Pathology associated with larval *eustrongylides* sp. (nematoda: Dioctophymatoidea) infection in *galaxias maculatus* (actinopterygii: Galaxiidae) from patagonia, Argentina

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**ABSTRACT**

Helminth infections within tissues tend to be subjected to a host response that can include encapsulation and melanization to isolate the parasite. The effectiveness of this response depends on the host species. During a survey of parasites of the native fish, *Galaxias maculatus*, we found conspicuous, strongly melanized exterior cysts located in the caudal peduncle of the fish. Dissection of these cysts exposed larval nematodes whose morphomertic features allowed their identification as *Eustrongylides* sp. Species of this genus are distributed worldwide. *Galaxias maculatus* was previously reported as second intermediate host to *Eustrongylides* sp. larvae, and the aquatic bird *Podiceps major* was cited as definitive host of *Eustrongylides tubifex* in Andean Patagonian lakes. The site of infection in the fish, and the host’s response are unique among infections by larvae of *Eustrongylides* sp. in fishes, so the objective of this study was to describe the histological injury caused by larvae of the genus *Eustrongylides* parasitizing *G. maculatus*. Samples of fish were taken near the shore of Gutiérrez lake, in northwestern Patagonia. Some larval morphometric features were described to confirm the identity of larvae to generic level. Histopathological analysis (tissues sectioned at 5–7 µm, stained with Hematoxylin and Eosin, and Masson’s Trichrome) of the caudal peduncle revealed almost complete disappearance of the epaxial musculature due to compressive atrophy and a chronic inflammatory response, associated mainly with a dense fibrotic capsule and an intense melanic deposit. This is the first description of the histopathology of an external cyst caused by *Eustrongylides* sp. larvae in fishes.

1. Introduction

Parasitic infections in fish can cause changes in behavior, vulnerability to predators, and a reduced capacity to resist stress (Coyner et al., 2001; Moore, 2002; Poulin, 2010; Hammond-Tooke et al., 2012). The presence of a parasite generates a host strategy of inactivation or elimination through cellular and humoral responses, which range from acute or chronic inflammation to severe necrosis (Feist and Longshaw, 2008; Mitchell et al., 2009). Helminth infections within tissues tend to be subjected to a host response that can include encapsulation and melanization to isolate the parasite; the effectiveness of this response depends on the fish species (Feist and Longshaw, 2008).

*Galaxias maculatus* (Jenyns, 1842) is a small prey fish with an exceptionally disjunct distribution, occurring in Australia, Tasmania, Lord Howe Island, New Zealand, Chatham Island, and the south of Chile and Argentina (McDowall, 2006). This fish was reported as host to *Eustrongylides* spp. in New Zealand (Hine, 1978), Australia (Pollard, 1974; Chapman et al., 2006), and Argentinean Patagonia (Brugni and Viozzi, 1999, 2003). Since the larval forms cannot be identified at species level, they are usually designated as *Eustrongylides* sp. *Brugni and Viozzi* (1999, 2003) found larvae of *Eustrongylides* sp. in the caudal peduncle musculature of *Galaxias maculatus* from Lake Gutiérrez and Lake Moreno, both located in Nahuel Huapi National Park. These larvae were used to perform experimental infections in chicks, obtaining immature adults which showed morphological characteristics similar to *E. tubifex* (Nitzsch, 1819) (Brugni and Viozzi, 1999), in accordance with the finding of an ovigerous female of *E. tubifex* in the proventriculus of *Podiceps major* from Moreno lake (Brugni and Viozzi, 2003).

The life cycle of *Eustrongylides* spp. involve an aquatic oligochaete, a fish, and a piscivorous bird. The larvae migrate from the digestive tract of the fish to the cavity or the musculature of the body wall, and have mostly been reported encysted in thin, white or yellowish-pink fibrous...
collagenous cysts located in mesenteries, fatty tissues, the coelomic cavity, liver, spleen, stomach wall, gonads, around the intestinal tract, and musculature (Paperna, 1974; Cooper et al., 1978; Measures, 1988; Kristmundsson and Helgason, 2007; Mitchell et al., 2009; Mir et al., 2012). In *G. maculatus* from Australia, *Eustrongylides* sp. larvae were reported encysted on the outer surface of the stomach, ovaries, and in the abdominal cavity (Pollard, 1974, Chapman et al., 2006). In Patagonia, the larvae have always been found in conspicuous, external, and strongly melanized cysts on the caudal peduncle of *G. maculatus* (Brugni and Viozzi, 1999, 2003; Viozzi et al., 2009), which is an atypical localization and fish reaction. Previous studies on the histopathology of infection by *Eustrongylides* species in different fish species were carried out mainly on larvae localized in the abdominal cavity (Paperna, 1974; Kennedy and Lie, 1976; Eiras and Rego, 1988; Measures, 1988; Mir et al., 2012; Kaur et al., 2012).

Since the site of infection and the host’s response are unique among infections by larvae of *Eustrongylides* sp. in fishes, the objective of this study was to describe the histological injury caused by larvae located in the caudal peduncle of *G. maculatus*.

2. Materials and methods

Specimens of *G. maculatus* were collected near the shore of Gutiérrez lake (41°10′S–71°24′W), northwestern Patagonia, in January and March 2018. Some fish were examined and the melanized cysts were dissected to observe the larvae. The larvae were fixed in 5% formalin and cleared in Aman’s Lactophenol. Morphometric characteristics were registered for third and fourth-stage larvae in order to confirm the generic identification (sensu Lichtenfels and Pilitt, 1986 and Measures, 1988). All measurements are expressed in micrometers (μm) unless otherwise stated.

Two types of fish from the March sample were selected for histological purposes: 1) specimens without cyst, 2) specimens with cyst. The caudal peduncules of infected and uninfected fish were fixed immediately postmortem in 10% buffered formalin, dehydrated in a graded series of alcohols, embedded in paraffin, sectioned at 5–7 μm, and stained with Hematoxylin and Eosin, and Masson’s Trichrome. Microphotographs were obtained with a Motic BA200 microscope.

3. Results

A total of 600 fish were collected (42–59 mm long), twelve of which were infected in the caudal peduncle by encysted larvae of *Eustrongylides* sp. (prevalence = 2%). Ten fish showed one conspicuous, strongly melanized cyst, while the remaining infected fish had two cysts with only a single larva in each (mean intensity = 1.2) (Fig. 1 A). The cysts measured up to 5 mm in length and up to 3.9 mm in height, representing about 8% of the fish’s total length. The cyst harbored 1 live, reddish larva of the third or fourth stage (Fig. 1 B). Cysts were not found in any other site on the body; however, when the host dies, the larva sometimes leaves the cyst and migrates through the musculature (Fig. 1 C).

All the larvae dissected from the January samples (4) were in the third stage of development, while the larvae dissected from the March sample (6) were in the fourth stage. The longest fourth-stage larva found measured 38 mm (Table 1). The remaining measurements of the third- and fourth-stage larvae are shown in Table 1. The body is dark red; the anterior end is conical and has two circles of labial papillae, with 6 papillae in each circle. The papillae of the exterior circle are larger than those of the inner circle and have wide bases with nipplelike apices, while the papillae of the inner circle have spinelike apices (Fig. 2 A). The posterior extremity is rounded and the anus and genital primordia are located terminally. Three cuticle layers are visible at the posterior extremity of the fourth-stage larvae (Fig. 2 B).

The cross sections of a non-parasitized fish show the hypaxial and epaxial striated musculature intact around the spinal cord (Fig. 3 A). In

![Image](image1.png)

**Table 1** Morphometrics of third- and fourth-stage larval *Eustrongylides* sp. from Galaxias maculatus from Gutiérrez Lake.

|                     | Third-stage larvae | Fourth-stage larvae |
|---------------------|--------------------|---------------------|
|                     | mean   | sd    | min  | max  | mean | sd  | min | max  |
| Total length (mm)   | 16.3   | 1.5   | 15   | 18   | 34.4 | 3.4 | 30  | 38   |
| Width at never ring | 144.0  | 17.9  | 125  | 168  | 171.2| 9.5 | 156 | 182  |
| Papillae of external circle from anterior end | 37.3 | 6.2 | 29 | 43 | 70.5 | 8.7 | 58 | 84 |
| Nerve ring from anterior end | 121.3 | 11.9 | 108 | 137 | 204.5| 13.0| 180 | 216 |
| Buccal cavity length | 72.5   | 9.0   | 60   | 79   | 142.8| 19.8| 120 | 175  |
| Oesophagus length (mm) | 5.2   | 0.7   | 5    | 6    | 12.7 | 2.0 | 10  | 15   |

**Fig. 1.** A. specimens of Galaxias maculatus showing melanized cysts located in the caudal peduncle. Bar = 15 mm; B. larva of *Eustrongylides* sp. emerging from the cyst. Bar = 1.5 mm μm; C. caudal peduncle of Galaxias maculatus showing 2 cysts, and a larva migrating through the musculature. Bar = 2.5 mm.

**Fig. 2.** Fourth-stage larva of *Eustrongylides* sp. from Galaxias maculatus. A. anterior end. Bar = 40 μm. Internal and external labial papillae (arrows); B. caudal extremity of male. Bar = 100 μm. Note three cuticles, outer second stage, middle third stage, and inner fourth stage (arrows).
transverse sections at one end of the cyst, it can be seen that the injury extends through the epaxial muscles (Fig. 3B). The dorsal portion of the vertical septum is deviated from the normal position due to displacement caused by the cyst, which takes the place of the musculature. The melanic zone is distinguishable from normal muscle (Fig. 3B).

The section in the middle of the cyst shows several portions of the worm immersed in liquid, and surrounded by a fibrous cyst larger than the original muscular mass. In addition an almost total disappearance of the epaxial musculature, due to compressive atrophy, is observed (Fig. 3 C). High magnification shows a predominant collagenous cyst surrounding the parasite (Masson’s Trichrome stain), with few inflammatory cells (Fig. 3D). Although the cross sections show a chronic response, the parasites were not affected (Fig. 3E). Within the fibrosis, melanin deposits are observed around necrotic tissue (Fig. 3F). Encapsulation is associated with hemorrhage immediately surrounding the encapsulated parasites. The presence of erythrocytes and melanomacrophagic centers (MMCs) can be observed (Fig. 3G).

4. Discussion

The presence of 2 rings of 6 labial papillae and the rounded posterior extremity with terminal anus and genital primordial, along with the morphometrical features allowed us to assign the larvae to the genus *Eustrongylides* (Lichtenfels and Pilitt, 1986; Measures, 1988). The infection site, shape and measurements of these larvae are similar to those found in previous reports for the same host and environment (Brugni and Viozzi, 1999, 2003).

In *G. maculatus* from Victoria, Australia, Pollard (1974) registered a maximum number of 44 larval *Eustrongylides* sp. (sensu the author, probably *E. gadopsis*) per fish, and indicated inhibition or prevention of egg laying by female fish due to larvae encysted either in the vent region or within the ovary itself. These larvae have usually been observed coiled in somewhat flattened, thin-walled, fibrous connective-tissue cysts attached to various organs within the body cavity of the fish, but most frequently attached to the outer surface of the stomach (Pollard, 1974). In *G. maculatus* from southwestern Australia, Chapman et al. (2006) reported larval *Eustrongylides gadopsis* encysted in the inner body.
wall. In the present study, the site of infection of *G. maculatus* with *Eustrongylides* sp. was invariably the skeletal musculature of the caudal musculature, which is large in size relative to the mosquito body size, and probably produces a physical and metabolic burden that may reduce and inhibit the fish’s ability to avoid capture. The same kind of effect may occur in the case of *G. maculatus*, where the strong encapsulation does not alter the viability of the parasite. The worms can sometimes emerge from the encapsulation and migrate to the musculature, causing hemorrhage in the muscles, as found for *G. maculatus* (Jenyns, 1842) in southwestern Australia. Ecol. Freshw. Fish 15, 559–564.

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