Bioelectrical Impedance Analysis for Monitoring Fluid and Body Cell Mass Changes in Patients Undergoing Cardiopulmonary Bypass

Mustafa Göz¹, MD; Cemil Sert², MD; Abdussamet Hazar³, MD; Mehmet Salih Aydin¹, MD; Nazım Kankılıç¹, MD

Abstract

Objective: To evaluate preoperative and postoperative body fluid distribution with a bioelectrical impedance analyzer in patients undergoing cardiopulmonary bypass.

Methods: Fifteen adult patients undergoing cardiopulmonary bypass were included in this study. Total body fluid changes, basal metabolism rates, body fat masses, lean body masses, and total cell masses were recorded. The patients' values were measured before anesthesia, after anesthesia, after sternotomy, at the 5th, 30th, and 60th minutes of cardiopulmonary bypass, and on the 1st, 3rd, and 5th postoperative days. All values were compared with preoperative values.

Results: Total body fluid changed significantly after cardiopulmonary bypass ($P<0.01$). Metabolic velocity significantly changed compared to preoperative measurements ($P<0.05$). Fat mass and lean body mass also changed significantly. Body mass index and phase angle did not change significantly ($P>0.05$).

Conclusion: Changes in body fluids during and after cardiopulmonary bypass are inevitable. The increase in total body weight shows that this fluid load shifts to the extracellular space during bypass and the fluid load in this area passes into the intravascular area in the early postoperative period. This may cause edema and dysfunction in the major organs. Therefore, the fluid balance should be adjusted very carefully, especially during the bypass phase and the early postoperative period.

Keywords: Bioelectrical Impedance. Fat Mass. Lean Body Mass. Total Body Water. Cardiopulmonary Bypass.

INTRODUCTION

Cardiac surgery with cardiopulmonary bypass leads to increase in total body fluid due to the prime solution and intravenous fluids. Additionally, interaction of extracorporeal circulation and prime volume with the body fluids causes a systematic inflammation[1]. Thus, the water retention in vital areas appears to cause notable clinical symptoms[1]. Therefore, perioperative fluid flow observation is closely related to prediction of morbidity.

Bioelectrical impedance analysis (BIA) is a non-invasive method to determine body fluid distribution[2]. This technique involves releasing a very little current in the tissues[2]. In a clinic, a single frequency of 50 kHz is used to make BIA measurements. These high-frequency electrical currents pass through both intra and extracellular fluids to calculate total body water (TBW). However, currents < 5 kHz of frequency proceed through the extracellular space and provide clinically useful information regarding circulation changes. Despite the use of BIA in medical...
Cardiac surgery with cardiopulmonary bypass is interrelated with a high load of fluids that causes an increase in total body fluids due to the pump solution and intravenous fluids\(^3\). Thus, it causes reduction of both hemoglobin and hemodilution\(^3\). Following the cardiac surgery, patients are usually given diuretic preparation to lower the TBW\(^3\). These are clinically kept track of.

In this study, body fluid changes, basic metabolic rate, body cell mass, fat-free body mass, and fat mass (FM) changes are studied before, during, and after the cardiac surgery by using BIA.

### METHODS

#### Choosing Patients

After approval by the Clinic Committee of Ethics, 15 patients who had undergone adult open cardiovascular surgery were included in the study. Patients with systemic inflammatory disease, infection, recurrent cardiac surgery, overweight (body mass index [BMI] > 30), underweight (BMI < 18.5), and very low ejection fraction (≤ 30%) were excluded from the study.

#### Anesthesia and Cardiopulmonary Bypass Technique

In order to help to control the high blood pressure and tachycardia before the surgery, 0.05-0.1 mg/kg of intravenous midazolam was applied. In induction, cases were endotracheally intubated after gaining enough amount of muscular relaxation with 3-10 mg/kg of fentanyl, 0.2-0.3 mg/kg of etomidate, and 0.6 mg/kg of rocuronium. Endotracheal intubation and anesthesia were performed in supine position. Then, central venous catheterization and left radial artery monitoring were performed. Nasopharyngeal and rectal heat probes were applied. During anesthesia, 5-channel electrocardiogram and blood oxygen saturation and end-tidal carbon dioxide monitoring were performed.

Dideco Compac Flo (Sorin Group, Italy) membrane oxygenator and Sasan Containerless Tubing Set (Sasan Medical Supplies Inc., Ankara, Turkey) were used in cardiopulmonary bypass. As the prime solution, 1200 cc of Lactated Ringer, 100 cc of 20% Mannitol, 20 cc of NaHCO3, 1 cc of heparin, and 1 g of cefazolin were used. All the cases were kept in mild hypothermia (26-32°C). Perfusion pressure was kept control of (60-70 mmHg). Full cardiac flow was maintained in 2.4 l/m². Throughout the operation, activated clotting time was kept > 480 sec. Blood gas analyses were checked with alphasat mechanism. Until all the cases’ rectal heat levels reached 37.5°C, they were heated by keeping a 10°C gradient between water and blood heat. Immediately after reaching adequate cardiac performance, the operation was terminated by exiting the cardiopulmonary bypass.

#### Tracking the Body Fluids and BIA Measurements

A Biodynamic P450 Bioelectrical Impedance Analyzer (Biodynamics Corporation, Washington, USA) was utilized before, during, and five days after the cardiopulmonary bypass in order to study and evaluate the fluid change, basic metabolic pace, body FM, and total cell mass. All patients’ hands and feet were cleared for measurements. Two electrodes were connected to the right hand and right foot. Stimulating electrodes were attached to the dorsal sides of the second and third metacarpal and metatarsal joints. The recording electrodes were connected to the dorsal sides of the hands and feet. The patients received a current of 500uA. The current was too small to be felt by the patients. Measurements were made before, during, and after anesthesia, after sternotomy, at the 5th, 30th, and 60th minutes of cardiopulmonary bypass, and on the 1st, 3rd, and 5th postoperative days. Daily fluid balance was monitored in all cases until the preoperative period and the postoperative 5th day.

#### Statistical Analysis

The Statistical Package for the Social Sciences software (SPSS, Chicago, Illinois, USA), version 11.5, was used for statistical analysis. The measurements were compared with preoperative values by using Wilcoxon test and statistical analyses were performed. \(P\)-values < 0.05 were considered significant.

#### RESULTS

The patients’ general characteristics are shown in Table 1. TBW showed statistically significant changes in the post-bypass stage \((P<0.01)\) (Figure 1). However, the amount of extracellular and intracellular fluids did not change significantly. The amount of extracellular fluid increased 60 minutes after bypass and one day after bypass, and then returned to normal (Figure 1). The level of intracellular fluid also increased on the 1st and 3rd days after bypass and returned to normal on the 5th postoperative day (Figure 1).

### Table 1. Patients’ general characteristics.

| Age (years) | 58±4.59 |
| Males (n [%]) | 9 (%60) |
| Females (n [%]) | 6 (%40) |
| Ejection fraction (%) | 46.26±7.48 |
| CCT (min) | 64.7±31.1 |
| CPBT (min) | 108.3±24.6 |
| ICU stay (days) | 3.21±1.01 |
| Hospital stay (days) | 8.4±4.21 |
| CABG (n [%]) | 8 (%65.3) |
| MVR (n [%]) | 3 (%20) |
| AVR (n [%]) | 4 (%26.6) |

AVR=aortic valve replacement; CABG=coronary artery bypass grafting; CCT=cross-clamp time; CPBT=cardiopulmonary bypass time; ICU=intensive care unit; MVR=mitral valve replacement
Basal metabolic rate (BMR) significantly changed compared to preoperative measurements ($P<0.05$). This change was bigger after the 60-minute post-bypass measurements ($P<0.01$) (Figure 1). Fat mass (FM) changed significantly ($P<0.05$) (Figure 2). Lean body mass (LBM) also changed significantly ($P<0.05$). This change started after the 5th minute post-bypass and returned to normal on the 1st postoperative day (Figure 2). Total cell mass changed significantly. This change occurred after the 5th minute post-bypass measurements (Figure 2). BMI and phase angle did not change significantly ($P>0.05$) (Figure 2).

**DISCUSSION**

It is observed that total body fluid increases significantly after cardiac surgery. However, it is an important problem when this increase of fluid starts exactly after the operation and when it returns to normal. It is also important how this is reflected in the extracellular and intracellular fluid changes and how BMR and body composition values change after cardiac surgery. Studies on this subject are very limited. Since we could not find a detailed study, we recorded the measurements up to the 5th day and our study was made detailed. Significant correlations between various body composition methods, such as isotope dilution, total-body potassium count, dual radiographic absorptiometry, or densitometry, have been shown. BIA is a body composition measurement method. It is used in many ways and in many disease groups. BIA can be easily performed to determine fat-free mass, fluid deposition, peritoneal or hemodialysis fluid exchange, perioperative fluid deposition, and body weight in weak, obese, and sick people.

David Bracco et al. measured body FM and body fluid mass before and after the operation in 22 patients undergoing cardiopulmonary bypass. They observed an increase of 8-10% in fat-free mass. They also observed an increase in fluid mass. The first mechanism is that it triggered edema formation due to surgical translocation and tissue trauma. The second mechanism is...
is macro and microcirculation changes caused by intestinal permeability changes\cite{20}. Fiogbé E et al.\cite{21} investigated the effect of water aerobic exercise training on heart rate autonomic modulation and body composition in the rehabilitation of patients with coronary artery disease. In BIA evaluation, it was observed that body compositions did not change with disease or rehabilitation. Contrary to that study, this change is seen in patients who underwent cardiovascular surgery in our study.

It is important to measure the amount of intracellular and extracellular fluids separately in BIA, because intracellular edema may be an indicator of organ dysfunction and may lead to organ failure\cite{22,23}. In recent studies, it was emphasized that BIA may be useful in determining the prognosis and treatment of patients with heart failure\cite{23-25}. It is not possible to detect this by methods other than BIA. In our study, intracellular fluid showed no significant change. However, it was different during the follow-up period. Intracellular fluid returned to baseline at the 5th postoperative day. The amount of extracellular fluid did not change significantly, but it slightly increased. Compared to the preoperative measurements, it showed a significant increase in the measurement after sternotomy. It returned to the normal level in the last measurement. This reduction should be seen as a result of the accumulation of total fluid, mainly in the non-fat body mass.

We searched the BMR parameter in other cardiac surgery studies, but it was not found in any of them. In this study, BMR values showed a significant increase compared to preoperative values. This increase in BMR that begins after anesthesia and continues until the end of the operation may seem contradictory with the desired effects of the hypothermia applied, but it shows that the trauma caused by cardiopulmonary bypass cannot be completely prevented despite the retarding effect of hypothermia. It is important in terms of the fact that the patient will need more energy for cell renewal and circulation in the surgical field. This is important for the patient’s diet. Phase angle reflects whether the permeability of the cell membrane has changed. This parameter did not change in our study and this shows that there was no change in the cell membrane level. The fact that the total cell mass increased significantly five minutes after the bypass was due to the acceleration of cell production from the bone marrow in response to the reduced body resistance in this stage.

---

**Fig. 2** – Changes in fat mass, lean body mass, body cell mass, and phase angle levels after cardiopulmonary bypass.
LBM values showed a significant increase in the postoperative period. This increase in LBM values may be the result of an increase in total body fluid and a decrease in FM. The increase in total body mass is due to the increase in total body fluid. We believe there are two reasons for this increase in total body mass. The first is the increase in extracellular fluid during cardiopulmonary bypass. The second is due to the passage of this fluid to the intracellular region after cardiopulmonary bypass and in the early postoperative period.

CONCLUSION

This study clearly demonstrates that the exchange of body fluids during and after cardiopulmonary bypass is inevitable. In general, the increase in TBW causes an increase in fluid volume in the extracellular compartment during cardiopulmonary bypass. However, in the early postoperative period this increase is shifting to the intracellular region. This may be seen as organ dysfunction due to edema in the organs. Therefore, edema and secondary organ dysfunctions should be closely monitored in the early postoperative period. Increased fluid in the intracellular area should be withdrawn into the vascular bed and appropriate treatment methods should be applied to remove it from the body through diuresis. Also, we believe that BIA, which is a non-invasive procedure, can be safely used in operations where cardiac balance should be closely monitored during and after cardiopulmonary bypass (heart valve patients, low ejection fraction patients, elderly patients, pediatric patients, etc.). Future studies with different patient groups on this subject will increase our knowledge.

REFERENCES

1. Feyrer R, Harig F, Cennjevar R, Mahmoud O, Doreth M, et al. Bioelectrical impedance analysis in cardiac surgery. Internet J Thoracic Cardiovasc Surg. 2002;5(2):[about 12 screens].

2. Chumlea WC, Guo SS. Bioelectrical impedance and body composition: present status and future directions. Nutr Rev. 1994;52(4):123-31. doi:10.1111/j.1753-4887.1994.tb01404.x.

3. Gagnon R, Gagner M, Duplessis S. [Variations of body composition by bioelectrical impedancemetry after major surgery]. Ann Chin. 1994;48(8):708-16. French.

4. Chioléro RL, Gay LJ, Cotting J, Gurtner C, Schutz Y. Assessment of changes in body water by bioimpedance in actually ill surgical patients. Intensive Care Med. 1992;18(6):322-6. Erratum in: Intensive Care Med 1992;18(8):498. doi:10.1007/bf01694359.

5. Zillikens MC, van den Berg JW, Wilson JH, Swart GR. Whole-body and segmental bioelectrical-impedance analysis in patients with cirrhosis of the liver: change after treatment of ascites. Am J Clin Nutr. 1992;55(3):621-5. doi:10.1093/ajcn/55.3.621.

6. Holt TL, Cui C, Thomas BJ, Ward LC, Quirk PC, Crawford D, et al. Clinical applicability of bioelectric impedance to measure body composition in health and disease. Nutrition. 1994;10(3):221-4.

7. Leiter LA. Use of bioelectrical impedance analysis measurements in patients with diabetes. The diabetes control and complications trial research group. Am J Clin Nutr. 1996;64(3 Suppl):S55-85. doi:10.1093/ajcn/64.3.5155.

8. Schols AM, Wouters EF, Soeters PB, Westerterp KR. Body composition by bioelectrical-impedance analysis compared with deuterium dilution and skinfold anthropometry in patients with chronic obstructive pulmonary disease. Am J Clin Nutr. 1991;53(2):421-4. doi:10.1093/ajcn/53.2.421.

9. McNeill G, Fowler PA, Maughan RJ, McGaw BA, Fuller MF, Gvozdanovic D, et al. Body fat in lean and overweight women estimated by six methods. Br J Nutr. 1991;65(2):95-103. doi:10.1079/bjn19910072.

10. Bracco D, Thiébaud D, Chioléro RL, Landry M, Burckhardt P, Schutz Y. Segmental body composition assessed by bioelectrical impedance analysis and DEXA in humans. J Appl Physiol (1985). 1996;81(6):2580-7. doi:10.1152/jappl.1996.81.6.2580.

11. Stewart SP, Bramley PN, Heighton R, Green JH, Horsman A, Losowski MS, et al. Estimation of body composition from bioelectrical impedance of body segments: comparison with dual-energy-x-ray absorptiometry. Br J Nutr. 1993;69(3):645-55. doi:10.1079/bjn19930066.

12. Segal KR, Van Loan M, Fitzgerald PI, Hodgdon JA, Van Itallie TB. Lean body mass estimation by bioelectrical impedance analysis: a four-site cross-validation study. Am J Clin Nutr. 1988;47(1):7-14. doi:10.1093/ajcn/47.1.7.

13. Kushner RF. Bioelectrical impedance analysis: a review of principles and applications. J Am Coll Nutr. 1992;11(2):199-209.

14. Jebb SA, Elia M. Techniques for the measurement of body composition: a practical guide. Int J Obes Relat Metab Disord. 1993;17(11):611-21.

15. Lukaski HC, Bolonchuk WW, Hall CB, Siders WA. Validation of tetrapolar bioelectrical-impedance method to assess human body composition. J Appl Physiol (1985). 1986;60(4):1327-32. doi:10.1152/jappl.1986.60.4.1327.

16. Roos AN, Westendorp RG, Frölich M, Meinders AE. Weight changes in critically ill patients evaluated by fluid balances and impedance measurements. Crit Care Med. 1993;21(6):871-7. doi:10.1097/00003246-199306000-00015.

17. Meguid MM, Lukaski HC, Tripp MD, Rosenberg JM, Parker FB Jr. Rapid bedside method to assess changes in postoperative fluid status with bioelectrical impedance analysis. Surgery. 1992;112(3):502-8.

18. Robert S, Zarowitz BJ, Hyzy R, Eichenhorn M, Peterson EL, Popovich J Jr. Bioelectrical impedance assessment of nutritional status in critically

No financial support.

No conflict of interest.

Authors’ roles & responsibilities

MG Substantial contributions to the conception or design of the work; or the acquisition, analysis, or interpretation of data for the work; final approval of the version to be published

CS Substantial contributions to the conception or design of the work; or the acquisition, analysis, or interpretation of data for the work; final approval of the version to be published

AH Substantial contributions to the conception or design of the work; or the acquisition, analysis, or interpretation of data for the work; final approval of the version to be published

MSA Substantial contributions to the conception or design of the work; or the acquisition, analysis, or interpretation of data for the work; final approval of the version to be published

NK Substantial contributions to the conception or design of the work; or the acquisition, analysis, or interpretation of data for the work; final approval of the version to be published
ill patients. Am J Clin Nutr. 1993;57(6):840-4. doi:10.1093/ajcn/57.6.840.
19. Bracco D, Revelly JP, Berger MM, Chioléro RL. Bedside determination of fluid accumulation after cardiac surgery using segmental bioelectrical impedance. Crit Care Med. 1998;26(6):1065-70. doi:10.1097/00003246-199806000-00029.
20. Ohri SK, Bjarnason I, Pathi V, Somasundaram S, Bowles CT, Keogh BE, et al: Cardiopulmonary bypass impairs small intestinal transport and increases gut permeability. Ann Thorac Surg. 1993;55(5):1080-6. doi:10.1016/0003-4975(93)90011-6.
21. Fiogbé E, Ferreira R, Sindorf MAG, Tavares SA, de Souza KP, de Castro Cesar M, et al. Water exercise in coronary artery disease patients, effects on heart rate variability, and body composition: a randomized controlled trial. Physiother Res Int. 2018;23(3):e1713. doi:10.1002/pri.1713.
22. Häussinger D, Lang F, Gerok W. Regulation of cell function by the cellular hydration state. Am J Physiol. 1994;267(3 Pt 1):E343-55. doi:10.1152/ajpendo.1994.267.3.E343.
23. Sakaguchi T, Yasumura K, Nishida H, Inoue H, Furukawa T, Shinouchi K, et al. Quantitative assessment of fluid accumulation using bioelectrical impedance analysis in patients with acute decompensated heart failure. Circ J. 2015;79(12):2616-22. doi:10.1253/circj.CJ-15-0723.
24. Núñez J, Mascarell B, Stubbe H, Ventura S, Bonanad C, Bodí V, et al. Bioelectrical impedance vector analysis and clinical outcomes in patients with acute heart failure. J Cardiovasc Med (Hagerstown). 2016;17(4):283-90. doi:10.2459/JCM.0000000000000208.
25. Weyer S, Zink MD, Wartzek T, Leicht L, Mischke K, Vollmer T, et al. Bioelectrical impedance spectroscopy as a fluid management system in heart failure. Physiol Meas. 2014;35(6):917-30. doi:10.1088/0967-3334/35/6/917.