Bone and joint infection

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Bone and joint infections include septic arthritis, prosthetic joint infections, osteomyelitis, spinal infections (discitis, vertebral osteomyelitis and epidural abscess) and diabetic foot osteomyelitis. All of these may present through the acute medical take. This article discusses the pathogenesis of infection and highlights the importance of taking a careful history and fully examining the patient. It also emphasises the importance of early surgical intervention in many cases. Consideration of alternative diagnoses, appropriate imaging and high-quality microbiological sampling is important to allow appropriate and targeted antimicrobial therapy. This article makes some suggestions as to empiric antibiotic choice; however, therapy should be guided by local antimicrobial policies and infection specialists. Involvement of a multidisciplinary team is essential for optimal outcomes.

Introduction

Bone and joint infections cause serious morbidity and pose significant management challenges. They may cause acute sepsis with bone and joint destruction, chronic pain, discharging wounds and permanent disability. With expanding populations and increasing age, bone and joint infections, especially those involving devices, will have a growing impact on healthcare resources. For effective management, well-coordinated multidisciplinary working is important.

General considerations

Pathogens can gain access into bone and joints through the bloodstream (haematogenous route) or via direct inoculation from a contiguous focus of infection. Acute haematogenous infections are most common in children and the elderly. The presence of foreign material such as implanted devices or dead bone significantly reduces the number of organisms required to cause infection. Foreign or non-viable material also allows normal skin commensals, such as coagulase-negative Staphylococcus spp, to become significant pathogens. Micro-organisms adhere to the inert surface where they are relatively protected from the blood supply, immune processes and antibiotics. Organisms, such as Staphylococcus spp can produce extracellular polymeric substance (EPS). Micro-organisms embedded in EPS create a biofilm on the inert surfaces. By communicating with each other they are able to up and downregulate gene expression enabling regulation of growth and adaptation to the environment. Biofilm formation is an important mechanism for bacterial survival in chronic bone and joint infection.

Key points

In suspected bone and joint infections quality microbiological sampling is important. In those with features of sepsis or acute skin and soft tissue infection (SSTI), blood cultures and, where possible, aspirates should be taken immediately followed by prompt empiric antibiotic therapy. In other cases antibiotic therapy should wait until intraoperative samples are taken. In all acute bone and joint infections, orthopaedic surgeons, infection specialists and radiologists should be involved early.

Acute septic arthritis should be managed with diagnostic aspiration followed by prompt arthroscopy or arthrotomy and washout in conjunction with antibiotics. In some cases, where surgery may not be possible, serial closed joint aspirations might be an alternative option.

Acute haematogenous osteomyelitis needs prompt surgical drainage if there is a purulent collection. Other cases can be managed by antibiotics alone but with repeat imaging if there is failure to settle.

Patients with possible spinal infection need blood cultures, prompt MRI and spinal surgery review. In stable patients not requiring surgical intervention, a radiological biopsy should be considered to direct antimicrobial therapy. Tuberculosis, Brucella, Nocardia and/or fungal cultures should be requested in patients with appropriate risk factors.

In diabetic foot infections it is important to recognise severe limb-threatening infections that require urgent surgical management. Signs of this are systemic sepsis, poor glycemic control, gas in soft tissues, abscess and infection from an ulcer tracking deeply through the foot to another site (eg plantar ulcer to dorsum of foot).

Keywords: bone and joint infection, osteomyelitis, septic arthritis, prosthetic joint infection, biofilm, diabetic foot infection, discitis, vertebral osteomyelitis

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Chronically established bone and joint infection can be persistent, evolve or relapse, even in the face of prolonged antimicrobial therapy. Biofilm has important implications for diagnostics, as well as surgical and antibiotic management.

**The ‘hot’ joint**

An inflamed ‘hot’ joint has a wide differential including inflammatory causes (septic arthritis, reactive, rheumatoid arthritis, spondyloarthropathies, SLE, gout or pseudo-gout) and non-inflammatory causes (degenerative joint diseases, trauma, avascular necrosis and Charcot’s arthropathy). Septic arthritis is important to exclude, as delayed or inadequate treatment can lead onto cartilage and then joint destruction.

**Acute septic arthritis**

Acute septic arthritis can affect any joint. It most commonly affects the knee but may also involve wrists, ankles, hips and the symphysis pubis. Polyarticular septic arthritis is more common in patients with inflammatory joint disease or overwhelming sepsis. Injecting drug use is a risk factor for septic arthritis of the sternoclavicular, sternomanubral or sacroiliac joints, often also associated with endocarditis.

The presentation can be similar to other inflammatory causes, such as crystal arthropathy, with an acutely painful, swollen, warm, red joint and a reduced range of movement. Features suggesting septic arthritis include fever and/or chills and absence of prior history or risk factors for gout, but dual pathology can occur. Risk factors for septic arthritis include extremes of age, bacteraemia, inflammatory joint disease, diabetes, intravenous drug use, alcoholism, immunosuppression, malignancy, recent trauma, intra-articular injections, arthroscopy or a prosthetic joint. The clinical history should include a sexual history (for gonococcal arthritis). A history of tick bite in an endemic area may raise the possibility of Lyme arthritis.

Blood cultures may be positive in up to 50% of cases but are usually negative in gonococcal septic arthritis. Plain X-rays should be done to exclude other causes of hot joint, but are usually normal in early septic arthritis. Ultrasound is sensitive for detecting joint effusions and synovitis. Synovial fluid should be aspirated and examined for leucocytes, urate and pyrophosphate crystals and by Gram stain and culture. A semiquantitative synovial leucocyte count can differentiate inflammatory from non-inflammatory causes but cannot differentiate infection from other inflammatory causes. Gram stain has low sensitivity (<75%) especially in gonococcal arthritis (<25%). Cultures are also often negative in the latter and, if suspected, molecular diagnostics (16S polymerase chain reaction) on synovial fluid may be considered and a urethral swab or urine should be sent for culture / nucleic acid amplification testing. Rectal and pharyngeal swabs may be indicated. If Lyme arthritis is suspected diagnosis is by serology. Diagnostic joint aspiration should be performed before antibiotics are given, except when the patient is acutely septic and aspiration delayed. Table 1 highlights the common bacterial causes and example antibiotic choices. All cases of suspected acute septic arthritis should be referred urgently to orthopaedics. Prompt arthroscopic or open washout is generally recommended despite the absence of good quality clinical trials. Observational studies show that in patients where surgery may be high risk, a more conservative approach of repeated aspirations may be effective. There is no good evidence to guide duration of antibiotic therapy for septic arthritis in adults. If synovial fluid Gram stain and cultures are negative in gonococcal arthritis, synovial fluid should be cultured. If negative, other organisms such as anaerobes, fungi, or Bartonella henselae should be considered.

**Table 1. Suggested empiric antibiotics for native joint septic arthritis in adults (but consult local guidelines).**

| Patient group                               | Suggested empiric antibiotic choice (after blood cultures and joint aspirate) |
|---------------------------------------------|--------------------------------------------------------------------------------|
| No specific risk factors                    | Possible organisms                                      | No known drug allergies | Penicillin allergy (non-severe eg rash) | Penicillin allergy (severe eg anaphylaxis) |
| Sterile or non-infectious arthritis         | Staphylococcus ssp, beta-haemolytic streptococci | IV fluoxacinil          | IV anti-staphylococcal cephalosporin (eg cefuroxime) | Clindamycin |
| Frail, recurrent UTIs, end-stage renal failure, recent abdominal surgery? | Aerobic Gram-negative rods | IV co-amoxiclav | IV 3rd generation cephalosporin (eg ceftriaxone) | Clindamycin plus ciprofloxacin |
| MRSA risk                                   | Metillin-resistant S aureus | Add IV glycopeptide⁶⁷   | Add IV glycopeptide⁶⁷ | Add IV glycopeptide⁶⁷ |
| Suspected gonococcal septic arthritis       | *Neisseria gonorrhoeae* | IV 3rd generation cephalosporin (eg ceftriaxone) | IV 3rd generation cephalosporin (eg ceftriaxone) | Clindamycin plus ciprofloxacin |
| Intravenous drug usage                      | S aureus. Less likely *Pseudomonas aeruginosa*, Fungal | IV fluoxacinil | IV anti-staphylococcal cephalosporin (eg cefuroxime) | Clindamycin |
| Known colonised with multidrug resistant organism | MRSA, ESBL, CPE etc | Discuss with microbiology |

Table adapted from ¹⁴

⁶⁷Glycopeptides include vancomycin and teicoplanin. These should be used at high doses in bone and joint infection ie vancomycin at 10–12 mg/kg and teicoplanin at 10 mg/kg. Modifications to dosing will need to be made in the setting of low body weight and/or impaired renal function.

⁷ESBL = extended-spectrum beta-lactamases; CPE = carbapenemase-producing enterobacteriaceae; IV = intravenous; MRSA = Metillin resistant Staphylococcus aureus
Infections involving prosthetic joints (PJI) should always be referred to orthopaedics, who should also involve infection specialists. Acute PJI occurs either postoperatively (up to 3 months after the initial arthroplasty) or through haematogenous spread after a period in which the prosthesis has been sound. For acute PJIs, blood cultures, plain X-ray, ultrasound and aspiration of the joint are the initial diagnostic modalities. The prosthesis at this stage is usually sound (not loose) and the most appropriate management is a DAIR procedure (a radical open Debridement with exchange of modular components but Retention of the Implant followed by Antibiotics). This needs to be done by an orthopaedic surgeon experienced in managing PJIs. Joint aspiration / arthroscopic washout may be required as an interim measure to drain pus if the relevant expertise is not immediately available. More chronic infections (usually presenting as increasing pain, loose prosthesis or discharging sinus) may be managed by revision of the prosthesis or excision arthroplasty. When the prosthesis is retained, a DAIR may be considered. Expert management of the soft tissues and dead space is important and may require plastic surgery input eg a muscle flap. When surgery is performed for chronically infected prosthetic joints, this should be done off antibiotics and multiple samples taken using separate instruments for separate sites, for microbiology and histology. Postoperative antibiotics need to be managed by infection specialists and may be prolonged.

**Osteomyelitis**

In children the most common site for acute haematogenous osteomyelitis is the growing end of long bones. In adults, it is the spine. The most common organism is *S aureus* but other pathogens such as beta-haemolytic *Streptococcus* spp, *Haemophilus influenzae*, *Kingella kingae* or *Mycobacterium tuberculosis* are possible. In patients with sickle cell disease, osteomyelitis is commonly due to *Salmonella* spp.

Osteomyelitis in children may relapse decades later in adulthood. Osteomyelitis may also occur in relation to infected fracture fixation devices. Infection may then contribute to delayed or non-union of the fracture. Figure 1 demonstrates the pathogenesis of osteomyelitis.

Acute osteomyelitis usually presents with fever and pain at the site of infection. Other skeletal sites should be examined as multifocal osteomyelitis can occur. Blood cultures and plain films should be performed. The purpose of plain films is to look for other causes of pain (eg fracture) and evidence of periosteal reaction or lucency. The imaging modality of choice for osteomyelitis, however, is MRI, which may show bone oedema, abscess formation, and periosteal reaction. If infection is chronic there may also be evidence of a sinus, cloacae, periostitis, sequestrum and/or involucrum (Fig 2). Occasionally, chronic disease is noted through the shaft of the femur, as well as visible overlying soft tissue deformity.
pathological fractures occur. If there are soft tissue collections, prompt aspiration for microbiology is appropriate. In acute osteomyelitis, many patients will respond to antibiotic treatment but those with evidence of pus on imaging require urgent surgical drainage / decompression. When osteomyelitis is chronic and/or related to a fracture fixation device it needs management by a multidisciplinary team experienced in the management of bone infections. Ideal management is with a combination of surgical resection with meticulous intraoperative sampling with/without stabilisation, with/without reconstruction, soft tissue management by plastic surgeons and appropriately managed postoperative antibiotics.

**Spinal infections (vertebral osteomyelitis, discitis, epidural abscesses)**

Spinal infections are most commonly either haematogenous or postsurgical. Haematogenous infections are most commonly due to *S. aureus*, *Streptococcus* spp, *aerobic Gram-negative bacilli* and *M. tuberculosis* to *postsurgical*. Haematogenous infections are most commonly due to *S. aureus*. *Streptococcus* spp, *aerobic Gram-negative bacilli* and *M. tuberculosis* should also be considered as should *Brucella* spp in endemic areas and fungi in immunocompromised patients. All patients with known fever and weight loss and/or bacteraemia and/or endocarditis should have prompt spinal imaging if they have new or worsening back pain. Imaging, usually by MRI (or computed tomography if MRI is contraindicated), should look for evidence of epidural abscess, discitis, vertebral osteomyelitis and, critically, for evidence of cord / cauda equina compression or vertebral instability requiring urgent surgical intervention. Epidural collections usually require surgical drainage. Unless blood cultures have already revealed a causative organism, deep microbiological samples should be obtained. This can be with intraoperative samples if surgery is indicated, or by radiological biopsy in other cases. This should be done urgently and, where possible, antibiotics withheld until after the biopsy has been taken (antibiotic therapy should not be delayed however in septic/ unstable patients). Cases should be discussed with microbiology to ensure the relevant tests are done in the laboratory.

Native vertebral osteomyelitis should be managed with 6 weeks of appropriately targeted therapy (possibly longer in complicated cases or where organisms such as *Brucella* spp are identified). Duration of therapy for epidural abscesses depends upon whether it was surgical drained and clinical/radiological response.

**Diabetic foot osteomyelitis**

Diabetic foot infections usually occur following skin ulceration in patients with neuropathy and/or vascular insufficiency. Infections can go on to cause adjacent osteomyelitis. In severe infections this can rapidly become limb and life threatening. It is essential for all patients, especially diabetics, presenting through acute medical services to have a full foot examination including a severity based on the Infectious Diseases Society of America severity score as follows:

- **Uninfected**: No symptoms or signs of infection present (symptoms/signs defined as at least two of local swelling or induration, erythema, local tenderness or pain, local warmth, purulent discharge)
- **Mild**: Local infection involving only the skin and subcutaneous tissue. If erythema, must be >0.5–1 cm around the ulcer. Exclude other causes of inflammatory response of the skin
- **Moderate**: Local infection (as above) with erythema >2 cm, or involving structures deeper than skin and subcutaneous tissue, and no systemic inflammatory response signs
- **Severe**: Local infection with signs of systemic inflammatory response with two or more of temperature >38°C or <36°C, heart rate >90 beats/min, respiratory rate >20 breaths/min or PaCO2 <32 mmHg, white blood cell count >12,000 or <4000 cells/μL, or ≥10% immature (band) forms

### Table 2. Diabetic foot infections: pathogens and suggested empiric antimicrobial therapy (but consult local guidelines)

| Severity* | Usual pathogens | Treatment |
|-----------|-----------------|-----------|
| Uninfected | Metillin-sensitive *Staphylococcus aureus* (MSSA); *Streptococcus* spp | No known drug allergy |
| Mild (usually treated with oral agents) | Metillin-resistant *S. aureus* (MRSA) | Penicillin allergy (non-severe eg rash) |
| Moderate (may be treated with oral or initial parenteral agents) | *MSSA; Streptococcus* spp; Enterobacteriaceae; anaerobes | Penicillin allergy (severe eg anaphylaxis) |
| Risk of *Pseudomonas aeruginosa* | | |
| MRSA | | |
| Severe (usually treated with parenteral agents) | *MRSA; Enterobacteriaceae; Pseudomonas; anaerobes* | |

*Severity based on the Infectious Diseases Society of America severity score as follows:*

- **Uninfected**: No symptoms or signs of infection present (symptoms/signs defined as at least two of local swelling or induration, erythema, local tenderness or pain, local warmth, purulent discharge)
- **Mild**: Local infection involving only the skin and subcutaneous tissue. If erythema, must be >0.5–1 cm around the ulcer. Exclude other causes of inflammatory response of the skin
- **Moderate**: Local infection (as above) with erythema >2 cm, or involving structures deeper than skin and subcutaneous tissue, and no systemic inflammatory response signs
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the removal of dressings. An audit (England and Wales) in 2015 showed that two-thirds of diabetic inpatients did not have a specific diabetic foot risk examination while inpatient.17

Urgent surgical intervention may be required in those patients with abscesses, necrotising soft tissue infections and/or uncontrollable sepsis. Clinical evidence of pus tracking from one area (eg ulcer) to another may be indicative of deep spreading infection. Less urgent surgery may be required for those with substantial non-viable tissue or extensive bone or joint involvement. An early vascular assessment and involvement of a vascular surgeon to consider revascularisation in appropriate cases, especially those with critical ischaemia, is essential. Table 2 gives a guide to empiric antibiotic management based on the severity of disease. Tissue sampling can be helpful for antimicrobial management. Superficial swabs often represent colonisation only, whereas deep tissue curettages/bone sampling can allow for appropriately targeted treatment.

Conclusions

Bone and joint infections can present through the acute medical take. It is important to take a careful history and remove any dressings during initial assessment and to recognise when urgent surgical intervention is required. Microbiological sampling should be done in all cases, to allow for targeted antimicrobial management. Comorbidities must be adequately managed. Involvement of the multidisciplinary team is essential.

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