Review Article

The Role of Natural Products as Inhibitors of JAK/STAT Signaling Pathways in Glioblastoma Treatment

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The permeability of glioblastoma, as well as its escaping the immune system, makes them one of the most deadly human malignancies. By avoiding programmed cell death (apoptosis), unlimited cell growth and metastatic ability could dramatically affect the immune system. Genetic mutations, epigenetic changes, and overexpression of oncogenes can cause this process. On the other hand, the blood-brain barrier (BBB) and intratumor heterogeneity are important factors causing resistance to therapy. Several signaling pathways have been identified in this field, including the Janus tyrosine kinase (JAK) converter and signal transducer and activator of transcription (STAT) activator pathways, which are closely related. In addition, the JAK/STAT signaling pathway contributes to a wide array of tumorigenesis functions, including replication, anti-apoptosis, angiogenesis, and immune suppression. Introducing this pathway as the main tumorigenesis and treatment resistance center can give a better understanding of how it operates. In light of this, it is an important goal in treating many disorders, particularly cancer. The inhibition of this signaling pathway is being considered an approach to the treatment of glioblastoma. The use of natural products alternatively to conventional therapies is another area of research interest among researchers. Some natural products that originate from plants or natural sources can interfere with JAK/STAT signaling in human malignant cells, also by stopping the progression and phosphorylation of JAK/STAT, inducing apoptosis, and stopping the cell cycle. Natural products are a viable alternative to conventional chemotherapy because of their cost-effectiveness, wide availability, and almost no side effects.

1. Introduction

Compared to other types of brain tumors, glioblastomas are very aggressive. Glioblastoma is a primary brain tumor that arises from glial tissue and is among the most common and deadly of all brain tumors due to its high permeability, immune system evasion ability, and molecular heterogeneity [1]. Despite many attempts, it has proven to be highly resistant to conventional treatment, and its recurrence has made many treatments seem ineffective. Among the reasons for resistance to treatment are the blood-brain barrier (BBB) as well as intratumor heterogeneity [2, 3]. Abnormal tumor cells have some features that complicate tumors and make them ineffective to treat. Among these are high proliferation
rates, escape mechanisms from the immune system, resistance to programmed cell death (apoptosis), angiogenesis, and maintenance of cell signaling to induce repetitive immortality, invasive activity, and metastasis [4]. If any of the above mechanisms is stopped, there will be advancements in cancer treatment. The biological cell survival abilities of tumors can be caused by genetic mutations, epigenetic changes, and overexpression of oncogenes [5]. Tumor development is caused by many factors, one of the most important factors is the oncogenic signaling pathway. In this field, many signaling pathways have been identified. One of the most important of them is the Janus tyrosine kinase (JAK) converter and transcription signaling (STAT) activator pathway, which is closely related and manages the replication, antiapoptotic, angiogenic, and immunosuppressive functions in tumor microenvironment [6–8]. It is believed that this route is responsible for the main source of tumor formation and resistance to treatments [2]. Therefore, blocking the abilities of this signaling pathway is seen as a potential approach to treat glioblastoma. In addition to conventional treatments, a new approach called “natural products” is being considered by researchers [5, 9]. The therapeutic effects of these bioactive compounds, which are mainly found in plants, have yet to be independently verified in the case of cancer [5, 10–15]. Nevertheless, studies and preclinical data have shown that natural compounds in plants can affect the signaling pathways and proliferation of malignancies in humans [11]. They can also strengthen and sensitize the immune system against cytotoxic agents. Furthermore, studies have found that this method has fewer side effects than conventional chemotherapy methods, and it is less toxic to organs such as the liver, heart, and kidneys [16]. The current research on this topic has been reviewed, and some of these bioactive elements have been excerpted in relation to their ability for affecting JAK/STAT signaling pathways. The purpose of this study is to collect and suggest the preventive factors and possible natural treatments for glioblastoma.

2. JAK/STAT Signaling Pathways and Glioblastoma

JAK/STAT are closely related to each other and together can promote growth, proliferation, survival, inflammation, invasion, blood vessel formation, and progression of multiple tumors [17]. The four main members of JAK are JAK1, JAK2, JAK3, and TYK2. STATs fall into seven major groups: STAT1, STAT2, STAT3, STAT4, STAT5a, STAT5b, and STAT6 [18–20]. The STAT protein, which is dormant in the cytoplasm, is the initiator of this pathway [21]. Following receptor dimerization, JAK proteins are activated. It is important to note that cytoplasmic domains of receptors contain tyrosine residues. After phosphorylation of these residues, JAK is activated to transphosphorylate the remaining tyrosine residues to form a binding site for the SH-2 domain of STAT proteins. As STAT molecules are transferred from the cytoplasm to the nucleus, they phosphorylate target genes by binding to DNA regulatory elements (DREs) [22, 23]. A change in this signaling pathway can influence pathological processes due to its important role in the regulation of biological processes because a malfunctioning of Janus kinase, JAK signal converter, and STAT signaling can lead to tumor formation [17]. Glioblastoma is a prevalent primary brain tumor in humans, and a highly infiltrative and extremely aggressive astrocytoma, which is characterized by resistance to apoptosis by radiation and chemotherapeutic treatments [7, 24]. Several studies indicate that JAK/STAT signaling pathways are involved in the progression, migration, and invasion of glioblastoma. For instance, a study found that glioma cells and other immune cells secrete IL-8, which promotes tumor migration, invasion, and mesenchymal transition by activating STAT1/hypoxia-inducible factor-1α (HIF-1α)/Snail pathway [25]. In glioblastoma, STAT5 signaling plays a role in proliferation and invasion, which are linked with tumorigenesis [26, 27]. Additionally, patients with glioblastoma have constitutively activated STAT3 and secreted IL-6 levels that are correlated with tumor grade [24]. The glioma cell-secreted granulocyte macrophage colony-stimulating factor (GM-CSF) activates STAT5 signaling in myeloid-derived suppressor cells (MDSCs), so Bcl2a1 is expressed and IRF8 is downregulated and they inhibit apoptosis and stimulate proliferation, respectively. Among all STAT family members, STAT3 has the most comprehensive oncogenic activity and immune suppressive role in glioblastoma. In malignant gliomas, aberrant STAT3 signaling is primarily the result of dysregulated upstream pathways, which drive proliferation, neovascularization, apoptosis resistance, and immune escape [2]. The interactions of tumor cells, reactive astrocytes, and microglia in glioblastoma lead to high-level expression of TGF-β and IL-10, which promote a positive feedback loop for STAT3 signaling and generate an immunosuppressive cytokine milieu [28]. A number of cellular populations, including reactive astrocytes, express IL-6 within the TME, which increases STAT3 signaling via JAK activation [29]. Upregulation of JAK/STAT gene targets, such as cytokines, cytokine receptors, and JAKs, is associated with a poor prognosis in classical glioblastoma [30]. The signaling pathway with phosphorylation of JAK2/STAT3 is associated with the management and enhancement of cancer cell proliferation, as well as resistance to radiation therapy that is one of the key roles of STAT signaling [31]. To inhibit STAT signaling, proteins like protein tyrosine phosphatases (PTP), cytokine signaling suppressors (SOCS), and active STAT protein inhibitors (PIAS) act as endogenous inhibitors [32]. So, targeting this pathway can suppress the expression of target genes that control essential cell functions and help to treat cancer by causing cell death (apoptosis) [33].Modulating and controlling STAT can be an effective strategy for protecting cells and tissues from malignant tumors [34].

Research on natural products has been boomed over the past few years, and the evidence that improper activation of STATs may cause malignancies in humans has prompted researchers to respond positively to this signaling pathway’s link to natural products [35]. Especially, since natural agents have demonstrated strong anticancer activity across a wide range of mechanisms and recent studies suggest phytochemicals can inhibit the JAK/STAT pathway and growth of cancer cells [5].
3. Natural Products with Inhibitory Effects on JAK/STAT Signaling Pathways

Since phytochemicals have been studied in a variety of cancers, some of these natural compounds have inhibited countless inflammatory pathways. By reducing the production of certain cytokines, some of these compounds are capable of inhibiting the STAT3 phosphorylation pathway and others act directly as inhibitors of JAK [36, 37]. In other research, the factors that control the SH2 domain and limit STAT dimerization have been investigated [38]. Several plant groups have been studied, including phenols (including resveratrol, curcumin, bergamottin, capillarisin, emodin, garcinol, cardamomin, castacin, and apigenin that inhibit JAK1,2 as well as STAT3), steroids (including diosgenin, ergosterol peroxide, and guggulsterone that inhibit JAK1/ JAK2 and DNA binding activity as well as STAT3), and terpenoids (including cucurbitacin, andrographolide, betulinic acid, cryptotanshinone, celastrol, oridonin, and alantolactone that inhibit JAK1 and JAK2 as well as STAT3 and STAT5) and these studies have yielded promising results for cancer prevention and therapy [5]. In the next section, we described the mechanisms and anticancer effects of some phytochemicals that act as inhibitors of the JAK/STAT signaling pathways by focusing on their therapeutic roles in glioblastoma (Figure 1).

3.1. Phenolics and Polyphenols. There are over 8,000 components of these plant compounds in nature, all of them contain hydroxyl groups and aromatic rings. These metabolites are water-soluble and exist in various vegetables and fruits. They can also control various biochemical and pharmacological effects, such as anti-inflammatory mechanisms, immune-modulating, and antioxidant effects, as well as regulation of some signaling pathways [5, 39].

3.1.1. Resveratrol. Resveratrol is an anti-inflammatory phytoalexin with antioxidant properties [40]. Its chemical and anticancer effects have been shown in multiple studies. Resveratrol can be found in peanuts, grapes, and berries, also known as trans-3,5,40-trihydroxystilbene. In a number of studies, it has been demonstrated that resveratrol administration is effective in glioblastoma treatment through inhibition of cancer cell proliferation, migration, and viability via inhibition of various molecular pathways [41]. The apoptotic and amplifying activities of this compound are related to the inhibition of JAK/STAT signaling, as it can prevent STAT1 phosphorylation by inhibiting JAK and STAT3, which reduces the activity of antiapoptotic genes and induces tumor cell death. Src/STAT and JAK/STAT pathways can both be blocked by resveratrol, that is, demonstrating its antitumor activity [5, 42, 43]. Recent research has shown that resveratrol can increase ROS generation and induce oxidation-related cellular lesions in U251 cells (malignant glioblastoma cell lines) by activating a ROS-related mitochondrial signaling pathway [44]. On glioblastoma cell lines LN18 and U87, Song et al. indicated that resveratrol reduced epithelial to mesenchymal transition (EMT), expression of β-catenin, and decreased the expression of stem cell markers (Twist, Snail, MMP-2, MMP-9, Slug, and Smad) [45] (Table 1).

3.1.2. Curcumin. Curcumin, a member of the ginger family (Zingiberaceae) is a diarylheptanoid compound, which is derived from turmeric’s rhizome. For many years, it has been used to treat chronic illnesses such as neoplastic diseases. There has been extensive evidence that curcumin has anti-inflammatory, antitumor, and antioxidant properties. Furthermore, it improves drug resistance in cancer therapy, so its consumption may overcome the resistance to gefitinib or erlotinib in humans with non-small-cell lung cancer (NSCLC) as a potential epidermal growth factor receptor tyrosine kinase inhibitor (EGFR-TKI) [46, 47].

By inhibiting tumor microenvironment mechanisms such as inflammation, angiogenesis, and metastasis, it can prevent the progression of malignant tumors. Additionally, it is capable of influencing and inhibiting molecular signaling mechanisms [48]. The anticancer activity of curcumin has been linked to its ability to modulate oncogenes (egr-1, c-Myc, Bcl-XL, NF-κB, and p53), transcription factors (NF-κB, STAT3, and AP-1), and protein enzymes (COX and LOX) [49–51].

There are numerous studies in which it has been shown that curcumin inhibits STAT3 through induction of ROS or RANK gene activity in glioblastoma cells, or by suppressing JAK1,2/STAT3 phosphorylation through downregulation of MMP-9, c-Myc, Ki-67, and Snail, suppressing cell proliferation, migration, and invasion by inducing G2/M cell cycle arrest [52–56] (Table 1).

So, by suppressing STAT3, curcumin disrupts the JAK/STAT signaling pathway (Figure 2). Various studies have shown that this plant compound can suppress proliferative
Table 1: Natural products inhibitory effects on JAK/STAT and related signaling pathways as therapeutic strategy for glioblastoma and other cancers.

| Natural compound | Type of natural compound | Chemical formula | Cancer/cell type or animal model studied | Used dosage | Function/mechanism | Ref. |
|------------------|--------------------------|------------------|------------------------------------------|-------------|--------------------|-----|
| Resveratrol      | Polyphenols              | C_{14}H_{12}O_{3} | U251 cell line                           | 100 μM      | Preventing STAT1 phosphorylation by inhibiting JAK and STAT3 and apoptotic genes induction.      | [5, 44, 45] |
|                  |                          |                  | LN18 and U87 cell lines                  | 20 μM-40 μM | Reduction of epithelial to mesenchymal transition (EMT), expression of β-catenin and decreased the expression of stem cell markers (Twist, Snail, MMP-2, MMP-9, Slug, and Smad). |      |
| Curcumin         | Glioblastoma cells       | C_{21}H_{20}O_{6} | Non-small-cell lung cancer (NSCLC)        | 1 μM-20 μM  | Inhibiting tumor microenvironments such as inflammation, angiogenesis, and metastasis. | [46–51, 52, 53–56] |
|                  |                          |                  | Glioblastoma cells                       | 10 μM-70 μM | Inhibiting STAT3 through induction of ROS or RANK gene activity by suppressing JAK1, 2/STAT3 phosphorylation through downregulation of MMP-9, c-Myc, Ki-67 and Snail. | [46–51, 52, 53–56] |
|                  |                          |                  | A549, U87, and U251 cell lines           | 2 μM and 10 μM | Wound-healing, migration, invasion by inducing G2/M cell cycle arrest. | [5, 56, 59–62] |
| Bergamottin      | Glioblastoma cells       | C_{21}H_{22}O_{4} | A549, U87, and U251 cell lines           | 2 μM and 10 μM | Inhibiting of STAT3 transcription by acting as a phytoestrogen. | [5, 56, 59–62] |
| Polyphenols      |                          | C_{20}H_{20}O_{4} | Glioblastoma cells                       | 2 μM and 20 μM | Inhibiting of NF-κB and IL-6-induced STAT3 activity and activating caspases 3 and 9 for stimulating apoptosis. Modulating the expression of phorbol-12-myristate-13-acetate-induced protein 1 (PMAIP1, alternative known as Noxa) and p53 tumor cells survival factors. | [5, 65, 66, 70] |
| Bavachin         | Microglia, macrophages,  | C_{22}H_{18}O_{11} | Glioblastoma cells                       | —           | Inhibiting the expression of iNOS, COX-2, and mPGES-1 and the production of nitric oxide (NO), matrix metalloproteinases (MMPs) and prostaglandin E2 (PGE2). |      |
| Epigallocatechin | and chondrocytes         |                  | Cholangiocarcinoma (CCA) cells           | 1 μM-50 μM  | Decreasing the HIF-1α activity and regulated transcription of genes related to energy metabolism, such as glucose transporter type 1 (GLUT1) and hexokinase 2 needed for survival of cancer cells. Inhibiting the progression of tumors by affecting the expression of cell cycle regulatory proteins, inhibiting JAK3/STAT3 signaling, and activating lethal caspases and apoptosis. | [72–76] |
| gallate (EGCG)   |                          |                  | —                                        | —           | Telomere shortening, elevating DNA damage through phosphorylation of γ-H2AX histone, micronuclei and telomere dysfunction. | [72–76] |

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Oxidative Medicine and Cellular Longevity
| Natural compound | Type of natural compound | Cancer/cell type or animal model studied | Used dosage | Function/mechanism | Ref. |
|------------------|-------------------------|----------------------------------------|-------------|--------------------|-----|
| Colorectal carcinoma | | | 5 μM | Inhibiting the angiogenesis via the Janus kinase/STAT3/IL-8 pathway in administration of curcumin. | |
| | | | 50 mg/kg | | |
| | | | Glioblastoma cultures | | |
| | | | 500 μM | | |
| PDX mouse model | | | | | |
| Induction of autophagy and apoptosis. | | | | | |
| Chalcones | C_{15}H_{12}O | Glioblastoma stem cells | 20 μM-40 μM | Inducing apoptosis by inhibiting STAT3 phosphorylation and activating caspases 8 and 9 and releasing ROS, changing the mitochondrial membrane potential and releasing the cytochrome C. | [77–80] |
| | 20 μg/mL | | | | |
| | | | Inhibiting the STAT3 phosphorylation, blocking STAT3 nuclear transport, and attenuating the expression of downstream genes including VEGF, survivin, Bcl-XL, and Bcl-2. | | |
| | Cardamonin | C_{16}H_{14}O_{4} | Prostate cancer cells | 2.5 μM-40 μM | Decreasing STAT3 and STAT5A protein expression. | [83, 84, 86] |
| | Primary and recurrent glioblastoma cells | 10 μM-25 μM | Inhibiting the STAT3 acetylation and dimerization, and negatively affects the protein’s ability to bind to DNA. | | |
| | HCC cells | 10 μM and 25 μM | Targeting signaling molecules involved in apoptosis (X-IAP, cIAP, caspase 3/9, PARP cleavage, and NF-κB). | | |
| | Pancreatic cancer cells (BxPC-3) | | | | |
| | Breast cancer cell lines MDA-MB-231 and Prostat cancer cell line DU145 | 10 μM and 25 μM | Decreasing both total and phosphorylated STAT3. | | |
| | U-87 MG and GBM8401 cell lines | 2.5 μM-40 μM | Inhibiting proliferation, invasion, and migration of cancer cells by enhancing the hsa-miR-181d/STAT3 and hsa-miR-181d/STAT5A ratios, dose dependently. | [83, 84, 86] |
| | U87/MG mouse xenograft model | 1 mg/kg body (intraperitoneal injection) | Downregulation of total as well as pSTAT-3 (Tyr 705) in tumors of garcinol administered mice. | | |
| | MDA-MB-231 xenograft mouse model | 5 mg/day/animal (oral gavage) | | | |
| | Immunocompromised mouse model | 1 mg/kg (intraperitoneally) | | | |
| | GMB cells and MCF-7 cell line | 100 μg/mL | | | |
| Silibinin | C_{25}H_{32}O_{10} | Human umbilical vein endothelial cells (HUVECs) | 100 nM-100 μM | Reducing glioblastoma tumor growth by attenuating STAT3/5A expression, enhancing the Bax/Bcl-XL apoptotic ratio, and downregulating the Ki-67 proliferation index. | [10–12] |
| Chrysin | C_{13}H_{10}O_{4} | | | | |
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Oxidative Medicine and Cellular Longevity
| Natural compound | Type of natural compound | Chemical formula | Cancer/cell type or animal model studied | Used dosage | Function/mechanism | Ref. |
|------------------|--------------------------|------------------|-----------------------------------------|-------------|--------------------|-----|
| Apigenin         | C15H10O5                 | Glioblastoma cells (GBM8901 cells) | 25μM-100μM | Arresting the cell cycle arrest in the G1 phase due to increasing P21 (waf1/cip1) and activating the P38-MAPK | Inducing apoptosis and TNF-α and reducing MCL-1 and Bcl-XL through inhibition of JAK1/2 and STAT3 |
|                 |                          | C15H10O5         | Glioblastoma cells (U1242 MG and U87 MG cell lines) | 10μM-80μM | Altering cytokine profiles, which are important for regulating the immune response. | [96–101] |
| Rat C6 glioma cells | 1–100 μmol/L | Multiple cancer types such as glioblastoma | Various concentrations such as 1.5 μM-50 μM | Halting the G2/M phase of the cell cycle by activating caspases 8, 9, and 3 and inhibiting Fas/CD95 as well as STAT3/P53/P21 signaling (cucurbitacin E). | Sensitizes glioblastoma to t-AUCB by dual inhibition of Hsp27 and COX-2. | [112–115] |
| T98G and U87 | Quercetin C15H10O7 | Glioblastoma cell lines | 25 μM | Regulating the PI3K/Akt/mTOR signaling pathways, IL-6/STAT3 signaling pathways, modulation of apoptosis-related proteins, altering the intracellular pH (pHi), and MMP-2/9 and fibronectin expression. | | [103, 105–107, 126] |
| Terpenoids | Cucurbitacins C30H42O7 | T24 cell line | 250 nM-2000 nM | Halting the G2/M phase of the cell cycle by activating caspases 8, 9, and 3 and inhibiting Fas/CD95 as well as STAT3/P53/P21 signaling (cucurbitacin E). | | [112–115] |
|                 |                          | KS62 cells | 5 nM-80 nM | | | |
Table 1: Continued.

| Natural compound | Type of natural compound | Chemical formula | Cancer/cell type or animal model studied | Used dosage | Function/mechanism | Ref. |
|------------------|--------------------------|------------------|------------------------------------------|-------------|-------------------|-----|
| Steroids         | Diosgenin                | C_{27}H_{42}O_{3} | U87, U87-EGFR-WT, and U87-EGFRviii glioblastoma cell lines | 25 nmol/L-5000 nmol/L | Inhibiting STAT3 activation and Raf/MEK/ERK signaling pathways (Cucurbitacin B). |       |
|                  | Dioscin                  | C_{43}H_{72}O_{16} | U87MG cells                              | 100 nM      | Inhibiting the proliferation of glioma cells by decreasing p-JAK1, p-JAK2, p-STAT3, and p-STAT5 and VEGF-induced JAK2 and STAT3 activation levels (cucurbitacin I). |       |

Increasing apoptosis, ROS generation, DNA damage, and arrest of the S phase cell cycle.

Suppressing the STAT3 activation through JAK1, c-Src, and JAK2.

Inhibiting cell proliferation and cell cycle arrest in G0/G1 phase by decreasing the cyclin D protein.

[123–125]
and invasitve proteins as well [57]. Some preclinical trials have shown that curcumin increases the effectiveness of existing chemotherapeutic agents, which is a great benefit [58].

3.1.3. Bergamottin. Bergamottin is a furanocoumarin with anti-inflammatory and antioxidant properties isolated from grapes. This compound is found in bergamot, grapefruit, and lemon. Furanocoumarin is primarily used by plants to defend themselves against predators and is considered a natural pesticide. Additionally, it acts as an inhibitor of some cytochrome enzymes, such as P450 [59, 60]. By regulating the signal transducer and activating the STAT3 transcription signaling pathway, which is linked to tumor progression, this combination can negatively regulate the cell cycle and activate apoptosis. This suppression is achieved by inhibiting phosphorylation of activated kinases of JAK1, JAK2, C-Src, and SHP-1 and suppressing STAT3 and its downstream products including Bcl-2, Bcl-XL, cyclin D1, survivin, COX-2, IAP-1, MMPs, P53, etc.

Also, it was revealed that treatment of human glioma cells with bergamottin significantly inhibited wound-healing migration and Matrigel invasion of human glioma cells, relative to untreated cells by inhibition of EMT, JNK, PI3K, Akt, NF-κB, STAT3, Rac1, and mTOR kinases, and MMP-9 production in A549, U87, and U251 cell lines [63, 64] (Table 1).

3.1.4. Bavachin. Bavachin inhibits STAT3 transcription by acting as a phytoestrogen. This compound is obtained from plants such as Psoralea corylifolia. By inhibiting NF-κB and IL-6-induced STAT3 activity and activating caspases 3 and 9, it can stimulate apoptosis [65]. Bavachin also modulates the expression of phorbol-12-myristate-13-acetate-induced protein 1 (PMAIP1, alternative known as Noxa) and p53 which play a role in tumor cells’ survival [5, 66].

Bavachin also suppresses the inflammation caused by LPS, which causes the release of (nitric oxide) NO, (prostaglandin E2) PGE 2, as well as IL-6 (produced by macrophage M1) and (NOD-, LRR-, and pyrin domain-containing protein 3) NLRP3 (in macrophages). In response to the release of NLRP3, the proinflammatory cytokine IL-1β is activated, which contributes to pyroptosis [67–69]. Bavachin also decreases HIF-1α activity, the main oxygen sensor within cells, under hypoxia in a concentration-dependent manner and reduces HIF-1-regulated transcription of genes related to energy metabolisms, such as glucose transporter type 1 (GLUT1) and hexokinase 2 [70] (Table 1).

3.1.5. Epigallocatechin Gallate (EGCG). Catechins in green tea act as potent antioxidants and prevent the progression of tumors by acting as antiangiogenic agents. By affecting the expression of cell cycle regulatory proteins, inhibiting JAK3/STAT3 signaling, and activating lethal caspases, this compound induces apoptosis and cell proliferation as an
EGFR-TKIs. In this line, EGCG, also known as epigallocatechin-3-gallate, inhibits the EGFR dimerization and its activation or even binds to EGFR to inhibit the EGFR phosphorylation. Moreover, EGCG suppresses glioblastoma development by telomere shortening, elevating DNA damage through phosphorylation of γ-H2AX histone, micronuclei, and telomere dysfunction [51, 71].

In addition, it inhibits carcinogenesis by affecting a wide range of signaling pathways, including Wnt, MAPK, Notch, and PI3K/Akt. The low cost and high immunity of this compound make it a great candidate for preventing cancer. This nontoxic natural agent can be used to treat malignancies in humans, either alone or in combination with other treatments [72, 73].

Furthermore, EGCG with quercetin can block the JAK/STAT pathway, ultimately making STAT1 and STAT3 inactive in cholangiocarcinoma (CCA) cells [71]. A dose-dependent reduction of phosphorylated STAT1/3 was observed when this combination was administered. In addition, this study revealed that CCA cells’ proliferation and migration were also impaired [74]. In a colorectal carcinoma microenvironment and in a PDX mouse model, administration of curcumin and EGCG together inhibited angiogenesis via the Janus kinase/EGCG pathway [75]. In a study, Grube et al. indicated that after 500 μM concentration of EGCG, strong induction of autophagy and apoptosis was observed in glioblastoma cultures, whereas its 100 nM concentration leads to elevate accumulation of autophagic vacuoles and reactive oxygen species production as a stress response in the first 12 h of treatment (Table 1). This data indicates that although green tea may have chemopreventive properties, it is not directly cytotoxic [76].

3.1.6. Chalcones. Chalcones are phenolic compounds in the flavonoid family. Among the fruits and vegetables that contain chalcones are oranges, strawberries, potatoes, and bean sprouts, as well as some spices, like licorice. Through their antioxidant, cytotoxic, and cell signaling modulatory properties, these phytochemicals are involved in the reduction of malignant tumors progression. In addition, this phenolic compound induces apoptosis [5]. Unsaturated flavonoids induce apoptosis by inhibiting STAT3 phosphorylation and activating caspases 8 and 9. As a result of releasing ROS and changing the mitochondrial membrane potential, cytochrome C is released and the death process is initiated [77]. Researchers have studied cardamonin (CAR), a chalcone isolated from plants of the Zingiberaceae, Asteraceae, Piperaceae, Polygonaceae, and many other families for its antitumor effects [5]. In the presence of silibinin activity, JAK2 inhibition with CAR reduced STAT3 phosphorylation but did not inhibit it. On the other hand, inhibition of JAK1 completely inhibits STAT3 phosphorylation and dimerization and negatively affects the basic mechanism of garcinol [82]. In order to investigate the molecular mechanism of garcinol’s action on pancreatic cancer cells, signaling molecules involved in apoptosis (X-IAP, cIAP, caspase 3/9, and PARP cleavage) were targeted, as well as NF-κB, VEGF, IL-8, and PGE2, which involve in chemo-resistance in pancreatic tumors. The antiproliferative, proapoptotic, antimeiotic, and antiangiogenic effects of garcinol in pancreatic cancer cells were significantly enhanced relative to those in untreated cells [83]. Study results showed that garcinol inhibited both total and phosphorylated STAT3 in breast, pancreatic, and prostate cancer cell lines and reduced cell invasion in these cancer cell lines in a dose-dependent manner [84]. A study by Liu et al. showed that garcinol inhibited proliferation, invasion, and migration in glioblastoma U-87 MG and GBM8401 cells dose dependently, which was mediated by increasing the hsa-miR-181d/STAT3 and hsa-miR-181d/STAT5A ratios in glioblastoma cells [85]. Garcinol reduced glioblastoma tumor growth in an immunocompromised mouse model by reducing STAT3/STAT5A expression, enhancing the Bax/Bcl-XL apoptotic ratio, and downregulating the Ki-67 proliferation index, in vivo [86] (Table 1).

3.1.7. Silibinin. Silibinin is a nonsteroidal anti-inflammatory drug that targets inflammation and positively affects cellular and noncellular components [87]. It is a plant compound derived from milk thistle that was previously used for the treatment of chronic liver disease. Moreover, it is an antioxidant/anti-inflammatory flavonoid that offers an appealing strategy for the treatment of cancer [88]. Several studies have demonstrated the benefits of this phytochemical in combination with conventional chemotherapy treatments, including a reduction in adverse neurological, cardiac, and renal effects [5]. In the presence of silibinin activity, JAK2 inhibitors reduced STAT3 phosphorylation but did not inhibit it. On the other hand, inhibition of JAK1 completely inhibits STAT3 phosphorylation and activates caspase 9 and 3, which cause apoptosis [89]. The downregulation of miR-21 by silibinin appears to activate the p53 pathway, as well as several genes related to intrinsic and extrinsic apoptosis. In glioblastoma cells, it has been reported that miR-21 inhibition by antisense oligonucleotides increased caspase activation and cell death (Table 1). Together, these results suggest that silibinin might induce apoptosis and cell cycle arrest in part by inhibiting miR-21 and miR-155 as onco-miRs and may be able to have a positive impact on eliminating human malignancies [89, 90].
3.1.9. Chrysin. It has been demonstrated that chrysin is a naturally occurring flavonoid found primarily in propolis and honey. As a result of its beneficial biological properties (anti-inflammatory, antitumor, antioxidant, and anti-estrogen), this compound, known as 5,7-dihydroxyflavone, is included in the list of natural products for anticancer. The compound has drawn the most attention due to its low toxicity, which inhibits angiogenesis, metastasis, and tumor growth [91, 92]. This combination has inhibited the signaling pathway of Akt and STAT in preclinical trials [5]. Based on the data, chrysin appears to be able to downregulate soluble IL-6 receptor (IL-6R), glycoprotein 130 (gp130), phosphorylated JAK1, and STAT3 levels and VEGF in human umbilical vein endothelial cells (HUVECs) [93]. Chrysin exerts antitumor activity via direct interaction with multiple molecular targets and modulation of signal transduction pathways involved in cellular metabolism (AMPK/Akt/ERK/PPAR) and inflammation (NF-κB, P38/MAPK, TBK1, and Wnt/β-catenin) [94] (Table 1). During chrysin treatment, glioma cells were arrested in the G1 phase due to increased P21 (waf1/cip1) and activation of P38-MAPK [95]. Chrysin displayed greater antiglioblastoma activity in GBM8901 cells compared to other compounds (PWE, pinocembrin, and tiliroside), and it reduced growth in a time-dependent manner from 25 to 100 μM in GBM8901 cells. However, the chrysin compound did not cause damage to other glial cell lines; this is suggesting that it might be able to display specific antiglioblastoma properties without damaging normal cells [96].

3.1.10. Apigenin. This flavonoid can be found in fruits (e.g., citrus and apple), vegetables (e.g., parsley and celery leaves), and some medicinal plants (chamomile, thyme, oregano, lemon balm, and yarrow). It has been proven to have antitumor properties and is considered by researchers. Apigenin inhibits angiogenesis by reducing the glucose uptake of cancer cells and inhibiting adhesion molecules [97]. The antitumor activity of this compound is through inhibition of Akt and STAT in preclinical trials [5]. Based on the data, chrysin appears to be able to regulate multiple enzyme expressions [106] (Figure 2). Curcurbitacin I (JSI-124) potently inhibits G2/M arrest in glioblastoma cells and decreased levels of Akt, mTOR, ERK, STAT3, and S6K proteins [99, 100]. Researchers found that apigenin reduced survival, growth, proliferation, and migration of rat C6 glioma cells by altering their cytokine profiles, which are important for regulating the immune response [101] (Table 1). Taking the drug in combination with other flavonoids also led to synergistic activity by decreasing MMP-2 expression and increasing fibronectin, laminin, and glial fibrillary acidic protein (GFAP) expressions [102]. By regulating the levels of TNF-α, caspases, and apoptotic proteins, apigenin with hydroxygenkwanin provided enhanced antiglioma activity [103].

3.1.11. Quercetin. Quercetin (chemically known as 3,3',4',5,7-pentahydroxy flavone) is a polyphenolic flavonoid found in a variety of fruits and vegetables [104]. Because of its anti-inflammatory, antioxidant, and antitumor effects, quercetin has been extensively studied in vitro and in vivo in multiple cancer models including lung, gastric, cervical, breast, prostate, and colon [105–108]. Quercetin has the ability to impact apoptosis and arrest phase G1 cell cycle in tumor cells, through its interaction with cell cycle regulators, including cyclin-dependent kinase-(CDK-)4 and cyclin D1, activating p53, cytochrome c release, and also inducing caspase 9 and caspase 3 release [109]. Furthermore, quercetin can be viewed as a potential multitarget agent and a potential PI3K inhibitor [110] (Table 1). By chemoprotecting normal cells from chemotherapy and radiotherapy, this phytochemical could provide a significant advantage for anticancer treatment [106]. In a variety of cellular and animal models as well as in humans, quercetin modulates signaling pathways and gene expression to exert antioxidant, anti-inflammatory, and antitumor properties [111]. The antitumor effects of this natural drug for the treatment of glioblastoma may occur mainly through the regulation of PI3K/Akt/mTOR signaling pathways, IL-6/STAT3 signaling pathways, modulation of apoptosis-related proteins, alteration of intracellular pH (pHi), and MMP-2/9 and fibronectin expressions [106] (Figure 2).

3.2. Terpenoids. Compounds called isoprenoids are second-ary plant metabolites that are abundant in nature. Several terpenoids have been used to treat cancer, including taxol [112]. Phytochemicals such as this combination have antifungal, antimicrobial, antiviral, antiparasitic, and immunomodulating properties. Research has shown that terpenoids inhibit the growth of cancer cells without causing any toxicity to normal human cells. It has been identified that various types of these compounds (cucurbitacins, andrographolide, raptotanshinone, nimidolide, etc.) can inhibit JAK/STAT signaling in human malignancies [5]. Here, we described curcurbitacin's effects on glioblastoma treatment by targeting JAK/STAT signaling pathways, while it seems that future investigations are required to determine the therapeutic effects of other terpenoids (e.g., andrographolide, raptotanshinone, and nimidolide) on glioblastoma by inhibiting JAK/STAT signaling pathways.

3.2.1. Cucurbitacin. It is a four-ring triterpene compound that is highly oxidized. It can inhibit some signaling pathways (JAK/STAT3, Wnt, PI3K/Akt, and MAPK) and has strong anticancer activity. The mechanism of its action is antiproliferation, inhibition of growth and mutation, and increase of cell cycle arrest [113]. Analogues of curcurbitacins range from curcurbitacin A to cocurbitacin T. The curcurbitacin E compound inhibits the cell cycle and induces apoptosis in T24 cells (human bladder carcinoma cell line). It is believed that this curcurbitacin plays a role in halting the G2/M phase of the cell cycle by activating caspases 8, 9, 3 and Fas/CD95 and inhibiting STAT3/P53/P21 signaling [114]. Curcurbitin B inhibits STAT3 activation and Raf/MEK/ERK signaling pathways in K562 cells (leukemia cell line) [115]. According to a study, curcurbitin I markedly decreased p-JAK1, p-JAK2, p-STAT, and p-STAT5 levels and inhibited proliferation of glioblastoma cells [116] (Figures 1 and 2). Curcurbitacin I (JSI-124) potently inhibits
VEGF-induced JAK2 and STAT3 activation. Coculturing HUVECs with glioblastoma cells increased their migration rate significantly. However, coculturing JSI-124-treated glioblastoma cells and HUVECs led to a marked decrease in HUVEC migration. This compound can inhibit HUVEC tubular formation induced by U87MG cells [117, 118] (Table 1). It seems that cucurbitacin will be increasingly used in the near future to target oncogenic pathways therapeutically.

3.3. Steroids. Steroids come from phytosterols, a class of compounds similar to cholesterol but differing in the number of hydrocarbon chains [119]. This phytochemical has been shown to have beneficial effects on diabetes and heart disease. According to new research, phytosterol consumption is directly connected to cancer prevention. The human body can be protected by them, and they can prevent cell proliferation and metastasis [119]. The compositions in these compounds affect hormone-dependent endocrine glands to exert their antitumor effects [120]. The next paragraph is about one drug from this phytochemical family.

3.3.1. Diosgenin. The compound in subject is a steroid sapogenin found in legumes, potatoes, and certain vegetables [121]. Also, fenugreek contains large amounts of diosgenin. In both preclinical and clinical studies, the therapeutic effects of fenugreek (antidiabetic, antihyperlipidemic, antiobesity, anticancer, anti-inflammatory, antioxidant, antifungal, and antibacterial) have been demonstrated [122]. Both in vitro and in vivo studies have shown that dioscin (the structural analog of diosgenin) is effective against glioblastoma. Dioscin treatment increased apoptosis, ROS generation, DNA damage, and arrest of the S phase cell cycle in C6 glioma cells [123]. STAT3 is affected by JAK1, c-Src, and JAK2 activation; these pathways are also suppressed by diosgenin [124] (Figure 2, Table 1). A decrease in cyclin D protein is caused by diosgenin, which inhibits cell proliferation and stops G0/G1 phase [125]. A study found that diosgenin decreased STAT3-regulated gene expression and inhibited hepatocellular carcinoma (HCC) proliferation. Diosgenin appears to be a novel STAT3 activation pathway inhibitor with potential application to treatment of HCC and other cancers [124].

4. Some Other Kinds of Natural Products or Diets Which Might Be Useful to Reduce Glioblastoma

4.1. Extra Virgin Olive Oil and Other Oils. Nowadays, more attention is paid to the use of natural products for the treatment of glioblastoma, but there are still many natural compounds with promising antioxidant/anti-inflammatory properties through the JAK-STAT pathway that have been overlooked.

There are several studies that examined the effects of some kinds of natural products as useful diets to reduce glioblastoma complications. A study by Lamy et al. on olive oil compounds and their effect on glioblastoma cells has revealed that four compounds contained in extra virgin olive oil including hydroxytyrosol (HT), oleuropein (OL), oleic acid (OA), and tyrosol (Tyr) can inhibit tumor necrosis factor-α (TNF-α) induced expression of cyclooxygenase-2 (COX-2) in a U-87 MG cell line model as a human glioblastoma cell. Furthermore, among these combinations, Tyr and OA could significantly inhibit TNF-α-induced COX-2 gene and protein expression, as well as prostaglandin E2 (PGE2) secretion, prevent TNF-α-induced JNK and ERK phosphorylation and also reduce human brain microvascular endothelial cells (HBMECs) migration [127]. Another examination of the systemic lupus erythematosus (SLE) model in mice demonstrated that dietary extra virgin olive oil ameliorates kidney injury through MAPK activation and suppression of JAK/STAT and NF-κB pathways [128]. Based on traditional Persian medicine, citral, a major component of lemon balm essential oil, induces apoptosis in GBM cells that express active multidrug resistance-associated protein 1 (MRP1) [129].

Currently, a natural combination of the wild olive tree which is called acbeuche (ACE) in Spain is being suggested for attenuating ocular oxidative stress induced by arterial hypertension. This study has revealed that ACE oil-enriched diet reduced nicotinamide adenine dinucleotide phosphate (NADPH) oxidase activity in gene and protein levels and improved alterations in bioavailability of nitric oxide and antioxidant enzyme profile in the retinas of hypertensive mice [130]. The use of a diet with extra virgin olive oil containing Hydroxytyrosol-acetate (HTy-Ac) has been shown to reduce serum synovial and cartilage biomarkers such as cartilage oligomeric matrix protein (COMP) and MMP-3 through the activation of the nuclear factor (erythroid-derived 2-) like 2/heme oxygenase (Nrf2/HO-1) pathway and the inhibition of relevant signaling pathways like JAK-STAT, MAPKs, and NF-κB in a model of collagen-induced arthritis (CIA) in mice [131]. However, approximately there is no information about testing the ACE potential therapeutic properties for glioblastoma and it remains elusive. Therefore, it is necessary to conduct preclinical and clinical studies in this field.

4.2. Withania somnifera and Withaferin-A. In addition, there are other interesting plants with properties in regulation of the JAK/STAT pathway that could be used to treat glioblastoma. In diverse preclinical cancer models, Withania somnifera as a magic plant and its extracts like withaferin-A (WA), indicated anti-inflammatory activities and regulated several signaling pathways such as JAK/STAT, NF-κB, and AP-1. The compound is suggested for treating cancer patients with drug resistance. However, clinical trials are testing this natural product for cancer therapy [132]. The development of steroidal lactone, WA as an anticancer agent which is obtained from the roots and leaves of withania somnifera, may provide a novel approach for treating glioblastoma since it modulates several oncogenic pathways simultaneously. Some WA properties which are collected from studies on glioblastoma are antiproliferative activities and modulating MAPK, JAK/STAT, and Akt/mTOR signaling, induction of apoptosis and cell cycle arrest, modulation of HSP60/HSP70 chaperones, and effect on DNA damage and repair mechanisms [133–136].
4.3. Blueberries and Malvidin. The benefits of blueberries, rich in anthocyanins, have gained considerable attention as functional foods. The major constituent of blueberry, malvidin, has been shown to abolish the JAK/STAT3 pathway, downregulate cell proliferation, and induce mitochondrial-mediated apoptosis in a hamster model of oral oncogenesis. In addition, the combination of S3I-201, a STAT3 inhibitor, with blueberry and malvidin was more effective in STAT3 inhibition respective to the single agent [137]. The compound resveratrol, found in blueberries, inhibits the growth of cells, induces apoptosis, stops G0/G1-phase cell cycle arrest, and blocks STAT3/JAK2 signaling [138]. Malvidin could inhibit EMT in glioblastoma cells (U-87MG) by signaling through TGF-β/SMAD2 [139]. It sounds that berries and their combinations are precious sources for glioblastoma treatment. But, despite the above studies, it is still essential to conduct more precise clinical studies on the effect of blueberries compounds and malvidin on glioblastoma cells through the JAK/STAT pathway.

4.4. Vitamin C and E. Other natural products with antioxidant/anti-inflammatory properties that can be used for the treatment of glioblastoma are vitamins which have received more attention today [140]. The results of experiments performed on glioblastoma cell lines and tumor xenografts suggest that high doses of ascorbate (vitamin C) induce cytotoxicity and oxidative stress in malignant cells selectively in comparison to normal cells. At present, only a few numbers of studies are available that test ascorbate as a potential therapeutic agent for glioblastoma patients. Nevertheless, the combination of pharmaceutical ascorbate with radiation and temozolomide has the potential to improve the patient’s treatment performance and quality of life [141]. It has been shown by Yang et al. that methotrexate (MTX), a clinical drug for cancer treatment, alone or in combination with vitamins C and E inhibits the growth of GBM cells [142]. Vitamin C appears to regulate JAK/STAT, TRAIL, TGF/SMAD, and microRNAs in different cancers according to scattered scientific evidence [143]. It seems that antioxidant vitamins such as vitamin C and E could be used as early, unconventional, and inexpensive therapy for the treatment of glioblastoma. In order to confirm these findings, larger cohorts and placebo-controlled trials should be administered.

5. Future Directions and Conclusions

The median survival rate remains bleak for glioblastoma, as the most common malignant brain tumor in adults. Even though numerous attempts have been made, very few FDA-approved drugs are available for its treatment, and they do not work for everyone [144]. Chemotherapy, which significantly destabilizes the cell metabolism and signaling network, can also lead to drug resistance in glioblastoma treatment [8, 145]. The majority of drugs used in cancer therapy are natural products or their structural relatives. Combinatorial chemistry and high-throughput screening technologies are used by the pharmaceutical industry to obtain these drugs [146, 147]. Thus, more subtle use of nature-derived templates combined with organic chemistry will be much more likely to produce selective analogs [148]. Glioblastoma patients have failed most chemotherapy trials partly due to poor drug penetration through the BBB. Consequently, natural products may alter the function of BBB components and therefore alter their permeability [149]. An understanding of glioblastoma biology and the microenvironment may allow researchers to develop more precise target-specific approaches using natural products (raw or modified formulations) to prevent and treat glioblastoma while minimizing severe side effects and off-target risks associated with conventional chemotherapies as well as improving treatment efficacy. Nevertheless, the difficulties associated with the development of these anticancer agents include poor solubility, resistance to development, and damaging side effects. To overcome these difficulties, a nanotechnology platform has become successful in recent times in providing “nano or other modified natural products” and providing a new dimension and face to natural products for the treatment of cancer [150, 151].

Targeting JAK/STAT signaling pathways has been established to be a practical therapeutic approach for a variety of cancers, including glioblastoma. This review discussed multiple natural products with therapeutic effects on glioblastoma by targeting JAK/STAT signaling pathways. These natural products can be combined with other common JAK/STAT inhibitors to reduce their required dosage and toxicity. However, more investigations are needed to examine other natural products with JAK/STAT inhibitory effects for glioblastoma and their combinational therapy with other conventional therapies.

Data Availability

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors’ Contributions

Conceptualization and design were performed by H.Z and H.F. Validation, investigation, data curation, and writing were performed by H.F, H.S, C.T, H.Z, and M.NA. Review and editing were performed by H.Z, H.S, and M.NA. H.S, H.F, H.Z, MNA, R.M, and R.F revised the study. The study was supervised by H.Z and M.NA. All the authors read and approved the final manuscript. Hanieh Fahmideh and Hooriyeh Shapourian contributed equally to this work.

References

[1] M. Karami Fath, N. Karimfar, A. Fazlollahpour Naghibi et al., "Revisiting characteristics of oncogenic extrachromosomal DNA as mobile enhancers on neuroblastoma and glioma cancers," Cancer Cell International, vol. 22, no. 1, pp. 1-14, 2022.
A. R. Amin, O. Kucuk, F. R. Khuri, and D. M. Shin, “The role and therapeutic targeting of JAK/STAT signaling in glioblastoma,” Cancers, vol. 13, no. 3, p. 437, 2021.

H. Zalpoor, H. Shapourian, A. Akbari, S. Shalveh, and L. Haghshenas, “Increased neuropilin-1 expression by COVID-19: a possible cause of long-term neurological complications and progression of primary brain tumors,” Human Cell, vol. 1-3, 2022.

D. Hanahan and R. A. Weinberg, “Hallmarks of cancer: the next generation,” Cell, vol. 144, no. 5, pp. 646–674, 2011.

S. Bose, S. Banerjee, A. Mondal et al., “Targeting the JAK/STAT signaling pathway using phytocompounds for cancer prevention and therapy,” Cell, vol. 9, no. 6, p. 1451, 2020.

N. Joshi, F. Hajizadeh, E. A. Dezfouli et al., “Silencing STAT3 enhances sensitivity of cancer cells to doxorubicin and inhibits tumor progression,” Life Sciences, vol. 275, article 119369, 2021.

M. Karami Fath, K. Babakhaniyan, M. Zokaei et al., “Anticancer peptide-based therapeutic strategies in solid tumors,” Cellular & Molecular Biology Letters, vol. 27, no. 1, pp. 1–26, 2022.

M. Karami Fath, A. Azargoonjahromi, A. Kiani et al., “The role of epigenetic modifications in drug resistance and treatment of breast cancer,” Cellular & Molecular Biology Letters, vol. 27, no. 1, pp. 1–25, 2022.

R. V. Javid, F. Safizadeh, M. Rastegary et al., “Effects of pomegranate juice with and without aerobic training on glycemic control and lipid profile in women with type 2,” Diabetes, vol. 6, 2020.

Y.-J. Surh, “Cancer chemoprevention with dietary phytochemicals,” Nature Reviews Cancer, vol. 3, no. 10, pp. 768–780, 2003.

M. L. McCullough and E. L. Giovannucci, “Diet and cancer prevention,” Oncogene, vol. 23, no. 38, pp. 6349–6364, 2004.

G. L. Russo, "Ins and outs of dietary phytochemicals in cancer chemoprevention," Biochemical Pharmacology, vol. 74, no. 4, article 17382300, pp. 533–544, 2007.

A. R. Amin, O. Kucuk, F. R. Khuri, and D. M. Shin, "Perspectives for cancer prevention with natural compounds," Journal of Clinical Oncology, vol. 27, no. 16, pp. 2712–2725, 2009.

G. G. Meadows, "Diet, nutrients, phytochemicals, and cancer metastasis suppressor genes," Cancer and Metastasis Reviews, vol. 31, no. 3, pp. 441–454, 2012.

W. Fernando and H. V. Rupasinghe, "Anticancer properties of phytochemicals present in medicinal plants of North America," Using Old Solutions to New Problems—Natural Drug Discovery in the 21st Century, pp. 159–180, Intech, 2013.

J. K. Patra, G. Das, S. Bose et al., "Star anise (Illicium verum): chemical compounds, antiviral properties, and clinical relevance," Phytotherapy Research, vol. 34, no. 6, pp. 1248–1267, 2020.

M. Ashrafizadeh, H. Rafei, R. Mohammadinejad, E. G. Afshar, T. Farkhondeh, and S. Samarghandian, "Potential therapeutic effects of curcumin mediated by JAK/STAT signaling pathway: a review," Phytotherapy Research, vol. 34, no. 8, pp. 1745–1760, 2020.

S. Tabassum, R. Abbasi, N. Ahmad, and A. A. Farooqi, "Targeting of JAK/STAT signaling in breast cancer: therapeutic strategies to overcome drug resistance," Breast Cancer Metastasis and Drug Resistance, vol. 1152, pp. 271–281, 2019.

F. Seif, M. Khoshmirsafa, H. Aazami, M. Mohsenzadegan, G. Sedighi, and M. Bahar, “The role of JAK-STAT signaling pathway and its regulators in the fate of T helper cells,” Cell Communication and Signaling, vol. 15, no. 1, pp. 1–13, 2017.

F. Seif, H. Aazami, M. Khoshmirsafa et al., “JAK inhibition as a new treatment strategy for patients with COVID-19,” International Archives of Allergy and Immunology, vol. 181, no. 6, pp. 467–475, 2020.

M. Gadina, C. Johnson, D. Schwartz et al., “Translational and clinical advances in JAK-STAT biology: the present and future of jakinibs,” Journal of Leukocyte Biology, vol. 104, no. 3, pp. 499–514, 2018.

N. C. Reich, “STATs get their move on,” Jak-stat, vol. 2, no. 4, article e27080, 2013.

Y. Verhoeven, S. Tilborgs, J. Jacobs et al., “The potential and controversy of targeting STAT family members in cancer,” in Seminars in Cancer Biology, pp. 41–56, Elsevier, 2020.

H. Zalpoor, A. Akbari, A. Samei et al., “The roles of Eph receptors, neuropilin-1, P2X7, and CD147 in COVID-19-associated neurodegenerative diseases: inflammosome and Jak inhibitors as potential promising therapies,” Cellular & Molecular Biology Letters, vol. 27, no. 1, pp. 1–21, 2022.

C. W. Duarte, C. D. Willey, D. Zhi et al., “Expression signature of IFN/STAT1 signaling genes predicts poor survival outcome in glioblastoma multiforme in a subtype-specific manner,” PloS One, vol. 7, no. 1, article e29653, 2012.

S. Cao, C. Wang, Q. Zheng et al., “STAT5 regulates glioma cell invasion by pathways dependent and independent of STAT5 DNA binding,” Neuroscience Letters, vol. 487, no. 2, pp. 228–233, 2011.

H. Zalpoor, A. Akbari, and M. Nabi-Afjadi, “Ephrin (Eph) receptor and downstream signaling pathways: a promising potential targeted therapy for COVID-19 and associated cancers and diseases,” Human Cell, vol. 35, no. 3, pp. 952–954, 2022.

D. Henrik Heiland, V. M. Ravi, S. P. Behringer et al., “Tumor-associated reactive astrocytes aid the evolution of immunosuppressive environment in glioblastoma,” Nature Communications, vol. 10, no. 1, pp. 1–12, 2019.

Q. Liu, G. Li, R. Li et al., “IL-6 promotion of glioblastoma cell invasion and angiogenesis in U251 and T98G cell lines,” Journal of Neuro-Oncology, vol. 100, no. 2, pp. 165–176, 2010.

A. K. Park, P. Kim, L. Y. Ballester, Y. Esquenazi, and Z. Zhao, “Subtype-specific signaling pathways and genomic aberrations associated with prognosis of glioblastoma,” Neuro-Oncology, vol. 21, no. 1, pp. 59–70, 2019.

S.-Y. Park, C.-J. Lee, J.-H. Choi et al., “The JAK2/STAT3/CCND2 Axis promotes colorectal cancer stem cell persistence and radioresistance,” Journal of Experimental & Clinical Cancer Research, vol. 38, no. 1, pp. 1–18, 2019.

M. David, H. E. Chen, H. E. Chen, S. Goelz, A. C. Larner, and B. G. Neel, “Differential regulation of the alpha/beta interferon-stimulated Jak/Stat pathway by the SH2 domain-containing tyrosine phosphatase SHPPT1,” Molecular and Cellular Biology, vol. 5, no. 12, pp. 7050–7058, 1995.

X. Lin, H. Chen, and X. Zhan, “mir-203 regulates JAK-STAT pathway in affecting pancreatic cancer cells proliferation and apoptosis by targeting SOCS3,” European Review for Medical and Pharmacological Sciences, vol. 23, no. 16, pp. 6906–6913, 2019.

A. M. Heeren, I. F. van Luijk, J. Lakeman et al., “Neo-adjuvant cisplatin and paclitaxel modulate tumor-infiltrating T cells in...
patients with cervical cancer,” *Cancer Immunology, Immunotherapy*, vol. 68, no. 11, pp. 1759–1767, 2019.

[35] S. R. Walker and D. A. Frank, “STAT signaling in the pathogenesis and treatment of cancer,” in *Signaling Pathways in Cancer Pathogenesis and Therapy*, pp. 95–108, Springer, 2012.

[36] B. B. Aggarwal, A. B. Kunnumakkara, K. B. Harikumar et al., “Signal transducer and activator of transcription-3, inflammation, and cancer: how intimate is the relationship?,” *Annals of the New York Academy of Sciences*, vol. 1171, no. 1, pp. 59–76, 2009.

[37] S. Fletcher, J. A. Drewry, V. M. Shahani, B. D. Page, and P. T. Gunning, “Molecular disruption of oncogenic signal transducer and activator of transcription 3 (STAT3) protein,” *Biochemistry and Cell Biology*, vol. 87, no. 6, pp. 825–833, 2009.

[38] W. Zhao, S. Jaganathan, and J. Turksan, “A cell-permeable Stat3 SH2 domain mimetic inhibits Stat3 activation and induces antitumor cell effects in vitro,” *Journal of Biological Chemistry*, vol. 285, no. 46, pp. 35855–35865, 2010.

[39] H. Zalpoor, M. Bakhtiyari, H. Shapourian, P. Rostampour, E. Madan, S. Prasad, P. Roy, J. George, and Y. Shukla, “Hesperetin as an anti-SARS-CoV-2 agent can inhibit COVID-19-associated cancer progression by suppressing intracellular signaling pathways,” *Inflammopharmacology*, vol. 22, pp. 1–7, 2022.

[40] M. Alipour, N. Nasiri, F. Kazemi, F. Zare, and S. Sharifzadeh, “Resveratrol plus low-dose hydroxyurea compared to high-dose hydroxyurea alone is more effective in γ-globin gene expression and ROS reduction in K562 cells,” *Natural Product Research*, vol. 1-5, pp. 1–5, 2022.

[41] M. Ashrafizadeh, R. Mohammadinejad, T. Farkhondeh, and S. Samarghandian, “Protective effect of resveratrol against glioblastoma: a review,” *Anti-Cancer Agents in Medicinal Chemistry* (Formerly Current Medicinal Chemistry-Anti-Cancer Agents), vol. 21, no. 10, pp. 1216–1227, 2021.

[42] E. Madan, S. Prasad, P. Roy, J. George, and Y. Shukla, “Regulation of apoptosis by resveratrol through JAK/STAT and mitochondria mediated pathway in human epidermoid carcinoma A431 cells,” *Biochemical and Biophysical Research Communications*, vol. 377, no. 4, pp. 1232–1237, 2008.

[43] A. K. Panda, D. Chakraborty, I. Sarkar, T. Khan, and G. Sa, “New insights into therapeutic activity and anticancer properties of curcumin,” *Journal of Experimental Pharmacology*, vol. 9, pp. 31–45, 2017.

[44] H. J. Mehta, V. Patel, and R. T. Sadikot, “Curcumin and lung cancer—a review,” *Targeted Oncology*, vol. 9, no. 4, pp. 295–310, 2014.

[45] Y. Liang, T. Zhang, and J. Zhang, “Natural tyrosine kinase inhibitors acting on the epidermal growth factor receptor: their relevance for cancer therapy,” *Pharmacological Research*, vol. 161, article 105164, 2020.

[46] S. C. Wong, M. N. A. Kamarudin, and R. Naidu, “Anticancer mechanism of curcumin on human glioblastoma,” *Nutrients*, vol. 13, no. 3, p. 950, 2021.

[47] Z. C. Gersev, G. A. Rodriguez, E. Barbarie et al., “Curcumin decreases malignant characteristics of glioblastoma stem cells via induction of reactive oxygen species,” *BMC Cancer*, vol. 17, no. 1, pp. 1–11, 2017.

[48] B. Wu, X. Yao, X. Nie, and R. Xu, “Epigenetic reactivation of RANK in glioblastoma cells by curcumin: involvement of STAT3 inhibition,” *DNA Cell Biology*, vol. 32, no. 6, 2013.

[49] J. Weissenberger, M. Priester, C. Bernreuther et al., “Dietary curcumin attenuates glioma growth in a syngeneic mouse model by inhibition of the JAK1, 2/STAT3 signalling pathway,” *Clinical Cancer Research*, vol. 16, no. 23, pp. 5781–5795, 2010.

[50] M. A. Aziz, M. S. Sarvar, T. Akter et al., “Polyphenolic molecules targeting STAT3 pathway for the treatment of cancer,” *Life Sciences*, vol. 268, article 118999, 2021.

[51] C.-L. Yang, Y.-Y. Liu, Y.-G. Ma et al., “Curcumin blocks small cell lung cancer cells migration, invasion, angiogenesis, cell cycle and neoplasia through Janus kinase-STAT3 signalling pathway,” *PLoS One*, vol. 7, no. 5, article e37960, 2012.

[52] D. Sinha, J. Biswas, B. Seng, B. B. Aggarwal, and A. Bishayee, “Chemopreventive and chemotherapeutic potential of curcumin in breast cancer,” *Current Drug Targets*, vol. 13, no. 14, pp. 1799–1819, 2012.

[53] D. J. Wagstaff, “Dietary exposure to furanocoumarins,” *Regulatory Toxicology and Pharmacology*, vol. 14, no. 3, pp. 261–272, 1991.

[54] L. C. Dolan, R. A. Matulka, and G. A. Burdock, “Naturally occurring food toxins,” *Toxins*, vol. 2, no. 9, pp. 2289–2332, 2010.

[55] S.-M. Kim, J. H. Lee, G. Sehri et al., “Bergamottin, a natural furanocoumarin obtained from grapefruit juice induces chemosensitization and apoptosis through the inhibition of STAT3 signaling pathway in tumor cells,” *Cancer Letters*, vol. 354, no. 1, pp. 153–163, 2014.

[56] J.-H. Ko, F. Arfuso, G. Sehri, and K. S. Ahn, “Pharmacological utilization of bergamottin, derived from grapefruits, in cancer prevention and therapy,” *International Journal of Molecular Sciences*, vol. 19, no. 12, p. 4048, 2018.

[57] W. Luo, Z. Song, H. Sun, J. Liang, and S. Zhao, “Bergamottin, a natural furanocoumarin abundantly present in grapefruit juice, suppresses the invasiveness of human glioma cells via inactivation of Rac1 signaling,” *Oncology Letters*, vol. 15, no. 3, pp. 3259–3266, 2018.

[58] S. Ahmed, H. Khan, M. Aschner, H. Mirzaei, E. Küpeli Akkol, and R. Capasso, “Anticancer potential of furanocoumarins: mechanistic and therapeutic aspects,” *International Journal of Molecular Sciences*, vol. 21, no. 16, p. 5622, 2020.
[94] A. Atiq and I. Parhar, “Anti-neoplastic potential of flavonoids and polysaccharide phytochemicals in glioblastoma,” Molecules, vol. 25, no. 21, p. 4895, 2020.

[95] M.-S. Weng, Y.-S. Ho, and J.-K. Lin, “Chrysins induces G1 phase cell cycle arrest in C6 glioma cells through inducing p21WAF1/CIP1 expression: involvement of p38 mitogen-activated protein kinase,” Biochemical Pharmacology, vol. 69, no. 12, pp. 1815–1827, 2005.

[96] A. Mishra, P. S. Mishra, R. Bandopadhyay et al., “Neuroprotective potential of chrysin: mechanistic insights and therapeutic potential for neurological disorders,” Molecules, vol. 26, no. 21, p. 6456, 2021.

[97] A. Kowalczyk, A. Bodalska, M. Miranowicz, and S. A. Ahmed, D. Parama, E. Daimari et al., “Anti-Cancer Drugs,” vol. 267, article 118814, 2021.

[98] T. A. Stump, B. N. Santee, L. P. Williams et al., “The antiproiferative and apoptotic effects of apigenin on glioblastoma cells,” Journal of Pharmacy and Pharmacology, vol. 69, no. 7, pp. 907–916, 2017.

[99] P. L. Coelho, M. N. Oliveira, A. B. da Silva et al., “The flavonoid apigenin from Croton betulaster Mull inhibits proliferation, induces differentiation and regulates the inflammatory profile of glioma cells,” Anti-Cancer Drugs, vol. 27, no. 10, pp. 960–969, 2016.

[100] B. Santos, M. Oliveira, P. Coelho et al., “Flavonoids suppress human glioblastoma cell growth by inhibiting cell metabolism, migration, and by regulating extracellular matrix proteins and metalloproteinases expression,” Chemico-Biological Interactions, vol. 242, pp. 123–138, 2015.

[101] Y. Wang, Y. S. Xu, L. H. Yin et al., “Synergistic anti-glioma effect of Hydroxyxegkenwain and Apigenin in vitro,” Chemico-Biological Interactions, vol. 206, no. 2, pp. 346–355, 2013.

[102] Y. Li, J. Yao, C. Han et al., “Quercetin, inflammation and immunity,” Nutrients, vol. 8, no. 3, p. 167, 2016.

[103] N. S. Srivastava and R. A. K. Srivastava, “Curcumin and quercetin synergistically inhibit cancer cell proliferation in multiple cancer cells and modulate Wnt/β-catenin signaling and apoptotic pathways in A375 cells,” Phytomedicine, vol. 52, pp. 117–128, 2019.

[104] E. Tavana, H. Mollazadeh, E. Mohtashami et al., “Quercetin: a promising phytochemical for the treatment of glioblastoma multiforme,” BioFactors, vol. 46, no. 3, pp. 356–366, 2020.

[105] H. Zalpooor, M. Bakhitiari, M. Liaghat, M. Nabi-Afjadi, and M. Ganjilkhani-Hakemi, “Quercetin potential effects against SARS-CoV-2 infection and COVID-19-associated cancer progression by inhibiting mTOR and hypoxia-inducible factor-1α (HIF-1α),” Phytotherapy Research, vol. 36, 2022.

[106] H. Zalpooor, M. Nabi-Afjadi, R. Forghanesifdavjani et al., “Quercetin as a JAK–STAT inhibitor: a potential role in solid tumors and neurodegenerative diseases,” Cellular & Molecular Biology Letters, vol. 27, no. 1, p. 60, 2022.

[107] E. Önyay Uçar, A. Şengelen, E. Mertoğlu, M. Pekmez, and N. Arda, “Suppression of HSP70 expression by quercetin and its therapeutic potential against cancer,” in HSP70 in Human Diseases and Disorders, pp. 361–379, Springer, 2018.

[108] L. Jia, S. Huang, X. Yin, Y. Yan, Y. Guo, and L. Han, “Quercetin suppresses the mobility of breast cancer by suppressing glycolysis through Akt-mTOR pathway mediated autophagy induction,” Life Sciences, vol. 208, pp. 123–130, 2018.

[109] M. Lesjak, I. Beara, N. Simin et al., “Antioxidant and anti-inflammatory activities of quercetin and its derivatives,” Journal of Functional Foods, vol. 40, pp. 68–75, 2018.

[110] R. J. Thoppill and A. Bishayee, “Terpenoids as potential chemopreventive and therapeutic agents in liver cancer,” World Journal of Hepatology, vol. 3, no. 9, pp. 228–249, 2011.

[111] Y. Cai, X. Fang, C. He et al., “Curcubitanins: a systematic review of the phytochemistry and anticancer activity,” The American Journal of Chinese Medicine, vol. 43, no. 7, pp. 1331–1350, 2015.

[112] W.-W. Huang, J.-S. Yang, M.-W. Lin et al., “Curcubitan C induces G2/M phase arrest through STAT3/p53/p21 signaling and provokes apoptosis via Fas/CD95 and mitochondria-dependent pathways in human bladder cancer T24 cells,” Evidence-Based Complementary and Alternative Medicine, vol. 2012, Article ID 952762, 11 pages, 2012.

[113] K. T. Chan, K. Li, S. L. Liu, K. H. Chu, M. Toh, and W. D. Xie, “Curcubitin B inhibits STAT3 and the Raf/MEK/ERK pathway in leukemia cell line K562,” Cancer Letters, vol. 289, no. 1, pp. 46–52, 2010.

[114] D. R. Premkumar, E. P. Jane, and I. F. Pollack, “Curcubatin C inhibits Aurora kinase A, Aurora kinase B and survivin, induces defects in cell cycle progression and promotes ABT-737-induced cell death in a caspase-independent manner in malignant human glioma cells,” Cancer Biology & Therapy, vol. 16, no. 2, pp. 233–243, 2015.

[115] G. Yuan, S. Yan, H. Xue, P. Zhang, J. Sun, and G. Li, “JSI-124 suppresses invasion and angiogenesis of glioblastoma cells in vitro,” PLoS One, vol. 10, no. 3, article e0118894, 2015.

[116] X. Lin and A. A. Farooqui, “Curcubitan C mediated regulation of deregulated oncogenic signaling cascades and non-coding RNAs in different cancers: spotlight on JAK/STAT, Wnt/β-catenin, mTOR, TRAIL-mediated pathways,” in Seminars in Cancer Biology, Elsevier, 2021.

[117] F. Blanco-Vaca, L. Cedó, and J. Julve, “Phytosterols in cancer: from molecular mechanisms to preventive and therapeutic potentials,” Current Medicinal Chemistry, vol. 26, no. 37, pp. 6735–6749, 2019.

[118] L. Jiang, X. Zhao, J. Xu et al., “The protective effect of dietary phytosterols on cancer risk: a systematic meta-analysis,” Journal of Oncology, vol. 2019, Article ID 7479518, 11 pages, 2019.

[119] J. Raju and R. Mehta, “Cancer chemopreventive and therapeutic effects of diosgenin, a food saponin,” Nutrition and Cancer, vol. 61, no. 1, pp. 27–35, 2009.

[120] K. C. Nagulapalli Venkata, A. Swaroop, D. Bagchi, and A. Bishayee, “A small plant with big benefits: fenugreek (Trigonella foenum-graecum Linn.) for disease prevention and health promotion,” Molecular Nutrition & Food Research, vol. 61, no. 6, article 1600950, 2017.

[121] D. Parama, M. Boruah, K. Yachna et al., “Diosgenin, a steroidal saponin, and its analogs: effective therapies against different chronic diseases,” Life Sciences, vol. 260, article 118182, 2020.
B. Javadi, P. Kumar, R. Sharma, and N. Garg, “Diosgenin, a steroidal saponin, inhibits STAT3 signaling pathway leading to suppression of proliferation and chemosensitization of human hepatocellular carcinoma cells,” Cancer Letters, vol. 292, no. 2, pp. 197–207, 2010.

D. Bian, Z. Li, H. Ma et al., “Effects of diosgenin on cell proliferation induced by IFN-1 in primary human thyrocytes,” Archives of Pharmacal Research, vol. 34, no. 6, pp. 997–1005, 2011.

J. Li, C. Tang, L. Li, R. Li, and Y. Fan, “Quercetin sensitizes glioblastoma to t-AUCB by dual inhibition of Hsp27 and COX-2 in vitro and in vivo,” Journal of Experimental & Clinical Cancer Research, vol. 35, no. 1, pp. 1–11, 2016.

S. Lamy, A. B. Saad, A. Zgheib, and B. Annabi, “Olive oil compounds inhibit the paracrine regulation of TNF-α-induced endothelial cell migration through reduced glioblastoma cell cyclooxygenase-2 expression,” The Journal of Nutritional Biochemistry, vol. 27, pp. 136–145, 2016.

M. Aparicio-Soto, M. Sánchez-Hidalgo, A. Cárdeno et al., “Dietary extra virgin olive oil attenuates kidney injury in pristane-induced SLE model via activation of HO-1/Nrf-2 and MAPK activation,” The Journal of Nutritional Biochemistry, vol. 27, pp. 278–288, 2016.

B. Javadi, “Diet therapy for cancer prevention and treatment based on traditional Persian medicine,” Nutrition and Cancer, vol. 70, no. 3, pp. 376–403, 2018.

A. Santana-Garrido, C. Reyes-Goya, M. C. Pérez-Camino, H. André, A. Mate, and C. M. Vázquez, “Retinoprotective effect of wild olive (Acebuche) oil-enriched diet against ocular oxidative stress induced by arterial hypertension,” Antioxidants, vol. 9, no. 9, pp. 885, 2020.

M. A. Rosillo, M. Sánchez-Hidalgo, A. González-Benjumea, J. G. Fernández-Bolaños, E. Lubberts, and C. Alarcón-de-la-Lastra, “Preventive effects of dietary hydroxytyrosol acetate, an extra virgin olive oil polyphenol in murine collagen-induced arthritis,” Molecular Nutrition & Food Research, vol. 59, no. 12, pp. 2537–2546, 2015.

P. Kumar, R. Sharma, and N. Garg, “Withania somnifera-A magic plant targeting multiple pathways in cancer related inflammation,” Phytomedicine, vol. 101, article 154137, 2022.

J. Dhami, E. Chang, and S. S. Gambhir, “Withaferin A and its potential role in glioblastoma (GBM),” Journal of Neuro-Oncology, vol. 131, no. 2, pp. 201–211, 2017.

N. Shah, H. Kataria, S. C. Kaul, T. Ishii, G. Kaur, and R. Wadhwa, “Effect of the alcoholic extract of Ashwagandha leaves and its components on proliferation, migration, and differentiation of glioblastoma cells: combinatorial approach for enhanced differentiation,” Cancer Science, vol. 100, no. 9, pp. 1740–1747, 2009.

P. Grogan, K. Sleder, S. Stecklein, and M. Cohen, “Vassobia breviflora root-extract withaferin a as a novel cytotoxic and synergistic agent against malignant gliomas,” Journal of Surgical Research, vol. 165, no. 2, p. 311, 2011.

F. Khomari, M. Nabi-Afjadi, S. Yarahmadi, H. Eskandari, and E. Bahreini, “Effects of cell proosteosis network on the survival of SARS-CoV-2,” Biological Procedures Online, vol. 23, no. 1, pp. 1–10, 2021.

A. B. Baba, R. Nivetha, I. Chattopadhyay, and S. Nagini, “Blueberry and malvidin inhibit cell cycle progression and induce mitochondrial-mediated apoptosis by abrogating the JAK/STAT-3 signalling pathway,” Food and Chemical Toxicology, vol. 109, Part 1, pp. 534–543, 2017.

S. A. Richard, “The therapeutic potential of resveratrol in gliomas,” Advances in Bioscience and Clinical Medicine, vol. 7, no. 2, pp. 44–59, 2019.

A. Ouanouki, S. Lamy, and B. Annabi, “Anticyanidins inhibit epithelial–mesenchymal transition through a TGFβ/Smad2 signaling pathway in glioblastoma cells,” Molecular Carcinogenesis, vol. 56, no. 3, pp. 1088–1099, 2017.

M. Aparicio-Soto, M. Sánchez-Hidalgo, A. González-Benjumea, J. Dhami, E. Chang, and S. S. Gambhir, “The effect of vitamin D, magnesium and zinc supplements on interferon signaling pathways and their relationship to control SARS-CoV-2 infection,” Clinical and Molecular Allergy, vol. 19, no. 1, pp. 1–10, 2021.

E. Bahreini, “Vitamin C as an anticancer agent: regulation of signaling pathways,” Current Topics in Medicinal Chemistry, vol. 20, no. 21, pp. 1868–1875, 2020.

M. A. Dymova, E. V. Kuligina, and V. A. Richter, “Molecular mechanisms of drug resistance in glioblastoma,” International Journal of Molecular Sciences, vol. 22, no. 12, p. 6385, 2021.

E. Muratspahi, M. Freissmuth, and C. W. Gruber, “Nature-derived peptides: a growing niche for GPCR ligand discovery,” Trends in Pharmacological Sciences, vol. 40, no. 5, pp. 309–326, 2019.

S. K. Saxena, V. K. Maurya, S. Ansari et al., “Introductory chapter: high-throughput screening—a new tool for precision medicine,” High-Throughput Screening for Drug Discovery, p. 3, 2022.

E. Muratspahi, M. Freissmuth, and C. W. Gruber, “Nature-derived peptides: a growing niche for GPCR ligand discovery,” Trends in Pharmacological Sciences, vol. 40, no. 5, pp. 309–326, 2019.

R. Vengoji, M. A. Macha, S. K. Batra, and N. A. Shonka, “Natural products: a hope for glioblastoma patients,” Oncotarget, vol. 9, no. 31, pp. 22194–22219, 2018.

R. Saka and N. Chella, “Nanotechnology for delivery of natural therapeutic substances: a review,” Environmental Chemistry Letters, vol. 19, no. 2, pp. 1097–1106, 2021.

F. Sousa, D. Ferreira, S. Reis, and P. Costa, “Current insights on antifungal therapy: novel nanotechnology approaches for drug delivery systems and new drugs from natural sources,” Pharmaceuticals, vol. 13, no. 9, p. 248, 2020.