Supporting information

Glycyrrhetinic acid improves insulin resistance by regulating the balance between the Ras/MAPK and PI3K/Akt pathways

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1. Abbreviations

DMEM: Dulbecco’s modified eagle medium
NH2-MMs: Fe3O4 amino magnetic microspheres
EDCI: 1-[3-(Dimethylamino)propyl]-3-ethylcarbodiimide hydrochloride
HOBt: N-hydroxybenzotrizole
Sulfo-SADP: [sodium1-((3-((4-azidophenyl)disulfanyl)propanoyl)oxy)-2,5-dioxopyrroolidine-3-sulfonate]
N3-tag: 3-azido-7-hydroxy-2H-chromen-2-one
FBS: Fetal bovine serum
PBS: Phosphate-buffered saline
PAGE: Polyacrylamide gel electrophoresis
SDS: Sodium dodecyl sulfate
DTT: DL-dithiothreitol

2. Target prediction of GA

To screen the potential target proteins of GA, the top 20 protein targets (determined based on fit value using PharmMapper were software) were analyzed using bioinformatics tools. Next, the interacting proteins were analyzed using String 10.0 (http://www.string-db.org/). We used AutoDock 4.2 software to conduct molecular docking studies to evaluate the interaction targets and GA. At last, we performed capture assays to verification of target proteins.

Reverse docking of GA was performed using the PharmMapper server (http://59.78.96.61/pharmmapper/). We prepared a 3D structure of GA in the sdf format using the ChemBio3D Ultra 13.0 software (PerkinElmer Inc., U.S.A.), and submitted the sdf file to the PharmMapper server with the choice of “human protein targets only” and the maximum generated conformations set to 300. In our study, we employed the 2010 version. The first 20 candidate targets of GA are listed in (Supplementary Table S1). Next, the interacting proteins were analyzed using String 10.0 (http://www.string-db.org/). Three targets in the insulin-related signaling pathways, namely, HRAS, PRKCA (PKCα) and MAP2K1 (MEK1). In addition, two targets in the steroid hormone biosynthesis signaling pathways, HSD11B1 and HSD17B1.
The three-dimensional structures of the MAP2K1, PKCa, HRas, HSD11B1 and HSD17B1 proteins (PDB ID code 4U81, 3IW4, 4EFL, 3CZR and 3HB4) were obtained from the Protein Data Bank (http://www.rcsb.org/pdb). The structures of the MAP2K1, PKCa, HRas, HSD11B1 and HSD17B1 proteins were constructed and minimized using the SYBYL software (Chemical Computing Group, Inc.); then, AutoDock version 4.2 (Olson Laboratory, La Jolla, CA) was applied to perform a docking study using a hybrid Lamarckian genetic algorithm (LGA). The number of LGA runs was set to 30. The step size parameters of quaternion and torsion were 30. The binding energies of the GA target proteins HRAS, PKCa, MEK1, HSD11B1 and HSD17B1 were -7.47, -8.77, -9.93, -10.31 and -10.12 kcal/mol, respectively.

Table S1. Top 20 proteins output from prediction of PharmMapper server.

| No. | Fit | Num | Feature | Name                                                                 |
|-----|-----|-----|---------|----------------------------------------------------------------------|
| 1   | 6.737 | 10 | 9       | Corticosteroid 11-beta-dehydrogenase isozyme 1                      |
| 2   | 5.537 | 10 | 7       | Transhyretin                                                         |
| 3   | 5.083 | 7  | 10      | Amine oxidase [flavin-containing] B                                  |
| 4   | 4.862 | 10 | 8       | Cellular retinoic acid-binding protein 2                            |
| 5   | 4.788 | 8  | 9       | Retinol-binding protein                                              |
| 6   | 4.660 | 9  | 11      | Dihydroorotate dehydrogenase (quinone), mitochondrial               |
| 7   | 4.630 | 15 | 6       | GTPase HRas                                                          |
| 8   | 4.606 | 10 | 14      | Retinoic acid receptor RXR-beta                                      |
| 9   | 4.581 | 16 | 13      | Estradiol 17-beta-dehydrogenase 1                                   |
| 10  | 4.562 | 13 | 5       | Vitamin D3 receptor                                                  |
| 11  | 4.554 | 9  | 12      | Dual specificity mitogen-activated protein kinase kinase 1          |
| 12  | 4.459 | 12 | 4       | Alpha-tocopherol transfer protein                                    |
| 13  | 4.412 | 7  | 10      | Lanosterol synthase                                                  |
| 14  | 4.400 | 11 | 8       | Methionine aminopeptidase 2                                          |
| 15  | 4.335 | 6  | 9       | Glutathione S-transferase A1                                         |
| 16  | 4.333 | 8  | 2       | Androgen receptor                                                    |
| 17  | 4.297 | 10 | 3       | Retinoic acid receptor gamma                                         |
| 18  | 4.281 | 10 | 11      | Nuclear receptor subfamily 1 group 1 member 2                        |
| 19  | 4.251 | 8  | 7       | Retinoic acid receptor RXR-alpha                                     |
| 20  | 4.175 | 9  | 1       | Protein kinase C alpha type                                          |

3. Synthesis section

**General Chemical Reagents and Methods**

All reagents purchased for synthesis. Thin-layer chromatography (TLC) was performed on silica gel GF254 plates with detection using shortwave UV light (λ=254 nm) and staining with 10% phosphomolybdic acid in EtOH, followed by heating on a hotplate. Flash chromatography was performed with silica gel (100-200 mesh) with EtOAc/petroleum ether or CH2Cl2/MeOH as eluent. 1H and 13C NMR spectra were recorded on a Bruker AV 400 spectrometer at 400 MHz (1H NMR) and 100 MHz (13C NMR), using CDCl3 as solvents. Coupling constants are displayed in Hertz.

Figure S1. Synthetic route for compound 2 (Alkynyl-GA). Reagents and conditions: (a) EDCI, HOBt, triethylamine, dichloromethane; (b) mono-Propargylamine, dichloromethane.
Compound 2 Synthesis (Alkynyl-GA)

To a solution of compound 1 (0.9414 g, 2.0 mmol) in 20 ml of dichloromethane. EDCI (0.4601g, 2.4mmol), HOBt (0.3243 g, 2.4 mmol) was added. The mixture was stirred for 0.5 h at 0 °C. Then triethylamine (8.0 mmol, 1115 µL) was added dropwise and stirred for 0.5 h at 0 °C. Then mono-propargylamine (3.0 mmol, 206 µL) was added and the mixture was stirred for 1 h at 0 °C and stirred at room temperature for 2 days. The reaction was quenched with water and extracted with dichloromethane (3×25 mL). The combined organic layers were dried over Na2SO4, and concentrated to give a crude product. The crude product was solubled in 3 mL methylene chloride and purified by column chromatography on silica gel (dichloromethane: methanol = 30:1) to get the desired products compound 2, a white solid (0.4814 g, 47.8%) (Figure S3). ¹H NMR (400 MHz, CDCl₃) δ 5.83 (s, 1H), 5.66 (s, 1H), 4.04 (dd, J = 39.7, 17.6 Hz, 2H), 3.21 (d, J = 5.4 Hz, 1H), 2.76 (d, J = 12.9 Hz, 1H), 2.27 (d, J = 33.0 Hz, 2H), 2.12 (s, 1H), 2.00 (d, J = 14.6 Hz, 1H), 1.91 (d, J = 9.3 Hz, 1H), 1.86 – 1.70 (m, 3H), 1.58 (d, J = 13.1 Hz, 4H), 1.52 – 1.30 (m, 9H), 1.27 – 1.07 (m, 10H), 1.05 – 0.88 (m, 5H), 0.79 (s, 5H), 0.67 (d, J = 11.5 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 200.4, 175.7, 169.3, 128.8, 79.9, 79.0, 71.9, 62.0, 55.1, 48.2, 45.6, 43.7, 43.4, 42.0, 39.3, 37.5, 37.3, 32.9, 32.1, 31.6, 29.5, 28.6, 28.3, 27.5, 26.7, 26.6, 23.6, 18.9, 17.7, 16.6, 15.8 (Figure S2). HRMS (ESI⁺): m/z [M+Na]+ calculated for C33H49O3: 507.75; found: 530.3608. (Figure S3).
Figure S2. The NMR data of Alkynyl-GA, (a) $^1$H NMR spectrum of Alkynyl-GA probe (400 MHz, CDCl$_3$) and (b) $^{13}$C NMR spectrum of Alkynyl-GA probe (100 MHz, CDCl$_3$).
Figure S3. The HRMS data for alkynyl-GA. (a) The spectra of GA and (b) alkynyl-GA.

Probe1 characterization section

Fe₃O₄ amino magnetic microspheres (NH₂-MMs) were purchased from Tianjin baseline chromtech research centre (Tianjin, China). The synthetic route for Probe1 was shown in (Figure S4).

Figure S4. Synthetic route for Probe 1 (compound 5). Reagents and conditions: (a) MMs-NH₂, borate buffer, DMSO; (b) CuBr, DIPEA, methanol; (c) DTT, methanol.

Azide modified-MMs Synthesis (Compound 4)

5 mL NH₂-MMs (25 mg, 5 mg/mL) were suspended in 2 mL borate buffer and Sulfo-SADP (compound 3) (0.5 mg, 11 µmol) were added in sequence, the mixture was shocked at room temperature for 12 h. After that, the azide modified-MMs was enriched through magnetic separation and washed with water for three times. The gathered azide modified-MMs (Compound 4) was used directly for next steps.

Probe 1 (Compound 5)

CuBr (10 mg, 0.7 mmol) was dissolved in degassed methanol (3 mL) under argon atmosphere and DIPEA (35 µL, 0.18 mmol) was added. The resulting yellowish suspension was degassed for 30 min under a stream of argon and with exclusion of light. Alkynyl-modified GA probe compound 2 (5 mg, 10 µmol) was dissolved in degassed methanol (0.5 mL) and treated with 1 mL of the freshly prepared suspension of CuBr-DIPEA and azide modified-MMs (25 mg, 5 mg/mL). The reaction mixture was shocked at room temperature with exclusion of light for 24 h. Then the GA-modified functionalized MMs were separated with magnet and washed three times each with methanol and water. The gathered GA-modified functionalized MMs (Compound 5) was used directly for next steps.
To a solution of probe1, probe1 (25 mg, 5 mg/mL) in 1 mL methanol, DTT (100 mM) were added. The reaction mixture was shocked at room temperature for 1h. Then the was removed the MMs with magnet and get compound 6

Probe 2 characterization section in (Figure S5)

Figure S5. Synthetic route for click chemistry product probe2 (compound 8). Reagents and conditions: (a) CuSO4, sodium ascorbic acid, DMSO, Alkynyl-GA.

Compound 7 (N3-tag)

The synthesis of Compound 7, N3-tag, (3-azido-7-hydroxy-2H-chromen-2-one) was delegated to Wuxi App Tec (Beijing, China).

Compound 8 (probe 2)