Physiological Measurement

On the suitability of laser-Doppler flowmetry for capturing microvascular blood flow dynamics from darkly pigmented skin

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Abstract

Objective: To assess the performance of laser Doppler flowmetry (LDF) in measuring blood perfusion from darkly-pigmented skin, i.e. skin with high melanin concentration. LDF provides for the noninvasive monitoring of microvascular blood flow dynamics. It has been used extensively on light-skinned subjects, i.e. on skin with low melanin concentration, in both the healthy and pathological states. Because the optical properties of human skin might affect the reliability of optically-based diagnostic equipment, the effectiveness of LDF needs to be checked and evaluated on dark skin, too, if this method is to be useful in global healthcare. Approach: Thirteen dark-skinned subjects and ten light-skinned subjects were included in the study. Microvascular blood flow dynamics was measured on both the right and left ankles using LDF with a laser diode of wavelength 780 nm. The characteristics of time-varying blood flow oscillations were investigated by wavelet analysis, nonlinear mode decomposition and wavelet phase coherence. An electrocardiogram (ECG), skin temperature, and respiratory effort were measured simultaneously with the LDF for each subject. Main results: No significant differences were observed between the groups in the mean blood perfusion \( (p > 0.1) \), or wavelet power \( (p > 0.6) \). The instantaneous heart rate (IHR), extracted from the LDF at each of the recording sites, and from the ECG, did not differ significantly between the groups \( (p > 0.8) \). Nor did the wavelet power of the IHR differ \( (p > 0.8) \) between the groups. The only significant difference found between the groups lay in left/right ankle blood flow coherence near the cardiac frequency, attributable to known ethnic physiological differences. Significance: These results indicate that high melanin concentrations in skin exert no significant influence on the ability of LDF to monitor microvascular blood flow dynamics when using a laser diode of wavelength 780 nm. Hence LDF can help in the diagnosis and exploration of the pathogenesis of diseases such as diabetes, hypertension, or malaria in darkly pigmented patients across sub-Saharan Africa.

1. Introduction

Non-invasive optical techniques in biomedicine have made notable advances in recent decades (Peng et al 2008, Vo-Dinh 2014, Tuchin 2016). One example is laser Doppler flowmetry (LDF). It has been shown to give results comparable to those from other methods of evaluating skin microvascular blood flow, and it possesses the particular advantage of continuous detection of microvascular blood flow in a volume of tissue, as opposed to axial flow in a single vessel (Nitzan et al 1988).

LDF provides a simple and non-invasive approach for assessing the dynamical properties of the skin microcirculation, and it can be applied in both the healthy and pathological states (Stefanovska et al 1999). In combination with appropriate time-series analysis, it can yield valuable insights into the dynamics of microvascular blood flow. Its working principle depends on the Doppler shift in the frequency of light reflected from moving red blood corpuscles (erythrocytes). So it relies on the passage of incident light through the skin, twice. Some knowledge of the skin’s optical properties is therefore required.

As illustrated in figure 1, human skin (Kanitakis 2002) is made up of several layers, of which the melanin chromophores responsible for skin pigmentation reside in the epidermal layer (Costin and Hearing 2007).
Epidermal melanin consists of eumelanin (black-brown pigment) and pheomelanin (yellow-reddish pigment), whose main responsibility is skin photo-protection (Anderson and Parrish 1981, Costin and Hearing 2007).

LDF monitoring of blood flow is known to work efficiently in the case of light skin (Nilsson et al 1980, Kvernmo et al 1999, Stefanovska et al 1999, Söderström et al 2003), whose optical properties enable light to penetrate more easily, but the question arises as to what happens in the case of darkly pigmented skin (Fredriksson et al 2009, Karsten and Smit 2012)? The interaction of light with skin mostly happens just below the surface, i.e. in the stratum corneum. As shown in figure 1, some of the coherent laser light shone on the skin surface gets scattered from the stratum corneum, referred to as surface scattering. Some of the light propagates to deeper skin layers such as the dermis and epidermis. Absorption and scattering by the melanin and erythrocytes occur in the epidermis and dermis respectively. Unlike the epidermis and dermis, the stratum corneum layer is usually colourless and absorbs only a small portion of incident light.

Melanin absorbs light within the visible and near-infra-red (near-IR) parts of the optical spectrum. The absorption decreases with increasing wavelength, so that near-IR in the wavelength range 780–800 nm is best suited to non-invasive diagnosis and treatment (Peng et al 2008). The fact that dark skins contain about twice the concentration of epidermal melanin, compared to lighter skins, inevitably means that less laser light reaches the deeper segments of dark skin (Peng et al 2008).

Extensive investigations of the absorption and reflection of light in from human skin began in 1911 with the spectrographic studies of Hasselbalch (1911). More recently, considerable effort has been devoted to quantifying the absorption spectrum of human skin, particularly darkly pigmented skins (Hardy et al 1956, Kollias and Baqer 1985, 1986, Kollias 1994, Lister et al 2012). There has been significant disagreement in the results, perhaps due to challenges in conducting the experimental measurements. For example, the studies of both Pauli and Ivancevic (1927) and Cartwright (1930) reported that a substantial proportion of IR radiation penetrates deeply into the body through the skin, whereas Blaine (1932) and Hardy and Muschenheim (1934) reported a negligible proportion.

In an attempt to determine the effect of melanin on spectroscopic signal formation, Kollias and Baqer (1985) studied the absorption characteristics of melanin in the wavelength range 620–720 nm and observed that remittance spectra measured from vitiligo-involved skin and normal skin did not differ, from which they concluded that both skins have similar absorption. In a comparable study by Kollias and Baqer (1986), the remittance spectroscopy parameters measured from normal skin and amelanotic skin in the range 620–720 nm were found to be strongly correlated.

More recently, a model of fluorescence spectra from biological tissue based on the Monte Carlo approach has predicted the effect of melanin concentration on a spectroscopy signal (Dremin and Dunaev 2016). The use of near-IR diode lasers of relatively long wavelength (670, 780, and 810–850 nm) was shown to improve optical penetration (Murray et al 2004).
Melanin and hemoglobin are known to dominate the absorption of light in skin within the visible range. About 4%–7% of visible light is reflected from the skin surface, regardless of skin pigmentation and wavelength (Anderson and Parrish 1981, Takiwaki 1998) (figure 1). Unlike hemoglobin which mainly absorbs light within the dermis, melanin present within the epidermis layer exhibits an absorption spectrum extending from the ultraviolet region to the IR (Lister et al 2012). Several studies (Kollias and Baquer 1985, and Kollias 1994) based on diffuse reflectance spectroscopy have investigated the absorption spectrum of melanin in vivo, with most of the data being analysed empirically. These studies have yielded a qualitative understanding of melanin absorption characteristics. Nonetheless, data analysis and the realisation (Karsten and Smit 2012, Dremin and Dunæv 2016) that a high epidermal melanin concentration attenuates laser light transmitted to the deeper skin layers, calls for a more extensive investigation.

Note that, because LDF computes the erythrocyte speed from the spectral broadening (see below) of the incident light, which is independent of the light’s absolute intensity, the method might be expected to function effectively regardless of the degree of attenuation along the signal path. However, this conclusion can only hold true if two conditions are fulfilled. First, the signal/noise ratio of the light returned to the detector must be sufficient for the spectral broadening to be determined reliably by the measurement algorithm. Secondly, the incidence of frequency-dependent scattering process along the optical return path that would modify the shape of the spectrum must be insignificant. In practice, therefore, the only convincing way forward is empirical: to test LDF on dark-skinned subjects and look for significant differences in the results compared to those obtained from comparable light-skinned subjects.

The present study therefore tests the hypothesis that, in the case of dark skin, the attenuation of the incident laser light reaching (and scattering back from) the dermis is not sufficient to prevent LDF functioning effectively. A relatively long wavelength (780 nm) was chosen with the intention of minimising the attenuation. The paper describes a detailed experimental study comparing the LDF flux measured for both darkly and lightly pigmented skin. The time-varying oscillations in the microvascular blood flow dynamics were checked and compared for the two pigmentation. They are known (Kvernmo et al 1999, Stefanovska et al 1999, Soderstrom et al 2003, Shigai et al 2010, Aalkjær et al 2011) to include components ranging from the cardiac frequency at ~1 Hz in healthy humans down to endothelium-related oscillations with frequencies of ~0.01 Hz. Non-linear time series analysis (Stefanovska et al 1999, Clemson and Stefanovska 2014, Iatsenko et al 2015b, Clemson et al 2016) was used to identify the oscillatory components in the signals. Specifically, the frequency interval from 0.0095 to 2 Hz was examined and categorized into six intervals: interval I (0.6–2 Hz) related to cardiac activity; interval II (0.145–0.6 Hz) related to respiratory activity; interval III (0.052–0.145 Hz) related to microvascular smooth muscle cell activity; interval IV (0.021–0.052 Hz) related to microvascular innervation; and intervals V & VI (0.0095–0.021 Hz and 0.005–0.0095 Hz, respectively) related to endothelial activity, both nitric oxide (NO) dependent and independent.

2. Methodology

2.1. Measurement of the skin blood perfusion

2.1.1. Laser Doppler flowmetry

LDF provides a continuous measurement of microcirculation in the skin, thus reflecting perfusion in capillaries, arterioles, venules and dermal vascular plexus. The LDF (moorLAB, Moor Instruments Ltd, UK) used in the present study transmits a near-IR laser light from temperature stabilized laser diodes operating at a wavelength of 780 nm and with a maximum power of 2.5 mW into the skin through an MP1-V2 probe (Moor Instruments Ltd, UK), which has two optical fibres. A time constant of 0.1 s was selected and the LDF processor bandwidth was between 18 Hz and 22.5 KHz. A flexible probe holder (PH1-V2, Moor Instruments Ltd, UK) was attached to the skin using double-sided adhesive discs. One fibre delivers light to the site under observation, while the backscattered (reflected) light is collected by the other fibre as shown in figure 1. According to the Doppler principle, the light reflected from moving red blood cells is shifted in frequency by an amount related to the blood flow in the illuminated volume of tissue—the frequency shift is proportional to red cell speed while the frequency of light reflected from stationary cells and tissue remains unchanged (Nilsson et al 1980). The difference between incident light and the Doppler-shifted back-scattered light gives the LDF signal, known as the blood perfusion signal. The LDF output is semi-quantitative and is expressed in perfusion units (PU) of output voltage (typically 1 PU = 10 mV) (Nilsson et al 1980). Skin perfusion on both left and right ankles was measured at a sampling frequency of 40 Hz. Throughout this paper we will therefore refer either to Doppler flow or blood perfusion.

2.1.2. Experimental protocol

The protocol of this study was approved by the Faculty of Science and Technology Research Ethics Committee, Lancaster University UK. Thirteen healthy dark-skinned subjects, born in sub-Saharan Africa, without known ancestors of non-African origin, with a high melanin concentration in their skins, and ten light-skinned Caucasian
subjects of European origin with low melanin concentration, between the ages of 18–27 years, were recruited. All subjects were male. Their anthropometric data are given in table 1. Written informed consent was obtained from all participants. Volunteers were asked to abstain from food, coffee, and alcohol for three hours prior to the experimental measurements. LDF was used for non-invasive and simultaneous measurement of the skin blood flow, on the outer sides of the left (LA) and right (RA) ankles (lateral malleolus). An electrocardiogram (ECG) was used to record the electrical activity of the heart with a sampling frequency of 1000 Hz. ECG was measured using a bipolar precordial lead. The electrodes were attached on both shoulders and the lowest left rib, as this maximizes the sharpness of the R-peak. Using an elastic belt fastened across the chest and fitted with a Biopac TSD201 Respiratory Effort Transducer (Biopac Systems Inc., CA, USA), the respiration was also measured. Skin temperature was monitored using two high sensitivity, low heat capacity thermistors—YSI 709B Thermilinear sensors (YSI Inc, Yellow Springs, OH, USA) of 8.5 mm diameter, which were taped on the skin. The thermistors were positioned outside the left ankles, over the lateral malleolus, close to the LDF probes.

The time series were recorded simultaneously using a signal conditioning system (Cardiosignals, Institute Jožef Stefan, Slovenia) and recordings lasted for 30 min with the volunteers lying relaxed. The recorded data are publicly available (see acknowledgements).

Blood pressure was measured prior to the initiation of signal acquisition. A Digital Automatic Blood Pressure Monitor (Omron, M10-IT) was used, wrapping a cuff on the subjects upper right arm while the subject was seated. The subject then moved to a supine position on a comfortable bed, where the necessary sensors were installed. In this way, subjects were in a supine position for 15–20 min of acclimatisation, before the recordings started. The bed and the equipment were housed within a well-ventilated Faraday cage, with a controlled ambient temperature of 20 °C–21 °C and constant low illumination. The equipment was either battery supplied or plugged in to the electrical supply via a mains filter.

2.1.3. Statistical analysis
Application of the Lilliefors test for normality (Lilliefors 1967) showed that there were no consistent normal distributions of data among the groups being compared. Non-parametric statistical tests were therefore used, meaning that no assumptions were made about any underlying distributions, thus allowing robust conclusions to be drawn. The Kruskal–Wallis ANOVA test (Kruskal and Wallis 1952) was used when all IHRs (derived from the LA and RA blood flows and from the ECG) were compared. The Kruskal–Wallis test checks whether two or more independent sets of data originate from the same distribution, and it does not assume normal distributions. Where significance is found, pairs of groups are tested either by the Wilcoxon signed rank test for paired data, or by the Wilcoxon rank sum test for unpaired data (Wilcoxon 1945). The latter was used to test for possibly significant differences between blood flow measured from dark-skinned and light-skinned subjects, respectively, as the corresponding time-series do not match. The Wilcoxon rank sum test is used to determine whether two unmatched samples come from similar distributions, whilst the sign rank test requires that the samples are matched. In all cases, $p < 0.05$ was considered as being statistically significant.

3. Time series analysis
Recorded signals can be analysed to extract statistical properties of the data, or to investigate potential oscillatory characteristics, by quantifying the amplitude and power of oscillations and their phase characteristics. In this study, we particularly focus on characterization of time-varying oscillatory properties, resolving the dynamics with an optimal time localization and frequency resolution.

| Table 1. Anthropometric data of subjects measured, median values, ranges [25th and 75th percentiles] and significance. |
|------------------------------------------------------------------------------------------------------------------|
|                                                                                                                  |
| Dark-skinned                                                                                                    |
| Light-skinned                                                                                                    |
| $p$                                                                                                              |
| Subjects ($n = 13$)                                                                                               |
| Subjects ($n = 13$)                                                                                               |
| Age (years)                                                                                                      |
| 21.0 [20.0 24.0]                                                                                                  |
| 22.0 [19.0 25.0]                                                                                                  |
| 0.66                                                               |
| Body mass index (kg/m$^2$)                                                                                       |
| 22.60 [20.05 23.98]                                                                                              |
| 23.15 [21.60 24.70]                                                                                              |
| 0.34                                                               |
| Skin temperature (°C)                                                                                            |
| 29.89 [27.98 31.10]                                                                                              |
| 30.13 [28.96 30.45]                                                                                              |
| 0.90                                                               |
| Instantaneous heart rate (Hz)                                                                                     |
| 0.99 [0.91 1.05]                                                                                                  |
| 0.99 [0.83 1.17]                                                                                                  |
| 1                                                                  |
| Instantaneous respiratory rate (Hz)                                                                               |
| 0.28 [0.27 0.29]                                                                                                  |
| 0.26 [0.24 0.27]                                                                                                  |
| 0.06                                                               |
| Systolic BP (mm Hg)                                                                                              |
| 115.0 [111.8 126.0]                                                                                              |
| 118.5 [109.0 126.0]                                                                                              |
| 0.85                                                               |
| Diastolic BP (mm Hg)                                                                                             |
| 72.0 [68.0 75.3]                                                                                                 |
| 76.5 [67.0 77.0]                                                                                                 |
| 0.64                                                               |
3.1. Wavelet analysis

The effective visualisation of the oscillations present in LDF generated signals greatly depends on the method used for their analysis.

Traditionally, representations of time series in the frequency domain are obtained with the Fourier transform, which constitutes a periodic function in terms of sines and cosines. This makes it suitable for analysing time series whose components are strictly periodic in nature, but it is unsuitable for LDF signals whose components are inherently non-periodic. The limitations of the Fourier transform can partly be addressed by use of the short-time Fourier transform through windowing, i.e. by dividing the time series into shorter time-windows within which there is not much time variation so that the fast Fourier transform (FFT) can usefully be calculated; in practice, this is usually done by sliding a window across the whole signal.

In the short-time Fourier transform, however, the spectral resolution depends strongly on the length of the window, with short windows leading to poor frequency resolution but good time localisation, and vice versa. This method may therefore fail when dealing with non-stationary physiological signals with varying frequencies, such as the LDF time series considered in this study. Its limitations can be overcome by use of wavelet analysis (Stefanovska et al 1999) which, by using an adaptive window length that simultaneously analyses time series at each moment in time, provides both optimal frequency resolution and good time localisation (Iatsenko et al 2013). These features are particularly important for the analysis of slowly-changing oscillatory dynamics over a broad frequency interval, of the kind giving rise to an LDF signal.

Wavelet analysis is a scale-independent method comprising an adaptive window length allowing low frequencies to be analysed using longer wavelets, and higher frequencies with shorter wavelets. The continuous wavelet transform \( W_s(t, f) \) of a signal \( f(t) \) is defined as

\[
W_s(t, f) = |s|^{-1/2} \int_{-\infty}^{\infty} \psi \left( \frac{u-f}{s} \right) f(u) du \tag{1}
\]

where \( s \) is a scaling factor, \( t \) is the temporal position on the signal, and the wavelet function is built by scaling and translating a chosen mother wavelet \( \psi \). In this study the complex Morlet wavelet

\[
\psi(u) = \frac{1}{\sqrt{\pi}} \left( e^{-i\omega_0 u} - e^{-\omega_0^2 u^2/2} \right) e^{-u^2/2} \tag{2}
\]

was chosen because it gives very good joint time-frequency resolution (Stefanovska et al 1999).

3.2. Extracting the instantaneous heart frequency

The instantaneous heart rate (IHR) was extracted using both time-frequency and time domain analysis techniques. The methods used to estimate heart frequency include nonlinear mode decomposition (NMD) (Iatsenko et al 2015), a technique that decomposes a signal into set of components, or modes. Using NMD, the instantaneous frequency of the heart beat was extracted from the wavelet transform of the ECG, thus yielding the IHR. Similarly, the IHR was also derived from the LDF signal using the same technique. Note that in the literature (Malik et al 1996, Iatsenko et al 2013) IHR is mostly referred to as HRV or, occasionally, as IHF.

3.3. Wavelet phase coherence

The coherence \( W_{PC_{s1,s2}}(f) \) between the two signals \( s_{1,2}(t) \) is determined through their WTs \( W_{s_1,s_2}(t, f) \) as

\[
W_{PC_{s1,s2}}(f) = \left| \frac{1}{T} \int_0^T e^{i\arg[W_{s_1}(t, f)W_{s_2}^*(t, f)]]} dt \right| \tag{3}
\]

and it reflects the extent to which the phases (and thus the underlying activities) of these signals at frequency \( f \) are correlated (Bandrivskyy et al 2004, Sheppard et al 2012). Unlike the usual coherence measures, wavelet phase coherence does not take into account the amplitude dynamics of the signals. This is appropriate because (i) the amplitudes of most physiological signals are subject to artefacts and noise, and (ii) the relationships between the amplitudes of common physiological oscillations in different signals can be complicated and nonlinear, but in all cases the relationship between their phases remains the same (up to a constant phase shift).

3.4. Effective (or significant) coherence

The wavelet phase coherence between the oscillations in the two blood flow signals is computed by evaluating the difference between the wavelet transform phases of the signals at each frequency, and at each moment in time, as given by equation (3). The oscillations are considered to be coherent at any given frequency if their phase shifts remain unchanged (with a coherence value ranging between 0 and 1); otherwise they are said to be incoherent. Coherence does not provide information about synchronization between oscillations, as this would require them to be coupled, which is not necessary for coherence to exist. However, information on possible synchronization between oscillations, particularly at the smaller ratios, e.g. 1:1 synchronization, can be obtained...
by estimating the wavelet phase coherence. Note, however, that the coherence computed in the first instance does not necessarily reflect a genuine phase relationship and requires careful evaluation. The problem arises because some of the coherence values obtained can be less than zero (although formally coherence values range between 0 and 1). These negative coherence values are then subtracted. Following this procedure, the very low frequency oscillations may appear to have a coherence close to 1, because of bias resulting from the use of recordings that are too short to encompass the content at low frequencies.

To minimise random effects giving rise to apparent (but spurious) coherence, whether at low or high frequency, we checked/tested the significance of the computed coherence using the method of surrogates (Schreiber and Schmitz 2000, Lancaster et al. 2018)—by setting as a null hypothesis that, for all frequencies, the phases in the signals are independent. We used iterative amplitude-adjusted Fourier transform (IAAFT) surrogates to estimate the significance level of the apparent coherence, thereby removing the bias associated with the power spectrum of the more commonly used amplitude-adjusted Fourier transform (AAFT) surrogates. First, the IAAFT surrogates are constructed by randomizing all the properties of the signals in question, whilst keeping only the phases unshuffled. Subsequently, this is accomplished in an iterative fashion, simply by using the appropriate value and re-scaling the distribution to substitute Fourier amplitudes, which allows us to obtain resemblance between the distributions and power spectra of the surrogates and the original signals. At each frequency we took the coherence threshold to be 95% of the highest value of 100 random realisations of IAAFT surrogates.

Finally, the effective/significant coherence was estimated by subtracting the 95th percentile of the 100 surrogate values, thus giving the extent to which the phases of the two signals at each frequency are correlated.

4. Results

The median and interquartile ranges of the instantaneous respiratory rate and IHR for both dark-skinned and light-skinned groups of subjects are presented in Table 2. As in the case of the anthropometric data (Table 1), there are no statistically significant differences between the groups.

Typical recordings of LDF blood flow time series simultaneously recorded from the right and left ankles of volunteers in both groups, together with their respective time-frequency representations, are presented in Figure 2. No differences in skin perfusion or fluctuations between the groups were evident. Table 2 presents the median and interquartile ranges of the blood flow of dark and light-skinned groups. Although the values for dark-skinned group are slightly lower, no statistically significant differences were found in mean blood perfusion between the two groups ($p > 0.1$).

The IHR values calculated from the LDF time series recordings measured on the ankles did not differ between groups, as shown in Table 2. Figures 3(a) and (c) illustrate how the IHR was extracted from the ECG and (b) and (d) from LDF blood flow signals; and the intra-group comparison between the IHRs (figures 3(e) and (f)). Comparisons made between mean values of the IHR signals derived from both the ECG and LDF data (measured from both LA and RA) (Table 2) revealed no statistically significant differences ($p > 0.7$).

Figure 4 summarises the quantitative analyses of the oscillations in LDF blood flow and in the IHRs derived from both ECG and LDF. No statistically significant differences in time-averaged wavelet power across the frequency intervals were observed in the LA and RA (figures 4(a) and (b)) blood flows between the dark- and light-skinned groups ($p = 0.95$ and $p = 0.62$ respectively). The 25th and 75th percentiles (indicated in dark green for dark skinned and light green for light skinned subjects) of both LA and RA LDF blood flow spectrum between groups are presented in figures 4(a)(i) and (b)(ii). There is no obvious difference in inter-subject variations between the two groups.

Intra-group comparison of the time-averaged power of the IHR derived from ECG, LA and RA LDF blood flows revealed no significant differences between light-skinned (figure 4(c)) and dark-skinned (figure 4(d)) groups: $p = 0.99$ and $p = 0.21$ respectively as obtained by the Kruskal–Wallis test. Figure 4(e) shows no significant difference in the inter-group comparison of the time averaged power for IHR derived from (i) ECG, (ii) LA LDF and (iii) RA LDF, $p = 0.64$, $p = 0.37$, and $p = 0.20$, respectively.

### Table 2. LDF skin perfusion and IHR (derived from LDF) of measured subjects, median values and ranges [25th and 75th percentiles].

|                         | Dark-skinned | Light-skinned | $p$   |
|-------------------------|--------------|---------------|-------|
| Blood flow (RA) (AU)    | 6.12 [5.00 6.89] | 9.30 [5.39 12.58] | 0.11  |
| Blood flow (LA) (AU)    | 7.54 [5.50 11.11] | 9.06 [7.01 11.75] | 0.37  |
| IHR (from ECG) (Hz)     | 0.99 [0.91 1.05] | 0.99 [0.83 1.18] | 0.98  |
| IHR (from RA) (Hz)      | 0.99 [0.93 1.11] | 0.99 [0.84 1.17] | 0.78  |
| IHR (from LA) (Hz)      | 1.00 [0.95 1.10] | 1.00 [0.83 1.18] | 0.73  |
Wavelet phase coherence between LA and RA blood flows from each group are shown in figure 4(f). Compared to the dark-skinned cohort, the light-skinned group exhibits a significantly higher coherence in the 0.6–2 Hz frequency interval, corresponding to cardiac activity. The coherence was significantly lower in the dark-skinned group also near 0.1 Hz.

5. Discussion

Analysis of the spectrally broadened signal due to backscattering of the incident 780 nm coherent laser light from darkly- and lightly-pigmented skins has provided a measure of the flux, which is proportional to the product of the average speed of the moving erythrocytes and their concentration. It has enabled the hypothesis underlying this study—that dark skin colour (melanin concentration) does not attenuate the light sufficiently to vitiate LDF as a method for measuring blood flow—to be tested.
Our study sheds additional light on the long-running debate about the significance for LDF of the optical difference between Caucasian and non-Caucasian skin, a question that has been under discussion ever since the early days of the technique (Leahy et al 1999). Dark skin colour significantly attenuates the incident laser light that reaches the deeper skin tissue (Goldman et al 1963). Fredriksson et al (2009) used Monte Carlo simulations of light propagation in tissue, for wavelengths between 543 and 780 nm, to show that skin pigmentation is expected to have a negligible effect on the measurement depth. The wavelength of the light certainly influences the optical penetration, irrespectively of the skin colour. Zhao and Fairchild (1998) showed that, in the wavelength interval 532–1064 nm, laser light of longer wavelength penetrates more deeply into skin. Also using Monte Carlo simulations (Ash et al 2017) it was recently confirmed that an increase in the wavelength of incident light increases its penetration, consistent with the fact that melanin absorption decreases monotonically with increasing wavelength (Keilhauer and Delori 2006). This arises because the scattering and absorption coefficients of melanin and other chromophores inside the skin are wavelength-dependent. In this study we tested one of the commonest wavelengths used for LDF measurements.

Our results show that LDF can provide reliable information about the dynamical properties of blood flow in darkly pigmented skin: a significant fraction of the laser light is able to penetrate, interact with the moving erythrocytes, and be back-scattered to reach the detector. By quantifying the power of oscillations and their phase characteristics, our findings show that the oscillatory characteristics of LDF time-series recordings from darkly pigmented skin did not differ significantly from those measured from light skins. This indicates that the intensity of 780 nm light penetrating to 1.15 mm below the epidermis is sufficient for LDF to gather information about the

Figure 4. Group median time-averaged spectral power calculated from the wavelet transforms of LDF signals recorded for 30 min in dark-skinned (green) and light-skinned (red) groups for (a) the left ankle (LA) and (b) the right ankle (RA). In neither case was any statistically significant difference seen at any frequency. The insets ((a)-(i)) and ((b)-(ii)) are provided to give an idea of inter-subject variability, which was similar in both groups. They show the 25th and 75th percentiles of the individual spectra from dark-skinned (dark green) and light-skinned (light green) groups. (c) and (d) show IHR time-averaged wavelet power for dark-skinned (green) and light-skinned groups (red). The box-plots represent the time-averaged wavelet power for IHR derived from ECG (first), LA LDF (second) and RA LDF (third box). The Kruskal–Wallis test revealed no significant differences among the three IHR powers (p = 0.21 for the light-skinned group and p = 0.99 for the dark-skinned group). (e) Comparison between groups: (i) IHR derived from ECG, (ii) IHR derived from LA LDF and (iii) IHR derived from RA LDF. No significant differences are obtained for any of the three comparisons. (f) Median wavelet phase coherence between left and right ankle blood flow for dark-skinned (green) and light-skinned (red) groups. Light-gray indicates frequencies at which there are significant differences (p < 0.05) as determined by the Wilcoxon rank-sum test.
blood flow dynamics (Braverman 1997, 2000). Our findings differ from some of those reported earlier (Goldman et al. 1963, Karsten and Smit 2012, Dremin and Dunaev 2016), because the wavelength of the laser diode in the present study differed, although the exact depth to which the LDF method can be effective remains unknown (Braverman et al 1992). One expects differences in the amplitude of the flux measured from different skins, due to possible difference in the density of erythrocytes between individuals, given that the flux (the Doppler-shifted signal) depends on the erythrocyte concentration. In this study, however, the average values of Doppler perfusion did not differ to a statistically significant extent between the darkly- and lightly-pigmented skins.

Wavelet analyses of LDF blood flow measured from dark and light skins produced closely similar results. The cardiac activity could clearly be observed in each case. Oscillations throughout the full 0.0095–2 Hz frequency interval investigated were the same, suggesting that the fluctuations in laser Doppler perfusion are being properly captured from dark-skin. This is contrary to earlier reports that reproducible estimation of laser Doppler perfusion in darkly pigmented skin is difficult (Bonner and Nossal 1990).

Two methodological issues deserve comment. First, the exact melanin concentrations were not measured directly. However, we have considered the two extreme cases of (a) darkly-skinned subjects that were born and raised in sub-Saharan Africa, without known ancestors of non-African origin, and (b) white-skinned Caucasians of European origin. It is well known that of all skins, the sub-Saharan black Africans’ skin possesses the highest concentration of melanin. It is packaged into larger singly-dispersed melanosomes of \( \sim 1 \mu m \), whereas in lighter skin types the melanosomes are known to be smaller with diameters of \( \sim 0.5\mu m \) (Alaluf et al 2002). Secondly, the group sizes are relatively small. We made every possible effort to make the groups homogenous, including healthy, non-obese males in a very narrow age-range (see table 1). Additionally, we would comment that a large number of studies with similar numbers of participants have been conducted with clinically and physiologically relevant results, e.g. Kvernmo et al (1999), Stefanovska et al (1999), Söderström et al (2003), Sheppard et al (2011) and Ticcinelli et al (2017).

The significantly higher coherence around the cardiac frequency interval in the light-skinned group, compared to the dark-skinned one, may result from the known ethnic disparity in cardiac autonomic modulation between white-skinned and black-skinned people (Urbina et al 1998, Guzzetti et al 2000, Esco et al 2010). It could also be associated with the ethnic differences in left ventricular wall thickness (Hinderliter et al 1996). Also, it is known that nitric oxide (NO) and endothelium-derived hyperpolarizing factor are of crucial importance in resting blood flow dynamics and that black-skinned subjects of African origin exhibit reduced NO compared to Caucasians (Cardillo et al 1998, Kalinowski et al 2004, Ozkor et al 2014, Kim et al 2018). So the significant difference in blood flow coherence observed here may reasonably be attributed to the physiological differences underlying the ethnic disparities (Berardesca et al 1991, Hill et al 2015) between the two groups of subjects, and we will provide a fuller analysis and discussion of these differences elsewhere.

The IHR values from the laser Doppler signals exhibited no significant differences between dark-skinned and light-skinned groups. Similarly, we obtained no significant difference in the spectral power over the 0.005–0.6 Hz frequency interval. The IHRs derived from ECG were considered as the reference signal in these comparisons. Although the curve of IHR does not match perfectly between groups, the observations still prove that the fluctuations in Doppler perfusion measured in high melanin skin are physiologically meaningful. The fact that the observed microcirculatory dynamics is similar for both groups shows that frequency-dependent scattering processes along the optical return path are insignificant.

6. Conclusion

Our investigation has shown that, with illumination derived from a laser diode of wavelength 780 nm, LDF provides an effective method of studying blood flow dynamics, even in darkly pigmented skin. We found no evidence that the greater light attenuation in the latter case has an adverse effect on measurements, and we were able to obtain the same information about the microcirculatory dynamics regardless of skin pigmentation.

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