HPV genotyping and p16/Ki-67 test significantly improve detection rate of high-grade cervical squamous intraepithelial lesion

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Abstract

Introduction: Liquid-based cytology allows to apply modern and specific analyses of hrHPV genotyping in p16/Ki-67 test. All of these together could raise accuracy ratio for high-grade squamous intraepithelial lesion above 90%. The purpose of this study was to evaluate the diagnostic accuracy of LBC, hrHPV testing, and p16/Ki-67 testing in diagnosis of high-grade cervical intraepithelial lesions.

Material and methods: The study consisted of 176 women, out of which 50 presented with HSIL (CIN2) SCC (cervical intraepithelial lesion grade 2 squamous cell carcinoma). 126 women with a negative Pap test were pooled into the second group of the study. All patients were resampled for LBC, HPV genotyping, and for the p16/Ki-67 test. The research was carried out between May and December 2017, and second sampling were taken from 1 to 4 months.

Results: We reported a strong correlation between positive Pap test and hrHPV (p < 0.05) that met accuracy close to 90%. We noted correlations between a positive p16/Ki-67 with a positive Pap test: p < 0.001; 66% sensitivity (95% CI: 51.2–78.8%), 87.8% specificity (95% CI: 75.2–95.4%), 76.8% accuracy (95% CI: 67.2–84.7%), and OR 13.9 (95% CI: 4.9–39.2), especially HSIL and HPV16: p < 0.001; sensitivity (95% CI) 64.0, specificity (95% CI) 98.4, accuracy (95% CI) 88.6, OR (95% CI) 109.3.

Conclusions: The results of our study indicate hrHPV genotyping as a good biomarker for the triage of patients with an abnormal cytological report. In our opinion, the hrHPV test reaches the highest level of sensitivity, specificity, and accuracy, and should be considered as crucial diagnostic test in cervical screening.

Key words: cervical cancer, cancer screening, Ki67/p16, liquid base cytology, human papillomavirus.
Introduction

Over 500,000 new cases of cervical cancer are reported worldwide each year, with about 27,000 fatal outcomes [1]. However, a recent meta-analysis restricted to the population of China illustrates the clinical aspect of cervical cancer treatment strategy, and although an additional innovative pelvic hyperthermia was applied, the 3-year survival rate remained unchanged (OR = 1.17) [2]. In order to decrease cervical cancer incidence ratio, the screening programs have been undergoing modification for the last years. National guidelines determine the type of program, terminology, monitoring, and quality control. The implementation of such a program must fulfill many conditions such as design, planning, feasibility testing, piloting and trial launching, scaling up health service delivery, running of full-scale programme, and finally sustainability [3]. In 1988, during a meeting of the National Institutes of Health in Bethesda, Maryland, USA, a new system for reporting cervical cytology was introduced and approved – the Bethesda system (TBS), which is still a fundamental algorithm for patients’ management on daily basis. In 2001 and most recently in 2014, TBS was modified, and ‘undetermined significance’ category and liquid-based cytology (LBC) principles were introduced [4–6]. In clinical practice, the introduction of the ASC-US, ASC-H (‘atypical squamous cells cannot exclude HSIL’), and LSIL categories significantly increased the number of ambiguous results of Pap smears that needed further verification, which in turn greatly increased the cost of screening but also led to improve high-grade detection ratio [7]. At that time, the American Society for Colposcopy and Cervical Pathology (ASCCP) has accepted the strategy for clinical treatment according to the TBS [8]. This was the first time when reporting terminology was correlated with HPV biology and clinical management. The recent cutting-edge solutions like LBC, computer-assisted imaging, and HPV-testing have been implemented over the past decade; however, looking from time perspective, only hrHPV testing significantly improved the LBC detection ratio [9–12]. The LBC has significantly decreased the ratio of inadequate smears, improved the detection of low-grade squamous intraepithelial lesion (LSIL) and ASC-US, and increased the number of colposcopy and recent intraepithelial neoplasia diagnosis. Moreover, this allowed to keep a high detection rate of HSIL, irrespectively of patient age [13–15]. Recently, the molecular status of hrHPV, especially HPV type 16 or 18 testing and prophylactic HPV vaccines, have been added to cervical cancer screening program and management guidelines. Liquid-based cytology is a technique, which has taken over conventional cytology (CC) in some countries. In Europe, Denmark and the Netherlands have implemented LBC-based screening as first [13–16]. Technically, LBC assay takes about 50% of sample volume to prepare slides, and the remaining sample, stored in the vial as a cell suspension, can be used for immunocytochemistry, molecular testing of infectious agents, DNA cytometry, and DNA ploidy analysis.

The p16/Ki-67 test provides an insight to HPV E6 and E7 oncoproteins underlying molecular disturbances. They inhibit the suppressor genes p53 and Rb leading to their inactivation, respectively. In effect, the connection between E7 and Rb gene promotes cell proliferation that could be visualized by cytoplasmic p16 overload and Ki-67 protein accumulation in nuclear compartment. The p16/Ki-67 and hrHPV-positive tests greatly correlate with high-grade cervical intraepithelial lesion, reaching accuracy ratio over 90% [11, 17, 18].

This way of screening could be much more efficient than CC assuming that the specificity and sensitivity ratio of LBC and hrHPV co-tests stands for over 90%. Recently ended the ATHENA trial covering above 42,000 U.S. women ≥ 25 years compared sensitivity/ specificity of Pap cytology (ThinPrep®), hybrid strategy (cytology and HPV test), and HPV as primary screening. Conclusions pointed hrHPV testing as the most well-balanced and efficient [9, 10].

To reach high detection rate of HSIL, in many papers, a lot of effort has been devoted to quality control and reproducibility of molecular solutions used for HPV typing. Conclusions were mostly similar: keeping full validation from the stage of sampling through storage, adequate equipment, and analysis are crucial [19–22]. However, the relatively high-cost of testing (roughly 40 USD per singular DNA-HPV or singular p16/Ki-67 test) was pointed out and a national screening provider should face new reality by employing such modern techniques.

The aim of this study was to evaluate the diagnostic accuracy and value of LBC, hrHPV testing, and dual-stained p16/Ki-67 test in the diagnosis of high-grade cervical intraepithelial lesions including cervical cancer.

Material and methods

This regional, Polish multicenter prospective study included patients from the Clinic of Gynecology and Obstetrics, Complex Voivodship Hospital and the Department of Gynecology, Holy Cross Oncology Centre in Kielce who had a CC performed. All enrolled patients were divided into two groups. The including criteria for group 1 were as follows: negative CC, negative previous screening history, and negative gynecological examination including colposcopy. Group 2 consisted of women with positive CC with cytological report covering
ASUS-US, ASC-H, LSIL, HSIL, and SCC. As inclusion criteria included high-grade cervical neoplasia (HSIL) confirmed by cervical biopsy as spectrum lesion ranging from CIN2 to SCC, the time from CC to second sampling was from 1 to 4 months and no treatment to date. Patients who did not fulfill the criteria were excluded.

All enrolled patients were reinvited for additional sampling for LBC, hrHPV molecular test, and p16/Ki-67 test. The research was carried out between May 2017 and December 2017, and a written consent was obtained from all participants.

The LBC were performed in the Pathology Department of Jan Kochanowski, University in Kielce using the LBC SurePathTM system. Cytological findings were categorized according to the updated 2014 TBS protocol [6]. The total pool of 176 cases were tested for the presence of DNA hrHPV by Cobas 4800 platform (Roche Diagnostic©) using typical brushing and preservation in PreservCyt® (Roche Diagnostic©). All PCR assays were performed using Cobas® 4800 commercial test (LOT Y08899). The Cobas HPV test was employed to detect 14 hrHPV genotypes. The test specifically identifies HPV16 and HPV18, while concurrently detecting other hrHPV (31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, and 68) at clinically relevant infection levels. All group No 2 patients and additional patients from group No 1 with negative cytology but positive Cobas test were assayed with TriPath SurePath™ (BD) unstained and tested immunocytochemically with the p16/Ki-67 test (CINtec PLUS, Roche Diagnostic© kit C.N 605-100, p16 IgG2a, Ki-67 IgG) according to fully validated protocol on Benchmark XT (Roche Diagnostic©). All employed solutions met the criteria of CE IVD (EU directive for the 'in-vitro-diagnostic medical devices') and FDA (the Food and Drug Agency), and are used in many national screening programs worldwide. SurePath® LBC and hrHPV Cobas® tests were performed in every case. One patient was excluded from the study due to an invalid PCR test.

LBC and p16/Ki-67 immunocytochemical reactions were analyzed by three independent pathologists (PL, MK, AN-G) blinded to the clinical data. All observed data was tabulated and then statistically analyzed. All inter-observer discrepancies were collectively reanalyzed and to final calculations consensus results were used.

Statistical analysis

Categorical data was expressed as number and percentage. Chi-square ($\chi^2$) test or Fisher’s exact test were applied to compare proportions. Numerical variables were presented as median and interquartile range and compared by Mann-Whitney $U$ test. Interobserver variabilities were assessed by Cohen’s $k$ coefficient. Sensitivity, specificity, accuracy, and odds ratio (OR) with 95% confidence intervals (95% CI) were also calculated. A two tailed $p$-value < 0.05 was considered as statistically significant. All statistical analyses were performed using R (version 3.1.2; the R Foundation for Statistical Computing, Vienna, Austria) or Statistica (StatSoft, Inc., 2014, version 12).

**Results**

One hundred and seventy five women ranging from 21 to 67 years of age (average age: 37.5; STD, 10.4; median, 34) were included in the analysis. Group 1 consisted of 126 women, age ranging from 21 to 67 (average: 38.3 years). Group 2 included 50 patients, with age ranging from 23 to 59 (average: 35.3 years). There were no statistical differences between the groups according to age ($p = 0.14$). Applied solution resulted in 0.5% ratio of unsatisfactory slides and 8% HSIL under-diagnosis by CC versus LBC. Blinded to clinical data pathologists reached 62% specificity to detect HSIL. The remaining 19 patients were coded as non-HSIL spectrum lesions. Table I presents the comparison of HPV infection and Pap test findings. Every categorized HPV type strongly associated with positive cytology confirming its strong value as biomarker for intraepithelial lesion. Inside both LSIL and unequivocal pattern group (ASC-US, ASC-H, AGUS), 84% frequency ratio of hrHPV infection was found.

Table II presents high percentage of sensitivity, specificity, and accuracy for hrHPV. In HPV16 and generally all types of HPV infection met accuracy close to 90%. Moreover, we observed high specificity of all types of HPV infection as predictors of cell abnormality as cancer prodromal sign.

To evaluate the utility of p16/Ki-67 in HSIL, we compared 64 pooled cases including 50 women with positive Pap test and 14 hrHPV-positive patients with negative Pap test; CIN excluded by biopsy (grey zone). The control group consisted of 36 triple negative (negative cytology, clinical examination, and Cobas test) cases. Full inter-observer agreement reached 90.9% (90/99), while Cohen’s $k$ coefficient was 0.807. Strong correlations were observed between a positive p16/Ki-67 and a positive Pap test (66% sensitivity (95% CI: 51.2–78.8%), 87.8% specificity (95% CI: 75.2–95.4%), 76.8% accuracy (95% CI: 67.2–84.7%), and OR = 13.9 (95% CI: 4.9–39.2; $p < 0.001$), especially HSIL and HPV16 (sensitivity (95% CI) 64.0, specificity (95% CI) 98.4, accuracy (95% CI) 88.6, OR (95% CI) 109.3; $p < 0.001$). HPV 16 was found more frequent in the HSIL ($p = 0.026$) and collaterally positive p16/Ki-67 test proved of strong diagnostic predictor of HSIL ($p = 0.002$, Table III).

The positive Pap test, confirmed by biopsy as CIN2-SCC, could be the golden standard to esti-
mate HSIL detection. Our data concerning p16/Ki-67 test efficiency was as follows: 66% sensitivity (95% CI: 51.2–78.8%), 87.8% specificity (95% CI: 75.2–95.4%), 76.8% accuracy (95% CI: 67.2–84.7%), and OR = 13.9 (95% CI: 4.9–39.2). The highest percentage of positive p16/Ki-67 tests was observed in patients with positive cytology and positive hrHPV (p < 0.001) (Figure 1).

**Discussion**

The past decade has been fruitful in attempts to improve cervical cancer screening productivity. It seems that CC has been irretrievably replaced by much more efficient LBC and additional molecular and immunological tests.

In our SurePath study, we met 0.5% unsatisfactory slides and 8% HSIL under-diagnosis by CC versus LBC. The Dutch study by Rozemeijer et al. published a large-cohort studies demonstrating the advantage of LBC over CC. The authors compared the sensitivity of CC, SurePath, and ThinPrep methods. They observed a 14% improvement in the detection of CIN I and CIN II using SurePath vs. CC. ThinPrep did not improve the CIN detection rate. In more recent study, HR at 0.81 was associ-

### Table I. Comparison of HPV genotype with HSIL and non-HSIL lesions

| Variable          | All (n = 175) | Positive Pap test (n = 50) | Negative Pap test (n = 125) | P-value* |
|-------------------|---------------|----------------------------|----------------------------|----------|
|                   | Collectively as non HSIL (n = 19) | HSIL (n = 31) |                   |          |
| HPV16:            |               |                            |                            | < 0.001  |
| Positive          | 34 (19.4)     | 4 (20)                     | 0                          |          |
| Negative          | 141 (80.6)    |                            | 123 (98.4)                 |          |
| HPV18:            |               |                            |                            | 0.022    |
| Positive          | 3 (1.71)      | 1 (10)                     | 0                          | 0 (0.00) |
| Negative          | 172 (98.3)    |                            | 125 (100)                  |          |
| HPV findings:     |               |                            |                            | < 0.001  |
| HPV DNA not detectable | 119 (68.0) | 1 (10)                     | 2 (20)                     |          |
| Single HPV type   | 47 (26.9)     | 9 (90)                     | 3 (40)                     |          |
| HPV coinfection   | 9 (5.14)      | 0                          | 1 (25)                     |          |

*P-value corresponds to comparison HSIL and non-HSIL versus negative Pap group; Fisher’s exact test was used if any expected counts were less than 5.

### Table II. Estimation of Cobas HPV categories diagnostic value

| Parameter                              | HPV16          | HPV18          | HPV other       | HPV-positive including coinfection* |
|----------------------------------------|----------------|----------------|-----------------|-------------------------------------|
| Sensitivity (95% CI)                   | 64.0 (49.2–77.1) | 6 (1.3–16.5)  | 30.0 (17.9–44.6) | 84.0 (70.9–92.8)                    |
| Specificity (95% CI)                   | 98.4 (94.3–99.8) | 100 (97.1–100.0) | 89.6 (82.9–94.3) | 88.8 (81.9–93.7)                   |
| Accuracy (95% CI)                      | 88.6 (82.9–92.9) | 73.1 (65.9–79.6) | 72.6 (65.3–79.0) | 87.4 (81.6–92.0)                   |
| OR (95% CI)                            | 109.3 (24.1–495.8) | –              | 3.7 (1.6–8.5)   | 41.6 (16.3–106.4)                  |

*Cases with HPV coinfection also many types infection in category 'HPV other'.  **not applicable due to specificity = 100%
HPV genotyping and p16/Ki-67 test significantly improve detection rate of high-grade cervical squamous intraepithelial lesion. We noted a strong correlation between HPV16 and ‘HPV other’ with positive cytological findings. Furthermore, a positive p16/Ki-67 test strongly correlated with HPV16, particularly with HSIL ($p < 0.0001$). These results confirm the crucial role of HPV genotyping and p16/Ki-67 test in improving the detection rate of high-grade cervical squamous intraepithelial lesion.

Table III. Comparison of HSIL and non-HSIL detection rate according to HPV genotypes and p16/Ki-67 test

| Variable | Control group (n = 36) | Targeted group (n = 31) | P-value* |
|----------|------------------------|------------------------|----------|
|          | Grey zone (n = 14)     | HSIL (n = 31)          | Collectively as non-HSIL (n = 19) |
|          |                        |                        | ASC-US (n = 10) | ASC-H (n = 5) | LSIL (n = 4) |
| Age median | 34.0 (29.0; 39.5) | 33.0 (27.5; 37.0) | 35.0 (31.5; 42.0) | 0.133 |
|          | Q1; Q3                 |                        |                        | Mann-Whitney U test |
| HPV16:   |                        |                        |                  | 0.026 |
| Negative | 18 (36.0)             | 7 (22.6)               |                      |            |
| Positive | 32 (64.0)             | 24 (77.4)              | 2 (10)              | 0 2 (50)    |
| HPV18:   |                        |                        |                  | 1.000 |
| Negative | 47 (94.0)             | 29 (93.5)              |                      |            |
| Positive | 3 (6.0)               | 2 (6.45)               | 1 (10)              | 0 0        |
| HPV other: |                        |                        |                  | 0.611 |
| Negative | 35 (70.0)             | 23 (74.2)              |                      |            |
| Positive | 15 (30.0)             | 8 (25.8)               | 6 (60)              | 3 (60) 2 (50) |
| p16/Ki-67: |                        |                        |                  | 0.002 |
| Negative | 17 (34.0)             | 5 (16.1)               | 9 4 0              |            |
| Positive | 33 (66.0)             | 26 (83.9)              | 1 (36.8)            | 1 4        |
| HPV and p16/Ki-67**: |                  |                      |                  | 0.003** |
| HPV-negative |                        |                        |                  |            |
| p16/Ki-67-negative | 40 | 2 (6.45) | 1 (10) | 2 (40) 0 |
| HPV-negative |                        |                        |                  |            |
| p16/Ki-67-positive | 3 | 1 (3.23) | 0 1 0 |
| HPV-positive |                        |                        |                  |            |
| p16/Ki-67-negative | 15 | 3 (9.68) | 9 (90) | 0 0 |
| HPV-positive |                        |                        |                  |            |
| p16/Ki-67-positive | 34 | 25 (80.6) | 0 2 (40) | 4 (100) |

*Comparative analysis of HSIL, non-HSIL with double negative group (negative Pap test with negative HPV test). **Comparative analysis of HSIL detection by p16/Ki-67 test according to HPV-positive test.

Figure 1. SurePath based on the p16/Ki-67 immunocytochemical assay: strongly positive reaction with LSIL pattern. Brown cytoplasmic chromogen corresponds to p16 overload (white arrow); nuclear compartment red chromogen is a proof of high Ki-67 protein accumulation and proliferative activity (black arrow); red arrow depicts unaffected cell (case of HPV multi-infection), magnification 40×
role of HPV16 of cancer detecting. Our results confirmed the efficiency of the p16/Ki-67 test in triaging HPV16-positive cases (p = 0.026). Similar observations were reported by Rokita et al. who found a very high diagnostic value of the p16/Ki-67 test in triaging patients with a positive Pap smear (ASC-US, ASC-H, LSIL) in cervical cancer screening [18]. In recent research from China, authors also reported a high correlation between HSIL and p16/Ki-67 in HPV16-positive women [11]. A fresh substudy nested in the ATHENA trial was also reported the utility p16/Ki-67. Wright et al. noted higher sensitivity of the p16/Ki-67 test compared to Pap cytology for triaging HPV-positive women (p < 0.0001) [9]. The same results were reported by Posatti-Resende et al. and Ordí et al. [17,23].

Many molecular analyses for the HPV infection detection are available. The most widely studied include HC2® (Qiagen), Cobas 4800® (Roche), Clart® (Genomica), Aptima® (Hologic), and recently launched Oncoclarity on Viper (BD®). However, some discrepancies between these assays exist and there is a wide agreement about their clinical usefulness. Except for Clart, the rest of the assays have been FDA-approved for screening as reliable and high throughput solutions [19,21,22]. It is important to note that the preservative fluid, storage time, and additional long-term preservatives have an impact on the reliability of the results [20]. In our opinion, performing p16/Ki-67 dual staining tests is useful, even crucial, for the correct categorization of hrHPV-positive patients with unequivocal cytological report, especially LSIL, ASC-US, or ASC-H. Barron et al. published a paper devoted to triaging LSIL-H (ASC-H) using hrHPV typing. They concluded that a Pap smear categorized as LSIL-H has a significant lower hrHPV ratio than all stage above CIN2 (CIN2+) [24]. Our cohort reported 5 cases of ASC-H with a 60% frequency of ‘HPV other’ infection. The most recent interesting paper published by Samimi et al. reported a problem with a cytological pattern detection of HPV infection. The authors concluded the HSIL cytological pattern might be obscured in cases of multigenotype hrHPV infection and in fact, true HSIL cases could mimic unequivocal Pap patterns (ASC-US, ASC-H, LSIL) [25]. In our cohort, we found only one biopsy, which confirmed Semimi’s thesis. Specifically, we discovered a case of CIN2 with HPV co-infection strongly positive with p16/Ki-67, but interestingly, with persistent LSIL cytological pattern. Whether this is nuclear abnormality masking or sampling error remains unclear (Figure 1).

In general, we found 16% hrHPV coinfection and 2 (6%) cases of HSIL hrHPV-negative on Cobas platform, proving that rarer hrHPV genotypes could be involved but they are not matched by commercial assays. According to our results, we found only 3 cases of isolated HPV18 infection with no statistical value, and the subgroup ‘HPV other’ did not demonstrate p16/Ki-67 overexpression (p = 0.502). This indirectly confirms the leading role of HPV16 what is in line with many observers. There is a question: does HPV16 E7 oncoprotein act the same way as other HPV ones?

All our calculations concerning p16/Ki-67 demonstrated a 66% sensitivity and 87.8% specificity for CIN2+ (HSIL), which are in line with other studies [9,11,17,18].

Our research was based on 50-item group and the results are consistent with other studies. The small cohort is an expected weakness of that analysis and there is a need for additional large-cohort study on Polish population. In addition, we did not reach statistical correlation with HPV18 and ‘HPV other’ what need further investigation, especially in light of mentioned E7 oncoprotein. Our study is considered the first paper presenting the LBC-HPV-p16/Ki-67 aspect in Poland. Undoubtedly, the strong side of our study was methodology used and technical aspect in compliance with CE IVD and FDA principles.

In conclusion, the results of our study indicate hrHPV genotyping and dual-stained p16/Ki-67 testing as good biomarkers for the triage of patients with an abnormal cytological report. We noted a high efficiency of SurePath® and its reliability for molecular and immunological applications. In our opinion, the hrHPV test reaches the highest level of sensitivity, specificity, and accuracy, and should be considered as a crucial diagnostic test in cervical screening. Assuming the advantage of LBC over CC and consequent higher diagnostic ratio, this method of screening should rapidly replace the CC testing. Only high-quality solutions provide globally comparable results and should be incorporated in a national screening program.

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Conflict of interest

The authors declare no conflict of interest.

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