Case Report

Transient Pneumonitis as a Possible Adverse Reaction to the BNT162b2 COVID-19 mRNA Vaccine in a Patient with Rheumatoid Arthritis: A Case Report and Review of the Literature

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The coronavirus disease (COVID-19) pandemic caused by severe acute respiratory syndrome coronavirus 2 has led to rapid progress in vaccine development to prevent the spread of the disease. Although COVID-19 vaccines have excellent effectiveness in reducing morbidity and disease severity with minor adverse reactions, some patients develop late hypersensitivity events as autoimmune reactions such as rheumatoid arthritis, lupus nephritis, and vasculitis following COVID-19 vaccination [3–6]. Herein, we describe a case of pneumonitis following COVID-19 mRNA vaccination in a patient with rheumatoid arthritis, which resolved spontaneously.

1. Introduction

The development of coronavirus disease (COVID-19) vaccines has progressed rapidly. Several vaccines have been reported to be effective in reducing morbidity and disease severity [1, 2]. In Japan, the mRNA vaccines BNT162b2 (Pfizer-BioNTech) and mRNA-1273 (Moderna) have been approved. Although minor adverse reactions are common, few serious adverse reactions have been reported [1, 2]. In addition, there have been late hypersensitivity events as autoimmune reactions such as rheumatoid arthritis (RA), lupus nephritis, and vasculitis following COVID-19 vaccination [3–6]. Herein, we describe a case of pneumonitis following COVID-19 mRNA vaccine, thought to be due to an autoimmune reaction, in a patient with RA, which resolved spontaneously.

2. Case Presentation

A 76-year-old woman with stable RA developed malaise and a high-grade fever (>39.0 °C) 5 days after receiving a second dose of BNT162b2 mRNA vaccine. Ten days after vaccination, she developed a persistent cough and dyspnea. Seventeen days after vaccination, she was admitted to another hospital because of persistent fever. Blood tests showed a white blood cell (WBC) count of 4990/μL and lactate dehydrogenase (LDH) and C-reactive protein (CRP) levels of 477 U/L and 6.4 mg/dL, respectively. Although the antigen test for SARS, serum B-D glucan, and blood culture was negative, tests for *Mycoplasma*, *Chlamydia*, and sputum culture were not examined. Chest computed tomography (CT) showed infiltration and ground glass opacities (GGOs) in all lung fields (Figure 1(a)). The patient was provisionally diagnosed with interstitial lung disease (ILD) and was transferred to our hospital 21 days after vaccination.

Prior to the onset of her symptoms, the patient’s RA had been stable on methotrexate treatment (12 mg/week), which she had taken for 15 years. On admission, her body temperature, blood pressure, pulse, respiratory rate, and SpO2 were 36.5°C, 110/60 mmHg, 65 beats/min, 18 breaths/min, and 98% (in room air), respectively. Her arthritis was stable and showed no changes of note since a previous check-up one month earlier. The results of blood tests were as follows: WBC, 5420/μL; red
blood cells (RBC), $377 \times 10^4/\mu L$; hemoglobin (Hb), 11.2 g/dL; hematocrit, 35.5%; platelets, $40.4 \times 10^4/\mu L$; CRP 1.47 mg/dL; LDH, 311 U/L; Krebs von den Lungen-6 (KL-6), 452.4 U/mL; and surfactant protein D (SP-D), 226 ng/mL. Her rheumatoid factor, anti-cyclic citrullinated peptide antibody, and antinuclear antibody were negative.

Tests for Mycoplasma, Chlamydia, and tuberculosis (T-Spot), and antigen tests for Cryptococcus, Candida, Aspergillus, cytomegalovirus antigen, and SARS-CoV-2 antigen were negative. Serum β-D glucan, blood culture, and polymerase chain reaction testing of respiratory samples for Pneumocystis jirovecii DNA were also negative. Chest CT showed a significant improvement compared to the chest CT performed 4 days earlier (Figure 1(b)). The patient was treated conservatively and did not receive antibiotics or immunosuppressive drugs during hospitalization. Her cough, dyspnea, and fever resolved spontaneously, and she was discharged from hospital after 7 days. Her final diagnosis was transient pneumonitis induced by a COVID-19 mRNA vaccine. At the 3-month follow-up, she had not experienced a relapse of pneumonitis and her blood test results had improved: CRP, 0.06 mg/dL (from 1.47 mg/dL); LDH, 189 U/L (from 311 U/L); KL-6, 165.1 U/mL (from 452.4 U/mL); and SP-D, 102 ng/mL (from 226 ng/mL).

3. Discussion

To the best of our knowledge, this is the first report of transient pneumonitis following COVID-19 mRNA vaccination in a patient with RA. The patient’s CT scan showed bilateral GGOs, which was consistent withILD. The diagnosis of ILD in patients with RA is challenging because ILD has several causes, including bacterial pneumonia, viral pneumonia, Pneumocystis pneumonia, RA, and certain drugs. Drug-induced ILD is particularly difficult to diagnose because the clinical, radiological, and histological findings are non-specific and other causes of ILD must be excluded [11]. Bronchoscopy is required to diagnose drug-induced ILD. However, the Japan Society for Respiratory Endoscopy recommends that unnecessary bronchoscopy should be avoided during the COVID-19 pandemic [7]. In our patient, we did not perform bronchoscopy based on the Japan Society for Respiratory Endoscopy recommendation because blood tests did not show any sign of infection, including Mycoplasma, Chlamydia, or Pneumocystis infection, and her blood culture was negative. The patient had been taking the same drugs for 15 years, and the only new medication that she had received was the COVID-19 mRNA vaccine. Furthermore, her symptoms developed 5 days after the vaccination and the pneumonitis improved spontaneously with decreasing serum KL-6 and SP-D, without any relapse. These findings suggest that the COVID-19 mRNA vaccine induced transient pneumonitis. However, several investigators reported that the serum KL-6 and SP-D is associated with pulmonary diseases with alveolar inflammation, not only in ILD but also in respiratory infections, such as Pneumocystis pneumonia, COVID-19, and viral bronchitis [8–10, 12]. We could not exclude all respiratory diseases including infectious pneumonia because of the lack of bronchoscopic confirmation and tests for all respiratory infections with increasing KL-6 and SP-D.

The BNT162b2 mRNA vaccine contains RNA, lipids including polyethylene glycol (PEG), and a buffer, but unlike
other vaccines, it contains no adjuvants or preservatives. Compared to conventional vaccines, mRNA vaccines have several advantages: they are non-infectious, do not contain cellular components, induce cellular immunity, do not require adjuvants, are simple to produce, and are inexpensive [2, 13]. The spike protein of the virus adsorbs to cell membranes covered with lipid bilayers and receptor-binding proteins on the body’s immune cells, which react with these proteins to produce neutralising antibodies. PEG, one of the components of the vaccine, can cause anaphylactic reactions [14]. The overall risk of anaphylaxis following vaccination with COVID-19 mRNA vaccines is low, and almost all adverse reactions are minor [15]. However, severe adverse reactions, including ILD, have been reported following vaccination [16,17].

We conducted a literature search for studies published up to October 2021 to determine the characteristics of patients who developed pneumonitis after the COVID-19 mRNA vaccination and found four case reports (Table 1) [16–19]. All patients had received the BNT162b2 vaccine. Of the five patients (the four previously reported cases and our patient), pneumonitis developed in two patients after the first dose and in three patients after the second dose. All patients developed the symptoms within five days after vaccination. The median age was 66 years (range, 60–86 years), and four of the five patients were male. Chest CT showed bilateral GGOs in all patients. Four patients received immunosuppressive therapy, and all patients survived. Our patient developed symptoms five days after vaccination with the BNT162b2 vaccine, which is consistent with the other cases. Notably, unlike the other cases, our patient’s pneumonitis resolved spontaneously without the use of immunosuppressive therapy.

The mechanism of pneumonitis development following COVID-19 vaccination is unclear. In contrast, in patients with pneumonitis after influenza vaccination, two major mechanisms are involved: cytotoxicity of the vaccine itself and an autoimmune response to the vaccine [20]. As with patients who develop pneumonitis following COVID-19 vaccination, almost all patients with pneumonitis after influenza vaccination develop bilateral GGOs on chest CT within 10 days and recover after immunosuppressive therapy [16,21]. The risk factors for developing pneumonitis after vaccination are unknown. According to the World Health Organization, there is no association between vaccination and the development of autoimmune diseases in patients with no underlying disease [22]. Several investigators have reported that systemic rheumatic diseases (SRD) did not worsen after influenza, pneumococcal, or human papillomavirus vaccination [23–25]. According to the 2019 European League against Rheumatic Diseases (EULAR) recommendations for vaccination in adult patients with autoimmune inflammatory rheumatic diseases, influenza vaccination is recommended for patients with RA [26]. In addition, the American College of Rheumatology and EULAR recommend COVID-19 vaccination for patients with SRD.

It is difficult to predict severe adverse reactions to COVID-19 mRNA vaccines because they are the first mRNA vaccines to be developed. Although serious adverse reactions, such as thrombosis with thrombocytopenia, myocarditis, and Guillain–Barré syndrome, have been reported after COVID-19 vaccination [27–29], risk factors for these adverse reactions have not been identified. Further prospective studies are warranted to investigate severe adverse reactions following COVID-19 vaccination, including pneumonitis.

In conclusion, COVID-19 mRNA vaccines may induce autoimmune diseases, including pneumonitis, and clinicians should be aware of the possibility of these diseases following COVID-19 vaccination.

| Case | 1 | 2 | 3 | 4 | 5 |
|------|---|---|---|---|---|
| Age (years) | 86 | 65 | 60 | 66 | 76 |
| Sex | M | M | M | M | F |
| Comorbidities | HT, DM, CKD | OMI, HL, HT | ACO, HT | None | RA, HT, myoma |
| Smoking | Never | Ex-smoker | Ex-smoker | NS | Never |
| Allergy history | NS | None | None | NS | None |
| Vaccine type | Pfizer-BioNTech | Pfizer-BioNTech | Pfizer-BioNTech | Pfizer-BioNTech | Pfizer-BioNTech |
| Dose | First | First | Second | Second | Second |
| Time from vaccination to symptom onset (days) | 1 | 5 | 4 | 2 | 5 |
| Symptoms | Fever, cough, dyspnea | Fever, dyspnea | Dyspnea | Fever, cough, dyspnea | Fever, cough |
| CRP (mg/dL) | 11.4 | 5.5 | 10.9 | 8.7 | 6.4 |
| Bilateral GGO on chest CT | Yes | Yes | Yes | Yes | Yes |
| Immunosuppressive therapy | Yes | Yes | Yes | Yes | No |
| Outcome | Survived | Survived | Survived | Survived | Survived |
| Reference | [7] | [8] | [9] | [10] | Current case |

Note. ACO: asthma and chronic obstructive pulmonary disease overlap; CKD: chronic kidney disease; CRP: C-reactive protein; CT: computed tomography; DM: diabetes mellitus; F: female; GGO: grand glass opacity; HL: hyperlipidemia; HT: hypertension; ILD: interstitial lung disease; M: male; NS: not specified; OMI: old myocardial infarction; RA: rheumatoid arthritis.

Table 1: Characteristics of patients with ILD after COVID-19 vaccination.
Data Availability

All relevant data are included within the article.

Consent

Written informed consent for this case report has been obtained from the patient.

Conflicts of Interest

The authors have no conflicts of interest to declare.

Authors’ Contributions

All authors approved the final version of this article. YO had full access to all of the data. SO was responsible for the organization and coordination of the case.

References

[1] F. P. Polack, S. J. Thomas, N. Kitchin et al., “Safety and Efficacy of the BNT162b2 mRNA Covid-19 vaccine,” New England Journal of Medicine, vol. 383, pp. 2603–2615, 2020.
[2] L. R. Baden, H. M. El Sahly, B. Essink et al., “Efficacy and safety of the mRNA-1273 SARS-CoV-2 vaccine,” New England Journal of Medicine, vol. 384, no. 5, pp. 403–416, 2021.
[3] C. Baimukhamedov, S. Makhmudov, and A. Botabekova, “Seropositive rheumatoid arthritis after vaccination against SARS-CoV-2 infection,” International Journal of Rheumatic Diseases, vol. 24, no. 11, pp. 1440-1441, 2021.
[4] M. F. Zavala-Miranda, S. G. González-Ibarra, A. A. Pérez-Arias, N. O. Uribe-UrIBE, and J. M. Mejía-Vilet, “New-onset systemic lupus erythematosus beginning as class V lupus nephritis after COVID-19 vaccination,” Kidney International, vol. 100, no. 6, pp. 1340-1341, 2021.
[5] A. Prabhahar, G. S. R. S. N. K. Naidu, P. Chauhan et al., “ANCA-associated vasculitis following ChAdOx1 nCoV19 vaccination: case-based review,” Rheumatology International, vol. 42, no. 4, pp. 749–752, 2022.
[6] M. E. Grossman, G. Appel, A. J. Little, and C. J. Ko, “Post-COVID-19 vaccination IgA vasculitis in an adult,” Journal of Cutaneous Pathology, vol. 49, no. 4, pp. 385–387, 2022.
[7] The Japan Society for Respiratory Endoscopy, “Urgent Recommendations on Coronavirus Disease on 2019 (COVID-19),” 2019, http://www.jsre.org/info/200217_corona_teigen.pdf.
[8] D. T. Arnold, C. Donald, M. Lyon et al., “Krebs von den Lungen 6 (KL-6) as a marker for disease severity and persistent radiological abnormalities following COVID-19 infection at 12 weeks,” PLoS One, vol. 16, no. 4, Article ID e0249607, 2021.
[9] N. Nathani, G. D. Perkins, W. Tunnicliffe, N. Murphy, M. Manji, and D. R. Thickett, “Krebs von Lugenh 6 antigen is a marker of alveolar inflammation but not of infection in patients with acute respiratory distress syndrome,” Critical Care, vol. 12, no. 1, 2008.
[10] H. Nakamura, M. Tateyama, D. Tasato et al., “Clinical utility of serum beta-D-glucan and KL-6 levels in Pneumocystis jirovecii pneumonia,” Internal Medicine, vol. 48, no. 4, pp. 195–202, 2009.
[11] M. Schwablmaier, W. Behr, T. Haeckel, B. Märkl, W. Foegg, and T. Berghaus, “Drug induced interstitial lung disease,” The Open Respiratory Medicine Journal, vol. 6, pp. 63–74, 2012.
[12] Y. Kawasaki, Y. Aoyagi, Y. Abe et al., “Serum KL-6 levels as a biomarker of lung injury in respiratory syncytial virus bronchiolitis,” Journal of Medical Virology, vol. 81, no. 12, pp. 2104–2108, 2009.
[13] A. Wadhwa, A. Aljabbari, A. Lokras, C. Foged, and A. Thakur, “Opportunities and challenges in the delivery of mRNA-based vaccines,” Pharmaceutics, vol. 12, no. 2, p. E102, 2020.
[14] K. G. Blumenthal, L. B. Robinson, C. A. Camargo et al., “Acute allergic reactions to mRNA COVID-19 vaccines,” JAMA, vol. 325, no. 15, pp. 1562–1565, 2021.
[15] M. H. Kim, S. Y. Lee, S. E. Lee et al., “Anaphylaxis to iodinated contrast media: clinical characteristics related with development of anaphylactic shock,” PLoS One, vol. 9, no. 6, Article ID e100154, 2014.
[16] J. Y. Park, J. H. Kim, I. J. Lee et al., “COVID-19 vaccine-related interstitial lung disease: a case study,” Thorax, vol. 77, no. 1, pp. 102–104, 2022.
[17] S. Matsuizaki, H. Kamiya, I. Inohishima, Y. Hirasesawa, O. Tago, and M. Arai, “COVID-19 mRNA vaccine-induced pneumonitis,” Internal Medicine, vol. 61, no. 1, pp. 8310–8316, 2022.
[18] A. Yoshifuji, K. Ishioka, Y. Masuzawa et al., “COVID-19 vaccine induced interstitial lung disease,” Journal of Infection and Chemotherapy, vol. 28, no. 1, pp. 95–98, 2022.
[19] A. Kono, R. Yoshioka, P. Hawk et al., “A case of severe interstitial lung disease after COVID-19 vaccination,” QJM: International Journal of Medicine, vol. 114, no. 11, pp. 805–806, 2022.
[20] O. Matsuno, “Drug-induced interstitial lung disease: mechanisms and best diagnostic approaches,” Respiratory Research, vol. 13, 2012.
[21] T. Okusaki and K. Fukuwara, “Exacerbation of connective tissue disease-associated interstitial lung disease due to influenza vaccination,” Respiratory Medicine Case Reports, vol. 33, Article ID 101463, 2021.
[22] L. Conklin, A. Hviid, W. A. Orenstein, A. J. Pollard, M. Wharton, and P. Zuber, “Vaccine safety issues at the turn of the 21st century,” BMJ Glob Health, vol. 6, no. Suppl 2, Article ID e004898, 2021.
[23] A. Chalmers, D. Scheifele, C. Patterson et al., “Immunization of patients with rheumatoid arthritis against influenza: a study of vaccine safety and immunogenicity,” Journal of Rheumatology, vol. 21, no. 7, pp. 1203–1206, 1994.
[24] F. Del Porto, B. Laganà, R. Biselli et al., “Influenza vaccine administration in patients with systemic lupus erythematosus and rheumatoid arthritis. Safety and immunogenicity,” Vaccine, vol. 24, no. 16, pp. 3217–3223, 2006.
[25] O. Elkahay, D. Paran, D. Caspi et al., “Immunogenicity and safety of pneumococcal vaccination in patients with rheumatoid arthritis or systemic lupus erythematosus,” Clinical Infectious Diseases, vol. 34, no. 2, pp. 147–153, 2002.
[26] V. Furer, C. Rondaan, M. W. Heijstek et al., “2019 update of EULAR recommendations for vaccination in adult patients with autoimmune inflammatory rheumatic diseases,” Annals of the Rheumatic Diseases, vol. 79, no. 1, pp. 39–52, 2020.
[27] H. Akiyama, S. Kakiuchi, J. Rikitake et al., “Immune thrombocytopenia associated with Pfizer-BioNTech’s BNT162b2 mRNA COVID-19 vaccine,” IDCases, vol. 25, Article ID e01245, 2021.

[28] B. Singh, P. Kaur, L. Cedeno et al., “COVID-19 mRNA vaccine and myocarditis,” European Journal of Case Report International Medicine, vol. 8, no. 7, Article ID 002681, 2022.

[29] M. García-Grimshaw, A. Michel-Chávez, J. M. Vera-Zertuche et al., “Guillain–Barré syndrome is infrequent among recipients of the BNT162b2 mRNA COVID-19 vaccine,” Clinical Immunology, vol. 230, Article ID 108818, 2021.