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Laser irradiated phenothiazines: New potential treatment for COVID-19 explored by molecular docking

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ABSTRACT

The worldwide infection with the new Severe Acute Respiratory Syndrome coronavirus 2 (SARS-CoV-2) demands urgently new potent treatment(s). In this study we predict, using molecular docking, the binding affinity of 15 phenothiazines (antihistaminic and antipsychotic drugs) when interacting with the main protease (M\textsuperscript{pro}) of SARS-CoV-2. Additionally, we tested the binding affinity of photoproducts identified after irradiation of phenothiazines with Nd:YAG laser beam at 266 nm respectively 355 nm. Our results reveal that thioridazine and its identified photoproducts (mesoridazine and sulforidazine) have high biological activity on the virus M\textsuperscript{pro}. This shows that thioridazine and its two photoproducts might represent new potent medicines to be used for treatment in this outbreak. Such results recommend these medicines for further tests on cell cultures infected with SARS-CoV-2 or animal model. The transition to human subjects of the suggested treatment will be smooth due to the fact that the drugs are already available on the market.

1. Introduction

A virus is an infectious entity that needs, for reproducing, a host cell in a living organism. Viruses constitute a ubiquitous biological entity on our planet [1]. Coronavirus disease 2019 (COVID-19) is caused by the new SARS-CoV-2 [2]. WHO report published on 4th of June 2020 (Situation Report-136) shows that there are 3,416,828 confirmed cases worldwide [3]. Since WHO-81 report from 10 April, the number of cases rise accelerated from 92,798 deaths to 157,847 on 20 April [3,4]. The massive number of persons infected by SARS-CoV-2 demands new, more potent treatments.

Drug repositioning is a viable approach instead of developing new drugs in the context of new infectious diseases; this leads to compounds that are de-risked and have lower development costs [5]. In silico studies are a good alternative in an emergent situation to repose a chemical compound [6]. Molecular docking approach is a fast, low-cost procedure used to predict at molecular level the interaction between a ligand and a target [7].

Phenothiazines are chemicals with high antipathogen proprieties against pathogens such as: (i) Protozoa \textit{Plasmodium falciparum} (protozoan responsible for malaria) [8], genus Leishmania (that cause Leishmaniasis) [9], (ii) several resistant bacterial strains (Multidrug-resistant \textit{Mycobacterium tuberculosis} and Methicillin-resistant \textit{Staphylococcus aureus}) and (iii) viruses such as those responsible for hepatitis B, HIV, TBEV, arenavirus, herpesvirus etc. [9]. Moreover, in previous epidemies with SARS-CoV-1 and MERS viruses, chlorpromazine (CPZ) shows high inhibitory activity and was recommended as a candidate for testes as a broad-spectrum antiviral [10]. Here, our aim is to propose the antiviral role of phenothiazines against COVID-19.

Phenothiazines like Thioridazine (TZ) and CPZ generate, by exposure to laser radiation, photoproducts that are more efficient in inhibition of bacterial strains than non-irradiated drugs [11]. To find the mechanism of action of TZ after irradiation, we have identified the generated photoproducts. In a previous study, TZ was irradiated up to 11 min with 355 nm Nd:YAG pulsed laser beam which has full-time width at half maximum 5 ns and the average pulse energy on the sample 30 mJ at a repetition rate of 10 pps. Using chromatographic methods (HPLC-MS) we have identified two TZ photoproducts.
### Table 1
The 2D/3D chemical structure of compounds from Phenothiazines class and TZ and CPZ photoproducts that resulted during laser irradiation; 2D chemical structure, lowest EFEB (kcal/mol) for each compound resulted after 100 runs using molecular docking simulation, predicted KI (nM) and pKI values are also presented. PZ – the acronym of promazine.

| Compound | 2D chemical structure | 3D chemical structure | Lowest EFEB kcal/mol | K<sub>I</sub> nM | pK<sub>I</sub> |
|----------|-----------------------|-----------------------|----------------------|----------------|------------|
| TZ       | ![TZ 2D chemical structure](image1) | ![TZ 3D chemical structure](image2) | -9.0 | 244 | 6.6 |
| SPZ      | ![SPZ 2D chemical structure](image3) | ![SPZ 3D chemical structure](image4) | -10.1 | 37 | 7.4 |
| MSO      | ![MSO 2D chemical structure](image5) | ![MSO 3D chemical structure](image6) | -9.4 | 121 | 6.9 |
| CPZ      | ![CPZ 2D chemical structure](image7) | ![CPZ 3D chemical structure](image8) | -8.3 | 785 | 6.1 |

(continued on next page)
Table 1 (continued)

| Compound | Structure | pKa | Log D | Log P |
|----------|-----------|-----|-------|-------|
| CPZ-SO   | ![Structure CPZ-SO](image1.png) | -7.3 | 4310  | 5.3   |
| 2-HO-PZ-SO | ![Structure 2-HO-PZ-SO](image2.png) | -7.1 | 5830  | 5.2   |
| 2-HO-PZ  | ![Structure 2-HO-PZ](image3.png) | -7.8 | 1630  | 5.7   |
| PZ       | ![Structure PZ](image4.png)      | -6.9 | 8240  | 5.0   |

(continued on next page)
| Compound          | pKa  | IC50  | nH | 
|-------------------|------|-------|----|
| PZ-SO             | -7.1 | 6130  | 5.2|
| P1                | -7.3 | 4480  | 5.3|
| P2                | -6.9 | 8020  | 5.0|
| prochlorperazine  | -8.4 | 595   | 6.2|
| fluphenazine      | -8.9 | 274   | 6.5|

(continued on next page)
Table 1 (continued)

| Compound          | Structure 1 | Structure 2 | 
|-------------------|-------------|-------------|
| levomepromazine   | ![levomepromazine](image1.png) | ![levomepromazine](image2.png) | -7.1 5540 5.2 |
| trifluoperazine   | ![trifluoperazine](image3.png) | ![trifluoperazine](image4.png) | -8.2 890 6.0 |
| alimemazine       | ![alimemazine](image5.png) | ![alimemazine](image6.png) | -7.2 4960 5.3 |
| hydroxyethylprometazine | ![hydroxyethylprometazine](image7.png) | ![hydroxyethylprometazine](image8.png) | -7.5 2990 5.5 |
| isothipendyl      | ![isothipendyl](image9.png) | ![isothipendyl](image10.png) | -6.7 1098 4.9 |

(continued on next page)
| Compound     | Structure 1 | Structure 2 | Log P | Octanol/Water | Octanol/Hexane |
|--------------|-------------|-------------|-------|---------------|---------------|
| Mequitazine  | ![Structure](image1) | ![Structure](image2) | -8.3  | 764           | 6.1           |
| Methdilazine | ![Structure](image3) | ![Structure](image4) | -7.5  | 2970          | 5.5           |
| Perphenazine | ![Structure](image5) | ![Structure](image6) | -9.0  | 227           | 6.6           |
| Promethazine | ![Structure](image7) | ![Structure](image8) | -6.8  | 9120          | 5.0           |
| Thiazinam    | ![Structure](image9) | ![Structure](image10) | -6.6  | 1324          | 4.8           |

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mesoridazine (MSO) and sulphoridazine (SPZ), those photoproducts are as well TZ metabolites. Molecular structure modifications occur through preferential oxidation in (-SCH3) radical zone [12].

CPZ photoproducts were obtained after irradiation with an Nd:YAG pulsed laser beam at 266 nm with an average pulse energy of 6.5 mJ. The CPZ samples at 2 mg/mL in ultrapure water irradiated 1, 5, 15, 30, 60, 120,180 and 240 min were analysed using LC-TOF/MS. Results showed seven CPZ generated photoproducts: promazine (PZ), P1, P2, 2-hydroxypromazinesulfoxide (2-HO-PZ-SO), chlorpromazine sulfoxide (CPZ-SO), promazine sulfoxide (PZ-SO) and 2-hidroxipromazine (2-HO-PZ) [13]. We mention that the photoproducts MSO and SPZ may also be used as antipsychotics.

The replications of coronaviruses are blocked by inhibiting the protease of the virus [14]. For identifying a treatment of COVID-19 disease, we used molecular docking procedure to predict the inhibitory activity (against SARS-CoV-2) Mpro of some compounds from phenothiazine drug class. This study also predicts the binding affinity of TZ and CPZ known photoproducts generated by laser irradiation with the Mpro.

2. Materials and Methods

2.1. Identify the Viral Protease

To simulate the interaction between SARS-CoV-2 and drugs from the class of phenothiazines including TZ and CPZ photoproducts we have used the virus Mpro from RCSB Protein Data Bank: PDB code 6LU7 [15]. The structure of the enzyme was obtained using X-ray diffraction at a resolution of 2.16 Å [15]. We have chosen the Mpro of the virus due to the essential role of this enzyme in proteolytic maturation of the nonstructural proteins. This enzyme represents a potential target in the development of antiviral drugs [16].

2.2. Molecular Docking Protocol

We have prepared the molecule for docking studies by deleting water, adding hydrogen, merging the non-polar hydrogen and adding Gasteiger partial charges [17]. The small molecules were drawn and optimized geometrically using Discovery studio visualizer. After optimization, we have saved the molecules in .mol format [18]. After geometrical optimization, we have used OpenBabel software [19] to shape the molecule in .pdbqt format. For molecular docking approach, we have used Autodock 4.2.6 software [20].

Grid-box was selected to contain only the protease situs identified by amino acids: His 41, Met 49, Phe 140, Asn142 Gly 143, Cys 145, His 163, His 164, Glu 166 and His 172 [21,22]. Covalent grid parameters had an energy barrier height of 1000 and the half-width of 5.00 Å. Total grid Pts per map is 257,725. The grid has the number of points in dimensions: x-60; y-64 and z-64 and the cartesian coordinates of central grid point of maps were set to x = −11.86, y = 17.38, z = 68.99. The spacing of grid points was set to 0.375 Å. For docking simulation, we generated 100 conformations for each ligand; the used search parameter to identify the binding conformation of the ligands was Genetic Algorithm and the output was saved as Lamarckian.

3. Results and Discussions

Generally, a molecular docking result is represented by Estimated Free Energy of Binding (EFEB). Higher biological activity of a drug is correlated with lower free energy of binding [23–25]. Here, EFEB was calculated as EFEB = (1) + (2) + (3)–(4) using Autodock 4.2.6 software where:

(1) is Final Intermolecular Energy vdW + H-bond + desolv Energy Electrostatic Energy.
(2) represents Final Total Internal Energy.
(3) is Torsional Free Energy.
(4) represents unbound System’s Energy [=(2)].

Furthermore, the estimated inhibition constant $K_i$ was predicted by Autodock 4.2.6 at 298.15 K; a lower $K_i$ is correlated with high biological activity (p$K_i$) [26]. For improved data analysis, we converted $K_i$ values (nM) in p$K_i$ by applying the logarithm function: $pK_i = \log (1/K_i)$ (M)). A p$K_i$ value of 4 or lower will define a compound with no
biological activity.

TZ presents an estimated Inhibition Constant (Ki) of 244.97 nM (Table 1), this energy being lower than EFEB of remdesivir (~7.80 kcal/mol) which is predicted with autodockVina [22].

We have compared our results with remdesivir antiviral drug due to improvements showed in patients infected with SARS-CoV-2 [27]; moreover, remdesivir interacts with the enzyme in the main situs of interaction [22], same as MSO (Fig. 1).

Photoproducts resulted after laser irradiation of TZ present higher biological activities than TZ (Table 1).

From Table 1 it results that SPZ presents the highest biological activity on SARS-CoV-2 Mpro (pKi = 7.42) and it is in close contact at a VDW scaling factor closer than 1. Dotted green line underlines an h-bond interaction between SPZ and His163. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

The highest EFEB from 100 runs was ~8.48 kcal/mol and the lowest ~10.12 kcal/mol.

MSO is the photoproduct with the second-highest biological activity (Table 1). It is similar with SPZ and interacts with the Mpro of the virus in the same binding situs and has hydrogen bond interaction with amino acid residue of His 163 (Fig. 2). SPZ highest binding energy is ~8.72 kcal/mol and the lowest ~9.43 kcal/mol (Table 1); it forms 9 distinct conformational clusters, and the number of multi-member conformational clusters found is 7.

TZ highest binding energy is ~7.06 kcal/mol and the lowest is ~9.02 kcal/mol (Table 1). The number of distinct conformational clusters is 8 and the number of multi-member conformational clusters that were found is 3. The cluster with the largest number of population (46) had a mean binding energy of ~8.22 kcal/mol. Perphenazine antipsychotic drug has the lowest binding energy (~9.06 kcal/mol) similar with TZ (Table 1).

CPZ (pKi 6.10) and fluphenazine (pKi 6.56) exhibit a medium biological activity against SARS-CoV-2 Mpro (Table 1). CPZ identified photoproducts had a lower biological activity than CPZ. In Table 1 are presented CPZ, fluphenazine and CPZ photoproducts biological activities where other results for tested phenothiazines show medium-low biological activity and are also presented. Compounds that have no biological activity or low biological activities are: (i) the antihistaminic drugs promethazine, thiazinam and isothipendyl; (ii) CPZ photoproducts PZ and P2 compound.

Table 2

| Cluster Rank | Lowest Binding Energy kcal/mol | Run | Mean Binding Energy kcal/mol | Number in Cluster |
|--------------|--------------------------------|-----|-------------------------------|------------------|
| 1            | −10.12                         | 40  | −9.71                         | 43               |
| 2            | −9.80                          | 55  | −9.55                         | 29               |
| 3            | −9.70                          | 65  | −9.70                         | 1                |
| 4            | −9.54                          | 2   | −9.42                         | 5                |
| 5            | −9.54                          | 99  | −9.42                         | 7                |
| 6            | −9.27                          | 90  | −9.25                         | 9                |
| 7            | −8.63                          | 64  | −8.58                         | 6                |

4. Conclusions

Animal models infected with SARS-CoV-2 do not reproduce the same characteristic symptoms as in humans, not even in non-human primates. The animal model, that shows the clinical symptomatology of COVID-19 is represented by a transgenic mouse, that expresses human angiotensin-converting enzyme 2. The appearance of clinically observed symptomatology in the trans-genic mouse suggests that this receptor might be responsible for the access of the virus in the human cell [28].

Due to the lack of animal models, and the urgent need of treatment, molecular docking studies are suitable to speed up the process of finding compounds that present possible inhibitory activity.

Our results suggest that TZ and TZ photoproducts obtained by laser irradiation, have significant biological activity on SARS-CoV-2 Mpro and could be used in a potent treatment in COVID-19 disease.

According to our previous experience [29], irradiated drugs show better antibacterial activity than non-irradiated drugs and even than photoproducts tested separately. Moreover, the mixture of two obtained photoproducts has improved results compared to a single compound use.

Ergo, we recommend experimental validation of TZ, TZ photoproducts (MSO and SPZ) and irradiated TZ in interaction with SARS-CoV-2, due to the possible increased activity of a combination of such compounds. The use of the “cocktails” of irradiated TZ and its photoproducts might possibly show increased activity on SARS-CoV-2 targets.

Authorship Statement

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All persons who meet authorship criteria are listed as authors, and all authors certify that they have participated sufficiently in the work to take public responsibility for the content, including participation in the concept, design, analysis, writing, or revision of the manuscript. Furthermore, each author certifies that this material or similar material has not been and will not be submitted to or published in any other publication before its appearance in the Journal of Photochemistry & Photobiology, B: Biology.
Declaration of Competing Interest

The authors do not declare any conflict of interest.

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