Allium-sativum and bakuchiol combination: A natural alternative to Chlorhexidine for oral infections?

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ABSTRACT

Objective: Chlorhexidine mouthrinses are considered a gold standard as an adjunct treatment of oral infections. However, owing to its toxicity, discoloration of tooth surface and the emerging prevalence of drug-resistant species, attention is being given to exploring natural alternatives to the drug.

Methods: The experiment was carried out in Azra Naheed Center for Research and Development (ANCRD), Superior University, Lahore, Pakistan from September 2018 till May 2019. Biofilms and planktonic cells of C. albicans alone and in combination with streptococci were subjected to chlorhexidine, allium sativum and bakuchiol individually and to allium-bakuchiol combination. Kirby-Bauer test, antifungal susceptibility testing, CFU count and drug synergy assessment was done on planktonic cells. Dynamic biofilms were formed to mimic conditions similar to oral cavity and CFU was determined.

Results: MIC of all three agents was higher against mixed species when compared to single species planktonic cells and biofilm. Allium sativum and bakuchiol demonstrated synergistic effects. The decrease in CFU count and minimum biofilm reduction to salivary pellicle caused by allium sativum-bakuchiol was comparable to that of chlorhexidine.

Conclusion: Thus, allium sativum-bakuchiol combination demonstrated antimicrobial effects similar to chlorhexidine against planktonic cells and dynamic biofilm. It could serve as a possible natural, economical alternative to chlorhexidine mouthrinses usually recommended in dental clinics. However, in vivo studies are required to determine the correct dosage of these agents.

KEYWORDS: Allium sativum, Bakuchiol, Candida, Chlorhexidine, Streptococci.

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INTRODUCTION

Chlorhexidine mouth rinses (0.2% w/v) are considered a gold standard as an adjunct treatment of oral bacterial and fungal infections in dental clinics.¹ However, the therapeutic applications of chlorhexidine become limited owing to its toxicity, discoloration of tooth surface and the emerging prevalence of drug-resistant candidal species.¹ Antimicrobial resistance is a consequence of imprudent use of antimicrobial agents and develops when a pathogenic organism mutates or obtains a resistance gene.² Candida albicans infections are becoming an escalating challenge for doctors as they are opportunistic microorganisms colonizing the human oral cavity.² Streptococcus
mitis and Streptococcus sanguinis, commensals of oral cavity have been co-isolated with C. albicans from several infections and from various biomaterial surfaces including implants, dentures, voice prostheses, feeding tubes and catheters. This type of polymicrobial growth may not only alter the inherent virulence of the species, but the treatments normally effective against monomicrobial species infections may also be rendered futile. When such infections are not cured by traditional therapy, doctors shift to steroids and harsher measures.

Natural plant derived products can be used because of antimicrobial properties and minimal side effects on human health. Bakuchiol, derived from leaves of Psoralea glansdulosa (Culen), is commonly used in folk medicine for the treatment of skin diseases caused by bacteria and fungi. Allium sativum (garlic) belonging to Liliaceae family, also has antibacterial activity against many common pathogenic bacteria. Preparing drugs in combination is common in modern medicine to enhance the pharmaceutic efficacy and decrease the side effects from the excessive use of drugs.

The aim of present research is to compare the effects of 0.2% w/v chlorhexidine, A. sativum extract and bakuchiol extract on planktonic and biofilm form of Candida albicans. The latter will be tested both alone and in a mixed species biofilm including Streptococcus mitis and Streptococcus sanguinis, which are early colonizers of oral cavity.

METHODS

Inoculum and Media: The experiment was carried out in Azra Naheed Center for Research and Development, Superior University, Lahore from September 2018 till May 2019 with the approval of the Institutional Research Ethics Board (Ref: 40550/AN/SU, dated March 7, 2017). Written informed consent was obtained from the individuals who participated in this study. C. albicans (ATCC 14053), S. sanguinis (BAA 1455) and S. mitis (ATCC 49456) strains were used for the formation of in vitro biofilms. A general purpose medium tryptic Soy Broth (Merck) was used as nutrient media.

The Kirby-Bauer Susceptibility Test: The antimicrobial activity of chlorhexidine (CHX) (Merck), Bakuchiol (Bk) (Merck) and Allium sativum (As) (Merck) (the last two used both alone and in combination) was analyzed by disc-diffusion susceptibility test proposed by Kirby-Bauer. Single and mixed species suspension was swabbed uniformly over a Mueller Hinton agar (MHA) plate. Sterile 6mm paper discs (Whatman®, USA) saturated with 100µL of CHX (0.2% w/v), As (50µg/mL) and Bk (50µg/mL) each were placed on the inoculated agar surface. For As + Bk, 50 µL of each was used. A blank paper disc inoculated with sterile distilled water was used as negative control. All plates were incubated at 37°C for 24 h. The inhibition zone created around the disc was measured to estimate the antimicrobial susceptibility for each group. The experiment was repeated in triplicate with biological and technical variants (n = 9) and mean value was recorded.

Antifungal susceptibility test of planktonic cells: Based on the results obtained from Kirby-Bauer test the minimum inhibitory concentration (MICs) of CHX, As and Bk for single and mixed species were determined using the Clinical and Laboratory Standards Institute broth microdilution method (CLSI-BMD) described in NCCLS guidelines. A 100 µL of single species (candida alone) and mixed (candida with both streptococci) (ratio of 1:1:1 at similar respective concentrations of 1 x 10⁶ cells / mL) species inoculum prepared in TSB was poured in 96-well plates, followed by 100 µL of different concentrations of CHX, As and Bk (125, 100, 64, 50, 32, 16, 8, 4, 2 µg / mL respectively. Tryptic soy broth, free of antimicrobial agent or active compounds was used as positive control and TSB without addition of microorganism was used as negative control. After 24hours’ incubation at 37°C, organism growth was assessed by optical density OD₅₇₀nm using a spectrophotometer (FC-Bios µQuant).

The MIC was determined as the minimum concentration of antimicrobial agent that inhibited ≥ 50% of microbial growth in comparison to positive control using a modified Gompertz model. The experiment was performed three times to ensure reliable and reproducible results.

Determination of drug synergy against planktonic cells: Combination of As with Bk was assessed for synergistic affect against planktonic single and mixed species using the checkerboard microdilution method. Different concentrations of antifungal combination (125, 100, 64, 50, 32, 16, 8, 4, 2 µg / mL) at 100 µL volume were poured in a sterile microtiter plate. Following which, 100µL of single and mixed species Candida suspension of 10⁶ cells/mL was added. The plates were then incubated at 37°C overnight and assessed in spectrophotometer). MIC and Fractional inhibitory concentration (FIC) was determined using the MIC values of As + Bk. The sum of individual FICs; FIC index was determined using the following formula:
FIC\textsubscript{i} = \frac{\text{MIC(As\textsubscript{comb})}}{\text{MIC(As\textsubscript{alone})}} + \frac{\text{MIC(Bk\textsubscript{comb})}}{\text{MIC(Bk\textsubscript{alone})}}.

FIC\textsubscript{i} ≤ 0.5 = synergy; FIC\textsubscript{i} > 0.5 - ≤ 1 = partial synergy; FIC\textsubscript{i} > 1 - ≤ 4 = indifference; and > 4, antagonism.

**Production of dynamic biofilm:** Biofilm was formed using Nordini’s artificial mouth model (NAM) defined by Rahim et al. (2008)\textsuperscript{10} by using glass beads to mimic tooth surface. Continuous salivary flow and 37°C temperature was maintained for pellicle formation. After which microorganisms were allowed to grow and form biofilm for 24 h. The biofilm was treated with MIC concentration of CHX, As, Bk and As + Bk combination respectively and CFU was calculated using formula:

\[
\text{Total CFU/mL} = \frac{\text{number of formed colonies}}{\text{dilution factor} \times \text{volume used (mL)}}
\]

Mean of CFU/mL were calculated in triplicates. The percentage reduction of microbial population in adhesion to salivary pellicle after treatment was calculated as follows:

\[
\text{Percentage decrease in adherence} = \frac{W \times 100}{Y}
\]

Where “W” is the mean microbial population treated with As + Bk combination and “Y” is the mean microbial population subjected to distilled water (negative control).

**Statistical analysis:** All results were calculated and expressed as mean values with standard deviation (±SD) from three technical and biological variants that were performed in triplicate (n = 9). SPSS software (version 18.0) was used to perform statistical analyses. Independent \(t\)-test was used to distinguish data between single and mixed species group. ANOVA was used to compare differences between groups treated with various drugs. \(P\) value of less than 0.05 was deliberated as statistically significant.

**RESULTS**

**The Kirby-Bauer susceptibility test:** All agents created inhibition zones on MH agar. CHX created larger inhibition zone in comparison to As and Bk alone. However, the combination of As and Bk exhibited the largest inhibitory zone in single and mixed species (Table-I) indicating high antimicrobial activity.

**Antimicrobial susceptibility of Planktonic cells:** MIC values of CHX, As and Bk was substantially high for mixed species making mixed species susceptible to chlorhexidine only at high dosage. The combination of As and Bk showed synergistic effect against single and mixed species group and was affective against microbes at low dosage (\(\sum\ FIC\ index = 0.5\)) (Table-II).

**Table-I:** The mean diameter of inhibition zone produced by antimicrobials on single and mixed species.

| Groups                  | Antimicrobials used alone | Combination of agents |
|-------------------------|---------------------------|-----------------------|
|                         | As (50µg/ml)              | Bk (50µg/ml)         | CHX (0.2% w/v) | As + Bk (50µg/ml+50µg/mL) | Sterile dH\textsubscript{2}O |
| Single species (C. albicans alone) | 19 | 20 | 22 | 25 | NS |
| Mixed species (Candida and bacteria) | 16 | 18 | 18 | 22 | NS |

NS: no sensitivity.

**Table-II:** Mean antimicrobial susceptibility values of single (Candida albicans) and mixed (Candida-bacteria) species to chlorhexidine, allium sativum, bakuchiol and A. sativum-bakuchiol combination.

| Species               | Chlorhexidine (MIC) (µg/mL) | Allium sativum (MIC) (µg/mL) | Bakuchiol (MIC) (µg/mL) | As+Bk (MIC) (µg/mL) | FIC index |
|-----------------------|-----------------------------|------------------------------|-------------------------|---------------------|-----------|
| C. albicans alone     | 4                           | 8                            | 25                      | 4\textsubscript{a} + 12.5\textsubscript{b} | 0.5= SYN |
| Mixed species         | 32                          | 16                           | 50                      | 8\textsubscript{a} + 12.5\textsubscript{b} | 0.5= SYN |

**Note:** The MIC\textsubscript{50} was determined as the minimum concentration of antimicrobial agent that inhibited ≥ 50% of microbial growth in comparison to drug free control. Fractional inhibitory concentration (FIC) was calculated to determine the synergistic interaction between A. sativum and bakuchiol, *SYN: Synergistic.
Effect of Allium sativum + Bakuchiol on Biofilm:

There was higher microbial count in mixed species in comparison to single species. CHX was extremely effective against single species. However, for mixed species, both As+Bk and CHX were equally effective against microorganisms (p>0.05) (Fig. 1).

**DISCUSSION**

*C. albicans* is known to form synergistic interactions with a number of bacteria, collectively causing diseases like oral candidiasis, denture stomatitis, periodontitis which are resistant to conventional antifungal treatment. The increased resistance to commonly prescribed antifungals motivate search for new ways of combating polymicrobial biofilms.

According to previous studies, CSH of planktonic microbial cells positively correlate with formation of biofilm. Difference in cell surface physicochemical properties and presence of carbohydrate moiety may impact microbial cell coherence to hydrophobic interface. The metabolites and crude extracts derived from herbs and plants are important in the search for new antifungal agents. *Bakuchiol* has been widely used for skincare regimens, as anti-wrinkle and anti-aging agent and has previously been linked with antimicrobial properties. It has been reported that garlic extract (*Allium sativum*) can inhibit growth of both Gram-positive and Gram-negative bacteria. The garlic cloves consist of sulfur containing chemicals like allicin, alliin, and ajoene. When the garlic cloves are cut or crushed they release the enzyme alliinase which converts alliin to allicin which is responsible for antibacterial activity. The antibacterial property of garlic extract was evaluated in previous studies and can inhibit the bacterial growth when used at higher concentration. Similarly, *Bakuchiol* has been studied for its action against *Candida albicans* and non albicans species. Katsura et al., 2001 has studied the anti-adherence effect of *bakuchiol* on various oral bacteria. The mechanism of action of this compound is yet to be studied in detail. But neither of these agents have been studied against fungal-bacterial mixed species. In the current research, exposure of Candida and mixed species planktonic cells to As+Bk revealed significant decrease in the CSH percentage in comparison to untreated samples. The current research was carried out to assess the antimicrobial activity of *Allium sativum* and *Bakuchiol* combination against candida-bacteria mixed species planktonic and biofilm cells. The results were compared with *Allium sativum*, *Bakuchiol* and chlorhexidine used alone.

It was also noted that CHX required a greater MIC against mixed species in comparison to single species. This indicates a stronger interaction between cells when Candida and bacteria are grown together. This interkingdom bond requires larger dose to overcome microbial growth. The FIC index of As+Bk showed synergy in planktonic cells. The effect of agents was further analyzed by assessing their action against single and mixed species biofilms. Since microorganisms form salivary biofilms in oral cavity and the complexity that is orchestrated in those biofilms govern their virulence and pathogenicity in the oral soft and hard tissue. The results indicate that As+Bk can be as effective as chlorhexidine against mixed species biofilms.

This study revealed that *Allium sativum* and *Bakuchiol* exposure had a considerable effect towards polymicrobial interactions of Candida-bacterial cells. *Allium sativum* and *Bakuchiol* could have altered cell surface area and caused impairment of cell hydrophobicity which might have led to reduced adhesion to hydrophobic surface. Clinical trials are needed for confirmation of the in-vitro beneficial antimicrobial effect and any needed dosage adjustment, these herbal and locally available agents could be effectively incorporated in the formulation of mouthwashes, gels or / and lozanges for the treatment of oral candidal and adjunctive infections.
CONCLUSION

Within the limitations of this study, the following could be concluded: *Allium sativum* and *Bakuchiol* individually, had antifungal and antimicrobial properties against *C. albicans* alone and candida-bacteria mixed species. The combined effect of *A. sativum* and *bakuchiol* extracts in terms of antimicrobial effects was comparable to that of the chlorhexidine, thus indicating their possible future employment as an alternative to chlorhexidine itself.

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REFERENCES

1. Karpinski TM, Szkaradkiewicz AK. Chlorhexidine−pharmaco-biological activity and application. Eur Rev Med Pharmacol Sci. 2015;19(7):1321-1326.
2. Pfaller MA, Diekema DJ. Epidemiology of invasive candidiasis: a persistent public health problem. Clin Microbiol Rev. 2007;20(1):133-163. doi: 10.1128/CMR.00029-06
3. Díaz PI, Xie Z, Sobue T, Thompson A, Biyikoglu B, Ricker A, et al. Synergistic interaction between Candida albicans and commensal oral streptococci in a novel in vitro mucosal model. Infect Immun. 2012;80(2):620-632. doi: 10.1128/IAI.05896-11
4. Willems HM, Xu Z, Peters BM. Polymicrobial Biofilm Studies: From Basic Science to Biofilm Control. Curr Oral Microbiol Rep. 2016;3(1):36-44. doi: 10.1007/s40496-016-0078-y
5. Nordin MA, Abdul Razak F, Himratul-Aznita WH. Assessment of Antifungal Activity of Bakuchiol on Oral-Associated Candida spp. Evid-Based Compl Alt Med. 2015;2015:918624. doi: 10.1155/2015/918624
6. Mozaffari Nejad AS, Shabani S, Bayat M, Hosseini SE. Antibacterial Effect of Garlic Aqueous Extract on Staphylococcus aureus in Hamburger. Jundishapur J Microbiol. 2014;7(11):e13134. doi: 10.5812/jjm.13134
7. Bauer AW, Kirby WM, Sherris JC, Turck M. Antibiotic susceptibility testing by a standardized single disk method. Am J Clin Pathol. 1966;45(4):493-496. doi: 10.1093/ajcp/45.4.493
8. Thornberry C. NCCLS standards for antimicrobial susceptibility tests. Lab Med. 2016;14(9):549-553. doi: 10.1093/labmed/14.9.549
9. Rand KH, Houck HJ, Brown P, Bennett D. Reproducibility of the microdilution checkerboard method for antibiotic synergy. Antimicrob Agents Chemother. 1993;37(3):613-615. doi: 10.1128/aac.37.3.613
10. Rahim ZH, Fathilah AR, Irwan S, Wan Nordin Hasnor WI. An artificial mouth system (NAM model) for oral biofilm research. Res J Microbiol. 2008;3(6):466-473. doi: 10.3923/jm.2008.466.473
11. Desai JV. Candida albicans Hyphae: From Growth Initiation to Invasion. J Fungi 2018;4(1):10. doi: 10.3390/jof4010010
12. Berzhi E, Sciola R, Biazzoni C, Cirasola D, Cappelletti L, Vizzi L, et al. Cell surface hydrophobicity: a predictor of biofilm production in Candida isolates? J Med Microbiol 2011;60(Pt 5):689-690. doi: 10.1099/jmm.0.026898-0
13. Chaudhuri RK, Bojanowski K. Bakuchiol: a retinol-like functional compound revealed by gene expression profiling and clinically proven to have anti-aging effects. Int J Cosmet Sci 2014;36(3):221-230. doi: 10.1111/ics.12117
14. Katsura H, Tsukiyama RI, Suzuki A, Kobayashi M. In vitro antimicrobial activities of bakuchiol against oral microorganisms. Antimicrob Agents Chemother. 2001;45(11):3909-3913. doi: 10.1128/AAC.45.11.3909-3913.2001
15. Martins N, Petropoulos S, Ferreira IC. Chemical composition and bioactive compounds of garlic (*Allium sativum* L.) as affected by pre- and post-harvest conditions: A review. Food Chem 2016;211:41-50. doi: 10.1016/j.foodchem.2016.05.029
16. Fani MM, Kohanteb J, Dayagh M. Inhibitory activity of garlic (*Allium sativum*) extract on multidrug-resistant Streptococcus mutans. J Indian Soc Pedod Prev Dent. 2007;25(4):164-168. doi: 10.4103/0970-4398.37011
17. Thomas A, Thakur S, Habib R. Comparison of Antimicrobial Efficacy of Green Tea, Garlic with Lime, and Sodium Fluoride Mouth Rinses against Streptococcus mutans, Lactobacilli species, and Candida albicans in Children: A Randomized Double-blind Controlled Clinical Trial. Int J Clin Pediatr Dent. 2017;10(3):234-239. doi: 10.5005/jp-journals-10005-1442
18. Varposhti M, Entezari F, Feizabadi MM. Synergistic interactions in mixed-species biofilms of pathogenic bacteria from the respiratory tract. Rev Soc Bras Med Trop. 2014;47(5):649-652. doi: 10.1590/0037-8682-0262-2013
19. BinAhmed S, Hasane A, Wang Z, Mansurov A, Romero-Vargas Castrillon S. Bacterial Adhesion to Ultrafiltration Membranes: Role of Hydrophilicity, Natural Organic Matter, and Cell-Surface Macromolecules. Environ Sci Technol. 2018;52(1):162-172. doi: 10.1021/acs.est.7b03682

Authors’ Contribution:

AF did data collection, manuscript writing & statistical analysis.

WHHA was responsible for data accuracy and integrity of the work.

PSAR edited and approved the manuscript.