A transmigratory cup in leukocyte diapedesis both through individual vascular endothelial cells and between them

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The basic route and mechanisms for leukocyte migration across the endothelium remain poorly defined. We provide definitive evidence for transcellular (i.e., through individual endothelial cells) diapedesis in vitro and demonstrate that virtually all, both para- and transcellular, diapedesis occurs in the context of a novel “cuplike” transmigratory structure. This endothelial structure was comprised of highly intercellular adhesion molecule-1– and vascular cell adhesion molecule-1–enriched vertical microvilli-like projections that surrounded transmigrating leukocytes and drove redistribution of their integrins into linear tracks oriented parallel to the direction of diapedesis. Disruption of projections was highly correlated with inhibition of transmigration. These findings suggest a novel mechanism, the “transmigratory cup”, by which the endothelium provides directional guidance to leukocytes for extravasation.

Introduction

Extravasation of blood leukocytes is critical for immune surveillance and a crucial first step in the development of inflammation and atherosclerosis (Springer, 1994; Ross, 1999). Two well-characterized processes, selectin-mediated rolling and integrin-mediated firm adhesion, cooperate to promote accumulation of leukocytes on the luminal surface of the vascular endothelium (Luscinskas et al., 1994; Springer, 1994). Subsequently, leukocytes actively migrate, in amoeboid fashion, across the endothelial monolayer into the interstitium, a process referred to as transendothelial migration (TEM) or diapedesis (Luscinskas et al., 1994; Springer, 1994). A variety of adhesion molecules have been identified that are important for this process. These include on leukocytes the integrins leukocyte function-associated molecule-1 (LFA-1) (αLβ2), Mac-1 (αMβ2), and very late antigen-4 (VLA-4) (α4β1), and on endothelium the immunoglobulin superfamily members intercellular adhesion molecules 1 and 2 (ICAM-1 and ICAM-2), vascular cell adhesion molecule-1 (VCAM-1), platelet/endothelial cell adhesion molecule-1 (PECAM-1), and the junctional adhesion molecules, as well as the proline-rich glycoprotein CD99 (Oppenheimer-Marks et al., 1991; Luscinskas et al., 1994; Greenwood et al., 1995; Muller, 2001; Aurrand-Lions et al., 2002). However, the central issues of how the proper leukocyte directionality to cross the endothelium is established, and the nature of the route of transendothelial passage, remain unclear and controversial (Kvietys and Sandig, 2001; Muller, 2001).

Shortly after arrest, most leukocytes spread and begin to migrate laterally over the apical surface of the endothelium (Luu et al., 1999). At some point, usually within several minutes (Luu et al., 1999), leukocytes make the critical “decision” to migrate in the direction perpendicular to the plane of the endothelium and extravasate. It has been suggested that shear forces provided by the luminal circulation may help to establish this directionality (Cinamon et al., 2001), though robust diapedesis is also observed in many systems in the absence of shear. In addition, junctionally localized gradients of chemoattractants and adhesion molecules, such as PECAM-1, CD99, and the junctional adhesion molecules, have been proposed to direct leukocytes to endothelial junctions and to provide the requisite traction to drive diapedesis at these locations (Bianchi et al., 1997; Muller, 2001, 2003; Aurrand-Lions et al., 2002). Furthermore, intra-endothelial cell signaling events are thought to facilitate leukocyte passage by transiently and locally down-regulating the integrity of the endothelial adherence junctions (Huang et al., 1993; Bianchi et al., 1997; Adamson et al., 1999; Etienne-
structures" that were predicted to be inhibitory for diapedesis injections were described explicitly as firm adhesion "docking of these studies was any relationship between projections and round a portion of the adherent leukocytes. However, in neither VCAM-1–enriched upright microvilli-like projections that sur- reiro et al., 2002; Carman et al., 2003) have demonstrated that, et al., 2000; Strey et al., 2002). Recently, we and others (Bar- occhi et al., 1997; Muller, 2001, 2003; Aur- and-Lions, 2002). In the absence of conclusive observa- tions made in the widely used in vitro cultured endothelial mono- layer systems, the idea of nonjunctional TEM directly through individual endothelial cells, i.e., the transcellular route of TEM, has remained largely disregarded (Kvietys and Sandig, 2001; Muller, 2001). However, some of the very first EM studies to ad- dress the nature of TEM in vivo provided evidence for the pre- dominance of the transcellular route of diapedesis (Williamson and Grisham, 1961; Marchesi and Gowans, 1964). Conclusive proof for transcellular migration in vivo has been provided by both serial-section transmission (Marchesi and Gowans, 1964; Cho and De Bruyn, 1986; Greenwood et al., 1994; Feng et al., 1998, 2002) and scanning (Faustmann and Dermietzel, 1985; Cho and De Bruyn, 1986; Hoshi and Ushiki, 1999) EM studies. Clearly, such events should be independent of junctional chemo- attractant and/or adhesion-molecule gradients and would not re- quire mechanisms for separation of adherence junctions. Proactive roles for the endothelium in facilitating diapa- desis have been suggested by a variety of studies. For example, disruption of the cytoskeleton or blockade of intracellular calcium in the endothelium significantly reduces the rate and ex- tent of leukocyte diapedesis without altering firm adhesion (Huang et al., 1993; Adamson et al., 1999; Etienne-Manneville et al., 2000; Strey et al., 2002). Recently, we and others (Bar- reiro et al., 2002; Carman et al., 2003) have demonstrated that, as a consequence of binding to LFA-1 and VLA-4, respect- ively, the endothelium proactively generates ICAM-1– and VCAM-1–enriched upright microvilli-like projections that sur- round a portion of the adherent leukocytes. However, in neither of these studies was any relationship between projections and leukocyte TEM characterized. Rather, in one of these, the pro- jections were described explicitly as firm adhesion "docking structures" that were predicted to be inhibitory for diapedesis and thus speculated to undergo disassembly or disruption be- fore diapedesis could proceed (Barreiro et al., 2002). Alterna- tively, we directly demonstrated that inhibition of projection formation did not alter the strength of adhesion and, based on this and the kinetics of endothelial projection formation, hypothesized a role for these structures in leukocyte diapedesis (Carman et al., 2003).

Here, we make extensive use of high resolution fluores- cence imaging techniques to explore the route and mecha- nisms for diapedesis. We demonstrate unambiguously that leukocytes can use the transcellular route of diapedesis. Fur- thermore, both paracellular and transcellular TEM are associ- ated with highly ICAM-1– and VCAM-1–enriched endothelial projections which appear to play a role in guiding leukocyte extravasation.

**Results**

**Leukocytes transmigrate through both para- and transcellular routes**

To investigate the mechanism and route of leukocyte diape- desis, we established an in vitro system for imaging TEM of primary leukocytes. TNF-α–activated human umbilical vein endothelial cell (HUVEC) monolayers were treated with monocyte chemoattractant protein-1 (MCP-1) before addition of monocytes, with platelet activating factor (PAF) before ad- dition of neutrophils, or with SDF-1 before addition of lympho- cytes. Chemoattractant not associated with the monolayers was removed by washing and leukocytes were then incubated with the monolayers for various times followed by fixation and staining with antibodies to ICAM-1, LFA-1, and VE-cadherin, a specific marker for endothelial adherence junctions (Lampug- nani et al., 1992). Samples were analyzed by high resolution, serial-sectioning confocal microscopy, coupled to digital de- convolution and three-dimensional image reconstruction, and leukocyte TEM was assessed according to the criteria described in Materials and methods.

At time periods of 10 min, significant numbers of all three classes of leukocytes were observed to be in the process of TEM. Remarkably, considerable fractions of TEM events were found to take place at sites clearly distinct from cell–cell junctions, demonstrating unambiguous use of the transcellular route of TEM (Fig. 1; Fig. S1, cell 2; Fig. S2, available at http://www.jcb.org/cgi/content/full/jcb.200404129/DC1). To analyze this statistically, at least 100 leukocytes in randomly selected fields from each of at least three separate experiments were imaged in all apical to basal planes. This revealed that 7 ± 1%, 5 ± 2%, and 11 ± 4% of the transmigrating monocytes, neutrophils, and lymphocytes, respectively, used transcellular routes. The remainder of leukocytes transmi- grated at sites closely juxtaposed to endothelial cell–cell junc- tions, and were scored as paracellular TEM (Fig. 2; Fig. S1, cell 1; Fig. S2). Among the TEM events scored as paracellular, were many whose appearance was suggestive of a trans- cellular route, yet whose TEM passage was too close to endo- thelial cell–cell junctions for an unambiguous determination of transcellular TEM (Fig. S2 C).
The nature of the transcellular pore itself remains to be elucidated. However, as an initial step, we characterized the distribution of endothelial caveolin-1, a structural constituent of caveolea. These experiments demonstrate a partial, nonetheless unambiguous, association of caveolin-1 with the transcellular pore (Fig. 3), which is consistent with a potential relationship between caveolae and pore formation.

**Leukocyte transmigration is highly associated with endothelial “cuplike” structures enriched in ICAM-1**

The overall appearance of the endothelium and of the leukocytes was not significantly different among para- and transcellular TEM events and among monocytes, neutrophils, and lymphocytes (Figs. 1 and 2; Figs. S1 and S2). Most importantly, virtually all TEM events shared a previously unrecognized characteristic of being associated with upright microvilli-like ICAM-1–enriched endothelial projections that formed a cup-like structure that surrounded the site of diapedesis in a largely symmetrical fashion (Figs. 1 and 2; Figs. S1 and S2). This architecture was not altered when leukocytes were subjected to a wall shear stress of 4 dyne/cm² in a laminar flow chamber during transmigration (Fig. S3, available at http://www.jcb.org/cgi/content/full/jcb.200404129/DC1).

Close investigation of leukocyte β2 integrin, the common subunit of the ICAM-1 receptors LFA-1 and Mac-1, revealed regions of increased density that formed linear clusters oriented parallel to the direction of diapedesis, which colocalized with the ICAM-1 projections (Fig. 1 E; Fig. 2 E; and Fig. 6 C, bottom). Separate analysis of αL and αM subunits revealed similar results (unpublished data). As demonstrated previously (Carman et al., 2003), leukocytes not associated with projections failed to exhibit linear integrin clusters (unpublished data).

To quantify the relationship between the cup-like structures and diapedesis, populations of leukocytes were scored using a system similar to one described previously (Sandig et al., 1997), as either adherent but not transmigrating (i.e., completely apical), in TEM stage-1 (TEM-1; 1–25% across the endothelium), TEM-2 (25–75% across the endothelium), TEM-3 (75–99% across the endothelium) or under the endothelium (100% below the endothelium). In addition, each cell was scored for the presence of significant ICAM-1–enriched projections of 1 μm in length or greater, and as transmigrating via either a para- or transcellular route.

Over a 60-min time course, the fraction of monocytes that had completed diapedesis steadily increased (Fig. 4 A). At all time points the population of cells that were in any stage of TEM (TEM-1, -2, or -3) was highly associated with ICAM-1 projections (average projection-positive fraction was 96%). Consistent with our previous findings (Carman et al., 2003), a proportion of cells on the apical surface of the endothelium that had not yet initiated diapedesis were also associated with
ICAM-1 projections. However, the projection-positive fraction of these cells averaged only 41% (P value vs. transmigrating cells of < 0.0001) over the 60-min time course. Using a single time point, a similar analysis was performed on neutrophils and lymphocytes (Fig. 4 B), which demonstrated a nearly complete association of TEM events with ICAM-1 projections (neutrophils, 94%; lymphocytes, 98%) and a significantly lower percentage of adherent, nontransmigrating leukocytes associated with projections (neutrophils, 22% (P < 0.0001); lymphocytes, 23% (P < 0.0001)). This situation was similar under shear conditions (4 dyne/cm²), with the majority of transmigrating monocytes (94 ± 2%), neutrophils, (87 ± 6%) and lymphocytes (91 ± 2) associated with projections.

Of the ICAM-1 projection-positive population of transmigrating leukocytes, over all time points, 97% were associated with cuplike projections that were largely symmetrical, encircling at least 240° of the circumference of the leukocytes (See Fig. 6, A–C, bottom; Fig. S4, A–C, available at http://www.jcb.org/cgi/content/full/jcb.200404129/DC1). In contrast, of the projection-positive, apically adherent, nontransmigrating leukocytes associated with projections (neutrophils, 22% (P < 0.0001); lymphocytes, 23% (P < 0.0001)). This situation was similar under shear conditions (4 dyne/cm²), with the majority of transmigrating monocytes (94 ± 2%), neutrophils, (87 ± 6%) and lymphocytes (91 ± 2) associated with projections.

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The endothelial cuplike structures are highly enriched in ICAM-1 and VCAM-1, but not ICAM-2, PECAM-1, or VE-cadherin

To investigate the localization of other relevant endothelial adhesion molecules, ICAM-1 was visualized concomitantly with β2 integrin and either VCAM-1, ICAM-2, PECAM-1, or VE-cadherin. VCAM-1 was highly coenriched with ICAM-1 in
the endothelial projections surrounding both adherent (not depicted) and transmigrating leukocytes (Fig. 6, A and C; Video 1, available at http://www.jcb.org/cgi/content/full/jcb.200404129/DC1). Conversely, VE-cadherin was localized to endothelial cell–cell junctions and was absent from projections (Fig. 6 B and Fig. S4 C). Interestingly, although in the majority of the paracellular TEM events VE-cadherin staining was interrupted and absent at the site of diapedesis (Fig. 7 B; Fig. S2, A and B; Fig. S4 C), in many cases the VE-cadherin was continuous and arched around one side of the TEM passage (Fig. 2, 6B, Fig. S1, Figure 3.

Caveolin-1 is partially associated with the transcellular migration pore. Human lymphocytes were incubated for 10 min with TNF-α-activated, SDF-1–pretreated HUVECs transfected with either caveolin-1-GFP (A and B) or GFP-caveolin-1 (C and D) followed by fixation and staining. β2 (blue), ICAM-1 (IC1, red), and caveolin-1 (ca1; green) are shown for representative transcellular migration events. Confocal sections that encompass the TEM passage (Fig. 1, F and H) are projected as top views. Bar, 5 μm.

Figure 4. ICAM-1 projections are highly associated with transmigrating cells. Monocytes (A) and neutrophils and lymphocytes (B) were incubated with TNF-α-activated and either MCP-1–, PAF–, or SDF-1–pretreated HUVEC monolayers, respectively, for the indicated number of minutes (m). Cells were fixed and stained for ICAM-1, LFA-1, and VE-cadherin. In each of three to eight separate experiments, a minimum of 100 leukocytes from randomly selected fields for each time point were carefully analyzed in all apical to basal planes and scored for the presence of ICAM-1–enriched projections of 1 μm in length or greater and as being either apically adherent (A), in the process of TEM (including both para- and transcellular events and including TEM stages 1–3) (T) or under (U) the HUVEC monolayer. Each bar represents the percentage of total cells scored at each indicated time point that were found in each of the three categories (i.e., A, T, or U). The black portion of each bar is the fraction of cells scored positive for associated ICAM-1 projections. The gray portion of each bar is the fraction of the cells that were negative for the presence of ICAM-1 projections. Values represent mean ± SEM (n = 3–8).
ICAM-2 and PECAM-1 exhibited only modest localization to the endothelial projections (Fig. S4, A and B). Functional roles for these molecules in this context cannot be excluded.

Given the strong enrichment of VCAM-1 to the projections, we also investigated the distribution of the leukocyte integrin VLA-4 (α4β1) on lymphocytes, the receptor for VCAM-1. These results revealed that α4, as with β2, αL, and αM, was distributed to linear clusters on leukocytes that colocalized with the ICAM-1/VCAM-1–enriched endothelial projections (Fig. 6 D).

At initial stages of TEM, ICAM-1 projections are clearly distinct from the TEM passage

Leukocytes at initial stages of TEM (TEM-1) looked much like adherent cells (Carman et al., 2003) surrounded by a perimeter of vertical endothelial ICAM-1–enriched projections (Fig. 6 A and Fig. 7; Fig. S1, cell 2 and Fig. S3, B and C). However, at the very bottom of the leukocyte/endothelial interface, usually at the center of the perimeter of ICAM-1 projections, was a small circular/ovoid gap (~0.5-2 μm in diameter) completely devoid of ICAM-1 staining in all confocal sections (the TEM passage; Fig. 6 A and Fig. 7; Fig. S1, cell 2 and Fig. S3, B and C). LFA-1 was clearly detected within the TEM passage extending down to the most basal sections indicating a portion of the leukocyte was spread underneath the endothelium. Thus, at initial stages of TEM the vertical endothelial projections are clearly distinct from the TEM passage itself and rather form a larger perimeter that encircles the location of passage formation, as though targeting this location for diapedesis. This feature was constant for both para- and transcellular routes of TEM and for monocytes (Fig. 6 A and 7, Fig. S1, cell 2 and Fig. S3 B), neutrophils (Fig. S3 C) and lymphocytes (not depicted).

ICAM-1 projections remain associated with leukocytes through completion of diapedesis

At the late stages of TEM (TEM-3) the distal ends of the ICAM-1 projections remained well attached to the apical portion of the leukocytes with significant LFA-1 clustered underneath them (Fig. S5, A–C, available at http://www.jcb.org/cgi/content/full/jcb.200404129/DC1). At the very final stages or just after completion of TEM, the ICAM-1 projections remained attached to the LFA-1–enriched uropod of the subendothelial monocytes, neutrophils and lymphocytes. Thus, the ICAM-1 projections were often observed extending from the apical surface, through the nearly closed paracellular or transcellular TEM passage, into the basal/sub-endothelial space (Fig. S5, D–I), an assessment confirmed by parallel experiments using interference reflection microscopy (IRM; not depicted).

Conditions that attenuate formation of ICAM-1–enriched projections decrease TEM

To assess the functional role of the ICAM-1–enriched projections in TEM, we scored the relative proportions of monocytes that were adherent, transmigrating or under the HUVEC monolayer after treatment with reagents (colchicine or 1,2Bis(2-amino-
nophenoxy)-ethane-N,N',N'-tetraacetic acid tetrakis(acetoxyethyl ester); BAPTA-AM) shown previously to inhibit projections (Carman et al., 2003). Compared with controls, colchicine and BAPTA-AM significantly reduced both para- and transcellular TEM (3-4-fold; Fig. 8 A), which is consistent with previous studies (Huang et al., 1993; Etienne-Manneville et al., 2000; J. Greenwood and P. Adamson, personal communication). Whereas 50% of adherent cells had ICAM-1 projections on the control monolayers, only 16% and 18% of the adherent cells on colchicine and BAPTA-AM treated monolayers, respectively, were associated with projections (Fig. 8 A). However, among the leukocytes that were in the process of TEM, the majority (96%, 91%, and 90% for control, colchicine, and BAPTA-AM–treated samples, respectively) were associated with ICAM-1 projections, regardless of HUVEC pretreatment (Fig. 8 A).
We demonstrated previously that the endothelial projections both were enriched in, and required, F-actin (Carman et al., 2003). To assess the role of Rho family GTPases in projection formation, endothelial cells were pretreated with either Clostridium difficile toxin-B, an inhibitor of Rho, Rac, and CDC42, or Clostridium botulinum C3 transferase, a selective inhibitor of Rho, under conditions that promoted similar decreases in actin stress fiber content, as assessed by phalloidin-FITC staining (unpublished data). For toxin-B this was associated with an approximately twofold reduction in both total projections and TEM (Fig. 8 B), whereas, C3 had no effect on either projections, as shown previously (Carman et al., 2003), or TEM in this setting (Fig. 8 B). As above (Fig. 8 A), in all cases the majority of TEM events (91%, 91%, and 93% for control,
in some cases apparent transcellular TEM passages were too current possible we were able to clearly visualize many trans-endothelium, to directly resolve the transendothelial passage.

with visualization of leukocyte integrins and their ligands on cadherin (or PECAM-1) to discern junctions concomitantly this work is primarily a consequence of visualization of VE- cadherin (or PECAM-1) to discern junctions concomitantly with increased with decreased efficiency of diapedesis. The finding that the relative low percentages of transcellular diapedesis observed here in vitro, compared with some in vivo studies (Feng et al., 2002), correlates with the relatively low levels of vesiculo-vacuolar organelle and caveole in cultured endothelium compared with endothelium in vivo (Vasile et al., 1999).

We and others (Barreiro et al., 2002; Carman et al., 2003) have recently demonstrated that the endothelium proactively forms ICAM-1 and VCAM-1–enriched microvilli-like projections that form cuplike structures around populations of adherent leukocytes in an LFA-1– and VLA-4–dependent manner. These studies did not address whether projections were associated with diapedesis. However, in one of them it was interpreted that the endothelial projection structures likely functioned in adhesion strengthening and were incompatible with diapedesis and therefore would be required to disassemble before diapedesis could proceed (Barreiro et al., 2002).

To the contrary, based on both our previous (Carman et al., 2003) and current studies, we conclude that any functional role attributed to the endothelial projections would most likely lie in transmigration. The kinetics of projection formation are on the order of minutes (Carman et al., 2003), which is consistent with the time scale of diapedesis, but not with those of either rolling interactions (fractions of seconds) or firm adhesion (tens of seconds; Luu et al., 1999). Moreover, our analysis here demonstrates that transmigrating leukocytes represent the predominant projection-associated leukocyte population with an essentially complete correlation between TEM and the presence of projections. Finally, disruption of endothelial projections failed to alter the strength of the adhesions formed (Carman et al., 2003), but, as demonstrated here, correlated strongly with decreased efficiency of diapedesis. The finding that the subpopulations of projection-associated cells and transmigrating cells were essentially identical no matter whether monocytes, neutrophils, or lymphocytes were examined, or whether inhibitors were used, and that projections were highly associated with cells even at the very earliest detectable stages of TEM, suggests that projection formation is essential to the mechanism of diapedesis.

Based on this and the previously noted similarity to the phagocytic cup (Barreiro et al., 2002), we propose the nomenclature “transmigratory cup” to describe this novel cell–cell in-
terface architecture. Significantly, a variety of previous EM studies have made observations, including “endothelial microvilli” embracing transmigrating leukocytes, which is consistent with the formation of transmigratory cups in vivo (Williamson and Grisham, 1961; Faustmann and Dermietzel, 1985; Raine et al., 1990; Fujita et al., 1991).

We previously provided conclusive evidence that projection formation is the proactive response of the endothelium and does not require force or tension generation from the leukocyte (Carman et al., 2003). However, based on our analysis of asymmetrically distributed projections found associated with the trailing edge of laterally migrating leukocytes, we conclude that forces generated by leukocytes can modify the endothelial projections. Because asymmetric lateral projections were associated only with laterally migrating cells, and TEM was associated only with symmetric, cuplike endothelial projections, diapedesis would seem to require a transition from a highly polarized phenotype and rapid lateral migration to greater symmetry (in the x-y plane) and reduced lateral migration.

As discussed above, an important issue for TEM is how leukocytes establish directionality during diapedesis. The current model is that junctionally localized chemoattractant and adhesion molecule gradients provide the principle driving force for this (Bianchi et al., 1997; Aurrand-Lions et al., 2002; Muller, 2003). If we consider the transcellular migration events observed in this work, which occur at sites distant from endothelial junctions, we must conclude that an alternate “driving force” for TEM exists. The endothelial projections represent an attractive candidate for this. These vertical ICAM-1/VCAM-1–enriched microvilli-like projections form a cuplike traction structure that is aligned perpendicular to the plane of the endothelium and parallel to the direction of diapedesis, providing a physical basis for oriented migration. The finding that at sites opposing the endothelial projections, leukocyte integrins formed linear clusters reminiscent of fibrillar adhesions, on which cells are known exert forces during migration (Dzamba et al., 1994), is consistent with this idea. Given that chemoattractants are well known to be critical for TEM (Springer, 1994; Cunnon et al., 2001; Worthylake and Burridge, 2001) we envision that they cooperate with the ICAM-1– and VCAM-1–enriched transmigratory guidance structure in establishing proper directionality and, therefore, efficient TEM. In this regard, it is interesting to note that two prototypical chemokines, IL-8 and RANTES, have been shown to be distributed apically on endothelial microvilli in vivo (Middleton et al., 1997). Importantly, because the transmigratory cups were observed to be associated with virtually all TEM events, any functional consequences attributed to them should extend to both para- and transcellular TEM. In the case of paracellular TEM, projections would be anticipated to cooperate with other mechanisms previously shown to be functionally important (Bianchi et al., 1997; Aurrand-Lions et al., 2002; Muller, 2003).

Our studies demonstrate that leukocytes use transcellular, in addition to paracellular, routes for diapedesis. Independent of route, TEM events were highly associated with the transmigratory cup, in which ICAM-1– and VCAM-1–enriched vertical endothelial projections surround the site of leukocyte diapedesis. The architecture of this cup, and the high degree of correlation between these structures and the act of diapedesis, suggest that the transmigratory cup represents an important mode of directional guidance for TEM. The finding that both para- and transcellular TEM events by monocytes, neutrophils, and lymphocytes are associated with topologically similar transmigratory cups, demonstrates that this structure represents a general feature of TEM.

Materials and methods

Antibodies and reagents

Sources for antibodies to human v-actin (T522/4), aM (CBRM1/23), β2 (CBR-LFA1/7), ICAM-1 (CBR-IC1/11), and ICAM-2 (CBR-IC2/11) have been described previously (Klickstein and Springer, 1995; Lu et al., 2001). Anti–CD14-FITC and control IgG2a-FITC were obtained from Immunotech. Anti–VE-cadherin (clone 55-7H1) and MCP-1 were obtained from BD Biosciences. monoclonal anti–ICAM-1 (clone 7.2R) and anti–PECAM-1 (clone 9C11) and polyvalent anti–VCAM-1 and SDF-1 were obtained from R&D Systems. Fab fragments were prepared via papain cleavage according to the manufacturer’s instructions (Pierce Chemical Co.). Antibody and Fab conjugation to Cy3, Cy5 (Amersham Biosciences), or Alexa 488 (Molecular Probes) bis-reactive dyes were according to manufacturer’s instructions. PAF was obtained from Calbiochem. BAPTA-AM, colchicine, cytochalasin, and heparin were obtained from Sigma-Aldrich. Clostridium difficile toxin B was obtained from List Biochemicals. Clostridium botulinum C3 transferase was a gift provided by J. Greenwood and P. Adams (University College of London, London, UK).

Cells and cell culture

HUVECs, neutrophils, peripheral blood mononuclear cells, and monocytes were purified and cultured as described previously (Carman et al., 2003). Lymphocytes were prepared by adsorbing the monocytes to gelatin-coated plates and culturing nonadherent lymphocytes in RPMI supplemented with 10% FBS and PHA (1 μg/ml) for 3 d, followed by culture in IL-2 (20 ng/ml) for 3–6 d. Flow cytometric analysis demonstrated that these cells were 97% CD3 positive. HUVEC transient transfection was performed by Amaxa nucleofection according to the manufacturer’s instructions (Amaxa Inc.). DNA expression constructs for Caveolin-1-GFP and GFP-caveolin-1 were provided by M. Lisanti (Albert Einstein College of Medicine, Bronx, NY).

Adhesion and TEM experiments

For most fluorescence microscopy experiments, HUVECs were plated at 90% confluence on fibronectin (10 μg/ml)-coated polystyrene or glass surfaces and cultured for 48–72 h before use. HUVECs grown on gelatin or Matrigel gave results indistinguishable from those on fibronectin. Before each experiment, HUVECs were activated for 12 h with TNF-α (100 ng/ml). Immediately before addition of leukocytes, HUVECs were incubated with either PAF (100 nm), MCP-1 (200 ng/ml), or SDF-1 (200 ng/ml) at 37°C for 20 min. Where indicated, HUVECs were also preincubated with BAPTA-AM (20 μM, 1 h), colchicine (10 μM, 20 min), toxin-B (100 ng/ml, 1 h), C3 transferase (50 μg/ml, 16 h), or appropriate dilutions of vehicle (DMSO or PBS) concomitant with chemoattractant. In all cases, HUVECs were washed three times before addition of leukocytes with HBSS supplemented with 20 mM Hepes, pH 7.2, and 1% human serum albumin (buffer A). Freshly isolated neutrophils or monocytes or IL-2 cultured lymphocytes were washed and resuspended in buffer A and then added to HUVEC monolayers and incubated at 37°C for the indicated times. Cells were imaged live in Bioptechs chambers maintained at 37°C, or fixed in 3.7% formaldehyde in PBS for 5 min, stained for β2 (IAF1/7-Cy3 or Cy5; 20 μg/ml), aM (TS22/4-Cy3; 20 μg/ml), a4 (R7.2-488, 20 μg/ml), ICAM-1 (IC1/11-488, IC1/11-Fab-488, or IC1/11-Cy3; 20 μg/ml), ICAM-2 (IC2/1-488 or Cy3; 20 μg/ml), VCAM-1 (polyvalent anti–VCAM-1-Cy3; 20 μg/ml), PECAM-1 (9G11-Cy3), VE-cadherin (55-7H1-Cy3 or Cy5; 20 μg/ml) in PBS for 20 min at RT and then washed three times with PBS before imaging. For live-cell experiments, endothelial cells were preincubated with nonfunction blocking IC1/11-Fab-488 (20 μg/ml) for 20 min and then washed before addition of leukocytes that were prelabeled with 5(6)-carboxy-2′,7′-dichloromethyl SNARF-1, acetate (Molecular Probes) according to the manufacturer’s instructions.
For shear experiments, HUVEC monolayers were assembled as the lower wall in a parallel-wall flow chamber in a 37°C warm room. Leukocytes, in buffer A, were infused at 0.3 dyne/cm² for 30 s, to allow accumulation, then subjected to 4 dyne/cm² shear force for 10 min and finally fixed by perfusion of buffer A/3.7% formaldehyde under the same shear. Cells were stained as above.

Image acquisition and processing

Wide field differential interference contrast, IRM, and fluorescence imaging was conducted on an Axiovert S200 epifluorescence microscope (Carl Zeiss Microimaging, Inc.), using a 63× oil objective, coupled to a Hamamatsu Orca CCD. Confocal imaging was performed with a Radiance 2000 Laser-scanning confocal system (Bio-Rad Laboratories) on a microscope (model BX50WV; Olympus) with a 100× water immersion objective. For serial Z-stacks the section thickness was between 0.1 and 0.3 μm. Image processing, including background subtraction and digital deconvolution, was performed with Openlab software (Improvision). Three dimensional reconstruction and projection of Z-stacks was performed with VolView software (Vay Tek). Images were then exported to Photoshop or Quotikta software for preparation of final images or videos, respectively.

Criteria for determination of leukocyte TEM

Criteria for determination of leukocyte TEM were based on analysis of the relative distribution of ICAM-1, LFA-1, and VE-cadherin fluorescence in both the x-y and the z dimensions. The point of leukocyte penetration through the endothelium (the TEM passage) was defined by the region in the x-y dimension completely devoid of ICAM-1 fluorescence in all z-planes. The TEM passage contained leukocyte LFA-1 fluorescence that extended down to the most basal sections at the substrate. LFA-1 fluorescence adjacent to the TEM passage was that which indicated region where TEM was defined by parallel experiments that compared the localization of HUVEC stress fibers with leukocyte LFA-1 as described previously (Sandig et al., 1997). Further validation for our method came from IRM, which confirmed that portions of leukocytes determined by ICAM-1/LFA-1 fluorescence to have penetrated the endothelium were in close contact with the substrate, whereas the apical portions of the leukocytes were confirmed not to be in contact with the substrate. TEM adjacent to VE-cadherin-stained endothelial cell–cell junctions was scored as paracellular; TEM where no part of the TEM passage was within 1 μm of a junction was scored as transcellular.

Cells were scored as either adherent but not transmigrating, in TEM stage 1–2, or 1–3 or under the endothelium. In addition, each cell was scored for the presence of significant ICAM-1–enriched vertical projections of 1 μm in length or greater. Projections associated with only one side of a leukocyte were termed “asymmetric” or “tetherlike,” whereas those that were associated with at least 240° of the circumference of a leukocyte were termed “symmetric” or “cuplike.”

Online supplemental material

Figs. S1–S5 show further examples of both para- and transcellular transmigration with monocytes, neutrophils, and lymphocytes under static (1–2) and shear (3) conditions. Fig. S4 depicts distribution of ICAM-2, PE-CAM-1, and VE-cadherin relative to transmigration cups. Fig. S5 shows ICAM-1 projections associated with subendothelial leukocytes. Video 1 shows three-dimensional rotation of the images depicted in Fig. 6 C. Online supplemental materials are available at http://www.jcb.org/cgi/content/full/jcb.200404129/DC1.

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