Effects of perioperative dexmedetomidine infusion on renal function and microcirculation in kidney transplant recipients: a randomised controlled trial

Yin-Chin Wangª, Ming-Jiuh Wangª, Chih-Yuan Leeb, Chien-Chia Chenb, Ching-Tang Chiua, Anne Chaoa, Wing-Sum Chanª, Meng-Kun Tsaib,d and Yu-Chang Yehª

ªDepartment of Anesthesiology, National Taiwan University Hospital, Taipei, Taiwan; bDepartment of Surgery, National Taiwan University Hospital, Taipei, Taiwan; cDepartment of Anesthesiology, Far Eastern Memorial Hospital, New Taipei, Taiwan; dDepartment of Surgery, National Taiwan University Hospital, Hsin-Chu Branch, Hsinchu City, Taiwan

ABSTRACT

Objective: Ischemia-reperfusion injury affects postoperative transplanted kidney function in kidney transplant recipients. Dexmedetomidine was reported to attenuate ischemia-reperfusion injury and improve microcirculation, but its propensity to cause bradycardia and hypotension may adversely affect microcirculation. This study investigated the effect of dexmedetomidine on postoperative renal function and sublingual microcirculation in kidney recipients.

Methods: The enrolled kidney transplant recipients were randomly allocated to the control group or dexmedetomidine group. After anaesthesia induction, patients in the dexmedetomidine group received dexmedetomidine infusion until 2 h after surgery. Sublingual microcirculation was recorded using an incident dark-field video microscope and analysed. The primary outcomes were the creatinine level on a postoperative day 2 and total vessel density at 2 h after surgery.

Results: A total of 60 kidney recipients were analysed, and the creatinine levels on postoperative day 2 were significantly lower in the dexmedetomidine group than in the control group (1.5 (1.1–2.4) vs. 2.2 (1.7–3.0) mg/dL, median difference 0.6 (95% CI, –0.7 to –0.5) mg/dL, p = .018). On a postoperative day 7, the creatinine levels did not differ significantly between the two groups. Total vessel density at 2 h after surgery did not differ significantly between the two groups.

Conclusion: We found that early postoperative renal function was better in kidney transplant recipients receiving dexmedetomidine infusion, but total vessel density was not significantly different between the intervention and control groups.

KEY MESSAGES

- Ischemia-reperfusion injury affects postoperative transplanted kidney function, and dexmedetomidine was reported to attenuate ischemia-reperfusion injury and improve microcirculation in other clinical conditions.
- This study showed that early postoperative renal function was better in kidney transplant recipients receiving dexmedetomidine.
- Dexmedetomidine’s side effect of bradycardia and hypotension may affect microcirculation, our results revealed that the perioperative sublingual microcirculation did not differ significantly in kidney transplant recipients receiving dexmedetomidine.

Introduction

End-stage kidney disease remains a global health concern; patients undergoing dialysis experience a lower quality of life and suffer increased morbidity and mortality. Kidney transplantation is the definitive treatment for patients on dialysis. However, ischemia-reperfusion injury to the transplanted kidneys may affect their postoperative function after kidney transplantation [1,2]. Moreover, renal microcirculation is a key issue related to acute and chronic kidney diseases.
Renal microvascular dysfunction includes alterations in endothelial barrier permeability, exaggerated inflammation, and impairment of endothelium-dependent vasorelaxation [3]. Our previous study revealed the alteration of sublingual microcirculatory dysfunction in patients on dialysis, and the severity of this alteration was lower in patients who receive a kidney transplant [4]. Surgical stress may affect the pre-existing microcirculatory dysfunction of patients undergoing kidney transplantation.

Dexmedetomidine is a highly selective alpha-2 agonist with sedative and analgesic effects [5]. It modulates inflammation by enhancing parasympathetic tone while reducing sympathetic tone [6,7]. Dexmedetomidine has been reported to confer renal protection effects in patients undergoing coronary artery bypass surgery [8,9]. The protective effects of dexmedetomidine against ischemia-reperfusion injury have been described in many studies [10–12]. Moreover, the sympatholysis effect of dexmedetomidine induced vasodilation [5], and our previous animal study showed that dexmedetomidine prevented alterations of intestinal microcirculation in rats with surgical stress and pain [13]. However, the most common side effects of dexmedetomidine are bradycardia and hypotension. Low cardiac output and low perfusion pressure may deteriorate microcirculation [14,15]. We hypothesised that dexmedetomidine could attenuate the ischemia-reperfusion injuries and preserve transplanted kidneys’ function. In addition, the issue that the effects of dexmedetomidine on perioperative microcirculation were protective or detrimental remained unknown. Thus, this study investigated postoperative renal function and perioperative sublingual microcirculation in patients undergoing kidney transplantation.

Methods

Patients

This prospective, randomised, controlled, single-blinded, open-label study was approved by the Research Ethics Committee of National Taiwan University Hospital, Taipei, Taiwan (Ethical Committee number 201512039MINB). This study was registered on the ClinicalTrials.gov protocol registration system (ID: NCT02707809). Patients undergoing kidney transplants were evaluated for eligibility. We excluded patients younger than 20 years or older than 70 years, and those with an allergy to dexmedetomidine, refractory bradycardia (heart rate below 60 beats per minute after treatment), and severe atrioventricular block (Mobitz type II and III). Written informed consent was obtained from all participants. The study was conducted and reported in accordance with the CONSORT recommendations [16]. The histidine-tryptophan-ketoglutarate solution was used for flushing and perfusion of the donor kidney, and the donor’s kidney was kept in static cold storage before transplant surgery.

Anaesthesia protocol and dexmedetomidine infusion protocol

The patients were allocated to a control group or dexmedetomidine group through randomised allocation using sealed opaque envelopes. The patients underwent sublingual microcirculation measurement before the induction of anaesthesia (T1). From before anaesthesia induction until 2 h after surgery, a non-invasive cardiac output monitoring system (NICOM, Cheetah Medical, Newton Centre, MA, USA) was used for measuring cardiac index, stroke volume index, and stroke volume variation. General anaesthesia was induced with fentanyl 1.5–2.0 µg/kg, propofol 1.5–3.0 mg/kg, glycopyrrolate 0.2 mg and cisatracurium 0.15–0.20 mg/kg and maintained with desflurane at an end-tidal concentration 4.2%–7.2%. Ventilator settings were as follows: fraction of inspired oxygen 40%; tidal volume 6–8 mL/kg; positive end-expiratory pressure level, 5 cm H2O; and end-tidal carbon dioxide partial pressure 35–40 mm Hg.

After anaesthesia induction, patients in the dexmedetomidine group received dexmedetomidine infusion until 2 h after surgery. To ensure patient safety, the titration rate of dexmedetomidine infusion was separately determined for each patient, and the infusion rate was adjusted in a range of 0.1–0.7 µg kg⁻¹ h⁻¹ according to the individual patient’s responses in terms of cardiac index, blood pressure, and heart rate. Fluid supplementation (in the range of 30–40 mL kg⁻¹ of predicted body weight), ephedrine, and norepinephrine were used to maintain appropriate mean arterial pressure (MAP 70–100 mm Hg), heart rate (>60 beats per minute), cardiac index (>2.5 L/min/m²), and stroke volume variation (<13%). Patients undergoing ABO-incompatible kidney transplantation received 6 units of fresh frozen plasma during the operation. Patients in the control group received conventional management with fluid supplementation (30–40 mL/kg of predicted body weight), ephedrine, and norepinephrine to meet the same hemodynamic goal. During operation, the patients received methylprednisolone (10 mg/kg) for immunosuppression. At the end of the surgery, intravenous morphine 0.1–0.2 mg/kg was
administered for postoperative analgesia. Extubation was performed in the operation room, and the patients were transferred to the post-anaesthesia care unit. After arrival at the post-anaesthesia care unit, the dexmedetomidine infusion rate was reduced by one-quarter of the initial rate every 30 min and discontinued after 2 h. After blood sample and microcirculation examinations were conducted, the patients were discharged from the post-anaesthesia care unit. In the general ward, postoperative creatinine level was measured once daily in the early morning during the follow-up period.

Preoperative desensitisation and immunosuppressive therapy for ABO-incompatible kidney transplantation and postoperative immunosuppressive therapy

For ABO-incompatible kidney transplantation, preoperative desensitisation with rituximab and double filtration plasmapheresis and preoperative immunosuppressive therapy with tacrolimus, mycophenolate mofetil, and methylprednisolone were performed according to our published protocol [17]. Postoperative immunosuppressive therapy with tacrolimus, mycophenolate mofetil, and methylprednisolone was administered according to the post kidney transplant care protocol [17].

Microcirculation examinations

Sublingual microcirculation images were captured using an incident dark-field video microscope (CytoCam, Braedius Medical, Huizen, the Netherlands). At each time point, six video sequences (length: 6 s each) were recorded at different sublingual sites, and three sequences with appropriate image quality were selected for analysis by a single observer who was blinded to the grouping. The analysis was performed using the semi-automated analysis software package Automated Vascular Analysis 3.0 (AVA, Academic Medical Centre, University of Amsterdam, Amsterdam, the Netherlands). Total vessel density (TVD, length of small vessels [less than 20 μm] in a 1-mm² area [mm/mm²]), perfused vessel density (PVD, length of perfused small vessels in a 1-mm² area [mm/mm²]), the proportion of perfused small vessels (PPV, PVD divided by TVD), and microvascular flow index (MFI, average of predominant flow classification [0–3] in four quadrants) were calculated according to round table conference guidelines [18,19].

Timing of microcirculation examination, blood sample collection, and laboratory data recording

In addition to T1, perioperative microcirculation was examined at eight other time points in this study: T2, 1 h after anaesthesia induction; T3, 2 h after anaesthesia induction; T4, after ureterovesical anastomosis; T5, the end of surgery; T6, 2 h after surgery; POD1, postoperative day 1; POD2, postoperative day 2; and POD7, postoperative day 7. Blood samples were collected at T1, POD1, and POD2 to measure endocan, diamine oxidase, and neutrophil gelatinase-associated lipocalin (NGAL) levels. Urine samples were collected at T6, POD 1, and POD 2 for measuring urine NGAL levels. The blood urea nitrogen and creatinine levels on the day before surgery and at T6, POD1, POD2, POD3, and POD7 were recorded. Arterial blood gas analysis, electrolyte, and lactate levels were measured at T1, T3, T4, T5, and T6.

Primary outcomes, sample size analysis, and other exploratory variables

The first primary outcome of this study was the difference in serum creatinine levels on postoperative day 2 between the two groups. According to our preliminary data, allocating 30 participants to each group provides sufficient power to detect a mean difference of 0.75 mg/dL in creatinine level between the two groups, with an α level of 0.05 (two-tailed) and a β level of 0.2 (80% power), assuming a controlled mean creatinine level of 2.5 mg/dL with a standard deviation of 1.0. The other primary outcome of this study was the difference in TVD at T6 between the two groups. According to our previous study [4], allocating 30 participants to each group provides sufficient power to detect a mean difference of 2.1 mm/mm² in TVD between the two groups, with an α level of 0.05 (two-tailed) and a β level of 0.2 (80% power), assuming a controlled mean TVD of 22.8 mm/mm² with a standard deviation of 2.7. Other exploratory variables included differences between the two groups in terms of hemodynamic variables; other microcirculation variables; lactate, blood urea nitrogen, creatinine, endocan, diamine oxidase, serum NGAL, and urine NGAL levels; tacrolimus level; and daily urine output at different time points.

Randomisation and blinding methods

Block randomisation with a block size of 10 was used to randomise patients into the two groups and ensure a balance in sample size across the groups over time.
Randomised allocation was performed by a research nurse in another research team in our department who was not otherwise involved in this study. She placed the sheet with the computerised random numbers generated from Excel software into sealed, opaque envelopes. The research nurse opened the sealed envelope before the induction of anaesthesia and confirmed the randomised number (0 = control, 1 = treatment, filled the checkbox in a case report form) and co-signed on the envelope with a colleague in the operation room. The envelope and randomised numbers were kept with the patient’s consent for reference. The video sequences of microcirculation were numbered, and the single observer who examined the microcirculation was blinded to the grouping.

**Statistical analysis**

All data were analysed using SPSS 20 (IBM SPSS, Chicago, IL, USA). Normality was examined using the Shapiro-Wilk test. Normally distributed data were presented as means (standard deviation) and analysed using $t$ test. Non-normally distributed data were presented as medians (interquartile range) and analysed using Mann-Whitney $U$ test. Categorical data were analysed using a Fisher’s exact test. A $p$ value $> .05$ was considered significant. The bootstrap method was used for calculating the median difference (95% confidence interval [CI]) for the comparisons of primary outcomes and lactate level.

**Results**

**Patient characteristics, operation durations, and intraoperative medications**

From August 2016 to March 2019, 71 patients undergoing kidney transplantation were assessed for eligibility, and 60 patients were enrolled (Figure 1). Patient characteristics, operation durations, and intraoperative medications are shown in Table 1. Fifty-eight patients received living kidney transplantation, and two patients in the control group received cadaveric kidney transplantation. The intraoperative infusion rate of dexmedetomidine was 0.19 (0.15–0.25) μg/kg/h for patients in the dexmedetomidine group. The end-tidal

![Figure 1. Consort flowchart of patient recruitment.](image-url)
concentration of desflurane did not differ significantly between the two groups during the operation.

**Primary outcomes**

Creatinine levels at POD2 were significantly lower in the dexmedetomidine group than in the control group (1.5 (1.1–2.4) vs. 2.2 (1.7–3.0) mg/dL, median difference −0.6 (95% CI, −0.7 to −0.5) mg/dL, p = .018) (Table 2). After exclusion of the creatinine levels of the one patient with nephrectomy (6.6 mg/dL) in the dexmedetomidine group and two patients with a cadaveric kidney transplant (3.6 and 2.9 mg/dL, respectively) in the control group. The creatinine levels at POD 2 remained significantly lower in the dexmedetomidine group than in the control group (1.5 (1.1–2.4) vs. 2.2 (1.6–2.7) mg/dL, median difference −0.6 (95% CI, −0.7 to −0.5) mg/dL, p = .016). TVD at T6 did not differ significantly between the dexmedetomidine and control groups (24.0 (22.8–24.9) vs. 24.1 (22.5–25.1) mm/mm², median difference −0.1 (95% CI −0.2–0.1) mm/mm², p = .918).

**Other exploratory variables**

**Macrocirculation variables**

Hemodynamic variables are shown in Figure 2. Heart rate at T6 was lower in the dexmedetomidine group than in the control group (81 (71–89) vs. 89 (79–96) beats per minute, p = .015). MAP did not differ significantly between the two groups. Cardiac indexes at T5 and T6 were lower in the dexmedetomidine group than in the control group (T5, 2.9 (2.4–3.5) vs. 3.3 (2.8–4.0) L/min/m², p = .017; T6, 2.8 (2.2–3.3) vs. 3.1 (2.7–3.8) L/min/m², p = .04). Stroke volume index at T2 was lower in the dexmedetomidine group than in the control group (35 (28–39) vs. 40 (32–49) mL/m²/beat, p = .035). Daily urine output did not differ significantly between the two groups at each time point (Table 2).

**Microcirculatory variables**

Images depicting the sublingual microcirculation of several patients at 2 h after surgery (T6) are shown in Figure 3. TVD and PVD did not differ significantly between the two groups (Figure 4). PPV and MFI did not differ significantly between the two groups.

**Laboratory data variables**

In addition to POD2, creatinine levels were higher in the control group than in the dexmedetomidine group at POD1 and POD3 (Table 2). Blood urea nitrogen levels were higher in the control group than in the dexmedetomidine group at POD2 and POD3. Lactate levels were significantly lower in the dexmedetomidine group than in the control group at T4, T5, and T6 (T6, 1.0 (0.7–1.3) vs. 1.4 (1.0–1.8) mmol/L, median difference −0.4 (95% CI −0.4 to −0.3) mmol/L, p = .003). Other laboratory data variables, including endocan, diamine oxidase, serum NGAL and urine NGAL levels did not differ significantly between the two groups. Tacrolimus levels at POD3 and POD7 did not differ significantly between the control and dexmedetomidine groups (POD3, 7.6 (4.9–11.5) vs. 6.7 (5.1–10.3) ng/mL,

### Table 1. Patient characteristics, operation duration, and perioperative management.

| Group          | Control | Dexmedetomidine |
|----------------|---------|-----------------|
| n              | 30      | 30              |
| Female, n (%)  | 10 (33%)| 12 (40%)        |
| Age (years)    | 43 (34–53) | 48 (27–56) |
| Weight (kg)    | 69.6 (14.5) | 61.3 (10.7) |
| Height (cm)    | 168 (10) | 166 (10)       |
| Hemodialysis, n (%) | 17 (57%) | 19 (63%)    |
| Peritoneal dialysis, n (%) | 13 (43%) | 11 (37%)    |
| Medical history|         |                 |
| Hypertension, n (%) | 16 (53%) | 25 (83%)   |
| Diabetes mellitus, n (%) | 6 (20%) | 4 (13%)   |
| Coronary artery disease, n (%) | 0 (0%) | 5 (17%)   |
| Preoperative BUN (mg/dL) | 65 (29) | 68 (23)   |
| Preoperative creatinine (mg/dL) | 11.5 (4.5) | 11.3 (4.1) |
| ABO-incompatible transplantation | 9 (30%) | 8 (27%)      |
| Operation duration (min) | 196 (180–234) | 217 (189–243) |
| Intraoperative management|         |                 |
| Dexmedetomidine (µg/kg/h) | − | 0.19 (0.15–0.25) |
| Fluid supplement (mL) | 2650 (2075–3000) | 2250 (1775–2600) |
| Norepinephrine use, n (%) | 19 (63%) | 17 (57%)    |
| Ephedrine use, n (%) | 8 (27%) | 10 (33%)    |
| Furosemide use, n (%) | 6 (20%) | 6 (20%)     |
| Postoperative 2 h at PACU |     |                 |
| Fluid supplement (mL) | 300 (175–300) | 300 (238–500) |

Values are presented as number, number (%), means (standard deviation), medians (interquartile range). BUN, blood urea nitrogen; PACU, post-anaesthesia care unit.
Table 2. Laboratory data, urine output, and organ injury markers.

| Group                  | Control     | Dexmedetomidine | P values |
|------------------------|-------------|-----------------|----------|
|                        | N=30        | N=30            |          |
| Lactate (mmol/L)       |             |                 |          |
| After induction of anaesthesia | 0.9 (0.8–1.2) | 0.8 (0.7–1.0)  | .157     |
| T3                     | 1.3 (1.0–1.7) | 0.9 (0.8–1.3)  | .091     |
| T4                     | 1.2 (1.0–1.6) | 1.0 (0.7–1.3)  | .032     |
| T5                     | 1.5 (1.0–1.7) | 1.0 (0.8–1.3)  | .007     |
| T6                     | 1.4 (1.0–1.8) | 1.0 (0.7–1.3)  | .003     |
| Creatinine (mg/dL)     |             |                 |          |
| Postoperative day 1    | 5.4 (4.2–6.9) | 3.8 (2.7–5.5)  | .050     |
| Postoperative day 2    | 2.2 (1.7–3.0) | 1.5 (1.1–2.4)  | .018     |
| Postoperative day 3    | 1.6 (1.3–2.1) | 1.3 (0.9–1.7)  | .024     |
| Postoperative day 7    | 1.3 (1.1–1.6) | 1.1 (0.9–1.5)  | .278     |
| Blood urea nitrogen (mg/dL) |         |                 |          |
| Postoperative day 1    | 49.3 (35.6–60.2) | 40.1 (26.3–51.0) | .061     |
| Postoperative day 2    | 33.2 (24.4–43.1) | 21.7 (15.7–32.9) | .008     |
| Postoperative day 3    | 28.1 (23.6–40.3) | 21.7 (17.8–32.2) | .034     |
| Postoperative day 7    | 31.5 (23.9–42.3) | 31.8 (26.7–35.8) | .796     |
| Urine output (mL)      |             |                 |          |
| Postoperative day 1    | 5035 (3581–8700) | 7005 (3558–9795) | .246     |
| Postoperative day 2    | 3740 (2784–5141) | 4355 (3105–7743) | .196     |
| Postoperative day 3    | 3640 (2758–4655) | 3915 (3070–5465) | .304     |
| Serum NGAL (ng/mL)     |             |                 |          |
| After induction of anaesthesia | 1854 (915–4119) | 1444 (649–3986) | .409     |
| Postoperative day 1    | 2161 (375–7963) | 2086 (682–4679) | .566     |
| Postoperative day 2    | 931 (482–5210)  | 672 (517–1968)  | .459     |
| Urine NGAL (ng/mL)     |             |                 |          |
| T6                     | 528 (247–855)  | 420 (300–1139)  | .486     |
| Postoperative day 1    | 184 (71–425)   | 171 (59–493)    | .877     |
| Postoperative day 2    | 141 (77–308)   | 98 (3–262)      | .299     |
| Endocan (ng/mL)        |             |                 |          |
| After induction of anaesthesia | 0.91 (0.48–1.60) | 0.99 (0.62–2.99) | .824     |
| Postoperative day 1    | 0.78 (0.45–1.59) | 0.93 (0.55–1.59) | .932     |
| Postoperative day 2    | 0.79 (0.43–1.54) | 0.91 (0.62–2.15) | .265     |
| Diamine oxidase (U/L)  |             |                 |          |
| After induction of anaesthesia | 2.6 (1.0–5.9)  | 3.3 (1.7–7.5)   | .165     |
| Postoperative day 1    | 5.2 (2.6–10.3)  | 6.1 (1.4–11.2)  | .744     |
| Postoperative day 2    | 4.5 (2.3–8.6)   | 5.9 (3.8–8.3)   | .447     |

Values are median (interquartile range). NGAL, neutrophil gelatinase-associated lipocalin; T3, 2 h after anaesthesia induction; T4, after ureterovesical anastomosis; T5, the end of surgery; T6, 2 h after surgery.

$P = .631$; POD7, 6.5 (5.0–8.5) vs. 7.0 (5.8–8.5) ng/mL, $P = .610$.

Post-hoc analysis

During the manuscript reviewing process, a post-hoc analysis was requested. Three patients were excluded from the post-hoc analysis as follows: two patients in the control group received cadaveric kidney transplantation and one patient in the dexmedetomidine group underwent nephrectomy of the transplanted kidney 10 days after the operation for acute rejection and infarction of the transplanted kidney. After exclusion of the three patients, the patient characteristics, operation durations, and intraoperative medications are shown in Supplementary Table 1. The creatinine levels on POD 2 remained significantly lower in the dexmedetomidine group than in the control group (1.5 (1.1–2.4) vs 2.2 (1.6–2.7) mg/dL, median difference −0.6 (95% CI, −0.1–−0.3) mmol/L, $P = .016$). TVD at T6 did not differ significantly between the dexmedetomidine and control groups (24.1 (22.9–24.6) vs. 24.0 (22.9–25.1) mm/mm², median difference 0.1 (95% CI, −0.1–−0.3) mmol/L, $P = .804$). Hemodynamic variables are shown in Supplementary Figure 1. TVD and PVD did not differ significantly between the two groups (Supplementary Figure 2). PPV and MFI did not differ significantly between the two groups.

In addition to POD2, creatinine levels were higher in the control group than in the dexmedetomidine group at POD1 and POD3 (Supplementary Table 2). Blood urea nitrogen levels were higher in the control group than in the dexmedetomidine group on POD2 and POD3. Lactate levels were significantly lower in the dexmedetomidine group than in the control group at T6 (0.7–1.4) vs. 1.3 (1.0–1.9) mmol/L, median difference −0.3 (95% CI, −0.4 to −0.3) mmol/L, $P = .008$).

Discussion

In this study, we observed that creatinine and blood urea nitrogen levels on POD2 were lower in the dexmedetomidine group than in the control group. However, the creatinine level on POD7 did not differ
significantly between the two groups. Moreover, sublingual microcirculation did not differ significantly between the two groups at 2 h after surgery.

Several potential mechanisms may explain the finding that early postoperative renal function after dexmedetomidine infusion was better in this study. First, several studies have reported that dexmedetomidine improved regional perfusion [13,20,21]. The favourable effect of such perfusion on mesenteric microcirculation had been reported in several studies [13,20]. Second, dexmedetomidine has been reported to attenuate inflammation and reduce ischemic–reperfusion injury in several studies [22–24]. However, we did not observe significant differences in the kidney and intestinal injury markers between the two groups. The intraoperative administration of high-dose steroids for immunosuppression in kidney transplant recipients may have attenuated the difference in inflammation and ischemia-reperfusion injury between the two groups. Third, dexmedetomidine’s effect on polyuria has been reported in several studies [8,25]. Because this study did not aim to investigate differences in daily urine output, our non-significantly higher daily urine output at POD1 and POD2 after dexmedetomidine treatment may suggest that further studies are warranted to investigate dexmedetomidine’s effect on urine output after kidney transplantation. In addition, a retrospective cohort study reported that perioperative dexmedetomidine use reduced the incidence of delayed graft function, risk of infection, risk of acute rejection, overall complication, and length of hospital stay [26].

Two factors may explain how microcirculation could be preserved after dexmedetomidine treatment at 2 h after surgery. First, to ensure patient safety, we did not apply a fixed dexmedetomidine infusion rate during the operation; we used blood pressure, heart rate, and cardiac index to determine the dexmedetomidine infusion rate for each patient. Mohamed et al. reported that high dexmedetomidine infusion (0.5 μg

Figure 2. Hemodynamic variables at each time point. CI, cardiac index; HR, heart rate; MAP, mean arterial pressure; SVI, stroke volume index. *P < .05 indicates significant differences between the two groups determined using the Mann-Whitney U test. T1, before anesthesia induction; T2, 1 h after anesthesia induction; T3, 2 h after anesthesia induction; T4, after ureterovesical anastomosis; T5, the end of surgery; T6, 2 h after surgery.
kg\(^{-1}\) h\(^{-1}\)) improved sublingual microcirculation variables in patients undergoing on-pump coronary artery bypass graft surgery [27]. Second, it has been reported that increased MAP improves microcirculation and reduces the incidence of renal failure in patients with sepsis [28,29]. In our previous study, we found that MAP was moderately positively correlated to microcirculation in patients on dialysis [4]. Because of adequate fluid supplements and medications, the median MAP of both groups in our study exceeded 80 mm Hg from T1 to T6.

Although a concomitant analysis of sublingual microcirculation mirrored the findings of a contrast-enhanced ultrasound examination of the kidney in a septic animal study [30], the correlation between sublingual microcirculation and microcirculation of transplanted kidney remains unknown. Please notice that our results of sublingual microcirculation could not directly reflect the change in the microcirculation of the transplanted kidney. Further studies are warranted to apply other advanced imaging techniques to investigate the microcirculation on the surface or inside of the transplanted kidney. These include a full-field laser perfusion imager by employing the laser speckle contrast imaging technique [21], contrast-enhanced ultrasound [31], and magnetic resonance imaging technique [32].

Preoperative desensitisation and immunosuppressive therapy in ABO-incompatible kidney recipients might attenuate the anti-inflammatory effect of dexmedetomidine during the operation. The randomisation design of this study prevented the significantly unequal number of patients with ABO-incompatible transplantation between the two groups. Moreover, tacrolimus was used in this study, and it had the potential to affect renal function. We did not observe that tacrolimus levels were significantly different between the two groups.

**Limitations**

This study has several limitations. First, this was a single-centre study. Different protocols of fluid supplementation, target MAP, and cardiac output goal may have different influences on the effects of dexmedetomidine on renal function and microcirculation. Many concurrent interventions during kidney transplants make it difficult to identify a mechanism for the observed effects. Second, the use of dexmedetomidine was not blinded. There were two reasons for not blinding. One was the safety issue of living kidney transplantation, and we aimed to maintain adequate cardiac output for all participants. The other was the difficulty to blind the bradycardia effect of...
dexmedetomidine. Third, the sample size of this study was not designed and powered to detect the difference between the two groups on POD7. Further studies are required to investigate the longer effect of dexmedetomidine. Fourth, two patients with cadaver kidney transplants in the control group were excluded from the analysis, their creatinine levels on postoperative day 2 were 3.6 and 2.9 mg/dL respectively. Further study is required to investigate the effect of dexmedetomidine in patients with cadaver kidney transplantation. Fifth, there are many components to performing the postoperative bundle care of kidney transplant recipients, and further studies are required to investigate other interventions after kidney transplantation for early recovery or long-term preservation of renal function.

Conclusions

In conclusion, we found that early postoperative renal function was better in kidney transplant recipients receiving perioperative dexmedetomidine infusion. The sublingual microcirculation in kidney transplant recipients of the dexmedetomidine group was preserved by maintaining adequate cardiac output and mean arterial pressure. Further studies are warranted to investigate the mechanism and effects of dexmedetomidine on other postoperative clinical outcomes in kidney transplant recipients.

Acknowledgements

The authors would like to thank all participants of the National Taiwan University Hospital Center of Microcirculation Medical Research (NCMMR), including Prof. Wei-Zen Sun, Prof. Ya-Jung Cheng, Dr. Chih-Min Liu, and Dr. Po-Yuan Shih. Furthermore, the authors thank Mr. Roger Lien (technician, MicroStar Instruments Co, Ltd, Taipei, Taiwan) and Ms. Yun-Ping Yang for technical assistance related to microcirculation analysis. This manuscript was edited by Wallace Academic Editing.

Author contributions

All authors have made substantial contributions to this work and have approved the final version of the manuscript. Concept and design: YCW, MJW, Cyl, CCC, MKT, YCY.
Acquisition of data: YCW, CYL, CCC, YCY. Statistical analysis: YCW, WSC, YCY. Interpretation of data: YCW, CYL, CTC, AC, WSC, MKT, YCY. Writing original draft: YCW, YCY. Writing review and editing: all authors.

**Disclosure statement**

Dr. Yeh reported receiving grants from the Ministry of Science and Technology, Taiwan. The other authors declare no competing interests.

**Funding**

This study was supported by a grant from the Ministry of Science and Technology, Taiwan (MOST 105-2314-B-002-037-MY2).

**ORCID**

Yu-Chang Yeh  http://orcid.org/0000-0001-5143-5520

**Data availability statement**

The data that support the findings of this study are available from the corresponding author upon reasonable request.

**References**

[1] Perico N, Cattaneo D, Sayegh MH, et al. Delayed graft function in kidney transplantation. Lancet. 2004;364(9447):1814–1827.

[2] Smith SF, Hosgood SA, Nicholson ML. Ischemia-reperfusion injury in renal transplantation: 3 key signaling pathways in tubular epithelial cells. Kidney Int. 2019;95(1):50–56.

[3] Zafrañi L, Ince C. Microcirculation in acute and chronic kidney diseases. Am J Kidney Dis. 2015;66(6):1083–1094.

[4] Yeh YC, Chao A, Lee CY, et al. An observational study of microcirculation in dialysis patients and kidney transplant recipients. Eur J Clin Invest. 2017;47(9):630–637.

[5] Khan ZP, Ferguson CN, Jones RM. Alpha-2 and imidazoline receptor agonists. Their pharmacology and therapeutic role. Anaesthesia. 1999;54(2):146–165.

[6] Kamibayashi T, Hayashi Y, Mammoto T, et al. Role of the vagus nerve in the antidysrhythmic effect of dexmedetomidine on halothane/epinephrine dysrsytisms in dogs. Anesthesiology. 1995;83(5):992–999.

[7] Tracey KJ. The inflammatory reflex. Nature. 2002;420(6917):853–859.

[8] Leino K, Hynynen M, Jalonen J, et al. Renal effects of dexmedetomidine during coronary artery bypass surgery: a randomized placebo-controlled study. BMC Anesthesiol. 2011;11(1):9.

[9] Kulka PJ, Tryba M, Zenz M. Preoperative alpha2-adrenergic receptor agonists prevent the deterioration of renal function after cardiac surgery: results of a randomized, controlled trial. Critical Care Med. 1996;24(6):947–952.

[10] Kilic K, Hanci V, Selek S, et al. The effects of dexmedetomidine on mesenteric arterial occlusion-associated gut ischemia and reperfusion-induced gut and kidney injury in rabbits. J Surg Res. 2012;178(1):223–232.

[11] Gu J, Sun P, Zhao H, et al. Dexmedetomidine provides renoprotection against ischemia-reperfusion injury in mice. Crit Care. 2011;15(3):R153.

[12] Wang ZX, Huang CY, Hua YP, et al. Dexmedetomidine reduces intestinal and hepatic injury after hepatectomy with inflow occlusion under general anaesthesia: a randomized controlled trial. British J Anaesth. 2014;112(6):1055–1064.

[13] Yeh YC, Sun WZ, Ko WJ, et al. Dexmedetomidine prevents alterations of intestinal microcirculation that are induced by surgical stress and pain in a novel rat model. Anesth Analg. 2012;115(1):46–53.

[14] Xu JY, Ma SQ, Pan C, et al. A high mean arterial pressure target is associated with improved microcirculation in septic shock patients with previous hypertension: a prospective open label study. Crit Care. 2015;19:130.

[15] Tripodaki ES, Tasoulias A, Koliopoulos A, et al. Microcirculation and macrocirculation in cardiac surgical patients. Crit Care Res Pract. 2012;2012:1–9.

[16] Schulz KF, Altman DG, Moher D. CONSORT 2010 statement: updated guidelines for reporting parallel group randomised trials. BMJ. 2010;340:c332.

[17] Tsai MK, Wu MS, Yang CY, et al. B cells and immunoglobulin in ABO-incompatible renal transplant patients receiving rituximab and double filtration plasmapheresis. J Formos Med Assoc. 2015;114(4):353–358.

[18] De Backer D, Hollenberg S, Boerma C, et al. How to evaluate the microcirculation: report of a round table conference. Crit Care. 2007;11(5):R101.

[19] Ince C, Boerma EC, Cecconi M, et al. Second consensus on the assessment of sublingual microcirculation in critically ill patients: results from a task force of the European Society of Intensive Care Medicine. Intensive Care Med. 2018;44(3):281–299.

[20] Miranda ML, Balarini MM, Bouskela E. Dexmedetomidine attenuates the microcirculatory derangements evoked by experimental sepsis. Anesthesiology. 2015;122(3):619–630.

[21] Yeh YC, Wu CY, Cheng YJ, et al. Effects of dexmedetomidine on intestinal microcirculation and intestinal epithelial barrier in endotoxemic rats. Anesthesiology. 2016;125(2):355–367.

[22] Bao N, Dai D. Dexmedetomidine protects against ischemia and reperfusion-induced kidney injury in rats. Mediators Inflamm. 2020;2020:2120971.

[23] Liu YE, Tong CC, Zhang YB, et al. Effect of dexmedetomidine on rats with renal ischemia-reperfusion injury and the expression of tight junction protein in kidney. Int J Clin Exp Med. 2015;8(10):18751–18757.

[24] Wang K, Wu M, Xu J, et al. Effects of dexmedetomidine on perioperative stress, inflammation, and immune function: systematic review and meta-analysis. Br J Anaesth. 2019;123(6):777–794.
[25] Villela NR, do Nascimento Junior P, de Carvalho LR, et al. Effects of dexmedetomidine on renal system and on vasopressin plasma levels. Experimental study in dogs. Rev Bras Anestesiol. 2005;55(4):429–440.

[26] Chen J, Perez R, de Mattos AM, et al. Perioperative dexmedetomidine improves outcomes of kidney transplant. Clin Transl Sci. 2020;13(6):1279–1287.

[27] Mohamed H, Hosny H, Tawadros Md P, et al. Effect of dexmedetomidine infusion on sublingual microcirculation in patients undergoing on-pump coronary artery bypass graft surgery: a prospective randomized trial. J Cardiothorac Vasc Anesth. 2019;33(2):334–340.

[28] Fiorese Coimbra KT, de Freitas FGR, Bafi AT, et al. Effect of increasing blood pressure with noradrenaline on the microcirculation of patients with septic shock and previous arterial hypertension. Critical Care Med. 2019;47(8):1033–1040.

[29] Beloncle F, Radermacher P, Guerin C, et al. Mean arterial pressure target in patients with septic shock. Minerva Anestesiol. 2016;82(7):777–784.

[30] Lima A, van Rooij T, Ergin B, et al. Dynamic contrast-enhanced ultrasound identifies microcirculatory alterations in sepsis-induced acute kidney injury. Crit Care Med. 2018;46(8):1284–1292.

[31] Schneider AG, Hofmann L, Wuerzner G, et al. Renal perfusion evaluation with contrast-enhanced ultrasonography. Nephrol Dial Transplant. 2012;27(2):674–681.

[32] Ganesh T, Estrada M, Yeger H, et al. A non-invasive magnetic resonance imaging approach for assessment of real-time microcirculation dynamics. Sci Rep. 2017;7(1):7468.