INTRODUCTION

Diabetes mellitus represents a major public health threat worldwide. A serious complication of diabetes is the development of foot ulcers which, when they become infected, are the most common cause of diabetes-related hospital admissions and a leading cause of the lower extremity amputation. Diabetes with ulcers commonly experience infection with Gram-positive organisms such as *Staphylococcus aureus*, *Streptococcus pyogenes*, and *Enterococcus faecalis*, and Gram-negative organisms such as *Pseudomonas aeruginosa*, *Escherichia coli*, *Klebsiella pneumoniae*, and *Serratia species* [1]. There is a continuous need of the development of new effective antimicrobial drugs because of the emergence of new infectious diseases. Herbs and plants have been in use as a source of therapeutical compounds traditional medicinal system since ancient time. Hence, we have chosen a plant which has number of antimicrobial compound [2] such as *Calotropis gigantea* which is normally identified as weed plant is a wasteland plant belonging to Asclepiadaceae family. It is a 3–4 m tall shrub with milky latex. Bark ash colored, leaves opposite, decussate, sessile or sub sessile; flower 2–4 cm across, purplish white, complete, fruit follicles, seed numerous, broadly ovate, and plano-convex. At present, the flowering parts of the plant prescribed to heal various bacterial infections. Hydrogel base is a critical factor to produce preparations having good physical properties and assist the process of wound healing in patients. This paper provides basic information on the antimicrobial activities of *C. gigantea* against bacterial pathogens which are often associated in foot ulcer in diabetes and the formulation of hydrogel.

METHODS

Selection and authentication of plant species

The plant *Calotropis Gigantea* was chosen and the extracts from its leaves were used for the complete work, which was authenticated by the Department of Botany, Govt Brennen College, Thalassery, Kerala. The authentication number is BSI/SRC-3/27/2019.

Isolation and purification of active factors from the selected plants

The leaves of the plant were collected, shade dried, and ground to form a fine powder. The powder was then subjected to solvent extraction using ethanol, butanol, and acetone. A 10% W/V of the powder is mixed in each of the selected solvents, and left as such in a shaker incubator for 24 h. After incubation the content is filtered using a Whatman filter paper and the filtrate mixture containing the specified active factor was dried out to concentrate the active factor [3]. The hydrogel obtained from powdered dried leaves may have potential in various drug delivery systems [4]. The dried powder is used further for estimating the presence of various phytochemical compounds and in the production of hydrogel.

Phytochemical constituent’s determination

The following phytochemicals such as tannins, alkaloids, flavonoids, saponins, steroids, terpenoids, carbohydrate, and proteins were determined by the methods described by Mikail and Venkitachalapathi Kalaiselvi [5,6].

Determination of antimicrobial activity against the selected clinical specimens

Microbes were pm-cured from KMCH Clinical Laboratory, Coimbatore. Among the organism based on the predominance in causing foot ulcer the organisms mentioned in the following was used for further antibacterial analysis such as *Staphylococcus aureus*, *Pseudomonas aeroginosa*, *Escherichia coli*, *Klebsiella pneumoniae*, *Streptococcus pyogenes*, *Enterococcus faecalis*, and *Serratia sp*.

Development of hydrogel containing extracts of *Calotropis gigantea*

Required quantity of carbopol-934 was slowly sprinkled with continuous stirring into measured quantum of water to get a uniform dispersion and then kept overnight for hydration. The accurately weighed amounts of plant extract along with other additives were poured in the fixed amount of hydrated Carbopol-934 dispersion (after incubation) with constant mechanical stirring. The solution is neutralized by slowly adding triethanolamine solution with constant stirring until the gel is formed. The quantity of ingredients is gradually increased up to three formulations (F1, F2, and F3) to detect the strength of the formulation effectively. Hydrogel formulations were made with different quantity of ingredients as mentioned in Table 1.
Evaluation of properties of developed hydrogel [7]
The physical appearance was visually checked for the texture of hydrogel formulations and observations were recorded. The color of the formulations was checked against white and black background and documented. For analyzing the pH of the developed hydrogel, 10% w/v of the gel was dispersed in distilled water and stored for 2 h. This solution was used for the measurement of pH of developed hydrogel. The measurement of pH of formulation was carried out in triplicate and the average values are recorded.

Gel is spreading on the skin at a particular area called spreadability. It is calculated using the formula:

\[ S = \frac{m \cdot l}{t} \]

Where, \( m \) = weight tied to upper slide, \( l \) = length moved on glass slide, and \( t \) = time taken to separate the slides completely from each other. Spreadability of the formulation was recorded. The hydrogel formulation whose spreadability had to be determined was placed over one slide. A second slide was placed over the slide in such a way that the formulation was sandwiched between them across the length of the slide (6 cm). Exactly 100 gm of weight was placed up on the upper slide so that the gel formulation between the two slides was traced uniformly to form a thin layer. The weight was removed and the excess of the gel formulation adhering to the slides was scrapped off the bottom of the slide was attached on the apparatus board and the upper portion of the slide was tied with a string. Phenolics are essential metabolites which are playing vital role in the antioxidant activity [8-10]. 20 g load was applied by means of a simple pulley. The time taken for the upper slide to travel the distance of 6 cm and separate away from the lower slide under the direction of the weight was noted. This step was repeated for 6 times and the average was calculated for gel formulation.

Exactly 1 gm of gel was accurately weighed and transferred to 100 ml volumetric flask to which about 70 ml of methanol was added and stirred followed by making up volume to 100 ml with methanol. The content was filtered using filter paper 1 ml filtrate was pipette out and diluted with methanol. Then the extract was estimated by using spectrophotometer.

Statistical analysis
All values are reported as mean ± SEM the statistical differences between groups were determined by the Duncan multiple range test and analysis of variance also applied. A value of p<0.05 was considered significant. Statistical analysis was performed using the SAS for Windows software.

RESULTS AND DISCUSSION
The extracts of Calotropis gigantea is said to have antidiabetic activity [8] and hence if the plant extract is capable of having an antibacterial activity too [9] the issue surrounding the treatment of diabetic foot ulcer would have additional leverage.

Table 1: Hydrogel formulations with various quantity of ingredients

| Ingredient            | F1 (in g) | F2 (in g) | F3 (in g) |
|-----------------------|-----------|-----------|-----------|
| C. gigantea extract   | 1         | 1.5       | 2         |
| Carbopol 934          | 1.5       | 1.5       | 1.5       |
| Polyethylene glycol   | 5         | 5         | 5         |
| Triethanolamine       | 1.5       | 1.5       | 1.5       |
| Distilled Water (in ml)| 50       | 50       | 50        |

F1, F2, F3- Formulations

Table 2: Phytochemical profile of Calotropis gigantea leaf extract

| S. No | Constituents | Ethanolic extract | Butanolic extract | Acetone extract |
|-------|--------------|-------------------|-------------------|-----------------|
| 1.    | Alkaloid     | ++                | +                 | +               |
| 2.    | Flavonoids   | ++                | +                 | -               |
| 3.    | Terpenoids   | +                 | +                 | -               |
| 4.    | Steroids     | +                 | +                 | -               |
| 5.    | Tannins      | +                 | +                 | -               |
| 6.    | Saponins     | +                 | ++                | -               |
| 7.    | Phenols      | +                 | +                 | -               |
| 8.    | Carbohydrate | -                 | +                 | -               |
| 9.    | Proteins     | -                 | -                 | -               |

+: Presence, -: Absence, ++: Excessive

Fig. 1: Zone of inhibition by the ethanolic C. gigantea

Fig. 2: Zone of inhibition by But anolic extract of extract of C. gigantea

Fig. 3: Hydrogel F3 formulation
Table 3: Antibacterial activity of Calotropis gigantea leaf-ethanol extract

| S. No. | Organism            | Zone of Inhibition in mm |
|--------|---------------------|--------------------------|
|        | ethanol leaf extract | Chl (20 µg) | DMSO and water |
|        | 20 µg | 10 µg | 0.5 µg | 20 µg | 10 µg | 0.5 µg |
| 1.     | Staphylococcus aureus | 17±0.02 | 14±0.02 | 13±0.02 | 21±0.07 | Nil |
| 2.     | Pseudomonas aeruginosa | 16±0.03 | 15±0.03 | 12±0.01 | 17±0.04 | Nil |
| 3.     | Escherichia coli | 16±0.04 | 14±0.03 | 12±0.02 | 17±0.02 | Nil |
| 4.     | Klebsiella pneumonia | 17±0.03 | 16±0.02 | 14±0.02 | 20±0.06 | Nil |
| 5.     | Streptococcus sp. | 22±0.06 | 19±0.04 | 16±0.03 | 14±0.02 | Nil |
| 6.     | Enterococcus faecalis | 9±0.01 | NOD | NOD | 15±0.03 | Nil |
| 7.     | Serratia sp. | 10±0.02 | 9±0.02 | NOD | 18±0.02 | Nil |

Table 4: Antibacterial activity of Calotropis gigantea butanol leaf extract

| S. No. | Organism            | Zone of Inhibition in mm |
|--------|---------------------|--------------------------|
|        | butanol leaf extract | Chl (20 µg) | DMSO and water |
|        | 20 µg | 10 µg | 0.5 µg | 20 µg | 10 µg | 0.5 µg |
| 1.     | Staphylococcus aureus | 12±0.02 | 10±0.02 | 9±0.02 | 2±0.14 | Nil |
| 2.     | Pseudomonas aeruginosa | 17±0.03 | 15±0.03 | 14±0.07 | 17±0.11 | Nil |
| 3.     | Escherichia coli | 19±0.08 | 17±0.02 | 15±0.03 | 17±0.11 | Nil |
| 4.     | Klebsiella pneumonia | 19±0.02 | 18±0.04 | 16±0.03 | 18±0.16 | Nil |
| 5.     | Streptococcus sp. | 14±0.03 | 13±0.02 | 12±0.01 | 14±0.09 | Nil |
| 6.     | Enterococcus faecalis | 12±0.02 | 10±0.03 | 9±0.08 | 15±0.12 | Nil |
| 7.     | Serratia sp. | 8±0.03 | NOD | NOD | 18±0.15 | Nil |

Table 5: Antibacterial activity of Calotropis gigantea acetone leaf extract

| S. No. | Organism            | Zone of Inhibition in mm |
|--------|---------------------|--------------------------|
|        | acetone leaf extract | Chl (20 µg) | DMSO and water |
|        | 20 µg | 10 µg | 0.5 µg | 20 µg | 10 µg | 0.5 µg |
| 1.     | Staphylococcus aureus | 18±0.04 | 10±0.09 | 6±0.09 | 12±0.12 | Nil |
| 2.     | Pseudomonas aeruginosa | 9±0.06 | 8±0.06 | NOD | 17±0.15 | Nil |
| 3.     | Escherichia coli | 10±0.09 | 8±0.03 | 9±0.05 | 17±0.09 | Nil |
| 4.     | Klebsiella pneumonia | 9±0.07 | 4±0.02 | NOD | 20±0.16 | Nil |
| 5.     | Streptococcus pyogenes | 9±0.06 | 4±0.03 | NOD | 14±0.12 | Nil |
| 6.     | Enterococcus faecalis | NOD | NOD | NOD | 15±0.10 | Nil |
| 7.     | Serratia sp. | NOD | NOD | NOD | 18±0.11 | Nil |

Table 6: Physical parameters of formulation

| S. No. | Formulation | Color | Odor | Homogeneity | pH |
|--------|-------------|-------|------|-------------|----|
| 1.     | F1          | Light green | Stable | Good | 7.2±0.03 |
| 2.     | F2          | Greenish | Stable | Good | 7.0±0.05 |
| 3.     | F3          | Greenish | Stable | Good | 6.8±0.02 |

Table 7: Spreadability of the Formulation

| S. No. | Formulation | Mean time | Spreadability % | drug content |
|--------|-------------|-----------|-----------------|--------------|
| 1.     | F1          | 5.8±0.03 | 25.8±0.21 | 61.2±0.38 |
| 2.     | F2          | 5.5±0.02 | 29.09±0.17 | 68.3±0.42 |
| 3.     | F3          | 5.0±0.04 | 34.0±0.28 | 75.2±0.55 |

Isolation and purification of active phytochemicals from the calotropis gigantean

The ethanolic and butanolic leaf extracts evident the excessive quantity of alkaloid, flavonoid and saponins than the acetone extract which evidenced the presence of alkaloid alone. This supports the earlier findings of Ogbulie [11] that ethanol and butanol are the best solvent for the extraction of plant bioactive principles of medicinal importance (Table 2).

Antibacterial activity of Calotropis gigantea leaf extract against clinical isolates

The result showed that there is no antimicrobial activity of the leaf water extract and the leaf DMSO extract (Table 3). The ethanol extract of C. gigantea leaves exhibited the antibacterial activity against seven clinical isolates of bacteria (Tables 3 and Fig. 1) and the results were expressed as mean ± standard deviation (n=3). Extract showed maximum antibacterial activity against Streptococcus sp. (22±0.06) and the lowest activity against Enterococcus faecalis (9±0.01). This result supports the earlier studies of Meenakshi Sharma [12].

The zone of inhibition (Table 4 and Fig. 2) of butanol leaf extract against seven clinical isolates was ranged from 8 mm to 19 mm, 10 mm to 18 mm, 9 mm to 16 mm for 20 µg, 10 µg, 0.5 µg, respectively. In which the wide zone of inhibition (19±0.02) against Klebsiella pneumoniae than the positive control antibiotic chloramphenicol, whereas the other organisms showed a lesser zone of inhibition than the control antibiotic.
Zone of inhibition not includes the diameter of the well

The antibacterial activity of acetone exact reveals a different scenario against clinical isolates. Even though acetone is a disinfectant on its own their bactericidal property, it has been evidenced when it used as solvent. The zone of inhibition (Table 5) of acetone leaf extract against seven clinical isolates was ranged from 9 mm to 18 mm, 4 mm to 10 mm, 6 mm to mm for 20 µg, 10 µg, and 0.5 µg, respectively. In which the wide zone of inhibition (18±0.94) against Staphylococcus aureus than the positive control antibiotic chloramphenico. However, there were no zone of inhibition observed against Enterococcus faecalis and Serratia sp. However, the leaf extract of Calotrops gigantea was found to effective against the foot ulcer causing organisms such as Staphylococcus aureus, Klebsiella pneumoniae, and Streptococcus sp. [13].

Development of hydrogel and evaluation of their properties

The hydro gel was light greenish in color with stable odor and at pH 7.2. It also showed good consistency and homogeneity (Fig. 3). Spreadability and drug content of gel was measured and tabulated (Tables 6 and 7).

Development of hydrogel and evaluation of their properties

In the present study, the ethanolic extract of C. gigantea was revealed more effective than other extract preparation. This might be due to the polar nature of the solvent, that is, ethanol, which resulted in leaching of more active ingredients during extraction for antimicrobial active substance from Calotrops compared to other solvents. Since only one more study is there on the preparation of hydro gel using Calotrops gigantea leaf extract. From the result it is clear that all the gel formulation shows good gelling properties such as homogeneity and consistency. The pH (Table 7) of all formulations was in the range of compatible (6.8±0.02) with normal pH range of the skin. The drug content released was also above average. Thus the gel formulation has all the desirable properties of an ideal gel formulation. The result shows that the hydrogel developed from Calotrops gigantea has probable solutions that could pave way for the new drug development against the foot ulcer occurring in the diabetes. Our results support overall in vitro antibacterial activity of the extracts that may prove to be of clinical importance in improving the management of bacterial recurrent infection in diabetic patients.

CONCLUSION

The medicinal property of Calotrops gigantea was best studied. It evident the antibacterial activity against bacteria which has been isolated from the foot ulcer of diabetic. Hydrogel formulation was tried and prepared from the leaf extract of Calotrops gigantea. Foot ulcer causing organisms such as Staphylococcus aureus, Streptococcus sp, and Klebsiella pneumoniae growth were inhibited significantly.

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AUTHORS CONTRIBUTION

All the authors contributed to the preparation of the final manuscript.

CONFLICT OF INTEREST

There is no conflict of interest

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