Peripheral effects of vagus nerve stimulation on anxiety and extinction of conditioned fear in rats

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Vagus nerve stimulation (VNS) enhances extinction of conditioned fear in rats. Previous findings support the hypothesis that VNS effects on extinction are due to enhanced consolidation of extinction memories through promotion of plasticity in extinction-related brain pathways however, alternative explanations are plausible. According to one hypothesis, VNS may produce a hedonic effect and enhance extinction through counter-conditioning. According to another hypothesis, VNS reduces anxiety during exposure and this weakens the association of conditioned stimuli with aversive conditioned responses. The present set of experiments (1) used conditioned place preference (CPP) to identify potential rewarding effects associated with VNS and (2) examined the peripheral effects of VNS on anxiety and extinction enhancement. Male Sprague–Dawley rats were surgically implanted with cuff electrodes around the vagus nerve and subjected to a CPP task in which VNS and sham stimulation were each paired with one of two distinct contexts over the course of 5 d. Following this procedure, rats did not show a place preference, suggesting that VNS is not rewarding or aversive.

The role of the peripheral parasympathetic system in the anxiolytic effect of VNS on the elevated plus maze was examined by blocking peripheral muscarinic receptors with intraperitoneal administration of methyl scopolamine prior to VNS. Methyl scopolamine blocked the VNS-induced reduction in anxiety but did not interfere with VNS enhancement of extinction of conditioned fear, indicating that the anxiety-reducing effect of VNS is not necessary for the extinction enhancement.

Exposure-based therapies are considered the “gold standard” approach to the treatment of posttraumatic stress disorder (PTSD) (Rauch et al. 2012). Rather than treating symptoms alone, exposure therapy provides opportunities to extinguish conditioned fear associations (Bouton 1988; Ressler et al. 2004; Davis et al. 2006; Powers et al. 2009). However, exposure therapy is not effective in all patients and many drop out (Schottenbauer et al. 2008; Garcia et al. 2011; Najavits 2015) or relapse after completing therapy (Boschen et al. 2009; Vervliet et al. 2013). We have attempted to target the same mechanisms that contribute to the enhancement of emotionally arousing memories in order to promote extinction memories that can compete with the memory of the trauma. During times of heightened emotional arousal, the brain signals the adrenal gland to release epinephrine into the bloodstream. This adrenergic response results in improvements in the ability to fight, or flee from danger. The increase in epinephrine also appears to contribute to the enhancement of emotionally arousing memories (Gold et al. 1975, 1977; Cahill and Alkire 2003). However, because epinephrine does not freely cross the blood–brain barrier, it cannot directly influence memory substrates in the brain. One pathway by which epinephrine may interact with the brain is through the 10th cranial nerve. The vagus nerve responds to peripheral administration of epinephrine (Miyashita and Williams 2006; Chen and Williams 2012) and stimulation of the vagus nerve increases norepinephrine levels in the amygdala (Hassert et al. 2004), and enhances memory in rats (Clark et al. 1995, 1998) and in humans (Clark et al. 1999). Blocking norepinephrine receptors in the amygdala prevents the memory enhancing effects of epinephrine administration (Liang et al. 1986), indicating that the vagus nerve serves as a bridge between the peripheral stress response and noradrenergic signaling in the brain that modulates the consolidation of emotionally arousing memories.

Vagus nerve stimulation (VNS) enhances the consolidation of extinction memories in rats (Peña et al. 2013; Alvarez-Dieppa et al. 2016; Noble et al. 2018), and it accelerates extinction of conditioned fear and attenuates reinstatement in a rat model of PTSD (Noble et al. 2017), suggesting that VNS could be used to augment the effects of exposure-based therapies in treatment of PTSD and other disorders. VNS may provide an added benefit of reducing anxiety during therapy. Chronic VNS reduces anxiety in rats (Furmaga et al. 2011; Shah et al. 2016) and in humans (George et al. 2008), and we recently found that short-term administration of VNS increased the time that rats spent in the open arms of an elevated plus maze (EPM) (Noble et al. 2019). The vagus nerve is part of the parasympathetic peripheral nervous system and it is called the “vagal brake” because it counteracts the sympathetic effects of stress on the heart and other organs. Although the left cervical vagus nerve is composed of 80% afferent fibers and only 20% efferent fibers, it is possible that peripherally mediated effects contribute to VNS enhancement of extinction memory. The peripheral parasympathetic effects of VNS may interfere with sympathetic responses to conditioned stimuli (CS); potentially altering the association of trauma reminders with the expected stress response. In addition to signaling the brain during times of heightened stress, the vagus nerve relays information from the gut to the brain.
In a recent study, optical stimulation of the right vagal sensory ganglion in mice promoted self-stimulation behavior, drove a conditioned place and flavor preference, and increased dopamine release from the substantia nigra in mice, implicating the vagus nerve in gut-induce reward (Han et al. 2018). It remains to be seen whether electrical stimulation of the left cervical vagus nerve using extinction-enhancing parameters is rewarding in rats.

Although extensive evidence indicates that VNS enhances memory consolidation and synaptic plasticity in extinction-related brain circuitry (Clark et al. 1998; Hassert et al. 2004; Chen and Williams 2012; Peña et al. 2014), alternative hypotheses remain plausible. One alternative explanation is counter-conditioning produced by rewarding properties of VNS (Pearce and Dickinson 1975). A second alternative hypothesis is a VNS-induced immediate anxiolytic effect during exposure to conditioned cues, which blunts physiological responses to conditioned cues, diminishing their potency. Here, we describe research designed to determine whether the VNS parameters that enhance extinction are rewarding and to test the role of peripheral effects of VNS on anxiety and extinction of conditioned fear.

Results

**VNS does not induce conditioned place preference**

To examine the potential of VNS to serve as a rewarding stimulus that replaces the negative CS association with a positive association (counter-conditioning), a conditioned place preference (CPP) test was used to assess potential hedonic effects of VNS. Twelve rats were used in this experiment. Each one was placed in a rectangular Plexiglas apparatus with two distinct, opposing compartments (Fig. 1A), and it was permitted to explore both compartments (places) for 10 min. Time spent in each of the two opposing compartments was used to determine place preference. Paired samples t-tests indicated no significant differences across time spent in the two opposite compartments in the CPP apparatus during the initial preference test (before pairing; \( t_{11} = 0.51, P = 0.62 \)). For CPP training, each of the 12 rats was given VNS or sham stimulation in one of the two compartments and the opposite treatment in the other compartment 2 h later. The compartment that was paired with VNS and the order of VNS and sham stimulation were counterbalanced across the 12 rats. On each of 5 d, one compartment was paired with four 30-sec trains of VNS over the course of 20 min and the other compartment was paired with four trains of sham stimulation over 20 min. Twenty-four hours after the last day of training, rats were again permitted to explore the open apparatus and time spent in each compartment was measured. No significant differences were seen in time spent in one compartment or the other during the CPP test after pairing (\( t_{11} = 0.61, P = 0.55 \); Fig. 1C). Naïve rats were used in a follow-up CPP test to validate the approach by pairing one side of the CPP apparatus with food. No preference for either side was displayed during the Initial Preference Test (before pairing; \( t_{5} = 0.57, P = 0.60 \)). On the CPP test after food pairing, time spent in the food-paired compartment was significantly greater than time spent in the unpaired compartment (Food Side vs. Empty Side, \( t_{4} = 5.34, P < 0.005 \); Fig. 1C). Taken together, these results indicate that a valid measure of CPP did not reveal that VNS is rewarding.

**Peripheral vagal blockade reverses VNS-induced anxiety reduction**

To examine the role of the peripheral parasympathetic nervous system in VNS effects on anxiety, rats were given intraperitoneal injections of a peripherally acting muscarinic acetylcholine receptor antagonist or saline before they were given a single train of VNS or sham stimulation, and then tested on the elevated plus maze (EPM). Acetylcholine binding to muscarinic receptors is the principle signaling mechanism between postganglionic parasympathetic nerves and effector targets. The muscarinic antagonist scopolamine methyl-bromide (methyl-scop) does not cross the blood–brain barrier, so it is used as a peripheral “vagal blockade” (Friberg et al. 1989; Carnevali et al. 2011). Rats were given I.P. injections of either saline or methyl-scop (0.1 mg/kg) 15 min before a single train of VNS or sham stimulation. Ten minutes after stimulation, they were allowed to explore the EPM for 10 min. More time in the open arms was considered an indication of less anxiety (Pellow et al. 1985). Figure 2 depicts the time spent in the open arms. A two-way ANOVA indicated a significant interaction of drug (methyl-scop or saline) versus stimulation (VNS or sham; \( F_{1,32} = 15.04, P = 0.0005 \)) for time spent in the open arms. Main effects of drug \( F(1,32) = 22.86; p < 0.0001 \) and stimulation \( F(1,32) = 10.78; p = 0.0025 \) were significant. A Tukey post-hoc test for multiple comparisons revealed significant differences between saline/VNS (\( M = 248.5, SD = 95.56 \)) versus saline/sham (\( M = 102.5, SD = 35.89 \)); saline/VNS versus methyl-scop/sham (\( M = 84.12, SD = 57.01 \)); and saline/VNS versus methyl-scop/VNS (\( M = 71.99, SD = 35.83 \)). Methyl-scop/VNS was not different from saline/sham or methyl-scop/sham and methyl-scop/sham was not different from saline/sham. Consistent with our previous study, VNS was sufficient to increase time spent in the open arms of the EPM in saline-treated rats (Noble et al. 2019), but we did not observe this anxiolytic effect in methyl-scop-treated rats. There was not a significant interaction \( (F_{1,32} = 0.08, P = 0.7758) \), drug \( (F_{1,32} = 2.148, P = \)
VNS-induced enhancement of extinction is not affected by blockade of peripheral muscarinic receptors

Thirty-six rats were given 2 d of auditory fear conditioning (AFC), followed by a preextinction conditioned fear response test on Day 3. A two-tailed t-test revealed no significant differences in preextinction freezing measured 24 h after fear conditioning (t(32) = 1.500, P = 0.1435), and before stimulation. To test whether the peripheral actions of VNS are required for VNS-dependent enhancement of fear extinction, methyl-scop or saline was administered 15 min prior to VNS or sham-paired extinction. Figure 3 depicts the effects of pre-VNS methyl-scop in rats submitted to extinction training. A two-way repeated measures ANOVA indicated no significant interaction between factors of drug and stimulation on the extinction freezing (F(1, 31) = 35.94, P = 0.5332). However, there was a main effect of stimulation on postextinction freezing (F(1, 31) = 52.20, P < 0.0001). Consistent with previous findings, a Tukey’s post-hoc test showed a significant decrease in freezing in VNS versus sham rats in both the saline- (sham M = 64.67, SD = 13.51; VNS M = 24.26, SD = 17.57) and methyl-scop-treated groups (sham M = 69.87, SD = 25.94; VNS M = 22.08, SD = 13.47). These findings indicate that VNS-dependent enhancement of fear extinction does not require a peripherally mediated parasympathetic effect.

Discussion

VNS enhances extinction of conditioned fear in rats. Although there is evidence that VNS promotes plasticity in extinction-related brain networks and enhances consolidation of extinction memory, two alternative hypotheses were investigated here: (1) Does VNS produce a counter-conditioning effect by replacing an aversive unconditioned stimulus with a rewarding unconditioned stimulus? (2) Do descending peripheral vagal fibers contribute to the VNS effects on anxiety and extinction? We found that rats did not show a conditioned preference or aversion to a place that was associated with VNS, suggesting that the VNS parameters used to enhance extinction are not rewarding. Consistent with our previous findings, in saline-treated rats, VNS increased time spent in the open arms of the EPM. This VNS-induced anxiolytic effect was blocked in rats given I.P. injections of the peripherally active muscarinic antagonist methyl-scop. In contrast, administration of methyl-scop did not block the VNS-induced enhancement of extinction of conditioned fear.

There was not a significant difference in time spent in either place in the CPP apparatus, indicating that stimulation of the left cervical vagus is neither rewarding nor aversive in rats. We previously found that VNS enhances extinction even when it is only administered during half of the exposures to the CS (Peña et al. 2013; Noble et al. 2017). Taken together with the present CPP
results, these findings do not support the hypothesis that a counter-conditioning effect can explain VNS-induced extinction enhancement.

We found that systemic administration of the peripheral parasympathetic muscarinic receptor antagonist methyl-scop was sufficient to block the anxiolytic effect of VNS on the EPM. These results indicate that the anxiolytic effect of VNS involves peripheral parasympathetic activity, and they support the hypothesis that VNS may reduce conditioned fear responding by interfering with the anxiety response during extinction training. Consistent with this hypothesis, chronic VNS reduces expression of anxiety in rats and in humans (George et al. 2008; Furnaga et al. 2011; Shah et al. 2016) and we previously found that pairing VNS with extinction reduced anxiety and arousal 1 wk later in a rat model of PTSD (Noble et al. 2017). However, administration of VNS that was not given during extinction training did not reduce conditioned fear responding (Peña et al. 2013), indicating that VNS effects on extinction were due to enhancement of extinction-related plasticity rather than a generalized anxiolytic effect. Although it is not likely that the reduction in conditioned fear is due to a lasting anxiolytic effect of VNS, it is possible that VNS administered during exposure to conditioned cues attenuates the conditioned fear response during extinction training.

We hypothesized that the anxiolytic effects of parasympathetic engagement may potentiate the effects of VNS on extinction of conditioned fear because VNS prevents the expected sympathetic nervous system response to the CS. Although peripheral administration of methyl-scop blocked the anxiolytic effect of VNS on the EPM, it had no effect on VNS-induced enhancement of extinction of conditioned fear. Based on the inference that the anxiolytic effects of VNS are dependent on peripheral parasympathetic signaling, the present findings do not support the hypothesis that the anxiolytic effects of VNS are necessary for enhancement of extinction. The present findings, by exclusion, suggest that VNS enhances extinction through the promotion of extinction-related plasticity and consolidation of extinction memory (Clark et al. 1995, 1998, 1999).

The pairing of VNS with training promotes brain plasticity (Engineer et al. 2011; Hays et al. 2013) and VNS increases levels of brain-derived neurotrophic factor (BDNF) in the rat brain (Furnaga et al. 2012). Several studies have demonstrated a role for BDNF in the medial prefrontal cortex (mPFC) in extinction of conditioned fear in rats (Peters et al. 2010; Rosas-Vidal et al. 2014, 2018). Our previous findings indicate that VNS administration during extinction influences synaptic plasticity in the extinction-related pathway between the infralimbic region of the prefrontal cortex and the basolateral amygdala (Peña et al. 2014), and VNS pairing with extinction increases expression of plasticity-related proteins GluN2B and phosphorylated CaMKII in the basolateral amygdala (Alvarez-Dieppa et al. 2016). GluN2B receptor binding to the endogenous ligand in the basolateral amygdala is essential for extinction of conditioned fear (Sotres-Bayón et al. 2007; Alvarez-Dieppa et al. 2016). This evidence, together with the results presented here, indicate that VNS enhances extinction of conditioned fear by influencing synaptic plasticity that supports the extinction memory.

The United States Food and Drug Administration approved the use of VNS for the prevention of seizures in humans in 1997 and it was approved for the treatment of depression in 2005. Our previous findings indicate that VNS delivered during exposure to CS in the absence of the unconditioned stimulus, produces targeted plasticity and reverses extinction impairments in rat models of PTSD (Peña et al. 2014; Noble et al. 2017; Souza et al. 2019). Pairing extinction with VNS also attenuates fear renewal and reinstatement and produces lasting benefits on tests of anxiety, arousal, and avoidance symptoms (Noble et al. 2017; Souza et al. 2019). The present findings indicate that VNS may also counteract the conditioned peripheral sympathetic response experienced in exposure therapy. Some have demonstrated that anxiolytic treatments interfere with progress in exposure therapy (Rothbaum et al. 2014). This may be due to their impairing effects on memory consolidation (Veselis et al. 1997). For example, the benzodiazepine alprazolam produces amnestic effects in normal subjects and in patients with agoraphobia (Block and Berchou 1984; Curran et al. 1994). Unlike these anxiolytic drugs, VNS enhances memory consolidation. Therefore, VNS is an attractive adjuvant to exposure-based therapies because it offers a unique combination of memory consolidation-enhancing and anxiety-reducing effects that may improve tolerability and reduce dropout.

Materials and Methods

Animals

All procedures were carried out in accordance with the NIH Guide for the Care and Use of Laboratory Animals and were approved by the Institutional Animal Care and Use Committee of the University of Texas at Dallas. Ninety-four male Sprague–Dawley rats (Charles River) weighing 225–250 g were housed on a 12 h light–dark cycle (lights on at 7:00 a.m.) with access to food and water ad libitum. To habituate rats to handling, they were handled by an experimenter for 5 min per day for 5 d before the start of every experiment.

Cuff electrode production

Platinum–iridium wire electrodes were glued to the inside of bio-compatible micro-renathane cuffs (1.25 mm inner diameter, 2.5 mm outer diameter, 4.0 mm long). Omnetics four-pin connectors were fixed on top of the skull. These connectors made contact with the platinum–iridium wires in the cuff, and with the cable that was plugged into the head cap, on one end, and an AM systems stimulator on the other end. For details, see Childs et al. (2015).

Vagus nerve stimulation and surgery

The left vagus nerve was located at the cervical level in anesthetized rats (2% isoflurane at an oxygen flow rate of 600–800 mL/min) and isolated from other tissue. The platinum–iridium electrode cuff was wrapped around the nerve. Electrode leads were tunneled subcutaneously to the top of the head and connected to an Omnetics connector that was affixed to the skull with acrylic. To test the efficacy of the cuff electrode after implantation, current (0.8 mA, 1 s) was applied to the cuff electrode while the rat was still anesthetized and breathing rate was observed. A brief cessation of breathing following VNS was used to indicate that the cuff electrode was properly placed and functioning. If the cessation of breathing was not observed, the cuff was adjusted or replaced. For sham rats, the vagus nerve was isolated from the other tissue and an Omnetics connector was affixed to the skull, but the platinum electrode was not implanted. Rats were given 1 wk to recover from surgery before training and testing.

For VNS administration, the head cap was connected to the stimulator for both VNS- and sham-treated rats. For a visual demonstration of surgical and VNS details, see Childs et al. (2015). Stimulation was delivered at 0.4 mA, 20 Hz, 100 μsec pulse width, for a duration of 30 sec. These parameters were previously used to enhance extinction of conditioned fear in rats (Noble et al. 2017, 2019).

Conditioned place preference (VNS vs. sham)

Twelve rats were used for this experiment. During an initial place preference test, each rat was placed in a rectangular Plexiglas apparatus (72 × 25 cm; Allied Plastics LLC; Fig. 1). The walls of the apparatus were 40 cm tall and the top was open. The apparatus was divided into three small compartments by two removable inner
Mechanisms of VNS enhancement of extinction

Conditioned place preference (food vs. no food)
To validate the CPP protocol, six additional rats underwent a control test using an appetitive stimulus (Froot Loops). These rats did not undergo surgery. Six Froot Loops were placed in each home cage every day for 5 d. For 3 d, rats were given free access to all of the compartments of the CPP apparatus for a total of 10 min. On the third day, the time spent in each compartment was recorded. During CPP training, one compartment contained six Froot Loops and the other contained no food. Again, every rat was exposed for 20 min to each side every day for 5 d. The side that was paired with Froot Loops was counterbalanced. Twenty-four hours after the fifth day of place pairing, interior walls were removed and rats were given free access to all compartments of the CPP apparatus for 10 min. No food was present on the maze.

Elevated plus maze
Rats were placed in the central part of an elevated plus-shaped maze with two walls (50 cm tall) on opposing arms and no walls on the other two arms. The arms were 10 cm wide, 50 cm long, and elevated 55 cm from the ground. During a 10-min test, time spent in the open arms, time spent in the closed arms, and time spent in the center of the maze were recorded. Greater time spent in the open arms was used as an indication of reduced anxiety and less time spent in the open arms was considered a measure of increased anxiety (Pellow et al. 1985; Walf and Frye 2007).

Thirty-six rats were treated with intraperitoneal injections of 0.1 mg/kg methyl-scop dissolved in sterile saline, or saline alone. The dose of methyl-scop was selected from a preliminary dose-response study indicating that 0.1 mg/kg (without VNS or sham stimulation) was the highest dose that did not affect EPM performance. Fifteen minutes after the injection, rats were given VNS (n=9/group) or sham (n=9/group) stimulation in their home cages and then placed in the center platform of an EPM 10 min later.

Auditory fear conditioning
Thirty-four rats were exposed to four pretones (9 kHz, 70 dB, lasting 30 sec) in order to determine baseline levels of freezing in response to the sound. Following these four tone exposures, eight tones were played and a single 1 sec, 0.4 mA footshock overlapped with each tone. The timing of the footshock was randomized during the 30-sec tone presentation and the tones were presented at a random inter-stimulus interval of between 120 and 240 sec. Eight tone-1 sec footshock pairings were given again 24 h later. No pretones were given on the second day of fear conditioning. All AFC occurred in Context A (electric grid floor, no olfactory cue). A conditioned fear response test was given 24 h after the 2 d of AFC. Fear responses to the CS were measured in Context B (Context A chamber with Plexiglas insert on floor and addition of peppermint oil odor cue). The 30-sec tone was presented four times with an interval of 120–240 sec and no footshock was administered. Freezing during tone presentation was recorded using a camera located on the side wall of the training apparatus. Two independent experimenters who were blind to conditions measured freezing in response to the tone and scores were averaged.

Anxiolytic contributions to extinction
Extinction trials were given in Context B on the following day. Rats received an intraperitoneal injection of either methyl-scop (0.1 mg/ml; n=9/group) or saline (n=9/group). Fifteen minutes following the injection, rats underwent extinction training where four presentations of the CS were paired with VNS, or sham stimulation, as in our previous studies (Peña et al. 2014; Childs et al. 2015; Álvarez-Dieppa et al. 2016). The day after extinction training, conditioned fear responses to the CS were measured again in Context B (Fig. 3a).

Statistical analyses
Time spent in the paired compartment during the CPP tests, time spent in the open arms and moving on the EPM, and time spent freezing in the extinction context were converted to percentages of total exposure time. CPP results were analyzed using a paired sample t-test. A two-tailed t-test was used to compare conditioned fear responses (% time spent freezing) in sham and VNS groups before extinction training. Data for peripheral vagal blockade experiments were analyzed using a two-way ANOVA to test for significant effects of drug (methyl-scop vs. saline) and stimulation (VNS vs. sham), and an interaction between the two variables, followed by a Tukey’s post-hoc test for multiple comparisons. Statistically significant effects were defined as those with P values that were <0.05.

Two rats were excluded from analysis for failure to express conditioned fear following AFC (freezing less than 50 percent of exposure time). Exclusion of these rats did not alter statistical comparisons.

Competing interest statement
This work has not been published and has not been submitted for publication elsewhere while under consideration. L.J.N., A.C., K.K.C., and R.R.S. declare no potential conflicts of interest. C.K.M. is an
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