Review Article
Diagnostic Utility of CSF Tau and Aβ42 in Dementia: A Meta-Analysis

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Received 3 December 2010; Revised 5 September 2011; Accepted 6 September 2011

1. Introduction

With the increase in life expectancy, Alzheimer’s disease, considered as disease of aging population, has become a major public health problem adding burden to societal costs each year for chronic care and lost productivity in developed and developing countries [1]. Presently, diagnosis of AD is primarily based on the exclusion of other causes of dementia on clinical trials. However, it is not of much help due to overlapping of clinical features of AD with other dementias in early stages. Attempts have been made in the last 4-5 decades to develop and validate specific biological markers which are able to detect the fundamental neuropathological changes occurring in AD in its early stage with high sensitivity (≥80%) and distinguish it from other dementias [2]. Based on these studies, the combination of total tau and amyloid β42 (Aβ42) was identified as being among the most promising and informative AD markers to be of use in early diagnosis and as surrogate biomarkers in CSF [2–4]. Increased levels of tau in CSF have been suggested to reflect neuronal and axonal degeneration [5] whereas reduced CSF levels of Aβ42 might reflect extracellular accumulation of Aβ42 into insoluble senile plaques in the AD brain [6, 7]. However, high CSF concentration of total tau has also been reported in mild cognitive impairment [8], as well as in vascular dementia (VaD) [2, 9–11]. The comorbidity of AD and other dementias can further generate problems. Though studies have reported highly accurate differentiation between AD and normal controls with high sensitivity (50–94%) and specificity (83–100%) [12], CSF-based differentiation of AD with VaD remains a challenge with specificity of 48% only [13]. Also, AD can be associated with other neurodegenerative diseases like Lewy body disease and progressive supranuclear palsy.

In view of the necessity of identifying biomarkers for differentiation of AD from other dementia disorders, the combination of CSF tau and Aβ42 has been advocated as diagnostic marker. However, before the diagnostic utility of CSF tau and Aβ42 concentration for diagnosis of AD can be established, it is crucial to assess whether the effect sizes of the published studies reported in the last fifteen years show consistent trends in their levels or not in AD cases as compared to controls. The consistency in effect size of tau and Aβ42 levels of these studies was explored by advanced
2 Materials and Methods

2.1 Search Strategy. Meta-analysis was performed for CSF tau and \( A\beta_{42} \) levels, by calculating and combining the effect sizes (raw mean difference), their standard error, and 95% confidence interval after extracting mean, standard deviation, and sample size from 11 CSF studies. 11 studies out of 60 studies for CSF tau levels and 7 studies out of 41 studies were selected for CSF \( A\beta_{42} \) levels in dementias from 1996 to 2009 using keywords CSF biomarkers in Alzheimer’s disease, tau levels in Alzheimer’s disease, and \( A\beta_{42} \) levels in Alzheimer’s disease.

2.2 Exclusion Criteria. Studies were excluded if they were not in English language, did not provide data for controls, did not mention the diagnostic criteria for AD, and did not report the SD of the mean CSF tau and \( A\beta_{42} \) level or their units were not in pg/mL. Also, studies where tau and \( A\beta_{42} \) were measured by methods other than sandwich enzyme linked immunosorbant assay were excluded. Only those studies were undertaken in which CSF tau and \( A\beta_{42} \) levels were measured in AD and other dementias (vascular dementia, Parkinson’s disease-associated dementia, Lewy body dementia) in controls using Innogenetics, Belgium. All the studies included had interday and intraday variation less than 10%. Other-dementias group was named dementia due to other causes (DOC). All the values were expressed in pg/mL.

2.3 Statistical Analysis. The raw mean difference (unstandardized) was used for calculating the effect size of each study. The heterogeneity in effect size of both parameters (tau and \( A\beta_{42} \)), which may occur from study to study and is one of the serious issues of any meta-analysis, was explored with the help of three statistical tools: (1) test of significance (Q statistics), (2) between-studies variance \( (I^2) \), and (3) degree of heterogeneity \( (P^2) \). An appropriate model (fixed effect versus random effect model) for getting the pooled effect size was decided after evaluating the heterogeneity in effect size of all the included studies.

The graphical method (Forest plot) has been applied for studying the variability between the effect sizes of individual studies. In Forest plot, each study effect size and respective confidence interval (CI) were plotted on one set of axis along with pooled estimate of effect size, together with its CI.

All the statistical analysis was done using the Meta-analysis report software (Version: Beta3.13) downloaded from the web site on 05.10.10.

3 Results

3.1 Meta-Analysis of CSF Tau Level in Dementia. A meta-analysis was performed in 11 published studies of CSF tau in AD and DOC, selected from total of 60 studies. Out of total published studies included in meta-analysis of CSF tau levels in DOC 5 published studies were of vascular dementia (VaD) [7, 18, 22–24], 2 of Parkinson’s disease-associated dementia (PDD) [15, 20], 1 of mixed dementia [16], and the rest were non-AD dementia (non-ADD) [17, 19]. Out of 11 studies, only two studies in AD [15, 23] and one study in DOC [23] were having matched controls.

Table 1 shows that details of 11 included studies along with mean, SD, and sample size (N) of CSF tau level for cases and controls. Effect size (raw mean difference), standard error, 95% confidence interval and weight for each included study were calculated. None of studies show effect size zero in both group (Alzheimer’s disease and DOC).

Table 2 shows that median sample size (case + control) in 11 Alzheimer’s disease studies of CSF tau level was 67 (range: 30–142) whereas it was 49 (range: 26–134) in dementia-due-to-other-causes studies. The median effect size (raw mean difference) of tau level in Alzheimer’s disease studies was 429 pg/mL (range: 32 to 910 pg/mL) whereas in dementia-due-to-other-causes studies it was 69 pg/mL (range: −53 to 518 pg/mL). In Alzheimer’s disease, CSF tau level is quite high as compared to DOC.

Table 3 shows the estimated pooled effect size of CSF tau level and its 95% confidence interval (CI) of 11 included studies for both groups. Between-studies variance, degree of heterogeneity with 95% CI, Q statistics, degree of freedom, and \( P \) value were also shown in the same tables. From Table 3, it can be seen that between-studies variance, degree of heterogeneity, and Q statistics all three approaches clearly indicate that variability in CSF tau estimates across the studies was too high and random effect model for pooling of CSF tau estimates is the only choice. After applying random effect model for pooling of estimates, the pooled estimate of CSF tau in Alzheimer’s disease studies was 414.073 pg/mL (CI: 237.170–590.975) while it was quiet low as 83.500 pg/mL (CI: 36.406–130.595) in dementia-due-to-other-causes studies.

Forest plot (Figure 1(a)) shows that out of 11 Alzheimer’s disease studies, effect size (raw mean difference) of 6 studies [13, 15, 18, 20, 22, 23] was more than pooled estimate of effect size, and in 5 studies [16, 17, 19, 21, 24], it was below the pooled effect size, whereas out of 11 dementia-due-to-other-causes studies (Figure 1(b)), 5 studies were having...
| S. no. | Author's name and year | Country where the studies were conducted | Tau level (pg/mL) (AD and control) | Effect size (pg/mL) | 95% CI | Weight | Tau level (pg/mL) (DOC and control) | Effect size (pg/mL) | 95% CI | Weight |
|-------|------------------------|------------------------------------------|------------------------------------|--------------------|--------|--------|------------------------------------|--------------------|--------|--------|
|       |                        |                                          | N | Diseased | M^*_d | SD | N | Control | M^*_c | SD | RMD^* | SE | N | Diseased | M^*_d | SD | N | Control | M^*_c | SD | RMD^* | SE | N | Diseased | M^*_d | SD | N | Control | M^*_c | SD | RMD^* | SE |
| 1     | Arai et al. (1995) [15] | Japan                                    | 19 | 919     | 349   | 19 | 9  | 4.5  | 910.00 | 80.07 | 7   | 77.2  | 45.5 | 19 | 9  | 4.5  | 68.20 | 17.23 | 34.43–101.97 | 0.215 |
| 2     | Tapiola et al. (1998) [16] | Finland                                  | 81 | 524     | 351   | 33 | 293 | 140  | 231.00 | 45.99 | 0.015 | 40  | 403  | 248  | 33 | 293 | 140  | 110.00 | 46.17 | 19.51–200.49 | 0.030 |
| 3     | Galasko et al. (1998) [17] | USA                                      | 82 | 663     | 481   | 60 | 387 | 167  | 276.00 | 57.33 | 0.010 | 74  | 456  | 383  | 60 | 387 | 167  | 69.00  | 49.47 | −27.96–165.96 | 0.026 |
| 4     | Andreasen et al. (1998) [18] | Sweden                                   | 43 | 796     | 382   | 18 | 190 | 57   | 606.00 | 59.78 | 0.009 | 21  | 708  | 382  | 18 | 190 | 57   | 518.00 | 84.44 | 352.51–683.49 | 0.009 |
| 5     | Kanai et al. (1998) [19] | Japan                                    | 32 | 489     | 297.5 | 41 | 217 | 128  | 272.00 | 56.26 | 0.010 | 33  | 267  | 146  | 41 | 217 | 128  | 50.00  | 32.34 | −13.38–113.38 | 0.061 |
| 6     | Andreasen et al. (2001) [7] | Sweden                                   | 105 | 759    | 417   | 18 | 264 | 102  | 495.00 | 47.27 | 0.014 | 23  | 461  | 280  | 18 | 264 | 102  | 197.00 | 63.14 | 73.25–320.75 | 0.016 |
| 7     | Sjögren et al. (2002) [20] | Sweden                                   | 19 | 919     | 349   | 17 | 342 | 116  | 577.00 | 84.87 | 0.004 | 15  | 306  | 65   | 17 | 342 | 116  | −36.00 | 32.76 | −100.21–28.21 | 0.059 |
| 8     | Grossman et al. (2005) [21] | US                                       | 17 | 534.6   | 303.5 | 13 | 260.4 | 93.8  | 274.20 | 78.07 | 0.005 | 73  | 207.1 | 270.3 | 13 | 260.4 | 93.8  | −53.30 | 40.95 | −133.58–26.98 | 0.038 |
| 9     | de Jong et al. (2006) [22] | The Netherlands                          | 61 | 613     | 326   | 30 | 184 | 89   | 429.00 | 44.79 | 0.016 | 50  | 303  | 307  | 30 | 184 | 89   | 119.00 | 46.36 | 28.14–209.86 | 0.030 |
| 10    | Bibl et al. (2007) [23] | Germany                                  | 15 | 700     | 480   | 15 | 200 | 130  | 500.00 | 128.40 | 0.002 | 15  | 340  | 280  | 15 | 200 | 130  | 140.00 | 79.71 | −16.22–296.22 | 0.010 |
| 11    | Ravaglia et al. (2008) [24] | Italy                                    | 31 | 56.3    | 32.7  | 36 | 24.28 | 5.9   | 32.02  | 5.96  | 0.909 | 13  | 63.5  | 40.3  | 36 | 24.28 | 5.9   | 39.22  | 11.22 | 17.23–61.21 | 0.506 |

*Mean in diseased group, **Mean in control group and raw mean difference.
individual effect size more than pooled estimate of effect size [13, 16, 18, 22, 23], while in 6 studies, individual effect size was below the pooled effect size [15, 17, 19–21, 24].

### 3.2. Meta-Analysis of CSF Amyloid $\beta_{42}$ Level in Dementia

A meta-analysis was performed in 7 published studies of CSF tau in AD and DOC, selected from total of 41 studies. Out of total published studies included in meta-analysis of CSF $\beta_{42}$ levels in DOC 3 published studies were of vascular dementia (VaD) [13, 22, 23], 1 of Parkinson’s disease-associated dementia (PDD) [20], 1 of semantic dementia [21], and the rest were non-AD dementia (non-ADD) [17, 19].

Table 4 shows that details of 7 included studies along with mean, SD, and sample size (N) of CSF $\beta_{42}$ level for cases and controls. Effect size (raw mean difference), standard error, 95% confidence interval, and weight for each included study were calculated. None of studies show effect size zero in both group (Alzheimer’s disease and dementia due to other causes).

Table 5 shows that median sample size (case + control) in 7 Alzheimer’s disease studies of CSF $\beta_{42}$ level was 73 (range: 30–142) whereas it was 41 (range: 26–134) in dementia-due-to-other causes studies. The median effect size (raw mean difference) of $\beta_{42}$ level in Alzheimer’s disease studies was $-442$ pg/mL (range: $-652$ to $-41.200$ pg/mL) whereas in dementia-due-to-other causes studies it was $-193$ pg/mL (range: $-356$ to $-33$ pg/mL). In Alzheimer’s disease, median CSF $\beta_{42}$ level is quite low as compared to DOC.

Table 6 shows the pooled effect size of CSF $\beta_{42}$ level and its 95% confidence interval (CI) of 7 included studies. Between-studies variance, degree of heterogeneity with 95% CI, Q statistics, degree of freedom, and $P$ value were also shown in the same tables. From Table 6, it can be seen that between-studies variance, degree of heterogeneity, and Q statistics all three approaches clearly indicate that variability in CSF $\beta_{42}$ estimates across the studies was too high and random effect model for pooling of CSF $\beta_{42}$ estimates is the only choice. After applying random effect model for pooling of estimates, the pooled estimate of CSF $\beta_{42}$ in Alzheimer’s disease studies was $-363.926$ pg/mL (CI: $-542.007$ to $-185.845$) while it was quiet high as $-170.743$ pg/mL (CI: $-256.912$ to $-84.574$) in DOC studies.

Forest plot (Figure 2(a)) shows that out of 7 Alzheimer’s disease studies, effect size (raw mean difference) of 2 studies was more than pooled estimate of effect size [19, 21] and, in 05 studies, it was less than the pooled effect size [13, 17, 20, 22, 23]. Of 7 DOC studies (Figure 2(b)), 02 studies were also having individual effect size more than pooled estimate of effect size [19, 21] while in 05 studies individual effect size was below the pooled effect size [13, 17, 20, 22, 23].

Funnel plots showed clear existence of publication bias in both groups.

### 4. Discussion

Diagnostic markers for AD have been sought for many years for its early diagnosis and differentiating it from other types of dementias. Distinguishing between the two most common forms of dementias, AD and VaD, is one of the most challenging differential diagnoses in geriatrics OPD and is very crucial also because the therapeutic strategies are very different. With the development of cholinergic-based treatments for AD, there is great emphasis on the need for its early and accurate diagnosis to allow initiation of therapy when it will be of most benefit to the patients [17]. CSF biomarkers tau and $\beta_{42}$ combined together have emerged as such diagnostic markers for early diagnosis of AD. Although combination of CSF tau and $\beta_{42}$ yields a highly accurate differentiation between AD and controls, CSF-based differentiation of AD from other dementias especially VaD
remains highly challenging as it has low specificity for the same [13].

4.1. Meta-Analysis of CSF Tau Level. A meta-analysis performed in 11 published studies of CSF tau in different types of dementias showed high levels of tau in AD in all the studies whereas 2 studies showed low levels in DOC as compared to controls. A number of studies have found a significant increase in CSF tau levels in AD and DOC as compared with normal ageing [25–27].

Table 2 shows that raw mean difference of CSF tau levels in AD group was much higher than that in DOC group signifying that levels of tau in CSF in AD cases rise much higher than in DOC. This finding supports the studies showing much higher sensitivity and specificity of CSF tau levels in distinguishing the AD patients from control subjects as compared to non-AD dementias [16] and VaD [13].

The assessment of the heterogeneity in meta-analysis is a crucial issue because the presence versus absence of true heterogeneity (between-studies variability) can affect the choice of statistical model. There can be two sources of variability leading to heterogeneity in a set of studies in a meta-analysis. First, variability due to sampling error (within-study variability) is always present in a meta-analysis because every single study uses different samples. The other source of variation is due to heterogeneity among the population effect sizes estimated by the individual studies (between-studies variability). It may be due to variation in the characteristics of the samples, treatment and the design quality of the study [27]. In the present study, three statistical methods were applied to explore the true heterogeneity. They
Table 4: Effect size, its 95% CI, and % weight calculations for each selected study of CSF Aβ42 in Alzheimer’s disease and dementia due to other causes.

| S. no. | Author’s name and year | Country where the studies were conducted | Aβ42 level (pg/mL) (AD and control) | Effect size (pg/mL) 95% CI Weight | Aβ42 level (pg/mL) (DOC and control) | Effect size (pg/mL) 95% CI Weight |
|--------|------------------------|------------------------------------------|-------------------------------------|----------------------------------|-------------------------------------|---------------------------------|
| 1      | Galasko et al. (1998)  | USA                                      | 82 833 379 60 1485 473 −652.000 74.031 | −797.70−506.90 0.012 74 1129 464 60 1485 473 −356.000 81.475 | 1129 464 60 1485 473 −356.000 81.475 | 0.012 |
| 2      | Kanai et al. (1998)    | Japan                                    | 32 109.9 73.2 41 242 180 −132.100 30.947 | −192.75−71.45 0.069 33 209 180 41 242 180 −33.000 42.096 | 209 180 41 242 180 −33.000 42.096 | 0.069 |
| 3      | Andreasen et al. (2001) | Sweden                                   | 105 523 180 18 897 242 −374.000 59.684 | −490.98−257.02 0.019 23 704 321 18 897 242 −193.000 87.941 | 704 321 18 897 242 −193.000 87.941 | 0.019 |
| 4      | Sjögren et al. (2002)  | Sweden                                    | 19 411 99 17 853 161 −442.000 45.173 | −530.54−353.46 0.033 15 680 96 17 853 161 −173.000 46.251 | 680 96 17 853 161 −173.000 46.251 | 0.033 |
| 5      | Grossman et al. (2005) | USA                                       | 17 54 15.1 13 95.2 29.7 −41.200 9.015 | −58.87−23.53 0.819 13 44 14.1 13 95.2 29.7 −51.200 9.118 | 44 14.1 13 95.2 29.7 −51.200 9.118 | 0.819 |
| 6      | de Jong et al. (2006) | The Netherlands                          | 61 419 128 30 869 207 −450.000 41.193 | −530.74−369.26 0.039 25 655 220 30 869 207 −214.000 58.003 | 655 220 30 869 207 −214.000 58.003 | 0.039 |
| 7      | Bibl et al. (2007)     | Germany                                   | 15 310 90 15 810 320 −500.000 85.829 | −668.22−331.78 0.009 15 480 220 15 810 320 −330.000 100.266 | 480 220 15 810 320 −330.000 100.266 | 0.009 |

∗Mean in diseased group, **Mean in control group and raw mean difference.
strongly recommend the presence of heterogeneity in a set of 11 tau studies (Table 3). The test of significance \((P = 0.000)\) suggests the presence of true heterogeneity among the effect size of all 11 studies. The magnitude of the true heterogeneity due to between-studies variance \(\left(T^2 = 85013.958 \text{ for AD versus controls, } & 4343.165 \text{ for DOC versus controls}\right)\) was much higher and contributed to 97.9% in AD versus control and 82.4% in DOC versus control (degree of heterogeneity \(I^2\)) in total heterogeneity. All the studies selected, fulfilling the defined inclusion criteria, report a significant difference in CSF tau levels in AD and DOC participants versus controls. Despite the uniform pattern of CSF tau level changes in AD and DOC as compared to control, there was wide range of effect size among 11 articles under study in AD group \((32.02–910.00 \text{ pg/mL})\) as compared to DOC group \((-53.30–518.00 \text{ pg/mL})\). There were large differences in baseline levels across studies; 6 studies’ in AD versus control \([13, 15, 18, 20, 22, 23]\) and 05 studies in DOC versus control have effect size more than pooled estimate \([13, 16, 18, 22, 23]\), signify that dementia patients having higher CSF tau levels than control. However, the increase in CSF tau levels in AD was much higher than in DOC as compared to control in most studies. Only Ravaglia et al. 2008 \([24]\) found low effect size (raw mean difference) of tau levels in DOC as compared to AD. A number of confounding factors are responsible for the heterogeneity and wide range of CI of effect size found in the studies undergoing meta-analysis. First, very few studies could be included for meta-analysis as per the inclusion criteria in the present study; second, except one study \([17]\) all of the studies had very low sample size for DOC group. Also most of the studies were not age-, sex-matched, and of 11 AD studies, 7 studies were having more number of cases than controls \([13, 16–18, 20–22]\) while in 4 studies the number of control was either equal or more than the number of cases \([15, 19, 23, 24]\). Similarly, in DOC group, 6 studies were having more number of cases than controls \([13, 16–18, 21, 22]\). There are inconsistent findings reported showing discrepancies in the effect of these on CSF tau levels in AD versus controls. Some investigators have reported increase of tau with age \([15, 19]\) and a negative correlation with sex \([17]\) with a tendency for females to have higher tau levels than males whereas others found tau levels unaffected by age, age at onset, and AD duration, severity, and rate of progression and equally increased in early and late disease, in mild and severe disease \([17, 19, 21]\). Another confounding factor is variation in freezer shelf life introducing variation in storage conditions for CSF.

### 4.2. Meta-Analysis of Amyloid \(\beta_{42}\) Level

All the published studies undertaken in the present meta-analysis showed low CSF \(\beta_{42}\) levels in AD and DOC as compared to controls. However, one study effect size had wider confidence interval with upper limit more than zero in both AD and DOC \([21]\). Table 5 shows that median effect size of \(\beta_{42}\) levels in CSF in AD is \(-442 \text{ pg/mL}\) as compared to \(-193 \text{ pg/mL}\) in DOC which indicates that decrease in CSF \(\beta_{42}\) in AD is much higher than in DOC group as compared to controls. 

It is supported by the studies done in last two decades reporting low CSF \(\beta_{42}\) levels in early stages of AD with high degree of sensitivity and specificity \([25]\) as compared to other dementias-LBD \([13]\) and VaD \([13]\).

Out of 7 studies undertaken, 5 studies in AD versus control group had more number of cases than controls \([13, 17, 20–22]\) while the other 2 studies had either equal number of controls or less than cases \([19, 23]\), whereas in DOC versus control group, only 2 studies had more number of cases than controls \([13, 17]\) and the other 5 studies had either more or equal number of controls than cases \([19–23]\).

Same three statistical techniques as have been applied in CSF tau studies (mentioned above) were used to determine the true heterogeneity (between-studies variability) in effect size of 7 included studies (Table 6). The test of significance \((P = 0.000)\) rejects the null hypothesis of similar effect

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**Table 5:** Descriptive statistics of CSF \(\beta_{42}\) studies in Alzheimer’s disease and dementia due to other causes.

| CSF \(\beta_{42}\) level (pg/mL) | Alzheimer’s disease | Dementia due to other causes |
|----------------------------------|---------------------|----------------------------|
| Sample size                      | Median              | Range                      |
|                                  | 73                  | 30–142                     |
| Effect size                      | -442                | -652–41,200                |
|                                  |                     | -193                       |
|                                  |                     | -356–33                    |

| Statistics                        | Alzheimer’s disease | Dementia due to other causes |
|-----------------------------------|---------------------|----------------------------|
| Pooled estimate of CSF \(\beta_{42}\) (pg/mL) | -363.926            | -170.743                   |
| 95% CI of pooled estimate (pg/mL)    | -542.007–185.845    | -256.912–84.574            |
| Between-studies variance \(T^2\)     | 54859.742           | 9755.39                    |
| Degree of heterogeneity \(I^2\)      | 97.8%               | 83.7%                      |
| 95% CI of \(I^2\)                  | 96.8%–98.5%         | 69.7%–91.7%                |
| Q statistics                       | 271.051             | 36.780                     |
| DF                                | 6                   | 6                          |
| \(P\) value                       | 0.000               | 0.000                      |
size in all the studies, and the estimated between-studies variance (between studies variance $I^2 = 54859.742$ for AD versus controls and $9755.039$ for DOC versus controls) was very much high and between-studies variance contributes the most in total heterogeneity (degree of heterogeneity $I^2 = 97.8$, CI: $96.8\%–98.5\%$ in AD and degree of heterogeneity $I^2 = 83.7$, CI: $67.9\%–91.7\%$ in DOC). The heterogeneity in effect size of these 07 published studies could be due to different population, small sample size and lack of standardization of assay methods [26]. Though in AD group all 7 studies showed significant reductions in CSF $\text{A}^{\beta}_{42}$ levels as compared to control participants, the result of the present study was unequivocal and the range of effect size of 7 studies was quite large ($-652$ to $-41.2$ pg/mL). However, the fall in DOC group was much lower than in AD when compared to the control with reduced range of effect size of 7 studies undertaken ($-33$ to $-356$ pg/mL). Due to wide range of effect size of these studies, 2 studies effect size was more [19, 21] and that of 5 studies was less than the pooled estimate [17, 20, 22, 23] whereas 1 study [13] was on the vertical line passing through the pooled effect size. Similarly, in DOC group, 2 studies effect size was more [19, 21] and that of 5 studies was less than the pooled estimate [13, 17, 22, 23] whereas 1 study [20] was on the vertical line passing through the pooled effect size.

It has been a challenging task to explain the wide variation in results in the different laboratories due to technical problems in the studies included for studying the diagnostic utility of CSF biomarkers tau and $\text{A}^{\beta}_{42}$ in dementia. To overcome this, sensitivity analysis was conducted to explore the sensitivity level of each study. It was observed that there was not much difference in overall effect size on exclusion of one study at a time. Another major problem with this meta-analysis study is the presence of “publication bias.” Several lines of evidence show [27] that studies that report relatively high effect sizes are more likely to be published than

![Figure 2](image-url)
studies that report lower effect sizes. In the present study, Funnel plot analysis was done which showed the presence of publication bias for both parameters in AD as well as DOC group supporting the hypothesis that any bias in the literature is likely to be reflected in the meta-analysis study as only statistically significant results are more likely to be published.

### 4.3. Limitation

During the present meta-analysis, a number of factors was observed which could be causing the limitation in developing hypothesis/coming to conclusion regarding using CSF tau and Aβ42 as diagnostic markers in differential diagnosis of AD from dementias due to other causes. One limitation encountered during the present study was simply the amount of effort and expertise meta-analysis takes. Also, lack of complete information in some studies and high level of heterogeneity in individual study effect size for both parameters (tau & Aβ42) added to the limitation. Variation in CSF tau and Aβ42 levels in AD, DOC, and controls may be due to the variability introduced during sample collection and storage like using glass or polystyrene tubes18 than polypropylene tubes, prolonged storage of CSF samples in frozen state or repeated freezing and thawing of CSF, errors introduced due to inadequate study plan like small sample size, less number of controls taken in the study as compared to cases, or case and controls being not age and sex matched [24].

### 5. Conclusion

Based on our findings of the present meta-analysis, it can be concluded that the combination of high tau and low Aβ42 is highly specific for AD and might be useful in screening out the suspected cases of AD, from other types of dementia. However, due to the limited number of studies having large number of sample size with age- and sex-matched samples in control and cases available, the use of these biomarkers has to be verified in further prospective studies.

### References

[1] R. Agarwal, S. S. Kushwaha, and M. Gupta, “Role of biomarkers in diagnosis of Alzheimer’s disease,” *Indian Journal of Medical Biochemistry*, vol. 10, no. 1, pp. 25–30, 2006.

[2] F. Hulstaert, K. Blennow, A. Ivanou et al., “Improved discrimination of AD patients using β-amyloid(1–42) and tau levels in CSF,” *Neurology*, vol. 52, no. 8, pp. 1555–1562, 1999.

[3] “Consensus report of the working group on: Biological Markers of Alzheimer’s Disease. The Ronald & Nacy Reagan Research Institute of the Alzheimer’s association & the National Institute on Aging Working Group,” *Neurobiology of Aging*, vol. 19, no. 2, pp. 109–116, 1998.

[4] R. A. Frank, D. Galasko, H. Hampel et al., “Biological markers for therapeutic trials in Alzheimer’s disease: proceedings of the biological markers working group; NIA initiative on neuroimaging in Alzheimer’s disease,” *Neurobiology of Aging*, vol. 24, no. 4, pp. 521–536, 2003.

[5] T. Tapiola, M. Overmyr, M. Lehtovirta et al., “The level of cerebrospinal fluid tau correlates with neurofibrillary tangles in Alzheimer’s disease,” *NeuroReport*, vol. 8, no. 18, pp. 3961–3963, 1997.

[6] C. Mulder, P. Scheltens, J. J. Visser, G. I. van Kamp, and R. B. H. Schutgens, “Genetic and biochemical markers for Alzheimer’s disease: recent developments,” *Annals of Clinical Biochemistry*, vol. 37, no. 5, pp. 593–607, 2000.

[7] N. Andreasen, L. Minthon, P. Davidson et al., “Evaluation of CSF-tau and CSF-Aβ42 as diagnostic markers for Alzheimer disease in clinical practice,” *Archives of Neurology*, vol. 58, no. 3, pp. 373–379, 2001.

[8] N. Andreasen, E. Vanmechelen, H. Vanderstichele, P. Davidson, and K. Blennow, “Cerebrospinal fluid levels of total-tau, phospho-tau and Aβ42 predicts development of Alzheimer’s disease in patients with mild cognitive impairment,” *Acta Neurologica Scandinavica, Supplement*, vol. 107, no. 179, pp. 47–51, 2003.

[9] K. Blennow, A. Wallin, H. Agren, C. Spenger, J. Siegfried, and E. Vanmechelen, “Tau protein in cerebrospinal fluid: a biochemical marker for axonal degeneration in Alzheimer disease?” *Molecular and Chemical Neurpathology*, vol. 26, no. 3, pp. 231–245, 1995.

[10] N. Andreasen, C. Hesse, P. Davidsson et al., “Cerebrospinal fluid β-amyloid(1–42) in Alzheimer disease: differences between early- and late-onset Alzheimer disease and stability during the course of disease,” *Archives of Neurology*, vol. 56, no. 6, pp. 673–680, 1999.

[11] M. Otto, J. Wiltfang, H. Tumani et al., “Elevated levels of tau-protein in cerebrospinal fluid of patients with Creutzfeldt-Jakob disease,” *Neuroscience Letters*, vol. 225, no. 3, pp. 210–212, 1997.

[12] M. M. Verbeek, D. de Jong, and H. P. H. Kreimer, “Brain-specific proteins in cerebrospinal fluid for the diagnosis of neurodegenerative diseases,” *Annals of Clinical Biochemistry*, vol. 40, no. 1, pp. 25–40, 2003.

[13] N. Andreasen, L. Minthon, P. Davidsson et al., “Evaluation of CSF-tau and CSF-Aβ42 as diagnostic markers for Alzheimer disease in clinical practice,” *Archives of Neurology*, vol. 58, no. 3, pp. 373–379, 2001.

[14] T. B. Huedo-Medina, J. Sánchez-Meca, F. Marín-Martínez, and J. Botella, “Assessing heterogeneity in meta-analysis: Q statistic or I² Index?” *Psychological Methods*, vol. 11, no. 2, pp. 193–206, 2006.

[15] H. Arai, M. Terajima, M. Miura et al., “Tau in cerebrospinal fluid: a potential diagnostic marker in Alzheimer’s disease,” *Annals of Neurology*, vol. 38, no. 4, pp. 649–652, 1995.

[16] T. Tapiola, M. Lehtovirta, J. Ramberg et al., “CSF tau is related to apolipoprotein E genotype in early Alzheimer’s disease,” *Neurology*, vol. 50, no. 1, pp. 169–174, 1998.

[17] D. Galasko, C. M. Clark, L. Chang et al., “High cerebrospinal fluid tau and low amyloid β42 levels in the clinical diagnosis of Alzheimer disease and relation to apolipoprotein E genotype,” *Archives of Neurology*, vol. 55, no. 7, pp. 937–945, 1998.

[18] N. Andreasen, E. Vanmechelen, A. Van de Voorde et al., “Cerebrospinal fluid tau protein as a biochemical marker for axonal degeneration in Alzheimer disease?” *Archives of Neurology*, vol. 64, no. 3, pp. 298–305, 1998.

[19] M. Kanai, E. Matsubara, K. Isoe et al., “Longitudinal study of cerebrospinal fluid levels of tau, Aβ1-40, and Aβ1-42(43) in Alzheimer’s disease: a study in Japan,” *Annals of Neurology*, vol. 44, no. 1, pp. 17–26, 1998.

[20] M. Sjögren, P. Davidsson, A. Wallin et al., “Decreased CSF-β-amyloid 42 in Alzheimer’s disease and amyotrophic lateral sclerosis may reflect mismetabolism of β-amyloid induced
by disparate mechanisms,” *Dementia and Geriatric Cognitive Disorders*, vol. 13, no. 2, pp. 112–118, 2002.

[21] M. Grossman, J. Farmer, S. Leight et al., “Cerebrospinal fluid profile in frontotemporal dementia and Alzheimer’s disease,” *Annals of Neurology*, vol. 57, no. 5, pp. 721–729, 2005.

[22] D. de Jong, R. W. M. M. Jansen, B. P. H. Kremer, and M. M. Verbeek, “Cerebrospinal fluid amyloid β42/phosphorylated tau ratio discriminates between Alzheimer’s disease and vascular dementia,” *Journals of Gerontology: Series A*, vol. 61, no. 7, pp. 755–758, 2006.

[23] M. Bibl, H. Esselmann, B. Mollenhauer et al., “Blood-based neurochemical diagnosis of vascular dementia: a pilot study,” *Journal of Neurochemistry*, vol. 103, no. 2, pp. 467–474, 2007.

[24] S. Ravaglia, P. Bini, E. Sinforiani et al., “Cerebrospinal fluid levels of tau phosphorylated at threonine 181 in patients with Alzheimer’s disease and vascular dementia,” *Neurological Sciences*, vol. 29, no. 6, pp. 417–423, 2008.

[25] E. Kapaki, G. P. Paraskevas, I. Zalonis, and C. Zournas, “CSF tau protein and β-amyloid (1–42) in Alzheimer’s disease diagnosis: discrimination from normal ageing and other dementias in the Greek population,” *European Journal of Neurology*, vol. 10, no. 2, pp. 119–128, 2003.

[26] A. Maddalena, A. Papassotiropoulos, B. Müller-Tillmanns et al., “Biochemical diagnosis of Alzheimer disease by measuring the cerebrospinal fluid ratio of phosphorylated tau protein to β-amyloid peptide1–42,” *Archives of Neurology*, vol. 60, no. 9, pp. 1202–1206, 2003.

[27] M. Borenstein, L. V. Higgins, and H. R. Rothstein, *Introduction to Meta-Analysis*, John Wiley & Sons, New York, NY, USA, 2009.