Impacts of 1-Methylcyclopropene and Controlled Atmosphere Established during Conditioning on Development of Bitter Pit in ‘Honeycrisp’ Apples

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Abstract. ‘Honeycrisp’ apples are susceptible to develop the physiological disorder bitter pit. This disorder typically develops during storage, but preharvest lesion can also develop. ‘Honeycrisp’ is also chilling sensitive, and fruit is typically held at 10–20 °C after harvest for up to 7 days to reduce development of chilling injury (CI) during subsequent cold storage. This temperature conditioning period followed by a lower storage temperature (2–4 °C) reduces CI risk but can exacerbate bitter pit development. Bitter pit development can be impacted in other apple cultivars by the use of controlled atmosphere (CA) storage and/or 1-methylcyclopropene (1-MCP). Studies were conducted to evaluate efficacy of CA and/or 1-MCP to manage ‘Honeycrisp’ bitter pit development. Apples from multiple lots, obtained at commercial harvest, were held at 10 °C for 7 days and then cooled to 3 °C. Half the fruit was exposed to 42 μmol·L⁻¹ 1-MCP the day of receipt while held at 10 °C. Fruit were stored in air or CA (3 kPa O₂, 0.5 kPa CO₂ for 2 days, then 1.5 kPa O₂, 0.5 kPa CO₂) established after 1 day at 10 °C or after 7 days at 10 °C plus 2 days at 3 °C. Fruit treated with 1-MCP and/or stored in CA developed less bitter pit compared with untreated fruit stored in air, and bitter pit incidence was lowest for 1-MCP-treated fruit with CA established during conditioning. Development of diffusion flesh browning (DFB) and cavities, reported to occur during ‘Honeycrisp’ CA storage, was observed in some lots. Incidence of these disorders was not enhanced by establishing CA 2 days compared with 9 days after harvest. 1-MCP and CA slowed peel color change, loss of soluble solids content (SSC) and titratable acidity (TA), and reduced ethylene production and respiration rate. The results indicate potential for the postharvest management of bitter pit development in ‘Honeycrisp’ apple through use of 1-MCP and/or CA storage.

High consumer demand for ‘Honeycrisp’ apples has supported a favorable price structure for producers (Warner, 2012) despite production and postharvest challenges across production regions (Tong, 2016). One of the challenges is managing susceptibility to bitter pit (Bedford, 2001; Rosenberger et al., 2001). Bitter pit symptoms include brown, dry areas typically just below the peel, typically ranging in size from 1 to 5 mm with adjacent peel also brown. Fruit nutrient content (Rosenberger et al., 2004; Telias et al., 2006), fruit maturity at harvest (Prange et al., 2011), fruit size (Prange et al., 2011; Telias et al., 2006), and crop load (Telias et al., 2006) are factors that impact risk of bitter pit development. Preharvest calcium application (Biggs and Peck, 2015; Peryea et al., 2007; Rosenberger et al., 2004), crop load management (DeLong et al., 2006; Robinson and Lopez, 2012), and harvest maturity management (Prange et al., 2011) can contribute to reduced bitter pit development on the tree and during subsequent storage. Bitter pit symptoms are often absent at harvest and develop during the postharvest period in storage (Ferguson and Watkins, 1989; McAlpine, 1912). Storage in a low-oxygen CA can reduce bitter pit development (Hewett, 1984; Sharples, 1982; Webster and Forsyth, 1979) while impacts of postharvest 1-MCP application have been mixed (Calvo and Candan, 2010; Gogo et al., 2015; Mirzaee et al., 2015).

‘Honeycrisp’ is also chilling sensitive with both peel (soft scald) and cortex (soggy breakdown) tissues at risk of injury development during cold storage (Watkins et al., 2004). Chilling sensitivity is influenced by orchard environment before harvest (Lachapelle et al., 2013; Moran et al., 2009) and fruit maturity at harvest (Watkins et al., 2005). CI risk can be reduced by preharvest application of 1-MCP (DeEll and Elshani-Moghaddam, 2010) and postharvest temperature conditioning (DeLong et al., 2006; Moran et al., 2009; Watkins et al., 2004).

CA storage of ‘Honeycrisp’ can provide some reduction in the rate of fruit ripening (Contreras et al., 2014; DeEll et al., 2015; DeLong et al., 2006; Watkins and Nock, 2012) as can postharvest application of 1-MCP (Contreras et al., 2014; DeEll, 2010; DeEll et al., 2015; Watkins and Nock, 2012). Disorder development is reduced with increased prestorage conditioning temperature, with prestorage diphenylamine application (Contreras et al., 2014), or by delayed establishment of CA conditions (DeEll et al., 2016). However, delaying CA storage can enhance development of other disorders including bitter pit and peel blotch (DeEll et al., 2016). Postharvest 1-MCP application can also enhance the development of CO₂ injury for fruit stored in CA (Lum et al., 2016; Watkins and Nock, 2012).

Although ‘Honeycrisp’ is susceptible to postharvest disorders that are exacerbated by CA, few incidents of CA injury have been reported in the U.S. Pacific Northwest (I. Hanrahan and J.P. Mattheis, personal communication) where 1-MCP and CA are widely used for ‘Honeycrisp’. As an exploration of the potential for postharvest CA and 1-MCP protocols to address bitter pit, the objective of this study was to assess impacts of 1-MCP and CA imposed during or after temperature conditioning of ‘Honeycrisp’ on bitter pit development and fruit quality.

Materials and Methods

Plant material. ‘Honeycrisp’ (Malus ×domestica Borkh.) apples were obtained from five commercial orchards in central Washington State over three harvest seasons (2013–15). Apples were commercially mature as determined by the grower and were obtained in bulk bins the day of harvest. Apples absent of external defects were placed onto pressed fiber trays and the trays placed into cardboard boxes. Packaged boxes were moved into one of two storage rooms at 10 °C, one for controls, the other for 1-MCP treatment. Fruit were exposed to 42 μmol·L⁻¹ 1-MCP (AgroFresh, Inc., Spring House, PA) in an 800-L metal cabinet for
16 h. The chamber was opened at the end of the treatment period, and 4 h later, boxes were removed from the treatment chamber and transferred to the other 10 °C storage room. All fruit was held for 7 d at 10 °C and then the room temperature was reduced to 3 °C. For CA storage, fruit on trays were placed into 0.14-m³ chambers and a CA of 3 kPa O₂ and 0.5 kPa CO₂ was established 24 h after receipt (CA 1 d) or after an additional 8 d (CA 9 d) consisting of 6 d at 10 °C then 2 d at 3 °C. Two days after CA establishment, O₂ was reduced to 2 kPa. Static CA gas concentrations were established using N₂ generated from a membrane gas separator (Permea, St. Louis, MO), and air and CO₂ from compressed gas cylinders. CA chamber gas concentrations were assessed and adjusted as needed using a commercial control system (Empire Controls, Chelan, WA).

Harvest maturity and fruit quality assessment. Starch content was rated visually using a 1–6 scale (Hanrahan, 2012) on 18 fruit at harvest after staining an equatorial section with a 30-mm solution of I-KI. Peel hue was measured on each fruit at a disorder-free location with minimal red color with a chromameter (CR-300, Minolta, Japan) using CIE illuminant C. Values were converted to hue as described by Hunter and Harold (1987). Firmness was measured on two pared surfaces at the equator of each fruit with a MTD1 analyzer (Mohr and Associates, Richland, WA) fitted with an 11-mm-diameter probe. SSC and TA were assessed using freshly prepared juice from nine replicates (prepared by combining juice from two fruit) per storage treatment combination using a refractometer (Atago N1; Atago, Tokyo, Japan) and a TIM850 titrator (Radiometer, Lyon, France), respectively. Juice was titrated to pH 8.2 using 0.1 N KOH. Internal ethylene concentration (IEC) was analyzed as described previously (Lumpkin et al., 2014). Ethylene and CO₂ were analyzed in gas samples obtained from four replicates of five fruit held in 3.79-L glass jars sealed with teflon lids with two gas ports. Jars were purged with air at 1.76 mL·s⁻¹ for 1 h, then 1 mL gas samples were collected from the outlet port and analyzed using a Hewlett Packard 5890 gas chromatograph (Agilent, Palo Alto, CA) equipped with a 0.5-m, 3.2-mm-i.d. stainless steel column packed with Porapak Q (Supelco, Bellefonte, PA), a methanizer (John Booker & Co., Austin, TX), and a flame ionization detector. The N₂ carrier, H₂, and air flows were 0.5, 0.5, and 5 mL·s⁻¹, respectively. Oven, injector, and detector temperatures were 35, 100, and 300 °C, respectively. The methanizer temperature was 290 °C controlled by an Instrumentation Temperature Controller (Valco Instruments, Inc., Houston, TX) with an H₂ flow of 0.5 mL·s⁻¹.

Fruit disorders (bitter pit, peel blotch, DFB, cavitation, soft scald, soggy breakdown, and greasiness) were reported as incidence. Bitter pit was identified as surface lesions <5 mm in diameter with underlying brown, corky tissue. Irregularly shaped peel areas of rough, brown tissue >5 mm were considered peel blotch (Fig. 1). Brown internal tissues outside the core with diffuse, brown to grayish color were considered DFB. Soft scald and soggy breakdown were rated based on symptoms described previously (Watkins et al., 2004). In years 1 and 2, external disorders were assessed while fruit (n = 72, year 1; n = 108, year 2) was in cold storage at 1, 2, 3 (year 2 only), 4, and 7 months after harvest. At 7 months in all years, fruit were removed from storage and held 7 d at 20 °C before further analyses. The combined incidence of bitter pit, peel blotch, DFB, and cavity development is reported as total non-chilling disorders and represents the percentage of fruit in each 1-MCP/ atmosphere combination with at least one of these disorders.

Experimental design and statistical analysis. The experiment was conducted using a completely random design with three replicates of six individual fruit at harvest and for each 1-MCP/storage atmosphere combination. All statistical analyses were conducted using SAS 9.4 (SAS Institute, Raleigh, NC). Disorder incidence data (percentage) was arcsine square root transformed before analysis. Significant treatment differences for fruit quality attributes and storage disorders were identified using analysis of variance and treatment means separated using Tukey’s honestly significant difference, P < 0.05, 0.01, or 0.001.

Results

At harvest, all lots had IEC values greater than 0.07 mmol·m⁻³, 4.5 starch rating, 54 N, 98 °C·h, 0.522% TA, and 11.6% SSC (Table 1).

| Lot | IEC (mmol·m⁻³) | Starch (1–6) | Firmness (N) | Peel (%h) | TA (%) | SSC (%) |
|-----|----------------|--------------|--------------|-----------|-------|--------|
| A   | 0.21 ± 0.08    | 4.9 ± 0.2    | 65.9 ± 1.6   | 104 ± 2.0 | 0.640 ± 0.016 | 13.8 ± 0.2 |
| B   | 0.52 ± 0.20    | 4.6 ± 0.2    | 61.6 ± 1.5   | 103 ± 1.2 | 0.530 ± 0.020 | 14.4 ± 0.3 |
| C   | 1.50 ± 0.12    | 5.3 ± 0.1    | 57.9 ± 1.0   | 108 ± 1.0 | 0.841 ± 0.037 | 11.7 ± 0.1 |
| D   | 2.00 ± 0.20    | 5.0 ± 0.1    | 63.2 ± 1.5   | 104 ± 1.9 | 0.523 ± 0.018 | 13.5 ± 0.1 |
| E   | 3.53 ± 0.18    | 5.2 ± 0.2    | 54.7 ± 1.2   | 103 ± 1.8 | 0.634 ± 0.020 | 14.0 ± 0.1 |
| F   | 0.08 ± 0.02    | 5.0 ± 0.1    | 55.2 ± 0.9   | 105 ± 1.4 | 0.549 ± 0.010 | 12.7 ± 0.1 |
| G   | 3.30 ± 0.09    | 5.8 ± 0.1    | 56.5 ± 0.9   | 98 ± 2.5  | 0.529 ± 0.011 | 12.5 ± 0.0 |

IEC = internal ethylene concentration; TA = titratable acidity; SSC = soluble solids content. Values are means ± standard error, n = 18 [IEC, starch pattern index, firmness, peel hue (%)], n = 9 (TA, SSC).

Fig. 1. Bitter pit (left apple) and peel blotch (right apple) symptoms on ‘Honeycrisp’ apples.
did not have cavities was stored in air without 1-MCP treatment. Lots B (year 1) and C had an atmosphere main effect where fruit stored in CA had higher incidence compared with fruit stored in air.

The combined incidence of bitter pit, DFB, and cavity development was within 5% of that for bitter pit for most treatment combinations for lots A, C, D, and E. Exceptions were lot A in year 2 where 10% of the fruit stored in CA following 1-MCP application had DFB and/or cavities compared with 17% that had bitter pit alone; lot B in year 1 where all treatment combinations except air control had 5% or more combined disorder incidence compared with bitter pit alone; lot B in year 2 where control CA after 1 or 9 d had greater than 5% more disorders compared with bitter pit alone. The increased total incidence of bitter pit, DFB, and cavities was typically due to cavity development also in fruit not having bitter pit.

Soft scald and soggy breakdown incidence were low, and consistent treatment effects were not observed. Peel greasiness was affected by atmosphere (lot A in year 2, lot E) or atmosphere × 1-MCP (lot B in year 2) where fruit stored in air had the highest incidence.

* Peel hue was lower (yellower) in fruit stored in air for all lots. CA established 1 d after harvest resulted in the highest or among the highest peel hue values for lots C, D, and E with fruit stored in air having lower values. For all lots, an atmosphere × 1-MCP interaction existed for SSC with control fruit stored in air having the lowest or among a group with the lowest values. Lots A–D had an atmosphere × 1-MCP interaction for TA with CA 1 d after receipt having the highest or among the highest values.

Discussion

Reduced bitter pit development in ‘Honeycrisp’ apple when fruit were stored in CA rather than air is consistent with previous research for other cultivars (Hewett, 1984; Sharples, 1982; Webster and Forsyth, 1979) but not in ‘Honeycrisp’ where bitter pit incidence exceeded 40% in air-stored fruit (Contreras et al., 2014). The lack of bitter pit reduction during ‘Honeycrisp’ CA storage may be due to different atmosphere composition and/or CA establishment protocols. The absence of an effect of CA storage on bitter pit incidence in orchard lots with low bitter pit incidence is consistent with Contreras et al. (2014) and DeEll et al. (2015). Anaerobiosis before cold storage has also resulted in decreased bitter pit development (Pesis et al., 2010), but the CA conditions imposed in the work reported here are unlikely to have created anaerobic conditions based on a low-oxygen limit of 0.33 kPa O2 for ‘Honeycrisp’ at 10 °C (Wright et al., 2010). The reduction in bitter pit

Table 2. ‘Honeycrisp’ apple fruit quality and disorders 7 months after harvest, lot A for two harvest years.

| Yr | Atmosphere 1-MCP | BP (%) | PB (%) | DFB (%) | Cavity (%) | TNCD (%) | SSld (%) | SB (%) | Greasy (%) | Peel (h) | SSC (%) | TA (%) | Firm (N) | C2H4 (mol·g⁻¹·s⁻¹) | CO2 (mol·g⁻¹·s⁻¹) |
|----|------------------|--------|--------|---------|------------|---------|---------|-------|----------|---------|--------|-------|--------|----------------|------------------|
| 1  | Control: CA 9 d  | 61     | 0      | 0       | 3         | 63      | 0       | 0     | 0        | 96.9    | 14.1   | 0.584 | 60.8   | 0.53              | 87 b              |
|    | CA 1 d           | 39     | 1      | 0       | 0         | 39      | 0       | 0     | 0        | 99.4    | 14.8   | 0.347 | 60.4   | 0.69              | 94 b              |
|    | 1-MCP: CA 9 d    | 66     | 5      | 2       | 0         | 67      | 0       | 0     | 0        | 91.8    | 13.9   | 0.43  | 63.8   | 0.43              | 95 b              |
|    | CA 1 d           | 40     | 5      | 0       | 0         | 40      | 0       | 0     | 0        | 96.5    | 14.3   | 0.42  | 62.4   | 0.09              | 112 a             |
|    | Air              | 65     | 5      | 1       | 0         | 66      | 0       | 0     | 0        | 91.8    | 14.3   | 0.43  | 62.4   | 0.09              | 112 a             |
|    | 1-MCP            | 66     | 5      | 3       | 0         | 66      | 0       | 0     | 0        | 91.8    | 14.3   | 0.43  | 62.4   | 0.09              | 112 a             |
|    | Air              | 36     | 0      | 1       | 0         | 37      | 0       | 0     | 0        | 91.2    | 14.3   | 0.43  | 62.4   | 0.09              | 112 a             |
|    | 1-MCP            | 36     | 0      | 1       | 0         | 37      | 0       | 0     | 0        | 91.2    | 14.3   | 0.43  | 62.4   | 0.09              | 112 a             |

BP = bitter pit; PB = peel blotch; DFB = diffuse flesh browning; TNCD = total incidence of non-chilling disorders; bitter pit, diffuse flesh breakdown, cavities; SSld = soft scald; SB = soggy breakdown; greasy = peel greasiness; firm = firmness; CA = controlled atmosphere; 1-MCP = 1-methylcyclopropene; TA = titratable acidity; SSC = soluble solids content. Fruit were held at 10 °C for 7 d and 3 °C thereafter. Fruit at 10 °C were exposed to 0 or 4.2 µmol·m⁻²·s⁻¹ 1-MCP for 16 h. Apples were stored in air or a controlled atmosphere (CA) established 1 or 9 d after 1-MCP exposure. Storage temperature was decreased to 3 °C 7 d after receipt. Values are means (n = 72, year 1; n = 108, year 2) ± se.
development following 1-MCP application at harvest or in response to CA storage indicates that limited ethylene action resulting from 1-MCP treatment (Sisler and Blankenship, 1996) and/or a low-O2 environment (Burg and Burg, 1965) soon after harvest may contribute to reduced bitter pit development. These and previous results (Hewett, 1984; Shariples, 1982; Webster and Forsyth, 1979) indicate a possible role for ethylene in bitter pit development. Reduced bitter pit incidence following 1-MCP treatment and/or CA storage was most evident in lots with high disorder incidence in control fruit stored in air. Significant main effects for 1-MCP and atmosphere indicate that both factors can influence bitter pit development. When main effects for 1-MCP and CA were significant but the interaction was not significant, the effect of each independent variable was consistent for all interaction means followed by different lowercase letters are significantly different. *n = 10 for C2H4 and CO2 production.

### Table 3. 'Honeycrisp' apple fruit quality and disorders 7 months after harvest, lots B, C, and D for one harvest year.

| Lot | Atmosphere | 1-MCP | BP | PB | DFB | Cavity | TNCd | SSld | SB | Greasy | Peel | SSC | TA | Firm | C2H4 | CO2 |
|-----|-------------|-------|----|----|-----|--------|------|------|----|--------|------|-----|----|------|------|-----|
| C   | Control: CA 9 d | 17  | 3  | 4  | 17  | 31     | 3    | 0    | 0  | 96.9  | 11.7 | 0.244 | 55.2 | 0.49 | b   | 76   |
|     | SA 9 d      | 8   | 3  | 1  | 6   | 14    | 0    | 0    | 0  | 99.4  | 11.8 | 0.341 | 58.9 | 0.55 | b   | 68   |
|     | Air         | 7   | 0  | 0  | 4   | 11    | 6    | 0    | 0  | 92.8  | 10.9 | 0.161 | 58.6 | 0.76 | b   | 99   |
|     | 1-MCP: CA 9 d | 7   | 7  | 2  | 18  | 24    | 13   | 0    | 3  | 96.5  | 12.1 | 0.304 | 57.2 | 0.06 | c   | 45   |
|     | Air         | 8   | 8  | 0  | 6   | 13    | 3    | 0    | 0  | 91.8  | 11.9 | 0.322 | 56.8 | 0.58 | c   | 44   |
|     | Atmosphere  | NS  | NS | NS | NS  | NS    | NS   | NS   | NS | NS     | NS   | NS   | NS   | NS   | NS   | NS |
| C   | 1-MCP       | NS  | *** | NS | NS | NS   | NS    | NS   | NS | NS     | NS   | NS   | NS   | NS   | *** | *** |
|     | Atmosphere  | NS  | NS | NS | NS | NS    | NS   | NS   | NS | NS     | NS   | NS   | NS   | NS   | NS   | NS |
| D   | Control: CA 9 d | 11  | 0  | 1  | 13  | 22    | 9    | 19   | 42  | 94.9  | 13.2 | 0.325 | 61.1 | 0.54 | a   | 73   |
|     | SA 9 d      | 3   | 0  | 0  | 7   | 10    | 11   | 14   | 0   | c 96.4 | 12.9 | 0.335 | 60.7 | 0.52 | a   | 62   |
|     | Air         | 11  | 0  | 0  | 0   | 11    | 3    | 0    | 0  | 94.4  | 12.9 | 0.251 | 61.8 | 0.65 | a   | 91   |
|     | 1-MCP: CA 9 d | 13  | 0  | 0  | 3   | 13    | 8    | 11   | 0   | c 97.5 | 13.7 | 0.373 | 59.6 | 0.10 | b   | 34   |
|     | Air         | 13  | 0  | 0  | 6   | 14    | 1    | 19   | 0   | c 92.4 | 13.5 | 0.346 | 57.2 | 0.04 | b   | 45   |
|     | Atmosphere  | NS  | NS | NS | NS | NS    | NS   | NS   | NS | NS     | NS   | NS   | NS   | NS   | NS   | NS |
| D   | 1-MCP       | NS  | NS | NS | NS | NS    | NS   | NS   | NS | NS     | NS   | NS   | NS   | NS   | NS   | NS |
|     | Atmosphere  | NS  | NS | NS | NS | NS    | NS   | NS   | NS | NS     | NS   | NS   | NS   | NS   | NS   | NS |

**BP = bitter pit; PB = peel blotch; DFB = diffuse flesh browning; TNCd = total incidence of non-chilling disorders; Cavity = cavities; SSld = soft scald; SB = soggy breakdown; Greasy = peel greasiness; Firm = firmness; CA = controlled atmosphere; 1-MCP = 1-methylcyclopropene; TA = titratable acidity; SSC = soluble solids content. Fruit were stored in air or a CA (3 kPa O2 for 2 d, then 2 kPa; 0.5 kPa CO2 throughout) imposed 1 or 9 d after receipt. 1-MCP × atmosphere means followed by different lower case letters are significantly different. n = 72 (year 1) or 108 (year 2) for disorders; n = 18 for peel and firmness. NS, *; **, *** Nonsignificant or significant main effects and interactions at P < 0.05, 0.01, or 0.001, respectively.**
10 °C (0.33 kPa) reported for ‘Honeycrisp’ (Wright et al., 2010), therefore DFB and cavities are unlikely a result of low-oxygen injury. Application of diphenylamine before imposition of CA provides protection from CA injury development (Contreras et al., 2014) and may be a means to reduce the risk of CA injury when using a postharvest protocol with CA established during conditioning.

The use of 1-MCP and/or CA resulted in enhanced fruit quality compared with untreated fruit stored in air including reduced peel greasiness and yellow color development and enhanced retention of SSC and TA, consistent with previous reports for this cultivar (Contreras et al., 2014; DeEll et al., 2015; DeLong et al., 2006; Watkins and Nock, 2012). These results are consistent with slower fruit ripening reflected in lower IEC or ethylene production and lower respiration rate. In a number of instances, rapid CA with or without previous 1-MCP treatment enhanced this fruit response compared with CA initiated later, providing additional reduction in fruit ripening and potential benefit in quality retention after storage. Rapid establishment of CA conditions for apples has long been demonstrated to enhance preservation of fruit quality of some cultivars compared with delayed CA establishment (Fidler and North, 1967; Lau, 1982).

Réduction of fruit physiological disorders that develop during storage contributes to enhanced orchard productivity and energy use efficiency in the postharvest supply chain. The postharvest protocols evaluated in this study are applicable without development of additional physical resources at commercial apple warehouses currently operating CA facilities. While an enhanced risk of physiologic disorder development due to CA storage was not observed during this study, regardless of when CA was established, it to what extent risk of injury from CA imposed during or after conditioning exists within the larger population of ‘Honeycrisp’ fruit from multiple orchards and production areas.

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The variable bitter pit response is consistent with other aspects of apple fruit ripening for which 1-MCP impacts are dependent on multiple factors including cultivar, 1-MCP treatment protocol, storage conditions, and storage duration (DeEll et al., 2002; Watkins et al., 2000). Peel blotch occurred in both control and 1-MCP-treated fruit but incidence was often higher for 1-MCP-treated fruit. Peel blotch incidence was always lower compared with bitter pit incidence. In fact, all fruit that had peel blotch also had bitter pit, therefore peel blotch was not an additional source of loss. The exclusive presence of peel blotch in the subset of fruit with bitter pit suggests a possible link between the two disorders, particularly when ethylene action has been inhibited by 1-MCP. The metabolic events that result in bitter pit may be altered following 1-MCP treatment such that disorder symptom development is more likely to progress to peel blotch. Zaanella et al. (2005) previously reported bitter pit damage was more pronounced but incidence was unchanged in 1-MCP-treated fruit compared with controls. Calvo and Candan (2010) reported that 1-MCP-treated ‘Granny Smith’ apples from an early harvest had increased lenticel blotch incidence relative to control fruit; however, lenticel blotch symptoms are localized around lenticels while peel blotch reported here is not lenticel specific.

Conditioning ‘Honeycrisp’ fruit at a higher than final storage temperature for a short period after harvest reduces CI, but this protocol can exacerbate bitter pit development (Watkins et al., 2004). Reports documenting ‘Honeycrisp’ fruit response to CA storage indicate a risk of internal disorders similar to CO2 injury (Contreras et al., 2014; DeEll et al., 2015, 2016; Watkins and Nock, 2012). Under the conditions of these experiments, the low-oxygen CA set point (3 kPa decreased to 2 kPa after 2 d) was above the low-oxygen limit at
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