Dasatinib enhances tumor growth in gemcitabine-resistant orthotopic bladder cancer xenografts

Stefan Vallo1,2, Martin Michaelis3, Kilian M. Gust2,4, Peter C. Black4, Florian Rothweiler1, Hans-Michael Kvasnicka5, Roman A. Blaheta2, Maximilian P. Brandt2, Felix Wezel6, Axel Haferkamp2 and Jindrich Cinatl Jr.1*

Abstract

Background: Systemic chemotherapy with gemcitabine and cisplatin is standard of care for patients with metastatic urothelial bladder cancer. However, resistance formation is common after initial response. The protein Src is known as a proto-oncogene, which is overexpressed in various human cancers. Since there are controversial reports about the role of Src in bladder cancer, we evaluated the efficacy of the Src kinase inhibitor dasatinib in the urothelial bladder cancer cell line RT112 and its gemcitabine-resistant sub-line RT112‘GEMCI20 in vitro and in vivo.

Methods: RT112 urothelial cancer cells were adapted to growth in the presence of 20 ng/ml gemcitabine (RT112‘GEMCI20) by continuous cultivation at increasing drug concentrations. Cell viability was determined by MTT assay, cell growth kinetics were determined by cell count, protein levels were measured by western blot, and cell migration was evaluated by scratch assays. In vivo tumor growth was tested in a murine orthotopic xenograft model using bioluminescent imaging.

Results: Dasatinib exerted similar effects on Src signaling in RT112 and RT112‘GEMCI20 cells but RT112‘GEMCI20 cells were less sensitive to dasatinib-induced anti-cancer effects (half maximal inhibitory concentration (IC50) of dasatinib in RT112 cells: 349.2 ± 67.2 nM; IC50 of dasatinib in RT112‘GEMCI20 cells: 1081.1 ± 377.2 nM). Dasatinib inhibited migration of chemo-naive and gemcitabine-resistant cells. Most strikingly, dasatinib treatment reduced RT112 tumor growth and muscle invasion in orthotopic xenografts, while it was associated with increased size and muscle-invasive growth in RT112‘GEMCI20 tumors.

Conclusion: Dasatinib should be considered with care for the treatment of urothelial cancer, in particular for therapy-refractory cases.

Keywords: Acquired resistance, Cancer cell line collection, Dasatinib, Gemcitabine, Orthotopic xenograft model, Urothelial bladder cancer
In other studies, however, Src inhibition increased bladder cancer cell migration and metastasis formation [11, 12]. A recently published phase II trial did not demonstrate clinical benefit of single-agent dasatinib in unselected patients with muscle-invasive urothelial carcinoma of the bladder [13].

Here, we investigated the effects of dasatinib on RT112 urothelial bladder cancer cells and RT112 cells with acquired resistance to gemcitabine (RT112'GEMC120) in cell culture and in an orthotopic xenograft model.

**Methods**

**Drugs**

Gemcitabine (Gemzar®) was obtained from Lilly (Indianapolis, IN, USA). Dasatinib was obtained from Absource Diagnostics (München, Germany).

**Cell lines and lentiviral transduction**

RT112 cells were obtained from the American Type Culture Collection (Manassas, VA, USA). Cells were grown in Iscove’s modified Dulbecco’s medium supplemented with 10% fetal calf serum (FCS, Gibco, Karlsruhe, Germany). RT112 cells were adapted to growth in presence of 20 ng/ml gemcitabine by continuous cultivation in the presence of increasing drug concentrations as previously described [14] resulting in the gemcitabine-resistant subline RT112'GEMC120. For in vivo studies, RT112 and RT112'GEMC120 cell lines underwent transduction with a lentiviral construct carrying the luciferase firefly gene for in vivo imaging resulting in cell lines RT112luc and RT112'GEMC120luc. The luciferase plasmid contained a blasticidin-resistance gene enabling positive selection with 10 mg/ml blasticidin (Life Technologies GmbH, Darmstadt, Germany). Cell lines were controlled for in vitro luciferase activity and cell number was correlated with bioluminescence using the Xenogen IVIS Spectrum (Caliper Lifesciences, Hopkinton, MA, USA) as previously described [15].

**Viability assay**

Cell viability was tested by 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) dye reduction assay after 120 h incubation modified as described before [16]. All experiments were performed at least in triplicate.

**Cell proliferation**

At day 0, 4000 cells/ml were plated in culture flasks. Cell numbers were determined every 24 h for 6 consecutive days using the automated cell counter Countess® (Life Technologies GmbH) after trypan blue staining. Doubling time (DT) was calculated using the formula DT = culture time/cell doubling. Cell doubling = ln(Nf/ Ni)/ln2, where Ni represents seeded cells number and Nf the harvested cells number [17]. All experiments were performed at least in triplicate.

**Western blot**

Cells were lysed in Triton X sample buffer and separated by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE). Proteins were detected using specific antibodies against β-actin (#A5441, Sigma-Aldrich, St. Louis, MO, USA), Src (#2123, Cell Signaling, Cambridge, UK), phosphorylated Src (Tyr416, #2101, Cell Signaling), Akt (#9272, Cell Signaling), and phosphorylated Akt (Thr308, #2965, Cell Signaling and Ser473, #04-736, EMD-Millipore, Billerica, MA, USA). Protein bands were visualized by enhanced chemiluminescence using a commercially available kit (GE Healthcare, Little Chalfont, UK). Pixel density of western blots is given in percentage compared to untreated cell line (100%). All experiments were performed at least in triplicate.

**Wound healing migration assay**

RT112 or RT112'GEMC120 cells were plated onto six-well plates and allowed to form a confluent monolayer. The cell monolayer was then scratched in a straight line to make a ‘scratch wound’ with a 0.2 ml pipette tip and the cell debris was removed by washing the cells with phosphate-buffered saline. Images of the closure of the scratch were captured at 0, 6, and 24 h. Quantification of wound repair was obtained from six measurements of every treatment from three independent experiments.

**Ethics statement**

All animal procedures were performed according to the guidelines of the Canadian Council on Animal Care and approved by the Institutional Review Board of the University of British Columbia (approval #A10-0295).

**Intramuscular orthotopic xenograft murine model**

Animal experiments were performed as described previously [15, 18]. Mice were subdivided into four treatment arms: RT112luc control treatment (n = 15), RT112luc dasatinib treatment (n = 15), RT112'GEMC120luc control treatment (n = 15), and RT112'GEMC120luc dasatinib treatment (n = 14). Six-week-old male nude mice (Harlan Laboratories, Indianapolis, IN, USA) were anesthetized with isoflurane (2 Vol. %) and analgesia was provided by subcutaneous injection with buprenorphine.
and meloxicam (Boehringer Ingelheim, Burlington, ON, Canada). After disinfection of the abdominal wall with chlorhexidine, a low transverse laparotomy was made and the urinary bladder was extracorporealized. 50 μl of a cell suspension containing 5 × 10^5 cells were directly injected into the bladder wall. The incision was closed with suture. Bioluminescence was used to quantify tumor burden and was measured on the xenogen IVIS spectrum imaging system after intraperitoneal injection of 200 μg/kg luciferin (Caliper Lifesciences). Images were taken at 10 and 14 min after luciferin injection and the average counts were used for statistical analysis. Evaluation of bioluminescence in this orthotopic model showed that 10 × 10^8 photons/s correlate to 10 mm^3 tumor volume [19]. Bioluminescence imaging was performed on day 5 after tumor inoculation and mice were divided into equal treatment groups based on tumor burden. Treatment started the following day and imaging was repeated every 4 days. Necropsy was performed after 4 weeks. Dasatinib was administered by oral gavage with an in vivo relevant dose of 20 mg/kg body weight (BW) twice daily [20] in sodium citrate/citric acid (Sigma-Aldrich, St. Louis, MO, USA). Vehicle treatment was prepared and administered in an identical manner, but without dasatinib. All experiments were performed at least in triplicate.

Haematoxylin and eosin staining (H&E)
All samples were fixed in buffered 4 % formalin (pH 7.4) and embedded in paraffin. We used standard procedures for deparaffinization and rehydration. Slides were cut at a microtome into 3 μm slices. H&E staining were performed according to standard.

Statistical analysis
Results are expressed as mean ± standard deviation (SD) of at least three independent experiments. For statistical analysis student’s t test, analysis of variance (ANOVA), and Student–Newman–Keuls-Test were performed whenever applicable. Significance was defined at values of p ≤ 0.05.

Results
Growth characteristics and sensitivity to gemcitabine of RT112 and RT112 rGEMCI20 cells
RT112 and RT112 rGEMCI20 cells showed similar growth kinetics with doubling times of 23.28 ± 2.88 h in RT112 cells and 25.2 ± 3.12 h in RT112 rGEMCI20 cells. Transduction with the luciferase plasmid, which is needed to monitor in vivo tumor growth, did not alter cell growth kinetics (doubling time RT112luc: 24.72 ± 5.04 h; doubling time RT112GEMCI20luc: 27.36 ± 3.6 h) (Fig. 1a; Table 1). RT112GEMCI20 cells (IC₅₀ = 125.40 ± 29.78 ng/ml) displayed a 77-fold increased resistance to gemcitabine compared to parental RT112 cells (IC₅₀ = 1.63 ± 0.55 ng/ml). There was a similar sensitivity to gemcitabine in the luciferase-transduced cell lines RT112luc and RT112GEMCI20luc compared to RT112 and RT112GEMCI20 [RT112luc (IC₅₀: 1.94 ± 0.24) and RT112GEMCI20luc (IC₅₀: 114.39 ± 14.52)] (Table 1).

Growth characteristics of RT112luc and RT112 rGEMCI20luc tumors in vivo
In vivo, growth of RT112luc and RT112 rGEMCI20luc tumors was investigated in an established murine orthotopic xenograft model [15, 18, 19, 21]. RT112GEMCI20luc cells were used for the first time in this orthotopic model. Nude mice received either 5 × 10^5 RT112luc or RT112GEMCI20luc cells directly injected into the bladder wall. Tumor take rates were 90 % (27 out of 30 animals) for RT112luc cells and 100 % (30 out of 30 animals) for RT112GEMCI20luc cells. RT112GEMCI20luc xenografts grew substantially slower than RT112luc xenografts resulting in a 6-fold smaller average tumor volume at day 25 (p < 0.0001) (Fig. 1b).

In vitro effects of dasatinib on RT112 and RT112 rGEMCI20 cells
RT112GEMCI20 cells (IC₅₀: 1081.1 ± 239.2 nM) showed a 31-fold decreased sensitivity to dasatinib compared to parental RT112 cells (IC₅₀: 349.2 ± 67.2 nM). The concentration-dependent drug response from a representative experiment is shown in Fig. 1c.

In cell culture, dasatinib caused in RT112 and in RT112GEMCI20 cells a dose-dependent reduction of Src phosphorylation. Phosphorylation of Akt (Thr308) was also reduced in both cell lines with a stronger inhibition in the chemo-naive cell line. No inhibition of pAkt (Ser473) was detected (Fig. 2a, b).

Since Src inhibition was described to increase migration in urothelial cancer [12], we evaluated migratory behavior using a wound healing scratch assay in RT112 and RT112GEMCI20 cells with and without dasatinib treatment. Results indicated a similar time-dependent migration inhibition in response to dasatinib in RT112 and RT112GEMCI20 cells resulting in >60 % inhibition of migration in both cell lines relative to untreated control (p < 0.0001) (Fig. 2c, d).
Effects of dasatinib treatment in RT112 luc and RT112’GEMCI20 luc xenografts

Dasatinib treatment using 20 mg/kg twice daily [20] caused a reduction of RT112luc xenograft size by 39% (p = 0.036) relative to untreated control at day 25. In contrast, dasatinib treatment induced a 4-fold increase in RT112’GEMCI20luc tumor size compared to vehicle treated RT112’GEMCI20luc xenografts at day 25 (p < 0.001) (Fig. 3).

Tumors were sampled and H&E staining was performed at day 25. Untreated RT112luc tumors displayed muscle-invasive growth (pT2) (Fig. 4a). In contrast, no signs of muscle-invasive growth (pT1) were detected in dasatinib-treated RT112luc tumors (Fig. 4b). The opposite pattern was observed in RT112’GEMCI20luc tumors. Untreated RT112’GEMCI20luc tumors did not show muscle-invasive growth (pT1) (Fig. 4c), while dasatinib-treated RT112’GEMCI20luc xenografts displayed muscle invasive growth (pT2) (p < 0.001). In dasatinib-treated RT112’GEMCI20luc xenografts, the bladder muscle was completely infiltrated by the tumor, and the bladder wall was not clearly visible anymore (Fig. 4d).
Discussion

In this study, we compared the effects of the Src inhibitor dasatinib on the urothelial cancer cell line RT112 and its gemcitabine-resistant sub-line RT112’GEMCl20 in cell culture and in an orthotopic bladder cancer xenograft model in mice.

In cell culture, both cell lines displayed similar growth kinetics. Dasatinib inhibited Src phosphorylation in RT112 and RT112’GEMCl20 cells at low nanomolar concentrations similar to those that had already been described to affect Src phosphorylation [9]. While dasatinib had previously been shown to interfere with the phosphorylation of Akt (Thr308 and Ser473) in squamous cell lung cancer [22], we only detected inhibition of phosphorylation of Akt (Thr308). The reasons for this may be the consequence of cell type-specific differences.

Fig. 2  a Representative western blots showing cellular levels of Src, pSrc, Akt, pAkt, and β-actin in RT112 and RT112’GEMCl20 cells after 24 h incubation with dasatinib.  b Pixel density of western blots is given in percentage compared to untreated cell line (100 %). One of three independent experiments is shown here. *p ≤ 0.05 vs untreated cell line.  c Wound healing migration assay. Effects of dasatinib (100 nM) treatment on RT112 or RT112’GEMCl20 cells were evaluated. Images of the closure of the scratch were captured at 0, 6, and 24 h. Representative images of scratch wound assays were shown (magnification ×100).  d Quantification of wound repair was obtained from six measurements of every treatment from three independent experiments.
between the investigated models. Although dasatinib exerted similar effects on Src signaling in RT112 and RT112’GEMCI20 cells, its effects on cell viability differed between the two cell lines. The effective concentrations of dasatinib in RT112 cells (IC50 of 349.2 ± 67.2 nM) were in the range of those previously described for the treatment of urothelial cancer cells [9]. However, in RT112’GEMCI20 cells the IC50 value was about 3-fold higher (1081.1 ± 239.2 nM). This suggests that RT112’GEMCI20 cells have acquired resistance mechanisms that interfere with antitumoral effects of dasatinib downstream of Src signaling. In accordance, dasatinib treatment resulted in reduced inhibition of the phosphorylation of the Src downstream kinase Akt (Thr 308) in RT112’GEMCI20 cells compared to RT112 cells.

In nude mice, RT112’GEMCI20luc cells formed about 6-fold smaller tumors than RT112luc cells. The reasons for this remain unclear. However, it is known that different cell lines may differ in their interaction with the tissue environment in animal models resulting in discrepancies in the growth kinetics [23, 24].

The most striking differences were found after oral dasatinib treatment of mice bearing orthotopic RT112luc or RT112’GEMCI20luc xenografts. Dasatinib treatment of RT112luc bladder tumors resulted in a significant reduction of tumor size relative to untreated control. In contrast, dasatinib treatment of RT112’GEMCI20luc bladder tumors resulted in a dramatic increase of tumor growth. In addition, the dasatinib treatment prevented muscle-invasive growth of RT112luc xenografts, but strongly induced muscle-invasive growth of RT112’GEMCI20luc xenografts. The mechanistic reasons underlying this discrepancy remain elusive.

Wu et al. [11] had reported that loss of Src increases metastasis formation in a bladder cancer model. Similar to this report, Thomas et al. [12] showed that Src enhances urothelial cancer cell migration and metastasis formation. However, dasatinib inhibited RT112 and RT112’GEMCI20 cell migration in a similar manner, similarly to other studies that reported on the effects of Src inhibition on cancer cell migration in models from different cancer entities [25–28]. Therefore, it seems unlikely that the dasatinib-induced increased invasive growth of RT112’GEMCI20luc cells in the xenograft model may be caused by dasatinib-induced enhanced RT112’GEMCI20luc cell migration.

![Fig. 3 Tumor growth kinetics of murine orthotopic a RT112luc and b RT112’GEMCI20luc xenografts. Dasatinib was given per oral gavage at a dose of 20 mg/kg body weight (BW) twice daily. Control animals were vehicle-treated in the same way. 10 × 10^6 photons/s correlate to 10 mm^3 tumor volume (21). c Representative photographs of treated and untreated RT112luc and RT112’GEMCI20luc xenografts with fluorescent bladder tumors.](image-url)
Conclusions
We present the first study that investigated the effects of dasatinib on urothelial cancer cells with acquired resistance to gemcitabine. In cell culture, gemcitabine-resistant RT112 cells were less sensitive to dasatinib than parental RT112 cells. Notably, parental RT112 cells and gemcitabine-resistant RT112 cells displayed an unexpected opposite response to dasatinib in an orthotopic xenograft model in mice. While dasatinib inhibited tumor growth and muscle invasion by parental RT112 cells, it increased tumor growth and muscle invasion by gemcitabine-resistant RT112 cells. Thus, our data do not generally support the use of dasatinib for the treatment of urothelial cancer, in particular not for therapy-refractory cases after first-line chemotherapy with gemcitabine. However, further studies will need to show whether similar effects are obtained in additional models of (acquired gemcitabine resistance in) urothelial cancer. If such studies suggested that some urothelial cancer diseases may benefit from dasatinib therapy, biomarkers would need to be identified that enable the prediction of the response of individual urothelial cancer diseases to dasatinib.

Abbreviations
MTT: 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; IC50: half maximal inhibitory concentration; GC: gemcitabine and cisplatin; DT: doubling time; H&E: haematoxylin and eosin staining; SD: standard deviation; ANOVA: analysis of variance.

Authors’ contributions
SV, KG: data collection, project development, data analysis, manuscript writing. MM, PB, JC: project development, data analysis, manuscript writing. FR: data analysis, data analysis. HK: data collection, manuscript writing. RB, MB, FW: data analysis, manuscript writing. AH: data analysis, project development. All authors read and approved the final manuscript.
Author details

1 Institute of Medical Virology, Goethe University Frankfurt, Paul-Ehrlich-Str. 40, 60596 Frankfurt am Main, Germany. 2 Department of Urology, Goethe University Frankfurt, Frankfurt, Germany. 3 Centre for Molecular Processing and School of Biosciences, University of Kent, Canterbury, UK. 4 Department of Urologic Sciences, Vancouver Prostate Centre, University of British Columbia, Vancouver, Canada. 5 Dr. Senckenberg Institute of Pathology, Goethe University Frankfurt, Frankfurt am Main, Germany. 6 Department of Urology, University Hospital Ulm, Ulm, Germany.

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Competing interests

The authors declare that they have no competing interests.

Availability of data and materials

All data and further information about “Methods” section are available via the corresponding author (Cinatl@em.uni-frankfurt.de).

Ethics

All animal procedures were performed according to the guidelines of the Canadian Council on Animal Care and approved by the Institutional Review Board of the University of British Columbia (approval #A10-0295).

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References

1. Pectasides D, Pectasides M, Economopoulos T. Systemic chemotherapy in locally advanced and/or metastatic bladder cancer. Cancer Treat Rev. 2006;32(6):456–70.
2. von der Maase H, Sengelov L, Roberts JT, Ricci S, Dogliotti L, Oliver T, Moore MJ, Zimmerman A, Arning M. Long-term survival results of a randomized trial comparing gemcitabine plus cisplatin, with methotrexate, vinblastine, doxorubicin, plus cisplatin in patients with bladder cancer. J Clin Oncol. 2005;23(21):4602–8.
3. Sonpavde G, Galsky MD, Hutson TE. Current optimal chemotherapy for advanced urothelial cancer. Expert Rev Anticancer Ther. 2008;8(1):51–61.
4. Choueiri TK, Ross RW, Jacobus S, Quinn DI, Vaishampayan U, Yu EY, Arora S, Majid S, Shahryari V, Chen Y, Deng G, Yamamura S, Boyer B, Bourgeois Y, Poupon MF. Src kinase contributes to the metastatic phenotype of the 53 mutated multi-drug-resistant cancer cells. Cell Death Dis. 2011;2:e243.
5. Irby RB, Yeatman TJ. Role of Src expression and activation in human carcinomas of the breast. Cell. 2000;101(1):1040–4.
6. Broder MS, DeGregori J, Anderson SM, Reymond ME. Inhibiting tyrosine phosphorylation of protein kinase Cβ (PKCβ) protects the salivary gland from radiation damage. J Biol Chem. 2014;289(15):10900–8.
7. Sonpavde G, Morrissey SC, et al. Adaptation of cancer cells from different entities to the MDM2 inhibitor nutlin-3 results in the emergence of p53-mutated multi-drug-resistant cancer cells. Cell Death Dis. 2011;2:6243.
8. Buettner R, Mesa T, Vultur A, Lee F, Jove R. Inhibition of Src family kinases in p53-mutated multi-drug-resistant cancer cells. Cell Death Dis. 2011;2:e243.
9. Hadaschik BA, Black PC, Sea JC, Metwalli AR, Dinney CP, Gleave ME. Sorafenib inhibited angiogenesis and tumour growth and spread in a mouse model of orthotopic bladder cancer using transurethral tumour inoculation and bimolecular imaging. BJU Int. 2007;100(6):1377–84.
10. Hadaschik BA, Black PC, Sea JC, Metwalli AR, Dinney CP, Gleave ME. Sorafenib inhibited angiogenesis and tumour growth and spread in a mouse model of orthotopic bladder cancer using transurethral tumour inoculation and bimolecular imaging. BJU Int. 2007;100(6):1377–84.
11. Hadaschik BA, Black PC, Sea JC, Metwalli AR, Dinney CP, Gleave ME. Sorafenib inhibited angiogenesis and tumour growth and spread in a mouse model of orthotopic bladder cancer using transurethral tumour inoculation and bimolecular imaging. BJU Int. 2007;100(6):1377–84.
12. Rutigliano L, Iavarone A, Czerniak B, Dinney CP, Black PC. Fibroblast growth factor receptor 1 is a rational therapeutic target in bladder cancer. Mol Cancer Ther. 2013;12(7):1245–54.
13. Hadaschik BA, Black PC, Sea JC, Metwalli AR, Dinney CP, Gleave ME. Sorafenib inhibited angiogenesis and tumour growth and spread in a mouse model of orthotopic bladder cancer using transurethral tumour inoculation and bimolecular imaging. BJU Int. 2007;100(6):1377–84.
14. Hadaschik BA, Black PC, Sea JC, Metwalli AR, Dinney CP, Gleave ME. Sorafenib inhibited angiogenesis and tumour growth and spread in a mouse model of orthotopic bladder cancer using transurethral tumour inoculation and bimolecular imaging. BJU Int. 2007;100(6):1377–84.
15. Hadaschik BA, Black PC, Sea JC, Metwalli AR, Dinney CP, Gleave ME. Sorafenib inhibited angiogenesis and tumour growth and spread in a mouse model of orthotopic bladder cancer using transurethral tumour inoculation and bimolecular imaging. BJU Int. 2007;100(6):1377–84.
16. Hadaschik BA, Black PC, Sea JC, Metwalli AR, Dinney CP, Gleave ME. Sorafenib inhibited angiogenesis and tumour growth and spread in a mouse model of orthotopic bladder cancer using transurethral tumour inoculation and bimolecular imaging. BJU Int. 2007;100(6):1377–84.
17. Hadaschik BA, Black PC, Sea JC, Metwalli AR, Dinney CP, Gleave ME. Sorafenib inhibited angiogenesis and tumour growth and spread in a mouse model of orthotopic bladder cancer using transurethral tumour inoculation and bimolecular imaging. BJU Int. 2007;100(6):1377–84.