Uracil DNA glycosylase (UDG) activities in *Bradyrhizobium diazoefficiens*: characterization of a new class of UDG with broad substrate specificity

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ABSTRACT

Repair of uracils in DNA is initiated by uracil DNA glycosylases (UDGs). Family 1 UDGs (Ung) are the most efficient and ubiquitous proteins having an exquisite specificity for uracils in DNA. Ung are characterized by motifs A (GQDPY) and B (HPSPLS) sequences. We report a novel dimeric UDG, Blr0248 (BdiUng) from *Bradyrhizobium diazoefficiens*. Although BdiUng contains the motif A (GQDPA), it has low sequence identity to known UDGs. BdiUng prefers single stranded DNA and excises uracil, 5-hydroxymethyl-uracil or xanthine from it. BdiUng is impervious to inhibition by AP DNA, and Ugi protein that specifically inhibits family 1 UDGs. Crystal structure of BdiUng shows similarity with the family 4 UDGs in its overall fold but with family 1 UDGs in key active site residues. However, instead of a classical motif B, BdiUng has a uniquely extended protrusion explaining the lack of Ugi inhibition. Structural and mutational analyses of BdiUng have revealed the basis for the accommodation of diverse substrates into its substrate binding pocket. Phylogenetically, BdiUng belongs to a new UDG family. *Bradyrhizobium diazoefficiens* presents a unique scenario where the presence of at least four families of UDGs may compensate for the absence of an efficient family 1 homologue.

INTRODUCTION

Uracil (as dUMP) can be incorporated into DNA by DNA polymerases (from dUTP, forming an A:U pair) or arise by deamination of cytosines (resulting in G:U pairs) (1,2). If unrepaired, A:U base pairs can hinder recognition by DNA binding proteins (3,4) while G:U base pairs can cause G:C to A:T mutations in the subsequent rounds of DNA replication, posing a threat to genomic integrity. Enzymes of the uracil DNA glycosylase (UDG) superfamily initiate base excision repair to remove uracil residues (5–7). Five distinct families of UDGs have so far been established which differ in their primary amino acid sequences and substrate specificities. However, UDGs possess a conserved α/β structural fold and seemingly a common evolutionary origin (8,9). Ubiquitously found family 1 UDGs (Ung), exemplified by the well-studied ‘master catalyst’ *Eco*Ung, are the most efficient of UDGs and show an exquisite specificity for uracil in DNA (10). Conserved orthologues of family 1 are found in nearly all organisms studied, including many viruses (5,10). This UDG family is characterized by motif A (GQDPY) responsible for catalysis and motif B (HPSPLS) implicated in the stabilization of the enzyme substrate complex (11–14). Interestingly, phage PBS-1 or 2 that infects *Bacillus subtilis* encodes a protein inhibitor, Ugi that forms a physiologically irreversible non-covalent complex in 1:1 stoichiometry specifically with family 1 UDGs to allow occurrence of uracil in their genomes (15,16).

While *Eco*Ung was the first UDG to be identified and extensively studied, several UDGs have been reported since then (5,17–21). Family 2 UDGs referred to as Mug/TDG have GINPG as motif A and MPSSSAR as motif B. While Mug/TDG excise uracil from double stranded (ds) DNA

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(G:U and A:U pairs) with a low efficiency, they also excise thymine from T:G pairs (18, 22, 24). GMNPG and HP-SPRN define motif A and motif B for family 3 UDGs (SMUG). SMUGs excise uracil from single stranded (ss) DNA with higher efficiency than from dsDNA (19, 25). Family 4 and family 5 UDG members contain a 4Fe–4S cluster. Motif A and Motif B are represented by GEx(A/G)P and HPAAVL, respectively, for family 4, and by GLAPA and HPSPLN for family 5 (20, 21, 26, 27). While family 4 UDGs act on ssDNA and dsDNA substrates, UDG activity of family 5 is enhanced on dsDNA substrates.

Members of Rhizobiaceae are an important group of soil bacteria that are recognized for their ability to fix atmospheric nitrogen through a symbiotic association with root nodules of leguminous plants. Formation and maintenance of root nodules in leguminous plants represents an interesting scenario of oxidative and nitrosative stress to invading rhizobial cells. To establish symbiotic associations with leguminous plants and to form nitrogen fixing root nodules through a symbiotic association with root nodules of leguminous plants, plants should efficiently overcome the initial defense response elicited by the plant. Similar to the leguminous plants, rhizobia should efficiently overcome the testing scenario of oxidative and nitrosative stress to invade root nodules in leguminous plants. Formation and maintenance of root nodules in leguminous plants represents an interesting scenario of oxidative and nitrosative stress to invading rhizobial cells. To establish symbiotic associations with leguminous plants, rhizobia should efficiently overcome the initial defense response elicited by the plant. Similar to the response to pathogens, plants also respond to rhizobial infection through an oxidative burst, with enhanced production of reactive oxygen species (ROS) (31–33). In addition to ROS, nodules also generate reactive nitrogen species (RNS) like nitric oxide (NO) and peroxynitrite (ONOO−) (42, 43).

Oxidative and nitrosative stress result in various kinds of lesions in DNA including deamination of cytosines, posing a threat to the genomic integrity of rhizobia (44, 45). Interestingly, their genome contains high G+C content making them inherently more susceptible to cytosine deamination events that produce uracil in the genome which can lead to transition mutations. Hence, one could expect these organisms to have evolved sophisticated strategies of DNA repair to cope with their hostile environment. However, aspects of DNA repair in rhizobium are currently under explored. In the present study, we used a member of α-proteobacteria, Bradyrhizobium diazoefficiens (previously B. japonicum) (46) as our model organism, which is well known for its ability to form nitrogen fixing root nodules through a symbiotic association with Glycine max (47, 48) to explore aspects of uracil excision repair and report the presence of at least four UDGs in this organism with a distinct absence of a classical family 1 UDG. Furthermore, we demonstrate that in Bradyrhizobium diazoefficiens there exists a unique base excision repair system catalyzed by a member of a new UDG family.

MATERIALS AND METHODS

Bacterial strains, plasmids, DNA oligomers, media and growth conditions

Bacterial strains and plasmids are listed in Supplementary Table S1. DNA oligomers are listed in Supplementary Table S2. Escherichia coli CC102 (49) was obtained from Coli Genetic Stock Centre (CGSC). E. coli strains were grown in Luria-Bertani broth (LB) or on LB containing 2% (w/v) agar (Difco, USA) at 37°C. Bradyrhizobium diazoefficiens was grown in TY medium (0.5% tryptone, 0.3% yeast extract, 6 mM CaCl2) or on TY with 2% agar at 30°C. Ampicillin (Amp), kanamycin (Kan) and chloramphenicol (Cm) were added to media at a concentration of 100, 25 and 30 μg ml−1 respectively.

Cell lysate preparation

Saturated cultures were obtained in 2 ml media by incubation at 37°C overnight for E. coli strains and at 30°C for 5–6 days for B. diazoefficiens and then subcultured (1% for E. coli and 10% for B. diazoefficiens) into 2 ml media and incubated until the OD600 reached ~0.6. Cells were pelleted, re-suspended in 200 μl TME (25 mM Tris–HCl pH 8.0, 2 mM β-mercaptoethanol, 1 mM Na2EDTA), sonicated with 2 s alternate pulses for 20 s, six to seven times with 50–60 s intervals and centrifuged at 13 000 rpm for 1.5 h on table top centrifuge at 4°C. The supernatant was collected, mixed with an equal volume of 2× storage buffer (80% glycerol, 1 mM β-mercaptoethanol, 20 mM Tris–HCl pH 8.0 and 200 mM NaCl) and stored at −20°C. Protein concentration in cell extracts was estimated by the Bradford assay using BSA as standard (50).

Cloning of putative UDGs

Bdi-ung ORF was PCR amplified from genomic DNA with forward (BdiUng Fp, 5′-GCATTTCATATGCTCACAG ATTTCG-3′) and reverse (BdiUng Rp, 5′-GGCCAAAGCT TCGGGTTTGCGGCCG-3′) primers containing NdeI and HindIII sites, respectively and Pfu DNA polymerase. The reaction was heated at 94°C for 4 min followed by 35 cycles of incubations at 94°C for 1 min, 50°C for 45 s and 70°C for 50 s. The PCR product (856 bp) was ligated into pTrcNdeIHis vector through NdeI and HindIII sites, such that a hexa-His tag is attached to the N-terminus of the protein. The clones were confirmed by DNA sequencing. Other putative UDG genes, including bll3023, bhr5068 and bhr6661, were also similarly cloned into pTrcNdeIHis vector (with hexa-His tag, through NdeI and HindIII sites).

Purification of proteins

For protein overexpression and purification, plasmid pTrcBdi-Ung was introduced into E. coli Δung strain by transformation. The E. coli TG1Δung::cm harbouring pTrcBdi-Ung was inoculated into 10 ml LB containing Amp and Cm and grown overnight. The inoculum was diluted 1:100 (v/v) (1%) into 600 ml fresh medium and grown at 37°C to an OD600 of ~0.6. The culture was induced with 0.1 mM isopropyl-β-D-thiogalactopyranoside (IPTG) and grown for another 4–5 h. Cells were harvested by centrifugation (7500 rpm, 4°C, 5 min), resuspended in buffer A (20 mM Tris–HCl pH 8.0, 500 mM NaCl, 10% glycerol v/v, 2 mM β-mercaptoethanol) containing 30 mM imidazole, sonicated and centrifuged at 29 000 rpm (Avanti™ J-30I, JA30.50 Ti) for 2 h 30 min at 4°C. The supernatant was loaded onto a 5 ml Ni-NTA column pre-equilibrated with buffer A, washed with 30 ml of buffer A and eluted with a linear gradient of 30 mM to 1 M imidazole in buffer A.
Aliquots from the fractions were analyzed on 15% SDS-PAGE, fractions containing pure protein were pooled, dialyzed against buffer A, then against storage buffer (20 mM Tris–HCl pH 8.0, 300 mM NaCl, 50% glycerol v/v, 2 mM β-mercaptoethanol), and stored at −20°C. Protein concentration was estimated by the Bradford assay using BSA as standard (50). Other UDGs were also purified using the same procedure, except that BL3023 was purified from E. coli TG1, cultured in the presence of 0.01% FeCl3.

DNA oligomers and their radiolabelling

The 24 nucleotide long ssU9 (ctcaagtgUagcatgcaagact) and the 37 nucleotide long tetra-loop hairpin GU9 (ctcaagtgUagcatgttggatctgccactgtg) containing uracil in the stem region, were used as ssDNA and dsDNA substrates, respectively. Other DNA oligomers are listed in Supplementary Table S2. DNA oligomers (10 pmol) were 5’ 32P-end labelled using 10 μCi of [γ-32P] ATP (6000 Ci/mmoll) and T4 polynucleotide kinase and purified on Sephadex G-50 minicolumns (1 ml). To generate duplexes (HmU and ssX9 with ssU9-Compl-G, Compl-T with G-oligo, and I with Compl-G), 5’ 32P-end labelled DNA oligomers (with modified bases) were annealed with complementary oligomers in approximately equimolar ratios in 40 mM Tris–HCl pH 7.5, 20 mM MgCl2 and 50 mM NaCl, heated at 90°C for 5 min and then slowly cooled to room temperature.

Base excision and inhibition assays

Appropriate amounts of cell extract/pure protein were incubated with 5’ 32P-end labelled DNA oligomers (~10000 cpm; ~10 nM concentration) in 10 μl reaction volumes in UDG buffer (50 mM Tris–HCl pH 8.0, 1 mM Na2EDTA, 1 mM DTT and 25 μg ml−1 of BSA) at 37°C for the indicated times. The reactions were stopped by the addition of 0.2 N NaOH and formamide dye (80% formamide, 0.05% each of bromophenol blue and xylene cyanol FF, 10 mM NaOH and 2 mM Na2EDTA) and boiled at 90°C for 15 min. Aliquots (20 μl) were analyzed on 15% polyacrylamide (19:1) 8 M urea gels, exposed to phosphorimaging screens and imaged using Fujiﬁlm analyzer. To see if BdiUng is inhibited by Ugi, BdiUng (1 μg; ~32 pmol of monomer) was pre-incubated with excess Ugi (1 μg; ~100 pmol) in 4 μl volume for 30 min before the reaction.

Gel filtration chromatography

Proteins were subjected to gel filtration chromatography using Superdex 75 (GE Healthcare) column equilibrated with a buffer containing 20 mM Tris–HCl pH 8.0, 150 mM NaCl, 2 mM β-mercaptoethanol, 10% glycerol. Elution was performed with the same buffer at a flow rate of 1 ml/min and the absorbance at 280 nm was recorded to obtain the elution profile.

EMSA for checking Ugi binding

Ugi (~8 μg, ~850 pmol) was incubated with BdiUng (5 μg, ~160 pmol of monomer) or EcoUng (2 μg, ~80 pmol) in 25 μl volumes in UDG buffer at room temperature for 20 min and then stored on ice (4°C) for 15 min. Loading dye (5 μl) containing 30% (v/v) glycerol, 0.25% (w/v) bromophenol blue and 0.25% (w/v) xylene cyanol FF was added, and the samples were analyzed on 15% native PAGE (29:1 crosslinking, pH 6.8). After electrophoresis, the gel was stained using Coomassie brilliant blue.

Impact of uracil and AP DNA on activity of BdiUng

To check the effect of free uracil and AP DNA on uracil excision enzymes, varying concentrations of free uracil or unlabelled ssF9 (ctcaagtgFagctgcaagact, where F is tetrahydrofuran, a stable mimic of AP site provided by Midland Certified Reagent Co.) were pre-incubated with 100 pg (~4 fmol) EcoUng or 10 ng (~0.32 pmol monomer) BdiUng at room temperature followed by incubation on ice for 15 min. The reaction mixtures were followed by addition of 5’ end 32P-labelled ssU9 (~0.1 pmol) and incubated at 37°C for 30 min for uracil excision.

Generation of site directed mutation in BdiUng, and its characterization

PCR based method (see supplementary material) was used to generate A59Y mutant of BdiUng. Mutant protein was purified from E. coli TG1Δung::cm strain using Ni-NTA column chromatography as described for the wild type BdiUng.

Generation of ung knockout in CC102 strain

A P1 phage lysate generated using E. coli MG1655Δung::kan was used to transduce the Δung::kan allele into E. coli strain CC102. Transfer of the Δung::kan allele into the transductants was verified by diagnostic PCR (with EcoUng-up-Fp and EcoUng-dn-Rp primers which flank the ung locus) and uracil excision assays (Supplementary Figure S1).

Assay for reversion of Lac− to Lac+ in E. coli

E. coli CC102 with a mutation in lacZ was used to assay for reversion to Lac+. Isolated colonies of this strain and its Δung::kan derivative were transformed with pTrc99c or EcoUng/BdiUng expression constructs. Cells were inoculated in replicates of ten in 2 ml LB supplemented with Amp and grown for 24 h with shaking at 37°C. Serial dilutions of saturated cultures were made and 50 μl of 10−4 dilution was plated onto M9 minimal media plates containing 0.2% glucose for viable counts. The saturated culture (1.5 ml) was spun down and plated onto M9 minimal media containing 0.2% lactose for counts of Lac+ revertants. The plates were incubated at 37°C until colonies appeared (24 h for glucose plates and 72 h for lactose plates). The reversion frequency was obtained by dividing the number of revertants (per ml culture) by the number of bacteria plated (viable count).

Crystallization and refinement

The purified BdiUng was dialyzed against 20 mM Tris–HCl, 50 mM NaCl, 5 mM β-mercaptoethanol, pH 8.5 and...
RESULTS

Identification of UDG homologues and Blr0248 (BdiUng) as putative UDGs

Analysis of B. diazoefficiens USDA110 genome through blast searches indicated the absence of a conserved family 1 homologue in this organism. Further analysis based on motif based BLAST (NCBI) searches revealed many putative UDGs (Table 1), among which most could be classified into established families of UDGs: bfr6661 (family 2 MUG), bll2523 (family 4 UdgA), blr5068 (family 5 UdgB) and bll3023 (family 4 UdgX). Of particular interest among the identified ORFs was blr0248 which codes for a 272 amino acid long protein Q89XRO_BRAJA containing a putative motif A (GQDPA) similar to family 1 UDGs (referred to as BdiUng). However, BdiUng lacked the motif B of family 1 UDGs and showed low sequence similarity to established UDG families (Supplementary Figure S2). Homology searches through BLAST revealed several homologues of BdiUng from many organisms (Figure 1), which form the putative members of a new family of UDGs. Homologues of BdiUng were found in various lineages of bacteria including actinobacteria (Streptacidiphilus rugosus, Kitasatospora azatica, Intrasporangium calvum, Streptomyces xylophagus, Frankia sp. EuI1c and Janibacter sp. HTCC2649) and β-proteobacteria (Burkholderia ubonensis, Chromobacterium haemolyticum, Nitrosospira sp. NpAV). Evolutionary analysis based on neighbor joining method (Supplementary Figure S3) showed that these bacteria possess a new family of UDGs related to family 1.

Cloning and purification of putative UDGs

We cloned ORFs of blr0248, bfr6661, blr5068 and bll3023 from B. diazoefficiens USDA110 into the pTrcNdeIHis expression vector along with a 20 amino acid long presequence containing a hexa-His tag. All UDGs other than Bll3023 were purified from an E. coli TG1ΔUng::cm strain to ensure that the UDG preparations were free from EcoUng contamination (Supplementary Figure S4). Since UdgX proteins are toxic in ung− strains (56), Bll3023 was purified from E. coli TG1. As shown in Supplementary Figure S5, Bll3023, Bfr6661, Blr5068 possessed uracil excision activities, which corresponded to their known classes of proteins, and were not pursued any further.

BdiUng: a novel Ugi insensitive UDG with broad substrate specificity

The activity of purified BdiUng was analyzed on uracil containing ssDNA and dsDNA oligomers (ssU9 and GU9) and was found to be active on both the substrates (Figure 2A and B, compare lanes 2 with 1). BdiUng reactions with/without preincubation with Ugi showed product bands of similar intensity (compare lanes 2 and 3). However, under the same conditions, preincubation of EcoUng with Ugi completely inhibited uracil excision activity (compare lanes 5 with 4 and 1). These observations suggest that Ugi does not inhibit BdiUng to any detectable levels and reveals a fundamental difference of BdiUng to family 1 UDG members.

Although the activity of BdiUng was not inhibited by Ugi, it was still possible that it formed a complex with Ugi (like family 1 UDGs). Therefore, electrophoretic mobility shift assays were carried out using native PAGE after preincubation of BdiUng or EcoUng with Ugi (Figure 2C). As a control, EcoUng–Ugi complex formed in vivo was also loaded (lane 4). While pre-incubation of EcoUng with Ugi resulted in a shift in the migration of EcoUng in native PAGE (compare lane 2 with lanes 1 and 3), BdiUng did not show any shift in its migration (nor a new band) upon pre-incubation with Ugi (compare lanes 5 and 6) suggesting the absence of complex formation. The EcoUng–Ugi (lane 2) complex migrated to the same position for purified EcoUng–Ugi complex (lane 4) formed in vivo (57).

Even though the bioinformatics and biochemical analyses with BdiUng suggested that B. diazoefficiens lacks a classical family 1 UDG, to rule out other unidentified proteins as family 1 UDG (Ung), we decided to carryout UDG assays with cell free extracts prepared from B. diazoefficiens (Figure 2D). As expected, the assays revealed UDG activity (compare lane 5 with 1). However, the activity was not diminished by Ugi (compare lane 5 with lanes 6 and 7). As a control, UDG activity of E. coli cell free extracts was inhibited by Ugi (compare lane 2 with lanes 3 and 4). These observations suggest that B. diazoefficiens lacks Ugi sensitive UDG activity.

To check if BdiUng is inhibited by either of the reaction products (uracil, or AP DNA) we employed a uracil excision assay with 5´end 32P-labelled ssU9, in the presence of varying concentrations of uracil or a tetrahydrofuran-containing DNA (ssF9), a stable mimic of AP DNA (Figure 3). While free uracil caused a 5-fold reduction in BdiUng activity at 10 mM concentration (panels i and ii), ssF9, in spite of its high concentrations (of up to 200 μM) did not significantly inhibit BdiUng (panels iii and iv). As a control, both the products inhibited uracil excision activity of EcoUng (panels i–iv).

Substrate specificity of BdiUng

To investigate if BdiUng is highly specific towards uracil in DNA (like family 1 UDGs, Ung) or relaxed in substrate
Figure 1. Multiple sequence alignment of BdiUng with putative homologues. The protein sequence of BdiUng was aligned against the protein sequences of FUDG (from Frankia sp. EuI1c), SrUDG (from Streptacidiphilus rugosus), KaUDG (from Kitasatospora azatica) and NsUng (from Nitrosospira sp. NpAV) using clustal omega. The schematic representation was obtained by using BOXSHADE (www.ch.embnet.org). Identical residues are shaded in black while similar residues in grey. Putative motif A and B are additionally shaded in red and blue, respectively.

Table 1. Putative UDGs identified from Bradyrhizobium diazoefficiens through genome analysis

| Gene   | Protein   | Motif                       |
|--------|-----------|-----------------------------|
| bbl0248| Q89XRO_BRAJA | GQDPA                      |
| bbl2523| Q89S82_BRAJA | UDG F4 TTUDGA like         |
| bbl3023| Q89QV1_BRAJA | UDG F4 TTUDGA like         |
| bbl5068| Q89K49_BRAJA | UDG F5 TTUDGB like         |
| bbl6661| Q89FNB_BRAJA | UDG F2 MUG                 |

choice like UDGs belonging to some other families, we used 5' end 32P-labelled DNA oligomers containing different base modifications. BdiUng utilized DNA oligomers containing xanthine (X) (Figure 4A, compare lane 8 with 7) and a uracil analogue 5-hydroxymethyluracil (HmU) (Figure 4A, compare lane 10 with 9) in ssDNA as substrates. However, it did not utilize DNA oligomer containing inosine (I) in ssDNA or dsDNA contexts to any detectable levels (Figure 4A and B). Thymine DNA glycosylase activity was also not detected (over the control reaction) using a G:T pair containing dsDNA (Figure 4B, lanes 3 and 4).

To determine the minimum substrate from which BdiUng could excise uracil, we used 5' end 32P-labelled DNA oligomers having a different number of bases on the 5' and 3' sides of the uracil (Figure 5A). EcoUng whose activity on these substrates has been well characterized (51) was used as control. With reference to EcoUng, BdiUng activity was detectable on oligomers having one phosphorylated nucleotide 5' of the uracil (compare lane 9 with 8) and the two nucleotides 3' of it (compare lane 3 with 2). However, excision of uracil from the 5' terminal or the penultimate position (from the 3' end) was undetectable. Thus, the minimum size substrate required for detectable excision of uracil by BdiUng is pNUNN (Figure 5A).

Genomes with a higher G+C content are inherently prone to forming stable intra-molecular structures such as the hairpin loops due to various topological changes during physiological processes like transcription and replica-
Figure 2. Uracil excision assays with single stranded and double stranded substrates. Assays were carried out at 37°C for 30 min with 1 μg of BdiUng (∼32 pmol monomer) or EcoUng (∼4 pmol) with (+) or without (−) pre-incubation with 1 μg of Ugi (∼100 pmol) as described in Materials and Methods using ∼0.1 pmol of ssU9 (A) or GU9 (B) DNAs, and resolved on 8 M urea PAGE (15%). (C) Electrophoretic mobility shift assay to analyze complex formation of BdiUng with Ugi. After incubation of enzyme with/without Ugi, reaction products were resolved on 15% native PAGE (29:1 crosslinking, pH 6.8). While complex EcoUng-Ugi complex formed is found to migrate adjacent to pure complex control, no band indicative of BdiUng-Ugi complex could be seen. (D) Uracil excision assay on ssU9 with 2 μg of B. diazoefficiens cell free extracts with/without pre-incubation with varying amounts of Ugi. E. coli TG1 cell free extract (2 μg) was used as a control. Reaction products were resolved on 8 M urea PAGE (15%).

The hairpin like structures formed on single stranded regions may lead to the appearance of unpaired C residues highly prone to deamination. In addition, in the loop structures such as the tetraloops, uracils may be fixed into extra-helical (flipped out) positions (52,58). Earlier studies have shown inefficient excision of uracils from the tetraloops by EcoUng (59). However, the uracil excision activity of Ung from Mycobacterium smegmatis, a G+C rich organism (like B. diazoefficiens) was found to be more efficient than that of EcoUng (60). Thus, we checked the uracil excision activity of BdiUng on four hairpin DNA structures that contained uracil at different positions of a tetraloop hairpin (Figure 5B). BdiUng excised uracil from all the loop positions. However, unlike EcoUng, which showed a significant decrease in excision of uracil from the U1-hairpin, U2-hairpin and U3-hairpin substrates when compared with the excision of uracil from ssDNA (compare lanes 5, 8 and 11 with lane 2), BdiUng was not as severely compromised on the same hairpin substrates when compared with its activity on ssDNA (compare lanes 6, 9, 12 with lane 3). Excision of uracil by BdiUng from U4-hairpin was as efficient as its excision of ssDNA (compare lane 15 with 3).

Overall structure of BdiUng

The structure of BdiUng was determined using the single anomalous dispersion (SAD) method. The detailed statistics for the structure are described in Table 2. In agreement with the gel filtration chromatography results (Supplementary Figure S6), the overall structure is a dimer with a typical α/β/α sandwich fold of UDG proteins for each monomer such that a central β-sheet is layered by α-helices on both sides (Figure 6A and B). Using the Dali server, the nearest structural homologue occurred with a family 4 UDG from Sulfolobus tokodaii (sequence identity, 12%, Supplementary Figure S7) with a Z-score of 14.4 and a 1.1 Å rmsd over 31 Cα atoms (61). However, unlike family 4 UDGs, BdiUng lacks the conserved cysteine residues for metal chelation and hence does not contain an iron-sulfur cluster [4Fe–S] in the structure, which is a chara-
Figure 3. Uracil excision assays for product inhibition. Uracil excision assay using ∼0.1 pmol 5’ end 32P-labelled ssU9 substrate with the indicated amounts of EcoUng or BdiUng in the presence of varying concentrations of uracil (i and ii), and ssF9 (a stable AP DNA mimic harbouring tetrahydrofuran at position 9) (iii and iv) were carried out as described in Materials and Methods. Reactions were resolved on 8M urea PAGE (15%) and imaged. The estimated values (%) of substrate (S) to product (P) conversion \( \frac{P}{(S+P)} \times 100 \) are plotted against the concentrations of the inhibitor in the reaction. The data were fitted with the one site-total of saturation binding equations \( Y = \frac{B_{\text{max}} \times X}{K_d + X} + N_S \times X + \text{background} \) (where \( Y \) is specific binding/activity, \( B_{\text{max}} \) is the maximum specific binding/activity in the same units as \( Y \), \( K_d \) is the equilibrium binding constant in the same units as \( X \), \( N_S \) is the slope of nonspecific binding in \( Y \) unit divided by \( X \) unit and background is the amount of nonspecific binding) using GraphPad Prism software. Amounts of product formed in control (without free uracil/AP DNA) reaction in panels i and iii were scaled to 100%, and the remaining activities shown relative to this reference to generate plots in panels ii and iv.

Figure 4. BdiUng shows broad substrate specificity. Excision assays using BdiUng (1 µg, ∼32 pmol monomer) with (A) ssDNA, and (B) dsDNA containing modified bases (∼0.1 pmol each); inosine (I), xanthine (X) and 5-hydroxymethyluracil (HmU) were carried out at 37°C for 30 min as described in Materials and Methods. The reactions were resolved on 8 M urea PAGE (15%) and imaged.
Table 2. Data collection and structure refinement statistics

| Ligand         | BdiUng native | Selenomethionine-labelled | Uracil bound BdiUng | Xanthine bound BdiUng |
|----------------|----------------|--------------------------|---------------------|-----------------------|
| X-ray source   | 7A, PAL        | 7A, PAL                  | Home source         | 5C, PAL               |
| Space group    | P2₁2₁₂₁         | P2₁2₁₂₁                  | C121                | P2₁2₁₂₁               |
| Unit cell dimension |               |                          |                     |                       |
| a, b, c (Å)    | 70.65, 90.03, 255.67 | 70.95, 90.06, 256.26     | 209.53, 89.63,143.78 | 70.02, 89.90, 255.34  |
| α, β, γ (°)    | 90.00, 90.00, 90.00 | 90.00, 90.00, 90.00      | 90.00, 90.00, 90.00 | 90.00, 90.00, 90.00   |
| Resolution (Å) | 1.95            | 2.28                     | 2.86                | 2.80                  |
| Rsym (%)       | 13.2 (56.7)     | 6.5 (60.8)               | 14.0 (33.3)         | 14.0 (52.8)           |
| I/σ(I)         | 13.34 (1.78)    | 21.39 (9.41)             | 09.67 (3.43)        | 15.13 (4.59)          |
| Completeness (%)| 98.71           | 91.73                    | 83.20               | 99.91                 |
| Redundancy     | 40.0 (3.2)      | 7.0 (6.5)                | 2.7 (3)             | 7.0 (6.3)             |
| Refinement     |                |                          |                     |                       |
| Resolution (Å) | 1.95            | 2.28                     | 2.86                | 2.80                  |
| Unique reflection | 117958         | 70019                    | 50734               | 40422                 |
| Rwork/Rfree (%)| 24.98/28.93     | 18.20/21.08              | 25.25/30.21         | 22.47/24.67           |
| Bond lengths (Å)/angles (°) | 0.009/1.30 | 0.010/1.22               | 0.009/1.00          | 0.015/1.00            |
| Average B values (Å²) | 47.40      | 33.20                    | 24.80               | 41.10                 |
| Ramachandran plot (%) |            |                          |                     |                       |
| Favoured region | 96.60          | 96.50                    | 95.00               | 96.50                 |
| Allowed region | 3.40           | 3.50                     | 5.00                | 3.50                  |
| PDB code       | 5GN2           | 5GN3                     | 5GNW                | 5GRK                  |

aNumbers in parentheses are statistics from the highest-resolution shell.

bRsym = Σ|Iobs| - |Iavg| / Σ|Iobs|, where Iobs is the observed intensity of individual reflection and Iavg is the average over symmetry equivalents.

Figure 5. Excision of uracil from different structural contexts. Uracil excision assays with DNA oligomers (~0.1 pmol each) containing a different number of nucleotides on either side of uracil (A) or with the hairpin DNA oligomers containing uracil in different positions in tetraloop (B) were carried out at 37°C as described in Materials and Methods. (A) Reactions were carried out with 1 µg of either protein (~40 pmol EcoUng or ~32 pmol BdiUng monomer) for 30 min. More than one product band is observed in some reactions because incomplete alkaline cleavage of AP site (first reaction leaves a semialdehdye residue on 3’ end of 5’ product and the second reaction removes it, producing a 3’ phosphate on the 5’ product; products are marked with asterisks). (B) Reactions were performed for 30 min with 100 pg (4 fmol) and 500 ng (~16 pmol monomer) of EcoUng and BdiUng, respectively. Reaction products were resolved on 8 M urea PAGE (15%).

DNA is substituted by VGT residues in BdiUng and positioned in a groove that appears to bind DNA. Residues 90–94, YVMVN, constitute the fourth strand of the core β-sheet corresponding to VLLLN motif in family 1 whereas GS motif involved in ‘holding’ DNA substrate is substituted by GG residues in BdiUng. Interestingly, the motif B of HPSPLS residues, commonly called the Leu-intercalation loop, shows a significantly different shape and composition in BdiUng. This motif is substituted by residues 168–178 in BdiUng and forms an irregularly extended helical geometry that connects further to a long helix α6 at the C-terminal region. Due to the significantly extended protruding architecture of this region (Figure 7B(ii)), the uracil binding pocket is located distant from the tip of this region and no residues previously known for intercalation function such as leucine, arginine or tyrosine were identified in this motif. The higher B-factor of this region, 33.1 compared to 25.3, suggests the flexible characteristic of this region. This unique architecture is not observed in any other UDG family known to date and could be a signature motif in this family of BdiUng. The leucine intercalation loop in family 1 UDGs provides the major binding surface for the inhibitor protein Ugi, which enables docking to the minor-groove in DNA. The altered geometry and surface potential of this region in BdiUng also explains the absence of the binding affinity to the Ugi molecule (Supplementary Figure S10).

Active site

The uracil binding pocket in the active site is lined with residues such as Q56, P58, A59, E62, L69, G71, A73, M92 and N94 (Figure 8A, Supplementary Figure S12). Residues surrounding the O2 and O4 of uracil moieties are conserved similarly to family 1 UDG members. Essential residues for
catalysis in family 1 UDGs such as H187 and D64 are also conserved in BdiUng (represented by H168 and D57), suggesting a catalytic mechanism for uracil cleavage similar to family 1 UDGs. The invariant aromatic residue that provides stable \( \pi-\pi \) stacking interaction with uracil ring in most UDGs, such as F62 in Ung, is absent and replaced by M92. Interestingly, the region surrounding uracil position 5 is differently shaped with a significantly larger cavity. The presence of A59 instead of a bulky residue (Y66 in EcoUng) in motif A provides sufficient cavity space in the active site. Analysis of the uracil binding pocket between the apo and the uracil bound form reveals no significant conformational changes in the cavity region indicating that a structural change may not be required for the recognition of the uracil base by BdiUng. Given that the L191 of EcoUng forms stable interactions with residues in the hydrophobic pocket of Ugi, the protruding loop of BdiUng, mainly constituted of polar residues such as E173, S174, S175, H177, D178, and S179, would disrupt any stable interaction with hydrophobic pocket of Ugi (Supplementary Figure S10). Similarly, a dimeric Ung protein from vaccinia virus, that has been reported to be resistant to Ugi inhibition, contains polar Arg at the position corresponding to L191 of EcoUng affecting the stable hydrophobic interactions with Ugi (Supplementary Figure S11) (62).

**Mutational analysis of BdiUng**

In spite of the very low homology with family 1 UDGs, the crystal structure of UDGs revealed a conservation of most of the key residues implicated in catalysis by family 1 UDGs.
UDGs (Figure 8A ii). The presence of Y66 in EcoUng allows uracil (with no bulky substituents at position 5 of the pyrimidine ring) but not the 5-hydroxymethyluracil (with a bulky substituent at position 5 of the pyrimidine ring), or the xanthine (a purine) in the active site pocket. Thus, to test the importance of A59 in BdiUng in conferring broad substrate specificity of excising uracil, xanthine, and 5-hydroxymethyluracil, we generated the A59Y mutant and purified the mutant protein from the TG1 Δung::cm strain (Supplementary Figure S4B). Interestingly, the A59Y mutation led to a decrease in its activity on xanthine, and 5-hydroxymethyluracil, while the wild-type and promoter architecture efficiently (Figure 10). However, under the same conditions, BdiUng did not significantly change Lac− to Lac+ reversion. This observation suggests that BdiUng does not substitute for the deficiency of EcoUng in E. coli in reducing C to T mutations.

**DISCUSSION**

Uracil DNA glycosylases are essential to prevent incorporation of uracil in DNA. The high G+C content in the root nodule bacterium B. diazoefficiens could predispose this organism to the accumulation of uracils in DNA, especially in cells exposed to reactive oxygen and nitrogen species collectively referred to as ROS/RNS. Both ROS (31–41) and RNS species (such as NO and ONOO−) (42,43) are produced in nitrogen fixing nodule environments. B. diazoefficiens contains five different UDGs (four belonging to families 2, 4 and 5) with a distinct absence of the classical family 1 UDG, which were thought to be ubiquitously present in all organisms since they provide one of the most efficient DNA repair mechanisms.

Interestingly, in the case of *Drosophila* also, while the earlier studies have shown the presence of UDG activity (63), the genome sequencing has now revealed lack of the classical family 1 UDG (UNG) protein (64). Thus, the presence of weak UDG activity in these organisms may be attributed to the other members of the UDG family (64). In fact, re-
Figure 8. Active site of BdiUng. (A) (i) The active site of BdiUng is shown along with uracil depicted as spheres. Motif A and H168 are highlighted. (ii) The ligand binding cavity of BdiUng and E. coli Ung (PDB Id: 1EUI) show high similarity. Overlap of BdiUng (grey) and E. coli Ung (blue) active site. The residues are labelled for BdiUng. (B) Model structures for mutation A59Y (i) BdiUng active site showing the A59Y mutation. (ii) Surface diagram of BdiUng showing the steric hindrance to substituents at C5 of uracil molecule caused by the A59Y mutation.

Figure 9. Activities of BdiUng A59Y mutant. The excision assays were carried out with oligomers (~0.1 pmol each) containing different lesions (uracil in ssU9, xanthine in ssX9 and 5-hydroxymethyluracil in HmU) using BdiUng and BdiUng A59Y mutant (each 1 µg, ~32 pmol monomer) at 37°C for 30 min as described in Materials and Methods. Products were resolved on 8 M urea PAGE (15%).

Figure 10. Mutation frequency (Lac+ reversion) of E. coli CC102 strain and its Δung::kan derivatives harboring plasmid pTrc99c or its derivatives containing EcoUng or BdiUng. Mutation frequency values are represented as mean ± SD from 10 independent colonies (replicates), calculated as the ratio of a number of colonies that appeared on minimal lactose plates compared to minimal glucose plates (per 1 ml culture). In Δung::kan strain, provision of BdiUng did not significantly change the C to T mutation frequency (comparing 3 and 5, P value = 0.21).

cent findings have shown that the lack of the highly efficient family 1 UDG (UNG) in Drosophila and the Holometabola in general, could be for an important physiological purpose (65). Developmental patterns in these organisms show a strong correlation between the presence of uracil in their genomes and the lowered level of expression of dUTPase, an enzyme that hydrolyses dUTP to dUMP to avoid its incorporation in DNA. Thus, the lack of family 1 UDG (UNG)
activity in these organisms is physiologically relevant in allowing occurrence of uracil in DNA (66). Whether the lack of highly efficient family 1 UDG in *B. diazoefficiens* contributes to its specialized physiology during nodulation or otherwise is not known.

To study the role of UDGs in *B. diazoefficiens*, we have identified and characterized a novel uracil DNA glycosylase from *B. diazoefficiens* (Q89XRO_BRAJA, referred to as *Bdi*Ung, encoded by bll248) containing a motif (QGDPA) similar to the active site motif A (GQDPY) of family 1 UDGs. *Bdi*Ung was found to be active on both ssDNA and dsDNA containing uracil with pNUNN as the minimum substrate required for detectable activity. *Bdi*Ung did not interact with Ugi and was therefore resistant to Ugi mediated inhibition in uracil excision. Further, unlike family 1 UDGs, the *Bdi*Ung showed broader substrate specificity and excised 5-hydroxymethyl uracil and xanthine present in ssDNA. The co-crystal structures of *Bdi*Ung with uracil, and xanthine provide a rationale for the relaxed substrate specificity (Supplementary Figure S12). While *Bdi*Ung was inhibited by free uracil, AP DNA (ssF9) did not inhibit its activity to any significant level even at 200 μM concentration (Figure 3).

Substrate specificity of *Eco*Ung is due to the well-tailored active site pocket made to specifically fit uracil (13). The efficient recognition of uracil is proposed to occur through a ‘pinch, push, plug, and pull’ mechanism (68–70). Initial backbone phosphate contacts pinch the DNA backbone. Then a conserved Leu (L191 in *Eco*Ung) acts as a wedge/plug to retain the flipped out uracil into the active site of the enzyme (67,68). Many specific hydrogen bonds and stacking interactions stabilize the uracil in extra-helical conformation within the active site (pull) (70). The side chain of Tyr in the GQDPA motif is in van der Waals’ interaction with C5 of uracil, thus blocking thymine (with a methyl group on C5) and bulky purine rings (11,13,71).

While *Bdi*Ung exhibited no significant sequence similarity with any of the established families of UDGs, the crystal structure of the protein revealed that the overall fold is similar to family 4 UDGs in the absence of the characteristic 4Fe-S cluster and that most of the key amino-acid residues implicated in *Eco*Ung catalysis of family 1 UDG are present in the active site. Structural alignment of *Bdi*Ung with *Eco*Ung showed that the GQDPA motif of *Bdi*Ung was located in a position analogous to motif A of *Eco*Ung which is involved in activation of a water molecule that performs a nucleophilic attack on C1’ of ribose during catalysis. *Bdi*Ung also contains Asn and His residues similar to those in *Eco*Ung, which form specific contacts with a uracil residue in the active site. In spite of the drastically different architecture of the active site periphery compared to *Eco*Ung, the similarity observed with respect to the key active site residues implies *Bdi*Ung catalysis to be similar to that of *Eco*Ung. But the substitution of the classical Leu intercalation loop by the extended protruding architecture with no apparent functional residue similar to L191 of *Eco*Ung (implicated in pushing the uracil/plugging the duplex void) could be the reason for its reduced activity on dsDNA as well as the absence of its complex formation with Ugi (68,69).

A conserved Tyr in the motif A of Ung proteins prevents entry of bulky bases and position 5- substituted uracil into the active site of the family 1, Ung proteins. In *Bdi*Ung the Tyr residue of motif A is substituted by Ala explaining its broad substrate specificity. Consistent with this observation, *Bdi*Ung A59Y mutant revealed a much decreased activity on 5-hydroxymethyl uracil and xanthine containing ssDNAs (Figure 9). Still, the active site seems carefully designed to exclude any of the normal bases. Most interestingly, while all the UDGs studied so far show a conserved aromatic residue (Phe) in the active site that provides stability to the flipped out uracil in the active site, no such supporting aromatic residue was seen in the active site of *Bdi*Ung.

As revealed by the crystal structure and biochemical analyses, *Bdi*Ung is a dimeric protein. UNG from vaccinia virus was also characterized as a dimer (72). However, the dimeric interfaces of the two enzymes are significantly different, in that the *Bdi*Ung dimerizes primarily by the C-terminal extended hairpin strand with each active site of the monomer, facing the same side of the dimer, whereas the vaccinia virus UNG dimerizes through the main α/β/α sandwich fold with two active sites positioned opposite (Supplementary Figure S13). The functional significance of the dimerization is not clear at this stage and may need further investigation.

From the analysis of the impact of *Bdi*Ung in the mutation frequency, using *ung* strains of *E. coli* CC102 (monitoring Lac⁺ reversion frequency), we found that *Bdi*Ung cannot carry out a physiological function in rescuing C to T mutations similar to that of *Eco*Ung in *E. coli*. We attribute this to the poor efficiency of uracil excision from dsDNA and also to the fact that these studies were carried out in a heterologous system with a relatively much higher growth rate. Besides, the much higher efficiency with which *Bdi*Ung acts on ssDNA compared to dsDNA and the higher relative efficiencies of uracil excision from loop regions of DNA tells us that it is a single strand specific UDG. This could be further supported by the surface analysis of *Bdi*Ung. The unique protruding loop yields a narrow DNA binding groove at the uracil binding pocket, a suitable architecture for ssDNA substrate interaction (Supplementary Figure S9).

Interestingly, most organisms with homologues of *Bdi*Ung possessed genomes with high G+C contents (Supplementary Table S3). Organisms having G+C rich genomes are intrinsically prone to accumulating hairpin-loops in their genome. The relatively less compromising efficiency of *Bdi*Ung on uracil found in loop regions of DNA (as compared to ssU9), along with the presence of its homologues in high G+C containing organisms might be pointing to a physiological function of this family of UDGs.

In addition to *Bdi*Ung, we have identified and characterized three other UDGs from *B. diazoefficiens*. Besides a family 2 homologue (Bll6661) and a family 5 homologue (Bll5068), the organism also contains a homologue of the UdgX protein, which we recently identified as a novel uracil–DNA binding protein in *M. smegmatis* (56). Bll5068 was found to efficiently excise uracil from ssDNA and dsDNA. Further, Bll2523 was identified as a gene coding for a putative family 4 homologue. The unusually high number of
UDGs and the presence of novel UDGs like *BdiUng* might compensate for the lack of a family 1 UDG homologue in this slow growing organism.

**SUPPLEMENTARY DATA**

Supplementary Data are available at NAR Online.

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