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Biological response to geochemical and hydrological processes in a shallow submarine cave

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Abstract

The Croatian coastal karst abounds in submerged caves that host a variety of environmental conditions depending on the geomorphology, depth and submarine groundwater discharge. One example is the Y-Cave, a shallow, mostly submerged, horizontal cave on Dugi Otok Island, on the eastern Adriatic coast. This study was aimed at examining the temporal and spatial variability of the marine cave environment, including temperature, salinity, light intensity, cave morphology and hydrodynamism, along with the dissolulutional effect caused by the mixing of sea and freshwater. The general distribution of organisms in the Y-Cave was positively correlated to the light gradient and reduced water circulation, thus the highest species diversity and abundance were recorded in the front part of the cave. The phylum Porifera was the most dominant group, and the pariferan species diversity in the cave ranks among the ten highest in the Mediterranean. The middle part of the cave, although completely dark, hosts an abundant population of the gastropod Homalopoma sanguineum and clusters of the gregarious brachiopod Novocrania anomala, whose presence could be connected to tidal hydrodynamics. The absence/scarcity of sessile marine organisms and pronounced corrosion marks at shallow depths inside the cave suggest a freshwater impact in the upper layers of the water column. A year long experiment with carbonate tablets revealed three different, independent ongoing processes affected by the position in the cave: bioaccumulation, dissolution and mechanical erosion. The results of long-term temperature readings also revealed water column stratification within the cave, which was not disturbed by either tidal or wave action. The shallow, partly submerged and relatively small Y-Cave is characterised by a suite of complex environmental conditions, which, together with the resulting distribution of organisms, are unique to this cave.

Keywords: Submerged cave, karst, mixing corrosion, biodiversity, Porifera, Adriatic Sea.

Introduction

Caves are natural underground voids usually formed in karst. They exhibit a unique environment generally characterized by stable environmental conditions throughout the year – primarily air temperature (equivalent to the mean annual surface atmosphere temperature) and relative humidity (approaching 100%). They are oligotrophic, aphotic subterranean habitats inhabited by scarce cave biota adapted to reduced light and near-constant temperature regimes, and are dependent on periodic inputs of food, usually swept in by floods (Gillieson, 1996). Once drowned by the rising sea, submerged caves within the marine environment also host relatively stable conditions regarding temperature and water chemistry. Throughout the transition of the cave from completely subaerial to an entirely marine environment, extreme conditions occur due to the interaction of different geochemical and hydrogeological settings. In particular, in the coastal zone, groundwater and sea levels oscillate in near synchrony, so as sea-levels rise, coastal caves experience a transition through vadose, littoral, anchialine and (sub)marine conditions (Surić et al., 2005; Van Hengstum et al., 2011). One of the most pronounced geochemical effects is produced by the interaction of fresh water and sea water, known as mixing corrosion (introduced as Mischungkorrossion by Bögli (1964)). The mixing of two solutions saturated at different partial pressures of CO₂ yields a third solution that is undersaturated with respect to calcite (Frank et al., 1998; Kaufmann & Dreybrodt, 2007) (Fig. 1). After a relatively long time underground, groundwater (H₂O-CO₂-CaCO₃ solution) that percolates within the epikarst and emerges in the cave is commonly supersaturated with respect to calcite (Fairchild & Baker, 2012), enabling the precipitation of speleothems. Sea water is also supersaturated with CaCO₃ down to the first 500 m (Kennett, 1982), enabling marine organisms to build their shells. The mixing of these two waters takes place in the coastal zone at the interface where denser sea water intrudes into the karst aquifer as a wedge under a fresh groundwater lens, and it is responsible for the dis-
solutions effects that influence carbonate in caves within the mixing zone, regardless of whether it is primary limestone bedrock, secondary deposited flowstone or organogenic carbonate produced by marine organisms.

As dissolution is one of the main processes of karstification, different methods have been used in order to assess its temporal and spatial dimensions, as outlined in Gabrovšek (2009). One such method is the weight loss measurements of “standard” limestone tablets exposed to dissolution in different environments, though the very few data published have exclusively regarded dissolution in continental environments (Gams, 1985; Plan, 2005; Zhang, 2011; Buzjak et al., 2013; Krklec et al., 2013 and references therein). Dissolution at the groundwater-sea-water interface has not previously been recorded using the method of limestone tablets, despite the suggestion that dissolution may be a principal mechanism of cave and karst development in the coastal zone (Myrloie & Myroie, 2007). With more than 235 known submerged caves (Surić et al., 2010), and thousands likely yet to be explored, the Croatian submerged karst offers exceptional potential for such studies.

From the biological point of view, the variety of coastal and marine geomorphic features offers a great diversity of submerged karst habitats (Bakran-Petricioli & Petricioli, 2008), such as completely or partially submerged pits, caves and submarine passages. Marine caves have unique faunistic and ecological characteristics, and the organisms and communities inhabiting them exhibit distinctive zonation (Pérès & Picard, 1964; Riedl, 1966; Harmelin et al., 1985; Bianchi & Morri, 1994; Bianchi, et al. 1996; Harmelin, 1997). The decreasing species richness, abundance and biomass from the entrance towards the end of marine caves is explained by a gradient of environmental factors, primarily the attenuation of light and hydrodynamics (Riedl, 1966; Cinelli et al., 1977; Harmelin et al., 1985; Gili et al., 1986; Balduzzi et al., 1989; Rosso et al., 2012). However, a study by Zabala et al. (1989) showed in an examined cave in the Western Mediterranean that zonation cannot be explained simply with a decrease in water motion and food supply. Moscatello & Belmonte (2007) also noted that although the innermost portion of a cave in the southern Adriatic showed benthic faunal depletion, a rich presence of plankton was recorded there. Recent studies using stable isotope signatures to investigate cave mysid feeding strategies have shed more light on resource partitioning in marine caves (Rastorgueff et al., 2011).

Classically, two distinctive bioenoses could be recognized within marine caves (Pérès & Picard, 1964): the biocoenosis of semi-dark caves (biocoenose des grottes semi-obscurces, GSO) and the biocoenosis of caves and ducts in total darkness (biocoenose des grottes et boyaux à obscurité totale, GO). GSO is located near the cave entrances and is characterized by high species diversity and biomass, dominated by massive sponges, cnidarians (class Anthozoa) and branched bryozoans. GO occupies the innermost parts of the caves, where the biodiversity and biomass are much lower, and the fauna is characterized by encrusting sponges, bryozoans and sessile polychaetes. In the latter bioenoses, rocks are usually barren and blackened by iron and manganese oxides of bacterial origin. Completely dark caves, especially those that trap cold seawater, can be considered extensions of the bathyal zone. They are often inhabited by deep sea organisms and are thus very interesting scientifically (Harmelin et al., 1985; Vacelet et al., 1994; Bakran-Petricioli et al., 2007; Janssen et al., 2013).

Different studies have shown that within caves there are transitions between the bioenoses of semi-dark and dark caves. Laborel & Vacelet (1959) described a transition zone between GSO and GO where sponges and stony corals are still present, though species more characteristic for GSO begin to disappear, and species specific to GO begin to appear. Harmelin et al. (1985) described a transition zone between the two bioenoses, which is dominated by bryozoan nodules. Similar structures have been found in other caves (Rosso et al., 2013).

A spatial variation in benthic assemblages was recorded in the span of a few meters within a marine cave (Benedetti-Cecchi et al., 1996). The study of two caves in the north-western Mediterranean revealed that the distinct morphology of the cave, i.e. its depth and geographic location, actually influence water movement, food availability and light penetration, and their interplay causes differences in the communities (Martí et al., 2004). Inconsistent spatial and temporal changes in assemblages, as a result of multiple and interactive ecological processes, were also recorded in three shallow marine caves in southern Italy (Bussotti et al., 2006).

Marine caves are recognized as habitats important for conservation. They have been included in the Habitats Directive as Special Areas of Conservation (SAC) and Special Protection Areas (SPA) under the European Union’s Habitats Directive.
tats Directive of the European Union (Council Directive 92/43/EEC) and are also protected in Croatia (Bakran-Petricioli, 2011). Their poor resilience and vulnerability to different disturbances from diving to climate change has already been well documented (Chevaldonné & Lejeusne, 2003; Parravicini et al., 2010; Guarnieri et al., 2012; Giakoumi et al., 2013). This study provides a report of the settings where hydrologically and geomorphologically induced geochemical features significantly influence biological distribution in the Y-Cave, a shallow coastal cave partially submerged by seawater on the eastern Adriatic coast. The aim of this study was to determine the distribution of benthic organisms and communities along the cave conduits in relation to cave morphology, temperature, light intensity, hydrodynamics and geochemical settings, and to assess how the dissolutional effect in the mixing zone influences not only biological components, but also the cave environment as a whole.

**Study area and environmental settings**

The eastern Adriatic coast is part of the Dinaric Classical karst formed within the up to 8 km thick succession of shallow-water carbonate sediments deposited mostly during the Mesozoic and the Paleogene (Vlahović et al., 2005). The emersion of the carbonate bedrock along with intensive folding and faulting generated by the Alpine orogeny was followed by overall karstification. A vast part of that karstified area was subsequently submerged by the last late Pleistocene–Holocene sea transgression, transforming originally subaerial karst features (karrens, dolines, karstic springs, etc.) into submarine ones, including numerous caves (Pikelj & Juračić, 2013). Presently, of more than 235 known submerged caves, more than half are completely marine (eualine), whilst environmental conditions in others (mainly coastal caves) vary depending on depth, tide and seaward freshwater discharge (Surić et al., 2010). One such cave is the Y-Cave (44°03'27.3" N; 14°59'04.8" E), located in the central Adriatic on the south-western coast of Dugi Otok Island near Brbišćica Cove (Fig. 2), with the entrance at 12 m below the mean sea level (Juračić et al., 2002). It is a shallow, almost completely submerged cave, except one small chamber with an air pocket (profile C-C’ in Fig. 2) developed above the junction of the two main channels. The cave is formed in well-bedded Upper Cretaceous (Turonian) fossiliferous limestone (Fuček et al., 1991), and its speleogenesis and simple morphology was predetermined by tectonics, i.e. two fissures striking 0–180° and 64–244° that were subsequently enlarged by groundwater flow. In a later phase of speleogenesis, speleothems were precipitated, and finally, the late Pleis-
tocene–Holocene sea-level rise partially submerged the Y-Cave, along with several other similar features in the surroundings of Bribišćica Cove.

The intensive karstification of the carbonate bedrock of Dugi Otok Island resulted in the absence of surface run-off and a fast, direct infiltration of meteoric water into the underground. Infiltration is discrete and occurs through open fractures, leading to a relatively short mean residence time of groundwater in the karst underground (Terzić et al., 2007). Due to the shape of the island (45 km long and only 2 km wide in places; total surface of 114.44 km²) (Fig. 2), it is unrealistic to expect a large groundwater body. Unlike the northern part of the island (Terzić et al., 2007), the aquifer near the Y-Cave is not protected from sea intrusion, and normal groundwater-seawater contact of the unconfined coastal karst aquifer can be expected. In a coastal cave, this is marked by a thin freshwater lens overlaying the seawater wedge that intrudes into the aquifer under the freshwater (Terzić et al., 2010). Such submarine groundwater discharge (SGD) usually has significant biogeochemical importance through the provision to nutrients to specialized coastal habitats (Bokuniewicz et al., 2003; Burnett et al., 2003; Sanfilippo et al., 2014), as in the case of the Y-Cave.

Although the Adriatic is a landlocked sea, it is influenced by dynamic meteorological conditions (Cushman-Roisin et al., 2001). The south-eastern wind (sirocco) with a sufficiently long wind fetch can generate waves with a maximum recorded height of 10.8 m (Leder et al., 1998). Yet, the highest wave impact on the south-western coasts is generated when the waves produced by the SE wind sirocco interfere with those of the S wind (ostro) or SW wind (libeccio), during a change of the synoptical situation above the Adriatic Sea, primarily during winter. Given the location of the Y-Cave on the SW coast of the island facing the open sea (Fig. 2), relatively high wave energy can be expected.

Generally, a mixed semidiurnal tide type prevails in the Adriatic Sea, although the tide maintains a diurnal character in proximity of the semidiurnal amphidromic point which is positioned on the Ancona - Šibenik line (Lovato et al., 2010), which is in relatively proximity to the Y-Cave. The tidal range from the adjacent Zadar tide gauge is 29 cm (HHI, 2012). The interaction of all these geological, geomorphic, hydrogeological, meteorological and oceanological properties, along with site-specific light intensity and temperature settings, produces a spectrum of different habitats populated by characteristic marine biocenoses.

Materials and Methods

The first phase of the Y-Cave research (2003–2006) encompassed detailed cave mapping of underground conduits and cavities using standard topographical/speleological surveying techniques, and biological mapping by numerous SCUBA speleo-dives. In order to protect the cave biota, most of the species were noted and determined in situ, and others were sampled and preserved in a 96% solution of ethyl alcohol for subsequent determination in the laboratory. Sampling was performed in the least destructive manner possible, i.e. only problematic species or species unable to be determined in situ were sampled. Therefore, the relative abundance of species was estimated rather than determined quantitatively. Sponges were identified using light microscopy of skeletal characters according to the current taxonomic literature. When necessary, for higher taxonomic categories, Systema Porifera (Hooper & Van Soest, 2002) was consulted. For corals and bryozoans hydrogen peroxide (H₂O₂ conc. 6%) or sodium hypochlorite (NaOCl conc. 6%) were used for the degradation of organic matter. The scientific names are listed according to the European Register of Marine Species (Costello et al., 2014) and the World Porifera database (Van Soest et al., 2014).

For photo documentation, two underwater cameras were used: analogue Nikonos V and Canon A75 with an underwater case. The photos from the Nikonos V were digitalised and all photos were analysed in order to obtain additional data and to complement the list of species (Appendix, Table 2).

A simple species diversity index, Sorensen’s Quotient of Similarity (Q/S=2j/(a+b)∗100; where a is the total number of species in one part of the cave, b is the total number of species in the other part of the cave and j is the number of species common to both parts), was calculated for the front, middle and end part of the Y-Cave.

In order to estimate the environmental conditions that govern the distribution of marine organisms, thermohaline properties and light intensity were measured at specific sites along the cave channel. Water temperature was measured with small stow away temperature data loggers (Onset Computers®, ±0.2°C accuracy) with a set sampling interval of 16 min. The loggers were placed at selected positions and depths in the cave as shown in Figure 3 and Table 1. The measurement started on August 27, 2003 and ended on July 4, 2004 (for loggers 2, 4, 6) and on October 8, 2004 (for loggers 1, 3, 5).

Light intensity was measured during eleven days using Onset Computers® light data loggers that were set at ten positions along the cave (Fig. 3). In four light data loggers, the photosensitive cell was facing upwards (data loggers with a black dot in Fig. 3) while in the remaining six loggers, the cell was positioned to face the entrance of the cave. In the entrance part of the cave, three pairs of loggers (2 & 3, 4 & 5, 6 & 7) with different sensor cell orientations were set to determine whether the position of the photosensitive cell affects the measurement. Light intensity was recorded every two minutes during the period from June 19 to 30, 2006.

Thermohaline properties of the water column inside the Y-Cave were measured with the Idronaut® Ocean Seven 316 CTD probe on August 27, 2003. The probe
was programmed to record data every two seconds. The data was processed with the REDAS® (Real Time Data Acquisition Software). The diver (D. Petricioli) carried the probe along the cave and made six vertical profiles with it at specific locations, carefully lowering the probe from the ceiling towards the bottom. Vertical profile no. 1 was performed at the cave entrance from the water surface down to 12.5 m depth; profile no. 2 was performed inside the cave entrance from 2 to 12 m depth; profile no. 3 was performed at the turning point (junction of two main channels), approximately at section C-C’; vertical profiles no. 4 to 6 were performed in the inner part of the cave: no. 4 approximately at section E-E’, no. 5 approximately at section G-G’ and no. 6 approximately at section

| Measurement site | Location within the cave                  | Depth (m) | $\Delta m = m_{init} - m_{fin}$ (g) | $R$ (μg/a) |
|------------------|-------------------------------------------|-----------|-----------------------------------|------------|
| 1                | Entrance of the cave - bottom             | 11.5      | -0.2051                           | -20.9750   |
|                  |                                           |           | -0.2207                           | -22.5703   |
|                  |                                           |           | -0.1453                           | -14.8594   |
| 2                | Middle portion of the entrance part - ceiling | 2.9      | -0.0036                           | -0.3682    |
|                  |                                           |           | -0.0028                           | -0.2863    |
|                  |                                           |           | 0.0011                            | 0.1125     |
| 3                | Chamber with air pocket and speleothems   | 0.7       | 0.0022                            | 0.2250     |
|                  |                                           |           | 0.0008                            | 0.0818     |
|                  |                                           |           | 0.0054                            | 0.5522     |
| 4                | Central portion of the cave - bottom      | 5.1       | -0.0068                           | -0.6954    |
|                  |                                           |           | -0.0047                           | -0.4807    |
|                  |                                           |           | 0.0173                            | 1.7692     |
| 5                | End of the cave - ceiling                 | 3.9       | -0.0054                           | -0.5522    |
|                  |                                           |           | -0.0095                           | -0.9715    |
|                  |                                           |           | 0.0026                            | -0.2659    |
| 6                | End of the cave - bottom                  | 8.5       | 0.007                             | 0.7159     |
|                  |                                           |           | 0.0333                            | 3.4055     |
|                  |                                           |           | 0.2013                            | 20.5863    |

**Table 1.** Mass difference ($\Delta m$) of the limestone tablets and dissolution rate ($R$) at 6 different positions within the Y-Cave after the 1-year exposure period. Measurement positions are marked on Figure 3. Note that positive values indicate dissolution, while negative values indicate the increase of the tablet mass.

**Fig. 3:** The positions of temperature and light intensity data loggers inside the Y-Cave (dark circles - temperature data loggers; white circles - light intensity data loggers). Four loggers with their photosensitive cell facing upwards are marked with a black dot; the remaining cells were positioned to face the entrance of the cave.
H-H’ (see Fig. 2 for the position of the sections). An additional vertical profile was performed from the surface down to 29 m depth in front of the cave.

Behind the B-B’ cross-section (Fig. 2), at 5.5 m depth, an unusual pink coloured, coarse sediment was sampled. The sample was fixed in 96% ethyl alcohol and further separated in the laboratory into four categories combining two criteria: origin (terrestrial/organogenic) and size (diameter less than/greater than 4 mm). The shells of the marine gastropod Homalopoma sanguineum were also separated due to their unusually high abundance. The mass and volume fractions of each category were determined after drying (70°C for 3 days).

In order to assess the carbonate dissolutional effect within different submerged parts of the cave, sets of limestone tablets (coupled with temperature loggers) were positioned and left exposed for more than one year (from August 27, 2003 to October 8, 2004) at six distinct sites as shown in Figure 3. The tablets were made of a borehole core of Cretaceous (Senonian) limestone from a quarry near Lipica (Karst plateau, Slovenia), and measured 41.1 mm in diameter and 3–5 mm in thickness. Four holes, 3 mm in diameter, were drilled in each tablet for fixation in the field. The tablets were precisely pre-weighted and tied in series of three for each measuring point. After the 1-year exposure to the biogeochemical and hydrodynamic influence of groundwater and/or seawater, the changes in tablet mass were again precisely measured.

Dissolution rates were calculated following the expression (1) (after Plan, 2005 modified by Buzjak et al., 2013): 

\[ R = \frac{(m_{\text{fin}} - m_{\text{fin}})}{A \times \rho} \times \frac{365}{t} \]  

where \( R \) is the dissolution rate (μm/a); \( (m_{\text{fin}} - m_{\text{ini}}) \) is the mass difference (g); \( \rho \) is the limestone density (2.68 g/cm³), \( t \) is the time span (408 days) and \( A \) is the area of the initial soluble surface (32.64 cm²). The exposed surface \( A \) (cm²) was calculated using the expression (2) taking into consideration the surface of the tablet and of the four holes:

\[ A = 2r_1 \pi H + r_2 \pi H + r_1 \pi (H - r_2) + 8r_2 \pi (H - r_2) \]  

where \( r_1 \) is the radius of the tablet (cm), \( r_2 \) is the radius of the hole (cm) and \( H \) is the thickness of the tablet (cm). The dissolution rate unit μm/a (mm/ka) is the most common, and can be read as m³/km²/a (Plan, 2005) when it stands for the denudation rate.

Results

Marine organisms and communities

Within the Y-Cave, two typical marine cave biocoenoses were identified. A well-developed GSO (Fig. 4, A-E) with high species diversity and biomass dominated from the entrance to approximately section C-C’ (Fig. 2). The GO with low species diversity and biomass, is present deeper in the cave (Fig. 4, H), approximately from the section E-E’ to the end of the cave. The middle portion of the cave could be characterized as a transition from GSO to GO (Fig. 4, F–G).

In total, 139 taxa of marine organisms were recorded in the Y-Cave, among which sponges dominated with 56 taxa (Appendix, Table 1). Three classes of this phylum were recorded: Calcarea (6 taxa), Demospongiae (47 species) and Homoscleromorpha (3 species). Among the 83 taxa of other organisms, 5 algal species were recorded (only in the entrance part; Chlorophyta 2 and Rhodophyta 3), 1 benthic foraminifer, 9 species of cnidarians (all belonging to Anthozoa), 1 echiuran species, 23 mollusc taxa (15 gastropods, 7 bivalves and 1 cephalopod), 4 polychaete taxa, 6 crustacean species, 7 bryozoan species, 2 brachiopod species, 9 echinoderm species, 1 ascidian species, and 15 fish species. The majority of taxa (122) were observed in the front part of the cave, while 42 taxa were noted in the middle part of the cave (section C-C’ to approximately section E-E’), and only 20 species were noted in the innermost part (from section E-E’ towards the end of the cave). Furthermore, 93 taxa were noted only in the front part of the cave, 6 only in the middle part and 4 only in the end part. Also, 20 taxa were found in both the front and the middle part of the cave and 7 in the middle and the end part of the cave, while 9 sponge species were present throughout the cave with decreasing abundance and body size towards the end (Appendix, Table 1). Of these, Chondrosia reniformis and Petrosia (Petrosia) ficiformis were completely white inside the cave due to the lack of light and therefore the absence of endosymbiotic algae that give them their characteristic colouration (Fig. 4 E). In the middle and end parts of the cave, virtually no sessile marine organisms were noted on walls shallower than 2 m depth (Fig. 11).

The end part of the cave was characterized by low species richness and low relative abundance and by distinct faunal depletion (Fig. 4 H; Appendix, Table 2) that is typical for GO. Barren rock was easily seen and the fauna consisted of scarce populations of small and/or encrusting forms of demosponges and homoscleromorphs, small calcareous sponges, and small serpulids.

Three sponge species with cave affinities characterize the inner dark part of the Y-Cave: Discodermia polymorpha (rare), Myrmekiderma speleaeum (common) and Placospongia decoritans (common; Fig. 4 G). Other common sponge species inside the dark part of the cave were: calcareous Amphoriscus chrysalis, Sycon elegans and Sycon sp., demosponges Chondrosia reniformis, Ciona schmidtii, Diplastrella bistellata, Petrosia (Petrosia) ficiformis, Spirastrella cunctatrix, Spongia (Spongia) virgulosa, and homoscleromorphs Corticium candlabrum and Plakinabowerbanki. None of these species showed any particular dominance in respect to the others.

In terms of species diversity, the middle and end parts of the cave were the most similar (50.0% of similarity), the front and middle parts were less similar (35.4%) and the front and
**Fig. 4:** Living communities inside the Y-Cave: A) the entrance part of the cave, vertical wall, depth 9 m, biocenosis of semi-dark caves (GSO, see text for explanation of acronym) dominated by numerous sponge species; B) the entrance part of the cave, ceiling, depth 6 m, GSO dominated by scleractinian coral *Leptopsammia pruvoti*; C) the entrance part of the cave, overhang, depth 7 m, GSO dominated by scleractinian coral *Madracis pharensis*; D) the entrance part of the cave, vertical wall (near the bottom), depth 9 m, GSO, a large specimen of the orange sponge *Agelas oroides* dominates the photo; E) the middle part of the cave, in front of the section C-C’, bottom, depth 10 m, a massive white specimen of the sponge *Chondrosia reniformis*; F) the middle part of the cave, between sections C-C’ and D-D’, vertical wall and overhang, depth 5 m, the transition from GSO to biocenosis of caves and ducts in total darkness (GO, see text for explanation of acronym), the community is dominated by serpulids; G) the middle part of the cave, between sections C-C’ and D-D’, vertical wall, depth 5 m, transition from GSO to GO, a dense population of brachiopod *Novocrania anomala*, encrusting sponge *Placospongia decorticans* and serpulids; H) the end part of the cave, near the section G-G’, vertical wall with overhang and horizontal shelf, depth 6 m, GO with scarce calcareous sponges and serpulids (see Fig. 2 for position of the sections).
end parts of the cave were the least similar (12.7%).

Within the Y-Cave, 15 fish species were noted, virtually all in the front part of the cave (14 of 15; Appendix, Table 2). The only fish species recorded at the end part of the cave was the speleophilic species Grammonus ater. Only two species could be considered common inside the cave: Apogon imberbis and the speleophilic Thorogobius ephippatus and they were noted in the front and middle parts of the cave.

**Temperature, salinity and light intensity**

The annual variations of water temperature in the entrance part of the cave and at five positions inside the cave are shown in Figure 5. Temperature readings revealed a complex seasonal distribution of water layers in the cave from August 2003 to July/October 2004: the retention of warm water (>15°C) in the shallower parts of the cave during winter (loggers 2, 3 and 5), the existence and retention of a cold water (18–22°C) layer inside the chamber with speleothems and air pocket during summer 2004 (logger 3), the retention of colder water in deeper parts inside the cave throughout the year (logger 6), and rapid periodical changes of water temperature in the middle part of the cave during winter (logger 4).

Since there was a prominent winter temperature fluctuation recorded by logger 4 (Fig. 5) at a depth of 5.1 m in the middle part of the cave, this was compared to the tide records of the nearest tide gauge at the port of Zadar for the period from February 2–8, 2004. Figure 6 clearly shows inverse fluctuations, i.e. a temperature increase of approximately 3°C with each low tide, and a drop with each high tide.

Vertical temperature profiles outside, at the entrance, and inside the Y-Cave, taken with the CTD probe in summer 2003, revealed that the thermocline outside the cave and in the entrance part was at around 10 m depth, which is approximately 5 m deeper than in the inner part of the cave (Fig. 7). At the end part of the cave (at sections G-G' and H-H'; Fig. 2) the thermocline was at around 5 m and a sudden temperature drop of 7°C occurred within one meter depth.

Vertical temperature and salinity profiles recorded with the CTD probe during the dive inside the Y-Cave on August 27, 2003 are presented in Figure 8. At the time of recording, the salinity throughout the cave showed a fully marine environment. Although salinity oscillations were minimal, approximately 1 PSU, it was notable that in all vertical profiles, except for profile no. 3, salinity rose slightly with each dive.

![Temperature data logger T1](image1)

![Temperature data logger T2](image2)

![Temperature data logger T3](image3)

![Temperature data logger T4](image4)

![Temperature data logger T5](image5)

![Temperature data logger T6](image6)

**Fig. 5:** The annual variation of temperature along the Y-Cave (from August 23–27, 2003 to July 4/October 8, 2004). Measurement positions are given in Fig. 3 and depths in Table 1.
The light intensity measurement taken over a period of 11 days in June 2006 showed that the orientation of the photosensitive cell did not yield any prominent differences in the measurement, and that virtually no light penetrated into the cave after logger 4 (see position in Fig. 3). Figure 9 shows the results of the light intensity measurements at only the most representative positions, i.e. logger 1 at the entrance, logger 4 in the middle part and logger 10 at the end part of the cave.

Cave sediment

The mass and volume fractions of four sediment categories in the coarse sediment sample collected behind section B-B', at 5.5 m depth inside the Y-Cave, are shown in Figure 10. The mass fraction of detrital terrigenous sediment (gravel >4 mm) was the highest (48.8%). The volume fractions of the four categories were very similar to the mass fractions. The shells of the gastropod *Homalopoma sanguineum* comprised a significant portion of the sediment (mass fraction 10.3%, volume fraction 13.0%).

Dissolutional effect

The potential presence of the aggressive SGD was primarily recognized through the corrosion marks on speleothems and bedrock (Fig. 11). In addition, there was a significant absence/scarcity of sessile marine organisms near the sea surface in the chamber with speleothems and air pocket and on the cave ceiling throughout the entire inner part of the cave (Fig. 11, B-D). The results of the subsequent 1-year dissolution experiment on the limestone tablets are shown in Table 1 and Figure 12. Since the methodology refers to the amount of carbonate dissolution, positive values suggest an enhanced mass loss while negative values indicate an increase in tablet mass. The most pronounced
differences were the mass gain at site 1 at the entrance of the cave (Fig. 12, A) attributed to bioaccumulation, and the mass loss at site 6 at the end of the cave (Fig. 12, C) where all three tablets were visibly abraded. The absolute dissolution values are not comparable to any previous studies, as this was the first deployment of “standard” limestone tablets in a submarine environment.

**Effect of hydrodynamics**

The impact of the hydrodynamics within the cave is seen in the sediment particle size. The sediment on the bottom is coarser behind the section B-B′ (Fig. 13) in the middle and at the very end of the cave (section H-H′) than the sediment at the entrance (section A-A′) and in the side channel in the middle of the cave (section C-C′).

**Discussion**

In order to protect the cave biota in the Y-Cave, the relative abundance of the species was estimated rather than determined quantitatively and sampling was performed only as necessary. Although justified from the conservation point of view, this methodology has its limitations as it cannot assure that the sampling effort was uniform for all cave areas or all groups of organisms. However, even with the limited precision obtained with this methodology, a comparison of the collected data could be made and conclusions could be drawn.

The poriferan species diversity in the Y-Cave (56 taxa identified out of 139 in total; Appendix, Table 2) ranks among the ten highest when compared with other studied Mediterranean caves (Gerovasileiou & Voultsiadou, 2012, see Table S4). Mediterranean marine caves have been recognized as an important sponge biodiversity reservoir, with 45.7% of all recorded marine poriferan species also found in caves (311 of 681 species; Gerovasileiou & Voultsiadou, 2012). In a previous study, a total of 77 sponge species were found in 7 marine caves along the eastern Adriatic coast (Bakran-Petricioli et al., 2012).

The distribution of sponges throughout the Y-Cave is positively correlated with the light gradient: the highest species diversity (45 of 56 taxa) was recorded in the front part of the cave where there is some light penetration (Fig. 9). That part of the cave, with over 100% biological cover (Fig. 4 A-D) and 122 taxa recorded, can be characterized as a typical GSO. In the middle and end parts of the cave (Fig. 4 E-H), which have virtually no light (Fig. 9), sponge diversity is much lower (21 taxa in the middle and 18 taxa in the end part) as is the relative abundance of sponge species. A decreasing number and abundance of sponge species along the horizontal cave axis has also been noted in many other superficial Mediterranean marine caves with blind ends (with a single entrance) (Cinelli et al., 1977; Balduzzi et al., 1989; Corriero et al., 2000; Arko-Pijevac et al., 2001; Gerovasileiou & Voultsiadou, 2012). It is widely
accepted that such a distribution is strongly correlated not only with light attenuation but also with reduced water circulation. Harmelin (1969) stated that in dark tunnels (ducts open on both sides) with constant water renewal, a typical GSO spreads into total darkness.

The middle and the end parts of the cave, which are characterized by an absence of light (Fig. 9), were the most similar in terms of their species diversity (taking all spe-
cies into account Q/S = 50.0%; taking only sponge species into account Q/S = 76.9%). Both parts could generally be assigned as GO due to their species composition, lower species diversity and abundance (Fig. 4 E-H). However, due to their biological peculiarities (discussed below), we decided to separate them into two zones: the middle part of the cave between sections C-C’ and E-E’ (Fig. 2), which we considered to be a transition zone from GSO to GO, and the end part of the cave from approximately section E-E’ to the end of the cave, which we considered to be GO.

In the middle part of the cave, the diversity and relative abundance of species was higher than in the end part of the cave (42 vs. 20 taxa). A typical nodular type of bryozoan facies, considered as a transition from GSO to GO (Harmelin et al., 1985), was noted only in small patches on vertical walls, between the sections C-C’ and D-D’. The community on most vertical surfaces and overhangs in that area of the cave was dominated by serpulids (Fig. 4 F).

In the middle part of the cave, an abundant population of the small pink gastropod *Homalopoma sanguineum* (family Colloniidae) was noted (Appendix, Table 2) on the rocky walls and on individuals of the sponge *Spongia (Spongia) virgultosa*. The analysis of the coarse sediment collected behind the B-B’ cross-section (at the beginning of the middle part of the cave; Fig. 2), at 5.5 m depth, showed that it contained a significant proportion of *Homalopoma* shells (Fig. 10 C). Arko-Pijevac et al. (2001) found that *H. sanguineum* dominated rock fissures below 2 m depth in a narrow vertical channel (“chimney”) open to air in a marine cave in the northern Adriatic. Dantart & Luque (1994) stated that this species is common in marine caves (often more abundant in the interior) and also in coralligenous habitats of the Mediterranean. Although usually considered an algal herbivore, these results suggest that *H. sanguineum* can maintain abundant populations inside marine caves tens of meters from algal beds, likely feeding on sponges and organic detritus. This finding is in agreement with the results of an analysis of the gastropod digestive tract content by Dantart & Luque (1994), which contained diatom tests, sponge spicules, fragments of hydroids and some unidentifiable detritus.

The discovery of a gregarious settlement of brachiopod *Novocrania anomala* on the walls in the middle part of the cave, between sections C-C’ and D-D’ (Fig. 4 G) is another interesting result. According to Jackson (2000), this species is gonochoristic and has lecithotrophic larvae. The fertilization is external and the eggs settle on the bottom. Free swimming larvae are fully developed within three days and settle on the substrate, cementing their ventral valves onto it within days. The species is an active filter feeder and is thought to prefer water currents at rates of less than one knot. This could explain the gregarious settlement of *N. anomala* in the Y-Cave. Gregarious settlements of only one or two species in cryptic habitats (lava caves) have been noted for some other species of brachiopods (Logan et al., 2007). The brachiopod *N. anomala*, albeit not very common, is distributed along the eastern coast of the Adriatic Sea (Logan, 2003; Legac, 2012). However, such a gregarious settlement like the one found in this cave has not previously been reported for this species. Although Logan (2003) and Legac (2012) consider the specimens found in the northern Adriatic to be a separate species (*Novocrania turbinata*), a recent study by Emig (2014) showed that *N. turbinata* is actually a synonym of *N. anomala*, the latter being the valid name for this species (Costello et al., 2014).

In their research on fish assemblages in three shallow marine caves with a single entrance (similar morphology to the Y-Cave), Bussotti et al. (2002) recorded a total of 19 fish species (per cave: 12, 12 and 10 species). They noted a decrease of fish species richness and abundance along the horizontal axes of the caves (except for *A. imberbis*). Due to the methodology used in our research, fish assemblages within the Y-Cave can not be discussed in detail, the species recorded (10 common species), their number and relative abundance along the cave are in concordance with the results of Bussotti et al. (2002).

Virtually no sessile marine organisms were noted on the walls within the Y-Cave shallower than 2 m depth (Fig. 11). This absence could be attributed to the freshwater influence in the upper layers of the water column inside the cave. Comparable observations have been made in submarine caves from SE Sicily (Guido et al., 2013; Sanfilippo et al., 2014). Although freshwater discharge was noted from the fissure at the entrance of the cave at 6 m depth (February 2006) there are no notable signs of freshwater impact on the benthic populations of GSO that inhabit the ceiling and overhangs in the entrance part (Fig. 4 B-C, depths 6 and 7 m respectively). Further in the cave, at shallower depths, the absence/scarcity of benthic organisms and pronounced corrosion marks suggest a freshwater influence. The existence and retention of colder surface water layers inside the chamber with speleothems and air pocket during most of summer 2004 (Fig. 5, logger 3) could only be explained by the less sa-
line character of that layer. Divers also noted a brackish layer approximately 10 cm thick on the sea surface inside the chamber in June 2006. Freshwater impact (flow, retention, mixing with sea water) in the upper layers of the water column inside shallow karstic caves could be a very important ecological factor that shapes the living communities within the caves, but which is often disregarded (Harmelin, 1986; Moscatello & Belmonte, 2007).

The dissolutive effect of SGD measured through differences in tablet masses after one year of exposure within the cave (Fig. 12, Table 1) led to the identification of three different, independent processes: i) bioaccumulation; ii) dissolution and iii) mechanical erosion. These processes are controlled by the position in the cave in relation to the distance from the entrance and the absolute depth that consequently generates different environmental conditions. An outline of each is provided below:

i) Bioaccumulation, owing to marine organisms that build carbonate skeletons, was pronounced in the entrance part of the cave and likely though to a smaller extent, along the inner, but not shallow part of the cave. After the 1-year exposure period, the limestone tablets were populated by serpulids, enrusting byozoans and fouling bivalves. The abundance of bioconstructing organisms such as stony corals, branched and enrusting byozoans, and serpulids, is much higher in the front part of the cave. Bioconstruction is common in the karst environment and plays an important role in shaping limestone coasts (De Waele & Furiani, 2013), cave interiors (Surić, 2005), and is an essential component of biokarst. Biokarst refers to depositional and erosional features produced by direct biological action (Viles, 1988; Bonacci et al., 2009) and it is not unusual to have overlapping depositional and erosional components. Bioerosional processes also take part in the Y-Cave, as the endolithic bivalves Lithophaga lithophaga and Roccellaria dubia, and boring sponges from the genus Cliona were observed in the front and the middle parts of the cave.

ii) The dissolutive effect, i.e. mixing corrosion, was noticed in the shallowest part of the cave. According to the recorded geochemical activity of freshwater and the position of the cave, the development of the Y-Cave would fit into the Carbonate Island Karst Model (CIKM), which integrates various components that control cave and karst development on carbonate islands (Mylroie & Mylroie, 2007). The most pronounced features within the CIKM are the flank margin caves that develop by mixing at the boundary between saltwater and the overlying freshwater lens, and also at the contact of the vadose and phreatic zones at the top of the freshwater lens. The two zones where mixing occurs are superimposed at the distal margin of the freshwater lens, where the lens thins and creates an additive geochemical effect (Mylroie & Mylroie, 2007; Otoničar et al., 2010). The Y-Cave is located within this zone. However, one of the principles of the CIKM is that the karst is eogenetic (i.e. diagenetically immature, young carbonate rocks that have never been buried below the zone of meteoric diagenesis) or highly tectonized telogenetic karst (diagenetically mature, sometimes recrystallized rock with a lack of significant primary porosity) (Mylroie & Mylroie, 2007; Mylroie et al., 2008). The Croatian coastal karst is generally telogenetic and flank margin caves are not expected to be found. The Plava Grotta Cave (Cres Island) is an exception: it was formed in talus breccias facies, which provides three-dimensional porosity and permeability structure that acts hydraulically similar to the high primary porosity and permeability of young eogenetic carbonate rocks (Otoničar et al., 2010). Still, the bedrock of the Y-Cave is neither highly tectonized nor breccia, so dissolution is not pronounced enough to form a maze-like cave or larger chambers. However, it is strong enough to affect the bedrock surface, and thus might be an important factor in the biological species distribution.

iii) Mechanical erosion due to the impact of hydrodynamism is pronounced at the very end of the cave, almost 90 meters from the entrance. The relatively wide submerged cave entrance (7 m) at 12 m depth faces the open sea (Fig. 2). The aforementioned high wave energy produces a hydraulic impact inside the cave but, due to the cave morphology, the impact is not of the same intensity in all parts of the cave. This is registered in the different sediment particles size (coarser sediment inside the cave and at the end of it than at the entrance or in the side channel). The impact of abrasive forces was additionally seen in the abraded limestone tablets at the end of the cave (Fig. 12 C, Table 1) and even more so by the rounded gravel and polished surrounding bedrock 22 m from the entrance, as shown in Figures 10 and 13. A similar abrasion effect observed on shells was mentioned by Driscoll (1976). These visible consequences of a pronounced hydraulic impact coincide with the narrowing of the cave channels at these sites. This phenomenon is known as the “water ram” but it has not previously been described within karstic caves. Interestingly, the abraded carbonate tablets (coupled with temperature logger 6) were within the bottom cold water layer which persists in the cave during most of the year (Fig. 5, logger 6), meaning that the wave energy is transferred in such a way that it does not mix the water column, thus maintaining the water layers. That can also explain the scarcity of organisms in the marine layer at the end of the cave despite the pronounced hydrodynamism – although wave energy is transferred, there is actually no food transport that reaches the end of the cave.

The stratification of the water column within the cave throughout most of the year is shown in Figure 5. First, the retention of a colder water layer, especially throughout the summer, happens along the bottom inside the cave (Fig. 5, logger 6). There was a pronounced thermocline, with a sudden temperature drop of 7°C in the summer at approximately 5 m depth (Fig. 7), which could clearly be attributed to the cave morphology (Fig. 2). The vertical profiles showed that the salinity of the cold water layer
was similar to that of the open sea (Fig. 8, profiles 4, 5 and 6). This phenomenon could have a significant impact on marine biota as deep sea sponges have been found elsewhere in cold water lenses on the bottom of shallow caves (Bakran-Petricioli et al., 2007). Second, the retention of cold, apparently brackish waters on the sea surface in the speleothem chamber during summer (Fig. 5, logger 3) is in accordance with the absence of marine organisms shallower than 2 m depth inside the cave (Fig. 11) and suggests a freshwater impact. Third, the retention of warmer water in the upper parts of the cave in the winter was noted (Fig. 5, loggers 2, 3 and 5), which could also suggest the possible mixing of freshwater and seawater.

The freshwater outflow from the Y-Cave certainly results in compensating seawater inflow though its extent cannot be estimated. This phenomenon was reported earlier in the stratified karstic estuary of the Krka River (Legović et al., 1991), and in the freshwater outlet of a hydropower plant in Croatia (Legović et al., 2003). The compensating inflow of seawater could bring more food to the marine organisms within the cave.

The year round measurement of water temperature revealed a peculiar phenomenon in the middle of the cave: rapid periodical changes of water temperature at 5 m depth during the winter (Fig. 5, logger 4). These oscillations are undoubtedly connected to the tides, as the temperature rises with each low tide and the temperature drops with each high tide, by approximately 3°C (Fig. 6). It can be presumed that with each high tide, a wedge of outside seawater, which is less dense than the cold water in the cave but denser than the upper layer (and likely less saline), penetrates into the cave, at least to its middle part. This daily tidal dynamic could bring more food to marine organisms in the middle part of the cave (evidenced by the dense population of the gastropod *Homalopoma sanguineum* and the gregarious settlement of the brachiopod *Novocrania anomala*), though it is not strong enough to disturb the inner stratification of the cave.

The temperature oscillations recorded during the summer months at the entrance of the cave at 11.5 m depth (Fig. 5, logger 1) are due to a phenomenon that was first recorded in the southern Adriatic. Novosel et al. (2004) found that marine life within the thermocline layer is exposed to large high-frequency oscillations connected partly to an internal diurnal tide. Similar oscillations could be noticed inside the cave at loggers 2, 4 and 5 (depths 2.9 m, 5.1 m and 3.9 m respectively) but not at loggers 3 (depth 0.7 m) and 6 (depth 8.5 m). Those summer oscillations did not disturb the stratification of the water column inside the cave.

**Conclusions**

The distribution of living organisms along the Y-Cave, together with the results of the measured abiotic parameters, clearly reflects the complex environmental conditions that shape this shallow, partly submerged coastal cave at a small (meter) scale. The phylum Porifera was the dominant group of organisms and the poriferan species diversity in the Y-Cave ranked among the ten highest of all other investigated Mediterranean caves. The general distribution of organisms along the Y-Cave is positively correlated to the light gradient and reduced water circulation, thus the highest species diversity and abundance were recorded in the front part of the cave. The strong hydrodynamism evidenced at the end of cave is produced by wave energy transfer inside the cave and does not necessarily mean that there is transport of food (water circulation) all the way to the end of the cave.

Although the middle and end parts of the cave, both characterized by an absence of light, were the most similar, the biological peculiarities of the former, such as an abundant population of the gastropod *Homalopoma sanguineum* and a gregarious settlement of the brachiopod *Novocrania anomala*, are interpreted to be caused by differences in environmental factors, e.g. tidally generated hydrodynamics.

The absence/scarcity of sessile marine organisms and pronounced corrosion marks at shallow depths inside the Y-Cave suggest a freshwater impact (flow, retention, mixing with sea water) in the upper layers of the water column. That impact, often disregarded, plays an important role in the distribution of organisms in the cave, either due to unfavourable salinity or a corrosive impact.

The one-year experiment with carbonate tablets, aimed at detecting the dissolutional effect of seawater and freshwater mixing, revealed three different, independent ongoing processes controlled by the position in the cave in terms of distance from the entrance, absolute depth and cave morphology: i) bioaccumulation due to the construction of carbonate skeletons that was most pronounced in the entrance part; ii) dissolution due to mixing corrosion observed in the shallowest part of the cave; and iii) mechanical erosion due to the impact of hydrodynamism pronounced in different parts of the cave and associated with the cave morphology.

The results also revealed water column stratification within the cave: i) retention of a colder water layer, especially throughout the summer, along the bottom of the cave interior, which is associated with the cave morphology, ii) retention of cold, apparently brackish water, on the sea surface in the speleothem chamber during summer, iii) retention of warmer water in upper parts of the cave in winter which could be attributed to the possible mixing of freshwater and seawater. Although the cave is shallow and exposed to the open sea, neither tidal nor wave energy, both observed at particular sites within the cave, disturb the stratification.

In addition to the hydrologically induced geochemical changes, even small differences in the morphology of a simple, shallow marine cave could have ecological implications that could easily be overlooked without simultaneous long-term measurements of key abiotic parameters, such as temperature, salinity, light, and hydro-
dynamism in distinct spots inside the cave. The present study shows that the resulting distribution of organisms within the Y-Cave is unique to the cave and confirms the necessity of considering this fact where the conservation and management of marine caves are concerned.

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APPENDIX

Table 2. List of the taxa collected and/or identified from photos in the Y-Cave, Dugi Otok Island, Croatia with their relative abundance (front part encompasses part of the cave from the entrance to approximately section C-C’, middle part encompasses from the section C-C’ to the section E-E’ and the end part encompasses from the section E-E’ to the end of the cave, see Figure 2 for the position of sections; relative abundance: cc - abundant species, c - common species, + - rare species).

| Taxa                                    | front part of the cave | middle part of the cave | end part of the cave |
|-----------------------------------------|-------------------------|--------------------------|-----------------------|
| **Chlorophyta**                         |                         |                          |                       |
| *Palmophyllum crassum* (Naccari) Rabenhorst, 1868 | c                       |                          |                       |
| *Valonia utricularis* (Roth) C. Agardh, 1823 | +                       |                          |                       |
| **Rhodophyta**                          |                         |                          |                       |
| *Lithophyllum stictaeforme* (Areschoug) Hauck, 1877 | +                       |                          |                       |
| *Peyssonella rubra* (Greville) J. Agardh, 1851 | c                       |                          |                       |
| *Peyssonella squamaria* (S.G.Gmelin) Decaisne, 1842 | c                       |                          |                       |
| **Foraminifera**                        |                         |                          |                       |
| *Miniacina miniacea* (Pallas, 1766)      | c                       | c                        |                       |
| **Porifera**                            |                         |                          |                       |
| *Amphoriscus chrysalis* (Schmidt, 1864)  | c                       |                          |                       |
| *Clathrina clathrus* (Schmidt, 1864)     | c                       | c                        |                       |
| *Clathrina coriacea* (Montagu, 1814)    | c                       |                          |                       |
| *Sycon elegans* (Bowerbank, 1845)       | c                       | +                        |                       |
| *Sycon raphanus* Schmidt, 1862          | +                       | +                        |                       |
| *Sycon sp.*                              | +                       | c                        |                       |
| *Acanthella acuta* Schmidt, 1862         | c                       |                          |                       |
| *Agelas oroides* (Schmidt, 1864)        | cc                      |                          |                       |
| *Axinella cannabina* (Esper, 1794)       | +                       |                          |                       |
| *Axinella damicornis* (Esper, 1794)      | +                       |                          |                       |
| *Axinella verrucosa* (Esper, 1794)       | c                       |                          |                       |
| *Caminella intuta* (Topsent, 1892)      | +                       |                          |                       |
| *Chondrosia reniformis* Nardo, 1847      | c                       | c                        | +                     |
| *Cliona celata* Grant, 1826              | +                       |                          |                       |
| *Cliona rhodensis* Rützler & Bromley, 1981 | +                      |                          |                       |
| *Cliona schmidtii* Ridley, 1881         | +                       |                          |                       |
| *Crambe crambe* (Schmidt, 1862)         | c                       |                          |                       |
| *Dictyonella incisa* (Schmidt, 1880)     | c                       |                          |                       |
| *Diplastrella bistellata* (Schmidt, 1862) | c                       | c                        | c                     |
| *Discodermia polymorpha* Pisera & Vacelet, 2011 | +                      |                          |                       |
| *Dysidea avara* (Schmidt, 1862)         | +                       |                          |                       |
| *Dysidea fragilis* (Montagu, 1814)      | c                       |                          |                       |
| *Eurypon clavatum* (Bowerbank, 1866)    | +                       |                          |                       |
| *Geodia conchilega* Schmidt, 1862        | +                       |                          |                       |
| *Haliclona* (Halichocoma) fulva* (Topsent, 1893) | cc                   |                          |                       |
| *Haliclona* (Soestella) mucosa* (Griessinger, 1971) | cc                  |                          |                       |
| *Hemimycera columnella* (Bowerbank, 1874) | cc                     | +                        |                       |
| *Hymedesmia* (Stylopus) coriacea* (Fristedt, 1885) | +                  |                          |                       |
| *Ircinia dendroides* (Schmidt, 1862)     | cc                      |                          |                       |
| *Ircinia oros* (Schmidt, 1864)          | c                       |                          |                       |
| *Ircinia variabilis* (Schmidt, 1862)     | cc                      |                          |                       |
| *Jaspis johnstoni* (Schmidt, 1862)      | +                       | +                        | +                     |
| *Merlia normani* Kirkpatrick, 1908      | +                       |                          |                       |
| *Myrmekioderma speculum* (Pulitzer-Finali, 1983) | +                |                          |                       |
| *Penares euastrum* (Schmidt, 1868)      | c                       |                          |                       |
| *Petrosia* (Petro sia) ficiformis* (Poi ret, 1789) | cc                  | c                        | +                     |
| *Phorbas tenacior* (Topsent, 1925)      | cc                      |                          |                       |
| *Placospofia decoricans* (Hanitsch, 1895) | c                      | +                        |                       |
| *Pleraplysilla spinifera* (Schulze, 1879) | +                      |                          |                       |
| *Raspaciona aculeata* (Johnston, 1842)  | +                       |                          |                       |
| *Reniera* spp.                          | +                       |                          |                       |

(continued)
| Taxa                                           | front part of the cave | middle part of the cave | end part of the cave |
|------------------------------------------------|------------------------|-------------------------|----------------------|
| Rhabderemia spinosa Topsent, 1896              | +                      | +                       | +                    |
| Sarcothragus foetidus Schmidt, 1862            | c                      |                         |                      |
| Sarcothragus spinosulus Schmidt, 1862          | c                      |                         |                      |
| Scalarispongia scalaris (Schmidt, 1862)        | cc                     |                         |                      |
| Spirastrella cunctatrix Schmidt, 1868          | cc                     | c                       | +                    |
| Spongia (Spongia) officinalis Linnaeus, 1759   | +                      |                         |                      |
| Spongia (Spongia) virgulosa (Schmidt, 1868)    | +                      | c                       | +                    |
| Spongosorites flavens Pulitzer-Finali, 1983    | +                      |                         |                      |
| Terpios gelatinosa (Bowerbank, 1866)           | +                      |                         |                      |
| Tehya aurantium (Pallas, 1766)                | +                      | +                       |                      |
| Tehya sp.                                      |                        |                         |                      |
| Timea sp.                                      | +                      |                         |                      |
| Corticium candelabrum Schmidt, 1862            | c                      | c                       | +                    |
| Oscarella lobularis (Schmidt, 1862)            | c                      |                         |                      |
| Plakina bowerbanki (Sara, 1960)               | +                      | c                       | +                    |

**Cnidaria - Anthozoa**

| Taxa                                           | front part of the cave | middle part of the cave | end part of the cave |
|------------------------------------------------|------------------------|-------------------------|----------------------|
| Balanophyllia (Balanophyllia) europaea (Risso, 1826) | +                      |                         |                      |
| Caryophyllia (Caryophyllia) inornata (Duncan, 1878) | +                      |                         |                      |
| Caryophyllia (Caryophyllia) smithii Stokes & Broderip, 1828 | +                      |                         |                      |
| Cerianthus membranaceus (Spallanzani, 1784)        | +                      |                         |                      |
| Isarachanthus nocturnus (Hartog,1977)             | +                      |                         |                      |
| Leptopsammia pruvoni Lacaze-Duthiers, 1897        | cc                     | c                       | +                    |
| Maddracis pharensis (Heller, 1868)               | cc                     |                         |                      |
| Parazoanthus axinellae (Schmidt, 1862)            | c                      |                         |                      |
| Polycyathus muellerae (Abel, 1959)               | c                      |                         |                      |

**Echiura**

| Taxa                                           | front part of the cave | middle part of the cave | end part of the cave |
|------------------------------------------------|------------------------|-------------------------|----------------------|
| Bonellia viridis Rolando, 1821                 | +                      | +                       |                      |

**Mollusca - Gastropoda**

| Taxa                                           | front part of the cave | middle part of the cave | end part of the cave |
|------------------------------------------------|------------------------|-------------------------|----------------------|
| Armina tigrina Rafinesque, 1814                 | +                      |                         |                      |
| Berthella aurantiaca (Risso, 1818)              | +                      |                         |                      |
| Bolma rugosa (Linnaeus, 1767)                   | +                      |                         |                      |
| Callioctoma zizyphinum (Linnaeus, 1758)         |                        |                         | +                    |
| Chromodoris sp.                                 |                        |                         |                      |
| Clanculus corallinus (Gmelin,1791)              | +                      |                         |                      |
| Conus ventricosus Gmelin, 1791                  | +                      |                         |                      |
| Emarginula rosea Bell, 1824                     | +                      |                         |                      |
| Flabellina affinis (Gmelin, 1791)               | +                      |                         |                      |
| Haliotis tuberculata Linnaeus, 1758             | +                      |                         |                      |
| Hexaplex trunculus (Linnaeus, 1758)             | +                      |                         |                      |
| Homalopoma sanguineum (Linnaeus, 1758)          |                         | c                       | cc                   |
| Luria lurida (Linnaeus, 1758)                   | +                      |                         |                      |
| Muricopsis cristata (Brocchi,1814)              | +                      |                         |                      |
| Peltodoris atromaculata Bergh, 1880             | +                      |                         |                      |

**Mollusca - Bivalvia**

| Taxa                                           | front part of the cave | middle part of the cave | end part of the cave |
|------------------------------------------------|------------------------|-------------------------|----------------------|
| Arca noae Linnaeus, 1758                        | +                      |                         |                      |
| Barbatia barbata (Linnaeus, 1758)               | +                      |                         |                      |
| Chama gryphoides Linnaeus, 1758                  | +                      |                         |                      |
| Lithophaga lithophaga (Linnaeus, 1758)          | c                      |                         | +                    |
| Musculus costatus (Risso,1826)                  | +                      |                         |                      |
| Roccellaria dubia (Pennant, 1777)               | c                      |                         | +                    |
| Spondylus gaederopus Linnaeus, 1758             | +                      |                         |                      |

**Mollusca - Cephalopoda**

| Taxa                                           | front part of the cave | middle part of the cave | end part of the cave |
|------------------------------------------------|------------------------|-------------------------|----------------------|
| Octopus vulgaris Cuvier, 1797                   | +                      |                         |                      |

**Annelida - Polychaeta**

| Taxa                                           | front part of the cave | middle part of the cave | end part of the cave |
|------------------------------------------------|------------------------|-------------------------|----------------------|
| Eupolymnia nebulosa (Montagu, 1818)            | +                      |                         |                      |
| Protula sp.                                    | c                      |                         | +                    |
| Taxa                                      | front part of the cave | middle part of the cave | end part of the cave |
|-------------------------------------------|------------------------|-------------------------|----------------------|
| Serpula sp.                               | c                      |                         | +                    |
| Serpulidae indet.                         |                        | c                       |                      |
| Galathea strigosa (Linnaeus, 1761)        |                        |                         | +                    |
| Hemimysis margalefi Alcaraz, Riera & Gili, 1986 |                        |                         | + c                  |
| Homarus gammarus (Linnaeus, 1758)         |                        |                         | + c                  |
| Palaemon serratus (Pennant, 1777)         |                        | +                       | c                    |
| Palinurus elephas (Fabricius, 1787)       |                        | +                       | + c                  |
| Stenopus spinosus Risso, 1827             |                        |                         | +                    |
| Arthropoda - Crustacea                    |                        |                         |                      |
| Adeonella polystomella (Reuss, 1848)      |                        |                         | + c                  |
| Crisia signoidea Waters, 1916             |                        | +                       | c                    |
| Frondipora verrucosa (Lamouroux, 1821)    |                        |                         | +                    |
| Margaretta cereoides (Ellis & Solander, 1786) |                        |                         | +                    |
| Myriapora truncata (Pallas, 1766)         |                        |                         | +                    |
| Schizotheca serratimargo (Hincks, 1886)   |                        |                         | + c                  |
| Scrupocellaria scrupea Busk, 1852         |                        |                         | + c                  |
| Bryozoa                                   |                        |                         |                      |
| Joania cordata (Risso, 1826)              |                        |                         | +                    |
| Novocrania anomala (Müller, 1776)         |                        |                         | cc                   |
| Brachiopoda                               |                        |                         |                      |
| Antedon mediterranea (Lamarck, 1816)      |                        |                         | +                    |
| Arbacia lixula (Linnaeus, 1758)           |                        |                         | +                    |
| Echinaster (Echinaster) sepositus (Retzius, 1783) |                        |                         | +                    |
| Hacelia attenuata Gray, 1840              |                        |                         | +                    |
| Holothuria (Panningothuria) forskali Delle Chiaje, 1823 |                        |                         | +                    |
| Marthasterias glacialis (Linnaeus, 1758)  |                        |                         | +                    |
| Ophidiaster ophidianus (Lamarck, 1816)    |                        |                         | +                    |
| Ophiothrix fragilis (Abildgaard, in O.F. Müller, 1789) |                        |                         | +                    |
| Paracentrotus lividus (Lamarck, 1816)     |                        |                         | +                    |
| Chordata - Ascidiae                       |                        |                         |                      |
| Halocynthia papillosa (Linnaeus, 1767)    |                        | +                       | +                    |
| Chordata - Actinopterygii                 |                        |                         |                      |
| Apogon imberbis (Linnaeus, 1758)          |                        | c                       | c                    |
| Chromis chromis (Linnaeus, 1758)          |                        | +                       | +                    |
| Conger conger (Linnaeus, 1758)            |                        | +                       | + c                  |
| Diplodus vulgaris (Geoffroy Saint-Hilaire, 1817) |                        |                         | +                    |
| Gobius cruentatus Gmelin, 1789            |                        |                         | +                    |
| Grammonus ater (Risso, 1810)              |                        |                         | +                    |
| Mullus barbatus barbatus Linnaeus, 1758   |                        |                         | +                    |
| Oblada melanura (Linnaeus, 1758)          |                        | +                       | + c                  |
| Parablennius rouxi (Cocco, 1833)          |                        |                         | +                    |
| Phycis phycis (Linnaeus, 1766)            |                        | +                       | + c                  |
| Sciaena umbra Linnaeus, 1758              |                        |                         | +                    |
| Scorpaena notata Rafinesque, 1810         |                        |                         | +                    |
| Serranus scriba (Linnaeus, 1758)          |                        |                         | +                    |
| Spicara maena (Linnaeus, 1758)            |                        |                         | +                    |
| Thorogobius ephippiatus (Lowe, 1839)      |                        | c                       | c                    |
| Total no. of taxa:                        | **122**                | **42**                  | **20**               |