New Data on Variously Directed Dose-Response Relationships and the Combined Action Types for Different Outcomes of in Vitro Nanoparticle Cytotoxicity

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Abstract
Spherical selenium-oxide and copper-oxide nanoparticles (SeO-NP with mean diameter 51 ± 14 nm and CuO-NP with mean diameter 21 ± 4 nm) were found to be cytotoxic for human fibroblast-like cells in vitro, as judged by decreased ATP-dependent luminescence. Compared with SeO-NP, CuO-NP produced a somewhat stronger effect of this kind. Along with cell hypertrophy developing in response to certain doses of SeO-NP and CuO-NP, our experiment also revealed doses causing a decrease in cell and cell-nucleus sizes. We observed both monotonic and different variants of nonmonotonic dose-response relationship. For the latter, we have succeeded in constructing adequate mathematical expressions based on the generalized hormesis paradigm that we had considered previously in respect of CdS-NP and PbS-NP cytotoxicity for cardiomyocytes. It was demonstrated as well that combined toxicity of SeO-NP and CuO-NP is of different types depending on the outcome.

Keywords
nanoparticles, selenium-oxide, copper-oxide, dose-response relationships, hormesis

Introduction
Previously, we demonstrated\textsuperscript{1} that dose-response relationships in lead sulfide and cadmium sulfide nanoparticles (PbS-NP and CdS-NP) cytotoxicity studied on a culture of cardiomyocytes could be monotonic for some effects and nonmonotonic for others. The relationship of the first type means that as the impact intensifies, the effect grows stronger or weaker in the entire range of doses. In the relationship of the second type, the direction of the effect changes at least twice with growth in impact, and it may be termed as hormesis in its broader sense considered in detail in.\textsuperscript{2,3} In these publications of ours, one can find a sufficiently comprehensive review the relevant scientific literature dealing with the hormesis paradigm.

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Besides, it was demonstrated that the type of combined action of the above nanoparticles depends on the toxicity outcome it is characterized for.

The purpose of this paper is to demonstrate that the above features of dose-response relationship are not a special case which holds only for the above nanoparticle species and cell type. To this end, we have analyzed results of experiments on a culture of fibroblast-like cells exposed to selenium-oxide (SeO-NP) or copper-oxide (CuO-NP) nanoparticles.

Materials and Methods

Preparation and Characterization of Nanoparticles

Suspensions of nanoparticles (NP) were prepared by laser ablation of 99.9% pure CuO and SeO targets in deionized water and visualized under a scanning electron microscope (SEM), Merlin (Carl Zeiss, Germany). The size distribution functions were obtained by a statistical analysis of the SEM images of several hundreds of respective NPs. The chemical nature of these NPs was characterized using the energy-dispersive X-ray spectroscopy.

Cell Line Characterization, in vitro Exposure Technique and Parameters, Cytotoxicity Estimates

Cell Cultures. The experiments were performed on a FELCH-104 stable cell line from BioloT Ltd. (Saint-Petersburg, Russia), which presents a culture of fibroblast-like cells derived from an 8-week human embryo. The cell culture was kept at 37°C under a 5% CO₂ atmosphere in a DMEM medium containing L-glutamine, 1 g/L glucose, 10% embryonic bovine serum, and .5% gentamicin. We waited till a monolayer has been formed having investigated the preparation under inverted microscope each 24 hours after seeding the cells. For assessing NP cytotoxicity, cells were seeded in a 96-well plate (TPP Techno Plastic Products AG, Trasadingen, Switzerland), 70 000 cells per well in 100 mcL medium, and maintained under standard conditions for 48 hours until a monolayer was obtained. Then suspensions of CuO and SeO nanoparticles (diluted to a needed concentration with DMEM medium) were added to the wells and incubated for 24 hours under standard conditions before performing an ATP assay. The final concentration of each NP type in the medium was 25-50-100 mcg/mcL. Nanoparticles (ether of one species or their combination) were added to the culture medium in these concentrations.

Cytotoxicity Effects Assessment. To estimate quantitatively cytotoxic effects produced by the nanoparticle species in the concentrations used, we determined the ATP content of the culture by the luminescent signal and measured cell and cell-nucleus size.

The Bioluminescence Assay

An ATP bioluminescence assay was performed using CellTiter-Glo reagents (Promega Corporation, USA). A working solution was obtained by reconstituting lyophilized CellTiter-Glo Substrate in CellTiter-Glo Buffer and warmed up to room temperature on a water bath. A 100 mcL portion of this solution was added to each well, and then the plate was rotated in one plane for 2 minutes to cause cell lysis. Upon incubation for 10 minutes at room temperature, we measured cell luminescence using an LM-01T luminometer with Kilia software (Immunotech, Beckman Coulter Company, Praha, Czech Republic). The measurement results were presented in relative luminescence units (RLU).

Cell Size Estimation

For morphometric study, we used microphotographs of cells suspended in the DMEM. To this end, 10 mcL of suspension were put on the microscope slide. In this way, 10 samples for each experimental condition were prepared and 8–10 cells in each sample were measured. For size measurements, the cells were removed from the plate and transferred onto a glass slide to perform morphometry under an optical microscope, 3D Cell Explorer (Nanolive, Switzerland). The professional image processing program ImageJ 1.48v (by Wayne Rasband, National Institutes of Health, USA) was used to measure the cell and cell-nucleus area in mc㎡.

The assaying techniques used are described in more detail in our previous publication.

Mathematical Description of the Experimental Results

In order to identify the type of dose-response relationship, experimental data for a particular outcome index should be approximated with an appropriate functional expression. However, the choice of approximating functions is not uniquely determined even where the dependence of the response on agent dose is monotonic. The problem becomes even more complex where the dependence is nonmonotonic.

Monotonic dose-response relationships are often described with the Hill function (1), proportional to the cumulative function of the log-logistic distribution

\[ y = b_0 + \frac{b_1}{1 + (b_2x)^{b_3}} \]  \hspace{1cm} (1)

where \( b_0, \ldots, b_3 \) are parameters determined with the least squares method by experimental data. Here, variable y represents the outcome and the variable x is the acting agent doses.

Another mathematical model for monotonic dependence presents a hyperbolic function (2) associated with the Michaelis–Menten equation, which is used, for example, to describe the rate of enzymatic reactions.
Here, variable y also represents the outcome and the variable x is the acting agent doses.

However, often, these functions are not enough to ensure good approximation of experimental data even in the case of monotonic dose-response dependence.

It is still more difficult to find a suitable mathematical model for a nonmonotonic dose-response relationship. Although there are lots of functional expressions for describing relationships of this type, none of them is universal, being confined to a certain limited area of research only.

At the same time, nonmonotonic relationship is often associated with manifestations of hormesis in cases where the effect is directed oppositely in 2 adjacent intervals of doses. This renders the theoretical generalization of different variants of a nonmonotonic dose-response dependence a lot more complex problem since this type of dependence can feature more than 2 phases of this kind.1,2,8,10 The objective of approximating such multiphase relationships of a hormesis-associated type becomes a difficult challenge which requires using more complex power1,2 or special functions.3

Like in the previous studies (for instance and many others), the type of combined cytotoxicity was estimated in a model based on the Response Surface Methodology (for example). In this methodology, equation (3) describing the response surface

\[ Y = f(x_1, x_2) \]

where \( Y \) is a quantitative effect (outcome) of a toxic exposure; \( x_1 \) and \( x_2 \) are the doses of the toxicants participating in the combination; \( f(x_1, x_2) \) is a regression equation with some numeric parameters. By virtually dissecting the response surface (3) at several model exposure levels, one obtains isoboles visually characterizing the type of two-factor combined action.

Today, the RSM is one of the most important general methods used in the analysis of combined effects produced by mixtures of bioactive substances, including toxic ones. This method enables the potentialities of effective experimental design to be used for approximating a response function. Constructing such approximation requires choosing an analytical model whose parameters would be determined by fitting to experimental data using the ordinary least squares method.

The quality of approximation by the proposed models was served experimental values well. However, often these functions are not enough to ensure good approximation of experimental data even in the case of monotonic dose-response dependence.

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The quality of approximation by the proposed models was served experimental values well.
Figure 1. SEM visualization of (A) SeO-NPs and (B) CuO-NPs (left panels) and respective size distribution functions (right panels).

Figure 2. Approximation by model
\[ y = 3.703 + 2.119 \sin(0.025x) - 0.200 \sin(0.035x) + 0.781 \sin(0.045x) + 0.822 \sin(0.055x) - 1.279 \sin(0.065x) \] of the reduction in ATP-dependent luminescence in the cell culture under exposure to various doses of SeO-NP. The abscissa plots the acting concentrations of nanoparticles in the incubation medium, mcg/mL; the ordinate plots the RLU values. The dots indicate the mean values with the standard error of the mean.
Figure 3. Approximation by model $y = 3.706 - (0.012x + 2.488/1 + e^{0.269(25 - x)})$ of the reduction in ATP-dependent luminescence in the cell culture under exposure to various doses of CuO-NP. The abscissa plots the acting concentrations of nanoparticles in the incubation medium, mcg/mL; the ordinate plots the RLU values. The dots indicate the mean values with the standard error of the mean.

Figure 4. The approximation of the cell area by the model $y = 144.57 - 35.06 \sin(3x/50) - 21.44 \sin(7x/100) - 9.08 \sin(2x/25) - 5.23 \sin(9x/100) - 2.03 \sin(x/10)$ in the cell culture under exposure to various doses of SeO-NP. The abscissa plots the acting concentrations of nanoparticles in the incubation medium, mcg/mL; the ordinate plots the values of the cell area in mcm$^2$. The dots indicate the mean values with the standard error of the mean.
In a similar assessment, the dependence of cell-nucleus area on dose for both SeO and CuO nanoparticles proved to be of the same type, a three-phase one, which is likely to reflect the dissimilarity of the mechanisms responsible for the increase in cell and cell-nucleus size.

The quality of approximation by the proposed models of dose-response relationships for both luminescence and morphometric indices, as estimated by the coefficient of determination (see Mathematical Description of the Experimental Results), proved to be quite high for all models (both usual and adjusted coefficients were no lower than .6). The evaluation of the statistical significance of the model parameters also shows their high significance ($P < .001$). In addition, it is obvious from the presented Figures 6 and 7 that the proposed dose-response models describe the observed experimental values well.

It should be noted that in a study involving exposure to CdS and PbS nanoparticles, the dose-dependent suppression of cell viability as measured by ATP-luminescence was also monotonic, while the dependence of the morphometric characteristic proved to be nonmonotonic by the hormesis-associated type despite the fact that two studies of ours involved NPs of different compositions and cells of essentially different type (cardiomyocytes and fibroblasts). We do not yet have sufficient experimental data to regard this coincidence as a regular feature and make suggestions concerning its mechanisms. However, it would be reasonable to make a mental note of it and revisit this issue when new data become available.

At the same time, it should be stressed that, even if the cell hypotrophy may seem an expected and better understood effect of cytotoxic impacts, it is not for the first time that we found that it was typical only for certain dose ranges while under the impact of other doses, on the contrary, the cell hypertrophy was evident. In particular, we observed the same in our in vitro experiments on cardiomyocytes under toxic impacts of CdS-NP and PbS-NP. The possible molecular or other mechanisms of this paradox are no more clear than the mechanisms of hormesis phenomenon in general and need special investigation for each particular response. Let us mention in this connection that, as it was many times demonstrated long ago (e.g., 15, 16) the products of macrophages breakdown caused by different agents have both in vivo and in vitro a stimulating effect on these cells differentiation and functions, so that a resulting response can be directed oppositely depending on a balance between a damage and its compensation.

In the same vein, the data that we have gained so far enable us to suggest that the type of dose-response relationship, whether monotonic or nonmonotonic, is not predetermined by the cell type or chemical nature of the impacting nanoparticles; rather, it depends on the impact’s outcome assessed as a response. The same can be said about the studied nanoparticles’ combined action type.
Figure 6. Approximation of cell-nucleus area by the model
\[ y = 34.60 - 8.08 \sin(3x/100) - 1.76 \sin(x/25) + 0.65 \sin(x/20) - 4.13 \sin(3x/50) - 9.21 \sin(7x/100) \] in the cell culture under exposure to various doses of SeO-NP. The abscissa plots the acting concentrations of nanoparticles in the incubation medium, mcg/mL; the ordinate plots the values of the cell area in mcm.² The dots indicate the mean values with the standard error of the mean.

Figure 7. Approximation of cell-nucleus area by the model
\[ y = 34.60 - 18.98 \sin(3x/50) - 4.78 \sin(7x/100) + 1.90 \sin(2x/25) + 2.39 \sin(9x/100) - 1.49 \sin(x/10) \] in the cell culture under exposure to various doses of CuO-NP. The abscissa plots the acting concentrations of nanoparticles in the incubation medium, mcg/mL; the ordinate plots the values of the cell area in mcm.² The dots indicate the mean values with the standard error of the mean.
Modeling of Combined Action

As may be seen from isoboles presented by the Figure 8 ATP-dependent luminescence, the combined action type proved the same as in the case of CdS-NP plus PbS-NP combination.1 Meantime, for the same combination, this type assessed for a morphometric index was different as compared with the present study.

In general, if considered together with our previously described experiments, which gave principally similar results, this study suggests once more that: (a) dose-response relationships for one and the same toxic agent but for different outcomes can be of both monotonic and nonmonotonic type, the latter corresponding to the hormesis paradigm in its generalized form; (b) a diversity of types of joint action characteristic of one and the same pair of toxic agents is one of the important assertions of the general theory of combined toxicity.

Conclusions

(1) Selenium-oxide and copper-oxide nanoparticles when acting on fibroblast-like cells in vitro display cytotoxicity manifesting itself as a decrease in ATP-dependent luminescence, CuO-NP producing a somewhat stronger effect than SeO-NP.

(2) Along with cell hypertrophy under the action of certain doses of SeO-NP and CuO-NP, our experiment also revealed doses causing a decrease in cell and cell-nucleus size.

(3) We obtained both monotonic and different variants of nonmonotonic dose-response relationships, and for the latter we managed to construct adequate mathematical expressions based on the generalized hormesis paradigm considered by us previously in respect of the cytotoxicity of CdS-NP and PbS-NP for cardiomyocytes.

(4) In general, the analysis of data obtained in the above-considered studies enable us to suggest that variability of dose-response relationship types displayed by different nanoparticle cytotoxicity effects is a common rule not accidental which makes it worthy of collecting further experimental evidence for establishing it as a common nanotoxicological rule.

(5) It was demonstrated as well that combined toxicity of SeO-NP and CuO-NP is also of different types depending on the outcome for which it is assessed.

Declaration of Conflicting Interests

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Figure 8. Isobolograms characterizing the combined toxic action of CuO-NP and SeO-NP on an fibroblast-like cell culture as estimated by its effects on (A) ATP-dependent luminescence (additivity), (B) cell area and (C) cell nuclear area (superadditivity). Numbers at the axes are respective NP concentrations in mcg/mL; numbers at the isoboles are the values of the effect to which they correspond.
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