THE STUDY OF THE VOLATILE OILS COMPOSITION OBTAINED FROM VEGETATIVE AND GENERATIVE ORGANS OF BALLOTA NIGRA L.

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Key words: black horehound; Ballota nigra; gas chromatography-mass spectrometry (GC-MS); volatile oil component composition

The article presents the data of the pharmacognostic study of the aerial part of black horehound (Ballota nigra L.). The aim of the work is to determine the qualitative composition and the quantitative content of the volatile components from vegetative and generative organs of black horehound. Leaves, flowers and stems of black horehound were harvested at the flowering phase in the Kharkiv region. For the first time volatile compounds of black horehound aerial parts have been investigated by GC/MS. The component composition of volatile substances in horehound herb has been studied using an Agilent Technologies 6890N chromatograph with a mass spectrometry detector 5973N. The components have been identified according to mass spectra NIST 05 and WILEY 2007 libraries together with the programmes for identifying AMDIS and NIST. In vegetative and generative organs of horehound 48 components have been identified. Germacrene D (40.2 mg/kg), hexahydrofarnesylacetone (167.42 mg/kg), 2,6-trimethyl-4-methylene-2H-pyran (172.3 mg/kg), myristic (271 3 mg/kg), pentadecanoic (182.1 mg/kg), palmitoleic (306.4 mg/kg) and palmitic (1620.6 mg/kg) acids prevail in vegetative organs. Farnesylacetone (68.8 mg/kg), verbenone (35.7 mg/kg), myristic (187.5 mg/kg), palmitic acid (656.5 mg/kg) and palmitoleic (196.9 mg/kg) acids are the major components in generative organs. Therefore, hexahydrofarnesylacetone, palmitic and palmitoleic acids, and 2,6-trimethyl-4-methylene-2H-pyran are predominant components in the raw material studied.

Materials and Methods

The analysis of the essential oil of black horehound leaves and flowers was performed using a chromatograph Agilent Technology HP6890 GC with a mass spectrometric detector 5973N.

A weighed portion of the crushed raw material (1.0 g) was placed in a 20 ml “Agilent” vial, to which the internal standard (50 μg of tridecane) and 10 ml of water were added. Volatile components of the sample were steamed for 2 hours using a reflux condenser with air cooling. Substances adsorbed on the inner surface of the reflux after cooling of the system were washed into a dry 10 ml vial by slow adding of 3 ml of ultrapure pentane. The washes were concentrated by blowing (100 ml/min) with ultrapure nitrogen until the residual volume of the extract was 10 μl, then it was completely withdrawn by a chromatographic syringe. The further concentration of the sample to the volume of 2 ml was carried out in the syringe. The injection of the sample was performed in a splitless mode; it allowed to introduce the sample without division loss and significantly (10-20 fold) increase sensitivity of the chromatographic process. The input sample rate was 1.2 ml/min for 0.2 minutes. The chromatographic conditions were as follows: capillary chromatographic column DB-5 (30 m×0.25 mm); the carrier gas – helium, the carrier gas speed – 1.2 ml/min; the thermostat temperature – from 50 to 320°C at the rate of 4 °/min; the heater temperature of the sample introduction – 250°C. From the aqueous extract vola-
The component composition of vegetative and generative organs of black horehound

| Retention time, min | Compound                              | Corollas, mg/kg | Calyx, mg/kg | Leaves, mg/kg | Stems, mg/kg |
|---------------------|---------------------------------------|-----------------|--------------|---------------|--------------|
| 5.31                | Benzaldehyde                          | 3.9             | –            | –             | 5.1          |
| 6.26                | 1-Octen-3-ol                          | 3.5             | –            | –             | 10.4         |
| 6.54                | 2,3,5-Trimethylpyrazine                | –               | –            | –             | 7.9          |
| 6.68                | Octanal                               | –               | 10.9         | –             | –            |
| 7.4                 | Phenylacetaldehyde                    | 9.3             | –            | 31.3          | –            |
| 7.49                | Benzyl alcohol                        | –               | –            | –             | 9.8          |
| 7.79                | Limonene                              | –               | 10.3         | –             | –            |
| 9.01                | Tetramethylpyrazine                   | –               | –            | –             | 26.5         |
| 9.17                | 2,5-Dimethyl-3-acetylfuran            | 5.1             | –            | –             | –            |
| 9.69                | Nonanal                               | –               | 21.5         | –             | –            |
| 9.74                | Linalool                              | 4.9             | –            | –             | –            |
| 10.04               | 2,2,6-Trimethyl-4-methylene-2H-pyran  | 172.3           | –            | 14.9          | 42.5         |
| 11.07               | Verbenol                              | –               | –            | –             | 7.1          |
| 12.34               | Methylsalicilate                      | 4.6             | –            | –             | 313          |
| 12.49               | α-Terpineol                           | 4               | –            | –             | 6.2          |
| 12.64               | Verbenone                             | –               | –            | 35.7          | 3.2          |
| 12.99               | Decanal                               | 8.9             | 71           | 27.1          | –            |
| 14.59               | Isopiperitenone                       | –               | 8.7          | –             | –            |
| 14.84               | Ethylsalicilate                       | –               | –            | 7.6           | –            |
| 16.17               | 4-Vinyl-2-methoxyphenol               | –               | –            | –             | 36.1         |
| 16.31               | Dodecanal                             | –               | 519          | –             | –            |
| 17.58               | Eugenol                               | 26.3            | –            | 12.2          | 18.9         |
| 18.76               | Caprinic acid                         | 39              | –            | –             | –            |
| 18.98               | Methyleugenol                         | 26.3            | 39.6         | 12.8          | –            |
| 20.04               | β-Caryophyllene                       | 26.8            | –            | 2             | –            |
| 20.75               | Geranylacetone                        | 8.6             | 10.9         | 12.5          | 3.8          |
| 21.03               | Gumulene                              | 11.3            | –            | –             | 2.2          |
| 21.53               | β-Ionone-epoxyside                    | –               | –            | 36.4          | 5.4          |
| 21.63               | β-Ionone                              | –               | –            | 3.3           | –            |
| 21.83               | Germacrene D                          | 40.2            | –            | –             | 8.2          |
| 21.89               | Tetradecane                           | –               | 6.8          | –             | –            |
| 22.09               | Dihydroactinidiolide                  | –               | –            | 43.8          | –            |
| 22.94               | Pentadecane                           | –               | 53.9         | –             | –            |
| 23.47               | Elemicin                              | –               | 14           | –             | –            |
| 23.66               | Elemol                                | 18.5            | –            | –             | –            |
| 24.39               | Spatulenol                            | 15.4            | –            | –             | –            |
| 24.48               | Caryophyllene oxyde                   | 57.1            | –            | 6.8           | –            |
| 24.56               | Lauric acid                           | –               | 67.1         | –             | –            |
| 25.19               | Benzophenone                          | –               | 11.7         | –             | –            |
| 26.07               | Trans-isoelemic                      | –               | 67.1         | –             | –            |
| 28.6                | 11-Tetradecenic acid                  | –               | –            | –             | 4.9          |
| 29.05               | Miristic acid                         | 100.6           | 271.3        | 187.5         | 41.9         |
| 29.97               | Farnesole                             | –               | –            | 32.2          | –            |
| 30.27               | Hexahydrofarnesylacetone              | 167.4           | 60.3         | –             | –            |
| 30.62               | Pentadecanoic acid                    | 50.7            | 182.1        | 121.8         | –            |
| 31.19               | Farnesylacetone                       | –               | –            | 68.8          | –            |
| 31.71               | Palmitoleic acid                      | 44.5            | 306.4        | 196.9         | 34.7         |
| 32.2                | Palmitic acid                         | 573.9           | 1620.6       | 656.5         | 130.9        |
tile compounds were extracted with methylene chloride and further analyzed as described above.

Substances were identified by comparison of the mass spectra obtained with the data from mass spectra libraries NIST05 and WILEY 2007 in combination with the programmes for identification AMDIS and NIST. The content of components in the herbal drug (mg/kg) is given in Table.

**Results and Discussion**

Qualitative and quantitative compositions of essential oils of various organs of horehound are given in mg/kg (Table). As the result of the research 48 components of essential oil have been identified and quantified in generative and vegetative organs. Quantitatively sesquiterpenoids and substances of different origin (aldehydes, hydrocarbons, and fatty acids) are predominant, in smaller concentration monoterpenoids and aromatic compounds have been found. Germacrene D (40.2 mg/kg), hexahydrofarnesylacetone (167.42 mg/kg), 2,6-trimethyl-4-methylene-2H-pyran (172.3 mg/kg), myristic (271.3 mg/kg), pentadecanoic (182.1 mg/kg), palmitoleic (306.4 mg/kg) and palmitic (1620.6 mg/kg) acids prevail in vegetative organs. Farnesylacetone (68.8 mg/kg), verbene (35.7 mg/kg), myristic (187.5 mg/kg), palmitic acid (656.5 mg/kg) and palmitoleic (196.9 mg/kg) acids are the major components in generative organs.

**CONCLUSIONS**

1. The Chromato-mass spectrometric research of volatile substances of vegetative and generative organs of black horehound has been carried out; 48 components have been identified and quantified.

2. The predominant components in the raw material studied are hexahydrofarnesylacetone, palmitic acid, palmitoleic acid and 2,6-trimethyl-4-methylene-2H-pyran.

3. The results obtained will be used for further phytochemical research of these herbal drugs.

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ИССЛЕДОВАНИЕ СОСТАВА ЭФИРНОГО МАСЛА ВЕГЕТАТИВНЫХ И ГЕНЕРАТИВНЫХ ОРГАНОВ BALLOTA NIGRA L.
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Ключевые слова: Белокудренник черный; Ballota nigra L.; хромато-масс-спектрометрия; компонентный состав эфирного масла

Проведены данные фармакогностического изучения надземной части белокудренника черного (Ballota nigra L.). Целью работы было определение качественного состава и количественного содержания летучих компонентов вегетативных и генеративных органов белокудренника черного. В качестве сырья использовали листья, цветки и стебли, заготовленные в фазу цветения в Харьковской области. Впервые методом хромато-масс-спектromетрии проведено исследование летучих веществ надземной части белокудренника черного. Компонентный состав летучих веществ травы белокудренника исследовали на хроматографе Agilent Technologies 6890N с масс-спектрометрическим детектором 5973N. Компоненты идентифицировали с помощью библиотеки масс-спектров NIST 05 и WILEY 2007 совместно с программами для идентификации AMDIS и NIST. В вегетативных и генеративных органах белокудренника идентифицировано 48 компонентов. В вегетативных органах преобладают гермакрен D (40,2 мг/кг), гексагидрофарнезилациетон (167,4 мг/кг), 2,6-триметил-4-метилен-2Н-пиран (172,3 мг/кг), миристиновая (271,3 мг/кг), пентадекановая (182,1 мг/кг), пальмитолеиновая (306,4 мг/кг) и пальмитиновая кислоты (1620,6 мг/кг). В генеративных органах растения преобладают фарнезилациетон (68,8 мг/кг), вербенон (35,7 мг/кг), миристиновая (187,5 мг/кг), пальмитиновая (656,5 мг/кг) и пальмитолеиновая (196,9 мг/кг) кислоты. Доминирующими компонентами в исследуемом сырье являются гексагидрофарнезилациетон, пальмитиновая и пальмитолеиновая кислоты, а также 2,6-триметил-4-метилен-2Н-пиран.