A monocyte-TNF-endothelial activation axis in sickle transgenic mice: Therapeutic benefit from TNF blockade

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1 | INTRODUCTION

A chronic and robust systemic inflammatory state is a striking feature and pathogenic factor in sickle cell anemia (SCA).1 Hence, identification of the core vector(s) underlying inflammation’s evolution and perpetuation should identify useful therapeutic targets. As general underlying processes, attention has focused upon vascular occlusion as the initiator of ischemia/reperfusion injury (I/R) pathophysiology2 and upon hemolysis as a source of toxic heme.3,4 Beyond this, however, the role of specific mediators as antecedent agents remains opaque in its intricacy. Indeed, available data on SCA do not even enable parsing potential mediators into those acting proximately versus more distally.

The literature on SCA, however, does document abnormal activation of blood monocytes and their ability to activate and/or damage vascular endothelial cells in vitro.5–17 This suggests monocyte prominence in clinical disease genesis, if only because monocyte/macrophages are dominant generators of pro-inflammatory cytokines in the broad context of inflammation’s generative role in vascular disease generally.18 The present studies implicate a disease causing vector extending from peripheral blood monocytes (PBM) to the vascular endothelium, with the bridging mediator being tumor necrosis factor (TNF, aka TNFα).

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Our focus upon TNF stems from its roles as a "sentinel cytokine," largely post-transcriptionally regulated, and as an acute-phase initiator of oxidant species and NFκB-driven (and other) responses that have defensive, beneficial roles.19 Yet, its obverse, maladaptive potential can be realized when TNF is produced in excess and/or absent appropriate resolution. Then, its pleiotropic effects can induce multi-faceted inflammatory pathology. Thus, it is the striking clinical benefit of TNF blockade in rheumatoid arthritis and other chronic inflammatory diseases19,20 that prompts our interest in this approach to the stubborn chronicity of the SCA inflammatory state. Yet, the perplexing complexity of TNF biology makes efficacy of TNF-blocking agents impossible to predict with assurance.

This deserves exploration because many well-known TNF effects are directly relevant to pathobiology of clinical sickle disease. To illustrate, we simply focus upon the vascular endothelium, the blood/tissue interface of enormous importance in multiple biological processes. Most globally harmful, TNF causes degradation of the glycocalyx,21 thus jeopardizing its critical roles that include: mediation of shear-dependent functions (e.g., NO production); anchorage of surface enzymes; and repelling potentially adherent blood cells. Separately, TNF jeopardizes NO bioavailability by activating both endothelial arginase (starving eNOS of its required substrate, arginine22) and endothelial NADPH oxidase (depleting tetrahydrobiopterin to provoke superoxide generation by eNOS23). Experimentally, TNF induces endothelial adhesion molecule expression to promote RBC adhesion24 and vasoocclusion.25 TNF exerts many additional adverse effects upon and beyond endothelial cells.

At the level of clinical disease, TNF plays a prominent causative role in organ diseases of general medicine, and these may be instructive regarding their counterparts in SCA. Examples include TNF’s role in: pulmonary hypertension;26 asthma;27 sleep apnea;28 left ventricular dysfunction;29 cognitive, neuropsychiatric and neurologic impairments;30 and pain syndromes.31,32 For most of these organ manifestations within general medicine, TNF blockade using etanercept has yielded clinical improvement.

Therefore, TNF is a therapeutic target that should be considered in SCA. The studies reported here examined effects of the TNF blocker, etanercept, utilizing three sickle transgenic mouse models that exhibit a systemic inflammatory state mimicking that of human sickle disease.33,34 The resulting data create a framework within which this intervention can be envisioned in the sickle disease context.

Note that, due to complexity and variety of experiments, interpretation of individual experiment sets is included in Results section, so that Discussion can address the broader issues. (The data reported here were presented, in preliminary form, at meetings of the American Society of Hematology, 2007–2013).

2 | MATERIALS/METHODS

Some Methods are presented in greater detail in Supporting Information Methods.

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2.1 | Drugs

Etanercept, a chimeric fusion of human IgG1 Fc domain and the 75 kDa extracellular portion of human TNFR2,35 acts as a " decoy " by binding TNF. It is known to block TNF in murine experimental inflammatory disease.56 It also binds the lymphotoxin family, less understood mediators that use the same receptors and mimic TNF itself. Our etanercept dosing (3–10 mg/kg) falls in the lower end of the range used during its preclinical development. We estimate that our highest dose would provide ~10^-fold molar excess over murine blood TNF level.37 By comparison, clinical trials of etanercept for human rheumatoid arthritis used dosing that could have achieved a ~10^-fold excess over human TNF level.

Infliximab is a chimeric, mouse/human hybrid monoclonal specific for TNF. Dosing here (10 mg/kg) would achieve the same fold-excess as for our highest-dose etanercept. The typical human dose is 3 mg/kg, given IV, every few weeks.

Anakinra, a recombinant form of the human IL-1 receptor antagonist, blocks signaling by IL-1α and IL-1β. We examined it because of prior use of an (undefined) IL-1 blocker by Kaul et al.38 Dosing here was 10 mg/kg, with unknown ratio to IL-1 receptor. Anakinra is known to impede experimental murine inflammation.39

Scrambled Control Peptide, obtained via custom synthesis, contains the 12 amino acids of the terminal end of the TNFR2 in scrambled order. For simplicity and to economize on animal consumption, this was always given at 0.1 mg/kg per dose, on same schedule as the other agents.

2.2 | Mice

Mice were housed and bred in our institution’s specific-pathogen-free facility, with routine surveillance testing, to avoid confounding infectious disease. Studies were done with the approval of our Institutional Animal Care and Use Committee. We used NY1DD, S+SAntilles and SS-Berk sickle transgenic mice exhibiting, respectively, mild to moderate to severe phenotypes.34 For NY1DD and S+SAntilles mice we used C57BL/6 controls. For mixed background BERK mice, our breeding strategy ensured background homogenization between SS-BERK and their AA-BERK controls.

2.2.1 | Cross-breeding

We separately crossedbred knockout states for NFκB(p50)14 and Egr-140 into NY1DD mice, using the same strategy we previously described.14 Additionally, we crossedbred the NFκB(p50) knockout into SAntilles mice. Then NFκB(p50)-/- NY1DD and NFκB(p50)-/- SAntilles were crossedbred to ultimately obtain NFκB(p50)-/- S+SAntilles mice. Such knockout transfers always included >10 backcrosses against wild-type C57BL/6.

2.2.2 | Marrow transplantation

As described,14 we previously conducted reciprocal marrow transplantsations between wild-type and NFκB(p50)-/- NY1DD to obtain NY1DD mice having NFκB(p50)-/- in blood cells but not endothelium/tissue, or conversely having NFκB(p50)-/- in endothelium/tissue but not blood cells. Using the same strategy, we here created NY1DD mice that were Egr-1-/- in blood cells but not endothelium/tissue, or vice versa.
2.3 | Study protocols

We tested the impact of drugs, transcription factor knockouts, or transfusion of PBMC using animals at 3.5–4 months of age, unless otherwise indicated. Experiments used pooled animals from multiple litters, with pups from any one litter divided into both test and control groups, using both males and females. Drug treatment studies always included parallel controls receiving scrambled peptide (on same schedule as the test drugs). Drugs were given by intra-peritoneal (ip) injection for short-term experiments and by sub-cutaneous (sq) injection for long-term experiments.

2.3.1 | Ischemia/reperfusion (I/R) model

Some NY1DD mice were subjected to hypoxia/reoxygenation (H/R) stress: 3 hr at normobaric 8% O2, followed by reoxygenation (return to room air, typically overnight). This rapidly triggers actual ischemia/reperfusion (I/R) in sickle – but not normal– mice. Some animals were pretreated with etanercept or infliximab or anakinra, at 10 mg/kg.

2.3.2 | Transfusion of PBMC (peripheral blood mononuclear cells)

Using cardiac blood from pooled donors to isolate PBMC, we transfused 4 × 10⁶ PBMC into naive recipient mice. Some PBMC preparations were monocyte depleted. Some PBMC donor mice were post-H/R; some donors or recipient mice were pretreated with etanercept at 10 mg/kg.

2.3.3 | Long-term therapeutic studies

We treated S₅SAntilles with etanercept (3 mg/kg, sq, once weekly, for 13 weeks), while BERK mice received higher dosing (3 mg/kg for 3 weeks or 6 mg/kg for 6 weeks, sq, twice weekly).

2.4 | Endpoints examined

2.4.1 | Blood counts and inflammatory markers

We performed CBC on cardiac blood using an analyzer standardized for murine blood. We used ELISA kits to assess plasma biomarker levels, and FACS to detect monocyte activation.

2.4.2 | Endothelial cell activation

We monitored expression of Tissue Factor (TF; expressed as the percent of pulmonary veins positive) and vascular cell adhesion molecule 1 (VCAM-1; expressed as percent of 50 ± 10 μm microvessels having >50% circumferential positivity).

2.4.3 | Vascular stasis

Using S₅SAntilles mice with implanted dorsal skinfold chambers, we determined the percentage of previously-flowing microvessels that developed vascular stasis after provocation with either H/R or infusion of hemin. Some were pretreated with etanercept.

2.4.4 | Dermal Cytokines/neuropeptides

We tested cytokine/neuropeptide levels in conditioned medium from skin biopsies (obtained from long-term etanercept-treated SS-BERK mice) that were incubated ex vivo for 24 hours, as described.

2.4.5 | Peri-vascular inflammatory aggregates

We used antibodies to CD11b and von Willebrand Factor (vWF) to identify CD11b+ cells that were perivascular. We discovered this abnormality only retrospectively, so retrievable data are from frozen lung sections that were not prepared with prospective intent to quantify lesion prevalence.

2.4.6 | Pulmonary arteriolar muscularization

The same caveat applies here as well. We used antibodies to murine smooth muscle actin (SMA) and vWF to score 50 ± 10 μm microvessels as non-muscular, partially muscular, or fully muscular. A single value for each mouse was derived from at least 50 vessels from three separated sections.

2.4.7 | Measurement of RVSP (right ventricular mean systolic pressure)

Using isoflurane anesthesia, standardized mechanical ventilation, and a para-sternal lateral thoracotomy, we placed a right ventricular pressure-transducing catheter to capture waveform data that enabled calculation of RVSP. We measured multiple parameters to assess RV mass.

2.5 | Statistics

We made data comparisons using t testing, paired or unpaired, ANOVA as indicated.

3 | RESULTS

We studied transgenic sickle mouse models having phenotypes ranging from mild to severe: NY1DD, S₅SAntilles and SS-BERK. We identified endothelial activation by monitoring endothelial expression level of tissue factor (TF) and VCAM-1. Studies utilized TNF blocker etanercept (E), with a control peptide (CP) always examined in parallel; the latter uniformly failed to exert any effect compared to no treatment. Therefore, to simplify graphics, we focus upon only the most important comparisons. An interpretation of individual experiment sets is included within Results, so Discussion can address the broader issues.

4 | IDENTIFICATION OF A MONOCYTE-TNF-ENDOTHELIAL ACTIVATION AXIS

4.1 | Activated monocytes cause endothelial activation in sickle mice (Figure 1)

Using all three sickle mouse models, we demonstrated the importance of peripheral blood monocytes (PBMC) via transfusion experiments in which peripheral blood mononuclear cells (PBMC) were infused into appropriate control recipient mice.

To illustrate context, our relevant previously published experiment is illustrated here as Figure 1A. When infused into naive (i.e., unstressed) NY1DD recipients, the PBMC harvested from control NY1DD did not activate endothelial TF (bar 1); but infusion of PBMC from H/R-stressed NY1DD donors did do so (bar 2). This activating effect was ameliorated if that donor PBMC population was selectively...
depleted of PBM (bar 3). Thus, PBM activation accounts for the endothelial activation state that is triggered when NY1DD mice are experimentally stressed with H/R exposure to induce I/R.2,14,34 Therefore, we here sought to corroborate this using sickle mice that spontaneously exhibit an inflammatory state, i.e., not requiring challenge by a stressor.

4.1.1 | S1SAntilles mice
Infusion of PBMC from S1SAntilles donors (Figure 1B, bar 2), but not infusion of PBMC from C57BL6 control donors (Figure 1B, bar 1), activated TF in C57BL6 recipient mice. This activating effect was lost if the S1SAntilles donors concurrently were NFκB(p50)-/- (Figure 1B, bar 3). This is supportive because we previously had demonstrated, using NY1DD mice, that NFκB(p50)-/- eliminates ability of PBMC to activate endothelium.14

4.1.2 | SS-BERK mice
Using BERK mice, we also identified the activating role of PBMC on endothelial TF (Figure 1C, as well as endothelial VCAM-1 (Figure 1D). In AA-BERK recipients, activation of both endothelial antigens resulted from infusion of PBMC from SS-BERK donors (bars 2) but not from AA-BERK control donors (bars 1).

4.2 | Endothelial activation is TNF-dependent in sickle mice
We identified involvement of TNF first using the H/R stressed NY1DD mice that developed increased expression of both TF and VCAM-1 (Figure 2A, bars 2), compared to baseline expression in unstressed control NY1DD (bars 1). This activation was unaffected by...
pretreatment with control peptide (CP, bars 3) or heat-inactivated etanercept (bars 4). However, pretreatment with intact etanercept clearly exerted dose-dependent inhibition (bars 5 and 6), also seen using the TNF-specific inhibitor, infliximab (bars 8). By comparison, the IL-1 receptor inhibitor, anakinra, was a less potent inhibitor of TF than was etanercept; but for VCAM-1 inhibition, anakinra exhibited potency equivalent to etanercept (bars 7).

4.3 | Relationship between PBM and TNF in endothelial activation

Because the biology of TNF is so complex (see Discussion), we examined this apparent PBM-TNF link in more detail, using PBMC transfusions and the BERK model. AA-BERK recipient mice that received PBMC from AA-BERK donors exhibited the lower baseline expression of TF (Figure 1C) and VCAM-1 (Figure 1D) (bars 1). But recipients that received PBMC from SS-BERK donors developed activation with higher expression of both TF and VCAM-1 (bars 2). These starting data enable interpretation of the following experiment.

4.3.1 | TF

When we pretreated the SS-BERK donors of with etanercept, their PBMC still induced endothelial TF expression in the AA-BERK recipients (Figure 1C, bar 3). However, when we pretreated the AA-BERK recipients with etanercept, the subsequently transfused SS-BERK PBMC were unable to activate endothelial TF in those recipients (bar 5). Thus, etanercept’s blockade of endothelial TF expression apparently requires the drug to be present when the active PBMC are present. In turn, this implies that etanercept acts either by binding sTNF in plasma or by engaging with tmTNF on endothelial cells themselves.

4.3.2 | VCAM-1

When we examined the same mice for endothelial VCAM-1 expression, a different result emerged. The elevation of recipient VCAM-1 caused by transfusion of SS-BERK PBMC (Figure 1D, bar 2) was ameliorated by etanercept pretreatment of either the PBMC donor (bar 3) or the PBMC recipient (bar 5). Thus, etanercept’s blockade of endothelial VCAM-1 expression presumably results from the drug modifying the PBMC themselves. One possibility is that etanercept could engage tmTNF on the PBMC, with a consequent down-regulating effect on their ability to produce TNF, an effect that persisted when the PBMC were then transfused.

4.4 | Divergence of TF versus VCAM-1 activating mechanisms

Thus, results described so far include two suggestions that activation of endothelial TF and VCAM-1 expression in the sickle I/R context may involve somewhat differing mechanisms. First, the results of treating mice with the TNF blocker etanercept vs. the IL-1 blocker anakinra suggest that endothelial TF and VCAM-1 may have non-identical underlying mechanisms (Figure 2A,B). Second, the data probing the PBMC/etanercept relationship suggested that etanercept may be targeting different features of TNF biology in its inhibition of TF vs VCAM-1 (Figure 1C,D). For this reason, we here describe ancillary experiments that bear on mechanistic divergence of etanercept action and its potential implications for therapeutics.

We focused upon NFκB and Egr-1 because these are dominant regulators of TF expression.

4.4.1 | Egr-1

Using the post-H/R NY1DD model, we observed that Egr-1-/- eliminated the increased endothelial TF expression induced by sickle I/R. This was true both if Egr-1-/- was in endothelium/tissue but not blood cells, and if Egr-1-/- was in blood cells but no endothelium/tissue (Supporting Information Figure S1). These findings are consistent, respectively, with Egr-1’s known prominence as a regulator of TF expression and with its suspected role in monocyte TNF expression.

4.4.2 | NFκB(p50)

We previously reported that the impact of PBM on endothelial TF expression actually requires a NFκB(p50) dependent gene within blood cells. It is relevant that NFκB(p50) is believed to participate indirectly in PBM production of TNF. Interestingly, however, we have now observed that NFκB(p50)-/- in sickle mice actually causes increased expression of endothelial VCAM-1 (Supporting Information Figure S1). This can be explained simply by the NFκB p50/p65 heterodimer’s less robust promotion of gene expression, compared to that of the NFκB p65/p65 homodimer. Hence, a knockout of p50 effectively removes a braking effect.

4.4.3 | NFκB inhibitors

These data suggest it may be advisable to avoid NFκB inhibitors that are truly specific for the p50 component. Fortunately, most are not p50 specific. Nonetheless, we here examined the VCAM-1 expression impact of the NFκB inhibitors we previously identified as effective inhibitors of endothelial TF in sickle mice. We find several of those agents to be effective also in ameliorating the increased VCAM-1 in post-H/R NY1DD mice: lovastatin, andrographolide, curcumin, sulphasalazine and histone deacetylase inhibitors (Supporting Information Table S1).

This VCAM-1 expression scoring was enabled by our discovery, during the course of these studies of an unsuspected, regional heterogeneity in the dynamism of VCAM-1’s activation in sickle mice. VCAM-1 responses, both increases and inhibitions, were detectable only by focusing on vessels 50 ± 10 μm diameter.

5 | THERAPEUTIC BENEFIT FROM TNF blockade

To assess etanercept in a therapeutic context, we conducted long-term studies giving etanercept vs. control peptide to S/S-BERK mice. Doses, schedules and length of treatment are described in Methods. Etanercept exerted a beneficial impact upon nearly all of the endpoints evaluated as being relevant to human sickle disease.
FIGURE 2  TNF blockade ameliorates endothelial activation in sickle mice. Panel A. The low TF expression of unstressed NY1DD mice (bar 1) is increased by H/R exposure (bar 2). Pretreatment with control peptide (bar 3) or with heat inactivated etanercept (bar 4) had no effect, but etanercept exerted dose-dependent inhibition (bar 5 and 6). IL-1 blocker anakinra had very little effect (bar 7), but TNF-specific blocker infliximab strongly inhibited TF (bar 8). For Panel A, n = 8,8,4,10,3,8,5. Panel B shows inhibition of VCAM-1 expression in those same NY1DD mice. For Panel B, n = 10,10,8,3,8,3,5,4. Panels C and D. S+SAntilles mice had elevated TF and VCAM-1 (bars 2), compared to their C57BL controls (bars 1). After long-term treatment (13 weeks), this was inhibited by etanercept (bars 4) but not control peptide (bars 3). For Panel C, n = 7,4,6,11. For Panel D, n = 7,4,5,6. Panels E and F. SS-BERK mice had elevated TF and VCAM-1 (bars 2), compared to their AA-BERK controls controls (bars 1). Etanercept treatment for 3 weeks (bars 4) or 6 weeks (bars 6) inhibited TF and VCAM-1 expression; control peptide did not (bars 3 and 5). For Panels E and F, n = 6,4,5,6,3,4.
5.1 | Inflammatory biomarkers (Table 1)

In general, blood biomarkers of inflammation responded favorably to etanercept, with significant improvements in: soluble VCAM-1 (sVCAM-1), a marker of endothelial activation; SAP (serum amyloid P), the murine equivalent of human CRP; TNF itself; and MCP-1 (monocyte chemotactic protein 1) elaborated, e.g., by endothelial cells in response to TNF. Serum neopterin, a product of monocyte activation, was unhelpful. Blood monocyte activation status (by FACS) revealed a response to etanercept in the proportion of monocytes labeling positively for TNF; but proportions of CD11b+ or IL-6+ did not respond.

Long-term treatment of S+5Antilles mice with etanercept also improved blood Hb and platelet count. An abnormally low platelet count is a feature of murine sickle models (Supporting Information Table S2), as it is in some humans with sickle cell anemia.

5.2 | Pulmonary endothelial activation state

Etanercept was an effective inhibitor of abnormal pulmonary endothelial cell expression of TF and VCAM-1 in all three sickle mouse models models (Figure 2): the acute I/R triggered in NY1DD by H/R exposure (panels A and B); and the spontaneous, chronic I/R state of S+5Antilles (panels C and D) and SS-BERK (panels E and F). Notably, the SS-BERK mice revealed that the suppression of TF expression increased further with longer duration of etanercept treatment.

5.3 | Neuro-inflammatory mediators and pain

In vitro incubation of skin biopsies from SS-BERK mice that had been treated long-term with control peptide or etanercept enabled quantitative measurement of neuro-inflammatory mediator elaboration from actual tissue. As previously reported, such data actually correlate with pain behaviors exhibited by sickle mice.41 Biopsies from the etanercept-treated mice exhibited decreased elaboration of IL-6, MCP-1, substance P and CGRP (calcitonin gene-related peptide) (Supporting Information Figure S2). Although not achieving significance due to small number of animals available, levels of IL-1β, IL-1α, IFNγ, and MIP-1α suggested a downwards trend.

We know that the biologies of acute and chronic pain both involve inflammation having complicated, bidirectional effects between systemic and neuro-inflammatory substances.31,32 Pain behaviors could not be tested here, as the number of available animals was far too small. Yet, the present data suggest that therapy with etanercept might benefit pain.

5.4 | Histopathology

In contrast to an absence of pathology in C57BL6 control mice, livers of S+5Antilles mice displayed acute or chronic coagulative necrosis, indicating ischemia or prior infarction. This was ameliorated both by long-term treatment with etanercept and, revealingly, by presence of the NFxBp50/- state (Supporting Information Table S2).

5.5 | Vascular occlusion

To assess etanercept impact upon acute vascular flow deficiency, we studied S+5Antilles mice bearing dermal windows (to enable microvascular flow visualization). We pretreated them with etanercept or control peptide and then challenged them with an insult known to trigger vascular stasis, either H/R2,14,34 or infusion of hemin.3 The stasis triggered by hemin infusion was not blunted significantly (Figure 3B). This is consistent with our current understanding of these two stasis induction mechanisms. H/R exposure triggers I/R14,34 which involves TNF elaboration in its very earliest stages. In contrast, hemin directly perturbs endothelium, causing stasis via TNF-independent mechanisms in addition to TNF elaboration.3

5.6 | Pulmonary arterial disease

We examined three surrogate markers for presence of pulmonary hypertension.

5.6.1 | Peri-vascular inflammatory aggregates

Staining for CD11b+ evinced the presence of perivascular inflammatory cell aggregates (examples shown in Supporting Information Figure S3).
Since the available material allowed only semi-quantitation, we applied three different measures (Table 2). Each parameter revealed increased prevalence of these inflammatory aggregates in S\textsuperscript{SAntilles} compared to C57Bl6 control, and each was notably diminished in response to etanercept. Because such perivascular inflammatory aggregates are part of the histopathology of human pulmonary hypertension,\textsuperscript{48} this result prompted the following additional studies.

5.6.2 | Pulmonary arteriolar muscularization

Staining for smooth muscle actin (SMA) (examples shown in Supporting Information Figure S3) suggested a trend towards increasing muscularization of \(\sim 50\mu\)m pulmonary arterial vessels as animals matured from age 1 to 4 months, seen for both C57BL6 and S\textsuperscript{SAntilles} mice (Figure 4A). However, at both ages, muscularization was greater for sickle than for normal mice. Notably, after 3 months of etanercept therapy (starting at age 1 month) the muscularization in S\textsuperscript{SAntilles} mice was significantly reduced. Indeed, it appears the etanercept may have both prevented further muscularization and somewhat reduced existing muscularization.

5.6.3 | Elevated right ventricular pressure

As a surrogate measure for possible elevation of pulmonary mean arterial pressure (i.e., pulmonary hypertension), we found that right ventricular (mean) systolic pressure (RVSP) was elevated in S\textsuperscript{SAntilles} compared to C57BL6 controls. Treatment with etanercept for 6 weeks (started at weaning) eliminated RVSP elevation in sickle mice (Figure 4B). After 13 weeks of therapy, however, the differential between groups was preserved, but the statistical significance was lost. This loss may simply be because, as inspection indicates (Figure 4B), inter-individual variability increased markedly as age advanced (as often happens in clinical complications of human SCA). The small number of animals precluded resolution of this.

For both treatment durations, the accompanying necropsy measures of right heart weights (RV/RV\textsubscript{1}LV; RV/tibia length; RV/body weight) did not reveal a significant change in response to etanercept in this study.

6 | DISCUSSION

The present studies identify a monocyte-TNF-endothelial activation axis promotive of pleiotropic disturbances and damage. Results were consistent for the sickle mice of two different genetic backgrounds,

### TABLE 2 Prevalence of peri-vascular inflammatory aggregates (CD11b\textsuperscript{+} cells)\textsuperscript{a}

| Semi-quantification method \textsuperscript{b} | C57BL6 | S\textsuperscript{SAntilles} | control | unRx | CP | E |
| --- | --- | --- | --- | --- | --- | --- |
| mice with large aggregates (n) | 0/5 | 4/6 | 4/6 | 0/11 |
| vessels with smaller aggregate (n)\textsuperscript{b} | 9 ± 8 | 47 ± 9 | 45 ± 6 | 22 ± 18 |

\textsuperscript{a}S\textsuperscript{SAntilles} mice were untreated (unRx) or treated either with control peptide (CP) or etanercept (E) for 13 weeks. Data for small aggregates and cells/vessel are expressed as mean ± SD. Definitions: large and small aggregates were >20 cells or 3–20 cells, respectively, around any given vessel. Above 20, they were too numerous to be discretely identified or counted.

\textsuperscript{b}Untreated S\textsuperscript{SAntilles} vs. C57, \(P = .00034\). CP vs. E treated S\textsuperscript{SAntilles} mice, \(P = .0038\).

\textsuperscript{c}Untreated S\textsuperscript{SAntilles} vs. C57, \(P = 2.4 \times 10^{-5}\). CP vs. E treated S\textsuperscript{SAntilles}, \(P = .042\).
and they were similar for both experimentally triggered I/R (NY1DD mice) and mice having chronic spontaneous I/R (SS-Antilles and SS-BERK mice). We identified a substantial ameliorating effect of the TNF blocker, etanercept, upon nearly all endpoints evaluated, a diverse group of clinical features shared by sickle mice and humans. Thus, interruption of this monocyte-TNF-endothelial vector offers an interesting therapeutic targeting option.

6.1 | Interpreting results of TNF blockade

The complexity of TNF biology renders interpretation of experiments challenging.19,20 Signaling derives from both soluble TNF (sTNF) and cell-bound trans-membrane TNF (tmTNF). The sTNF activates via both receptors, TNFR1 and TNFR2; mTNF activates only via latter. One cell’s tmTNF can receive outside-in activating signals if it engages another cell’s TNFR. Conversely, a TNF blocking antibody may induce positive or negative signaling within a cell upon engaging its tmTNF.49 Finally, plasma contains released TNFRs that can bind TNF. Despite this bewildering intricacy, its net impact and TNF-dependence are experimentally demonstrable. In the present studies, the impact of etanercept was uniformly beneficial.

6.2 | Evidence for a TNF role in sickle cell anemia

The SCA inflammatory state guarantees participation of TNF, the question being with what primacy. sTNF levels often are elevated in SCA,50,51 but this itself may not be very useful information, as has been discovered in the classical TNF-dependent disease, rheumatoid arthritis (RA). Nonetheless, the very spectrum of clinical complications in SCA is consistent with TNF being a participating inflammatory instigator (per Introduction).

Actually, RA provides an instructive paradigmatic context. Since humans having the TNF(-308) promoter GG polymorphism are predisposed to develop inflammatory RA,52 it is interesting that SCA children having that G allele are predisposed to develop large vessel stroke53 that is caused by an inflammatory vasculopathy in the Circle of Willis.1 Indeed, we found that endothelial cells from sickle children with Circle of Willis disease exhibited an exaggerated NFκB activation response to stimulation with TNF/IL-1.54 Thus, current concepts about TNF and inflammation generally, and disease consequent to the sickle mutation specifically, are consistent with the conclusions we draw from the present studies.

6.3 | Is TNF actually the most proximate mediator?

This really is the wrong question. Inflammatory signaling, rather than a strictly hierarchical cascade, is a vast and intricate network. To illustrate, Kaul et al. preliminarily reported that blockade of IL-1βameliorated endothelial activation and inflammation in a sickle mouse model (probing only VCAM-1).38 This is perfectly consistent with our present observations. Moreover, TNF and IL-1β exert overlapping effects, and each induces production of the other. Thus, our results reveal only that the participation of TNF is substantial enough that its inhibition is unambiguously effective and beneficial.

Our results do beg the question: How are sickle monocytes activated in the first place? This is not yet answerable, as many mediators within the sickle milieu are capable of doing so; it seems likely that multiple agents are activating. Nonetheless, we find three TLR4 ligands to be highly likely actors in SCA: free heme from hemolysis, HMGB1 (high mobility group box 1) released during I/R, and heparan sulfate released from endothelial glyocalyx. Each of these ligands can be expected to
exert activating effects via TLR4 on monocytes, leading to TNF release; and TNF actually enhances activity of this TLR4 pathway.55 This would seem to predict the very cyclicity and apparent perpetuity of inflammation that is evident in SCA.

6.4 | Why TNF blockade should work in sickle disease

The remarkably uniform efficacy of etanercept seen here is, we suspect, not because it is exerting “anti-inflammatory” effects in the traditional sense. Rather, it is impeding the actual triggers that comprise the very inception of I/R pathobiology.2 If so, TNF blockade would inhibit — perhaps even halt — the very driver of the perpetual inflammatory process responsible for the vascular dysfunction of SCA. If this I/R model of SCA pathophysiology is valid, TNF can be expected to be ameliorative.

6.5 | What next?

The unavoidable uncertainty inherent in the complexity of TNF biology also complicates therapeutic predictability. Beyond data such as ours, only advancing to pilot study in humans can answer whether or not etanercept may exert benefit for SCA. This, however, may require our field to create solutions to several barriers, as we expect efficacy of etanercept might be exerted over longer rather than shorter timescales. Additionally, nature and mechanism of benefit could be different for acute intervention vs. chronic prevention.

Finally, we are not arguing that etanercept itself would be an ideal therapeutic, because it is expensive and must be administered subcutaneously, although it could be deployed immediately in health systems that can deal with such issues. As alternatives, ongoing development of small molecule TNF blockers may eventually offer very wide applicability and accessibility. Meanwhile, it would be helpful for us to glean insights about pathophysiology that would derive from a focused pilot test of etanercept in humans with SCA. Attempting that would present several significant challenges for which solutions have been proposed elsewhere. Indeed, that might identify additional opportunities for productive therapeutic targeting.

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CONFLICT OF INTEREST

The authors have no conflicts of interest.

AUTHOR CONTRIBUTIONS

Conceived, designed, supervised and guided study: AS, AS, KG, GV, RPH. Conducted experiments: AS, AS, JB, LV, LM, KAN, GOS. Provided critical models, reagents and expertise: RJK, RP, NM, RPH. Wrote or edited manuscript: AS, AS, NM, GOS, JB, RPH.

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SUPPORTING INFORMATION

Additional Supporting Information may be found online in the supporting information tab for this article.

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