RESEARCH ARTICLE

HYPERHOMOCYSTINEMIA IN EPILEPTIC PATIENTS: THE ROLE OF THFR POLYMORPHISM AND ANTI-EPILEPTIC DRUGS.

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Manuscript Info

Abstract

Hyperhomocystinemia despite can be seen in small percentage of normal population but associated with increased vascular diseases in epileptics. Hyperhomocystinemia in epileptic patients was supposed to develop primarily from a complex interplay between the chronic intake of enzyme inducing AEDs and genetically determined mutations of MTHFR. Ninety two patients with epilepsy between the ages of 18 and 40 years consented to participate in the study in order to study analyses the effect of a C677T polymorphism of MTHFR and AEDs on tHcy levels in patients with epilepsy treated for at least 6 months with monotherapy including the traditional and newer AEDs.

Introduction:

Homocysteine (Hcy) which is a sulfur-amino acid, considered as an intermediate product of methionine metabolism and is normally at plasma total (t) concentrations <15 umol/L. Its higher values more than 15 umol/L i.e. hyperhomocysteinemia are considered abnormal and this can be seen in about 5% of the general population. Higher percentages, from 10–20%, are observed in epileptic patients Hcy(Caccamo et al., 2004; Belcastro et al., 2007).

Hyperhomocystinemia is associated with increased vascular disease risk in epileptics (Williams and Schalinske, 2010). Furthermore, it was shown to enhance seizure activity in those patients (Balderelli et al., 2010).

There are conflicting data about the role of methylene tetrahydro folate reductase (MTHFR) polymorphisms (particularly the C677T and A1298C) as determining factor of high total homocysteine plasma (tHcy) levels in epileptic patients (Ono et al., 2002). MTHFR is a key enzyme in the production of 5-methyltetrahydrofolate (a major circulating form of folate in human), which is the primary methyl donor for Hcyremethylation to methionine. The C677T mutation of the MTHFR gene decreases the activity of this enzyme leading to decreased folate levels in serum and subsequent accumulation of homocysteine(Jacques et al.,1996).

A relationship between increased tHcy levels and antiepileptic drugs (AEDS) administration, has also been recognized in the last decade (Sener et al.,2006; Kurul et al.,2007). Antiepileptic drugs, in fact, deplete folate and other B vitamins (necessary cofactors in the metabolic pathways of Hcy). So, hyperhomocystinemia in epileptic patients was supposed to develop primarily from a complex interplay between the chronic intake of enzyme inducing AEDs and genetically determined mutations of MTHFR (Caccamo et al., 2004; Belcastro et al., 2007).

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The present study analyses the effect of a C677T polymorphism of MTHFR and AEDs on tHcy levels in patients with epilepsy treated for at least 6 months with monotherapy including the traditional and newer AEDs.

**Subjects And Method:-**

Subjects were consecutively recruited from epileptic patients under treatment by AEDS monotherapy (including new and traditional) at Zagazig university hospitals and out-patient clinic, from August 2016 through Feb 2017. Ninety two patients with epilepsy between the ages of 18 and 40 years consented to participate in the study. We excluded those with neurometabolic or systemic diseases such as thyroid or renal dysfunction, chronic therapy with drugs other than AEDs and current or previous treatment with folic acid and/or other vitamins. Compliance and adherence to therapy were assessed using Morisky medication adherence scale.

Ninety two healthy control who had no clinical and/or laboratory evidence of metabolic and/or endocrine diseases were included. Controls were recruited from outpatients of the Neurology Department, Zagazig University. These subjects had been referred for non-epileptic problems e.g. episodic headache, minor head trauma, and dizziness. Exclusion criteria were identical to those of the patient group. Multivitamin users were also excluded from the study.

All patients and controls underwent the following examinations: complete blood count, liver and kidney function tests and serum levels of folic acid and tHcy. DNA was extracted from peripheral blood using the QIAamp DNA Blood Mini Kit (Qiagen, Hilden, Germany) following the manufacturer’s recommendations and then stored at −20°C.

After blood samples were obtained, plasma was immediately separated by centrifugation at 1000g for 15 min. Blood samples were immediately processed in order to prevent artefactual variations of tHcy, due to the products of in vitro erythrocyte metabolism. Samples were stored at −20°C until analysis. Plasma total levels of Homocysteine and folate were assessed by use of commercially available kit for Human Homocysteine ELISA (MyBioSource, Inc., San Diego, USA), Human Folic Acid ELISA Kit (MyBioSource, Inc., San Diego, USA) respectively. Plasma tHcy concentrations were considered elevated when they exceeded 15 umol/L, and <4 ng/mL folate. (Myers et al., 2009 ; Benoist, 2008).

Genotyping for C677T MTHFR polymorphisms was carried out on DNA extracted from white cells of freshly drawn peripheral blood samples. In each patient, isolation of DNA was performed by using the QIAamp DNA Blood Mini Kit (Qiagen, Hilden, Germany) following the manufacturer’s recommendations.

The MTHFR C677T (rs1801133) mutation was detected using the ABI StepOne TM Real Time PCR System (Applied Biosystems) and analyzed using Step one software v2.3 (Applied Biosystems), using the TaqMan assay. The final volume for each reaction was 20 μL, containing 10 μLTaqMan Genotyping Master Mix, 1.0 μLTaqMan probes mix, 20 ng genomic DNA and reaching the final volume with RNase-free water. PCR temperature profile consisting of denaturation at 95°C for 10min followed by 40 cycles of denaturation at 95°C for 15seconds and then annealing and elongation at 65°C for 1minute.

**Statistical Analysis:-**

Comparison of discrete variables (sex, MTHFR C677T genotype distribution) was made using the chi square test. Student’s t-test for unpaired data was used to compare tHcy concentrations of patients and controls while folate levels were investigated using the Mann-Whitney, ANOVA F-test was used to compare tHcy levels among the three MTHFR genotypes in patients and controls. Kruskal-Wallis test was implemented to explore the variation of plasma folate levels across the three MTHFR genotypes in patients and controls. Multiple logistic regression analysis was used to analyze factors of influence on tHcy concentrations, odds ratio was calculated for each variable. SPSS (Statistical Package for Social Science) version 18 was used for statistical analysis (SPSS, Chicago). p < 0.05 values were considered significant.

**Results:-**

In table 1 we summarized demographic and clinical characteristics of the study subjects. A total of 92 epileptic patients (53 male, 39 female) with mean age (30.5 ± 8.5 years) were included in the study; in addition to 92 healthy age and sex matched controls. All patients were on monotherapy, the AEDs used in the study group were as follows:phenytoin (n = 11), carbamazepine (n = 17), valproic acid (n = 20), topiramate (n =10), oxcarbazepin (n = 14), lamotrigin (n = 10) and levetiracetam (n=10). Mean tHcy values were significantly higher in patients than in
controls (17.3 ± 3.2 vs 6.9 ± 2.2 ; p < 0.001), while folic acid concentrations were significantly lower in patients than those of controls (7.3 ± 2.5 vs 10.0 ± 4.2 ; p<0.001) and these differences remained significant even if patients and controls were grouped according to the MTHFR genotypes. Our patients showed the following distribution of MTHFR polymorphism: CT (41.3 %), CC (29.3 %), and TT (21.7%). Of epileptic patients involved in this study, carriers of the TT677 MTHFR genotype had a significant increase in tHcy ( F = 280 , p < 0.0001) and significant decrease in folate levels (KW = 3.23 , P= 0.04) ( Table 2). Multiple logistic regression data analysis showed a significant negative correlation between folic acid serum levels and tHcy levels (p < 0.001); in addition, TT677 MTHFR genotype and type of AEDs therapy, proved to be a significant risk factor for high tHcy levels in epileptic patients (p < 0.001) (Table 3). 

A significant difference in plasma tHcy values was also seen across patients stratified on the basis of pharmacogenetics. Hyperhomocysteinemia was observed in patients on carbamazepine (CBZ), phenobarbital (PB), topiramate (TPM), and oxcarbazepine (OXC) therapy, especially in TT677 MTHFR genotype. Plasma tHcy levels in patients taking LTG and LEV therapy were significantly lower than those observed in the other drug groups and not significantly different from those observed in the controls and in the VPA group (figure1).

Table 1: Clinical and Biochemical Characteristics of Epileptic Patients Receiving AEDs and Healthy Controls.

| Variable                      | Epileptic Patients (n = 92) | Controls (n = 92) | Test | p value | OR |
|-------------------------------|-----------------------------|------------------|------|---------|----|
| **Demographic data:**         |                             |                  |      |         |    |
| Sex ratio (male/female)       | 53/39                       | 51/41            | χ²=0.09 | 0.77 NS | ---- |
| Age (yr)                      | 30.5 ± 8.5                  | 32.1 ± 6.8       | t=1.41 | 0.16 NS | ---- |
| **Clinical data:**            |                             |                  |      |         |    |
| Duration of epilepsy (yr)     | 8.4 ± 4.10                  | NA               | --   | ---     | ---- |
| Seizure types                 |                             |                  |      |         |    |
| Primary generalized           | (n) 66                      | NA               | ---  | -----   | ---- |
| Temporal focal                | (n) 11                      | NA               | ---  | -----   | ---- |
| Extratemporal focal           | (n) 15                      | NA               | ---  | -----   | ---- |
| Duration of therapy           | 6.5 ± 2.4                   | 6.9 ± 2.2        | t=25.69 | <0.001** | ---- |
| tHcy (µmol/L)                 | 17.3 ± 3.2                  | 12.9 ± 2.3       | MW=5.9 | <0.001** | ---- |
| Folate (ng/mL)                | 7.3 ± 2.5                   | 10.0 ± 4.2       |      |         |    |
| **Genetic data:**             |                             |                  |      |         |    |
| CC677 MTHFR genotype          | 27 (29.3%)                  | 34 (37%)         | χ²=1.2 | 0.27 NS | 1.41(0.76-2.62) |
| CT677 MTHFR genotype          | 32 (34.8%)                  | 38 (41.3%)       | χ²=0.83 | 0.36 NS | 1.32(0.73-2.40) |
| TT677 MTHFR genotype          | 33 (35.9%)                  | 20 (21.7%)       | χ²=4.48 | 0.03*  | 2.01(1.05-3.87) |
| t: Independent sample t test, χ²: Chi square test, MW: Mann Whitney test, NA: not applicable

Table 2: Levels of tHcy and Plasma Folate in Epileptic Patients receiving AEDs and Healthy Controls According to MTHFR Genotype:

| Genotype          | Epileptic Patients (n = 92) | Healthy Controls (n = 92) | Test     | p value | OR  |
|-------------------|-----------------------------|---------------------------|----------|---------|-----|
| CC677 MTHFR tHcy (µmol/L) |                             |                           | t        | 9.63   |      |
|                   | (n = 27)                    | (n = 34)                  | 16.5 ± 3.3 a |         |      |
|                   |                             |                           | 9.7 ± 2.2 c |         |      |
| CT677 MTHFR tHcy (µmol/L)  |                             |                           | t        | 4.61   |     |
|                   | (n = 32)                    | (n = 38)                  | 16.0 ± 3.3 a |         |      |
|                   |                             |                           | 12.9 ± 2.3 c |         |      |
| TT677 MTHFR tHcy (µmol/L)  |                             |                           | t        | 18.89  |     |
|                   | (n = 33)                    | (n = 21)                  | 33.8 ± 3.6 b |         |      |
|                   |                             |                           | 15.6 ± 3.2 d |         |      |
| F                 |                             |                           |          | <0.001** |     |
| P                 |                             |                           |          | <0.001** |     |
| CC677 MTHFR Folate (ng/mL) |                             |                           | MW       | 0.40   | 0.70 NS |
|                   | (n = 27)                    | (n = 34)                  | 8.5 ± 5.1 a |         |      |
|                   |                             |                           | 9.0 ± 4.9 c |         |      |
| CT677 MTHFR Folate (ng/mL) |                             |                           | MW       | 0.47   | 0.46 NS |
|                   | (n = 32)                    | (n = 38)                  | 8.6 ± 3.9 a |         |      |
|                   |                             |                           | 9.3±4.0 c  |         |      |
| TT677 MTHFR Folate (ng/mL) |                             |                           | MW       | 3.16   | 0.003** |
|                   | (n = 33)                    | (n = 20)                  | 6.4 ± 4.9 b |         |      |
|                   |                             |                           | 10.1 ± 2.3 c |         |      |
Table 3: Multiple regression analysis showing the association of AEDs, TT677 genotype, vitamin status, and demographic variables with plasma tHcy

|                  | Beta | B   | SE  | OR  | p-value |
|------------------|------|-----|-----|-----|---------|
| TT677 MTHFR      | 0.34 | 0.31| 0.03| 9.8 | <0.001**|
| Folate           | 0.45 | 0.09| 0.01| 11.1| <0.001**|
| Age              | 0.02 | 0.00| 0.03| 0.60| 0.50    |
| Sex              | 0.03 | 0.02| 0.03| 0.8 | 0.41    |
| Therapy          |      |     |     |     |         |
| OXC              | 0.10 | 0.12| 0.05| 2.3 | 0.002** |
| TPM              | 0.20 | 0.25| 0.06| 4.1 | <0.001**|
| LEV              | 0.08 | 0.1 | 0.05| 1.7 | 0.08    |
| LTG              | 0.04 | 0.05| 0.4 | 0.06| 0.80    |
| CBZ              | 0.29 | 0.34| 0.05| 6.9 | <0.001**|
| PHT              | 0.41 | 0.21| 0.04| 5.43| <0.001**|
| VPA              | 0.65 | 0.22| 1.90| 0.42| 0.07    |
| Therapy duration | 1.05 | 0.29| 3.79| 0.07| 0.95    |

SE: standard error

Fig 1: Mean tHcy levels in epileptic patients receiving different AEDS (traditional and new) and the control group according to MTHFR genotype.

Discussion: The interest of the medical community in the amino acid Hcy stems from the observation that elevated plasma Hcy has been associated with cardiovascular disease in multiple large-scale epidemiologic studies and, consequently, it is considered as an independent risk factor for atherosclerosis (Humphrey et al., 2008). An additional aspect to be considered, and strictly associated to hyper-tHcy, is the AED-induced folate deficiency, a condition that itself may have different clinical implications (Reynolds, 2006; McNulty & Scott, 2008).
We observed a higher mean tHcy and higher prevalence of hyperhomocysteinemia in epileptic patients receiving AEDs than in the controls. The present data confirm previously published data by (Verrotti et al., 2000; Caccamo et al., 2004; Belcastro et al., 2007; Tan et al., 2009) who showed that epileptic patients on chronic AED therapy are more prone than the general population to develop hyper-tHcy and low folate levels.

An elevated plasma level of homocysteine can be induced by either genetic or environmental factors. Because MTHFR 677 C --* T mutation is known to be associated with reduced enzyme activity and AEDs decrease MTHFR enzyme activity, we examined whether the combined presence of MTHFR 677 C --* T mutation and AEDs intake in epileptic patients causes elevated tHcy. In the population studied, the frequency of homozygosity for MTHFR 677 TT mutation in epileptic patients was higher than that in the controls. In epileptic patients receiving AEDs, hyperhomocysteinemia was more frequent in the MTHFR TT genotype versus CT or CC genotypes in agreement with Yoo et al. (1999) Ashfield et al. (2002) and Belcastro et al. (2010). This can be explained by the fact that individuals with the 677TT genotype, have approximately 30% the MTHFR enzyme activity of those with the 677CC genotype as reported by Rozen, (1997) and Rielly et al. (2014). They also reported that low folate affects individuals with the 677TT genotype to a greater extent than those with the 677CC/CT genotypes and these results were parallel to ours. So, 677TT individuals with lower plasma folate levels are at higher risk for elevated plasma homocysteine levels than those with 677CC/CT genotype. In this context, patients carrying the 677TT genotype had a more pronounced drop in tHcy and a bigger increase in serum folate concentrations than those with the 677CC or 677CT genotype after switching from a low-folate to a high-folate diet (Silaste et al., 2001) or after supplementation with folic acid (Foehr et al., 2002).

In our patients with MTHFR TT homozygotes had a higher tHcy and lower plasma folate in the phenytoin and carbamazepine group, but not in the valproic acid group and that correlates with the findings of Yoo et al., 1999. In keeping with other studies (Ono et al., 2002; Vurucu et al., 2008) duration of therapy has no clear effect on tHcy levels.

The new finding obtained from this study was that TPM and OXC among of the recent AEDs can cause hyper-tHcy as the old ones, whereas lamotrigine (LTG) and levetiracetam (LEV) are devoid of this effect. In this context, our data might have practical implications, in the case of well-controlled patients on those medications, a supplement of folate could be prescribed to normalize plasma tHcy levels which also match with previous recommendations of Huemer et al., 2005; Reynolds, 2006; Belcastro et al., 2007.

With respect to genetic variation, a common MTHFR 677 C --* T mutation is a determinant of hyperhomocysteinemia in epileptic patients receiving anticonvulsants, suggesting that a gene-drug interaction induces hyperhomocysteinemia. Since hyper-tHcy is a modifiable risk factor for vascular disease and a sensitive marker for folate status, we suggest that epileptic patients receiving anticonvulsants should be monitored for tHcy levels.

Epileptic patients receiving AEDs may have a higher folate requirement to maintain a normal tHcy level, especially homozygotes for the MTHFR 677 TT mutation.

In the view of the previous findings, we recommend if patients are not satisfactorily controlled by PTH, CBZ, OXC and or TPM, a switch to another drug, like LTG or LEV among the new AEDs, or VPA among the older ones, could be taken into consideration, as indicated by the present data and those of other studies (Gidal et al., 2005; Mintzer et al., 2009). But if our patients are controlled and adherent to the medications which incriminated in hyperhomocysteinemia, add on folate supplementations would be helpful in normalization of its levels.

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