Programmed cell death 4 (PDCD4) expression during multistep Barrett’s carcinogenesis

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ABSTRACT

Aim To test the contribution of programmed cell death 4 (PDCD4) tumour suppressor gene in Barrett’s carcinogenesis.

Methods PDCD4 immunohistochemical expression was assessed in 88 biopsy samples obtained from histologically proven long-segment Barrett’s mucosa (BM); 25 non-intestinal columnar metaplasia, 25 intestinal metaplasia (IM), 16 low-grade intraepithelial neoplasia (LG-IEN), 12 high-grade IEN (HG-IEN) and 10 Barrett’s adenocarcinoma (BAC). As controls, 25 additional samples of native oesophageal mucosa (N) were obtained from patients with dyspepsia. To further support the data, the expression levels of miR-21, an important PDCD4 expression regulator, in 14 N, 5 HG-IEN and 11 BAC samples were determined by quantitative real-time PCR analysis.

Results PDCD4 immunostaining decreased progressively and significantly with the progression of the phenotypic changes occurring during Barrett’s carcinogenesis (p<0.001). Normal basal squamous epithelial layers featured strong PDCD4 nuclear immunoreaction (mostly coexisting with weak–moderate cytoplasmic staining). Non-intestinal columnar metaplasia and intestinal metaplasia preserved a strong nuclear immunostaining; conversely, a significant decrease in PDCD4 nuclear expression was seen in dysplastic (LG-IEN and HG-IEN) and neoplastic lesions. Weak–moderate cytoplasmic immunostaining was evident in cases of LG-IEN, while HG-IEN and BAC samples showed weak cytoplasmic or no protein expression. As expected, miR-21 expression was significantly upregulated in HG-IEN and BAC samples, consistently with PDCD4 dysregulation.

Conclusions These data support a significant role for PDCD4 downregulation in the progression of BM to BAC, and confirm miR-21 as a negative regulator of PDCD4 in vivo. Further efforts are needed to validate PDCD4 as a potential prognostic marker in patients with Barrett’s oesophagus.

INTRODUCTION

In many western countries, the incidence of Barrett’s adenocarcinoma (BAC) has dramatically increased over the last two decades. Barrett’s oesophagus (BE) is defined as the replacement of squamous oesophageal epithelium by columnar intestinalised mucosa (Barrett’s mucosa, BM) and this metaplastic transformation is recognised as the cancerisation field in which BAC develops.1–3

Many investigators have focused on which histopathological characteristics of BE have malignant potential, and which factors promote Barrett’s carcinogenesis. The histological diagnosis of dysplasia (defined as low-grade or high-grade intraepithelial neoplasia [IEN or NIIN]) is currently considered the only biomarker for defining high-risk BE populations.4–6 The identification of additional molecular markers of cancer progression might support strategies for cancer secondary prevention and/or provide a biological rationale for targeted therapies.

A growing number of reports have pointed to PDCD4 (programmed cell death 4) as a new tumour suppressor gene.7–13 PDCD4 is a 64 kDa protein involved in the apoptotic machinery, which suppresses cell transformation, tumorigenesis and invasion.7–13 Different mechanisms have been implicated in the control of the steady-state and subcellular location of PDCD4. Among others, the oncogenic microRNA miR-21 (hsa-miR-21) has been shown to specifically target the PDCD4 3’-UTR, which negatively regulates PDCD4 expression.14–15

PDCD4 expression is significantly downregulated in various human cancers, as well as in cancer cell lines, and this has been associated with a poor patient prognosis.16–24 PDCD4 protein levels have been found to be inversely correlated with miR-21 expression in oesophageal squamous cell carcinoma cell lines,17 and we have shown that PDCD4 expression is significantly downregulated in oesophageal cancers (adenocarcinoma and squamous cell carcinoma histotypes) and it predicts patient outcome.25

To test the role of PDCD4 in contributing to oesophageal carcinogenesis, we investigated PDCD4 immunohistochemical expression in Barrett’s carcinogenesis. We also examined miR-21 expression levels in high-grade IEN (HG-IEN) and BAC samples by quantitative real-time PCR (qRT-PCR analysis).

MATERIALS AND METHODS

cDNA microarray analysis

The Oncomine database and gene microarray analysis tool, a repository for published cDNA microarray data (http://www.oncomine.org/),26 27 was explored (on 15 December 2009) for PDCD4 mRNA expression in non-neoplastic oesophageal tissues, BM and primary BAC. Oncomine algorithms were used to perform a statistical analysis of the differences in PDCD4 expression, since it allows for multiple comparisons among different studies.26–28 Only studies with analytical results with a p<0.05 were considered.

Patients

The cases in the present study were retrospectively collected from the files of the Veneto Region’s multicentre Barrett’s Oesophagus Registry (EBR;
Padova Unit,29 selecting cases of histologically proven, long-segment BE. A total of 88 biopsy samples of oesophageal mucosa were obtained from different patients with BE, that is, 25 with non-intestinal columnar metaplasia (cardiac-type columnar metaplasia), 25 with intestinal metaplasia (Barrett’s mucosa), 16 with low-grade intraepithelial neoplasia (LG-IEN), 12 with high-grade IEN (HG-IEN), and 10 with BAc. Another 25 native oesophageal mucosa samples (N) were obtained from patients with dyspepsia who had no gastro-oesophageal disease at endoscopy/histology (from the files at the Surgical Pathology Unit of the University of Padova). For the qRT-PCR study, 2 mm cores were obtained from paraffin blocks selected from 14 consecutive patients who had undergone oesophagectomy at the Department of Gastroenterological and Surgical Sciences, University of Padova, for HG-IEN and/or BAc, giving a total of 14 N, 5 HG-IEN and 11 BAc samples. None of the patients had received neoadjuvant therapy. Written consent was obtained from all the patients involved in the study.

Histological and immunohistochemical study
All biopsy specimens were immediately fixed in 10% buffered formalin and embedded in paraffin wax. Serial histological sections 4–6 µm thick were obtained from each paraffin wax block and stained with H&E and Alcian-PAS. The original diagnosis was confirmed on the histological evidence in all cases. Immunohistochemical staining was done automatically (Ventana Benchmark XT system; Ventana, Touchstone, Arizona, USA)30 for PDCD4 (1:100; catalogue no. HPA001032; Atlas Antibodies, Stockholm, Sweden) according to the manufacturer’s instructions. Sections were lightly counterstained with haematoxylin. Appropriate positive and negative controls were run concurrently. In cancer samples, the presence of positive stromal and inflammatory cells served as an internal control. PDCD4 expression was jointly scored by two pathologists (MF and MR) who were unaware of the patients’ clinical history. Consistent with Mudduluru et al.,22 nuclear and cytoplasmic immunoreactions were considered. Nuclear PDCD4 staining results were classified in four groups according to the percentage of positively stained nuclei: score 0, none; score 1, ≤50%; score 2, 30–70%; score 3, ≥70%. Cytoplasmic PDCD4 expression was scored by staining intensity (score 0, none; score 1, weak; score 2, intermediate; score 3, strong). A total score was also calculated as the sum of the nuclear and cytoplasmic scores, and cases were divided into four groups, defined as: negative (total score 0), weak staining (total score 1 or 2), intermediate staining (total score 3 or 4), and strong staining (total score 5 or 6).

Quantitative real-time PCR
Tissue cores were deparaffinised with xylene at 50°C for 3 min. Total RNA extraction was done using the RecoverAll kit (Ambion, Austin, Texas, USA) according to the manufacturer’s instructions. The NCodeTM miRNA qRT-PCR method (Invitrogen, Carlsbad, California, USA) was used to detect and quantify mature miR-21 (primer sequence: 5’-CGGTAGCT-TATCGACTGATGGTA-3’) on real-time PCR instruments according to the manufacturer’s instructions (Applied
Biosystems, Foster City, California, USA). Normalisation was done with the small nuclear RNA U6B (RNU6B; Invitrogen). PCR reactions were run in triplicate, including no-template controls. The data were analysed using the comparative CT method.

**Statistical analysis**

Kruskal–Wallis test, Student t test and Pearson's correlation tests were used to analyse differences and correlations between groups; p values <0.05 were considered significant. All the statistical analyses were done with Stata software (Stata Corporation, College Station, Texas, USA).

**RESULTS**

The *PDCD4* gene is downregulated in Barrett's carcinogenesis

*PDCD4* gene expression was analysed by checking different publicly available BE microarray studies using the Oncomine database and gene microarray data analysis tools.\(^26\)\(^-\)\(^27\) The analysis considered the mRNA expression levels for each of the studies involved, and the significance of the expression of the gene across the studies considered was also taken into account. In the three independent data sets of human Barrett's oesophageal carcinogenesis considered, *PDCD4* mRNA expression levels decreased significantly when primary BAc was compared versus BE and versus native squamous epithelium (Pearson's correlation p=1.98×10\(^{-4}\))\(^{31\text{–}33}\) (figure 1).

**PDCD4 expression is downregulated in BAc**

The normal basal squamous epithelial layer (proliferative zone) consistently featured strong PDCD4 nuclear expression (mostly coexisting with weak–moderate cytoplasmic staining; figures 2 and 3). In normal and cancer tissue samples, non-cancerous and non-epithelial cells (fibroblasts, lymphocytes, smooth muscle cells and endothelia) always featured PDCD4 nuclear expression (this finding was considered as a positive internal control and it did not interfere with the histological assessment).\(^19\)\(^\)\(^22\)\(^\)\(^34\)\(^\)\(^35\) PDCD4 nuclear downregulation was consistently found in dysplastic and neoplastic lesions (Kruskal–Wallis p<0.001). In particular, moderate–strong nuclear staining was always associated with non-intestinal columnar metaplasia and intestinal metaplasia (scoring 2/3 in 24/25 and 25/25 cases, respectively), whereas intraepithelial (LG and HG) and invasive neoplasia showed a significantly lower PDCD4 nuclear expression (scores 2/3: 5/16, 0/12 and 0/10, respectively). Weak–moderate cytoplasmic staining was apparent in LG-IEN (scores 1/2: 15/16); HG-IEN and BAc samples showed weak cytoplasmic or no protein expression (score 1: 10/12 and 4/10, respectively). Overall, PDCD4 levels decreased progressively and significantly with the dedifferentiation of the lesions considered (Kruskal–Wallis p<0.001).

**miR-21 is overexpressed in HG-IEN and BAc**

It has been demonstrated that *PDCD4* is regulated directly by the oncomiR miR-21, with a 100% match sequence at nt228-249 (figure 4A).\(^14\)\(^-\)\(^15\) The *PDCD4* 3'-UTR miR-21 target sequence is highly conserved among different species,\(^14\)\(^-\)\(^24\) so we investigated miR-21 expression in preneoplastic and neoplastic Barrett’s lesions. When we determined miR-21 expression levels in 14 N, 5 HG-IEN and 11 BAc samples (figure 4B), a significant miR-21 upregulation was found in the HG-IEN (with a 2.3-fold increase; t test p<0.001) and BAc (with a twofold increase; t test p<0.001).

**DISCUSSION**

The progression of BM to BAc passes through well-established histological changes, in which dysplasia is considered the most advanced phenotypic change before invasive cancer sets in.\(^1\)\(^\)\(^2\)\(^-\)\(^3\)\(^6\) However, not all dysplastic lesions progress to cancer, and a better understanding of the molecular biology of this disease may lead to improvements in the diagnosis, treatment and prognosis of patients with BE.\(^3\)\(^7\)

*PDCD4* has been characterised as a new tumour suppressor gene, which is downregulated in several human malignancies.\(^19\)\(^-\)\(^24\) Its overexpression inhibits neoplastic transformation in both cancer cell lines and in vivo models.\(^10\)\(^\)\(^38\)\(^\)\(^39\)

Low PDCD4 protein levels (assessed by western blot and qRT-PCR analyses) have been observed in oesophageal squamous cell carcinoma cell lines.\(^18\) In a large series of oesophageal tumours, we recently found that PDCD4 protein was completely lost or significantly reduced in almost 60% of the tumour samples

Figure 3 Representative PDCD4 immunostainings of non-cancer oesophageal tissues and preneoplastic/neoplastic lesions in the Barrett’s carcinogenic cascade. In native squamous oesophageal epithelium, normal oesophageal basal cells feature strong nuclear and weak–moderate cytoplasmic staining. Non-intestinal columnar metaplasia (GM) and intestinal metaplasia (IM) preserved a strong nuclear immunostaining. Weak–moderate cytoplasmic immunostaining was seen in cases of low-grade intraepithelial neoplasia (HG-IEN), whereas in high-grade intraepithelial neoplasia (HG-IEN) and Barrett’s adenocarcinoma (BAc) samples showed weak cytoplasmic or no protein expression. Note the presence in the HG-IEN and BAc examples of PDCD4-nuclear positive residual GM/IM glands (black arrows) (original magnifications ×20 and ×40).
Programmed cell death 4 (PDCD4) is a 64 kDa protein that is downregulated in different human cancers, including oesophageal tumours.

The oncogenic microRNA miR-21 (hsa-miR-21) has been demonstrated to specifically target the PDCD4 3’-UTR, negatively regulating PDCD4 expression.

PDCD4 expression is significantly downregulated during Barrett’s carcinogenesis.

PDCD4 and miR-21 are novel molecular targets of Barrett’s carcinogenesis.

![Figure 4](image_url)
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ELISA in patient’s CSF was non-suggestive, but IgM was high. The negative serological results could be false-negative, as the specificity of the test is low. The sensitivity and specificity for IgM and IgG antibody range from 93 to 100% and 77 to 90%, respectively. PCR detection of Toxoplasma DNA in CSF increases both the sensitivity and specificity with a positive predictive value of 100%.

Various distribution patterns of intracerebral and periventricular calcification and diffuse hydrocephalus have been described on CT, but the occurrence of periventricular cysts has not been reported in the literature. Congenital toxoplasmosis has been extensively studied in murine models to elucidate the mode of infection and its effects. The lesions result from (1) destruction of parasitised cells, mainly tachyzoites, (2) tissue necrosis from rupture of cysts, (3) infarction necrosis due to vascular involvement and (4) periaqueductal and periventricular vasculitis with necrosis. The histopathological lesions herein predominantly revealed extensive periaqueductal and periventricular involvement with formation of cysts, which has not been previously described. The lesions were chronic in nature with numerous cortical and periventricular cysts, and the formation of subependymal cysts herniating into the ventricles can reveal jelly-like coagulum within its lumen. Demonstration of T gondii cysts within the tissue sections, or smears of the ventricular fluid or detection of its DNA amplification in the blood, body fluids or tissue sections is diagnostic.

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CORRECTION

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Matteo Fassan, Marco Pizzi, Giorgio Battaglia, Luciano Giacomelli, Paola Parente, Paolo Bocus, Ermanno Ancona, Massimo Rugge. Programmed cell death 4 (PDCD4) expression during multistep Barrett’s carcinogenesis. J Clin Pathol 2010;63:692—6. The sentence on page 694 should read: The normal basal layer featured moderate cytoplasmic staining, never associated with PDCD4 nuclear expression. The supra-basal squamous layers consistently featured PDCD4 nuclear expression (also coexisting with weak/moderate cytoplasmic staining; figures 2 and 3).

The legend to Figure 3 should read: In native squamous oesophageal epithelium, normal oesophageal supra-basal cells feature strong nuclear and concomitant weak/moderate cytoplasmic staining.