Diagnostic accuracy of adenosine deaminase for tuberculous peritonitis: a meta-analysis

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Abstract

Introduction: Tuberculous peritonitis remains a diagnostic challenge for clinicians. Many studies have investigated the usefulness of adenosine deaminase (ADA) in ascites for the diagnosis of tuberculous peritonitis; however, the overall diagnostic accuracy of ADA for tuberculous peritonitis remains unclear. The aim of the present meta-analysis was to determine the overall accuracy of ADA measurements in the diagnosis of tuberculous peritonitis.

Material and methods: We performed a systematic search in PubMed and Embase to identify published studies that evaluated the diagnostic role of ADA for tuberculous peritonitis. Quality was assessed according to standardized Quality Assessment of Diagnostic Accuracy Studies criteria. Sensitivity, specificity and other measures of accuracy of ADA assay in order to diagnose tuberculous peritonitis were pooled using random effects models. Summary receiver operating characteristic curve (SROC) was used to summarize overall test performance.

Results: Sixteen studies met inclusion criteria for the present meta-analysis. The pooled sensitivity and specificity for diagnosing tuberculous peritonitis were 0.93 (95% CI: 0.89–0.95) and 0.96 (95% CI: 0.94–0.97), respectively. The positive likelihood ratio was 15.80 (95% CI: 10.87–22.95), negative likelihood ratio was 0.09 (95% CI: 0.05–0.16) and diagnostic odds ratio was 249.28 (95% CI: 113.11–549.39). The area under the SROC was 0.98.

Conclusions: Ascitic ADA determination is a relatively sensitive and specific test for the diagnosis of tuberculous peritonitis. Measurement of ADA in ascites is thus likely to be a useful diagnostic method for tuberculous peritonitis.

Key words: tuberculous peritonitis, adenosine deaminase, meta-analysis.
ods utilized, and acid-fast stained smears are disappointingly insensitive [5, 6]. Caseous granulomas of peritoneal biopsies obtained by invasive laparoscopy or laparotomy are helpful for rapid primitive diagnosis, but the procedures may not be available in all level hospitals and well tolerated, and they may increase rates of morbidity and mortality [7, 8]. The high mortality rate in untreated patients warrants a quick and noninvasive test for screening TBP.

Adenosine deaminase (ADA) is a purine-degrading enzyme that catalyzes the deamination of adenosine in an irreversible manner, which results in the production of inosine. Adenosine deaminase levels in body fluids can be measured rapidly, and they might provide an alternative for the diagnosis of TB [9, 10]. Several studies reported the use of ADA in the diagnosis of TB in other fluids including meningeal, pleural, and pericardial effusions, suggesting that increasing ADA activity relates to the intensity of stimulation and the maturation state of the lymphocyte, due to the immune cellular response against Mycobacterium tuberculosis [10–12].

In fact, quite a lot of studies have investigated the diagnostic role of ascitic ADA for TBP. Considering the controversy about the current role of ADA as a diagnostic tool for TBP, the present meta-analysis aims to determine the overall diagnostic accuracy of ADA for TBP.

Material and methods

The present meta-analysis was performed according to the guidelines of the preferred reporting items for systematic reviews and meta-analysis (PRISMA) statement and with methods recommended by the Cochrane Diagnostic Test Accuracy Working Group [13, 14].

Study identification

To identify studies that evaluated the diagnostic accuracy of ADA for TBP, two independent reviewers performed a search of PubMed (Medline) and Embase up to October, 2012. The search key words included “ADA or adenosine deaminase”, “tuberculosis or tuberculous”, and “peritonitis”. In addition, we obtained additional articles by citation tracking of review articles and original articles.

Study selection

We set the inclusion criteria as follows: measurement of ascitic ADA in human subjects; detailed diagnostic criteria for TBP; studies provided both the sensitivity and specificity of ADA assay; at least 20 participants (10 patients and 10 controls).

Exclusion criteria: no control group; limited participants; non-English publications; publications with limited information to calculate sensitivity and specificity of ADA.

The articles that were finally included in the meta-analysis were reviewed independently by two different reviewers and discrepancies in the interpretation were resolved by consensus.

Data extraction

The final set of articles was assessed independently by two reviewers, who were blinded to the article details, and the differences between them were solved by consensus. The following data from each publication were retrieved: author, publication year, participants, gold standard for TBP diagnosis, ADA assay method, sensitivity and specificity data, methodological quality, study design. If no data on the above information were presented in the primary studies, we marked it with "Not Available, NA”.

Assessment of study quality

To assess trial methodology, included publications were reviewed independently by two authors and given a quality score by using the QUADAS (quality assessment for studies of diagnostic accuracy, an evidence-based quality assessment tool to be used in systematic reviews of diagnostic accuracy studies, maximum score 14) tools [15].

Data synthesis and statistical analysis

The standard methods recommended for diagnostic accuracy meta-analyses were used in the present study [16]. The following indexes of test accuracy were computed for each study: sensitivity, specificity, positive likelihood ratio (PLR), negative likelihood ratio (NLR), and diagnostic odds ratio (DOR). The analysis was based on a summary receiver operating characteristic (SROC) curve [17]. Heterogeneity was evaluated by using the $\chi^2$ test and $I^2$ test. The random effects model was performed to synthesize data when heterogeneity was present ($p < 0.05$ and $I^2 > 50$); otherwise the fixed effects model was used. Since publication bias is of concern for meta-analyses of diagnostic studies, we tested for the potential presence of this bias using Deeks’ funnel plots [18].

All analyses were performed using two statistical software programs (Meta-DiSc for Windows; XI Cochrane Colloquium, Barcelona, Spain and Stata, version 12; Stata Corporation, College Station, TX, USA). All statistical tests were two-sided, and significance was set at $p < 0.05$.

Results

After independent review, sixteen studies in fifteen publications with 1574 subjects on the use of ADA in patients with ascites were considered eligi-
ble for inclusion in the present meta-analysis [19–33]. The major reasons for excluding other studies were as follows: non-diagnostic studies or studies cannot reconstruct the diagnostic 2 by 2 table; limited samples, or mixed with other serous effusions.

**Study characteristics and quality report**

Of the included studies, the average sample size in the 16 studies was 49 (41–368). For most studies, the diagnosis of TBP was based on bacteriological or histological examinations or both, four studies included some patients who were diagnosed with TBP based on clinical diagnosis, including clinical presentation, pleural fluid analysis, radiology and the responsiveness to anti-tuberculosis chemotherapy [20, 22, 29, 32]. For ADA assay method, the Giusti method was applied in 11 studies and non-Giusti methods were used in 5 studies. The cut-off value was ≥ 30 IU/l in 14 studies (30–40 IU/l), and < 30 IU/l in 2 studies; one was 21 IU/l [19], and the other one was 7 IU/l [26]. The quality of the sixteen studies was generally high with ten studies having QUADAS scores ≥ 10. The clinical summary of these studies, along with the QUADAS scores, are outlined in Table I.

**Diagnostic accuracy for TBP**

Heterogeneity examination is performed to choose the appropriate calculation model; the heterogeneity analysis showed $I^2$ of 54.1% for sensitivity and 59.2% for specificity, suggesting significant heterogeneity among included studies; thus the random effects model approach was selected for the present meta-analysis. The forest plots of the sensitivity and specificity for ADA assays in diagnosing TBP are shown in Figures 1 and 2, respectively. The pooled sensitivity was 0.93 (95% CI: 0.89–0.95), specificity was 0.96 (95% CI: 0.94–0.97), The PLR was 15.80 (95% CI: 10.87–22.95), the NLR was 0.09 (95% CI: 0.05–0.16) and the DOR was 249.28 (95% CI: 113.11–549.39).

Figure 3 shows the SROC plotting the true-positive against the false-positive rates of individual studies. The area under the curve (AUC) was 0.98, indicating that the level of overall accuracy was high.

We conducted a sub-group analysis, for the 11 studies that determined ascitic ADA with the Giusti method. The pooled sensitivity was 0.96 (95% CI: 0.93–0.98), specificity was 0.96 (95% CI: 0.94–0.97), PLR was 13.82 (95% CI: 12.58–26.68), NLR was 0.07 (95% CI: 0.04–0.11) and the DOR was 509.44 (95% CI: 227.66–1139.95). The AUC was 0.99. Thus, the Giusti method may be a suitable method for determination of ascitic ADA.

**Publication bias**

Deeks’ funnel plot asymmetry test was used to evaluate potential publication bias. The statistical-

Discussion

Tuberculous peritonitis is still a public health problem in endemic regions of the world. It may be fatal but is medically cured if diagnosed in a timely fashion. To make an early and accurate diagnosis is of great importance for its prognosis. Adenosine deaminase is a well-known diagnostic marker for tuberculosis. In fact, Riquelme et al. conducted a meta-analysis to analyze the diagnostic role ADA for TBP. According to his inclusion criteria, only four publications were included [34]. Several years have passed, and some new studies have been added, so we conducted this updated meta-analysis. According to our inclusion criteria, we included the most recent published studies, and included studies using a non-Giusti method. The number of included studies provides enough evidence to support the diagnostic power of ADA for TBP. We found a summary AUC of 0.98, a summary estimate of 0.93 for sensitivity and 0.96 for specificity. It seems that ADA assay plays a valuable role in the diagnosis of TBP. Both Riquelme’s and our studies support the proposition that ADA determination is a discriminating test for diagnosing TBP [34].

The SROC curve has been recommended to represent the overall performance of a diagnostic study, which shows the trade-off between sensitivity and specificity, based on data from a meta-analysis [35]. The AUC and index Q(•) are recognized as potentially useful summaries of the curve. Q-value, the intersection point of the SROC curve with a diagonal line from the left upper corner to the right lower corner of the ROC space, which corresponds to the highest common value of sensitivity and specificity for the test, represents an overall measure of the discriminatory power of a test. In the present study, the Q value was 0.94, and the maximum joint sensitivity and specificity of ADA for TBP was 0.94. The AUC also represents the overall accuracy of the diagnostic study. The AUC represents an analytical summary of test performance and displays the trade-off between specificity and sensitivity. If the AUC is 1, it means the ADA test differentiates perfectly between TBP and non-TBP patients. An AUC of greater than 0.9 indicates high diagnostic accuracy. In the present meta-analysis, the AUC was 0.98, suggesting that the level of overall accuracy of ascitic ADA for TBP is high.

Apart from SROC, we also examined other diagnostic indexes. DOR is a single indicator of diagnostic accuracy that combines the data from sensitivity and specificity into a single number [36]. It is defined as the ratio of the odds of positive test results in the diseased relative to the odds of pos-
### Table 1: Clinical summary of included studies

| Author                  | Year | Sample size | Gold standard | ADA assay method | Cut-off value [IU/l] | TP | FP | FN | TN | QUADAS | Study design |
|-------------------------|------|-------------|---------------|------------------|---------------------|----|----|----|----|--------|--------------|
| Kang et al.             | 2012 | 27          | 25            | B + HP           | NA                  | 21 | 25 | 4  | 2  | 21     | 12 R         |
| Saleh et al.            | 2012 | 14          | 27            | B + CD           | Giusti 35           | 14 | 2  | 0  | 25 | 11     | P            |
| Hong et al.             | 2011 | 35          | 17            | B + HP           | NA                  | 30 | 31 | 3  | 4  | 14     | 8 R          |
| Bandopadhyay et al.     | 2006 | 36          | 60            | B + HP + CD      | Giusti 33           | 32 | 0  | 4  | 60 | 10     | P            |
| Sharma et al.           | 2006 | 31          | 88            | B + HP           | Giusti 37           | 30 | 5  | 1  | 83 | 11     | P            |
| Burgess et al.          | 2001 | 18          | 160           | B + HP           | Giusti 30           | 17 | 13 | 1  | 147| 10     | P            |
| Sathar et al.           | 1999 | 23          | 22            | HP               | Kinetic enzyme-coupled assay 30 | 22 | 0  | 1  | 22 | 9      | P            |
| Sathar et al.           | 1996 | 17          | 351           | B + HP           | NA                  | 7  | 10 | 16 | 7  | 335    | 10 R         |
| Sathar et al.           | 1995 | 28          | 53            | B + HP           | Giusti 30           | 26 | 2  | 2  | 51 | 9      | P            |
| Fernandez-Rodriguez et al. | 1991 | 12          | 96            | B + HP           | Slaats 32           | 10 | 0  | 2  | 96 | 10     | P            |
| Riera et al.            | 1991 | 16          | 70            | B + HP + CD      | Giusti 40           | 16 | 2  | 0  | 68 | 10     | P            |
| Bhargava et al.         | 1990 | 17          | 70            | HP               | Giusti 36           | 17 | 2  | 0  | 68 | 9      | P            |
| Dwivedi et al.          | 1990 | 19          | 30            | B + HP           | Giusti 33           | 19 | 1  | 0  | 29 | 9      | P            |
| Voigt et al. (1)        | 1989 | 41          | 41            | B                | Giusti 32.3         | 39 | 1  | 2  | 40 | 11     | R            |
| Voigt et al. (2)        | 1989 | 11          | 53            | B + HP + CD      | Giusti 32.3         | 11 | 2  | 0  | 51 | 11     | P            |
| Martinez-Vazquez et al. | 1986 | 10          | 56            | B + HP           | Giusti 35           | 10 | 0  | 0  | 56 | 7      | R            |

**TBP** – tuberculous peritonitis, **B** – bacteriology, **HP** – histopathology, **CD** – clinical diagnosis, **NA** – not available, **TP** – true positive, **FP** – false positive, **FN** – false negative, **TN** – true negative, **QUADAS** – quality assessment for studies of diagnostic accuracy, **P** – prospective, **R** – retrospective
D iagnostic accuracy of adenosine deaminase for tuberculous peritonitis: a meta-analysis

Positive test results in the non-diseased. The value of DOR ranges from 0 to infinity, with higher values indicating better discriminatory test performance. In the present meta-analysis, the pooled DOR was 249.28, suggesting that ADA assays seemed to be useful in the diagnosis of TBP. Since the SROC curve and DOR are not easy to interpret and use in clinical practice, likelihood ratios are considered more clinically meaningful [37]. The PLR was 15.80, indicating that patients with TBP have about 16-fold higher chance of being ADA assay-positive compared with non-TBP subjects. The NLR was 0.09; it means that if the ADA assay result was negative, the probability that this subject has TBP is only 9%, which is low enough to exclude TBP.

Our meta-analysis suggests that ADA determination plays a valuable role in diagnosing TBP. The reported sensitivities varied among studies; only two studies had sensitivity less than 0.85, and the pooled sensitivity was 0.93. Ascitic ADA assay is suitable as a routine screening tool for TBP. Physicians may argue that ADA assay results may be affected by liver cirrhosis. Liao et al confirmed that even with lower ascites ADA activity in patients

Figure 1. Forest plots of pooled sensitivity of ADA for the diagnosis of TBP. The point estimates of sensitivity from each study are shown as solid circles. Error bars indicate 95% confidence intervals

Figure 2. Forest plots of pooled specificity of ADA for the diagnosis of TBP. The point estimates of specificity from each study are shown as solid circles. Error bars indicate 95% confidence intervals
with liver cirrhosis, ascites ADA levels could be significantly elevated resulting from strong immune responses when cirrhotic patients suffer from TBP [38]. Considering its high sensitivity and specificity, ascitic ADA may be useful in the differential diagnosis of TBP. In patients with underlying cirrhosis, concomitant cirrhosis should not limit its clinical utility [38]. Although peritoneal biopsies obtained by laparoscopy or laparotomy are valuable for rapid diagnosis of TBP, these invasive procedures may not be available in all hospitals and increase mortality [7,8]. Thus, the importance of the ADA test is that it not only provides high diagnostic accuracy, but also guides the inclusion of patients who might benefit from further invasive procedures.

Based on the evidence compiled in this meta-analysis, ascitic ADA measurement plays a critical role in the diagnosis of TBP. It is likely to be a useful diagnostic tool for TBP. In addition, the results of ascitic ADA assays should be interpreted in parallel with clinical findings and the results of traditional tests such as microbiologic examination and peritoneal biopsy. It should be noted that there are currently no ascitic markers (including ADA) which are specific for TBP. Further studies aim to investigate the diagnostic performance of other ascitic markers, such as interferon-γ [23], or combined diagnostic accuracy of different ascitic markers should be performed.

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Diagnostic accuracy of adenosine deaminase for tuberculous peritonitis: a meta-analysis

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