pH-Dependent Degradation of Diclofenac by a Tunnel-Structured Manganese Oxide

Ching-Yao Hu 1, Yu-Jung Liu 2 and Wen-Hui Kuan 3,4,*

1 School of Public Health, Taipei Medical University, Taipei 110, Taiwan; cyhu@tmu.edu.tw
2 Graduate Institute of Environmental Engineering, National Taiwan University, Taipei 106, Taiwan; yujungliu77@gmail.com
3 Department of Safety, Health and Environmental Engineering, Ming Chi University of Technology, New Taipei 24301, Taiwan
4 Chronic Diseases and Health Promotion Research Center, Chang Gung University of Science and Technology, Chiayi 61363, Taiwan
* Correspondence: whkuan@mail.mcut.edu.tw; Tel.: +886-2-29089899 (ext. 4653); Fax: +886-29041914

Received: 25 June 2020; Accepted: 3 August 2020; Published: 5 August 2020

Abstract: The mechanism of diclofenac (DIC) degradation by tunnel-structured γ-MnO 2, with superior oxidative and catalytic abilities, was determined in terms of solution pH. High-performance liquid chromatography with mass spectroscopy (HPLC–MS) was used to identify intermediates and final products of DIC degradation. DIC can be efficiently oxidized by γ-MnO 2 in an acidic medium, and the removal rate decreased significantly under neutral and alkaline conditions. The developed model can successfully fit DIC degradation kinetics and demonstrates electron transfer control under acidic conditions and precursor complex formation control mechanism under neutral to alkaline conditions, in which the pH extent for two mechanisms exactly corresponds to the distribution percentage of ionized species of DIC. We also found surface reactive sites (S rxn), a key parameter in the kinetic model for mechanism determination, to be exactly a function of solution pH and MnO 2 dosage. The main products of oxidation with a highly active hydroxylation pathway on the tunnel-structured Mn-oxide are 5-iminoquinone DIC, hydroxyl-DIC, and 2,6-dichloro-N-o-tolylbenzenamine.

Keywords: diclofenac (DIC); pH-dependent degradation mechanism; reactive site; tunnel-structured manganese oxide; γ-MnO 2

1. Introduction

Diclofenac (DIC), one of the most commonly used nonsteroidal anti-inflammatory drugs (NSAIDs) worldwide, is discharged in large amounts from wastewater treatment plants because of its high hydrophilic nature [1] and low biodegradability [2]. Thus, DIC is widely found in the aquatic environment in a range from ng/L to µg/L and is one of the most frequently detected pharmaceutical and personal care products in water [3,4]. No evidence suggests that DIC is harmful to humans; however, it might be toxic to aquatic organisms and harmful to embryos, infants, children, and adults with low immunity and being sensitive to pharmaceuticals [5–8]. Most of the evidence was focused on its adverse effects on the aquatic and terrestrial organisms, which might cause ecological damage [9–11]. Besides, the transformation products of diclofenac might be more toxic than diclofenac [12,13], which needs to be investigated further.

Manganese oxides (Mn-oxides) are effective natural oxidants for organic pollutants including phenols [14], chlorophenol [14–17], aliphatic amines [18] and anilines [16,19] in soils and sediments. In past decades, Mn-oxides have been used to remove antibacterials and compounds with phenolic and fluoroquinolonic moieties [20,21], triazine, aromatic N-oxides [22], tetracyclines [23], and estrogenic...
compounds such as the synthetic hormone 17R-ethinylestradiol [24,25] and as an alternative treatment for wastewater or groundwater containing DIC because Mn-oxides are cheap and operated-friendly [26,27]. These compounds may be endocrine disruptors (EDS) and precursors of harmful disinfection byproducts such as haloacetonitriles, haloacetamides, and nitrogenous heterocyclic [28].

In general, Mn-oxides are classified into a layer phase and tunnel structure with edge-sharing and corner-sharing octahedral MnO$_6$, respectively. Studies have commonly used layer-structured birnessite because of its high sorption capacity for target pollutants to remove organic pollutants [15,20,24]. Recently, tunnel-structured Mn-oxides have received considerable attention because of their excellent catalytic oxidative capacity for organic pollutant degradation; however, their mechanisms and feasibility remain unclear and must be investigated [29,30]. γ-MnO$_2$, a tunnel-structured Mn-oxide, normally contains a combination of pyrolusite (1 $\times$ 1 tunnel) and ramsdellite (1 $\times$ 2 tunnel) and has been confirmed to be environmentally friendly without apparent cell toxicity [31,32]. To the best of the authors’ knowledge, this is the first study investigating the action of γ-MnO$_2$ on DIC degradation.

The ionized and acid form of a weak acid such as DIC significantly alters its adsorptive behavior between solution and solid surface, i.e., pollutants adsorption onto oxides from water and analytes separation in high-performance liquid chromatography (HPLC) column. The ionization of a weak acid such as DIC for which forms of the acid exist under different pH values [33]. The extent of adsorption is, as with anions of a weak acid, strongly dependent on pH and favored by lower pH values [34]. In HPLC if the pH of the sample solution and the eluent is not well-matched with each other and around the pKa of the organic acid deformed peaks will appear and then mislead the HPLC analysis conclusions [35,36]. Therefore, the pH could influence the mechanism of the antibiotic interaction with the manganese oxide while those issues have not been systematically addressed.

This study investigated the oxidative degradation of DIC on γ-MnO$_2$ suspensions by varying the key operating parameter pH, which highly influenced the surface features and redox potential of Mn-oxides [37] and the charge density of the chemical form of DIC [38]. Therefore, degradative mechanisms, intermediates, and final products were investigated in terms of pH and compared with the performances of other structures of Mn-oxides presented in the literature.

2. Materials and Methods

2.1. Materials

The sodium diclofenac (CAS 15307-86-5) with a purity of $>$98.5% was purchased from Sigma-A. All other chemicals used in this study were of analytical grade and purchased from Sigma A (St. Louis, MO, USA), J. T. Baker (Phillipsburg, NJ, USA) and Riedel-deHean (Seelze, Germany). Mn-oxide purchased from Tosoh was characterized as tunnel-structured (or molecular sieve) γ-MnO$_2$ (JCPDS 14-0644, PANalytical X’Pert Pro MRD diffractometer) with the Brunauer–Emmett–Teller (BET) specific surface area of 45.6 m$^2$ g$^{-1}$ (Micrometrics ASAP, USA, 2010) and pH$_{zpc}$ of 5.1 (Dispersion Technology, USA, DT-1200).

2.2. Batch Experiments

Experiments were conducted as a function of pH (4–9). For each batch system, various amounts of a γ-MnO$_2$ suspension solution were added to 15-mL glass centrifuge tubes. In the solution, 0.005 M NaH$_2$PO$_4$ and NaH$_2$BO$_3$ were added as a buffer. Various proportions of 0.1 M HCl and NaOH were used to adjust pH to the designed value within a controlled range ($\pm 0.07$). After each reaction course, the solution pH was remeasured to confirm that it remained within the controlled range. For simplification, the pH value is indicated as the designed value in the following paragraphs. The initial concentration of sodium diclofenac (CAS 15307-86-5) prepared was 100 $\mu$M, which was confirmed to be completely soluble, with a water solubility of $10^{-5.3}$–$10^{-1.78}$ M [39,40]. The centrifuge tubes were covered with an aluminum foil to prevent light exposure. The suspensions were rotated
at 25 °C through end-over-end rotations at 10 rpm for a specific time in kinetic trials and 24 h in thermodynamic tests. All experiments were conducted in duplicate. Moreover, controls (no MnO₂ powder) were established using a similar preparatory process to account for sorption on glass tubes and other reactions in the solution.

2.3. Sample Preparation and Analysis

After reactions, the suspensions were centrifuged (Pico 17, Thermo Scientific, Waltham, MA, USA) at 8000 rpm for 40 min, and then the supernatant was quantified using high-performance liquid chromatography (HPLC, L-7200, Hitachi, Japan) with a diode array (DAD) detector (L-7455, Hitachi) at 270 nm. Chromatographic separation was conducted using an RP-18 column (150 µm × 4.6 µm and an internal diameter of 5 µm) purchased from Mightysi with an eluent comprising 60% acetonitrile and 40% acidified water (25 mM phosphoric acid). The flow rate and injection volume were 1 mL min⁻¹ and 20 µL, respectively.

2.4. Identification of Oxidation Products

Major oxidation products were identified using HPLC with mass spectrometry (HPLC–MS). The HPLC–MS system comprised an Agilent 1100 Series liquid chromatography system (LC, Agilent, Palo Alto, CA, USA) with a CTC PAL auto-sampler (CTC Analytica, Carrboro, NC, USA) separation module interfaced with an API 4000 triple quadrupole mass spectrometer (Applied Biosystems AB/MDS Sciex, Foster City, CA, USA). The LC column was a Luna Polar RP (150 mm × 2.1 mm internal diameter) column purchased from Phenomenex (Torrance, CA, USA). The flow rate and injection volume were 0.5 mL min⁻¹ and 10 µL, respectively. An HPLC gradient was established by mixing two mobile phases: acetonitrile and deionized water, with 10 mM formic acid. Chromatographic separation was achieved with the following gradient: 0 to 1 min: 0% acetonitrile; 1–5 min: linear-gradient to 100% acetonitrile; 5–10 min: 100% acetonitrile; 10 to 10.1 min: 0% acetonitrile; and 10.1–15 min: 0% acetonitrile. Mass spectrometer parameters operated in a positive ion mode were as follows: curtain gas, 20 psi; ion source gas 1, 30 psi; ion source gas 2, 40 psi; source temperature, 500 °C; entrance potential, 10 V; and nebulizer current, 5 µA, and the interface heater was turned on. Positive ions in the range of 100–500 m/z were scanned at a cycle time of 1 s. The data obtained were processed with Analyst 1.4.2 software.

3. Results and Discussions

3.1. DIC Degradation Kinetics on the Tunnel-Structured Mn-Oxide

Figure 1 presents the kinetic data of DIC degradation on the tunnel-structured Mn-oxide denoted as dot symbols. Interfacial reactions between DIC and γ-MnO₂ were highly pH-dependent and initially involved a rapid removal of DIC, followed by gradual slowdown and eventual approach to a plateau. In the acidic medium (pH 4–6), the gradual slowdown period was longer than that in neutral and alkaline conditions (pH 7–9) within the tested pH range. The complicated and multistep reactions between the organic micropollutant interface and Mn-oxides result in limitations of kinetics studies; therefore, only initial reaction rates have been explored in most studies and have been generally characterized with a pseudo-first-order degradation model [27,41]. However, the pseudo-first-order kinetics may not satisfy conditions for the later stage of the interfacial reaction. In general, the interfacial reaction can be initiated by the formation of a precursor complex between the Mn(IV) of oxide surface and target organic pollutants, subsequently followed by electron transfer within the precursor complex, redox product formation (Equations (1)–(5)). Either formation of the precursor complex (Equation (1)) or electron transfer within the precursor complex (Equation (2)) is likely to be the rate-limiting step [42]. The formation of redox products, including the surface Mn(III) and Mn(II) (Equation (4)), and further oxidation or combination of organic radicals to form products (Equation (5)), was rapid because of the unstable nature of intermediates.
An integrated kinetic model [42] was applied to examine the DIC reaction over γ-MnO₂. The kinetic equation can be expressed as follows:

$$\frac{d[≡ Mn^{IV} \cdots DIC]}{dt} = k_1[≡ Mn^{IV}] [DIC] - k_{-1} [≡ Mn^{IV} \cdots DIC] - k_2 [≡ Mn^{IV} \cdots DIC]$$

(6)

Total reactive surface sites (S) for DIC degradation can be represented as follows:

$$S = [≡ Mn^{IV}] + [≡ Mn^{IV} \cdots DIC] + [≡ Mn^{III} \cdots DIC] + [≡ Mn^{III}] + [≡ Mn^{II}]$$

(7)

Both $≡ Mn^{III} \cdots DIC$ and $≡ Mn^{III}$ are negligible in Equation (7) because of their high instability. The $≡ Mn^{II}$ concentration can be calculated with the concentration difference between parent DIC at initial ($C_0$) and specific (C) times on account of two electrons transferred from parent DIC to Mn-oxide [43]. To verify the rate-limited step and degradation mechanism, $k_1C >> k_{-1} + k_2$ and $k_{-1} + k_2 >> k_1$, respectively, were assumed for electron transfer control and precursor complex formation control, and the analytical solution of Equation (6) for electron transfer control kinetic model is as follows:

$$C = (C_0 - S) + Se^{-k_{et}t},$$

(8)

where $k_{et}$ equals to $k_2$ and represents the rate constant of the electron transfer control mechanism.

The analytical solution for the precursor complex formation control model is as follows:

$$C = \frac{S - C_0}{\frac{k_{pf}}{k_{-1}k_{2}}[S-C_0]^t - 1},$$

(9)

where $k_{pf}$ equals to $\frac{k_1k_2}{k_{-1}k_{2}}$ and denotes the rate constant of the precursor formation control mechanism.
3.2. Correlation between pH and Oxidative Kinetic Constants

The pH of the solution shifted the degradation kinetics from electron-transfer control to precursor complex-formation control (Figure 1). The electron-transfer control mechanism model successfully described degradation evolution with time under acidic conditions (pH 4–6), whereas under neutral-to-alkaline conditions (pH 7–9), the precursor complex-formation control mechanism was highly fitting to the experimental data (high r-value in Table 1). As Table 1 indicates, the pHs (7–9) with precursor complex-formation control mechanism exactly correspond to the DIC existing as 100% ionized species. Figure S1 (supplementary materials) displayed the DIC species distribution versus solution pH based on the calculation of DIC’s pKa 4.15 [43].

When pH was higher than 7, the ionized species account for 100% of DIC in solution. Since the formation of precursor complex of DIC with the γ-MnO₂ surface is coupled with a release of OH⁻ ions [34], the anionic species of DIC confront with the competition of OH⁻ ions for the surface sites under pH 7–9. Thus, adsorption was not favored by higher pH value (7–9) and the precursor complex formation becomes the control mechanism.

Moreover, the rate constant (k̄et or k̄pf) and surface reaction site (S) for DIC degradation decreased when pH increased (Table 1). The inverse relationship between k and pH for DIC could be partially attributed to a decrease in the reduction potential of MnO₂ with an increase in pH (Equation (10)).

$$
\text{MnO}_2 + 4H^+ + 2e^- \leftrightarrow Mn^{2+} + 2H_2O \quad E_H = E_H^0 + 0.0296 \log \frac{[\text{MnO}_2][H^+]^4}{[Mn^{2+}]}.
$$

Table 1. Fitting constants (k and S) of the kinetic model under various pH.

| pH  | k̄et or k̄pf | S (µM) | r   | Kinetic Model | Ionized Species of DIC (%) † |
|-----|--------------|--------|-----|--------------|-------------------------------|
| 4.0 | 0.998        | 62.3   | 0.96| et †         | 41.5                          |
| 5.0 | 0.623        | 41.1   | 0.94| et           | 87.6                          |
| 6.0 | 0.887        | 18.6   | 0.91| et           | 98.6                          |
| 7.0 | 0.590        | 11.8   | 0.93| pf †         | 100                           |
| 8.0 | 0.588        | 11.0   | 0.97| pf           | 100                           |
| 9.0 | 0.541        | 9.7    | 0.96| pf           | 100                           |

† et: electron-transfer-limited mechanism; # pf: precursor complex-formation-limited mechanism; † calculated by the DIC’s pKa 4.15.
In addition to reducing the potential, solution pH alters the amounts of surface reactive sites (S, Table 1). Under acidic conditions, a large amount of S was expected because of the relatively strong affinity of anionic DIC (pKa = 4.15 [44]) species on the surface of net positively charged MnO₂. Consequently, electron transfer was limited against sufficient active reaction sites for DIC within the tested acidic pH (4–6). When pH increased, electrostatic attraction between the net negatively charged surface and anionic DIC species decreased. Furthermore, OH⁻ strongly competed against DIC for surface-bound Mn(IV). A lower amount of S at higher pH represented insufficient active reaction sites for DIC attachment, leading to the removal mechanism to shift to precursor complex-formation control mechanism. In addition, each component was actually derived from the initial dosage of MnO₂ (Equation (7)); therefore, surface reaction sites were presumed to be functions of initial dosage of MnO₂ and solution pH:

\[ S = \left[ \text{MnO}_2 \right] \left[ H^+ \right]^n. \]  

To present the H⁺ concentration as pH, the log form can be written as follows:

\[ \log S = \log \left[ \text{MnO}_2 \right] - n \cdot \log \text{pH}. \]  

To investigate the influence of pH on S and the kinetic mechanism shift, the log S values extracted from Table 1 and the log value of original MnO₂ loading (log 1150 = 3.06) were plotted as a function of pH (Figure 2). The influencing order (n) of pH was determined as the slope of the straight line of correlation and was equal to 0.24. The correlation coefficient (r) of 0.95 corroborates the presumption that the amounts of surface reactive sites are the function of solid oxidant loadings and solution pH.

\[ \text{Figure 2. Surface active sites (S) relative to solution pH.} \]

Compared with the Mn-oxide dosage yield (total mole of DIC removal per mole of MnO₂ dosage), that for DIC removal by employing γ-MnO₂ in this study was 0.07, which falls in relatively higher than 4.49 × 10⁻⁴–0.14 reported in studies on DIC degradation using other structured Mn-oxides [27,45,46], at 24 h under similar pH conditions. Despite varying the DIC concentration and Mn-oxide dosage from µM to mM in these studies, degradation efficiencies could be compared in a unified manner when the oxide dosage yield was introduced. The remarkable differences in dosage yields indicated that the structure of Mn-oxides substantially influenced their degradative capacity toward DIC, and striking differences were observed for their sorption, oxidative, catalytic, and electrochemical properties [47–49]. The higher oxide dosage yield of γ-MnO₂ could be ascribed to the higher amounts of more flexible corner-shared MnO₆ sites dominated in Mn-oxide bulk, which may facilitate oxidation for target pollutant degradation [49–51].
3.3. Identification of Oxidation Products Using HPLC–MS

HPLC–MS was used to determine the M/Z ratio of parent DIC, oxidation intermediates (reaction time of 2 h), and products (reaction time of 24 h), and Figures 3–5, respectively, present their MS chromatograms. Figure 3a has a peak with a very pronounced tailing and this phenomenon should be due to the pH mismatch effect mentioned in a previous study [35]. This fact should not affect the identification of the oxidation intermediates and products because the pronounced tail did not appear after reaction (because the concentration of DIC decreased significantly) and most of the compounds did not appear in this region. System peaks were observed in Figure 4 (Figure 4a,b) and Figure S2. This phenomenon reflected some compounds which are strongly absorbed to the stationary phase were generated after reaction [36]. These compounds cannot be identified using this effluent procedure.

Under neutral-to-alkaline conditions, no intermediates were detected at a reaction interval of 2 h, and compared with acidic conditions, fewer oxidative products were obtained at 24 h according to MS analysis results (Figure S2). Because of the relatively low degradation of DIC under neutral and alkaline conditions, the MS analyses of intermediates and final products were mainly conducted under the acidic condition.

Because of the ionic nature of DIC, two electrospray ionization (ESI) methods, ESI+ and ESI−, were employed to study degradation products, and Table 2 presents the results. Under acidic conditions, three intermediates (I₁, I₂, and I₃) were formed after 2 h of the reaction, and four final products (F₁, F₂, F₃, and F₄) were obtained after a day of the reaction.

Figure 3. High-performance liquid chromatography (HPLC)–mass spectroscopy (MS) chromatographic patterns of DIC standard. (a) Total ion chromatogram (TIC) in the ESI+ mode, (b) TIC in the ESI− mode, (c) MS patterns in the ESI+ mode, and (d) MS patterns in the ESI− mode.
Figure 4. HPLC–MS chromatographic patterns of degradation intermediates. (a) TIC in the ESI+ mode, (b) TIC in the ESI− mode, (c) MS patterns in the ESI+ mode, and (d) MS patterns in the ESI− mode under a pH of 5 for a reaction time of 2 h with an initial MnO₂ loading of 200 mg/L.
Figure 5. HPLC–MS chromatographic patterns of degradation products. (a) TIC in the ESI+ mode, (b) TIC in the ESI− mode, (c) MS patterns in the ESI+ mode, and (d) MS patterns in the ESI− mode under a pH of 5 for a reaction time of 24 h with an initial MnO$_2$ loading of 200 mg/L.
According to Monteagudo et al. [52,53], I₁ (RT = 2.09, m/z = 346) correspond to tri-hydroxyl-DIC (m/z = 346) or di-hydroxyl-DIC (m/z = 328). I₂ (RT = 2.48, m/z = 298) should be a hydrolyzed-decarboxylated DIC (296 – 14 + 16 = 298). The molecular weight of I₃ (RT = 2.92, m/z = 597) is considerably higher than that of DIC. Moreover, I₃ exhibited numerous isotopic peaks, and its intensity ratio of (M + 1)/Z to (M + 3)/Z was approximately 3:4, which revealed that these compounds contained four chlorine atoms. Thus, I₃ should be a dimer of 5-iminoquinone DIC (m/z = 308) and another intermediate. This finding indicated that polymerization or dimerization, which was found during the reaction of other aromatic compounds with Mn-oxides, may occur during DIC degradation by γ-MnO₂. Similar results were reported by Huguet et al. [26].

F₁ (RT = 3.02, m/z = 503) is a new product, and its molecular weight is substantially higher than that of DIC. Therefore, it should be a transformation product of I₁. F₂ (RT = 3.58, m/z = 308) and F₃ (RT = 3.92, m/z = 312) correspond to 5-iminoquinone DIC and hydroxyl-DIC, respectively, which have been reported in literature [26]. The peak of hydroxyl-DIC (F₃) split into two and the m/z ratio (255) in negative mode was considerably lower than the m/z ratio (312) in the positive mode. The split of the peak could be attributed to the different sites of the hydroxyl group of the compound (structure isomers) leading to different hydrophilicity, and the observed difference of m/z ratio for F₃ between in positive mode and negative mode should result from the carbon chain (–CH₂COOH, M = 57) broke during ionization. F₄ (RT = 2.59, m/z = 250) should be a decarboxylation product of DIC (2,6-dichloro-N-o-tolylbenzenamine), which was reported by Martínez et al. [54].

The intensity of F₃ was much higher than that of F₂, and multiple hydroxyl intermediates (I₁) were found. According to studies, decarboxylation, hydroxylation, and dimerization are the three main pathways of DIC transformation by Mn-oxides [26]. The pathways of DIC transformation by γ-MnO₂ are as similar as those of birnessite or other natural manganese oxides [26], and compared with the layer-structured birnessite that is widely used in studies, hydroxylation of DIC by γ-MnO₂ was more active than those by other pathways. This phenomenon could corroborate that the large amounts of highly flexible corner-shared MnO₆ may provide abundant reactive hydroxyl groups and facilitate oxidation for target pollutant degradation [50,51]. Therefore, the dimerization products of DIC obtained through γ-MnO₂ are highly hydrophilic and can be detected without extraction. Hydroxylation intermediates were not detected after 1 day because they were oxidized to smaller or hydrophilic compounds due to further hydroxylation.

| Compound | RT (min) | Model | m/z            | Possible Structure | Ref |
|----------|----------|-------|----------------|-------------------|-----|
| DIC      | 3.18     | ESI+  | 296/298 (3/2)  | ![DIC structure](attachment) |     |
|          |          |       | 278/280, 250/252, 215/217 | |     |
|          |          |       | 250/252 (3/2)  |                   |     |
| F₁       | 2.09     | ESI+  | 346/348 (3/2)  | Tri or Di—hydroxyl DIC | [52,53] |
|          |          |       | 328/330, 284/286, 244/246, 162/164 | |     |
| I₂       | 2.48     | ESI+  | 298/300 (3/2)  | ![I₂ structure](attachment) |     |
|          |          |       | 267, 244       |                   |     |
|          |          | ESI−  | 266/264 (3/2)  |                   |     |
|          |          |       | 228, 230       |                   |     |
| I₃       | 2.92     | ESI+  | 597/599 (3/4)  | Dimer             |     |
|          |          |       | 575/577 (3/4), 551/553 (3/4), 507/509 (3/4), 308, 267, 253 | |     |
Table 2. Cont.

| Compound | RT (min) | Model | m/z            | Possible Structure | Ref     |
|----------|---------|-------|----------------|--------------------|---------|
| F1       | 3.02    | ESI+  | 503 461, 425, 409, 373, 329, 308, 267 | Dimer             | [26,55] |
| F2       | 3.58    | ESI+  | 308, 267 |                    |         |
| F3       | 3.93    | ESI+  | 312          |                    | [26,55] |
| F4       | 2.59    | ESI−  | 250/252 (3/2) 214 |                | [54]    |

RT: retention time; bold number: parent ion.

4. Conclusions

This study demonstrated that the pH of media highly influences DIC oxidative degradation on the tunnel-structured Mn-oxide (γ-MnO₂). The reduction potential of Mn-oxide, the number of surface reactive sites (S), and electrostatic affinity between DIC and γ-MnO₂ increase with a decrease in pH value. Consequently, the electron-transfer control mechanism model successfully described degradation evolution with time under acidic conditions (pH = 4–6). While under neutral-to-alkaline conditions (pH = 7–9), the precursor complex-formation control mechanism was highly fitting to the experimental data. At pH 7–9 the anionic species account for 100% DIC in solution and hence confront with the competition of OH⁻ ions for the complex formation on the γ-MnO₂ surface. In contrast, the acid form of DIC with a substantial ratio under pH 4–6 is favored for the surface complex formation with less competition. The results of the analysis of oxidative intermediates and products by using HPLC–MS revealed decarboxylation, hydroxylation, and dimerization as the three main pathways of DIC transformation by γ-MnO₂. Although the oxidation products obtained by γ-MnO₂ are similar to those obtained by other Mn-oxides, hydroxylation of DIC by γ-MnO₂ is more active than other pathways because of an abundance of flexible corner-shared MnO₆ for target pollutant degradation.

Supplementary Materials: The following are available online: http://www.mdpi.com/2073-4441/12/8/2203/s1, Figure S1: DIC species distribution versus solution. HA and A⁻ represent the acid and ionized form of DIC, respectively. The black and gray line were calculated based on pKa = 4.15 of DIC [44]. When pH higher than 7, the ionized species (A⁻) accounts for 100% of DIC in solution pH. Figure S2: LC–MS chromatographic patterns of degradation intermediates (a) TIC in ESI⁺ mode (b) MS patterns in ESI⁺ mode under pH 7.0 for reaction time = 24 h with initial MnO₂ loading 200 mg.

Author Contributions: C.-Y.H. conceived and designed the experiments. Y.-J.L. performed the experiments. C.-Y.H. and W.-H.K. analyzed the data and wrote the paper. All authors have read and agreed to the published version of the manuscript.

Funding: This research received the funding from Ministry of Science and Technology of the Republic of China, Contract No. NSC 98-2221-E-038 -004, MOST 105-2221-E-131-001-MY3 and MOST 108-2221-E-131-024-MY3.

Conflicts of Interest: The authors declare no conflict of interest.
References

1. Ternes, T.A.; Herrmann, N.; Bonerz, M.; Knacker, T.; Siegrist, H.; Joss, A. A rapid method to measure the solid-water distribution coefficient (K-d) for pharmaceuticals and musk fragrances in sewage sludge. *Water Res.* 2004, 38, 4075–4084. [CrossRef] [PubMed]

2. Joss, A.; Zabczynski, S.; Gobel, A.; Hoffmann, B.; Loffler, D.; McArdell, C.S.; Ternes, T.A.; Thomsen, A.; Siegrist, H. Biological degradation of pharmaceuticals in municipal wastewater treatment: Proposing a classification scheme. *Water Res.* 2006, 40, 1686–1696. [CrossRef] [PubMed]

3. Camacho-Munoz, D.; Martin, J.; Santos, J.L.; Aparicio, I.; Alonso, E. Occurrence, temporal evolution and risk assessment of pharmaceutically active compounds in Donana Park (Spain). *J. Hazard. Mater.* 2010, 183, 602–608. [CrossRef] [PubMed]

4. Comeau, F.; Surette, C.; Brun, G.L.; Losier, R. The occurrence of acidic drugs and caffeine in sewage effluents and receiving waters from three coastal watersheds in Atlantic Canada. *Sci. Total Environ.* 2008, 393, 132–146. [CrossRef] [PubMed]

5. Schwab, B.W.; Hayes, E.P.; Fiori, J.M.; Mastrocco, F.J.; Roden, N.M.; Cragin, D.; Meyerhoff, R.D.; D’Aco, V.J.; Anderson, P.D. Human pharmaceuticals in US surface waters: A human health risk assessment. *Regul. Toxicol. Pharmacol.* 2005, 42, 296–312. [CrossRef]

6. Sidlova, P.; Podlipna, R.; Vanek, T. Cytotoxic pharmaceuticals in the environment. *Chem. Listy* 2011, 105, 8–14.

7. Tauxe-Wuersch, A.; de Alencastro, L.F.; Grandjean, D.; Tarradellas, J. Occurrence of several acidic drugs in sewage treatment plants in Switzerland and risk assessment. *Water Res.* 2005, 39, 1761–1772. [CrossRef]

8. Weber, S.; Khan, S.; Hollender, J. Human risk assessment of organic contaminants in reclaimed wastewater used for irrigation. *Desalination* 2006, 187, 53–64. [CrossRef]

9. Lonappan, L.; Brar, S.K.; Das, R.K.; Verma, M.; Surampalli, R.Y. Diclofenac and its transformation products: Environmental occurrence and toxicity—A review. *Environ. Int.* 2016, 96, 127–138. [CrossRef]

10. Oaks, J.L.; Gilbert, M.; Virani, M.Z.; Watson, R.T.; Meteyer, C.U.; Rideout, B.A.; Shivaprasad, H.L.; Ahmed, S.; Iqbal Chaudhry, M.J.; Arshad, M.; et al. Diclofenac residues as the cause of vulture population decline in Pakistan. *Nature* 2004, 427, 630–633. [CrossRef]

11. Taggart, M.A.; Senacha, K.R.; Green, R.E.; Jhala, Y.V.; Raghavan, B.; Rahmani, A.R.; Cuthbert, R.; Pain, D.J.; Meharg, A.A. Diclofenac residues in carcasses of domestic ungulates available to vultures in India. *Environ. Int.* 2007, 33, 759–765. [CrossRef] [PubMed]

12. Schmitt-Jansen, M.; Bartels, P.; Adler, N.; Altenburger, R. Phytotoxicity assessment of diclofenac and its phototransformation products. *Anal. Bioanal. Chem.* 2007, 387, 1389–1396. [CrossRef] [PubMed]

13. Stülten, D.; Zühlke, S.; Lamshöft, M.; Spitterler, M. Occurrence of diclofenac and selected metabolites in sewage effluents. *Sci. Total Environ.* 2008, 405, 310–316. [CrossRef]

14. Stone, A.T. Reductive dissolution of manganese(III/IV) oxides by substituted phenols. *Environ. Sci. Technol.* 1987, 21, 979–988. [CrossRef] [PubMed]

15. Zhang, H.C.; Huang, C.H. Oxidative transformation of triclosan and chlorophene by manganese oxides. *Environ. Sci. Technol.* 2003, 37, 2421–2430. [CrossRef] [PubMed]

16. Park, J.W.; Dec, J.; Kim, J.E.; Bollag, J.M. Effect of humic constituents on the transformation of chlorinated phenols and anilines in the presence of oxidoreductive enzymes or birnessite. *Environ. Sci. Technol.* 1999, 33, 2028–2034. [CrossRef]

17. Ulrich, H.J.; Stone, A.T. The oxidation of chlorophenols adsorbed to manganese oxide surfaces. *Environ. Sci. Technol.* 1989, 23, 421–428. [CrossRef]

18. Li, H.; Lee, L.S.; Jafvert, C.T.; Gravel, J.G. Effect of substitution on irreversible binding and transformation of aromatic amines with soils in aqueous systems. *Environ. Sci. Technol.* 2000, 34, 3674–3680. [CrossRef]

19. Pizzigallo, M.D.R.; Ruggiero, P.; Crecchio, C.; Mascolo, G. Oxidation of chloroanilines at metal oxide surfaces. *J. Agric. Food Chem.* 1998, 46, 2049–2054. [CrossRef]

20. Lin, K.; Liu, W.; Gan, J. Oxidative removal of bisphenol A by manganese dioxide: Efficacy, products, and pathways. *Environ. Sci. Technol.* 2009, 43, 3860–3864. [CrossRef]

21. Lu, Z.J.; Lin, K.D.; Gan, J. Oxidation of bisphenol F (BPF) by manganese dioxide. *Environ. Pollut.* 2011, 159, 2546–2551. [CrossRef] [PubMed]
22. Sarmah, P.; Dutta, D.K. Manganese mediated aqueous reduction of aromatic nitro compounds to amines. J. Chem. Res. 2003, 236–237. [CrossRef]
23. Rubert, K.F.; Pedersen, J.A. Kinetics of oxytetracycline reaction with a hydrous manganese oxide. Environ. Sci. Technol. 2006, 40, 7211–7212. [CrossRef] [PubMed]
24. Sabirova, J.S.; Cloetens, L.F.F.; Vanhaecke, L.; Forrez, I.; Verstraete, W.; Boon, N. Manganese-oxidizing bacteria mediate the degradation of 17α-ethinylestradiol. Microb. Biotechnol. 2008, 1, 507–512. [CrossRef] [PubMed]
25. Xu, L.; Xu, C.; Zhao, M.; Qiu, Y.; Sheng, G.D. Oxidative removal of aqueous steroid estrogens by manganese oxides. Water Res. 2008, 42, 5038–5044. [CrossRef]
26. Huguet, M.; Debode, M.; Papot, S.; Gallard, H. Oxidative decarboxylation of diclofenac by manganese oxide bed filter. Water Res. 2013, 47, 5400–5408. [CrossRef]
27. Forrez, I.; Carballa, M.; Verbeeken, K.; Vanhaecke, L.; Schlusener, M.; Ternes, T.; Boon, N.; Verstraete, W. Diclofenac Oxidation by Biogenic Manganese Oxides. Environ. Sci. Technol. 2010, 44, 3449–3454. [CrossRef]
28. Nihemaiti, M.; Le Roux, J.; Hoppe-Jones, C.; Reckhow, D.A.; Croué, J.-P. Formation of haloacetonitriles, haloacetamides, and nitrogenous heterocyclic byproducts by chloramination of phenolic compounds. Environ. Sci. Technol. 2017, 51, 655–663. [CrossRef]
29. Chen, T.; Dou, H.; Li, X.; Tang, X.; Li, J.; Hao, J. Tunnel structure effect of manganese oxides in complete oxidation of formaldehyde. Microporous Mesoporous Mater. 2009, 122, 270–274. [CrossRef]
30. Luo, J.; Zhang, Q.; Garcia-Martinez, J.; Suib, S.L. Adsorptive and acidic properties, reversible lattice oxygen evolution, and catalytic mechanism of cryptomelane-type manganese oxides as oxidation catalysts. J. Am. Chem. Soc. 2008, 130, 3198–3207. [CrossRef]
31. Wu, C.; Xie, W.; Zhang, M.; Bai, L.; Yang, J.; Xie, Y. Environmentally friendly γ-MnO2 hexagon-based nanoarchitectures: Structural understanding and their energy-saving applications. Chem. A Eur. J. 2009, 15, 492–500. [CrossRef] [PubMed]
32. Zhang, J.; Li, Y.; Wang, L.; Zhang, C.; He, H. Catalytic oxidation of formaldehyde over manganese oxides with different crystal structures. Catal. Sci. Technol. 2015, 5, 2305–2313. [CrossRef]
33. Silberberg, M. Chemistry: The Molecular Nature of Matter and Change with Advanced Topics; McGraw-Hill: New York, NY, USA, 2018.
34. Stumm, W. Chemistry of the Solid-Water Interface: Processes at the Mineral-Water and Particle-Water Interface in Natural Systems; John Wiley & Son Inc.: New Jersey, NJ, USA, 1992.
35. Samuelsson, J.; Forssén, P.; Forstnå, T. Sample conditions to avoid pH distortion in RP-LC. J. Sep. Sci. 2013, 36, 3769–3775. [CrossRef] [PubMed]
36. Forstnå, T.; Forssén, P.; Westerlund, D. System peaks and their impact in liquid chromatography. TrAC Trends Anal. Chem. 2016, 81, 42–50. [CrossRef]
37. Xyla, A.G.; Sulzberger, B.; Luther III, G.W.; Hering, J.G.; van Cappellen, P.; Stumm, W. Reductive dissolution of manganese (III, IV)(hydr) oxides by oxalate: The effect of pH and light. Langmuir 1992, 8, 95–103. [CrossRef]
38. De Oliveira, T.; Guégan, R.; Thiebault, T.; Le Milbeau, C.; Muller, F.; Teixeira, V.; Giovanela, M.; Boussafir, M. Adsorption of diclofenac onto organoclays: Effects of surfactant and environmental (pH and temperature) conditions. J. Hazard. Mater. 2017, 323, 558–566. [CrossRef]
39. Das, R.N.; Roy, K. QSAR with extended topochemical atom (ETA) indices. 4. Modeling aqueous solubility of drug like molecules and agrochemicals following OECD guidelines. Struct. Chem. 2013, 24, 303–331. [CrossRef]
40. Loos, R. Analytical Methods for Possible WFD 1st Watch List Substances; European Commission, Joint Research Center, Institute for Environment and Sustainability, Publications Office of the European Union: Luxembourg, 2015.
41. Zhao, J.; Wang, Q.; Fu, Y.; Peng, B.; Zhou, G. Kinetics and mechanism of diclofenac removal using ferrate (VI): Roles of Fe(3+), Fe(2+), and Mn(2+). Environ. Sci. Pollut. Res. 2018, 25, 22998–23008. [CrossRef]
42. Zhang, H.C.; Chen, W.R.; Huang, C.H. Kinetic modeling of oxidation of antibacterial agents by manganese oxide. Environ. Sci. Technol. 2008, 42, 5548–5554. [CrossRef]
43. Zhang, H.C.; Huang, C.H. Oxidative transformation of fluoroquinolone antibacterial agents and structurally related amines by manganese oxide. Environ. Sci. Technol. 2005, 39, 4474–4483. [CrossRef]
44. Bui, T.X.; Choi, H. Influence of ionic strength, anions, cations, and natural organic matter on the adsorption of pharmaceuticals to silica. Chemosphere 2010, 80, 681–686. [CrossRef] [PubMed]
45. Li, J.; Zhang, T.; Ye, M. Heterogeneous oxidation of diclofenac in the presence of α-MnO2 nanorods: Influence of operating factors and mechanism. *Water Sci. Technol.* 2015, 71, 1340–1346. [CrossRef] [PubMed]

46. Liu, W.; Sutton, N.B.; Rijnaarts, H.H.; Langenhoff, A.A.J.E.S.; Research, P. Anoxic conditions are beneficial for abiotic diclofenac removal from water with manganese oxide (MnO2). *Environ. Sci. Pollut. Res. Int.* 2018, 25, 10141–10147. [CrossRef] [PubMed]

47. Su, D.; Ahn, H.-J.; Wang, G. Hydrothermal synthesis of α-MnO2 and β-MnO2 nanorods as high capacity cathode materials for sodium ion batteries. *J. Mater. Chem. A* 2013, 1, 4845–4850. [CrossRef]

48. Wang, Y.; Feng, X.; Villalobos, M.; Tan, W.; Liu, F. Sorption behavior of heavy metals on birnessite: Relationship with its Mn average oxidation state and implications for types of sorption sites. *Chem. Geol.* 2012, 292, 25–34. [CrossRef]

49. Smith, P.F.; Deibert, B.J.; Kaushik, S.; Gardner, G.; Hwang, S.; Al-Sharab, J.F.; Garfunkel, E.; Fabris, L.; Li, J.; et al. Coordination Geometry and Oxidation State Requirements of Corner-Sharing MnO6 Octahedra for Water Oxidation Catalysis: An Investigation of Manganite (γ-MnOOH). *ACS Catal.* 2016, 6, 2089–2099. [CrossRef]

50. Li, G.; Li, K.; Liu, A.; Yang, P.; Du, Y.; Zhu, M. 3D flower-like β-MnO2/reduced graphene oxide nanocomposites for catalytic ozonation of dichloroacetic acid. *Sci. Rep.* 2017, 7, 43643. [CrossRef]

51. Kasprzyk-Hordern, B.; Ziolkł, M.; Nawrocki, J. Catalytic ozonation and methods of enhancing molecular ozone reactions in water treatment. *Appl. Catal. B Environ.* 2003, 46, 639–669. [CrossRef]

52. Monteagudo, J.M.; El-taliawy, H.; Durán, A.; Caro, G.; Bester, K. Sono-activated persulfate oxidation of diclofenac: Degradation, kinetics, pathway and contribution of the different radicals involved. *J. Hazard. Mater.* 2018, 357, 457–465. [CrossRef]

53. Yu, H.; Nie, E.; Xu, J.; Yan, S.; Cooper, W.J.; Song, W. Degradation of diclofenac by advanced oxidation and reduction processes: Kinetic studies, degradation pathways and toxicity assessments. *Water Res.* 2013, 47, 1909–1918. [CrossRef]

54. Martínez, C.; Canle, L.M.; Fernández, M.I.; Santaballa, J.A.; Faria, J. Aqueous degradation of diclofenac by heterogeneous photocatalysis using nanostructured materials. *Appl. Catal. B Environ.* 2011, 107, 110–118. [CrossRef]

55. Soufan, M.; Deborde, M.; Legube, B. Aqueous chlorination of diclofenac: Kinetic study and transformation products identification. *Water Res.* 2012, 46, 3377–3386. [CrossRef] [PubMed]

© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).