Association of serum deiodinase type 2 level with chronic obstructive pulmonary disease in the Polish population

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INTRODUCTION

Chronic obstructive pulmonary disease (COPD) is an important cause of mortality and morbidity worldwide and is becoming a major public health problem (López-Campos et al., 2016). The disease is a result of many complex gene-environment interactions and is characterized by an unclear and multidirectional background; it is also considered to be a multicomponent disease (Mayer & Newman, 2001).

Large number of data have demonstrated that a chronic and systemic inflammation reaction and their components may participate in the patomechanism of COPD, and that signs of inflammation are also characteristic for individuals suffering from this disease (Sinden & Stockley, 2010).

Inflammation is orchestrated by many factors. One of them is the adipose tissue, which produces mediators, such as adipokines, involved in many chronic inflammatory diseases, including COPD (Fantuzzi, 2005).

Chemerin, which is found in the liver, the white fat tissue, lung, heart, and immune cells, is one of the newly discovered adipokines, also known as adipocytokines (Fatima et al., 2014). Chemerin plays a role of an immunomodulatory factor, the regulation of which is under the influence of, for example, IL-1, IL-6, TNF-α and other proinflammatory molecules. As a Chemerin as a ligand of chemokine receptors participates in inflammation from its first acute phase and in its reduction. The process through which chemerin is involved in inflammation is the induction of chemokinesis of immune cells, such as natural killers, macrophages, dendritic cells, and neutrophils (Marianti & Roncucci, 2015). Increased levels of chemerin concentration in serum are observed in various diseases of inflammatory nature (Weigert et al., 2010; De Palma et al., 2011; Kaneko et al., 2011). The mediatory role of chemerin observed in the immune responses may be associated with the pathogenesis of asthma and the inflammatory process itself (Zhou et al., 2018). Levels of chemerin in the synovial fluid and synovial membranes have been shown to be increased in the knees of osteoarthritis patients and those levels proportionately correlate with the severity of the disease (Ma et al., 2015).

Thyroid hormones (THs) are other key factors affecting almost all systems of the human body. Thus, special attention has been once again recently paid to the THs and THs-synthesis related enzymes – iodothyronine deiodinases (DIOs) – as important immune targets and modifiers of an inflammatory response (Korle 1999). There are also...
findings that both the THs and DIOs may modulate the immune system and influence inflammatory processes by affecting phagocytosis, chemotaxis, oxidative stress or cytokine production (De Vito et al., 2012).

Deiodinase type 2 (DIO2) is an activating enzyme that converts the T4 prohormone into triiodothyronine T3 through deiodination at the phenolic ring. This enzyme is responsible for local cellular demands, rather than for circulating T3. Presence of DIO2 is observed in the central nervous system, pituitary, placenta and skin. Regulation of DIO2 expression and function is a complex process. For example, mRNA levels for DIO2 and its enzymatic activity are controlled by the mechanism of posttranslational regulation (Korle, 1999). DIO2 is stimulated by corticosteroids and growth factors (Courtin et al., 1990; Coppola et al., 2005). Some reports indicate a highly inducible nature of DIO2 during inflammation by the nuclear factor-xB after treatment with bacterial lipopolysaccharide (LPS). In addition, its expression is similar to that of pro-inflammatory cytokines (Quan et al., 1998; Zeold et al., 2006). A study conducted by Kwakkel and others (Kwakkel et al., 2014) confirmed that DIO2 is upregulated during acute and chronic inflammation in macrophages and is correlated with IL-1 independently of the T3 serum levels. This may suggest that this enzyme plays a special role in the immune response. Therefore, DIO2 loss of function has significant metabolic implications, as D2KO mice have been shown to gain more weight and adipose tissue, and that tissue also being a known source of the pro-inflammatory molecules (Marishi et al., 2011).

In COPD, as in many other diseases, different biomarkers play a potential role in risk stratification, prediction of treatment responsiveness, monitoring of the disease and new drug investigation. Biomarkers are widely investigated in the sputum and blood of patients. A number of previous studies have examined concentration of different biomarkers in COPD; however, to the best of our knowledge, chemerin and deiodinase type 2 have not been studied in the serum of patients diagnosed with COPD. Markers involved in inflammation may play a crucial role in the development, mechanism and management of a disease, such as COPD. Measurements of inflammatory and immunomodulatory processes that play a key role in the development of COPD.

In addition, it would appear that DIO2 plays a role in the determination of TH levels, with its inappropriate serum concentrations found in some patients diagnosed with COPD. Reduced TH serum levels have been shown in patients with acute exacerbations of COPD with subsequent recovery of THs at the rate to various degrees, as the primary disease improves (Coskun et al., 2009; Cheng et al., 2016). Whereas chemerin (adipokine) is a product of adipose tissue and there are data that adipokines may function as the link between adipocytes and COPD (Stanciu et al., 2009).

The purpose of this study was to determine whether the serum chemerin and DIO2 levels are linked with COPD, and to assess the role of these molecules as potential novel markers and/or risk factors of this disease.

**MATERIALS AND METHODS**

This study was carried out on a group of 80 subjects, including patients diagnosed with COPD and healthy volunteers— all of Caucasian ethnicity. The study was conducted at the Norbert Barlicki University Clinical Hospital No. 1 (Łódź, Poland), Department of Pulmonology and Allergology of the Medical University of Łódź, Poland, and later at an outpatient clinic. All of the patients underwent detailed diagnostics, which included a clinical examination, spirometry, gasometry, and anthropometry.

The group of patients with COPD comprised 50 people diagnosed in compliance with the Global Initiative for Chronic Obstructive Lung Disease (GOLD) document from 2015. All the patients were ex-smokers. Smoking history in the study group totaled between 25 and 40 years, and the average number of packs of cigarettes per year was 25.

A total of 30 subjects, matched for age and sex, were selected among healthy community individuals invited to take part in the study based on the absence of diagnostic criteria.

Lung function was tested in clinically stable patients with the use of the LUNGTEST 1000 spirometer in a sitting position after a minimum of 15-minute rest. The patients were instructed to avoid short-acting β2-agonists at least 6 hours before testing and long-acting β2-agonists at least 12 hours prior to the lung function test. Gasometry was measured based on arterial blood according to the required standards, and the test was performed using the Corning 348 blood gas analyzer (Ciba Corning, United States).

Body weight and height were measured for all of the patients with an intention to calculate the body mass index (BMI), while the waist and hip circumference enabled the calculation of the waist-to-hip ratio (WHR).

Exacerbations in the course of the study were excluded based on laboratory tests (blood cell count with a smear, C-reactive protein – CRP) and clinical presentation (absence of dyspnea, normal body temperature, no green sputum expectoration). Lung tumors were excluded using radiological modalities (X-ray, computed tomography).

All of the procedures were reviewed and approved by the Local Bioethics Committee. Written informed consent was obtained from all the participants of the study.

The venous blood samples were collected into sterile tubes (2×5 ml) without additives and stored at room temperature until formation of surface clot (about 30 minutes); then, the samples were centrifuged for 15 minutes at approximately 1000×g. After centrifugation, the serum was removed and aliquots were stored at −80°C. Sera samples were collected from patients participating in the study in the morning, not less than 12 hours after their last meal.

Chemerin and DIO2 were assessed using a commercially available Human DIO2 ELISA Kit (MyBioSource, San Diego, CA, USA) and Human Chemerin Quantikine ELISA Kit (R&D Systems, McKinley Place, MN, USA). All calculations were performed according to the instructions and protocols provided by the manufacturers. The absorbance of the samples was measured using Multiskan Ascent Microplate Photometer (Thermo Lab-systems) at λ=450 nm. Analytical curves of the analyzed proteins were assessed to determine protein concentration. Serum DIO2 protein was presented in U/L, while serum chemerin protein levels in ng/ml. The detection range for human deiodinase type 2, manufactured by Mybiosource was 1.56 U/L to 50 U/L. The detection range for human chemerin assay manufactured by R&D Systems was 48.0 to 142 ng/ml. Both, the Intra-assay CV(%) and Inter-assay CV(%) were less than 15 [CV(%) =SD/mean ×100].
Statistical analysis. All of the data analyses were performed using Statistica (version 12.0). A statistical analysis of the collected material included calculation of both, the descriptive and inferential statistics. The results are presented as percentages (%) or means (M) with standard deviation (± S.D.). The chi-square test and Mann-Whitney U test were used to compare demographic variables (gender and age) between the patients and controls. The comparison of DIO1 and DIO3 concentration between the subjects with COPD and the controls was performed using the non-parametric Mann-Whitney U test. Pearson’s correlation was calculated to evaluate the relationships between the analyzed protein levels and other variables. Statistical significance was defined as \( p < 0.05 \) for all analyses.

RESULTS

The group of COPD patients consisted of 19 (38%) females and 31 (62%) males, with mean age of 65.7±8.2 years (range: 50–81 years). Amongst the 30 volunteers from the control group, 16 (53.33%) were female and 14 (46.67%) male, with mean age of 62.7±7 years (range: 52–74 years). The baseline characteristics of the subjects, together with anthropometric and spirometric data, are summarized in Table 1. There were no differences with respect to age (\( z = 1.61, p = 0.1 \)), gender (\( \chi^2 = 1.79, p = 0.1808 \)), and BMI (\( z = 0.79, p = 0.43 \)). Differences between the COPD patients and the controls were observed in terms of WHR (\( z = 3.81, p = 0.000137 \)). A significant reduction in FEV1, FV1/VC, and FVC in the COPD patients was acknowledged after a comparison with the controls, with \( z = –7.45108, p < 0.001 \), \( z = –7.05856, p < 0.0001 \), and \( z = –7.19838, p < 0.0001 \), respectively.

The obtained results confirmed that DIO2 levels were significantly higher in the serum of the patients suffering from COPD compared to the healthy controls. Detailed data are presented in Table 2.

Table 1. Baseline characteristics of the study population, including anthropometric and spirometric data

| Parameters                  | Healthy controls | COPD Patients |
|-----------------------------|------------------|---------------|
| Age (years)                 | 62.7             | 65.7          |
| Age women (years)           | 62.2             | 66.1          |
| Age men (years)             | 63.2             | 65.5          |
| BMI (kg/m2)                 | 25.0             | 27.6          |
| WHR                         | 0.8              | 1.0           |
| FEV1                        | 102.8            | 104.0         |
| FEV1/VC                     | 97.5             | 98.0          |
| FVC                         | 109.4            | 109.4         |

Table 2. Statistical analysis describing differences in the protein levels between controls and COPD patients

| Variable            | Healthy controls | COPD Patients | Statistical Analysis |
|---------------------|------------------|---------------|----------------------|
| DIO2 U/L            |                  |               |                      |
| Mean                | 13.1             | 50.3          | z=6.15               |
| Median              | 10.5             | 52.0          | p<0.00001            |
| Minimum             | 0.1              | 3.74          |                      |
| Maximum             | 69               | 92.7          |                      |
| Standard deviation  | 13.1             | 23.2          |                      |
| Chemerin ng/ml      |                  |               |                      |
| Mean                | 100.701          | 107.559       | z=–0.61              |
| Median              | 96.063           | 86.278        | p=0.54               |
| Minimum             | 4.693            | 5.506         |                      |
| Maximum             | 272.19           | 579.641       |                      |
| Standard deviation  | 53.805.5         | 86.696        |                      |
| HS-CRP mg/L         |                  |               |                      |
| Mean                | 3.816            | 9.721         | z=5.62               |
| Median              | 3.824            | 8.919         | p<0.00001            |
| Minimum             | 1.14             | 2.487         |                      |
| Maximum             | 9.39             | 22.807        |                      |
| Standard deviation  | 1.959            | 4.911         |                      |

COPD, chronic obstructive pulmonary disease; DIO2, deiodinase type 2; HS-CRP, high sensitive C, reactive protein

DISCUSSION

Currently, the focus of many research studies is placed on the significance of biomarkers in respiratory dysfunc-
tions. This study was intended to test serum concentrations. The authors compared serum molecule levels, such as deiodinase type 2 and chemerin, in patients diagnosed with COPD. According to the latest scientific data, DIO2 is not only involved in TH synthesis, but also in the immune- and inflammation-related mechanisms (Kwakkel et al., 2014). The investigated molecules may play an important role in COPD, i.e. a disease with inflammatory mechanisms. This study demonstrated that DIO2 and chemerin are detectable in the serum of patients with COPD and in healthy controls. The main finding of this study is that DIO2 serum levels are highly increased in the patients suffering from COPD. To the best of our knowledge, and due to the fact that we were not able to find any other data on DIO2 concentration in COPD, this is the first study addressing and investigating this particular issue.

The investigated molecule was of interest in terms of other pulmonary-related diseases. Barca-Mayo and others (Barca-Mayo et al., 2011) had investigated the role of DIO2 in response to acute lung injury (ALI) in epithelial and pulmonary endothelial cells. Similar to our results, but as the DIO2 immunoreactivity, chemerin was known to increase. This suggests that higher levels of DIO2 may play a protective role against the lung injury, as the injury was more severe in D2 knock-out mice (D2KO) (Barca-Mayo et al., 2011). A significant increase in the gene and protein levels was also found in the lung tissue in a murine ALI model. In addition, an allele of the Thr92Ala single nucleotide polymorphism, related to a higher expression level, had a protective function in severe sepsis and severe sepsis-associated ALI (Ma et al., 2011). The protective effect may be explained in a multidirectional manner. One mechanism may include the effects of DIO2 on the TH levels. It is widely known that a severe illness, such as for example acute lung injury, is characterized by a non-thyroidal illness known as decreased T4 and T3 serum levels (Pappa et al., 2011). DIO2 may work as a compensatory mechanism during the injury when hepatic/renal DIO1 production is reduced. An increase in DIO2 may be considered a protective mechanism against inflammation. DIO2 could be upregulated as part of the inflammatory pathway via NF-κB, which is also strongly involved in acute lung injury and COPD (de Vries et al., 2014). Marked up-regulation of DIO2 expression was observed by Kwakkel and others (Kwakkel et al., 2014) during acute and chronic inflammation. These researchers had suggest that DIO2 is important for macrophage phagocytic capacity. Increased DIO2 levels also result in higher TH levels, which are known to enhance the following elements of inflammation: respiratory bursts, iNOS activation, cytokine production, and bacterial killing (Chen et al., 2012). The local role of DIO2 in thyroid hormone transformation must be of importance for the inflammatory response and favors the innate immune response, as DIO2 deficiency results in proinflammatory expression (Barca-Mayo et al., 2011; Kwakkel et al., 2014). Wittmann and others (Wittmann et al., 2014), who had observed a cell type-specific and highly inducible nature of DIO2 expression by inflammation, claimed that DIO2 was involved in inflammation. Moreover, according to Chen and others (Chen et al., 2012), suppression of selenoproteins, such as DIO2, resulted in strong pro-inflammatory effects with increased expression of interleukin-1 and cyclooxygenase type 2.

To shortly sum up, an increase in DIO2 levels in COPD may by a complex issue, associated with TH synthesis, modulation and control of the immune function. Chemerin was yet another molecule of interest in our study. This adipokine demonstrates immunomodulating activity and is regulated by proinflammatory cytokines, such as TNF-α, IL-6, IL1β (Kaur et al., 2010).

The results of our study did not confirm any significant differences in the chemerin levels between patients suffering from COPD and the healthy controls. According to the best of our knowledge, based on the available literature, there are only three studies estimating chemerin in COPD. The results of these studies are not in line with our results, hence an open discussion and further exploration of the role of chemerin in COPD are necessary. Li and others (Li et al., 2016) used ELISA to detect plasma chemerin levels in COPD. These researchers had observed that chemerin levels might be a prognostic factor for COPD and might reflect lipid metabolism. The patients enrolled in that study were divided into two groups, taking into account the body mass index (thin group with BMI≤18.5 kg/m² and normal group with BMI≥18.5 kg/m²). The collected data had shown differences in the chemerin levels between the patients and healthy controls. When compared with the control group, plasma levels of chemerin were elevated in the COPD group during acute exacerbation and remission stages. The plasma levels of chemerin in the normal group were

| Healthy controls | n | r | Patients with COPD | n | R |
|------------------|---|---|-------------------|---|---|
| DIO2 & age       | 30 | 0.236575 | 50 | -0.058321 |
| DIO2 & BMI       | 30 | 0.129686 | 50 | 0.181011 |
| DIO2 & WHR       | 30 | 0.096869 | 50 | 0.009871 |
| DIO2 & chemerin  | 30 | 0.031367 | 50 | -0.058315 |
| DIO2 & HS-CRP    | 30 | -0.203578 | 50 | -0.080361 |
| DIO2 & FEV1      | 30 | -0.148490 | 50 | -0.045653 |
| DIO2 & FEV1/VC   | 30 | -0.059227 | 50 | 0.215806 |
| DIO2 & FVC       | 30 | -0.005650 | 50 | -0.129634 |
| Chemerin & age   | 30 | 0.351187 | 50 | -0.001541 |
| Chemerin & BMI   | 30 | -0.306040 | 50 | -0.095915 |
| Chemerin & WHR   | 30 | -0.246334 | 50 | 0.071330 |
| Chemerin & DIO2  | 30 | 0.031367 | 50 | -0.058315 |
| Chemerin & HS-CRP| 30 | 0.269543 | 50 | -0.095915 |
| Chemerin & FEV1  | 30 | -0.014631 | 50 | -0.127662 |
| Chemerin & FEV1/VC| 30 | -0.286143 | 50 | -0.191124 |
| Chemerin & FVC   | 30 | 0.365811 | 50 | -0.005087 |
lower in comparison with the thin group. The plasma levels of chemerin in patients suffering from COPD, who were hospitalized for half a year or that had died, were higher than the plasma levels in COPD patients without hospitalization. In addition, there was a correlation between chemerin and lipid levels, such as the total cholesterol, triglyceride, and high-density protein levels. Similarly, an increase in the plasma levels of chemerin in patients diagnosed with COPD and the relationship with total cholesterol and triglyceride levels were confirmed in the second study by Boyuk and others (Boyuk et al., 2015).

The involvement of chemerin and chemerin-related signal was also confirmed in a murine cigarette smoke (CS) COPD. The obtained results revealed that subacute and chronic CS exposure caused increased protein levels of the ChemR23 ligand and chemerin in the bronchoalveolar lavage (BAL) fluid of wild-type (WT) mice. Higher levels of the above-indicated molecules were positively correlated with massive accumulation of inflammatory cells, including neutrophils, monocytes or dendritic cells. In contrast, the inflammatory process was diminished in the BAL fluid and lungs of ChemR23 knockout mice (Demoor et al., 2011).

Results confirm that the two molecules investigated in our study are induced by the proinflammatory cytokines characteristic for COPD, such as TNF-α, IL-6, IL-1β, and are positively correlated (Kaur et al., 2010; Kwakkel et al., 2014). Therefore, we aimed to find if there is a correlation between DIO2 and chemerin levels in COPD. Nevertheless, the obtained results did not reveal any correlation between the molecules and their role in the mechanism involved in COPD. We have not observed correlation between DIO2, chemerin and the severity of the disease. Nevertheless, COPD is characterized by persistent inflammation. Increased levels of DIO2 may result in a higher production and release of proinflammatory cytokines, promoting the occurrence and development of an inflammatory process.

LIMITATIONS

The study presented here has several limitations that need to be noted. Patients in this study continued to take their prescribed COPD medications during the entire period of the study period. Nevertheless, a comprehensive search of literature relevant to the subject matter did not yield any information that could link DIO2/chemerin levels with various pharmacotherapies used for treatment of COPD. Yet another notable limitation of this study pertains to the lack of significant information about the patients’ thyroid hormone status. Undoubtedly, data demonstrating the patients’ TH serum concentrations could add valuable information about the possible association(s) between DIO2 and the thyroid hormones.

CONCLUSION

COPD is considered to be a disease of systemic inflammation. Identification of biomarkers related to this process is a developing field. Chemerin and DIO2 are significantly linked to inflammation. The study presented here confirmed an association of circulating DIO2 in COPD. While recognizing that more studies need to be conducted in this subject matter, the data presented here continues to cautiously support our initial hypothesis that DIO2 may play a role in COPD, including in its inflammatory mechanism. However, other mechanisms might also be considered as responsible for that process. Further studies are necessary to investigate the biological function of DIO2 in COPD and its role in the occurrence and development of the disease. Nevertheless, we did not confirm the role of chemerin in the disease in question. Further research and studies on larger populations are needed to prove the results for both molecules.

Conflict of interest

The authors declare no conflict of interests.

REFERENCES

Barea-Mayo O, Liao XH, DiCosmo C, Dumitrescu A, Moreno-Vinasco I, Wade MS, Sammanhi S, Mirzapazova T, Garcia JG, Refetoff S, Weiss RE (2011) Role of type 2 deiodinase in response to acute lung injury (ALI) in mice. Proc Natl Acad Sci U S A 108: E1321–E1329. https://doi.org/10.1073/pnas.1009926108

Boyuk B, Guzel EC, Atalay H, Guzel S, Mutlu LC, Kucukaycigin V (2015) Relationship between plasma chemerin levels and disease severity in COPD patients. Clin Respir J 9: 468–474. https://doi.org/10.1183/20754781.001264

Chen Y, Spilimber M, Wang X, Altenbaucher G, Hagner M, Berglund P, Gao Y, Lu T, Jonsson AB, Spilimber S (2012) Thyroid hormone enhances nitric oxide-mediated bacterial clearance and promotes survival after meningococcal infection. PLoS One 7: e44145. https://doi.org/10.1371/journal.pone.0044145

Coppola A, Meli R, Diano S (2005) Increase in circulating corticosterone and leptin levels elevates hypothalamic deiodinase type 2 in fasted rats. Endocrinology 146: 2827–2833. https://doi.org/10.1210/en.2004-1361

Coutin F, Gavaret JM, Toru-Debalhaube D, Pierre M (1990) Induction of 5'-deiodinase activity in rat astroglial cells by acidic fibroblast growth factor. Brain Res Dev Brain Res 53: 247–242. https://doi.org/10.1016/0165-3806(90)90012-N

De Palma G, Castellano G, Del Pretre A, Sozanni S, Fiore N, Loverre A, Hermenier M, Gesualdo I, Grandalano G, Bchena FP (2011) The possible role of ChemR23/Chemerin axis in the recruitment of dendritic cells in lupus nephritis. Kidney Int 79: 1228–1235. https://doi.org/10.1038/ki.2011.32

De Vito P, Balducci V, Leone S, Pecorari Z, Mangino G, Davis Pj, Davis FB, Affarbins E, Luly P, Pedersen ZJ, Incenpi S (2012) Nongenomic effects of thyroid hormones on the immune system cells: New targets, old players. Steroids 77: 988–995. https://doi.org/10.1016/j.steroids.2012.02.018

de Vries EM, Kwakkel J, Eggels I, Kalsbeek A, Barrett P, Fliers E, Boelen A (2014) NFκB signaling is essential for the lipopolysaccharide-induced increase of type 2 deiodinase in tanyocytes. Endocrinology 155: 2000–2008. https://doi.org/10.1210/endo.2013-2018

Demoor T, Bracke KR, Dupont LL, Plantinga M, Bondou B, Roy MO, Lannoy V, Lambrecht BN, Brusselle GG, Joos GF (2011) The role of ChemR23 in the induction and regulation of cigarette smoke-induced inflammation. J Immunol 186: 5457–5467. https://doi.org/10.4049/jimmunol.1005862

Fantuzzi G (2005) Adipose tissue, adipokines, and inflammation. J Allergy Clin Immunol 115: 911–919. https://doi.org/10.1016/j.jaci.2004.09.019

Fatima SS, Rehman R, Bajg M, Khan TA (2014) New roles of the multidimensional adipokine: chemerin. Peptides 62: 15–20. https://doi.org/10.1016/j.peptides.2014.09.019

Kandelko K, Miyabe Y, Takayasu A, Fukuoka S, Miyabe C, Ebisawa M, Yokoyma W, Watanabe K, Imai T, Muramoto K, Terashima Y, Sugihara T, Matsushima K, Miyasaka N, Nakani T (2011) Chemerin activates fibroblast-like synoviocytes in patients with rheumatoid arthritis. Arthritis Res Ther 13: R1. https://doi.org/10.1016/j.10.1210/en.2013-2018

Kaur J, Adya R, Tan BK, Chen J, Randeva HS (2010) Identification of chemerin receptor (ChemR23) in human endothelial cells: New targets, old players. J Allergy Clin Immunol 115: 911–919. https://doi.org/10.1016/j.jaci.2004.09.019

Köhler J (1999) Local activation and inactivation of thyroid hormones: the deiodinase family. Mol Cell Endocrinol 151: 103–119. https://doi.org/10.1016/S0303-7207(99)00480-4

Kwakkel J, Sarovsveva OV, de Vries EM, Stap J, Fliers E, Boelen A (2014) A novel role for the thyroid hormone-activating enzyme type 2 deiodinase in the inflammatory response of macrophages. Endocrinology 155: 2725–2734. https://doi.org/10.1210/en.2013-1866

Li C, Yan L, Song J (2016) Plasma level of chemerin in COPD patients and the relationship between chemerin and lipid metabolism. Zhong nan da xue xue bao. Yi xue ban = Journal of Central South University. Medical Sciences 41: 676–83. https://doi.org/10.1187/jssm.1672-7347.2016.07.003 (in Chinese)
López-Campos JL, Tan W, Soriano JB (2016) Global burden of COPD. Respirology 21: 14–23. http://doi.org/10.1111/resp.12660

Ma J, Niu DS, Wan NJ, Qin Y, Guo CJ (2015) Elevated chemerin levels in synovial fluid and synovial membrane from patients with knee osteoarthritis. Int J Clin Exp Pathol 8: 13393–13398. PMCID: 4680491

Ma SF, Xie L, Pino-Yanes M, Sammani S, Wade MS, Lestou E, Siegler J, Wang T, Infusino G, Kintses R, Flores C, Zhou T, Prabhakar BS, Moreno-Vinasco I, Villar J, Jacobson JR, Dudek SM, Garacia JG (2011) Type 2 deiodinase and host responses of sepsis and acute lung injury. Am J Respir Cell Mol Biol 45: 1203–1211. https://doi.org/10.1165/rcmb.2011-0179OC

Mariani F, Roncucci L (2015) Chemerin/chemR23 axis in inflammation onset and resolution. Inflamm Res 64: 85–95. https://doi.org/10.1007/s00011-014-0792-7

Marsili A, Aguayo-Mazzucato C, Chen T, Kumar A, Chung M, Lansford EP, Harney JW, Van Tran T, Gianetti E, Ramadan W, Chou C, Bonner-Weir S, Larsen PR, Silva JE, Zavacki AM (2011). Mice with a targeted deletion of the type 2 deiodinase are insulin resistant and susceptible to diet induced obesity. PLoS One 6: e28832. https://doi.org/10.1371/journal.pone.0028832

Mayer AS, Newman LS (2001) Genetic and environmental modulation of chronic obstructive pulmonary disease. Respir Physiol 128: 3–11. https://doi.org/10.1016/S0034-5687(01)00258-4

Pappa TA, Vagenakis AG, Alevizaki M (2011) The nonthyroidal illness syndrome in the non-critically ill patient. Eur J Clin Invest 41: 12–20. https://doi.org/10.1111/j.1365-2362.2010.02859.x

Quan N, Whiteside M, Herkenthon M (1998) Time course and localization patterns of interleukin-1beta messenger RNA expression in brain and pituitary after peripheral administration of lipopolysaccharide. Neuroscience 83: 281–93. https://doi.org/10.1016/S0306-4522(97)00350-3

Sinden NJ, Stockley RA (2010) Systemic inflammation and comorbidity in COPD: a result of “overspill” of inflammatory mediators from the lungs? Review of the evidence. Thorax 65: 930–936. https://doi.org/10.1136/thx.2009.130260

Weigert J, Obermeier F, Neumeier M, Wanninger J, Fillansky M, Bauer S, Awanisid C, Rogler G, Ott C, Schäffler A, Schölmich J, Buechler C (2010) Circulating levels of chemerin and adiponectin are higher in ulcerative colitis and chemerin is elevated in Crohn’s disease. Inflamm Bowel Dis 16: 630–637. https://doi.org/10.1002/ibd.21091

Wittmann G, Harney JW, Singru PS, Nouriel SS, Reed Larsen P, Lechan RM (2014) Inflammation-inducible type 2 deiodinase expression in the leptomeninges, choroid plexus, and at brain blood vessels in male rodents. Endocrinology 155: 2009–2019. https://doi.org/10.1210/en.2013-2154

Zeöld A, Doleschall M, Haffner MC, Capelo LP, Ménynhert J, Liposits Z, da Silva WS, Bianco AC, Kaeshskies I, Fekete C, Gereben B (2006) Characterization of the nuclear factor-kappa B responsiveness of the human dio2 gene. Endocrinology 147: 4419–4429. https://doi.org/10.1210/en.2005-1608

Zhou Q, Fu Y, Hu L, Li Q, Jin M, Jiang F (2018) Relationship of circulating chemerin and omentin levels with Th17 and Th9 cell immune responses in patients with asthma. J Asthma 55: 579–587. https://doi.org/10.1080/02770903.2017.1355378