Controls over carbon storage and turnover in high-latitude soils

Hobbie, SE
Schimel, JP
Trumbore, SE
et al.

2000-12-01

10.1046/j.1365-2486.2000.06021.x

4.0
Controls over carbon storage and turnover in high-latitude soils

SARAH E. HOBBIIE,* JOSHUA P. SCHIMEL,‡ SUSAN E. TRUMBORE¶ and JAMES R. RANDERSOn§
*Department of Ecology, Evolution, and Behavior, University of Minnesota, 100 Ecology Bldg, 1987 Upper Buford Circle, St. Paul, MN 55108, ‡Ecology, Evolution, and Marine Biology, University of California, Santa Barbara, CA 93106, ¶Geochemistry Department, University of California, Irvine, CA 92717-3100, §Center for Atmospheric Sciences, University of California, Berkeley, CA 94702, Institute for Arctic Biology, University of Alaska, Fairbanks, AK 99775, USA

Abstract

Despite the importance of Arctic and boreal regions in the present carbon cycle, estimates of annual high-latitude carbon fluxes vary in sign and magnitude. Without accurate estimates of current carbon fluxes from Arctic and boreal ecosystems, predicting the response of these systems to global change is daunting. A number of factors control carbon turnover in high-latitude soils, but because they are unique to northern systems, they are mostly ignored by biogeochemical models used to predict the response of these systems to global change. Here, we review those factors. First, many northern systems are dominated by mosses, whose extremely slow decomposition is not predicted by commonly used indices of litter quality. Second, cold temperature, permafrost, waterlogging, and substrate quality interact to stabilize soil organic matter, but the relative importance of these factors, and how they respond to climate change, is unknown. Third, recent evidence suggests that biological activity occurring over winter can contribute significantly to annual soil carbon fluxes. However, the controls over this winter activity remain poorly understood. Finally, processes at the landscape scale, such as fire, permafrost dynamics, and drainage, control regional carbon fluxes, complicating the extrapolation of site-level measurements to regional scales.

Keywords: Arctic, boreal, carbon, decomposition, global change, soil

Introduction

Understanding the factors that control fluxes of carbon from arctic and boreal soils is essential for predicting how carbon fluxes in these regions will respond to global change. Arctic and boreal regions have long been recognized as potentially important players in the global carbon cycle because of the large stocks of carbon stored in northern soils. Estimates of arctic and boreal soil carbon pools vary considerably; they range from 90 to 290 Pg (1015 g) of carbon in upland boreal forest soils, 120–460 Pg of carbon in peatland soils, and 60–190 Pg of carbon in arctic tundra soils (Schlesinger 1977; Post et al. 1982; Post et al. 1985; Oechel 1988; Gorham 1991; Chapin & Matthews 1992). In total, this represents 20–60% of the global soil carbon pool, and 1–2 orders of magnitude more carbon than is emitted annually to the atmosphere from fossil fuel burning and deforestation. Yet the magnitude and even the sign of feedbacks from northern regions to climate change induced by rising concentrations of CO2 in the atmosphere are disputed (Lashof 1989; Post 1990; Shaver et al. 1992; Oechel et al. 1993; Goulden et al. 1998).

Despite the importance of arctic and boreal ecosystems in the global carbon cycle, our understanding of controls over carbon storage in these systems is lacking. For example, observations of atmospheric δ13CO2 suggest a large terrestrial sink (>3 Pg C yr⁻¹ in 1992 and 1993) in temperate and boreal regions of the northern hemisphere (Ciais et al. 1995). In contrast, estimates of temperate, boreal, and arctic carbon uptake based on forest inventories, forward modelling simulations, and regional extrapolations yield a range of source and sink estimates with a much smaller absolute magnitude (Dixon et al. 1994; Schimel et al. 1995; Houghton 1996; McGuire et al. 1996).
Identifying why these estimates of terrestrial carbon storage differ is critical for predicting future concentrations of atmospheric CO2 over the next several centuries. The discrepancy among carbon flux estimates arises in part from our lack of understanding about the controls over carbon residence times in mid- and high-latitude ecosystems. Within forest stands, rates of carbon turnover are regulated by species composition, allocation, litter decomposition, and fine root turnover (Vogt et al. 1986). At the regional scale, residence times of carbon are also determined by mortality events such as fires, harvesting, and insect outbreaks (Kurz & Apps 1994; Harden et al. 1997). The δ13CO2, forest inventory, and forward modelling approaches to calculating terrestrial carbon fluxes depend on accurate descriptions of these processes.

In the δ13CO2 approach, rates of terrestrial carbon turnover determine the isotopic disequilibrium of the gross exchange between the land and the atmosphere. Disequilibria are created in all terrestrial ecosystems by decreasing atmospheric δ13CO2 driven by fossil fuel emissions (that are relatively depleted in δ13C). Because of this, carbon released through respiration can be significantly enriched in δ13C compared to that currently fixed from the atmosphere if the residence time of carbon in plants and soils is long enough (Tans et al. 1993; Fung et al. 1997). Terrestrial CO2 uptake also causes atmospheric CO2 to become enriched in δ13C because of discrimination against δ13C during photosynthesis (Lloyd & Farquhar 1994). Because disequilibria and terrestrial sinks both cause the atmosphere to become relatively enriched in δ13C, they can both potentially explain the recent atmospheric δ13CO2 trend (Francey et al. 1995). In other words, if analyses of atmospheric δ13CO2 (e.g. Ciais et al. 1995) had assumed slower turnover of terrestrial carbon, they would have obtained a smaller estimate of a boreal carbon sink.

In the forest inventory approach, carbon turnover dynamics determine the length of the recovery (and uptake) period following a disturbance. During the 19th and early 20th centuries, large tracts of forest were cleared across Europe and the east coast of North America (Houghton 1994; Kohlmaier et al. 1995). With low rates of slash and soil decomposition and carbon stocks that were decimated at the time of disturbance, many of these previously cleared forests may currently function as carbon sinks (Kauppi et al. 1992). There is evidence, however, for increasing rates of disturbance and thus a mean annual release of carbon in other temperate and boreal regions, including the west coast of North America (Kasischke et al. 1999).

In forward modelling analyses, the magnitude of the carbon sink is directly proportional to the rate of change in NPP and the residence time of carbon with ecosystems (Thompson et al. 1996). In recent decades, increasing levels of atmospheric CO2 (Keeling et al. 1989) and nitrogen deposition (Holland et al. 1997) may have stimulated rates of net primary production (NPP) in high-latitude ecosystems (Melillo et al. 1996). A critical component of forward modelling simulations is the response of decomposition to temperature, and in particular, the response during winter months when low but persistent rates of respiration under the snow pack can influence seasonal and annual carbon balances (McGuire et al. 2000b).

Accurate estimates of carbon residence times in soils are necessary for accurate descriptions of the present carbon balance of high-latitude ecosystems. In this paper we review the major controls over carbon turnover through soils in arctic and boreal ecosystems, highlighting those controls that are unique to high latitudes and pointing out how current biogeochemical models might be misrepresenting these processes for northern systems.

Controls over litter decomposition

As in other regions, the major controls over decomposition of fresh litter in arctic and boreal regions include temperature, moisture, and substrate quality. Numerous studies have demonstrated the importance of cold temperature in limiting rates of decomposition in arctic and boreal systems. In laboratory incubations, litter decomposition rates increase from subzero temperatures (−5°C) up to 25°C (Flanagan & Veum 1974; Rosswall 1974; Heal et al. 1981; Klein & Schimel 1995). Optimal moistures average near 400% of dry weight, but decrease at higher temperatures (Flanagan & Veum 1974). Across broad arctic-boreal regions, both mean annual temperature and actual evapotranspiration are good predictors of decomposition rates (Johansson et al. 1995).

Substrate quality, particularly carbon chemistry, is also a strong limitation to decomposition rate in arctic and boreal ecosystems. Decomposition is generally slower for litter with high lignin concentrations and low concentrations of soluble carbohydrates (Van Cleve 1974; Heal et al. 1981; Hobbie 1996). Large differences occur among plant growth forms and among plant organs within species. Woody stems, mosses, lichens, and evergreen leaf litter decompose slowly, while deciduous and forb leaf litter decompose rapidly (Heal & French 1974; Moore 1984; Hobbie 1996; Shaver et al. 1997).

Several studies have suggested that the influence of substrate quality over decomposition can be large relative to the effects of temperature in arctic and boreal systems, perhaps reflecting the diversity of growth forms...
found in these regions. Differences in decomposition rate among Alaskan tundra species were sometimes larger than the effects of a 6°C warming (Hobbie 1996).

Similarly, faster decomposition rates in birch than in spruce forests in Alaskan taiga were caused primarily by the better substrate quality of birch litter rather than by the warmer soils in birch forests (Flanagan & Van Cleve 1983).

To evaluate the broad-scale controls over decomposition rate in arctic and boreal regions, we searched for general patterns among a number of published decomposition studies (Appendix). To allow comparison, we restricted our analysis to those studies that measured decomposition rates of senesced tissues using the litter bag technique. Studies of decomposition of organs other than leaves were rare, so we analysed only leaf litter decomposition (for the vascular species). From most studies we were able to extract the following information: latitude and longitude, mean annual temperature (MAT), mean annual precipitation (MAP), the dominant vegetation of the study, and the species studied. When climate data were not included or readily obtained from citations, we estimated MAT from nearby studies. Topographic position (i.e. upland or lowland) was determined from descriptions of the vegetation type. Bogs, forested bogs, mires, fens, laggs, and wet meadow tundra were considered lowlands. Forests, and heath, tussock, and shrub tundras were considered uplands. Insufficient information was available to include analyses of litter chemistry.

Our results are consistent with laboratory and field studies and suggest that temperature, topography (i.e. the degree of inundation), and vegetation composition are the primary controls over litter decomposition at the regional scale. Decomposition rate was positively affected by MAT but MAP explained less than 15% of the variation in decomposition rate (Fig. 1, F_{1,111} = 13.07, P < 0.001; R^2 = 0.11; % mass loss = 23.01 + 1.02 × MAT). In contrast, decomposition rate was not significantly related to MAP (F_{1,90} = 1.12, P = 0.29, R^2 = 0.01). So, despite relatively low precipitation rates, cold temperatures are the primary climatic factor limiting decomposition at high latitudes.

Whether decomposition occurred in upland or lowland ecosystems also significantly affected rates of litter mass loss. Litter in upland ecosystems decomposed more rapidly than litter in ecosystems characterized by poor drainage (Fig. 1a). In an analysis of covariance (ANCOVA) with topographic position as a categorical variable and MAT as a covariate, both variables were significant (F_{1,107} = 13.26, P < 0.001 and F_{1,107} = 19.96, P < 0.001, respectively; R^2 = 0.21; slopes were homogeneous). Slow decomposition in poorly drained areas likely results from low oxygen availability in saturated soils with little water flow.

Substrate quality was just as important as temperature in explaining variation in decomposition rates in arctic and boreal regions. To determine the importance of vegetation composition relative to abiotic factors, we grouped species into the dominant growth forms found in high-latitude ecosystems [mosses, evergreens, deciduous species, and herbs (graminoids and forbs); insufficient data existed to include lichens], because species within these growth forms have previously been shown to decompose similarly (Hobbie 1996). Together, growth form and MAT explained greater than 50% of the variation in decomposition rate (Fig. 1b). In an ANCOVA including growth form as a categorical variable and MAT as a covariate, both variables were significant (F_{3,102} = 31.09, P < 0.001 and F_{1,102} = 52.38, P < 0.001, respectively; R^2 = 0.54; slopes were homogeneous). Overall, litter of deciduous and herbaceous species decomposed more quickly than other litter types (Tukey’s HSD, P < 0.001) but had similar rates of decomposition to each other (Tukey’s HSD, P = 0.49). Evergreen litter decomposed significantly more slowly than that of deciduous and herbaceous species (Tukey’s HSD, P < 0.001), and moss litter decomposed even more slowly than that of evergreens (Tukey’s HSD, P = 0.02).

Decomposition rates of the vascular growth forms probably reflect differences in both carbon and nutrient chemistry, because evergreen species tend to have both higher lignin and lower nutrient concentrations than do deciduous species (Coley et al. 1985). Despite the strong association of moss with poorly drained ecosystems (80% of the cases of moss decomposition studied here were in lowland ecosystems), their slow decomposition appears to be a function of their poor substrate quality. When we narrowed our analysis to lowland ecosystems and compared moss decomposition with that of deciduous and herbaceous litter, we still found that mosses decompose significantly more slowly (Fig 1c). In an ANCOVA that compared just herbaceous and deciduous litter with that of moss litter decomposing within lowland ecosystems and included MAT as a covariate, both growth form and MAT were significant (F_{1,27} = 34.60, P < 0.001 and F_{1,27} = 16.00, P < 0.001, respectively; R^2 = 0.58; slopes were homogeneous). The reason why mosses decompose so slowly is elusive but widely observed (Clymo & Hayward 1982; Hobbie 1996). Slow decomposition of Sphagnum spp. may result from their low nutrient concentrations (Coulson & Butterfield 1978) or from recalcitrant carbon compounds (Karunen & Ekman 1982; Chapin et al. 1986; Johnson & Damman 1991). In addition, Sphagnum spp. are known to produce compounds with antimicrobial properties (Painter 1991; Verhoeven & Toth 1995).
Differences in decomposition rates among growth forms were large relative to the effect of temperature. Litter of deciduous species decomposed twice as quickly as moss litter. Temperature would have to increase by approximately 5°C to cause a comparable doubling of decomposition rate. Thus, understanding variation in carbon storage across landscapes or predicting how growth form abundance will change, respectively. Although for the vascular plants these growth-form differences in decomposition might be summarized accurately by some index of litter quality (e.g. lignin concentration), slow moss decomposition is not predicted by its carbon chemistry (Hobbie 1996). Current biogeochemical models that use litter quality indices appropriate for vascular plants to predict decomposition rates may overestimate rates of moss-derived peat decomposition in response to climate warming (e.g. Schimel et al. 1994), perhaps contributing to the discrepancy among estimates in high-latitude carbon sinks discussed earlier.

Recent reviews suggest that the dominant growth forms in arctic ecosystems make logical functional groups in which to lump species with respect to their influence on biogeochemical processes (Chapin et al. 1995, 1997). Our analysis provides evidence that across arctic and boreal regions, growth forms are distinct in their effect on decomposition and may provide a useful way of simplifying species for inclusion in ecosystem models. Boreal and arctic ecosystems differ from other regions because growth forms with such contrasting effects on biogeochemical processes, such as mosses and deciduous species, can dominate adjacent landscapes. Also, different growth forms can dominate the same landscape during different stages of succession (e.g. postfire or postflood). Notably, we did not consider decomposition of woody litter in our analysis. Depending on allocation, the slow decomposition of woody litter may reduce the overall decomposition rates of litter from deciduous and evergreen shrubs and trees (Hobbie 1996).

Accumulation, storage and decomposition of soil organic matter

Most of the carbon stored in northern soils has decomposition rates that are slower than those found in fresh litter (k<0.1 y⁻¹) as revealed by radiocarbon data (e.g. Clymo 1984; Trumbore & Harden 1997; Clymo et al. 1998) and chronosequence studies (Harden et al. 1992). Slow decomposition rates of soil organic matter result from several factors. First, recalcitrant compounds may dominate soils because of selective preservation of components that decompose slowly, like mosses, as decomposers break down more labile material. Secondly, recalcitrant compounds may form because of microbial processing (humification) or by fire (charcoal). Thirdly, persistent cold soil temperatures associated with permafrost, as well as anaerobic conditions, slow in situ decomposition rates. For example, the largest carbon inventories exist in poorly drained soils and wetlands (Ping et al. 1997), where high moss abundance, low oxygen, and cold temperatures combine to retard decay. In boreal uplands, large stores of decomposed and

Fig. 1 (a) Litter mass loss after one year vs. MAT in upland and lowland ecosystems. (b) Litter mass loss after one year vs. MAT for different dominant growth forms. Lines represent regression lines obtained for individual growth forms (g, graminoid and forb; e, evergreen; d, deciduous; m, moss). (c) Litter mass loss after one year vs. MAT in lowland ecosystems only.
charred material are found below the litter layer and above the mineral soil (Harden et al. 1997) and decomposition rates in this humic layer are 10-fold slower than those of the overlying litter (Trumbore & Harden 1997). In areas dominated by permafrost, cryoturbation (mixing of soils by freeze-thaw processes) may be an additional important process by which organic matter from surface horizons is stabilized in permanently frozen horizons, making it inaccessible to microbes. Where carbon stocks in permafrost have been measured, they are large and can double estimates of soil carbon based on carbon measured in the active layer that thaws during the growing season (Michaelson et al. 1996). The long-term storage of this nonlitter carbon contrasts with lower-latitude systems where interaction with mineral surfaces (approximated by soil texture) is a dominant control of the storage and dynamics of carbon on millennial time scales. Indeed some biogeochemical models assume that soil organic matter decomposition is a function of soil texture (e.g. CENTURY, Parton et al. 1987). However, soil organic matter decomposition may be unrelated to soil texture in ecosystems where much of the decomposition occurs in an organic horizon with little mineral content.

In wetland ecosystems, stores of carbon in deep, humified layers are so large that even at very slow rates of decomposition, the flux of CO₂ derived from these layers contributes significantly to annual soil respiration (Tolonen & Turunen 1996; Clymo et al. 1998). This may also be true in moderately to poorly drained upland soils in boreal regions (Trumbore & Harden 1997). In a black spruce/moss stand in central Manitoba, an overall increase in soil respiration in the late summer/early fall was linked to the timing of thawing and warming of deep humified organic matter (Goulden et al. 1998). Radiocarbon measurements in soil CO₂ confirmed that much of the CO₂ respired in fall and winter months was derived from decomposition of organic matter that was between 50 and several hundred years old (Winston et al. 1997). Hence the lag time between carbon fixation and decomposition for a significant portion of the annual net carbon flux may be quite long, which may increase estimates of isotope disequilibrium in boreal regions. Year-to-year differences in soil respiration may be caused by climate variability as it impacts the timing and degree of deep soil thaw in upland soils (Goulden et al. 1998).

Incubation of these organic materials shows that decomposition rates may increase dramatically if the soils are dried or warmed (Billings et al. 1982; Moore & Knowles 1989; Goulden et al. 1998). If high-latitude regions dry or thaw as a result of future climate change, net decomposition of the vast stores of deep carbon could result in a large positive feedback to rising atmospheric CO₂. Alternatively, if nutrients associated with mineralized carbon stimulate plant production, high-latitudes could prove sinks for carbon (Shaver et al. 1992). Indeed, models used to predict future carbon balance with climate warming that consider thawing of carbon stored in permafrost are extremely sensitive to assumptions about the amount of carbon stored in permafrost and its carbon to nutrient ratio (Waëlbroeck et al. 1997). Yet, little is known about the characteristics of frozen carbon pools for large regions. A better understanding of the importance of limitation by substrate quality vs. limitation by abiotic conditions (permafrost or waterlogging) is required to predict the storage of carbon in soil organic matter for high-latitude regions and its response to changes in temperature and precipitation.

**Winter decomposition**

A unique aspect of arctic and boreal systems is their long, cold winter. Microbially decomposition processes during this period have long been assumed to be insignificant in the annual cycling of carbon and nutrients (e.g. Steudler et al. 1989). Yet, recent data indicate that litter decomposition and soil respiration occur over winter in both arctic and boreal ecosystems (Sommerfeld et al. 1993; Zimov et al. 1993; Clein & Schimel 1995; Wagener 1995; Brooks et al. 1996; Hobbie & Chapin 1996; Zimov et al. 1996; Oechel et al. 1997; Winston et al. 1997; Fahnestock et al. 1998; Fahnestock et al. 1999; Grogan & Chapin 1999). In fact, winter fluxes of CO₂ are substantial in annual carbon budgets (Table 1) and likely influence both the magnitude and direction of annual carbon fluxes.

Winter activity accounts for roughly 20% of annual soil respiration, although estimates range between 3 and 50% across different arctic and alpine tundra and boreal forest communities (Table 1). Many of the methods used to estimate winter fluxes are problematic. For example, atmospheric CO₂ may be concentrated in freezing snow and transported to the base of the snowpack (Williams 1949), resulting in overestimates of net efflux based on CO₂ concentration profiles in snow. Nevertheless, all of the methods used to date indicate net fluxes of CO₂ during the winter, although the magnitude of these fluxes is uncertain. Winter CO₂ fluxes vary both spatially and temporally. They are highest and most variable during the autumn–winter and winter–spring transitional periods as soils are subjected to repeated freeze–thaw events, but can continue through winter (Jones et al. 1997; Oechel et al. 1997; Fahnestock et al. 1998; Fahnestock et al. 1999; Grogan & Chapin 1999).

Our understanding of the processes involved in winter CO₂ efflux remains poor. We know little about the microbial processes controlling winter decomposition and CO₂ fluxes: do they operate similarly, but more slowly, to those in the summer, or do they behave quite differently? For example, CO₂ during the summer comes

© 2000 Blackwell Science Ltd, *Global Change Biology*, 6 (Suppl. 1), 196–210
primarily from root respiration and decay of plant detritus in the surface organic horizon (J.P. Schimel, unpubl. data, Grogan & Chapin 1999), but during the winter it appears that microbial respiration may be driven by soluble material remaining in water films or deeper in the profile or into the mineral soils (Clein & Schimel 1995). If this is the case, then models of carbon dynamics developed for summer activity would be fundamentally flawed in modelling winter activity.

Litter mass and N loss appear to occur primarily during winter in arctic systems (Hobbie & Chapin 1996), though it was unclear from that work whether the winter mass and N loss were biological or associated with spring leaching. Wagener (1995), however, showed that litter decomposition occurred in the taiga before snowmelt, suggesting that overwinter decomposition may also result from microbial activity. Several laboratory studies have shown that significant microbial activity can occur in soils at temperatures in the range of −5 to −7°C (Flanagan & Bunnell 1980; Coxson & Parkinson 1987; Clein & Schimel 1995). Soil temperatures in this range are common in the tundra during early and late winter, and throughout the winter in areas where snow insulates the soil.

The controls on this activity in frozen soils, however, are complex. Temperature responses vary in different tundra and taiga soils, with Q10 responses ranging from 1 to 9.8 over the range from −5°C to −2°C (Clein & Schimel 1995). The temperature below which activity ceases also varies among soils. Brooks et al. (1996) reported activity down to −5°C for two alpine sites, while Mosier et al. (1991) found that respiration only continued in a short-grass steppe down to about −2°C. In a treeline site no CO2 production occurred when the soil was below 0°C (Brooks et al. 1996). This critical temperature can even vary between nearby sites: the alpine and treeline sites described by Brooks et al. (1996) were within 100 m of each other.

Decomposition during the winter appears to be controlled by both physical and biological factors. Physical factors include the thermal regime of the soil and the quantity of unfrozen water it contains. How large a thawed layer exists in the soil and how long into the winter it exists is critical in controlling winter decomposition activity (Zimov et al. 1993). Perhaps equally important is the amount of unfrozen water that remains in ‘frozen’ soil, as substantial microbial activity can occur in films of liquid water that remain on soil particles even after the bulk of the water has frozen. For example, Clein & Schimel (1995) showed significantly less respiration in a white spruce taiga soil when that soil was incubated frozen at 10% vs. 50% of water holding capacity. The amount of unfrozen water that remains in soil or litter at a particular temperature depends on soil composition (particle size distribution) and chemistry. There are several existing models for predicting the amount of unfrozen water in soil (Lovell 1957; Nakano & Brown 1971), but it is not yet clear which is most appropriate for modelling microbial activity in frozen soils, particularly organic soils. This area needs further study. It is also likely that the physical and biological factors interrelate with each other, as microbes may produce enough heat to affect the soil thermal regime (Zimov et al. 1996).

Because different soils appear to have variable responses to increased temperature, these soils must differ in other factors that control microbial activity during the winter. For example, the nature of the available substrates is probably an important control on winter activity and none of the few studies at subzero temperatures evaluated microbial substrate use. During the summer, respiration in arctic and boreal regions is

| Vegetation Type           | Magnitude of wintertime fluxes                  | Method                   | Reference                  |
|---------------------------|------------------------------------------------|--------------------------|----------------------------|
| Tussock tundra            | 61% of net annual CO2 exchange                 | Flux chambers            | Oechel et al. 1997        |
| Wet sedge                 | 81% of net annual CO2 exchange                 | "                        | "                         |
| Forest tundra             | 41% of net annual CO2 exchange                 | Snow CO2 profiles        | Zimov et al. 1996         |
| Mixed types               | 18% of total annual soil respiration           | "                        | Zimov et al. 1996; Fahnstock et al. 1998 |
| Shrub                     | 7% of total annual soil respiration            | Snow CO2 profiles        | "                         |
| Tussock                   | 21% of total annual soil respiration           | "                        | "                         |
| Wet meadow                | 17% of total annual soil respiration           | "                        | "                         |
| Various types             | >50% of total annual soil respiration          | Soda lime traps          | Grogan & Chapin 1999      |
| Tussocks                  | 34% of total annual soil respiration           | Extrapolation of lab data| Clein & Schimel 1995      |
| Intertussock              | 12% of total annual soil respiration           | "                        | "                         |
| Wet meadow                | 21% of total annual soil respiration           | "                        | "                         |
| Black spruce/ jack pine forests | 3−17% of total annual soil respiration | Flux chambers             | Winston et al. 1997; Savage et al. 1997 |
almost certainly dominated by activity in the surface organic horizon, because the bulk of the decomposable litter and organic matter is found there. However, during winter, it is quite possible that much of the decomposition activity occurs deeper in the profile in warmer (and possibly unfrozen) horizons. In areas where there are not deep peat accumulations, activity in the mineral soil may become important. Dissolved organic carbon (DOC) fractions may also be more important than nondissolved ones in supplying carbon to microbes even within the organic horizons (Clein & Schimel 1995). In tussock tundra the $^{13}$C of respired CO$_2$ varied by up to 5% between summer and winter (Welker pers. comm.), indicating changes in substrate use. Dramatic seasonal changes in the radiocarbon in soil CO$_2$ in boreal forest soils demonstrate a significant shift in substrate age, with older carbon respired in the fall and winter (Winston et al. 1997). Thus, it seems likely that there are major shifts in the zone of microbial activity and of substrates between summer and winter. Models of annual activity must account for these shifts to accurately model winter activity.

Basic thermodynamics also supports the idea that substrate use may change as temperature drops. The fundamental control on the temperature response of biochemical reactions is the activation energy of the reaction, as described by the Arrhenius equation. The higher the activation energy, the higher the temperature sensitivity, and the comparatively slower the rates that would occur at low temperature. Metabolizing complex, aromatic structures should have a higher activation energy and temperature sensitivity than metabolizing simpler, carbohydrate-based structures. Equally important, temperature can affect the activation energy of enzymatic reactions by causing changes in enzyme conformation. Linkins et al. (1984) noted a dramatic increase in the activation energy of endocellulase in a tundra soil below 5°C, which would greatly reduce cellulose use at low temperature.

**Landscape controls over soil carbon storage**

A number of factors in arctic and boreal regions interact to control soil carbon storage at the landscape scale. Fires are uncommon in tundra (Wein 1976), but in boreal regions, the timing and spatial distribution of fires (as well as the distribution of wetland areas that do not burn) control whether landscapes are sources or sinks for carbon. Fire is an especially important control over carbon storage in soils of upland boreal forests. These forest stands are carbon sources to the atmosphere during and in the years immediately following fire, and carbon sinks as trees and mosses regrow (e.g. Kasischke et al. 1995; Harden et al. 1997; Rapalee et al. 1998). Approximately equal amounts of carbon are lost to burning in fires and during decomposition in the years immediately following fires, when less shade and reduced insulating ground cover combine to increase soil temperatures (Kasischke et al. 1995). Between burning events, decomposition rates of mosses are slow enough that large stores of carbon (several kg C m$^{-2}$) accumulate in moss and detrital layers. For ecosystems where fire is the principal mode of disturbance, the loss of carbon during burning must be considered along with decomposition rates as a control on the amount of accumulated litter material.

Topographic position is an important factor controlling carbon storage in both arctic and boreal landscapes because it influences the drainage and formation of wetland areas. In tundra, soil carbon stocks are greatest in low-lying poorly drained wet meadow tundra and in tussock tundra that forms on stable, gentle slopes with relatively shallow thaw (Giblin et al. 1991; Michaelson et al. 1996). Wetlands can be significant sinks of carbon, accumulating on average about 30 g C m$^{-2}$ y$^{-1}$ (Gorham 1991). Small landscape elements, such as beaver ponds (Roulet et al. 1997), or local areas of permafrost collapse, may act as large local sources of carbon to the atmosphere that offset carbon gains in the remainder of the landscape. Drainage of northern wetlands results in a short-term release of carbon, as water tables drop and decomposable substrates are exposed to oxygen (Billings et al. 1982; Moore & Dalva 1993; Schurpali et al. 1996; Waddington & Roulet 1996).

**Conclusions**

Despite the importance of arctic and boreal systems in the present carbon cycle, controls over carbon storage in northern soils are not well-understood. Biogeochemical models used to predict the response of ecosystems to changing climate may be inadequate for use at high latitudes, where a number of unique factors strongly influence soil carbon turnover:

1. Differences in litter decomposition between growth forms are large relative to the effects of abiotic factors such as temperature. Of particular importance is the slow decomposition rate of moss litter, which is not predicted by commonly used indices of litter quality.

2. Cold temperature, anaerobiosis, and permafrost interact with substrate quality to determine the degree of organic matter stabilization. The relative importance of these factors, and how they will respond to climate change, is unknown.

3. Recent evidence suggests that biological activity occurring in frozen soils can contribute significantly to annual soil carbon fluxes. Yet the controls over this activity remain poorly understood.
Landscape-level processes, such as fire and inundation, can be just as important as processes within landscape units in controlling regional carbon fluxes.

By explicitly considering these unique aspects of soil carbon turnover in high-latitude systems, we may be able to refine our understanding of present carbon fluxes and better predict the influence of these systems on the future climate.

Acknowledgements

This work resulted from a workshop (Arctic Boreal Processes that Feed Back to Climate: Extrapolation and Synthesis) conducted at the National Center for Ecological Analysis and Synthesis, a Center funded by NSF (Grant #DEB-94-21535), the University of California–Santa Barbara, the California Resources Agency, and the California Environmental Protection Agency.

References

Berg B, Ekbohm G (1991) Litter mass-loss rates and decomposition patterns in some needle and leaf litter types. Long-term decomposition in a Scots pine forest. VII. Canadian Journal of Botany, 69, 1449–1456.
Berg B, McLaughey C, Johansson M-B (1993) Litter mass-loss rates in late stages of decomposition at some climatically and nutritionally different pine sites. Long-term decomposition in a Scots pine forest. VIII. Canadian Journal of Botany, 71, 680–692.
Billings WD, Luken JO, Mortensen DA, Peterson KM (1982) Arctic tundra: a source or sink for atmospheric carbon dioxide in a changing environment? Oecologia, 53, 7–11.
Brooks PD, Williams MW, Schmidt SK (1996) Microbial activity under alpine snowpacks, Niwot Ridge, Colorado. Biogeochemistry, 32, 93–113.
Chapin FS III, Matthews E (1992) Boreal carbon pools: approaches and constraints in global extrapolations. In: Global Change and Carbon Cycling in Northern Ecosystems (eds Vinson T, Kolchugina T). EPA, Corvallis.
Chapin FS III, McKendrick JD, Johnson DA (1986) Seasonal changes in carbon fractions in Alaskan tundra plants of differing growth form: Implications for herbivores. Journal of Ecology, 74, 707–731.
Chapin FS III, Hobbie SE, Bret-Harte MS, Bonan G (1995) Causes and consequences of plant functional diversity in arctic ecosystems. In: Arctic and Alpine Biodiversity: Patterns, Causes and Ecosystem Consequences (eds Chapin FS III, Körner C), pp. 225–237. Springer, Berlin.
Chapin FS III, McFadden JP, Hobbie SE (1997) The role of arctic vegetation in ecosystem and global processes. In: Ecology of Arctic Environments (eds Woodin SJ, Marquiss M), pp. 121–136. Blackwell Science, Oxford.
Clais P, Tans PP, Trolier M, White JMC, Francey RJ (1995) A large Northern hemisphere terrestrial CO2 sink indicated by the $^{13}{\text{C}}/{^{12}{\text{C}}}$ ratio of atmospheric CO2. Science, 269, 1098–1102.
Clymo RS (1984) The limits to peat growth. Transactions of the Royal Society of London, B303, 605–645.
Clymo RS, Haywood PM (1982) The ecology of Sphagnum. In: Bryophyte Ecology (ed. Smith AJE), pp. 229–289. Chapman & Hall, London.
Clymo RS, Turunen J, Tolonen K (1998) Carbon accumulation in peatland. Oikos, 81, 368–388.
Coley PD, Bryant JP, Chapin FS III (1985) Resource availability and plant anti-herbivore defense. Science, 230, 895–899.
Coulson JC, Butterfield J (1978) An investigation of the biotic factors determining the rates of plant decomposition on blanket bog. Journal of Ecology, 66, 631–650.
Coxson DS, Parkinson D (1987) Winter respiratory activity in Aspen woodland forest floor litter and soils. Soil Biology and Biochemistry, 19, 49–59.
Dixon RK, Brown S, Houghton RA, Solomon AM, Trexler MC, Wisniewski J (1994) Carbon pools and flux of global forest ecosystems. Science, 263, 185–190.
Fahnestock JT, Jones MH, Brooks PD, Walker DA, Welker JM (1998) Winter and early spring CO2 efflux from tundra communities of northern Alaska. Journal of Geophysical Research Atmospheres, 103, 29,023–29,027.
Fahnestock JT, Jones MH, Welker JM (1999) Wintertime CO2 efflux from arctic soils: implications for annual carbon budgets. Global Biogeochemical Cycles, 13, 775–779.
Finer L, Messier C, De Grandpre L (1997) Fine-root dynamics in mixed boreal conifer-broad-leafed forest stands at different successional stages after fire. Canadian Journal of Forest Research, 27, 304–314.
Flanagan PW, Veum AK (1974) Relationships between respiration, weight loss, temperature, and moisture in organic residues on tundra. In: Soil Organisms and Decomposition in Tundra (eds Holding AJ et al.), pp. 249–277. Tundra Biome Steering Committee, Stockholm.
Flanagan PW, Bunnell FL (1980) Microflora activities and decomposition. In: An Arctic Ecosystem: the Coastal Tundra at Barrow, Alaska (eds Brown J et al.), pp. 291–334. Dowden, Hutchinson and Ross, Stroudsburg, PA.
Flanagan PW, Van Cleeve K (1983) Nutrient cycling in relation to decomposition and organic matter quality in taiga ecosystems. Canadian Journal of Forest Research, 13, 795–817.
Francey RJ, Tans PP, Allison CE, Enting IG, White JWC, Trolier M (1995) Changes in oceanic and terrestrial carbon uptake since 1982. Nature, 373, 326–333.
French DD (1974) Classification of IBP Tundra Biome sites based on climate and soil properties. In: Soil Organisms and Decomposition in Tundra (eds Holding AJ et al.), pp. 3–25. Tundra Biome Steering Committee, Stockholm.
Fung IY, Berry JA, Field CB et al. (1997) Carbon 13 exchanges between the atmosphere and biosphere. Global Biogeochemical Cycles, 11, 507–533.
Giblin AE, Nadelhoffer KJ, Shaver GR, Laundre JA, McKerrow AJ (1991) Biogeochemical diversity along a riverside toposequence in arctic Alaska. Ecological Monographs, 61, 415–435.
Goehm E (1991) Northern peatlands: role in the carbon cycle and probable responses to climatic warming. Ecological Applications, 1, 182–195.
Goulden ML, Wofsy SC, Harden JW et al. (1998) Sensitivity of boreal forest carbon balance to soil thaw. Science, 279, 214–217.
Grigal DF, McColl JG (1977) Litter decomposition following
forest fire in northeastern Minnesota. *Journal of Applied Ecology*, **14**, 531–538.

Grogan P, Chapin FS III (1999) Arctic soil respiration: effects of climate and vegetation depend on season. *Ecosystems*, **2**, 451–459.

Harden JW, Sundquist ET, Stallard RF, Mark RK (1992) Dynamics of soil carbon during deglaciation of the Laurentide ice sheet. *Science*, **258**, 1921–1926.

Harden J, O’Neill KP, Trumbore SE, Veldhuis H, Stocks BJ (1997) Moss and soil contributions to the annual net carbon flux in a maturing boreal forest. *Journal of Geophysical Research Atmospheres*, **102**, 817–828.

Heal OW, French DD (1974) Decomposition of organic matter in tundra. In: *Soil Organisms and Decomposition in Tundra* (eds Holding AJ et al.), pp. 279–310. Tundra Bione Steering Committee, Stockholm.

Heal OW, Flanagan PW, French DD, Maclean SF Jr (1981) Decomposition and accumulation of organic matter in tundra. In: *Tundra Ecosystems: a Comparative Analysis* (eds Bliss LC et al.), pp. 387–633. Cambridge University Press, Cambridge.

Hobbie SE (1996) Temperature and plant species control over litter decomposition in Alaskan tundra. *Ecological Monographs*, **66**, 503–522.

Hobbie SE, Chapin FS III (1996) Winter regulation of tundra litter carbon and nitrogen dynamics. *Biogeochemistry*, **35**, 327–522.

Holland EA, Braswell BH, Lambarque J-F et al. (1997) The spatial distribution of atmospheric nitrogen deposition and its impact on carbon uptake by terrestrial ecosystems. *Journal of Geophysical Research*, **102**, 15,849–15,866.

Houghton RA (1994) The worldwide extent of land-use change. *Bioscience*, **44**, 305–313.

Houghton RA (1996) Terrestrial sources and sinks of carbon inferred from terrestrial data. *Tellus*, **48B**, 420–432.

Huang WZ, Schoenau JJ (1997) Mass loss measurements and statistical models to predict decomposition of leaf litter in a boreal aspen forest. *Communications in Soil Science and Plant Analysis*, **28**, 863–874.

Johansson M-J, Berg B, Meentemeyer V (1995) Litter mass-loss rates in late stages of decomposition in a climatic transect of pine forests. Long-term decomposition in a Scots pine forest. IX. *Canadian Journal of Botany*, **73**, 1509–1521.

Johnson LC, Damman AWH (1991) Species controlled *Sphagnum* decay on a South Swedish raised bog. *Oikos*, **61**, 234–242.

Jones MH, Fahnstock JT, Welker JM (1997) Winter CO2 efflux in the Kuparuk River watershed, Alaska. *Bulletin of the Ecological Society of America*, **78**, 265.

Karunen P, Ekman R (1982) Age-dependent content of poly-merized lipids in *Sphagnum fuscum*. *Physiologia Plantarum*, **54**, 162–166.

Kasischke ES, Christensen NL Jr, Stocks BJ (1995) Fire, global warming, and the carbon balance of boreal forests. *Ecological Applications*, **5**, 437–451.

Kasischke ES, Bergen K, Fininnmore R et al. (1999) Satellite imagery gives clear picture of Russia’s boreal forest fires. *EOS, Transactions of the American Geophysical Union*, **80**, 141–147.

Kauppi PE, Mielikäinen K, Kuusela K (1992) Biomass and carbon budget of European forests. 1971–90. *Science*, **256**, 70–74.

Keeling CD, Bacastrow RB, Carter AF et al. (1989) A three-dimensional model of atmospheric CO2 transport based on observed winds: 1. Analysis of observational data. In: *Aspects of Climate Variability in the Pacific and the Western Americas* (ed. Peterson DH), pp. 165–236. American Geophysical Union, Washington, D. C.

Kohlmair GH, Hager C, Wurth G et al. (1995) Effects of age class distribution of the temperate and boreal forests on the global CO2 source-sink function. *Tellus*, **47B**, 212–231.

Kurz WA, Apps MJ (1994) The carbon budget of Canadian forest: a sensitivity analysis of changes in disturbance regimes, growth rates, and decomposition rates. *Environmental Pollution*, **83**, 55–61.

Lashof DA (1989) The dynamic greenhouse: feedback processes that may influence future concentrations of atmospheric trace gases and climatic change. *Climate Change*, **14**, 213–242.

Linkins AE, Melillo JM, Sinsabaugh RL (1984) Factors affecting cellulase activity in terrestrial and aquatic ecosystems. In: *Current Perspectives in Microbial Ecology* (eds Klug MJ, Reddy CA), pp. 572–579. American Society for Microbiology, Washington, D. C.

Lloyd J, Farquhar GD (1994) 13C discrimination during CO2 assimilation by the terrestrial biosphere. *Ecology*, **99**, 201–215.

Lovell CIW (1957) Temperature effects on phase composition and strength of partially-frozen soil. *Highway Research Board Bulletin*, **168**, 74–95.

McGuire AD, Clein JS, Melillo JM et al. (2000a) Modeling carbon responses of tundra ecosystems to historical and projected climate: sensitivity of pan-Arctic carbon storage to temporal and spatial variation in climate. *Global Change Biology*, **6** (Suppl. 1), 141–159.

McGuire AD, Melillo JM, Randerson JT et al. (2000b) Modeling the effects of snowpack on heterotrophic respiration across northern temperate and high latitude regions: comparison with measurements of atmospheric carbon dioxide in high latitudes. *Biogeochemistry*, in press.

Melillo JM, Prentice IC, Farquhar GD, Schultz E-D, Sala OE (1996) Terrestrial biotic responses to environmental change and feedbacks to climate. In: *Climate Change 1995 the Science of Climate Change Contribution of Working Group 1 to the Second Assessment Report of the Intergovernmental Panel on Climate Change* (eds Bolin B et al.), pp. 445–481. Cambridge University Press, Cambridge.

Michaelson GJ, Ping CL, Kimber JM (1996) Carbon storage and distribution in tundra soils of arctic Alaska, U.S.A. *Arctic and Alpine Research*, **28**, 414–424.

Moore TR (1984) Litter decomposition in a subarctic spruce-lichen woodland, Eastern Canada. *Ecology*, **65**, 299–308.

Moore TR, Knowles R (1989) The influence of water table levels on methane and carbon dioxide emissions from peatland soils. *Revue Canadienne de la Science Du Sol*, **69**, 33–38.

Moore TR, Dalva M (1993) Influence of temperature and water table position on carbon dioxide and methane emissions from columns of peatland soils. *Journal of Soil Science*, **44**, 651–664.

Mosier A, Schimel D, Valentine D, Bronson K, Parton W (1991) Methane and nitrous oxide fluxes in native, fertilized and cultivated grasslands. *Nature*, **350**, 330–332.

Nakano Y, Brown J (1971) Effect of a freezing zone of finite width on the thermal regime of soils. *Water Resources Research*, **7**, 1226–1233.

Nakatsubo T, Uchida M, Horikoshi T, Nakane K (1997) Comparative study of themass loss rate of moss litter in
boreal and subalpine forests in relation to temperature. *Ecological Research*, **12**, 47–54.

Oechel WC (1989) Nutrient and water flux in a small arctic watershed: an overview. *Holartic Ecology*, **12**, 229–237.

Oechel WC, Hastings SJ, Vourlitis GL, Jenkins M, Riechers G, Gruulke N (1993) Recent change of Arctic tundra ecosystems from a net carbon dioxide sink to a source. *Nature*, **361**, 520–523.

Oechel WC, Vourlitis GL, Hastings SJ (1997) Cold season CO2 emission from arctic soils. *Global Biogeochemical Cycles*, **11**, 163–172.

Oechel WC, Vourlitis GL, Verfaillie Jr J et al. (2000) A scaling approach for quantifying the net CO2 flux of the Kuparuk River Basin, Alaska. *Global Change Biology*, **6** (Suppl. 1), 160–173.

Painter TG (1991) Lindow Man, Tollund Man, and other peat-bog bodies: the preservative and antimicrobial action of sphagnum, a reactive glycuronoglycan with tanning and sequestering properties. *Carbohydrate Polymers*, **15**, 123–142.

Parton WJ, Schimel DS, Cole CV, Ojima DS (1987) Analysis of factors controlling soil organic matter levels in Great Plains grasslands. *Soil Science Society of America Journal*, **51**, 1173–1179.

Ping CL, Michaelson J, Kimble JM (1997) Carbon storage along a latitudinal transect in Alaska. *Nutrient Cycling in Agroecosystems*, **49**, 235–242.

Post WM (1990) Report of a Workshop on Climate Feedbacks and the Role of Peatlands, Tundra, and Boreal Ecosystems in the Global Carbon Cycle. Oak Ridge National Laboratory, Oak Ridge, TN, USA.

Post WM, Emanuel WR, Zinke PJ, Stangenberger AG (1982) Soil carbon pools and world life zones. *Nature*, **298**, 156–159.

Post WM, Pastor J, Zinke PJ, Stangenberger AG (1985) Global patterns of soil nitrogen storage. *Nature*, **317**, 613–616.

Prescott CE, Parkinson D (1985) Effects of sulphur pollution on rates of litter decomposition in a pine forest. *Canadian Journal of Botany*, **63**, 1436–1443.

Rapalae G, Trombore SE, Davidson EA, Harden JW, Veldius H (1998) Scanning soil C pools and fluxes in a boreal forest landscape. *Global Biogeochemical Cycles*, **12**, 687–701.

Reader RJ, Stewart JM (1972) The relationship between net primary production and accumulation for a peatland in southeastern Manitoba. *Ecology*, **53**, 1024–1037.

Robinson CH, Wookey PA, Parsons AN, Potter JA, Callaghan TV, Lee J (1995) Responses of plant litter decomposition and nitrogen mineralisation to simulated environmental change in a high arctic polar semi-desert and a subarctic dwarf shrub heath. *Oikos*, **74**, 503–512.

Rochefort L, Vitt DH, Bayley SE (1990) Growth, production, and decomposition dynamics of *Sphagnum* under natural and experimentally acidified conditions. *Ecology*, **71**, 1986–2000.

Rosswall T (1974) Cellulose decomposition studies on the tundra. In: *Soil Organisms and Decomposition in Tundra* (eds Holding AJ et al.). Tundra Biome Steering Committee, Stockholm.

Roulet NT, Crill PM, Comer NT, Dove A, Zepp RG (1997) CO2 and CH4 flux between a boreal beaver pond and the atmosphere. *Journal of Geophysical Research Atmospheres*, **102**, 29,313–29319.

Savage K, Moore TR, Crill PM (1997) Methane and carbon dioxide exchanges between the atmosphere and northern boreal forest soils. *Journal of Geophysical Research Atmospheres*, **102**, 29,279–29,288.

Schimel DS, Braswell BH, Holland EA et al. (1994) Climatic, edaphic, and biotic controls over storage and turnover of carbon in soils. *Global Biogeochemical Cycles*, **8**, 279–293.

Schimel DS, Enting IG, Heimann M et al. (1995) CO2 and the carbon cycle. In: *Climate Change 1994: Radiative Forcing of Climate Change and an Evaluation of IPCC IS92 Emission Scenarios* (eds Houghton JT et al.), pp. 35–71. Cambridge University Press, Cambridge, UK.

Schlesinger WH (1977) Carbon balance in terrestrial detritus. *Annual Review of Ecology and Systematics*, **8**, 51–81.

Schurpali NJ, Verma SC, Kim J (1996) Carbon dioxide exchange in a peatland ecosystem. *Journal of Geophysical Research Atmospheres*, **100**, 14,319–14,326.

Shaver GR, Billings WD, Chapin FS III et al. (1992) Global change and the carbon balance of arctic ecosystems. *Bioscience*, **42**, 433–441.

Shaver GR, Giblin AE, Nadelhofer KJ, Rastetter EB (1997) Plant functional types and ecosystem change in arctic tundras. In: *Plant Functional Types: Their Relevance to Ecosystem Properties and Global Change* (eds Smith TM et al.), pp. 153–173. Cambridge University Press, Cambridge.

Sommerfeld RA, Mosier AR, Musselman RC (1993) Carbon dioxide, methane, and nitrous oxide flux through a Wyoming snowpack and implications for global budgets. *Nature*, **361**, 140–142.

Steedler PA, Bowden RD, Mellilo JM, Aber JD (1989) Influence of nitrogen fertilization on methane uptake in temperate forest soils. *Nature*, **341**, 314–315.

Tans PP, Berry JA, Keeling RF (1993) Oceanic 13C/12C observations: a new window on ocean CO2 uptake. *Global Biogeochemical Cycles*, **7**, 353–368.

Thompson MV, Randerson JT, Malmstrom CM, Field CB (1996) Change in net primary production and heterotrophic respiration: how much is necessary to sustain the terrestrial carbon sink? *Global Biogeochemical Cycles*, **10**, 711–726.

Tolonen K, Turunen J (1996) Accumulation rates of carbon in mires in Finland and implications for climate change. *Holocene*, **6**, 171–178.

Trumbore SE, Harden JW (1997) Input, accumulation, and turnover of carbon in soils of the BOREAS northern study area. *Journal of Geophysical Research Atmospheres*, **102**, 28,805–28,816.

Van Cleve K (1971) Energy- and weight-loss functions for decomposing foliage in birch and aspen forests in interior Alaska. *Ecology*, **52**, 720–723.

Van Cleve K (1974) Organic matter quality in relation to decomposition. In: *Soil Organisms and Decomposition in Tundra* (eds Holding AJ et al.), pp. 311–324. Tundra Biome Steering Committee, Stockholm.

Verhoeven JTA, Toth E (1995) Decomposition of Carex and *Sphagnum* litter in fens: effects of litter quality and inhibition by living tissue homogenates. *Soil Biology and Biochemistry*, **27**, 271–275.

Vogt KA, Grier CC, Vogt DJ (1986) Production, turnover and nutrient dynamics of above- and belowground detritus of world forests. *Advances in Ecological Research*, **15**, 303–375.

Waddington JM, Roulet NT (1996) Atmosphere-wetland carbon exchanges: scale dependency of CO2 and CH4 exchange on...
the developmental topography of a peatland. *Global Biogeochemical Cycles*, 10, 233–245.

Waelbroeck C, Monfray P, Oechel WC, Hastings S, Vourlitis G (1997) The impact of permafrost thawing on the carbon dynamics of tundra. *Geophysical Research Letters*, 24, 229–232.

Wagener SM (1995) Ecology of birch litter decomposition and forest floor processes in the Alaskan taiga. PhD thesis, University of Alaska, Fairbanks, AK.

Wein RW (1976) Frequency and characteristics of arctic tundra fires. *Arctic*, 29, 213–222.

Williams JE (1949) Chemical weathering at low temperatures. *Geophysical Review*, 39, 129–135.

Winston G, Sundquist ET, Stephen BB, Trumbore SE (1997) Winter CO₂ fluxes in a boreal forest. *Journal of Geophysical Research Atmospheres*, 102, 28,795–28,804.

Wookey PA, Robinson CH, Parsons AN, Welker JM, Callaghan TV, Lee JA (1995) Environmental constraints on the growth, photosynthesis and reproductive development of *Dryas octopetala* at a high Arctic polar semi-desert, Svalbard. *Oecologia*, 102, 478–489.

Zimov SA, Zimova GM, Daviodov SP et al. (1993) Winter biotic activity and production of CO₂ in Siberian soils: a factor in the greenhouse effect. *Journal of Geophysical Research*, 98D, 5017–5023.

Zimov SA, Davidov SP, Voropaev YV et al. (1996) Siberian CO₂ efflux in winter as a CO₂ source and cause of seasonality in atmospheric CO₂. *Climatic Change*, 33, 111–120.
## Appendix

Sources of decomposition data used analyses of controls over litter decomposition in arctic and boreal ecosystems. Dashes indicate no information available.

| Location                  | MAP (mm) | MAT (°C) | Site                                           | Topographic position | Growth form | Litter source | Mass loss (% after 1 y) | Reference                      | Notes                          |
|---------------------------|----------|----------|------------------------------------------------|-----------------------|-------------|---------------|-------------------------|--------------------------------|--------------------------------|
| 54° 50′ N, 95° 30′ W      | 700      | 2.8      | Little Sioux Wildfire, MN, USA                  | upland                | deciduous   | *Populus tremuloides* | 41.075                  | [Grigal & McColl](1977)          | [Climatology Working Group](1977) |
| 49° 53′ N, 95° 54′ W      | 613      | 2.1      | Elma, Manit., Canada                           | lowland               | moss        | *Salix bebbiana*    | 29                      | [Reader & Stewart](1972)          |                                |
| 53° 38′ N, 106° 13′ W     | 456      | -0.2     | Pr. Albert Natl. Park, Sask., Canada           | upland                | deciduous   | *Populus tremuloides* | 38.5                  | [Huang & Schoenau](1997)         |                                |
| 53° 57′ N, 105° 30′ W     | 623      | 0.6      | Candle Lake, Sask., Canada                     | lowland               | moss        | *Hylocomium splendens* | 12.1                  | [Nakatsubo et al.](1997)        |                                |
| 54° 15′ N, 116° W         | 540      | -0.2     | Whitecourt, Alberta, Canada                    |                       | evergreen   | *Pinus banksiana × P. contorta* | 12.2                  | [Prescott & Parkinson](1985)     | [Nakatsubo et al.](1997)        |
| Location | MAP (mm) | MAT (°C) | Site | Topographic position | Growth form | Litter source | Mass loss (% after 1 y) | Reference | Notes |
|----------|----------|----------|------|----------------------|-------------|--------------|------------------------|-----------|-------|
| 54°43'N, 66°42'E | 785 | -0.2 | Schefferville, Quebec, Canada |  |  |  | 13.9 | Moore (1984) | MAT from Finer et al. (1997) |
| 56°36'N, 13°15'E | 1070 | 6.8 | Mastocka, Sweden |  |  |  | 42.2 | Berg et al. (1993) |  |
| 57°10'N, 19°50'E | 702.5 | 6 | Akhelt Mire, Sweden | lowland | moss | Sphagnum cuspidatum | 19.4 | Johnson & Damman (1991) |  |
| 58°04'N, 14°08'E | 595 | 5.1 | Sanskjon, Sweden | upland | evergreen | Picea mariana | 11.4 | Berg et al. (1993) |  |
| 58°06'N, 13°17'E | 600 | 5.4 | Tveten, Sweden |  |  |  | 30 | Berg et al. (1993) |  |
| 58°28'N, 13°39'E | 550 | 5.4 | Remningstorp, Sweden |  |  |  | 36.3 | Berg et al. (1993) |  |
| 58°33'N, 15°51'E | 525 | 6.1 | Grensholm, Sweden |  |  |  | 42 | Berg et al. (1993) |  |
| 59°07'N, 15°44'E | 550 | 5.5 | Dimbo, Sweden |  |  |  | 43.7 | Berg et al. (1993) |  |
| 59°31'N, 17°16'E | 470 | 5.3 | Kungs-Husby, Sweden |  |  |  | 38.1 | Berg et al. (1993) |  |
| 59°49'N, 16°33'E | 500 | 5 | Tomta, Sweden |  |  |  | 36.9 | Berg et al. (1993) |  |
| 60°17'N, 7°40'E | 700 | -3 | Hardangervidda, Norway |  |  |  | 24 | Heal & French (1974) | location, MAP, MAT, elev. from French (1974), Rosswall (1974) |
| 60°18'N, 7°41'E | 752 | -2.1 |  | lowland | deciduous |  | 10 |  |  |
| 60°25'N, 7°21'E | 1013 | 0 |  | upland | herbaceous |  | 28 |  |  |
| 60°49'N, 16°30'E | 609 | 3.8 | Ivantjarnsheden, Sweden |  |  |  | 40.9 | Berg & Ekbohm (1991) |  |
| 63°13'N, 14°28'E | 455 | 2 | Vastbyn, Sweden |  |  |  | 36.4 | Berg et al. (1993) |  |
| Location | MAP (mm) | MAT (°C) | Site | Topographic position | Growth form | Litter source | Mass loss (% after 1 y) | Reference | Notes |
|----------|---------|---------|------|----------------------|-------------|--------------|------------------------|-----------|-------|
| 64°21'N, 20°53'E | 595 | 1.2 | Norrileden, Sweden | "" | "" | "" | 17.4 | Flanagan & Van Cleve (1983) | "" |
| 64°53'N, 147°45'W | 305 | -3.6 | Fairbanks, AK, USA | "" | "" | "" | 40.6 | Van Cleve (1971) | "" |
| 65°47'N, 20°37'E | 700 | 1 | Manjerv 3:1, Sweden | "" | evergreen | "" | 19.8 | Berg et al. (1993) | "" |
| 66°08'N, 20°53'E | 650 | -1.3 | Harads, Sweden | "" | "" | "" | 11.1 | "" | "" |
| 68°21'N, 18°49'E | 300 | -1 | Abisko, Sweden | "" | "" | "" | 39.2 | Robinson et al. (1995) | "" |
| 68°38'N, 149°34'W | 300 | -7 | Toolik Lake, AK, USA | "" | "" | "" | 23.2 | Shaver et al. (1997) | "" |
| Location          | MAP (mm) | MAT (°C) | Site        | Topographic position | Growth form | Litter source     | Mass loss (% after 1 y) | Reference         | Notes                  |
|-------------------|----------|----------|-------------|----------------------|-------------|-------------------|------------------------|---------------------|------------------------|
| 69°45'N, 27°01'E  | 420      | -2.4     | Kevo, Finland | evergreen            |            | Ledum palustre    | 32.8                   | Shaver et al. (1997) | location, MAP, MAT,   |
|                   |          |          |             | herbaceous           | moss        | Enophorium vaginatum | 9.0                    |                     | elev. from French     |
|                   |          |          |             |                      | Sphagnum spp.| Aulacomnium turgidum | 12.9                   |                     | (1974), Rosswall (1974)|
| 71°20'N, 156°39'W | 108      | -12.4    | Barrow, AK, USA | evergreen            | lowland     | Pinus sylvestris   | 15                     |                     |                        |
|                   |          |          |             | herbaceous           |             | Dupontia fischeri | 16                     |                     |                        |
|                   |          |          |             |                      |             | Enophorium angustifolium | 6                     |                     |                        |
|                   |          |          |             |                      |             | Carex aquatilis    | 5                      |                     |                        |
| 75°N, 95°W        | 196      | -15      | Devon Island, Canada | upland         |             | Carex stans      | 20                     | Robinson et al. (1995)| climate data from     |
| 78°56'N, 11°50'E  | 371      | -6       | Svalbard, Norway |           |              | Salix polaris     | 12.7                   |                     | Wookey et al. (1995)  |

1http://www.soils.agri.umn.edu/research/climatology/