Protease nexin-1 (PN-1) is a serine protease inhibitor belonging to the serpin superfamily. This study was undertaken to investigate the regulatory role of PN-1 in the pathogenesis of intervertebral disk (IVD) degeneration. Expression of PN-1 was detected in human IVD tissue of varying grades. Expression of both PN-1 mRNA and protein was significantly decreased in degenerated IVD, and the expression levels of PN-1 were correlated with the grade of disc degeneration. Moreover, a decrease in PN-1 expression in primary NP cells was confirmed. On induction by IL-1β, the expression of PN-1 in NP cells was decreased at day 7, 14, and 21, as shown by western blot analysis and immunofluorescence staining. PN-1 administration decreased IL-1β-induced MMPs and ADAMTS production and the loss of Agg and Col II in NP cell cultures through the ERK1/2/NF-kB signaling pathway. The changes in PN-1 expression are involved in the pathogenesis of IVD degeneration. Our findings indicate that PN-1 administration could antagonize IL-1β-induced MMPs and ADAMTS, potentially preventing degeneration of IVD tissue. This study also revealed new insights into the regulation of PN-1 expression via the ERK1/2/NF-kB signaling pathway and the role of PN-1 in the pathogenesis of IVD degeneration.

Low-back pain (LBP) is among the leading causes of the most costly musculoskeletal problems in adults worldwide. Intervertebral disc degeneration (IDD) is one of the most common disorders reported in orthopedic practice resulting in LBP; the magnitude of the issue is intensified by the increasing adult population, attaining an overall cost exceeding $100 billion per year in the United States1,2. Advances in the research on disc physiology and the etiology of IDD have been made, and a strong association between IDD and LBP has been shown3,4. The pathogenesis of IDD is very complicated and remains poorly understood. To date, possible etiological factors in the pathogenesis of IDD have been identified as aberrant, cell-mediated, age- and genetic-dependent molecular degeneration processes5.

An IVD consists mainly of a central nucleus pulposus (NP) and radially aligned annulus fibrosus (AF), both of which play a key role in spinal column articulation, force coordination, and cushioning against axial load6. During degeneration, the composition and structure of the IVD are altered, resulting in impaired biomechanical function7,8. Papers published in the medical literature9-13 have emphasized that the expression of matrix metalloproteinases (MMPs) and the disintegrin and metalloproteinase with thrombospondin motifs (ADAMTS) genes have essential roles in the degeneration of human IVDs. During the pathogenesis of IDD, degeneration of the extracellular matrix (ECM) is initiated by proteolytic enzymes, including MMP-1, -3, -7, -9, and -13, as well as ADAMTS-4 and ADAMTS-514,15. At a steady state condition, the activity of MMPs is very low in IVD tissue; however, this activity can be up-regulated by inflammatory cytokines16. IL-1β, TNF-α, and other pro-inflammatory cytokines are increased in IDD, which induces MMP production and decreases the synthesis of ECM components17-20.

Protease nexin-1(PN-1) is a serine protease inhibitor with a unique structure shared by most serpins, belonging to the serpin superfamily. The reactive center loop of PN-1 is located near the carboxy-terminal end of the

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serpin domain, which is necessary for its inhibitory activity\(^{21,22}\). PN-1 can inactivate several proteases, including plasmin, plasminogen, and urokinase, preventing cartilage degradation\(^{23,24}\). As is known, the plasmin/plasminogen enzymatic cascades play an important role in cartilage catabolism, which is mediated by activated matrix metalloproteinases (MMP). IVD tissues share pathophysiological characteristics with osteoarthritis (OA)\(^{26}\). Various types of proteases are directly involved in ECM degradation; however, MMPs are considered the major enzymes\(^{27}\). Given the role of serine proteases in OA pathology, the endogenous serine protease inhibitor PN-1 could share a similar role in ECM degeneration involving the activated plasminogen/plasmin and MMP systems in IVD tissue.

Here, we hypothesize that the expression of PN-1 decreases during IDD, which is related to altered disc-cell function and subsequent characteristic features of IDD. Thus, this study aimed to investigate the expression of PN-1 during IDD, and to determine the induction of its regulation by pro-inflammatory cytokine TNF-\(\alpha\) and IL-1\(\beta\). Then, we examined the effects of PN-1 on the expression and activity of MMPs and ADAMTS in NP cells. Finally, the associated signaling pathway was investigated, focusing on the activation of ERK1/2/NF-\(\kappa\)B.

Materials and Methods

This study complies with accepted ethical standards for human and animal research. The research has been approved by the Ethics Committee of Tongji medical college, and written informed consent was obtained from all participants.

Materials. Recombinant human PN-1 was purchased from R&D Systems (2980-PI-010). Recombinant human IL-1\(\beta\) (AF-200-01B-10), TNF-\(\alpha\) (AF-300-01A-10), and TGF\(\beta\)1 (AF-100-21C-10) were obtained from Peprotech. Polyclonal antibodies to PN-1 (ab154591), MMP13 (ab39012), ADAMTS5 (ab153656), and Aggrecan (ab37778) were purchased from Abcam. Mouse anti-human fibronectin N-terminal monoclonal antibody (mAB10936) was obtained from Chemicon (San Francisco, CA, USA). Polyclonal antibodies against COL2 (sc-7764) and P-P65 (sc-33020) were obtained from Santa Cruz. Polyclonal antibodies against P38 (BS1681), P-P38 (BS6381), P-ERK (BS5016), and MMP9(BS6893) were obtained from Biozvorld. P65 (10745-1-AP), and ERK (16443-1-AP) were purchased from ProteinTech Group. The anti-MMP3 (14351) antibody was purchased from CST and the anti-ADAMTS4 (EAP1002) antibody was obtained from Elabscience. All fluorescence-conjugated secondary antibodies, anti-IgG horseradish peroxidase (HRP)-conjugated antibodies, and the LaminB antibody were obtained from BosterBio. DAPI was purchased from Beyotime Biotechnology.

Patient tissue samples. Degenerated IVD tissues were obtained from 32 symptomatic patients undergoing disc excision and spinal fusion surgery. Non-degenerated disc samples were obtained from the intervertebral discs of eight scoliosis patients undergoing deformity correction surgery. The samples were classified by the degree of IVD degeneration according to the magnetic resonance imaging (MRI) classification of disc degeneration\(^{28}\). Degeneration grades were assigned as follows: grade 1–2, non-degenerated (normal disc height); grade 3, mild degeneration (normal to slightly decrease in disc height); grade 4, moderate degeneration (normal of moderately decrease in disc height); and grade 5, severe degeneration (collapsed disc space). Additionally, IVD cryosections from each group were stained with Alcian blue.

Isolation and Culture of NP Cells. Normal human NP cells were isolated from the discs in younger scoliosis patients undergoing deformity correction surgery (Union Hospital, Tongji Medical College). Briefly, NP tissues were minced into small fragments and enzymatically digested in 0.2% type II collagenase and 0.25% trypsin for 3 hours. After being filtered and washed in PBS, the cells were seeded and cultured in growth medium (DMEM/F-12 supplied with 20% fetal bovine serum, 50 U/mL penicillin, and 50 \(\mu\)g/mL streptomycin) in a 5% CO\(_2\) incubator. The cells were passaged two to three times for use in the following experiments.

Stimulation of NP cells. NPs were seeded and cultured in 12-well dishes and allowed to reach 100% confluence. After being serum-starved for 2 hours, the cells were incubated in growth medium alone or in stimulatory medium containing IL-1\(\beta\) (10 ng/mL), TNF-\(\alpha\) (50 ng/mL), or TGF\(\beta\)1 (1 ng/mL) for 7, 14, and 21 days, and the medium was changed every 3 days. In control cultures, the medium was replaced at the indicated times. Each treatment was performed in three different wells. In another experiment, to test the signaling pathways involved in NP cells, PN-1 and/or IL-1\(\beta\) at different concentrations were used to treat cells for 24 h, and inhibitors of NF-\(\kappa\)B (Caffeic Acid Phenethyl Ester) and ERK (GDC-0994) were used.

ELISA. PN-1 is a secreted protein that acts in the ECM, and therefore, the protein analysis was performed on cell-conditioned media. The level of PN-1 was detected by ELISA from the stored supernatant of each group at different time points after induction, according to the manufacturer's procedures (USCN, SED381H).

Immunohistochemistry. Immunohistochemistry (IHC) for PN-1 expression was performed on degenerated IVD tissue. To detect protein expression, antigen retrieval was performed on the IVD cryosections by incubation in 0.8% hyaluronidase at 37\(^\circ\)C for 60 min. The sections were washed gently with PBS for 5 min, and then blocked in 0.5% goat serum for 40 min at room temperature. Subsequently, the samples were incubated with polyclonal anti-PN-1 (1:100), or control rabbit IgG (2 \(\mu\)g/mL) at 4\(^\circ\)C overnight, followed by washing with PBS. Then secondary antibody conjugated to horseradish peroxidase (diluted 1:3000) was applied for 20–30 min at room temperature. After washing, the sections were incubated with diaminobenzidine (DAB; Solarbio, DA1010) reaction solution until color was detected, followed by counterstaining with hematoxylin, and images were captured using a microscope (Olympus).
**Results**

**PN-1 gene expression in human NP tissue.** PN-1 mRNA expression levels in human IVD tissue with differing grades of disc degeneration were normalized to β-actin and presented as $2^{-\Delta\Delta Ct}$. As seen in Fig. 1A, there was significant expression of PN-1 in non-degenerated IVD tissue (Grade 2), but expression was markedly weakened in degenerated IVD tissue. FN, a protein that induces intervertebral disc degeneration, was increased in the IVD samples with increased degrees of disc degeneration (Fig. 1B). Western blot analysis was performed to detect the protein expression of PN-1, FN, and FN-fs in IVD with varying degrees of disc degeneration (Fig. 1C). As the photomicrographs show, the expression of PN-1 was similar to that seen by PCR (Fig. 1D), indicating that gene expression of PN-1 decreased during the process of disc degeneration. FN-fs play important roles in the development of human disc degeneration. In the study, FN and FN-f were analyzed by western blot, which showed that the highest amount of FN-fs emerged in the moderately degenerative discs (grades III) and retained a high level during IDD (Fig. 1E).

**Histological examination for PN-1 expression in human NP tissue.** The degree of degeneration of the IVD tissue was confirmed by Alcian blue staining. IHC was used to confirm the expression of PN-1 in human IVD tissue. As shown in Fig. 2, the extent of PN-1 expression by NP cells was clearly decreased in the degenerated IVD sample. Thus, the expression of PN-1 is inversely correlated with the grade of disc degeneration, in that a higher degenerative degree correlated to a lower expression level of PN-1 (Fig. 2).

**PN-1 Expression in NP cells in response to IL-1β stimulation.** To determine the expression of PN-1 mRNA at the cellular level under degeneration, NP cells were cultured with IL-1β, and mRNA was detected using RT-PCR at different time points (10, 30, and 60 min, and 6, 12, and 24 h). As Fig. 3 shows, administration of IL-1β resulted in a significant decrease in PN-1 mRNA expression at 24 h (Fig. 3A,B). Cell lysates from different
time points were collected and analyzed by western blot, which also confirmed a significant decrease of PN-1 expression at 24 h (Fig. 3C,D). The analysis of PN-1 expression levels on cell-conditioned media was performed by an ELISA, which indicated that PN-1 concentration increased significantly in NP cells in a time-dependent manner after IL-1β induction (Fig. 3E). Therefore, IL-1β induction could decrease the gene expression of PN-1 in NP cells, and the decrease in cytosolic PN-1 protein expression is due to an increase in protein secretion in the culture media.

Pro-inflammatory cytokines IL-1β and TNF-α decrease PN-1 expression in NP cells. To detect the expression of PN-1 regulated by IL-1β and TNF-α, human NP cells were cultured with IL-1β (10 ng/mL), TNF-α (50 ng/mL), IL-1β+TGF-β1 (10 ng/mL), and TNF-α+TGF-β1, and detected by western blotting and immunofluorescent staining. The protein expression levels in different groups were examined at different time points (Fig. 4A). Western blot analysis showed a significant decrease in the PN-1 protein level in NP cells after administration of IL-1β and TNF-α on days 7 (Fig. 4B) and 14 (Fig. 4C). There was still a trend of decreased PN-1 expression at day 21 (Fig. 1D), but it was not as evident as on days 7 and 14. The results also indicated that TGF-β1
Immunofluorescent staining for PN-1 expression was performed in cells cultured with IL-1β and TNF-α at the corresponding time points, which confirmed the decreased expression of PN-1 in NP cells in comparison with the control group at day 7 (Fig. 4E), 14 (Fig. 4F), and 21 (Fig. 4G).

PN-1 inhibits IL-1β-induced MMP production in NP cells. To investigate the effects of PN-1 on the protein expression of matrix degrading enzymes in IVD, NP cells were cultured in the presence of IL-1β with or without PN-1, and proteins were detected by western blotting. Changes in MMP-3, ADAMTS-4, MMP-13, MMP-9, ADAMTS-5, Agg, and COL2 levels were observed (Fig. 5A). Specifically, IL-1β treatment resulted in a significant up-regulation of MMP protein expression after 24 h stimulation in human NP cells. Interestingly, recombinant PN-1 was able to reverse the production of MMPs induced by IL-1β in human NP cells. Compared to IL-1β stimulation alone, PN-1 administration could reverse the expression of MMP-3 (Fig. 5B), ADAMTS-4 (Fig. 5C), MMP-13 (Fig. 5D), MMP-9 (Fig. 5E), and ADAMTS-5 (Fig. 5F) in a dose-dependent manner. The optimal concentration of PN-1 was 100 ng/mL. We detected the expressions of Agg and COL2. The results showed that IL-1β treatment down-regulated expressions of Agg and COL2, while PN-1 administration could attenuate the effects, reaching the highest levels at the optimal dose of 100 ng/mL.

PN-1 inhibits IL-1β-induced gelatinolytic activities of MMPs. MMPs are enzymes that degrade components of the ECM, and among them, MMP-2 and -9 are the most effective gelatinolytic MMPs. Cell gelatinolytic activity in conditioned cell media has been performed successfully. The zymographic pattern (Fig. 6A) revealed gelatinolytic activity in different groups, expected for MMP-2 and MMP-9. The active band for MMP-9

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**Figure 2. PN-1 protein expression in human IVD tissue.** Representative images of PN-1 protein from paraffin-embedded IVD tissue sections (n = 12, 3 for each grade of IVD) that was detected by immunohistochemistry staining. Grade 2 represents a non-degenerated IVD, whereas grades 3, 4, and 5 signify mild, moderate, and severe degeneration, respectively. (Alcian blue staining 100×; IHC 100×, 400×).
(83 kDa) was reduced 0.36 and 0.29 fold after PN-1 administration, compared to after administration of IL-1β alone, at 30 and 60 min (Fig. 6B). Similarly, MMP-2 (62 kDa) was found to be reduced 0.54 and 0.22 fold (Fig. 6C). These results indicated that IL-1β causes a significant up-regulation of MMP-9/2 expression and gelatinolytic activity, while exogeneous PN-1 could invert the effects in human NP cells.

PN-1 inhibits IL-1β-induced MMP production through ERK1/2/NF-κB activation in NP cells. To investigate the effects of PN-1 on the signaling pathway of IL-1β-induced IVD degeneration, we treated human NP cells with IL-1β, with or without specific inhibitors of NF-κB (Caffeic Acid Phenethyl Ester) and ERK (GDC-0994). Levels of phosphorylated and unphosphorylated P38, ERK1/2, and P65 proteins were determined by western blotting (Fig. 7). The levels of unphosphorylated P38 were similar in both groups at different time points. IL-1β stimulation induced increased levels of phosphorylated p38, which was significantly inhibited in NP cells treated by recombinant PN-1 addition (Fig. 7B). ERK1 and ERK2 contribute to cytokine dependent induction of intervertebral disc degeneration, and hence, activate the inflammatory-related signaling molecule NF-κB. To obtain further insights into the underlying mechanism, NF-κB signaling was also investigated. We observed that IL-1β induced increased levels of phosphorylated ERK1/2, which was significantly blocked by the addition of recombinant PN-1 (Fig. 7C,D). Hence, we sought to analyze whether PN-1 was able to modulate the phosphorylated levels of downstream effector P65, besides NF-κB. As shown in Fig. 7E, and as expected, IL-1β was able to increase the phosphorylated levels of P65, and the addition of recombinant PN-1 reduced the phosphorylated levels of P65. Finally, an ERK inhibitor was used, which showed that the phosphorylated levels of ERK1/2 were reduced.
partially blocked by recombinant PN-1 in NP cells (Fig. 7G,H); an NF-κB inhibitor was also used, and as expected, the results showed that levels of P-P65 were significantly blocked by PN-1 addition. Therefore, the ERK1/2/NF-κB signaling pathways were involved in the regulation of PN-1 in the pathogenesis of IVD degeneration.

PN-1 inhibits IL-1β-induced MMP production through ERK1/2/NF-κB signal pathways. To investigate the effect of the PN-1 signaling pathway on the expression of matrix degrading enzymes in IVD, NP cells were cultured in the presence of IL-1β with or without specific inhibitors of NF-kB (Caffeic Acid Phenethyl Ester) and ERK (GDC-0994), and MMP proteins were detected by western blotting (Fig. 8A). Specifically, IL-1β administration could up-regulate the production of MMPs in NP cells after stimulation with PN-1, ERK inhibitor or NF-kB inhibitor. MMP-3 (Fig. 8B), ADAMTS-4 (Fig. 8C), MMP-13 (Fig. 8D), MMP-9 (Fig. 8E), and ADAMTS-5 (Fig. 8F) levels were significantly inhibited. Meanwhile, expressions of Agg (Fig. 8G) and COL2 (Fig. 8H) were up-regulated for the corresponding groups. Therefore, the results showed that IL-1β could up-regulate the expression of MMPs through both ERK1/2/NF-κB signaling pathways in NP cells, which could be suppressed by exogenous PN-1.

Discussion
Our findings from clinical samples provide evidence for our hypothesis that PN-1 is expressed in human IVD tissue and is highly regulated during the process of IDD in vivo and in vitro. These results also indicate that PN-1 could inhibit the activation of MMPs and ADAMTS through the ERK1/2/NF-κB signaling pathway. In all, these findings provide insights into the molecular mechanisms of IDD. Pro-inflammatory cytokines such as IL-1β and TNF-α, influence the degeneration of intervertebral discs, through protease pathways.

PN-1, a 45- to 50-kDa glycoprotein, is encoded by the SERPINE2 gene on human chromosome 2q99-q35. PN-1 is a member of the serine protease inhibitor (serpin) family, involved in tissue remodeling, cellular invasion, matrix degradation, and tumor growth. PN-1 is barely detectable in plasma, but is found in many organs, such as the brain, heart, kidneys, lungs, spleen, muscle, and cartilage, and is produced by most cell types, such as...
Glial cells, smooth muscle cells (SMCs), endothelial cells, and fibroblasts. PN-1 primarily localizes at the cell surface by binding to glycosaminoglycans (GAGs), which trap and potentiate PN-1 activity within the pericellular space, so that proteolysis does not lead to widespread protein destruction. Therefore, the activity of proteases on cells is regulated by PN-1 bound to cell-surface GAGs. Furthermore, PN-1 plays an important role in the process of proteolysis in an irreversible and highly complex manner through the inhibition of proteases.

Figure 5. Comparison of immunoblotting for quantitative analysis of relative protein change in NP cells. Cells were treated with IL-1β (10 ng/mL) alone or combined with PN-1 at different concentrations (0, 10, 50, 100, or 200 ng/mL). (A) Equal amounts of protein extract were resolved by SDS-PAGE, and MMP-3, MMP-9, MMP-13, ADAMTS-4, ADAMTS-5, aggrecan, and COL2 proteins were detected by western blot analysis. (B–H) Densitometric analysis shows the suppression of MMP3 (B), ADAMTS-4 (C), MMP-13 (D), MMP-9 (E), and ADAMTS-5 (F) Protein levels in NP cells treated by IL-1β and PN-1. (G,H) Densitometric analysis shows the promotion effect of Aggrecan (G) and COL2 (H) Protein levels in NP cells treated by IL-1β and PN-1. Data are representative of three independent experiments, and p-values are shown: *p < 0.05; **p < 0.01 compared to IL-1β stimulation alone.

Figure 6. Gelatinolytic activity of matrix metalloproteinase-2 and -9 in NP cells. (A) Molecular markers indicate active MMP-9 and -2 as 92 and 62 kDa proteins, respectively. (B) Zymographic band densities for MMP-9 were quantified. (C) Zymographic band densities for MMP-2 were quantified. Data are representative of three independent experiments, and p-values are shown: *p < 0.05; **p < 0.01, compared to IL-1β stimulation alone.
Osteoarthritis (OA), a joint disease caused by cartilage loss, is strongly associated with a net loss of aggregan and collagen breakdown caused by an imbalance in ECM homeostasis. The function of articular cartilage depends on the molecular composition of the ECM, which mainly consists of collagen II/IX/XI fibrils and proteoglycan-glycosaminoglycan networks of aggregan and hyaluronan. ECM components are essential in regulating chondrocyte metabolism and function, and play a crucial role in the ability of the tissue to withstand compressive forces and respond to mechanical loading. In the cartilage of OA patients, there is a significant increase in the level of serine protease Htra1 (high temperature factor A-1), which mediates proteolysis of aggregan and contributes to OA pathology.

IVD tissues share pathophysiological characteristics with OA; however, little is known about the involvement of serine proteases in the pathogenesis of disc degeneration. Type I and II collagen fibrils make up almost 20% and 70% of the collagen network in the dry weight of the nucleus pulposus and annulus fibrosus, respectively. Previous studies have confirmed that IDD is associated with fibrosis of the NP, in which the collagen meshwork is gradually replaced by fibrils of abnormal size and rigidity, resulting in altered gene expression for matrix molecules, degradation enzymes, and catabolic cytokines. In addition, alterations in PG lead to a decrease in hydration, loss of structural integrity, and an inability to withstand loads. The molecular mechanisms of collagen fibrilogenesis are complicated, but highly related to mechanical stress, collagen genetic types, and aging. Data are available for structural proteins of IVD, such as aggregan and collagen; however, little is known about the longevity of serine proteases from this tissue. The imbalance between serine proteases and serpins may have an effect on the degeneration of IVD; thus, expression of PN-1 is important to gain further insight into the mechanisms of IDD.

Figure 7. The suppressive effect of PN-1 on the IL-1β-induced NF-κB pathway. (A) NP cells were stimulated with IL-1β alone or combined with PN-1. Cells from different groups were collected and whole-cell protein was extracted at 10, 30, and 60 min time points. The phosphorylated and unphosphorylated levels of P38, ERK1/2, and P65 were detected by western blot analysis. (B–E) Densitometric analysis shows the suppression of P-P38 (B), P-ERK-1 (C), P-ERK-2 (D), and P-P65 (E) protein levels in NP cells treated with IL-1β and PN-1. F. NP cells were stimulated with IL-1β alone or in combination with PN-1 or ERK inhibitor. The phosphorylated and unphosphorylated levels of ERK1/2 were detected by western blot analysis. (G–H) Densitometric analysis shows the suppression of P-ERK-1 (G) and P-ERK-2 (H) protein levels in NP cells treated with IL-1β alone or combined with PN-1 or NF-kB inhibitor. The levels of phosphorylated and unphosphorylated P65 were detected by western blot analysis.

Nucleoprotein extracts were prepared and analyzed for the expression of P-P65. (I) NP cells were stimulated with IL-1β alone or combined with PN-1 or NF-kB inhibitors. Data were obtained from three independent experiments. GADPH/Lamin B were used as the loading control. *p < 0.05; **p < 0.01, compared to IL-1β stimulation alone.
The present study was designed to investigate the changes of PN-1 expression in degenerated IVD of different grades using RT-PCR and IHC. Investigations in human IVD tissue have demonstrated a decreased expression of PN-1 following IDD. Moreover, we found that PN-1 mRNA and protein levels were coordinately regulated during the progression of IDD in a stage-specific manner. Inflammatory cytokines, such as IL-1β and TNF-α, have been known to induce degeneration of IVD. Based on in vitro experiments, changes in MMP protein and mRNA expression levels have been detected by western blotting and RT-PCR, and activity of MMPs have been examined by gelatin zymography. IL-1β, TNF-α, and other pro-inflammatory cytokines have also been shown to induce MMP production in NP cells. As our results indicated, when stimulated with IL-1β, the secretion of MMPs and ADAMTS by human NP cells increased, whereas PN-1 administration could antagonize these proteases and potentially preserve IVD tissue from degeneration.

Fibronectin (FN) and its fragments (FN-fs) have been found to accumulate during disc degeneration and acceleration of IVD degeneration in rabbits. Within human degenerated-disc tissue samples, a marked increase in FN and FN-fs has been observed; thus, they are considered an integral part of the underlying pathology of IDD. PN-1 activity is at least partly regulated by the extracellular matrix component fibronectin, and fibronectin and its fragments have been found to accumulate during disc degeneration and acceleration of IVD degeneration in rabbits. Within human degenerated-disc tissue samples, a marked increase in FN and FN-fs has been observed; thus, they are considered an integral part of the underlying pathology of IDD.

Nuclear factor kappa B (NF-κB) is a family of transcription factors, which becomes activated in response to inflammation, damage, and stress. MMPs have been identified as NF-κB target genes in IVD cells responsible for ECM degradation, including MMP-1, MMP-2, MMP-3, MMP-9, and MMP-13. NF-κB play a role in the regulation of MMP expression.
central role in MMP and ADAMTS expression in NP cells stimulated by IL-1β. Activation of the NF-κB signaling pathway results in increased matrix-degrading enzyme activity in the NP, which is an important catabolic pathway involved in IDD. Therefore, NF-κB pathway has been fully investigated in human NP cells stimulated with TNF-α.

Disc degeneration is characterized by the loss of Agg and COL2. Agg is a type of proteoglycan responsible for the normal structure of discs, while COL2 is an important component of the ECM, helping the IVD bear pressure. In this study, IVD degeneration induced by IL-1β lead to loss of Agg and COL2 in NP cells, which was associated with upregulation of MMPs and ADAMTS. PN-1 intervention reversed the downregulation of Agg and COL2 and the upregulation of MMPs, implying that PN-1 could partially antagonize NF-κB pathway-specific transcription factors and contribute to maintaining matrix integrity of the IVD. Moreover, we found that PN-1 could inhibit IL-1β-induced activation of ERK1/2 in NP cells. Therefore, PN-1 intervention can potentially reverse the expression of MMPs and ADAMTS through the ERK1/2/NF-κB signaling pathway in IDD. Agg and COL2 are necessary for the rebuilding of IVD, which also can be upregulated through the pathway by PN-1.

In conclusion, under inflammatory conditions, activated ERK1/2/NF-κB contributes to the pathogenesis of IDD, manifested by the upregulation of MMPs and ADAMTS and degraded disc matrix macromolecules Agg and collagen II. PN-1 is involved in pathogenesis, at least in part by suppressing IL-1β-induced activation of ERK1/2/NF-κB and its downstream targets. Thus, decreased expression of PN-1 may serve as an important endogenous mechanism to increase the deleterious effects of excessive serine protease activity within the IVD.

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Author Contributions

X.W. designed the study and drafted the manuscript W.L. and Y.S. carried out the immunoassays and participated in collecting clinical samples Z.D., S.Y. and Z.S. participated in the design of the study, did the statistical analysis and helped revise the manuscript Y.G., S.L. and K.W. carried out the molecular mechanism studies and Gelatin zymography C.Y. conceived the study, participated in designing the study and helped revise the manuscript All authors read and approved the final manuscript.
Additional Information

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