Comprehensive analysis of a long noncoding RNA-associated competing endogenous RNA network in colorectal cancer

Qiaowei Fan1,2
Bingrong Liu1

1Department of Gastroenterology and Hepatology, The Second Affiliated Hospital of Harbin Medical University, Harbin, Heilongjiang, People’s Republic of China; 2Translational Medicine Research and Cooperation Center of Northern China, Heilongjiang Academy of Medical Sciences, Harbin, Heilongjiang, People’s Republic of China

Purpose: This study was aimed to develop a lncRNA-associated competing endogenous RNA (ceRNA) network to provide further understanding of the ceRNA regulatory mechanism and pathogenesis in colorectal cancer (CRC).

Patients and methods: Expression profiles of mRNAs, lncRNAs, and miRNAs, and clinical information for CRC patients were obtained from The Cancer Genome Atlas. The differentially expressed mRNAs, lncRNAs, and miRNAs (referred to as “DEmRNAs”, “DElncRNAs”, and “DEmiRNAs”, respectively) were screened out between 539 CRC samples and 11 normal samples. The interactions between DElncRNAs and DEmiRNAs were predicted by miRcode. The DEmiRNAs targeted by the DEmiRNAs were retrieved according to TargetScan, miRTarBase, and miRDB. The lncRNA–miRNA–mRNA ceRNA network was constructed based on the DEmiRNA–DElncRNA and DEmiRNA–DEmRNA interactions. Functional enrichment analysis revealed the biological processes and pathways of DEmiRNAs involved in the development of CRC. Key lncRNAs were further analyzed for their associations with overall survival and clinical features of CRC patients.

Results: A total of 1,767 DEmRNAs, 608 DElncRNAs, and 283 DEmiRNAs were identified as CRC-specific RNAs. Three hundred eighty-two DEmiRNA–DElncRNA interactions and 68 DEmiRNA–DEmRNA interactions were recognized according to the relevant databases. The lncRNA–miRNA–mRNA ceRNA network was constructed using 25 DEmiRNAs, 52 DEmRNAs, and 64 DElncRNAs. Two DElncRNAs, five DEmiRNAs, and six DEmRNAs were demonstrated to be related to the prognosis of CRC patients. Four DElncRNAs were found to be associated with clinical features. Twenty-eight Gene Ontology terms and 10 Kyoto Encyclopedia of Genes and Genomes pathways were found to be significantly enriched by the DEmiRNAs in the ceRNA network.

Conclusion: Our results showed cancer-specific mRNA, lncRNA, and miRNA expression patterns and enabled us to construct an lncRNA-associated ceRNA network that provided new insights into the molecular mechanisms of CRC. Key RNA transcripts related to the overall survival and clinical features were also found with promising potential as biomarkers for diagnosis, survival prediction, and classification of CRC.

Keywords: colorectal cancer, competing endogenous RNA network, long noncoding RNA, survival analysis

Introduction
Noncoding RNAs (ncRNAs) are RNAs that lack protein-coding functions.1 On the basis of their characteristics, ncRNAs can be divided into several classes, including lncRNAs, miRNAs, small interfering RNAs, small nuclear RNAs, and ribosomal RNAs.1 miRNAs are a class of small ncRNAs with important regulatory roles, ~22 nt
in length. They can posttranscriptionally regulate gene expression through binding to miRNA response elements (MREs) on their target transcripts. A single miRNA can regulate up to hundreds of target genes, and each gene may also be modulated by multiple miRNAs. Aberrantly expressed miRNAs are associated with many diseases including cancer.

In 2011, Salmena et al presented a competing endogenous RNA (ceRNA) hypothesis, which proposed a new “language” among different types of RNA transcripts. The hypothesis postulates that RNA molecules can communicate with each other through shared MREs. MRNAs, lncRNAs, and other RNA transcripts could act as endogenous miRNA sponges to inhibit miRNA function, thereby impact the multiple targets of multiple miRNAs. RNA transcripts sharing one or more MREs can actively regulate their respective expression levels by competing for a limited pool of miRNAs. This ceRNA crosstalk forms a large-scale regulatory network across the transcriptome, including protein-coding and noncoding RNAs. LncRNAs, a class of ncRNAs 200 nt in length, are suggested to participate in various biological processes (BPs) by regulating gene expression at transcriptional, posttranscriptional, and epigenetic levels. Recent studies have provided emerging support for the ceRNA activity of lncRNAs involved in the progression of various types of cancer. LncRNA-associated ceRNA networks have been constructed and analyzed in lung squamous cell carcinoma, papillary renal cell carcinoma, hepatocellular carcinoma, and gastric cancer. However, there have been few integrated analyses of lncRNA-associated ceRNA networks in colorectal cancer (CRC).

In the present study, we conducted a comprehensive analysis of the mRNA, lncRNA, and miRNA expression profiles in CRC and constructed a CRC-specific lncRNA-associated ceRNA network using a large cohort from The Cancer Genome Atlas (TCGA). To the best of our knowledge, our study is the first to establish an lncRNA-associated ceRNA network in CRC using a combined dataset of colon adenocarcinoma and rectum adenocarcinoma from TCGA. The results are expected to help to further elucidate the lncRNA–miRNA–mRNA crosstalk in CRC and generate insight into the molecular mechanisms involved in the tumorigenesis and progression of CRC.

**Patients and methods**

**Selection of CRC patients**

The mRNA, lncRNA, and miRNA expression quantification files and the corresponding clinical information for CRC patients were obtained from TCGA (https://cancergenome.nih.gov). The inclusion criteria were as follows: 1) patients with complete clinical information including age, gender, pathological stage, TNM stage, survival status, and survival time; 2) patients with complete RNA-Seq data of mRNAs and lncRNAs, and complete miRNA-Seq data; 3) patients with a follow-up survival time of ≤2,000 days. After filtering with the inclusion criteria, a total of 539 CRC patients and 11 normal controls were finally enrolled in the study. The clinicopathological characteristics of the CRC patients are summarized in Table 1.

**Expression profile analysis of mRNAs, lncRNAs, and miRNAs in CRC and normal tissues**

The differentially expressed mRNAs, lncRNAs, and miRNAs (hereafter referred to as “DEmRNAs”, “DElncRNAs”, and “DEmiRNAs”, respectively) in CRC and normal tissues were screened out individually using the edgeR package in the R language with the threshold of

| Clinicopathological characteristics | Patients (N=539) |
|-----------------------------------|-----------------|
| Age category (years) | n | % |
| <65 | 220 | 40.82 |
| ≥65 | 319 | 59.18 |
| Gender | | |
| Female | 252 | 46.75 |
| Male | 287 | 53.25 |
| Pathological stage | | |
| Stage I | 99 | 18.37 |
| Stage II | 192 | 35.62 |
| Stage III | 163 | 30.24 |
| Stage IV | 85 | 15.77 |
| Pathologic T | | |
| Tis | 1 | 0.18 |
| T1 | 18 | 3.34 |
| T2 | 95 | 17.63 |
| T3 | 360 | 66.79 |
| T4 | 65 | 12.06 |
| Pathologic N | | |
| N0 | 301 | 55.85 |
| N1 | 138 | 25.60 |
| N2 | 100 | 18.55 |
| Pathologic M | | |
| M0 | 404 | 74.95 |
| M1 | 83 | 15.40 |
| Mx | 52 | 9.65 |
| Survival status | | |
| Alive | 440 | 81.63 |
| Dead | 99 | 18.37 |

**Table 1** Clinicopathological characteristics of 539 CRC patients

**Abbreviation:** CRC, colorectal cancer.
\[ \text{log}_2 \text{fold change (log}_2 \text{FC}) > 2 \] and false discovery rate (FDR) < 0.01. Hierarchical clustering analysis was then performed using the pheatmap package in R (Version 1.0.8) to evaluate the specificity of the differentially expressed RNAs.

Construction of the ceRNA network

The interactions between DElncRNAs and DEmiRNAs were predicted by miRcode. The mRNAs targeted by the DEmiRNAs were retrieved according to TargetScan, miRTarBase, and miRDB. Only mRNAs recognized by all three databases were considered as candidate mRNAs and intersected with the DEmiRNAs to screen out the DEmiRNAs targeted by the DEmiRNAs. Then, the lncRNA–miRNA–mRNA ceRNA network was constructed based on the DEmiRNA–DElncRNA and DEmiRNA–DEmRNA interactions and visualized using the Cytoscape software. A flowchart of the lncRNA–miRNA–mRNA ceRNA network analysis is displayed in Figure 1. Linear regression analysis was performed to evaluate the correlation between ceRNA expression levels.

Survival analysis

For each of the DEmiRNAs, DElncRNAs, and DEmiRNAs in the ceRNA network, the CRC patients were classified into either a high expression group or a low expression group using the median expression value of the specific RNA as the cutoff value. The differences in patients’ survival time between the high expression group and the low expression group were then evaluated by Kaplan–Meier survival curve and log-rank test analysis to find the mRNAs, IncRNAs, and miRNAs that were significantly associated with the survival of CRC patients. \( P < 0.05 \) was set as the threshold for statistical significance.

The associations between key IncRNAs and clinical features

Based on the bioinformatics analysis and the ceRNA network, key IncRNAs were screened out. The IncRNAs were

![Flowchart of the lncRNA–miRNA–mRNA ceRNA network analysis.](image)

Figure 1 Flowchart of the lncRNA–miRNA–mRNA ceRNA network analysis.
then analyzed to determine whether they were differentially expressed between different clinical feature categories, including gender (male vs female), T stage (T3 + T4 vs Tis + T1 + T2), N stage (N1 + N2 vs N0), M stage (M1 vs M0), and pathological stage (stage III + IV vs stage I + II). Differentially expressed key lncRNAs with $|\log_{2} FC|>1$ and FDR $<0.05$ were considered significantly associated with the relevant clinical feature.

Functional enrichment analysis

Functional enrichment analysis was performed for the DEmRNAs in the ceRNA network to reveal the functional implications of these mRNAs in the tumorigenesis of CRC. Gene Ontology (GO) functional enrichment analysis was conducted using the Database for Annotation, Visualization and Integrated Discovery (DAVID, Version 6.8) online tool. Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analysis was carried out using the KEGG Orthology-Based Annotation System (KOBAS). $P<0.05$ was set as the statistical significance threshold criterion for both GO and KEGG enrichment analysis.

Results

Cancer-specific mRNAs, lncRNAs, and miRNAs in CRC

According to the cutoff threshold, a total of 1,767 DEmRNAs (including 854 upregulated and 913 downregulated) were screened out between CRC tissues and normal tissues. The expressions of 608 DElncRNAs (including 370 upregulated and 238 downregulated) and 283 DEmiRNAs (including 201 upregulated and 82 downregulated) were altered significantly in CRC tissues compared with normal tissues. Unsupervised hierarchical clustering analysis was performed for the top 100 upregulated and top 100 downregulated mRNAs and lncRNAs (owing to the large amount of DEmRNAs and DElncRNAs), respectively, and all the DEmiRNAs. The heatmaps showed that the CRC samples could be clearly distinguished from the normal samples with respect to the expression of these differentially expressed RNAs (Figure 2A–C).

DElncRNA–DEmiRNA interactions predicted by miRcode

The lncRNAs that are targeted by miRNAs were predicted by miRcode. Only interactions between DElncRNAs and DEmiRNAs were selected for ceRNA network construction. As a result, 479 interactions between 67 DElncRNAs (including KCNQ1OT1, ADAMTS9-AS2, and AGAP11) and 32 DEmiRNAs (including hsa-mir-106a, hsa-mir-141, and hsa-mir-143) were identified for further analysis.

Prediction of DEmiRNAs targeted by DEmiRNAs

The 32 DEmiRNAs recognized in the above step were mapped into TargetScan, miRTarBase, and miRDB.
 databases to search for their target mRNAs. A total of 1,416 mRNAs were found to be targeted by these DEmiRNAs in all three databases. These candidate mRNAs were then intersected with the DEmiRNAs, and 52 of the 1,416 mRNAs were identified as the DEmiRNAs targeted by the DEmiRNAs. Among the 32 candidate DEmiRNAs, only 25 were found to interact with these 52 DEmiRNAs, the remaining seven DEmiRNAs without specific DEmRNA targets were excluded from further analysis. To highlight the ceRNA features of the network, only DEmiRNAs interacting with both DEmiRNAs and DElncRNAs were introduced into the ceRNA network. Finally, 25 DEmiRNAs, 52 DEmiRNAs, and 64 DElncRNAs were selected for ceRNA network construction.

**Figure 2** Heatmaps based on the differentially expressed RNAs in CRC and normal tissues.

**Notes:** (A) The heatmap with top 100 upregulated and top 100 downregulated mRNAs. (B) The heatmap with top 100 upregulated and top 100 downregulated lncRNAs. (C) The heatmap with all the DEmiRNAs.

**Abbreviation:** CRC, colorectal cancer.
The ceRNA network in CRC

Based on the above data, an lncRNA–miRNA–mRNA ceRNA network consisting of 141 molecules and 450 interactions (382 DEmiRNA–DElncRNA interactions and 68 DEmiRNA–DEmRNA interactions) was constructed. The top 11 DElncRNAs targeted by most DEmiRNAs are shown in Table 2. All the 64 DElncRNAs in the ceRNA network with their name, Ensembl ID, log2 FC, and FDR are listed in Table S1. All the 25 DEmiRNAs with their target DEmRNAs in the ceRNA network are shown in Table 3. The network was visualized with Cytoscape and is displayed in Figure 3. According to the ceRNA network, there is a possibility of indirect interactions between DElncRNAs and DEmRNAs. To confirm our finding, linear regression analysis was performed. The results suggested a good positive correlation between ceRNA expression levels (Figures 4A–I and 5A–H). For example, mediated by hsa-mir-106a, LINC00461 interacted with CADM2, CFL2, and FAM129A while mediated by hsa-mir-424, LINC00461 interacted with TMEM100 and TPM2 (Figure 4A, E, D, F, and H). Mediated by hsa-mir-152, LINC00484 interacted with BMP3 and KLF4 while mediated by hsa-mir-141, LINC00484 interacted with PHLPP2 (Figure 5 A, D, and C).

Table 2 The top 11 DElncRNAs putatively targeted by most DEmiRNAs in the ceRNA network

| DElncRNAs | DEmiRNAs |
|-----------|-----------|
| KCNQ1OT1  | hsa-mir-106a, hsa-mir-141, hsa-mir-143, hsa-mir-145, hsa-mir-150, hsa-mir-152, hsa-mir-17, hsa-mir-182, hsa-mir-192, hsa-mir-193b, hsa-mir-206, hsa-mir-217, hsa-mir-301b, hsa-mir-32, hsa-mir-338, hsa-mir-372, hsa-mir-375, hsa-mir-424, hsa-mir-429, hsa-mir-454, hsa-mir-96, hsa-mir-98 |
| ADAMTS9-AS2 | hsa-mir-106a, hsa-mir-141, hsa-mir-143, hsa-mir-145, hsa-mir-150, hsa-mir-152, hsa-mir-182, hsa-mir-192, hsa-mir-193b, hsa-mir-223, hsa-mir-301b, hsa-mir-32, hsa-mir-338, hsa-mir-372, hsa-mir-375, hsa-mir-454, hsa-mir-96, hsa-mir-98 |
| DLX6-AS1 | hsa-mir-106a, hsa-mir-141, hsa-mir-143, hsa-mir-144, hsa-mir-145, hsa-mir-150, hsa-mir-152, hsa-mir-17, hsa-mir-192, hsa-mir-193b, hsa-mir-206, hsa-mir-223, hsa-mir-338, hsa-mir-372, hsa-mir-424, hsa-mir-429 |
| AGAPI1 | hsa-mir-106a, hsa-mir-141, hsa-mir-143, hsa-mir-145, hsa-mir-150, hsa-mir-152, hsa-mir-182, hsa-mir-192, hsa-mir-193b, hsa-mir-206, hsa-mir-21, hsa-mir-217, hsa-mir-372, hsa-mir-375, hsa-mir-424, hsa-mir-96 |
| AC010336.2 | hsa-mir-106a, hsa-mir-141, hsa-mir-143, hsa-mir-144, hsa-mir-145, hsa-mir-150, hsa-mir-192, hsa-mir-32, hsa-mir-338, hsa-mir-372, hsa-mir-424, hsa-mir-96 |
| LINC00461 | hsa-mir-106a, hsa-mir-141, hsa-mir-143, hsa-mir-144, hsa-mir-145, hsa-mir-150, hsa-mir-192, hsa-mir-32, hsa-mir-338, hsa-mir-372, hsa-mir-424, hsa-mir-96 |
| LINC00484 | hsa-mir-106a, hsa-mir-141, hsa-mir-143, hsa-mir-145, hsa-mir-152, hsa-mir-206, hsa-mir-217, hsa-mir-223, hsa-mir-32, hsa-mir-338, hsa-mir-372, hsa-mir-424, hsa-mir-96 |
| LIFR-AS1 | hsa-mir-106a, hsa-mir-141, hsa-mir-143, hsa-mir-150, hsa-mir-182, hsa-mir-192, hsa-mir-193b, hsa-mir-206, hsa-mir-32, hsa-mir-372, hsa-mir-375, hsa-mir-96 |
| LINC00330 | hsa-mir-106a, hsa-mir-141, hsa-mir-143, hsa-mir-145, hsa-mir-150, hsa-mir-152, hsa-mir-182, hsa-mir-206, hsa-mir-217, hsa-mir-372, hsa-mir-424, hsa-mir-454, hsa-mir-96 |
| FAM95B1 | hsa-mir-106a, hsa-mir-143, hsa-mir-145, hsa-mir-150, hsa-mir-152, hsa-mir-206, hsa-mir-301b, hsa-mir-372, hsa-mir-424, hsa-mir-98 |
| PVT1 | hsa-mir-106a, hsa-mir-145, hsa-mir-150, hsa-mir-17, hsa-mir-21, hsa-mir-217, hsa-mir-372, hsa-mir-424 |

Table 3 The 25 DEmiRNAs with their target DEmRNAs in the ceRNA network

| DEmiRNAs | DEmRNAs |
|----------|---------|
| hsa-mir-106a | CFL2, FOXQ1, FAM129A, CADM2, FJX1 |
| hsa-mir-141 | ELAVL4, PHLPP2, EPAH7, KIAA1549 |
| hsa-mir-143 | COL1A1, SERPINE1 |
| hsa-mir-144 | KCNQ5, GRIK3 |
| hsa-mir-145 | SERPINE1 |
| hsa-mir-150 | EREG, HILPDA, SLC7A11 |
| hsa-mir-152 | BMP3, NPTX1, KLF4 |
| hsa-mir-17 | FAM46C, FOXQ1, FJX1, CADM2, CYBRD1, FAM129A, CFL2 |
| hsa-mir-182 | CHL1, NPTX1, ULBP2 |
| hsa-mir-192 | GRHL1 |
| hsa-mir-193b | PLAU |
| hsa-mir-206 | STC2, SFRP1 |
| hsa-mir-21 | TGFBI |
| hsa-mir-217 | DACH1 |
| hsa-mir-223 | EPB4I1L3 |
| hsa-mir-301b | RBM20, CFL2 |
| hsa-mir-32 | PHLPP2, PBLD, PAX9, UGP2 |
| hsa-mir-338 | NOVAI |
| hsa-mir-372 | CADM2, TMEM100, SLC7A11, MIXL1 |
| hsa-mir-375 | ELAVL4 |
| hsa-mir-424 | PSAT1, TMEM100, PHLPP2, CBX2, AXIN2, TPM2 |
| hsa-mir-429 | PMAIP1 |
| hsa-mir-454 | CFL2, RBM20 |
| hsa-mir-96 | ALK, TRIB3, SLC1A1 |
| hsa-mir-98 | HAND1, IGF2BP1, PRSSI22, RGS16, TRIM71, IGF2BP3, CPA4 |
Figure 3 The lncRNA–mirRNA–mRNA ceRNA network in CRC.
Notes: Red rounds represent upregulated DEmRNAs and blue rounds represent downregulated DEmRNAs. Red squares represent upregulated DElncRNAs and blue squares represent downregulated DElncRNAs. Green triangles represent upregulated DEmirnas and yellow triangles represent downregulated DEmirnas.
Abbreviation: CRC, colorectal cancer.

Important RNAs associated with the prognosis of CRC patients
Kaplan–Meier survival curve and log-rank test analysis were performed for each of the DElncRNAs, DEmiRNAs, and DEmRNAs in the ceRNA network to find the RNAs significantly associated with the survival of CRC patients. According to the results, two DElncRNAs (AP004609.1 and C2orf48), five DEmiRNAs (hsa-mir-96, hsa-mir-144, hsa-mir-145, hsa-mir-193b, and hsa-mir-375), and six DEmRNAs (FAM46C, FJX1, HAND1, SERPINE1, TPM2, and ULBP2) were identified as prognosis-related RNAs \( (p<0.05, \text{Figures 6A and B, 7A–E, and 8A–F}) \). Among these, hsa-mir-144, and hsa-mir-375 (DEmiRNA) and FAM46C (DEmRNA) appeared to be protective, as patients with higher expression levels of these RNAs tended to have longer overall survival, compared with those with lower expression levels of these RNAs (Figures 7B and E, and 8A). The remaining two DElncRNAs, three DEmiRNAs, and five DEmRNAs were considered risky, as their high expression was associated with shorter overall survival of CRC patients.
Figure 4: Linear regression of circRNA expression levels for LINC00461.

Notes: LINC00461 vs (A) CADM2, (B) GRIK3, (C) ELAVL4, (D) FAM129A, (E) CFL2, (F) TMEM100, (G) NOVA1, (H) TPM2, and (I) MIXL1. The red line represents the linear model fitted by the dots in each figure, while green lines represent the 95% CI. "Cor" represents the correlation between the expressions of LINC00461 and the specific mRNA.

Figure 5: Linear regression of circRNA expression levels for LINC00484.

Notes: LINC00484 vs (A) BMP3, (B) UGP2, (C) PHLPP2, (D) KLF4, (E) PBLD, (F) EPB41L3, (G) TMEM100, and (H) SFRP1. The red line represents the linear model fitted by the dots in each figure while green lines represent the 95% CI. "Cor" represents the correlation between the expressions of LINC00484 and the specific mRNA.
Key lncRNAs relevant to clinical features
The 64 DElncRNAs from the ceRNA network were further analyzed to find their associations with clinical features. The lncRNA expression profiles of CRC patients were compared between different genders, T, N, and M (tumor, node, and metastasis) stages, and pathological stages. Four key DElncRNAs were found to be significantly differentially expressed in clinical feature comparisons. AC110491.1 was significantly
altered between genders (male vs female). H19 and HULC were not only differentially expressed in advanced-stage patients (stage III + IV) compared with earlier-stage patients (stage I + II), but also appeared to be associated with N stage (N1 + N2 vs N0) and M stage (M1 vs M0). As well as H19 and HULC, CLDN10-AS1 was also indicated to be an M stage-associated DElncRNA. The key DElncRNAs relevant to clinical features are summarized in Table 4.

**Functional enrichment analysis**

Functional enrichment analysis revealed that a total of 28 GO terms, including 20 BP terms, 4 cellular component terms, and 4 molecular function terms were enriched by the DEmRNAs in the ceRNA network. Some of the GO terms were concerned with regulation of transcription, such as GO:0045892~negative regulation of transcription, DNA-templated and GO:0001190~transcriptional activator activity, RNA polymerase II transcription factor binding (Table 5).

The KEGG pathway enrichment analysis was also performed for the DEmRNAs. Ten KEGG pathways were found to be significantly enriched by the DEmRNAs in the ceRNA network, among which hsa04115: p53 signaling pathway, hsa05206: MicroRNAs in cancer, hsa04310: Wnt signaling pathway, and hsa04390: Hippo signaling pathway were closely associated with the pathogenesis of cancer (Table 6).

**Discussion**

CRC is one of the most common malignancies worldwide, ranking the third in men and the second in women. Despite numerous studies endeavoring to reveal the mechanisms of CRC, the exact pathogenesis has remained to be elucidated. In recent years, a novel hypothesis has been proposed that different RNA transcripts could interact with each

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**Table 4** The correlations between DElncRNAs from the ceRNA network and clinical features of CRC patients

| Comparisons                        | Upregulated | Downregulated |
|------------------------------------|-------------|---------------|
| Gender (male vs female)            |             | AC110491.1    |
| N stage (N1 + N2 vs N0)            | H19         | HULC          |
| M stage (M1 vs M0)                 | CLDN10-AS1, H19 | HULC          |
| Pathological stage (III + IV vs I + II) | H19         | HULC          |

**Figure 8** Kaplan–Meier survival curves for the six DEmRNAs significantly associated with overall survival of CRC patients.

**Note:** Kaplan–Meier survival curves for (A) FAM46C, (B) FJX1, (C) HAND1, (D) SERPINE1, (E) TPM2, and (F) ULBP2.

Abbreviation: CRC, colorectal cancer.
other through shared MREs. This is called “the ceRNA hypothesis” and presents a new pattern of gene expression regulation that could be used to further understand the mechanisms of various diseases including cancer.

In the present study, we identified DElncRNAs, DEmiRNAs, and DEmRNAs between CRC tissues and normal tissues, using a large cohort from the TCGA database. Then, an lncRNA–miRNA–mRNA ceRNA network was constructed, which provided an integrated view of the ceRNA regulatory crosstalk among these CRC-specific RNA transcripts. The DElncRNAs in the ceRNA network were analyzed for their associations with the survival and clinical features of CRC patients. Functional enrichment analysis further revealed the BPs and pathways associated with the DEmRNAs involved in the development of CRC.

Dysregulated lncRNAs have been observed in various cancers, and some of them have been demonstrated to be associated with patient survival. For instance, decreased expression of the lncRNA MLLT4 antisense RNA 1 indicates poor prognosis in gastric cancer patients, whereas overexpression of the lncRNA linc-UBC1 is associated with poor prognosis in CRC. In the present study, two DElncRNAs from the ceRNA network, namely AP004609.1 and C2orf48, were found to be significantly associated with the survival of patients with CRC. Both of them could be considered life threatening, as patients with high expression levels of these two lncRNAs tended to have shorter survival times than patients with low expression. Although little is known about the functions of these two DElncRNAs, our results indicate the prognostic potential of the lncRNAs in survival prediction and provide new directions for further investigation of the two lncRNAs involved in CRC.

With respect to the associations between DElncRNAs and clinical features, we found that four DElncRNAs were related to clinical features, including gender, N stage, M stage, and tumor pathological stage. Among the four DElncRNAs, H19, a widely studied lncRNA, has been suggested as an indicator of tumor progression in multiple types of cancers, including CRC. H19 has been reported to indicate poor prognosis and promote tumor growth in CRC. Overexpression of H19 is associated with decreased overall survival rates in CRC.

### Table 5 The top 10 predominant BP terms and all the four CC terms and MF terms in GO functional enrichment analysis

| Categories | Terms | Count | P-value |
|------------|-------|-------|---------|
| GO BP      | GO:0045892–negative regulation of transcription, DNA-templated | 7 | 3.16 E–03 |
|            | GO:0048147–negative regulation of fibroblast proliferation | 3 | 3.68 E–03 |
|            | GO:0001525–angiogenesis | 5 | 4.02 E–03 |
|            | GO:0043434–response to peptide hormone | 3 | 7.30 E–03 |
|            | GO:0071363–cellular response to growth factor stimulus | 3 | 7.62 E–03 |
|            | GO:0050680–negative regulation of epithelial cell proliferation | 3 | 0.012 |
|            | GO:0017148–negative regulation of translation | 3 | 0.012 |
|            | GO:0001957–intramembranous ossification | 2 | 0.017 |
|            | GO:0042035–regulation of cytokine biosynthetic process | 2 | 0.020 |
|            | GO:0060346–bone trabecula formation | 2 | 0.023 |
| GO CC      | GO:0005615–extracellular space | 13 | 1.79 E–04 |
|            | GO:0070062–extracellular exosome | 14 | 0.031 |
|            | GO:0005886–plasma membrane | 18 | 0.036 |
|            | GO:0031012–extracellular matrix | 4 | 0.045 |
| GO MF      | GO:0003730–mRNA 3′-UTR binding | 3 | 8.90 E–03 |
|            | GO:0045182–translation regulator activity | 2 | 0.017 |
|            | GO:0048027–mRNA 5′-UTR binding | 2 | 0.026 |
|            | GO:001190–transcriptional activator activity, RNA polymerase II transcription factor binding | 2 | 0.045 |

**Abbreviations:** BP, biological process; CC, cellular component; GO, Gene Ontology; MF, molecular function; UTr, untranslated region.

### Table 6 KEGG pathways enriched by the DEmRNAs in the ceRNA network

| ID        | Terms                               | P-value |
|-----------|-------------------------------------|---------|
| hsa04550  | Signaling pathways regulating pluripotency of stem cells | 3.51 E–03 |
| hsa04115  | p53 signaling pathway               | 9.51 E–03 |
| hsa04610  | Complement and coagulation cascades  | 0.012   |
| hsa00750  | Vitamin B6 metabolism               | 0.014   |
| hsa04974  | Protein digestion and absorption    | 0.016   |
| hsa05206  | MicroRNAs in cancer                 | 0.019   |
| hsa04933  | AGE-RAGE signaling pathway in diabetic complications | 0.020 |
| hsa04724  | Glutamatergic synapse               | 0.024   |
| hsa04310  | Wnt signaling pathway               | 0.036   |
| hsa04390  | Hippo signaling pathway             | 0.041   |

**Abbreviation:** KEGG, Kyoto Encyclopedia of Genes and Genomes.
patients, as well as increased viability and migration in colon cancer cells. In our study, the expression of H19 was significantly upregulated in advanced tumor stages (stages N1/N2, M1, and stage III/IV) compared with earlier tumor stages (stages N0, M0, and stage I/II). The findings of our study were consistent with previous reports and further confirmed the important function of H19 in the progression of CRC.

MiRNAs, the hub elements of the ceRNA network, exhibited key roles in the crosstalk among different RNA transcripts. Mediated by hsa-mir-424, LINC00461 interacted with TMEM100 and TPM2 while LINC00484 interacted with TMEM100 and PHLP2 in the present CRC-specific ceRNA network. Mediated by hsa-mir-141, LINC00461 interacted with ELAVL4 while LINC00484 interacted with PHLP2. Both hsa-mir-424 and hsa-mir-141 were demonstrated to be involved in the pathogenesis of CRC.41,42 We also found five miRNAs to be significantly associated with the prognosis in CRC. High expression of hsa-mir-96, hsa-mir-145, and hsa-mir-193b indicated an unfavorable clinical outcome, while upregulation of hsa-mir-144 and hsa-mir-375 suggested longer survival times of patients with CRC. All the key DEmiRNAs show promising potential as biomarkers for survival prediction in CRC.

mRNAs comprise another important part of the ceRNA network, through being directly targeted by miRNAs or having indirect interactions with lncRNAs mediated by miRNAs. Similar to lncRNAs and miRNAs, some mRNAs were also found to be associated with survival of CRC patients, such as the protective mRNA FAM46C, and the risky mRNAs including FJX1, HAND1, SERPINE1, TPM2, and ULBP2. Furthermore, functional enrichment analysis revealed that DEMRNAs were enriched in some BPs and pathways related to transcription regulation and cancer, such as negative regulation of transcription, the p53 signaling pathway, and the Wnt signaling pathway.43,44 The close correlation between the enriched pathways and the ceRNA network demonstrates the credibility of our results.

Conclusion

The present study identified CRC-specific lncRNAs, miRNAs, and mRNAs using a large cohort from the TCGA database. We successfully constructed an lncRNA-associated ceRNA network, which provided insight into the newly proposed crosstalk among different types of RNA transcripts. Key lncRNAs were analyzed to determine significant associations with overall survival and clinical features of CRC patients. Hub miRNAs also showed promising potential as prognostic biomarkers in CRC. Our study provides better understanding of the ceRNA regulatory mechanism and helps to elucidate the pathogenesis involved in the tumorigenesis and progression of CRC.

Disclosure

The authors report no conflicts of interest in this work.

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## Supplementary material

### Table S1 All the 64 DEIncRNAs in the ceRNA network

| DEIncRNAs | Ensembl ID | log_FC | FDR  | DEIncRNAs | Ensembl ID | log_FC | FDR  |
|-----------|------------|--------|------|-----------|------------|--------|------|
| ABCA9-AS1 | ENSG00000231749 | 3.9864 | 1.50E–03 | ERVMER61-1 | ENSG00000230426 | 6.2730 | 4.00E–03 |
| AC007384.1 | ENSG00000237513 | –2.4025 | 6.92E–14 | FAM95B1 | ENSG00000223839 | –2.0743 | 4.64E–06 |
| AC009065.1 | ENSG00000259780 | 2.4120 | 7.23E–03 | FRMD6-AS2 | ENSG00000258537 | –3.1025 | 7.09E–09 |
| AC009093.1 | ENSG00000259807 | 3.0580 | 8.28E–05 | GAS6-AS1 | ENSG00000233695 | 3.5499 | 1.09E–07 |
| AC009336.1 | ENSG00000272729 | –2.0597 | 6.03E–06 | H19 | ENSG00000130600 | 4.1017 | 5.86E–04 |
| AC010336.2 | ENSG00000260500 | –2.7897 | 2.57E–14 | HCG23 | ENSG00000228962 | –2.5508 | 3.36E–13 |
| AC012640.1 | ENSG000000248968 | 2.4459 | 2.90E–04 | HULC | ENSG000000251164 | 6.8799 | 6.67E–03 |
| AC016745.1 | ENSG00000234997 | 3.6141 | 4.61E–03 | JAZF1-AS1 | ENSG0000024336 | –2.6263 | 2.05E–13 |
| AC020907.1 | ENSG00000179066 | 3.8949 | 3.35E–03 | KCNQ1-ASI | ENSG00000229414 | 2.53016 | 6.52E–03 |
| AC064836.2 | ENSG00000272966 | 2.5934 | 4.09E–03 | KCNQ1OT1 | ENSG00000269821 | 2.0209 | 1.35E–05 |
| AC084262.1 | ENSG000000260253 | 3.2217 | 4.57E–03 | LIFR-AS1 | ENSG00000244968 | –2.3659 | 2.13E–13 |
| AC110491.1 | ENSG000000261292 | –4.8260 | 3.22E–22 | LINC00092 | ENSG00000225194 | –2.8438 | 8.99E–30 |
| ADAMTS9-AS1 | ENSG00000241158 | –3.9253 | 2.66E–28 | LINC00330 | ENSG00000235097 | –2.9422 | 1.32E–07 |
| ADAMTS9-AS2 | ENSG000000241684 | –2.8713 | 5.15E–21 | LINC00402 | ENSG00000235532 | –2.7266 | 1.25E–09 |
| AGAP11 | ENSG00000271880 | –2.1666 | 1.83E–07 | LINC00460 | ENSG00000233532 | 6.2479 | 1.30E–08 |
| AL138995.1 | ENSG00000027763 | –2.4069 | 1.42E–17 | LINC00461 | ENSG00000245526 | –2.9817 | 1.73E–13 |
| AL161431.1 | ENSG000000275216 | 5.7502 | 7.72E–07 | LINC00473 | ENSG00000223414 | –3.3201 | 6.64E–17 |
| AL360004.1 | ENSG00000196979 | –2.3594 | 1.39E–06 | LINC00484 | ENSG00000229694 | –3.1173 | 2.08E–16 |
| AL512652.1 | ENSG000000275485 | 2.02862 | 2.74E–04 | LINC00488 | ENSG00000241381 | –3.1742 | 1.39E–10 |
| AL513123.1 | ENSG000000236347 | 3.0582 | 5.80E–05 | LINC00507 | ENSG00000256193 | –4.11785 | 5.34E–24 |
| AL590483.1 | ENSG000000229960 | 2.8803 | 7.85E–06 | LMOT-AS1 | ENSG00000261105 | 2.4135 | 6.21E–05 |
| AP000553.1 | ENSG000000272954 | 3.3636 | 2.14E–05 | MIR22HG | ENSG00000186594 | –2.0251 | 9.64E–22 |
| AP002478.1 | ENSG000000266401 | 4.3909 | 1.13E–04 | MYO16-AS1 | ENSG00000236242 | 3.8592 | 6.26E–03 |
| AP004609.1 | ENSG000000245869 | –2.0890 | 7.59E–05 | PCAT1 | ENSG00000253438 | 2.4776 | 4.83E–07 |
| C17orf77 | ENSG00000182352 | 4.1557 | 1.43E–03 | POU6F2-AS1 | ENSG00000242212 | 5.8205 | 4.29E–03 |
| C20orf166-AS1 | ENSG000000174403 | –3.4682 | 8.37E–20 | PVT1 | ENSG00000249859 | 2.6328 | 2.82E–18 |
| C2orf48 | ENSG000000163009 | 2.2264 | 3.27E–05 | PVRN1 | ENSG00000259095 | –2.2617 | 1.55E–04 |
| C7orf69 | ENSG000000136275 | 3.2131 | 9.71E–03 | RBMS3-AS3 | ENSG00000235904 | –2.7161 | 1.60E–13 |
| CLDN10-AS1 | ENSG00000223392 | 4.5630 | 6.72E–04 | SFTA1P | ENSG00000225383 | –2.7508 | 1.59E–11 |
| CRNDE | ENSG000000245694 | 4.4129 | 1.47E–12 | ST7-OT4 | ENSG00000214188 | 2.4463 | 6.43E–04 |
| DLX6-AS1 | ENSG000000231764 | 4.3820 | 1.38E–03 | UCA1 | ENSG00000214049 | 3.9110 | 1.06E–04 |
| EGOT | ENSG000000235947 | 2.3163 | 2.49E–03 | WASIR2 | ENSG00000231439 | 2.1913 | 8.49E–03 |

Abbreviations: FDR, false discovery rate; log_FC, log fold change.