Gene Profiles in a Smoke-Induced COPD Mouse Lung Model Following Treatment with Mesenchymal Stem Cells

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Mesenchymal stem cells (MSCs) effectively reduce airway inflammation and regenerate the alveolus in cigarette- and elastase-induced chronic obstructive pulmonary disease (COPD) animal models. The effects of stem cells are thought to be paracrine and immune-modulatory because very few stem cells remain in the lung one day after their systemic injection, which has been demonstrated previously. In this report, we analyzed the gene expression profiles to compare mouse lungs with chronic exposure to cigarette smoke with non-exposed lungs. Gene expression profiling was also conducted in a mouse lung tissue with chronic exposure to cigarette smoke following the systemic injection of human cord blood-derived mesenchymal stem cells (hCB-MSCs). Globally, 834 genes were differentially expressed after systemic injection of hCB-MSCs. Seven and 21 genes, respectively, were up-and down-regulated on days 1, 4, and 14 after HCB-MSC injection. The Hbb and Hba, genes with oxygen transport and antioxidant functions, were increased on days 1 and 14. A serine protease inhibitor was also increased at a similar time point after injection of hCB-MSCs. Gene Ontology analysis indicated that the levels of genes related to immune responses, metabolic processes, and blood vessel development were altered, indicating host responses after hCB-MSC injection. These gene expression changes suggest that MSCs induce a regeneration mechanism against COPD induced by cigarette smoke. These analyses provide basic data for understanding the regeneration mechanisms promoted by hCB-MSCs in cigarette smoke-induced COPD.

INTRODUCTION

Chronic obstructive pulmonary disease (COPD) is characterized by airway inflammation and alveolar wall destruction (emphysema) that result from exposure to cigarette smoke (Rabe et al., 2007). The developmental mechanism of COPD has been suggested to involve cigarette smoke-induced airway inflammation, oxidative stress, and alveolar destruction from an imbalance between proteases and anti-proteases (MacNee and Tudor, 2009; Yoshida and Tudor, 2007). Although current treatments are available to inhibit airway inflammation and relax contracted airways, there is no therapy that regenerates destroyed alveoli (Aaron et al., 2007; Calverley et al., 2007). Thus, recent studies to improve COPD treatment have focused on the control of anti-oxidative and anti-protease activities.

Cell therapy using various stem cells has shown beneficial therapeutic effects in animal models of COPD (Huh et al., 2011; Katsha et al., 2011; Schweitzer et al., 2011). Our recent studies showed that MSCs from the bone marrow and cord blood successfully improved the emphysema component of COPD (Huh et al., 2011). Recently, we reported that most stem cells injected systemically disappeared within one day, when the stem cells were tracked by fluorescent labeling or with human-based alu sequence quantification (Kim et al., 2014). The early disappearance of infused stem cells has also been observed in other studies (Lee et al., 2009). Most stem cell biologists agree that stem cells can differentiate into bone, adipose tissue, and cartilage in vitro, but exert their therapeutic effects via paracrine and immunomodulatory functions in vivo (Murphy et al., 2013). Stem cells may deliver some signals to host cells, inducing a regenerative mechanism against alveolar destruction in the COPD lung. Therefore, studies focused on changes in gene expression profiles in host cells may be important for understanding MSCs regeneration mechanisms in destroyed alveoli.

Previous studies have analyzed gene profiles in the presence and absence of cigarette exposure. However, no studies have examined the changes in gene profiles following MSC injection in a cigarette smoke-induced COPD model. Therefore, the goal of the current study was to analyze the changes in gene expression profiles in the cigarette smoke-induced COPD mouse lung following systemic injection of human cord blood-derived (hCB)-MSCs over time on days 1, 4, and 14. Using a mouse WG-6 expression bead array based on Illumina, we...
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Fig. 1. Heat map of DEG with p-value < 0.01. (A) and (B) indicates normal air and cigarette exposure, respectively (n = 8, each group). Each gene is represented on the right side of the heat map.

compared gene expression profiles following cigarette exposure and non-exposure. Next, we compared the gene expression profiles of two groups, an hCB-MSC-injected group and a non-injected group, both of which were induced to have COPD lungs by a 6-month exposure to cigarette smoke before systemic injection of hCB-MSCs.

MATERIALS AND METHODS

Animal experiments
C57BL/6J mice were purchased from OrientBio (Korea). Mice were bred in specific pathogen-free facilities at the Asan Medical Center. All live mouse experiments were approved by the Institutional Animal Care and Use Committee of the Asan Medical Center (Korea). Seven-week-old female C57BL/6J mice were exposed to commercial cigarette smoke (Eighty Eight Lights, KT&G, Daejeon, Korea) 5 days per week for 6 months as we described previously (Huh et al., 2011). Briefly, 30-40 mice were settled in an inhalation box (50 × 40 × 30 cm) connected to a pump, and exposed to 12 cigarettes. Control mice were bred in cages with clean room air. Human cord blood MSCs were provided by MEDIPOST Co., Ltd (Korea). hCB-MSCs were cultured with alpha-minimum essential medium (Gibco Life Technologies, USA) and subcultured using 0.25% trypsin-EDTA (Gibco Life Technologies). Mice exposed to 6 months of cigarette smoke were intravenously injected with 5 × 10^4 hCB-MSCs. Mice were then sacrificed and their lungs were collected on days 1, 4, and 14 following injection of hCB-MSCs. RNA later was used to maintain RNA stability during mouse lung collection.

RNA isolation
Total mRNA from lung tissue was prepared using Trizol (Invitrogen Life Technologies, USA) and purified using RNeasy columns (Qiagen, Germany) according to the manufacturer’s protocol. The purity and quantity of the isolated RNA were evaluated using aN1000 spectrophotometer (NanoDrop Technologies Inc., USA), and a quality check of the RNA was performed using a 2100 Bioanalyzer (Agilent Technologies, USA). Only RNA samples with an RNA integrity number >7.5 were used for the microarray experiment.

Microarray experiments
RNA was amplified and purified using the AmbionIllumina RNA amplification kit (Ambion, USA) according to the manufacturer’s instructions. Biotinylated cRNA (1.5 μg) was hybridized to each MouseWG-6 expression bead array (Illumina, Inc., USA), and the array signal was detected using Amersham fluorolink streptavidin-Cy3 (GE Healthcare Biosciences, UK). Arrays were scanned with an Illumina bead array reader confocal scanner.

Immunohistochemistry
The lung was inflated with 0.5% low-melting agarose and fixed using 4% formalin, and then embedded in paraffin and cut into 4-μm-thick sections for immunohistochemistry. The sections were stained using an Immunohistochemistry accessory kit (Bethyl Laboratories, Inc., USA) with anti-Egr-1 antibody (1:250, Abcam, UK) and then observed by microscopy.

Data analysis
All data analysis of differentially expressed genes was conducted using R 2.15.1. Raw expression data was log2-transformed and normalized by a quantile method. Statistical significance of the expression data was determined using a t-test, and we restricted our interest to the gene list with p-values compared to cigarette exposure or non-exposure. The genes analyzed included those show a 1.3-fold change between the smoke-exposed group and smoke + hCB-MSCs injected group at each
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RESULTS

Gene expression profiles following chronic exposure to cigarette smoke

A t-test was performed to identify differentially expressed genes (DEGs) based on their p-values. Forty-nine and 231 DEGs had p-values of < 0.01 and < 0.05, respectively. Figure 1 shows the heat map indicating that 45 genes were up-regulated and 4 genes were down-regulated in the cigarette-exposed group compared to the non-exposed group. The top 10 genes are listed in Table 1 and 231 of DEGs were subjected to GO evaluation using DAVID. Immune response, inflammatory response, and apoptosis (cell death) were enriched in up-regulated DEGs. Down-regulated DEGs were enriched in cell and biological adhesion (Table 2).

Gene expression profiles following injection of hCB-MSCs

A t-test was performed to select genes that were significantly up- or down-regulated following injection of hCB-MSCs. We identified genes showing a > 1.3-fold change that were significantly different between the hCB-MSC-injected group and non-injected group. A total of 834 genes were differently expressed between the two groups; 392, 270, and 368 showed significant changes in expression at 1, 4, and 14 days after injection of hCB-MSCs, respectively. Figure 2 shows the summary and heat map of the t-test and the number of up- and down-regulated genes in the 1.3-fold change group. Some representative genes were subjected to quantitative PCR (qPCR) for technical validation and most microarray data matched with the qPCR results (Fig. 3).

Figure 4 shows the genes specifically altered by injection of hCB-MSCs at each time point and in common between the
COPD-related GO terms such as immune response, inflammatory response, wounding, chemotaxis, and cell adhesion (Yang et al., 2013). Our results also generated these COPD-related GO terms (Table 4). The top 20 up- and down-regulated genes over time are listed in Supplementary Table 2 and 3. Cyp1b1, a representative gene up-regulated by smoke exposure in mice, was decreased by MSC injection on day 14. Hematopoiesis-, immune response-, and cholesterol-related genes were up-regulated on day 1. Down-regulated genes on day 1 were related to transcription, metabolic process, and blood vasculature development (Supplementary Table 4). Supplementary Table 5 shows the results of GO term analysis for up- and down-regulated genes on day 4. Up-regulated genes on day 4 were related to wound healing, immune responses, and blood circulation. Blood and tube development-, transcription-, and metabolic process-related genes were down-regulated on day 4. The results of GO term analysis for up- and down-regulated genes on day 14 are shown in Supplementary Table 6. In particular, the GO term categories of blood vessel and vasculature development appeared in both up- and down-regulated genes.

**DISCUSSION**

In the present study, we analyzed the gene expression profiles following cigarette exposure or non-exposure as well as hCB-MSC-injected or non-injected mice. We focused on the changes in gene profiles between hCB-MSC-injected mice following hCB-MSC injection compared to controls. DEGs on day 14 after injection of hCB-MSCs were related to cell growth regulation. Our results suggest that immune responses, oxidative stress, and transcription were regulated at a nearly stage after hCB-MSC injection (on days 1 and 4), and that blood vessel development and cell growth regulation predominated at a later stage after hCB-MSC injection (on day 14).

In previous reports, MSCs showed regenerative properties in animal models with smoke- and elastase-induced COPD (Huh et al., 2011; Katsha et al., 2011; Schweitzer et al., 2011). Recently, we reported that systemically injected MSCs were observed for only 1 day after cell injection in an elastase-induced emphysema model (Kim et al., 2014). Other reports failed to detect injected MSCs; therefore, stem cell biologists have attempted to explain the therapeutic effect of mesenchymal stem cells in these models in the context of the regulation of host cells or host microenvironment (Henning et al., 2014; Liang et al., 2014; Suga et al., 2014). To understand the detailed mechanisms of stem cell therapy, understanding the changes in host genes following MSC injection is required. Thus, we focused on the host changes based on the profiles of DEGs in this study.

Previous studies analyzed the gene expression profiles in the lungs of smoke-exposed mice (Halappanavar et al., 2009; Rangasamy et al., 2009). Inflammatory signaling was up-regulated following cigarette exposure. We also detected up-regulation of inflammatory-related genes such as GPR109A (HCA2), TNFAIP2, IL-8RA, and CCL6 (Table 1) (Chen et al., 2008; Rangasamy et al., 2009; Zandi-Nejad et al., 2013). A previous report showed that most genes altered by exposure to
smoke returned to a basal level of expression 6 weeks after smoking cessation (Rangasamy et al., 2009). Temporal alterations in the gene expression profile during the progression of emphysema have also been analyzed in the lungs of mice exposed to smoke for 5 h, 8 days, 1.5 months, and 6 months. Genes up-regulated by smoke exposure are categorized as exposed to smoke for 5 h, 8 days, 1.5 months, and 6 months. Temporal alterations in the gene expression profile during the progression of smoking cessation (Rangasamy et al., 2009). Temporal alterations in the gene expression profile during the progression of smoking cessation (Rangasamy et al., 2009). Genes involved in xenobiotic metabolic processes including Ahrr, Cyp1a1, and Cyp1b1 are up-regulated on day 4 after injection of hCB-MSCs. This finding can be explained by AHRR repression of Cyp1a1 through binding to its xenobiotic response element sequence. The expression of Cyp1b1 was transiently increased (up-regulated on day 4 and down-regulated on day 14 after injection of hCB-MSCs). Hbb and Hba, which were highly up-regulated on days 1 and 14 after injection of hCB-MSCs, are involved in oxygen transport from the lung to various peripheral tissues and are expressed in alveolar cells to perform antioxidant functions (Ahmad and Mahmood, 2016; Grek et al., 2011; Huffman et al., 2000; Macari and Lowrey, 2011; Newton et al., 2006).

Serpin is an inhibitor of serine proteases with proteolytic activity, particularly against elastase (Benarafa et al., 2007). The elastase versus anti-elastase hypothesis is one of the most important mechanistic theories for the development of smoking-induced emphysema (Shapiro, 1995; Stone, 1983). An imbalance between elastase and anti-elastase resulting from overexpression of elastase compared with anti-elastase causes alveolar destruction and airspace enlargement. Macrophages and neutrophil-derived elastase-deficient mice are significantly protected from the development of emphysema compared to wild-type mice (Hautamaki et al., 1997; Shapiro et al., 2003).

Therefore, MSCs may have regenerative roles that reestablish a balance between elastase and anti-elastase by up-regulating Serpin expression. Egr1-deficient mice and lung fibroblasts exhibit lower metalloproteinase activity after inflammatory stimuli (Banks et al., 2005). This previous report suggests that EGR-1, by regulating metalloproteins, is important in the development of COPD. We found that Egr1 was downregulated following injection of hCB-MSCs at all time points analyzed and that this alteration may inhibit metalloproteinase activity. Collectively, according to the cigarette smoke emphysema model, MSC up-regulate anti-oxidant functions and down-regulate proteinase activity.

There were some limitations to this study. One limitation is that the differential change in gene expression profiles did not involve exposure to normal air and exposure to normal air plus hCB-MSCs. Therefore, further studies of mice exposed to normal air and the use of injected cell controls are needed to correctly identify the detailed regeneration mechanisms of stem cells. Another limitation is that our microarray experiment used whole lungs. Although we determined organ-level changes in molecular signals following the injection of hCB-MSCs, it remains unclear how the gene profiles change in individual cells in the lung such as bronchial and alveolar epithelial cells, fibroblasts, and endothelial cells. More advanced techniques are needed to analyze the gene expression profiles using sections of the lung tissues or specific lung cell types.

This is the first study to determine the time-dependent molecular changes induced by treatment with hCB-MSCs in a cigarette smoke-induced COPD model. MSCs systemically injected into an animal disappear within several days; it is thought that these cells rarely differentiate into tissue resident cells in the lungs, such as epithelial cells. Therefore, studies focused on molecular changes in host tissues may reveal the regenerative mechanisms of MSCs. Our present report improves the understanding of the regenerative mechanisms induced by injection of hCB-MSCs based on the gene expression profile changes in a cigarette smoke-induced COPD animal model.

Note: Supplementary information is available on the Molecules and Cells website (www.molcells.org).

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