HLA System and its Participation in Recurrent Pregnancy Loss

Laiba Ajmal1, Sidra Ajmal2, Maleeha Ajmal3, Gul Nawaz3, Rabail Hassan Toor4, Hooria Younas1, Tassaduq Hussain Sheikh5 and Raazia Tasadduq1*

1Department of Biochemistry, Kinnaird College for Women, 93 Jail Road, GOR 1, Lahore, Pakistan
2Department of Chemistry and Biochemistry, University of Oklahoma, Norman, Oklahoma, United States
3Internal Medicine, Marshfield Clinic Health System, Marshfield, Wisconsin, United States
4School of Biological Sciences, University of Punjab, Quaid-I-Azam Campus, Lahore, 54590, Pakistan
5Department of Anatomy, Central Park Medical College, Lahore, Pakistan

ABSTRACT

A successful pregnancy is a biological paradox as the maternal immune system accepts and tolerates the semi allogenic fetus. Continued tolerance of the fetus relies on an optimal immune privileged environment that allows the developing fetus to avoid recognition and rejection. Any dysregulation in the crosstalk between the host’s immune system and the fetal allograft would result in rejection of the fetus, subsequently leading to miscarriages. The functioning of human leukocyte antigen (HLA) system at the maternal-fetal interface plays a pivotal role in acceptance of the fetus. Alterations in the modulated activity of HLA system influence the fetal tolerance. Therefore, this review will provide an insight on how the working of HLA at the maternal-fetal interface contributes towards recurrent pregnancy losses (RPL). Antigen hiding is a phenomenon that contributes towards immune evasion by the fetal cells as the placental barrier lacks HLA expression. However, the HLA molecules that are expressed by the extravillous trophoblast modulate maternal immune activity towards a protective function, leading to acceptance of the fetus. Further studies encompassing the functioning of HLA system at maternal fetal interface would prove to be beneficial for the affected couples.

INTRODUCTION

A successful pregnancy requires the mother and fetus, two genetically different entities, to coexist (de Luca Brunori et al., 2003). The mother’s immune system, that is in proximity with the fetal antigens, tolerates the semi allogenic fetus and does not reject it (Hunt and Orr, 1993). The maternal immune system is not silent but recognizes the fetal antigens and actively diverges towards a protective response (Leber et al., 2021; Aghaeepour et al., 2017; Colucci, 2019). Originally, recurrent pregnancy loss (RPL), affecting 1-2% women, had been described as three or more repetitive fetal losses before the completion of twentieth week of gestation (Baek et al., 2007). Recent classification defines RPL as loss of two or more sequential pregnancies, thus increasing the percentage of the affected couples to 5% (Ford and Schust, 2009). Despite the various etiological factors for RPL, in many couples the causative factor remains unknown (Branch et al., 2010; Tasadduq et al., 2021). Unexplained recurrent pregnancy losses are believed to occur due to immune responses that induces maternal allogenic immune reaction against the fetus (Rocklin et al., 1982). Dysregulation in the maternal fetal immune crosstalk is associated with gestational complications and if the protective and tolerant immune response fails, pathological pregnancy state such as miscarriage might occur (Moghraby et al., 2010).

Abbreviations

APC, Antigen Presenting Cell; GM-CSF, Granulocyte-Macrophage Colony Stimulating Factor; HLA, Human Leukocyte Antigen; ILT, Immunoglobin Like Transcript; KIR, Killer Immunoglobulin Like Receptors; NK, Natural Killer; NKG, Natural Killer Group; Treg Cells, Regulatory T cells; RM, Recurrent Miscarriage; RPL, Recurrent Pregnancy Losses; uNK, Uterine Natural Killer.
Perturbations in the activity of the human leukocyte antigen (HLA) system come under the umbrella term of immune system etiology of RPL since the expression of HLA genes and their interlinked regulatory factors play a vital role at feto maternal interface and may lead to fetal loss (Christiansen, 2013). The villous (syncytiotrophoblast) trophoblast cells at the placenta protect the fetus from possible maternal rejection by acting as an immunological barrier, as these lack the expression of HLA antigens (Christiansen, 2013; Sibai, 1991). However, alleles of HLA-C, HLA-E and HLA-G manifest on the extravillous trophoblast cells (Christiansen, 2013). In the light of the existing evidence, this review explores the functioning of the HLA system at the placental trophoblast that may contribute towards rejection of the fetus.

**ROLE OF HLA SYSTEM IN RPL**

**HLA-C, and its duality at the maternal fetal interface**

HLA-C is the only classical HLA class I antigen expressed on trophoblast cells and is believed to support placentation (Singh et al., 2019). HLA-C not only interacts with natural killer (NK) cells but also accounts for the autologous identification of the fetal cells. Expressed on the extravillous trophoblasts (EVT), HLA-C binds to the killer immunoglobulin like receptors (KIRs) on NK cells and promotes trophoblast invasion (Christiansen, 2013). KIRs regulate the activity of NK cells by conferring either an inhibitory or activating signal. The KIR genes are categorized into two haplotypes, A and B. The most frequently occurring KIR A haplotype consists of inhibitory genes whereas KIR B haplotype, that is more genetically variable, is the activating KIR. Two allotype groups recognized for HLA-C are HLA-C1 and HLA-C2. HLA-C1 is involved in the activation of KIR2DS2 and inhibition of KIR2DL2/3 receptors whereas HLA-C2 activates KIR2DS1 and inhibits KIR2DL1 receptors by acting as ligands (Christiansen, 2013).

The KIR B haplotype is also believed to confer protection to the mother from gestational dysfunctions and hence its absence can increase the risk of complications (Hiby et al., 2010). This haplotype consists of KIR2DS1 receptor which upon binding to its ligand (HLA-C2) stimulates NK cells to secrete chemokines such as GM-CSF that support trophoblast invasion (Bari et al., 2009) (Fig. 1). In contrast, females with homozygous KIR A haplotype and heterozygous HLA-C1/HLA-C2 with fetal cells having homozygous C2 are at an increased risk of complicated pregnancy (Obeagu, 2015). This is because KIR AA genotype encodes two copies of the KIR2DL1, the inhibitory KIR for HLA-C2 that interacts with the fetal HLA-C2 allotype, inhibiting the NK cells and results in defective placentation (Hiby et al., 2004, 2014). Furthermore, decreased occurrence of KIR2DS1 has been observed in women undergoing repetitive miscarriages (Christiansen, 2013). It has been noted that when both the partners in RPL patients have the HLA-C2, KIR2DS1 expression is lowered (Wang et al., 2007). These findings implicate the role of KIR2DS1 in activation of NK cells to produce trophoblast supporting cytokines. Furthermore, women with history of preeclampsia and RPL have greater chance of carrying the KIR (AA) genotype with concurrent fetal HLA-C2 expression. Hiby et al. (2010) reported occurrence of RPL due to variations in paternally inherited HLA-C2.

![Fig. 1. Modulation of maternal immune response by HLA-C is influenced by its interaction with the activating or inhibitory KIR.](image)

Among the HLAs that are expressed on the trophoblast, HLA-C is considered to have the greatest number of polymorphisms (Christiansen, 2013). Some of these polymorphisms have been reported to have deleterious effects on the trophoblast proliferation by inhibiting the NK cells, consequently restricting growth, and leading to RPL (Christiansen, 2013; Hiby et al., 2010; Nielsen et al., 2010; Moffett et al., 2015).

HLA-C also establishes maternal tolerance by modulating the activity of CD8+ T cells and regulatory T cells (Treg cells). The decidual CD8+ T cells express significant levels of the co-inhibitory molecules; Cytotoxic T lymphocyte associated protein 4, the programmed cell death 1 and lymphocyte activation gene 3 (van der Zwan et al., 2018). As the maternal uterine CD+8 T cells interact with the paternal HLA-C molecules, these co-inhibitory cellular molecules also interact with their respective co-receptors on fetal cells and downregulate...
the cytotoxic activity of the CD8+T cells (van der Zwan et al., 2018). HLA-C also induce maternal Treg cells to release anti-inflammatory mediators and thus prevent inflammatory reactions in response to paternal antigens (Tilburgs et al., 2006). Decreased number and activity of Treg cells has been found in women suffering from RPL and preeclampsia, both in the decidua and peripheral blood (Jin et al., 2009). Moreover, it has been speculated that the Treg differentiation occurs after the maternal T cell recognition of the fetal HLA-C but the mechanisms pertaining to this phenomenon remain unexplored.

**HLA-E modulator of NK response**

HLA-E is involved in maternal immunosuppression at the fetal-maternal interface and hence might also contribute to RPL (Tripathi et al., 2006). HLA-E is recognized by the CD94/NKG2A inhibitory receptors and the CD94/NKG2C activating receptors. Since all the extravillous trophoblast cells have HLA-E expression and the decidual NK cells express the CD94/NKG2, the interactions between these aids the trophoblast cells in evading the NK cell mediated killing and hence provide protection from maternal immune response (Trowsdale and Moffett, 2008). Hence, it is the degree of the functioning of the activating and inhibitory NK receptors that dictate the NK cell response towards fetus (Strauss-Albee et al., 2014; Le Gars et al., 2019).

The surface expression of HLA-E is dependent on a conserved nonamer that is derived from the signal sequence of class I HLA (Lee et al., 1998). The specificity of this nonamer is important as a highly restricted repertoire can be loaded on HLA-E (Tripathi et al., 2006). HLA-E presents signal peptides of HLA-A, B, C and G to inhibitory receptor NKG2 on NK cells (Braud et al., 1998). In other words, HLA-E specifically interacts with NKG2A to inhibit the NK cells (Carretero et al., 1998). This result in subsequent target cell protection by the interactions of HLA-E/peptide and CD94/NKG2 receptor on NK cells (Trowsdale and Moffett, 2008).

It has been elucidated that the NK cells with the CD94/NKG2A have a greater affinity for the HLA-E/peptide than the CD94/NKG2C (Llano et al., 1998; Vales-Gomez et al., 1999). In addition, this affinity is highly dependent on the nonamer peptide complexed with HLA-E (O’Callaghan et al., 1998). Interactions have postulated that it is the specific CD94/NKG2 mediated recognition of the HLA-E that plays a role in the cellular communications at maternal-fetal interface (Llano et al., 1998). In the placenta, the leader peptides for HLA-E are restricted to HLA-G and HLA-C due to its hydrophobic characteristics (O’Challaghan et al., 1998). Moreover, it has been established that the HLA-E preferentially presents the HLA-G leader peptides (Llano et al., 1998).

Bioimaging of trophoblast cell lines have shown increased co-localization of HLA-G and HLA-E reflecting upon their dependent surface expression (Jabeen et al., 2013). The HLA-E/HLA-G complex inhibits the cytotoxic activity of the NK cells by interacting with CD94/NKG2A and may also activate the NK cells to produce supporting chemokines by interacting with CD94/NKG2C (Ishitani et al., 2006) (Fig. 2).

**Fig. 2.** Effect of interactions between HLA-E and NKG2 in normal pregnancy and pregnancy loss.

HLA-E is recognized by the CD94/NKG2A and modulates the cytotoxic activity of NK cell. Decreased expression of NKG2A on uNK cells has been associated with recurrent miscarriages (Karami et al., 2012). Interaction between HLA-E and NKG2A inhibits the cytolytic pathway of NK cells and instead promote endometrial invasion. On the other hand, lack of this interaction due to loss of NKG2A would result in activation of the cytolytic pathway and lysis of the extravillous trophoblast (Karami et al., 2012; Beytamouni and Ghanem, 2016) (Fig. 2). Women with repeated pregnancies have increased population of memory uNK cells that express higher levels of NKG2C. Therefore, interaction between NKG2C and HLA-E result in increased secretion of INF-γ and VEGFα that promote vascularization and placentation (Gamliel et al., 2018).
HLA-E is a non-classical HLA class I gene with two alleles of practical importance, E*0101 and E*0103, that are distinguished on the basis of non-synonymous substitution from arginine to glycine at codon 107 (Grimsley and Ober, 1997; Strong et al., 2003). Compromised activity of NK cells to protect against maternal immune system has been associated with decreased expression and stability of HLA-E allele 0101 (Tripathi et al., 2006). In Egyptian RPL population, the frequency of allele 0101 had been found to be greater and similar results have been reported from African, Indian, Caucasian and Hispanic populations (Tripathi et al., 2006; Grimsley and Ober, 1997; Mosaad et al., 2011). This association is found to be stronger in homozygous RPL women (Tilburgs et al., 2006). The increased frequency of the 0101 allele in RPL women can be connected to the varying levels of the protein as the surface levels of HLA-E are lower when cells have the 0101 allele than when they have the 0103 allele (Strong et al., 2003). In conjunction is the dependence of HLA-E with the HLA-G nonamer. The E*0103 has a greater affinity for the HLA-G derived nonamer than the E*0101, the complex also having greater thermal stability supporting the decreased expression of HLA-E when the allele E*0101 is present (Strong et al., 2003). This links the allelic distribution with the etiology of RPL. However, the greater frequency of 0103 has been documented in Chinese and Japanese populations (Grimsley and Ober, 1997; Kimkong et al., 2003). Hence, the association between HLA-E and fetal loss remains inconclusive due to conflicting results from different populations (Strong et al., 2003). Therefore, further investigations are needed to explore the linkage between the different allelic frequencies of HLA-E and recurrent miscarriages (Dhal and Hviid, 2012).

**HLA-G, a non-classical mediator of immune tolerance at maternal-fetal interface**

HLA-G, another non-classical HLA molecule, is expressed on the fetal placental cells and contributes towards implantation and maternal acceptance of the fetus. HLA-G mRNA levels are associated with increased embryo cleavage rate along with successful implantation of the embryo (Hviid et al., 2004). A maternal immune response is not generated against the placenta at the fetomaternal interface, even though the cytotoxic lymphocyte does not express the class I molecules, HLA-A and HLA-B. Normally the absence of class I HLA molecules is recognized by NK cells which attack the cells lacking the class I molecules (Munz et al., 1997). The HLA-G interacts with the decidual immune cell population and supports fetal tolerance by inhibiting the cytotoxic activity of NK cells and CD8+ T cells; suppress the proliferation of CD4+ T cells; reduces the B cell activity and stimulate the proliferation and activity of T regulatory cells (Hviid et al., 2004) (Fig. 3). HLA-G is able to execute these functions by communicating with cells harboring the receptors LILRB1 (inhibitory receptors on APC), CD160 (receptor on T lymphocytes, NK and endothelial cells) and KIR2DL4 (receptor on NK cells) (Hviid et al., 2004; Persson et al., 2020).

![Fig. 3. HLA-G, a mediator of cross-talk between EVT and maternal immune cells.](image-url)

The uNK cells of women with repeated pregnancies are characterized by increased expression of immunoglobulin like transcript 2 (ILT-2), which is alternatively known as LILRB1 (Gamliel et al., 2018). In such women increased secretion of INF-γ and VEGFα has been observed upon interaction of ILT-2, an inhibitory receptor on NK cells, with HLA-G and NKG2C with HLA-E (Gamliel et al., 2018). Increased production of INF-γ and VEGFα by uNK cells in multigravid women support vascularization and maintenance of the decidua (Gamliel et al., 2018). Hence it has been demonstrated that the ILT-2 acts as an ‘activating’ receptor in the decidua and its interaction with HLA-G supports pregnancy due to increased expression of INF-γ and VEGFα (Gamliel et al., 2018).

Another fascinating way in which HLA-G induce tolerance is via its interaction with inhibitory receptors, immunoglobulin like transcript 2 (ILT-2) and 4 (ILT-4). These receptors are present on dendritic cells, monocytes, and macrophages (Colonna et al., 1998). HLA-G binds
preferentially with these inhibitory receptors and thus results in prevention of cytotoxic activity of CD8+ T cells, as CD8 T cells and ILT-2/4 engage in competition for binding with HLA-G (Shiroishi et al., 2003). Moreover, both membrane bound and soluble HLA-G inhibit the allopregnancy of T cells through interactions with ILT-2 and ILT-4 (Naji et al., 2007).

Soluble HLA-G is produced by fetal trophoblast cells which is taken up by the NK cells and transported to the endosome containing KIR2DL4. Binding of soluble HLA-G and KIR2DL4 in endosome leads to cellular senescence of NK cells and concurrent release of cytokines that promote vascular remodeling (Rajagopalan, 2014). The significance of the KIR2DL4 can be deduced by the findings that the uNK cells of fertile women have a higher protein level of this receptor than the women experiencing RPL (Yan et al., 2007). Interaction of HLA-G and KIR2DL4 does not stimulate the cytotoxic activity of NK cells but activates the production and release plethora of chemokine and cytokine such as IL-6, IL-8, IL-1β, INF-γ and TNF-α that support vascularization (Li et al., 2009; Rajagopalan and Long, 2012; Benson and Caligiuri, 2018).

Decreased levels of HLA-G expression and polymorphisms associated with HLA-G loci have been implicated with pregnancy complications such as recurrent miscarriages and preeclampsia. Mothers with successful pregnancies have been found to have a greater amount of soluble HLA-G in the serum in comparison to women having downregulated HLA-G and KIR2DL4 cells (Athanassakis et al., 1999; Vianna et al., 2016). It has been reported that women with decreased concentrations of soluble HLA-G are at a greater risk of developing placental abruption during pregnancy (Durminova et al., 2013).

As HLA-G gene expression is implicated in successful implantation, various polymorphisms of the gene that result in decreased production of HLA-G have been studied as a possible cause of RPL (Grimslay and Ober, 1997; Fritz and Speroff, 2010; Cecati et al., 2011). A homozygous 14-bp insert in exon 8 in HLA-G gene results in unstable mRNA leading to lower production of soluble HLA-G (Singh et al., 2019; Takakuwa et al., 1999). This polymorphism has been associated with repetitive fetal loss and unsuccessful IVF cycles (Hvid et al., 2004). Moreover, decreased expression of HLA-G is reported to suppress the decidual NK cells and could lead to RPL (Guo et al., 2017). Greater fetal loss rates have been associated with polymorphisms 725C/G, 1573T/C and 1746C/A in the promoter region of the HLA-G (Ober et al., 2003; Yazdani et al., 2018). Moreover, polymorphisms of HLA-G with concurrence of HY antibodies in male offspring can lead to not only reduced birth weight but RPL as well (Krieg and Westphal, 2015). Considering this, observing HLA-G protein levels of can be used as a diagnostic tool for pathogenesis of gestational complications.

**HLA-D**

Class II HLA constitute HLA-DR, HLA-DQ and HLA-DP and are involved in allograft rejection (Ober, 1998). Class II antigens present peptides to CD4+ T cells and are predominantly expressed by macrophages, dendritic cells, and B cells (Owen et al., 2013). As the HLA class II antigens are absent at the feto-maternal interface, this allows continued survival of the fetus as the mother’s immune system is unable to recognize it as a foreign entity.

In normal pregnancies, the trophoblast cells not only lack HLA-DR4 antigens but also the HLA-DR4 enhancing interferons (Hunt and Orr, 1993). In contrast, trophoblast cells have been reported to express the HLA-DR4 in complicated pregnancies such as preeclampsia (Labarrere and Faulk, 1995). Enhanced production of cytokines such as IL-4, INFγ and TNF have also been observed due to the dysfunctional endothelial state (Omu et al., 1995) which augment the class II HLA expression (Omu et al., 1998).

Association between class II HLA and retarded intrauterine growth has been established with occurrence of HLADR4 and HLAD1w2 haplotypes (Omu et al., 1998). In women with RPL, increased occurrence of HLA-DRB1 allele, HLA-DR and HLA-DQB1 has been observed indicating possible association between pregnancy loss and HLA-D (Cecati et al., 2011; Aruna et al., 2011; Gharesi-Fard et al., 2014). HLA-DR also has been observed in the syncytiotrophoblast cells in placenta of preeclampsia women (Tersigni et al., 2018). HLA-DQB1 allele 03:03:02 has been implicated with a greater risk of miscarriages in South Indian women (Takakuwa et al., 1999). Moreover, increased expression of HLA-DQ2/DQ8 has been noted in RPL women. The HLA-DQ2/DQ8 haplotype code proteins that are involved with peptide presentation to T-lymphocytes (D’ippolito et al., 2016).

An intriguing finding has been reported by the studies that linkage disequilibrium exists between HLA-G*010102 and HLA-DR3. An increased risk for RPL had been established with HLA-DR3/G*010102 and HLA-DR1/G*010102 haplotypes than with the independent occurrence of HLA-DR3, HLA-DR1 and HLA-G*010102 (14-bp insertion in the 3’UTR) alleles (Hvid and Christiansen, 2005). Moreover, family studies indicate the occurrence of linkage disequilibrium between the genes of HLA-DR1 and HLA-DR3 or genes in linkage disequilibrium to these effect the risk of RPL (Christiansen et al., 1999). This can be explained by the occurrence of increased secretion of cytokines such as TNF-α (harmful
for normal pregnancy), predisposed by the HLA class II alleles due to linkage disequilibrium with TNF-α alleles (Christiansen et al., 1999; Pociot et al., 1993).

Patients with family history of pregnancy losses are at a higher risk of suffering from RPL (Zhang et al., 2010). Moreover, siblings of patients suffering from idiopathic recurrent miscarriages share genetic factors such as HLA class II alleles and HLA-G 14-bp insert, that increase their risk of miscarriage (Kolte et al., 2011). Female relatives of patients having the alleles HLA-DRB1*01 or HLA-DRB1*03 are considered at a risk for pregnancy losses especially because HLA-DRB1*03 has been associated with RPL, with the disposition worsened in women with secondary RPL or higher pregnancy losses (Christiansen et al., 1995; Kruse et al., 2004). Therefore, a significant association of certain HLA-D alleles has been linked with gestational complications.

**Role of HLA-antibodies**

The extravillous trophoblast, that is in direct contact with the maternal cells expresses the non-classical HLA-E and HLA-G along with classical HLA-C. The chimeric cells in the peripheral circulation direct the production of HLA antibodies as these express all class I and class II HLA antigens. About 30% of healthy women form HLA antibodies during gestation and the titer of antibodies increase till 28 weeks of pregnancy. In addition, the antibodies may persist till the conception of the next pregnancy (Regan et al., 1991; van Kampen et al., 2001). The presence of these antibodies has been reported to be higher during the first trimester among multiparous women compared to nulliparous women (Meuleman et al., 2016).

The occurrence of HLA antibodies in early pregnancy is related to decreased probability of live birth in women with RPL (Nielsen et al., 2010). Antibody dependent rejection of the fetus might occur because of antibody mediated complement fixation against the paternal HLA antigens. In women with RPL, a greater deposition of C4d, a marker of the antibody dependent complement activation, has been reported at the maternal fetal interface (Meuleman et al., 2016). Circulating maternal anti-HLA class I antibodies have been associated with C4d appearing in fetal umbilical cord endothelium in women with premature births (Lee et al., 2011). Moreover, the frequency of complement fixing antibodies is greater in patients with a history of complicated birth (Moffett et al., 2015). Therefore, it can be inferred that the concurrent increase in complement fixation mediated by HLA-antibodies at the trophoblast may lead to the repetitive pregnancy losses (Meuleman et al., 2016).

It has been reported that the probability of secondary recurrent miscarriage (RM) is higher for pregnancies preceded by the delivery of a boy. Other studies have reported a higher amount of HLA antibodies in women who had delivered a boy prior to the miscarriage (Nielsen et al., 2008, 2010). Mounting of an abnormal immune reaction against male specific minor histocompatibility antigens (H-Y antigens) has been described as one of the contributing factors towards fetal loss (Nielsen et al., 2009).

Patients suffering from secondary RM have increased allogeneic or immunologically incompatible fetal cells appearing in the circulation of the mother. The higher frequency of the antibodies in individuals having a firstborn son and/or history of miscarriages can be linked to the increased fetal cell microchimerism (Wang et al., 2007). The fetal cells persist in mother’s circulation for years after delivery and their number has been found to be greater in secondary RM patients with a prior boy (Nielsen et al., 2010).

The occurrence of HLA antibodies is not implicated with any adverse effects in normal fertile population (Regan et al., 1991). However, the prevalence of class I HLA antibodies has been found to be higher in patients who suffer from placental abruption and have been associated with increased activity of monocytes in the fetal circulation. This leads to the heightened immune response against the developing fetus (Steinborn et al., 2004).

The detrimental effects of HLA antibodies in RPL patients have been attributed to the dysfunctions of different components of the immune system. The placental mesenchyme can be stimulated to express the class II HLA molecules in response to INFγ. As a result, maternal HLA antibodies can target the HLA-C or the HLA-DR/HLA-DQ (Christiansen et al., 2006). Inflammatory response due to repetitive pregnancy loss at the maternal-fetal interface damages the trophoblast barrier, allowing the anti-HLA antibodies to target the fetal tissue (Christiansen et al., 2006). It is probable that women with RPL might have different subtypes of anti-HLA antibodies in contrast to normal obstetric women (Nielsen et al., 2010). The maternal immune response diverges from fetal tolerance to fetal rejection as more amount of pro-inflammatory mediators were reported to be present in RPL patients when fetal antigens were presented to maternal APCs (Steinman et al., 2003). Increased microchimerism can also cause engraftment of cells leading to increase in mother’s anti-HLA cell activity which supports the cytotoxic T cells (Llano et al., 1998; van Kampen et al., 2001). The association of HLA-antibodies with RPL considering specific immunogenic dysfunctions needs to be further investigated to establish a concrete relation.
HLA SHARING: A WAY OF PREVENTING HOMOZYGOSITY

For successful implantation, the maternal immune system needs to be adapted for pregnancy and this is aided by HLA sharing (de Luca Brunori et al., 2003; Colbern et al., 1994). Studies have backed the notion that HLA compatibility among couples lead to greater RPL rates as inbred populations have been found to be more prone to RPL (Fritz and Speroff, 2010; Regan et al., 1991). HLA alleles have been shown to possess positive linkage disequilibrium. Increased incidence of identical HLA-A and HLA-B alleles were found in families with increased occurrence of RPL (Christiansen et al., 1989).

During pregnancy, the mother’s immune system initiates an alloimmune response to maintain pregnancy the activation of which is mediated by HLA molecules. In the case when both the partners share the same HLA antigens possible tissue rejection sequence against the fetus is activated and hence it is believed that repeated miscarriages occur in these situations (Gharesi-Fard et al., 2014).

Maternal hypo responsiveness is invoked against paternal antigens in pregnancy, a consequence of excessive HLA antigenic sharing among couples and resulting in pregnancy failure (Beer et al., 1981). The lack of fetal tolerant inducing response has been linked to failure in maternal recognition of the fetal tissues, a consequence of the compatibility of the HLA system among couples (Beer et al., 1981; Ho et al., 1990). Hence, HLA incompatibility and not histocompatibility as presumed in earlier studies, confers a certain reproductive advantage (Beer et al., 1981; Hauck and Ober, 1991).

An excessive HLA sharing has been observed in primary and secondary RM patients with the primary RM patients sharing the HLA-A and HLA-DQ antigens and both the primary and secondary RM patients sharing three or more HLA-A, HLA-B, HLA-DR and HLA-DQ antigens (Ober et al., 1993). It has been observed that partners who share HLA-DQA1 alleles have a significant chance of miscarriage early in pregnancy (Ober et al., 1993). The HLA-DQ might interfere with implantation and hence results in loss of fertility. This is due to the persistent exposure of the fetal cells to the maternal uNK cells that have enough cytotoxic activity to be detrimental to the trophoblast (van der Zwan et al., 2018).

CONCLUSION

A regulated functioning of the immune system at the maternal fetal interface is a prerequisite for a healthy pregnancy. Certain immunomodulatory compounds establish fetal tolerance, in which HLA system plays a pivotal role. HLA antigens interact with maternal immune system and modulate its activity to hide and accept the developing fetus. Hence, any dysfunction in HLA family leads to fetal rejection and may cause RPL. For couples with undetermined etiology of RPL, HLA typing and testing at cellular and molecular levels will be beneficial and provide better prognosis. Future laboratory and clinical investigations must consider HLA driven mechanisms and variables that promote fetal tolerance and could be manipulated to affect the pregnancy outcome.

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