Procalcitonin in Preterm Neonates: A Different Threshold and Prolonged Interpretation

Blandine Bianco¹, Bérengère François-Garret¹, Marine Butin², Cyril Dalmasso³, Florence Casagrande¹, Mostafa Mokhtari⁴ and Sergio Eleni Dit Trolli²*

¹ Neonatal Intensive Care Unit, CHU de Nice, Archet 2 Hospital, Nice, France, ² Neonatal Intensive Care Unit, Hospices Civils de Lyon, Hôpital Femme-Mère-Enfant, Bron, France, ³ Laboratoire de Mathématiques et Modélisation d’Evry (LaMMe), Université d’Evry Val d’Essonne, UMR CNRS 8071, Evry, France, ⁴ Neonatal Intensive Care Unit, Assistance Publique-Hôpitaux de Paris, Kremlin-Bicêtre Hospital, University Paris Sud, Kremlin-Bicêtre, France

Objectives: To evaluate the positive threshold of PCT for neonates of <32 weeks of gestation for the diagnosis of early-onset sepsis and to determine if the level of PCT collected within 6 h of life could be used.

Design: Retrospective and bicentric study from May 2016 to April 2018.

Setting: Two groups were established, neonates evaluated for PCT at birth (CordPCT) and within 6 h of life (delPCT).

Patients: Two hundred and sixty neonates of <32 weeks of gestation born in Nice and South Paris (Bicêtre) University Hospitals, had been evaluated for PCT level.

Main Outcomes Measures: The value of the PCT positive threshold was determined for the total population and each groups thanks ROC curves.

Results: The threshold level of PCT for the total population was 0.98 ng/mL. The threshold value of cordPCT group was 1.00 vs. 0.98 ng/mL for delPCT group. The area under the Receiver Operating Characteristics curve for PCT sampled in delPCT group was significantly higher than in cordPCT group (0.94 compared to 0.75).

Conclusions: The threshold level of PCT was higher in this cohort of neonates of <32 weeks of gestation compared to the value generally described for term neonates. The secondary sampling PCT level seems to be usable in screening algorithm for early-onset neonatal sepsis.

Keywords: procalcitonin, early-onset sepsis, preterm, biomarker, threshold

INTRODUCTION

Early-onset neonatal sepsis (EOS) is defined as a systemic infection occurring within the first 72 h of life (1). The incidence of EOS among premature neonates <1,500 g is 8–26 for 1,000 births (2). Infection is responsible for the high morbidity and mortality for preterm infants (3), due to the immaturity of their innate immune system (4). Diagnosis is still difficult when considering the perinatal infection risk factors, such as non-specific clinical symptoms (5, 6) biomarkers of low sensitivity and blood cultures that are often negative (7).
The majority of extremely premature neonates [78.6% of neonates of <1,500 g and 87% of <1,000 g (8)] receive large spectrum antibiotic therapy immediately in postnatal, which is often justified within a context of maternal infection and non-specific or absent neonatal clinical symptoms (9). Antibiotic therapy has short-term consequences: delay in colonization of the intestinal microbiota, decrease in the diversity in favor of bacterial pathogens (Enterobacter, Enterococcus and Streptococcus) (10–12), increase in the risk of delayed bacterial infection, necrotizing enterocolitis and death (13). Long-term modifications to the intestinal microbiota result in an increased risk of developing diabetes, allergies (asthma, eczema) and chronic inflammatory bowel disease (14).

The prohormone procalcitonin (PCT) is more sensitive than C-reactive protein (CRP) and the leucocyte count in diagnosis of EOS (15–17). However, a few issues concerning PCT must be understood when interpreting the clinical relevance of the level: the physiological increase during the first few days of life (18–20) and the kinetic differences for preterm infants (21) [delayed plasmatic peak and prolonged return to the basal level (19, 22)]. The PCT level of the umbilical cord avoids the postnatal physiological increase. The threshold positive level for PCT of the umbilical cord is 0.6 ng/mL (17) and is the same for neonates, whatever the term of birth. Only a few studies have evaluated the level of PCT of the umbilical cord in premature neonates (21, 23) and only one regarding neonates of <28 weeks (24). Neonates <32 weeks represent a small percentage of babies included in these studies. The data are discordant concerning the possible effect of gestational age on PCT (22, 25, 26). Defining the positive threshold value for neonates of <32 weeks will improve the diagnosis of EOS in this population by better targeting newborns at risk and limiting unnecessary antibiotic therapy.

Most studies concentrate on the PCT of the umbilical cord due to its high post-natal physiological level. A physiologic increase in the PCT level occurs after the first 6 h of life (27). The aim of this work is also to assess PCT before the first 6 h of life during admission to the critical care unit and during intensive neonatal care.

The main objective of this study was to determine the positive threshold level of PCT in premature neonates <32 weeks. The secondary objective was to compare the level of PCT obtained during the first 6 h of life with the level of the umbilical cord at birth.

**Inclusion and Exclusion Criteria**
All the premature neonates born in one of the two centers who presented infection risk factor(s) and sampled for PCT at birth or in the first 6 h of life were included in this study. Infection risk factors have been defined by the ≫ Haute Autorité de Santé (HAS) ≫ (28) and the ≫ Société Française de Néonatalogie (SFN) ≫ (29). Neonates without PCT evaluation, without infection risk factors, born outside of a hospital with a neonatal critical care unit or with malformations diagnosed antepartum or with chromosomal anomalies were not included. Neonates were excluded if the infectious status had not been determined due to incomplete clinical or biological data.

**PCT Laboratory Test**
A sample of whole blood of 500 µL was obtained from an umbilical cord vein (from the cord or after introducing a catheter into the umbilical vein) or from a peripheral vein after introducing a peripheral venous line. Microvette® 500 Hep-Li-Gel (SARSTEDT, Nümbrecht, Germany) sampling tubes containing lithium heparin and a separating gel were used. The PCT level was evaluated by sandwich immunoassay and electro chemiluminescence. The Nice University Hospital used a ADVIA Centaur CP (SIEMENS HEALTHINEERS, Erlangen, Germany) system with the ADVIA Centaur BRAHMS PCT reactive. The Bicêtre University Hospital used a COBAS® 8000 (ROCHE DIAGNOSTICS, Bâle, Switerland) system with the Elecsys BRAHMS PCT reactive. Comparative tests for PCT using the different methods using the BRAHMS reactive were performed. The result of analysis of PCT with the different reactive were the same (correlation coefficient close to one).

**EOS Classification**
The classification of the neonates was made retrospectively according to the infectious status (certain, probable or absent infection) by two neonatalogists blinded to the results of the PCT level. In the case of disagreement a third neonatalogist was consulted concerning the infectious status of the infant.

Infection was certain if a central sample was positive (blood culture and/or lumbar puncture). Infection was probable if the neonate had symptoms and/or had a level of CRP ≥ 10 mg/mL (30, 31), which may be associated to a positive gastric fluid culture and/or an anomaly in blood count and leukocytic formula (leukocytes < 5,000/mm³ or > 25,000/mm³ or > 25,000/mm³ or thrombocytopenia < 150,000/mm³). A neonate is symptomatic if it presented with the following: hypothermia/hyperthermia, tachycardia/ bradycardia, arterial hypotension/poor perfusion, apnea, respiratory distress, lethargy, seizures or digestive intolerance. Premature neonates showing only respiratory and/or digestive symptoms due to frequent symptoms of respiratory distress and feeding difficulties during the 1st days of life were differentiated.

**Patient Classification**
The threshold level of PCT was determined for the entire population (≪ allPCT ≫), and then for two distinct groups depending on the time of sampling for PCT analysis: at birth (≪ cordPCT ≫) and after birth during admission into neonatal intensive care (≪ dePCT ≫).
Statistical Analysis
A descriptive analysis of the characteristics of the population expressed as frequencies (percentages) for quantitative variables and medians (minimal-maximal values) for quantitative variables is provided. The non-parametric Wilcoxon and the Fisher exact tests were used to compare the characteristics of the population and the level of PCT (patients infected and not infected in the groups <allPCT>, <cordPCT> and <delPCT>). The Receiver Operating Characteristics (ROC) curves were calculated to determine the positive threshold value for the different populations and their sensitivity, specificity, and positive and negative predictive values. The areas under the curve (AUC) were compared using the method of DeLong et al. (32). The level of significance was set at 5% (a result with a p-value below 5% was considered as statistically significative). The statistics and the ROC curves were obtained using R software version 3.6.0.

RESULTS
Description of the Population
Five hundred and eleven premature neonates <32 weeks were taken into care in type III centers, Nice and Paris (Bicêtre) University Hospitals, between 1 May 2016 and 30 April 2018. Samples for PCT were obtained from 273 neonates presenting with infection risk factor(s), 13 were excluded because of the lack of data concerning the infectious status (Figure 1). Two hundred and sixty neonates were included into the statistical analysis; 173 samples for PCT were obtained at birth (group <cordPCT>) and 87 in the first 6 h of life (groupe <delPCT>). Among the 260 neonates, 26 had an infection (18 probable and eight certain infections). The clinical characteristics of the included neonates are presented in Table 1.
TABLE 1 | Baseline characteristics of patients and their mothers in the “allPCT,” “cordPCT” and “delPCT” groups (the statistical analysis compared the “cordPCT” and “delPCT” groups).

|                                | < allPCT > | < cordPCT > | < delPCT > | p-value |
|--------------------------------|------------|-------------|------------|---------|
| Gestational age (weeks)        |            |             |            | 0.06    |
| Median (IQR)                   | 28.9 (27; 30) | 29.1 (27; 31) | 28.6 (27.1; 29.9) |          |
| Min; max                       | 23.9; 31.9 | 24.4; 31.9 | 23.9; 31.9 |          |
| Male sex, No. (%)              | 146 (56.2) | 97 (56.1)  | 49 (56.3)  | 1.00    |
| Birth weight (g)               |            |             |            | 0.04    |
| Median (IQR)                   | 1200 (970; 1450) | 1240 (970; 1470) | 1130 (855; 1355) |          |
| Min; max                       | 520; 2240 | 540; 2240 | 520; 1820 |          |
| Antenatal glucocorticoids exposure, No. (%) | 245 (94.2) | 165 (85.4) | 80 (92) | 0.27 |
| Magnesium sulfate, No. (%)     | 210 (80.8) | 145 (83.8) | 65 (74.7) | 0.13 |
| Premature rupture of membranes, N (%) | 170 (65.4) | 119 (68.8) | 51 (58.6) | 0.13 |
| Prolonged rupture of membranes (≥ 12 h), No. (%) | 143 (55) | 102 (59) | 41 (47.1) | 0.13 |
| Cesarean, No. (%)              | 105 (40.4) | 61 (35.3)  | 44 (50.6)  | 0.02 |
| Clinical chorioamnionitis, No. (%) | 128 (49.2) | 86 (49.7) | 42 (48.3) | 0.90 |
| Prenatal antibiotic exposure, No. (%) | 203 (78.1) | 147 (85) | 56 (64.4) | <0.001 |
| Pre eclampsia, No. (%)         | 3 (1.2)    | 2 (1.2)    | 1 (1.1)    | 1.00 |
| Gestational diabetes, No. (%)  | 32 (12.3)  | 21 (12.1)  | 11 (12.6)  | 1.00 |
| Intrauterine growth retardation, No. (%) | 5 (1.9) | 2 (1.2) | 3 (3.4) | 0.34 |
| Threat of preterm birth, No. (%) | 221 (85) | 150 (86.7) | 71 (81.6) | 0.28 |
| Fetal rhythm abnormalities, No. (%) | 96 (36.9) | 64 (37) | 32 (36.8) | 1.00 |
| Appgar at 5 min < 7, No. (%)   | 37 (14.2)  | 21 (12.1)  | 16 (18.4)  | 0.19 |
| pH at birth                    | 7.32 (7.28; 7.32) | 7.32 (7.28; 7.37) | 7.32 (7.28; 7.38) | 0.79 |
| Min; max                       | 6.88; 7.54 | 7.05; 7.54 | 6.88; 7.49 |          |
| Multiple pregnancy, No. (%)    | 96 (36.9)  | 58 (33.5)  | 38 (43.7)  | 0.10 |
| Positive vaginal swab, No. (%) | 137 (52.7) | 94 (54.3) | 43 (49.4) | 0.41 |
| Infection, No. (%)             | 28 (10)    | 19 (11)    | 7 (8)      | 0.52 |
| Certain, No. (%)               | 8 (3.1)    | 6 (3.5)    | 2 (2.3)    | 1.00 |
| Probable, No. (%)              | 18 (6.9)   | 13 (7.5)   | 5 (5.7)    | 1.00 |
| Postnatal antibiotic exposure, No. (%) | 195 (75) | 135 (78) | 60 (69) | 0.13 |
| Death, No. (%)                 | 41 (15.8)  | 23 (13.3)  | 18 (20.7)  | 0.11 |
| Due to early-onset sepsis, No. (%) | 9 (3.5) | 6 (3.5) | 3 (3.4) | 0.48 |

IQR, Interquartile range. Bold values indicates statistically significant difference.

TABLE 2 | Pathogens of certain and probable infections in the < allPCT >, < cordPCT > and < delPCT > groups.

| Infection, No. (%) | < allPCT > | < cordPCT > | < delPCT > | p-value |
|--------------------|------------|-------------|------------|---------|
| Certain infection, No. (%) | 8 (3.1) | 6 (3.5) | 2 (2.3) |         |
| Escherichia coli, No. | 3 | 2 | 1 |         |
| Capnocytophaga spithogens, No. | 1 | 1 | 0 |         |
| Corynebacterium aurinurocosm, No. | 1 | 0 | 1 |         |
| Enterobacter cloacae, No. | 1 | 1 | 0 |         |
| Enterococcus faecalis, No. | 1 | 1 | 0 |         |
| Streptococcus mitis, No. | 1 | 1 | 0 |         |
| Probable infection, No. (%) | 18 (6.9) | 13 (7.5) | 5 (5.7) |         |
| Escherichia coli, No. | 10 | 6 | 4 |         |
| Streptococcus agalactiae, No. | 1 | 1 | 0 |         |
| Streptococcus mitis, No. | 1 | 1 | 0 |         |
| Others Streptococcus, No. | 1 | 1 | 0 |         |
| Candida albicans, No. | 1 | 1 | 0 |         |
| Citrobacter koseri, No. | 1 | 1 | 0 |         |
| Enterococcus faecalis, No. | 1 | 1 | 0 |         |
| Negative, No. | 2 | 1 | 1 |         |

Bacteria of certain infections were detected in hemocultures or cerebrospinal fluid cultures. Pathogens of probable infections were detected in newborns (gastric fluid) and/or in their mothers (vaginal swab, placenta, amniotic fluid, endocol, urine).

PCT and EOS (“allPCT” Group)

The median PCT of infected neonates was significantly higher compared to non-infected neonates (3.45 vs. 0.33 ng/mL, $p < 0.001$). Escherichia coli was the pathogen detected in the majority of the certain and probable infections (Table 2).

Analysis of the ROC curve gave a positive threshold value of PCT of 0.98 ng/mL; sensitivity of 0.65 (IC 95% 0.50–0.85), specificity of 0.90 (IC 95% 0.57–0.97), predictive positive value of 0.42 (IC 95% 0.17–0.67) and negative predictive value of 0.96 (IC 95% 0.94–0.98) (Table 3).

Cord Blood PCT and EOS (“cordPCT” Group)

Among the 260 samples obtained for PCT evaluation, 173 were from umbilical cord blood. Nineteen neonates in group < cordPCT > presented a probable or certain EOS. The average PCT of the umbilical cord of infected neonates was significantly higher compared to non-infected neonates (3.02 vs. 0.32 ng/mL, $p < 0.001$).

The threshold positive PCT level of the umbilical cord was 1 ng/mL; sensitivity of 0.63 (IC 95% 0.47–0.84), specificity of 0.89 (IC 95% 0.5–0.98), predictive positive value of 0.42 (IC 95% 0.15–0.82) and predictive negative value of 0.95 (IC 95% 0.93–0.98) (Table 3).
TABLE 3 | Diagnostic values for procalcitonin (PCT) in the “allPCT” vs. “cordPCT” vs. “delPCT” groups.

| Threshold (ng/mL) | “allPCT” | “cordPCT” | “delPCT” |
|------------------|----------|-----------|----------|
| Sensitivity (95% CI) | 0.65 (0.5–0.85) | 0.63 (0.47–0.84) | 0.86 (0.71–1) |
| Specificity (95% CI) | 0.90 (0.57–0.97) | 0.89 (0.5–0.98) | 0.93 (0.66–1) |
| Positive predictive value (95% CI) | 0.42 (0.17–0.67) | 0.42 (0.15–0.82) | 0.50 (0.21–1) |
| Negative predictive value (95% CI) | 0.96 (0.94–0.98) | 0.95 (0.93–0.98) | 0.99 (0.97–1) |

PCT Level During the 1st h of Life and EOS (“delPCT” Group)

The 87 blood samples of “delPCT” group were obtained up to 3 h and 39 min after birth. The median hour of sampling was 60 min (20–219). Seven neonates presented a probable or certain EOS. The median PCT level in the 1st h of life of the infected neonate was significantly higher compared to non-infected neonates (9.09 vs. 0.36 ng/mL, p < 0.001).

The threshold value determined from the ROC curve was 0.98 ng/mL when samples were obtained in the first 6 h of life; sensitivity of 0.86 (IC 95% 0.71–1.00), specificity of 0.93 (IC 95% 0.66–1.00), predictive positive value of 0.50 (IC 95% 0.21–1.00) and predictive negative value of 0.99 (IC 95% 0.97–1.00) (Table 3).

Comparison of the Levels of PCT in the “cordPCT” Group vs. “delPCT” Group

The threshold values of “cordPCT” group and “delPCT” group were similar (respectively, 1.00 and 0.98 ng/mL). Their diagnostic values are given in Table 3.

The AUC for PCT levels obtained during the first 6 h of life were significantly higher than those of cord blood PCT; 0.94 (IC 95% 0.85–1.00) vs. 0.75 (IC 95% 0.61–0.89; p = 0.03) (Figure 2).

DISCUSSION

The main result of this study shows that the positive threshold value for PCT in a population of <32 weeks newborns was higher than that usually reported in the literature (17). One study reported a higher basal PCT level in preterm neonates compared to term neonates irrespective of the presence of infection (25). Several explanations can be given: possible association of PCT, the precursor of calcitonin with osteogenesis and metabolism of calcium (25) and frequent respiratory distress and hypoxemia in premature neonates, which are factors that increase the secretion of PCT (26, 33). The higher level of PCT reported in our study may be explained the selection of newborn with infection risk factors: 55% had prolonged membrane rupture, 49.2% was born in context of clinical chorioamnionitis, 85% presented a threat of preterm birth and 52.7% had a positive vaginal swab. This infection and/or inflammation of the intra-uterine results in an increase in the level of pro-inflammatory cytokines, such as interleukin-6 (IL-6) in the plasma of the fetus, which defines the syndrome of fetal inflammatory response (34). This pro-inflammatory cytokine along with interleukin-1β, Tumor Necrosis Factor α and endotoxines stimulate the production of PCT by the fetus (35).

However, the specificity and negative predictive value were excellent, allowing diagnosis of non-infected neonates and thus limiting prescription of antibiotics. As for the majority of studies, the sensitivity and the positive predictive value for PCT was not as good, which does not allow its use as a positive diagnostic tool (17, 36) and can be explained by a non-specific increase in PCT within a non-infectious context of asphyxia or gestational diabetes (19).

Studies concerning PCT have selected non-homogeneous populations (gestational age, number and criteria of inclusion) making difficult the comparison of our study with those of the literature. Chiesa et al. determined the value of cord PCT at 1 ng/mL for 134 neonates of an average of 33.8 weeks (31). Joram et al. obtained a threshold value of 0.6 ng/mL for a population of 812 premature neonates presenting with infection risk factors, in which 300 were <32 weeks (36.9%) (17). A recent study found a cord blood PCT threshold value of 0.7 ng/mL in a cohort of 186 extremely preterm neonates (sensitivity of 69% and specificity of 70%) and showed that premature neonates of <28 weeks may have a cord PCT threshold value higher than that used previously (24). Several studies focusing more on premature newborns remain contradictory: PCT higher, faster and longer than in the newborn at term (21) or on the contrary rather lower (22). However, our population is more premature than in these studies, which makes it original.

Further studies are required to determine the specific threshold value of premature neonates, a population at a high risk of EOS.

The physiological increase in PCT during the early days of life naturally led the majority of studies to concentrate on cord blood PCT. The urgent care of neonates in the delivery room means that the level of PCT is not always determined at birth, for example due to coagulated, insufficient or impossible to obtain cord samples. The neonatologists question the interpretation of PCT levels evaluated from samples obtained a few hours after birth due to its postnatal physiological increase. In our study the PCT threshold value in the “delPCT” group was similar to that of the “cordPCT” group (0.98 and 1.00 ng/mL, respectively). The specificity and negative predictive value of PCT in the 1st h of life remained excellent and delayed sampling at the peak plasma level during immediate postnatal infection may explain the increase in the AUC and its sensitivity. In our study samples were obtained at a maximum of 219 min of life. So it is not possible to extrapolate further to sampling between the fourth and 6th h of life. Our study is the first to our knowledge, to investigate the use in clinical practice of PCT levels sampled within 4 h of life.

This work has some limitations. Firstly the methodology used (retrospective, neonates with infection risk factors) may explain...
the higher PCT threshold values of our population compared to those of the literature and the high frequency of infection. Another limit is the low absolute number of proven infections that can modify the threshold established, which makes these results difficult to generalize for the moment. The low number of extreme premature newborns (<28 GA) also limited us in the interpretation of the results of this subgroup.

To confirm our findings, it is necessary to consider longitudinal prospective studies in order to define, as is the case in the surveillance of neonatal jaundice, thresholds according to the term of birth.

In conclusion, this study reports a threshold value of PCT that seems to be higher in a population of premature neonates of <32 weeks compared to full term neonates. It shows that the PCT level evaluated during the first 4 h of life was an appropriate and reliable tool in the screening for EOS.

What is already known on this topic?

- The PCT cord test is widely used in the decision algorithm for the management of neonatal early onset sepsis
- Neonatal early onset sepsis remain responsible for significant morbidity and mortality in newborns, especially premature infants

- Limiting neonatal antibiotics exposure requires effective and safe screening strategies

What this study adds?

- A better definition of the positivity threshold of the PCT assay in the very preterm newborn
- The possibility of using and interpreting the dosage of PCT not taken from the cord, up to 4 h after birth
- The need to continue the studies in a broader way to refine the interpretation according to the term of birth.

DATA AVAILABILITY STATEMENT

The original contributions generated for this study are included in the article/supplementary material, further inquiries can be directed to the corresponding author/s.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by a favorable opinion from the ethics committee (CERNI): Approval number: 2020-67. Written
informed consent for participation was not provided by the participants’ legal guardians/next of kin because: Retrospective data study.

**AUTHOR CONTRIBUTIONS**

BB had contributed to the conception, the design of the work, the acquisition, analysis, interpretation of data, and drafted the work.

**REFERENCES**

1. Stoll BJ, Gordon T, Korones SB, Shankaran S, Tyson JE, Bauer CR, et al. Early-onset sepsis in very low birth weight neonates: a report from the National Institute of Child Health and Human Development Neonatal Research Network. J Pediatr. (1996) 129:72–80. doi: 10.1016/S0022-3476(96)70192-0

2. Simonsen KA, Anderson-Berry AL, Delair SF, Davies HD. Early-onset neonatal sepsis. Clin Microbiol Rev. (2014) 27:21–47. doi: 10.1128/CMR.00031-13

3. Hordijk CP, Fort P, Clark RH, Watt K, Benjamin DK Jr, Smith PB, et al. Early and late onset sepsis in very-low-birth-weight infants from a large group of neonatal intensive care units. Early Hum Dev. (2012) 88(Suppl. 2):S69–74. doi: 10.1016/j.ijedy.2012.07.019

4. Sharma AA, Jen R, Butler A, Lavoie PM. The developing human preterm neonatal immune system: a case for more research in this area. Clin Immunol Orlando Fl. (2012) 145:61–8. doi: 10.1016/j.clim.2012.08.006

5. Cottineau M, Launay E, Branger B, Caillon J, Muller JB, Boscher S, et al. Diagnostic value of suspicion criteria for early-onset neonatal bacterial infection: report ten years after the Anaes recommendations. Arch Pediatr Organes Off Soc Francaise Pediatr. (2014) 21:187–93. doi: 10.1016/j.arped.2013.11.011

6. Shane AL, Sánchez PJ, Stoll BJ. Neonatal sepsis. Lancet. (2017) 390:1770–80. doi: 10.1016/S0140-6736(17)31002-4

7. Buttery JP. Blood cultures in newborns and children: optimising an everyday test. Arch Dis Child Fetal Neonatal Ed. (2002) 87:F25–8. doi: 10.1136/fn.87.1.F25

8. Flannery DD, Ross RK, Mukhopadhyay S, Tribble AC, Puopolo KM, Gerber JS. Temporal trends and center variation in early antibiotic use among premature infants. JAMA Netw Open. (2018) 1:e180164. doi: 10.1001/jamanetworkopen.2018.0164

9. Ramasethu J, Kawakita T. Antibiotic stewardship in perinatal and neonatal care. Semin Fetal Neonatal Med. (2017) 22:278–83. doi: 10.1016/j.siny.2017.07.001

10. Greenwood C, Morrow AL, Lagomarcino AL, Altaye M, Taft DH, Yu Z, et al. Early empiric antibiotic use in preterm infants is associated with lower bacterial diversity and higher relative abundance of Enterobacter. J Pediatr. (2014) 165:23–9. doi: 10.1016/j.jpeds.2014.01.010

11. Madan JC, Salari RC, Saxena D, Davidson L, O’Toole GA, Moore JH, et al. Gut microbial colonisation in prematute neonates predicts neonatal sepsis. Arch Dis Child Fetal Neonatal Ed. (2012) 97:F456–62. doi: 10.1136/adc.2011-301373

12. Zhu D, Xiao S, Yu J, Ai Q, He Y, Cheng C, et al. Effects of one-week empirical antibiotic therapy on the early development of gut microbiota and metabolites in preterm infants. Sci Rep. (2017) 7:8025. doi: 10.1038/s41598-017-0 8530-9

13. Kuppala VS, Meinzen-Derr J, Morrow AL, Schibler KR. Prolonged initial empirical antibiotic treatment is associated with adverse outcomes in premature infants. J Pediatr. (2011) 159:720–5. doi: 10.1016/j.jpeds.2011.05.033

14. Altenberg L, Kelsen J. Advances in gut microbiome research and relevance to pediatric diseases. J Pediatr. (2016) 178:16–24. doi: 10.1016/j.jpeds.2016.08.044

15. Mishra UK, Jacobs SE, Doyle LW, Garland SM. Newer approaches to the diagnosis of early onset neonatal sepsis. Arch Dis Child Fetal Neonatal Ed. (2006) 91:F208–12. doi: 10.1136/adc.2004.064188

16. Joram N, Boscher C, Denizot S, Loubersac V, Winer N, Roze JC, et al. Umbilical cord blood procalcitonin and C reactive protein concentrations as markers for early diagnosis of very early onset neonatal infection. Arch Dis Child Fetal Neonatal Ed. (2006) 91:F65–6. doi: 10.1136/adc.2005.074245

17. Joram N, Muller J-B, Denizot S, Orsonneau JL, Caillon J, Roze JC, et al. Umbilical cord blood procalcitonin level in early neonatal infections: a 4-year university hospital cohort study. Eur J Clin Microbiol Infect Dis. (2011) 30:1105–13. doi: 10.1007/s10099-011-1187-0

18. Sachse C, Dressler F, Henkel E. Increased serum procalcitonin in newborn infants without infection. Clin Chem. (1998) 44:1343–4. doi: 10.1093/clinchim/44.6.1343

19. Chiesa C, Panero A, Rossi N, Stucagnino M, Giusti MD, Osborn JF, et al. Reliability of procalcitonin concentrations for the diagnosis of sepsis in critically ill neonates. Clin Infect Dis. (1998) 26:664–72. doi: 10.1086/541576

20. Monneret G, Labaune JM, Isaac C, Bienvenu F, Putet G, Bienvenu J. Procalcitonin and C-reactive protein levels in neonatal infections. Acta Paediatr. (1997) 86:209–12. doi: 10.1111/j.1651-2227.1997.tb8870.x

21. Chiesa C, Natale F, Pascone R, Osborn JF, Pacifico L, Bonci E, et al. C reactive protein and procalcitonin: reference intervals for preterm and term newborns during the early neonatal period. Clin Chim Acta. (2011) 412:1053–9. doi: 10.1016/j.cca.2011.02.029

22. Turner D, Hammerman C, Rudensky B, Schlesinger Y, Goia C, Schimmel MS. Procalcitonin in preterm infants during the first few days of life: introducing an age related nomogram. Arch Dis Child Fetal Neonatal Ed. (2006) 91:F283–6. doi: 10.1136/adc.2005.085449

23. Steinberger E, Hofer N, Resch B. Cord blood procalcitonin and Interleukin-6 are highly sensitive and specific in the prediction of early-onset sepsis in preterm infants. Scand J Clin Lab Invest. (2014) 74:432–6. doi: 10.1111/scj.12654.2014.900696

24. Prerot A, Baud O, Colella M, Tahi L, Bonacorsi S, Alberti C, et al. Cord blood procalcitonin level and early-onset sepsis in extremely preterm infants. Eur J Clin Microbiol Infect Dis. (2019) 38:1651–7. doi: 10.1007/s10096-019-3593-0

25. Fukuzumi N, Osawa K, Sato I, Iwatsi S, Ishino R, Hayashi N, et al. Age-specific percentile-based reference curve of serum procalcitonin concentrations in Japanese preterm infants. Sci Rep. (2016) 6:23871. doi: 10.1038/srep23871

26. Lapillonne A, Basson E, Monneret G, Bienvenu J, Salle BL. Lack of specificity of procalcitonin for sepsis diagnosis in premature infants. Lancet Lond Engl. (1998) 351:1211–2. doi: 10.1016/S0140-6736(05)79165-0

27. Stocker M, Fontana M, El Helou S, Wegscheider K, Berger TM. Use of procalcitonin-guided decision-making to shorten antibiotic therapy in suspected neonatal early-onset sepsis: prospective randomized intervention trial. Neonatology. (2010) 97:165–74. doi: 10.1159/000241296

28. Diagnostic and traitement curatif de l’infection bactérienne précoce du nouveau-né. Arch Pédiatr. (2003) 10:489–96. doi: 10.1016/S0929-693X(03)00165-3

29. Prise en charge du nouveau-né à risque d’infection néonatale bactérienne précoce (≥ 34 SA). J Pédiatrie Puériculture. (2017) 30:284–91. doi: 10.1016/j.jpp.2017.10.002

30. Mathers NJ, Pohlandt F. Diagnostic utility of C-reactive protein in neonatal infection. Eur J Pediatr. (1987) 146:147–51. doi: 10.1007/BF02343221

31. Chiesa C, Pellegrini G, Panero A, Osborn JF, Signore F, Assumma M, et al. C-reactive protein, interleukin-6, and procalcitonin in the immediate postnatal
period: influence of illness severity, risk status, antenatal and perinatal complications, and infection. *Clin Chem.* (2003) 49:60–8. doi: 10.1373/49.1.60

32. DeLong ER, DeLong DM, Clarke-Pearson DL. Comparing the areas under two or more correlated receiver operating characteristic curves: a nonparametric approach. *Biometrics.* (1988) 44:837–45. doi: 10.2307/253195

33. Monneret G, Labaune JM, Isaac C, Bienvenu F, Putet G, Bienvenu J. Increased serum procalcitonin levels are not specific to sepsis in neonates. *Clin Infect Dis.* (1998) 27:1559–61. doi: 10.1086/517758

34. Chiesa C, Pacifico L, Natale F, Hofer N, Osborn JF, Resch B. Fetal and early neonatal interleukin-6 response. *Cytokine.* (2015) 76:1–12. doi: 10.1016/j.cyto.2015.03.015

35. Schuetz P, Albrich W, Mueller B. Procalcitonin for diagnosis of infection and guide to antibiotic decisions: past, present and future. *BMC Med.* (2011) 9:107. doi: 10.1186/1741-7015-9-107

36. Kordek A, Halasa M, Podraza W. Early detection of an early onset infection in the neonate based on measurements of procalcitonin and C-reactive protein concentrations in cord blood. *Clin Chem Lab Med.* (2008) 46:1143–8. doi: 10.1515/CCLM.2008.214

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

*Copyright © 2021 Bianco, François-Garret, Batin, Dalmasso, Casagrande, Mokhtari and Eleni Dit Trolli. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.*