Seven Functional Polymorphisms in the \textit{CETP} Gene and Myocardial Infarction Risk: A Meta-Analysis and Meta-Regression

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Abstract

Objective: This meta-analysis aims to evaluate the relationships between seven functional polymorphisms in the \textit{CETP} gene and myocardial infarction (MI) risk.

Method: The PubMed, CISCOM, CINAHL, Web of Science, Google Scholar, EBSCO, Cochrane Library, and CBM databases were searched for relevant articles published before March 1st, 2013 without any language restrictions. Meta-analysis was conducted using the STATA 12.0 software.

Results: Nine case-control studies with a total 8,623 MI cases and 8,564 healthy subjects met the inclusion criteria. The results of our meta-analysis suggested that \textit{CETP} rs708272 (C>T) polymorphism might be correlated with an increased risk of MI, especially among Caucasians. Furthermore, we observed that \textit{CETP} rs1800775 (C>A) polymorphism might increase the risk of MI. Nevertheless, no similar findings were found for \textit{CETP} rs5882 (A>G), rs2303790 (A>G), rs1800776 (C>A), rs12149545 (G>A), and rs4783961 (G>A) polymorphisms.

Conclusion: The current meta-analysis suggests that \textit{CETP} rs708272 (C>T) and rs1800775 (C>A) polymorphisms may contribute to MI susceptibility, especially among Caucasians. Thus, \textit{CETP} rs708272 and rs1800775 polymorphisms may be promising and potential biomarkers for early diagnosis of MI.

Introduction

Myocardial infarction (MI) remains the leading cause of death and disability worldwide, accounting for up to 40% of all deaths [1]. Due to high mortality and disability rates, MI is becoming a global epidemiological health concern [2]. Rupturing of coronary atherosclerotic plaque with consequent platelet aggregation and thrombus formation is the major cause of MI [3–5]. Many intrinsic and extrinsic risk factors for MI have been established, including dyslipidemia, hypertension, smoking, obesity, etc. [6,7]. Atherogenic dyslipidemia is usually characterized by three lipid abnormalities: increases in plasma triglyceride, small low density cholesterol (LDL-C) and very low density lipoprotein cholesterol (VLDL-C) levels, and decreased high-density lipoprotein cholesterol (HDL-C) levels [8–10]. Although the exact cellular and molecular mechanisms leading to the development of MI remain unclear, it is believed that functionally relevant mutations in the dyslipidemia-related genes may function to increased susceptibility to MI [11].

Cholesteryl ester transfer protein (CETP) is a plasma protein that mediates the exchange of neutral lipids, including cholesteryl esters and triglycerides between plasma lipoproteins [12]. CETP plays a critical role in reverse cholesteryl transport of cholesteryl esters and triglycerides from HDL-C to LDL-C and VLDL-C [13,14]. It is well established that HDL-C has a protective role against cardiovascular disease [15]. Plasma HDL particles play an important role in removing cellular cholesterol and delivering it to the liver for re-utilization [16]. Furthermore, it should be noted that levels of HDL-C is significantly negatively correlated with arterial stenosis whose occurrence is strongly associated with the phenomenon of plaque rupture [17]. Thus higher levels of HDL-C tend to have fewer problems with cardiovascular diseases such as MI, while those with low HDL-C cholesterol levels may easily suffer from MI [18,19]. Variation in CETP activity could influence HDL-C levels and thus contribute to increased susceptibility to cardiovascular disease such as MI [20]. Genetic and epigenetic changes in the \textit{CETP} gene may enhance plasma cholesteryl ester formation and lead to low HDL-C levels and...
The included studies must meet all four of the following criteria: (1) the study design must be clinical cohort or case-control study that focused on the relationships of CETP genetic polymorphisms with the risk of MI; (2) all patients met the diagnostic criteria for MI; (3) the genotype frequencies of healthy controls should follow the Hardy-Weinberg equilibrium (HWE); (4) the study must provide sufficient information about the genotype frequencies. If the study could not meet the inclusion criteria, it would be excluded. The most recent or the largest sample size publication was included when the authors published several studies using the same subjects. The PRISMA checklist is available in Checklist S1.

**Selection criteria**

Figure 1. Flow chart shows study selection procedure.

**Materials and Methods**

**Search strategy**

The PubMed, CISCOM, CINAHL, Web of Science, Google Scholar, EBSCO, Cochrane Library, and CBM databases were searched for relevant articles published before March 1st, 2013 without any language restrictions. The following keywords and MeSH terms were used: (“SNP” or “mutation” or “genetic polymorphism” or “variation” or “polymorphism” or “single nucleotide polymorphism” or “variant”) and (“myocardial infarction” or “myocardial infarct” or “MI” or “AMF” or “heart attacks”) and (“cholesterol ester transfer protein” or “CETP” or “cholesterol ester exchange protein” or “CE transport protein”). We also performed a manual search of the relevant articles to find other potential articles.
| First author [Ref] | Year | Country | Ethnicity | Number | Gender (M/F) | Age (years) | Source | Genotype method | SNP | NOS score |
|--------------------|------|---------|-----------|--------|-------------|-------------|--------|----------------|-----|-----------|
| Li et al [36]      | 1999 | China   | Asian     | 102    | 102         | 81/21       | HB     | PCR-RFLP       | rs2303790 (A>G) | 24 |
| Zhang et al [35]   | 2005 | China   | Asian     | 50     | 94          | 35/15       | HB     | PCR-RFLP       | rs708272 (G>A)  | 26 |
| Eiriksdottir et al [29] | 2001 | Iceland | Caucasian | 388    | 749         | 388/0       | PB     | PCR-RFLP       | rs708272 (G>A)  | 30 |
| Liu et al [33]     | 2002 | USA     | Caucasian | 384    | 384         | 384/0       | HB     | PCR-RFLP       | rs1800775 (C>A) | 9  |
| Andrikopoulos et al [34] | 2004 | Greek   | Caucasian | 1625   | 735         | -           | PB     | PCR-RFLP       | rs5882 (A>G)    | 27 |
| Keavney et al [30] | 2004 | UK      | Caucasian | 4442   | 3273        | 2892/1550   | PB     | PCR-RFLP       | rs708272 (G>A)  | 35 |
| Tobin et al [31]   | 2004 | UK      | Caucasian | 547    | 505         | 372/175     | PB     | PCR-RFLP       | rs1800775 (C>A) | 33 |
| Zee et al [45]     | 2006 | USA     | Caucasian | 523    | 2092        | 523/0       | PB     | Direct sequencing | rs1800775 (C>A) | 33 |
| Meiner et al [32]  | 2008 | USA     | Caucasian | 561    | 629         | 314/237     | PB     | Direct sequencing | rs12149545 (G>A) | 34 |

M = male, F = female, PB = population-based, HB = hospital-based, PCR = polymerase chain reaction, RFLP = restriction fragment length polymorphism, SNP = single nucleotide polymorphism, NOS = Newcastle-Ottawa quality assessment scale.

doi:10.1371/journal.pone.0088118.t001
Relevant data were systematically extracted from all included studies by two observers by using a standardized form. The researchers collected the following data: language of publication, publication year of article, the first author’s surname, geographical location, design of study, sample size, the source of the subjects, genotype frequencies, source of samples, genotyping method, evidence of HWE, etc.

Quality assessment
Methodological quality was evaluated separately by two observers using the Newcastle-Ottawa Scale (NOS) criteria [37]. The NOS criteria included three aspects: (1) subject selection: 0–4; (2) comparability of subject: 0–2; (3) clinical outcome: 0–3. NOS scores ranged from 0 to 9; and a score ≥7 indicate a good quality. The NOS criteria are available in File S1.

Statistical analysis
The STATA version 12.0 (Stata Corp, College Station, TX, USA) software was used for meta-analysis. We calculated crude odds ratio (OR) with their 95% confidence interval (95%CI) to evaluate their relationships under 5 genetic models. Genotype frequencies of healthy controls were tested for the HWE using the $\chi^2$ test. The statistical significance of pooled ORs was assessed by the Z test. The Cochran’s Q-statistic and $I^2$ test were used to evaluate potential heterogeneity between studies [38–40]. If Q-test shows a $P<0.05$ or $I^2$ test exhibits >50% which indicates significant heterogeneity, the random-effect model was conducted, or else the fixed-effects model was used. We also performed subgroup and meta-regression analyses to investigate potential sources of heterogeneity. We conducted a sensitivity analysis to assess the influence of single studies on the overall ORs. Begger's

**Figure 2. Forest plot of the relationships between CETP rs708272 (C>T) polymorphism and myocardial infarction risk under the allele and dominant models.**

doi:10.1371/journal.pone.0088118.g002

| Study ID          | OR (95% CI)   | Weight % |
|-------------------|---------------|----------|
| A                 |               |          |
| Zhang et al (2005)| 1.35 (0.82, 2.22) | 1.24     |
| Eiriksdottir et al (2001) | 1.13 (0.94, 1.35) | 10.66   |
| Liu et al (2002)  | 0.95 (0.78, 1.17) | 9.09     |
| Keavney et al (2004) | 1.53 (1.43, 1.64) | 66.63   |
| Meiner et al (2008) | 1.16 (0.98, 1.37) | 12.39   |
| **Overall** (I2 = 87.2%, $P < 0.001$) | 1.39 (1.31, 1.47) | 100.00   |

| Study ID          | OR (95% CI)   | Weight % |
|-------------------|---------------|----------|
| B                 |               |          |
| Zhang et al (2005)| 1.57 (0.77, 3.20) | 1.23     |
| Eiriksdottir et al (2001) | 1.20 (0.93, 1.56) | 10.56   |
| Liu et al (2002)  | 0.96 (0.71, 1.31) | 8.57     |
| Keavney et al (2004) | 1.70 (1.55, 1.86) | 68.27   |
| Meiner et al (2008) | 1.34 (1.05, 1.71) | 11.41   |
| **Overall** (I2 = 78.0%, $P = 0.001$) | 1.54 (1.42, 1.67) | 100.00   |

Data extraction
Relevant data were systematically extracted from all included studies by two observers by using a standardized form. The researchers collected the following data: language of publication, publication year of article, the first author’s surname, geographical location, design of study, sample size, the source of the subjects, genotype frequencies, source of samples, genotyping method, evidence of HWE, etc.
funnel plots and Egger’s linear regression test were used to investigate publication bias [41].

Results

Baseline characteristics of included studies

Initially, the searched keywords identified 90 articles. We reviewed the titles and abstracts of all articles and excluded 44 articles; full texts were also reviewed and 34 articles were further excluded. Three other studies were excluded due to no sufficient data about seven common SNPs in the CETP gene [42–44]. Figure 1 shows the selection process of eligible articles. Finally, 9 case-control studies with a total 8,623 MI cases and 8,564 healthy subjects met our inclusion criteria for qualitative data analysis [29–36,45]. Population-based controls were used in 6 studies, and hospital-based controls were used in 3 studies. Overall, seven studies were conducted among Caucasians and two studies among Asians. Polymerase chain reaction-restriction fragment length polymorphism (PCR-RELP) method was conducted in 7 studies, and 2 studies used direct sequencing method. Seven common polymorphisms in the CETP gene were assessed, including rs708272 (C>T), rs1800775 (C>A), rs5882 (A>G), rs2303790 (A>G), rs1800776 (C>A), rs12149545 (G>A), and rs4783961 (G>A); and among these, rs708272 (C>T) and rs1800775 (C>A)
Table 2. Meta-analysis of the associations between CETP rs708272 (G>A) and rs1800775 (C>A) polymorphisms and endometrial cancer risk.

| Subgroups          | rs708272 (G>A) | rs1800775 (C>A) |
|--------------------|----------------|----------------|
|                    | M allele vs. W allele | WM+MM vs. WW | MM vs. WW+WM | MM vs. WW homozgyous model | MM vs. WM heterozygous model |
| Overall            | 1.39 [1.31, 1.47] | 1.54 [1.42, 1.67] | 1.52 [1.37, 1.70] | 1.87 [1.66, 2.11] | 1.29 [1.15, 1.45] |
| Ethnicity          |                |                |                |                |                |
| Caucasians         | 1.39 [1.31, 1.47] | 1.54 [1.42, 1.67] | 1.53 [1.37, 1.70] | 1.87 [1.66, 2.11] | 1.30 [1.15, 1.46] |
| Asians             | 1.35 [0.82, 2.22] | 0.243           | 0.216           | 0.594           | 1.57 [0.82, 2.22] |
| Country            |                |                |                |                |                |
| China              | 1.35 [0.82, 2.22] | 0.243           | 0.216           | 0.594           | 1.57 [0.82, 2.22] |
| Iceland            | 1.13 [0.94, 1.35] | 0.185           | 0.162           | 0.497           | 1.13 [0.94, 1.35] |
| USA                | 1.07 [0.94, 1.22] | 0.290           | 0.088           | 0.852           | 1.07 [0.94, 1.22] |
| UK                 | 1.53 [1.43, 1.64] | <0.001          | 1.70 [1.55, 1.86] | <0.001          | 1.83 [1.60, 2.10] |
| Source of controls |                |                |                |                |                |
| Population-based   | 1.16 [1.03, 1.30] | 0.017           | 1.29 [1.09, 1.53] | 0.004           | 1.09 [0.87, 1.36] |
| Hospital-based     | 1.46 [1.37, 1.56] | <0.001          | 1.62 [1.48, 1.77] | <0.001          | 1.69 [1.49, 1.92] |
| Genotype methods   |                |                |                |                |                |
| PCR-RFLP           | 1.42 [1.34, 1.51] | <0.001          | 1.57 [1.44, 1.70] | <0.001          | 1.61 [1.43, 1.81] |
| DNA sequencing      | 1.16 [0.98, 1.37] | 0.081           | 1.34 [1.05, 1.70] | 0.018           | 1.04 [0.76, 1.41] |

| Overall            | 1.13 [1.05, 1.22] | 0.002           | 1.34 [1.18, 1.53] | <0.001          | 1.03 [0.90, 1.17] |
| Country            |                |                |                |                |                |
| Iceland            | 1.31 [1.10, 1.56] | 0.003           | 1.74 [1.30, 2.33] | <0.001          | 1.18 [0.88, 1.58] |
| USA                | 1.21 [1.09, 1.34] | <0.001          | 0.89 [0.67, 1.18] | <0.001          | 1.15 [0.97, 1.36] |
| UK                 | 0.83 [0.70, 0.98] | 0.028           | 1.43 [1.21, 1.69] | 0.015           | 0.66 [0.50, 0.88] |
| Genotype methods   |                |                |                |                |                |
| PCR-RFLP           | 1.03 [0.92, 1.17] | 0.592           | 1.24 [1.01, 1.51] | 0.038           | 0.88 [0.72, 1.08] |
| DNA sequencing      | 1.21 [1.09, 1.34] | <0.001          | 1.43 [1.21, 1.69] | <0.001          | 1.15 [0.97, 1.36] |

OR = odds ratios, 95%CI = 95% confidence interval, W = wild allele, M = mutant allele, WW = wild homozygote, WM = heterozygote, MM = mutant homozygote, PCR = polymerase chain reaction, RFLP = restriction fragment length polymorphism.

DOI:10.1371/journal.pone.0088118.t002
were the most common SNPs. None of the studies deviated from the HWE (all \( P > 0.05 \)). NOS scores of all included studies were \( 5 \). We summarized the study characteristics and methodological quality in Table 1. The genotypic distributions of CETP genetic polymorphisms are shown in File S2.

**Quantitative data synthesis**

The relationships of \( \text{CETP}\, \text{rs708272 (C>T)} \) polymorphism with the risk of MI were reported in 5 studies. The heterogeneity obviously existed (\( P < 0.05 \)), so the random effects model was conducted. Our meta-analysis results revealed that \( \text{CETP}\, \text{rs708272} \) polymorphism may increase the risk of MI (T allele vs. C allele: \( \text{OR} = 1.39, 95\% \text{CI}: 1.31–1.56, P = 0.001 \); CT+TT vs. CC: \( \text{OR} = 1.52, 95\% \text{CI}: 1.37–1.70, P < 0.001 \); TT vs. CC: \( \text{OR} = 1.87, 95\% \text{CI}: 1.66–2.11, P < 0.001 \); TT vs. CT: \( \text{OR} = 1.29, 95\% \text{CI}: 1.15–1.45, P = 0.001 \) (Figure 2). Among different ethnic subgroups, the results revealed positive correlations between \( \text{CETP}\, \text{rs708272 (C>T)} \) polymorphism and an increased risk of MI among Caucasians (Figure 3), but not among Asians (all \( P > 0.05 \)). The results of subgroup analyses also suggested that \( \text{CETP}\, \text{rs708272 (C>T)} \) polymorphism was associated with increased risk of MI in the UK, population-based, hospital-based, PCR-RFLP and direct sequencing subgroups (as shown in Table 2). However, \( \text{CETP}\, \text{rs708272 (C>T)} \) polymorphism showed no association with MI susceptibility in studies conducted in China, Iceland and USA.

There were 4 studies that referred to the relationships of \( \text{CETP}\, \text{rs1800775 (C>A)} \) polymorphism with MI risk. Since heterogeneity was significantly observed (\( P < 0.05 \)), the random effects model was used. Meta-analysis of these studies indicated positive correlations of \( \text{CETP}\, \text{rs1800775 (C>A)} \) polymorphism with an increased risk of MI (A allele vs. C allele: \( \text{OR} = 1.13, 95\% \text{CI}: 1.05–1.22, P = 0.002 \); CA+AA vs. CC: \( \text{OR} = 1.34, 95\% \text{CI}: 1.18–1.53, P < 0.001 \); AA vs. CC: \( \text{OR} = 1.27, 95\% \text{CI}: 1.08–1.49, P = 0.004 \) (Figure 4). We also conducted subgroup analyses by country and genotype; the results indicated that \( \text{CETP}\, \text{rs1800775 (C>A)} \) polymorphism might increase susceptibility to MI in most subgroups (as shown in Table 2).

The relationships of rs5882 (A>G), rs2303790 (A>G), rs1800776 (C>A), rs12149545 (G>A), and rs4783961 (G>A)
Table 3. Meta-analysis of the associations between five common polymorphisms in CETP gene and MI risk.

| SNP ID   | M allele vs. W allele (allele model) | OR [95%CI] | P     | M allele vs. WM (heterozygous model) | OR [95%CI] | P     | WM vs. WW (homozygous model) | OR [95%CI] | P     | MM vs. WW (dominant model) | OR [95%CI] | P     | MM vs. WM (recessive model) | OR [95%CI] | P     |
|----------|------------------------------------|------------|-------|-------------------------------------|------------|-------|--------------------------------|------------|-------|-------------------------|------------|-------|--------------------------|------------|-------|
| rs2303790 (A>G) | 1.52 [0.42, 5.45] | 0.525 |       |                                     |            |       |                                     |            |       | -                        |            |       | -                        |            |       |
| rs5882 (A>G)    | 0.96 [0.88, 1.05]    | 0.343 |       |                                     |            |       |                                     |            |       | 0.93 [0.78, 1.13]    | 0.461 |       | 0.89 [0.72, 1.11]    | 0.365 |       |
| rs1800776 (C>A) | 1.10 [0.90, 1.36]    | 0.337 |       |                                     |            |       |                                     |            |       | 1.20 [0.97, 1.48]    | 0.151 |       | 1.36 [1.10, 1.66]    | 0.07  |       |
| rs12149545 (G>A) | 0.93 [0.76, 1.12]    | 0.486 |       |                                     |            |       |                                     |            |       | 0.92 [0.74, 1.14]    | 0.493 |       | 0.94 [0.74, 1.22]    | 0.38  |       |
| rs4783961 (G>A) | 0.99 [0.76, 1.26]    | 0.941 |       |                                     |            |       |                                     |            |       | 0.89 [0.76, 1.07]    | 0.284 |       | 0.95 [0.77, 1.15]    | 0.367 |       |

OR = odds ratios, 95%CI = 95% confidence interval, W = wild allele, M = mutant allele, WW = wild homozygote, WM = heterozygote, MM = mutant homozygote, SNP= single nucleotide polymorphism.

doi:10.1371/journal.pone.0088118.t003

Discussion

CETP, a hydrophobic glycoprotein secreted mainly by the liver, catalyzes the transfer of cholesteryl esters from HDL to other lipoproteins and influences plasma HDL-C levels [46,47]. Previous studies have demonstrated a protective effect of HDL-C against cardiovascular disease by inhibiting lipoprotein oxidation [8,48,49]. High plasma levels of CETP are correlated with low HDL-C levels, and it has been implicated as a strong risk factor for cardiovascular disease, including MI [50]. Although MI is one of the most common heritable cardiovascular diseases, the fundamental molecular pathways remain undefined [51,52]. Thus, it was speculated that CETP genetic variations may be involved in the development of MI [45,53]. The CETP gene has been mapped to locus 16q21 encoding cholesteryl ester transfer protein [23]. Common polymorphisms of CETP gene may result in the over-expression of this protein and a subsequent decrease of HDL-C levels, thus contributing to the incidence of MI [21]. Indeed, several studies have demonstrated positive correlations of CETP genetic polymorphisms with an increased risk of MI [29–32], but the controversy still persists.

In the present meta-analysis, our findings revealed that CETP rs708272 (C>T) polymorphism might increase the risk of MI, especially among Caucasians, while similar results were not observed among Asians. There also existed positive correlations of CETP rs1800775 (C>A) polymorphism with an increased risk of MI among Caucasians. Although ethnic differences in to the risk of MI are well known, potential molecular mechanism is not fully understood. One possible reason for ethnic difference might be that CETP gene mutations might affect cholesteryl ester synthesis and result in low HDL-C levels, thereby possibly explaining interindividual differences in the incidence of MI [21]. Another likely explanation for this difference could be that large differences in common SNPs that influence the risk of MI are mostly due to genetic drift and natural selection [54]. The results of subgroup analyses demonstrated positive correlations of CETP rs708272 (C>T) polymorphism with an increased risk of MI in the UK, population-based, hospital-based, PCR-RFLP and direct sequencing subgroups, indicating that country, source of controls and genotype method may be the potential sources of heterogeneity. However, our meta-regression analyses indicated that only ethnicity was the major source of heterogeneity. These disparate results may be due to small sample size resulting in substantial errors from estimation. Nevertheless, we observed no associations between the other 5 common polymorphisms in the CETP gene and MI risk. In short, the results of our meta-analysis were
consistent with previous studies that CETP genetic polymorphisms may be closely linked to the risk of MI, suggesting that CETP genetic polymorphism could be useful and promising biomarkers for early diagnosis of MI.

The current meta-analysis also had many limitations that should be acknowledged. First, our results had lacked sufficient statistical power to assess the correlations between CETP genetic polymorphisms and MI risk. Secondly, meta-analysis is a retrospective study that may lead to subject selection bias, and thereby affecting
the reliability of our results [55]. Thirdly, our meta-analysis failed to obtain original data from the included studies, which may limit further evaluation of potential role of CETP genetic polymorphisms in the development of MI. Although our study has many limitations, this is the first meta-analysis focusing on the relationships between CETP genetic polymorphisms and the risk of MI. Furthermore, we performed a highly sensitive literature search strategy for electronic databases. A manual search of the reference lists from the relevant articles was also conducted to find other potential articles. The selection process of eligible articles was based on strict inclusion and exclusion criteria. Importantly, rigorous statistical analysis of SNP data provided a basis for pooling of information from individual studies.

In conclusion, our findings provide empirical evidence that CETP rs708272 (C>T) and rs1800775 (C>A) polymorphisms may contribute to MI susceptibility, especially among Caucasians. Thus, CETP rs708272 and rs1800775 polymorphisms may be promising and potential biomarkers for early diagnosis of MI. However, due to the limitations mentioned above, more researches with larger sample size are needed to provide a more representative statistical analysis precisely.

Figure 6. Beggar’s funnel plots of the relationships of CETP rs708272 (C>T) and rs1800775 (C>A) polymorphisms with myocardial infarction risk. Each point represents a separate study for the indicated association. Log[OR], natural logarithm of OR. Horizontal line, mean magnitude of the effect. doi:10.1371/journal.pone.0088118.g006
Table 4. Univariate and multivariate meta-regression analyses of potential source of heterogeneity.

| Heterogeneity factors | rs708272 (C>T) | rs1800775 (C>A) |
|-----------------------|---------------|-----------------|
|                       | | |
| Publication year | | |
| Univariate | 0.032 [-0.058, 0.124] | -0.009 [-0.141, 0.122] |
| Multivariate | 0.014 [-0.113, 0.143] | -0.003 [-0.131, 0.126] |
| Ethnicity | | |
| Univariate | 0.182 [-0.700, 1.065] | -0.542 [-0.877, 0.209] |
| Multivariate | 0.500 [-0.912, 1.911] | -0.559 [-0.993, 0.125] |
| Country | | |
| Univariate | 0.185 [0.073, 0.297] | -0.039 [-0.446, 0.368] |
| Multivariate | 0.160 [-0.309, 0.628] | -0.150 [-2.237, 1.935] |
| Source of controls | | |
| Univariate | -0.021 [-0.506, 0.465] | 0.355 [-0.114, 0.824] |
| Multivariate | 0.192 [-0.195, 0.579] | 0.362 [-0.296, 1.021] |
| Genotype methods | | |
| Univariate | -0.013 [-0.600, 0.574] | -0.147 [-0.790, 0.496] |
| Multivariate | 1.189 [0.324, 2.054] | -0.768 [-2.000, 0.463] |

SE = standard error, 95%CI = 95% confidence interval.

Supporting Information

Checklist S1 The PRISMA Checklist. (DOC)

File S1 The Newcastle-Ottawa quality assessment scale. (DOC)

File S2 The genotypic distributions of CETP genetic polymorphisms in the case and control groups. (XLS)

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Acknowledgments

We would like to acknowledge the reviewers for their helpful comments on this paper.

Author Contributions

Conceived and designed the experiments: QW SBZ YZJ. Performed the experiments: LW MML. Analyzed the data: YW CM. Contributed reagents/materials/analysis tools: QW SBZ. Wrote the paper: QW. Final approval of manuscript: YZJ.
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