Accurate Inference of Tree Topologies from Multiple Sequence Alignments Using Deep Learning

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Abstract.—Reconstructing the phylogenetic relationships between species is one of the most formidable tasks in evolutionary biology. Multiple methods exist to reconstruct phylogenetic trees, each with their own strengths and weaknesses. Both simulation and empirical studies have identified several “zones” of parameter space where accuracy of some methods can plummet, even for four-taxon trees. Further, some methods can have undesirable statistical properties such as statistical inconsistency and/or the tendency to be positively misleading (i.e. assert strong support for the incorrect tree topology).

Recently, deep learning techniques have made inroads on a number of both new and longstanding problems in biological research. In this study, we designed a deep convolutional neural network (CNN) to infer quartet topologies from multiple sequence alignments. This CNN can readily be trained to make inferences using both gapped and ungapped data. We show that our approach is highly accurate on simulated data, often outperforming traditional methods, and is remarkably robust to bias-inducing regions of parameter space such as the Felsenstein zone and the Farris zone. We also demonstrate that the confidence scores produced by our CNN can more accurately assess support for the chosen topology than bootstrap and posterior probability scores from traditional methods. Although numerous practical challenges remain, these findings suggest that the deep learning approaches such as ours have the potential to produce more accurate phylogenetic inferences.

[Supervised machine learning, convolutional neuronal network, phylogenetics.

Reconstructing the historical relationships among biological sequences remains one of the most important challenges in evolutionary biology. To reconstruct a tree, it is first necessary to acquire orthologous (Koonin 2005) and/or paralogous (Hohl and Ragan 2007; Bonham-Carter et al. 2014) sequences from a sample of species or populations. Generally, the next step involves producing a multiple sequence alignment (MSA) to compare orthologous sequence characters, although in recent years a number of methods that bypass explicit orthology and alignment searches have been proposed (Hohli and Ragan 2007; Bonham-Carter et al. 2014). Finally, any one of a plethora of tree reconstruction methods can be employed to infer a tree from an MSA. Determining the topology of a tree has been shown to be NP-hard for at least some formulations of the problem (Roch 2006), and the space of possible solutions grows double-factorially with increasing numbers of sequences (Felsenstein 1978b). We are thus limited to methods that utilize various heuristics and/or make assumptions about the process of sequence evolution to efficiently traverse tree space, including maximum parsimony (MP; Farris 1970; Fitch 1971), distance-based approaches such as neighbor-joining (NJ; Saitou and Nei 1987), maximum likelihood (ML; Felsenstein 1981), and Bayesian inference (B: Rannala and Yang 1996; Li et al. 2000; Huelsenbeck and Ronquist 2001). Although none of these methods are guaranteed to produce the true tree topology, one can evaluate and compare their effectiveness.

Supervised machine learning algorithms represent an alternative framework for drawing inferences from data, including sequence data. These methods have been successfully applied to various areas of biological research (Tarca et al. 2007), and have recently been introduced to evolutionary biology—population genetics in particular (reviewed in Schrider and Kern 2018). In brief, given some multivariate observation \( x \) that is associated with a response variable \( y \), supervised machine learning seeks to create a function \( f(x) \) that predicts \( y \). This is achieved through a process called training, wherein a “training set” of observations with known response variables are examined by an algorithm that creates and tunes the function \( f \) to minimize the disparity between \( f(x) \) and \( y \) on this set. Typically, \( x \) is summarized by a vector of values, or features. Convolutional neural networks (CNNs; Lecun et al. 1998), a class of deep learning algorithms (Goodfellow et al. 2016), are able to learn to extract informative features from data arranged in a matrix or higher-dimensional tensor to make a prediction; thus CNNs do not require a predefined feature vector. CNNs have proved extremely effective in analyzing image data (Krizhevsky et al. 2012) and have recently been shown to achieve impressive accuracy on a number of tasks in population genetic inference when applied to MSAs (Chan et al. 2018; Flagel et al. 2018). CNNs and other deep learning methods may thus prove to make sizeable gains in a number of inference tasks involving biological sequence data.

In this study, we propose a novel approach applying CNNs to phylogenetic inference. We show that CNNs can be trained to extract phylogenetic signal from an MSA and use it to accurately reconstruct unrooted gene tree topologies (i.e. here our \( y \) is a tree topology).
when given an alignment of four sequences. We adopt a classification approach, wherein the CNN is trained to discriminate among discrete data classes which in this case correspond to the three possible tree topologies. We find that our CNN approach is highly accurate on simulated data, can naturally include indel information, is fairly robust to classical biases affecting tree reconstruction and has several other benefits over existing methods. Below, we describe our methodology and the CNN’s performance in detail before concluding with a brief discussion of limitations, potential extensions, and outstanding practical challenges for adapting deep neural networks to phylogenomics.

**Methods**

**MSA Simulations**

For the 4-taxon case, there exist exactly three possible unrooted tree topologies: ((A,B),(C,D)), ((A,C),(B,D)), and ((A,D),(B,C)) in Newick notation. We simulated an equal number of MSAs from each of these topologies using INDELible (Fletcher and Yang 2009), which allows for the inclusion of indels. The MSA length was set to 1000 sites unless stated otherwise. To construct a training set consisting of trees with a variety of branch-length configurations we sampled uniformly from a "branch-length space" (BL-space) that encompasses a wide array of branch lengths and nodal rotations (see Supplementary Text and Fig. S1 available on Dryad at http://dx.doi.org/10.5061/dryad.ct2895s). Tree branch lengths are expressed in expected number of substitutions per site.

With the tree topology and branch lengths in hand, sequences were generated under a randomly assigned Markov model of nucleotide substitution using INDELible. We used five nested models of increasing complexity: JC (Jukes and Cantor 1969), TIM, TIMet (Posada 2003), GTR (Tavaré 1986), and UNREST (Yang 1994) accommodating rate heterogeneity along the sequence by drawing the shape parameter of a continuous gamma distribution (+I) from $U(0,4)$. We also allowed the proportion of invariable sites ($\pi_{inv}$) in an MSA to vary, drawing this parameter (+I) from $U(0,1)$. We note that the average expected substitutions per site (i.e. the branch length) is specified for variable sites only, not for the entire MSA. For example, when simulating an MSA with $\pi_{inv}=0.5$, the specified branch lengths refer only to the 50% of sites that are free to vary whereas the remaining sites have branch lengths of zero. Substitution rate parameters and nucleotide frequencies were drawn from $U(0,3)$ and a four-category Dirichlet distribution ($\alpha = 30$), respectively. Finally, when including indels in the simulation, we used identical fixed insertion and deletion rates ($\lambda_{I} = \lambda_{D} = 0.01$) and drew indel lengths from a Zipf distribution with $\alpha = 1.5$, similar to empirical estimates (Vialle et al. 2018), and a maximum indel size of 50. For each topology class, we simulated either $5 \times 10^5$, $1.5 \times 10^6$, or $3 \times 10^6$ MSAs for training and 1.5 $\times 10^4$ MSAs for the validation and test data sets. In addition, we simulated test sets consisting of 3000 trees from each of 13 regions of the tree-shape space that may potentially cause biased or more erroneous tree topology inference, including the Farris and Felsenstein zones that have long been known to negatively affect performance of classical methods (Swofford et al. 2001).

The simulation parameters for each of these were the same as those stated above with branch lengths drawn from the distributions shown in Table 1. In addition, we simulated trees within so-called “twisted” Farris zone (McTavish et al. 2015) with asymmetrical branch lengths as well as trees from different regions that may potentially cause tree estimation to be less accurate. Throughout this article, we denote sister branches or their branch lengths on the left side of the tree as $B_1$ and $B_2$, those on the right side as $B_3$ and $B_4$, and the internal branch as $B_5$. The overall pipeline is summarized in Figure 1.

The internal branch length ($B_5$) for the Farris and Felsenstein zones was drawn from $U(0,0.05)$ (Table 1). To investigate effect of internal branch length on each method’s performance in more detail, we also created variants of the Farris and Felsenstein zones, called “extended” where $B_5$ was drawn from the interval $U(0,0.5)$. Previous simulation studies suggest that methodological biases primarily arise when an internal branch appears to be very short, i.e. internal branch length $\leq 0.05$ (Swofford et al. 2001). However, as these studies followed overly simplified simulation approaches (e.g. MSA simulation under JC model only; Siddall 1998), this conclusion may not hold for MSA data sets generated using more complex simulation strategies that incorporate various substitution models, rate heterogeneity, varying proportions of invariant sites and indels as we do in this study.

**CNN Architecture and Training**

This section describes the structure of our neural network and training procedure, and thus many of the concepts, herein, may be foreign to some readers. We refer those interested in an accessible overview of CNNs in the context of sequence alignments to Flagel et al. (2018). We implemented our CNN in Python using version 2.2.4 of the Keras API (https://keras.io/) with TensorFlow 1.12.0 (Abadi et al. 2016) as the backend. To extract phylogenetically informative features from an MSA, we used a 1D convolution-pooling strategy. We represented each MSA as a matrix with rows corresponding to sequences and columns to sites in the alignment, and with each character in the alignment encoded as a number (‘A’:0, ‘T’:1, ‘C’:2, ‘G’:3, ‘-‘:4), and encoded the MSAs’ associated topology as a numeric class label. As indels result in MSAs of varying lengths and our CNN requires all input matrices to have equal size, shorter alignments were extended by “padding” the MSA with additional columns wherein each value was arbitrarily set to -15 until the MSA’s...
length matched that of the longest alignment in our simulated data. Our CNN consisted of eight convolution layers with ReLU activation (Nair and Hinton 2010) and each followed by an average-pooling step, one 1024-node fully connected layer with ReLU activation and an output layer with three nodes corresponding to the three possible topology classes. We used softmax as the activation function for our output layer to assign a class membership probability to each of the three classes. We used categorical cross-entropy as our loss (i.e. error) function. During training we used the adaptive moment estimation (Adam) optimizer (Kingma and Ba 2014). We set the filter size for the first convolution operation to $4 \times 1$, reasoning that it would potentially capture a phylogenetic signal by striding across the MSA and examining one column of the alignment at a time. The total number of filters was set to 1024 for the first and second convolutional layer and set to 128 for the remaining convolutional layers. We decided upon 1024 filters because this number is greater than the number of possible columns in an MSA with four DNA sequences allowing gap characters, and we had decided (arbitrarily) to limit these values to powers of two. Additional network hyperparameters were chosen somewhat arbitrarily after assessing performance of

![Diagram](image-url)

**Figure 1.** Summary of the simulation and testing procedures. The top branch of the diagram represents the steps that were used to simulate data sets with various branch-length configurations (Table 1) to test each method’s performance. The bottom branch of the diagram represents the steps that were used to simulate training and validation data sets to train the CNN, and the initial test data set was used to estimate accuracy for each method. The training, validation and test data sets were simulated using the procedure described in Supplementary Text. BI = Bayesian inference; BS = bootstrap; CNN = convolutional neuronal network; CPP = class membership posterior probability; ML = maximum likelihood; MP = maximum parsimony; MSA = multiple sequence alignment; NJ = neighbor-joining; PP = posterior probabilities. The tree shows the branch labeling convention that was used throughout the article, where numbers in circles correspond to branches B1, B2, B3, B4, and B5.

**Table 1.** Branch-length distributions for test sets designed to assess accuracy and the presence of bias in topology inference

| Region                        | $B_1$ | $B_2$ | $B_3$ | $B_4$ | $B_5$ |
|-------------------------------|-------|-------|-------|-------|-------|
| Truncated exponential         | $U(0,0.05)$ | $U(0,0.05)$ | $U(0,0.05)$ | $U(0,0.05)$ | $U(0,0.05)$ |
| Farris zone                   | $U(0,0.5)$ | $U(0,0.05)$ | $U(0,0.05)$ | $U(0,0.05)$ | $U(0,0.05)$ |
| "Extended" Farris zone        | $U(0,0.5)$ | $U(0,0.05)$ | $U(0,0.05)$ | $U(0,0.05)$ | $U(0,0.05)$ |
| Felsenstein zone              | $U(0,0.5)$ | $U(0,0.05)$ | $U(0,0.05)$ | $U(0,0.05)$ | $U(0,0.05)$ |
| Long branches                 | $U(0,0.5)$ | $U(0,0.05)$ | $U(0,0.05)$ | $U(0,0.05)$ | $U(0,0.05)$ |
| Extra-long branches           | $U(0.5,1)$ | $U(0.5,1)$ | $U(0.5,1)$ | $U(0.5,1)$ | $U(0.5,1)$ |
| Single short branch           | $U(0,0.05)$ | $U(0,0.05)$ | $U(0,0.05)$ | $U(0,0.05)$ | $U(0,0.05)$ |
| Short branches                | $U(0,0.05)$ | $U(0,0.05)$ | $U(0,0.05)$ | $U(0,0.05)$ | $U(0,0.05)$ |
| Extra-short branches          | All $U(0,0.01)$ | $U(0,0.01)$ | $U(0,0.01)$ | $U(0,0.01)$ | $U(0,0.01)$ |
| BL-space                      | Sampled from $w_1 \text{Beta}(a=b=0.1) + w_2 \text{Beta}(a=b=0.5) + w_3 \text{Beta}(a=b=1)$ as described in Supplementary Text |

Notes: $B_1$ and $B_2$ and $B_3$ and $B_4$ denote sister branches on the left and right sides of a tree, respectively, whereas $B_5$ denotes the internal branch.

TExp denotes truncated exponential distribution. Beta denotes beta distribution. A mixture of beta components with equal weights (i.e. $w_1 = w_2 = w_3 = 1/3$) was used to generate the space from which samples were drawn as described in the Supplementary Text. $U$ denotes uniform distribution. $B_1 + B_5$ is a sum of two uniform random variables which follows the Irwin–Hall distribution.
several different network architectures. We also used average-pooling rather than the more common max-pooling operation given that Flagel et al. (2018) found this approach to be effective when applied to population genetic alignments. The first average-pooling operation size was set to $1 \times 1$ (i.e. no pooling), then all the subsequent convolution steps had equal sizes of $1 \times 2$ and pooling steps had equal sizes of $1 \times 4$, $1 \times 4$, $1 \times 4$, $2$, $1 \times 2$, and $1 \times 1$. We had also experimented with fewer layers but noticed that “deeper” CNN architectures, i.e. with more convolutional layers, showed better accuracy.

To avoid possible internal covariate shift and model overfitting, we used batch normalization (Ioffe and Szegedy 2015) followed by dropout regularization with a rate of 0.2 for each convolution layer as well as for the hidden layer. During training in most of the cases, we used a minibatch size of 150 MSAs. The detailed network architectures along with the training time are in Supplementary Text available on Dryad. The training procedure was run for a number of iterations determined by the stopping criterion (improvement in validation loss of the current iteration over the best prior iteration <0.001) and patience value (the number of consecutive iterations that must satisfy the stopping criterion before halting training). We observed that the results of our training runs were fairly stable, with a given CNN architecture consistently achieving the same final accuracy on the validation set. To further examine the performance during training, we visualized the surface of the loss function and training trajectories across the CNN’s parameter space (i.e. the possible values of all weights in the network) as described below. This revealed that there appear to be many readily reachable local minima of the loss function that all have roughly equivalent values (Supplementary Fig. S2 available on Dryad), lending further support to the notion that training results should be fairly stable, at least for the data sets and architectures examined here.

**Testing Procedures**

We sought to compare the performance of CNN vs. conventional tree inference methods, namely MP, NJ, ML, and BI using our simulated test data sets. MP reconstructions were performed using PHYLIP’s dnparas program v3.696 (Felsenstein 1989) in the interface provided by SeaView v4.7 (Gouy et al. 2010). ML analysis was implemented in IQ-TREE v1.6.5 (Nguyen et al. 2015) restricting model selection to only the models that were used to simulate MSAs, i.e. JC, TIM, TIMf, GTR, and UNREST (equivalent to the 12.12 model in IQ-TREE) with $+\Gamma$ and/or $+\alpha$, where the continuous rate gamma distribution was approximated by discrete gamma with eight categories (G8). For each MSA, ML search was performed exhaustively evaluating each of the three possible unrooted topologies. The NJ trees were generated using the distance function of the best-fit model in IQ-TREE. Note that ML and NJ approaches infer a rooted tree under the UNREST model, thus every such estimated tree was unrooted for further comparisons. Bayesian tree reconstruction was carried out in ExaBayes v1.4.1 (Aberer et al. 2014) under default parameters starting with a random initial topology. Three independent MCMC chains were run for $10^6$ generations each with two coupled chains. Tree support was assessed by standard nonparametric bootstrap (Felsenstein 1985) for MP, NJ, ML, and CNN, and by posterior split probability for BI. We also assessed tree support from the CNN by simply using the estimated posterior class membership probabilities emitted by the output layer. Then, for each method we calculated topological accuracy as the number of correctly returned topologies divided by the total number of inferences performed. See Supplementary Text available on Dryad for exact commands that were used to infer topologies.

**RESULTS AND DISCUSSION**

**CNNs Are More Accurate Than Standard Methods on Gapped and Ungapped MSAs within BL-Space**

Initially, we trained our CNN on gapped and ungapped training sets each comprised of $1.5 \times 10^4$ MSAs (5 $\times 10^3$ MSAs for each of our three possible topologies, with each MSA having 1000 sites prior to adding indels). Consequently, a training set generated in the same manner as our training data. We compared the accuracy of our CNN on simulated test data ($1.5 \times 10^4$ MSAs per topology) to that of several other standard tree estimation methods, namely MP, NJ, ML, and BI. Strikingly, the CNN was substantially more accurate than the other methods (McNemar test, $P < 1 \times 10^{-10}$ for each comparison with the CNN) when applied to gapped MSAs (Fig. 2). However, this was not the case when testing methods on ungapped MSAs: here accuracy of the CNN (0.765), which was in this case also trained on ungapped MSAs, was nearly identical to the best performing method, BI (accuracy = 0.765, McNemar test, $P = 0.918$). Nevertheless, the CNN’s accuracy improved with increasing amounts of training data and obtained higher accuracy (0.777; Fig. 2) than all other methods when trained on either $1.5 \times 10^5$ or $3 \times 10^5$ ungapped MSAs per topology. The difference with the next best method (BI) was not significant (McNemar test, $P = 0.08$) with $1.5 \times 10^5$ training examples but significant (McNemar test, $P = 0.013$) with $3 \times 10^5$ training examples. Although the results of this initial evaluation within our BI-space are encouraging, we must bear in mind that accuracy can vary within this space. Moreover, our BI-space was constructed in part to expose CNN to many branch-length configurations including some extreme configurations which one may not expect to commonly encounter in real data sets. Thus in the next section, we test each method on a number of more narrowly defined tree branch-length configurations to gain a more detailed picture of our method’s strengths and weaknesses.
We also investigated the relationship among several simulation parameters and inference accuracy. When sequences are very closely related, the MSA may consist largely of invariant sites, which are phylogenetically uninformative. One of the parameters of our simulations was the fraction of sites that were not allowed to vary (\(p_{\text{inv}}\); see Methods section). We found that for each method, misclassified MSAs from the gapped test set had a larger value of \(p_{\text{inv}}\) on average (Wilcoxon rank sum test, \(P < 1 \times 10^{-10}\) for each standard method and \(P = 6.135 \times 10^{-3}\) for the CNN), but this difference is far less pronounced for the CNN (Fig. 3a). However, in the case of the ungapped test set, MSAs misclassified by all methods excepting the CNN showed a similar bias toward larger values of \(p_{\text{inv}}\) on average (Wilcoxon rank sum test, \(P < 1 \times 10^{-10}\) for each method; Fig. 3b). The gamma shape parameter \(\alpha\), which governs mutation rate heterogeneity across the MSA does not impact the CNN’s ability to infer the correct topology (Wilcoxon rank sum test, \(P = 0.91\)) whereas for all other methods misclassified MSAs had elevated values of \(\alpha\) (Wilcoxon rank sum test, \(P < 1 \times 10^{-4}\) for all standard methods; Fig. 3c). Again, in the case of ungapped data each method showed a similar bias, with larger \(\alpha\) in misclassified versus correctly classified examples (Wilcoxon rank sum test, \(P < 1 \times 10^{-5}\) for all methods; Fig. 3d). Together these results suggest that CNNs are more robust to higher values of \(p_{\text{inv}}\) and \(\alpha\) than traditional methods for gapped MSAs. In addition, we note that the DNA substitution model does not affect any method’s performance (Fisher’s exact test, \(P > 0.05\) for all methods).

Overall these results suggest that our CNN can effectively extract phylogenetic signal. This is especially so for the CNN trained on gapped MSAs, which easily outperformed the other methods, all of which treat gaps as missing data. Despite their potential to provide unambiguous phylogenetic information (Rokas and Holland 2000; Dessimoz and Gil 2010) due to reduced homoplasy relative to substitutions (Ashkenazy et al. 2014), approaches incorporating indels are controversial and not generally utilized in practice (Ogden and Rosenberg 2007; Warnow 2012). When our CNN is trained on gapped data it learns to detect phylogenetic signal from both indels and substitutions present in an MSA. To further examine this property of the CNN, we simulated MSAs of length 1000 using trees with branch lengths drawn from \(U(0, 0.5)\) with no substitutions (i.e. \(p_{\text{inv}} = 1\) allowing only indels. Under these conditions, all standard methods fail to achieve consistency (Warnow 2012; Truszkowski and Goldman 2016), and MP, NJ, ML, and BI all had accuracy of \(<0.33\) or lower—no better than random guessing. In contrast, the CNN’s accuracy on this set was 0.927. To test a scenario toward the opposite extreme wherein every site is informative, we simulated MSAs of length 1000 allowing all sites to vary (\(p_{\text{inv}} = 0\)) under the JC model, including indels, and using trees with all branch lengths drawn from \(U(0, 0.5)\). Under these conditions, all methods exhibited very high accuracy (MP: 0.962, NJ: 0.976, ML: 0.975, BI: 0.978, and CNN: 0.959).

These results further suggest that our CNN, when trained on gapped MSAs, is in principle able to incorporate indel information into its phylogenetic inference. Because the CNN implicitly constructs substitution and indel process models by examining the simulated training data, alignment error could impact inference in practice. This could be handled...
in one of two ways: 1) by incorporating alignment errors into the training set (e.g. by running a sequence aligner on training data prior to training), or 2) judging the reliability of indels within the MSA and masking particular sites or omitting entire MSAs appearing to be unreliable (e.g. following Castresana 2000; Misof and Misof 2009; Ashkenazy et al. 2014; Fujimoto et al. 2016). Another potential problem is model misspecification: accuracy could suffer if indel rates/lengths differ dramatically between simulated training data and real data sets. Future work will have to investigate strategies to train a network that is robust to such misspecification, perhaps by incorporating a variety of indel models into training; additional steps to train neural networks to be insensitive to the proclivities of empirical data may also be necessary (discussed in Concluding Remarks section below).

**Topology Estimation under Branch-Length Heterogeneity**

There exist several regions in the branch-length parameter space where common methods fail to produce the correct topology. The bias known as long-branch attraction (LBA) is the inability of MP (Felsenstein 1978a) and possibly Bayesian (Kolaczkowski and Thornton 2009; Susko 2015) methods to estimate the correct topology in the presence of long nonadjacent branches (i.e. the Felsenstein zone; Huelsenbeck and Hillis 1993): these methods tend to erroneously place such long branches together and thus favor an incorrect topology (Table 1; Figs. 4c and 5c). Another region in the branch-length space, called the Farris zone ( Siddall 1998), causes ML methods to exhibit reduced accuracy on topologies that have adjacent long branches and causes MP to correctly infer trees more often as a result of LBA which, in this case, biases it toward the correct topology (Table 1; Figs. 4a and 5a); we note that this bias will diminish when using longer alignments due to ML’s property of consistency (Swofford et al. 2001). In addition, we assessed the performance of all methods in the asymmetrical (“twisted”) Farris zone (Table 1; Figs. 4b and 5b), where a single incorrect topology is favored by NJ if the corrected distance function is convex (McTavish et al. 2015). We also examined trees with a short internal branch (≤0.05) but external branches lengths varying along the interval [0, 0.5] (Table 1; Figs. 4d and 5d).

We primarily focused on these four regions of known bias (Figs. 4 and 5) but also examined several other regions of potential bias on test sets of gapped (Table 1; Supplementary Fig. S3 available on Dryad) and ungapped (Table 1; Supplementary Fig. S4 available on Dryad) MSAs. The accuracy estimates for each region are summarized in Table 2. In most of these regions, our CNN exhibited accuracy roughly matching or exceeding that of the best performing traditional method, whereas each other method failed catastrophically in at least one of these regions (Figs. 4 and 5 and Supplementary Figs. S3, S4 available on Dryad). The two exceptions where the CNN exhibited markedly reduced accuracy were the Felsenstein zone (Figs. 4 and 5; Table 2) and the “extra-short branches” region where all branch lengths vary uniformly along [0, 0.01] (Table 2; Supplementary Figs. S3 and S4 available on Dryad). Specifically, in the Felsenstein zone, the CNN’s accuracy is 0.464 (gapped) and 0.461 (ungapped), above the random expectation of 1/3, and well below that of ML [0.723 (gapped) and 0.719 (ungapped)].

**FIGURE 3.** Differences in parameter values between correct and incorrect classifications for each method. The distribution of the parameter governing the fraction of invariable sites (pinv) for correctly and incorrectly inferred trees for a) gapped and b) ungapped MSAs. The distribution of gamma rate parameter (α) for correctly and incorrectly inferred trees for c) gapped and d) ungapped MSAs. The black dots of each violin plot indicate median of the distribution. These results were obtained using 15,000 simulated MSAs from BL-space. BI = Bayesian inference; CNN = convolutional neuronal network; ML = maximum likelihood; MP = maximum parsimony; NJ = neighbor-joining.
FIGURE 4. Accuracy within regions with elevated branch-length heterogeneity for gapped MSAs. The four columns summarize results in a) the Farris zone, b) the twisted Farris zone, c) the Felsenstein zone, and d) trees with short internal branches. The top row illustrates these tree branch-length configurations, and the second row shows accuracy of each method’s performance in each zone (with violins representing accuracy on bootstrap replicates from the test set). Panels three through seven show the locations (gray dots) and densities (heat map) of incorrectly inferred topologies for each method plotted in two-dimensional space with B5 length on the x-axis and B1 + B2 (labeled as “B1 + B2”), B1 + B3 (“B1 + B3”), B1 + B2 + B3 (“B1 + B2 + B3”) on the y-axis for Farris, twisted Farris, Felsenstein, and short internal branch regions, respectively. See Table 1 for branch-length simulation details. The color scale shows the estimated density of the incorrectly inferred topologies at each point in the branch-length parameter space, with brighter colors representing a greater density of errors. BI = Bayesian inference; CNN = convolutional neuronal network; ML = maximum likelihood; MP = maximum parsimony; NJ = neighbor-joining.
FIGURE 5. Accuracy within regions with elevated branch-length heterogeneity for ungapped MSAs. The four columns summarize results in a) the Farris zone, b) the twisted Farris zone, c) the Felsenstein zone, and d) trees with short internal branches. The top row illustrates these tree branch-length configurations, and the second row shows accuracy of each method’s performance in each zone (with violins representing accuracy on bootstrap replicates from the test set). Panels three through seven show the locations (gray dots) and densities (heat map) of incorrectly inferred topologies for each method plotted in two-dimensional space with $B_5$ length on the x-axis and $B_1+B_2$ labeled as “$B_1+B_2$”, $B_1+B_2$ (“$B_{1,2}$”), $B_1+B_2+B_3$ (“$B_{1,2,3}$”), and $B_1+B_2+B_3+B_4$ (“$B_{1,2,3,4}$”)) on the y-axis for Farris, twisted Farris, Felsenstein, and short internal branch regions, respectively. See Table 1 for branch-length simulation details. The color scale shows the estimated density of the incorrectly inferred topologies at each point in the branch-length parameter space, with brighter colors representing a greater density of errors. BI = Bayesian inference; CNN = convolutional neuronal network; ML = maximum likelihood; MP = maximum parsimony; NJ = neighbor-joining.
(ungapped); Figs. 4c and 5c). However, CNN’s accuracy could be notably improved by adding additional MSAs simulated from the Felsenstein zone or the “extra-short branches” region: performing additional training iterations on 1000 MSAs per topology from these regions boosted the CNN’s accuracy to 0.753 for gapped MSAs and to 0.771 for ungapped MSAs for the Felsenstein zone, and to 0.882 (gapped) and 0.904 (ungapped) for the “extra-short branches” region, respectively (Table 2).

This suggests that it is possible to fix a known “blind spot” of the CNN by simply simulating additional relevant training data and performing more iterations of the training algorithm. The CNN shows superior accuracy in each of the remaining regions of potential bias (Supplementary Fig. S3 available on Dryad) for gapped MSAs compared with all other methods. Overall, on ungapped data, the CNN’s performance roughly matched that of BI across zones (Supplementary Fig. S4 available on Dryad), with the exception of the regions mentioned above. These two methods both exhibit acceptable but not exceptional accuracy in each of these different zones (Table 2), perhaps with the exception of the Felsenstein zone where all methods other than ML have low accuracy (≤0.57 vs. 0.719 for ML), unless the CNN is provided with additional training data as described above.

In addition, we tested performance of all methods on shorter gapped and ungapped MSAs of length 500 with one quadruplet region specified in Table 2 (see Supplementary Table S1 available on Dryad for accuracy estimates). Overall, accuracies were lower for all methods, but with qualitatively similar rankings across methods. Further, we showed that accuracy increases with increasing MSA size for the CNN which is potentially indicative of statistical consistency (Supplementary Table S2 and Text available on Dryad), though we do not prove this here.

Although we find that overall the CNN performs quite well on the test sets examined above, it is important to stress that a method’s overall utility is not fully captured by a single accuracy estimate from one test set, or even a suite of test sets as we have done here. Each of the test sets we examined is a subspace of the BL-space that we defined in the Supplementary Text available on Dryad, and there are an infinite number of such sets that one could examine. Moreover, partitioning this space into a number of equally sized sets small enough to comprehensively assess how performance varies across the entire BL-space is a monumental task. For example, if one defined a BL-space and discretized it into 1% increments for all five branches, one would need to simulate 100^5 data sets with all possible branch-length combinations to traverse the entire parameter space. The unfeasibility of such an experiment was previously acknowledged by (Huelsenbeck, 1995), thus we cannot rule out the possibility that our CNN’s performance is compromised in certain regions of parameter space especially if examples from such regions were underrepresented during training phase. Indeed, this appears be the cause of our initially poor performance on the Felsenstein and the “extra-short branches” zones, which was rectified through targeted additional training.

### Analysis of Phylogenetic Reliability Measures

Testing phylogenetic hypotheses usually involves calculation of some reliability measures such as bootstrap support (BS) for MP, NJ, and ML or posterior probabilities (PP) in the context of BI. Despite the fact that their direct comparison is somewhat problematic due to their fundamental differences, one can ask which measures are more prone to support false tree topologies (Douady et al. 2003; Huelsenbeck and Rannala 2004). For our CNN, we used two confidence measures: BS for the chosen topology (obtained by applying the trained CNN to each MSA in a set of bootstrap replicates), and the class membership posterior probability (CPP) estimates produced automatically by the CNN which can also be interpreted as a measure of topological reliability. We find that high support (BS > 0.95, or PP > 0.95 in the case of BI) is occasionally assigned to incorrectly classified trees by several methods: 0.052 of misclassifications by MP, 0.05 by NJ, 0.003 by ML, 0.042 by BI, and 0.038 by the CNN for gapped MSAs (Fig. 6a) and 0.056 by MP, 0.054 by NJ, 0.0036 by ML, 0.048 by BI, and 0.034 by the CNN for ungapped MSAs (Fig. 6b). Thus, a very high reliability measure cutoff may be necessary to filter out positively misleading cases. On the other hand, the CPP estimates show a larger disparity between scores assigned to correctly versus incorrectly classified topologies than do other measures (Fig. 6), and only 0.0004 and 0.0038 of incorrectly classified gapped and ungapped MSAs, respectively, having a CPP above 0.95. In other words, the CNN can avoid incorrect classifications even when imposing a relatively relaxed posterior probability cutoff, and thus simultaneously recover a large fraction of correctly classified topologies while producing very few misclassifications. This is further illustrated by precision-recall curves (Supplementary Fig. S5 available on Dryad), which show that compared with other methods the CNN achieves high sensitivity while retaining a high positive predictive value on gapped alignments. Moreover, for our four-taxon topologies the interpretation of CPP is straightforward: the most unreliable tree estimates receive CPP of ~1/3, which is equivalent to randomly picking one of the three possible topologies.

### Concluding Remarks

We used a CNN to infer unrooted quartet topologies through classification. Our results demonstrate that CNNs perform this task quite well and have a number of desirable qualities. First and foremost, under many scenarios our CNN resolves quartets with higher accuracy than existing methods (Table 2). The CNN is also relatively insensitive to regions of the tree branch-length heterogeneity (e.g. the Felsenstein and
Farris zones) that are known to confound certain methods, perhaps because the training set included trees with a variety of branch-length configurations (see Supplementary Text available on Dryad). Another advantage of using CNNs is that they produce posterior probability estimates for each tree topology; these PP are estimated during the classification process and thus require no additional computation. Also, we have shown that our CNN’s accuracy can be further improved by adding more training data sets, albeit most likely with diminishing returns (Fig. 2). Although training on large data sets can impose a computational burden up front, this training only needs to be completed once, after which the CNN can be used to infer tree topologies very rapidly: the CNN can infer the topologies of 3000 MSAs of length 1000 in ∼5 s using an NVIDIA Tesla V100-SXM2 GPU. Finally, CNNs have the advantage of being able to incorporate indels into their classification in a straightforward manner—in principle one need only include them in the simulated training data—which can improve accuracy by supplying more information to the classifier. However, future efforts will have to examine the robustness of inference to indel model misspecification, and the extent to which robustness can be improved by training under a mixture of indel rates and length distributions. More generally, this problem underscores the importance of considering ways in which empirical data may not match simulated data, and the consequences of such mismatches for inference. This problem is particularly challenging, as one may not always be able to predict the manner in which empirical data may differ substantially from model-based simulations, but could potentially be addressed by incorporating empirical data as well into the training process. For example, the CNN could be trained on simulated MSAs, and then a data set consisting of real MSAs for which there is strong consensus around the true topology can be supplied to the CNN for additional training iterations. Thus, through this transfer learning procedure (Pan and Yang 2010), the CNN would first learn to accurately infer trees from simulated MSA, and then be tuned to recognize some of the idiosyncrasies of real MSAs.

Our efforts thus suggest that there is great potential for the application of deep learning techniques to inferring trees, which is central to questions in both phylogenetics (species tree inference) and population genomics (inferring ancestral recombination graphs along a recombinating chromosome (Rasmussen et al. 2014). Indeed, using the entire MSA as input would be most useful on recombining sequences—otherwise it is possible to collapse the MSA down to a vector of statistics (e.g. the number of occurrences of each distinct possible MSA column, if the number of sequences is small) that would sufficiently capture all of the relevant information from the input. Deep learning methods acting directly on the MSA could also be applied to additional phylogenetic problems, such as substitution model selection or tree inference from protein sequence data.

Although our findings are encouraging, additional advances are required for our approach to match the practical utility of conventional methods. First, we have only investigated the ability of a CNN to produce the correct tree topology but not the associated branch lengths. Artificial neural networks are capable of estimating real-valued parameters in addition to discrete class labels, so future work should investigate the possibility of using deep learning for inferring branch lengths in addition to the topology. Another limitation of our current approach is that it is limited to a small number of sequences/taxa. Future extensions could use quartet amalgamation methods...
(e.g. von Haeseler and Strimmer 1996; Reaz et al. 2014). Although such methods have not matched the accuracy of standard approaches such as ML in the past, because our method is able to resolve quartet topologies with greater accuracy and better calibrated confidence scores, it may be worth investigating whether quartet amalgamation combined with our CNN could compete with standard methods when applied to larger numbers of sequences. Alternatively, it may be possible to devise neural networks capable of directly predicting a tree topology. The latter could in principle be accomplished by treating the values of the final hidden layer of the network as entries in a distance matrix and using an output layer that builds a tree from this matrix (e.g. using NJ), and adopting a tree dissimilarity measure such as RF distance as the objective function to be minimized. However, this approach would also have a number of practical challenges that would have to be overcome (e.g. how to train networks that can handle different numbers of input sequences). One possible solution to this problem is to train a number of CNNs each designed to handle a fixed number of sequences, and then when given an input MSA select the CNN corresponding to the closest number of sequences greater than or equal to that of the input. The size of input can then be increased by adding duplicates of one row of the MSA until the desired number of sequences is reached. After inferring the topology for this input, the duplicate nodes can be pruned from the tree to produce the final topology.

Finally, to move from inferring gene trees to inferring species trees, our approach will have to be extended to handle tree discordance due to variation in evolutionary rates across loci, incomplete lineage sorting (Maddison and Knowles 2006; Degnan and Rosenberg 2009) and gene flow (Maddison 1997). Supervised learning approaches may prove to be well suited for this question, as arbitrarily complicated phylogenetic models combining multiple sources of discordance can readily be incorporated into the training process via simulation. Thus, although additional practical developments are necessary, applications of deep learning to problems in tree inference and phylogenomics are likely to be a fruitful endeavor.

**DATA AVAILABILITY**

R and Python scripts can be found at GitHub (https://github.com/SchriderLab/Tree_learning), also
archived on Zenodo (DOI: 10.5281/zenodo.3246669). Training, validation, and test MSA data sets are deposited on Figshare (https://doi.org/10.6084/m9.figshare.8279618.v2).

SUPPLEMENTARY MATERIAL

Data available from the Dryad Digital Repository: http://dx.doi.org/10.5061/dryad.ct2895s.

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