The prognostic significance of vasohibin-1 expression in patients with prostate cancer

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Background: We recently isolated vasohibin-1 (VASH1), a novel angiogenic molecule that is specifically expressed in activated vascular endothelial cells (ECs), and the status of VASH1 expression has been documented in various cancer angiogenesis. The aim of this study was to assess the prognostic value of VASH1 expression in prostate cancer (PCa).

Methods: In this study, we retrospectively analysed the clinical records and evaluated the VASH1 expression of tumour microvessels in 167 patients with PCa who underwent radical prostatectomy. We immunohistochemically examined the microvessels positive for anti-CD34 as microvessel density (MVD) and the microvessels with activated ECs positive for VASH1 density.

Results: We found that the VASH1 expression was restricted to ECs in the tumour stroma. VASH1 density was significantly associated with pathological T stage, Gleason score and MVD. The 5-year PSA recurrence-free survival rate was 58.8% in patients with higher VASH1 density (≥12 per mm²) and 89.1% in patients with lower VASH1 density (<12 per mm²), respectively (P<0.001). Microvessel density was not an independent predictor of PSA recurrence. Multivariate analysis revealed that high VASH1 density was an independent prognostic indicator of PSA recurrence (P=0.007, HR = 2.950).

Conclusion: VASH1 density represents a clinically relevant predictor of patient prognosis and can be a new biomarker that would provide additional prognostic information in PCa.

Prostate cancer (PCa) is one of the most commonly diagnosed malignant tumours in men and the second leading cause of cancer-related deaths in the United States (Jemal et al, 2010). Nowadays, there are still few effective therapeutic options for advanced PCa (Scher and Sawyers, 2005; Chen et al, 2008). One of the most troublesome aspects of PCa is that androgen-dependent PCa inevitably progresses to highly aggressive and life-threatening castration-resistant prostate cancer after androgen ablation therapy. The key molecular events associated with PCa progression remains to be elucidated.

Angiogenesis, which is the formation of new blood vessel networks, not only plays a role in human normal development but also in pathophysiological conditions such as inflammation and cancer (Folkman, 1971; Sato, 2003). The development and establishment of blood supply has an important role in the development and growth of cancer cells by providing oxygen, nutrients and growth factors. In addition to its traditional role, angiogenesis is one of the potential pathways that contribute to solid tumour progression and metastasis by allowing cancer cells direct access to vessels. In general, angiogenesis is regulated by a balance between stimulatory and inhibitory factors (Sato and Sonoda, 2007, Sato, 2012). Molecules such as CD34, von Willebrand factor and vascular endothelial-cadherin, which are specifically expressed in vascular endothelial cells (ECs), could serve as biomarkers of angiogenesis (Weidner et al, 1991). One of the parameters of angiogenesis in neoplasms is microvessel density.
(MVD). In PCa, several studies indicated that MVD served as a predictor of poorly differentiated tumours and biochemical failure after treatment (Weidner et al, 1993; Bettencourt et al, 1998; Borre et al, 1998; Rubin et al, 1999; Krupski et al, 2000; Josefsson et al, 2005; Concato et al, 2007, 2009; Kosaka et al, 2007). However, to date the prognostic role of MVD in PCa has not been fully characterised. This is because those angiogenic molecules are expressed in quiescent ECs as well as in activated ECs, and thus cannot reflect ‘angiogenic activity’ alone.

We recently isolated vasohibin-1 (VASH1), a novel angiogenic molecule, that is specifically expressed in ECs and upregulated by vascular endothelial growth factor (VEGF) and fibroblast growth factor 2 (FGF-2) (Watanabe et al, 2004; Kimura et al, 2009). Previous studies found that the expression of VASH1 was restricted to ECs of blood vessels in the tumour stroma and correlated with the expression of VEGF and FGF-2 in tumour cells (Hosaka et al, 2009). No one has ever characterised the expression of VASH1 in relation to tumour angiogenesis in PCa. In this study, we will evaluate whether the expression of VASH1 can serve as a more accurate biomarker of tumour angiogenesis compared with MVD.

We will examine the expression of VASH1 and MVD in PCa specimens acquired by primary surgery and investigate whether VASH1 expression reflects angiogenic activity in PCa and if this is related to the clinical outcome.

MATERIALS AND METHODS

Patients selection. Samples of archival paraffin-embedded tissue sections and clinicopathological features were obtained from 167 patients with PCa diagnosed and operated on at Keio University between January 2000 and December 2003. None of the patients had received hormonal treatment before the operation. The characteristics of these patients are shown in Table 1. After radical prostatectomy, patients were followed by serum PSA level and imaging studies. Prostate-specific antigen relapse was defined by an elevation of serum PSA level at three consecutive measurements. After radical prostatectomy, patients were followed by serum PSA level and imaging studies. Prostate-specific antigen relapse was defined by an elevation of serum PSA level at three consecutive measurements.

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Table 1. Correlation of clinicopathological parameters and MVD or VASH1 expression in the 167 study patients

| Characteristic          | No. of patients (%) | MVD (mean ± s.d.) | P-value | VASH1 density (mean ± s.d.) | P-value |
|-------------------------|---------------------|-------------------|---------|----------------------------|---------|
| Age (years)             |                     |                   |         |                           |         |
| <65                     | 99 (59.3)           | 133.1 ± 58.3      | 0.201   | 9.3 ± 7.4                  | 0.101   |
| ≥65                     | 68 (40.7)           | 115.6 ± 41.2      |         | 10.8 ± 7.2                 |         |
| PSA                     |                     |                   |         |                           |         |
| <15                     | 138 (82.6)          | 120.5 ± 47.5      | 0.588   | 9.4 ± 6.8                  | 0.139   |
| ≥15                     | 29 (17.4)           | 133.7 ± 67.9      |         | 12.3 ± 8.9                 |         |
| Gleason score           |                     |                   |         |                           |         |
| ≤6                      | 69 (41.3)           | 105.8 ± 42.2      | <0.001  | 7.5 ± 5.6                  | <0.001  |
| ≥7                      | 98 (58.7)           | 134.6 ± 54.4      |         | 11.6 ± 7.9                 |         |
| Pathological T stage    |                     |                   |         |                           |         |
| ≥pT2                    | 113 (67.7)          | 111.2 ± 46.3      | <0.001  | 8.6 ± 6.5                  | <0.001  |
| ≥pT3                    | 54 (32.3)           | 146.8 ± 54.3      |         | 12.6 ± 8.2                 |         |

Abbreviations: MVD = microvessel density; PSA = prostate-specific antigen; VASH1 = vasohibin-1.
Evaluation of immunostaining. Two authors independently evaluated immunoreactivity. They were blinded to the clinical course of the patients and the average of the numbers counted by the two investigators was used for subsequent analyses. Olympus IX71 (Olympus, Tokyo, Japan) was used for the analysis. The microvessels within the tumour were counted. Microvessels were identified based on their architecture, lumen lined by ECs and complemented by positivity of the ECs for anti-CD34 after scanning the immunostained section at low magnification (×40 and ×100). The areas with the highest number of distinctly highlighted microvessels were selected, and these were counted at high magnification (×200). We evaluated at least six areas in high-power fields and selected one area with the highest number of vessels. Any immunostained EC or cluster separated from adjacent vessels was counted as a single microvessel, even in the absence of vessel lumen. Each single count was defined as the highest number of microvessels identified at the ‘hot spot’ as shown previously (Mikami et al, 2006; Kosaka et al, 2007; Shirotaka et al, 2011; Yoshinaga et al, 2011). The highest number of microvessels in the hot spot was counted for MVD. Vasohibin-1-positive signals were counted in the ‘hot spot’ in which the highest number of vessels positive for anti-CD34 was identified. We regarded the number of VASH1-positive signals per mm² as ‘VASH1 density’ (Tamaki et al, 2009, 2010; Yoshinaga et al, 2011). The median values of MVD and VASH1 density were 119.5 and 9.9 per mm², respectively. We used a median MVD of ≤120 per mm² and a VASH1 density of ≥12 per mm² as the cutoff levels.

Statistical analysis. The associations between each clinicopathological parameter and VASH1 density of the tumour were analysed. These associations were validated using χ² test or Mann–Whitney U-test. Biochemical recurrence-free survival was estimated using the Kaplan–Meier method and was compared by using the log-rank test. Multivariate analysis was performed using the Cox proportional hazard model. Differences among groups were regarded as significant when P<0.05. These analyses were performed with the SPSS version 18.0 statistical software package (IBM corporation, New York, NY, USA).

RESULTS

Patient characteristics and VASH1 expression in PCa. Table 1 shows the clinicopathological characteristics of the patients and their association with MVD or VASH1 density in our study population. The median age of the patients was 66.6 years (range 46–75 years). Pathological T (pT) stage was ≤pT2 in 113 cases (67.7%) and ≥pT3 in 54 cases (32.3%). Gleason score (GS) was ≤6 in 69 cases (41.3%) and ≥7 in 98 cases (58.7%). During a median follow-up of 4.9 years, 48 patients (28.7%) experienced PSA recurrence. To elucidate the biological significance of VASH1 in PCa, we examined the expression of VASH1 by immunohistochemical staining (Figure 1). Although CD34 staining of vascular ECs in benign prostatic tissue adjacent to cancerous tissue was
positive as shown previously (Figure 1A), VASH1 staining of vascular ECs was negative or negligible (Figure 1B). Vasohibin-1 staining of vascular ECs was negative or negligible in low pT stage and low GS PCa (Figure 1B). Strong VASH1 staining was detected in many cases (Figure 1F). Strong VASH1 staining was observed in ECs of microvessels in the tumour lesion of high pT stage and low GS PCa (Figure 1D). On the other hand, in high pT stage PCa, strong VASH1 staining of vascular ECs was negative or negligible (Figure 1B). Vasohibin-1 was positive as shown previously (Figure 1A), VASH1 staining of vascular ECs was negative or negligible in low pT stage and low GS PCa (Figure 1B). Strong VASH1 staining was detected in many cases (Figure 1F). Strong VASH1 staining was observed in ECs of microvessels in the tumour lesion of high pT stage and low GS PCa (Figure 1D). On the other hand, in high pT stage PCa, strong VASH1 staining of vascular ECs was negative or negligible (Figure 1B). Vasohibin-1 was positive as shown previously (Figure 1A), VASH1 staining of vascular ECs was negative or negligible in low pT stage and low GS PCa (Figure 1B). Strong VASH1 staining was detected in many cases (Figure 1F). Strong VASH1 staining was observed in ECs of microvessels in the tumour lesion of high pT stage and low GS PCa (Figure 1D). On the other hand, in high pT stage PCa, strong VASH1 staining of vascular ECs was negative or negligible (Figure 1B). Vasohibin-1 was positive as shown previously (Figure 1A), VASH1 staining of vascular ECs was negative or negligible in low pT stage and low GS PCa (Figure 1B). Strong VASH1 staining was detected in many cases (Figure 1F). Strong VASH1 staining was observed in ECs of microvessels in the tumour lesion of high pT stage and low GS PCa (Figure 1D). On the other hand, in high pT stage PCa, strong VASH1 staining of vascular ECs was negative or negligible (Figure 1B). Vasohibin-1 was positive as shown previously (Figure 1A), VASH1 staining of vascular ECs was negative or negligible in low pT stage and low GS PCa (Figure 1B). Strong VASH1 staining was detected in many cases (Figure 1F). Strong VASH1 staining was observed in ECs of microvessels in the tumour lesion of high pT stage and low GS PCa (Figure 1D). On the other hand, in high pT stage PCa, strong VASH1 staining of vascular ECs was negative or negligible (Figure 1B). Vasohibin-1 was positive as shown previously (Figure 1A), VASH1 staining of vascular ECs was negative or negligible in low pT stage and low GS PCa (Figure 1B). Strong VASH1 staining was detected in many cases (Figure 1F). Strong VASH1 staining was observed in ECs of microvessels in the tumour lesion of high pT stage and low GS PCa (Figure 1D). On the other hand, in high pT stage PCa, strong VASH1 staining of vascular ECs was negative or negligible (Figure 1B). Vasohibin-1 was positive as shown previously (Figure 1A), VASH1 staining of vascular ECs was negative or negligible in low pT stage and low GS PCa (Figure 1B). Strong VASH1 staining was detected in many cases (Figure 1F). Strong VASH1 staining was observed in ECs of microvessels in the tumour lesion of high pT stage and low GS PCa (Figure 1D). On the other hand, in high pT stage PCa, strong VASH1 staining of vascular ECs was negative or negligible (Figure 1B). Vasohibin-1 was positive as shown previously (Figure 1A), VASH1 staining of vascular ECs was negative or negligible in low pT stage and low GS PCa (Figure 1B). Strong VASH1 staining was detected in many cases (Figure 1F). Strong VASH1 staining was observed in ECs of microvessels in the tumour lesion of high pT stage and low GS PCa (Figure 1D). On the other hand, in high pT stage PCa, strong VASH1 staining of vascular ECs was negative or negligible (Figure 1B).

Table 2. Univariate and multivariate analysis for PSA recurrence-free survival in 167 PCa patients

| Characteristic | Univariate | Multivariate |
|---------------|------------|-------------|
| Age (years)   |            |             |
| <65           | 0.525      |             |
| ≥65           |            |             |
| PSA           | <0.001     |             |
| <15           |            |             |
| ≥15           |            |             |
| Gleason score | 0.073      |             |
| ≤6            |            |             |
| >6            |            |             |
| Pathological T stage | <0.001 | <0.001 |
| ≥pT2         |            |             |
| ≥pT3         | 4.667 (2.358-9.234) |             |
| MVD          | <0.001     |             |
| <120 per mm² |            |             |
| ≥120 per mm² |            |             |
| VASH1 density | <0.001     | 0.007       |
| <12 per mm²  |            |             |
| ≥12 per mm²  | 2.950 (1.349-6.449) |             |

Table 3. Clinicopathological parameters in 167 patients according to the level of the VASH1 density

| Characteristic | No. of patients (%) | Patients with VASH1 density | Patients with VASH1 density | P-value |
|---------------|---------------------|-----------------------------|-----------------------------|---------|
| Age (years)   |                     |                             |                             |         |
| <65           | 53 (63.9)           | 46 (54.8)                   | 0.271                       |         |
| ≥65           | 30 (36.1)           | 38 (45.2)                   |                             |         |
| PSA           |                     |                             |                             |         |
| <15           | 72 (86.7)           | 66 (78.6)                   | 0.220                       |         |
| ≥15           | 11 (13.3)           | 18 (21.4)                   |                             |         |
| Gleason score |                     |                             |                             |         |
| ≤6            | 48 (57.8)           | 21 (25.0)                   | <0.001                      |         |
| >6            | 35 (42.2)           | 63 (75.0)                   |                             |         |
| Pathological T stage |             |                             |                             |         |
| ≥pT2          | 65 (78.3)           | 48 (57.1)                   | 0.005                       |         |
| ≥pT3          | 18 (21.7)           | 36 (42.9)                   |                             |         |
| MVD           |                     |                             |                             |         |
| <120 per mm² | 62 (74.7)           | 21 (25.0)                   | <0.001                      |         |
| ≥120 per mm² | 21 (25.3)           | 63 (75.0)                   |                             |         |

Abbreviations: CI = confidence interval; HR = hazard ratio; MVD = microvessel density; PCa = prostate cancer; PSA = prostate-specific antigen; VASH1 = vasohibin-1.

Risk stratification for PCa according to pT stage and VASH1 density. We distributed the patients into three different groups according to pT stage and VASH1 density, which were the two statistically significant variables found by the multivariate Cox regression analysis (Figure 2C). The relative risk of death was calculated with the formula, exp (1.540 × pT stage + 1.082 × VASH1 density) for PSA recurrence-free survival. In this equation, the pT stage equaled 1 if the pT stage was pT3 or more, and it equaled 0 if the pT stage was pT2 or less. VASH1 density equaled 1 if VASH1 density was ≥12 per mm² and 0 if <12 per mm². On the basis of the relative risk of death, patients with PCa were divided into three risk groups: low (relative risk of PSA recurrence = 1), intermediate (8.02-12.7 for PSA recurrence-free survival) and high (13.8 for PSA recurrence-free survival). According to the risk stratification for PCa based on prognostic factors, 65 patients (38.9%) were in the low-risk group (low pT stage and low VASH1 density), 66 patients (39.5%) were in the high-risk group (high pT stage and high VASH1 density) and 36 patients (21.6%) were in the intermediate-risk group (all others).

The 5-year PSA recurrence-free survival was 98.1% in the low-risk group, 69.7% in the intermediate-risk group and 33.7% in the high-risk group, respectively. The differences among the groups were statistically significant (P = 0.001). The 5-year Kaplan–Meier PSA recurrence-free survival rate was 58.8% in patients with high VASH1 density compared with 89.1% (P = 0.001) in their counterparts (Figure 2A).
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Those with high VASH1 density and 80.1% for those with low recurrence-free survival rate in the high MVD group was 43.1% for (25.0%) had low VASH1 density (Figure 3B). The 5-year PSA group, 63 patients (75.0%) had high VASH1 density and 21 patients (25.3%) had high MVD (Figure 3). In the low MVD group, 21 patients (25.3%) had high VASH1 density and 62 patients (74.7%) had low VASH1 density. The difference between analysis of MVD according to high or low VASH1 density.

Univariate analysis revealed that high MVD was one of the significant predictors of PSA recurrence (Figure 2B), while multivariate analysis showed that high MVD was not an independent predictor of PSA recurrence. Then, we re-evaluated univariate analysis of MVD according to high or low VASH1 density (Figure 3). In the low MVD group, 21 patients (25.3%) had high VASH1 density and 62 patients (74.7%) had low VASH1 density (Figure 3A). The 5-year PSA recurrence-free survival rate in the low MVD group was 79.9% for those with high VASH1 density and 94.9% for those with low VASH1 density. The difference between the groups was statistically significant (P<0.001). In the high MVD group, 63 patients (75.0%) had high VASH1 density and 21 patients (25.0%) had low VASH1 density (Figure 3B). The 5-year PSA recurrence-free survival rate in the high MVD group was 43.1% for those with high VASH1 density and 80.1% for those with low VASH1 density. The differences were significant (P<0.001). These results indicated that MVD in PCa could be subdivided into two subclasses for PSA recurrence according to VASH1 density.

and P<0.001 for intermediate- vs high-risk group in PSA recurrence-free survival).

Microvessel density could be divided into two subclasses for PSA recurrence according to VASH1 density in PCa patients. Univariate analysis revealed that high MVD was one of the significant predictors of PSA recurrence (Figure 2B), while multivariate analysis showed that high MVD was not an independent predictor of PSA recurrence. Then, we re-evaluated univariate analysis of MVD according to high or low VASH1 density (Figure 3). In the low MVD group, 21 patients (25.3%) had high VASH1 density and 62 patients (74.7%) had low VASH1 density (Figure 3A). The 5-year PSA recurrence-free survival rate in the low MVD group was 79.9% for those with high VASH1 density and 94.9% for those with low VASH1 density. The difference between the groups was statistically significant (P<0.001). In the high MVD group, 63 patients (75.0%) had high VASH1 density and 21 patients (25.0%) had low VASH1 density (Figure 3B). The 5-year PSA recurrence-free survival rate in the high MVD group was 43.1% for those with high VASH1 density and 80.1% for those with low VASH1 density. The differences were significant (P<0.001). These results indicated that MVD in PCa could be subdivided into two subclasses for PSA recurrence according to VASH1 density.

DISCUSSION

In this study, we retrospectively evaluated the impact of VASH1 expression by immunohistochemical staining in a series of patients with PCa treated in a single centre. Our results suggested that VASH1 expression was a prognostic indicator in addition to other standard factors such as pT stage. High VASH1 density was related to shorter patient PSA recurrence-free survival. To the best of our knowledge, this is the first study evaluating the prognostic value of VASH1 expression in patients with prostate cancer.

Angiogenesis has a critical role in tumour growth and metastasis (Folkman, 1971; Chung et al, 2010). Recent studies revealed significant roles for angiogenesis in the prediction of survival in patients with different malignancies (Sato, 2003, 2012; Chung et al, 2010). One of the biomarkers that could reflect angiogenic aggressiveness was MVD (Weidner et al, 1991; Bochner et al, 1995; Kosaka et al, 2007; Shirotake et al, 2011). Several studies on PCa indicated that the status of MVD was associated with clinical features, such as GS and pathological stage, and could be an independent prognostic factor of patient survival (Weidner et al, 1993; Bettencourt et al, 1998; Borre et al, 1998; Rubin et al, 1999; Krupski et al, 2000; Josefsson et al, 2005; Concato et al, 2007, 2009; Kosaka et al, 2007). However, to date evidence of the prognostic role of MVD in PCa is contradictory, suggesting that the prognostic impact of MVD might be controversial. In this study, univariate analysis revealed that there was significant association between MVD and PSA recurrence. However, multivariate analysis including VASH1 density showed that MVD did not have a prognostic significance in PCa progression. One of the reasons might be because MVD corresponds to the number of accomplished vessels and includes vessels without the potential of neovascularisation in PCa. Our previous reports showed that VASH1 has been isolated from VEGF-inducible genes in ECs present in newly formed blood vessels behind the sprouting front where angiogenesis terminates (Watanabe et al, 2004; Sato and Sonoda, 2007; Hosaka et al, 2009). Recently, we have reported that histologic evidence of VASH1 expression has been found in...
samples from patients with endometrial carcinoma (Yoshinaga et al, 2008), breast cancer (Tamaki et al, 2009, 2010) and cervical carcinoma (Yoshinaga et al, 2011). Our reports indicated that VASH1 expression was associated with tumour grade and histological type of carcinomas. Moreover, it was reported that VASH1 expression tended to be concordant with MVD, although partial dissociation was observed in some patients with breast carcinoma. VASH1 expression was significantly higher in invasive breast carcinoma, although no significant difference was observed in the level of MVD between patients with invasive disease or not (Tamaki et al, 2010). These studies suggested that an evaluation of the number of VASH1-positive vessels may become one of the prognostic biomarkers for metastasis and prognosis. These results indicate that VASH1 could become a new molecular biomarker of the angiogenic heterogeneity of tumours. In this study, we demonstrated that the prognostic value of MVD depends on the level of VASH1 density.

Our study demonstrated that PCa with a higher number of VASH1-positive vessels tended to have a poor prognosis. We found a significant correlation among VASH1 density, GS and pT stage. Multivariate analysis showed that high VASH1 density was an independent prognostic factor, suggesting that the status of VASH1 density could serve as a biomarker of the malignant potential of tumour angiogenesis. These results suggest that the level of VASH1 expression may influence the clinical course of prostate cancer progression. Moreover, VASH1 may become a molecular target in PCa patients.

Using VASH1 density and other independent indicators, we established a prognostic risk stratification for PCa. Patients were stratified into three groups according to statistical modelling based on the relative risk associated with the prognostic indicators derived from multivariate analysis. As shown in Figure 2C among patients with PCa, the prognosis of patients with higher pT stage and high VASH1 density was worse than that of the other groups (P<0.001). This stratification made it possible to predict PSA recurrence more accurately, suggesting more appropriate follow-up including PSA follow-up interval.

In conclusion, these results indicated that VASH1 can serve as a new biomarker for predicting PCa progression and could become a molecular target in PCa, especially for targeting tumour angiogenesis.

ACKNOWLEDGEMENTS

We thank colleagues from the Department of Vascular Biology, Institute of Development, Aging and Cancer, Tohoku University for their excellent technical assistance in purifying anti-human VASH1 mAb. This work was supported in part by grants-in-aid for Scientific Research from the Ministry of Education, Culture, Sports, Science, and Technology of Japan, by the Prostate Research Fund in Japan, the Japan Urological Association (5th Young Research Grant 2011 to TK) and the Cooperative Research Project Program 2011, 2012 of Joint Usage/Research Center at the Institute of Development, Aging and Cancer, Tohoku University.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

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