Looking into Limoncello: The Structure of the Italian Liquor Revealed by Small-Angle Neutron Scattering

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ABSTRACT: Limoncello, the Italian liquor based on lemon essential oils, is becoming increasingly popular around the world. This digestive is not only an iconic representative of Italian food culture, but it is also a complex colloidal system, made of essential oils, ethanol, sucrose, and water. Smell, aroma, taste, and appearance of Limoncello do, of course, depend on the components, in particular on the peculiar essential oil mixture. Accordingly, several studies are available in the literature investigating the composition of various Limoncellos. However, the microscopic structure plays an equally important role when it comes to the sensory properties of food and beverages. In this work, small-angle neutron scattering was used to probe the microscopic structure of Limoncello, revealing the presence of spontaneously formed 100 nm-sized droplets over a large range of composition and temperature. The results are not limited to this famous drink but can be extended to the rapidly developing formulations based on water-insoluble oils, water, and alcohols.

INTRODUCTION

It is no exaggeration to say that almost every festive meal in southern Italy is crowned with a frozen glass of Limoncello, the famous Italian liquor renowned for its flavor and neon-yellow color. The traditional recipe foresees that the citrus zest, obtained by scraping the outer part of the lemon peels, is kept macerating in 95% vol ethanol for several (4–8) weeks. The zest, or flavedo, contains most of the essential oils giving rise to the characteristic taste and color of the liquor. The ethanolic extract is then filtered and diluted with a sucrose/water syrup previously prepared. Usually, the liquor contains ca. 30% vol of alcohol and ca. 20% vol of sucrose. However, the preparation procedure and exact final composition vary from family to family. Limoncello is also largely produced on an industrial scale, with an estimate of 15 million liters commercialized in 2003.1 Three million liters were produced using the IGP lemon of Sorrento in 2017, according to the local consortium.

To assess the genuineness and quality of commercial Limoncellos, several analytical procedures have been developed.1–5 Different studies performed on commercial Limoncellos reveal that, in several cases, the final product has never been in direct contact with the lemon peel, but it is rather a mixture of citrus essential oils, alcohol, water, and sucrose.2,4 Multiple reasons are found for the rare employment of the traditional and time-consuming procedure, in addition to the obvious economic aspect. The composition of the lemon essential oil, extracted from the citrus flavedo, can vary significantly as a function of the harvest period and the growth location,6 making a production of the liquor with constant organoleptic properties difficult. Moreover, Crupi et al. found evidence that commercial liquors were prepared using terpeneless oils or essential oils enriched with citral,4 possibly to avoid or reduce oxidative processes and to enhance the liquor flavor and shelf-life.

In this work, the focus is put on the microscopic structure of Limoncello, rather than on its chemical composition. While the chemical composition is essentially determining the flavor of liquor, the microstructure of food and beverages strongly affects the sensory perception during tasting, on its shelf-life, and on the optical appearance.7–12 From a physicochemical
perspective, the Limoncello is a mixture of hydrophobic essential oils extracted from the lemon flavedo, ethanol, water, and sucrose. Similar ternary, or pseudo-ternary mixtures, formed by two partly miscible liquids (the water/sucrose syrup and the essential oil) and a common solvent, such as ethanol, exhibit strong composition fluctuations close to the phase-separation boundary,\textsuperscript{3,13–17} that is, the mixtures become inhomogeneous both at the microscopic and macroscopic level, thus turning often turbid. This effect is commonly observed upon water addition to anise-flavored spirits, such as pastis, absinthe, raki, or ouzo, hence the name of “ouzo-effect”\textsuperscript{18}. Furthermore, the effect is not limited to alcoholic beverages, it is also exploited for the development of reaction media and promising drug delivery systems.\textsuperscript{19–22} The size of the clusters strongly depends on the composition of the mixture and varies between a few nanometers in the so-called pre-ouzo region (thermodynamically stable and transparent) to micrometer-sized domains which exhibit slow phase separation in the ouzo region. It is noteworthy that there exists different designations in the literature to describe these systems, the two most common being surfactant-free emulsions, to distinguish them from the conventional cases of two immiscible liquids stabilized by surfactant molecules\textsuperscript{20,23} or ultraflexible microemulsions, to highlight the exceptionally low rigidity of the droplet interface.\textsuperscript{24,25}

The characterization of the microstructure of Limoncello was performed using small-angle neutron scattering (SANS), a very powerful technique for studying the structure of soft condensed matter in the range of Å ($10^{-10}$ m) to μm ($10^{-6}$ m), which was already successfully employed in similar ouzo-systems.\textsuperscript{21,25,26} However, because of the high complexity of real food products, the use of scattering techniques is mostly limited to simplified, abstracted representations thereof.\textsuperscript{27–30} In this study, the microstructure of “real” Limoncello samples made following a traditional receipt (apart from the use of deuterated compounds) is probed and compared to a model system comprising pure citrus essential oils. Isotopic labeling is a prerequisite for several techniques used in colloidal science, such as neutron scattering or nuclear magnetic resonance, and the effect of deuteration on the physicochemical properties of emulsion systems was investigated before.\textsuperscript{31,32} While phase boundaries are slightly affected by the isotopic labeling, negligible effects on the emulsion morphology, that is, average size and size distribution, have been reported.\textsuperscript{31,32}

\section*{RESULTS}

Before the small-angle scattering data can be analyzed and interpreted, the composition of the ethanolic extract has to be accessed. The essential oil extracted from citrus flavedo has a complex composition, and more than 60 volatile compounds have been identified.\textsuperscript{6} The main components of the essential oil are monoterpenes, oxygenated compounds such as aldehydes, and sesquiterpenes.\textsuperscript{1,3,3,4,3,3} Monoterpenes constitute 90\% wt of the mixtures and are the main components of the citrus essential oil, with the most representative compounds being limonene, $\beta$-pinene, and $\gamma$-terpinene. Oxygenated compounds, such as decanal, geraniol and neral—two isomers of citral—make up to 10\% of the mixture. Sesquiterpenes, such as $\beta$-bisabolene, constitute ca. 1\% of the essential oil mixture.

The amount of essential oil in the ethanolic extract was evaluated by proton nuclear magnetic resonance, $^1$H NMR (see the spectrum in Figure 1). A quantitative analysis was performed using tetramethylsilane (TMS) as an internal standard. Different intense peaks are observed: one at 0 ppm of the internal reference TMS; two broad peaks centered at 4.6 and 5.3 ppm, which are ascribed to the water proton and to the hydroxyl proton of ethanol, respectively;\textsuperscript{34,35} two singlets at 1.1 and 3.6 ppm, ascribed to residual light hydrogen of the ethanol (see the $^1$H NMR spectrum of deuterated ethanol in the Supporting Information); finally, a large number of peaks between 3.0 and 4.0 ppm and between 0.5 and 2.5 ppm, ascribed to the main components of the essential oil, are found.\textsuperscript{36,37} Accordingly, the amount of “essential-oil-protons” can be estimated by the integral over these two regions, and the oil content in the ethanolic extract estimated assuming a generic chemical formula of C$_{10}$H$_{16}$O$_x$ typical of terpenes. With this assumption, an oil content in the extract of ca. 0.28\% wt is estimated. Moreover, from the integrals of the two broad peaks at 4.6 and 5.3 ppm, the water content in the extract is estimated at 15\% wt, in good agreement with the weight loss of the lemon peel after infusion in alcohol and with literature data.\textsuperscript{38} It has to be remarked that the $^1$H NMR spectrum provides only an estimate of the amount of solubilized oil in the complex mixture, for example, the alkene hydrogens are buried under the large signal from hydroxyl groups.

Once the chemical composition of the ethanolic extract was identified, the mesoscopic structure of mixtures formed by the lemon essential oil, water, ethanol, and sucrose was studied by SANS. Samples along the dilution line of the ethanolic extract with the water/syrup were prepared (LM1 to LM5); the effect of the sucrose content in the liquor was also probed by preparing samples with constant ethanol/oil content and with variable water and sucrose amounts (LM5 to LM9). The location of the investigated samples within the pseudo-ternary phase diagram is shown in Figure 2. Furthermore, the effect of temperature within the range of 266 K (−7 °C) to 298 K (25 °C) was studied, to evaluate potential structural changes during consumption: from the chilled glass to our warm mouth. The ternary phase diagram water/ethanol/lemon essential oil is presented in Figure 2 and the phase diagram syrup/ethanol/lemon essential oil is provided in Figure S6 in the Supporting Information. Both phase diagrams are almost identical and in very close agreement with the phase diagram obtained with the pure limonene.\textsuperscript{39} The very large two-phase domain is due to the poor solubility of limonene in water (0.01 g/L). The samples are prepared in the water-rich part of the phase diagram and contained less than 0.3\% in mass of lemon oil.
A representative set of SANS curves is given in Figure 3, where the scattering intensity \( I(q) \) is plotted as a function of the scattering vector \( q \). All other recorded patterns are given in the Supporting Information. The ethanolic extract (sample LM1), located in the ethanol-rich corner of the phase diagram, is homogeneous at a molecular scale, as proven by the very weak scattering intensity. As expected, the scattering intensity increases over 4 orders of magnitude upon addition of water, as evidenced in the left-hand-side plot of Figure 3. All curves look qualitatively very similar, with the main feature being an extended \( q^{-4} \) power-law, typical for two-domain systems with a sharp interface. Typical examples are emulsions, marble, alloys presenting precipitate formation, or foams.\(^{29,40} \) The observed scattering arises from the variation of the atomic composition at the interface between the domains and the suspending medium. Performing a Porod analysis allows us to determine the specific amount of interface \( \Sigma \) present in the system:\(^{43} \)

\[
\Sigma = \frac{1}{2\pi \Delta \rho^2} \lim_{q \to \infty} I(q)q^4
\]

where \( \Delta \rho \) is the scattering length difference between the domains and the medium. In the approximation that the oil droplets are spherical, the specific interface is related to the Porod radius \( R_P \):

\[
\Sigma = \frac{3\phi}{R_P}
\]

with \( \phi \) being the volume fraction of the droplets. For the calculation of \( \Delta \rho \) and \( \phi \), we assumed that the essential oil droplets are suspended in a water/ethanol/sucrose mixture (see the Supporting Information for details). The scattering curves, however, present more features than a simple \( q^{-4} \) power law. In fact, an oscillation is visible in the low-\( q \) part of the \( I(q)q^4 \) representation (right-hand-side plot of Figure 3). This oscillation arises from the finite size of the essential oil droplets. Accordingly, the whole scattering pattern was described with a polydisperse sphere model, given by the following expression.
\[ I(q) = \Delta \rho^2 \int_0^{\infty} N \text{Pdf}(r)V(r)P(q, r)dr \text{ with } P(q, r) \]

\[ = \left[ \frac{3 \sin qr - qr \cos qr}{(qr)^3} \right] \]  

(3)

with \( N \) being the number of droplets per unit of volume and \( \text{Pdf}(r) \) the probability density function describing the relative abundance of a droplet of radius \( r \) and volume \( V(r) = 4/3\pi r^3 \). Herein, a lognormal distribution was chosen to describe the polydisperse sizes of the emulsion droplets. The probability density function, defined by two parameters, \( \mu \) and \( \sigma \), is given by

\[ \text{Pdf}(r) = \frac{1}{\sigma \sqrt{2\pi}} \exp \left[ -\frac{(\ln r - \ln \mu)^2}{2\sigma^2} \right] \]  

(4)

The main advantage of the use of a lognormal distribution for the description of SANS data from polydisperse systems is that negative values for the droplet radius are not allowed and that the function can be used without any need of introducing cut-offs.

In summary, there are two procedures to obtain the size of the lemon essential oil droplets in the Limoncello mixtures: in the former one, the size is deduced from the scattering at the oil/medium interface, assuming the oil is completely separated from the solution; in the latter one, the patterns are analyzed with a model of polydisperse spheres, whereby the parameters \( \mu \) and \( \sigma \) describing the size distribution and the droplet volume fraction \( \phi \) are optimized. In the right-hand-side plot of Figure 3, the Porod constant is indicated with the dashed line, while the polydisperse sphere model is indicated by a full line. Further details, calculated curves, and the obtained parameters are given in the Supporting Information. The Porod radius, calculated with eq 2, can be directly compared to the ratio of the third and second moment of the size distribution obtained from the full SANS analysis, given by

\[ R_p \equiv \frac{\langle R^3 \rangle}{\langle R^2 \rangle} = \mu \exp(2.5\sigma^2) \]  

(5)

The calculated radii are given in Figure 4 as a function of the water content, for all samples recorded at the different temperatures. The temperature-dependence of the volume fraction of the components and of the scattering contrast was taken into account (see the Supporting Information for full details). The results from the Porod analysis indicate that the total amount of interface in the systems increases with increasing water content, which results, within the assumption of full microphase-separation of the essential oil from the medium, in a decreasing radius of the droplets. In contrast, the full analysis of the scattering patterns indicates that the size of the droplets is rather constant of ca. 100 nm for all investigated samples, and that the volume fraction of the oil droplets increases with increasing water content (see Figure S14). Both analyses deliver the same, consistent results for the samples with the highest water content (\( >55\% \) wt).

The discrepancy between the droplet sizes obtained from the Porod analysis and those obtained from the fit with a polydisperse sphere model indicates that, at low water content, a significant amount of essential oil is molecularly dissolved in the solution and is not participating in the formation of the emulsified droplets. A wrong assumption on the volume fraction of oil droplets leads therefore to an overestimation of

\[ Q^* = \int_0^{\infty} I(q)q^2 dq = 2\pi^2 \Delta \rho^2 \phi(1 - \phi) \]  

(6)

The obtained volume fractions of oil droplets are fully consistent with the values obtained from the fit of the data with the polydisperse sphere model (see Figure S17 in the Supporting Information). This consistency further corroborates the validity of the results from the polydisperse sphere model analysis.

The study also reveals that temperature does not affect the microstructure of the Limoncello in the studied temperature range (\( -7 \) to \( 25 \) °C), except that the solubility of the essential oil in the mixed solvent increases with increasing temperature, in agreement with previous studies on (+)-limonene/water/ethanol mixtures. Surprisingly, the presence of large amounts of sucrose, up to 25% wt, does not alter the microstructure of the Limoncello samples. This observation is further confirmed by the fact that the presence of sucrose minimally affects the
phase behavior of the mixtures of essential oil/ethanol/water, as shown in the Supporting Information in Figure S6.

The size of the self-emulsified oil domains is relatively small when compared to other ternary oil/water/ethanol mixtures, where typical sizes of larger than 500 nm are found.\textsuperscript{25,29} The small size can be explained by the presence of surface active molecules, for example, lipids or proteins, extracted during the maceration of the lemon zest. It is likely that these components stabilize the oil/water interface leading to smaller oil droplets.\textsuperscript{30,47} To verify this hypothesis, additional SANS experiments were performed on pseudo-Limoncello samples prepared by mixing water, sucrose, ethanol, and lemon essential oil, thus avoiding the direct contact of ethanol and the lemon zest (Figure 5). The data were recorded on an extended $q$-range, thus covering also the Guinier region at low-$q$. They were analyzed with the same models described earlier in the text, the droplet radii are given in Figure 4 and show excellent agreement with the results from the “real” Limoncello samples. The finding rules out that the exceptionally small sizes found in these mixtures are caused by the presence of non-oily, surface-active components, such as lipids and proteins. In addition, the good agreement corroborates the correctness of the estimate of the essential oil content in the ethanolic extract used to prepare the “real” Limoncello samples and that the missing Guinier plateau does not lead to a misinterpretation of the scattering data.

Finally, the interfacial tension between lemon essential oil and water/sucrose/ethanol was determined by pending drop experiments. A low, but not extraordinary low, value of the interfacial tension of 4−5 mN/m (see the last paragraph of the Supporting Information) was found. Similar values were found between the oil-rich and oil-poor phase in water/ethanol/1-octanol mixtures.\textsuperscript{13}

### DISCUSSION

The microstructure of the Limoncello samples prepared in this work is surprisingly stable. Except for the ethanolic extract, in all samples the presence of polydisperse essential oil droplets with an average size of ca. 100 nm is demonstrated. Neither the presence of sucrose nor changes in temperature have an effect on the microstructure of the investigated Limoncello samples. The size of the oil-domains of ca. 100 nm is surprisingly small when compared to the typical size of ouzo-systems which ranges from 500 nm to several micrometers.\textsuperscript{25,29} The small size of the Limoncello droplets does not arise from the presence of surface-active contaminants, for example, lipids or proteins extracted during the maceration of the lemon zest. It is more likely that the complex mixture of the essential oil is able to stabilize droplets, whose size is intermediate between classical microemulsions (2−20 nm) and metastable ouzo-systems or classical oil/water emulsions (>500 nm).

While the size of the domains remains basically constant with temperature and composition, a continuous evolution in the amount of essential oil which participates in the droplet formation is observed. Sample LM1, with a water content of 15% wt (ethanol content of ca. 85% wt), is fully homogeneous, and no concentration fluctuations were observed. On the contrary, a complete microphase separation of the essential oils from the continuous phase is obtained for a water content larger than 60% wt. Interestingly, high-quality Limoncello samples have usually a composition close to the LMS sample, with a water content of ca. 50% wt. At this composition, 1/3 of the essential oil is molecularly dispersed in the continuous phase (Figure S16). The microstructure of Limoncello is fundamental for the different properties of the liquor: the droplet size will affect the optical appearance and time-stability of the drink. Moreover, droplet size and distribution of the active fragrance within the mixture is expected to have a pronounced effect on the vapor pressure of the active perfume molecules, and therefore on the smell of the liquor. In a recent work, Zemb et al. have shown that the vapor pressure of a model fragrance, styrylal acetate, dissolved in a water/ethanol mixture, reaches a maximum close to the boundary between the meta-stable ouzo region and the biphasic domain.\textsuperscript{17} A lower activity of the fragrance is observed in the homogeneous region, where it is well-dissolved as well as in the biphasic domain, where the active components are trapped in micrometer sized droplets. Further studies evidenced that the solubilization of the fragrance in micellar aggregates reduces their evaporation rate,\textsuperscript{48} in agreement with the finding that the evaporation of the active component from a surfactant stabilized air/water interface is limited by its diffusion through a depleted region close to the surface.\textsuperscript{49} According to these findings, it seems that Limoncello samples with a composition close to LMS present all the needed properties for a good digestive: the rather large fraction of free essential oils ensures a strong smell and taste of the liquor, which made it so popular. The typical droplet size of approx. 100 nm gives rise to the typical opalescent appearance but ensures both long-time stability and a rapid exchange between molecularly dissolved essential oil and small droplets. Noteworthy, a homemade Limoncello, prepared with hydrogenated water and ethanol close to the composition of LMS and the same lemons of the experiment reported herein, had an excellent taste.

### CONCLUSIONS

In this work, a structural analysis of real Limoncello samples was performed using SANS, shedding light for the first time on the mesoscopic structure of this very famous Italian liquor. The results show that the system is composed of polydisperse oil droplets dispersed in a continuous water/ethanol/sucrose medium. These droplets, whose size of ca. 100 nm shows little variation with temperature and overall composition, are in equilibrium with molecularly dissolved essential oil. Interestingly, at a system composition close to that of commercial...
Limoncellos, ca. 1/3 of the essential oil is molecularly dissolved and in equilibrium with the sub-micrometer large droplets. It is expected that the overall sensory experience of Limoncello: its smell, taste, and optical appearance, results from this delicate equilibrium.

However, the relevance of the study goes beyond the use of neutron scattering for the characterization of a tasty digestive. Essential oils, especially limonene, are experiencing quickly growing applications. In addition to the traditional use as flavors and fragrances, the oils extracted from citrus fruits are becoming increasingly relevant, for example, as green solvents or as mosquito-repellent formulations or as a medium for enzymatic and chemical reactions. To understand these findings open different configurations were used, with a wavelength of $\lambda = 6.0$ Å, sample-to-detector distances (SD) of 1.2, 8, and 34 m and collimation of 4, 8, and 34 m, respectively, covering a q-

### Table 1. Composition of Samples Used in This Work

| sample name | weight fraction | volume fraction at 298 K |
|-------------|----------------|-------------------------|
|             | oil (%) | $d_4$-EtOH (%) | sucrose (%) | water (%) | oil (%) | EtOH (%) | sucrose (%) | water (%) |
| LM1         | 0.28    | 84.7           | 0.0         | 15.0      | 0.30    | 86.1      | 0.0         | 13.6      |
| LM2         | 0.21    | 62.2           | 8.9         | 28.7      | 0.24    | 67.9      | 5.4         | 26.4      |
| LM3         | 0.14    | 42.1           | 16.8        | 41.0      | 0.18    | 49.3      | 10.9        | 39.6      |
| LM4         | 0.07    | 21.4           | 24.9        | 53.6      | 0.10    | 27.0      | 17.5        | 55.4      |
| LM5         | 0.10    | 31.7           | 20.8        | 47.3      | 0.14    | 38.6      | 14.1        | 47.2      |
| LM6         | 0.10    | 31.2           | 16.1        | 52.6      | 0.15    | 37.5      | 10.8        | 51.7      |
| LM7         | 0.10    | 31.7           | 10.9        | 57.3      | 0.15    | 37.4      | 7.2         | 55.3      |
| LM8         | 0.10    | 30.9           | 6.4         | 62.6      | 0.15    | 36.0      | 4.1         | 59.7      |
| LM9         | 0.10    | 31.6           | 0.0         | 68.3      | 0.15    | 36.1      | 0.0         | 63.8      |
| LM-E01      | 0.41    | 87.9           | 0.0         | 11.7      | 0.44    | 89.9      | 0.0         | 9.7       |
| LM-E02      | 0.31    | 66.2           | 8.0         | 25.6      | 0.36    | 72.2      | 4.8         | 22.6      |
| LM-E03      | 0.21    | 44.0           | 16.1        | 39.7      | 0.26    | 51.6      | 10.5        | 37.7      |
| LM-E04      | 0.15    | 33.0           | 20.2        | 46.7      | 0.20    | 40.1      | 13.7        | 46.1      |
| LM-E05      | 0.10    | 22.3           | 24.1        | 53.5      | 0.14    | 28.2      | 16.9        | 54.8      |

1LMx series are samples prepared from the ethanolic extract, and the amount of essential oil was evaluated from the $^1$H NMR spectrum shown in Figure 1. LM-EOx sample series denote samples prepared using cold-pressed lemon essential oil. The conversion from weight fractions to volume fractions was performed using the density values given in Table 2.

### MATERIALS AND METHODS

#### Materials and Sample Preparation

The samples were prepared using deuterated water (D content $>$99.8%) from Eurisotop (Gif-sur-Yvette, France), $d_4$-ethanol (99% purity) from Deutero (Kastellaun, Germany), commercial sucrose, and nontreated lemons from Sicily bought at a local market. The ethanolic extract was prepared by immersing 2.5 g of the outer part of lemon skins, the flavedo, in 12.6 g of pure fully deuterated ethanol. The lemon peels were kept macerating in the absolute deuterated alcohol for 4 weeks. After this period, a sucrose syrup was prepared by dissolving 6.6 g of sucrose in 13.2 g of D$_2$O. Samples were prepared by addition of the syrup and/or D$_2$O to the ethanol extract. Gentle shaking is sufficient for homogenization. The final composition of the samples is given in Table 1. The composition of the ethanolic extract was determined via $^1$H NMR as described before. Further samples were prepared using cold-pressed lemon essential oil from Arkpharma (Carros, France).

#### Phase Diagram Determination

The phase diagrams were visually determined. Binary mixtures of cold-pressed lemon essential oil and ethanol at different ratios were prepared by weighing the proper amounts of chemicals in 5 mL glass tubes. Small amounts of water, or of a 33% wt sucrose solution, were added gradually. The binary essential oil/ethanol mixtures are perfectly transparent. The transition to the pre-ouzo region is identified as the first signs of opalescence are detected. Further addition of water, or of a 33% wt sucrose solution, causes the system to turn highly turbid. The transition from translucent to turbid delimits the metastable ouzo-region. In the oil-rich part, further addition of water induces a sudden phase separation of the sample corresponding to the binodal line. The 2-phase domain is more difficult to observe in the oil-poor domain, where the phase separation only appears after several hours at rest.

#### Methods

Small-Angle Neutron Scattering. SANS patterns were recorded on D11 at the Institut Laue–Langevin (ILL) in Grenoble, France. Raw and reduced data are available at doi:10.5291/ILL-DATA.INTER−372. Three different configurations were used, with a wavelength of $\lambda = 6.0$ Å, sample-to-detector distances (SD) of 1.2, 8, and 34 m and collimation of 4, 8, and 34 m, respectively, covering a q-
range of 0.015−5 nm−1, where \( q = 4\pi \sin(\theta/2)/\lambda \) is the magnitude of the scattering vector and \( \theta \) is the scattering angle. An additional configuration was used for the samples prepared with cold-pressed lemon essential oil, with a sample-to-detector distance of 39 m, collimation of 40 m, wavelength \( \lambda = 12 \) Å, and removing the beam-stop, reaching a minimum \( q \) value of 0.004 nm−1. The differential cross sections in absolute scale were obtained by comparison with the scattering from a 1 mm water sample. The analysis was performed using the values of densities and scattering length densities \( \rho \) of the compounds reported in Table 2.

**Table 2. Density and Neutron Scattering Length Densities (\( \rho \)) at 298 K of the Components of the Mixtures Studied in This Work**

| compound                  | chemical formula | density/g cm\(^{-3}\) | \( \rho/10^{-4} \) nm\(^{-2}\) |
|---------------------------|------------------|------------------------|---------------------------------|
| light water               | H\(_2\)O         | 1.0                    | -0.56                           |
| heavy water               | D\(_2\)O         | 1.1                    | 6.33                            |
| \( d_x \) ethanol         | C\(_x\)D\(_x\)O   | 0.89                   | 6.10                            |
| sucrose (H)               | C\(_x\)H\(_{2x}\)O\(_y\) | 1.6                | 1.7                             |
| lemon essential oil (terpenes) | C\(_x\)H\(_{1x}\) | 0.84                   | 0.25                            |

“Density values of limonene and sucrose in water/ethanol mixtures are taken from refs.\(^*^\),\(^{**}\), respectively.

\(^*^\)H NMR. \(^1\)H NMR spectra were recorded on a 700 MHz Bruker AVANCE III HD spectrometer. Experiments were carried out at 25 °C.

**Interfacial Tension.** The interfacial tension between the citrus essential oil and the water/sucrose/ethanol solution was measured at room temperature using a drop shape analyzer DSA10 from Krüss. Drops of oil in heavy water were created using an inverted blunt needle of 0.72 mm in diameter immersed in a 10 mm (thickness) × 20 mm (width) × 40 mm (height) optical glass cuvette. The interfacial tension was calculated from the drop profile by the Krüss software using the Young–Laplace equation.

© ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acsomega.8b01858.

Additional sample characterization, further phase diagrams, further SANS patterns and their interpretation, and interfacial tension results (PDF).

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Notes

The authors declare no competing financial interest.

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