Influence of Eco-Friendly Control Strategies on the Germination of Mycotoxin Secreted *Fusarium Verticillioides* Infested Maize

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**ABSTRACT:** *Fusarium verticillioides* is a major seed borne fungal pathogen of maize causing rots and seedling blight, and secretion of toxigenic compounds in grains. This study aimed at reducing the seedling blight and mycotoxin contamination level of maize caused by *F. verticillioides* with environmentally friendly antagonistic microorganisms. One gram of ground mycelia of each of the species of *Trichoderma*, *B. subtilis* and *P. fluorescens* was suspended in 100, 200, 300 and 400 ml of distilled water to determine the inoculum concentrations. Maize seeds were treated with inoculum and planted at least 2 seeds per pot in replicates of 3 pots. Two grams of ground mycelial mat of the pathogen was added (at planting) per ten (10) kilogram of sterile soil. Germination percentage of these seeds was taken on the 14, 21, and 28 days after planting. Observations were on incidence and severity of the disease, from germination period till harvest. *T. pseudokoningii* at its C3 gave the best germination percentage, and *P. fluorescens* inoculum at C2 was the best for the control of the pathogen, which is comparatively effective as Benomyl against *F. verticillioides*. The use of ecofriendly control alternative like *T. pseudokoningii* and *P. fluorescens* should be used to in place Benomyl; it has no residual effect on the environment.

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Maize (*Zea mays* L) is the third most important cereal crop in the tropics (CIMMYT, 2000). It is an important grain crop in the world due to its ability to produce economic yields under the marginal production conditions of low soil fertility and management (FAO, 2005). Maize (*Zea mays* L.) is a food crop of economic significance and nutritional importance in human diet; animal feed, fodder crop and raw materials for industries. Demands for maize have always exceeded its supply in Nigeria, indicating the need for higher and better productivity in terms of quantity and quality (Smale et al., 2011). Maize is highly susceptible to both pests and parasites that cause pre- and post-harvest losses which leads to great economic losses in Nigeria (Samuel et al., 2011). Diseases of maize associated with *F. verticillioides* include seed rot, root rot, stalk rot, kernel or ear rot, and seedling blight (Bacon and Hinton, 1992). It is an important contributor to maize seedling disease, and for the induction of leaf lesions indicative of foliar maize diseases. Fusarium disease of maize is among the destructive diseases in many areas of the country. It reduces crop yield, and its mycotoxin secretion causes food contamination, which makes its consumption poisonous to human and livestock. The control of this disease is important to reduce contamination thereby producing cleaner, safer maize seeds; to enhance higher yields and invariably increase the farmers’ income. The use of environmentally friendly control strategies for the disease management is needed. These are beneficial antagonistic microorganisms isolated from maize rhizosphere to fight against the germination and seedling blight, which are *T. harzianum*, *T. pseudokoningii*, *P. fluorescens* and *B. subtilis* that were tested in this study to control the *F. verticillioides* infestation on maize germinability potentials.

**MATERIALS AND METHODS**

*Sample collection and preparation:* Test biocontrol organisms (*Trichoderma harzianum, T. pseudokoningii, Pseudomonas fluorescens* and *Bacillus subtilis*) used in this study (i.e. the antagonists) were from soil collected in maize growing areas (rhizosphere) of the Institute Agricultural Research and Training, IAR&T and the soil samples collected were taken to Phytopathology Laboratory of the Crop Protection and Environmental Biology Department, University of Ibadan (UI) for isolation of the organisms. These were maintained on antibacterial PDA by adding 0.67 g of streptomycin powder to 500 ml of PDA medium for the fungal growth and PDA for...
bacterial growth. The mycelial mats of the antagonists were cultured on Potato Dextrose Agar (PDA), dried in an oven at 30°C and blended in a sterilized Gallenkamp blender set (which had been thoroughly washed and rinsed with NaOCl) for 1 min. These mycelial mat powders were stored in sterilized Petri dishes and refrigerated at 4°C before use. One gram each of the species of Trichoderma, *B. subtilis* and *P. fluorescens* was suspended in 100, 200, 300 and 400ml of distilled water to determine the inoculum concentrations in each. The concentrations are *T. harzianum* (*C1 = 1.8 x 10^7, C2 = 1.7 x 10^10, C3 = 2.3 x 10^9, C4 = 5.3 x 10^8 spores/ml respectively), *T. pseudokoningii* (*C1 = 2.7 x 10^9, C2 = 5.3 x 10^8, C3 = 7.1 x 10^7, C4 = 1.5 x10^8 spores/ml respectively), *B. subtilis* and *P. fluorescens* (*C1 = 1.0 x 10^8, C2 = 1.0 x 10^9, C3 = 1.0 x10^7, C4 = 1.0 x 10^6 cfu/ml respectively*). The seeds were treated with these inoculum and planted at least 2 seeds per pot in replicates of 3 pots. Two grams of ground mycelial mat of the pathogen was added (at planting) per ten (10) kilogram of sterile soil, mixed thoroughly, to induce the infection. Seeds were also treated with benomyl (Benlate WP 50) a fungicide at a rate of 0.5g a.i./50g of seeds using the same method.

**Analysis and Statistical Evaluation**: The germination percentage of these seeds was counted on the 14, 21, and 28 days after planting. Observation was carried out till harvest. The experiment was laid out in a 2 x 4 factorial in Completely Randomised Design. Data collected were subjected to descriptive statistics and analysis was done using SAS ANOVA at *p* ≤ 0.05 to test for significant differences in treatment means.

**RESULTS AND DISCUSSION**

At 28 days after planting, *C3* of *T. pseudokoningii* had 75% maize germination which was higher and better than the benomyl treated plants that had 73% germination (Table 1). The *C4* of *T. pseudokoningii* had 60% germination and was weaker in germination compared with the control plants which had 72% germination of maize seeds planted in it. It was observed at 7 days after planting that the percentage germination for the control plants were 69% on the average. The control plants gave weak stands and had seedling blight which is symptomatic *F. verticillioides* infection. *T. pseudokoningii* at *C3* gave the best germination percentage which was not significantly different from the *C1*. The least effective of the concentrations of *T. pseudokoningii* inoculum was *C4* = 1.5 x 10^8 spores/ml. Benomyl was 60% germination of the maize seeds significantly higher than the rest of the treatments at 7 DAP, followed by *C2* and *C1* in efficacy. At the 21 and 28 DAP, *C2* (with 72.66% and 73.33%) was significantly different from the benomyl treatment, with 64% (Table 1). *P. fluorescens* at the end of the observation, *C2* (73.33%) and benomyl (64%) are significantly different from each other. *C1, C3 and C4* were significantly better than the control which has the pathogen, *F. verticillioides* only in the soil. The performance of *P. fluorescens* at *C2* was the highest i.e. it has the highest germination percentage in all but was not significantly different from *C1*. Statistically to summarize the effect of the antagonists’ inoculum on the germination of maize seeds planted in infested soil (Table 1). In the first trial, *T. harzianum’s C2* and benomyl are not significantly different from each other, but different from the control, *C1, C3 and C4*. *C1* of this antagonist did not act significantly different from control; control was not also different from *C3*. This suggests that for *T. harzianum* to be effective as an antagonist, nothing more than *C1* should be used and nothing less than *C3* should be used. But to have an efficient control as observed in the benomyl, *C1* should be used. *T. pseudokoningii* at its *C3* gave the best germination percentage which was not significantly different from the *C1* of it. The least effective of the concentrations of *T. pseudokoningii* inoculum was *C4*. From this, it is certain that for efficient control, to have good germination of maize on infested soils, *C3* should be used, and concentration above *C1* should not be used. *P. fluorescens* inoculum at *C2* was the best for the control of the pathogen causing seedling blight of maize seeds. *C2* was as effective as benomyl in this trial against *F. verticillioides*. For the *P. fluorescens*, all the concentrations used were effective, but dilutions less than *C4* should not be used in this kind of control, due to its insensitivity in *in vivo* state. *B. subtilis* was a weak bacterium against this pathogen in vivo. However, *C3* concentration can still be used. Benomyl was the most effective control, but for its residual effect on the environment, as a control measure, the use of environmentally friendly control alternatives like *T. harzianum*, *P. fluorescens* and *T. pseudokoningii* should be used. The efficient control of this pathogen; to have good germination of maize on infested soils, *C3* should be used, and concentrations above *C1* should not be avoided. *P. fluorescens* inoculum at *C2* was the best for the control of the pathogen causing seedling blight of maize seeds. *C2* was as effective as Benomyl in this trial against *F. verticillioides* (Table 1). For the *P. fluorescens*, all the concentrations used were effective, but dilutions less than *C4* should not be used due to its insensitivity in *in vivo* state. *B. subtilis* was a weak bacterium against this pathogen *in vivo*. However, *C3* concentration can still be used. Benomyl was the most effective control, but for its residual effect on the environment, the use
of environmentally friendly control alternatives like *P. fluorescens* and *T. pseudokoningii* are recommended.

### Table 1: The effect of the antagonists’ inoculum on maize seed germination (in %)

| Treatments | *T. harzianum* | *T. pseudokoningii* | *P. fluorescens* | *B. subtilis* |
|------------|----------------|---------------------|-----------------|--------------|
| 1st Trial  |                |                     |                 |              |
| C1         | 75.00b         | 74.00ab             | 63.33ab         | 60.33c       |
| C2         | 84.00a         | 70.00d              | 80.00a          | 72.33b       |
| C3         | 69.00cd        | 75.00a              | 65.00ab         | 60.00c       |
| C4         | 64.00d         | 60.00e              | 59.33ab         | 54.33d       |
| Benomyl    | 86.00a         | 73.00bc             | 86.00a          | 86.00a       |
| Control    | 70.66bc        | 72.00c              | 50.67b          | 70.66b       |
| Mean       | 74.77          | 70.66               | 67.38           | 67.26        |
| S.E        | 5.825          | 1.452               | 27.512          | 5.451        |
| 2nd Trial  |                |                     |                 |              |
| C1         | 63.66a         | 75.00b              | 65.00b          | 58.66b       |
| C2         | 65.00a         | 75.00b              | 73.33a          | 64.66a       |
| C3         | 58.00b         | 80.00a              | 61.33cd         | 54.66bc      |
| C4         | 48.00c         | 60.00e              | 59.33d          | 52.33c       |
| Benomyl    | 64.00a         | 73.00c              | 64.00bc         | 64.00a       |
| Control    | 40.00d         | 72.00d              | 40.00e          | 40.00d       |
| Mean       | 56.44          | 72.50               | 60.49           | 55.71        |
| S.E        | 2.334          | 0.726               | 2.994           | 5.303        |

C1, C2, C3 and C4 are based on individual concentrations. Means of 3 replicates (%). Means in the same column followed by the same letter are not significantly different at 0.05 level. Infected seedlings are usually stunted, wilting, chlorotic, and have pale green or purple leaves and poor roots (CIMMYT, 2004). Symptoms of *Fusarium* stalk rot in mature plants are difficult to distinguish from those of other stalk rots, but the internal tissues of affected stalks are usually reddish-brown and rotted. The discoloration may also be seen on the surface of the stalks near nodes. Stalks are weak and lodge easily. *T. pseudokoningii* had highest effect on maize germination, followed by *P. fluorescens*, and the least *B. subtilis*. Several researchers have reported that an application of fluorescent *Pseudomonas* to seed (Callan et al., 1990), soil and foliage (Praveen et al., 2000) also controlled several plant diseases. Lower disease incidence and resultant yield increase in seeds treated with microbial agents might be attributed to rapid multiplication of antagonists in the soil and its colonisation in the roots of seedlings, thereby preventing the establishment of the pathogens in the rhizosphere. Its effect on the phylloplane was also observed cause of its ability to control the foliar diseases. The present results revealed that *P. fluorescens* significantly controlled ear rot disease and also improved field emergence and grain yield in maize grains. Possibly both rhizosphere (to help the root systems) and phyllosphere populations of *P. fluorescens* helped to control disease. Both direct inhibition of the pathogen to systemically induced resistance in maize plants could be involved in control. Similar observations were also made on rice plants (Albouvette et al., 1993) in the case of *P. fluorescens*.

Increase in yield owing to *P. fluorescens* has been reported in several crops (Niranjan et al., 2004; Srinivas et al., 2005). The activities of species of *Trichoderma* showed good action in the control of the incidence of *Fusarium* ear rot disease on the field. The incidence of *F. verticillioides* was drastically reduced with the *Trichoderma* species compared to the untreated control. The action and ability of *Trichoderma* to act as mycoparasites of hyphae and resting structures of plant pathogens have been demonstrated both in *in vitro* and natural soil (Akinbode and Ikotun, 2011).

*Trichoderma* species are good sources of various enzymes such as exo- and endo-glucanases, cellulase, chitinase and growth stimulators (Mari Aidemark et al., 2010). This proven ability of species of *Trichoderma* to produce diffusible substances toxic to other fungi *in vitro* and even in organic substrates in soil (Mohiddin et al., 2010), strengthens and suggests the importance of the native *Trichoderma* in biocontrol. Other workers have found that there are great variabilities between the different strains of *Trichoderma* in their ability to colonize their rhizosphere. It was observed that there was an increase in susceptibility of the plants to the disease probably due to the absence of the organism in the maize plant phyllosphere; this brought the idea of spraying with the bioagents at 6 weeks after planting.

**Conclusions:** To control *F. verticillioides*, bioagents were isolated from maize rhizosphere; *T. harzianum, T. pseudokoningii, P. fluorescens* and *B. subtilis* that were used in this study. The severity of *Fusarium* ear rot disease of maize was greatly reduced by *T.
pseudokoningii and P. fluorescens which can serve as a good alternative to the synthetic fungicide, benomyl used. Environmentally friendly control strategies with the use of these two bioagents is possible to reduce contamination thereby producing cleaner, safer maize seeds; to enhance higher yields and invariably increase the farmers’ income. The potentials of these bioagents used in the experiment as mycoparasites of hyphae and resting structures of plant pathogens have been demonstrated both in in vitro and natural soil. The safe use of the organisms to control the Fumonisin secreting F. verticillioides in maize is important and should be encouraged for safer maize production.

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