Extrapolating evidence for molecularly targeted therapies from common to rare cancers: a scoping review of methodological guidance

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ABSTRACT

Objectives Cancer is increasingly classified according to biomarkers that drive tumour growth and therapies developed to target them. In rare biomarker-defined cancers, randomised controlled trials to adequately assess targeted therapies may be infeasible. Extrapolating existing evidence of targeted therapy from common cancers to rare cancers sharing the same biomarker may reduce evidence requirements for regulatory approval in rare cancers. It is unclear whether guidelines exist for extrapolation. We sought to identify methodological guidance for extrapolating evidence from targeted therapies used for common cancers to rare biomarker-defined cancers.

Design Scoping review.

Data sources Websites of health technology assessment agencies, regulatory bodies, research groups, scientific societies and industry. EBM Reviews—Cochrane Methodology Register and Health Technology Assessment, Embase and MEDLINE databases (1946 to 11 May 2022).

Eligibility criteria Papers proposing a framework or recommendations for extrapolating evidence for rare cancers, small populations and biomarker-defined cancers.

Data extraction and synthesis We extracted framework details where available and guidance for components of extrapolation. We used these components to structure and summarise recommendations.

Results We identified 23 papers. One paper provided an extrapolation framework but was not cancer specific. Extrapolation recommendations addressed six distinct components: strategies for grouping cancers as the same biomarker-defined disease; analytical validation requirements of a biomarker test to use across cancer types; strategies to generate control data when a randomised concurrent control arm is infeasible; sources to inform biomarker clinical utility assessment in the absence of prospective clinical evidence; requirements for surrogate endpoints chosen for the rare cancer; and assessing and augmenting safety data in the rare cancer.

Conclusions In the absence of an established framework, our recommendations for components of extrapolation can be used to guide discussions about interpreting evidence to support extrapolation. The review can inform the development of an extrapolation framework for biomarker-targeted therapies in rare cancers.

STRENGTHS AND LIMITATIONS OF THIS STUDY

⇒ This review of methodological guidance on extrapolation of evidence was based on a contemporary and comprehensive search including websites of regulatory bodies, health technology assessment agencies, oncology societies and scientific databases.

⇒ Key recommendations of extrapolation components have been derived from multidisciplinary fields including governmental regulation, health technology, clinical trials, epidemiology and molecular pathology.

⇒ Although a wide variety of sources were included, the search was limited to publicly available guidance published in English.

⇒ Standard terminology for extrapolation of trial evidence in this setting is not well established which may limit the sensitivity of the search strategy to identify methodological guidance.

INTRODUCTION

Historically, cancer diagnosis has focused on managing patients according to the site of their primary cancer, such as breast cancer or colorectal cancer. Yet, as understanding of the molecular alterations driving cancer growth progresses, therapeutic developments focus on testing novel agents that target these alterations.1-3 This is termed a precision oncology approach. Using this approach, patients may be grouped together and managed based on shared biomarker profiles, as opposed to by the location of their primary cancer.

Initial trials of new molecularly targeted treatments are typically conducted in one tissue type. When these trials demonstrate a therapeutic benefit within a biomarker-defined subpopulation, the challenge is determining whether the biomarker is also ‘predictive’ of therapeutic benefit (identifies a subpopulation that responds differently to a particular treatment) in cancers of other tissue types that share the same biomarker. However, the role of a biomarker...
in carcinogenesis, cancer progression and prediction of treatment effect may differ between cancer tissue types.4-7 Patient prognosis regardless of treatment (‘natural history’) is also likely to differ between cancer tissue types even if they share the same biomarker status.8-9 Thus, randomised comparisons of a new therapy versus other available therapies in a specific cancer tissue-type cohort is the ideal evidence to minimise bias for estimation of the relative treatment effect on clinical outcomes and to assess cost-effectiveness.10 11

In cancer tissue types of high prevalence where biomarker prevalence is also high, referred to herein as ‘common cancers’, it is feasible to conduct randomised controlled trials (RCTs) in biomarker-defined subpopulations within each cancer tissue type. However, in cancer tissue types of low prevalence, particularly where biomarker prevalence is also low, RCTs within biomarker-defined subpopulations may not be feasible or provide timely results. A ‘rare cancer’ is formally defined as incidence of the disease less than 6–15 cases per 100 000 persons per year.12-14 In this paper, we use ‘rare cancer’ to mean cancer tissue types where the biomarker-defined subpopulation is sufficiently small that RCTs are deemed infeasible.

To illustrate, the V600E mutation of the v-Raf murine sarcoma viral oncogene homologue B (BRAF) gene is one such molecular alteration affecting tumour cell proliferation that is shared across several different primary cancers, from common to rarer cancer types.15 In patients with BRAF V600E mutated metastatic melanoma, treatment with vemurafenib, a BRAF inhibitor, demonstrated improved overall survival (OS) compared with standard of care chemotherapy, dacarbazine, in a phase III RCT establishing the importance of determining tumour BRAF mutation status when determining treatment choice.16 17 In this example, melanoma is the common cancer expressing the molecular alteration. Beyond melanoma, vemurafenib was also tested in a basket trial, where patients are treated with the same treatment for different cancer tissue types sharing a common biomarker profile. In this instance, different cancer types all harbouring a BRAF V600 mutation (NCT01524978). Tumour response was seen in non-melanoma cancers including non-small cell lung cancer (NSCLC) (objective response rate (ORR) 42%) and Erdheim-Chester disease (ECD) (ORR 43%).7 In melanoma, prevalence of BRAF V600E mutation is approximately 40%-60%.18 Mutation prevalence in ECD is comparable (~50%)19; however, ECD is a rare disease with approximately 800 reported cases in the literature and the exact incidence remains unknown.20 NSCLC is a common cancer but BRAF V600E mutation prevalence is low (1%-2%).21 To date, randomised evidence to confirm vemurafenib activity in ECD and NSCLC is unavailable.

In small populations such as in this scenario, extrapolation from similar populations where robust clinical trial evidence for therapy exists may help to infer treatment effect and lower thresholds of evidence may be accepted for regulatory or reimbursement approval.22 Extrapolation of treatment evidence from adult to paediatric populations has been used in this way23 and guidelines exist.24 25 Extrapolation from common to rare cancers sharing the same predictive biomarker has also been documented without reference to established guidelines.26 However, determining the predictive value of a biomarker across different cancer types requires careful consideration of the appropriateness of extending the existing trial evidence beyond specific cancer tissue-type cohorts.

The aim of this scoping review is to inform the development of guidance on assessing the effectiveness of molecular targeted treatments across different cancer types defined by the same biomarker—and concomitantly the value of the biomarker to predict treatment benefit for a rare cancer, where randomised trial evidence is only available for a common cancer type. Specifically, we sought to identify guidelines outlining approaches for extrapolation of evidence for targeted therapies from common to rare cancers sharing the same biomarker. The findings were used to develop a framework for extrapolation to assist key stakeholders, such as regulatory and reimbursement bodies, researchers and clinicians. This work will assist interpretation of existing data and make standardised, transparent decisions on whether extrapolation from common to rare cancers is appropriate.

**METHODS**

**Objectives**

1. What methodological guidance frameworks exist for extrapolating evidence of relative treatment effect of targeted therapies versus standard of care treatment from common to rare cancers that share the same biomarker profile?
2. Specifically, do they address:
   a. When it is appropriate to extrapolate evidence and to what extent?
   b. What assumptions are made to extrapolate and what evidence is required to support these assumptions?
   c. What methods can be used to appropriately extrapolate evidence?

**Eligibility criteria**

We included papers proposing a framework outlining criteria or methods for extrapolating evidence for targeted therapies from common to rare cancers sharing the same biomarker profile. As literature specific to this context was limited, we extended our review to include guidance for using extrapolation in trials assessing targeted therapies matched to a biomarker regardless of cancer type (‘pan-cancer’ trials), rare cancers or small populations. No limit was set for year of publication. We excluded papers that only discussed challenges or limitations, rather than providing an explicit framework or recommendations. Language was limited to English only.
Information sources and search strategy

We searched websites of health technology assessment (HTA) agencies, regulatory bodies, research groups, scientific societies and industry. We also searched MEDLINE, Embase, EBM Reviews—Cochrane Methodology Register and Health Technology Assessment databases (1946 to 11 May 2022). The full search strategy is available in the online supplemental appendices 1 and 2. Additional papers from reference lists of included papers and from discussions with content experts were also screened. Two reviewers performed the website and database searches and screened papers for eligibility (DC, SC). One reviewer extracted and synthesised data from the included papers (DC).

Data extraction and synthesis

We extracted details from each paper of relevant frameworks and recommendations for evidence extrapolation. If a comprehensive framework does not exist, we then sought to identify, categorise and integrate recommendations relevant for extrapolation in this context into a single summary document. To organise this information, we defined six key evidence components for extrapolation based on evidence-based medicine principles for the evaluation of new tests and treatments27 28 as follows: disease definition, analytical validity of the biomarker test, control (‘standard of care’) data, biomarker actionability, endpoints and safety. In the absence of RCT evidence for the rare cancer, each component represents a critical assumption for extrapolation. We grouped individual recommendations for evidence to support these assumptions from each paper under the relevant component. We generated tables and figures to summarise the number of components addressed by each paper, recommendations for each component and potential evidence gaps that may limit extrapolation. We used the Preferred Reporting Items for Systematic Reviews and Meta-Analyses extension for Scoping Reviews checklist for reporting.

Patient and public involvement

As this study was a scoping review of published methodological guidance, it was not appropriate to involve patients or the public in the design, conduct, reporting or dissemination plans of our research.

RESULTS

The database search yielded 1345 citations. A further 31 papers were retrieved from other sources. Screening of titles and abstracts identified 109 papers for full-text review of which 23 papers were included for evidence synthesis22 29–50 (figure 1, online supplemental appendix 3).

Characteristics of included papers

The majority of included papers were published by academic research groups (n=11, 48%), followed by regulatory bodies (n=6, 26%), HTA organisations (n=3, 13%) and scientific societies (n=3, 13%) (table 1). Publication year ranged from 2006 to 2021. The main focus of these papers were small population trials (n=10), pan-cancer trials of biomarker targeted therapies (n=10), biomarker actionability (n=10) and analytical validity of the biomarker test (n=7) (online supplemental table...
| Author group          | Year | Extrapolation framework (n) | Components addressed (n) | Disease definition | Test analytical validity | Control data | Biomarker actionability | Endpoints | Safety |
|-----------------------|------|-----------------------------|--------------------------|--------------------|------------------------|--------------|-----------------------|-----------|--------|
| **Research group**    |      |                             |                          |                    |                        |              |                       |           |        |
| Seligson et al        | 2021 | 2                           | 1                        | 1                  |                        |              |                       |           |        |
| Dittrich              | 2020 | 3                           | 1                        | 1                  |                        |              |                       |           |        |
| Jørgensen             | 2020 | 3                           | 1                        | 1                  |                        |              |                       |           |        |
| Moscow et al          | 2018 | 4                           | 1                        | 1                  |                        |              |                       |           |        |
| Chakravarty et al     | 2017 | 1                           |                          | 1                  |                        |              |                       |           |        |
| Beckman et al         | 2016 | 3                           | 1                        | 1                  |                        |              |                       |           |        |
| Meric-Bernstam et al  | 2015 | 1                           |                          | 1                  |                        |              |                       |           |        |
| Van Allen et al       | 2014 | 1                           |                          | 1                  |                        |              |                       |           |        |
| Andre et al           | 2014 | 1                           |                          | 1                  |                        |              |                       |           |        |
| Vidwans et al         | 2014 | 1                           |                          | 1                  |                        |              |                       |           |        |
| Sharma and Schilsky   | 2011 | 2                           | 1                        | 1                  |                        |              |                       |           |        |
| **Regulatory body**   |      |                             |                          |                    |                        |              |                       |           |        |
| FDA: Human gene therapy for rare diseases | 2020 | 3                           | 1                        | 1                  |                        |              |                       |           |        |
| FDA: Rare diseases: common issues in drug development | 2019 | 4                           | 1                        | 1                  |                        |              |                       |           |        |
| FDA: Developing targeted therapies in low-frequency molecular subsets of a disease | 2018 | 4                           | 1                        | 1                  |                        |              |                       |           |        |
| Schuck et al (FDA)    | 2018 | 3                           | 1                        | 1                  |                        |              |                       |           |        |
| EMA: Concept paper on extrapolation of efficacy and safety in medicine development | 2013 | 1                           | 3                        | 1                  |                        |              |                       |           |        |
| EMA: Guideline on clinical trials in small populations | 2006 | 3                           | 1                        | 1                  |                        |              |                       |           |        |
| **HTA**               |      |                             |                          |                    |                        |              |                       |           |        |
| Lengliné et al (TC-HAS) | 2021 | 4                           | 1                        | 1                  |                        |              |                       |           |        |
| Morona et al (MSAC)   | 2020 | 4                           | 1                        | 1                  |                        |              |                       |           |        |
| Merlin et al (Adelaide HTA) | 2013 | 3                           | 1                        | 1                  |                        |              |                       |           |        |
| **Scientific society**|      |                             |                          |                    |                        |              |                       |           |        |
| Mateo et al (ESMO)    | 2018 | 3                           | 1                        | 1                  |                        |              |                       |           |        |
Only one paper outlined an explicit extrapolation framework but was not specific to targeted therapies for cancer. Biomarker actionability was the most commonly addressed component (n=14) (Tables 1 and 2, figure 2).

**Extrapolation framework**

Our search did not identify any published guidelines or framework explicitly outlining a comprehensive approach for extrapolating evidence for targeted therapies from common to rare cancers sharing the same biomarker. However, we identified papers reporting individual recommendations in relevant fields and incorporated these into one or more components to develop an extrapolation framework for assessing biomarker-targeted therapies in rare cancers. One paper by the European Medicines Agency (EMA) outlined an extrapolation framework for medicines in general and proposed a systematic approach for when, to what extent and how extrapolation can be applied for new medicines.22 Although this framework is not cancer specific, the same extrapolation principles can be applied to targeted treatments for biomarker-defined cancers. Elements of the EMA framework as well as recommendations made by the other papers are organised under the relevant component and summarised in Table 2 and discussed below.

**Defining the disease**

A principal assumption for extrapolation is that there is strong biological rationale that both cancers can be grouped as the same biomarker-defined disease, characterised by similar response to the targeted therapy being assessed. A quantitative assessment of the similarity of disease should be made and can be based on clinical, preclinical and mechanistic data sources. To define the disease, a companion diagnostic assay should be developed alongside the therapy that detects and reports all specific molecular variants, across cancer types, comprising the group expected to respond to the targeted therapy. For continuously measured biomarkers, the cut-offs to define the ‘biomarker-positive’ group should be determined for each cancer type.

**Analytical validity of the biomarker test**

Critical to defining the biomarker-defined disease is an analytically valid test that accurately, reproducibly and reliably identifies the biomarker. An assumption for extrapolation is that the biomarker test has been analytically validated against a reference or gold standard before the start of the clinical trial in both the common and rare cancers. A test with acceptable analytical validity in one cancer type may not be assumed to be valid in another cancer type. As biomarker prevalence may differ substantially between cancer types, an assessment of the positive predictive value (the proportion of biomarker ‘positive’ results correctly identifying the disease) and negative predictive value (the proportion of biomarker ‘negative’ results correctly identifying absence of disease) of the
### Table 2  Recommendations for extrapolation components

| Components | Author | Author group | Year |
|------------|--------|--------------|------|
| **Extrapolation framework** | | | |
| Presents a framework for a systematic approach to set out when, to what extent and how extrapolation can be applied to extend information from a source population to make inferences for another target population. Elements of the framework include (1) a systematic synthesis of available data informing quantitative hypotheses for the similarity of the conditions and predicted response to intervention in the source and target population, (2) a reduced set of additional evidence requirements in the target population and selection of the type of extrapolation strategy (no extrapolation, partial extrapolation or full extrapolation), (3) validation of the hypotheses by emerging clinical or preclinical data or revision of the appropriateness of extrapolation if emerging data are not supportive, (4) interpretation of the limited data in the target population in the context of extrapolation, and (5) methods to address uncertainty and mitigate risk. | EMA: Concept paper on extrapolation of efficacy and safety in medicine development\(^{22}\) | Regulatory body | 2013 |

| Disease definition | | | |
| Different types of evidence can support grouping strategies. Clinical studies are considered the strongest types of evidence. Preclinical studies and in silico (computational) evidence from a model system demonstrating similar drug effects within the proposed molecularly defined group are accepted as additional sources of evidence. Consistent treatment effects observed across more than one type of evidence increase the strength of evidence. | FDA: Developing targeted therapies in low-frequency molecular subsets of a disease\(^{43}\) | Regulatory body | 2018 |
| | | | |
| | Schuck et al (FDA)\(^{42}\) | Regulatory body | 2018 |
| | Morona et al (MSAC)\(^{45}\) | HTA | 2020 |
| | Offin et al (ASCO)\(^{41}\) | Scientific society | 2018 |
| | FDA: Human gene therapy for rare diseases\(^{46}\) | Regulatory body | 2020 |
| | FDA: Developing targeted therapies in low-frequency molecular subsets of a disease\(^{43}\) | Regulatory body | 2018 |
| | Schuck et al (FDA)\(^{42}\) | Regulatory body | 2018 |

| | EMA: Concept paper on extrapolation of efficacy and safety in medicine development\(^{22}\) | Regulatory body | 2013 |

| Assays used to define the disease should be designed to detect and report all specific variants comprising the group expected to respond to better understand the molecular spectrum of the disease rather than limiting the definition to ‘biomarker-positive’ or ‘biomarker-negative’. | | | |
| FDA: Human gene therapy for rare diseases\(^{46}\) | Regulatory body | 2020 |
| FDA: Developing targeted therapies in low-frequency molecular subsets of a disease\(^{43}\) | Regulatory body | 2018 |
| Schuck et al (FDA)\(^{42}\) | Regulatory body | 2018 |
| Merlin et al (Adelaide HTA)\(^{31}\) | HTA | 2013 |

Continued
### Table 2  Continued

| Components                                                                                     | Author                                      | Author group | Year |
|-----------------------------------------------------------------------------------------------|---------------------------------------------|--------------|------|
| Biomarker type may affect the generalisability of disease definition across cancer types. For binary or categorical biomarkers such as presence or absence of a particular mutation, the definitions of ‘biomarker-positive’ and ‘biomarker-negative’ may generalise across cancers. These biomarkers may also be able to be detected from liquid biopsy. However, for continuously measured biomarkers, such as protein expression, the cut-offs to define the ‘biomarker-positive’ group should be determined for each cancer type. This cut-off should be justified and could be provisionally informed by randomised exploratory studies stratified by the predictive biomarker. | Lengliné et al (TC-HAS)\(^{47}\)            | HTA          | 2021 |
|                                                                                               | Beckman et al\(^{36}\)                      | Research group | 2016 |
| A companion diagnostic test used to define the disease should be developed alongside clinical drug development to avoid the need for subsequent bridging studies. This test should be a central analytically validated assay to have an optimal definition of the biomarker-defined population. | Jørgensen\(^{49}\)                          | Research group | 2020 |

### Analytical validity of the biomarker test

The companion diagnostic test used should be analytically validated before the start of the clinical trial. Performance characteristics of the test (sensitivity, specificity, positive and negative predictive values) should be documented.

- Jørgensen\(^{49}\) Research group 2020
- Moscow et al\(^{40}\) Research group 2018
- Beckman et al\(^{36}\) Research group 2016
- Sharma and Schilsky\(^{30}\) Research group 2011
- FDA: Rare diseases: common issues in drug development\(^{44}\) Regulatory body 2019
- FDA: Developing targeted therapies in low-frequency molecular subsets of a disease\(^{43}\) Regulatory body 2018
- Schuck et al (FDA)\(^{42}\) Regulatory body 2018
- Lengliné et al (TC-HAS)\(^{47}\) HTA 2021
- Morona et al (MSAC)\(^{45}\) HTA 2020
- Merlin et al (Adelaide HTA)\(^{31}\) HTA 2013
- Mateo et al (ESMO)\(^{39}\) Scientific society 2018

For a test intended to be used across multiple cancer types, a central analytically validated assay should be used in order to avoid interassay variability.

- Jørgensen\(^{49}\) Research group 2020

For a test intended to be used across multiple cancer types, assessment of analytical accuracy of the assay in each cancer type against a reference or gold standard is required.

- Beckman et al\(^{36}\) Research group 2016
- Moscow et al\(^{40}\) Research group 2018
- Morona et al (MSAC)\(^{45}\) HTA 2020

Continued
Table 2  Continued

| Components                                                                 | Author                  | Author group | Year  |
|---------------------------------------------------------------------------|-------------------------|--------------|-------|
| A test with acceptable analytical validity in one cancer type may not be assumed to be valid in another cancer type. As an example, for biomarkers defined by protein expression, differences in tumour biology and tissue processing variables may alter the sensitivity of an assay. If a gold standard does not exist, concordance between the test used in the additional cancer type and the evidentiary standard (the test used in the pivotal trial that establishes the effectiveness of targeted treatment in the common cancer) should be demonstrated. Where the analytical sensitivity and specificity of the biomarker assay are the same between different tissue types, differences in biomarker prevalence alter the PPV and NPV (the proportion of biomarker ‘positive’ results correctly identifying the disease) and NPV (the proportion of biomarker ‘negative’ results correctly identifying absence of disease). As biomarker prevalence may differ substantially between cancer types, an assessment of the PPV and NPV of the test across the relevant prevalence range for the cancers being targeted is recommended. | Morona et al (MSAC)45  | HTA          | 2020  |
| Tumour molecular profiling technologies are rapidly advancing. Thus, both tests using different technology (eg, next generation sequencing as opposed to PCR or Sanger sequencing to detect a mutation) or different assay (eg, two different commercial PCR assays to detect BRAF V600E mutation) to be used require validation in the new cancer type. | Merlin et al (Adelaide HTA)31 | HTA          | 2013  |
| Standardisation of laboratory procedures for sample collection, storage and processing between laboratories currently organised according to cancer tissue type rather than test or technology type is recommended to improve interlaboratory concordance and test reproducibility. | Moscow et al49          | Research group | 2018  |
| Control (standard of care) data  If randomisation to a concurrent control arm is not feasible in the rare cancer group, alternative sources of control data will be required. Natural history data for the rare cancer can be used as a historical control. | Beckman et al96         | Research group | 2016  |
| Dittrich50                                                                 | Research group          | 2020         |
| Mateo et al39                                                               | Research group          | 2018         |
| Beckman et al96                                                             | Research group          | 2016         |
| Sharma and Schilsky33                                                      | Research group          | 2011         |
| FDA: Human gene therapy for rare diseases46                                  | Regulatory body         | 2020         |
| FDA: Rare diseases: common issues in drug development44                     | Regulatory body         | 2019         |
| EMA: Guideline on clinical trials in small populations29                    | Regulatory body         | 2006         |
| Lengliné et al (TC-HAS)47                                                  | HTA                     | 2021         |
| Morona et al (MSAC)45                                                      | HTA                     | 2020         |
| Continued                                                                 |                         |              |
### Table 2  Continued

| Components                                                                 | Author                     | Author group   | Year  |
|---------------------------------------------------------------------------|----------------------------|----------------|-------|
| Natural history data should be from the biomarker-defined population to   | Dittrich50                 | Research group | 2020  |
| account for any difference in prognosis between biomarker-positive and    |                            |                |       |
| biomarker-negative subgroups.                                             | Sharma and Schilsky30      | Research group | 2011  |
| Lengliné et al (TC-HAS)47                                                 | HTA                        |                | 2021  |
| Morona et al (MSAC)45                                                     | HTA                        |                | 2020  |
| Clinically and genetically annotated retrospective cohorts of patients    | Lengliné et al (TC-HAS)47   | HTA            | 2021  |
| with a rare cancer or cancer with rare genetic alterations should be      |                            |                |       |
| established. All data from basket trials should be recorded in registries.|                            |                |       |
| These data can be reused as a prespecified external control.              |                            |                |       |
| RWE (observational evidence derived from clinical use outside of an RCT)  | Mateo et al (ESMO)39       | Scientific society | 2018  |
| from genomic databases annotated with clinical outcome data can be used   |                            |                |       |
| to assemble historical control benchmarks.                               |                            |                |       |
| Concurrent prospective registry controls can mitigate bias from outcome   | Beckman et al36            | Research group | 2016  |
| differences as a result of stage migration and improved supportive       |                            |                |       |
| treatment or available therapies over time. Shared controls can be used   |                            |                |       |
| for multiple rare cancer types to reduce sample size requirements.        |                            |                |       |
| This will be limited to cancer types with the same control therapy.      |                            |                |       |
| Patients as their own controls where time to disease progression on       | FDA: Human gene therapy for | Regulatory body | 2020  |
| targeted therapy may be compared with that from their previous treatment  |                            |                |       |
| can be used.                                                              |                            |                |       |
| Effect size of treatment in the common cancer (reference case) can be     | Morona et al (MSAC)45      | HTA            | 2020  |
| benchmarked against prognostic data from biomarker-positive rare cancer   |                            |                |       |
| historical controls.                                                      |                            |                |       |
| The threshold to be met for the assessment of efficacy in the rare cancer | Dittrich50                 | Research group | 2020  |
| depends on the historical control rare of the biomarker-positive cancer   |                            |                |       |
| type.                                                                     |                            |                |       |

**Biomarker actionability**

Biomarker actionability in one cancer type may or may not extrapolate to another cancer type as molecular pathways driving cancer progression and/or resistance mechanisms can differ between cancer types despite sharing the same biomarker. Frameworks have been proposed that classify biomarkers or molecular alterations into tiers, according to actionability, and can inform this assessment in each cancer type.

- Chakravarty et al37  
- Meric-Bernstam et al35  
- Van Allen et al33  
- Vidwans et al34  
- Andre et al32  
- Mateo et al (ESMO)39  
- Li et al (AMP, ASCO, CAP)38  
- Seligson et al48

If prospective clinical evidence is unavailable in the rare cancer, other types of evidence are accepted to inform functional relevance and actionability of the biomarker and include:

- Preclinical evidence.

Continued
Table 2  Continued

| Components                                                                 | Author                                                                 | Author group | Year  |
|----------------------------------------------------------------------------|------------------------------------------------------------------------|--------------|-------|
| ► Biological plausibility and scientific rationale.                         | FDA: Developing targeted therapies in low-frequency molecular subsets of a disease<sup>43</sup> | Regulatory body | 2018  |
| ► EMA: Concept paper on extrapolation of efficacy and safety in medicine development<sup>42</sup> | Morona et al (MSAC)<sup>45</sup>                                      | HTA          | 2020  |
| ► Offin et al (ASCO)<sup>41</sup>                                           |                                                                       | Scientific society | 2018  |
| ► Systems biology.                                                          | Vidwans et al<sup>34</sup>                                              | Research group | 2014  |
| ► In silico evidence and modelling of pharmacokinetic and pharmacodynamic relationships. | Van Allen et al<sup>32</sup>                                            | Research group | 2014  |
| ► Schuck et al (FDA)<sup>42</sup>                                           | Morona et al (MSAC)                                                  | HTA          | 2020  |
| ‘Linked evidence approach’—linking evidence for the test, biomarker actionability and treatment outcomes from different sources, provided they are generated in an internally valid manner from similar patient populations, to build a chain of arguments as an alternative when RCT evidence is not feasible. | Merlin et al (Adelaide HTA)<sup>31</sup>                              | HTA          | 2013  |
| Data assessing the functional consequence of genomic alterations must be generated in the cancer type for which the therapy is being considered as the same alteration can have distinct epistatic interactions across different cancer types. | Dittrich<sup>50</sup>                                                | Research group | 2020  |
| Longitudinal, multiomic (genomic, RNA sequencing, methylomic, proteomic) assessment of cancers can be used to better predict treatment response and understand resistance mechanisms across cancer types and inform biomarker actionability in the rare cancer. | Seligson et al<sup>48</sup>                                           | Research group | 2021  |
| The type of molecular alteration may be important for whether actionability can be extrapolated to a different cancer type. Examples of therapies exhibiting activity across cancer types are either targeting the host immune system or kinase fusions. However, very few therapies targeting single nucleotide variants and amplifications have exhibited histology-agnostic activity. | Jørgensen<sup>49</sup>                                               | Research group | 2020  |
| Evidence of treatment effect in the biomarker-negative population is required to inform biomarker actionability. If this is limited in the rare cancer, if early studies addressed this question whereby patients for whom no signal of efficacy was subsequently excluded in a seamless oncology trial approach, these studies may reasonably be used as best available evidence in the biomarker-negative subgroup in the rare cancer. | Morona et al (MSAC)<sup>45</sup>                                      | HTA          | 2020  |
| Postapproval studies and RWE, including large non-randomised studies, should be used to confirm or refute regulatory decisions based on limited evidence supporting actionability in the rare cancer. | Seligson et al<sup>48</sup>                                           | Research group | 2021  |
| Moscow et al<sup>40</sup>                                                   | Schuck et al<sup>42</sup>                                              | Regulatory body | 2018  |
### Clinical trial endpoints

Data on clinically relevant outcomes may not be available in rare cancers. In this scenario, surrogate endpoints may need to be used for extrapolation. Recommendations on the choice of surrogate endpoint(s) include:

- Surrogate endpoints can be informed by understanding the disease pathophysiology, natural history and biological/pharmacological rationale for the treatment effect.
  - FDA: Human gene therapy for rare diseases  
  - EMA: Concept paper on extrapolation of efficacy and safety in medicine development  
  - EMA: Guideline on clinical trials in small populations  
  
- Endpoints not susceptible to subjective interpretation may reduce assessment bias which is particularly problematic for single-arm studies.
  - FDA: Rare diseases: common issues in drug development  
  - EMA: Guideline on clinical trials in small populations  

- Where the surrogate endpoint is measured in the laboratory, analytical validity of the assay in both cancers and clinical validity (the ability to define a clinical state) in the common cancer should be demonstrated.
  - FDA: Rare diseases: common issues in drug development  
  - EMA: Guideline on clinical trials in small populations  

- The surrogate should be reasonably likely to predict clinical benefit.
  - Moscow et al  
  - FDA: Human gene therapy for rare diseases  
  - EMA: Guideline on clinical trials in small populations  

- At a minimum, correlation between changes in the surrogate and clinical endpoint should be demonstrated in the common cancer.
  - FDA: Human gene therapy for rare diseases  

- When the primary endpoint is the rate of tumour response or shrinkage, its assessment must be standardised, and the duration of response, which is more relevant than tumour response rate alone, must also be analysed. Strong correlation between the treatment effect on tumour response and overall survival should be anticipated.
  - Lengliné et al (TC-HAS)  

| Components | Author | Author group | Year |
|------------|--------|--------------|------|
| Surrogate endpoints can be informed by understanding the disease pathophysiology, natural history and biological/pharmacological rationale for the treatment effect. | FDA: Human gene therapy for rare diseases | Regulatory body | 2020 |
| | EMA: Concept paper on extrapolation of efficacy and safety in medicine development | Regulatory body | 2013 |
| | EMA: Guideline on clinical trials in small populations | Regulatory body | 2006 |
| Endpoints not susceptible to subjective interpretation may reduce assessment bias which is particularly problematic for single-arm studies. | FDA: Rare diseases: common issues in drug development | Regulatory body | 2019 |
| | EMA: Guideline on clinical trials in small populations | Regulatory body | 2006 |
| Where the surrogate endpoint is measured in the laboratory, analytical validity of the assay in both cancers and clinical validity (the ability to define a clinical state) in the common cancer should be demonstrated. | FDA: Rare diseases: common issues in drug development | Regulatory body | 2019 |
| | EMA: Guideline on clinical trials in small populations | Regulatory body | 2006 |
| The surrogate should be reasonably likely to predict clinical benefit. | Moscow et al | Research group | 2018 |
| | FDA: Human gene therapy for rare diseases | Regulatory body | 2020 |
| | EMA: Guideline on clinical trials in small populations | Regulatory body | 2006 |
| At a minimum, correlation between changes in the surrogate and clinical endpoint should be demonstrated in the common cancer. | FDA: Human gene therapy for rare diseases | Regulatory body | 2020 |
| | Lengliné et al (TC-HAS) | HTA | 2021 |

When a targeted therapy has demonstrated efficacy and safety based on clinical endpoints in a population composed of various molecular subtypes all thought to result in the same clinical disease, treatment effect can be extrapolated to additional putatively similar molecular subtypes based on pharmacodynamic or surrogate endpoints.

- A single trial that pools different cancer-type cohorts together is only acceptable if tumour response rates are homogeneous. If tumour response is heterogeneous, separate trials for each cancer type are required.
  - Seligson et al  
  - Dittrich  

| Components | Author | Author group | Year |
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Continued
| Components | Author | Author group | Year |
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| Safety: When safety information is limited in the rare cancer, the following sources can augment safety data: (1) natural history studies, (2) auxiliary safety cohorts, (3) expanded access programmes and postmarketing studies in the rare cancer group, and (4) studies of the targeted therapy in other indications or studies of similar drugs. | Dittrich | Research group | 2020 |
| A quantitative assessment of existing data from clinical, preclinical and in vitro data: EMA: Concept paper on extrapolation of efficacy and safety in medicine development | EMA: Guideline on clinical trials in small populations | Regulatory body | 2006 |
| In the rare cancer, postmarketing safety studies or risk mitigation measures beyond labelling and routine pharmacovigilance are recommended. | FDA: Rare diseases: common issues in drug development | Regulatory body | 2019 |
| Risks associated with misclassification of the biomarker should also be considered when assessing safety. | Morona et al. (MSAC) | HTA | 2020 |
| AMP, Association for Molecular Pathology; ASCO, American Society of Clinical Oncology; CAP, College of American Pathologists; EMA, European Medicines Agency; ESMO, European Society for Medical Oncology; FDA, Food and Drug Administration; HTA, Health Technology Assessment organisations; MSAC, Medical Services Advisory Committee; NPV, negative predictive value; PPV, positive predictive value; RCT, randomised controlled trial; RWE, real-world evidence; TC-HAS, Transparency Committee of the French National Authority for Health (Haute Autorité de Santé). | | | |
test across the relevant prevalence range for the cancers being targeted is recommended.

Control (standard of care) data

An assumption made for extrapolation is that reliable control data are available for the biomarker-defined rare cancer. To compare the effect of a targeted therapy with the effect of non-targeted therapies and differentiate treatment effect from the prognosis (natural history) of the biomarker-positive cancer subgroup, outcome data of biomarker-positive patients treated with non-targeted therapy (control arm) are essential. Concurrent standard of care data from a well-conducted RCT are the ideal source of control data to minimise risk of bias. Such data are usually available for common but not rare cancers. If randomisation to a concurrent control arm is not feasible in the rare cancer group, alternative sources of control data will be required. Natural history data from the biomarker-defined subpopulation of the rare cancer can be used as a historical control. Data sources include clinically and genetically annotated cohorts of patients with rare cancers from registries, basket trials and real-world evidence (RWE). Concurrent prospective registry controls have the advantage of mitigating bias from outcome differences as a result of stage migration and improved supportive treatment or available therapies over time.

Biomarker actionability

The actionability of a biomarker is measured by the ability of decisions based on the biomarker to improve health outcomes. For a predictive biomarker, this depends on how well it predicts treatment effect (clinical validity of a biomarker). Assessment of both biomarker-positive and biomarker-negative groups receiving targeted therapy versus non-targeted therapy is required to differentiate between the prognostic value of the biomarker to predict survival (assessed by comparing clinical outcomes for the biomarker-positive vs negative groups receiving non-targeted therapy) and the predictive value of the biomarker to predict treatment effect (assessed by comparing the relative treatment effect of targeted vs non-targeted therapy for the biomarker-positive vs negative groups). Prospective clinical trial evidence is essential to establish biomarker actionability in the common cancer and if trial evidence is also available in the rare cancer, extrapolation of evidence is not required. However, if this is not available in the rare cancer and extrapolation is used, a key assumption is that the biomarker is similarly actionable in the rare cancer.

Even if biomarker actionability is established in one cancer type, actionability may or may not extrapolate to another cancer type as molecular pathways driving cancer progression and/or resistance mechanisms can differ between cancer types despite having the same biomarker. Frameworks classifying biomarkers or molecular alterations into tiers, according to actionability, can inform this assessment in each cancer type. These frameworks are based on the strength of evidence supporting biomarker actionability. Sources supporting tier classification in decreasing order of level of evidence include clinical practice guidelines, prospective clinical evidence, retrospective clinical evidence, observational evidence, RWE from large-scale cancer mutation databases and registry data. High tier associations in one cancer type may be downgraded in another cancer type if the level of evidence in the latter is lower. If prospective clinical evidence...
evidence is unavailable in the rare cancer, lower levels of evidence generated in the rare cancer tissue type are accepted to inform functional relevance and actionability of the biomarker, including preclinical evidence, biological rationale, systems biology and in silico evidence. Where regulatory decisions have been based on limited evidence supporting actionability in the rare cancer, postapproval studies should be used to confirm treatment benefit.

Clinical trial endpoints
Endpoints used for extrapolation are assumed to be applicable to both common and rare cancers. Progression-free survival (PFS) and OS are often used in common cancer trials to measure clinically relevant treatment outcomes, but PFS and OS data may not be available in rare cancers. In this scenario, other surrogate endpoints that are more sensitive or able to be measured earlier may be used to extrapolate treatment effect from the common cancer to the rare cancer. Extrapolation is accepted when treatment effects in both cancers, based on surrogate endpoints, are homogeneous. The surrogate endpoint chosen should reasonably be likely to predict clinical benefit. When the surrogate used is the rate of tumour shrinkage, assessment should be standardised, and the duration of response, which is more clinically relevant than tumour shrinkage rate alone, should also be analysed. Treatment results based on surrogate endpoints need confirmation of clinical benefit in postapproval studies.

Safety
Risk-benefit assessment of therapies requires data on both safety and effectiveness. However, decisions for extrapolating evidence for either safety or effectiveness are usually made independently. Clinical studies acceptable for extrapolating treatment effectiveness may not be applicable for extrapolating safety due to differences in endpoints chosen and study power requirements for rare adverse events. An assumption made for extrapolating safety data is that the safety profile of targeted therapy is the same in both the common and rare cancers. However, the safety profile of the same therapy may differ when used in different cancer tissue types.

The safety profile of therapies is generally informed by randomised concurrently controlled studies and supplemented by phase IV and RWE capturing use in broader populations compared with the highly selected populations recruited to phase III RCTs. While safety information is available in the common cancer it may be limited in the rare cancer. In this scenario, sources to augment safety data include natural history studies, auxiliary safety cohorts, expanded access programme and postmarketing studies. In the rare cancer, postmarketing safety studies or risk mitigation measures beyond labelling and routine pharmacovigilance are recommended.

In addition to the risks associated with treatment adverse events, risks associated with misclassification from the biomarker test and subsequent incorrect treatment recommendations should also be considered when assessing safety.

DISCUSSION
This scoping review identified 23 guidance papers published by research groups, regulatory bodies, HTA organisations and scientific societies of which only one paper presented an explicit extrapolation framework for medicines. Principles from this framework may be applied to our setting but there was no framework for extrapolation to generate evidence specifically for biomarker-targeted therapies in rare cancers. Each of the other 22 papers made recommendations for one or more components of extrapolation.

With rapid advances in molecular profiling technologies, cancers are increasingly partitioned into smaller subgroups according to potentially actionable biomarkers.52 With dwindling sample sizes available for recruitment to trials, the traditional trial design paradigm is becoming unfeasible for novel therapies targeted to these biomarker-defined rare cancers. Informally, extrapolation of evidence is already being used for assessing targeted therapies in biomarker-defined rare cancers for regulatory approval.56 Our findings of similar recommendations for individual components of extrapolation from different groups indicate broad consensus. However, these recommendations are not yet integrated into a comprehensive framework for judging whether extrapolation should be used and how. Additionally, no paper outlined trial design strategies for generating new evidence in rare cancers in the context of using prior evidence from common cancers sharing the same biomarker. Overall, statistical recommendations for extrapolation were limited though four papers raised the potential for incorporating adaptive designs including Bayesian approaches using prior information from the common cancer.22 42 48 50

Strengths and limitations
There is an immediate and growing need to explore novel methods to assess evidence for biomarker-defined rare cancers when RCTs are unfeasible. This is the first scoping review aiming to identify if and to what extent guidance exists on extrapolation to address this need. A key finding is there is no extrapolation framework specific for rare biomarker-defined cancers. This review is timely to summarise in a single document recommendations of commonly accepted components of extrapolation relevant to biomarker-targeted therapies for rare cancers to guide discussions and stimulate debate about best methods for addressing each.

There are potential limitations to our work. First, given we were unable to identify any papers that fit our a priori research question, it became necessary to take a pragmatic approach and broaden the eligibility criteria to include papers that addressed one or more components of extrapolation that would be useful to inform such a framework. Second, we relied on published or publicly
available guidance and may have missed unpublished documents. Third, we limited our search to documents in English due to practical restraints and may have missed extrapolation guidance published in other languages. Fourth, standard terminology for extrapolation of trial evidence in this setting is not well established which may limit the sensitivity of the search strategy to identify methodological guidance.

Application and future directions

This summary document has systematically identified distinct components for extrapolation, namely disease definition, analytical validity of the biomarker test, control (‘standard of care’) data, biomarker actionability, clinical trial endpoints and safety. This framework is specific for our setting of extrapolating data from common to rare cancers sharing the same biomarker treated with targeted therapy, and is not designed for other cancer and non-cancer settings.

This work is valuable to guide discussions between key stakeholders in drug development, regulatory approval and reimbursement when extrapolated evidence from other cancers is being used to interpret evidence for targeted therapies in rare cancers. This work is also relevant for clinical trialists designing future studies in rare cancers where extrapolated data from common cancers are used to justify the trial design. It can also inform trialists designing studies in common cancers so that data generated would be useful and easily applicable for extrapolation in the future. The work is also relevant for clinicians as it outlines important considerations when using extrapolated evidence from common to rare cancers to make targeted treatment recommendations for their patients.

Ongoing work will articulate criteria for assessing the level of uncertainty to promote standardised decision-making for clinical and regulatory decisions and facilitate transparent discussion between key stakeholders in the development and evaluation of molecular targeted therapies.

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