Evaluation of GENEDIA® malaria pf/pan antigen test relative to blood smear microscopy in a malaria diagnosis in Ethiopia.

Abnet Abebe*, Abeba Gebretsadik, Desalegn Nega, Adugna Woyesa
Ethiopian Public Health Institute, Addis Ababa, Ethiopia

Abstract

Background: Early and accurate diagnosis of malaria followed by prompt treatment is crucial to reduce morbidity and mortality in endemic regions. Presumptive treatment of malaria is widely practiced where microscopy or rapid diagnostic tests are not readily available. Introduction of rapid diagnostic tests (RDTs) for the treatment of malaria needs prior evaluation of their performance relative to reference methods. Hence, this study evaluated the diagnostic capacity of GENEDIA® Malaria Pf/pan Ag Rapid Test relative to blood smear for P. falciparum and P. vivax malaria in Ethiopia.

Methods: This was a cross-sectional study conducted from November to December 2014 on malaria blood samples collected from symptomatic patients who visited malaria control center in Adama, Oromia Region, Ethiopia. Thick and thin malaria blood films stained by 10% Giemsa working solution were examined microscopically under 100x magnifications for Plasmodium detection, species identification and determination of parasitaemia. The RDT was performed as per the manufacturer's instruction.

Results: A total of 417 febrile malaria suspected outpatients were diagnosed in the study center. Of which, 149(35.7%) by light microscopy (Pf =47; Pv =93; mixed Pf/Pv =9), and 151 (36.2%) by Genedia® RDT (Pf=20; Pv=87; Pf/pan=44) were positive for malaria. RDT detected 0.6% (n=27) mixed Pf/pv cases on samples that were P. falciparum confirmed by microscopy. The general sensitivity, specificity, positive and negative predictive value of GENEDIA® test relative to microscopy were respectively 95.3%, 96.6%, 94.0% and 97.4%.

Conclusion: The GENEDIA® Malaria Pf/pan Ag test showed good sensitivity and specificity to diagnose malaria in Ethiopia, with respect to the blood film microscopy.

Keywords: GENEDIA® malaria Ag test, Blood film microscopy, Diagnostic performance, Ethiopia.

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Introduction

WHO recommends parasitological confirmation of malaria through quality assured diagnosis in all settings before the start of treatment [1,2]. Treatment solely based on clinical suspicion should only be considered when parasitological diagnosis is not accessible [2]. Early diagnosis of malaria allows early implementation of effective therapy, thereby reducing morbidity and mortality. Microscopy of Giemsa-stained thick and thin blood films by skilled microscopists remain the standard laboratory method for the diagnosis of malaria; however, it needs expertise and electricity [3]. So commercially available RDTs may yield more accurate and fast results in remote areas with inexperienced professionals [4,5].

Malaria RDTs detect Plasmodium parasites in blood by an antibody-antigen reaction on a nitrocellulose strip [4,5]. This strip can be available as a self-standing strip or be embedded in a plastic cassette or occasionally in a cardboard format. Reactions on the nitrocellulose strip are visible as Cherry-red lines [6]. There are different RDTs that can be used to diagnose P. falciparum and other non-falciparum Plasmodium species. Malaria RDTs are qualitative immuno-chromatographic tests for the detection of antigens against all isotypes (IgG, IgM, IgA) in human serum, plasma or whole blood [6]. In different studies, RDTs incorporating the histidine-rich protein II (HRP2) and parasite lactate dehydrogenase (pLDH) antigens [7] have been compared and evaluated for their sensitivities and specificities in various epidemiological settings [8].

Different RDTs that have been evaluated in Ethiopia and in other parts of the world [9-12] showed good sensitivity and specificity of the devices in diagnosis of malaria. The GENEDIA® Malaria Pf/pan Ag test is a rapid and simple serological assay for qualitative determination of Pf specific HRP-2 and pan specific pLDH antigens in human whole blood which has not been evaluated for its performance in Ethiopia. Therefore, this study assessed the performance of GENEDIA® Malaria Pf/pan Ag test for the detection of P. falciparum and P. vivax infections in the Adama malaria control center, which is located in malarious areas of Oromia Regional State of Ethiopia.

Materials and Methods

Study area

This study was undertaken at Adama malaria control center, Oromia Regional State, Central Ethiopia. Adama is located at 8°33’N 39°16’E/ 8.55°N 39.27°E / 8.55; 39.27 at an elevation of
1712 meters, 99 km southeast of Addis Ababa. Oromia region has reported more than 17 million malaria risk population, 20-35% of outpatient consultations, 16% of hospital admissions, and 18-30% of all hospital deaths [13].

**Study participant**

Febrile patients who visited Adama malaria control center gave informed consent to participate in the study and enrolled by using convenient sampling method. Febrile patients from the non-malarious areas, who visited St. Paulo’s referral Hospital, Addis Ababa, participated in the study preferably to better check the specificity of the test.

**Study design**

A cross sectional study was undertaken in November and December 2014 at Adama malaria control center, Oromia Regional State, Ethiopia.

**Sample size determination**

The required sample size for this study was calculated using a formula for a single population proportion. By using 95% confidence level, 3% of margin of error, and 96% expected sensitivity level: 

\[ n_1 = \frac{Z^2 P (1-P)}{d^2} \text{ and } n_2 = n_1 / \text{expected sensitivity level} \]

\[ n_1 = 164; \text{ and } n_2 = n_1 / 0.96 = 164 / 0.96 = 170. \text{ Then } N = 170 + 164 = 334 \]

Where, \( P = \) prevalence estimated; \( d = \) margin of error; \( N = \) sample size of all screening febrile patients; \( n_1 = \) sample size of microscopically confirmed positive cases; \( n_2 = \) expected sample size of microscopically confirmed negative cases.

**Enrollment**

Inclusion criteria: All febrile patients who visited malaria control center

Exclusion criteria: pregnant women and severely ill patients.

**Sample collection**

Clinically malaria-suspected patients were diagnosed using blood smear microscopy and malaria RDTs at the same time. After obtaining the consent of participants, finger prick blood was taken from each febrile patient at the malaria control center and from individuals selected from St. Paulos referral Hospital (Negative controls).

**Microscopic examination:** Thick and thin blood smears were prepared for each participant on a single slide by using finger prick blood sample. Blood smear was air-dried and stained for 15 minutes with 10% Giemsa working solution, after the thin film was fixed with methanol. The blood films were microscopically examined under oil immersion objective. Parasitaemia was determined from thick smear while species identification was identified from thin smear. A minimum of one hundred high power fields was examined on a thick blood film before declaring a negative result. Parasite counting was done against 200 WBCs and multiplying by 8000 standard WBC count based on the guideline set by WHO. Discrepancy between the first and second microscopic readings was settled by a third senior microscopist whose readings were considered final [14,15].

**GENEDIA® malaria Pf/pan Ag test:** GENEDIA® Malaria Pf/pan Ag was conducted following the manufacturer’s instruction. Both the RDT device used and the BF slide, for each study subject, were labeled with similar identification code. 5 μL of fresh blood sample was added onto the sample pad of test device, followed by adding 4 drop of the assay diluents and finally the result was read within 20 minutes. The result was declared positive for the presence of pink bands in both the control and test lines (T1: P. falciparum; T2: P. vivax); and negative when pink color was appeared only on the control line. Mixed infections for \( P. \) falciparum and \( P. \) vivax was recorded when a control band and two test areas (T1 and T2) bands appeared pink concurrently. The test was considered invalid when no pink band appeared on control line, irrespective of the test line bands.

**Data management and analysis**

The data was entered using Microsoft Excel and analyzed using STATA 11. Sensitivity, specificity and predictive values of GENEDIA® Malaria Pf/pan Ag test was calculated using blood smear microscopy as the gold standard [14].

**Ethical considerations**

The study was ethically approved by the Scientific and Ethical Review Office (SERO) of Ethiopian Public Health Institute, Addis Ababa, Ethiopia.

**Results**

**Socio demography**

Diagnostic capacity of GENEDIA® Malaria Pf/pan rapid Ag test: A total of 417 febrile patients were diagnosed for malaria in the Malaria Control Center in Adama, Ethiopia. 149 (35.7%) were microscopically confirmed malaria positives; of which, 47 (31.5%) were positive for \( Pf \), 93 (62.4%) for \( Pv \) and 9 (6.4%) for mixed \( Pf/ \) \( Pv \) cases (Table 1).

GENEDIA® Malaria Pf /pan Ag RDT detected 151 (36.2%) malaria positive cases; of which 20 (13.2%) were Pf/infected, 87 (57.6%) were Pv infected and 44 (26.5%) were Pf/pan infected (Table 1). The RDT detected 0.6% (n=27) mixed Pf and Pv positive cases on samples that were Pf confirmed by microscopy (Table 2 and Figure 1).

The GENEDIA® RDT had the general sensitivity of 95.3%, specificity of 96.6%, positive predictive value of 94.0% and negative predictive value of 97.4%, relative to microscopy in malaria diagnosis.

**Discussion**

GENEDIA® Malaria Pf/pan Ag Test, a new alternative rapid diagnostic test, showed a sensitivity of 95.3% and specificity of 96.6% referred to microscopy in the current study. The performance of GENEDIA® RDT was quite higher compared to the previous RDTs in Belgium and Afghanistan [16,17] and almost similar to the finding from a study done in Malawi [8].

The high sensitivity of the GENEDIA® might have also been associated with the high parasite density. In the present study, parasite density was greater than 100 parasites/μL of blood as determined by the microscopic examination of the thick film.
Regarding The relationship between parasite density and RDT performance, previous studies have shown the sensitivity of the RDT is low when the parasite density is less than 100 parasite/µL of blood, more common in asymptomatic patients [18]. The discrepancy in sensitivity of rapid tests depending on:

The level of parasitemia, suggest the need to improve the quality of the test inpatients suffering from malaria with low parasitemia.

The specificity of GENEDIA® Malaria Pf/pan Ag test observed presently for diagnosing Plasmodium falciparum and Plasmodium vivax was also relatively higher than the specificity of similar RDT reported somewhere else [7,16]. The study done in Madagascar shows similar finding [19]. Highly sensitive rapid test can have low specificity and this situation is more commonly seen by the post-treatment persistence of HRP2 for about 1.5 month in Plasmodium falciparum patients [5]. Alternatively, the type of test used may be adversely affected by humidity and may show low specificity [20].

Table 1. Malaria confirmed cases by microscopy and RDTs from Adama malaria control Center, Central Ethiopia, November to December 2014.

| Diagnostic method          | Plasmodium species | Number positive (%) |
|----------------------------|--------------------|---------------------|
| Microscopy                 |                    |                     |
|                            | P. falciparum      | 47 (31.5)           |
|                            | P. vivax           | 93 (62.4)           |
|                            | Mixed              | 9 (6.0)             |
|                            | Total              | 149 (100.0)         |
| GENEDIA® Malaria Pf/pan Ag Rapid Test | |                     |
|                            | P. falciparum      | 20 (13.2)           |
|                            | P. vivax           | 87 (57.6)           |
|                            | *Mixed             | 44 (29.2)           |
|                            | Total              | 151 (100.0)         |

Note: *Mixed infection of P. falciparum and P. vivax.

Table 2. Performance of GENEDIA® Malaria Pf/pan Ag Test relative to microscopy in Adama malaria control center, Central Ethiopia, November to December 2014.

| GENEDIA® | Sensitivity | Specificity | PPV | NPV | Kappa |
|----------|-------------|-------------|-----|-----|-------|
| All Plasmodium | 95.3 (93.8-96.8) | 96.6 (95.1-98.1) | 94.0 (92.5-95.5) | 97.4 (95.9-98.9) | 0.81 |
| P. falciparum | 95.7 (94.2-97.2) | 98.1 (96.6-99.4) | 90.0 (88.6-91.5) | 99.4 (97.9-99.7) | 0.84 |
| P. vivax | 95.7 (94.2-96.9) | 98.1 (96.6-99.4) | 94.7 (93.2-95.9) | 98.5 (97.1-99.8) | 0.82 |

Abbreviations: PPV: Positive Predictive Value; NPV: Negative Predictive Value.

The quality of different types of malaria RDTs have been evaluated in different location of Ethiopia. The performance of Care Start™ malaria Pf/Pv Combo test which was evaluated in Wondogenet, South Ethiopia showed high sensitivity and specificity for Plasmodium falciparum (98%) and Plasmodium vivax (98.2%) [10] and this is almost similar to the current finding. We compare this investigation with kits evaluated in different countries because we couldn’t get similar studies in Ethiopia. Some tests showed less sensitivity and specificity to this research [21]. This study reveals that GENEDIA® Malaria Pf/pan Ag Rapid Test can be used as a rapid diagnostic kit for malaria in the Ethiopian setting like Care Start™ malaria Pf/Pv Combo test and others that were previously evaluated in different parts of the country.

Conclusion

GENEDIA® Malaria Pf/pan Ag Rapid Test showed very high sensitivity and specificity in the detection of Plasmodium vivax and Plasmodium falciparum malaria, which fulfills the WHO recommendation. We suggest further studies to evaluate the performance in different parts of malaria endemic areas.

Authors’ Contributions

AG& AA conceived& designed the study.
AG& AA acquired data.
AG, AA & DN participated in data analysis and interpretation.
AG &DN drafted the manuscript.
AA, AG & DN critically reviewed the manuscript.
All authors read and approved the final manuscript.

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*Correspondence to:
Abnet Abebe
Ethiopian Public Health Institute
Addis Ababa, P.O. BOX: 1242
Ethiopia
E-mail: abnetabas@gmail.com