Species richness and distribution patterns of the snake fauna of Rio Grande do Norte state, northeastern Brazil

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Abstract: The Neotropics are one of the richest regions in biodiversity globally. Still, much remains unknown about the mechanisms and processes responsible for the accumulation of species in this region. Among the many limitations on our current knowledge on the region’s biodiversity, understanding of community composition and species distributions is limited and greatly biased in many Brazilian regions. We present information on species composition, habitat use, geographic distribution, taxonomic accounts, and conservation of snakes from Rio Grande do Norte state, Northeastern Brazil. We compiled, from primary and literature data, 851 snake records from seven families and 47 species, with five new records for the state. Species are mainly terrestrial and semi-arboreal and associated with at least six vegetation types from Caatinga and Atlantic Forest domains. None of the species is listed in threatened categories of IUCN, while two species are listed in the Brazilian Threatened Fauna list. Our data covers 32.34% of the state’s area, a consequence of locally limited inventories and lack of long-term studies on snakes’ fauna. The richest areas within the state are near large cities, which lack protected areas for the species they harbor, and highlights the necessity of protective policies and conservation actions.

Key words: Atlantic Forest, Caatinga, conservation, Serpentes, Squamata.

INTRODUCTION

Biodiversity is not equally distributed across Earth, varying significantly among and within geographic regions and taxa. The Neotropical region is one of the most biodiverse areas in the world (Olson et al. 2001, Antonelli & Sanmartin 2011). Understanding the mechanisms and processes that generate and maintain the Neotropical hyper-diversity has been an important target in research and also a challenge since 19th century naturalists began to study the region. Nowadays, this effort is still a central goal for researchers in systematic, ecology, biogeography, and evolutionary biology (Rull 2011, Hughes et al. 2013). One fundamental and practical limit of biodiversity knowledge is that scientists often work with incomplete and/or unrepresentative datasets. For instance, knowledge about the identity and distribution of species is biased (the Wallacean Shortfall), which compromises our capacity to synthesize knowledge of existing biodiversity, leading to misidentification of ecological and evolutionary processes and inefficient conservation strategies (Lomolino 2004, Hortal et al. 2015).

Squamate reptiles comprise the world’s most diverse group of terrestrial vertebrates (Tonini et al. 2016, Uetz et al. 2020), and are arguably among the most neglected in conservation prioritization exercises (Schipper...
Within squamates, snakes comprise nearly 3,800 known species inhabiting temperate to tropical environments in varied habitats including terrestrial, marine, and freshwater areas (Wallach et al. 2014, Uetz et al. 2020). Snakes have a wide variety of ecological and morphological adaptations, and intriguing reproductive and dietary aspects and thus constitute an excellent system to investigate evolutionary processes (Cadle & Greene 1993). However, the lack of knowledge about taxonomy and geographic distribution, for example, has hindered the inclusion of snakes in most large-scale biodiversity and evolutionary studies (e.g. Pereira et al. 2010, Colston et al. 2013, Jenkins et al. 2015, Alencar et al. 2016, Card et al. 2016, Moura et al. 2016). Reliable estimates of the diversity and distribution of snakes would contribute to both global and regional understanding of evolutionary history of biotas and more accurate strategies for their conservation (Böhm et al. 2013, Meiri & Chapple 2016).

In a recent study, Guedes et al. (2018) provided an overview of the distribution of Neotropical snakes based on a large database of georeferenced records representing 886 species and 12 families spanning 27 countries. To our knowledge, it is the most extensive and complete database of snake distribution for the Neotropical region. That study identified Amazonia, the Andes, and some portions of Northeastern Brazil as poorly sampled (less than 100 records per one-degree grid cell). As a consequence, these areas also present low richness (Guedes et al. 2018). The state of Rio Grande do Norte (RN hereafter) is located in northeastern Brazil and is also assumed to have low sampling and low species richness of snakes. Thus, a large knowledge gap identified in the literature for this state represents a serious shortcoming of present literature.

Many herpetofaunal studies in RN have focused on amphibians and lizards, while snake studies are por based on punctual observations and do not address local or state diversity. Indeed, some studies focused on snake association with bromeliads (Jorge et al. 2020), and chromatic anomaly (Paredero & Passos 2020), while others reported endoparasites for two species (Almeida et al. 2008). In addition, accidents with venomous species and the regions where they occur have also been reported (Tavares et al. 2017, Costa et al. 2019). Otherwise, publications on the snakes of RN report single geographic distribution records (Brito P.S., unpublished data, Jorge & Freire 2011, Brito & Freire 2012, Santos et al. 2020, Aquino et al. 2020) or natural history notes (Souza & Freire 2008, Sales et al. 2013, Ribeiro et al. 2014).

Snakes have also been part of local herpetofaunal inventories (Lima-Verde 1971, 1976, Sales et al. 2009, Caldas et al. 2016, Calixto & Morato 2017, Coelho-Lima et al. 2020), and conservation inquiries such as how forest fragment characteristics (e.g., area, shape, isolation and matrix quality) affected reptiles in RN’s Atlantic Forest (Lion et al. 2016). Finally, records of snakes for RN are mostly contained in broad natural history compilations (Schmidt & Inger 1951, Guedes et al. 2014a, 2018) and taxonomic reviews (Thomas unpublished data, Myers & Cadle 1994, Arredondo unpublished data, Fernandes & Hamdan 2014, Pires et al. 2014, Montingelli et al. 2019). Costa & Bérnils (2018) compiled the list of snake species for all Brazilian states based on presence and absence and listed for the first time a total of 43 species for RN without any further information.

Herein, we provide an updated list of snake species recorded inside the political limits of RN state through primary data, complemented by scientific literature. Additionally, for each species we present (1) the geographic distribution within...
the state, (2) information on habitat use, and (3) we discuss previous identification mistakes of species occurring in the state reported in the literature.

MATERIALS AND METHODS

Study area

The Northeast region of Brazil comprises a massive portion of the country with an area of 1,558,000 km² (IBGE 2017). Rio Grande do Norte state is the northeasternmost state in Brazil, between latitudes 4.82 and 6.98 South, and longitudes 38.58 and 34.96 West (Figure 1), and borders the states of Ceará and Paraíba. The state covers an area of 52,811 km² corresponding to 0.62% of the Brazilian territory (Figure 1; IBGE 2017) and encompasses 167 municipalities arranged into four geographic mesoregions: Oeste Potiguar (62 municipalities), Central Potiguar (37 municipalities), Agreste Potiguar (43 municipalities), and Leste Potiguar (25 municipalities) (Figure 1; IDEMA 2008).

The natural landscapes in RN include a variety of geomorphological formations and phytosociomogies. The relief includes plains and Tabuleiros along the coastal plain, lowland areas such as the sublitorânea and sertaneja.
depressions, and mountain chains (e.g., Chapada da Serra Verde, Chapada do Apodi and Planalto da Borborema) (Diniz et al. 2015). The state is irrigated by sixteen river basins composed of seasonal rivers, lakes, and ponds. Two major river basins, Apodi-Mossorô and Piranhas-Assu, are responsible for the majority of the state’s water reserves (Nascimento et al. 2014). The climate type according to Köppen-Geiger ranges from Tropical savanna (Aw) to Arid steppe (BSh) and desert hot (BWh) (Peel et al. 2007), with an average temperature of 28°C. Precipitation is higher in the eastern coastal region (1,100 mm per year) and decreases below 500 mm per year towards the west (Diniz & Pereira 2015). Most of the state is within the Caatinga biome (95% of the state area), while a narrow portion on the east coast is composed by Atlantic Forest (5% of the state area) (Figure 1). Hence, the result is a vast vegetation mosaic composed of restinga sand dunes and mangroves (both with marine influence), seasonally dry formations such as caatinga bush, forest, and parkland vegetation, savanna-like fragments, and deciduous and semideciduous forests (IBGE 2012, SFB 2018).

Data source
We gathered snake distribution records by examining voucher specimens collected by us, housed in the collections of the Laboratório de Anfíbios e Répteis at the Universidade Federal do Rio Grande do Norte (LAR-UFRN), and the Coleção Herpetológica do Semiárido at the Universidade Federal Rural do Semi-Árido (CHSA-UFERSA). Additionally, we compiled snake records from the scientific literature such as species inventories, ecological studies, taxonomic reviews, natural history and geographic distribution papers, dissertations, thesis, and books. We considered all records available from each paper and compared records with other studies to make sure we did not use the same record twice. We did not consider Lima-Verde (1971, 1976) whose records were not detailed for municipalities. We also included in our list photographic records and confirmation of records based on direct observation by the authors. We followed Burbrink et al. (2020) for family taxonomy.

We defined snake habitats based on records and the phytoecological units in RN state established by the national forest inventory (SFB 2018). When possible, we also provided information about habitat use from the location where the snake was collected or observed, complemented with literature information. We plotted snake records on a shapefile with federal and state protected areas to evaluate the degree of protection of each species in the state. We generated distribution maps for each species with habitat information using Mapbiomas 3.0 shapefiles. Finally, to evaluate distribution patterns of species richness, we compiled snake richness and geographic coordinates from all recorded municipalities and produced a Kernel density map restricted to RN with a search radius of 0.2 in Qgis 3.8.2.

RESULTS
We obtained 851 records (450 primary and 401 from literature) of 47 species from seven families (Dipsadidae, Colubridae, Elapidae, Boidae, Viperidae, Typhlopidae, and Leptotyphlopidae) and 33 genera. Dipsadidae and Colubridae were the most represented in species richness with 28 species (58.3%) and 7 species (14.6%), respectively. We recorded five species for the first time in RN: Apostolepis longicaudata, Bothrops leucurus, Dipsas mikanii, Imantodes cenchoa, and Thamnodynastes sertanejo. Most snakes in RN are terrestrial (66.7%), followed by semiarboreal (25%), fossorial (18.8%), aquatic (16.7%) and arboreal (10.4%) species. Most
species were found active during only one period of the day (40.4% nocturnal, and 36.2% diurnal), while a small portion was found active on both periods (23.4%) (Table I and Figures 2–5).

Our database contains at least one snake record in 54 (32.34%) of the 167 municipalities in RN territory. When we consider the four mesoregions of the state, there were more records for municipalities in the Leste Potiguar (19 municipalities) with 277 snake occurrences, followed by Central Potiguar (14; 185), Oeste Potiguar (8; 67). The municipalities of Mossoró and Macaíba were the most representative in species richness (22 and 18 species, respectively), followed by Natal and Nísia Floresta (17 species each) (Figure 6A). The Kernel density map showed major richness on the Leste Potiguar and Mossoró regions (Oeste Potiguar), and additional scattered areas in João Câmara (Agreste Potiguar), Serra Negra do Norte (Central Potiguar), and Assú (Oeste Potiguar) (Figure 6B). Philodryas nattereri, Micrurus aff. ibiboboca, and Oxyrhopus trigeminus are widespread in RN and were recorded in 22, 16, and 15 municipalities, respectively (Figure 7). Detailed information of distribution maps for each species, sources, and snake species occurrences for all municipalities in RN are listed in the supporting information (Table SI and Figures S1–4).

Regarding the biomes, 660 records were from the Caatinga and 191 were from the Atlantic Forest. Twenty-two species were recorded in both biomes, while 13 species occurred only in Caatinga (Apostolepis longicaudata, Boiruna sertaneja, Crotalus durissus, Erythrolamprus miliaris, E. mossoroensis, Helicops angulatus, H. leoparдинus, Leptodeira annulata, Lygophis dilepis, Phimiphis guerini, Sibon nebulatus, Thamnodynastes almae, and T. sertanejo) and 12 others only in Atlantic Forest (Amerotyphlops amoipira, A. paucisquamus, Bothrops leucurus, Drymarchon corais, Erythrolamprus almadensis, Hydrodynastes gigas, Imantodes cenchoa, Palusophis bifossatus, Micrurus cf. corallinus, M. potyguara, Psomophis joberti, and Spilotes pullatus). We detected snakes in six phytosociognomies within RN. Snakes are predominantly reported for caatinga bush vegetation (37 species) and restinga sand dunes (29), followed by semideciduous forest (18), cerrado vegetation and caatinga forest vegetation (7 each), and caatinga parkland vegetation (5). Oxyrhopus trigeminus and Philodryas nattereri were detected in five vegetation types while other species were exclusive from restinga sand dunes (Amerotyphlops amoipira, Erythrolamprus almadensis, Bothrops leucurus, Micrurus potyguara, and Spilotes pullatus), caatinga bush vegetation (Micrurus aff. ibiboboca, Apostolepis longicaudata, Boiruna sertaneja, Erythrolamprus miliaris, E. mossoroensis, Helicops angulatus, H. leopar dinus, Leptodeira annulata, Phimiphis guerini, Thamnodynastes almae, and T. sertanejo), and semideciduous forest (Imantodes cenchoa and Psomophis joberti).

The IUCN lists Corallus hortulana, Bothrops erythromelas, Crotalus durissus, Erythrolamprus viridis, and Lygophis dilepis as Least Concern and Amerotyphlops amoipira as Data Deficient. The Red Book of Brazilian Threatened Fauna lists A. amoipira as endangered and A. paucisquamus as vulnerable (ICMBIO 2018). We recorded snake species inside the limits of three protected areas in the Leste Potiguar mesoregion (Environmental Protection Area of Apariqui-Una, EPA Bonfim/ Guanaíra, and Mata Estrela private reserve), one in the Central Potiguar mesoregion (Seridó Ecological Station), and one in the Oeste Potiguar mesoregion (Açu National Forest). Protected areas harbor 36 species, including three new records for RN, while Apostolepis longicaudata and Thamnodynastes sertanejo, both poorly known species (Curcio et al. 2011, Guedes et al. 2014a), were recorded only in unprotected areas.
Table I. Snake species recorded in Rio Grande do Norte state, Brazil. The table outlines total records for each species (TR), total municipalities of occurrence (MO), biomes (Caatinga – CA and/or Atlantic Forest – AF), habitats (restinga sand dunes – RSD, caatinga bush vegetation – CBV, caatinga forest vegetation – CFV, caatinga parkland vegetation – CPV, cerrado – CE, and semideciduous forest – SDF. Snakes habitats are as follows: aquatic (Aq), arboreal (Ar), fossorial (Fo), terrestrial (Te) and semiarboreal (SA). Activity period: diurnal (D) and nocturnal (N). (*) New state records.

| Species                      | TR  | MO | Biome      | Habitat | Habit | Activity |
|------------------------------|-----|----|------------|---------|-------|----------|
|                              |     |    |            | RSD     | CBV   | CFV      | CPV     | CE     | SDF |       |
| **Leptotyphlopidae**         |     |    |            |         |       |          |         |        |     |       |
| *Epictia borapeliotes*       | 32  | 7  | CA, AF     | X       | X     | Fo       | N       |        |     |       |
| (Vanzolini, 1996)            |     |    |            |         |       |          |         |        |     |       |
| ** Typhlopidae **            |     |    |            |         |       |          |         |        |     |       |
| *Amerotyphlops amoipira*     | 2   | 1  | AF         | X       |       | Fo       | N       |        |     |       |
| (Rodrigues & Juncá, 2002)    |     |    |            |         |       |          |         |        |     |       |
| *Amerotyphlops paucisquamus* | 6   | 4  | AF         | X       |       | X        | Fo      | N       |     |       |
| (Dixon & Hendricks, 1979)    |     |    |            |         |       |          |         |        |     |       |
| **Boidae**                   |     |    |            |         |       |          |         |        |     |       |
| *Boa constrictor* Linnaeus, 1758 | 22 | 7  | CA, AF     | X, X    |       | X        | Te, SA  | D, N    |     |       |
| *Corallus hortulana* Linnaeus, 1758 | 6 | 3  | CA, AF     | X       |       | X        | Ar      | N       |     |       |
| *Epicrates assisi* Machado, 1945 | 14 | 9  | CA, AF     | X, X    |       | X        | Te, SA  | N       |     |       |
| **Viperidae**                |     |    |            |         |       |          |         |        |     |       |
| *Bothrops erythromelas*      | 58  | 13 | CA, AF     | X, X    |       | X        | Te      | N       |     |       |
| Amaral, 1923                 |     |    |            |         |       |          |         |        |     |       |
| *Bothrops leucurus* Wagler in Spix 1824* | 1 | 1  | AF         | X       |       |          | Te      | N       |     |       |
| *Crotalus durissus* Wagler in Spix, 1824 | 14 | 8  | CA         | X, X    |       |          | Te      | N       |     |       |
| **Elapidae**                 |     |    |            |         |       |          |         |        |     |       |
| *Micrurus aff. ibiboboca*    | 38  | 16 | AF, CA     | X, X    |       | X        | Fo      | D, N    |     |       |
| (Jan, 1863)                  |     |    |            |         |       |          |         |        |     |       |
| *Micrurus cf. carallinus*    | 2   | 2  | AF         | X       |       | X        | Fo      | D       |     |       |
| (Merrem, 1820)               |     |    |            |         |       |          |         |        |     |       |
| *Micrurus potyguara* Pires, Da Silva Jr, Feitosa, Prudente, Pereira-Filho & Zaher, 2014 | 2 | 1  | AF         | X       |       |          | Fo      | D, N    |     |       |
| **Colubridae**               |     |    |            |         |       |          |         |        |     |       |
| *Chironius flavolineatus*    | 28  | 6  | CA, AF     | X, X    |       | X        | X       | SA      | D     |       |
| (Jan, 1863)                  |     |    |            |         |       |          |         |        |     |       |
| *Drymarchon corais* Boie, 1827 | 4 | 3  | AF         | X, X    |       | X        | Ar      | D       |     |       |
### Table I. Continuation

| Species                                      | Count | Number of records | Distribution | Notes |
|----------------------------------------------|-------|-------------------|--------------|-------|
| *Leptophis ahaetulla* (Linnaeus, 1758)       | 15    | 8                 | CA, AF       |       |
| *Oxybelis aeneus* (Wagler in Spix, 1824)    | 52    | 11                | CA, AF       |       |
| *Palosulis bifossatus* (Raddi, 1820)        | 5     | 2                 | AF           |       |
| *Spilotes pullatus* (Linnaeus, 1758)        | 3     | 2                 | AF           |       |
| *Tantilla melanocephala* (Linnaeus, 1758)   | 16    | 8                 | CA, AF       |       |
| **Dipsadidae**                              |       |                   |              |       |
| *Apostolepis cearensis* Gomes, 1915          | 23    | 8                 | CA, AF       |       |
| *Apostolepis longicaudata* Gomes in Amaral, 1921* | 1   | 1                 | CA           |       |
| *Boiruna sertaneja* Zaher, 1996             | 17    | 5                 | CA           |       |
| *Dipsas mikanii* (Schlegel, 1837)*          | 4     | 3                 | CA, AF       |       |
| *Erythrolamprus almadensis* (Wagler in Spix, 1824) | 1   | 1                 | AF           |       |
| *Erythrolamprus miliaris* (Linnaeus, 1758)  | 2     | 2                 | CA           |       |
| *Erythrolamprus mossoroensis* (Hoge & Lima-Verde, 1973) | 7   | 4                 | CA           |       |
| *Erythrolamprus poecilogyrus* (Wied, 1825)  | 28    | 9                 | CA, AF       |       |
| *Erythrolamprus viridis* (Gunter, 1862)     | 47    | 6                 | CA, AF       |       |
| *Helicops angulatus* (Linnaeus, 1758)       | 6     | 1                 | CA           |       |
| *Helicops leopardinus* (Schlegel, 1837)     | 1     | 1                 | CA           |       |
| *Hydrodynastes gigas* (Duméril, Bibron & Duméril, 1854) | 3   | 3                 | AF           |       |
| *Imantodes cenchoa* (Linnaeus, 1758)*       | 1     | 1                 | AF           |       |
| *Leptodeira annulata* (Linnaeus, 1758)      | 13    | 3                 | CA           |       |
| *Lygophis dilepis* (Cope, 1862)             | 38    | 10                | CA           |       |
### Table I. Continuation

| Species                                      | Count | Size | CA | AF | CA | X | X | X | Te | SA | Location |
|----------------------------------------------|-------|------|----|----|----|---|---|---|----|----|----------|
| *Oxyrhopus trigeminus* Duméril, Bibron & Duméril, 1854 | 73    | 15   | X  | X  | X  | X | X | X | Te | SA | D, N     |
| *Philodryas nattereri* Steindachner, 1870      | 130   | 21   | CA | AF | X  | X | X | X | X  | Te | SA      | D        |
| *Philodryas olfersii* (Liechtenstein, 1823)    | 43    | 12   | CA | AF | X  | X | X | X | X  | Te | SA      | D        |
| *Philodryas patagoniensis* (Girard, 1858)      | 8     | 4    | CA | AF | X  | X |   |   | X  |    | Te, D    |          |
| *Phimophis guerini* (Duméril, Bibron & Duméril, 1854) | 1     | 1    | CA |    | X  |   |   |   |    |    | Te, Fo   | N        |
| *Pseudoboa nigra* (Duméril, Bibron & Duméril, 1854) | 26    | 9    | CA | AF | X  | X |   |   |    |    | Te, D    | N        |
| *Psomophis joberti* (Sauvage, 1884)            | 1     | 1    | AF |    |    |   |   |   |    | Te, Fo | D        |
| *Sibon nebulatus* (Linnaeus, 1758)             | 2     | 2    | CA |    | X  |   |   |   |    | Ar | N        |
| *Taeniophallus occipitalis* (Jan, 1863)         | 8     | 6    | CA | AF | X  | X |   |   |    |    | Te, Fo   | D        |
| *Thamnodynastes almae* Franco & Ferreira, 2002 | 2     | 1    | CA |    | X  |   |   |   |    |    | Te, SA   | N        |
| *Thamnodynastes phoenix* Franco, Trevine, Montingelli & Zaher, 2017 | 28    | 9    | CA | AF | X  | X |   |   |    |    | Te, SA   | N        |
| *Thamnodynastes sertanejo* Bailey, Thomas & Da Silva, 2005* | 1     | 1    | CA |    | X  |   |   |   |    |    | Te, SA   | N        |
| *Xenodon merremii* (Wagler in Spix, 1824)      | 16    | 5    | CA | AF | X  | X |   |   |    |    | Te       | D        |
**Figure 2.** Snakes from Rio Grande do Norte state, Brazil. Leptotyphlopidae: (a) *Epictia borapeliotes*. Typhlopidae: (b) *Amerotyphlops amoipira*, (c) *A. paucisquamus*. Boidae: (d) *Boa constrictor*, (e) *Corallus hortulana*, (f) *Epicrates assisi*. Viperidae: (g) *Bothrops erythromelas*, (h) *Bothrops leucurus*, (i) *Crotalus durissus*. Elapidae: (j) *Micrurus cf. corallinus*, (k) *M. aff. ibiboboca*, and (l) *M. potyguara*. Photo b by D.J. Santana and photo h by J.L.G.S. Silveira.

**Figure 3.** Snakes from Rio Grande do Norte state, Brazil. Colubridae: (a) *Chironius flavolineatus*, (b) *Drymarchon corais*, (c) *Leptophis ahaetulla*, (d) *Oxybelis aeneus*, (e) *Palusophis bifossatus*, (f) *Spilotes pullatus*, (g) *Tantilla melanolephala*. Dipsadidae: (h) *Apostolepis cearensis*, (i) *A. longicaudata*, (j) *Boiruna sertaneja*, (k) *Dipsas mikanii*, and (l) *Erythrolamprus almadensis*. Photo e by D.J. Santana.
Figure 4. Snakes from Rio Grande do Norte state, Brazil. Dipsadidae: (a) Erythrolamprus miliaris, (b) E. mossoroensis, (c) E. poecilogyrus, (d) E. viridis, (e) Helicops angulatus, (f) H. leopardinus, (g) Hydrodynastes gigas, (h) Imantodes cenchoa, (i) Leptodeira annulata, (j) Lygophis dilepis, (k) Oxyrhopus trigeminus, (l) Philodryas nattereri. Photo d by B. Pontes.

Figure 5. Snakes from Rio Grande do Norte state, Brazil. Dipsadidae: (a) Philodryas olfersii, (b) P. patagoniensis, (c) Phimophis guerini, (d) Pseudoboa nigra, (e) Psomophis joberti, (f) Sibon nebulatus, (g) Taeniophallus occipitalis, (h) Thamnodynastes almae, (i) T. phoenix, (j) T. sertanejo, and (k) Xenodon merremii. Photos c and e by G.A. Pereira-Filho.
DISCUSSION

Snake richness and habitat use overview in Rio Grande do Norte

This is the largest compilation of snake records for Rio Grande do Norte to date, and we generate detailed maps of snake occurrences and patterns of species richness, along with quantifying sampling gaps. We also report new occurrences, geographic distribution extensions, and habitat information for all species. The species richness of RN is somewhat low when compared to other state compilations for Brazil, such as Bahia (131 spp.; Curcio et al. 2012, Hamdan & Lira-da-Silva 2012, Fernandes & Hamdan 2014), Rondônia (119 spp.; Bernarde et al. 2012, Passos et al. 2016), Mato Grosso do Sul (113 spp.; Ferreira et al. 2017), Ceará (67 spp.; Roberto & Loebmann 2016, Borjes-Nojosa et al. 2017), and Paraíba (63 spp.; Pereira-Filho et al. 2017).

Many factors can affect species richness, including diversity of habitats, area, and sampling effort (Bracewell et al. 2018). For example, Bahia and Mato Grosso do Sul states, while also being much larger than RN, harbor three distinct biomes of forested and open formations. Ceará state is also larger than RN, and although the Atlantic Forest does not reach the state, it harbors many relictual enclaves of humid forests on top of high-altitude areas maintained by orographic rainfall. Larger area and environmental complexity allow higher richness and support for endemic species to persist over time (Roberto & Loebmann 2016). Aside from environmental features, areas with high species richness are usually better sampled at local level (Guedes et al. 2018). Indeed, the low snake species richness in RN is very likely a sampling artifact resulting from the reduced number of studies for the group in RN compared to the neighboring states of Ceará and Paraíba (Santana et al. 2008, Loebmann & Haddad 2010, Mesquita et al. 2011, 2012, 2013, Pereira-Filho & Montingelli, 2011, França et al. 2012, Roberto & Loebmann, 2016, Costa et al. 2018, Sampaio et al. 2018).

Some species reported herein are known from single or old records for RN (e.g. Psomophis joberti, Erythrolamprus almadensis, Helicops leopardinus, H. angulatus). Most of these species are frequent in other regions (Guedes et al. 2014a, Marques et al. 2016, Mesquita et al. 2018, França et al. 2020) and the lack of recent records in RN reinforces the scarcity of snake studies in the state. Likewise, the five new species recorded for the state further suggests that the
state is still heavily understudied. Knowledge of snake species richness is concentrated in areas easily accessed by researchers, leading to a lack of snake records on most of the state’s territory, hindering our ability to correctly assess the overall diversity and conservation status of these animals in the state (Meyer et al. 2015, Oliveira et al. 2016).

Over 65% of the localities we list and 29% of the state’s area have three or fewer species registered. Furthermore, areas with the highest species richness are within/near the municipalities of two of the largest cities (Natal and Mossoró) or inside protected areas with research infrastructure (Açu National Forest and Seridó Ecological Station). Such correlation of sampling intensity near larger cities and research facilities is expected (Meyer et al. 2015, Oliveira et al. 2016).

Most species were recorded in Caatinga bush vegetation, which covers 84% of RN state (SFB 2018). Although restinga sand dune habitats cover around 4% of RN vegetation, this habitat is distributed along the seashore of RN where densely populated cities are found. Despite the sampling artifact, snake richness is high throughout Atlantic Forest habitats (Argôlo 2004, Santana et al. 2008, Pereira-Filho & Montingelli 2011, Marques et al. 2016, Guedes et al. 2018, Sampaio et al. 2018). This raises concerns, as the biome is threatened by ongoing anthropogenic actions and habitat loss (Joly et al. 2014) and is mostly restricted to small fragments embedded in sugar cane plantations in RN (Lion et al. 2016). Even though the Atlantic Forest covers a small portion of RN state, it showed similar richness to the Caatinga biome, reinforcing the conservation importance of this biome and its related ecosystems for biodiversity. Some species recorded in restinga sand dunes (e.g. *Erythrolamprus almadensis*, *Bothrops leucurus*, and *Spilotes pullatus*) are also common in this habitat in other regions (Miranda et al. 2012, Marques et al. 2016). *Oxyrhopus trigeminus* and *Philodryas nattereri* are generalist species and inhabit many vegetation types along their distributions (Pereira-Filho & Montingelli 2011, Guedes et al. 2014a, Marques et al. 2016), and were also recorded in almost all habitats in RN. All species exclusive to caatinga bush vegetation
are typical from open habitat formations and likely occur in this habitat (Guedes et al. 2014a). Likewise, *Imantodes cenchria* and *Psomophis joberti* were only found in semideciduous forest, as the former is an arboreal species typical from forests and the later occurs in many habitats from Caatinga, Cerrado, and Atlantic Forest (Moura et al. 2013, Marques et al. 2016, Mesquita et al. 2018).

**Geographic distribution records and taxonomic accounts**

Our data added five new species records for RN state. *Apostolepis longicaudata* is mostly reported for areas of cerrado vegetation (Curcio et al. 2011) and here it was recorded in João Câmara municipality (5.37346 S, 35.879398 W, WGS84, 188 m a.s.l.) where xeric vegetation prevails. This record extends its distribution to 237 km North of Cabaceiras, Paraíba state (Curcio et al. 2011, Pereira-Filho et al. 2017) and 1281 km East from Estreito, Maranhão state (França et al. 2018). *Bothrops leucurus* inhabits mainly forests (Argôlo 2004) and the nearest record is from REBIO Guaribas (Reserva Biológica, or Biological Reserve, a strict protection area in Brazil), between the municipalities of Rio Tinto and Mamanguape, Paraíba state (Mesquita et al. 2018), 38 km south from our record in Baía Formosa (6.381705 S, 35.016233 W, 63 m asl). We recorded *Dipsas mikanii* in Macaíba (5.866257 S, 35.32931 W, 42 m a.s.l.), Nísia Floresta (6.083561 S, 35.18251 W, 59 m a.s.l.), and Parnamirim (5.915981 S, 35.18419 W, 34 m a.s.l.). These records are 466 km Northeast of Barbalha, Ceará state (Roberto and Loebmann 2016) and 72 km North of Mamanguape, in Paraíba state (Mesquita et al. 2018). We also recorded *Imantodes cenchria* in Baía Formosa, 38 km northward from REBIO Guaribas in Paraíba state (Mesquita et al. 2018) and 495 km eastward from the municipality of Crato, Ceará state (Roberto and Loebmann, 2016). Finally, *Thamnodynastes sertanejo* is endemic from Caatinga, and our record in João Câmara (5.526461 S, 35.841056 W, 160 m a.s.l.) extends its distribution 423 km Northeast of Jati, Ceará state, and 320 km Northeast of Sertânia, Pernambuco state (Coelho et al. 2013).

Previous records within RN require clarification. Sales et al. (2009) reported *Epicrates cenchria* for sand dunes in the municipality of Natal without specifying its subspecies. Passos and Fernandes (2008) reviewed and elevated the subspecies of *Epicrates* to species. *Epicrates cenchria* occurs in Amazonia and Atlantic Forest in primary, ombrophilous, and disturbed forests (Martins & Oliveira 1998, Argólo 2004). Though the municipality of Natal is within the Atlantic Forest biome, its vegetation is strongly influenced by Caatinga and this record actually corresponds to *E. assisi* (R. Sales pers. comm.). Large patches of semideciduous forests are abundant on the southeast portion of the state, near the Paraíba state boundary, where we recorded typical Atlantic Forest species as *Corallus hortulana* and *Bothrops leucurus*. Schmidt & Inger (1951) and Freire et al. (2009) reported the occurrence of *Lygophis lineatus* but this species was restricted to Amazonia after review by Hoge (1952), while *L. dilepis* has a broad distribution in RN state, as already pointed by Guedes et al. (2014a).

The compilation of Costa & Bérnils (2018) reported *Micrurus lemniscatus carvalhui*, *Amerotyphlops brongersmianus* and *Erythrolamprus viridis viridis* for RN. *Micrurus lemniscatus carvalhui* was recorded by Schmidt & Inger (1951) in Nísia Floresta, but later review by Pires et al. (2014), who showed this to be a novel species, *M. potyguara*. As for *A. brongersmianus*, Lion et al. (2016) recorded it in Goianinha, but we examined the voucher specimen and scale counts (18 rows around the body and 214 middorsal scales) are typical of *A. paucisquamus* (Hedges et al. 2014). Finally,
Sales et al. (2009) reported *E. viridis* for the sand dunes in Natal and Costa & Bérnils (2018) considered the subspecies *E. v. viridis* because the record was from the Atlantic Forest biome (H. Costa pers. comm.). We believe this situation is similar to that of *Epicrates cenchria* / *E. assisi*, and the subspecies that must be considered for the state is *Erythrolamprus viridis prasinus*. As we did not examine its voucher, however, further investigation is needed to clarify this issue. Lastly, species like *E. almadensis* (Guedes et al. 2014a) and *Spilotes pullatus* (Sales et al. 2009) are reported here but are absent in the compilation of Costa & Bérnils (2018).

The holotype of *Micrurus ibiboboca* was described from the Atlantic Forest (Belmonte, Bahia state) with a diagnosis of 210 ventral scales and 23 subcaudal scales (Merrem 1820). Vanzolini et al. (1980) reported that specimens from the Caatinga were not properly described in the literature, since they obtained snakes with 219-243 ventral scales. Then, Argôlo (2004) examined specimens of *M. ibiboboca* from Southeast Bahia with 192-219 ventral scales and 16-26 subcaudal scales, which matched the holotype of *M. ibiboboca*. The segregation between specimens from Caatinga and Atlantic Forest was also highlighted in Guedes et al. (2014a), but Pires et al. (2014) treated them as a single entity. Ventral scales from specimens of RN ranged from 226 to 250 and subcaudal scales from 24 to 30 without proper segregation between both biomes. Until a more detailed taxonomic review of these species is available, we propose specimens from RN should be treated as *Micrurus aff. ibiboboca*.

**Conservation challenges for Rio Grande do Norte snakes**

The biggest conservation challenge for snake species in RN in face of rising habitat loss is to identify priority areas for conservation considering current sampling gaps. RN still holds 42% of its original vegetation coverage (SFB 2018). Though 6.2% of RN’s area corresponds to protected areas (SFB 2018, IDEMA 2019), most of these confer low protection (Environmental Protection Areas, or APAs) and just a small part of the Caatinga (55.37 km²) that occurs in the state is protected by two Strict Protection areas (SPA), Seridó Ecological Station and Furna Feia National Park.

Most snake species recorded for RN show broad distributions, occurring in other biomes and neighboring states (Guedes et al. 2014a). Additionally, most of them are also recorded in SPA along their distribution and also in RN (Guedes et al. 2014b, this work). Because of this, only two species (*Amerotyphlops amoipira* and *A. pausicquamus*) are listed by ICMBio in threatened categories for the State, while none are listed in IUCN (ICMBio 2018, IUCN 2019). Nevertheless, evidence suggests that some species should be monitored carefully in future assessments. The coral snake *Micrurus potyguara* was recently described for a small area of the Atlantic Forest of RN, Paraíba, and Pernambuco states (Pires et al. 2014). Its habitat is reported to be declining, which might lead the species to be listed in IUCN categories in the future, as has occurred with other squamate species in similar conditions recently (Fazolato et al. 2017, Rosário et al. 2019). *Erythrolamprus mossoroensis* is a Caatinga endemic that occurs in four municipalities of RN; although this species has a large range (Guedes et al. 2014a), most records are old, possibly indicating a low-density population or lack of faunal inventories. Similarly, *Boiruna sertaneja* is considered rare in the neighboring state of Paraíba (Pereira-Filho et al. 2017) and might be threatened by intrinsic aspects of the species (Pizzatto 2005), besides anthropogenic factors. It is urgent for RN state to evaluate the conservation status of
its flora and fauna, including snakes, and draft lists of endangered species to guide political and conservation actions aiming to maintain the remaining habitats.

Our data updates the current knowledge of snakes in RN and the necessity for a significant increase in sampling effort within many poorly sampled regions of the state. The low number of records compared to other regions is likely the result of limited local inventories and fewer long-term studies on the snake fauna. Several undersampled regions might gather similar richness to those observed in the East region of RN, and even produce new state records or undescribed species. Finally, those areas reported herein as the richest in snake species in RN require protective policies and conservation actions for the species they harbor.

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**SUPPLEMENTARY MATERIAL**

**Table S1.** Detailed distribution of snakes in RN state with the recorded municipality of each species and the source material and voucher.

**Figure S1.** Geographic distribution maps of snake species and soil cover in Rio Grande do Norte State, Brazil. Leptotyphlopidae: (a) *Epictia borapeliotes*; Typhlopidae: (b) *Amerotyphlops amoipira* and *A. paucisquamus*; Boidae: (c) *Boa constrictor*, (d) *Epicrates assisi* and *Corallus hortulanus*; Viperidae and Elapidae: (e) Bothrops erythromelas and *B. leucurus*, (f) *Crotalus durissus*, *Micrurus potyguara* and *M. cf. corallinus*. (g) *M. aff. ibiboboca*, and Colubridae: (h) *Chironius flavolineatus* and *Palusophis bifossatus*. Stars indicate new state records.

**Figure S2.** Geographic distribution maps of snake species and soil cover in Rio Grande do Norte State, Brazil. Colubridae and Dipsadidae: (a) *Psomophis joberti*, (b) *Erythrolamprus poecilogyrus* and *Psomophis joberti*, (c) *E. viridis*, (d) *Helicops leopardinus*, *Leptodeira annulata*, and *Sibon nebulatus*, (e) *Lygophis dilepis*, (f) *Oxyrhopus trigeminus*, (f) *Philodryas nattereri*, (G) *P. olfersii*, and (H) *Phimophis guerini* and *Philodryas patagoniensis*. Stars indicate new state records.

**Figure S3.** Geographic distribution maps of snake species and soil cover in Rio Grande do Norte State, Brazil. Colubridae and Dipsadidae: (a) *Erythrolamprus poecilogyrus* and *Psomophis joberti*, (b) *E. viridis*, (c) *Helicops leopardinus*, *Leptodeira annulata*, and *Sibon nebulatus*, (d) *Lygophis dilepis*, (e) *Oxyrhopus trigeminus*, (f) *Philodryas nattereri*, (G) *P. olfersii*, and (H) *Phimophis guerini* and *Philodryas patagoniensis*. Stars indicate new state records.

**Figure S4.** Geographic distribution maps of snake species and soil cover in Rio Grande do Norte State, Brazil. Dipsadidae: (a) *Pseudoboa nigra*, (b) *Taeniophallus nigritus*, (c) *Thamnodynastes occipitalis*, (d) *Thamnodynastes almae*, (T. phoenix and T. sertanejo), and (d) *Xenodon merremii*. Stars indicate new state records.

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Author contributions

RM designed the manuscript, reviewed the specimens housed at LAR-UFRN, gathered all data and performed data interpretation, elaborated figures of heat and distribution maps, tables, and wrote the manuscript. TBG helped in the conceptualization, gathered data, review the analyses, writing - original draft, review and edit final manuscript. FML gathered data from literature, elaborated figures and writing. DCP provided primary data, contributed to the writing of early versions and reviewed the final version of manuscript. WPS collected part of the data. AAG conceived the idea of the paper, collected field data, and participated in the production of the manuscript. All authors discussed and reviewed the final version of the manuscript.