Epigenetic Inheritance, Epimutation, and the Response to Selection

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Abstract

There has been minimal theoretical exploration of the role of epigenetic variation in the response to natural selection. Using a population genetic model, I derive formulae that characterize the response of epigenetic variation to selection over multiple generations. Unlike genetic models in which mutation rates are assumed to be low relative to the strength of selection, the response to selection decays quickly due to a rapid lowering of parent-offspring epiallelic correlation. This effect is separate from the slowing response caused by a reduction in epigenetic variation. These results suggest that epigenetic variation may be less responsive to natural selection than is genetic variation, even in cases where levels of heritability appear similar.

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Introduction

Although epigenetic variation has been observed in many wild populations [1–6] and can be inherited across meiotic generations [7–18], its role in phenotypic heritability and adaptive evolution is unclear. Theoreticians and empiricists often hint at the potential importance of epigenetic processes in adaptation [19,20], but Holeski et al. [21] note that “… no multigenerational experiments have evaluated the relative contribution of epigenetic inheritance in response to natural selection.” Despite this, recent results suggest that epigenetic variation can play a role in adaptation. Cropley et al. [22] selected for coat color on mice with induced epigenetic variability and found that the methylation-associated phenotype increased progressively over generations, as long as a dietary generator of epigenetic variation was present. In a review of epigenetic variation and inheritance in plants, Hirsch et al. [23] share the results of a currently unpublished selection experiment using Arabidopsis thaliana, in which a selected line and its genetically identical ancestor consistently differed in phenotype and in cytosine methylation status. In both of these systems, it appears that epigenetic variation responded to selective pressures.

Current population genetic theory doesn’t readily lend itself to an intuitive or analytical understanding of such an adaptive response over multiple generations. Day and Bonduriansky [24] present a very nice theoretical analysis of the transgenerational change in phenotype and genotype frequencies for an array of non-genetic inheritance models, but only derive formulae for change over a single generation. Geoghegan and Spencer [25] explore the properties of evolutionarily stable equilibria when mutation rates are environment-sensitive. However, they focus primarily on the characteristics of the equilibria as opposed to the dynamics of the adaptive response. Klironomos et al. [26] use simulation modeling to understand the response to selection on a fitness landscape where the phenotypic optimum can be achieved by either genetic or epigenetic variants. They find that epigenetic adaptation may occur rapidly as a transient process before genetic adaptation ultimately supersedes, although the authors do not analyze the rate of this process.

To develop an intuitive understanding of epigenetic variation’s role in sustained adaptive evolution, I derive analytical formulae to characterize the response to selection at an epigenetic locus over multiple generations. For simplicity, I focus on one of the simplest models of epigenetic inheritance: the case where the environment is homogeneous, epimutation rates are constant, and viability selection acts on variation at a single epigenetic locus. Unlike genetic mutation, rates of epimutation are poorly understood and may not be orders of magnitude lower than the strength of selection [18,27], so my analytical approximation does not ignore higher order mutation terms. The results suggest that higher potential mutation rates of epigenetic marks can lead to a rapid decay in the response to selection over multiple generations, with this response increasingly reduced in successive generations of selection.

Model

I consider a dynamic haploid model identical to that of Slatkin [28], where an epiallele can take one of two states, 0 or 1, in a particular generation. The individual contributes a fraction of offspring to the next generation that is proportional to its relative fitness. The distribution of states of an individual’s offspring are determined by the epimutation rates $\mu_{01}$ and $\mu_{10}$ (Figure 1). I define the parameter $\mu_{\text{sum}}$ to be the sum of epimutation rates, $\mu_{01} + \mu_{10}$, and assume that state 0 has fitness lower than state 1 by a factor $(1-s)$. Table 1 summarizes the epigenetic states, frequencies, and fitness values.

In a large population with a life cycle of selection, epimutation, and reproduction, the frequency of epiallele 1 in the next generation, $p'$, can be written in terms of the epiallele frequencies in the current generation as
Note that a diploid model in which the viability fitnesses of the heterozygote and the 00 homozygote are equal to \((1 - s)\) and \((1 - s)^2\), respectively, will produce an identical recursion in epiallele frequency. Thus the results presented here extend to the diploid case where the two epialleles within an individual mutate and affect fitness independently of one another.

Results

Epiallelic correlation

Consider the haploid population with epigenetic variation at a locus. I define \(K_P\) and \(K_O\) to be random variables for the epigenetic state of a random parent and one of their offspring. I assume that the population begins at the neutral epimutational equilibrium where the frequency of epiallele 1 is \(p_1 = \frac{\mu_0}{\mu_{sum}}\), and the frequency of epiallele 0 is \(p_0 = \frac{\mu_{10}}{\mu_{sum}}\), where \(\mu_{sum} = \mu_0 + \mu_{10}\). In this case, the covariance between parent and offspring is

\[
\text{Cov}(K_P, K_O) = E[K_P K_O] - E[K_P] E[K_O]
\]

\[
= \pi_1 (1 - \mu_{10}) - \pi_1^2
\]

\[
= \pi_1 (1 - \pi_1 - \mu_{10}).
\]

The variances of these two variables at neutral equilibrium are identical and equal to

\[
\text{Var}(K_P) = \text{Var}(K_O) = \pi_1 (1 - \pi_1).
\]

Combining these formulae, the parent-offspring correlation in epiallelic state is

\[
\rho = \frac{\text{Cov}(K_P, K_O)}{\sqrt{\text{Var}(K_P) \text{Var}(K_O)}} = \frac{\pi_1 (1 - \pi_1 - \mu_{10})}{\pi_1 (1 - \pi_1)}
\]

\[
= 1 - \frac{\mu_{10}}{1 - \pi_1}
\]

\[
= 1 - \frac{\mu_{10} - \mu_{01}}{1 - \pi_1}.
\]

The variances of these two variables at neutral equilibrium are identical and equal to

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= 1 - \frac{\mu_{10}}{1 - \pi_1}
\]

\[
= 1 - \frac{\mu_{10} - \mu_{01}}{1 - \pi_1}.
\]

Exact equilibrium equation

Equation (1) represents a linear fractional system with strictly positive terms, so it will converge to the unique stable equilibrium given by the larger solution to the quadratic equation defined when \(p' = p\). The solution is

\[
p = \frac{s - \mu_{10} - (1 - s)\mu_{01} + \sqrt{(s - \mu_{10} - (1 - s)\mu_{01})^2 + 4s(1 - s)\mu_{01}}}{2s}
\]

Because this solution does not readily lend intuition to the rate of adaptation nor to the relative roles of variance, selection strength, and mutation rate, the following results focus on an approximation to equation (1).
The exact recursion for the change in allele frequencies, equation (1), can be re-arranged as

\[ p' = p + \frac{pq(1-\mu_{\text{sum}})}{1-qs} - p\mu_{10} + q\mu_{01}. \]

Assuming that the selection coefficient \( s \) is relatively small, the recursion can be approximated by a Taylor expansion of equation (1) in which terms of order \( s^2 \) and higher are ignored. This yields

\[ p' \approx p + pq(1-\mu_{\text{sum}}) - p\mu_{10} + q\mu_{01}. \]

To assess the longer term response to selection, I assume that the initial allele frequencies in the population are at epimutation balance, where \( p \) at time \( t = 0 \) is equal to \( \pi_1 = \frac{\mu_{01}}{\mu_{\text{sum}}} \). From these initial conditions, the neutral mutational terms in the exact equation cancel each other.

\[ p(t) \approx \pi_1 + \pi_1(1-\pi_1)s((1-\mu_{\text{sum}}) + \cdots + (1-\mu_{\text{sum}})^t), \]

and

\[ \Delta p(t) \approx \pi_1(1-\pi_1)s(1-\mu_{\text{sum}})^t, \]

where \( \Delta p(t) \) is the change in allele frequency between generation \( t-1 \) and generation \( t \).

The approximate response to selection is proportional to three factors: the initial epiallelic variance \( \pi_1(1-\pi_1) \), the strength of selection \( s \), and a power of the initial parent-offspring epiallelic correlation \( (1-\mu_{\text{sum}})^t \). Because this final factor is raised to the \( t \)-th power, the exact epiallelic correlation will crucially determine whether there is a sustained response to selection. Given this recursion, the approximate equilibrium frequency of allele 1 at epimutation-selection balance is

\[ p \approx \pi_1 + \pi_1(1-\pi_1)\frac{s}{\mu_{\text{sum}}} (1-\mu_{\text{sum}})^t. \]

As mutation rates increase, the response to selection is weaker and there is more rapid convergence to the equilibrium (Figure 2). The approximation holds well as long as the strength of selection \( s \) is smaller than \( \mu_{\text{sum}} \). Alternatively, this approximate equilibrium can be derived from the exact equilibrium equation (2) by Taylor expanding around \( s=0 \) and ignoring terms of \( s^2 \) and higher. Because equation (1) also represents a diploid population in which the two epialleles act independently, these results apply in that case as well.

**Discussion**

In the absence of selection, non-environmentally-sensitive epigenetic variation contributes to parent-offspring phenotypic covariance in a manner nearly identical to that of genetic variation. There is simply one additional discounting factor.
(1 − μsum), which corresponds to the allelic correlation between parent and offspring at an epigenetic locus in a haploid population. This is analogous to the factor (1 − τ) presented by Tal et al. [29]. Under selection, the correlation factor (1 − μsum) characterizes the rate of decay of the response, and also plays a role in determining the epiallele frequencies at epimutation-selection balance. Because the factor (1 − μsum) is taken to progressively higher powers in subsequent generations of selection, an epigenetically controlled phenotype may demonstrate a reduced response to selection in comparison to a genetically controlled phenotype, even if the initial contributions to phenotypic heritability are similar.

In practice, this effect should be observable in an artificial selection experiment. Johannes et al. [13] performed a single-generation selection experiment with epigenetically variable lines. An extension of this work over multiple generations should shed light on these theoretical results, and help characterize responses to natural selection in wild populations. However, as opposed to the constant epimutation rates modeled here, environmentally-sensitive epimutation rates may yield a less predictable selective response. If the selective pressures also influence the rates of epimutation, mutational processes may serve to either magnify the reductive effect or counter it, depending on how the rates of epimutation are affected. Genetic variation may also interact with epigenetic variation in the response to selection. One possibility is that epigenetic variation plays a role in the initial response to selection, while genetic variation shifts more slowly but ultimately produces a phenotypic distribution in the population with less mutational load [26]. Further empirical and theoretical research on the epigenetic response to selection should help clarify how natural populations will respond to novel selective pressures: an issue of great concern as humans continue to rapidly modify landscapes around the world.

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Author Contributions

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References

1. Herrera CM, Bazaga P (2010) Epigenetic differentiation and relationship to adaptive genetic divergence in diverse populations of the viola vita caucalensis. New Phytol 187: 967–76.
2. Herrera CM, Bazaga P (2011) Untangling individual variation in natural populations: ecological, genetic and epigenetic correlates of long-term inequality in herbivory. Mol Ecol 20: 1675–89.
3. Herrera CM, Bazaga P (2013) Epigenetic correlates of plant phenotypic plasticity: DNA methylation differs between prickly and nonprickly leaves in heterophylyllus flex aquelimum (aqulifoliacae) trees. Bot J Linn Soc 171: 441–452.
4. Lira-Medeiros CF, Parisod C, Fernandes RA, Mata CS, Cardoso MA, et al. (2010) Epigenetic variation in mangrove plants occurring in contrasting natural environment. PLOS ONE 5: e10326.
5. Liu S, Sun K, Jiang T, Ho JP, Liu B, et al. (2012) Natural epigenetic variation in the female great roundback bat (hipposideros armiger) populations. Mol Genet Genomics 287: 643–50.
6. Silvaibe AB, Trountin C, Cortijo-S, Barau, J, Del Ben LEV, et al. (2013) Extensive natural epigenetic variation at a de novo originated gene. PLOS Genet 9: e1003437.
7. Acar M, Beckei A, van Oudenaarden A (2005) Enhancement of cellular memory by reducing stochastic transitions. Nature 435: 229–32.
8. Acar M, Mettetal JT, van Oudenaarden A (2008) Stochastic switching as a survival strategy in fluctuating environments. Nat Genet 40: 471–10.
9. Beaumont HJE, Gallee J, Kost C, Ferguson GC, Rainey PB (2009) Experimental evolution of bet hedging. Nature 462: 90–3.
10. Bender J, Fink GR (1995) Epigenetic control of an endogenous gene family is revealed by a novel blue fluorescent mutant of arabidopsis. Cell 83: 725–34.
11. Daxinger L, Whitelaw E (2012) Understanding transgenerational epigenetic inheritance, and selection in plant populations. Cold Spring Harb Symp Quant Biol 77: 97–104.
12. Day T, Bonduriansky R (2011) A unified approach to the evolutionary consequences of genetic and nongenetic inheritance. Am Nat 178: E10–36.
13. Geoghegan JL, Spencer HG (2012) Transgenerational defense induction and epigenetic inheritance in plants. Trends Ecol Evol 27: 618–26.
14. Greer EL, Maures TJ, Ucar D, Hauswirth AG, Mancini E, et al. (2013) Transgenerational epigenetic longevity in caenorhabdites elegans. Nature 479: 365–71.
15. Levy SF, Zie N, Siegal ML (2012) Bet hedging in yeast by heterogeneous, age-correlated expression of a stress protectant. PLoS Biol 10: e1001325.