ZAC1 and SSTR2 Are Downregulated in Non-Functioning Pituitary Adenomas but Not in somatotropinomas

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Abstract

Introduction: There are few data regarding ZAC1 expression in clinically non-functioning pituitary adenomas (NFPA). Because somatotropinomas and NFPA behave differently with respect to tumor shrinkage during somatostatin analogs (SA) therapy, we sought to compare the ZAC1 and somatostatin receptor (sstr) types 1, 2, 3 and 5 mRNA expression in these two pituitary adenoma subtypes and in normal human pituitaries.

Methods: ZAC1 and SSTR mRNA expression levels were evaluated using real-time RT-PCR (TaqMan) in 20 NFPA and compared with the expression levels in 23 somatotropinomas and five normal pituitaries. The NFPA invasiveness was evaluated using magnetic resonance imaging with Hardy’s modified criteria. Ki-67 and p53 were evaluated using immunohistochemistry.

Results: A total of 20 patients with NFPA [6 males, median age 56 years (range: 30–78)], 23 with acromegaly [12 males, median age 43 years (range: 24–57)] and five normal pituitaries [4 males, median age 48 years (range: 36–54)] were included. Four of the patients (20%) had Hardy’s grade 2 tumors; all of the others had Hardy’s grade 3 tumors. The Ki-67 median expression was 2.35 (range: 0.2–9.23), and only four of the tumors (20%) were positive for p53. The ZAC1 mRNA expression was significantly lower in NFPA than in somatotropinomas and in normal pituitaries (p<0.001 for both), as well as the SSTR2 (p=0.001 and 0.01, respectively). The SSTR3 expression was higher in the NFPA than in the somatotropinomas and in the normal pituitaries (p=0.03 and 0.02, respectively). No correlation was found between the ZAC1 mRNA expression and the tumor invasiveness, Ki-67 and p53.

Conclusion: ZAC1 and SSTR2 are underexpressed and SSTR3 is overexpressed in NFPA compared to those in somatotropinomas and in normal pituitaries, which might explain the lack of tumor shrinkage that is observed in response to commercially available SA therapy in patients with NFPA.

Introduction

Somatostatin receptors (SSTR) are G-protein-coupled receptors that are encoded by five separate genes (SSTR1–5) [1]. All of the five genes are expressed in normal adult human pituitaries, although SSTR4 mRNA has only been detected at extremely low levels [1]. In pituitary adenomas, SSTR expression has been reported to be highly variable within and between tumor subtypes [2,3]. In clinically non-functioning pituitary adenomas (NFPA), the SSTR2 and 3 mRNA transcripts were expressed in the majority of tumors, whereas the SSTR1, 4 and 5 mRNA transcripts were expressed in a subset of the tumors. The SSTR3 transcript showed the highest expression level, followed by the SSTR2 transcript; the SSTR1, 4 and 5 transcripts exhibited low levels of expression [2]. In somatotropinomas and normal pituitaries, the SSTR2...
and 5 mRNA transcripts and proteins were expressed in all of the samples [2–6]. The SSTR5 mRNA transcript exhibited the higher level of expression, followed by SSTR2, 3, 1 and 4 [2].

The somatostatin analogs octreotide and lanreotide bind preferentially to SSTR2 (but also to SSTR3 and SSTR5) and are the mainstay in the treatment of acromegaly [7]; however, they have shown a lack of efficacy in NFPA [8]. Studies in GH3 cells demonstrated that the antiproliferative action of octreotide is mediated by an induction of Zact1 (zinc finger protein which regulates apoptosis and cell cycle arrest) expression [9–12]. ZAC1 is a seven zinc finger protein that acts as both a transcription factor and a coregulator [13]. Its gene is located in the chromosome 6q24-25, which is a frequent site of loss of heterozygosity (LOH) in several tumors, such as pituitary adenomas [14], ovarian cancer [15], breast cancer [16], pheochromocytoma [17] and hemangioblastoma [18]. It is highly expressed in normal human pituitaries but shows reduced expression in pituitary adenomas, especially NFPA [14,19]. In this pituitary adenoma subtype, ZAC1 immunoreexpression has been found to be inversely correlated with tumor recurrence after surgery [20].

To better understand the lack of efficacy of the SSTR2 somatostatin analogs in NFPA, we evaluated the mRNA expression levels of ZAC1 and SSTR subtypes 1, 2, 3 and 5 in NFPA and compared them to those of somatotropinomas and normal pituitaries. In addition, we correlated these expression levels with those of the proliferative markers Ki-67 and p53 and with the invasiveness of the tumors.

Patients and Methods

This study was approved by the ethics committees of the Clementino Fraga Filho University Hospital/Medical School – Federal University of Rio de Janeiro and of the Ribeirao Preto School – University of São Paulo. This study has been conducted according to the principles expressed in the Declaration of Helsinki. All of the patients signed informed consent forms.

Patients and tumors

Tumor samples from drug-naïve unselected patients with NFPA were included. The diagnosis of NFPA was based on the presence of a magnetic resonance image (MRI) of the sella turcica with an adenoma visualized in the absence of symptoms suggesting hormone hypersecretion, in addition to biochemical confirmation of a normal or hypofunctioning pituitary. This diagnosis was confirmed by a histopathological examination, and immunohistochemistry revealed either negative immunostaining for all of the anterior pituitary hormones or positive immunostaining for LH, FSH and/or the alpha-subunit. We also studied tumor samples from drug-naive acromegalic patients and normal human pituitaries for comparison purposes. The diagnosis of acromegaly was based on the presence of classic clinical features associated with a lack of GH suppression to <1.0 µg/L during an oral glucose tolerance test and a high age-matched IGF-I level. A positive GH staining of the tumor specimen upon immunohistochemical examination was required to confirm the etiology. Five normal human pituitary glands were obtained during autopsy following accidental deaths, with no clinical or pathological evidence of endocrine disorders. The time between death and sample collection was 6–12 h. Pathological examination excluded the presence of pituitary adenoma/hyperplasia or metastasis.

The NFPA were classified as invasive or non-invasive according to radiological criteria using Hardy’s modified criteria [21] [19]. All of the tumors were evaluated using preoperative MRI, and all of the images were analyzed by the same radiologist (ELG), who was blinded to the RT-PCR and immunostaining results. Briefly, grade 1 tumors are microadenomas (<1 cm diameter), and grade 2 tumors are macroadenomas (>1 cm diameter) with or without suprasellar extension. Grade 3 tumors are locally invasive, with evidence of bone destruction and the presence of the tumor within the sphenoid and/or cavernous sinus. Grade 4 tumors exhibit central nervous system/extracranial spread with or without distant metastases. Grade 3 and 4 tumors are considered invasive.

Assessment of ZAC1 and SSTR1, 2, 3 and 5 mRNA expression levels using real-time RT-PCR

The total RNA was extracted from a 30-µg fresh tissue sample obtained during transtransphenoidal surgery using the RNeasy® Mini Kit (Qiagen, Valencia, CA). To avoid contamination by genomic DNA, the RNA samples were treated with an RNase-free DNase Set (Qiagen), as specified in the manufacturer’s protocol. The integrity of the total RNA was confirmed using standard agarose gel electrophoresis with ethidium bromide staining. The RNA quantification was performed using a spectrophotometer NanoDrop 1000 (NanoDrop Technologies, Wilmington, Delaware, USA) at a wavelength of 260 nm. The ratio of the absorbances at 260 nm and 280 nm was used to verify the purity of the RNA. The synthesis of first-strand cDNA suitable for PCR amplification was performed using the enzyme MultiScrib® kit and high capacity cDNA Reverse Transcription (Applied Biosystems, CA, USA) with 500 ng of RNA (final volume 20 µL), according to protocols supplied by the manufacturer.

To control for variations in the amount of sample and the RNA quality used in the RT reaction and the efficiency of the RT reaction, the expression levels of three housekeeping genes [TATA-binding protein (TBP), β-glucuronidase (GUS) and phosphoglycerate kinase-1 (PGK1)] were determined for each sample. To determine whether these genes were appropriate to be used as internal controls, the stability of expression was calculated using the GeNorm 3.3 visual basic application for Microsoft Excel, as previously developed and validated by Vandesompele et al. [22]. This program calculates the average pairwise variation of a particular gene with all of the other control genes, allowing the elimination of the worst-scoring control genes and recalculation. The geometric means of the copy numbers for the most stable genes are then used as a normalization factor. In the current study, the three housekeeping genes were confirmed as being stable.

The real-time RT-PCR reactions were performed using the TaqMan® method. The amplification reactions were made in duplicate, using 96-well plates with 0.5 µL of primers + probe, 5
μL of master mix, 2.5 μL of cDNA and 2.0 μL of DEPC water. The thermal cycling profile consisted of an initial temperature of 50°C for 2 minutes, followed by a denaturation at 95°C for 10 minutes and then 40 successive cycles at 95°C for 15 seconds and 60°C for 1 minute. The TaqMan® assays used in each reaction are identified in Table 1. The total RNA sample that was not reverse-transcribed was run to control for genomic contamination.

Immunohistochemistry

Immunohistochemistry for Ki-67 and p53 was performed according to the protocol previously described [25]. Briefly, the tissues were submitted to heat-mediated antigen retrieval in Dako® Retrieval buffer (Dako, Carpinteria, CA, USA), pH 9.0 in a steamer for 30 minutes for ki-67 and in 0.01 M citric acid buffer, pH 6.0 in a microwave for 10 minutes at 750 W for p53. The primary antibody incubation was performed overnight with antibodies directed against the Ki-67 antigen (1:100, Dako, MIB-1 clone, cat. number M-7240) and p53 (1:100, Santa Cruz Biotechnologies, Santa Cruz, CA, USA, DO1 clone, cat. number sc-126), which were detected using the EnVision peroxidase kit (Dako, K1491), followed by the chromogen substrate diaminobenzidine (Liquid DAB, Dako K3468). Quantification was performed by counting the immunolabeled and unlabeled nuclei and then calculating the percentage of positive cells (Labeling Index – LI).

Table 1. Quantitative real-time RT-PCR assays.

| Gene  | TaqMan assay          |
|-------|-----------------------|
| ZAC1  | Hs00414677_m1         |
| SSTR1 | Hs0265617_s1          |
| SSTR2 | Hs00990356_m1         |
| SSTR3 | Hs01066399_m1         |
| SSTR5 | Hs00990408_s1         |
| TBP   | Hs_00939627_m1        |
| GUS   | Hs_99999906_m1        |
| PGK1  |                      |

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Spearman’s correlation test. A p-value of less than 0.05 was considered significant.

Results

Patients and samples characteristics

Twenty patients with NFPA [6 males, median age 56 years old (30-78)], 23 with acromegaly [12 males, median age 43 years old (24-57)] and five normal pituitary tissues [4 males, median age 48 years old (36-54)] were included in the study. All of the patients with NFPA presented macroadenomas; two (10%) had null cell adenomas, and the others showed immunopositivity for LH, FSH and/or α-subunit (gonadotrophinomas). Four of the patients (20%) had Hardy’s grade 2 tumors, and all of the others had Hardy’s grade 3 tumors. All of the tumors exhibited immunostaining for Ki-67, with a variable level of expression.
Table 2. Demographic, ZAC1, somatostatin receptors subtypes and Ki-67 data in non-functioning pituitary adenomas.

| Age  | Sex | IHC | Hardy | ZAC1 | SSTR1 | SSTR2 | SSTR3 | SSTR5 | Ki-67 | p53 |
|------|-----|-----|-------|------|-------|-------|-------|-------|-------|-----|
| 1    | 54  | F   | G     | 2    | 0.004 | 0.016 | 6.924 | 0.009 | 0.010 | 0.20 | 0   |
| 2    | 74  | F   | G     | 2    | 0.002 | 7.103 | 0.102 | 15.084 | 4.387 | 1.16 | 0   |
| 3    | 30  | F   | G     | 2    | 0.039 | 43.72 | 0.066 | 3.540 | 34.534 | 5.66 | 0.26 |
| 4    | 50  | F   | G     | 2    | 0.028 | 0.038 | 0.231 | 2.312 | 0.055 | 2.71 | 0   |
| Median| 52  |     |       | 0.016 | 3.571 | 0.166 | 2.926 | 2.221 | 1.935 | 0   |

| Invasive | Sex | IHC | Hardy | ZAC1 | SSTR1 | SSTR2 | SSTR3 | SSTR5 | Ki-67 | p53 |
|----------|-----|-----|-------|------|-------|-------|-------|-------|-------|-----|
| 5        | 39  | F   | G     | 3    | 0.001 | 0.010 | 0.052 | 0.726 | 0.042 | 9.23 | 0.63 |
| 6        | 58  | F   | G     | 3    | 0.018 | 10.193 | 2.589 | 2.421 | 2.988 | 0.02 | 1.99 |
| 7        | 57  | F   | G     | 3    | 0.027 | 4.741 | 1.314 | 11.639 | 5.256 | 0.00 | 2.36 |
| 8        | 58  | M   | G     | 3    | 0.001 | 0.200 | 0.08  | 5.304 | 0.055 | 0.298 | 0.15 |
| 9        | 52  | M   | G     | 3    | 0.003 | 0.227 | 0.041 | 7.189 | 0.096 | 0.06 | 0.00 |
| 10       | 56  | F   | M     | 3    | 0.010 | 0.968 | 0.017 | 3.154 | 0.063 | 0.001 | 3.02 |
| 11       | 44  | F   | M     | 3    | 0.007 | 3.477 | 0.039 | 7.372 | 2.472 | 2.89 | 0.00 |
| 12       | 68  | M   | G     | 3    | 0.068 | 1.663 | 0.088 | 1.195 | 1.378 | 2.32 | 0.00 |
| 13       | 66  | F   | N     | 3    | 1.267 | 0.026 | 0.014 | 0.018 | 0.093 | 4.07 | 0.00 |
| 14       | 56  | F   | G     | 3    | 0.002 | 0.174 | 0.065 | 4.981 | 0.275 | 2.62 | 0.00 |
| 15       | 64  | M   | G     | 3    | 0.038 | 0.247 | 0.151 | 13.468 | 0.323 | 1.85 | 1.1 |
| 16       | 34  | F   | G     | 3    | 0.001 | 0.245 | 0.013 | 9.105 | 0.328 | 2.33 | 0.00 |
| 17       | 75  | M   | G     | 3    | 0.015 | 1.059 | 0.27  | 5.677 | 0.964 | 2.00 | 0.7 |
| 18       | 50  | M   | N     | 3    | 0.024 | 0.048 | 0.257 | 4.166 | 2.710 | 1.15 | 0.00 |
| 19       | 54  | F   | G     | 3    | 0.005 | 0.299 | 0.024 | 2.795 | 0.470 | 2.20 | 0.00 |
| 20       | 78  | F   | G     | 3    | 0.001 | 9.538 | 0.165 | 4.028 | 7.974 | 0.28 | 0.00 |
| Median   | 56.5|     |       | 0.008 | 0.273 | 0.073 | 4.574 | 0.587 | 3.25 | 0.00 |

Legend: F = female; M = male; IHC = immunohistochemistry for LH, FSH and α-subunit; G = gonadotropinoma; N = null cell adenoma; SSTR = somatostatin receptor

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In the acromegalic patients, two (8.7%) harbored a microadenoma. The median GH at diagnosis was 8.1 ng/mL (0.9-235), whereas only four (20%) of the tumors were positive for p53 (Table 2).

In the somatotropinomas, this expression was variable; the median was 0.739 and it varied from 0.12 to 3.321 (Table 3). The ZAC1 mRNA expression was different among the three groups (p < 0.001). The expression level was 106 times lower than in the somatotropinomas and 140 times lower than in the normal pituitaries (p values < 0.001 for both) [Table 4; Figure 1].

ZAC1 mRNA expression

The ZAC1 mRNA was expressed at very low levels in all but one of the NFPA, with a median of 0.007, varying from 0 to 1.267 (Table 2). In the somatotropinomas, this expression was variable; the median was 0.739 and it varied from 0.12 to 3.321 (Table 3). The ZAC1 mRNA expression was different among the three groups (p < 0.001). The expression level was 106 times lower than in the somatotropinomas and 140 times lower than in the normal pituitaries (p values < 0.001 for both) [Table 4; Figure 1].

No correlation was found between the ZAC1 and the SSTR2 or the other SSTR subtype mRNA expression levels in the NFPA.

Comparison of ZAC1 expression with Ki-67, p53 and invasiveness

No correlation was found between ZAC1 and Ki-67 or p53. In addition, the ZAC1 expression level was similar in the invasive and non-invasive NFPA.

Discussion

In this study, we demonstrated for the first time that ZAC1 mRNA was underexpressed in NFPA relative to normal human pituitaries and somatotropinomas using a quantitative technique (real-time RT-PCR). Similar data have been previously reported; however, that studies involved a limited number of patients and a semi-quantitative technique (conventional RT-PCR) [14,19] [12,17]. On the other hand, ZAC1 mRNA expression was similar in somatotropinomas and normal human pituitaries. In respect to the SSTR, lower SSTR2 and higher SSTR3 mRNA expression levels were found in the NFPA compared with both the normal pituitaries and the somatotropinomas. A previous study from our group also found a lower SSTR2 mRNA expression in NFPA in comparison to
expression levels in non-functioning pituitary adenomas, SSTR3

Table 3. Demographic, ZAC1 and somatostatin receptors subtypes data in somatotropinomas.

|   | Age | Sex | ZAC1 | SSTR1 | SSTR2 | SSTR3 | SSTR5 |
|---|-----|-----|------|-------|-------|-------|-------|
| 1 | 30  | F   | 1,151| 0,007 | 0,374 | 0,005 | 1,802 |
| 2 | 37  | F   | 0,785| 0,033 | 15,505| 0,11  | 0,005 |
| 3 | 36  | M   | 0,307| 0,038 | 0,122 | 324,759| 0,722 |
| 4 | 32  | M   | 0,384| 3,694 | 0,234 | 0,126 | 0,014 |
| 5 | 56  | F   | 0,352| 0,026 | 0,535 | 0,045 | 0,471 |
| 6 | 53  | F   | 3,321| 0,141 | 0,264 | 8,315 | 1,201 |
| 7 | 31  | F   | 0,059| 0,036 | 0,256 | 10,028| 0,66  |
| 8 | 42  | M   | 0,302| 0,018 | 2,869 | 0,454 | 0,012 |
| 9 | 57  | M   | 0,739| 0,09  | 0,321 | 12,857| 4,046 |
| 10| 50  | F   | 1,605| 0,034 | 1,026 | 4,101 | 0,337 |
| 11| 24  | M   | 0,429| 0,008 | 6,509 | 0,002 | 0,03  |
| 12| 42  | F   | 0,624| 0,092 | 1,586 | 2,094 | 0,943 |
| 13| 43  | F   | 1,041| 0,068 | 1,793 | 5,988 | 0,648 |
| 14| 37  | F   | 0,519| 0,232 | 0,04  | 0,988 | 0,526 |
| 15| 43  | M   | 1,966| 0,097 | 2,919 | 1,131 | 1,261 |
| 16| 36  | M   | 0,814| 1,012 | 0,319 | 1,629 | 3,362 |
| 17| 40  | F   | 0,062| 7,463 | 0,037 | 2,582 | 2,53  |
| 18| 55  | M   | 0,701| 1,502 | 0,199 | 0,159 | 2,233 |
| 19| 50  | M   | 0,12 | 5,146 | 1,727 | 1,444 | 0,534 |
| 20| 48  | M   | 0,936| 4,144 | 1,855 | 0,055 | 4,356 |
| 21| 49  | F   | 1,052| 5,898 | 1,017 | 0,169 | 3,012 |
| 22| 43  | M   | 1,466| 1,227 | 1,951 | 2,308 | 0,44  |
| 23| 46  | M   | 0,318| 0,077 | 22,949| 0,113 | 0,346 |
| Median | | | 0,739 | 0,092 | 0,107 | 1,131 | 0,66 |

Legend: F = female; M = male; SSTR = somatostatin receptor

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Tumor shrinkage has been observed in approximately 40% of somatotropinomas and in 11–13% of NFPA [8] after treatment with octreotide and lanreotide [26], which bind preferentially to SSTR2. This effect occurs despite the fact that the SSTR subtype with the highest level of expression in these tumors is SSTR3, which is the main SSTR subtype associated with apoptosis [27]. Theodoropoulou et al. [10] demonstrated that, in GH3 cells (a rat somatomammotroph pituitary cell line that expresses SSTR1 and 2), the anti-proliferative action of octreotide occurred by increasing Zac1 gene expression. In addition, Zac1 gene knockdown abolishes the anti-proliferative effect of octreotide. These data indicate that Zac1 is involved in the anti-proliferative action of somatostatin analogs and that its expression is necessary for this effect. Consequently, the low Zac1 expression in NFPA may explain, at least in part, the lack of response of these tumors to somatostatin analog treatment. However, apart from the SSTR or Zac1 expressions, differences in the pathogenesis of somatotropinomas and NFPA are also likely to account for the different response to somatostatin analogs in these adenoma subtypes.

On the other hand, Theodoropoulou et al. [28] recently also demonstrated in GH3 cells that Zac1 inhibition by siRNA suppressed Sstr2 expression. In contrast, increasing Zac1 expression by treatment with the PI3K inhibitors LY294002 and wortmannin led to an increase in Sstr2 expression. Although these data was evaluated in GH3 cells, which is not an appropriate model for NFPA, we hypothesize that, analogously, the low Zac1 expression might contribute to explain the low SSTR2 expression observed in NFPA. However, we did not find correlation between Zac1 and SSTR2 expression in NFPA or somatotropinomas.

The transfection of Zac1 in LLC-PK1 (porcine kidney epithelial) and SaO2-2 (human osteogenic sarcoma) cell lines caused a substantial reduction in cell proliferation, comparable with transfection with p53 [26]. Whereas the ablation of Zac1 gene expression using antisense treatment in AtT-20 (mouse corticotroph pituitary tumor) and TtT/GF (folliculo-stellate) cell lines stimulated DNA synthesis in 36–50% [29]. These data may indicate a possible theoretical role of the low Zac1 expression in the pathogenesis of NFPA, however, this hypothesis still needs to be further studied.

We examined the proliferative markers Ki-67 and p53 using immunohistochemistry, and no correlations were observed of these markers to the Zac1 expression levels in NFPA. This result may be explained by the very low Zac1 expression in all of the tumors except one, suggesting that low expression of Zac1 is a characteristic of NFPA as a group. Curiously, the tumor with the highest Zac1 expression also presented a high Ki-67 LI.

Noh et al. [20] analyzed several markers associated with cell proliferation or apoptosis in 35 NFPA and tumor recurrence. They found that recurrent NFPA had higher levels of Ki-67 and TUNEL labeling indexes. In addition, high level of expression of phospho-Akt, phospho-p44/42 MAPK and PTTG1 were associated with early recurrence. On the other hand, phospho-

Table 4. ZAC1 and somatostatin receptor subtype mRNA expression levels in non-functioning pituitary adenomas, somatotropinomas and normal pituitaries.

|   | NFPA | Somatotropinomas | Normal pituitaries | p value† |
|---|------|------------------|--------------------|---------|
| ZAC1 | 0.007 (0.1-2.671) | 0.739 (0.12-3.321) | 0.979 (0.313-2.089) | < 0.001 |
| SSTR1 | 0.273 (0.01-43.72) | 0.092 (0.007-7.463) | 0.903 (0.299-1.719) | NS |
| SSTR2 | 0.084 (0.013-6.924) | 1.017 (0.037-22.949) | 1.281 (0.272-1.672) | < 0.001 |
| SSTR3 | 0.497 (0.009-15.084) | 1.131 (0.002-324.759) | 1.171 (0.525-1.366) | < 0.001 |
| SSTR5 | 0.566 (0.01-34.534) | 0.66 (0.005-4.356) | 0.858 (0.487-2.033) | NS |

Data expressed in median (min-max).

The Kruskall-Wallis test (†) was used to compare the mRNA expression among the three groups.

The Mann-Whitney test was used for comparison between NFPA and normal pituitaries and somatotropinomas (a = < 0.001; b = 0.001; c = 0.01; d = 0.03; e = 0.02).

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CREB and ZAC1 expression levels were inversely associated with recurrence. We compared the ZAC1 expression levels with the tumor invasiveness, but no association was found. A previous study from our group already compared the levels of Ki-67 and p53 between invasive and non-invasive NFPA in a larger subset of tumors, which included some of the tumors described here (16 ACNF and 5 somatotropinomas), and found similar results [25]. This lack of an association between the ZAC1 levels and invasiveness may be due to the very low ZAC1 mRNA expression, as was the case for Ki-67 and p53. Another explanation is that the high frequency of invasive tumors in our series may have led to a type II error. A possible cause for this high frequency is that this series examined tumors that required operations. Smaller and non-invasive tumors frequently do not require operations, so in a series of operated tumors, the incidence of invasive tumors would be expected to increase. Another explanation may be that the study was held at a referral center, which receives more advanced and complicated cases.

As stated above, the SSTR3 is the main subtype associated with apoptosis [27], and the fact that NFPA express this receptor in a large amount may be of interest with the development of new SA, with broader spectrum of binding to SSTR, such as pasireotide, or SSTR3 specific. Florio et al. [30] demonstrated in vivo, using matrigel sponge model, that a SSTR3 specific agonist, L-796778, significantly inhibited angiogenesis. This effect was reversed by a SSTR3 antagonist, BN81658. Data about pasireotide treatment of patients with NFPA is still lacking. In this series, we did not find any difference in gender, age, ZAC1 or Ki-67 between adenomas with high or low SSTR3 expression (data not shown).

In conclusion, we found that ZAC1 and SSTR2 expression levels are reduced in NFPA compared to somatotropinomas and normal pituitaries. Despite the function of ZAC1 as a tumor suppressor gene, we did not find any correlations between its level of expression and those of proliferative markers or invasiveness. However, the low expression levels of ZAC1 and SSTR2 may be involved in the resistance to somatostatin analogs that is observed in these tumors, partially justifying the failure of tumor shrinkage that is observed in NFPA compared to somatotropinomas.

**Author Contributions**

Conceived and designed the experiments: MC MRG. Performed the experiments: LVN LEW LMC ABM NVM LK. Analyzed the data: LVN LEW. Contributed reagents/materials/analysis tools: ELG. Wrote the manuscript: LVN LEW.
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