**Tal2** expression is induced by all-trans retinoic acid in P19 cells prior to acquisition of neural fate

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TAL2 is a member of the basic helix-loop-helix family and is essential for the normal development of the mouse brain. However, the function of TAL2 during brain development is unclear. P19 cells are pluripotent mouse embryonal carcinoma cells that adopt neural fates upon exposure to all-trans retinoic acid (atRA) and culture in suspension. We found that the expression of Tal2 gene was induced in P19 cells after addition of atRA in suspension culture. Tal2 expression was detected within 3 h after the induction, and had nearly returned to basal levels by 24 h. When GFP-tagged TAL2 (GFP-TAL2) was expressed in P19 cells, we observed GFP-TAL2 in the nucleus. Moreover, we showed that atRA and retinoic acid receptor \(\alpha\) regulated Tal2 expression. These results demonstrate for the first time that atRA induces Tal2 expression in P19 cells, and suggest that TAL2 commits to the acquisition of neural fate in brain development.

**TAL2** is a transcription factor belonging to the basic helix-loop-helix (bHLH) family\(^1,2\). The Tal2 gene is transcribed in SUP-T3 cells derived from a human T-cell leukemia; these cells harbor the t(7;9)(q34;q32) chromosomal translocation\(^3\). A translocation between chromosomes 7 and 9 is associated with human T-cell acute lymphoblastic leukemia (T-ALL)\(^4,5\). Thus, TAL2 may be an oncogene that is activated as a consequence of the chromosomal translocation in T-ALL\(^6\). Moreover, Tal2 overexpression is confirmed in some lung adenocarcinoma tumor tissues\(^7\) and conversely, Tal2 expression is significantly lower in ovarian carcinoma tissue than that in normal tissues\(^8,9\).

Although aberrant TAL2 expression may be involved in development of some cancers, TAL2 is essential for the normal development of the mouse brain. Tal2 is expressed during embryogenesis, and sites of Tal2 expression include specific regions of the diencephalon, mesencephalon and metencephalon\(^10\). Tal2-null mutant mice are viable at birth and initially appear normal. However, they develop signs of runting and die between 13 and 32 days after birth; additionally, they display a prominent hydrocephalus at death\(^11\). These findings indicate that TAL2 plays a pivotal role in development of the mature central nervous system; nevertheless the function of TAL2 in the development processes remains unclear.

P19 cells are a line of pluripotent mouse embryonal carcinoma (EC) cells derived from a teratocarcinoma that formed following transplantation of a 7.5 day embryo into a testis\(^12\). P19 cells can be induced to differentiate into derivatives of three germ layers—endoderm, mesoderm, or ectoderm—depending on the inducers and the culture conditions. Two different treatments—addition of all-trans retinoic acid (atRA) and suspension culture for cell aggregation—are required to induce neural differentiation of P19 cells\(^12–14\). Various genes—including Neurod, Neurog1\(^15\) and Fgf8\(^16\)—that are implicated in neural development are induced in P19 cells following these treatments and during the subsequent neural differentiation. atRA is a biologically active form of vitamin A that functions as the activating ligand for three isoforms of retinoic acid receptor (RAR); these isoforms—\(\alpha\), \(\beta\), and \(\gamma\)—can each form heterodimers with retinoid X receptors (RXRs)\(^17\). These receptors transduce atRA signals, belong to the family of nuclear receptors, and induce the transcription of target genes when bound with atRA\(^18,19\). atRA plays an important role in early embryonic development and in differentiation of the nervous system including patterning of the brain, specification of neural cell fate, and stimulation of neurite outgrowth\(^20,21\). Additionally, atRA can induce EC cells, including P19 cells, and embryonic stem cells to adopt neural fates\(^22,23\). P19 cells have been used as model cells for the studies of neural differentiation, and these studies have led to the discovery of a number of genes that are important for neural development in vivo\(^22,24,25\). To understand how the neural fate is acquired in early embryonic
Results

Tal2 was transiently expressed in P19 cells introduced into the neural differentiation. We found that Tal2 expression was upregulated in P19 cells introduced into the neural differentiation for our study using DNA microarray experiments (unpublished data). To study the alteration of Tal2 expression in more detail here, we extracted RNA from P19 cells 0, 3, 6, 12, 18, 24, 36 and 48 h after addition of atRA in suspension culture, and investigated Tal2 expression in P19 cells (Fig. 1a). We also examined the expression of Pou5f1 gene (also known as Oct3/4), which encodes the transcription factor in pluripotent cells, and Ascl1 gene (also known as Mash1), which encodes a neurogenic bHLH transcription factor, to assess whether P19 cells were induced to neural cell fate. Pou5f1 expression decreases during neural differentiation of P19 cells26,27; contemporaneously, Ascl1 expression increases27,28. As previously reported, Pou5f1 expression decreased and Ascl1 expression increased contemporaneously in atRA-treated, aggregated P19 cells. This observation demonstrated that P19 cells lost the pluripotency by addition of atRA in suspension culture and were entered into neural cell fate. Tal2 expression increased within 3 h after treatment, and Tal2 mRNA levels was returned to the basal levels within 24 h after treatment. In contrast, the expression of all three genes—Tal2, Pou5f1, and Ascl1—was not altered by DMSO treatment in suspension culture (see Supplementary Fig. S1 online).

Next, we used real-time PCR assays to measure Tal2 expression in P19 cells (Fig. 1b). Tal2 expression increased 3 h and 6 h after treatment. A reduction in Tal2 expression from peak levels was first observed at 12 h and Tal2 expression was close to the basal levels at 24 h. These results indicate that Tal2 expression is altered in P19 cells for one day.

It has been reported that Tal2 expression was observed in embryonic head (E10.5 to E14.5)10. We also confirmed Tal2 expression in embryogenesis (Fig. 1c). For this purpose, we used a mouse cDNA panel and RT-PCR. As previously reported, Tal2 expression was evident at E11 and was not observed in adult brain. In addition, Tal2 was also hardly observed in E7, E15 and E17. The transient expression of Tal2 was confirmed in mouse embryogenesis, as well as Tal2 transiently expressed in P19 cells by addition of atRA in suspension culture.

Localization of Tal2 in P19 cells. To examine the behavior of Tal2 in P19 cells, we generated GFP-tagged Tal2 (GFP-TAL2) expression construct. This construct was introduced into P19 cells via transfection, and these cells were stained with Hoechst33258 48 h post transfection. We then used fluorescence microscopy to visualize these cells (Fig. 2). GFP-TAL2 expressed in P19 cells was detected in nuclei.

atRA induced the expression of Tal2 gene in P19 cells. atRA and cell aggregation are important for neural differentiation of P19 cells12–14. Thus, we examined how these treatments were involved in the expression of Tal2 gene. When P19 cells were treated with atRA and cultured as a monolayer in culture dishes, Tal2 expression was nearly the same as that in atRA-treated and cell-aggregated P19 cells (Fig. 3a). Additionally, the Tal2 gene was not induced by DMSO.
treatment in suspension culture of P19 cells (see Supplementary Fig. S1 online). These results indicate that the induction of Tal2 in P19 cells is associated with atRA.

atRA binds to the RARs (RARα, β, or γ), and RARs form heterodimers with RXRs (RXRα, β, or γ). atRA-bound RAR/RXR heterodimer can regulate the transcription of target genes. To elucidate the involvement of these receptors in Tal2 induction, we confirmed the expression of Rar genes and Rxr genes in P19 cells after addition of atRA in suspension culture (Fig. 3b). For these six genes, the expression of Rara, Rxra or Rxrb was not altered at 48 h. The expression of Rarg and Rxrg had decreased relative to basal levels. On the other hand, Rarb expression was not evident at 0 h, but it was evident within 3 h after treatment, as it has been reported that Rarb was induced by retinoic acid in P19 cells29,30. The induction of Tal2 and Rarb were similar in P19 cells, but Rarb expression was not altered from 3 h to 48 h while the expression of Tal2 gene had decreased from peak levels by 24 h.

Next, in order to investigate the relation between atRA and its receptors, we used four agonists instead of atRA; Am8031 (RARα agonist), adapalene32 (RARβ and RARγ agonist), AC-41848 (RARγ agonist) and methoprene acid33 (RXR pan-agonist). Am80 was able to induce Tal2 gene as well as atRA. Adapalene also induced Tal2 in P19 cells, but the induction of Tal2 by adapalene was weaker than that by atRA or Am80. In contrast, neither AC-41848 nor methoprene acid induced Tal2 expression (Fig. 3c). Moreover, we also assessed morphological changes of P19 cells that were treated with atRA, DMSO, or each agonist (Fig. 3d). P19 cells formed spherical aggregates called embryoid bodies in the presence of atRA, Am80 or adapalene. Whereas, DMSO, AC-41848 or methoprene acid did not formed embryoid bodies.

**Tal2 expression was induced by RARα.** Adapalene formed P19 cells into embryoid bodies, and weakly induced the expression of Tal2 compared with atRA or Am80. Moreover, it has been reported that
adapalene has stimulated RAR\(\alpha\) weakly\(^{32}\). Although the expression of Rarb gene was not observed at 0 h in P19 cells, RAR\(\alpha\) protein was expressed at 0 h. Based on these findings, we assumed that RAR\(\alpha\) had an intimate involvement in Tal2 induction in P19 cells.

We used RNA interference to suppress RAR\(\alpha\) expression in P19 cells, and examined the involvement of RAR\(\alpha\) and the induction of Tal2. The RNAi-mediated suppression of RAR\(\alpha\) in P19 cells was verified by Western blotting (Fig. 4a). Tal2 expression significantly decreased by RAR\(\alpha\) knockdown in P19 cells after addition of atRA in suspension culture (Fig. 4b). These results indicate that atRA–RAR\(\alpha\) complexes induce Tal2 gene in P19 cells.

**Discussion**

P19 EC cells are induced into neural differentiation by addition of atRA in suspension culture\(^{13,14}\). We explore the molecular mechanisms of the entry into neural differentiation using P19 cells. We found that Tal2 expression was altered rapidly and transiently in these cells (Fig. 1a and b). Interestingly, the transient expression of Tal2 gene was observed at the stage of organogenesis during embryonic development (Fig. 1c). Tal2 expression in embryonic head was reported previously\(^{10,11}\), and Tal2 is inferred to play a role at the early stage of brain development. Because P19 cells have been used as model cells for the studies of neural differentiation, a detailed investigation of the molecular mechanisms regulating Tal2 expression in P19 cells may be key to understanding the function of Tal2 in brain development.

We investigated the correlation of atRA, cell aggregation and Tal2 induction in P19 cells. As a result, atRA participated in Tal2 induction in P19 cells, but cell aggregation was not necessary for Tal2 induction (Fig. 3a). Next, we examined the involvement of RAR/RXR in the induction of Tal2 in P19 cells (Fig. 3c). Am80, a RAR\(\alpha\) agonist, was able to induce Tal2 expression in P19 cells. Adapalene also weakly induced Tal2 in P19 cells. Although adapalene is an agonist of RAR\(\beta\) and RAR\(\gamma\), it might also react to RAR\(\alpha\) in P19 cells. Indeed, RAR\(\alpha\) is less responsive to adapalene than RAR\(\beta\) and RAR\(\gamma\), and RAR\(\beta\) is no responsive to adapalene\(^{32}\). Rarb gene was not expressed in P19 cells at 0 h, and was induced by atRA as well as Tal2 gene. Therefore, adapalene is thought to stimulate the RAR\(\alpha\) signaling weakly and to induce the expression of Tal2 in P19 cells. Moreover, RAR\(\alpha\) knock down by RNA interference suppressed Tal2 expression in P19 cells (Fig. 4b). These results indicate that atRA–RAR\(\alpha\) complexes induce transcription of Tal2 gene in P19 cells.

The embryoid bodies were formed by the agonists, atRA, Am80 and adapalene, that induced the expression of Tal2 in P19 cells (Fig. 3d). These results raise the possibility that Tal2 is also involved in the formation of embryoid bodies of P19 cells. TAL2 induced by these agonists might activate genes such as Fox8\(^{34}\) as a transcription factor that are involved in cell aggregation.

The bHLH family to which TAL2 belongs regulates a variety of biological processes. The bHLH usually has two functionally distinct domains—the basic domain and the HLH domain. The bHLH proteins form dimers via HLH domains and bind DNA via the basic domains. These dimers activate target genes in the nucleus\(^2\). Therefore, Tal2, which is a bHLH protein, is probably involved in transcriptional activation of target genes in the nucleus. Indeed, the GFP-TAL2 fusion protein that we generated localized to nuclei (Fig. 2). Moreover, GFP-TAL2 was highly enriched in a part of each nucleus. In zebrafish, TAL2 is expressed in the lateral floor plate of the spinal cord\(^{35}\). The floor plate is an important signal center that specifies neurons and glia cells in the ventral neural tube and guides the trajectory of outgrowing axons. Tal2 acts upstream of the gad67 gene that is the GABA-synthesizing enzyme glutamic acid decarboxylase in Kolmer–Agduhr (KA\(^{+}\)) cells of the lateral floor plate\(^{36}\). However, it is not clear whether the mammalian spinal cord has KA\(^{+}\) cells, and Tal2 has not yet been observed in the mouse spinal cord.

atRA is important for embryogenesis and the differentiation of the nervous system, and we demonstrated for the first time that Tal2 expression was induced by atRA, which introduced P19 cells into neural differentiation. In addition, the transient expression of Tal2 was observed in organogenesis of development. These findings suggest that Tal2 plays a role in acquisition of neural cell fate as a transcription factor in neural differentiation. Because the neural differentiation of P19 cells in part mimics the development of the nervous system, P19 cells are useful for studying the molecular mechanism of Tal2 in neural differentiation. Further work is underway to clarify the function of Tal2 in neural differentiation using the differentiation system of P19 cells.

**Methods**

Cell culture and neural differentiation. P19C6, a subclone of P19 mouse embryonic carcinoma cell line, was used in this study\(^{35}\). P19C6 was provided by the RIKEN BRC through the National Bio-Resource Project of the MEXT, Japan. P19 cells were cultured in α-MEM (Sigma-Aldrich, MO) supplemented with 10% FBS (Life Technologies, CA) and 2 mM L-glutamine (Kanto Chemical, Japan). To induce neural differentiation, these cells were aggregated in a suspension culture dish (SUMILON, Japan) at a seeding density of 2 × 10\(^4\) cell/ml in the presence of 1 μM atRA (Sigma-Aldrich), which was dissolved in DMSO (Sigma-Aldrich)\(^{36}\). DMSO concentration in culture condition was 0.01%. To identify the RAR and RXR subtypes, 100 nM Am80 (RAR\(\alpha\) agonist), 100 nM adapalene (RAR\(\beta\) and RAR\(\gamma\) agonist), 100 nM AC-41848 (RAR\(\gamma\) agonist), or 100 nM methoprene acid (RXR pan-agonist) were used in place of atRA\(^{37}\). These agonists were purchased from Sigma-Aldrich.

RT-PCR and real-time PCR. RNeasy Mini Kit (QIAGEN, Germany) was used according to the manufacturer’s instructions to isolate total RNA from P19 cells. RT-PCR was performed as described previously\(^{36}\). In brief, reverse transcription was performed with 1 μg of total RNA and SuperScript III reverse transcriptase (Life Technologies), and gene-specific primer were used for amplification of target genes. The list of primers can be found as Supplementary Table S1 online. The Gapdh gene was used as an internal standard. To assess Tal2 expression in normal mouse embryogenesis, MTC Multiple Tissue cDNA Panels (TAKARA BIO, Japan) were used as templates.
Microscopy. GFP-fused Tal2 subcloned into pcAGGS vector44. Lipofectamine 2000 (Life Technologies) was used according to the manufacturer's instructions to transfect with the constructed vector into P19 cells. After the transfection, cells were washed with phosphate-buffered saline (PBS), and fixed via incubation with 1% glutaraldehyde (nacalai tesque, Japan) in PBS for 30 min. After washed with PBS, the cells were stained with Hoechst33258 for 30 min. After washed with PBS, the cells were visualized using BX-8000 (KEYENCE, Japan). Morphological changes of P19 cells were assessed at 48 h after atRA, DMSO, or four agonists treatment. A relief contrast microscope (OLYMPUS, Japan) was used to examine these cells.

RNA interference. We used pSilencer 3.1-H1 (Life Technologies) as an shRNA vector. The sequences of the shRNA duplexes used for RARα knockdown were as follows: RARα gene, sense 5'-GCAAGTACTACTGAACAACTCAAGGAATGTTGTCATCTTGATCITTTTTGGAA-3' and antisense 5'-TTCCAAAAGCAAGTACTACTGAACAACTCAAGGAATGTTGTCATCTTGATCITTTTTGGAA-3' and antisense 5'-TTCAAAAAGCAAGTACTACTGAACAACTCAAGGAATGTTGTCATCTTGATCITTTTTGGAA-3'.

Tohoku, Nakane, & Chiwa, S. Gene expression of MASH-1, MATH-1, neuroD and NSCL-2, basic helix-loop-helix proteins, during neural differentiation in P19 embryonal carcinoma cells. Tohoku. J. Exp. Med. 182, 327–336 (1996).

Pratt, M. A., Kralova, J. & McBurney, M. W. A dominant negative mutation of the alpha retinoic acid receptor gene in a retinoic acid-nons-responsive embryonal carcinoma cell. Mol. Cell. Biol. 10, 6445–6453 (1990).

Kruyt, F. A. et al. Transcriptional regulation of retinoic acid receptor beta in retinoic acid-sensitive and -resistant P19 embryocarcinoma cells. Mech. Dev. 33, 171–178 (1991).

Zheng, B. et al. Kruppel-like factor 4 inhibits proliferation by platelet-derived growth factor receptor beta-mediated, not by retinoic acid receptor alpha-mediated, phosphatidylinositol 3-kinase and ERK signaling in vascular smooth muscle cells. J. Biol. Chem. 284, 22773–22785 (2009).

Shroot, B. & Michel, S. Pharmacology and chemistry of adapalene. J. Am. Acad. Dermatol. 56, 59–103 (2007).

Harmon, M. A., Boehm, M. F., Heyman, R. A. & Mangelsdorf, D. J. Activation of mammalian retinoid X receptors by the insect growth regulator metaporphene. Proc. Natl. Acad. Sci. U. S. A. 92, 6157–6160 (1995).

Wang, C. et al. Cell aggregation-induced FGFlf8 elevation is essential for P19 cell neural differentiation. Mol. Biol. Cell. 17, 3075–3084 (2006).

Pinheiro, P., Gering, M. & Patient, R. The basic helix-loop-helix transcription factor, Tal2, marks the lateral floor plate of the spinal cord in zebrafish. Gene. Expr. Patterns. 4, 85–92 (2004).

Yang, L., Rastegar, S. & Strahle, U. Regulatory interactions specifying Kolmer–Busby–Weiskopf neurons: the P19 pathway. J. Comp. Neurol. 495, 22773–22785 (2010).

Gao, X. et al. A role of N-cadherin in neuronal differentiation of embryonic carcinoma P19 cells. Biochem. Biophys. Res. Commun. 284, 1098–1103 (2001).

Niwa, H., Yamamura, K. & Miyazaki, J. Efficient selection for high-expression transfectants with a novel eukaryotic vector. Gene. 108, 193–199 (1991).

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Author contributions

T.K., R.K. and H.M. conceived the project. T.K., R.K. and H.M. designed the experiments. T.K., R.K. and K.I. performed the experiments. T.K., S.T. and H.M. wrote the manuscript with discussion of all other authors. All authors reviewed the manuscript.

Additional information

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