The role of SOCS proteins in the development of virus-induced hepatocellular carcinoma

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Abstract

Background: Liver cancer has become one of the most common cancers and has a high mortality rate. Hepatocellular carcinoma is one of the most common liver cancers, and its occurrence and development process are associated with chronic hepatitis B virus (HBV) and hepatitis C virus (HCV) infections.

Main body

The serious consequences of chronic hepatitis virus infections are related to the viral invasion strategy. Furthermore, the viral escape mechanism has evolved during long-term struggles with the host. Studies have increasingly shown that suppressor of cytokine signaling (SOCS) proteins participate in the viral escape process. SOCS proteins play an important role in regulating cytokine signaling, particularly the Janus kinase-signal transducer and activator of transcription (JAK-STAT) signaling pathway. Cytokines stimulate the expression of SOCS proteins, in turn, SOCS proteins inhibit cytokine signaling by blocking the JAK-STAT signaling pathway, thereby achieving homeostasis. By utilizing SOCS proteins, chronic hepatitis virus infection may destroy the host's antiviral responses to achieve persistent infection.

Conclusions: This review provides recent knowledge regarding the role of SOCS proteins during chronic hepatitis virus infection and provides some new ideas for the future treatment of chronic hepatitis.

Keywords: SOCS, Hepatocellular carcinoma, Cytokine, JAK-STAT signaling pathway, Hepatitis virus

Background

Although substantial progress has been achieved in modern medical standards, unknown and existing viruses still pose serious risks to human and animal lives. For example, the epidemics of hepatitis B virus (HBV) present an escalating trend of global infection risk which causes a large number of people to suffer from chronic HBV. More than 257 million people worldwide are chronically infected with HBV, even until now, the virus has not been completely eradicated [1]. Unfortunately, such infection can only be controlled. However, what is even worse, is that the outcomes of these infected with HBV are often diagnosed as cirrhosis and hepatocellular carcinoma (HCC). HCV, another type of hepatitis virus that also poses serious threats to human health, its infection originates from an initial asymptomatic chronic infection which can lead to life-threatening cirrhosis or HCC [2]. The severity of these viral infections is due to the effective
invasion strategy of these viruses, such as adhesion, invasion, and proliferation [3]. Furthermore, the virus has never stopped evolving escape mechanisms during the long-struggling with the host which spared them a better chance to evade the host’s defense.

Some cytokines, such as Type I interferon, have direct or indirect antiviral effects on fighting virus infection. Cytokines constitute a class of signaling molecules secreted by cells that play critical regulatory roles in communication with surrounding cells [4]. Cytokines activate cell surface receptor complexes to regulate the activation, growth, differentiation, and apoptosis of cells [5]. These receptors are mainly divided into four categories. The first class of receptors primarily activates nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) and mitogen-activated protein (MAP) kinases, which are mainly involved in the tumor necrosis factor α family, including the interleukin-1 family (IL-1β, IL-18, and IL-33) and the IL-17 family [6]. The second class of receptors primarily activates the Janus kinase-signal transducer and activator of transcription (JAK-STAT) signaling pathway, and most cytokines function by activating this signaling pathway [7]. The third class of receptors comprises the transforming growth factor β receptor, which mainly activates the Smad family of transcription factors [8]. The final class of receptors includes growth factor receptors, which primarily function through the Ras intracellular signal-regulated kinase pathway [9]. Cytokines activate intracellular signaling pathways by binding the corresponding receptors, but this process does not occur continuously because of a negative feedback regulation mechanism, which is important for homeostasis.

Cytokine-inducible SH2 (CIS) and suppressor of cytokine signaling (SOCS) proteins play important roles in regulating cytokine signaling, particularly the JAK-STAT signaling pathway. Numerous studies have found that cytokines stimulate the expression of SOCS proteins; however, SOCS proteins inhibit cytokine signaling by blocking the JAK-STAT signaling pathway, thereby achieving homeostasis [10]. In addition to cytokines, other stimuli, such as lipopolysaccharides, bacteria, viruses, and chemokines, induce the expression of SOCS proteins [11]. Additionally, empirical evidences have shown that a close relationship exists between the SOCS family and cancer [12–14]. According to the International Agency for Research on Cancer, liver cancer is one of the most common cancers with a high mortality rate. HCC is one of the most common liver cancers, and its occurrence and development processes are associated with cirrhosis induced by HBV and HCV infections [15]. In addition, an increasing number of studies indicate that SOCS proteins are involved in the development of HCC [16–18]. Up to this point, this review aims to summarize recent knowledge regarding the role of SOCS proteins during hepatitis virus infection, as well as contribute some new ideas for the future treatment of hepatitis.

**Immune regulation by SOCS**

**CIS/SOCS family**

The CIS/SOCS family includes CIS and SOCS proteins, and can also further be subdivided into SOCS1, SOCS2, SOCS3, SOCS4, SOCS5, SOCS6, and SOCS7 [19, 20]. The proteins belonging to this family have similar structures: an SH2 domain is located in the central region, the amino terminus is of variable length and sequence, and the carboxy terminus is a module consisting of 40 amino acids, namely, the SOCS box [21]. These three-part structures have different functions; the SH2 domain recognizes and binds a cognate phospho-tyrosine residue, and different SOCS/CIS proteins use this domain to execute their regulatory functions. The amino-terminal region interacts with the substrate. However, the SOCS box interacts with elongin B, elongin C, and cullin 5, utilizes the RING-finger-domain-only protein (RBX2) to recruit E2 ubiquitin-transferase, and ubiquitinates Janus kinases (JAKs) and other cytokine receptors, ultimately targeting these proteins for proteasomal degradation (Fig. 1a) [17,
membrane receptor, i.e., the IFNα receptor (IFNAR), cytokines. Type I interferon binds a heterodimeric transmembrane receptor, which is composed of IFNAR1 and IFNAR2 subunits. Once IFNAR is bound by Type I interferon, the receptor-associated protein tyrosine kinases JAK1 and tyrosine kinase 2 (TYK2) are activated; the transcription factors STAT1 and STAT2 are then phosphorylated and form a dimer. Dimerized STAT1 and STAT2 assemble with IRF9 to form a trimolecular complex called IFN-stimulated gene factor 3 (ISGF3). Finally, ISGF3 translocates to the nucleus and binds its cognate DNA sequences called IFN-stimulated response elements (ISRE), thereby initiating the transcription of ISGs. [28–30]

Next, the SOCS proteins begin to express and play their roles in type I interferon signaling. CIS, SOCS1, SOCS2, and SOCS3 aim to execute their negative feedback regulation in JAK-STAT-mediated cytokine signaling, while SOCS4, SOCS5, SOCS6 and SOCS7 primarily focus on regulating the growth factor receptor signal. Among these proteins, SOCS1 and SOCS3 are the most widely studied [12]. SOCS1 and SOCS3 possess a unique kinase inhibitory region (KIR) at the amino terminus that inhibits the activity of JAK tyrosine kinase by acting as a pseudo substrate, meanwhile, the other SOCS proteins also directly or indirectly interact with JAKs or specific cytokine receptors using their respective structures, thereby degrading targeted proteins [31]. For example, SOCS1 and SOCS3 directly inhibit the activity of JAK tyrosine kinase through the KIR region, at the same time, SOCS proteins also block STAT by interacting with the phosphorylation region of the STAT receptor, thereby inhibiting cytokine signaling. Likewise, SOCS3 specifically inhibits STAT3 and STAT4 [17]. During the fine phase, the SOCS box region begins to recruit E2 ubiquitination transferase. Next, this complex ubiquitinylates the target proteins, marking them for proteasomal degradation. In addition, the SOCS proteins directly regulate the degradation of related proteins, such as E3 ubiquitin-ligation ligase [19].

In addition to regulating the JAK-STAT signaling pathway, SOCS proteins can also modulate the TLR signaling pathway. Similar to the negative feedback regulation of the JAK-STAT signaling pathway, SOCS proteins mainly bind key molecules in the Toll receptor signaling pathway and then use the SOCS box region to ubiquitinate and degrade the target molecule, thereby suppressing the signaling pathway. As SOCS1 and SOCS3 are amongst the most widely reported in the past studies. The findings have shown that SOCS1 and SOCS3 use the SH2 region to recognize and bind the myeloid differentiation factor 88 (MYD88)-adaptor-like (Mal) protein [32], TRAF3/6 [33] and a subunit of NF-kB p65 [34] that contains a phosphorylated tyrosine. In the same way, SOCS1 and SOCS3 bind and degrade IRF7, inhibiting the production of TLR7-mediated type I interferon [35].

SOCS proteins were originally shown to inhibit cytokine signaling through the JAK-STAT signaling pathway [27]. With more advanced research, SOCS proteins were found to play an important role in inflammation and the development and progression of cancers. The induction of the SOCS proteins is usually mediated by the JAK-STAT pathway. Following binding to cellular membrane surface receptors, cytokines and interferons activate and phosphorylate receptor-associated JAK proteins. Next, the activated JAK proteins phosphorylate the cytoplasmic domains of receptors. These phosphorylated receptors begin to recruit and activate STATs, enabling the dimerization of STATs. These dimerized complexes then enter the nucleus to initiate the transcription of different target genes, including the SOCS genes [17].

**Type I interferon signaling and the SOCS signaling pathway**

Innate immunity is the first-line defense against invading pathogens and plays an important role in maintaining the host’s safety. The innate immune system includes the following three classical pattern-recognition responses (PRRs): Toll-like receptors (TLRs), retinoic acid-inducible gene I-like receptors (RLRs), and nucleotide oligomerization domain-like receptors (NLRs). Among these receptors, TLRs and RLRs play a pivotal role in the production of type I interferon and various cytokines, and NLRP3, which is the most widely studied NLRs, regulates the maturation of IL-1β and IL-18 via the activation of caspase-1.

After recognition, TLRs immediately recruit Toll/interleukin-1 receptor (TIR)-containing adapters, such as MyD88 and TIR domain-containing adapters inducing IFN-β (TRIF), thus transmitting related signals to downstream molecules. Following the recognition of viral double-stranded RNA, the TLR3 signaling pathway is activated by the TRIF adaptor, and TNF receptor-associated factor 3 (TRAF3) is recruited in response. TRAF3 activates two related kinases, inducible IκB kinase (IKK) and TANK-binding kinase 1 (TBK1), which, in turn, mediate the phosphorylation of interferon regulatory factor 3/7 (IRF-3/7). Finally, IRF-3/7 enters the nucleus and stimulates the production of type I interferon and cytokines. Type I interferon binds a heterodimeric transmembrane receptor, i.e., the IFNα receptor (IFNAR), which is composed of IFNAR1 and IFNAR2 subunits. Once IFNAR is bound by Type I interferon, the receptor-associated protein tyrosine kinases JAK1 and tyrosine kinase 2 (TYK2) are activated; the transcription factors STAT1 and STAT2 are then phosphorylated and form a dimer.
regulates insulin signaling by binding insulin receptor substrate 1 and 2 (IRS1 and IRS2) proteins [36], and SOCS2 induces TRAF6 degradation (Fig. 1b) [37].

Role of SOCS in hepatic viral infection
SOCs signaling following the activation of innate immunity by viral infections
A complex balance system exists in the host that ensures that innate and adaptive immune responses are properly activated and terminated via the regulation of positive and negative signaling to clear invading viruses and regulate uncontrolled inflammation. However, persistent pathogen infection may disrupt this balanced mechanism to achieve long-term stable replication and proliferation in the host. Host cells recognize the components of a virus through PRRs and activate innate immune responses, while adaptive immunity plays an important role in preventing the reinvagination of pathogens. Once a virus invades the host, PRRs quickly recognize the virus, activate complex intracellular signaling pathways and target inflammatory and antiviral responses, ultimately removing the invading virus rapidly. TLRs not only activate the host’s innate immunity but also promote specific adaptive immunity by acting on antigen presenting cells. Among these processes, cytokine production is inseparable from the key signaling pathway, i.e., the JAK-STAT signaling pathway. The JAK-STAT signaling pathway regulates more than 50 downstream cytokines and growth factors as a central communication node in the immune system. This signaling pathway mainly comprises JAK and STAT proteins; JAKs can be subdivided into JAK1, JAK2, JAK3 and TYK2, while STATs can be subdivided into STAT1, STAT2, STAT3, STAT4, STAT5A, STAT5B and STAT6 [38]. As a negative feedback regulator of the JAK-STAT signaling pathway, SOCS proteins play an important role in viral infection. SOCS proteins regulate the viral-induced cytokine levels by initiating the JAK-STAT signaling cascade and interfering with cellular signaling by regulating the degradation of signaling proteins. These signaling proteins regulate immune cells to produce large amounts of pro- and anti-inflammatory cytokines, which further determine the host’s sensitivity to viral infections and host outcomes after viral infection [39].

HCV and SOCS proteins
HCV was the first virus used to explore the function of SOCS proteins, which were revealed to inhibit type I interferon signaling. Once HCV stably proliferates in the liver, the virus can compete with the antiviral response produced by the host and interact with various immune evasion mechanisms to damage the host’s antiviral response, such as the type I interferon response. A recent study revealed that HCV increased SOCS1 expression as the viral load increased [40]. HCV infection upregulates SOCS proteins via different mechanisms depending on different HCV proteins. In particular, The HCV protein p7 induces SOCS3 via STAT3 and extracellular signal-regulated kinase (ERK)-mediated pathways [41]. On the other hand, SOCS7 expression induced by the HCV core protein genotype 3a is independent of STAT3 and may be regulated by peroxisome proliferator-activated receptor gamma (PPAR-γ) activity [42]. However, the mechanism by which the HCV core protein induces STAT3 or PPAR-γ still remains uncertain. Therefore, such subject is worthy of further investigation. The related research outcomes can provide meaningful implications for the future treatment of HCV infection.

Due to the fact that HCV infection mainly evades host innate immunity by regulating SOCS proteins. Overexpressed SOCS1 protein also inhibits the antiviral effect of type I interferon [43] and type III interferon (IFN-λ) signaling [44, 45], which inhibits the IFNa/β-induced activation of the JAK/STAT signaling pathway by downregulating the phosphorylation of STAT1 and STAT3. SOCS1 and SOCS3 proteins inhibit IFN-α induced expression of the anti-viral proteins 2′,5′-OAS and MxA in Hep-G2 cells [43]. Hepatic SOCS3 expression is increased in patients infected with HCV genotype 1, which may be a mechanism by which SOCS3 reduces the biological response to IFN-α [46]. In addition to inhibiting innate immunity signaling, different HCV genotypes have different effects on adaptive immunity by regulating SOCS proteins. The HCV core protein differentially regulates T- and B-cell signaling by exploiting programmed death-1 (PD-1) and SOCS1 functions [47, 48]. A previous report provided evidence that the HCV core protein inhibits T-cell responses by interacting with gC1qR, inducing SOCS1/3 and suppressing STAT1/3 [49, 50]. However, B cells are abnormally activated during chronic HCV infection, characteristics associated with TNF- and Apo-L-related leucocyte-expressed ligand-1 (TALL-1) overexpression and SOCS1 suppression [51]. Another study revealed that patients with HCV genotype 3 sustained virological responses (SVR) have significantly higher expression of SOCS or lower expression of MHC class-II genes, which is associated with a diminished capacity to present antigens and a functionally defective state of dendritic cells [52]. However, a recent study revealed that the transcript levels of SOCS1 in peripheral blood mononuclear cells do not significantly differ between 100 HCV genotype 1 patients with SVR and non-SVR [53]. Another study revealed that a specific strain of HCV, i.e., genotype 1b, fails to activate the transcription...
of SOCS1, SOCS3, and SOCS7. Furthermore, amino acids 49 and 131 of the HCV core-encoding sequence mediate these transactivating effects [54].

Patients with chronic HCV infection often have multiple associated diseases, several of which have found to be linked to SOCS protein expression. Empirical epidemiological studies show strong evidences in which HCV infection is linked to type 2 diabetes, which is thought to originate from the downregulation of IRS1 and IRS2 by upregulating SOCS3 during HCV infection [55, 56]. Furthermore, the HCV core protein of genotype 3a promotes IRS1 (but not IRS2) degradation by upregulating SOCS7 in a human hepatoma cell line (Huh-7) [57]. Another report revealed that the HCV core protein in both transgenic mice or hepatoma cells downregulates IRS1 and IRS2 by upregulating SOCS3 [56]. These data suggest that HCV core-induced SOCS3 and SOCS7 promote the proteasomal degradation of IRS1 and IRS2 through ubiquitination, thereby contributing to HCV-induced insulin resistance. Furthermore, HCV suppresses proinflammatory TNF-α responses by inducing SOCS3 [58].

Recently, numerous reports have shown that miRNAs play an important role in hepatitis virus infections. In these reports, miR146a is upregulated in monocytes from HCV-infected patients [18], and miRNA221 is upregulated in serum from HCV chronic hepatitis patients and Huh7.5.1 cells infected with HCV [59]. Furthermore, miRNA164a increases IL-23, IL-10, and TGF-β1 expression by inhibiting SOCS1 and inducing STAT3 [18]. In addition, miRNA221 enhances the action of interferon against HCV by targeting SOCS1 and SOCS3 [59]. miRNA122 also regulates type I interferon by blocking SOCS1 [60]. These phenotypes demonstrate that the host reacts to HCV via certain miRNAs targeting SOCS proteins against HCV. However, HCV employs multiple strategies to evade host immune responses by hijacking different SOCS proteins (Table 1).

### HBV and SOCS proteins

In another hepatitis virus study, HBV infection increased SOCS3 expression in Huh7 cells, resulting in sustained STAT3 activation [61]. Additionally, the woodchuck model has been used to study HBV, and woodchuck hepatitis virus (WHV) elevates the mRNA levels of SOCS3 in the liver during chronic WHV infection [62]. Uncontrolled SOCS and STAT signaling result in more severe inflammation in the liver, which may be related to persistent HBV infections [61]. A recent report showed that an HBV antigen, i.e., HBeAg, augments miR-155 expression in macrophages, and increased miR-155 levels promote HBeAg-induced inflammatory cytokine production by suppressing SOCS1 expression in HepG2.2.15 cells [63]. However, another HBV antigen, i.e., HBsAg, significantly induces the expression of SOCS1, which may be a key factor in the HBsAg-regulated inhibition of TLR9-mediated signaling; furthermore, HBsAg binds BDCA2 receptors on the plasma membrane of pDCs, resulting in the inhibition of IFN-α production [61].

| Virus | SOCS protein | In vivo or In vitro | Viral proteins | Mechanism | References |
|-------|--------------|---------------------|----------------|-----------|------------|
| HCV   | SOCS1/3      | Hep-G2 cell         | HCV core protein | Inhibit IFN-α induced expression of 2′,5′-OAS and MxA | [40] |
|       | SOCS3        | patients            | HCV genotype 1  | Reduce the biological response to IFN-α | [43] |
|       | SOCS1        | PBMC                | HCV core protein | Dysregulate T- and B-cell signalling | [44–47] |
|       | SOCS         | Myeloid DCs        | HCV genotype 3  | Diminish capacity to present antigen | [49] |
|       | SOCS3/7      | Huh-7               | HCV genotype 3  | Downregulation of IRS1 and IRS2 | [52–54] |
| HBV   | SOCS3        | Huh-7 and liver specimens from HBV-infected patients | adenoviral AdHBV (genotype A) | Dysregulate STAT/SOCS-signalling | [58] |
|       | SOCS1        | HepG2.2.15 cells and PBMC | HBV antigen, HBeAg | Promote inflammatory cytokine production | [60] |
|       | SOCS1        | plasmacytoid dendritic cells | HBsAg | Inhibition of the IFN-α production | [61] |
|       | SOCS1/3      | HBV transgenic mice | CTP-HBcAg18-27-tapasin | Enhance T cell immune responses | [62–64] |

Viral proteins disrupt the host’s immune response by hijacking different SOCS proteins in vivo or in vitro.

*Table 1 Hepatitis viruses have developed multiple strategies to disrupt the immune response by hijacking the SOCS system*
example, how does HBsAg induce SOCS1 expression? HBsAg may rely on the same pathway as the HCV core protein, inducing SOCS proteins via STAT3 and ERK-mediated pathways. Notably, HBsAg and other HBV antigens more likely interact with certain proteins to induce SOCS proteins.

HBV inhibits the level of endogenous interferon and induces SOCS1 and SOCS3 expression; however, following interferon treatment, HBV replication is inhibited, and the expression levels of SOCS1 and SOCS3 begin to decrease, indicating that HBV may achieve sustained infection in vivo by upregulating the expression of the SOCS proteins [68]. SOCS1 expression is negatively correlated with the prognosis of patients with acute-on-chronic hepatitis B liver failure (ACHBLF); meanwhile, patients without SOCS1 methylation display a favorable response to glucocorticoid treatment [69]. Furthermore, HBV-X mutants enhance STAT3 activation, inhibit STAT1 expression and silence SOCS1 and SOCS3 expression, likely because of the atypical nuclear/perinuclear localization of the HBV-X mutants [70]. Furthermore, SOCS1 polymorphisms may affect the susceptibility and outcome of HBV infection [71], and CIS polymorphisms are associated with persistent HBV infection [72]. Most patients with chronic HBV infection develop HCC. Increasing evidence indicates that the X protein of HBV is closely related to HCC. In addition, the X gene of HBV downregulates the SOCS1 expression but increases SOCS-1 gene promoter CpG island methylation, leading to oncogenes activation and HCC development [73]. X. ZHANG et al. also found that the gene loss and epigenetic silencing of SOCS1 are strongly associated with HBV-related HCC [14]. However, the hypermethylation of CpG islands of SOCS-1 is not closely related to HBV-induced HCC [13]. Furthermore, the expression of SOCS family proteins is remarkably suppressed in the livers from HBV X protein transgenic mice relative to that in non-transgenic mice from the early to late stages of partial hepatectomy (PH) compared with non-PH mice [74]. The relationship between HCC and SOCS proteins remains largely unknown. In summary, innate and adaptive immunity may be disrupted during HBV infection by enhanced levels of SOCS proteins, leading to persistent HBV infection (Table 1).

**Other hepatitis viruses and SOCS proteins**

Only a few studies have reported the interactions between SOCS proteins and hepatitis A virus (HAV). However, in various animal models of HAV, SOCS proteins are involved in HAV infection. For example, previous reports have shown that ducks can be used as a small animal model for studying HAV, and duck hepatitis A virus (DHAV) can induce acute and chronic hepatitis in ducks [75]. During DHAV-1 infection, the liver expresses a large amount of SOCS1 and SOCS3, which may be involved in viral adaption [76]. Furthermore, DHAV-1 inhibits type I interferon signaling to assist viral adaption by increasing SOCS3 expression [77].

**SOCS proteins as therapeutic targets to treat chronic hepatitis**

Considering the relationships between chronic hepatitis and SOCS proteins, small-molecule antagonists of SOCS signaling and SOCS protein silencing could be viable strategies for improving the treatment of chronic hepatitis as well as effectively mitigating hepatitis virus infection. Waiboci et al. found that the KIR region of SOCS-1 binds directly to the phosphorylation site of JAK2 and they synthesized a peptide of pJAK2 (1001–1013, LPQDKEYYKVEKP) as an antagonist of SOCS1 [78]. This antagonist enhances innate and adaptive immune response by targeting the KIR of SOCS1 [79]. HBV and HCV evade host immune responses by hijacking SOCS proteins. Therefore, the SOCS1 antagonist may be valuable to treat chronic hepatitis. Furthermore, IL-7 downregulates SOCS3, resulting in amplified cytokine production, increases T-cell effector function and numbers, and attenuates viral persistent infection [80]. HBV infection disrupts innate and adaptive immunity by enhancing the levels of SOCS proteins, causing persistent HBV infection. These attributes of SOCS protein inhibition by an antagonist (pJAK2 peptide) or natural cytokines (IL-7) have profound implications for the therapeutic in the treatment of chronic hepatitis.

**Conclusions**

In recent years, we have enhanced our understanding of the structure and function of SOCS proteins by conducting numerous in-depth researches. By functioning as negative feedback regulators, SOCS proteins are involved in the participation of the life cycle of the host, especially during the process of hepatitis virus infection. Studying the role of SOCS proteins in hepatitis virus-induced hepatocellular carcinoma provides some new ideas for improving the future treatment of hepatitis.

**Abbreviations**

HCC: Hepatocellular carcinoma; HBV: Hepatitis B virus; HCV: Hepatitis C virus; NF-kB: Nuclear factor kappa-light-chain-enhancer of activated B cells; MAP: Mitogen-activated protein; SOCS: Suppressor of cytokine signaling; JAK-STAT: Janus kinase–signal transducer and activator of transcription; CIS: Cytokine-inducible SH2; RBX2: Ring-finger-domain-only protein; PRRs: Pattern-recognition responses; TLR: Toll-like receptor; RLRs: Retinoic acid-inducible gene I-like receptors; NLRs: Nucleotide oligomerization domain-like receptors; IKKi: Inducible IκB kinase; TBK1: TANK-binding kinase 1; IFNAR: IFNα receptor; ISGF3: IFN-stimulated gene factor 3; ISRE: IFN-stimulated response elements;
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Authors’ contributions
J.X. A.C. and M.W. conceptualized the project; J.X. wrote the paper; R.J., D.Z., M.L., S.C., X.Z., Q.Y., Y.W., S.Z., Q.L., Y.W., Z.X., Z.C., L.Z., Y.L., L.Z. and X.C reviewed and edited the paper.

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References
1. Zhang YY, Hu KQ. Rethinking the pathogenesis of hepatitis B virus (HBV) infection. J Med Virol. 2015;87:1899–99.
2. Martinello M, Hajirazaei A, Gerebely J, Dore B, Matthews GV. Management of acute HCV infection in the era of direct-acting antiviral therapy. Nat Rev Gastroenterol Hepatol. 2018;15:412–24.
3. Takeuchi O, Akira S. Innate immunity to virus infection. Immunol Rev. 2009;227:75–86.
4. Dinarello CA. Historical insights into cytokines. Eur J Immunol. 2007;37(Suppl1):534–45.
5. Vlasova-St Louis I, Bohjanen PR. Post-transcriptional regulation of cytokine and growth factor signaling in cancer. Cytokine Growth Factor Rev. 2017;33:83–93.
6. Wajant H, Henkler F, Scheurich P. The TNF- receptor-associated family: scaffold molecules for cytokine receptors, kinases and their regulators. Cell Signal. 2001;13:389–400.
7. Waldmann TA, Chen J. Disorders of the JAK/STAT pathway in T cell lymphopah pathogenesis: implications for immunotherapy. Annu Rev Immunol. 2017;35:533–50.
8. Budi EH, Duan D, Deryckx R. Transforming growth factor- β receptors and smads: regulatory complexity and functional versatility. Trends Cell Biol. 2017;27:658–72.
9. Yoshimura A, Ito M, Chikuma S, Akamuna T, Nakatsuika H. Negative Regulation of Cytokine Signaling in Immunity. Cold Spring Harb Perspect Biol. 2018;10:a028571.
10. Kershaw NJ, Murphy JM, Lucy IS, Nicola NA, Babon JJ. Regulation of Janus kinases by SOCS proteins. Biochem Soc Trans. 2013;41:1042–7.
11. Rakesh K, Agrawal DK. Controlling cytokine signaling by constitutive inhibitors. Biochem Pharmacol. 2005;70:649–57.
12. Inagaki-Ohara K, Kondo T, Ito M, Yoshimura A. SOCS, inflammation, and cancer. JAKSTAT. 2013;2:e24053.
13. Ko E, Kim SJ, Joh JW, Park CK, Park J, Kim DH. CpG island hypermethylation of SOCS-1 gene is inversely associated with HBV infection in hepatocellular carcinoma. Cancer Lett. 2008;271:240–50.
14. Zhang X, Wang J, Cheng J, Ding S, Li M, Sun S, Zhang L, Liu S, Chen X, Zhuang H, Lu F. An integrated analysis of SOCS1 down-regulation in HBV infection-related hepatocellular carcinoma. J Viral Hepat. 2014;21:264–71.
15. Baumert TF, Schuster C, Cosset FL, Duboisson J, Hofmann M, Tauxz N, Zeisel MB, Thirme R. Addressing the next challenges: A summary of the 22nd international symposium on hepatitis C virus and related viruses. J Hepatol. 2016;64:968–73.
16. Zhao LJ, He SF, Wang W, Ren H, Qi ZT. Interferon alpha antagonizes STAT3 and SOCS3 signaling triggered by hepatitis C virus. Cytokine. 2016;80:48–55.
17. Delgado-Ortega M, Marc D, Dupont J, Trapp S, Berri M, Meurens F. SOCS proteins in infectious diseases of mammals. Vet Immunol Immunopathol. 2013;151:1–19.
18. Ren JP, Ying RS, Cheng YQ, Wang L, El Gazzar M, Li FY, Ning SB, Moorman JR, Yao QZ. HCV-induced miR146a controls SOCS1/STAT3 and cytokine expression in monocytes to promote regulatory T-cell development. J Viral Hepat. 2016;23:755–66.
19. Yoshimura A, Naka T, Kudo M. SOCS proteins, cytokine signalling and immune regulation. Nat Rev Immunol. 2007;7:454–65.
20. Gisselbrecht S. The CIS/SOCS proteins: a family of cytokine-inducible regulators of signaling. Eur Cytokine Netw. 1999;10:463–70.
21. Masuhara M, Sakaguchi R, Yasukawa H, Mitsuji K, Wakioka T, Akiyama S, Sasaki A, Misawa H, et al. Cloning and characterization of novel CIS family genes. Biochem Biophys Res Commun. 1997;239:439–46.
22. Yoshimura A, Suzuki M, Sakaguchi R, Hanada T, Yasukawa H. SOCS, inflammation, and autoimmunity. Front Immunol. 2012;3:20.
23. Kamura T, Maenaka K, Kotoshiba S, Matsumoto M, Kohda D, Conaway RC, Conaway JW, Nakayama K. VHL-box and SOCS-box domains determine binding specificity for Cul2-Rbx1 and Cul5-Rbx2 modules of ubiquitin ligases. Genes Dev. 2004;18:3055–65.
24. Yoshimura A, Ohkubo T, Kikuchi T, Jenkins NA, Gilbert DJ, Copeland NG, Har a T, Miyajima A. A novel cytokine-inducible gene CIS encodes an SH2-containing protein that binds to tyrosine-phosphorylated interleukin 3 and erythropoietin receptors. Embo J. 1995;14:2816–26.
25. Matsumoto A, Masuhara M, Mitsuji K, Yokouchi M, Ohtsubo M, Misawa H, Miyajima A, Yoshimura A. CIS, a cytokine-inducible SH2 protein, is a target of the JAK-STAT pathway and modulates STAT3 activation. Blood. 1997;89:3148–54.
26. Louis C, Souza-forseca-Guiramaes F, Yang Y, Silva D, Kratina T, Dagle y L, Hiedehy-Zadeh S, Rautela J, Masters SL, Davis MJ, et al. NK cell-derived GM-CSF potentiates inflammatory arthritis and is negatively regulated by SOCS proteins. Nat Immunol. 2007;8:1076–84.
27. Starr R, Willison TA, Viney EM, Murray LJ, Rayner JR, Jenkins BJ, Gonda TJ, Alexander WS, Metcalfe D, Nicola NA, Hilton DJ. A family of cytokine-inducible inhibitors of signalling. Nature. 1997;387:917–21.
28. Kershaw NJ, Murphy JM, Lucy IS, Nicola NA, Babon JJ. Regulation of Janus kinases by SOCS proteins. Biochem Soc Trans. 2013;41:1042–7.
29. Ivashkiv LB, Donlin LT. Regulation of type I interferon responses. Nat Rev Immunol. 2009;9:227–38.
30. Martinello M, Hajirazaei A, Gerebely J, Dore B, Matthews GV. Management of acute HCV infection in the era of direct-acting antiviral therapy. Nat Rev Gastroenterol Hepatol. 2018;15:412–24.
1 negatively regulates Toll-like receptor signaling by mediating Mal degradation. Nat Immunol. 2006;7:148–55.

33. Frobose H, Ronn SG, Heding PE, Mendoza D, Cohen P, Mandrup-Poulsen T, Billestrup N. Suppressor of cytokine Signaling-3 inhibits interferleukin-1β signaling by targeting the Traf-6/Tak1 complex. Mol Endocrinol. 2006;20:1587–96.

34. Strebovsky J, Walker P, Lang R, Dalpke AH. Suppressor of cytokine signaling 1 (SOCS1) limits NF-kappaB signaling by decreasing p65 stability within the cell nucleus. Faseb J. 2011;25:863–74.

35. Yu CF, Peng WM, Schlee M, Barchet W, Eis-Hubinger AM, Kolanus W, Frobose H, Ronn SG, Heding PE, Mendoza H, Cohen P, Mandrup-Xie.

36. Yao ZQ, Waggoner SN, Cruise MW, Hall C, Xie X, Oldach DW, Hahn YS. Interaction of cytokine signaling 1 expression in peripheral blood mononuclear cells of hepatitis C genotype 1 patients. J Formos Med Assoc. 2016;115:440–4.

37. Pascarella S, Clement S, Guilloux K, Conzelmann S, Penin F, Negro F. Effects of hepatitis C virus on suppressor of cytokine signaling mRNA levels: comparison between different genotypes and core protein sequence analysis. J Med Virol. 2011;83:1005–15.

38. Ahmed QL, Manzoor S, Tarq M, Khalid M, Ashraf W, Parvaiz F, Imran M. Hepatitis C virus infection in vitro triggers endoplasmic reticulum stress and downregulates insulin receptor substrates 1 and 2 through upregulation of cytokine signaling suppressor 1. Acta Virol. 2014;58:238–44.

39. Kawaguchi T, Yoshida T, Harada M, Hisamoto T, Nagao Y, Ide T, Taniguchi E, Kumemura H, Hanada S, Maeyama M, et al. Hepatitis C virus downregulates insulin receptor substrates 1 and 2 through up-regulation of suppressor of cytokine signaling 3. Am J Pathol. 2004;165:1499–508.

40. Pazienza V, Clement S, Pugnale P, Conzelman S, Foti M, Mangia A, Negro F. The hepatitis C virus core protein of genotypes 3a and 1b downregulates insulin receptor substrate 1 through genotype-specific mechanisms. Hepatology. 2007;45:1164–71.

41. As C S A, S N, M S, J A E, X F, N S I. Hepatitis C virus (HCV)-induced suppression of cytokine signaling (SOCS) regulates proinflammatory TNF-α responses. J Leukoc Biol. 2014;96:255–63.

42. Xu G, Yang F, Ding CL, Wang J, Zhao P, Wang W, Ren H. MiR-221 accentuates IFNα anti-HCV effect by downregulating SOCS1 and SOCS3. Virology. 2014;462–463:343–50.

43. Li A, Song W, Qian J, Li Y, He J, Zhang Q, Li W, Zhai A, Kao W, Hu Y, et al. MiR-122 modulates type I interferon expression through blocking suppression of cytokine signaling 1. Int J Biochem Cell Biol. 2014;35:858–65.

44. Koeberlein B, zur Hausen A, Bektas N, Zentgraf H, Chin R, Nguyen LT, Kandolf R, Torresi J, Bock CT. Hepatitis B virus overexpresses suppressor of cytokine signaling-3 (SOCS3) thereby contributing to severity of inflammation in the liver. Viruses. 2010;1:4851–9.

45. Fletcher SP, Chin DJ, Ji Y, Iniguez AL, Taillon B, Swinnen DC, Ravindran P, Cheng DT, Bitter H, Lopatin U, et al. Transcriptomic analysis of the woodchuck model of chronic hepatitis B. Hepatology. 2012;56:820–30.

46. Wang W, Bian H, Li F, Li X, Zhang D, Sun S, Song S, Zhu Q, Ren W, Qin C, Qi J. HBsAg induces the expression of macrophage miR-155 to accelerate liver injury via promoting production of inflammatory cytokines. Cell Mol Life Sci. 2018;75:2627–41.

47. Xu Y, Hu Y, Shi B, Zhang X, Wang J, Zhang Z, Shen F, Zhang Q, Sun S, Yuan Z. HBsAg inhibits TRAIL-mediated activation and IFN-α production in plasmacytoid dendritic cells. Mol Immunol. 2009;46:2640–6.

48. Tang Y, Chen X, Zhang Y, Tang Z, Zhou M, Li D, Wang P, Zang G, Yu Y. Fusion protein of tapasin and hepatitis B core antigen 1827 enhances T helper cell type 1/2 cytokine ratio and antiviral immunity by inhibiting suppressors of cytokine signaling family members 1/3 in hepatitis B virus transgenic mice. Mol Med Rep. 2014;9:1171–8.

49. Chen X, Liu H, Tang Z, Yu Y, Zang G. The modification of Tapasin enhances hepatitis C T lymphocyte activity of intracellularly delivered CTL epitopes via cytoplasmic transduction peptide. Acta Biochim Biophys Sin (Shanghai). 2013;45:203–12.

50. Chen X, Tang Y, Zhang Y, Zhou M, Tang Z, Yu Y, Zang G. Tapasin modification on the intracellular epitope HBcAg18-27 enhances HBV-specific CTL immune response and inhibits hepatitis B virus replication in vivo. Lab Invest. 2014;94:4788–90.

51. Du LY, Cui XL, Chen EQ, Cheng X, Liu L, Tang H. Correlation between the suppressor of cytokine signaling-1 and 3 and hepatitis B virus: possible roles in the resistance to interferon treatment. Virol J. 2014;11:51.

52. JI Z, Ye F, ZH Zhou Y, CY D, S G, K W. Prognoses of patients with acute-on-chronic hepatitis B liver failure are closely associated with altered SOCS1 mRNA expression and cytokine production following glucocorticoid treatment. Cell Mol Immunol. 2014;11:396–404.

53. Bock CT, Toan NL, Koeberlein B, Le Song H, Chin R, Zentgraf H, Kandolf R, Torresi J. Subcellular mislocalization of mutant hepatitis B X proteins
contributes to modulation of STAT/ SOCS signaling in hepatocellular carcinoma. Intervirology. 2008;51:432–43.

71. Zhang P, Li F, Li N, Zhu Q, Yang C, Han Q, Chen J, Lv Y, Yu L, Wei P, Liu Z. Genetic variations of SOCS1 are associated with chronic hepatitis B virus infection. Hum Immunol. 2014;75:709–14.

72. Hu Z, Yang J, Wu Y, Xiong G, Wang Y, Yang J, Deng L. Polymorphisms in CISH gene are associated with persistent hepatitis B virus infection in Han Chinese population. PLoS ONE. 2014;9:e100826.

73. Fu X, Song X, Li Y, Tan D, Liu G. Hepatitis B virus X protein upregulates DNA methyltransferase 3A/3B and enhances SOCS-1 CpG island methylation. Mol Med Rep. 2016;13:301–8.

74. Teng CF, Chang HY, Tsai HW, Hsieh WC, Kuo YH, Su IJ, Lin YJ. Liver regeneration accelerates hepatitis B virus-related tumorigenesis of hepatocellular carcinoma. Mol Oncol. 2018;12:1175–87.

75. Ou X, Mao S, Cao J, Ma Y, Ma G, Cheng A, Wang M, Zhu D, Chen S, Jia R, et al. The neglected avian hepatotropic virus induces acute and chronic hepatitis in ducks: an alternative model for hepatology. Oncotarget. 2017;8:18398–51.

76. Xie J, Zeng Q, Wang M, Ou X, Ma Y, Cheng A, Zhao X-X, Liu M, Zhu D, Chen S, et al. Transcriptomic characterization of a chicken embryo model infected with duck hepatitis a virus type 1. Front Immunol. 2018;9.

77. Xie J, Wang M, Cheng A, Zhao X-X, Liu M, Zhu D, Chen S, Jia R, Yang Q, Wu Y, et al. DHAV-1 inhibits type I interferon signaling to assist viral adaption by increasing the expression of SOCS3. Front Immunol. 2019;10:731.

78. Waiboci LW, Ahmed CM, Mujtaba MG, Flowers LO, Martin JP, Hader MI, Johnson HM. Both the suppressor of cytokine signaling 1 (SOCS-1) kinase inhibitory region and SOCS-1 mimetic bind to JAK2 auto phosphorylation site: implications for the development of a SOCS-1 antagonist. J Immunol. 2007;178:5058–68.

79. Ahmed CM, Larkin J 3rd, Johnson HM. SOCS1 Mimetics and Antagonists: A Complementary Approach to Positive and Negative Regulation of Immune Function. Front Immunol. 2015;6:183.

80. Pellegrini M, Calzascia T, Toe JG, Preston SP, Lin AE, Elford AR, Shahinian A, Lang PA, Lang KS, Morre M, et al. IL-7 engages multiple mechanisms to overcome chronic viral infection and limit organ pathology. Cell. 2011;144:601–13.

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