Association of Vitamin D With Insulin Resistance and \(\beta\)-Cell Dysfunction in Subjects at Risk for Type 2 Diabetes

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OBJECTIVE — To examine cross-sectional associations of serum vitamin D [25-hydroxyvitamin D, 25(OH)D] concentration with insulin resistance (IR) and \(\beta\)-cell dysfunction in 712 subjects at risk for type 2 diabetes.

RESEARCH DESIGN AND METHODS — Serum 25(OH)D was determined using a chemiluminescence immunoassay. Insulin sensitivity/resistance were measured using the Matsuda insulin sensitivity index for oral glucose tolerance tests (ISOGTT) and homeostasis model assessment of insulin resistance HOMA-IR. \(\beta\)-Cell function was determined using both the insulinogenic index (IGI) divided by HOMA-IR (IGI/IR) and the insulin secretion sensitivity index-2 (ISSI-2).

RESULTS — Linear regression analyses indicated independent associations of 25(OH)D with ISOGTT and HOMA-IR (\(P = 0.004\), \(P = 0.003\), and \(P = 0.0011\), respectively) after adjusting for sociodemographics, physical activity, supplement use, parathyroid hormone, and BMI.

CONCLUSIONS — Vitamin D may play a role in the pathogenesis of type 2 diabetes, as 25(OH)D concentration was independently associated with both insulin sensitivity and \(\beta\)-cell function among individuals at risk of type 2 diabetes.

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RESEARCH DESIGN AND METHODS — A detailed methodology for this study has been described previously (8). Briefly, participants in the PROspective Metabolism and ISlet cell Evaluation (PROMISE) cohort were recruited from Toronto and London, Ontario, Canada, from 2004 to 2006. Participants were 30 years of age and older and at high risk for type 2 diabetes and/or metabolic syndrome (8). The current study includes 712 subjects, 92% of whom were free of diabetes based on oral glucose tolerance tests (OGTTs). None had known diabetes at the time of the assessments.

Fasting blood samples were collected and 75-g OGTTs were performed. Insulin sensitivity was quantified using the Matsuda insulin sensitivity index for oral glucose tolerance tests (ISOGTT) (9), and IR was measured using the homeostasis model assessment of insulin resistance (HOMA-IR) index (10). \(\beta\)-cell dysfunction was determined by dividing the insulinogenic index (IGI) by HOMA-IR (IGI/IR) (11) and by calculating the insulin secretion sensitivity index-2 (ISSI-2) (12).

Serum vitamin D, specifically 25-hydroxyvitamin D [25(OH)D], was measured using DiaSorin’s “25-OH vitamin D TOTAL” competitive chemiluminescence immunoassay on an automated LIAISON analyzer (Stillwater, MN). BMI and waist circumference were determined using standardized procedures (8). Parathyroid hormone (PTH) was measured using an electrochemiluminescence immunoassay on the Roche Modular E170 analyzer (Laval, QC). Structured questionnaires assessed self-reported ethnicity, smoking and physical activity, and included an open-ended question on current medication and supplement use. Season was defined using the participant’s date of clinical assessment and categorized as May–October (summer/early fall) and November–April (winter/early spring).

SAS Version 9.1 (Cary, NC) was used for all analyses. Natural logarithmic transformations were applied for all non-normally distributed variables. Univariate analyses, including \(x^2\) tests, analysis of variance (ANOVA), and Spearman correlation were conducted to assess the relationship between serum 25(OH)D and potential covariates. Multiple linear regression analyses were conducted to investigate the independent associations of 25(OH)D with measures of insulin sensitivity/resistance (ISOGTT and HOMA-IR) and \(\beta\)-cell dysfunction (IGI/IR and ISSI-2). Model 1 adjusted for sex, age, ethnicity and season; model 2 additionally

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Vitamin D, IR, and β-cell function in at-risk subjects

Table 1—Multiple linear regression analysis for associations of vitamin D with measures of insulin sensitivity/resistance and β-cell function

| Outcome per unit increase in baseline 25(OH)D | Model 1* | Model 2† | Model 3‡ |
|---------------------------------------------|----------|----------|----------|
|                                             | β (95% CI) | R²     | β (95% CI) | R²     | β (95% CI) | R²     |
| Insulin sensitivity/resistance measures     |          |        |          |        |          |        |
| IS-OGTT                                     | 0.009 (0.005–0.013) | 0.004 (0.002–0.006) | 0.003 (0.001–0.005) |
| HOMA-IR                                     | 0.008 (0.005–0.011) | 0.007 (0.005–0.010) | 0.006 (0.004–0.009) |
| β-cell function measures                    |          |        |          |        |          |        |
| IS-OGTT                                     | −0.010 (−0.012 to −0.007) | <0.0001 | −0.0001 | <0.0001 |
| HOMA-IR                                     | 0.008 (0.005–0.011) | <0.0001 | 0.0001 | <0.0001 |
| IGRI                                         | 0.005 (0.003–0.007) | 0.004 (0.003–0.006) | 0.003 (0.002–0.005) |
| ISSI-2                                       | 0.002 (0.000–0.004) | 0.001 (0.000–0.003) | 0.0001 |

β is the coefficient

*Model 1: adjusted for age, sex, season, ethnicity. †Model 2: adjusted as in model 1 plus supplements, total physical activity, parathyroid hormone.‡Model 3: adjusted as in model 2 plus BMI. §Log transformations.

RESULTS — The sample included 498 (69.9%) females and 462 (64.9%) Caucasians, and the mean age of the participants was 49.6 ± 10.0 years. The mean serum 25(OH)D concentration was 55.8 ± 22.90 nmol/l (range 10.0–161.0). Participant characteristics across quartiles of 25(OH)D concentration and correlations for continuous variables are presented (online Table A1, available in an online appendix at http://care.diabetesjournals.org/content/full/dc09-2321/DC1). A significant seasonal effect was evident, with higher 25(OH)D concentrations in the summer/early fall (n = 343; 59.11 ± 23.71 nmol/l) than in the winter/early spring (n = 351; 52.58 ± 21.64 nmol/l) (P = 0.0002). Univariate analyses indicated a significant positive association between 25(OH)D and IS-OGTT (r = 0.30, P < 0.0001), a significant negative association between 25(OH)D and HOMA-IR (r = −0.29, P < 0.0001), as well as significant positive associations between 25(OH)D and IGRI and ISSI-2 (r = 0.14, P = 0.0002) and ISSI-2 (r = 0.14, P = 0.0002).

In multivariate regression analyses, serum 25(OH)D was a significant independent predictor of insulin sensitivity (IS-OGTT and HOMA-IR) and β-cell function (IGRI and ISSI-2) across all models (Table 1). There was a slight attenuation of the association of 25(OH)D on measures of insulin sensitivity and β-cell function after additional adjustment for BMI, but the association remained significant.

We found a significant interaction by BMI, reflecting weaker magnitudes of association of 25(OH)D with measures of insulin sensitivity and β-cell function in obese individuals (BMI ≥30 kg/m²) (online Table A2).

CONCLUSIONS — This study demonstrated independent associations of 25(OH)D with both insulin sensitivity and β-cell function in subjects without known diabetes, the majority of whom were free of diabetes based on OGTTs. The major contribution of this study is the finding of an association between vitamin D and β-cell function. Previous studies assessing the association between 25(OH)D and β-cell function have yielded inconsistent results (2–4,13,14), possibly resulting from small sample sizes, the use of indirect measures of β-cell function (i.e., primarily fasting-based measures), and/or the lack of adjustment for background IR. In contrast, our study found a significant positive association between vitamin D and β-cell function, using validated measures of β-cell function which account for the hyperbolic relationship between insulin secretion and insulin sensitivity (12).

Although an inverse association between 25(OH)D and IR has been observed in previous studies (3–5,7), the majority of these studies relied primarily on simple fasting-based measures and most did not adjust for physical activity or PTH. In addition, negative findings have been reported, even when more direct measures of insulin sensitivity were used (2,6). Possible reasons for this discrepancy in findings may be due to small sample sizes or differences in study populations. The negative findings of Gulseth et al. (2) among those with the metabolic syndrome may be attributable to the sequestering of 25(OH)D in adipose tissue (15), resulting in reduced bioavailability of 25(OH)D. Similarly, we found a weaker association of vitamin D with IR and β-cell function in those with a BMI ≥30 kg/m².

Strengths of the current study include the measurement of serum 25(OH)D concentration, as well as the use of validated measures of both IR and β-cell dysfunction. In addition, the current study included a large, well-characterized multi-ethnic sample. Limitations include the cross-sectional design, and the lack of “gold standard” measures of IR and β-cell function, which are invasive and costly to use in large studies. Lastly, 25(OH)D was measured in blood samples obtained across different seasons, although we controlled for a seasonal effect and assessed potential interactions.

In conclusion, vitamin D was significantly related to IR and β-cell function in a multi-ethnic sample at risk for type 2 diabetes. Further research is needed on the prospective association between vitamin D and the underlying disorders of type 2 diabetes in large population-based studies.

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