Downregulation of interferon-γ-induced protein 10 in the tears of patients with Stevens-Johnson syndrome with severe ocular complications in the chronic stage

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ABSTRACT

Objectives Stevens-Johnson syndrome (SJS) and toxic epidermal necrolysis (TEN) are acute inflammatory vesiculobullous reactions of the skin and mucosa such as the ocular surface, oral cavity and genitals. Severe ocular complications (SOC) arise in some patients with SJS/TEN diagnosed by dermatologists. To investigate the pathophysiology of ocular surface inflammation in SJS/TEN with SOC in the chronic stage, we examined cytokines in the tears of patients with ocular surface diseases and healthy controls.

Participants SJS/TEN eyes in the chronic stage (n=30), healthy eyes (n=20, controls) and eyes (n>20) from patients with atopic keratoconjunctivitis representing different ocular surface inflammatory disorders.

Primary outcome measures Tear samples were collected on Schirmer’s measurement strips. To measure the level of various cytokines in the tears we used BD CBA Flex sets.

Study design An observational study (case–control study).

Results We recorded the level of interleukin (IL)-6, IL-8, eotaxin, macrophage inflammatory protein (MIP)-1β, RANTES (regulated on activation, normal T cell expressed and secreted), interferon gamma (IFN)-γ, monocyte chemoattractant protein-1, IFN-γ-induced protein 10 (IP-10) and total IgE. We found that compared with the controls, in SJS/TEN with SOC, IL-6, IL-8, eotaxin and MIP-1β were significantly upregulated while IP-10 was significantly downregulated. Compared with atopic keratoconjunctivitis, IP-10 was significantly downregulated in SJS/TEN with SOC; on the other hand, total IgE was significantly upregulated in atopic keratoconjunctivitis compared with SJS/TEN with SOC.

Conclusions IP-10 in tears may be a biomarker to distinguish between chronic SJS/TEN with SOC and other ocular inflammatory disorders such as atopic keratoconjunctivitis.

INTRODUCTION

Stevens-Johnson syndrome (SJS) is an acute inflammatory vesiculobullous reaction of the skin and mucosa, for example, the ocular surface, oral cavity, respiratory tract and genitals. In patients with extensive skin detachment and a poor prognosis, the condition is called toxic epidermal necrolysis (TEN). Approximately 40% of patients with SJS/TEN present with severe ocular lesions, for example, severe conjunctivitis with pseudomembrane and ocular surface epithelial defects in the acute stage.1

While dermatologists tend to see patients with SJS/TEN only in their acute stage, ophthalmologists usually encounter these patients in the chronic stage. Besides typical ocular surface findings in patients with SJS/TEN in the chronic stage such as scarring of the lid conjunctiva with severe dry eye, trichiasis and symblepharon, and often...
conjunctival invasion to the cornea, the ophthalmological diagnosis of SJS/TEN is based on a confirmed history of acute-onset high fever, serious mucocutaneous illness with skin eruptions and involvement of at least two mucosal sites including the ocular surface. In the chronic stage of SJS/TEN with severe ocular complications (SOC), patients may manifest mild ocular surface inflammation.

Elsewhere we reported that among various cytokines, the level of interleukin (IL)-6, IL-8 and monocyte chemoattractant protein-1 (MCP-1), but not of IL-1β, IL-5, eotaxin, interferon (IFN)-γ and macrophage inflammatory protein (MIP)-1α, was dramatically increased in the tear fluid and serum of a patient with SJS in the acute stage. Our search of the literature found no report on tear cytokines in patients with SJS/TEN in the chronic stage.

Identification of the ocular pathophysiology of chronic SJS/TEN using patient tears, and the detection of differences from other ocular surface diseases, may help elucidate the pathophysiological mechanism(s) underlying the onset of SJS/TEN with SOC. Therefore, we investigated the pathophysiology of ocular surface inflammation in patients with SJS/TEN in the chronic stage by studying various cytokines in their tears. We compared our findings with those made in healthy controls and in patients with other ocular surface inflammatory disorders such as atopic keratoconjunctivitis.

MATERIALS AND METHODS

Study design
An observational study (case–control study).

Subjects
Our study protocol was approved by the ethical review board of Kyoto Prefectural University of Medicine; all patients provided prior written informed consent. This study was performed in accordance with the guidelines of the Declaration of Helsinki. We included 40 eyes of patients with SJS/TEN in the chronic stage who had developed SJS/TEN with SOC more than 1 year earlier. We defined SOC as a condition manifesting pseudo-membrane and epithelial defects on the ocular surface (cornea and/or conjunctiva) in the acute stage, and ocular sequelae such as severe dry eye, trichiasis, symblepharon and conjunctival invasion into the cornea in the chronic stage.

We used tear samples from more than 30 eyes of patients who were seen between 2013 and 2016 by MU in the outpatient clinic of the Department of Ophthalmology, Kyoto Prefectural University of Medicine. The controls were tears from more than 20 eyes of healthy volunteers free of ocular disease. For comparative studies, we collected tears from more than 20 eyes of patients with atopic keratoconjunctivitis.

In patients with SJS with SOC, we recorded the titre of IL-8, eotaxin, IFN-γ, MCP-1, total IgE and IFN-γ-induced protein 10 (IP-10) in 40 eyes, the level of IL-6 and RANTES (regulated on activation, normal T cell expressed and secreted) in 36 eyes, and the MIP-1β level in 34 eyes. The level of eotaxin was assayed in 28 eyes of control subjects; IL-6, IL-8, RANTES, IFN-γ, MCP-1, and total IgE in 27, and IP-10 and MIP-1β in 23 and 22 such eyes, respectively. To determine the titre of IL-6, IL-8, eotaxin, RANTES, IFN-γ, MCP-1 and total IgE in patients with atopic keratoconjunctivitis we used 26 eyes; IP-10 and MIP-1β levels were determined in 22 and 20 such eyes, respectively.

Tear collection
Tears were collected on Schirmer’s measurement strips (Schirmer Tear Production Measuring Strips, Showa Yakuhin Kako, Tokyo, Japan) according to our previously reported method. They were then immersed in 100 μL Tris-buffered saline with Tween 20 (TBST) (DAKO, Japan) for 10 min at room temperature. Using BD CBA flex sets, we measured the titre of IL-6, IL-8, MCP-1, IFN-γ, IP-10, MIP-1β, eotaxin, RANTES and total IgE in 50 μL of TBST containing the tears. The tear volume on the Schirmer strips was calculated at 1 μL intervals using a standard curve obtained from 0 to 25 μL of distilled water.

Measurement of tear cytokines using CBA kits
The concentration of cytokines was measured with BD CBA flex sets and BD human soluble protein master buffer kits according to the manufacturer’s instructions (BD Bioscience-PharMingen, San Diego, CA). Briefly, 50 μL of mixed capture beads was admixed with 50 μL of the provided standards or the tear samples and incubated in the dark for 1 hour at room temperature. Then 50 μL of mixed detection reagents was added, the samples were incubated in the dark for 2 hours, washed, centrifuged and resuspended in 300 μL of wash buffer. A BD FACs Canto II flow cytometer was calibrated with set-up beads. Data were analysed as recommended by the manufacturer. Concentrations were obtained by interpolating the fluorescence intensity on an at least 7-point dilution standard curve supplied with the kit and calculated with BD FCAP Array software.

Data were expressed as the mean and as individual values and evaluated with Student’s t-test and one-way analysis of variance (ANOVA) using Microsoft Excel software. Differences of p<0.05 were considered to be statistically significant.

RESULTS
We analysed the level of IL-6, IL-8 and MCP-1 because we documented elsewhere that it was upregulated in tears from patients with SJS/TEN in the acute stage.
We also compared the titre of the Th1 cytokines IFN-γ, IP-10, RANTES and MIP-1β, and of eotaxin as another cytokine. Because IgE is a specific marker for allergic diseases, we also included total IgE in our study. We confirmed that these cytokines and IgE could be detected in tears. As our preliminary study using small tear samples to detect the titre of IFN-α, IL-4, IL-1β, IL-10, IL-12p70 and IL-17F showed that their detection was difficult they were not included in our analyses.

**Figure 1** Comparison of cytokine and IgE levels in tears from patients with SJS/TEN with SOC, healthy controls and patients with atopic keratoconjunctivitis. The cytokines were measured in several groups. *p<0.05, **p<0.005, ***p<0.0005. IFN-γ, interferon gamma; IL, interleukin; IP-10, IFN-γ-induced protein 10; MCP-1, monocyte chemoattractant protein-1; MIP-1β, macrophage inflammatory protein; N, number of eyes; RANTES, regulated on activation, normal T cell expressed and secreted; SJS, Stevens-Johnson syndrome; SOC, severe ocular complications; TEN, toxic epidermal necrolysis.
As shown in figure 1, our comparison of tear samples from patients with SJS/TEN with SOC and the controls revealed significant upregulation of IL-6 (p<0.0005), IL-8 (p<0.005), eotaxin (p<0.005), MIP-1β (p<0.005), RANTES (p<0.05) and IFN-γ (p<0.005) in the tears of patients with SJS/TEN with SOC. The level of MCP-1, which we reported to be upregulated in SJS/TEN with SOC in the acute stage, was similar in patients with SJS/TEN with SOC and the controls. The total IgE level was not significantly different in the patients and the controls. On the other hand, comparison of the IP-10 titre showed that it was significantly downregulated in SJS/TEN with SOC (p<0.005).

When we compared tear samples from patients with SJS/TEN with SOC and atopic keratoconjunctivitis, we found that IP-10 was significantly downregulated in SJS/TEN with SOC (p<0.05); total IgE, on the other hand, was significantly upregulated in atopic keratoconjunctivitis (p<0.0005). There was no significant difference between patients with SJS/TEN with SOC and patients with atopic keratoconjunctivitis with respect to the level of IL-6, IL-8, eotaxin, MIP-1β, RANTES, IFN-γ and MCP-1.ANOVA, applied as a multiple comparisons test, revealed that the titre of IL-6, IL-8, eotaxin and MIP-1β was significantly different between the three groups (all p<0.05), as was the total IgE titre (p<0.000000005).

DISCUSSION

In tears from patients with SJS/TEN with SOC and patients with atopic keratoconjunctivitis, the level of IL-6, IL-8, eotaxin and MIP-1β was upregulated. Their upregulation in both patient groups may be a common phenomenon in the presence of ocular surface inflammatory disorders. On the other hand, IP-10 was downregulated in the tears from patients with chronic SJS/TEN with SOC but not in the presence of atopic keratoconjunctivitis. Others reported that IP-10 was upregulated in the tears of patients with dry eye. Its downregulation may contribute specifically to the pathogenesis of the SJS with SOC. Like the significant upregulation of total IgE in patients with atopic keratoconjunctivitis, the downregulation of IP-10 in tears may be a biomarker to distinguish chronic SJS/TEN with SOC from other ocular inflammatory disorders. Assay of these serum cytokine levels showed that only RANTES and total IgE could be clearly detected (data not shown). This observation suggests tears are more useful than serum for the detection of biomarkers of ocular surface inflammatory disorders.

Elsewhere we reported that TLR3 polymorphisms are associated with SJS/TEN with SOC, and that IP-10 was strongly induced by polyI:C, a TLR3 ligand, in conjunctival epithelial cells. We document that in patients with SJS in the chronic stage, the tear IP-10 level was significantly downregulated compared with patients with atopic conjunctivitis and healthy controls.

However, IFN-γ was not downregulated although IP-10 and IFN-γ are both Th1 cytokines. As IFN-γ is primarily produced by immune cells such as lymphocytes, a difference in its dominant producers may account for the different IP-10 and IFN-γ levels.

The current study documents that IP-10 regulation is different in patients with SJS/TEN with SOC and patients with other ocular surface inflammatory disorders and suggests strongly that TLR3 contributes to the pathogenesis of SJS/TEN with SOC. To identify the mechanism of ocular surface inflammation of SJS, further investigations and analysis are needed.

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