Assessment of Matrix Metalloproteinases-8 and -9 in Gingival Crevicular Fluid of Smokers and Non-smokers with Chronic Periodontitis Using ELISA

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Abstract:
Background: Matrix metalloproteinases (MMP)-8 is increased in smokers with chronic periodontitis (CP). The levels of MMP-9 have not been assessed in the periodontal tissues of smokers. Therefore, the aim of this study is to estimate the concentrations of MMP-8 and -9 in the gingival crevicular fluid (GCF) of smokers and non-smokers with CP.

Materials and Methods: Plaque, bleeding on probing, probing pocket depth, and attachment level were assessed in 60 subjects who were equally divided into three groups. Group A consisted of smokers with CP, Group B - non-smokers with CP and Group C - periodontally healthy subjects. The concentrations of MMP-8 and -9 were determined from GCF samples by using enzyme-linked immunosorbent assay.

Results: A highly significant difference (P < 0.0001) was found in the MMP-9 concentration between the Groups A (27.56 ± 2.87 ng/ml) and B (17.16 ± 1.74 ng/ml) and between Groups A and C (14.13 ± 2.87 ng/ml). A significant difference (P < 0.002) was also found between the Groups B and C. A highly significant difference (P < 0.0001) in MMP-8 concentration was found between the A (68.96 ± 20.53 ng/ml) and C (7.05 ± 0.75 ng/ml) and also between Groups B (56.97 ± 12.70 ng/ml) and C (7.05 ± 0.75 ng/ml) and also between Groups A and C (14.13 ± 2.87 ng/ml) and C (7.05 ± 0.75 ng/ml).

Conclusion: Our findings of significantly elevated MMP-9 than MMP-8 from the diseased sites in smokers imply that MMP-9 is more significantly related to the increased periodontal destruction seen among smokers with CP.

Key Words: Chronic periodontitis, gingival crevicular fluid, matrix metalloproteinases-8, matrix metalloproteinases-9, smoking

Introduction
Matrix metalloproteinases (MMPs) represent a superfamily of proteases acting not only in physiological development and tissue remodeling, but also in pathological tissue destruction. Pathogens in microbial dental plaque are capable of stimulating host cells to increase their MMP release, which is considered among the indirect mechanisms of tissue destruction seen during periodontitis. MMPs are secreted in latent forms that can be activated by microbial-derived proteases or host inflammatory mediators such as tumor necrosis factor-α (TNF-α), interleukin-1β (IL-1β), and reactive oxygen species. MMPs can collectively degrade almost all components of extracellular matrix and basement membrane and their excess activity in the periodontium leads to periodontal tissue destruction.

MMP-8 (collagenase-2) is synthesized by differentiating granulocytes in the bone marrow and stored in specific granules of circulating neutrophils. However, the other cellular sources are sulcular epithelial cells, gingival and periodontal ligament fibroblasts, monocytes/macrophages, and plasma cells. MMP-8 is detected as a latent pro-enzyme in the gingival crevicular fluid (GCF) of shallow pockets, but in deep periodontal pockets it is converted to the active form. MMP-9, a gelatinase is secreted mainly by neutrophils and it is capable of degrading denatured interstitial collagens, gelatins, laminin, elastin, fibronectin and also collagens Type IV and Type VII. During active and progressing phases of periodontitis, MMP-8 and -9 levels in the GCF are significantly elevated.

Smoking is a major risk factor as well as a modifier for the progression of periodontal disease. Chronic smokers present with an altered response of specific MMPs-8 and -9. MMP-8 expression is higher in the periodontal tissue of smokers than non-smokers that indicate that smoking modulates the expression of MMP-8 in periodontal tissues.

This study which evaluated the levels of MMP-8 and -9, in smokers and non-smokers with periodontitis, was performed to assess the relationship between the expression levels of MMP-8 and -9 in the GCF.
Materials and Methods

Study population
Sixty subjects were selected from the patients who attended the clinics of the Department of Periodontics of SRM dental college, for the management of their dental and oral conditions. Subjects were classified into three groups - Group A (smokers with chronic periodontitis [CP]), Group B (non-smokers with CP) and Group C (clinically healthy periodontium) based on the smoking status and periodontal parameters. The study protocol was approved by the Ethical Committee of SRM University. All participants gave written informed consent in accordance with the Helsinki Declaration. The dental, as well as medical histories, were obtained from the patient. Pregnant and lactating women, patients with chronic inflammatory systemic diseases and patients with acute infections were excluded from the study. Systemically healthy patients between 18 and 45 years who had not received any antibiotic therapy and periodontal therapy in the last 6 months and having a minimum of 20 teeth (excluding third molars) with not more than two teeth missing in each quadrant were included in the study.

All patients in the smokers and non-smokers group with CP showed clinically detectable attachment loss and bone loss characterized by periodontal pocket formation and/or gingival recession with changes in density and height of subjacent alveolar bone as detected on the intraoral periapical radiographs. Each patient had at least one site with probing pocket depth (PPD) of ≥6 mm were diagnosed as CP in accordance with the clinical criteria stated in the consensus report of the World Workshop in periodontitis.10

Periodontal status evaluation
Clinical parameters assessed for the study were plaque index (PlI), sulcular bleeding index (SBI), bleeding on probing (BOP), PPD and clinical attachment level. The periodontal parameters were assessed at six sites per tooth excluding third molars. BOP (deemed positive if it occurred within 15 s after periodontal probing) and visible plaque accumulation were recorded dichotomously by visual examination. All measurements were performed by a single calibrated examiner using a Williams periodontal probe. GCF samples were obtained using extra-sulcular method before initiation of any periodontal intervention. The total levels of MMP-8 and -9 in GCF were measured using a commercially available enzyme-linked immunosorbent assay (Quantikine; R&D Systems Inc., Minneapolis, MN, USA) according to the manufacturer’s instructions.

MMP-8 and -9 analysis
All reagents and samples were brought to room temperature before use and prepared for the assay. 150 μl of assay diluents RD1-S2 was added to each well, followed by 50 μl of standard or sample to each well. The plate was covered and incubated for 2 h at room temperature (18-25°C) and was gently tapped to ensure thorough mixing. The cover was removed, and plate was washed four times using the wash buffer. After the last wash, the plate was inverted and blotted against clean paper towels. 200 μl of MMP-8 or -9 conjugate was added to all the wells.

The plate was then securely covered with a plate sealer and incubated for 1 h at room temperature. The plate was washed 4 times followed by the addition of 200 μl of substrate solution to each well. It was then incubated for 30 min at room temperature and protected from light. The optical density values for the plate were monitored, and the substrate reaction stopped before positive wells were no longer properly readable. The enzyme-substrate reaction was stopped quickly by pipetting 50 μl of stop solution to each well. The color in the wells changed from blue to yellow. The optical density of each well was determined within 30 min, using a microplate reader set to 450 nm as the primary wavelength. The levels of MMP-8 in the samples were estimated using the standard curve. The same procedure was followed for determining the levels of MMP-9 in the GCF samples using a standard curve. MMP-8 and -9 concentrations were obtained in ng/ml.

Statistical analysis
Mean and standard deviation was estimated from the sample for each study group. Mean values were compared among different study groups by using Kruskal–Wallis one-way ANOVA and then post-hoc (Tukey HSD) test. Spearman rank correlation analysis was done to assess the relationship between various study parameters. In the present study, \( P < 0.05 \) was considered as the level of significance. The statistical software SPSS version 17 was used for the analysis of the data.

RESULTS

The comparison of mean values of PI, sulcular bleeding index, pocket depth, MMP-8 and -9 among the three study groups are shown in Table 1.

PlI
There was no statistical significant difference (\( P > 0.025 \)) found in the PlI between Groups A and B. A very high statistical significant difference (\( P < 0.0001 \)) was found between Groups A and C and also between Groups B and C.

Table 1: Comparison of mean values of clinical parameters, MMP-8 and MMP-9 concentrations among the three study groups.

| Clinical parameters/ MMP concentration | Group A (Mean±SD) | Group B (Mean±SD) | Group C (Mean±SD) |
|---------------------------------------|------------------|------------------|------------------|
| PI                                    | 2.63±0.37        | 2.37±0.28        | 1.43±0.24        |
| SBI                                   | 1.7±0.47         | 2.2±0.41         | 0.18±0.87        |
| Pocket depth (mm)                     | 7.2±0.95         | 6.75±1.02        | 2.5±0.51         |
| MMP-8 (ng/ml)                         | 68.96±20.5       | 56.97±12.70      | 7.05±0.75        |
| MMP-9 (ng/ml)                         | 27.56±2.87       | 17.16±1.74       | 14.13±3.02       |

MMP: Matrix metalloproteinases, SD: Standard deviation, PI: Plaque index
**Sulcular bleeding index**
A statistically highly significant difference (P < 0.0001) was found between the study groups.

**PPD**
No statistical difference (P < 0.230) in the PPD between Groups A and B, but a statistically high significant difference (P < 0.0001) was found between Groups A and C and also between Groups B and C.

**MMP-8 concentration**
The mean MMP-8 concentration was 68.96 ± 20.53 ng/ml in Group A, 56.97 ± 12.70 ng/ml in Group B and 7.05 ± 0.75 ng/ml in Group C. A statistically mild significant difference (P < 0.023) was found in the MMP-8 concentration between Groups A and B, and statistically high significant difference (P < 0.0001) was found between Groups A and C and between Groups B and C (Table 2).

**MMP-9 concentration**
The mean MMP-9 concentration was 27.56 ± 2.87 ng/ml in Group A, 17.16 ± 1.74 ng/ml in Group B and 14.13 ± 3.02 ng/ml in Group C. A statistically highly significant difference (P < 0.0001) was found in the MMP-9 concentration between Groups A and B, between Groups A and C and a statistically significant difference (P < 0.002) was also found between Groups B and C (Table 2).

**Discussion**
In this study, we analyzed the GCF levels of MMP-8, and -9, in smokers and non-smokers with CP patients in an attempt to explain possible mechanisms of the detrimental effects of smoking on periodontal tissues. Tobacco smokers are reported to exhibit an increased susceptibility for development and progression of periodontal diseases. Knowing the fact that MMPs -8 and -9 are the major metalloproteinases that take part in connective tissue breakdown of periodontitis, the current study was performed to estimate and evaluate the possible role of these MMPs in periodontal disease in both smokers and non-smokers. MMP-9 is involved in bone resorption, playing a role in the subsequent digestion of denatured collagen I after being cleaved by collagenase. Although many studies have evolved to establish a relation between MMP-8 and -9 and the extent of periodontal diseases, there are no studies to date that evaluate the levels of MMP -9 in cohorts representing smokers. Knowing the fact that MMPs -8 and -9 are the major metalloproteinases that take part in connective tissue destruction in periodontal disease, the current study was performed to estimate and evaluate the possible role of these MMPs in periodontal disease in both smokers and non-smokers compared to the healthy cohorts devoid of periodontal disease.

The results of the current study showed that the mean MMP-8 concentrations were higher in smokers with CP when compared with non-smokers with CP and healthy patients.

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**Table 2:** Comparison of the levels of MMP-8 between the study groups.

| (I) Group          | (J) Group          | Mean difference (I-J) | Standard error | Significant | 95% confidence interval | Upper bound | Lower bound |
|--------------------|--------------------|-----------------------|----------------|-------------|-------------------------|-------------|-------------|
| Smokers with CP    | Non-smokers with CP| 11.99285*             | 4.410306       | 0.023       | 1.37                    | 22.60       |             |
| Healthy controls   |                    | 61.91050*             | 4.410306       | 0.0001      | 51.29                   | 72.52       |             |
| Non-smokers with CP| Smokers with CP    | -11.99285*            | 4.410306       | 0.023       | -22.60                  | -1.37       |             |
| Healthy controls   |                    | 49.91765*             | 4.410306       | 0.0001      | 39.30                   | 60.53       |             |
| Healthy controls   | Smokers with CP    | -61.91050*            | 4.410306       | 0.0001      | -72.52                  | -51.29      |             |
| Non-smokers with CP|                    | -49.91765*            | 4.410306       | 0.0001      | -60.53                  | -39.30      |             |

CP: Chronic periodontitis, MMP: Matrix metalloproteinases, *The mean difference is significant at the 0.05 level.
Table 3: Comparison of the levels of MMP-9 between the study groups.

| (I) Group          | (J) Group          | MMP-9       | 95% confidence interval |
|--------------------|--------------------|-------------|-------------------------|
|                    |                    | Mean difference (I-J) | Standard error | Significant | Upper bound | Lower bound |
| Smokers with CP    | Non-smokers with CP | 10.406300   | 0.826229  | 0.0001   | 8.41       | 12.39       |
| Healthy controls   |                    | 13.437250   | 0.826229  | 0.0001   | 11.44      | 15.42       |
| Non-smokers with CP| Smokers with CP    | -10.406300  | 0.826229  | 0.0001   | -12.39     | -8.41       |
| Healthy controls   |                    | 3.030950    | 0.826229  | 0.002    | 1.042      | 5.01        |
| Smokers with CP    | Healthy controls   | -13.437250  | 0.826229  | 0.0001   | -15.42     | -11.44      |
| Non-smokers with CP|                    | -3.030950   | 0.826229  | 0.002    | -5.01      | -1.04       |

CP: Chronic periodontitis, MMP: Matrix metalloproteinases

However, this increase in levels seen amongst smokers was not statistically significant (P < 0.023). This in accordance with the earlier studies conducted by Söder\textsuperscript{20,21} which identified that MMP-8 concentration was not significantly increased among smokers. A statistically high significant difference (P < 0.0001) was present between smokers with CP on comparison with healthy cohorts.

The GCF levels of MMP-9 in Group A and Group B was significantly elevated than healthy cohorts. However on comparing the MMP-9 levels between patients in Groups A and B, a statistically high significant difference (P < 0.0001) was noted. This shows that smokers with CP had increased levels of MMP-9 than the non-smokers with CP. This higher level of MMP-9 in the GCF of patients with CP indicates that MMP-9 may be a significant factor that modulates the increased periodontal breakdown that is evident amongst smokers. This study concludes that GCF levels of MMP-9 could be a very significant marker for periodontal breakdown.

This current study, as many as others has not found a significant difference between the levels of MMP-8 in smokers and non-smokers with CP, indicating that it may not be a significant marker of periodontal breakdown.\textsuperscript{20,21} This study that was carried out to identify the difference in the capacity of smokers and non-smokers to elicit markers of connective tissue breakdown a necessary feature required prior to the development of attachment loss. This study proposes that MMP-9 would be a more significant marker for impending periodontal attachment loss and more specifically in smokers. This will need to be borne out by longitudinal studies that correlate the levels of MMP-9 with ensuing attachment loss.

Further directions in the search for a clinically relevant biomarker will require that studies are conducted longitudinally to identify whether the MMP is in its active or inactive form for such assays to be more significant.

**Conclusion**

Our present finding of significantly higher GCF concentrations of MMP-9 in smokers with CP patients than their non-smokers and healthy cohort counterparts provides further support for the smoking associated risk for increased periodontal destruction.

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