Dogs and Humans Share a Common Susceptibility Gene SRBD1 for Glaucoma Risk

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Abstract

Glaucoma is a degenerative optic neuropathy that is associated with elevated intraocular pressure. Primary open angle glaucoma is the most common type of glaucoma in canines, and its highest incidence among dog breeds has been reported in Shiba-Inus, followed by Shih-Tzus. These breeds are known to have an abnormal iridocorneal angle and dysplastic prectinate ligament. However, the hereditary and genetic backgrounds of these dogs have not yet been clarified.

In this study, we investigated the association between polymorphisms of the glaucoma candidate genes, SRBD1, ELOVL5, and ADAMTS10, and glaucoma in Shiba-Inus and Shih-Tzus. We analyzed 11 polymorphisms in these three genes using direct DNA sequencing. Three SRBD1 SNPs, rs8655283, rs22018514 and rs22018513 were significantly associated with glaucoma in Shiba-Inus, while rs22018513, a synonymous SNP in exon 4, showed the strongest association ($P=0.00039$, OR = 3.03). Conditional analysis revealed that rs22018513 could account for most of the association of these SNPs with glaucoma in Shiba-Inus. In Shih-Tzus, only rs9172407 in the SRBD1 intron 1 was significantly associated with glaucoma ($P=0.0014$, OR = 5.25). There were no significant associations between the ELOVL5 or ADAMTS10 polymorphisms and glaucoma in Shiba-Inus and Shih-Tzus. The results showed that SRBD1 polymorphisms play an important role in glaucoma pathology in both Shiba-Inus and Shih-Tzus. SRBD1 polymorphisms have also been associated with normal- and high-tension glaucomas in humans. Therefore, SRBD1 may be a common susceptibility gene for glaucoma in humans and dogs. We anticipate that the nucleotide sequencing data from this study can be used in genetic testing to determine for the first time, the genetic status and susceptibility of glaucoma in dogs, with high precision. Moreover, canine glaucoma resulting from SRBD1 polymorphisms could be a useful animal model to study human glaucoma.

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Introduction

Glaucoma is a degenerative optic neuropathy comprising a group of eye disorders, including visual field defects, progressive loss of retinal ganglion cells, and degeneration of optic nerve axons, and is frequently associated with elevated intraocular pressure (IOP) [1]. Glaucoma is classified into three types: primary open angle glaucoma (POAG), primary closed angle glaucoma (PCAG), and primary congenital glaucoma (PCG) [1]. POAG is the most common type of glaucoma, and is usually associated with high IOP. Japanese populations, however, have a substantially higher incidence of normal tension glaucoma (NTG), a form of glaucoma in which optic nerve damage occurs even though the IOP is not elevated [2,3].

It is well known that glaucoma is genetically heterogeneous and many genes, such as CYP1B1, MYOC, OPTN, and OPTC, are linked to POAG and PCG in humans and/or dogs [4–9]. Recently, the Normal Tension Glaucoma Genetic Study Group of the Japan Glaucoma Society performed a genome-wide association study with NTG patients and controls in a Japanese population [2]. The study identified two new susceptibility genes for NTG, SRBD1 and ELOVL5, with strong statistical significance. Similarly, Mabuchi et al. also reported the association of an SRBD1 polymorphism with Japanese POAG patients, including late-onset NTG and high tension glaucoma [10].

Canine primary glaucoma has been investigated since almost 50 years ago [11], and high incidences have been reported in Beagles [12–14], Welsh Springer Spaniels [15], and other breeds [16,17]. Recent study reported the Gly661Arg variant in ADAMTS10 as the candidate disease-causing variant for POAG Beagles [12]. Kato et al. investigated the incidence of canine POAG, and reported that Shiba-Inus exhibited the highest incidence of glaucoma among 29 breeds, followed by Shih-Tzus [18]. They also reported that an abnormal iridocorneal angle and dysplastic prectinate ligament were associated with a high incidence of glaucoma in Shiba-Inus and Shih-Tzus. However, the hereditary and genetic backgrounds of glaucoma in these dogs have not yet been clarified.

In this study, to verify recent genetic findings, we investigated the association between glaucoma in Shiba-Inu and Shih-Tzu
Results

The average ages of glaucoma cases and controls were 8.5 ± 2.9 and 10.0 ± 3.0 years, respectively, in Shiba-Inu dogs. Those in Shih-Tzu dogs were 9.2 ± 2.0 and 10.1 ± 2.5 years old, respectively. We genotyped 11 polymorphisms in \textit{SRBD1}, \textit{ELOVL5}, and \textit{ADAMTS10} in 98 Shiba-Inus and 67 Shih-Tzu dogs using the direct DNA sequencing method (Table 1).

Table 2 shows the details of five single nucleotide polymorphisms (SNPs) in \textit{SRBD1}, including their genomic locations and allele frequencies in Shiba-Inus and Shih-Tzus. In Shiba-Inus, the most statistically significant association was observed for rs22018513 (\(P = 0.00039\); the G allele of rs22018513 had a 3.03-fold (95% CI = 1.62–5.65) increased risk of glaucoma, with a frequency of 78.6% in cases vs. 54.8% in controls. Significant associations were also observed for rs8655283 and rs22018514 in Shiba-Inus; the frequencies of the T allele of rs8655283 and the G allele of rs22018514 were significantly greater among glaucoma cases than among controls (rs8655283, 37.5% vs. 21.4%; rs22018514, 41.1% vs. 21.4%, \(P = 0.0037\), OR = 2.56, 95% CI = 1.34–4.36). In Shih-Tzus, we observed a significant association for rs9172407 (\(P = 0.0014\)) and the G allele of rs9172407 had a 5.25-fold (95% CI = 1.76–15.63) increased risk of glaucoma (25.9% in cases vs. 9.7% in controls). Significant associations were also not reached statistically significant for either breed.

Discussion

The aim of the present study was to assess the potential associations of polymorphisms in the candidate genes \textit{SRBD1}, \textit{ELOVL5}, and \textit{ADAMTS10}, with the development of canine glaucoma. To this end, we genotyped 11 polymorphisms of these genes in two breeds of dogs with cases of glaucoma or without, as controls. Here we report that the \textit{SRBD1} polymorphisms exhibited significant association with canine glaucoma, while the \textit{ELOVL5} and \textit{ADAMTS10} polymorphisms that were examined in this study were not associated with canine glaucoma. In Shiba-Inus, the strongest association with glaucoma in \textit{SRBD1} was observed at rs22018513, which is a synonymous SNP in exon 4. Two other SNPs, rs8655283 and rs22018514, were associated with glaucoma in Shiba-Inus, as well as rs22019922, also did not achieve statistically significant associations with glaucoma in Shih-Tzu. However, the odds-ratios of these variants in Shih-Tzus were suggestive of an association, with the T allele of rs8655283, the G allele of rs22018514, and the A allele of rs22019922, each having a 2.43 or 2.49-fold increased risk of glaucoma.

Table 1. Primer pairs for PCR of glaucoma-related genes.

| Gene       | Allele            | 5'-3' Forward | 5'-3' Reverse |
|------------|-------------------|---------------|---------------|
| SRBD1      | rs22019922        | TGTGTTGTGTTGTCAGCAAGT | TCACTCTTTTCCTCATCTCTC |
|            | rs8655283         | TTAGGATGAAACATCGGAGAC | TGGCGGATTATTGAACTAAC |
|            | rs22018513, rs22018514 | GCTATTGCTGATGTTGATTTG | TGAAGCGGAGGTGGCAAGG |
|            | rs9172407         | GTGAACCTGAAATGGCAAA | TTAACAGCTTCTCCGTCTCC |
| ELOVL5     | rs22226301        | AGTATGGTGTTGTCAGCAAGT | AGCAACGGCAGATAGTCTC |
|            | rs9194033         | AGTATGGTGTTGTCAGCAAGT | GCTCAGGTCAATGCAAGAG |
|            | rs22202438        | CATGCTGAACTCCTGTTGAG | GCTGGTCTGGATGAGTCTA |
|            | rs8643563         | AATGTATGGTGTTGAGAACAA | ACCACAGAGACCTCTACAA |
|            | rs22194174        | AATGTATGGTGTTGAGAACAA | ACCACAGAGACCTCTACAA |
| ADAMTS10   | Gly661Arg (rs6097365 G>A) | CACAGACAGCAAGGGAGT | GGGTTGGAAGTGGCAAGAG |

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Table 2. Primer pairs for PCR of glaucoma-related genes.

| Gene     | Allele            | 5'-3' Forward | 5'-3' Reverse |
|----------|-------------------|---------------|---------------|
| SRBD1    | rs22019922        | TGTGTTGTGTTGTCAGCAAGT | TCACTCTTTTCCTCATCTCTC |
|          | rs8655283         | TTAGGATGAAACATCGGAGAC | TGGCGGATTATTGAACTAAC |
|          | rs22018513, rs22018514 | GCTATTGCTGATGTTGATTTG | TGAAGCGGAGGTGGCAAGG |
|          | rs9172407         | GTGAACCTGAAATGGCAAA | TTAACAGCTTCTCCGTCTCC |
| ELOVL5   | rs22226301        | AGTATGGTGTTGTCAGCAAGT | AGCAACGGCAGATAGTCTC |
|          | rs9194033         | AGTATGGTGTTGTCAGCAAGT | GCTCAGGTCAATGCAAGAG |
|          | rs22202438        | CATGCTGAACTCCTGTTGAG | GCTGGTCTGGATGAGTCTA |
|          | rs8643563         | AATGTATGGTGTTGAGAACAA | ACCACAGAGACCTCTACAA |
|          | rs22194174        | AATGTATGGTGTTGAGAACAA | ACCACAGAGACCTCTACAA |
Table 2. Association analysis for five polymorphisms in the SRBD1 gene region for Shiba-Inu and Shih-Tzu dog breeds.

| SNP ID  | Chr. Position (CanFam2.0) | Allele | SNP Type  | Risk Allele | Breed | N   | Cases | Controls | Cases | Controls | Risk Allele Frequency (%) | P     | OR (95% CI) |
|---------|---------------------------|--------|-----------|-------------|--------|------|-------|----------|-------|----------|---------------------------|-------|-------------|
| rs22019922 | 10 50924623 | A/C   | Intron    | A           | Shiba-Inu | 56   | 42   | 8.9   | 7.1   | 0.65 | 1.27 (0.44–3.66) |
|         |              |       |           |             | Shih-Tzu  | 27   | 40   | 96.3 | 91.3  | 0.25 | 2.49 (0.50–12.49) |
|         |              |       |           |             | Overall   | 83   | 82   | 0.40  |       | 1.59 (0.66–3.80) |
| rs8655283 | 10 50989281 | C/T   | Intron    | T           | Shiba-Inu | 56   | 42   | 37.5 | 21.4  | 0.016 | 2.20 (1.15–4.20) |
|         |              |       |           |             | Shih-Tzu  | 27   | 40   | 92.6 | 83.8  | 0.13 | 2.43 (0.75–7.89) |
|         |              |       |           |             | Overall   | 83   | 82   | 0.0068 |       | 2.25 (1.28–3.97) |
| rs22018514 | 10 51,049,600 | C/G | Non-synonymous | G          | Shiba-Inu | 56   | 42   | 41.1 | 21.4  | 0.0037 | 2.56 (1.34–4.86) |
|         |              |       |           |             | Shih-Tzu  | 27   | 40   | 92.6 | 83.8  | 0.13 | 2.43 (0.75–7.89) |
|         |              |       |           |             | Overall   | 83   | 82   | 0.0018 |       | 2.52 (1.43–4.44) |
| rs22018513 | 10 51,049,604 | A/G | Synonymous | G          | Shiba-Inu | 56   | 42   | 78.6 | 54.8  | 0.00039 | 3.03 (1.62–5.65) |
|         |              |       |           |             | Shih-Tzu  | 27   | 40   | 94.4 | 93.8  | 0.87 | 1.13 (0.26–4.95) |
|         |              |       |           |             | Overall   | 83   | 82   | 0.0015 |       | 2.59 (1.46–4.61) |
| rs9172407 | 10 51062753 | A/G   | Intron    | G           | Shiba-Inu | 56   | 42   | 8.9  | 6.0   | 0.44 | 1.55 (0.51–4.71) |
|         |              |       |           |             | Shih-Tzu  | 27   | 40   | 25.9 | 6.3   | 0.0014 | 5.25 (1.76–15.63) |
|         |              |       |           |             | Overall   | 83   | 82   | 0.0074 |       | 2.90 (1.34–6.26) |

OR, odds ratio; CI, confidence interval.
Overall P values and ORs for meta-analysis were calculated using the Mantel-Haenzel method.
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Figure 1. Linkage disequilibrium (LD) plot of five SNPs of the SRBD1 gene. A) LD structure in Shiba-Inus. B) LD structure in Shih-Tzus. The D’ value and r² value (in parentheses) corresponding to each SNP pair are expressed as a percentage and shown within the respective square. The color scheme is based on D’ and LOD score values: bright red (LOD ≥2 and D’ = 1); shades of pink/red (LOD ≥2 and D’ < 1); blue (LOD <2 and D’ = 1); white (LOD <2 and D’ < 1).
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Table 3. Conditional logistic regression analysis of rs8655283, rs22018514 and rs22018513 in the SRBD1 gene for Shiba-Inus.

| SNP ID   | Risk Allele | Model  | P* | Covariates | rs8655283 | rs22018514 | rs22018513 |
|----------|-------------|--------|----|------------|-----------|------------|------------|
| rs8655283 | T           | Additive | 0.021 | –          | 0.92      | 0.15       |
| rs22018514 | G           | Additive | 0.00066 | 0.13       | –         | 0.10       |
| rs22018513 | G           | Additive | 0.00025 | 0.0010     | 0.0021    | –          |

*P values for each SNP under the recessive, additive, or dominant model that provided the best fit by logistic regression analysis. The lowest P value was selected as the best fit model. The indicated model showed the lowest P value for each SNP.

**P values adjusted for each SNP under the indicated model by conditional logistic regression analysis.

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Table 4. Association analysis for six polymorphisms in the ELOVL5 and ADAMTS10 gene regions for Shiba-Inu and Shih-Tzu dog breeds.

| SNP ID   | Chr. (CanFam2.0) | Gene | Allele | SNP Type | Risk Allele* | Breed | N | Risk Allele Frequency (%) | P | OR (95% CI) |
|----------|------------------|------|--------|----------|--------------|-------|---|--------------------------|---|-------------|
| rs22226301 | 12 20733716      | ELOVL5 | C/T   | 3'UTR    | T            | Shiba-Inu | 56 | 0.0 0.0 0.0 0.0 - -       |   |             |
| rs9194033  | 12 20739417      | ELOVL5 | A/G   | Intron   | G            | Shiba-Inu | 56 | 33.9 26.2 0.25 1.45 (0.78–2.70) |   |             |
| rs22202438 | 12 20743516      | ELOVL5 | A/G   | Synonymous | G            | Shiba-Inu | 56 | 35.7 28.6 0.29 1.39 (0.75–2.56) |   |             |
| rs8643563  | 12 20744701      | ELOVL5 | (-)/T | Frameshift coding | T          | Shiba-Inu | 56 | 0.0 0.0 0.0 0.0 - -       |   |             |
| rs22194174 | 12 20749077      | ELOVL5 | A/C   | Intron   | A            | Shiba-Inu | 56 | 0.0 0.0 0.0 0.0 - -       |   |             |
| Gly661Arg  | 20 56097365      | ADAMTS10 | A/G | Non-synonymous | A           | Shiba-Inu | 56 | 0.0 0.0 0.0 0.0 - -       |   |             |

OR, odds ratio; CI, confidence interval.
Overall P values and ORs for meta-analysis were calculated using the Mantel-Haenzel method.

*Risk allele is for Shiba-Inu dogs.

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region (http://asia.ensembl.org/Canis_familiaris/Transcript/Exons?db=core&g=ENSCAFG00000002554;r=10:47923593-47924593; t=ENSCAFG000000049409). SRBD1 transcript is reportedly expressed in the retinal ganglion cell and neuroblast layers in neonatal mouse tissue [2], but its localization in dogs remains unknown.

The present study found that a synonymous SNP (rs22018513) and intronic SNP (rs9172407) in SRBD1 were associated with glaucoma in Shiba-Inus and Shih-Tzus, respectively. Synonymous and intronic polymorphisms can significantly affect gene expression by various mechanisms and lead to the development of disease [22–24]. We did not compare SRBD1 mRNA levels in normal and affected dogs; however, a similar experiment has been conducted in humans [2]. Results from that study showed a significant correlation between increased SRBD1 expression and the NTG-associated risk allele of intronic SNP. These results suggest that rs22018513 and rs9172407 in canine SRBD1 could cause enhanced SRBD1 expression. Reports show that SRBD1 is indirectly involved in cell growth, general protein synthesis, induction of apoptosis, and maintaining homeostasis [2]. Therefore, we hypothesize that enhanced expression, leading to increased activity of SRBD1, which could induce apoptosis, could result in retinal ganglion cell death during the development of glaucoma.

ELOVL5 is a fatty acid condensing enzyme involved in the biosynthesis of long-chain polyunsaturated fatty acids [25], and is one of the candidate genes for retinitis pigmentosa [26]. The Normal Tension Glaucoma Genes Study Group of the Japan Glaucoma Society reported ELOVL5 as a new susceptibility gene for human NTG [2]. However, the present study did not show any significant association of ELOVL5 polymorphisms with canine glaucoma. The difference between our results and those reported by the study group may be due to the different types of glaucoma that were studied (human NTG without IOP elevation, and canine glaucoma with IOP elevation, respectively). In contrast, SRBD1 polymorphisms were associated with canine glaucoma and human glaucoma independent of IOP, suggesting that SRBD1 polymorphisms may affect a common disease condition in canine and human glaucoma. Therefore, detection of any common phenotypes in these glaucoma studies [2,10] is important because it will help to clarify how SRBD1 affects the development of glaucoma. Moreover, canine glaucoma resulting from SRBD1 polymorphisms could be used as an excellent genetic animal model for human glaucoma and contribute significantly to the development of novel diagnostic and therapeutic options for glaucoma.

Kato, et al., reported that the Beagle breed has the fourth-highest incidence of canine glaucoma, after Shiba-Inu, Shih-Tzu, and American Cocker Spaniel breeds [18]. Kuchety et al. recently reported that the Gly661Arg variant (56097365 G>A) of ADAMTS10 in Beagles with POAG is a candidate, predictive gene allele for canine POAG [12]. We did not observe an association between this 56097365 G>A variant and glaucoma in Shiba-Inus or Shih-Tzus, possibly because of breed-specific allelic differences between Beagles and Shiba-Inus or Shih-Tzus. More recently, Kuchety, et al., reported that the Gly661Arg variant was not found in any of the other dog breeds analyzed. (Shiba-Inu, Shih-Tzu, American Cocker Spaniels, Chihuahua, Australian Cattle Dog, Jack Russell Terrier, Jindo, Siberian Husky, and Yorkshire Terrier), suggesting that this allele is Beagle-specific, and that other genes may be associated with glaucoma in other breeds [27]. However, ADAMTS10 may be still a candidate gene for glaucoma in dogs, including Shiba-Inu and Shih-Tzu, because other ADAMTS10 variants have yet to be investigated for their association with canine glaucoma. The Ensembl database (http://asia.ensembl.org/index.html) shows 16 genetic polymorphisms in the canine ADAMTS10 gene region. Since the dog genome information is still incomplete, it is predicted that more even polymorphisms exist in the canine ADAMTS10 gene region. Therefore, it is necessary to perform a comprehensive genetic analysis of the region and clarify whether ADAMTS10 is a candidate gene for glaucoma not only in Beagles, but also in other breeds.

There are no genetic tests currently available to assist in glaucoma diagnosis, identification of people at risk, initiation of treatment, and timing of surgical intervention. We performed the present SNP analysis of candidate genes in Shiba-Inu and Shih-Tzu dog breeds for the possibility to help develop diagnostic, genetic analyses for glaucoma risk factors. However, the mechanism by which these genes contribute to the development of glaucoma remains to be determined. Future studies are expected to examine the roles of SRBD1 in humans and dogs, in an effort to determine whether genetic testing might not only help predict whether someone will develop glaucoma, but may also, perhaps, be a valuable prognostic factor for the clinical course of the disease, and/or predictive factor for its treatment. Despite new and improving diagnostic and therapeutic options for glaucoma, blindness resulting from glaucoma remains a major public health problem. These future experiments will help to optimize glaucoma treatment.

Materials and Methods

Ethics Statement

This study was performed as part of research approved by the Ethical Committee of Azabu University (Permit Number: 110408-2). Informed written consent was obtained from each dog owner. All procedures in this study were conducted in accordance with the Guide for the Care and Use of Laboratory Animals of Azabu University.

Diagnosis of Glaucoma

98 Japanese Shiba-Inu dogs and 67 Shih-Tzu dogs were recruited from the Veterinary Teaching Hospital at Azabu University. All dogs received complete ophthalmologic examinations using a hand-held slit-lamp biomicroscope (SL-14, Kowa, Tokyo, Japan), indirect ophthalmoscopy, and tonometry. After the application of topical anesthesia (oxybuprocaine hydrochloride, Santen, Osaka, Japan), IOP was measured by tonometry using the Tono-Pen XL (Mentor O&O Inc., Norwell, MA). 42 Shiba-Inus and 40 Shih-Tzus were diagnosed as normal (<25 mmHg IOP). 56 Shiba-Inus and 27 Shih-Tzus had elevated IOP (>25 mmHg) in at least one eye, and were diagnosed with glaucoma. Since glaucoma is a late-onset disorder, we did not recruit dogs younger than four years in the control group, in an attempt to exclude potential glaucomatous dogs.

DNA Preparation

Genomic DNA from glaucomatous and normal dogs was collected from peripheral blood and purified using a DNA whole blood spin kit (Fuji Film, Tokyo, Japan). The purity and concentration of DNA were examined using GeneQuant Pro (GE Healthcare, Cambridge, UK).

Determination of DNA Sequences

In the SRBD1 and ELOVL5 regions, a total of ten polymorphisms were selected to cover the entire gene regions (rs22019922, rs8655283, rs22018514, rs22018513 and rs9172407 in SRBD1; rs22226301, rs9194033, rs22202438, rs8643363 and rs22194174 in ELOVL5) (Table 2,4). In the ADAMTS10 gene
region, we selected the Gly661Arg variant (56097365 G>A) for analysis (Table 4). PCR primer pairs listed in Table 1 were used to amplify regions containing the SNPs mentioned above. The PCR products were electrophoretically separated on a 1% agarose gel, and the PCR product bands were cut and frozen at −20°C in Tris-EDTA buffer. The frozen samples were thawed, homogenized, and centrifuged at 15,000 g for 5 minutes. The supernatants were subjected to cycle sequencing using the Big Dye terminator sequencing kit (Life Technologies, Foster City, CA). The sequence data were analyzed using Sequence Scanner v. 1.0 (Life Technologies) and GENETYX-WIN v. 4.0 (Genetyx, Tokyo, Japan). We did not find any novel DNA

Statistical Analysis
Hardy-Weinberg equilibrium was tested for each SNP among glaucomatous and normal dogs. Differences in allele frequency

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