REVIEW

Selective phytochemicals targeting pancreatic stellate cells as new anti-fibrotic agents for chronic pancreatitis and pancreatic cancer

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Received 3 August 2019; received in revised form 23 September 2019; accepted 29 October 2019

KEY WORDS
Pancreatic stellate cells; Anti-fibrotic; Chronic pancreatitis;

Abstract Activated pancreatic stellate cells (PSCs) have been widely accepted as a key precursor of excessive pancreatic fibrosis, which is a crucial hallmark of chronic pancreatitis (CP) and its formidable associated disease, pancreatic cancer (PC). Hence, anti-fibrotic therapy has been identified as a novel therapeutic strategy for treating CP and PC by targeting PSCs. Most of the anti-fibrotic agents have been...
1. Introduction

Pancreatic stellate cells (PSCs), which are star-shaped fibroblasts, were only identified and characterized 20 years ago, despite research on stellate cells having begun in the eighteenth century \(^1,2\). PSCs are responsible for the synthesis and degradation of extracellular matrix (ECM) proteins, such as tissue inhibitors, matrix metalloproteinases (TIMPs), and metalloproteinases (MMPs). Thus, PSCs can regulate the pancreatic tissue functions and maintain the normal architecture of the pancreas by balancing fibrogenesis and the matrix degradation process\(^3\). They comprise about 4% of the local cells in the pancreas and are found in the periarcinar and interlobular spaces\(^3\). Furthermore, they play a pivotal role in the development of a desmoplastic reaction (a reaction associated with tumors that is characterized by the growth of dense fibrous or connective tissues around a tumor), which is the hallmark of chronic pancreatitis (CP) and pancreatic cancer (PC)\(^4\).

Quiescent PSCs are activated by pancreatic injury or inflammation to become myofibroblast-like cells, expressing alpha-smooth muscle actin (alpha-SMA) and various ECM proteins, growth factors, and cytokines through structural and functional changes\(^5\). Pancreatic damage and inflammation could expose PSCs to a variety of soluble factors, which act as regulators of PSCs activation, as evidenced by several in vitro studies. These factors are interleukin-1 (IL-1), IL-6, tumor necrosis factor-alpha (TNF-alpha), platelet-derived growth factor (PDGF), transforming growth factor (TGF-beta), activin A, and ethanol and its metabolites, which cause oxidative stress and pressure, as well as extensive changes in the composition and components of ECM\(^6\). Continuous PSC activation occurs if the inflammation and injury are perpetuated; for example, if the inflammation and injury are limited, PSCs might proceed to apoptosis or deactivate, so that fibrosis will not develop. Repeated and persistent pancreatic injury and inflammation are key to the initiation of fibrogenesis\(^5\). Furthermore, cigarette smoke components (cigarette smoke extract (CSE) and/or nicotine (NNK)) in clinically relevant concentrations can activate PSCs, as evidenced by increased migration, proliferation, and collagen production in the presence or absence of ethanol. Ultimately, these findings suggest that the development of alcoholic pancreatic fibrosis could be caused by the combined effects of alcohol and cigarette smoke components via PSCs’ mediation\(^7\). The comparative roles of PSCs in PC progression are clear. PSCs possess adequate capacity to interact with cancer cells and other stromal cells in order to multiply the stromata and promote the cancer progression. Activated PSCs play important roles in PC, including producing ECM proteins and regulating the formation of desmoplastic reaction, as well as promoting cancer cell proliferation, migration, invasion, angiogenesis, and chemoresistance\(^8\). In addition, PSCs stimulate angiogenesis, which is important for tumor growth and metastasis, disruption of the antitumor immune system, and indirect induction of immune cell dysfunction\(^9\), causing conventional chemotherapy resistance and severe treatment failure. Furthermore, chemoresistance in PC cells is caused by various molecular mechanisms, including epigenetics, post-translational modifications, altered key signaling pathways, epithelial–mesenchymal transition (EMT), and the involvement of cancer stem cells and the cellular and non-cellular components of the tumor microenvironment\(^8,10\). Survival rates in PC are only minimally increased, due to the poor responsiveness of pancreatic tumors to chemotherapy and radiation therapy, affected and regulated by the molecular targets (e.g., mucin 1 (MUC1))\(^11,12\). Furthermore, PSCs’ role in radiotherapy resistance, by activating the integrin-focal adhesion kinase (FAK) signaling in PC cells, has been confirmed\(^13\).

Targeting stroma cells, particularly PSCs, in conjunction with chemotherapy has become the focus of current PC treatment and the two interventions may reveal potent anticancer activities when they are used concurrently. The current chemotherapeutic regimes for CP and PC remain inadequate, and the more recent treatment protocols under trial are costly and toxic\(^14\). Therefore, it is urgently necessary to search for, and identify, an alternative, more economical treatment with lower toxicity to replace the current treatment. Phytochemicals have received increasing interest in the past decade, largely owing to their effectiveness for disease prevention and treatment\(^15\). The use of natural products, particularly plant-based remedies, is favored for combating fibrotic-induced diseases, compared to other complementary and alternative approaches; for example, Japanese herbal medicine (Saiko-keishito), polyphenol compounds (curcumin), antioxidants (vitamins A and E), protease inhibitor (camostat mesylate), and lovastatin (Monascus or Aspergillus-fermented rice and Dioscorea) are plant-derived pharmacological agents that have been shown to prevent and/or improve CP and PC\(^16\). In recent years, plant-derived products have undergone clinical trials to evaluate their efficacy as anti-fibrotic agents in the treatment of metastatic PC. Specifically, analogs of vitamins A and D have been claimed as potential anti-fibrotic agents against PSCs; for example, limited to phase I/II clinical trials involving vitamin analogs, which are abundant in medicinal plants and have proved to be promising for clinical application. The use of phytomedicines, as new anti-fibrotic agents, has been applied to a variety of complementary and alternative approaches. The aim of this review was to present a focused update on the selective new potential anti-fibrotic agents, including curcumin, resveratrol, rhein, emodin, green tea catechin derivatives, metformin, eruberin A, and ellagic acid, in combating PSC in CP and PC models. It aimed to describe the mechanism(s) of the phytochemicals used, either alone or in combination, and the associated molecular targets. Most of them were tested in PC models with similar mechanism of actions, and curcumin was tested intensively. Future research may explore the issues of bioavailability, drug design, and nano-formulation, in order to achieve successful clinical outcomes with promising activity and tolerability.

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paricalcitol, a synthetic vitamin D analog, is undergoing Phase I and II clinical trials, in combination with different conventional chemotherapeutic drugs, for treating metastatic PC. Other potential PC treatments in clinical settings involve reprogramming PSCs using vitamin A metabolites, such as all-trans retinoic acid or selective retinoic acid receptor beta (RAR-β) agonist. In addition, combinations of pirfenidone and N-acetylcysteine, or the use of pirfenidone alone, with regard to PC warrant more extensive studies in human subjects. The evidence has shown that plant-derived products have significant potential to act as anti-fibrotic agents in treating CP and PC and deserve intensive investigation using in vitro and in vivo models.

This paper presents a detailed review of the anti-fibrotic activity of selective potential phytochemicals, which are new and effective in treating CP and PC, by focusing on the PSCs evidenced in in vitro and in vivo models. Furthermore, it discusses the mechanism(s) that underlie the anti-fibrotic activity, the key molecules involved, and the concentrations used in the CP and PC models.

2. Pancreatic stellate cells

PSCs are the pluripotent cells, located between the pancreatic lobules and the surrounding area of acinar, that maintain the connective tissue architecture. PSCs have two phenotypes: quiescent and activated. In a normal human pancreas, PSCs comprise approximately 4%—7% of the parenchymal cells and contain cytoplasmic lipid droplets containing vitamin A in its quiescent form. Under normal physiological conditions, PSCs maintain their quiescence by expressing nestin, vimentin, glial fibrillary acidic protein (GFAP), and desmin. Furthermore, retinoids, sometimes in the form of retinyl palmitate, can be found in the cytosolic droplets of quiescent PSCs. These retinoids can be used as markers to differentiate them from the normal fibroblasts.

The activation of PSCs can be induced by pathologic conditions, such as CP and PC; hence, activated PSCs are responsible for the excessive fibrotic state in pancreatic pathology. The inactive PSCs are identified by the abundant vitamin A stored in the cytoplasm, while an injured pancreas lacks cytoplasmic vitamin A-storing lipid droplets. Activated PSCs have been identified using a variety of phenotypic parameters. They were found to be localized interlobularly in fibrotic areas adjacent to the carcinoma cells. Activated PSCs display a loss of fat droplets and a high mitotic index, with intense reticular endoplasmic reticulum (ER) and high motility and contraction. Receptors, such as platelet-derived growth factor receptor (PDGF-R), transforming growth factor beta receptor (TGFβ-R), and intercellular adhesion molecules (ICAM-1), are expressed by the activated PSCs, along with other receptors. Additionally, there is an enhanced expression of ECM proteins (collagens I, III and XI, fibronectin, and peroxin (a cell adhesion protein) that stimulates cancer cell growth), which forms the fibrous tissue, and an enhanced release of neutrophilic factors/transmitter, growth factors, and cytokines. Furthermore, activated PSCs also exhibit differential expression of multiple genes, including a 32.25-fold up-regulation of MMP-3 and a 2.25-fold down-regulation of the basement membrane component, collagen type IV-a1, which may contribute to restructuring of the ECM in the activated form.

The ECM is composed of collagen, fibronectin, and multiple soluble factors secreted by PSCs, which provide the structural support and promote differentiation, remodeling, and carcinogenesis. Ben-Harosh et al. reported that palmitate fatty acids significantly limited PSCs’ activation and fibrosis by halting their proliferation and migration capacity via the suppression of ER. By contrast, PSCs’ activation and differentiation were augmented following the treatment of oleate fatty acids and caerulein-induced stress with increased levels of ER stress markers (X-box binding protein 1 (Xbp1) and CCAAT-enhancer-binding protein homologous protein (CHOP)). Fibrous proteins, such as collagen, laminin, and fibronectin, and non-collagenous proteins, such as glycoproteins, proteoglycans, and glycosaminoglycans, together make-up the ECM constituting the stromal component. This abundant stromal reaction usually surrounds the island of cancer cells and accounts for 50%—80% of tumor volume.

The cytokines and growth factors secreted by PSCs promote the angiogenesis and the proliferation, migration, and invasion of the epithelial cancer cells that lead to metastasis. Soluble factors, especially IL-6, have been shown to be involved in transitioning non-invasive into invasive pancreatic ductal adenocarcinoma (PDAC). Apparently, PDAC is among the stroma-rich and fibrotic malignancies, leading to the conclusion that the ECM process plays a key role in the development of fibrosis and PDAC progression. It is characterized by the formation of dense fibrotic stromata (desmoplasia), formed by the activated PSCs.

Furthermore, the continuous interaction between PSCs and the ECM can lead to further PSC activation due to the resulting stiffness of the matrix formation. Such a process is fundamental to pathological fibrosis in both CP and PC.

3. Chronic pancreatitis

CP is a fibro-inflammatory disease that causes the pancreatic parenchyma to be progressively replaced by fibrous connective tissue, potentially leading to exocrine and endocrine pancreatic inadequacy. Continuing pancreatic damage is caused by oxidative stress or recurrent episodes of inflammation, leading to irreversible functional and morphological changes in the pancreas, which may or not be clinically evident, and resulting in the development of CP and PC. The activation of digestive enzymes in pancreatitis occurs before they are released into the small intestine and cause glandular damage, leading to CP by inducing a progressive, destructive inflammatory process that ends in the destruction of the pancreas. The clinical presentation of CP includes abdominal pain, steatorrhea, diabetes, weight loss, and obstructive jaundice, all of which are very similar to PDAC.

Nearly 70% of CP cases are caused by alcohol abuse, and the remaining cases are attributed to genetic disorders, pancreatic duct obstruction, recurrent acute pancreatitis (AP), autoimmune pancreatitis, or unknown mechanisms. Laboratory studies have highlighted a proliferation of reactive oxygen species (ROS) as the trigger and potentiator of inflammation, since they activate the signaling cascades that convert the damaged acinar cells into a production site for chemokines and cytokines. ROS have several physiological roles, including signal transduction, but an excess of ROS relative to the antioxidant capacity (electrophilic stress) is potentially harmful. Depending on the concentration, ROS facilitate mild to moderate levels of carcinogenesis and cancer progression, while the excessive ROS damage to cancer cells is dramatic and leads to cell death. Increased ROS are observed in CP, which increases the PC incidents via PSCs’ activation. Furthermore, the exocytosis blockade seems to be caused by the disruption of the methionine trans-sulphuration...
pathway that produces the essential methyl and thiol moieties. This condition occurs in both clinically acute and acute-on-chronic pancreatitis. When this physiological process is disrupted, it may result in the development of pathological fibrosis, with significant adverse effects on the anatomy and physiology of affected tissues. Abnormal deposition of fibrous tissue is a characteristic histological feature of two major diseases of the pancreas: CP and PC. Fibrosis is a sign that interstitial PSCs have been activated in CP, due to the increase in lipid peroxidation products and the release of mast cell degranulation products; hence, it is evident that PSCs are associated with CP, and that targeting PSCs maybe a promising treatment option for CP.

4. Pancreatic cancer

By 2030, it is expected that PC will be the second leading cause of deaths from types of cancer. Deaths result from PDAC, the major and most aggressive type of PC, and rank fourth among the cancer-related deaths in USA. PC is more prevalent among the elderly (occurring mainly between the ages of seventy and eighty) than in younger people, and less than 20% of patients have localized and potentially curable tumors. PC is very difficult to diagnose and often remains undetected until the disease has reached an advanced stage. The etiology of PC is poorly understood, but several factors are known to increase the risk. The preventable risk factors include cigarette smoking, obesity, and a high intake of animal fat, while non-preventable risk factors include CP, an inherited genetic predisposition, and cystic fibrosis.

Activated PSCs were found to produce the ECM proteins that comprised the pancreatic stroma. PC cells are closely interacting PSCs that cause an increase in ECM and fibrosis, in turn stimulating cancer cell proliferation and inhibiting cancer cell apoptosis. PC cells recruit PSCs to their immediate vicinity and promote a fibrogenic response in the PSCs. Inflammatory markers, including IL-6, increase in patients with PDAC. Tumor-associated macrophages are the main source of IL-6 in PC tissue, but IL-6 secreted by the activated PSCs has been reported to regulate PDAC phenotypes. CP is a risk factor for PC, but most cases of PC develop in patients without clinical symptoms of CP; however, there is evidence of inflammation in tissue samples. Kristen rat sarcoma virus oncogene homolog (KRAS) mutations and activations in PC cells have been seen, in mice models, to promote inflammatory signaling and precancerous lesion development. Subsequently, the inflammatory stimulus activates PSCs in the periacinar area, leading to the recruitment of immune cells (monocytes, T cells, neutrophils, macrophages, and mast cells). Despite significant advances in understanding tumor biology and developing novel therapies, survival rates remain discouraging. Targeted therapies based on advances in precision medicine, such as immunotherapy, engineered T-cells, tumor vaccines, myeloid-based immunotherapy, stromal modulating immunotherapy, and gene therapy, are nowadays available for use in dedicated healthcare centers, while others are still under preclinical investigation.

In view of the dominant role played by PSCs in the initiation and progression of PC, it is crucial to identify and develop an anti-fibrotic agent to fight PSCs in order to inhibit devastating PC.

5. The roles of pancreatic stellate cells in chronic pancreatitis and pancreatic cancer

One major similar property shared by CP and PC is that both possess a large proportion of stromata. Pancreatic stromata play an important role in hereditary PC, with most cases resulting from the progression of hereditary pancreatitis to CP. PSCs play a key role in supporting and promoting various aspects of PC, such as proliferation, migration, invasion, colony formation, and angiogenesis, in addition to other promoting factors. Liu et al. isolated, identified, and cultured human PSCs and discovered that activated PSCs are present in PC tissues. Their results further showed that PSCs can promote the invasive ability of PC cells and reduce the apoptosis rate induced by gemcitabine.

The EMT is necessary for many physiological developmental steps; however, it contributes to tumorigenesis (the broad spectrum of trans-differentiation in tumors) and to metastatic spread. During tumorigenesis, PSCs transform into active myofibroblast-like phenotypes, which are involved in several processes. They create a suitable microenvironment for facilitating most cases of cancer progression and invasion. PSCs secrete MMPs, including MMP2, MMP9, and MMP13, as well as TIMP1 and TIMP2, suggesting that PSCs contribute to maintaining the balance in the ECM in a healthy organ. However, they disrupt this balance upon their activation in pancreatitis and PC. The epidermal growth factor receptor (EGFR) pathway is involved in pancreatic fibrosis; the overexpression of heparin-binding epidermal growth factor-like HB-EGF in the pancreatic islet has been claimed as one of mechanisms contributing to the massive fibrotic state in cancer and CP. PSCs express EGFR (which is activated by HB-EGF), leading in an autocrine manner to an increase in PSCs' activation and migration, thereby modulating the stromata to support PC growth. Furthermore, Komar et al. demonstrated that the JAK/STAT pathway plays a prominent role in PSC proliferation and activation, secreted as an abundance of several immunomodulatory factors, including IL-6 and monocyte chemoattractant protein-1 (MCP-1). Inhibition of this pathway led to reduced caerulein-induced CP in vivo.

6. Application of phytochemicals as new anti-fibrotic agents in treating chronic pancreatitis and pancreatitis cancer

In recent decades, plant-based/herbal constituents and natural products have been widely used as complementary and alternative medicines to increase longevity and treat diseases. The phytochemicals extracted from the medicinal plants or herbs play important roles in preventing or treating CP and PC via different mechanistic pathways. For this review, we have selected several phytochemicals that have shown potential anti-fibrotic activity against PSCs in recent years. They are curcumin, resveratrol, rhein, emodin, green tea catechin derivatives, ellagic acid, embelin, eruberin A, and metformin, and their respective chemical structures are shown in Fig. 1. The anti-fibrotic activity of these phytochemicals, as well as their mechanistic actions, are elaborated in the following sections.

6.1. Curcumin

Curcumin belongs to the Zingiberaceae family and is a turmeric polyphenol derived from the rhizomes of Curcuma longa, which is cultivated in most parts of Southeast Asia. Curcumin is a lipophilic agent and stable in the acidic pH environment of the stomach. Curcumin is well-known for its medicinal value; in particular, its antioxidant and anti-inflammatory properties. It has been reported that curcumin is responsible for the suppression of cell proliferation, invasion, and angiogenesis.
Furthermore, it also induces apoptosis via deactivation of nuclear factor-kappa B (NF-κB) and its regulated gene products. Additionally, it can also suppress several inflammatory cytokines, such as TNF-α, interleukins (IL-1, IL-6, IL-8 and IL-1β), and cyclooxygenase-2 (COX-2). Curcumin has direct effects on pancreatic beta cells, which could contribute to the hypoglycemic effects of this compound and decreased beta cell volume, suggesting that this is another novel attribute of curcumin. Additionally, it increases the islet content of glutathione (GSH, a product of the modulatory subunit of gamma-glutamate cysteine ligase (γ-GCL)) and basal insulin secretion and protects them from oxidative stress.

Thus far, a few in vitro and in vivo studies have been performed to evaluate the anti-fibrotic activity of curcumin against PSCs in CP and/or PC models, and the mechanisms of their actions are depicted in Fig. 2. An in vitro study using obese mice livers showed that curcumin decreased inflammation in adipose liver steatosis through the phosphorylation of the signal transducer and activation of transcription 3 (STAT3) signaling, as well as reduction of the cytokine signaling 3 (SOCS3) suppressor and sterol regulatory element-binding protein-1c (SREBP-1c). These findings indicated the ability of curcumin to mediate anti-inflammatory effects for the treatment of liver steatosis. In activated PSCs, curcumin decreased the pancreatic beta cell volume, which could be associated with hypoglycemic effects.

In addition, curcumin inhibited PDGF-induced PSCs proliferation, and it reduced α-SMA gene expression, IL-1β and TNF-α-induced MCP-1 production, type I collagen production, and activator protein-1 (AP-1) activation in the activated PSCs. The activation of AMP-activated protein kinase (AMPK) by curcumin is important for the inhibition of differentiation in adipocytes and cancer cells. This results in the attenuation of peroxisome proliferator-activated receptor gamma in 3T3-L1 adipocytes, and decreased COX-2 expression. Schwer et al. provided the first evidence that curcumin can inactivate PSCs by inhibiting their proliferation. This action is mediated by a decrease in extracellular signal-regulated protein kinase 1 and 2 (ERK1/2) activation in parallel with heme oxygenase-1 (H O-1) up-regulation, increasing the level of cellular carbon monoxide and thereby activating P38 mitogen-activated protein kinases (MAPK), leading to a reduction of PSC proliferation. Moreover, curcumin and three phenolic compounds were shown to significantly suppress the mRNA and protein levels of several fibrotic mediators in primary PSCs activated by TGF-β, including α-SMA, type I collagen, and fibronectin, and the underlying mechanism was associated with the down-regulation of the NF-κB signaling pathway. These findings suggested that curcumin may serve as an anti-fibrotic agent for treating pancreatic fibrosis and PSC-related pathologies, including PDAC.

A newly synthesized curcumin analog (L49H37) was used as an intervention to target the stromal component of PC and compared to traditional curcumin. It was found that L49H37 was more potent for inducing PSCs’ apoptosis at a concentration 10 times lower than curcumin. The results showed that the anti-proliferative effect of L49H37 (2.5 μmol/L) significantly inhibited PSC proliferation compared to curcumin (25 μmol/L), as evidenced by the observation of changes in the cell cycle regulatory protein levels of the potent cyclin-dependent kinase inhibitor (CKI), P21WAF1/Cip1.

The efficacy of curcumin for clinical application has been tested in several clinical phase trials. A well-performed clinical trial revealed that combination therapy with gemcitabine-based chemotherapy and oral curcumin administration (8 g) proved to be feasible and safe for PC patients. In addition, a phase II clinical trial on 25 patients with advanced PC confirmed the safety and efficacy of curcumin, despite its low bioavailability. Both clinical studies have completed phase II trials. A randomized placebo-controlled pilot study.
A study involving 20 tropical pancreatitis patients showed that an oral combination of curcumin (500 mg) and piperine (5 mg) was effective in relieving pain and beneficially modulating the markers of oxidative stress, including malonyldialdehyde (MDA) and GSH. In advanced PC, a dose of 8 g of curcumin per day was administered for 2 months. This study discovered that curcumin was well-tolerated and signs of biological activity were found in most patients. Another clinical trial, involving 21 patients, showed stabilized disease progression in advanced PC after administration of an 8 g dose of curcumin per day. One patient maintained disease stabilization for 18 months and a second patient experienced a significant increase in serum cytokine levels, accompanied by brief, but marked, tumor regression (73%).

6.2. Rhein

Rhein is a natural anthraquinone derivative, extracted from the rhizomes of several traditional medicinal plants; for example, Rheum palmatum, also known as “da huang”, is commonly used as a purgative. Rhein has been reported to exert various pharmacological effects, such as antimicrobial, anti-inflammatory, anti-angiogenic, and anticancer effects. In fact, the anticancer activity of rhein has been tested in both in vitro and in vivo models; for instance, rhein inhibited PC cell growth by arresting the expression of hypoxia-inducible factor-1 alpha (HIF-1α) via the decrease in phosphorylation of phosphorylated protein kinases B (p-AKT) and ERK1/2 (p-ERK1/2) in vitro.

The anti-fibrotic activity of rhein has been reported in both CP and PC models, as evidenced in several studies, and the mechanisms of their actions are described in Fig. 3. Tsang et al. demonstrated that treatment with rhein (50 mg/kg/day) in a study involving 20 tropical pancreatitis patients, showed that an oral combination of curcumin (500 mg) and piperine (5 mg) was effective in relieving pain and beneficially modulating the markers of oxidative stress, including malonyldialdehyde (MDA) and GSH. In advanced PC, a dose of 8 g of curcumin per day was administered for 2 months. This study discovered that curcumin was well-tolerated and signs of biological activity were found in most patients. Another clinical trial, involving 21 patients, showed stabilized disease progression in advanced PC after administration of an 8 g dose of curcumin per day. One patient maintained disease stabilization for 18 months and a second patient experienced a significant increase in serum cytokine levels, accompanied by brief, but marked, tumor regression (73%).

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mouse model of cerulein-induced CP significantly attenuated fibrogenesis by decreasing the immunoreactivity of fibrotic activators, including α-SMA and TGF-β, in pancreatic tissues, followed by the reduction of fibronectin (FN1) and type 1 collagen deposition in the exocrine. In addition, rhein was found to suppress various fibrotic and tumorigenic markers, such as α-SMA, fibronectin, type I collagen, N-cadherin, and MMPs, in cultured PSCs and tested mammalian cells by modulating both the sonic hedgehog (SHH) and serine–threonine kinase signaling pathways[@2][@3]. Like other phenolic compounds, the underlying mechanisms of the anti-fibrotic and anti-tumorigenic effects of rhein are associated with the downregulation of the NF-κB and STAT3 signaling pathways[@4][@5]. The subunit of NF-κB (P65), which is involved in an inflammatory response, was significantly reduced by 20 μmol/L of rhein in PSCs[@6].

6.3. Green tea catechin derivatives

Green tea is also known as *Camellia sinensis*. In Asian countries, the leaves are common, popular, and widely consumed by the people for their health benefits or medicinal effects, such as antioxidant, anti-inflammatory, anti-proliferative, anti-atherosclerotic, and anti-cancer effects[@7][@8]. Green tea extracts contain both polyphenolic and non-phenolic components. The phenolic compound in green tea mainly consists of phenolic catechins, including (−)-epigallocatechin gallate (EGCG), (−)-epicatechin gallate (ECG), (−)-epicatechin (C), and (+) -catechin (GC). EGCG is one of the most important catechins, due to its high content and antioxidant activity.

EGCG has been shown to inhibit the PDGF-induced proliferation and migration of PSCs[@9]. EGCG was proved to inhibit the PDGF-induced tyrosine phosphorylation of the PDGF β-receptor, downstream activation of ERK, and phosphoinositol 3-kinase (PI3K)/AKT pathways. Furthermore, pre-treatment with EGCG inhibited the ethanol-induced activation of PSCs in vitro. Ethanol significantly increased the production of α-SMA protein (type-I procollagen), activated TGF-β1, and induced phosphorylation of P38 MAPK. Interestingly, treatment with EGCG significantly decreased the production of all these factors and thoroughly abolished P38 MAPK phosphorylation. In addition, ethanol-stimulated transforming of PSCs from normal quiescent phenotypes into myofibroblast-like phenotypes was inhibited by EGCG[@10].

6.4. Resveratrol

Resveratrol, or 3,5,4’-trihydroxy stilbene, is a polyphenolic stilbene compound that can be found in grapes, raspberries, blueberries, cocoa, and peanuts[@11]. It is synthesized in response to injury or attacks from pathogens of the plants[@12]. A large number of *in vitro* and *in vivo* studies have demonstrated the protective effects of resveratrol with regard to several pathological diseases, such as cardiovascular diseases[@13], diabetes[@14], neurological disorders[@15][@16], and different types of cancer[@17][@18]. The uses of resveratrol in treating PC were evidenced by several *in vitro* studies. It was shown to induce PC apoptosis by promoting caspase-3 activation, while remaining nontoxic to the normal pancreatic cells[@19]. Furthermore, resveratrol increased the chemosensitivity of PC cells by targeting nutrient-deprived autophagy factor-1 (NAF-1) via ROS/nuclear factor erythroid 2 (NRF2) signaling[@20]. Zhou et al.[@21] also reported that resveratrol could enhance the sensitivity of PC cells (MiaPaCa-2 and Panc-1) and decrease the markers of cancer stem cells *via* suppression of SREBP1 in both *in vitro* and *in vivo* models. Intriguingly, Yang et al.[@22] discovered that, in addition to acting as a tumor suppressant *via* BAX up-regulation, resveratrol can act as a tumor activator by up-regulating vascular endothelial growth factor B (VEGF-B) in Capan-2 cells.

The activation of PSCs is an important process in the development of pancreatic fibrogenesis, leading to CP and PC. Resveratrol was found to impede the activation, invasion, migration, and glycosylation of PSCs induced by ROS by down-regulating the expression of microRNA 21 (miR-21) and increasing the phosphatase and tensin homolog (PTEN) protein levels[@23]. In the same study, the results further demonstrated that resveratrol inhibited the invasion and migration of PC by suppressing ROS/ miR-21 mediated activation and glycosylation in PSCs. In addition, resveratrol was shown to suppress the PSCs’ viability *via* the reduction of several major fibrogenic mediators, such as α-SMA, type I collagen, and fibronectin, which are associated with down-regulation of the NF-κB signaling pathway[@24].

6.5. Emodin

Emodin (1,3,8-trihydroxy-6-methylanthraquinone), an important component of *Aloe vera*, can be extracted from members of the Polygonaceae plant family, such as *Palmatum Rheum*. Diverse studies have found that emodin exerts various pharmacological effects, including anti-inflammatory, anti-angiogenic, and anti-dyslipidemic actions, in addition to having anticancer potential in *in vitro* and *in vivo* studies[@25][@26][@27][@28]. Emodin has shown potent anticancer activity in PC, as evidenced in several *in vitro* and *in vivo* studies, by targeting cell proliferation and inducing apoptosis *via* various mechanisms[@29][@30]. Furthermore, the combination of emodin and existing chemotherapeutic drugs for PC further enhanced the chemosensitivity of PC cells[@31].

In the literature, only one study investigated the anti-fibrotic activity of emodin against PSCs. Treatment with emodin (4 μmol/L) significantly decreased the primary PSCs’ cell viability by down-regulating the mRNA and protein expression of several fibrotic mediators, including α-SMA, type I collagen, and fibronectin[@32]. The authors suggested that the concentration used in *in vitro* experiments should not be higher than 5 μmol/L.

6.6. Ellagic acid

Ellagic acid is a plant-derived non-flavonoid polyphenol that is mainly found in fruits and nuts, such as berries, grapes, pomegranates, and walnuts[@33]. In recent decades, the health benefits of ellagic acid have been reported and shown to be effective, not only for the treatment of chronic metabolic diseases, such as dyslipidemia, non-alcoholic fatty liver diseases, insulin resistance, and type-2 diabetes[@34][@35][@36], but also as anticancer and antitumor treatments[@37][@38][@39].

Several studies, in either CP and/or PC models, have evaluated the anti-fibrotic activity of ellagic acid, and its underlying mechanisms are depicted in Fig. 4. Treatment with ellagic acid in an experimental CP model, using Wistar Bonn/Kodori rats, significantly abolished the development of pancreatic fibrosis. The mRNA expression of α-SMA and TGF-β1 were markedly reduced. In addition, the infiltration of macrophages or monocytes and ROS production in isolated PSCs were significantly decreased in the
rats after ellagic acid treatment\(^{114}\), and ellagic acid inhibited the PSC proliferation and migration induced by PDGF-BB. Although the PDGF-\(\beta\) receptor protein levels did not change, ellagic acid inhibited the tyrosine phosphorylation of the receptors, such as ERK and AKT. Thus, ellagic acid diminished the activation of the downstream signaling pathway of RAF proto-oncogene serine/threonine-protein kinase (c-Raf)/MAPK/ERK and PI3K/AKT, which are important for PSC cell proliferation and migration\(^{115}\). Masamune et al.\(^{115}\) further reported that ellagic acid inactivated PSCs by attenuating the protein levels of \(\alpha\)-SMA and ECM pro-collagen types I and III. Moreover, ellagic acid treatment inhibited IL-1\(\beta\) and TNF-\(\alpha\)-induced MCP-1 in PSCs correlated with AP-1, but not NF-\(\kappa\)B. Ellagic acid was also found to prevent the transformation of PSCs from quiescent into myofibroblast-like phenotypes.

### 6.7. Embelin

Embelin is a naturally occurring benzoquinone that can be extracted from the fruits (berries) of the *Embelia ribes* Burm. plant (Myrsinaceae). It has been used for various traditional medicinal remedies in India. Embelin had been shown to exert different pharmacological effects, including as an anticanter agent via inhibition of cell migration, invasion, and induction of apoptosis in colon, lung, and lung cancer cells\(^{116}-118\).

To date, the anti-fibrotic activity of embelin has been indicated in a PC model\(^{113}\), with the results showing that embelin inhibited both PC cells and PSCs in a dose-dependent manner. Interestingly, embelin in combination with ellagic acid at low concentrations (0.5–3 \(\mu\)mol/L) synergistically increased apoptosis and reduced cell proliferation compared to the individual treatments. A similar observation was also made in relation to a subcutaneous xenograft mouse model of PC, in which embelin alone, or in combination with ellagic acid, significantly reduced a tumor’s size and cellularity. The mechanism underlying the action of embelin is via STAT3 dephosphorylation and the reduced expression of its downstream target, survivin, in PC cells\(^{113}\). In PC cells (MIA PaCa-2 and HPAF-II), embelin was demonstrated to stimulate apoptosis and inhibit cancer cell proliferation dose-dependently. Embelin reversed the anti-apoptotic effect of the X-linked apoptosis protein (XIAP) by preventing the interaction of XIAP with caspases, and it was also found to downregulate the expression of XIAP, survivin, inhibitor of apoptosis 1 and 2 (IAP1/2), tumor necrosis factor receptor-associated factor 1 (TRAF1), cellular FLICE (a FADD-like IL-1\(\beta\)-converting enzyme), inhibitory protein (cFILP), B-cell lymphoma 2 (BCL-2), and B-cell lymphoma-x (BCL-X) by suppressing the activation of NF-\(\kappa\)B\(^{113,119}\). In a xenograft mouse model of PC, embelin was reported to reduce tumor growth and the results also showed that embelin metabolized rapidly after oral administration in the xenograft mouse model\(^{113}\). Furthermore, embelin was found to inhibit cancer cell proliferation by down-regulating the SHH signaling pathway, with implications, not only for promoting cell proliferation, but also cell invasion, metastasis, and tumor growth. It was assumed that targeting SHH might inactivate PSCs, since the expression of SHH was reported to promote tumor growth via PSC-induced desmoplasia formation, as well as to affect the mobility and differentiation of PSCs\(^{120,121}\). The mechanisms of the actions underlying the anti-fibrotic activity of embelin are shown in Fig. 5.

### 6.8. Eruberin A

Eruberin A is a pure compound extracted from the fern plant, *Pronephrium penangianum*. It is an organic flavanol glycoside and...
Table 1  The mechanisms of actions of selective phytochemicals in inactivating PSCs in chronic pancreatitis and/or pancreatic cancer models.

| Phytochemical | Disease | Test model | Dosage | Metabolic responses/mechanisms | Ref. |
|---------------|---------|------------|--------|-------------------------------|------|
| Curcumin      | CP      | In vitro, TGF-β stimulated cultured primary PSCs (LTC-14) | 20 μmol/L | 1. Down-regulated NF-κB signaling by reducing subunit P65; 2. Inhibited the production of fibrogenic mediators (Acta 2, Col I and FNI) induced by TGF-β. | 58   |
|               | PC      | In vitro, cultured PSCs | 5–25 μmol/L | 1. Inhibited PDGF-induced proliferation; 2. Reduced gene expression of α-SMA, IL-1β, Col I and Col III; 3. Inhibited production of TNF-α-induced MCP-1. | 65   |
|               | PC      | In vitro, cultured human PSCs | 1–25 μmol/L | 1. Inhibited cell proliferation and induced cell apoptosis; 2. Increased phosphorylation of ERK1/2 at lower concentrations (1 and 10 μmol/L). | 69   |
| Rhein         | CP      | In vivo, cerulein-induced CP mouse model | 50 mg/kg/day | 1. Attenuated fibrogenesis by decreasing immunoreactivity of fibrotic activators (α-SMA and TGF-β) and reduction of fibronectin (FNI and type 1 collagen deposition). | 82   |
|               | CP      | In vitro, TGF-β stimulated cultured LTC-14 cells | 20 μmol/L | 1. Reduced P65 (subunit of NF-κB); 2. Inhibited the production of fibrogenic mediators (Acta 2, Col I and FNI); 3. Down-regulated NF-κB signaling pathway. | 58   |
|               | PC      | In vitro, human pancreatic cancer cells BxPC-3, PANC-1, Patu8988T and AsPC-1 | 60 μmol/L | Suppressed constitutive STAT3 tyrosine phosphorylation and induces apoptosis in pancreatic cancer cells. | 84   |
|               | In vivo, xenograft BALB/c female mice model | 60 mg/kg | 1. Inhibited tumor growth; 2. Reduced expression of p-STAT-3 and p-EGFR; 3. Downregulated STAT-3 signaling pathway. | 83   |
|               | PC      | In vitro, cultured PSCs and testing mammalian cells | 20 μmol/L | 1. Suppressed various fibrotic and tumorigenic markers (α-SMA, fibronectin, type I collagen, N-cadherin, and MMPs); 2. Modulating both sonic hedgehog (SHH) and serine–threonine kinase signaling pathways. | 89   |
| Green tea [[(−)−epigallo-
|               | PC      | In vitro, cultured PSCs | 25 μmol/L | 1. Inhibited ethanol-induced morphological changes of PSCs from normal quiescent-phenotype to myofibroblast-like; 2. Decreased production of α-SMA; suppressed type-I procollagen production; 3. Activated TGF-β secretion; 4. Abolished ethanol-induced increases in P38 MAP kinase phosphorylation. | 88   |
|               | PC      | In vitro, rat PSCs | 1–25 μmol/L | 1. Inhibited PDGF-induced proliferation and migration; 2. Inhibited cell cycle progression beyond G1 phase; 3. Inhibited PDGF-induced phosphorylation ERK and Akt. | 89   |
| Resveratrol   | PC      | In vitro, cultured MiaPaC-2 and PANC-1 cell lines | 50 μmol/L | 1. Increased sensitivity of PCs to gemcitabine; inhibited lipid synthesis; | 102  |
|               | PC      | In vivo, genetically engineered mouse | 50 mg/kg/day | 2. Rescued the stemness induced by gemcitabine via suppressing SREBP1. | 101  |

*Liver fibrosis

(continued on next page)
| Phytochemical | Disease | Test model | Dosage | Metabolic responses/mechanisms | Ref. |
|---------------|---------|------------|--------|--------------------------------|------|
| Emodin        | PC      | *In vitro*, TGF-β stimulated cultured LTC-14 cells | 4 μmol/L | 1. Reduced P65 (subunit of NF-κB); 2. Inhibited the production of fibrogenic mediators (*Acta2*, *Col α1*, and *Fln*); 3. Downregulated NF-κB signaling pathway. | 58   |
| Ellagic acid  | CP      | *In vivo*, male wistar Bonn/Kobori rats | 100 mg/kg/day | 1. Attenuated myeloperoxidase activity; 2. Decreased in collagen content, reduced TGF-β1 expression, and reduced the amount of α-SMA and macrophages monocytes (ED)-positive cells; 3. Inhibited TNF-β1 and platelet derived growth factor (PDGF)-induced reactive oxygen species (ROS). | 115  |
| PC            | *In vitro*, rat PSCs | 25 μg/mL | 1. Down-regulated α-SMA, collagen (α1(I) procollagen and α1(III) procollagen); 2. Inhibited transformation of quiescent freshly isolated PSCs into myofibroblast. | 116  |
| PC            | *In vitro*, MIA PaCa-2; HPAF-II cells | 10–30 μmol/L | 1. Decreased NF-κB transcriptional activity; 2. Stimulated apoptosis and inhibited proliferation in pancreatic cancer cells. | 114  |
| Emodin        | PC      | *In vitro*, AsPC-1, PANC-1, MIA PaCa-2 and Hs766T cell lines | 1–15 μmol/L | Inhibited cell growth; suppressed SHH signaling pathway. | 127  |
| Embelin       | PC      | *In vitro*, nude mice xenograft model of pancreatic cancer (AsPC-1) | 40 mg/kg | 1. Inhibited tumor cell proliferation and induced apoptosis; 2. Inhibited angiogenesis. | 114  |
| Metformin     | PC      | *In vitro*, geneti- cally engineered mouse model, KPC mice | 200 mg/kg/day | 1. Suppressed the growth and the progression of the tumor in PDAC; 2. Reduced production of α-SMA and ECM; 3. Anti-PSCs effect via the reduction of SHH expression, thus decreasing VEGF, tumor neoangiogenesis, and desmoplastic reaction. | 134  |
| Eruberin A    | PC      | *In vitro*, human pancreatic cancer cell AsPC-1, BxPc3, CFPAC-1, Panc-1 and SW1990 | 5 mmol/L | 1. Significantly reduced mRNA expression of CTGF, TGF-β1, and PDGF-A; 2. Suppressed secretion of TGF-β1 through activation of AMPK signaling pathway; 3. Inhibited invasion and migration ability of PSC. | 132  |
|               | PC      | *In vitro*, Balb/c nude mice orthotopic pancreatic cancer model | 100 mg/kg | 1. Reduced the α-SMA-positive cell and collagen in the tumor microenvironment; 2. Enhanced chemo sensitivity of gemcitabine by inhibited ECM deposition. |      |
|               | PC      | *In vitro*, rat LTC-14 cell line; human PDAC PANC-1 cell line | 20 μg/mL | 1. Anti-fibrotic effect; suppressed TGF-β1-induced fibrogenic mediator; 2. Downregulated the activation of NF-κB and SHH signaling components; 3. Suppressed the activation of PI3K/AKT signaling pathway. | 135  |
has been reported to exert antioxidant effects and reduce proinflammatory cytokine production and diabetic-associated oxidants, such as hydrogen peroxide\textsuperscript{122,123}. Eruberin A has also been demonstrated to have potent cytotoxic effects on L929 fibroblasts and HeLa cells\textsuperscript{124}. Thus far, only one study has investigated the anti-fibrotic activity of eruberin A against primary PSCs; namely, LTC-14. The growth rate of LTC-14 cells decreased after treatment with eruberin A in a dose-dependent manner and significantly suppressed the gene expression of the major fibrotic filaments and ECM mediators, including smooth muscle \(\alpha\) actin (\textit{Acta 2}), collagen type I-alpha 1 (\textit{Col I-a1}), and fibronectin 1 (\textit{FN1}) at 20 \(\mu\)g/mL; a concentration that did not cause cytotoxic effects\textsuperscript{125}. Similar results were observed in eruberin A-treated PANC-1 cells. Eruberin A inhibited NF-\(\kappa\)B activation and the SHH signaling components, and suppressed the activation of the PI3K/AKT pathway linked to inflammatory and fibrogenesis downstream cascades\textsuperscript{125}. A 20 \(\mu\)g/mL dose of eruberin A inhibited TGF-\(\beta\)-induced AKT phosphorylation and attenuated \(\alpha\)-SMA at both protein and cytoplasmic levels in LTC-14 cells and PANC-1 cells\textsuperscript{125}. The effects of eruberin A on PSCs and PC were less reported, so more research and investigations are needed to gain further understanding.

6.9. Metformin

Metformin is one of the guanidine derivatives that are rich in \textit{Galega officinalis} (a well-known European goat rue) and, in 1918, were shown to lower blood glucose levels\textsuperscript{126}. Metformin is an oral anti-diabetic medicine for type-2 diabetic patients and helps to control the amount of glucose produced by the liver. It has also been reported to exert anticancer effects on different cancer types, including breast, ovarian, pancreatic, and colon cancer, by modulating the inflammatory responses and cancer stem cells\textsuperscript{127–130}.

Metformin has been shown to inhibit desmoplasic reaction and enhance the chemosensitivity of PDAC towards gemcitabine by activating AMPK. Furthermore, \textit{in vitro} and \textit{in vivo} studies have demonstrated that metformin activated AMPK in both human PSCs and nude mice with subcutaneous pancreatic cancer, by up-regulating p-AMPK expression, significantly decreasing TGF-\(\beta\)-1, \(\alpha\)-SMA, and collagen in tumor microenvironments, and inhibiting PSC proliferation\textsuperscript{131,132}. In addition, metformin enhanced the sensitivity of PSCs and PC cells in response to gemcitabine treatment. Combined treatment with metformin and gemcitabine significantly reduced the expression of SHH by inhibiting the production of VEGF and tumor neovascularisation, thus improving the chemosensitivity of PC cells to gemcitabine\textsuperscript{131,133}. However, the resistance of PSCs to metformin can be increased by a higher glucose intake, and it was suggested that the tumor microenvironment plays a pivotal role in determining the effect of metformin\textsuperscript{134}. The anti-fibrotic activity of these phytochemicals, as well as their mechanistic actions, are summarized in Table 1.

7. Conclusions

Phytochemicals extracted from plant-based foods have shown multifaceted bioactivity in maintaining human health and disease prevention. In this present review, we have provided insight into several selective phytochemicals that have recently been evaluated for their anti-fibrotic activity, as well as the underlying mechanisms and key molecules involved, which may serve as potent and novel anti-fibrotic agents in targeting PSCs for their mediated CP and PC diseases. PSC activation can be stimulated by cell damage or an inflamed pancreas, inducing the PCs’ proliferation and secretion of cytokines and ECM proteins, supporting fibrosis formation, and stimulating cellular microenvironments that are favorable for the pathologic development of CP and PC. With regard to the reviewed phytochemicals, most studies examining their potential anti-fibrotic activity were carried out using PC models, and rhein and ellagic acid treatments were only tested in CP models. Additionally, curcumin has been tested intensively, compared to other selective phytochemicals. Intriguingly, it was noted that most of the selective phytochemicals exerted similar mechanisms of action, which underpinned their anti-fibrotic action against PSCs in both CP and PC models. Specifically, the selective phytochemicals were shown to deactivate the PSCs by decreasing their proliferation, via the regulation of ERK1/2, P38 MAPK, and the SHH and PI3K/AKT signaling pathways, to suppress PSC migration and fibrogenesis. Comparatively, rhein and ellagic acid treatment deactivated PSCs by reducing the expression of their fibrogenic markers (\(\alpha\)-SMA), and their soluble factors, such as ECM (fibronectin and collagens) and TGF-\(\beta\), which are associated with pancreatic fibrosis. Involvement of the above-mentioned mechanisms supported the potential of these selective phytochemicals to act as novel anti-fibrotic agents for combating CP and PC diseases by targeting PSCs.

Given their potential to serve as new anti-fibrotic agents, based on preclinical studies, it is important to consider a few issues relating to drug delivery and low bioavailability before using them in clinical settings. One of the treatment strategies for improving their therapeutic efficacy is to synthesize their analogs by modifying their chemical structures. Taking curcumin as an example, the curcumin analog L49H37 has been proved to exert potent anti-proliferative activity on PSCs at a concentration that is much lower than curcumin itself. To the best of our knowledge, all the synthetic analogs of the other selective phytochemicals, except eruberin A, have been synthesized and tested, mainly in cancer and other disease models\textsuperscript{135–140}. It is, therefore, crucial to test their treatment efficacy against PSCs in both CP and PC models. Furthermore, curcumin was reportedly subjected to a clinical trial and the advanced PC patients showed good tolerance and attested to its treatment efficacy. However, the issue of low bioavailability may impede its therapeutic efficacy. In recent years, the use of nanoparticle systems, including polymers, liposomes, and some special moieties, have emerged in relation to drug delivery to patients\textsuperscript{141}. With the exception of eruberin A, these selective phytochemicals have been encapsulated into nanoparticles and their therapeutic efficacy against various diseases, mainly cancers, has been tested\textsuperscript{142}. Of these tests, treatment of PC with curcumin, metformin, and ellagic acid showed promising outcomes\textsuperscript{143–145}. Given the economic and non-toxic properties of the selective phytochemicals, as well as their promising treatment outcomes with regard to PSCs in CP and PC models, further investigation of potential plants or their derived compounds should be encouraged, in order to isolate, identify, and evaluate their benefits, with the aim of discovering more potential anti-fibrotic agents to treat patients with CP-associated PC.

Acknowledgments

This work was supported by two research university grants, from University of Malaya (grant No. GPF002C-2018, Malaysia) and MAHSA University (grant No. RP165-05/19, Malaysia).
Author contributions
Puvanesswaray Ramakrishnan, Wei Mee Loh, Srinivasa Reddy Bonam, and Yuan Seng Wu contributed to the writing. Yuan Seng Wu conceived and designed the manuscript while, Yuan Seng Wu, Maw Shin Sim, Srinivasa Reddy Bonam, and Subash C.B. Gopinath revised the manuscript. Ismail M. Fareez, Rhanyee Mac Guard, and Maw Shin Sim provided vital guidance and insight for the writing.

Conflict of interests
The authors have no conflicts of interest to declare.

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