Clinical Approach to Mast Cell Activation Syndrome: A Practical Overview

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Abstract

The diagnosis of mast cell activation syndrome (MCAS) is defined by 3 criteria: (1) typical clinical signs and symptoms of acute, recurrent (episodic), and systemic mast cell activation (MCA); (2) increase in tryptase level to >20% + 2 ng/mL within 1-4 hours after onset of the acute crisis; and (3) response of MCA symptoms to antimediator therapy. Classification of MCAS requires highly sensitive and specific methodological approaches for the assessment of clonal bone marrow mast cells at low frequencies. The Spanish Network on Mastocytosis (REMA score) has been used successfully as a predictive model for selecting MCAS candidates for bone marrow studies based on a high probability of an underlying clonal mast cell disorder. In this article, we propose a diagnostic algorithm and focus on the practical evaluation and management of patients with suspected MCAS.

Key words: Anaphylaxis. Antimediator therapy. Mast cell activation syndrome. Mast cell mediator release–related symptoms. Tryptase.

Resumen

El diagnóstico de síndrome de activación mastocitaria (SAM) se basa en 3 criterios: 1) signos y síntomas específicos de activación mastocitaria aguda, recurrente y sistémica, 2) aumento de los valores de triptasa en un 20% + 2 ng/mL sobre el valor basal de cada individuo en el periodo comprendido entre 1-4 horas desde el inicio del cuadro agudo, y 3) resolución de los síntomas con tratamiento antimediador. Para realizar el diagnóstico de SAM, es preciso emplear métodos diagnósticos altamente sensibles y específicos capaces de detectar bajas cantidades de mastocitos en la médula ósea. El modelo predictivo de la Red Española de Mastocitosis (REMA score) resulta útil para identificar a los pacientes con mayor probabilidad de padecer una patología mastocitaria clonal y que, por tanto, requieren que se realice un estudio de médula ósea en el proceso diagnóstico. En este artículo, proponemos un algoritmo diagnóstico para SAM y abordamos el manejo de estos pacientes desde un punto de vista práctico en la consulta alergológica.

Palabras clave: Anafilaxia. Síndrome de activación mastocitaria. Síntomas secundarios a la liberación de mediadores mastocitarios. Tratamiento antimediador. Triptasa.
**Introduction**

Mast cells (MCs) are myeloid lineage–derived cells. They are present in connective tissue and play an important role as immunomodulatory and effector cells by releasing mediators that provoke clinically relevant reactions [1-3]. Correct diagnosis of mast cell activation syndrome (MCAS) is usually challenging for clinicians, since it is necessary to rule out conditions that mimic MCAS (eg, cardiovascular, endocrinologic, gastrointestinal, rheumatologic, and immunologic disorders) [4]. Symptoms of mast cell activation (MCA) are frequently nonspecific and can present in diverse physiologic and pathologic conditions. Thus, MCAS must be considered an unusual entity and may be diagnosed according to the following criteria [2,5,6]: (1) acute, recurrent (episodic), and systemic (involving at least 2 organ systems) signs and symptoms of MCA consistent with a diagnosis of anaphylaxis; (2) an increase in tryptase level from baseline to >20% + 2 ng/mL within 1-4 hours after onset of the reaction; and (3) clinical response to therapy with MC-stabilizing agents that target MC mediator production and secretion or receptor binding. All 3 criteria must be fulfilled to establish the diagnosis of MCAS. Despite their great utility, they also present some limitations. Furthermore, since MCAS and its diagnostic criteria are primarily reported for adults, further studies are required for the evaluation of this entity in children [7].

**Clinical Signs and Symptoms of MCAS [2,3,8]**

MCA symptoms are secondary to the release of various vasoactive and proinflammatory MC mediators such as histamine, prostaglandins (PGs), leukotrienes (LTs), proteases, platelet-activating factor (PAF), growth factors, and cytokines. Different combinations of these mediators may be involved in MCA-related symptoms, which can present as acute episodes and/or chronic symptoms. Symptoms of MCA range from mild to severe and may even be life-threatening. Moreover, the trigger may be known (IgE- or non–IgE-mediated) or unknown. Thus, MCAS must be considered an unusual entity and may be diagnosed according to the following criteria [2,5,6]: (1) acute, recurrent (episodic), and systemic (involving at least 2 organ systems) signs and symptoms of MCA consistent with a diagnosis of anaphylaxis; (2) an increase in tryptase level from baseline to >20% + 2 ng/mL within 1-4 hours after onset of the reaction; and (3) clinical response to therapy with MC-stabilizing agents that target MC mediator production and secretion or receptor binding. All 3 criteria must be fulfilled to establish the diagnosis of MCAS. Despite their great utility, they also present some limitations. Furthermore, since MCAS and its diagnostic criteria are primarily reported for adults, further studies are required for the evaluation of this entity in children [7].

**Monitoring of MC Mediators in the Clinical Diagnosis of MCAS**

The MC mediators used as biomarkers of MCA in various disorders (eg, allergen-triggered systemic anaphylaxis, systemic anaphylaxis associated with systemic mastocytosis [SM], aspirin-exacerbated respiratory disease) include tryptase (in serum or plasma), urine histamine metabolites (N-methylhistamine [N-MH] and N-methylimidazole acetic acid [MIAA]), and the urine metabolites of PGD2 and LTC4, i.e., 11ß-PGF2α and LTD4/LTE4, respectively [3,6,13]. Histamine and tryptase are both produced and stored in tissue MCs and blood basophils. However, MCs contain >100-fold higher levels of tryptase than basophils [14], and immature (leukemic) basophils express relatively low amounts of tryptase [15]. Indeed, other cells can release histamine (eg, neutrophils, platelets, histamine-secreting carcinoid tumor cells), which is metabolized rapidly (half-life, 1-2 minutes), thus reducing the utility of this mediator as a clinical biomarker [13]. Nevertheless, some authors consider histamine-specific metabolites (N-MH and MIAA) to be appropriate biomarkers of systemic histamine release from MCs or basophils [13,16], although in one study [17], the measurement of 24-hour urine N-MH was elevated in only 2 out of 25 MCAS patients and showed little clinical utility for diagnosing MCAS.

While MCs are the major source of the previously mentioned mediators [13], other cell types also release such substances. PGD2 is produced mainly by Tp2 lymphocytes, dendritic cells, megakaryocytes, and eosinophils; PGF2 is synthesized by the luteal endometrium, gestational tissues, human and primate granulosa cells, and hepatocytes; and LTC4 can be generated by basophils, eosinophils, monocytes, macrophages, and platelets [3,13]. Thus, of all these mediators, serum tryptase is considered the most accurate parameter for the evaluation of MCA [2] and is the biomarker used as a criterion for diagnosis of MCAS.

The commercially available ImmunoCAP Tryptase assay (ThermoFisher Scientific) detects total tryptase. Of
A relationship has been reported between increased serum tryptase levels—either acute tryptase or acute/basal ratio (preferable)—detected during perioperative anaphylaxis, as has underlying IgE-mediated anaphylaxis [25-27]. On the other hand, when anaphylaxis is triggered by foods, the acute/basal ratio is more informative than peak tryptase determinations (usually in the normal range) [3,28-30].

Regarding postmortem serum tryptase, levels can be elevated in nonanaphylactic causes of death (eg, myocardial infarction, asphyxia, and trauma) [31,32]. A recent study established the postmortem tryptase reference value in nonanaphylactic death as <23 μg/L [31]. In addition, tryptase levels can vary depending on perimortem and postmortem factors, including the sampling technique: it is recommended to take blood samples from a clamped femoral/external iliac vein [31].

Serum tryptase levels should be analyzed in the 1-4 hours after a suspicious MCA episode, and the result must be compared with the individual’s previous serum baseline tryptase and/or the baseline levels must be determined at least 24-48 hours after the resolution of the clinical event (following the >20%/+2 formula) [6,33,34]. Of note, the sensitivity of this tryptase algorithm decreases with decreasing clinical severity and with delayed blood extractions after the resolution of symptoms [2].

Tryptase can be analyzed in blood. However, measurement of other mediators (eg, urine) requires a period of 24 hours according to specific guidelines, including dietary restrictions (ie, histamine metabolites) [16,17]. Serum tryptase analysis immediately after an MC crisis may be difficult owing to logistic concerns (it is not routinely performed in the emergency department); therefore, in the authors’ opinion, it is advisable to provide specific written instructions for the determination of serum tryptase to patients at risk of presenting an acute MCA crisis who require attention in the emergency department.

Although metabolites of PGD2 and cys-LTs (LTC4, LTD4, and LTE4) can be measured in random urine specimens [13] and serum samples [35], the commercial assays necessary to perform these analyses may not be available in some clinical settings. In addition, levels can be elevated in various reactive conditions (cell source might be ambiguous) and in mild mediator-related symptoms [2,3,6]. Thus, their contribution in the diagnosis of MCAS is not well defined, although some authors report their usefulness in guiding treatment that blocks the production of MC mediators [13].

Other parameters, such as diaminooxidase and heparin, are not currently recommended as biomarkers of MCAS [3,6].

Hereditary α-protryptase is released constitutively from MCs into plasma [18], while specific release of β-tryptase during anaphylaxis has been reported [19]. Increased tryptase levels in anaphylaxis induced by insect stings frequently correlate with the magnitude of hypotension during the episode. Furthermore, hypotension in anaphylaxis elicited by Hymenoptera venom is highly suspicious for underlying c-MCD [3,9,20-24].

An on-demand schedule is used to control acute MCA episodes and, in the most severe cases, corresponds to acute treatment of systemic anaphylaxis [29]. It consists of the following: (1) removing the trigger when possible; (2) assessing the patient’s circulation, airway, breathing, mental status, and skin; (3) placing the patient in the supine position (or in a position of comfort if there is respiratory distress and/or vomiting), with the legs raised; (4) calling for help and (self-) injection of intramuscular epinephrine in the mid-anterolateral thigh. If necessary, supplemental oxygen, intravenous fluids, cardiopulmonary resuscitation, and continuous noninvasive monitoring can be considered. In addition, intramuscular epinephrine is indicated in laryngeal angioedema and in severe bronchospasm, which can also be treated with inhaled rapid-acting β2 agonists [3].

Second-line medications, such as H1 and H2 antihistamines and corticosteroids, are also recommended in the treatment of anaphylaxis or acute MCA episodes [8,40]. Patients at risk for such events, as well as their relatives and care providers, should carry an epinephrine injector and be trained in the treatment of acute episodes.

As regards the prevention of presentation of MCA-related symptoms, it is important to avoid or adequately manage the general and specific triggers that may elicit release of MC mediators (eg, Hymenoptera venom, nonsteroidal anti-inflammatory drugs, opioids, anesthetic procedures, iodinated contrast media) [3,40,41].

Furthermore, continuous antimediator therapy should be selected according to intensity and/or severity of MCA signs.
and symptoms. It is also important to evaluate possible MC mediator–related symptoms recorded between acute systemic MCA (anaphylactic) episodes.

Following previous recommendations, and based on the practice of mastocytosis experts [3,8,9,42-44], various drugs (alone or in specific combinations) are indicated, as follows: (1) oral sodium cromolyn (MC stabilizer); (2) scheduled or on-demand nonsteroidal H1 antihistamines (preferable) combined with a sedating antihistamine at night or on demand in selected highly symptomatic cases [44,45]; (3) scheduled H1 antihistamines; (4) scheduled leukotriene antagonists; and (5) corticosteroids for uncontrolled MC mediator–related symptoms. Table 1 shows a stepwise antimediator therapy approach for control of MC mediator–related symptoms.

Despite the lack of high-quality evidence based on well-designed, double-blind, and placebo-controlled randomized trials (DBPCRT) to support recommendations regarding choice of H1 antihistamine or dosing [46], interesting data have been reported on this topic. Desloratadine and ketotifen have MC-stabilizing properties [47,48]. In addition to its antimediator activity, loratadine inhibits spontaneous growth of neoplastic MCs in vitro [49]. PAF is a lipid-derived mediator involved in episodic hypotension and flushing in mastocytosis [50]. Rupatadine exerts an antagonistic effect against PAF receptor [51] and improves quality of life and MC-related symptoms (ie, itch, wheal, flare, flushing, tachycardia) in mastocytosis patients based on the results of a DBPCRT [50]. To a lesser extent, rupatadine and levocetirizine inhibit PAF degranulation induced in MCs in vitro [52]. H1 antihistamines are used specifically for treating gastric hypersecretion and peptic ulcer–related symptoms in patients with mastocytosis. They can also enhance the effect of H2 antihistamines used specifically for treating gastric hypersecretion and peptic ulcer–related symptoms in patients with mastocytosis.

Table 1. Stepwise Antimediator Therapy for MC Mediator–Related Symptoms in MCAS [3,8,40,42,43]

| Symptoms                      | Therapy for chronic/recurrent symptoms                                                                 | Therapy for acute episodes                                                                 |
|-------------------------------|--------------------------------------------------------------------------------------------------------|---------------------------------------------------------------------------------------------|
| Pruritus, urticaria, and/or angioedema | Nonsteroidal H1, up to 4-fold doses + Topical cromolyn if skin lesions or pruritus are restricted to small areas + Oral cromolyn + Omalizumab if uncontrolled CSU + Sedating H1 if uncontrolled episodes + Systemic corticosteroids | Nonsteroidal H1 + Topical cromolyn if skin lesions or pruritus are restricted to small areas + Systemic corticosteroids + Epinephrine if acute laryngeal angioedema |
| Flushing                      | Oral cromolyn + nonsteroidal H1 + H2 + ASA if previous tolerance is demonstrated + Sedating H1 + LT antagonists | Nonsteroidal H1 + Sedating H1 + Systemic corticosteroids                                      |
| Abdominal cramping and/or diarrhea | Oral cromolyn + Nonsteroidal H1 + H2 + Oral budesonide cycles + LT antagonists + COX-2-I if previous tolerance is demonstrated + Low-dose systemic corticosteroid cycles | Nonsteroidal H1 + Sedating H1 + Oral budesonide + Antispasmodic drugs if acute episodes of pain + Anti-diarrheal drugs; if acute, severe, and uncontrolled episodes of diarrhea + Low-dose systemic corticosteroids |
| Peptic symptoms               | Oral cromolyn + H2 + PPI + VIT, if hymenoptera venom allergy + Nonsteroidal H1 + H2, if idiopathic, stress-induced, or uncontrolled anaphylaxis + Anxiolytics or antidepressants if stress-induced anaphylaxis + Sedating H1 if stress-induced anaphylaxis + Omalizumab, if uncontrolled anaphylaxis or bad tolerance to VIT + IFNα + TKIα | H1 + PPI + Epinephrine + Nonsteroidal or sedating H1 + Systemic corticosteroids + H2 + Inhaled short β2 agonist if bronchospasm + Fluids + Vasoactive drugs |

Abbreviations: ASA, acetylsalicylic acid; COX2-I, cyclooxygenase 2 inhibitor; CSU, chronic spontaneous urticaria; H1, H1 antihistamine; H2, H2 antihistamine; LT, leukotriene; MC, mast cells, MCAS, mast cell activation syndrome; PPI, proton pump inhibitor; TKI, tyrosine-kinase inhibitor; VIT, venom immunotherapy.

*In selected unresponsive cases.
*Evaluate known tolerance if an opioid drug is prescribed.
*Avoidance of the specific trigger in food or drug anaphylaxis.
*Some cases may not require combination with cromolyn.
antihistamines and prove useful in patients with abdominal pain, diarrhea, and recurrent, severe episodes of MC mediator release [45,53-56]. The precise mechanism of action of the MC stabilizer sodium cromolyn remains unclear. However, it inhibits activation of MCs and release of mediators by MCs in vitro and in vivo, inhibits GTP-g-S–induced exocytosis of MCs, and modulates sensory nerve function [57,58]. Moreover, despite its limited systemic absorption [59], DBPCRTs have shown that oral sodium cromolyn is effective for controlling gastrointestinal symptoms such as abdominal pain, vomiting, and diarrhea, in addition to other clinical manifestations associated with the release of MC mediators, such as pruritus and flushing [54,60-64]. Based on its experience, the Spanish Network on Mastocytosis (REMA), recommends the use of oral disodium cromolyn as treatment for chronic/recurrent MCA symptoms such as flushing, abdominal cramping, diarrhea, and unprovoked anaphylaxis [8,44,53]. Standard doses of oral disodium cromolyn for the adult population are 600-800 mg/d, although this could be increased to 1600 mg/d if necessary [44]. Acetylsalicylic acid might control flushing and hypotension in selected cases with known tolerance to the drug and elevated urinary 11B-PGF_2α [3]. Celecoxib could be considered an option for intractable diarrhea in mastocytosis patients [65]. Montelukast has been shown to improve respiratory, cutaneous, and gastrointestinal symptoms in this population [66-68]. Systemic corticosteroids, which must be limited for long-term use owing to adverse effects, are recommended for refractory, acute, and/or severe MC mediator release–related symptoms. Corticosteroids can improve abdominal pain that is refractory to other options, and short cycles of low-doses prednisone (0.3 mg/kg/d) or oral budesonide (0.1 mg/kg/d) may be prescribed [8,44,54,69]. Indeed, anti-IgE therapy with omalizumab has proven useful in cases with anaphylaxis that does not respond to conventional antimediator therapy [3,8,9,40], as well as for reaching maintenance doses of venom immunotherapy (VIT) [70]. VIT is recommended in IgE-mediated Hymenoptera anaphylaxis requiring lifelong therapy [71,72]. Currently, there is no consensus regarding recommendations about stepwise antimediator therapy or regarding the number of drugs or specific combinations to establish lack of response. The REMA defines the lack of response to antimediator therapy after failure to at least a combination of oral cromolyn, H1, and H2 antihistamines, antileukotrienes, and acetylsalicylic acid (or other cyclooxygenase inhibitors) [8]. A provisional diagnosis of “possibly MCAS” may be established in patients who present with acute, systemic, and recurrent MC symptoms and a diagnostic increase in serum tryptase levels, in the absence of improvement with conventional antimediator therapy, which should be increased [2]. Bone mass loss, specifically osteoporosis, constitutes an important public and treatable health problem that should be evaluated in clonal MCs. It is a frequent finding in systemic mastocytosis (especially among patients without cutaneous involvement) [10,73], secondary to local MC infiltration and disturbances in bone remodeling owing to MC mediators such as IL-6, histamine, and heparin [74,75]. Calcium, vitamin D supplements, and bisphosphonates are usually prescribed [76]. On the other hand, denosumab [77] and interferon α-2b [78,79] might also be considered in patients with severe osteoporosis at risk of pathologic bone fractures who do not respond to conventional treatments.

Classification of MCAS

The current classification of MCAS establishes the following categories: (1) primary MCAS, where KIT-mutated, clonal (CD25+) MCs are detected (with or without an underlying diagnosis of mastocytosis); (2) secondary MCAS, in which usually an IgE-dependent allergy, another hypersensitivity reaction, or another immunologic disease that can evoke MCA is detected; and (3) idiopathic MCAS, with no detection of KIT-mutated MCs, other inflammatory disorders that may explain MCA, or a trigger for a hypersensitivity reaction [2]. According to the REMA experience, around 5% of patients presenting with anaphylaxis in the absence of the typical skin lesions for mastocytosis may have an underlying clonal MCAS (c-MCAS) [80]. When the diagnostic criteria of MCAS are fulfilled, the evaluation of the typical D816V KIT mutation (or other gain-of-function KIT mutations) [81] should be considered. Immunophenotyping of bone marrow MCs is usually necessary to confirm or rule out a primary (clonal) MCAS. Some REMA data suggest greater utility for detection of the D816V KIT mutation, as assessed by allele-specific oligonucleotide quantitative polymerase chain reaction (ASO-qPCR) in fluorescence-activated cell sorting (FACS) of purified bone marrow MCs rather than in peripheral blood or unfractionated bone marrow samples [8]. If there is no evidence of clonality (CD25+ bone marrow MCs and no KIT mutations), a nonclonal MCAS (nc-MCAS) should be considered, although the term nonclonal is based on the absence of currently detectable clonality [40]. Furthermore, whether clonal or not, co-occurrence of allergy or other underlying conditions according to the clinical features should be evaluated (Table 2) [8,9,40]. Thus, patients with an underlying c-MCD who experience IgE-mediated anaphylaxis after Hymenoptera sting could be more specifically categorized [8,10,40,82]. It is noteworthy that MCAS patients with associated skin lesions are usually c-MCD (either cutaneous mastocytosis or SM) [8]. Therefore, it has been recommended to use the diagnostic label ‘SY’ for symptoms (ie, indolent SM–ISM– ISM_SY–) in mastocytosis cases, with any form of MC requiring continuous antimediator therapy, even if the criteria for MCAS are not met [2,33].

Bone Marrow Aspirate and Biopsy: How and When?

The bone marrow study does not quantify activation of MCs and does not have to be performed before starting antimediator therapy. It is well known that mastocytosis (or c-MCD) patients present remarkable clinical heterogeneity in the severity of MC mediator–related symptoms [9]. Furthermore, cytoreductive and/or targeted therapies should
The evaluation of patients with suspected or confirmed MCAS should include a clinical, physical, and allergological work-up, together with a routine peripheral blood count and differential, routine biochemistry, and serum baseline tryptase (sBT). In addition, the bone marrow study is mandatory for the classification of MCAS (Table 2), and for diagnosis of underlying c-MCD in the absence of the typical skin lesions of mastocytosis (c-MCAS or SM) [9,40]. A relatively low MC burden in c-MCAS and indolent systemic mastocytosis without skin lesions of mastocytosis (ISMs–) has also been reported [9,40,73]. Highly sensitive and specific methodological approaches for the study of bone marrow MCs are required, including detailed cytological analysis of bone marrow smears, histology, immunochemistry, and flow cytometry–based immunophenotyping using specific gating strategies for detecting MCs present at low frequencies. Indeed, ASO-qPCR with unfractionated bone marrow and FACS with purified bone marrow are the techniques to be applied for detection of KIT mutations [81]. If the latter fails, the mutation could also be explored using peptide nucleic acid–mediated PCR clamping in FACS-purified bone marrow MCs. Finally, another useful option is sequencing of the whole KIT gene [8]. These complete methods are usually only available in high-efficiency reference centers for the diagnosis of c-MCD [83].

Table 2. Classification of Mast Cell Activation Syndromes [2,8]

| Molecular category | Recognized category | Diagnostic features of MCs | Underlying conditions |
|--------------------|---------------------|---------------------------|----------------------|
| Clonal MCs         | Primary             | D816V KIT mutationa and/or aberrant expression of CD25+ in MCs in BM (WHO minor SM criteria) | c-MCAS (or MMAS) |
|                    |                     | WHO criteria for SM are fulfilled | SM |
|                    |                     | Infiltration of skin by MCs, in the absence of WHO criteria for SMb | CM |
| Nonclonal MCs      | Secondary           | No KIT mutations detecteda Expression of CD25– in MCs in BM | IgE-mediated allergy, another hypersensitivity reaction, or another immunologic (autoimmune, inflammatory) disease that causes MCA |
| Idiopathic         |                     | No KIT mutations detecteda Expression of CD25– in MCs in BM | Neither primary nor secondary conditions are found |

Abbreviations: BM, bone marrow; CM, cutaneous mastocytosis; c-MCAS, clonal mast cell activation syndrome; MC, mast cells; MCA, mast cell activation; MCAS, mast cell activation syndrome; MMAS, monoclonal mast cell activation syndrome; SM, systemic mastocytosis; WHO, World Health Organization.

aOther gain-of-function KIT mutations are described [3,81].
bSkin MC infiltrate is accepted to be clonal MC proliferation.
cPotential existence of unknown molecular defects cannot be ruled out. Adapted with permission from Elsevier [8].

Some predictive models, such as the REMA score [73,84,85] or the National Institutes of Health clinical activity score (including allele-specific PCR the KIT D816V mutation in peripheral blood) [86], have proven to be useful for selecting MCAS candidates for bone marrow studies based on a high probability of underlying c-MCD. The REMA score (Table 3) is based on sex, symptoms and signs observed during the acute episodes, and sBT levels. Below, we provide 2 brief examples of how to use the REMA score to evaluate systemic acute episodes in adult patients in the absence of skin lesions of mastocytosis (in some cases, patients can experience various episodes with different MCA-related symptoms each time): (1) In the case of a 35-year-old man who presented with dizziness and loss of consciousness after a wasp sting, the allergology work-up showed sensitization to Polistes dominula venom and an sBT of 10 ng/mL. The REMA score was 4: male (+1), no urticaria, no pruritus and no angioedema (+1), syncope (+3), and sBT <15 ng/mL (–1). (2) A 35-year-old man who presented with generalized urticaria, throat swelling, bronchospasm, abdominal cramping, diarrhea with no identified trigger after a thorough allergology work-up, and an sBT of 22 ng/mL; the REMA score was –1: male (+1), urticaria and angioedema (–2), and sBT not applicable.

Table 3. REMA Score

| Variable | Scorea |
|----------|--------|
| Sex      |        |
| Male     | +1     |
| Female   | –1     |
| Clinical symptoms |    |
| No urticaria, no pruritus and no angioedema | +1 |
| Urticaria, pruritus and/or angioedema | –2 |
| Presyncpe or syncpe | +3 |
| sBT      |        |
| <15 ng/mL | –1 |
| >25 ng/mL | +2 |

Abbreviations: MCAS, mast cell activation syndrome; REMA, Spanish Network on Mastocytosis; sBT, serum basal tryptase. Score <2: low probability of clonal MCAS. Score ≥2: high probability of clonal MCAS. Sensitivity, 0.92; positive predictive value, 0.89; specificity, 0.81; negative predictive value, 0.87. Reproduced with permission from Elsevier and Karger [73,84].
The Figure proposes an algorithm for the diagnosis of MCAS. A REMA score ≥2 is highly specific and sensitive for ISMs− or c-MCAS, and a bone marrow study is indicated. On the other hand, a REMA score <2 usually indicates nonclonal disease in this situation. If sBT levels are <25-30 ng/mL, then the KIT D816V mutation should be identified in peripheral blood using ASOqPCR, and, if positive, a bone marrow study would be also indicated. Finally, the European Competence Network on Mastocytosis recommends performing a bone marrow study in cases with sBT ≥25-30 ng/mL. In these cases, SM and other clonal bone marrow diseases (eg, myeloproliferative neoplasm, myelodysplastic syndrome, or myeloid leukemia), renal failure, and genetic syndromes (eg, HtT) should also be evaluated [85].

**Conclusions**

This article provides recommendations on the diagnosis and management of MCAS according to the most recent studies and consensus guidelines. Diagnosis of MCAS is based on 2 clinical criteria and 1 analytical criterion (blood sample) that can be applied as part of clinical routine. The subsequent classification based on the presence of clonal or nonclonal bone marrow MCs is often only available in specialized centers. In addition, other underlying conditions (eg, allergy) should be evaluated according to the clinical picture in each case. Patients with MCAS are usually cared for by multidisciplinary teams owing to the marked heterogeneity in clinical presentation and the methodological approaches required for management and classification. Further studies and advances are necessary to determine the frequency of MCAS in adults and children and thus specify and standardize clinical recommendations on stepwise antimediator therapy. It is also essential to address other areas, such as defining the criteria for characterizing the lack of response to antimediator therapy, improving the measurement and monitoring of MC mediators other than tryptase, understanding the relevance of increased copy numbers of the TPSAB1 gene in MCAS, improving the efficiency in the detection of the KIT mutation in peripheral blood, and better characterizing nonclonal MCAS.

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**Conflicts of Interest**

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