Draft Genome Sequence of Pandrug-Resistant *Pseudomonas aeruginosa* SPA03, Isolated from a Patient with Benign Prostatic Hyperplasia

Chanakya Pachi Pulusu, a Balaram Khamari, a Manmath Lama, a Arun Sai Kumar Peketi, a Prakash Kumar, b Valakunja Nagaraja, c,d @Eswarappa Pradeep Bulagonda

aDepartment of Biosciences, Sri Sathyai Sai Institute of Higher Learning, Puttaparthi, India
bDepartment of Microbiology, Sri Sathyai Sai Institute of Higher Medical Sciences, Prasanthigram, India
cDepartment of Microbiology and Cell Biology, Indian Institute of Science, Bangalore, India
dJawaharlal Nehru Centre for Advanced Scientific Research, Bangalore, India

**ABSTRACT** The draft genome of pandrug-resistant *Pseudomonas aeruginosa* strain SPA03, which belongs to global high-risk sequence type 357 (ST357) and was isolated from a patient with benign prostatic hyperplasia, is presented in this report. The genome assembly was generated by combining short-read Illumina HiSeq-X Ten and long-read Oxford Nanopore Technologies MinION sequence data using the Unicycler assembler.

*Pseudomonas aeruginosa*, a member of the WHO priority pathogen list (1), is an opportunistic human pathogen that infects vulnerable patients (2, 3). Urinary tract infections (UTIs) due to *P. aeruginosa* are often associated with increased morbidity and mortality rates (4). Although several genomes of *P. aeruginosa* are available in the GenBank database, the emergence of pandrug-resistant (PDR) variants belonging to global high-risk clone sequence type 357 (ST357) warrants the sequencing of additional PDR clinical isolates.

Here, we present a draft genome sequence of PDR *P. aeruginosa* strain SPA03, which was isolated from the catheter urine of a 77-year-old male patient with benign prostatic hyperplasia. Ethical approval was obtained from the Sri Sathyai Sai Institute of Higher Learning Institutional Ethics Committee (approval number SSSIHL/IEC/PSN/BS/2014/03). A clean urine aspirate sample from the urinary catheter was cultured on sheep blood agar (HiMedia) for 24 h at 37°C. The isolate was identified as *P. aeruginosa* with matrix-assisted laser desorption ionization–time of flight mass spectrometry (MALDI-TOF MS) (bioMérieux), and this was confirmed by ribosomal multilocus sequence typing (MLST) (5). The strain was found to be PDR, exhibiting resistance to all tested antibiotics (Table 1), by Vitek 2 (AST N281 card).

Genomic DNA extracted with the Macherey-Nagel NucleoSpin DNA extraction kit from the culture of a single colony grown overnight in LB broth at 37°C was used for the preparation of an Illumina (paired-end) sequencing library and a Nanopore sequencing library (prepared using sheared DNA) with the NEBNext Ultra II DNA library preparation kit (E7645S) and the Nanopore ligation sequencing kit (SQK-LSK109), respectively. Quality and quantity were assessed using a Qubit 2 fluorometer and an Agilent Bioanalyzer DNA 1000 kit. An Illumina HiSeq-X Ten system was used to sequence the libraries, and demultiplexing was performed using bcl2fastq v2.2. A total of 4,722,619 read pairs were generated using 2 × 150-bp chemistry. Adapter sequences were removed using fastp v0.20.1 (6). Nanopore sequencing was performed with a FLO-MIN106 flow cell on a MinION device (Oxford Nanopore Technologies). Base calling and demultiplexing were performed using Albacore v2.0.1, and adapter sequences
were removed using Porechop v0.2.4 with default settings. The MinION sequencing run generated 90,337 reads, with an average length of 4,961 bp and a mean quality score of 8.9 (NanoStat) (7). The processed reads from Nanopore and Illumina sequencing were used to generate a hybrid de novo assembly using Unicycler v0.4.8 (8). Final assembly quality was assessed with QUAST (9). The SPA03 genome was annotated using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) v5.0 (10). Default parameters were used for all software unless otherwise specified.

The hybrid genome assembly consists of 9 contigs, with a total length of 6,829,266 bp (N_{50}, 5,225,585 bp; N_{75}, 5,225,585 bp), a GC content of 65.9%, and genome coverage of 116.13 x. A total of 6,307 genes were predicted by the NCBI PGAP, including 6,153 protein-coding sequences, 9 rRNAs, 62 tRNAs, 4 noncoding RNAs, and 79 pseudogenes. MLST analysis by MLST v2.0 (11) and serotyping by Past v1.0 (12) identified that the study genome belongs to ST357 and serogroup O11. ResFinder v4.1 (13) and CARD v3.1.1 (14) analyses revealed the presence of antibiotic resistance genes for aminoglycosides, β-lactams, fluoroquinolones, fosfomycin, chloramphenicol, rifampin, sulfonamides, and tetracyclines, along with multiple efflux pump genes.

In summary, we report the draft genome of PDR *P. aeruginosa* SPA03, which belongs to high-risk clone ST357. Early identification of such pathogens could help in investigating outbreaks and controlling infections. The whole-genome sequencing data for PDR clinical isolates would serve as a useful reference for genome assemblies and comparator strains for further research.

**Data availability.** The whole-genome sequence data and raw sequence data for *P. aeruginosa* strain SPA03 are available at NCBI under BioProject PRJNA689041, GenBank accession number GCA_016595165.1, and Sequence Read Archive (SRA) accession numbers SRX10461636 (Illumina raw sequence data) and SRX10461637 (MinION raw sequence data).

**ACKNOWLEDGMENTS**

We thank the Department of Mathematics and Computer Sciences, Sri Sathya Sai Institute of Higher Learning (SSSIHL), for access to the high-performance computing facility. This project was supported by an ICMR EMR grant (OMI/27/2020-ECD-I).

We acknowledge UGC-SAP-DRS-III, DST-FIST, and DBT-BIF, Government of India, for infrastructural support to the Department of Biosciences, SSSIHL, and ICMR-SRF, UGC-SRF, and NFST Fellowships from the Government of India to C.P.P., B.K., and M.L., respectively. V.N. is a J. C. Bose Fellow of the Department of Science and Technology, Government of India.

---

**TABLE 1 Antimicrobial susceptibility profile of *P. aeruginosa* SPA03**

| Antimicrobial            | MIC (μg/ml) | Resistance profile |
|--------------------------|-------------|--------------------|
| Ticarcillin-clavulanic acid | ≥128        | R                  |
| Piperacillin-tazobactam  | ≥128        | R                  |
| Ceftazidime              | ≥64         | R                  |
| Ceferazone-sulbactam     | ≥64         | R                  |
| Cefepime                 | ≥64         | R                  |
| Doripenem                | ≥8          | R                  |
| Imipenem                 | ≥16         | R                  |
| Meropenem                | ≥16         | R                  |
| Amikacin                 | ≥64         | R                  |
| Gentamicin               | ≥16         | R                  |
| Ciprofloxacin            | ≥4          | R                  |
| Levofloxacin             | ≥8          | R                  |
| Minocycline              | ≥16         | R                  |
| Tigecycline              | ≥8          | R                  |
| Colistin                 | 4           | R                  |
| Trimethoprim-sulfamethoxazole | ≥320       | R                  |

*a* Antibiotic susceptibility testing was performed using the Vitek 2 system according to Clinical and Laboratory Standards Institute (CLSI) guidelines (15).

*b* R, resistant.
REFERENCES

1. Tacconelli E, Carrara E, Savoldi A, Harbarth S, Mendelson M, Monnet DL, Pulcini C, Kahlemer G, Kluytmans J, Carmeli Y, Ouellette M, Outterson K, Patel J, Cavalieri M, Cox EM, Houchens CR, Grayson ML, Hansen P, Singh N, Theuretzbacher U, Magrini N. 2018. Discovery, research, and development of new antibiotics: the WHO priority list of antibiotic-resistant bacteria and tuberculosis. Lancet Infect Dis 18:318–327. https://doi.org/10.1016/S1473-3099(17)30753-3.

2. De Bentzmann S, Plésiat P. 2011. The Pseudomonas aeruginosa opportunistic pathogen and human infections. Environ Microbiol 13:1655–1665. https://doi.org/10.1111/j.1462-2920.2011.02469.x.

3. Dricoll JA, Brody SL, Kollef MH. 2007. The epidemiology, pathogenesis and treatment of Pseudomonas aeruginosa infections. Drugs 67:351–368. https://doi.org/10.2165/00003495-200767030-00003.

4. Lamas Ferreiro JL, Álvarez Otero J, González González L, Novoa Lamazares L, Arca Blanco A, Bermúdez Sanjurjo JR, Rodríguez Conde I, Fernández Someira M, de la Fuente Aguado J. 2017. Pseudomonas aeruginosa urinary tract infections in hospitalized patients: mortality and prognostic factors. PLoS One 12:e0178178. https://doi.org/10.1371/journal.pone.0178178.

5. Jolley KA, Bliss CM, Bennett JS, Bratcher HB, Brehony C, Colles FM, Wimalarathna H, Harrison OB, Sheppard SK, Cody AJ, Maiden MCJ. 2012. Ribosomal multilocus sequence typing: universal characterization of bacteria from domain to strain. Microbiology (Reading) 158:1005–1015. https://doi.org/10.1099/mic.0.055459-0.

6. Chen S, Zhou Y, Chen Y, Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ pre-processor. Bioinformatics 34:i884–i890. https://doi.org/10.1093/bioinformatics/bty560.

7. De Coster W, D’Hert S, Schultz DT, Cruts M, Van Broeckhoven C. 2018. NanoPack: visualizing and processing long-read sequencing data. Bioinformatics 34:2666–2669. https://doi.org/10.1093/bioinformatics/bty149.

8. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. https://doi.org/10.1371/journal.pcbi.1005595.

9. Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. Bioinformatics 29:1072–1075. https://doi.org/10.1093/bioinformatics/btt086.

10. Tatusova T, DiCuccio M, Badetdin A, Chetverin V, Nawrocki EP, Aslalovsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. https://doi.org/10.1093/nar/gkw569.

11. Larsen MV, Cosentino S, Rasmussen S, Friis C, Hasman H, Marvig RL, Jelsbak L, Sicheritz-Pontén T, Ussery DW, Aarestrup FM, Lund O. 2012. Multilocus sequence typing of total-genome-sequenced bacteria. J Clin Microbiol 50:1355–1361. https://doi.org/10.1128/JCM.00944-11.

12. Thrane SW, Taylor VL, Lund O, Lam JS, Jelsbak L. 2016. Application of whole-genome sequencing data for O-specific antigen analysis and in silico serotyping of Pseudomonas aeruginosa isolates. J Clin Microbiol 54:1782–1788. https://doi.org/10.1128/JCM.00349-16.

13. Bortolaia V, Kaas RS, Ruppe E, Roberts MC, Schwarz S, Cattoir V, Philippin A, Allesoe RL, Rebelo AR, Florensa AF, Fagelhauer L, Chakraborty T, Neumann B, Werner G, Bender J, Stigl K, Nguyen M, Coppenes J, Xavier BB, Malhotra-Kumar S, Westh H, Pinholt M, Anjum MF, Duggett NA, Kempf F, Nykäsenoja S, Olliola S, Wiekorek K, Amaro A, Clemence L, Mossong J, Losch R, Ramigau C, Lund O, Aarestrup FM. 2020. ResFinder 4.0 for predictions of phenotypes from genotypes. J Antimicrob Chemother 75:3491–3500. https://doi.org/10.1093/jac/dkaa345.

14. Alcock BP, Raphenya AR, Lau TTY, Tsang KK, Bouchard M, Edalatmand M, Huynh N, Nguyen AV, Cheng AA, Liu S, Min SY, Miroshnichenko A, Tran HK, Werfalli RE, Nasir JA, Oloni M, Speicher DJ, Florescu A, Singh B, Faltynek M, Hernandez-Koutoucheva A, Sharma AN, Bordeleau E, Pavlovski AC, Zubyk HL, Dooley D, Griffiths E, Maquire F, Winsor GL, Beiko RG, Brinkman FSL, Hisao WWL, Domelaar GV, McArthur AG. 2020. CARD 2020: antibiotic resistance surveillance with the Comprehensive Antibiotic Resistance Database. Nucleic Acids Res 48:DS17–DS25. https://doi.org/10.1093/nar/gkz935.

15. Clinical and Laboratory Standards Institute. 2020. Performance standards for antimicrobial susceptibility testing, 30th ed. CLSI supplement M100. Clinical and Laboratory Standards Institute, Wayne, PA.