Clinical Application of Indocyanine Green Fluorescence Imaging in the Resection of Hepatoblastoma: A Single Institution’s Experiences

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Purpose: Indocyanine green (ICG) fluorescence imaging is becoming increasingly popular in adult oncologic surgery, but remains relatively uncommon in pediatric oncologic surgery. Herein, we report our experience with the use of ICG fluorescence imaging in the resection of hepatoblastoma (HB).

Patients and Methods: Hepatoblastoma patients who underwent liver resection with ICG fluorescence imaging between January 2020 and March 2021 were included in this study. Patients’ demographic data, clinical information, and detailed information of the use of ICG fluorescence imaging were retrospectively reviewed.

Results: Sixteen HB patients underwent ICG fluorescence imaging-guided liver resection. There were 11 males and 5 females, age ranged from 8 to 134 months. The initial alpha-fetoprotein ranged from 436 to 528,390 ng/ml. There were one pre-treatment extent of tumor stage I, nine stage II, four stage III, and two stage IV. Three patients underwent up-front hepatectomy, 13 patients received 2–8 cycles of platinum-based neoadjuvant chemotherapy and underwent delayed hepatectomy. ICG (0.5 mg/kg) was given intravenously 48–72 h prior to surgery. The operative time ranged from 180 to 400 min. All patients achieved negative surgical margins. In two patients, ICG identify additional lesions which were not detected in preoperative imaging.

Conclusion: ICG fluorescence imaging is useful in the resection of HB and may detect small lesions not shown in preoperative imaging.

Keywords: indocyanine green, fluorescence imaging, hepatoblastoma, children, hepatectomy

INTRODUCTION

Hepatoblastoma (HB) is the most common primary pediatric liver malignancy, which accounts for approximately 1% of all pediatric malignancies. The estimated incidence of HB is about 1–1.5 cases per million per year in children younger than 15 years (1–3). The management of HB is multi-disciplinary, consisting of chemotherapy and surgery. Complete surgical resection is key to
successful treatment of HB (4, 5). During the past several decades, major advances had been achieved in the surgical management of HB. Extended liver resection for pre-treatment extent (PRETEXT) III and PRETEXT IV patients has been investigated, with encouraging results. Liver transplantation for advanced HB is gaining more acceptance, with a 5-year overall survival rate of over 70% (6).

In recent years, indocyanine green (ICG) fluorescence imaging-guided liver resection has become increasingly popular (7, 8). Several studies have described the use of ICG in the resection of HB, with successful results. After intravenously injection, ICG is secreted into bile and washed out in hours within normal liver, whereas ICG secretion is inhibited in malignant liver tumor and metastasis focus. Based on this characteristic, ICG fluorescence imaging navigation becomes an effective technology in hepatobiliary surgery. It can be used to determine tumor margin, detect satellite lesions, and other many applications. Hereby, we summarize our experience with the use of ICG fluorescence imaging-guided HB resection.

PATIENTS AND METHODS

HB patients who underwent hepatectomy with the use of ICG fluorescence imaging between January 2020 and March 2021 were included in this study. All patients underwent computed tomography (CT) scanning, and a PRETEXT stage was assigned. Patients underwent either up-front hepatectomy or ultrasound-guided percutaneous biopsy with neoadjuvant chemotherapy based on clinical findings, especially the PRETEXT stage. The diagnosis of HB was established based on pathologic findings. Patients’ demographic, disease characteristics, surgical finding and ICG fluorescence imaging, pathologic diagnoses, and clinical outcome were reviewed and analyzed.

This study was approved by the Guangzhou Woman and Children’s Medical Center’s Institutional Review Board. All patients’ data were anonymized and deidentified prior to analyses. ICG (0.5 mg/kg) was given intravenously 48–72 h prior to surgery, which was in accordant with the recommendations in consensus guidelines for the use of fluorescence imaging in hepatobiliary surgery (9). Informed consent was obtained from the patients’ parents before ICG injection.

During the surgery, a near-infrared light camera was used to detect ICG fluorescence. Resection lines can be marked based on real-time ICG fluorescence imaging (Figure 1), and small satellite lesions can also detected. During and after the completion of liver resection, the resection plane and surgical margin were re-examined with ICG fluorescence imaging.

Statistical Analyses
The statistical analyses were performed using SAS software, version 9.4 (SAS Institute, Inc., Cary, NC, USA). The continuous variables are presented as the medians (ranges), while the categorical variables are presented as numbers (percentages).

RESULTS
Sixteen patients underwent ICG fluorescence imaging-guided hepatectomy (Table 1). There were 11 (68.8%) males and 5 (31.2%) females. The median age was 15 months (8–134 months). There were 1 (6.2%) cases of PRETEXT stage I, 9 (56.2%) cases of stage II, 4 (25.0%) cases of stage III, and 2 (12.5%) cases of stage IV. The median tumor diameter based on the preoperative CT images was 8 cm (2.9–11.0 cm). The median preoperative alpha-fetoprotein (AFP) value was 97,983.5 ng/ml (436–528,390 ng/ml). Three (18.7%) patients underwent up-front hepatectomy, and 13 (81.2%) received 2–8 cycles of platinum-based neoadjuvant chemotherapy prior to hepatectomy.

During surgery, the boundary between normal liver tissue and HB was presented under the real-time ICG fluorescence imaging in 16 patients (Figure 2). Multiple small lesions were detected by intraoperative ICG fluorescence imaging in two patients (patient 1 and patient 3, Table 1). These lesions were not found in preoperative contrast-enhanced CT with iopromide and contrast-enhanced MRI with Gd-DTPA-BMA, and were confirmed as HB tissue pathologically (Figure 3).
### TABLE 1 | Patients’ demographic and clinical information.

| Gender | Age (month) | PRETEXT Neoadjuvant chemotherapy (cycles) | POSTTEXT Tumor location | Tumor size (cm) | Operative time (minutes) | Volume of red blood cells transfusion (u) | Pathologic subtype |
|--------|-------------|------------------------------------------|-------------------------|----------------|--------------------------|------------------------------------------|-------------------|
| 1      | M           | 9                                       | III                     | S5, S6         | 6.5 x 6.4 x 8.5          | 265                                      | 1                 | EM                |
| 2      | M           | 8                                       | II                      | S5, S8         | 2.9 x 2.8 x 2.5          | 300                                      | 1                 | MEM               |
| 3      | F           | 12                                      | II                      | S3, S4         | 4.6 x 3.6 x 4.3          | 235                                      | 0.9               | MEM               |
| 4      | M           | 48                                      | II                      | S5, S6, S7     | 8.6 x 7.2 x 11           | 400                                      | 2                 | MEM               |
| 5      | M           | 12                                      | II                      | S5, S6         | 8.1 x 5.9 x 7.8          | 180                                      | 0                 | EM                |
| 6      | F           | 29                                      | III                     | S4, S5, S6     | –                        | 240                                      | 1                 | EV                |
| 7      | M           | 32                                      | II                      | S7, S8         | 6.9 x 5.9 x 8.4          | 295                                      | 0.8               | MEM               |
| 8      | M           | 16                                      | II                      | S4             | 3.1 x 2.9 x 2.0          | 260                                      | 0.2               | EV                |
| 9      | M           | 15                                      | III                     | S4, S6, S7, S8 | 6.8 x 5.9 x 9.3          | 360                                      | 1                 | MEM               |
| 10     | M           | 11                                      | I                       | –              | 4.9 x 4.6 x 4.1          | 192                                      | 0.5               | EM                |
| 11     | M           | 19                                      | II                      | S5, S6, S7, S8 | 7.1 x 5.1 x 9.3          | 340                                      | 0.8               | EM                |
| 12     | M           | 23                                      | IV                      | S2, S3, S4, S5, S6, S8 | 7.3 x 4.0 x 4.9          | 265                                      | 1.5               | EV                |
| 13     | F           | 39                                      | III                     | S5, S6         | 10.1 x 6.3 x 10.6        | 120                                      | 0                 | EV                |
| 14     | F           | 14                                      | III                     | S2, S3, S4, S7 | 4.6 x 3.9 x 2.7          | 205                                      | 1                 | EV                |
| 15     | M           | 132                                     | IV                      | S5, S6, S7, S8 | 8.0 x 4.1 x 4.8          | 279                                      | 1                 | MEM               |
| 16     | F           | 14                                      | III                     | S4, S5, S7, S8 | 4.2 x 2.7 x 3.0          | 270                                      | 1                 | EV                |

**PRETEXT, pre-treatment extent of disease system; POSTTEXT, post-treatment extent of disease system; M, male; F, female; EM, epithelial mixed; MEM, Mixed epithelial and mesenchymal; EV, Epithelial variants.**
ICG fluorescence imaging was used to re-examine the resection plane in situ and ex vivo, none of them show fluorescence (Figure 4). This was consistent with the pathologic findings of negative surgical margins in 16 patients.

The median operation time was 265 min (80–400 min). All patients achieved negative surgical margins. There were 6 (37.5%) cases of epithelial variants type, 6 (37.5%) cases of mixed epithelial and mesenchymal type, and 4 (25.0%) cases of epithelial mixed type. All patients had an uneventful recovery.

**DISCUSSION**

Our study showed that intraoperative ICG fluorescence imaging could be a valuable tool for HB surgery. It helps to determine the resection plane and assure a negative resection margin. Furthermore, it can detect small lesions not shown in preoperative imaging. Our results were consistent with several other studies, showing that ICG aids in determining the resection line and identifying residual tumors (10–12).

ICG has been used in clinic for more than 50 years, and the overall adverse effect rate is less than 0.01% (13). ICG is relatively safe, the lethal doses of ICG is up to 50–80 mg/kg, it is essentially nontoxic when injected at standard doses of less than 2 mg/kg (14). No patients have any adverse effect after ICG injection in our cohort.

ICG fluorescence imaging is widely used in surgical navigation, but its clinical application in pediatric surgery is still in its infancy (15). The use of preoperatively injected intravenous ICG and near-infrared imaging equipment has emerged as a novel therapy in cases of HB and metastatic resections (16–18). As is well known, ICG is retained in the tumor tissue much longer than in the normal liver parenchyma (19). By this principle, ICG can be used for imaging both primary HB and metastases. Using this specialized equipment, sites of primary HB and metastatic locations are able to be detected with green fluorescence (4). This modality can be used to help achieve a negative resection status, safely remove tumors from closely underlying vascular structures, as well as assess the degree of any remaining malignancy (4). The negative margin can be achieved and confirmed through repeatedly imaging the resection plane during hepatectomy, and imagining the resection margin after tumor removal. All 16 patients in our study achieved negative resection margin. In addition to delineate the resection line and assess the degree of any remaining malignancy. The identification of small lesions that were not detected in preoperative CT/MRI scanning under ICG fluorescence imaging was observed in our study, showing that the significant advantage of ICG fluorescence imaging is compared with preoperative imaging examination (20). We identified multiple small lesions in two cases with this method.

ICG fluorescence imaging is also useful in identifying small viable metastatic lung lesions. Kitagawa et al. reported that ICG can detect lung lesions as small as 0.062 mm in diameter, and all of the pathologically positive lesions were clearly fluorescence positive in a study of 10 patients (21).

However, ICG fluorescence imaging has some limitations. One of the limitations of ICG fluorescence imaging is its inability to probe deep tissue. The fluorescence emitted by ICG can only penetrate 5–10 mm of tissue. When the tumor goes beyond this depth, its fluorescence is undetectable on the surface (22, 23). Souzaki et al. previously reported that ICG failed to detect a tumor 12 mm from the lung surface (10). Additionally, although ICG is highly sensitive to tumors, its specificity is rather low. Cotoh’s study reported that strong fluorescence also can be displayed in the surface of focal cirrhosis and hepatic nodular hyperplasia, the false positive rate of these lesions can reach 40%–50% (24). Dysfunction liver can present fluorescence as well. Some non-tumor tissues
showing fluorescence are hard to avoid. When intraoperative ICG fluorescence imaging was significantly different from the preoperative imaging results, the possibility of false positive must be considered, and intraoperative frozen-section can be helpful in determining the nature of suspicious nodule. Further study is needed to find out how to reduce the false positive rate such as by developing high-performance near-infrared nanocomplexes formulated with ICG with high selectivity for tumors (25). This is a small case-series report, and further studies with a larger number of patients will be needed to validate the ICG fluorescence imaging technique.

CONCLUSION

In conclusion, ICG fluorescence imaging is useful in the resection of HB. This technique effectively improves the surgical efficiency and helps achieve complete resection of tumors.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/Supplementary Material, further inquiries can be directed to the corresponding author/s.

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ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Ethics Committee, Guangzhou Women and Children’s Medical Center. Written informed consent was obtained from the individual(s), and minor(s)’ legal guardian/next of kin, for the publication of any potentially identifiable images or data included in this article.

AUTHOR CONTRIBUTIONS

TY and YZ conceptualized and designed the study, YS and MZ drafted the initial manuscript, TY and YZ reviewed and revised the manuscript. YS, MZ, JL, CH, TT, JY, and JP collected data, carried out the initial analyses, and reviewed and revised the manuscript. TY coordinated and supervised data collection, and critically reviewed the manuscript for important intellectual content. All authors contributed to the article and approved the submitted version.

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