Hereditary Fructose Intolerance Diagnosed in Adulthood

Min Soo Kim¹, Jin Soo Moon¹, Man Jin Kim²,³, Moon-Woo Seong², Sung Sup Park², and Jae Sung Ko¹

¹Department of Pediatrics, Seoul National University College of Medicine, ²Department of Laboratory Medicine and ³Rare Disease Center, Seoul National University Hospital, Seoul National University College of Medicine, Seoul, Korea

INTRODUCTION

Hereditary fructose intolerance (HFI; OMIM# 229600) is a rare autosomal recessive disorder caused by a deficiency in aldolase B, usually diagnosed in childhood.¹,² After intake, fructose is phosphorylated to fructose 1-phosphate (F 1-P). F 1-P is then catabolized by aldolase B.³ In HFI patients, F 1-P accumulates and inhibits gluconeogenesis and glycogenolysis. Early manifestations of fructose intake include nausea, vomiting, abdominal pain, hypoglycemia, elevated liver enzymes, and faintness. Chronic exposure to fructose might cause failure to thrive, liver failure, renal failure, and, eventually, death.⁴ Severity depends on the timing and amount of fructose intake. Patients usually show their first symptoms in the infantile period when exposed to fructose and sucrose-containing foods.⁵ Recurrent symptoms make diagnosis possible in childhood; however, some remain undiagnosed until adulthood.⁵,⁶ Traditionally, HFI was diagnosed using a fructose tolerance test or an enzyme assay of liver or small intestine biopsy. Nowadays, diagnosis occurs through aldolase B gene analysis. Here we report a 41-year-old woman diagnosed with HFI in adulthood through gene analysis. This is the first report of a Korean HFI patient diagnosed in adulthood.

CASE REPORT

A 41-year-old woman visited an outpatient clinic due to repeated nausea and vomiting after the administration of fruits, sucrose, or fructose-containing foods. A lifelong history of aversion to sweets was revealed. She was breastfed until 12 months of age. Her mother tried giving her fruits, but she refused them. After being forced to eat fruits at the age of three, she showed symptoms of nausea, vomiting, and visual disturbance. The patient then continuously avoided sweets. In adulthood, after one sip of a beverage, nausea, vomiting, and diarrhea occurred. Two hours later,
she presented with a cold sweat and faintness. She had no family history of liver or genetic disease. Other family members were asymptomatic. Her physical examination was normal; no hepatomegaly or splenomegaly was found. Laboratory findings showed no abnormality as well. Her white blood cell count was 8,460/mm$^3$ (neutrophil 64%, lymphocytes 30.5%, monocyte 4.8%, eosinophil 0.5%, and basophil 0.2%), hemoglobin 14.1 g/dL, platelet 261,000 /mm$^3$, calcium 8.8 mg/dL, phosphorus 3.5 mg/dL, glucose 96 mg/dL, uric acid 3.0 mg/dL, total cholesterol 180 mg/dL, total protein 7.5 g/dL, albumin 4.5 g/dL, total bilirubin 0.7 mg/dL, alkaline phosphatase 61 IU/L, aspartate aminotransferase 19 IU/L, alanine aminotransferase 16 IU/L, hemoglobin 14.1 g/dL, platelet 261,000 /mm$^3$. Her height was 160 cm (50th percentile), and her weight was 50.2 kg (25th to 50th percentile). Her physical examination was normal; no hepatomegaly or splenomegaly was found. Laboratory findings showed no abnormality as well. Her white blood cell count was 8,460/mm$^3$ (neutrophil 64%, lymphocytes 30.5%, monocyte 4.8%, eosinophil 0.5%, and basophil 0.2%), hemoglobin 14.1 g/dL, platelet 261,000 /mm$^3$, calcium 8.8 mg/dL, phosphorus 3.5 mg/dL, glucose 96 mg/dL, uric acid 3.0 mg/dL, total cholesterol 180 mg/dL, total protein 7.5 g/dL, albumin 4.5 g/dL, total bilirubin 0.7 mg/dL, alkaline phosphatase 61 IU/L, aspartate aminotransferase 19 IU/L, alanine aminotransferase 16 IU/L, hemoglobin 14.1 g/dL, platelet 261,000 /mm$^3$. Through her history, HFI was suspected, and gene analysis of aldolase B was completed. Genomic DNA was extracted, and the Agilent SureSelectXT Human all Exon 50 Mb kit was used to target the exon regions. These targeted regions were sequenced using the Illumina HiSeq sequencing system with 100 bp paired-end reads. All sequence variants were confirmed by Sanger sequence analysis. The patient showed known heterozygous pathogenic nonsense (c.178C>T, p.Arg60Ter) variant and known pathogenic frameshift (c.360_363delCAAA, p.Asn120LysfsTer32) variant. She began a fructose-restricted diet after counseling with a dietician. HFI has an estimated incidence of 1:18,000 to 1:31,000. HFI is caused by a mutation in aldolase B, which results in the accumulation of F 1-P. HFI often manifests in young infants when fructose or sucrose-containing foods are first introduced after breastfeeding. Symptoms usually manifest as nausea, vomiting, and aversion to fructose-containing foods. Prolonged fructose ingestion may cause liver and renal failure; physical examination might reveal hepatomegaly and jaundice; the main consequence is depletion of phosphate due to the phosphorylation of fructose. As a result, patients show hypophosphatemia, hyperuricemia, hypermagnesemia, hypoglycemia, and acidosis after fructose loading. Lack of phosphate interrupts all cellular processes requiring phosphorylation or adenosine triphosphate, including glycogenolysis and gluconeogenesis. This explains why the administration of glucagon does not correct hypoglycemia (Fig. 1). Patients voluntarily avoid sweet foods. When fructose is restricted after the infantile period, HFI may remain undiagnosed until adulthood. In the 1970s, undiagnosed HFI patients were exposed to the danger of using fructose or sorbitol as a source of parenteral nutrition; several fatal cases were reported. Fructose and sorbitol infusions are no longer used. Nowadays, glucose and lipid solutions are preferred sources of energy.
as a possible diagnosis in a patient showing unpleasant symptoms after sugar loading. Symptoms may vary depending on the age of exposure and intensity of fructose intake, possibly as severe as acute liver failure. Cases of acute liver failure in neonates were reported when exposed to sucrose-containing formula; recurrent episodes of hepatitis have been reported in infancy. In adults, patients show a lifelong history of avoiding sweet fruits and nausea after small amounts of sweets. Therefore, thorough history taking is critical for HFI diagnosis. Formerly HFI was diagnosed through activity assay of aldolase B in tissue biopsy specimens or fructose tolerance test. Biopsy techniques have risks of pain, bleeding, and complications of sedation. Fructose tolerance tests induce acute symptoms of hypoglycemia, nausea, and vomiting. These invasive tests are unnecessary since genetic testing for HFI is safer, and should be frequently and promptly performed.

There have been two HFI Korean childhood cases; this is the first report of a Korean adult patient. One case was diagnosed through an enzyme assay of an intestinal and liver biopsy in 2002. The other case was diagnosed through genetic testing, which showed a c.758_759insT (p.V253fsX24) homozygote in 2012. More than 50 mutations are known to cause HFI, but frequencies of mutant alleles in East Asia are not well known. c.448G>C (p.Ala150Pro) is the most common mutation worldwide, other common mutations are c.524C>A (p.Ala175Asp), c.1005C>G (p.Asn335Lys), c.360_363delCAAA (p.Asn120LysfsTer32), and c.178C>T (p.Arg60Ter).

Interestingly, there is no report on the p.Ala150Pro mutation in Asian HFI patients. In Asia, three HFI cases diagnosed with genetic testing were reported. Besides one Korean report of novel frameshift mutation, the other two cases were reported in Japan and China, respectively. The Japanese patient was homozygote for a nonsense mutation c.720C>T (p.Cys240Ter) from consanguineous parents. The Chinese patient, whose parents were cousins, was homozygote for a frameshift mutation c.479_482delAACA (p.Arg60Ter).

These mutations are uncommon in Western countries. In Asia, few HFI cases were reported, genetic testing was rare, and no study was conducted on the frequency of HFI. Two mutations found in our patient, c.360_363delCAAAA and c.178C>T, are the third and fourth most common HFI mutations in America, respectively, and are also widespread in Europe. c.360_363delCAAAA and c.178C>T mutations both create premature stop codons, resulting in a truncated protein with deteriorated function. Frameshift and nonsense mutations cause more severe changes in protein structure than missense mutations. However, recent studies show that phenotypes do not differ from genotypes. Several patients are reported to have two null alleles without an exhibition of severe symptoms or unusual phenotypes. In this case, the patient also showed typical symptoms of classic HFI and excellent prognosis after fructose restriction. It implies that harboring two null alleles does not lead to a severe phenotype.

After the diagnosis of HFI has been made, an expert dietician should develop a strict restrictive diet. Careful advice on medication should also be provided since sucrose and sorbitol are common components in syrups and tablets. Recently, higher intrahepatic triglyceride content was reported in HFI patients than healthy control groups, even with a fructose-restricted diet. Nonalcoholic fatty liver disease is also a potential threat in HFI; regular follow-up is needed. Prognosis is excellent if patients maintain a strict exclusion diet. Therefore, repeated education is essential for HFI patients.

HFI is a rare, unfamiliar, and underdiagnosed disorder in Korea. HFI should be considered in individuals with characteristic discomforts and clinical manifestations following exposure to fructose, sucrose, or sorbitol. Knowing the symptoms and pathophysiology of HFI could alter the clinical course of HFI patients. Simply restricting fructose could dramatically change the patient’s outcome. Therefore, we report the first adult case of a Korean HFI patient to facilitate early recognition and treatment. Further studies will provide opportunities to determine the incidence of HFI in the Korean population.

CONFLICTS OF INTEREST

No potential conflict of interest relevant to this article was reported.

AUTHOR CONTRIBUTIONS

Data analysis and interpretation: M.J.K., M.W.S., S.S.P. Data acquisition, drafting of the manuscript, study concept and design: M.S.K. Administrative, technical, and material support: J.S.M. Study concept and design, critical revision of the manuscript for important intellectual content, and study supervision: J.S.K.

ORCID

Min Soo Kim https://orcid.org/0000-0001-7244-2359
Jin Soo Moon https://orcid.org/0000-0001-9760-297X
Man Jin Kim https://orcid.org/0000-0002-9345-6976
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