BIOCHEMICAL AND HISTOPATHOLOGICAL CHANGES OF TRADITIONAL SIDDHA FORMULATION ON CCL4 INDUCED LIVER FIBROSIS IN RATS

DIWAKAR MANICKAM*, SRI KAMATCHI PRIYA RAMAMOORTHY*, B. SANTHOSH KUMAR*, SAMU SUBRAMANIAM*, SHYAMA SUBRAMANIAM*

*Department of Biochemistry, Regenix Super Speciality Laboratories, Affiliated to University of Madras, Chennai, Tamilnadu, India, a,b,c
Lab Director, Head of the Department, Department of Biochemistry, Regenix Super Speciality Laboratories, Chennai, Tamilnadu, India, 
Consultant, Lab Services, Apollo Hospital, Chennai, Tamilnadu, India
Email: subramaniam@regenix.in

ABSTRACT

Objective: The main objective of this study was to evaluate the therapeutic activity of traditional Siddha formulation (TSF) on CCl4 induced liver fibrosis in rats. Methods: Plant materials were collected, shade dried and mixed in equal proportion. This mixture was used for the extraction process (kasayam). Twenty four male Wistar albino rats were randomly divided into four groups of six rats each. The normal control group was allowed free access to food and water. The drug control group was allowed to the daily intragastric administration of TSF dosage 400 mg/kg body weight/day. Liver damage was performed in the remaining two groups with an intraperitoneal injection of a 1.0 ml/kg body weight CCl4 and olive oil (2:3 v/v) mixture, weekly twice for 8 w. All rats, with the exception of the injury model group, were intragastrically administered TSF dosage 400 mg/kg body weight/day from 5th to end of the experiment of CCl4 intoxicated rats. The serum levels of Alamine Transaminase (ALT), Aspartate Transaminase (AST), Alkaline Phosphatase (ALP), Lactate Dehydrogenase (LDH), Y-Glutamyl Transferase (Y-GT), Albumin (ALB), Total Protein (TP) and Total Bilirubin (TB) and Hydroxyproline (HYP) in the rat livers were measured. Histopathological changes in the liver were assessed for each group using hematoxylin and eosin and Masson trichrome staining procedures. Results: At the end of the 8-week experimental period, histopathological examination and Masson trichrome staining demonstrated that TSF could attenuate inflammation, necrosis and reduce the score of liver fibrosis. Similarly, we found that the TSF administration significantly decreased the levels of biomarkers AST (136.25±3.88, p<0.01), ALT (64.81±4.68, p<0.01), ALP (132.21±8.30, p<0.01), LDH (1451.66±49.56, p<0.01), TB (0.87±0.04, p<0.01), Y-GT (12.81±1.74, p<0.01), HYP (8.50±7.67, p<0.01) and significantly increased ALB (3.73±0.24, p<0.01), TP (5.76±0.15, p<0.01), in associated with hepatic damage compared with CCl4 intoxicated group. Conclusion: TSF attenuates CCl4 intoxicated hepatic fibrosis in Wistar rats, which may be related to essential phytochemical ingredients rich in this formulation. These therapeutic activities were in the form of improving the serum biochemical parameters and a histopathological picture of the liver.

Keywords: Traditional Siddha formulation, Liver fibrosis, CCl4 and Vaithiyars

INTRODUCTION

Liver fibrosis is the scarring process, characterized by the loss of hepatocytes destruction, hepatic architecture, proliferation of hepatic fibroblast [1] to chronic injury, including alcohol, chronic viral hepatitis, nonalcoholic fatty liver diseases and drug exposure [2], results imbalance secretion and degrading of extracellular matrix including collagen, polysaccharide and glycoprotein which distorts liver architecture [3]. Advanced liver fibrosis leads to cirrhosis, liver failure and portal hypertension [4].

Carbon tetrachloride is a widely used potent hepatotoxin for induction of hepatocellular damage involving enhanced inflammatory process, recruitment of inflammatory cells [5], hepatocyte apoptosis [6], increase oxidative stress [7], oxidative stress plays important role in the development of chronic and degenerative diseases [8]. CCl4 induced liver fibrosis model similar to all important properties of liver disease. Qualitative analysis of TSF showed the presence of various beneficial phytochemicals like flavonoids, alkaloids, terpenoids, saponin, steroids and rich in polyphenols [14]. The aim of the present study was to observe and document the therapeutic activity of TSF on liver fibrosis induced by CCl4 in Wistar rats.

Siddha medicine is practiced mainly by institutionally trained and non-institutionally trained practitioner. Non-institutionally Siddha practitioner named as Siddha Vaithiyars got knowledge through hereditary family training or teachers (Aasaan). A Large number of Siddha formulation includes plant, herbomineral, animal kingdom based medicine passed among the family member of the practitioner and rarely to a lot of cures and formulations are secret [13].

Traditional formulation is a polyherbal Siddha formulation consists of seven botanicals used by the traditional Siddha medicine practice for liver disease. Siddha medicine is practiced mainly by institutionally trained and non-institutionally trained practitioner. Non-institutionally Siddha practitioner named as Siddha Vaithiyars got knowledge through hereditary family training or teachers (Aasaan). A Large number of Siddha formulation includes plant, herbomineral, animal kingdom based medicine passed among the family member of the practitioner and rarely to a lot of cures and formulations are secret [13].

Three traditional systems of medicine are practiced in India, they are Siddha, Ayurveda and Unani. Siddha system of healing and medication is more popular in south India used for many chronic ailments [12].

MATERIALS AND METHODS

Chemicals

CCl4 were purchased from Sigma–Aldrich Chemicals, USA. Trichloroacetic acid, hydroxyproline, HCl, chloramine-T and p-dimethylamino benzaldehyde were purchased from Sigma–Aldrich Chemicals, USA. All other chemicals were of the highest purity and analytical grade. CCl4 was diluted 2:3 ratio with olive oil.

Drug preparation and dose determination

TSF is a combination of seven botanicals include Sphagneticola calendulacea whole plant, Phyllanthus amarus whole plant, Terminalia
Animals

Institutional animal ethical committee (IAEC) at Saveetha University approved the experimental protocol of this study (SU/BRILAC/ RD/019/2014). Male albino Wistar rats (150-180 g) were obtained from the king Institute of preventive medicine, Chennai, India. All rats were housed in polyethylene cages under controlled laboratory condition of temperature 20-22 °C with 12 h dark and light cycle and provided with standard rat pellet and water, ad libitum.

Experimental design

After one week acclimatization period rats were randomly assigned into four groups (n = 6 per group). Group 1 (Normal control) was allowed free access to food and water for eight weeks. Group 2 (Model control) group received intra peritoneal injections of 1 ml/kg CCl4 and olive oil [2:3 v/v] mixture twice a week for eight weeks as described by Gup et al. [16]. Group 3 (Therapeutic model) received CC14 as per group 2 followed by TSF orally 400 mg/kg body weight daily from 5th week to the end of the experiment. Group 4 (Drug control) rats received TSF 400 mg/kg body weight daily for eight weeks. At the end of the experimental period, all the rats were anesthetized by ketamine hydrochloride intravenously 30 mg/kg body weight. A sample of liver and blood were collected for further analysis.

Determination of serum biomarker for liver damage

Serum level of Alanine Transaminase, Aspartate Transaminase, total protein, albumin, total bilirubin and total protein and albumin were measured using commercial kit available (BioSystems Diagnostics Pvt Ltd) according to manufacturer's instruction.

Hydroxyproline content in liver tissue

100 mg of liver tissue was taken for hydroxyproline determination according to Woessner, J. F 1961. 100 mg of tissue was homogenised with 100 ml of 5% trichloroacetic acid and kept at 90°C for 30 min to extract protein, DNA and collagen. The solution was filtered and the filtrate was used for estimation. Aliquots of the 5% trichloroacetic acid extract were hydrolyzed by adding hydrochloric acid to a final concentration of 6N in sealed tubes for 3 h at 150°C. After hydrolysis the sample was evaporated to dryness the residue was dissolved in water and made up to a known volume. Hydroxyproline oxidation was initiated by adding 1.0 ml chloramines-T to each tube, the tube content was mixed by shaking a few times and allowed to stand for 20 min at room temperature. The chloramines-T was then destroyed by adding 1.0 ml Perchloric acid to each tube. The content was mixed and allowed to stand for 5 min. Finally 1.0 ml p-dimethyl amino benzaldehyde solution was added and the mixture was shaken. The tubes were placed in a water bath for 20 min, then cooled with tap water for 5 min. The color developed was read on a Chemito UV spectrophotometer at 557 nm.

Histological examination

The liver specimen was fixed in 10% formalin solution, dehydrated with graded alcohol, embedded in paraffin, cut 5-micron thickness section and stained with hematoxylin and eosin. Histopathological changes of each liver slide were examined under a light microscope (Labomed). For collagen deposition, fixed section was stained with Masson trichrome using a standard protocol.

Statistical analysis

The data were expressed as the mean±SD for the six rats in each group. Statistically significant differences between the groups were calculated using SPSS. The value of p<0.05 was considered to the statically significant.

RESULTS

Effect of TSF on body weight

Alteration in the body weight and liver weight in the control and CCl4 intoxicated groups monitored during the progression of CCl4-induced liver fibrosis. Body weight and liver weight of the rats significantly decreased (p<0.01) in CCl4 treated rats, and significantly increased (p<0.01) in control and CCl4+TSF treated group shown in table 1.

Effect of TSF on liver function

The curative effect of TSF on biochemical markers of liver fibrosis in CCl4 induced rats showed in table 1. The rats treated with CCl4 had significantly (p<0.01) elevated levels of serum AST, ALT, ALP, LDH, YGT, Total bilirubin and significantly reduced (p<0.01) the level of total protein and albumin compared to the normal group. However, treatment with TSF after CCl4 intoxication significantly (p<0.01) reduced the elevated level of AST, ALT, ALP, LDH, YGT and Total bilirubin to near normal level and significantly increased (p<0.01) the total protein and albumin to normal level shown in table 1.

Table 1: Effect of TSF on liver function parameters in CCl4 induced rats

| Parameter       | Control group | Induced group | Therapeutic group | TSF control |
|-----------------|---------------|---------------|-------------------|-------------|
| Body Weight     | 243.3±8.16    | 169.1±5.84a   | 211.6±7.52a       | 253.3±6.05  |
| Liver Weight    | 7.3±0.16      | 5.8±0.25b     | 6.8±0.22c         | 7.8±0.17    |
| ALT             | 52.96±0.95    | 183.8±0.24a   | 64.8±1.48b        | 53.6±0.85   |
| AST             | 117.8±6.13    | 264.9±3.54a   | 136.2±3.68b       | 118.4±5.65  |
| ALP             | 111.6±3.07    | 297.7±6.46a   | 132.2±8.30b       | 103.8±6.70  |
| Total Protein   | 6.65±0.35     | 4.95±0.16c    | 5.76±0.15c        | 6.1±0.17    |
| Albumin         | 4.13±0.13     | 2.90±0.10d    | 3.73±0.24c        | 4.1±0.20    |
| Total Bilirubin | 0.50±0.04     | 1.31±0.07f    | 0.87±0.04c        | 0.5±0.44    |
| Y-GT            | 5.95±0.38     | 26.63±3.41f   | 12.81±1.74d       | 6.1±0.28    |
| HydroxyProline  | 3.46±0.16     | 17.76±0.55g   | 6.05±0.76f        | 3.1±0.27    |
| LDH             | 1150.6±50.85  | 2061.7±47.77  | 1451.6±49.56      | 11458.0±48.56 |

Values are expressed as mean±SD. (n=6). Statistical significance at p<0.05. Comparisons are made with ‘a’ control group and ‘b’ induced group. p<0.05, “p<0.01. Rat body weight and liver weight are expressed in grams. AST, ALT, ALP, LDH and y-GT activities are expressed as IU/l. Total protein and albumin are expressed in g/dl total bilirubin are expressed in mg/dl and hydroxyproline are expressed in the µg/g tissue.
Effect of TSF on hepatic hydroxyproline content

Hepatic hydroxyproline content was significantly (p<0.01) increased in the CCl4 group compared with the control group. Compared with the CCl4 group, the levels of hydroxyproline were significantly (p<0.01) decreased in the treatment group shown in Table 1.

Effect of TSF on histopathological examination

Control rats showed the normal histological appearance (Fig. 1A) whereas CCl4-induced group showed many pathological changes include infiltration of lymphocyte, portal inflammation (Fig. 1D), hepatocyte ballooning and hepatocyte fathery degeneration changes in both portal tract (Fig. 1B) and around the central vein (Fig. 1C), whereas Treatment with TSF groups displayed completely reversed the pathological changes of liver extensively when compare to CCl4 groups and almost similar to normal group (Fig. 1F).

Fig. 1: Microscopic examination of the liver in control and experimental groups determined by hematoxylin and eosin (original magnification 400x liver section in rats) staining is presented in A–H: (A): Control; (B): CCl4 control liver showing degenerative changes of hepatocyte in the periportal region (arrow); (C): CCl4 intoxicated liver showing hepatocyte degeneration around central vein (arrow mark); (D): Inflammatory cell infiltration in periportal region; (E): CCl4+TSF treated liver showing few inflammatory cell inflammation (Arrow mark); (F): CCl4+TSF treated liver showing a normal architecture with granulated cytoplasm; (G): TSF treated significantly ameliorated these pathological changes and exhibited normal architecture; (H): TSF control group showing the normal architecture of liver cells

TSF treatment decreases the infiltration of the inflammatory cell shown in Fig. 1E. The group treated with TSF showed a less destruction of the liver architecture without fibrosis shown in Fig. 1G. Masson trichrome staining with liver section exhibited collagen deposition increased in CCl4 group compared to control group shown in Fig. 2B. Treatment with TSF resulted in a reduced the degree of fibrosis appearance shown in Fig. 2C.

DISCUSSION

In the present study, TSF exhibited a curative effect, as demonstrated by a significant reduction in ALT and AST level and also confirmed by histopathological changes in the liver of rats treated with CCl4 induced liver fibrosis. Hepatotoxicity of CCl4 is attributed to free radical generation during its metabolism in the endoplasmic reticulum. The free radical thus formed can attack lipid on the membrane of the endoplasmic reticulum to elicit lipid peroxidation [17], disturbs cell homeostasis which finally result in cell death. This leads to hepatocyte damage and release of intracellular enzymes like AST and ALT [18]. Zimmerman et al. reported that the CCl4 increase the serum ALT and AST level by the mechanism of the membrane and mitochondrial damage [19]. The present study demonstrates that TSF administration attenuates oxidative stress, decreased inflammatory cell infiltration and fibrosis in liver treated with CCl4, evidenced by complete reversal of hepatic serum biomarkers, this indicate TSF have a strong membrane stabilizing activity so it prevent the leakage of intracellular enzyme to serum and also TSF possibly have to rejuvenate the hepatocyte and healing of liver parenchyma. The level of serum bilirubin may be found in hepatitis, hepatocellular damage and hemolytic jaundice [20]. CCl4 because extensively hepatocellular degeneration and blockage of a bile duct resulted in an increased total bilirubin level. The TSF treatment group had a significant decrease in total bilirubin level compared to untreated groups. Another marker for liver cell damage ALP (synthesized in the hepatocyte canalicul membrane) was found to be elevated in CCl4 induced liver fibrosis [21]. Serum ALP activity acts as a sensitive indicator of obstructive and space occupying lesion in the liver. This elevation is significantly suppressed by the supplementation of traditional formulation which reveals TSF have a membrane remodelling activity and clear the obstruction in hepatic biliary track, which will result if the ALP activity is decreased. Total protein and albumin produced by liver cell were decreased in CCl4 intoxicated rats. This clearly indicates an impaired synthesis of the liver cell due to liver injury [22]. The TSF treated rats showed an increase in total protein and albumin level in serum which indicates a recovery of liver damage.
content in liver tissue is increased in CCl₄ group due to excessive collagen deposition because scar formation disrupts architectural changes in the liver. Collagen is a sensitive indicator which reflects the stage of fibrosis in fibrous liver [23]. Our Masson trichrome staining results indicate TSF treated group significantly decreased the collagen content level in liver tissue. This reveals TSF has a strong anti-fibrotic activity and capacity to inhibit collagen deposition and degradation compared to CCl₄ induced group. In addition, above biochemical changes in liver function correlate with the histopathological changes observed from the microscopic examination of TSF treated rats. Lymphocyte infiltration in the portal areas, hepatocyte feathery degenerative changes in both portal track and central vein region were found in CCl₄ intoxicated rats. These histopathological changes were reversed to normal architecture by TSF treated rats. Hence our TSF is found to be a potent drug to treat liver fibrosis.

CONCLUSION

In conclusion, we showed that TSF significantly inhibited CCl₄ induced hepatic fibrosis, and its anti-fibrotic activity may be associated due to the spectrum of synergistically active phytochemicals. This natural Siddha formulation could be developed as a promising effective therapeutic drug for liver fibrosis. However, further analytical studies may be required to support the above conclusion.

CONFLICTS OF INTERESTS

All authors have none to declare

REFERENCES

1. Friedman SL. Hepatic stellate cells: protein, multifunctional, and endocrine cells of the liver. Physiol Rev 2008;88:125–72.
2. Gines P, Cardenas A, Arroyo V, Rodes J. Management of cirrhosis and ascites. N Engl J Med 2004;350:1646–54.
3. Friedman SL. Liver fibrosis: from bench to bedside. J Hepatol 2003;38:S58–53.
4. UE Lee, SL Friedman. Mechanisms of hepatic fibrogenesis. Best Practice Res Clin Gastroenterol 2011;25:195–206.
5. Weiler-Normann C, Herkl J, Lohse AW. Mouse models of liver fibrosis. Z Gastroenterol 2007;45:45–50.
6. Hamdy N, EI-Demerdash E. New therapeutic aspect for carvedilol: anti-fibrotic effects of carvedilol in chronic carbon tetrachloride-induced liver damage. Toxicol Appl Pharmaco 2012;261:292–9.
7. Shi J, Aisaki K, Ikawa Y, Wake K. Evidence of hepatocyte apoptosis in rat liver after the administration of carbon tetrachloride. Am J Pathol 1999;153:515–25.
8. Malang HM, Eichelbaum FM, Saul W, Sipos IG, Brown EA, Gillette JR. Nature of the protection against carbon tetrachloride-induced hepatotoxicity produced by pretreatment with dibenamine [N-(2-chloroethyl) dibenzylamine]. Biochem Pharmaco 1974;23:1479–91.
9. Valko M, Rhodes CJ, Moncol J, Izakovic M, Mazur M. Free radicals, metals and antioxidants in oxidative stress-induced cancer. Mini-review. Chem Biol Interact 2006;160:1–40.
10. Abdelaziz DHA, Ali SA. The protective effect of Phoenix dactylifera L. seeds against CCl₄-induced hepatotoxicity in rats. J Ethnopharmacol 2014;155:736–43.
11. Halliwell B, Gutteridge JMC. Free radicals in biology and medicine. 3rd edition. Oxford, UK: Clarendon Press; 1999.
12. Beckman KB, Ames BN. The free radical theory of aging matures. Physiol Rev 1998;78:547–81.
13. Methewsaran S, Pandikumar P, Chellaappandian M, Ignacimuthu S, Duraiappadhandi V, Logamanian M. Consensus analysis of sastric formulations used by non-institutionally trained siddha medical practitioners of Virudunagar and Tirunelveli districts of Tamil Nada, India J Ethnopharmacol 2014;153:290–6.
14. Sri kamatchi priya Ramamoorthy, Diwakar Manickam, Samu subramaniam, Shyama Subramaniam. Standardisation and phytochemical screening of traditional formulation. Int J Curr Pharm Res 2017;9:70–4.
15. Vaiidya Ratnam KS, Murugesu Mudaliar. Gunapadam. 1st edition. (Siddha Materia Medica); 1936. p. 229, 345, 459, 520, 720.
16. Guo-Yu Li, Hong-Ying Gao, Jian Huang, Jin Lu, Jing-Kai Gu, et al. Hepatoprotective effect of Cichorium intybus L traditional Ughur medicine, against carbon tetrachloride-induced hepatic fibrosis in rats. World J Gastroenterol 2014;20:4753–60.
17. Fiorucci S, Antonelli E, Distruiti E, Severino B, Fiorentina R, Baldoni M, et al. PAR1 antagonism protects against experimental liver fibrosis. Role of proteinase receptors in stellate cell activation. Hepatology 2004;39:365–75.
18. Shahjahan M, Sabitha KE, Jaimi M, Shyamala Devi CS. Effect of Solanum trilobatum against carbon tetrachloride induced hepatic damage in abino rats. Indian J Med Res 2004;120:194–8.
19. Zimmerman HJ, Kodera Y, West M. Effects of carbon tetrachloride. Poisoning on the plasma levels of cytoplasmic and mitochondrial enzymes in animals with nutritional fatty metamorphosis. J Lab Clin Med 1965;66:324–33.
20. Murray RK, Porphyris and bile pigments. In: Murray RK, Ganner DK, Mayes PA, Rodwell VW. editors. Harper’s illustrated biochemistry. 26th Ed. New York: McGraw-Hill Inc; 2003. p. 270–85.
21. Khan AA, Alahairy M. Hepatoprotective effects of camel milk against CCL4-induced hepatotoxicity in Rats. Asian J Biochem 2011;6:171–80.
22. Smyth R, Munday MR, York MJ, Clarke CJ, Dare T, Turton JA. Comprehensive Characterization of serum clinical chemistry parameters and the identification of urinary superoxide dismutase in a carbon tetrachloride-induced model of hepatic fibrosis in the female Hanover Wistar rat. Int J Exp Pathol 2007;98:361–76.
23. Wang Y, Cheng M, Zhang B, Nie F, Jiang H. Dietary supplementation of blueberry juice enhances hepatic expression of metallothionein and attenuates liver fibrosis in rats. PLoS One 2013;8:e85909.

How to cite this article

- Diwakar Manickam, Sri Kamatchi Priya Ramamoorthy, B. Santhosh Kumar, Samu Subramaniam, Shyama Subramaniam. Biochemical and histopathological changes of traditional Siddha formulation on CCl₄ induced liver fibrosis in rats. Int J Pharm Pharm Sci 2017;9(7):120–123.