Draft Genome Sequence of *Bacillus thuringiensis* Strain m401, Isolated from Honey in Argentina

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**ABSTRACT** We report here the 6,092,003-bp draft genome sequence of *Bacillus thuringiensis* strain m401, a tetracycline-resistant isolate recovered from honey. The isolate contained three plasmids of 8,307 bp, 9,934 bp, and 69,561 bp and a tetracycline resistance gene with high homology to *tet*45 in a contig of 236,180 bp.

Sporo-forming bacteria of *Bacillus*, and also *Paenibacillus larvae*, the causal agent of American foulbrood disease, are frequently present in honey (1, 2). This disease is controlled in beehives with oxytetracycline favoring the emergence of resistance within the bacterial population (3). Bacterial resistance to tetracyclines is mainly due to the presence of *tet* (tetracycline) and *otr* (oxytetracycline) genes frequently associated with mobile elements (4). Tetracycline-resistant strains of the genus *Bacillus* have been found in honey (5).

*Bacillus thuringiensis* strain m401 was recovered from honey samples from the province of Buenos Aires, Argentina (6). A 10-ml honey sample was homogenized at 40°C, diluted with phosphate buffer at pH 7.2, and centrifuged (3,500 × g; 45 min; 4°C). A 3-ml aliquot of the vortexed fluid-sediment mixture was heated to 80°C (10 min), and 10 μl was spread on Mueller-Hinton broth, yeast extract, potassium phosphate, glucose, and pyruvate (MYPGP) agar (6) and on Trypticase soy agar (TSA) (6) and incubated at 37°C and 30°C, from 48 h to 7 days. *Bacillus* species developed in the TSA plates.

Tetracycline resistance strain m401 has been used to study the mechanisms of tetracycline resistance transfer (7). Total DNA was obtained from overnight cultures grown on Mueller-Hinton broth at 32°C with aeration using the Wizard genomic DNA purification kit (Promega). The DNA solution was further purified using phenol/chloroform/isoamyl alcohol at a ratio of 25:24:1 (pH 8.0) and, subsequently, ethanol precipitation. The final pellet was resuspended in buffer (10 mM Tris-Cl, pH 8.5). The quality and quantity of DNA were assessed by gel electrophoresis using a NanoDrop 2000 UV-Vis spectrophotometer (Thermo Fisher Scientific, MA). A library was prepared using the TrueSeq DNA library preparation kit, and 2 × 100-bp paired-end sequencing was performed on an Illumina HiSeq 4000 sequencing system at Macrogen Co. (Seoul, South Korea). Raw reads were error corrected and *de novo* assembled using SPAdes 3.11.1 (8). In addition, raw reads were quality trimmed using Trimmomatic 0.36 (9) and *de novo* assembled with Geneious 9.1.2 (10) using medium-low sensitivity, enabling the option to circularize contigs with matching. The quality of the genome assembly was assessed using QUAST (11). Open reading frames (ORFs), tRNAs, and rRNAs were predicted using Glimmer 3 (12), tRNAscan-SE 2.0 (13), and RNAmmer 1.2 (14), respectively. The functional annotation was carried out using Blast2GO Basic (15).

Sequencing yielded 1,987,020,672 bases in 19,673,472 reads. The genome was assembled into 45 scaffolds (>500 bp; N₅₀, 391,966 bp) with an average coverage of 146×, comprising a total of 6,092,003 bp with a G+C content of 34.72%. The assembly included three circular contigs of 69,561 bp, 9,934 bp, and 8,307 bp. A total of 5,905 genes were annotated, including 3 rRNAs, 25 tRNAs, and 5,877 ORFs. InterPro accession
numbers, Gene Ontology identification numbers, and Enzyme Commission numbers were assigned to 4,764, 4,509, and 1,560 ORFs, respectively. A gene highly homologous to tet45 was detected in a contig of 236,180 bp.

**Data availability.** This whole-genome shotgun project was deposited at DDBJ/EMBL/GenBank under the accession number PYAP00000000. The version described here is the first version, PYAP01000000. Raw reads are available in the NCBI SRA under accession number SRX4662930.

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**REFERENCES**

1. Snowdon JA, Cliver DO. 1996. Microorganisms in honey. Int J Food Microbiol 31:1–26. https://doi.org/10.1016/0168-1605(96)00970-1.
2. Genersch E. 2010. American Foulbrood in honeybees and its causative agent, *Paenibacillus larvae*. J Invertebr Pathol 103:510–519. https://doi.org/10.1016/j.jip.2009.06.015.
3. Evans JD. 2003. Diverse origins of tetracycline resistance in the honey bee bacterial pathogen *Paenibacillus larvae*. J Invertebr Pathol 83:46–50. https://doi.org/10.1006/jip.2003.0039-9.
4. Roberts MC. 2011. Environmental macrolide-lincosamide-streptogramin and tetracycline resistant bacteria. Front Microbiol 2:40. https://doi.org/10.3389/fmicb.2011.00040.
5. López AC, De Ortuzar RV, Alippi AM. 2008. Tetracycline and oxytetacycline resistance determinants detected in *Bacillus cereus* strains isolated from honey samples. Rev Argent Microbiol 40:231–236.
6. Alippi AM, Reynaldi FJ, López AC, De Giusti MR, Aguilar OM. 2004. Molecular epidemiology of *Paenibacillus larvae larvae* and incidence of American foulbrood in Argentinian honeys from Buenos Aires Province. J Apic Res 43:135–143. https://doi.org/10.1080/0022-2011(2004)1110124.
7. Abrahamovich E. 2018. Estudios sobre la transferencia horizontal de resistencia a tetraciclina en bacterias esporuladas aisladas de colmenas de abejas melíferas. Tesis doctoral. Universidad Nacional de La Plata, La Plata, Buenos Aires, Argentina.
8. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
9. Bolger AM, Lohse M, Usadel B. 2014. Trimmmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10.1093/bioinformatics/btu170.
10. Keasey M, Moir R, Wilson A, Stones-Havas S, Cheung M, Surrock S, Buxton S, Cooper A, Markowitz S, Duran C, Thierer T, Ashton B, Meintjes P, Drummond A. 2012. Geniebus: an integrated and extendable desktop software platform for the organization and analysis of sequence data. Bioinformatics 28:1647–1649. https://doi.org/10.1093/bioinformatics/bts199.
11. Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. Bioinformatics 29:1072–1075. https://doi.org/10.1093/bioinformatics/btt086.
12. Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and endosymbiont DNA with Glimmer. Bioinformatics 23:673–679. https://doi.org/10.1093/bioinformatics/btm009.
13. Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. Nucleic Acids Res 25:955. https://doi.org/10.1093/nar/25.5.955.
14. Lagesen K, Hallin P, Rødland EA, Stærfeldt HH, Rogne S, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. Nucleic Acids Res 35:3100–3108. https://doi.org/10.1093/nar/gkm160.
15. Conesa A, Götz S, García-Gómez JM, Terol J, Talón M, Robles M. 2005. Blast2GO: a universal tool for annotation, visualization and analysis in functional genomics research. Bioinformatics 21:3674–3676. https://doi.org/10.1093/bioinformatics/bti610.