Maternal Lipids Are as Important as Glucose for Fetal Growth

Findings from the Pune Maternal Nutrition Study

OBJECTIVE—To study the relationship between maternal circulating fuels and neonatal size and compare the relative effects of glucose and lipids.

RESEARCH DESIGN AND METHODS—The Pune Maternal Nutrition Study (1993–1996) investigated the influence of maternal nutrition on fetal growth. We measured maternal body size and glucose and lipid concentrations during pregnancy and examined their relationship with birth size in full-term babies using correlation and regression techniques.

RESULTS—The mothers (n = 631) were young (mean age 21 years), short (mean height 151.9 cm), and thin (BMI 18.0 kg/m²) but were relatively more adipose (body fat 21.1%). Their diet was mostly vegetarian. Between 18 and 28 weeks gestation, fasting glucose concentrations remained stable, whereas total cholesterol and triglyceride concentrations increased and HDL-cholesterol concentrations decreased. The mean birth weight of the offspring was 2666 g. Total cholesterol and triglycerides at both 18 and 28 weeks and plasma glucose only at 28 weeks were associated directly with birth size. One SD higher maternal fasting glucose, cholesterol, and triglyceride concentrations at 28 weeks were associated with 37, 54, and 36 g higher birth weights, respectively (P < 0.05 for all). HDL-cholesterol concentrations were unrelated to newborn measurements. The results were similar if preterm deliveries also were included in the analysis (total n = 700).

CONCLUSIONS—Our results suggest an influence of maternal lipids on neonatal size in addition to the well-established effect of glucose. Further research should be directed at defining the clinical relevance of these findings.
concentrations of folate and vitamin C gave birth to larger newborns. This highlighted the role of maternal micronutrients in fetal growth. In this article, we report the relationship between maternal circulating fuels (glucose, total and HDL-cholesterol, and triglycerides) and neonatal size and compare the relative effects of glucose and lipids.

**RESEARCH DESIGN AND METHODS**—The design and methods of the PMNS have been described previously (4). In brief, we identified 2,675 married, nonpregnant women in six villages near Pune for possible enrollment in the study; 2,466 consented to take part (Fig. 1). Socioeconomic status (SES) was assessed using a standardized questionnaire (24), and a higher score represents higher status. Anthropometry (height, weight, circumferences, and skinfolds) was recorded every 3 months, and fat mass was calculated using four skinfolds (Durnin formula) (25). The women’s menstrual dates were recorded every month, and women missing two successive periods underwent an ultrasound examination to confirm pregnancy and assess gestational age (26). Pregnant women (n = 1,102) were identified and of these, women with a singleton pregnancy of <21 weeks’ gestation (n = 797) entered the study. Enrollment for the study started in September 1993, the first pregnancy was enrolled in June 1994, and the last delivery occurred in April 1996. Ethical permission for the study was granted by the KEM Hospital Ethics Committee.

**Maternal measurements during pregnancy**

At 18 ± 2 and 28 ± 2 weeks’ gestation the following information was collected: anthropometry, dietary intakes using a semi-weighed 24-h recall method and food frequency questionnaire (4), and physical activity (the conventional 24-h recall method was modified and made more objective by incorporating information on portion sizes, most of which were weighed at mealtimes by a trained field worker). Physical activity was assessed using a structured questionnaire to record the women’s daily workload, which mainly included farming and domestic activities (27). A weighted score was generated and was used in analysis; a higher score indicates higher physical activity. At 18 weeks’ gestation a fasting venous blood sample was collected. At 28 weeks, fasting and 2-h venous blood samples during an oral glucose tolerance test (OGTT; 75-g anhydrous glucose load) were collected. The following measurements were made using the fasting blood samples and standard enzymatic kits at both time points: plasma glucose, total cholesterol, HDL-cholesterol, and triglycerides. Glucose tolerance was classified by the then-prevalent World Health Organization 1985 criteria (28) (diabetes mellitus: fasting plasma glucose ≥7.77 mmol/L and/or 2-h plasma glucose ≥11.1 mmol/L; impaired glucose tolerance [IGT]: fasting plasma glucose <7.77 mmol/L and 2-h plasma glucose between 7.77 and 11.0 mmol/L).

**Neonatal anthropometry**

Babies were measured by one of five trained fieldworkers within 72 h of birth. Birth weight was measured to the nearest 50 g using a Salter spring balance (Salter Abbey, Suffolk, U.K.); crown-heel length was measured to the nearest 0.1 cm using a portable Pedobaby baby meter (ETS JMB, Brussels, Belgium). Ponderal index (PI) was calculated using the formula: PI = [birth weight (g) × 100 / (crown heel length (cm))^3]. Triceps and subscapular skinfold thicknesses were measured on the left side of the body to

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**Figure 1**—A flow diagram describing data collection and exclusions from the Pune Maternal Nutrition Study. LMP, last menstrual period; FFQ, food frequency questionnaire.
the nearest 0.2 mm using Harpenden skinfold calipers (CMS Instruments, London, U.K.). Occipito-frontal head circumference, mid-upper arm circumference (MUAC), and abdominal circumference (at the level of the umbilicus during expiration) were measured to the nearest 0.1 cm using fiberglass tapes (CMS Instruments). Placental weight was recorded to the nearest 5 g using Ishida scales after trimming the umbilical cord. Interobserver and intraobserver variation studies were conducted every 3 months to ensure the quality of these measurements.

Statistical methods
Data in tables are presented as mean ± SD. For statistical analysis, variables with skewed distributions (subscapular and triceps skinfolds) were log-transformed to satisfy assumptions of normality. Relationshps between maternal size before pregnancy and maternal fuels (glucose and lipids) during pregnancy, as well as the interrelationships between maternal fuels, were tested using Pearson correlation coefficients adjusting for SES, parity, and maternal age. We performed regression analyses to study the univariate associations of Z-standardized maternal plasma glucose and lipid concentrations with neonatal measurements, adjusting for gestation at the time of measurements, sex, SES, parity, maternal age, and maternal BMI before pregnancy. We show the effect of a 1-SD change in each maternal “fuel” on the birth size measurement in original units. Finally, we constructed multivariate models in which glucose and lipids were included together to examine independent associations. Analyses were carried out using STATA version 11.2 (Stata Corp, College Station, TX).

RESULTS

Measurements before pregnancy
The characteristics of the mothers are shown in Table 1. These rural mothers were young, short, and thin but had relatively high adiposity (body fat %). None of them smoked tobacco or drank alcohol, but one-fourth used non-smoked tobacco (for chewing or as a tooth powder). The majority of women (68%) belonged to subsistence farming families. One-third of the women were lacto-vegetarian and only 15% of women ate nonvegetarian foods more than once every alternate day. The portion sizes of nonvegetarian foods were small (<120 g/day for chicken, fish, and meat dishes and ~60 g/day for eggs).

Measurements during pregnancy
Mean maternal weight gain from before pregnancy was 2.0 kg (SD 2.8 kg) up to 18 weeks’ gestation and 5.6 (SD 2.9) up to 28 weeks’ gestation. In 221 women we also had a weight measurement near delivery (35.1 [SD 1.1] weeks’ gestation), and the mean weight gain was 7.7 kg (SD 3.2 kg).

Dietary intake and physical activity
Mean daily maternal energy and protein intakes at 18 and 28 weeks’ gestation were 7.4 and 7.0 MJ and 45.4 and 43.5 g,

Table 1—Maternal body size and biochemical characteristics

|                              | Before pregnancy (n = 631), mean (SD) | At 18 weeks | At 28 weeks |
|------------------------------|---------------------------------------|-------------|-------------|
|                              | N = 631 | Mean (SD) | N = 631 | Mean (SD) | N = 631 | Mean (SD) | P* |
| Age (years)                  | 21.4 (3.6) | — | — | — | — | — | — |
| Primiparous, n (%)           | 226 (35.8) | — | — | — | — | — | — |
| Weight (kg)                  | 41.7 (5.1) | 631 | 43.7 (5.0) | 592 | 47.3 (5.2) | 0.0001 | — |
| Height (cm)                  | 151.9 (5.1) | — | — | — | — | — | — |
| BMI (kg/m²)                  | 18.0 (1.9) | 631 | 18.9 (1.8) | 592 | 20.4 (1.9) | 0.0001 | — |
| Waist circumference (cm)     | 60.7 (5.6) | — | — | — | — | — | — |
| Waist-to-hip ratio           | 0.74 (0.06) | — | — | — | — | — | — |
| Fat percentage               | 21.1 (4.3) | — | — | — | — | — | — |
| Plasma glucose/lipids        | — | — | — | — | — | — | — |
| Fasting glucose (mmol/L)     | — | 611 | 3.96 (0.65) | 564 | 3.97 (0.64) | 0.894 | — |
| 2-h glucose (OGTT) (mmol/L)  | — | 609 | 4.11 (0.85) | 562 | 4.38 (1.08) | 0.0001 | — |
| Total cholesterol (mmol/L)   | — | 595 | 1.12 (0.28) | 554 | 1.08 (0.28) | 0.001 | — |
| HDL-cholesterol (mmol/L)     | — | 610 | 1.09 (0.36) | 562 | 1.51 (0.52) | 0.0001 | — |
| Triglyceride (mmol/L)        | — | — | — | — | — | — | — |
| Intake and physical activity | — | 624 | 7.4 (2.1) | 607 | 7.0 (2.0) | 0.0001 | — |
| Total energy (MJ)            | — | 624 | 317.4 (89.3) | 607 | 302.8 (87.7) | 0.0001 | — |
| Total carbohydrate (g)       | — | 624 | 34.8 (14.7) | 607 | 32.4 (13.9) | 0.001 | — |
| Total fat (g)                | — | 624 | 45.4 (14.0) | 607 | 43.5 (13.5) | 0.002 | — |
| Total protein (g)            | — | 624 | 73.5 (25.7) | 608 | 65.4 (25.9) | 0.0001 | — |

*P values show the difference between 18 and 28 weeks and were derived by paired t tests, performed using the maximum number of paired samples available.
respectively (4). The difference of 5.7% in the energy intake was statistically significant. These values were lower compared with the recommended daily allowances for Indian pregnant women given by the Indian Council of Medical Research at that time (29). Carbohydrates were the main energy source (72%), whereas 10 and 18% of energy was derived from protein and fat, respectively. These women had relatively high levels of physical activity; a woman with an average physical activity score (73.5) spent more than 8 h performing domestic activities that included cooking, washing clothes and utensils, sweeping the house, and fetching water and firewood (27). Two-thirds of women did farm work for more than 4 h/day.

Glycemia
Mean fasting plasma glucose concentrations were similar at 18 and 28 weeks (Table 1). At 28 weeks, OGTT results were available for 492 women. Using World Health Organization 1985 criteria (28), there were two women with impaired glucose tolerance and one with diabetes. They were given dietary advice and their subsequent glucose concentrations were normal; none required antidiabetic medication.

Lipids
Between 18 and 28 weeks’ gestation, plasma total cholesterol concentrations increased by 16.9% and triglyceride concentrations by 38.4%, whereas HDL-cholesterol concentrations decreased by 3.9%. Using the National Cholesterol Education Program criteria in the nonpregnant state (30), 10% of women had hypercholesterolemia (≥5.18 mmol/L) at 18 weeks’ and 34% at 28 weeks’ gestation, whereas 6% had hypertriglyceridemia (≥1.69 mmol/L) at 18 weeks’ and 31% at 28 weeks’ gestation. Low HDL-cholesterol concentrations (<1.29 mmol/L) were present in 74% of women at 18 weeks’ and 77% at 28 weeks’ gestation.

Interrelationships between glucose and lipids
Fasting plasma glucose concentrations were not significantly associated with lipid concentrations at 18 and 28 weeks. At 28 weeks’ gestation, 2-h plasma glucose was directly related to total cholesterol and triglyceride concentrations ($r = 0.34$ and $0.31$, respectively; $P < 0.001$ for both gestations).

Table 2—Characteristics of full-term babies (N = 631)

| Characteristics         | Mean (SD)   |
|-------------------------|-------------|
| Gestational age (weeks) | 39.4 (1.7)  |
| Birth weight (g)        | 2,666 (355) |
| Crown-heel length (cm)  | 47.7 (1.9)  |
| Ponderal index (kg/m²)  | 2.4 (0.2)   |
| Skinfolds               |             |
| Subscapular (mm)        | 4.2 (0.9)   |
| Triceps (mm)            | 4.2 (0.8)   |
| Circumferences          |             |
| Head (cm)               | 33.1 (1.2)  |
| Abdomen (cm)            | 28.6 (1.9)  |
| Mid-upper arm (mm)      | 9.7 (0.9)   |
| Placental weight (g)    | 360.3 (76.4) |

Interrelationships between maternal weight, food intake, physical activity, and circulating glucose and lipids
Maternal BMI before pregnancy was inversely related to HDL-cholesterol concentrations ($r = -0.11$; $P < 0.01$ for both gestations) and directly related to triglycerides ($r = 0.11$; $P = 0.008$ at 28 weeks’ gestation). Waist circumference showed similar relationships as maternal BMI. Weight gain at 28 weeks’ gestation was directly related to plasma total cholesterol ($r = 0.18$; $P = 0.001$) and triglyceride concentrations ($r = 0.11$; $P = 0.006$). Macronutrient intakes were not associated with any fuels. Physical activity at 28 weeks was inversely associated with 2-h glucose ($r = -0.14$; $P = 0.002$) and total cholesterol ($r = -0.12$; $P = 0.005$) concentrations. All these associations were independent of maternal age, SES, and parity.

Birth measurements
Mean birth weight was 2,666 g, mean length was 47.7 cm, and mean PI was 2.4 kg/m² (Table 2). Twenty-eight percent of babies had a low birth weight (<2.5 kg). Mean placental weight was low at 360 g.

Relationships of maternal weight, glucose, and lipids during pregnancy with neonatal size
Weight at different time points during pregnancy was directly associated with all newborn measurements ($r = 0.08–0.35$; $P < 0.05$ for all). Weight gain up to 18 weeks was unrelated to any of the newborn measurements. Weight gain up to 28 weeks was directly related to all newborn measurements with the exception of placental weight ($r = 0.09–0.16$; $P < 0.05$ for all). The mean gestation was 39.4 (SD 1.7) weeks, and maternal fuels were unrelated to duration of pregnancy.

At 18 weeks’ gestation, plasma glucose and HDL-cholesterol concentrations were unrelated to newborn size. Maternal total cholesterol concentration was directly associated with birth weight, abdominal circumference and MUAC, and placental weight; a 1-SD-higher total cholesterol was associated with a 39-g-higher birth weight. Maternal triglyceride concentration was directly associated with abdominal circumference and placental weight (Supplementary Fig. 1).

At 28 weeks’ gestation, fasting plasma glucose concentration was directly associated with birth weight and MUAC; a 1-SD-higher fasting glucose concentration was associated with a 37-g-higher birth weight. Two-hour plasma glucose concentration was directly associated with abdominal circumference and MUAC. Plasma cholesterol concentration was directly associated with all newborn measurements except head circumference; a 1-SD-higher maternal cholesterol concentration was associated with a 54-g-higher birth weight. Maternal triglyceride concentration was directly associated with birth weight, birth length, skinfold thicknesses, and abdominal circumference; a 1-SD-higher maternal triglyceride concentration was associated with a 36-g-higher birth weight.

Although in general the effects of 28-week maternal measurements seemed to be stronger than those of 18-week measurements (Table 3), and although the strongest effects seemed to be with cholesterol, overlapping CIs indicate that the associations were broadly similar in magnitude at both gestational time points and that the associations of glucose, cholesterol, and triglycerides were of similar strength.

Multivariate analysis
In a multivariate model with fasting glucose, total and HDL-cholesterol, and triglyceride concentrations at 18 weeks, maternal total cholesterol concentration was directly associated with birth weight, abdominal circumference, and MUAC (Table 4). There were no independent
## Table 3—Univariate analyses of maternal glucose and lipids at 18 and 28 weeks' gestation as predictors of neonatal size

|                | Birth weight (g) | Length (cm) | SS + TR (mm) | Head circumference (cm) | Abdominal circumference (cm) | Mid-upper-arm circumference (cm) | Placental weight (g) |
|----------------|-----------------|-------------|--------------|-------------------------|-----------------------------|---------------------------------|---------------------|
| **18 Weeks**   |                 |             |              |                         |                             |                                 |                     |
| Fasting glucose | 1.66 (−29.76 to 29.96) | 0.02 (−0.14 to 0.18) | 0.027 (−0.11 to 0.14) | −0.07 (−0.19 to 0.05) | −0.11 (−0.27 to 0.05) | −0.05 (−0.12 to 0.02) | 2.66 (−3.73 to 9.05) |
| Cholesterol    | 39.07 (10.57–67.58) | 0.11 (−0.05 to 0.28) | 0.06 (−0.07 to 0.19) | 0.08 (−0.04 to 0.20) | 0.29 (0.13–0.45) | 0.11 (0.04–0.19) | 6.64 (0.19–13.09) |
| HDL-cholesterol| 17.57 (−11.64 to 46.77) | 0.09 (−0.07 to 0.26) | 0.0001 (−0.13 to 0.13) | 0.12 (−0.001 to 0.25) | 0.14 (−0.03 to 0.30) | 0.07 (−0.01 to 0.14) | 2.07 (−4.45 to 8.64) |
| Triglyceride   | 14.76 (−13.34 to 42.86) | 0.03 (−0.13 to 0.19) | −0.02 (−0.14 to 0.11) | 0.01 (−0.11 to 0.13) | 0.18 (0.03–0.34) | 0.05 (−0.03 to 0.12) | 6.44 (0.14–12.75) |
| **28 Weeks**   |                 |             |              |                         |                             |                                 |                     |
| Fasting glucose | 37.25 (6.94–67.56) | 0.12 (−0.06 to 0.29) | 0.02 (−0.12 to 0.15) | −0.05 (−0.17 to 0.08) | 0.07 (−0.10 to 0.24) | 0.09 (0.02–0.17) | 5.38 (−1.57 to 12.33) |
| 2-h glucose    | 25.02 (−8.54 to 58.58) | 0.11 (−0.09 to 0.30) | 0.09 (−0.05 to 0.25) | 0.03 (−0.12 to 0.17) | 0.24 (0.05–0.42) | 0.12 (0.04–0.21) | 4.69 (−2.89 to 12.26) |
| Cholesterol    | 54.34 (24.85–83.88) | 0.21 (0.04–0.38) | 0.15 (0.02–0.28) | 0.04 (−0.08 to 0.17) | 0.29 (0.13–0.46) | 0.12 (0.04–0.19) | 9.39 (2.64–16.14) |
| HDL-cholesterol| −8.89 (−38.72 to 20.95) | −0.05 (−0.22 to 0.12) | −0.03 (−0.16 to 0.10) | 0.02 (−0.11 to 0.14) | 0.03 (−0.05 to 0.10) | −0.40 (−0.76 to 6.45) |                     |
| Triglyceride   | 36.27 (4.32–68.23) | 0.24 (0.06–0.42) | 0.15 (0.01–0.29) | 0.08 (−0.06 to 0.21) | 0.20 (0.02–0.38) | 0.07 (−0.01 to 0.16) | 5.22 (−2.01 to 12.52) |

Glucose and lipids at 18 and 28 weeks are expressed as SDs. Values are regression coefficients (β) (95% CI), representing the unit change in neonatal body size per 1 SD change in maternal fuels. All models were adjusted for gestation, sex of the baby, parity, SES, and maternal age, BMI before pregnancy, and total energy intake at the time of measurements.
All maternal fuels are entered together in each model. Glucose and lipids at 18 and 28 weeks are expressed as SDs. Values are regression coefficients (95% CI) representing the unit change in neonatal size per 1 SD change in maternal fuels. All models were adjusted for gestation, sex of the baby, parity, SES, and maternal age, BMI before pregnancy, and total energy intake at the time of measurements.

Table 4: Multivariate analyses of maternal glucose and lipids at 18 and 28 weeks as predictors of neonatal size

| Week/Model | BMI (kg/m²) | Head (cm) | Abdominal (cm) | Hilp (cm) | Weight (g) | Length (cm) | 50-75th (mm) |
|------------|-------------|-----------|----------------|----------|------------|-------------|-------------|
| 18 Weeks model 1 | 0.74 (0.18) | 10.77 (2.68) | 0.12 (0.02) | 2.24 (0.04) | 0.06 (0.01) | 0.09 (0.03) | 0.09 (0.03) |
| 28 Weeks model 1 | 0.74 (0.18) | 10.77 (2.68) | 0.12 (0.02) | 2.24 (0.04) | 0.06 (0.01) | 0.09 (0.03) | 0.09 (0.03) |
| 18 Weeks model 2 | 0.74 (0.18) | 10.77 (2.68) | 0.12 (0.02) | 2.24 (0.04) | 0.06 (0.01) | 0.09 (0.03) | 0.09 (0.03) |
| 28 Weeks model 2 | 0.74 (0.18) | 10.77 (2.68) | 0.12 (0.02) | 2.24 (0.04) | 0.06 (0.01) | 0.09 (0.03) | 0.09 (0.03) |

Note: The size of the effect of all placental weight was predicted by plasma glucose at 18 weeks (18) and triglycerides (18 weeks) but not other maternal fuels.
Maternal metabolism and neonatal size

maternal triglycerides to be a stronger correlate of fetal size and body fat than maternal BMI, glucose, and insulin (18,38). HDL-cholesterol has been inversely related to birth weight in overweight and obese women but not in normal-weight women (34).

The majority of these studies were of well-nourished women in high-income countries with nonvegetarian food habits and predominantly diabetic pregnancies. In rural Indian mothers with very low BMI, fasting glucose concentration remained similar but total cholesterol and triglyceride concentrations rose by 16.9% and 38.4%, respectively, and they were as strong predictors of neonatal size as glucose concentrations, if not stronger. Thus, the pattern of associations in Indian undernourished rural women might indicate metabolic adjustments to maintain fetal growth. The placenta transfers glucose from mother to baby by both passive and facilitated diffusion (39). Cholesterol is directly transferred, although the mechanisms are poorly understood (15), while triglycerides are broken down into fatty acids and glycerol, transported across the placenta, and reconstituted before transfer to the fetus (7,40). Glucose and fatty acids are used for energy production by the placenta and the fetus. Cholesterol and fatty acids are used as structural elements of cell membranes and contribute to synthesis of hormones and other messengers by the fetus and placenta. Low concentrations of arachidonic acid precursors and high concentrations of trans fatty acids in both early and late pregnancy have been associated with restriction of fetal growth (41,42), thought to be especially important in high-risk pregnancies, such as adolescent pregnancies (43).

Thus, maternal fuels contribute to both structural and functional requirements of fetal growth, and our findings highlight the importance of maternal lipids in this relationship. Humans have the highest body fat percentage (15%) at birth of all mammals, including sea lions (5%) and pigs (~2%), suggesting an evolutionary advantage of adiposity during the perinatal period (44); it contributes to energy stores, thermal insulation, and even attracting the mothers to feed (45). We have shown that these rural Indian babies, despite their low weights, are relatively more adipose compared with English babies (46). The relatively high adiposity of our rural mothers despite a low BMI also suggests an advantage of the “thin-fat” phenotype in reproduction. On the other hand, higher triglyceride concentrations during early pregnancy increase the risk of gestational diabetes and postpartum diabetes (40,47-48), and in an obese or diabetic pregnancy, maternal dyslipidemia contributes to fetal macrosomia. This suggests a continuum and the need for further research to decide on treatment strategies to achieve a balance between fetal growth and morbidity. Maternal hypercholesterolemia increases the risk of early atherogenesis in the fetus (fatty streaks in the aorta) both in animals and human beings (16,23). There is no information on the effect of treating these severe lipid abnormalities on the growth and future health of the fetus.

Ours is the first study relating maternal lipid concentrations to newborn size in normoglycemic, undernourished, rural Indian women with relatively low fat and cholesterol intakes and contributes a piece to solving a complex puzzle. The women were representative of the village population. Measurements of glucose and lipids were made twice, during mid- and late pregnancy, and included an OGTT in late pregnancy in 492 women. We also collected high-quality data on maternal food intake and physical activity and detailed newborn anthropometry, allowing us to construct multivariate models to examine the association between maternal fuels and newborn size and allowing for potential confounding factors. However, this is an observational study and therefore we cannot be sure of the causality of associations. We did not measure circulating fatty acids because of insufficient stored samples, and therefore we are unable to comment on the effect of individual fatty acids.

In summary, we demonstrate in rural, undernourished, normoglycemic Indian pregnant women a significant association between maternal circulating lipids and fetal growth, which was at least as strong as that of glucose. It will be important to study these associations in other populations with different rates of obesity and glycemic levels, for example, in the HAPO study. A study of future maternal and fetal risk associated with pregnancy lipids will help understand the importance of our findings over the life course. Such a study is now in progress.

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S.R.K. performed statistical analysis and wrote the manuscript. K.K. wrote and edited the manuscript. S.R.R. collected data and edited the manuscript. S.D.C., T.M.D., A.J.B., V.A.S., and D.S.B. collected data. C.H.D.F. and C.S.Y. planned the research, guided the analysis, and wrote and edited the manuscript. C.H.D.F. and C.S.Y. are the guarantors of this work and, as such, had full access to all the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis.

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