Mast Cell Activation Syndrome as a Significant Comorbidity in Sickle Cell Disease

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Abstract: Some sickle cell anemia (SCA) patients suffer significantly worse phenotypes than others. Causes of such disparities are incompletely understood. Comorbid chronic inflammation likely is a factor. Recently, mast cell (MC) activation (creating an inflammatory state) was found to be a significant factor in sickle pathobiology and pain in a murine SCA model. Also, a new realm of relatively noncytoproliferative MC disease termed MC activation syndrome (MCAS) has been identified recently. MCAS has not previously been described in SCA. Some SCA patients experience pain patterns and other morbidities more congruent with MCAS than traditional SCA pathobiology (eg, vasoocclusion). Presented here are 32 poor-phenotype SCA patients who met MCAS diagnostic criteria; all improved with MCAS-targeted therapy. As hydroxyurea benefits some MCAS patients (particularly SCA-like pain), its benefit in SCA may be partly attributable to treatment of unrecognized MCAS. Further study will better characterize MCAS in SCA and identify optimal therapy.

Key Indexing Terms: sickle cell anemia; mast cell activation syndrome; KIT mutations; pain; hydroxyurea. [Am J Med Sci 2014;348(6):460–464.]

The contrast between the mutational homogeneity (in beta globin) and clinical heterogeneity in sickle cell anemia (SCA) has long been recognized. Putative factors associated with higher rates of painful vasoocclusive crises include higher hemoglobin concentration, lower hemoglobin F (HbF) concentration, higher hemolysis rate, higher blood viscosity and neutrophil activation, among others. Putative factors associated with higher mortality risk include vasoocclusive crises, acute chest syndrome, renal failure, seizures, lower hemoglobin concentration, lower HbF concentration and leukocytosis. Nevertheless, there remains substantial variability in crisis rates among SCA patients sharing similar levels of these factors; some endure frequent crises, whereas others suffer few crises—and some suffer none at all. Primary and emergency care physicians and hematologists know well the “poor-phenotype” minority of their SCA population who disproportionately present with crises and other SCA complications. In 1 study, the 5.2% of SCA patients who averaged 3 or more pain crises per year accounted for 32.9% of the SCA pain crises treated by physicians at hospitals.

One group of factors proposed to account for the clinical heterogeneity of SCA is genetic polymorphisms affecting not only aspects of hemoglobin production other than hemoglobin S production (eg, upregulation of HbF production, alpha thalassemia) but also other systems impacted by erythrocyte sickling. Another factor that may affect SCA clinical heterogeneity is inflammation, which might be consequential to the repeated vasoocclusive crises of SCA and/or other specific inflammatory ailments.

Inflammation is a complex milieu of humoral and cellular factors. Although granulocytes and lymphocytes are often considered among these cellular factors, the role of the mast cell (MC) has been less commonly appreciated. Recently, MC activation was identified as a key factor in the pathobiology and pain of SCA in a murine model. On the clinical front, there also has been recognition recently that the spectrum of primary MC disease extends beyond the various forms (eg, cutaneous, systemic) of the proliferative disease of mastocytosis to the relatively nonproliferative MC activation syndrome (MCAS). The clonal origins of mastocytosis and other myeloproliferative neoplasms (MPNs) have been appreciated for some time; more recently, the heterogeneity of these mutations across patients, and the complexity of the mutation set in any given patient, are being increasingly recognized. Similarly, there are preliminary data suggesting substantial intra-individual mutational complexity and interindividual mutational heterogeneity in MCAS.

Reported here for the first time is the presence of MCAS in a cohort of poor-phenotype sickle cell disease (SCD) patients.

PATIENTS AND METHODS

In the course of their routine clinical care, after recognition that some of their symptoms were more easily attributable to MCAS, 38 patients followed by the author for poor-phenotype SCA (mostly genotype SS; leading to at least 3 emergency department presentations and/or hospitalizations for sickle cell crises per year for the previous 5 years, and/or engaged in a treatment program of chronic red cell transfusions or hydroxyurea [HU] to mitigate frequent crises) were diagnosed by MCAS as described in recent reviews. In brief, testing included serum tryptase and chromogranin A (CgA) levels, plasma histamine and heparin and prostaglandin D2 (PGD2) levels, and spot and 24-hour urinary PGD2 and N-methylhistamine (NMH) levels. Patients were cautioned to avoid nonsteroidal anti-inflammatory drugs (potentially reducing prostaglandin production) and proton pump inhibitors (potentially escalating CgA production) for at least 5 days before specimen acquisition, and all samples were chilled on ice immediately on acquisition (the 24-hour urine samples were kept continuously refrigerated throughout collection) and kept chilled throughout handling and transport.

RESULTS

Thirty-two of the 38 evaluated poor-phenotype SCA patients (84%) were found to meet the current proposed diagnostic criteria for MCAS. This cohort of SCA/MCAS patients is summarized described in Table 1 and in more detail in Supplemental Digital Content 1 (see Table...
of the major sources of heterogeneity of clinical phenotype among SCA patients who all share the same point mutation in beta globin have long been investigated and debated. As noted above, a number of potential contributing factors have been identified, and yet it has generally been appreciated that a significant portion of the cause of poor-phenotype SCD (especially variants expected to be less morbid than SCA) remains poorly accounted. Inflammation has been proposed as a significant contributor to the morbidity and heterogeneity of such patients, but a common recurring cause of such inflammation has not yet been identified.

Recently, Vincent et al.14 found that in a murine model of SCA, MC activation underlies sickle pathophysiology leading to inflammation, vascular dysfunction, pain, and requirement for high doses of morphine, raising the questions of whether a similar process is present in sickle cell patients and whether therapies targeted at MC mediator production or action might help ameliorate some of the morbidities of SCD. The present series provides preliminary data that MC activation indeed is present in some poor-phenotype SCD patients, including not only those with SCA but also those with other sickle cell variants, and that therapy in such patients targeted at their MC activation can change the acuity of at least some of their clinical manifestations that traditionally have been attributed exclusively to their SCD.

Of hematopoietic origin, MCs are found in all human tissues, especially at the environmental interfaces and perivascular/perineural sites. They serve largely as sentinels of environmental change and bodily insults and respond by releasing variable assortments and levels of molecular mediators that directly and indirectly influence behavior in other (local and distant) cells and tissues to respond to changes/insults so as to maintain, or homeostasis. The transmembrane tyrosine kinase receptor KIT is the dominant MC regulatory element, shown to be critical for key MC functions including survival, differentiation, chemotaxis, and activation.33

Traditionally, MC disease has been thought to be principally a matter of neoplastic burdens of MCs (ie, mastocytosis), with symptoms resulting principally from an accompanying inappropriate release of mediators from these excessive MCs. Nearly a quarter century ago, though, the notion was first advanced that there might be forms of MC disease manifesting inappropriate mediator release with little to no accompanying MC cytoproliferation. This theory appeared validated when the first recognized cases of what is now called MCAS were published in 2007.14,33,34 MCAS typically causes chronic multisystem polymorbidity of a generally inflammatory theme. Different patterns of aberrant expression of the large MC mediator repertoire in different MCAS patients make for markedly heterogeneous—and thus diagnostically challenging—presentations (see Table, Supplemental Digital Content 2, http://links.lww.com/MAJ/A61). The cause of such heterogeneity of mediator expression in MCAS is not yet clear. Provocatively, though, Molderings et al.14,15 have repeatedly found a broad array of (presumably mostly constitutively activating) mutations scattered across all domains of KIT in small cohorts of MCAS patients, with most of their studied patients bearing multiple mutations in no yet-apparent recurring patterns. (Interestingly, too, the MC KIT

Given these new biological and clinical insights, proposals have emerged to consider all MC diseases, including mastocytosis and MCAS, under the umbrella term of MC activation disease (MCAD). It has also been proposed that the assorted systemic MCAD variants and clinical phenotypes represent not distinct disease entities but instead varying presentations of a common generic root process of MC dysfunction. Despite its rarity, mastocytosis is fairly readily recognizable because of its distinctive clinicopathological presentation, whereas MCAS, although suspected to be far more prevalent in the whole population than mastocytosis, is more challenging to recognize in large part because of the heterogeneity of its variant presentations, some of which are already discretely recognized (eg, idiopathic anaphylaxis, cryopyrin-associated periodic syndrome), but most not.
TABLE 1. Summary characteristics of 32 poor-phenotype SCA (and variant) patients with comorbid MCAS

| Age at time of MCAS diagnosis, yr | Range | Mean | Median |
|----------------------------------|-------|------|--------|
| Gender, n (%)                    |       |      |        |
| Male                             | 16 (50) |      |        |
| Female                           | 16 (50) |      |        |
| SCD variant, n (%)               |       |      |        |
| SS                               | 25 (78) |      |        |
| SC                               | 3 (9)  |      |        |
| S-β0 thalassemia                 | 3 (9)  |      |        |
| S-β+ thalassemia                 | 1 (3)  |      |        |
| CKD at time of MCAS diagnosis, n (%) |     |      |        |
| None                             | 23 (72) |      |        |
| Stage 1                          | 0 (0)  |      |        |
| Stage 2                          | 0 (0)  |      |        |
| Stage 3A                         | 1 (3)  |      |        |
| Stage 3B                         | 1 (3)  |      |        |
| Stage 4                          | 0 (0)  |      |        |
| Stage 5                          | 7 (22) |      |        |

| MC mediators (normal range) at time of MCAS diagnosis | Range | Mean | Median | Patients with nonelevated level, n (%) | Patients with elevated level, n (%) |
|------------------------------------------------------|-------|------|--------|---------------------------------------|------------------------------------|
| Serum tryptase (0.4–10.9 ng/mL, N = 32)              | 2.2–26.4 | 8.4  | 6.0    | 23 (72)                              | 9 (28)                             |
| CKD present at time of MCAS diagnosis (n = 9)         | 4.5–26.4 | 15.2 | 12.8   | 3 (33)                               | 6 (67)                             |
| CKD absent at time of MCAS diagnosis (n = 23)          | 2.2–12.7 | 5.4  | 4.6    | 22 (96)                              | 1 (4)                              |
| Plasma histamine (0–6 nmol/L, n = 23)                | 2–20 | 7.7  | 7      | 12 (52)                              | 11 (48)                            |
| Plasma prostaglandin D2 (35–115 pg/mL, n = 13)       | 64–342 | 144  | 132    | 5 (38)                               | 8 (62)                             |
| Urinary prostaglandin D2 (100–280 pg/mL, n = 31)     | 25–1287 | 382  | 304    | 14 (45)                              | 17 (55)                            |
| 24-hr collection (n = 19)                            | 25–1287 | 437  | 355    | 5 (26)                               | 14 (74)                            |
| Random/spot collection (n = 12)                       | 63–810 | 294  | 227    | 8 (67)                               | 4 (33)                             |
| Urinary N-methylhistamine (30–200 μg/g Cr, n = 30)    | 29–274 | 106  | 90     | 28 (93)                              | 2 (7)                              |
| 24-hr collection (n = 16)                             | 29–274 | 111  | 95     | 14 (87)                              | 2 (13)                             |
| Random/spot collection (n = 14)                       | 29–231 | 99   | 84     | 13 (93)                              | 1 (7)                              |
| Plasma heparin (0.00–0.02 anti-Factor-Xa units/mL,18 n = 5) | 0.02–0.09 | 0.06 | 0.07    | 1 (20)                               | 4 (80)                             |
| Serum chromogranin A (0–50 ng/mL, n = 28)            | 17–3746 | 533  | 58     | 14 (50)                              | 14 (50)                            |
| CKD present at time of MCAS diagnosis (n = 9)         | 126–3746 | 1553 | 1792   | 0 (0)                                | 9 (100)                            |
| CKD absent at time of MCAS diagnosis (n = 19)         | 17–129 | 50   | 40     | 22 (79)                              | 6 (21)                             |

Therapeutic result (N = 27, others LTFU), n (%)  
Complete response | 5 (19)  
Partial response | 22 (81)  

| 5 yr before diagnosis of MCAS | Since MCAS diagnosis (mean 31 mo) |
|-------------------------------|----------------------------------|
| Hospital days (n = 21, others LTFU) | | |
| Total                         | 501 | 400 |
| Annual mean                   | 4.9 | 19.0 |
| Emergency department visits (n = 21, others LTFU) | | |
| Total                         | 329 | 214 |
| Annual mean                   | 3.3 | 10.2 |

SCA, sickle cell anemia; MCAS, MC activation syndrome; SCD, sickle cell disease; CKD, chronic kidney disease; MC, mast cell; LTFU, lost to follow-up.
Such clinical heterogeneity of MCAD, including MCAS, and the similarity of the pain syndromes in SCA and MCAD may challenge health care providers to recognize MCAD in sickle cell patients from symptoms alone. As MC mediator testing is ordered only after clinical suspicion of MCAD is established, markers of MCAD in routinely obtained laboratory testing would be useful. It is interesting that in the present series, as has been reported for the MCAS patient population in general,\textsuperscript{40} not only was mild relative monocytosis frequently found (long described as a component of the inflammation seen in many sickle cell patients), but there also was frequent finding of mild relative eosinophilia and basophilia, abnormalities perhaps more suggestive of inflammation driven by MC activation than by neutrophil or monocyte/macrophage activation from vasoocclusion-driven inflammation. Modest elevations in the prothrombin time and activated partial thromboplastin time were also frequently seen in the present series.

The present case series suggests for the first time the presence of MCAS as a factor in the morbidity of poor-phenotype SCA. As hypothesized by Vincent et al\textsuperscript{45,46} several patients in the present series seem to have been significantly helped by therapy targeted at MC mediator production or action. Although it is interesting that current high-end estimates of the portion of the general population harboring MCAS (14\%–17\%\textsuperscript{41,42}) roughly correspond to the portion of the SCA population suffering a poor phenotype of SCA, specific prospective studies will be needed to confirm the portion of the SCA population truly harboring MCAS.

Many MCAD patients suffer migratory soft tissue and/or bone pain, which frequently responds poorly to typical (narcotic and non-narcotic) analgesics and atypical analgesics such as antidepressants and anticonvulsants. HU has long been recognized to effectively modulate the illness of SCA,\textsuperscript{47} including reducing pain crises, but its known mechanisms of action have not yet satisfactorily accounted in full for its observed effects in SCA.\textsuperscript{48} HU also has been used for decades to help occasional mastocytosis patients, provocatively sometimes reducing symptoms without clearly reducing tumor burden, and recently its utility in MCAS was also reported.\textsuperscript{49} Thus, it is possible that another mechanism by which HU helps in SCA is to ameliorate comorbid MCAS.

Noncompliance seems to have limited the benefit many of the patients in the present series might have achieved from MCAS-directed therapy. Noncompliance is common in chronic illness and seems to be caused by many factors including the degree of chronicity of the illness. That SCA patients often are chronically ill since infancy poses a significant challenge to educating those with comorbid MCAS of the value of compliance with MCAS-directed therapy. Earlier diagnosis and effective therapy of comorbid MCAS in SCA may be a route toward reducing health care resource utilization and noncompliance.

As noted above, MCAD’s heterogeneity and other issues pose diagnostic challenges (discussed in detail in recent reviews\textsuperscript{6,13}), and with no predictors of therapeutic response yet identified, this heterogeneity poses therapeutic challenges, too. Nevertheless, with a wide range of therapies (see Table, Supplemental Digital Content 4, http://links.lww.com/MAJ/A61) having shown benefit in assorted MCAD patients, it seems likely that a patient, persistent and methodical approach to a series of medication trials would yield benefit in some poor-phenotype SCA patients also found to harbor MCAS. If MCAS is confirmed to largely be a clonal illness, it may be possible before long—much as is developing in many other disease areas—to follow diagnosis of MCAS (whether in SCA or other settings) by determination, through whole exome or genome sequencing of isolated peripheral blood MCs, of the patient’s particular MC mutational pattern(s), which then might yield therapeutic insights leading to more efficient determination of effective therapy than afforded by the present trial-and-error approach.

CONCLUSIONS

Poor-phenotype SCD patients should be evaluated for possible comorbid MCAS. If found, MCAS should be treated, although biologically informed approaches to such treatment are limited at present and thus the path to effective individualized therapy can be challenging. Further study is needed to better characterize the prevalence and nature of MCAS in SCD and to define efficient approaches to identifying effective therapy.

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