IRF6 polymorphisms in Mexican patients with non-syndromic cleft lip

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ABSTRACT

Cleft lip with or without cleft palate (CL/P) is one of the most common birth defects; it is a multifactorial disease affecting > 1/1,000 live births in Europe, and its etiology is largely unknown, although it is very likely genetic and environmental factors contribute to this malformation. Orofacial development is a complex process involving many genes and signaling pathways. Mutations in the gene for the interferon regulatory factor 6 (IRF6) cause a hereditary dominant malformation syndrome including CL/P, and polymorphisms are associated with non-syndromic CL/P (MIM 119530). Five SNPs at the locus with high heterozygosity in Caucasian populations were chosen for the present research due to their very strong association with CL/P. A case–parent trio study was performed using 292 samples from Mexico. Association with the rs1319435-C/C genotype (P = 0.02) was found in patients (73) as compared to pseudocontrols (219), while the genotype rs1319435-T/C was related with protection (P = 0.041) in the triad design. Significant over-transmission of the G allele for marker rs2235375 (P = 0.049) was found. Only the TACGT haplotype was diminished in the affected child, either in single (P = 0.0208) or double (P = 0.0208) dose. The pairwise analysis showed rs2235543 and rs2235371 were in strong linkage.
Introduction

Cleft lip with or without cleft palate (CL/P) is a common congenital structural anomaly notable for significant lifelong morbidity and its complex etiology. The prevalence of orofacial clefts varies from 1/500 to 1/2,500 births depending on geographic origin [Vanderas, 1987], racial and ethnic backgrounds [Croen et al., 1998; Tolarova and Cervenka, 1998], and socioeconomic status [Murray et al., 1997]. Non-syndromic CL/P (MIM 119530) is a complex multifactorial disease affecting 1/1,000 live births in Europe [Carinci et al., 2003]. The causes of orofacial clefts involve genetic and environmental factors [Schutte and Murray, 1999]. Epidemiological data in different populations have shown the prevalence of CP is generally lower than that of CL/P and families at high risk for one type of cleft are not at increased risk for the other type. A further subdivision of orofacial clefts into syndromic vs. isolated forms depends on whether additional structural and/or developmental anomalies occur with the cleft [Jugessur and Murray, 2005]. Orofacial development is a complex process involving many genes and signaling pathways [Murray and Schutte, 2004]. Alterations in one or more genes could cause CL/P.

Interferon regulatory factor 6 (IRF6) on 1q32.2 is consistent in its contribution to orofacial clefting among the large number of candidate genes and mutations in this gene cause a dominant malformations syndrome including CL/P and polymorphic markers strongly associated with CL/P [Jugessur et al., 2008; Rahimov et al., 2008].

IRF6 belongs to a family of nine transcription factors that share a high conserved winged helix DNA binding domain and a less conserved protein-binding domain [Kondo et al., 2002]. IRF6 mutations may produce a non-functional protein leading to haplo-insufficiency, affecting the DNA binding domain and cause a dominant negative effect, resulting in severe phenotypes [Houdayer et al., 2001]. Also mutations in the IRF6 transcriptional activation domain may inhibit transcriptional activation, conferring different effects on the function of IRF6 [Little et al., 2009].

A common polymorphic variant, in which isoleucine is substituted for valine at amino acid position 274 (V274I) in the protein-binding domain of IRF6, was identified in an analysis of IRF6 mutations [Kondo et al., 2002]. A family study showed highly significant transmission disequilibrium for this V274I variant with CL/P [Zucchero et al., 2004]. In an Italian population, four different SNPs were analyzed and revealed strong evidence of linkage disequilibrium between CL/P and the markers rs2013162 and rs2235375 [Scapoli et al., 2005].

In the present study, we analyzed five IRF6 SNPs (rs1319435, rs2013162, rs2235375, rs2235371, rs2235543) in 68 Mexican families to test for association with non-syndromic CL/P.

Materials and methods

Sample study

Our study sample consisted of 73 families including father, mother, and two brothers or sisters (without CL/P) of the affected child, corresponding to 292 subjects. All participants were recruited between 2009 and 2011, and all patients were screened for the presence of associated anomalies or syndromes by expert geneticists, and only those determined to have isolated cleft lip with or without cleft palate were included. This study was approved by the local Ethics and Research Committees of the Hospital General “Dr. Manuel Gea González”. Written informed consent was obtained from each person or legal representative of children. DNA was extracted from peripheral blood [Sambrook et al., 2001]. Polymerase chain reaction (PCR) of the IRF6 gene was performed using the primers described by Zucchero et al., 2004 and Scapoli et al., 2005. PCR was performed following manufacturer indications (Epicentre Biotechnologies, USA), and alleles were determined by dot-blot and specific digoxigenin-11-ddUTP-labeled oligonucleotide probes and visualized by
chemiluminescence [Bignon and Fernandez-Vina, 1995]. Hybridization results were interpreted by direct observation, considering positive as a strong black dot [Jimenez-Gonzalez et al., 2012].

**Statistical analysis**

Two approaches were performed: cases and pseudocontrols, according to the Falk and Rubinstein, 1987, and Spielman et al., 1993 description, “two parental genes not transmitted to their diseased offspring could be used as the control sample.” This would assure that both samples of the case’s genes and its matched control genes (i.e., nontransmitted parental genes) would come from the same genetic population. The second one consisted in a triad analysis considering child, mother, and father.

The data were analyzed by bivariate and multivariate statistics. Allele frequencies (AF) and genotype frequencies (GF) were calculated by direct counting of alleles and genotypes and were compared between the patients and the pseudocontrols using a Chi-squared test or a two-tailed Fisher’s exact test when the expected frequency in at least one cell was less than 5. Relative risk for alleles, genotypes, and haplotypes were calculated as an odds ratio (OR) according to Woolf formula [Woolf, 1955], using a 2 × 2 contingency table for each instance. Ninety-five percent confidence intervals (95%CI) were obtained by using Cornfield’s approximation. A log-linear method to test for asymmetric distribution of a particular variant allele among affected offspring and their biologic parents assuming Mendelian inheritance with or without Hardy–Weinberg equilibrium (HWE) was performed [Weinberg et al., 1998; Wilcox et al., 1998; Weinberg, 1999], using the program HAPLIN (version 5.3) [Gjessing and Lie, 2006]. This program was chosen because it was specifically designed to analyze genetic risk factors in offspring-parent triads and case–control collections, based on log-linear modeling, implementing a full maximum-likelihood model for estimation. It computes

| Alleles and genotypes | Affected (n = 73; %) | Non-affected (n = 219; %) | P       | OR (95%CI)* |
|-----------------------|----------------------|---------------------------|---------|-------------|
| rs1319435             |                      |                           |         |             |
| T                     | 60                   | 61                        | 0.89    | 0.97 (0.66–1.43) |
| C                     | 40                   | 39                        | 0.89    | 1.02 (0.69–1.50) |
| T/T                   | 29                   | 24                        | 0.43    | 1.27 (0.70–2.30) |
| T/C                   | 63                   | 74                        | 0.08    | 0.61 (0.35–1.07) |
| C/C                   | 8                    | 2                         | 0.02    | 3.84 (1.12–12.78) |
| rs2013162             |                      |                           |         |             |
| A                     | 62                   | 63                        | 0.81    | 0.95 (0.65–1.40) |
| C                     | 38                   | 37                        | 0.81    | 1.04 (0.71–1.54) |
| A/A                   | 23                   | 25                        | 0.71    | 0.89 (0.48–1.66) |
| A/C                   | 77                   | 75                        | 0.71    | 1.13 (0.60–2.10) |
| rs2235375             |                      |                           |         |             |
| C                     | 45                   | 50                        | 0.25    | 0.80 (0.55–1.17) |
| G                     | 55                   | 50                        | 0.25    | 1.25 (0.85–1.82) |
| C/C                   | 12                   | 21                        | 0.09    | 0.52 (0.24–1.12) |
| C/G                   | 64                   | 58                        | 0.30    | 1.34 (0.77–2.32) |
| G/G                   | 23                   | 21                        | 0.72    | 1.12 (0.60–2.11) |
| rs2235371 (V274I)     |                      |                           |         |             |
| A                     | 56                   | 56                        | 0.89    | 1.03 (0.70–1.50) |
| G                     | 44                   | 44                        | 0.89    | 0.98 (0.67–1.42) |
| A/A                   | 12                   | 13                        | 0.97    | 0.98 (0.44–2.20) |
| A/G                   | 88                   | 86                        | 0.74    | 1.15 (0.52–2.55) |
| G/G                   | 0                    | 1                         | 0.64    | 0.49 (0.02–9.89) |
| rs2235543             |                      |                           |         |             |
| T                     | 52                   | 51                        | 0.81    | 1.04 (0.72–1.52) |
| C                     | 48                   | 49                        | 0.81    | 0.96 (0.66–1.39) |
| T/T                   | 4                    | 3                         | 0.57    | 1.50 (0.37–6.16) |
| T/C                   | 96                   | 96                        | 0.88    | 0.90 (0.23–3.48) |
| C/C                   | 0                    | 1                         | 0.85    | 0.74 (0.03–16.60) |

A character in bold and italics indicates association.
explicit estimates of relative risks with asymptotic standard errors and confidence intervals. It uses the expectation-maximization (EM) algorithm to impute genotypes that are missing [Jugessur et al., 2012]. The most frequent haplotype was used as reference for estimating the effect in a multiplicative model.

Linkage disequilibrium (LD) was determined using Haploview (version 4.2) to reconstruct haplotypes and to estimate the relative risk associated with a single or double dose of each haplotype among the mother–father–child triads. Association analyses were performed under additive or genotype-wise models using family-based conditional logistic regression analysis [Cordell et al., 2004; Barrett et al., 2005; Gjessing and Lie, 2006].

To establish the most informative model of Mendelian association (dominant, co-dominant, recessive, overdominant, or log additive), the online SNPStats program was used in case–pseudocontrols and triads (Solé et al., 2006).

Results

Allelic and genotypic frequencies of all IRF6 polymorphisms, showing P, OR, and 95% CI, obtained by the bivariate analysis in patients and pseudocontrols, are summarized in Table 1. The most frequent alleles were rs1319435-T, rs2013162-A, rs2235375-G, rs223571-A, and rs2235543-T, while the more frequent genotypes were the heterozygous ones of each SNP. No association was found with any allele when considered alone. However, the genotype rs1319435-C/C showed increased risk ($P = 0.02$, OR (95% CI) = 3.84 (1.12–12.78), while the C/C genotype at rs2235375 showed a marginal association with a protective effect $P = 0.09$, 0.52 (0.24–1.12). Allele and genotype frequencies in these triads (mother–father case) showed no significant differences with any individual allele, but the genotype rs1319435-T/C was significantly

| Allele/genotype | Cases ($n = 68$; %) | Parents ($n = 136$; %) | $P$ | OR (95% CI)* |
|----------------|-------------------|------------------------|-----|--------------|
| rs1319435 | | | | |
| T | 61 | 60 | 0.83 | 1.05 (0.69–1.60) |
| C | 39 | 40 | 0.83 | 0.95 (0.62–1.46) |
| T/T | 31 | 23 | 0.21 | 1.51 (0.78–2.90) |
| T/C | 60 | 74 | 0.04 | 0.53 (0.28–0.98) |
| C/C | 9 | 3 | 0.07 | 3.19 (0.87–11.73) |
| rs2013162 | | | | |
| A | 62 | 62 | 0.66 | 0.91 (0.59–1.39) |
| C | 38 | 38 | 0.66 | 1.10 (0.72–1.68) |
| A/A | 24 | 25 | 0.49 | 0.78 (0.39–1.57) |
| A/C | 76 | 75 | 0.49 | 1.27 (0.64–2.55) |
| rs2235375 | | | | |
| C | 43 | 50 | 0.13 | 0.72 (0.48–1.10) |
| G | 57 | 50 | 0.13 | 1.39 (0.91–2.10) |
| C/C | 12 | 19 | 0.10 | 0.50 (0.21–1.16) |
| C/G | 63 | 62 | 0.71 | 1.12 (0.62–2.03) |
| G/G | 25 | 19 | 0.29 | 1.44 (0.73–2.83) |
| rs2235371 (V274I) | | | | |
| A | 56 | 57 | 0.93 | 1.02 (0.67–1.54) |
| G | 44 | 43 | 0.93 | 0.98 (0.65–1.49) |
| A/A | 12 | 14 | 0.85 | 0.92 (0.39–2.17) |
| A/G | 88 | 85 | 0.95 | 0.97 (0.44–2.17) |
| G/G | 0 | 1 | 0.53 | 1.88 (0.26–13.67) |
| rs2235543 | | | | |
| T | 52 | 51 | 0.84 | 1.04 (0.69–1.58) |
| C | 48 | 49 | 0.84 | 0.96 (0.63–1.45) |
| T/T | 4 | 2 | 0.40 | 1.98 (0.39–10.11) |
| T/C | 96 | 98 | 0.40 | 0.50 (0.10–2.57) |

A character in bold and italics indicates association.
increased among parents (Table 2). Table 3 shows overtransmitted alleles in the child obtained with Haploview v4.2, being allele rs2235375-G significant ($P = 0.049$).

The haplotype frequencies of the child and the mother in a single dose (one copy) or double dose (two copies) were obtained with HAPLIN v5.3 and are shown in Fig. 1, where OR are presented as relative risk (RR), and 15 haplotypes were dropped from the analysis because their frequency was less than 1%. Seventeen 5 SNP haplotypes were found for both mother and child, using CCGGT as the reference haplotype. TACGT haplotype was underrepresented in the affected child, either in single ($P = 0.0208$; RR (95% CI) = 0.157 (0.032–0.762)) or double ($P = 0.0208$; RR (95% CI) = 0.0247 (0.00102–0.58)) dose. Heterozygotes for the haplotype TAGGT showed a slight excess ($P = 0.0674$; RR (95% CI) = 6.95 (0.856–57.3)).

### Table 3

| SNP          | Overtransmitted | T:U* | Chi-square | $P$ value |
|--------------|-----------------|------|------------|-----------|
| 1 rs2235543  | T               | 68:65| 0.07       | 0.794     |
| 2 rs2235371  | G               | 60:56| 0.14       | 0.710     |
| 3 rs2235375  | G               | 51:33| 3.86       | 0.049     |
| 4 rs2013162  | C               | 52:50| 0.04       | 0.843     |
| 5 rs1319435  | T               | 52:49| 0.09       | 0.765     |

* Ratio of transmissions to non-transmissions of the overtransmitted allele. A character in bold and italics indicates association.

Fig. 1. Estimated relative risks of cleft lip with or without cleft palate (CL/P) in 68 Mexican children carrying one or two copies of the each haplotype. Upper half estimates based on haplotypes in the child; lower half estimates based on haplotypes in the mother. Vertical bars, 95% confidence intervals are on a logarithmic scale.
The pairwise LD ($D'$ and $r^2$) for all 5 SNPs is presented in Fig. 2. Two SNPs were in coding regions: rs2235371 in exon 7 producing a change of the amino acid valine to isoleucine at codon 274 (V274I) and rs2013162 in exon 5 is a silent variant (Ser153Ser), which are noted in green in Fig. 2. SNP rs2235543 is also intragenic, located in intron 6, rs1319435 is 70 kb 5' of IRF6 and rs2235543 is 100 kb 3' of IRF6 [Scapoli et al., 2005]. The LD plot in Fig. 2 indicates alleles at rs2235543 and rs2235371 are in strong LD among parents in these trios, while rs2235543, rs2235375, and rs1319435 seem be almost independent of one another. However, rs2235543 with rs2013162 and rs2235543 with rs1319435 also showed strong LD.

Genetic models were fitted for each SNP; odd ratios (OR) and 95% confidence intervals (CI) were calculated to check the relative risk for association with the online SNPStats program (Solé et al., 2006). The results are shown in the Table 4. No inheritance model was found for rs2013162 in the case–pseudocontrol approach, as well as for rs2013162 and rs2235543 in triads.

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**Table 4**

SNP model inheritance for case–pseudocontrols and triads.

| SNP         | Model       | Genotype | OR (CI 95%) | P value |
|-------------|-------------|----------|-------------|---------|
| Case-pseudocontrols | | | | |
| rs2235543   | Log additive | —        | 1.76 (0.48–6.39) | 0.40    |
| rs2235371   | Co-dominant  | G/A      | 0.97 (0.43–2.17) | 0.41    |
| rs2235375   | Recessive    | C/C      | 1.92 (0.89–4.16) | 0.08    |
| rs1319435   | Recessive    | C/C      | 0.26 (0.08–0.89) | 0.035   |
| Triads      | | | | |
| rs2235371   | Co-dominant  | G/A      | 0.81 (0.34–1.97) | 0.60    |
| rs2235375   | Recessive    | C/C      | 1.77 (0.76–4.16) | 0.17    |
| rs1319435   | Overdominant | C/T      | 1.90 (1.02–3.53) | 0.043   |

A character in bold and italics indicates association.
Discussion

The objective in this study was to test for evidence of association and linkage between five SNPs in the IRF6 gene and to calculate the risk of cleft lip with or without cleft palate. We used a population based case–pseudocontrol and case triad study of CL/P in a Mexican sample. Two studies that analyzed Asian and South American populations [Zucchero et al., 2004, Srichomthong et al., 2005] showed a significant association between CL/P and the rs2235371 (V274I) polymorphism in IRF6; in the Zucchero et al. (2004) study, no association with IRF6 was found between cleft palate alone and the V allele in the South American population, but in the South American and Asian groups, the results were highly significant (P < 0.001) for the association between cleft lip or palate and the V allele and also between cleft lip alone and the V allele. In the bivariate analysis of our study, this polymorphism alone did not show association with CL/P. However, an association with increased risk (P = 0.02; OR [95%CI] = 3.84 [1.12–12.78]) was seen with the homozygous genotype rs1319435-C/C in the case–pseudocontrol comparison, while the rs1319435-T/C genotype in the triad analysis was underrepresented among cases (P = 0.04; OR [95%CI] = 0.53 [0.28–0.98]).

Vieira et al. (2002) describe that Amerindian-specific haplotype D of mtDNA had a high frequency. Afterward, Vieira et al. (2007) investigated the specific maternal origin association with IRF6. Individuals with mtDNA haplotype D did not show association with the IRF6 alleles (P = 0.259), but individuals with mtDNA haplotypes other than haplotype D showed a trend for association with IRF6 (P = 0.08). Nevertheless, when they stratified by mitochondrial haplotype and for CL/P, the results indicated an association of IRF6 and risk of NSOFC among individuals with mitochondrial DNA haplotype other than haplotype D (P = 0.023).

On the other hand, admixture analysis of Mexican mestizos from different regions of the country has been carried out with different genetic systems. Few analyses have described the maternal lineages variability (mtDNA) throughout the Mexican territory. Martínez-Cortés et al. (2013) studied the mtDNA variation in 10 populations from different regions of Mexico. The maternal diversity estimated in the 742 Mexican mestizos studied was defined by nine haplogroups and paragroups. In the whole population sample Native American haplogroups A, B, C, and D were prevalent (92.9%), with frequencies of 47, 23.7, 15.9, and 6.2, respectively. In the Martínez-Cortés et al., 2013 work, the haplogroup D had the lowest total frequency (6.2%) without a clear geographical pattern. In our study, mtDNA was not analyzed, but if we considered the results of Vieira et al. (2007) and Martínez-Cortés et al. (2013), one would think that similar results could be found in our CL/P population. This kind of analysis would be important to perform in the next future, since in this moment it is not possible.

Since we had families with only one affected child, no model free linkage approach could be done because it is necessary the inclusion of multcase families for the calculation of identical-by-descent allele-sharing-based method.

The fitted models obtained by the SNPStats program were the recessive model for rs1319435 in case–pseudocontrols, which was associated with a protective effect (P = 0.035 OR (95% CI) = 0.26 (0.08–0.89)), and the overdominant model for rs1319435 in triads, that was significantly increased in affected child (P = 0.043 OR (95% CI) = 1.90 (1.02–3.53)).

Linkage disequilibrium between markers rs2235543, rs2235375, and rs1319435 was low in this population. TDT analysis showed a significant under-transmission of the C common allele and significant over-transmission of the G allele at marker rs2235375 (P = 0.0495) similar to the data of Huang et al., 2009. SNP rs2235375 showed the strongest association with over-transmission of the G allele with CL/P in our population, as reported in European-American, Taiwanese, Singaporean, Korean, and Western Chinese case–parent triads in a genome wide TDT analysis; however, the SNP rs2013162 was also significant in the same populations, except for Western Chinese [Huang et al., 2009; Park et al., 2007] and Mexican (this study).

IRF6 genotypes influence the risk of CL/P during development, and environment and ethnic factors should be considered as triggers since they could interact with the IRF6 pathway, altering fetal cell differentiation during palate formation. Our results agree with Park et al. (2007), who pointed that “significant results observed from SNPs other than rs2235371 (p.V274I) suggest that rs2235371 itself is not causal, but rather in LD with some causal mutation in IRF6.” Additionally, the lack of association of the rs2235371 SNP and CL/P may be influenced by the relatively small sample size analyzed in this work, although we calculated the statistical power for the case–pseudocontrol and triads association, being 0.62 and 0.53, respectively, and these values are under the rejection level of the null hypothesis, it is important to describe the behavior...
of these polymorphisms in a sample of the Mexican population, to increase the knowledge of CL/P for further extensive analysis.

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