Accounting for age of onset and family history improves power in genome-wide association studies

Authors
Emil M. Pedersen, Esben Agerbo, Oleguer Plana-Ripoll, ..., John J. McGrath, Florian Privé, Bjarni J. Vilhjálmsson

Correspondence
bjv@econ.au.dk (B.J.V.), emp@ph.au.dk (E.M.P.)
Accounting for age of onset and family history improves power in genome-wide association studies

Emil M. Pedersen,1,2,* Esben Agerbo,1,2,3 Oleguer Plana-Ripoll,1 Jakob Grove,2,4,5,6 Julie W. Dreier,1,3 Katherine L. Musliner,1,2,3 Marie Bækvad-Hansen,2,10 Georgios Athanasiadis,11 Andrew Schork,2,11 Jonas Bybjerg-Grauholm,2,10 David M. Hougaard,2,10 Thomas Nerup,2,11,12 Merete Nordentoft,2,13 Ole Mørsk,2,14 Søren Dalsgaard,1 Jakob Christensen,1,8,9 Anders D. Børglum,2,6,7 Preben B. Mortensen,1,2,3 John J. McGrath,1,15,16 Florian Prive´,1,17 and Bjarni J. Vilhjálmsson1,2,4,17,*

Summary

Genome-wide association studies (GWASs) have revolutionized human genetics, allowing researchers to identify thousands of disease-related genes and possible drug targets. However, case-control status does not account for the fact that not all controls may have lived through their period of risk for the disorder of interest. This can be quantified by examining the age-of-onset distribution and the age of the controls or the age of onset for cases. The age-of-onset distribution may also depend on information such as sex and birth year. In addition, family history is not routinely included in the assessment of control status. Here, we present LT-FH++, an extension of the liability threshold model conditioned on family history (LT-FH), which jointly accounts for age of onset and sex as well as family history. Using simulations, we show that, when family history and the age-of-onset distribution are available, the proposed approach yields statistically significant power gains over LT-FH and large power gains over genome-wide association study by proxy (GWAX). We applied our method to four psychiatric disorders available in the iPSYCH data and to mortality in the UK Biobank and found 20 genome-wide significant associations with LT-FH++, compared to ten for LT-FH and eight for a standard case-control GWAS. As more genetic data with linked electronic health records become available to researchers, we expect methods that account for additional health information, such as LT-FH++, to become even more beneficial.

Introduction

Identifying the genetic variants underlying diseases and traits is a hallmark of human genetics. In recent years, large meta-analyses of genome-wide association studies (GWASs) have identified thousands of genetic variants for common diseases,1–7 including psychiatric disorders,8–12 revealing a remarkably complex and polygenic genetic architecture for most traits. International research collaboration where GWAS summary statistics have been shared in large consortia has been vital to this success, allowing researchers to obtain large sample sizes needed to study polygenic diseases. Novel advances in computational methods have also contributed to this success by enabling researchers to do more with less data.13–17 Yet, for most of these traits and diseases, only a small fraction of the estimated heritable variation has been identified in GWASs,18,19 highlighting the need for even larger samples and more powerful analysis methods.

Currently, most case-control GWASs are conducted with a regression model where the outcome is the case-control status or occasionally the age of onset of disease.20 In this paper, we have opted for using the phrase age of onset over age at first diagnosis because they commonly refer to the same underlying thing, i.e., when a diagnosis is given. Recently, researchers have proposed several methods that leverage additional information to improve the power to detect genetic associations without having to increase the number of genotyped individuals. These include multivariate methods that leverage shared environmental or genetic correlations between traits and diseases21–25 as well as methods that account for age of onset.26–29 Perhaps the most fruitful development has come from methods that leverage family information to increase statistical power to identify associations, such as genome-wide association study by proxy (GWAX)30,31 and liability-threshold-model-based approach.32 The liability threshold model conditioned on family history (LT-FH)32 estimates the

1National Centre for Register-Based Research, Aarhus University, 8210 Aarhus, Denmark; 2Lundbeck Foundation Initiative for Integrative Psychiatric Research, 8210 Aarhus, Denmark; 3Centre for Integrated Register-Based Research at Aarhus University, 8210 Aarhus, Denmark; 4Biogenomics Research Centre, Aarhus University, 8000 Aarhus, Denmark; 5Department of Biomedicine and Center for Integrative Sequencing, Aarhus University, 8000 Aarhus, Denmark; 6Department of Biomedicine, Aarhus University, 8000 Aarhus, Denmark; 7Department of Biomedicine, Human Genetics, Aarhus University, 8000 Aarhus, Denmark; 8Department of Neurology, Aarhus University Hospital, 8200 Aarhus, Denmark; 9Department of Clinical Medicine, Aarhus University, 8200 Aarhus, Denmark; 10Department of Neonatal Screening, Department for Congenital Disorders, Statens Serum Institut, 2300 Copenhagen, Denmark; 11Institute of Biological Psychiatry, MRC Ctr. Hs, St. Mary’s Hospital, 2800 Roskilde, Denmark; 12Department of Clinical Medicine, University of Copenhagen, 2200 Copenhagen, Denmark; 13Mental Health Services in the Capital Region of Denmark, Mental Health Center Copenhagen, University of Copenhagen, 2100 Copenhagen, Denmark; 14Psychosis Research Unit, Aarhus University Hospital, 8245 Risskov, Denmark; 15Queensland Brain Institute, University of Queensland, St Lucia, QLD 4072, Australia; 16Queensland Centre for Mental Health Research, The Park Centre for Mental Health, Wacol, QLD 4076, Australia; 17These authors contributed equally
*Correspondence: bjv@econ.au.dk (B.J.V.), emp@ph.au.dk (E.M.P.) https://doi.org/10.1016/j.ajhg.2022.01.009. © 2022 The Authors. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/).
posterior mean genetic liability under the liability threshold model conditional on the case-control status of the individual, parents, and siblings. Here, “family history” refers to the case-control status of all family members, i.e., parents and siblings. As for GWAX, it considers any individual with a family member who has the disorder being studied as a case, increasing the number of cases. The GWAX phenotype remains a case-control phenotype. Although both GWAX and LT-FH can lead to power increases over case-control GWAS on real data, they achieve it in two different ways. It has been shown that GWAX can lead to a reduction in power when compared to a case-control GWAS; if the in-sample disease prevalence is high, however, LT-FH consistently provides an increase in power compared to case-control GWAS and GWAX. This power improvement in LT-FH stems from two main sources. First, it distills family information and the individual’s case-control status into a genetic liability estimate, resulting in a more informative outcome than the case-control status alone, to be used in GWASs. Second, it also allows researchers to include more individuals in their analysis. For instance, when studying breast cancer, we can derive the posterior genetic liability for genotyped males conditional on the family history for their mothers and sisters and thus include them in the GWAS.

However, family members often span a large age range, which can affect the expected disease prevalence because of changes in diagnostic methods and criteria over time. We refer to such differences in prevalence by birth year as “birth cohort effects.” For instance, in the iPSYCH (Lundbeck Foundation Initiative for Integrative Psychiatric Research) data, where genotyped individuals are born after 1980, we expect severe right censoring for many diagnoses. Survival models are routinely used in epidemiology to model time-to-event data in order to account for right censoring, time at risk, and age of onset as well as cohort effects. They can be used to improve genomic prediction and predict disease progression and have also been shown to provide up to 10% increase in power to detect genetic variants in GWASs when compared to standard logistic regression. Recently, computationally efficient survival models for GWASs have been proposed: both Cox regression and frailty models that can control for population and family structure in large samples. However, to the best of our knowledge, these advanced time-to-event GWAS methods cannot account for family history (without genotype information for family members) to boost statistical power, as observed for LT-FH. Furthermore, LT-FH posterior liability estimates cannot be used directly as an outcome in survival analysis, as these are not binary and, more fundamentally, survival models are based on a different generative model than the liability threshold model. Hujoel et al. proposed an approach to address this problem by accounting for age of onset in the genotyped individuals by linearly shifting the threshold for the genetic liabilities based on observed in-sample prevalence in different age groups but did not observe any improvements in power. We believe that this approach was unsuccessful in part because the in-sample estimate of the prevalence is subject to both a survival and selection bias and does not properly reflect prevalence in the population.

In this paper, we propose LT-FH++, a method that extends the model underlying LT-FH to account for information such as right censoring, age of onset, sex, and cohort effects. We achieve this by using a personalized threshold for each person (including family members), conditional on available information as well as general population incidence rates by age, sex, and birth year. LT-FH++ has been implemented into an R package (see data and code availability), which utilizes a Gibbs sampler implemented in C++ through the Rcpp R package. The personalized thresholds are made possible by replacing the Monte Carlo sampling used by Hujoel et al. with a much more efficient Gibbs sampler. The Gibbs sampler allows us to estimate the posterior mean genetic liability for each individual independent of one another, thereby making it highly scalable.

First, we perform a GWAS with the standard case-control phenotype as well as GWAX, LT-FH, and LT-FH++ outcomes for simulated data with the liability threshold model as the generative model. For real-world application, we analyzed mortality in the UK Biobank and four psychiatric disorders in the iPSYCH cohort.

Material and methods

Model

The underlying model is identical to the one used in LT-FH; as a result the model will only briefly be presented here, and the main differences will be elaborated on. Under the liability threshold model, each individual has a liability, $\ell$, which follows the standard normal distribution. An individual will be considered a case, increasing the number of cases. The threshold, $T$, is determined from the prevalence of the dichotomous disorder, such that $P(\ell \geq T) = K$, where $K$ denotes the prevalence in the population.

LT-FH builds on this idea, and for a single individual, the liability is assumed to be further decomposed into a genetic and environmental component, $\ell = \ell_g + \ell_e$. Both $\ell_g$ and $\ell_e$ are normally distributed and independent. We have

$$\ell_e \sim N(0, h^2), \ell_g \sim N(0, 1 - h^2)$$

Here, $h^2$ is the heritability on the liability scale. The LT-FH setup extends this idea to include parents and siblings. It considers a multivariate normal distribution given by

$$\ell = (\ell, \ell_p, \ell_{p1}, \ell_{p2}, \ell_s) \sim N(0, \Sigma)$$

$$\Sigma = \begin{bmatrix} h^2 & h^2 & 0.5h^2 & 0.5h^2 & 0.5h^2 \\ h^2 & 1 & 0.5h^2 & 0.5h^2 & 0.5h^2 \\ 0.5h^2 & 0.5h^2 & 1 & 0 & 0.5h^2 \\ 0.5h^2 & 0.5h^2 & 0 & 1 & 0.5h^2 \\ 0.5h^2 & 0.5h^2 & 0.5h^2 & 0.5h^2 & 1 \end{bmatrix}$$

Here, $\ell_g$ denotes the full liability for the individual (denoted $\ell$ for a single individual above), and $\ell_e$ denotes the genetic component
of this liability, \( \theta_p \) and \( \theta_p \) denotes the full liability of each parent, while \( \theta_s \) denotes those of the sibling. The example above includes one sibling only, but in theory any number of siblings could be included in the model. We are interested in estimating the posterior mean genetic liability for each individual conditional on family information:

\[
E[\theta_s | Z] = (\theta_{s1}, \theta_{s2}, \theta_{s3}, \theta_{s4})^T.
\]

Here, \( Z \) denotes the vector of status for the family, consequently a restriction is placed on each individual's full liability. In the case of everyone’s having the disorder, we would consider the space \( \{0 \leq R(i) \leq T_i \text{ for all } i\} \), where \( i \) denotes a family member, \( T_i \) denotes the family member's threshold, and \( \theta_s \) denotes their full liability. In LT-FH, the thresholds are the same for all children (the offspring and any siblings), and another threshold is used for all parents.

The choice of thresholds is where LT-FH++ starts to differentiate itself from LT-FH. In short, the liability thresholds are personalized, such that every individual, sibling, or parent has a potentially unique threshold that is determined by their age, birth year, and sex. Furthermore, we adopt an age-dependent liability threshold model, where the threshold is dynamic in the sense that it decreases as a population grows older. This idea is illustrated in Figure 1A, where the threshold decreases as time progresses for a population, with marks for ages 15, 25, 35, 50, and 60. This model assumes that the threshold decreases continuously as time progresses, and these marks can be seen as snapshots in time, where an individual who was diagnosed at one of the marks had an assumed (fixed) liability equal to said mark. This age-dependent liability threshold model allows us to be very precise with the liability for cases when an accurate age of onset is available. If an accurate estimate of age of onset is not available, then the threshold can still be personalized on the basis of other available information, with the modification that we do not fix the full liability but integrate over all liabilities above the personalized threshold. Interestingly, the age-dependent liability threshold model can be thought of as a survival model (see below).

Another point where LT-FH++ differs from LT-FH is in how siblings are included. LT-FH includes the siblings by specifying the number of siblings and assigns a single case-control status to the siblings with the condition that at least one sibling has the disorder. However, a more fine-grained inclusion of the siblings, where each sibling is added individually, is not available. LT-FH++ expects each individual and their family members to be added separately, such that information on each individual can be accounted for.

**Relationship with survival analysis**

In survival analysis GWASs, the risk for becoming a case in a time interval depends on the covariates in the model. This is reflected by a hazard rate \( h(t|x) \), which describes the event rate. In our context, it would refer to the rate for becoming a case. This rate depends on both time \( t \), and covariates of the model \( x \), e.g., genotypes. The hazard rate (also referred to as the intensity) can be approximated by \( h(t|x) = P(T(t + dt) \leq T(t) | x) \) where \( dt \) is a small change in time. \(^{10} \) \( T(t) \) is the threshold for being a case at time \( t \), and \( \theta \) is the full liability of an individual. This means that the hazard rate is proportional to the probability an event occurs within a time interval \( (t, t + dt) \), given that no event had occurred earlier. For different types of survival analyses, we can estimate this probability by using the hazard rate, e.g., for a Cox proportional hazards model where we aim to estimate the effect of a genotype \( x \) on the hazard rate, it becomes \( P(T(t + dt) \leq T(t) > x) = d\lambda(t|x) = d\lambda_0(t) \exp(\beta x) \). To keep notation simpler, we will denote the genetic liability of individual \( i \) as \( g_i \), instead of \( \theta_s \), and if we furthermore assume that the genetic component for an individual of a case-control outcome contributes to the hazard rate such that \( \lambda(t|g_i) = \lambda_0(t) \exp(g_i) = \lambda_0(t) \exp(\beta x) \), where \( x \) denotes the genotype of the individual and \( \beta \) their true effects (in the Cox-regression model). Conceptually, this means that individuals with higher than average genetic risk, i.e., \( g_i > 0 \), will be at higher risk to become cases throughout their lives, irrespective of age. These high-risk individuals will on average also have earlier age of onset.

To understand how this model relates to the proposed age-dependent liability threshold model, we can derive the same probability to approximate the corresponding hazard rate. Under the LT-FH++ model, the probability for an individual \( i \) to be diagnosed (become a case) within a time interval \( dt \) can be written as \( P(T(t + dt) \leq \theta_i | T(t) > \theta_i) \), where \( t \) again denotes the age of the individual and \( T(t) \) now denotes the age-dependent liability threshold. We note that \( T(t) \) is a monotonic decreasing function as the prevalence of a case-status (i.e., cumulative lifetime incidence proportion) always increases with age (conditional on birth year and sex). Furthermore, \( \theta_i \) denotes the full liability of the individual and \( g_i \) the genetic component of that liability (which is generally on a different scale than a genetic component in Cox regression). The liability threshold model assumes that the liability of an individual consists of genetic and environmental components, i.e., \( g_i = g_i + \epsilon_i \). It also assumes that these are independent, follow a Gaussian distribution, and have variance \( \sigma^2 = h^2 + 1 - h^2 \), respectively. Hence using these, we can expand the probability of being diagnosed within a time interval \( dt \) further as follows:

\[
P(T(t + dt) \leq \theta_i | T(t) > \theta_i, g_i) = P(T(t + dt) \leq \theta_i | T(t) > \theta_i, \hat{g}_i) \times (P(T(t) > \theta_i | g_i))^{-1} = \left( 1 - \phi \left( \frac{T(t)}{\sqrt{1 - h^2}} \right) \right) \times \left( 1 - \phi \left( \frac{T(t + dt) - \theta_i}{\sqrt{1 - h^2}} \right) \right) \times \left( \phi \left( \frac{T(t)}{\sqrt{1 - h^2}} \right) \right). \]

Plotting this function for different thresholds and genetic liability values shows that the probability for being diagnosed within the time interval, and thus the hazard rate, increases linearly as a function of the genetic liability when \( g_i \) is near \( T(t) \) or larger. We compare this probability with the corresponding Cox regression probability assuming a base incidence rate of \( \lambda_0(t) = \alpha \), where \( \alpha \) is determined by the prevalence. These two probabilities, which are proportional to the hazard rate, are plotted as a function of \( g_i \) in Figure S50, illustrating how the hazard rates of the two models depend on \( g_i \). We note that the two models share the properties that individuals with higher than average genetic risk will, on average, be more likely to become cases within any time interval and have earlier age of onset.

It may seem counterintuitive that a deterministic model such as the age-dependent liability threshold model, where the liability is constant throughout life, can be recast as a survival analysis model. The reason for this is that although the outcome of the age-dependent liability threshold model is always known.
given the liability, one never observes this liability. Hence, the environmental term, which can be thought of as capturing various environmental effects as well as chance events and other non-genetic effects, leads to a non-deterministic survival analysis model.

**Sampling strategy**

If we consider an individual with disease status available for both parents, but no siblings, then we have a total of six unique ways to configure the status vector, $Z$, when disregarding other information because the scenario where a single parent is a case can happen in two ways. LT-FH estimates the posterior mean genetic liability for each of these configurations by sampling a large number of observations from the multivariate normal distribution described above. The observations are then grouped into these six unique configurations, and the genetic liabilities are estimated by averaging genetic liabilities within each configuration. This strategy works well when there are a limited number of configurations but becomes infeasible when the number of configurations becomes too large.

LT-FH++ cannot efficiently use the same sampling strategy because the personalized thresholds increase the number of potential configurations such that the strategy becomes intractable. Instead LT-FH++ considers each family as a unique configuration because it uses individualized thresholds. To derive the posterior means efficiently, we use a Gibbs sampler to sample from a

**Figure 1. Overview of LT-FH++ and illustration of the differences between LT-FH and LT-FH++**

(A and B) An age-dependent liability threshold model with different thresholds marked (A). The marks correspond to the prevalence at the age of 80 years (10%), 50 years (6%), 35 years (3.5%), 25 years (2%), and 15 years (1%). The posterior mean estimate of the liability is obtained by integrating over the liability space spanned by the genotyped individual and their family members (B). Here, we consider a brother and a mother, where the contour lines indicate the joint multivariate liability density of the mother and the brother (assuming a heritability of 0.5). Using fixed population prevalence for males and females (dashed lines), and assuming mother and brother are cases, LT-FH integrates over the blue shaded area to estimate the genetic liability. In contrast LT-FH++ considers the age of onset, sex, and birth year for family members to obtain a more precise genetic liability estimate highlighted by the red dot. In short, the additional information collapses the area to integrate to a single value.

(C) An overview of how LT-FH++ GWAS works and what information it accounts for. In contrast to LT-FH, which accounts for the case-control status of the genotyped individual and family history, LT-FH++ also uses population prevalence information to account for gender, age, and birth year of family members. As with LT-FH, the predicted liabilities are then used as a continuous outcome in a GWAS via BOLT-LMM.
truncated multivariate normal distribution.\textsuperscript{30} The truncation points in the truncated multivariate normal distribution are the personalized thresholds. Sampling for all individuals is fast, requires far fewer observations, and can be easily parallelized across individuals, as each family is independent from each other.

**Practical considerations for LT-FH++ GWAS**

When deriving the posterior mean genetic liabilities, it is important to ensure that the genotyped family members do not have shared family members, as that can otherwise lead to individuals’ being more correlated than expected given their genetic similarity.\textsuperscript{32} This can cause problems in subsequent GWAS analysis and lead to inflation of false positive rates. We therefore recommend only applying LT-FH++ to unrelated individuals, where the relatedness threshold is stringent enough to ensure that no genotyped pair of individuals have common family members.

As LT-FH++ reports effects on a genetic liability scale, these can be hard to interpret. However, the general strategy proposed by Huijoe et al.\textsuperscript{32} can be used to transform these to per-allele observed-scale effect sizes for non-standardized phenotypes.

LT-FH++ has several ways to deal with missing information. If age-of-onset information is missing for an individual, the threshold used for that individual will correspond to the average prevalence (the LT-FH threshold). If age-of-onset information is available for the family members, their threshold can still be personalized. The estimated genetic liability under LT-FH++ with no age-of-onset information available for an individual and their family members but complete family history information would be identical to the LT-FH estimate. If case-control status is missing for the genotyped individual, we integrate over the entire range of liabilities for this individual. If case-control status is missing for family members, we exclude these from the analysis. For example, if the case-control status is known for one parent but not the other parent, we exclude the second parent from the analysis. Finally, age-of-onset information acts as an additional level of fine-tuning in the age-dependent liability threshold model. In our analysis, the threshold depends on sex, birth year, and age or age of onset, but if less information is available, e.g., no sex, then an estimate of the threshold could still be based on the birth year and age or age of onset. Similarly, if prevalence estimates are known for a given (categorical) risk factor (e.g., smoking status), then LT-FH++ can account for this additional risk factor (also in family members).

**Prevalence information**

The age-dependent prevalence of attention deficit-hyperactivity disorder (ADHD [MIM: 143465]), autism spectrum disorder (ASD [MIM: 209850]), depression (DEP [MIM: 608516]), and schizophrenia (SCZ [MIM: 181500]) was obtained through Danish national population-based registers. For these estimates, we included all 9,251,071 persons living in Denmark at some point between January 1, 1969 and December 31, 2016. Each individual in the study was followed from birth, immigration to Denmark, or January 1, 1969 (whichever happened last) until death, emigration from Denmark, or December 31, 2016 (whichever happened first). All dates were obtained from the Danish Civil Registration System,\textsuperscript{21} which has maintained information on all residents since 1968, including sex, date of birth, continuously updated information on vital status, and a unique personal identification number that can be used to link information from various national registers. Information on mental disorders was obtained from the Danish Psychiatric Central Research Register,\textsuperscript{42} which contains data on all admissions to psychiatric inpatient facilities since 1969 and visits to outpatient psychiatric departments and emergency departments since 1995. The diagnostic system used was the Danish modification of the International Classification of Diseases, Eighth Revision (ICD-8) from 1969 to 1993, and Tenth Revision (ICD-10) from 1994 onward. The specific disorders were identified with the following ICD-8 and ICD-10 codes: ADHD (308.01 and F90.0), autism (299.00, 299.01, 299.02, 299.03 and F84.0, F81.4, F84.5, F84.8, F84.9), depression (296.09, 296.29, 298.09, 300.49 and F32, F33), and schizophrenia (295.x9 excluding 295.79 and F20). For each individual in the study, the date of onset for each disorder was defined as the date of first contact with the psychiatric care system (inpatient, outpatient, or emergency visit). All analyses were done separately for each sex and for each birth year. The cumulative incidence function for each disorder was estimated with the Aalen-Johansen approach considering death and emigration as competing events.\textsuperscript{43} The cumulative incidence over age is interpreted as the proportion of persons diagnosed with the specific disorder before a certain age.

**Personalized thresholds**

With the cumulative incidence rate tables, we are able to assign personalized thresholds to everyone with sufficient information available. Examples of cumulative incidence rate curves can be seen in Figures S21, S27, S32, S38, and S44. Under the liability threshold model, sex, birth year, and age for controls or age of onset for cases can uniquely determine the threshold for an individual. On the basis of this information, a proportion is assigned to them, which is transformed to an individual's threshold through the inverse normal cumulative distribution function.

For controls, it has allowed us to tailor the threshold in the liability threshold model to each individual, similar to what is seen in Figure 1A, where the threshold is decreasing as an individual is getting older. In short, the older a control is, the larger a proportion of the possible liabilities in the liability threshold model can be excluded as no longer attainable. For cases, the tailored threshold means we are able to very accurately estimate what a person's full liability is for a given disorder under the liability threshold model. Because the full liability can be accurately estimated for a case by the assigned threshold, we will fix the full liability of a case to be the threshold in the model.

**Simulation details**

For the simulations, we simulated 100,000 unrelated individuals each with 100,000 independent single-nucleotide polymorphisms (SNPs). We simulated two parents and between zero and two sibs. The parents’ genotypes were drawn from a binomial distribution with probability parameters equal to the allele frequency (AF) of the corresponding variant. The variant AF was drawn from a uniform distribution on the interval (0.01, 0.49). The parents’ genotypes were either 0, 1, or 2; we defined the child's genotypes as the average between the genotypes of both parents, rounding values of 0.5 or 1.5 up or down with equal probability. Allele effect sizes were drawn from $\mathcal{N}(0, h^2/C)$, where $C$ was the number of causal SNPs and $h^2$ denoted the heritability. Case-control status was assigned with a liability threshold model.

The default simulation setup consisted of causal SNPs assigned to positions at random, two different prevalences, 5% and 10%, $C$ set to 1,000, and a sex-specific prevalence of 8% for men and 2% for women. When the prevalence was 10%, these sex-specific
prevalences were doubled. To generate the age of onset, we assumed that the cumulative incidence curve followed a logistic function because it resembles real-world cumulative incidence rates for some traits, see Figures S21, S27, S32, S38, and S44. The logistic function is given by

\[ T(x) = \frac{L}{1 + e^{K(x-x_0)}} \]

where \( L \) denotes the maximal attainable prevalence value, \( k \) is the growth rate, and \( x_0 \) denotes the age (in years) at which \( K = L/2 \), which is the midpoint of the curve, i.e., median age of onset. Due to the properties of the function, the lifetime prevalence will only be approximately \( L \) (only slightly smaller). These parameters resulted in an age of onset that was largely normally distributed around the median age, \( x_0 \). The cumulative incidence rate curve allows us to obtain the expected prevalence at each age, which we can then translate into a threshold in the liability threshold model, i.e., an earlier diagnosis indicates higher liability for the trait. We fix the lifetime prevalence \( L \) in the combined population and the corresponding sex-specific lifetime prevalences. We then assigned each individual a male or female sex with equal probability. In our simulation, we assumed males were four times as likely to be cases than females. For the two lifetime prevalences (5% and 10%), this corresponded to 8% and 16% prevalence among males (liability thresholds \( T_{\text{male}} = 1.41 \) and \( T_{\text{male}} = 0.99 \)) and 2% and 4% prevalence among females (liability thresholds \( T_{\text{female}} = 2.05 \) and \( T_{\text{female}} = 1.75 \)). E.g., with an overall prevalence of 5%, we used \( L = 0.08 \) for males and \( L = 0.02 \) for females. We also set \( k \) to 1/8 and \( x_0 \) to 60 such that 90% of cases have an age of onset between 36.5 and 83.5.

A family consisted of one offspring, two parents, and zero to two siblings. The age of the cases was set to the age of onset. The age of onset was assigned by taking the inverse of the logistic function on the full liability’s quantile under the standard normal distribution. Individuals with an age lower than their age of onset would normally be considered controls because they had not yet had the time to develop the disorder. However, setting high liability individuals to controls because age of onset was later than age was decided against to properly fix the number of cases to the prevalence in the simulated data. For controls, the offspring’s age was uniformly distributed between 10 and 60. The parents’ age was set to the age of the child plus a uniform draw between 20 and 35, allowing for up to 95 year olds. The threshold was assigned with the logistic function with the age and sex as inputs. For simplicity, birth year was not modeled. Finally, we simulated sample ascertainment by downsampling controls such that cases and controls had equal proportions (50% each). For 5% prevalence, this resulted in a sample size of 10,000 and 20,000 individuals when using a prevalence of 10%.

### GWAS in UK Biobank

We restricted individuals to the White British group (field 222006) and to the individuals used for computing the principal components (PCs) in the UK Biobank (field 222020). These individuals are unrelated and have passed some quality control (see section S3 of Bycroft et al.44). This resulted in 337,475 individuals. Table 1 shows a breakdown of how many people are cases and controls for the genotyped individuals and parents. We used the genotyped SNPs for the UK Biobank participants as model SNPs in BOLT-LMM,38 after removing SNPs with minor allele frequency (MAF) < 0.01, missing call rate > 0.01, and Hardy-Weinberg equilibrium p value < 1 × 10⁻¹⁰, which left us with a total of 504,138 SNPs. When performing the GWAS, we used the imputed SNPs in bgen files and removed SNPs with an MAF < 0.005 or info score < 0.6, which resulted in 11,335,564 SNPs. We used BOLT-LMM v2.3.2 with age, sex, and the first 16 PCs as covariates. The three mortality outcomes used in the UK Biobank were case-control status, LT-FH, and LT-FH++. We considered the binary death outcome as the case-control phenotype, and LT-FH and LT-FH++ further utilize the mortality status of both parents but no siblings. The UK Biobank data was downloaded on the March 17, 2020.

LT-FH++ and LT-FH require prevalence information, which was acquired from the Office for National Statistics (ONS). Mortality rates for England and Wales were available from 1841 to the present day. The same information was available for all of the United Kingdom (UK), but only from the 1950’s onward. Because England is the most populous country in the UK, we believe these mortality rate estimates are a good proxy for all of the UK. From the mortality rates provided by ONS, we calculated the cumulative incidence curves for death for each birth year from 1841 onward and for both sexes. We used this information to calculate the personalized thresholds in LT-FH++, accounting for birth year, sex, and current age or age of death.

To determine the birth year of the parents in the UK Biobank, we assumed they were 25 years older than their child (for which year of birth is available in data field 34). The resulting estimated birth year was then used in the prevalence curves to get the liability thresholds for each parent. Age of death for the parents are available in data fields 1807 and 3526.

Note that, in LT-FH, it is not possible to adjust for sex, age, or cohort effects at the individual level, but two different thresholds can be specified, one for all parents and one for all children. Therefore, we assumed the same age for all children and the same age for all parents when running LT-FH. We used the last recorded death as the endpoint, which happened in 2018, and assumed all children were 55 years old and parents were 85 years old. This translated into an assumed birth year of 1963 and 1933, respectively. On the basis of these birth years, we found the prevalence of death for these birth years and ages in the survival curve and averaged the sex-specific prevalences. For LT-FH, we also considered thresholds on the basis of prevalence estimated in the UK Biobank participants and their parents, however we did not see any significantly different results when comparing to the population-based prevalence estimates (results not shown). A heritability of 20% was used for LT-FH and LT-FH++.

### GWAS in iPSYCH

The iPSYCH cohort has recently received a second wave of genotyped individuals, increasing the number of genotyped individuals from ~80,000 to ~143,000.55 The two iPSYCH waves have been imputed separately with the Ricopili pipeline.46 After
combining the two waves and removing any SNP with missingness > 0.1 or MAF < 0.01, we have a total of 4,706,774 SNPs. When performing a GWAS, we restrict the analysis to individuals classified as controls in the iPSYCH design and individuals diagnosed with the analyzed phenotype, even when using LT-FH or LT-FH+++. We filtered for relatedness with a 0.0884 KING-relatedness cutoff and restricted the analysis to a genetically homogeneous group of individuals by calculating a Mahalanobis distance based on the first 16 PCs and keeping individuals within a log-distance of 4.5. For a breakdown of the number of individuals included in each GWAS and the number of cases and controls, see Table 2. We used BOLT-LMM v2.3.2 to perform the GWAS with sex, age, wave, and the first 20 PCs as covariates. LT-FH and LT-FH+++ require an estimate for the heritability; we used 75% for ADHD, 83% for autism, 37% for depression, and 75% for schizophrenia. See prevalence information for details on how the cumulative incidence curves were derived.

When assessing power between outcomes, we considered SNPs that are in the iPSYCH cohort and have been found to be significantly associated with the psychiatric disorder being analyzed in the largest publicly available meta-analyzed GWAS. We used PLINK to perform linkage disequilibrium (LD) clumping on the external summary statistics. We used PLINK’s default parameters, except for the significance thresholds. The PLINK p value threshold we used was 5x10^-6 for both the index SNPs and the clumped SNPs. We used the default window size of 250 kb and the LD threshold of 0.5.

### Results

#### Overview of methods

The LT-FH+++ method proposed here extends the LT-FH method to account for additional information for family members, such as age, sex, and cohort effects for case-control outcomes. LT-FH assumes a liability threshold model, where every individual has an underlying liability for the outcome but only becomes a case if the liability exceeds a given threshold, which is determined by the sample or population prevalence. It further assumes that the covariance structure depends on the heritability and relatedness coefficient between each individual, which is a reasonable assumption for polygenic case-control diseases. Under these assumptions, LT-FH estimates the posterior mean genetic liability conditional on the case-control status of the genotyped individual and their family members via a Monte Carlo sampling. The posterior mean genetic liability is then used as the continuous outcome in a GWAS, e.g., with BOLT-LMM.

In LT-FH++, we introduce an “age-dependent liability threshold model” to capture the effect of age and replace the Monte Carlo sampling with a much more computationally efficient Gibbs sampler. Illustrated in Figure 1A, the age-dependent liability threshold model extends the liability threshold model by assuming that the threshold for becoming a case at a given age corresponds to the prevalence of the disease at that age. Interestingly, this model can be viewed as a type of survival analysis (see material and methods). We can then account for additional information, such as birth year and sex, by further conditioning the disease prevalence on this information. This leads to an individualized disease liability threshold for each person, including family members, which in practice requires us to be able to estimate separate genetic liabilities for each individual. This is made possible by replacing the Monte Carlo strategy of LT-FH with the computationally efficient Gibbs sampler that can sample from multivariate truncated Gaussian distributions to obtain personalized genetic liability estimates. As illustrated in Figure 1B, this results in more precise genetic liability estimates for LT-FH+++ under the model compared to LT-FH, which for a population translates also into more variable genetic liability estimates (see Figure S1). Thus, in order to reap the full benefit of LT-FH++, it requires prevalence information to be available by age, sex, and birth year. Fortunately, such information is often partially or fully available on a population level, e.g., in the Danish registers. The use of population prevalence information also allows LT-FH+++ to estimate the genetic liability on a population scale, which may also reduce the risk of ascertainment and selection bias. We summarize the information that LT-FH+++ can account for and the two-step procedure of estimating individual genetic liabilities and performing GWASs on these in Figure 1C.

#### Simulation results

We examined the performance of LT-FH+++ by using both simulated and real data. We simulated 100,000 unrelated
individuals each with 100,000 independent SNPs and their family (two parents and 0–2 siblings). We generated case-control outcomes under the liability threshold model and assigned age of onset by assuming the prevalence followed a logistic curve as a function of age (see materials and methods for simulation details). We first considered the simulations for families with no siblings. We benchmarked LT-FH++ against case-control status and LT-FH. The results for 5% prevalence are shown in Figure 2, and the results for 10% prevalence can be found in Figure S12. We simulated sample ascertainment by downsampling controls such that cases and controls had equal proportions (50% each), which translated into a total of 10,000 individuals for a 5% prevalence and 20,000 individuals for a 10% prevalence. The simulation results confirmed the increase in power (number of causal SNPs detected) of LT-FH over standard GWASs when accounting for family history. When also accounting for sex differences and age in LT-FH++, we observed a further increase in power, especially when the cases were ascertained (downsampling controls). Averaging over ten simulations, LT-FH had a power improvement over standard GWASs between 14% and 54%, where less power improvement was observed when downsampling controls. In contrast, the average power increase for LT-FH++ and standard GWASs was between 34% and 61%. Without downsampling controls, the relative improvements of LT-FH++ over LT-FH for a 5% and 10% prevalence were 4% and 5%, respectively. However, when downsampling controls, we observed an improvement of 18% for a 5% prevalence and 15% for a 10% prevalence. In Table S14, p values for various tests of difference between LT-FH and LT-FH++ can be seen. All tests showed a significant difference between them, in favor of LT-FH++. In Table S15, the absolute and relative difference in the number of causal SNPs detected within each simulated dataset and for each phenotype compared to LT-FH is shown. When simulating families with two siblings, we observed an increase in mean power and causal test statistics (across the ten simulations) compared to families with no siblings, but the relative improvement of LT-FH++ over LT-FH remained the same (results not shown). We also assessed the robustness of LT-FH++ by misspecifying model hyper-parameters, i.e., the heritability and prevalence parameters. Simulated heritability was 50%, and when misspecifying it, we used 25% and 75%. For the prevalence, we used simulated values of either 5% or 10% and used either half or double of the true value to assess the impact of misspecifying this parameter. We found in, e.g., a prevalence of 5% or 20% when the true prevalence was 10%. In Figures S4, S5, S13, and S14, we see similar results as in Figure 2 with nearly identical mean null $\chi^2$ statistics, mean causal $\chi^2$ statistics, and power. LT-FH++ is therefore robust to misspecification of heritability and prevalence.

To better understand when one could expect gain from accounting for age of onset and family history, we performed additional simulations where we varied the number of individuals N as well as the completeness/missingness of the family history and age-of-onset information (see Figures S6–S11 and S15–S20 and Table S13). We found that the relative gain in statistical power of using LT-FH++ instead of LT-FH was largely constant when varying sample...
size, completeness of family history, and age-of-onset information. As expected, the power decreased for both LT-FH++ and LT-FH when family information was missing. However, the relative power gain of LT-FH++ over LT-FH increased when family information was missing or when the cases were ascertained. In short, one can expect to gain the most when the in-sample prevalence is high either among participants or in the family history.

Lastly, we have performed simulations for computation time. The results are shown in Figures S2 and S3 and the numbers are available in Table S13. In short, LT-FH++ scales linearly with sample size, and using 32 cores, it can estimate posterior genetic liabilities for 350,000 individuals in less than 25 min. All computation time simulations were performed on genomeDK.

Analysis of mortality in the UK Biobank
To evaluate the performance of LT-FH++ on real data, we chose mortality in the UK Biobank, as this is the only outcome available where we have age information for family members, i.e., we have age or age of death for mothers and fathers. We then obtained population prevalence information from the Office for National Statistics (ONS), which provides mortality rates for England and Wales by sex and birth year (since 1841), and for the UK since 1950. This allowed us to obtain individualized prevalence thresholds for LT-FH++ for each genotyped individual and their parents (see material and methods for details). The mortality rates by age and sex are shown for each decade in Figure S21.

The Manhattan plots for standard case-control, LT-FH, and LT-FH++ GWASs can be found in Figure 3 (see material and methods for analysis details). When using the case-control phenotype as the outcome in GWASs, we did not observe any genome-wide significant SNPs. For LT-FH, we found two genome-wide significant associations, including a well-known association with mortality in APOE (MIM: 107741) and in HYKK (MIM: 614681), which is strongly associated with smoking behavior. These were also the two strongest associations found with LT-FH++, which additionally found eight other independent associated variants, where independence was assessed with GCTA-COJO. The ten identified variants are shown in Table S10, of which three variants have not previously been identified as associated with mortality or aging. One of these is near HLA-B (MIM: 142830), which is involved in immune response and has been found to be associated with white blood cell count and Psoriasis. The second association is near MYCPCP2 (MIM: 610392), which has previously been identified as being associated with chronotype, and the expression of this gene was recently found to increase with age and interact with the SARS-CoV-2 proteome. The third association was near ZBBX (MIM: 609118), which has been found to be associated with changes in DNA methylation with age.

Because we do not know the true causal variants for mortality, we cannot accurately estimate power. Power has a formal statistical definition that requires us to know
whether a SNP is causal or not. However, to approximate relative power gain (between methods) we considered a set of LD-pruned variants with a $p$ value below $5 \times 10^{-6}$ for at least one of the three compared outcomes. Assuming that these are enriched to be causal variants (or in strong linkage with causal variants), and that their null test statistics have similar inflation, then one can approximate relative power gain. We measure the increase in effective sample size by comparing $Z$ scores from both methods. We then refer to this increase in effective sample size as an increase in power because power increases with sample size. We also note that the GWAS Q-Q plots for mortality for all methods (Figures S24–S26) showed no sign of test statistics’ being inflated, suggesting that false-positive rates are similar across all methods. For LT-FH++, it leads to an estimated power increase of 42% over LT-FH. Because the $Z$ scores squared are the $\chi^2$ statistics, we opted to illustrate the power improvement of LT-FH++ over LT-FH through the $\chi^2$ statistics. We plotted the $\chi^2$ statistic for variants with a $p$ value below $5 \times 10^{-6}$ in Figure 4. LT-FH and LT-FH++ both had a large increase in power over case-control status, resulting in an estimated relative power increase of 110% and 187%, respectively: The $\chi^2$ statistics and $Z$ scores plots compared to case-control status can be found in Figures S22 and S23.

**Application to four psychiatric disorders in iPSYCH**

The iPSYCH data with linked Danish registers has age and age-of-onset information for all close family members of genotyped individuals. We considered four psychiatric disorders in the iPSYCH data: ADHD, autism, depression, and schizophrenia. For each of these, we obtained prevalences by birth year, age, and sex by using the same diagnostic criteria (see material and methods for details). As shown in Figures S27, S32, S38, and S44 the prevalence of psychiatric disorders strongly depends on birth year and sex, making it an appealing application of LT-FH++. We performed a GWAS of the three outcomes, case-control GWAS, LT-FH, and LT-FH++, for the four psychiatric disorders (see material and methods for analysis details). Across the four psychiatric disorders, we found ten genome-wide significant associations by using LT-FH++ compared to eight by using both LT-FH and case-control. Specifically for ADHD, LT-FH++ found seven significant associations, while case-control status and LT-FH found five. All three outcomes identified the same five variants, and LT-FH++ identified two additional variants for ADHD. One of these variants was on chromosome 11 near LINC02758 (MIM: 6187711), which was found to be associated with ADHD in a meta-analysis, and the other one was on chromosome 14 in AKAP6 (MIM: 604691), which has previously been identified as being associated with cognitive traits. The Manhattan plots for ADHD can be seen in Figure 5 for all three outcomes, i.e., case-control, LT-FH, and LT-FH++ (see material and methods for details). Manhattan plots for all three outcomes are very similar and no one outcome clearly outperforms the others. However, LT-FH++ does have two associations that were close to genome-wide significance with both LT-FH and case-control analysis but did not pass the significance threshold. Similarly, LT-FH++ and case-control have one SNP that is not found by LT-FH, but it is also close to the genome-wide significance threshold for LT-FH. In Figure 6, we show the $\chi^2$ statistics plot restricting to LD-clumped SNPs with a $p$ value threshold of $5 \times 10^{-6}$ for the index SNP and the clumped SNPs from the largest external meta-analyzed ADHD summary statistics (see material and methods for details). If one
method had clearly performed better than another, we would have expected to see a slope different from one, however this is not the case here. Overall, there is little power improvement by using either LT-FH or LT-FH++ over case-control GWAS for ADHD.

We performed a similar analysis for the three other iP-SYCH disorders analyzed, namely ASD, depression, and schizophrenia. The Manhattan, QQ, Z scores, and \( \chi^2 \) statistics plots can be found in Figures S28–S31, S33–S37, S39–S43, and S45–S49 for all iP-SYCH analysis. For depression and schizophrenia, we found no genome-wide significant hits for any method used and the Z scores and \( \chi^2 \) statistics indicate no difference in power between standard GWAS, LT-FH, and LT-FH++. For autism, we do see genome-wide significant hits: three for case-control GWAS and LT-FH++ and four for LT-FH. The SNP that is unique to LT-FH is also highly suggestive for case-control GWAS and LT-FH++. A table containing the COJO-independent SNPs can be found in Tables S11 and S12 for ADHD and ASD.

\[ \log_{10}(P) \]

| Chromosome | ADHD - LT-FH++ |
|------------|----------------|
| 1          | 2              |
| 3          | 4              |
| 5          | 6              |
| 7          | 8              |
| 9          | 10             |
| 11         | 12             |
| 13         | 14             |
| 15         | 16             |
| 17         | 18             |
| 19         | 20             |
| 21         | 22             |

Discussion

Several large genetic datasets with linked electronic health registries (EHRs) have emerged in recent years, e.g., the UK Biobank data,\(^{44}\) the iP-SYCH data,\(^{33}\) FinnGen, deCODE, and many more. As more genetic data is linked to EHRs, it is essential to develop statistical methods that make best use of all this information to decipher the genetics of common diseases. Here, we present a new and scalable method LT-FH++ for improving power in GWASs when family history and an age-of-onset distribution is available, which is typically the case in EHRs. We demonstrated the feasibility and relevance of the approach by using both simulations and real data applications. Using simulated case-control outcomes with a prevalence of 5% and 10%, we observed power gains of up to 18% compared to LT-FH and up to 61% compared to standard case-control status. We found that LT-FH++ provided the largest relative improvements when cases were ascertained (such that in-sample case-control ratio becomes larger than prevalence) and when prevalence was high. As age-of-onset information allows us to estimate individual liabilities for cases, it makes sense that the largest relative power gains for LT-FH++ are observed when the sample prevalence is high or when the prevalence in the family history is high. Furthermore, LT-FH++ can be applied to individuals with partial or missing family information, as well as individuals for which age and age-of-onset information was missing.

We acknowledge that not everyone has access to the same level of detailed health register data (e.g., Danish registries) or other electronic health records. Therefore, we would like to point out that it is not a requirement to estimate prevalence curves in the population that you are performing the analysis in. In some instances, prevalence rates can be found in publications or from public sites such as statistikbanken or the Office for National Statistics (UK). In practice, prevalence rates may have to be
Figure 6. The $\chi^2$ statistics from the GWAS of ADHD for each of the three methods (LT-FH++, LT-FH, and case-control GWAS) plotted against each other. The dots correspond to LD-clumped SNPs that have a p value below $5 \times 10^{-6}$ in the largest published meta-analysis and present in the iPSYCH cohort (see material and methods for details). The blue line indicates the linear regression line between two methods and the black line indicates the identity line. The slopes of the regression lines are not significantly different from one for any pair of methods.

approximated with external populations and subsequently used to assign the personalized thresholds in the internal population provided information such as sex, age of onset, and birth year is available in the internal and external data.

We applied LT-FH++ to study mortality in UK Biobank and four common psychiatric disorders in iPSYCH, all prevalent outcomes for which we had both family history available as well as age-of-onset distributions. This includes age, age of onset (for cases), cohort effects, and sex for both the genotyped individuals and family members. We also had access to public data for mortality incidence rates by age, sex, and birth year for England and Wales from 1840s to the present day. We compiled similar information for the four psychiatric disorders by the full Danish register data (see material and methods). For mortality in the UK Biobank data, we found ten independent associations when applying LT-FH++, compared to two with LT-FH and none with the case-control status. This result further underlines the importance of including other information in GWASs. The power increase of LT-FH over case-control status highlights the importance of family history, and the power increase of LT-FH++ over LT-FH highlights the importance of accounting for age of onset. The most significant association was found in APOE, which also harbored the only significant association in a recent survival model (frailty model) GWAS of mortality in the UK Biobank data. Most of the identified associations were in or near well-known disease-related genes and were largely concordant with the genome-wide associations found by Pilling et al. when performing a GWAS of combined mothers’ and fathers’ attained age.

We further applied LT-FH++ to the four common psychiatric disorders in the iPSYCH data. Combined, we found ten independent genome-wide significant associations with LT-FH++, compared to eight for LT-FH and case-control status. Compared to mortality, the observed power gain for the iPSYCH disorders was small despite having access to more information per individual. The discrepancy in performance when applied to the mortality in the UK Biobank and four common psychiatric disorders may have several reasons. First, case-control, LT-FH, and LT-FH++ performed similarly for each of the four common psychiatric disorders, and in the simulations, we saw a relative power increase when cases were ascertained through downsampling of controls; however, due to the lower overall sample size, the absolute power to detect causal SNPs also decreased significantly with sample size. We suspect a similar situation might be happening in the iPSYCH data. Second, because simulations showed the power improvement was larger when prevalence was higher and cases were ascertained, the difference may be explained by the prevalence differences. Death is a guarantee, while psychiatric disorders are not. Prevalence rates were far lower for the psychiatric disorders compared to mortality (see Tables 1 and 2), suggesting that less could be gained by accounting for family history and age of onset. Third, it is possible that the multivariate liability threshold model (underlying LT-FH and LT-FH++) may better fit mortality than psychiatric disorders. More specifically, the model makes several key assumptions. First, both LT-FH and LT-FH++ assumes that the heritability is known and that there is no environmental covariance between family members. In practice, one can often estimate the heritability in the sample or rely on published estimates. Second, it assumes that the population disease prevalence is known and (if relevant) provided for subgroups defined by age, birth year, and sex. However, simulations using LT-FH and LT-FH++ indicate that it is relatively robust to misspecification of these parameters. Third, the model assumes that the genetic architecture of the disease or trait in question does not vary by age of diagnosis, birth year, or differ between sexes. Some research suggests that this assumption is reasonable for many outcomes, including the four psychiatric disorders analyzed here, but these will generally not hold in practice. We note that case-control GWASs also assume this unless the analysis is stratified by these
subgroups. Fourth, LT-FH++ assumes that the threshold always decreases with age. The intuition behind this is that the disease prevalence is the cumulative incidence, which by definition always increases with age, and the threshold is the upper quantile of the inverse standard normal at the age-specific prevalence. An individual then only becomes a case if their liability becomes larger than the prevalence threshold, as it decreases with time. A consequence of this assumption is that early-onset cases generally have higher disease liabilities than late-onset cases, which is also the expectation in survival model analysis if the hazard rate is positively correlated with the genetic risk. The correlation between genetic risk and earlier age of onset has been observed for several common diseases, e.g., Alzheimer disease (MIM: 104300), coronary artery disease (MIM: 608320), and prostate cancer (MIM: 176807). However, if the age of onset for a given disease is not heritable, or if the genetic correlation between the age of onset and disease outcome is weak, then we do not expect LT-FH++ to improve statistical power for identifying genetic variants. Indeed, this might be one possible explanation for why we do not observe improvements in power when applying LT-FH++ to iPSYCH data, although we note that polygenic risk scores have been found to contribute to hazard rates for psychiatric disorders in the iPSYCH data.

Conceptually, LT-FH++ combines two methods into one to improve power in genetic analyses, namely LT-FH, which is based on the liability threshold model and incorporates family history, and survival analysis, which can account for age and changes in prevalence over time and is routinely used to model time-to-event data. With family history and age-of-onset information available, we believe LT-FH++ will be an attractive method for improving power in many different genetic analyses, including GWASs and heritability analyses and for polygenic risk scores.

As more genetic datasets with linked health records and family information become available, e.g., in large national biobank projects, we expect the value of statistical methods that efficiently distill family history and individual health information into biological insight will only increase.

Data and code availability

iPSYCH is approved by the Danish Scientific Ethics Committee, the Danish Health Data Authority, the Danish Data Protection Agency, Statistics Denmark, and the Danish Neonatal Screening Biobank Steering Committee. UK Biobank received ethical approval from the NHS National Research Ethics Service North West (11/NW/0382). The present analyses were conducted under UK Biobank data application number 58024. The code used for LT-FH++ has been implemented into an R package, and it is available at https://github.com/EmilMiP/LTFHPlus. We have also reimplemented LT-FH in the package, where we utilize the Gibbs sampler to efficiently estimate the genetic liabilities, keeping the same input format as the original implementation. Summary statistics can be downloaded from https://drive.google.com/drive/folders/13Ityy7KuoXKUSyYu4Cf0ntSw5WOTLmD and mortality rates were found on https://www.ons.gov.uk/.

Supplemental information

Supplemental information can be found online at https://doi.org/10.1016/j.ajhg.2022.01.009.

Acknowledgments

We would like to thank Margaux Hujoel for useful discussions and allowing us to use the LT-FH++ name. We would like to thank Mark Daly for useful advice and helpful comments. E.P. and B.J.V. were supported by the Danish National Research Foundation (Niels Bohr Professorship to Prof. John McGrath). We also acknowledge the Lundbeck Foundation Initiative for Integrative Psychiatric Research, iPSYCH (R102-A9118, R155-2014-1724, and R248-2017-2003). B.J.V. was also supported by a Lundbeck Foundation fellowship (R335-2019-2339). High-performance computer capacity for handling and statistical analysis of iPSYCH data on the GenomeDK HPC facility was provided by the Center for Genomics and Personalized Medicine and the Centre for Integrative Sequencing, iSEQ, Aarhus University, Denmark (grant to A.D.B.). This research has been conducted with the UK Biobank Resource under application number 58024.

Declaratıon of interests

J.C. has received honoraria for serving on the Scientific Advisory Board of Union Chimique Belge (UCB) Nordic and Eisai AB and for giving lectures for UCB Nordic and Eisai as well as travel funds from UCB Nordic and funding by the Novo Nordisk Foundation (grant number: NNFI600C0019126), the Central Denmark Region, and the Danish Epilepsy Association.

Received: July 16, 2021
Accepted: January 7, 2022
Published: February 8, 2022

Web resources

GenomeDK, https://genome.au.dk/
LTFHPlus, https://github.com/EmilMiP/LTFHPlus
Office of National Statistics, https://www.ons.gov.uk/
Statistikbanken, https://www.statistikbanken.dk/

References

1. Nielsen, J.B., Thorolfsdottir, R.B., Fritsche, L.G., Zhou, W., Skov, M.W., Graham, S.E., Herron, T.J., McCarthy, S., Schmidt, E.M., Sveinbjornsson, G., et al. (2018). Biobank-driven genomic discovery yields new insight into atrial fibrillation biology. Nat. Genet. 50, 1234–1239.
2. Wuttke, M., Li, Y., Li, M., Sieber, K.B., Feitosa, M.F., Gorski, M., Tin, A., Wang, L., Chu, A.Y., Hoppmann, A., et al. (2019). A catalog of genetic loci associated with kidney function from analyses of a million individuals. Nat. Genet. 51, 957–972.
3. Mahajan, A., Taliun, D., Thurmer, M., Robertson, N.R., Torres, J.M., Rayner, N.W., Payne, A.J., Steinthorsdottir, V., Scott, R.A., Grarup, N., et al. (2018). Fine-mapping type 2 diabetes loci to single-variant resolution using high-density imputation and islet-specific epigenome maps. Nat. Genet. 50, 1505–1513.
4. Siewert, K.M., and Voight, B.F. (2018). Bivariate Genome-Wide Association Scan Identifies 6 Novel Loci Associated With Lipid
33. Pedersen, C.B., Bybjerg-Grauholm, J., Pedersen, M.G., Grove, J., Agerbo, E., Bækkvad-Hansen, M., Poulsen, J.B., Hansen, C.S., McGrath, J.J., Als, T.D., et al. (2018). The iPSYCH2012 case-cohort sample: new directions for unravelling genetic and environmental architectures of severe mental disorders. Mol. Psychiatry 23, 6–14.

34. Cox, D.R., and Oakes, D. (1984). Analysis of Survival Data (CRC Press).

35. Ojavee, S.E., Kousathanas, A., Trejo Banos, D., Orliac, E.J., Patxot, M., Läll, K., Mägi, R., Fischer, K., Kutilik, Z., and Robinson, M.R. (2021). Genomic architecture and prediction of censored time-to-event phenotypes with a Bayesian genome-wide analysis. Nat. Commun. 12, 2337.

36. Li, R., Chang, C., Jusotse, J.M., Tanigawa, Y., Qiang, J., Hastie, T., Rivas, M.A., and Tibshirani, R. (2020). Fast Lasso method for large-scale and ultrahigh-dimensional Cox model with applications to UK Biobank. Biostatistics, kxa038.

37. Eddelbuettel, D., and Francois, R. (2011). Rcpp: Seamless R and C++ Integration. J. Stat. Softw. 40, 1–18.

38. Loh, P.-R., Tucker, G., Bulik-Sullivan, B.K., Vilhjalmsson, B.J., Finucane, H.K., Salem, R.M., Chasman, D.I., Ridker, P.M., Neale, B.M., Berger, B., et al. (2015). Efficient Bayesian mixed-model analysis increases association power in large cohorts. Nat. Genet. 47, 284–290.

39. Kragh Andersen, P., Pohar Perme, M., van Houwelingen, H.C., Cook, R.J., Joly, P., Martinussen, T., Taylor, J.M.G., Abrahamowicz, M., and Therneau, T.M. (2021). Analysis of time-to-event for observational studies: Guidance to the use of intensity models. Stat. Med. 40, 185–211.

40. Wilhelm, S. (2015). Gibbs sampler for the truncated multivariate normal distribution. https://cran.r-project.org/web/packages/tmvtnorm/vignettes/GibbsSampler.pdf.

41. Pedersen, C.B. (2011). The Danish Civil Registration System. Scand. J. Public Health 39 (7, Suppl), 22–25.

42. Mors, O., Perto, G.P., and Mortensen, P.B. (2011). The Danish Psychiatric Central Research Register. Scand. J. Public Health 39 (7, Suppl), 54–57.

43. Hansen, S.N., Overgaard, M., Andersen, P.K., and Parner, E.T. (2017). Estimating a population cumulative incidence under calendar time trends. BMC Med. Res. Methodol. 17, 7.

44. Bycroft, C., Freeman, C., Petkova, D., Band, G., Elliott, L.T., Sharp, K., Motyer, A., Vukcevic, D., Delaneau, O., O’Connell, J., et al. (2018). The UK Biobank resource with deep phenotyping and genomic data. Nature 562, 203–209.

45. Bybjerg-Grauholm, J., Pedersen, C.B., Bækkvad-Hansen, M., Pedersen, M.G., Ademansen, D., Hansen, C.S., Agerbo, E., Grove, J., Als, T.D., Schork, A.J., et al. (2020). The iPSYCH2015 Case-Cohort sample: updated directions for unravelling genetic and environmental architectures of severe mental disorders. medRxiv. https://doi.org/10.1101/2020.11.30.20237768.

46. Lam, M., Awasthi, S., Watson, H.J., Goldstein, J., Panagiotaropoulos, G., Trubetskoy, V., Karlsson, R., Frei, O., Fan, C.C., De Witte, W., et al. (2020). RICOPILI: Rapid Imputation for COnsortias Pipline. Bioinformatics 36, 930–933.

47. Privé, F., Luu, K., Blum, M.G.B., McGrath, J.J., and Vilhjalmsson, B.J. (2020). Efficient toolkit implementing best practices for principal component analysis of population genetic data. Bioinformatics 36, 4449–4457.

48. Brikell, I., Kuja-Halkola, R., and Larsson, H. (2015). Heritability of attention-deficit hyperactivity disorder in adults. Am. J. Med. Genet. B. Neuropsychiatr. Genet. 168, 406–413.

49. Sandin, S., Lichtenstein, P., Kuja-Halkola, R., Hultman, C., Larsson, H., and Reichenberg, A. (2017). The Heritability of Autism Spectrum Disorder. JAMA 318, 1182–1184.

50. Fernandez-Pujals, A.M., Adams, M.J., Thomson, P., McKechnie, A.G., Blackwood, D.H., Smith, B.H., Dominiczak, A.F., Morris, A.D., Matthews, K., Campbell, A., et al. (2015). Epidemiology and Heritability of Major Depressive Disorder, Stratified by Age of Onset, Sex, and Illness Course in Generation Scotland: Scottish Family Health Study (GSSFHS). PLoS ONE 10, e0142197.

51. Hilker, R., Helenius, D., Fagerlund, B., Skyttea, A., Christensen, K., Werne, T.M., Nordentoft, M., and Glenthoj, B. (2018). Heritability of Schizophrenia and Schizophrenia Spectrum Based on the Nationwide Danish Twin Register. Biol. Psychiatry 83, 492–498.

52. Wray, N.R., Ripke, S., Mattheisen, M., Trzaskowski, M., Byrne, E.M., Abdellaouiti, A., Adams, M.J., Agerbo, E., Air, T.M., Andlauer, T.M.E., et al. (2018). Genome-wide association analyses identify 44 risk variants and refine the genetic architecture of major depression. Nat. Genet. 50, 668–681.

53. Falconer, D.S. (1965). The inheritance of liability to certain diseases, estimated from the incidence among relatives. Ann. Hum. Genet. 29, 51–76.

54. Fisher, R.A. (1918). The correlation between relatives on the supposition of mendelian inheritance. Trans. R. Soc. Edinb. 52, 899, 438.

55. Lee, S.H., Wray, N.R., Goddard, M.E., and Visscher, P.M. (2011). Estimating missing heritability for disease from genome-wide association studies. Am. J. Hum. Genet. 88, 294–305.

56. Thygesen, L.C., Daanes, C., Thaulow, I., and Brønnum-Hansen, H. (2011). Introduction to Danish (nationwide) registers on health and social issues: structure, access, legislation, and archiving. Scand. J. Public Health 39 (7, Suppl), 12–16.

57. Hayeck, T.J., Loh, P.-R., Pollack, S., Gusev, A., Patterson, N., Zaitlen, N.A., and Price, A.L. (2017). Mixed Model Association with Family-Biased Case-Control Ascertainment. Am. J. Hum. Genet. 100, 31–39.

58. Hayeck, T.J., Zaitlen, N.A., Loh, P.-R., Vilhjalmsson, B.S., Pollack, S., Gusev, A., Yang, J., Chen, G.-B., Goddard, M.E., Visscher, P.M., et al. (2015). Mixed model with correction for case-control ascertainment increases association power. Am. J. Hum. Genet. 96, 720–730.

59. So, H.-C., and Sham, P.C. (2010). A unifying framework for evaluating the predictive power of genetic variants based on the level of heritability explained. PLoS Genet. 6, e1001230.

60. Schächter, E., Faure-Delanef, L., Guénolé, F., Rouger, H., Froguel, P., Lesueur-Ginot, L., and Cohen, D. (1994). Genetic associations with human longevity at the APOE and ACE loci. Nat. Genet. 6, 29–32.

61. Yang, J., Ferreira, T., Morris, A.P., Medland, S.E., Madden, P.A.F., Heath, A.C., Martin, N.G., Montgomery, G.W., Weedon, M.N., Loos, R.J., et al. (2012). Conditional and joint multiple-SNP analysis of GWAS summary statistics identifies additional variants influencing complex traits. Nat. Genet. 44, 369–375, S1–S3.

62. Chen, M.-H., Raffield, L.M., Mousas, A., Sakaue, S., Huffman, J.E., Moscati, A., Trivedi, B., Jiang, T., Akbari, P., Vuckovic, D. et al. (2020). Trans-ethnic and Ancestry-Specific Blood-Cell Genetics in 746,667 Individuals from 5 Global Populations. Cell 182, 1198–1213.e14.
63. Tsoi, L.C., Spain, S.L., Knight, J., Ellinghaus, E., Stuart, P.E., Capon, F., Ding, J., Li, Y., Tejasvi, T., Gudjonsson, J.E., et al. (2012). Identification of 15 new psoriasis susceptibility loci highlights the role of innate immunity. Nat. Genet. 44, 1341–1348.

64. Jones, S.E., Lane, J.M., Wood, A.R., van Hees, V.T., Tyrrell, J., Beaumont, R.N., Jeffries, A.R., Dashi, H.S., Hillsdon, M., Ruth, K.S., et al. (2019). Genome-wide association analyses of chronotype in 697,828 individuals provides insights into circadian rhythms. Nat. Commun. 10, 343.

65. Chow, R.D., Majety, M., and Chen, S. (2021). The aging transcriptome and cellular landscape of the human lung in relation to SARS-CoV-2. Nat. Commun. 12, 4.

66. Zhang, Q., Marioni, R.E., Robinson, M.R., Higham, J., Sproul, D., Wray, N.R., Deary, I.J., McRae, A.F., and Visscher, P.M. (2018). Genotype effects contribute to variation in longitudinal methyolate patterns in older people. Genome Med. 10, 75.

67. Savage, J.E., Jansen, P.R., Stringer, S., Watanabe, K., Bryois, J., de Leeuw, C.A., Nagel, M., Awasthi, S., Barr, P.B., Coleman, J.R.I., et al. (2018). Genome-wide association meta-analysis in 269,867 individuals identifies new genetic and functional links to intelligence. Nat. Genet. 50, 912–919.

68. Lee, J.J., Wedow, R., Okbay, A., Kong, E., Maghzian, O., Zacher, M., Nguyen-Viet, T.A., Bowers, P., Sidorenko, J., Karlsson Linner, R., et al. (2018). Gene discovery and polygenic prediction from a genome-wide association study of educational attainment in 1.1 million individuals. Nat. Genet. 50, 1112–1121.

69. Pilling, L.C., Kuo, C.-L., Sicinski, K., Tomaszkaite, J., Kuchel, G.A., Harries, L.W., Herd, P., Wallace, R., Ferrucci, L., and Melzer, D. (2017). Human longevity: 25 genetic loci associated in 389,166 UK biobank participants. Aging (Albany N.Y.) 9, 2504–2520.

70. Martin, J., Khramtsova, E.A., Goleva, S.B., Biokland, G.A.M., Traglia, M., Walters, R.K., Hübel, C., Coleman, J.R.I., Breen, G., Borglum, A.D., et al. (2020). Examining sex-differentiated genetic effects across neuropsychiatric and behavioral traits. Biol. Psychiatry 89, 1127–1137.

71. Traglia, M., Breslin, D., Gusev, A., Adviento, B., Park, D.S., Melford, J.A., Zaitlen, N., and Weiss, L.A. (2017). Genetic Mechanisms Leading to Sex Differences Across Common Diseases and Anthropometric Traits. Genetics 205, 979–992.

72. Zhang, Q., Sidorenko, J., Couvy-Duchesne, B., Marioni, R.E., Wright, M.J., Goate, A.M., Marcara, E., Huang, K.-L., Porter, T., Laws, S.M., et al. (2020). Risk prediction of late-onset Alzheimer’s disease implies an oligogenic architecture. Nat. Commun. 11, 4799.

73. Mars, N., Koskela, J.T., Ripatti, P., Kiiskinen, T.T.J., Havulinna, A.S., Lindbohm, J.V., Ahola-Olli, A., Kurki, M., Karjalainen, J., Falta, P., et al. (2020). Polygenic and clinical risk scores and their impact on age at onset and prediction of cardiometabolic diseases and common cancers. Nat. Med. 26, 549–557.

74. Musliner, K.L., Krebs, M.D., Albíñana, C., Vilhjálmsson, B., Ågerbo, E., Zandi, P.P., Hougaard, D.M., Nordenfelt, M., Borglum, A.D., Werge, T., et al. (2020). Polygenic Risk and Progression to Bipolar or Psychotic Disorders Among Individuals Diagnosed With Unipolar Depression in Early Life. Am. J. Psychiatry 177, 936–943.

75. Ågerbo, E., Trabjerg, B.B., Borglum, A.D., Schork, A.J., Vilhjálmsson, B.J., Pedersen, C.B., Hakulinen, C., Albíñana, C., Hougaard, D.M., Grove, J., et al. (2021). Risk of Early-Onset Depression Associated With Polygenic Liability, Parental Psychiatric History, and Socioeconomic Status. JAMA Psychiatry 78, 387–397.

76. Ågerbo, E., Sullivan, P.F., Vilhjálmsson, B.J., Pedersen, C.B., Mors, O., Borglum, A.D., Hougaard, D.M., Hollegaard, M.V., Meier, S., Mattheisen, M., et al. (2015). Polygenic Risk Score, Parental Socioeconomic Status, Family History of Psychiatric Disorders, and the Risk for Schizophrenia: A Danish Population-Based Study and Meta-analysis. JAMA Psychiatry 72, 635–641.

77. Lencz, T., Backenroth, D., Green, A., Weissbröd, O., Zuk, O., and Carmi, S. (2020). Utility of polygenic embryo screening for disease depends on the selection strategy. bioRxiv. https://doi.org/10.1101/2020.11.05.370478.

78. Hujoel, M.L.A., Loh, P.-R., Neale, B.M., and Price, A.L. (2021). Incorporating family history of disease improves polygenic risk scores in diverse populations. bioRxiv. https://doi.org/10.1101/2021.04.15.439975.