Effect of Chaihu-Shugan-San on the mRNA expression of the 5-HT1A receptor and cellular proliferation in the hippocampus of epileptic rats with depression

PING YANG1*, LIANG LI2,3*, XUE-JUN LIU1, XIONG CAI2, MEI-ZHEN SUN4, JUN-FENG HE2, GUANG ZENG2 and HUI-YONG HUANG2

1Department of Psychiatrics, Hunan Brain Hospital, Changsha, Hunan 410007; 2Provincial Key Laboratory of TCM Diagnostics and 3Department of Anatomy, Hunan University of Chinese Medicine, Changsha, Hunan 410208; 4Department of Neurology, The First Hospital of Shanxi Medical University, Taiyuan, Shanxi 030001, P.R. China

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Abstract. Chaihu-Shugan-San (CHSGS) is a herbal preparation that has been shown to effectively relieve neurologic impairment and reduce depression. However, the efficacy of CHSGS in the treatment of patients with epilepsy with depression is unknown. Therefore, in the present study, adult rats were exposed to chronic mild stress following the establishment of chronic pilocarpine-induced epilepsy. Subsequently, a sucrose consumption test and open-field test (OFT) were performed to assess depression-like behavior. Rats were randomly divided into four groups: Control, model, fluoxetine (1.8 g/kg) and CHSGS (2.7 g/kg) groups. The control and model groups received normal saline. The mRNA expression levels of the 5-hydroxytryptamine 1A (5-HT1A) receptor and the number of 5-bromo-2’-deoxyuridine (BrdU)-labeled cells in the hippocampal dentate gyrus were detected using reverse transcription-quantitative polymerase chain reaction and immunohistochemical analysis, respectively. Treatment administration was conducted by once daily intragastric perfusion for 28 days. The mRNA expression levels of the 5-HT1A receptor, the number of BrdU-labeled cells in the hippocampal dentate gyrus, the consumption of sucrose, and frequency of vertical and horizontal movement scores in the OFT were enhanced in the fluoxetine and CHSGS groups compared with the model group (P<0.05). However, no statistically significant difference was detected between the fluoxetine and CHSGS groups. These data suggest that CHSGS is able to increase the expression of 5-HT1A receptor mRNA and cellular proliferation in the hippocampal dentate gyrus in epileptic rats with depression, and thus effectively improve certain symptoms of depression.

Introduction

The co-occurrence of epilepsy and depression is commonly observed in neurological psychiatry (1). Studies have revealed that the co-occurrence of epilepsy with depression is associated with the activation of 5-hydroxytryptamine 1A (5-HT1A) receptors (2,3). A previous study indicated that the 5-HT1A receptor agonist 8-hydroxy-dipropylaminotetralin (8-OH-DPAT) exerts anti-epileptic and anti-depressive effects in epileptic rats with depression, which involve the enhancement of the growth of hippocampal neurons (4). However, certain current treatments possess shortcomings, such as the antidepressant bupropion which may cause generalized seizures and central nervous system depression if administered in excess (5). Chinese medicinal herbs, such as Bupleurum chinense root and Pericarpium Citri Reticulatae appear to exhibit limited adverse reactions and therapeutic efficacy (6).

The present study employed chronic epileptic rats with depression induced by a combination of pilocarpine and chronic mild stress (CMS) to investigate the efficacy of Chaihu-Shugan-San (CHSGS). Detection methods included behavioral analysis, immunohistochemistry and reverse transcription-quantitative polymerase chain reaction (RT-qPCR). The expression levels of 5-HT1A receptor mRNA and numbers of 5-bromo-2’-deoxyuridine (BrdU)-labeled cells in the hippocampus were detected to clarify whether CHSGS improves cellular proliferation in the hippocampus by promoting the expression of 5-HT1A receptor mRNA. The results of this
study may support the use of CHSGS as a novel treatment for epilepsy with depression.

Materials and methods

Animals and experimental protocol. The present study was performed as a randomized, controlled animal study. The study was performed at the Anatomical Research Laboratory of Hunan University of Chinese Medicine (Changsha, China). A total of 60 male Sprague-Dawley (SD) rats (weight, 180-220 g; age, 8-9 weeks) were provided by the Laboratory Animal Center of Hunan University of Chinese Medicine. The animals were acclimatized to the environment for 2 weeks, while being trained to consume 1% (w/v) sucrose solution for taste adaptation. Based on the baseline scores of sucrose consumption test and the open-field test (OFT) during the adaptation period, 48 rats with similar behavior were randomly divided into four groups: Control, model, fluoxetine and CHSGS groups (n=12 per group). All procedures were performed in accordance with the Guidance Suggestions for the Care and Use of Laboratory Animals, formulated by the Ministry of Science and Technology of China (7).

Reagents. The CHSGS preparation contained the following: 6 g Bupleurum chinense root, 6 g Pericarpium Citri Reticulatae, 4.5 g Rhizoma Chuanxiong, 4.5 g Rhizoma Cyperi, 4.5 g Fructus Aurantii, 4.5 g Paeonia and 1.5 g Glycyrrhizae uralensis root. All CHSGS herbal components were obtained from the First Hospital of Hunan University of Chinese Medicine. The herbal components were identified by an expert to fulfill the quality requirements of the Pharmacopoeia of the People's Republic of China (8). CHSGS and its components were individually decocted in boiling water for 45 min, concentrated and vacuum-dried to form a paste, and were subsequently combined into a paste containing 8 g crude extracts per gram. This CHSGS paste was diluted with distilled water to a concentration of 1.0 g paste, and were subsequently combined. One bottle of sucrose solution was replaced with 100 ml water for 24 h. On day 3, rats were deprived of water and food for 24 h. On day 2, one bottle of sucrose solution was replaced with 100 ml water for 24 h. On day 3, rats were deprived of water and food for 24 h. On day 4, rats were deprived of water and food for 24 h. On day 5, rats were deprived of water and food for 24 h. On day 6, rats were deprived of water and food for 24 h. On day 7, rats were deprived of water and food for 24 h. On day 8, rats were deprived of water and food for 24 h. On day 9, rats were deprived of water and food for 24 h. On day 10, rats were deprived of water and food for 24 h. On day 11, rats were deprived of water and food for 24 h. On day 12, rats were deprived of water and food for 24 h. On day 13, rats were deprived of water and food for 24 h. On day 14, rats were deprived of water and food for 24 h. On day 15, rats were deprived of water and food for 24 h. On day 16, rats were deprived of water and food for 24 h. On day 17, rats were deprived of water and food for 24 h. On day 18, rats were deprived of water and food for 24 h. On day 19, rats were deprived of water and food for 24 h. On day 20, rats were deprived of water and food for 24 h. On day 21, rats were deprived of water and food for 24 h. On day 22, rats were deprived of water and food for 24 h. On day 23, rats were deprived of water and food for 24 h. On day 24, rats were deprived of water and food for 24 h. On day 25, rats were deprived of water and food for 24 h. On day 26, rats were deprived of water and food for 24 h. On day 27, rats were deprived of water and food for 24 h. On day 28, rats were deprived of water and food for 24 h. On day 29, rats were deprived of water and food for 24 h. On day 30, rats were deprived of water and food for 24 h. On day 31, rats were deprived of water and food for 24 h. On day 32, rats were deprived of water and food for 24 h. On day 33, rats were deprived of water and food for 24 h. On day 34, rats were deprived of water and food for 24 h. On day 35, rats were deprived of water and food for 24 h. On day 36, rats were deprived of water and food for 24 h. On day 37, rats were deprived of water and food for 24 h. On day 38, rats were deprived of water and food for 24 h. On day 39, rats were deprived of water and food for 24 h. On day 40, rats were deprived of water and food for 24 h. On day 41, rats were deprived of water and food for 24 h. On day 42, rats were deprived of water and food for 24 h. On day 43, rats were deprived of water and food for 24 h. On day 44, rats were deprived of water and food for 24 h. On day 45, rats were deprived of water and food for 24 h. On day 46, rats were deprived of water and food for 24 h. On day 47, rats were deprived of water and food for 24 h. On day 48, rats were deprived of water and food for 24 h. On day 49, rats were deprived of water and food for 24 h. On day 50, rats were deprived of water and food for 24 h. On day 51, rats were deprived of water and food for 24 h. On day 52, rats were deprived of water and food for 24 h. On day 53, rats were deprived of water and food for 24 h. On day 54, rats were deprived of water and food for 24 h. On day 55, rats were deprived of water and food for 24 h. On day 56, rats were deprived of water and food for 24 h. On day 57, rats were deprived of water and food for 24 h. On day 58, rats were deprived of water and food for 24 h. On day 59, rats were deprived of water and food for 24 h. On day 60, rats were deprived of water and food for 24 h.

Succrose consumption test. The sucrose consumption test was performed as described previously (10). The test was conducted over 3 days. On day 1, rats were housed individually and had free access to two 100-ml bottles of sucrose solution (1% w/v). Rats were trained to adapt to sucrose solution for 24 h. On day 2, one bottle of sucrose solution was replaced with 100 ml water for 24 h. On day 3, rats were deprived of water and food for 23 h, and subsequently received free access to two pre-weighed bottles containing 100 ml sucrose solution (1% w/v) and 100 ml water, respectively. After 1 h, the consumed volume from each bottle was recorded. The sucrose preference rate was calculated using the following formula: Sucrose preference rate (%) = Sucrose consumption/water consumption x 100. This test was conducted prior to division of the rats into groups, and following the 28-day treatment period.

OFT. The OFT was performed according to a previously described method (11), to measure spontaneous activity in rodents. Briefly, the floor of a 100x100x40-cm square arena was divided into 16 equal 25x25-cm squares. A single rat was placed in the center of the arena, and after 30 sec of adaptation the frequency of vertical and horizontal movement were recorded manually for 5 min. All rat movement was recorded using an XZ-10 camera (Olympus Corporation, Tokyo, Japan) located 40 cm above the arena. After each test, the arena was cleaned with 90% alcohol solution.

Epilepsy model establishment. Adult male rats in the model, fluoxetine and CHSGS groups received 127 mg/kg lithium chloride (62480; Sigma-Aldrich, St. Louis, MO, USA) intraperitoneally (i.p.). On the following day, 1 mg/kg methylscopolamine bromide (W131102; i.p.; Shanghai Tongyong Pharmaceutical Co., Ltd., Shanghai, China) was administered to limit the peripheral effects of the convulsant. Status epilepticus was induced by injecting 35 mg/kg pilocarpine (P1650000; Sigma-Aldrich) 30 min after the administration of methylscopolamine bromide. Animals were monitored throughout status epilepticus induction, and seizure severity was assessed according to the scale described by Racine (12). Animals that did not show clear signs of status epilepticus were excluded, since ≥1 h of status epilepticus is required to develop spontaneous recurrent epileptic seizures in the pilocarpine model of epilepsy. Status epilepticus occurred within 15-60 min and was characterized by continuous motor-limbic seizures accompanied by intermittent rearing and falling and occasional wild running spells. At 1 h after status epilepticus, the seizures were stopped with 4 mg/kg diazepam (AH131203; Shanghai Sunrise Pharmaceutical Co. Ltd., Shanghai, China) administered intramuscularly and/or 3 ml/kg chloral hydrate (i.p.; Anatomical Research Laboratory of Hunan University of Chinese Medicine).

Epilepsy with depression model establishment. Following the successful establishment of the epilepsy model, CMS was induced in accordance with a previously described method (13). The CMS regimen included seven different stressors, which were arranged in random order for 14 consecutive days as follows: Food and water deprivation (20 h, followed by sucrose consumption test); water deprivation (18 h, followed by 1 h exposure to empty bottles); 45° cage tilt (17 h); overnight illumination (lights on for a total of 36 h); soiled cage (200 ml of water in 100 g sawdust bedding, 21 h); swimming in 41°C water (5 min); and paired caging (2 h). At day 14 after the establishment of CMS, the model, fluoxetine and CHSGS groups were intragastrically perfused with 10 ml/kg normal saline, 1.8 g/kg fluoxetine and 2.7 g/kg CHSGS once per day for a total of 28 days.

5-Bromo-2′-deoxyuridine (BrdU) labeling and immunofluorescence staining. The thymidine analog BrdU (ED1100;
Table I. Primer sequences used for polymerase chain reaction.

| Primer               | Sequences (5’-3’)                                      | Product size (bp) |
|----------------------|--------------------------------------------------------|-------------------|
| 5-HT1A               | Forward: GCACCAGCTTAGGAACCTTCG<br>Reverse: CAGAGGAAGGTGCTCTTTGG | 206               |
| β-actin              | Forward: GTCAGGTCTATCCTATCGGCAAT<br>Reverse: AGAGGTCTTTACGGATGTCAACGT | 210               |

5-HT1A, 5-hydroxytryptamine 1A.

Sigma-Aldrich) was dissolved in saline with 0.007 M NaOH. The BrdU solution (50 mg/kg) was administered twice per day for 7 days following the experimental treatment with fluoxetine or CHSGS administration. Subsequently, 6 rats in each group were sacrificed 24 h after the final BrdU administration using 10% chloral hydrate, transfused intraperitoneally. Rats were perfused transcardially with cold 0.1 M phosphate-buffered saline (PBS) and 4% paraformaldehyde. The brains were removed, post-fixed for 18 h, and placed in 30% sucrose until they sank. Frozen coronal sections were cut at 25 µm using a cryostat (CM3050S; Leica Biosystems GmbH, Nußloch, Germany) and stored at -20°C in a cryoprotectant solution containing 25% ethylene glycol, 25% glycerol and 50% 0.1 M phosphate buffer (pH 7.4) for immunofluorescence staining.

The sections were subsequently incubated with 1% bovine serum albumin to block non-specific interactions. Following this step, the sections were incubated with mouse anti-rat BrdU monoclonal antibody (1:100; BM0201; Wuhan Boster Biological Technology, Ltd., Wuhan China) in PBS containing 0.3% Triton X-100 overnight at 4°C. Subsequently, the sections were incubated with goat anti-mouse IgG-Cy2 (1:100; SA1021; Wuhan Boster Biological Technology, Ltd.) for 2 h at room temperature. The stained sections were observed using a BX-51 confocal microscope (Olympus Corporation). BrdU-labeled cells were quantified using Image Pro software, version 6.0 (Media Cybernetics, Inc., Bethesda, MD, USA).

RT-qPCR to detect 5-HT1A receptor mRNA expression levels in the rat hippocampus. Following the behavioral tests, 6 rats in each group were sacrificed. Fresh hippocampus tissue was dissected and placed on ice. Specimens were removed from the liquid nitrogen. Total RNA was extracted using TRIzol (2214L; Beijing CoWin Biotech Co., Ltd., Beijing, China). RNA concentration was determined using a P-Class NanoPhotometer (5622; Implen GmbH, Munchen, Germany). The RT step was conducted using a GoScript™ Reverse Transcription System (0000112333; Promega Corporation, Madison, WI, USA), following the manufacturer’s instructions. Primers were synthesized by Wuhan Boster Biological Technology, Ltd. (Table 1), and a 2x SYBR GREEN-I Mix kit (0020140910; Beijing Biotekte Biotechnological Co., Ltd., Beijing, China) was used. Amplification reactions were conducted using a qPCR system (TL988; Xi’an Tianlong Science and Technology Co., Ltd., Xi’an, China).

The PCR cycling conditions were as follows: i) Pre-denaturation at 95°C for 10 min; ii) 40 cycles of denaturation at 95°C for 15 sec and annealing/extension at 60°C for 1 min; and iii) final extension at 60°C for 5 min. The specificity of PCR products was monitored via melting curve analysis, and the standard curves of the target and reference gene β-actin were prepared. The ratio of 5-HT1A to β-actin was considered as the relative expression of the target gene. Relative quantification of gene expression was determined using the 2^ΔΔCq method.

Statistical analysis. Statistical analysis was performed using SPSS software, version 17.0 (SPSS, Inc., Chicago, IL, USA). Measurement data are expressed as the mean ± standard error of the mean. Analysis of variance was used for intergroup comparisons. P<0.05 was considered to indicate a statistically significant difference.

Results

Animal grouping and treatments. A total of 60 SD rats were used. The baseline scores of the sucrose consumption test and OFT were determined, and then 48 SD rats with similar behavior were identified and randomly divided into four groups: Control, model, fluoxetine and CHSGS groups (n=12 per group).

Behavior of epileptic rats with depression. After 28 days of treatment, the consumption of sucrose water, the sucrose preference rate, and frequency of vertical and horizontal movement decreased in the model group compared with the control group (P<0.05). Compared with the control group, frequency of vertical movement decreased in the fluoxetine group (P<0.05), while frequency of vertical and horizontal movement decreased in the CHSGS group (P<0.05). Compared with the model group, all sucrose and OFT test parameters were increased in the fluoxetine and CHSGS groups (P<0.05). However, no statistically significant differences were observed between the fluoxetine and CHSGS groups (Fig. 1).

CHSGS promotes cellular proliferation in the hippocampal dentate gyrus. Immunofluorescence staining indicated the presence of BrdU-labeled cells in the hippocampal dentate gyrus (Fig. 2). After 28 days of treatment, the number of BrdU-labeled cells increased significantly in the hippocampal dentate gyrus of the model, fluoxetine and CHSGS groups compared with the control group (P<0.05). Furthermore, compared with the model group, the number of BrdU-labeled...
cells increased significantly in the hippocampal dentate gyrus in the fluoxetine and CHSGS groups (P<0.05). No significant difference was identified between the fluoxetine and CHSGS groups (P>0.05; Fig. 3).

**Discussion**

The reported prevalence of epilepsy in China ranges between 3.6 and 7.0% (14). The number of patients with active epilepsy was >9 million in 2010, and increases by ~400,000 each year. Recurrent epileptic attacks and long-term treatment can seriously reduce the life quality of the patients, and affect mentality to a certain extent. Evident psychological and behavioral disorders have been identified in large numbers of patients with epilepsy (15). The prevalence of depression in patients with epilepsy is reportedly 23.1%, which is higher compared with that in the general population. In addition, the lifetime prevalence of depression in patients with epilepsy is high, approaching 13% (16). Thus, the co-occurrence of epilepsy and depression is a common phenomenon.
In our previous study, epileptic rats with depression were used in a lithium pilocarpine-induced epileptic model (4). Thus, a combined model of epilepsy with depression was established. However, the model exhibited a number of shortcomings, such as the low success rate of model establishment of ~25%, high experimental expense and waste of resources. In the present study, the establishment of the combined model of epilepsy with depression has been improved compared with the previous model. The revised model includes a combination of the traditional lithium pilocarpine-induced epilepsy model and the CMS-induced depression model. The success rate of model establishment reached ~80% in the present study.

In our previous study, the 5-HT1A receptor agonist 8-OH-DPAT was used to improve depressive symptoms in epileptic rats with depression. However, the clinical application of 8-OH-DPAT is currently limited, which may be due to a number of factors. Firstly, although 5-HT1A receptor agonists, such as tandospirone, have good antidepressant effects (17), they are frequently used as antidepressant synergists (18). Secondly, the treatment compliance for traditional and newer antidepressant drugs has been adversely affected by their defects, including side effects and addictive properties. Certain antidepressant agents in clinical use, such as tricyclic antidepressants, bupropion and fluoxetine (19,20), have been reported to cause seizures in certain patients. Therefore, the selection of antidepressants for epileptic patients with depression is controversial.

According to Traditional Chinese Medicine (TCM) theory, epilepsy with depression belongs to the ‘depressed syndrome’ category of diseases (21) and the pathogenesis has a marked association with emotion. The liver fails to maintain the normal flow of ‘Qi’, which affects ‘pneuma’ transportation to the five ‘zang’ organs and the six ‘fu’ organs, resulting in corresponding emotional symptoms and potentially depression. The primary treatment method for depression according to the TCM tradition is the herbal preparation Shu Gan, which is hypothesized to conduct ‘Qi’ and release ‘liver depression’. According to TCM theory, the movement of ‘Qi’ is able to ‘free the blood’, and thus alleviate various forms of stagnancy. Therefore, in the present study a representative decoction of the ‘Shu Gan Li Qi’ method (CHSGS) was selected as a candidate treatment for epileptic rats with depression.

It has been reported that CHSGS reduces the expression of microRNA-125a and microRNA-182, which are associated with the onset of depression (22) in the hippocampus. Furthermore, CHSGS increases brain-derived neurotrophic factor and tropomyosin receptor kinase B levels in the amygdaloid nucleus, hippocampus and frontal lobe (23) and upregulates the expression of ERK1/2 mRNA in the hippocampus (24), thus improving the symptoms of depression in rat depression models. CHSGS appears to enhance the expression levels of B-cell lymphoma 2 (Bcl-2) and decrease those of Bcl-2-like protein 4 in the hippocampus and frontal cortex, thus inhibiting neuronal apoptosis in epileptic rats (25). In addition, CHSGS has been shown to decrease the expression of multidrug resistance protein P-glycoprotein and electroencephalograph epileptiform discharges in the hippocampus and temporal lobe cortex (26), which are associated with antiepileptic changes. In the clinical treatment of patients with post-stroke depression, the curative effect of CHSGS was found to be better compared with fluoxetine at relieving the symptoms of depression and stabilizing patient emotional state (27). CHSGS combined with the conventional antidepressant venlafaxine has been shown to significantly improve neurological functions and alleviate depressive symptoms in
stroke patients (28). In addition, Wang et al (29) conducted a meta-analysis which confirmed the efficacy and safety of CHSGS for the treatment of depression. In the present study, CHSGS was administered to epileptic rats with depression. No statistically significant difference in depressive behavior was observed between the CHSGS and fluoxetine groups. By contrast, compared with the model group, the behavioral data in CHSGS group exhibited statistically significant differences. These results suggest that CHSGS improves the symptoms of depression in epileptic rats with depression.

However, the underlying mechanism by which CHSGS ameliorates the depressive symptoms in epileptic rats with depression requires clarification. Recent studies and our previous study have indicated that the 5-HT1A receptor is associated with the pathogenesis of epilepsy with depression (4,30). The 5-HT1A receptor has become a key drug target for the treatment of epilepsy with depression (31). Therefore, it is possible that CHSGS improves depressive symptoms in rats by modifying the expression of the 5-HT1A receptor. In the present study, compared with the control group, the expression of 5-HT1A receptor mRNA was decreased in the model, fluoxetine and CHSGS groups. The expression of 5-HT1A receptor mRNA was enhanced in the fluoxetine and CHSGS groups compared with the model group. However, no significant difference in 5-HT1A receptor mRNA expression was identified between the fluoxetine and CHSGS groups. These results indicate that the expression of 5-HT1A receptor mRNA is reduced in epileptic rats with depression. Fluoxetine and CHSGS are able to increase the expression of 5-HT1A receptor mRNA; however, there is no discernible difference in their therapeutic efficacy.

The mechanism underlying the antiepileptic and antidepressive effect of increased 5-HT1A receptor expression remains unclear. Our previous study indicated that the 5-HT1A receptor is associated with neural plasticity and is crucially involved in the pathogenesis of epilepsy with depression (4). Therefore, we hypothesize that the 5-HT1A receptor exerts its antiepileptic and antidepressive effect by promoting neural plasticity. The adult hippocampus has the capability to generate new neurons throughout life, and this property contributes to functional plasticity under physiological and pathological conditions (32). Further research has shown that neural stem cells are present in the subgranular zone of the dentate gyrus throughout the lifetime of an individual (33). These cells proliferate and differentiate into neurons and glial cells, and subsequently migrate into the granule cell layer and finally into the molecular layer (33). Various antidepressants, including tricyclic antidepressants and selective serotonin reuptake inhibitors, are able to increase neuronal proliferation in the hippocampal dentate gyrus in adult rodents (34). A previous study has indicated that the activation of the 5-HT1A receptor is a critical step in the activation of seizure-induced cellular proliferation and survival in the dentate gyrus (35). This effect may attenuate depressive symptoms in epilepsy by enhancing neuronal plasticity.

In the present study, no statistically significant difference was detected between the CHSGS and fluoxetine groups with regard to the number of BrdU-labeled cells, indicating that the two agents have similar effects on cell proliferation. Cell proliferation in the CHSGS group was significantly greater than that in the model group, and the expression of 5-HT1A receptor mRNA was increased accordingly. Thus, the results of the present study indicate that CHSGS regulates neurogenesis in epileptic rats with depression by adjusting the expression levels of the 5-HT1A receptor. However, the exact mechanism by which CHSGS exerts its anti-epileptic and anti-depressive effect against epilepsy with depression remains unclear and requires further study.

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