Predicting environmentally suitable areas for *Anopheles superpictus* Grassi (s.l.), *Anopheles maculipennis* Meigen (s.l.) and *Anopheles sacharovi* Favre (Diptera: Culicidae) in Iran

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**Abstract**

**Background:** Malaria is an important mosquito-borne disease, transmitted to humans by *Anopheles* mosquitoes. The aim of this study was to gather all records of three main malaria vectors in Iran during the last decades, and to predict the current distribution and the environmental suitability for these species across the country.

**Methods:** All published documents on *An. superpictus* Grassi (s.l.), *An. maculipennis* Meigen (s.l.) and *An. sacharovi* Favre during 1970–2016 in Iran were obtained from different online data bases and academic libraries. A database was created in ArcMap 10.3. Ecology of these species was analyzed and the ecological niches were predicted using MaxEnt model.

**Results:** *Anopheles superpictus* (s.l.) is the most widespread malaria vector in Iran, and exists in both malaria endemic and non-endemic areas. Whereas *An. maculipennis* (s.l.) is reported from the northern and northwestern parts, *Anopheles sacharovi* is mostly found in the northwestern Iran, although there are some reports of this species in the western, southwestern and eastern parts. The area under receiver operating characteristic (ROC) curve (AUC) for training and testing data was calculated as 0.869 and 0.828, 0.939 and 0.915, and 0.921 and 0.979, for *An. superpictus* (s.l.), *An. maculipennis* (s.l.) and *An. sacharovi*, respectively. Jackknife test showed the environmental variable with highest gain in the predicting power of the model when used in isolation was annual precipitation for *An. superpictus* (s.l.) and *An. maculipennis* (s.l.), and precipitation of the driest quarter for *An. sacharovi*.

**Conclusions:** Despite this range, global warming may increase the potential risk for malaria transmission in some cleared-up areas, where these proven vectors are active. Mapping and prediction of spatial/temporal distribution of these vectors will be beneficial for decision makers to be aware of malaria transmission risk, especially in the western parts of the country.

**Keywords:** *Anopheles superpictus* (s.l.), *An. maculipennis* (s.l.), *An. sacharovi*, Ecology, Spatial distribution, Modeling, Iran

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Background

The incidence of malaria has been steadily decreasing in Iran from 16,747 to 632 during 2001–2015, with the majority of cases imported. Iran is now focused on the elimination of malaria, and the tools implemented are vector control based on long-lasting insecticide-treated nets (LLINs) and indoor residual spraying (IRS), as well as improved diagnosis and free of charge access to drugs via the health system. There is a heavy focus of vector control in the southeastern region of Iran because local transmission occurs in this area, noting that insecticide resistance to pyrethroids has been recently detected in this area [1–4]. Throughout Iran seven anopheline species are primary or secondary vectors of malaria: An. culicifacies Giles (s.l.), An. dthali Patton, An. fluviatilis James (s.l.), An. maculipennis Meigen (s.l.), An. sacharovi Favre, An. stephensi Liston and An. superpictus Grassi (s.l.) [5]. The relative importance of these vector species varies across the country, with An. stephensi, An. culicifacies (s.l.) and An. fluviatilis (s.l.) being relevant in southern Iran [6], and An. superpictus (s.l.), An. maculipennis (s.l.) and An. sacharovi being relevant across the central, northern and northwestern Iran [5]. It is noteworthy that An. sacharovi was known important vector during epidemics occurred in northwestern Iran two decades ago [7].

Anopheles superpictus (s.l.) is a Palaearctic species recorded from Afghanistan, Albania, Algeria, Armenia, Azerbaijan, Bosnia and Herzegovina, Bulgaria, Croatia, Cyprus, Egypt and France. In addition, the species is also present in Georgia, Greece, Iran, Iraq, Israel (and Gaza Strip & West Bank), Italy, Jordan, Kazakhstan, Kyrgyzstan, Lebanon, Libya, Macedonia, Pakistan, Russia, Spain, Tajikistan, Tunisia, Turkey, Turkmenistan, Uzbekistan and Yugoslavia [5, 8–13]. Anopheles superpictus (s.l.) has the most extensive geographical distribution in Iran [14–18]. This species was identified as the main vector of malaria in the central plateau of Iran, but a secondary vector in southern areas [5, 18]. Previous studies in the country found three genotypes of this species based on cytochrome c oxidase 1 (cox1) data, although there is no information concerning its distribution ranges in other countries [19]. Harbach [20] listed two species, A and B, in the complex according to internal transcribed spacer 2 (ITS2) data. Studies in Iran and other countries where this mosquito is distributed reported zoophilicity feeding behavior, but it can only feed on human in the absence of other animal baits. Human blood index of this species in Iran is reported as 11.4% [9, 21]. Under laboratory conditions, artificially infected An. superpictus (s.l.) has completed sporogonic stages of Plasmodium vivax within 11.7 days [22]. This species was found to be naturally infected by sporozoite in Iran with an infection rate of 0.65–4.7% [23]. Recent study showed a decrease in An. superpictus (s.l.) distribution within western Zagros region and an increasing trend in distribution in the southern boundary [24].

The Anopheles maculipennis Group includes seven species in Iran. There are no reliable morphological characters for distinguishing all species of this group. The most common characters to identify members of this group are egg pattern, polytene chromosome, isoenzymes and species-specific polymerase chain reaction (PCR) [25]. Indeed, in the keys, only An. sacharovi is separated from other species in Iran [26]. With a human blood index of about 5% and reported sporozoite rate of 0.33%, An. sacharovi had an important role in malaria transmission in the Caspian Sea littoral in past decades [21]. Recent studies reported this Anopheles species from Iran, especially in the northern and western parts [14, 16, 27–33]. Global warming in the past 40 years has led to changes in spatial distribution of this species group; thus, the distribution area of An. maculipennis (s.s.) is expanding at an average speed of about 30 km per year; between 2008–2009 [34].

Anopheles sacharovi is reported from coastal parts of Italy, Sardinia, Corsica, Croatia, Republic of Macedonia, Albania, Bulgaria, Romania, southern regions of the former USSR, Turkey, Lebanon, Israel, Jordan, Syria, Iraq and Iran [35]. It is the most important malaria vector in Turkey, northwest border of Iran [36]. Twenty years ago, an epidemic of malaria occurred in northwestern Iran, where An. sacharovi was identified as the vector [7]. This species is distributed from northwest to southeast of Iran, alongside the Zagros Mountains [5, 37]. A human blood index of 38.5% shows the high tendency of this Anopheles species to human hosts [7]. This species has a good potential for malaria transmission to humans, where the infected host is available.

Mapping the potential distribution of malaria vectors in Iran is an important issue, because the country is in the phase of malaria elimination. This approach has been considered by different researchers across malaria endemic/at risk areas in the world, although their methodologies were different [6, 38–40]. Modeling by means of climatic and environmental variables will help us to predict the current and future spatial distribution of targeted species. Ecological niche modeling using MaxEnt model [41] is a commonly used method for predicting the potential presence of different species including malaria vectors [6, 38]. MaxEnt uses only presence data. This is appropriate for species distribution modelling concerned with predicting areas of potential species occurrence, recognizing that absence data are rarely available or reliable. Furthermore, the predictive performance of MaxEnt is consistently competitive with the highest performing methods [41–43].

The aim of this study was to gather all records of three malaria vectors, i.e. Anopheles superpictus (s.l.), An. maculipennis (s.l.) and An. sacharovi, in Iran during the last decades, and to predict the current distribution and
environmental suitability of these species across the country.

Methods

Study area

Iran with an area of 1,648,000 km² is located in Middle East and bordered by Armenia, Azerbaijan, Caspian Sea and Turkmenistan in the north, Turkey and Iraq in the west, Persian Gulf and Oman Sea in the south, and Afghanistan and Pakistan in the east. Two main mountainous ridges in the north (Alborz) and the west (Zagros) affect the climate of Iran. Thus, seven main climates exist, i.e. arid, highly arid, absolutely arid, moderate semi-arid, slightly semi-arid, semi wet and wet (Koppen climate classification). Most areas in the country are affected and covered by three climates: arid, absolutely arid and moderate semi-arid (Fig. 1). These natural conditions and topography provide different ecological niches suitable for various plants and animals species, including mosquitoes.

Species data collection

In this study, various scientific databases were searched (Google Scholar, PubMed, SID, Ovid Medline, Web of Science, Irandoc and Magiran) using the scientific names of the targeted species, i.e. “An. superpictus”, “An. maculipennis” and “An. sacharovi”. All documents that were published during 1970–2016 were considered and reviewed. To avoid false data, one of the main inclusion criteria was to enter information which published by experts in the field of mosquito taxonomy. According to the reports, distribution data of target species were reported in different levels including village, city, county and province. Although we included the spatial distribution of these species at the county and province level, just the reported x and y coordinates were used for modeling.

Fig. 1 Iran climate map (Koppen climate classification) and its neighboring countries
Also, if a study reported a mosquito species at village level, we extracted the geographical coordinates of the village from the database of Iranian villages and used for mapping and modeling. All occurrence coordinates that used for mapping and modeling were included in Additional file 1.

**Environmental and bioclimatic data**

Species distribution models (SDMs), also known as bioclimatic envelope models, are the most widely used approach for predicting suitable habitats for different species. Bioclimatic variables are derived from the monthly temperature and rainfall values in order to generate more biologically meaningful variables. The bioclimatic variables represent annual trends, seasonality and extreme or limiting environmental factors. In this study, the bioclimatic data were downloaded from the WorldClim database. This dataset is available as individual rasters spanning the inhabited continents, presented as latitude/longitude coordinates in WGS84. All 19 bioclimatic variables with a spatial resolution of 1 km² were downloaded for the period of 1950–2000 and clipped using shapefile of Iran border as analysis mask (version1.4, http://www.worldclim.org/bioclim). Band collection statistics analysis in ArcMap that provides statistics for the multivariate analysis of a set of raster was used to find bioclimatic variables with less than 0.8 correlations. So, the included variables in the model were: Bio1, annual mean temperature (°C); Bio6, minimum temperature of coldest month (°C); Bio11, mean temperature of coldest quarter (°C); Bio12, annual precipitation (mm); Bio14, precipitation of the driest month (mm); Bio17, precipitation of the driest quarter (mm); Bio18, precipitation of warmest quarter (mm). Two environmental variables, altitude (m) and Normalized Difference Vegetation Index (NDVI), were also used for modeling. Altitude was derived from Digital Elevation Model (DEM) of Iran, whereas NDVI was acquired from June 2016 image of MODIS satellite. To run MaxEnt model it is required to all environmental raster layers have identical cell sizes and projections, so these two environmental layers were rescaled and re-projected in ArcMap 10.3 to match WorldClim layer resolution (1 km²) and projection (WGS 1984).

**Ecological niche modeling**

All selected environmental and bioclimatic variables for modeling, were converted to ASCII format for modeling in the next step by Maximum Entropy (MaxEnt) model version 3.3.3 [41, 42]. MaxEnt takes a list of species presence locations as input, often called presence-only data, as well as a set of environmental predictors such as precipitation, temperature, vegetation, etc. across a user-defined landscape that is divided into grid cells. From this landscape, MaxEnt extracts a sample of background locations that it contrasts against the presence locations. Presence is unknown at background locations [44]. Here the default MaxEnt selection of background points was kept.

The contribution of the environmental and bioclimatic variables was tested by Jackknife analysis in MaxEnt model to get alternate estimates of which variables are most important in the model [45]. All variables with no contribution (0 values) in the test were excluded from the final analysis. In MaxEnt, presences can be split into ‘training’ and ‘test’ data. Training data are used to create the predictive model; test data are used to assess model accuracy. In our study, 80% of the occurrence records was randomly selected by the model and used as training data. The remaining 20% were used as test data. Then the model output was two AUCs for training and test data, as we presented in the results.

**GIS-based analysis and mapping data**

Data records were mapped using ArcMap 10.3. Distribution points of Anopheles superpictus (s.l.), An. maculipennis (s.l.) and An. sacharovi in villages, where coordinates existed, and crossed with altitude and climate layers of Iran to find their distribution in different climates and altitudes. Prediction maps of environmental suitability for these species, produced by MaxEnt model, was then classified in ArcMap to obtain the area with a suitability of more than 60%. Then the main climatic variables including ranges, average annual temperature ± standard deviation, relative humidity and altitude were determined for this range.

**Results**

**Spatial distribution**

Of over 200 records, 159 documents had information on the fauna of Culicidae in Iran. These documents were obtained, reviewed and their results recorded in the database. Of this number, there were 72 master and PhD theses, 52 articles in English, 25 Persian language papers, 8 abstracts and two research projects. Among the 159 reviewed documents, 67, 45 and 27 had reported spatial distribution points of Anopheles superpictus (s.l.), An. maculipennis (s.l.) and An. sacharovi, respectively. In this study we considered the distribution of these species in three scales: province, county and village or city level (as points of collection). With this approach An. superpictus (s.l.) was found in 29 out of 31 provinces and 152 counties in Iran during the last five decades (Table 1). During this period this species was collected from 296 collection sites. Anopheles maculipennis (s.l.) was reported from 20 provinces, 122 counties and 163 collection sites in Iran during the study period. Finally, An. sacharovi was recorded in 18 provinces, 22 counties (Table 1) and 50 collection sites (Fig. 2).

Analysis of the three environmental variables in all recorded points of these malaria vectors in Iran showed...
that An. superpictus (s.l.) existed in an altitudinal range of 0–2716 m (mean ± standard deviation: 1124.80 ± 611.57 m) with annual mean temperature of 9.1–27.5 °C (16.7 ± 4.8 °C) and annual precipitation range of 54–940 mm (359 ± 169 mm). Anopheles maculipennis (s.l.) was also found in altitudinal range of -27–3483 m (1175 ± 752 m), annual mean temperature of 5.4–17.6 °C (13 ± 2.4 °C), and annual precipitation of 97–1496 mm (481 ± 309 mm). Finally, analysis of the environmental variables showed that An. sacharovi was found within an altitudinal range of 23–2113 m (7.9 ± 572 m), annual mean temperature of 8.8–25.4 °C (13.6 ± 3.4 °C), and annual precipitation range of 130–532 mm (349 ± 88 mm) in Iran.

Considering the climatic condition in the recorded points for these Anopheles species, An. superpictus (s.l.) was mostly found in moderate semi-arid (33%) and highly semi-arid (28%) climates whereas more than 44% of the occurrence records of An. maculipennis (s.l.) were in wet and semi-wet climates. However, no records of these species were reported from arid and absolutely arid climates. Also An. sacharovi collected mainly in moderate semi-arid climate (64.3%), while no record of its existence was found in wet and absolutely arid climates (Table 2).

### Ecological niche modeling

#### Anopheles superpictus (s.l.)

As shown in the model output, the environmental suitability for An. superpictus (s.l.) was predicted to be present in most western parts of Iran (Fig. 3a), although there are some areas of high environmental suitability in northeastern, southwestern and southern regions as well. Warmer colors (red) show areas with higher environmental suitability for An. superpictus (s.l.); white dots show the presence locations used for training, while violet dots show test locations. The area under receiver operating characteristic (ROC) curve (AUC) for training and testing data was calculated as 0.869 and 0.828, respectively.

Table 3 shows the contributed ratio of Bio 12 was more than other variables used in modeling of this species. The results of the jackknife test of variable importance showed the environmental variable with the highest gain in the predicting power of the model when used in isolation was Bio12 (annual precipitation), which appeared to have the most useful information by itself, while NDVI showed the most decreased gain in the predicting power of the model when omitted. This showed that environmental suitability for A. maculipennis was found to be improved with increasing annual precipitation (Fig. 4b).

On the other hand, more water availability would also increase the probability of presence.

#### Anopheles sacharovi

As shown by the model output again, environmental suitability for An. sacharovi (s.l.) was predicted to be greater in north and northwestern parts of Iran (Fig. 3b), although there are some areas of high environmental suitability in northeastern part as well. Warmer colors (red) show areas with higher environmental suitability for An. maculipennis (s.l.); white dots show the presence locations used for training, while violet dots show test locations. AUC for training and testing data was calculated as 0.939 and 0.915, respectively.

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Table 3 shows the contribution ratio for the different variables used in the model, i.e. which variables matter most for the species being modeled. As shown in the table, Bio 17 had the most contribution in modeling this species. The results of the jackknife test of variable importance showed that the environmental variable with highest gain in the predicting power of the model when used in isolation was Bio17 (Precipitation of the driest
Fig. 2 Spatial distribution of Anopheles superpictus (s.l.), An. maculipennis (s.l.) and An. sacharovi in Iran at the county (left) and village scales (right), 1970–2016.
quarter). This appeared to have the most useful information by itself in predicting suitable environments for this malaria vector, but altitude has been shown to decreased gain in the predicting power of the model most when omitted. Environmental suitability for *A. sacharovi* was found to be more abundant with increasing rainfall in the driest quarter (Fig. 4c). In other words, species distribution for this *Anopheles* is influenced by precipitation in summer.

**Discussion**

Species distribution models estimate the relationship between species records at sites and the environmental and/or spatial characteristics of those sites. MaxEnt
model uses presence-only data. These data are a valuable resource and potentially can be used to model the same ecological relationships as with presence-absence data, provided that biases can be dealt with and except for the non-identifiability of prevalence [43].

The AUC value in this study was 0.869, 0.939 and 0.921 for *An. superpictus* (s.l.), *An. maculipennis* (s.l.) and *An. sacharovi*, respectively. Previous studies on MaxEnt model considered predicting AUC > 0.75 as good value for suitable niche for these species [42, 46]. This means that MaxEnt prediction in our study is very good. A recent study in Iran reported AUC values of 0.943, 0.974 and 0.956 for *Anopheles stephensi*, *An. culicifacies* (s.l.) and *An. fluviatilis* (s.l.), respectively [6]. In other countries AUC values were reported between 0.77 and 0.99 for different *Anopheles* species [47–51]. This value is affected
by the ecology of the different species, number of training points for model and the variables used for modeling. However, some researchers believe AUC is a misleading measure of the performance of predictive distribution models [52].

According to the results of the Jackknife test, precipitation had the main role in modeling all three species (Fig. 4). Bio12 (annual precipitation) was the most important variable for predicting An. superpictus (s.l.) and An. maculipennis (s.l.), and Bio17 (precipitation of the driest quarter) was the most important variables for An. sacharovi. It is clear that increasing precipitation will increase the availability of larval habitats and also relative humidity. Therefore, mosquitoes will have more places to lay their eggs, increasing the population. Also, the longevity of adults will increase in higher relative humidity of the environment. Previous studies have shown that in southern parts of Iran with warmer

| Variable | An. maculipennis (s.l.) | An. sacharovi | An. superpictus (s.l.) |
|----------|-------------------------|---------------|-----------------------|
| Altitude | 5.7                     | 20.1          | 3.9                   |
| Bio 1    | 3.3                     | 0.1           | 22                    |
| Bio 6    | 1.1                     | 0.6           | 4                     |
| Bio 11   | 10.9                    | 6.5           | 2.3                   |
| Bio 12   | 66.7                    | 12.7          | 31.3                  |
| Bio 14   | 2.1                     | 0.1           | 2.2                   |
| Bio 17   | 0.6                     | 58.5          | 2.9                   |
| Bio 18   | 8                       | 0.7           | 16                    |
| NDVI     | 1.6                     | 0.7           | 15.4                  |

Fig. 4 Result of the jackknife test of variable importance (left) and the curve of the most important variable in the model (right) for three malaria vectors in Iran. **a** Anopheles superpictus (s.l.). **b** Anopheles maculipennis (s.l.). **c** Anopheles sacharovi
weather with low precipitation, temperature was a significantly effective variable in predicting the ecological niches of malaria vectors [6]. The difference between An. superpictus (s.l.) and the reported species may be due to their ecology. This Anopheles species is found to have a tendency for high altitudes with higher rainfall and lower temperatures. Comparing southern malaria vectors, other environmental variables have been reported to be important factors in modeling some Anopheles species in other countries. Slope and land cover was relevant for An. bellator and An. marajoara in Brazil [48], altitude, stream vegetation and soil were relevant for An. gambiae and An. sergentii in Saudi Arabia [47], whereas altitude was crucial for primary malaria vectors in northern South America [46] and for Anopheles albimanus in Mesoamerica and the Caribbean Basin [50]. In our study, the most important environmental variable in the model for An. superpictus (s.l.) was NDVI, which was followed by altitude; both had lower importance rather than precipitation.

Based on our study, An. superpictus (s.l.) can be found with > 60% probability in the areas with annual mean temperature of 8.9–27.6 °C and mean annual rainfall of 423.55 mm. Study on this species in Europe showed it is distributed in areas where the range of temperature was between 14.9–23.7 °C and the mean rainfall was reported to be 165 mm [53]. Study on the effect of different larval rearing temperatures on the biology of An. superpictus (s.l.) under the laboratory condition showed highest survival rate at 27 °C, while all larvae in 15 °C cohort died before reaching the third-instar stage [54]. This means that although 89% of eggs hatched at 15 °C, this species needs more environmentally suitable temperatures to complete its life-cycle successfully. Although increasing temperature from 27 to 35°C resulted in shorter larva-adult development, from 15 to 10.82 days, higher temperatures reduced survival rate of adults/larvae from 70 to 6% [54]. For An. maculipennis (s.l.), the life-cycle from egg to adult ranged from 45 days in 13.9 °C to 14 days in 24.8 °C. These values were > 60 days in 15 °C and 11.5 days in 25 °C for An. sacharovi [55]. Considering land cover, An. superpictus (s.l.) seems to be more distributed in Marshy areas, irrigated cropland and grassland in Europe [53] with plants, clear and flowing water breeding sites [9]. Irrigated croplands, rice fields and river edges are the main artificial and natural breeding sites for this species in Iran, therefore, it is expected that more densities of this Anopheles in areas having such land covers [2, 28, 56, 57]. This is consistent with findings from other studies in Europe [53, 55], while in Tajikistan this species preferred small bodies of water with rocky bottoms in riverbeds as well as on rice fields [11]. Anopheles maculipennis (s.l.) also prefers rice fields and river bed/margins in Iran [16, 23–33]. This was also in accordance with earlier studies found in Europe [55].

With regard to altitudinal distribution, and according to the location of the occurrence records, we found An. superpictus (s.l.) to within the range between 0–2716 m above sea level in Iran. The maximum recorded elevation for this species in Tajikistan was 2800 m [12]. This value was calculated as 3483 m and 2113 m for An. maculipennis (s.l.) and An. sacharovi, respectively, in Iran. Anopheles sacharovi was collected between 353–1126 m above sea level in the Sanliurfa province of Turkey [58]. These altitudes could be considered environmental suitable areas for studies of malaria vectors. However, due to global warming it is unlikely to find these species in higher altitude in future studies.

Climate change and global warming, as well as changes in land use such as dam construction, urban development and agricultural projects, have large influence on the distribution of various species of plants and animals. Therefore, having a database for potential vectors such as mosquitoes is very important. It will greatly contribute to the study of climate change and environmental effects on future distribution of these insects. Using different scenarios of climate change and existing data in the databases, it is possible to predict the temporal and spatial distribution of different mosquito species in Iran. In addition, it will enable us determine information gaps and research need in the field of Culicidae in Iran. This approach will guarantee success of the national malaria elimination program, especially considering the climatic change and adaptation. Therefore, modeling spatial and temporal distribution of all malaria vectors is a very important and essential step for successful vector control and elimination program.

The lack of accurate registration of geographical coordinates of collection site(s), a main challenge in the entomological studies, was quite obvious in this survey. Knowing the exact coordinates of the collection sites for each mosquito species will contribute to the use of mathematical models to analyze field results. It is important to note that we have used occurrence data from 1970 to 2016 and no temporal covariates. Although bioclimatic data were an average of 1950–2000, available in worldClim dataset, the values of the co-variates may have been very different at the species occurrence sites in 1970, and this is not accounted in our analysis.

Conclusions
Malaria still remains an important mosquito-borne disease in Iran, although indigenous cases have been limited to some foci in southeastern region. Because An. superpictus (s.l.), An. maculipennis (s.l.) and An. sacharovi, the main malaria vectors, are distributed more or less in different areas of the country, there is transmission risk from imported cases, even in cleared-up foci. Global warming is changing the areas at risk of malaria transmission globally and in Iran it seems northern areas will more prone to
malaria transmission, where the studied species exists. Planning vector control strategies is highly dependent on good understanding of the vector ecology, as well as preparing risk maps for their environmental suitability. It is also dependent on monitoring of insecticide resistance in regular intervals. Finally, environmental variables have direct effects on the temporal and spatial distribution of mosquitoes and their ability for malaria transmission. Therefore, results of this study will be beneficial for decision makers to plan and apply proper vector control measures at the right time and place for malaria vector control in Iran.

Additional file

Additional file 1: Coordinates for collection sites. (DOCX 17 kb)

Abbreviations

A: Arid; AA: Absolutely arid; AUC: Area under curve; CSP: Circumsporozoite proteins; DEM: Digital elevation model; EMRO: Eastern Mediterranean Region; HSA: Highly semi-arid; IRS: Indoor residual spraying; ITNs: Insecticide-treated nets; MaxEnt: Maximum entropy; MSA: Moderate semi-arid; NDVI: Normalized difference vegetation index; PCR: Polymerase chain reaction; ROC: Receiver operator characteristic; SSA: Slightly semi-arid; SID: Scientific information database; SW: Semi-wet; W: Wet

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Availability of data and materials

The data supporting the conclusions of this article are included within the article. The raw data will not be shared, because the authors do not have permission to publish raw data.

Authors’ contributions

AAHB, MMS and HV hypothesized the study. AAHB, SAH and KP collected the manuscript. All authors read and approved the final manuscript. AAHB drafted the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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References

1. World Health Organization. World malaria report. Geneva: WHO; 2016.
2. Hanafi-Bojd AA, Vatandoost H, Oshaghi MA, Haghdoot AA, Shahi M, Sedaghat MM, et al. Entomological and epidemiological attributes for malaria transmission and implementation of vector control in southern Iran. Acta Trop. 2012;121:85–92.
3. Vatandoost H, Hanafi-Bojd AA. Indication of pyrethroid resistance in the main malaria vector, Anopheles stephensi from Iran. Asian Pac J Trop Med. 2012;5:722–6.
4. Yenyan M, Basseri HR, Hanafi-Bojd AA, Edalat H, Safiari R. Bio-ecology of malaria vectors in an endemic area, Southeast of Iran. Asian Pac J Trop Med. 2016;9:32–8.
5. Hanafi-Bojd AA, Azari-Hamidian S, Hassan V, Zabihollah C. Spatio-temporal distribution of malaria vectors (Diptera: Culicidae) across different climatic zones of Iran. Asian Pac J Trop Med. 2011;4:498–504.
6. Pakdad K, Hanafi-Bojd AA, Vatandoost H, Sedaghat MM, Raesi A, Moghaddam AS, et al. Predicting the potential distribution of main malaria vectors Anopheles stephensi, An. culicifacies s.l and An. flavimanus s.l in Iran based on maximum entropy model. Acta Trop. 2017;169:83–9.
7. Yaghoubi-Ershadi MR, Namazi J, Piazak N. Bionomics of Anopheles sacharovi in Ardabil Province, northwestern Iran during a larval control program. Acta Trop. 2001;78:207–15.
8. Rowland M, Mohammed N, Rehman H, Hewitt S, Mendis C, Ahmad M, et al. Anopheline vectors and malaria transmission in eastern Afghanistan. Trans R Soc Trop Med Hyg. 2002;96:520–6.
9. Sinka ME, Bangs NJ, Manguin S, Coetzee M, Mbogo CM, Hemingway J, et al. The dominant Anopheles vectors of human malaria in Africa, Europe and the Middle East: occurrence data, distribution maps and bionomic précis. Parasit Vectors. 2010;3:117.
10. Hantosh HA, Hassan HM, Al-fatlawy A. Mosquito species geographical distribution in Iraq 2009. J Vector Borne Dis. 2012;49:33–5.
11. Habirov Z, Kadamov O, Iskandarov F, Komilova S, Cook S, McKister E, et al. Malaria and the Anopheles mosquitoes of Tajikistan. J Vector Ecol. 2012;37:419–27.
12. Habirov Z, Manilova E, Kadamov D, Komilova S, Harbach RE. ELISA Incrimination of Anopheles superpictus and Anopheles hysucanus (Diptera: Culicidae) as vectors of Plasmodium vivax (Haemosporida: Plasmodiidae) in Tajikistan. J Med Entomol. 2013;50:1298–302.
13. Khater EI, Sowilem MA, Sallam MF, Alsharwi AM. Ecology and habitat characterization of mosquitoes in Saudi Arabia. Trop Biomed. 2013;30:409–27.
14. Azari-Hamidian S, Abai MR, Arzamani K, Bakhshi H, Karami H, Ladonni H, Harbach RE. Mosquitoes (Diptera: Culicidae) of North Khorasan Province, northeastern Iran, and the zoogeographic affinities of the Iranian and Middle Asian mosquito fauna. J Entomol. 2011;8:204–17.
15. SaghafiPour A, Abai MR, Faraminia B, Nafar R, Ladonni H, Azari-Hamidian S. Mosquito (Diptera: Culicidae) fauna of Qom Province, Iran. J Arthropod Borne Dis. 2012;6:54–61.
16. Banafshi O, Abai MR, Ladonni H, Bakhshi H, Karami H, Azari-Hamidian S. The fauna and ecology of mosquito larvae (Diptera: Culicidae) in western Iran. Turkish J Zool. 2013;37:298–307.
17. Khoshdel-Nezamiahi F, Vatandoost H, Oshaghi MA, Azari-Hamidian S, Arabi Mianroodi R, Dabiri F, et al. Molecular characterization of mosquitoes (Diptera: Culicidae) in northwestern Iran by using RNA-ITS2. Jpn J Infect Dis. 2016;69:319–22.
18. Sedaghat MM, Harbach RE. An annotated checklist of the Anopheles mosquitoes (Diptera: Culicidae) in Iran. J Vector Ecol. 2005;30:272–6.
19. Oshaghi MA, Yaghobi-Ershadi MR, Shemshad K, Pedram M, Amani H. The Anopheles superpictus complex: introduction of a new malaria vector complex in Iran. Bull Soc Pathol Exot. 2008;101:29–34.
20. Harbach RE. The phylogeny and classification of Anopheles. In: Manguin S, editor. Anopheles mosquitoes - new insights into malaria vectors. Rijeka: InTech; 2013. p. 1–55.
21. Edrisian GH, Manouchehry AV, Hafizi A. Application of an enzyme-linked immunosorbent assay (ELISA) for determination of the human
blood index in anopheline mosquitoes collected in Iran. J Am Mosq Control Assoc. 1985;1:349–52.

22. Kasap H. Comparison of experimental infectivity and development of Plasmodium vivax in Anopheles sacharovi and An. superpictus in Turkey. Am J Trop Med Hyg. 1990;42:111–7.

23. Faghhi MA. Malariaology and malaria eradication. Tehran: Tehran University Press; 1969 (In Persian).

24. Salahí-Moghaddam A, Khooshdel A, Dalbei H, Pakkad K, Nutafqa GG, Sedaghat MM. Spatial changes in the distribution of malaria vectors during the past 5 decades in Iran. Acta Trop. 2017;166:45–53.

25. Azari-Hamidian S. Checklist of Iranian mosquitoes (Diptera: Culicidae). J Vect Ecol. 2007;32:235–42.

26. Azari-Hamidian S, Habach RE. Keys to the adult females and fourth-instar larvae of the mosquitoes of Iran (Diptera: Culicidae). Zootaxa. 2009;2078:1–33.

27. Azari-Hamidian S. Larval habitat characteristics of the genus Anopheles (Diptera: Culicidae) and a checklist of mosquitoes in Guilan province, northern Iran. J Anthropol Borne Dis. 2011;5:37–53.

28. Amani H, Yaghoobi-Ershadi MR, Kassiri H. The ecology and larval habitats characteristics of anopheline mosquitoes (Diptera: Culicidae) in Aligudarz County (Luristan Province, western Iran). Asian Pac J Trop Biomed. 2014;4:233–41.

29. Lodonnì H, Azari-Hamidian S, Alizadeh M, Abai MR, Bakhshi H. The fauna, habitats, and affinity indices of mosquito larvae (Diptera: Culicidae) in central Iran. NW J Zool. 2015;11:76–85.

30. Nikooak SH, Moosa-Kazemi SH, Yaghoobi-Ershadi MR, Vandatoost H, Oshaghi MA, Azai A, et al. Fauna and larval habitat characteristics of mosquitoes in Neka County, northern Iran. J Anthropol Borne Dis. 2015;9:233–66.

31. Nikooak SH, Fazeli-Dinar M, Azari-Hamidian S, Mousavinbas SN, Alanji M, Ziapour SP, et al. Correlation between mosquito larval density and their habitat physicochemical characteristics in Mazandaran Province, northern Iran. PLOS Negl Trop Dis. 2017;11:e0005935.

32. Sofizadeh A, Edatal H, Abai MR, Hanafi-Bojd AA. FAUNA and some biological characteristics of Anopheles mosquitoes (Diptera: Culicidae) in Kalahe County, Golestan Province, northeast of Iran. Asian Pac J Trop Biomed. 2016;6:730–4.

33. Sofizadeh A, Moosa-Kazemi SH, Dehghani H. Larval habitats characteristics of mosquitoes (Diptera: Culicidae) in North-East of Iran. J Vector Ecol. 2007;32:235–42.

34. Novikov YM, Vaulin OV. Expansion of Anopheles maculipennis s.s. (Diptera: Culicidae) to northeastern Europe and northwestern Asia causes and consequences. Parasit Vectors. 2014;7:389.

35. Becker N, Petric D, Zgomba M, Boase C, Madon M, Dahl C, et al. Mosquitoes and their control. 2nd ed. Berlin: Springer Verlag; 2010.

36. Alten B, Caglar SS, Simsek FM, Kaynas S. Effect of insecticide-treated bednets for malaria control in Southeast Anatolia - Turkey. J Vector Ecol. 2003;28:97–107.

37. Barghamadi Z, Moosa-Kazemi SH, Pirmohammadi M. Mosquito species diversity (Diptera: Culicidae) and larval habitat characteristics in Kohgiluye a Boyerahmad Province of Iran, 2012. Armaghane Danesh J. 2014;19:67–77 (In Persian).

38. Kulkarni MA, Desrochers RE, Kerr JT. High resolution niche models of malaria vectors in northern Tanzania: a new capacity to predict malaria risk? PLoS One. 2010;5:e9395.

39. Drake JM, Beier JC. Ecological niche and potential distribution of Anopheles arabiensis in Africa in 2050. Malar J. 2014;13:213.

40. Almeida OP, Montalvo PR, Ortiz WM, Toulkeridis T. Modeling of the ecology and distribution of the genus Anopheles in Africa in 2050. Malar J. 2014;13:213.

41. Alimi TO, Fuller DO, Qualls WA, Herrera SV, Arevalo-Herrera M, Quinones ML, et al. Predicting potential ranges of primary malaria vectors and malaria in northern South America based on projected changes in climate, land cover and human population. Parasit Vectors. 2015;8:4311.

42. Al Ahmed AM, Naeem M, Kheir SM, Sallam MF. Ecological distribution modeling of two malaria mosquito vectors using geographical information system in Al-Baha Province, Kingdom of Saudi Arabia. Pakistan J Zool. 2015;47:797–806.

43. Laporta GZ, Ramos DG, Ribeiro MC, Sallam MA. Habitat suitability of Anopheles vector species and association with human malaria in the Atlantic Forest in south-eastern Brazil. Mem Inst Oswaldo Cruz. 2011;106(Suppl. 1):239–46.

44. Ma A, Wang JF, Wang DQ, Ren ZP. Prediction of potential distribution of Anopheles sinensis in China based on MaxEnt. Chin J Vector Biol Control. 2014;25:393–8.

45. Fuller DO, Alimi T, Herrera S, Beier JC, Quiñones ML. Spatial association between malaria vector species richness and malaria in Colombia. Acta Trop. 2016;158:197–200.

46. Ren Z, Wang D, Ma A, Hwang J, Bennett A, Sturrock HJW, et al. Predicting malaria vector distribution under climate changes scenarios China: challenges for malaria elimination. Sci Rep. 2016;6:20694.

47. Lobo JM, Jiménez-Valverde A, Real R. A misleading measure of the performance of predictive distribution models. Glob Ecol Biogeogr. 2008;17:145–51.

48. Gaardbo Kuhn K, Campbell-Lendrum DH, Davies CR. A continental risk map for malaria mosquito (Diptera: Culicidae) vectors in Europe. J Med Entomol. 2002;39:621–30.

49. Aytekin S, Aytekin AM, Alten B. Effect of different larval rearing temperatures on the productivity (Ro) and morphology of the malaria vector Anopheles superpictus Grassi (Diptera: Culicidae) using geometric morphometrics. J Vector Ecol. 2009;34:42–42.

50. Setten TH, Takken W. Anophelism without malaria in Europe: a review of the ecology and distribution of the genus Anopheles in Europe. Wageningen: Wageningen Agricultural University; 1994.

51. Soleimani-Ahmadi M, Vandatoost H, Zare M, Turki H, Alizadeh A. Topographical distribution of anopheine mosquitoes in an area under elimination programme in the south of Iran. Malar J. 2015;14:262.

52. Soleimani-Ahmadi M, Vandatoost H, Hanafi-Bojd AA, Zare M, Safari R, Mojahedi A, et al. Environmental characteristics of anopheine mosquito larval habitats in a malaria endemic area in Iran. Asian Pac J Trop Med. 2013;6:510–5.

53. Yurttas H, Alten B. Geographic differentiation of life table attributes among Anopheles sacharovi (Diptera: Culicidae) populations in Turkey. J Vector Ecol. 2006;31:275–84.