Case Report: ANCA-Associated Vasculitis Presenting With Rhabdomyolysis and Pauci-Immune Crescentic Glomerulonephritis After Pfizer-BioNTech COVID-19 mRNA Vaccination

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As the coronavirus disease 2019 (COVID-19) pandemic is ongoing and new variants of severe acute respiratory syndrome coronavirus type 2 (SARS-CoV-2) are emerging, there is an urgent need for COVID-19 vaccines to control disease outbreaks by herd immunity. Surveillance of rare safety issues related to these vaccines is progressing, since more granular data emerge with regard to adverse events of COVID-19 vaccines during post-marketing surveillance. Interestingly, four cases of anti-neutrophil cytoplasmic antibody (ANCA)-associated vasculitis (AAV) presenting with pauci-immune crescentic glomerulonephritis (GN) after COVID-19 mRNA vaccination have already been reported. We here expand our current knowledge of this rare but important association and report a case of AAV presenting with massive rhabdomyolysis and pauci-immune crescentic GN after Pfizer-BioNTech COVID-19 mRNA vaccination. As huge vaccination programs are ongoing worldwide, post-marketing surveillance systems must continue to assess vaccine safety important for the detection of any events associated with COVID-19 vaccination. This is especially relevant in complex diseases where diagnosis is often challenging, as in our patient with AAV presenting with massive rhabdomyolysis and pauci-immune crescentic GN.

Keywords: coronavirus disease 2019 (COVID-19), vaccination, anti-neutrophil cytoplasmic antibody (ANCA), ANCA-associated vasculitis (AAV), rhabdomyolysis, acute kidney injury (AKI), pauci-immune crescentic glomerulonephritis (GN)
INTRODUCTION

As the coronavirus disease 2019 (COVID-19) pandemic is ongoing and new variants of severe acute respiratory syndrome coronavirus type 2 (SARS-CoV-2) are emerging, there is an urgent need for COVID-19 vaccines to control disease outbreaks by herd immunity (1). The use of novel vaccines containing a nucleoside-modified messenger ribonucleic acid (mRNA) or a viral deoxyribonucleic acid (DNA) vector that encodes the viral spike (S) glycoprotein of SARS-CoV-2 has already been approved. Large clinical trials have shown that these COVID-19 vaccines are safe and effective. Common adverse events include mild to moderate reactions at the injection site, fever, fatigue, body aches, and headache (2). Surveillance of rare safety data emerge with regard to adverse events of COVID-19 vaccines during post-marketing surveillance (3). Anti-neutrophil cytoplasmic antibody (ANCA)-associated vasculitis (AAV) is a small vessel vasculitis hallmarked by the presence of antibodies against autoantigens in cytoplasmic granules of neutrophils (4). AAV presents as granulomatosis with polyangiitis (GPA), microscopic polyangiitis (MPA), and eosinophilic granulomatosis with polyangiitis (EGPA) (5). Generally, renal manifestations in AAV are estimated at 80% among all cases mainly manifesting as ANCA-associated glomerulonephritis (ANCA GN), and the overall prevalence does not seem to differ substantially between MPO-ANCA and PR3-ANCA AAV (6). Interestingly, five cases of renal AAV presenting with pauci-immune crescentic ANCA GN after COVID-19 mRNA vaccination have already been reported (7–10). We here expand our current knowledge of this rare but important association and report a case of AAV presenting with massive rhabdomyolysis and pauci-immune crescentic GN after Pfizer-BioNTech COVID-19 mRNA vaccination.

CASE REPORT

A 79-year-old Caucasian female with a past medical history of hypertension, degenerative disc disease, and no documented history of COVID-19 received two doses of Pfizer-BioNTech COVID-19 mRNA vaccination. Two weeks thereafter, the patient presented to our emergency department with weakness and upper thigh pain. Vital parameters were stable, and physical examination was unremarkable. The patient had no allergies and denied illicit drug use. External routine laboratory assessments obtained 1 week prior to admission were normal for serum creatinine of 0.71 mg/dl (reference range: 0.5–0.95), estimated glomerular filtration rate (eGFR) of 84.4 ml/min/1.73 m², and urinalysis with the absence of hematuria or proteinuria. Repeat reverse transcription polymerase chain reaction (RT-PCR) testing for SARS-CoV-2 RNA from nasopharyngeal swabs was negative. Laboratory assessments at admission showed massive rhabdomyolysis with creatinine kinase (CK) levels of 14,243 U/L (reference range: 29–168), myoglobinemia of >12,000 μg/L (reference range: ≤106, Figure 1A), and acute kidney injury (AKI) with serum creatinine levels of 1.38 mg/dl (reference range: 0.7–1.2, Figure 1B) and an estimated glomerular filtration rate (eGFR) of 33.5 ml/min/1.73 m². Urinary analysis revealed leukocyturia, hematuria (no dysmorphic erythrocytes), few renal tubular epithelial cells, and nephrotic range proteinuria of >18,000 mg/g creatinine and albuminuria of <5,000 mg/g creatinine (reference range: <30 mg/g, Figure 1C). The patient received intravenous crystalloids with decreasing CK levels and myoglobinemia (Figure 1A). However, progressive deterioration of kidney function with worsening of serum creatinine levels up to 6.57 mg/dl (reference range: 0.7–1.2 mg/dl, Figure 1B) and an eGFR of <15 ml/min/1.73 m² occurred. ANCA immunofluorescence (IF) was positive at 1:1,000 (reference range: <1:10) with elevated MPO-ANCA levels >134 IU/ml (reference range: <3.5 IU/ml), while myositis antibodies, complement levels, and other serologic parameters were all tested negative (Table 1). Because of leukocytosis, a white blood differential was conducted revealing prominent peripheral blood eosinophilia (Table 1).

Based on suspected MPO-positive AAV, the patient received a steroid pulse with intravenous methylprednisolone for 3 days (250 mg per day) and oral prednisone 1 mg/kg daily thereafter (60 mg per day, Figure 1D). A kidney biopsy confirmed severe acute tubular injury with pauci-immune crescentic GN and interstitial nephritis: cellular crescents in 1/15 (6.7%) glomeruli, global glomerular sclerosis in 2/15 (13.3%), mild (5%) interstitial fibrosis and tubular atrophy (IF/TA), interstitial inflammation (25%) with prominent eosinophilic infiltration, and severe acute tubular injury with myoglobin casts (Figure 2 and Table 2). According to histopathological scoring, focal class ANCA GN and intermediate risk ANCA renal risk score (ARRS) were present (Table 2) (11, 12). Based on the diagnosis of AAV presenting with massive rhabdomyolysis and pauci-immune crescentic GN, intravenous cyclophosphamide (CYC) was initiated at 10 mg/kg (per CYCLOPS trial dosing, Figure 1D) (13). Thereafter, kidney function normalized without requirement of dialysis and proteinuria decreased to 1,603 mg/g creatinine and albuminuria to 351 mg/g creatinine (reference range: <30 mg/g, Figures 1B, C). Repeat serological testing confirmed that ANCA IF turned negative. Thereafter, oral prednisone was tapered down (currently 50 mg per day), and we do not plan to repeat administration of intravenous cyclophosphamide because rhabdomyolysis ceased and kidney function normalized.

DISCUSSION

To our knowledge, we here present the first case of AAV presenting with massive rhabdomyolysis and pauci-immune crescentic GN after Pfizer-BioNTech COVID-19 mRNA vaccination. With millions of people being vaccinated for COVID-19, rare reports of adverse events are emerging. In this case, the temporal association between Pfizer-BioNTech
FIGURE 1 | Timeline of the case after admission. (A–C) Time course of CK, myoglobin, plasma creatinine, and levels of uPCR and uACR. (D) Time of treatment regimens and kidney biopsy. CK, creatinine kinase; CYC, cyclophosphamide; uACR, urinary albumin-to-creatinine ratio; uPCR, urinary protein-to-creatinine ratio.
COVID-19 mRNA vaccination and AAV presenting with rhabdomyolysis and pauci-immune crescentic GN suggests a neutrophilic immune response to mRNA as a potential trigger. This patient initially presented with upper thigh pain due to massive rhabdomyolysis after Pfizer-BioNTech COVID-19 mRNA vaccination. Rhabdomyolysis has been described in the context of COVID-19, and a direct viral tropism to myocytes has been suggested (14, 15). However, detection of SARS-CoV-2 infection in skeletal muscle cells has not been established yet (16). Rhabdomyolysis secondary to vaccination has previously been reported, mostly in the context of influenza vaccination (17, 18). In association with COVID-19 mRNA vaccination, the onset of fatigue, myalgias, and arthralgias following mRNA vaccination has been reported in a considerable subset of patients (19). Additionally, there is evidence that COVID-19 mRNA vaccination can directly induce myositis at the injection site, as previously observed in the deltoid muscle (20). In addition to rhabdomyolysis, we observed pauci-immune crescentic GN accompanied by MPO-ANCA autoantibodies after COVID-19 mRNA vaccination. To date, five cases of pauci-immune crescentic ANCA GN after the second dose of COVID-19 mRNA vaccination in all cases have been reported (7-10). In our case, kidney biopsy showed myoglobin casts due to massive rhabdomyolysis and pauci-immune crescentic GN, and it is likely that both contributed to deterioration of kidney function. It has already been reported that the first COVID-19 mRNA vaccination primes the innate immune system to mount a more potent response after the second booster immunization (21). It is possible that the enhanced immune response especially observed after the second dose of COVID-19 mRNA vaccination could be responsible for triggering the observed MPO-ANCA autoantibodies. Causal links between immune system activation by viral infections and AAV have been suggested due to onset of AAV predominantly during the winter (22, 23). Toll-like receptors (TLRs) are expressed on leukocytes and play crucial roles in the recognition of viral antigens, facilitating immune system activation and inflammation. Predominant TLR-2 and TLR-9 activation can stimulate autoimmunity in AAV, previously been described in the context of MPO-ANCA autoantibodies (24). Interestingly, TLR-2 activation in immunodominant cytotoxic T lymphocytes in response to SARS-CoV-2 S glycoprotein (as also produced by COVID-19 vaccines) has already been described (25). With regard to vaccination, there is some discussion about the relationship between vaccination and AAV recurrence in patients with pre-existing autoimmune disease after influenza vaccination as very rare but significant side effects (26). The temporal relationship could be explained theoretically, including molecular mimicry, polyclonal activation, or transient systemic proinflammatory cytokine responses that potentially provoke autoimmune diseases in genetically predisposed individuals (27). Interestingly, increased production of ANCA autoantibodies has already been described in response to viral mRNA-based influenza and rabies vaccines (27). Moreover, AAV and autoimmune reactions have been reported in the context of COVID-19, implicating a direct reaction to viral RNA (28-30). Therefore, the occurrence of AAV in the context of COVID-19 mRNA as compared with non-mRNA vaccines would be of great relevance. Huge vaccination programs are ongoing worldwide, and post-marketing surveillance systems must continue to assess vaccine safety important for the detection of any events associated with COVID-19 vaccination. This is especially relevant in complex diseases where diagnosis is often challenging, as in our patient with AAV presenting with massive rhabdomyolysis and pauci-immune crescentic GN. The limitation of this case report is only a temporal relationship between COVID-19 mRNA vaccination and onset of AAV. However, AAV onset in association with COVID-19

### Table 1: Serologic parameters after admission.

| Parameter                     | Value      | Reference range |
|-------------------------------|------------|-----------------|
| HIV Ag/Ab—titer              | Neg        | Neg             |
| HBsAg—titer                  | Neg        | Neg             |
| Anti-HCV—titer               | Neg        | Neg             |
| Rheumatoid factor—IU/ml      | <10        | <15            |
| Complement C3c—g/L           | 0.97       | 0.82–1.93       |
| Complement C4—g/L            | 0.19       | 0.15–0.67       |
| ANCA IF                       | 1:1,000    | <1:100          |
| PR3-ANCA—IU/ml               | <0.2       | <2             |
| MPO-ANCA—IU/ml               | >134       | <3.5            |
| ENA screen                   | <0.1       | <0.7            |
| Anti-DFS70—U/ml              | <0.6       | <7             |
| Anti-ds-DNA—IU/ml            | 4.4        | <15            |
| Histones—U/ml                | 7.5        | 20             |
| ANA IF                        | 1:320      | <1:100          |
| ROS2—blot                    | Neg        | Neg             |
| PM-Scl-100—blot              | Neg        | Neg             |
| PM-Scl-75—blot               | Neg        | Neg             |
| Ku—blot                      | Neg        | Neg             |
| SRP—blot                     | Neg        | Neg             |
| PL-7—blot                    | Neg        | Neg             |
| PL-12—blot                   | Neg        | Neg             |
| EJ—blot                      | Neg        | Neg             |
| OJ—blot                      | Neg        | Neg             |
| JO1—blot                     | Neg        | Neg             |
| Mi alpha—blot                | Neg        | Neg             |
| Mi-2 beta—blot               | Neg        | Neg             |
| TIF1 gamma—blot              | Neg        | Neg             |
| MDA-5—blot                   | Neg        | Neg             |
| NXP2—blot                    | Neg        | Neg             |
| SAE1—blot                    | Neg        | Neg             |
| White blood differential      |            |                 |
| Leukocytes—1,000/µl          | 22.9       | 4–11            |
| Lymphocytes—%                | 4.7        | 20–45           |
| Monocytes—%                  | 4.5        | 3–13             |
| Eosinophils—%                | 23.3       | ≤5              |
| Basophils—%                  | 0.2        | ≤2              |
| Neutrophils—%                | 67.3       | 40–76           |

ANA, antinuclear antibodies; ANCA, anti-neutrophil cytoplasmic antibody; ds-DNA, double stranded-DNA; DSF70, dense-fine-spectled 70; ENA, Ej, EJ, IF 1; KU, Ku; MPO, myeloperoxidase; Neg, negative; NXP2, nuclear matrix protein 2; OJ, isoleucine; PM-Scl, PL-7, threonine; PL-12, alanine, polymyositis-scleroderma; PR3, proteinase 3; SAE1, small ubiquitin-like modifier activating enzyme; SRP, signal recognition particle; TIF1, transcriptional intermediary factor 1.
mRNA vaccination has independently been observed before and requires further investigation with regard to the mechanisms linking autoimmunity to COVID-19 vaccines (7–9). Fortunately, treatment of AAV is possible and caution in such cases is warranted with regard to early testing if clinical symptoms are compatible with AAV in principle.

**DATA AVAILABILITY STATEMENT**

The original contributions presented in the study are included in the article/supplementary material. Further inquiries can be directed to the corresponding author.

**ETHICS STATEMENT**

Ethical review and approval was not required for the study on human participants in accordance with the local legislation and institutional requirements. The patients/participants provided their written informed consent to participate in this study.

**AUTHOR CONTRIBUTIONS**

BT was directly involved in the treatment of the patient, conceived the case report, collected and analyzed the data, and wrote the manuscript. SH evaluated kidney biopsy findings and edited the manuscript. All authors contributed to the article and approved the submitted version.

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