**Lactobacillus plantarum ZLP001: In vitro Assessment of Antioxidant Capacity and Effect on Growth Performance and Antioxidant Status in Weaning Piglets**

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**ABSTRACT:** The objective of this study was to evaluate the antioxidant capacity of *Lactobacillus plantarum* ZLP001 and its effects on growth performance and antioxidant status in weaning piglets. The survival in hydrogen peroxide and free radical-scavenging activity of *Lactobacillus plantarum* ZLP001 were analysed in vitro. The *Lactobacillus plantarum* ZLP001 showed high viability in 1.0 mmol/L hydrogen peroxide and high scavenging ability against hydroxyl, superoxide anion, and DPPH (1,1-diphenyl-2-picrylhydrazyl) radicals which was dose dependent. Ninety-six weaning piglets were selected (7.45±0.79 kg) and divided into three groups comprising of negative control without any supplementation, treatment group with supplemented 6.8×10^7 *Lactobacillus plantarum* ZLP001 CFU/g of diet, and positive control with antibiotic treatment (chlorotetracycline, 80 mg/kg diet). The results showed that *Lactobacillus plantarum* ZLP001 supplementation enhanced feed conversion rates in piglets compared with control (p<0.05). Supplementation of *Lactobacillus plantarum* ZLP001 increased the concentration of superoxide dismutase (p<0.05), glutathione peroxidase (p<0.01) and catalase in serum (p<0.10), while decreased the concentration of malondialdehyde (p<0.05). The present study implies that the strain *Lactobacillus plantarum* ZLP001 had high antioxidant ability and its supplementation improved the growth performance and antioxidant status of weaning piglets, so it can be considered useful to alleviate oxidative stress and increase productive performance of pigs. (Key Words: Probiotic, Antioxidant Capacity, Weaning Piglet)

**INTRODUCTION**

Piglets are faced with many new stressors during the post-weaning period which can lead to risk of diarrhea, reduced growth rate, changes in gut morphology and microbiology, and increased susceptibility to disease and death (Hampson, 1994). Many studies have shown that direct-fed *Lactobacillus* preparation plays an important role in the improving of gut microflora balance and consequently in the better health condition of piglets. The functional effects of *Lactobacillus* such as protection against infections, stimulation of immune system, reduction of incidence of diarrhea, and improvement of production performance have been demonstrated in many studies (Ouwehand et al., 2002; Koninkx and Malago, 2008).

Several in vivo studies have demonstrated that some *Lactobacillus* strains can improve the antioxidant status of rats and humans (Kaizu et al., 1993; Kullisaar et al., 2003). Therefore, many investigations have focused on antioxidant properties of lactic acid bacteria and their role in health and disease recently (Ito et al., 2003; Kim et al., 2006). There are only few studies about the effects of *Lactobacillus* on the antioxidant status of pigs (Wang et al., 2009), especially on weaning piglets. However, it is known that an abnormality in the antioxidant defence system can increase the susceptibility of pigs to stress, resulting in decreased performance and reduced immune function (Duthie et al., 1989; Lauridsen et al., 1999).

The main objective of this study was to estimate the viability in hydrogen peroxide and the free radicals scavenging ability of *Lactobacillus plantarum* ZLP001 isolated from gastrointestinal mucosa of a healthy weaning piglet. In addition, the effects of this *Lactobacillus* strain on weaning piglet production performance and oxidant status were also determined.
MATERIALS AND METHODS

Bacterial strain
A probiotic lactic acid bacterial strain, Lactobacillus plantarum ZLP001 was originally isolated from the gastrointestinal tract of a healthy weaning piglet in our laboratory. The strain was identified through standard morphological, biochemical and physiological tests, and by 16s rRNA gene sequence analysis by the China Center of Industrial Culture Collection (Beijing, China). Stock cultures in MRS broth were mixed with 20% of sterilized glycerol (v/v) at a concentration of 4:1 and stored at -80°C.

Resistance to hydrogen peroxide
The Lactobacillus plantarum ZLP001 (concentration of 10⁵ CFU/ml) was incubated with 1.0 mmol/L hydrogen peroxide at 37°C. The hydrogen peroxide concentration was determined by spectrophotometer at 230 nm absorption using the molar extinction coefficient for hydrogen peroxide. At 30-min time intervals, the aliquots were removed and plated onto MRS plates after serial ten-fold dilution and the number of viable cells was estimated. All plates were incubated at 37°C for 48 h. The effect of 1.0 mmol/L hydrogen peroxide on the survival of Lactobacillus plantarum ZLP001 was tested at 30-min time intervals within 240 min.

Determination of scavenging capacity
The scavenging activities of Lactobacillus plantarum ZLP001 were assessed against the hydroxyl radical, superoxide radical, and DPPH. To this aim the Lactobacillus plantarum ZLP001 cells were harvested after 20 h of incubation. The bacterial cells were washed three times with isotonic saline and re-suspended in 0.9% NaCl. The bacterial counts in the cell pellet were adjusted to 10⁷, 10⁸, 10⁹, and 10¹⁰ CFU/ml.

The scavenging capacity of hydroxyl radical was estimated according to the method of de Avellar et al. (2004) modified as follows: 1 ml of 1,10-phenanthroline (0.75 mmol/L), 2 ml of phosphate buffer (pH 7.4) and 1 ml of FeSO₄ (0.75 mmol/L) were thoroughly mixed. Then, 1 ml of hydrogen peroxide (0.12%) and 1 ml of the Lactobacillus plantarum ZLP001 intact cells were added. The mixture was incubated at 37°C for 90 min, and the absorbance was measured at 536 nm. The scavenging ability was measured as follows: (∆A₄₀₀-∆A₅₁₄)/∆A₅₁₄×100; where ∆A₄₀₀ represents the absorbance of the control solution including 1,10-phenanthroline, FeSO₄, and hydrogen peroxide; ∆A₅₁₄ represents the absorbance of the blank solution including 1,10-phenanthroline and FeSO₄.

The assessment of the scavenging capacity of superoxide anion radical by Lactobacillus plantarum ZLP001 was carried out according to the methods of Zhao et al. (2003) modified as follows: the Tris-HCl buffer (1 mol/L, pH 8.2, including 0.1 mmol/L EDTA) 3 ml and Lactobacillus plantarum ZLP001 intact cells 1 ml were mixed and incubated for 25 min in 25°C water bath. Then, 40 μL of 45 mmol/L pyrogallol preheated at 25°C was added. The controls included only Tris-HCl buffer and deionized water. The absorbance of the sample and control were measured at 325 nm every 30 s for 5 min. The scavenging activity was measured as follows: (∆A₄₇₀-∆A₄₅₀)/∆A₄₇₀×100; where ∆A₄₇₀ and ∆A₄₅₀ are the speed of pyrogallol autoxidation before and after the addition of the sample and deionized water, respectively.

The scavenging of DPPH by Lactobacillus plantarum ZLP001 was analyzed by a modified method utilized by Shimada et al. (1992). Eight tenths of a milliliter of intact cells and 1 ml of freshly prepared DPPH solution (0.2 mmol/L in ethanol) were mixed and reacted in the dark for 30 min. Blank samples contained deionized water. The scavenging DPPH was then monitored by measuring the decrease in absorbance at 517 nm after centrifugation at 6,000 g for 10 min. The scavenging ability was defined as follows: (1-A₅₁₇(sample)/A₅₁₇(blank))×100.

In vivo assessment of antioxidant effects in weaning piglets
Ninety-six piglets (Landrace×Yorkshire, 48 males and 48 females) with 7.45±0.79 kg initial body weight (BW) were selected from Beijing Jingdongyu Farm (Beijing City, China). The pigs were weaned at 28 d of age and randomly allotted to three groups by initial BW. Four replicates per treatment were made and eight pigs per pen were allocated. Each pen was 1.65×1.45 m² with mesh floor, a feeder and a water nipple. The pig barn was maintained at 25 to 28°C. All pigs had free access to feed and water throughout the 4-wk feeding trial. The basal diet mainly contained maize and soybean meal, and the nutrient contents met or exceeded nutrient requirements recommended by NRC (1998). The dietary treatments consisted of the basal diet with no additive served as a negative control, the basal diet with antibiotic (chlorotetracycline, 80 mg/kg diet) served as a positive control, and the basal diet with freeze dried Lactobacillus plantarum ZLP001 at 6.8×10⁷ CFU/g of diet. Details on the ingredient composition and chemical analysis of the diets are shown in Table 1.

Piglets were weighed and the feed intake per pen was recorded every week in order to calculate the average daily weight gain, average daily feed intake, and feed conversion ratio. At the end of the experiment, blood was collected from four pigs per pen (pig closer to the average weight of the pen) by anterior vena cava puncture using 10-ml anticoagulant-free vacutainer tubes. The serum was
**Results**

**Effect of hydrogen peroxide on the survival of Lactobacillus plantarum ZLP001**

As showed in Figure 1, the survival of Lactobacillus plantarum ZLP001 decreased along with the incubation time, however Lactobacillus plantarum ZLP001 still maintained over 95% of the original viable cell numbers after 240 min of incubation (8.46 vs 8.11 log CFU/ml at 0 and 240 min, respectively).

Free radical scavenging activities of Lactobacillus plantarum ZLP001

The scavenging activities of Lactobacillus plantarum ZLP001 against the hydroxyl radical, superoxide radical, and DPPH are shown in Figure 2. It can be seen that the free radical scavenging activity of Lactobacillus plantarum ZLP001 against all three radicals increased in dose-dependent manner. In particular, Lactobacillus plantarum ZLP001 exhibited obvious inhibitory effects especially on the generation of the hydroxyl radical (83.6% of scavenging activity) and superoxide radical (78.5% of scavenging activity) with the 10⁰ CFU/ml concentration. For the DPPH radical, Lactobacillus plantarum ZLP001 showed lower scavenging activity than the other two radicals. Only 51.1% scavenging activity was exhibited at 10⁹ CFU/ml and...
almost no scavenging activity at 10⁶ CFU/ml.

**Effect of Lactobacillus plantarum ZLP001 on growth performance of weaning piglets**

The effects of dietary *Lactobacillus plantarum* ZLP001 supplementation on growth performance of weaning piglets are shown in Table 2. Over the 4 wks feeding trial, piglets of antibiotic and probiotics groups were consumed higher feeds than no additive group (p<0.05). The piglets received diets containing *Lactobacillus plantarum* ZLP001 supplements had the same daily weight gain compared with the antibiotic group (p>0.05) and significant higher than no additive group (p<0.05). The feed conversion ratio in probiotic and antibiotic group were significantly lower (p<0.05) than those of no additive group.

**Effect of Lactobacillus plantarum ZLP001 on antioxidant status in piglets serum**

The effects of *Lactobacillus plantarum* ZLP001 supplementation on antioxidant enzyme activity and malondialdehyde levels in serum of piglets are shown in Table 2. The activity of superoxide dismutase and glutathione peroxidase in *Lactobacillus plantarum* ZLP001 supplementation was significantly higher (p<0.05 and p<0.01, respectively) compared with antibiotic and no additive treatments. The malondialdehyde concentration was significantly decreased (p<0.05) for pigs fed *Lactobacillus plantarum* ZLP001 compared with pigs fed no additive feed. There were no significant differences (p>0.05) among the treatments regarding the activities of catalase in serum of piglets.

**DISCUSSION**

*Lactobacillus* plays significant role in the natural microflora of the pig intestinal tract. They create a healthy balance between beneficial and potentially harmful microorganisms in the gut ecosystem when they are present in sufficient numbers. The use of these cultures in pig diets is desirable for their potential probiotic effects.

The *Lactobacillus plantarum* ZLP001 had high viability in the presence of 1.0 mmol/L hydrogen peroxide. In the present study, the cells were viable exceed 95% after 240 min of incubation. It was not unexpected that *Lactobacillus* cells have high viability in hydrogen peroxide, because hydrogen peroxide is one of the antimicrobial compounds produced by *Lactobacilli* when it plays a role in the gastrointestinal tract (Ouwehand and Vesterlund, 1998). Our result was similar with the findings of Wang et al. (2009) who found that over 90% of *Lact. fermentum* cells were viable after incubation in an environment of 1.0 mmol/L hydrogen peroxide. However, other researchers also found that *Lactobacilli* have high viability only 180 min in the presence of 1.0 mmol/L hydrogen peroxide (Kullisaar et al., 2002). The reason for this discrepancy may be due to differences of *Lactobacillus* strains used or the initial number of *Lactobacillus*.

According to free radical chain reaction proposed by Farmer et al. (1942), free radicals formed in the process of incomplete reduction of oxygen can attack and damage biological molecules. The consequences of a free radical chain reaction can result in serious damage to living organisms. The production of free radicals and many diseases are closely related (Halliwell and Gutteridge, 1984; Halliwell and Gutteridge, 1989). Therefore, the ability of *Lactobacillus plantarum* ZLP001 to scavenge hydroxyl, superoxide, and DPPH radicals were determined in this study. The *Lactobacillus plantarum* ZLP001 showed significant ability to scavenge the hydroxyl, superoxide, and DPPH radicals and supported previous findings on scavenging ability of *Lactobacillus* to free radicals in *vitro* (Lin and Chang, 2000; Kullisaar et al., 2002). The effect of scavenging on these three radicals were all showed to be dose dependent, in fact with the concentration of 10⁶ SEM.

**Table 2.** Effect of *Lactobacillus plantarum* ZLP001 supplementation on growth performance of weaning piglets and superoxide dismutase, glutathione peroxidase, malondialdehyde, and catalase concentrations in the serum

| Parameters                        | Dietary treatment       | SEM* | p value |
|-----------------------------------|-------------------------|------|---------|
|                                   | No additive             |      |         |
|                                   | Antibiotic              |      |         |
|                                   | *Lactobacillus plantarum* ZLP001 (6.8×10⁷ CFU/g of diet) | | |
| Growth performance                |                         |      |         |
| Average daily feed intake (g/d)   | 612ᵇ                   | 651ᵃ | 660ᵃ    | 13.1 | 0.035 |
| Average daily gain (g/d)          | 381ᵇ                   | 419ᵃ | 436ᵇ    | 10.2 | 0.022 |
| Feed conversion ratio             | 1.61ᵃ                  | 1.5ᵇ | 1.5ᵇ²   | 0.019 | 0.017 |
| Antioxidant status                |                         |      |         |
| Superoxide dismutase (U/ml)       | 112.9ᵇ                 | 122.2ᵇ | 139.8ᵃ   | 5.52 | 0.043 |
| Glutathione peroxidase (U/ml)     | 401.5ᶜ                 | 432.1ᵇ | 489.2ᵃ   | 7.15 | 0.007 |
| Malondialdehyde (nmol/ml)         | 6.23ᵃ                  | 5.49ᵇ | 4.10ᵇ   | 0.482 | 0.025 |
| Catalase (U/ml)                   | 5.16                   | 5.71  | 6.06    | 0.386 | 0.092 |

* Standard error of the mean.ᵃᵇᶜ: Means with different superscript within the same row differ significantly (p<0.05).
CFU/ml consistently exhibiting the highest and with the concentration of $10^6$ CFU/ml consistently exhibiting the lowest scavenging ability. This tendency was consistent with studies by Wang et al. (2009) using Lact. Fermentum.

The in vitro studies for evaluating Lactobacillus antioxidant properties are far from the requirements in practice. The strain for the in vivo trial was selected primarily for the survival in environment of hydrogen peroxide and free radical scavenging activities. In this study, the selected Lactobacillus plantarum ZLP001 showed high viability in hydrogen peroxide and high free radical scavenging ability. Therefore, the in vivo trial was carried out to estimate the effects of Lactobacillus plantarum ZLP001 supplementation in weaning piglets. It was demonstrated that weaning in piglets is related to oxidative stress (Sauerwein et al., 2007). This stress can result in reduced performance and increased susceptibility to disease and death (Hampson, 1994; Cromwell, 2002). In this study, Lactobacillus plantarum ZLP001 supplementation group showed significant higher productivity effect compared with the control group. These results indicated that in terms of feed consumption, the probiotic groups consumed 2.3% and 6.1% less than the antibiotic group and no additive group to achieve the same weight respectively. Similar observations were obtained by Francisco et al. (1995) and Chang et al. (2001) that selected probiotic strains had increasing effect on feed conversion rate in piglets. This may be partly owing to the antioxidative effect of Lactobacilli strains that alleviating the oxidative stress of weaning piglets.

The antioxidant systems in the body including numerous antioxidant enzymes (i.e. superoxide dismutase, glutathione peroxidase, and catalase) that are employed to protect the body from oxidative stress (Zhan et al., 2006). The superoxide dismutase enzyme is used to ending the cascade of electron stealing which were caused by the superoxide radicals, and then alleviate the damage to the cell. Superoxide radicals are converted by superoxide dismutase to less toxic hydrogen peroxide which is further processed to form non-toxic water by either glutathione peroxidase or catalase. In this study, the antioxidant status of weaning piglet was evaluated through measuring these enzymes in serum. At the end of the experiment phase, the superoxide dismutase and glutathione peroxidase in the serum of pigs fed Lactobacillus plantarum ZLP001 was significantly higher (p<0.01) than for pigs fed the antibiotic diet and basal diet, which consisted with the studies by Wang et al. (2009) on growing-finishing pigs using Lact. fermentum. There was no significant difference (p>0.05) among three treatments on catalase concentration in the serum in this study. The serum catalase levels were 17.4% higher for piglets fed Lactobacillus plantarum ZLP001 than for piglets fed the negative control. The high activities of these enzymes will decrease the concentrations of oxygen radicals in body, thereby alleviating the oxidative damage to living organisms. Furthermore, malondialdehyde is also one of the most frequently used indicators of oxidative stress in an organism (Janero, 1990; Nielsen et al., 1997). Cellular biomembranes are one of the major targets of reactive oxygen species, where they induce lipid peroxidation resulting in the production of malondialdehyde, while malondialdehyde can react with biomolecules and exert cytotoxic and genotoxic effects (Ardestani and Yazdanparast, 2007). In this study, it was observed that malondialdehyde levels in serum of weaning piglets were significantly reduced (p<0.05) for piglets fed Lactobacillus plantarum ZLP001 compared with the basal diet. These results demonstrated that there was improvement in the antioxidant defence system for weaning piglets fed Lactobacillus plantarum ZLP001.

**CONCLUSION**

The results of our study suggest the antioxidative potential of Lactobacillus plantarum ZLP001. The strain used in the present study exhibited significant in vitro free radical-scavenging capacity, which was dose dependent. Supplementation in the diet improved the growth performance and antioxidant status of pigs as exhibited by improved feed conversion ratio and increased antioxidant enzymes, and decreased malondialdehyde concentration in the serum. This study suggests that Lactobacillus plantarum ZLP001 is a highly antioxidative bacterial strain, which could be used in the future to alleviate oxidative stress and thereby increase growth performance. However, the exact mechanism through which Lactobacillus plantarum may play an important role in scavenging of free radical remains uncertain and further research is also needed to fully determine the exact antioxidative mechanism of this strain through which the antioxidative effects are achieved.

**REFERENCES**

Ardestani, A. and R. Yazdanparast. 2007. Antioxidant and free radical scavenging potential of Achillea santolina extracts. Food Chem. 104:21-29.

Chang, Y. H., J. K. Kim, H. J. Kim, W. Y. Kim, Y. B. Kim and Y. H. Park. 2001. Selection of a potential probiotics Lactobacillus strain and subsequent in vivo studies. Antonie Van Leeuwenhoek 80:193-199.

Cromwell, G. L. 2002. Why and how antibiotics are used in swine production. Anim. Biotechnol. 13:7-27.

de Avellar, I. G., M. M. Magalhaes, A. B. Silva, L. L. Souza, A. C. Leitao and M. Hermes-Lima. 2004. Reevaluating the role of 1,10-phenanthroline in oxidative reactions involving ferrous ions and DNA damage. Biochim. Biophys. Acta. 1675:46-53.

Duthie, G. G., J. R. Arthur, F. Nicol and M. Walker. 1989. Increased indices of lipid peroxidation in stress-susceptible
pigs and effects of vitamin E. Res. Vet. Sci. 46:226-230.
Farmer, E. H., G. F. Bloomfield, A. Sundralingam and D. A. Sutton. 1942. The course and mechanism of autoxidation reactions in olefinic and polyolefinic substances, including rubber. Trans. Faraday Soc. 38:348-356.
Francisco, T., R. Juan, F. Erenia and L. R. Maria. 1995. Response of piglets to oral administration of lactic acid bacteria. J. Food Prot. 58:1369-1374.
Halliwell, B. and J. M. C. Gutteridge. 1984. Oxygen toxicity, oxygen radicals, transition metals and disease. Biochem. J. 219:1-4.
Halliwell, B. and J. M. C. Gutteridge. 1989. Free radical tissue damage: Protective role of antioxidant nutrients. FASEB J. 1:441-445.
Hampson, D. J. 1994. Postweaning Escherichia coli diarrhea in pigs. Escherichia coli in Domestic Animals and Humans (Ed. C. L. Gyles) pp. 171-791. CAB International. London, England.
Ito, M., K. Ohishi, Y. Yoshida, W. Yokoi and H. Sawada. 2003. Antioxidative effects of lactic acid bacteria on the colonic mucosa of iron-overloaded mice. J. Agric. Food Chem. 51:4456-4460.
Janero, D. R. 1990. Malondialdehyde and thiobarbituric acid reactivity as diagnostic indices of lipid peroxidation and peroxidative tissue injury. Free Radic. Biol. Med. 9:515-540.
Kaizu, H., M. Sasaki, H. Nakajima and Y. Suzuki. 1993. Effect of antioxidative lactic acid bacteria on rats fed a diet deficient in vitamin E. J. Dairy Sci. 76:2493-2499.
Kim, H. S., H. S. Chae, S. G. Jeong, J. S. Ham, S. K. Im, C. N. Ahn and J. M. Lee. 2006. In vitro antioxidative properties of Lactobacillus. Asian-Aust. J. Anim. Sci. 19:262-265.
Koninkx, J., F. J. G. and J. J. Malago. 2008. The protective potency of probiotic bacteria and their microbial products against enteric infections. Folia Microbiol. 53:189-194.
Kullisaar, T., E. Songisepp, M. Mikelsaar, K. Zilmer, T. Vihalem and M. Zilmer. 2003. Antioxidative properties of fermented goats’ milk decreases oxidative stressmediated atherogenicity in human subjects. Br. J. Nutr. 90:449-456.
Kullisaar, T., M. Zilmer, M. Mikelsaar, T. Vihalem, H. Annuk, C. Kairane and A. Kilk. 2002. Two antioxidative lactobacilli strains as promising probiotics. Int. J. Food Microbiol. 72:215-224.
Lauridsen, C., S. Hojsgaard and M. T. Sorensen. 1999. Influence of dietary rapeseed oil, vitamin E, and copper on the performance and the antioxidative and oxidative status of pigs. J. Anim. Sci. 77:906-916.
Lin, M. Y. and F. J. Chang. 2000. Antioxidative effect of intestinal bacteria Bifidobacterium longum ATCC 15708 and Lactobacillus acidophilus ATCC 4356. Dig. Dis. Sci. 45:1617-1622.
Nielsen, F., B. B. Mikkelsen, J. B. Nielsen, H. R. Anderson and P. Grandjean. 1997. Plasma malondialdehyde as biomarker for oxidative stress: reference interval and effects of lifestyle factors. Clin. Chem. 43:1209-1214.
NRC. 1998. Nutrient Requirements of Swine, 10th Edition. National Academic Press. Washington, DC, USA.
Ouwehand, A. C., S. Salminen and E. Isolauri. 2002. Probiotics: an overview of beneficial effects. Antonie Van Leeuwenhoek 82:279-289.
Ouwehand, A. C. and S. Vesterlund. 1998. Antimicrobial components from lactic acid bacteria. Lactic Acid Bacteria (Ed. S. Salminen, A. Wright and A. Ouwehand) pp. 375-388.
Marcel Dekker Inc. New York, USA.
SAS. 1997. SAS/STAT user’s guide, Version 9. SAS Institute Inc. Cary, NC, USA.
Sauerwein, H., S. Schmitz and S. Hiss. 2007. Effects of a dietary application of a yeast cell wall extract on innate and acquired immunity, on oxidative status and growth performance in weaning piglets and on the ileal epithelium in fattened pigs. J. Anim. Physiol. Anim. Nutr. 91:369-380.
Shimada, K. K. Fujikawa, K. Yahara and T. Nakamura. 1992. Antioxidative properties of xanthan on the autoxidation of soybean oil in cyclodextrin emulsion. J. Agric. Food Chem. 40:945-948.
Wang, A. N., X. W. Yi, H. F. Yu, B. Dong and S. Y. Qiao. 2009. Free radical scavenging activity of Lactobacillus fermentum in vitro and its antioxidative effect on growing-finishing pigs. J. Appl. Microbiol. 107:1140-1148.
Zhan, X. A., M. Wang, Z. R. Xu, W. F. Li and J. X. Li. 2006. Effects of fluoride on hepatic antioxidant system and transcription of Cu/Zn SOD gene in young pigs. J. Trace Elem. Med. Biol. 20:83-87.
Zhao, Y. P., W. L. Yu and D. P. Wang. 2003. Chemiluminescence determination of free radical scavenging abilities of ‘tea pigments’ and comparison with ‘tea polyphenols’. Food Chem. 80:115-118.