Descriptions of two new genera of Maruinini (Diptera, Psychodidae, Psychodinae) from the Mitaraka range of French Guiana

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ABSTRACT
Little information is available regarding the non-phlebotomine Psychodidae Newman, 1834 of French Guiana. Psychodid specimens were sorted from samples collected during the recent “Our Planet Reviewed” Guyane-2015 expedition in the Mitaraka range. Males and females of two genera of tribe Maruinini Enderlein, 1937, Polletomyia subulata n. gen., n. sp. and Myiomystax trilineata n. gen., n. sp., are described and illustrated. Characters pertinent to assessing relationships among genera of Maruinini are discussed.

RÉSUMÉ
Description de nouveaux genres de Maruinini (Diptera, Psychodidae, Psychodinae) dans les environs du Mitaraka en Guyane.
Peu d’informations sont disponibles concernant les Psychodidae Newman, 1834 de Guyane autres que les phlébotomes. Des spécimens de psychodides ont été triés à partir d’échantillons prélevés lors de la récente expédition « La Planète revisitée » Guyane-2015 dans la chaîne de Mitaraka. Les mâles et les femelles de deux genres de la tribu Maruinini Enderlein, 1937, Polletomyia subulata n. gen., n. sp. et Myiomystax trilineata n. gen., n. sp., sont décrits et illustrés. Les caractères pertinents pour évaluer les relations entre les genres de Maruinini sont discutés.

KEY WORDS
Neotropical Region, biodiversity, new genus, new species.

MOTS CLÉS
Région néotropicale, biodiversité, genres nouveaux, espèces nouvelles.

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INTRODUCTION

Psychodidae Newman, 1834 (moth flies and sand flies), currently with six recognized subfamilies, are cosmopolitan in distribution, occupying a broad range of niches (Curler & Moulton 2012). In the Neotropical region, subfamily Phlebotominae Rondani, 1840 is best known due to their significance as vectors of disease agents such as Leishmania Ross, 1903 parasites (Young & Duncan 1994; Galati 2019). The remaining subfamilies have received relatively little attention. In particular, a majority of the non-phlebotomine psychodid fauna of French Guiana remains unknown. Indeed, Psychodinae Newman, 1834 is the only other subfamily reported from the country; Quate & Brown (2004) provided details of these records. In summary, a total of four genera and 12 species of tribe Maruinini Enderlein, 1937 are known to occur in French Guiana: Alepia Enderlein, 1937 is represented by four species (A. piscicatuda Quate & Brown, 2004; A. diocula Quate & Brown, 2004; A. fruticos Quate & Brown, 2004; and A. bisululata Duckhouse, 1968); Balbagathis Quate, 1996 by one species (B. agrestis Quate & Brown, 2004); Tonnoina Enderlein, 1937 by three species (T. sicilis Quate & Brown, 2004; T. didyma Quate & Brown, 2004; T. cavernicola Quate & Brown, 2004); and Caenobrunettia Wagner, 1981 by four species (C. subdititsia Quate & Brown, 2004; C. laelvia Quate, 1996; C. echiocflagellata Wagner, 1981; and C. fraudulenta Quate & Brown, 2004). Recently, an expedition in the Mitaraka range of French Guiana Amazonian Park resulted in the collection of nearly 400 specimens of Psychodidae, including two undescribed genera of Maruinini. These two genera are described and illustrated here.

MATERIAL AND METHODS

STUDY AREA

All Psychodidae examined as part of this study were collected during the “Our Planet Reviewed” Guyane-2015 expedition in the Mitaraka range, in the core area of the French Guiana Amazonian Park, organized by the MNHN and Pro-Natura international (Touroult et al. 2018).

COLLECTION AND OBSERVATION

Most specimens were collected by pan traps (yellow, white or blue pans) while few were also sampled in a Malaise trap or randomly using a sweep net. The biological sampling benefited from the access and benefit sharing agreement “APA973-1”, which is reported in each sample label. All specimens were preserved in 70% or 95% ethanol, subsequently macerated in cold 8% potassium hydroxide (KOH), dehydrated in ethanol, transferred to xylene and finally mounted in Canada balsam. Observations and measurements were completed using an Olympus BX50 compound microscope equipped with differential interference contrast, and a Wild M5 stereomicroscope fitted with an ocular micrometer. Drawings were rendered with the aid of a drawing tube on the Olympus system. Photomicrographs were captured using a Nikon DS-Fi1 camera mounted on a Nikon Eclipse E800 compound microscope. Focus stacks were created using Zerene Stacker (Build T-2018-07-19-1515) and images were optimized for color and clarity using Adobe Lightroom® CC and Adobe Photoshop® CS6.

MEASUREMENTS

Head width was measured at the point of greatest width of the eyes. Adult head height was measured from the vertex to the anterior margin of the clypeus. Approximate wing length and width were measured at the points of greatest length and width of the dissected wing. Sample sizes are provided before each description with measurements in millimeters presented as a mean followed by a range in parentheses.

TERMINOLOGY

General terminology follows Cumming & Wood (2017), except interpretation of wing venation, which is that of Krzemieniecki & Krzemieńska (2003). Most Psychodidae have hyaline sensilla, variable in shape, inserted on some or all their antennal flagellomeres; these are termed “ascoids” (Kwifte & Wagner 2017). Specific characters of the male terminalia (e.g. gonocoxal apodemes) are described here according to the interpretation of Duckhouse (1990) as followed by Quate & Brown (2004). The elongate lobes articulated posteriorly with the epandrium in Psychodinae are referred to here as “epandrial claspers” sensu Dos Santos & Curler (2014). Both “lateral arm of subependrial sclerite” and “dorsal paramere” are terms that have not been applied previously for Psychodidae, but are used here provisionally to describe specific parts of the male genitalia.

TAXONOMY

Specimens examined during this study were identified or determined as new to science based on keys and generic diagnoses provided by Quate & Brown (2004). Nonetheless, the tribal classification used in that work (i.e., Setomimini Vaillant, 1982) is not followed here, as Setomimini is a junior synonym of Maruinini. Duckhouse (1990) provided a diagnosis for Maruinini.

ABBREVIATIONS

Male genitalia

aed  aedeagus;
dpr  dorsal paramere;
ecl  epandrial clasper;
eja  ejaculatory apodeme;
gca  gonocoxal apodeme;
gcx  gonostyle;
gst  gonostyle;
hdp  hypandrium;
lar  lateral arm of subependrial sclerite;
prm  paramere;
ses  subependrial sclerite;
ten  tenacula.

Female genitalia

st8  abdominal sternum 8;
oviduct.

General morphology

asc  ascoid;
New genera of Psychodidae from Mitaraka

Genus Polletomyia n. gen.

TYPE SPECIES. — Polletomyia subulata n. gen., n. sp.

ILLUSTRATIONS. — Head: Fig. 1A; Wing: Fig. 1B; Male genitalia: Fig. 1E-G.

ETYMOLOGY. — This genus is named in honor of Dr Marc Pollet, who collected the type specimens, forwarded them to the author and provided supporting information during the course of this study.

DIAGNOSIS. — The following combination of characters is diagnostic for the genus: male head: antennal flagellomeres fusiform, with paired, unbranched ascoids on f4 f7 (Fig. 1C), single ascoids on f8 f1; male terminalia: bilaterally symmetrical; aedeagus comprised of four movable, blade-like sclerites surrounded by membranous sheath; outer aedeagal sclerites sinuous, spreading laterally when extended posteriorly (Fig. 1F); gonostyli with stout, conical setae inserted apically (Fig. 1E-G); epandrial claspers each with single, conical, spiniform seta inserted apically; tenacula setiform.

DISTRIBUTION. — Polletomyia n. gen. is currently known only from the Mitaraka range of French Guiana.

HABITAT. — Little information regarding specific habitat is available; however, all specimens were collected in lowland rainforest.

REMARKS
The relatively small size of Polletomyia n. gen., as well as their lanceolate wings and dark coloration make it possible to differentiate them in a sample comprised of many psychodid genera and species. These superficial characters also make it possible to associate males and females, as the latter are otherwise lacking in characters that readily differentiate them from other psychodine females. Indeed, if further species of Polletomyia n. gen. are discovered, it may become difficult or impossible to differentiate females within the genus. Despite the relative ease of differentiating them in a bulk sample, accurate diagnosis of this genus is dependent on examination of slide-mounted male terminalia.

RESULTS
Most of the approximately 400 specimens collected from Mitaraka represent undescribed taxa. Nonetheless, a majority of this material was not included in the present report, but will be treated in subsequent papers. Adequate series of males and females of two undescribed genera of Maruinini were among the collected material, allowing for the following descriptions.

Subfamily PSYCHODINAE Newman, 1834
Tribe MARUINI Enderlein, 1937

Genus Polletomyia n. gen.

CoWin venation
CuA anterior cubital vein;
M medial vein;
R radial vein;
Sc subcostal vein.

Repository
MNHN Muséum national d’Histoire naturelle, Paris;
LACM Natural History Museum of Los Angeles County, Los Angeles, CA.

Collecting methods
BPT blue pan trap;
YPT yellow pan trap;
WPT white pan trap;
SLAM square Malaise trap;
SW sweep net.

DESCRIPTION
See below, Polletomyia subulata n. gen., n. sp.

Polletomyia subulata n. gen., n. sp.
(Figs 1; 2; 5A)

TYPE LOCALITY. — Guyane, Mitaraka.

TYPE MATERIAL. — Holotype. French Guiana • ♂; Mitaraka, site MIT-C-RBF2; 02°14’03.4”N, 54°26’53.0”W; 29 m a.s.l.; lowland rainforest; 27.II-6.III.2015; Marc Pollet leg. (sample code: MITARAKA/125), YPT; specimen dissected, mounted on slide; MNHN.
Fig. 1. — Polletomyia subulata n. gen., n. sp.: A, male head, frontal view; B, wing; C, male antennal flagellomeres 3–4, dorsal view; D, male antennal flagellomeres 12–14, dorsal view; E, male terminalia, epandrium removed, dorsal view; F, male terminalia, epandrium removed, aedeagal structure expanded, dorsal view; G, male terminalia, epandrium removed, aedeagal structure partly expanded, ventral view. Abbreviations: see Material and methods. Scale bars: A, 0.25 mm; B, 0.5 mm; C–G, 0.1 mm.
Fig. 2. — Polletomyia subulata n. gen., n. sp.: A, male terminalia, lateral view; B, male epandrium, proctiger and epandrial claspers, dorsal view; C, female terminalia, external structure, ventral view; D, female terminalia, internal structure, ventral view. Abbreviations: see Material and methods. Scale bars: 0.1 mm.
Paratypes. French Guiana • 8 ♂, 2 ♀; same data as holotype; YPT; MNHN (4 ♂, 1 ♀), LACM (4 ♂ LACM 340591-340594, 1 ♀ LACM 340595) • 2 ♂, 1 ♀; site MIT-C-RBF2; 02°14’03.4”N, 54°26’53.0”W; 29 m a.s.l.; lowland rainforest; 27.II-5.III.2015; Marc Pollet leg. (sample code: MITARAKA/126); BPT; LACM (♂ LACM 340600-34061; ♀ LACM 340602) • 3 ♂, 2 ♀; site MIT-C-RBF1; 02°14’10.8”N, 54°26’49.5”W; 25 m a.s.l.; lowland rainforest; 27.II-8.III.2015; Marc Pollet leg. (sample code: MITARAKA/133); YPT; MNHN (2 ♂, 1 ♀), LACM (1 ♂ LACM 340596, 1 ♀ LACM 340597) • 2 ♂; site MIT-C-RBF1; 02°14’10.8”N, 54°26’49.5”W; 25 m a.s.l.; lowland rainforest; 27.II-8.III.2015; Marc Pollet leg. (sample code: MITARAKA/134); WPT; MNHN • 1 ♀; site MIT-C-RBF1; 02°14’10.8”N, 54°26’49.5”W; 25 m a.s.l.; lowland rainforest; 27.II-8.III.2015; Marc Pollet leg. (sample code: MITARAKA/135); BPT; MNHN • 3 ♂; site MIT-C-RBF2; 02°14’03.4”N, 54°26’53.0”W; 29 m a.s.l.; lowland rainforest; 5-10.III.2015; Marc Pollet leg. (sample code: MITARAKA/137); BPT; MNHN (2 ♂), LACM (1 ♂ LACM 340598) • 1 ♂; site MIT-A-RBF2; 02°14’12.5”N, 54°27’08.1”W; 28 m a.s.l.; lowland rainforest; 4-10.III.2015; Marc Pollet leg. (sample code: MITARAKA/149); BPT; LACM 340599.

ETYMOLOGY. — The specific epithet is derived form the Latin word "subulatus", meaning awl-shaped, in reference to the prominent, spiny apex of each gonostylius.

DIAGNOSIS. — See above, Polletomyia n. gen.

DISTRIBUTION. — Currently known from multiple localities in the Mitaraka range of French Guiana.

HABITAT. — Little information regarding specific habitat is available; however, all specimens were collected in lowland rainforest.

DESCRIPTION

Male
Measurements, (N = 5) head width 0.37 mm (0.34-0.40), head height 0.33 mm (0.30-0.34), wing length 1.70 mm (1.64-1.76), wing width 0.44 mm. Head with vertex extended slightly posteriorly, eyebridge comprised of 3 facet rows, contiguous but slightly constricted at median. Antennae: scape cylindrical, about 1.5 times longer than wide; pedicel globular; flagellomeres fusiform, with paired setae on f4 f7, single setae on f8 f11. Frons and clypeus distinctly divided; frontal scar patch bilobed posteriorly, contiguous mediad. Mouthparts inconspicuous, obscured by clypeus, extending slightly beyond apex of palp segment 1; palp typical of Psychodinae, with apical segment annulated; palp segment proportions: 1, 2, 2.2, 18.18; labellum about as wide as anterior margin of clypeus. Wing: strongly lanceolate; radial and medial fork positioned at a level basal to the apex of CuA; R3 terminating in C, posterior to wing apex; M3 incomplete at base; CuA conspicuously widened at base. Male terminalia: bilaterally symmetrical; hypandrium flat, band-like, arched posteriorly, fused with gonocoxites laterally; hypandrium in some specimens appearing folded along medial side (Fig. 1E); intersegmental membrane connecting between anterior margin of hypandrium and posterior margin of abdominal segment 8, covering base of ejaculatory apodeme; ejaculatory apodeme dorsoventrally compressed at base, dome-like at center, articulating posteriorly with aedeagal sclerites; aedeagus comprised of four blade-like sclerites (Fig. 1E-G) surrounded by a tubular membrane; innermost aedeagal sclerites pithyform, expanding dorsally in extended aedeagus; outer sclerites sinuous, longer than inner sclerites, expanding lateroventrally in extended aedeagus (Fig. 1F); parameres dorsoventrally compressed, flat, with pointed, hook-like projection posteromedially; gonostyli tapered gradually from base to apex, curved ventrally along apical half, appearing pitted ventrally; gonocoxites reniform, about two times as long as wide, setose, with setae alveoli laterally and dorsally; gonocoxal apodemes inflated, triangular in dorsoventral view, bulbous in lateral view, with dorsal paramere extending from posterior margin; subepandrial sclerite narrow, band-like, with lateral arms claw-shaped in dorsoventral view, linked to base of epandrial clasps; epandrium rectangular, narrowing posteriorly; epandrial clasps elongate, tapered, curved dorsally from base to apex, with single, setiform tenaculum inserted dorsoapically.

Female
Measurements, (N = 3) head width 0.34 mm (0.32-0.36), head height 0.33 mm (0.32-0.34), wing length 1.84 mm (1.76-1.88), wing width 0.48 mm (0.44-0.52). Head and wing generally as in male; palp segment proportions: 1, 2.50, 3, 3. Terminalia: cerci elongate, heavily sclerotized, spatulate in lateral view, with numerous setae inserted on medial surface; posterior margin of sternum 8 bilobed, oviduct complex, ornate internally (Fig. 2D).

Genus Myiomystax n. gen.

TYPE SPECIES. — Myiomystax trilineata n. gen., n. sp.

ILLUSTRATIONS. — Wing: Fig. 3A; Head: Fig. 3B; Male genitalia: Fig. 3C-D.

ETYMOLOGY. — This genus name is derived from the Latin “myia”, meaning fly and “mystax”, meaning mustache, in reference to the appearance of the fly having a prominent mustache.

DIAGNOSIS. — Male head: antennal flagellomeres 1-5 elongate, cylindrical; all flagellomeres lacking setae; clypeus elongate, expanded laterally, with lateral margins overlapping medial margins of eyes; palp conspicuously elongate, with segment 1 hook-like, segment 4 inflated apically, capitale; palp segments with clusters of spinulate setae inserted dorsally and ventrally; palps folded behind head at rest. Male terminalia: bilaterally symmetrical; gonocoxites glabrous, without setae alveoli; aedeagal sclerites paired, laterally compressed, spatulate. Female head: maxillary palp not elongated. Female terminalia: posterior margin of sternum 8 trilobed, with median lobe wider than flanking lobes; oviduct with microtrichia internally on membrane.

DISTRIBUTION. — Myiomystax n. gen. is currently known only from the Mitaraka range of French Guiana.

HABITAT. — Little information regarding specific habitat is available; however, all specimens were collected in lowland rainforest.
Fig. 3. — Myiomystax trilineata n. gen., n. sp.: A, wing; B, male head, frontal view; C, male epandrium, proctiger and epandrial claspers, dorsal view; D, male terminalia, epandrium removed, dorsal view. Abbreviations: see Material and methods. Scale bars: A, 0.5 mm; B, 0.25 mm; C-D, 0.1 mm.
Fig. 4. — *Myiomystax trilineata* n. gen., n. sp.: A, male terminalia, lateral view; B, female mouthparts and maxillary palpi, frontal view; C, female terminalia, external structure, ventral view; D, female terminalia, internal structure and egg, ventral view. Abbreviations: see Material and methods. Scale bars: 0.1 mm.
**Remarks**

In contrast to *Polletomyia* n. gen., the relatively large size and light coloration of *Myiomystax* n. gen. make it possible to easily differentiate them in samples containing many psychodid genera. Moreover, while both genera have lanceolate wings, the latter is more distinctive due to several veins being darkly pigmented (Fig. 5). The unique wing of *Myiomystax* n. gen. also makes it easy to associate males and females of this genus. It remains to be determined whether these characteristics of the wing will be present in the wings of congeners, if any are discovered. Although less important compared to those of *Polletomyia* n. gen., accurate diagnosis of this genus is still dependent on examination of slide-mounted male terminalia.

*Myiomystax* n. gen. males have a bilaterally symmetrical terminalia, with an aedeagus that extends as an open-close mechanism, much like *Polletomyia* n. gen. and *Maruina*. However, they differ from the other genera by having paired aedeagal sclerites as opposed to two pairs (as in *Polletomyia* n. gen.) or multiple aedeagal spines (as in *Maruina*). *Myiomystax* n. gen. males are also unique among these genera in that their gonocoxites lack setae.

**Description**

See below, *Myiomystax trilineata* n. gen., n. sp.

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**Myiomystax trilineata** n. gen., n. sp.

(Figs 3; 4; 5B)

**Type locality.** — Guyane, Mitaraka.

**Type material.** — Holotype, French Guiana • ♀; Mitaraka, site MIT-C-RBF2; 02°14’03.4”N, 54°26’53.0”W; 29 m a.s.l.; lowland rainforest; 27.II-6.III.2015; YPT; Marc Pollet leg. (sample code: MITARAKA/125); specimen dissected, mounted on slide; MNHN.

Paratypes, French Guiana • 1 ♀; site MIT-C-RBF2; 02°14’03.4”N, 54°26’53.0”W; 29 m a.s.l.; lowland rainforest; 27.II.2015; Marc Pollet leg. (sample code: MITARAKA/018); SW; LACM 340595 • 4 ♂, 2 ♀; same data as holotype; YPT; MNHN (2 ♂, 1 ♀), LACM (2 ♂ LACM 340633-340604, 1 ♀ LACM 340605) • 1 ♂, 2 ♀; site MIT-C-RBF1; 02’14’10.8”N, 54°26’49.5”W; 25 m a.s.l.; lowland rainforest; 27.II-8.III.2015; Marc Pollet leg. (sample code: MITARAKA/133); YPT; MNHN (♂, 1 ♀), LACM (1 ♀ LACM 340606) • 9 ♂, 5 ♀; site MIT-C-RBF2; 02°14’03.4”N, 54°26’53.0”W; 29 m a.s.l.; lowland rainforest; 6-10.III.2015; Marc Pollet leg. (sample code: MITARAKA/136); YPT; MNHN (4 ♂, 3 ♀), LACM (5 ♂ LACM 340607-340611, 2 ♀ LACM 340612-340613) • 1 ♂; site MIT-C-RBF2; 02°14’03.4”N, 54°26’53.0”W; 29 m a.s.l.; tropical wet forest (bas fond); 27.II-2.III.2015; Marc Pollet leg. (sample code: MITARAKA/184); SLAM; MNHN • 1 ♀; site different sites near base camp and along trails; 14.III.2015; Julien Touroult & Eddy Poirier leg. (sample code: MITARAKA/191); SLAM; MNHN.

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**Fig. 5.** Photomicrographs of male wings: A, *Polletomyia subulata* n. gen., n. sp.; B, *Myiomystax trilineata* n. gen., n. sp. Scale bars: 0.5 mm


**ETYMOLOGY.** — The specific epithet is derived form the Latin word “linea”, meaning line, in reference to the three darkly pigmented wing veins present in this species.

**DIAGNOSIS.** — See above, Myiomystax n. gen.

**DISTRIBUTION.** — Currently known from multiple localities in the Mitaraka range of French Guiana.

**HABITAT.** — Little information regarding specific habitat is available; however, all specimens were collected in lowland rainforest.

**DESCRIPTION**

**Male**

Measurements, (N = 4) head width 0.41 mm (0.38-0.44), head height 0.47 mm (0.44-0.48), wing length 2.17 mm (2.08-2.24), wing width 0.54 mm (0.52-0.56). Head with vertex extended slightly posteriorly, eyebridge comprised of 3 facet rows, contiguous but strongly constricted at median. Frons and clypeus distinctly divided; frontal scar patch bilobed posteriorly, contiguous medially; clypeus elongated laterally, with lateral margins overlapping medial margins of eyes. Antennae: scape cylindrical, about 2.5 times longer than wide; pedicel globular; flagellomeres 1-5 elongate, cylindrical; flagellomeres 6-11 fusiform; flagellomeres 12-14 reduced, beak-like; flagellomere 14 with apical process digitiform, setose, approximately as long as node; all flagellomeres lacking ascoids. Mouthparts extending slightly beyond base of palp segment 1; labellum about as wide as anterior margin of clypeus; palpi conspicuously elongate, with segment 1 hook-like; segment 2 clavate, inflating from base to apex; segment 3 elongate, inflated at midlength; segment 4 inflated apically, capitulate; palp segments with clusters of spatulate setae inserted dorsally and ventrally; palps folded behind head at rest. Wing: strongly lanceolate; radial fork positioned slightly distal to apex of CuA; medial fork positioned at the level of apex of CuA; R3 terminating in C posterior to wing apex; wing veins R1, R3, and M4 pigmented brown, appearing more heavily sclerotized than other wing veins (Fig. 5B); costal vein with light pigment between apices of R1 and R3, M1 and M4; pigmented veins contrasting against other lightly colored wing veins. Male terminalia: bilaterally symmetrical; hypandrium flat, band-like, arched posteriorly, fused with gonocoxites laterally; intersegmental membrane connecting between anterior margin of hypandrium and posterior margin of abdominal segment 8, covering base of ejaculatory apodeme; ejaculatory apodeme dorsoventrally compressed anterolaterally, dome-like at center, articulating ventrally with gonocoxal apodemes, posteriorly with aedeagal sclerites; aedeagus comprised of paired, laterally compressed sclerites, spatulate in lateral view; spatulate sclerites enveloped by tubular membrane; parameres not differentiated from aedeagus; gonostyli tapered, sinuous from base to apex (Fig. 3D), digitiform apically, setose, with paired, elongate setae inserted mediadly near apex (Figs 3D; 4A); gonocoxites glabrous, without setae alveoli (Figs 3D; 4A), cylindrical, about two times as long as wide; gonocoxal apodemes inflated, rounded in dorsoventral view, bulbous in lateral view, linked to subependial sclerite anteromedially; subependial sclerite narrow, divided medially, with puckered seam at median, with posterolateral arms ribbon-like, linked to base of epandrial claspers; epandrium rectangular along posterior half in dorsoventral view, anterior half narrowing abruptly; epandrial claspers elongate, tapered, curved dorsally from base to apex, with single, spatulate tenaculum inserted dorsoapically.

**Female**

Measurements, (N = 4) head width 0.44 mm (0.42-0.46), head height 0.41 mm (0.40-0.42), wing length 2.16 mm (2.04-2.36), wing width 0.53 mm (0.52-0.56). Head mostly as in male. Antennae with flagellomeres 1 and 2 elongate, cylindrical, shorter than in male; apical flagellomeres missing in specimens observed. Mouthparts as in male; maxillary palpi more typical of Psychodinae, segments not elongated; palp segment proportions: 1, 1.30, 1, 2.30. Wing venation as in male. Terminalia: cerci conical in ventral view, with ventral surface covered by setulae, numerous setae inserted laterally and ventrally; posterior margin of sternum 8 trifoliated, with median lobe wider than flanking lobes; oviduct with microtrichia internally on membrane.

**DISCUSSION**

Males of Maruinini exhibit geometry in their genitalia that is recognizable among most of the tribe's constituent genera. In addition to their unique general shape, specific structures such as the ejaculatory apodeme and the gonocoxal apodemes are interlinked in a way that distinguishes them from other Psychodinae. Genitalia morphology most similar to that described and illustrated above can also be found in genera such as Australopericoma Vaillant, 1971 (e.g. Bravo 2007: fig. 9), Alepia (e.g. Bravo 2008: figs 8-9), Tonnoira (e.g. Bravo et al. 2008: figs 16-18) and Maruina (e.g. Camico et al. 2019: figs 43, 44). In particular, the “ball and socket” construction, as discussed by Duckhouse (1990), can be identified; however, to see these structures clearly, it is necessary to examine both dissected and whole terminalia in dorsoventral and lateral view.

Another interesting character state in both genera described here is the bilateral symmetry of the male terminalia. In most genera of Maruinini, at least the aedeagus and parameres are strikingly asymmetrical; in some species, the gonopods are asymmetrical as well. Exceptions are as mentioned above. Prior to the description of Polletomyia n. gen. and Myiomystax n. gen., Maruina was the only genus of Maruinini of which all described species are known to have symmetrical genitalia. Furthermore, Maruina is the only other genus of Maruiniini in which an open and close mechanism involving aedeagal and parameral sclerites has been reported (Vaillant 1963: fig. 2; Camico et al. 2019: figs 43-44).

Specimens of Polletomyia n. gen. and Myiomystax n. gen. collected during this study died with parts of their terminalia fixed in different positions. As shown previously for Maruina, it is evident that the aedeagal and parameral complex has developed into an evertable, or extendable (i.e., open and close) mechanism. Unfortunately, it is unclear how this mechanism works. The movable (i.e., extendable and expandable) nature of the aedeagus in P. subulata n. gen., n. sp. appears to be
dependent on the anterior-posterior pivoting movement of the ejaculatory apodeme. As in some other Maruiniini, the parameral sclerites appear to serve as a more stationary framework for the opening and closing aedeagus. This indicates that the parameres are serving to stabilize the terminalia in one position while the flies are in copula. Accurate descriptions of these structures depend on being able to differentiate the constituent sclerites. Unfortunately, in many groups of Psychodidae, fusions between the aedeagal and parameral sclerites have apparently occurred, making it difficult or impossible to tease apart these structures. Regardless, these are genus or species-level characters and thus, are of lesser importance for understanding relationships at a higher level. Despite the challenges posed by some characters, others (e.g. ejaculatory apodeme; gonocoxal apodemes) are less ambiguous and can be traced across genera. When ample material is available, Maruiniini specimens should be prepared using various methods, such as clearing with cedarwood oil. This method eliminates fat body while leaving musculature intact, which will be necessary to fully understand the structure of the ejaculatory apodeme and how it interacts with the gonocoxal apodemes. This, in turn, could elucidate the process by which the aedeagal and parameral sclerites extend and retract.

Information on Neotropical Maruiniini (= Setomimini) was synthesized in part by Quate & Brown (2004), and many species of this group have since been described. Despite this, no phylogenetic revision of the group has been attempted. Many characters of the male and female genitalia are rich in information, but there is a need for clarification of homologies and identification of synapomorphies at higher levels rather than focusing only on autapomorphies found in genera and species.

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