The promise of stem cells in the therapy of Alzheimer’s disease

Chunmei Yue and Naihe Jing*

Abstract

Alzheimer’s disease (AD), a common neurodegenerative disorder associated with gradually to dramatic neuronal death, synaptic loss and dementia, is considered to be one of the most obscure and intractable brain disorders in medicine. Currently, there is no therapy clinically available to induce marked symptomatic relief in AD patients. In recent years, the proof-of-concept studies using stem cell-based approaches in transgenic AD animal models provide new hope to develop stem cell-based therapies for the effective treatment of AD. The degeneration of basal forebrain cholinergic neurons (BFCNs) and the resultant cholinergic abnormalities in the brain contribute substantially to the cognitive decline of AD patients. The approaches using stem cell-derived BFCNs as donor cells need to be developed, and to provide proof of principle that this subtype-specific neurons can induce functional recovery of AD animal models. With the continuous scientific advances in both academic and industrial fields, the potentials of stem cells in cellular neuroprotection and cell replacement in vivo have been elucidated, and stem cell-based therapy for repairing degenerative brains of AD is promising.

Keywords: Alzheimer’s disease, Stem cell-based therapy, Basal forebrain cholinergic neurons, Cognitive impairment, Embryonic stem cells

Alzheimer’s disease (AD), a multifactorial syndrome, is believed to be the most common form of neurodegenerative dementia. Previous studies have demonstrated that AD is associated with several distinct neuropathological features, mainly as extracellular Aβ plaques, intracellular neurofibrillary tangles and neuron lost. Currently, there is no effective therapies available for the treatment of AD, and the widely used medicine, including cholinesterase inhibitors, have very modest clinical effects in treating the symptoms of AD, such as language loss, cognitive deficits and behavioral impairments. It was concluded that AD epitomized the mechanistic ignorance and therapeutic nihilism that pervaded the study of neurodegeneration in humans [1]. In 2000, approximately 4.5 million people suffered from AD in the America, which is expected to 13.2 million by the year 2050 [2]. The AD patients by no means will become one of the largest public health challenges and place heavy demands on the health care system.

The stem cells hold potentials to develop cell replacement therapies for various neurodegenerative disease, including AD. The subtype-specific neurons from stem cells might be the most ideal donor cells to replace the same type of neurons lost through disease. The embryo stem cell (ESC)-derived dopaminergic neurons were shown to correct functional deficits in animal models of Parkinson’s disease [3,4]. The attempts to develop stem cell-based therapies for AD have used several types of cells in AD animal models, such as adult neural stem cells [5], ESC-derived neural precursor cells [6], a couple of mesenchymal stem cells [7,8], and astrocytes [9]. These studies had demonstrated that transplanted cells partially rescue the cognitive deficits of AD animals. Among the widespread neuron and synaptic loss in AD brain, the degeneration of basal forebrain cholinergic neurons (BFCNs) and severe devastation of basal forebrain cholinergic innervation of cerebral cortex and hippocampus in the brain of AD patients are thought to play essential role in memory impairment, cognitive deficits, and finally lead to dementia [10,11]. The replacement procedures using stem cell-derived BFCNs as donor cells
need to be estabilished, and the functional efficacy of grafted BFCN needs to be determined.

The purpose of this short review is to briefly summarize the continuous advances in the development of stem cell-based approaches for AD in both academic and industrial fields, and carefully predict the promising potentials of stem cells for repairing degenerative brains of AD.

**The scientific advances in academic research**

Recently, multiple attempts have used different types of stem cells as donor cells and investigated the effects of stem cell transplantation on relieving neuropathological symptoms and restoring cognitive function of transgenic AD animal models. Both cultured adult and neonatal mouse astrocytes are transplanted into the hippocampus of ApdE9 AD mice. Seven days later, these astrocytes are found mainly near Aβ deposits and internalize human Aβ immunoreactive material in vivo. This study supports the role of astrocytes as active Aβ clearing cells in the brain, which may have important implications for future development of therapeutic strategies for AD [9]. Adult neural stem cells from newborn GFP transgenic mice have been transplanted into the hippocampus of aged 3xTg-AD mice, a model that recapitulates many of the salient features of AD. The neural stem cell transplantation induces a robust enhancement of BDNF-mediated hippocampal synaptic density and rescues the spatial learning and memory deficits of AD mice, without altering Aβ deposits. This study suggests that modulation of neurotrophin levels could provide a viable approach in the development of stem cell-based therapies to treat AD in future [5]. The human umbilical cord blood-derived mesenchymal stem cells are transplanted into hippocampus of AD mice, which rescues memory deficits of host mice by reducing neuronal apoptosis [7]. The bone marrow-derived mesenchymal stem cells are transplanted into hippocampus of AD mice, which reduces Aβ deposits and rescues memory deficits of host mice [8]. The mouse embryonic stem cell-derived cholinergic motor neuron precursors are transplanted into the right NBM of rats with unilateral NBM lesion and memory deficits, and promote the memory recovery of host rats [6]. The human iPSC-derived neuronal precursors from SHH/RA treatment are transplanted into the hippocampus of AD transgenic mice and restore spatial memory of AD mice [12].

In summary, different types of donor cells that are transplanted mainly into hippocampus of AD model animals exhibit distinct effect on neuropathological symptoms relief and cognitive function restoration (Table 1). All these studies suggest the perspective possibility of using stem cells to develop therapeutics for AD.

**The promising application of basal forebrain cholinergic neurons (BFCNs) in stem cell-based treatment of AD**

There are two major types of cholinergic neurons in the brain, the projection neurons in the basal forebrain, known as BFCNs, and interneurons in the striatum. In rodent brain, BFCNs originating in the most ventral regions of the developing telencephalon at embryonic day 11, display highly discrete locations and possess striking anatomic heterogeneities [13]. In primate brain, the BFCNs situate in the medial septal nucleus, vertical limb of the diagonal band of Broca and nucleus basalis of Meynert (NBM), and typically innervate all parts of cerebral cortex, hippocampus, entorhinal cortex and amygdala [14]. The BFCNs play essential roles in various aspects of cognitive function, such as learning, memory and attention, and the cholinergic blockade disrupts the cognitive function of normal humans [10,15,16]. Numerous studies show the severe devastation of basal forebrain cholinergic innervation and resultant declined cholinergic neurotransmission in the brains of AD patients, and even in the early stage of AD patients [17–20]. Also, the most severely affected areas in AD brain are within the temporal lobes, especially within the hippocampus and entorhinal cortex

| Donor cells                          | AD models                          | Graft site       | Behavior recovery    | Possible mechanism                | Key refs  |
|--------------------------------------|------------------------------------|------------------|----------------------|----------------------------------|-----------|
| Mouse astrocytes                     | ApdE9 transgenic mouse             | Hippocampus      | Not known            | Aβ deposits clearing              | [9]       |
| Mouse adult NSCs                     | 3xTg transgenic mouse              | Hippocampus      | Learning and memory  | BDNF induction                    | [5]       |
| Mouse cholinergic motor neuron precursors | Rat with NBM lesion              | NBM              | memory recovery      | Not Known                        | [6]       |
| Human UCB-MSCs                       | Not known                          | Hippocampus      | Memory               | Reducing neuronal apoptosis       | [7]       |
| Mouse BM-MSCs                        | APP/PS1 transgenic mouse           | Hippocampus      | Memory               | reducing Aβ deposits              | [8]       |
| Human iPSC-derived motor neuron precursors | PDAPP transgenic mouse         | Hippocampus      | Spatial Memory       | Not Known                        | [12]      |

NSCs, neural stem cells, NBM, nucleus basalis of Meynert, UCB-MSCs, umbilical cord blood-derived mesenchymal stem cells, BM-MSCs, bone marrow-derived mesenchymal stem cells, PDAPP, PDGF promoter driven amyloid precursor protein transgenic mice.
[21,22]. These studies point out that the degeneration of BFCNs essentially contribute to the cognitive deficits and pathogenesis of AD, suggesting that BFCNs might be an ideal type of donor cells to ameliorate the cognitive symptoms associated with AD. Even if the hypothesis by using BFCNs as donor cells for treating AD is exciting, none of the studies listed above has attempted to test if BFCNs can restore cholinergic function and alleviate cognitive deficits in AD model animals. Indeed, BFCNs cannot be stably generated from pluripotent stem cells, either mouse or human embryonic stem cells (ESCs) for the transplantation experiments. Up to date, the optimal strategy directing the differentiation of pluripotent stem cells into BFCNs in vitro has not been established, which is mainly due to the unclear molecular basis of the differentiation and development of BFCNs in vivo. A number of endogenous neurotrophic factors, such as nerve growth factor (NGF), brain-derived neurotrophic factor (BDNF), basic fibroblast growth factor (bFGF) and bone morphogenetic protein 9 (BMP9), have been reported to participate and promote the survival, growth, and differentiation of cholinergic neurons, and probably BFCNs in the brain [23–25]. The various transcription factors, such as Mash1, Islet1, Nkx2.1, Lhx8, and Olig2, act hierarchically to determine the cell fate and function coordinately to maintain the cell identity of striatal cholinergic neurons, but the dynamics expression and genetic determinants of these factors in the development of BFCNs are incompletely understood [26–29].

Due to the unclear elucidation of key developmental factors and extracellular signals that control the development of BFCNs in vivo, the successful derivation of BFCNs from mouse ESCs has not been reported, and the studies on differentiation of human ESCs to BFCNs has been sparse. Few years ago, Bissonnette and colleagues [30] reported the derivation of a predominantly pure BFCN population from human ESCs. They used diffusible ligands present in forebrain, including RA in the early stage of neural induction in their method. These human ESC-derived BFCN neurons express typical cholinergic neuronal markers, and stably and functionally engraft in murine ex vivo hippocampal slices through generating electrophysiologically functional cholinergic synapses and firing action potentials. Two years later, Liu and colleagues have developed a method for differentiating hESCs to a nearly homogeneous population of MGE-like progenitors [31]. Following their differentiation protocols, these MGE-like progenitors finally give rise to mixed cell population containing nearly equal amount of GABAergic interneurons and BFCNs. After transplantation into the hippocampus of severe combined immunodeficiency (SCID) mice in which BFCNs and some GABA neurons in the medial septum had been destroyed by mP75-saporin, human MGE-like progenitors produced BFCNs that synaptically connected with endogenous neurons. More excitingly, mice transplanted with MGE-like progenitors showed improvements in learning and memory deficits. The function of ESC-derived BFCNs has been tested from the murine ex vivo hippocampal slices to the severe combined immunodeficiency mice with medial septum lesion, indicating that one step has moved forward on the road of exploring the subtype-specific neuron-based cell therapy for AD. Recently, we have successfully differentiated both mouse and human ESCs into BFCNs through a highly pure population of BFCN progenitors. Both mouse and human ESC-derived BFCN progenitors are transplanted into the NBM of transgenic AD model mice, 5XFAD and APP/PS1, and specifically differentiated into mature and functional cholinergic neurons in vivo. These exogenous cholinergic neurons exhibited typical basal forebrain cholinergic projection and migration patterns, and morphologically and functionally incorporate into the endogenous projection system. Importantly, AD mice with transplanted BFCN progenitors exhibited improved learning and referenced memory abilities in the behavior test, demonstrating the feasibility of using ESC-derived BFCNs for the development of stem cell therapy for AD (unpublished data).

It is well accepted that the degeneration of BFCNs and resultant devastation cholinergic innervation spread a large area of brain, from basal forebrain to hippocampus and entorhinal cortex in temporal lobe. However, the time that BFCNs start to degenerate through disease processing remains unclear, and mechanism underlying the BFCN degeneration needs to be elucidated. These unknown events make it more difficult to optimize the cell replacement procedures using BFCNs. The optimal transplantation site remains to be determined. The survival, proliferation, cell fate specification and differentiation of grafted BFCN progenitors need to be fine controlled. The biochemical functions, migration, projection and

| Table 2 The advantages and disadvantages of using BFCNs as donor cells in the stem cell-based approaches for AD |
| Advantage | Disadvantage |
|-----------------|-----------------|
| Subtype-specific replacing the lost endogenous BFCNs in basal forebrain | The standard preparation of homogeneous human BFCN progenitors |
| Repairing the dysfunctions of basal forebrain cholinergic system by releasing acetylcholine, neurotrophic factors or cytokines | The efficient cholinergic specification of BFCN progenitors, and long term survival of cholinergic derivatives in vivo |
| Functionally integrating into the endogenous basal forebrain neural circuities | The disabilities of long-distance inervation from the basal forebrain to hippocampus and cortex |
| Rescuing the innervation track along the host cholinergic projections through out the basal forebrain | The uneven and patchy migration through the basal forebrain to hippocampus and cortex |
integration of exogenous BFCNs remain largely unknown. Obviously, the procedures of stem cell therapies using ESC-derived BFCNs for the treatment of AD are far from optimal, and the challenges ahead will be tremendous (Table 2).

The translational perspective in stem-cell based therapy of AD in industry

Even with the continuous progress in developing stem cell-based therapy for AD in academic field, the challenges in the early stage of stem-cell based therapy to treat neurodegenerative disease, including AD, are obviously insurmountable. However, the transition from proof-of-concept studies in AD animals to human clinical trials in AD patients has already been underway in the industrial field, which comes sooner than one might think [32]. In 1999, based on their groundbreaking finding of cell surface markers for adult human neural stem cell, the scientists of StemCells Inc. have successfully isolated a highly purified, expandable population of neural stem cells from human brain tissue by using monoclonal antibodies against the cell surface markers. Then, the human neural stem cells have been prepared under controlled conditions and cGMP standards and named HuCNS-SC cells. The rigorous preclinical studies have shown that HuCNS-SC cells can survive long-term with no evidence of tumor formation or adverse effects, and engraft, migrate, differentiate into neurons, astrocytes and oligodendrocytes. In 2011, StemCells established collaborations with some famous AD research groups in the world to study the therapeutic potential of HuCNS-SC cells in AD. The preliminary results showed HuCNS-SC cells can survive in brain of AD animal models, and StemCells Inc. hopes to advance toward clinical testing of HuCNS-SC cells in AD patients. Different from StemCell, the NeuralStem has devoted to isolate and expand regionally specific cells. NeuralStem has announced their preclinical study by using engineered human cortex-derived neural stem cells for AD. The neural stem cells can secrete human insulin-like growth factor 1 and rescue spatial learning and memory deficits after transplanted into an animal model of AD.

Although the stem cell-based therapy for AD has commenced, it is still in the earlier stage compared to development of stem cell therapy candidates and technological platforms for other brain or spinal cord disorders. The clinical trial of human ESC-derived oligodendrocyte progenitors from Geron for spinal cord injury has been authorized by FDA, which is a milestone for the stem cell therapy field. The clinical-grade neural stem cell CTX from ReNeuron has been approved for the phase II clinical trials for treatment of stroke. With the continued progress and mutual involvement of scientific, ethical, regulatory, translational and clinical fields, more and more companies start to involve in and make efforts to develop rational stem cell-based therapies for the treatment of AD.

The progresses in both scientific and industrial fields have provided proof of principle that the stem cells can induce marked improvements in AD animals, and may lead to valuable improvement in clinical trials. But, there are still different kinds of challenges ahead before stem cell-based therapy can move to the clinic as a treatment for AD: a) the standard preparation of cells suitable for transplantation has not been achieved, b) the present cell replacement procedures are far from optimal, c) it is still unclear the mechanism underlying symptomatic relief upon cell transplantation, d) it still needs to be determined the chronic inflammatory and immune response due to the cell grafts. Even though, the continued understanding in cell biology of donor cells, the advances in stem cell transplantation and involvement of clinical activities will facilitate the development of stem cell-based therapeutic strategies for AD, and the perspective of stem cell therapy for the treatment of AD is expected.

Competing interests
The authors declare that they have no competing interests.

Author contributions
CM wrote the manuscript. NJ conceived the whole study and supervised the manuscript writing. Both authors read and approved the final manuscript.

Acknowledgement
This work was supported in part by the “Strategic Priority Research Program” of the Chinese Academy of Sciences (XDA01010201), National Natural Science Foundation of China (91219933, 31430058), National Key Basic Research and Development Program of China (2014CB964904, 2015CB964500).

Received: 2 March 2015 Accepted: 14 April 2015
Published online: 28 April 2015

Reference
1. Selkoe DJ. Translating cell biology into therapeutic advances in Alzheimer’s disease. Nature. 1999;399:A23–A31.
2. Hebert LE, Scherr PA, Bienias JL, Bennett DA, Evans DA. Alzheimer disease in the US population: prevalence estimates using the 2000 census. Arch Neurol. 2003;60:1119–22.
3. Kim JH, Auerbach JM, Rodriguez-Gomez JA, Velasco I, Gavin D, Lumelsky N, et al. Dopamine neurons derived from embryonic stem cells function in an animal model of Parkinson’s disease. Nature. 2002;418:504–6.
4. Kriks S, Shim JW, Piao J, Ganat YM, Wakeman DR, Xie Z, et al. Dopamine neurons derived from human ES cells efficiently engraft in animal models of Parkinson’s disease. Nature. 2011;480:547–51.
5. Burton-Jones M, Kitazawa M, Martinez-Costa H, Castello NA, Muller FJ, Loring JD, et al. Neural stem cells improve cognition via BDNF in a transgenic model of Alzheimer disease. Proc Natl Acad Sci U S A. 2009;106:13594–9.
6. Moghadam FH, Alaie H, Karbalaie K, Tanhaei S, Nam Esfahani MH, Baharvand H. Transplantation of primed or unprimed mouse embryonic stem cell-derived neural precursor cells improves cognitive function in Alzheimerian rats. Differentiation. 2009;78:59–68.
7. Lu LF, Lee JH, Lee H, Shin JW, Carter JE, Sakamoto T, et al. The therapeutic potential of human umbilical cord blood-derived mesenchymal stem cells in Alzheimer’s disease. Neurosci Lett. 2010;481:30–5.
8. Lee JH, Lee JY, Kim D, Schuchman EH, Carter JE, Bae JS. Intracerebral transplantation of bone marrow-derived mesenchymal stem cells reduces amyloid-beta deposition and rescues memory deficits in Alzheimer’s disease mice by modulation of immune responses. Stem Cells. 2010;28:329–43.
9. Pihlaja R, Koistinaho J, Malm T, Sikkila H, Vainio S, Koistinaho M. Transplanted astrocytes internalize deposited beta-amyloid peptides in a transgenic mouse model of Alzheimer’s disease. Glia. 2000;35:154–63.

10. Bartus RT, Dean 3rd RL, Beer B, Lippa AS. The cholinergic hypothesis of geriatric memory dysfunction. Science. 1982;217:408–14.

11. Geula C, Mesulam MM. Cholinergic systems in Alzheimer’s disease. In: Terry RD et al., editors. Alzheimer disease. 2nd ed. Philadelphia, PA: Lippincott, Williams and Wilkins; 1999. p. 69–292.

12. Fujiwara N, Shimizu J, Takai K, Arimitsu N, Saito A, Kono T, et al. Restoration of spatial memory dysfunction of human APP transgenic mice by transplantation of neuronal precursors derived from human IPS cells. Neurosci Lett. 2013;557:29–34.

13. Mesulam MM, Mufson EJ, Wainer BH, Levey AI. Central cholinergic pathways in the rat: an overview based on an alternative nomenclature (Ch1-Ch6). Neuroscience. 1983;10:1185–201.

14. Mesulam MM, Geula C. Nucleus basalis (Ch4) and cortical cholinergic innervation in the human brain: observations based on the distribution of acetylcholinesterase and choline acetyltransferase. J Comp Neurol. 1982;216:40–60.

15. Drachman DA, Leavitt J. Human memory and the cholinergic system. A relationship to aging? Arch Neurol. 1974;30:113–21.

16. Drachman DA, Sahakian BJ. Memory and cognitive function in the elderly. A preliminary trial of physostigmine. Arch Neurol. 1980;37:674–5.

17. Coyle JT, Price DL, Delong MR. Alzheimer’s disease: a disorder of cortical cholinergic innervation. Science. 1982;219:1184–90.

18. Peason RC, Softinov MV, Cuello AC, Powell TP, Eckenstein F, Esiri MM, et al. Persistence of cholinergic neurons in the basal nucleus in a brain with senile dementia of the Alzheimer’s type demonstrated by immunohistochemical staining for choline acetyltransferase. Brain Res. 1983;289:375–9.

19. Whitehouse PJ, Price DL, Struble RG, Clark AW, Coyle JT, Delon MR. Alzheimer’s disease and senile dementia: loss of neurons in the basal forebrain. Science. 1982;215:1237–9.

20. Whitehouse PJ, Struble RG, Clark AW, Price DL. Alzheimer disease: plaques, tangles, and the basal forebrain. Ann Neurol. 1982;12:494.

21. Pappas BA, Bayley P, Bui BK, Hansen LA, Thal LJ. Choline acetyltransferase activity and cognitive domain scores of Alzheimer’s patients. Neurobiol Aging. 2000;21:11–7.

22. Perry EK, Tomlinson BE, Blessed G, Bergmann K, Gibson PH, Perry RH. Correlation of cholinergic abnormalities with senile plaques and mental test scores in senile dementia. Br Med J. 1978;2:1457–9.

23. Krusel B, Michel PP, Schwabe JS, Hefyi F. Selective and nonselective stimulation of central cholinergic and dopaminergic development in vitro by nerve growth factor, basic fibroblast growth factor, epidermal growth factor, insulin and the insulin-like growth factors I and II. J Neurosci. 1990;10:558–70.

24. Krusel B, Winslow JW, Rosenthal A, Burton LE, Seid DP, Nikolics K, et al. Promotion of central cholinergic and dopaminergic neuron differentiation by brain-derived neurotrophic factor but not neurotrophin 3. Proc Natl Acad Sci U S A. 1991;88:961–5.

25. Lopez-Coviella I, Follette MT, Mello T, Kovacheva VP, Slack BE, Diesl V, et al. Bone morphogenetic protein 9 induces the transcriptome of basal forebrain cholinergic neurons. Proc Natl Acad Sci U S A. 2005;102:10698–9.

26. Elbaty Y, Gan L. The LM-homeobox gene lsx-1 is required for the development of restricted forebrain cholinergic neurons. J Neurosci. 2006;26:3291–7.

27. Furusho M, Ono K, Takebayashi H, Masahira N, Kagawa T, Ikeda K, et al. Bone morphogenetic protein 9 induces the transcriptome of basal forebrain cholinergic neurons. Proc Natl Acad Sci U S A. 2006;103:7959–64.

28. Mori T, Yuxing Z, Takechi M, Hagiwara S, et al. The LIM homeobox gene, L3/Lhx8, is necessary for proper development of basal forebrain cholinergic neurons. Eur J Neurosci. 2004;19:3129–31.

29. Sussel L, Marin O, Kimura S, Rubenstein JL. Loss of Nkx2.1 homeobox gene expression results in a ventral to dorsal molecular reorganization within the basal telencephalon: evidence for a transformation of the pallidum into the eminence. Development. 1999;126:3359–70.

30. Bissonnette C, Lyass L, Bhattacharyya BJ, Belmadani A, Miller RJ, Kessler JA. The controlled generation of functional basal forebrain cholinergic neurons from human embryonic stem cells. Stem Cells. 2011;29:802–11.

31. Liu Y, Weick JP, Liu H, Krenck R, Zhang X, Ma L, et al. Medial ganglionic eminence-like cells derived from human embryonic stem cells correct learning and memory deficits. Nat Biotechnol. 2013;31:440–7.

32. Huhn S. Cellular Therapy for CNS Disorders: A Translational Perspective. 2009. http://www.stemcellsciences.com/Therapeutic-Programs/CNS-Program.htm