INTRODUCTION

A free radical is a relatively unstable molecule with an atom in its outer orbit having one or more unpaired electrons. The molecule that loses the pair becomes unstable and radical. Therefore, this molecule always tries to find its electron pair by seizing electrons from other molecules\(^1\). Free radicals can damage collagen and elastin, which are a protein that keeps skin moisturized, smooth, flexible, and elastic. Excessive free radical levels trigger various degenerative diseases and conditions in the skin, such as premature aging, wrinkles, erythema, skin cancer, and others\(^3\).

Aging is a process that is a certainty, but premature aging is something that is not expected. The skin's premature aging is characterized by thinning skin, dry skin, wrinkles, and uneven coloration\(^5\). One of the causes of premature aging is free radial\(^6\). One of the ways used to prevent premature aging is to use cosmetics that contain antioxidants. These antioxidants can inhibit the development of oxidation reactions by binding to free radicals and highly reactive molecules to inhibit cell damage\(^7\).

One of the cosmetic dosage forms that have developed lately to prevent premature aging is serum. The serum has the advantage of having a high concentration of active ingredients so that the skin more quickly absorbs the effect, can provide a more relaxing effect, and is easier to spread on the surface of the skin because the viscosity is not too high\(^8\).

Research on antioxidant activity from medicinal plants in serum form has been carried out, including by Mardhiani...
by testing the formulation and stability of the serum from green coffee (Coffea canephora var. Robusta) extract as an antioxidant where the results showed an IC₅₀ value of 68.89 µg/mL, which is classified as a strong antioxidant.

Another plant whose antioxidant activity has been studied is turmeric (Curcuma domestica). Curcuma domestica is a type of spice with nutritious compounds called curcuminoids consisting of curcumin, desmethoxycurcumin, and bisdemethoxycurcumin, which have various important pharmacological activity such as antioxidants.

Curcuma domestica rhizome extract in cosmetics has been widely used, and there is some clinical evidence showing that preparations containing C. domestica are either oral or topical provide benefits to treating various skin diseases and overall skin health. In other studies, it was found that the hexane fraction and ethyl acetate fraction from the ethanol extract of C. domestica rhizome were included in the non-toxic category. Based on studies that have been carried out regarding C. domestica and curcumin’s safety evaluation, it is known that it does not cause toxic effects at doses. Therefore, C. domestica and curcumin can be developed in modern medicine for the therapy of various diseases.

Naturally, at least 1% of the collagen in the human body is lost every year, and at the age of 40, humans do not produce collagen anymore, so the loss of collagen reaches 35-40%. Therefore, collagen from outside the body is needed, which has an anti-aging activity to prevent aging from occurring faster. Collagen has a vital role in the food, biomedical, pharmaceutical, and cosmetic industries; therefore, collagen is needed for the skin.

Research on collagen has been carried out by Budiarti by testing collagen from chicken bone waste (Gallus gallus domesticus) against anti-aging activity in vitro, where the results obtained by stirring for six hours have a smaller particle size (1.34 µm) compared to eight hours (1.80 µm) stirring. Collagen with a size of 1.34 µm showed the best activity, namely antioxidant activity against 2,2-diphenyl-1-picrylhydrazyl (DPPH) of 24.7% and tyrosinase inhibitor of 26.77%. Based on the antioxidant, anti-glycation, and antityrosinase activities of collagen by immersing 0.10 M NaOH and stirring for six hours, it has anti-aging properties.

Antioxidant testing can be done in several ways, one of which is using the DPPH method. The DPPH method is one of the most popular methods because it is practical and sensitive. Antioxidant testing using the DPPH method can be seen from the IC₅₀ value. The research aims to see the effectiveness of the combination between C. domestica rhizome extract and collagen from fish skin in serum by determining the antioxidant using the DPPH method.

MATERIALS AND METHODS

Materials
The materials used were C. domestica rhizome obtained in Pekanbaru and determined at the Botanical Laboratory, Universitas Riau, collagen, ethanol 96% (Merck), methanol (Merck), demineralized water (Brataco), disodium EDTA (Merck), NaCl (Merck), DPPH (Merck), natrosol hydroxyethyl cellulose, glycerin, DMDM hydantoin, and ethoxydiglycol. The instrument used were analytical scales (Boeco), ultrasonic bath (Branson), rotary evaporator (Buchi R-14), micropipette, viscometer (Scorex), microplate (Bertholt Tristar), and microplate reader (Bertholt Tristar).

Methods
Extraction
Curcuma domestica rhizome was blanched by boiling at 100°C for five minutes with a medium of 0.05% citric acid solution. Each treatment sample was subjected to an extraction process of 200 g of macerated samples for 72 hours using 96% ethanol solvent and then sonicated...
using an ultrasonic bath. The solvent of macerate was collected, and it was evaporated using a rotary evaporator until a thick extract was obtained.

**Serum formulation**

The serum made was based on the results of research by Mardhiani. Variations were made to the concentrations of *C. domestica* and collagen extracts, each with varying levels of 0, 0.5, 0.8, and 1.1; as well as 0 and 2, respectively. The serum formulation in this study was presented in Table I.

### Table I. Serum formulation

| Ingredients                  | Concentration (%) | F0  | F1  | F2  | F3  | F4  | F5  |
|------------------------------|-------------------|-----|-----|-----|-----|-----|-----|
| *Curcuma domestica* extract  |                   | -   | -   | 0.5 | 0.5 | 0.8 | 1.1 |
| Collagen                     |                   | -   | 2   | -   | 2   | -   | 2   |
| Natrosol                     |                   | 0.75| 0.75| 0.75| 0.75| 0.75| 0.75|
| Glycerin                     |                   | 10  | 10  | 10  | 10  | 10  | 10  |
| DMDM                         |                   | 0.05| 0.05| 0.05| 0.05| 0.05| 0.05|
| Ethocydiglycol               |                   | 2   | 2   | 2   | 2   | 2   | 2   |
| Demineralized water          |                   | Ad  | Ad  | Ad  | Ad  | Ad  | Ad  |
| Ad                           |                   | 100 | 100 | 100 | 100 | 100 | 100 |

F0 = negative control; F1 = collagen 2%; F2 = ethanol extract 0.5%; F3 = *C. domestica* ethanol extract 0.5% + collagen 2%; F4 = *C. domestica* ethanol extract 0.8% + collagen 2%; F5 = *C. domestica* ethanol extract 1.1% + collagen 2%

**Physical evaluation of serum**

The physical evaluation of serum could be carried out with several tests, including:

1. **Organoleptic test:** The organoleptic test was intended to see the physical appearance of the preparation, which includes shapes, colors, and smells.

2. **Homogeneity test:** Homogeneity checks were carried out using an object-glass, with a certain amount of the serum applied to a piece of glass or other suitable transparent material. The preparation must exhibit a homogeneous arrangement and did not show any coarse grains.

3. **pH test:** pH determination was carried out using a pH meter.

4. **Irritation test:** The test was carried out using a closed patch test on human skin, with 1 g of serum is taken, then applied to the inner arm with a diameter of 2 cm, covered with a bandage, left plastered for 24 hours, and observed symptoms such as redness and itching of the skin. This irritation test was carried out on three panelists for one formula.

5. **Viscosity test:** The serum’s viscosity was measured by placing the sample in a viscometer until the spindle is submerged.

### Antioxidant activity test using DPPH

DPPH solution was made with a concentration of 80 μg. Then, a main standard solution of the sample was made with a 1000 μg/mL concentration. Determination of the serum’s antioxidant activity was carried out using a microplate reader with the DPPH method at a wavelength of 520 nm. The mixture was incubated in a dark place for 30 minutes and then measured its absorbance at a wavelength of 520 nm using 80 μg/mL DPPH 80 μg/mL, while for the blank used 50 mL methanol absolute.

### Data analysis

The absorbance measurements using a microplate reader were used to calculate the percentage of DPPH free radical reduction. The percentage of DPPH free radical reduction was calculated using the following equation:

\[
\% \text{inhibition} = \frac{\text{ABS control} - \text{ABS sample}}{\text{ABS control}} \times 100
\]

ABS control : Absorbance of DPPH + methanol - Absorbance of methanol

ABS sample : (Absorbance of sample) - Absorbance of methanol

### RESULTS AND DISCUSSION

The sample used was fresh *C. domestica* rhizome. The middle part of the clean sample was taken and blanched at 100°C for five minutes. This blanching process was able to increase antioxidant activity by changing fewer active compounds to be active. In the blanching process, it was suspected that there would be degradation of
The finished serum formulation was followed by a physical evaluation test of the preparation. The color and smell test obtained results following Table II. As for the form, all were in gel form. Furthermore, the pH test for serum in the six formulas was still in the skin pH range of 4.5-6.5. If serum has a pH too acidic, it will cause skin irritation. On the other hand, if it is too alkaline, it can cause scaly skin. The homogeneity test indicated that all formulas are homogeneous, marked by the absence of coarse grains at the time of testing. The serum's viscosity test showed that the results still met the serum's viscosity requirements by SNI 16-4399-1996; the standard viscosity value for serum was 6000-50000 cP or 6-50 Pa.s. Viscosity testing aims to determine the viscosity value of a substance. The higher the viscosity value, the higher the substance's viscosity level. Also, viscosity is related to the ability of a liquid to flow. Viscosity is inversely proportional to dispersibility, in which the higher the viscosity, the lower the dispersibility. Viscosity also determines the length of the skin's adhesion supply so that the supply can adhere well. In this study, the lowest viscosity obtained at F0 (base) was 30860 cP; collagen's addition increased the preparation's viscosity, which was 34250 cP. The formula containing a combination of collagen and ethanol extract of C. domestica rhizome had decreased viscosity value, increasing the dispersibility of the preparation. In the irritation test on ten panelists, there were no allergic reactions such as redness, itching, heat, and swelling; it could be seen in Table III.

Antioxidant testing of C. domestica rhizome serum was carried out using the DPPH radical absorption method. DPPH is a purple radical compound that has one unpaired atom. The antioxidant activity is indicated by the DPPH absorption measurement that reacts to antioxidant compounds at a maximum wavelength range of 515-520 nm. The DPPH was a free purple radical molecule that turns into a stable yellow compound caused by the reaction to antioxidants, in which antioxidants give one electron to DPPH so that there was a reduction in DPPH free radicals. The sample's antioxidant activity caused a color change in the DPPH solution, which was initially dark purple to pale yellow and colorless. The test results of the % inhibition
value obtained in the ethanol extract serum formulation of *C. domestica* rhizome with the addition of fish skin collagen were shown in Table IV.

**Table IV.** The % inhibition value of the serum.

| Formula | P1   | P2   | P3   | % Inhibition ± SD |
|---------|------|------|------|------------------|
| F0      | 93.494 | 87.732 | 90.335 | 90.526 ± 1.87    |
| F1      | 70.260 | 70.074 | 68.960 | 60.805 ± 0.94    |
| F2      | 60.967 | 59.108 | 60.223 | 60.124 ± 0.94    |
| F3      | 45.540 | 45.725 | 45.539 | 45.635 ± 3.92    |
| F4      | 34.572 | 38.290 | 36.803 | 36.594 ± 2.89    |

The % inhibition value shows that the higher the extract concentration, the higher the antioxidant activity. At F0 and F1, the % inhibition value increased. This finding shows that the collagen used had a supporting role in increasing antioxidant activity, although only partially. The addition of collagen aims to help prevent premature aging because collagen had anti-aging activity. Collagen with a smaller size would have a higher antioxidant activity36. Furthermore, the F2 test was carried out using only turmeric rhizome extract, where *C. domestica* had various functions, one of which was as an antioxidant. Following Pratiwi and Wardaniati31, one of the natural antioxidants was found in the *C. domestica* plant because *C. domestica* rhizomes contain active compounds with medicinal properties. It was called curcuminoids, which belong to the group of phenolic compounds.

The extract and collagen were combined into F3, F4, and F5 to increase the % inhibition value obtained. At F5 with an extract concentration of 1.1%, it shows % inhibition of 90.526%. A higher % inhibition value results in lower absorbance and high antioxidant activity. It could be said that the extract used was able to inhibit free radicals plus the addition of collagen so that free radical inhibition increased. The % inhibition result increased due to differences in extract concentration and collagen's addition in the formula. This study also used a negative control to determine whether there was an effect of each ingredient used in the manufacture of serum and was also carried out on serum-containing only one active ingredient, which aims to determine which active ingredient in serum had antioxidant activity.

**CONCLUSION**

From this research, it was concluded that the serum formula used the active ingredient of *C. domestica* rhizome extract with the addition of collagen had a % inhibition value that inhibited free radicals. The collagen used affected increasing antioxidant activity apart from the extract. At F5, the concentration of 1.1% extract indicated the highest % inhibition value in inhibiting free radicals, which was 90.526%.

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**REFERENCES**

1. Di Meo S, Venditti P. Evolution of the Knowledge of Free Radicals and Other Oxidants. Oxid Med Cell Longev. 2020;2020:9829176. doi:10.1155/2020/9829176

2. Phaniendra A, Jestadi DB, Periyasamy L. Free Radicals: Properties, Sources, Targets, and Their Implication in Various Diseases. Indian J Clin Biochem. 2015;30(1):11-26. doi:10.1007/s12291-014-0446-0

3. Silva SAM, Michniak-Kohn B, Leonardi GR. An overview about oxidation in clinical practice of skin aging. An Bras Dermatol. 2017;92(3):367-74. doi:10.1590/abd1806-4841.20175481

4. Polšák B, Dahnane R. Free Radicals and Extrinsic Skin Aging. Dermatol Res Pract. 2012;2012:135206. doi:10.1155/2012/135206
5. Zhang S, Duan E. Fighting against Skin Aging. The Way from Bench to Bedside. Cell Transplant. 2018;27(5):29-38. doi:10.1177/0963689717725755

6. Liguori I, Russo G, Curcio F, Bulli G, Aran L, Della-Morte D, et al. Oxidative stress, aging, and diseases. Clin Interv Aging. 2018;13:752-72. doi:10.2147/CIA.S158513

7. Lobo V, Patil A, Phatak A, Chandra N. Free radicals, antioxidants and functional foods: Impact on human health. Pharmacogn Rev. 2010;4(8):118-26. doi:10.4103/0973-7847.70902

8. Garre A, Narda M, Valderas-Martinez P, Piquero J, Granger C. Antiaging effects of a novel facial serum containing L-Ascorbic acid, proteoglycans, and proteoglycan-stimulating tripeptide: ex vivo skin explant studies and in vivo clinical studies in women. Clin Cosmet Investig Dermatol. 2018;11:253-63. doi:10.2147/CCID.S161352

9. Mardhiani YD, Yulianti H, Azhary DP, Rusdiana T. Formulasi dan Stabilitas Sediaan Serum dari Ekstrak Kopi Hijau (Coffea canephora var. Robusta) sebagai Antioksidan. Indones Nat Res Pharm J. 2017;2(2):19-33.

10. Amalraj A, Pius A, Gopi S, Gopi S. Biological activities of curcuminoids, other biomolecules from turmeric and their derivatives – A review. J Trad Complement Med. 2017;7(2):205-33. doi:10.1016/j.jtcme.2016.05.005

11. Vaughn AR, Branum A, Sivamani RK. Effects of Turmeric (Curcuma longa) on Skin Health: A Systematic Review of the Clinical Evidence. Phytother Res. 2016;30(8):1243-64. doi:10.1002/ptr.5640

12. Winarsih W, Wintarsih I, Sulistyawati NP, Wahyudina I. Uji Toksisitas Akut Ekstrak Rimpang Kunyit pada Mencit: Kajian Histopatologis Lambung, Hati dan Ginjal. Jurnal Veteriner Jurnal Kedokteran Hewan Indonesia. 2012;13(4):402-9.

13. Hewlings SJ, Kalman DS. Curcumin: A Review of Its’ Effects on Human Health. Foods. 2017;6(10):92. doi:10.3390/foods6100092

14. Aguirre-Cruz G, León-López A, Cruz-Gómez V, Jiménez-Alvarado R, Aguirre-Alvarez G. Collagen Hydrolysates for Skin Protection: Oral Administration and Topical Formulation. Antioxidants. 2020;9(2):181. doi:10.3390/antiox9020181

15. Bolke L, Schlippe G, Gerß J, Voss W. A Collagen Supplement Improves Skin Hydration, Elasticity, Roughness, and Density: Results of a Randomized, Placebo-Controlled, Blind Study. Nutrients. 2019;11(10):2494. doi:10.3390/nu11102494

16. Budiarti E, Budiarti P, Aristi MA, Batubara I. Kolagen dari Limbah Tulang Ayam (Gallus gallus domesticus) terhadap Aktivitas Anti Aging secara In Vitro. Alchemy Jurnal Penelitian Kimia. 2019;15(1):44-56. doi:10.20961/alchemy.15.1.23046.44-56

17. Molyneux P. The use of the stable free radical diphenylpicrylhydrazyl (DPPH) for estimating antioxidant activity. Songklanarin J Sci Technol. 2004;26(2):211-9.

18. Fitriani L, Saputra F, Melisa M, Zaini E. Studi Awal Sediaan Gel Ekstrak Etanol Kayu Angin (Usnea sp) untuk Penyembuhan Luka Bakar. JSFK Jurnal Sains Farmasi Klinis. 2018;5(2):83-7. doi:10.25077/jsfk.5.2.83-87.2018

19. Sarlina S, Rakaz AR, Tandah MR. Uji Aktivitas Antibakteri Sediaan Gel Ekstrak Daun Sereh (Cymbopogon nardus L. Rendle) terhadap Bakteri Staphylococcus aureus Penyebab Jerawat. Jurnal Farmasi Galenika Galenika J Pharm. 2017;3(2):143-9. doi:10.22487/24428744.0.v.0.8770

20. Almurdani M, Jose C, Teruna HY. Uji Aktivitas Antioksidan dan Toksisitas Ekstrak Akar Tanaman Amaranthus spinosus. Indones Chemia Acta. 2013;4(1):7-11.

21. Pujimulyani D, Raharjo S, Marsono Y, Santosu O. Aktivitas Antioksidan dan Kadar Senyawa Fenolik pada Kunir Putih (Curcuma mangga Val) Segar dan Setelah Blanching. Agritech. 2010;30(2):68-74. doi:10.22146/agritech.9675

22. Jantan I, Saputri FC, Qaisar MN, Buang F. Correlation between Chemical Composition of Curcuma domestica and Curcuma xanthorrhiza and Their Antioxidant Effect on Human Low-Density Lipoprotein Oxidation. Evid Based Complement Alternat Med. 2012;2012:438356. doi:10.1155/2012/438356

23. Maharani BC, Lindriati T, Diniyah N. Pengaruh Variasi Waktu Blanching dan Konsentrasi Asam Sitrat Terhadap Karakteristik dan Aktivitas Ekstrak Pigmen Ubi Jalar Ungu (Ipomoea batatas L.). JP2 Jurnal Penelitian Pangan Indones J Food Res. 2016;1(1):60-7.
24. Hasrawati A, Hardianti H, Qama A, Wais M. Pengembangan Ekstrak Etanol Limbah Biji Pepaya (Carica papaya L) Sebagai Serum Antijerawat. Jurnal Fitofarmaka Indonesia. 2020;7(1):1-8. doi:10.33096/jffi.v7i1.458

25. Yumas M. Formulasi Sediaan Krim Wajah Berbahan Aktif Ekstra Metanol Biji Kakao Non Fermentasi (Theobroma cacao L) Kombinasi Madu Lebah. Jurnal Industri Hasil Perkebunan. 2016;11(2):75-87. doi:10.33104/jihp.v11i2.3414

26. Directorate General of Drug and Food Control, Ministry of Health, Republic of Indonesia. Formularium Kosmetika Indonesia. Jakarta, Indonesia: Ministry of Health, Republic of Indonesia; 1985.

27. Badan Standarisasi Nasional. Sediaan Tabir Surya. SNI 16-4399-1996. Jakarta, Indonesia: Badan Standarisasi Nasional; 1996.

28. Ardana M, Aeyni V, Ibrahim A. Formulasi dan Optimasi Basis Gel HPMC (Hidroxy Propyl Methyl Cellulose) dengan Berbagai Variasi Konsentrasi. J Trop Pharm Chem. 2015;3(2):101-8. doi:10.25026/jtpc.v3i2.95

29. Tambunan S, Sulaiman TNS. Gel Formulation of Lemongrass Essential Oil with HPMC and Carbopol Bases. Majalah Farmaseutik. 2018;14(2):87-95. doi:10.22146/farmaseutik.v14i2.42598

30. Kedare SB, Singh RP. Genesis and development of DPPH method of antioxidant assay. J Food Sci Technol. 2011;48(4):412-22. doi:10.1007/s13197-011-0251-1

31. Pratiwi D, Wardaniati I. Pengaruh Variasi Perlakuan (Segar dan Simplisia) Rimpang Kunyit (Curcuma domestica) terhadap Aktivitas Antioksidan dan Kadar Fenol Total. Jurnal Farmasi Higea. 2019;11(2):159-65.