Serotype Distribution of Plasmid-encoded Virulence Profiles, and Identification of \textit{espP} and \textit{subAB} Alleles in Verotoxigenic \textit{Escherichia coli}

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Authors' contributions

This work was carried out the collaboration of all authors. Authors AMS and AVB participated in the planning of the study, drafting and critical review of the manuscript and supervised the laboratory work. Authors AMS, AVB, TF, JG and JSC participated in the data collection and data entry. Author PMAL participated in the planning of the study and critical review of the manuscript. All authors read and approved the final manuscript.

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ABSTRACT

\textbf{Aims:} This study was designed to characterize verotoxigenic \textit{Escherichia coli} (VTEC), important food-borne pathogens, in relation to virulence genes found in large plasmids harboured by disease-associated strains. Our aim was to detect these genes and possible combinations of them, and to establish if some kind of relationship exists between these profiles and different serotypes.

\textbf{Study Design:} Amplification of genes and allelic variants by PCR.

\textbf{Place and Duration of Study:} Laboratorio de Inmunocquímica y Biotecnología (FCV-UNCPBA, Argentina), between June 2010 and July 2013.

\textbf{Methodology:} 208 VTEC isolates belonging to 49 serotypes were characterized for the presence of plasmid-encoded genes: \textit{epeA} (serine-protease), \textit{espP} (extracellular serine protease) and its variants, \textit{katP} (periplasmic catalase-peroxidase), \textit{stcE} (zinc metalloprotease) and \textit{subA} (subtilase)}
cytotoxin) and its variants.

**Results:** The most frequently detected gene was espP (87%), followed by ehxA (47%), saa (20%) and katP (17%), whereas epeA, subA and stcE ranged from 7 to 12%. Taking into account these genes, twelve profiles were observed, ranging from the presence of zero to five of the genes. Some serotypes presented only one plasmid profile whereas others, such as O20:H19, showed up to four different profiles. Functional differences have been previously found among EspP subtypes, and we found that several VTEC isolated from bovines or foods contained the espPxα, which is one of the alleles possibly associated with severe human disease. Considering subtilase cytotoxin gene subtypes, only subAB1 was identified.

**Conclusion:** Our results show that VTEC strains can possess different combinations of plasmid-encoded virulence genes and even strains belonging to the same serotype can differ in relation to their plasmid genetic composition. In relation to allelic variants, LEE-positive serotypes showed for espP gene the allele α or the γ, and the detected subAB allele differed from the prevalent allelic variant subAB2 carried by strains circulating in sheep and humans in European countries.

**Keywords:** Plasmid profiles; verotoxigenic Escherichia coli; virulence; espP alleles; subAB alleles.

### 1. INTRODUCTION

The verotoxigenic *Escherichia coli* (VTEC) group, also called Shiga toxin-producing *Escherichia coli* (STEC), is very diverse, being *E. coli* O157:H7 the most common serotype associated with sporadic cases and large outbreaks of hemolytic uremic syndrome (HUS). However, there is growing concern about the risk to human health associated with non-O157 VTEC serotypes. Mobile genetic elements (MGEs) have had a profound influence on the emergence and spread of pathogenic bacteria. Examples of MGEs are plasmids, which confer determinants of pathogenicity. All clinical isolates of *E. coli* O157:H7 harbor a large plasmid called pO157, and plasmids that share some virulence factors are commonly found in disease-associated non-O157 VTEC, including O103:H2, O26:H11, O26:NM, O111:NM and O113:H21 serotypes [1-3].

Among the recognized plasmid-encoded virulence markers is the EHEC (enterohemorrhagic *Escherichia coli*)-enterohemolysin (EHEC-HlyA or EhxA), which was the first described virulence factor of pO157 [4]. Furthermore, other plasmid-encoded proteins have been proposed as virulence factors in VTEC.

The STEC autoagglutinating adhesin encoded on pO113 by saa, has been suggested to be unique to LEE (locus for enterocyte effacement)-negative VTEC isolates [5-7].

Another gene, katP, encoding for a catalase-peroxidase was identified on pO157 [8]. This enzyme may help *E. coli* O157:H7 to colonize host intestines by reducing oxidative stress. It has been demonstrated that KatP is a peroxide scavenger that may protect *E. coli* O157:H7 EDL933 from oxidative damage during aerobic growth in an OxyR (peroxide-inducible transcriptional regulator)-dependent manner [9].

The protein EspP belongs to the serine protease autotransporters of Enterobacteriaceae (SPATE) family, able to cleave pepsin A and human coagulation factor V, contributing to the pathology of hemorrhagic colitis [10]. Dziva and colleagues [11] reported that EspP influences intestinal colonization of calves and adherence to bovine primary intestinal epithelial cells. In addition, it has been reported that this protease is able to cleave complement factors, and therefore could interfere with host innate immune response [12]. EspP can be subtyped into five groups and functional differences have also been found among them, being EspPα and EspPγ proteolytically active [13,14].

The zinc metalloprotease StcE is secreted through the type II secretion system encoded on pO157 [15]. StcE cleaves the C1 esterase inhibitor, which is a host regulator of the classical complement pathway. Furthermore, it has been demonstrated that it can contribute to intimate adherence of *E. coli* O157:H7 to Hep2 cells in vitro [16].

The subtilase cytotoxin (SubAB) is an AB5 toxin described in certain VTEC strains that usually lack the LEE and has shown to be cytotoxic to Vero cells and lethal for mice [17,18]. This toxin was first described in a VTEC O113:H21 strain implicated in an outbreak of HUS in Australia, and it was encoded by two cotranscribed genes (subA and subB), on a large virulence plasmid (pO113) [17], which also carries the saa gene.
But recently, it has been reported that some SubAB-positive strains harbor subAB genes in the chromosome and next to the gene tia, which encodes an invasion factor previously described in enterotoxigenic E. coli [19].

The gen epeA, found in the pO113 transfer region, encodes an SPATE protein that may aid colonization and adherence to the host intestine through mucinolytic activity. The presence of epeA appeared to be also specifically associated with LEE-negative isolates of EHEC [2].

Our aim was to characterize a selection of VTEC strains isolated mainly from cattle and meat and representing many serotypes, in relation to several plasmid-borne virulence factors genes, to analyze the different combinations of these genes and to establish if some kind of relationship exists between these profiles and the serotypes. Also, in relation with the espP and subA genes, our aim was to investigate the occurrence of different subtypes.

2. MATERIALS AND METHODS

A total of 208 VTEC isolates obtained from cattle and raw meat, with the exception of two O157:H7 human ones, were investigated. The isolates belonging to 49 serotypes are part of the culture collection of the Laboratorio de Inmunocuímica y Biotecnología (FCV-UNCPBA, Argentina). They had all been previously analyzed in relation to the presence of genes encoding for chromosomal virulence factors, verotoxin 1 and 2 (vtx1, vtx2), intimin (eae), and to plasmid-encoded factors, enterohaemolysin (ehxA) and STEC autoagglutinating adhesin (saa) [6, 20-22]. In the present study, the isolates were characterized for the presence of plasmid-encoded genes: epeA (serine-protease), espP (extracellular serine protease) and its variants, katP (periplasmic catalase-peroxidase), stcE (zinc metalloprotease) and subA (subtilase cytotoxin) and its variants. The epeA gene was detected by a monoplex PCR [2] and the remaining ones, by a pentavalent PCR assay [23] which also detects ehxA. PCR amplification of epeA was performed under the following conditions: denaturing step of 5 min at 94 °C and 30 cycles of 30 s at 94 °C, 1 min at 46 °C, and 1 min at 72 °C, followed by 5 min at 72 °C for elongation. Plasmid virulence profiles were defined taking into account the results obtained in the present study and those previously found for saa.

The strains positive for espP or subA were further characterized as follows. Determination of espP alleles α, β, γ and δ was done by PCR as described by Brockmeyer et al. [13] and subtype ε, as described by Bielaszewskà et al. [14]. For subA-positive strains, the presence of subtilase cytotoxin alleles (subAB1 and subAB2) plus the tia gene were assessed as described by Michelacci et al. [24].

3. RESULTS

In order to characterize the plasmid virulence profile of 208 VTEC strains isolated mainly from cattle and raw meat, we determined the presence of particular virulence genes that have been reported to be located on large plasmids of VTEC.

The most commonly found gene was espP, present in 182 (87%) of the VTEC strains tested, and the second most frequent gene was ehxA (47%). Both were detected either alone or in combination with other plasmid genes. The gene katP was found in 36 strains (17%) whereas epeA, subA and stcE were present in frequencies ranging from 7 to 12%, but they were never detected as the only plasmid gene. Those strains that were positive for subA harbored neither stcE nor katP; stcE was present only in O157:H7 isolates and katP appeared only in LEE-positive strains, never simultaneously with saa.

Taking into account the seven encoded-plasmid genes, twelve virulence profiles were observed, ranging from the presence of zero to the presence of five of the genes, being the profile espP (negative for the other 6 genes) the most prevalent one (98 isolates), followed by katP espP ehxA (22 isolates) and epeA espP subA ehxA saa (19 isolates) (Fig. 1). Most LEE-positive non-O157 serotypes were grouped in two profiles: katP espP ehxA and espP ehxA, whereas the profile katP espP stcE ehxA was characteristic of O157:H7 and only present in those isolates. Some serotypes presented only one plasmid profile whereas others, such as O20:H19, showed up to 4 different profiles. In nine VTEC isolates it was not possible to detect any of the 7 plasmid genes (Table 1). Most of the studied isolates harbored vtx2, alone or in combination with vtx1 (data previously obtained). Some of the few isolates which carried only vtx1 are those belonging to O8:H16 and these ones presented a particular profile, espP saa.
Fig. 1. Relationships of plasmid virulence profiles for VTEC isolates

Minimum-spanning tree of plasmid profiles generated with Bionumerics software v6.6 with single and double locus variance priority rules and the Dice similarity coefficient calculated for binary characters. Each circle represents a different virulence profile, and the letter in the circle indicates the virulence profile type. The circle size corresponds to the number of isolates with an identical virulence profile. Number of isolates included in each profile is indicated in parentheses.

Table 1. Plasmid virulence profiles of VTEC, frequencies and serotypes

| Profiles                        | %   | Serotypes*          |
|---------------------------------|-----|---------------------|
| espP (A)                        | 47.1| O15:H21 (1), O20:H7 (2), O20:H19 (2), O112:H2 (1), O113:NM (7), O113:H21 (3), O117:H7 (11), O162:H7 (1), O171:NM (5), O171:H2 (11), O171:HNT (1), O174:H21 (13), O178:H19 (5), O185:H7 (1), O194:H21 (13), ONT:NM (16), ONT:H7 (4), ONT:H8 (1), ONT:H21 (12), ONT:HNT (1) |
| espP saa (B)                    | 1.4 | O8:H16 (3)          |
| espP ehxA (C)                   | 5.3 | O5:NM (4), O8:H19 (1), O20:HNT (1), O26:H11 (2), O103:NM (2), O103:H2 (1) |
| espP ehxA saa (D)               | 4.3 | O20:H19 (1), O22:H8 (1), O88:H21 (1), O91:H21 (4), O174:H21 (1), ONT:H19 (1) |
| ehxA (E)                        | 5.8 | O8:H19 (8), O25:H19 (1), O111:NM (1), O175:H8 (2) |
| ehxA saa (F)                    | 1.9 | O22:H8 (1), ONT:H21 (2), ONT:HNT (1) |
| katP espP ehxA (G)              | 10.6| O26:H11 (5), O38:H19 (1), O118:H16 (1), O145:NM (10), O146:NM (1), O165:NM (1), O177:NM (3) |
| katP espP stcE ehxA (H)         | 6.8 | O157:H7 (14)        |
| espP subA ehxA saa (I)          | 2.9 | O20:H19 (1), O116:H21 (1), O141:H7 (1), O141:H8 (1), O178:H19 (1), ONT:H8 (1) |
| epe espP subA ehxA saa (J)      | 9.1 | O2:H5 (1), O20:H19 (4), O39:H49 (3), O79:H9 (3), O113:H21 (3), O120:H19 (1), O141:H8 (1), O178:H19 (1), ONT:H7 (1), ONT:H19 (1) |
| epe ehxA saa (K)                | 0.5 | ONT:H19 (1)         |
| Negative for the 7 genes (L)    | 4.3 | O2:NM (1), O22:H8 (1), O113:NM (3), O145:NM (2), O146:H21 (1), O174:H21 (1) |

*LEE-positive serotypes are in bold. Number of isolates is indicated in parentheses after each serotype.
All espP and subA-positive strains were subjected to further analysis to identify particular subtypes. With regard to espP, the γ subtype was the most frequently found, being present in 62 isolates belonging to 25 serotypes, including both LEE-positive and LEE-negative ones. The α subtype was found in 35 isolates from 9 serotypes, all of them LEE-positive; the β subtype, in 36 isolates of 14 serotypes. Meanwhile subtype δ was not detected in our strain collection and subtype ε was only found in the unique O15:H21 isolate. It is noticeable that it was not possible to assign alleles to 48 isolates, in particular, to most of the isolates belonging to the serogroup O171 and to serotype O174:H21 (Table 2).

Taking into account the serotypes that comprised more than one isolate and the espP alleles identified, several of them (O5:NM, O39:H49, O79:H19, O141:H8, O157:H7 and O177:NM) presented a same subtype among the isolates. On the contrary, others, such as O8:H16, O20:H19, O26:H11, O113:NM, O113:H21, O117:H7 and O178:H19, presented more than one known espP allele. Within each serotype, no association could be found between the origin of the isolates and the espP subtype (data not shown).

In 23 of the 25 subA-positive strains, the subAB allele could be identified. All of them (n = 23) were positive for the prototype of SubAB (SubAB)α, and, conversely, negative for the recently identified new variant of SubAB (SubAB)γ and for tia, both genes proposed to be located in a same PAI. The remaining two isolates, in which the subAB subtype could not be identified, belonged to O79:H19 and O120:H19 serotypes.

4. DISCUSSION

Our results showed that virulence plasmids of VTEC are highly variable. No isolates contained all seven markers, but isolates belonging to O2:H5, O20:H19, O39:H49, O79:H19, O113:H21, O120:H19, O141:H8, O178:H19, ONT:H7 and ONT:H19 did possess 5 of the genes, epeA, espP, subA, ehxA and sac. On the other hand, in some isolates belonging to serotypes O2:NM, O22:H8, O113:NM, O145:NM, O146:H21 and O174:H21 we could not detect any of these plasmid genes. It is plausible that these isolates had lost the large virulence plasmid since MGEs can be easily exchanged among strains or lost, and other isolates belonging to some of these serotypes did have one or more plasmid genes. As a consequence, VTEC clones with different genetic assets in regard to virulence genes are generated.

The most prevalent genes were espP and ehxA, similarly to Bosilevac et al. [25], but in contrast to other studies in which all VTEC isolates that were positive for espP were also positive for ehxA [26], we detected the profile espP as the most frequent.

The profile katP espP stcE ehxA was exclusively found among O157:H7 strains, being stcE only present in this serotype, in concordance with other studies [27]. The plasmids from O157:H7 have been reported to be highly similar [28], but a diversity of marker combinations has been found among plasmids from O103:H2 [29-31].

| Serotype (**) | espP subtype (**) | Serotype | espP subtype | Serotype | espP subtype |
|--------------|------------------|----------|--------------|----------|--------------|
| O2: H5 (1)   | γ                | O103:NM (2) | α (1) n (1) | O162: H7 (1) | γ           |
| O5: NM (4)   | γ                | O103: H2 (1) | α           | O165: NM (1) | γ           |
| O8: H16 (3)  | β (1) γ (2)     | O112: H2 (1) | γ           | O171: NM (5) | γ (1) n (4) |
| O8: H19 (1)  | n                | O113: NM (7) | β (2) γ (5) | O171: H2 (11) | γ (2) n (9) |
| O15: H21 (1) | ε                | O113: H21 (6) | β (4) γ (2) | O171: HNT (1) | n           |
| O20 : HNT (1)| γ                | O116: H21 (1) | n           | O174: H21 (14) | γ (5) n (9) |
| O20: H7 (2)  | n                | O117: H11 (11) | β (4) γ (6) n (1) | O177: NM (3) | α           |
| O20: H19 (8) | β (5) γ (1) n (2)| O118: H16 (1) | α           | O178: H19 (7) | β (2) γ (2) n (3) |
| O22: H8 (1)  | γ                | O120: H19 (1) | γ           | O185: H7 (1) | β           |
| O26: H11 (7) | α (4) γ (2) n (1) | O141: H7 (1) | n           | ONT: NM (16) | β (4) γ (4) n (8) |
| O38: H39 (1) | α                | O141: H8 (2) | β           | ONT: H7 (5) | β (2) γ (3) |
| O39: H49 (3) | β                | O145: NM (10) | α (9) n (1) | ONT: H8 (2) | β (1) n (1) |
| O79: H19 (3) | β                | O146: NM (1) | α           | ONT: H19 (2) | γ (1) n (1) |
| O88: H21 (1) | γ                | O157: H7 (14) | α           | ONT: H21 (12) | β (2) γ (10) |
| O91: H21 (4) | γ (3) n (1)      |            |              | ONT: HNT (1) | γ           |

(*) number of analyzed espP-positive isolates, (**) number of isolates is only indicated when not all the isolates of a serotype presented the same espP allele; n: non identified espP subtype. LEE-positive serotypes are in bold.
and we detected the profile espP ehxA, not common for this serotype.

SubAB, the subtilase cytotoxin, has been detected mostly in LEE-negative VTEC strains [32 and papers cited in it]. The subAB genes may be mobilized by a plasmid that also carries saa gene, or alternatively, by a putative pathogenicity island (PAI) together with tia [19]. Allelic variants of subAB, differentially associated with these two MGEs have been described but there are few studies that identify them and analyze their distribution among VTEC strains. In Spain, the variant subABγ was found to predominate among VTEC isolated from large game animals, their meats, small ruminants and possibly related strains from human [33-35,24]. Furthermore, this allele was also detected in almost all subAB-positive human strains from Denmark [24], and in deer meat strains from Germany [36]. However, all the subAB-positive isolates that could be subtyped in this study (obtained from cattle and derived meat) possessed subABα. This allele was also detected in most of the subAB-positive VTEC strains isolated from cattle in Spain [33]. Recently, two allelic variants for subABγ have been described (subABγ2,1 and subABγ2,2) [36]. Further studies from different geographical areas and VTEC reservoirs are needed to determine if there exists an association between specific animal reservoirs and subAB alleles.

EspP is a serine protease expressed during human infection that can degrade human coagulation factor V. From the five subtypes of espP identified in VTEC [13,14], only EspPα and EspPγ were able to cleave coagulation factor V [37]. This study revealed an association among some serogroups and particular espP subtype(s).

LEE-positive serotypes showed the allele α or the γ. Despite that in previous studies espPα was found particularly in isolates from HUS patients, we detected this allele in several strains isolated from bovines or foods. Particularly interesting is the observation that serogroup O103, considered to be highly pathogenic and reported as frequently negative for espP [13,38], resulted espP-positive and carried the allele α. On the other hand, espPγ was detected both in LEE-positive and in LEE-negative isolates, which was also found by other authors [13]. The subtype δ was not detected in our strain collection, neither in isolates from Austria [37]. In a significant proportion of espP-positive isolates we were not able to determine the espP allele, especially in O171 and O174:H21 strains, suggesting the presence of novel alleles.

Before many of the VTEC plasmid genes were described, Brunder et al. [39] detected a considerable variation among megaplasmids. They are clearly mosaics, and most putative virulence genes are flanked by mobile genetic determinants, which probably facilitate their transfer. However, partial deletions of these mobile elements, as shown with the regions that flank the sfp operon in pO165, can stabilize particular virulence loci [14].

VTEC virulence is a multifactorial trait and the genetic markers to define it have yet to be determined [25]. Brandt et al. [40] remarked that serotyping is not an absolute predictor of risk to human health, and instead suggested to examine the virulence gene content of a strain.

5. CONCLUSION

In the present study, the characterization of VTEC strains belonging to 49 serotypes revealed a great diversity of virulence profiles in regard to plasmid encoded genes and, noticeably, intraserotype differences were also identified. The finding of different combinations of virulence factors in non-O157:H7 serotypes traditionally not associated with disease should alert on the desirability to search for these specific virulence factors in routine diagnostic.

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COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Gyles C, Johnson R, Gao A, Ziebell K, Pierard D, Aleksic S, et al. Association of enterohemorrhagic Escherichia coli hemolysin with serotypes of Shiga-like-toxin-producing Escherichia coli of human and bovine origin. Appl Environ Microbiol. 1998;64:4134–4141.

2. Leyton DL, Sloan J, Hill RE, Doughty S, Hartland EL. Transfer region of pO113 from enterohemorrhagic Escherichia coli.
similarity with R64 and identification of a novel plasmid-encoded autotransporter, EpeA. Infect Immun. 2003;71:6307–6319.

3. Fratamico PM, Yan X, Caprioli A, Esposito G, Needleman DS, Pepe T, et al. The complete DNA sequence and analysis of the virulence plasmid and of five additional plasmids carried by Shiga toxin-producing *Escherichia coli* O26:H11 strain H30. Int J Med Microbiol. 2011;301:192–203.

4. Schmidt H, Beutin L, Karch H. Molecular analysis of the plasmid encoded hemolysin of *Escherichia coli* O157:H7 strain EDL 933. Infect Immun. 1995;63:1055–1061.

5. Paton AW, Srimanote P, Woodrow MC, Paton JC. Characterization of Saa, a novel autoagglutinating adhesin produced by locus of enterocyte effacement-negative Shiga toxigenic *Escherichia coli* strains that are virulent for humans. Infect Immun. 2001;69:6999-7009.

6. Lucchesi PM, Krüger A, Parma AE. Distribution of saa gene variants in verocytotoxigenic *Escherichia coli* isolated from cattle and food. Res Microbiol. 2006;157:263-266.

7. Cergole-Novella MC, Nishimura LS, Dos Santos LF, Irino K, Vaz TM, Bergamini AM, et al. Distribution of virulence profiles related to new toxins and putative adhesins in Shiga toxin-producing *Escherichia coli* isolated from diverse sources in Brazil. FEMS Microbiol Lett. 2007;274:329–334.

8. Brunder W, Schmidt H, Karch H. KatP, a novel catalase-peroxidase encoded by the large plasmid of enterohaemorrhagic *Escherichia coli* O157:H7. Microbiology. 1996;142:3305–3315.

9. Uhlich GA. KatP contributes to OxyR-regulated hydrogen peroxide resistance in *Escherichia coli* serotype O157:H7. Microbiology. 2009;155:3589–3598.

10. Brunder W, Schmidt H, Karch H. EspP, a novel extracellular serine protease of enterohaemorrhagic *Escherichia coli* O157:H7 cleaves human coagulation factor V. Mol Microbiol. 1997;24:767-778.

11. Dziva F, Mahajan A, Cameron P, Currie C, McKendrick IJ, Wallis TS, et al. EspP, a type V-secreted serine protease of enterohaemorrhagic *Escherichia coli* O157:H7, influences intestinal colonization of calves and adherence to bovine primary intestinal epithelial cells. FEMS Microbiol Lett. 2007;271:258-264.

12. Orth D, Ehrlenbach S, Brockmeyer J, Khan AB, Huber G, Karch H, et al. EspP, a serine protease of enterohemorrhagic *Escherichia coli*, impairs complement activation by cleaving complement factors C3/C3b and C5. Infect Immun. 2010;78:4294-4301.

13. Brockmeyer J, Bielaszewska M, Fruth A, Bonn ML, Mellmann A, Humph U-H, et al. Subtypes of the plasmid-encoded serine protease EspP in Shiga toxin-producing *Escherichia coli*: distribution, secretion, and proteolytic activity. Appl Environ Microbiol. 2007;73:6351–6359.

14. Bielaszewska M, Stoewe F, Fruth A, Zhang W, Prager R, Brockmeyer J, et al. Shiga toxin, cytolethal distending toxin, and hemolysin repertoires in clinical *Escherichia coli* O91 isolates. J Clin Microbiol. 2009;47:2061–2066.

15. Lathem WW, Grys TE, Witowski SE, Torres AG, Kaper JB, Tarr PI, et al. StcE, a metalloprotease secreted by *Escherichia coli* O157:H7, specifically cleaves C1 esterase inhibitor. Mol Microbiol. 2002;45:277-288.

16. Lathem WW, Bergsbaken T, Welch RA. Potentiation of C1 esterase inhibitor by StcE, a metalloprotease secreted by *Escherichia coli* O157:H7. J Exp Med. 2004;199:1077-1087.

17. Paton AW, Srimanote P, Talbot UM, Wang H, Paton JC. A new family of potent AB(5) cytotoxins produced by Shiga toxigenic *Escherichia coli*. J Exp Med. 2004;200:35–46.

18. Morinaga N, Yahiro K, Matsuura G, Watanabe N, Moss J, Noda M. Two distinct cytotoxic activities of subtilase cytotoxin produced by Shiga-toxigenic *Escherichia coli*. Infect Immun. 2007;75:488-496.

19. Tozzoli R, Caprioli A, Cappannella S, Michelacci V, Marziano ML, Morabito S. Production of the subtilase AB5 cytotoxin by Shiga toxin negative *Escherichia coli*. J Clin Microbiol. 2010;48:178–183.

20. Parma AE, Sanz ME, Blanco JE, Blanco J, Viñas MR, Blanco M, et al. Virulence genotypes and serotypes of verotoxigenic *Escherichia coli* isolated from cattle and foods in Argentina. Importance in public health. Eur J Epidemiol. 2000;16:757-762.
21. Padola NL, Sanz ME, Blanco JE, Blanco M, Blanco J, Etcheverría AI, et al. Serotypes and virulence genes of bovine Shigatoxigenic Escherichia coli (STEC) isolated from a feedlot in Argentina. Vet Microbiol. 2004;100:3–9.

22. Sanz ME, Villalobo C, Elichiribehety E, Arroyo G. Prevalencia de Escherichia coli verotoxigénico en productos cárnicos de la ciudad de Tandil. La Ind Cárnica Lat. 2007;146:56-58.

23. Bustamante AV, Sanso AM, Lucchesi PMA, Parma AE. Multiplex PCR assay for the detection of five putative virulence genes encoded in Verotoxigenic Escherichia coli plasmids. Curr Microbiol. 2011;62:1411-1415.

24. Michelacci V, Tozzoli R, Caprioli A, Martínez R, Scheutz F, Grande L, et al. A new pathogenicity island carrying an allelic variant of the Subtilase cytotoxin is common among Shiga toxin producing Escherichia coli of human and ovine origin. Clin Microbiol Infect. 2013;19:49-56.

25. Bosilevac JM, Koohmaraie M. Prevalence and characterization of non-O157 Shiga toxin-producing Escherichia coli isolates from commercial ground beef in the United States. Appl Environ Microbiol. 2011;77:2103–2112.

26. Bai X, Zhao A, Lan R, Xin Y, Xie H, Meng Q, et al. Shiga Toxin-Producing Escherichia coli in Yaks (Bos grunniens) from the Qinghai-Tibetan Plateau, China. PLoS One. 2013;8:e65537.

27. Yan X, Fratamico PM, Needleman DS, Bayles DO. DNA sequence and analysis of a 90.1-kb plasmid in Shiga toxin-producing Escherichia coli (STEC) O145:NM. Plasmid. 2012;68:25-32.

28. Johnson TJ, Nolan LK. Pathogenomics of the Virulence Plasmids of Escherichia coli. Microbiol. Mol Biol Rev. 2009;73:750-774.

29. Karama M, Johnson RP, Holtslander R, Gyles CL. Phenotypic and genotypic characterization of verotoxin-producing Escherichia coli O103:H2 isolates from cattle and humans. J Clin Microbiol. 2008;46:3569–3575.

30. Prager R, Liesegang A, Voigt W, Rabsch W, Fruth A, Tschäpe H. Clonal diversity of Shiga toxin-producing Escherichia coli O103:H2/H(-) in Germany. Infect Genet Evol. 2002;1:265–275.

31. Beutin L, Kaufluss S, Herold S, Oswald E, Schmidt H. Genetic analysis of enteropathogenic and enterohemorrhagic Escherichia coli serogroup O103 strains by molecular typing of virulence and housekeeping genes and pulsed-field gel electrophoresis. J Clin Microbiol. 2005;43:1552–1563.

32. Granobles CV, Sanso AM, Krüger A, Suárez L, Lucchesi PMA, Parma AE. Occurrence of subtilase cytotoxin and relation with other virulence factors in verocytotoxigenic Escherichia coli isolated from food and cattle in Argentina. Braz J Microbiol. 2011;42:711-715.

33. Orden JA, Horcajo P, de la Fuente R, Ruiz-Santa-Quiteria JA, Dominguez-Bernal G, et al. Subtilase cytotoxin-coding genes in verotoxin-producing Escherichia coli strains from sheep and goats differ from those from cattle. Appl Environ Microbiol. 2011;77:8259–8264.

34. Sánchez S, Beristain X, Martínez R, García A, Martín C, Vidal D, et al. Subtilase cytotoxin encoding genes are present in human, sheep and deer intimin-negative, Shiga toxin-producing Escherichia coli O128:H2. Vet Microbiol. 2012;159:531–535.

35. Sánchez S, Díaz-Sánchez S, Martínez R, Llorente MT, Herrera-León S, Vidal D. The new allelic variant of the subtilase cytotoxin (subAB2) is common among Shiga toxin-producing Escherichia coli strains from large game animals and their meat and meat products. Vet Microbiol. 2013;166:645-649.

36. Funk J, Stoeber H, Hauser E, Schmidt H. Molecular analysis of subtilase cytotoxin genes of food-borne Shiga toxin-producing Escherichia coli reveals a new allelic subAB variant. BMC Microbiol. 2013;13:230.

37. Khan AB, Naim A, Orth D, Grif K, Mohsin M, Prager R, et al. Serine protease espP subtype α, but not β or γ, of Shiga toxin-producing Escherichia coli is associated with highly pathogenic serogroups. Int J Med Microbiol. 2009;299:247–254.

38. Bugarel M, Beutin L, Martin A, Gill A, Fach P. Micro-array for the identification of Shiga toxin-producing Escherichia coli (STEC) seropathotypes associated with Hemorrhagic Colitis and Hemolytic Uremic Syndrome in humans. Int J Food Microbiol. 2010;142:318–329.
39. Brunder W, Schmidt H, Frosch M, Karch H. The large plasmids of Shiga-toxin-producing \textit{Escherichia coli} (STEC) are highly variable genetic elements. Microbiology. 1999;145:1005–1014.

40. Brandt SM, King N, Cornelius AJ, Premaratne A, Besser TE, On SLW. Molecular risk assessment and epidemiological typing of Shiga toxin-producing \textit{Escherichia coli} by using a novel PCR binary typing system. Appl Environ Microbiol. 2011;77:2458-2470.

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