Integrative transcriptome data mining for identification of core lncRNAs in breast cancer

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ABSTRACT

Background. Cumulative evidence suggests that long non-coding RNAs (lncRNAs) play an important role in tumorigenesis. This study aims to identify lncRNAs that can serve as new biomarkers for breast cancer diagnosis or screening.

Methods. First, the linear fitting method was used to identify differentially expressed genes from the breast cancer RNA expression profiles in The Cancer Genome Atlas (TCGA). Next, the diagnostic value of all differentially expressed lncRNAs was evaluated using a receiver operating characteristic (ROC) curve. Then, the top ten lncRNAs with the highest diagnostic value were selected as core genes for clinical characteristics and prognosis analysis. Furthermore, core lncRNA-mRNA co-expression networks based on weighted gene co-expression network analysis (WGCNA) were constructed, and functional enrichment analysis was performed using the Database for Annotation, Visualization and Integrated Discovery (DAVID). The differential expression level and diagnostic value of core lncRNAs were further evaluated by using independent data set from Gene Expression Omnibus (GEO). Finally, the expression status and prognostic value of core lncRNAs in various tumors were analyzed based on Gene Expression Profiling Interactive Analysis (GEPIA).

Results. Seven core lncRNAs (LINC00478, PGM5-AS1, AL035610.1, MIR143HG, RP11-175K6.1, AC005550.4, and MIR497HG) have good single-factor diagnostic value for breast cancer. AC093850.2 has a prognostic value for breast cancer. AC005550.4 and MIR497HG can better distinguish breast cancer patients in early-stage from the advanced-stage. Low expression of MAGI2-AS3, LINC00478, AL035610.1, MIR143HG, and MIR145 may be associated with lymph node metastasis in breast cancer.

Conclusion. Our study provides candidate biomarkers for the diagnosis and prognosis of breast cancer, as well as a bioinformatics basis for the further elucidation of the molecular pathological mechanism of breast cancer.

Subjects Bioinformatics, Oncology

Keywords IncRNA, Breast cancer, Biomarkers, Bioinformatics, WGCNA
INTRODUCTION

Long non-coding RNAs (lncRNAs) are a type of non-coding RNA molecules with a length of more than 200 nucleotides (McFadden & Hargrove, 2016). Studies have shown that lncRNAs are involved in a variety of biological processes (e.g., genetic imprinting, cell differentiation, apoptosis, cell proliferation, and immune response) (Zhu et al., 2018). The value of lncRNAs in the diagnosis and prognosis of tumors has now been widely recognized (Yang et al., 2016).

Breast cancer is the most common malignant tumor in women (Bertoli, Cava & Castiglioni, 2015). In recent years, the clinical management of breast cancer has improved, but the cancer is still a serious threat to women’s health due to its poor prognosis and high mortality rate (Pelosi et al., 2014). With the rapid development of tumor genomics, the discovery of highly specific biomarkers has become an intrinsic requirement for clinical diagnosis and treatment of breast cancer. Numerous transcripts of lncRNAs provide a wealth of materials for our search for quality biomarkers.

The Cancer Genome Atlas (TCGA), as the most authoritative cancer genome database, provides abundant raw data for large-scale tumor researches (Gao et al., 2019). A series of highly sensitive and specific tumor biomarkers based on TCGA data mining have been discovered and applied with the assistance of various assistive technologies (Gao et al., 2018). WGCNA, featuring soft threshold settings and a weighted network, can efficiently identify highly synergistically altered gene sets (Langfelder & Horvath, 2007; Sutherland et al., 2016). The construction of key target co-expression networks based on WGCNA provides methodological support for exploring the molecular mechanisms of diseases.

In this study, we used the limma package for identifying differentially expressed lncRNAs in breast cancer. The ROC curve was used to evaluate and select the top ten differentially expressed lncRNAs with the highest diagnostic value as core genes. We then comprehensively analyzed the clinical features and prognostic value of these core genes in breast cancer. The WGCNA co-expression networks and their functional enrichment analysis assisted us in exploring the role of core genes and their key targets in breast cancer. GEO’s independent data set was used to evaluate the expression level and diagnostic value of core lncRNAs. Based on GEPIA, we explored the expression status and prognostic value of core genes in a variety of tumors. Through this research, we hope to identify high-quality biomarkers for breast cancer from the perspective of lncRNAs, and provide a bioinformatics basis for further elucidation of the molecular pathogenesis of breast cancer.

MATERIALS & METHODS

Downloading breast cancer RNA expression profile data from TCGA

We downloaded high-throughput data of breast cancer RNA-Seq from TCGA (Wu et al., 2016. The access time for the TCGA data is February 1, 2019, and the species is human. These RNA-Seq data were derived from 837 breast cancer tumor samples and 105 adjacent non-cancerous breast tissue samples. This study strictly adheres to the TCGA publication guidelines.
Identification of differentially expressed lncRNAs in breast cancer

The breast cancer RNA-Seq data we downloaded covers 12,727 lncRNAs described by NCBI (http://www.ncbi.nlm.nih.gov) or Ensembl (http://asia.ensembl.org/index.html). We used the limma package (in R language) to identify differentially expressed lncRNAs ($P < 0.05$ and $|\log_{2}FC| > 1$) (Ritchie et al., 2015). The limma package is an excellent analytical statistical tool for differential expression analysis of gene expression chip data. ROC curves were used to assess the diagnostic effect of all differentially expressed lncRNAs in breast cancer (Patel et al., 2019). ROC curves were plotted based on the expression value of each core lncRNA and the type of samples (either breast cancer tissue or non-cancerous breast tissue). According to the expression value of each core lncRNA, we classified the 837 breast cancer tumor samples and 105 adjacent non-cancerous breast tissue samples (downloaded from TCGA) into the breast cancer group and non-breast cancer group. The results of the grouping according to the classification were compared with the actual species of the sample (breast cancer tissue or non-breast cancer tissue) to obtain a false positive rate and a true positive rate. These rates were respectively represented on the abscissa ($X$ axis) and the ordinate ($Y$ axis) to plot the ROC curves. The area under the curve (AUC) was calculated, and the statistical standard of AUC > 0.7 and $P < 0.05$ was used (the higher the AUC, the better the diagnostic effect). The top 10 lncRNAs of most diagnostic values were further analyzed as core lncRNAs.

Clinical characteristics analysis and prognostic value evaluation of core genes

In order to get a more accurate result, a Student’s t-test was also used to evaluate the differential expression level of core lncRNAs in breast cancer tissues and non-cancer breast tissues. A Student’s t-test can highlight a difference between two groups. (Ryu et al., 2018). Similarly, differences in the expression of core genes between different clinical groups were evaluated. K-M analysis (Gökbuget et al., 2018) was used to assess the prognostic value of core lncRNAs. The sample data for the K-M survival analysis was derived from the corresponding clinical data (including survival time and survival status) of the 837 breast cancer tumor samples originally downloaded from TCGA. According to the median value of each core lncRNA expression value in R language, the patients were divided into the high expression group and low expression group. Genes of two groups of patients with significantly different overall survival (OS) rate ($P < 0.05$) were selected as prognostic factor. The $p$-value of logrank <0.05 was statistically significant. In order to select better prognostic biomarkers, we also added univariate cox analysis (Zhao et al., 2018b) to these core lncRNAs.

WGCNA-based construction of core lncRNA-mRNA co-expression network and its functional enrichment analysis

The core lncRNA-mRNA co-expression networks based on WGCNA were constructed to explore the molecular mechanism of core genes in breast cancer (Chen et al., 2015). The threshold is 0.02. We used Cytoscape to visualize the co-expression networks. The DAVID database (david.ncifcrf.gov) was used for GO analysis of co-expression networks. The widely used GO analysis can perform functional enrichment analysis of gene collections.
from three aspects: biology pathway (BP), cell component (CC), and molecular function (MF) (Chai et al., 2016). $P < 0.05$ was considered statistically significant.

**Evaluation of core IncRNAs using GEO data sets**

We downloaded the data of GEO’s independent data set GSE125677. GSE125677 contained IncRNAs sequencing data derived from three paired breast cancer and their adjacent normal tissues, and the identified 10 core IncRNAs were included. Independent sample t-test was conducted to analyze differential expression levels of core IncRNAs between breast cancer tumor and adjacent breast tissues (Al-Hashel et al., 2018). A test result with $|t|>2$ and $p < 0.05$ was considered statistically significant. ROC curves were used to assess the diagnostic effect of core IncRNAs in breast cancer. Patel et al. (2019). AUC is the area under the curve of the ROC curve. The higher the AUC, the better the diagnostic effect is considered to be. ROC curves were plotted based on the expression value of each core IncRNA and the type of samples, namely either breast cancer tissue or non-cancerous breast tissue. An IncRNA with AUC >0.7 and $P < 0.05$ is considered to have a good diagnostic value for breast cancer.

**GEPIA-based analysis of expression patterns and prognostic value of core IncRNAs in a variety of tumors**

GEPIA was developed by Zefang Tang, Chenwei Li and Boxi Kang of Zhang’s Lab in Peking University (Tang et al., 2017). The RNA-Seq datasets GEPIA used is based on the UCSC Xena project (http://xena.ucsc.edu), which computed all expression raw data with a standard pipeline. Therefore, with GEPIA we could efficiently obtain information about the core IncRNAs from TCGA database. We used GEPIA to analyze the differential expression of core IncRNAs in 31 tumor tissues and adjacent non-cancerous tissues. A difference or result with $P < 0.01$ and $|\log_2 FC|>1$ was considered statistically significant. Meanwhile, we assessed the single-factor prognostic value of core IncRNAs in these 30 tumors other than breast cancer. $P < 0.05$ was considered statistically significant.

**RESULTS**

**Differentially expressed IncRNAs selected from breast cancer RNA expression data in TCGA**

We identified 276 differentially expressed IncRNAs in breast cancer (Fig. 1), among which 164 were up-regulated and 112 were down-regulated. The statistical criteria were $P$ value $< 0.05$ and $|\log_2 FC|>1$. The top 22 in the results of ROC analysis of differentially expressed IncRNAs was shown in Table S1 ($P < 0.05$ and AUC $>0.95$). The following top 10 IncRNAs with highest diagnostic value were used as core genes for subsequent analysis: AC093850.2, MAGI2-AS3, LINC00478, PGM5-AS1, AL035610.1, MIR143HG, RP11-175K6.1, AC005550.4, MIR497HG, and MIR145 (see Table 1 for details).

**An analysis of clinical characteristics of core IncRNAs in breast cancer**

The level of AC093850.2 in breast cancer was significantly higher than that in adjacent breast tissues. In contrast, nine other IncRNAs were down-regulated in breast cancer tissues; and
the low expression of MIR145 was most significant ($|\log_2\text{FC}|>16$) (Fig. 2). These ten core lncRNAs had high diagnostic value for distinguishing between breast cancer tissues and non-cancer tissues, with the smallest AUC = 0.969 (Fig. 3). K-M survival analysis showed that AC093850.2 was significantly correlated with breast cancer survival ($P = 0.0075$, Zhang et al. (2019), PeerJ, DOI 10.7717/peerj.7821).
Figure 2  Differential expression analysis of the core genes between breast cancer and para-tumorous breast tissues. Deep yellow column: breast cancer tissues (BRCA). Blue column: breast para-tumorous tissue (pT). X axis: tissue types. Y axis: Normalized expression. (A) AC093850.2; (B) MAGI2-AS3; (C) LINC00478; (D) PGM5-AS1; (E) AL035610.1; (F) MIR143HG; (G) RP11-175K6.1; (H) AC005550.4; (I) MIR497HG; (J) MIR145. ***P < 0.001.

Figure 3  ROC curves of the core lncRNAs in breast cancer. Deep yellow: sensitive curve. Blue: identify line. X axis: 1-specificity. Y axis: sensitivity. AUC: area Under Curve. The higher the AUC, the better the diagnostic effect. (A) AC093850.2; (B) MAGI2-AS3; (C) LINC00478; (D) PGM5-AS1; (E) AL035610.1; (F) MIR143HG; (G) RP11-175K6.1; (H) AC005550.4; (I) MIR497HG; (J) MIR145.

Fig. 4). Figure 4 shows the K-M curves and contains the p-value of logrank. In addition, univariate cox analysis also showed that AC093850.2 was an independent prognostic indicator for breast cancer (P = 0.000482235, Table S2). Several core lncRNAs were closely related to some clinical parameters of breast cancer. The main results are shown in Table 2 and Fig. 5 (P < 0.05, |t| ≥ 1.987). The expression levels of AC005550.4 and MIR497HG can distinguish the breast cancer patients in early-stage from the advanced-stage. Compared with the clinical group of breast cancer without lymph nodal involvement, these five core lncRNAs (MAGI2-AS3, LINC00478, AL035610.1, MIR143HG, and MIR145) had lower expression level in the group with lymph nodal involvement.
Figure 4  K-M curves of the core lncRNAs in breast cancer. The prognostic significance of the core lncRNAs in patients with breast cancer, according to the Kaplan–Meier analysis (in R) of data from TCGA. The deep yellow lines represent patients with high gene expression, and blue lines represent patients with a low gene expression. (A) AC093850.2; (B) MAGI2-AS3; (C) LINC00478; (D) PGM5-AS1; (E) AL035610.1; (F) MIR143HG; (G) RP11-175K6.1; (H) AC005550.4; (I) MIR497HG; (J) MIR145.

Table 2  Relationship between the expression of core lncRNAs and different clinical parameters in breast cancer from TCGA.

| lncRNA factor | Gender (female/male) | T (T1/2 vs. T3/4) | N (no/yes) | M (no/yes) | Pathological stage (I/II vs. III/IV) | Targeted molecular therapy (no/yes) |
|---------------|----------------------|------------------|------------|------------|-------------------------------------|-----------------------------------|
|               | t                     | P                | t          | P          | t                    | P                | t              | P                |
| AC093850.2    | −1.675                | 0.132            | 1.322      | 0.188      | −1.217               | 0.224            | 0.764          | 0.459            |
| MAGI2-AS3     | 0.418                 | 0.686            | 0.908      | 0.365      | −2.824               | 0.005            | −0.32          | 0.754            |
| LINC00478     | 5.277                 | 0.0003           | −1.559     | 0.122      | −3.199               | 0.0014           | −0.463         | 0.651            |
| PGM5-AS1      | 1.245                 | 0.235            | 1.651      | 0.099      | −0.544               | 0.586            | 0.719          | 0.48             |
| AL035610.1    | 4.638                 | 0.0007           | 0.1461     | 0.884      | −1.987               | 0.048            | 0.338          | 0.741            |
| MIR143HG      | 2.471                 | 0.03             | −0.52      | 0.604      | −2.882               | 0.004            | −0.452         | 0.659            |
| RP11-175K6.1  | 0.209                 | 0.839            | 1.195      | 0.234      | −1.559               | 0.12             | 0.347          | 0.734            |
| AC005550.4    | 5.514                 | 0.03E−08         | 2.194      | 0.029      | 0.978                | 0.329            | 1.738          | 0.102            |
| MIR497HG      | 1.479                 | 0.175            | −2.257     | 0.026      | −0.62                | 0.535            | 0.087          | 0.932            |
| MIR145        | 0.455                 | 0.66             | −0.037     | 0.971      | −2.545               | 0.012            | −0.437         | 0.67             |

Notes.

T, tumor stage; N, lymph node; M, metastasis.
P < 0.05 and | t | > 2 are statistically significant.

The roles of core lncRNAs in breast cancer based on WGCNA network

The results show that six core genes (AC005550.4, AL035610.1, PGM5-AS1, RP11-175K6.1, MAGI2-AS3, and MIR497HG), especially the first three, in the lncRNA-mRNA WGCNA networks were closely related. In addition, AC093850.2 and MIR143HG had a large number of co-expressed mRNAs (Fig. 6). The GO analysis showed that AC005550.4 and AL035610.1 were significantly enriched in biology pathways such as glucose metabolism, lipid metabolism, and alcohol metabolism. Meanwhile, co-expressed mRNAs of RP11-175K6.1 were significantly enriched in biology pathways including protein kinase B
Figure 5  Relationship between the expression of core lncRNAs and different clinical parameters in breast cancer from TCGA. The statistical significance of differences was calculated by the Student’s t-test. \( P < 0.05, |t| > 1.987 \). (A) Gender: deep yellow, female; blue, Male. (B) Lymph Node: deep yellow, no; blue, yes. (C) Targeted Molecular Therapy: deep yellow, no; blue, yes. (D) Tumor Stage: deep yellow, T1-T2; blue, T3-T4.

Evaluation of the expression and diagnostic value of all core lncRNAs
in GEO independent datasets

Information of all core lncRNAs can be obtained from GSE125677. With the exception of the high expression of AC093850.2 in breast cancer tissues, the other nine lncRNAs
were lowly expressed in cancer tissues. Moreover, the ROC curves of these core lncRNAs indicated they had a good diagnostic value for distinguishing breast cancer patients from the healthy population. This supported our findings of the analysis based on the TCGA database (see Table 3 and Fig. 7).
Table 3  Validation of expression and diagnostic value of core lncRNAs in breast cancer based on GEO dataset (GSE125677).

| Variable  | n  | pT Mean   | SD    | BC Mean   | SD    | T-test t | P     | ROC AUC | SE    | 95% CI       | P       |
|-----------|----|-----------|-------|-----------|-------|----------|-------|---------|-------|--------------|---------|
| AC093850.2| 6  | 0.34478499| 0.94650167| 6  | −0.344785| 0.8436006| 1.6316 | 0.1225  | 0.852 | 0.0997       | 0.656–1.000 | 0.0004236 |
| MAGI2-AS3 | 6  | −0.4193923| 0.6252397 | 6  | 0.4193923| 1.0432272| −2.069 | 0.0589  | 0.79  | 0.1184       | 0.558–1.000 | 0.01428  |
| LINC00478 | 6  | −0.4452022| 0.9184215 | 6  | 0.4452022| 0.7652063| −2.2345| 0.04056 | 0.852 | 0.0907       | 0.674–1.000 | 0.0001052 |
| PGM5-AS1  | 6  | −0.4562647| 0.638028  | 6  | 0.4562647| 0.99976049| −2.3082| 0.03728 | 0.827 | 0.113        | 0.606–1.000 | 0.003783  |
| AL035610.1 | 6  | −0.7851672| 0.49833236| 6  | 0.7851672| 0.4894569| −6.7445| <0.0001 | 1    | 0           | 1.000–1.000 | <0.0001  |
| MIR143HG  | 6  | −0.4520927| 0.6770228 | 6  | 0.4520927| 0.9781449| −2.2802| 0.03849 | 0.753 | 0.0918       | 0.519–0.987 | 0.034    |
| RP11-175K6.1 | 6  | −0.6691243| 0.6003143 | 6  | 0.6691243| 0.7122052| −4.3102| <0.0001 | 0.914 | 0.0682       | 0.780–1.000 | <0.0001  |
| AC005550.4 | 6  | −0.559967 | 0.9184722 | 6  | 0.55996702| 0.57086945| −3.1068| 0.008093 | 0.827 | 0.1189       | 0.594–1.000 | 0.00593  |
| MIR497HG  | 6  | −0.8101302| 0.5172673 | 6  | 0.8101302| 0.3615728| −7.702< 0.0001 | 1    | 0           | 1.000–1.000 | <0.0001  |
| MIR145    | 6  | −0.3768513| 0.7582684 | 6  | 0.3768513| 0.9901974| 1.813  | 0.08992 | 0.728 | 0.1245       | 0.484–0.973 | 0.06665  |

Notes.

pT, para-tumor tissue; BC, breast cancer.
Table 4: Differential expression of seven core lncRNAs in tumor tissues and para-tumor tissues and their prognostic value in 31 tumors based on GEPIA. Acronyms for these diseases are provided by GEPIA (http://gepia.cancer-pku.cn/detail.php?gene=&clicktag=boxplot##).

| core lncRNAs   | In which diseases the core gene is up-regulation ($P < 0.01$) | In which diseases the core gene is down-regulation ($P < 0.01$) | In which diseases the core gene have prognostic value ($P < 0.05$) |
|----------------|-------------------------------------------------------------|-------------------------------------------------------------|-------------------------------------------------------------|
| AC093850.2     | BRCA/LUAD/LUSC/PAAD                                          | BRCA/ACC/BLCA/CESC/CHOL/COAD/ESCA/KICH/KIRP/LIHC/LUAD/     | GBM/KICH/LUSC/MESO/STAD/                                     |
| MAGI2-AS3      | PAAD/SKCM/THYM                                               | BRCA/ACC/BLCA/CESC/CHOL/COAD/ESCA/KICH/KIRP/LIHC/LUAD/     | BLCA/LGG/LUSC/STAD/UVM                                      |
| PGM5-AS1       |                                                             |                                                            |                                                            |
| AL035610.1     | KIRC/KIRP/OV                                                | BRCA/ESCA/HNSC/KICH/SARC                                   | LGG/MESO                                                    |
| MIR143HG       |                                                             |                                                            |                                                            |
| RP11-175K6.1   | DLBC/GBM                                                    | BRCA/BLCA/CESC/SKCM/TGCA/UVCEC/                           | KIRC/UCRC                                                   |
| MIR497HG       | DLBC/KIRC/LGG/THYM                                           | BRCA/ACC/BLCA/CESC/COAD/ESCA/KICH/LUAD/LUSC/PRAD/READ/    | BLCA/GBM/KICH/LUAD/SKCM/UVM                                 |

Notes.
Full names of these diseases can be found in Table S5.

Expression pattern and prognostic value of core lncRNAs in various tumors based on GEPIA

According to GEPIA, seven core lncRNAs (AC093850.2, MAGI2-AS3, PGM5-AS1, AL035610.1, MIR143HG, RP11-175K6.1, and MIR497HG) were differentially expressed in various tumors. Meanwhile, these seven core lncRNAs had a good single-factor prognostic value for a variety of tumors other than breast cancer. Details are shown in Table 4.

DISCUSSION

With the rapid development of molecular genomics, highly specific diagnostic and prognostic biomarkers have provided molecular evidence for improvements in conventional diagnostic and therapy protocols. More and more lncRNAs are recognized as oncogenes or tumor suppressors. As a complement to the coding genes and miRNAs, lncRNAs have brought new hope for the diagnosis and prognosis of breast cancer. In this study, from the perspective of lncRNAs, we selected biomarkers that are closely related to the diagnosis, clinical features, and prognosis of breast cancer.

lncRNAs have an advantage in the application of diagnosis (Richard & Eichhorn, 2018), and the detection of specific lncRNAs has been applied to the early diagnosis of cancers (Zhang & Tang, 2018). In this study, the 10 differentially expressed core lncRNAs (AC093850.2, MAGI2-AS3, LINC00478, PGM5-AS1, AL035610.1, MIR143HG, RP11-175K6.1, AC005550.4, MIR497HG, and MIR145) selected have high diagnostic value for breast cancer. This suggests that they were closely linked to the risk of breast cancer. After further evaluation using the independent data set (GSE125677) in GEO, we gained results (see Table 3) showing that the differential expression levels and diagnostic value of the
following seven core lncRNAs all reached statistical standards: LINC00478, PGM5-AS1, AL035610.1, MIR143HG, RP11-175K6.1, AC005550.4, and MIR497HG. However, a shortcoming of this study should be acknowledged, that is, the sample size of the data set used for evaluation is small. It is necessary to actively search for data sets of larger sample size in the follow-up studies in order to further explore and improve related biological experiments.

Further, the analysis of the clinical characteristics of the core lncRNAs showed that AC005550.4 and MIR497HG could better distinguish between early and late stages of breast cancer: the expression level of AC005550.4 in the early stage of breast cancer was higher than the late stage; and the expression level of MIR497HG in early breast cancer was lower than that in advanced stage. It is worth noting that, compared with those in the clinical group of breast cancer without lymph node metastasis, the five core lncRNAs (MAGI2-AS3, LINC00478, AL035610.1, MIR143HG, and MIR145) were expressed at lower levels due to lymph node metastasis in breast cancer. Lymphatic metastasis is generally recognized as an important indicator for prognosis (Yamashita et al., 1997), so that results support that the low expression of MAGI2-AS3, LINC00478, AL035610.1, MIR143HG, and MIR145 is associated with metastasis and progression of breast cancer. It has been reported that the down-regulation of MIR145 is closely correlated with the invasion behavior of malignant breast tumors (Radojicic et al., 2011).

There are many reports on MIR143HG, LINC00478, and MIR145. It has been reported that miR-143 (host gene: MIR143HG) reduces cell proliferation and migration, and the host gene MIR143HG can be regulated by miR-143 (Du et al., 2016). Zhao et al. (2018a) used miRNA microarray and RT-qPCR data to confirm that the expression of miR-143/145 and its host gene MIR143HG were down-regulated in tumor tissues of HBV-related liver cancer patients. LINC00478 was lowly expressed in cancer tissues of oral and oropharyngeal squamous cell carcinoma, which was verified by three data sets GSE42743, GSE9844, and GSE6791 (Feng et al., 2017). One of the introns of LINC00478, Let-7c, can exert a tumor suppressor effect by targeting oncogenes (Kumar & Purohit, 2014). It has also been reported that LINC00478 binds to androgen to down-regulate the expression of the miR-99a/let7c/125b-2 cluster produced by introns of LINC00478, then consequently prostate cancer growth factors IGF1R (targeted directly by miR-99a/let7c/125b-2 cluster) derepresses, triggering a series of downstream signaling cascades, which may contribute to prostate cancer progression (Sun et al., 2014). In our study, MIR145 was significantly down-regulated in breast cancer tissues (|log2FC|>16). MIR145 with Ensembl ID ENSG00000269936 (see Table 1) is miRNA-145′s host gene, and miRNA-145 could be used as a candidate biomarker for breast cancer diagnosis (Xiong et al., 2017). It has been reported that miR-SNP rs353291 (located in miRNA-145) is associated with the white population’s susceptibility to breast cancer (Chacon-Cortes et al., 2015). Studies on breast cancer cell lines have shown that miRNA-145 has a regulatory effect on genes that regulate apoptosis (Wang et al., 2009). A study found that the promoter region of MIR145 is hypermethylated in breast cancer clinical samples and breast cancer cell lines, and miRNA-145 is clearly down-regulated, which can inhibit breast cancer cells migration and invasion by directly targeting angiopoietin 2 gene (ANGPT2) (Liu et al., 2017).
The functional enrichment analysis of MIR145 co-expressed mRNAs also showed it has a significant correlation with biology pathways including negative regulation of blood vessel endothelial cell migration.

In addition, we also discovered a small number of reports on AC093850.2, MAGI2-AS3, and PGM5-AS1. A report has shown that MAGI2-AS3 inhibited the proliferation of breast cancer (Liu et al., 2017). MAGI2-AS3 has been reported to be a hub gene of the triple negative breast cancer IncRNA-mRNA co-expression network, and triple-negative breast cancer patients with elevated levels of MAGI2-AS3 have better recurrence-free survival rate (HR = 0.51) (Du et al., 2016). There are also reports that MAGI2-AS3 was down-regulated in cancer tissues of patients with bladder cancer, and it had low expression level and consequently poor prognosis (Wang et al., 2018a). As an immune-related IncRNA, PGM5-AS1 has prognostic value for regenerative glioma. The expression of PGM5-AS1 in the high-risk group was higher than that in the low-risk group (Wang et al., 2018b). Moreover, patients with high expression of PGM5-AS1 had lower OS (Zhu et al., 2017). At present we have not seen reports of four IncRNAs such as AL035610.1, RP11-175K6.1, AC005550.4, and MIR497HG. According to our functional enrichment analysis, AC093850.2 co-expressed mRNAs were closely related to biology pathways such as collagen metabolic process and glycosaminoglycan binding. Very similar to AC005550.4 co-expressed mRNAs, AL035610.1 co-expressed mRNAs are mainly involved in biological pathways including the metabolism of carbohydrates, lipids, alcohol, coenzyme metabolism, and coping peptides. RP11-175K6.1, closely related to the regulation of circadian rhythm, participated in the protein kinase B signaling pathway, regulated angiogenesis, and controlled the response of cells to steroid stimulation. These results provide guidance for further studies of the action mechanism of these IncRNAs in breast cancer.

It is worth noting that the six IncRNAs including MAGI2-AS3, PGM5-AS1, AL035610.1, RP11-175K6.1, AC005550.4, and MIR497HG were closely related in the WGCNA network, and we preliminarily deduced that their functions are similar, and this provides a good molecular function module for exploration of the disease mechanisms of breast cancer. These new IncRNAs (AL035610.1, RP11-175K6.1, AC005550.4, and MIR497HG) and the less frequently reported IncRNAs (AC093850.2, MAGI2-AS3, and PGM5-AS1) provide us with new ideas for understanding breast cancer. However, their mechanism of action remains to be explored, and their clinical potential in breast cancer needs further validation.

The K-M survival analysis and univariate cox analysis in this study showed that the core IncRNA AC093850.2, whose expression was significantly up-regulated in breast cancer (log2FC>6.5), has a certain prognostic value for breast cancer patients. Guo-Wei Huang (Huang et al., 2018) reported a result validated in a GEO dataset (GSE53624) that AC093850.2 was closely associated with OS and DFS in patients with esophageal squamous cell carcinoma. They found that with elevated AC093850.2 levels, the patient’s overall survival became shorter, and the relapse happened earlier. Moreover, their results indicate that knockdown of AC093850.2 had an inhibitory effect on both cell proliferation and cell migration. Other studies have shown that AC093850.2 is associated with survival in patients with invasive breast carcinoma (Zhao et al., 2018c).
To further understand the roles of core lncRNAs in tumors, we analyzed the expression levels of core lncRNAs in 31 tumor tissues and adjacent non-cancerous tissues based on GEPIA. According to the results of GEPIA, the expression pattern (up- or down-regulation) of AC093850.2, PGM5-AS1, and MIR143HG in other cancers was consistent with that in breast cancer. MAGI2-AS3, MIR143HG, and MIR497HG had significant differences in the expression in a variety of cancers. Based on these results, we deduced that they might have primarily been involved in the underlying pathological processes of cancer development. In particular, the expression of MIR143HG was significantly down-regulated in 16 cancers including breast cancer, suggesting that MIR143HG may be an important tumor suppressor gene. At the same time, we evaluated the single-factor prognostic value of core lncRNA in various tumors based on GEPIA and found 7 core lncRNAs (AC093850.2, MAGI2-AS3, PGM5-AS1, AL035610.1, MIR143HG, RP11-175K6.1, MIR497HG) has a good prognostic value for a variety of tumors other than breast cancer. This greatly expands our understanding of these core lncRNAs and provides a bioinformatics basis for further in-depth researches.

CONCLUSION

In summary, our results indicate that seven core lncRNAs (LINC00478, PGM5-AS1, AL035610.1, MIR143HG, RP11-175K6.1, AC005550.4, and MIR497HG) have good single-factor diagnostic value for breast cancer, while AC093850.2 has a certain prognostic value for breast cancer. AC005550.4 and MIR497HG could better distinguish the breast cancer patients in early-stage from the advanced-stage. Low expression of MAGI2-AS3, LINC00478, AL035610.1, MIR143HG, and MIR145 may be associated with lymph node metastasis in breast cancer. These findings not only provide potential biomarkers for the future diagnosis and prognosis of breast cancer, but also provide a new perspective for elucidating the molecular mechanisms underlying.

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The authors declare there are no competing interests.

Author Contributions
- Xiaoming Zhang, Cun Liu and Changgang Sun conceived and designed the experiments, authored or reviewed drafts of the paper, approved the final draft.
- Jing Zhuang, Lijuan Liu and Xia Ding analyzed the data, prepared figures and/or tables, approved the final draft.
- Zhengguo He analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Xiaoran Ma analyzed the data, contributed reagents/materials/analysis tools, prepared figures and/or tables, approved the final draft.
- Jie Li analyzed the data, contributed reagents/materials/analysis tools, approved the final draft.

Data Availability
The following information was supplied regarding data availability:
In this article, we used data from TCGA (https://portal.gdc.cancer.gov/; search terms: TCGA-BRCA) and GEO: GSE125677.

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REFERENCES
Al-Hashel JY, Ahmed SF, Alshawaf FJ, Alroughani R. 2018. Use of traditional medicine for primary headache disorders in Kuwait. The Journal of Headache and Pain 19(1):Article 118 DOI 10.1186/s10194-018-0950-3.
Bertoli G, Cava C, Castiglioni I. 2015. MicroRNAs: new biomarkers for diagnosis, prognosis, therapy prediction and therapeutic tools for breast cancer. Theraanostics 5(10):1122–1143 DOI 10.7150/thno.11543.
Chacon-Cortes D, Smith RA, Haupt LM, Lea RA, Youl PH, Griffiths LR. 2015. Genetic association analysis of miRNA SNPs implicates MIR145 in breast cancer susceptibility. BMC Medical Genetics 16:107 DOI 10.1186/s12881-015-0248-0.
Chai Y, Wang G, Fan L, Zhao M. 2016. A proteomic analysis of mushroom polysaccharide-treated HepG2 cells. Scientific Reports 6:23565 DOI 10.1038/srep23565.
Chen Y, Ni H, Zhao Y, Chen K, Li M, Li C, Zhu X, Fu Q. 2015. Potential role of lncRNAs in contributing to pathogenesis of intervertebral disc degeneration based on microarray data. Medical Science Monitor 21:3449–3458 DOI 10.12659/MSM.894638.
Du C, Shen Z, Zang R, Xie H, Li H, Chen P, Hang B, Xu X, Tang W, Xia Y. 2016. Negative feedback circuitry between MIR143HG and RBM24 in Hirschsprung disease. Biochimica et Biophysica Acta/General Subjects 1862(11):2127–2136 DOI 10.1016/j.bbadis.2016.08.017.
Feng L, Houck JR, Lohavanichbutr P, Chen C. 2017. Transcriptome analysis reveals differentially expressed lncRNAs between oral squamous cell carcinoma and healthy oral mucosa. *Oncotarget* 8(19):31521–31531 DOI 10.18632/oncotarget.16358.

Gao C, Zhou C, Zhuang J, Liu L, Liu C, Li H, Liu G, Wei J, Sun C. 2018. MicroRNA expression in cervical cancer: novel diagnostic and prognostic biomarkers. *Journal of Cellular Biochemistry* 119:7080–7090 DOI 10.1002/jcb.27029.

Gao C, Zhuang J, Zhou C, Ma K, Zhao M, Liu C, Liu L, Li H, Feng F, Sun C. 2019. Prognostic value of aberrantly expressed methylation gene profiles in lung squamous cell carcinoma: a study based on The Cancer Genome Atlas. *Journal of Cellular Physiology* 234:6519–6528 DOI 10.1002/jcp.27389.

Gökbuget N, Dombret H, Bonifacio M, Reichle A, Graux C, Faul C, Diedrich H, Topp MS, Brüggemann M, Horst H-A, Havelange V, Stiegmaier J, Wessels H, Haddad V, Benjamin JE, Zugmaier G, Nagorsen D, Bargou RC. 2018. Blinatumomab for minimal residual disease in adults with B-cell precursor acute lymphoblastic leukemia. *Blood* 131:1522–1531 DOI 10.1182/blood-2017-08-798322.

Huang G-W, Xue Y-J, Wu Z-Y, Xu X-E, Wu J-Y, Zhu Y, He J-Z, Li C-Q, Li E-M, Xu L-Y. 2018. A three-lncRNA signature predicts overall survival and disease-free survival in patients with esophageal squamous cell carcinoma. *BMC Cancer* 18 DOI 10.1186/s12885-018-4058-6.

Kumar A, Purohit R. 2014. Use of long term molecular dynamics simulation in predicting cancer associated SNPs. *PLOS Computational Biology* 10:e1003318 DOI 10.1371/journal.pcbi.1003318.

Langfelder P, Horvath S. 2007. Eigengene networks for studying the relationships between co-expression modules. *BMC Systems Biology* 1:54 DOI 10.1186/1752-0509-1-54.

Liu S-Y, Li X-Y, Chen W-Q, Hu H, Luo B, Shi Y-X, Wu T-W, Li Y, Kong Q-Z, Lu H-D. 2017. Demethylation of the MIR145 promoter suppresses migration and invasion in breast cancer. *Oncotarget* 8(37):61731–61741 DOI 10.18632/oncotarget.18686.

McFadden EJ, Hargrove AE. 2016. Biochemical methods to investigate lncRNA and the influence of lncRNA: protein complexes on chromatin. *Biochemistry* 55:1615–1630 DOI 10.1021/acs.biochem.5b01141.

Patel DA, Higginbotham T, Slaughter JC, Aslam M, Yuksel E, Katzka D, Gyawali CP, Mashi M, Pandolfino J, Vaezi MF. 2019. Development and validation of a mucosal impedance contour analysis system to distinguish esophageal disorders. *Gastroenterology* 156(6):1617–1626 DOI 10.1053/j.gastro.2019.01.253.

Pelosi A, Careccia S, Sagrestani G, Nanni S, Manni I, Schinzari V, Martens JH, Farsetti A, Stunnenberg HG, Gentileschi MP. 2014. Dual promoter usage as regulatory mechanism of let-7c expression in leukemic and solid tumors. *Molecular Cancer Research* 12:878–889 DOI 10.1158/1541-7786.MCR-13-0410.

Radojicic J, Zaravinos A, Vrekoussis T, Kafousi M, Spandidos DA, Stathopoulos EN. 2011. MicroRNA expression analysis in triple-negative (ER, PR and Her2/neu) breast cancer. *Cell Cycle* 10:507–517 DOI 10.4161/cc.10.3.14754.

Richard JLC, Eichhorn PJA. 2018. Deciphering the roles of lncRNAs in breast development and disease. *Oncotarget* 9(28):20179–20212 DOI 10.18632/oncotarget.24591.
Ritchie ME, Phipson B, Wu D, Hu Y, Law CW, Shi W, Smyth GK. 2015. limma powers differential expression analyses for RNA-sequencing and microarray studies. Nucleic Acids Research 43(7):e47–e47 DOI 10.1093/nar/gkv007.

Ryu H, Lim H, Choi G, Park Y-J, Cho M, Na H, Ahn CW, Kim YC, Kim W-U, Lee S-H, Chung Y. 2018. Atherogenic dyslipidemia promotes autoimmune follicular helper T cell responses via IL-27. Nature Immunology 19(6):583–593 DOI 10.1038/s41590-018-0102-6.

Sun D, Layer R, Mueller AC, Cichewicz MA, Negishi M, Paschal BM, Dutta A. 2014. Regulation of several androgen-induced genes through the repression of the miR-99a/let-7c/miR-125b-2 miRNA cluster in prostate cancer cells. Oncogene 33(11):1448–1457 DOI 10.1038/onc.2013.77.

Sutherland JJ, Jolly RA, Goldstein KM, Stevens JL. 2016. Assessing concordance of drug-induced transcriptional response in rodent liver and cultured hepatocytes. PLOS Computational Biology 12:e1004847 DOI 10.1371/journal.pcbi.1004847.

Tang Z, Li C, Kang B, Gao G, Li C, Zhang Z. 2017. GEPIA: a web server for cancer and normal gene expression profiling and interactive analyses. Nucleic Acids Research 45:W98–W102 DOI 10.1093/nar/gkx247.

Wang F, Zu Y, Zhu S, Yang Y, Huang W, Xie H, Li G. 2018a. Long noncoding RNA MAGI2-AS3 regulates CCDC19 expression by sponging miR-15b-5p and suppresses bladder cancer progression. Biochemical and Biophysical Research Communications 507:231–235 DOI 10.1016/j.bbrc.2018.11.013.

Wang S, Bian C, Yang Z, Bo Y, Li J, Zeng L, Zhou H, Zhao RC. 2009. miR-145 inhibits breast cancer cell growth through RTKN. International Journal of Oncology 34:1461–1466.

Wang W, Zhao Z, Yang F, Wang H, Wu F, Liang T, Yan X, Li J, Lan Q, Wang J. 2018b. An immune-related lncRNA signature for patients with anaplastic gliomas. Journal of Neuro-Oncology 136:263–271 DOI 10.1007/s11060-017-2667-6.

Wu X, Ruan L, Yang Y, Mei Q. 2016. Identification of crucial regulatory relationships between long non-coding RNAs and protein-coding genes in lung squamous cell carcinoma. Molecular and Cellular Probes 30:146–152 DOI 10.1016/j.mcp.2016.02.009.

Xiong D-D, Lv J, Wei K-L, Feng Z-B, Chen J-T, Liu K-C, Chen G, Luo D-Z. 2017. A nine-miRNA signature as a potential diagnostic marker for breast carcinoma: an integrated study of 1,110 cases. Oncology Reports 37:3297–3304 DOI 10.3892/or.2017.5600.

Yamashita H, Noguchi S, Murakami N, Kawamoto H, Watanabe S. 1997. Extracapsular invasion of lymph node metastasis is an indicator of distant metastasis and poor prognosis in patients with thyroid papillary carcinoma. Cancer 80:2268–2272 DOI 10.1002/(SICI)1097-0142(19971215)80:12<2268::AID-CNCR8>3.0.CO;2-Q.

Yang F, Liu Y-H, Dong S-Y, Ma R-M, Bhandari A, Zhang X-H, Wang O-C. 2016. A novel long non-coding RNA FGF14-AS2 is correlated with progression and prognosis in breast cancer. Biochemical and Biophysical Research Communications 470:479–483 DOI 10.1016/j.bbrc.2016.01.147.
Zhang Y, Tang L. 2018. The application of lncRNAs in cancer treatment and diagnosis. Recent Patents on Anti-Cancer Drug Discovery 13:292–301 DOI 10.2174/1574892813666180226121819.

Zhao Q, Sun X, Liu C, Li T, Cui J, Qin C. 2018a. Expression of the microRNA-143/145 cluster is decreased in hepatitis B virus-associated hepatocellular carcinoma and may serve as a biomarker for tumorigenesis in patients with chronic hepatitis B. Oncology Letters 15:6115–6122 DOI 10.3892/ol.2018.8117.

Zhao R, Zhang Y, Zhang X, Yang Y, Zheng X, Li X, Liu Y, Zhang Y. 2018b. Exosomal long noncoding RNA HOTTIP as potential novel diagnostic and prognostic biomarker test for gastric cancer. Molecular Cancer 17(1):Article 68 DOI 10.1186/s12943-018-0817-x.

Zhao W, Geng D, Li S, Chen Z, Sun M. 2018c. Lnc RNA HOTAIR influences cell growth, migration, invasion, and apoptosis via the miR-20a-5p/HMGA 2 axis in breast cancer. Cancer Medicine 7:842–855 DOI 10.1002/cam4.1353.

Zhu H, Yu J, Zhu H, Guo Y, Feng S. 2017. Identification of key lncRNAs in colorectal cancer progression based on associated protein–protein interaction analysis. World Journal of Surgical Oncology 15:Article 153 DOI 10.1186/s12957-017-1211-7.

Zhu N, Hou J, Wu Y, Liu J, Li G, Zhao W, Ma G, Chen B, Song Y. 2018. Integrated analysis of a competing endogenous RNA network reveals key lncRNAs as potential prognostic biomarkers for human bladder cancer. Medicine 97(35):e11887 DOI 10.1097/md.0000000000011887.