Physicochemical Properties and Trace Elements of Soils in Mambilla Plateau of Northeastern Nigeria

Isaac Ernest¹, Maina Humphrey Mayi², Donatus Raymond Bwano°, Riki Yohanna Emmanuel⁴, Joseph Bala²

¹Department of Chemistry, Taraba State University, Jalingo, Nigeria
²Department of Chemistry, Modibbo Adama University of Technology, Yola, Nigeria
°Department of Chemical Sciences, Federal University Wukari, Wukari, Nigeria
⁴Department of Basic Sciences, College of Agriculture, Jalingo, Nigeria

Email address:
donatus_r28@yahoo.com (D. R. Bwano)
°Corresponding author

To cite this article:
Isaac Ernest, Maina Humphrey Mayi, Donatus Raymond Bwano, Riki Emmanuel Yohanna, Joseph Bala. Physicochemical Properties and Trace Elements of Soils in Mambilla Plateau of Northeastern Nigeria. *World Journal of Applied Chemistry*. Vol. 3, No. 3, 2018, pp. 72-82. doi: 10.11648/j.wjac.20180303.11

Received: June 6, 2018; Accepted: August 29, 2018; Published: October 13, 2018

Abstract: The soil samples from Mambilla Plateau were analyzed to determine the physicochemical properties of elements present by energy dispersive X-Ray Fluorescence. The physical and chemical parameters determined were colour, pH, electrical conductivity, specific gravity and elemental identifications of Cr₂O₃, CuO, NiO and V₂O₅ respectively. The pH values for soils of the study area are high between 3.96-4.30 being acidic, electrical conductivity values ranged from 81.79 – 238 mS/cm with higher electrical conductivity value at N1 having 238 mS/cm and lower electrical conductivity at S1 having 81.79 uS/cm respectively. The soil are yellow, red and gray; with hue steps of 2.5YR, 5YR, 7.5YR, and 10YR, a step values between 3-7 and the chroma range of 1- 6. The specific gravity which also is the relative density range from 2.00-2.50. The percentage elemental contents identified for trace elements decreased in the order V₂O₅ > Cr₂O₃ > NiO > CuO. The highest percentage concentration of element found was V₂O₅ at N4 0.29% while, CuO showed minimum concentration of 0.01%. The entire soils indicated high acidity levels due to the pH results tested, with spatial variation of the elements oxides contents higher at N1, N2, N3, N4 and decreasing through E1, E2, E3 and E4 to S1, S2, S3 and S4 at lower concentrations.

Keywords: Mambilla, Physicochemical, Trace Element, Soil, Standard Notation, Northeastern Nigeria

1. Introduction

Soil is the topmost layer of the earth surface that is diverse in complexity with mixtures of minerals and organic materials, aqueous and gaseous components, which are capable of supporting plants and animals’ life. [1-3]. Soil survey generates host of information on soils and other environmental factors. In the past, these soils information were primarily used to develop land use plans, to support increased crop production. However, the use of soil information for non-agricultural purpose is on the increased. There is increased awareness of the use of soil information as an essential input in the evaluation, prediction of minerals deposits and effect of land use on environmental degradation and sustainability. In Nigeria, like most developing countries, one primary constraint to sustainable and successful agricultural programme is the lack of knowledge about the soil resources and how to manage them. [4]. There is increasing need for information on soils as a means to its maximum uses, greater and healthy food production. The variation in soil properties due to land use and management, and their consequences to the production capacity; has been subject of research in the past for a range of climatic and edaphic conditions. [5]. A study by Amuyou et al. [6] on spatial variability of soil properties, observed topographic influence over the distribution of soil physical and chemical properties such as pH, organic carbon, exchangeable elements, cation exchange capacity and total nitrogen. [6].
The plateau covers an area of over 9,389 square kilometers (3,625 square meters) with daytime temperature that hardly exceeds 25°C (77.0°F). [14]. The Mambilla Plateau is selected for the study, in view of its peerless splendor and natural architecture and landscaping with hilly deep gorges. The climate of the Plateau is comparatively cold, associated with frequent and heavy rainfall due to orographic activities on the Plateau. The vegetation on the plateau comprises low grasses with trees being noticeably absent except for manmade forest [14, 15]. There was no known research conducted to assess the nutrient and pollutant elements in this abundant tourism potential. As at the time of this study Sardauna Local Government Area of Taraba State has no established information on status as well as the variability of soil physical and chemical properties in this natural architecture and landscaping of the terrain. This study was set to fill the gap and initiate further quest for mineral exploration and toxicity studies of soils in the area. The results of this study will ascertain the authenticity of high speculation of mineral deposit including gold, silver, and precious stones in the Plateau. The result from this research would assist in exploiting, correcting and preserving the enriching features of the plateau for the maximum agricultural and mineral utilization, enhancement of revenue earnings for the state in particular and the country at large. The energy dispersive X-ray fluorescence (EDXRF) is a multi-element analytical instrument that is highly sensitive Virendra and Agrawal. [16]. It will be used to achieve this analytical task.

2. Materials and Methods

2.1. Materials (Instruments/Apparatus)

Munsell soil colour chart, funnels, beakers, filter papers, 2mm sieve, glass rod, volumetric flask and analytical balance, measuring cylinder, pestle and mortar, Sensio pH meter, Sensio Electrical conductivity meter, and MiniPal4 energy-dispersive x-ray fluorescence (EDXRF) spectrometer model PW 4052/47B. Additionally the following tools were used: cutlass, hoe, measuring rule, clean dry polyethylene bags and corer.

2.2. The Study Area

The study area, Mambila Plateau is found in the southeastern part of Taraba state of Nigeria in Sardauna Local Government Area. It measures about 96km (60m) along its curved length, it is 40km (25m) wide and bounded by an escarpment that is about 900m (2,953 ft) high in some places. The plateau covers an area of over 9,389 square kilometers (3,625 square meters) with daytime temperature that hardly exceeds 25°C (77.0°F). [14].
2.3. Sampling and Sample Preparation

The methods of Virendra and Agrawal [16], ISO [17], Rayment and Higginson [8] were used with some modifications in the process of soil sampling. Samples of soils were collected starting from the foot of the mountain after clearing grasses with cutlass, the digging was done with a simple hoe at (0 – 25) cm depth, and a ruler used for the measurement. For each location, three soil samples weighing about 500 g were collected at three random points using the corer. Sampling began from Mayo-Selbe Village, at a distance of 3km interval moving through hilly parts with deep gorges from one panoramic view to the other, to the first tip settlement Maisamari village. The sampling process continued through Nguroje, Yelwa, Lekkitaba, Mayondaga, Tunga Shaibu, Kakara, Mbanga, Mayodale, Papa settlements and Gembu town. A total of twelve settlements with thirty six (36) bulk samples was collected, bagged in clean dry polyethylene and transported to the laboratory. The pre analysis treatment of all the bulk samples collected was done immediately at the laboratory. All particles in the soils like stones, pieces of wood, particle rocks, gravels, organic debris were removed. The soils were sieved through a 2 mm mesh, mixed and quartered to obtain a total of (12) representative samples one for each location, oven dried and finally introduced into a clean dry polyethylene bag [17, 18]. The soil samples were coded and labelled as follows: The northern settlements N₁, N₂, N₃, N₄; eastern settlements E₁, E₂, E₃, E₄ while the southern part are S₁, S₂, S₃, and S₄ respectively.
2.4. Determination of Soil Colour and Analytical Studies of Soil

The colour of each soil sample was determined by comparing the soil colour in its field condition to a series of colour clips in a booklet called soil colour chart using the methods of Munsell. [19, 20]. Each chip was described uniquely by its position on the three axes of colours; hue or spectral colour, value or degree of darkness, and chroma, purity or strength of colour. The soil sample paste was held behind the page of the chips solution so that the soil colour matches any of the colours of chips seen through the holes punched on the page, then the colour that resonates was recorded as its colour both in name and standard notation.

2.5. The pH Measurement

The method of Onanuga and Shehzad et al. [21, 22] was adopted, about 5g of each air-dried and sieved soil samples were added to 40cm³ distilled water and stir with glass rod for 5 minutes, for equilibration of the contents. The sensio pH meter was standardized using standard buffer 4.0, 7.0 and 10.0 at 25°C with an accuracy of ±0.02 pH units for the electrode calibration. The pH of the aqueous sample suspension was read off after inserting the sensio pH meter electrode into each soil suspension and gently swirling the beaker so as to ensure accurate reading from equal surrounding condition. The reading was taken after it had attained stability, and test were repeated to obtain an average data of three readings at 25°C.

2.6. Electrical Conductivity Measurement

This was carried out as described by Onanuga and Shehzad et al. [21, 22]. The Sensio electrical conductivity meter was checked and the electrode cleansed using distilled and deionized water, and dipped into the aqueous sample solution to test for dissociated charge ions present. The reading were then taken at a time the pointer becomes steady. All readings were replicated trice to obtain average
value.

2.7. Specific Gravity Measurement

The specific gravity of the sample was determined using a specific gravity bottle as described by a bureau of Indian standards [17, 23]. About 12 grams of the sample was used. The specific gravity of the sample was calculated from using the formula below:

\[ S.G = \frac{(W_2 - W_1)}{((W_4 - W_1)(W_3 - W_2))} \]

Where \( W_1 \) weight of bottle; \( W_2 \) weight of bottle + sample; \( W_3 \) weight of bottle + sample + water; and \( W_4 \) weight of bottle + water.

3. Results and Discussions

Table 1. Physicochemical properties of soil samples collected.

| Location code | Soil colour | Standard Notation | Specific gravity | pH   | Conductivity mS/cm |
|---------------|-------------|-------------------|------------------|------|--------------------|
| E1            | drb         | 5YR3/4            | 2.21             | 4.12 | 93.0               |
| E2            | rb          | 5YR5/4            | 2.12             | 4.26 | 89.30              |
| E3            | b           | 7.5YR4/4          | 2.33             | 4.24 | 153.60             |
| E4            | drg         | 5YR4/2            | 2.50             | 4.11 | 136.70             |
| Range         |             |                   | 2.12-2.50        | 4.11-4.26 | 89.30-153.60       |
| Mean SD       |             |                   | 2.29±0.14        | 4.18±0.07 | 118.15±29.68      |
| N1            | lg          | 10YR7/2           | 2.35             | 4.52 | 238.00             |
| N2            | drb         | 5YR3/4            | 2.00             | 3.93 | 183.20             |
| N3            | yr          | 5YR5/6            | 2.09             | 4.09 | 174.30             |
| N4            | rb          | 5YR5/4            | 2.31             | 3.62 | 93.00              |
| Range         |             |                   | 2.00-2.35        | 3.62-4.52 | 93-238.00       |
| Mean SD       |             |                   | 2.19±0.15        | 4.04±0.32 | 172.13±57.78      |
| S1            | wr          | 2.5YR5/2          | 2.24             | 4.29 | 81.79              |
| S2            | yr          | 5YR5/6            | 2.36             | 4.30 | 101.90             |
| S3            | g           | 5YR6/1            | 2.39             | 4.16 | 120.10             |
| S4            | sb          | 7.5YR5/6          | 2.37             | 3.96 | 100.20             |
| Range         |             |                   | 2.24-2.39        | 3.96-4.30 | 81.79-120.10    |
| Mean SD       |             |                   | 2.34±0.09        | 4.18±0.13 | 100.10±46.17     |

b = brown; drb = dark reddish brown; drg = dark reddish gray; g = gray; lg = light gray; sb = strong brown; rb = reddish brown; wr = weak red; yr = yellow red;

Figure 3. The pH values for soils.
3.1. Physicochemical Properties

Table 1 presents the physicochemical properties of the sampled soils studied. The colour of the soil varied from yellow to red and gray with soil hues of 2.5YR, 5YR, 7.5YR and 10YR. The entire soil also showed value of 3 to 7 and chroma range of 1-6. The soil colour of eastern samples points showed the hue difference of one unit (2.5) interval at E3 7.5YR4/4 against E1 5YR3/4 and E2 5YR5/4 and E4 5YR4/2 that are all reddish yellow in standard notation, the hue is 5, value is 3-5 and chroma 2, 4. The ordinary mixed brown colours of dark reddish brown, reddish brown, brown and dark reddish gray of these soils sampled were perhaps due to high presence of magnetite content in the soil. [24]. Likewise the northern sampled points had a hue 5 and 10, value 3, 5, and 7 and chroma 2, 4, and 6 with mixed colours. N1 10YR7/2 showed gray colour, N2 5YR3/4 indicated dark reddish brown, as sample N3 5YR5/6 is yellowish red, N4 5YR5/4 was reddish brown. The Samples S1 2.5YR5/2 and S2 5YR5/6 were, weak red and reddish yellow colours respectively. S3 5YR6/1 was gray, but S4 7.5YR5/6 strong brown. The hue is 2.5, 5 and 7.5, value is between 5, 6 and chroma 1, 2 and 6 respectively. The yellow and reddish colours displayed by some samples may be due to high iron oxides present in the soil samples. [25, 26]. The development and distribution of colours may be due to the extensive and various minerals that results from chemical and biological weathering, especially redox reaction. The primary minerals in soil parent material are weathered and thus the elements combine into new and colourful compounds, forming secondary minerals with a yellow or red colour, the soil organic matter also decomposes into black and brown compounds. [27]. Based on these results, the difference in hue chroma is by 1 and 2 that indicates the chroma contrast as prominent as classified by Munsell. [20]. Figure 3 present the pH of soils from all sampled points analyzed for the study, gave an entire range of pH 3.62 – 4.52. The eastern locations have pH range between 4.11-4.26 at E1-E4, with an average of 4.18. The northern samples are a range of pH 3.63-4.52 with mean value of 4.04 also indicating strong acid soil. The southern soil samples are also acidic with a pH range of 3.96-4.30. The result of this study is in tandem with a pH of 4.44-5.59 reported by Imadojemu et al. [28]. Jarrod [29] reported that, a mineral soil of pH 6.0 -7.0 and pH 5.5 for organic soils will be good for most agricultural crops. Furthermore, soils with high acidity can lead to toxic amounts of aluminium and manganese. Haynes and Naidu [30], also reported that soil pH has effects on plant growth, metal ion solubility, microbial activity and clay dispersion. Moeckel et al. [26] established that, water moving through the soil removes the bases compounds and substitutes them with hydrogen ions making the soil acidic and unsuitable for agriculture as a result of the leaching process. It was observed that the low farming activities in the area could be attributed to the soil status. In Figure 4, generally the values for electrical conductivity of the soil samples tested showed the lowest and highest conductivity range of 81.79 - 238 mS/cm respectively. The Electrical conductivity is a numerical expression of the ability of an aqueous solution to carry an electrical current at equilibrium in a water suspension of soil, sludge or bio-waste. [31].
northern location is the most conductive with \( N_1 \) tested 238 mSc/cm and \( N_2 \) 93 mSc/cm. In the southern parts \( S_1 \) is 81.79 mSc/cm while \( S_2 \) gave 120.10 mSc/cm. Reihm et al. [32] emphasized that, the guideline for a suitable electrical conductivity that support productive soil is between 200-1200 mSc/cm. Similarly Pravin et al. [33] stated that soil conductivity correlates with soil properties that affect soil texture, organic matter level, crop productivity, cation exchange capacity, drainage conditions and salinity, and subsoil characteristics. In addition soil with conductivity less than 200 uS/cm will have no enough elements to support crop production; thus show a sterile soil with little microbial activity. [34, 35, 36]. On the other hand, Oleveira et al. [37]

3.2. Percentage Concentrations of Trace Elements

Table 2 shows the trace elements analyzed at various samples sites. The trace metal oxides detected in this study were \( \text{Cr}_2\text{O}_3 \), \( \text{CuO} \), \( \text{NiO} \) and \( \text{V}_2\text{O}_5 \). The concentration of trace elements in this study indicated decreased order from vanadium with the highest concentration \( \text{V}_2\text{O}_5 > \text{Cr}_2\text{O}_3 > \text{NiO} > \text{CuO} \) to copper showing the minimum concentration. However, other elements like \( \text{Pb}, \text{Cd}, \text{Hg}, \text{As}, \text{Zn} \) were not detected in any of the samples studied; perhaps their availability might be at low concentration that are below detection limit of the instrument used for the analysis. One possible explanation is that the high pH level of the soil could have reduced their presence through precipitation or complex formation with Fe-Mn oxyhydroxides. [39]. As seen in Figure 6 above, the concentration of chromium detected in the samples was in the range 0.04% - 0.25% (400-2500 µg/g). The percentage mean concentrations for \( \text{Cr} \) were 0.15% ±0.06 (1500 µg/g) in both eastern and northern locations. While \( N_1 \) showed the highest concentration at 0.25% (2500 µg/g), \( S_1 \) was the lowest Cr concentration with 0.04% (400 µg/g). These are higher than the (11, 50, 100, 150, 250) mg/kg stipulated standards on agricultural soil by US EPA [40], EPAA [41], WHO [42], EU [43], and CME [44] respectively. Similarly, Lago-Vila et al. [45] on analysis of quarry site soil stated a higher \( \text{Cr} \) level 1300 µg/g in a tested soil. Fosu-Mensah et al [46] on the contrary established a lower chromium level 56.0±1.15 µg/g in soils and vegetation of Korle lagoon. Sexana [27] observes that soils contamination could be attributed to the parent materials of the soils. This reflects contamination from non-irrigated sources from soil parent material, minerals waste derived fertilizers and some fungicides/pesticides commonly applied in agricultural practices around the areas [10, 36, 47, 48].

Figure 7 shows the concentration of copper analysed in the sampled locations ranging from 0.01% – 0.17% (100-1700µg/g). This element recorded significant lower levels of detection compare to other elements in the study, as seen evident by negligible or below detection concentration from the samples sites at \( N_1-N_4 \) and \( E_1 \). The samples \( S_4 \) and \( S_3 \) measured 0.01% (100 µg/g) as that was within the guidelines of FAO/WHO/EU [49, 42, 43]. The concentrations of copper at \( S_1, S_2, E_1, E_2 \) and \( E_4 \) indicated (200<400<1000<1700) µg/g accordingly was higher than the guidelines for \( \text{Cu} \) in agricultural soils, also higher value of \( \text{Cu} \) 12,000 µg/g was relates that when soil conductivity is greater than 3000 uS/cm is saline. Figure 5 illustrates the specific gravity of the soils ranging from 2.00 g/cm\(^3\) to 2.500g/cm\(^3\), with mean values of 2.29±0.14 from the eastern sample locations; 2.19±0.15 from the northern sample locations and 2.34±0.09 in the southern sample points respectively. In a study on soils of Morbi, Khoriya et al. [38] stated a lower values of bulk density1.20-1.66 g/cm\(^3\) and higher values for the particle density of the sampled soils between 2.04 to 3.44 g/cm\(^3\). This result agrees with the specific gravity of soils made up of mineral particles with very low organic matter content stated by Sarka et al. [24].

Table 2. Percentage (%) concentration of trace elements in soils.

| Location code | \( \text{Cr}_2\text{O}_3 \) | \( \text{CuO} \) | \( \text{NiO} \) | \( \text{V}_2\text{O}_5 \) |
|--------------|----------------|----------------|----------------|----------------|
| \( E_1 \)    | 0.16           | 0.10           | 0.11           | 0.23           |
| \( E_2 \)    | 0.13           | 0.10           | 0.12           | 0.15           |
| \( E_3 \)    | 0.11           | ND             | 0.08           | 0.18           |
| \( E_4 \)    | 0.21           | 0.17           | 0.23           | 0.10           |
| Range        | 0.11-0.21      | 0.10-0.17      | 0.08-0.23      | 0.10-0.23      |
| Mean SD      | 0.15±0.06      | 0.12±0.03      | 0.14±0.06      | 0.17±0.05      |
| \( N_1 \)    | 0.08           | ND             | 0.10           | 0.07           |
| \( N_2 \)    | 0.13           | ND             | 0.12           | 0.20           |
| \( N_3 \)    | 0.12           | ND             | 0.17           | 0.20           |
| \( N_4 \)    | 0.25           | ND             | 0.19           | 0.29           |
| Range        | 0.08-0.25      | 0.10-0.19      | 0.07-0.29      | 0.19±0.02      |
| Mean SD      | 0.15±0.06      | 0.15±0.04      | 0.19±0.02      |                |
| \( S_1 \)    | 0.04           | 0.02           | 0.08           | 0.08           |
| \( S_2 \)    | 0.06           | 0.04           | 0.04           | 0.09           |
| \( S_3 \)    | 0.05           | 0.01           | 0.03           | 0.10           |
| \( S_4 \)    | 0.10           | 0.01           | 0.08           | 0.10           |
| Range        | 0.04-0.10      | 0.07           | 0.03-0.08      | 0.08-0.10      |
| Mean SD      | 0.06±0.02      | 0.07±0.11      | 0.06±0.02      | 0.09±0.01      |

Detection: limit >0.001; ND = Not Detection; SD=Standard deviation
reported [45] in serpentine quarry. The average concentration of the availability of copper in the soil may be also as a function of the soil pH and texture [26]. The finer-texture mineral soils generally contain the highest amount of copper, the lowest concentrations are associated with the organic or peat soils. Therefore, as soil pH increases, the availability of these nutrients decreases [24].

Figure 8 Nickel was also detected in all samples in a range 0.03% – 0.23% (300-2300) µg/g. The concentration were higher in most samples from N1-N4 and E1-E2. The location E4 yielded the highest 2300 µg/g of Ni analyzed, while S1 was low at 300 µg/g. These concentrations are all above the established guidelines for nickel 45-50 µg/g levels in agricultural soil [42, 43, 44].

Figure 9 presents the concentration of vanadium between 0.07 - 0.29% (700-2900) µg/g. The study revealed N4 sample with 2900 µg/g, as the highest concentrations of an element detected and N1 700 µg/g as the least of vanadium in this study. These values exceeded the FAO established allowed limits of vanadium 75 µg/g in agricultural soil [49]. Other studies that established levels of vanadium above threshold includes, Rasheed et al. [50] reported 490 µg/g, Jayawardana et al gave 820 µg/g [51], and Hernandez and Rodriguez stated 645 µg/g of vanadium accordingly [52]. Manga et al. stated 654 µg/g [53] Kabata-Pendias wrote 460 µg/g as highest vanadium level in metal soil investigation studied. [54]. They further stated, that the behaviour of trace metals in soil environment also depend on their origin due to pedogenic process that transformed lithogenic and anthropogenic metals, which are greatly control by soil properties. In addition, vanadium amounts in soils such as loamy and silty as well as ferralitic can be above that of parent materials [54]. The high content of trace elements detected in the studied samples so far are indications of a mineralized bedrocks occurrence of ores resulting to the geogenic anomalies in the soil, Alloway [55], or in binding nature with other minerals like bauxite and carnotile [56]. Therefore, the appreciable excess concentration of these trace elements studied, will have toxic effects and posed human health risks on other sources like groundwater used and plants grown in those locations.

![Figure 6. Percentage concentration of CrO$_3$ in soil.](image)

![Figure 7. Percentage concentration of CuO in soils.](image)

![Figure 8. Percentage concentration of NiO in soils.](image)
3.3. Statistical Data Analysis

The data analyzed at $F_{3,39} = 2.85 \ p<0.05$. There is no significant variations in the elements concentration and the pH of the studied soils.

4. Conclusion

From this study, the following conclusions can be drawn from data of the physicochemical parameters of the soil samples obtained in this study; colour, pH, specific gravity and electrical conductivity showed that the Mambilla plateau was developed on a highly weathered soil. This may in other way, decrease its organic matter content owing to the soil acidic pH which may greatly decreased the availability of soluble bases such calcium, magnesium, potassium and nitrogen in the soil. The trace elements analyzed were absent in some of the soil samples which perhaps is due to their trace amounts in the location below the detection limit of the instrument used, or as a result of scavenging capabilities of other minerals. The soil formed on the plateau is generally slightly acidic and has high amounts of all the detected elements beyond guidelines may be toxic to plants and human health. The observed stunted growth in plants cultivated, low grasses as if they were trimmed and poor agricultural yields in the area is an indication of poor soil fertility. In addition, further study be conducted to assess the bioavailability and potential human risks of the soil components in the environment as it relates to sources of groundwater and presence in plants cultivated in the locations.

References

[1] Ayoub A. S, McGaw B. A, Shand C. A, Midwood A. J (2003). Photoavailability of Cd and Zn in soil estimated by stable isotope exchange and chemical extraction. Plant and Soil Vol. 252 No. 2, 291-300.

[2] Raymond R. Weil, Nyle C. Brady (2016). Nature and Properties of Soils. 15th Edition. Pearson Education. Columbus.

[3] Abaje I. B (2007). Introduction to soil and vegetation. Personal Touch Productions, Kaduna. 1-56.

[4] Raji, B. A., Chude, V. O. (2007). “Compilation of Existing Soil Series in North-Western Nigeria” Department of Soil Sevens. Ahmadu Bello University Zaria, Nigeria.

[5] Fasina, S. A. (2004), “Influence of Land Utilization of an alisol in South-Western Nigeria”, Journal of Sustainable Agriculture and Environment 6 (2): 171-178.

[6] Amoyou U. A, Eze E. B, Essoka P. A, Effiong J, Egbai O. O (2013). Spatial variability of soil properties in Obudu Mountain Region of southeastern Nigeria. International Journal of Humanities and Social Science Vol. 3 No. 145-149.

[7] Owonubi, A., Raji, B. A. and Oduzze, A. C. (2006), “Morphological and physical properties of soils of a Toposequence in Samaru”, Zaria, Ahmadu Bello University, Nigeria.

[8] Udo, B. I., Ndaeyo N. U. and Harold, K. O. (2006), “Fertility Status and Variability of some soil Developed from different parent materials in the humid region of Nigeria. Department of soil science, University of Uyo, Nigeria.

[9] Stolt, M. H. Bake, J. and Simpson, T. W. (1993), “Soil Landscape Relation in Virginia”. Soil Science Society, American Journal of Soil 57: 414 – 420.

[10] Mokuna, D. L. (1987), “Soil Variability of five Landforms in Michigan”. Soil Survey and Evaluation 7 (1): 25 – 30.

[11] Yu, K. N., Yeung, Z. L. L, Lee, L. Y. L. Stokesa, M. J and Kwok, R. C. W (2002). Determination of Multi-element profiles of soil using energy dispersive X-ray fluorescence (EDXRF), Appl. Radiat. Isot. 57, 279–284.

[12] Buol, S. W., Hole, F. D. and McCracken, R. J. (1973), “Soil Genesis and Classification 1st edition, Ames, I. A: Iowa State University Press 15BN 0-8138-1460-X.

[13] Boul, W. Stanley, Randall J. Southhard, Robert C. Graham, Paul A. McDaniel (2003). Soil genesis and classification. 5th Edition. Wiley Publication.

[14] Bami Yuno (2013). The Mambilla Region in African History (ms ed) Nzakachia: Unpublished p360.

[15] Mambilla Plateau (2011). Nigeria Online Tourism Magazine. GggGoodlife.com.ng.

[16] Virendra Singh, H. M. Agrawal (2012). EDXRF Analysis of Soil Samples to Study the Role of Trace Elements in Optimizing the Yield, International Journal of Modern Engineering Research (IJMER) Vol. 2, Issue. 4, 1454-1458.
[17] ISO (2006) ISO 11464 (2006) Soil quality–pretreatment of samples for physico-chemical Analysis.

[18] Rayment, G. E., Higginson, F. R. (2002). Australian Laboratory Handbook of Soil and Water Chemical Methods, Melbourne, In kata Press.

[19] Munsell, (1973), “Munsell soil Colour Chart” Macbeth Division of Kollomorgen Corporation Baltimore, Maryland 21218, USA.

[20] Munsell, (2009), Soil Colors charts with genuine Munsell color chips, Produced by Munsell color, 4300 44th street Grand Rapids MI, 49512.

[21] Onanuga, K. V. (2008), “Chemical Analysis of Nutrients and Pollutant Elements Contents of soils in two commercial farms in Adamawa state”, Federal University of Technology Yola, Nigeria. Unpublished M-Tech Thesis.

[22] Shehzad Ahmed, Hameed-ur-Rehman, Baharullah, Khatta, Nisar Ahmad, Anum Urooj, Noor Ul Akbar, Muhammad Masab, Wajid Ullah, Ali Rehman and Azam Khan Durrani (2015). Soils of an European Mediterranean area by multivariate analysis. Chemosphere 65:863–872.

[23] ASTM (2008) C127 - 15 Standard Test Method for Relative Density (Specific Gravity) and Absorption of Coarse Aggregate, ASTM International, West Conshohoken, PA, 2015, http://www.astm.org/cgi-bin/resolv.

[24] Sarka, J. P. Landey, R. J. Kalbande, A. R. and Mondol, C. (2011). Characteristic and Classification of Soils of Kathiawar Region of Gujarat as influenced by Topography. Agropedology, Journal of Soil Science, Ahmadu Bello University, Zaria, 83-90.

[25] Brady N. C and Weil, R. R. (2003)."The Nature and Properties of Soils," Pearson Education (Asia).

[26] Moeckel C., Nizzetto L., Di Guardo A., Steinens E., Freppas M., Filippa G., Camporini P., Benner J., Jones, K. C. (2008), “Persistent Organic Pollutants in Boreal and Arid Regions of New Zealand. Sci Total Environ 355:31–47.

[27] Oliveira, F C; Mattiazzo, M E; Marciano, C R; Rossetto, R. (2002). Organic carbon, electrical conductivity, pH, and CEC changes in a typical hapludox after repeated amendments. Braz. J. Soil Sci. 26:505 – 519.

[28] Khorajiya, K. U., Kaneria, S. C., Jadeja, A. S., Chapadiya, F. and Rajani, A. V., Surendranath, R., Jawaharlal, M. and Anitha, K., (2017). Evaluation of Physical Properties of Irrigated Soils of Morbi District of Saurashtra Region of Gujarat Int. J. Pure App. Biosci. 5 (4):DOI:10.18782/2320-7051.5189.

[29] Chuan M. C, Shu G. Y, Liu J. C (1996). Solubility of heavy metals in a contaminated soil: Effects of redox potential and pH. Water, Air and Soil Pollution. Vol. 90 No. 3-4, 543-556.

[30] United States Environmental Protection Agency (US EPA) (2012). Reviewed Supplemental guidance for developing soil screening levels for superfund sites. Office of Solid Waste and Emergency Response, Washington D. C http://www.epa.gov/perfund/health/commedia/soil/index.htm.

[31] Environmental Protection Authority of Australia (EPAA) (2012). Classification and management of contaminated soil for disposal. Information Bulletin 105. Hobart, TAS 7001 Australia.

[32] World Health Organization (WHO) (2015). Global Health Observatory data repository. Burden of disease. Lead attributable daily. http://apps.who.int/gho/data/node.home.

[33] European Environmental Agency (EEA) (2007). Progress in management of contaminated sites (CSI 015/LSI 003, http://www.eea.europa.eu/data-and-maps/indicators.

[34] Canadian Ministry of the Environment (CME). (2009). Soil, Ground Water, Sediment Standards for use under Part XV. 1 of the Environmental Protection Act.

[35] World Health Organization (WHO) (2015). Global Health Observatory data repository. Burden of disease. Lead attributable daily. http://apps.who.int/gho/data/node.home.

[36] United States Environmental Protection Agency (US EPA) (2012). Reviewed Supplemental guidance for developing soil screening levels for superfund sites. Office of Solid Waste and Emergency Response, Washington D. C http://www.epa.gov/perfund/health/commedia/soil/index.htm.

[37] Environmental Protection Authority of Australia (EPAA) (2012). Classification and management of contaminated soil for disposal. Information Bulletin 105. Hobart, TAS 7001 Australia.

[38] World Health Organization (WHO) (2015). Global Health Observatory data repository. Burden of disease. Lead attributable daily. http://apps.who.int/gho/data/node.home.

[39] European Environmental Agency (EEA) (2007). Progress in management of contaminated sites (CSI 015/LSI 003, http://www.eea.europa.eu/data-and-maps/indicators.

[40] Canadian Ministry of the Environment (CME). (2009). Soil, Ground Water, Sediment Standards for use under Part XV. 1 of the Environmental Protection Act.
[47] Mico´ C, Recatala´ L, Peris M, Sa´nchez J (2006). Assessing heavy metal sources in Agricultural soils of an European Mediterranean area by multivariate analysis. Chemosphere 65:863– 872.

[48] Tamene Fite Duressa, Seyoum Leta (2015). Determination of Levels of As, Cd, Cr, Hg and Pb in Soils and Some Vegetables Taken from River Mojo Water Irrigated Farmland at Koka Village, Oromia State, East Ethiopia. International Journal of Sciences: Basic and Applied Research Volume 21, No 2, 352-372.

[49] Food and Agricultural Organisation FAO (2006): Guidelines for soil description, Food and Agriculture Organization of the United Nations, Rome.

[50] Rasheed M. A, Lakshmi M, Rao P. L. S, Kalpana M. S, Dayal A. M and Patil D. J (2013). Geochemical evidences of trace metal anomalies for finding hydrocarbon micro seepage in the petroliferous regions of the Tatipaka and Pasarlapudi areas of Krishna Godavari Basin, India. Pet. Sci. (2013) 10:19-29. DOI 10.1007/s12182-013-0245-x.

[51] Jayawardana D. T, Pitawala H. M. T. G. A, Ishiga H (2014). Geochemical evidence for the accumulation of vanadium in soils of chronic kidney disease areas in Sri Lanka. Environ Earth Sci. Vol. 73. Issue 9, 5415-5424. DOI 10.1007/s12665-014-3796-2.

[52] Hernandez, H. & Rodriguez, R. (2012). Geochemical evidence for the origin of vanadium in an urban environment. Environ Monit. Assess. Volume 184, Issue 9, 5327–5342 DOI/10.1007/s10661-011-2343-9.

[53] Veronica E. Manga, Christopher M. Agyingi, Cheo E. Suh (2014). Trace Elements Soil Quality Status of Mt. Cameroon Soils. Advances in Geology, volume 2014, 1-8 DOI/10.1155/2014/894103.

[54] Aliana Pendias-Kabata, Henryk Pendias (2001). Trace Elements in Soils and Plants. Third Edition CRC Press Boca Raton, London 239-249.

[55] Alloway B. J (2012). Heavy metals in soils: Trace Metals and metalloids in soils and their bio-availability. Third Edition. Springer Science.

[56] Merian, E., Anke, M., Ihnat, M and Stoeppler, M (2004). Elements and their Compounds in the Methods. Melbourne, Australia: Inkata Press. 145-148.