A deep learning framework for high-throughput mechanism-driven phenotype compound screening and its application to COVID-19 drug repurposing

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Abstract

Phenotype-based compound screening has advantages over target-based drug discovery, but is unscalable and lacks understanding of mechanism. Chemical-induced gene expression profile provides a mechanistic signature of phenotypic response. However, the use of such data is limited by their sparseness, unreliability, and relatively low throughput. Few methods can perform phenotype-based de novo chemical compound screening. Here, we propose a mechanism-driven neural network-based method DeepCE, which utilizes graph neural network and multi-head

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Author contributions

LX and PZ conceived the project. THP, YQ and JZ extracted and preprocessed the data. THP and PZ developed the method. THP conducted the experiments. THP, YQ, and JZ analyzed the results. THP, YQ, LX, and PZ wrote the manuscript. All authors read and approved the final manuscript.

Competing interests

The authors declare no competing interests.

Data availability

The Bayesian-based peak deconvolution LINCS L1000 dataset is available at https://github.com/njpipeorgan/L1000-bayesian. The training, development, and testing gene expression sets used in our study, gene expression profiles generated from DeepCE for all drugs in DrugBank are available at https://github.com/pth1993/DeepCE.

Code availability

DeepCE source code and its usage instructions are available in Github (https://github.com/pth1993/DeepCE) and Zenodo (doi.org/10.5281/zenodo.3978774).
attention mechanism to model chemical substructure-gene and gene-gene associations, for predicting the differential gene expression profile perturbed by de novo chemicals. Moreover, we propose a novel data augmentation method which extracts useful information from unreliable experiments in L1000 dataset. The experimental results show that DeepCE achieves superior performances to state-of-the-art methods. The effectiveness of gene expression profiles generated from DeepCE is further supported by comparing them with observed data for downstream classification tasks. To demonstrate the value of DeepCE, we apply it to drug repurposing of COVID-19, and generate novel lead compounds consistent with clinical evidence. Thus, DeepCE provides a potentially powerful framework for robust predictive modeling by utilizing noisy omics data and screening novel chemicals for the modulation of a systemic response to disease.

Target-based high-throughput screening dominates conventional drug discovery process. It has been the focus of computer-aided drug discovery for decades including recent application of deep learning. However, the readout from the modulation of a single protein by a chemical is poorly correlated with organism-level therapeutic effect or side effect. As a result, the failure rate from a lead compound generated from the target-based screening to approved drug is high. Phenotype-based screening has created renewed interests for identifying cell-active compounds but suffered from low-throughput and difficulty in target deconvolution. Therefore, a high-throughput, mechanism-driven phenotype compound screening method will no doubt facilitate drug discovery and development.

Gene expression profiling has been widely used to characterize cellular and organismal phenotypes. Systematic analysis of genome-wide gene expression of chemical perturbations on human cell lines has led to significant improvements in drug discovery and systems pharmacology. In particular, it can be applied to drug repurposing\textsuperscript{1–4}, discovering drug mechanisms\textsuperscript{5}, lead identification\textsuperscript{6}, and predicting side effects for pre-clinical compounds\textsuperscript{7}. The use of genome-wide chemical-induced gene expression was initially made possible by the appearance of Connectivity Map (CMap)\textsuperscript{8}, which consists of gene expression profiles of five human cancer cell lines perturbed by \textasciitilde 1300 compounds after 6h. However, the limited data availability across cell types restricts the performances of these analyses which heavily depend on the coverage of chemicals and human cell lines. To overcome this limitation, a novel gene expression profiling method, L1000, which is the extension of CMap project, was developed by NIH library of integrated network-based cellular signatures (LINCS) program\textsuperscript{9}. After Phase I of LINCS program, L1000 dataset consists of \textasciitilde 1,400,000 gene expression profiles on the responses of \textasciitilde 50 human cell lines to one of \textasciitilde 20,000 compounds across a range of concentrations. Recently, L1000 dataset and its normalization versions\textsuperscript{10} are widely used in drug repurposing and discovery\textsuperscript{11, 12}. Despite these successes, there are several major problems when utilizing L1000 dataset. First, although the number of gene expression profiles is much larger than that in CMap, many missing expression values remain in the vast combinatorial space of chemicals and cell lines. Second, there are hundreds of millions of drug-like purchasable chemicals which are potential drug candidates\textsuperscript{13}. It is infeasible to experimentally test all of these chemicals for their chemical-induced gene expression profiles across multiple cell lines. Finally, due to various experimental problems (e.g. batch effect), many experiment measurements are not reliable (as shown in Supplementary Figure 1). These serious obstacles will limit the effectiveness
and scope of utilizing L1000 dataset in drug discovery. Therefore, predicting gene expression values for unmeasured and unreliable experiments are necessary.

Missing entries in the combinatorial space is the problem of not only L1000 dataset but also other gene expression profiling datasets. Before the appearance of L1000 dataset, several methods of imputing missing values have been proposed for gene expression datasets. We categorize these methods into two main approaches depending on the dependence of other information besides gene expression data. The first approach does not use any additional information. Works following this approach include k nearest neighbor (kNN)\(^{14}\), singular value decomposition\(^{14}\), least mean square\(^{15-17}\), Bayesian principal component analysis\(^{18}\), Gaussian mixture clustering\(^{19}\), and support vector regression\(^{20}\). The second approach uses additional information to predict expression profiles. For example, chemical structures are used to predict chemical-induced gene expressions but that work does not consider cell-specific information\(^{21}\).

The approaches described above are designed for matrix-structured data (i.e. gene × experiment) while L1000 dataset is formulated as tensor-structured data (i.e. gene × chemical × cell × doses × time) so they cannot be applied to capture high-dimensional associations that help to impute missing values for L1000 dataset. Recently, several methods are proposed to predict gene expression profiles in L1000 dataset. In particular, to deal with high-dimensional structured data, an extension of linear regression model named polyadic regression is developed to capture interactions emerging across features\(^{22}\). Matrix completion methods are also adapted to handle tensor-structured gene expression data\(^{23, 24}\).

Above methods for L1000 dataset just focus on imputing the missing values of some gene expression profiles or the whole gene expression profiles of some missing experiments. They are not very useful in the real setting of drug discovery where the chemical-induced gene expression profile of new chemicals needs to be identified. This motivates us to solve a more practical but more challenging problem: predicting gene expression profiles for de novo chemicals (i.e. chemicals that do not appear in training data). Solving this problem is necessary because it helps to infer gene expression profiles of new chemicals without conducting experiments that require time and human resources. More importantly, this problem can be expanded to predict gene expression profiles for new cell lines which can be difficult for measuring in in vitro environment. However, current computational approaches for predicting gene expression values for L1000 dataset cannot work well in de novo setting. In particular, tensor completion approach cannot predict gene expression profiles for new chemicals because of the inaccessibility to chemical features. Polyadic regression, theoretically, can predict gene expression profiles for high-dimensional data in de novo chemical setting because of using chemical features. However, in practice, it is not feasible because of huge computational resources required for handling high-dimensional data (i.e. this method fails when applied to more than 3-dimensional data). Therefore, there is a strong incentive to develop a new and effective method that exploits high-dimensional data for predicting gene expression profiles for de novo chemical setting.

To address the aforementioned problems, we design a mechanism-driven neural network-based model, DeepCE\(^{25}\), which captures high-dimensional associations among biological
features as well as non-linear relationships between biological features and outputs to predict gene expression profiles given a new chemical compound. Our proposed DeepCE significantly outperforms the state-of-the-art models for predicting gene expression profiles in L1000 dataset not only for de novo chemical setting but also for traditional imputation setting. Several novelties in the architecture of model contribute to the success of DeepCE. First, we leverage graph convolutional network to automatically extract chemical substructure features from data. Second, attention mechanism is used to capture associations among chemical substructures and genes, and among genes in cell lines. Finally, gene expression values layer which has similar architecture to the 1st layer in the of all L1000 genes are predicted simultaneously from hidden features by multi-output multi-layer feed-forward neural network. Besides developing this neural network-based model, we propose a data augmentation method by which we can extract useful information from unreliable experiments in L1000 dataset to improve the prediction performance of our model. We also verify the effectiveness of DeepCE by comparing the performances of several classification models trained on gene expression profiles generated from DeepCE and those trained on original gene expression profiles in L1000 dataset for two downstream tasks: drug-target and disease predictions. Finally, we assess the value of our proposed method for the challenge and urgent problem, finding treatment for COVID-19, by in silico screening all chemical compounds in DrugBank against COVID-19 patient clinical phenotypes. The prioritized lead compounds are consistent with existing clinical evidences. To our knowledge, it is the first work of phenotype-based drug repurposing for COVID-19. The source code of DeepCE and the generated gene expression profiles of all chemical compounds in DrugBank are publicly available for research purpose, which could make significant a contribution to drug discovery and development in particular, and computational chemistry and biology research in general.

**Chemical-induced Gene Expression Prediction Models and Datasets**

In the section, we present datasets used in our study and our proposed model, DeepCE, as well as baseline models for predicting gene expression profiles including linear models, vanilla neural network, k-nearest neighbor, and tensor-train weight optimization models. The general framework of training and testing these computational models for L1000 gene expression profile prediction is shown in Figure 1. Basically, computational models take L1000 experimental information (i.e. chemical compound, cell line, time stamp, and chemical dose size) from L1000 dataset as inputs, transform them into numerical representations, and then predict L1000 gene expression profiles based on these representations. The details of the numerical feature transformation process for chemical and biological objects used in our study and model implementation of DeepCE and other baselines are shown in Supplementary Notes. Moreover, in this section, we present the data augmentation method that extracts useful information from unreliable experiment in L1000 dataset to improve the prediction performance of our models, and the evaluation method for our models.
Datasets

In the following paragraphs, we present the details and the usages of several biological datasets in our study including L1000, STRING, DrugBank, and transcriptome data of COVID-19 patients. We also give the summary of these datasets in Supplementary Table 1.

Bayesian-based peak deconvolution L1000 dataset

After the original version of L1000 dataset was released\(^9\), many efforts have been made to improve the quality of this dataset. For example, instead of using k-means clustering algorithm as in the original version, some works propose to use Gaussian mixture model to enhance the accuracy of peak deconvolution step\(^{26, 27}\). One work, in another way, develops a multivariate method called Characteristic Direction to compute gene signatures instead of using the moderated Z-score as in the original version\(^{10}\). In our study, we conduct experiments on Bayesian-based peak deconvolution L1000 dataset which has been shown to generate more robust z-score profiles from L1000 assay data, and therefore, gives better representation for perturbagens\(^{28}\). In particular, we train and evaluate our proposed methods on level 5 data of this dataset. The gene expression profiles result from experiments of 7 most frequent cell lines and 6 most frequent chemical dose sizes in L1000 dataset are used to construct our gene expression dataset. We then select high-quality experiments from our dataset and split into high-quality training set, and development and testing set. We also construct the original training set by keeping unreliable experiments in our gene expression dataset and the augmented training set generated by our data augmented algorithm. The details of constructing these sets are described in Supplementary Notes. The statistics of these training, development, and testing sets are shown in Supplementary Table 2.

STRING database for human protein-protein interactions

STRING\(^{29}\) is a multi-source database of protein-protein interactions. These interactions which can be known or predicted, direct (physical) or indirect (functional) are collected from five main sources including genomics context prediction, high-throughput lab experiments, conserved co-expression, automated text-mining, and previous knowledge databases. In our setting, we extract the human protein-protein interaction network (i.e. ~ 19,000 nodes (proteins) and ~ 12,000,000 edges (interactions)) from this database to compute vector representations for L1000 genes. The drug-target vector representations for chemical compounds used in our study are also computed from this human protein-protein interaction network. The details of generating these representations from STRING database are shown in Supplementary Notes.

DrugBank database for drug-target interaction and disease predictions

DrugBank is a well-known, comprehensive database used in many bioinformatics and cheminformatics tasks\(^{30}\). This database consists of information about drugs and their targets. In our experiments, we extract ATC labels derived from the first level of ATC tree and targets of drugs appeared in L1000 dataset from DrugBank. There are 698 drug targets and 14 ATC labels in the extracted dataset. We select the most frequent ATC labels and drug targets based on their frequencies on this dataset as the labels of drugs to form drug-target prediction and ATC prediction datasets. These datasets are used to evaluate the performance...
of gene expression profiles generated from our models. We also predict gene expression profiles for all drugs in DrugBank and use them to screen potential candidates for COVID-19 treatment.

Patient expression in response to SARS-CoV-2 infection

Patient expression datasets for this study are downloaded from National Genomics Data Center (NGDC) (PRJCA002273)\textsuperscript{31} and National Center for Biotechnology Information (NCBI) (GSE147507)\textsuperscript{32}. While the former includes 8 SARS-CoV-2 patients and 12 healthy samples, the latter has only 1 SARS-CoV-2 patient and 2 healthy samples. For each dataset, we use expression profiles from both SARS-CoV-2 patients and healthy negative controls for differential expression analysis. Thus, the first dataset can be considered as population-based gene expression analysis while the second dataset is patient-specific gene expression analysis. DESeq\textsuperscript{233} package is used to generate the differential gene expression profiles of the patients. Not all L1000 genes appear in the result of DESeq2 package so we only consider genes appear in both L1000 dataset and DEseq2 package when comparing with chemical-induced gene expression profiles.

Overall architecture of DeepCE

Our neural network-based model for L1000 gene expression profile prediction, DeepCE, consists of several components as follows. First, we use graph convolutional network to learn numerical representation for chemical compound from its graph structure and feed-forward neural network to learn numerical representations for cell line and chemical dose size. We also use numerical representations for L1000 genes which are derived from the human protein-protein interaction network (described in Supplementary Notes). After that, these vector representations are put into the interaction component to capture high-level feature associations including chemical substructure-gene and gene-gene feature associations. Finally, the prediction component takes the outputs of the interaction component as inputs to predict the gene expression values for all L1000 genes simultaneously. The overall architecture of DeepCE and its hyperparameters used in our experiments are shown in Figure 2 and Supplementary Table 4 respectively. The following paragraphs describe each component of DeepCE in detail.

Graph convolutional network for neural fingerprint

Recently, data-driven chemical fingerprints are shown to be more effective than predefined chemical fingerprints (e.g. PubChem, ECFP) for many biological prediction problems. Therefore, we propose to use graph convolutional network (GCN) to capture the chemical substructure information. The original GCN model for chemical fingerprint\textsuperscript{34} takes a graph structure of chemical compound as input and update vector representations for each node (atom) in graph (chemical compound) from its neighborhoods by convolutional operation. Thus, the vector for each node after convolutional operation can be seen as the representation of chemical substructures. The final vector which is the sum of vectors of every node is used as the chemical fingerprint. GCN model used in our experiments is primarily based on that model but with a minor modification. In particular, we output vector representations for every nodes instead of one vector representation for the chemical compound because we want to model the associations of chemical substructure features with
gene features. In our settings, we use the GCN model with 2 convolutional layers (radius $R = 2$). It means that the output vector from GCN for each atom represents the chemical substructure which is a span of 2-hop distance from that atom. The initial representations for atoms and bonds are multi-hot vectors that capture the symbol, degree, number of Hydro neighborhoods, and aromaticity of atoms, and type of bonds that have lengths of 62 and 6, respectively. The details of GCN model used in our experiments are shown in Supplementary Algorithm 1.

**Multi-head attention for gene-gene and chemical substructure-gene feature associations**

Attention mechanisms where an element of one set selectively focuses on a subset of another set (attention) or its set (self-attention) based on attention weights are used widely in neural network-based models and effectively applied to many AI tasks including computer vision and natural language processing. In our experiments, we propose to apply the attention method named multi-head attention for modeling associations among gene features, and among gene and chemical substructure features. Multi-head attention was first proposed in Transformer model which achieves state-of-the-art results for many natural language processing tasks\(^{35}\). Basically, each element in sets can be represented by a set of three vectors query, key, and value. An individual attention module is a function of mapping queries and sets of key-value pairs to output matrix computed by:

$$\text{Attention}(Q, K, V) = \text{softmax}\left(\frac{QK^T}{\sqrt{d_k}}\right)V$$

where $Q, K, V$ are matrices (sets) of queries, keys, values respectively and $d_k$ is a scaling factor. Multi-head attention focuses on different representation subspaces by concatenating several individual attention modules:

$$\text{MultiHead}(Q, K, V) = \text{concat}(\text{head}_1, \ldots, \text{head}_h)w^o$$

where $\text{head}_i = \text{Attention}(QW_i^Q, KW_i^K, VW_i^V)$.

This multi-head attention mechanism is the main ingredient used to construct the interaction component of DeepCE. In particular, the interaction component consists of two identical layers where outputs of the first layer are used as inputs for the second layer. For each layer, we use 2 separate multi-head attention modules with 4 heads for each module to model associations among genes in gene set and among elements in gene set and chemical substructure set. Length of query, key, value vectors is set at 512. Outputs from these two multi-head attention modules are concatenated and put into normalization layer followed by feed-forward layer and another normalization layer. The abstract architecture of interaction component is shown in Figure 2.

**Multi-output prediction**

The multi-output prediction component which is a 2-layer feed-forward neural network with ReLU activation function takes input as the concatenation of chemical neural fingerprint,
gene feature generated by interaction component, cell line and chemical dose size features to predict gene expression values for all L1000 genes together as follows:

$$Y = W_2(ReLU(W_1X + b_1)) + b_2$$

where $W_1, W_2, b_1, b_2$ are weight matrices and bias vectors of this network. The output size of this feed-forward neural network is set at 978 which is the number of L1000 genes.

**Objective function**

The objective function used in DeepCE model is mean squared error (MSE) between predicted and ground-truth gene expression values and is computed as follows:

$$\text{loss}_{\text{DeepCE}}(\Theta) = \frac{1}{NM} \sum_{i=1}^{N} \sum_{j=1}^{M} (z_{i,j} - y_{i,j})^2$$

where $\Theta$ are the set of parameters in DeepCE model. $N$ and $M$ the number of gene expression profiles in a batch and number of L1000 genes respectively. $Z_{i,j}$ and $y_{i,j}$ are ground-truth and predicted gene expression values of $j^{th}$ gene in $i^{th}$ gene expression profile.

**Baseline Models**

In this section, we describe several baseline models used in our experiments including linear models, vanilla neural network, k-nearest neighbor, and tensor-train weight optimization.

**Linear models**

We experiment with multi-output linear regression model and its regularization versions including Lasso regression (L1 regularization) and ridge regression (L2 regularization) models. Like DeepCE, input for these models is the concatenation of numerical representations for chemical, gene, cell line, and chemical dose size features but we use predefined chemical fingerprints and drug-target features instead of data-driven representations derived from GCN for chemicals. The details of these representations are described in Supplementary Notes. Multi-output linear models can be seen as 1-layer feed-forward neural network without activation function.

**Vanilla neural network**

The vanilla neural network used in our experiments can be seen as the simpler version of DeepCE model that does not include the interaction network component for modeling gene-gene and gene-chemical substructure feature associations and GCN for generating neural fingerprints. Input for this vanilla neural network is similar to its for linear models. The following layers in this network are similar to the prediction network component in DeepCE model which is a 2-layer feed-forward neural network with ReLU activation function.
K-nearest neighbor

We also propose a k-nearest neighbor-based approach for gene expression prediction for de novo chemical setting. In particular, gene expression profile for a new chemical compound in one particular setting (i.e. cell line, chemical dose size) is generated by averaging gene expression profiles of its nearest neighborhoods in the training set in the same setting. In our research, we experiment with different numbers of neighborhoods from 1 to 15 and different similarity measures including cosine, correlation, Jaccard and Tanimoto similarity as well as euclidean distance.

Tensor-train weight optimization

Tensor-train weight optimization (TT-WOPT) is a tensor completion approach proposed to retrieve missing values in tensor data from existing values. It has been shown to be effective for predicting missing values of L1000 dataset which can be formulated as a tensor-structure object without using additional information\(^\text{24}\). In our research, we conduct experiments to compare it with our proposed model, especially in de novo chemical setting. Because this model does not require additional information so input for it is L1000 gene expression values formulated as a tensor.

Data augmentation

From Supplementary Figure 1, we can see that only a small number of experiments in L1000 dataset are reliable (i.e. APC score $\geq 0.7$) so it would be wasteful if we cannot exploit useful information from a large number of unreliable experiments. It will be shown in the Results section (i.e. Table 1) that simply adding unreliable experiments to the high-quality training set (original training set) makes the performances of our models worse. Thus, we propose the data augmentation method by which we can effectively exploit unreliable experiments to improve the performances of our models. We argue that although an experiment (level 5 data) is unreliable, not all its bio-replicates experiments (level 4 data) are also unreliable and we will extract these reliable bio-replicate experiments by our proposed data augmentation method. The basic idea is that we, first, train our model on the high-quality training set, and then, generate predicted gene expression profiles for unreliable experiments. These predicted gene expression profiles are compared with their bio-replicate gene expression profiles and we incorporate bio-replicate gene expression profiles that have the similarity scores with their predicted gene expression profiles larger than the threshold. Supplementary Algorithm 2 presents this data augmentation method in detail. In our settings, the similarity score is Pearson correlation.

Performance evaluation

Pearson correlation coefficient is used as the main metric to evaluate performances of models in our experiments. Correlation scores which measure the relationship between ground-truth and predicted gene expression profiles have been shown to be more effective than error measures for microarray data analysis\(^\text{36, 37}\). Moreover, using Pearson correlation allows us to conduct unbiased evaluation for our models which are optimized for mean squared error. We calculate the average Pearson correlation for a dataset as follows:
\[ r = \frac{1}{N} \sum_{i=1}^{N} \frac{\sum_{j=1}^{M} (z_{i,j} - \bar{z}_i)(y_{i,j} - \bar{y}_i)}{\sqrt{\sum_{j=1}^{M} (z_{i,j} - \bar{z}_i)^2} \sqrt{\sum_{j=1}^{M} (y_{i,j} - \bar{y}_i)^2}} \]

where \( N \) and \( M \) are number of gene expression profiles in the dataset and number of L1000 genes respectively. \( z_{i,j}, y_{i,j}, \bar{z}_i, \bar{y}_i \) are ground-truth and predicted gene expression values of \( j^{th} \) gene in \( i^{th} \) gene expression profile and ground-truth and predicted mean values of \( i^{th} \) gene expression profile.

Besides Pearson correlation, we also report the performances of models by other metrics including root mean squared error (RMSE), gene set enrichment analysis (GSEA)\(^{38, 39} \), and Precision@k. While Pearson correlation and RMSE captures the variations among all L1000 genes, GSEA and P@k (including both positive and negative P@k) only focus on the most significant up and down regulated genes. Thus, using multiple metrics allow us to measure the performances of models in different aspects. The details of these additional metrics are shown in the Supplementary Notes.

Furthermore, we use area under the receiver operating characteristic curve (AUC) to verify the effectiveness of these predicted profiles for downstream binary classification tasks including drug-target and ATC code predictions.

**Results and Discussions**

The results and discussions below are mainly based on Pearson correlation, and we also observe the same patterns by using other metrics.

**DeepCE significantly outperforms baseline models in the novel chemical setting**

In this experiment, we compare DeepCE model and its simpler variants constructed by removing either the whole interaction component or just one part of its (i.e. chemical substructure-gene or gene-gene feature association modules) with several baseline models including vanilla neural network, kNN, linear models, and TT-WOPT. While TT-WOPT predicts output based on gene expression values only, other models learn the relationship between experimental information and gene expression profiles to make predictions. For DeepCE, we use neural fingerprints while for other models, we use predefined fingerprints including PubChem and circular (ECFP6) fingerprints, and drug-target information including latent target interaction profile (LTIP)\(^{40} \) and our proposed drug-target feature to represent chemicals. All models are trained on the high-quality training set and are evaluated on the test set.

As listed in Table 1, DeepCE model and its variants achieve order-of-magnitude improvements over baseline models. In particular, DeepCE model significantly outperforms other models including vanilla neural network, kNN, linear models, and TT-WOPT by achieving a Pearson correlation of 0.4907 on the testing set (paired t-test, \( p – value < 4.63 \times 10^{-15} \)). Comparing to its simpler variants whose interaction components are removed, DeepCE also achieves better performance, indicating that the effectiveness of modeling
chemical substructure-gene and gene-gene feature associations. Specifically, the performance of DeepCE decreases to 0.4620, 0.4477, and 0.4418 when removing chemical substructure-gene feature association part (Deep CE·drug-gene attn), gene-gene feature association part (Deep CE·gene-gene attn), and the whole interaction component (Deep CE·attn) (paired t-test, p-value < 2.25 × 10^{-5}), respectively. We also delve deeper into the performance of DeepCE by looking it over cell lines, chemical dose sizes, and L1000 genes. The results of this analysis are shown in Supplementary Figures 2 and 3. For baseline models, vanilla neural networks and kNN achieve pretty good performances. Linear models including linear regression, Lasso, and ridge regression do not work well for our problem. It indicates that the linear relationship is not sufficient to model the dependencies among variables in this dataset. TT-WOPT, which does not leverage additional features besides gene expression values to make the predictions, as we expect, does not work well for de novo chemical setting. In particular, it achieves a Pearson correlation of 0.0144 which is similar to randomness. We also provide error estimate for these performances by conducting 5-fold cross-validation on the high-quality dataset. The results are shown in Supplementary Table 5.

DeepCE outperforms the state-of-the-art methods in the imputation setting

We further investigate the performance of DeepCE for the traditional imputation setting that does not require chemicals in the testing set to be different from chemicals in the training set, and compare it with TT-WOPT which has been shown to be effective for this setting. To do that, we randomly split the high-quality dataset to the new training, development, and testing sets and conduct the experiment on these sets. Note that, at this time, we split the dataset by gene expression profile instead of chemical compound. The details of the training, development, and testing set for imputation setting are shown in Supplementary Table 3.

For the traditional imputation setting, we observe DeepCE outperforms TT-WOPT with a large margin. In particular, DeepCE achieves a Pearson correlation of 0.7010 compared to its of 0.5113 of TT-WOPT. This result indicates that DeepCE consistently achieves the best performances for both de novo chemical and traditional imputation settings by effectively leveraging features of chemical and biological objects including chemical compounds and genes.

Chemical similarity has an impact on prediction performance

To investigate thoroughly the prediction performance of our models, we investigate the impact of chemical similarity between testing set and training set. In particular, we compute the distance between one experiment in the testing set and its nearest neighbors experiments in the training set which are induced by the most similar chemicals (i.e. determined by comparing their fingerprints with the fingerprint of the chemical compound induced the experiment in the testing set) on the same cell line. The distance between the two experiments is the Tanimoto coefficient of PubChem fingerprints of their two chemicals, and the distance between the experiment on the testing with its nearest neighbor experiments in the training set is the average of distances between that experiment and each of its nearest neighbors. After computing the distances to the training set for all experiments on the testing set, we sort them by the ascending order and compare the Pearson correlation scores of these
experiments. We calculate the average Pearson correlation scores of all experiments in the testing set that have their distances to the training set smaller than the first quartile (Q1), from Q1 to the second quartile (Q2), from Q2 to the third quartile (Q3), and larger than Q3 of the sorted list. Figure 3 shows the average Pearson correlation scores with these distances of three models including DeepCE, vanilla neural network, and kNN. From this figure, we can see the same pattern for all models that the prediction performances are higher when the experiments in the testing set are more similar to their nearest neighbor experiments on the training set. We also recognize that DeepCE achieves better performances than vanilla neural network and kNN for all distance categories, especially for experiments that have their distances to the training set smaller than Q1.

**Data quality has a significant impact on prediction performance**

Besides sparseness problem, L1000 dataset also includes many unreliable gene expression profiles. To investigate the impact of noisy profiles on the prediction performances of our models, we train two baseline models including neural network and kNN on different training sets generated by filtering unreliable gene expression profiles with different average Pearson correlation (APC) thresholds varying from −1 (original training set) to 0.7 (high-quality training set). Chemical feature used in this experiment is PubChem fingerprint.

As shown in Figure 4, all models have the same pattern. Starting at the threshold of 0.1, they achieve better performances on the testing set when the threshold is higher and the best setting is training our models on the high-quality training set (i.e. Pearson correlation of 0.3923 for vanilla neural network and 0.3903 for kNN). For training on the original training set and other training sets generated by filtering unreliable experiments with thresholds < 0.1, the ground-truth and predicted gene expression profiles are uncorrelated showing the randomness of the model predictions. These results indicate that unreliable data has a severely negative impact on prediction performances and removing this part from the dataset is necessary for achieving good performances.

**A novel data augmentation method improves the model performance**

We propose the data augmentation method (described in detail in Supplementary Algorithm 2) to effectively exploit useful information from unreliable gene expression profiles. In this experiment, we evaluate the impact of this method on our models. In particular, DeepCE trained on high-quality training set are used to generate gene expression profiles and the threshold for selecting bio-replicate profiles is 0.5 which is similar to the performance of DeepCE. The statistics of this augmented training set are shown in Supplementary Table 1.

The experimental results for training vanilla neural network, kNN, and DeepCE on the augmented training set are shown in Table 1. We can see that the performances of all models trained on this augmented training set are improved in most cases. For example, the Pearson correlation of DeepCE is increase from 0.4907 to 0.5014 (paired t-test, \( p - value < 0.05 \)). These results indicate that information extracted from unreliable gene expression profiles by our data augmentation method is effective for gene expression prediction.
Selection of chemical feature affects model performance

In this experiment, we investigate the effectiveness of several chemical feature representations for our models. Models used in this experiment are vanilla neural network for PubChem, ECFP fingerprints, our proposed drug-target features, and LTIP, and DeepCE model without interaction component for neural fingerprint. These models are trained on the high-quality training set. We also create random chemical features by generating random binary vectors whose size is similar to PubChem fingerprint from discrete uniform distribution.

Table 1 shows the performances measured by Pearson correlation of these models with different chemical feature representations. First, chemical features achieve much better performances than the random feature, indicating that chemical features capture important information about chemicals which is useful for predicting gene expression profiles. Second, DeepCE which uses neural fingerprint achieves the Pearson correlation of 0.4418 which is the best performance compared to other settings (paired t-test, \(P - value < 4.89 \times 10^{-5}\)). For other chemical features, biological-based features including drug-target feature and LTIP achieves slightly better performances than chemical-based features including PubChem and ECFP fingerprints. All of these observations are verified by the paired t-tests with \(P - values < 0.01\). In fact, most of the \(P - values\) are much less than 0.01.

We also conduct an ablation study to investigate the impact of other features (i.e. cell line, dose size) to the predictive performance by removing them from the feature vectors. The results in Supplementary Table 6 show that removing these features decreases the performance of DeepCE and the worst scenario is when removing both cell line and dose size information.

DeepCE is effective in predictive down-stream tasks

In this section, we design an experiment to answer a question about whether these predicted gene expression profiles can provide added values for downstream prediction tasks, especially in the case that original gene expression profiles in L1000 dataset are unreliable. We first extract gene expression profiles of chemicals that do not have reliable experiments in L1000 dataset (original feature set) as well as use DeepCE model trained on high quality training set to generate gene expression profiles for these drugs (predicted feature set). We then use these sets as the features for drugs to train classification models for two tasks: ATC code and drug-target predictions. The details of constructing these datasets are presented in Supplementary Notes and Supplementary Table 7. Finally, we train 4 popular classification models including logistic regression (LR), support vector machine (SVM), k-nearest neighbor (kNN), and decision tree (DT) using 14 different versions of chemical features (7 cell-specific features for each original and predicted feature sets) for 14 binary classification tasks (i.e. 10 ATC codes and 4 drug-targets). For each experiment setting, we use 5-fold cross-validation and report the average results.

The differences in AUC between training classification models with predicted and original feature sets for drug-target and ATC prediction tasks are shown in Extended Data Figure 1. The improvements in AUC when using predicted features instead of original features are
recognized in all cell-specific profiles (Extended Data Figure 1a), all classification models (Extended Data Figure 1b), 8/10 ATC codes (Extended Data Figure 1c), and 3/4 drug-targets (Extended Data Figure 1d), and these improvements are significant (paired t-test, $P$-value $< 4.87 \times 10^{-5}$). The details of AUC scores for predicted and original features for each setting (i.e. per model, cell line, ATC code, and drug-target) are shown in Supplementary Table 8. These results indicate that we can substitute unreliable gene expression profiles in L1000 dataset with gene expression profiles generated from DeepCE to achieve better performances on downstream prediction tasks.

**Drug repurposing for COVID-19**

To further demonstrate the value of DeepCE, we use chemical-induced gene expression profile to discover potential drugs for COVID-19 treatment. Because the disease state and symptom of COVID-19 patients vary dramatically depending on many factors such as age, gender, and underlying conditions, etc., we, therefore, evaluate the drug repurposing for COVID-19 task under 2 settings including population-based (group of patients) and individual-based (individual patient) analysis. In particular, we first use the trained DeepCE on the high-quality part of L1000 dataset to generate predicted gene expression profiles for all of 11179 drugs in Drugbank database at the largest chemical dose size. For patient gene expression profiles, we use SARS-COV-2 gene expression datasets from NCBI and NGDC to calculate the differential gene expression profiles of the patients under population-based and individual-based settings respectively. Specifically, DESeq2 package is used to generate the patient profiles from 8 SARS-CoV-2 patients and 12 healthy samples (population-based) and from 1 SARS-CoV-2 patient and 2 healthy samples (individual-based). We then screen drugs in Drugbank by computing Spearman’s rank-order correlation scores between their gene expression profiles with the patient gene expression profiles and select drugs that give the most negative scores as the potential drugs. Here, we incorporate the gene expression profiles of A549 - the cancerous lung tissue - beside the main 7 cell lines in the high-quality dataset. Besides the predicted profiles, we also include the gene expression profiles extracted from the high-quality part of L1000 dataset. For each cell line, we extract the top 100 drugs that have the most negative correlation scores with the patient profile as the potential drugs. Finally, we output drugs that are potential drugs for COVID-19 treatment at all cell lines as the result of our screening process.


The results for the population-based and individual-based drug repurposing are shown in Table 2 and Extended Data Figure 2 respectively. COVID-19 induced acute respiratory failure is thought to be related to both direct viral pathogenicity and dysregulated inflammatory host response. As shown in Table 2, among the 10 drugs we identified for population-based analysis, 3 drugs are antiviral drugs used in Hepatitis C and two drugs are immunosuppressive agents. In particular, Voclorsporin and Cyclosporine are immunosuppressant and calcineurin inhibitors that share similar structures. Cyclosporine has been used to prevent organ rejection and to treat T cell-associated autoimmune diseases, and recently shows potential in preventing uncontrolled inflammatory response, SARS-CoV-2 replication, and acute lung injury caused by COVID-19. Besides that, calcineurin inhibitors have been shown to be promising treatment for severe COVID-19 cases. Alisporivir which is a nonimmunosuppressive analogue of cyclosporine with potent

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cyclophilin inhibition properties is shown effective reducing in SARS-CoV-2 RNA production in Vero E6 cells\textsuperscript{47}. Moreover, Valspodar inhibits P-glycoprotein, which affects the transportation of immunosuppressive agents, and Ceftobiprole Medocaril is used in hospital-acquired and community-acquired pneumonia\textsuperscript{48}.

For individual-based analysis, among the 15 drugs we identified (Extended Data Figure 2), 9 drugs are antiviral drugs and 7 of them are used in treating Hepatitis C as NS5A inhibitor. They are similar to the top-ranked drugs that are identified from the population-based analysis. Especially, two of Hepatitis C treatment, Elbasvir and Velpatasvir, have been shown as potential candidates for COVID-19 treatment by using other approaches\textsuperscript{49–51}. Moreover, two drugs show anti-inflammatory or immune regulating function, and have the potential in regulate immune response under COVID-19 infection. Laniquidar can suppress the function of P-glycoprotein 1 and affect transportation of immunosuppressive agents. The population-based analysis also recognizes the drugs with the similar mode of actions. AMG-487 targets Chemokine receptor CXCR3, which can regulate leukocyte trafficking. It is noted that all potential drugs here are not available in L1000 dataset, showing the effectiveness of DeepCE for phenotype compound screening under both population-based and individual-based settings.

**Conclusion**

Deep learning has attracted a great attention in drug discovery. Past and existing efforts mainly focus on accelerating compound screening against a single target\textsuperscript{52}. However, such one-drug-one-gene paradigm is proved to be less successful in tracking complex diseases. A systematic compound screening approach, which both takes information on biological system into consideration and uses chemical-induced systematic response as readouts, will provide new opportunities on discovering safe and effective therapeutics that module the biological system. In this study, we have proposed DeepCE - a novel and robust neural network-based model for predicting chemical-induced gene expression profiles from chemical and biological objects, especially in de novo chemical setting. Our model achieves state-of-the-art results of predicting gene expression profiles compared to other models not only in de novo chemical setting but also in the traditional setting. In addition, we have addressed the unreliable measurement problem of L1000 dataset by introducing the data augmentation method to effectively exploit useful information from unreliable gene expression profiles to improve the prediction performances of our models. Furthermore, the downstream prediction task evaluation shows that training classification models with gene expression profiles generated from DeepCE achieves better performances than training them with unreliable gene expression profiles in L1000 dataset, indicating the added values of DeepCE for downstream prediction. Finally, DeepCE is shown to be effective in the challenge and urgent problem, finding treatment for COVID-19, by in silico screening all chemical compounds in DrugBank against COVID-19 patient clinical phenotypes (i.e. comparing chemical-induced gene expression profiles generated from DeepCE with the patient profiles). In summary, DeepCE could be a powerful tool for phenotype-based compound screening.
Extended Data Fig. 1. Improvement of predicted profiles over original profiles in AUC.
(a) Per cell-specific profile, across experiments for different classification tasks and models.
(b) Per model, across experiments for different cell-specific profiles and classification tasks.
(c) Per ATC code, across experiments for different cell-specific profiles and models.
(d) Per drug-target, across experiments for different cell-specific profiles and models.
| Drug             | Structure | Status     | Known Uses                        |
|------------------|-----------|------------|-----------------------------------|
| Elbasvir         | ![Structure](image1.png) | Approved   | Hepatitis C, NS5A inhibitor       |
| Pibrentasvir     | ![Structure](image2.png) | Approved   | Hepatitis C, NS5A inhibitor       |
| Velpatasvir      | ![Structure](image3.png) | Approved   | Hepatitis C, NS5A inhibitor       |
| Ruzasvir         | ![Structure](image4.png) | Investigational | Hepatitis C, NS5A inhibitor     |
| Samatasvir       | ![Structure](image5.png) | Investigational | Hepatitis C, NS5A inhibitor     |
| Odalasvir        | ![Structure](image6.png) | Investigational | Hepatitis C, NS5A inhibitor     |
| Cobopasvir       | ![Structure](image7.png) | Investigational | Hepatitis C, NS5A inhibitor     |
| Baloxavir Marboxil | ![Structure](image8.png) | Approved   | Influenza A and B                 |
| Metocurine       | ![Structure](image9.png) | Approved   | Muscle relaxant                   |
| Dactinomycin     | ![Structure](image10.png) | Approved   | Cancer                            |
| Laniquidar       | ![Structure](image11.png) | Investigational | Cancer, P-glycoprotein inhibitor |
| Tadalafil        | ![Structure](image12.png) | Approved   | Erectile Dysfunction, PDE5 inhibitors |
| GE-2270A         | ![Structure](image13.png) | Experimental | Antibiotic                        |
| SD146            | ![Structure](image14.png) | Experimental | Binds HIV-1 protease              |
| AMG-487          | ![Structure](image15.png) | Experimental | CXCR3 antagonist                   |

**Extended Data Fig. 2.**
The chemical structures, status, and known uses of potential drugs for COVID-19 treatment (i.e. drugs appeared in top 100 drugs for all 8 cell lines when comparing their cell-specific predicted gene expression profiles with the individual-based patient profile by Spearman’s correlation.

**Supplementary Material**
Refer to Web version on PubMed Central for supplementary material.
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Figure 1.
General framework of training computational models for L1000 gene expression profile prediction and using them for downstream application (i.e. drug repurposing for COVID-19 treatment). The objective for the learning process is minimizing the loss between predicted profiles and ground-truth profiles in L1000 dataset. After training, models is used for generating profiles for new chemicals in external molecular database (i.e. DrugBank). These profiles are then used for in silico screening (i.e. comparing with patient gene expression) to find potential drugs for COVID-19 treatment.
Figure 2.
Overall architecture of DeepCE (The details of 2nd layer which has similar architecture to the 1st layer in the interaction network are omitted to save space)
Figure 3.
Performances of DeepCE, vanilla neural network, and kNN with different distances among chemicals in the training and testing sets
Figure 4.
Pearson correlation scores of vanilla neural network and kNN trained on training sets generated by filtering unreliable experiments with different APC thresholds.
## Table 1.

Performances (i.e. Pearson correlation, RMSE, GSEA, positive and negative Precision@100) on testing set of vanilla neural network, kNN, linear models with different chemical features, TTWOPT, and DeepCE with its simpler variants trained with different training sets

| Training sets | Models       | Features | Pearson | RMSE   | GSEA   | Pos P@100 | Neg P@100 |
|---------------|--------------|----------|---------|--------|--------|-----------|-----------|
|               | Vanilla Neural Network | PubChem  | 0.1101  | -      | -      | -         | -         |
|               |               | ECFP     | 0.0705  | -      | -      | -         | -         |
|               |               | Drug-target | 0.1076  | -      | -      | -         | -         |
|               |               | LTIP     | 0.0770  | -      | -      | -         | -         |
| Original      | kNN          | PubChem  | 0.0844  | -      | -      | -         | -         |
|               |               | ECFP     | 0.1469  | -      | -      | -         | -         |
|               |               | Drug-target | 0.1811  | -      | -      | -         | -         |
|               |               | LTIP     | 0.1231  | -      | -      | -         | -         |
| High-quality  | Vanilla Neural Network | PubChem  | 0.3929  | 1.8413 | 0.3853 | 0.2230   | 0.2622   |
|               |               | ECFP     | 0.4105  | 1.8218 | 0.4049 | 0.2353   | 0.2690   |
|               |               | Drug-target | 0.4270  | 1.8002 | 0.4098 | 0.2334   | 0.2788   |
|               |               | LTIP     | 0.4259  | 1.7843 | 0.4168 | 0.2361   | 0.2798   |
|               |               | Random   | 0.3129  | 1.9152 | 0.3299 | 0.1729   | 0.2284   |
|               | kNN          | PubChem  | 0.3903  | 1.8464 | 0.3877 | 0.2089   | 0.2606   |
|               |               | ECFP     | 0.3991  | 1.8264 | 0.4041 | 0.2186   | 0.2639   |
|               |               | Drug-target | 0.3907  | 1.8375 | 0.4105 | 0.2182   | 0.2625   |
|               |               | LTIP     | 0.3922  | 1.8388 | 0.3959 | 0.2176   | 0.2578   |
|               | Linear Regression | PubChem  | 0.1762  | 1.9821 | 0.2184 | 0.1220   | 0.1956   |
|               |               | ECFP     | 0.1770  | 1.9916 | 0.2227 | 0.1232   | 0.1956   |
|               |               | Drug-target | 0.1763  | 1.9768 | 0.2216 | 0.1240   | 0.1957   |
|               |               | LTIP     | 0.1764  | 1.9769 | 0.2232 | 0.1230   | 0.1956   |
|               | Lasso        | PubChem  | 0.1761  | 1.9775 | 0.2160 | 0.1203   | 0.1935   |
|               |               | ECFP     | 0.1770  | 1.9763 | 0.2237 | 0.1198   | 0.1961   |
|               |               | Drug-target | 0.1764  | 1.9764 | 0.2177 | 0.1209   | 0.1935   |
|               |               | LTIP     | 0.1764  | 1.9764 | 0.2177 | 0.1213   | 0.1916   |
| Training sets | Models | Features | Pearson | RMSE | GSEA | Pos P@100 | Neg P@100 |
|---------------|--------|----------|---------|------|------|-----------|-----------|
| Ridge Regression | PubChem | 0.1762 | 1.9809 | 0.2185 | 0.1220 | 0.1961 |
| | ECFP | 0.1770 | 1.9839 | 0.2254 | 0.1236 | 0.1953 |
| | Drug-target | 0.1764 | 1.9764 | 0.2221 | 0.1232 | 0.1956 |
| | LTIP | 0.1764 | 1.9762 | 0.2237 | 0.1215 | 0.1953 |
| | TT-WOPT | N/A | 0.0133 | 1.9695 | 0.0121 | 0.1228 | 0.1342 |
| | Deep CE | - atm | 0.4418 | 1.7738 | 0.4088 | 0.2435 | 0.2827 |
| | | Neural FP | 0.4620 | 1.7418 | 0.4493 | 0.2667 | 0.3088 |
| | | - drug-gene atm | 0.4477 | 1.7711 | 0.4244 | 0.2784 | 0.2961 |
| | | Neural FP | 0.4907 | 1.6889 | 0.4656 | 0.2885 | 0.3195 |
| | | Deep CE | 0.4204 | 1.8140 | 0.3932 | 0.2282 | 0.2736 |
| | | | 0.4177 | 1.8102 | 0.4171 | 0.2191 | 0.2783 |
| | | | 0.4302 | 1.8092 | 0.4263 | 0.2130 | 0.2785 |
| | | | 0.4299 | 1.7819 | 0.4237 | 0.2259 | 0.2810 |
| | Vanilla Neural Network | PubChem | 0.3973 | 1.8392 | 0.3927 | 0.2023 | 0.2615 |
| | | ECFP | 0.4121 | 1.8020 | 0.4204 | 0.2202 | 0.2809 |
| | | Drug-target | 0.4023 | 1.8072 | 0.4011 | 0.2232 | 0.2794 |
| | | LTIP | 0.4016 | 1.8223 | 0.3924 | 0.2184 | 0.2650 |
| | Augmented | PubChem | 0.5014 | 1.6810 | 0.4735 | 0.2940 | 0.3249 |
| | | ECFP | 0.5014 | 1.6810 | 0.4735 | 0.2940 | 0.3249 |
| | | Drug-target | 0.5014 | 1.6810 | 0.4735 | 0.2940 | 0.3249 |
| | | LTIP | 0.5014 | 1.6810 | 0.4735 | 0.2940 | 0.3249 |
The chemical structures, status, and known uses of potential drugs for COVID-19 treatment (i.e. drugs appeared in top 100 drugs for all 8 cell lines when comparing their cell-specific predicted gene expression profiles with the population-based patient profiles by Spearman’s correlation. Experimental and investigational drugs are those at the preclinical or animal testing stage and in human clinical trials, respectively.

| Drug          | Structure | Status              | Known Uses                  |
|---------------|-----------|---------------------|------------------------------|
| Faldaprevir   | ![Faldaprevir Structure](image1) | Investigational | Hepatitis C, Protease Inhibitors |
| Alisporivir   | ![Alisporivir Structure](image2) | Investigational | Hepatitis C                  |
| NIM811        | ![NIM811 Structure](image3) | Investigational | Hepatitis C                  |
| Ceftobiprole medocaril | ![Ceftobiprole medocaril Structure](image4) | Experimental, Investigational | Antibiotics, Pneumonia       |
| Anidulafungin | ![Anidulafungin Structure](image5) | Approved, Investigational | Antifungal                  |
| Drug         | Structure | Known Uses                                      | Status       |
|--------------|-----------|-------------------------------------------------|--------------|
| Oteseconazole|           | Antifungal                                      | Investigational |
| Voclosporin  |           | Immunosuppressants, Calcineurin Inhibitor       | Investigational |
| Cyclosporine |           | Immunosuppressants, Calcineurin Inhibitor       | Approved, Investigational |
| Valspodar    |           | Cancer, P-glycoprotein Inhibitor                | Investigational |
| Evacetrapib  |           | Cardiovascular, CETP Inhibitor                 | Investigational |