Research Article

The Role of the NMDA Receptor in the Anticonvulsant Effect of Ellagic Acid in Pentylentetrazole-Induced Seizures in Male Mice

Mohammad Rahimi-Madiseh, Zahra Lorigooini, Shakiba Nasiri Boroujeni, Marziyeh Taji, and Hossein Amini-Khoei

Medical Plants Research Center, Basic Health Sciences Institute, Shahrekord University of Medical Sciences, Shahrekord, Iran

Correspondence should be addressed to Hossein Amini-Khoei; aminikhoyi@gmail.com

Received 24 April 2021; Revised 15 February 2022; Accepted 13 April 2022; Published 11 May 2022

Academic Editor: Giuseppe Biagini

Copyright © 2022 Mohammad Rahimi-Madiseh et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Background and Aim. Epilepsy is the most common neurological disorder after stroke. Ellagic acid (EA) has been shown to possess neuroprotective effects. The N-methyl-D-aspartate receptor (NMDA-R) is involved in the pathophysiology of seizure. We aimed to evaluate the possible involvement of NMDA-R in the anticonvulsant effect of EA in pentylenetetrazole-(PTZ-) induced seizures in male mice.

Methods. In this experimental study, 64 mice were divided into 8 groups and received the following: normal saline; EA at doses of 6.25, 12.5, and 25 mg/kg; NMDA agonist at a dose of 75 mg/kg; NMDA antagonist (ketamine) at a dose of 0.5 mg/kg; an effective dose of EA plus NMDA agonist; and a subeffective dose of EA plus ketamine. We induced seizure using intravenous administration of PTZ. 60 minutes before induction of seizure, drugs were administrated. Duration lasts to seizure-induced was measured. Finally, the gene expression of NMDA receptor subunits (Nr2a and Nr2b) was assessed in the prefrontal cortex.

Results. Results showed that EA increased the seizure threshold and decreased the expression of Nr2a and Nr2b. We determined that ketamine potentiated and NMDA attenuated the effects of subeffective and effective doses of EA. Conclusion. EA probably via attenuation of the NMDA-R pathway possesses an anticonvulsant effect in PTZ-induced seizure in mice.

1. Introduction

Seizure is a common disease in the world [1]. The seizure occurs when an abnormal electrical activity manifests in the central nervous system (CNS). Various etiologies are involved in the pathophysiology of seizure [2]. According to the researcher’s findings, 30 to 40% of patients are resistant to all treatments and do not respond adequately to the standard anticonvulsants [3]. Despite the introduction of new drugs in recent decades, these are accompanied by several side effects [4]. Studies have shown that changes in gabaergic and glutamatergic synaptic transmission have a pivotal role in seizure development. Seizures occur when the activity of excitatory neurotransmitters (glutamate) increases or the activity of inhibitory neurotransmitters (GABA) decreases [5].

Glutamate is a nonessential dicarboxylic amino acid in various brain structures [6]. As a significant stimulatory neurotransmitter, glutamate participates in several physiological functions such as brain development, synaptic flexibility, memory, and learning [5, 7]. An increase in the glutamate concentration in the synaptic space and extracellular fluid leads to neuron damage [8]. N-Methyl-D-aspartate (NMDA) is one of the ionotropic glutamate receptors. Activation of NMDA receptor causes accumulation of calcium ions in neurons [9, 10]. Studies have shown an increase in the activation and expression of NMDA receptor subunits (Nr2a and Nr2b) was assessed in the prefrontal cortex. Results. Results showed that EA increased the seizure threshold and decreased the expression of Nr2a and Nr2b. We determined that ketamine potentiated and NMDA attenuated the effects of subeffective and effective doses of EA. Conclusion. EA probably via attenuation of the NMDA-R pathway possesses an anticonvulsant effect in PTZ-induced seizure in mice.
of NMDA receptors decreases status epilepticus-induced neuronal cell death [16]. In this regard, researchers have demonstrated that seizure increased the expression of NR2a and NR2b in the brain [17].

Recently, the central concept in seizure management research is finding agents with neuroprotective activities [9, 10]. Ellagic acid (EA), with a molecular weight of 302 g/mol, is a polyphenolic compound with antioxidant properties found in fruits like pomegranate, raspberry, and berry [18]. Previous studies have reported several pharmacological properties for EA, including antibacterial, anti-inflammatory, immunoregulatory, and antitumor effects [19–22]. It has been suggested that EA exhibits a neuroprotective effect and protects the brain from oxidative and inflammatory challenges [11, 23, 24]. Preclinical examinations determined antidepressant-like effects for EA [25, 26].

Considering the role of glutamate and NMDA receptors in the pathophysiology of seizure and several pharmacological effects of EA, especially its neuroprotective effects, in this study, we aimed to evaluate the possible involvement of NMDA-R in the anticonvulsant effect of EA in pentylenetetrazole- (PTZ-) induced seizures in male mice.

2. Material and Methods

2.1. Study Design. 64 male NMRI mice were kept under standard laboratory conditions (12-hour light/dark cycle, 22 ± 2°C, and free access to water and food). All stages of the present experimentation were carried out following the regulations of the university and the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health (Ethics code: IR.SKUMS.REC.1397.312) and Guide for the Care and Use of Laboratory Animals (8th edition, National Academies Press). Full efforts were made to diminish animals’ use and improve their wellbeing. Mice were randomly divided into 8 groups (n = 8). Group 1 received normal saline (considered as a control group), groups 2–4 received EA at doses of 6.25, 12.5, and 25 mg/kg, respectively, group 5 received NMDA agonist at the dose of 7.5 mg/kg, group 6 received NMDA antagonist (ketamine) at the dose of 0.5 mg/kg, group 7 received the effective dose of EA plus NMDA agonist, and group 8 received the subeffective dose of EA plus ketamine. All drugs were administrated intraperitoneally (i.p.). 60 minutes after the treatments, pentylenetetrazole (PTZ) was injected intravenously at an 80 mg/kg dose to induce seizures. The dose and time of drug administrations were chosen based on previous studies and our pilot study [27–29].

2.2. Induction of Seizures by PTZ. In order to inject the PTZ, a needle gauge 30 was attached to the mice’s tail vein. After fixing the mice’s tail, the PTZ (0.5%) was injected at a 1 mL/min rate by a seizure pump. The injection stopped as soon as the clonus of the anterior limb was seen. The minimum dose of PTZ for seizure was considered the dose of seizure threshold. In this method, the seizure threshold was dependent on the PTZ dose and was related to time. PTZ was injected intravenously at a dose of 80 mg/kg 60 minutes after treatments [27].

2.3. Real-Time PCR Analysis for Expression of NMDA Receptors in the Prefrontal Cortex. At the end of the study, animals were sacrificed, the prefrontal cortex was isolated, and the gene expression of NMDA receptor subunits (Nr2a and Nr2b) was examined by real-time PCR. Firstly, total RNA using TRizol reagent (Invitrogen) was extracted from the prefrontal cortex. Alterations in the mRNA levels of genes were determined using qRT-PCR after the reverse transcription of 1 μg of RNA from each sample using the PrimeScript RT reagent kit (Takara Bio, Inc., Otsu, Japan). qRT-PCR was done on a light cycler device (Roche Diagnostics, Mannheim, Germany) using SYBR Premix Ex Taq technology (Takara Bio). Thermal cycling conditions included an initial activation step for 30 s at 95°C afterward 45 cycles, a denaturation step for 5 s at 95°C, and a combined annealing/extension step for 20 s at 60°C. Melting curve analysis was performed to certify whether all primers yielded a single PCR product. The genes and their primers are listed in Table 1. H2afz was used as a house-keeping gene (normalizer), and alterations in the expression of each target mRNA
in comparison with B2m were measured based on the 2-ΔΔCt relative expression formula, as described in our previous publication [30, 31].

2.4. Statistical Analysis. Statistical analysis of data was performed using GraphPad Prism 8 software. One-way ANOVA was used to determine the significant differences between the treatments, and the Tukey post hoc test compared the means. Data were recorded as mean ± standard deviation, and $P < 0.05$ was considered statistically significant.

3. Results

3.1. Effects on the Seizure Threshold. The results showed (Figure 1) that EA at doses of 12.5 and 25 mg/kg significantly increased the seizure threshold in comparison to the control group ($P < 0.001$). We showed that ketamine significantly increased the seizure threshold compared to the control group ($P < 0.001$). Administration of a subeffective dose of EA (6.25 mg/kg) plus ketamine significantly increased the seizure threshold in comparison to the group that received a subeffective dose of EA alone ($P < 0.001$). Furthermore, coinjection of NMDA agonist plus an effective dose of EA (25 mg/kg) significantly decreased the seizure threshold in comparison to the group that received an effective dose of EA alone ($P < 0.05$).

3.2. Effect on Gene Expression of NMDA Receptors in the Prefrontal Cortex. According to the results (Figure 2(a)), EA at a dose of 25 mg/kg significantly decreased the gene expression of Nr2a in comparison to the control group ($P < 0.05$). Ketamine as an NMDA receptor antagonist significantly reduced the gene expression of Nr2a in comparison to the control group ($P < 0.05$). We found that coadministration of ketamine plus the subeffective dose of EA (6.25 mg/kg) significantly decreased the gene expression of Nr2a in comparison to the group that received a subeffective dose of EA alone ($P < 0.05$).

The present study showed that EA at doses of 12.5 and 25 mg/kg significantly decreased the gene expression of Nr2b compared to the control group ($P < 0.05$, Figure 2(b)). Simultaneous injection of the subeffective dose of EA (6.25 mg/kg) with ketamine significantly reduced the expression of the Nr2b gene compared to the group that received the subeffective dose of EA alone ($P < 0.05$). Moreover, coinjection of the effective dose of EA (25 mg/kg) plus NMDA agonist significantly increased the Nr2b gene in comparison to the group that received an effective dose of EA alone ($P < 0.05$).

4. Discussion

The present study showed that EA possessed the anticonvulsant effect and increased the threshold of PTZ-induced seizures in mice. We demonstrated that inhibition of NMDA receptor using ketamine potentiated the anticonvulsant effect of a subeffective dose of EA. Findings determined that coadministration of NMDA agonist with the effective dose of EA attenuated the anticonvulsant effect of an effective dose of EA. Our data showed that EA decreased the gene expression of Nr2a and Nr2b subunits of NMDA receptors in the prefrontal cortex.

It has been well-determined that the glutamatergic system and NMDA receptors are involved in the pathophysiology of seizures [32]. In this concept, it has been determined that in subsequent seizures, the concentration of glutamate increased in the synaptic space and extracellular fluid, which through its excitatory toxicity leads to neural damages [33]. The molecular basis of this cytotoxicity is not well understood; however, there is some agreement that the accumulation of calcium ions within neurons following activation of NMDA receptors leads to neuronal damages [34, 35].
Activation of NMDA receptors induces long-term alteration of synaptic connections and alteration of neuronal circuits, which may involve the pathogenesis of seizure [16]. It has been determined that NMDA receptor antagonists exerted anticonvulsant effects [12, 13]. Moreover, previous studies have demonstrated that agonists of NMDA receptors increase the severity of seizures and diminish the anticonvulsant effect of some anticonvulsants [36].

In this regard, past studies have shown that changes in the expression of NMDA receptor subunits, including NR2a and NR2b, play a vital role in the pathophysiology of seizures [37]. In this concept, animal studies showed that seizures increased the expression of Nr2a and Nr2b subunits of the NMDA receptors in the brain [38]. Previous studies have demonstrated that selective NR2b blockers significantly attenuate seizures and increase seizure threshold [39]. In a study by Mathern et al., on patients with seizures, an increase in mRNA levels and change in the composition of ionotropic glutamate receptors in the temporal lobe of the brain were observed. It has been suggested that these changes may play a role in neuronal stimulation, neuronal synchronization, and seizures [40]. In our study, in line with aforementioned previous studies, induction of seizures by PTZ leads to an increase in gene expression of NMDA receptor subunits including Nr2a and Nr2b in the prefrontal cortex.

EA is a polyphenolic compound found in some fruits [41]. Ample evidence reported neuroprotective effects for EA in various neurological models [42–44]. Lorigooini et al. showed that EA by attenuating the NMDA receptors possessed antidepressant-like effects in mice [45]. In a study conducted by Girish et al., it has been determined that EA via activation of the gabaergic system improves learning and memory [46]. Previously, it has been demonstrated that EA possessed anti-inflammatory effects and reduced oxidative stress [47]. Dhingra and Jangra showed that EA could attenuate the seizures in PTZ-induced seizures in mice [48].

Furthermore, ameliorative effects of EA on maximal electroshock and PTZ-induced seizures in mice have been reported [49]. Recently, EA has been shown that modulating oxidative stress and inflammatory cytokines ameliorates PTZ-induced seizures in mice [50]. However, the exact mechanisms involved in the anticonvulsant effect of EA have not been fully determined. In line with the aforementioned studies, we found that EA increased the seizure threshold in PTZ-induced seizures in mice.

This study found that following treatment with EA, the gene expression of Nr2a and Nr2b subunits of NMDA receptors in the prefrontal cortex significantly decreased. They were indicating that attenuation of NMDA receptors may mediate the anticonvulsant effects of EA. To examine this hypothesis, we treated mice with an EA plus agonist and/or antagonist of the NMDA receptor. Findings showed that ketamine (the NMDA antagonist) potentiated while NMDA agonist attenuated the anticonvulsant effects of subeffective and effective doses of EA, respectively. These results determined that the NMDA receptor, partially at least, is involved in the anticonvulsant effects of EA.

5. Conclusion

Our results showed that EA exerts an anticonvulsant effect. We found that the anticonvulsant effect of EA, partially at least, mediated through attenuation of NMDA receptors.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors have no conflicts of interest to declare regarding the study described in this article and its preparation.

Authors’ Contributions

Mohammad Rahimi-Madiseh designed the experiments, performed the experiments, and wrote the paper. Zahra Lorigooini performed the experiments and wrote the paper. Shabakir Nasiri Boroujeni analyzed and interpreted the data and performed the experiments. Marziyeh Tajfi performed the experiments. Hossein Amini-Khoei contributed to the idea, designed the experiments, prepared reagents and materials, analyzed data, and wrote the paper. Mohammad Rahimi-Madiseh and Zahra Lorigooini are co-first authors.

Acknowledgments

This study was supported by a research grant (No. 3944) from Shahrekord University of Medical Sciences, Shahrekord, Iran. The authors are thankful to Mr. Amin Soltani and Mrs. Elham Bijad for contributing to this study.

References

[1] I. Bhandari, K. K. Malla, P. Ghimire, and B. Bhandari, “Clinical profile of patients admitted with seizure disorder in a tertiary care hospital of central Nepal,” Journal of College of Medical Sciences-Nepal, vol. 17, no. 2, pp. 129–135, 2021.

[2] H. Tekgul, K. Gauvreau, J. Soul et al., “The current etiologic profile and neurodevelopmental outcome of seizures in term newborn infants,” Pediatrics, vol. 117, no. 4, pp. 1270–1280, 2006.

[3] W. Löscher, H. Klitgaard, R. E. Twyman, and D. Schmidt, “Modern antiepileptic drug development has failed to deliver: ways out of the current dilemma,” Epilepsia, vol. 52, no. 4, pp. 657–678, 2011.

[4] W. Löscher and D. Schmidt, “New avenues for anti-epileptic drug discovery and development,” Nature Reviews Drug Discovery, vol. 12, no. 10, pp. 757–776, 2013.

[5] H. Bradford, “Glutamate, GABA and epilepsy,” Progress in Neurobiology, vol. 47, no. 6, pp. 477–511, 1995.

[6] J. D. Fernstrom, “Monosodium glutamate in the diet does not raise brain glutamate concentrations or disrupt brain functions,” Annals of Nutrition and Metabolism, vol. 73, Supplement 5, pp. 43–52, 2018.
D. M. Lovinger, “Neurotransmitter roles in synaptic modulation, plasticity and learning in the dorsal striatum,” *Neuropsychopharmacology*, vol. 58, no. 7, pp. 951–961, 2010.

S. He, X. Zhang, and S. Qu, “Glutamate, glutamate transporters, and circadian rhythm sleep disorders in neurodegenerative diseases,” *ACS Chemical Neuroscience*, vol. 10, pp. 175–181, 2018.

H. Franke and H. Kittner, “Morphological alterations of neurons and astrocytes and changes in emotional behavior in pentylenetetrazol-kindled rats,” *Pharmacology Biochemistry and Behavior*, vol. 70, no. 2-3, pp. 291–303, 2001.

G. Mahaveer, B. Jagriti, and D. S. Arya, “Hydroalcoholic extract of Emblica officinalis Gaertn. affords protection against PTZ-induced seizures, oxidative stress and cognitive impairment in rats,” *Indian Journal of Experimental Biology*, vol. 1, pp. 474–478, 2010.

A. Sarkaki, Y. Farbood, M. Dolatshahi, S. M. T. Mansouri, and A. Khodadadi, “Neuroprotective effects of ellagic acid in a rat model of Parkinson’s disease,” *Acta Medica Iranica*, vol. 54, no. 8, pp. 494–502, 2016.

E. N. Bum, M. Schmutz, C. Meyer et al., “Anticonvulsant properties of the methanolic extract of *Cyperus articulatus* (Cyperaceae),” *Journal of Ethnopharmacology*, vol. 76, no. 2, pp. 145–150, 2001.

R. M. Jafari, M. H. Ghahremani, N. Rahimi et al., “The anticonvulsant activity and cerebral protection of chronic lithium chloride via NMDA receptor/nitric oxide and phospho-ERK,” *Brain Research Bulletin*, vol. 137, pp. 1–9, 2018.

L. Lumley, J. Niquet, B. Marrero-Rosado et al., “Treatment of acetylcholinesterase inhibitor-induced seizures with polytherapy targeting GABA and glutamate receptors,” *Neuropsychopharmacology*, vol. 185, article 108444, 2021.

A. A. Kovalenko, M. V. Zakharova, O. E. Zubareva, A. P. Schwarz, T. Y. Postnikova, and A. V. Zaitsev, “Alterations in mRNA and protein expression of glutamate receptor subunits following pentylenetetrazole-induced acute seizures in young rats,” *Neuroscience*, vol. 468, pp. 1–15, 2021.

Q. Chen, S. He, X.-L. Hu et al., “Differential roles of NR2A and NR2B-containing NMDA receptors in activity-dependent brain-derived neurotrophic factor gene regulation and limbic epileptogenesis,” *Journal of Neuroscience*, vol. 27, no. 3, pp. 542–552, 2007.

L. Wearick-Silva, A. Sebben, Z. Costa-Ferro, D. R. Marinovic, and M. Nunes, “Undernourishment and recurrent seizures early in life impair long-term potentiation and alter NMDAR and AMPAR expression in rat hippocampus,” *International Journal of Developmental Neuroscience*, vol. 75, no. 1, pp. 13–18, 2019.

S.-Y. Chen, K. Zheng, Z. Q. Wang, and Z.-Q. Wang, “Neuroprotective effects of ellagic acid on neonatal hypoxic brain injury via inhibition of inflammatory mediators and down-regulation of JNK/p38 MAPK activation,” *Tropical Journal of Pharmaceutical Research*, vol. 15, no. 2, pp. 241–251, 2016.

O. Herrera-Calderon, R. Santiváñez-Acosta, B. Pari- Olarte, E. Enciso-Roca, V. M. C. Montes, and J. L. A. Acevedo, “Anticonvulsant effect of ethanolic extract of *Cyperus articulatus* L. leaves on pentylenetetrazol induced seizure in mice,” *Journal of Traditional and Complementary Medicine*, vol. 8, no. 1, pp. 95–99, 2018.

P. Chen, F. Chen, and B. Zhou, “Antioxidative, anti-inflammatory and anti-apoptotic effects of ellagic acid in liver and brain of rats treated by D-galactose,” *Scientific Reports*, vol. 8, no. 1, pp. 1–10, 2018.

J.-Y. Park, J. Y. Lee, K. H. Seo et al., “Comparison of whitening effect of Rubus coreanusfruit according to maturity,” *Nutrition and Health*, vol. 53, no. 2, pp. 121–128, 2020.

O. M. Ali, A. A. Bekhit, S. N. Khattab et al., “Synthesis of lactoferin mesoporous silica nanoparticles for pemetrexed/ellagic acid synergetic breast cancer therapy,” *Colloids and Surfaces B: Biointerfaces*, vol. 188, article 110824, 2020.

E. Uzar, H. Alp, M. U. Cevik et al., “Ellagic acid attenuates oxidative stress on brain and sciatic nerve and improves histopathology of brain in streptozotocin-induced diabetic rats,” *Neurological Sciences*, vol. 33, no. 3, pp. 567–574, 2012.

Y. Farbood, A. Sarkaki, M. Dolatshahi, S. M. T. Mansouri, and A. Khodadadi, “Ellagic acid protects the brain against 6-hydroxydopamine induced neuroinflammation in a rat model of Parkinson’s disease,” *Basic and Clinical Neuroscience*, vol. 6, no. 2, pp. 83–89, 2015.

C. Girish, V. Raj, J. Arya, and S. Balakrishnan, “Evidence for the involvement of the monoaminergic system, but not the opioid system in the antidepressant-like activity of ellagic acid in mice,” *European Journal of Pharmacology*, vol. 682, no. 1-3, pp. 118–125, 2012.

D. Dhirenga and R. Chhillar, “Antidepressant-like activity of ellagic acid in unstressed and acute immobilization-induced stressed mice,” *Pharmacological Reports*, vol. 64, no. 4, pp. 796–807, 2012.

S. Saeidi, H. Azhdari Zarmehri, E. Erami, and A. Khodadadi, “Ellagic acid protects the brain against 6-hydroxydopamine-induced neuroinflammation,” *Acta Medica Iranica*, vol. 10, pp. eaau2357, 2018.

S. Moradi, A. Haj-Mirzaian, H. Amini-Khoei et al., “NMDA receptor antagonists attenuate the proconvulsant effect of juvenile social isolation in male mice,” *Brain Research Bulletin*, vol. 121, pp. 158–168, 2016.

A. Haj-Mirzaian, S. Amir, H. Amini-Khoei et al., “Involvement of NO/NMDA-R pathway in the behavioral despair induced by amphetamine withdrawal,” *Brain Research Bulletin*, vol. 139, pp. 81–90, 2018.

P. Galecki, E. Galecka, M. Maes et al., “The expression of genes encoding for COX-2, MPO, iNOS, and sPLA2-IIA in patients with recurrent depressive disorder,” *Journal of Affective Disorders*, vol. 138, no. 3, pp. 360–366, 2012.

A. Nouri, F. Hashemzadeh, A. Soltani, E. Saghaei, and H. Amini-Khoei, “Progesterone exerts antidepressant-like effect in a mouse model of maternal separation stress through mitigation of neuroinflammatory response and oxidative stress,” *Pharmaceutical Biology*, vol. 58, no. 1, pp. 64–71, 2020.

Y. Wang, X. Tian, D. Xu et al., “GPR40 modulates epileptic seizure and NMDA receptor function,” *Science Advances*, vol. 4, no. 10, p. eaau2357, 2018.

D. V. Amakhin, S. L. Malkin, J. L. Ergina et al., “Alterations in properties of glutamatergic transmission in the temporal cortex and hippocampus following pilocarpine-induced acute seizures in Wistar rats,” *Frontiers in Cellular Neuroscience*, vol. 11, p. 264, 2017.

A. Kłodzińska, M. Bijak, E. Chojnacka-Wojcik et al., “Roles of group II metabotropic glutamate receptors in modulation of seizure activity,” *Naunyn-Schmiedeberg’s Archives of Pharmacology*, vol. 361, pp. 283–288, 2000.
[35] Q. Gu and C. Wang, “The NMDA receptors: physiology and neurotoxicity in the developing brain,” in *Handbook of Developmental Neurotoxicology*, pp. 207–214, Elsevier, 2018.

[36] K. A. Edwards and S. L. Zup, “Serotonin pretreatment abolishes sex-specific NMDA-induced seizure behavior in developing rats,” *Neuroscience*, vol. 463, pp. 184–196, 2021.

[37] J. Sánchez-Hernández, P. Aguiler, J. Manjarrez-Marmolejo, and J. Franco-Pérez, “Fructose ingestion modifies NMDA receptors and exacerbates the seizures induced by kainic acid,” *Neuroscience Letters*, vol. 772, p. 136476, 2022.

[38] S. Gascón, M. Sobrado, J. M. Roda, A. Rodriguez-Pena, and M. Díaz-Guerra, “Excitotoxicity and focal cerebral ischemia induce truncation of the NR2A and NR2B subunits of the NMDA receptor and cleavage of the scaffolding protein PSD-95,” *Molecular Psychiatry*, vol. 13, no. 1, pp. 99–114, 2008.

[39] X. Zhu, J. Dong, K. Shen et al., “NMDA receptor NR2B subunits contribute to PTZ-kindling-induced hippocampal astrogliosis and oxidative stress,” *Brain Research Bulletin*, vol. 114, pp. 70–78, 2015.

[40] G. W. Mathern, J. K. Pretorius, H. I. Kornblum et al., “Human hippocampal AMPA and NMDA mRNA levels in temporal lobe epilepsy patients,” *Brain: A Journal of Neurology*, vol. 120, no. 11, pp. 1937–1959, 1997.

[41] A. Gupta, A. K. Singh, R. Kumar, S. Jamieson, A. K. Pandey, and A. Bishayee, “Neuroprotective potential of ellagic acid: a critical review,” *Advances in Nutrition*, vol. 12, no. 4, pp. 1211–1238, 2021.

[42] M. Goudarzi, S. Amiri, A. Nesari, A. Hosseinzadeh, E. Mansouri, and S. Mehrzadi, “The possible neuroprotective effect of ellagic acid on sodium arsenate-induced neurotoxicity in rats,” *Life Sciences*, vol. 198, pp. 38–45, 2018.

[43] M. Goudarzi, M. A. Mombeini, I. Fatemi et al., “Neuroprotective effects of Ellagic acid against acrylamide-induced neurotoxicity in rats,” *Neurological Research*, vol. 41, no. 5, pp. 419–428, 2019.

[44] A. B. Jha, S. S. Panchal, and A. Shah, “Ellagic acid: insights into its neuroprotective and cognitive enhancement effects in sporadic Alzheimer’s disease,” *Pharmacology Biochemistry and Behavior*, vol. 175, pp. 33–46, 2018.

[45] Z. Lorigooini, N. Salimi, A. Soltani, and H. Amini-Khoei, “Implication of NMDA-NO pathway in the antidepressant-like effect of ellagic acid in male mice,” *Neuropeptides*, vol. 76, article 101928, pp. 10–19, 2019.

[46] C. Girish, V. Raj, J. Arya, and S. Balakrishnan, “Involvement of the GABAergic system in the anxiolytic-like effect of the flavonoid ellagic acid in mice,” *European Journal of Pharmacology*, vol. 710, no. 1-3, pp. 49–58, 2013.

[47] N. Seeram, R. Lee, M. Hardy, and D. Heber, “Rapid large scale purification of ellagitannins from pomegranate husk, a byproduct of the commercial juice industry,” *Stroke*, vol. 41, pp. 49–55, 2005.