Introduction

Carbon nanotubes (CNTs) are carbon-based nanomaterials that have specific physicochemical and electronic properties [1] that make them applicable to various fields such as lubricating oils, fuel cells, drug delivery systems, and next generation semiconductors [2-4]. CNTs are often functionalized with hydrophilic ligands, as the commercial application of CNTs is often limited by their low solubility and dispersibility in organic or inorganic solutions [5]. To overcome this difficulty, the material surface is functionalized by introducing hydrophilic chemical groups, which lead to higher dispersibility while maintaining the unique properties of pristine CNTs [6]. Functionalized CNTs are therefore widely used as additives, catalysts, sensors, absorbents, intracellular carriers, electrodes, and imaging agents [7,8]. As the use of CNTs in applications and their commercialization has increased, human and environmental exposure to CNTs have also dramatically increased [9,10]. Increased exposure to CNTs has raised concerns over their potential toxicity for human health in the case of occupational and environmental exposure [11]. The potential toxicity of CNTs has been attributed to their physicochemical characteristics including length, diameter, structural defects, surface area, tendency to agglomerate, dispersibility in solution, the presence and nature of catalyst residues, as well as surface chemistry [12-16]. The structure of CNTs is similar to that of asbestos. The similarities in size and...
shape between CNTs and asbestos fibers have led to concerns about major toxicity of CNTs. Several studies have reported that the high-aspect-ratio (i.e., length divided by diameter) of CNTs can induce pulmonary toxicity [17,18]. As seen with asbestos, high-aspect-ratio multi walled CNTs (MWCNTs) have greater toxicity and potential to induce mesothelioma than low-aspect-ratio MWCNTs [19].

Most of the previous toxicity studies of CNTs have focused on respiratory exposure pathways in vivo or in vitro assays with pulmonary cells, due to the similarity in the structure of CNTs with that of asbestos [19,20]. As potential mechanisms of toxicity of CNTs, most studies emphasize oxidative stress, inflammation, and fibrosis, etc. [20,21]. However, only limited studies are available on the ecotoxicity of CNT [22,23], much less on C. elegans [24,25]. Another research gap in CNT toxicity lies in the potential hazard of functionalized CNTs. Though functionalized CNTs have wide application potential, most CNT toxicity studies have been conducted on pristine CNTs [26,27].

C. elegans is an emerging model in environmental toxicology, both for mechanistic studies as well as high-throughput screening. Various special features such as its well characterized genome, genetic manipulability, ease of maintenance, short and prolific life cycle, transparent and small body size, etc. have led to an increasing use of C. elegans in toxicology [28]. Most of all, a rich collection of available mutants has made it possible to establish mutant screening assays which are not only sensitive and useful in estimating the effects of toxicants but may also shed light on the underlying mechanisms [28,29]. Recently, C. elegans has also been used increasingly as a model for investigating the toxicity of nanomaterials [30].

It has been well documented that physicochemical properties play an important role in the uptake of nanomaterials and subsequently in their toxicity. As described above, aspect ratio is considered an important factor in determining the toxicity of CNTs. Therefore, in this study we investigated whether the outer diameter (OD) size and tube length of hydroxylated (OH)- MWCNTs affected their uptake and toxicity in C. elegans using loss-of-function mutants. Representative genes for uptake and possible toxic mechanism were selected and loss-of-function

**Table 1. The mutant strains used in this study**

| Genotype | Description |
|----------|-------------|
| rme-1 (b1045) V | RME-1 is required for endocytosis, specifically for modulating endocytic transport through the endosomal recycling compartment (ERC) |
| rme-2 (b1008) IV | RME-2 functions as the yolk receptor, whose activity is required for yolk uptake during oogenesis |
| rme-8 (b1023) I | RME-8 is required for both receptor-mediated endocytosis and pinocytosis (fluid-phase endocytosis) in coelomocytes and other cell types; rme-8 is also generally required for development and viability |
| sod-3 (gk235) X | SOD-3 encodes an iron/manganese superoxide dismutase, predicted to be mitochondrial, that might defend against oxidative stress and promote a normal lifespan; sod-3 mRNA levels are diminished by mutation of daf-16 and chromatin immunoprecipitation (ChIP) studies demonstrate that DAF-16 can directly bind the sod-3 promoter |
| ctl-2 (ek1137) II | CTL-2 exhibits catalase and peroxidase activity in vitro, and thus likely functions in vivo as an antioxidant enzyme that protects cells from reactive oxygen species; ctl-2 activity is required for a normal lifespan as well as for the extended lifespan seen in daf-2 mutant animals; in addition, ctl-2 is required for normal egg-laying capacity and for normal peroxisomal morphology |
| cyp-35A2 (gk317) V | CYP-35A2 encodes one of over 80 C. elegans cytochrome P450s, NADPH-dependent monooxygenases that metabolize endogenous and exogenous compounds; RT-PCR experiments indicate that cyp-35A2 mRNA is upregulated in response to treatment with xenobiotics |
| pmk-1 (km25) IV | PMK-1 encodes a mitogen-activated protein kinase (MAPK), orthologous to human p38 MAPK (OMIM:600289), that is required for eliciting gonadal programmed cell death in response to Salmonella enterica infection; PMK-1 lies upstream of CED-9, a negative regulator of apoptosis, in the programmed cell death pathway |
| daf-16 (mu86) I | DAF-16 encodes the sole C. elegans forkhead box 0 (FOXO) homologue; DAF-16 functions as a transcription factor that acts in the insulin/IGF-1-mediated signaling (IIS) pathway that regulates dauer formation, longevity, fat metabolism, stress response, and innate immunity; DAF-16 regulates various processes through isoform-specific expression, isoform-specific regulation by different AKT kinases, and differential regulation of target genes |
| CEP-1 (gk138) I | CEP-1 encodes an ortholog of the human tumor suppressor p53 (OMIM:191170, mutated in Li-Fraumeni syndrome) that promotes DNA damage-induced apoptosis, is required for normal meiotic segregation in the germ line, and affects sensitivity to hypoxia-induced lethality and longevity in response to starvation |
| hif-1 (la4) V | HIF-1 encodes an ortholog of the mammalian hypoxia-induced factor HIF-1, and is required for survival in hypoxic environments (1% oxygen) that have no effect on wild-type C. elegans |
| mtl-2 (gk125) V | MTL-2 functions in metal detoxification and stress adaptation; in addition, mtl-2 plays a role in regulating growth and fertility; mtl-2 expression is induced in larval and adult intestinal cells following exposure to cadmium or heat shock |
| hsp-16.2 (gk249) V | HSP-16.2 encodes a 16-kD heat shock protein (HSP) that is a member of the hsp16/hsp20/alphaB-crystallin (HSP16) family of heat shock proteins. HSP-16.2 has been shown to interact with intracellular human beta amyloid peptide, a primary component of the extracellular plaques found in Alzheimer’s disease; HSP-16.2 likely functions as a passive ligand temporarily preventing unfolded proteins from aggregating |
| eat-2 (ad465) II | EAT-2 functions postsynaptically in pharyngeal muscle to regulate the rate of pharyngeal pumping; eat-2 is also required for a normal life span and defecation |

*From WormBase http://www.wormbase.org/*
mutants of these genes were exposed to OH-MWCNTs and their responses compared with that of wildtype. We interpreted the arbitrary response of each mutant compared to wildtype as an indicator of how these genes are involved in the toxicity of different types of MWCNTs.

Materials and Methods

Growth of C. elegans and Treatment with Multi Walled Carbon Nanotubes

C. elegans were grown in Petri dishes on nematode growth medium and fed with OP50 strain Escherichia coli. Worms were incubated at 20°C, with young adults (3 days old) from an age-synchronized culture then being used in all the experiments. Wildtype and mutants were provided by the Caenorhabditis Genetics Center (www.CGC.org) at the University of Minnesota. Descriptions of the selected C. elegans mutants are in Table 1.

Multi-wall Carbon Nanotubes

Three different types of OH-MWCNTs were purchased from Cheaptube (www.cheaptube.com). Detailed physicochemical properties of the OH-MWCNTs are presented in Tables 2 and 3. Before exposure to C. elegans, each type of MWCNTs was sonicated for 1 hour using a sonicator (Branson-5210; Branson Inc., Danbury, CT, USA) in distilled water. Starting from stock solutions, experimental concentrations of MWCNTs were prepared in K-medium (0.032 M KCl and 0.051 M NaCl) for C. elegans exposure. The shape and crystal structure of the MWCNTs were determined using a LIBRA 120 transmission electron microscope (TEM; Carl Zeiss, Oberkochen, Baden-Württemberg, Germany). The size distribution and zeta potential of MWCNTs in C. elegans K-media was measured using an electrophoretic light scattering spectrophotometer (ELS; ELS-8000; Otsuka Electronics, Osaka, Japan).

C. elegans Mortality Test

Wildtype and mutant C. elegans were exposed to pristine and OH-MWCNTs at a concentration of 500 mg/L in K-media with E. coli OP50 as food. Twenty young adult worms were transferred to 96-well tissue culture plates containing 100 μL of the test solution in each. The worms were exposed for 48 hours at 20°C.

Statistical Analysis

Data are presented in arbitrary units compared to the control, with statistical differences between the wildtype and mutants relating to survival determined by an analysis of variance test, with a Dunnett’s multiple comparison test. All statistical tests were performed with the SPSS version 12.0 KO (SPSS Inc., Chicago, IL, USA).

Results

Outer Diameter Size-dependent Toxicity

Physicochemical Properties

We examined the physicochemical properties of two different type of OH-MWCNTs (Table 2). The OH-MWCNTs had the same tube length (10 to 30 μm) with different OD sizes (Small-OD-MWCNTs, 8 to 15 nm; Large-OD-MWCNTs 20 to 30 nm). The Small-MWCNTs had a greater aspect ratio range (666.7 to 3750) than the Large-MWCNTs (333.3 to 1500). The shape and structure of the MWCNTs were monitored with TEM and their hydrodynamic diameter (HDD) was measured using ELS. Their HDD ranged from 5000 to 9000 nm in K-media. When two different OH-MWCNTs were dispersed in K-media, the HDD of the Small-MWCNTs (5000 to 7000 nm)

Table 2. Physicochemical properties of hydroxylated-multi walled carbon nanotubes (OH-MWCNTs) with different sized outer diameters (ODs)

| OD size of OH-MWCNTs (nm) | Tube length (μm) | Purity (%) | Aspect ratio | Zeta potential (mV) | Hydrodynamic diameters (nm) | Transmission electron microscope |
|--------------------------|------------------|------------|--------------|---------------------|---------------------------|---------------------------------|
| 8-15 (Small-MWCNTs)      | 10-30            | 95         | 666.7-3750   | -31.5               | 5000-7000                  | ![Image](image1.png) |
| 20-30 (Large-MWCNTs)     |                  |            | 333.3-1500   | -22.3               | 6000-9000                  | ![Image](image2.png) |
was less than that of the Large-MWCNTs (6000 to 9000 nm). Their dispersity was affected by the structural properties (OD and HDD) of OH-MWCNTs in aqueous phase. The zeta potentials of the MWCNTs, which indicate their ionic stability in aquatic media, were measured in K-media. The OH-MWCNTs had negative zeta potentials, -30.8 mV in the case of Small-MWCNTs and -22.3 mV in the case of Large-MWCNTs.

**Mechanism of Uptake and Toxicity**

To identify whether the OD sizes of the OH-MWCN affected their uptake and toxicity in *C. elegans*, we selected loss-of-function mutants of genes involved in uptake (i.e., *rme-1, -2, -8* [endocytosis], *eat-2* [pharyngeal pumping defect]) and representative toxicity (i.e., *sod-3, ctl-2* [oxidative stress], *cyt35a2* [xenobiotic metabolism], *pmk-1, daf-16* [DNA damage], *hif-1* [hypoxia response], *mtl-2* [metal stress], and *hsp-16.2* [general stress]) and their responses were compared.

The comparative toxicity of OH-MWCNTs with different ODs showed Small-MWCNTs was more toxic than Large-MWCNTs in *wildtype* *C. elegans* (Table 4). Interestingly a smaller HDD was found in K-media exposed to Small-MWCNTs than in media exposed to their Large counterpart (Table 2). Among uptake-related mutants, *rme-1 (b1045)* and *rme-8 (b1023)* tended to show a more sensitive response than *wildtype* in response to Small-MWCNTs exposure, while the toxicity of Big-MWCNTs did not affect. The toxicity by MWCNTs was completely rescued in *eat-2 (ad465)* mutant, which suggests that ingestion is the main route of MWCNTs uptake.

Exposure to Small-MWCNTs led to a decrease in survival for the toxic response related mutant, *pmk-1 (km25), sod-3 (gk235), cyt35a2 (gk317), cep-1 (gk138), mtl-2 (gk125), and hsp-16.2 (gk249)* comparing to exposure with Big-MWCNTs. Especially, the *sod-3 (gk235), cyt35a2 (gk317), and mtl-2 (gk125)* mutants were more sensitive than *wildtype* to Small-MWCNTs exposure.

**Table 3.** Physicochemical properties of hydroxylated-multi walled carbon nanotubes (OH-MWCNTs) with different tube lengths

| Tube length of OH-MWCNTs (μm) | 0.5-2.0 (Short-MWCNTs) | 8-15 | 10-30 (Long-MWCNTs) |
|--------------------------------|-------------------------|------|---------------------|
| Outer diameter size (nm)       | 8-15                    | 8-15 | 8-15                |
| Purity (%)                     | 95                      | 95   | 95                  |
| Aspect ratio                   | 33.3-250                | 666.7-3750 | 666.7-3750 |
| Zeta potential (mV)            | -31.5                   | -30.8| -30.8               |
| Hydrodynamic diameters (nm)    | 700-1200                | 5000-7000 | 5000-7000 |
| Transmission electron microscope|                         |      |                     |

**Tube Length Dependent Toxicity**

**Physicochemical Properties**

Table 3 described the properties of OH-MWCNTs with different tube length. OH-MWCNTs have a same OD size (8 to 15 nm) and different tube length (Short-MWCNTs, 0.5 to 2.0 μm; Long-MWCNTs, 10 to 30 μm). Long-MWCNTs have a higher aspect ratio ranges (666.7 to 3750) than Short-MWCNTs (33.3 to 250). The shape and structure of MWCNTs were monitored with TEM, the HDD was measured using ELS. When two different OH-MWCNTs were dispersed in K-media, HDD of Short-MWCNTs (700 to 1200 nm) was smaller than that Long-MWCNTs (5000 to 7000 nm). The zeta potentials of MWCNTs had negative zeta potentials, both MWCNTs were observed very similar zeta potentials, -31.5 mV and -30.8 mV.

**Mechanism of uptake and toxicity**

Comparing the toxicities of Short-MWCNTs and Long-MWCNTs showed that MWCNTs with a shorter tube length were more toxic than those with a longer tube length in *wildtype* *C. elegans* (Table 4). In terms of the uptake mechanism, the survival of the *eat-2 (ad465)* mutant increased when compared with *wildtype* exposed to Short-MWCNTs and Long-MWCNTs. This result suggests that the possible uptake route of MWCNTs is via ingestion. Among uptake-related mutants, *rme-1 (b1045)* and *rme-8 (b1023)* tended to show a more sensitive response than *wildtype* in response to Short- MWCNTs and Long-MWCNTs. Among toxic response related mutants, the *pmk-1 (km25)* and *cep-1 (gk138)* mutants were more sensitive than *wildtype* to
Table 4. Survival of wildtype and uptake and toxicity related mutants of *C. elegans* exposed to multi walled carbon nanotubes (MWCNTs) with different sized ODs and tube lengths

| Strain          | OD size of OH-MWCNTs (nm) | Tube length of MWCNTs (μm) |
|-----------------|---------------------------|---------------------------|
|                 | 8-15                      | 20-30                     |
|                 | 0.5-2.0                   | 10-30                     |
| Wildtype        | N2                        |                           |
| Uptake          |                           |                           |
| Endocytosis     | rme-1 (b1045)             | 79.5±11.3                 |
|                 | rme-2 (b1008)             | 96.1±18.2*                |
|                 | rme-8 (b1023)             | 53.0±19.5*                |
| Pharyngeal pumping defect | eat-2 (ad465) | 100.0±0.0                 |
| Toxicity        |                           |                           |
| Immune response | pmk-1 (km25)              | 66.6±31.6                 |
|                 | def-16 (mu86)             | 86.1±10.7                 |
| Oxidative stress| sod-3 (gk235)             | 50.2±23.8                 |
|                 | ctrl-2 (ok1137)           | 97.2±2.0                  |
| DNA damage      | cep-1 (gk130)             | 79.0±22.9                 |
| Xenobiotic metabolism | cyp35a2 (gk317) | 46.1±26.9                 |
| Hoxia response  | hif-1 (ia4)               | 96.1±4.8                  |
| Metal stress    | mtl-2 (gk125)             | 37.2±30.7                 |
| General stress  | hsp-16.2 (gk249)          | 66.1±24.8                 |

Values are presented as the mean± standard error compared to control (control=100, n=3) by two-tailed *t*-test. OH-MWCNTs, hydroxylated-MWCNTs; ODs, outer diameters. 
*From Eom HJ et al. Chem Biol Interact 2015;239:153-163 [33].
*p<0.05, **p<0.01.

Short-MWCNTs exposure, whereas sod-3 (gk235), cyp35a2 (gk317) and mtl-2 (gk125) mutants were more sensitive than wildtype to Long-MWCNTs exposure. This result showed that each mutant had a different sensitivity to MWCNTs of different lengths.

**Discussion**

Many studies have reported that endocytosis/phagocytosis is the cellular uptake mechanism of CNTs [31]. We investigated whether endocytosis was involved in the uptake mechanism of MWCNTs in *C. elegans* using endocytosis defective mutants such as rme-1,-2,-8. Receptor-mediated endocytosis (RME) is called clathrin-dependent endocytosis as the most common pathway and has been demonstrated to be the internalization mechanism of CNTs [31]. Unexpectedly, all three rme mutants showed a sensitive response to Short-MWCNTs and rme-1 (b1028) and rme-8 (b1023) tended to show a more sensitive response than wildtype in response to Long-MWCNTs (Table 4). Chen et al. [32] reported that amide-single walled CNTs inhibited endocytosis may result in decreased *C. elegans* growth due to reduced nutrient uptake. Thus, we suggest that endocytosis via the RME pathway may not be involved in the uptake of OH-MWCNTs. In addition, we previously demonstrated phagocytosis as the uptake mechanism of MWCNTs [33]. Finally, toxicity was completely rescued in a pharyngeal pumping mutant, eat-2 (ad465), which strongly suggests that ingestion is the main route of MWCNTs uptake.

We hypothesized that the OD and tube length of MWCNTs plays a role in the toxicity of MWCNTs. Only a few studies have evaluated the role of their diameter in the toxicity of CNTs [14,34]. Nagai et al. [35] compared the toxicity of two different MWCNTs that had similar hydrophobicity, surface reactivity and length, but different diameters (9.4 and 70 nm). They found that the thin MWCNTs were significantly more toxic than the thicker ones, both *in vitro* and *in vivo*. Another study also reported that thin MWCNTs showed cytotoxicity and inflammatory and carcinogenic properties. In the current study, the MWCNTs with a smaller OD appeared more toxic than the larger one in *wildtype C. elegans* (Table 4). Moreover, in the mutant test, the Small-MWCNTs significantly increased the mortality of sod-3 (gk235), cyp35a2 (gk317), and mtl-2 (gk249) mutants compared to wildtype, suggesting that the toxicity of MWCNTs with smaller OD sizes might be related to oxidative stress, xenobiotic metabolism, and metal stress. However, further more in depth study would be needed to understand the mechanism.

Regarding the length of MWCNTs, several studies have shown that CNTs with longer lengths and larger diameters have greater toxicity than smaller ones [34]. Another group has also shown that short CNTs do not result in damage to cells. Despite the fact that most studies have shown that longer and wider CNTs lead to greater toxicity, some researchers have found the opposite [36,37]. In the present study, we found the MWCNTs with a shorter tube length were more toxic to *wildtype C. elegans*; however, mutant analysis provided a hint that the Short- and Long-MWCNTs might exert their toxicities via distinct mechanisms (Table 4). For example, Short-MWCNTs decreased the survival of pmk-1 (km25) and cep-1 (138) mutants compared to...
wildtype, which represented genes in the mitogen-activated protein kinase pathway and DNA damage repair, respectively, while Long-MWCNTs affected the sod-3 (gk235), cyp35a2 (gk317), and mtl-2 (gk249) mutants. However, here again further study would be needed in order to understand the mechanisms involved.

Aspect ratio (length-to-diameter ratio) is known to affect MWCNT’s toxicity. We therefore investigated toxicity according to the aspect ratio. Many studies have reported that high aspect ratio (long and thin) MWCNTs show more toxicity than low aspect ratio MWCNTs [19,38]. Our results comparing OD sizes support this, as Small-MWCNTs, which had a high aspect ratio, were more toxic than their Large counterpart (Tables 2 and 4). However, the comparison of tube lengths showed the opposite, as the short MWCNTs, which had a low aspect ratio, were more toxic than the long one, which had a high aspect ratio (Tables 3 and 4). In Table 5 we summarize the relationship between aspect ratio and toxicity of CNTs observed here as well as reported previously by other groups. Taken together, CNTs with a high aspect ratio are generally more toxic than those with a low aspect ratio, but this was not always observed, probably due to other factors that affect CNTs toxicity.

Our mutant analysis provides an insight into the mechanism of uptake and toxicity, as we found that each mutant had a different sensitivity different MWCNTs. However, we cannot yet fully explain the variation in the responses of the different mutants. More evidence is needed to fully understand the mechanism of toxicity of MWCNTs.

In conclusion, we have demonstrated that decreasing the OD size and tube length of MWCNTs increased their toxicity to C. elegans, which suggests that diameter and length are important parameters to be considered in CNT-induced toxicity. This and other information will help in designing safer CNTs.

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Conflict of Interest

The authors have no conflicts of interest with material presented in this paper.

Table 5. Effect of aspect ratio on the toxicity of multi walled carbon nanotubes (MWCNTs)

| Carbon nanotube | Tube length (μm) | Diameter (nm) | Aspect ratio | System | Toxic endpoint | Toxicity | Reference |
|----------------|-----------------|---------------|--------------|--------|----------------|---------|-----------|
| MWCNT          | 0.1-1           | 9.4           | High         | MH-S cells | Cytotoxicity    | High    | 14        |
|                | 1-3             | 70            | Low          |        |                |         |           |
| MWCNT          | 0.1-1           | 9.4           | High         | Wistar rats | Inflammation   | High    | 14        |
|                | 1-3             | 70            | Low          |        |                |         |           |
| MWCNT          | 5-15            | 20-60         | High         | A549 cells | DNA damage     | High    | 34        |
|                | 1-2             | 60-100        | Medium       |        |                |         |           |
|                | 2-1             | <10           | Low          |        |                |         |           |
| MWCNT          | 5-15            | 20-60         | High         | C578L/6 mice | Inflammation | High    | 34        |
|                | 1-2             | 60-100        | Medium       |        |                |         |           |
|                | 2-1             | <10           | Low          |        |                |         |           |
| MWCNT          | 4.6±0.10        | 52.4±0.72     | High         | Human peritoneal Mesothelial cells | Cytotoxicity | High    | 35        |
|                | 4.88±0.10       | 116.2±1.6     | Low          |        |                |         |           |
| MWCNT          | 4.6±0.10        | 52.4±0.72     | High         | Rat | Carcinogenicity | High    | 38        |
|                | 4.88±0.10       | 116.2±1.6     | Low          |        |                |         |           |
| MWCNT          | 10              | 10-15         | High         | CHO-K1 cells | Cytotoxicity | High    | 36        |
|                | 0.15            | 10-16         | Low          |        |                |         |           |
| BSA-MWCNTs     | 0.8±0.5         | 19.9±8.25     | High         | Zebrafish | Developmental toxicity | High    | 39        |
|                | 0.2±0.1         | 19.9±8.25     | Low          |        |                |         |           |
| AT-MWCNT       | 77±31           | 17±6          | High         | Escherichia coli | Toxicity assays | Low    | 40        |
| f-MWCNT        | 4.1±3.7         | 19.7          | Low          | K12 |                |         |           |
| MWCNTs         | 10-30           | 8-15          | High         | C. elegans | Mortality | High    | This study |
|                | 20-30           | Low           |          |                |         |           |
| MWCNTs         | 10-30           | 8-15          | High         | C. elegans | Mortality | Low     | This study |
|                | 0.5-20          | 8-15          | Low          |                |         |           |

BSA-MWCNTs, bovine serum albumin functionalized MWCNTs; AT-MWCNT, acid treated MWCNT; f-MWCNT, functionalized MWCNT; MH-S, murine alveolar macrophages; A549 cells, human alveolar carcinoma epithelial cells; CHO-K1, Chinese hamster ovary cells.
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