Autochthonous probiotic mixture improves biometrical parameters of larvae of *Piaractus mesopotamicus* (Caracidae, Characiforme, Teleostei)

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ABSTRACT: Probiotics are a novel alternative to antibiotics as growth factors. Previously, our group isolated, selected and tested in vivo, eight autochthonous strains. They showed no significant effects when administered individually. However, the best doses, stages and ways of administration were combined in a multi strain formula. The aim of this research was to evaluate the effect of this probiotic product on the growth and survival of *Piaractus mesopotamicus* larvae. The administration was implemented during egg incubation and endogenous feeding period (5), during larvae exogenous feeding period (10) and all along the experiment (15). A group without microorganisms was used as control. The probiotic generates significant increments of mean weight and not significant increases of survival and biomass in two of the three tested stages. These results demonstrate the effectiveness of an autochthonous probiotic formula for the culture of this native fish species.

Key words: Lactic acid bacteria, Bacillus, Pacú, larviculture.

INTRODUCTION

Fishing capture is diminishing since 1996 (FAO 2016), while worldwide fish consumption is expected to increase until 2030 (WORLD BANK, 2013). The generated gap is fulfilled by increments in aquaculture (FAO 2009 and 2008). *Piaractus mesopotamicus* (Pacú) is a native fish species from Parana River and the most produced fish in Argentinian aquaculture. In the last years its production stagnated, probably due to deficits of animals for fattening, breeding and reproduction. Low mean weight and survival values could be due to a low knowledge of the staff, high stress conditions of animals and scarce technology applied during production process (MINISTERIO DE AGRICULTURA, GANADERÍA, PESCA Y ALIMENTOS DE LA NACIÓN ARGENTINA, 2015).

Antibiotics were used as growth factors to increment productivity. However, their use could produce residual drugs and affect consumers and expose not only the environment but also untargeted animals to these drugs (SMITH, 2008). Worldwide Organizations prohibited the use of antibiotics as growth promoters in animals for human consumption (EFSA, 2008), generating the need of novel strategies to increment productivity, while avoiding the use of chemotherapeutic and related drugs. Probiotics emerged as a putative solution.
They are defined for aquaculture as “a live microbial adjunct which has a beneficial effect on the host by modifying the host associated or ambient microbial community, by supporting an improved use of feed or enhancing its nutritional value, by stimulating the host response or by improving the quality of its ambient environment” (VERSCHUERE et al., 2000). The composition of the probiotic product is a critical aspect. Commercial probiotics have shown relative ineffectiveness, probably for including non-autochthonous strains. Isolates from non-aquatic animals have low possibilities to survive and adhere in the intestinal environment of fish (RIDHA & AZAD, 2015). Another aspect to consider is that several authors confirm that the administration of more than one strain could generate higher benefits by the complementation of beneficial properties or synergic effect (DE et al., 2014).

Previously, our group isolated, selected and tested in vivo eight autochthonous strains: four Enterococcus faecium, three Bacillus subtilis and one Pediococcus acidilactici. The results of these tests allowed us to select the most efficient doses of each microorganism for the formulation of a multi strain probiotic product for future assays (GUIDOLI et al., 2015, 2016a and 2016b).

The aim of this research was to determine the viability of the selected bacteria in the aquaculture environment and live food suspension and to evaluate the effect of the multi strain probiotic product on the biometrical parameters of P. mesopotamicus eggs and larvae under intensive culture system.

MATERIALS AND METHODS

The following methods and techniques were planned in order to prove the hypothesis that the administration of a multi-strain probiotic, formulated with strains able to resist the environmental conditions and included in an adequate dose, improves the biometrical parameters of larvae of Piaractus mesopotamicus.

Facilities, Animals and microorganisms

The present study was carried out in the facilities of the Instituto de Ictiología del Nordeste (INICNE) and Cátedra de Microbiología from the Facultad de Ciencias Veterinarias of the Universidad Nacional del Nordeste in the province of Corrientes, Argentina.

The larvae were obtained by controlled reproduction of breeders maintained in the facilities of INICNE. Spawning was induced by injection of pituitary extract from Prochilodus lineatus. Sexual gametes were obtained by stripping, immediately mixed, suspended in fresh water, washed and re-suspended to obtain the desired concentration (GÓMEZ et al., 2014).

Bacterial strains were isolated from Piaractus mesopotamicus, selected by their in-vitro expression of beneficial properties, genetically identified, registered in culture collections, tested in vivo individually and protected by patent aims (CONICET & UNNE, 2013; GUIDOLI et al., 2015, 2016a and 2016b). Lactic acid bacteria (LAB) and Bacillus cells were grown at 37°C in LAPTg (1% yeast extract, 1.5% peptone, 1% tryptone, 1% glucose, 0.1% Tween 80; pH 7.2) and Nutrient Broth (Britania©), respectively. Bacterial cells were obtained by centrifugation, and re-suspended in saline solution at the desired concentrations. The multi-strain suspension (MIX) contains the eight strains combined in previously determined concentrations (GUIDOLI et al., 2016a and 2016b) (Table 1).

Table 1 - Autochthonous putative probiotics and their doses in the MIX suspension.

| Gen Bank Accession no. | Strain | Dose in MIX (CFU L⁻¹) |
|------------------------|--------|----------------------|
| KJ740152               | E. faecium CRL 1937 | 6x10⁷ |
| KJ740153               | E. faecium CRL 1938 | 6x10⁷ |
| KJ740154               | P. acidilactici CRL1939 | 6x10⁴ |
| KJ740156               | E. faecium CRL 1941 | 6x10⁴ |
| KJ740155               | E. faecium CRL 1940 | 6x10⁴ |
| KJ754388               | B. subtilis A252    | 6x10⁷ |
| KJ740157               | B. subtilis A253    | 6x10⁷ |
| KJ740158               | B. subtilis A254    | 6x10⁷ |

Ciência Rural, v.48, n.7, 2018.
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Characterization of bacteria in the experimental units and the live food suspension  
Aliquots of 10mL sterile fishbowls’ water were inoculated, individually, with $1 \times 10^{10}$ CFU mL$^{-1}$ of each strain and maintained at room temperature for 4h. Viability was evaluated every hour by the technique of serial dilutions, subjected to subsequent inoculation in specific media and incubation at 37°C for 24h. The number of viable cells was expressed as the number of colony forming units (CFU) per mL of sample (CFU mL$^{-1}$). The same procedure was applied to tubes containing 10mL of a suspension containing 2,000 *Artemia* sp. mL$^{-1}$ in order to evaluate the bacteria viability in the pellet and in the supernatant of the centrifuged suspensions. Both assays were performed by triplicate.

Experimental design  
Experimental units consisted of 5-L plastic fishbowls, with constant water exchange and 300 fecundated eggs. There were a control group and three different treatments: a 5-days period administration ranging from eggs fertilization to the beginning of exogenous feeding - treatment 5-, a 10-days period ranging from the beginning of exogenous feeding to the end of the assay -treatment 10- and a 15-days period ranging from eggs fertilization to the end of the assay - treatment 15-. Bacteria were administered four times a day, directly into water the first 5 days and together with live food (*Artemia* sp.), after a 2-hours co-incubation, from day 6 up to day 15. At the end of the experiment animals were counted and weighed in order to obtain values of survival -expressed as percentage-, mean weight and total produced biomass. For microbial evaluations, three larvae of each replica were, individually, washed three times with sterile water and homogenized in 2mL of sterile saline solution. The obtained suspension was plated into LAPTg agar for LAB and Nutrient agar, with or without previous heating at 80°C for 15 minutes, for *Bacillus* and general microorganisms, respectively. Results were expressed as CFU animal$^{-1}$. Finally, *macro and microscopic evaluations* were performed to assess the normal development and behavior of animals. Histological evaluations were implemented to detect adverse effects in the normal tissue structures.

Results were analyzed by means of a one-way analysis of variance (ANOVA), using a completely randomized design with one control, three treatments and three replications ($n=12$). Duncan’s multiple range test was used as a post hoc test to compare differences between means. Statistical significance was settled at a probability value of $p<0.05$. The software used for data analysis was Statistica 6.0 for Microsoft Windows.

Ethics and biosafety  
Approved by protocols 0019/14-2011-02204 and 14-2012-03865 by the Ethics and Biosafety Committee - Facultad de Ciencias Veterinarias - Universidad Nacional del Nordeste, Argentina.

RESULTS AND DISCUSSION  
The probiotic treatment proposed in this paper was formulated from previously obtained results of *in vivo* tests, based on the idea that multi strain autochthonous probiotic products could generate higher benefits by complementation of beneficial properties (DE et al., 2014). The experimental design was planned to fit with the normal procedures performed in any aquaculture production in order to facilitate the use of the potential probiotic product. *Artemia* sp. not only has a proper size and great nutritional value but also is an adequate vehicle to deliver other substances such as nutrients, antimicrobials, vaccines and probiotics (AZIMIRAD et al., 2016; JAMALI et al., 2015). Thus, in the first stage of this investigation, it was proved that all strains remain viable in the water of the experimental units with no significant modifications in the number of viable cells after 4 hours. Same results were obtained in the pellet-*Artemia* nauplii- and supernatant -water of the suspension- of the live feed suspension. Figure 1 shows that there are no significant differences -$p>0.05$- between the affinity of each strain and the supernatant of the *Artemia* suspension. However, the adherence to the *Artemia* nauplii shows significant differences -$p<0.05$- with *E. faecium* CRL 1940, and *B. subtilis* A252, A253 and A254 presenting the highest values. These results indicated the survival of selected strains in the environment of fishes and their ability to adhere to the vehicle used to deliver the microorganisms.

The results of the *in vivo* tests show that all treatments induce higher average values of mean weight, survival and produced biomass than in the control group. However, only the values of mean weight presented significant differences -$p<0.05$-, with groups administered in stages 10 and 15 differentiating significantly from control and administered-in-stage-5 groups. These increments
represent a 109.24% and 103.07% for stages 10 and 15, respectively, referred to control group (Figure 2A). Survival increases, non-significantly, 2.00, 2.04 and 2.20 times compared with control values in treatments 5, 10 and 15, respectively (Figure 2B). Produced biomass in all treatments is also higher than in control group, with increments of 140.15%, 419.30% and 394.59% for stages 5, 10 and 15 respectively and without significance (Figure 2C). Literature in the area shows in vitro evaluations, modifications of immunological parameters and in vivo effects of potentially probiotic strains in animals challenged with pathogens. However, only a few articles describe probiotics able to increase biometrical parameters under non-challenge conditions (IBRAHEM, 2015). Our findings are consistent with those obtained by ABD EL-RHMAN et al. (2009), ZHOU et al. (2010), JATOBA et al. (2011) and ABUMOURAD et al. (2014), who described significant increments in the mean weight of Nile tilapia after the administration of different probiotics, with no modifications in the survival rate.

Microbial evaluations showed no significant differences \( p > 0.05 \) for total microorganisms between groups. Sporulated microorganisms are absent in the control group while its number increases significantly \( p < 0.05 \) in treated groups. Conversely, the LAB count is significantly higher \( p < 0.05 \) in the control than in treated groups (Figure 2D). In opposition to our findings, AZIMIRAD et al. (2016) reported significant increments in both, total microorganisms and LAB counts, accompanied by significant increments in weight gain when administering Pediococcus acidilactici and fructose saccharide to angelfish (Pterophyllum scalare). Increment in the number of sporulated microorganisms after its
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administration to aquatic animals was previously described by our group (GUIDOLI et al., 2016a). Also, JAMALI et al. (2015) and CHAI et al. (2016) demonstrated that shrimps administered with higher doses of *Bacillus* strains presented higher counts of sporulated microorganisms in their guts. In this research the differences in the number of *Bacillus* cannot be related with the increment in the biometrical parameters. However, the studies in this research do not differentiate between genera, species or strains. Thus, the relative proportions of species or strains in each group of microorganisms could vary between treatments without being detected with the applied techniques. Further studies must be performed in order to monitor the proportions of the specific genera or species used in this research.

The development, behavior and histological structure of major organs, mainly those related to the digestive system (Figure 3), showed no differences between the control and treated groups.

There are no previous research stating the improvement of biometrical parameters of *Piaractus mesopotamicus*, the most produced native species in Argentinian aquaculture, by the administration of autochthonous probiotics. Results of the present work, allowed us to conclude that the use of the multi strain probiotic formula “MIX” would increment the survival, mean weight and produced biomass of *Piaractus mesopotamicus* under intensive production systems. These increments would not only raise the national production by supporting an increase of two important biological productivity factors such as growth and survival but also by encouraging other investors to participate in a more profitable entrepreneurship.

Figure 2 - Mean weight (A), Survival (B), Biomass (C) and Microbial determinations (D) of *Piaractus mesopotamicus* larvae after bacterial treatment with MIX suspension during the first 5, 10 and 15 days of life respectively. Vertical bars indicate Standard Error of means (SE). Different letters indicated significant differences between treatments.
DECLARATION OF CONFLICT OF INTEREST

The authors declare no conflict of interest. The founding sponsors had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, and in the decision to publish the results.

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Figure 3 - Histology of stomach (*), liver (#) and intestine (X) of control (a) compared with animals treated with MIX suspension during the first 5 (b), 10 (c) and 15 (d) days of life respectively. Bars indicate 100µm.
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