Hypoxia inducible factor 2α (HIF2α/EPAS1) is associated with development of pulmonary hypertension in severe congenital diaphragmatic hernia patients

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Abstract
We show that hypoxia inducible factor 2α (HIF2α) is highly expressed in patients with pulmonary hypertension (PH). HIF2α is expressed in every patient with congenital diaphragmatic hernia, while only half of the controls express HIF2α. Our data suggest that HIF2α is a link between hypoxia and the development of PH.

Keywords
hypoxia inducible factor, pulmonary hypertension, pulmonary vascular development and epithelium

Introduction
Idiopathic pulmonary hypertension in newborns (PHN) is a life-threatening condition and requires intensive clinical support.1,2 PHN is frequently associated with congenital disorders, such as congenital heart diseases and congenital diaphragmatic hernia (CDH).3 The underlying causes of PHN are still largely unknown, although several potential genetic and epigenetic mechanisms have been described.2,3 Prolonged exposure to hypoxia causes pulmonary arterial hypertension (PAH), and hypoxia inducible factors (HIF) are the key component of the cellular response to hypoxia.4 HIFs are heterodimeric transcription factors composed of two subunits, a stable HIF1β and one of three oxygen-sensitive subunits HIF1α, HIF2α, or HIF3α.4

HIFs have been associated with lung development and PH.5 In humans, genome-wide studies showed a positive correlation of HIF2α with adaptation to hypobaric hypoxia in Tibetan highlanders.6,7 In rodents, both Hif1α and Hif2α appeared to modulate hypoxia-induced PH in gene ablation studies.5–12 Two reports described that endothelial-specific ablation of prolyl-4-hydroxylase 2 (PHD2), which targets HIFs for degradation by hydroxylating proline residues in HIF2α under normoxic conditions, results in a HIF2α-dependent adult PAH.10,11 Using an endothelial-specific inactivation of HIF2α, Cowburn et al. showed that HIF2α is involved in hypoxia-induced PAH.12 Since PAH is different from PHN, we examined HIF2α protein expression in a developmental series of normal lung tissue, as well as in the lungs of a cohort of CDH patients and patients with idiopathic PH.

Materials and methods
Human lung tissue collection
With the approval of the Erasmus MC Medical Ethical Committee and the informed consent of parents, lung...
tissue was obtained from the archives of the Department of Pathology, Erasmus MC (Rotterdam).

Immunohistochemistry

Tissue micro arrays were constructed as described. Paraffin embedded lung material was sectioned, blocked, and incubated overnight at 4°C (HIF2α, Genetex). The Envision kit (Dako) was used and images were taken with a charge-coupled device camera attached to an Olympus BX41 microscope.

Results

HIF2α was expressed in half of the fetal lungs that were analyzed at 15–36 weeks of gestation. In addition, lungs of neonates born after week 37 were also positive for HIF2α (Fig. 1a). HIF2α was localized in the nucleus of proximal and distal airway epithelial cells and alveolar epithelial cells (Fig. 1a, arrowheads). HIF2α is also highly expressed in alveolar type II cells in lungs as we and others have shown before in mouse lungs.9,14

Next, we analyzed the expression of HIF2α in lungs from a cohort of CDH patients of different gestational ages until...
The HIF2α is prominently present in all CDH cases analyzed, contrasting the expression of HIF2α in normal, unaffected lungs (Fig. 1a, Table 1). The sites of HIF2α expression were in both alveolar epithelial cells (Fig. 1a, arrowheads) and all endothelial cells of the blood vessels (Fig. 1a, arrows). Moreover, in some CDH lungs, the level of HIF2α appeared very high as indicated by the intense staining (Fig. 1).

We also analyzed 11 lungs of neonates with idiopathic PH and age-matched controls (Fig. 1b). In the neonatal control human lung, only few cells are positive for HIF2α (Fig. 1b, control). Clear staining patterns were observed in the endothelial cells of the vessels of patients with PH (Fig. 1b, arrows) and in alveolar epithelial cells (Fig. 1b, arrowheads), although not all PHN cases expressed HIF2α (Fig. 1b). Our data suggest that expression of HIF2α is maintained in some of the clinical cases of PH, supporting the relative hypoxia of these patients. Finally, we also analyzed the expression of one of the HIF2α targets, VEGF, and found comparable numbers of positive samples as for the HIF2α staining (Table 1).

### Discussion

Previously, we showed a gradual increase of HIF2α messenger RNA (mRNA) during gestation, but no differences were detected in expression levels between CDH patients and controls.15,16 Here, we report an increased expression of HIF2α protein in CDH patients compared to controls. Even before birth, we found significantly higher expression of HIF2α in CDH patients. Our data suggest that high levels of HIF2α correlate with PHN and CDH-associated PH. Since prolonged exposure to hypoxia results in PH,5 and the lungs of patients with PH are under constant hypoxic conditions, it may result in elevated levels of HIF2α. Although all CDH cases tested were clearly positive for HIF2α, not all cases of PHN expressed high levels of HIF2α. This most likely reflects the patient variability and the heterogeneity of PHN.

In all fetal CDH cases, HIF2α was expressed in the lung and at much higher levels than controls. Moreover, HIF2α is also highly expressed in the lungs of patients suffering from PHN. This suggests that early in gestation, the lungs of CDH patients are already intrinsically different from control lungs, and high levels of HIF2α may contribute to postnatal PH, which develops in a significant number of patients with CDH after birth.

Prolonged hypoxia, which is also observed in PHN patients, results in structural changes of the pulmonary vasculature characterized by a thickening of the vascular wall of small pulmonary arteries leading to an increased vascular resistance and a worsening of gas exchange.2 We recently showed that the vascular smooth muscle cells in the developing CDH lung had a different expression pattern of contractile components compared to control lungs, suggesting that these cells prematurely differentiate.17 Furthermore, the expression of VEGF-A mRNA was increased in the lungs from CDH patients at the canalicular stage, while a significant decrease in the expression of VEGF-A mRNA was observed in the alveolar stage of lung development in CDH patients. However, the spatial distribution of VEGF-A was not different between control and CDH lungs.16,18 Here, we report that HIF2α is expressed in more CDH lungs than in control lungs, and that the expression of VEGF appears to correlate with HIF2α expression. The site of expression does not differ between control and CDH lungs, as previously described. Additionally, we previously showed that a significant increase in Hif2α in epithelial alveolar type II cells did not induce an increased expression of Vegf-A.14

Although the mechanisms of HIF2α activity and its role in the development of PH are still incompletely understood, it may be a putative target for future therapies. In this respect, it is interesting to investigate the potential of the Hif2α-specific competitor, FM19G11.19

In summary, we showed high expression levels of HIF2α in the lungs of all CDH patients at different gestational ages and after birth, while only half of the age-matched controls showed expression of HIF2α. From these results, we suggest that HIF2α is associated with the development of PH in CDH, and possibly also with PHN.

### Declaration of conflicting interests

The author(s) declare that there is no conflict of interest.

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| Table 1. CDH patient characteristics and the age-matched control lungs. |
|------------------|----------------|----------------|----------------|----------------|----------------|
| Developmental stage | Gestational age (weeks) | Postnatal age | Birth weight (g) | HIF2α-positive (n/total n) | VEGF-positive (n/total n) |
|-------------------|-------------------|---------------|-----------------|----------------------|----------------------|
| Control           | Premature         | 31.5 (15–36)  | 1 h (0–24 h)    | 2000 (57–2700)       | 5/9                   |
|                   | Term              | 39.5 (38–41)  | 36 h (1 h–1 week)| 3220 (2490–3950)     | 4/8                   |
| CDH               | Premature         | 31.5 (15–36)  | 1 h (0–48 h)    | 1032 (30–2515)       | 7/7                   |
|                   | Term              | 39 (37–40)    | 7 h (1 h–3 days)| 2835 (2000–3800)     | 6/6                   |

Indicated are the average age in weeks, the average postnatal age, birth weight, and the number of HIF2α-positive and VEGF-positive samples per total samples tested.
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