Genetic Variants of IDE-KIF11-HHEX at 10q23.33 Associated with Type 2 Diabetes Risk: A Fine-Mapping Study in Chinese Population

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Abstract

Background: Genome-wide association studies (GWAS) in populations of European ancestry have mapped a type 2 diabetes susceptibility region to chromosome 10q23.33 containing IDE, KIF11 and HHEX genes (IDE-KIF11-HHEX), which has also been replicated in Chinese populations. However, the functional relevance for genetic variants at this locus is still unclear. It is critical to systematically assess the relationship of genetic variants in this region with the risk of type 2 diabetes.

Methodology/Principal Findings: A fine-mapping study was conducted by genotyping fourteen tagging single-nucleotide polymorphisms (SNPs) in a 290-kb linkage disequilibrium (LD) region using a two-stage case-control study of type 2 diabetes in a Chinese Han population. Suggestive associations (P<0.05) observed from 1,200 cases and 1,200 controls in the first stage were further replicated in 1,725 cases and 2,081 controls in the second stage. Seven tagging SNPs were consistently associated with type 2 diabetes in both stages (P<0.05), with combined odds ratios (ORs) ranging from 1.14 to 1.33 in the combined analysis. The most significant locus was rs7923837 [OR = 1.33, 95% confidence interval (CI): 1.21–1.47] at the 3′-flanking region of HHEX gene. SNP rs1111875 was found to be another partially independent locus (OR = 1.23, 95% CI: 1.13–1.35) in this region that was associated with type 2 diabetes risk. A cumulative effect of rs7923837 and rs1111875 was observed with individuals carrying 1, 2, and 3 or 4 risk alleles having a 1.27, 1.44, and 1.73-fold increased risk, respectively, for type 2 diabetes (P for trend = 4.1E-10).

Conclusions/Significance: Our results confirm that genetic variants of the IDE-KIF11-HHEX region at 10q23.33 contribute to type 2 diabetes susceptibility and suggest that rs7923837 may represent the strongest signal related to type 2 diabetes risk in the Chinese Han population.

Introduction

Type 2 diabetes is one of major public health problems around the world with an affected number of 240 million in 2007, with an expected rapid increase to 380 million by 2025 [1]. In China, about 92.4 million adults (≥20 years old) are affected with diabetes while about 148.2 million adults are in a status of prediabetes [2]. Type 2 diabetes is a serious metabolic disorder, characterized by insulin resistance and relative insulin deficiency, which results from both genetic and environmental factors. The increasing prevalence of type 2 diabetes is largely attributed to environmental factors acting on genetically susceptible individuals. Therefore, it is important to understand the susceptibility genes for type 2 diabetes to facilitate risk assessment, primary prevention, early detection and treatment.

Genome-wide association study (GWAS) has made spectacular progress in identifying susceptibility genes involved in type 2 diabetes. In 2007, the first GWAS of type 2 diabetes conducted in a French case-control cohort identified a type 2 diabetes related locus on chromosome 10q23.33, which is located in a gene cluster including the insulin-degrading enzyme (IDE), the kinesin-interacting factor 11 (KIF11), and the hematopoietically expressed homeobox (HHEX) [3]. This association has been consistently replicated in the subsequent studies [4–9], including several studies in Chinese populations [10–17]. HHEX encodes a transcription factor with a key role for pancreatic development [18]. Reduction of IDE activity by a pharmacological inhibitor increases islet amyloid polypeptide (amylin) accumulation and amylin-mediated cytotoxicity in cultured β-cells [19], whereas IDE ablation causes glucose intolerance in knockout mice [20]. Therefore, both IDE and HHEX genes are strong candidates as susceptibility genes modulating the pathogenesis of type 2 diabetes.
The IDE-KIF11-HHEX locus spans a 290 kb region in linkage equilibrium (LD) (Figure S1). Two variants at the 3’-flanking region of the HHEX gene, rs7923837 and rs1111873, have been associated with type 2 diabetes risk as lead single-nucleotide polymorphisms (SNPs) in original GWAS in populations of European ancestry. These two SNPs do not reside within the coding or putative regulatory regions of any known genes. It is still an open topic which variant(s) at IDE-KIF11-HHEX locus is (are) causal, especially in different populations. Herein, with the aim to systematically evaluate the relationship between genetic variants at 10q23.33 and type 2 diabetes risk and provide evidence for causal variant(s) in this gene region, we conducted a fine-mapping study including 14 tagging SNPs at 10q23.33 in a large, two-stage case-control study with a total of 2,925 cases and 3,281 controls in a Chinese population.

Methods

Ethics statement
This study was approved by the Ethical Committee of Nanjing Medical University. Written informed consent was obtained from each participant before investigation.

Study population
In this study, we performed a two-stage case-control study. The first-stage (discovery-phase) fine-mapping analysis was designed to discover the suggestive variants associated with type 2 diabetes in a Chinese population consisting of 1,200 cases and 1,200 controls from a community based cohort study of type 2 diabetes in Wuxi, a city of southern Jiangsu Province, China. In this study, eligible subjects aged over 30 years old were enrolled in 2007 and the baseline information, including demographic, disease history, family history of diabetes was obtained and a detailed clinical examination was conducted. Anthropometric variables including height, weight, waist and hip circumference and blood pressure were measured. Body mass index (BMI) was calculated as weight (in kilograms) divided by the square of height (in meters). Ten hours overnight fasting blood samples were drawn for measurements of fasting blood glucose (FBG) and lipids in all subjects. Glucose concentration and lipids were measured on an OLYMPUS (C2734-Au640) automatic analyzer in the central laboratory of Wuxi Center for Disease Control and Prevention, which was authorized to perform laboratory tests according to the international quality standard ISO/IEC 17025. At baseline, 1,200 subjects who had a history of type 2 diabetes and were on medical treatment for type 2 diabetes were selected as type 2 diabetes cases. The control subjects were selected from those without history of diabetes, hypertension, coronary heart disease, stroke, cancer and with a FBG<5.6 mmol/l at both baseline and follow-up in 2009 and were frequency-matched to the cases on sex (n = 1,200).

The second-stage (replication-phase) was to confirm the results observed in the first-stage, and consisted of 1,725 cases and 2,081 controls derived from a community-based cross-sectional survey on chronic non-communicable diseases in 2009 in Nantong, a city in middle Jiangsu Province, China. All study participants aged over 30 years were interviewed face-to-face by trained interviewers using a pre-tested questionnaire including demographic, behaviors, disease history, and family history of diabetes. Anthropometric parameters and blood pressure were measured. Over 10-hour fasting blood samples were collected for measurement of blood glucose and lipids. Subjects were considered to have type 2 diabetes if they had a history of type 2 diabetes or if their fasting glucose was 7.0 mmol/l or higher. Eventually, 1,725 subjects were recruited as cases. Meanwhile, we selected 2,081 subjects without history of diabetes, hypertension, coronary heart disease, stroke, cancer and with a FBG<5.6 mmol/l as controls from the same population and also frequency-matched to the cases on sex.

Individuals with the following conditions were excluded from the study at both stages: malnutrition (BMI<18.5 kg/m^2); physical disabilities or psychological disorder; obesity caused by other diseases (such as inlet cell tumor, Cushing’s syndrome, polycystic ovary syndrome, hypogonadism, hypothyroidism) or by medication (such as glucocorticoid, oral contraceptive); cancer; current diagnosis of any communicable disease. All subjects in this study were unrelated ethnic Han Chinese. All blood specimens were stored at the Key Molecular Epidemiology Laboratory, Department of Epidemiology and Biostatistics, School of Public Health, Nanjing Medical University. Genomic DNA was extracted from a leukocyte pellet by protease K digestion and was followed by phenol-chloroform extraction and ethanol precipitation.

SNPs selection
We selected both potentially functional SNPs and haplotype-tagging SNPs (htSNP) in the IDE-KIF11-HHEX locus at 10q23.33 (chr10:94199856–94489557). Firstly, to infer potentially functional SNPs, we searched common (MAF≥0.10) SNPs located in the 5’-flanking region, 5’untranslated region (5’UTR), exon, and 3’UTR of IDE, KIF11 and HHEX genes using NCBI dbSNP database and 8 SNPs (rs7078243, rs1044153, rs11595187, rs11187083, rs2297743, rs3758505, rs7099761, rs4646954) were found. As there isn’t any information on rs1044153, rs7099761, rs4646954 in HapMap database, we conducted LD analysis on the other 5 SNPs (rs7078243,rs11595187, rs11187083, rs2297743, rs3758505). Among them, 3 SNPs (rs2297743, rs7078243, rs11187083) were finally selected for this study, because the other 2 SNPs (rs3758505 and rs11595187) are in high LD with rs2297743 (r^2 = 0.97 and r^2 = 0.96, respectively). Secondly, we directly included three SNPs (rs111875, rs5015480, rs7923837) which were significantly associated with type 2 diabetes risk in previous GWAS.

Then we used the block-based tagging strategy to find tagging SNPs using Haploview 4.2 software according to the HapMap database (http://www.hapmap.org/), phase II Nov08, on NCBI B36 assembly, dbSNP b126; population: Chinese Han population (CHB) and Japanese population (JPT); MAF≥0.10, Hardy-Weinberg equilibrium P≥0.05 and call rate ≥95%) on the basis of pairwise LD r^2 threshold of 0.8. After forcing including the above 6 SNPs (rs2297743, rs7078243, rs11187083, rs111875, rs5015480, rs7923837) as tagging SNPs into Haploview software, the additional 13 SNPs (rs11187096, rs4646957, rs7911264, rs2488073, rs947591, rs10882084, rs11187146, rs7910605, rs2488075, rs7918084, rs11187094, rs6583826, rs4933236) that met the above criteria in this IDE-KIF11-HHEX region were selected. As a result, a total of 19 SNPs were selected for genotyping in the discovery stage (Figure S1).

Genotype determination
In the discovery stage, genotyping was performed using the TaqMan OpenArray Genotyping System (Life Technologies, Carlsbad, USA), a medium-throughput genotyping platform. DNA samples with standardized concentration were loaded and amplified on 48-sample arrays following the manufacturer’s protocol. For quality control, the equal amounts of cases and controls and two no template controls (NTCs) were simultaneously detected in each chip. Four SNPs (rs2297743, rs11187096, rs2488073 and rs7918084) were excluded because of deficiencies in probes design in the chip and the SNP rs6583826 was excluded.
from analysis because of low call rate (<80.0%). The overall call rate for the remaining 14 SNPs was 90.9%, with a call rate >97.0% for each locus.

In the replication stage with 1,725 type 2 diabetes cases and 2,081 controls, iPLEX Sequenom MassARRAY platform (Sequenom, Inc) was used to genotype the 11 SNPs that were significant in the discovery stage. Genotyping was conducted blindly and two NTCS in each 384-sample plate were used for quality control. The overall call rate of this stage was 99.4%, with a call rate >99% individually.

Except for rs7078243 in the discovery stage (P=0.048) and rs11107994 in the replication stage (P=0.002), the genotype distribution of other SNPs were all in Hardy-Weinberg equilibrium.

Statistical analyses

χ² or Student’s t tests were used to examine the differences in the distributions of characteristics between type 2 diabetes cases and controls. Hardy-Weinberg equilibrium was tested using a likelihood ratio test. LD between SNPs was evaluated using Haploview version 4.2. Genotype distributions between cases and controls were compared using logistic regression under the additive genetic model with adjustment for age, sex and BMI as confounding factors. The combined effect of multiple SNPs on the risk of type 2 diabetes was determined by logistic regression after categorizing the participants into groups according to the number of the risk alleles carried. Individuals with no risk alleles served as the reference group. Cochran’s χ² -based Q-statistic was performed to assess heterogeneity in subgroups. Meta-analysis of 9 studies [10–17 and this study] in Chinese populations was conducted to estimate the pooled effect size. Heterogeneity of the 9 studies was assessed with the Cochran’s χ² -based Q-statistic. The random-effects model was adopted when heterogeneity existed; otherwise the fixed-effects model was appropriate. Combined ORs were calculated using the Mantel-Haenszel (fixed-effects) and DerSimonian and Laird (random-effects) tests [21,22]. The significant P value of overall ORs was determined using the Z-test. All analyses were performed using the PLINK 1.07 and Stata software (version 11.1; StataCorp LP, College Station, Texas).

Results

The characteristics of the study populations are presented in Table 1. No significant differences were observed in the distributions of sex in both stages and the combined analysis (P>0.05). Type 2 diabetes cases were in a higher age than controls (P<0.01) and had significantly higher levels of BMI, FBG, triglycerides (TG), total cholesterol (TC) and significantly lower level of high density lipoprotein-cholesterol (HDL-C) as compared with controls in both stages and combined (P<0.001).

Among the 14 SNPs analyzed in the first stage (Table 2), 11 SNPs were associated with type 2 diabetes risk with a P value less than 0.05, rs4646957, rs7910605, rs11187094, rs7078243, rs7911264, rs11187185, rs5015480, rs11187146, rs7923837, rs2488075, and rs947591. The 11 suggestive SNPs were further to be tested in the second stage with additional 1,725 type 2 diabetes cases and 2,081 controls. As shown in Table 2, there were 7 SNPs (rs4646957, rs11187185, rs5015480, rs11187146, rs7923837, rs2488075, and rs947591) showed similar associations as observed in the first stage and had a P value less than 0.05. We further combined the results of two stages for the 7 SNPs that were consistently associated with type 2 diabetes in both stages (P<0.05). As presented in Table 3, all of the 7 SNPs showed significant associations with type 2 diabetes risk with effect size (OR) ranging from 1.14 to 1.33. The most significant locus was rs7923837 after the two stages were combined with an OR of 1.33 (95% CI: 1.21–1.47).

As shown in Figure S2, a moderate LD (r²: 0.19–0.64) is indicated between the most significant SNP rs7923837 and the other 6 SNPs (i.e. rs4646957, rs11187185, rs5015480, rs11187146, rs2488075 and rs947591) that were consistently associated with the risk of type 2 diabetes in the present study. After being conditioned by rs7923837, rs11187185 was the only SNP still with a P value<0.05 (Table 4), suggesting that the effect of rs11187185 could not be fully explained by rs7923837. In contrast, the SNP rs7923837 remained significant after being conditioned by any of the other SNPs (Table 4). We then combined rs11187185 and rs7923837 genotypes to test their joint effects on type 2 diabetes risk. A significant increased risk of type 2 diabetes was detected as the number of risk alleles increased (P for trend =4.1E-10, Table 5). Compared to those without carrying any risk allele, individuals carrying one, two, and three or four risk alleles had a 1.27, 1.44 and 1.73-fold increased risk for developing type 2 diabetes, respectively, while individuals carrying one or more risk alleles had a 1.39-fold increased risk for type 2 diabetes.

Stratification analyses using pooled case-control sets in additive genetic model showed that the associations between the two SNPs (rs1111875 and rs7923837) and type 2 diabetes risk were significant in subgroups stratified by age, sex, BMI, or stage, with ORs ranging from 1.15 to 1.44, and the associations had no significant difference between the subgroups (P>0.05 for heterogeneity test, Table S1).

To estimate the pooled effect size of IDE-KIF11-HHEX variants on type 2 diabetes risk, we conducted a meta-analysis of the three reported SNPs (rs1111875, rs5015480 and rs7923837) in a total of 14,141 cases and 15,725 controls for rs1111875, 9,208 cases and 11,363 controls for rs5015480, and 8,184 cases and 10,358 controls for rs7923837, respectively, in Chinese populations [10–17 and this study]. There was no heterogeneity of ORs across studies of rs1111875 and rs5015480, but there lied heterogeneity among studies of rs7923837 (P=0.044). The pooled ORs for type 2 diabetes were significant for rs1111875 (pooled OR = 1.16, 95% CI = 1.11–1.20, P<0.0001), rs5015480 (pooled OR = 1.18, 95% CI = 1.15–1.25, P<0.0001) in the fixed-effects model, and for rs7923837 (pooled OR = 1.19, 95% CI = 1.08–1.30, P<0.0001) in the random-effects model (Figure S3).

Discussion

The current study represents the first fine-mapping study with an effort to comprehensively investigate the relationship between IDE-KIF11-HHEX locus and type 2 diabetes risk in this Chinese population. This is also the largest study in terms of sample size in a Chinese population to date. We confirmed the associations of genetic variants at IDE-KIF11-HHEX gene region with risk of type 2 diabetes in Chinese Han population. The tagging SNP rs7923837 was found to be the variant with the strongest effect in our population, whereas rs1111875 showed an independent effect on type 2 diabetes that could not be fully interpreted by rs7923837. These findings provide new insights into the genetic variants at IDE-KIF11-HHEX and susceptibility of type 2 diabetes, and allow for a more stable estimation of effect size on this locus facilitating genetic risk prediction in the future.

Since Sladek et al firstly reported the association of genetic variants at IDE-KIF11-HHEX with type 2 diabetes risk in 2007 [3], several studies have replicated the significant association in Chinese population [10–16]. Among those studies, the SNP rs1111875 was included in six studies and the reported ORs
Table 1. Characteristics of the study populations.

| Variables | Stage I | Stage II | Combined all |
|-----------|---------|---------|-------------|
|           | Case    | Control | P | Cases | Controls | P | Cases | Controls | P |
| n (% male) | 1200 (39.8) | 1200 (39.8) | 0.980 | 1725 (35.9) | 2081 (36.3) | 0.805 | 2925 (37.5) | 3281 (37.6) | 0.953 |
| Age (years) | 57.4±9.77 | 56.4±8.02 | 0.006 | 58.7±10.31 | 56.6±10.81 | 1.1E-9 | 58.2±10.11 | 56.5±9.88 | 1.2E-10 |
| BMI (kg/m²) | 24.92±3.42 | 22.64±2.87 | 6.66-66 | 25.13±3.55 | 21.82±2.44 | 7.9E-220 | 25.05±3.50 | 22.12±2.63 | 1.2E-275 |
| FBG (mmol/l) | 8.98±3.52 | 4.51±0.47 | 4.4E-306 | 9.07±3.30 | 4.53±0.56 | <0.001f | 9.03±3.52 | 4.52±0.53 | <0.001f |
| TG (mmol/l) | 2.55±2.46 | 1.32±0.45 | 5.2E-62 | 2.58±2.48 | 0.96±0.39 | 9.7E-170 | 2.57±2.47 | 1.09±0.45 | 6.4E-227 |
| TC (mmol/l) | 5.39±1.62 | 4.64±0.71 | 1.2E-46 | 4.70±1.23 | 4.26±0.82 | 2.0E-38 | 4.98±1.44 | 4.39±0.80 | 3.7E-85 |
| HDL-C (mmol/l) | 1.37±0.34 | 1.53±0.31 | 7.5E-30 | 1.51±0.55 | 1.68±0.40 | 3.6E-26 | 1.46±0.48 | 1.62±0.38 | 2.0E-51 |

Data are means ± SD or numbers (percentage).

Table 2. Summary results of associations between 14 tagging SNPs at 10q23.33 and risk of type 2 diabetes in two stages.

| SNP | Position | Location | /minor allele | MAFcase/control | OR (95%CI)b | Pb | MAFcase/control | OR (95%CI)b | Pb |
|-----|----------|----------|---------------|-----------------|-------------|-----|-----------------|-------------|-----|
| rs4649597 | 94219982 | IDE (intron) | C/T | 0.256/0.224 | 1.22 (1.06–1.41) | 0.007 | 0.250/0.231 | 1.16 (1.02–1.31) | 0.023 |
| rs7910605 | 94229773 | IDE (intron) | A/G | 0.181/0.160 | 1.20 (1.02–1.41) | 0.033 | 0.175/0.161 | 1.12 (0.97–1.29) | 0.119 |
| rs10882084 | 94319093 | intergene | A/G | 0.106/0.093 | 1.21 (0.98–1.49) | 0.073 | – | – | – |
| rs11187083 | 94342485 | KIF11 (5’ near A/T gene) | 0.111/0.109 | 1.06 (0.87–1.29) | 0.553 | – | – | – | – |
| rs11187094 | 94358158 | KIF11 (intron) | A/G | 0.463/0.418 | 1.26 (1.11–1.42) | 3.0E-4 | 0.405/0.396 | 1.05 (0.95–1.17) | 0.339 |
| rs7078243 | 94400243 | KIF11 (3’UTR) | C/A | 0.202/0.157 | 1.42 (1.20–1.67) | 3.0E-5 | 0.181/0.168 | 1.08 (0.94–1.24) | 0.297 |
| rs7911264 | 94426831 | intergene | C/T | 0.241/0.199 | 1.34 (1.15–1.56) | 1.5E-4 | 0.224/0.208 | 1.09 (0.96–1.24) | 0.184 |
| rs11187857 | 94452862 | intergene | T/C | 0.299/0.251 | 1.33 (1.16–1.53) | 4.8E-5 | 0.285/0.261 | 1.15 (1.02–1.30) | 0.019 |
| rs5015480 | 94455539 | intergene | T/C | 0.186/0.160 | 1.27 (1.08–1.49) | 0.005 | 0.184/0.163 | 1.23 (1.07–1.42) | 0.004 |
| rs11187146 | 94468335 | intergene | C/G | 0.358/0.331 | 1.14 (1.01–1.30) | 0.042 | 0.350/0.325 | 1.14 (1.02–1.28) | 0.018 |
| rs7923837 | 94471897 | intergene | A/G | 0.235/0.194 | 1.33 (1.15–1.54) | 1.7E-4 | 0.235/0.199 | 1.34 (1.18–1.52) | 1.0E-5 |
| rs2488075 | 94480154 | intergene | T/C | 0.169/0.139 | 1.32 (1.12–1.56) | 0.001 | 0.171/0.140 | 1.34 (1.15–1.55) | 1.1E-4 |
| rs947591 | 94485733 | intergene | C/A | 0.185/0.158 | 1.28 (1.09–1.50) | 0.003 | 0.187/0.162 | 1.21 (1.06–1.40) | 0.007 |
| rs4933236 | 94488416 | intergene | C/T | 0.340/0.322 | 1.09 (0.95–1.23) | 0.214 | – | – | – |

*Molecular allele frequency (MAF) in cases and controls.

bOdds ratio (OR) and P value derived from logistic regression with a adjustment for age, sex and BMI assuming an additive genetic model.

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In this fine-mapping study, we identified the most significant signal of rs7923837 and additional independent signal of rs1111875, both of which are located at the telomeric end of a 290-kb LD block on chromosome 10. As shown in Figure S1, these two SNPs are both in the 3′-flanking region of HHEX gene. The HHEX gene encodes a transcription factor that is involved in Wnt signaling, a fundamental pathway for cell growth and development [25], and has been shown to regulate β-cell development and/or function through the activation of hepatocyte nuclear factor 1α [26]. Recently, Pivovarova et al. found that rs7923837 and rs1111875 were associated with altered capacity of β-cell secretion and proposed that genetic variants at IDE-KIF11-HHEX locus might mediate the type 2 diabetes risk by modulating β-cell secretory capacity and β-cell mass [27]. Several studies also reported associations between rs1111875 and rs7923837 and fasting insulin secretion, insulin sensitivity and insulin secretion response following a glucose load [28–32]. Therefore, HHEX was suggested as the most likely causal candidate gene at 10q23.33 for type 2 diabetes [33]. Nevertheless, IDE is also another strong candidate gene. Reduction of IDE activity by a pharmacological inhibitor increases islet amyloid polypeptide (amylin) accumulation and amylin-mediated cytotoxicity in cultured β-cells [19] and IDE ablation causes glucose intolerance in knockout mice [20]. To now, it is still hard to say which genetic variant(s) in this locus is (are) causal and how they function in the pathogenesis of type 2 diabetes, though our findings provide additional evidence to refine the potential functional variants. Further functional and resequencing studies are warranted to clarify the biological mechanisms of IDE-KIF11-HHEX locus on type 2 diabetes and to identify additional variants to narrow down the fine-mapped region.

In summary, our fine-mapping study with a two-stage case-control design and large sample size confirmed the association of IDE-KIF11-HHEX locus at 10q23.33 with susceptibility to type 2 diabetes. Our results suggest that tagging SNP rs7923837 may represent the most significant signal in this region in Chinese Han population.
Table 5. Combined effects of rs7923837 and rs1111875 on type 2 diabetes risk.

| Risk allele number | Cases | Controls | OR(95%CI) | P     |
|-------------------|-------|----------|-----------|-------|
| 0                 | 1129  | 1480     | 1         |       |
| 1                 | 810   | 873      | 1.27(1.10–1.46) | 7.5E-4 |
| 2                 | 681   | 675      | 1.44(1.24–1.67) | 1.1E-6 |
| 3–4               | 266   | 224      | 1.73(1.39–2.15) | 8.2E-7 |

rs7923837-G and rs1111875-C were assumed as risk alleles.

Supporting Information

Figure S1 The 290-kb linkage disequilibrium analysis on 10q23.33 and tagging single-nucleotide polymorphisms (chr10:94199856–94489557). The IDE-KIF11-HHEX locus at 10q23.33 (chr10:94199856–94489557) splits into 5 linkage disequilibrium (LD) blocks in Asians (Chinese & Japanese). 19 single-nucleotide polymorphisms (SNPs) (seen at the black arrows) were selected for genotyping in the discovery stage, including 13 haplotype-tagging SNPs (htSNP), chose with criteria of minor allele frequency (MAF) ≥0.10, Hardy-Weinberg equilibrium (P>0.05), and call rate ≥95% on the basis of pairwise LD r² threshold of 0.8, 3 potentially functional SNPs and 3 SNPs previously reported by GWAS of type 2 diabetes. (DOC)

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Figure S2 Linkage disequilibrium analysis of 7 single-nucleotide polymorphisms consistently associated with type 2 diabetes risks. Linkage disequilibrium (LD) strength in controls was shown in the diamonds represented by r² values. A moderate LD (r²: 0.19–0.64) was indicated between the most significant SNP rs7923837 and the other significant 6 SNPs, with r² value being 0.19 for rs4646957, 0.20 for rs1111875, 0.43 for rs5015480, 0.50 for rs11187146, 0.64 for rs2488075, 0.46 for rs947591 respectively. (DOC)

Figure S3 Meta-analysis of 3 single-nucleotide polymorphisms from IDE-KIF11-HHEX locus with type 2 diabetes in Chinese populations. The pooled odds ratios (ORs) for type 2 diabetes were significant for rs1111875 (pooled OR = 1.16, P<0.001), rs5015480 (pooled OR = 1.18, P<0.001) in the fixed-effects model, and for rs7923837 (pooled OR = 1.19, P<0.001) in the random-effects mode. (DOC)

Table S1 Stratification analysis for two independent SNPs (rs7923837 and rs1111875) and risk of type 2 diabetes in additive genetic model. (DOC)

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Author Contributions

Conceived and designed the experiments: YQ GJ ZH HS. Performed the experiments: YQ FL MD YL HI JC CS. Analyzed the data: YQ FL GJ. Contributed reagents/materials/analysis tools: YQ CS HS. Wrote the paper: YQ FL GJ HS.

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