Chemical and pharmacological evaluations on the extract of *Scutellaria baicalensis* Georgi (Huang-Qin) prepared by various extraction methods

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Abstract

**Background:** This study reported a comprehensive approach (comparing the extraction yields, chemical profiles, antioxidant properties and CYP450-inhibitory effects) to evaluated the effectiveness of various extraction methods [microwave-assisted extraction using water (MAE-W), heat reflux extraction using water (HRE-W), ultrasonic extraction using 70% ethanol and ultrasonic extraction using ethanol (UE-E)] for Huang-Qin (HQ), the dried root of *Scutellaria baicalensis* Georgi.

**Results:** The HQ extraction efficiency by MAE-W was the best. The chemical profiles of extracts obtained using HRE-W and MAE-W were similar; whereas more flavones but less flavone glycosides were detected in the UE-E extract. There was no difference in the antioxidant properties among different extracts. In vitro human liver microsome assays illustrated that all extracts possessed herb–drug interaction potentials but the UE-E extract are shown with a potent interaction with CYP3A4-metabolized drugs.

**Conclusion:** MAE-W is a favorable method for the preparation of HQ extracts based on extraction yield, pharmacological properties and safety.

**Keywords:** *Scutellaria baicalensis* Georgi, Extraction efficiency, Antioxidant properties, Herb–drug interaction

Background

There are different widely available and affordable extraction methods such as heat reflux extraction (HRE), microwave-assisted extraction (MAE) and ultrasonic extraction (UE) for routine herbal extraction.

Comparisons always focus on the yields of crude extracts of the same herb using different extraction methods, but more importantly comparing the pharmacological/toxicological effects of extracts obtained are lacking. In this study, Huang-Qin (HQ), the root of *Scutellaria baicalensis* Georgi, was chosen as the model to evaluate the effectiveness of different extraction methods because its chemical properties and major components have been well studied and information is available in the literature for comparison. A successful search for a cost-efficient (i.e. a better yield of active component with similar/improved therapeutic activities using the same amount of herb) extraction method for this herb can lay down the foundation for future use of this method for other more...
expensive herbs with favourable outcomes both financially and therapeutically.

HQ is a widely used medicinal plant for treatment of various diseases (Chien et al. 2010; Shang et al. 2010); it is also used as food supplement for health-care purposes because of its antioxidant properties (Chan et al. 2008). HQ is rich in flavones and flavone glycosides (baicaerin, wogonin and their glycosides), while baicalin and wogonoside are generally believed to be its major active ingredients. Flavones and flavone glycosides found in HQ are responsible for its reported anti-cancer, anti-inflammatory (Li-WeiBer 2009), vasorelaxant (Huang et al. 2005), anti-viral and other biological activities (Chan et al. 2011; Cui et al. 2010; Liu et al. 2011; Zhang et al. 2015). Currently, there are more than 87 formulations and concoctions containing HQ extract indexed in the Chinese Pharmacopoeia (Comission 2010). As mentioned in the Chinese Pharmacopoeia, HQ is routinely prepared using heat reflux extraction using water (HRE-W) procedures and baicalin is used as reference for quality control purpose. In addition, heat reflux extraction using ethanol (HRE-E), ultrasonic extraction (UE) with organic solvents (and water) and microwave-assisted extraction (MAE) (Horvath et al. 2005; You et al. 2010) are alternative extraction methods that in general provide better yields of HQ crude extract. However, simply looking at the yield of extract obtained is bias because extraction methods making use of ethanol/organic solvent in general result in better yields of extract, but with more hydrophobic products. Moreover, as observed in a Danshen (Salvia Miltiorrhiza) study, extracts prepared by different extraction methods have varied pharmacological effects (Zhou et al. 2012a). Unfortunately, extraction using organic solvent is not a common household practice, and more importantly unwanted or even toxic substances could accumulate in the extract at levels which are high enough to pose serious health hazards, but may exist only in minimal levels when water is used.

The aim of this study was to quantify and compare the yields of HQ extracts given by different extraction methods, namely conventional heat reflux extraction using water (HRE-W), microwave-assisted extraction using water (MAE-W) and ultrasonic extraction using ethanol (70 % vol/vol) (UE-E) or water (UE-W) as solvent. In addition, the chemical profiles of different HQ extracts were characterized, their anti-oxidant potentials were estimated, and the magnitudes of CYP450s-mediated herb–drug interactions were evaluated.

Methods
Reagents and materials
HQ herb was purchased from a TCM distributor (Man-San-Tong, Macao, China) and ground into fine powder for extraction. The voucher sample (Lot. No. 11-803) of HQ herb was authenticated by a botanist, Ms. Huan Zhang, and kept in Prof. Yiu Wa Kwan’s laboratory, School of Biomedical Sciences, Faculty of Medicine, The Chinese University of Hong Kong, Hong Kong. Authentic standards (purity > 95 %) of baicalein and baicalin were purchased from Nanjing Zelang Co. (Nanjing, China); those of wogonoside and wogonin were from Hong Kong Jockey Club Institute of Chinese Medicine (Hong Kong, China). Pooled human liver microsomes (HLMs) were supplied by GenTest Corporation (Woburn, MA, USA). 6-hydroxyflavone, trifluoroacetic acid (TFA, HPLC grade) and other chemicals were purchased from Sigma-Aldrich Chemicals Co. (St. Louis, MO, USA).

Extraction procedures
In order to make an accurate/meaningful comparison, 50 g of ground HQ was immersed in different solvents using various extraction methods. Heat reflux extraction using water (HRE-W) was performed according to the Chinese Pharmacopoeia (Comission 2010) and HQ powder was extracted (100 °C, 2 h). For microwave-assisted extraction using water (MAE-W), HQ powder was put into a 500-ml flask inside a MAS-II microwave oven (Shanghai SINEO Microwave Co., China) with microwave power of 300 W connected to a water-jacket condenser to alter the atmospheric pressure inside the flask if necessary and heated at 100 °C for 2 h. For ultrasonic extraction (UE) using water (UE-W) or ethanol (UE-E), HQ powder was extracted (1 h) at 45 °C with either water or ethanol (70 % vol/vol) with the same ultrasonic frequency (40 kHz) and power (250 W) (Branson 8510 ultrasonicator, Branson, USA). All herbal mixtures obtained using different extraction methods were centrifuged at 5000g for 20 min before the supernatant of each sample was carefully collected. Supernatant obtained from the UE-E method was condensed under reduced pressure to remove ethanol. All extracts collected were then freeze-dried (Scanlab Coolsafe 110-4 Freeze-dryer, LaboGene, Denmark) and weighed for extraction yield calculation before they were ground into powder. All ground powders were stored at −80 °C in a light-proof container before use. Three independent extractions were performed, and these three different batches of samples were individually evaluated in the following assays.

Identification and quantification of chemical profiles
Chemical analysis was carried out using an Agilent 1100 LC/MSD Trap (Agilent, Germany) equipped with a binary pump, an online degasser, an auto-sampler, a Diode Array Detector (DAD) and an APCI source using positive and negative ionization modes. Chromatography was performed at 30 °C with a Grace Alltima C18 column
Different concentrations of FeCl₃ were used to induce cell death in rat heart H9c2 cells (Chan et al. 2010). H9c2 cells were cultured in high glucose DMEM with 10% FBS, 100 units/ml penicillin and 100 μg/ml streptomycin, and seeded into a 96-well plate at a density of 1.5 × 10⁴ cells per well overnight before they were treated with HQ extracts (62.5–500 μg/ml, diluted from 50 mg/ml stock in DMSO) for 24 h. After treatment, the cells were washed twice with warmed PBS, and then 500 μM H₂O₂ diluted in DMEM was added and incubated for 6 h, followed by the addition of 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide (MTT, 0.5 mg/ml in medium) for 3 h. Crystals formed were dissolved in DMSO and the absorbance of the solution was measured at 570 nm. The DMSO (1%)-treated (without H₂O₂ or HQ extracts) group served as solvent control, whereas the H₂O₂-treated (plus 1% DMSO) group was the positive model for determination of cell death. The protective effects of HQ extracts against H₂O₂-induced cell death were estimated and compared based on the calculated EC₅₀ values of different HQ extracts.

Inhibition study on CYP450 isoforms
The inhibitory effects of HQ extracts on the activities of human CYP1A2, CYP2C9, CYP2D6 and CYP3A4 in pooled human liver microsomes (HLMs) were studied using enzymatic reactions with probe substrates (50 μM) as follows: CYP1A2, phenacetin O-deethylation; CYP2C9, tolbutamide 4-hydroxylation; CYP2D6, dextromethorphan O-demethylation; CYP3A4, testosterone 6β-hydroxylation. Incubation, samples preparation and subsequent HPLC analysis were carried out as previously described (Wang et al. 2010; Zhou et al. 2012b). Series of concentration of HQ extracts (1–1000 μg/ml) were prepared and the estimated inhibitory potency was expressed as IC₅₀ value. To estimate the potential in vivo CYP enzymes inhibition caused by HQ extracts, the volume per dose index (VDI, liter per dose unit) was employed and VDI was calculated as the clinical dose unit of HQ extract divided by IC₅₀ value, as reported (Strandell et al. 2004).

Statistical analysis
Data were statistically analyzed with one-way ANOVA followed by Brunnett’s Post Hoc using Prism v4.0 (GraphPad Inc., CA) and the results were presented as mean ± SEM. P value <0.05 was considered statistically significant.

Results and discussion
Based on the dry weights i.e. the yields of the HQ crude extracts obtained (Table 1), MAE-W was the most effective extraction method (crude extract: 390.8 ± 16.6 mg/g) whereas HRE-W was less effective (crude extract: 250 mm × 4.6 mm, 5 μm) protected by a C18 guard column (10 mm × 4.6 mm, 5 μm). The mobile phase constituted (A) 0.05% TFA water (vol/vol) and (B) acetonitrile in a gradient elution: 20% B at 0–10 min, 20–55% B at 15–45 min, 55–80% B at 45–50 min, and 80% B at 50–55 min (Horvath et al. 2005). Injection volume was 50 μl, and flow rate was set at 0.7 ml/min with detection wavelength at 280 nm and reference wavelength at 580 nm. The interface and mass spectrum detector (MSD) parameters employed were as follows: nebulizer pressure, 50 psi; dry gas, 5.0 l/min; dry gas temperature, 350 °C; vaporizer temperature: 400 °C; scan range 100–1100 m/z; target mass, 500 m/z; compound stability, 100%. Stock solutions (10 mg/ml) of all authentic standards were prepared in methanol of analytical grade, and stored at 4 °C before use. Samples of HQ extracts were freshly prepared in water (10 mg/ml). Authentic standards and extract samples were diluted (methanol–water, 50:50, vol/vol) to the desired concentrations. 6-hydroxy-flavone was used as the internal standard (I.S.), and 10 μl of I.S. (50 μg/ml) was used with 50 μl of individual sample. The quantitative method used in this study was validated based on the detection of baicalin, baicalein, wogonoside and wogoninin.

Evaluation of antioxidant properties
The free radicals scavenging capacities (SC) were measured and compared using the DPPH assay (Chan et al. 2010). Briefly, 5 μl of individual extract (0.625–10 mg/ml) was mixed with 195 μl of DPPH solution (24 mg/l) in a 96-well plate. The reaction was allowed to proceed in the dark for 30 min before the absorbance of the resultant solution was measured at 515 nm using a Micro-plate Absorbance Reader (Bio-Rad, CA, USA). Ascorbic acid (Sigma-Aldrich Chemicals Co., MO, USA) in distilled water served as the positive control. SC₅₀ was estimated as the concentration of sample which scavenges 50% of free radicals.

The FRAP assay was performed in a 96-well plate as described ( Firuzi et al. 2005). Briefly, 5 μl of each sample was mixed with 150 μl of freshly prepared FRAP reagent (250 mM acetate buffer, pH 3.6; 8.3 mM 2,4,6-tripryridyl-s-triazine dissolved in 40 mM HCl and 16.7 mM FeCl₃·6H₂O solution). After incubation at 37 °C for 30 min, the absorbance of the solution was measured at 593 nm. Different concentrations of Fe²⁺ standard solution were used for the construction of the standard curve, and ascorbic acid was used as the positive control.

Comparison of the protective effects of HQ extracts on hydrogen peroxide-induced cell death
The protective effects of HQ extracts on hydrogen peroxide (H₂O₂)-induced cell death were evaluated using rat heart H9c2 cells (Chan et al. 2010). H9c2 cells were cultured in high glucose DMEM with 10% FBS, 100 units/ml penicillin and 100 μg/ml streptomycin, and seeded into a 96-well plate at a density of 1.5 × 10⁴ cells per well overnight before they were treated with HQ extracts (62.5–500 μg/ml, diluted from 50 mg/ml stock in DMSO) for 24 h. After treatment, the cells were washed twice with warmed PBS, and then 500 μM H₂O₂ diluted in DMEM was added and incubated for 6 h, followed by the addition of 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide (MTT, 0.5 mg/ml in medium) for 3 h. Crystals formed were dissolved in DMSO and the absorbance of the solution was measured at 570 nm. The DMSO (1%)-treated (without H₂O₂ or HQ extracts) group served as solvent control, whereas the H₂O₂-treated (plus 1% DMSO) group was the positive model for determination of cell death. The protective effects of HQ extracts against H₂O₂-induced cell death were estimated and compared based on the calculated EC₅₀ values of different HQ extracts.
obtained when ethanol (70 % vol/vol) was used as sol-
vent, whereas only 275.0 ± 9.0 μg/mg of crude extract was obtained when water was used. Our results clearly illustrate that among all the extraction methods employed in this study, MAE-W was the most efficient method for herbal extraction based on the yield of HQ crude extract.

Figure 1 illustrates the HPLC–UV (at 280 nm) of base peak chromatographic (BPC) profiles of authentic standards used. Nine major HPLC peaks were detected and identified as flavones in the HPLC chromatogram. Compounds 3, 6, 7 and 8 were identified as baicalin, wogonoside, baicalein and wogonin, respectively. Other components were identified using APCI-MS methods (Table 2). The glycoside (e.g. baicalin) exhibited a significant MS signal in both positive and negative modes, whereas aglycone (e.g. baicalein) was observed only in positive mode. In the MS spectra, all analyzed compounds exhibited a noticeable protonated [M + H]+ ion in positive mode. However, in negative mode, no consistent pattern was detected. The O-linked glycosides such as baicalein and wogonoside exhibited an intensive adduct ion [M + TFA]− which produced a [M − H]− ion in MS2. The multi-stage MS of [M + H]+ or [M − H]− of O-linked glycosides provided information about the loss of sugar moiety, and the structure of aglycones was formed. Compounds 4 and 5 were O-linked glycosides, and they were identified as norwogonin-7-O-glucuronide (norwogonin-7-O-GlcA) and chrysin-7-O-glucuronide (chrysin-7-O-GlcA) respectively as previously reported (Han et al. 2007; Liu et al. 2009). On the other hand, Compounds 1 and 2 demonstrated a dominant pseudo-molecular ion [M − H]− in their respective MS spectra, illustrating the existence of [M − H − 60]−, [M − H − 90]− and [M − H − 120]− ions, which are the characteristics of typical C-linked glycosides (Han et al. 2007; Liu et al. 2009). Moreover, the MSn data suggested that Compound 1 was chrysin-6-C-arabinosyl-8-C-glucoside (chrysin-6-C-Ara-8-C-Glc) and Compound 2 was chrysin-6-C-glucosyl-8-C-arabinoside (chrysin-6-C-Glc-8-C-Ara). Compound 9 was an isomer of Compound 8 (wogonin) and it was probably oroxylin A according to the retention time and MS fragments.

Table 1 Summary of the yields of crude extracts (dry weight, mg) obtained from ground Huang-Qin (50 g) using different extraction methods

| Extraction methods                              | Abbreviation | Crude extract (mg/g herb) |
|------------------------------------------------|--------------|--------------------------|
| Heat reflux extraction using water              | HRE-W        | 297.4 ± 28.4             |
| Microwave-assisted extraction using water       | MAE-W        | 390.8 ± 16.6***          |
| Ultrasonic extraction using ethanol (70 % vol/vol) | UE-E         | 327.4 ± 19.4             |
| Ultrasonic extraction using water               | UE-W         | 275.0 ± 90               |

Results are expressed as mean ± SEM (n = 3)

*** P < 0.001 compared to HRE-W

297.4 ± 28.4 mg/g) (i.e. MAE-E versus HRE-W: ~30 % increase). Using the UE method (a widely-used efficient method in a previous quality-control study on HQ) (Li et al. 2009), 327.4 ± 19.4 mg/g of HQ crude extract was obtained when ethanol (70 % vol/vol) was used as solvent, whereas only 275.0 ± 9.0 μg/mg of crude extract was obtained when water was used. Our results clearly illustrate that among all the extraction methods employed in this study, MAE-W was the most efficient method for herbal extraction based on the yield of HQ crude extract.

The chemical profiles and quantity of four marker compounds in HQ extracts obtained are shown in Fig. 2a. Our results demonstrated that MAE-W (baicalin, 190.1 ± 4.4 μg/mg; wogonoside, 37.7 ± 1.7 μg/mg; baicalein, 21.8 ± 2.1 μg/mg; wogonin, 7.1 ± 0.2 μg/mg) and HRE-W (baicalin, 186.0 ± 6.4 μg/mg; wogonoside, 37.8 ± 1.5 μg/mg; baicalein, 22.0 ± 1.8 μg/mg; wogonin, 7.3 ± 0.1 μg/mg) methods gave similar results in terms of the quantity of these compounds. In addition, these two extraction methods did not affect the chemical profiles of the HQ extracts obtained with a constant ratio of 5:1 for baicalin/wogonoside. On the other hand, HQ extract contained less baicalin (163.0 ± 6.3 μg/mg) but more baicalein (48.2 ± 7.6 μg/mg) and wogonin (15.6 ± 1.5 μg/mg) when UE-E was used. These results are consistent with the observations of some previous studies on HQ extraction using organic solvent (Horvath et al. 2005; Lin et al. 1996). However, when water was used as solvent in the UE procedures, HQ extract obtained had the least amount of baicalin (0.8 ± 0.3 μg/mg) and wogonoside (0.8 ± 0.3 μg/mg) among all extraction methods used in this study.

Besides, there were no significant differences in the amount of water-soluble components (e.g. chrysin-6-C-Ara-8-C-Glc, chrysin-6-C-Glc-8-C-Ara, norwogonin-7-O-GlcA and chrysin-7-O-GlcA) and lipid components (like oroxylin A) present in the extracts prepared by MAE-W and HRE-W (Fig. 2b). As expected, using UE with ethanol, the yield of most water-soluble components were lower compared with those prepared by MAE-W and HRE-W (e.g. 19 % decrease of baicalin), whereas higher yields of the major lipid-soluble components (e.g. baicalein, 119 %; wogonin, 79 %; oroxylin A, 197 %) were observed probably due to the higher solubility of these compounds in ethanol. When water was used as solvent in UE, the least amount of these identified compounds
could be obtained. These results therefore suggested that based on the profiles and quantity of all chemicals detected in the extracts, the MAE-W method is as good as the commonly used HRW-E method, whereas the efficiency and effectiveness of the UE method is highly dependent on the solvent used.

Baicalein, wogonin and their glycosides contribute significantly to the total weight of HQ (Su et al. 2008),
and these flavones are strong antioxidants. It is therefore feasible to utilize both the chemical profiles and the antioxidant properties of HQ extract for quality control purposes for all HQ-containing products in the market (Boyle et al. 2011). Table 3 illustrates that there were no significant differences in the free radicals scavenging activities (as estimated by DPPH assay) of extracts prepared by the extraction methods (SC_{50}: MAE-W, 1.046 ± 0.064 mg/ml; HRE-W, 0.958 ± 0.032 mg/ml; UE-E, 0.988 ± 0.057 mg/ml). However, extract prepared by the UE-W method has the weakest scavenging activity (SC_{50}: 3.899 ± 0.175 mg/ml), whereas ascorbic acid is the strongest scavenger (SC_{50}: 0.107 ± 0.004 mg/ml).

The results obtained from the FRAP assay demonstrated a similar trend of antioxidant properties of HQ extracts prepared by different extraction methods (Table 3).

Due to the relatively low yield (the lowest yield among all extraction methods) of bioactive components and antioxidant activity of the extract obtained, water extract of HQ obtained from UE was therefore not evaluated. As illustrated in Table 4, all HQ extracts (HRWE, MAE-W and UE-E) demonstrated a similar protective effect (EC_{50} of 176.4 ± 14.5, 190.6 ± 12.1 and 165.7 ± 19.9 μg/ml, respectively) against H_{2}O_{2}-induced cell death as determined by MTT assay. Interestingly, these cell-based results were comparable to the results of antioxidant/free radicals scavenging assays as discussed in previous sections.

There is little information in the literature that compares the hepatic enzymes inhibitory effects of HQ extracts prepared by different methods. Some HQ flavones, which are abundant especially when using ethanol as solvent for extraction, demonstrated substantial inhibition of human hepatic drug metabolizing enzymes cytochrome P450s (CYP 450s) such as CYP2C9 (Si et al. 2009), CYP3A4, CYP1A2 (Kim et al. 2002; Li et al. 2011) and CYP2D6 (Mo et al. 2012). Therefore, it is important to evaluate whether the extract obtained using MAE-W method has different inhibitory profiles compared with other extraction methods on hepatic CYP450 enzymes. HQ extract obtained by the HRE-W method was used as reference for comparison.

Extracts obtained by MAE-W demonstrated a moderate inhibition of CYP1A2 and CYP2C9 (as indicated by 9 and 20 % decrease in IC_{50} values, respectively) whereas an “enhancement” rather than an inhibition (i.e. ~ 40 %

### Table 2 Characteristics of major ingredients in Huang-Qin extract by HPLC–DAD–APCI–MS

| Peak no. | f_{R} (min) | UV \( \lambda_{max} \) | MS\(^{a} \) (m/z) and relative abundance (% base peak) | Identification |
|----------|-------------|-------------------------|------------------------------------------------|----------------|
| 1        | 14.0        | 215, 275, 318           | APCI⁺—MS: 549(100); MS\(^{2}\) 549: 531(70), 513(60), 495(90), 483(20), 465(45), 429(10), 411(100), 393(30), 363(10); MS\(^{3}\) [411]: 393(100), 375(60), 363(70), 333(80), 309(30), 279(28) | Chrysin-6-C-arabinosyl-8-C-glucoside\(^{c}\) |
| 2        | 17.6        | 215, 275, 318           | APCI⁺—MS: 549(100); MS\(^{2}\) 549: 531(100), 513(30), 495(20), 483(45), 465(20), 453(18), 441(18), 411(10), 393(30), 381(10), MS\(^{3}\) [531]: 513(80), 495(60), 483(100), 465(75), 453(70), 441(60), 411(12), 393(30), 381(50), 363(40) | Chrysin-6-C-glucosyl-8-C-arabinoside\(^{c}\) |
| 3        | 27.3        | 215, 277, 316           | APCI⁺—MS: 447(100), 271(20); MS\(^{2}\) 447[447]: 271(100); MS\(^{3}\) [271]: 253(40), 241(100) | Baicalin\(^{b}\) |
| 4        | 29.7        | 215, 280, 360           | APCI⁺—MS: 447(100), 271(20); MS\(^{2}\) 447[447]: 271(100) | Norwogonin-7-O-glucuronide\(^{c}\) |
| 5        | 30.8        | 215, 273, 312           | APCI⁺—MS: 431(100), 255(20); MS\(^{2}\) 431[431]: 255(100) | Chrysin-7-O-glucuronide\(^{c}\) |
| 6        | 32.3        | 215, 275, 349           | APCI⁺—MS: 461(100); MS\(^{2}\) 461: 285(100); MS\(^{3}\) [285]: 270(100) | Wogonoside\(^{b}\) |
| 7        | 40.1        | 215, 275, 322           | APCI⁺—MS: 271(100); MS\(^{2}\) [271]: 241(100) | Baicalein\(^{b}\) |
| 8        | 48.0        | 210, 275                | APCI⁺—MS: 285(100); MS\(^{2}\) [285]: 270(100) | Wogonin\(^{b}\) |
| 9        | 50.5        | 215, 273, 318           | APCI⁺—MS: 285(100); MS\(^{2}\) [285]: 270(100) | Oroxylin A\(^{c}\) |

\(^{a}\) The peak number is the same as indicated in Fig. 1  
\(^{b}\) Identities of compounds were established by comparison with the authentic compounds  
\(^{c}\) On-line identification by HPLC–MS\(^{n}\) analysis
increase in IC$_{50}$ value) of CYP3A4 activity was observed (Table 5). There was a similar magnitude of CYP2D6 inhibition caused by HQ extracts obtained by MAE-W and HRE-W. On the other hand, extract obtained by UE-E had a marked inhibition on CYP1A2, CYP2C9 and CYP2D6 (as illustrated by 63 to 71 % decrease in IC$_{50}$ values) and only a weak inhibition (~39 % decrease in IC$_{50}$) of CYP3A4 was observed. Previous studies reported that HQ extract caused CYP450s inhibition mainly through the flavones and flavones glycosides present in the extract (Kim et al. 2002; Li et al. 2011). However, CYP1A2 inhibition by flavone glycosides of HQ extracts has not been reported.

Volume per dose index (VDI) has been employed for the estimation of possible in vivo herb–drug interactions in in vitro incubation using human pooled liver

![Fig. 2](https://example.com/fig2.png) Effects of extraction methods on a the contents of marker compounds and b the estimated contents of other identified ingredients in Huang-Qin extracts. Results are expressed as mean ± SEM (n = 3). **P < 0.01 and ***P < 0.001 compared to HRE-W
microsomes. As proposed by Strandell et al. (2004), when VDI is greater than 4 l (i.e. human blood volume), it is likely that in vivo herb–drug interactions occurred with the participation of hepatic CYP enzymes. According to the National Drug Standards (Traditional Chinese Medicine Prescription, Volume 2) (Ministry of Health, PR of China), the dose unit of HQ tablets is about 650 mg HQ extract in which 117 mg is baicalin. Based on this dose unit and the respective IC50 values, the VDI of individual CYP450 isoenzymes were estimated (Table 5). The estimated VDIs of all HQ extracts examined on CYP1A2, CYP2C9 and CYP2D6 enzymes were greater than 4 l per dose unit. More importantly, based on the estimated VDIs, only the extract obtained using UE-E method illustrated a significant in vivo herb–drug interaction with CYP3A4-metabolized Western drugs. Taken together, our results clearly demonstrated that HQ extracts, irrespective of the extraction methods/solvents used, possess significant herb–drug interactions with CYPs-metabolized Western medicines.

Recent pharmacological and toxicological studies indicated that water-soluble components of HQ extract exhibit a variety of therapeutically useful activities with relatively low toxicity (Yimam et al. 2010), whereas long-term toxicity (e.g. damage to the bile duct) of extracts prepared using organic solvent has been reported (Ong et al. 2004) although herbal extraction using organic solvent (e.g. UE-E in this study which possesses a significant herb–drug interaction with CYP3A4-metabolized drugs) is not a common practice for human consumption. Taken together, our results illustrate that MAE-W provided an effective and efficient procedure in terms of a higher yield of crude extract collected than the conventional HRE-W, and more importantly HQ extract prepared by this procedure has similar pharmacological activities with a lesser magnitude of herb–drug interactions when compared with extracts prepared using other organic-solvent extraction methods. It remains to be determined whether this favorable cost-efficient extraction method could be used in preparation of extracts of more expensive Chinese herb such as Cordyceps for therapeutic uses without affecting their pharmacological effects.

Table 3: Summary of the antioxidant potential of Huang-Qin extracts prepared by different extraction methods as estimated by DPPH assay (SC50, mg/ml) and FRAP assay (μmol Fe2+/mg extract)

| Samples  | DPPH assay (SC50, mg/ml) | FRAP assay (μmol Fe2+/mg extract) |
|----------|--------------------------|----------------------------------|
| Ascorbic acid | 0.107 ± 0.004*** | 2278 ± 15*** |
| HRE-W | 0.958 ± 0.032 | 1249 ± 131 |
| MAE-W | 1.046 ± 0.064 | 1306 ± 71 |
| UE-E | 0.988 ± 0.057 | 1359 ± 29 |
| UE-W | 3.899 ± 0.175*** | 5416.6 ± 98*** |

Results are expressed as mean ± SEM (n = 3)

*** P < 0.001 compared to HRE-W

Table 4: Summary of the protective effects [expressed as EC50 (μg/ml)] of Huang-Qin extracts prepared by different extraction methods against H2O2-induced cell death (estimated by MTT assay) using rat heart H9c2 cells

| Samples | EC50 (μg/ml) |
|---------|-------------|
| HRE-W | 1764.4 ± 14.5 |
| MAE-W | 1906.0 ± 12.1 |
| UE-E | 1657.8 ± 19.9 |

Results are expressed as mean ± SEM (n = 3)

Table 5: Summary of the inhibitory effects [expressed as IC50 (μg/ml) and volume per dose index (VDI, L/dose unit)] of Huang-Qin (HQ) extracts prepared by different extraction methods on model probes’ metabolism

| Samples | CYP1A2 | CYP2C9 | CYP2D6 | CYP3A4 |
|---------|--------|--------|--------|--------|
|        | IC50   | VDI    | IC50   | VDI    | IC50   | VDI    | IC50   | VDI    |
| HRE-W  | 8.13 ± 0.08 | 80.0   | 38.87 ± 0.60 | 16.7   | 131.3 ± 5.35 | 4.95   | 174.1 ± 4.86 | 3.74   |
| MAE-W  | 7.42 ± 0.20* | 87.6   | 30.71 ± 0.84*** | 21.2   | 135.3 ± 6.43 | 4.80   | 243.0 ± 11.81** | 2.68   |
| UE-E   | 3.00 ± 0.12*** | 216.7  | 11.25 ± 0.66*** | 57.8   | 38.29 ± 1.28*** | 17.0   | 106.6 ± 4.53** | 6.10   |

Positive controls

|       | 0.640 | –      | 0.138 | –      | 0.292 | –      | 0.047 | –      |

Results are expressed as mean ± SEM (n = 3)

* P < 0.05; ** P < 0.01 and *** P < 0.001 compared to HRE-W

Furafylline for CYP1A2; Sulfaphenazole for CYP2C9; Quinidine for CYP2D6; Ketoconazole for CYP3A4. Results of the positive controls included in this Table for comparison have been published in our previous work (Wang et al. 2010; Zhou et al. 2012b)
Horváth CR, Matsos PA, Saxena PK (2005) Identification and quantification of eight flavonoids in root and shoot tissues of the medicinal plant Huang-Qin (Scutellaria baicalensis Georgi) using high-performance liquid chromatography with diode array and mass spectrometric detection. J Chromatogr A 1062(2):199–207. doi:10.1016/j.jchroma.2001.02.050

Huang Y, Tsang SY, Yao X, Chen ZY (2005) Biological properties of bacosin in cardiovascular system. Curr Drug Targets Cardiovasc Haematol Disord 5(2):177–184. doi:10.2174/1568006054386206

Kim JY, Lee S, Kim DH, Kim BR, Park R, Lee BM (2002) Effects of flavonoids isolated from Scutellaria radix on cytochrome P-450 activities in human liver microsomes. J Toxicol Environ Health A 65(6–7):373–381. doi:10.1080/109365102901228994

Li C, Zhou L, Lin G, Zuo Z (2009) Contents of major bioactive flavones in proprietary traditional Chinese medicine products and reference herb of Radix Scutellariae. J Pharm Biomed Anal 50(3):298–306. doi:10.1016/j.jpba.2009.04.028

Li T, Li N, Guo Q, Ji H, Zhao D, Xie S, Li X, Qiu Z, Han D, Chen X, You Q (2011) Inhibitory effects of Wogonin on catalytic activity of cytochrome P450 enzyme in human liver microsomes. Eur J Drug Metab Pharmacokinet 36(4):249–256. doi:10.1016/j.sjpbi.2011.09.002

Lin SJ, Tieng HH, Wen KC, Suen TT (1996) Determination of gentiopicroside, mangiferin, palmitate, berberine, baicalin, wogonin and glycyrrhizin in the traditional Chinese medicinal preparation Sann-Joong-Kuey-Jan-Tang by high-performance liquid chromatography. J Chromatogr 730(1):17–23. doi:10.1016/0021-9673(95)00761-7

Li‑Weber M (2009) New therapeutic aspects of flavones: the anticancer properties of Scutellaria and its main active constituents Wogonin, Baicalein and Baicalin. Cancer Treat Rev 35(1):57–68. doi:10.1016/j.ctrv.2008.09.005

Mo SL, Liu WF, Chen Y, Luo HB, Sun LB, Chen XM, Zhou ZW, Sneed KB, Li CG, Du YM, Liang J, Zhou SF (2012) Ligand- and protein-based modeling studies of the inhibitors of human cytochrome P450 2D6 and a virtual screening for potential inhibitors from the Chinese herbal medicine, Scutellaria baicalensis (Huangqin, Baiqal Skullcap). Comb Chem High Throughput Screen 15(1):36–80. doi:10.2174/13862071298280826

Ong ES, Len SM, Lee AC, Chui P, Chooi KF (2004) Proteomic analysis of mouse liver for the evaluation of effects of Scutellaria radix by liquid chromatography with tandem mass spectrometry. Rapid Commun Mass Spectrom 18(1):2522–2530. doi:10.1002/rcm.1654

Shang X, He X, Li M, Zhang R, Fan P, Zhang Q, Jia Z (2010) The genus Scutellaria as an ethnopharmacological and phytochemical resource. J Ethnopharmacol 128(2):279–313. doi:10.1016/j.jep.2010.01.006

Si D, Wang Y, Zhou YH, Guo Y, Wang J, Zhou H, Li ZS, Fawcett JP (2009) Mechanism of CYP3A9 inhibition by flavones and flavonols. Drug Metab Dispos 37(3):629–634. doi:10.1124/dmd.108.023416

Strandell J, Neil A, Carlin G (2004) An approach to the in vitro evaluation for potential inhibitors from the Chinese herbal medicine, Scutellaria baicalensis (Huangqin, Baiqal Skullcap) for potential inhibitors from the Chinese herbal medicine, Scutellaria baicalensis (Huangqin, Baiqal Skullcap).

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Zhang SF, Dong YC, Zhang XF, Wu XG, Cheng JJ, Guan LH, Shang YZ (2015) Flavonoids from Scutellaria attenuate okadaic acid-induced neuronal damage in rats. Brain Injury 29(11):1376–1382. doi:10.3109/02699052.2015.1042053

Zhou X, Chan SW, Tseng HL, Deng Y, Hoi PM, Choi PS, Or PM, Yang JM, Lam FF, Lee SM, Leung GP, Kong SK, Ho HP, Kwan YW, Yeung JH (2012a) Danshensu is the major marker for the antioxidant and vasorelaxation effects of Danshen (Salvia miltiorrhiza) water-extracts produced by different heat water-extractions. Phytomedicine 19(14):1263–1269. doi:10.1016/j.phymed.2012.08.011

Zhou X, Wang Y, Or PM, Wan DC, Kwan YW, Yeung JH (2012b) Molecular docking and enzyme kinetic studies of dihydrotanshinone on metabolism of a model CYP2D6 probe substrate in human liver microsomes. Phytomedicine 19(7):648–657. doi:10.1016/j.phymed.2012.01.005