Article

Synthesis of Nano-Praseodymium Oxide for Cataluminescence Sensing of Acetophenone in Exhaled Breath

Qian-Chun Zhang 1,*, Wu-Li Yan 2, Li Jiang 1, Yu-Guo Zheng 1, Jing-Xin Wang 2 and Run-Kun Zhang 2,*

1 School of Biology and Chemistry, Key Laboratory of Chemical Synthesis and Environmental Pollution Control-Remediation Technology of Guizhou Province, Xingyi Normal University for Nationalities, Xingyi 562400, China; jiangli@xynun.edu.cn (L.J.); zhengyuguo@xynun.edu.cn (Y.-G.Z.)
2 School of Public Health, Guangdong Pharmaceutical University, Guangzhou 510310, China; yanwuli@163.com (W.-L.Y.); wjx@gdpu.edu.cn (J.-X.W.)
* Correspondence: zhangqianchun@xynun.edu.cn (Q.-C.Z.); zhangrk@gdpu.edu.cn (R.-K.Z.);
Tel.: +86-589-3296359 (Q.-C.Z.); +86-020-34055201 (R.-K.Z.)

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Abstract: In this work, we successfully developed a novel and sensitive gas sensor for the determination of trace acetophenone based on its cataluminescence (CTL) emission on the surface of nano-praseodymium oxide (nano-Pr$_6$O$_{11}$). The effects of working conditions such as temperature, flow rate, and detecting wavelength on the CTL sensing were investigated in detail. Under the optimized conditions, the sensor exhibited linear response to the acetophenone in the range of 15–280 mg/m$^3$ (2.8–52 ppm), with a correlation coefficient ($R^2$) of 0.9968 and a limit of detection ($S/N=3$) of 4 mg/m$^3$ (0.7 ppm). The selectivity of the sensor was also investigated, no or weak response to other compounds, such as alcohols (methanol, ethanol, n-propanol, iso-propanol, n-butanol), aldehyde (formaldehyde and acetaldehyde), benzenes (toluene, o-xylene, m-xylene, p-xylene), n-pentane, ethyl acetate, ammonia, carbon monoxide, carbon dioxide. Finally, the present sensor was applied to the determination of acetophenone in human exhaled breath samples. The results showed that the sensor has promising application in clinical breath analysis.

Keywords: cataluminescence; sensor; acetophenone; Pr$_6$O$_{11}$

1. Introduction

In recent years, the development of cataluminescence (CTL)-based gas sensors have drawn extensive interest, mainly because of the high sensitivity, fast response, long-term stability, and simplicity of the device [1–3]. CTL is defined as the light emission during the catalytic oxidation reaction on the surface of catalytic solid material [4,5]. This phenomenon was firstly reported by Breysse et al. in 1976 [6]. After that, Analytical chemists have made great efforts to promote the development of CTL. Nowadays, CCL was proved to be a powerful technology for rapid analysis, which shows promising applications in environmental analysis [7–9], food analysis [10–12], clinical breath analysis [2,13,14], and biochemical analysis [15,16].

Sensing material plays an important role in a sensor system, it interacts with analytes to induce detectable signals. The characteristic of the sensing material directly affects the sensitivity, selectivity and stability of the sensor. In 2002, Zhang et al. first introduced nanomaterials as sensing material into the design of CTL gas sensor, they demonstrated that nanomaterials can greatly enhance CTL performances, due to the pleasing advantages of high surface areas and high reaction activity for...
nanomaterials [17]. Since then, many nanomaterials were prepared and used for design of CCL sensors. For instance, nanomaterial ZnSiO2-based sensor for ethyne [18], nanosized NaYF4:Er-based sensor for ketone [19], sensor based on α-MoO3 nanobelts for diethyl ether [20], sensor based on nano-MgO2 for iso-butanol [21], sensor based on nanomaterial g-C3N4 for carbon monoxide [22], nanomaterial NiO for H2S sensor [23], nanosized ZnO-based sensor for propionaldehyde [24].

There are abundant volatile organic compounds (VOCs) in human exhaled breath, some of them have been identified as biomarkers of diseases [25,26]. Analysis of concerned VOCs in exhaled breath has been recognized as a useful method for diagnosis of disease such as diabetes [27–29], asthma [30,31], lung cancer [32–34] and breast cancer [35,36]. The most outstanding advantage of breath analysis is that the pain-free and noninvasive collection of sample represents minimal risk to the human subjects [13,37]. Gas chromatography or chromatography/mass spectroscopy (GC/MS) are the traditional methods for breath analysis. However, these methods require complicated, bulky and expensive instruments, and their measuring procedures are also relatively tedious [38,39]. Sensors for breath analysis have attract wide attention because of their simple and compact device fabrication, as well as easy operation [40,41]. Examples include a WO3-based sensor for diagnosis of diseases [42], carbon nanotube (CNT)-based sensors functionalized with ionic liquids for infectious diseases [43], surface-enhanced Raman scattering sensor based on gold nanoparticles on reduced graphene oxide for gastric cancer [44].

Exhaled acetophenone in human breath is considered as an important biomarkers of breast cancers patients [45,46]. In the present work, nano-praseodymium oxide (nano-Pr6O11) was synthesized by hydrothermal method. The prepared nano-Pr6O11 exhibited strong CTL response to acetophenone, but no or weak responses to other compounds. Thus, the nano-Pr6O11 was used as sensing material to design CTL sensor for acetophenone. The sensor showed high sensitivity to acetophenone with a limit of detection of 4 mg/m3 (0.7 ppm). We demonstrated that the proposed sensor can be used for analysis of acetophenone in human exhaled breath. To the best of our knowledge, this work is first report of a CTL sensor for acetophenone.

2. Results and Discussion

2.1. Characterization

The physical-chemical characteristics such as morphology, size, and crystal form of nanomaterial directly affect the CTL behaviors. The surface morphology of the prepared Pr6O11 was investigated by SEM. As Figure 1a shows, quasi-hexagonal nanoparticles having about 50 nm diameters were obtained. Figure 1b shows the X-ray power diffraction patterns of Pr6O11. The diffraction peaks at 28.2°, 32.7°, 47.7°, 56.7°, 59.3°, 69.8°, 77.0°, 79.4° can be attributed to the (111), (200), (220), (311), (222), (400), (331), (420) planes. All of diffraction peaks are well indexed to a Pr6O11 (JCPDS.NO.41-1219) according to the standard card.
2.2. Selectivity of the CTL Sensor Based on Nano-Pr$_6$O$_{11}$

The selectivity of the CTL sensor based on nano-Pr$_6$O$_{11}$ was investigated. In a total of 18 kinds of chemical compounds include acetophenone, iso-propanol, benzophenone, n-propanol, n-butanol, methanol, ethanol, formaldehyde, acetaldehyde, n-pentane, ethyl acetate, toluene, o-xylene, m-xylene, p-xylene, ammonia, carbon monoxide, carbon dioxide, at concentration of 100 mg/m$^3$ were tested. As shown in Figure 2, acetophenone produce strong CTL intensity, iso-propanol, n-propanol and n-butanol just induce weak response. The intensities of iso-propanol, benzophenone, n-propanol and n-butanol are 5.5%, 4.1%, 3.1% and 2.6% of the acetophenone. In addition, no detectable CTL signals were observed for other compounds. Under the same conditions, ceramic heater without coating with nano-Pr$_6$O$_{11}$ was used to detection of acetophenone, no CTL signal was detected, indicating that appropriate nanomaterial is essential to the CTL emission of acetophenone.

It was reported that CTL responses are different for a given chemical compound on different nanomaterials, and the same nanomaterial shows dissimilar CTL responses to different chemical compounds. Although the detail reaction mechanism of CTL still remains uncertain, it is now well recognized that CTL emission must meet some requirements. One of the essential requirements is that the chemical compound can be oxidized to form a CTL intermediate, then the intermediate can produce photoemission when it returns to the ground state during the reaction process. It means that not all the chemical compounds can emit CTL on the same nanomaterial, even if they could be oxidized on the surface of nanomaterial. Although it is difficult for us to clarify the mechanism behind the reaction of acetophenone on nano-Pr$_6$O$_{11}$ surface, the above results demonstrate that the prepared nano-Pr$_6$O$_{11}$ exhibits satisfactory selectivity for sensing of acetophenone.

2.3. Optimization of Working Temperature

During the CTL sensing process, working temperature has a great effect on the CTL response. The effect of working temperature on the CTL response was investigated by changing the working temperature from 199 to 278 °C. As shown in Figure 3, the CTL intensity increases with temperature from 199 to 258 °C and decreases when the temperature over 258 °C. This phenomenon may be attributed to the reaction rate increases with increasing working temperature, but higher working temperature accelerate molecular motion that may results in quenching of CTL intensity. In addition,
higher working temperature emits stronger thermal radiation, leading to higher background noise is recorded. These factors determine the CTL signal and the S/N (signal-to-noise ratio) increase firstly then decrease with increment of working temperature. Because S/N reaches the maximum at 248 °C. Therefore, working temperature of 248 °C was chosen for further exploration.

![Figure 3](image1.png)

**Figure 3.** Effect of temperature on the cataluminescence (CTL) intensity and the S/N. Air flow rate, 230 mL/min; detecting wavelength, 425 nm, concentration, 100 mg/m³. Error bars stand for ±S.D.

### 2.4. Optimization of Air Flow Rate

The effect of the air flow rate on the CTL intensity was investigated by changing the flow rate in a rage of 180–250 mL/min. As shown in Figure 4, the CTL intensity increases gradually with the air flow rate from 180–230 mL/min, and reaches the maximum at 230 mL/min. Whereas, the CTL intensity becomes decline when the flow rate over 230 mL/min. This trend possibly results from the total reaction rate at low flow rate is controlled by the diffusion rate of acetophenone, and thereby CTL intensity increases with increasing flow rate. However, when the flow rate exceeds a certain degree, higher flow rate leads to insufficient reaction time, resulting in decrease in CTL intensity. Therefore, the optimal flow rate of 230 mL/min was selected for sensing of acetophenone.

![Figure 4](image2.png)

**Figure 4.** Effect of air flow rate on the CTL intensity. Temperature, 248 °C, detecting wavelength: 425 nm, concentration, 100 mg/m³. Error bars stand for ±S.D.
2.5. Optimization of Detecting Wavelength

In order to investigate the effect of detecting wavelength on CTL response, a series of optical filters, including 380, 400, 425, 440, 460, 490, 535, 555 and 575 nm were selected in sequence to detect acetophenone using nano-Pr$_6$O$_{11}$ as sensing martial. The CTL emission from the catalytic oxidation of acetophenone on the surfaces of nano-Pr$_6$O$_{11}$ and the S/N values at different emission wavelength are shown in Figure 5. It can be seen that the CTL intensity gradually increases with the wavelength from 380–460 nm and then decreases when the wavelength over 460 nm. The maximum emission wavelength is observed at 460 nm. However, the background noise emitted by thermal radiation increases dramatically with the wavelength, and the maximum S/N is observed at 425 nm. As a result, 425 nm was selected as the detecting wavelength for sensing of acetophenone.

![Figure 5](image)

**Figure 5.** CTL spectra emission on nano-praseodymium oxide. Temperature, 248 °C; air flow rate, 230 mL/min; concentration, 100 mg/m$^3$. Error bars stand for ±S.D.

2.6. CTL Response Profile and Analytical Characteristics

The CTL response profiles of acetophenone on the surface of nano-Pr$_6$O$_{11}$ were investigated by sensing of acetophenone at different concentrations under the above-optimized conditions. As shown in Figure 6a, the shape of the CTL response profiles is similar to each other. The CTL intensity sharply increases from the baseline to maximum value within 5 s after sample injection, the time of the CTL intensity decays from maximum value to baseline is about 20 s. These results demonstrate the capacities of rapid response and fast recovery of the sensor for acetophenone.

![Figure 6](image)

**Figure 6.** (a) The CTL response profiles. (b) The calibration curve for acetophenone. Temperature, 248 °C; air flow rate, 230 mL/min; wavelength, 425 nm. Error bars stand for ±S.D.
Under the optimized conditions, the CTL intensity is proportional to the concentration of acetophenone and exhibits a linear range of 15–280 mg/m$^3$ (2.8–52 ppm), and a detection limit of 4 mg/m$^3$ (0.7 ppm) is obtained at S/N = 3. The linear regression equation is $Y = 100.9X - 1356$ (correlation coefficient $R^2 = 0.9968$), where $Y$ is the relative CTL intensity, $X$ is the concentration of acetophenone (Figure 6b). Compared to the previously reported sensors for acetophenone [47,48], the present sensor shows wider linear range and lower LOD. The detail can be seen in Table 1.

### Table 1. Comparison of the analytical characteristics of the sensors for acetophenone.

| Principle                  | Sensing Materials                                    | Linear Range (ppm) | LOD (ppm) | References |
|----------------------------|------------------------------------------------------|--------------------|-----------|------------|
| CTL                        | Nano-Pr$_6$O$_{11}$                                   | 2.8–50             | 0.7       | Present work |
| Electrochemistry           | 1-Octyl, 3-methylimidazolium tetrafluoroborate        | 5–80               | 2.0       | [47]       |
| Quartz microbalance        | Macrocyclic oligolactams                             | 5–40               | 2.0       | [48]       |

2.7. Sample Analysis

Acetophenone in exhaled breath is identified as a biomarker of breast cancer. In order to probe the potential application of the designed sensor, 100 mL of exhaled breath samples from three volunteers were collected in sampling bags, then were detected by the CTL sensor based on nano-Pr$_6$O$_{11}$. However, no signal was measured, possibly due to the concentration of acetophenone was too low to be detectable. The three samples were spiked with different concentrations of acetophenone, subsequently, the spiked sample were measured by the sensor for recovery analysis, the results are shown in Table 2. The recoveries of acetophenone in the three samples are 106.7–113.1%, the RSDs are 3.2–4.0%. The satisfactory results demonstrate the potential of the sensor for real sample analysis.

### Table 2. Determination of acetophenone in the exhaled breath samples spiked with acetophenone by the designed sensor ($n$ = 3).

| Sample No. | Spiked Concentration (mg/m$^3$) | Measured Concentration (mg/m$^3$) | Recovery (%) | RSD (%) |
|------------|---------------------------------|-----------------------------------|--------------|---------|
| 1          | 20.0                            | 21.9 ± 0.9                        | 109.5        | 4.0     |
| 2          | 25.0                            | 26.7 ± 0.9                        | 106.7        | 3.2     |
| 3          | 30.0                            | 33.9 ± 1.3                        | 113.1        | 3.8     |

3. Experimental Section

3.1. Materials and Instrumentation

Praseodymium nitrate hexahydrate, urea, acetophenone, iso-propanol, n-propanol, n-butanol, methanol, ethanol, formaldehyde, acetaldehyde, n-pentane, ethyl acetate, toluene, o-xylene, m-xylene, p-xylene, ammonia were purchased from Aladdin Reagent Co. Ltd. (Shanghai, China). Carbon monoxide and carbon dioxide were purchased from The National Standard of Material Resources Network (Beijing, China).

The morphology of Pr$_6$O$_{11}$was characterized by using a scanning electron microscopy (Helios G4 CX). X-ray diffraction patterns were recorded on a Rigaku Ultima IV X-Ray Diffractometer using CuK$\alpha$ radiation with a scan speed of 5$^\circ$/min ranging from 10$^\circ$ to 80$^\circ$. (\(\lambda = 1.54\ \text{Å},\) operated on 40 mA and 40 kV current). BPCL ultra-weak luminescence analyzer (Guangzhou Microphotonics Technologies Co., Ltd., Guangzhou, China) was used to measured luminescent signal. The luminescence analyzer equipped with a photomultiplier detector, a high voltage of 680 V was applied to the photomultiplier, the Data acquisition rate was set as 0.5 s.
3.2. Synthesis of Praseodymium Oxide Nanoparticles

1.0 g of praseodymium nitrate hexahydrate and 6.5 g of urea were dissolved in 240 mL deionized water under ultrasonically vibrating for 20 min. The resulting solution was dried at 100 °C for 2 h. Subsequently, the obtained gels were rinsed with deionized water and ethanol. Finally, the product was calcined at 500 °C for 4 h in air to obtain the target products.

3.3. Procedure for Sensing

The CTL sensor for acetophenone was fabricated according to our previous work [13,49]. In brief, 3.0 mL of deionized water was mixed with 0.5 g of solid catalyst, then the suspended liquid containing catalyst was dripped onto the surface of a ceramic heater and was heated to 450 °C for 20 min. The ceramic heater coating with catalyst was put into a lab-made quartz tube that with a gas inlet and outlet. A voltage regulator was used to control the temperature of the ceramic heater by adjusting the output voltage. An air pump was used to support air carrier, and the flow rare was controlled by a flowmeter. The CTL signal was measured and processed by the BPCL ultra-weak luminescence analyzer. The detecting wavelength was selected by choosing optical filter. At the beginning of each series of experiments, the CTL sensor based on praseodymium oxide was heated to 450 °C for 10 min in air to activate the catalyst, and eliminate the influence of previous absorbates. Then the temperature of the sensor was reduced to a certain value for subsequent detection.

4. Conclusions

In summary, we synthesized nano-Pr$_6$O$_{11}$ via a simple hydrothermal method, and demonstrated that the prepared nanomaterial can be used as sensing material for CTL detection of acetophenone. The CTL sensor exhibited good selectivity, fast response, and high sensitivity for sensing acetophenone. The sensor was applied to the determination of acetophenone in exhaled breath samples, satisfactory recoveries in the range of 106.7–113.1% were obtained. This work provides a simple, rapid, and sensitivity method for sensing of acetophenone in many cases, especially in exhaled breath.

Author Contributions: Q.-C.Z. and R.-K.Z. conceived and designed the experiment, and wrote the paper. W.-L.Y., L.J. and Y.-G.Z. performed the experiments and analyzed the data. J.-X.W. helped with characterization of nanomaterial. All authors have discussed and approved the manuscript.

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References

1. Hu, J.X.; Zhang, L.C.; Yi, L. Recent advances in cataluminescence gas sensor: Materials and methodologies. *Appl. Spectrosc. Rev.* 2018, 1–19. [CrossRef]
2. Na, N.; Liu, H.Y.; Han, J.Y.; Han, F.F.; Liu, H.L.; Ouyang, J. Plasma-Assisted cataluminescence sensor array for gaseous hydrocarbons discrimination. *Anal. Chem.* 2012, 84, 4830–4836. [CrossRef] [PubMed]
3. Han, F.F.; Yang, Y.H.; Han, J.Y.; Ouyang, J.; Na, N. Room-temperature cataluminescence from CO oxidation in a non-thermal plasma-assisted catalysis system. *J. Hazard. Mater.* 2015, 293, 1–6. [CrossRef] [PubMed]
4. Zhang, R.K.; Cao, X.A.; Liu, Y.H.; Chang, X.Y. A new method for identifying compounds by luminescent response profiles on a cataluminescence based sensor. *Anal. Chem.* 2011, 83, 8975–8983. [CrossRef] [PubMed]
5. Zhang, L.C.; Song, H.J.; Su, Y.Y.; Lv, Y. Advances in nanomaterial-assisted cataluminescence and its sensing applications. *TrAC Trends Anal. Chem.* 2015, 67, 107–127. [CrossRef]
6. Breysse, M.; Claudel, B.; Faure, L.; Guerin, M.; Williams, R.J.; Wolkenstein, T. Chemiluminescence during the catalysis of carbon monoxide oxidation on a thoria surface. *J. Catal.* 1976, 45, 137–144. [CrossRef]
7. Zhang, L.J.; He, N.; Shi, W.Y.; Lu, C. A cataluminescence sensor with fast response to diethyl ether based on layered double oxide nanoparticles. *Anal. Bioanal. Chem.* 2016, 408, 8787–8793. [CrossRef]
8. Zhang, L.J.; Wang, S.; Yuan, Z.Q.; Lu, C. A controllable selective cataluminescence sensor for diethyl ether using mesoporous TiO2 nanoparticles. Sens. Actuators B 2016, 230, 242–249. [CrossRef]

9. Han, J.Y.; Han, F.F.; Ouyang, J.; He, L.X.; Zhang, Y.T.; Na, N. Low temperature CO sensor based on cataluminescence from plasma-assisted catalytic oxidation on Ag doped alkaline-earth nanomaterials. Nanoscale 2014, 6, 3069–3072. [CrossRef]

10. Li, Z.H.; Wei, X.; Chao, L. Hydrotalcite-supported gold nanoparticle catalysts as a low temperature cataluminescence sensing platform. Sens. Actuators B 2015, 219, 354–360. [CrossRef]

11. Li, M.; Chen, J.Y.; Hu, Y.F.; Li, G.K. Titanium dioxide-Yttrium(III)-oxide composite based cataluminescence gas Sensor for fast detection of propylene oxide. Chin. J. Anal. Chem. 2019, 47, 191–196. [CrossRef]

12. Zhang, R.K.; Li, G.G.; Hu, Y.F. Simple and excellent selective chemiluminescence-based Cs2 on-line detection system for rapid analysis of sulfur-containing compounds in complex samples. Anal. Chem. 2015, 87, 5649–5655. [CrossRef] [PubMed]

13. Zhang, R.K.; Huang, W.T.; Li, G.K.; Hu, Y.F. Noninvasive strategy based on real-time in vivo cataluminescence monitoring for clinical breath analysis. Anal. Chem. 2017, 89, 3353–3361. [CrossRef] [PubMed]

14. Wu, Y.Y.; Wen, F.; Liu, D.; Kong, H.; Zhang, C.H.; Zhang, S.C. Analysis of 2-propanol in exhaled breath using in situ enrichment and cataluminescence detection. Luminescence 2011, 26, 125–129. [CrossRef] [PubMed]

15. Kong, H.; Liu, D.; Zhang, S.C.; Zhang, X.R. Protein sensing and cell discrimination using a sensor array based on nanomaterial-assisted chemiluminescence. Anal. Chem. 2011, 83, 1867–1870. [CrossRef]

16. Kong, H.; Wang, H.; Zhang, S.C.; Zhang, X.R. A thermochemiluminescence array for recognition of protein subtypes and their denatured shapes. Analyst 2011, 136, 3643–3648. [CrossRef]

17. Zhu, Y.F.; Shi, J.J.; Zhang, Z.Y.; Zhang, C.; Zhang, X.R. Development of a gas sensor utilizing chemiluminescence on nanosized titanium dioxide. Anal. Chem. 2002, 74, 120–124. [CrossRef]

18. Peng, C.H.; Shao, K.; Long, Z.; Ouyang, J.; Na, N. A plasma-assisted cataluminescence sensor for ethyne detection. Anal. Bioanal. Chem. 2016, 408, 1–8. [CrossRef]

19. Tang, J.; Song, H.J.; Zeng, B.R.; Zhang, L.C.; Lv, Y. Cataluminescence gas sensor for ketones based on nanosized NaYF4: Er. Sens. Actuators B 2016, 222, 300–306. [CrossRef]

20. Zhen, Y.Z.; Zhang, H.M.; Fu, F.; Zhang, Y.T. A cataluminescence sensor based on α-MoO3 nanobelts with low working temperature for the detection of diethyl ether. J. Mater. Sci. Mater. Electron. 2019, 30, 1–7. [CrossRef]

21. Meng, F.F.; Lu, Z.Y.; Zhang, R.K.; Li, G.K. Cataluminescence sensor for highly sensitive and selective detection of iso-butanol. Talanta 2019, 194, 910–918. [CrossRef] [PubMed]

22. Li, L.; Deng, D.Y.; Huang, S.X.; Song, H.J.; Xu, K.L.; Zhang, L.C.; Lv, Y. UV-assisted Cataluminescence Sensor for Carbon Monoxide based on Oxygen Functionalized g-C3N4 Nanomaterials. Anal. Chem. 2018, 90, 9598–9605. [CrossRef] [PubMed]

23. Yu, K.L.; Hu, J.X.; Li, X.H.; Zhang, L.C.; Lv, Y. Camellia-like NiO: A novel cataluminescence sensing material for H2S. Sens. Actuators B 2019, 288, 243–250. [CrossRef]

24. Cher, Y.X.; Zhang, Q.C.; Li, Y.H.; Xu, Z.M.; Long, W.R. A cataluminescence sensor for propionaldehyde based on the use of nanosized zirconium dioxide. Microchem. Acta 2014, 181, 1152–1132. [CrossRef]

25. Konvalina, G.; Haick, H. Sensors for breath testing: From nanomaterials to comprehensive disease detection. Acc. Chem. Res. 2014, 47, 66–76. [CrossRef] [PubMed]

26. Kim, K.H.; Jahan, S.A.; Kabir, E. A review of breath analysis for diagnosis of human health. TrAC Trends Anal. Chem. 2012, 33, 1–8. [CrossRef]

27. Saidi, T.; Zaim, O.; Moufid, M.; El Bari, N.; Ionescu, R.; Bouchikhi, B. Exhaled breath analysis using electronic nose and gas chromatography–mass spectrometry for non-invasive diagnosis of chronic kidney disease, diabetes mellitus and healthy subjects. Sens. Actuators B 2018, 257, 178–188. [CrossRef]

28. Worrall, A.D.; Qian, Z.; Bernstein, J.A.; Angelopoulos, A.P. Water-resistant polymeric acid membrane catalyst for acetone detection in the exhaled breath of diabetics. Anal. Chem. 2018, 90, 1819–1826. [CrossRef]

29. Neerincx, A.H.; Vijverberg, S.J.H.; Bos, L.D.J.; Brinkman, P.; van der Schee, M.P.; de Vries, R.; Sterk, P.J.; Maitland-van Der Zee, A. Breathomics from exhaled volatile organic compounds in pediatric asthma. Pediatr. Pulmonol. 2017, 52, 1616–1627. [CrossRef]
31. Brinkman, P.; van de Pol, M.A.; Gerritsen, M.G.; Bes, L.D.; Dekker, T.; Smids, B.S.; Sinha, A.; Majoor, C.J.; Snoebeor, M.M.; Knobel, H.H.; et al. Exhaled breath profiles in the monitoring of loss of control and clinical recovery in asthma. Clin. Exp. Allergy 2017, 47, 1159–1169. [CrossRef] [PubMed]

32. Peng, G.; Tisch, U.; Adams, O.; Hakim, M.; Shehada, N.; Broza, Y.Y.; Billan, S.; Abdah-Bortnyak, R.; Kuten, A.; Haick, H. Diagnosing lung cancer in exhaled breath using gold nanoparticles. Nat. Nanotechnol. 2009, 4, 669–673. [CrossRef] [PubMed]

33. Katwal, G.; Paulose, M.; Rusakova, I.A.; Martinez, J.E.; Varghese, O.K. Rapid growth of zinc oxide nanotube-nanowire hybrid architectures and their use in breast cancer related organics detection. Nano Lett. 2016, 16, 3014–3021. [CrossRef]

34. Sagin, K.; Ling, C.; Murray, E.P.; Mainardi, D.S. Kinetics of nitric oxide and oxygen gases on porous Y-stabilized SrCO$_3$ based sensors. Molecules 2018, 23, 2432–2437. [CrossRef] [PubMed]

35. Silva, C.L.; Perestrelo, R.; Silva, P.; Tomás, H.; Cámara, J.S. Volatile metabolomic signature of human breast cancer cell lines. Sci. Rep. 2017, 7, 43969. [CrossRef]

36. Rudnicka, J.; Kowalkowski, T.; Buszewski, B. Searching for selected VOCs in human breath samples as potential markers of lung cancer. Lung Cancer 2019, 135, 123–129. [CrossRef] [PubMed]

37. Hakim, M.; Broza, Y.Y.; Barash, O.; Peled, N.; Phillips, M.; Amann, A.; Haick, H. Volatile organic compounds of lung cancer and possible biochemical pathways. Chem. Rev. 2012, 112, 5949–5966. [CrossRef]

38. Silva, C.L.; Perestrelo, R.; Silva, P.; Tomás, H.; Cámara, J.S. Volatile metabolomic signature of human breast cancer cell lines. Sci. Rep. 2017, 7, 43969. [CrossRef]

39. Martínez-Aquino, C.; Costero, M.A.; Gil, S.; Gaviña, P. A New environmentally-friendly colorimetric probe for formaldehyde gas detection under real conditions. Molecules 2018, 23, 2646. [CrossRef]

40. Martinet-Aquino, C.; Costero, M.A.; Gil, S.; Gaviña, P. A New environmentally-friendly colorimetric probe for formaldehyde gas detection under real conditions. Molecules 2018, 23, 2646. [CrossRef]

41. Hengwei, L.; Minseok, J.; Suslick, K.S. Preoxidation for colorimetric sensor array detection of VOCs. J. Am. Chem. Soc. 2011, 133, 16786.

42. Park, C.H.; Schroeder, V.; Kim, B.J.; Swager, T.M. Ionic liquid-carbon nanotube sensor arrays for human breath related volatile organic compounds. ACS Sens. 2018, 3, 2432–2437. [CrossRef] [PubMed]

43. Choi, S.; Fuchs, F.; Demadrille, R.; Grévin, B.; Jang, B.; Lee, S.; Lee, J.; Tuller, H.L.; Kim, I. Fast responding exhaled-breath sensors using WO$_3$ hemitubes functionalized by graphene-based electronic sensitizers for diagnosis of diseases. ACS Appl. Mater. Interfaces 2014, 6, 9061–9070. [CrossRef] [PubMed]

44. Chen, Y.S.; Zhang, Y.X.; Pan, F.; Liu, J.; Wang, K.; Zhang, C.L.; Cheng, S.L.; Lu, L.G.; Zhang, W.; Zhang, Z.; et al. Breath analysis based on surface-enhanced raman scattering sensors distinguishes early and advanced gastric cancer patients from healthy persons. ACS Nano 2016, 10, 8169–8179. [CrossRef]

45. Peng, G.; Hakim, M.; Broza, Y.Y.; Billan, S.; Abdah-Bortnyak, R.; Kuten, A.; Tisch, U.; Haick, H. Detection of lung, breast, colorectal, and prostate cancers from exhaled breath using a single array of nanosensors. Br. J. Cancer 2010, 103, 542–551. [CrossRef]

46. Michael, P.; David, B.J.; Cataneo, R.N.; Jan, H.; Kaplan, P.D.; Lalissang, R.I.; Philippe, L.; Lobbes, M.B.I.; Mayur, M.; Nadine, P. Rapid point-of-care breath test for biomarkers of breast cancer and abnormal mammograms. PLoS ONE 2014, 9, e90226.

47. Gębićki, J.; Kloskowski, A.; Chrzanoski, W. Prototype of electrochemical sensor for measurements of volatile organic compounds in gases. Sens. Actuators B 2013, 177, 1173–1179. [CrossRef]

48. Muhr, E.; Leicht, O.; González, S.S.; Thanbichler, M.; Heider, J. A fluorescent bioreporter for acetophenone and 1-phenylethanol derived from a specifically induced catabolic operon. Front. Microbiol. 2016, 6, 1561–1572. [CrossRef]

49. Zhang, Q.C.; Meng, F.F.; Lin, C.; Wang, X.Y.; Zhang, G.Y. A sensitive cataluminescence-based sensor using a SrCO$_3$/graphene composite for n-propanol. RSC Adv. 2015, 5, 57482–57489. [CrossRef]

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