Bisleuconothine A protects periodontal tissue via the regulation of RANKL expression and infiltration of inflammatory cells

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Abstract

Purpose: To investigate the protective effect of bisleuconothine A on periodontal tissue in rats and the mechanism involved.

Methods: Adult male Sprague Dawley rats (n = 32) weighing 180 - 200 g (mean weight, 190 ± 10 g) were randomly assigned to four groups of eight rats each: control group, periodontitis group, bisleuconothine A (50 mg/kg) group and bisleuconothine A (100 mg/kg) group. Rats in the treatment groups received bisleuconothine intraperitoneally for two weeks. Periodontitis was induced in the rats using standard procedures. Serum and tissue samples were used for biochemical analysis. Alveolar bone loss was measured in rat maxillae, while the activity of bone alkaline phosphatase (BALP) was determined in serum. Tumor necrosis factor α (TNF-α), interleukin-1β (IL-1β) and interleukin-6 (IL-6) were determined in gingival tissue using enzyme-linked immunosorbent assay (ELISA) kit. Gene and protein expressions of receptor activator of nuclear factor kappa-β ligand (RANKL), osteoprotegerin (OPG), and matrix metallopeptidase-9 (MMP-9) were measured in gingival tissue using real-time quantitative polymerase chain reaction (qRT-PCR) and Western blotting, respectively.

Results: Bisleuconothine A treatment significantly and dose-dependently reduced alveolar bone loss, as well as serum levels of TNF-α, IL-1β and IL-6, but increased BALP activity in periodontitis rats (p < 0.05). It also significantly and dose-dependently reduced mRNA expressions of RANKL and MMP-9, but significantly increased OPG mRNA expression (p < 0.05). Similarly, treatment with bisleuconothine A significantly and dose-dependently down-regulated RANKL, p-NF-kB, p-IkBα and iNOS proteins in gingival tissue of periodontitis rats (p < 0.05). The results of histopathological examination indicated that bisleuconothine A treatment significantly reversed histological changes in periodontal tissues of periodontitis rats. It also significantly reduced the degree of polymorphonuclear (PMN) cell infiltration in periodontal tissue.

Conclusion: The results obtained show that bisleuconothine A protects periodontal tissue via the regulation of RANKL expression and infiltration of inflammatory cells.

Keywords: Bisleuconothine A, Expression, Inflammation, Periodontitis, RANKL

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INTRODUCTION

Periodontitis is a chronic inflammatory and infectious disease in which the integrity of gum tissue is disrupted [1]. Inflammation-induced damage to gum connective tissues reduces the anchoring ability of the teeth and may lead to tooth loss [2]. Environmental and genetic factors contribute significantly to periodontal disease via alterations in inflammatory/immune responses [3]. Studies have shown that inflammatory cytokines such as IL-6 and IL-8 activate matrix metalloproteinases (MMPs), thereby promoting osteoclastogenesis [4]. Receptor activator of nuclear factor kappa-B ligand (RANKL), also known as tumor necrosis factor ligand superfamily member 11, tumor necrosis factor (TNF)-related activation-induced cytokine, OPG ligand or osteoclast differentiation factor, regulates cell proliferation by modifying Id4, Id2 and cyclin D1 protein levels. In humans, it is encoded by the TNFSF11 gene. On binding to its receptor, RANKL induces osteoclastogenesis [5]. The binding of RANKL to OPG reduces its activity. Osteoclastogenesis is regulated by maintaining RANKL/OPG ratio. Studies have shown that cytokines decrease OPG expression, thereby stimulating RANKL and ultimately tooth loss [6].

Although surgery and mechanical therapy are used for the management of periodontal disease, they are however ineffective. Adjuvant therapies such as non-steroidal anti-inflammatory drugs (NSAIDs) and antibiotics are also used for the treatment of periodontitis. Their clinical application is however limited due to some setbacks. Thus, the development of new therapeutic approaches for effective management of periodontitis has become necessary. Over the last few decades, alternative medicine has shown great potential in the treatment of several diseases including periodontitis.

Bisleuconothine A is a bisindole alkaloid isolated from *Leuconotis griffithii* [7]. Bisindole alkaloids have received considerable attention for their potential biological properties, including antitumor, antimalarial, anti-inflammatory and antibacterial effects [8-11]. The antitumor effect of bisleuconothine A is exerted via Wnt and AKT/mTOR signalling pathways [12]. These pathways are regulated by inflammatory cytokines [13]. Bisleuconothine A regulates these pathways via modulation of cytokine expressions. This study investigated the protective effect of bisleuconothine A on periodontal tissue in rats and the mechanism involved.

EXPERIMENTAL

Rats

Adult male Sprague Dawley rats weighing 180 - 200 g (mean weight, 190 ± 10 g) were used for this study. They were housed in metal cages under standard conditions and had free access to standard feed and water. The rats were exposed to 12-h light/12-h dark cycle, and maintained at a temperature of 25 °C and 65 % humidity. The rats were acclimatized to the laboratory conditions for one week prior to commencement of the study. The study protocol was approved by the Institutional Animal Ethics Committee of Stomatology Hospital, Zhejiang University School of Medicine, China (approval no. IAEC/SH/ZUSM/2017/08), and the study procedures were carried out according to the guidelines of the International Association for the Assessment and Accreditation of Laboratory Animal Care (AAALAC) [14].

Experimental design

The rats (n = 32) were randomly assigned to four groups of eight rats each: control group, periodontitis group, bisleuconothine A (50 mg/kg bwt) group and bisleuconothine A (100 mg/kg bwt) group. Rats in the treatment groups received bisleuconothine intraperitoneally for two weeks.

Preparation of rat model of periodontitis

Periodontitis was induced in the rats using standard procedures [15]. The rats were anesthetized via intraperitoneal administration of 50 mg/kg bwt pentobarbital and a suture was made around each cervix of the first molar (M1) on bilateral sides in the maxilla in the open mouth, using Hashimoto’s gag. Mesial site of M1 was knotted with the suture, but in control group M1 was not ligated with the suture.

Blood sample collection

At the end of the 14th day, the rats were fasted overnight and euthanized. Blood was collected from the rats via cardiac puncture and centrifuged at 3000 rpm for 10 min to obtain serum. The excised bone tissue and serum were used for biochemical analysis.

Determination of BALP activity

At the end of the 14th day, the rats were fasted overnight and euthanized. Blood was collected from the rats via cardiac puncture and centrifuged at 3000 rpm for 10 min to obtain serum. The excised bone tissue and serum were used for biochemical analysis.

Total alkaline phosphatase (ALP) activity was determined in rat serum using ALP assay kit.
Aliquot of serum was subjected to thermo-activation for 10 min at 56 °C, and ALP activity was measured in the heated serum. The activity of BALP was obtained by taking the difference between ALP activities in unheated serum and heated serum.

**Determination of alveolar bone loss**

Maxillae isolated from the rats were stained with 1 % methylene blue after an initial fixation in 10 % formalin solution. Bone loss was determined by measuring the length of each molar. Alveolar bone loss was estimated using sum of the measurements from buccal tooth surfaces and the difference between the values of the right maxilla and those of left maxilla. Imaging software was then used to measure the distance, and standardized digital photography was used for the morphometric analysis of alveolar bone.

**Assessment of levels of cytokines in gingival tissue**

Gingival tissues were isolated from each rat and homogenized with phosphate buffer using mechanical homogenizer. Levels of IL-1β, IL-6 and TNF-α were determined in the resultant tissue homogenate using ELISA kit.

**Quantitative real-time polymerase chain reaction (qRT-PCR)**

The mRNA levels of OPG, RANKL, and MMP-9 were determined in gingival tissue using qRT-PCR. Each tissue was trypsinized using 0.05 trypsin and the resultant cell suspension was subjected to qRT-PCR. Trizol RNA extraction reagent was used to extract total RNA from cells of each group, while cDNA synthesis kit was used to perform cDNA synthesis reaction according to the instructions of the manufacturer. Light Cycler 1536 RT-PCR detection system was used for the estimation of the mRNA expressions of the genes. Variation in the cDNA content was normalized using β-actin. The PCR reaction mixture (20 μL) consisted of 6.4 μL of dH2O, 1.6 μL of gene-specific primer (10 μM), 2 μL of synthesized cDNA and 10 μL of SYBR Premix Ex Taq™ II. The Ct value of U6 was taken as the internal parameter, and 2^−ΔΔCt was used to calculate the relative expression levels of the proteins.

**Western blotting**

The cell suspension was washed with phosphate-buffered saline (PBS) and NP40 protein lysis buffer was used to lyse the cells. The lysate was centrifuged at 12,000 rpm for 10 min at 4 °C, and the protein concentration of the supernatant was determined using DC protein assay kit. A portion of total cell protein (30 μg) from each sample was separated on 10 % sodium dodecyl sulphate (SDS)-polyacrylamide gel electrophoresis and transferred to a fixed polyvinylidene fluoride membrane at 110 V and 90 °C for 120 min. Subsequently, non-fat milk powder (3 %) in Tris-buffered saline containing 0.2 % Tween-20 (TBS-T) was added with gentle shaking at 37 °C and incubated to block non-specific binding of the blot. Incubation of the blots was performed overnight at 4 °C with primary antibodies of RANKL, iNOS, IkB-α, NF-κB and β-actin, each at a dilution of 1 to 1000. Then, the membrane was washed thrice with TBS-T and further incubated with horseradish peroxidase-conjugated goat anti-rabbit IgG secondary antibody for 1 h at room temperature. The blot was developed using an X-ray film. Grayscale analysis of the bands was performed using Bio-rad gel imaging system. The respective protein expression levels were normalized to that of β-actin which was used as a standard.

**Histopathological examination of gingival tissue**

Tissue specimen from rat molars was separated out and fixed in 10 % formalin solution. It was subsequently treated for a period of four weeks with EDTA to allow for decalcification. The tissue was thereafter embedded in paraffin. Serial sections, each of 4 μm thickness, were made using a microtome and stained with hematoxylin and eosin according to standard method, and examined under light microscope. Different tissue regions were selected with the help of morphometric lens in 0.25 mm² microscopic field, and the mean was calculated. ImageJ image analysis software was used to determine the infiltration of inflammatory PMN cells into the gingival tissue of periodontitis rats.

**Statistical analysis**

Data are expressed as mean ± SD. Statistical analysis was performed using GraphPad prism (version 6.1). Groups were compared using Student’s t-test. Statistical significance was assumed at p < 0.05.

**RESULTS**

**Effect of bisleuconothine A on alveolar bone loss**

As shown in Figure 1 A and B, treatment with bisleuconothine A significantly and dose-
dependently reduced alveolar bone loss in periodontitis rats \((p < 0.05)\).

**Figure 1:** Alveolar bone loss in periodontitis rats. (A): Analysis of alveolar bone loss; (B): Morphometric observation of alveolar bone. @@@\(p < 0.05\), when compared with control group; **\(p < 0.05\), when compared with periodontitis group

**Activity of BALP**

Bisleuconothine A treatment significantly and dose-dependently increased the activity of BALP in periodontitis rats \((p < 0.05; \text{Figure 2})\).

**Figure 2:** Effect of bisleuconothine A on BALP activity; @@@\(p < 0.05\), when compared with control group; **\(p < 0.05\), when compared with periodontitis group

**Effect of bisleuconothine A on levels of cytokines in serum of periodontitis rats**

Treatment with bisleuconothine A significantly and dose-dependently reduced the levels of TNF-\(\alpha\), IL-1\(\beta\) and IL-6 in serum of periodontitis rats \((p < 0.05)\). These results are shown in Figure 3 A - C.

**Figure 3:** Levels of inflammatory cytokines in serum of periodontitis rats. (A): Level of TNF-\(\alpha\) in serum of periodontitis rats; (B): Level of IL-1\(\beta\) in serum of periodontitis rats; (C): Level of IL-6 in serum of periodontitis rats; @@@\(p < 0.05\), when compared with control group; **\(p < 0.05\), when compared with periodontitis group

**Levels of expression of RANKL, OPG and MMP-9 in periodontitis rats**

Bisleuconothine A treatment significantly and dose-dependently reduced mRNA expressions of RANKL and MMP-9, but significantly increased OPG mRNA expression \((p < 0.05; \text{Figure 4})\).

**Figure 4:** Effect of bisleuconothine A on mRNA expressions of RANKL, OPG and MMP-9 in gingival tissue of periodontitis rats; @@@\(p < 0.05\), when compared with control group; **\(p < 0.05\), when compared with periodontitis group

**Protein expressions of RANKL, p-NF-\(k\)B, p-I\(k\)B\(\alpha\) and iNOS in gingival tissue of periodontitis rats**

As shown in Figure 5, treatment with bisleuconothine A significantly and dose-dependently down-regulated the protein expressions of RANKL, p-NF-\(k\)B, p-I\(k\)B\(\alpha\) and iNOS in gingival tissue of periodontitis rats \((p < 0.05)\).
DISCUSSION

Periodontitis is a chronic inflammatory disorder caused by an imbalance between host defense mechanism and subgingival microbiota [16]. It refers to inflammation of gums and supporting structures of the teeth. It is one of the most common human diseases. Periodontitis is caused by certain bacteria (known as periodontal bacteria) and by the local inflammation triggered by them.

The available treatment option for periodontitis is associated with several limitations. This study investigated the protective effect of bisleuconothine A on periodontal tissue in rats and the mechanism involved.

In periodontitis, translocation of immune cells to the periodontal tissue promotes the release of inflammatory cytokines [17]. Cytokines such as IL-1β and TNF-α participate in collagen degradation and bone resorption by activating collagenolytic enzymes and osteoclasts [18]. Reduction in cytokine levels has been shown to prevent/delay periodontitis. Thus, anti-inflammatory drugs are normally used for its treatment.

In this study, bisleuconothine A treatment significantly and dose-dependently reduced the levels of inflammatory cytokines in gingival tissue of periodontitis rats. In periodontitis, gingival tissues are destroyed due to increased secretion of IL-6 and TNF-α, a process mediated by RANKL/OPG [19]. It has been reported that in periodontitis, the expression of RANKL is increased, while that of OPG is reduced in gingival tissue, with subsequent tooth loss [20]. In this study, bisleuconothine A treatment significantly reversed the altered expressions of OPG and RANKL in gingival tissue of periodontitis rats. These results are in agreement with those of previous studies [19,20]. The results of histopathological examination which showed that bisleuconothine A treatment significantly reversed histological changes in periodontal tissues of periodontitis rats provided supportive evidence for results from biochemical assays. The treatment also significantly reduced the degree of PMN cell infiltrating in the periodontal tissue.

Oxidative stress has also been implicated in the pathogenesis of periodontitis [21]. The results of this study showed that bisleuconothine A treatment significantly and dose-dependently down-regulated the protein expressions of RANKL, p-NF-kB, p-IkBα and iNOS in gingival tissue of periodontitis rats.
CONCLUSION

The results obtained in this study show that bisleuconothine A protects periodontal tissue via the regulation of RANKL expression and infiltration of inflammatory cells. Thus, bisleuconothine A may have clinical application in the treatment of periodontitis.

DECLARATIONS

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Conflict of interest

No conflict of interest is associated with this study.

Contribution of authors

We declare that this work was done by the authors named in this article and all liabilities pertaining to claims relating to the content of this article will be borne by the authors. Fang Wang conducted the experimental work and literature review related to presented work. Ping Sun perform the statistical analysis, and Qiang Sun supervised the presented work and written the manuscript.

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