Detection of Mercury in Water using Filter Paper Based Channel and Colorimetric-Android Readout

Gurushree Dindorkar, Vishal Rathee, Suresh Balpande, Jayu Kalambe

Abstract: The heavy metals dissolved in water are exceptionally unsafe to human and marine health which causes numerous medical complications. This paper demonstrates the use of a Microfluidic paper-based channel (µPADD) and an easy-to-use colorimetric android based application for the accurate detection of heavy metal Mercury (Hg²⁺) in water. Gold Nanoparticles (AuNPs) functionalized with Papain and 2,6-pyridinedicarboxylic acid is used to detect Hg for further colorimetric analysis. Droplet-based Microfluidic channel in star shape with a paper-based stencil, a hydrophobic barrier and a hydrophilic channel using Polyvinyl Alcohol (PVA) was created for this. Colorimetric detection is used to create a database which is used to calibrate the color range for other unknown quantities of Mercury (Hg²⁺) present in water. This experimental database was used to create a user-friendly Android Application based display. The real-time android application was calibrated to quantify mercury concentrations from 0.1g / L to 0.001mg / L by observing a change in color from red to blue. This built platform can be utilized as a basic low-cost and portable system for various other fluid testing.

Keywords: Mercury, Colorimetric Detection, Gold Nanoparticles, µPADs based detection.

I. INTRODUCTION

Dealing with Heavy metal pollutants is an intense issue in making the environment pollution free for over a decade. Different heavy metals such as Mercury (Hg), Lead (Pb), Cadmium (Cd), Arsenic (As), Nickel (Ni) can be exceptionally unsafe to bio-system and biological activities because they can cause severe medical problems in living things. Accessibility of harmful heavy metal ions in water or aquatic ecosystems has genuine health impacts like osteoporosis, Alzheimer’s disease, mental retardation in addition to that these ions are carcinogenic. Hence detection of such harmful metals proved to be a good topic of interest [1]. There are different methodologies that are used for detecting the presence of heavy metals in water. These methods require huge instruments and laboratories that are costly in nature, leading to large investments, need skilled labors for its operation and an enormous amount of reagents and solvents that need proper storage and handling facility. Some of the most commonly employed methods are Atomic Absorption Spectrometry (AAS), Surface Enhanced Raman Spectroscopy (SERS), Inductively Coupled Plasma Mass Spectrometry (ICP-MS), and Fluorescence Spectrometer [2][3][4]. Thus there arises a need for generating resources and facilities that are cost-effective, maneuverable and simple to operate requiring lesser amounts of reagents and have quicker recognition and display system.

In recent times, Gold Nanoparticles (AuNP) are getting colossal consideration for its widespread utility in applications for detection and sensing metals, because of its colorimetric sensing properties. Gold Nanoparticles (AuNP) are exceedingly powerful for colorimetric sensing on account of its high visible range extinction coefficient and color-tunable activity that relies upon the interparticle size [5][6][7]. Recently, extensive research is being garnered for heavy metal detection, particularly with Gold Nanoparticles being a significant factor in nanoscale materials and devices. The AuNP-based colorimetric detector is commonly used to detect small to very small concentrations of heavy metal ions [8][9][10][11].

In this paper, we have discussed a simple colorimetric process for the detection of mercury ions (Hg²⁺) in water, using the Papain and 2,6-pyridinedicarboxylic acid functionalized AuNPs. An experiment where only papain was considered for the detection of Mercury ions demonstrated that it immediately reacts with Hg²⁺ and the color changes from red to blue. However, papain was also found to demonstrate color-changing response to other metals like (Lead) Pb²⁺ and (Copper) Cu²⁺ in the AuNP solution. Thus, to attain extremely high selectivity for Hg²⁺ ions we have used 2,6-pyridinedicarboxylic acid along with papain. Thus, in its presence, Papain and Hg²⁺ react together, to form Au-S whereas Pb²⁺ and Cu²⁺ are isolated.

II. MATERIALS AND METHODS

A. Materials

Chloroauric acid (HAuCl₄) in powder form, Papain papaya extract, 2,6-Pyridinedicarboxylic Acid-99% Pure (PDCA), Mercuric Sulphate, Polyvinyl Alcohol (PVA-powder)-98-99% Hydrolysis. The reagents did not require further cleansing and all the solutions were prepared in adequate quantity of Deionized Water (DI) water.

B. Preparation of AuNPs

AuNPs were processed using the HAuCl₄ Citrate Reduction Method [12][13]. All the glassware were first cleaned by acetone and washed with DI water and were vacuum-dried completely at 60°C for 15
min. The aqueous solution of HAuCl₄ (0.5mM, 200ml) was first prepared by dissolving HAuCl₄ powder in distilled water and stirred till pale yellow color appeared. This 200ml solution of HAuCl₄ was then boiled in a beaker till dense vapor appears and then the aqueous solution is stirred rapidly for another 10 min. 20ml of 38.8mM Trisodium Citrate solution is then poured in the boiling mixture and stirred continuously till color changes. As the trisodium citrate is added to the solution, the pale yellow solution first becomes colorless (Figure 1a) and on further boiling, the solution turns blue. Gradually the blue color changes to purple and then to Ruby Red. This red solution is stirred further at room temperature for another 15 minutes to obtain homogenous color and then left overnight to cool at room temperature (Figure 1b).

![Image](image-url)

Figure 1: (a) Aqueous solution of HAuCl₄ being boiled and stirred. (b) Red-colored Gold Nanoparticle (AuNPs) Solution.

C. Preparation of Hg²⁺ Sample

Multiple aqueous samples containing mercury (Hg) with varied concentrations were prepared for testing [14]. Initially, the stock standard solution of mercury (1.0 gm/L) was prepared by dissolving 0.148g, HgSO₄ in 10ml of water along with mercuric sulphate. Since mercuric sulphate does not dissolve in pure water and a yellow precipitate is formed when mixed with water, 0.83 mL of concentrated H₂SO₄ is added to the solution that produces an exothermic reaction and the solution becomes colorless. Different concentrations (0.1g, 0.05g, 0.025g, 0.0125g, 0.25mg, 0.125mg, 0.56mg, 0.78mg, 0.39mg, 0.19mg, 0.09mg, 0.04mg, 0.02mg, 0.006mg, 0.003mg, 0.001mg) of Hg working solutions were prepared by further diluting the stock solution with DI water. Numerous colorimetric tests were carried out on this stock solution using μPADs. Stock solution was also used to spike the actual samples of water with known levels of mercury for checking its presence in it.

D. Preparation of P-PDCA-AuNPs

Papain only functionalized AuNPs does not give high selectivity to Mercury and are also responsive to Cadmium and Lead. To achieve exclusivity and high sensitivity of AuNPs to mercury, nanoparticles are functionalized with papain and 2,6-Pyridinedicarboxylic Acid [15]. Solid papain is dissolved in water to prepare P-PDCA-AuNPs solution (0.1μM). Papain is added in abundance to AuNPs ensuring that AuNPs are completely functionalized. 2,6-pyridinedicarboxylic acid (PDCA) based water solution (0.5mM) is also prepared using Deionized water. Papain and 2,6-pyridinedicarboxylic (PDCA) acid solutions are then added to the AuNP solution, one at a time which requires settling time to form a homogenous mixture. First, the Papain based aqueous solution was added to the sample containing Hg and the resultant solution after being stirred for 30 minutes was kept in dark for around 6 hours. PDCA solution was then added to the P-AuNPS and the mixture stirred again for 30 minutes and kept away from light for another 6 hours. This Functionalized P-PDCA-AuNPs were used as a reagent for the colorimetric testing to determine the presence of Mercury in water.

E. Fabrication of μPADs

μPADs using stencil-based channels and PDMS are already developed [16][17][18]. But here an approach involving Polyvinyl Alcohol-based barriers on the paper stencil for the generation of μPADs was employed for its simplicity and non-reactiveness with sample and reagents. The μPADs were developed by depositing the PVA solution as a hydrophobic barrier thus preventing the diverging of the sample and reagents on paper. For this, the PVA solution was prepared by mixing 25 grams of polyvinyl alcohol (PVA) in 1000 grams of water. The solution was stirred on a magnetic stirrer for 120 min to get uniform viscous solution which was subsequently sieved using a 25 μm sieve to remove suspended particles. Thus μPADs without costly tools were designed and developed in various shapes using a paper stencil.

The stepwise fabrication of μPADs is shown in Figure 2.

![Image](image-url)

Figure 2: (A) Stencil printed on printing paper. (B) Cut-out of the Stencil (C) Stencil on chromatographic paper (D) Final fabricated μPAD with PVA Layer.

The designed paper mask was accomplished by first printing an example structure carefully onto a trace printing paper using an ink-jet laser printer (Canon LBP 2900B), trailed by carving along the designed pattern. The μPADs were developed in a star-shaped pattern, with 8 test zones each of 7 mm diameter. The middle control area, with a diameter of 9 mm, was used to connect each of the seven test points by channels. The width of the channel was kept 2 mm so as to permit non-delayed concurrent responses in all sample zones for different concentrations of samples [19].
The printed design was cut along the borders and kept on a chromatograph Whatman filter paper, and thick aqueous PVA (Polyvinyl Alcohol) was then deposited along the borders, to draw the hydrophobic barriers which would prevent spreading of sample and reagents. The PVA was coated in such a way so that even if the sample and reagents infiltrates the chromatography paper from both sides (i.e. back and front), a well defined and distinct barrier would exist between the test points. The PVA coated filter paper was allowed to dry on a hot plate at 80°C for 5mins. Fabricated structure with hydrophobic and hydrophilic channels is shown in Figure 3.

![Figure 3: Fabricated μPAD with the PVA barrier.](image1)

**F. Colorimetric Detection using μPADs**

Colorimetric detection of Hg was carried out to estimate the amount of mercury present in water using μPADs which requires very small quantities of chemicals [21]. A series of test samples containing different concentrations of Hg were prepared and analyzed with AuNPs solution and the corresponding color changes were recorded. Different samples containing Hg within a range of 0.1g to 0.001mg were poured on different test points of μPADs.

![Figure 4: μPAD with different concentrations of Hg.](image2)

To counter the effect of error in the absence of a high-resolution camera and determine the effectiveness of this whole process in the presence of impurities with mercury in water, different water samples from various sources were collected in larger quantities (ml) from the nearby areas. The impure water samples were stored in test tubes at room temperature and spiked by adding known quantities of mercury solutions [15][20] for further analysis (Figure 5). Colorimetric tests were carried out after adding P-PDCA-AuNPs solution to the contaminated water samples and stirring it for some time to get uniform color change. The images of the colorimetric tests were used to prepare a database that would help predict the concentration of unknown quantities of mercury present in the water.

![Figure 5: (a) Absence of Hg in water Sample. (b) presence of Hg in the water sample.](image3)

**G. Color detection using ImageJ Software.**

ImageJ software was used to find the color intensity of recorded samples deposited on the μPADs and thus calculate the near exact quantity of mercury (Hg²⁺) content present. The mean color intensity of each zone on the μPAD was determined using the measurement tool from the image toolbar of ImageJ. Initially, the area of interest demonstrating color change is marked by using a circle tool from the tool tray and then the average color intensity of the selected area is easily obtained. Figure 6 shows the test sample calibration curve, i.e. mg/L to mean color intensity graph obtained using ImageJ.

![Figure 6: Calibration curve: detection of Mercury with varied color intensity of samples.](image4)
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I. Hg Detection through Android Application
An android application based on colorimetric detection was formulated which could display the concentration of mercury present in the contaminated water. The android application could detect the color intensity of the samples from the picture taken using a phone camera, analyze it based on the preferred threshold values from the database [22] and then display the output message showing the exact quantity of Mercury (Hg$^{2+}$) present in water. It is also customized to instruct whether the water is safe for drinking or not. Figure 7 shows the App screen displaying a pretested sample with 0.05g of mercury present.

![Figure 7: a) Home page of Android Application b) Message buttons and Selected Sample c) Message box d) Mercury concentration.](image)

The application's versatility lies in the fact that it can efficiently work using a standard phone camera with a resolution of 8 MP, f/2.2 (focal length), 1.12μm (pixel rate), that was also used to prepare the database. The average value of colored pixels present on the test zone of the µPADs was determined using an image processing algorithm which was used to set up the database. It was revalidated that the sensitivity and selectivity of AuNPs rely on a lot of factors such as particle size, concentration, nature, density, pH and temperature [23][24].

III. RESULTS AND CONCLUSION
In this paper, a simple, rapid, cost-effective method of water analysis determining the presence and proportion of mercury is demonstrated. A user-friendly display technique is also proposed and implemented. Using the fact that many metal ions react with the carboxylic group and amino acids like PDCA and increase their selectivity and sensitivity, 2,6-pyridinedicarboxylic acid and Papain was used for functionalizing AuNPs for the detection of mercury (Hg$^{2+}$). As the concentration of mercury increases, the color of the solution changed from red to violet and then to blackish blue (Figure 8).

![Figure 8: Water samples with mercury in the range of 0.1g to 0.001mg](image)

The color variation as observed in µPADs and even in the test tube sample was marred by the proportions of the sample, the quantity of reagents used, the physical dimensions of the µPADs, the resolution of the camera and the environmental conditions. Thus, when the concentration of Mercury (Hg$^{2+}$) was increased above 0.1g/L, no significant color change was observed indicating that most all the AuNPs were already aggregated. The system showed the capability to detect harmful proportions of mercury surrounding 0.002mg/L mark as provided by WHO and can also be calibrated employing suitable changes in reagents to detect other harmful chemicals and metals in fluids.

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