Does exercise influence the susceptibility to arterial thrombosis? An integrative perspective

Nørregaard Olsen, Line; Fischer, Mads; Evans, Phillip Adrian; Gliemann, Lasse; Hellsten, Ylva

Published in:
Frontiers in Physiology

DOI:
10.3389/fphys.2021.636027

Publication date:
2021

Document version
Publisher's PDF, also known as Version of record

Document license:
CC BY

Citation for published version (APA):
Nørregaard Olsen, L., Fischer, M., Evans, P. A., Gliemann, L., & Hellsten, Y. (2021). Does exercise influence the susceptibility to arterial thrombosis? An integrative perspective. Frontiers in Physiology, 12, [636027]. https://doi.org/10.3389/fphys.2021.636027
Does Exercise Influence the Susceptibility to Arterial Thrombosis? An Integrative Perspective

Line Nørregaard Olsen†, Mads Fischer†, Phillip Adrian Evans‡, Lasse Gliemann† and Ylva Hellsten‡*

† Department of Nutrition, Exercise and Sports, University of Copenhagen, Copenhagen, Denmark, ‡ Haemostasis Biomedical Research Unit, Welsh Centre for Emergency Medicine Research, Morriston Hospital, SBU Health Board, Swansea, United Kingdom, § College of Medicine, Swansea University, Swansea, United Kingdom

Arterial thrombosis is the primary cause of death worldwide, with the most important risk factors being smoking, unhealthy diet, and physical inactivity. However, although there are clear indications in the literature of beneficial effects of physical activity in lowering the risk of cardiovascular events, exercise can be considered a double-edged sword in that physical exertion can induce an immediate pro-thrombotic environment. Epidemiological studies show an increased risk of cardiovascular events after acute exercise, a risk, which appear to be particularly apparent in individuals with lifestyle-related disease. Factors that cause the increased susceptibility to arterial thrombosis with exercise are both chemical and mechanical in nature and include circulating catecholamines and vascular shear stress. Exercise intensity plays a marked role on such parameters, and evidence in the literature accordingly points at a greater susceptibility to thrombus formation at high compared to light and moderate intensity exercise. Of importance is, however, that the susceptibility to arterial thrombosis appears to be lower in exercise-conditioned individuals compared to sedentary individuals. There is currently limited data on the role of acute and chronic exercise on the susceptibility to arterial thrombosis, and many studies include incomplete assessments of thrombogenic clotting profile. Thus, further studies on the role of exercise, involving valid biomarkers, are clearly warranted.

Keywords: physical activity, exercise, thrombogenicity, blood clots, platelet reactivity, clot microstructure, plasma biomarkers

INTRODUCTION

Thrombosis is the formation of a blood clot inside a blood vessel, leading to obstruction of blood flow in the arterial or venous circulatory system (Mackman, 2008). Acute arterial thrombosis is the cause of myocardial infarction and stroke, which collectively are the most common causes of death in developed countries (Mozaffarian et al., 2016). The risk of thrombosis and the consequent cardiovascular events are closely coupled to aging and lifestyle factors such as a diet, smoking, and physical inactivity.
A primary trigger of arterial thrombosis is rupture of an atherosclerotic plaque, which leads to a rapid recruitment of platelets to the injured site, through interaction of platelet surface receptors with collagen and other proteins exposed at the site of injury in the vessel wall. Subsequent activation of the coagulation cascade, leading to generation of thrombin and fibrin, results in clot formation and occlusion of the artery (Mackman, 2008). The ultimate microstructure of the formed blood clot is of significance; less dense clots can be dissolved naturally through the process of fibrinolysis, whereas more dense blood clots cannot easily be dissolved, and consequently, dense blood clots are associated with higher risk of thromboembolic events (Collet et al., 2000; Mills et al., 2002). Thus, the severity of a blood clot is closely associated with the structure of the fibrin network, which is supported by the finding that the rate of fibrinolysis is dependent on the structure formed by the fibrin fibers (Collet et al., 2006). Therefore, the potential for thrombosis formation, the density of the thrombus, and the efficacy of fibrinolysis are of critical importance in evaluating the risk of arterial thrombosis.

Lifestyle changes, such as regular physical activity, have protective effects on cardiovascular disease, such as acute myocardial infarction and stroke (Buckley et al., 2019) and habitual physical activity is accordingly recommended for populations at risk (Arnett et al., 2019; Pelliccia et al., 2020). Nevertheless, there is a risk-benefit paradox with regard to physical activity; acute exercise influences the coagulation system with a consequent transient increase in the risk of arterial thrombosis, with the magnitude of risk being influenced by physical fitness and relative exercise intensity (Thompson et al., 2007, 2020). This brief review covers some of the known effects of acute and habitual exercise on risk markers of arterial thrombosis. We first provide a short overview over experimental methods used to assess the integrated risk of arterial thrombosis, followed by a discussion on the influence of acute exercise and exercise training on biomarkers of arterial thrombosis.

**BIOMARKERS INDICATING THE SUSCEPTIBILITY TO ARTERIAL THROMBOSIS**

**Platelet Reactivity**

Platelets play an important role for many physiological and pathophysiological processes including healing of injured blood vessels and the development of arterial thrombosis (Nachman and Rafii, 2008; de Groot et al., 2012). Platelets circulate with the blood in an inactivated state until they come into contact with activating molecules, such as circulating epinephrine and collagen exposed upon injury to the vessel wall. The activated platelets adhere and initiate a tightly regulated process leading to the formation of a hemostatic plug (de Groot et al., 2012).

Evaluation of platelet aggregation can include platelet number, morphology, and function (Vinholt et al., 2014) and there are several available methods for these assessments as described in detail elsewhere (Vinholt et al., 2014). The present review only focuses on the assessment of platelet reactivity. For this assessment, light transmission aggregometry is often used (Born, 1962; O’Brien, 1962; Vinholt et al., 2017), and reactivity is determined by assessing the propensity of platelets to aggregate in response to different concentrations of platelet receptor agonists (Algahtani and Heptinstall, 2017). Physiologically relevant agonists for the assay are collagen, adenosine diphosphate (ADP), and thrombin receptor-activating peptide (TRAP) (Born, 1962; Davidson et al., 1988; Lundberg Slingsby et al., 2017; Vinholt et al., 2017).

Although light transmission aggregometry is a useful method for the assessment of platelet function, a few aspects are useful to keep in mind when interpreting the data. One limitation of assessing platelet function per se as a marker of blood clot susceptibility is that it only provides indication of one, albeit important, step in hemostasis. Methodological limitations include preparation time of platelet rich plasma to be used in the assay, which may influence aggregation, and that the measurement is made in vitro, without immediate influence of hemodynamic factors. Nevertheless, with careful methodological considerations and in combination with other markers of thrombogenicity, the platelet aggregometry method provides a useful indication of the susceptibility to thrombosis, and the method also has the advantage that a large number of samples, and many different agonists, can be tested simultaneously (Salehi et al., 2014; Paniccia et al., 2015; Vinholt et al., 2017).

**Plasma Markers of Thrombogenicity**

Various markers of hemostasis and fibrinolysis have been identified as independent cardiovascular risk factors (Koenig and Ernst, 2000) and have been widely used in clinical practice. Standard coagulation assays include assessment of activated partial thromboplastin time (APTT), prothrombin time (PT), and thrombin time (TT) (Feng et al., 2014). The assays are functional and evaluate the rate of clot formation when the coagulation cascade has been activated. Other commonly used markers of thrombogenicity are related to fibrin, which plays an essential role in the microstructure of the blood clot. Blood clots can be dissolved by the fibrinolytic system through degradation of fibrin. The key enzyme for degradation of fibrin is plasmin, which is converted from circulating plasminogen by plasminogen activators: tissue-plasminogen activator (t-PA) and urokinase-plasminogen activator (u-PA). The t-PA and u-PA can be inhibited by plasminogen-activator inhibitor-1 (PAI-1) and plasminogen-activator inhibitor-2 (PAI-2).

Although standard coagulation markers remain the mainstay of pathway analysis (PT, APTT), they have several limitations in assessing diseases and treatment thereof. The markers only indicate isolated parts of the plasma-based coagulation pathways. Furthermore, many of these pathway markers do not take into account the effect exerted by the cellular components in blood (such as platelets, and white and red blood cells). Furthermore, many of the tests are carried out in physiologically altered blood (in tubes with, e.g., heparin or sodium citrate). The testing process varies from laboratory to laboratory often with different reference ranges, making interpretation in various disease states difficult (Ebner et al., 2018). In addition, many of
the hypercoagulable disease states have a marked inflammatory component and response due to endothelial change and damage, which can neither be measured nor assessed with these tests. In most circumstances, this makes their overall utility in many diseases mainly an adjunct to clinical care.

**Clot Microstructure**

Due to the need for a global marker to assess hemostatic competency in hypercoagulable states and their treatment, recent studies have focused on the potential of clot microstructure and quality as a more global and accurate measurement. This has led to the development of a potential and exciting new marker, fractal dimension, which has been seen as an improved marker of coagulation that has the ability to assess the direct and indirect effect of coagulation in one simple and immediate test in unaltered whole blood (Kopytek et al., 2019).

Fractal dimension provides information about the structure of the fibrin network of a developing blood clot (Evans et al., 2010) and is thus a functional biomarker of hemostasis and clot microstructure. Fractal dimension provides a description of the incipient clot, which is the initial templating structure of the clot, which leads to its ongoing mechanical structure and strength (Figure 1) (Curtis et al., 2011, 2013). Low values of fractal dimension are equivalent to a weak and less dense clot with low number of branches, whereas high values are equivalent to a strong and dense clot with a high number of complex structured branches, which is more difficult to break down therapeutically (Evans et al., 2010; Sabra et al., 2017). In acute vascular inflammatory disease such as ischemic stroke and myocardial infarction, there is considerable over-production of mass and cross-linking at this templating phase, which leads to a very strong and abnormally structured clot, as compared to clot formation in healthy individuals (Figures 1, 2). In contrast to standard and conventional coagulation measurements like PT or APTT, fractal dimension is performed in untreated blood within minutes after blood withdrawal and provides a rapid measurement of the integrated hemostatic property and thereby an immediate indication of the risk of cardiovascular events (Sabra et al., 2017). Limitations include the requirement of relatively expensive equipment, and as with platelet aggregometry, the measurement is made in vitro without physiological hemodynamic impact. Also, the method is still relatively new, and although the method has been used in a large number of subjects (Evans et al., 2010; Lawrence et al., 2014), normal ranges for gel point and fractal dimension in different populations have yet to be defined.

**ACUTE EXERCISE AND ARTERIAL THROMBOSIS**

Paradoxically, although regular exercise or a physically active lifestyle protect against cardiovascular disease (Albert et al., 2000; Lee et al., 2003; Sattelmair et al., 2011), a single acute bout of physical exertion might trigger an acute myocardial infarct or stroke (Lee et al., 2003). The increased risk is reported to last up to 30–60 min after termination of exercise (Röcker et al., 1986; Mittleman et al., 1993; Albert et al., 2000; Buckley et al., 2019), with one study reporting the effect to persist for up to 2 h (von Klot et al., 2008). However, an important aspect is that, whereas in a sedentary individual the risk of a myocardial infarct increases ~50- to 100-fold with acute vigorous physical activity, in an individual accustomed to exercise the increased risk is much lower (~2- to 5-fold) (Figure 2) (Bärtsch, 1999; Thompson et al., 2007). This would suggest that once an individual is accustomed to exercise, the susceptibility to thrombosis by acute exercise is substantially reduced. This aspect is further discussed below in the section on exercise training.

Exercise intensity is likely to influence the susceptibility to arterial thrombosis and may also affect the time required for returning to resting hemostasis (Supplementary Table 1) (Hegde et al., 2001; Menzel and Hilberg, 2011). As such, low to moderate intensity exercise elicits low or no increase in plasma thrombotic markers and platelet reactivity (Hegde et al., 2001; Davies et al., 2016; Lundberg Slingsby et al., 2018), whereas strenuous exercise leads to a substantial increase (Hegde et al., 2001; Davies et al., 2016). The increased risk of myocardial infarct correlates with high self-reported physical exertion, with a score representing vigorous exertion resulting in a >5-fold increase in the relative risk of onset of a myocardial infarct in the 1–2 h following exercise (Buckley et al., 2019).

One reason for the increased relative risk of arterial thrombosis with intense exercise could be elevated vascular shear stress (Badiei et al., 2015) due to the higher blood flows with intense exercise (Hathcock, 2006; Chen et al., 2010; Badiei et al., 2015). High shear stress levels can promote thrombosis as the frictional force may cause tearing or rupture of an atherosclerotic plaque (Hallqvist et al., 2000; Thompson et al., 2007). Also, many platelet activating factors are influenced by increased exercise intensities, such as catecholamines (Dimsdale and Moss, 1980; Lawrence et al., 2018), temperature (Meyer et al., 2013; Lawrence et al., 2016), blood osmolality (Lawrence et al., 2014), and central hypovolemia (Meyer et al., 2013). In addition, aspects such as endothelial alterations, which influence platelet adhesion and thrombin generation, may also be affected by exercise and lead to an increased susceptibility to thrombosis.

Catecholamines promote thrombosis by activation of platelets and an increase in clot microstructure (Mittleman et al., 1995; Thompson et al., 2007; Davies et al., 2016; Lawrence et al., 2018). In a recent study, tyramine infusion, leading to endogenous noradrenalin formation, was found to increase fractal dimension, indicating a denser incipient clot microstructure (Lawrence et al., 2018). Accordingly, compared to moderate intensity exercise, intense exercise with higher plasma norepinephrine concentrations was shown to be associated with significantly higher levels of thrombotic plasma markers (Menzel and Hilberg, 2011). Nevertheless, single leg knee extensor exercise, which has a limited impact on sympathetic activity, significantly increases fractal dimension suggesting that local hemodynamic changes such as shear stress can influence clot microstructure independently of catecholamines (Lawrence et al., 2018). Combined these studies provide support for that both catecholamines and hemodynamic changes during exercise
can influence the microstructure of developing blood clots, but further studies elucidating their precise role are required.

Data on exercise-induced effects in healthy individuals may differ from data in patient groups for whom the exercise-induced thrombogenicity is likely to be more critical. For example, although patients with either hypertension or coronary artery disease do not seem to differ in basal platelet reactivity or markers of thrombosis compared to healthy controls (Petidis et al., 2008; Hong et al., 2009), these patient groups may present increased catecholamine levels during and after exercise (Petidis et al., 2008) with a potential impact on thrombogenicity. Nevertheless, findings on exercise-induced thrombogenicity in coronary artery disease patients are inconsistent (Mehta and Mehta, 1982; Pamukcu et al., 2005; Aurigemma et al., 2007; Petidis et al., 2008), likely due to variations in criteria for medication and co-morbidities of the populations.

It should be emphasized that despite the apparent immediate changes in thrombogenicity after exercise, the risk of hospitalization during clinical graded exercise testing is approximately one in 10,000 (Myers et al., 2009). In healthy adults, between 30 and 60 years of age, this risk of a cardiac event is estimated to be ~0.2 and 0.3 per 10,000 person-hours of exercise for women and men, respectively (Gibbons et al., 1980). Moreover, in one of the few studies assessing platelet reactivity following acute low intensity one-leg knee extensor exercise, a significant decrease in epinephrine-induced platelet aggregation was observed in moderately and highly trained, but not sedentary, middle-aged male subjects (Lundberg Slingsby et al., 2018). This finding suggests that, in individuals accustomed to exercise, the risk of arterial thrombosis after exercise of lower intensities may even be lower than at rest. Also, plasma fibrinolysis has been shown to increase after acute exercise as a result of increased release of t-PA from endothelial cells in the vasculature (Rankinen et al., 1995; Gunga et al., 2002; Sumann et al., 2007). The increase in t-PA occurs with different exercise modalities but appears to be higher with higher exercise intensity (Handa et al., 1992; El-Sayed, 1993; Hegde et al., 2001; Menzel and Hildberg, 2011).
INFLUENCE OF EXERCISE TRAINING ON SUSCEPTIBILITY TO ARTERIAL THROMBOSIS

It is well established that regular physical activity protects against cardiovascular disease (Buckley et al., 2019), and physical activity for ≥ 30 minutes per day, 5 days a week, is advised (Arnett et al., 2019; Pelliccia et al., 2020). One of the beneficial effects of regular physical activity on arterial thrombi formation is likely through a reduction in basal platelet reactivity (Lundberg Slingsby et al., 2018; Heber et al., 2020). Whereas untrained individuals display an increased level of adrenaline-induced platelet aggregation in response to an acute bout of exercise, platelet aggregation remains unaffected in moderately trained or well-trained individuals (Lundberg Slingsby et al., 2018). Similar to that in healthy individuals, exercise training in patients with coronary artery disease has been found to decrease platelet reactivity, and inclusion of high intensity exercise sessions has been reported to be more beneficial than moderate exercise training alone (Heber et al., 2020). This effect of reduced platelet reactivity with the added high intensity exercise sessions is also present immediately after acute exercise (Supplementary Table 3) (Heber et al., 2020).

Currently, only a limited amount of data exists regarding the influence of exercise training on the thrombogenic clotting profile (Supplementary Tables 2, 3) (Stratton et al., 1991; van den Burg et al., 2000; Lundberg Slingsby et al., 2017, 2018; Heber et al., 2020). In these studies, exercise training consistently resulted in reduced platelet reactivity and a reduced level of plasma markers indicating thrombogenicity (Rauramaa et al., 1986; El-Sayed et al., 1995; Wang et al., 1995, 1997; van den Burg et al., 2000; Lundberg Slingsby et al., 2017; Heber et al., 2020). Interestingly, in late premenopausal women, a period of aerobic interval training by cycling, reduced platelet reactivity, whereas this effect was not observed in recently postmenopausal women (Lundberg Slingsby et al., 2017). The reason for this discrepancy is unclear. It should be noted that both groups of women were found to have improved endothelial function as well as enhanced sensitivity to platelet inhibition by prostacyclin. Improved endothelial function is associated with enhanced formation of prostacyclin...
and nitric oxide (Taddei et al., 2006; Flammer et al., 2012) which both are potent inhibitors of platelet reactivity (Jin et al., 2005; Yau et al., 2015). Several studies, including studies on pre- and postmenopausal women, have shown that exercise training leads to increased expression of enzymes related to prostacyclin synthesis and nitric oxide formation as well as increased plasma levels of prostacyclin and nitric oxide metabolites (Hellsten et al., 2012; Cocks et al., 2013; Nyberg et al., 2017).

Thus, although numerous studies have examined the effect of regular physical activity on cardiovascular risk factors, the specific influence of physical activity on thrombogenicity has not been well investigated, and in particular, the role of exercise intensity and volume is lacking. Future studies should therefore aim to assess the impact of differentiated training modalities to evaluate the influence on plasma markers of hemostasis, platelet aggregation, and clot microstructure.

**CONCLUSION**

Exercise may be considered a double-edged sword since, on one hand, acute exercise can be a direct cause of a thrombotic event, and on the other hand, exercise training is a potent intervention for lowering the risk of cardiovascular events. Although further studies are required to unravel the influence of different exercise modalities, existing literature points at a greater risk of enhanced thrombogenicity with high, rather than light to moderate intensity exercise. Thus, for patients at risk, the safer recommendation would accordingly be to initiate exercise programs at low to moderate intensities. We propose that studies combining platelet reactivity assay, clinical measures of hemostatic markers, and the novel functional measure of clot microstructure will provide a new level of detailed prediction of the susceptibility to harmful arterial blood clots.

**AUTHOR CONTRIBUTIONS**

LO and MF drafted the manuscript. LG, PE, and YH critically revised and contributed to the content of the manuscript and approved its final version. All authors contributed to the article and approved the submitted version.

**FUNDING**

LO was supported by a grant from the Nordea Foundation. MF and LG were supported by a grant from the Eva and Henry Fraenkel’s Memorial Fund. PE was supported by a visiting professor grant from the University of Copenhagen, Faculty of Science.

**SUPPLEMENTARY MATERIAL**

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fphys.2021.636027/full#supplementary-material

**Supplementary Table 1** | Influence of an acute exercise bout on biomarkers of thrombogenicity.

**Supplementary Table 2** | The effect of different training protocols on biomarkers of thrombogenicity.

**Supplementary Table 3** | Influence of an acute exercise bout on biomarkers of thrombogenicity in patient groups compared to healthy controls.

**REFERENCES**

Albert, C. M., Mittleman, M. A., Chae, C. U., Lee, I. M., Hennekens, C. H., and Manson, J. E. (2000). Triggering of sudden death from cardiac causes by vigorous exertion. *N. Engl. J. Med.* 343, 1355–1361. doi: 10.1056/NEJM200011093431902

Alghani, M., and Heptinstall, S. (2017). Novel strategies for assessing platelet reactivity. *Future Cardiol.* 13, 33–47. doi: 10.2217/fca-2016-0054

Arnett, D. K., Blumenthal, R. S., Albert, M. A., Buroker, A. R., Goldberg, Z. D., Hahn, E. J., et al. (2019). 2019 ACC/AHA guideline on the primary prevention of cardiovascular disease: a report of the American College of Cardiology/American Heart Association task force on clinical practice guidelines. *Circulation* 140, e596–e646. doi: 10.1161/CIR.0000000000000678

Aurigemma, G., Fattorossi, A., Sestito, A., Sgueglia, G. A., Farnetti, S., Buzzonetti, A., et al. (2007). Relationship between changes in platelet reactivity and changes in platelet receptor expression induced by physical exercise. *Thromb. Res.* 120, 901–909. doi: 10.1016/j.thromres.2007.01.009

Badiei, N., Sowedan, A. M., Curtis, D. J., Brown, M. R., Lawrence, M. I., Campbell, A. L., et al. (2015). Effects of unidirectional flow shear stresses on the formation, fractal microstructure and rigidity of incipient whole blood clots and fibrin gels. *Clin. Hemorheol. Microcirc.* 60, 451–464. doi: 10.3233/CH-151924

Bärtch, P. (1999). Platelet activation with exercise and risk of cardiac events. *Lancet* 354, 1747–1748. doi: 10.1016/S0140-6736(99)0259-3

Born, G. V. (1962). Aggregation of blood platelets by adenosine diphosphate and its reversal. *Nature* 194, 927–929. doi: 10.1038/194927b0

Buckley, T., Soo Hoo, S. Y., Shaw, E., Hansen, P. S., Fethney, J., and Toftler, G. H. (2019). Triggering of acute coronary occlusion by episodes of vigorous physical exertion. *Heart Lung Circ.* 28, 1773–1779. doi: 10.1016/j.hlc.2018.11.001

Chen, Y. W., Chen, J. K., and Wang, J. S. (2010). Strenuous exercise promotes shear-induced thrombin generation by increasing the shedding of procoagulant microparticles from platelets. *Thromb. Haemost.* 104, 293–301. doi: 10.1160/TH09-09-0633

Cocks, M., Shaw, C. S., Shepherd, S. O., Fisher, J. P., Ranasinghe, A. M., Barker, T. A., et al. (2013). Sprint interval and endurance training are equally effective in increasing muscle microvascular density and eNOS content in sedentary males. *J. Physiol.* 591, 641–656. doi: 10.1113/jphysiol.2012.239566

Collet, J. P., Allali, Y., Lesty, C., Tanguy, M. L., Silvain, J., Ankri, A., et al. (2006). Altered fibrin architecture is associated with hypofibrinolysis and premature coronary atherothrombosis. *Arterioscler. Thromb. Vasc. Biol.* 26, 2567–2573. doi: 10.1161/01.ATV.0000241589.52950.4c

Collet, J. P., Park, D., Lesty, C., Soria, J., Soria, C., Montalescot, G., et al. (2000). Influence of fibrin network conformation and fibrin fiber diameter on fibrinolysis speed: dynamic and structural approaches by confocal microscopy. *Arterioscler. Thromb. Vasc. Biol.* 20, 1354–1361. doi: 10.1161/01.ATV.20.5.1354

Collins, P., Ford, I., Ball, D., Macaulay, E., Greaves, M., and Brittenden, J. (2006). Rheometrical and molecular dynamics simulation studies of platelet activation and fibrin gels. *Heart Lung Circ.* 15, 17–26. doi: 10.1016/j.hlc.2005.10.011

Curtis, D. J., Brown, M. R., Hawkins, K., Evans, P. A., Lawrence, M. J., Rees, P., et al. (2011). Rheometrical and molecular dynamics simulation studies of incipient clot formation in fibrin-thrombin gels: an activation limited method.
aggregation approach. J. Nonnewton. Fluid Mech. 166, 932–938. doi: 10.1016/j.
jnnfm.2011.04.016
Curts, D. J., Williams, P. R., Badiei, N., Campbell, A. L., Hawkins, K., Evans, P. A., et al. (2013). A study of microstructural templating in fibrin–thrombin gel networks by spectral and viscoelastic analysis. Soft Matter 9, 4883–4889. doi: 10.1039/c3sm50263e
Davidson, J. F., Colvin, B. T., Barrowcliffe, T. W., Dawson, D. W., Machin, S. I., Poller, L., et al. (1988). Guidelines on platelet function testing. The British society for haematology BCSH haemostasis and thrombosis task force. J. Clin. Pathol. 41, 1322–1330. doi: 10.1136/jcp.41.12.1322
Davies, N. A., Lwyd, O., Brugniaux, J. V., Davies, G. R., Marley, C. J., Hudson, D., et al. (2016). Effects of exercise intensity on clot microstructure and mechanical properties in healthy individuals. Thromb. Res. 143, 130–136. doi: 10.1016/j.
thromres.2016.1005.1018
de Groot, P. G., Urbanus, R. T., and Roest, M. (2012). Platelet interaction with the vessel wall. Handb. Exp. Pharmacol. 210, 87–110. doi: 10.1007/978-3-642-29423-5_4
Dimsdale, J. E., and Moss, J. (1980). Plasma catecholamines in stress and exercise. Int. J. Sports Med. 11, 123–127. doi: 10.1055/s-20000300018017
Ebner, M., Birschmann, I., Peter, A., Härtig, F., Spencer, C., Kuhn, J., et al. (2018). Limitations of specific coagulation tests for direct oral anticoagulants: a critical analysis. J. Am. Heart Assoc. 7:e009807. doi: 10.1161/JAHA.118.009807
El-Sayed, M. S. (1993). Fibrinolytic and hemostatic parameter response after resistance exercise. Med. Sci. Sports Exerc. 25, 597–602. doi: 10.1249/00005768-199305000-00011
El-Sayed, M. S., Lin, X., and Rattu, A. J. (1995). Blood coagulation and fibrinolysis levels. Int. J. Sports Med. 16, 23–26. doi: 10.1055/s-2002-35070
Evans, P. A., Hawkins, K., Morris, R. H., Thirumalai, N., Munro, R., Wakeman, L., et al. (2018). The effect of tyramine infusion and exercise on blood flow, coagulation and clot microstructure in healthy individuals. Thromb. Res. 170, 32–37. doi: 10.1016/j.thromres.2018.1007.1025
Lawrence, M. J., Kumar, S., Hawkins, K., Boden, S., Rutt, H., Mills, G., et al. (2014). A new structural biomarker that quantifies and predicts changes in clot strength and quality in a model of progressive haemodilution. Thromb. Res. 134, 488–494. doi: 10.1016/j.thromres.2014.05.039
Lawrence, M. J., Marsden, N., Motahakori, K., Morris, R. H., Davies, G., Hawkins, K., et al. (2016). The effect of temperature on clot microstructure and strength in healthy volunteers. Anesth. Analg. 122, 21–26. doi: 10.1213/ANE.00000000009992
Lee, C. D., Folsom, A. R., and Blair, S. N. (2003). Physical activity and stroke risk: a meta-analysis. Stroke 34, 2475–2481. doi: 10.1161/01.STR.0000091843.02517.9D
Lundberg Slingsby, M. H., Gliemann, J., Thrane, M., Ryter, N., Englund, J., Chan, M. V., et al. (2018). Platelet responses to pharmacological and physiological interventions in middle-aged men with different habitual physical activity levels. Acta Physiol. (Oxf.) 223, e13028. doi: 10.1111/apha.13028
Lundberg Slingsby, M. H., Nyberg, M., Englund, J., Mandrup, C. M., Friikke-Schmidt, R., Kirkby, N. S., et al. (2017). Aerobic exercise training lowers platelet reactivity and improves platelet sensitivity to prostacyclin in pre- and postmenopausal women. J. Thromb. Haemost. 15, 2419–2431. doi: 10.1111/jth.13866
Mackness, N. (2008). Triggers, targets and treatments for thrombosis. Nature 451, 914–918. doi: 10.1038/nature06797
Mehta, J., and Mehta, P. (1982). Comparison of platelet function during exercise in normal subjects and coronary artery disease patients: potential role of platelet activation in myocardial ischemia. Am. J. Cardiol. 50, 103–108. doi: 10.1038/jnc.1982.203
Mills, J. D., Ariëns, R. A., Mansfield, M. W., and Grant, P. J. (2002). Altered fibrin clot structure in the healthy relatives of patients with premature coronary artery disease. Circulation 106, 1938–1942. doi: 10.1161/01.CIR.0000033221.730826
Mittleman, M. A., Maclure, M., Tofler, G. H., Mulry, R. P., Tofler, G. H., Jacobs, S. C., et al. (1993). Triggering of acute myocardial infarction onset by episodes of anger. Determinants of myocardial infarction onset study investigators. Circulation 92, 1720–1725. doi: 10.1161/01.CIR.92.7.1720
Mittleman, M. A., Maclure, M., Tofler, G. H., Sherwood, J. B., Goldberg, R. J., and Muller, J. E. (1993). Triggering of acute myocardial infarction by heavy physical exertion. Protection against triggering by regular exertion. Determinants of myocardial infarction onset study investigators. N. Engl. J. Med. 329, 1677–1683. doi: 10.1056/NEJM199312023292301
Mozaffarian, D., Benjamin, E. J., Go, A. S., Arnett, D. K., Blaha, M. J., Cushman, M., et al. (2016). Executive summary: heart disease and stroke statistics–2016 update: a report from the American Heart Association. *Circulation* 133, 447–454. doi: 10.1161/CIR.0000000000000366

Myers, J., Arena, R., Franklin, B., Pina, I., Kraus, W. E., McInnis, K., et al. (2009). Recommendations for clinical exercise laboratories: a scientific statement from the american heart association. *Circulation* 119, 3144–3161. doi: 10.1161/CIRCULATIONAHA.109.192520

Nachman, R. L., and Rafii, S. (2008). Platelets, petechiae, and preservation of the vascular wall. *N. Engl. J. Med.* 359, 1261–1270. doi: 10.1056/NEJMra0800887

Nyberg, M., Egelund, J., Mandrup, C. M., Andersen, C. B., Hansen, K., Herigel, I. F., et al. (2017). Leg vascular and skeletal muscle mitochondrial adaptations to aerobic high-intensity exercise training are enhanced in the early postmenopausal phase. *J. Physiol.* 595, 2969–2983. doi: 10.1113/JP273871

O'Brien, J. R. (1962). Platelet aggregation: part I some of the adenosine phosphates, thrombin, and cocaine upon platelet adhesiveness. *J. Clin. Pathol.* 15, 446–452. doi: 10.1136/jcp.15.5.446

Pamukcu, B., Oflaz, H., Acar, R. D., Umman, S., Koylan, N., Umman, B., et al. (2015). Platelet function tests: a scientific statement from the American heart association. *Circulation* 119, 3144–3161. doi: 10.1161/CIRCULATIONAHA.109.192520

Pelliccia, A., Sharma, S., Gati, S., Bäck, M., Börjesson, M., Caselli, S., et al. (2020). 2020 ESC guidelines on sports cardiology and exercise in patients with cardiovascular disease: the task force on sports cardiology and exercise in patients with cardiovascular disease of the European Society of Cardiology (ESC). *Eur. Heart J.* 42, 17–96.

Petidis, K., Douma, S., Doumas, M., Basagiannis, I., Vagiatziis, K., and Zamboulis, C. (2008). The interaction of vasoactive substances during exercise modulates platelet aggregation in hypertension and coronary artery disease. *BMC Cardiovasc. Disord.* 8:11. doi: 10.1186/1471-2261-8-11

Rankinen, T., Viitasä, S., Penttilä, I., and Rauramaa, R. (1995). Acute dynamic exercise increases fibrinolytic activity. *Thromb. Haemost.* 73, 281–286. doi: 10.1055/s-0038-1653765

Rauramaa, R., Salonen, J. T., Seppänen, K., Salonen, R., Venäläinen, J. M., Ihnaininen, M., et al. (1986). Inhibition of platelet aggregability by moderate-intensity physical exercise: a randomized clinical trial in overweight men. *Circulation* 74, 939–944. doi: 10.1161/01.CIR.74.5.939

Röcker, L., Drygas, W. K., and Heyduck, B. (1986). Blood platelet activation and increase in thrombin activity following a marathon race. *Eur. J. Appl. Physiol. Occup. Physiol.* 55, 374–380. doi: 10.1007/BF00427736

Sabra, A., Lawrence, M. J., Aubrey, R., Obaid, D., Chase, A., Smith, D., et al. (2017). Characterisation of clot microstructure properties in stable coronary artery disease. *Open Heart* 4:e000562. doi: 10.1136/openhrt-2016-000562

Salehi, S. H., Fatemi, M. J., Asadi, K., Shoa, S., Ghazarian, A. D., and Samimi, R. (2014). Electrical injury in construction workers: a special focus on injury with electrical power. *Burns* 40, 300–304. doi: 10.1016/j.burns.2013.05.019

Sattelmair, J., Pertman, J., Ding, E. L., Kohl, H. W. III, Haskell, W., and Lee, I. M. (2011). Dose response between physical activity and risk of coronary heart disease: a meta-analysis. *Circulation* 124, 789–795. doi: 10.1161/CIRCULATIONAHA.110.101701

Stratton, J. R., Chandler, W. L., Schwartz, R. S., Cerqueira, M. D., Levy, W. C., Kahn, S. E., et al. (1991). Effects of physical conditioning on fibrinolytic variables and fibrinogen in young and old healthy adults. *Circulation* 83, 1692–1697. doi: 10.1161/01.CIR.83.5.1692

Suman, G., Fries, D., Griesmacher, A., Falkensammer, G., Küngler, A., Koller, A., et al. (2007). Blood coagulation activation and fibrinolysis during a downhill marathon run. *Blood Coagul. Fibrinolysis* 18, 435–440. doi: 10.1097/MBC.0b013e328136c19b

Taddei, S., Virdis, A., Ghidoni, L., Versari, D., and Salvetti, A. (2006). Endothelium, aging, and hypertension. *Curr. Hypertens. Rep.* 8, 84–89. doi: 10.1007/s11906-006-0045-4

Thompson, P. D., Franklin, B. A., Balady, G. J., Blair, S. N., Corrado, D., Estes, N. A. III, et al. (2007). Exercise and acute cardiovascular events placing the risks into perspective: a scientific statement from the American Heart Association Council on Nutrition, physical activity, and metabolism and the council on clinical cardiology. *Circulation* 115, 2358–2368. doi: 10.1161/CIRCULATIONAHA.107.181485

Thompson, W. R., Saltis, R., Joy, E., Javorksi, C. A., Stuhr, R. M., and Trilk, J. L. (2020). Exercise Is Medicine. *Am. J. Lifestyle Med.* 14, 511–523. doi: 10.1177/1559827620912192

van den Burg, P. J., Hospers, J. E., Mosterd, W. L., Bouma, B. N., and Huisveld, I. A. (2000). Aging, physical conditioning, and exercise-induced changes in hemostatic factors and reaction products. *J. Appl. Physiol.* (1985) 88, 1558–1564. doi: 10.1152/jappl.2000.88.5.1558

Vinhol, P. J., Hvas, A. M., and Nybo, M. (2014). An overview of platelet indices and methods for evaluating platelet function in thrombocytopenic patients. *Eur. J. Haematol.* 92, 367–376. doi: 10.1111/ejh.12262

Vinhol, P. J., Nybo, M., Nielsen, C. B., and Hvas, A. M. (2017). Light transmission aggregometry using pre-coated microriter plates and a Victor X5 plate reader. *PLoS One* 12:e0185675. doi: 10.1371/journal.pone.0185675

von Klot, S., Mittleman, M. A., Dockery, D. W., Heier, M., Meisinger, C., Hörmann, N. A. III, et al. (2007). Exercise and acute cardiovascular events placing the risks into perspective: a scientific statement from the American Heart Association Council on Nutrition, physical activity, and metabolism and the council on clinical cardiology. *Circulation* 119, 3144–3161. doi: 10.1161/CIRCULATIONAHA.107.181485

Wang, J. S., Chen, H. I. (1995). Effects of exercise training and deconditioning on platelet function in men. *Arterioscler. Thromb. Vasc. Biol.* 15, 1668–1674. doi: 10.1161/01.ATV.15.10.1668

Wang, J. S., Chen, H. I. (1997). Effects of chronic exercise and deconditioning on platelet function in women. *J. Appl. Physiol.* (1985) 83, 2080–2085. doi: 10.1152/jappl.1997.83.6.2080

Yau, J. W., Teoh, H., and Verma, S. (2015). Endothelial cell control of thrombosis. *BMC Cardiovasc. Disord.* 15:130. doi: 10.1186/s12872-015-0124-z

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Olsen, Fischer, Evans, Gliemann and Hellsten. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.