Evaluation of Salinity Tolerance of Prairie Junegrass, a Potential Low-maintenance Turfgrass Species

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Abstract. Prairie junegrass (Koeleria macrantha) is a perennial, cool-season, native grass that has shown potential for use as a turfgrass species in the northern Great Plains; however, limited information is available on its salinity tolerance. In this study, salinity tolerance of four junegrass populations from North America (Colorado, Minnesota, Nebraska, and North Dakota) and two improved turf-type cultivars from Europe (‘Barleria’ and ‘Barkoel’) was evaluated and compared with kentucky bluegrass (Poa pratensis), perennial ryegrass (Lolium perenne), sheep fescue (Festuca ovina), hard fescue (F. brevifila), and tall fescue (F. arundinacea). Salinity tolerance was determined based on the predicted salinity level causing 50% reduction of final germination rate (PSLF) and daily germination rate (PSLD) as well as electrolyte leakage (EL), tissue dry weight (DW), and visual quality (VQ) of mature plants. All populations of prairie junegrass showed similar salt tolerance with an average of PSLF and PSLD being 7.1 and 5.3 g L⁻¹ NaCl, respectively, comparable to kentucky bluegrass and hard and sheep fescue but lower than tall fescue and perennial ryegrass. Larger variations were observed in VQ in the junegrasses compared with EL and DW, in which ‘Barleria’ from the European population showed the highest VQ, following two salt-tolerant grasses, tall fescue and sheep fescue. Nebraska population was the least salt-tolerant within the species but still exhibited similar or higher tolerance than kentucky bluegrass and perennial ryegrass cv. Arctic Green. Overall, junegrass was more salt-sensitive during germination but more tolerant to salinity when mature. Salinity tolerance of junegrass may be further improved through turfgrass breeding because salinity tolerance varied in different populations.

Prairie junegrass or junegrass (Koeleria macrantha), a cool-season, perennial, short-grass prairie species, is native to North America (Robertson, 1974). It possesses many characteristics ideal for low-maintenance turfgrass use such as a slow vertical growth rate (Dixon, 2000; Sovali and Bender, 2006), moderate drought tolerance (McKernan et al., 2001), and adaptation to dry and sandy soil (Pammell et al., 1901–1904). Mintenko et al. (2002) evaluated 28 native grasses, including an improved turf-type prairie junegrass (‘Barkoel’) from Europe and three prairie junegrass populations collected from Alberta, Iran, and Minnesota. ‘Barkoel’ junegrass yielded some of the highest quality ratings and the populations from Alberta and Minnesota were among the grasses with the earliest spring green-up in the study. Clark and Watkins (2010b) reported that 30 of 48 prairie junegrass accessions provided adequate turf quality (5.0 or greater) after a 3-year mowed spaced plant field evaluation under low-input conditions in St. Paul, MN. Furthermore, prairie junegrass has the potential to produce sufficient seeds to be economically available for the turf industry (Clark and Watkins, 2010a), a major limiting factor causing reduced application of buffalograss (Buchloe dactyloides), another native grass with similar ideal characteristics of xericaphytic turf (Riordan and Browning, 2003).

Low-input turfgrass must meet visual and functional requirement under minimum care. Salinity is a major stress that reduces turf appearance and inhibits plant growth in the Great Plains. For instance, high soil sodium and salinity affects 1,900,000 and 700,000 acres of land in North Dakota, respectively (Seelig, 2000). One of the most common methods to address salt damage is use of species and cultivars that exhibit salinity tolerance. Mintenko and Smith (2001) reported prairie junegrass was tolerant to moderate soil salinity levels (4 to 8 ds m⁻¹) at maturity but more sensitive to salinity during germination; however, ‘Barkoel’ was the only cultivar evaluated in their study. Large variations have been reported in the characteristics of seed production and turf quality in prairie junegrass germplasm (Clark and Watkins, 2010a, 2010b). Therefore, the objective of this study was to evaluate relative salinity tolerance of four native prairie junegrass populations (Colorado, Minnesota, Nebraska, and North Dakota) along with the European cvs. Barleria and Barkoel and then compare their performance with several commonly used cool-season turfgrass species. These results can then be used by turfgrass breeders in germplasm improvement programs aimed at developing salt-tolerant low-input turfgrass cultivars.

Materials and Methods

Plant material. Five populations of junegrass, one tall fescue, one sheep fescue, one perennial ryegrass, and two kentucky bluegrasses were used in this study (Table 1). The North American junegrass germplasm was collected from Colorado (Weld county), Minnesota (Wabasha county), Nebraska (Sioux and Dawes counties), and North Dakota (Billings county) as seed from native stands in either 2005 or 2006. Plants were grown from seed and placed into a breeding nursery after the collection. Each breeding nursery contained only germplasm collected from one state to avoid outcrossing with germplasm from different collection areas. Seed from these nurseries was harvested and bulked and labeled as Colorado, Minnesota, and Nebraska populations. Material representing the North Dakota population was harvested from a single plant originated from seed collected in Billings county, ND. This single plant was representative of the entire population because the population exhibited uniformity in both unmowed nurseries and mowed turf evaluations. Seeds of ‘Barkoel’ and ‘Barleria’, the European population, were provided by Barenbrug USA.

The sheep fescue used in this study is a selection developed at the University of Minnesota for use on roadsides in cold climates where salinity stress is problematic. The hard fescue is an advanced population from the University of Minnesota breeding program that has been selected for use as low-input turf in Minnesota; however, it has not been screened for salinity tolerance. ‘Arctic Green’ perennial ryegrass was released by the University of Minnesota in 2008 for improved winter-hardiness and crown rust resistance. Other cultivars and selections were identified based on their performance (i.e., acceptable quality under low-maintenance management) in turfgrass trials in Minnesota and/or North Dakota.

Seed germination. Seeds of the grasses were surface-sterilized following the method of Wang and Zhang (2010). Seeds were submerged in 70% ethanol for 1 min followed by submergence in 2% sodium hypochlorite solution...

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for 20 min. Seeds were then rinsed three times with deionized/distilled water (ddH₂O) and air-dried in a laminar-flow hood. Thirty-six seeds of each grass were placed in a 100 × 15-mm petri dish containing 20 mL of 1% agar salinized with 0, 5, 10, 15, or 20 g L⁻¹ NaCl and the electrical conductivity (EC) levels of the salt solutions were 0.0, 8.6, 15.3, 20.4, and 25.7 dS m⁻¹, respectively, when measured with an EC meter (Model 1054; VWR International LLC, West Chester, PA). Petri dishes were placed in an incubator (Model 1-35 VL; Percival Scientific, Perry, IA) at 20/15 °C (day/night) under fluorescent light (36 μmol m⁻² s⁻¹) with a 15/9-h (light/dark) photoperiod for 30 d (Mintenko and Smith, 2001).

This experiment was conducted in a completely randomized design consisting of three replications of a 12 (grass) × 5 (salinity level) factorial arrangement. The number of seeds germinating per dish was recorded three times a week. Seed germination was defined as an emerged shoot visible under 2× magnification (McCarty and Dudeck, 1993). Final germination rate (FGR) and daily germination rate (DGR) were calculated using FGR(%) = [100 × (%/m)/36] and DGR(%) = [100 × (n/D)/36], where n was the number of new seeds germinated at each counting and D was the number of days accumulated up to that counting (Wang and Zhang, 2010). To avoid the differences that may be attributed by seed size or seedling vigor, FGR was standardized by calculating as a percentage of the control value (Horst and Dunning, 1989). The experiment was conducted from 29 Jan. to 1 Mar. (Study I) and repeated from 9 Apr. to 10 May 2010 (Study II). Final germination rate and DGR were transformed using arcsine and square root, respectively, before the analysis of variance. All data were subjected to PROC GLM (SAS, 2004) and means were separated by calculating as a percentage of the control size or seedling vigor, FGR was standardized by the number of days accumulated up to that counting (McCarty and Dudeck, 1993). Final germination rate was observed between each soaking. No additional irrigation was applied to the plants and no drought stress symptom (i.e., leaf wilting) was observed between each soaking.

The experiment was arranged in a split-plot design with salinity level being the whole-plot factor (arranged in a randomized complete block design with three replicates) and grass being the subplot factor. Data were collected on tissue EL, DW, and VQ at 2 and 5 weeks after treatment (WAT). Electrolyte leakage was measured following the method of Su et al. (2007). Briefly, five fully expanded leaves were collected from each container and rinsed three times with ddH₂O. The leaves were then cut into ≈2-cm segments and placed in a test tube containing 20 mL ddH₂O. Conductivity (C₁) was measured with an EC meter after the test tubes were agitated on a shaker (Model 6010; Eberbach Corp., Ann Arbor, MI) for 24 h. The leaf samples were then autoclaved at 121 °C for 20 min and the test tubes were agitated for another 24 h followed by a second conductivity measurement (C₂). Electrolyte leakage (%) was calculated as EL = C₁/C₂ × 100 with lower EL indicating a higher tolerance to stress. Tissue DW was recorded after the clippings were oven-dried at 60 °C for 48 h. Tissue EL and DW of each grass under saline conditions were expressed as a percentage of the control value of the same grass to minimize the difference attributed to variable growing habits in different species (Horst and Beadle, 1984). Visual quality was evaluated with a 1 to 9 scale, in which 1 = dead grass, 5 = acceptable, and 9 = best quality based on color, texture, and uniformity (Emmons, 2000). The experiment was conducted from 10 Mar. to 14 Apr. (Study I) and repeated from 17 May to 21 June 2010 (Study II). Salinity levels were reduced to 0, 3, 6, 9, or 12 g L⁻¹ of NaCl (EC = 1.3, 6.5, 10.7, 14.8, or 18.5 dS m⁻¹, respectively) in Study II as a result of a rapid DW decline at the highest saline level (20 g L⁻¹) observed during Study I. Data were subjected to PROC GLM (SAS, 2004) and means were separated with Fisher’s protected least significant difference at P ≤ 0.05. PROC CORR was applied to evaluate the relationships among EL, DW, and VQ.

### Results

#### Seed germination.

Statistical analysis showed that Studies I and II were homogeneous (data not shown); thus, the results were combined before the analyses of variance and regression. Salinity had an adverse effect on FGR and DGR when data were pooled across grasses (Fig. 1). Final germination rate was reduced from 100% in the control treatment to 76%, 25%, 11%, and 3% as salinity increased from 5 to 10, 15, and 20 g L⁻¹ NaCl, respectively (Fig. 1A). Similarly,

### Vegetative growth.

Thirty plugs (5 × 5 cm) of each grass initiated from a single seed were transplanted into D20 deepots (8 cm-diam., 20 cm-deep; Stuewe and Sons, Inc., Tangent, OR) with Miracle-Gro potting mix (Scotts Company LLC, Marysville, OH) on 15 July 2009. Grasses were bowed at 5 cm once a week and hand-watered every other day until water drained freely from the bottom. Fertilizer (13.0N–0.0P–22.0K; Anderson Golf Products, Maumee, OH) was applied at 12.5 kg ha⁻¹ N once each month. Grasses were exposed to saline conditions after a 3-month establishment period in the greenhouse. Saline conditions were comprised of NaCl at 0.5, 10, 15, or 20 g L⁻¹ dissolved in tap water with EC at 1.3, 9.8, 16.3, 22.3, or 27.5 dS m⁻¹, respectively. Grasses were soaked in the salt solution one time each week in a 30-L tank (59 × 47 × 39 cm) for 2 h and then drained freely. The tank was refilled with the fresh solution of the same concentration after each soaking. No additional irrigation was applied to the plants and no drought stress symptom was observed between each soaking.

The experiment was arranged in a split-plot design with salinity level being the whole-plot factor (arranged in a randomized complete block design with three replicates) and grass being the subplot factor. Data were collected on tissue EL, DW, and VQ at 2 and 5 weeks after treatment (WAT). Electrolyte leakage was measured following the method of Su et al. (2007). Briefly, five fully expanded leaves were collected from each container and rinsed three times with ddH₂O. The leaves were then cut into ≈2-cm segments and placed in a test tube containing 20 mL ddH₂O. Conductivity (C₁) was measured with an EC meter after the test tubes were agitated on a shaker (Model 6010; Eberbach Corp., Ann Arbor, MI) for 24 h. The leaf samples were then autoclaved at 121 °C for 20 min and the test tubes were agitated for another 24 h followed by a second conductivity measurement (C₂). Electrolyte leakage (%) was calculated as EL = C₁/C₂ × 100 with lower EL indicating a higher tolerance to stress. Tissue DW was recorded after the clippings were oven-dried at 60 °C for 48 h. Tissue EL and DW of each grass under saline conditions were expressed as a percentage of the control value of the same grass to minimize the difference attributed to variable growing habits in different species (Horst and Beadle, 1984). Visual quality was evaluated with a 1 to 9 scale, in which 1 = dead grass, 5 = acceptable, and 9 = best quality based on color, texture, and uniformity (Emmons, 2000). The experiment was conducted from 10 Mar. to 14 Apr. (Study I) and repeated from 17 May to 21 June 2010 (Study II). Salinity levels were reduced to 0, 3, 6, 9, or 12 g L⁻¹ of NaCl (EC = 1.3, 6.5, 10.7, 14.8, or 18.5 dS m⁻¹, respectively) in Study II as a result of a rapid DW decline at the highest saline level (20 g L⁻¹) observed during Study I. Data were subjected to PROC GLM (SAS, 2004) and means were separated with Fisher’s protected least significant difference at P ≤ 0.05. PROC CORR was applied to evaluate the relationships among EL, DW, and VQ.

### Table 1. Predicted salinity levels (g L⁻¹ NaCl) causing 50% reduction in final germination rate and daily germination rate in prairie junegrass and other cool-season turfgrasses.

| Grass              | PSLF(3) (g L⁻¹ NaCl) | PSLD (g L⁻¹ NaCl) |
|--------------------|----------------------|-------------------|
| Tall fescue—‘Falcon IV’ | 14.7 a'              | 10.8 a            |
| Perennial ryegrass—‘Arctic Green’ | 13.3 a              | 9.8 a             |
| Hard fescue—‘MN-HD1’ | 8.9 b                | 6.2 b             |
| Prairie junegrass—‘Barkoel’ | 8.6 bc               | 5.2 bc            |
| Prairie junegrass—‘Barlera’ | 7.9 bcd              | 6.6 b             |
| Kentucky bluegrass—‘Langara’ | 7.6 bc              | 5.4 bc            |
| Prairie junegrass—North Dakota | 7.6 bcd      | 4.2 e             |
| Sheep fescue—‘67135’ | 7.6 bcd              | 4.2 e             |
| Kentucky bluegrass—‘Park’ | 7.2 bcd              | 5.1 b             |
| Prairie junegrass—Minnesota | 6.6 cde         | 5.3 b             |
| Prairie junegrass—Nebraska | 6.6 de              | 4.9 b             |
| Prairie junegrass—Colorado | 5.5 bc              | 5.5 b             |

1PSLF = the predicted salinity level causing 50% reduction of final germination rate; PSLD = the predicted salinity level causing 50% reduction of daily germination rate.
2Means in each column followed by the same letter are not significantly different at the P ≤ 0.05 level.
DGR was reduced from 9.5%/d to 5.4 (43% reduction), 1.9 (80%), 0.7 (93%), and 0.1%/d (99%) at the same saline levels (Fig. 1B).

Tall fescue and perennial ryegrass had the highest PSLF when data were pooled across saline levels (14.7 and 13.3 g L⁻¹ NaCl, respectively), whereas other grasses required lower saline levels, ranging from 8.9 to 5.5 g L⁻¹ NaCl to cause 50% reduction in FGR (Table 1). Within prairie junegrasses, ‘Barkoel’ had the highest PSLF (8.6 g L⁻¹), 36% higher than that of Colorado population (5.5 g L⁻¹) (Table 1). Regression analysis showed that tall fescue and perennial ryegrass required similar saline levels to reduce DGR by 50%, indicating their similar tolerance to salt stress (Table 1). An average saline level of 5.3 g L⁻¹ NaCl was required to cause 50% reduction in DGR in other grasses, which was 51% or 46% lower than that of tall fescue or perennial ryegrass, respectively (Table 1). Similarly, significant differences in PSLF were observed between prairie junegrass entries with ‘Barleria’ and the population of North Dakota being the most salt-tolerant and salt-sensitive, respectively (Table 1).

**Vegetative growth.** The results of the two studies are presented separately as salt levels varied. Significant differences in EL, DW, and VQ of both grasses and salinity levels were observed. However, the interaction between grass and saline level was only significant for VQ in both studies (data not shown).

**Leaf electrolyte leakage.** Electrolyte leakage significantly increased by 61%, 160%, 200%, and 291% when salinity raised from 0 to 5, 10, 15, and 20 g L⁻¹, respectively, at 2 WAT in Study I (Fig. 2I). By 5 WAT, EL in the grasses exposed to the highest saline level (20 g L⁻¹) reached 1119% (Fig. 2I). Similarly, EL was elevated as the saline level increased and saline stress progressed in Study II (Fig. 2IV). Among all of the grasses, tall fescue had the lowest EL in both studies, except 5 WAT in Study I (Table 2), indicating its higher salt tolerance. In Study I, junegrass population of Minnesota was the most salt-sensitive grass with the highest EL at 2 and 5 WAT, whereas ‘Barleria’ junegrass showed significantly high EL at 5 WAT (Table 2). In Study II, ‘Park’ kentucky bluegrass and ‘Barkoel’ prairie junegrass had the highest EL at 2 and 5 WAT, respectively (Table 2).

**Leaf dry weight.** Leaf DW was significantly reduced when the saline level was higher than 5 and 6 g L⁻¹ NaCl in Studies I and II, respectively, at 2 WAT when data were pooled across the grasses (Fig. 2II, V). At 5 WAT, DW in all treatments, excluding the 5 g L⁻¹ NaCl treatment in Study I and 3 g L⁻¹ in Study II, dropped to less than 50% of the control. Tissue DW was reduced to less than 10% of the control at 5 WAT when the saline was higher than 9 to 10 g L⁻¹ (Fig. 2II, V). Significant differences in DW were observed in the grasses in Study I ‘Barleria’ and ‘Barkoel’, the two European prairie junegrass cultivars, had the highest DW at 2 WAT followed by sheep and hard fescue (Table 3). However, hard fescue had the highest DW by 5 WAT in Study I and it was the only grass with DW higher than 50% of control. In Study II, no significant difference in DW of all grasses at both 2 and 5 WAT was observed (Table 3), which may be a result of the reduced salinity levels applied.

**Fig. 2.** Salinity effects on electrolyte leakage (%), tissue dry weight (%), and visual quality across all grasses in Study I (I–III) and Study II (IV–VI). Electrolyte leakage and tissue dry weight under saline conditions were expressed as a percentage of the control (NaCl = 0 g L⁻¹). Upper and lower case letters represented significant differences at $P \leq 0.05$ level for 2 and 5 weeks after treatment (WAT), respectively.

**Table 2.** Electrolyte leakage in prairie junegrass and other cool-season turfgrasses across all salinity levels at 2 and 5 weeks after treatment (WAT).

| Grass                        | Study I   | Study II  |
|------------------------------|-----------|-----------|
|                              | 2 WAT     | 5 WAT     | 2 WAT     | 5 WAT     |
| Tall fescue—‘Falcon IV’      | 165 c d   | 776 abc   | 322 d     | 396 c     |
| Hard fescue—‘MN-HD1’         | 187 c     | 770 abc   | 317 d     | 660 b     |
| Sheep fescue—‘67135’         | 206 bc    | 547 c     | 365 cd    | 419 bc    |
| Prairie junegrass—‘Barleria’ | 212 b     | 864 a     | 364 cd    | 399 bc    |
| Prairie junegrass—‘Barkoel’  | 230 b     | 606 bc    | 431 bcd   | 961 a     |
| Kentucky bluegrass—‘Lanagara’| 234 bc    | 834 ab    | 366 cd    | 639 b     |
| Prairie junegrass—Colorado   | 242 bc    | 688 abc   | 425 bcd   | 677 b     |
| Kentucky bluegrass—‘Park’    | 243 bc    | 614 bc    | 755 a     | 737 ab    |
| Prairie junegrass—North Dakota| 252 bc   | 784 ab    | 358 ed    | 591 bc    |
| Perennial ryegrass—‘Arctic Green’ | 253 bc | 674 abc   | 544 b     | 597 bc    |
| Prairie junegrass—Nebraska   | 295 ab    | 832 ab    | 519 bc    | 767 ab    |
| Prairie junegrass—Minnesota  | 387 a     | 898 a     | 363 cd    | 586 bc    |

*Means in each column followed by the same letter are not significantly different at the $P \leq 0.05$ level.
*Saline levels were 0, 5, 10, 15, and 20 NaCl g L⁻¹ in Study I and 0, 3, 6, 9, and 12 NaCl g L⁻¹ in Study II.
*Data under saline conditions were expressed as a percentage of the control (NaCl = 0 g L⁻¹).
Visual quality. When data were pooled across the grasses, no adequate quality (quality 5.0 or greater) was observed at salinity levels higher than 5 and 6 g L⁻¹ at 2 WAT in Study I and Study II, respectively (Fig. 2I, VI). All grasses under saline conditions showed a poor appearance by 5 WAT in both studies (Fig. 2II, VI). Tall fescue, sheep fescue, hard fescue, and ‘Barleria’ prairie junegrass showed acceptable performance (i.e., the grass materials were able to survive and provide acceptable quality under regular mowing conditions) at 2 WAT in Study I when data were pooled across salinity levels (Table 4). As saline conditions extended, all grasses in Study I exhibited VQ ratings less than 5.0 at 5 WAT. Seven grasses had acceptable quality at 2 WAT in Study II compared with four grasses in Study I, which may be the result of the reduced saline levels in Study II. However, similar to Study I, no grasses showed adequate appearance (quality greater than 5.0) at 5 WAT in Study II. Significant interactions between salinity level and grass entry were observed in VQ on all sampling dates, except 2 WAT in Study I (Fig. 3).

Discussion

Salinity reduced FGR and DGR; however, its impact is greater on DGR than on FGR because higher reduction rate was observed on DGR at low saline levels (Fig. 1). Furthermore, PSLD of all grasses was lower than PSLF, except of prairie junegrass–Colorado (Table 1). Similar results were reported by Dai et al. (2009) and Wang and Zhang (2010). Therefore, DGR and PSLD can be used as more sensitive indicators of relative saline tolerance during germination than FGR and PSLF with higher DGR and PSLD suggesting higher tolerance.

Electrolyte leakage (an indicator of cell membrane stability) and tissue DW (or yield) (an indicator of overall vigor) have been widely used for screening tolerance of drought-related stresses in plants, including sorghum (Sorghum bicolor) (Sullivan and Ross, 1979), wheat (Triticum aestivum) (Blum and Ebrecon, 1981), tomato (Lycopersicon esculentum) (Chen et al., 1982), and turfgrass (Marcum, 1998; Su et al., 2007). Under salinity stress, plants are not only exposed to ion toxicity and imbalance, but also water deficiencies that are the most common and serious consequences of salt stress (Murns, 2002; Tabaei-Aghdaei et al., 2000). Marcum (2001) evaluated the salinity tolerance of 35 bentgrass cultivars and reported that tissue dry weight (root or shoot) was an effective parameter for predicting salinity tolerance along with percentage of green leaf area and root length. Farooq and Azam (2006) reported significant correlations between EL and salt stress injury (relative water content, Na⁺ and K⁺ leaf content, and yield) in wheat; thus, EL is also a reliable technique for screening salt tolerance. Turfgrass with high stress tolerance must demonstrate not only functional quality, but also visual quality. Mintenko and Smith (2001) suggested that visual quality was more

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### Table 3. Tissue dry weight in prairie junegrass and other cool-season turfgrasses across all salinity levels at 2 and 5 weeks after treatment (WAT).

| Grass                  | Study I | Study II |
|------------------------|---------|----------|
|                        | 2 WAT   | 5 WAT    | 2 WAT   | 5 WAT    |
| Prairie junegrass—Barleria' | 97 a ² 37 b | 139 a 56 a |
| Prairie junegrass—Barkoel' | 95 a 27 bc | 96 a 35 a |
| Sheep fescue—‘67135’ | 86 ab 59 a | 66 a 66 a |
| Hard fescue—‘MN-HD1’ | 75 abc 35 b | 71 a 47 a |
| Kentucky bluegrass—‘Langara’ | 66 bcd 32 bc | 74 a 37 a |
| Perennial ryegrass—‘Arctic Green’ | 61 b cde 31 bc | 83 a 44 a |
| Kentucky bluegrass—‘Park’ | 56 cde 34 bc | 92 a 55 a |
| Tall fescue—‘Falcon IV’ | 54 cde 34 bc | 73 a 53 a |
| Prairie junegrass—Nebraska | 47 cde 23 bc | 60 a 34 a |
| Prairie junegrass—Colorado | 45 de 28 bc | 66 a 36 a |
| Prairie junegrass—Minnesota | 42 de 25 bc | 64 a 34 a |
| Prairie junegrass—North Dakota | 33 e 20 e | 49 a 45 a |

²Saline levels were 0, 5, 10, 15, and 20 NaCl g L⁻¹ in Study I and 0, 3, 6, 9, and 12 NaCl g L⁻¹ in Study II.
³Means in each column followed by the same letter are not significantly different at the P ≤ 0.05 level.

### Table 4. Visual quality in prairie junegrass and other cool-season turfgrasses across all salinity levels at 2 and 5 weeks after treatment (WAT).

| Grass                  | Study I | Study II |
|------------------------|---------|----------|
|                        | 2 WAT   | 5 WAT    | 2 WAT   | 5 WAT    |
| Tall fescue—‘Falcon IV’ | 5.73 a 3.60 a | 6.33 a 4.60 a |
| Sheep fescue—‘67135’ | 5.67 a 2.87 bc | 5.40 c 3.73 b |
| Hard fescue—‘MN-HD1’ | 5.60 a 3.00 b | 4.47 ef g 2.67 ef |
| Prairie junegrass—‘Barleria’ | 5.40 ab 2.93 bc | 5.67 b 3.27 cd |
| Prairie junegrass—‘Barkoel’ | 4.87 bcd 2.53 cd | 5.67 b 3.47 bc |
| Prairie junegrass—Colorado | 4.93 bc 2.53 cd | 5.67 bc 3.07 cde |
| Prairie junegrass—North Dakota | 4.73 cd 2.07 e | 5.08 cd 3.01 cde |
| Prairie junegrass—Nebraska | 4.67 cd 2.27 de | 4.93 def 2.80 ef |
| Prairie junegrass—Minnesota | 4.40 cd 2.13 de | 6.40 a 3.33 e |
| Perennial ryegrass—‘Arctic Green’ | 4.27 de 2.07 e | 3.93 g 2.53 f |
| Kentucky bluegrass—‘Park’ | 3.67 ef 2.07 e | 4.40 fg 2.87 def |
| Kentucky bluegrass—‘Langara’ | 3.53 f 2.07 e | 4.00 g 2.80 ef |

²Saline levels were 0, 5, 10, 15, and 20 NaCl g L⁻¹ in Study I and 0, 3, 6, 9, and 12 NaCl g L⁻¹ in Study II.
³Means in each column followed by the same letter are not significantly different at the P ≤ 0.05 level.

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Fig. 3. Visual quality of six representative grasses as influenced by salinity at 2 and 5 weeks after treatment (WAT) in Study I (A–C) and Study II (D–F). The selected grasses included ‘Falcon IV’ tall fescue (TF), ‘67135’ sheep fescue (SF), ‘Barleria’ prairie junegrass (PJB), Nebraska prairie junegrass population (PJN), ‘Park’ kentucky bluegrass (KBP), and ‘Arctic Green’ perennial ryegrass (PR). Vertical bars represent least significant difference (LSD) at the P ≤ 0.05 level.
important than other characteristics such as yield resulting from the aesthetic function of turfgrass. Regrowth rates only need to be adequate rather than optimal levels under stress conditions. In their evaluation, alkali-grass (*Puccinellia nuttalliana*), blue grama (*Bouteloua gracilis*), Idaho bentgrass (*Agrostis idahoensis*), and *Barkoel* prairie junegrass all showed reduced growth under severe soil salinity, but alkali-grass was the only grass with adequate quality, indicating relative higher saline tolerance.

In the present study, EL increased as salinity levels increased in the reverse relationship of DW and VQ under saline condition (Fig. 2). Similar to previous studies, EL, DW, and VQ were significantly correlated (Table 5), suggesting that all aforementioned parameters can be used to quantify relative salinity tolerance. However, use of a single parameter may not be sufficient. For example, DW was similar in tall fescue and junegrass–Nebraska populations (Table 3), whereas VQ was significantly higher in tall fescue than in *Nebraska* junegrass (Table 4) when data were pooled across saline levels. Similarly, Almansouri et al. (1999) reported no significant differences in relative water content and shoot DW between ‘Belikh’ (salt-resistant) and ‘Cando’ (salt-sensitive) durum wheat (*T. durum*) under saline conditions, but ‘Belikh’ showed a higher K/Na ratio. No grass showed superior salinity tolerance in EL, DW, and VQ when data were pooled across saline levels (Tables 2–4), which might be the result of the high salinity levels we applied, especially in Study I (up to 20 g L\(^{-1}\) NaCl). Lunt et al. (1961) reported 50% of shoot reduction in tall fescue cv. Alta at 160 mEq L\(^{-1}\) (water EC = 14 dS m\(^{-1}\)), similar to the moderate saline level (15 g L\(^{-1}\) NaCl) in Study I. The highest saline level (20 g L\(^{-1}\) NaCl) applied in Study I might have caused severe stress on all grasses, including relatively saline-tolerant ones such as tall fescue. As salinity levels were reduced in Study II, relative salinity tolerance in the grasses were further separated, in which tall fescue and kentucky bluegrass were consistently ranked salt-tolerant and salt-sensitive in EL and VQ (Tables 2 and 4) as shown in other studies (Harivandi et al., 1992; Marcum, 2007). For example, tall fescue provided acceptable quality (greater than 5.0) up to 12 g L\(^{-1}\) NaCl by 2 WAT and 6 g L\(^{-1}\) NaCl by 5 WAT (Fig. 3C–D). In contrast, poor appearance of kentucky bluegrass (VQ less than 5.0) was observed at saline levels equal to or higher than 9 g L\(^{-1}\) NaCl at 2 WAT (Fig. 3C); by 5 WAT, VQ decreased to 3.3 at 3 g L\(^{-1}\) NaCl (Fig. 3D). Almansouri et al. (1999) suggested that quantified parameters and final salinity level, besides genotype and environmental conditions, may affect the ranking of relative salinity tolerance in plants.

‘Arctic Green’ perennial ryegrass showed high salinity tolerance, similar to tall fescue, during germination in the present study (Table 1). However, its relative salt tolerance was comparable to kentucky bluegrass at vegetative growth, contradictory to previous findings that perennial ryegrass was more salt-tolerant (Harivandi et al., 1992; Marcum, 1992; Marcus, 1997). Marcar (1987) reported that relative salinity tolerance of Wimmera (*L. rigidum*), Italian (*L. multiflorum*), and perennial ryegrass ranked differently for germinating and mature plants. Differences in perennial ryegrass and kentucky bluegrass cultivars in the studies may also contribute to the inconsistent results, because large variations of salinity tolerance exist in perennial ryegrass and kentucky bluegrass cultivars (Marcum, 2007). Greub et al. (1985) reported that perennial ryegrass cvs. Common and NK 200 and kentucky bluegrass cvs. Park, Pennstar, and Nugget showed similar VQ under saline condition, although the ryegrasses had higher DW than the bluegrasses.

Limited information is available on salinity tolerance on sheep and hard fescue. Sheep fescue cv. 67135 and hard fescue cv. MN-HD1 evaluated in the present study were developed at the University of Minnesota. Although no testing has confirmed its tolerance to salinity stress, ‘67135’ was originally developed for use on roadsides in northern climates where road salt is a source of physiological stress on turf. Sheep fescue ‘67135’ showed similar EL, DW, and VQ as tall fescue; however, sheep fescue did not show acceptable VQ under saline conditions at 5 WAT in Study II, whereas tall fescue had acceptable quality at saline levels up to 6 g L\(^{-1}\) (Tables 2–4; Fig. 3). Similarly, ‘MN-HD1’ hard fescue showed a similar salinity tolerance as tall fescue in Study I; however, its VQ was inadequate in Study II (Table 4).

The present study was the first attempt to evaluate the salt tolerance of different populations in prairie junegrass. The results showed that prairie junegrass required average salinity levels of 7.1 and 5.3 g L\(^{-1}\) NaCl to cause 50% reduction of FGR and DGR, similar to that of kentucky bluegrass (Table 1). Within junegrasses, ‘Barleria’ was the most salt-tolerant and ‘North Dakota’ was the most salt-sensitive based on PSLF. Limited variations were observed in all the grasses in vegetative growth except VQ (Tables 2–4; Fig. 3). ‘Barleria’ junegrass along with tall fescue, sheep fescue, and hard fescue provided acceptable quality at 2 WAT in Study I.

As salinity levels reduced in Study II, all junegrasses except the Nebraska population had adequate quality (greater than 5.0) at 2 WAT, similar to tall fescue and sheep fescue but higher than kentucky bluegrass and perennial ryegrass (Table 4). Furthermore, ‘Barleria’ junegrass exhibited high VQ (greater than 5.0) under up to 12 g L\(^{-1}\) NaCl at 2 WAT in Study II; so did tall fescue and sheep fescue (Fig. 3C). The decrease of VQ in ‘Barleria’ junegrass was slower and more gradual than in other prairie junegrass, kentucky bluegrass, and perennial ryegrass as saline exposure extended, although VQ in ‘Barleria’ junegrass was unacceptable (less than 5.0) at all saline levels at 5 WAT (Fig. 3D). Mintenok and Smith (2001) reported that ‘Barkoel’ prairie junegrass was tolerant to slight to moderate saline levels (soil EC = 4 to 8 dS m\(^{-1}\)), slightly lower than the susceptibility of tall fescue (soil EC = 6 to 10 dS m\(^{-1}\)) (Harivandi et al., 1992). The results of this research indicated that salinity tolerance of some junegrass populations increased from germination to the mature stage and its salt tolerance may be further improved through turfgrass breeding because salinity tolerance varied in different junegrass populations.

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**Table 5. Correlation coefficients and associated probability levels for electrolyte leakage (EL), tissue dry weight (DW), and visual quality (VQ) across grasses, saline levels, and the duration of saline exposure in Studies I and II.**

|        | Study I     | Study II    |
|--------|-------------|-------------|
|        | Viable DW   | EL          | DW          | EL          |
| EL     | 0.60        | <0.0001     | -0.47       | <0.0001     |
| VQ     | 0.68        | <0.0001     | 0.57        | <0.0001     |

*Saline levels were 0, 5, 10, 15, and 20 NaCl g L\(^{-1}\) in Study I and 0, 3, 6, 9, and 12 NaCl g L\(^{-1}\) in Study II.*

*Data in EL and DW under saline conditions were expressed as a percentage of the control (NaCl = 0 g L\(^{-1}\)).*
