A Review of the Potential Interaction of Selenium and Iodine on Placental and Child Health

Nahal Habibi 1, Jessica A. Grieger 2,∗ and Tina Bianco-Miotto 1,∗

1 School of Agriculture, Food and Wine, Waite Research Institute, and Robinson Research Institute, University of Adelaide, Adelaide 5005, Australia; nahal.habibi@adelaide.edu.au
2 Adelaide Medical School, and Robinson Research Institute, University of Adelaide, Adelaide 5005, Australia
* Correspondence: jessica.grieger@adelaide.edu.au (J.A.G.); tina.bianco@adelaide.edu.au (T.B.-M.)

Received: 10 July 2020; Accepted: 28 August 2020; Published: 2 September 2020

Abstract: A healthy pregnancy is important for the growth and development of a baby. An adverse pregnancy outcome is associated with increased chronic disease risk for the mother and offspring. An optimal diet both before and during pregnancy is essential to support the health of the mother and offspring. A key mediator of the effect of maternal nutrition factors on pregnancy outcomes is the placenta. Complicated pregnancies are characterized by increased oxidative stress in the placenta. Selenium and iodine are micronutrients that are involved in oxidative stress in placental cells. To date, there has been no comprehensive review investigating the potential synergistic effect of iodine and selenium in the placenta and how maternal deficiencies may be associated with increased oxidative stress and hence adverse pregnancy outcomes. We undertook a hypothesis-generating review on selenium and iodine, to look at how they may relate to pregnancy complications through oxidative stress. We propose how they may work together to impact pregnancy and placental health and explore how deficiencies in these micronutrients during pregnancy may impact the future health of offspring.

Keywords: iodine; micronutrients; selenium; pregnancy; placenta; oxidative stress

1. Introduction

Pregnancy is characterized by a pro-inflammatory, hyperlipidemic and hyperinsulinemic state [1]. A normal healthy pregnancy is important for the growth and development of the baby and for the lifelong health of the mother and offspring [2]. To provide a suitable condition for the developing fetus, the maternal body undergoes major biochemical, physiological and anatomic adaptations [3], which typically revert back to the condition they were in prior to pregnancy [4].

Human pregnancy is also associated with an increase in oxidative stress markers, due to increased placental mitochondrial activity and the production of reactive oxygen species, which have pronounced effects on placental function [5]. The placenta is the interface between the mother and baby. It mediates nutrient and waste exchange to support a healthy pregnancy. Factors such as poor maternal diet or obesity impact placental health and function, such as proliferation, invasion and apoptosis of trophoblast cells [6,7]. This can have a profound effect on fetal growth and development, and pregnancy success [6,7]. As such, a balance of antioxidant and oxidant status is key to a healthy placenta and hence a successful pregnancy outcome for mother and baby.

There is a clear role for an optimal diet both before and during pregnancy to support the health of the mother and offspring [8–12]. Higher dietary scores for a high-protein/fruit pattern before pregnancy, was associated with decreased likelihood of preterm birth (adjusted odds ratio; 95% confidence interval, OR: 0.31; 95% CI: 0.13, 0.72), whereas the reverse direction was apparent for the high-fat/sugar/takeaway pattern (adjusted OR: 1.54; 95% CI: 1.10, 2.15) [13]. Increasing dietary quality,
calculated by the Alternative Healthy Eating Index 10 during pregnancy, was associated with a decrease in the likelihood of delivering a small-for-gestational-age (SGA) baby [14]. Supplementation with vitamins and minerals may also have a positive effect on infant birthweight [15]. Some micronutrients, such as selenium and iodine, are essential micronutrients because the body cannot produce them and the mother needs to obtain them from her diet or from supplemental intake. Deficiencies in iodine can lead to thyroid hormone deficiency, and therefore improper neurodevelopment and mental retardation in the fetus [16]. Selenium deficiency is associated with pregnancy-induced hypertensive disorders [17], miscarriage [18], preterm birth [19], and gestational diabetes [20,21]. Importantly, the key mediator of the effect of maternal nutrition factors on pregnancy outcomes is the placenta. However, such pregnancy outcomes do not just end after pregnancy, but can have lifelong consequences for a mother and her offspring. This is depicted by the developmental origins of health and disease (DOHaD) hypothesis, which emphasizes the importance of fetal life exposure to maternal factors, such as diet, in the development of chronic diseases later in life [22]. Yet, while maternal nutrition and the placenta are important factors regarding pregnancy success, there is a paucity of information examining multiple nutrients in combination, and the potential synergistic effect they may have in pregnancy.

**Figure 1.** Deficiencies (↓) in selenium and iodine result in placental oxidative stress which may contribute to adverse pregnancy outcomes and hence increased (↑) risk of chronic disease in offspring.

Adverse pregnancy outcomes are associated with increased oxidative stress in the placenta [23] and since selenium and iodine are involved in oxidative stress in placental cells [24], it is important to investigate the role of both micronutrients. To the best of our knowledge, no study has reviewed current evidence regarding the potential synergistic effect of iodine and selenium in the placenta. Our knowledge from other organs, such as the thyroid, highlights the importance of these two micronutrients and their combined impact on oxidative stress [25]. We identified studies that investigated the association between maternal selenium and iodine status with pregnancy complications using the following keywords: “maternal blood selenium concentration AND pregnancy complication” and “maternal urinary iodine concentration AND pregnancy complication” in PubMed. The last search was performed in July 2020. In addition, any relevant studies identified from the reference list of selected studies were also included. The exclusion criteria were: (1) review papers, (2) unavailable full text, (3) non-English language, (4) animal studies, and (5) taking any medication during pregnancy. Pregnancy outcomes were defined as healthy if there was no complication or complicated, if there was...
one of the following pregnancy complications: preeclampsia (PE), pregnancy-induced hypertension, miscarriage, gestational diabetes mellitus (GDM), preterm birth (PTB), intrauterine growth restriction (IUGR), or premature rupture of membrane (PROM). In this review we outline how selenium and iodine may relate to pregnancy complications through oxidative stress, and we propose how they may work together to impact pregnancy and placental health (Figure 1). We also explore how deficiencies in these two micronutrients during pregnancy may impact on the future health of the offspring, which has not been previously reviewed as a combination.

2. Oxidative Stress and the Importance of Antioxidants

Oxidative stress occurs when free radicals are generated to a level higher than the physiological level and the antioxidant system cannot neutralize them [26,27]. Free radicals, namely reactive oxygen species (ROS), are unstable molecules that have unpaired electrons and can donate or accept an electron to be stabilized [28]. As a result, the new molecule is unstable and formation of free radicals continues. This chain reaction can cause oxidative damage to nucleic acids, lipids, carbohydrates and proteins, which results in the decaying of tissues and the disruption of homeostasis [27]. The production of ROS is a continuous phenomenon in cells and is essential for cell-signaling, however, excessive ROS levels are detrimental to cells [27,29].

To stop the accumulation and damage of ROS, free radicals need to be neutralized by pairing unpaired electrons. This is how the antioxidant system can protect the cell against oxidative damage [30]. Antioxidants typically work in three ways, including the prevention of ROS formation, the interception of a ROS chain reaction and the repairing of damaged molecules [31,32]. The antioxidant defense system is made up of enzymes and non-enzyme components. Glutathione peroxidase (GPx), catalase, superoxide dismutase and thioredoxin reductase (TRx) are some of the enzymes [33] but they need cofactors such as micronutrients to work properly. For example, superoxide dismutase requires manganese or copper and zinc, while GPx and TRx need selenium to function properly. Therefore, micronutrient deficiencies can reduce the ability of the antioxidant system to protect cells against free radicals [34]. Vitamin C, vitamin E and beta-carotene are other micronutrients involved in the antioxidant defense system [35,36].

**Oxidative Stress and Pregnancy Complications**

Pregnancy complications are adverse outcomes of pregnancy such as preeclampsia (PE), gestational diabetes mellitus (GDM), intrauterine growth restriction (IUGR), preterm birth (PTB), and premature rupture of membrane (PROM). They can increase morbidity and mortality rates for both the mother and her offspring [37–40].

Preeclampsia is a pregnancy-specific, multisystem disorder, presenting as hypertension with new onset of one or more of proteinuria or renal, liver, neurological or hematological complications, or uteroplacental dysfunction after 20 weeks’ gestation [41]. Patients with a history of PE have around a 2-fold increased risk for vascular diseases such as hypertension and ischemic heart disease [42], stroke [42] and chronic kidney disease [43], at 10–15 years post-partum. Infants of mothers who had PE are also at an increased risk of small-for-gestational-age [44] and perinatal mortality [45]. Children exposed to preeclamptic pregnancies have increased hospitalizations due to infectious and parasitic, nutritional and metabolic diseases of the respiratory system, blood, and blood-forming organs at ages 1–13, 16, 18–21 and 24 years compared to unexposed children [46]. Moreover, women who experience PE in their first pregnancy have a higher risk of myocardial infarction and cardiovascular death compared to non-PE mothers [47]. Currently, delivery of the fetus and placenta is the only treatment [41,42,45,48,49].

The pathogenesis of PE involves improper placental development as a result of dysfunctional proliferation, migration, and invasion of placenta-derived extravillous cytotrophoblast cells into the uterine vasculature, along with maternal endothelial and vascular dysfunction [48]. This leads to placental hypoxia and subsequent reperfusion, resulting in oxidative stress and inflammation [50,51].
Similar to PE, oxidative stress also contributes to PTB, IUGR and PROM. IUGR refers to a fetus that is smaller than expected for their gestational age. One of the most common causes of IUGR is uteroplacental insufficiency in which the placenta is unable to provide the developing fetus with sufficient nutrients and oxygen. The growth-restricted offspring have a 2–6 fold increased risk of developing chronic diseases such as type 2 diabetes mellitus, coronary heart disease and chronic kidney disease later in life [52].

The same placental dysfunction seen in PE is also seen in IUGR with deficient spiral artery remodeling resulting in malperfusion [53]. The malperfusion results in oxidative stress within the placenta and overwhelms the antioxidant system. In addition, the enzymes which act as antioxidants require micronutrients to work and maternal diets deficient in these micronutrients impact on the placenta’s ability to combat oxidative stress [53].

3. Micronutrients and Pregnancy

To compensate for the higher demand of growing maternal tissues and fetus and also for hemodilution that occurs in pregnancy, a higher dietary intake of many micronutrients and trace elements are recommended [4,54]. In developed countries, despite the availability of suitable nutritional foods, many pregnant women have an imbalanced diet that puts them at risk of an inadequate intake of micronutrients like folate, vitamin D, vitamin B12, iron and iodine [55,56]. There is considerable evidence that shows that deficiency of micronutrients such as iodine, selenium, zinc, vitamin E, folate and iron, adversely impacts maternal and fetal health, and pregnancy outcome [19,57–69]. One of the reasons for this association may be the bio-functionality of vitamins and minerals in pregnancy, besides their classical roles in health and disease in the general population [56]. Although maternal micronutrient intake through the entire gestational period can affect fetal growth and development, the peri-implantation stage and placental development are critical windows for programming a healthy birth and future life [70,71].

3.1. Selenium

Selenium is an essential trace element that plays a pivotal role in the antioxidant defense system, cell cycle, and immune function, because of its contribution to selenoproteins [72]. Selenoproteins such as glutathione peroxidases (I, II, III, IV, and VI); thioredoxin reductases (I, II, and III); and selenoprotein H, P, and W, have antioxidant properties and are involved in managing ROS production. Selenoproteins also possess a vast range of functions such as protein folding, signaling, lipid biosynthesis, cell cycle, and calcium regulation [73–79].

Selenium is found in a broad range of foods including brazil nuts, wholegrain foods and cereals, fish, beef, eggs, and some fortified ready-to-eat breakfast cereals. The recommended dietary intake of selenium has been calculated by the amount of selenium needed to maximize the synthesis of glutathione peroxidase (GPx) [80]. During pregnancy, it is 60 micrograms per day in European countries [81] and 65 micrograms per day in Australia and New Zealand [82]. The requirement for selenium is higher during pregnancy, up to 4 micrograms per day, due to fetal requirements [83].

Globally, up to one in seven people have a low selenium dietary intake [84]. Selenium levels can be influenced by multiple factors including drinking water, soil, plant and animal tissue content of selenium [84], as well as different intakes in different regions. For example, the average intake of selenium in Eastern Europe is lower than that of Western Europe [85]. In addition, in New Zealand [85] and most areas of China [86], selenium deficiency is evident. In the Middle East, selenium intake is dependent on socio-economic status [85].

While studies across different countries report various levels of selenium during pregnancy (Table 1), using the data reviewed by Mariath et al. (2011) [87], Perkins and Vanderlelie (2016) proposed that selenium levels below 45 micrograms per litre can be dangerous and can be associated with adverse birth outcomes. They further concluded that selenium levels above 95 micrograms per litre can be considered as seleno-sufficient because the majority of selenoproteins can be maximally
expressed at this level [57]. In addition to the measurement of maternal selenium levels, markers of functional effects of selenium may provide additional information. For instance, assessment of selenoproteins such as GPX3 and SEPP1 are suitable to investigate the functional effects of selenium such as antioxidant activity, nutritional selenium deficiency, and evaluating responses of deficient individuals to selenium supplementation [88]. However, more studies would be needed to determine whether these selenoproteins provide additional information for interpreting clinical outcomes in addition to, or instead of, maternal selenium levels. The time point of selenium assessment can also affect the interpretation of the data, since pregnant women that were selenium sufficient in the first trimester have been shown to become selenium deficient later in pregnancy [89].

| References (Ref.) | Country Year | Sample Size (Women) | Time of Sampling | Sample Type | Selenium Concentration (µg/L) |
|------------------|--------------|---------------------|------------------|-------------|-----------------------------|
| [90]             | Indonesia 2019 | 25 pregnant         | Delivery         | Serum       | Median ± SD: 78.42 ± 16.30   |
| [91]             | Turkey 2019    | 30 pregnant         | Delivery         | Blood, urine, amniotic fluid | Maternal blood: 78.98 (72.36–84.14) |
|                  |               |                     |                  |             | Maternal urine: 23.44 (19.66–26.69) |
|                  |               |                     |                  |             | Amniotic fluid: 26.00 (22.56–29.88) |
| [92]             | Australia 2018 | 558 pregnant        | 15 ± 1 weeks’ gestation | Plasma    | Mean ± SD: 71.93 ± 11.05     |
| [93]             | Sudan 2014     | 31 pregnant         | Not reported     | Serum       | Median (25–75th quartile): 204 (68–541) |
| [94]             | Iran 2011      | 40 pregnant         | 34–39 weeks’ gestation | Blood     | Mean ± SD: 58.51 ± 11.85     |
| [95]             | UK 2008        | 27 pregnant         | Delivery         | Blood       | Mean ± SD: 58.4 ± 14.9       |
|                  |               | 22 non-pregnant     |                  |             | Non-pregnant: 69.8 ± 11.7    |
| [96]             | Hungary 2008   | 20 pregnant         | 24–28 weeks’ gestation | Serum    | Mean ± SD: 40.5 ± 8.03       |
|                  |               | 24 non-pregnant     |                  |             | Non-pregnant: 77.4 ± 14.82   |
| [97]             | Turkey 2005    | 28 pregnant         | 28–39 weeks’ gestation | Serum     | Mean ± SD: 87.50 ± 10.96     |
|                  |               | 25 non-pregnant     |                  |             | Non-pregnant: 109.0 ± 6.34   |
| [98]             | Kuwait 2004    | 15 pregnant         | Delivery         | Blood       | Mean ± SEM: Maternal vein: 102.3 ± 3.1 |
|                  |               |                     |                  |             | Umbilical artery: 85.4 ± 4.2 |
|                  |               |                     |                  |             | Umbilical vein: 82.6 ± 4.1   |
| [99]             | USA 2004       | 22 pregnant         | 12 and 34 weeks’ gestation | Plasma  | Mean ± SD: 12 weeks’ gestation: 126.0 ± 15.0 |
|                  |               |                     |                  |             | 34 weeks’ gestation: 111.0 ± 12.0 |
| [100]            | Poland 2001    | 36 pregnant         | Before or within 12 hours after uterine curettage | Whole blood and plasma | Mean ± SD: Whole blood: 74.1 ± 11.6 |
|                  |               | 28 non-pregnant     |                  |             | Non-pregnant: 90.5 ± 11.2    |
|                  |               | 36 pregnant         |                  |             | Plasm: 54.6 ± 11.1           |
|                  |               | 28 non-pregnant     |                  |             | Non-pregnant: 66.1 ± 13.1    |

Table 1. Selenium levels in women of reproductive age.

SD, Standard deviation; SEM, standard error of the mean; µg/L, micrograms per litre.

3.2. Selenium and Pregnancy Complications

Selenium is necessary for human reproduction [100]. We have recently shown that in 1060 Australian pregnant women, those who had lower selenium concentrations (<0.95 micromol per litre) took a longer time to conceive (1.19 (1.01–1.40)), an equivalent of around 0.6 months, and they were at greater risk of infertility (1.46 (1.06–2.03)) [101]. Deficiency in selenium, measured by lower concentrations either in plasma, toenail, red blood cells or hair, has also been associated with a higher risk of pregnancy complications such as pregnancy-induced hypertension, PTB, impaired glucose tolerance and GDM [19–21,67,97,99,102,103] (Table 2). Comparatively, selenium supplementation was associated with a lower incidence of PE and PROM [104–106]. Importantly, inflammatory indicators such as high sensitivity C-reactive protein were higher in selenium-deficient pregnant women and antioxidant defense indicators namely GPx, catalase, thiobarbituric acid reactive substances (TBARS), superoxide dismutase, and glutathione S-transferase were altered [20,95,99,107,108] (Table 2).
This supports that selenium deficiency may contribute to higher oxidative stress levels, inflammation, and subsequent pregnancy complications (Table 2).

### 3.3. Iodine

Iodine is a vital trace element required for thyroid hormone synthesis and plays a fundamental role in fetal brain development [109,110]. Maternal iodine deficiency during pregnancy is associated with insufficient neurodevelopment, and defective intellectual skills such as poorer school performance and language delay as well as attention-deficit/hyperactivity disorder in offspring later in life [111,112]. While iodine deficiency is described as the most common cause of a child’s central nervous system maldevelopment [113], mandatory iodine fortification programs such as salt iodization have helped address this common health issue among women of child-bearing age globally [114–117]. In Australia, although this preventive program seems to be successful among the general population, 16–44 year old pregnant women’s median urinary iodine concentrations (MUIC: 116 micrograms per litre) still indicate insufficient iodine intake [118]. In New Zealand, MUIC of 68 micrograms per litre indicates a mild iodine deficiency in 18–44 year old women post-mandatory iodine fortification [118]. A similar situation has been reported in other countries with iodine fortification such as Austria, Croatia, Egypt, and Iran, all reporting that pregnant women’s iodine status was insufficient [119]. Pregnancy iodine status should be at an optimal level to avoid the potentially harmful consequences of iodine deficiency [120].

Iodine intake during pregnancy should cover the needs of both the mother and her developing fetus. Thus, it is recommended that dietary intake increases from 150 micrograms per day for a non-pregnant woman to 220 micrograms per day during pregnancy [82]. The World Health Organization recommends a daily iodine supplement of 250 micrograms per day during pregnancy, or an annual dose of iodized oil supplement of 400 milligrams per year [121]. While maternal dietary intake of iodine can impact placental iodine content, and therefore control the effect of iodine on fetal thyroid gland activity [122], placental iodine accumulation plays a significant role in iodine availability to the fetus [123].

In thyroid hormone production, I\(_2\) oxidation occurs to form iodine (I\(_2\)). This reaction uses H\(_2\)O\(_2\) and thyroid peroxidase and inhibits H\(_2\)O\(_2\) accumulation or its decomposition to a hydroxyl radical [25,124]. A normal level of thyroid hormone exerts a negative feedback on thyroid-stimulating hormone (TSH). Iodine deficiency is associated with several health issues including a reduction in thyroid hormone production [125], which results in the absence of the negative feedback on TSH. Therefore, a cascade of signals and reactions, including increasing TSH secretion, occurs [126]. TSH stimulates H\(_2\)O\(_2\) generation for I\(^-\) oxidation but in severe iodine deficiency this process does not occur, resulting in thyroid hormone insufficiency [125,126]. Therefore, TSH continues to increase H\(_2\)O\(_2\) generation, which can be higher than the antioxidant capacity of GPx; H\(_2\)O\(_2\) will accumulate and more ROS is produced, resulting in oxidative stress and apoptosis [126].

Iodide is the ionic state of iodine, occurring when iodine forms a salt with another element, such as potassium. Iodide may have an ancestral antioxidant function in various iodide-concentrating cells not only similar to thyroid cells where iodine consumes H\(_2\)O\(_2\), but also because iodide can reduce the lipid peroxidation rate by reacting with double bonds of the cell membrane polyunsaturated fatty acids and make iodolipids that will be less reactive to ROS [127].
Table 2. Association between maternal selenium levels or selenoproteins and pregnancy complications.

| Ref. | Country Year | Sample Size (Pregnant Women) | Time of Sampling | Sample Type and Assays | Outcomes |
|------|--------------|------------------------------|------------------|------------------------|----------|
| [128] | Norway 2020  | 2638 (2558 term, 80 PTB)    | 17–18 weeks      | Blood Se              | No association between blood Se and PTB risk with adjustment for iodine intake (population had moderate iodine deficiency) |
| [90]  | Indonesia 2019 | 51 (25 term, 26 PTB)      | Delivery          | Serum, placental and cord blood Se | Lower Se in the placenta and cord blood from PTB compared to term deliveries ($p < 0.05$) |
| [91]  | Turkey 2019  | 50 (30 term, 20 PTB)       | Delivery          | Blood, urine and amniotic fluid Se | Lower blood, urine and amniotic fluid Se in PTB compared to term ($p < 0.05$) |
| [92]  | Australia 2018 | 1065 (480 healthy, 585 complicated) | 15 ± 1 weeks      | Plasma Se, C-reactive protein | No difference between complicated and healthy pregnancies. No association between Se and C-reactive protein. |
| [129] | Iran 2017    | 60 at risk for IUGR (30 Se-supplementation, 30 placebo); RCT | Week 17 and end of intervention (week 27) | Plasma total antioxidant capacity, glutathione Serum C-reactive protein | Higher total antioxidant capacity and glutathione and lower C-reactive protein after 100 µg/d Se supplementation |
| [130] | South Africa 2017 | 66 (23 healthy, 43 PE) | Delivery          | Serum and hair Se | Lower Se concentration in PE, no difference in hair Se between healthy and PE |
| [131] | Bangladesh 2015 | 74 PE (52 mild, 22 severe 118 normotensive) | ≥20 weeks         | Serum Se | Lower Se concentration in mild and severe PE, lower Se in severe PE compared to mild PE |
| [132] | Iran 2015    | 65 with GDM (32 Se-supplementation 33 placebo) | Week 24 and end of intervention (week 28) | Plasma MDA and glutathione | Higher glutathione and lower MDA after 200 µg/d Se-supplementation |
| [133] | Australia 2015 | 716 (472 healthy, 244 PE) | 15 ± 1 weeks      | Plasma Se | No difference in plasma Se between healthy and PE |
| [104] | UK 2015      | 230 primiparous 60 µg/d selenium yeast or placebo | (1) 12 and 35 weeks (2) 16 weeks | (1) Whole-blood Se (2) Toenail Se | After Se supplementation, low toenail Se was associated with decreased OR for PE/PIH (OR 0.30, 95% CI 0.09, 1.00, $p = 0.049$) |
| [93]  | Sudan 2014   | 62 (31 healthy,31 GDM)     | Not reported       | Serum Se | No difference in Se level between healthy and GDM pregnancies |
| [106] | UK 2014      | 229 primiparous (115 Se-treated, 114 placebo) 60 µg/d selenium yeast | (1) 12 and 35 weeks (2) 35 weeks | (1) Whole-blood Se (2) selenoprotein P | Higher Se and selenoprotein P in Se-treated group at 35 weeks. Reduced PE/PH odds in all Se-treated participants (OR 0.350, 95% CI 0.126, 0.974; $p = 0.044$) |
| [134] | Iran 2013    | 76 (38 healthy, 38 PE)     | 24 weeks–2 days after delivery | Plasma Se | Lower Se in women with PE compared to healthy pregnancies |
| [135] | UK 2013      | 50 (27 healthy, 23 PE)     | Delivery          | Serum Se, placental deiodinase mRNA and enzyme activity | Lower Se in women with PE, correlation between placental deiodinase III mRNA expression and its enzyme activity only in PE |
| [136] | Indonesia 2013 | 71 (46 healthy, 25 miscarriage) | 8–20 weeks        | Serum Se, GPx | Lower Se in women with miscarriage, no difference in GPx activity |
| [94]  | Iran 2011    | 80 (40 healthy, 40 PE)     | 34–39 weeks       | Plasma Se | Lower Se in women with PE |
| Ref. | Country Year | Sample Size (Pregnant Women) | Time of Sampling | Sample Type and Assays | Outcomes |
|------|--------------|-----------------------------|-----------------|------------------------|----------|
| [19] Netherlands 2011 | 1129 (60 PTB, 21 PROM, 13 PE) | 12 weeks | Serum Se | Higher risk for PTB with lower Se (OR 2.18, 95% CI 1.25–3.77) |
| [105] Iran 2010 | 166 primigravid (83 Se supplement, 83 placebo) | 60 µg/d selenium yeast | 1st trimester and delivery | Serum Se | Increased Se (p < 0.01) and lower incidence of PROM (p < 0.001) in 100 µg/d of selenium yeast-supplemented group |
| [95] UK 2008 | 74 (27 healthy, 25 PE, 22 non-pregnant age-matched) | Delivery | Serum, plasma, umbilical venous Se. Plasma and umbilical venous TBARS. Plasma and placental GPx. | Decreasing trend of plasma Se from non-pregnant to normal pregnant and PE; Lower serum Se and plasma GPx in PE compared to healthy pregnancies; higher level of maternal and umbilical venous TBARS in PE group; Lower placental GPx in PE |
| [20] Hungary 2008 | 61 (20 healthy, 17 GDM, 24 healthy non-pregnant) | 24–28 weeks | Serum Se, high sensitivity C-reactive protein | Negative correlation between serum Se and high-sensitive C-reactive protein |
| [21] Turkey 2008 | 180 (101 healthy, 30 GDM, 49 glucose intolerance) | 24–28 weeks | Serum Se | Lower Se in GDM and glucose intolerance compared to healthy pregnancies |
| [137] Kuwait 2007 | 20 obese (10 GDM, 10 control) | Delivery | Blood Se, GPx, SOD and total antioxidant enzyme activity | Higher SOD activity in maternal vein, umbilical artery and umbilical vein blood of control obese women |
| [96] Turkey 2005 | 85 (32 PE, 28 healthy pregnant, 25 non-pregnant) | 28–39 weeks | Serum and placenta MDA, SOD and catalase in erythrocytes. Placental GSH and GPx. Serum Se | Lower serum Se, erythrocyte SOD activity, and placental GPx and higher erythrocyte catalase activity in PE; negative correlation between placental MDA level and serum Se in PE |
| [67] Italy 2005 | 504 (210 gestational hyperglycemic, 294 normoglycemic) | 24–28 weeks | Serum Se Dietary intake of Se | Lower dietary intake of Se in gestational hyperglycemic women; lower serum Se in women with impaired glucose tolerance; negative association between Se (OR 0.92, 95% CI = 0.87 to 0.95, p < 0.0001) and gestational hyperglycemia |
| [97] Kuwait 2004 | 30 (15 healthy, 15 GDM) | Delivery | Blood Se from maternal vein, umbilical artery and umbilical vein | Lower maternal vein Se in GDM compared to healthy pregnancies |
| Ref. | Country     | Year | Sample Size (Pregnant Women) | Time of Sampling  | Sample Type and Assays                                                                 | Outcomes                                                                                                                                 |
|------|-------------|------|------------------------------|-------------------|----------------------------------------------------------------------------------------|-----------------------------------------------------------------------------------------------------------------------------------------|
| [108]| Turkey      | 2003 | 36 (16 healthy, 9 PIH and PE, 3 IDDP; 3 GDM, 3 OP, 2 PAP) | 3rd trimester and immediately after delivery | GST, GPx and catalase activity and TBARS in maternal erythrocyte, plasma, and umbilical cord blood | Higher erythrocyte GPx activity and increased plasma TBARS in PIH and IDDP; higher cord blood GST activity (2-3 fold) in PE and PIH and IDDP compared to maternal activity before delivery; lower cord blood GPx activity compared to before delivery in PE and PIH; lower cord blood GPx (in PE and PIH, IDDP) and catalase activity (in PE and PIH) compared to maternal values; higher plasma TBARS in PE and PIH and IDDP in the antenatal period; lower cord blood erythrocytic TBARS in PE and PIH compared to maternal value |
| [102]| UK          | 2003 | 106 (53 healthy, 53 PE)      | Not reported      | Toenail Se                                                                             | Lower Se in PE; more severe PE (delivery <32 weeks') with lower Se within the PE group; higher PE risk in the bottom tertile of Se (OR 4.4, 95% CI 1.6–14.9) |
| [103]| China       | 2001 | 251 (98 IGT, 46 GDM, 90 healthy, 17 healthy non-pregnant) | 20 and 42 weeks   | Serum Se                                                                               | Lower Se at 33–42 weeks’ than at 20–33 weeks’ in all pregnant women; lower Se in IGT and GDM; lower Se in healthy pregnancies compared to healthy non-pregnant |

CI, Confidence Interval; GDM, Gestational diabetes mellitus; GPx, Glutathione Peroxidase; GSH, Glutathione; GST, Glutathione S-transferase; Hct, Haematocrit; IDDP, Insulin-Dependent Diabetes mellitus in Pregnancy; IGT, Impaired glucose tolerance; IUGR, Intrauterine growth restriction; MDA, Malondialdehyde; µg/d, micrograms per day; OP, Oligohydramniotic Pregnancy; OR, Odds Ratio; PAP, Pregnancy with Abruptio Placentae; PE, Preeclampsia; PIH, Pregnancy-induced hypertension; PROM, Premature rupture of membrane; PTB, Preterm birth; RBC, Red blood cell; RCT, Randomized controlled trial; Se, selenium; SOE, superoxide dismutase; TBARS, Thiobarbituric acid reactive substances.
3.4. Iodine and Pregnancy Complications

There is limited but supporting evidence that iodine contributes to the antioxidant system. In a small sample of 74 women, there was decreased total antioxidant status and superoxide dismutase activity in women with mild iodine deficiency in the 2nd and 3rd trimesters of pregnancy compared to pregnant women with optimal iodine levels [138]. Iodine sufficient pregnant women, as indicated by more than 150 micrograms per litre urinary iodine concentration, had higher superoxide dismutase enzyme activity compared to iodine-deficient pregnant women [66]. In vitro studies support this and have shown that iodine supplementation reduces ROS production in a dose-dependent manner [139]. While to date there is no study about the potential antioxidant effect of iodine in the placenta, studies have shown a role for iodine in the antioxidant system of other organs such as thyroid, breast, stomach and eye [25,140–143]. We have recently shown in a placental cell line that treatment with iodine resulted in a lower lipid peroxidation compared to control upon induction of oxidative stress [24], further supporting a potential role of iodine as an antioxidant in the placenta.

Importantly, several studies have shown that iodine-deficient pregnant women are at an increased risk of pregnancy complications such as maternal high blood pressure, PE, IUGR and PTB [64–66,138,144,145] (Table 3). Iodine assessment methods can significantly impact the iodine measurement [146]. Iodine intake can be assessed indirectly by thyroid hormone level, urine spot or 24 h urinary samples [146]. Although spot urinary samples in population studies with more than 500 people is a more feasible approach and provides a good estimate of iodine status, it is not a valid measure for individual iodine status; 24 h urine analysis is a more appropriate assessment among individuals or in studies with smaller sample sizes [146]. Adjusting for urinary creatinine to avoid the influence of fluid intake also improves the reliability of the measurement of iodine status [146,147]. However, regardless of the assays used to measure iodine, it is clear that iodine deficiency is associated with pregnancy complications (Table 3). Because complications in pregnancy are exacerbated by increased oxidative stress, the role of iodine as an antioxidant may positively impact pregnant complications.

3.5. Potential Synergistic Effects of Selenium and Iodine

To date, most studies have investigated the impact of micronutrients on oxidative stress in the placenta, separately. Unfortunately, this is unlikely to provide more information on potential interactions or synergistic effects than if they were assessed in combination. In particular, selenium and iodine are essential micronutrients that may affect oxidative stress synergistically. Deiodinases are selenocysteine-containing enzymes that can regulate thyroid hormone bioavailability by removing iodide from different positions on the tyrosine ring. There are various deiodinases in different tissues (deiodinase I, II, III), however type III is dominant in the placenta. It inactivates the T3 (triiodothyronine) and T4 (thyroxine) hormones by removing an inorganic iodine from their inner ring and converting them to T2 (diiodothyronine) and reverse T3 (rT3), respectively [148,149]. Deiodinase type II can produce T3 from T4, and increase the bioavailability of T3 [150]. Thus, the activity of both deiodinase II and deiodinase III leads to the release of iodine in the placenta [151].

Selenoprotein expression in endocrine tissues is precisely controlled to be maintained, even in Se deficiency, and deiodinases seem to be higher than GPxs hierarchically in some tissues [152]. It is also unclear whether there are similar protective mechanisms for deiodinase in the placenta because of the lack of systematic investigation of the placental deiodinases and other selenoproteins. Only one study has reported that placental deiodinase III mRNA expression and its enzyme activity were correlated in PE but not in normotensive pregnant women [135]. In this study, maternal selenium levels were significantly lower in PE compared to normotensive women. This suggests that in normotensive pregnant women where selenium level is optimal, translation of deiodinase III is conserved, while in PE women with low selenium levels, deiodinase III enzyme regulation is altered; therefore iodine metabolism may be affected [135]. However, further studies are required and in larger sample sizes. This will help identify whether and how fetal adaptations occur, in order to maximize iodide uptake.
**Table 3. Iodine status and pregnancy complications.**

| Ref. | Country Year | Sample Size (Pregnant Women) | Time of Sampling | Sample Type and Assays | Outcome |
|------|--------------|------------------------------|------------------|------------------------|---------|
| [153] | UK 2018      | 3182 (3140 with child alive at 1st year, 42 pregnancy/infant loss) | Not reported | Urinary iodine-to-creatinine ratio (spot urine) | No association between iodine status and pregnancy complications or infant loss |
| [69]  | China 2018   | 1569 euthyroid and primapara | 1st trimester | Urinary iodine concentration (spot urine) | Mild iodine deficiency (urinary iodine 100–150 µg/L) was an independent risk factor for GDM (OR 1.669, 95% CI 1.114–2.501, \( p < 0.05 \)). More than adequate and excessive iodine (urinary iodine \( \geq 250 \) µg/L) was an independent risk factor for macrosomia (OR = 2.116, 95% CI 1.218–3.676, \( p < 0.05 \)) |
| [154] | China 2018   | 2347                         | 1st, 2nd, 3rd trimester | Urinary iodine concentration (spot urine) | Lower incidences of PE in pregnant women with UIC 150–249 µg/L compared to the reference group of UIC < 50 µg/L (OR = 0.12, 95% CI 0.01–0.87, \( p < 0.05 \)) |
| [66]  | Mexico 2017  | 57 (37 normotensive, 20 HPD) | 3rd trimester | Urinary iodine concentration (spot urine), SOD, CAT, TBARS | Significant association between iodine deficiency and hypertensive disease of pregnancy (HPD); lower level of urinary iodine, SOD and CAT and higher level of TBARS in HPD compared to normotensive |
| [64]  | Thailand 2016| 390                          | Each trimester | Urinary iodine concentration (spot urine) | Higher PTB risk (OR 2.69, 95% CI 1.38–5.24, \( p = 0.004 \)) and low birthweight (OR 2.66, 95% CI 1.40–5.05, \( p = 0.003 \)) among urinary iodine < 150 µg/L |
| [155] | Argentina 2012| 77                           | Not reported | Urinary iodine concentration (morning and evening urine samples, placental weight, placental index) | Higher risk of lower placental weight in iodine deficiency (urinary iodine < 150 µg/L) (OR 3, 95% CI 1.06–8.5) |
| [156] | Turkey 2010  | 58 (40 severe PE, 18 healthy) | Not reported | Urinary iodine concentration (spot urine), thyroid hormone levels (T3, T4, TSH, fT3, fT4), blood magnesium | Positive correlation between urinary iodine and blood magnesium level in PE; higher T3 and fT3 levels in PE |
| [144] | Turkey 2009  | 40 (24 severe PE, 16 healthy) | Not reported for blood | Serum protein-bound iodine | Lower level of serum protein-bound iodine in maternal blood in PE; higher serum protein-bound iodine level in umbilical cord blood of infants in severe PE |
### Table 3. Cont.

| Ref. | Country Year | Sample Size (Pregnant Women) | Time of Sampling | Sample Type and Assays | Outcome |
|------|--------------|------------------------------|------------------|------------------------|---------|
| [65] | Turkey 2007  | 35 (20 severe PE, 15 healthy) | Not reported for blood | Placental tissue iodine content, blood magnesium | Lower placental iodine in PE; positive correlation between placental iodine and blood magnesium level in PE |
| [145] | Senegal 2000 | 882 (462 pregnant, 420 non-pregnant) | Not reported | Urinary iodine concentration (spot urine), rate of miscarriage and stillbirth | Higher risk of miscarriage and stillbirth in iodine deficiency; highest rate in severe iodine deficiency (urinary iodine ≤ 20 µg/L) (OR 3.64, 95% CI 2.92–4.55) |
| [157] | China 1997  | >60,000 Iodine supplementation to water | Not reported | Neonatal and infant mortality rate after iodine supplementation | Large reduction in both neonatal and infant mortality with iodine supplementation of water among all population in three severely iodine-deficient townships |

CAT, Catalase; CI, Confidence Interval; fT3, Free triiodothyronine; fT4, Free thyroxine; GDM, Gestational diabetes mellitus; IUGR, Intra uterine growth restriction; µg/L, micrograms per litre; OR, Odds Ratio; PE, Preeclampsia; PIH, Pregnancy-induced hypertension; PTB, Preterm birth; SOD, Superoxide dismutase; T3, triiodothyronine; T4, thyroxine; TBARS, Thiobarbituric acid reactive substances; TSH, Thyroid-stimulating hormone; UIC, Urinary iodine concentration.
4. Impact of Maternal Selenium and Iodine, via Oxidative Stress, on Child Health

Maternal deficiencies in selenium or iodine may result in oxidative stress in the placenta, which may impact on future offspring health through developmental origins of health and disease [158–161]. Numerous studies support that an adverse in utero environment contributes to future chronic disease risk in adult offspring (reviewed in [162]) and that this may be mediated by the placenta [159,163].

It is known that during a healthy pregnancy there is a large amount of oxidative stress, especially since the placenta initially develops in a hypoxic environment with maternal blood flow established at approximately 10 weeks of gestation [164]. During this time, reactive oxygen species are produced and the antioxidant system combats this, however, if there are deficiencies in micronutrients involved in this antioxidant system, then it is likely that oxidative stress will occur which will lead to damaged placenta and potentially pregnancy complications. These pregnancy complications are associated with future chronic disease risk in the offspring. Several animal studies have shown that the use of maternal antioxidant supplementation can prevent placental oxidative stress and the associated programming of cardiovascular disease risk to the offspring (reviewed in [165]). Human randomized controlled trials with antioxidants have not shown improvements in pregnancy complications and some have in fact been associated with an increased risk (reviewed in [166]). However, what needs to be considered is the micronutrient status of the mother as deficiencies in elements such as selenium and iodine, which are required for antioxidant enzymes, may diminish the effectiveness of simply adding antioxidant supplements to the maternal diet.

5. Conclusions

This is the first comprehensive review examining the potential synergistic effects of selenium and iodine (Figure 2). In addition, we have discussed whether the association between maternal selenium and iodine status as measured in biological specimens is associated with pregnancy complications due to their roles in oxidative stress. In future, studies that assess maternal dietary intake of selenium and iodine should also be examined. Iodothyronine deiodinases are selenoenzymes involved in thyroid hormone metabolism. The incorporation of selenium into deiodinases causes it to play an essential role in the metabolism of thyroid hormones and in the release of iodide. In addition, iodine deficiencies result in greater production of H$_2$O$_2$, which requires the selenoenzyme GPx to remove the excess H$_2$O$_2$. Thus, selenium and iodine may have some combined effects that should be investigated. Maternal diet is essential for the health of the placenta and baby, and deficiencies in micronutrients impact placental health potentially via oxidative stress pathways. This in turn not only increases the risk of an adverse pregnancy outcome, but is associated with poor future health outcomes for mother and offspring. Supplementation with antioxidants is not necessarily the key if the underlying selenium and iodine levels are low, as antioxidants require these and other micronutrients for optimal activity. Therefore, to address adverse pregnancy outcomes and the impact they have on future offspring health, a better understanding of the role of each micronutrient, alone and in combination, in placental development and hence pregnancy success is needed.
Acknowledgments: We acknowledge support from Claire Roberts, Anthony Perkins and Jo Zhou.

Author Contributions: Conceptualization, N.H., J.A.G. and T.B.-M.; methodology, N.H.; formal analysis, N.H.; investigation, N.H., J.A.G. and T.B.-M.; writing—original draft preparation, N.H.; writing—review and editing, N.H., J.A.G. and T.B.-M; supervision, J.A.G. and T.B.-M. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Acknowledgments: We acknowledge support from Claire Roberts, Anthony Perkins and Jo Zhou.

Conflicts of Interest: The authors declare no conflict of interest.

References
1. Denison, F.C.; Roberts, K.A.; Barr, S.M.; Norman, J.E. Obesity, pregnancy, inflammation, and vascular function. Reproduction 2010, 140, 373–385. [CrossRef] [PubMed]
2. Williams, D. Pregnancy: A stress test for life. Curr. Opin. Obstet. Gynecol. 2003, 15, 465–471. [CrossRef] [PubMed]
3. Lockitch, G. Clinical biochemistry of pregnancy. Crit. Rev. Clin. Lab. Sci. 1997, 34, 67–139. [CrossRef] [PubMed]
4. Soma-Pillay, P.; Nelson-Piercy, C.; Tolppanen, H.; Mebazaa, A. Physiological changes in pregnancy. Cardiovasc. J. Afr. 2016, 27, 89–94. [CrossRef] [PubMed]
5. Myatt, L.; Cui, X. Oxidative stress in the placenta. Histochem. Cell Biol. 2004, 122, 369–382. [CrossRef]
6. Ji, L.; Brkic, J.; Liu, M.; Fu, G.; Peng, C.; Wang, Y.L. Placental trophoblast cell differentiation: Physiological regulation and pathological relevance to preeclampsia. Mol. Asp. Med. 2013, 34, 981–1023. [CrossRef]
7. Wong, F.; Cox, B.J. Cellular analysis of trophoblast and placenta. Placenta 2017, 59 (Suppl. 1), S2–S7. [CrossRef]
8. Raghavan, R.; Dreibelbis, C.; Kingshipp, B.L.; Wong, Y.P.; Abrams, B.; Gernand, A.D.; Rasmussen, K.M.; Siega-Riz, A.M.; Stang, J.; Casavale, K.O.; et al. Dietary patterns before and during pregnancy and maternal outcomes: A systematic review. Am. J. Clin. Nutr. 2019, 109, 705S–728S. [CrossRef]
31. Zaragoza, A.; Diez-Fernandez, C.; Alvarez, A.M.; Andres, D.; Cascales, M. Effect of N-acetylcysteine and defereroxamine on endogenous antioxidant defense system gene expression in a rat hepatocyte model of cocaine cytotoxicity. *Biochim. Biophys. Acta* **2000**, *1496*, 183–195. [CrossRef]
32. Augustin, W.; Wiswedel, I.; Noack, H.; Reinheckel, T.; Reichelt, O. Role of endogenous and exogenous antioxidants in the defence against functional damage and lipid peroxidation in rat liver mitochondria. *Mol. Cell Biochem.* **1997**, *174*, 199–205. [CrossRef] [PubMed]
33. Jeeva, J.S.; Sunitha, J.; Ananthalakshmi, R.; Rajkumari, S.; Ramesh, M.; Krishnan, R. Enzymatic antioxidants and its role in oral diseases. *J. Pharm. Bioallied Sci.* **2015**, *7*, S331–S333. [CrossRef]
34. Leeson, P. Long term cardiovascular outcomes for mother and child. *Pregnancy Hypertens.* **2016**, *6*, 76. [CrossRef]
35. Neiger, R. Long-Term Effects of Pregnancy Complications on Maternal Health: A Review. *J. Clin. Med.* **2017**, *6*, 76. [CrossRef]
36. Padayatty, S.J.; Katz, A.; Wang, Y.; Eck, P.; Kwon, O.; Lee, J.H.; Chen, S.; Corpe, C.; Dutta, A.; Dutta, S.K.; et al. Vitamin C as an antioxidant: Evaluation of its role in disease prevention. *J. Am. Coll. Nutr.* **1999**, *18*, 426–433. [CrossRef] [PubMed]
37. Pisaneschi, S.; Boldrini, A.; Genazzani, A.R.; Coceani, F.; Simoncini, T. Feto-placental vascular dysfunction as a prenatal determinant of adult cardiovascular disease. *Intern. Emerg. Med.* **2013**, *8* (Suppl. 1), S41–S45. [CrossRef]
38. Tranquilli, A.L.; Dekker, G.; Magee, L.; Roberts, J.; Sibai, B.M.; Steyn, W.; Zeeman, G.G.; Brown, M.A. The classification, diagnosis and management of the hypertensive disorders of pregnancy: A revised statement from the ISHHP. *Pregnancy Hypertens.* **2014**, *4*, 97–104. [CrossRef]
39. van Esch, J.J.A.; van Heijst, A.F.; de Haan, A.F.J.; van der Heijden, O.W.H. Early-onset preeclampsia is associated with perinatal mortality and severe neonatal morbidity. *J. Matern.-Fetal Neonatal Med.* **2017**, *29*, 2789–2794. [CrossRef] [PubMed]
40. Wu, C.S.; Nohr, E.A.; Bech, B.H.; Vestergaard, M.; Catov, J.M.; Olsen, J. Health of children born to mothers who had preeclampsia: A population-based cohort study. *Am. J. Obstet. Gynecol.* **2009**, *201*, 269-e1. [CrossRef]
41. Bellamy, L.; Casas, J.P.; Hingorani, A.D.; Williams, D.J. Pre-eclampsia and risk of cardiovascular disease and cancer in later life: Systematic review and meta-analysis. *BMJ* **2007**, *335*, 974. [CrossRef] [PubMed]
42. Ayansina, D.; Black, C.; Hall, S.J.; Marks, A.; Millar, C.; Prescott, G.J.; Wilde, K.; Bhattacharya, S. Long term effects of gestational hypertension and pre-eclampsia on kidney function: Record linkage study. *Pregnancy Hypertens.* **2016**, *6*, 344–349. [CrossRef] [PubMed]
43. Gastrich, M.D.; Zinonos, S.; Bachmann, G.; Cosgrove, N.M.; Cabrera, J.; Cheng, J.Q.; Kostis, J.B. Preeclamptic Women Are at Significantly Higher Risk of Future Cardiovascular Outcomes over a 15-Year Period. *Clin. Nutr. Metab. Care* **2014**, *17*, 249–254. [CrossRef] [PubMed]
44. Roberts, J.M.; Escudero, C. The placenta in preeclampsia. *Pregnancy Hypertens.* **2012**, *2*, 72–83. [CrossRef]
45. Roberts, J.M.; Hubel, C.A. Oxidative stress in preeclampsia. *Am. J. Obstet. Gynecol.* **2004**, *190*, 1177–1178. [CrossRef] [PubMed]
54. Gernand, A.D.; Schulze, K.J.; Stewart, C.P.; West, K.P., Jr.; Christian, P. Micronutrient deficiencies in pregnancy worldwide: Health effects and prevention. *Nat. Rev. Endocrinol* 2016, 12, 274–289. [CrossRef] [PubMed]
55. Parisi, F.; Laoreti, A.; Cetin, I. Multiple micronutrient needs in pregnancy in industrialized countries. *Ann. Nutr. Metab.* 2014, 65, 13–21. [CrossRef]
56. Berti, C.; Biesalski, H.K.; Gartner, R.; Lapillonne, A.; Pietrzik, K.; Poston, L.; Redman, C.; Koletzko, B.; Cetin, I. Micronutrients in pregnancy: Current knowledge and unresolved questions. *Clin. Nutr.* 2011, 30, 689–701. [CrossRef]
57. Perkins, A.V.; Vanderlelie, J.J. Multiple micronutrient supplementation and birth outcomes: The potential importance of selenium. *Placenta* 2016, 48 (Suppl. 1), S61–S65. [CrossRef]
58. Cogswell, M.E.; Parvanta, I.; Ickes, L.; Yip, R.; Brittenham, G.M. Iron supplementation during pregnancy, anemia, and birth weight: A randomized controlled trial. *Am. J. Clin. Nutr.* 2003, 78, 773–781. [CrossRef]
59. Christian, P.; Khattri, S.K.; Katz, J.; Pradhan, E.K.; LeClerq, S.C.; Shrestha, S.R.; Adhikari, R.K.; Sommer, A.; West, K.P., Jr. Effects of alternative maternal micronutrient supplements on low birth weight in rural Nepal: Double blind randomised community trial. *BMJ* 2003, 326, 571. [CrossRef]
60. Smiths, I.J.; Essed, G.G. Short interpregnancy intervals and unfavourable pregnancy outcome: Role of folate depletion. *Lancet* 2001, 358, 2074–2077. [CrossRef]
61. Siega-Riz, A.M.; Savitz, D.A.; Zeisel, S.H.; Thorp, J.M.; Herring, A. Second trimester folate status and preterm birth. *Am. J. Obstet. Gynecol.* 2004, 191, 1851–1857. [CrossRef]
62. Watanabe, H.; Fukuoaka, H.; Sugiyama, T.; Nagai, Y.; Ogasawara, K.; Yoshiike, N. Dietary folate intake during pregnancy and birth weight in Japan. *Eur. J. Nutr.* 2008, 47, 341–347. [CrossRef] [PubMed]
63. Lindblad, B.; Zaman, S.; Malik, A.; Martin, H.; Ekstrom, A.M.; Amu, S.; Holmgren, A.; Norman, M. Folate, vitamin B12, and homocysteine levels in South Asian women with growth-retarded fetuses. *Acta Obstet. Gynecol. Scand.* 2005, 84, 1055–1061. [CrossRef] [PubMed]
64. Bo, S.; Lezo, A.; Menato, G.; Gallo, M.L.; Bardelli, C.; Signorile, A.; Berutti, C.; Massobrio, M.; Pagano, G.F. Gestational hyperglycemia, zinc, selenium, and antioxidant vitamins. *Nutrition* 2005, 21, 186–191. [CrossRef] [PubMed]
65. Kharb, S. Lipid peroxidation in pregnancy with preeclampsia and diabetes. *Gynecol. Obstet. Investig.* 2000, 50, 113–116. [CrossRef]
66. Xia, Y.; Sun, H.; Li, C.; Li, Y.; Peng, S.; Fan, C.; Teng, W.; Shan, Z. Effect of Iodine Nutrition on Pregnancy Outcomes in an Iodine-Sufficient Area in China. *Biol. Trace Elem. Res.* 2018, 182, 231–237. [CrossRef]
67. Nafee, T.M.; Farrell, W.E.; Carroll, W.D.; Fryer, A.A.; Ismail, K.M. Epigenetic control of fetal gene expression. *Br. J. Obstet. Gynaecol.* 2008, 115, 158–168. [CrossRef]
68. Waterland, R.A.; Jirtle, R.L. Early nutrition, epigenetic changes at transposons and imprinted genes, and enhanced susceptibility to adult chronic diseases. *Nutrition* 2004, 20, 63–68. [CrossRef]
69. Labunskyy, V.M.; Hatfield, D.L.; Gladyshev, V.N. Selenoproteins: Molecular pathways and physiological roles. *Physiol. Rev.* 2014, 94, 739–777. [CrossRef] [PubMed]
70. Schchedrina, V.A.; Zhang, Y.; Labunskyy, V.M.; Hatfield, D.L.; Gladyshev, V.N. Structure-function relations, physiologica roles, and evolution of mammalian ER-resident selenoproteins. *Antioxidants Redox Signal.* 2010, 12, 839–849. [CrossRef]
71. Grumolato, L.; Ghzili, H.; Montero-Hadjadje, M.; Gasman, S.; Lesage, J.; Tanguy, Y.; Galas, L.; Ait-Ali, D.; Leprince, J.; Guerineau, N.C.; et al. Selenoprotein T is a PACAP-regulated gene involved in intracellular Ca2+ mobilization and neuroendocrine secretion. *FASEB J.* 2008, 22, 1756–1768. [CrossRef]
72. Steinbrenner, H.; Speckmann, B.; Klotz, L.O. Selenoproteins: Antioxidant selenoenzymes and beyond. *Arch. Biochem. Biophys.* 2016, 595, 113–119. [CrossRef] [PubMed]
73. Arteel, G.E.; Mostert, V.; Oubrahim, H.; Briviba, K.; Abel, J.; Sies, H. Protection by selenoprotein P in human plasma against peroxynitrite-mediated oxidation and nitration. *Biol. Chem.* 1998, 379, 1201–1205. [PubMed]
77. Saito, Y.; Hayashi, T.; Tanaka, A.; Watanabe, Y.; Suzuki, M.; Saito, E.; Takahashi, K. Selenoprotein P in human plasma as an extracellular phospholipid hydroperoxide glutathione peroxidase. Isolation and enzymatic characterization of human selenoprotein p. *J. Biol. Chem.* 1999, 274, 2866–2871. [CrossRef] [PubMed]
78. Labunskyy, V.M.; Hatfield, D.L.; Gladyshev, V.N. The Sep15 protein family: Roles in disulfide bond formation and quality control in the endoplasmic reticulum. *IUBMB Life* 2007, 59, 1–5. [CrossRef] [PubMed]
79. Zeng, H. Selenium as an essential micronutrient: Roles in cell cycle and apoptosis. *Molecules* 2009, 14, 1263–1278. [CrossRef]
80. Thomson, C.D. Assessment of requirements for selenium and adequacy of selenium status: A review. *Eur. J. Clin. Nutr.* 2004, 58, 391–402. [CrossRef]
81. Kipp, A.P.; Strohm, D.; Brigelius-Flohe, R.; Schomburg, L.; Bechthold, A.; Leschik-Bonnet, E.; Heseker, H.; German Nutrition, S. Revised reference values for selenium intake. *J. Trace Elem. Med. Biol.* 2015, 32, 195–199. [CrossRef]
82. National Health and Medical Research Council. Nutrient Reference Values for Australia and New Zealand. Available online: https://www.nrv.gov.au/nutrients (accessed on 14 April 2020).
83. Food and Nutrition Board, Institute of Medicine. *Dietary Reference Intakes for Vitamin C, Vitamin E, Selenium, and Carotenoids; a Report of the Panel on Dietary Antioxidants and Related Compounds, Subcommittees on Upper Level References of Nutrients and of Interpretation and Use of Dietary Reference Intakes, and the Standing Committee on the Scientific Evaluation of Dietary Reference Intakes*; National Academy Press: Washington, DC, USA, 2000.
84. Jones, G.D.; Droz, B.; Greve, P.; Gottschalk, P.; Poffet, D.; McGrath, S.P.; Seneviratne, S.I.; Smith, P.; Winkel, L.H. Selenium deficiency risk predicted to increase under future climate change. *Proc. Natl. Acad. Sci. USA* 2017, 114, 2848–2853. [CrossRef] [PubMed]
85. Shreenath, A.P.; Ameer, M.A.; Dooley, J. Selenium Deficiency. In *Treasure Island (FL)*; StatPearls: Petersburg, FL, USA, 2020.
86. Li, S.; Banuelos, G.S.; Wu, L.; Shi, W. The changing selenium nutritional status of Chinese residents. *Nutrients* 2014, 6, 1103–1114. [CrossRef] [PubMed]
87. Mariath, A.B.; Bergamaschi, D.P.; Rondo, P.H.; Tanaka, A.C.; Hinnig Pde, F.; Abbade, J.F.; Diniz, S.G. The possible role of selenium status in adverse pregnancy outcomes. *Br. J. Nutr.* 2011, 105, 1418–1428. [CrossRef] [PubMed]
88. Xia, Y.; Hill, K.E.; Li, P.; Xu, J.; Zhou, D.; Motley, A.K.; Wang, L.; Byrne, D.W.; Burk, R.F. Optimization of selenoprotein P and other plasma selenium biomarkers for the assessment of the selenium nutritional requirement: A placebo-controlled, double-blind study of selenomethionine supplementation in selenium-deficient Chinese subjects. *Am. J. Clin. Nutr.* 2010, 92, 525–531. [CrossRef] [PubMed]
89. Ambroziak, U.; Hybsier, S.; Shahnazaryan, U.; Krasnodebska-Kiljanska, M.; Rijntjes, E.; Bartoszewicz, Z.; Bednarczuk, T.; Schomburg, L. Severe selenium deficits in pregnant women irrespective of autoimmune thyroid disease in an area with marginal selenium intake. *J. Trace Elem. Med. Biol.* 2017, 44, 186–191. [CrossRef] [PubMed]
90. Irwinda, R.; Wibowo, N.; Putri, A.S. The Concentration of Micronutrients and Heavy Metals in Maternal Serum, Placenta, and Cord Blood: A Cross-Sectional Study in Preterm Birth. *J. Pregnancy* 2019, 2019, 5062365. [CrossRef] [PubMed]
91. Yildirim, E.; Derici, M.K.; Demir, E.; Apaydin, H.; Kocak, O.; Kan, O.; Gorkem, U. Is the Concentration of Cadmium, Lead, Mercury, and Selenium Related to Preterm Birth? *Biol. Trace Elem. Res.* 2019, 191, 306–312. [CrossRef]
92. Wilson, R.L.; Bianco-Miotto, T.; Leemaqz, S.Y.; Roberts, C.T. Early pregnancy maternal trace mineral status and the association with adverse pregnancy outcome in a cohort of Australian women. *J. Trace Elem. Med. Biol.* 2018, 46, 103–109. [CrossRef]
93. Hamdan, H.Z.; Elbashir, L.M.; Hamdan, S.Z.; Elhassan, E.M.; Adam, I. Zinc and selenium levels in women with gestational diabetes mellitus at Medani Hospital, Sudan. *J. Obstet. Gynaecol.* 2014, 34, 567–570. [CrossRef]
94. Maleki, A.; Fard, M.K.; Zadeh, D.H.; Mamegani, M.A.; Abasaizadeh, S.; Mazloomzadeh, S. The relationship between plasma level of Se and preeclampsia. *Hypertens. Pregnancy* 2011, 30, 180–187. [CrossRef]
95. Mistry, H.D.; Wilson, V.; Ramsay, M.M.; Symonds, M.E.; Broughton Pipkin, F. Reduced selenium concentrations and glutathione peroxidase activity in preeclamptic pregnancies. *Hypertension* 2008, 52, 881–888. [CrossRef] [PubMed]
96. Atamer, Y.; Kocyigit, Y.; Yokus, B.; Atamer, A.; Erden, A.C. Lipid peroxidation, antioxidant defense, status of trace metals and leptin levels in preeclampsia. *Eur. J. Obstet. Gynecol. Reprod. Biol.* 2005, 119, 60–66. [CrossRef]

97. Al-Saleh, E.; Nandakumaran, M.; Al-Shammari, M.; Al-Harouny, A. Maternal-fetal status of copper, iron, molybdenum, selenium and zinc in patients with gestational diabetes. *J. Matern.-Fetal Neonatal Med.* 2004, 16, 15–21. [CrossRef] [PubMed]

98. Hawkes, W.C.; Alkan, Z.; Lang, K.; King, J.C. Plasma selenium decrease during pregnancy is associated with glucose intolerance. *Biol. Trace Elem. Res.* 2004, 100, 19–29. [CrossRef]

99. Zachara, B.A.; Dobrzynski, W.; Trafikowska, U.; Szymanski, W. Blood selenium and glutathione peroxidases in miscarriage. *Br. J. Obstet. Gynaecol.* 2001, 108, 244–247. [CrossRef]

100. Ceko, M.J.; O’Leary, S.; Harris, H.H.; Hummitzsch, K.; Rodgers, R.J. Trace Elements in Ovaries: Measurement and Physiology. *Biol. Reprod.* 2016, 94, 86. [CrossRef]

101. Grieger, J.A.; Grzeskowiak, L.E.; Wilson, R.L.; Bianco-Miotto, T.; Leemaqz, S.Y.; Jankovic-Karasoulos, T.; Perkins, A.V.; Norman, R.J.; Dekker, G.A.; Roberts, C.T. Maternal Selenium, Copper and Zinc Concentrations in Early Pregnancy, and the Association with Fertility. *Nutrients* 2019, 11. [CrossRef]

102. Rayman, M.P.; Bode, P.; Redman, C.W. Low selenium status is associated with the occurrence of the pregnancy disease preeclampsia in women from the United Kingdom. *Am. J. Obstet. Gynecol.* 2003, 189, 1343–1349. [CrossRef]

103. Tan, M.; Sheng, L.; Qian, Y.; Ge, Y.; Wang, Y.; Zhang, H.; Jiang, M.; Zhang, G. Changes of serum selenium in pregnant women with gestational diabetes mellitus. *Biol. Trace Elem. Res.* 2001, 83, 231–237. [CrossRef]

104. Rayman, M.P.; Bath, S.C.; Westaway, J.; Williams, P.; Mao, J.; Vanderlelie, J.J.; Perkins, A.V.; Redman, C.W. Selenium status in U.K. pregnant women and its relationship with hypertensive conditions of pregnancy. *Br. J. Nutr.* 2015, 113, 249–258. [CrossRef]

105. Tara, F.; Rayman, M.P.; Boskabadi, H.; Ghayour-Mobarhan, M.; Sahebkar, A.; Yazarlu, O.; Ouladan, S.; Tavallaei, S.; Azimi-Nezhad, M.; Shakeri, M.T.; et al. Selenium supplementation and premature (pre-labour) rupture of membranes: A randomised double-blind placebo-controlled trial. *J. Obstet. Gynaecol.* 2010, 30, 30–34. [CrossRef] [PubMed]

106. Rayman, M.P.; Searle, E.; Kelly, L.; Johnsen, S.; Bodman-Smith, K.; Bath, S.C.; Mao, J.; Redman, C.W. Effect of selenium on markers of risk of pre-eclampsia in UK pregnant women: A randomised, controlled pilot trial. *Br. J. Nutr.* 2014, 112, 99–111. [CrossRef] [PubMed]

107. Dobrzynski, W.; Trafikowska, U.; Trafikowska, A.; Pilecki, A.; Szymanski, W.; Zachara, B.A. Decreased selenium concentration in maternal and cord blood in preterm compared with term delivery. *Analyst* 1998, 123, 93–97. [CrossRef]

108. Orhan, H.; Onderoglu, L.; Yucel, A.; Sahin, G. Circulating biomarkers of oxidative stress in complicated pregnancies. *Arch. Gynecol. Obstet.* 2003, 267, 189–195. [CrossRef]

109. Morreale de Escobar, G.; Obregon, M.J.; Escobar del Rey, F. Role of thyroid hormone during early brain development. *Eur J. Endocrinol* 2004, 151 (Suppl. 3), U25–U37. [CrossRef]

110. Bernal, J. Thyroid hormones and brain development. *Vitam. Horm.* 2005, 71, 95–122. [CrossRef]

111. Abel, M.H.; Brandlilstuen, R.E.; Caspersen, I.H.; Aase, H.; Torheim, L.E.; Meltzer, H.M.; Brantsaeter, A.L. Language delay and poorer school performance in children of mothers with inadequate iodine intake in pregnancy: Results from follow-up at 8 years in the Norwegian Mother and Child Cohort Study. *Eur J. Nutr.* 2019, 58, 3047–3058. [CrossRef]

112. Abel, M.H.; Caspersen, I.H.; Meltzer, H.M.; Haugen, M.; Brandlilstuen, R.E.; Aase, H.; Alexander, J.; Torheim, L.E.; Brantsaeter, A.L. Suboptimal Maternal Iodine Intake Is Associated with Impaired Child Neurodevelopment at 3 Years of Age in the Norwegian Mother and Child Cohort Study. *J. Nutr.* 2017, 147, 1314–1324. [CrossRef]

113. World Health Organization. *Assessment of Iodine Deficiency Disorders and Monitoring Their Elimination: A Guide for Programme Managers*; World Health Organization: Geneva, Switzerland, 2007.

114. Azizi, F.; Sheikholeslam, R.; Hedayati, M.; Mirmiran, P.; Malekafzali, H.; Kimiagar, M.; Pajouhi, M. Sustainable control of iodine deficiency in Iran: Beneficial results of the implementation of the mandatory law on salt iodization. *J. Endocrinol Invest.* 2002, 25, 409–413. [CrossRef]

115. Charlton, K.; Probst, Y.; Kiene, G. Dietary Iodine Intake of the Australian Population after Introduction of a Mandatory Iodine Fortification Programme. *Nutrients* 2016, 8, 701. [CrossRef]
116. Pandav, C.S.; Yadav, K.; Srivastava, R.; Pandav, R.; Karmarkar, M.G. Iodine deficiency disorders (IDD) control in India. *Indian J. Med. Res.* 2013, 138, 418–433. [PubMed]

117. Pearce, E.N.; Andersson, M.; Zimmermann, M.B. Global iodine nutrition: Where do we stand in 2013? *Thyroid* 2013, 23, 523–528. [CrossRef]

118. Australian Institute of Health and Welfare. Monitoring the Health Impacts of Mandatory Folic Acid and Iodine Fortification. Available online: www.aihw.gov.au (accessed on 14 April 2020).

119. Iodine Global Network. Global Scorecard of Iodine Nutrition in 2016. Available online: https://www.ign.org/cm_data/Scorecard_2016_SAC_PW.pdf (accessed on 20 April 2020).

120. Stagnaro-Green, A.; Pearce, E.N. Iodine and pregnancy: A call to action. *Lancet* 2013, 382, 292–293. [CrossRef]

121. World Health Organization. Iodine Supplementation in Pregnant and Lactating Women. Available online: www.who.int/elena/titles/guidance_summaries/iodine_pregnancy/en/ (accessed on 20 April 2020).

122. Burns, R.; Azizi, F.; Hedaya, M.; Mirmiran, P.; O’Herlihy, C.; Smyth, P.P. Is placental iodine content related to dietary iodine intake? *Clin. Endocrinol.* 2011, 75, 261–264. [CrossRef]

123. Burns, R.; O’Herlihy, C.; Smyth, P.P. The placenta as a compensatory iodine storage organ. *Thyroid* 2011, 21, 541–546. [CrossRef]

124. Schomburg, L.; Kohrle, J. On the importance of selenium and iodine metabolism for thyroid hormone biosynthesis and human health. *Mol. Nutr. Food Res.* 2008, 52, 1235–1246. [CrossRef] [PubMed]

125. Zimmermann, M.B.; Boelaert, K. Iodine deficiency and thyroid disorders. *Lancet Diabetes Endocrinol.* 2015, 3, 286–295. [CrossRef]

126. Song, Y.; Driessens, N.; Costa, M.; De Deken, X.; Detours, V.; Corwilain, B.; Maenhaut, C.; Miot, F.; Van Sande, J.; Many, M.C.; et al. Roles of hydrogen peroxide in thyroid physiology and disease. *J. Clin. Endocrinol. Metab.* 2007, 92, 3764–3773. [CrossRef] [PubMed]

127. Venturi, S. Is there a role for iodine in breast diseases? *Breast* 2001, 10, 379–382. [CrossRef]

128. Barman, M.; Brantsaeter, A.L.; Nilsson, S.; Haugen, M.; Lundh, T.; Combs, G.F.; Zhang, G.; Muglia, L.J.; Meltzer, H.M.; Jacobsson, B.; et al. Maternal dietary selenium intake is associated with increased gestational length and decreased risk of preterm delivery. *Br. J. Nutr.* 2020, 123, 209–219. [CrossRef] [PubMed]

129. Mesdaghinia, E.; Rahavi, A.; Bahmani, F.; Sharifi, N.; Asemi, Z. Clinical and Metabolic Response to Selenium Supplementation in Pregnant Women at Risk for Intrauterine Growth Restriction: Randomized, Double-Blind, Placebo-Controlled Trial. *Biomed. Trace Elem. Res.* 2017, 178, 14–21. [CrossRef] [PubMed]

130. Maduray, K.; Moodley, J.; Soobramoney, C.; Moodley, R.; Naicker, T. Elemental analysis of serum and hair subjects. *J. Trace Elem. Med. Biol.* 2013, 2678 20 of 22, 106–113. [CrossRef]

131. Asemi, Z.; Jamilian, M.; Mesdaghinia, E.; Esmaillzadeh, A. Effects of selenium supplementation on glucose homeostasis, inflammation, and oxidative stress in gestational diabetes: Randomized, double-blind, placebo-controlled trial. *Nutrition* 2015, 31, 1235–1242. [CrossRef] [PubMed]

132. Mistry, H.D.; Gill, C.A.; Kurlak, L.O.; Seed, P.T.; Hesketh, J.E.; Meplan, C.; Schomburg, L.; Chappell, L.C.; Morgan, L.; Poston, L.; et al. Association between maternal micronutrient status, oxidative stress, and common genetic variants in antioxidant enzymes at 15 weeks gestation in nulliparous women who subsequently develop preeclampsia. *Free Radic. Biol. Med.* 2015, 78, 147–155. [CrossRef] [PubMed]

133. Ghaemi, S.Z.; Forouharni, S.; Dabbaghmanesh, M.H.; Sayadi, M.; Bakbshayeshkaram, M.; Vaziri, F.; Tavana, Z. A prospective study of selenium concentration and risk of preeclampsia in pregnant Iranian women: A nested case-control study. *Biomed. Trace Elem. Res.* 2013, 123, 174–179. [CrossRef]

134. Ghaemi, S.Z.; Forouharni, S.; Dabbaghmanesh, M.H.; Sayadi, M.; Bakbshayeshkaram, M.; Vaziri, F.; Tavana, Z. A prospective study of selenium concentration and risk of preeclampsia in pregnant Iranian women: A nested case-control study. *Biomed. Trace Elem. Res.* 2013, 152, 174–179. [CrossRef]

135. Kurlak, L.O.; Mistry, H.D.; Kaptein, E.; Visser, T.J.; Broughton Pipkin, F. Thyroid hormones and their placental deiodination in normal and pre-eclamptic pregnancy. *Placenta* 2013, 34, 395–400. [CrossRef]

136. Abdulah, R.; Noerjasin, H.; Septiani, L.; Defi, I.R.; Suradji, E.W.; Puspitasari, I.M.; Barliana, M.I.; Yamazaki, C.; Nakazawa, M.; Koyama, H. Reduced serum selenium concentration in miscarriage incidence of Indonesian subjects. *Biomed. Trace Elem. Res.* 2013, 154, 1–6. [CrossRef]

137. Al-Saleh, E.; Nandakumaran, M.; Al-Rashdan, I.; Al-Harmi, J.; Al-Shammari, M. Maternal-foetal status of copper, iron, molybdenum, selenium and zinc in obese gestational diabetic pregnancies. *Acta Diabetol.* 2007, 44, 106–113. [CrossRef]
138. Vidal, Z.E.; Rufino, S.C.; Tlaxcalteco, E.H.; Trejo, C.H.; Campos, R.M.; Meza, M.N.; Rodriguez, R.C.; Arroyo-Helguera, O. Oxidative stress increased in pregnant women with iodine deficiency. *Biol. Trace Elem. Res.* 2014, 157, 211–217. [CrossRef]

139. Olivo-Vidal, Z.E.; Rodriguez, R.C.; Arroyo-Helguera, O. Iodine Affects Differentiation and Migration Process in Trophoblastic Cells. *Biol. Trace Elem. Res.* 2016, 169, 180–188. [CrossRef]

140. Elstner, E.F.; Adamczyk, R.; Kroner, R.; Furch, A. The uptake of potassium iodide and its effect as an antioxidant in isolated rabbit eyes. *Ophthalmologica* 1985, 191, 122–126. [CrossRef]

141. Rieger, G.; Winkler, R.; Buchberger, W.; Moser, M. Iodine distribution in a porcine eye model following iophterhesis. *Ophthalmologica* 1995, 209, 84–87. [CrossRef]

142. Vejbjerg, P.; Knudsen, N.; Perrild, H.; Laurberg, P.; Andersen, S.; Rasmussen, L.B.; Ovesen, L.; Jorgensen, T. Estimation of iodine intake from various urinary iodine measurements in population studies. *Thyroid* 2009, 19, 1281–1286. [CrossRef]

143. Maier, J.; van Steeg, H.; van Oostrom, C.; Paschke, R.; Weiss, R.E.; Krohn, K. Iodine deficiency activates antioxidant genes and causes DNA damage in the thyroid gland of rats and mice. *Biochim. Biophys. Acta* 2007, 1773, 990–999. [CrossRef]

144. Borecki, B.; Gulaboglu, M.; Gu, M. Iodine and magnesium levels in maternal and umbilical cord blood of preeclamptic and normal pregnant women. *Biol. Trace Elem. Res.* 2009, 129, 1–8. [CrossRef]

145. Dillon, J.C.; Milliez, J. Reproductive failure in women living in iodine deficient areas of West Africa. *Br. J. Obstet. Gynaecol.* 2000, 107, 631–636. [CrossRef]

146. Gulaboglu, M.; Borekci, B.; Delibas, I. Urine iodine levels in preeclamptic and normal pregnant women. *Biol. Trace Elem. Res.* 2010, 136, 249–257. [CrossRef]

147. DeLong, G.R.; Leslie, P.W.; Wang, S.H.; Jiang, X.M.; Zhang, M.L.; Rakeman, M.; Jiang, J.Y.; Ma, T.; Cao, X.Y. Effect on infant mortality of iodination of irrigation water in a severely iodine-deficient area of China. *Lancet* 1997, 350, 771–773. [CrossRef]

148. Barker, D.J.; Gluckman, P.D.; Godfrey, K.M.; Harding, J.E.; Owens, J.A.; Robinson, J.S. Fetal nutrition and cardiovascular disease in adult life. *Lancet* 1993, 341, 938–941. [CrossRef]

149. Barker, D.J.; Gluckman, P.D.; Godfrey, K.M.; Harding, J.E.; Owens, J.A.; Robinson, J.S. Placental adaptations to micronutrient dysregulation in the programming of chronic disease. *Clin. Exp. Pharmacol. Physiol.* 2018, 45, 871–884. [CrossRef]
160. Hofstee, P.; Bartha, L.A.; McKeating, D.R.; Radenkovic, F.; McEnroe, G.; Fisher, J.J.; Holland, O.J.; Vanderlelie, J.J.; Perkins, A.V.; Cuffe, J.S.M. Maternal selenium deficiency during pregnancy in mice increases thyroid hormone concentrations, alters placental function and reduces fetal growth. J. Physiol. 2019, 597, 5597–5617. [CrossRef] [PubMed]

161. Hofstee, P.; McKeating, D.R.; Bartha, L.A.; Anderson, S.T.; Perkins, A.V.; Cuffe, J.S.M. Maternal Selenium Deficiency in Mice Alters Offspring Glucose Metabolism and Thyroid Status in a Sexually Dimorphic Manner. Nutrients 2020, 12, 267. [CrossRef] [PubMed]

162. Hsu, C.N.; Tain, Y.L. The Good, the Bad, and the Ugly of Pregnancy Nutrients and Developmental Programming of Adult Disease. Nutrients 2019, 11, 894. [CrossRef] [PubMed]

163. Chavatte-Palmer, P.; Tarrade, A.; Rousseau-Ralliard, D. Diet before and during Pregnancy and Offspring Health: The Importance of Animal Models and What Can Be Learned from Them. Int. J. Environ. Res. Public Health 2016, 13, 586. [CrossRef] [PubMed]

164. Burton, G.J.; Cindrova-Davies, T.; Yung, H.W.; Jauniaux, E. Oxygen and development of the human placenta. Reproduction 2020. [CrossRef]

165. Aljunaidy, M.M.; Morton, J.S.; Cooke, C.M.; Davidge, S.T. Prenatal hypoxia and placental oxidative stress: Linkages to developmental origins of cardiovascular disease. Am. J. Physiol. Regul. Integr. Comp. Physiol. 2017, 313, R395–R399. [CrossRef] [PubMed]

166. Morton, J.S.; Cooke, C.L.; Davidge, S.T. In Utero Origins of Hypertension: Mechanisms and Targets for Therapy. Physiol. Rev. 2016, 96, 549–603. [CrossRef]