Original Research Article

Effect of Sodium Substitution on Lactic Acid Bacteria and Total Bacterial Population in Mango Pickle

Arghya Mani¹*, Prodyut Kumar Paul² and Ivan Wilson³

¹Department of Post-harvest Technology, BCKV, Mohanpur, India
²Department of Pomology and Post-harvest Technology, UBKV, Coochbehar, India
³Warner School of Food and Dairy, SHUATS, Allahabad, India

*Corresponding author

ABSTRACT

An attempt has been made to prepare mango pickle using different salt compositions of NaCl, KCl and CaCl₂ and evaluate the pickle samples for total bacterial population and Lactic acid bacteria populations. The purpose of salt mixture was to reduce Na consumption. The results showed that the total bacterial population tends to increase with storage. Minimum bacterial population was observed at 0 day and maximum at 210th day of storage. Among different treatments, T₁ and T₂ showed maximum bacterial population of 164 and 160 respectively, whereas minimum bacterial colonies was observed in T₅ (147.67). It clearly indicates that CaCl₂ has a negative influence on bacterial population. The Lactic Acid Bacteria population shows a reverse trend and tends to decline with storage time. At 0 day, T₁ shows highest population of LAB (162.67) followed by T₂ (139.67), T₅ (137.67) and T₄ (135.67); whereas T₅ showed lowest LAB population (114.33). At 210th day, T₅ showed highest LAB population (50.67) followed by T₁ (47). T₅ supported lowest LAB population (25) followed by T₃ (30.33), T₂ (32) and T₄ (34). This clearly shows that NaCl and KCl have positive influence on LAB population. A conclusion can be drawn from this that the incorporation of CaCl₂ in the salt mixture which is to be used for curing purpose directly helps to suppress the bacterial population even at 210th day (7 month) of storage. But it also affects the growth of desirable Lactic Acid Bacteria (LAB) in the pickle. Hence, a salt mixture with 50% NaCl, 25% KCl and 25% CaCl₂ can be used for pickle preparation.

Keywords
Lactic acid bacteria,
Salt substitution,
Bacterial population,
LAB.

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Introduction

Mango pickle is one of the oldest preserved food products which is made from unripe mango. The term pickle is derived from the Dutch word ‘Pekel”, meaning brine (Wikipedia, 2017). Pickles are made through the natural fermentation of fruits and vegetables, and besides having nutritional value, pickles also act as a food accompaniments and palatability enhancers (Saviti and Bhill, 2007). The process of pickling involves fermentation which is a primitive preservation method primarily used to enable the long-term storage of foods. Fermentation is a slow decomposition process of organic substances induced by microorganisms or enzymes that essentially convert carbohydrates to alcohols or organic acids (FAO, 1998). When the fermentation
term is used in case of fruits and vegetables, it is known as pickling. Out of the various approaches to fermentation, lactic acid fermentation, using natural micro flora or lactic acid bacterial (LAB) cultures, is employed throughout the world. Lactic acid (LA) fermentation of vegetables and fruits is a common practice to maintain and improve the nutritional and sensory features of food commodities (Cagno et al., 2013; Karovicova and Kohajdova, 2003). Salt is an indispensable part of our food habit. Salts not only improve the taste, but it also has a big role in human nutrition. NaCl is one of the most commonly employed agents for food conservation, allowing considerable increase in storage time by reducing water activity. Salt (sodium chloride) is the oldest food seasoning, which provides one of the important basic human tastes (saltiness) and preserves foods to extend the shelf life. Salt mainly consists of two elements: sodium and chloride. In the pickling industry, salt has historically been used for directing the fermentation of cucumbers, radishes, and carrots (Thompson et al., 1979; Hudson and Buescher, 1985; Fleming et al., 1995; Mcfeeters et al., 1996). Common salt contains Na⁺ (Cation) and Cl⁻ (Anion). Na⁺ (Cation) is mainly responsible for the saltiness in the food. Sodium is a vital element required in small amounts by the human body, as it helps to control homeostasis and nerve impulses (Starr and McMillan, 2006). Sodium chloride is an essential in food as it improves the preservative, technological and sensory quality of food (Brady, 2002). The extra intake of sodium present in salt might lead to conditions such as hypertension and high blood pressure. Approximately one quarter of the world’s population suffers from this condition (WHO, 2011). High sodium intake is increasing the risk of heart attack and high blood pressure (Doyle, 2008). Results for sodium intake and its effects on human blood pressure were derived from scientific research, animal studies and other human surveys (Doyle, 2008; Kesteloot and Joossens, 1988; Meneton et al., 2005). The mechanism of the effect of salt on blood pressure could be due to the rise in plasma sodium or to the increase in extracellular fluid volume. Higher dietary sodium intake is also related to bone disease (Doyle, 2008). Pickling is done in presence of high concentration of salt solution in which the fruit pieces are dipped to ensure fermentation. Pickles contain salt at about 15-20% levels making it one of the high salt containing foods. The biggest drawback with pickles is the presence of high concentration of sodium ion (Na⁺) which may lead to adverse effects on human health and on food business. Many food products have been launched in its low salt version. The only form of salt that does not contain sodium is the low-sodium alternatives are the replacement with Potassium, Magnesium and Calcium ion instead of sodium. Eating excess salt raises the amount of sodium in our bloodstream and disturbs the delicate balance, reducing the ability of our kidneys to remove the water. The partial substitution of NaCl by KCl or CaCl₂ seems to provide an alternative for reducing sodium content. Increased potassium intake is reported to protect stroke, high blood pressure, heart rhythm problems, kidney failure, and even osteoporosis (Hall, 2003). The additional use of KCl and CaCl₂ to partially replace NaCl could be helpful in reducing sodium content (Gillette, 1985). However, the use of KCl is mainly limited by its bitter and astringent taste (Reddy and Marth, 1991). Some people have reported a metallic after taste and therefore choose not to use KCl in food. But a mixed concentration of Na, K and Ca can help to reduce the total salt intake in our body. Therefore, the present work has been undertaken to investigate the possibility of replacing sodium chloride by potassium and Calcium salts and develop low sodium mango pickle, to study the effect of
partial NaCl substitution on processing parameters of pickles, to study the effect of NaCl substitution of sensory properties of pickle and to optimize salt mixture components for low sodium mango pickles without affecting its physiochemical, biochemical, microbiological and sensory qualities.

Materials and Methods

Site of experiment

The present study was conducted in Post Graduate Laboratory, Department of Pomology and Post-Harvest Technology, Faculty of Horticulture and Central Instrumentation Centre Lab of Uttar Banga Krishi Viswavidyalaya, Pundibari, Coochbehar, West Bengal.

Source of pickling materials

The fruits were fresh, unripe and were free from pests, diseases and blemishes. Fazli variety was procured from local market since it is known to have high acid content. The chemical purchased were of Laboratory grade.

Design for deciding the salt mixture for pickle preparation in the experiment

The following design was used for deciding the salt mixture for pickle preparation in the experiment.

Design: Randomized Block Design (RBD)
Software used: SPSS

Procedure for mango pickle preparation

Pickles were prepared by using the standardized procedure. The prepared pickles were stored in glass jars which were cleaned properly and were sterilized in boiling water at room temperature. During the entire storage period it was ensured that the pickle was stored in aerated, dry and hygienic conditions.

Lactic acid bacteria and total plate count upto 210th day of pickle storage

The entire microbiological aspects of the experiment were performed in Post Graduate Lab, Department of Plant Pathology, UBKV. Microbiological analysis for the pickle was carried out by the method of Ranganna (1977). All the enumerations of Bacteria and Lactobacillus were carried out following serial dilution technique using specific media. Plates were incubated at 34±1 °C for 48 hours and colony forming units (CFU/g) were recorded. Observations for microbial count were made at prescribed intervals. The principle behind this is that the population of total bacterial population tends to decline with the decimal reduction in the concentration of the sample analyzed. Usually in a culture the microbial population was expected to be higher in $10^{-1}$ which tends to decline with $10^{-2}$, $10^{-3}$, $10^{-4}$, $10^{-5}$ and $10^{-6}$ sample concentration. Higher the sample concentration higher would be the expected microbial concentration. As the colonies tends to coalesce or merge at higher concentration hence the readings for Total Plate Count and Total Bacterial Population Count has been studied at $10^{-5}$ concentration.

Results and Discussion

Table 2 shows the effect of different salt proportion on Lactic Acid Bacteria population in the mango pickle sample which has been studied upto 210th day of preparation. Varying proportion of NaCl, KCl and CaCl$_2$ has been used for the curing purpose. At the 0 day of storage, T$_1$ shows highest population of Lactic acid bacteria (162.67) followed by T$_2$ (139.67), T$_6$ (137.67) and T$_4$ (135.67) whereas T$_3$ showed lowest LAB population (114.33). This can be attributed due to the fact that KCl and CaCl$_2$ have a negative response on any
sort of microbial growth. At 30th day of storage, T1 have highest LAB population (154) followed by T2 (137.67), T6 (135.33) and T4 (133.33). T3 have lowest LAB population (111) followed by T3 (114.67). Similar trend in the LAB population dynamics has been observed at 60th, 90th, 120th, 150th and 180th day. At 210th day, T6 showed highest LAB population (50.67) followed by T1 (47). T5 supported lowest LAB population (25) followed by T3 (30.33), T2 (32) and T4 (34). At 210th day of storage, LAB population using T1 and T6 salt proportion is at par. This clearly shows that NaCl and KCl have positive influence on LAB population whereas addition of CaCl2 in salt mixture resulted in reduction in LAB population. This can be due to the unique ability of CaCl2 to reduce the water activity of salt cured mango pieces.

Table 3 shows the effect of different salt proportions on the total bacterial population in mango pickle sample at ambient storage up to 210th days. The total plate count showed significant variation of total bacterial population among different treatments. At 0 day, highest bacterial population was observed in T1 (7.67) followed by T2 (7), T4 (6) and T6 (5.33). Lowest bacterial population was observed in T5 (4.33) followed by T3 (5). T1 was at par with T2 which clearly indicates that the presence of higher amount of NaCl and no CaCl2 provides suitable environment for bacteria to survive. Similar trend was observed at 30th, 60th, 90th, 120th, 150th and 180th day of treatment. At 210th day of treatment, the pickle prepared using salt composition T1 showed maximum bacterial colonies (164). High bacterial population was also observed in T2 (160). Comparatively lower bacterial population was observed in T5 (147.67) followed by T6 (150). The reason behind lowest bacterial population in T5 and lower bacterial population at T6 can be attributed to the salt composition that was used during the curing procedure. T5 salt mixture contain 50% CaCl2 because of which the pickle prepared was having lowest bacterial count. CaCl2 is known to be a good curing agent that can substantially reduce the available water in tissue of pickle pieces thus reducing the water activity (aw) and resulting in lower total bacterial population.

**Fig.1** Lactic Acid Bacteria population (log CFU) under ambient storage conditions
Fig. 2 Total Bacterial population (log CFU) under ambient storage conditions

Table 1 Treatment details showing different salt proportion used for curing purpose

| Treatment | NaCl (%) | KCl (%) | CaCl₂ (%) |
|-----------|----------|---------|-----------|
| T1        | 100      | 0       | 0         |
| T2        | 50       | 50      | 0         |
| T3        | 50       | 0       | 50        |
| T4        | 0        | 100     | 0         |
| T5        | 0        | 50      | 50        |
| T6        | 50       | 25      | 25        |

Table 2 Lactic Acid Bacteria population (log CFU) under ambient storage conditions

| Treatments | 0 day | 30th day | 60th day | 90th day | 120th day | 150th day | 180th day | 210th day |
|------------|-------|----------|----------|----------|-----------|-----------|-----------|-----------|
| T1         | 162.67| 154      | 150      | 131.67   | 114       | 93.67     | 74.33     | 47        |
| T2         | 139.67| 137.67   | 135.33   | 119.67   | 102       | 85        | 64.33     | 32        |
| T3         | 119.33| 114.67   | 111.33   | 97.33    | 84        | 77.33     | 58        | 30.33     |
| T4         | 135.67| 133.33   | 130.33   | 103.67   | 89.33     | 79.33     | 64        | 34        |
| T5         | 114.33| 111      | 108      | 94.67    | 80.67     | 72.33     | 56        | 25        |
| T6         | 137.67| 135.33   | 132      | 109.33   | 95.33     | 84        | 71.33     | 50.67     |
| C.D.       | 17.776| 15.795   | 13.335   | 12.231   | 11.526    | 7.229     | 5.201     | 3.314     |
| SE(m)      | 5.569 | 4.949    | 4.178    | 3.832    | 3.611     | 2.265     | 1.63      | 1.038     |
| C.V.       | 7.151 | 6.543    | 5.661    | 6.068    | 6.638     | 4.787     | 4.365     | 4.926     |
**Table 3** Total Bacterial population (log CFU) under ambient storage conditions

| Treatments | 0 day | 30th day | 60th day | 90th day | 120th day | 150th day | 180th day | 210th day |
|------------|-------|----------|----------|----------|-----------|-----------|----------|----------|
| T1         | 7.67  | 18       | 42       | 70.67    | 100.33    | 119.33    | 142.33   | 164      |
| T2         | 7     | 22       | 41       | 68.67    | 95        | 113.67    | 137.67   | 160      |
| T3         | 5     | 20       | 38.33    | 64.33    | 84        | 106.67    | 128.33   | 153      |
| T4         | 6     | 17       | 42.67    | 68.33    | 93.33     | 112.33    | 137      | 155.33   |
| T5         | 4.33  | 22       | 34.67    | 62.67    | 80.67     | 103.67    | 124.33   | 147.67   |
| T6         | 5.33  | 16       | 39.67    | 64       | 86.33     | 110.67    | 130      | 150      |
| C.D.       | 1.637 | 1.99     | 1.332    | 1.046    | 2.621     | 2.808     | 1.893    | 1.748    |
| SE(m)      | 0.513 | 0.624    | 0.417    | 0.328    | 0.821     | 0.88      | 0.593    | 0.548    |
| C.V.       | 15.083| 5.635    | 1.819    | 0.854    | 1.581     | 1.372     | 0.771    | 0.612    |

**Summary**

The mango pickle which was prepared after curing with different salt mixture was evaluated for total bacterial population and lactic acid bacteria (LAB) population. The results showed that the total bacterial population tends to increase with storage. Hence minimum bacterial population was observed at 0 day and maximum at 210th day of storage. Among different treatments, T1 and T2 showed maximum bacterial population of 164 and 160 respectively, whereas minimum bacterial colonies was observed in T5 (147.67). It clearly indicates that CaCl₂ has a negative influence on bacterial population.

The Lactic Acid Bacteria population shows a reverse trend and tends to decline with time. At 0 day of storage, T1 shows highest population of Lactic acid bacteria (162.67) followed by T2 (139.67), T6 (137.67) and T4 (135.67) whereas T5 showed lowest LAB population (114.33). At 210th day, T6 showed highest LAB population (50.67) followed by T1 (47). T5 supported lowest LAB population (25) followed by T3 (30.33), T2 (32) and T4 (34). This clearly shows that NaCl and KCl have positive influence on LAB population.

A conclusion can be drawn from this that the incorporation of CaCl₂ in the salt mixture which is to be used for curing purpose directly helps to suppress the bacterial population even at 210th day (7 month) of storage. But it also affects the growth of desirable Lactic Acid Bacteria (LAB) in the pickle. Hence, a salt mixture with 50% NaCl, 25% KCl and 25% CaCl₂ can be used for pickle preparation. This salt mixture would not only help to minimize Sodium (Na) consumption but can also ensure a balanced microbial population throughout its ambient storage period to satisfactory level.

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