Tumor-Associated Macrophages in Tumor Immunity

Yueyun Pan¹,², Yinda Yu², Xiaojian Wang²* and Ting Zhang¹,²*

¹ Department of Radiation Oncology, Second Affiliated Hospital, School of Medicine, Zhejiang University, Hangzhou, Zhejiang, China, ² Institute of Immunology, School of Medicine, Zhejiang University, Hangzhou, Zhejiang, China

Tumor-associated macrophages (TAMs) represent one of the main tumor-infiltrating immune cell types and are generally categorized into either of two functionally contrasting subtypes, namely classical activated M1 macrophages and alternatively activated M2 macrophages. The former typically exerts anti-tumor functions, including directly mediating cytotoxicity and antibody-dependent cell-mediated cytotoxicity (ADCC) to kill tumor cells; the latter can promote the occurrence and metastasis of tumor cells, inhibit T cell-mediated anti-tumor immune response, promote tumor angiogenesis, and lead to tumor progression. Both M1 and M2 macrophages have high degree of plasticity and thus can be converted into each other upon tumor microenvironment changes or therapeutic interventions. As the relationship between TAMs and malignant tumors becoming clearer, TAMs have become a promising target for developing new cancer treatment. In this review, we summarize the origin and types of TAMs, TAMs interaction with tumors and tumor microenvironment, and up-to-date treatment strategies targeting TAMs.

Keywords: tumor-associated macrophages, regulation, immunosuppression, tumor microenvironment, tumor therapy

INTRODUCTION

Macrophages play critical roles in both innate and adaptive immunity and are known for their remarkable phenotypic heterogeneity and functional diversity. Embryonic hematopoietic stem cells in a variety of tissues during fetal development and differentiate into tissue-specific resident macrophages, including Kupffer cells in the liver, alveolar macrophages in the lung, and osteoclasts in bone tissue. After birth, bone marrow-derived precursors in particular circulating monocytes can also differentiate into macrophages in steady state or during tissue inflammation (1). Macrophages are involved in tissue and systemic inflammation and immunity, as well as tissue reconstruction. They have a wide range of functions, including phagocytosis, antigen presentation, defense against microbial cytotoxicity, and secretion of cytokines, complement components, etc. (2). It is worth noting that the broad biological activities of macrophages often have diametrically opposite characteristics, such as inflammatory response and anti-inflammatory activity; immunogenic and inducing immune tolerance; causing tissue destruction and repairing (3).

Tumor-associated macrophages (TAMs) are macrophages that participate in the formation of the tumor microenvironment. TAMs are widely present in various tumors (4). TAMs can promote tumor growth, invasion, metastasis, and drug resistance (5). It has been proposed that functional
difference of macrophages is closely related to the plasticity of macrophages, and its functional phenotype is regulated by molecules in tumor microenvironments.

In this review, we discuss the origins and types of TAMs, the interaction between tumors and the tumor microenvironment, and review the emerging strategies for cancer treatment via targeting TAMs.

**ORIGINS AND TYPES OF TAMs**

**Origins**

For a long period of time, it is believed that macrophages in tumors are exclusively recruited from the periphery by chemotaxis and generated by monocytic precursors in the local environment. However, more recent evidence shows that at least certain tumors, and generated by monocytic precursors in the local environment. Despite this belief, there have been studies showing that monocytic-derived but not embryonic-derived resident macrophages are capable in supporting the growing body of TAMs in the inflammatory environment of tumor, the potentially different roles of monocytic- versus embryonic-derived TAMs on tumor development and/or progress remains an intriguing question that is largely unanswered (2).

M-MDSCs (monocyte-related myeloid-derived suppressor cells) are currently known as another main circulating precursor of TAMs. MDSCs are a type of myeloid leukocytes that is related to immunosuppression (7). Based on surface markers Ly6C+/Ly6C- and Ly6C-/Ly6G+, MDSCs can be divided into monocyte (M)-related and granulocyte (G)-related MDSC. Among them, M-MDSCs are induced into TAMs by various chemokines (8).

It is all known that macrophages derive from bone marrow-derived monocytic precursors. In tumors, TAMs mainly originate from bone marrow monocytes, but recent evidence suggests that, recruitment of circulating monocyes is essential for TAMs accumulation. Circulating inflammatory monocytes could be recruited by multiple chemokines (CCL2 and CCL5) and cytokines (CSF-1 and members of the VEGF family) to tumor (9). Tumor growth can also induce the differentiation of CCR2+ monocytes into TAMs (10).

Furthermore, complement components, particularly C5a, are an important mediator of the recruitment and functional polarization of TAMs (11). Indeed, such chemokines do more than attractants do because they activate transcription programs that help macrophages tilt toward the functional of a particular phenotype (12). At the same time, CSF-1 is a monocyte attractant, as well as macrophage survival and polarization signals, which drive TAM to immunosuppressive differentiation M2 macrophages (13). Unlike CSF-1, GM-CSF activates macrophage function associated with antitumor activity (14).

**Types**

Macrophages undergo specific differentiation in different tissue environments, and can be divided into two different polarization states: M1 type macrophages (M1) and M2 type macrophages (M2).

M1 can respond to dangerous signals transmitted by bacterial products or IFN-γ, which attracting and activating cells of the adaptive immune system; an important feature of M1 is that it can express nitric oxide synthase (iNOS) and reactive oxygen species (ROS) (15–17) and cytokine IL-12 (18). M1 also has the function of engulfing and killing target cells.

M2 expresses a large number of scavenger receptors, which is related to the high-intensity expression of IL-10, IL-1β, VEGF and matrix metalloprotein (MMP) (19, 20). M2 has the function of removing debris, promoting angiogenesis, tissue reconstruction and injury reparments, as well as promoting tumorigenesis and development (4).

It is worth noting that the polarization of macrophages into M2 appears to be oversimplified. Some people have classified M2 macrophages into M2a (induced by IL-4 or IL-13), M2b (induced by immune complexes combined with IL-1β or LPS) and M2c (induced by IL-10, TGFB, or glucocorticoid), and M2d (conventional M2 macrophages that exert immunosuppression) (21, 22).

**THE ROLE OF TAMs IN TUMOR PROGRESS**

Current studies have shown that TAM population is in a state of constant transition between the two forms of M1 and M2 type. The proportion of each form is determined by the type and concentration of different signals in the tumor environment (Figure 1).

**M1 Macrophages and Tumor Suppression**

M1-type macrophages have anti-tumor effects, which can distinguish tumor cells from normal cells. By identifying tumor cells and ultimately killing tumor cells, studies have found that M1 type macrophages have two different effects on killing tumor cells mechanism. M1 type macrophages directly mediate cytotoxicity to kill tumor cells: macrophage-mediated cytotoxicity is a slow process (generally requires 1 to 3 days) and involves multiple mechanisms. For example, macrophages release tumor killing molecules such as ROS and NO, which have cytotoxic effects on tumor cells (23). The other is antibody-dependent cell-mediated cytotoxicity (ADCC) killing tumor cells: ADCC requires less time to kill tumor cells (generally within a few hours) and requires the participation of anti-tumor antibodies (24).

**M2 Macrophages Promote Tumor Cell Proliferation and Invasion**

TAM infiltration is closely related to tumor cell proliferation. Many studies have shown that TAMs can express a variety of cytokines that stimulate tumor cell proliferation and survival, including epithelial growth factor (EGF), platelet-derived growth factor (PDGF), TGF-β1, hepatocyte growth factor (HGF), and epithelial growth ligands of the factor receptor (EGFR) family.
FIGURE 1 | A schematic representation of the roles of tumor-associated macrophages (TAMs) in tumor progression. TAMs can mediate immune response, tumor cell proliferation and invasion, angiogenesis and metastasis. MMP, matrix metalloprotein; M-MDSCs, monocyte-related myeloid-derived suppressor cells; CSF1, macrophage colony-stimulating factor; VEGF, vascular endothelial growth factor; ROS, reactive oxygen species; INOS, nitric oxide synthases; LIF, leukocytosis induced factor.
and basic fibroblast growth factor (BFGF) (25). The ligands of the EGFR family play an important role in tumorigenesis, especially breast and lung cancers. Members of this family can form homo- or heterodimers on the cell surface, mediating the transduction of cell proliferation signals. In all, TAMs are an important cell source for EGF secretion in tumor tissues (25).

As for invasion, in glioma cells, extracellular adenosine deaminase protein cat eye syndrome critical region protein 1 (CECR1) has been shown to regulate the maturation of macrophages. CECR1 is induced by M2-like TAM secretory effects activate MAPK signaling and stimulate the proliferation and migration of glioma cells (26). Another investigation shows that a positive feedback loop of CCL5 and CCL18 between TAMs and myofibroblast is constituted to drive the malignant invasion of phyllodes tumor (PT). CCL5 binds to CCR5, and activates the AKT signal to recruit and repolarize TAMs. TAMs release CCL18 to further induce the invasion of malignant PTs by differentiating the mesenchymal fibroblasts to myofibroblast, causing the malignancy of PTs (27).

**TAMs Promote Tumor Metastasis**

Tumor metastasis is an important feature of poor prognosis after tumor therapy. The main reason for tumor cell migration and metastasis is the degradation and damage of tumor tissue endothelial cell basement membrane. It has been reported that activated TAMs exert a direct effect on promoting metastasis via directly producing soluble factors (28). M2 macrophages can destroy matrix membrane of endothelial cells by secreting matrix metalloproteinases (MMPs), serine proteases, cathepsins, and decompose various collagen and other components of extracellular matrix, thereby helping the migration of tumor cells and tumor stromal cells (19, 20). Epithelial-mesenchymal transition (EMT) is the basis of tumor metastasis (29). This process enables tumor cells to acquire the ability to migrate and endows them with the properties of stem cells (30). Besides, cytokines produced by tumor cells also promote the differentiation process of TAMs, thus forming a positive feedback loop between TAMs and EMT (31).

**M2 Macrophages Promoting Angiogenesis**

TAMs are enriched in hypoxic areas with poor blood supply (1). Proangiogenic effects by TAMs involves the coordinated regulation of a wide range of cytokines, including BFGF, VEGF, IL-1, IL-8, TNF-α, MMP-9, MMP-2, and nitric oxide (NO). The coordinated expression of these molecules promotes the proliferation of endothelial cells, matrix remodeling and vascularization in time and space. Macrophages can release the angiogenic molecules and express a series of enzymes involved in the regulation of angiogenesis, including MMP-2, MMP-7, MMP-9, MMP-12, and cyclooxygenase-2 (20, 32).

However, metabolism still exists in angiogenesis, and it is still unknown whether changes in metabolism affect these functions. Hypoxic TAM strongly up-regulates the expression of mTOR’s negative regulator REDD1. REDD1-mediated mTOR inhibition can hinder glycolysis in TAM and reduce its excessive angiogenic response, thereby forming abnormal blood vessels (33).

**Immune Regulation by TAMs**

TAM can regulate the killing effect of T cells and NK cells on tumor cells. M1 macrophages increased the number of total and activated natural killer (NK) cells in fibrotic liver, released TNF-related apoptosis-inducing ligand (TRAIL), and induced HSC apoptosis (34). HCC-derived exosomes induced macrophages to upregulate the expression of IFN-γ and TNF-α in T cells, while the expression of inhibitory receptors PD-1 and CTLA-4 was upregulated (35). In mesothelioma, the macrophages isolated from pleural effusion showed the M2 phenotype were negatively correlated with T cells in vivo, which emphasized the use of macrophages as treatments in mesothelioma Target potential (36).

In addition to these functions, TAMs can also directly inhibit CD8+ T-cell proliferation through metabolism of L-arginine via arginase 1, iNOS, oxygen radicals or nitrogen species (37–39). Besides, TAMs recruit Tregs through CCL22 (40), which further suppress the antitumor immune response of T-cells. Conditional TAM ablation blocks Treg cell recruitment and inhibits tumor growth by lowering the CCL20 level of xenograft mice (41).

Substantial evidence indicates that the inflammatory reaction at a tumor site can promote tumor growth and progression. Inflammation and immune evasion are considered as hallmarks of cancer. It has been reported that TAMs can also contribute to cancer-related inflammation that leads to tumorigenesis by generation of inflammatory Th subset such as TFH (42). Toll-like receptor 4 (TLR4)-induced monocyte inflammation is important for induction of IL21+ TFH-like cells, which operate in IL21-IFNγ-dependent pathways to induce plasma cell differentiation and thereby create ideal conditions for M2b macrophage and cancer progression (42) (Figure 1). These suggest that strategies to influence functional activities of inflammatory cells may benefit anticancer therapy.

**FACTORS REGULATING TAMs FUNCTIONS**

TAMs are a collection of multiple cell types with a wide range of functional effects under steady state and pathological conditions. This diversity is regulated by many different factors, such as the tumor cell-derived soluble molecules, tumor metabolic alterations, other immune cells and other factors (Figure 2).

**Tumor Cell-Derived Soluble Molecules**

TAMs can be activated and polarized by tumor cell-derived soluble molecules, thereby promoting tumor progression and metastasis. Tumor cells secrete the sonic hedgehog (SHH), and tumor-derived SHH drives TAM M2 polarization. Hh-dependent polarization of TAM suppresses the recruitment of CD8+ T cells to TME via inhibiting CXCL9 and CXCL10, mediating TAM immunosuppression mechanism (43). In addition, kynurenine produced by glioblastoma cells can activate the aromatic hydrocarbon receptor (AHR) in TAMs, and AHR can drive KLF4 expression and inhibit NF-κB...
activation in TAMs, which regulate TAM function and T cell immunity (44). Cancer cells can also release succinate into their microenvironment and activate the succinate receptor (SUCNR1) signal, thereby polarizing macrophages to TAMs (45). Meanwhile, there is a positive correlation between the expression of osteopontin (OPN) in tumor cells and TAMs infiltration. OPN promotes chemotaxis migration and activation of TAMs (46). Also, when mucin MUC1 is expressed on cancer cells and is decorated with multiple short, sialylated O-linked glycans (MUC1-ST), which will induce TAM to express M2-like phenotype (47).

**Tumor Metabolic Alterations**

It is worth noting that macrophage polarization is correlated with distinct metabolic characteristics pertaining to glucose metabolism (48, 49), lipid metabolism (50), and glutamine metabolism (51). Such metabolic alterations can also determine the phenotype and function of TAMs in promoting the cancer progression (52).

Cancer cells can utilize metabolic byproducts to take the control of tumor-infiltrating immune cells to their own benefit. For example, lactate secreted by glycolysis in cancer cells, which transfers the polarization of TAMs from a pro-inflammatory (M1-like) to an anti-inflammatory (M2-like) phenotype (53, 54). Another research shows that membrane cholesterol efflux drives TAM reprogramming and tumor progression. Ovarian cancer cells promote membrane cholesterol efflux, and increased cholesterol efflux promotes IL-4 mediated signaling in TAMs, which will promote tumor invasion and metastasis (55). In addition, glutamate-ammonia ligase (GLUL) favors M2-like TAMs polarization by catalyzing the conversion of glutamate into glutamine, and GLUL inhibition can transfer M2-like TAMs into M1-like phenotype by increasing glycolytic flux and succinate availability (51).

**Regulated by Immune Cells**

TAMs can be regulated by other immune cells, such as Treg cells, MDSCs and B cells. IFN-γ is the main cytokines responsible for inhibiting M2-like TAM. Treg cells can inhibit IFN-γ secreted by CD8+ T cells, which will prevent the activation of fatty acid synthesis that mediated by sterol regulatory element binding protein 1 (SREBP1) in immunosuppressive M2-like TAM. Therefore, Treg cells indirectly but selectively maintain M2-like TAM metabolic adaptability, mitochondrial integrity and survival rate (56). In addition, MDSCs also regulate TAM differentiation and promote tumor proliferation by downregulation of STAT3 (57). Besides, B cells are the key factors determining the tumor promoting function of TAMs. B cells can induce M2b macrophage polarization in human HCC (58), as well as suppress other immune cells, such as CD8+ T cells and M1 macrophages in the tumor microenvironment and promote the proliferation of cancer cell (59). Depletion of B cells prevented generation of M2b, increased the activity of anti-tumor T cell response, and reduced tumor growth.
Regulation by Other Factors
There are also some other factors of tumor microenvironment that can regulate TAMs function. Autophagy in the tumor microenvironment can provide essential nutrients, nucleotides, and amino acids to the tumor cells, facilitating tumor growth (60). Autophagy proteins in myeloid cells in the tumor microenvironment help to activate TAM by influencing LAP and mediate immunosuppression of T lymphocytes (61). In non-alcoholic fatty liver disease (NAFLD), NLRC4 contributes to the polarization of TAM to M2 type and the production of IL-1β and VEGF, thereby promoting the growth of tumor (62). Moreover, C-Maf transcription factor is the main regulator of cancer-promoting TAM polarization. C-Maf can promote the immunosuppressive activity of TAMs and control its metabolic process (63).

TARGETING TAMs FOR CANCER TREATMENT
TAMs are one of the most important components of the tumor immunosuppression microenvironment with high degree of plasticity. TAMs have both M1 and M2 type and have the potential ability of repolarization to M1 type macrophages. Therefore, targeting TAMs is a new cancer treatment strategy, including limiting monocytes recruitment, targeting TAMs activation, reprogramming TAMs into anti-tumor activity, and targeting TAMs specific markers (Figure 2).

Limiting Monocyte Recruitment
One of the strategies for targeting TAMs is to block monocyte recruit to tumor tissue. Tumor cells recruit CCR2-expressing monocytes from the peripheral blood to the tumor site by releasing CCL2 and these recruit CCR2-expressing monocytes will finally mature into TAMs, which accelerate the tumor progress. Thus, targeting CCL2-CCR2 axis is a very effective method of cancer therapy. Blocking the CCL2-CCR2 axis could greatly reduce the incidence of tumors by preventing TAMs recruitment and enhance the anti-tumor efficacy of CD8+ T cells in the tumor microenvironment (64).

CSF1 signaling pathway plays a key role in the production of bone marrow monocytes and the polarization of TAMs in tumor tissues. CSF1 produced by tumor cells caused down-regulation of granulocyte-specific chemokine expression in HDAC2-mediated cancer-associated fibroblasts (CAF), thereby limiting the migration of monocytes to tumors. The combination of CSF1R inhibitor and CXCR2 antagonist can prevent granulocytes from infiltrating the tumor, showing a strong anti-tumor effect (65). Also, combination of anti-PD-1 and anti-CSF1R antibodies induces melanoma regression in mice (66).

Targeting TAM Activation
Targeted activation of TAMs is an effective tumor treatment method. One of them is inhibiting TAMs from promoting tumor cell activation. Epidermal cell growth factor (EGF) secreted by TAM activates EGFR on tumor cells, which in turn upregulates VEGF (vascular endothelial growth factor)/VEGFR signaling in surrounding tumor cells, thereby promoting the proliferation and migration of tumor cells. EGFR blockade or ICAM-1 (intercellular adhesion molecule) antibody neutralization in TAM reduced the occurrence of ovarian cancer in mice (25).

Another effective tumor treatment method is blocking inhibitory receptor signals on TAMs that promote phagocytosis and antigen presentation function. Tumor cells highly express CD47, which restricts the ability of macrophages to engulf tumor cells through the signal regulatory protein alpha (SIRPα) -CD47 signal. The destruction of the SIRPα-CD47 signal axis is effective against various brain tumors including glioblastoma multiforme (GBM) by inducing tumor phagocytosis (67). Leukocyte immunoglobulin-like receptor subfamily B (LILRB) family is a class of inhibitory receptors expressed by myeloid cells, and its ligands are MHCI-like molecules (68). LILRB1 is up-regulated on the surface of TAM, and the MHCI-like component β2-microglobulin expressed by cancer cells can directly protect it from being engulfed. Therefore, blocking MHC I molecules or LILRB1 can enhance TAM phagocytosis (69).

Targeting pre-tumor myeloid cells at the metabolic level is another therapeutic strategy. Immunosuppressive phenotype of TAMs is controlled by long-chain fatty acid metabolism (especially unsaturated fatty acids), which makes BMDMs polarized into M2 phenotypes with strong inhibitory ability. Therefore, chemical inhibitors can effectively block TAM polarization in vitro and tumor growth in vivo (70).

Reprogramming TAMs Into Anti-Tumor Activity
One of the key characteristics of macrophages is their plasticity, which allows them to change the phenotype according to the tumor microenvironment. Therefore, reprogramming TAMs into an anti-tumor phenotype is a very promising tumor treatment strategy. Anti-tumor macrophages (M1 type) have abilities to clear and destroy tumor cells. RP-182 can selectively induce conformational switching of the mannose receptor CD206 expressed on TAM expressing the M2 phenotype, reprogramming M2-like TAM into anti-tumor M1-like TAM phenotype (71). Another finding shows that serine/threonine protein kinase 1 (RIPI) interacting with receptors in TAMs in pancreatic ductal adenocarcinoma (PDA) is up-regulated. Targeting RIPI, which act as a checkpoint kinase, reprogram TAM toward MHCIIhi TNFα+ IFNγ+ phenotype (72).

Targeting Immune Inhibitory Molecules on TAMs
Targeting immune inhibitory molecules on TAMs is also an effective method. Blocking of MerTK leads to the accumulation of apoptotic cells in tumor cells and triggers a type I interferon response. MerTK blockade increases tumor immunogenicity and enhances anti-tumor immunity. Treatment of tumor-bearing mice with anti-MerTK antibodies can stimulate T cell activation and synergize with anti-PD-1 or anti-PD-L1 therapy (73). PD-1-PD-L1 therapy can also work by direct action on macrophages. Both mouse and human TAM express PD-1. The
expression of TAM PD-1 is negatively correlated with the phagocytic ability against tumor cells, and blocking PD-1-PD-L1 in vivo will increase the phagocytosis of macrophages, reduce tumor growth, and rely on macrophage-dependent ways to prolong the survival of mice in cancer models (74).

CONCLUDING REMARKS

Under the effect of the tumor microenvironment, TAMs are tamed by tumor cells and has become a promoter of tumor growth. Studies have shown that TAMs have a significant role in promoting the development and progress of tumors. Therefore, how to inhibit the tumor-promoting roles of TAMs will provide new clues for future tumor therapy. However, a number of key questions remain to be answered, including mechanisms of TAM development, key factors that drive phenotypic changes of TAMs in the tumor microenvironment. Recent pre-clinical and clinical studies aiming at targeting TAMs for cancer treatment have shown inspiring results. TAM-targeting therapy represents a promising treatment of cancer patients in the future.

AUTHOR CONTRIBUTIONS

YP and YY analyzed the data and wrote the paper. XW and TZ edited the manuscripts. All authors contributed to the article and approved the submitted version.

REFERENCES

1. Cassetta L, Pollard JW. Targeting macrophages: therapeutic approaches in cancer. Nat Rev Drug Discov (2018) 17(12):887–904. doi: 10.1038/nrd.2018.169
2. Mantovani A, Marchesi F, Malesci A, Laghi L, Allavena P. Tumour-associated macrophages as treatment targets in oncology. Nat Rev Clin Oncol (2017) 14(7):399–416. doi: 10.1038/nrclinonc.2016.217
3. Noy R, Pollard JW. Tumor-associated macrophages: from mechanisms to therapy. Immunity (2014) 41(1):49–61. doi: 10.1016/j.immuni.2014.06.010
4. Ngambenjawong C, Gustafson HH, Pun SH. Progress in tumor-associated macrophage (TAM)-targeted therapeutics. Adv Drug Deliv Rev (2017) 114:206–21. doi: 10.1016/j.addr.2017.04.010
5. Li XL, Pan Q, Cao HX, Xin FZ, Zhao ZH, Yang RX, et al. Lipotoxic Macrophage activation and polarization: nomenclature and experimental characterization standards. Nat Commun (2016) 7:12150. doi: 10.1038/ncomms12150
6. Zhu C, Kros JM, Cheng C, Mustafa D. The contribution of tumor-associated macrophages in glioma neo-angiogenesis and implications for anti-angiogenic strategies. Neuro-oncology (2017) 19(11):1435–46. doi: 10.1093/neuonc/nox081
7. Peglia F, Perego M, Gabrilovich D. Myeloid-derived suppressor cells coming of age. Nat Immunol (2018) 19(2):108–19. doi: 10.1038/s41590-017-0022-x
8. Bronte V, Brandau S, Chen SH, Colombo MP, Frey AB, Greten TF, et al. Recommendations for myeloid-derived suppressor cell nomenclature and characterization standards. Nat Commun (2016) 7:12150. doi: 10.1038/ncomms12150
9. Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. Nature (2008) 454(7203):436–44. doi: 10.1038/nature07025
10. Franklin RA, Liao W, Sarkar A, Kim MV, Bivona MR, Liu K, et al. The cellular and molecular origin of tumor-associated macrophages. Science (2014) 344(6186):921–5. doi: 10.1126/science.1252510
11. Bonavita E, Galdiero MR, Jaillon S, Mantovani A. Phagocytes as Corrupted Policemen in Cancer-Related Inflammation. Adv Cancer Res (2015) 128:141–71. doi: 10.1016/bs.acr.2015.04.013
12. Kitamura T, Qian BZ, Soong D, Cassetta L, Noy R, Sugano G, et al. CCL2-induced chemokine cascade promotes breast cancer metastasis by enhancing retention of metastasis-associated macrophages. J Exp Med (2015) 212(7):1043–59. doi: 10.1084/jem.20141836
13. Pyontek SM, Akkari L, Schuhmacher AJ, Bowman RL, Sevenich L, Quail DF, et al. CSF-1R inhibition alters macrophage polarization and blocks glioma progression. Nat Med (2013) 19(10):1264–72. doi: 10.1038/nm.3337
14. Murray PJ, Allen JE, Biswas SK, Fisher EA, Gilroy DW, Goedt S, et al. Macrophage activation and polarization: nomenclature and experimental guidelines. Immunity (2014) 41(1):14–20. doi: 10.1016/j.immuni.2014.06.008
15. Liu X, Pan Q, Cao HX, Xin FZ, Zhao ZH, Yang RX, et al. Lipotoxic Hepatocyte-Derived Exosomal miR-192-5p Activates Macrophages Through Rictor/Akt/Forkhead Box Transcription Factor O1 Signaling in Nonalcoholic Fatty Liver Disease. Hepatology (2020) 72(2):454–69. doi: 10.1002/hep.31050
29. Dongre A, Weinberg RA. New insights into the mechanisms of epithelial-mesenchymal transition and implications for cancer. *Nat Rev Mol Cell Biol* (2020) 21(2):69–84. doi:10.1038/s41580-018-0080-4

30. Song W, Mazzieri R, Yang T, Gobe GC. Translational Significance for Tumor Metastasis of Tumor-Associated Macrophages and Epithelial-Mesenchymal Transition. *Front Immunol* (2017) 8:1106. doi:10.3389/fimmu.2017.01106

31. Chaffer CL, San Juan BP, Lim E, Weinberg RA. EMT, cell plasticity and metastasis. *Cancer Metastasis Rev* (2016) 35(4):645–54. doi:10.1007/s10555-016-9648-7

32. Egawa M, Mukai K, Yoshikawa S, Iki M, Mukaida N, Kawano Y, et al. Inflammatory monocytes recruited to allergic skin acquire an anti-inflammatory M2 phenotype via basophil-derived interleukin-4. *Immunity* (2013) 38(3):570–80. doi:10.1016/j.immuni.2012.11.014

33. Wenes M, Shang M, Di Matteo M, Govea J, Martin-Perez R, Sernes J, et al. Macrophage Metabolism Controls Tumor Blood Vessel Morphogenesis and Metastasis. *Cell Metab* (2016) 24(5):701–15. doi:10.1016/j.cmet.2016.09.008

34. Ma PF, Gao CC, Yi J, Zhao JL, Liang SQ, Zhao Y, et al. Cytometry with T1-polarized macromolecules ameliorates liver fibrosis by modulating immune microenvironment in mice. *J Hepatol* (2017) 67(4):770–9. doi:10.1016/j.jhep.2017.05.022

35. Yin C, Han Q, Xu D, Zheng B, Zhao X, Zhang J. SALL4-mediated upregulation of exosomal miR-146a-5p drives T-cell exhaustion by M2 tumor-associated macrophages in HCC. *Oncoimmunology* (2019) 8 (7):1601479. doi:10.1080/2162402X.2019.1601479

36. Lievense LA, Cornelissen R, Bezemer K, Kaijen-Lambers ME, Hegmans JP, Aerts JG. Pleural Effusion of Patients with Malignant Mesothelioma Induces Macrophage-Mediated T Cell Suppression. *J Thorac Oncol* (2016) 11(10):1755–64. doi:10.1097/JTO.0000000000000921

37. Movahedi K, Laoui D, Gysemans C, Baeten M, Stange-atrice R, et al. Macrophage Metabolism Controls Tumor Blood Vessel Morphogenesis and Metastasis. *Cell Metab* (2016) 24(5):701–15. doi:10.1016/j.cmet.2016.09.008

38. Molon B, Ugel S, Del Pozzo F, Soldani C, Zilio S, Avella D, et al. Chemokine CXCL10 Promotes Tumor Associated Macrophage Polarization to a Regulatory Phenotype and Suppresses Tumor Inflammation and Metastasis. *Nature* (2014) 512(7509):559–63. doi:10.1038/nature13490

39. Goossens P, Rodriguez-Vita J, Eterodot A, Masse M, Rastoin O, Gourand V, et al. Membrane Cholesterol Efflux Drives Tumor-Associated Macrophage Reprogramming and Tumor Progression. *Cell Metab* (2019) 29(6):1376–89.e4. doi:10.1016/j.cmet.2019.02.016

40. Liu C, Chikina M, Deshpande R, Menk AV, Wang T, Tabib T, et al. Treg Cells Promote the SREBP1-Dependent Metabolic Fitness of Tumor-Promoting Macrophages via Repression of CD8α+ T Cell-Derived Interferon-γ. *Immunity* (2017) 51(2):381–97.e6. doi:10.1016/j.immuni.2016.09.017

41. Kumar V, Cheng P, Condamine T, Mony S, Languino LR, McCaffrey JC, et al. CD45 Phosphatase Inhibits STAT3 Transcription Factor Activity in Myeloid Cells and Promotes Tumor-Associated Macrophage Differentiation. *Immunity* (2016) 44(2):303–15. doi:10.1016/j.immuni.2016.01.014

42. Liu RX, Wei Y, Zeng QH, Chan KW, Xiao Z, Zhao XY, et al. Chemokine (C-X-C motif) receptor 3-positive B cells link interleukin-17 inflammation to protumorigenic macrophage polarization in human hepatocellular carcinoma. *Hepatology* (2013) 56(6):1729–39. doi:10.1002/hep.26020

43. Bogdanova K, Panov D, Daskalova M, et al. Translational Significance for Tumor Metastasis of Tumor-Associated Macrophages and Epithelial-Mesenchymal Transition. *Front Immunol* (2017) 8:1106. doi:10.3389/fimmu.2017.01106

44. Chaffer CL, San Juan BP, Lim E, Weinberg RA. EMT, cell plasticity and metastasis. *Cancer Metastasis Rev* (2016) 35(4):645–54. doi:10.1007/s10555-016-9648-7

45. Wu JY, Huang TW, Hsieh YT, Wang YP, Zhu LY, Zhaung SM, et al. Glycotoxic activation of peritoneal monocytes fosters immune privilege via the PFKFB3-PD-L1 axis in human hepatocellular carcinoma. *J Hepatol* (2019) 71(2):333–43. doi:10.1016/j.jhep.2019.04.007

46. Hao J, Yan F, Zhang Y, Triplett A, Zhang Y, Schultz DA, et al. Expression of Adipocyte/Macrophage Fatty Acid-Binding Protein in Tumor-Associated Macrophages Promotes Breast Cancer Progression. *Cancer Res* (2018) 78(9):2343–55. doi:10.1158/0008-5472.CAN-17-2465

47. Palmeiri K, Meng A, Martinez-Perez R, Quinto A, Riera-Domingo C, De Tullio G, et al. Pharmacologic or Genetic Targeting of Glutaminase Skews Macrophages toward an M1-like Phenotype and Inhibits Tumor Metastasis. *Cell Rep* (2017) 20(7):1654–66. doi:10.1016/j.celrep.2017.07.054

48. Vitale I, Manic G, Coussens LM, Kroemer G, Galluzzi L. Macrophages and Metabolism in the Tumor Microenvironment. *Cell Metab* (2019) 30(1):36–50. doi:10.1016/j.cmet.2019.06.001

49. Chen P, Zhu H, Xiong H, Kolar MJ, Chu Q, Saghatelian A, et al. Gpr132 sensing of lactate mediates macrophage-microglia interplay to promote breast cancer metastasis. *Proc Natl Acad Sci U S A* (2017) 114(3):580–5. doi:10.1073/pnas.1614351114

50. Roghanian A, Fraser C, Kleyman M, Chen J, B Cells Promote Pancreatic Tumorigenesis. *Cancer Discov* (2016) 6(3):230–2. doi:10.1158/2159-8290.CD-16-0100

51. Sousa CM, Bancur D, Wang X, Halbrook C, Stromer MH, Zhang L, et al. Pancreatic stellate cells support tumor metabolism through autophagic alamin secretion. *Nature* (2016) 536(7617):479–83. doi:10.1038/nature19084

52. Cunha LD, Yang M, Carter R, Guy C, Harris L, Crawford JC, et al. LC3-Associated Phagocytosis in Myeloid Cells Promotes Tumor Immune Tolerance. *Cell* (2018) 175(2):429–41.e16. doi:10.1016/j.cell.2018.08.061

53. Ohashi K, Wang Z, Yang YM, Billet S, Tu W, Pimienta M, et al. NOD-like receptor C4 Inflammasome Regulates the Growth of Colon Cancer Liver Metastasis in NF-κB. *Hepatology (Baltimore Md)* (2019) 70(5):1582–99. doi:10.1002/hep.30693

54. Conojo-Garcia JR, Rodriguez-PC. c-Maf: a bad influence in the education of macrophages. *J Clin Investig* (2020) 130(4):1629–31. doi:10.1172/JCI135444

55. Yang H, Zhang Q, Xu M, Wang L, Chen X, Feng Y, et al. CCL2-CCL2 axis recruits tumor associated macrophages to induce immune evasion through PD-1 signaling in esophageal carcinogenesis. *Mol Cancer* (2020) 19(1):41. doi:10.1186/s12935-020-01165-x

56. Kumar V, Donbrziedly R, Marvel D, Condamine T, Wang F, Lavilla-Alonso S, et al. Cancer-Associated Fibroblasts Neutralize the Anti-tumor Effect of CSF1 Receptor Blockade by Inducing PMN-MDSC Infiltration of Tumors. *Cancer Cell* (2017) 32(3):654–68.e5. doi:10.1016/j.ccell.2017.10.005
66. Neubert NJ, Schmittnaegel M, Bordry N, Nassiri S, Wald N, Martignier C, et al. T cell-induced CSF1 promotes melanoma resistance to PD1 blockade. Sci Trans Med (2018) 10(436):eaa3311. doi: 10.1126/scitranslmed.aan3311

67. Hutter G, Theruvath J, Graef CM, Zhang M, Schoen MK, Manz EM, et al. Microglia are effector cells of CD47-SIRPα antiphagocytic axis disruption against glioblastoma. Proc Natl Acad Sci U S A (2019) 116(3):997–1006. doi: 10.1073/pnas.1721434116

68. Hudson LE, Allen RL. Leukocyte Ig-Like Receptors - A Model for MHC Class I Disease Associations. Front Immunol (2016) 7:281. doi: 10.3389/fimmu.2016.00281

69. Barkal AA, Weiskopf K, Kao KS, Gordon SR, Rosental B, Yiu YY, et al. Engagement of MHC class I by the inhibitory receptor LILRB1 suppresses macrophages and is a target of cancer immunotherapy. Nat Immunol (2018) 19(1):76–84. doi: 10.1038/s41590-017-0004-z

70. Wu H, Han Y, Rodriguez Sillé Y, Deng H, Siddiqui S, Treese C, et al. Lipid droplet-dependent fatty acid metabolism controls the immune suppressive phenotype of tumor-associated macrophages. EMBO Mol Med (2019) 11(11): e10698. doi: 10.15252/emmm.201910698

71. Jaynes JM, Sable R, Ronzetti M, Bautista W, Knots Z, Abisoye-Ogunniyan A, et al. Mannose receptor (CD206) activation in tumor-associated macrophages enhances adaptive and innate antitumor immune responses. Sci Trans Med (2020) 12(530):eaa6337. doi: 10.1126/scitranslmed.aax6337

72. Wang W, Marinis JM, Beal AM, Savadkar S, Wu Y, Khan M, et al. RIP1 Kinase Drives Macrophage-Mediated Adaptive Immune Tolerance in Pancreatic Cancer. Cancer Cell (2018) 34(5):757–74.e7. doi: 10.1016/j.ccell.2018.10.006

73. Zhou Y, Fei M, Zhang G, Liang WC, Lin W, Wu Y, et al. Blockade of the Phagocytic Receptor MerTK on Tumor-Associated Macrophages Enhances P2X7R-Dependent STING Activation by Tumor-Derived cGAMP. Immunity (2020) 52(2):357–73.e9. doi: 10.1016/j.immuni.2020.01.014

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Pan, Yu, Wang and Zhang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.