Real, Spain (E. Perez-Ramirez, U. Höfe)

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References

1. Bofill D, Domingo C, Cardehosu N, Zara-
goza J, de Ory F, Minguell S, et al. Human West Nile virus infection, Catalonia, Spain. Emerg Infect Dis. 2006;12:1163–4.

2. Kaptoul D, Viladrach PF, Domingo C, Niubó J, Martínnez-Yélamos S, de Ory F, et al. West Nile virus in Spain: reports of the first diagnosed case (in Spain) in a human with aseptic meningitis. Scand J Infect Dis. 2007;39:70–93. DOI: 10.1080/0365540600740553

3. Figuerola J, Soriguer R, Rojo G, Gómez-Tejedor C, Jiménez-Claver MA. Seroconversion in wild birds and local circulation of West Nile virus, Spain. Emerg Infect Dis. 2007;13:1915–7.

4. Figuerola J, Jiménez-Claver MA, Rojo G, Gómez-Tejedor C, Soriguer R. Prevalence of West Nile virus neutralizing antibodies in colonial aquatic birds in southern Spain. Avian Pathol. 2007;36:209–12. DOI: 10.1080/00365540600743239

5. Höfe U, Blanco JM, Crespo E, Naranjo V, Jiménez-Claver MA, Sanchez A, et al. West Nile virus in the endangered Spanish imperial eagle. Vet Microbiol. 2008;129:171–8. DOI: 10.1016/j.vetmic.2007.11.006

6. Ward CL, Dempsey MH, Ring CJ, Kempson RE, Zhang L, Gor D, et al. Design and performance testing of quantitative real time PCR assays for influenza A and B viral load measurement. J Clin Virol. 2004;32:179–88. DOI: 10.1016/S1386-6532(03)00122-7

7. Credan JL, Graham DA, McCullough SJ. Detection and differentiation of pathogenicity of avian paramyxovirus serotype 1 from field cases using one-step reverse transcriptase polymerase chain reaction. Avian Pathol. 2002;51:493–9. DOI: 10.1080/030794502100005860

8. Jiménez-Claver MA, Agüero M, Rojo G, Gómez-Tejedor C. A new fluorescent real-time RT-PCR assay for detection of lineage 1 and lineage 2 West Nile viruses. J Vet Diagn Invest. 2006;18:459–62.

9. Scaramozzino N, Crance JM, Jouan A, DeBriel DA, Stolf F, Garin D. Comparison of flavivirus universal primer pairs and development of a rapid, highly sensitive heminested reverse transcription-PCR assay for detection of flaviviruses targeted to a conserved region of the NS5 gene sequences. J Clin Microbiol. 2001;39:1922–7. DOI: 10.1128/JCM.39.5.1922-1927.2001

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LETTERS

Merkel Cell Polyomavirus and Merkel Cell Carcinoma, France

To the Editor: Merkel cell carcinoma (MCC) is a primary cutaneous neuroendocrine tumor. This aggressive skin cancer is uncommon but increasing in frequency. During 1986–2001, incidence rate tripled; average annual increase was 8% (1). MCC shares epidemiologic features with Kaposi sarcoma, a malignant tumor associated with human herpesvirus 8 infection (2). In particular, MCC affects predominantly patients with B-cell lymphoid tumors (3). In France, 60% of MCC patients have a history of psoriasis, which may support the hypothesis of an infectious origin of MCC.

A new polyomavirus, provisionally named Merkel cell polyomavirus (MCPyV), has been recently identified in tumor tissue from patients with MCC. Furthermore, clonal integration of viral DNA within the tumor genome was observed in most of the cases (7). To assess the implication of MCPyV in MCC, we tested tumor biopsy samples collected from 9 patients with MCC. Patient median age was 65 years, and 2 patients were immunocompromised (patient 1 had a lymphoma, which was treated with rituximab; patient 7 had psoriatic rheumatism, which was treated with corticosteroids and methotrexate). As controls, biopsy samples from 15 patients with diverse proliferative or inflammatory skin or mucosal lesions were tested (Table).

DNA was extracted from fresh tissue samples by using the QIAamp DNA Mini Kit (QIAGEN, Courtaboeuf, France) according to the manufacturer’s instructions. Paraffin was removed from previously formaldehyde-fixed, paraffin-embedded biopsy samples with xylene, and the samples were rehydrated with decreasing concentrations of ethanol. The extracts were tested for MCPyV DNA by PCR using 3 sets of primers initially described by Feng et al. (7) to target the predicted T-antigen (LT1 and LT3 primer pairs) and the viral capsid (VP1 primer pair) coding regions. Extracted DNA (5 μL) was added to 45 μL of the reaction mixture, which contained 5 μL 10× PCR buffer, 10 μL 5× Q-solution (QIAGEN), 2.5 mmol/L MgCl2, 200 μmol/L each dNTP, 2.5 units Taq DNA polymerase (QIAGEN), and 15 pmol of each primer. Touchdown PCR conditions were as follows: 95°C for 5 min followed by 35 cycles of denaturation at 95°C for 30 s; annealing at 61°C (10 cycles), 57°C (15 cycles) for 30 s; extension at 72°C for 1 min; and a final extension step at 72°C for 10 min. Amplification products were subjected to electrophoresis in a 2% agarose, 1× Tris-borate-EDTA gel stained with ethidium bromide and examined under UV light. The sizes of the fragments amplified with the LT1, LT3, and VP1 primers were 439, 308, and 351 bp, respectively. A negative control was included in each experiment; positive samples were confirmed by analyzing a second stored sample aliquot, and the amplified fragments were sequenced by using the same primers used for the amplification. The sequences were submitted to GenBank under accession numbers AM992895–AM992906. Total DNA level in sample extracts was measured by using the LightCycler control DNA kit targeting the β-globin gene (Roche Berlin, Germany). Results showed 100% of positivity.

Table

| Patient | Age | Gender | Histology | MSP108 | MSP560 |
|---------|-----|--------|-----------|--------|--------|
| 1       | 73  | M         | +         | +      | +      |
| 2       | 74  | F         | +         | +      | +      |
| 3       | 68  | M         | +         | +      | +      |
| 4       | 70  | M         | +         | +      | +      |
| 5       | 65  | M         | +         | +      | +      |
| 6       | 67  | M         | +         | +      | +      |
| 7       | 69  | M         | +         | +      | +      |
| 8       | 62  | M         | +         | +      | +      |
| 9       | 64  | M         | +         | +      | +      |
| 10      | 71  | F         | +         | +      | +      |

MSP108 and MSP560 are specific to MCPyV and not to other polyomaviruses. Therefore, the observed positivity confirms the presence of MCPyV in these MCC cases. These results suggest that MCPyV infection may play a role in MCC carcinogenesis, either as a risk factor or as a causative agent.
MCPyV sequences were detected in 8 of the 9 patient samples and in none of the control samples (Table). Results for all 8 patients were positive with the LT3 primer pair, whereas they were positive for only 5 with the VP1 primer pair and only 1 with the LT1 primer pair (Table). Because the LT1, VP1, and LT3 primer pairs generate the longer, intermediate, and shorter DNA fragments, respectively, the difference in sensitivity could result from the deleterious effect of formaldehyde fixation on DNA; this effect would increase with the length of the fragment to be amplified. The negative result obtained for patient 8 might suggest that some MCC patients are not infected with MCPyV. This explanation is in accordance with the findings of Feng et al., who reported 80% prevalence of MCPyV in patients with MCC (7). Nevertheless, the single negative result observed in our study might alternatively be explained by insufficient tissue or by DNA degradation through the fixation and embedding process. Indeed, the level of β-globin gene DNA was much lower in the sample from this patient (441 pg/μL) than in samples from the other patients (median 13,500 pg/μL, interquartile range 8,902–19,750 pg/μL).

As observed with human papillomaviruses, a gene disruption caused by viral DNA integration into the host genome might be an alternative hypothesis to explain the lack of amplification of an MCPyV genome region (8). Sequences of the amplified PCR product were 99%–100% identical to those reported by Feng et al. (7), which indicates that this virus is genetically stable.

In summary, we detected MCPyV DNA sequences in 8 of 9 tumor samples from patients with MCC but in none of 15 control samples. Our results confirm the likely association of MCPyV with MCC. The epidemiologic characteristics as well as the carcinogenic role played by this newly discovered virus need to be more thoroughly investigated.

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References

1. Hodgson NC. Merkel cell carcinoma: changing incidence trends. J Surg Oncol. 2005;89:1–4. DOI: 10.1002/jso.20167
2. Chang Y, Cesarman E, Pessin MS, Lee F, Culpepper J, Knowles DM, et al. Identification of herpesvirus-like DNA sequences in AIDS-associated Kaposi’s sarcoma. Science. 1994;266:1865–9. DOI: 10.1126/science.7997879
3. Penn I, First MR. Merkel’s cell carcinoma in organ recipients: report of 41 cases. Transplantation. 1999;68:1717–21. DOI: 10.1097/00007890-199912150-00015

Table. Merkel cell polyomavirus from 9 patients with Merkel cell carcinoma and 15 control patients, France*

| Patient no. | Age, y | Sex | Diagnosis | Sample | LT1 | LT3 | VP1 |
|------------|--------|-----|-----------|--------|-----|-----|-----|
| 1          | 57     | M   | MCC, primary | Fresh biopsy, buttock | +   | +   | +   |
| 2          | 61     | F   | MCC, primary | FFPE biopsy, buttock | –   | +   | –   |
| 3          | 57     | F   | MCC, primary | FFPE biopsy, lower eyelid | –   | +   | +   |
| 4          | 82     | F   | MCC, primary | FFPE biopsy, cheek | –   | +   | –   |
| 5          | 78     | M   | MCC, metastatic | Fresh biopsy, jugular lymph node | –   | +   | +   |
| 6          | 80     | F   | MCC, primary | Fresh biopsy, upper eyelid | –   | +   | +   |
| 7          | 65     | M   | MCC, primary | FFPE biopsy, temple | –   | +   | –   |
| 8          | 81     | M   | MCC, primary | FFPE biopsy, forearm | –   | –   | –   |
| 9          | 60     | M   | MCC, primary | FFPE biopsy, forearm | +   | +   | –   |
| 10         | 61     | M   | Hyperkeratosis | Fresh biopsy, foot | –   | –   | –   |
| 11         | 49     | M   | Seborrheic keratosis, penis | Fresh biopsy, penis shaft | –   | –   | –   |
| 12         | 60     | M   | Nonspecific lesion, esophagus | Fresh biopsy, lesion | –   | –   | –   |
| 13         | 40     | M   | Nasal papilloma | Fresh biopsy, papilloma | –   | –   | –   |
| 14         | 58     | M   | Anal condylomas | Fresh biopsy, condyloma | –   | –   | –   |
| 15         | 44     | M   | Epidermodysplasia verruciformis | Fresh biopsy, skin | –   | –   | –   |
| 16         | 19     | M   | Cutaneous warts | Fresh biopsy, wart | –   | –   | –   |
| 17         | 57     | F   | Cutaneous warts | Fresh biopsy, wart | –   | –   | –   |
| 18         | 64     | F   | Cutaneous nodule | Fresh biopsy, skin | –   | –   | –   |
| 19         | 6      | F   | Pharyngeal papillomatosis | Fresh biopsy, pharynx | –   | –   | –   |
| 20         | 27     | M   | Vocal cord poly | Fresh biopsy, polyp | –   | –   | –   |
| 21         | 58     | M   | Lichen | Fresh biopsy, skin | –   | –   | –   |
| 22         | 63     | M   | Skin inflammation | Fresh biopsy, skin | –   | –   | –   |
| 23         | 72     | M   | Skin inflammation | Fresh biopsy, skin | –   | –   | –   |
| 24         | 57     | M   | Skin inflammation | Fresh biopsy, skin | –   | –   | –   |

*MCC, Merkel cell carcinoma; +, positive Merkel cell polyomavirus PCR amplification; FFPE, formaldehyde-fixed paraffin-embedded; –, negative Merkel cell polyomavirus PCR amplification.
Worldwide Prevalence of Head Lice

To the Editor: Pediculosis capitis has been well-known since antiquity (1). Human infestation can result in psychological frustration for parents and children (2); furthermore, preventive and therapeutic practices, such as head shaving and the “no-nit” policy of excluding infected children from school, can also induce social stress.

We sought to synthesize the available evidence regarding the worldwide prevalence of lice infestation in the 21st century by conducting a literature search of PubMed and Scopus databases in which we searched for the term pediculosis. We also searched Google for the terms head lice/pediculosis capitis and individual country names and evaluated references of the articles and reports retrieved through this search. Eligible studies were archived from January 1, 2000, to January 18, 2008.

We retrieved 55 studies (online Technical Appendix, available from www.cdc.gov/EID/content/14/9/1493-Techapp.pdf). Most studies referred to schoolchildren, but some involved refugees, urban slums, child labor, jails, orphanages, and fishing communities.

Most studies had been conducted in Asia; Turkey was overrepresented. Prevalence varied from 0.7% to 59% and was higher in girls and women. Of the 29 studies, 24 involved schoolchildren; the other studies involved refugee children, child laborers, the general population, street children, jail inmates, and children accompanying their mothers in prison.

In Europe, prevalence varied from 0.48% to 22.4%. However, 1 study reported a much higher annual incidence (37.4%) in England (3). A study in the Ukraine showed increasing adult representation in the overall affected population (4). Six studies involved schoolchildren; the remaining studies involved refugees, homeless persons, and the general population.

Data from Africa, with the exception of 1 study in South Africa, were derived from Egypt. Prevalence varied from 0% to 58.9% and was higher in females. The study in South Africa (5) challenges the generally accepted concept that head lice infestation refers to lower socioeconomic status; of 2 schools, 1 in a low socioeconomic status area, populated by black students only, and the other in a high socioeconomic status area, populated by students of various races, head lice infestation was found only in the second school, solely among white pupils. Of 6 studies in Egypt, 4 involved diverse populations: urban poor preschool children, orphanage children, and the general population.

Most studies in the Americas were conducted in Brazil, although we also found data from the United States, Cuba, and Argentina. Prevalence varied from 3.6% to 61.4% and was higher in females. Of 7 studies, 4 involved populations other than schoolchildren to some extent: urban slum residents, fishing community residents, adolescents and adults sampled randomly from the general population, elderly nursing home residents, and persons living with repeatedly infested children. A recent study in Brazil (6) noted that prevalence rates determined by visual inspection are twice that of rates determined by hair analysis.

Only 1 study has been performed in Oceania. This study in Australia reported prevalence of 13% and that girls were more likely to have active infection.

Our review shows that pediculosis capitis is widespread throughout the world and does not discriminate on socioeconomic status grounds. The traditional perception of head lice as a parasitosis exclusively associated with schoolchildren of low socioeconomic status is challenged by some of the reports (online Technical Appendix).