Spinal Intradural, Extramedullary Ependymoma with Astrocytoma Component: A Case Report and Review of the Literature

1. Introduction

Most adult ependymomas are spinal lesions, with the vast majority arising in an intradural, intramedullary location [1]. Intradural, extramedullary ependymomas are exceptionally rare tumors that have been reported in the literature [2, 3]. We report a case of an intradural, extramedullary ependymoma with an astrocytoma component. To the best of our knowledge, this has not been described in the literature before. Both the imaging and pathologic features are described here.

2. Case Report

A 56-year-old female presented to an outside hospital for several months of low back pain, bilateral hip pain, and leg pain/weakness. She subsequently had an episode of severe low back pain that resulted in difficulty breathing and loss of consciousness. Review of systems revealed several episodes of incontinence. Physical exam was remarkable for 5/5 strength in all extremities, 2+ reflexes in the lower extremities, and a negative Babinski sign.

Magnetic resonance imaging of the lumbar spine performed with and without intravenous contrast revealed a 2.1 cm × 1.5 cm × 1.3 cm intradural, extramedullary mass at L1-L2 which compressed the filum terminale. Precontrast MRI showed intrinsic T1 hypointensity and heterogeneous T2 hyperintensity (Figures 1(a) and 1(b)). Postcontrast imaging showed homogenous postcontrast enhancement (Figures 1(c) and 1(d)). The presumed diagnosis was schwannoma.

Given the patient's symptoms, she underwent elective operative resection of the mass. A posterior L1-L2 laminectomy was performed. A tan, fleshy intradural tumor was identified (Figure 2). Most of the cauda equina was ventral to the tumor. Several small vessels were seen feeding the tumor, which were coagulated. Several small nerve rootlets were also seen leading into the tumor and the tumor had grown around...
one of the rootlets. The tumor did not appear to arise from any one particular rootlet and was distinct from the filum terminale. The nerve rootlets were dissected free of the tumor and the tumor was removed en bloc. The patient tolerated the procedure well and there were no complications.

Gross examination revealed an oval shape soft nodule with a smooth surface, which appeared encapsulated with a delicate membrane. Permanent sections showed cellular biphasic glial neoplasm: moderately cellular tumor with numerous perivascular pseudorosettes and less cellular areas with prominent eosinophilic fascicular component (Figure 3(a)). The cellular areas with pseudorosettes were consistent with ependymoma. The cells in the fascicular component were similar to the cells forming perivascular arrangements, but the cytoplasm was more eosinophilic and fibrillary. Many Rosenthal fibers were noted within the fascicular component, along with giant cell-like tumor cells and cystic change (Figure 3(b)). Focally, within the same pseudorosette, some cells show ependymal features, while others are more eosinophilic and fibrillary with Rosenthal fibers (Figure 4(a)). The fascicular component with Rosenthal fibers was representative of astrocytic lesion, reactive and/or tumor. Macro- and microcyst formation and Rosenthal fibers, along with giant tumor cells, which were present in fascicular component, are the known features of astrocytic tumor. There was no necrosis and the mitotic count was low. Interestingly, the periphery of the lesion consisted of ependymal component and central portion of the lesion had fibrillary and ependymal components intermingled (Figure 4(b)). These findings are consistent with a low grade glial lesion.

The lesional cells, both components, showed strong, diffuse immunopositivity with glial fibrillary acidic protein (GFAP) and S-100 (more in fascicular component). Synaptophysin was staining stronger in the ependymal component (Figure 5(a)). The Ki-67 proliferation rate was low, around 1-2%, in both components. The epithelial membrane antigen (EMA) showed cytoplasmic dot-like positivity in many cells of ependymal component (Figure 5(b)).
In summary, the microscopic examination revealed low grade glial neoplasm, with histologic features of ependymoma and pilocytic astrocytoma, closely intermixed, and represented collision tumor.

The tumor was analyzed for mutation using a massively parallel (“next generation” or NGS) sequencing approach, performed by Foundation Medicine, INC, Cambridge, MA. This test assesses genomic alterations including base pair substitutions, insertions and deletions, and copy number alterations and select gene rearrangements for 315 cancer-related genes and introns from 28 genes often rearranged or altered in cancer, including NF1, NF2, IDH1, and TP53 [4]. The mutation in MUTYH G382D was identified in our sample. Normal tissue was not available for control testing. In addition, gene fusion testing was performed for the BRAF-KIAA1549 gene via FISH analysis, and the result was negative.
The patient had no postoperative complications. At one-month follow-up, she had resolution of lower extremity pain and tingling. At one-year imaging and clinical follow-up, patient had no evidence of tumor recurrence.

3. Discussion

Ependymoma (WHO grade II) is the most common intramedullary intradural spinal tumor followed by pilocytic astrocytoma. Although uncommon, spinal ependymomas (WHO grade II) have been described in an extramedullary, intradural location. To our knowledge, 23 such cases have been described [2, 3, 5–13]. The majority of these were in the thoracic spine, with four isolated to the lumbar spine [3, 6, 8]. The typical patient on presentation is a middle-aged female with spinal compressive symptoms of pain and/or myelopathy. All the previously described tumors were diagnosed as ependymomas. While a few reported cases showed higher grade components [9, 14], there were no cases of a collision/composite tumor.

Other extramedullary tumors of glial origin have been reported. Singh et al. described an astrocytoma in the conus region [15]. Kumar et al. reported a cystic pilocytic astrocytoma of the cauda equina, though that patient also had intracranial neoplasms and therefore likely had an underlying cancer-causing mutation such as NFI or NF2 [16]. Both of these cases were primary extramedullary tumors with no intramedullary component.

Our case showed an extramedullary low grade glial tumor with histologic characteristics of an ependymoma and astrocytic lesion. Additionally, a fascicular component with Rosenthal fibers and cystic change was seen, consistent with pilocytic astrocytoma, and was intermixed with ependymoma component. This leads us to speculate the possibility that the lesion has dual origin: ependymal and astrocytic. We considered the option that the astrocytic component might represent piloid gliosis within a WHO grade II ependymoma. However, this was discounted as the astrocytic component was not focal or at periphery of the lesion, which one would expect for reactive change/piloid gliosis, but rather interspersed throughout and comprised at least half of the entire lesion. In addition, the presence of ependymal and fibillary differentiation within the same pseudorosette further supports the hypothesis of compound tumor.

There are no absolute histologic criteria or immunohistochemical markers to differentiate piloid gliosis from pilocytic astrocytoma. Recent studies have shown that some pilocytic astrocytomas carry duplication at chromosome band 7q34 containing a BRAF-KIAA1549-gene fusion [17]. Although our specimen was negative for the KIAA1549-gene fusion transcript, the frequency of this is significantly lower in the adult population (<10% in above 40-patient populations). Therefore, the sensitivity of this diagnostic marker is weakened, and a negative FISH result does not rule out the possibility of the fascicular component of the lesion being pilocytic astrocytoma [18]. However, MUTYH G382D was identified by next-generation sequencing (NGS). To date, no association has been found between the MUTYH gene and glial neoplasia, though the G382D variant has been implicated in adenomatous polyposis [19]. Zadnik et al. identified several genes associated with astrocytoma, including CDK2NA, H2F3A, IDH1, NFI, TP53, ATRX, and PTEN [20]. All of these were tested by NGS and no mutations were found. Zadnik et al. also identified multiple genes associated with ependymoma, HOXB5, and PLA2G5, though neither of those were tested by NGS.

Several cases of mixed astrocytoma and ependymoma have been described, though none have been identified in the spine. Molnár and Hegedüs encountered an intraventricular ependymoma in the setting of a partially removed infiltrating astrocytoma in the frontal lobe [21]. However, this was felt to be a collision tumor between two distinct tumor origins. Kondziolka and Bilbao described a mixed ependymoma-astrocytoma in the parietal lobe, with features of a subependymoma but far from any ependymal lining [22].

There are case reports of ependymomas to have mesenchymal differentiation, for example, cartilage formation, which can be seen in adults and children [23, 24]. It is possible that we have a case where ependymal cells differentiate into astrocytic lineage.

4. Conclusion

To our knowledge, no cases of mixed ependymoma-astrocytoma have been reported. A tumor of glial origin should be included in the differential of an extramedullary, intradural mass, and based on our findings, a composite tumor should be considered in such a case.

Competing Interests

The authors declare that they have no competing interests.

References

[1] J. E. Marks and S. J. Adler, "A comparative study of ependymomas by site of origin," International Journal of Radiation Oncology, Biology, Physics, vol. 8, no. 1, pp. 37–43, 1982.
[2] M. J. Kinsman, J. D. Callahan, E. M. Hattab, and A. A. Cohen-Gadol, "Extramedullary spinal ependymoma: a diagnostic challenge and review of the literature," Clinical Neurology and Neurosurgery, vol. 113, no. 8, pp. 661–664, 2011.
[3] P. Tripathy, D. Mohapatra, and S. Mohapatra, "Primary intradural extramedullary ependymoma: report of two cases and review of the literature," Neurologiai Neurochirurgia Polska, vol. 45, no. 4, pp. 397–401, 2011.
[4] Foundation Medicine, Technical Information and Test Overview, http://foundationone.com/ONE-I-001-20140804_(nobleed)TechnicalSpecs.pdf.
[5] C. M. Bonfield, D. Amin, R. L. Hamilton, and P. C. Gerszten, "Extramedullary ependymoma near the conus medullaris with lumbar nerve root attachment: case report," Neurosurgery, vol. 68, no. 3, pp. E831–E834, 2011.
[6] N. Fuentes Rodriguez, M. de la Paz Rivero, J. A. Prince López, and J. H. Salas Rubio, "Ependimoma intradural extramedullar primario," Revista Cubana de Medicina Militar, vol. 33, no. 1, 2004.
[7] J. Graça, N. Gütlaçlı, N. D’Haene, J. Brotchi, I. Salmon, and D. Baleriaux, “Cystic extramedullary ependymoma,” *American Journal of Neuroradiology*, vol. 27, no. 4, pp. 818–821, 2006.

[8] E. A. Junes, J. N. Stávale, R. D. C. C. Pessoa et al., “Multifocal intradural extramedullary ependymoma: case report,” *Journal of Neurosurgery: Spine*, vol. 14, no. 1, pp. 65–70, 2011.

[9] T. Ikata, S. Katoh, A. Inou, and M. Takahashi, “Intradural extramedullary ependymoma: a case report,” *Spine*, vol. 20, no. 18, pp. 2036–2038, 1995.

[10] M. H. Li, S. Holtås, and E.-M. Larsson, “MR imaging of intradural extramedullary tumors,” *Acta Radiologica*, vol. 33, no. 3, pp. 207–212, 1992.

[11] T. Moriwaki, K. Iwatsuki, Y.-i. Ohnishi, M. Umegaki, M. Ishihara, and T. Yoshimine, “Intradural extramedullary spinal ependymoma: a case report of malignant transformation occurring,” *Asian Spine Journal*, vol. 7, no. 2, pp. 139–142, 2013.

[12] B. Oliver, A. de Castro, M. A. Sarmiento, C. Argüello, and M. G. Blázquez, “Dorsal extramedullary ependymoma,” *Archivos de Neurobiologia*, vol. 44, no. 4, pp. 215–224, 1981.

[13] M. Schuurmans, J. A. L. Vanneste, M. J. T. Verstegen, and W. R. van Furth, “Spinal extramedullary anaplastic ependymoma with spinal and intracranial metastases,” *Journal of Neuro-Oncology*, vol. 79, no. 1, pp. 57–59, 2006.

[14] A. Cerase, C. Venturi, G. Oliveri, D. De Falco, and C. Miracco, “Intradural extramedullary spinal anaplastic ependymoma. Case illustration,” *Journal of Neurosurgery: Spine*, vol. 5, no. 5, p. 476, 2006.

[15] P. Singh, A. P. Singh, T. Rajaram, and A. K. Sabhikhi, “Extradural astrocytoma of conus region: a short report,” *Neurology India*, vol. 49, no. 1, pp. 97–99, 2001.

[16] A. Kumar, A. Garg, M. C. Sharma, B. S. Sharma, and S. S. Kale, “Giant cystic intradural extramedullary pilocytic astrocytoma of Cauda equina,” *Journal of Neurosciences in Rural Practice*, vol. 4, no. 4, pp. 453–456, 2013.

[17] M. Hasselblatt, B. Riesmeier, B. Lechtape et al., “BRAF-KIAA1549 fusion transcripts are less frequent in pilocytic astrocytomas diagnosed in adults,” *Neuropathology and Applied Neurobiology*, vol. 37, no. 7, pp. 803–806, 2011.

[18] A. J. Sievert, S.-S. Lang, K. L. Boucher et al., “Paradoxical activation and RAF inhibitor resistance of BRAF protein kinase fusions characterizing pediatric astrocytomas,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 110, no. 15, pp. 5957–5962, 2013.

[19] V. Gismondi, M. Meta, L. Bonelli et al., “Prevalence of the Y165C, G382D and 1395delGGA germline mutations of the MYH gene in Italian patients with adenomatous polyposis coli and colorectal adenomas,” *International Journal of Cancer*, vol. 109, no. 5, pp. 680–684, 2004.

[20] P. L. Zadnik, Z. L. Gokaslan, P. C. Burger, and C. Bettegowda, “Spinal cord tumors: advances in genetics and their implications for treatment,” *Nature Reviews Neurology*, vol. 9, no. 5, pp. 257–266, 2013.

[21] P. Molnár and K. Hegedűs, “Adjacent astrocytoma and ependymoma: dependent mixed neoplasia or unusual collision tumors?” *Surgical Neurology*, vol. 22, no. 5, pp. 455–460, 1984.

[22] D. Kondziolka and J. M. Bilbao, “Mixed ependymoma-astrocytoma (subependymoma?) of the cerebral cortex,” *Acta Neuropathologica*, vol. 76, no. 6, pp. 633–637, 1988.

[23] A. Coli, M. Novello, L. Massimi, M. Caldarelli, V. Ranucci, and L. Lauriola, “Cartilage differentiation in ependymoma: histogenetic considerations on a new case,” *Child’s Nervous System*, vol. 30, no. 7, pp. 1301–1305, 2014.

[24] M.-M. Ruchoux, J. J. Kepes, P. Dhellemmes et al., “Lipomatous differentiation in ependymomas: a report on three cases and comparison changes reported in other central nervous system neoplasms of neuroectodermal origin,” *American Journal of Surgical Pathology*, vol. 22, no. 3, pp. 338–346, 1998.