Microbial ‘old friends’, immunoregulation and socioeconomic status

Summary

The immune system evolved to require input from at least three sources that we collectively term the ‘old friends’: (i) the commensal microbiotas transmitted by mothers and other family members; (ii) organisms from the natural environment that modulate and diversify the commensal microbiotas; and (iii) the ‘old’ infections that could persist in small isolated hunter-gatherer groups as relatively harmless subclinical infections or carrier states. These categories of organism had to be tolerated and co-evolved roles in the development and regulation of the immune system. By contrast, the ‘crowd infections’ (such as childhood virus infections) evolved later, when urbanization led to large communities. They did not evolve immunoregulatory roles because they either killed the host or induced solid immunity, and could not persist in hunter-gatherer groups. Because the western lifestyle and medical practice deplete the ‘old’ infections (for example helminths), immunoregulatory disorders have increased, and the immune system has become more dependent upon microbiotas and the natural environment. However, urbanization maintains exposure to the crowd infections that lack immunoregulatory roles, while accelerating loss of exposure to the natural environment. This effect is most pronounced in individuals of low socioeconomic status (SES) who lack rural second homes and rural holidays. Interestingly, large epidemiological studies indicate that the health benefits of living close to green spaces are most pronounced for individuals of low SES. Here we discuss the immunoregulatory role of the natural environment, and how this may interact with, and modulate, the proinflammatory effects of psychosocial stressors in low SES individuals.

Keywords: host–pathogen interactions, inflammation, regulatory T cells
FOCUS ON HYGIENE HYPOTHESIS AND APPROACHES TO MODULATING THE MICROBIOME

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hypothesis or the ‘old friends’ mechanism [2]. The ‘old friends’ mechanism should now be seen as one component of the broad spectrum of interactions between mammals and their microbial environment [3]. At one end of the spectrum are the endosymbiotic events that led to the organelles in mammalian eukaryotic cells. There are also endogenous viruses and retrotransposons that lurk in our tissues and form major components of our DNA [4]. Next, in terms of their interdependency with human physiology, and particularly important in the context of this paper, are the various microbiotas (gut, skin, airway, oropharyngeal, genitourinary). These microbiotas perform significant parts of mammalian metabolism and contain about 100-fold more genes than does the human genome [5]. Metabolomic analyses show that much of ‘our’ metabolism is microbial [6]. The commensal microbiotas are also involved in development of mammalian organ systems, including gut, immune system, bone and brain (reviewed in [3]). The brain provides an example relevant to this paper. The brain of germ-free mice has altered chemistry and gene expression, and the animals behave abnormally [7]. The hypothalamic–pituitary–adrenal (HPA) axis of germ-free animals is also abnormal, manifested as altered central nervous system (CNS) gene expression and abnormal responses to stress [8,9]. To correct these abnormalities it is necessary to reconstitute the gut microbiota with appropriate organisms within the first 6 weeks of life [7,8]. Moving along the spectrum of human–microbe interdependency, we next have to consider our encounters with harmless organisms from the natural environment, present in large quantities in air, soil and water [10]. Finally, we come to the infections, the type of microbial interaction initially implicated by the hygienic hypothesis.

This paper is focused upon four questions: (i) why do some microbial exposures regulate the immune system (in addition to triggering its development in the neonate); (ii) which categories of organism have this immunoregulatory property; (iii) what relevance does this immunoregulation have to depression and stress resilience; and (iv) how do these mechanisms interact with the clear influence of socio-economic status (SES) on many aspects of health? We emphasize that immunoregulation by microbial exposures is no longer a ‘hypothesis’. We will not discuss the existence of these effects, but rather their relative importance, because they certainly do not act alone.

Which microbial exposures regulate the immune system, and why do they do it?

At birth the immune system has genetically inherited mechanisms, but it lacks data. It has some knowledge of self, acquired as lymphocytes mature in the thymus, and minimal knowledge of the outside world, transferred from the mother across the placenta. After birth it needs microbial exposures for at least three reasons. The first two are somewhat obvious and are part of the microbe-dependent signals for the development of the immune system mentioned in the previous paragraph. Exposure to a broad biodiversity of organisms builds up a memory of diverse molecular structures that accelerates subsequent rapid recognition of novel dangerous organisms [11,12]: this works because all life forms share fundamental molecular building blocks [3]. Secondly, microbial components such as peptidoglycans and lipopolysaccharide (LPS) taken in from the gut maintain background activation of the innate immune system [13].

It is less obvious that microbial exposures also play a role in setting up the control mechanisms that stop the immune system from causing inappropriate inflammatory responses. Thus, the system needs to develop a network of regulatory pathways and regulatory T cells (Treg) that stop inappropriate immune attacks on self, harmless allergens and gut contents, including harmless microorganisms (Fig. 1). These are the pathological targets involved in autoimmune diseases [14,15], allergic disorders [16] and inflammatory bowel diseases (IBD) [17], respectively, all of which are increasing in prevalence in high-income countries. These down-regulatory mechanisms must also be able to shut off inflammatory responses that are no longer needed, because chronically raised background inflammation, easily documented as raised C-reactive protein (CRP) or interleukin (IL)-6, correlates with a multitude of health problems, including cardiovascular disease, depression and reduced stress resilience [18,19], all of which are increasing problems in modern urbanized high-income countries. In high-income communities there are often persistent high background levels of CRP in the absence of any demonstrable reason for ongoing inflammation. In sharp contrast, a longitudinal study in a low-income country revealed that large peaks of CRP accompanied episodes of infection (therefore, cross-sectional studies suggest higher CRP levels in low- than in high-income settings), but after resolution of the infection CRP levels fell to values close to zero [20]. Thus, in the underdeveloped low-income setting, inflammation appears when needed but is completely shut off when not needed, whereas in high-income urban communities there can be a failure to shut off unwanted inflammation.

Which categories of organism have an immunoregulatory role?

Which categories of organism have an immunoregulatory role? The short answer is that the immunoregulatory organisms are those with which we co-evolved, and that had to be tolerated. These include the microbiotas (gut, airway, skin, genitourinary, oropharyngeal) which, as commensals performing crucial physiological functions, clearly must not be attacked [12,21–23]. We also evolved to tolerate harmless organisms (bacteria, archaea, fungi, viruses, protozoa, etc.) from the natural environment in water, air and soil because
these were inevitably taken into the body daily in large quantities (some of these might, in fact, become incorporated into the microbiota, as discussed later). Finally, we evolved to tolerate a specific subset of infections from our evolutionary past, sometimes referred to as the ‘old infections’ [24,25]. The crucial feature of the ‘old infections’ was their ability to persist in small isolated human hunter-gatherer groups, because they caused persistent non-fatal carrier states or subclinical disease (Fig. 2). These ‘old infections’ include ‘paleolithic’ strains of *Mycobacterium tuberculosis* that were less pathogenic that modern ones [25], helminths including gut parasites, but also blood nematodes that never enter the gut [26] and *Helicobacter pylori* (reviewed in [2]). Infections such as blood nematodes are not always harmless but once established, attempts by the immune system to eliminate them merely cause pointless immunopathology, leading to complications such as elephantiasis [26].

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**Fig. 1.** The immune system requires ‘educational’ input. The microbiota of others, organisms from the natural environment and other tolerated organisms (such as helminths) with which we co-evolved are required to expand the effector and regulatory branches of the immune system. During subsequent encounters with pathogens, danger signals generated by tissue damage enhance effector mechanisms and attenuate regulatory pathways to permit an appropriate immune response. Adequate background levels of regulatory T cells and dendritic cells and other regulatory mechanisms are required to maintain suppression of responses to ‘forbidden targets’ and to switch off inflammation completely when the danger is eliminated, so that proinflammatory mediators do not continue to circulate.

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**Fig. 2.** A simplified scheme that divides organisms into categories based on whether or not they co-evolved with humans and needed to be tolerated, and so developed immunoregulatory roles within the immune system. The ‘old’ infections were able to persist in isolated hunter-gatherer groups as carrier states and latent infections, and so evolved the ability to modulate the immune system. The microbiota also needed to be tolerated, and an unknown subset of organisms within the commensal microbiota is derived from the environment, including animal sources. The crowd infections evolved relatively recently and either kill the host or immunize, and constitute the only category that is increased rather than decreased in high-income settings. Epidemiological studies show that the crowd infections are not immunoregulatory.

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**Table 1.** Microbial exposures and high-income countries.

| ‘Old’ infections | High-income countries | Known effects on immune system |
|------------------|-----------------------|--------------------------------|
| can persist in small hunter-gatherer groups | mostly lost | inflammation progressing to life-long equilibrium with host |
| (helminths, gut infections and blood nematodes), S. Typhimurium, *H. pylori*, *Mycobacteria*, toxoplasmosis, etc. | | Treg adjuvant effect |
| **Commensal microbiota** | expanded Treg population, regulatory B cells, anti-inflammatory cytokines |
| (skin, gut, airway, oropharyngeal, genitourinary) | | background activation of innate immune system |
| **Environmental microbiota** | expanded via TLR-2 or TGF-β receptor, modulate ratio of Treg to Th17, drive regulatory DC, anti-inflammatory short chain fatty acids, etc. |
| (animals, soil, spores, air, plants – rhizosphere, phyllosphere) | | increased diversity and exposure |
| **Crowd infections** | increased...especially in inner cities | inflammation |
| (kill or immunize, cannot persist in hunter-gatherer groups, recently evolved childhood virus infections: measles, etc.) | | elimination some susceptible genotypes (eg HLA types); kill host or elicit solid immunity |

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Mechanisms of immunoregulation

We are only just beginning to understand some of the mechanisms involved in this microbe-driven immunoregulation. Some ‘old friends’ (including members of the human gut microbiota such as Bacteroides fragilis), or molecules that they secrete, are known to specifically expand Treg populations [22,27–29], or to cause dendritic cells (DC) to switch to regulatory DC that preferentially drive immunoregulation [30,31]. The latter mechanism implies a ‘Treg adjuvant’ effect. For example, when patients suffering from early relapsing multiple sclerosis (MS) become infected with helminths the disease stops progressing, and circulating myelin-recognizing Treg appear in the peripheral blood [32,33], an exciting observation that has led to formal clinical trials [34]. Release of immunoregulatory molecules is not confined to organisms in the gut, given that such activities have also been isolated from blood nematodes [35,36]. Another recent suggestion is that a certain level of the proinflammatory cytokine tumour necrosis factor (TNF)-α is required to drive apoptosis of potentially autoreactive T cells, and that in the absence of sufficient TNF-releasing infections such T cells can persist to cause type 1 diabetes (T1D) or multiple sclerosis [37]. Numerous other regulatory mechanisms are discussed in recent publications [22]. The relevance of these regulatory mechanisms for chronic inflammatory disorders is now well supported by data from animal models. Given their regulatory effects, it is not surprising that ‘old friends’ can be shown to drive immunoregulation, and to block or treat pathophysiology in models of allergies, autoimmune disease and IBD [29,38,39].

Non-immunoregulatory crowd infections

These old infections are very different from the ‘crowd infections’ that started to infect man after the First Epide-miological Transition, when the Neolithic revolution led to agriculture and large settlements and eventually to urbanization (category 4 in Fig. 2). The ‘crowd infections’ are mainly viruses such as measles that could not persist in sparsely distributed hunter-gatherer bands, because they either killed the host or induced solid immunity [24]. Crowd infections need large populations and networks of social contacts, so that the infection can return to cause an epidemic when herd immunity declines [24]. Such populations did not exist until well after 10 000 BCE, when agriculture and permanent settlements initiated the transition to larger population densities. Thus humans did not co-evolve with the crowd diseases, and the crowd diseases did not need to be tolerated (they killed the host or generated solid immunity) so, as anticipated, they play little role in setting up immunoregulatory pathways. Epidemiological studies have confirmed that the crowd infections do not protect children from allergic disorders [40–42] and often, in fact, trigger them [43]. Crowd infections also fail to protect from autoimmunity and IBD [44–47]. Thus the crowd infections do not explain the original ground-breaking observations of Strachan that led to the coining of the misleading term ‘hygiene hypothesis’ [1]. The crowd infections can, of course, impact upon human evolution. For instance, they may eliminate certain susceptible genotypes or major histocompatibility complex (MHC) phenotypes. However, sporadic dangerous infections do not co-evolve essential ongoing roles in mammalian physiology; but the ‘old friends’ (the old infections, and the commensal and environmental microbiota) that accompanied human evolution do have crucial immunoregulatory roles that are disturbed by the modern urban lifestyle. What aspects of that lifestyle are important?

Loss of contact with ‘old friends’ in high-income countries

It is of interest to summarize the lifestyle changes that reduce contact with the ‘old friends’ in high-income settings where chronic inflammatory disorders are increasing [48], because it casts light upon the relative immunoregulatory importance of the different groups of organisms. First, as reviewed in detail elsewhere [19], the chronic inflammatory disorders tend to increase when populations immigrate from low-income to developed high-income countries, and within any given country they tend to be more common in urban than in rural communities. This is equally true for psychiatric disorders, including autism, schizophrenia and depression (reviewed in [19]). It is obvious that these lifestyle changes will deplete the old infections, listed as category 1 in Fig. 1. For example, helminths are almost eliminated from high-income cities.

Perhaps of greater interest, and less fully understood, are the lifestyle changes that reduce contact with the commensal microbiota of others (category 2 in Fig. 2) and the microbiota of the natural environment (category 3 in Fig. 2).

Caesarean section, birth order, antibiotics and diet

Birth by caesarean section causes development of the gut microbiota to be delayed and to take an unusual course [49] (Fig. 3). This has been linked to increased T1D [50,51] and asthma [52,53], while increased contact with maternal microbiota may be protective [54]. Caesarean birth also increases the risk of coeliac disease [55], but there is little or no effect on the risk of IBD [55–57]. Birth order also modulates the gut microbiota [58], and this probably accounts for the protection against allergic disorders attributable to having older siblings [58] reported originally by Strachan [1].

Antibiotics, particularly excessive antibiotic use during pregnancy or the neonatal period, induces changes in the
microbiota that have been implicated in asthma [59,60], cow’s milk allergy [61], irritable bowel syndrome [62], IBD [63] and obesity [64].

Typical modern western diets have also been shown to exert profound effects on the microbiota [65]. For example, the gut microbiota of Italians is very different from that of people living in traditional villages in Burkina Faso [66], and the authors attributed this difference to diet because the African microbiomes contained genes for the hydrolysis of cellulose and xylan, abundant in African but not in Italian diets [66]. Similarly, in faeces from 98 Americans, enterotypes (bacterial ecosystems in the gut microbiota) correlated with long-term dietary habits, particularly the consumption of animal fat versus carbohydrates [67].

The natural environment and animals

In addition to factors discussed thus far, the composition of the commensal microbiota also depends upon input from the natural environment (reviewed in [10]). The skin microbiota from individuals living close to agricultural land in Finland was more diverse than that from individuals living close to urban centres, and was associated with reduced atopic sensitization [21]. Moreover, the ability of exposure to the farming environment early in life to protect from asthma seems to correlate closely with airborne fungal and bacterial biodiversity in children’s bedrooms [68]. Similarly, the biodiversity of the gut microbiota was greater in Amazonian Amerindians than in Malawians, while the biodiversity of gut microbiota from the United States was lower than that of either of these low-income rural groups [69]. Clearly, diet could play a role, but recent experiments with piglets suggest the importance of environmental inputs. The gut microbiota of piglets that were housed in a natural outdoor environment was rich in Firmicutes, particularly Lactobacillus strains, whereas the hygienic indoor piglets had reduced Lactobacilli and more potentially pathogenic phylotypes [70]. Moreover, biopsies of the gut mucosa revealed that these ‘indoor’ piglets had increased type 1 interferon activity, increased MHC class 1 and up-regulation of many chemokines [70], implying a more inflammatory state in the guts of animals whose microbiota had not been modified and diversified by exposure to the natural environment. In mice [71] and humans, a correlation between reduced gut microbial biodiversity and poor control of inflammation is a common finding [72–74]. It therefore seems likely that microbial inputs are required to maintain diversity of the gut microbiota, and that such diversity plays a role in the regulation of inflammation, although at present the relationship between environmental strains and colonizing strains is not documented (discussed in [10]). Data on gut bacteria are accumulating rapidly, so for these organisms the issue may soon be addressed, but data for bacteriophages and other viruses or for archaea and fungi are only just beginning to appear [23,75].

Animals, faeces and spores

Contact with animals might provide part of the explanation for this environmental input. The protective effect of the farming environment was noted in the 19th century [76], and has been associated with cowsheds [77]. Contact with dogs, with which humans have co-evolved for many millennia [78], also protects from allergic disorders [79,80], and people seem to share their microbiota via dogs [81], which greatly increase the microbial biodiversity of the home [82,83]. In a developing country the presence of animal faeces in the home correlated with better ability to control background inflammation (CRP levels) in adulthood [20], and in Russian Karelia (where the prevalence of childhood atopy is four times lower and T1D is six times lower than in Finnish Karelia), house dust contained a sevenfold higher number of clones of animal-associated species than was present in Finnish Karelian house dust [84].
It is interesting to speculate here that these effects have something to do with faeces and spores. Approximately one-third of the bacteria in the gut microbiota are spore-forming, and spores are readily demonstrable in human faeces [85]. Spores are remarkably resistant and can remain viable for thousands, possibly millions of years (reviewed in [86]). It has been calculated that many billions of tonnes of animal and human faeces are generated every year, so faeces-derived spores accumulate in the natural environment. Human faeces average up to $10^4$ spores/g, while soil contains approximately $10^6$ spores/g [87]. However, spores in soil have tended to be studied by environmental microbiologists and ecologists, and the soil has been regarded as the natural habitat of spore-forming organisms such as *Bacillus* spp., despite awareness that many of them can germinate and replicate in the intestinal tracts of insects and other animals [86] [some of these are toxic to the insect, and so are used as biological insecticides (reviewed in [86])]. Recently it has been reported that spores of *Bacillus subtilis* can germinate in the small bowels of mice and rabbits [88–90]. Moreover, after germination they replicated in the small bowel and then respored as they entered the colon. The same occurs in humans. *B. subtilis* strains were obtained from biopsies of human ileum and from faecal samples [85]. Most of these strains are able to form biofilms, sporulate anaerobically and secrete antimicrobials, properties that could facilitate survival in the gut [85]. There is therefore a growing view that *B. subtilis* and other environmental spore-forming species are gut commensals rather than soil microorganisms [87]. This might be very relevant to the ‘old friends’ mechanism, particularly to the clear importance of exposure to animals, agricultural land and green spaces. For example, *B. subtilis* is an important stimulus for development of the gut-associated lymphoid tissue (GALT) in rabbits, and sporulation of live bacilli within the GALT is considered critical to this process [90]. Interestingly, *B. subtilis* is capable of driving GALT in synergy with *B. fragilis*, an organism that also drives formation of murine GALT, and secretes a polysaccharide antigen that drives proliferation of Treg [22].

**Inflammation and SES**

Of course, this does not mean that we can attribute all urban health deficits to lack of immunoregulatory microbial exposures, so to what extent can we do so? Many factors are linked to increased background levels of inflammation (measured as CRP or IL-6) in people of low SES. These include unhealthy behaviours such as smoking, drugs, poor diet and obesity, as well as psychosocial stressors such as violence, distrust, neglect, poverty and crime [97–99]. For example, CRP levels of 3 mg/l or more are found in increased percentages of children living in areas with high levels of poverty or crime [100].

At least some of this predisposition to inflammation is developmental, and is established during the perinatal period. Perinatal stress results in adults who themselves show exaggerated inflammatory responses to stress [101–104]. For example, maltreated children develop higher levels of IL-6 in response to a standardized social stressor (the Trier Social Stress Test; TSST) when tested as adults in comparison to a non-maltreated control group [101,105], and maltreated children tend to have higher levels of CRP 20 years later [103].

Interestingly, negative life events during the first years of life, whether they affect the child directly or indirectly via traumatic experiences of the mother, also predispose to some of the chronic inflammatory disorders that are increasing in high-income countries and that are associated more traditionally with the ‘hygiene hypothesis’. For example, the autoimmune disease T1D is increased in low SES children ([106,107], reviewed in [108]). Low SES children also tend to have more severe asthma, accompanied by increased expression of proinflammatory pathways [109].

In view of the large literature on urban–rural differences in chronic inflammatory and psychiatric disorders (reviewed in [19]), and on the protective effects of close exposure [77,96] or even mere proximity [21] to the farming environment or to airborne microbial biodiversity [68], it is strange that the likely connection between the ‘green space’ effect and the ‘hygiene hypothesis’ or ‘old friends’ mechanism has only recently been made. Urban populations are heavily exposed to the crowd infections, but as pointed out above, we did not co-evolve with the crowd infections, they did not co-evolve roles in setting up our immunoregulatory mechanisms and exposure to them does not protect against the chronic inflammatory disorders [40–47]. Conversely, urban populations are deprived of exposure to the natural and agricultural environment. This is particularly true of low SES individuals who perhaps do not have rural secondary homes or holiday travel to rural settings. This point, which needs verification by epidemiologists, could explain why the beneficial effect of proximity to green space is strikingly more marked at the lower end of the socioeconomic scale [91–93].
In some settings IBD is also more common in low SES children [110]. Similarly SES-associated inflammation partly explains the SES-related incidence of type 2 diabetes [111]. These observations imply a broad-based immunoregulatory problem in low SES communities.

There are several explanations for this effect. First, prenatal stress causes long-term alterations in the HPA axis function, both in animal models [112,113] and in humans exposed to prenatal [114] or early childhood stress [115], or to a childhood background of low SES [97]. Secondly, prenatal stress has effects on the microbiota of rats and rhesus monkeys that persist into adulthood [116,117], and it is likely that the same is true in humans, although this has not, to our knowledge, been documented. However, we do know that in humans fluctuations in the microbiota early after surgery may lead to an increased risk of immunoregulatory failure, manifested in this clinical group as graft-versus-host disease [118], and changes in the microbiota of severely stressed critically ill humans are rapid and prolonged [119]. The nature of the microbiota, as discussed earlier, modulates development of the brain and HPA axis [7–9] and the microbiota plays a major role in the inflammatory response to stress [9]. Much emphasis has been placed upon the role of poor diet and obesity in the health deficits associated with low SES settings, and indeed obesity has proinflammatory effects, but much of this is again mediated via the microbiota [120]. These, and no doubt other mechanisms that are beyond the scope of this review, cause low SES urban children to have poor regulation of inflammation that predisposes them to cardiovascular disease, depression and metabolic syndrome [97–99,111] and other chronic inflammatory disorders [108–110].

The ‘old friends’ mechanism and inflammation in low SES settings

Does the ‘old friends’ mechanism have anything to do with the increased inflammation observed in low SES settings? There are a number of ways in which these consequences of low SES might overlap with, and be modified or exaggerated by, the ‘old friends’ mechanism. For example, the release of inflammatory mediators is subject to negative immunoregulatory feedback. Therefore, poor immunoregulation will lead to exaggerated inflammatory responses. Because low SES inner-city children lack exposure to all the categories of immunoregulatory organisms except the microbiota of others, but are bombarded by the non-immunoregulatory crowd infections, they are at risk for frequent inflammatory responses (Fig. 3). If these occur during pregnancy, the increased levels and persistence of inflammatory mediators may predispose to developmental abnormalities of the brain, such as those that underlie some cases of schizophrenia or autism [121] and other neurological disorders [122]. Essentially, all the common crowd infections are associated epidemiologically with these neurological/psychiatric disorders when the infection occurs during pregnancy [121–123], and abnormal brain development is seen when inflammation is provoked during pregnancy in monkeys [124] or rodents [125]. A similar mechanism might explain the associations of low SES with T1D ([106,107], reviewed in [108]), asthma [109] and IBD, as mentioned above [110].

An immunoregulatory deficit is equally relevant to inflammation driven by psychosocial stressors. In the absence of effective priming of immunoregulation by ‘old friends’, a given level of psychosocial stress might be expected to cause a greater release of inflammatory mediators (Fig. 4), which predispose towards [18,126], and can drive, psychiatric symptoms [127,128], whereas exposure to ‘old friends’ might be expected to reduce cytokine levels and so increase stress resilience [19]. Recent studies in the Philippines have found that even a childhood trauma as severe as maternal deprivation can fail to result in a raised background CRP in adulthood in those individuals who were heavily exposed to a microbe-rich environment and animal faces in childhood [129]. In US citizens such adverse childhood events tend to have serious consequences for later health, as outlined above [101–105]. Similarly, recent psychosocial stress did not cause detectable rise in CRP in these adults who had received heavy microbial exposures as infants [129]. In a fascinating further twist it was found that, in this community, depressive symptoms were not
associated with raised biomarkers of inflammation [130], whereas depressive symptoms are often associated with raised biomarkers of inflammation in high-income countries [126,127,131]. In view of the growing awareness of the relevance of contact with animals and spores outlined earlier, this finding is intriguing and suggests that in low-income settings exposure to animal-derived microbes might improve regulation of inflammation, and so increase stress-resilience, although this observation clearly needs to be confirmed in other populations.

Conclusions and critique

Much confusion has arisen from the assumption that multiple older siblings provide protection from allergic disorders because they transmit the common childhood infections [1], whereas it now seems more likely that this protection is attributable to exchange of commensal microbiota [58]. This paper is an attempt to show that a broad evolutionary and biological approach can provide a framework that makes sense of several paradoxes, including the fact that inner-city children, in whom childhood infections are rife, tend to have more rather than fewer inflammatory disorders, including allergies. If we consider our microbiological encounters in terms of our evolutionary history, and the nature of our relationships to each category of organism, we can see that low SES urban children are at the extreme end of a spectrum of loss of the organisms with which we co-evolved (Fig. 3). A second objective of the paper is to point out that these effects have little or nothing to do with hygiene [132]. Indeed, increased hygiene in the low SES home might help to protect from the crowd infections, while also protecting from ‘new enemies’ such as gastroenteritis-inducing Escherichia coli strains and Campylobacter. Our third objective is to illustrate how the low SES lifestyle, including the psychosocial stressors that it entails, will synergize with the consequences of the ‘old friends’ mechanism to increase susceptibility to a broad range of inflammation-dependent health problems. We hope that these insights will help to focus attention on the need for increased hygiene, coupled with innovative design for homes and sustainable cities that promote appropriate microbial exposures. Meanwhile, there is enormous need for a greater understanding of the relationship between organisms in the natural environment and those that colonize us so that we can optimize the design of urban green spaces.

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Disclosure

C. L. R. reports the following activities for the previous 2 years: advisory board participation and related travel funds for Pamlab, Lilly and North American Center for Continuing Education; development and presentation of disease state slides for Pamlab, Pfizer and Johnson & Johnson, as well as related travel funds for these activities; development of continuing medical education material for North American Center for Continuing Education and for CME Incite.

References

1 Strachan DP. Hay fever, hygiene, and household size. BMJ 1989; 299:1259–60.
2 Rook GAW. 99th Dahlem conference on infection, inflammation and chronic inflammatory disorders: Darwinian medicine and the ‘hygiene’ or ‘old friends’ hypothesis. Clin Exp Immunol 2010; 160:70–9.
3 McFall-Ngai M, Hadfield MG, Bosch TC et al. Animals in a bacterial world, a new imperative for the life sciences. Proc Natl Acad Sci USA 2013; 110:3229–36.
4 Holmes EC. The evolution of endogenous viral elements. Cell Host Microbe 2011; 10:568–77.
5 O’Hara AM, Shanahan F. The gut flora as a forgotten organ. EMBO Rep 2006; 7:688–93.
6 Wikoff WR, Anfora AT, Liu J et al. Metabolomics analysis reveals large effects of gut microbiota on mammalian blood metabolites. Proc Natl Acad Sci USA 2009; 106:3698–703.
7 Heijtz RD, Wang S, Anuar F et al. Normal gut microbiota modulates brain development and behavior. Proc Natl Acad Sci USA 2011; 108:3047–52.
8 Sudo N, Chida Y, Aiba Y et al. Postnatal microbial colonization programs the hypothalamic–pituitary–adrenal system for stress response in mice. J Physiol 2004; 558:263–75.
9 Bailey MT, Dowd SE, Galley JD, Hufnagle AR, Allen RG, Lyte M. Exposure to a social stressor alters the structure of the intestinal microbiota: implications for stressor-induced immunomodulation. Brain Behav Immun 2011; 25:397–407.
10 Rook GAW. Regulation of the immune system by biodiversity from the natural environment: an ecosystem service essential to health. Proc Natl Acad Sci USA 2013; 110:18360–7.
11 Su LF, Kidd BA, Han A, Kotzin JJ, Virus-specific DMM. CD4(+) memory-phenotype T cells are abundant in unexposed adults. Immunity 2013; 38:373–83.
12 Naik S, Bouladoux N, Wilhelm C et al. Compartmentalized control of skin immunity by resident commensals. Science 2012; 337:1115–9.
13 Clarke TB, Davis KM, Lysenko ES, Zhou AY, Yu Y, Weiser JN. Recognition of peptidoglycan from the microbiota by Nod1 enhances systemic innate immunity. Nat Med 2010; 16:228–31.

14 Fleming JO. Helminth therapy and multiple sclerosis. Int J Parasitol 2013; 43:259–74.

15 Stene LC, Nafsstad P. Relation between occurrence of type 1 diabetes and asthma. Lancet 2001; 357:607.

16 Eder W, Ege MJ, von Mutius E. The asthma epidemic. N Engl J Med 2006; 355:226–35.

17 Elliott DE, Summers RW, Weinstock JV. Helminths and the modulation of mucosal inflammation. Curr Opin Gastroenterol 2005; 21:51–8.

18 Gimeno D, Kivimaki M, Brunner EJ et al. Associations of C-reactive protein and interleukin-6 with cognitive symptoms of depression: 12-year follow-up of the Whitehall II study. Psychol Med 2009; 39:413–23.

19 Rook GAW, Lowry CA, Raison CL. Microbial Old Friends, immunoregulation and SES. Out-of-Africa migration and modern humans. Nat Genet 2013; 45:1176–82.

20 Babu S, Blauvelt CP, Kumaraswami V, Nutman TB. Regulatory T cells induced by live parasites impair both Th1 and Th2 pathways in patent lymphatic filariasis: implications for parasite persistence. J Immunol 2006; 176:3248–56.

21 Hanski I, von Hertzen L, Fyhruistr N et al. Environmental biodiversity, human microbiota, and allergy are interrelated. Proc Natl Acad Sci USA 2012; 109:8334–9.

22 Round JL, Lee SM, Li J et al. The Toll-like receptor 2 pathway establishes colonization by a commensal of the human microbiota. Science 2011; 332:974–7.

23 Hoffmann C, Dolfie S, Grunberg S et al. Archaea and fungi of the human gut microbiome: correlations with diet and bacterial residents. PLOS ONE 2013; 8:e66019.

24 Wolfe ND, Dunavan CP, Diamond J. Origins of major human infectious diseases. Nature 2007; 447:279–83.

25 Comas I, Coscolla M, Liao T et al. Out-of-Africa migration and Neolithic coexpansion of Mycobacterium tuberculosis with modern humans. Nat Genet 2013; 45:1176–82.

26 Babu S, Blauvelt CP, Kumaaraswami V, Nutman TB. Regulatory networks induced by live parasites impair both Th1 and Th2 pathways in patent lymphatic filariasis: implications for parasite persistence. J Immunol 2006; 176:3248–56.

27 Grainger JR, Smith KA, Hewitson JP et al. Helminth secretions induce de novo T cell Foxp3 expression and regulatory function through the TGF-beta pathway. J Exp Med 2010; 207:2331–41.

28 Atarashi K, Tanoue T, Shima T et al. Induction of colonic regulatory T cells by indigenous clostridium species. Science 2011; 331:337–41.

29 Karimi K, Imman MD, Bienenscho J, Forsythe P. Lactobacillus reuteri-induced regulatory T cells protect against an allergic airway response in mice. Am J Respir Crit Care Med 2009; 179:186–93.

30 Smith H, Engering A, van der Kleij D et al. Selective probiotic bacteria induce IL-10-producing regulatory T cells in vitro by modulating dendritic cell function through dendritic cell-specific intercellular adhesion molecule 3-grabbing nonintegrin. J Allergy Clin Immunol 2005; 115:1260–7.

31 Hart AL, Lammers K, Brigid P et al. Modulation of human dendritic cell phenotype and function by probiotic bacteria. Gut 2004; 53:1602–9.

32 Correale J, Farez M. Association between parasite infection and immune responses in multiple sclerosis. Ann Neurol 2007; 61:97–108.

33 Correale J, Farez MF. The impact of parasite infections on the course of multiple sclerosis. J Neuroimmunol 2011; 233:4–11.

34 Fleming J, Isaac A, Lee J et al. Probiotic helminth administration in relapsing–remitting multiple sclerosis: a phase 1 study. Mult Scler 2011; 17:743–54.

35 Harnett MM, Melendez AJ, Harnett W. The therapeutic potential of the filamentous nematode-derived immunodulator, ES-62 in inflammatory disease. Clin Exp Immunol 2010; 159:256–67.

36 Kron MA, Metwali A, Vodanovic-Jankovic S, Elliott D. Nematode AsnRS resolves intestinal inflammation in murine T-cell transfer colitis. Clin Vaccine Immunol 2012; 20:276–81.

37 Faustman DL, Wang L, Okubo Y et al. Proof-of-concept, randomized, controlled clinical trial of bacillus Calmette–Guerin for treatment of long-term type 1 diabetes. PLOS ONE 2012; 7:e41756.

38 Round JL, Mazmanian SK. The gut microbiota shapes intestinal immune responses during health and disease. Nat Rev Immunol 2009; 9:313–23.

39 Osada Y, Kanazawa T. Parasitic helminths: new weapons against immunological disorders. J Biomed Biotechnol 2010; 2010:743–58.

40 Benn CS, Melbye M, Wohlfahrt J, Bjerkaas T, Aaby P. Cohort study of sibling effect, infectious diseases, and risk of atopic dermatitis during first 18 months of life. Br Med J 2004; 328:1223–8.

41 Dunder T, Tapiainen T, Pokka T, Uhari M. Infections in child day care centers and later development of asthma, allergic rhinitis, and atopic dermatitis: prospective follow-up survey 12 years after controlled randomized hygiene intervention. Arch Pediatr Adolesc Med 2007; 161:972–7.

42 Bremner SA, Carey IM, DeWilde S et al. Infections presenting for clinical care in early life and later risk of hay fever in two UK birth cohorts. Allergy 2008; 63:274–83.

43 Yoo J, Tcheurekdjian H, Lynch SV, Cabana M, Boushey HA. Microbial manipulation of immune function for asthma prevention: inferences from clinical trials. Proc Am Thorac Soc 2007; 4:277–82.

44 Cardwell CR, Carson DJ, Yarnell J, Shields MD, Patterson CC. Atopy, home environment and the risk of childhood-onset type 1 diabetes: a population-based case–control study. Pediatr Diabetes 2008; 9:191–6.

45 Amre DK, Lambrette P, Law L et al. Investigating the hygiene hypothesis as a risk factor in pediatric onset Crohn’s disease: a case–control study. Am J Gastroenterol 2006; 101:1005–11.

46 Bernstein CN, Rawsthorne P, Cheang M, Blanchard JF. A population-based case–control study of potential risk factors for IBD. Am J Gastroenterol 2006; 101:993–1002.

47 Koloski NA, Breit L, Radford-Smith G. Hygiene hypothesis in inflammatory bowel disease: a critical review of the literature. World J Gastroenterol 2008; 14:665–73.

48 Bach JF. The effect of infections on susceptibility to autoimmune and allergic diseases. N Engl J Med 2002; 347:911–20.

49 Dominguez-Bello MG, Costello EK, Contreras M et al. Delivery mode shapes the acquisition and structure of the initial microbiota across multiple body habitats in newborns. Proc Natl Acad Sci USA 2010; 107:11971–5.
FOCUS ON HYGIENE HYPOTHESIS AND APPROACHES TO MODULATING THE MICROBIOME

G. A. W. Rook et al.

50. Bonifacio E, Warncke K, Winkler C, Wallner M, Ziegler AG. Cesarean section and interferon-induced helicase gene polymorphisms combine to increase childhood type 1 diabetes risk. Diabetologia 2011; 54:3000–6.

51. Cardwell CR, Stene LC, Joner G et al. Cesarean section is associated with an increased risk of childhood-onset type 1 diabetes mellitus: a meta-analysis of observational studies. Diabetologia 2008; 51:726–35.

52. Thavagnanam S, Fleming J, Bromley A, Shields MD, Cardwell CR. A meta-analysis of the association between cesarean section and childhood asthma. Clin Exp Allergy 2008; 38:629–33.

53. Guibas GV, Moschonis G, Xepapadaki P et al. Conception via in vitro fertilization and delivery by Cesarean section are associated with paediatric asthma incidence. Clin Exp Allergy 2013; 43:1058–66.

54. Hesselmar B, Sjoberg F, Findeisen A et al. Cesarean section and interferon-induced helicase gene polymorphisms combine to increase childhood type 1 diabetes risk. Diabetes 2011; 60:3300–6.

55. Cardwell CR, Stene LC, Joner G et al. Cesarean section is associated with an increased risk of childhood-onset type 1 diabetes mellitus: a meta-analysis of observational studies. Diabetologia 2008; 51:726–35.

56. Bager P, Simonsen J, Nielsen NM, Frisch M. Cesarean section and childhood asthma. J Allergy Clin Immunol 2013; 132:1829–37.

57. Decker E, Engelmann G, Findeisen A et al. Cesarean delivery is associated with celiac disease but not inflammatory bowel disease in children. Pediatrics 2010; 125:e1433–40.

58. Penders J, Stobberingh EE et al. Establishment of the intestinal microbiota and its role for atopic dermatitis in early childhood. J Allergy Clin Immunol 2013; 132:601–7.

59. Metsala J, Lundqvist A, Virta LJ, Kaila M, Gissler M, Virtanen SM. Environmentally-acquired Bacillus cereus (hay-fever and hay-asthma). London: Baillière Tindall and Cox, 1873.

60. Russell SL, Gold MJ, Hartmann M et al. Infant antibiotic exposures and early-life body mass. Int J Obes (Lond) 2013; 37:16–23.

61. Metcalfe GR, Brown M, Bauman JE. Diet, gut microbiota and immunity. Nat Immunol 2011; 12:5–9.

62. VillaRreal AA, Aberg-Ber J, Benrud R, Gundrum JD. Use of broad-spectrum antibiotics and the development of irritable bowel syndrome. Wisconsin Med J 2012; 111:17–20.

63. Shaw SY, Blanchard JD, Bernstein CN. Association between the use of antibiotics in the first year of life and pediatric inflammatory bowel disease. Am J Gastroenterol 2010; 105:2687–92.

64. Trasande L, Blustein J, Liu M, Corwin E, Cox LM, Blaser MJ. Infant antibiotic exposures and early-life body mass. Int J Obes (Lond) 2013; 37:16–23.

65. Maslow KM, Mackay CR. Diet, gut microbiota and immune responses. Nat Immunol 2011; 12:5–9.

66. De Filippo C, Cavaliere D, Di Paola M et al. Impact of diet in shaping gut microbiota revealed by a comparative study in children from Europe and rural Africa. Proc Natl Acad Sci USA 2010; 107:14691–6.

67. Wu GD, Chen J, Hoffman C et al. Linking long-term dietary patterns with gut microbial enterotypes. Science 2011; 334:105–8.

68. Ege MJ, Mayer M, Normand AC et al. Exposure to environmental microorganisms and childhood asthma. N Engl J Med 2011; 364:701–9.
FOCUS ON HYGIENE HYPOTHESIS AND APPROACHES TO MODULATING THE MICROBIOME

Microbes, immunoregulation and SES

89 Tam NK, Uyen NQ, Hong HA et al. The intestinal life cycle of Bacillus subtilis and close relatives. J Bacteriol 2006; 188:2692–700.
90 Rhee KJ, Sethupathi P, Driks A, Lanning DK, Knight KL. Role of commensal bacteria in development of gut-associated lymphoid tissues and preimmune antibody repertoire. J Immunol 2004; 172:1118–24.
91 Maas J, Verheij RA, Groenewegen PP, de Vries S, Spreeuwenga P. Green space, urbanity, and health: how strong is the relation? J Epidemiol Community Health 2006; 60:387–92.
92 Mitchell R, Popham F. Effect of exposure to natural environment on health inequalities: an observational population study. Lancet 2008; 372:1655–60.
93 Dadvand P, de Nazelle A, Figueras F et al. Green space, health inequality and pregnancy. Environ Int 2012; 40:110–5.
94 Aspinall P, Mavros P, Coyne R, Roe J. The urban brain: analysing outdoor physical activity with mobile EEG. Br J Sports Med 2013; Mar 6. [Epub ahead of print].
95 Morris DW. Adaptation and habitat selection in the eco-evolutionary process. Proc Biol Sci 2011; 278:2401–11.
96 Radon K, Windstetter D, Poluda AL, Mueller B, von Mutius E, Koletzko S. Contact with farm animals in early life and juvenile inflammatory bowel disease: a case–control study. Pediatrics 2007; 120:354–61.
97 Miller GE, Chen E, Fok AK et al. Low early-life social class leaves a biological residue manifested by decreased glucocorticoid and increased proinflammatory signaling. Proc Natl Acad Sci USA 2009; 106:14716–21.
98 Chen E, Miller GE. Socioeconomic status and health: mediating and moderating factors. Annu Rev Clin Psychol 2013; 9:723–49.
99 Seaman T, Epel E, Gruenewald T, Karlamangla A, McEwen BS. Socio-economic differentials in peripheral biology: cumulative allostatic load. Ann NY Acad Sci 2010; 1186:223–39.
100 Broyles SL, Staiano AE, Dzabha KT, Gupta AK, Sothern M, Katzmarzyk PT. Elevated C-reactive protein in children from risky neighborhoods: evidence for a stress pathway linking neighborhoods and inflammation in children. PLOS ONE 2012; 7:e45419.
101 Carpenter LL, Gawuga CE, Tyrka AR, Lee JK, Anderson GM, Price LH. Association between plasma IL-6 response to acute stress and early-life adversity in healthy adults. Neuropsychopharmacology 2010; 35:2617–23.
102 Danese A, Moffitt TE, Pariante CM, Ambler A, Poulton R, Caspi A. Elevated inflammation levels in depressed adults with a history of childhood maltreatment. Arch Gen Psychiatry 2008; 65:409–15.
103 Danese A, Pariante CM, Caspi A, Taylor A, Poulton R. Childhood maltreatment predicts adult inflammation in a life-course study. Proc Natl Acad Sci USA 2007; 104:1139–24.
104 Entringer S, Kumsta R, Nelson EL, Hellhammer DH, Wadhwa PD, Wust S. Influence of prenatal psychosocial stress on cytokine production in adult women. Dev Psychobiol 2008; 50:579–87.
105 Pace TW, Miletzko TC, Alaghe O et al. Increased stress-induced inflammatory responses in male patients with major depression and increased early life stress. Am J Psychiatry 2006; 163:1630–3.
106 Sepa A, Frodi A, Ludvigsson J. Mothers’ experiences of serious life events increase the risk of diabetes-related autoimmunity in their children. Diabetes Care 2005; 28:2394–9.
107 Vlajinac H, Sipetic S, Marinkovic J, Bjekic M, Kocev N, Sajic S. The Belgrade childhood diabetes study – comparison of children with type 1 diabetes with their siblings. Paediatr Perinat Epidemiol 2006; 20:238–43.
108 Peng H, Hagopian W. Environmental factors in the development of Type 1 diabetes. Rev Endocr Metab Disord 2006; 7:149–62.
109 Chen E, Miller GE, Walker HA, Arevalo JM, Sung CY, Cole SW. Genome-wide transcriptional profiling linked to social class in asthma. Thorax 2009; 64:38–43.
110 Ekborg A, Adami HO, Helmick CG, Jonzon A, Zack MM. Perinatal risk factors for inflammatory bowel disease: a case–control study. Am J Epidemiol 1990; 132:1111–9.
111 Stringhini S, Batty GD, Bovet P et al. Association of lifetime socioeconomic status with chronic inflammation and type 2 diabetes risk: the Whitehall II prospective cohort study. PLoS Med 2013; 10:e1001479.
112 Kapoor A, Dunn E, Kostaki A, Andrews MH, Matthews SG. Fetal programming of hypothalamo–pituitary–adrenal function: prenatal stress and glucocorticoids. J Physiol 2006; 572:31–44.
113 Weinstock M. The potential influence of maternal stress hormones on development and mental health of the offspring. Brain Behav Immun 2005; 19:296–308.
114 Entringer S, Kumsta R, Hellhammer DH, Wadhwa PD, Wust S. Prenatal exposure to maternal psychosocial stress and HPA axis regulation in young adults. Horm Behav 2009; 55:292–8.
115 Heim C, Newport DJ, Heit S et al. Pituitary–adrenal and autonomic responses to stress in women after sexual and physical abuse in childhood. JAMA 2000; 284:592–7.
116 O’Mahony SM, Marchesi JR, Scully P et al. Early life stress alters behavior, immunity, and microbiota in rats: implications for irritable bowel syndrome and psychiatric illnesses. Biol Psychiatry 2009; 65:263–7.
117 Bailey MT, Lubach GR, Coe CL. Prenatal stress alters bacterial colonization of the gut in infant monkeys. J Pediatr Gastroenterol Nutr 2004; 38:414–21.
118 Jeng RR, Ubeda C, Taur Y et al. Regulation of intestinal inflammation by microbiota following allogeneic bone marrow transplantation. J Exp Med 2012; 209:903–11.
119 Hayakawa M, Asahara T, Henzan N et al. Dramatic changes of the gut flora immediately after severe and sudden insults. Dig Dis Sci 2011; 56:2361–5.
120 Karlsson F, Tremaroli V, Nielsen J, Backhed F. Assessing the human gut microbiota in metabolic diseases. Diabetes 2013; 62:3341–9.
121 Meyer U, Felden J, Dammann O. Schizophrenia and autism: both shared and disorder-specific pathogenesis via perinatal inflammation? Pediart Res 2011; 69:26R–33R.
122 Hagberg H, Gressens P, Mallard C. Inflammation during fetal and neonatal life: implications for neurologic and neuropsychiatric disease in children and adults. Ann Neurol 2012; 71:444–57.
123 Brown AS, Derkits EJ. Prenatal infection and schizophrenia: a review of epidemiologic and translational studies. Am J Psychiatry 2010; 167:261–80.
124 Willette AA, Lubach GR, Knickmeyer RC et al. Brain enlargement and increased behavioral and cytokine reactivity in infant monkeys following acute prenatal endotoxemia. Behav Brain Res 2011; 219:108–15.
125 Smith SE, Li J, Garbett K, Mirmics K, Patterson PH. Maternal immune activation alters fetal brain development through interleukin-6. J Neurosci 2007; 27:10695–702.
FOCUS ON HYGIENE HYPOTHESIS AND APPROACHES TO MODULATING THE MICROBIOME

G. A. W. Rook et al.

126 Valkanova V, Ebmeier KP, Allan CL. CRP, IL-6 and depression: a systematic review and meta-analysis of longitudinal studies. J Affect Disord 2013; 150:736–44.

127 Raison CL, Lowry CA, Rook GAW. Inflammation, sanitation and consternation: loss of contact with co-evolved, tolerogenic microorganisms and the pathophysiology and treatment of major depression. Arch Gen Psychiatry 2010; 67:1211–24.

128 Raison CL, Rutherford RE, Woolwine BJ et al. A randomized controlled trial of the tumor necrosis factor antagonist infliximab for treatment-resistant depression: the role of baseline inflammatory biomarkers. Arch Gen Psychiatry 2012; Sep 3:1–11.

129 McDade TW, Hoke M, Borja JB, Adair LS, Kuzawa CW. Do environments in infancy moderate the association between stress and inflammation in adulthood? Preliminary evidence from a birth cohort in the Philippines. Brain Behav Immun 2012; 31:23–30.

130 McDade TW, Borja JB, Adair L, Kuzawa CW. Depressive symptoms are not associated with inflammation in younger and older adults in the Philippines. Evol Med Public Health 2012; 2013(1):18–23.

131 Howren MB, Lamkin DM, Suls J. Associations of depression with C-reactive protein, IL-1, and IL-6: a meta-analysis. Psychosom Med 2009; 71:171–86.

132 Stanwell-Smith R, Bloomfield SF, Rook GAW. The hygiene hypothesis and its implications for home hygiene, lifestyle and public health. International Scientific Forum on Home Hygiene 2012. Available at: http://www.ifh-homehygiene.org/reviews-hygiene-hypothesis (accessed 10 February 2014).