OPTICAL TECHNOLOGY DEVELOPMENTS IN BIOMEDICINE:
HISTORY, CURRENT AND FUTURE

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Abstract

Biomedical optics is a rapidly emerging field for medical imaging and diagnostics. This paper reviews several biomedical optical technologies that have been developed and translated for either clinical or pre-clinical applications. Specifically, we focus on the following technologies: 1) near-infrared spectroscopy and tomography, 2) optical coherence tomography, 3) fluorescence spectroscopy and imaging, and 4) optical molecular imaging. There representative biomedical applications are also discussed here.
1. Introduction

Optical technologies are currently emerging as promising tools for medical imaging and diagnostics. Optics has several advantages, including non-ionizing radiation, low-cost, portable, and high molecular and biochemical specificity. These advantages enable functional imaging using light and open up new opportunities for light-based applications in clinical medicine.

This paper reviews several optical technologies that have been developed and translated for either clinical or pre-clinical applications. Specifically, we focus on the following technologies: 1) near-infrared spectroscopy and tomography, 2) optical coherence tomography, 3) fluorescence spectroscopy and imaging, and 4) optical molecular imaging. There representative biomedical applications are also discussed here.

2. Review of Optical Technology

2.1 Near-infrared (NIR) Spectroscopy and Tomography

Historical Perspective and Technology Development

Biological molecules have unique absorption spectra against a range of light wavelengths, thus can be detected with accurate concentration by spectroscopy. The spectroscopy system was pioneered in the Cambridge University [1, 2]. However, biological sample is usually opaque and therefore the light absorption spectra would be disrupted due to scattering. In 1950, Britton Chance invented “double beam spectrometer” using two wavelengths in the visible region with a small spectral interval to eliminate the effect of scattering [3]. This
double beam concept was adapted to the optical spectroscopy used for biological systems even up to date for medical use.

Pulse oximeter may be the first use of optics for human in vivo, which utilizes near-infrared light to monitor arterial hemoglobin oxygen saturation. It was first made by Takuo Aoyagi in a Japanese company, Nihon Kouden in 1972 [4]. Because arterial pulse induces changes in arterial blood volume between systolic and diastolic heart contraction, light intensity difference between these two conditions is only caused by arterial blood. Thus arterial blood oxygen saturation can be quantified with a simple linear equation [5], ignoring scattering effects of tissue. This concept is similar to the “double beam spectroscopy” dated back to 1940s, when Glenn Milliken tried to observe differences of light transmitting intensity through human tissues using green and red color filters to measure oxygenation in human tissue [6].

The first demonstration of NIR light on human tissue in vivo was reported by Franz Jobsis in 1977 [7]. Jobsis demonstrated that NIR light can carry information of not only hemoglobin but mitochondrial chromophore, cytochrome a,a3 in the neonatal brain measured non-invasively. Since then, many papers were published along the line of proving tissue oxygenation and mitochondrial redox states by means of those hemoglobin and cytochrome a,a3 signals in the NIR region in many animal models and human tissues. Many researchers use continuous-wave (CW) technology as the system is simple, low-cost, and robust. Figure 1 shows a representative CW near-infrared (NIR) imaging system [8] with three-wavelength light emitting diode (LED) at 760 nm, 805 nm, and 850 nm, and 8 silicon photodiode detectors. Many companies such as Somanetics have commercialized this technology to measure tissue hemoglobin saturation. Since only intensity attenuation is measured with CW system, it is difficult to separate scattering coefficient from absorption coefficient in the tissue.
Figure 1
A photograph of the whole apparatus (a) illustrates the handheld puck or probe, the coupling to the circuit box which contains the drivers for the LED, the amplifiers for the detectors, the digitally controlled gain adjustment amplifier, the electronic switch which decodes the light pulses and stores the information in a memory capacitor, the second set of switches which sample the memory capacitor at a rate compatible with the computer analog-to-digital converter (ADC), and (b) Handheld puck. From Ref. [8], with permission.

Time-Resolved Spectroscopy (TRS) technology gave a solution for this problem of absolute quantification of chemical concentrations in the turbid media such as the in vivo human investigation in 1988-1989 by B. Chance [9, 10]. TRS machines are commercially available then in 1993 by Hamamatsu, and it has been made for many applications [11, 12]. Alternatively frequency-domain (FD) NIR spectroscopy (NIRS) can be also used for quantization [13-15]. FD technology is available commercially by ISS, Inc. These two technologies have been used for obtaining more accurate information from the turbid tissue, namely absorption and reduced scattering coefficients.

NIRS can be extended to imaging mode by using multiple source-detector channels. One way to form an image is using back-projection and interpolation algorithms. This approach, sometimes referred to as diffuse optical topography, can provide a quick and good estimate of 2D spatial distribution of the optical properties of interested. The drawback of this relatively simpler approach is that the tissue optical properties are not reconstructed with good accuracy, and the spatial resolution is lower [16]. Another approach is to perform 3D
tomographic reconstruction, therefore, is referred to as diffuse optical tomography (DOT). In principle, DOT is similar to other tomographic schemes such as X-ray computerized tomography, and involves image reconstruction by solving the inverse problem [17]. DOT can accurately reconstructed the spatially-resolved changes in optical properties in tissue.

**Clinical Applications of NIR**

NIR diffuse optical spectroscopy and imaging techniques have been applied to numerous clinical applications. Here we focus on three main application areas, namely: 1) breast cancer detection and characterization; 2) functional brain imaging; and 3) imaging the skeletal muscles.

**Breast Imaging**

Breast cancer is the most commonly diagnosed cancer among women in the United States and worldwide. Early detection through mammography and clinical breast exams is essential for effective breast cancer screening. For women between the ages of 50-69, regular mammograms can reduce the chance of death from breast cancer by approximately 30% [18]. X-ray mammography may miss up to 25% of breast tumors in women in their 40s, and about 10% of women over age 50. Other imaging techniques, such as magnetic resonance imaging (MRI) and ultrasound (US), have been developed for breast cancer detection and staging without using X-rays [19, 20]. In general, mammography, MRI and US provide mostly anatomic information, rather than quantitative tissue function and composition [21]. Positron Emission Tomography (PET) could provide the metabolic information, but requires the injection of exogenous radionuclides [22].
Compared with those modalities, NIR diffuse optical imaging has its own merits of non-ionizing, economic and biochemical specificity. The use of light in breast cancer detection dates back to the 1920s [23]. In the past two decades, with the development of advanced light sources and detector, as well as modeling of light propagation in tissue, the application of diffuse optical imaging for breast imaging (often referred to as “optical mammography”) have been developing rapidly. The development of tumor is generally associated with increased vascularization (also called “angiogenesis”) [24, 25] and lower oxygen partial pressure (pO$_2$) [26, 27]. NIR light is sensitive to intrinsic chromophores such as oxygenated and deoxygenated hemoglobin (HbO$_2$ and Hb), water (H$_2$O) and lipids [28, 29]. Therefore, NIR diffuse optical spectroscopy and imaging can provide sensitive and specific physiological information for breast cancer diagnosis [30, 31]. Nioka et al introduced endogenous contrast NIR imaging of the human breast in 1994 [32]. Blood volume and oxygen saturation are two important parameters. Studies indicate that there are two to four folds of contrast between normal and tumor regions for blood volume, and oxygen saturation in the tumor is generally lower than normal [21, 29, 31]. There exist variations in normal breast tissue optical properties. For example, Durduran et al reported the averaged blood volume and oxygen saturation on healthy female breast tissues are $34 \pm 9 \text{ } \mu \text{M}$ and $68 \pm 8 \%$, respectively [33]. These baseline values are important to assess the potential contrasts available for diffuse optical imaging to discriminate healthy and diseased tissues. The scattering properties of tissue also contain important information for lesion diagnosis. The scattering coefficients are related to the tissue structure properties and the concentration or size of organelles [34].

The clinical niche for NIR diffuse optical spectroscopy and imaging in breast cancer are tumor detection in pre-menopausal women and monitoring neoadjuvant chemotherapy [35-37]. CW systems are relatively inexpensive and compact. It can be interfaced with a
handheld probe to image the breast. Using the handheld puck shown in Figure 1, Chance et al reported a 6 year, two-site study on 116 patients of whom 44 patients had confirmed malignancy by biopsy and histopathology [8]. The absorbance increments of the cancerous regions are referred to the mirror image location on the contra-lateral breast. This technique was able to distinguish cancer from non-cancer breasts with 96% sensitivity and 93% specificity. In another pilot clinical trial of 48 patients, Xu et al used a portable handheld NIR imaging device, “P-Scan”, a pre-commercial prototype of ViOptix Inc. (Fremont, CA, USA), to image suspicious breast lesions identified on diagnostic clinical ultrasound (US) [38]. An external mechanical compression was applied to breast tissue to dynamically record the oxygen saturation and hemoglobin concentration.

Indocyanine Green (ICG) is an organic dye that has been approved by US Food and Drug Administration (FDA) for clinical use. ICG can be used to enhance the tumor-to-normal contrast to aid in the detection of lesions in the breast, as first demonstrated by Nioka et al [39]. Figure 2 shows an example of CW DOT system for imaging the uptake of ICG by breast tumors [40]. This instrument employs a circular configuration with 16 sources and 16 detectors (Figure 2A) to collect light in parallel on the surface of the tumor-bearing breast. ICG was injected by bolus, and the absorption changes induced by ICG were recorded dynamically (Figure 2B). DOT successfully localized the tumor regions, which was in good agreement with a priori information (Figure 2C). A two-compartment model composed of plasma and extracellular-extravascular space was used to analyze the pharmacokinetics of ICG. Moreover, different dynamical features were observed for different pathologies. The malignant cases exhibited slower rate constants (uptake and outflow) compared to healthy tissue. Further studies enabled direct forming of the pharmacokinetics-rate image by DOT, and found statistically different rates from the tumor region compared to those outside the tumor region [41]. These results demonstrated that in vivo pharmacokinetics of ICG in breast
tumors could be a useful diagnostic tool for differentiation of benign and malignant pathologies. Schmitz et al developed a more sophisticated CW system for bi-lateral breast imaging, DYnamic Near-Infrared Optical Tomography (DYNOT) system (NIRx Medical Technologies) [42]. This system enables simultaneous imaging of hemodynamics within both breasts.

![Figure 2](image)

**Figure 2**  
(A) CW imager configuration. The sources are sequentially shining upon the breast and so the configuration is equivalent to a fan-beam configuration. (B) The dashed curve represents the intensity drop associated to the ICG-uptake for source 6 and detector 3 (not capturing mass area). (C) Differential absorption reconstruction for the time selected in (B). From Ref. [40], with permission.

Frequency-domain measurement can quantify the concentration of chromophores. Culver et al developed a hybrid CW/frequency-domain breast imaging system which combines the benefits of high-speed and low-cost of CW techniques with more accurate quantitative nature of frequency-domain techniques [43]. Using this system, Choe et al [44] found that malignant cancers (n=41) showed significantly higher total hemoglobin, oxy-hemoglobin concentration, and scattering compared to normal tissue. Benign tumors (n=10) did not exhibit statistical significance in the tumor-to-normal ratios of any parameter. These results demonstrate that benign and malignant lesions can be distinguished by quantitative three-dimensional DOT. Such a system also has been applied to monitor neoadjuvant
chemotherapy [45]. DOT revealed tumor shrinkage and decrease in relative total hemoglobin concentration during the course of chemotherapy, therefore demonstrated the potential for monitoring physiological parameters of breast lesions during chemotherapy. Tromberg’s group at UC Irvine also developed a handheld NIR diffuse optical spectroscopic imaging (DOSI) system for breast cancer detection and monitoring neoadjuvant chemotherapy [37].

Time-domain DOT systems have been also developed for breast imaging. Ntziachristos et al developed a time-domain imaging system using time-correlated single photon counting (TCSPC) technique, and demonstrated concurrent MRI and DOT imaging of breast after contrast enhancement using ICG [11]. Other prototype instruments have been developed by groups at Politecnico di Milano, Italy [46-48] and Physikalisch-Technische-Bundesanstalt of Berlin, Germany [12, 49-51], as part of Optimamm, a consortium funded by the European Union, and have acquired data from more than 300 clinical cases. They reported successful identification of 80%-85% mammographically identified lesions. A prototype time-domain DOT breast imaging system has been developed by Advanced Research Technologies (ART, Canada) [52]. Initial results suggested that optical imaging can discriminate benign and malignant tumors, therefore, held great clinical promise for breast cancer imaging.

One of the recent trends in NIR DOT is to combine with other imaging modalities such as X-ray CT, MRI or US, which can provide high spatial resolution map of tissue structures. Those maps can be used as a priori information to improve the reconstruction of DOT images [53, 54]. Representative multi-modality breast imaging systems include the combined DOT and X-ray mammography system developed at the Massachusetts General Hospital [55, 56], the combined DOT and MRI multi-modal imaging system developed at Dartmouth College [53, 57], and the combined ultrasound and frequency-domain diffuse optical imaging/tomography systems [58-61].
**Brain Imaging**

Since Seiji Ogawa’s discovery that deoxyhemoglobin signal changes in NMR can detect brain cognition in early 1990s [62], researchers are interested in using NIR light to detect brain function [63-65]. NIRS can quantify the concentration of both Hb and HbO₂, thereby revealing the blood volume and oxygenation saturation changes associated with brain functions.

NIR diffuse optical imaging (DOI) has found widespread applications in clinical settings [66, 67]. One major research area is to understand how the brain functions. DOI offers unique capability to non-invasively monitor the functional activations in vivo without disturbing the organ. Various applications such as visual responses [68, 69], somatosensory responses [70], auditory responses [71], language stimuli [72, 73], and problem solving [74] have been explored. Another important area for DOI brain imaging is to diagnose and monitor the diseases such as stroke [75, 76], epilepsy [77], brain injury [78], and post-traumatic stress disorder [79]. Optical techniques are well-suited for early detection of hemorrhage [80], and discrimination between ischemic and hemorrhagic stroke leading to a better management of the patient treatment [81].

Commercial CW brain imaging systems have been developed by Hitachi Medical Corporation (Tokyo, Japan) [82, 83]. This optical topography system (ETG-100) uses 8 laser diodes at 780 nm and another 8 at 830 nm. 8 avalanche photodiodes (APDs) are used to detect the signals. Multiple channels are encoded by different frequencies from 1 to 6.5 kHz. The Hitachi system has been applied to investigate healthy brain functions such as language development [72], the emotional response to music [84], cognitive functions [73, 85], and brain development in newborn infants [86, 87], as well as pathological conditions such as
epilepsy [77], post-traumatic stress disorder [79], and cognitive function study in patients with motor neuron disease [88]. The clinical applications of the Hitachi system have been quite successful despite using a simple CW system and relatively simple image reconstruction method. Other companies such as Shimadzu Corporation (Japan) also developed optical topography system from brain imaging [89].

Franceschini et al reported the development of the CW imaging system (CW4 and CW5) at the Massachusetts General Hospital [90, 91]. The newer system (CW5) employs 32 sources and 32 detectors to cover most of the areas in the adult head, which enables simultaneous collection of optical signals from prefrontal, sensorimotor, and visual cortices in both hemispheres. Using this system, they investigated the spatiotemporal patterns of physiological signals during rest. This information will help to understand the physiological signals and develop signal processing algorithms to distinguish them from the functional activation signals [91]. White et al applied a high-performance, high-density CW DOT system [69] to map resting-state networks in the human brain [92], which enables studies of the functional connectivity of different cortical regions. These studies demonstrated that high-density DOT has become a practical and powerful tool for mapping function in the human cortex.

ISS Inc. has developed a commercial frequency-domain brain imaging system (Imagent™). Frequency-domain DOI has been shown to measure the hemodynamic (slow) signals [93, 94] and neuronal (fast) signals [95, 96]. Especially, the fast signals are thought to be associated with the neuronal scattering changes, which will induce phase delay in the modulated diffuse photons. Therefore frequency-domain imaging system is required to measure the phase delay which indicates the event-related optical signals (EROS). Time-domain imaging systems have been actively developed for brain imaging, especially for tomographic imaging of whole brain. Hintz et al reported the early development of time-
domain optical tomographic system for neonatal brain imaging by reconstructing measurements of mean photon flight time [97]. More recently, the group at University College London (UCL) has developed a 32-channel time-resolved optical imaging system [98] for 3D neonatal brain imaging. This system has been successfully used to image the brain of a premature infant with a cerebral hemorrhage [99] and monitor the blood volume and oxygenation changes in the newborn infant brain during ventilation [100].

Muscle Imaging

Non-invasive monitoring of muscle tissues using light can be dated back to the 1930s by Millikan [101]. Since then, optical imaging of muscles has received steadily increased interests. Optical methods can probe hemoglobin, myoglobin, blood flow, and metabolism, therefore provide an ideal means for monitoring muscle functions under different physiological or pathological conditions [102-106].

Using CW imaging system similar to that shown in Figure 1, Lin et al demonstrated fast imaging of blood volume and oxygenation changes in peroneus and gastrocnemius muscles during exercise [107]. This 8-channel imager can differentiate the regions corresponding to extensors and flexors since they show different responses during exercise. Using a higher density (200-channel) CW imager which covers 45 cm × 15 cm² area, Hamaoka et al recently showed DOI of quadriceps muscles before, during, and after exercise (see Figure 3) [108]. These results demonstrated the spatial differences within muscles during exercise and recovery, which would be an important tool to study muscular physiology.

Maris et al used frequency-domain NIR optical topography system to map the differences in the hemoglobin concentration in finger extensor muscle during exercise [109]. Yu et al later demonstrated a hybrid frequency-domain diffuse reflectance spectroscopy
(DRS) and diffuse correlation spectroscopy (DCS) system for simultaneous monitoring of muscle hemodynamics and blood flow [110]. DRS can quantify the total hemoglobin concentration and oxygenation saturation, while DCS, an emerging extension of diffuse optical imaging techniques [111, 112], quantifies the relative blood flow in deep tissues. Together, this hybrid technique provides a method for evaluation of muscle microcirculation and metabolism in vivo.

![Figure 3](image)

**Figure 3**

NIR DOI images from the quadriceps muscles before, during, and after intermittent isometric knee-extension exercise. The top left image indicates the approximate location of specific muscles. Contractions 1, 3, and 5 indicate images obtained during a series of 3-s duration contractions at 50% of maximum voluntary contraction (MVC), with one second rest in between. The 15-s contraction shows data at the end of a continuous 15-s MVC. The recovery image was obtained 10 s after the last contraction. These images demonstrate the spatial differences seen within muscles during exercise and recovery. \( \Delta \text{O}_2\text{Hb} \): oxygenated hemoglobin and myoglobin; \( \Delta \text{HHb} \): deoxygenated hemoglobin and myoglobin; \( \Delta \text{tHb} \): total hemoglobin and myoglobin. From Ref. [108], with permission.

Time-domain methods have also been extended into muscle imaging. Hillman et al have used the 32-channel time-domain DOT system to measure the absorption changes of
human forearm in response to finger flexion exercise [113]. Zhao et al also developed a time-resolved (TR) DOT system and demonstrated the capability of imaging the forearm during hand-gripping test [114]. The group at Milan has developed a compact and fast multi-channel TR DOI system to image the calf muscle oxygenation and hemoglobin content during dynamic plantar flexion exercise [115]. These results strengthen the role of DOI as a powerful tool for investigating the spatial and temporal features of muscle physiology. These above results clearly demonstrated that NIR diffuse optical imaging has been widely used for imaging muscle functions and diseases. Although it is difficult to decouple the relative contributions from hemoglobin and myoglobin in the muscle [116, 117], DOI will continue to play an important role in imaging muscle functions for athletic training [118, 119] and disease diagnostics [120].

2.2 Optical Coherence Tomography (OCT)

Principle and Instrumentation of OCT

OCT is an emerging medical imaging technology which enables cross-sectional imaging of tissue microstructure in situ and in real time [121]. OCT can achieve 1-10 µm resolutions and 1-2 mm penetration depths, approaching those of standard excisional biopsy and histopathology, but without the need to remove and process tissue specimens [122]. OCT is analogous to ultrasound imaging, except that imaging is performed by measuring the echo time delay and intensity of back-reflected or backscattered light rather than sound. OCT imaging can be performed fiber-optically using delivery devices such as handheld probes, endoscopes, catheters, laparoscopes, and needles which enable non-invasive or minimally invasive internal body imaging [123].
OCT measurements are performed using a Michelson interferometer with a low coherence length light source. One arm of the interferometer illuminates the light on the tissue and collects the backscattered light (typically referred to as “sample arm”). Another arm of the interferometer has a reference path delay which is scanned as a function of time (typically referred to as “reference arm”). Optical interference between the light from the sample and reference arms occurs only when the optical delays match to within the coherence length of the light source. This technique is generally referred to as “time-domain OCT”. Alternatively, OCT interference signals can be detected in frequency or Fourier domain. In Fourier-domain OCT, the reference mirror position is fixed, and echoes of light are obtained by Fourier transforming the interference spectrum. These techniques are somewhat analogous to Fourier transform spectroscopy and have a significant sensitivity and speed advantage compared to time-domain OCT because they measure the optical echo signals from different depths along the entire axial scan simultaneously rather than sequentially. Fourier-domain detection enables 10-100 folds improvement in detection sensitivity and speed over the time-domain configuration [124-126]. These advances not only greatly improve the performance of OCT, but enables three-dimensional OCT (3D-OCT) imaging in vivo. Fourier-domain OCT can be performed using two complementary techniques, known as spectral / Fourier domain OCT and swept source / Fourier domain OCT (SS-OCT, also known as Optical Frequency Domain Imaging, OFDI). Three-dimensional imaging of biological tissue in vivo enabled by Fourier-domain OCT promises to have a powerful impact in disease diagnosis [127, 128].

To image internal organs, miniaturized catheter/endoscope imaging devices have been developed for intraluminal and intravascular imaging [129, 130]. Other imaging devices such as laparoscopes [131, 132] and needle imaging device have been developed to enable solid organ imaging [133, 134]. Nowadays, various OCT imaging probes have been developed for
different clinical applications. Development of such devices facilitates the translation of OCT to clinical applications and allows clinicians to use the enhanced imaging capabilities of this technique to benefit the patients.

Clinical Applications of OCT

OCT was first demonstrated in 1991 [121]. Since then, OCT has rapidly developed as a non-invasive biomedical imaging modality that enables cross-sectional visualization of tissue microstructures in vivo [135-138]. The resolution of OCT is one to two orders of magnitude higher than conventional ultrasound, approaching that of histopathology, thereby allowing architectural morphology to be visualized in situ and in real time. OCT enables imaging of structures in which biopsy would be hazardous or impossible, and promise to reduce the sampling errors associated with excisional biopsy. OCT has been increasingly translated from bench to various clinical applications including ophthalmology [139-145], cardiology [146-149], gastroenterology [150-156], dermatology [157], dentistry [158, 159], urology [160-163], gynecology [164-166], among others. The most developed clinical OCT applications are those focusing on ophthalmic, cardiovascular, and oncologic imaging.

Ophthalmic Imaging

OCT was first applied for imaging in the eye [167, 168]. To date, OCT has made the largest and most significant clinical impact in ophthalmology. OCT provides cross-sectional views of eye with high resolution, thereby allowing detailed structures to be discerned. The non-contact and non-invasive nature of OCT enables a new way to image the structures in the anterior eye and retina, and reveal the information not available through standard
ophthalmoscopes [169, 170]. Ophthalmic OCT was first commercialized by Carl Zeiss Meditec, Inc., and is now considered superior to the current standard of care for the evaluation of many retinal diseases. Over 6000 units of Status OCT™ system has been sold to date, and it is estimated that more than 37,000 OCT scans are performed daily in the U.S. With the development of high-speed OCT using spectral/Fourier domain methods, several companies have introduced new OCT instruments into the ophthalmic market in the past few years. The increased availability of high performance OCT will enable a wide range of clinical studies.

The high axial resolution of OCT makes it an ideal imaging modality for the evaluation of fine features such as intra-retinal layers and the vitreal-retinal interface. OCT has been demonstrated for the detection and monitoring of a variety of macular diseases [171], including macular edema [172-175], macular holes [142, 143, 176], central serous chorioretinopathy [177], age-related macular degeneration and choroidal neovascularization [178]. OCT can also image and quantify the retinal nerve fiber layer thickness, which is a predictor for early glaucoma. Quantitative nerve fiber layer thickness has been measured with OCT, and correlated with the optic nerve structure and function [179-184].

The increasing impact in clinical medicine promotes the rapid development in OCT imaging technologies, which dramatically enhance the imaging performance and clinical utilities of OCT. A comprehensive review of the state-of-the-art ophthalmic OCT has been provided elsewhere [185]. Here we provide a concise overview of these technology advances and their translation into ophthalmic applications.

One of the key parameters in OCT imaging is axial resolution. This is of particular interests in retinal imaging owing to the layered structures of the retina. Enhanced axial resolution enables better visualization of the intraretinal structural details and more accurate
diagnosis of diseases. The axial resolution of OCT is inverse proportional to the bandwidth of the low-coherence light source. Therefore, increasing the bandwidth of the light source enables the improvement in axial resolution [186, 187]. Ultrahigh resolution (UHR) OCT achieves superior axial image resolutions of 2-3 μm as compared to ~ 10 μm in standard OCT systems using superluminescent diode (SLD), thereby enabling the visualization of intraretinal structures [188]. UHR OCT is a key technology advance towards achieving non-invasive optical biopsy of the human retina. UHR OCT technology has been investigated in clinical settings to assess its clinical utility. Cross-sectional studies in ~1,000 eyes with different pathologies demonstrated unprecedented visualization of all major intraretinal layers especially the photoreceptor layer [145, 189-193]. All intraretinal layers, especially the inner and outer photoreceptor segment, are significantly better visualized by UHR OCT (see Figure 4). These studies demonstrated visualization of photoreceptor layer impairment in macular pathologies such as macular holes, central serous chorioretinopathy, age related macular degeneration, foveomacular dystrophies, Stargardt’s dystrophy and retinitis pigmentosa [185]. Therefore, UHR OCT holds strong potential to enhance early diagnosis by detecting subtle morphological changes in a wide range of retinal diseases.
Transverse resolution is also an important imaging parameter. The transverse resolution is determined by the numerical aperture (NA) of the focusing lens. For ophthalmic retinal imaging, the cornea and lens act as the “imaging objective”. In practice, ocular aberrations limit the minimum focused spot size on the retina. Adaptive optics (AO) is a promising approach to correct ocular aberrations in order to decrease the spot size on the retina to improve the OCT transverse resolution [194, 195]. Combining AO with UHR OCT provides isotropic high resolution in the 3D dataset, thereby enabling cellular resolution retinal imaging [196-199]. Ultrahigh transverse resolution imaging can also be achieved by using high NA objectives (also called optical coherence microscopy - OCM) [200]. Parallel detection using full-field OCM has been demonstrated on cellular-level imaging of cornea and retina tissues [201, 202]. This technology has shown strong promises in clinical translation for in vivo ophthalmic imaging.

Imaging speed is another critical parameter in clinical OCT imaging. High speed imaging not only reduces the motion artifacts, but enables comprehensive visualization of the three-dimensional structures. Fourier-domain OCT is a key enabling technology which dramatically increase the OCT imaging speed for three-dimensional (3D) imaging in vivo. Spectral OCT, commonly operates at 800 nm, has been rapidly developed and translated into retinal imaging. The first demonstration of retinal imaging using spectral OCT was
performed in 2002 [203], and high-speed video-rate imaging was achieved 2003 [204, 205]. Three-dimensional ultrahigh-resolution ophthalmic imaging in vivo has been demonstrated on numerous clinical studies [206-210]. Three-dimensional OCT provides quantitative measurement of intraretinal layers for early diagnosis of diseases such as glaucoma or diabetic retinopathy, and enables assessment of disease progression or response to therapy. Figure 5 shows an example of topographic information of optical disk similar to that obtained by scanning laser tomography system. In addition, spectral OCT systems working at 1000 nm range promise to improve the penetration depth for better visualization of choroidal tissues [211, 212].

![Figure 5](image)

**Figure 5**

*Topography using three-dimensional, ultrahigh-resolution OCT. Quantitative topography using ultrahigh-resolution 3D-OCT (A) of a normal human optic disk, as compared to those performed by Heidelberg retinal tomography (Heidelberg Engineering, Germany) (B). From Ref. [213], with permission.*

Other technological advancements have been applied to ophthalmic imaging as well. Doppler OCT [214-216] enables measuring of blood flow velocity in the tissue.
Spectral/Fourier domain OCT with Doppler flow imaging has been demonstrated [217-219]. Three-dimensional Doppler OCT enables visualization of 3D retinal vasculature (also called optical coherence angiography [220, 221]). Another functional OCT method, polarization-sensitive (PS) OCT [222], enables detection of depth-resolved tissue birefringence and scattering properties. PS-OCT has been used for imaging the retinal nerve fiber layer (RNFL) changes in glaucoma patients [223]. This method has been applied to image both the anterior and posterior eye imaging [224-227]. Recently, significant advances have been made on detecting OCT scattering signals due to functional responses of the retina (also called optophysiology) [228]. Functional responses have been observed in vivo on human subjects [229]. This technique could detect functional impairment before morphological changes. Lastly, multi-modal technology combining OCT with other optical imaging modalities, such as scanning laser ophthalmoscopy (SLO) and fluorescence angiography promises to integrate the information from different methods and enhance the diagnostic capability [230].

Cardiovascular Imaging

Another major area for OCT clinical application is cardiology. The potential of OCT for cardiology imaging has been extensively investigated over the past decade [146-148, 231-239]. Compared to intravascular ultrasound (IVUS), OCT has an order of magnitude higher resolution, therefore enables visualization of fine structures in the luminal wall (such as the differentiation of intimal, medial, and adventitial layers). Several technological advances, including catheter-based imaging probes and high-speed OCT imaging, have enabled the translation of OCT cardiovascular imaging from the bench to the bedside. Development of small-diameter fiber catheters facilitates the manual feeding of the imaging catheters through the vasculature. In intravascular OCT imaging, blood will significantly attenuate the signal,
which can be alleviated by saline flush or temporary vascular occlusion with balloon. To scan a long segment of an artery within a short interval of saline flush or blood occlusion, high-speed imaging is critical. Recent advancement in high-speed Fourier domain OCT provides exciting new avenue to diagnose vascular diseases and guide the intravascular interventions in real time [240].

OCT has unique ability to visualize atherosclerotic lesions in microscopic details, and in particular, to detect the vulnerable plaques which has high risks of rupture [148, 234, 236, 241, 242]. OCT can distinguish the characteristic morphology of vulnerable plaques including a thin fibrous cap and a large lipid pool, and is able to quantify the increase of macrophage activity [137, 241]. The potential of OCT imaging of vulnerable plaques was first investigated on in vitro studies and demonstrated the capability of OCT in identifying clinically relevant architectural morphology including fibrous caps, lipid-laden pools, and calcifications [243]. The specific features of different types of atherosclerotic plaques (fibrous, lipid, and calcified) can be clearly identified with histological correlation [244]. OCT images of fibrous plaques were characterized by homogeneous, signal-rich regions; fibrocalcific plaques by well-delineated, signal-poor regions with sharp borders; and lipid-rich plaques by signal-poor regions with diffuse borders. The detection sensitivity and specificity for different types of plaques are: 71-79% and 97-98% for fibrous plaques; 95-96% and 97% for fibrocalcific plaques; and 90-94% and 90-92% for lipid-rich plaques. These results represent an important step in validating intravascular OCT imaging and provide a basis for interpretation of intracoronary OCT images obtained from patients. The first in vivo coronary imaging study in humans was performed using a 1-mm-diameter (3.0 F) catheter, demonstrating the ability of intravascular OCT to visualize the coronary plaques [234], followed by subsequent study to characterize different coronary atherosclerotic plaques in vivo [149]. Intravascular OCT imaging has been investigated extensively for in vivo
diagnostics of vulnerable plaques [245-248]. Identifying vulnerable plaques in situ could potentially allow cardiologists to develop therapeutic strategies, predict the vulnerability of plaques, and monitor structural changes after intervention.

OCT has also been demonstrated to be able to quantify the activated macrophage content in vivo [148]. It has been demonstrated that there was a high degree of positive correlation between OCT measurements and histological measurements of fibrous cap macrophage density. The unique capabilities of OCT for fibrous cap characterization suggest that this technology could be well suited for identifying vulnerable plaques in patients. Other quantitative analyses including the OCT signal attenuation and layer thickness changes has been undertaken for tissue characterization of OCT imaging of coronary arteries [249-251]. Recent study using both the attenuation and backscattering properties showed enhanced contrast and better tissue characterization [251]. These findings hold strong potentials for future computer-aided diagnosis of atherosclerotic plaques and better detection of thin-cap fibroatheroma (TCFA) [251].

Another potential application for intravascular OCT is to monitor therapeutic interventions, such as stent deployment [147, 252-257]. OCT provides a clear view of the stent struts and their positions relative to the wall, thereby giving surgeons a real-time assessment of stent apposition, tissue prolapse, and wall dissections. It has been demonstrated that OCT can be used to visualize stent integrity, neointimal formation, and neovascularization [238, 253]. Recent developments in drug-eluting stents promises to prevent the in-stent restenosis. Intravascular OCT imaging enables evaluation of neointimal coverage and earlier detection of excessive regrowth of the inner layer of vessel [258, 259].

In addition to high-resolution and high-speed imaging of morphology, OCT can also provide functional information for improved understanding and assessment of disease. For
example, polarization-sensitive (PS) OCT has shown the potential to image intrinsic tissue birefringence changes, which can be used to differentiate fibrous and calcified plaques [260-262]. PS-OCT will significantly improve our ability to evaluate plaque stability in patients. Optical coherence elastography (OCE), another functional extension of OCT for imaging tissue biomechanical properties, has been investigated to evaluate the mechanical properties of arterial walls and plaques [263, 264]. Furthermore, a dual-modality device that combines the anatomical imaging capabilities of OCT with the functional capabilities of laser-induced fluorescence (LIF) spectroscopy has been applied for imaging normal and atherosclerotic portions of aorta wall [265]. Such dual-modal approach is desirable in cardiac functional imaging [266], where the structural heterogeneities influence the arrhythmia induction, stabilization, and termination. The comprehensive information offered by both modalities promises to characterize the patterns of functional signals and their correlation with structures in great details, thereby providing new insights into the structure-function relationships.

**Oncology Imaging**

Cancer imaging is also a promising area of OCT application. Many cancers arise from the epithelial layers, and demonstrate disruption of normal architectural morphology of tissues. The resolution and imaging field-of-view of OCT is approaching those of standard biopsy and histopathology, therefore OCT represents a potential method for “optical biopsy” of the tissue in situ, which can guide the excision biopsy to improve the sampling accuracy. OCT has shown promises in detecting structural alterations associated with malignancies including those arising in the breast [267-271], bladder [163, 272-274], brain [275-277], gastrointestinal [154, 155, 278, 279], respiratory [280] and reproductive [281] tracts, skin [282], larynx [283, 284], and oral cavity [285, 286].
Development of endoscopic OCT greatly facilitates imaging of cancers raised from internal organs in situ. OCT has been demonstrated for detection of specialized intestinal metaplasia in Barrett's esophagus [150-152, 287] and transmural inflammation in inflammatory bowel disease (IBD) patients [288]. Recently, OCT has shown the promise for detection of high-grade dysplasia in Barrett's esophagus. Evans et al reported 83% sensitivity and 75% specificity for detecting high-grade dysplasia and intramucosal carcinoma with blinded scoring of OCT images from 55 patients using a numeric scoring system based on the surface maturation and glandular architecture [155]. Isenberg et al reported 68% sensitivity and 82% specificity, with 78% accuracy for detection of dysplasia from 33 patients with Barrett's esophagus [154]. Using ultrahigh-resolution (UHR) endoscopic OCT imaging with 4 μm axial resolution, Chen et al demonstrated in vivo clinical imaging in a cross-sectional study of 50 patients [156]. Real-time endoscopic OCT imaging was performed using a 1.8 mm diameter OCT catheter probe introduced into the accessory channel of a standard endoscope. Figure 6 A-B shows a representative UHR OCT image of normal esophagus which exhibits a characteristic layered architecture and its corresponding histology. C-D shows a representative UHR OCT image of Barrett's esophagus and the associated histology. The layered architecture in normal esophagus is replaced by glandular structures. Low-backscattering Barrett's glands are frequently observed within the mucosa, with interlaced regions of high-backscattering connective tissue corresponding to the lamina propria. E-F shows an example UHR OCT image of high-grade dysplasia and the corresponding histology. OCT images of high-grade dysplasia are characterized by irregular, distorted, and cribriform or villiform glandular architecture and are more heterogeneous than those of non-dysplastic Barrett's epithelium. G-H shows an example UHR OCT image of adenocarcinoma and the representative histology. Irregular infiltrating glandular morphology can be visualized with OCT. Ultrahigh resolution OCT images showed good correlation of architectural
morphology with histological findings. Enhanced image resolution and reduced speckle size enable ultrahigh resolution OCT to visualize tissue architectural heterogeneity more clearly than standard resolution OCT. Future clinical studies are needed to investigate the role of ultrahigh resolution OCT in detection of early neoplastic lesions.

Figure 6

*In vivo ultrahigh resolution endoscopic OCT image (top row) and the corresponding histology (bottom row). A-B: normal esophagus; C-D: Barrett’s esophagus; E-F: High-grade dysplasia; G-H: Adenocarcinoma. From Ref. [156], with permission.*

Using Fourier-domain methods, endoscopic 3D-OCT imaging in vivo has recently been demonstrated [127, 289-291]. 3D OCT enables comprehensive assessment of early structural changes associated with diseases. With further technology refinement, higher resolution and imaging speed will be available in the near future. The unique capability of high-resolution imaging with large field of view promises to enable more complete characterization of tissue microscopic features and open new possibilities for improving identification of early neoplastic changes.

Bladder cancer is another promising candidate for endoscopic OCT imaging. Recently, a 32 patient study showed OCT has high detection accuracy for real-time imaging and staging of bladder cancer adjunct to white light cystoscopy (90% sensitivity and 89%
specificity for tumor confined to the mucosa, and 100% sensitivity and 90% specificity for muscle-invasive tumors) [274]. Another clinical study with 24 patients reported an overall sensitivity of 100%, specificity of 89%, and diagnostic accuracy of 92% for OCT imaging of superficial bladder transitional-cell carcinoma (TCC) [163]. Computer-aided recognition of bladder cancer using OCT and texture analysis is under investigation to improve the clinical utility of OCT [273].

OCT also holds promises for detection of cancers in the solid organ such as breast. In a pathological laboratory study, ultrahigh resolution OCT imaging of human breast specimens was performed in 119 freshly excised specimens from 35 women with 3.5 μm axial resolution [268]. Microstructures of normal breast parenchyma, including glands, lobules, and ducts, as well as stromal changes associated with infiltrating cancer were visible from OCT images. Furthermore, fibrocystic changes and benign fibroadenomas were identified. Imaging of ductal carcinoma in situ (DCIS) revealed microcalcifications. Figure 7 shows an example of OCT images of the human breast. Figure 7 A shows an OCT image of normal fibroadipose tissue with the corresponding histology shown in B. Fibrous stroma appears heterogeneous and highly scattering, whereas adipocytes appear low scattering, with individual, well-circumscribed scattering borders. Figure 7 C shows an OCT image of DCIS lesions in lobules. Tumor cells within lobules appear uniformly low scattering. Dilatation and architectural distortion of the lobules is visible. A microcalcification (C) within the lobules appears highly scattering with pronounced shadowing. D shows the corresponding histology. E-F shows an OCT image of infiltrating ductal carcinoma and the corresponding histology. Highly scattering regions corresponding to tongues of invasive cancer are visible. G-H shows an OCT image of a solid variant infiltrative lobular carcinoma with histology. Regions with densely infiltrating tumor cells appear low scattering and homogeneous, with isolated regions of entrapped fat. Using quantitative signal analyses including slope, standard deviation, and
spatial frequency, high sensitive tissue classification has been shown with normal and tumor tissues [269, 292]. A recent 37 patient study used OCT to image surgical margins of lumpectomy specimens, yielding a sensitivity of 100% and specificity of 82% [270]. These results clearly demonstrated that OCT is a strong candidate for future clinical adoption for image-guided interventions of breast cancer including guiding breast biopsy and intra-operative margin assessment in breast-conserving surgeries. The potential of OCT in detecting cancers in other solid organs such as prostate cancer [161] and kidney cancer [293, 294] are also under investigation.

**Figure 7**

Ultrahigh resolution OCT image (top row) and the corresponding histology (Hematoxylin-eosin stain, bottom row) of human breast tissues ex vivo. (A) OCT image of normal fibroadipose tissue. (B) Histologic specimen corresponding to OCT image. (C) OCT image of DCIS lesions in lobules. (D) Histologic specimen corresponding to OCT image. (E) OCT image of infiltrating ductal carcinoma. (F) Histologic specimen corresponding to OCT image. (G) OCT image of a solid variant infiltrative lobular carcinoma. (H) Histologic specimen corresponding to OCT image. From Ref. [268], with permission.
Multi-modal imaging combining OCT with other imaging techniques such as fluorescence has been actively investigated to improve the sensitivity and specificity for cancer detection. Previous studies combining OCT and laser-induced fluorescence (LIF) spectroscopy showed improved identification of tumor boundaries in the cervix [295, 296]. Tumlinson et al. developed a combined OCT and LIF imaging catheter for in vivo imaging of mouse colon [297, 298]. This miniaturized 2-mm-in-diameter catheter has been applied to longitudinally monitor disease progression in the mouse colon, and is able to identify colorectal adenomas in murine models [299]. In an ex vivo study of murine GI tracts, Hariri et al. showed that OCT and LIF provided complementary information for the detection of dysplasia and inflammatory bowel disease (IBD) of the intestines [300]. Pan et al. showed that fluorescence-guided endoscopic OCT could enhance the efficiency and sensitivity of early bladder cancer diagnosis [301]. In a rat model study, they demonstrated that the specificity of fluorescence detection of transitional cell carcinoma was significantly enhanced by fluorescence-guided OCT (53% vs. 93%), and the sensitivity of fluorescence detection also improved by combination with OCT (79% vs. 100%) [302]. A recent clinical study involving 138 volunteers and 10 patients with lung cancer has been performed to investigate the feasibility of OCT characterization of preneoplastic changes in the bronchial epithelium identified by autofluorescence bronchoscopy [280]. The preliminary data suggested that autofluorescence bronchoscopy-guided OCT imaging of bronchial lesions is promising for in vivo imaging of preneoplastic bronchial lesions.

Other Applications

In addition to ophthalmology, cardiology, and cancer imaging, which are the most developed fields in OCT, novel applications are constantly being explored. OCT has shown promises in
imaging cartilage diseases [303-306]. Dental OCT has also been developed extensively [159, 307-313]. Neurosurgical guidance using OCT represents another exciting new application [314]. Non-destructive evaluation of transplant kidney status using OCT [315, 316] is actively under investigation. Further research is needed to evaluate the role of OCT in a variety of clinical areas.

2.3 Fluorescent Techniques: Spectroscopy and Tomography

Principle and Instrumentation

Fluorescence techniques use fluorescence from either endogenous molecules (i.e., autofluorescence) or exogenous dyes to probe the biochemical and pathological status of the tissue [317, 318]. Typical fluorescence system illuminates the tissue with light, which excites fluorescent molecules (fluorophores) within the tissue. The emitted fluorescence light, typically at longer wavelengths than the excitation light, is collected and analyzed [319, 320]. Common endogenous fluorophores include connective tissues (collagen, elastin), cellular metabolism related coenzymes (reduced nicotinamide adenine dinucleotide (NADH), flavin adenine dinucleotide (FAD), and flavin mononucleotide (FMN)), by-products of heme biosynthesis (porphyrins), among others [321]. Several exogenous dyes also have been approved for clinical use, such as ICG [11, 40, 230, 322, 323], fluorescein [175, 324], 5-aminolevulinic acid (5-ALA) [325-327], among others. Table 1 lists the optical properties of some commonly-used fluorophores [328].

Table 1. Optical Properties of Representative Fluorophores.

| Fluorophore | Excitation (nm) | Emission (nm) |
|-------------|----------------|---------------|

80
There are three types of fluorescence systems: point-probe spectroscopy, wide-field imaging, and tomography. Point-probe spectroscopy system is designed to obtain the wavelength-dependent optical properties of tissue at a single spatial location. A typical point-probe spectroscopy instrument consists of a light source, a fiber-optic delivery probe containing both the excitation and collection fibers, and a spectrometer as the detector. The excitation fiber illuminates the tissue volume and the closely spaced detection fibers collect the scattered and fluorescence light. The detected fluorescence light (after long-pass emission filter) is dispersed in wavelength by a spectrograph, and the spectrum is recorded and analyzed. The separation between the illumination and detection fibers is usually on the order of millimeters, with the total diameter of the optical probe being small enough to pass through the accessory channel of a standard endoscope. The tissue sampled in point-probe spectroscopy system is usually restricted to a small area comparable to the extent of conventional excisional biopsy. Fluorescence spectroscopy systems can be extended into wide-field imaging mode. Wide-field techniques can image over a larger surface area. The light source provides a wide-field illumination on the tissue sample and a CCD camera is used as the detector to generate high-quality images [329]. A series of emission filters can be used to acquire wavelength dependent fluorescence signals. Wide-field planar imaging
typically does not contain depth-resolved information. In addition, the detected fluorescence intensities are non-linearly attenuated from different imaging depths [330-332], which could lead to a surface weighted images (in other words, signals are bias towards the surface lesions). In contrast, fluorescence tomography can reconstruct spatially-resolved fluorophore distribution, using similar approaches as diffuse optical tomography (DOT). The excitation light is illuminated from multiple source positions (through either fiber-optic coupling or scanning beam spot), and multiple detectors (such as CCD cameras) collect the fluorescence light propagating through different paths. Using image reconstruction similar to computerized tomography (CT) or DOT, the fluorophore distribution inside the tissue can be reconstructed.

**Clinical Applications**

One of the major clinical applications for fluorescence systems is early cancer detection. Several biochemical and morphologic factors may correlate with fluorescence signal changes in neoplastic lesions, including increased absorption of hemoglobin and loss of spectral contributions from submucosal connective tissues [333]. Abnormal thickening of epithelial tissue may cause attenuation of the excitation light, leading to further decreases in the fluorescence intensity. Therefore, the peak wavelength and intensity of the fluorescence spectra can be used to differentiate normal versus diseased tissues due to the changes in the concentration and distribution of metabolically related fluorophores and alterations of the tissue microstructures [317, 334, 335]. Here we focus on the applications in cancers in the gastrointestinal (GI) tracts, brain, and breast.

*Point-Probe Fluorescence Spectroscopy*
Point-probe fluorescence spectroscopy instruments can be integrated into a fiber-optic catheter device for endoscopic applications. Previous studies have shown that fluorescence spectroscopy can increase the detection rate of high-grade dysplasia (HGD) in Barrett’s esophagus [317, 334]. Using 330 nm excitation, Bourg-Heckly et al found 86% sensitivity and 95% specificity for differentiating neoplastic tissue (HGD and intramucosal carcinoma) from normal and Barrett’s mucosa in 24 patients by analyzing the fluorescence intensity ratio at 390 nm and 550 nm [334]. Panjehpour et al proposed another method called differential normalized fluorescence (DNF), where the measured fluorescence spectrum is subtracted by a baseline value (obtained by averaging the total-intensity-normalized spectra from the normal tissues) [317]. Based on the DNF intensity at 480 nm, a sensitivity of 90% and a specificity 96% for detection of HGD in non-dysplastic BE mucosa was reported from 36 patients [317]. Using the intensity ratio of green (500-549nm) to red (667-700nm) fluorescence and the intensity of blue excitation (477 nm) as two parameters for tissue classification, Mayinger et al found 97% sensitivity and 95% specificity for diagnosis of esophageal carcinoma on 9 patients [336]. Using a similar approach, they reported 84% sensitivity and 87% specificity for the diagnosis of gastric adenocarcinoma on 15 patients [337]. Fluorescence spectroscopy can also accurately identify dysplasia associated with adenomatous polyps in the colon [318, 338]. Using the probability distribution of the fluorescence intensity at 460nm and the intensity ratio of I_{680nm}/I_{660nm} as the diagnostic parameters, Cothren et al reported 90% sensitivity, 95% specificity, and 90% positive predictive value (PPV) for detection of colonic dysplasia in a study with 57 patients [318]. Mayinger et al also applied light-induced autofluorescence spectroscopy for the diagnosis of colorectal cancer and adenoma [339]. In a study with 11 patients, they found 96% sensitivity and 93% specificity for rectal cancer detection, and 98% sensitivity and 89% specificity for diagnosis of dysplastic adenomas [339].
The fluorescence signal is often influenced by wavelength-dependent tissue scattering and absorption. To overcome this limitation, methods have been developed to extract the intrinsic fluorescence spectroscopy (IFS) signal by measuring the fluorescence and reflectance spectra with same illumination/detection geometry to cancel these unwanted wavelength-dependent effects [340, 341]. Using IFS, Georgakoudi et al reported 100% sensitivity and 97% specificity in differentiating HGD from low-grade dysplasia (LGD) and non-dysplastic BE on 16 patients [342].

To enhance the fluorescence detection capability for early lesions, exogenous contrast agents that can selectively accumulate in neoplastic tissues can be applied. One of the most widely used exogenous contrast agents is 5-aminolevulinic acid (5-ALA), which is converted intracellularly into protoporphyrin IX (PpIX). PpIX has greater production and retention in neoplastic cells due to the increased metabolic rate and the reduced ferrochelatase activity which converts PpIX to heme [329, 343]. As a result, the characteristic red fluorescence of PpIX is increased in neoplastic tissues [327]. Clinical studies using point fluorescence spectroscopy showed oral administration of 5-ALA can detect HGD from non-dysplastic BE [344]. Using PpIX fluorescence alone, 77% sensitivity and 71% specificity were reported in a study with 20 patients, whereas 100% sensitivity and 100% specificity can be achieved by using the fluorescence intensity ratio \( I_{635nm}/I_{480nm} \) [344]. To further differentiate the PpIX fluorescence from autofluorescence, Ortner et al invented time-gated fluorescence spectroscopy utilizing the long fluorescence decay time of PpIX to suppress the autofluorescence background [345]. In this approach, nanosecond excitation pulses is used to excite the tissue, and then the ratio of 20 ns delayed PpIX fluorescence intensity to the immediate autofluorescence intensity is calculated. Using this method, it was possible to differentiate LGD from non-dysplastic BE, and dysplasia can be detected at a rate of 2.8 times higher compared to white-light screening endoscopy [345].
Fluorescence spectroscopy can be combined with other spectroscopic techniques to provide complementary information about the biochemical and morphological state of tissue. Georgakoudi et al demonstrated that superior results for differentiating dysplastic from non-dysplastic epithelium can be achieved by combining fluorescence, reflectance, and light-scattering spectroscopies [342]. In such a multi-modality approach, fluorescence spectroscopy (IFS) provides the biochemical information, diffuse reflectance spectroscopy (DRS) reveals morphologic information about the bulk tissue, and light-scattering spectroscopy (LSS) indicates the nuclei size and density information. Tri-modal spectroscopy has been shown to detect HGD from LGD and non-dysplastic BE with 100% sensitivity and 100% specificity on 40 sites from 16 patients [342]. The results from tri-modal spectroscopy are better than those obtained using individual modalities alone as tri-modal spectroscopy combines the advantages of each modality. Such multi-modality methods can be extended to imaging modes to enable rapid surveillance of large tissue areas.

Wide-Field Fluorescence Imaging

One example of the clinically used wide-field fluorescence systems is autofluorescence endoscopy, which can be implemented by using fiber-coupled wide-field excitation and CCD cameras for detection. There are several approaches to generate fluorescence images. Wang et al used a long-pass filter (> 400 nm) to select the fluorescent light, and an intensified charge injection device (CID) camera to capture the fluorescence image [346]. To account for the non-uniform illumination/detection geometry, they applied a moving average algorithm to the acquired image. The area of dysplasia was determined by the fluorescence intensity below a certain threshold. Another approach used two filters and intensified CCD (ICCD) cameras to detect the fluorescence in the green (490-560 nm) and red (>630 nm) wavelength ranges,
and the ratio between these two channels \( \frac{I_{\text{red}}}{I_{\text{green}}} \) was used to create pseudo-color images in real time [347, 348]. In this “laser-induced fluorescence” (LIF) system, normal mucosa usually appears cyan (blue-greenish), whereas neoplastic lesions appear red due to the higher red/green fluorescence intensity ratio. Newer generations of fluorescence endoscopy systems incorporate total autofluorescence and both green and red reflectance into the imaging algorithm. In this case, non-dysplastic mucosa appears green whereas neoplastic lesion appears blue-purple [349].

Fluorescence imaging can accurately identify dysplasia associated with adenomatous polyps in ex vivo colon specimens [346]. In an in vivo study of 30 patients, dysplasia was identified with a sensitivity of 83% [350]. Fluorescence endoscopy with LIF mode (LIFE) has been found to enhance the ability to localize small neoplastic lesions in the bronchus [351] and the GI tracts [347, 348, 352]. However, a randomized crossover study on 50 patients showed that LIFE was not superior to standard video endoscopy in detecting early neoplasia in Barrett’s esophagus (sensitivity for the diagnosis of HGD / early-stage cancer in targeted biopsy were both only 62%) [353].

As in point-probe fluorescence spectroscopy, using exogenous contrast agents such as 5-ALA promises to enhance the detection capability in wide-field fluorescence imaging. Endlicher et al found a sensitivity of >80% for dysplasia detection, while the specificity was only between 27% to 56% in a study of 47 patients [326]. Messmann et al also used 5-ALA to evaluate the detection of low and high-grade dysplasia in ulcerative colitis patients, and reported a sensitivity of 87-100%, while the specificity was only in the range of 51-62% in a study of 37 patients [325]. The high false positive rates are most likely associated with inflamed tissue or fecal materials [327, 354].
Using video autofluorescence imaging (AFI) system with both green and red reflectance, Kara et al recently demonstrated that AFI detected more dysplastic and neoplastic regions in Barrett’s esophagus than conventional endoscopy [349]. In a cohort of 60 patients with Barrett’s esophagus, 22 patients were diagnosed with high-grade intraepithelial neoplasia (HGIN), and among them, 7 patients were detected solely by AFI after high-resolution endoscopy (HRE) had not shown any suspicious lesions. However, the positive predictive value (PPV) from per lesion analysis was only 49%. Further technology development and combination of other imaging modalities will improve the specificity and PPV.

One of such promising modalities is narrow-band imaging (NBI). NBI utilizes a set of optical filters to allow only narrow wavelength regions of blue, green, and red light to sequentially illuminate the tissue [355-357]. Blue light penetrates only superficially, whereas red light penetrates into deeper layers. In addition, blue and green wavelengths are strongly absorbed by hemoglobin. Therefore, NBI enhances mucosal surface contrast and capillary patterns allowing detailed inspection of the mucosal and vascular patterns with high resolution and contrast without the use of exogenous dyes to improve visualization and diagnosis [358]. When combining AFI with NBI, AFI has high sensitivity in detecting dysplasia, and therefore can first scan large areas of mucosal surface to identify possible regions of neoplasia. However, AFI is often associated with high false-positive rate. NBI, on the other hand, can provide magnified inspection of mucosal patterns for detection of dysplasia. Therefore, the combination of AFI and NBI can provide complementary information for more accurate detection of early neoplasia. Kara et al performed a cross-sectional study on 20 patients with BE using endoscopic video AFI followed by NBI [359]. AFI identified all HGIN lesions (100% sensitivity), however, the false positive rate was high.
(19 in 47 lesions, 40%). Using NBI, the false positive rate dropped to 10% (5/47). Figure 8 shows an example of the detection of HGIN lesions using the combined modality.

![Image of Barrett's esophagus lesion with high-grade intraepithelial neoplasia (HGIN) detected with autofluorescence imaging (AFI) and narrow band imaging (NBI).](image)

**Figure 8**

The images of a Barrett's esophagus lesion with high-grade intraepithelial neoplasia (HGIN) detected with autofluorescence imaging (AFI) and narrow band imaging (NBI). (A) During inspection with white light, this area was not judged as suspicious. (B) The area around the small squamous island in the middle of the image showed a blue violet autofluorescence imaging color. (C) and (D) With NBI, irregular and disrupted mucosal patterns were found. The histopathology confirmed the presence of HGIN. From Ref. [359], with permission.

Wide-field imaging also enables image-guided resection of tumor and tumor margin assessment. Keller et al demonstrated autofluorescence and reflectance spectral imaging as a valuable tool for examining the superficial margin status of excised breast tissue specimens, with 85% sensitivity and 96% specificity [360]. 5-ALA-induced PpIX fluorescence imaging
has found great success clinically in fluorescence-guided resection of malignant glioma [361, 362]. Fluorescence-guided resection improves the results of glioma surgery for gross total resection and patient survival. Several clinical trials are undergoing to further validate its clinical utility [361-365]. Another FDA approved fluorescence contrast agent is ICG. Troyan et al recently demonstrated successful 6-patient pilot human clinical trial in breast cancer sentinel lymph node mapping using an intra-operative NIR fluorescence imaging system (FLARE™) [366]. Figure 9 shows the photo of the system and an example of SLM identification using peri-tumoral injection of ICG. These results demonstrate successful clinical translation of a new NIR fluorescence imaging system for image-guided oncologic surgery. Intra-operative fluorescence imaging of ICG lymphatic draining promises to guide resection or biopsy of various forms of cancers (including breast cancer [367-371], skin cancer [372, 373], gastric cancer [374, 375], and rectal cancer [376]), monitor the perineal wound contamination in abdominoperineal resection (APR) [377], as well as to interrogate the difference between normal and abnormal lymphatic structure and function [378-382].
**Figure 9**

(A) Portable FLARE™ imaging system and satellite monitor stand deployed in the operating room. (B) NIR fluorescent sentinel lymph node (SLN) mapping in a woman with breast cancer. Color video images (left), 800 nm NIR fluorescence images (middle) and a pseudo-colored (lime green) merge of the two (right) after injection (Inj.) of 10 μM ICG:HSA. The single SLN identified and resected for this patient. Shown are flow through a lymphatic channel (LC) and position of the SLN (arrow; top row), identification of the SLN (arrow; middle row), and a zoomed image of the SLN (arrow) during resection (bottom row). 800 nm camera exposure time was 200 msec. From Ref. [366], with permission.

**Fluorescence Tomography**

Fluorescence tomography can provide cross-sectional and volumetric views of biological tissue, therefore it is a promising tool to quantitatively characterize lesions and monitor therapy. Corlu et al reported the first human study of fluorescence tomographic imaging of breast tumor with ICG contrast enhancement [323]. Figure 10 shows the tomographic reconstruction of hemoglobin concentration, blood oxygen saturation, scattering coefficient and fluorescence signal. The reconstructed images demonstrated significant tumor contrast compared to typical endogenous optical contrast in breast, such as hemoglobin concentration and scattering coefficients obtained with traditional diffuse optical tomography (DOT). Successful fluorescence tomography in human represents a critical step towards application of molecularly-targeting probes for future clinical translation.
**Figure 10**

(A) Total hemoglobin concentration, blood oxygen saturation, $\mu'_s$ (786nm) and fluorescence image slices at $y = 5$ cm. (B) Iso-surface plot of THC, $\mu'_s$ (786nm) and fluorescence at isovales of three standard deviations above their respective means correspond to tumor location. Outline designates the border of the breast modeled as an ellipsoid using the breast photo taken with the CCD camera. From Ref. [323], with permission.

**Other Techniques and Applications**

Besides the spectrally-resolved steady-state fluorescence measurement techniques described above, time-resolved fluorescence measurement techniques [383, 384] are evolving and currently under investigation as a potential tool for clinical diagnosis and surgical guidance. Different from the intensity-based steady-state methods, time-resolved methods measure the fluorescence intensity decay properties (lifetime) to provide additional information of tissue. There are several advantages of using time-resolved fluorescence systems to investigate biological tissues [385]:

1. Biomolecules with overlapping fluorescence emission spectra but with different fluorescence decay times can be discriminated.
2. The measurements are sensitive to various parameters of the biological microenvironment, including pH, ion concentration and binding, enzymatic activity, and temperature, thus allowing these variables to be analyzed.

3. Time-resolved measurements are more robust to changes in fluorescence excitation-collection geometry; presence of endogenous absorbers (e.g., hemoglobin); photobleaching; and changes in fluorophore concentration and location depth, light scattering, and excitation intensity.

Time-resolved fluorescence techniques have been applied to detect atherosclerotic plaques and shown great promise in providing diagnostic information for high-risk atherosclerotic plaques [385]. In addition, applications such as tumor detection [386, 387] and image-guided tumor surgery [388] are actively pursued. With further technical development and pilot studies, this method holds strong promise on clinical translation.

2.4 Optical Molecular Imaging

*Probe and Instrumentation*

Nonspecific contrast agents (such as ALA and ICG) passively accumulate in diseased tissues without a specific molecular targeting moiety, therefore are subject to low sensitivity and specificity. Recently, there have been great advances in targeted imaging of basic molecular processes such as gene expression, enzyme activity, and disease-specific molecular interactions in vivo [389-391]. Molecular imaging promises early detection and in situ characterization of diseases with high sensitivity and specificity [392-395]. Optical imaging in conjunction with near-infrared fluorescent imaging probes has been developed to improve early disease detection [396, 397]. Generally, there are two major strategies for molecular-
targeting contrast agents [398], including active targeting (using targeted molecular probes with high affinity for specific disease-related targets) and selective activation of the image probe at target tissues.

**Affinity-Based Molecular Probes**

Affinity-based molecular probes utilize the molecular selectivity of diseased cells to differentiate normal from abnormal tissues. Compared to normal cells, diseased cells tend to over-express certain specific molecular biomarkers, therefore providing a means for imaging contrast. A variety of targeting moieties can be used to deliver the reporter dyes to the diseased tissue. Representative strategies for reporter dye-labeled molecular targeting contrast agents are listed in Table 2. Common approaches include: monoclonal antibodies [399, 400], protein ligands [401-404], small peptides [405-407], and non-peptide ligands [408-410].

**Table 2.** Representative Affinity-Based Molecular Probes.

| Targeting Moiety                  | Biomarker          | Reference          |
|-----------------------------------|--------------------|--------------------|
| Monoclonal Antibodies (mAb)       | VEGFR, VEGFR2      | [399, 400]         |
| Protein Ligands                   | VEGFR              | [401, 402]         |
|                                   | EGFR               | [403, 404]         |
| Small Peptides                    | Somatostatin       | [405-407, 411, 412]|
|                                   | Integrin           | [413-420]          |
|                                   | Bombesin           | [421-423]          |
| Non-Peptide Ligands               | Folate receptor    | [424-426]          |
VEGFR: vascular endothelial growth factor receptor; EGFR: epidermal growth factor receptor

**Activatable Molecular Probes**

Activatable probes are initially administered in a quenched (non-fluorescent) state, and the fluorescence signals increase when the probes are activated by specific biomolecules or environment in the disease tissue. Figure 11 shows an example of enzyme activatable imaging probe. Originally, the fluorophores are stacked together on a polymeric support, which leads to the quenching of fluorescence signals through Forster resonance energy transfer (FRET). The peptide linker is recognized by a specific proteolytic enzyme. Upon linker cleavage by the enzyme, the fluorophores are detached, which leads to dequenching and increases the fluorescence signal. Diseases tissues tend to have higher level of certain enzyme classes, therefore activatable probes provide an imaging contrast to differentiate normal from diseased tissues. Examples of activation mechanisms include [432]: enzymes, nucleic acids, ions and reactive oxygen species (ROS). Table 3 listed some representative activatable molecular imaging probes.
Figure 11
Principle of enzyme activated molecular probe.
**Table 3.** Representative Activatable Molecular Probes.

| Activation Mechanism | Biomarker | Reference |
|----------------------|-----------|-----------|
| Enzymes              | Cathepsin | [433, 434]|
|                      | MMP       | [435-437]|
| Nucleic Acids        | RNA       | [438, 439]|
| Ions                 | Calcium   | [440]     |
|                      | pH        | [441, 442]|
| ROS                  | H$_2$O$_2$| [443, 444]|

MMP: metalloproteinase; ROS: reactive oxygen species

**Optical Molecular Imaging Systems**

Optical molecular imaging systems are essentially similar to those used in fluorescence imaging. Fluorescence imaging can be performed with different resolutions and penetration depths ranging from microscopy to tomography. Fluorescence reflectance imaging (FRI) is commonly used for two-dimensional mapping of superficial fluorophore distributions. Fluorescence molecular tomography (FMT) [332] enables three-dimensional quantification of fluorescence signals inside scattering tissues. Using near-infrared fluorophores, deeper penetration can be achieved. In addition, tissue auto-fluorescence is reduced at longer wavelength, thereby improving target to background ratio.

Miniaturized endoscope devices can be developed for molecular imaging inside luminal structures in vivo. For example, Funovics et al developed a miniaturized 2.7 F (0.8
mm in diameter) fiber-optic sensor for laparoscopic imaging of enzyme activity and gene expression in vivo [445]. This device includes a dichroic mirror, a bandpass filter, and two independent cameras permitting simultaneous recording of white-light and fluorescent images. Zhu et al also demonstrated a one-dimensional near-infrared fluorescence imaging catheter for the detection of atherosclerotic plaque in human carotid plaque specimens in vitro [446]. The endoscopic devices will enable the intraluminal molecular imaging of GI tracts and vessels for early diseases identification.

Applications and Clinical Translation

Oncology

Numerous biomarkers for cancers have been identified to facilitate early detection [447]. Antibodies to these markers have high specificity, but their in vivo use has been limited by immunogenicity [448]. In contrast, peptides are typically less immunogenic, non-toxic, and relatively easier for mass production. Kelly et al developed fluorescent affinity ligands derived from a phage library specific for colon cancer, and demonstrated a 7-fold higher contrast than a control in orthotopic colonic tumors (HT29) using a two-channel miniaturized near-infrared fluorescent endoscopy [396]. Hsiung et al screened phage display peptide libraries against fresh human colonic adenomas for high-affinity ligands with preferential binding to premalignant tissue [449]. Furthermore, they conjugated the peptide with fluorescein and topically applied to human patients undergoing colonoscopy. In vivo fluorescence confocal microendoscopy images showed stronger binding of fluorescent affinity ligands to dysplastic tissues than normal (see Figure 12), with an overall sensitivity of 81% and specificity of 82%. Phage display based molecular imaging and targeted therapy represents a promising diagnostic and therapeutic approach for early detection of colorectal
cancer [450] and other cancers, such as prostate cancer [451], cholangiocarcinoma [452], hepatocarcinoma [453], and melanoma [454], among others.

![In vivo confocal fluorescence images of peptide binding. (a) Binding to dysplastic colon polyp. (b) Binding to adjacent normal mucosa. (c,d) Histology of dysplastic colon polyp (c) and normal mucosa (d) stained with H&E. Scale bars, 20 µm. From Ref. [449], with permission.]

In addition, proteolytic enzymes have been shown to play an essential role in tumor growth, including high cell turnover, invasion, and angiogenesis [455]. Cathepsin B in particular has been shown to be up-regulated in areas of inflammation, necrosis, angiogenesis [455], focal invasion of colorectal carcinomas [456], and dysplastic adenomas [457, 458]. Marten et al applied the fluorescent enzyme-cleavable and activatable cathepsin B sensing probe, which is non-fluorescent at injection and locally activated after target interaction, to the APC\textsuperscript{min} mouse model [397]. Ex vivo fluorescent imaging showed increased detection sensitivity and specificity, and the smallest lesion detected measured about 50 µm [397]. In vivo imaging of enzyme activity for colorectal cancer detection can be achieved using catheter-based microendoscope [445, 459]. Protease activity concentration (PAC), quantified by fluorescence molecular tomography (FMT), has been demonstrated to be a unique in vivo
diagnostic parameter for tumor detection and chemotherapy monitoring for brain glioma [460]. Another family of proteases, the matrix metalloproteinase (MMP), also shows higher expression in cancers than normal tissues. Studies have indicated that MMP-2 degrades the extracellular matrix and is involved in tumor infiltration and angiogenesis [461, 462]. Clinical studies also show a correlation between the level of MMP-2 expression and poor disease outcome [463]. Therefore, in vivo imaging and quantification of MMP-2 expression would be important in characterization of tumors. Bremer et al developed an activatable probe which can sense the MMP activity in vivo using near-infrared optical imaging [435, 436].

Mucins (glycoproteins that cover the surfaces of epithelial cells and aid the epithelia in homeostatic and metabolic functions) represent another promising target for molecular imaging. Colorectal tissues are abundantly supplied with mucins throughout the mucosa; however, the adenoma to carcinoma transformation of cancerous cells alters O-glycans mucinous expression [464]. During carcinoma transformation of cells, O-glycans mucinous expression is altered in tumor tissues [465]. An earlier study found out that “an exposed carbohydrate structure that is not normally present in human tissues is expressed in the mucin produced by malignant colonic epithelium” [466]. Another study also reported the levels of mature Muc1 mucins were significantly higher in carcinoma tissues than those in normal mucosa (p<0.001) [467]. A more recent study showed 100% Muc1 expression in colonic adenocarcinomas and 76% expression in adenomas, relative to 29% Muc1 expression by mucosa within 2 cm of the cancer margin, and 0% expression by normal mucosa > 2 cm from the cancer margin [468]. Previous studies demonstrated that α-L-fucose binding lectin Ulex europaeus agglutinin-1 (UEA-1) showed positive binding in human colorectal specimens of adenocarcinomas, adenomas, and polyposis coli, but not in the normal epithelium [469, 470]. In addition, increased UEA-1 reactivity in polyposis patients with a familial history of large bowel carcinoma has been reported [471]. Furthermore, it was reported that there was an 83%
positive rate of UEA-1 binding on apical surfaces of human carcinoma cells, compared with a 0% positive rate of UEA-1 binding on non-neoplastic mucosa adjacent to the carcinoma [472].

Roney et al developed UEA-1 conjugated polymerized liposomes for fluorescence molecular imaging (FMI) [473]. Figure 13 shows the results of the OCT/FMI imaging of APC\textsuperscript{min} mouse intestine ex vivo [474]. Polyps are visible in OCT images (Figures 13A-E) and histology (F-I). Fluorescence intensities (13K) are higher around the polypoid areas. This indicates the preferential accumulation and targeted binding of UEA-1 conjugated contrast agents to the polyps (13L).

![Figure 13](image)

**Figure 13**

Co-registered OCT/FMI of intestinal polyps ex vivo. (A) OCT en face surface profile image. (B-E) Cross-sectional OCT images corresponding to lines 1-4 in (A) and corresponding histology (F-I). (J) Tissue scattering coefficient ($\mu_s$) image. Polyps show higher scattering coefficients. (K) Fluorescence image using the contrast agents targeting to \(\alpha\)-L-fucose overexpressed in the mucin of polyp regions. (L) Fused scattering coefficient and fluorescence image. Bar = 1 mm. From Ref. [474], with permission.

Hypermetabolism is another hallmark of cancer and has been utilized clinically for radiolabeled fluorodeoxy glucose (\(^{18}\)FDG) based positron emission tomography (PET)
imaging for cancer detection [475]. The analogous fluorescent deoxy glucose (2-NBDG) has shown enhanced fluorescence in neoplastic tissues compared to normal tissues [408]. NIR fluorescence labeled glucose analogs also showed in vivo tumor-to-normal contrast enhancement [409, 410, 428-430, 476] and efficient photodynamic therapy [431]. Although the mechanism of cellular uptake is still under debate [477], metabolism-based molecular probe holds the promise of addressing the universal nature of cancer cells (not limited by specific cancer types) and could be an optical surrogate of FDG [394].

The expression of folate receptor-α (FR-α) is increased in 90-95% of epithelial ovarian cancers (EOC) [478, 479], and therefore could be a promising target for molecular imaging. Recently, van Dam et al performed the first in-human intra-operative fluorescence-guided ovarian cancer surgery using tumor-specific FR-α-targeted fluorescence contrast agent [480]. This work shows the potential application of molecular imaging in patients with ovarian cancer for improved intra-operative surgical guidance.

**Skeletal Disease**

Optical molecular imaging is also able to image osteoblastic activity. The development and integrity of the skeleton requires hydroxyapatite (HA) deposition by osteoblasts. Zaheer et al developed a NIR fluorescent bisphosphonate derivative which specifically binds to HA, thereby revealing osteoblastic activity in living animals [481]. Recently, Kovar et al synthesized a NIR fluorescent tetracycline derivative which binds specifically to differentiated mineralized osteoblasts [482]. Those agents hold promises in imaging bone development and mineralization, osteoblastic metastasis, and bone remodeling process.

Using FMT, Zilberman et al imaged fracture repair on murine models with implantation of mesenchymal stem cells overexpressing the osteogenic gene BMP2 [483].
Real-time imaging and quantification of bone formation and remodeling was performed using bisphosphonate imaging agent (OsteoSense\textsuperscript{TM}). Higher fluorescence signals was found at implantation sites, indicating fluorescence molecular imaging has the potential for quantitative evaluation of bone regeneration and tissue engineering.

Optical molecular imaging is also able to image and characterize arthritis, including rheumatoid arthritis (RA) and osteoarthritis (OA). Chen et al developed a NIR fluorophore-conjugated folate as probe for RA imaging [484]. A fluorescence signal intensity ratio of 2.3 between arthritic and normal joint was detected 12 and 24 hours after folate injection, which shows the potential for early diagnosis of RA. In addition, proteinase activities are altered during arthritis. Wunder et al used an activatable probe to detect proteinase activity in joints [485]. Similar approach also performed by Lai et al to measure the proteinase activity in OA instead of RA [486]. Together, these studies demonstrated the potential of optical molecular imaging as a means for mechanistic study and clinical applications for orthopedic research [487].

\textit{Cardiovascular Disease}

Optical molecular imaging can visualize molecular targets rather than anatomic structures therefore helps to elucidate the underpinning molecular and cellular mechanisms associated with cardiovascular diseases in vivo. NIR fluorescence molecular imaging has been applied to image atherosclerosis in vivo [488]. Chen et al. used FMT to image protease activity (ProSense\textsuperscript{TM}) in atheromata [489]. Non-invasive FMT detected the fluorescence signal in the atherosclerotic aorta in vivo and correlated well with ex vivo FRI images. This study demonstrated the feasibility of FMT to visualize augmented plaque protease activity. Deguchi et al investigated MMP-2 and MMP-9 activity in atherosclerosis using FMT [490].
Augmented NIR fluorescence (NIRF) signals were detected and co-localized with macrophage accumulation. As macrophages contribute pivotally to cardiovascular diseases, in vivo imaging of macrophages and protease activity would provide an important means to understanding the pathophysiology, evaluating the effects of interventions, and ultimately aiding clinical care [491].

To facilitate the clinical translation of molecular cardiovascular imaging, Jaffer et al developed an intravascular NIRF molecular sensing catheter based on the OCT imaging catheter [492]. This one-dimensional intravascular fluorescence catheter can detect cysteine protease activity using ProSense™ in real time. This device could aid in the detection of inflammation and high-risk plaques in small arteries. To provide a two-dimensional (2D) imaging, Jaffer et al also developed a 2D intravascular NIRF imaging catheter using rational and pullback design [493]. In atherosclerosis, 2D NIRF imaging provided insight into the spatial distribution of plaque protease activity. In stent-implanted vessels, 2D NIRF imaging indicated an edge-based pattern of stent induced arterial inflammation. Figure 14 shows in vivo multimodality x-ray angiography, IVUS, and intravascular NIRF imaging [493]. In stent-injured vessels, increased in vivo NIRF signal localized at the edges of the implanted stents, and particularly at the leading or distal stent edge (Fig. 14C), suggesting that stent-based injury occurred at sharp transition points. The continued development of molecular-sensing imaging agents and minimally-invasive intravascular fluorescence imaging device promise to provide high-resolution in vivo spatial mapping of inflammation-regulated protease activity in vivo.
Figure 14

Representative multimodality NIRF molecular and IVUS anatomical imaging of arterial inflammation at day 7 following coronary stent implantation. (A) Angiogram of an implanted bare-metal stent in the abdominal aorta. Dotted rectangle denotes stent position. (B, C) Longitudinal IVUS and NIRF catheter pullbacks demonstrate NIRF signal within the stent. NIRF signal collection was performed through blood without flushing, in 3.5-mm diameter vessels. From Ref. [493].
3. Conclusion

This paper provides an overview of several emerging optical technologies, including their principles and translation to various biomedical applications. The continuous technology development in advanced light sources, miniaturized imaging devices, and detection methods, will motivate future clinical investigations to determine optics method’s utility in medicine. The future technology development will be strengthened by the translational research from the bench to the bedside, and the clinical feedback will shape the new technology advancement. In addition, a close academia-industry synergy will facilitate the technology transfer and bring the new optical technologies to the hands of the clinicians. With such inter-disciplinary collaboration, optical methods are promised to significantly impact the future practice in clinical medicine.
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References

1. H. Hartridge, and F. J. W. Roughton, "A Method of Measuring the Velocity of Very Rapid Chemical Reactions," Proc. R. Soc. A. 104, 376-394 (1923).

2. F. J. W. Roughton, and G. A. Millikan, "Photoelectric Methods of Measuring the Velocity of Rapid Reactions. I. General Principles and Controls," Proc. R. Soc. A. 155, 258-269 (1936).

3. B. Chance, "Rapid and Sensitive Spectrophotometry. III. A Double Beam Apparatus," Rev. Sci. Instrum. 22, 634-638 (1951).

4. J. W. Severinghaus, "Takuo Aoyagi: discovery of pulse oximetry," Anesth Analg 105, S1-4 (2007).

5. J. Moyle, Pulse Oximetry (Principles and Practice) (BMJ Books, London, 2002).

6. G. A. Millikan, "The Oximeter, an Instrument for Measuring Continuously the Oxygen Saturation of Arterial Blood in Man," Rev. Sci. Instrum. 13, 434-444 (1942).

7. F. F. Jobsis, "Noninvasive, infrared monitoring of cerebral and myocardial oxygen sufficiency and circulatory parameters," Science 198, 1264-1267 (1977).

8. B. Chance, S. Nioka, J. Zhang, E. F. Conant, E. Hwang, S. Briest, S. G. Orel, M. D. Schnall, and B. J. Czerniecki, "Breast cancer detection based on incremental biochemical and physiological properties of breast cancers: a six-year, two-site study," Acad Radiol 12, 925-933 (2005).

9. B. Chance, J. S. Leigh, H. Miyake, D. S. Smith, S. Nioka, R. Greenfeld, M. Finander, K. Kaufmann, W. Levy, M. Young, and et al., "Comparison of time-resolved and unresolved measurements of deoxyhemoglobin in brain," Proceedings of the National Academy of Sciences of the United States of America 85, 4971-4975 (1988).

10. M. S. Patterson, B. Chance, and B. C. Wilson, "Time resolved reflectance and transmittance for the non-invasive measurement of tissue optical properties," Applied optics 28, 2331-2336 (1989).

11. V. Ntziachristos, A. G. Yodh, M. Schnall, and B. Chance, "Concurrent MRI and diffuse optical tomography of breast after indocyanine green enhancement," P Natl Acad Sci USA 97, 2767-2772 (2000).

12. D. Grosenick, K. T. Moesta, H. Wabnitz, J. Mucke, C. Stroszczyński, R. Macdonald, P. M. Schlag, and H. Rinneberg, "Time-domain optical mammography: initial clinical results on detection and characterization of breast tumors," Applied optics 42, 3170-3186 (2003).

13. J. B. Fishkin, and E. Gratton, "Propagation of photon-density waves in strongly scattering media containing an absorbing semi-infinite plane bounded by a straight edge," J Opt Soc Am A 10, 127-140 (1993).
14. S. J. Madsen, E. R. Anderson, R. C. Haskell, and B. J. Tromberg, "Portable, high-bandwidth frequency-domain photon migration instrument for tissue spectroscopy," Optics letters 19, 1934-1936 (1994).

15. Y. Chen, X. Intes, D. R. Tailor, R. R. Regatte, H. Ma, V. Ntziachristos, J. S. Leigh, R. Reddy, and B. Chance, "Probing rat brain oxygenation with near-infrared spectroscopy (NIRS) and magnetic resonance imaging (MRI)," Oxygen Transport to Tissue Volume Xxiii 510, 199-204 (2003).

16. A. H. Hielscher, A. Y. Bluestone, G. S. Abdoulaev, A. D. Klose, J. Lasker, M. Stewart, U. Netz, and J. Beuthan, "Near-infrared diffuse optical tomography," Disease markers 18, 313-337 (2002).

17. S. R. Arridge, "Optical tomography in medical imaging," Inverse Probl 15, R41-R93 (1999).

18. M. M. Roberts, F. E. Alexander, T. J. Anderson, U. Chetty, P. T. Donnan, P. Forrest, W. Hepburn, A. Huggins, A. E. Kirkpatrick, J. Lamb, and et al., "Edinburgh trial of screening for breast cancer: mortality at seven years," Lancet 335, 241-246 (1990).

19. M. J. Cross, S. E. Harms, J. H. Cheek, G. N. Peters, and R. C. Jones, "New horizons in the diagnosis and treatment of breast cancer using magnetic resonance imaging," Am J Surg 166, 749-753; discussion 753-745 (1993).

20. P. Forouhi, J. S. Walsh, T. J. Anderson, and U. Chetty, "Ultrasonography as a method of measuring breast tumour size and monitoring response to primary systemic treatment," Br J Surg 81, 223-225 (1994).

21. A. E. Cerussi, A. J. Berger, F. Bevilacqua, N. Shah, D. Jakubowski, J. Butler, R. F. Holcombe, and B. J. Tromberg, "Sources of absorption and scattering contrast for near-infrared optical mammography," Academic radiology 8, 211-218 (2001).

22. M. Tatsumi, C. Cohade, K. A. Mourtzikos, E. K. Fishman, and R. L. Wahl, "Initial experience with FDG-PET/CT in the evaluation of breast cancer," Eur J Nucl Med Mol Imaging 33, 254-262 (2006).

23. M. Cutler, "Transillumination as an aid in the diagnosis of breast lesions," Surg. Gynecol. Obstet. 48, 721-729 (1929).

24. J. Folkman, "Angiogenesis and breast cancer," J Clin Oncol 12, 441-443 (1994).

25. A. Rice, and C. M. Quinn, "Angiogenesis, thrombospondin, and ductal carcinoma in situ of the breast," J Clin Pathol 55, 569-574 (2002).

26. P. Vaupel, F. Kallinowski, and P. Okunieff, "BLOOD-FLOW, OXYGEN AND NUTRIENT SUPPLY, AND METABOLIC MICROENVIRONMENT OF HUMAN-TUMORS - A REVIEW," Cancer research 49, 6449-6465 (1989).
27. P. Vaupel, K. Schlenger, C. Knoop, and M. Hockel, "OXYGENATION OF HUMAN TUMORS - EVALUATION OF TISSUE OXYGEN DISTRIBUTION IN BREAST CANCERS BY COMPUTERIZED O2 TENSION MEASUREMENTS," Cancer research 51, 3316-3322 (1991).

28. V. Quaresima, S. J. Matcher, and M. Ferrari, "Identification and quantification of intrinsic optical contrast for near-infrared mammography," Photochemistry and photobiology 67, 4-14 (1998).

29. T. O. McBride, B. W. Pogue, E. D. Gerety, S. B. Poplack, U. L. Osterberg, and K. D. Paulsen, "Spectroscopic diffuse optical tomography for the quantitative assessment of hemoglobin concentration and oxygen saturation in breast tissue," Applied optics 38, 5480-5490 (1999).

30. B. J. Tromberg, O. Coquoz, J. B. Fishkin, T. Pham, E. R. Anderson, J. Butler, M. Cahn, J. D. Gross, V. Venugopalan, and D. Pham, "Non-invasive measurements of breast tissue optical properties using frequency-domain photon migration," Philos Trans R Soc Lond B Biol Sci 352, 661-668 (1997).

31. B. J. Tromberg, N. Shah, R. Lanning, A. Cerussi, J. Espinoza, T. Pham, L. Svaasand, and J. Butler, "Non-invasive in vivo characterization of breast tumors using photon migration spectroscopy," Neoplasia 2, 26-40 (2000).

32. S. Nioka, M. Miwa, S. Orel, M. Shnall, M. Haïda, S. Zhao, and B. Chance, "Optical imaging of human breast cancer," Adv Exp Med Biol 361, 171-179 (1994).

33. T. Durduran, R. Choe, J. P. Culver, L. Zubkov, M. J. Holboke, J. Giammarco, B. Chance, and A. G. Yodh, "Bulk optical properties of healthy female breast tissue," Physics in medicine and biology 47, 2847-2861 (2002).

34. V. Ntziachristos, and B. Chance, "Probing physiology and molecular function using optical imaging: applications to breast cancer," Breast Cancer Res 3, 41-46 (2001).

35. N. Shah, A. E. Cerussi, D. Jakubowski, D. Hsiang, J. Butler, and B. J. Tromberg, "The role of diffuse optical spectroscopy in the clinical management of breast cancer," Disease markers 19, 95-105 (2003).

36. B. J. Tromberg, A. Cerussi, N. Shah, M. Compton, A. Durkin, D. Hsiang, J. Butler, and R. Mehta, "Imaging in breast cancer: diffuse optics in breast cancer: detecting tumors in pre-menopausal women and monitoring neoadjuvant chemotherapy," Breast Cancer Res 7, 279-285 (2005).

37. A. Cerussi, D. Hsiang, N. Shah, R. Mehta, A. Durkin, J. Butler, and B. J. Tromberg, "Predicting response to breast cancer neoadjuvant chemotherapy using diffuse optical spectroscopy," Proceedings of the National Academy of Sciences of the United States of America 104, 4014-4019 (2007).
38. R. X. Xu, D. C. Young, J. J. Mao, and S. P. Povoski, "A prospective pilot clinical trial evaluating the utility of a dynamic near-infrared imaging device for characterizing suspicious breast lesions," Breast Cancer Res 9, R88 (2007).

39. S. Nioka, Y. Yung, M. Shnall, S. Zhao, S. Orel, C. Xie, B. Chance, and L. Solin, "Optical imaging of breast tumor by means of continuous waves," Adv Exp Med Biol 411, 227-232 (1997).

40. X. Intes, J. Ripoll, Y. Chen, S. Nioka, A. G. Yodh, and B. Chance, "In vivo continuous-wave optical breast imaging enhanced with Indocyanine Green," Medical Physics 30, 1039-1047 (2003).

41. B. Alacam, B. Yazici, X. Intes, S. Nioka, and B. Chance, "Pharmacokinetic-rate images of indocyanine green for breast tumors using near-infrared optical methods," Physics in medicine and biology 53, 837-859 (2008).

42. C. H. Schmitz, D. P. Klemer, R. Hardin, M. S. Katz, Y. Pei, H. L. Graber, M. B. Levin, R. D. Levina, N. A. Franco, W. B. Solomon, and R. L. Barbour, "Design and implementation of dynamic near-infrared optical tomographic imaging instrumentation for simultaneous dual-breast measurements," Applied optics 44, 2140-2153 (2005).

43. J. P. Culver, R. Choe, M. J. Holboke, L. Zubkov, T. Durduran, A. Slemp, V. Ntziachristos, B. Chance, and A. G. Yodh, "Three-dimensional diffuse optical tomography in the parallel plane transmission geometry: evaluation of a hybrid frequency domain/continuous wave clinical system for breast imaging," Medical physics 30, 235-247 (2003).

44. R. Choe, S. D. Konecky, A. Corlu, K. Lee, T. Durduran, D. R. Busch, S. Pathak, B. J. Czerniecki, J. Tchou, D. L. Fraker, A. DeMichele, B. Chance, S. R. Arridge, M. Schweiger, J. P. Culver, M. D. Schnall, M. E. Putt, M. A. Rosen, and A. G. Yodh, "Differentiation of benign and malignant breast tumors by in-vivo three-dimensional parallel-plate diffuse optical tomography," Journal of biomedical optics 14, 024020 (2009).

45. R. Choe, A. Corlu, K. Lee, T. Durduran, S. D. Konecky, M. Grosicka-Koptyra, S. R. Arridge, B. J. Czerniecki, D. L. Fraker, A. Demichele, B. Chance, M. A. Rosen, and A. G. Yodh, "Diffuse optical tomography of breast cancer during neoadjuvant chemotherapy: a case study with comparison to MRI," Medical physics 32, 1128-1139 (2005).

46. P. Taroni, G. Danesini, A. Torricelli, A. Pifferi, L. Spinelli, and R. Cubeddu, "Clinical trial of time-resolved scanning optical mammography at 4 wavelengths between 683 and 975 nm," Journal of biomedical optics 9, 464-473 (2004).

47. P. Taroni, L. Spinelli, A. Torricelli, A. Pifferi, G. M. Danesini, and R. Cubeddu, "Multi-wavelength time domain optical mammography," Technology in cancer research & treatment 4, 527-538 (2005).
48. P. Taroni, A. Torricelli, L. Spinelli, A. Pifferi, F. Arpaia, G. Danesini, and R. Cubeddu, "Time-resolved optical mammography between 637 and 985 nm: clinical study on the detection and identification of breast lesions," Physics in medicine and biology 50, 2469-2488 (2005).

49. D. Grosenick, K. T. Moesta, M. Moller, J. Mucke, H. Wabnitz, B. Gebauer, C. Stroszczynski, B. Wassermann, P. M. Schlag, and H. Rinneberg, "Time-domain scanning optical mammography: I. Recording and assessment of mammograms of 154 patients," Physics in medicine and biology 50, 2429-2449 (2005).

50. D. Grosenick, H. Wabnitz, K. T. Moesta, J. Mucke, M. Moller, C. Stroszczynski, J. Stossel, B. Wassermann, P. M. Schlag, and H. Rinneberg, "Concentration and oxygen saturation of haemoglobin of 50 breast tumours determined by time-domain optical mammography," Physics in medicine and biology 49, 1165-1181 (2004).

51. D. Grosenick, H. Wabnitz, K. T. Moesta, J. Mucke, P. M. Schlag, and H. Rinneberg, "Time-domain scanning optical mammography: II. Optical properties and tissue parameters of 87 carcinomas," Physics in medicine and biology 50, 2451-2468 (2005).

52. X. Intes, "Time-domain optical mammography SoftScan: initial results," Academic radiology 12, 934-947 (2005).

53. B. A. Brooksby, H. Dehghani, B. W. Pogue, and K. D. Paulsen, "Near-infrared (NIR) tomography breast image reconstruction with a priori structural information from MRI: Algorithm development for reconstructing heterogeneities," IEEE Journal of Selected Topics in Quantum Electronics 9, 199-209 (2003).

54. A. Li, E. L. Miller, M. E. Kilmer, T. J. Brukilacchio, T. Chaves, J. Stott, Q. Zhang, T. Wu, M. Chorlton, R. H. Moore, D. B. Kopans, and D. A. Boas, "Tomographic optical breast imaging guided by three-dimensional mammography," Applied optics 42, 5181-5190 (2003).

55. Q. Fang, S. A. Carp, J. Selb, G. Boverman, Q. Zhang, D. B. Kopans, R. H. Moore, E. L. Miller, D. H. Brooks, and D. A. Boas, "Combined optical imaging and mammography of the healthy breast: optical contrast derived from breast structure and compression," IEEE transactions on medical imaging 28, 30-42 (2009).

56. Q. Q. Fang, J. Selb, S. A. Carp, G. Boverman, E. L. Miller, D. H. Brooks, R. H. Moore, D. B. Kopans, and D. A. Boas, "Combined Optical and X-ray Tomosynthesis Breast Imaging," Radiology 258, 89-97 (2011).

57. B. Brooksby, S. D. Jiang, H. Dehghani, B. W. Pogue, K. D. Paulsen, C. Kogel, M. Doyley, J. B. Weaver, and S. P. Poplack, "Magnetic resonance-guided near-infrared tomography of the breast," Review of Scientific Instruments 75, 5262-5270 (2004).

58. M. J. Holboke, B. J. Tromberg, X. Li, N. Shah, J. Fishkin, D. Kidney, J. Butler, B. Chance, and A. G. Yodh, "Three-dimensional diffuse optical mammography with ultrasound localization in a human subject," Journal of Biomedical Optics 5, 237-247 (2000).
59. Q. Zhu, E. B. Cronin, A. A. Currier, H. S. Vine, M. M. Huang, N. G. Chen, and C. Xu, "Benign versus malignant breast masses: Optical differentiation with US-guided optical imaging reconstruction," Radiology 237, 57-66 (2005).

60. Q. Zhu, S. H. Kurtzman, P. Hegde, S. Tannenbaum, M. Kane, M. M. Huang, N. G. Chen, B. Jagjivan, and K. Zarfos, "Utilizing optical tomography with ultrasound localization to image heterogeneous hemoglobin distribution in large breast cancers," Neoplasia 7, 263-270 (2005).

61. Q. Zhu, "Optical tomography with ultrasound localization: Initial clinical results and technical challenges," Technology in cancer research & treatment 4, 235-244 (2005).

62. S. Ogawa, T. M. Lee, A. R. Kay, and D. W. Tank, "Brain magnetic resonance imaging with contrast dependent on blood oxygenation," Proceedings of the National Academy of Sciences of the United States of America 87, 9868-9872 (1990).

63. B. Chance, Z. Zhuang, C. UnAh, C. Alter, and L. Lipton, "Cognition-activated low-frequency modulation of light absorption in human brain," Proceedings of the National Academy of Sciences of the United States of America 90, 3770-3774 (1993).

64. A. Villringer, and B. Chance, "Non-invasive optical spectroscopy and imaging of human brain function," Trends Neurosci 20, 435-442 (1997).

65. Y. Hoshi, B. H. Tsou, V. A. Billock, M. Tanosaki, Y. Iguchi, M. Shimada, T. Shinba, Y. Yamada, and I. Oda, "Spatiotemporal characteristics of hemodynamic changes in the human lateral prefrontal cortex during working memory tasks," Neuroimage 20, 1493-1504 (2003).

66. M. Wolf, M. Ferrari, and V. Quaresima, "Progress of near-infrared spectroscopy and topography for brain and muscle clinical applications," Journal of biomedical optics 12, 062104 (2007).

67. E. M. C. Hillman, "Optical brain imaging in vivo: techniques and applications from animal to man," Journal of biomedical optics 12, 051402 (2007).

68. K. Takahashi, S. Ogata, Y. Atsumi, R. Yamamoto, S. Shiotsuka, A. Maki, Y. Yamashita, T. Yamamoto, H. Koizumi, H. Hirasawa, and M. Igawa, "Activation of the visual cortex imaged by 24-channel near-infrared spectroscopy," Journal of biomedical optics 5, 93-96 (2000).

69. B. W. Zeff, B. R. White, H. Dehghani, B. L. Schlaggar, and J. P. Culver, "Retinotopic mapping of adult human visual cortex with high-density diffuse optical tomography," Proceedings of the National Academy of Sciences of the United States of America 104, 12169-12174 (2007).

70. L. Becerra, W. Harris, M. Grant, E. George, D. Boas, and D. Borsook, "Diffuse optical tomography activation in the somatosensory cortex: specific activation by painful vs. non-painful thermal stimuli," PLoS One 4, e8016 (2009).
71. H. Sato, T. Takeuchi, and K. L. Sakai, "Temporal cortex activation during speech recognition: an optical topography study," Cognition 73, B55-66 (1999).

72. E. Watanabe, A. Maki, F. Kawaguchi, K. Takashiro, Y. Yamashita, H. Koizumi, and Y. Mayanagi, "Non-invasive assessment of language dominance with near-infrared spectroscopic mapping," Neurosci Lett 256, 49-52 (1998).

73. M. Pena, A. Maki, D. Kovacic, G. Dehaene-Lambertz, H. Koizumi, F. Bouquet, and J. Mehler, "Sounds and silence: an optical topography study of language recognition at birth," Proceedings of the National Academy of Sciences of the United States of America 100, 11702-11705 (2003).

74. B. Chance, S. Nioka, and Z. Zhao, "A wearable brain imager," IEEE Eng Med Biol Mag 26, 30-37 (2007).

75. P. J. Kirkpatrick, J. Lam, P. Al-Rawi, P. Smielewski, and M. Czosnyka, "Defining thresholds for critical ischemia by using near-infrared spectroscopy in the adult brain," J Neurosurg 89, 389-394 (1998).

76. T. Durduran, C. Zhou, B. L. Edlow, G. Yu, R. Choe, M. N. Kim, B. L. Cucchiara, M. E. Putt, Q. Shah, S. E. Kasner, J. H. Greenberg, A. G. Yodh, and J. A. Detre, "Transcranial optical monitoring of cerebrovascular hemodynamics in acute stroke patients," Optics express 17, 3884-3902 (2009).

77. E. Watanabe, Y. Nagahori, and Y. Mayanagi, "Focus diagnosis of epilepsy using near-infrared spectroscopy," Epilepsia 43 Suppl 9, 50-55 (2002).

78. M. N. Kim, T. Durduran, S. Frangos, B. L. Edlow, E. M. Buckley, H. E. Moss, C. Zhou, G. Yu, R. Choe, E. Maloney-Wilensky, R. L. Wolf, M. S. Grady, J. H. Greenberg, J. M. Levine, A. G. Yodh, J. A. Detre, and W. A. Kofke, "Noninvasive Measurement of Cerebral Blood Flow and Blood Oxygenation Using Near-Infrared and Diffuse Correlation Spectroscopies in Critically Brain-Injured Adults," Neurocrit Care 12, 173-180(2009).

79. K. Matsuo, T. Kato, K. Taneichi, A. Matsumoto, T. Ohtani, T. Hamamoto, H. Yamasue, Y. Sakano, T. Sasaki, M. Sadamatsu, A. Iwanami, N. Asukai, and N. Kato, "Activation of the prefrontal cortex to trauma-related stimuli measured by near-infrared spectroscopy in posttraumatic stress disorder due to terrorism," Psychophysiology 40, 492-500 (2003).

80. M. R. Stankovic, D. Maulik, W. Rosenfeld, P. G. Stubblefield, A. D. Kofinas, E. Gratton, M. A. Franceschini, S. Fantini, and D. M. Hueber, "Role of frequency domain optical spectroscopy in the detection of neonatal brain hemorrhage--a newborn piglet study," J Matern Fetal Med 9, 142-149 (2000).

81. X. Intes, and B. Chance, "Non-PET functional imaging techniques: optical," Radiol Clin North Am 43, 221-234 (2005).
82. A. Maki, Y. Yamashita, Y. Ito, E. Watanabe, Y. Mayanagi, and H. Koizumi, "Spatial and temporal analysis of human motor activity using noninvasive NIR topography," Medical Physics 22, 1997-2005 (1995).

83. H. Koizumi, T. Yamamoto, A. Maki, Y. Yamashita, H. Sato, H. Kawaguchi, and N. Ichikawa, "Optical topography: practical problems and new applications," Applied optics 42, 3054-3062 (2003).

84. M. Suda, K. Morimoto, A. Obata, H. Koizumi, and A. Maki, "Emotional responses to music: towards scientific perspectives on music therapy," Neuroreport 19, 75-78 (2008).

85. S. Tsujimoto, T. Yamamoto, H. Kawaguchi, H. Koizumi, and T. Sawaguchi, "Prefrontal cortical activation associated with working memory in adults and preschool children: an event-related optical topography study," Cereb Cortex 14, 703-712 (2004).

86. G. Taga, K. Asakawa, A. Maki, Y. Konishi, and H. Koizumi, "Brain imaging in awake infants by near-infrared optical topography," Proceedings of the National Academy of Sciences of the United States of America 100, 10722-10727 (2003).

87. G. Taga, Y. Konishi, A. Maki, T. Tachibana, M. Fujiwara, and H. Koizumi, "Spontaneous oscillation of oxy- and deoxy- hemoglobin changes with a phase difference throughout the occipital cortex of newborn infants observed using non-invasive optical topography," Neurosci Lett 282, 101-104 (2000).

88. Y. Fuchino, M. Nagao, T. Katura, M. Bando, M. Naito, A. Maki, K. Nakamura, H. Hayashi, H. Koizumi, and T. Yoro, "High cognitive function of an ALS patient in the totally locked-in state," Neurosci Lett 435, 85-89 (2008).

89. T. Kusaka, K. Kawada, K. Okubo, K. Nagano, M. Namba, H. Okada, T. Imai, K. Isobe, and S. Itoh, "Noninvasive optical imaging in the visual cortex in young infants," Hum Brain Mapp 22, 122-132 (2004).

90. M. A. Franceschini, and D. A. Boas, "Noninvasive measurement of neuronal activity with near-infrared optical imaging," NeuroImage 21, 372-386 (2004).

91. M. A. Franceschini, D. K. Joseph, T. J. Huppert, S. G. Diamond, and D. A. Boas, "Diffuse optical imaging of the whole head," Journal of biomedical optics 11, 054007 (2006).

92. B. R. White, A. Z. Snyder, A. L. Cohen, S. E. Petersen, M. E. Raichle, B. L. Schlaggar, and J. P. Culver, "Resting-state functional connectivity in the human brain revealed with diffuse optical tomography," NeuroImage 47, 148-156 (2009).

93. M. A. Franceschini, V. Toronov, M. Filiaci, E. Gratton, and S. Fantini, "On-line optical imaging of the human brain with 160-ms temporal resolution," Optics express 6, 49-57 (2000).
94. V. Toronov, M. A. Franceschini, M. Filiaci, S. Fantini, M. Wolf, A. Michalos, and E. Gratton, "Near-infrared study of fluctuations in cerebral hemodynamics during rest and motor stimulation: temporal analysis and spatial mapping," Medical physics 27, 801-815 (2000).

95. G. Gratton, and M. Fabiani, "Shedding light on brain function: the event-related optical signal," Trends Cogn Sci 5, 357-363 (2001).

96. C. Y. Tse, C. L. Lee, J. Sullivan, S. M. Garnsey, G. S. Dell, M. Fabiani, and G. Gratton, "Imaging cortical dynamics of language processing with the event-related optical signal," Proceedings of the National Academy of Sciences of the United States of America 104, 17157-17162 (2007).

97. S. R. Hintz, W. F. Cheong, J. P. van Houten, D. K. Stevenson, and D. A. Benaron, "Bedside imaging of intracranial hemorrhage in the neonate using light: comparison with ultrasound, computed tomography, and magnetic resonance imaging," Pediatr Res 45, 54-59 (1999).

98. F. E. W. Schmidt, M. E. Fry, E. M. C. Hillman, J. C. Hebden, and D. T. Delpy, "A 32-channel time-resolved instrument for medical optical tomography," Rev. Sci. Instrum. 71, 256-265 (2000).

99. J. C. Hebden, A. Gibson, R. M. Yusof, N. Everdell, E. M. Hillman, D. T. Delpy, S. R. Arridge, T. Austin, J. H. Meek, and J. S. Wyatt, "Three-dimensional optical tomography of the premature infant brain," Physics in medicine and biology 47, 4155-4166 (2002).

100. J. C. Hebden, A. Gibson, T. Austin, R. M. Yusof, N. Everdell, D. T. Delpy, S. R. Arridge, J. H. Meek, and J. S. Wyatt, "Imaging changes in blood volume and oxygenation in the newborn infant brain using three-dimensional optical tomography," Physics in medicine and biology 49, 1117-1130 (2004).

101. G. A. Millikan, "Experiments on Muscle Haemoglobin in vivo; The Instantaneous Measurement of Muscle Metabolism," Proceedings of the Royal Society of London B: Biological Sciences 123, 218-241 (1937).

102. B. Chance, and W. Bank, "Genetic disease of mitochondrial function evaluated by NMR and NIR spectroscopy of skeletal tissue," Biochimica et biophysica acta 1271, 7-14 (1995).

103. S. Matsui, N. Tamura, T. Hirakawa, S. Kobayashi, N. Takekoshi, and E. Murakami, "Assessment of working skeletal muscle oxygenation in patients with chronic heart failure," Am Heart J 129, 690-695 (1995).

104. V. Quaresima, R. Lepanto, and M. Ferrari, "The use of near infrared spectroscopy in sports medicine," J Sports Med Phys Fitness 43, 1-13 (2003).

105. J. M. Kowalchuk, H. B. Rossiter, S. A. Ward, and B. J. Whipp, "The effect of resistive breathing on leg muscle oxygenation using near-infrared spectroscopy during exercise in men," Exp Physiol 87, 601-611 (2002).
106. U. Wolf, M. Wolf, J. H. Choi, M. Levi, D. Choudhury, S. Hull, D. Coussirat, L. A. Paunescu, L. P. Safonova, A. Michalos, W. W. Mantulin, and E. Gratton, "Localized irregularities in hemoglobin flow and oxygenation in calf muscle in patients with peripheral vascular disease detected with near-infrared spectrophotometry," J Vasc Surg 37, 1017-1026 (2003).

107. Y. Q. Lin, G. Lech, S. Nioka, X. Intes, and B. Chance, "Noninvasive, low-noise, fast imaging of blood volume and deoxygenation changes in muscles using light-emitting diode continuous-wave imager," Rev. Sci. Instrum. 73, 3065-3074 (2002).

108. T. Hamaoka, K. K. McCully, V. Quaresima, K. Yamamoto, and B. Chance, "Near-infrared spectroscopy/imaging for monitoring muscle oxygenation and oxidative metabolism in healthy and diseased humans," Journal of biomedical optics 12, 062105 (2007).

109. M. Maris, E. Gratton, J. Maier, W. Mantulin, and B. Chance, "Functional near-infrared imaging of deoxygenated hemoglobin during exercise of the finger extensor muscles using the frequency-domain technique," Bioimaging 2, 174-183 (1994).

110. G. Yu, T. Durduran, G. Lech, C. Zhou, B. Chance, E. R. Mohler, 3rd, and A. G. Yodh, "Time-dependent blood flow and oxygenation in human skeletal muscles measured with noninvasive near-infrared diffuse optical spectroscopies," Journal of biomedical optics 10, 024027 (2005).

111. D. A. Boas, L. E. Campbell, and A. G. Yodh, "Scattering and Imaging with Diffusing Temporal Field Correlations," Phys Rev Lett 75, 1855-1858 (1995).

112. G. Yu, T. F. Floyd, T. Durduran, C. Zhou, J. Wang, J. A. Detre, and A. G. Yodh, "Validation of diffuse correlation spectroscopy for muscle blood flow with concurrent arterial spin labeled perfusion MRI," Optics express 15, 1064-1075 (2007).

113. E. M. Hillman, J. C. Hebden, M. Schweiger, H. Dehghani, F. E. Schmidt, D. T. Delpy, and S. R. Arridge, "Time resolved optical tomography of the human forearm," Physics in medicine and biology 46, 1117-1130 (2001).

114. H. Zhao, F. Gao, Y. Tanikawa, and Y. Yamada, "Time-resolved diffuse optical tomography and its application to in vitro and in vivo imaging," Journal of biomedical optics 12, 062107 (2007).

115. A. Torricelli, V. Quaresima, A. Pifferi, G. Biscotti, L. Spinelli, P. Taroni, M. Ferrari, and R. Cubeddu, "Mapping of calf muscle oxygenation and haemoglobin content during dynamic plantar flexion exercise by multi-channel time-resolved near-infrared spectroscopy," Physics in medicine and biology 49, 685-699 (2004).

116. D. M. Mancini, L. Bolinger, H. Li, K. Kendrick, B. Chance, and J. R. Wilson, "Validation of near-infrared spectroscopy in humans," J Appl Physiol 77, 2740-2747 (1994).
117. T. K. Tran, N. Sailasuta, U. Kreutzer, R. Hurd, Y. Chung, P. Mole, S. Kuno, and T. Jue, "Comparative analysis of NMR and NIRS measurements of intracellular PO2 in human skeletal muscle," The American journal of physiology 276, R1682-1690 (1999).

118. M. Ferrari, T. Binzoni, and V. Quaresima, "Oxidative metabolism in muscle," Philos Trans R Soc Lond B Biol Sci 352, 677-683 (1997).

119. L. Szmedra, J. Im, S. Nioka, B. Chance, and K. W. Rundell, "Hemoglobin/myoglobin oxygen desaturation during Alpine skiing," Med Sci Sports Exerc 33, 232-236 (2001).

120. K. K. McCully, C. Halber, and J. D. Posner, "Exercise-induced changes in oxygen saturation in the calf muscles of elderly subjects with peripheral vascular disease," J Gerontol 49, B128-134 (1994).

121. D. Huang, E. A. Swanson, C. P. Lin, J. S. Schuman, W. G. Stinson, W. Chang, M. R. Hee, T. Flotte, K. Gregory, C. A. Puliafito, and J. G. Fujimoto, "Optical coherence tomography," Science 254, 1178-1181 (1991).

122. J. G. Fujimoto, M. E. Brezinski, G. J. Tearney, S. A. Boppart, B. Bouma, M. R. Hee, J. F. Southern, and E. A. Swanson, "Optical biopsy and imaging using optical coherence tomography," Nature medicine 1, 970-972 (1995).

123. Z. Yaqoob, J. Wu, E. J. McDowell, X. Heng, and C. Yang, "Methods and application areas of endoscopic optical coherence tomography," J Biomed Opt 11, 063001 (2006).

124. M. A. Choma, M. V. Sarunic, C. H. Yang, and J. A. Izatt, "Sensitivity advantage of swept source and Fourier domain optical coherence tomography," Optics Express 11, 2183-2189 (2003).

125. J. F. de Boer, B. Cense, B. H. Park, M. C. Pierce, G. J. Tearney, and B. E. Bouma, "Improved signal-to-noise ratio in spectral-domain compared with time-domain optical coherence tomography," Optics letters 28, 2067-2069 (2003).

126. R. Leitgeb, C. K. Hitzenberger, and A. F. Fercher, "Performance of Fourier domain vs. time domain optical coherence tomography," Optics Express 11, 889-894 (2003).

127. D. C. Adler, Y. Chen, R. Huber, J. Schmitt, J. Connolly, and J. G. Fujimoto, "Three-dimensional endomicroscopy using optical coherence tomography," Nature Photonics 1, 709-716 (2007).

128. B. J. Vakoc, M. Shishko, S. H. Yun, W. Y. Oh, M. J. Suter, A. E. Desjardins, J. A. Evans, N. S. Nishioka, G. J. Tearney, and B. E. Bouma, "Comprehensive esophageal microscopy by using optical frequency-domain imaging (with video)," Gastrointestinal endoscopy 65, 898-905 (2007).

129. G. J. Tearney, M. E. Brezinski, B. E. Bouma, S. A. Boppart, C. Pitvis, J. F. Southern, and J. G. Fujimoto, "In vivo endoscopic optical biopsy with optical coherence tomography," Science 276, 2037-2039 (1997).
130. P. R. Herz, Y. Chen, A. D. Aguirre, J. G. Fujimoto, H. Mashimo, J. Schmitt, A. Koski, J. Goodnow, and C. Petersen, "Ultrahigh resolution optical biopsy with endoscopic optical coherence tomography," Optics Express 12, 3532-3542 (2004).

131. S. A. Boppart, B. E. Bouma, C. Pitris, G. J. Tearney, J. G. Fujimoto, and M. E. Brezinski, "Forward-imaging instruments for optical coherence tomography," Optics letters 22, 1618-1620 (1997).

132. L. P. Hariri, G. T. Bonnema, K. Schmidt, A. M. Winkler, V. Korde, K. D. Hatch, J. R. Davis, M. A. Brewer, and J. K. Barton, "Laparoscopic optical coherence tomography imaging of human ovarian cancer," Gynecol Oncol 114, 188-194 (2009).

133. X. Li, C. Chudoba, T. Ko, C. Pitris, and J. G. Fujimoto, "Imaging needle for optical coherence tomography," Optics letters 25, 1520-1522 (2000).

134. S. Han, M. V. Sarunic, J. Wu, M. Humayun, and C. Yang, "Handheld forward-imaging needle endoscope for ophthalmic optical coherence tomography inspection," J Biomed Opt 13, 020505 (2008).

135. A. F. Fercher, W. Drexler, C. K. Hitzenberger, and T. Lasser, "Optical coherence tomography-principles and applications," Reports on Progress in Physics 66, 239-303 (2003).

136. J. G. Fujimoto, "Optical coherence tomography for ultrahigh resolution in vivo imaging," Nat Biotechnol 21, 1361-1367 (2003).

137. B. E. Bouma, and G. J. Tearney, "Clinical imaging with optical coherence tomography," Academic radiology 9, 942-953 (2002).

138. J. Walther, M. Gaertner, P. Cimalla, A. Burkhardt, L. Kirsten, S. Meissner, and E. Koch, "Optical coherence tomography in biomedical research," Anal Bioanal Chem 400, 2721-2743 (2011).

139. M. R. Hee, J. A. Izatt, E. A. Swanson, D. Huang, J. S. Schuman, C. P. Lin, C. A. Puliafito, and J. G. Fujimoto, "Optical coherence tomography of the human retina," Arch Ophthalmol 113, 325-332 (1995).

140. A. Gaudric, B. Haouchine, P. Massin, M. Paques, P. Blain, and A. Erginay, "Macular hole formation: new data provided by optical coherence tomography," Arch Ophthalmol 117, 744-751 (1999).

141. R. J. Antcliff, D. J. Spalton, M. R. Stanford, E. M. Graham, T. J. ffytche, and J. Marshall, "Intravitreal triamcinolone for uveitic cystoid macular edema: an optical coherence tomography study," Ophthalmology 108, 765-772 (2001).

142. M. N. Apostolopoulos, C. N. Koutsandrea, M. N. Moschos, D. A. Alonistiotis, A. E. Papaspyrou, J. A. Mallias, T. E. Kyriaki, P. G. Theodossiadas, and G. P. Theodossiadi, "Evaluation of successful macular hole surgery by optical coherence tomography and multifocal electroretinography," American journal of ophthalmology 134, 667-674 (2002).
143. H. Sato, R. Kawasaki, and H. Yamashita, "Observation of idiopathic full-thickness macular hole closure in early postoperative period as evaluated by optical coherence tomography," American journal of ophthalmology 136, 185-187 (2003).

144. J. S. Schuman, C. A. Puliafito, and J. G. Fujimoto, Optical coherence tomography of ocular diseases (2nd Edition) (Slack Inc., Thorofare, NJ, 2004).

145. T. H. Ko, A. J. Witkin, J. G. Fujimoto, A. Chan, A. H. Rogers, C. R. Baumal, J. S. Schuman, W. Drexler, E. Reichel, and J. S. Duker, "Ultrahigh-resolution optical coherence tomography of surgically closed macular holes," Arch Ophthalmol 124, 827-836 (2006).

146. M. E. Brezinski, G. J. Tearney, B. E. Bouma, S. A. Boppart, M. R. Hee, E. A. Swanson, J. F. Southern, and J. G. Fujimoto, "Imaging of coronary artery microstructure (in vitro) with optical coherence tomography," The American journal of cardiology 77, 92-93 (1996).

147. I. K. Jang, G. Tearney, and B. Bouma, "Visualization of tissue prolapse between coronary stent struts by optical coherence tomography: comparison with intravascular ultrasound," Circulation 104, 2754 (2001).

148. G. J. Tearney, H. Yabushita, S. L. Houser, H. T. Aretz, I. K. Jang, K. H. Schlendorf, C. R. Kauffman, M. Shishkov, E. F. Halpern, and B. E. Bouma, "Quantification of macrophage content in atherosclerotic plaques by optical coherence tomography," Circulation 107, 113-119 (2003).

149. I. K. Jang, G. J. Tearney, B. MacNeill, M. Takano, F. Moselewski, N. Iftima, M. Shishkov, S. Houser, H. T. Aretz, E. F. Halpern, and B. E. Bouma, "In vivo characterization of coronary atherosclerotic plaque by use of optical coherence tomography," Circulation 111, 1551-1555 (2005).

150. B. E. Bouma, G. J. Tearney, C. C. Compton, and N. S. Nishioka, "High-resolution imaging of the human esophagus and stomach in vivo using optical coherence tomography," Gastrointest Endosc 51, 467-474 (2000).

151. M. V. Sivak, Jr., K. Kobayashi, J. A. Izatt, A. M. Rollins, R. Ung-Runyawee, A. Chak, R. C. Wong, G. A. Isenberg, and J. Willis, "High-resolution endoscopic imaging of the GI tract using optical coherence tomography," Gastrointest Endosc 51, 474-479 (2000).

152. X. D. Li, S. A. Boppart, J. Van Dam, H. Mashimo, M. Mutinga, W. Drexler, M. Klein, C. Pitris, M. L. Krinsky, M. E. Brezinski, and J. G. Fujimoto, "Optical coherence tomography: advanced technology for the endoscopic imaging of Barrett's esophagus," Endoscopy 32, 921-930 (2000).

153. V. X. Yang, S. J. Tang, M. L. Gordon, B. Qi, G. Gardiner, M. Cirocco, P. Kortan, G. B. Haber, G. Kandel, I. A. Vitkin, B. C. Wilson, and N. E. Marcon, "Endoscopic Doppler optical coherence tomography in the human GI tract: initial experience," Gastrointestinal endoscopy 61, 879-890 (2005).
154. G. Isenberg, M. V. Sivak, A. Chak, R. C. K. Wong, J. E. Willis, B. Wolf, D. Y. Rowland, A. Das, and A. Rollins, "Accuracy of endoscopic optical coherence tomography in the detection of dysplasia in Barrett's esophagus: a prospective, double-blinded study," Gastrointest Endosc 62, 825-831 (2005).

155. J. A. Evans, J. M. Poneres, B. E. Bouma, J. Bressner, E. F. Halpern, M. Shishkov, G. Y. Lauwers, M. Mino-Kenudson, N. S. Nishioka, and G. J. Tearney, "Optical coherence tomography to identify intramucosal carcinoma and high-grade dysplasia in Barrett's esophagus," Clin Gastroenterol H 4, 38-43 (2006).

156. Y. Chen, A. D. Aguirre, P. L. Hsiung, S. Desai, P. R. Herz, M. Pedrosa, Q. Huang, M. Figueiredo, S. W. Huang, A. Koski, J. M. Schmitt, J. G. Fujimoto, and H. Mashimo, "Ultrahigh resolution optical coherence tomography of Barrett's esophagus: preliminary descriptive clinical study correlating images with histology," Endoscopy 39, 599-605 (2007).

157. J. Welzel, C. Reinhardt, E. Lankenau, C. Winter, and H. H. Wolff, "Changes in function and morphology of normal human skin: evaluation using optical coherence tomography," Br J Dermatol 150, 220-225 (2004).

158. L. L. Otis, R. I. al-Sadhan, J. Meiers, and D. Redford-Badwal, "Identification of occlusal sealants using optical coherence tomography," The Journal of clinical dentistry 14, 7-10 (2003).

159. L. S. de Melo, R. E. de Araujo, A. Z. Freitas, D. Zezell, N. D. Vieira, J. Girkin, A. Hall, M. T. Carvalho, and A. S. Gomes, "Evaluation of enamel dental restoration interface by optical coherence tomography," J Biomed Opt 10, 064027 (2005).

160. G. J. Tearney, M. E. Brezinski, J. F. Southern, B. E. Bouma, S. A. Boppart, and J. G. Fujimoto, "Optical biopsy in human urologic tissue using optical coherence tomography," The Journal of urology 157, 1915-1919 (1997).

161. A. V. D'Amico, M. Weinstein, X. Li, J. P. Richie, and J. Fujimoto, "Optical coherence tomography as a method for identifying benign and malignant microscopic structures in the prostate gland," Urology 55, 783-787 (2000).

162. E. V. Zagaynova, O. S. Streltsova, N. D. Gladkova, L. B. Snopova, G. V. Gelikonov, F. I. Feldchtein, and A. N. Morozov, "In vivo optical coherence tomography feasibility for bladder disease," The Journal of urology 167, 1492-1496 (2002).

163. M. J. Manyak, N. D. Gladkova, J. H. Makari, A. M. Schwartz, E. V. Zagaynova, L. Zolfaghari, J. M. Zara, R. Iksanov, and F. I. Feldchtein, "Evaluation of superficial bladder transitional-cell carcinoma by optical coherence tomography," Journal of Endourology 19, 570-574 (2005).

164. C. Pitris, A. Goodman, S. A. Boppart, J. J. Libus, J. G. Fujimoto, and M. E. Brezinski, "High-resolution imaging of gynecologic neoplasms using optical coherence tomography," Obstet Gynecol 93, 135-139 (1999).
165. S. A. Boppart, A. Goodman, J. Libus, C. Pitris, C. A. Jesser, M. E. Brezinski, and J. G. Fujimoto, "High resolution imaging of endometriosis and ovarian carcinoma with optical coherence tomography: feasibility for laparoscopic-based imaging," British journal of obstetrics and gynaecology 106, 1071-1077 (1999).

166. M. A. Brewer, U. Utzinger, J. K. Barton, J. B. Hoying, N. D. Kirkpatrick, W. R. Brands, J. R. Davis, K. Hunt, S. J. Stevens, and A. F. Gmitro, "Imaging of the ovary," Technology in cancer research & treatment 3, 617-627 (2004).

167. E. A. Swanson, J. A. Izatt, M. R. Hee, D. Huang, C. P. Lin, J. S. Schuman, C. A. Puliafito, and J. G. Fujimoto, "In vivo retinal imaging by optical coherence tomography," Optics letters 18, 1864-1866 (1993).

168. A. F. Fercher, C. K. Hitzenberger, W. Drexler, G. Kamp, and H. Sattmann, "In vivo optical coherence tomography," American journal of ophthalmology 116, 113-114 (1993).

169. J. A. Izatt, M. R. Hee, E. A. Swanson, C. P. Lin, D. Huang, J. S. Schuman, C. A. Puliafito, and J. G. Fujimoto, "Micrometer-scale resolution imaging of the anterior eye in vivo with optical coherence tomography," Arch Ophthalmol 112, 1584-1589 (1994).

170. C. A. Puliafito, M. R. Hee, C. P. Lin, E. Reichel, J. S. Schuman, J. S. Duker, J. A. Izatt, E. A. Swanson, and J. G. Fujimoto, "Imaging of macular diseases with optical coherence tomography," Ophthalmology 102, 217-229 (1995).

171. R. A. Costa, M. Skaf, L. A. Melo, Jr., D. Calucci, J. A. Cardillo, J. C. Castro, D. Huang, and M. Wojtkowski, "Retinal assessment using optical coherence tomography," Progress in retinal and eye research 25, 325-353 (2006).

172. M. R. Hee, C. A. Puliafito, C. Wong, J. S. Duker, E. Reichel, B. Rutledge, J. S. Schuman, E. A. Swanson, and J. G. Fujimoto, "Quantitative assessment of macular edema with optical coherence tomography," Arch Ophthalmol 113, 1019-1029 (1995).

173. T. Otani, S. Kishi, and Y. Maruyama, "Patterns of diabetic macular edema with optical coherence tomography," American Journal of Ophthalmology 127, 688-693 (1999).

174. P. Massin, C. Allouch, B. Haouchine, F. Metge, M. Paques, L. Tanguy, A. Erginay, and A. Gaudric, "Optical coherence tomography of idiopathic macular epiretinal membranes before and after surgery," American journal of ophthalmology 130, 732-739 (2000).

175. R. J. Antcliff, M. R. Stanford, D. S. Chauhan, E. M. Graham, D. J. Spalton, J. S. Shilling, T. J. Ffytche, and J. Marshall, "Comparison between optical coherence tomography and fundus fluorescein angiography for the detection of cystoid macular edema in patients with uveitis," Ophthalmology 107, 593-599 (2000).

176. J. M. Jumper, R. P. Gallemore, B. W. McCuen, 2nd, and C. A. Toth, "Features of macular hole closure in the early postoperative period using optical coherence tomography," Retina 20, 232-237 (2000).
177. M. R. Hee, C. A. Puliafito, C. Wong, E. Reichel, J. S. Duker, J. S. Schuman, E. A. Swanson, and J. G. Fujimoto, "Optical coherence tomography of central serous chorioretinopathy," American Journal of Ophthalmology 120, 65-74 (1995).

178. M. R. Hee, C. R. Baumal, C. A. Puliafito, J. S. Duker, E. Reichel, J. R. Wilkins, J. G. Coker, J. S. Schuman, E. A. Swanson, and J. G. Fujimoto, "Optical coherence tomography of age-related macular degeneration and choroidal neovascularization," Ophthalmology 103, 1260-1270 (1996).

179. J. S. Schuman, M. R. Hee, C. A. Puliafito, C. Wong, T. Pedut-Kloizman, C. P. Lin, E. Hertzmark, J. A. Izatt, E. A. Swanson, and J. G. Fujimoto, "Quantification of nerve fiber layer thickness in normal and glaucomatous eyes using optical coherence tomography," Arch Ophthalmol 113, 586-596 (1995).

180. E. Z. Blumenthal, J. M. Williams, R. N. Weinreb, C. A. Girkin, C. C. Berry, and L. M. Zangwill, "Reproducibility of nerve fiber layer thickness measurements by use of optical coherence tomography," Ophthalmology 107, 2278-2282 (2000).

181. P. Carpineto, M. Ciancaglini, E. Zuppardi, G. Falconio, E. Doronzo, and L. Mastropasqua, "Reliability of nerve fiber layer thickness measurements using optical coherence tomography in normal and glaucomatous eyes," Ophthalmology 110, 190-195 (2003).

182. L. A. Paunescu, J. S. Schuman, L. L. Price, P. C. Stark, S. Beaton, H. Ishikawa, G. Wollstein, and J. G. Fujimoto, "Reproducibility of nerve fiber thickness, macular thickness, and optic nerve head measurements using StratusOCT," Investigative Ophthalmology and Visual Science 45, 1716-1724 (2004).

183. F. A. Medeiros, L. M. Zangwill, C. Bowd, R. M. Vessani, R. Susanna, Jr., and R. N. Weinreb, "Evaluation of retinal nerve fiber layer, optic nerve head, and macular thickness measurements for glaucoma detection using optical coherence tomography," Am J Ophthalmol 139, 44-55 (2005).

184. M. Mujat, R. Chan, B. Cense, B. Park, C. Joo, T. Akkin, T. Chen, and J. de Boer, "Retinal nerve fiber layer thickness map determined from optical coherence tomography images," Opt Express 13, 9480-9491 (2005).

185. W. Drexler, and J. G. Fujimoto, "State-of-the-art retinal optical coherence tomography," Progress in retinal and eye research 27, 45-88 (2008).

186. W. Drexler, "Ultrahigh-resolution optical coherence tomography," Journal of Biomedical Optics 9, 47-74 (2004).

187. W. Drexler, "Methodological advancements. Ultrahigh-resolution optical coherence tomography," Ophthalmologe 101, 804-812 (2004).
188. W. Drexler, U. Morgner, R. K. Ghanta, F. X. Kärtner, J. S. Schuman, and J. G. Fujimoto, "Ultrahigh-resolution ophthalmic optical coherence tomography," Nature medicine 7, 502-507 (2001).

189. W. Drexler, H. Sattmann, B. Hermann, T. H. Ko, M. Stur, A. Unterhuber, C. Scholda, O. Findl, M. Wirtitsch, J. G. Fujimoto, and A. F. Fercher, "Enhanced visualization of macular pathology with the use of ultrahigh-resolution optical coherence tomography," Arch Ophthalmol 121, 695-706 (2003).

190. G. Wollstein, L. A. Paunescu, T. H. Ko, J. G. Fujimoto, A. Kowalevicz, I. Hartl, S. Beaton, H. Ishikawa, C. Mattox, O. Singh, J. Duker, W. Drexler, and J. S. Schuman, "Ultrahigh-resolution optical coherence tomography in glaucoma," Ophthalmology 112, 229-237 (2005).

191. T. H. Ko, J. G. Fujimoto, J. S. Schuman, L. A. Paunescu, A. M. Kowalevicz, I. Hartl, W. Drexler, G. Wollstein, H. Ishikawa, and J. S. Duker, "Comparison of Ultrahigh- and Standard-Resolution Optical Coherence Tomography for Imaging Macular Pathology," Ophthalmology (2005).

192. L. A. Paunescu, T. H. Ko, J. S. Duker, A. Chan, W. Drexler, J. S. Schuman, and J. G. Fujimoto, "Idiopathic juxtafoveal retinal telangiectasis - New findings by ultrahigh-resolution optical coherence tomography," Ophthalmology 113, 48-57 (2006).

193. A. J. Witkin, T. H. Ko, J. G. Fujimoto, A. Chan, W. Drexler, J. S. Schuman, E. Reichel, and J. S. Duker, "Ultra-high resolution optical coherence tomography assessment of photoreceptors in retinitis pigmentosa and related diseases," Am J Ophthalmol 142, 945-952 (2006).

194. Y. Zhang, J. Rha, R. Jonnal, and D. Miller, "Adaptive optics parallel spectral domain optical coherence tomography for imaging the living retina," Opt Express 13, 4792-4811 (2005).

195. R. J. Zawadzki, S. M. Jones, S. S. Olivier, M. Zhao, B. A. Bower, J. A. Izatt, S. Choi, S. Laut, and J. S. Werner, "Adaptive-optics optical coherence tomography for high-resolution and high-speed 3D retinal in vivo imaging," Opt Express 13, 8532-8546 (2005).

196. E. J. Fernandez, B. Povazay, B. Hermann, A. Unterhuber, H. Sattmann, P. M. Prieto, R. Leitgeb, P. Ahnelt, P. Artal, and W. Drexler, "Three-dimensional adaptive optics ultrahigh-resolution optical coherence tomography using a liquid crystal spatial light modulator," Vision research 45, 3432-3444 (2005).

197. Y. Zhang, B. Cense, J. Rha, R. S. Jonnal, W. Gao, R. J. Zawadzki, J. S. Werner, S. Jones, S. Olivier, and D. T. Miller, "High-speed volumetric imaging of cone photoreceptors with adaptive optics spectral-domain optical coherence tomography," Opt Express 14, 4380-4394 (2006).
198. R. J. Zawadzki, S. S. Choi, A. R. Fuller, J. W. Evans, B. Hamann, and J. S. Werner, "Cellular resolution volumetric in vivo retinal imaging with adaptive optics-optical coherence tomography," Opt Express 17, 4084-4094 (2009).

199. B. Cense, E. Koperda, J. M. Brown, O. P. Kocaoglu, W. Gao, R. S. Jonnal, and D. T. Miller, "Volumetric retinal imaging with ultrahigh-resolution spectral-domain optical coherence tomography and adaptive optics using two broadband light sources," Opt Express 17, 4095-4111 (2009).

200. J. A. Izatt, M. R. Hee, G. M. Owen, E. A. Swanson, and J. G. Fujimoto, "Optical coherence microscopy in scattering media," Optics letters 19, 590-592 (1994).

201. K. Grieve, A. Dubois, M. Simonutti, M. Paques, J. Sahel, J. F. Le Gargasson, and C. Boccara, "In vivo anterior segment imaging in the rat eye with high speed white light full-field optical coherence tomography," Opt Express 13, 6286-6295 (2005).

202. M. Akiba, N. Maeda, K. Yumikake, T. Soma, K. Nishida, Y. Tano, and K. P. Chan, "Ultrahigh-resolution imaging of human donor cornea using full-field optical coherence tomography," J Biomed Opt 12, 041202 (2007).

203. M. Wojtkowski, R. Leitgeb, A. Kowalczyk, T. Bajraszewski, and A. F. Fercher, "In vivo human retinal imaging by Fourier domain optical coherence tomography," Journal of Biomedical Optics 7, 457-463 (2002).

204. N. Nassif, B. Cense, B. H. Park, S. H. Yun, T. C. Chen, B. E. Bouma, G. J. Tearney, and J. F. de Boer, "In vivo human retinal imaging by ultrahigh-speed spectral domain optical coherence tomography," Optics letters 29, 480-482 (2004).

205. N. A. Nassif, B. Cense, B. H. Park, M. C. Pierce, S. H. Yun, B. E. Bouma, G. J. Tearney, T. C. Chen, and J. F. de Boer, "In vivo high-resolution video-rate spectral-domain optical coherence tomography of the human retina and optic nerve," Optics Express 12, 367-376 (2004).

206. M. Wojtkowski, V. J. Srinivasan, T. H. Ko, J. G. Fujimoto, A. Kowalevicz, and J. S. Duker, "Ultrahigh resolution, high speed, Fourier domain optical coherence tomography and methods for dispersion compensation," Optics Express 12, 2404-2422 (2004).

207. B. Cense, N. Nassif, T. C. Chen, M. C. Pierce, S. Yun, B. H. Park, B. Bouma, G. Tearney, and J. F. de Boer, "Ultrahigh-resolution high-speed retinal imaging using spectral-domain optical coherence tomography," Optics Express 12, 2435-2447 (2004).

208. R. A. Leitgeb, W. Drexler, A. Unterhuber, B. Hermann, T. Bajraszewski, T. Le, A. Stingl, and A. F. Fercher, "Ultrahigh resolution Fourier domain optical coherence tomography," Optics Express 12, 2156-2165 (2004).

209. Y. Chen, L. N. Vuong, J. Liu, J. Ho, V. J. Srinivasan, I. Gorczynska, A. J. Witkin, J. S. Duker, J. Schuman, and J. G. Fujimoto, "Three-dimensional ultrahigh resolution optical
coherence tomography imaging of age-related macular degeneration," Opt Express 17, 4046-4060 (2009).

210. G. Vizzeri, M. Balasubramanian, C. Bowd, R. N. Weinreb, F. A. Medeiros, and L. M. Zangwill, "Spectral domain-optical coherence tomography to detect localized retinal nerve fiber layer defects in glaucomatous eyes," Opt Express 17, 4004-4018 (2009).

211. B. Povazay, B. Hermann, A. Unterhuber, B. Hofer, H. Sattmann, F. Zeiler, J. E. Morgan, C. Falkner-Radler, C. Glittenberg, S. Blinder, and W. Drexler, "Three-dimensional optical coherence tomography at 1050 nm versus 800 nm in retinal pathologies: enhanced performance and choroidal penetration in cataract patients," J Biomed Opt 12, 041211 (2007).

212. B. Povazay, B. Hofer, C. Torti, B. Hermann, A. R. Tumlinson, M. Esmaeelpour, C. A. Egan, A. C. Bird, and W. Drexler, "Impact of enhanced resolution, speed and penetration on three-dimensional retinal optical coherence tomography," Opt Express 17, 4134-4150 (2009).

213. M. Wojtkowski, V. Srinivasan, J. G. Fujimoto, T. Ko, J. S. Schuman, A. Kowelczyk, and J. S. Duker, "Three-dimensional retinal imaging with high-speed ultrahigh-resolution optical coherence tomography," Ophthalmology 112, 1734-1746 (2005).

214. Z. Chen, T. E. Milner, D. Dave, and J. S. Nelson, "Optical Doppler tomographic imaging of fluid flow velocity in highly scattering media," Optics letters 22, 64-66 (1997).

215. J. A. Izatt, M. D. Kulkami, S. Yazdanfar, J. K. Barton, and A. J. Welch, "In vivo bidirectional color Doppler flow imaging of picoliter blood volumes using optical coherence tomography," Optics letters 22, 1439-1441 (1997).

216. Y. Zhao, Z. Chen, C. Saxer, S. Xiang, J. F. de Boer, and J. S. Nelson, "Phase-resolved optical coherence tomography and optical Doppler tomography for imaging blood flow in human skin with fast scanning speed and high velocity sensitivity," Optics letters 25, 114-116 (2000).

217. Y. Wang, B. A. Bower, J. A. Izatt, O. Tan, and D. Huang, "Retinal blood flow measurement by circumpapillary Fourier domain Doppler optical coherence tomography," J Biomed Opt 13, 064003 (2008).

218. Y. K. Tao, K. M. Kennedy, and J. A. Izatt, "Velocity-resolved 3D retinal microvessel imaging using single-pass flow imaging spectral domain optical coherence tomography," Opt Express 17, 4177-4188 (2009).

219. T. Schmoll, C. Kolbitsch, and R. A. Leitgeb, "Ultra-high-speed volumetric tomography of human retinal blood flow," Opt Express 17, 4166-4176 (2009).

220. S. Makita, Y. Hong, M. Yamanari, T. Yatagai, and Y. Yasuno, "Optical coherence angiography," Opt Express 14, 7821-7840 (2006).
221. C. Kolbitsch, T. Schmoll, and R. A. Leitgeb, "Histogram-based filtering for quantitative 3D retinal angiography," Journal of biophotonics 2, 416-425 (2009).

222. J. F. De Boer, T. E. Milner, M. J. C. van Gemert, and J. S. Nelson, "Two-dimensional birefringence imaging in biological tissue by polarization-sensitive optical coherence tomography," Optics letters 22, 934-936 (1997).

223. B. Cense, T. C. Chen, B. H. Park, M. C. Pierce, and J. F. de Boer, "Thickness and birefringence of healthy retinal nerve fiber layer tissue measured with polarization-sensitive optical coherence tomography," Investigative ophthalmology & visual science 45, 2606-2612 (2004).

224. E. Gotzinger, M. Pircher, M. Sticker, A. F. Fercher, and C. K. Hitzenberger, "Measurement and imaging of birefringent properties of the human cornea with phase-resolved, polarization-sensitive optical coherence tomography," Journal of Biomedical Optics 9, 94-102 (2004).

225. M. Pircher, E. Gotzinger, O. Findl, S. Michels, W. Geitzenauer, C. Leydolt, U. Schmidt-Erfurth, and C. K. Hitzenberger, "Human macula investigated in vivo with polarization-sensitive optical coherence tomography," Investigative ophthalmology & visual science 47, 5487-5494 (2006).

226. M. Mujat, B. H. Park, B. Cense, T. C. Chen, and J. F. de Boer, "Autocalibration of spectral-domain optical coherence tomography spectrometers for in vivo quantitative retinal nerve fiber layer birefringence determination," J Biomed Opt 12, 041205 (2007).

227. Y. Yasuno, M. Yamanari, K. Kawana, T. Oshika, and M. Miura, "Investigation of post-glaucoma-surgery structures by three-dimensional and polarization sensitive anterior eye segment optical coherence tomography," Opt Express 17, 3980-3996 (2009).

228. K. Bizheva, R. Pflug, B. Hermann, B. Povazay, H. Sattmann, P. Qiu, E. Anger, H. Reitsamer, S. Popov, J. R. Taylor, A. Unterhuber, P. Ahnelt, and W. Drexler, "Optophysiology: depth-resolved probing of retinal physiology with functional ultrahigh-resolution optical coherence tomography," Proceedings of the National Academy of Sciences of the United States of America 103, 5066-5071 (2006).

229. V. J. Srinivasan, Y. Chen, J. S. Duker, and J. G. Fujimoto, "In vivo functional imaging of intrinsic scattering changes in the human retina with high-speed ultrahigh resolution OCT," Opt Express 17, 3861-3877 (2009).

230. R. B. Rosen, M. Hathaway, J. Rogers, J. Pedro, P. Garcia, G. M. Dobre, and A. G. Podoleanu, "Simultaneous OCT/SLO/ICG imaging," Investigative ophthalmology & visual science 50, 851-860 (2009).

231. G. J. Tearney, M. E. Brezinski, S. A. Boppart, B. E. Bouma, N. Weissman, J. F. Southern, E. A. Swanson, and J. G. Fujimoto, "Catheter-based optical imaging of a human coronary artery," Circulation 94, 3013 (1996).
232. J. G. Fujimoto, S. A. Boppart, G. J. Tearney, B. E. Bouma, C. Pitris, and M. E. Brezinski, "High resolution in vivo intra-arterial imaging with optical coherence tomography," Heart 82, 128-133 (1999).

233. E. Grube, U. Gerckens, L. Buellesfeld, and P. J. Fitzgerald, "Images in cardiovascular medicine. Intracoronary imaging with optical coherence tomography: a new high-resolution technology providing striking visualization in the coronary artery," Circulation 106, 2409-2410 (2002).

234. I. K. Jang, B. E. Bouma, D. H. Kang, S. J. Park, S. W. Park, K. B. Seung, K. B. Choi, M. Shishkov, K. Schlendorf, E. Pomerantsev, S. L. Houser, H. T. Aretz, and G. J. Tearney, "Visualization of coronary atherosclerotic plaques in patients using optical coherence tomography: comparison with intravascular ultrasound," Journal of the American College of Cardiology 39, 604-609 (2002).

235. N. A. Patel, X. Li, D. L. Stamper, J. G. Fujimoto, and M. E. Brezinski, "Guidance of aortic ablation using optical coherence tomography," The international journal of cardiovascular imaging 19, 171-178 (2003).

236. I. K. Jang, B. Bouma, B. MacNeill, M. Takano, M. Shishkov, N. Iftima, and G. J. Tearney, "In-vivo coronary plaque characteristics in patients with various clinical presentations using Optical Coherence Tomography," Circulation 108, 373-373 (2003).

237. B. D. MacNeill, I. K. Jang, B. E. Bouma, N. Iftimia, M. Takano, H. Yabushita, M. Shishkov, C. R. Kaufman, S. L. Houser, H. T. Aretz, D. DeJoseph, E. F. Halpern, and G. J. Tearney, "Focal and multi-focal plaque distributions in patients with macrophage acute and stable presentations of coronary artery disease," Journal of the American College of Cardiology 44, 972-979 (2004).

238. E. Regar, H. M. van Beusekom, W. J. van der Giessen, and P. W. Serruys, "Images in cardiovascular medicine. Optical coherence tomography findings at 5-year follow-up after coronary stent implantation," Circulation 112, e345-346 (2005).

239. R. Bhindi, S. M. Munir, and K. M. Channon, "Images in cardiovascular medicine. Optical coherence tomography in the setting of an acute anterior myocardial infarction," Circulation 116, e366-367 (2007).

240. S. H. Yun, G. J. Tearney, B. J. Vakoc, M. Shishkov, W. Y. Oh, A. E. Desjardins, M. J. Suter, R. C. Chan, J. A. Evans, I. K. Jang, N. S. Nishioka, J. F. de Boer, and B. E. Bouma, "Comprehensive volumetric optical microscopy in vivo," Nature medicine 12, 1429-1433 (2006).

241. M. Brezinski, "Characterizing arterial plaque with optical coherence tomography," Current opinion in cardiology 17, 648-655 (2002).
242. H. Yabushita, B. E. Bouma, S. L. Houser, H. T. Aretz, I. K. Jang, K. H. Schlendorf, C. R. Kauffman, M. Shishkov, E. F. Halpern, and G. J. Tearney, "Monitoring atherosclerotic plaque evolution in vivo by optical coherence tomography," Circulation 108, 416-416 (2003).

243. M. E. Brezinski, G. J. Tearney, B. E. Bouma, J. A. Izatt, M. R. Hee, E. A. Swanson, J. F. Southern, and J. G. Fujimoto, "Optical coherence tomography for optical biopsy. Properties and demonstration of vascular pathology," Circulation 93, 1206-1213 (1996).

244. H. Yabushita, B. E. Bouma, S. L. Houser, H. T. Aretz, I. K. Jang, K. H. Schlendorf, C. R. Kauffman, M. Shishkov, D. H. Kang, E. F. Halpern, and G. J. Tearney, "Characterization of human atherosclerosis by optical coherence tomography," Circulation 106, 1640-1645 (2002).

245. G. J. Tearney, I. K. Jang, and B. E. Bouma, "Optical coherence tomography for imaging the vulnerable plaque," J Biomed Opt 11, 021002 (2006).

246. F. Saia, J. Schaar, E. Regar, G. Rodriguez, P. J. De Feyter, F. Mastik, A. Marzocchi, C. Marrozzini, P. Ortolani, T. Palmerini, A. Branzi, A. F. van der Steen, and P. W. Serruys, "Clinical imaging of the vulnerable plaque in the coronary arteries: new intracoronary diagnostic methods," Journal of cardiovascular medicine 7, 21-28 (2006).

247. M. E. Brezinski, "Optical coherence tomography for identifying unstable coronary plaque," International journal of cardiology 107, 154-165 (2006).

248. T. Kubo, T. Imanishi, S. Takarada, A. Kuroi, S. Ueno, T. Yamano, T. Tanimoto, Y. Matsuo, T. Masho, H. Kitabata, K. Tsuda, Y. Tomobuchi, and T. Akasaka, "Assessment of culprit lesion morphology in acute myocardial infarction: ability of optical coherence tomography compared with intravascular ultrasound and coronary angiography," Journal of the American College of Cardiology 50, 933-939 (2007).

249. F. J. van der Meer, D. Faber, D. M. Sassoon, M. C. Aalders, G. Pasterkamp, and T. G. van Leeuwen, "Localized measurement of optical attenuation coefficients of atherosclerotic plaque constituents by quantitative optical coherence tomography," IEEE Trans Med Imaging 24, 1369-1376 (2005).

250. T. Kume, T. Akasaka, T. Kawamoto, H. Okura, N. Watanabe, E. Toyota, Y. Neishi, R. Sukmawan, Y. Sadahira, and K. Yoshida, "Measurement of the thickness of the fibrous cap by optical coherence tomography," American heart journal 152, 755.e1-755.e4 (2006).

251. C. Xu, J. M. Schmitt, S. G. Carlier, and R. Virmani, "Characterization of atherosclerosis plaques by measuring both backscattering and attenuation coefficients in optical coherence tomography," J Biomed Opt 13, 034003 (2008).

252. B. E. Bouma, G. J. Tearney, H. Yabushita, M. Shishkov, C. R. Kauffman, D. DeJoseph Gauthier, B. D. MacNeill, S. L. Houser, H. T. Aretz, E. F. Halpern, and I. K. Jang, "Evaluation of intracoronary stenting by intravascular optical coherence tomography," Heart 89, 317-320 (2003).
253. T. Kume, T. Akasaka, T. Kawamoto, N. Watanabe, E. Toyota, R. Sukmawan, Y. Sadahira, and K. Yoshida, "Visualization of neointima formation by optical coherence tomography," International heart journal 46, 1133-1136 (2005).

254. S. Ito, M. Itoh, and M. Suzuki, "Intracoronary imaging with optical coherence tomography after cutting balloon angioplasty for in-stent restenosis," J Invasive Cardiol. 17, 369-370. (2005).

255. Y. Kawase, K. H. Kozo, Y. Ryuichi, J. Mcgreor, R. Hajjar, I. K. Jang, and M. Hayase, "In vivo volumetric analysis of coronary stent using optical coherence tomography with a novel balloon occlusion-flushing catheter: a comparison with intravascular ultrasound," Ultrasound in Med. Biol. 31, 1343-1349 (2005).

256. O. C. Raffel, J. C. Hannan, and I. K. Jang, "Coronary stent malapposition as a post-stenotic aneurysm detected by optical coherence tomography," J. Invasive. Cardiol. 18, 561-562 (2006).

257. P. Barlis, G. van Soest, P. W. Serruys, and E. Regar, "Intracoronary optical coherence tomography and the evaluation of stents," Expert review of medical devices 6, 157-167 (2009).

258. M. Takano, S. Inami, I. K. Jang, M. Yamamoto, D. Murakami, K. Seimiya, T. Ohba, and K. Mizuno, "Evaluation by optical coherence tomography of neointimal coverage of sirolimus-eluting stent three months after implantation," Am. J. Cardiol. 99, 1033-1038 (2007).

259. D. Matsumoto, J. Shite, T. Shinke, H. Otake, Y. Tanino, D. Ogasawara, T. Sawada, O. L. Paredes, K. Hirata, and M. Yokoyama, "Neointimal coverage of sirolimus-eluting stents at 6-month follow-up: evaluated by optical coherence tomography," Eur. Heart J. 28, 961-967 (2007).

260. S. D. Giattina, B. K. Courtney, P. R. Herz, M. Harman, S. Shortkroff, D. L. Stamper, B. Liu, J. G. Fujimoto, and M. E. Brezinski, "Assessment of coronary plaque collagen with polarization sensitive optical coherence tomography (PS-OCT)," Int. J. Cardiol. 107, 400-409 (2006).

261. W. C. Kuo, M. W. Hsiung, J. J. Shyu, N. K. Chou, and P. N. Yang, "Assessment of arterial characteristics in human atherosclerosis by extracting optical properties from polarization-sensitive optical coherence tomography," Opt Express 16, 8117-8125 (2008).

262. S. K. Nadkarni, M. C. Pierce, B. H. Park, J. F. de Boer, P. Whittaker, B. E. Bouma, J. E. Bressner, E. Halpern, S. L. House, and G. J. Tearney, "Measurement of collagen and smooth muscle cell content in atherosclerotic plaques using polarization-sensitive optical coherence tomography," Journal of the American College of Cardiology 49, 1474-1481 (2007).
263. A. H. Chau, R. C. Chan, M. Shishkov, B. MacNeill, N. Iftimia, G. J. Tearney, R. D. Kamm, B. E. Bouma, and M. R. Kazaempur-Mofrad, "Mechanical analysis of atherosclerotic plaques based on optical coherence tomography," Annals of Biomedical Engineering 32, 1494-1503 (2004).

264. J. Rogowska, N. Patel, S. Plummer, and M. E. Brezinski, "Quantitative optical coherence tomographic elastography: method for assessing arterial mechanical properties," The British journal of radiology 79, 707-711 (2006).

265. J. K. Barton, F. Guzman, and A. Tumlinson, "Dual modality instrument for simultaneous optical coherence tomography imaging and fluorescence spectroscopy," J Biomed Opt 9, 618-623 (2004).

266. W. J. Hucker, C. M. Ripplinger, C. P. Fleming, V. V. Fedorov, A. M. Rollins, and I. R. Efimov, "Bimodal biophotonic imaging of the structure-function relationship in cardiac tissue," J Biomed Opt 13, 054012 (2008).

267. S. A. Boppart, W. Luo, D. L. Marks, and K. W. Singletary, "Optical coherence tomography: feasibility for basic research and image-guided surgery of breast cancer," Breast Cancer Res Treat 84, 85-97 (2004).

268. P. L. Hsiung, D. R. Phatak, Y. Chen, A. D. Aguirre, J. G. Fujimoto, and J. L. Connolly, "Benign and malignant lesion in the human breast depicted with ultrahigh resolution and dimensional optical coherence tomography," Radiology 244, 865-874 (2007).

269. B. D. Goldberg, N. V. Iftimia, J. E. Bressner, M. B. Pitman, E. Halpern, B. E. Bouma, and G. J. Tearney, "Automated algorithm for differentiation of human breast tissue using low coherence interferometry for fine needle aspiration biopsy guidance," J Biomed Opt 13, 014014 (2008).

270. F. T. Nguyen, A. M. Zysk, E. J. Chaney, J. G. Kotynek, U. J. Oliphant, F. J. Bellafiore, K. M. Rowland, P. A. Johnson, and S. A. Boppart, "Intraoperative evaluation of breast tumor margins with optical coherence tomography," Cancer Res 69, 8790-8796 (2009).

271. C. Zhou, D. W. Cohen, Y. Wang, H. C. Lee, A. E. Mondelblatt, T. H. Tsai, A. D. Aguirre, J. G. Fujimoto, and J. L. Connolly, "Integrated optical coherence tomography and microscopy for ex vivo multiscale evaluation of human breast tissues," Cancer Res 70, 10071-10079 (2010).

272. Z. J. Yuan, Z. G. Wang, R. B. Pan, J. X. Liu, H. Cohen, and Y. T. Pan, "High-resolution imaging diagnosis and staging of bladder cancer: comparison between optical coherence tomography and high-frequency ultrasound," Journal of Biomedical Optics 13, 054007 (2008).
273. C. A. Lingley-Papadopoulos, M. H. Loew, M. J. Manyak, and J. M. Zara, "Computer recognition of cancer in the urinary bladder using optical coherence tomography and texture analysis," Journal of Biomedical Optics 13, 024003 (2008).

274. S. P. Lerner, A. C. Goh, N. J. Tresser, and S. S. Shen, "Optical coherence tomography as an adjunct to white light cystoscopy for intravesical real-time imaging and staging of bladder cancer," Urology 72, 133-137 (2008).

275. S. A. Boppart, M. E. Brezinski, C. Pitr is, and J. G. Fujimoto, "Optical coherence tomography for neurosurgical imaging of human intracortical melanoma," Neurosurgery 43, 834-841 (1998).

276. K. Bizheva, A. Unterhuber, B. Hermann, B. Povazay, H. Sattmann, A. F. Fercher, W. Drexler, M. Preusser, H. Budka, A. Stingl, and T. Le, "Imaging ex vivo healthy and pathological human brain tissue with ultra-high-resolution optical coherence tomography," Journal of Biomedical Optics 10, 11006 (2005).

277. H. J. Bohringer, D. Boller, J. Leppert, U. Knopp, E. Lankenau, E. Reusche, G. Huttman, and A. Giese, "Time-domain and spectral-domain optical coherence tomography in the analysis of brain tumor tissue," Lasers in surgery and medicine 38, 588-597 (2006).

278. X. Qi, M. V. Sivak, G. Isenberg, J. E. Willis, and A. M. Rollins, "Computer-aided diagnosis of dysplasia in Barrett's esophagus using endoscopic optical coherence tomography," Journal of Biomedical Optics 11, 044010 (2006).

279. P. A. Testoni, and B. Mangiavillano, "Optical coherence tomography in detection of dysplasia and cancer of the gastrointestinal tract and bilio-pancreatic ductal system," World Journal of Gastroenterology 14, 6444-6452 (2008).

280. S. Lam, B. Standish, C. Baldwin, A. McWilliams, J. leRiche, A. Gazdar, A. I. Vitkin, V. Yang, N. Ikeda, and C. MacAulay, "In vivo optical coherence tomography imaging of preinvasive bronchial lesions," Clin Cancer Res 14, 2006-2011 (2008).

281. P. F. Escobar, L. Rojas-Espaillat, S. Tisci, C. Enerson, J. Brainard, J. Smith, N. J. Tresser, F. I. Feldchtein, L. B. Rojas, and J. L. Belinson, "Optical coherence tomography as a diagnostic aid to visual inspection and colposcopy for preinvasive and invasive cancer of the uterine cervix," International Journal of Gynecological Cancer 16, 1815-1822 (2006).

282. M. Mogensen, T. M. Joergensen, B. M. Nurnberg, H. A. Morsy, J. B. Thomson, L. Thrane, and G. B. E. Jemec, "Assessment of Optical Coherence Tomography Imaging in the Diagnosis of Non-Melanoma Skin Cancer and Benign Lesions Versus Normal Skin: Observer-Blinded Evaluation by Dermatologists and Pathologists," Dermatologic Surgery 35, 965-972 (2009).

283. A. V. Shakhov, A. B. Terentjeva, V. A. Kamensky, L. B. Snopova, V. M. Gelikonov, F. I. Feldchtein, and A. M. Sergeev, "Optical coherence tomography monitoring for laser surgery of laryngeal carcinoma," J Surg Oncol 77, 253-258 (2001).
284. W. B. Armstrong, J. M. Ridgway, D. E. Vokes, S. Guo, J. Perez, R. P. Jackson, M. Gu, J. P. Su, R. L. Crumley, T. Y. Shibuya, U. Mahmood, Z. P. Chen, and B. J. F. Wong, "Optical coherence tomography of laryngeal cancer," Laryngoscope 116, 1107-1113 (2006).

285. W. G. Jung, J. Zhang, J. R. Chung, P. Wilder-Smith, M. Brenner, J. S. Nelson, and Z. P. Chen, "Advances in oral cancer detection using optical coherence tomography," IEEE Journal of Selected Topics in Quantum Electronics 11, 811-817 (2005).

286. M. T. Tsai, H. C. Lee, C. W. Lu, Y. M. Wang, C. K. Lee, C. C. Yang, and C. P. Chiang, "Delineation of an oral cancer lesion with swept-source optical coherence tomography," Journal of Biomedical Optics 13, 044012 (2008).

287. J. M. Poneros, S. Brand, B. E. Bouma, G. J. Tearney, C. C. Compton, and N. S. Nishioka, "Diagnosis of specialized intestinal metaplasia by optical coherence tomography," Gastroenterology 120, 7-12 (2001).

288. B. Shen, G. Zuccaro, Jr, T. L. Gramlich, N. Gladkova, P. Trolli, M. Kareta, C. P. Delaney, J. T. Connor, B. A. Lashner, C. L. Bevins, F. Feldchtein, F. H. Remzi, M. L. Bambrick, and V. W. Fazio, "In vivo colonoscopic optical coherence tomography for transmural inflammation in inflammatory bowel disease," Clin Gastroenterol Hepatol 2, 1080-1087 (2004).

289. H. L. Fu, Y. Leng, M. J. Cobb, K. Hsu, J. H. Hwang, and X. Li, "Flexible miniature compound lens design for high-resolution optical coherence tomography balloon imaging catheter," J Biomed Opt 13, 060502 (2008).

290. M. J. Suter, B. J. Vakoc, P. S. Yachimski, M. Shishkov, G. Y. Lauwers, M. Mino-Kenudson, B. E. Bouma, N. S. Nishioka, and G. J. Tearney, "Comprehensive microscopy of the esophagus in human patients with optical frequency domain imaging," Gastrointestinal endoscopy 68, 745-753 (2008).

291. D. C. Adler, C. Zhou, T. H. Tsai, J. Schmitt, Q. Huang, H. Mashimo, and J. G. Fujimoto, "Three-dimensional endomicroscopy of the human colon using optical coherence tomography," Opt Express 17, 784-796 (2009).

292. A. M. Zysk, and S. A. Boppart, "Computational methods for analysis of human breast tumor tissue in optical coherence tomography images," Journal of Biomedical Optics 11, 054015 (2006).

293. J. A. Linehan, E. R. Bracamonte, L. P. Hariri, M. H. Sokoloff, P. S. Rice, J. K. Barton, and M. M. Nguyen, "Feasibility of optical coherence tomography imaging to characterize renal neoplasms: limitations in resolution and depth of penetration," BJU Int (2011).

294. K. Barwari, D. M. de Bruin, E. C. Cauberg, D. J. Faber, T. G. van Leeuwen, H. Wijkstra, J. de la Rosette, and M. P. Laguna, "Advanced diagnostics in renal mass using optical coherence tomography: a preliminary report," J Endourol 25, 311-315 (2011).
295. R. V. Kuranov, V. V. Sapozhnikova, H. M. Shakhova, V. M. Gelikonov, E. V. Zagainova, and S. A. Petrova, "Combined application of optical methods to increase the information content of optical coherent tomography in diagnostics of neoplastic processes," Quantum Electronics 32, 993-998 (2002).

296. V. V. Sapozhnikova, N. M. Shakhova, V. A. Kamensky, S. A. Petrova, L. B. Snopova, and R. V. Kuranov, "Capabilities of fluorescence spectroscopy using 5-ALA and optical coherence tomography for diagnosis of neoplastic processes in the uterine cervix and vulva," Laser Phys 15, 1664-1673 (2005).

297. A. R. Tumlinson, L. P. Hariri, U. Utzinger, and J. K. Barton, "Miniature endoscope for simultaneous optical coherence tomography and laser-induced fluorescence measurement," Applied optics 43, 113-121 (2004).

298. J. B. McNally, N. D. Kirkpatrick, L. P. Hariri, A. R. Tumlinson, D. G. Besselsen, E. W. Gerner, U. Utzinger, and J. K. Barton, "Task-based imaging of colon cancer in the Apc(Min+/+) mouse model," Applied optics 45, 3049-3062 (2006).

299. L. P. Hariri, A. R. Tumlinson, D. G. Besselsen, U. Utzinger, E. W. Gerner, and J. K. Barton, "Endoscopic optical coherence tomography and laser-induced fluorescence spectroscopy in a murine colon cancer model," Lasers in surgery and medicine 38, 305-313 (2006).

300. L. P. Hariri, A. R. Tumlinson, N. H. Wade, D. G. Besselsen, U. Utzinger, E. W. Gerner, and J. K. Barton, "Ex vivo optical coherence tomography and laser-induced fluorescence spectroscopy imaging of murine gastrointestinal tract," Comparative Med. 57, 175-185 (2007).

301. Y. T. Pan, T. Q. Xie, C. W. Du, S. Bastacky, S. Meyers, and M. L. Zeidel, "Enhancing early bladder cancer detection with fluorescence-guided endoscopic optical coherence tomography," Optics letters 28, 2485-2487 (2003).

302. Z. G. Wang, D. B. Durand, M. Schoenberg, and Y. T. Pan, "Fluorescence guided optical coherence tomography for the diagnosis of early bladder cancer in a rat model," J Urology 174, 2376-2381 (2005).

303. W. Drexler, D. Stamper, C. Jesser, X. Li, C. Pitris, K. Saunders, S. Martin, M. B. Lodge, J. G. Fujimoto, and M. E. Brezinski, "Correlation of collagen organization with polarization sensitive imaging of in vitro cartilage: implications for osteoarthritis," The Journal of rheumatology 28, 1311-1318 (2001).

304. Y. Pan, Z. Li, T. Xie, and C. R. Chu, "Hand-held arthroscopic optical coherence tomography for in vivo high-resolution imaging of articular cartilage," Journal of biomedical optics 8, 648-654 (2003).
305. X. Li, S. Martin, C. Pitris, R. Ghanta, D. L. Stamper, M. Harman, J. G. Fujimoto, and M. E. Brezinski, "High-resolution optical coherence tomographic imaging of osteoarthritic cartilage during open knee surgery," Arthritis Res Ther 7, R318-323 (2005).

306. T. Xie, S. Guo, J. Zhang, Z. Chen, and G. M. Peavy, "Determination of characteristics of degenerative joint disease using optical coherence tomography and polarization sensitive optical coherence tomography," Lasers in surgery and medicine 38, 852-865 (2006).

307. B. T. Amaechi, A. G. Podoleanu, G. N. Komarov, J. A. Rogers, S. M. Higham, and D. A. Jackson, "Application of optical coherence tomography for imaging and assessment of early dental caries lesions," Laser Phys 13, 703-710 (2003).

308. A. K. Braz, B. B. Kyotoku, R. Braz, and A. S. Gomes, "Evaluation of crack propagation in dental composites by optical coherence tomography," Dent Mater 25, 74-79 (2009).

309. R. S. Jones, C. L. Darling, J. D. Featherstone, and D. Fried, "Remineralization of in vitro dental caries assessed with polarization-sensitive optical coherence tomography," J Biomed Opt 11, 014016 (2006).

310. H. Kakuma, K. Ohbayashi, and Y. Arakawa, "Optical imaging of hard and soft dental tissues using discretely swept optical frequency domain reflectometry optical coherence tomography at wavelengths from 1560 to 1600 nm," J Biomed Opt 13, 014012 (2008).

311. L. L. Otis, M. J. Everett, U. S. Sathyam, and B. W. Colston, Jr., "Optical coherence tomography: a new imaging technology for dentistry," J Am Dent Assoc 131, 511-514 (2000).

312. C. Sinescu, M. L. Negrutiu, C. Todea, C. Balabuc, L. Filip, R. Rominu, A. Bradu, M. Hughes, and A. G. Podoleanu, "Quality assessment of dental treatments using en-face optical coherence tomography," J Biomed Opt 13, 054065 (2008).

313. C. H. Wilder-Smith, P. Wilder-Smith, H. Kawakami-Wong, J. Voronets, K. Osann, and A. Lussi, "Quantification of Dental Erosions in Patients With GERD Using Optical Coherence Tomography Before and After Double-Blind, Randomized Treatment With Esomeprazole or Placebo," The American journal of gastroenterology (2009).

314. M. S. Jafri, S. Farhang, R. S. Tang, N. Desai, P. S. Fishman, R. G. Rohwer, C. M. Tang, and J. M. Schmitt, "Optical coherence tomography in the diagnosis and treatment of neurological disorders," J Biomed Opt 10, 051603 (2005).

315. Q. Li, M. L. Onozato, P. M. Andrews, C. W. Chen, A. Paek, R. Naphas, S. Yuan, J. Jiang, A. Cable, and Y. Chen, "Automated quantification of microstructural dimensions of the human kidney using optical coherence tomography (OCT)," Opt Express 17, 16000-16016 (2009).

316. M. L. Onozato, P. M. Andrews, Q. Li, J. Jiang, A. Cable, and Y. Chen, "Optical coherence tomography of human kidney," The Journal of urology 183, 2090-2094 (2010).
317. M. Panjehpour, B. F. Overholt, T. VoDinh, R. C. Haggitt, D. H. Edwards, and F. P. Buckley, "Endoscopic fluorescence detection of high-grade dysplasia in Barrett's esophagus," Gastroenterology 111, 93-101 (1996).

318. R. M. Cothren, M. V. Sivak, J. VanDam, R. E. Petras, M. Fitzmaurice, J. M. Crawford, J. Wu, J. F. Brennan, R. P. Rava, R. Manoharan, and M. S. Feld, "Detection of dysplasia at colonoscopy using laser-induced fluorescence: A blinded study," Gastrointest Endosc 44, 168-176 (1996).

319. G. I. Zonios, R. M. Cothren, J. T. Arendt, J. Wu, J. VanDam, J. M. Crawford, R. Manoharan, and M. S. Feld, "Morphological model of human colon tissue fluorescence," Ieee Transactions on Biomedical Engineering 43, 113-122 (1996).

320. K. T. Schomacker, J. K. Frisoli, C. C. Compton, T. J. Flotte, J. M. Richter, N. S. Nishioka, and T. F. Deutsch, "Ultraviolet Laser-Induced Fluorescence of Colonic Tissue - Basic Biology and Diagnostic Potential," Lasers in surgery and medicine 12, 63-78 (1992).

321. R. S. Dacosta, B. C. Wilson, and N. E. Marcon, "New optical technologies for earlier endoscopic diagnosis of premalignant gastrointestinal lesions," Journal of gastroenterology and hepatology 17 Suppl, S85-104 (2002).

322. T. Desmettre, J. M. Devoisselle, and S. Mordon, "Fluorescence properties and metabolic features of indocyanine green (ICG) as related to angiography," Survey of ophthalmology 45, 15-27 (2000).

323. A. Corlu, R. Choe, T. Durduran, M. A. Rosen, M. Schweiger, S. R. Arridge, M. D. Schnall, and A. G. Yodh, "Three-dimensional in vivo fluorescence diffuse optical tomography of breast cancer in humans," Optics Express 15, 6696-6716 (2007).

324. S. W. Kang, C. Y. Park, and D. I. Ham, "The correlation between fluorescein angiographic and optical coherence tomographic features in clinically significant diabetic macular edema," Am J Ophthalmol 137, 313-322 (2004).

325. H. Messmann, E. Endlicher, G. Freunek, P. Rummele, J. Scholmerich, and R. Knuchel, "Fluorescence endoscopy for the detection of low and high grade dysplasia in ulcerative colitis using systemic or local 5-aminolaevulinic acid sensitisation," Gut 52, 1003-1007 (2003).

326. E. Endlicher, R. Knuchel, T. Hauser, R. M. Szeimies, J. Scholmerich, and H. Messmann, "Endoscopic fluorescence detection of low and high grade dysplasia in Barrett's oesophagus using systemic or local 5-aminolaevulinic acid sensitisation," Gut 48, 314-319 (2001).

327. H. Messmann, R. Knuchel, W. Baumler, A. Holstege, and J. Scholmerich, "Endoscopic fluorescence detection of dysplasia in patients with Barrett's esophagus, ulcerative colitis, or adenomatous polyps after 5-aminolevulinic acid-induced protoporphyrin IX sensitization," Gastrointest Endosc 49, 97-101 (1999).
328. D. Roblyer, R. A. Schwarz, and R. Richards-Kortum, "Fluorescence Spectroscopy," in *Handbook of Biomedical Optics*, D. A. Boas, C. Pitris, and N. Ramanujam, eds. (CRC Press, Boca Raton, FL, 2011), pp. 217-232.

329. B. C. Wilson, "Detection and treatment of dysplasia in Barrett's esophagus: a pivotal challenge in translating biophotonics from bench to bedside," J Biomed Opt *12*, 051401 (2007).

330. W. Yu, R. M. Sandoval, and B. A. Molitoris, "Rapid determination of renal filtration function using an optical ratiometric imaging approach," American journal of physiology *292*, F1873-1880 (2007).

331. V. E. Centonze, and J. G. White, "Multiphoton excitation provides optical sections from deeper within scattering specimens than confocal imaging," Biophysical journal *75*, 2015-2024 (1998).

332. V. Ntziachristos, C. H. Tung, C. Bremer, and R. Weissleder, "Fluorescence molecular tomography resolves protease activity in vivo," Nature medicine *8*, 757-760 (2002).

333. P. R. Pfau, and M. V. Sivak, "Endoscopic diagnostics," Gastroenterology *120*, 763-781 (2001).

334. G. Bourg-Heckly, J. Blais, J. J. Padilla, O. Bourdon, J. Etienne, F. Guillemin, and L. Lafay, "Endoscopic ultraviolet-induced autofluorescence spectroscopy of the esophagus: Tissue characterization and potential for early cancer diagnosis," Endoscopy *32*, 756-765 (2000).

335. K. Izuishi, H. Tajiri, T. Fujii, N. Boku, A. Ohtsu, T. Ohnishi, M. Ryu, T. Kinoshita, and S. Yoshida, "The histological basis of detection of adenoma and cancer in the colon by autofluorescence endoscopic imaging," Endoscopy *31*, 511-516 (1999).

336. B. Mayinger, P. Horner, M. Jordan, C. Gerlach, T. Horbach, W. Hohenberger, and E. G. Hahn, "Light-induced autofluorescence spectroscopy for the endoscopic detection of esophageal cancer," Gastrointest Endosc *54*, 195-201 (2001).

337. B. Mayinger, M. Jordan, T. Horbach, P. Horner, C. Gerlach, S. Mueller, W. Hohenberger, and E. G. Hahn, "Evaluation of in vivo endoscopic autofluorescence spectroscopy in gastric cancer," Gastrointest Endosc *59*, 191-198 (2004).

338. R. Richards-Kortum, R. P. Rava, R. E. Petras, M. Fitzmaurice, M. Sivak, and M. S. Feld, "Spectroscopic Diagnosis of Colonic Dysplasia," Photochemistry and photobiology *53*, 777-786 (1991).

339. B. Mayinger, M. Jordan, P. Horner, C. Gerlach, S. Muehldorfer, B. R. Bittorf, K. E. Matzel, W. Hohenberger, E. G. Hahn, and K. Guenther, "Endoscopic light-induced autofluorescence spectroscopy for the diagnosis of colorectal cancer and adenoma," Journal of Photochemistry and Photobiology B-Biology *70*, 13-20 (2003).
340. J. Wu, M. S. Feld, and R. P. Rava, "Analytical Model for Extracting Intrinsic Fluorescence in Turbid Media," Applied optics 32, 3585-3595 (1993).

341. Q. G. Zhang, M. G. Muller, J. Wu, and M. S. Feld, "Turbidity-free fluorescence spectroscopy of biological tissue," Optics letters 25, 1451-1453 (2000).

342. I. Georgakoudi, B. C. Jacobson, J. Van Dam, V. Backman, M. B. Wallace, M. G. Muller, Q. Zhang, K. Badizadegan, D. Sun, G. A. Thomas, L. T. Perelman, and M. S. Feld, "Fluorescence, reflectance, and light-scattering spectroscopy for evaluating dysplasia in patients with Barrett's esophagus," Gastroenterology 120, 1620-1629 (2001).

343. Q. Peng, T. Warloe, K. Berg, J. Moan, M. Kongshaug, K. E. Giercksky, and J. M. Nesland, "5-aminolevulinic acid-based photodynamic therapy - Clinical research and future challenges," Cancer 79, 2282-2308 (1997).

344. S. Brand, T. D. Wang, K. T. Schomacker, J. M. Poneros, G. Y. Lauwers, C. C. Compton, M. C. Pedrosa, and N. S. Nishioka, "Detection of high-grade dysplasia in Barrett's esophagus by spectroscopy measurement of 5-aminolevulinic acid-induced protoporphyrin IX fluorescence," Gastrointest Endosc 56, 479-487 (2002).

345. M. A. Ortner, B. Ebert, E. Hein, K. Zumbusch, D. Nolte, U. Sukowski, J. Weber-Eibel, B. Fleige, M. Dietel, M. Stolte, G. Oberhuber, R. Porschen, B. Klump, H. Hortnagl, H. Lochs, and H. Rinneberg, "Time gated fluorescence spectroscopy in Barrett's oesophagus," Gut 52, 28-33 (2003).

346. T. D. Wang, J. VanDam, J. M. Crawford, E. A. Preisinger, Y. Wang, and M. S. Feld, "Fluorescence endoscopic imaging of human colonic adenomas," Gastroenterology 111, 1182-1191 (1996).

347. H. S. Zeng, A. Weiss, R. Cline, and C. E. MacAulay, "Real-time endoscopic fluorescence imaging for early cancer detection in the gastrointestinal tract," Bioimaging 6, 151-165 (1998).

348. J. Haringsma, G. N. J. Tytgat, H. Yano, H. Iishi, M. Tatsuta, T. Ogihara, H. Watanabe, N. Sato, N. Marcon, B. C. Wilson, and R. W. Cline, "Autofluorescence endoscopy: feasibility of detection of GI neoplasms unapparent to white light endoscopy with an evolving technology," Gastrointest Endosc 53, 642-650 (2001).

349. M. A. Kara, F. P. Peters, F. J. W. ten Kate, S. J. van Deventer, P. Fockens, and J. J. G. H. M. Bergman, "Endoscopic video autofluorescence imaging may improve the detection of early neoplasia in patients with Barrett's esophagus," Gastrointest Endosc 61, 679-685 (2005).

350. T. D. Wang, J. M. Crawford, M. S. Feld, Y. Wang, I. Itzkan, and J. Van Dam, "In vivo identification of colonic dysplasia using fluorescence endoscopic imaging," Gastrointest Endosc 49, 447-455 (1999).
351. S. Lam, T. Kennedy, M. Unger, Y. E. Miller, D. Gelmont, V. Rusch, B. Gipe, D. Howard, J. C. LeRiche, A. Coldman, and A. F. Gazdar, "Localization of bronchial intraepithelial neoplastic lesions by fluorescence bronchoscopy," Chest 113, 696-702 (1998).

352. K. Niepsuj, G. Niepsuj, W. Cebula, W. Zieleznik, M. Adamek, A. Sielanczyk, J. Adameczyk, J. Kurek, and A. Sieron, "Autofluorescence endoscopy for detection of high-grade dysplasia in short-segment Barrett's esophagus," Gastrointest Endosc 58, 715-719 (2003).

353. M. A. Kara, M. E. Smits, W. D. Rosmolen, A. C. Bultje, F. J. W. ten Kate, P. Fockens, G. N. J. Tytgat, and J. J. G. H. M. Bergman, "A randomized crossover study comparing light-induced fluorescence endoscopy with standard videendoscopy for the detection of early neoplasia in Barrett's esophagus," Gastrointest Endosc 61, 671-678 (2005).

354. T. Stepinae, C. Felley, P. Jornod, N. Lange, T. Gabrecht, C. Fontolliet, P. Grosjean, G. vanMelle, H. van den Bergh, P. Monnier, G. Wagnieres, and G. Dorta, "Endoscopic fluorescence detection of intraepithelial neoplasia in Barrett's esophagus after oral administration of aminolevulinic acid," Endoscopy 35, 663-668 (2003).

355. T. Yoshida, H. Inoue, S. Usui, H. Satodate, N. Fukami, and S. E. Kudo, "Narrow-band imaging system with magnifying endoscopy for superficial esophageal lesions," Gastrointestinal endoscopy 59, 288-295 (2004).

356. K. Gono, T. Obi, M. Yamaguchi, N. Ohyama, H. Machida, Y. Sano, S. Yoshida, Y. Hamamoto, and T. Endo, "Appearance of enhanced tissue features in narrow-band endoscopic imaging," J Biomed Opt 9, 568-577 (2004).

357. M. A. Kara, and J. J. G. H. M. Bergman, "Autofluorescence imaging and narrow-band imaging for the detection of early neoplasia in patients with Barrett's esophagus," Endoscopy 38, 627-631 (2006).

358. Y. Hamamoto, T. Endo, K. Nosho, Y. Arimura, M. Sato, and K. Imai, "Usefulness of narrow-band imaging endoscopy for diagnosis of Barrett's esophagus," J Gastroenterol 39, 14-20 (2004).

359. M. A. Kara, F. P. Peters, P. Fockens, F. J. W. ten Kate, and J. J. G. H. M. Bergman, "Endoscopic video-autofluorescence imaging followed by narrow band imaging for detecting early neoplasia in Barrett's esophagus," Gastrointest Endosc 64, 176-185 (2006).

360. M. D. Keller, S. K. Majumder, M. C. Kelley, I. M. Meszoely, F. I. Boulos, G. M. Olivares, and A. Mahadevan-Jansen, "Autofluorescence and diffuse reflectance spectroscopy and spectral imaging for breast surgical margin analysis," Lasers in surgery and medicine 42, 15-23 (2010).

361. W. Stummer, U. Pichlmeier, T. Meinel, O. D. Wiestler, F. Zanella, and H. J. Reulen, "Fluorescence-guided surgery with 5-aminolevulinic acid for resection of malignant glioma: a randomised controlled multicentre phase III trial," Lancet Oncol 7, 392-401 (2006).
362. M. Hefti, G. von Campe, M. Moschopoulos, A. Siegner, H. Looser, and H. Landolt, "5-aminolevulinic acid induced protoporphyrin IX fluorescence in high-grade glioma surgery: a one-year experience at a single institution," Swiss Med Wkly 138, 180-185 (2008).

363. W. Stummer, S. Stocker, S. Wagner, H. Stepp, C. Fritsch, C. Goetz, A. E. Goetz, R. Kiefmann, and H. J. Reulen, "Intraoperative detection of malignant gliomas by 5-aminolevulinic acid-induced porphyrin fluorescence," Neurosurgery 42, 518-525 (1998).

364. D. W. Roberts, P. A. Valdes, B. T. Harris, K. M. Fontaine, A. Hartov, X. Fan, S. Ji, S. S. Lollis, B. W. Pogue, F. Leblond, T. D. Tosteson, B. C. Wilson, and K. D. Paulsen, "Coregistered fluorescence-enhanced tumor resection of malignant glioma: relationships between delta-aminolevulinic acid-induced protoporphyrin IX fluorescence, magnetic resonance imaging enhancement, and neuropathological parameters. Clinical article," J Neurosurg 114, 595-603 (2010).

365. P. A. Valdes, F. Leblond, A. Kim, B. T. Harris, B. C. Wilson, X. Fan, T. D. Tosteson, A. Hartov, S. Ji, K. Erkmen, N. E. Simmons, K. D. Paulsen, and D. W. Roberts, "Quantitative fluorescence in intracranial tumor: implications for ALA-induced PpIX as an intraoperative biomarker," J Neurosurg 115, 11-17 (2010).

366. S. L. Troyan, V. Kianzad, S. L. Gibbs-Strauss, S. Gioux, A. Matsui, R. Oketokoun, L. Ngo, A. Khamene, F. Azar, and J. V. Frangioni, "The FLARE intraoperative near-infrared fluorescence imaging system: a first-in-human clinical trial in breast cancer sentinel lymph node mapping," Ann Surg Oncol 16, 2943-2952 (2009).

367. T. Kitai, T. Inomoto, M. Miwa, and T. Shikayama, "Fluorescence navigation with indocyanine green for detecting sentinel lymph nodes in breast cancer," Breast Cancer 12, 211-215 (2005).

368. N. Tagaya, H. Aoyagi, A. Nakagawa, A. Abe, Y. Iwasaki, M. Tachibana, and K. Kubota, "A novel approach for sentinel lymph node identification using fluorescence imaging and image overlay navigation surgery in patients with breast cancer," World J Surg 35, 154-158 (2011).

369. N. Tagaya, R. Yamazaki, A. Nakagawa, A. Abe, K. Hamada, K. Kubota, and T. Oyama, "Intraoperative identification of sentinel lymph nodes by near-infrared fluorescence imaging in patients with breast cancer," American Journal of Surgery 195, 850-853 (2008).

370. E. M. Sevick-Muraca, R. Sharma, J. C. Rasmussen, M. V. Marshall, J. A. Wendt, H. Q. Pham, E. Bonefas, J. P. Houston, L. Sampath, K. E. Adams, D. K. Blanchard, R. E. Fisher, S. B. Chiang, R. Elledge, and M. E. Mawad, "Imaging of lymph flow in breast cancer patients after microdose administration of a near-infrared fluorophore: feasibility study," Radiology 246, 734-741 (2008).

371. T. Kitai, and M. Kawashima, "Transcutaneous detection and direct approach to the sentinel node using axillary compression technique in ICG fluorescence-navigated sentinel node biopsy for breast cancer," Breast Cancer (in press).
372. M. Fujiwara, T. Mizukami, A. Suzuki, and H. Fukamizu, "Sentinel lymph node detection in skin cancer patients using real-time fluorescence navigation with indocyanine green: preliminary experience," J Plast Reconstr Aesthet Surg 62, e373-378 (2009).

373. Y. Fujisawa, Y. Nakamura, Y. Kawachi, and F. Otsuka, "A custom-made, low-cost intraoperative fluorescence navigation system with indocyanine green for sentinel lymph node biopsy in skin cancer," Dermatology 222, 261-268 (2011).

374. W. Kelder, H. Nimura, N. Takahashi, N. Mitsumori, G. M. van Dam, and K. Yanaga, "Sentinel node mapping with indocyanine green (ICG) and infrared ray detection in early gastric cancer: an accurate method that enables a limited lymphadenectomy," Eur J Surg Oncol 36, 552-558 (2010).

375. I. Miyashiro, K. Kishi, M. Yano, K. Tanaka, M. Motoori, M. Ohue, H. Ohigashi, A. Takenaka, Y. Tomita, and O. Ishikawa, "Laparoscopic detection of sentinel node in gastric cancer surgery by indocyanine green fluorescence imaging," Surg Endosc 25, 1672-1676 (2011).

376. S. Noura, M. Ohue, Y. Seki, K. Tanaka, M. Motoori, K. Kishi, I. Miyashiro, H. Ohigashi, M. Yano, O. Ishikawa, and Y. Miyamoto, "Feasibility of a lateral region sentinel node biopsy of lower rectal cancer guided by indocyanine green using a near-infrared camera system," Ann Surg Oncol 17, 144-151 (2010).

377. T. Kitai, M. Kawashima, H. Fujii, S. Mashima, and Y. Shimahara, "Indocyanine Green Fluorescence Monitoring of Perineal Wound Contamination in Abdominoperineal Resection: A Preliminary Report," Surgery Today 41, 1037-1040 (2011).

378. J. C. Rasmussen, I. C. Tan, M. V. Marshall, K. E. Adams, S. Kwon, C. E. Fife, E. A. Maus, L. A. Smith, K. R. Covington, and E. M. Sevick-Muraca, "Human Lymphatic Architecture and Dynamic Transport Imaged Using Near-infrared Fluorescence," Transl Oncol 3, 362-372 (2010).

379. J. C. Rasmussen, I. C. Tan, M. V. Marshall, C. E. Fife, and E. M. Sevick-Muraca, "Lymphatic imaging in humans with near-infrared fluorescence," Current Opinion in Biotechnology 20, 74-82 (2009).

380. R. Sharma, W. Wang, J. C. Rasmussen, A. Joshi, J. P. Houston, K. E. Adams, A. Cameron, S. Ke, S. Kwon, M. E. Mawad, and E. M. Sevick-Muraca, "Quantitative imaging of lymph function," Am J Physiol Heart Circ Physiol 292, H3109-3118 (2007).

381. R. Sharma, J. A. Wendt, J. C. Rasmussen, K. E. Adams, M. V. Marshall, and E. M. Sevick-Muraca, "New horizons for imaging lymphatic function," Ann N Y Acad Sci 1131, 13-36 (2008).

382. I. C. Tan, E. A. Maus, J. C. Rasmussen, M. V. Marshall, K. E. Adams, C. E. Fife, L. A. Smith, W. Chan, and E. M. Sevick-Muraca, "Assessment of lymphatic contractile function
after manual lymphatic drainage using near-infrared fluorescence imaging," Arch Phys Med Rehabil 92, 756-764 e751 (2011).

383. B. B. Das, F. Liu, and R. R. Alfano, "Time-resolved fluorescence and photon migration studies in biomedical and model random media," Reports on Progress in Physics 60, 227-292 (1997).

384. R. Cubeddu, D. Comelli, C. D'Andrea, P. Taroni, and G. Valentini, "Time-resolved fluorescence imaging in biology and medicine," Journal of Physics D-Applied Physics 35, R61-R76 (2002).

385. L. Marcu, "Fluorescence lifetime in cardiovascular diagnostics," Journal of Biomedical Optics 15 (2010).

386. L. Marcu, Y. Sun, J. Phipps, D. S. Elson, H. Stoy, S. Tinling, J. Meier, B. Poirier, F. S. Chuang, and D. G. Farwell, "Fluorescence lifetime imaging microscopy: in vivo application to diagnosis of oral carcinoma," Optics letters 34, 2081-2083 (2009).

387. J. D. Meier, H. Xie, Y. Sun, N. Hatami, B. Poirier, L. Marcu, and D. G. Farwell, "Time-resolved laser-induced fluorescence spectroscopy as a diagnostic instrument in head and neck carcinoma," Otolaryngol Head Neck Surg 142, 838-844 (2010).

388. L. Marcu, Y. H. Sun, N. Hatami, M. Yee, J. Phipps, D. S. Elson, F. Gorin, and R. J. Schrot, "Fluorescence lifetime imaging microscopy for brain tumor image-guided surgery," Journal of Biomedical Optics 15, 056022 (2010).

389. R. Weissleder, and U. Mahmood, "Molecular imaging," Radiology 219, 316-333 (2001).

390. C. H. Contag, and M. H. Bachmann, "Advances in vivo bioluminescence imaging of gene expression," Annu Rev Biomed Eng 4, 235-260 (2002).

391. T. F. Massoud, and S. S. Gambhir, "Molecular imaging in living subjects: seeing fundamental biological processes in a new light," Gene Dev 17, 545-580 (2003).

392. U. Mahmood, and R. Weissleder, "Near-infrared optical imaging of proteases in cancer," Mol Cancer Ther 2, 489-496 (2003).

393. R. G. Blasberg, "Molecular imaging and cancer," Mol Cancer Ther 2, 335-343 (2003).

394. S. Achilefu, "Lighting up tumors with receptor-specific optical molecular probes," Technology in cancer research & treatment 3, 393-409 (2004).

395. R. Weissleder, "Molecular Imaging in Cancer," Science 312, 1168-1171 (2006).

396. K. Kelly, H. Alencar, M. Funovics, U. Mahmood, and R. Weissleder, "Detection of invasive colon cancer using a novel, targeted, library-derived fluorescent peptide," Cancer Res 64, 6247-6251 (2004).
397. K. Marten, C. Bremer, K. Khazaie, M. Sameni, B. Sloane, C. H. Tung, and R. Weissleder, "Detection of dysplastic intestinal adenomas using enzyme-sensing molecular beacons in mice," Gastroenterology 122, 406-414 (2002).

398. W. J. Akers, and S. Achilefu, "Molecular Reporter Systems for Optical Imaging," in Handbook of Biomedical Optics, D. A. Boas, C. Pitris, and N. Ramanujam, eds. (CRC Press, Boca Raton, FL, 2011), pp. 673-696.

399. K. P. Withrow, J. R. Newman, J. B. Skipper, J. P. Gleysteen, J. S. Magnuson, K. Zinn, and E. L. Rosenthal, "Assessment of bevacizumab conjugated to Cy5.5 for detection of head and neck cancer xenografts," Technology in cancer research & treatment 7, 61-66 (2008).

400. J. Virostko, J. Xie, D. E. Hallahan, C. L. Arteaga, J. C. Gore, and H. C. Manning, "A molecular imaging paradigm to rapidly profile response to angiogenesis-directed therapy in small animals," Mol Imaging Biol 11, 204-212 (2009).

401. M. V. Backer, Z. Levashova, V. Patel, B. T. Jehning, K. Claffey, F. G. Blankenberg, and J. M. Backer, "Molecular imaging of VEGF receptors in angiogenic vasculature with single-chain VEGF-based probes," Nature medicine 13, 504-509 (2007).

402. H. Wang, K. Chen, G. Niu, and X. Chen, "Site-specifically biotinylated VEGF(121) for near-infrared fluorescence imaging of tumor angiogenesis," Mol Pharm 6, 285-294 (2009).

403. S. Ke, X. Wen, M. Gurfinkel, C. Charmsangavej, S. Wallace, E. M. Sevick-Muraca, and C. Li, "Near-infrared optical imaging of epidermal growth factor receptor in breast cancer xenografts," Cancer Res 63, 7870-7875 (2003).

404. J. L. Kovar, M. A. Johnson, W. M. Volcheck, J. Chen, and M. A. Simpson, "Hyaluronidase expression induces prostate tumor metastasis in an orthotopic mouse model," Am J Pathol 169, 1415-1426 (2006).

405. A. Becker, C. Hessenius, S. Bhargava, C. Grotzinger, K. Licha, J. Schneider-Mergener, B. Wiedenmann, and W. Semmler, "Cyanine dye labeled vasoactive intestinal peptide and somatostatin analog for optical detection of gastroenteropancreatic tumors," Ann NY Acad Sci 921, 275-278 (2000).

406. A. Becker, C. Hessenius, K. Licha, B. Ebert, U. Sukowski, W. Semmler, B. Wiedenmann, and C. Grotzinger, "Receptor-targeted optical imaging of tumors with near-infrared fluorescent ligands," Nat Biotechnol 19, 327-331 (2001).

407. S. Achilefu, R. B. Dorshow, J. E. Bugaj, and R. Rajagopalan, "Novel receptor-targeted fluorescent contrast agents for in vivo tumor imaging," Investigative radiology 35, 479-485 (2000).
408. N. Nitin, A. L. Carlson, T. Muldoon, A. K. El-Naggar, A. Gillenwater, and R. Richards-Kortum, "Molecular imaging of glucose uptake in oral neoplasia following topical application of fluorescently labeled deoxy-glucose," Int J Cancer 124, 2634-2642 (2009).

409. Y. Chen, G. Zheng, Z. H. Zhang, D. Blessington, M. Zhang, H. Li, Q. Liu, L. Zhou, X. Intes, S. Achilefu, and B. Chance, "Metabolism-enhanced tumor localization by fluorescence imaging: in vivo animal studies," Optics letters 28, 2070-2072 (2003).

410. G. Zheng, Y. Chen, X. Intes, B. Chance, and J. D. Glickson, "Contrast-enhanced near-infrared (NIR) optical imaging for subsurface cancer detection," Journal of Porphyrins and Phthalocyanines 8, 1106-1117 (2004).

411. G. Kostenich, N. Livnah, T. A. Bonasera, T. Yechezkel, Y. Salitra, P. Litman, S. Kimel, and A. Orenstein, "Targeting small-cell lung cancer with novel fluorescent analogs of somatostatin," Lung Cancer 50, 319-328 (2005).

412. G. Kostenich, M. Oron-Herman, S. Kimel, N. Livnah, I. Tsarfaty, and A. Orenstein, "Diagnostic targeting of colon cancer using a novel fluorescent somatostatin conjugate in a mouse xenograft model," Int J Cancer 122, 2044-2049 (2008).

413. X. Chen, P. S. Conti, and R. A. Moats, "In vivo near-infrared fluorescence imaging of integrin alphavbeta3 in brain tumor xenografts," Cancer Res 64, 8009-8014 (2004).

414. S. Achilefu, S. Bloch, M. A. Markiewicz, T. Zhong, Y. Ye, R. B. Dorshow, B. Chance, and K. Liang, "Synergistic effects of light-emitting probes and peptides for targeting and monitoring integrin expression," Proceedings of the National Academy of Sciences of the United States of America 102, 7976-7981 (2005).

415. Z. Cheng, Y. Wu, Z. Xiong, S. S. Gambhir, and X. Chen, "Near-infrared fluorescent RGD peptides for optical imaging of integrin alphavbeta3 expression in living mice," Bioconjug Chem 16, 1433-1441 (2005).

416. S. Kwon, S. Ke, J. P. Houston, W. Wang, Q. Wu, C. Li, and E. M. Sevick-Muraca, "Imaging dose-dependent pharmacokinetics of an RGD-fluorescent dye conjugate targeted to alpha v beta 3 receptor expressed in Kaposi's sarcoma," Mol Imaging 4, 75-87 (2005).

417. Y. Ye, S. Bloch, B. Xu, and S. Achilefu, "Design, synthesis, and evaluation of near infrared fluorescent multimeric RGD peptides for targeting tumors," J Med Chem 49, 2268-2275 (2006).

418. Z. H. Jin, J. Razkin, V. Josserand, D. Boturyn, A. Grichine, I. Texier, M. C. Favrot, P. Dumy, and J. L. Coll, "In vivo noninvasive optical imaging of receptor-mediated RGD internalization using self-quenched Cy5-labeled RAFT-c(-RGDFK-)(4)," Mol Imaging 6, 43-55 (2007).

419. B. R. Smith, Z. Cheng, A. De, A. L. Koh, R. Sinclair, and S. S. Gambhir, "Real-time intravital imaging of RGD-quantum dot binding to luminal endothelium in mouse tumor neovasculature," Nano letters 8, 2599-2606 (2008).
420. W. J. Mulder, K. Castermans, J. R. van Beijnum, M. G. Oude Egbrink, P. T. Chin, Z. A. Fayad, C. W. Lowik, E. L. Kaijzel, I. Que, G. Storm, G. J. Strijkers, A. W. Griffioen, and K. Nicolay, "Molecular imaging of tumor angiogenesis using alphavbeta3-integrin targeted multimodal quantum dots," Angiogenesis 12, 17-24 (2009).

421. J. E. Bugaj, S. Achilefu, R. B. Dorshow, and R. Rajagopalan, "Novel fluorescent contrast agents for optical imaging of in vivo tumors based on a receptor-targeted dye-peptide conjugate platform," J Biomed Opt 6, 122-133 (2001).

422. S. Achilefu, H. N. Jimenez, R. B. Dorshow, J. E. Bugaj, E. G. Webb, R. R. Wilhelm, R. Rajagopalan, J. Johler, and J. L. Erion, "Synthesis, in vitro receptor binding, and in vivo evaluation of fluorescein and carbocyanine peptide-based optical contrast agents," J Med Chem 45, 2003-2015 (2002).

423. L. Ma, P. Yu, B. Veerendra, T. L. Rold, L. Retzloff, A. Prasanphanich, G. Sieckman, T. J. Hoffman, W. A. Volkert, and C. J. Smith, "In vitro and in vivo evaluation of Alexa Fluor 680-bombesin[7-14]NH2 peptide conjugate, a high-affinity fluorescent probe with high selectivity for the gastrin-releasing peptide receptor," Mol Imaging 6, 171-180 (2007).

424. E. I. Sega, and P. S. Low, "Tumor detection using folate receptor-targeted imaging agents," Cancer metastasis reviews 27, 655-664 (2008).

425. J. A. Reddy, V. M. Allagadda, and C. P. Leamon, "Targeting therapeutic and imaging agents to folate receptor positive tumors," Curr Pharm Biotechnol 6, 131-150 (2005).

426. W. T. Chen, K. Khazaie, G. Zhang, R. Weissleder, and C. H. Tung, "Detection of dysplastic intestinal adenomas using a fluorescent folate imaging probe," Mol Imaging 4, 67-74 (2005).

427. E. M. Barbosa, S. Nonogaki, M. L. Katayama, M. A. Folgueira, V. F. Alves, and M. M. Brentani, "Vitamin D3 modulation of plasminogen activator inhibitor type-1 in human breast carcinomas under organ culture," Virchows Arch 444, 175-182 (2004).

428. Y. Ye, S. Bloch, J. Kao, and S. Achilefu, "Multivalent carbocyanine molecular probes: synthesis and applications," Bioconjug Chem 16, 51-61 (2005).

429. C. Li, T. R. Greenwood, Z. M. Bhujwalla, and K. Glunde, "Synthesis and characterization of glucosamine-bound near-infrared probes for optical imaging," Org Lett 8, 3623-3626 (2006).

430. C. Li, T. R. Greenwood, and K. Glunde, "Glucosamine-bound near-infrared fluorescent probes with lysosomal specificity for breast tumor imaging," Neoplasia 10, 389-398 (2008).

431. M. Zhang, Z. Zhang, D. Blessington, H. Li, T. M. Busch, V. Madrak, J. Miles, B. Chance, J. D. Glickson, and G. Zheng, "Pyropheophorbide 2-deoxyglucosamide: a new photosensitizer targeting glucose transporters," Bioconjug Chem 14, 709-714 (2003).
432. J. F. Lovell, and G. Zheng, "Activatable smart probes for molecular optical imaging and therapy," Journal of Innovative Optical Health Sciences 1, 45-61 (2008).

433. G. Blum, G. von Degenfeld, M. J. Merchant, H. M. Blau, and M. Bogoy, "Noninvasive optical imaging of cysteine protease activity using fluorescently quenched activity-based probes," Nat Chem Biol 3, 668-677 (2007).

434. F. A. Jaffer, D. E. Kim, L. Quinti, C. H. Tung, E. Aikawa, A. N. Pande, R. H. Kohler, G. P. Shi, P. Libby, and R. Weissleder, "Optical visualization of cathepsin K activity in atherosclerosis with a novel, protease-activatable fluorescence sensor," Circulation 115, 2292-2298 (2007).

435. C. Bremer, S. Bredow, U. Mahmood, R. Weissleder, and C. H. Tung, "Optical imaging of matrix metalloproteinase-2 activity in tumors: feasibility study in a mouse model," Radiology 221, 523-529 (2001).

436. C. Bremer, C. H. Tung, and R. Weissleder, "In vivo molecular target assessment of matrix metalloproteinase inhibition," Nature medicine 7, 743-748 (2001).

437. C. Bremer, C. H. Tung, and R. Weissleder, "Molecular imaging of MMP expression and therapeutic MMP inhibition," Acad Radiol 9 Suppl 2, S314-315 (2002).

438. C. Molenaar, S. A. Marras, J. C. Slats, J. C. Truffert, M. Lemaitre, A. K. Raap, R. W. Dirks, and H. J. Tanke, "Linear 2' O-Methyl RNA probes for the visualization of RNA in living cells," Nucleic Acids Res 29, E89-89 (2001).

439. D. P. Bratu, B. J. Cha, M. M. Mhlanga, F. R. Kramer, and S. Tyagi, "Visualizing the distribution and transport of mRNAs in living cells," Proceedings of the National Academy of Sciences of the United States of America 100, 13308-13313 (2003).

440. A. Miyawaki, J. Llopis, R. Heim, J. M. McCaffery, J. A. Adams, M. Ikura, and R. Y. Tsien, "Fluorescent indicators for Ca2+ based on green fluorescent proteins and calmodulin," Nature 388, 882-887 (1997).

441. G. Miesenbock, D. A. De Angelis, and J. E. Rothman, "Visualizing secretion and synaptic transmission with pH-sensitive green fluorescent proteins," Nature 394, 192-195 (1998).

442. Y. Urano, D. Asanuma, Y. Hama, Y. Koyama, T. Barrett, M. Kamiya, T. Nagano, T. Watanabe, A. Hasegawa, P. L. Choyke, and H. Kobayashi, "Selective molecular imaging of viable cancer cells with pH-activatable fluorescence probes," Nature medicine 15, 104-109 (2009).

443. V. V. Belousov, A. F. Fradkov, K. A. Lukyanov, D. B. Staroverov, K. S. Shakhbazov, A. V. Terskikh, and S. Lukyanov, "Genetically encoded fluorescent indicator for intracellular hydrogen peroxide," Nat Methods 3, 281-286 (2006).
444. E. W. Miller, O. Tulyathan, E. Y. Isacoff, and C. J. Chang, "Molecular imaging of hydrogen peroxide produced for cell signaling," Nat Chem Biol 3, 263-267 (2007).

445. M. A. Funovics, R. Weissleder, and U. Mahmood, "Catheter-based in vivo imaging of enzyme activity and gene expression: Feasibility study in mice," Radiology 231, 659-666 (2004).

446. B. H. Zhu, F. A. Jaffer, V. Ntziachristos, and R. Weissleder, "Development of a near infrared fluorescence catheter: operating characteristics and feasibility for atherosclerotic plaque detection," Journal of Physics D-Applied Physics 38, 2701-2707 (2005).

447. G. Garcea, R. A. Sharma, A. Dennison, W. P. Steward, A. Gescher, and D. P. Berry, "Molecular biomarkers of colorectal carcinogenesis and their role in surveillance and early intervention," Eur J Cancer 39, 1041-1052 (2003).

448. S. J. Goldsmith, "Receptor imaging: competitive or complementary to antibody imaging?," Semin Nucl Med 27, 85-93 (1997).

449. P. L. Hsiung, J. Hardy, S. Friedland, R. Soetikno, C. B. Du, A. P. Wu, P. Sahbaie, J. M. Crawford, A. W. Lowe, C. H. Contag, and T. D. Wang, "Detection of colonic dysplasia in vivo using a targeted heptapeptide and confocal microendoscopy," Nature medicine 14, 454-458 (2008).

450. S. J. Miller, B. P. Joshi, Y. Feng, A. Gaustad, E. R. Fearon, and T. D. Wang, "In vivo fluorescence-based endoscopic detection of colon dysplasia in the mouse using a novel peptide probe," PLoS One 6, e17384 (2011).

451. J. R. Newton, K. A. Kelly, U. Mahmood, R. Weissleder, and S. L. Deutscher, "In vivo selection of phage for the optical Imaging of PC-3 human prostate carcinoma in mice," Neoplasia 8, 772-780 (2006).

452. H. Kitahara, J. Masumoto, A. L. Parker, F. Maruta, N. Kubo, A. Shimizu, N. Akita, S. Miwa, N. Kobayashi, J. Nakayama, and S. Miyagawa, "COP35, a cholangiocarcinoma-binding oligopeptide, interacts with the clathrin heavy chain accompanied by GRP78," Mol Cancer Res 9, 688-701 (2011).

453. B. Zhang, Y. Zhang, J. Wang, J. Chen, Y. Pan, L. Ren, Z. Hu, J. Zhao, M. Liao, and S. Wang, "Screening and identification of a targeting peptide to hepatocarcinoma from a phage display peptide library," Mol Med 13, 246-254 (2007).

454. J. R. Newton, Y. Miao, S. L. Deutscher, and T. P. Quinn, "Melanoma imaging with pretargeted bivalent bacteriophage," J Nucl Med 48, 429-436 (2007).

455. J. E. Koblinski, M. Ahram, and B. F. Sloane, "Unraveling the role of proteases in cancer," Clinica Chimica Acta 291, 113-135 (2000).

456. M. R. Emmertbuck, M. J. Roth, Z. P. Zhuang, E. Campo, J. R. Rozhin, B. F. Sloane, L. A. Liotta, and W. G. Stetlerstevenson, "Increased Gelatinase a (Mmp-2) and Cathepsin-B
Activity in Invasive Tumor Regions of Human Colon-Cancer Samples," American Journal of Pathology **145**, 1285-1290 (1994).

457. L. G. M. Hazen, F. E. Bleeker, B. Lauritzen, S. Bahns, J. Song, A. Jonker, B. E. M. Van Driel, H. Lyon, U. Hansen, A. Kohler, and C. J. F. Van Noorden, "Comparative localization of cathepsin B protein and activity in colorectal cancer," Journal of Histochemistry & Cytochemistry **48**, 1421-1430 (2000).

458. L. Herszenyi, M. Plebani, P. Carraro, M. De Paoli, G. Roveroni, R. Cardin, Z. Tulassay, R. Naccarato, and F. Farinati, "The role of cysteine and serine proteases in colorectal carcinoma," Cancer **86**, 1135-1142 (1999).

459. R. Upadhyay, R. A. Sheth, R. Weissleder, and U. Mahmood, "Quantitative real-time catheter-based fluorescence molecular imaging in mice," Radiology **245**, 523-531 (2007).

460. C. M. McCann, P. Waterman, J. L. Figueiredo, E. Aikawa, R. Weissleder, and J. W. Chen, "Combined magnetic resonance and fluorescence imaging of the living mouse brain reveals glioma response to chemotherapy," Neuroimage **45**, 360-369 (2009).

461. J. Fang, Y. Shing, D. Wiederschain, L. Yan, C. Butterfield, G. Jackson, J. Harper, G. Tamvakopoulos, and M. A. Moses, "Matrix metalloproteinase-2 is required for the switch to the angiogenic phenotype in a tumor model," Proceedings of the National Academy of Sciences of the United States of America **97**, 3884-3889 (2000).

462. J. Folkman, "Angiogenic zip code," Nat Biotechnol **17**, 749 (1999).

463. G. I. Murray, M. E. Duncan, P. O'Neil, W. T. Melvin, and J. E. Fothergill, "Matrix metalloproteinase-1 is associated with poor prognosis in colorectal cancer," Nature medicine **2**, 461-462 (1996).

464. R. Dahiyia, S. H. Itzkowitz, J. C. Byrd, and Y. S. Kim, "Mucin oligosaccharide biosynthesis in human colonic cancerous tissues and cell lines," Cancer **70**, 1467-1476 (1992).

465. Y. S. Kim, R. Isaacs, and J. M. Perdomo, "Alterations of membrane glycopeptides in human colonic adenocarcinoma," Proceedings of the National Academy of Sciences of the United States of America **71**, 4869-4873 (1974).

466. C. R. Boland, C. K. Montgomery, and Y. S. Kim, "Alterations in human colonic mucin occurring with cellular differentiation and malignant transformation," Proceedings of the National Academy of Sciences of the United States of America **79**, 2051-2055 (1982).

467. S. Nakamori, D. M. Ota, K. R. Cleary, K. Shirotani, and T. Irimura, "MUC1 mucin expression as a marker of progression and metastasis of human colorectal carcinoma," Gastroenterology **106**, 353-361 (1994).
468. P. J. Limburg, D. A. Ahlquist, J. A. Gilbert, J. J. Harrington, G. G. Klee, and P. C. Roche, "Immunodiscrimination of colorectal neoplasia using MUC1 antibodies: discrepant findings in tissue versus stool," Dig Dis Sci 45, 494-499 (2000).

469. S. Yonezawa, T. Nakamura, S. Tanaka, and E. Sato, "Glycoconjugate with Ulex europaeus agglutinin-I-binding sites in normal mucosa, adenoma, and carcinoma of the human large bowel," J Natl Cancer Inst 69, 777-785 (1982).

470. S. Yonezawa, T. Nakamura, S. Tanaka, K. Maruta, M. Nishi, and E. Sato, "Binding of Ulex europaeus agglutinin-I in polyposis coli: comparative study with solitary adenoma in the sigmoid colon and rectum," J Natl Cancer Inst 71, 19-24 (1983).

471. T. Kuroki, A. Kubota, Y. Miki, T. Yamamura, and J. Utsunomiya, "Lectin staining of neoplastic and normal background colorectal mucosa in nonpolyposis and polyposis patients," Dis Colon Rectum 34, 679-684 (1991).

472. M. Watanabe, H. Ohtani, M. Tanaka, and H. Ikehara, "Appearance of Ulex europaeus agglutinin-I and Griffonia simplicifolia agglutinin-I binding sites on cancer cells in sigmoid colonic polyps," Acta Pathol Jpn 42, 800-806 (1992).

473. C. A. Roney, B. Xu, J. Xie, S. Yuan, J. Wierwille, C. W. Chen, Y. Chen, G. L. Griffiths, and R. M. Summers, "Rh-I-UEA-1 Polymerized Liposomes Target and Image Adenomatous Polyps in the APCMin/+ Mouse Using Optical Colonography," Mol Imaging 10, 305-316 (2011).

474. S. Yuan, C. A. Roney, J. Wierwille, C. W. Chen, B. Xu, G. Griffiths, J. Jiang, H. Ma, A. Cable, R. M. Summers, and Y. Chen, "Co-registered optical coherence tomography and fluorescence molecular imaging for simultaneous morphological and molecular imaging," Phys Med Biol 55, 191-206 (2010).

475. E. K. Pauwels, E. J. Sturm, E. Bombardieri, F. J. Cleton, and M. P. Stokkel, "Positron-emission tomography with [18F]fluorodeoxyglucose. Part I. Biochemical uptake mechanism and its implication for clinical studies," J Cancer Res Clin Oncol 126, 549-559 (2000).

476. H. Zhou, K. Luby-Phelps, B. E. Mickey, A. A. Habib, R. P. Mason, and D. Zhao, "Dynamic near-infrared optical imaging of 2-deoxyglucose uptake by intracranial glioma of athymic mice," PLoS One 4, e8051 (2009).

477. Z. Cheng, J. Levi, Z. Xiong, O. Gheyssens, S. Keren, X. Chen, and S. S. Gambhir, "Near-infrared fluorescent deoxyglucose analogue for tumor optical imaging in cell culture and living mice," Bioconjug Chem 17, 662-669 (2006).

478. K. R. Kalli, A. L. Oberg, G. L. Keeney, T. J. Christianson, P. S. Low, K. L. Knutson, and L. C. Hartmann, "Folate receptor alpha as a tumor target in epithelial ovarian cancer," Gynecol Oncol 108, 619-626 (2008).
479. S. Markert, S. Lassmann, B. Gabriel, M. Klar, M. Werner, G. Gitsch, F. Kratz, and A. Hasenburg, "Alpha-folate receptor expression in epithelial ovarian carcinoma and non-neoplastic ovarian tissue," Anticancer Res 28, 3567-3572 (2008).

480. G. M. van Dam, G. Themelis, L. M. Crane, N. J. Harlaar, R. G. Pleijhuis, W. Kelder, A. Sarantopoulos, J. S. de Jong, H. J. Arts, A. G. van der Zee, J. Bart, P. S. Low, and V. Ntziachristos, "Intraoperative tumor-specific fluorescence imaging in ovarian cancer by folate receptor-α targeting: first in-human results," Nature medicine (in press).

481. A. Zaheer, R. E. Lenkinski, A. Mahmood, A. G. Jones, L. C. Cantley, and J. V. Frangioni, "In vivo near-infrared fluorescence imaging of osteoblastic activity," Nat Biotechnol 19, 1148-1154 (2001).

482. J. L. Kovar, X. Xu, D. Draney, A. Cupp, M. A. Simpson, and D. M. Olive, "Near-infrared-labeled tetracycline derivative is an effective marker of bone deposition in mice," Anal Biochem 416, 167-173 (2011).

483. Y. Zilberman, I. Kallai, Y. Gañni, G. Pelled, S. Kossodo, W. Yared, and D. Gazit, "Fluorescence molecular tomography enables in vivo visualization and quantification of nonunion fracture repair induced by genetically engineered mesenchymal stem cells," J Orthop Res 26, 522-530 (2008).

484. W. T. Chen, U. Mahmood, R. Weissleder, and C. H. Tung, "Arthritis imaging using a near-infrared fluorescence folate-targeted probe," Arthritis Res Ther 7, R310-317 (2005).

485. A. Wunder, C. H. Tung, U. Muller-Ladner, R. Weissleder, and U. Mahmood, "In vivo imaging of protease activity in arthritis: a novel approach for monitoring treatment response," Arthritis Rheum 50, 2459-2465 (2004).

486. W. F. Lai, C. H. Chang, Y. Tang, R. Bronson, and C. H. Tung, "Early diagnosis of osteoarthritis using cathepsin B sensitive near-infrared fluorescent probes," Osteoarthritis Cartilage 12, 239-244 (2004).

487. P. Mayer-Kuckuk, and A. L. Boskey, "Molecular imaging promotes progress in orthopedic research," Bone 39, 965-977 (2006).

488. F. A. Jaffer, P. Libby, and R. Weissleder, "Optical and multimodality molecular imaging: insights into atherosclerosis," Arterioscler Thromb Vasc Biol 29, 1017-1024 (2009).

489. J. Chen, C. H. Tung, U. Mahmood, V. Ntziachristos, R. Gyurko, M. C. Fishman, P. L. Huang, and R. Weissleder, "In vivo imaging of proteolytic activity in atherosclerosis," Circulation 105, 2766-2771 (2002).

490. J. O. Deguchi, M. Aikawa, C. H. Tung, E. Aikawa, D. E. Kim, V. Ntziachristos, R. Weissleder, and P. Libby, "Inflammation in atherosclerosis: visualizing matrix metalloproteinase action in macrophages in vivo," Circulation 114, 55-62 (2006).
491. T. Quillard, K. Croce, F. A. Jaffer, R. Weissleder, and P. Libby, "Molecular imaging of macrophage protease activity in cardiovascular inflammation in vivo," Thromb Haemost 105, 828-836 (2011).

492. F. A. Jaffer, C. Vinegoni, M. C. John, E. Aikawa, H. K. Gold, A. V. Finn, V. Ntziachristos, P. Libby, and R. Weissleder, "Real-time catheter molecular sensing of inflammation in proteolytically active atherosclerosis," Circulation 118, 1802-1809 (2008).

493. F. A. Jaffer, M. A. Calfon, A. Rosenthal, G. Mallas, R. N. Razansky, A. Mauskapf, R. Weissleder, P. Libby, and V. Ntziachristos, "Two-dimensional intravascular near-infrared fluorescence molecular imaging of inflammation in atherosclerosis and stent-induced vascular injury," Journal of the American College of Cardiology 57, 2516-2526 (2011).