Research Article

A highly significant association between Cathepsin S gene polymorphisms rs12068264 and chronic obstructive pulmonary disease susceptibility in Han Chinese population

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Introduction

Chronic obstructive pulmonary disease (COPD) is a major cause of deaths worldwide and is characterized by an irreversible progressive decline in lung function and chronic inflammatory response [1]. The prevalence is approximately 10% in the elderly and this disease is expected to become the third leading cause of deaths by 2030 in the world [2]. COPD is regarded as a systemic disease. Numerous studies demonstrated that oxidative stress, protease–antiprotease imbalance, as well as inflammation played important roles in COPD pathogenesis [3–5]. It is well reported that long-term exposure to cigarette smoke, a predominant risk stimuli for COPD development, leads to cellular oxidative stress and inflammatory response in the airways as well as lungs [3]. Interestingly, there are only 15–20% of smokers reaching COPD, suggesting that genetic factors are implicated in the pathogenesis of the disorder except for environmental factors [6]. The molecular mechanisms underlying COPD pathogenesis are complex and it is very important to identify the genetic susceptibility factor of the development of COPD for early prevention, early detection, and the early treatment of this disease.
Table 1 Basic SNP information of CTSS and SIRT1 genes examined in the study

| Gene  | SNP ID   | Chr. | Chr. position | SNP property | Alleles | Allele 1 | Allele 2 |
|-------|----------|------|---------------|--------------|---------|----------|---------|
| CTSS  | rs12068264 | 1    | 150727329     | intron3      | C/T     | C        | T       |
| CTSS  | rs11576175 | 1    | 150727394     | intron3      | G/A     | G        | A       |
| SIRT1 | rs2273773  | 10   | 696666598     | synon        | C/T     | C        | T       |
| SIRT1 | rs16924934 | 10   | 69664933      | intron3      | G/A     | G        | A       |
| SIRT1 | rs3818291  | 10   | 69672999      | intron7      | G/A     | G        | A       |

Abbreviation: Chr., chromosome.

Increasing evidence suggested that a variety of factors including imbalance of proteases exert a vital role in the development and progression of COPD [4,7,8]. Proteases are able to cleave proteins into smaller fragments, which have regulatory roles in many pathological processes such as apoptosis, oxidative stress, and inflammatory response [7]. Therefore, dysregulated proteases are involved in the pathogenesis of various diseases including COPD. Proteinases like neutrophil elastase, matrix metalloproteinases, and cysteine proteases are closely associated with development of emphysema and destruction of airways in COPD [9].

Cathepsin S (CTSS) is a cysteine proteinase of the papain superfamily, which is mainly expressed in professional antigen-presenting cells related to COPD-like macrophages, microglia, B lymphocytes, as well as dendritic cells [10,11]. This enzyme plays a crucial action in assembling MHC class II–Ag complexes and promotes antigen processing and presentation and subsequently induces the activation of CD4+ T cells [12]. CTSS levels were elevated in bronchoalveolar lavage fluid and plasma of COPD patients compared with the healthy volunteers [11,14]. Moreover, CTSS is implicated in pathogenesis of alveolar remodeling and pulmonary emphysema in interferon-γ-treated mouse [15,16]. Collectively, CTSS plays an influential role in the pathogenesis of COPD.

Sir2uin-1 (SIRT1) is the best characterized member of silent information regulator (Sir2) family in mammals and is an NAD-dependent protein deacetylase [16]. This enzyme controls numerous cellular responses involved in regulating homeostasis [17] and it is demonstrated to have inhibitory roles in ageing and inflammation [18-20]. SIRT1 is reported to decrease in lungs of COPD patients, the well-known ageing-related and inflammation-associated diseases [1,21], initially reported by Rajendraosozhan et al. [20]. In addition, through deacetylating multiple substrate proteins like Forkhead box O proteins (FOXOs), p53, and nuclear factor-kB, SIRT1 exerts an important role in the cellular responses to oxidative stress, cellular senescence, and modulation of inflammation in COPD [22]. These observations indicated that SIRT1 has important functions in the progression of COPD.

COPD is a complicated disease and is at least partly determined by gene(s). Increasing evidence suggested that single nucleotide polymorphisms (SNP) distribution of genes implicated in COPD such as IL-13, FAM13A, and VEGFA genes play key roles in the COPD development [23-25]. However, the SNP-caused effects of many genes involved in COPD susceptibility and progression remain largely unknown. Therefore, the purpose of the present study was to investigate the associations between CTSS and SIRT1 SNPs and the susceptibility of COPD in a Chinese population.

Materials and methods

Study subjects

Cases (182) and controls (203) were selected as our research objects in the hospital-based case–control study. The control samples were obtained from medical examination center of Affiliated Hospital of Guangdong Medical University and cases were collected from Affiliated Hospital of Guangdong Medical University as well as The Second People’s Hospital of Zhanjiang from June 2015 to June 2017. All the cases were diagnosed newly with forced expiratory volume in 1 s (FEV1) for the reason that FEV1 over forced vital capacity (FEV1/FVC) < 0.7 has become the gold criteria for diagnosing COPD [26,27]. The controls who were free from a history of cancer and respiratory diseases were randomly selected in the same hospital during the same period. The present study was approved by the institutional review board of Guangdong Medical University.

Candidate SNP selection and genotyping

The characteristics of two SNPs (rs12068264 and rs11576175) of CTSS gene and three SNPs (rs2273773, rs16924934, and rs3818291) of SIRT1 gene selected for the test are shown in Table 1. Whole blood samples (5 ml) were collected from each control and COPD patients during a laboratory examination. Genomic DNA was extracted from whole...
blood using a commercially available DNA isolation kit (Tiangen Biochemical Technology, Beijing, China) following the manufacturer’s protocol. SNPs were used based on their frequencies and positions, as well as their relationship with infectious diseases documented previously. SNP genotyping was performed using an improved multiplex ligation detection reaction (iMLDR) technique (Genesky Biotechnologies Inc., Shanghai, China).

Statistical analysis
All statistical analyses were two-tailed and conducted with SPSS 18.0. A value of *P* < 0.05 was defined as the criterion of statistical significance. Hardy–Weinberg equilibrium (HWE) was tested by a goodness-of-fit *χ²* test. Student’s *t* test was performed in continuous variables and the Pearson’s *χ²* was performed in categorical variables between cases and controls. The odds ratios (OR) and 95% confidence intervals (95% CI) were calculated to estimate the association between SNP and the risk of COPD by logistic regression analysis after adjustment of age and gender. In addition, the haplotype analysis of CTSS and SIRT1 polymorphisms was determined using the Haploview 4.2 program. Only those haplotypes with frequencies greater than 3% were further analyzed.

Results

**CTSS and SIRT1 SNPs genotype and allele frequencies**
Genotype distributions and allele frequencies of CTSS and SIRT1 SNPs in the COPD group and the control group are listed in Table 2. Genotype distributions of all polymorphisms were found to conform to HWE (*P* > 0.05), indicating that our subjects presented the source population well. A statistical difference was observed between the patients with COPD and the controls in the distribution of CTSS rs12068264 genotypes (*P* = 0.048). The prevalence of rs12068264 T-allele frequencies in CTSS gene was significantly higher in COPD patients than in controls (T compared with C: OR = 1.351, 95% CI: 1.008–1.811, *P* = 0.044). In addition, for the other polymorphisms, no significant differences were observed in the genotype and allele frequencies between the COPD patients and the control groups.

**Association analysis of CTSS and SIRT1 polymorphisms and risk of COPD in dominant and recessive model**
The age and gender analysis showed COPD patients had older age than the controls (71.67 ± 10.43 compared with 55.48 ± 10.62, *P* < 0.001) and the COPD cases and control subjects exhibited significant difference in gender (145 males and 37 females compared with 111 males and 92 females, *P* < 0.001). Subsequently, to investigate the association between the CTSS and SIRT1 SNPs associated with COPD, we compared the genotype frequencies of every polymorphism between groups under the dominant and recessive genetic models using logistic regression analysis.

### Table 2 Frequencies of genotype and allele of CTSS and SIRT1 SNPs in the control and COPD groups

| SNP        | Group | Genotype number | P    | HWE P | Allele freq (%) | P | OR (95% CI) |
|------------|-------|-----------------|------|-------|-----------------|---|-------------|
| rs12068264 | Control | TT              | 21   | 0.048 | 0.531           | 0.3416 | 0.6584 | 0.044 | 1.351 (1.008–1.811) |
|            | COPD   | TC              | 96   | 0.531 | 34.16%          | 65.84% | 0.044  | 1.351 (1.008–1.811) |
| rs12068264 | Control | CC              | 85   | 0.222 | 41.21%          | 58.79% | 0.044  | 1.351 (1.008–1.811) |
| rs12068264 | COPD   | TG              | 67   | 0.222 | 41.21%          | 58.79% | 0.044  | 1.351 (1.008–1.811) |
| rs11576175 | Control | AG              | 10   | 0.506 | 20.2%           | 79.8%  | 0.234  | 0.801 (0.555–1.515) |
| rs11576175 | COPD   | GG              | 131  | 0.506 | 20.2%           | 79.8%  | 0.234  | 0.801 (0.555–1.515) |
| rs16924934 | Control | AA              | 10   | 0.506 | 20.2%           | 79.8%  | 0.234  | 0.801 (0.555–1.515) |
| rs16924934 | COPD   | GG              | 131  | 0.506 | 20.2%           | 79.8%  | 0.234  | 0.801 (0.555–1.515) |
| rs2273773  | Control | CC              | 10   | 0.208 | 23.89%          | 76.11% | 0.084  | 1.326 (0.962–1.828) |
| rs2273773  | COPD   | CT              | 116  | 0.208 | 23.89%          | 76.11% | 0.084  | 1.326 (0.962–1.828) |
| rs3818291  | Control | AA              | 15   | 0.860 | 29.4%           | 70.6%  | 0.044  | 1.351 (1.008–1.811) |
| rs3818291  | COPD   | AG              | 90   | 0.860 | 29.4%           | 70.6%  | 0.044  | 1.351 (1.008–1.811) |
| rs3818291  | Control | 8               | 55   | 0.860 | 29.4%           | 70.6%  | 0.044  | 1.351 (1.008–1.811) |
| rs3818291  | COPD   | 5               | 140  | 0.860 | 29.4%           | 70.6%  | 0.044  | 1.351 (1.008–1.811) |

Abbreviation: Freq, frequency. NA, not available.
Table 3 Logistic regression analysis of CTSS and SIRT1 polymorphisms and risk of COPD in dominant and recessive model

| SNP Group | Dominant OR (95% CI)* | P* | Recessive OR (95% CI)* | P* |
|-----------|------------------------|----|------------------------|----|
| rs12068264 | CTSS TT + TC CC | 1.014 (0.596–1.723) | 0.960 | 2.30 (1.06–4.989) | 0.035 |
| Control    | 117 85                  | 21 181 | 35 147 |
| COPD       | 115 67                  |                |                |
| rs11576175 | SIRT1 AA + AG GG | 0.760 (0.435–1.327) | 0.334 | 0.505 (0.124–2.055) | 0.340 |
| Control    | 72 131                  | 10 193 | 6 175 |
| COPD       | 55 126                  |                |                |
| rs16924934 | CTSS GG + GA AA | 0.953 (0.545–1.669) | 0.867 | 0.556 (0.137–2.267) | 0.413 |
| Control    | 60 143                  | 6 197 | 5 177 |
| COPD       | 57 125                  |                |                |
| rs2273773  | CTSS CC + CT TT | 1.000          | 0.000 | 0.000          | 0.000 |
| Control    | 87 116                  | 10 193 | 15 167 |
| COPD       | 92 90                   |                |                |
| rs3818291  | SIRT1 AA + AG GG | 1.000          | 0.000 | 0.000          | 0.000 |
| Control    | 63 140                  | 8 195 | 3 179 |
| COPD       | 59 123                  |                |                |

*OR and P-value was adjusted by age and gender.

Table 4 Clinical characteristics of the patients with COPD and distribution by genotypes of the rs12068264

| Parameters       | TT               | TC + CC             | P    |
|------------------|------------------|---------------------|------|
| Duration of illness (years) | 8.871 ± 4.16   | 9.73 ± 7.12         | 0.124|
| Years of smoke (years)       | 33.37 ± 11.57   | 33.02 ± 10.97       | 0.590|
| WBC (10^9) count           | 8.11 ± 3.30     | 9.70 ± 5.37         | 0.082|
| N%                           | 69.49 ± 12.17   | 71.85 ± 12.67       | 0.747|
| PCT (ng/ml)                 | 0.55 ± 0.87     | 0.28 ± 0.93         | 0.043|
| CRP (mg/l)                  | 8.31 ± 4.46     | 5.57 ± 4.06         | 0.486|
| FEV1/FVC                   | 59.4 ± 11.74    | 57.12 ± 14.96       | 0.057|
| FEV1%                       | 53.33 ± 21.54   | 53.77 ± 23.43       | 0.508|
| Age at onset (years)        |                 |                     |      |
| <60                          | 3               | 11                  |      |
| ≥60                          | 30              | 88                  | 1.000|
| Gender                      |                 |                     |      |
| Male                         | 29              | 78                  | 0.248|
| Female                      | 4               | 21                  |      |
| Pulmonary heart disease     |                 |                     |      |
| No                           | 28              | 86                  | 0.946|
| Yes                          | 3               | 12                  |      |
| Combined severe pneumonia   |                 |                     |      |
| No                           | 15              | 53                  | 0.580|
| Yes                          | 16              | 45                  |      |

For every SNP, if the frequency of one allele was relatively lower than that of another allele, it was defined as minor allele, which was recognized as a risk allele compared with a wild-type allele. As shown in Table 3, the minor allele T at rs12068264 of CTSS gene was associated with an increased COPD risk under a recessive model (TT compared with TC + CC: OR = 2.30, 95% CI: 1.06–4.989, P = 0.035). However, no significant associations were observed between the other SNPs and the risk of COPD in genotype comparisons under different genetic models.

**Correlation analysis between rs12068264 of CTSS gene and clinical characteristics of COPD patients**

For further analysis, we also focused on the description of general clinical characteristics of 132 patients with COPD by genotype and we found that the FEV1/FVC ratio and FEV1/predicted of these COPD subjects were 57.69 ± 14.22, 53.66 ± 22.89, respectively. As shown in Table 4, the homozygous carriers of the TT genotype for the rs12068264...
polymorphism was associated with higher levels of procalcitonin (PCT) compared with the presence of the C variant ($P=0.043$). Nevertheless, there were no significant associations of the genotype of rs12068264 with duration of illness, years of smoke, age at onset, gender, and the prevalence of pulmonary heart disease and combined severe pneumonia. Moreover, the association between rs12068264 and the levels of white blood cell (WBC) count, neutrophil granulocyte ratio (N%), C-reactive protein (CRP), FEV1/FVC, FEV1/predicted did not show significance.

### Haplotype analysis of CTSS and SIRT1 polymorphisms

We also performed a haplotype analysis according to two SNPs of CTSS gene (rs12068264 and rs11576175) and three SNPs of SIRT1 gene (rs16924934, rs2273773, and rs3818291). As shown in Table 5, there are three major haplotypes of CTSS and four major haplotypes of SIRT1 polymorphisms with a frequency above 3% in our samples. Analysis of the haplotype frequencies in both CTSS and SIRT1 polymorphisms showed no significant differences between patients with COPD and the controls.

### Discussion

In the current study, we analyzed the associations between SNPs in the CTSS as well as SIRT1 gene and COPD risk in Han Chinese population. We found that rs12068264 of CTSS was associated with the risk of COPD in our population. The T allele frequency of rs12068264 was significantly higher in COPD patients than in controls. Furthermore, the rs12068264 was associated with an increased risk of COPD based on recessive models. We also found that the homozygous carriers of the TT genotype showed higher levels of PCT compared with the presence of the C variant in the rs12068264 polymorphism. However, haplotype analysis demonstrated that the haplotype frequencies of CTSS and SIRT1 genes had no significant differences between the patients with COPD and the control groups.

COPD is a major global health concern [28]. The development and progression of COPD is involved in an inflammatory cell profile including macrophages, neutrophils as well as T lymphocytes. It is of great importance for the early diagnosis of COPD due to the remediable characteristics at initial stage of this illness. It is widely accepted that oxidative stress, proteinases, and pro-inflammatory mediators are involved in the pathogenesis of COPD [3-5]. CTSS and SIRT1 are reported to play crucial roles in COPD development and therefore can serve as potential biomarkers for COPD [14,22,29]. In this study, we evaluated the potential correlations of the two polymorphisms (rs12068264 and rs11576175) of CTSS gene and three polymorphisms (rs2273773, rs16924934, and rs3818291) of SIRT1 gene with COPD in Han Chinese population. A previous study investigated the common functional polymorphisms (rs7534124, rs35989725, rs16827671, and rs34495036) in the 5’-flanking region in the CTSS promoter and found their possible association with pulmonary emphysema in the Japanese population [30]. Our study found that the T allele frequencies at rs12068264 in CTSS gene exhibited significantly higher in COPD patients than in controls, indicating that the T allele of rs12068264 may increase the susceptibility of COPD. We also found that there were significant differences in the age and gender in the current case–control hierarchy, therefore we used the prominent phenotypes for the correction of logic regression. Our binary logistic regression analysis verified that the minor allele T at rs12068264 was related to an increased COPD risk in a recessive model. The statistically significant associated with the SNPs rs12068264 is in the third intron of the CTSS gene, which is usually removed during the gene-spooling process. Although they cause no apparent functional change, intronic SNPs may modulate gene function by affecting the regulation of gene expression [31]. Nevertheless, no significant associations were found between the other SNP rs11576175 of CTSS gene as well as three SNPs of SIRT1 gene and COPD risk. A previous study found that AG genotype of rs11576175 had lower risks
of metabolic disorders in a Han Chinese population [32]. However, no significant association was observed between SNP rs11576175 of CTSS and risk of COPD in the present study. This inconsistency may contribute to the different roles of SNP rs11576175 in the pathogenesis of the two different disorders. Furthermore, the genotypes frequencies at rs2273773 in SIRT1 had no significant difference between the COPD patients and controls, which is not in line with the previous observation reported by Kalcmci et al. [34] that there is a significant difference between TT, TC, and CC genotypes of rs2273773 in the same different group in Muğla province. These conflicting findings may result from some factors, such as clinical heterogeneity, different sample sizes and ethnic differences.

Subsequently, we further analyzed the association between general clinical characteristics of COPD patients and rs12068264 polymorphism. The recorded clinical characteristics include duration of illness, years of smoke, age at onset, gender, and the prevalence of pulmonary heart disease and combined severe pneumonia, WBC count, N%, PCT, CRP, FEV1/predicted, FEV1/FVC. Serum PCT is reported to serve as an inflammatory biomarker and become the only increased biomarker in the systemic bacterial infections. Moreover, it is well accepted that the values of serum PCT have been applied to diagnose local bacterial infections with increased sensitivity of detection methods and reduced diagnostic threshold (0.5 μg/l was used as a diagnostic threshold for bacterial infection) [33]. We observed that the TT genotype had significantly higher PCT levels than the C variant. However, there were other infection indexes, such as WBC, CRP, neutrophil proportion etc. The levels of these indexes were similar between TT homozygotes and C allele carriers. This inconsistency might be related to PCT that exhibits higher specificity and sensitivity than other indexes in the diagnosis of bacterial infection. But we cannot rule out the false positive results due to the small sample size in the present study. FEV1/FVC ratio and FEV1/predicted have been commonly used to confirm the diagnosis of COPD. All patients had FEV1 values <80% of predicted, and thus were diagnosed with moderate to severe COPD based on the Global Initiative for Chronic Obstructive Lung Disease [1]. The classification of COPD severity is as following: mild = FEV1 ≥ 80% of predicted; moderate = FEV1 ≥50 to <80% of predicted; severe = FEV1 ≥30 to <50% of predicted; and very severe = FEV1 ≤ 30% of predicted. The value of FEV1/predicted in COPD patients is more than 30% but less than 50% in the present study, which suggest that the COPD patients are in moderate stage. Nevertheless, rs12068264 genotype demonstrated no significant associations with duration of illness, years of smoke, age at onset, gender, and the prevalence of pulmonary heart disease and combined severe pneumonia, WBC count, N%, CRP, FEV1/FVC, and FEV1/predicted. We also carried out the haplotype analysis and the data demonstrated that the haplotype of CTSS and SIRT1 gene had no significant associations with the risk of COPD progression.

Our current study has several limitations. First, this was a hospital-based, case–control study, therefore, a selection bias was unavoidable and the subjects are not fully representative of the general population. Second, the polymorphisms rs12068264 studied, which were in non-functional area, may not offer a comprehensive view of the genetic variability of CTSS. Third, the present study did not investigate whether the SNP regulate CTSS gene transcription and protein translation. Fourth, the association between CTSS gene rs12068264 polymorphism and clinical symptoms need to be evaluated in future studies. Fifth, the sample size is relatively small. Thus, false positive results could not be excluded. Finally, the mechanism underlying the association between the polymorphisms at rs12068264 of CTSS and the susceptibility of COPD is still unclear. Therefore, further studies with a larger study subjects and functional experiments are required to be performed.

In conclusion, the current study finds that CTSS gene rs12068264 polymorphism may be associated with the risk of COPD in a Han Chinese population, this finding is expected to lead to better COPD prevention and therapy strategies.

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Competing interests
The authors declare that there are no competing interests associated with the manuscript.

Author contribution
All authors contributed significantly to this work. G.L., Z.-y.L., and Y.-h.W. conducted the research design. S.-l.G., C.-y.L., Y.-h.W., L.-w.Y., B.-a.Z., and Z.-g.C. collected all the samples and the data. W.-m.Y., Z.-q.S., J.-f.C. provided the samples. S.-l.G. prepared all the tables and wrote the paper. G.L. revised the paper. All authors reviewed the manuscript. In addition, all authors have read and approved the manuscript.
Abbreviations

COPD, chronic obstructive pulmonary disease; CRP, C-reactive protein; CTSS, cathepsin S; FEV1, forced expiratory volume in 1 s; FVC, forced vital capacity; HWE, Hardy–Weinberg equilibrium; N%, neutrophil granulocyte ratio; OR, odds ratio; PCT, procalcitonin; SIRT1, Sirtuin-1; SNP, single nucleotide polymorphism; WBC, white blood cell; 95% CI, 95% confidence interval.

References

1 Vestbo, J., Hurd, S.S., Agusti, A.G., Jones, R.W., Vogelmeier, C., Anzueto, A. et al. (2013) Global strategy for the diagnosis, management, and prevention of chronic obstructive pulmonary disease: GOLD executive summary. Am. J. Respir. Crit. Care Med. 187, 347–365, https://doi.org/10.1164/rccm.201204-0596PP

2 Shergis, J.L., Di, Y.M., Zhang, A.L., Vahos, R., Hellilwelt, R., Ye, J.M. et al. (2014) Therapeutic potential of Panax ginseng and ginsenosides in the treatment of chronic obstructive pulmonary disease. Complement. Ther. Med. 22, 944–953, https://doi.org/10.1016/j.ctim.2014.08.006

3 Barnes, P.J. and Celi, B.R. (2009) Systemic manifestations and comorbidities of COPD. Eur. Respir. J. 33, 1165–1185, https://doi.org/10.1183/09031936.00128008

4 Fischer, B.M., Pavilsko, E. and Vomyov, J.A. (2011) Pathogenic triad in COPD: oxidative stress, protease-antiprotease imbalance, and inflammation. Int. J. Chron. Obstruct. Pulmon. Dis. 6, 413–421, https://doi.org/10.2147/COPD.S10770

5 Conti, V., Corbi, G., Manzo, V., Pelaia, G., Filippelli, A. and Vatrelia, A. (2015) Sirtuin 1 and aging theory for chronic obstructive pulmonary disease. Anal. Cell Pathol. (Amst.) 2015, 897327

6 Bosse, Y. (2009) Genetics of chronic obstructive pulmonary disease: a succinct review, future avenues and prospective clinical applications. Pharmacogenomics 10, 655–667, https://doi.org/10.2217/pgs.09.10

7 Pandey, K.C., De, S. and Mishra, P.K. (2017) Role of proteases in chronic obstructive pulmonary disease. Fron. Pharmacol. 8, 512, https://doi.org/10.3389/fphar.2017.00512

8 Lomas, D.A. (2016) Does protease-antiprotease imbalance explain chronic obstructive pulmonary disease? Ann. Am. Thorac. Soc. 13 (Suppl. 2), S130–S137

9 Owen, C.A. (2008) Roles for proteinases in the pathogenesis of chronic obstructive pulmonary disease... Int. J. Chron. Obstruct. Pulmon. Dis. 3, 253–268, https://doi.org/10.2147/COPD.S2089

10 Petanceska, S., Canoll, P. and Devi, L.A. (1996) Expression of rat cathepsin S in phagocytic cells. J. Biol. Chem. 271, 4403–4409, https://doi.org/10.1074/jbc.271.8.4403

11 Zavantnig-Bergant, V., Seknik, A., Golouh, R., Turk, V. and Kos, J. (2001) Immunohistochemical localisation of cathepsin S, cathepsin L and MHC class II-associated p41 isoform in invariant chain in human lymph node tissue. Biol. Chem. 382, 795–804

12 Andrault, P.M., Samsonov, S.A., Weber, G., Coquet, L., Nazmi, K., Bolscher, J.G. et al. (2015) Antimicrobial peptide LL-37 is both a substrate of cathepsin S and K and a selective inhibitor of cathepsin L. Biochemistry 54, 2785–2798, https://doi.org/10.1021/bi500231

13 Geraghty, F., Greene, C.M., O’Mahony, M., O’Neill, S.J., Taggart, C.C. and McEvaney, N.G. (2007) Secretory leucocyte protease inhibitor inhibits interferon-gamma-induced cathepsin S expression. J. Biol. Chem. 282, 33389–33395, https://doi.org/10.1074/jbc.M706884200

14 Nakajima, T., Nakamura, H., Owen, C.A., Yoshida, S., Tsuduki, K., Chubachi, S. et al. (2016) Plasma cathepsin S and cathepsin S/Cystatin C ratios are potential biomarkers for COPD. Dis. Markers 2016, 4093870, https://doi.org/10.1155/2016/4093870

15 Wang, Z., Zheng, T., Zhu, Z., Horner, R.J., Riese, R.J., Chapman, J.R. et al. (2006) Interferon gamma induction of pulmonary emphysema in the adult murine lung. J. Exp. Med. 192, 1587–1600, https://doi.org/10.1084/jem.192.11.1587

16 Zheng, T., Kang, M.J., Crothers, K., Zhu, Z., Liu, W., Lee, C.G. et al. (2005) Role of cathepsin S-dependent epithelial cell apoptosis in IFN-gamma-induced alveolar remodeling and pulmonary emphysema. J. Immunol. 174, 8106–8115, https://doi.org/10.4049/jimmunol.174.12.8106

17 Guarente, L. and Franklin, H. (2011) Epstein Lecture: Sirtuins, aging, and medicine. N. Engl. J. Med. 364, 2235–2244, https://doi.org/10.1056/NEJMr1100831

18 Yeung, F., Hoberg, J.E., Ramsey, C.S., Keller, M.D., Jones, D.R., Frye, R.A. et al. (2004) Modulation of NF-kappaB-dependent transcription and cell survival by the SIRT1 deacetylase. EMBO J. 23, 2369–2380, https://doi.org/10.1038/sj.emboj.7600244

19 Kitada, M., Ogura, Y. and Koya, D. (2016) The protective role of Sirt1 in vascular tissue: its relationship to vascular aging and atherosclerosis. Aging (Albany, N.Y.) 8, 2290–2307, https://doi.org/10.18632/aging.101068

20 Rajendrasozhan, S., Yang, S.R., Kinnula, V.L. and Rahman, I. (2008) SIRT1, an antiinflammatory and antiaging protein, is decreased in lungs of patients with chronic obstructive pulmonary disease. Am. J. Respir. Crit. Care Med. 177, 861–870, https://doi.org/10.1164/rccm.200708-1269OC

21 Mercado, N., Ito, K. and Barnes, P.J. (2015) Accelerated ageing of the lung in COPD: new concepts. Thorax 70, 482–489, https://doi.org/10.1136/thoraxjnl-2014-206084

22 Yao, H. and Rahman, I. (2012) Perspectives on translational and therapeutic aspects of SIRT1 in inflammaging and senescence. Biochem. Pharmacol. 84, 1332–1339, https://doi.org/10.1016/j.bcp.2012.06.031

23 Liao, N., Zhao, H., Chen, M.L. and Xie, Z.F. (2017) Association of the IL-13 polymorphisms rs1800925 and rs20541 with chronic obstructive pulmonary disease risk: an updated meta-analysis. Medicine (Baltimore) 96, e5556, https://doi.org/10.1097/MD.0000000000005855

24 Wang, B., Liang, B., Yang, J., Xiao, J., Ma, C., Xu, S. et al. (2013) Association of FAM13A polymorphisms with COPD and COPD-related phenotypes in Han Chinese. Clin. Genet. 683, 168–1688, https://doi.org/10.1111/cgen.12013

25 Baz-Davila, R., Espinoza-Jimenez, A., Rodriguez-Perez Mdel, C., Zulueta, J., Varo, N., Montejo, A. et al. (2016) Role of HIF1A, VEGFA and VEGFR2 SNPs in the susceptibility and progression of COPD in a Spanish population. PLoS ONE 11, e0154998, https://doi.org/10.1371/journal.pone.0154998

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26 Broekhuizen, B.D., Sachs, A.P., Hoes, A.W., Verheij, T.J. and Moons, K.G. (2012) Diagnostic management of chronic obstructive pulmonary disease. *Netherlands J. Med.* **70**, 6–11

27 Kocaal, H., Saygi, A., Sariman, N., Alici, E., Yurtlu, S., Yilmaz, H. et al. (2013) Evaluation of clinical and functional parameters in female subjects with biomass smoke exposure. *Respir. Care* **58**, 424–430

28 Postma, D.S., Kerkhof, M., Boezen, H.M. and Koppelman, G.H. (2011) Asthma and chronic obstructive pulmonary disease: common genes, common environments? *Am. J. Respir. Crit. Care Med.* **183**, 1588–1594, [https://doi.org/10.1164/rccm.201011-1796PP](https://doi.org/10.1164/rccm.201011-1796PP)

29 Yanagisawa, S., Papaioannou, A.I., Papaporyriou, A., Baker, J.R., Vuppusetty, C., Loukides, S. et al. (2017) Decreased serum Sirtuin-1 in COPD. *Chest* **152**, 343–352, [https://doi.org/10.1016/j.chest.2017.05.004](https://doi.org/10.1016/j.chest.2017.05.004)

30 Minematsu, N., Nakamura, H., Furuuchi, M., Nakajima, T., Takahashi, S., Tsujimura, S. et al. (2008) Common functional polymorphisms in the cathepsin S promoter in Japanese subjects: possible contribution to pulmonary emphysema. *Respirology* **13**, 498–504, [https://doi.org/10.1111/j.1440-1843.2008.01280.x](https://doi.org/10.1111/j.1440-1843.2008.01280.x)

31 Korb, M., Ke, Y. and Johnson, L.F. (1993) Stimulation of gene expression by introns: conversion of an inhibitory intron to a stimulatory intron by alteration of the splice donor sequence. *Nucleic Acids Res.* **21**, 5901–5908, [https://doi.org/10.1093/nar/21.25.5901](https://doi.org/10.1093/nar/21.25.5901)

32 Ou, Z., Wang, G., Li, Q., Ma, Z., Lin, D., Dai, M. et al. (2013) The genetic polymorphisms of cathepsin S were associated with metabolic disorders in a Chinese Han population. *Gene* **526**, 385–389, [https://doi.org/10.1016/j.gene.2013.05.028](https://doi.org/10.1016/j.gene.2013.05.028)

33 Gao, D., Chen, X., Wu, H., Wei, H. and Wu, J. (2017) The levels of serum pro-calcitonin and high-sensitivity C-reactive protein in the early diagnosis of chronic obstructive pulmonary disease during acute exacerbation. *Exp. Ther. Med.* **14**, 193–198, [https://doi.org/10.3892/etm.2017.4496](https://doi.org/10.3892/etm.2017.4496)

34 Kalemci, S., Edgunlu, T.G., Kara, M., Turkcu, U.O., Cetin, E.S. and Zeybek, A. (2014) Sirtuin gene polymorphisms are associated with chronic obstructive pulmonary disease in patients in Mugla province. *Kardiochir Torakochirurgia Pol* **11**, 306–310, [https://doi.org/10.5114/ktp.2014.45682](https://doi.org/10.5114/ktp.2014.45682)