Phytochemical Screening of Medicinal Plants of the Kakamega Country, Kenya Commonly Used Against Cancer

Dominic O Ochwang’I*, Charles N Kimwele1, Jemimah A Oduma1, Peter K Gathumbi2, Stephen G Kiama1 and Thomas Effertth**

1Department of Veterinary Anatomy and Physiology, University of Nairobi, Nairobi, Kenya
2Department of Pharmaceutical Biology, Institute of Pharmacy and Biochemistry, Johannes Gutenberg University, Staudinger Weg 5, 55128 Mainz, Germany

Abstract

Phytochemical constituents of 35 selected anticancer medicinal plant extracts were analyzed by thin layer chromatography (TLC) for alkaloids, anthraquinones, xanthines, valepotriates, cardiac glycosides, flavonoids, essential oils, coumarins, lignans, saponins and arbutin compounds. These plants sourced from Kakamega tropical rain forest have been previously reported to be used for cancer treatment in the Kakamega County, Kenya. The medicinal plants investigated were Harungana madagascariensis Lam. ex poir, Fuerstia africana T.C.E. Fr., Sida rhomboloba L., Zanthoxylum rubescens Hook. f, Bridelia micrantha (Hochst.) Baill, Juniperus procera Endl., Tragia brevipes Pax, Phyllanthus sapialis, Conyza sumatrensis, Momordica foetida Schumach., Synsepalum cerasiferum Synonym: Afroseraisia seraisera (Welw.) Aubrev, Aloe volkensii Engl, Aeschynomene abyssinica (A. Rich.) Vatke, Futumia africana Benth. Cyphostemon serplus (A. Rich), Ipomoea carica (L.), Spathodea campanulata P. Beauv. ssp. nilotica (Seem), Abrus precatorius L. ssp africanus Verdc., Triumfetta rhomboidea Jacq., Psyrax schimperiana (A. Rich), Ficus thornningii, Rothea myrocytoides (Hochst. Steane and Mabb), Croton macrostachyus Delile, Vernonia lasiopus O Hoffin, Albizia gummi-fera (J.F. Gmel.), Zanthoxylum gilletii (De Wild) P.G. Waterman, Microglossa pyriformis (Lam.) Kuntze, Senna didymobotrya (Fresem.) Irwin and Barneby, Trichilia emetica Vahl, Entada abyssinica Steud.ex A.Rich., Shirakaiosis elliptica (Hochst.) Esser Synonym: Sapium ellipticum (Hochst.kraus) Pax, Ocimum gratissimum L. Suave wild O. tomentosum oliv., Prunus africana (Hook.f.) Kalkman, Phyllanthus fischeri Pax and Olea hotch sp. Hochstetter. The phytochemical distribution in the 35 plants included: 71.4% alkaloids, 57.1% anthraquinones, 94.2% xanthines, 82.8% valepotriates, 94.2% cardioactive glycosides, 82.8% flavonoids, 77.1% essential oils, 85.7% coumarin drugs, 68.5% lignans, 80% saponins and 62.85% arbutin drugs. Our findings provided evidence that crude organic and inorganic solvent extracts of these tested plants contain medicinally important bioactive compounds and provide a rational basis for their use in traditional medicine.

Keywords: Phytochemicals; Alkaloids; Flavonoids; Saponins

Introduction

A majority of important bioactive compounds that produce desirable physiologic activity have been derived from plants. These compounds include but are not limited to phenolic compounds, alkaloids, tannins and flavonoids [1]. These compounds are diverse secondary metabolites that can be utilized for drug development in human and veterinary medicine [2]. Plant products derived from barks, leaves, flowers, roots, fruits or seeds have been part of phytotherapies from the ancient days [3]. Lead compounds with great pharmacological features have been synthesized from these plants [4]. In the present investigation, the phytochemical constituents of 35 selected herbal extracts were analyzed for alkaloids, anthraquinones, xanthines, valepotriates, cardiac glycosides, flavonoids, essential oils, coumarins, lignans, saponins and arbutin drugs by thin layer chromatography. These medicinal plants have been previously reported to have anticancer properties [5].

Materials and Methods

Extraction preparation

Organic and aqueous extracts were prepared using dichloromethane, methanol and water. Ten grams of powdered plant material was extracted by cold maceration in a dichloromethane-methanol mixture at a ratio of 1:1 for 72 h at room temperature. The extract was reduced in a rotavapor and thereafter in an oven at 40°C. The yield of each extract was determined and the extract preserved at 4°C. For aqueous extraction, the plant powder was macerated in distilled water at a ratio of 1:6 (w/v). The suspension was rotated on a shaker for 24 h at room temperature and filtered using cotton wool and Whatman filter paper No 1. The aqueous extracts were freeze dried and the resultant lyophilized residues stored at room temperature.

Thin layer chromatography (TLC)

The phytochemical analysis of extracts of commonly used medicinal plants was carried out using TLC as described by Wagner. Extract aliquots of 2.5 mg were dissolved in 500 µL of methanol. Silica gel 60F 254-aluminium foils (0.2 mm layer thickness) (Millipore Corporation, Germany) were used. The TLC plates were fan-dried shortly before applying the substances with starting points in a height of 1-1.5 cm. The distance between the starting points was 0.5 cm and 1 cm to the plate boundary. It proved reasonable to apply the substances not only as points, but also as bar marks. The glass capillaries used to apply the solution corresponded to 1.2 cm liquid column of 5 µL. The...
plate was well dried after every application. The developing chamber with super plasticizer was lined with filter paper (the super plasticizer moves faster). The development of the TLC plates took place, if the chamber was saturated. The super plasticizer was completely flashed off, if the plate was developed by fan-drying or putting them in a drying cabinet before detection and taking chromatogram pictures.

**Spray solutions**

Spray solutions were prepared and used for the various analyses. Glass spray heads were used, if the solution contained sulphuric acid. These solutions were as follows:

- **Anisic aldehyde-sulfuric acid-reagent:** Anisic aldehyde (0.5 mL) was slowly mixed with 10 mL acetic acid, 85 mL MeOH and 5 mL conc. sulphuric acid in that order. The solution was kept in the refrigerator.
- **Dragendorff reagent:** Basic bismuth nitrate (1.7 g) and 20 g tartaric acid were suspended in 40 mL water. Potassium iodide (16 g) was dissolved in 40 mL water and added to the bismuth nitrate suspension. The resultant liquid was stirred for 1 h and filtrated. The stock solution was kept several days in brown glass bottles in the refrigerator and protected from light. The spray solution was prepared by mixing 5 mL stock solution with 15 mL water.
- **Ethanol KOH reagent:** A 5% solution of KOH in EtOH 50% (V/V) was prepared, which comprised of 2.5 g KOH dissolved in 25 mL water and 25 mL ethanol.
- **Iodine-platinate reagent:** Potassium hexachloroplatinatein (0.15 g) was dissolved in 50 mL water, while heating. Then, 50 mL of a 6% potassium iodine solution were added and the solution kept in the refrigerator.
- **Potassium permanganate-sulfuric acid reagent:** KMnO₄ (1 g) was carefully dissolved in 30 mL conc. sulphuric acid and cooled with ice to avoid the generation of explosive manganhexoxide.
- **Natural substance-PEG reagent:** Solution A: A 1% solution of natural substance reagent A (diphenylboronic acid-aminooethyl ester) in MeOH was prepared and was kept in the refrigerator. Solution B: A 5% solution of polyethylene glycol 4000 in EtOH was prepared. Solutions A and B were sprayed one after the other onto the TLC plate. The PEG solution increased the detection sensitivity.
- **Hydrochloric acid-glacial acetic acid reagent:** Forty milliliters conc. hydrochloric acid were mixed with 10 mL glacial acetic acid.
- **Vanillin sulfuric acid reagent:** Solution A: A 5% solution of conc. sulfuric acid in EtOH was prepared (5 mL acid mixed with 95 mL ethanol). Solution B: A 1% solution of vanillin in EtOH was prepared (1 g vanillin mixed with 99 mL ethanol). Solution A was first sprayed onto the TLC plate and immediately thereafter, solution B was sprayed.
- **Thioglycosides:** The reference solutions consisted of 1 mg aloin and 1 mg franguline dissolved in at 5 μL. Super plasticizer consisting of EtOAc / MeOH / H₂O 50 / 8.5 / 6.5 was used with a running height of 15 cm. The plate was dried (max. 10 min) and immediately sprayed with freshly prepared solution of nitrosodimethyl aniline in pyridine (0.1%). The chromatogram was immediately evaluated. Ethanol KOH reagent was sprayed onto it afterwards and put into a 100-105°C warm drying cabinet for 15 min. The plate was evaluated in daylight and in UV light (365 nm).
- **Xanthines:** The reference solutions were caffeine, theophylline and theobromine dissolved in 300 μL methanol applied at a volume of 5 μL per point. Super plasticizer consisting of toluol / isopropanol / NH₃ conc. 15 / 30 / 5 was used with a running height of 15 cm. The dry TLC plate was evaluated under UV light (254 nm) and sprayed with potassium permanganate-sulfuric acid reagent. It was subsequently fan-dried and put into a 50°C warm drying cabinet for 15 min. Dragendorff reagent was sprayed onto the TLC plate afterwards and evaluated immediately in daylight.
- **Valepotriates:** The reference solution was Valmanc®. The sugar coat of one tablet Valmanc® was removed and half pulverized in a mortar. Ether (1.5 mL) was added to this powder in an Eppendorf vessel. An aliquot of 10 μL was applied per point as bands 15 mm length. Super plasticizer of toluol / EtOAc: 37.5 / 12.5 was used with a running height of 15 cm. The dry TLC plate was evaluated in UV light (254 nm) and sprayed with hydrochloric acid-acetic acid reagent. The plate was heated under supervision in a 110°C warm drying cabinet for 10 min. The evaluation occurred in daylight and in UV light (365 nm). The zone’s colours deepened while heating the TLC plate. This was the reason, why continuous supervision was important.

**TLC analyses**

A total of 35 plant extract samples were investigated, 10 samples per plate with bands of 10 mm length together with their corresponding references for each compound class.

**Alkaloids:** The reference compounds used included: Cinchonidin, cinchonin and chinidium dissolved in 500 μL of methanol applied at a final volume of 5 μL at a point. A super plasticizer of toluol/EtOAc/DEA: 5/4/1 was used with a running height of 12 cm. The plate was dried to remove DEA (fan-dry or put into a drying cabinet at 100°C). The chromatogram was evaluated under UV light (254 or 365 nm) and afterwards sprayed with 10% methanolic sulphuric acid (V/V) onto the plate. The chromatogram was evaluated again under UV light (365 nm) and sprayed with iodine-platinate-reagent onto the plate and thereafter evaluated in daylight.

**Anthraquinones:** The reference solutions were 1 mg aloin and 1 mg franguline dissolved in at 5 μL. Super plasticizer consisting of EtOAc / MeOH / H₂O 50 / 8.5 / 6.5 was used with a running height of 15 cm. The plate was dried (max. 10 min) and immediately sprayed with freshly prepared solution of nitrosodimethyl aniline in pyridine (0.1%). The chromatogram was immediately evaluated. Ethanol KOH reagent was sprayed onto it afterwards and put into a 100-105°C warm drying cabinet for 15 min. The plate was evaluated in daylight and in UV light (365 nm).

**Flavonoids:** The reference solution consisted of 1 mg of rutin, chlorogenic acid and hyperoside dissolved together in 1 mL MeOH. Also, 1 mg of caffeic acid dissolved in 1 mL MeOH was used. Aliquots of 25-30 μL of the herbal drug extracts and reference solutions were applied as bands of 10 mm length. Super plasticizer of EtOAc / formic acid / acetic acid / H₂O 50 / 5.5 / 5.5 / 13.5 was used. The first three components were mixed and water was slowly added while shaking the mixture with a running height of 15 cm. The super plasticizer was completely fan-dried or dried by putting the plate into a 100°C warm drying cabinet. The chromatogram was evaluated in UV light (254 and 365 nm). It was controlled after 15-20 min. Afterwards, natural substance-PEG reagent was sprayed onto the plate and evaluated in daylight.

**Essential oils:** A solution of 5 mg or μL anethole, carvone, 1, 8-cineol (eucalyptol), eugenol, menthol and thymol in 1.0 mL toluol
was used as reference solution. Super plasticizer of toluol / EtOAc 93 / 7 was used with a running height of 6 cm. After evaluating the dry TLC plate in UV light (254 and 365 nm), vanillin-sulfuric acid reagent was sprayed onto it and heated in a 110°C warm drying cabinet for 5-10 min under supervision. The evaluation occurred in daylight.

**Coumarin drugs:** The reference solution was caffeic acid dissolved in methanol with super plasticizer of toluene-ether 1:1 saturated with 10% acetic acid and a running height of 15 cm. Detection and evaluation occurred under UV light (365 nm) without further chemical treatment.

**Lignans:** The reference solution of rutin, chlorogenic acid and hyperoside dissolved in methanol was used with super plasticizer consisting of chloroform-methanol-water (70:30:4) and a running height of 15 cm. Detection and evaluation was done after drying the plate and spraying with PEG reagent and evaluating under UV light (365 nm).

**Saponins:** As reference solution, aescin dissolved in methanol with super plasticizer of chloroform-acetic acid-methanol-water; 60:32:12:8 with a running height of 15 cm was used. Detection and evaluation was done after drying the plate in a warm cabinet, watching under UV light (254 and 365 nm) and spraying with anisaldehyde-sulphuric acid reagent.

**Arbutin drugs:** The reference solutions included arbutin and rutin in 300 µL methanol with a super plasticizer of ethyl acetate-methanol-water (100/13.5/10) and a running height of 15 cm. After drying the plate in a warm cabinet and evaluating under UV light (256 and 354 nm), natural products polyethylene glycol reagent (NP/PEG) was sprayed and evaluated under UV light (365 nm).

### Results

**Phytochemical screening results**

TLC results of phytochemical constituents of 35 medicinal plants were summarized in Table 1 representing the designated sample ID, the botanical name of each medicinal plant and the various phytochemical constituents, i.e., alkaloids, anthraquinones, xanthines, valepotriates, cardiac glycosides, flavonoids, essential oils, coumarin drugs, lignans, saponins and arbutin drugs. The results revealed the presence of medically active compounds in the plant extracts. A synopsis of the frequency of all compounds detected in our set of 35 plants has been compiled according to the chemical class they belong to. As can be seen in Figure 1, xanthines, cardioactive glycosides and coumarins were the most frequent compounds (85.7% to 94.2%), while lignans, arbutin drugs, and anthraquinones were the least frequent phytochemicals (57.1% to 68.5%). All other chemical classes were present in intermediate frequencies. As can be seen in Figure 2, Representative TLC chromatograms of phytochemicals of medicinal plants from the Kakamega County, Kenya.

(a) **Alkaloids:** The predominantly light blue fluorescence of alkaloids was intensified by treatment with 10% methanolic sulphuric acid. Other alkaloids showed bright yellow and yellow green fluorescence.

(b) **Anthraquinones:** The bands showed characteristic yellow and blue fluorescence. All major compounds such as aloins or aloinosides showed quenching in UV light (254 nm). Treatment with KOH reagent intensified the yellow fluorescence of aloin and aloinosides as well as the blue fluorescence of aloe resins.

(c) **Xanthines:** Chromatograms showed red, blue and green fluorescence at UV light (360 nm).

(d) **Valepotriates:** The ingredients of Valmane® reference solution showed Rf-values of 0.70 for valtrate/isovaltrate, 0.60 for didrovaltrate, 0.55 for acevaltrate, and 0.40 forIVHD. The chromatogram showed blue and brown fluorences.

(e) **Cardioactive glycosides:** After spraying with conc. sulphuric acid and heating for 1-3 min at 100°C, blue, brown, green and yellowish fluorences appeared at 365 nm and appeared brown or blue in normal daylight.

(f) **Flavonoids:** The chromatograms showed yellow-orange or yellow-green and blue fluorences. Spraying with PEG reagent strengthened the fluorescence signals.

(g) **Essential oils:** At a wavelength of 254 nm UV light, the fluorescence was quenched and bands appeared as dark zones against a light green fluorescent background of the TLC plate. After spraying with vanillin sulphuric acid essential oil, strong blue, green, red and brown stainings appeared.

(h) **Coumarin drugs:** The chromatograms showed characteristic bright blue, blue-green, yellow green and violet-blue fluorescence of coumarins at a wavelength of 365 nm UV light.

(i) **Lignans:** The chromatograms showed blue and blue-green fluorescence zones under UV light of 365 nm. The blue fluorescence was strengthened after spraying with PEG reagent.

(j) **Saponins:** The chromatograms showed grey blue, yellow brown and red fluorescence bands at UV light (365 nm) after spraying with anisaldehyde-sulphuric acid reagent.

(k) **Arbutin drugs:** Distinct grey and blue zones were observed in Figure 2.

### Discussion

In the present study, we investigated the phytochemical constituents of 35 selected medicinal plants used in the Kakamega County, Kenya for the treatment of cancer. Phytochemical analyses of the present study revealed the presence of bioactive constituents known to possess important pharmacological effects [6]. Analyses of the plant extracts revealed the presence of alkaloids, anthraquinones, xanthines, valepotriates, cardioactive glycosides, flavonoids, essential oils, coumarins, lignans, saponins, and arbutin drugs. Different reference compounds were used as controls. In total, 71.4% of the medicinal plants contained alkaloids, 57.1% anthraquinones, 94.2% xanthines, 82.8% valepotriates, 94.2% cardioactive glycosides, 82.8% flavonoids, 77.1% essential oils, 85.7% coumarin drugs, 68.5% lignans, 80% saponins, and 62.8% arbutin drugs. These results show that the majority of the selected anti-cancer medicinal plants have a wide range of biologically important phytochemicals with cardioactive glycosides and xanthines as most abundant compounds. Phenolic compounds such as flavonoids have been previously shown to have anti-apoptosis, anti-aging, anti-carcinogenic, anti-inflammatory, anti-atherosclerotic, and cardiovascular protective activities [7]. Flavonoids in plants comprise a vast array of biologically active compounds which have been used in traditional medicine for many years and have majorly antioxidant and anti proliferative effects especially against chronic inflammatory and allergic diseases, breast cancer and coronary artery disease [8]. Ogawa et al. has reported evidence of flavonoids having antimutagenic activity of quercetin that was shown to inhibit the mutagenic activity of benzo[a]pyrene, a polyaromatic hydrocarbon carcinogen [9]. Flavonoids have also been found to be an active antimicrobial substance against a variety of microorganisms in vitro.

---

Citation: Ochwang’I DO, Kimwele CN, Oduma JA, Gathumbi PK, Kiama SG, et al. (2016) Phytochemical Screening of Medicinal Plants of the Kakamega Country, Kenya Commonly Used against Cancer. Med Aromat Plants (Los Angel) 5: 277. doi: 10.4172/2167-0412.1000277
Figure 1: Frequency of phytochemical constituents: 71.4% alkaloids, 57.1% anthraquinones, 94.2% xanthines, 82.8% valepotriates, 94.2% cardioactive glycosides, 82.8% flavonoids, 77.1% essential oils, 85.7% coumarin drugs, 68.5% lignans, 80% saponins, and 62.85% arbutin drugs.

Figure 2: Representative TLC chromatograms of phytochemicals of medicinal plants from the Kakamega County, Kenya. (a) Alkaloids, (b) Anthraquinones, (c) Xanthines, (d) Valepotriates, (e) Cardiac glycosides, (f) Flavonoids, (g) Essential oils, (h) Coumarin drugs, (i) Lignans, (j) Saponins, (k) Arbutin drugs.
Citation: Ochwang’l DO, Kimwele CN, Oduma JA, Gathumbi PK, Kiama SG, et al. (2016) Phytochemical Screening of Medicinal Plants of the Kakamega Country, Kenya Commonly Used against Cancer. Med Aromat Plants (Los Angel) 5: 277. doi: 10.4172/2167-0412.1000277

| Sample ID | Plant Species (Botanical family)                  | Alkaloids | Anthraquinones | Xanthines | VIPapotinates | Cardioactive Glycosides | Flavonoids | Essential Oils | Coumarin Drugs | Lignans | Saponins | Arbutin Drugs |
|-----------|---------------------------------------------------|-----------|----------------|-----------|---------------|--------------------------|------------|---------------|----------------|---------|----------|----------------|
| 1         | Harungana madagascariensis Lam. ex poir (Stem bark) | ++        | –              | +++       | +             | ++                       | –          | ++            | –              | –       | –        | –              |
| 2         | Fuerstia africana T.C.E. Fr. (Whole plant)       | –         | +              | ++        | ++            | ++                       | ++         | ++            | –              | –       | –        | –              |
| 3         | Sida rhombifolia L. (Leaves)                     | +         | –              | +++       | +             | ++                       | –          | –             | ++             | –       | –        | –              |
| 4         | Zanthoxylum rubescens Hook. f (Stem bark)        | +++       | +              | +++       | ++            | ++                       | ++         | ++            | –              | –       | –        | –              |
| 5         | Bridelia micrantha (Hochst.) Balli (Stem bark)   | +         | –              | –          | –             | –                        | –          | –             | –              | –       | –        | –              |
| 6         | Juniperus procera Endl. (Stem bark)              | +++       | +              | +++       | ++            | ++                       | ++         | ++            | ++             | –       | –        | –              |
| 7         | Tragia brevipes Pax (Leaves)                     | ++        | –              | +++       | ++            | ++                       | ++         | ++            | –              | –       | –        | –              |
| 8         | Phypanthus sapialis (Leaves)                     | +++       | +              | +++       | +++           | –                        | –          | +++           | +++            | ++      | –        | –              |
| 9         | Conyza sumatrensis (Retz.) E.H Walker (Leaves)   | +         | –              | +++       | ++            | ++                       | –          | +++           | +++            | ++      | –        | –              |
| 10        | Momordica foetida Schumach. (Whole aerial plant) | ++        | –              | +++       | +             | +++                       | +++        | ++            | –              | –       | –        | –              |
| 11        | Synsepalum cerasiferum Synonym: Afrosercalisia cerasifera (Welw.) Aubrev (Stem bark) | –         | –              | –          | +             | +                        | –          | –             | +              | –       | –        | –              |
| 12        | Aloe volkensii Engl. (Leaves)                    | +         | +              | +++       | ++            | ++                       | –          | ++            | ++             | –       | –        | –              |
| 13        | Aeschynomene abyscinica (A. Rich.) Vatke (Leaves) | +         | –              | +++       | +++           | +++                      | ++         | –             | +++            | –       | –        | –              |
| 14        | Futumia africana Benth. (Leaves)                 | ++        | –              | +++       | ++            | –                        | +          | –             | –              | –       | –        | –              |
| 15        | Cyphostemma serpens (A. Rich) (Roots)            | +         | +              | +++       | +++           | ++                       | –          | +++           | +++            | ++      | –        | –              |
| 16        | Ipomoea cairica (L.) (Leaves)                    | ++        | –              | +++       | +++           | ++                       | +++        | +++           | –              | –       | –        | –              |
| 17        | Spathodea campanulata P. Beauv. ssp. nilotica (Seem) (Stem bark) | –         | –              | +++       | ++            | +                        | –          | ++            | –              | –       | –        | –              |
| 18        | Abrus precatorius L. ssp africanus Verdc. (Roots and seeds) | ++        | –              | +++       | ++            | ++                       | +++        | +++           | +++            | –       | –        | –              |
| 19        | Triumfetta rhomboidea Jacq. (Leaves)             | +         | ++             | +++       | +++           | ++                       | +++        | ++            | +++            | ++      | –        | –              |
| 20        | Psidrax schimperiana (A. Rich) (Stem bark)       | ++        | ++             | +++       | ++            | +                        | ++         | ++            | ++             | –       | –        | –              |
| 21        | Ficus thonningii (Leaves and stem bark)          | +++       | +              | +++       | ++            | –                        | –          | +++           | –              | –       | –        | –              |
| 22        | Rotheca myricoides (Hochst. Steane and Mabb.) (Whole plant) | +         | +              | +++       | +++           | –                        | +          | –             | +++            | –       | –        | –              |
| 23        | Croton macrostachyus Delle (Stem bark)            | –         | –              | +++       | ++            | +                        | –          | +             | +              | –       | –        | –              |
| 24        | Vernonia lasiopus O Hoffin (Stem bark)           | –         | ++             | –          | –             | –                        | –          | –             | –              | –       | –        | –              |
with their activity postulated to be due to their ability to complex with extracellular and soluble proteins of bacterial cell wall [10]. Besides they have shown effectiveness as antioxidants and strong anticancer activities [11].

Essential oils do not represent a chemically consistent class of substances. They are all characterized by high volatility and aromatic smell. Most of the molecules consist of mono- and sesquiterpenes. Phenylpropane base units and sulfur and nitrogen containing substances (for example anthranilic acid esters) were also found. A plant’s essential oil is composed of many individual components with their activity postulated to be due to their ability to complex with extracellular and soluble proteins of bacterial cell wall [10]. Besides they have shown effectiveness as antioxidants and strong anticancer activities [11].

Table 1: Thin layer chromatography of medicinal plants commonly used for cancer treatment in the Kakamega County, Kenya.

| Plant Name | TLC | Staining | Antimicrobial | Antioxidant | Anti-Inflammatory | Anti-Cancer |
|------------|-----|----------|---------------|-------------|------------------|-------------|
| Albizia gummifera (J. F. Gmel.) (Leaves and Stem bark) | + | _ | ++++ | +++ | + | ++++ |
| Zanthoxylum gillettii (De Wild.) P. G. Waterman (Stem bark) | ++++ | ++++ | ++++ | ++++ | + | ++++ |
| Microglossa pyrifolia (Lam.) Kunze (Leaves) | _ | ++++ | ++++ | ++++ | + | ++++ |
| Senna didymobotrya (Fresen.) Irwin and Barneby (Leaves) | +++ | + | ++++ | ++++ | + | ++++ |
| Trichilia emetica Vahl (Stem bark) | _ | ++++ | ++++ | ++++ | + | ++++ |
| Entada abyssinica Steud, ex A. Rich. (Stem bark) | ++ | + | ++++ | ++++ | + | ++++ |
| Shirakopsis elliptica (Hochst.) Esser Synonym: Sapium ellipticum (Hochst. kraus) Pax (Stem bark) | _ | ++++ | ++++ | ++++ | + | ++++ |
| Ocimum gratissimum L. Suave wild O. tomentosum oliv. (Leaves) | ++ | + | ++++ | ++++ | + | ++++ |
| Prunus africana (Hook.f.) Kakman (Stem bark) | _ | ++ | ++++ | ++++ | + | ++++ |
| Phyllanthus fistcheni Pax (Leaves and roots) | ++ | + | ++++ | ++++ | + | ++++ |
| Olea hotch spp. Hochstetteri (Stem bark) | _ | + | ++++ | ++++ | + | ++++ |

Cytotoxicity, analgesic, antispasmodic and antibacterial are the main activities that have been associated with alkaloids [27]. The biogenesis principle of amino acid (biogenetic amine) and non-amine represents the common basis of all alkaloids. The classification of alkaloids is based on the heterocycle’s structure or on the biogenetic origin. Majority of this compound are derived from the amino acids lysine, tryptophan, tyrosine and arginine (Wagner and Bladt). Catharanthus roseus (L.) G. Don (also known as Madagascar periwinkle) leaves contain of two indole alkaloids, vinblastine and vincristine, both of which are well-established in medical oncology since decades [28]. Vinca alkaloids kill cancer cells by inhibiting tubulin polymerization during mitotic spindle formation [29].

Cardioactive glycosides are known to lower the blood pressure [30]. A group of glycosidic plant ingredients is called cardioactive steroid glycosides and cause specific myocardial effects owing to special structural characteristics. Cardiac glycosides (CGs) are natural compounds that are potent inhibitors of the plasma membrane Na+/K+-ATPase leading indirectly to the intracellular accumulation of Ca2+ ions which can lead to myocardial contractility. CGs tends to exert potent antinflammatory effects by increasing the immunogenicity of dying cancer cells [31]. Digitoxigenin can be classified as a cardioactive steroid aglycone’s prototype. Especially the cis-trans-cis connection of the steroid skeletal structure’s rings A-B-C, which is derived from the pregnan’s C21-frame (hydroxylated at C3 and C14, wearing an unsaturated lactone ring at C17), causes the cardiac effects (Wagner and Bladt). Digitals has been shown to block cell proliferation and in non-toxic concentrations has been shown to induce apoptosis in different malignant cell lines [32].
Anthraquinone derivatives are the characteristic ingredients of hydroxyanthraquinone drugs. They usually contain C- and O-bound sugar moieties. The sugar’s chemical bonding to the aglycone’s different C-atoms leads to more variations (Wagner and Bladt). The most abundant anthraquinone of rhubarb, a traditional Chinese medicine, emodin, which has been seen to have the capacity of inhibiting cellular proliferation, apoptosis induction and prevention of metastasis reported to act through tyrosine kinases, phosphoinositide 3-kinase (PI3K), protein kinase C (PKC), nuclear factor-kappa B (NF-κB), and mitogen-activated protein kinase (MAPK) signaling cascades [33].

Valepotriates are triesters of a terpenoid, tricyclic alcohol with the structure of an iridoid cyclopenta-c pyran with an attached epoxide ring (Regrett 1996). They have been seen to exhibit weak to moderate cytotoxicity against the human metastatic prostate cancer cell line, PC-3M [34].

The results obtained in this study thus suggest that the identified phytochemical compounds may be bioactive constituents against cancer and represent a valuable reservoir of compounds deserving substantial medicinal merit.

Conclusion

The present study shows the presence of medicinally important constituents in medicinal plants of the Kakamega County, Kenya, which could serve as valuable sources for useful drugs. We postulate that the anticancer activity is linked to the various constituents of the plants as cross-referenced with published data. In traditional medicine practice, these plants are strongly recommended and further work should be carried out to isolate, purify, and characterize the active constituents responsible for the activity of these plants.

Acknowledgements

The present study received financial support from Carnegie Corporation of New York through Region 4 Initiative in Science and Education (RISE AFNET) under Prof. Kiama Stephen and Johannes Gutenberg University, Germany. I am grateful to Prof. Dr. Thomas Effert (Institute of Pharmacy and Biochemistry, Johannes Gutenberg University, Germany) for providing the environment, resources and expertise that made this work successful. Further appreciation goes to Doris Rohr, Janine Naß and Ilona Zirbs in Prof. Effert’s laboratory for their material expertise. I appreciate the support of my university supervisors Prof. Charles Kimwele, Prof. Jenimah Oduma, Prof. Peter Gathumbi and Prof. Stephen Kiama.

References

1. Hill AF (1952) Economic Botany. A textbook of useful plants and plant products. 2nd edn. McGraw-Hill Book Company Inc., New York, USA.
2. Vasu K, Goud JV, Suryam A, Singara S, Chary MA (2009) Biomolecular and phytochemical analyses of three aquatic angiosperms. Afr. J. Microbiol. Res. 3: 418-421.
3. Criagg GM, David JN (2001) Natural product drug discovery in the next millennium. J. Pharm. Biol. 39: 8-17.
4. Pareek J, Chanda S (2007) Antibacterial and phytochemical studies on twelve species of Indian medicinal plants. Afr. J. Biomed. Res. 10: 175-181.
5. Ochwang’I DO, Kimwele CN, Oduma JA, Gathumbi PK, Mbaria JM, et al. (2014) Medicinal plants used in treatment and management of cancer in Kakamega County, Kenya. Journal of Ethnopharmacology 151: 1040-1055.
6. Sofowora A (1993) Medicinal plants and traditional medicine in Africa. Spectrum Books Ltd., Ibadan, Nigeria, pp: 191-289.
7. Han X, Shen T, Lou H (2007) Dietry polyphenols and their biological significance. Int. J. Mol. Sci., pp: 950-988.
8. Elliott M Jr, Chilkan K, Theoharis T (2000) The Effects of Plant Flavonoids on Mammalian Cells: Implications for Inflammation, Heart Disease and Cancer. Pharmacol Rev 52: 673-751.
9. Ogawa S, Hiyarayama T, Mohara M, Tokuda M, Hirai K, et al. (1985) The effect of quercetin on the mutagenicity of 2-acetylanilinofluorene and benzol[b]pyrene in Salmonella typhimurium strains. Mutat Res 142: 103-107.
10. Marjorie C (1996) Plant products as antimicrobial agents. Clinical Microbiol Rev 12: 564-562.
11. Salah N, Miller NJ, Pagange G, Tijburg L, Bolwell GP, et al. (1995) Polyphenolic flavonoids as scavenger of aqueous phase radicals as chai breaking antioxidant. Arch. Biochem. 2: 339-346.
12. Sylvestre M, Pichettea A, Longtina A, Nagaub F, Legault J (2006) Essential oil analysis and anticancer activity of leaf essential oil of Croton flaves L. from Guadeloupe. Journal of Ethnopharmacology 103: 99-102.
13. Lacy A, O’Kennedy R (2004) Studies on Courmarins and Courmarin-Related Compounds to Determine their Therapeutic Role in the Treatment of Cancer. Current Pharmaceutical Design 10: 3797-3811.
14. Musa MA, Cooperwood JS, Khan MOF (2008) A Review of Courmarin Derivatives in Pharmacotherapy of Breast Cancer. Current Medicinal Chemistry 15: 2664-2679.
15. MacRae WD, Neil GH (1984) Biological activities of lignans, 23: 1207-1220.
16. Hiroso T, Gohk M, Oka K (1994) Natural flavonoids and lignans are potent cytokatic agents against human leukemia HL-60 cells. Life Sciences 55: 1081-1089.
17. Francis G, Kerem Z, Harinder PS, Becker K (2002) The biological action of saponins in animal systems: a review. British Journal of Nutrition 88: 587-605.
18. Just MJ, Recio MC, Giner RM, Cueller MJ, Manez S (1998) Antiinflammatory activity of unusual lupine saponins from Bupleurum fractescens. 64: 404-407.
19. Kuznetzova TA, Anisimov MM, Popov AM (1982) A comparative study in vitro of physiological activity of triterpene glycosides of marine invertebrates of echinoderm type. Comparative Biochemistry and Physiology 73C: 41-43.
20. Rao AV, Sung MK (1995) Saponins as anticancerogens. Journal of Nutrition 125: 717-724.
21. Konoshima T, Takasaki M, Tokuda M, Nishino H, Duch M (1998) Anti-tumor-promoting activity of majonoside-R2 from Vietnamese ginger. Panax vietnamensis HA et GRUSHV (I). Biological and Pharmaceutical Bulletin 21: 834-838.
22. Marino SD, Iorizzi M, Palagiano E, Zollo F, Roussakis C (1998) Starfish saponins. 55. Isolation, structure elucidation, and biological activity of steroid oligoglycosides from an aquatic starfish of the family Asterinidae. Journal of Natural Products 61: 1319-1327.
23. Mimaki Y, Kuroda M, Kameyama A, Yokosuka A, Sashida Y, et al. (1998) Steroidal saponins from the underground parts of Ruscus aculeatus and their cytostatic activity on HL-60 cells. Phytochemistry 48: 485-493.
24. Podoval I, Elas M, Cieszka K (1998) In vitro antifungal and cytotoxic activity of triterpene saponosides and quinoid pigments from Lysimachia vulgaris L. Phytotherapy Research 12: 70-73.
25. Mupoo K, Haridas V, Hoffmann JJ, Wachter GA, Hutter LK, et al. (2001) Triterpenoid saponins from Acacia victoriae (Benth) decrease tumor cell proliferation and induce apoptosis. Cancer Research 61: 5496-5490.
26. Hanusek M, Ganesh P, Wolaksz E, Amizcen TJ, Staja TJ, et al. (2001) Avicins, a family of Triterpenoid saponins from Acacia victoriae (Benth), suppress H-ras mutations and aneuploidy in a murine skin carcinogenesis model. Proceedings of the National Academy of Sciences USA 98: 11551-11556.
27. Stray F (1998) The Natural Guide to Medicinal Herbs and Plants. Tiger Books International, London, p: 12-16.
28. Noble RL (1990) The discovery of the vinca alkaloids - chemotherapeutic agents against cancer. Biochemistry and Cell Biology 68: 1344-1351.
29. Jordan MA, Thrower D, Wilson L (1991) Mechanism of Inhibition of Cell Proliferation by Vinca Alkaloids. Cancer Res 51: 2212.
30. Nyarko AA, Addy ME (1990) Effects of aqueous extract of Adenia cissampeloides on blood pressure and serum analyte of hypertensive patients. Phytotherapy Res 4: 25-28.
31. Mengler L, Vacchelli E, Kepp O, Eggemort A, Tartour E, et al. (2013) Cardiac glycosides and cancer therapy (Trial watch). OncoImmunology 2: e23082.
32. Hau X (1999) Digiltin is a potential anticancer agent for several types of cancer. Medical Hypotheses 53: 543-548.
33. Huang Q, Lu G, Shen HM, Chung MCM. Ong CN (2007) Anti-cancer properties of anthraquinones from rhubarb. Med Res Rev 27: 609-630.
34. Xu YM, McLaughlin SP, Gunatahka AAL (2007) Oxygenlvelarbitrates A-D, Diene Valepotriates from Valeriana sortifolia. J Nat Prod 70: 2045-2048.