Analysis of the Cancer Genome Atlas Data Reveals Novel Putative ncRNAs Targets in Hepatocellular Carcinoma

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Hepatocellular carcinoma (HCC) is the prevalent type of primary liver malignancy. Different noncoding RNAs (ncRNAs) that negatively regulate gene expression, such as the microRNAs and the long ncRNAs (lncRNAs), have been associated with cell invasiveness and cell dissemination, tumor recurrence, and metastasis in HCC. To evaluate which regulatory ncRNAs might be good candidates to disrupt HCC proliferation pathways, we performed both unsupervised and supervised analyses of HCC expression data, comparing samples of solid tumor tissue (TP) and adjacent tissue (NT) of a set of patients, focusing on ncRNAs and searching for common mechanisms that may shed light in future therapeutic options. All analyses were performed using the R software. Differential expression (total RNA and miRNA) and enrichment analyses (Gene Ontology + Pathways) were performed using the package TCGABiolinks. As a result, we improved the set of lncRNAs that could be the target of future studies in HCC, highlighting the potential of FAM170B-AS1 and TTN-AS1.

1. Introduction

Epidemiologic data from the International Agency for Research on Cancer of the World Health Organization reveals that liver cancer comprises 5.6% of worldwide cancer incidence and 9.1% of all cancer-associated mortality [1]. Hepatocellular carcinoma (HCC) is the most prevalent type of primary liver malignancy [2]. The high lethality of HCC can be attributed to the lack of diagnostic markers for an early detection and late stages high heterogeneity [3]. HCC has been epidemiologically associated with chronic Hepatitis B Virus (HBV) or Hepatitis C Virus (HCV) [4], as well as alcoholic and nonalcoholic fatty liver disease, which are its major risk factors [2]. Currently, the most effective treatment is either surgical tumor resection or liver transplantation [5].

Multiple studies have shown the potential of different microRNAs (miRNAs) as prognostic and diagnostic biomarkers in many types of cancer, including HCC [6–8]. miRNAs are noncoding RNAs that negatively regulate gene expression by leading mRNAs to target degradation or translational repression after binding to its 3'UTR (for review see [9]). In cancer, their role has been either as tumor suppressors or as enhancers (oncomiRs) [10].

In HCC, different miRNAs have been associated with cell invasiveness by repressing TET gene expression, leading to silencing of several invasion-suppressors via hypermethylation [11], and cell dissemination by regulating differentiation, hence increasing metastatic potential [12]. They are even implicated in improvement of HBV and HCV viral replication and tumor-supporting mechanisms [13, 14]. This
multifaceted miRNA capacity of influencing in the HCC environment proves the importance of studies describing expression profiles of miRNAs during tumor occurrence.

Another class of noncoding (nc) RNAs, the long ncRNAs (lncRNAs), are > 200 nucleotides' RNA molecules with multiple regulatory roles that cannot be inferred by their sequence. These roles comprise, among others, chromatin organization affecting the gene expression [15]. HOTAIR, an antisense lncRNA, has been associated with HCC recurrence and metastasis [16]. HULC and FTX (HCC) are also upregulated in tumoral samples [17].

Here, differently from previous works that focused on viral infection (HBV or HCV) comparing primary solid tumor tissue (TP) and adjacent tissue (NT) [6, 8, 18, 19], or focused on the mutation found [20], we in silico compared TP and NT of a set of patients in sense to discover the pathways that differentiate both groups of samples and the regulatory ncRNAs and their putative targets. As a result, we improve the set of lncRNAs that could be the target of future studies.

2. Material and Methods

All analyses were performed using the R software (v. 3.4.0) [21]. The differential expression (mRNA and miRNA) analysis was performed using the package TCGA-Bioliqns (v. 2.7.1) [22]. First, we downloaded HCC harmonized data (hg38) from The Cancer Genome Atlas (TCGA) using the function GDC download with the option legacy = FALSE. We analyzed a total of 41 participants that have expression data of both primary solid tumor and adjacent tissue samples. It is worth noticing that in the database adjacent tissue is referred to as normal; however this is hardly the case as all patients were cirrhotic. Thus we use the term adjacent, as this is not a sample from a normal liver. To select these individuals, we used only the participant ID of the TCGA barcode as query sample from a normal liver. To select these individuals, we focused on the mutation found [20], we in silico compared TP and NT of a set of patients in sense to discover the pathways that differentiated both groups of samples and the regulatory ncRNAs and their putative targets. As a result, we improve the set of lncRNAs that could be the target of future studies.

2.1. Data Source

TCGA-BC-A10Q

2.2. Data Preprocessing

Preprocessing involved the following steps: (1) Normalization of each sample with the option legacy = FALSE of the function TCGA-Bioliqns, (2) filtering out low expression genes (log2 expression value < 10), and (3) filtering out samples with less than 75% of expression values.

2.3. Data Analysis

We found a total of 1739 DEGs in total RNA-seq among tumor and normal samples. From these, 1276 were upregulated in tumor (Figure 1(a), Figure S1A, and Table S1). miRNA differential expression (DE) revealed 234 DE miRNAs, of which 169 were upregulated in tumor (Figure 1(b), Figure S1B, and Table S1). Other noncoding regulatory RNAs resulted in 92 pre-miRNAs (73 upregulated in tumor) and 122 lncRNAs (90 upregulated in tumor) (Figure 1(c), Table S2). Clusters with approximately unbiased grouping support p values (%) (au – red values) of 95 were considered as statistically significant groups.

For the differentially expressed transcripts, we performed a Spearman correlation to detect which regulatory RNAs are negatively correlated with other RNAs. We accepted those with r < -0.8 and p value < 0.05 as statistically significantly correlated. These correlated transcripts were used as interactions to input the network on Cytoscape (v. 3.5.1) [26], where the edges represent the statistically significant r values. The miRNAs and their putative targets were used to predict their interaction using the online software TargetScan (release 7.1) [27]. Interactions not found in TargetScan were also tested in miRDB [28] and TarBase (v. 8) [29]. The interactions found by either TargetScan or TarBase were confirmed by two other tools: miRWalk v. 3.0 (http://mirwalk.umm.uni-heidelberg.de/) [30] considering a binding probability cut-off of 0.8, and mirDIP v. 4.1 (http://ophid.utoronto.ca/mirDIP/index.jsp) [31, 32] considering a "medium" cut-off of scores. Gene Ontology Biological Processes of the proteins associated with the network were evaluated using the Cytoscape plugin BINGO [33]. For the interest in lncRNAs, we performed a supervised prediction model using the Area Under the Curve of the Receiver Operating Characteristic (AUC-ROC) using the package pROC v. 1.11.0 [34].

3. Results and Discussion

In this study we performed a supervised analysis of HCC expression data focusing on ncRNAs searching for common mechanisms that may shed light in future therapeutic options. The majority of statistically significant differentially expressed ncRNAs are higher expressed on tumor samples, suggesting that these RNAs are necessary to tumor progression/maintenance. Additionally, tumor samples showed a more diverse expression profile in comparison to those from adjacent tissues. Such pattern has been reported also for gastric [35] and colorectal cancers [36].

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The enrichment analysis (Gene Ontology + Pathways) revealed that the most represented pathways in differentially expressed transcripts from total RNA-seq are involved in bile metabolism, fear behavioral response, and immune-related categories (Figure 2). To infer putative expression relationship, we plotted a network based on Spearman’s correlation,
Figure 1: Heat maps of differentially expressed noncoding regulatory RNAs. (a) miRNA. (b) Pre-miRNA. (c) LncRNA. Tumor samples in brown and adjacent samples in blue. Hierarchical clusterization based on transcript log10 scale expression.

considering only the negative interactions. These interactions involved a total of 18 highly correlated regulatory ncRNAs of all types (miRNA, pre-miRNA, and LncRNA) with their putative targets (Figure 3(a)). In the case of miRNAs and pre-miRNAs, the miRNA-target interactions were predicted as explained in the Material and Methods. These highly negative targets are most involved in programmed cell death, immune response, and Molybdenum cofactor biosynthesis processes (Figure 3(b)). For the LncRNAs in the network, we calculated the AUC-ROC values and found four LncRNAs with potential to correctly discriminate TP and NT samples: **CCND2-ASI** (AUC = 0.792, 95% confidence interval: 0.6834-0.8903), **FAM170B-ASI** (AUC = 0.917, 95% confidence interval: 0.8387-0.9758), **TTN-ASI** (AUC = 0.901, 95% confidence interval: 0.84-0.9539), and **SYNPR-ASI** (AUC = 0.939, 95% confidence interval: 0.8798-0.9823).

The DEGs’ enrichment analysis suggested that bile metabolism and fear behavioral response immune-related categories are the most represented pathways. Immune-related categories are usually disrupted in cancer. For example, **CD274**, upregulated in our TP samples, confers immune resistance to tumor cells by the inactivating cytotoxic T-cell
Table 1: Top ten differentially expressed genes (DEGs) from total RNA-seq and microRNA RNA-seq.

| Transcript | Total RNA-seq DEGs |  |
|------------|-------------------|---|
|            | Upregulated in tumor | Fold change | Downregulated in tumor | Fold change |
| SIRT1      | 15.32119          |  |
| RPL28      | 11.29295          |  |
| LINC01613† | 10.83190          |  |
| CACNB3     | 10.71749          |  |
| CREM       | 10.54143          |  |
| GOLGA8B    | 10.32588          |  |
| SLC46A2†   | 10.16996          |  |
| LINC00449† | 9.78697           |  |
| CFAP44-AS1†| 9.67638           |  |

| Transcript | DE miRNAs |  |
|------------|-----------|---|
|            | Upregulated in tumor | Fold change | Downregulated in tumor | Fold change |
| hsa-mir-767| 8.99037   |  |
| hsa-mir-105-2| 8.90799 |  |
| hsa-mir-891a| 8.86653  |  |
| hsa-mir-105-1| 8.19189 |  |
| hsa-mir-3923| 7.94124  |  |
| hsa-mir-520f| 7.91259  |  |
| hsa-mir-520c| 7.63784  |  |
| hsa-mir-518e| 7.37922  |  |
| hsa-mir-520b| 7.17289  |  |
| hsa-mir-520a| 7.00665  |  |

†putative regulatory IncRNA.

Figure 2: Enrichment analysis of differentially expressed transcripts from total RNA data. GO: Gene Ontology. The red lines represent the ratio of genes found for the pathway over the total number of genes for that specific pathway. Inside each bar, n: number of genes. Bar sizes are in agreement to the -log10 of the FDR of the enriched ontology/pathway.
[37]. DACT1, which encodes for an antagonist of beta catenin 1, and DVL2, a dishevelled protein family member, are respectively, down- and upregulated in tumor tissues, suggesting that the Wnt signaling is active [38]. Additionally, CDK14 and GSK3B are upregulated, reinforcing the Wnt signaling activation, which is related to cell polarity category [39]. This signaling pathway has been associated with malignant transformation [40].

GO group classified as fear behavioral response includes a series of genes neurotransmitters (such as glutamate, dopamine, and serotonin receptors), which comes as no surprise since many studies have shown the impact of serotonin, GABA, and sympathetic neurotransmitters in hepatocyte proliferation [41–43]. It also includes MECP2 and transcription factors associated with chromatin remodeling. Finally, bile acids are also known to act as potential carcinogens and deregulation of bile acids homeostasis has been linked to HCC formation [44].

Another transcription factor, FXR2, is supposed to act as heterodimer (or larger complexes) with TP53 or FXR1, suppressing tumor development. However, TP53 and FXRI expressions were not detected after normalization process. Still, FXR2 can interact as homodimer or as a larger complex [45]. The absence of FXRI expression could be a consequence of GSK3B upregulation, once FXRI phosphorylation by GSK3B leads to FXRI downregulation [46].

From the negative correlation network (Figure 3), we can highlight the immune-related categories, as occurred in the DEGs enrichment analysis. The expression of NAT1 has been recently reported to show high expression in breast cancer and be associated with steroid biosynthetic pathway genes [47]. Here, NAT1 is also upregulated in TP samples. This gene's
expression is also negatively correlated with two lncRNAs, both antisense RNAs: SYNPR-ASI and CCND2-ASI. CCND2-ASI is known to promote glioma cell proliferation by activating Wnt/β-catenin signaling [48], but it is downregulated in HCC. CASP1, usually downregulated in cancer cells once it promotes apoptosis [49], showed a high expression pattern in TP samples and is negatively correlated with CCND2-ASI. NLRP1 expression is associated with tumor inflammamasomes and suppression of apoptosis in metastatic melanoma [50]. However, NLRP1 is downregulated in our tumor data and negatively correlated with the pre-miRNA MIR3667. This miRNA is known to disrupt the oncogenetic activity of PCAT-1/ MYC in prostate cancer [51] and thus its low expression in TP samples is expected. GRIK2, correlated with tumor progress [52], ERCI, which is upregulated in TP samples and its expression is associated with tumor progression once it is necessary to focal adhesion turnover [53], and PRDX3, whose overexpression is highly connected to prostate cancer [54] by protecting tumoral cells from oxidative stress [55], are all upregulated in TP samples of HCC and negatively correlated with the expression of the antisense RNA FAM170B-ASI. ERCI is also negatively correlated with SYNPR-ASI, hsa-mir-139, which is known to play antitumoral roles in HCC [56], and the pre-miRNA MIR320A plays antitumoral roles in breast cancer [57]. SLC17A8 is upregulated in prostate cancer [58] and in HCC and negatively correlated with SYNPR-ASI. HSD3B7 is associated with bile acid and did not change its expression in CTNNB1 mutated HCC samples [59]. However, here HSD3B7 is upregulated in TP samples and negatively correlated with LINC01493.

NLRP1/CASP1 form a complex that induces pyroptosis [60], a cell death dependent on CASP1 and associated with many pathological conditions, including cancer [61]. Bearing in mind that we can not interpret gene expression as active protein production or enzymatic activity, still it seems like pyroptosis pathway is disrupted in HCC in comparison to other cancer types and that CCND2-ASI might play a role by regulating CASP1 expression in this process.

AQP9 overexpression decreased the PIK3CB levels in normal tissues, reducing the cell proliferative potential by increasing FOXO1 levels and reducing PCNA expression [62]. In HCC, AQP9 levels are low [63] inducing PIK3CB activity and cell proliferation [62]. In agreement with these authors, AQP9 is downregulated in our TP samples profile, while PIK3CB is upregulated. PCNA is also upregulated but did not pass the logFC cut-off. AQP9 is negatively correlated with TTN-ASI, which was recently described as an oncogene highly expressed in esophageal squamous cell carcinoma progression and metastasis [64].

Hierarchical cluster analysis of the differentially expressed total transcripts, miRNAs, pre-miRNAs, and lncRNAs, shows that statistically significant groups are created in all cases, discriminating most adjacent from tumoral samples. This kind of distinction was not found when trying to differentiate samples also by viral types (HBV or HCV) (data not shown). It is worth noticing that DEG, but especially ncRNA analysis, was able to perfectly discriminate between TP and NT, although it was not able to separate HBV and HCV-infected samples. This suggests that the mechanisms depicted here are common to HCC regardless of its causative injury. Even though risk factors for HCC are well-known, it remains as an important cause of death worldwide. Although tumor surveillance in cirrhotics is highly recommended by international guidelines [65], late diagnosis is quite common. Moreover, advanced liver disease and parenchymal dysfunction further prevent curative therapies [66].

4. Conclusions

Our data suggests that neither HBV nor HCV infection changes overall gene expression (including those genes encoding for ncRNAs) in TP samples. Pyroptosis pathway is misregulated in HCC if compared to other cancer types and the lncRNA CCND2-ASI might be involved in this misregulation, revealing a singular characteristic of HCC. Additionally, FAM170B-ASI and TTN-ASI emerge as new candidates to tests to disrupt HCC homeostasis by turning cancer cells susceptible to oxidative stress or affecting cancer cell proliferation, respectively. Also, these lncRNAs show remarkable expression signatures, differentiating TP from NT samples with high AUC-ROC values.

Data Availability

All data used in this work is publicly available at The Cancer Genome Atlas (TCGA) database <https://cancer-genome.nih.gov/>.

Conflicts of Interest

The authors declare that there are no conflicts of interest regarding the publication of this paper.

Authors’ Contributions

Tiago Falcon and Martiela Freitas contributed equally to this work.

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Supplementary Materials

Figure S1: volcano plot representing differentially expressed transcripts (primary solid tumor x normal tissues). (A.) Total RNA. (B.) MicroRNA. Blue dots: upregulated transcripts in tumor samples. Red dots: downregulated transcripts in tumor samples. Black dots: statistically nonsignificant expressed transcripts. logFC: log2 fold change. Vertical gray lines: cut-off of logFC < -1 and logFC > 1. Horizontal purple line: FDR cut-off of 0.01. Table S1: differentially expressed data from total RNA. logFC: log2 fold change. Table S2:
differentially expressed miRNAs. logFC: log2 fold change.

(Supplementary Materials)

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