Retrospective study of Dana Farber Consortium Protocol in newly diagnosed Egyptian adolescents and young adults with acute lymphoblastic leukemia: Tanta experience

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Abstract

Background: Intensive acute lymphoblastic leukemia (ALL) regimens in children improve the 5-year event-free survival (EFS) to reach ~ 90%. Adolescents and young adults (AYA) have EFS (30% to 45%). Young AYA ALL patients treated with pediatric chemotherapy protocols such as Dana Farber Consortium Protocol (DFCP) experience a better prognosis. This study aimed to assess the efficacy [EFS and overall survival (OS)] and the toxicity of DFCP in the treatment of Egyptian AYA with newly diagnosed ALL. A retrospective study was performed on 41 patients with newly diagnosed ALL (15 and 39 years) who were treated with DFCP. EFS and OS were estimated using the Kaplan-Meier method.

Results: Thirty-eight patients (92.68%) achieved complete remission (CR). Eleven patients (26.83%) relapsed. Ten (24.39%) patients died. One, two, and three years of EFS were 75.61%, 72.91%, and 67.51% respectively. One, two, and three years OS were 85.3%, 77.26%, and 74.39% respectively. Neutropenia was the most common adverse event observed in 100% of patients.

Conclusion: DFCP can be considered as an effective ALL protocol for the AYA group of patients with good CR, EFS, and OS rates. DFCP seemed to be feasible in AYA despite the toxicities experienced.

Keywords: Acute lymphoblastic leukemia (ALL), Dana Farber Consortium Protocol (DFCP), Adolescents and young adults (AYA), Event-free survival (EFS), Overall survival (OS)
patient biology differences [6, 7]. The identification of special characters of AYA with ALL has led to an improvement of EFS, with EFS now ~ 70% [8]. Young AYA (15–21 years at diagnosis) ALL patients treated with pediatric chemotherapy protocols such as Dana Farber Consortium Protocol (DFCP) experience better prognosis when compared with AYA treated with adult protocols [9].

So, this study aimed to assess the efficacy [Event free survival (EFS) and overall survival (OS)] and the toxicity of the DFCP in the treatment of Egyptian adolescents and young adults with newly diagnosed acute lymphoblastic leukemia.

Methods

Patients

This was a retrospective study performed on 41 patients with newly diagnosed ALL who were treated with DFCP selected consecutively and diagnosed between January 2014 and December 2017 at Tanta Cancer Center (acute leukemia unit) and Hematology Unit, Internal Medicine Department, Faculty of Medicine, Tanta University.

The diagnosis of ALL was based on the characteristic presence of 20% or more lymphoblast in bone marrow (BM) examination and was confirmed by immunophenotyping and cytogenetic and/or molecular studies [10].

Eligibility criteria

The patients included in this study were newly diagnosed ALL with no prior chemotherapy, aged between 15 and 39 years.

ALL patients with performance status Eastern Cooperative Oncology Group (ECOG) (3 or more) and patients with chronic renal, hepatic, or uncontrolled severe cardiovascular disease and pregnant women were excluded. Patients with an uncontrolled active infection, human immunodeficiency virus, active psychiatric illness, cerebrovascular accident or hemorrhage, or prior history of pancreatitis were also excluded.

Treatment protocol: (Table 1)

All patients received DFCP with supportive therapy [11]. Imatinib or dasatinib was added to the protocol in the case of Philadelphia positive [12].

Data collection

Data were collected by reviewing patients’ records. Records with incomplete data (four patients) were omitted from the study. Every patient has a file with a private code number to ensure the privacy of our patients. All patients’ data were handled according to ethical standards in accordance with the Declaration of Helsinki. Signed informed consents from all alive and still followed up included patients were taken before the starting of data collection.

Data that were collected included age, sex, history, physical examination, and initial laboratory investigations including complete blood count (CBC), BM aspiration, immunophenotyping, cytogenetics study, and BCR-ABL1 test.

Follow-up

While patients on therapy complete blood counts and BM aspirations with other routine investigations were done for assessing the patient’s response. The patients were followed up weekly by complete blood counts for 33 weeks during phase II induction, central nervous system (CNS) prophylaxis, and phase III intensifications therapy. After the completion, patients were followed up by the complete physical examination, blood cell counts, and routine chemistry; every month during the first year. Thereafter, every 3 months in the 2nd year then half-yearly from the 3rd year. BM aspiration was done every 6 months or as needed clinical for 5 years. BCR-ABL1 was done periodically every 3 months for Philadelphia positive patients [8].

Minimal residual disease (MRD) was evaluated post-induction and those who achieved more than 0.1% were referred for allogeneic BMT with Philadelphia positive and relapsed patients [8].

Response to treatment [8]

Complete remission

Less than 5% blasts in BM, no peripheral blood blasts, absolute neutrophil count at least 1 × 10^9/L, platelet counts of at least 100 × 10^9/L, and no extra-medullary disease, if only platelets count and/or absolute neutrophil count not reach to the target complete remission (CR) with incomplete blood count recovery.

Refractory disease

Failure to achieve CR at the end of induction.

Relapsed disease

The reappearance of blasts in the blood or bone marrow (> 5%) or any extra-medullary site after CR.

Outcomes

Event-free survival (EFS) was calculated from the date of ALL diagnosis till the date of treatment failure, ALL relapse, last follow-up, or death. Overall survival (OS) was calculated from the date of ALL diagnosis to the date of last follow-up or death.

Statistical analysis

The collected data were analyzed using SPSS version 23 software (SPSS Inc, Chicago, ILL Company). Non-
parametric distributed quantitative data are expressed as median and range. Categorical variables are expressed as numbers and percentages. Cumulative Incidence of EFS and OS were estimated using the Kaplan-Meier method. \( P \) value \( \leq 0.05 \) was considered statistically significant.

### Results

#### Patients’ characteristics: (Table 2)

Forty-one patients with newly diagnosed ALL were included in our study with a median age of the patients was 28 years (range 18–39 years). Male was the predominant

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**Table 1** Dana Farber Consortium Protocol used in the study [11, 12]

| Phase                          | Duration | Drug               | Dose            | Route of administration | Days | Notes                                           |
|-------------------------------|----------|--------------------|-----------------|-------------------------|------|------------------------------------------------|
| Phase I induction             | 4 weeks  | Prednisone         | 40 mg/m²        | Oral                    | D1-29| Maximum dose 2 mg                               |
|                               |          | Vincristine        | 1.4 mg/m²       | Intravenous             | D1,8,15,22 |                                           |
|                               |          | Doxorubicin        | 30 mg/m²        | Intravenous             | D1,2 |                                           |
|                               |          | Methotrexate       | 1.5 g/m²        | Intravenous             | D3   | leucovorin rescue                               |
|                               |          | L-asparaginase     | 25,000 IU/m²    | Intramuscular           | D5   |                                               |
|                               |          | Cytarabine         | 40 mg           | Intrathecal             | D1   |                                               |
|                               |          | Methotrexate       | 12 mg           | Intrathecal             | D15,29|                                               |
|                               |          | Hydrocortisone     | 15 mg           |                         |      |                                               |
|                               |          | IMatinib           | 600 mg          |                         | D3-15| Ph +ve patients only                           |
| Phase II induction and CNS therapy | 3 weeks | Vincristine        | 1.4 mg/m²       | Intravenous             | D1   | Maximum dose 2 mg                               |
|                               |          | Doxorubicin        | 30 mg/m²        | Intravenous             | D1   |                                               |
|                               |          | 6-mercaptopurine   | 50 mg/m²        | Oral                    | D1-14|                                               |
|                               |          | Methotrexate       | 12 mg           | Intrathecal             | D1, 4, 8, 11 |                                           |
|                               |          | Cytarabine         | 40 mg           | Intrathecal             | D1, 4, 8, 11 |                                           |
|                               |          | Hydrocortisone     | 15 mg           |                         |      |                                               |
|                               |          | Cranial irradiation| 1200 centigray  |                         | Over 8 days | With very high initial total leucocytic count or CNS infiltration unless patient will have BMT |
| Phase III Intensification therapy | 10 cycles (cycle every 3 weeks) | Dexamethasone | 9 mg/m² twice daily | Oral | D1-5 |                                               |
|                               |          | 6-mercaptopurine   | 50 mg/m²        | Oral                    | D1-14|                                               |
|                               |          | Vincristine        | 1.4 mg/m²       | Intravenous             | D1   | Maximum dose 2 mg                               |
|                               |          | L-asparaginase     | 12,500 IU/m²    | Intramuscular           | D1, 8, 15 |                                           |
|                               |          | Doxorubicin        | 30 mg/m²        | Intramuscular           | D1   | In the first 7 cycles                           |
|                               |          | Methotrexate       | 30 mg/m²        | Intramuscular or oral   | D2, 9, 16 | In last 3 cycles                               |
|                               |          | Methotrexate       | 12 mg           | Intrathecal             | D1   | 6th cycle only                                 |
|                               |          | Cytarabine         | 40 mg           | Intrathecal             | D1   |                                               |
|                               |          | Hydrocortisone     | 15 mg           |                         |      |                                               |
| Phase IV Continuation therapy | 24 cycles (cycle every 3 weeks) | Dexamethasone | 6 mg/m² twice daily | Oral | D1-5 |                                               |
|                               |          | 6-mercaptopurine   | 50 mg/m²        | Oral                    | D1-14|                                               |
|                               |          | Vincristine        | 1.4 mg/m²       | Intravenous             | D1   | Maximum dose 2 mg                               |
|                               |          | Methotrexate       | 30 mg/m²        | Intramuscular or oral   | Weekly|                                               |
|                               |          | Methotrexate       | 12 mg           | Intrathecal             | Every 18 weeks at the start of the cycle | Every 18 weeks at the start of the cycle |
|                               |          | Cytarabine         | 40 mg           |                         |      |                                               |
|                               |          | Hydrocortisone     | 15 mg           |                         |      |                                               |

*BM*T bone marrow transplantation, CNS central nervous system, D day, Ph Philadelphia
gender as (68.29%) were males. Immunophenotyping study revealed that 30 patients (73.17%) were B-ALL. The cytogenetic study revealed 27 patients (65.85%) had normal karyotyping and eight patients (19.51%) were Philadelphia chromosome-positive. Allogeneic BMT from siblings was done for 10 patients (3 Philadelphia positive, 5 relapsed, and 2 with MRD). The remaining 5 Philadelphia chromosome-positive patients were not subjected to allogeneic BMT due to the lack of donors, these patients received tyrosine kinase inhibitors (imatinib or dasatinib).

Outcome results: (Table 3)
Thirty-eight patients (92.68%) achieved complete remission (CR), only 3 (7.32%) were refractory. Eleven patients (26.83%) relapsed (seven in the 1st year, three in the 2nd year, and one after that). Two patients out of the eleven relapsed patients had CNS relapse manifested by facial palsy and persistent headache with severe vomiting. Ten (24.39%) patients died (three just after induction after they became refractory, and seven of them died after relapse).

The median follow-up period was 42 months (95% CI, 27.981–38.068). One-, two-, and three-year event-free survivals (EFS) were 75.61%, 72.91%, and 67.51% respectively (Fig. 1). One-, two-, and three-year overall survivals (OS) were 85.3%, 77.26%, and 74.39% respectively (Fig. 2).

Toxicity: (Table 4)
All the patients (100%) had DFCP side effects; some patients had more than one side effect. No major side effects were observed that necessitated treatment discontinuation. Neutropenia was the most common adverse event observed in 41 patients (100%). Many other adverse events were also developed in patients under DFCP in the form of thrombocytopenia, febrile neutropenia, infections, hyperglycemia, mucositis, venous thromboembolism, neuropathy, hepatotoxicity, pancreatitis, and avascular necrosis. No other long-term side effects were developed during the follow-up period. No patient developed an allergy.

Raw data of all patients of the study were mentioned in (Table 5).

Discussion
Adolescent and young adult (AYA) ALL patients represent a special patient group, as they may receive chemotherapy based on either pediatric or adult chemotherapy protocol [13]. The favorable outcomes found in children, coupled with unfavorable outcomes noticed in AYA, generated the idea for practical use of pediatric protocols among AYA. Also, many novel drugs such as Blinatumomab, inotuzomab, and chimeric antigen receptor T cell have been established to improve outcomes in poorly responding, relapsed, or refractory B cell ALL [14].

In ALL, AYA patients have poorer EFS and OS compared to children, because AYA tends to have unfavorable characteristics, such as T cell phenotype, more incidence of Philadelphia chromosome (9;22), and less occurrence of favorable chromosomal abnormalities such as hyperdiploidy [15]. In our study, immunophenotyping study revealed 26.83% of our patients were T-ALL, and 19.51% were Philadelphia chromosome-positive. These results coincide to a great extent with literatures, as in adults, T-ALL accounts for about 25% of cases, also Philadelphia chromosome in adults ALL can range from 15 to 50% increasing with age [16].

Several retrospective studies have shown that AYA patients who received pediatric protocols have better outcomes (CR 90–99% and EFS 63–80%) when compared with AYA patients who received adult protocols (CR 80–94% and EFS 34–71%) [17]. Although, one study by Usvasalo et al. [17] showed no superiority of both pediatric and adult protocols regarding CR and EFS.

| Table 2 | Characteristics of the study population |
|-----------------|-----------------|
| Variables | Median | Range |
| Age (years) | 28 | 18–39 |
| Hemoglobin (g/dl) | 9 | 5–14 |
| Total leucocytic count x10⁹/L | 14 | 1–138 |
| Platelets x10⁹/L | 60 | 8–253 |
| Bone marrow blast (%) | 73 | 22–96 |
| Variables | Number (%) |
| Sex | Male 28 (68.29%) Female 13 (31.71%) |
| Immunophenotyping | B-ALL 30 (73.17%) T-ALL 11 (26.83%) |
| Cytogenetics | Normal 27 (65.85%) Abnormal 14 (34.15%) |
| Philadelphia chromosome | Positive 8 (19.51%) Negative 33 (80.49%) |

| Table 3 | Therapy outcomes of the study population |
|-----------------|-----------------|
| Variables | Number (%) |
| Response | Complete remission (CR) 38 (92.68%) Refractory 3 (7.32%) |
| Relapse | Relapse 11 (26.83%) Non-relapse 30 (73.17%) |
| Survival | Living 31 (75.61%) Dead 10 (24.39%) |
Several factors may clarify better outcomes in pediatric protocols. Firstly, pediatric protocols have more non-myelosuppressive drugs with more activity on leukemic cells particularly during the BM suppression phase induced by anti-metabolites and anthracyclines. Moreover, in pediatric protocols, CNS prophylaxis as intrathecal methotrexate was administered earlier, more frequently, and for a longer time as CNS is the shelter for blast cells. Also, the maintenance therapy period is shorter in adult protocols [18, 19].

In our study, forty-one patients received the pediatric protocol DFCP, Thirty-eight patients (92.68%) achieved complete CR, eleven patients (26.83%) relapsed, and ten (24.39%) patients died. One-, two-, and three-year EFS were 75.61%, 72.91%, and 67.51% respectively. One-,
two-, and three-year OS were 85.3%, 77.26%, and 74.39% respectively.

These results coincide to a great extent with some differences reported in many studies. DeAngelo et al. [20] used DFCP for the treatment of 74 adult patients (18–50 years) with ALL. Eighty-four percent of the patients achieved CR, with 2-year EFS and OS of 72.5% and 77.1%, respectively.

Barry et al. [11] treated 51 de novo ALL patients (15–18 years) with DFCP, and found that the 5-year EFS and OS were 77.5% and 81% respectively. Also, Storing et al. [21] used a modified DFCP in 68 patients (17–71 years), 82% of the patients achieved CR, with 3-year EFS and OS of 77% and 65%, respectively.

Furthermore, Al-Khabori et al. [22] conducted a retrospective study on T-ALL patients, 32 AYA patients treated with a DFCP. Ninety-three percent of the patients achieved CR with 3-year relapse-free survival and OS 88% and 83% respectively. Besides, DeAngelo et al. [23] enrolled 92 patients (18–50 years) for a median follow-up period (4.5 years). Eighty-five percent achieved CR, with 4-year EFS and OS 69% and 67% respectively.

Lastly, Alabdulwahab et al. [24] enrolled 38 patients with a median age (19 years) or a median follow-up period (22 months). Then, 92.1% achieved CR, with 1- and 3-year EFS were 80% and 68%, respectively, and 1- and 3-year OS were 88% and 72%, respectively.

As ALL therapies have more intensified, more toxicities have been increased especially in AYA patients. The toxicities of intensified protocols increase in both incidence and severity due to hormonal changes, body weight changes, and different chemotherapy metabolism [25]. Researchers are focused on decreasing early and late toxicities by good use of supportive care to improve OS [26].

Pancreatitis and venous thromboembolism are usually linked with L-asparaginase use in DFCP but may be related to increased steroid dose and the use of dexamethasone [27, 28]. Avascular necrosis incidence was more experienced in DFCP trials using dexamethasone [29]. Hyperglycemia risk is more likely in the AYA patients of ALL due to adult hormonal changes, and is usually associated with immune suppression and the risk for infections [30].

In our study, all patients (100%) had developed side effects. Neutropenia was the most common adverse event observed in 100% of the patients. Many other adverse events were also developed in patients under DFCP in the form of thrombocytopenia (70.73%), febrile neutropenia (68.29%), infections (24.39%), hyperglycemia (24.39%), mucositis (21.95%), venous thromboembolism (9.76%), neuropathy (9.76%), hepatotoxicity (9.76%), pancreatitis (7.31%), and avascular necrosis (7.31%). No patient developed an allergy.

These results match with a great extent with some differences that were reported in many studies. DeAngelo et al. [20] shown in their study that the incidence of venous thromboembolism (19%) and pancreatitis (13%) but this drug-related toxicity was controllable.

DeAngelo et al. [23] demonstrated toxicities of DFCP included thrombocytopenia (82%), hepatic toxicity (62%), infection (61%), hyperglycemia (45%), febrile neutropenia (33%), thrombosis (17%), pancreatitis (11%), stomatitis (11%), bone fractures (8%), osteonecrosis (5%), allergy (5%), and CNS complications (5%).

Alabdulwahab et al. [24] demonstrated toxicities of DFCP included febrile neutropenia (100%), sepsis (29%), pneumonia (26%), typhilitis (21%), myopathy (13%), pancreatitis (13%), osteonecrosis (7.8%), neurological toxicity (5%), and severe liver failure together with renal failure (2.6%). There was no venous thromboembolism was recorded apart from (7%) who had central catheter-related thrombosis.

The discrepancy between results of our study and other previous studies could be explained by variations in number and age of patients, follow-up time, ALL phenotype (B or T), Philadelphia chromosome incidence, the occurrence of favorable, and unfavorable chromosomal abnormalities, ethnic differences, and regimens of chemotherapy used as DFCP has many modifications. To our knowledge, this is the first study to collect data from Egyptian ALL patients. The present study had some limitations such as a small number of patients and a short time of follow-up. Also, our study was retrospective; therefore, unrecognized biases might be considered. To

| Table 4 Toxicity of Dana Farber Consortium Protocol in the study population |
|-----------------------------|-----------------|-----------------|
| Toxicity                   | Number | Percentage (%) |
| Neutropenia                 | 41     | 100             |
| Thrombocytopenia            | 29     | 70.73           |
| Febrile neutropenia         | 28     | 68.29           |
| Infection                  | 27     | 65.85           |
| Hyperglycemia              | 10     | 24.39           |
| Mucositis                  | 9      | 21.95           |
| Venous thromboembolism     | 4      | 9.76            |
| Neutropathy                | 4      | 9.76            |
| Hepatotoxicity             | 4      | 9.76            |
| Pancreatitis               | 3      | 7.31            |
| Avascular necrosis         | 3      | 7.31            |
| No | Age (years) | Sex | Total leucocytic count x10⁹/L | Hemoglobin (g/dl) | Platelets x10⁹/L | Bone marrow blast (%) | Immunophenotyping | Philadelphia chromosome | Cytogenetics | Response | Relapsed | FATE |
|----|-------------|-----|-------------------------------|------------------|-----------------|----------------------|------------------|-----------------------|-------------|----------|----------|------|
| 1  | 31          | M   | 4                             | 9                | 35              | 65                   | B                | Negative              | Normal      | Refractory| N/A      | Dead |
| 2  | 23          | M   | 13                            | 13               | 223             | 66                   | B                | Negative              | Abnormal    | Refractory| N/A      | Dead |
| 3  | 20          | F   | 129                           | 8.3              | 50              | 47                   | T                | Negative              | Abnormal    | CR       | No       | Alive |
| 4  | 31          | M   | 77                            | 10               | 55              | 96                   | B                | Negative              | Normal      | CR       | No       | Alive |
| 5  | 24          | M   | 3                             | 8.6              | 25              | 80                   | B                | Negative              | Normal      | CR       | No       | Alive |
| 6  | 32          | M   | 14                            | 12               | 18              | 72                   | B                | Negative              | Abnormal    | CR       | Yes      | Dead |
| 7  | 32          | M   | 75                            | 13               | 8               | 82                   | B                | Positive              | Abnormal    | CR       | Yes      | Dead |
| 8  | 21          | M   | 97                            | 7.6              | 53              | 70                   | B                | Positive              | Abnormal    | CR       | No       | Alive |
| 9  | 29          | F   | 24                            | 7.7              | 31              | 95                   | B                | Negative              | Normal      | CR       | No       | Alive |
| 10 | 26          | M   | 7                             | 13.6             | 206             | 42                   | T                | Negative              | Normal      | CR       | No       | Alive |
| 11 | 39          | M   | 138                           | 12.5             | 180             | 57                   | T                | Negative              | Normal      | CR       | Yes      | Alive |
| 12 | 27          | F   | 4                             | 9.2              | 27              | 95                   | T                | Negative              | Normal      | CR       | No       | Alive |
| 13 | 33          | M   | 3                             | 9                | 190             | 42                   | T                | Negative              | Normal      | CR       | Yes      | Dead |
| 14 | 18          | M   | 10                            | 11               | 98              | 25                   | T                | Negative              | Normal      | CR       | No       | Alive |
| 15 | 20          | M   | 9                             | 14               | 224             | 27                   | T                | Negative              | Normal      | CR       | No       | Alive |
| 16 | 28          | F   | 4                             | 9                | 160             | 80                   | T                | Negative              | Normal      | CR       | Yes      | Alive |
| 17 | 21          | M   | 9                             | 9                | 89              | 80                   | B                | Positive              | Abnormal    | CR       | No       | Alive |
| 18 | 26          | M   | 3                             | 5                | 80              | 91                   | B                | Positive              | Abnormal    | CR       | No       | Alive |
| 19 | 20          | M   | 7                             | 9.5              | 33              | 95                   | B                | Negative              | Normal      | CR       | No       | Alive |
| 20 | 27          | M   | 41                            | 8.7              | 50              | 94                   | B                | Negative              | Abnormal    | CR       | No       | Alive |
| 21 | 33          | M   | 1                             | 10               | 141             | 80                   | T                | Negative              | Normal      | CR       | Yes      | Dead |
| 22 | 22          | M   | 6                             | 7                | 196             | 83                   | T                | Negative              | Normal      | CR       | Yes      | Alive |
| 23 | 31          | M   | 5                             | 8                | 80              | 67                   | B                | Negative              | Normal      | CR       | Yes      | Dead |
| 24 | 23          | M   | 17                            | 8                | 53              | 90                   | T                | Negative              | Normal      | CR       | No       | Alive |
| 25 | 22          | M   | 126                           | 6.3              | 65              | 90                   | T                | Negative              | Normal      | CR       | No       | Alive |
| 26 | 35          | M   | 5                             | 6                | 60              | 85                   | B                | Positive              | Abnormal    | CR       | No       | Alive |
| 27 | 35          | M   | 137                           | 8.4              | 46              | 80                   | B                | Negative              | Normal      | CR       | No       | Alive |
| 28 | 34          | F   | 88.2                          | 8.3              | 57              | 90                   | B                | Negative              | Normal      | CR       | No       | Alive |
| 29 | 32          | M   | 10.3                          | 5.4              | 104             | 67                   | B                | Negative              | Normal      | CR       | No       | Alive |
| 30 | 30          | F   | 90                            | 7.2              | 60              | 87                   | B                | Positive              | Abnormal    | CR       | Yes      | Dead |
| 31 | 39          | F   | 1                             | 8.3              | 54              | 23                   | B                | Negative              | Abnormal    | Refractory| N/A     | Dead |
| 32 | 33          | F   | 43                            | 9.6              | 13              | 73                   | B                | Negative              | Normal      | CR       | No       | Alive |
| No | Age (years) | Sex | Total leucocytic count x10^9/L | Hemoglobin (g/dl) | Platelets x10^9/L | Bone marrow blast (%) | Immunophenotyping | Philadelphia chromosome | Cytogenetics | Response | Relapsed | FATE |
|----|-------------|-----|---------------------------------|-------------------|------------------|---------------------|--------------------|-----------------------|--------------|-----------|----------|------|
| 33 | 39          | M   | 16                              | 13.6              | 253              | 22                  | B                  | Negative              | Normal       | CR        | Yes      | Alive |
| 34 | 23          | M   | 138                             | 8.7               | 46               | 90                  | B                  | Negative              | Normal       | CR        | No       | Alive |
| 35 | 19          | M   | 1.7                             | 9.1               | 17               | 41                  | B                  | Negative              | Normal       | CR        | No       | Alive |
| 36 | 30          | F   | 52.9                            | 8.5               | 40               | 90                  | B                  | Negative              | Abnormal     | CR        | No       | Alive |
| 37 | 25          | F   | 11.3                            | 10.7              | 83               | 57                  | B                  | Negative              | Normal       | CR        | No       | Alive |
| 38 | 31          | F   | 38                              | 8.9               | 55               | 37                  | B                  | Positive              | Abnormal     | CR        | Yes      | Dead  |
| 39 | 33          | F   | 44                              | 7.4               | 94               | 34                  | B                  | Positive              | Abnormal     | CR        | No       | Alive |
| 40 | 27          | F   | 63                              | 10                | 121              | 61                  | B                  | Negative              | Normal       | CR        | No       | Alive |
| 41 | 19          | M   | 19                              | 9.3               | 112              | 49                  | B                  | Negative              | Normal       | CR        | No       | Alive |

**CR** complete remission, **F** female, **M** male, **N/A** not applicable
overcome these limitations, further larger, longer, prospective, and multicenter studies are necessary.

Conclusion
DFCP can be considered as an effective ALL protocol for the AYA group of patients with a good complete remission, event-free survival, and overall survival rates. DFCP seemed to be feasible in AYA despite the toxicities experienced which overcame by good supportive care and temporary cessation of some drugs.

Abbreviations
ALL: Acute lymphoblastic leukemia; AYA: Adolescents and young adults; BCR-ABL1: Fusion between BCR gene (Breakpoint Cluster Region) and ABL1 gene (ABelson murine Leukemia virus); BM: Bone marrow; BMT: Bone marrow transplantation; CBC: Complete blood count; CNS: Central nervous system; CR: Complete remission; DFCP: Dana Farber Consortium Protocol; ECOG: Eastern Co-operative Oncology Group; EFS: Event-free survival; MRD: Minimal residual disease; OS: Overall survival; SPSS: Statistical Package for the Social Sciences; t: Translocation

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Authors’ contributions
HEAE and TAE performed concept, design, definition of intellectual content, and data acquisition. TAE performed statistical analysis and data analysis. HEAE, HEE, BBE, SMG, and TAE performed clinical studies, literature search, and manuscript review. HEAE, HEE, and TAE performed manuscript preparation, and manuscript editing. All authors read and approved the final manuscript.

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Availability of data and materials
All data generated or analyzed during this study are included in this published article.

Declarations
Ethics approval and consent to participate
The study was approved by the research ethical committee of faculty of medicine, Tanta University. Institutional Review Board (IRB) for human studies (Approval code is 34071). Our study conforms to provisions of the Declaration of Helsinki. Informed written consent to participate in this study was provided by all participants (or their parent or legal guardian) before the starting of data collection.

Consent for publication
Informed written consent for publication was obtained from all participants (or their parent or legal guardian) before the starting this study.

Competing interests
The authors declare that they have no competing interests.

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References
1. Jabbour EJ, Faderl S, Kantarjian HM. Adult acute lymphoblastic leukemia. Mayo Clin Proc. 2005;80(11):1517–27.
2. Esparzo SD, Sakamoto KM. Topics in pediatric leukemia—acute lymphoblastic leukemia. MedGenMed. 2005;7(1):23.
3. Ma H, Sun H, Sun X. Survival improvement by decade of patients aged 0-14 years with acute lymphoblastic leukemia: a SEER analysis. Sci Rep. 2014;4:4227.
4. Surveillance, Epidemiology, and End Results Program. National Institutes of Health. Available at: https://seer.cancer.gov/statfacts/html/alyl.html. Accessed: 20 Feb 2020.
5. Pulle D, Gondos A, Brenner H. Improvement in survival in younger patients with acute lymphoblastic leukemia from the 1980s to the early 21st century. Blood. 2009;113(7):1408–11.
6. Stock W, Johnson JL, Stone RM, Kolitz JE, Powell BL, Wetzler M, et al. Dose intensification of daunorubicin and cytarabine during treatment of adult acute lymphoblastic leukemia: results of cancer and leukemia group B study 19802 (published correction appears in Cancer. 2014;120(14):2222). Cancer. 2013;119(1):990–8.
7. Bleyer A, Barr R. Cancer in young adults 20 to 39 years of age: overview. Semin Oncol. 2009;36(3):194–206.
8. NCCN Clinical Practice Guidelines in Oncology: acute lymphoblastic leukemia. National Comprehensive Cancer Network. Available at http://www.nccn.org/professionals/physician_gls/pdf/allp.pdf. Version 1.2014 — June 6, 2014. (Last accessed: 10 Oct 2014).
9. Ssegel SE, Stock W, Johnson RH, Advani A, Muffly L, Douer D, et al. Pediatric-inspired treatment regimens for adolescents and young adults with Philadelphia chromosome-negative acute lymphoblastic leukemia: a review. JAMA Oncol. 2018;4(5):725–34.
10. Chiaretti S, Zini G, Basan R. Diagnosis and subclassification of acute lymphoblastic leukemia. MedGenMed. 2005;7(1):23.
11. Barry E, DeAngelo DJ, Neuberg D, Stevenson K, Koh ML, Asselin BL, et al. Favorable outcome for adolescents with acute lymphoblastic leukemia treated on Dana-Farber Cancer Institute Acute Lymphoblastic Leukemia Consortium Protocols. J Clin Oncol. 2007;25(7):813–9.
12. de Labarthe A, Rousselet P, Huguet-Rigal F, Delabesse E, Witz F, Maury S, et al. Group for research on adult acute lymphoblastic leukemia (GRAALL). Imatinib combined with induction or consolidation chemotherapy in patients with de novo Philadelphia chromosome-positive acute lymphoblastic leukemia: results of the GRAALL-2003 study. Blood. 2007;109(4):1408–13.
13. Stock W. Adolescents and young adults with acute lymphoblastic leukemia. Hematol Am Soc Hematol Educ Program. 2010;2010:21–9.
14. Bydding ME, Jabbour EJ, O’Brien SM, Kantarjian HM. Acute lymphoblastic leukemia in adolescents and young adults. Cancer. 2017;123(13):2398–403.
15. Harrison CJ. Cytogenetics of paediatric and adolescent acute lymphoblastic leukaemia. Br J Haematol. 2009;144(2):147–56.
16. Arber DA, Orazi A, Hasserjian R, Thiele J, Borowitz MJ, Le Beau MM, et al. The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia. Blood. 2016;127(23):405–13.
17. Ribera JM, Ribera J, Genescà E. Treatment of adolescent and young adults with acute lymphoblastic leukemia. Mediterr J Hematol Infect Dis. 2014;6(1):e2014073.
18. Lukenbill J, Advani AS. The treatment of adolescents and young adults with acute lymphoblastic leukemia. MedGenMed. 2015;29(3):526–34.
19. Elashtokhy AS, Elsayed HG, Sherisher MA, Elbjeirami WM, Alseldin N. The Dana Farber Consortium protocol for the treatment of adolescents...
and young adults with acute lymphoblastic leukemia: a single institution experience in Saudi Arabia. Clin Lymphoma, Myeloma Leuk. 2017;17(5):320–5.

25. Gramatges MM, Rabin KR. The adolescent and young adult with cancer: state of the art–acute leukemias. Curr Oncol Rep. 2013;15(4):317–24.

26. Silverman LB, Gelber RD, Dalton VK, Asselin BL, Barr RD, Clavell LA, et al. Improved outcome for children with acute lymphoblastic leukemia: results of Dana-Farber Consortium Protocol 91-01. Blood. 2001;97(5):1211–8.

27. Knoderer HM, Robarge J, Folkhart DA. Predicting asparaginase associated pancreatitis. Pediatr Blood Cancer. 2007;49(5):634–9.

28. Athale UH, Siciliano SA, Crowther M, Barr RD, Chan AK. Thromboembolism in children with acute lymphoblastic leukemia treated on Dana-Farber Cancer Institute protocols: effect of age and risk stratification of disease. Br J Haematol. 2005;129(6):803–10.

29. Strauss AJ, Su JT, Dalton VM, Gelber RD, Sallan SE, Silverman LB. Bony morbidity in children treated for acute lymphoblastic leukemia. J Clin Oncol. 2001;19(12):3066–72.

30. Sonabend RY, McKay SV, Okcu MF, Yan J, Haymond MW, Margolin JF. Hyperglycemia during induction therapy is associated with increased infectious complications in childhood acute lymphocytic leukemia. Pediatr Blood Cancer. 2008;51(3):387–92.

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