Topical use of tranexamic acid: Are there concerns for cytotoxicity?

Ioannis Gkiatas, Aristeidis-Panagiotis Kontokostopoulos, Spyridon E Tsirigkakis, Ioannis Kostas-Agnantis, Ioannis Gelalis, Anastasios Korompilias, Emilios Pakos

Abstract

Tranexamic acid (TXA) has revolutionized modern blood management in orthopaedic surgery, especially in total joint arthroplasty, by significantly reducing blood loss and transfusion rates. It is an antifibrinolytic agent and a synthetic derivative of the amino acid lysine, which can inhibit the activation of plasminogen and the fibrin breakdown process. The administration of TXA can be intravenous (IV), topical, and oral. In patients where the IV administration is contraindicated, topical use is preferred. Topical administration of the drug theoretically increases concentration at the operative site with reduced systemic exposure, reduces cost, and gives the surgeon the control of the administration. According to recent studies, topical administration of TXA is not inferior compared to IV administration, in terms of safety and efficacy. However, there are concerns regarding the possible toxicity in the cartilage tissue with the topical use of TXA mainly in hemiarthroplasty operations of the hip, unilateral knee arthroplasties, total knee arthroplasties where the patella is not resurfaced, and other intraarticular procedures, like anterior cruciate ligament reconstruction. The purpose of the present review is to present all the recent updates on the use of TXA focusing on the toxicity on chondrocytes and the articular cartilage that may or may not be provoked by the topical use of TXA.

Key Words: Tranexamic acid; Topical use; Cytotoxicity; Orthopaedic surgery

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Core Tip: Tranexamic acid (TXA) is an antifibrinolytic agent and is associated with decreased blood loss in surgical procedures. It is widely used in major orthopaedic procedures in order to decrease blood transfusion needs. TXA can be administered intravenously; however, topical administration of the drug increases concentration at the operative site. There are concerns that this increased concentration may cause toxicity in the cartilage tissue. In this review, we present the recent literature regarding the cytotoxic effects of the topical administration of TXA.

INTRODUCTION

Tranexamic acid (TXA) is an antifibrinolytic agent and a synthetic derivative of the amino acid lysine, which can inhibit the activation of plasminogen and the fibrin breakdown process[1]. Plasminogen is a glycoprotein pro-enzyme which is produced in the liver. Plasminogen molecules are folded into loops and lysine binding sites are located at the end of these loops. These lysine binding sites on plasminogen are connected with lysine residues on fibrin. Tissue plasminogen activator is also released by endothelial cells and binds to fibrin, causing the conversion of plasminogen to plasmin. Plasmin cleaves fibrin into small fragments like D-dimers. This abruption causes the exposure of more lysine residues to plasminogen, therefore accelerating fibrinolysis[2]. The fact that TXA is a synthetic derivative of the amino acid lysine causes the attachment of the lysine residues to the lysine binding sites on plasminogen, thus preventing the binding of plasminogen to fibrin and blocking its activation to plasmin[2,3]. At higher concentrations, TXA is also a noncompetitive inhibitor of plasmin, therefore it stabilizes the clot formation[4]. Numerous studies conducted in animal tissue samples showed that TXA increases thrombus formation in a dose-dependent manner[5]. Plasminogen receptors are also located on endothelial cells, monocytes, lymphocytes, and platelets, which indicates the role of plasminogen in inflammation. Thus, TXA may have another role, by inhibiting this procedure, too[6].

TXA is associated with decreased blood loss in surgical procedures and inhibits the activation of plasminogen to a greater extent in comparison with another major antifibrinolytic agent, epsilon-aminocaproic acid (EACA)[6,7]. TXA does not seem to be associated with a major difference in transfusion rates when compared with EACA[6].

After administration of a mean TXA intravenous dose of 10 mg/kg on healthy individuals, peak concentration of the drug is observed at 1 h after administration and its half-life is approximately 2 h, with 90% excreted at 24 h. TXA remains in serum for approximately 8 h and in tissues for 17 h. Nevertheless, these pharmacokinetic factors may be different in patients that have been through major trauma and the efficacious TXA levels in these patients may differ with those in healthy individuals[7].

Nevertheless, there are concerns that topical use of TXA may be toxic for chondrocytes and surrounding soft tissue, as stated in numerous studies[3,4,8-11]. The current manuscript is a narrative review and its purpose is to present the recent literature regarding the effect of the topical use of TXA, whether it can cause toxicity on chondrocytes and articular cartilage, and if there is a standard safe level of the drug at which it can be harmless and efficacious.

USE OF TRANEXAMIC ACID (TXA) IN ORTHOPEDIC SURGERY

TXA can be effective to reduce blood loss in major surgical procedures and severe traumatic conditions. Thus, TXA can decrease the blood transfusion needs without increasing the risk of thromboembolic events. Many studies have supported its use in major orthopedic procedures[8,9,12-17]. Fillingham et al [9] showed that TXA administration can reduce blood loss during total knee arthroplasty by approximately 100 mL, compared to placebo. Yi et al[8] have also shown that a group of patients that received TXA after total hip arthroplasty had a lower blood transfusion rate in comparison with the placebo group. These patients also had a lower mean total blood loss varying form approximately 200 mL to 350 mL in comparison with the placebo group[8]. Also, Li et al[12] proved that a group of patients treated with TXA after spinal surgery had significantly lower blood loss and the number of patients that required blood transfusion was notably reduced, irrespective of using TXA at high or low dosage[10]. Generally, TXA has demonstrated efficacy in hip[8], knee[9], and shoulder[14] arthroplasties as well as in spinal surgery[12] and anterior cruciate ligament reconstruction[13]. Farrow et al[15] found moderate quality evidence that TXA administration can reduce blood transfusion rate in patients undergoing hip fracture surgery, so the effectiveness of TXA in this surgical procedure is under investigation.

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Numerous studies have also proved its efficacy during trauma operations based on data from the CRASH-2 trial and showed that TXA can reduce the mortality rates in trauma patients due to bleeding [16,17]. Few studies suggested the lack of potential benefit from the use of TXA in major orthopaedic procedures. For example, Lack et al. [18] concluded that there are no significant differences in blood loss and transfusion rates between TXA and placebo. However, the number of the studies supporting this conclusion is very limited and this heterogeneity can potentially be attributed to the variety of methods used for calculating the blood loss.

**CONTRAINDICATIONS**

Despite its wide use in major orthopaedic operations, TXA cannot be used in certain cases. More specifically, there are absolute and relative contraindications for TXA administration. Hypersensitivity and allergy to TXA as well as active thromboembolic disease and fibrinolytic conditions with consumption coagulopathy are the absolute contraindications for TXA [19].

The relative contraindications must consider the association between risk and benefit of TXA [19]. Relative contraindications include renal dysfunction, thrombosis disorders, and preexisting coagulopathy or oral anticoagulants [19]. TXA is also contraindicated in patients with a high risk of seizures [19,20]. Pregnancy is another contraindication, but TXA can be administered when it is vitally indicated [20].

**TXA ADMINISTRATION AND SAFE LEVELS**

TXA can be administrated through the intravenous (IV) and oral route as well as topical. However, IV and oral TXA administration is associated with an increased risk of systematic complications, such as thromboembolic events [21]. Ker et al. [22] showed that although IV TXA can minimize blood loss during surgeries, it is arguable whether it can cause major thromboembolic events in the future. In addition, since TXA is a derivative of the amino acid lysine and goes through ionization in physiologic environment, it is expected that its absorption through membranes may be inadequate. Therefore, its bioavailability will be moderate and higher doses of the drug may be required in order to reach the desirable levels of TXA in articular cartilage, thus causing major systematic complications [23]. On the other hand, topical administration of the drug may have many benefits like increased concentration at the operative site with reduced systemic exposure, cost reduction, and surgeon control [24]. It has been also shown that topical administration is not inferior to IV administration, in terms of safety and efficacy [25,26]. Abdel et al. [26] and Xu et al. [27] showed that IV administration of TXA has no advantage over topical administration of the drug as they both caused lower rates of blood transfusion and these two groups had no clinically significant differences. However, many authors investigated the combined route of administration of TXA, suggesting that combining both topical and IV administration of the drug could potentially have a beneficial effect in reducing blood loss during major orthopaedic procedures [28]. It is not clear yet whether this combined route can lead to a better result, concerning the blood loss and the blood transfusion rates [29]. Dong et al. [28] suggested that combined topical and intravenous administration can further reduce blood loss while Li et al. [29] did not support this conclusion and suggested that this technique has no additional benefits.

TXA can be administered topically using various techniques [30]. Intra-articular or peri-articular application is the most common technique of TXA administration [30]. Some surgeons perform a perioperative wash of the drug in order to avoid its topical side effects [30]. Clamping drainage is another proposed technique which creates an intra-articular tamponade, thus producing temporary haemostasis and increasing the tissue contact time with the drug [31].

TXA was supported as a safe drug for topical use in a few studies (Table 1). In the study of Degirmenci et al. [1], an in vivo study on experimental animals, TXA seemed to have a significant contribution to the healing of osteochondral defects. This was observed not only macroscopically but also histopathologically, where samples treated with TXA had clearly better recovery than the control groups. More specifically, the macroscopic examination of the animal samples was performed by observing any abnormality of the location, color, shape, and size of the tissue in situ. 4 and 8 wk after TXA administration. The group that received TXA seemed to have better recovery, after both 4 and 8 wk, in comparison with the control group. Histopathologically, the group that received TXA had higher regeneration rate of subchondral tissue and cartilage tissue repair rate than the control group. No problems were mentioned in terms of safety of the drug. Ambra et al. [3], in a study carried out on experimental animal samples, also supported the efficacy and safety of the drug. Images of full-thickness cartilage sections showed a similar number of live cells, as well as a minimal presence of dead cells, no matter the duration of incubation and the concentrations of TXA administered. Moreover, this study suggested that TXA doses up to 4 mg/mL for up to 6 h did not seem to have any detrimental effect on cell viability. This study was carried out on experimental animal tissue; however, samples of the animals that were chosen are the closest to the human samples. Also, TXA doses were adapted.
Table 1 Characteristics and outcomes of studies that considered tranexamic acid as a safe drug

| Ref.                     | Tissue                                | Doses                        | Time of exposure | Outcome                              |
|--------------------------|---------------------------------------|------------------------------|------------------|---------------------------------------|
| Ambra et al[1], 2017     | Animal samples (Yucatan minipigs)      | 1, 2, and 4 mg/mL.           | 1, 3, and 6 h    | No effect on cell viability           |
| Marmotti et al[4], 2016  | Human samples (cartilage and synovial tissues) | 7 mg/mL in both cartilage and synovial tissues | 2 wk for cartilage tissue and 48 h for synovial tissue | No effect on synoviocyte morphology |
| Degirmenci et al[1], 2019 | Animal samples (rabbits)               | Not mentioned                | Not mentioned    | Healing rate was improved after TXA administration and no detrimental effects were observed |
| Bolam et al[29], 2020    | Human and animal samples               | 1, 2, 4, 20, 25, 50, and 100 mg/mL | 10 min, 1 h, 2 h, 4 h, 24 h, and 48 h | No effect on cell viability and morphology when treated at doses lower than 20 mg/mL. |
| Wang et al[33], 2021     | Human samples (fibroblast cells)       | 12, 5, 25, 50, and 100 mg/mL | 10 min, 6 h, 12 h, 24 h, and 48 h | No effect on fibroblast cells in a time- and dose-dependent way |

TXA: Tranexamic acid.

depending on the thickness of each tissue sample.

Marmotti et al[4] conducted another study, in which human tissue samples were harvested. This study also supported the hypothesis that TXA is a safe drug. Low doses of TXA (7 mg/mL) were administered for 48 h and the samples were assessed after short (immediately after exposure) and long time of exposure (1, 2, and 6 wk). The drug was completely safe and did not have any effect on cells’ ability to migrate. Also, cells treated with TXA did not seem to have any effects in terms of morphology, no matter the time of exposure, in comparison with the control group. In this study, a cell phenotype analysis was also conducted between the group that received TXA and the control group. Results showed that TXA exposure had no significant effects on cell phenotype after both long term and brief time of exposure. A further biochemical analysis investigated any possible alteration in glycosaminoglycan (GAG) concentration but no significant difference was noted between the TXA-treated group and the control group. Bolam et al[30] supported in their scoping review that TXA is a safe drug, when administered topically. More specifically, they included 15 studies that were considered eligible for their review. Some of these studies used human tissue samples while others used animal tissue samples. The authors concluded that there is a dose-dependent effect of TXA on chondrocytes, tenocytes, synoviocytes, and periosteum-derived cells. Both human and animal tissue samples were exposed to varying doses of the drug (1, 2, 4, 20, 25, 50, and 100 mg/mL) for different times of exposure (from 10 min to 48 h). The cultures that were exposed to TXA doses higher than 20 mg/mL had reduced cell viability and altered cell morphology. Thus, treatment with doses lower than 20 mg/mL had no significant detrimental effects. This conclusion supports the safety of the drug in a dose-dependent way.

Wang et al[32] studied the effects of topical administration of TXA on human fibroblast cells. These cells where cultured for a brief (10 min) and long time (6, 12, 24, and 48 h) after exposure to different drug concentrations (12.5, 25, 50, and 100 mg/mL). The authors suggested that there is a time and dose-dependent way in which TXA affects the fibroblast cells. More specifically, the samples had no detrimental effects when exposed to a high concentration of TXA (100 mg/mL) but for a brief time (10 min) or when exposed to a lower concentration of TXA (12.5 mg/mL) for a long time (from 6 to 48 h). However, high concentrations of the drug for a long period of time had clear effects concerning cell viability.

**HOW TXA AFFECTS CELL CYCLE**

In eukaryotic cells, the cell cycle has five different phases, G0, G1, S, G2, and M in order. In G0 phase, there is a cell cycle arrest, while in G1 phase, cells are gaining mass and activating growth signals. G2 is also a phase where cells are growing and organizing their genome. In S phase, the duplication of the chromosomes occurs, and in M-phase, the duplicated chromosomes and the cell are divided[33]. In order to investigate the way in which TXA affects the biological functions of cells, Goderecci et al[34] assessed the effects of the drug on the cell cycle. Cells incubated with TXA for only 10 min did not seem to have any significant alterations in terms of cell cycle profile. However, when cells were treated with TXA for 24 h or 48 h, there were significant changes. The number of cells in the G0/G1 phase was decreased, while there was an increase of cells in the G2/M phase and a significant increase of cells in the S phase, when compared to the control group and the group incubated for 10 min. As for the cells treated with TXA for 10 min and incubated with fresh medium for 24 h, there was an increased number of cells in the G0/G1 phase and a considerable decrease of cells in the G2/M phase, approaching the cell cycle profile of cells in the control group. Another study that investigated the potential mechanism by
which TXA could cause cytotoxicity, suggests that a caspase-3-dependent apoptotic mechanism could be responsible for the detrimental effects of TXA in human cells[10].

**TXA CYTOTOXICITY**

However, intraarticular TXA administration leads to exposure of intact articular cartilage to the drug. Consequently, there have been many concerns that TXA may cause detrimental effects on articular cartilage and have cytotoxic effects on chondrocytes, especially since chondrocyte death is related with the development of further cartilage degeneration and osteoarthritis[3].

TXA can cause massive cytotoxicity (Table 2). McLean et al[10] conducted a study where human tendon tissue, synovial tissue, and cartilage tissue were obtained during major orthopaedic surgeries and TXA seemed to have significant effects on each kind of tissue[26]. All samples were exposed to a wide range of TXA concentrations (1, 50, and 100 mg/mL) for 4 h, 16 h, or 24 h[10]. A significant reduction in cell viability as well as high rates of apoptosis were observed in all types of tissue in a time- and dose-dependent way. McLean et al[10] tried to identify potential mechanisms that may cause these high rates of apoptosis and they came to the conclusion that increased amount of apoptotic proteins occurred in tenocytes and chondrocytes, in comparison with the control group, thus suggesting that TXA-mediated cell death may be caused by apoptotic proteins like caspase-3. Lower doses had a lower impact on cells but still influenced cell function. Moreover, the authors reported that a high percentage of cell death occurred in the control group, most likely due to tissue drying by surgical harvesting and completion of graft preparation and also due to age-related changes, as most of the samples were harvested from elderly patients. However, they did not seem to have any major impact on the study outcome as these factors affected every sample group.

Parker et al[11] also proved the drug’s cytotoxic effects by exposing human chondrocytes on 2D and 3D cultures to TXA (10, 20, and 40 mg/mL for 3 h and 6 h). On 2D cultures, chondrocytes were seeded onto numerous tissue culture plastic trays while on 3D cultures the authors tried to simulate the 3D nature of articular cartilage by using chondrocyte-laden gelatine-methacryloyl (GelMA) hydrogels. However, this study also suggests that there may be a safe dose for the drug. More specifically, on the 2D cell-culture model, the drug had significant effects on cell viability at doses between 20 mg/mL and 40 mg/mL. On the contrary, cells treated with doses lower than 20 mg/mL did not seem to be affected by the drug. In 3D cultures, cell viability was decreased in a time- and dose-dependent way. Furthermore, Parker et al[11] inspected the effect of TXA on glycosaminoglycan GAG concentration and found no significant differences between the control group and the groups exposed to the drug, thus suggesting that high doses of TXA can cause cell death but they do not affect the quality of the cartilage tissue. Another study, carried out by Tuttle et al[24] on murine and bovine chondrocytes, also reached the conclusion that TXA can be cytotoxic. Treatment with high doses of TXA (50 and 100 mg/mL) had clear detrimental effects on chondrocyte viability in a time-dependent way. However, cells treated with 25 mg/mL had no considerable decrease in cell viability. Moreover, in this study it was observed that there was a small but significant loss of GAG in groups that were treated with TXA in comparison with the control groups, in contrary to the conclusion of Parker et al[11] suggesting that TXA does not affect GAG concentrations. These studies did not take into consideration all the pharmacological factors that normally take place inside the human joint. The time of cell exposure to TXA in an *in vitro* study is longer than the time of exposure that theoretically occurs *in vivo* where other kinds of cells (inflammatory cells and platelets) could have a potential role in the drug’s clearance. As a result, McLean et al[10] suggested that the *in vitro* nature of these studies might have an influence on the results.

Another interesting finding is that washing the drug out of the cultures after a few minutes of exposure seems to decrease TXA’s cytotoxicity[34]. According to Goderecci et al[34], when the drug was washed out of the cultures, only the highest dose of TXA (100 mg/mL) affected cell cycle profile and cell viability. However, 20, 50, and 70 mg/mL of TXA did not have any detrimental effect on cell function when the drug was washed out after 10 min of incubation. These samples were compared with other samples that were exposed to same TXA concentrations for 24 h and 48 h without washing the drug out, in which TXA had significant effects at doses higher than 50 mg/mL. Tuttle et al[24] observed the same outcome when animal tissue samples were incubated with TXA for different amounts of time. Samples were incubated with various TXA concentrations for three different time points, 8 h, 24 h, and 48 h. Samples incubated with high TXA levels presented complete cell death, no matter the time of exposure. However, at low levels of TXA, cell viability was notably lower in samples incubated for 24 h when compared with samples incubated for 8 h. Complete cell death was observed in the 48 h sample, thus supporting the conclusion that the amount of time that cells are exposed to TXA is a major factor concerning the cytotoxicity of the drug.

A few studies observed the effects of TXA in different kind of tissue samples. Marmotti et al[3] carried out a study, in which human cartilage and synovial biopsies were obtained from patients during major orthopedic procedures. Each kind of tissue was incubated with the same concentration of TXA for the same amount of time. TXA had the same effects on those groups, and it showed no cytotoxicity on both samples and was completely safe for all types of cells. Other investigators harvested synovial and...
Table 2 Characteristics and outcomes of studies that proved cytotoxic effects of tranexamic acid

| Ref. | Tissue | Doses | Time of exposure | Outcome |
|------|--------|-------|------------------|---------|
| Tuttle et al [22], 2015 | Animal samples (bovine and murine) | 25, 50, and 100 mg/mL in murine chondrocytes and 100 mg/mL in bovine chondrocytes | 8, 24, and 48 h for both bovine and murine explants | Cell viability was notably decreased as TXA dose and incubation time were increased |
| Parker et al [28], 2018 | Human samples (chondrocytes on 2D and 3D models) | 5, 10, 20, and 40 mg/mL in 2D cultures and 10, 20, and 40 mg/mL in 3D cultures | 3, 6 and 12 h for the 2D models and 3 h for the 3D models | Cell viability, cellular metabolic activity, and number of attached cells were considerably decreased in a dose-dependent way |
| Coderecci et al[29], 2019 | Human samples (chondrocytes) | 20, 50, 70, and 100 mg/mL | 10 min, 24 h, and 48 h | Chondrocyte viability and cell cycle profile were affected in a dose- and time-dependent way |
| McLean et al [27], 2019 | Human samples (tendon, synovial, and cartilage tissues) | 0, 1, 50, and 100 mg/mL | 1, 4, and 24 h | Cell viability and apoptosis rate were affected at higher doses |

TXA: Tranexamic acid.

cartilage samples from patients who were also treated with the same TXA doses for the same amount of time[10]. Apoptosis rate was approximately the same for all three types of tissue. Viability was decreased in all types of cells at different time points for each dose level; however, the detrimental result was the same in all tissue samples. Thus, TXA seems to interact, in the same way, with all types of tissue and it has the same effects in all types of cells.

**EFFECTS OF TXA ON GLYCOSAMINOGLYCAN CONCENTRATION**

Marmotti et al[4] proved in their study that TXA did not cause any significant change in GAG concentration by conducting a biochemical analysis. Moreover, Parker et al[11] supported this conclusion as they noted no significant difference in GAG concentration between the TXA-treated cells and the placebo-treated cells and thus suggested that TXA had no effect on the quality of cartilage tissue irrespective of the drug’s toxicity to chondrocytes[27]. However, Tuttle et al[24] observed in their study that TXA had a small but significant effect on GAG concentration, in contrary to the conclusions of Marmotti et al[4] and Parker et al[11]. This differentiation may be due to the nonidentical period for which cells were exposed to TXA, thus causing a different effect on GAG concentration in each study. Parker et al[11] used a shorter and more clinically appropriate length of exposure to the drug to come to a more accurate conclusion[27].

**EFFECTS OF TXA DEPENDING ON PATIENT AGE**

TXA may have different effects on chondrocytes depending on the age of patients. Marmotti et al[4] obtained human biopsies from young donors (patients between 14 and 40 years old) in order to inspect the short- and long-term effects of TXA (7 mg/mL) on these samples. Biopsies from young donors did not seem to develop any kind of damage in terms of morphologic and immunophenotypic characteristics no matter the time of exposure. However, McLean et al[10] harvested tissue samples from older patients (specific age not provided) undergoing total knee and hip arthroplasties. In this study, just like previously, biopsies were checked for any effect of TXA after a short and a long period of incubation. Cell viability was significantly reduced after 24 h of exposure to TXA, regardless of the TXA dose. This suggests that TXA may be safer for younger patients and cause more serious detrimental effects in older patients.

**FUTURE PERSPECTIVES**

While many studies have been carried out on tissue samples, it is important to carry out in vivo studies in order to support the data that were obtained from in vitro studies, considering all the pharmacological interactions inside the human joint such as drug clearance and tissue distribution, and observe the development of any side effects. Additionally, more studies are needed to confirm the possibility that there is a safe TXA dose for topical use.

Safety of the drug is quite important as well as effectiveness of the drug. Studies should not only suggest the safe doses of TXA but also inspect the healing process at each dose level and if TXA
contributes in the recovery process. Moreover, more data is needed to clarify the molecular mechanism of TXA cytotoxicity. This could help us understand the process through which TXA causes detrimental effects in human cells and make clear its molecular correlation with cell apoptosis.

Effects of TXA on GAG concentration are only discussed in a limited number of studies. This is an interesting topic that should be furtherly discussed. Many studies suggest that TXA is cytotoxic, but we should also clarify whether TXA causes any significant differences in GAG concentration. This will help us to conclude whether the drug affects the quality of cartilage tissue, irrespective of any cytotoxic effects that the drug may have.

A topic that is not greatly discussed and should be furtherly investigated is the cost vs benefit of a potential therapeutic intervention with topical TXA. Clinical studies should be carried out to investigate whether the efficacy of topical use of TXA is as great as the cost of the drug and if there is a point to use the drug topically instead of IV.

**CONCLUSION**

Given the low evidence of the available literature and the lack of clinical data, there seems to be no clear outcome considering the effects of the drug on chondrocytes, as some studies suggest that TXA is clearly cytotoxic[10,11,24,34] while others support the safety of the drug[1,3,4]. Most of these studies came to the conclusion that TXA can have effects on chondrocytes in a time- and dose-dependent way[10,11,24,34]. Higher doses can cause detrimental effects on cells especially when they are exposed to the drug for a long period of time[24,34]. Other studies suggest that TXA seems to have the same effects on every type of tissue inside the human joint and each type of cell interacts in the same way with the drug[4,10]. Nevertheless, some studies proved that there seems to be a safe TXA level at which the healing process can continue without any effects on the joint function[1,3,4,11,24]. There are indications that high doses of TXA can affect the cell cycle profile, but this suggestion needs further investigation[34]. The suggestion that the effects of TXA depend on the age group of the patients also needs further investigation[4,10].

**FOOTNOTES**

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**Country/Territory of origin:** Greece

**ORCID number:** Aristeidis-Paragiotis Kontokostopoulos 0000-0001-8326-8376.

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**REFERENCES**

1. **Degirmenci E, Ozturan KE, Sahin AA, Yilmaz F, Kaya YE. Effects of tranexamic acid on the recovery of osteochondral defects treated by microfracture and acellular matrix scaffold: an experimental study. J Orthop Surg Surg 2019; 14: 105 [PMID: 30929060 DOI: 10.1186/s13018-019-1144-7]**

2. **Cesarman-Maus G, Hajjar KA. Molecular mechanisms of fibrinolysis. Br J Haematol 2005; 129: 307-321 [PMID: 15842654 DOI: 10.1111/j.1365-2141.2005.05444.x]**

3. **Ambra LF, de Girolamo L, Niu W, Phan A, Spector M, Gomoll AH. No effect of topical application of tranexamic acid on articular cartilage. Knee Surg Sports Traumatol Arthrose 2019; 27: 931-935 [PMID: 29119286 DOI: 10.1007/s00167-019-05320-6]**

4. **Marmotti A, Mattia S, Mangiavini L, Bonasia DE, Bruzzone M, Dettoni F, Rosso F, Blonna D, Rossi R, Castoldi F, Peretti GM. Tranexamic acid effects on cartilage and synovial tissue: an in vitro study for a possible safe intra-articular use. J Biol Regul Homeost Agents 2016; 30: 33-40 [PMID: 28002898]**
Tranexamic acid cytotoxicity

Gkiatas I et al. Tranexamic acid cytotoxicity: a clinical review. *Anaesthesiol Intensive Ther* 2015; 47: 339-350 [PMID: 25797505 DOI: 10.5603/AIT.2015.0011]

Bradley KE, Ryan SP, Penrose CT, Grant SA, Wellman SS, Attarian DE, Green CL, Risoli T Jr, Bolognesi MP. Tranexamic acid or epsilon-aminocaproic acid in total joint arthroplasty? *Bone Jt J* 2019; 101-B: 1093-1099 [PMID: 31474134 DOI: 10.1016/j.bjtj.2018-1096.R1]

Ramirez RJ, Spinella PC, Bochicchio GV. Tranexamic Acid Update in Trauma. *Crit Care Clin* 2017; 33: 85-99 [PMID: 27604541 DOI: 10.1016/j.ccc.2016.08.004]

Yi Z, Bie S, Jing Y, Zongke Z, Pengke F,FIXING P. Tranexamic Acid Administration in Primary Total Hip Arthroplasty: A Randomized Controlled Trial of Intravenous Combined with Topical Versus Single-Dose Intravenous Administration. *J Bone Joint Surg Am* 2016; 98: 983-991 [PMID: 27307358 DOI: 10.2106/JBJS.15.00638]

Fillingham YA, Ramkumar DB, Jevsevar DS, Yates AJ, Shores PA, Mullen K, Bini SA, Clarke HD, Schemitsch E, Johnson RL, Memtsoudis SG, Sayeed SA, Sah AP, Della Valle CJ. The Efficacy of Tranexamic Acid in Total Knee Arthroplasty: A Network Meta-Analysis. *J Arthroplasty* 2018; 33: 3090-3098.e1 [PMID: 29805161 DOI: 10.1016/j.arth.2018.04.043]

McLean M, McCall K, Smith IDM, Blyth M, Kitson SM, Crowe LAN, Leach WJ, Rooney BP, Spencer SJ, Mullen M, Campton JL, McInnes IB, Akbar M, Millar NL. Tranexamic acid toxicity in human periarticular tissues. *Bone Joint Res* 2019; 8: 11-18 [PMID: 30800295 DOI: 10.2106/2016-3758.81.BJR-2018-0181.R1]

Parker JD, Lim KS, Kieser DC, Woodfield TB, Hooper GJ. Is tranexamic acid toxic to articular cartilage when administered topically? *Bone Jt J* 2018; 100-B: 404-412 [PMID: 29589496 DOI: 10.1302/0301-620X.100B3.BJR-2017-1135.R1]

Li ZJ, Fu X, Xing D, Zhang HF, Zang JC, Ma XL. Tranexamic acid effective and safe in spinal surgery? *Eur Spine J* 2013; 22: 1950-1957 [PMID: 23567623 DOI: 10.1007/s00586-013-2774-9]

Karaaslan F, Karaöglu S, Yurdakul E. Reducing Intra-articular Hemarthrosis After Arthroscopic Anterior Cruciate Ligament Reconstruction by the Administration of Intravenous Tranexamic Acid: A Prospective, Randomized Controlled Trial. *Am J Sports Med* 2015; 43: 2720-2726 [PMID: 26337246 DOI: 10.1177/0363546515599629]

Gillespie R, Shishani Y, Joseph S, Streit JF, Goebzie R. Neer Award 2015: A randomized, prospective evaluation on the effectiveness of tranexamic acid in reducing blood loss of total shoulder arthroplasty. *J Shoulder Elbow Surg* 2015; 24: 1679-1684 [PMID: 26480877 DOI: 10.1016/j.jse.2015.07.029]

Farrow LS, Smith TO, Ashcroft GP, Myint PK. A systematic review of tranexamic acid in hip fracture surgery. *Br J Clin Pharmacol* 2016; 82: 1458-1470 [PMID: 27492116 DOI: 10.1111/bcp.13079]

CRASH-2 trial collaborators. Shakur H, Roberts I, Bautista R, Caballero J, Coats T, Dewan Y, El-Sayed H, Gigoeiashvili T, Gupta S, Herrera J, Hunt B, Iribhogbe P, Izziertelor M, Khamis H, Komolafe E, Marrero MA, Mejia-Mantilla J, Miranda J, Morales C, Olaomi O, Oddashi F, Perel P, Peto R, Ramana PV, Ravi RR, Yuthakasemset S. Effects of tranexamic acid on death, vascular occlusive events, and blood transfusion in trauma patients with significant haemorrhage (CRASH-2): a randomised, placebo-controlled trial. *Lancet* 2010; 376: 23-32 [PMID: 20554319 DOI: 10.1016/S0140-6736(10)60835-5]

Roberts I, Perel P, Prieto-Merino D, Shakur H, Coats T, Hunt BJ, Lecky F, Brohi K, Willett K; CRASH-2 Collaborators. Effect of tranexamic acid on mortality in patients with traumatic bleeding: prespecified analysis of data from randomised controlled trial. *BMJ* 2012; 345: e5839 [PMID: 22968527 DOI: 10.1136/bmj.e5839]

Lack WD, Crist BD, Seymour RB, Harvin W, Karunakar MA; TXA Study Group. Effect of Tranexamic Acid on Transfusion: A Randomized Clinical Trial in Acetabular Fracture Surgery. *J Orthop Trauma* 2017; 31: 526-530 [PMID: 28938283 DOI: 10.1097/BOT.0000000000000968]

Goothie SM, Faraoni D. Tranexamic acid and perioperative bleeding in children: what do we still need to know? *Curr Opin Anaesthesiol* 2016; 29: 343-352 [PMID: 28093114 DOI: 10.1097/ACO.0000000000000726]

Pabhi R, Fries D, Schöchl H, Streif W, Toller W. Tranexamic acid for treatment and prophylaxis of bleeding and hyperfibrinolysis. *Wien Klin Wochenschr* 2017; 129: 303-316 [PMID: 28432428 DOI: 10.1007/s00508-017-1194-y]

Kim C, Park SS, Davey JR. Tranexamic acid for the prevention and management of orthopedic surgical hemorrhage: current evidence. *J Blood Med* 2015; 6: 239-249 [PMID: 26345147 DOI: 10.2147/JBM.S61915]

Ker K, Edwards P, Perel P, Shakur H, Roberts I. Tranexamic acid usage on surgical bleeding: prospective review and cumulative meta-analysis. *BMJ* 2012; 344: e3054 [PMID: 22611614 DOI: 10.1136/bmj.e3054]

Karaman R, Ghareeb H, Dajani KK, Scerano L, Hallak H, Abu-Lafi S, Mecca G, Bufo SA. Design, synthesis and in vitro kinetic study of tranexamic acid prodrugs for the treatment of bleeding conditions. *J Comput Aided Mol Des* 2013; 27: 615-635 [PMID: 23881217 DOI: 10.1007/s10822-013-9666-2]

Tuttle JR, Feltman PR, Ritterman SA, Ehrlich MG. Effects of Tranexamic Acid Cytotoxicity on In Vitro Chondrocytes. *Am J Orthop (Belle Mead NJ)* 2015; 44: E497-E502 [PMID: 26665251]

Gomez-Barrena E, Ortega-Andreu M, Padilla-Eguiluz NG, Perez-Chrzanowska H, Figueredo-Zalve R. Topical intra-articular compared with intravenous tranexamic acid to reduce blood loss in primary total knee replacement: a double-blind, randomized, controlled, noninferiority clinical trial. *J Bone Joint Surg Am* 2014; 96: 1937-1944 [PMID: 25471907 DOI: 10.2106/JBJS.N.00060]

Abdel MP, Chalmers BP, Taunton MJ, Pagnano MW, Trousdale RT, Sierra RJ, Lee YY, Boettner F, Su EP, Haas SB, Figgie MP, Mayman DJ. Intravenous Versus Topical Tranexamic Acid in Total Knee Arthroplasty: Both Effective in a Randomized Clinical Trial of 640 Patients. *J Bone Joint Surg Am* 2018; 100: 1023-1029 [PMID: 29916929 DOI: 10.2106/JBJS.17.00908]

Xu JW, Qiang H, Li TL, Wang Y, Wei XX, Li F. Efficacy of topical vs intravenous tranexamic acid in reducing blood loss and promoting wound healing in bone surgery: A systematic review and meta-analysis. *World J Clin Cases* 2021; 9: 4210-4220 [PMID: 34141783 DOI: 10.12998/wjcc.v9.i17.4210]

Dong Y, Liang J, Tong B, Shen J, Zhao H, Li Q. Combined topical and intravenous administration of tranexamic acid further reduces postoperative blood loss in adolescent idiopathic scoliosis patients undergoing spinal fusion surgery: a randomized controlled trial. *BMC Musculoskelet Disord* 2021; 22: 663 [PMID: 34372818 DOI: 10.1186/s12891-021-04562-5]
29 Li S, Lu Q, Guo X, Zhang M, Miao Z, Luo D, Liu P. Intravenous Combined with Topical Tranexamic Acid Administration Has No Additional Benefits Compared with Intravenous Administration Alone in High Tibial Osteotomy: A Retrospective Case-Control Study. Orthop Surg 2020; 12: 515-523 [PMID: 32162488 DOI: 10.1111/os.12652]

30 Bolam SM, O'Regan-Brown A, Paul Monk A, Musson DS, Cornish J, Munro JT. Toxicity of tranexamic acid (TXA) to intra-articular tissue in orthopaedic surgery: a scoping review. Knee Surg Sports Traumatol Arthrosc 2021; 29: 1862-1871 [PMID: 32860523 DOI: 10.1007/s00167-020-06219-7]

31 Stucinskas J, Tarsasevicius S, Cebatorius A, Robertsson O, Smailys A, Wingstrand H. Conventional drainage vs four hour clamping drainage after total knee arthroplasty in severe osteoarthritis: a prospective, randomised trial. Int Orthop 2009; 33: 1275-1278 [PMID: 18925394 DOI: 10.1007/s00264-008-0662-4]

32 Wang F, Wang SG, Yang Q, Nan LP, Cai TC, Wu DS, Zhang L. Cytotoxicity and Effect of Topical Application of Tranexamic Acid on Human Fibroblast in Spine Surgery. World Neurosurg 2021; 153: e380-e391 [PMID: 34224885 DOI: 10.1016/j.wneu.2021.06.125]

33 Barnum KJ, O'Connell MJ. Cell cycle regulation by checkpoints. Methods Mol Biol 2014; 1170: 29-40 [PMID: 24906307 DOI: 10.1007/978-1-4939-0888-2_2]

34 Goderecci R, Giusti I, Necozione S, Cinque B, D'Ascenzo S, Dolo V, Calvisi V. Short exposure to tranexamic acid does not affect, in vitro, the viability of human chondrocytes. Eur J Med Res 2019; 24: 15 [PMID: 30795796 DOI: 10.1186/s40001-019-0373-x]
