**α-Synuclein structural features inhibit harmful polyunsaturated fatty acid oxidation, suggesting roles in neuroprotection**

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**α-Synuclein (aS)** is a cytosolic protein abundant in presynaptic nerve terminals in Parkinson disease (PD) and is a major component of intracellular Lewy bodies, the pathological hallmark of neurodegenerative disorders such as PD. Accordingly, the relationships between aS structure, its interaction with lipids, and its involvement in neurodegeneration have attracted great interest. Previously, we reported on the interaction of aS with brain polyunsaturated fatty acids, in particular docosahexaenoic acid (DHA). aS acquires an α-helical secondary structure in the presence of DHA and, in turn, affects DHA structural and aggregative properties. Moreover, aS forms a covalent adduct with DHA. Here, we provide evidence that His-50 is the main site of this covalent modification. To better understand the role of His-50, we analyzed the effect of DHA on aS-derived species: a naturally occurring variant, H50Q; an oxidized aS in which all methionines are sulfoxides (aS4ox); a fully lysine-alkylated aS (acetyl-aS); and aS fibrils, testing their ability to be chemically modified by DHA. We show, by mass spectrometry and spectroscopic techniques, that H50Q and aS4ox are modified by DHA, whereas acetyl-aS is not. We correlated this modification with aS structural features, and we suggest a possible functional role of aS in sequestering the early peroxidation products of fatty acids, thereby reducing the level of highly reactive lipid species. Finally, we show that fibrillar aS loses almost 80% of its scavenging activity, thus lacking a potentially protective function. Our findings linking aS scavenging activity with brain lipid composition suggest a possible etiological mechanism in some neurodegenerative disorders.

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The abbreviations used are: aS, α-synuclein; PD, Parkinson’s disease; NAC, non-Aβ component; PUFA, polyunsaturated fatty acid; DHA, Docosahexaenoic acid; AA, arachidonic acid; 4HNE, 4-hydroxy-2-nonenal; ESI, electrospray ionization.
radical-mediated mechanism (16–18). Each oxidation mechanism yields specific products, in particular hydroperoxides, hydroxide positional isomers, and free radicals that are highly reactive (19). The relative susceptibility of PUFAs to oxidation depends on the reaction milieu as well as their inherent structure. The biological role of PUFAs peroxidation products has received a great deal of attention because when it was discovered that α,β-unsaturated aldehydes, the advanced lipid peroxidation end products, can react with sulfhydryl groups of proteins, forming stable adducts able to affect several metabolic processes (20).

Lipid peroxidation is involved in several human diseases such as atherosclerosis, cancer, diabetes, acute lung injury, as well as neurodegenerative disorders including PD (21), Huntington disease (22), and Alzheimer disease (23). Therefore, efforts have been devoted to understanding the mechanism of lipid peroxidation and preventing the deleterious effects of this process. Interestingly, it was shown that 4-hydroxy-2-nonenal (4HNE), a product of lipid peroxidation, forms protein adducts in Lewy bodies in neocortical and brain neurons (24). 4HNE was also found to alter dopamine transport, contributing to the loss of bodies in neocortical and brain neurons (24). 4HNE was also found to alter dopamine transport, contributing to the loss of

In the present work, we show by mass spectrometry analysis that the early radical products of DHA and AA autoxidation react with aS, producing a covalent modification on the protein. Histidine (His) at position 50 seems to be the major target of this reaction and the four Met are oxidized. To understand if the role of His would be specific in this sort of scavenger activity of free radicals, we analyze DHA in the presence of a pathogenic aS variant presenting the H50Q missense mutation (34–36).

Our in vitro data suggest that aS might be able to exert a sequestering/scavenging activity versus DHA and AA early radical autoxidation products that is modulated by different factors. First, the mutual structural interaction between aS and PUFAs is an essential prerequisite for this activity, because it provides a stabilizing effect on the fatty acid physical state. Second, His and Lys collaborate in sequestering the early peroxidation products. In this way, the propagation of the oxidative process of fatty acids is hampered, and concentration of the highly reactive species is reduced.

### Results

#### DHA and AA autoxidation process

DHA and AA samples were incubated at pH 7 and 37 °C under shaking up to 24 h, in the absence and presence of aS (P/lipid molar ratio 1:50). Aliquots from DHA samples were analyzed by direct injection in electrospray ionization (ESI)-MS (Fig. 2). The ESI-MS spectrum of DHA just after its solubilization is reported in Fig. 2A. It presents a specific pattern of several signals (37): the most peculiar are those at mass/charge values of 329.2, 351.2, and 367.2 corresponding to a protonated molecular DHA (monoisotopic mass 328.2 Da), an adduct with sodium (328.2 Da + 22), and an oxidized form of this adduct (328.2 Da + 22 + 16). Moreover, other minor signals due to in-cone fragmentation (m/z 311.2, 269.2) are also visible. After 24 h of incubation (Fig. 2B), the signals at m/z 329.2, 351.2, and 367.2, characteristics of DHA, disappear and new species are evident in the spectrum at m/z values of 381.3, 397.2, and 415.2 corresponding to hydroperoxide derivatives of DHA, which generate during the propagation phase of the fatty acids autoxidation (Scheme 1). The species at m/z 415.2 is the most intense signal and it has been identified by MS-MS analysis, as a dihydroperoxide derivative of DHA (Table 1). Then the fatty acid was incubated in the presence of aS recording the mass spectra up to 24 h (Fig. 2D). The 24-h spectrum is quite similar to that obtained at time 0 (Fig. 2A and C) and the oxidized species do not form in the presence of the protein. As control, DHA upon

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**Oxidative modifications of α-synuclein**

![Figure 1. Amino acid sequence of aS. Schematic representation of the aS sequence highlighting methionine, lysine, and histidine residues in red, blue, and green, respectively (top). Scheme of the three major domains of the protein: the N-terminal region involved in lipid and membrane binding, the NAC domain (61–95) responsible for aggregation properties of the protein, and the acidic C-terminal region, able to transiently interact with the N-terminal and the NAC domains modulating the aggregation propensity of the protein (bottom).](image-url)
Oxidative modifications of α-synuclein

**Table 1**

| MH<sup>+</sup> | Species | Description |
|---------------|---------|-------------|
| 329.2 | DHA (C<sub>22</sub>H<sub>32</sub>O<sub>2</sub>) | Docosahexaenoic acid (monoisotopic mass 328.2) |
| 351.2 | DHA + Na | Sodium adduct of DHA |
| 367.2 | DHAOH + Na | Sodium adduct of hydroxy DHA |
| 381.3 | DHAAOO + Na | Sodium adduct of peroxo radical DHA or DHA(O'<sup>·</sup>) + Na |
| 397.2 | DHA(OO')<sup>·</sup> + Na | Sodium adduct of alkoxy-peroxy radical DHA or DHA(OO'<sup>·</sup>) + Na |
| 415.2 | DHA(OO')<sub>2</sub> + Na | Sodium adduct of dihydroperoxy DHA |
| 395.3 | AA (C<sub>20</sub>H<sub>28</sub>O<sub>2</sub>) | Arachidonic acid (monoisotopic mass 304.2) |
| 327.3 | AA + Na | Sodium adduct of AA |
| 343.3 | AAOH + Na | Sodium adduct of hydroxy AA |
| 349.3 | AA + 2Na | Di-sodium adduct of AA |
| 373.3 | AA(OOH)<sup>·</sup> + Na + 18 | Sodium adduct of dihydroperoxy AA dehydrated |
| 391.3 | AA(OOH)<sup>·</sup> + Na | Sodium adduct of dihydroperoxy AA |
| 413.3 | AA(OOH)<sub>2</sub> + 2Na | Di-sodium adduct of dihydroperoxy AA |

**Table 2**

| Molecular mass (Da) | PUFA | Found<sup>a</sup> | Calculated<sup>b</sup> | Protein species |
|---------------------|------|------------------|-----------------------|----------------|
| 14459.9 (±0.1)     | aS   | 14460.1          | aS + 10x              |
| 14475.7 (±0.8)     | aS   | 14476.1          | aS + 20x              |
| 14492.2 (±0.1)     | aS   | 14492.1          | aS + 30x              |
| 14508.3 (±0.1)     | aS   | 14508.1          | aS + 40x              |
| 14802.2 (±0.1)     | aS   | 14802.1          | aS + 10x + 326        |
| 14817.1 (±0.7)     | aS   | 14818.1          | aS + 20x + 326        |
| 14835.3 (±0.5)     | aS   | 14834.1          | aS + 30x + 326        |
| 14849.9 (±0.7)     | aS   | 14850.1          | aS + 40x + 326        |
| 14865.2 (±0.9)     | aS   | 14866.1          | aS + 30x + 358        |
| 14880.8 (±0.9)     | aS   | 14882.1          | aS + 40x + 358        |
| 14747.2 (±1.4)     | aS   | 14746.1          | aS + 286              |
| 14765.1 (±0.8)     | aS   | 14764.1          | aS + 304              |
| 14781.2 (±0.9)     | aS   | 14780.1          | aS + 10x + 304        |
| 14794.8 (±0.3)     | aS   | 14796.1          | aS + 20x + 304        |

<sup>a</sup> Experimental molecular masses determined by ESI-MS.  
<sup>b</sup> Masses calculated from the amino acid sequence of aS.

hydroxyl form are evident (m/z 305.3, 327.3, and 343.3). Also the products of partial degradation of AA (37) at m/z 259.2, 269.2, and 287.2 are visible. After 24 h of incubation, AA forms several species including the di-hydroperoxy derivative (m/z 391.3), a sign of autoxidation process (Fig. 2F). In Fig. 2, G and H, the same spectra are recorded in the presence of aS, showing that in this case the early autoxidation products do not form within 24 h of incubation with the protein. In Table 1, the species of DHA and AA identified by MS analysis are listed.

**Chemical modification of aS**

The mass analysis of aS samples analyzed in the presence of DHA or AA are reported in Table 2. In particular, species containing oxidations (+16 Da or multiple of 16) and covalent adducts with DHA (+326 Da) or AA (+304 Da) are present. For the purpose of the present work, the samples were analyzed only up to 24 h of incubation, because aS aggregates and forms oligomers upon prolonged incubation in the presence of fatty acids, as previously observed, and other modifications can occur (29).

To identify the site of modification in the aS sequence, fingerprinting analysis by trypsin and endoproteinase GluC was performed. These proteases selectively cleave the peptide bonds involving basic and glutamic acid residues, respectively (Fig. 3). A RP-HPLC fraction enriched of the chemically modified aS with DHA was digested by trypsin, using an enzyme to substrate ratio (E/S) of 1:100, or by GluC, using an E/S of 1:50.

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24 h at 37 °C in the presence of aS added immediately before the MS analysis, was analyzed to show that the higher molecular weight DHA species are still detectable.

The same experiment was conducted using AA as a fatty acid. In Fig. 2E, the m/z spectrum of AA is reported, where the signals corresponding to molecular AA, its sodium adduct, and its

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The proteolytic fragments were purified by RP-HPLC (Fig. 3, A and B) and identified by MS (Tables 3 and 4). In the tryptic mixture, fragments 1–6, 1–60, 81–140, and 98–140, containing Met residues are present also in oxidized form (+16 Da), whereas fragments corresponding to sequences 1–60, 35–80, 46–80, and 46–58 were found with a mass increase of 326 Da. The residues, susceptible of modification, present in all the detected modified peptides are His and Lys. His as a nucleophile is stronger than Lys, and it is consequently more prone to be modified. To confirm this hypothesis, MS-MS analysis of the modified fragments was performed. In Fig. 3C, the MS-MS spectrum of the peptide-(46–58) + 326, the shortest modified peptide, is reported. The analysis was conducted on its double charged ion at m/z 811.4. The spectrum does not show signals relative to the series of y- or b-ions, but products of partial degradation (m/z 754.33, 762.95, and 781.89) together with a strong ion at m/z 648.39 corresponding to the double charged ion of the peptide-(46–58) without the covalent adduct (Fig. 3C). The species at m/z 781.89 is particularly interesting, because it is produced for removal of the moiety CH$_2$COOH (59 Da), partially confirming the identity of the modified peptide. From the other hand, it was not possible to obtain direct evidence of which specific amino acid has reacted with the fatty acid, because the energy used for the collision of the double charged ions produces ions corresponding to the peptide without modification (m/z 648.39). Indeed, the MS-MS spectrum of the ion at m/z 648.39 confirms the identity of the peptide (Fig. 3D). In the proteolytic mixture of modified aS with DHA by GluC, an increase of 326 Da was found on the mass of peptides corresponding to 36–61 and 29–57 after a 5-min reaction (data not shown) and to 47–57 after a 70-min reaction (Fig. 3B, Table 4). This last peptide, which does not contain Lys, provides direct evidence that His-50 is modified, even if, also in this case, a MS-MS spectrum was not obtained. Moreover, no fragments containing only Lys (and not His) were found modified.

Interaction of H50Q, aS-4ox, and acetyl-aS with DHA

To understand the specific role of His as a site of modification of aS in the presence of fatty acids, the interaction with DHA was studied testing the variant in which the only His residue in position 50 is replaced by Gln. The role of Met was scrutinized by using aS with oxidized Met (tetraoxidized aS, aS4ox). Acetylation was used to abrogate the effect of the 15 Lys (acetyl-aS). In Fig. 4, the m/z spectra of DHA after 24 h of incubation in the presence of these three aS species are reported. They show that DHA early autoxidation products are not present if the fatty acid is incubated with H50Q (Fig. 4A) and aS4ox (Fig. 4B), which in turn are chemically modified by DHA and oxidized aS4ox (Fig. 4C).
The modified peptides are highlighted in bold.

Table 3
Chemical characterization of fragments corresponding to the peaks of the chromatogram relative to the fragmentation of aS/DHA with trypsin shown in Fig. 4A

| RT (min) | Molecular mass (Da) | Peptide species |
|---------|---------------------|-----------------|
| 11.2    | 830.43 (±0.01)     | 292.05          |
| 11.3    | 1294.99 (±0.05)    | 247.02          |
| 11.4    | 1524.16 (±0.01)    | 247.02          |
| 11.5    | 1620.8 (±0.1)      | 247.02          |
| 11.6    | 1754.37 (±0.02)    | 247.02          |
| 11.7    | 1179.93 (±0.05)    | 247.02          |
| 11.8    | 1406.9 (±0.1)      | 247.02          |
| 14.1    | 785.88 (±0.11)     | 247.02          |
| 14.2    | 801.78 (±0.12)     | 247.02          |
| 14.3    | 950.7 (±0.1)       | 247.02          |
| 14.4    | 2157.17 (±0.04)    | 247.02          |
| 14.5    | 769.88 (±0.1)      | 247.02          |
| 14.6    | 1607.16 (±0.01)    | 247.02          |
| 15.1    | 2148.14 (±0.11)    | 247.02          |
| 15.2    | 1928.41 (±0.05)    | 247.02          |
| 15.3    | 1478.05 (±0.04)    | 247.02          |
| 15.4    | 4923.16 (±0.1)     | 247.02          |
| 15.5    | 4847.06 (±0.38)    | 247.02          |
| 15.6    | 6159.38 (±0.1)     | 247.02          |
| 15.7    | 6167.3 (±0.5)      | 247.02          |
| 15.8    | 6472.55 (±0.75)    | 247.02          |
| 15.9    | 4958.66 (±0.01)    | 247.02          |
| 15.10   | 4288.74 (±0.34)    | 247.02          |
| 15.11   | 4300.5 (±0.1)      | 247.02          |
| 19.3    | 6434.93 (±0.04)    | 247.02          |
| 19.4    | 6451.22 (±0.46)    | 247.02          |
| 19.5    | 6419.65 (±0.13)    | 247.02          |
| 19.6    | 3434.52 (±0.05)    | 247.02          |
| 19.7    | 3760.8 (±0.1)      | 247.02          |
| 19.8    | 3664.32 (±0.01)    | 247.02          |

* Peptides are listed in order of retention times (RT).
* Experimental molecular masses determined by ESI-MS.
* Molecular masses calculated from aS amino acid sequence.
* The modified peptides are highlighted in bold.

Table 4
Chemical characterization of peptide species corresponding to the peaks of the chromatogram relative to the fragmentation of aS/DHA with GluC, after 70-min incubation, shown in Fig. 4B

| RT (min) | Molecular mass (Da) | Peptide species |
|---------|---------------------|-----------------|
| 4.5     | 703.21 (±0.01)     | 292.05          |
| 5.2     | 607.2 (±0.02)      | 292.05          |
| 10.8    | 859.52 (±0.01)     | 292.05          |
| 11.7    | 970.36 (±0.01)     | 292.05          |
| 12.5    | 1029.61 (±0.01)    | 292.05          |
| 12.6    | 1045.72 (±0.02)    | 292.05          |
| 12.7    | 1544.85 (±0.01)    | 292.05          |
| 12.8    | 1998.61 (±0.02)    | 292.05          |
| 12.9    | 1457.01 (±0.01)    | 292.05          |
| 13.2    | 954.35 (±0.01)     | 292.05          |
| 13.3    | 1069.62 (±0.01)    | 292.05          |
| 13.4    | 1515.7 (±0.1)      | 292.05          |
| 13.5    | 1982.8 (±0.01)     | 292.05          |
| 13.6    | 2022.8 (±0.1)      | 292.05          |
| 13.7    | 1717.9 (±0.02)     | 292.05          |
| 13.8    | 2142.2 (±0.1)      | 292.05          |
| 13.9    | 912.32 (±0.03)     | 292.05          |
| 14.2    | 1037.56 (±0.01)    | 292.05          |
| 14.3    | 2006.69 (±0.02)    | 292.05          |
| 14.4    | 2112.08 (±0.01)    | 292.05          |
| 14.5    | 1363.62 (±0.02)    | 292.05          |
| 14.6    | 2189.82 (±0.51)    | 292.05          |
| 14.7    | 3083.89 (±0.03)    | 292.05          |
| 14.8    | 2504.04 (±0.05)    | 292.05          |
| 14.9    | 2127.16 (±0.01)    | 292.05          |
| 15.0    | 3633.11 (±0.11)    | 292.05          |
| 15.1    | 4795.66 (±0.02)    | 292.05          |

* Peptides are listed in order of retention times (RT).
* Experimental molecular masses determined by ESI-MS.
* Molecular masses calculated from aS amino acid sequence.
* The modified peptides are highlighted in bold.

Figure 4. Mass/charge spectra of DHA in the presence of H50Q, aS4ox, and acetyl-aS. Positive ESI-MS spectra of DHA recorded upon incubation for 24 h at 37 °C and 300 rpm in the presence of H50Q (A), tetraoxidized aS (aS4ox, B), and alkylated aS (acetyl-aS, C). Proteins were added to realize a protein/lipid ratio of 1:50. Measurements are conducted as described in the legend to Fig. 2.

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Table 5
Molecular masses of H50Q, aS4ox, and acetyl-aS in the presence of DHA

| Protein       | Molecular mass (Da) | Protein species |
|---------------|---------------------|----------------|
| H50Q          | 14455.1 + 0.1       | H50Q           |
| aS4ox         | 14524.1 + 0.7       | aS4ox          |
| Acetyl-aS     | 14966.4 + 1.1       | 12 acetyl      |

* Experimental molecular masses determined by ESI-MS.
* Molecular masses calculated from the amino acid sequence of aS species.

ified (see “Discussion”). Thus, lacking the preferential site of modification (His-50), it is reasonable to expect more than one site to undergo modification with DHA, because potentially all of the 15 Lys can be modified with the same chemical propensity. To identify the site(s) of modification, a RP-HPLC fraction enriched of the chemically modified H50Q with DHA was digested by using endoproteinase GluC, using an enzyme to substrate ratio of 1:50. The mixture was analyzed by RP-HPLC (Fig. 5). As in the case of aS/DHA, peptides that contain Met are present also in oxidized form (Table 6). The smallest fragment that has been found modified by DHA corresponds to the sequence 58–83 (found mass 2939.7 Da, calculated 2613.98 + 326 Da). This species contains 3 Lys residues (in position 58, 60, and 80). Lys in position 60 is reported to be, after His-50, the target of the reaction with 4HNE (38). However, we detected more than one adduct with DHA, therefore different lysines can be involved.

Structural interaction with the fatty acids

Far UV-CD measurements were performed (Fig. 6), to correlate the ability of the protein to be modified with their structural features deriving from the interaction with DHA. H50Q and aS4ox maintain the property observed in aS to undergo structural transition acquiring the α-helical secondary structure in the presence of the polyunsaturated fatty acid (Fig. 6, A–C). Indeed, the characteristic signals at 208 and 222 nm in the far UV-CD spectra of the protein species in the presence of DHA are visible. The acetylation of lysine residues and the N-terminal amino group cause the loss of this ability and acetyl-aS remains unfolded in the presence of the fatty acid (Fig. 6D). Interestingly, this correlates with the loss of scavenger activity observed in Fig. 4C, whereas all the other interacting species maintain their ability to slow down DHA oxidation.

DHA and aS fibrils

MS spectra of DHA were recorded in the presence of preformed WT aS fibrils (Fig. 7A). After 24 h of incubation (Fig. 7B), the mass/charge signals at 329.2, 351.2, and 367.2 are still evident, but the presence of the signal at m/z 415.2 shows that DHA undergoes partial oxidation. aS fibrils were further oxidized as observed by MS analysis (not shown).

Figure 5. Fingerprinting analysis of modified H50Q by GluC. RP-HPLC analysis of the proteolytic mixture of modified H50Q with DHA. The analysis was conducted on a Vydac C18 column (4.6 × 250 mm); The Separations Group, Hesperia, CA), eluted with a gradient of water, 0.1% TFA versus acetonitrile, 0.085% TFA from 5 to 25% in 5 min, from 25 to 28% in 13 min, from 28 to 39% in 3 min, and from 39 to 45% in 21 min at a flow rate of 1 ml/min. The numbers close to the peaks refer to the main proteolytic fragments. The sequence of the peptide containing the chemical modification is reported.

Table 6
Chemical characterization of fragments corresponding to the peaks of the chromatogram relative to the fragmentation of H50Q/DHA with GluC shown in Fig. 5

| RT (min) | Found (Da) | Calculated (Da) | Peptide species |
|---------|------------|-----------------|----------------|
| 10.6    | 859.5 + 0.1 | 859.48          | 21–28          |
| 11.6    | 1028.4 + 0.1 | 1028.55         | 47–57          |
| 1046.4 + 0.2 | 1046.39     | 115–123 + 1 ox |
| 1068.4 + 0.2 | 1070.42     | 132–140         |
| 970.4 + 0.5 | 970.36      | 124–131 + 1 ox |
| 12.3    | 615.3 + 0.1 | 615.32          | 14–20          |
| 12.9    | 1030.4 + 0.1 | 1030.39         | 115–123        |
| 1456.8 + 0.2 | 1456.79     | 14–28           |
| 1200.6 + 0.2 | 1199.46     | 131–140         |
| 13.1    | 954.4 + 0.2 | 954.36          | 124–131        |
| 1070.4 + 0.2 | 1070.42     | 132–140         |
| 13.5    | 1514.8 + 0.2 | 1514.83         | 47–61          |
| 13.7    | 1179.6 + 0.1 | 1179.65         | 36–46          |
| 14.1    | 912.5 + 0.1 | 912.46          | 106–114        |
| 15.0    | 2191.3 + 0.2 | 2190.19         | 36–57          |
| 15.3    | 2939.7 + 0.2 | 2940.48         | 58–83 + 326    |
| 15.8    | 3086.3 + 0.1 | 3086.41         | 84–114         |
| 16.8    | 1926.1 + 0.9 | 1924.84         | 106–123        |
| 18.8    | 4101.1 + 0.8 | 4099.46         | 84–123         |
| 19.4    | 2614.5 + 0.1 | 2614.98         | 58–83          |
| 21.1    | 2128.3 + 0.5 | 2127.15         | 62–83          |
| 29.5    | 5684.8 + 0.8 | 5683.37         | 58–114         |
| 30.1    | 4789.6 + 0.9 | 4788.48         | 36–83          |
| 30.3    | 3626.3 + 0.5 | 3626.12         | 47–83          |
| 37.9    | 9733.9 + 1.1 | 9729.97         | 47–140 + 20x   |
| 38.5    | 8676.1 + 0.5 | 8676.56         | 47–131 + 20x   |
| 39.8    | 6964.3 + 1.5 | 6961.89         | 36–105         |
| 40.3    | 7290.3 + 1.1 | 7297.89         | 36–105 + 326   |
| 40.6    | 6696.6 + 1.1 | 6696.42         | 58–123         |

* Peptides are listed in order of retention times (RT).
* Experimental molecular masses determined by ESI-MS.
* Molecular masses calculated from aS amino acid sequence.
* The modified peptides are highlighted in bold.
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The process of unsaturated fatty acids peroxidation is a free-radical chain reaction that consists of three partially overlapping phases of radical reactions: initiation, propagation, and termination (17) (Scheme 1). Initiation takes place by abstraction of a hydrogen radical from the PUFA (FAH). During the propagation, the resulting lipid free radical (FA') reacts with molecular oxygen to generate a peroxyl radical (FAOO''), which is the dominant radical present in the chain. It can react with another FAH to form hydroperoxides (FAOOH) and FA', the early products of autoxidation. Successively, the hydroperoxides may decompose in alkoxy (FAO') and hydro (HO') derivatives. In vitro, the presence of trace metals or sources of energy (i.e., light or heat) can initiate the process (19). PUFAs easily undergo autoxidation because of the intrinsic structure of the radical intermediates, where electrons are highly delocalized. Normally, such radicals can interact with proteins giving rise to a wide range of compounds able to affect many physiological events (41). Here, we show that the propagation chain reaction is hampered in the presence of aS, and mutually, the protein results chemically modified with the formation of covalent adducts.

Fingerprinting analysis of the modified species and relative reactivities toward oxidative modifications suggested that the residue that is likely modified in aS is His-50. These data agree with the specific reactivity of lipid peroxidation products, such as allylic radicals, which involve preferentially in the order cysteine (absent in aS), histidine and lysine residues (42), and 4HNE, which, in aS, involves specifically His-50 (43). The imidazole moiety is nucleophilic and can react with one of the early products of autoxidation of DHA or AA. The propagation step in the PUFAs oxidation is hampered and the consequent breakdown into lipid-derived aldehydes and ketones is avoided, indeed, under our experimental conditions, the reaction terminates with the sequestration of the DHA or AA early reactive species by aS and the formation of a 4-hydroxy-2(E)-nonenal moiety was not observed.

Interestingly, Zhu et al. (44) proposed for aS an antioxidant activity of protection of lipids in neuronal membranes. They suggested a mechanism mediated by the four Met residues. This fact is corroborated by the finding that oxidized Met(s) can be repaired in vivo by intracellular methionine sulfoxide reductase (45). Our data show that the scavenging activity of DHA radicals is not exclusively correlated to Met residues, because aS40x is still able to sequester these radicals. In view of the fact that His-50 is the only His residue in aS and, chemically, the preferential modification site by lipid radicals (42), the importance of this residue in the scavenger activity was investigated by using a natural aS variant containing a Gln instead of His in position 50 (H50Q) (34–36). We showed that, despite the lack of His-50, this variant is able to protect DHA. The behavior of H50Q in the presence of DHA is very similar to that of WT aS, and the protein acquires α-helical secondary structure upon incubation with DHA. It was recently found that the H50Q mutation does not affect aS subcellular localization, the structure of free or membrane-bound aS monomer nor its capacity for repair of oxidized Met(s) (37). This fact is corroborated by the finding that oxidized Met(s) can be repaired in vivo by intracellular methionine sulfoxide reductase (45). Our data show that the scavenging activity of DHA radicals is not exclusively correlated to Met residues, because aS40x is still able to sequester these radicals. In view of the fact that His-50 is the only His residue in aS and, chemically, the preferential modification site by lipid radicals (42), the importance of this residue in the scavenger activity was investigated by using a natural aS variant containing a Gln instead of His in position 50 (H50Q) (34–36). We showed that, despite the lack of His-50, this variant is able to protect DHA. The behavior of H50Q in the presence of DHA is very similar to that of WT aS, and the protein acquires α-helical secondary structure upon incubation with DHA. It was recently found that the H50Q mutation does not affect aS subcellular localization, the structure of free or membrane-bound aS monomer nor its capacity for repair of oxidized Met(s) (37). This fact is corroborated by the finding that oxidized Met(s) can be repaired in vivo by intracellular methionine sulfoxide reductase (45). Our data show that the scavenging activity of DHA radicals is not exclusively correlated to Met residues, because aS40x is still able to sequester these radicals. In view of the fact that His-50 is the only His residue in aS and, chemically, the preferential modification site by lipid radicals (42), the importance of this residue in the scavenger activity was investigated by using a natural aS variant containing a Gln instead of His in position 50 (H50Q) (34–36). We showed that, despite the lack of His-50, this variant is able to protect DHA. The behavior of H50Q in the presence of DHA is very similar to that of WT aS, and the protein acquires α-helical secondary structure upon incubation with DHA. It was recently found that the H50Q mutation does not affect aS subcellular localization, the structure of free or membrane-bound aS monomer nor its capacity for repair of oxidized Met(s) (37). This fact is corroborated by the finding that oxidized Met(s) can be repaired in vivo by intracellular methionine sulfoxide reductase (45). Our data show that the scavenging activity of DHA radicals is not exclusively correlated to Met residues, because aS40x is still able to sequester these radicals. In view of the fact that His-50 is the only His residue in aS and, chemically, the preferential modification site by lipid radicals (42), the importance of this residue in the scavenger activity was investigated by using a natural aS variant containing a Gln instead of His in position 50 (H50Q) (34–36). We showed that, despite the lack of His-50, this variant is able to protect DHA. The behavior of H50Q in the presence of DHA is very similar to that of WT aS, and the protein acquires α-helical secondary structure upon incubation with DHA. It was recently found that the H50Q mutation does not affect aS subcellular localization, the structure of free or membrane-bound aS monomer nor its capacity...
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to be phosphorylated in vitro (46). Moreover, the substitution of His-50 with Gln accelerates the conversion of αS into β-sheet-rich oligomers (35) and increases αS secretion and extracellular toxicity (46). If incubated with DHA, H50Q is modified and a mass increase of 326 Da (or a multiple of it) is still detectable, indicating the formation of covalent adduct(s) with the fatty acid. Lacking His, the modifications can reasonably occur at the level of the lysine residues, which, as nucleophiles, are vulnerable to modification by lipid peroxidation-derived electrophiles (42). A possible candidate is Lys in position 60 that was reported modified in the presence of 4HNE (38).

Our data show also that the structural interaction is an important requisite for αS scavenger activity. The analysis of the structural properties of αS40x by CD studies has shown that the presence of methionine sulfoxide does not alter the ability of αS to acquire α-helical secondary structure upon interaction with DHA, as already observed with other lipids (30). On the contrary, the extensive acetylation does not allow any interaction with the fatty acid and acetyl-αS does not sequester DHA radicals. This behavior suggests an important role for Lys residues in the repeats of αS.

A further consideration can be done on the ability of αS fibrils to partially protect DHA. The fold of αS fibrils comprises a central core of cross-β structure (residues 35–94 according to Vilar et al. (47), and residues 38–96 according to Comellas et al. (48) flanked by flexible regions. In both models, the N-terminal domain containing the first and second repeats is not engaged in the fibril structure. Zarbiv et al. (49) demonstrated that lysines belonging to these repeats are essential for lipid interaction. Therefore, αS fibrils could still interact with DHA. The reduced activity could be due to the fact that not the entire N-terminal region, but likely only the first 25 residues, not engaged in the structure of fibril, are suitable for lipid interaction. This is consistent with the observation that the first 25 residues have the highest membrane affinity (50), with amino acids 6–25 likely firmly anchored to the membrane (51). Comparing the spectra of DHA obtained in the presence of αS as a monomer and αS as fibril, just evaluating the height of the signals at m/z 415 in a ratio with those due to not oxidized DHA, roughly fibrils lose 80% of the scavenging activity of αS. It is not unexpected that αS, accumulating as fibrils in Lewy bodies, does not maintain the scavenging activity. In vivo, fibril formation could be further detrimental, because it is known that monomeric αS decreases in PD (52), probably captured by the fibrillar aggregates.

In our previous works, we have shown that αS and its N-terminal fragments (such as 1–52) reduce the aggregative concentration of DHA and cause a resealing of the lipid oil droplets with formation of a microemulsion (27, 53). In aqueous buffer, DHA forms an emulsion and shows a strong tendency to oxidize, generating monohydroxides and dihydroperoxides derivatives. In a bulk phase or in an inorganic solvent, this tendency is partially reduced (54). According to Miyashita (54), we can hypothesize that αS protects the fatty acid from oxidation acting also as an emulsifier. Indeed, when αS is not able to interact with lipids or even to reseal the fatty acid droplets, no oxidant scavenger effect is observable. This is evident with acetyl-αS that lost the ability to interact with DHA (Fig. 6) and to reseal DHA droplets (data not shown). However, αS does not behave as an inert emulsifier, but it undergoes oxidative modifications. Upon interaction with DHA, αS acquires a conformation that promotes its oxidation serving as terminal of the lipid oxidation process.

Several studies reported that αS is involved in the maintenance of the correct levels of PUFAs (55). It was also proposed that αS may play a protective role of neurons exposed to oxidative stress preventing oxidation of PUFAs (33). Indeed, lipid peroxidation increases with aging and is linked to PD (21). In neurons exposed to oxidative stress or obtained from patients affected by PD, the levels of PUFAs in a non-esterified form were found higher than normal (15). Interestingly, it was also reported that in cultured cells a PUFA treatment induces the appearance of toxic αS oligomers (56). Soluble oligomers are also triggered by oxidation of αS Met residues (57). In vivo, methionine sulfoxide are repaired by intracellular reductase. Very recently, it was observed that when the two C-terminal Met are not efficiently repaired by endogenous cellular enzymes, they remain oxidized. This results in the accumulation of oxidized αS, with high tendency to form oligomers, and that shows impaired phosphorylation (58). The cellular impact of this depends on the eventual toxicity of the oligomeric species. If toxic oligomers accumulate in cells, the formation of oxidized or partially oxidized αS becomes a significant contributing factor to PD. The protective role of αS toward PUFAs and the tendency of the protein to aggregate in their presence are apparently conflicting. Therefore, it is plausible to speculate that the protein/PUFAs ratio is a discriminant factor determining the effect of the interaction between αS and PUFAs. Under physiological conditions, αS can participate actively to the control of oxidative homeostasis of the intracellular environment, protecting the free fatty acids from oxidation and maintaining their correct level. Environmental factors can affect this equilibrium, by either increasing oxidative stress or changing DHA concentration. PD has been previously linked to oxidative stress, which can increase lipid oxidative species to an extent that does not allow the normal biological turnover of αS, leading to the accumulation of modified protein and to increased propensity to its oligomerization. Moreover, also the DHA concentration has been reported to increase in neuronal membranes of PD patients and also in general aging (55). Higher concentrations of DHA prevents αS amyloid aggregation, probably by the α-helix structure acquisition, but the formation of stable toxic oligomers is observed (27–29, 55). Because oligomers might represent the most toxic species, the increased DHA concentration in the cell could accelerate toxicity rather than having a protective role. Otherwise, we found that fibrils, in comparison to soluble αS, have a reduced scavenging activity. Determination of the real intracellular proportion between soluble and fibrillar αS will allow in the future to properly frame these findings within the complexity of the pathophysiology of the disease.

In conclusion, αS sequence is tailored to efficiently exert an oxidant scavenger activity, accordingly with recent hypothesis (33). The role of Met has been extensively described (30, 45). Lysine residues could play a significant role. Their distribution makes the αS interaction with PUFAs and other lipids effective.
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This deep interaction has specific consequences on the physical state of PUFA s, which are emulsified and resided as well as on the reactive species that are sequestered. Lysine and histidine residues can mediate the capturing action for its intrinsic chemical reactivity, hampering the propagation of the PUFA s oxidation process. This proposed scavenging mechanism could not be assumed as specific of αS. Its biological significance resides in the recognized role of the protein in PD, and specifically in the reported evidences of its interaction with PUFA and its involvement in their metabolism (11–16). αS is mainly a presynaptic protein, and it has been reported to be able to bind plasma membranes, lipid rafts, inner nuclear membrane, and mitochondrial membranes. These interactions are considered critical for the physiological function of αS (10). DHA is an essential PUFA, fundamental for the functions and maintenance of the nervous system, and deficits in DHA are associated with cognitive decline during aging and in neurodegenerative disease (59). αS has been repeatedly reported to interact with PUFA: endogenous cytosolic levels of αS can buffer arachidonic acid affecting SNARE assembly on cellular membrane (60). Elevated PUFA levels have been detected in PD or Lewy body dementia-affected human brains, whereas the levels of certain PUFA s were decreased in the brains of mice genetically deleted of αS (55). This mechanism of scavenging activity could provide a link between αS, altered lipid composition, and PD development.

Experimental procedures

Protein expression and purification

Expression and purification of recombinant αS and αS variant were obtained as previously described (29, 35). Protein concentrations were determined by absorption measurements at 280 nm using a double-beam Lambda-20 spectrophotometer from PerkinElmer Life Sciences. The extinction coefficient for αS and H50Q at 280 nm was 5960 M⁻¹ cm⁻¹ (61). DHA and AA were purchased from Sigma. All other chemicals were of analytical reagent grade and were obtained from Sigma or Fluka (Buchs, Switzerland). Aliquots of DHA and AA samples were obtained as previously described (27). DHA under this condition forms oil droplets, as previously reported (27). Aliquots of the solution after preparation and 24 h of incubation (37 °C, 300 rpm) were diluted 10 times in 50% H₂O, 49.9% acetonitrile, 0.1% HCOOH, and analyzed by an ESI-mass spectrometer with a Q-TOF analyzer (Micro) from Waters (Manchester, UK). The measurements were conducted at a capillary voltage of 2.7 kV and a cone voltage of 35 V. The molecular masses of protein samples were estimated using the Mass-Lynx software 4.1 (Micromass).

CD spectroscopy

Circular dichroism (CD) spectra in the far UV were recorded on a Jasco J-710 spectropolarimeter (Tokyo, Japan), using a 1-mm path length quartz cell and a protein concentration of 5–7 μM. The mean residue ellipticity [θ]MRW (deg cm² dmol⁻¹) was calculated from the formula [θ]MRW = (θobs/10) (MRW/le), where θobs is the observed ellipticity in deg, MRW is the mean residue molecular weight of the protein, l the optical path length in cm, and c the protein concentration in g/ml. The spectra were recorded in PBS buffer, pH 7.4.

Fingerprinting analysis

Proteolysis with trypsin was conducted using an E/S ratio of 1:100 (by weight), in PBS, pH 7.4, and the reaction was quenched by acidification with TFA in water (4%, v/v). Proteolysis with endoproteinase GluC was performed at E/S ratio of 1:50 (by weight). RP-HPLC of proteolytic mixture was conducted on a Vydac C18 column (4.6 × 250 mm; The Separations Group, Hesperia, CA), eluted with a gradient of water, 0.1% TFA versus acetonitrile, 0.085% TFA from 5 to 25% in 5 min, from 25 to 28% in 13 min, from 28 to 39% in 3 min, and from 39 to 45% in 21 min at a flow rate of 1 ml/min. The peptides were identified by mass spectrometry. MS-MS analysis was conducted on the selected ions using collision energies from 15 to 30 V.

Author contributions—P. P. D. L. conceived and supervised the project; G. D. F. and C. F. designed and performed the experiments; G. D. F., C. F., C. P., V. B., and P. P. D. L. analyzed the data; G. D. F., C. F., and P. P. D. L. wrote the paper; V. B. critically revised the paper; and R. S. and A. H. V. S. supervised parts of the project. All authors analyzed the results and approved the final version of the manuscript.
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