Inflammatory Pseudotumor-Like Follicular Dendritic Cell Sarcoma: Imaging Features And radiologic-pathologic Correlation

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Research Article

Keywords: Inflammatory pseudotumor-like follicular dendritic cell sarcoma, Computed tomography, Histopathology, Magnetic resonance imaging, Ultrasonography

DOI: https://doi.org/10.21203/rs.3.rs-109388/v1

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Abstract

Background: Inflammatory pseudotumor-like follicular dendritic cell sarcoma (IPT-like FDCS) is a rare type of tumor. The purpose of this study is to reveal the radiological characteristics of IPT-like FDCS by radiologic-pathologic correlation.

Results: We analyzed two cases of IPT-like FDCS in liver, nine cases in spleen, and two cases in both liver and spleen concomitantly. IPT-like FDCS presented as well-defined iso- or hypodense masses on unenhanced CT images in both liver and spleen. Hyperintensities on T1-weighted images and hypointensities on T2-weighted images with hypointense rings were characteristic features in splenic cases. “Halo signs” were observed in two out of three liver tumors. Hepatic lesions showed significant enhancement, whereas splenic lesions showed only mild enhancement. Delayed annular enhancement was observed in both liver and spleen cases. On ultrasonographic examinations, IPT-like FDCS presented as hypoechoic lesions with enhancement similar to that observed in CT. Hyaline fibrous pseudocapsules, which correlated with hypointensities on T2-weighted images, were microscopically observed at the tumor edge. IPT-like FDCS was characterized by an abundance of small blood vessels and capillaries. Capillaries were also found in the fibrous capsule of some IPT-like FDCSs, which may explain the delayed annular enhancement.

Conclusion: The manifestations of IPT-like FDCS in the liver and spleen showed differences that warrant they be approached differently during diagnosis. Characteristic radiological findings of IPT-like FDCS included different enhancement patterns between liver and spleen tumors and rim-like hypointensities on T2-weighted images, as well as annular enhancement, on CT and MR images. These imaging features correlated with tumor pathology.

Background

Follicular dendritic cell sarcoma (FDCS) is an extremely rare type of tumor. While it usually arises from dendritic cells in lymph nodes, a wide variety of extranodal sites may also be involved. There are no more than 100 FDCS cases in the literature, mostly case reports, especially of extranodal forms [1–3]. The improved lymphoid hematopoietic classification system was published in 2001 and the popularization of FDCS-specific immune markers, such as CD21 and CD35, FDCS has been further recognized and reported [4,5].Inflammatory pseudotumor-like FDCS (IPT-like FDCS) is a special subtype of FDCS that develops specially in the liver and spleen. Compared to classical FDCS, IPT-like FDCS is more uncommon [6,7] with different histologic appearance and behavior [8]. Due to the inertia of IPT-like FDCS, surgical resection can achieve good long-term results [8]. Therefore, accurate diagnosis prior to operation is particularly important. To our knowledge, no comprehensive imaging analysis of IPT-like FDCS exists, especially regarding the analysis of radiological-pathological correlations. The purpose of our study was to describe computed tomography (CT), ultrasound, and magnetic resonance imaging (MRI) findings of IPT-like FDCS and relate them to their respective tumor pathological analysis, to contribute to the understanding of this uncommon tumor.
Methods

Study population

This study was approved by the institutional review board and complied with the guidelines of the Healthcare Insurance Portability and Accountability Act. The requirement for written informed consent was waived because of the retrospective study design.

All IPT-like FDCS cases from the 2010–2019 period were retrieved from multiple centers and were confirmed by the local pathological center. In total, 13 patients were included in this retrospective study. The following clinical and pathological data were collected: age, sex, anatomic tumor locations, tumor size, clinical symptoms, surgical and pathological findings, treatment outcomes, and follow-up time. In all patients, preoperative images were reviewed.

Examination methods

Contrast-enhanced MRI was performed in 11 cases, contrast-enhanced CT was performed in eight cases, while both contrast-enhanced MRI and CT were performed in six cases. Ultrasound was performed in four cases, of which one case was also examined by contrast-enhanced ultrasound imaging. Contrast-enhanced CT was performed on a 256-row CT (Revolution; GE Healthcare, Milwaukee, WI, USA) or 16-row multi-detector computed tomography (Brilliance 16; Philips, Eindhoven, the Netherlands). For multiphasic contrast-enhanced CT scanning, an 80–100-mL intravenous iopromide bolus (Ultravist; Bayer HealthCare Pharmaceuticals, Berlin, Germany) with 300 mg iodine/mL was injected at a flow rate of 4–6 mL/s, followed by an injection of 30–40 mL saline solution. The scanning was delayed by 20, 60, 90, and 120 s for the arterial, portal vein, hepatic vein, and delayed phases, respectively. A 1.5-T MR scanner (Ingenia; Philips) or 3.0-T MR scanner (Discovery 750; GE Healthcare) with corresponding 8-channel abdominal phased-array coils were used for MRIs. Conventional axial spin-echo sequence T1-weighted images, axial fast spin-echo sequence with fat suppression T2-weighted images, and single excitation of spin-echo diffusion-weighted images were acquired. For multiphasic contrast-enhanced MR scanning, a 15-mL intravenous bolus of gadopentetate (Magnevist; Bayer HealthCare Pharmaceuticals) was injected into the elbow vein at a flow rate of 2.5 mL/s before axial and coronal T1 high-resolution isotropic volume excitation scanning. The contrast scan was delayed by 25, 80, 110, and 200 s for the arterial, portal, hepatic venous, and delayed phases, respectively. The Philips S2000 or Toshiba Aplio500 systems were used for ultrasound imaging. The frequency of the convex array probe was 3.5–5.0 MHz.

Image assessment

All images were retrospectively reviewed by two senior radiologists (with 15 and 20 years of experience, respectively) who were blinded to the patients’ clinical information until after a consensus was reached. The radiological findings were analyzed considering the following criteria: location of lesions; number of lesions; shape; size (maximum diameters on axial, coronal, or sagittal images); tumor density/intensity on unenhanced CT and MRI (the signal of the tumor-lesioned organ was defined as isodense and
isointense, respectively); contrast enhancement pattern (signal intensities or densities similar or higher to those of adjacent organs were defined as mildly or significantly enhanced); “halo sign” (a thin hyperintense ring surrounding the tumor in T2-weighted images); and pseudocapsules (on T2-weighted images, a thin hypointense ring surrounding the tumor).

Ultrasound examinations were performed in two hospitals by senior ultrasound physicians with 16 years of experience each. The images were retrospectively analyzed by another senior ultrasound physician with 25 years of experience. The physicians agreed on the following signs: tumor location, size, shape, boundary, internal echo, relationship with adjacent organs, color Doppler blood flow, and enhancement intensity and mode of the tumor on contrast-enhanced ultrasound.

Pathological examination

In all 13 cases, the tumors were resected and tissue samples were examined using hematoxylin and eosin staining as well as immunohistochemical EnVision staining. The antibodies used are listed in Table 1. One pathologist with 13 years of experience analyzed the histopathologic specimens after surgery. Appropriate positive and negative controls were evaluated simultaneously. In situ hybridization for Epstein–Barr virus (EBV)-encoded RNA (EBER) was performed in the 13 cases using an EBV probe in situ hybridization kit (Novocastra, Newcastle upon Tyne, UK). The manufacturer’s instructions were followed with no modification, and a known positive control was used to ascertain the sensitivity of the assay.

Statistical analysis

Statistical analyses were performed using SPSS (version 20.0, IBM, New York, USA). Student’s t-test was used to compare the patients’ age at onset and the maximum tumor diameters in the liver and spleen. The chi-square test was used for qualitative variables, such as sex, presence or absence of symptoms, density, signal intensity, enhancement pattern, presence or absence of pseudocapsules, and “halo signs.” p<0.05 was considered significant.

Results

Clinical characteristics

Our patient cohort included seven men and six women aged 36–88 years. Three patients presented with abdominal distension and epigastric pain, and two with low-level fever. The other eight asymptomatic cases were detected during health checkups. In these eight cases, physical examinations revealed no signs of tenderness or rebound tenderness. Only one case had an increased white blood cell count before surgery. The white blood cell count and C-reactive protein levels were high at 19.5×10^9/L and 31.1 mg/L, respectively, whereas alpha-fetoprotein, carcinoembryonic antigen, carbohydrate antigen 125, and carbohydrate antigen 19-9 were within the normal range. Both hepatitis B virus surface antigen and antibody were negative. All tumors were resected without radiological recurrence during the follow-up periods of 10–24 months.
The clinical manifestations of IPT-like FDCS and comparison in liver and spleen are listed in Tables 2 and 3. No statistically significant differences were found for either sex, clinical manifestation, median age, or the number of nodules.

**Multi-detector computed tomography findings**

The imaging characteristics of IPT-like FDCS differed slightly between the liver and spleen (Table 4). Most unenhanced CTs showed a circular or quasi-round, slightly hypodense mass with a clear boundary. The tumor density was homogeneous in five cases. Obvious necrosis was found in three cases, and no calcification or bleeding was detected. The tumors in the liver were slightly enhanced in the arterial phase, decreased in the portal and delayed phases, and showed slightly lower densities than the liver parenchyma (Fig. 1). Splenic tumors showed mild continuous enhancement in five cases. Because the tumor enhancement was lower than that of the parenchyma, the lesions were always hypodense, and annular enhancement was seen in the delayed phase (Fig. 2 a-d). In the other two cases, the tumors in the spleen were significantly enhanced, and one of them showed honeycomb changes. The solid part showed heterogeneous enhancement after contrast administration sparing the central necrotic part (Fig. 3).

**MRI findings**

All liver tumors demonstrated slightly hypointense signals on T1-weighted imaging (Fig. 4a) and slightly hyperintense signals on T2-weighted imaging, with unclear edges and “halo sign” present in 2 of 3 cases (67%) (Fig. 4b). The diffusion-weighted imaging sequence showed slightly high or high signal intensities (Fig. 4c). The liver lesions demonstrated substantial enhancements from the center to the periphery in the arterial phase (Fig. 4d). The enhancement amplitude of the lesions in the portal, venous, and delayed phases tended to be homogeneous and decreased in varying degrees, and annular enhancements could be seen (Fig. 4e,f). Seven of the nine cases with splenic lesions showed slight hyperintensities on T1-weighted imaging (Fig. 5a), slight hypointensities on T2-weighted imaging (Fig. 5b), slightly high or high signal intensities on diffusion-weighted imaging (Fig. 5c), and a hypointense ring on T2-weighted imaging could be seen at the lesion margin (Fig. 5b). There was mild-to-moderate heterogeneous enhancement; after enhancement, the amplitude was always lower than that of the normal spleen tissue, and annular enhancement could also be seen in the delayed phase (Fig. 5d-g). Significant differences between liver and spleen IPT-like FDCSs were found when comparing T1 and T2 signal intensities, enhancement patterns, and the presence of “halo signs” (Table 4).

**Ultrasonographic findings**

Two-dimensional ultrasound demonstrated round-like lesions inside the spleen, with clear boundaries in two cases (Fig. 2e), unclear boundaries in the other two cases, and homogeneous and heterogeneous hypoechogenicity in two cases each. Patchy, slightly hyperechoic, and small dark areas could be seen in one case, and a small dark area could be seen in another case. Strip or short-rod vascular signals could be seen in tumors with a pulse wave registered in the center and at the margins on color Doppler, and
their velocity ranged from 90 to 140 cm/s. Only one case underwent contrast-enhanced ultrasound imaging and showed slight and rapid heterogeneous enhancement from the periphery to the center. Not only did the enhancement appear slightly earlier in the tumor than in the surrounding tissue, but it disappeared earlier as well. The number of cases examined by ultrasound was too low for statistical analyses.

Pathology results

The histology of the IPT-like FDCS in both liver and spleen is very similar to that of classical FDCS with a more prominent inflammatory component. Macroscopically, masses were round or nodular with diameters ranging from 3.5 to 12.0 cm (average 6.5 cm). Sections were solid and grayish-white with clear boundaries, and fibrous pseudocapsules with vitreous degeneration could be seen (Fig. 5i) pushing the surrounding tissue. Necrosis was detected in the center of the larger lesions in three cases. Histologically, the tumors were composed of fusiform or epithelioid cells, arranged in a whirlpool, diffuse sheet, or bundle shape, and fibrous pseudocapsule with vitreous degeneration was confirmed (Fig. 5h). The tumor cells showed a syncytial shape with an unclear boundary, and the rich cytoplasm was reddish. The tumors were mainly composed of an admixture of lymphocytes, plasma cells, and spindle cells. Nuclei were mild-to-moderate atypical, large, round or oval, vacuolar or finely granular, sometimes with pseudoinclusion bodies. Mitosis varied, and multinucleated giant cells were seen in some cases. IPT-like FDCS in the liver had a rich distribution of small blood vessels and capillary networks (Fig. 4g,h). The vascular distribution could also be seen in the fibrous capsule of some IPT-like FDCSs (Fig. 2f). Immunophenotypically, the tumor cells were strongly positive for CD21 and CD35 in 13 cases and for CD23 in three cases. Clusterin staining showed strong positive expression in the cytoplasm in three cases. Some tumor cells were positive for CD68, smooth muscle actin, vimentin, or epidermal growth factor receptor. The Ki-67 index was 5–30%. All cases were positive for the Epstein–Barr encoding region.

Treatment outcomes and follow-up

All 13 patients underwent surgical excision without adjuvant chemotherapy. The tumors were resected, and splenectomy and/or right or left hepatectomy and wedge resection of the liver were performed, accordingly. All cases were followed up postoperatively for periods ranging from 10 months to two years and did not exhibit local recurrence or distant metastasis.

Discussion

IPT-like FDCS is more common in adults. Compared with classical FDCS, IPT-like FDCS appears more in females; however, there was no statistically significant difference regarding sex in our study. Most patients had no clinical symptoms or these were unspecific. In our study population, only two patients had prodromal symptoms of low-level fever, and three patients had symptoms of abdominal distension and abdominal pain, whereas eight patients had no obvious typical symptoms. This agrees with previously reported findings [7]. While IPT-like FDCS is very similar in morphology to IPTs, it has the same cytological morphology and immunophenotype as classical FDCS; that is, tumor cells are scattered in an
inflammatory background, including lymphocytes and plasma cells. In our study, IPT-like FDCS tumor cells expressed CD21 and CD35, while partly expressing CD68, smooth muscle actin, vimentin, and clusterin. Studies have shown that almost all cases of IPT-like FDCS are associated with the Epstein–Barr virus [6–9]. All cases in our cohort were positive for CD21, CD35, and the Epstein–Barr encoding region.

FDCS has a low degree of invasion but high local recurrence and metastasis rates, making it a moderately malignant tumor. IPT-like FDCS is more inert, and surgical resection is the first choice of treatment. In patients with tumor recurrence or patients who cannot be treated by surgery, chemotherapy or radiotherapy can be used [10]. All 13 patients showed no tumor recurrence upon follow-up, which is similar to reports from previous studies [11].

To date, most imaging findings had been reported as individual cases or small cohorts; summaries of imaging findings are scarce. To our knowledge, this study represents the largest radiologic-pathologic correlation study of IPT-like FDCS cases. The images of IPT-like FDCS in the liver and spleen revealed both similarities and differences. In both intrahepatic and intrasplenic tumors, the density was often homogeneous and hypodense on CT. We speculate that slow tumor growth, sparse tumor cells, and abundant blood supply may be the reasons behind this observation. Seven of eight cases showed slight hypodensities by CT; only three tumors (>5 cm in diameter) had a central hypodense necrotic area. After contrast medium injection and enhancement of the solid components, the enhancement of IPT-like FDCS in the spleen was found to be weaker than that in the liver, which may be related to the obvious enhancement of the spleen itself. In MRI scans, IPT-like FDCS was more characteristic. Intrasplicenic IPT-like FDCS showed relative hypointensities on T2-weighted images due to the hyperintense signals on T2-weighted images of the spleen itself. Our study confirmed that the pseudocapsule formed at the tumor boundaries in nine cases presented as hypointense signals in the T2-weighted image sequence, because it mainly comprised collagen fibers, which was consistent with other studies [12]. Our study further found that the enhancement of the tumor pseudocapsule in the delayed phase was due to the blood vessel distribution in the pseudocapsule of the tumor. IPT-like FDCS has slightly hyperintense or hyperintense signals on diffusion-weighted imaging, which may be closely related to the higher cell density. The spleen is rich in blood sinuses, which may cause an obvious T2 shine-through effect; the spleen parenchyma has hyperintense signals on diffusion-weighted imaging, whereas the IPT-like FDCS signal in the spleen is relatively lower. The mild-to-moderate contrast enhancement of tumors in the liver may be related to the abundance of micro-vessels and inflammatory cells. The “halo sign” could be seen only in a certain proportion of IPT-like FDCSs of the liver, which is consistent with the incomplete tumor capsule and the infiltration of plasma cells and lymphocytes in the peritumoral edema, as seen under the microscope. IPT-like FDCSs of the spleen displayed homogeneous or heterogeneous hypoechoic lesions consistent with ultrasonographic findings in other studies [3]. The patchy hyperechoic area detected in the tumor was related to the infiltration of many inflammatory cells [13,14]. The abundant strip or short-rod blood flow signals in the periphery, and the circular enhancement around lesions in contrast-enhanced ultrasonography, were consistent with the blood vessel distribution in the pseudocapsule of the tumor in the pathological findings. Therefore, the imaging findings of IPT-like FDCS in the liver and spleen have their own specific characteristics and need to be treated differently in terms of diagnosis.
Although the imaging manifestations of the tumors vary, IPT-like FDCS contrast enhancement can be divided into three types according to the enhancement characteristics, and this is closely related to the tumor's histopathological features. (a) Centrifugal enhancement type: following contrast agent administration, the center of the tumor was substantially enhanced in the arterial phase, further enhanced in the portal and venous phases, the scope of the enhancement was enlarged, and the edge of the tumor was enhanced in the delayed phase. In our study, this type was only found in the liver (Fig. 4). (b) Marginal enhancement type: the boundary of this type was clear on a plain scan, with uniform density or signal intensity on CT and MRI, respectively. Mild enhancement of a thin ring in the arterial and portal phases, followed by continuous enhancement in the delayed phase, was seen. Pathologically, the center of this type lacked blood supply. This tumor type contained many collagen fibers, and a pseudocapsule was sometimes formed by fibrous tissue and infiltrating inflammatory cells (Fig. 5). (c) Grid enhancement type: on plain CT scan, honeycomb-like hypodensities were visible in tumors with a clear boundary. Multiple areas of hypointensities on T1-weighted imaging and hyperintensities on T2-weighted imaging were observed inside the tumor, with an isointense septum. Obvious continuous enhancement changes were observed around the tumor and its internal septum. Hypodense areas on CT indicated necrosis, whereas iso- or slightly hyperdense areas indicated the proliferation of fibrous tissue. This tumor type was often larger, the capsule was incomplete, and its biological behavior suggested that it was more aggressive (Fig. 3).

In summary, IPT-like FDCS is a rare tumor mainly observed in the liver and spleen. By associating imaging and pathology findings, our comparative study of 13 IPT-like FDCS cases identified characteristic imaging features of this type of tumor in the liver and spleen. Although these tumors are usually indolent, surgical resection is still the fundamental treatment and requires postoperative follow-up.

Our study had some limitations. The relatively small sample size is the most important. IPT-like FDCS is an uncommon tumor; moreover, patients with IPT-like FDCS may never undergo diagnostic imaging because they are asymptomatic. Second, owing to the retrospective nature of this study, we did not obtain both multi-detector CT and MR images in the majority of patients. Additionally, different MRI scanners and techniques were used. However, these issues are unavoidable due to the rarity of this tumor type, and this limitation should not have significantly affected the radiological characteristics studied.

Conclusion

We believe a complete CT and MRI examination may be of great help in the diagnosis of IPT-like FDCS. Consideration should be given to a possible diagnosis of IPT-like FDCS when single or multiple masses are found with different patterns of enhancement on CT and MRI, a “halo sign,” the presence of a pseudocapsule appearance, and annular enhancement in the delayed phase.

Abbreviations
CT: Computer tomography   MRI: Magnetic resonance imaging   IPT-like FDCS: Inflammatory pseudotumor-like follicular dendritic cell sarcoma;

**Declarations**

**Ethics approval and consent to participate**

The Institutional Review Board of Ningbo Medical Center Lihuili Hospital approved this study and waived the requirement for written informed consent due to its retrospective nature.

**Consent for publication**

Not applicable.

**Availability of data and materials**

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

**Competing interests**

The authors declare that they have no competing interests.

**Funding**

This study was funded by the Medical and Health Research Foundation of Zhejiang (grant no. 2019KY610). The funding sponsors estimated the study’s feasibility but had no role in data collection, analysis, interpretation of data, or the decision to submit the manuscript for publication.

**Authors’ contributions**

XLB and GSS conceived and designed the study. XLB, GR, and GSS conducted the study and collected important background data. XLB and GSS drafted the manuscript. All authors have read and approved the final manuscript.

**Acknowledgements**

We are grateful to Dr. Rong Ge and his laboratory staff for their valuable contribution to the study, as well as Dr. Feng Fu, Dr. Feng Mao, and Dr. Saijun Chen for reviewing the radiological images used in this study.

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### Tables

**Table1**

**Antibodies of immunostaining**
| Antibodies     | Source    | Dilution | pretreatment |
|---------------|-----------|----------|--------------|
| CD 21         | Dako      | 1:50     | HT           |
| CD 35         | Dako      | 1:25     | PC           |
| CD 23         | Dako      | 1:50     | PC           |
| ALK 1         | Novocastra| 1:25     | HT           |
| CD 30         | Dako      | 1:20     | PC           |
| EMA           | Dako      | 1:50     | HT           |
| CD 68         | Dako      | 1:20     | TP           |
| S-100 protein | Dako      | 1:800    | HT           |
| SMA           | Dako      | 1:500    | No pretreatment |
| desmin        | Dako      | 1:50     | No pretreatment |
| CD 1a         | Novocastra| 1:20     | HT           |
| CD 3          | Dako      | 1:800    | PC           |
| CD 20         | Dako      | 1:1600   | HT           |

HT, heating at 95 °C in citrate buffer 10mmol/L PH 6.0 for 30 min; PC, pressure cooking in EDTA for 2 minutes; TP, trypsinization with 0.1% trypsin for 20 min at room temperature.

Table 2

Comparisons of clinical manifestation of inflammatory pseudotumor-like follicular dendritic cell sarcomas of liver and spleen.
|                                      | Liver (n=2) | Spleen (n=9) | Liver& spleen (n=2) | P-value |
|--------------------------------------|-------------|--------------|---------------------|---------|
| Sex                                  |             |              |                     | 0.706   |
| Male                                 | 2           | 4            | 1                   |         |
| Female                               | 0           | 5            | 1                   |         |
| Clinical manifestation               |             |              |                     | 0.119   |
| Fever or abdominal discomfort        | 1           | 2            | 2                   |         |
| Physical examination                 | 1           | 7            | 0                   |         |
| Median age (years)                   | 67(53–81)   | 59(36–88)    | 56(46–66)           | 0.983   |
| Number of nodules                    |             |              |                     | 0.052   |
| Single                               | 1           | 8            | 0                   |         |
| Multiple                             | 1           | 1            | 2                   |         |

**Table 3**

Clinical information of the inflammatory pseudotumor-like follicular dendritic cell sarcomas in the liver and spleen.
| Case | Age (years) | Sex | Location | Symptoms | Treatment | Follow-up (months) | Examination method |
|------|-------------|-----|----------|----------|-----------|-------------------|--------------------|
| 1    | 81          | Male| Liver    | No       | Excision  | 12                | CT and MRI         |
| 2    | 53          | Male| Liver    | Abdominal distension | Excision  | 24                | MRI, ultrasound    |
| 3    | 76          | Female| Spleen  | No       | Excision  | 10                | MRI                |
| 4    | 49          | Female| Spleen  | No       | Excision  | 24                | CT and MRI         |
| 5    | 73          | Male| Spleen   | No       | Excision  | 18                | CT, ultrasound     |
| 6    | 66          | Female| Liver and spleen | Epigastric pain | Excision  | 20                | CT, ultrasound     |
| 7    | 55          | Male| Spleen   | No       | Excision  | 20                | CT and MRI         |
| 8    | 46          | Male| Liver and spleen | Fever       | Excision  | 24                | CT and MRI Ultrasound |
| 9    | 62          | Female| Spleen  | No       | Excision  | 18                | MRI                |
| 10   | 43          | Female| Spleen  | Abdominal distension | Excision  | 24                | CT and MRI         |
| 11   | 36          | Male| Spleen   | Fever    | Excision  | 24                | CT and MRI         |
| 12   | 41          | Female| Spleen  | No       | Excision  | 17                | MRI                |
| 13   | 88          | Male| Spleen   | No       | Excision  | 12                | MRI                |

CT: computed tomography; MRI: magnetic resonance imaging

**Table 4**

Imaging characteristics of inflammatory pseudotumor-like follicular dendritic cell sarcomasin the liver and spleen.
|                      | Liver       | Spleen     | P-value |
|----------------------|-------------|------------|---------|
| Diameter (cm)        | 3.6±1.1     | 5.2±2.6    | 0.12    |
| Density              |             | 1.0        |         |
| Iso                  | 0(0%)       | 1(14%)     |         |
| Hypo                 | 3(100%)     | 6(86%)     |         |
| T1 signal intensity  |             |            | 0.045*  |
| Hypo                 | 3(100%)     | 2(22%)     |         |
| Hyper                | 0(0%)       | 7(78%)     |         |
| T2 signal intensity  |             |            | 0.045*  |
| Hypo                 | 0(0%)       | 7(78%)     |         |
| Hyper                | 3(100%)     | 2(22%)     |         |
| Enhancement pattern  |             |            | 0.033*  |
| Significantly enhanced| 3           | 1          |         |
| Mildly enhanced      | 1           | 10         |         |
| Pseudocapsule        | 2/3(66%)    | 7/9(78%)   | 0.637   |
| Necrosis             | 1/4(25%)    | 4/11(36%)  | 0.462   |
| “Halo sign”          | 2/3(67%)    | 0/9(0%)    | 0.045*  |

*p<0.05

**Figures**
A 64-year-old male patient with inflammatory pseudotumor-like follicular dendritic cell sarcoma of the liver. (a) Plain computed tomography scan showing a round, slightly hypodense lesion with homogeneous density and no obvious necrosis, calcification, or bleeding in the caudate lobe of the liver. (b) Slight enhancement in the arterial phase. (c) and (d) Decreased enhancement in the portal phase and delayed phase, respectively.

Figure 1
A 64-year-old male patient with inflammatory pseudotumor-like follicular dendritic cell sarcoma of the liver. (a) Plain computed tomography scan showing a round, slightly hypodense lesion with homogeneous density and no obvious necrosis, calcification, or bleeding in the caudate lobe of the liver. (b) Slight enhancement in the arterial phase. (c) and (d) Decreased enhancement in the portal phase and delayed phase, respectively.

Figure 1
Figure 2

A 45-year-old male patient with inflammatory pseudotumor-like follicular dendritic cell sarcoma of the spleen. (a) Plain computed tomography scan showing a round slightly hypodense lesion of the spleen. Homogeneous density, no obvious signs of necrosis, calcification, or bleeding. (b) and (c) After contrast enhancement, the relatively hypodense lesion is observed in all phases. (d) Circular enhancement at the lesion edge in the delayed phase. (e) Ultrasound image, revealing a round-like lesion with clear boundary.
and homogeneous hypoechogenicity in the spleen. (f) Pathological picture of the lesion (hematoxylin and eosin [HE], 100×). A large number of inflammatory cells infiltrate the tumor (black star). The formation of a fibrous pseudocapsule (white arrow) with blood vessels in the fibrous capsule (black arrow) can be seen at the edge of the tumor.

Figure 2
A 45-year-old male patient with inflammatory pseudotumor-like follicular dendritic cell sarcoma of the spleen. (a) Plain computed tomography scan showing a round slightly hypodense lesion of the spleen. Homogeneous density, no obvious signs of necrosis, calcification, or bleeding. (b) and (c) After contrast enhancement, the relatively hypodense lesion is observed in all phases. (d) Circular enhancement at the lesion edge in the delayed phase. (e) Ultrasound image, revealing a round-like lesion with clear boundary and homogeneous hypoechogenicity in the spleen. (f) Pathological picture of the lesion (hematoxylin and eosin [HE], 100×). A large number of inflammatory cells infiltrate the tumor (black star). The formation of a fibrous pseudocapsule (white arrow) with blood vessels in the fibrous capsule (black arrow) can be seen at the edge of the tumor.

Figure 3

A 57-year-old female patient with multiple inflammatory pseudotumor-like follicular dendritic cell sarcomas in the liver and spleen. (a) The plain computed tomography scan reveals round-like hypodense masses with uneven densities in the spleen and liver. (b) After contrast enhancement, the masses show a honeycomb structure, and no enhancement can be seen in the central necrotic area.
Figure 3

A 57-year-old female patient with multiple inflammatory pseudotumor-like follicular dendritic cell sarcomas in the liver and spleen. (a) The plain computed tomography scan reveals round-like hypodense masses with uneven densities in the spleen and liver. (b) After contrast enhancement, the masses show a honeycomb structure, and no enhancement can be seen in the central necrotic area.
A 46-year-old male patient with inflammatory pseudotumor (IPT)-like follicular dendritic cell sarcoma (FDCS) in the liver. (a) The lesion shows a slightly lower signal intensity in the T1-weighted image sequence, and the boundary is not clear. (b) In the T2-weighted image sequence, the lesion is slightly hyperintense, and a “halo sign” can be seen at the edge of the lesion. (c) The lesion shows heterogeneous hyperintense signals in the diffusion-weighted imaging sequence. (d) The lesion shows an obvious central
nodular enhancement in the portal vein phase. (e) Progressive enhancement in the venous phase with relatively low enhancement at the lesion margin. (f) The enhancement of the lesion decreases in the delayed phase and a rim enhancement can be seen at the edge. (g) IPT-like FDCS of the liver, demonstrating abundant infiltration of inflammatory cells and the distribution of capillaries (hematoxylin and eosin [HE], 200×). (h) Small vessels can also be seen in the fibrous pseudocapsule of the tumor (black star; HE, 100×).

Figure 4
A 46-year-old male patient with inflammatory pseudotumor (IPT)-like follicular dendritic cell sarcoma (FDCS) in the liver. (a) The lesion shows a slightly lower signal intensity in the T1-weighted image sequence, and the boundary is not clear. (b) In the T2-weighted image sequence, the lesion is slightly hyperintense, and a "halo sign" can be seen at the edge of the lesion. (c) The lesion shows heterogeneous hyperintense signals in the diffusion-weighted imaging sequence. (d) The lesion shows an obvious central nodular enhancement in the portal vein phase. (e) Progressive enhancement in the venous phase with relatively low enhancement at the lesion margin. (f) The enhancement of the lesion decreases in the delayed phase and a rim enhancement can be seen at the edge. (g) IPT-like FDCS of the liver, demonstrating abundant infiltration of inflammatory cells and the distribution of capillaries (hematoxylin and eosin [HE], 200×). (h) Small vessels can also be seen in the fibrous pseudocapsule of the tumor (black star; HE, 100×).
Figure 5

A 37-year-old male patient with inflammatory pseudotumor-like follicular dendritic cell sarcoma of the spleen. (a) The mass shows a slight hyperintensity in the T1-weighted image sequence. (b) In the T2-weighted image sequence, the tumor shows a slightly hypointense signal, and its edge displays a hypointense annular signal representing the pseudocapsule. (c) The mass shows a slightly hyperintense signal on diffusion-weighted imaging. (d)-(g) The contrast enhancement of the mass is mild in each
phase, and the pseudocapsule shows delayed enhancement. (h) In the histopathological section, the tumor (black arrow) is surrounded by dense fibrous tissue (black star; hematoxylin and eosin, 100×).

(i) Macroscopic examination showing a grayish-white tumor with uniform texture and clear boundaries.

Figure 5

A 37-year-old male patient with inflammatory pseudotumor-like follicular dendritic cell sarcoma of the spleen. (a) The mass shows a slight hyperintensity in the T1-weighted image sequence. (b) In the T2-
weighted image sequence, the tumor shows a slightly hypointense signal, and its edge displays a hypointense annular signal representing the pseudocapsule. (c) The mass shows a slightly hyperintense signal on diffusion-weighted imaging. (d)-(g) The contrast enhancement of the mass is mild in each phase, and the pseudocapsule shows delayed enhancement. (h) In the histopathological section, the tumor (black arrow) is surrounded by dense fibrous tissue (black star; hematoxylin and eosin, 100×). (i) Macroscopic examination showing a grayish-white tumor with uniform texture and clear boundaries.