Relationship Between Metal Load (Selenium, Arsenic, Molybdenum) of Soil, Plant and Serum of Buffaloes

ZAFAR IQBAL KHAN¹*, KAFEEL AHMAD¹, NAUNAIN MEHMOOD², ASIA FARDOUS¹, YONGJUN YANG³, JING MA³, TASNEEM AHMAD⁴, FU CHEN³, SUMAIRA GONDAL¹, KINZA WAJID¹, IFRA SALEEM MALIK¹, MUDASRA MUNIR¹, IJZA RASOOL NOORKA⁵, MUHAMMAD FAHAD ULLAH⁶, HUMAYUN BASHIR¹, MOHAMMAD REZA KOUHKAN NEJAM⁷, ALI-REZA BAYAT⁷

¹ Department of Botany, University of Sargodha, Sargodha, 40100, Pakistan
² Department of Zoology, University of Sargodha, 40100, Sargodha, Pakistan
³ School of Environment Science and Spatial Informatics, China University of Mining and Technology, Xuzhou, China
⁴ Pakki Thatti R & D Farm, Toba Tek Singh, Pakistan
⁵ University College of Agriculture, University of Sargodha, Pakistan
⁶ Department of Earth Sciences, University of Sargodha, Sargodha, Pakistan
⁷ Milk Production, Production Systems, Natural Resources Institute Finland (Luke), 31600 Jokioinen, Finland

Abstract: Presence of hazardous minerals in three major components (soil, forage and buffalo serum) was evaluated for determining extent of metal contamination in Sargodha, Pakistan. Ten soil and forage samples for each season were taken from ten randomly selected sites. 30 grazing buffaloes of ‘niliravi’ breed (calves, non-lactating and lactating) were also sampled randomly from these sites for collection of serum. Heavy metal quantification via atomic absorption spectrophotometry was carried out for selenium, molybdenum and arsenic. Non-significant but positive correlation was found for selenium (Se) concentrations in soil, forage and serum and molybdenum (Mo) concentrations in soil and serum. Negative correlation (p>0.05) was obtained for Mo level between forage-serum and soil-forage which was indicative of molybdenum deficiency in the soil. The results suggested that the heavy metals remained below maximum permissible levels and were non-toxic for the ruminants of the study area.

Keywords: heavy metals, toxicity, soil, Niliravi buffalo, forage, serum

1. Introduction

Livestock productivity and nutritional status are dependent upon balanced provision of all the minerals and food components in diet [1]. With growing population and issues of food security, burden on agriculture and livestock sector has increased manifolds. A major share in coping with food scarcity issues is of livestock production field [2]. It is of utmost importance to assess nutritional and health status of ruminants to ensure maximum productivity. Animals need different metals in varying concentrations necessary for normal physiological functioning. Selenium (Se) as a constituent of glutathione peroxidase is an important part of cycle involving reduction of hydrogen peroxide. Se deficient animals exhibit hepatitis dietetica and liver necrosis, which is often fatal. Swine mulberry heart disease is triggered by low Se and vitamin E levels in the body causing pigs to die in young age [3]. Benefits of arsenic have been reported in literature for animal health and performance. Arsenic acts similarly to antibiotics in controlling harmful microbes in intestines [4].

Another essential micronutrient for animal and plant growth is molybdenum (Mo). In plants, Mo helps in nitrogen fixation and assimilation being a part of nitrate reductase [5].

The current study was aimed at investigating and evaluating the mineral status of forage and soil of livestock rich agricultural region of Pakistan in order to assess mineral bioavailability. Moreover, evaluation of the similar minerals in ruminants was also done in order to determine the health risk among ruminants (Niliravi buffalo) of that area. The information presented in this study would be

*email: zikhan11@gmail.com
helpful for livestock rearing areas of Pakistan and other Asian countries with similar environmental conditions.

2. Materials and methods

2.1. Study Site

The dairy farm of Fodder Research Institute (FRI), Sargodha and associated farm lands were selected as the study site. Sargodha is predominantly agricultural area of Pakistan with extended summer season and short winter season. Mean summer and winter temperatures have a great fluctuation ranging between 14 to 35°C. Farm lands were cultivated with different forage species, namely Sarso (*Brassica compestris*), oat (*Avenasariva*) and Barsem (*Trifolium alexander*). Soil and forage samples were taken from the farm lands while blood was collected from the Niliravi buffalo at the farm.

2.2. Sample collection

Ten randomly selected sites separated by the distance of 1 acre were chosen for the soil and forage sample collection. Soil samples (10) were collected from 12-15 cm depth, by stainless steel auger, partially containing all soil layers [6]. Likewise, the forage samples were collected from the farm lands during summer and winter seasons. The samples were given a wash in distilled water and dilute HCl for removal of dust residues and other contaminants. Moisture removal was achieved for soil and forage samples via air drying and subsequent oven drying at 50°C for 15 days.

Three cohorts of buffaloes (calves, non-lactating and lactating) were made for collection of blood samples during summer and winter. Ten buffaloes were sampled from each group and the blood was collected from the jugular vein with the help of sterilized needle. To prevent clotting, blood was quickly shifted to heparinized Na-Citrate vials. Serum separation from plasma was achieved after centrifugation (3000 rpm) for half hour. After successful separation, the serum was placed in labeled vials and stored in freezer till further use.

2.3. Sample preparation

All samples (soil, forage and serum) were digested following the protocol prescribed by Ugulu et al. [7]. Dry samples (forage and soil) were taken in 1g quantity while serum sample was taken in 1 mL quantity to be digested with H$_2$O$_2$ and H$_2$SO$_4$ (2:1). The sample, after complete digestion, was filtered using Whatman filter paper # 42. Final volume of 50 mL was made for each sample after adding required amount of double distilled water.

2.4. Mineral analysis

Mo contents of soil, forage and serum were determined using AAS equipped with D$_2$ corrector and graphite furnace. Fluorometric method was used along with injection hydride generation AAS to determine As and Se in the samples [8].

2.5. Statistical analysis

Data were subjected to the statistical analysis using the SPSS software for one-way ANOVA, correlation and bioconcentration factor (BCF) keeping significance level at 0.001, 0.01 and 0.05 [9].

3. Results and discussions

Mineral analysis

Soil

Sampling seasons had considerable (*p*<0.001) variation with respect to Se concentration in soil (Table 1). Se level varied between 0.361 and 0.465 mg/kg in summer and winter respectively (Figure 1). These values were slightly below the critical value for Se (0.5 mg/kg) in soils [10] indicating deficiency of Se at all locations. A similar study at adjoining city of Jhang, revealed high Se level
(0.70-0.94 mg/kg) is soil [11] in comparison to present investigation. Low Se level is also reported in other studies carried out by Khan et al. [12, 13] in Punjab, Pakistan. Current study Se values, on the other hand, were lower than the findings of Kunli et al. [14]. Excessive or lower amount of Se in animals depend upon the bioavailable amount of Se to the plants [15].

**Table 1.** One-way ANOVA for metal concentrations in soil, forage and serum during winter and summer seasons

| Source of variation | Degree Of freedom | Soil | Forage | Serum |
|---------------------|-------------------|------|--------|-------|
|                     | (df)              | Se   | As     | Mo    |
| Sampling Period     | 1                 | 0.045*** | 0.051** | 0.003** | 0.015*** | 0.001*** |
| Sampling Period     | 1                 | 0.078** | 0.037** | 0.001** | 0.006** | 0.003** |
| Error               | 18                | 0.002 | 0.014  | 0.002  | 0.002  | 0.001  |
| Error               | 18                | 0.020** | 3.698** | 1.568** | 5.778*** | 0.903** |

***p<0.001, **p<0.01, ns= non-significant

![Figure 1. Selenium level fluctuations in soil and forage at different sampling seasons](image)

Arsenic level in soil did not display much variation (*p*>0.05) with respect to the seasons (Table 1). There was slight seasonal variation in As concentration in soil (winter, 3.19 mg/kg; summer, 3.06 mg/kg) (Figure 2). Lower As levels were reported in current study compared to the critical concentration of 20 mg/kg [16]. Soil evaluation revealed exceptionally high As content in Jhang city (12.01 to 14.89 mg/kg) [11] compared to current study values. The concentration of other minerals (iron, aluminum, and phosphorous), soil type and *pH* of the soil [17, 18] also affect As level and mobility in soil layers. A drop in *pH* increases As leaching in soils [19]. Current As levels were safe for the grazing livestock of the area under study.

Molybdenum concentration exhibited negligible variation during the two sampling periods (Table 1) yielding non-significant results (*p*>0.05). Mean Mo soil concentration in winter was 0.968 mg/kg while it showed slight decrease during summer season (0.904 mg/kg) (Figure 3). Critical level of Mo in soils is estimated to be 1 mg/kg [20] and current results were very close to this limit. Soil optimal levels given for Mo (1-2 mg/kg) by McDowell [21] make this element highly bioavailable for plants which potentially absorb high amounts of Mo ranging between 10 and 20 mg/kg. On the other hand, Mo-deficient soils are rich in iron and have low *pH* and organic matter which affect its ability to retain Mo. Such soils require supplementation in the form of foliar sprays or seed treatments and application of fertilizers.
Figure 2. Arsenic level fluctuations in soil and forage at different sampling seasons

Figure 3. Molybdenum level fluctuations in soil and forage at different sampling seasons

Forage

The forage samples yielded non-significant variation ($p>0.05$) in terms of two sampling seasons (Table 1) for Se concentration. Exhibiting high mean value in winter (1.22 mg/kg), the Se mean level dropped to 0.94 mg/kg in summer (Figure 1). Critical limit in plants, set for Se, is 0.1 mg/kg [22] and the current values were considerably higher than this concentration. It could be deduced that plants absorbed quite high levels of Se from soil of the study area. McDowell et al. [23]and Khan et al. [12] obtained lower values for Se in plants compared to the current study whereas, similar observations have been reported in Colombia [24] and Pakistan [11, 25].

ANOVA analysis displayed non-significant variation ($p>0.05$) in the mean concentration of As in forage during winter and summer (Table 1). Winter mean level in forage (1.53 mg/kg) was higher than the summer mean level (1.36 mg/kg) (Figure 2). The forage plants had As values lower than the critical limit of 3 mg/kg [16]. High As values (1.82-1.91 mg/kg) in forage samples were exhibited in adjoining city of Jhang in another study [11]. Arsenic methylation in plants potentially reduces the risks associated with high As ingestion [26]. Higher levels of soil As result in decreased plant growth [27].

Insignificant change ($p>0.05$) in Mo forage concentration was observed in the sampling seasons (Table 1). Two sampling intervals depicted a range of 5.9 to 6.2 mg/kg Mo in forages under study; winter season values were relatively higher than values obtained for summer season (Figure 3). Current winter Mo levels in the plants exceeded the critical level (6 mg/kg) [28] of Mo in plants whereas, the mean Mo level in summer remained close to this limit. Around 65% of the samples displayed Mo deficiency in forages. Nearly similar observations were made by McDowell et al. [23],
Merkel et al. [29] and Hornick et al. [30] in Florida. Soil pH substantially affects the uptake of Mo by plants [31]. Metal bioavailability also increases with high organic content and moisture in soil [32].

**Serum**

Significant variation \((p<0.05)\) was observed in serum levels of Se in calves and non-lactating buffaloes while lactating group showed nearly similar Se concentration \((p>0.05)\) in both sampling seasons (Table 1). Mean Se levels varied between 0.017 and 0.028 mg/L in all buffalo cohorts in winter while in summer season, Se concentration remained between 0.012 and 0.024 mg/L (Figure 4) displaying high mean levels in winter season. Critical limit set for serum Se concentration (0.03 mg/L) [28] was higher than the present study findings. 90% of the samples displayed Se deficiency in lactating buffaloes while the other two groups (non-lactating and calves) were 100% deficient for Se. High Se level may result in multiple toxicities in the grazing animals such as skin cracks, emaciation, gangrene of extremities, hoof detachment; high Se level in buffaloes may even be fatal [33]. Deore et al. [34] found lower Se levels in their study on animals in comparison to current study. Similar Se levels were exhibited for buffalo plasma (0.02 mg/kg) at Jhang [11]. Selenium is a beneficial trace element for animal bodies and plays role in growth, fertility, antibody production and function of lymphocytes and neutrophils. Rats and pigs having Se deficiency exhibit hepatic apoptosis; ruminants, on the other hand display different clinical signs of poor antibody response, impaired immunity, embryonic mortality, white muscle disease [35].

**Figure 4.** Selenium level fluctuations in serum at different sampling seasons

Heterogeneous distribution of As in Niliravi buffaloes was evident through significant variation \((p<0.05)\) with respect to sampling seasons in dry buffaloes, whereas, the lactating buffaloes and calves exhibited non-significant variation \((p>0.05)\) during the two sampling intervals (Table 1). Mean winter levels (0.023-0.069 mg/L) were higher than the summer season (0.020-0.064 mg/L) for as (Figure 5). Current study values were below the toxic limit (1 ppm) for As [36]. All samples from the three buffalo cohorts were 100% deficient for As. Kinacid and Hodgson [37] found lower mean values for
as while Anke et al. [38] observed higher mean concentrations for As in comparison to current observations. Buffaloes at Jhang city had higher As content (0.91-0.94 mg/kg) in their blood plasma [11] in contrary to current findings. No imminent threat was found for studied livestock with respect to As concentration.

The concentration of Mo varied significantly \((p<0.05)\) in all the buffalo categories with respect to sampling intervals (Table 1). Following the pattern of high serum levels in winter season, the mean concentration of Mo varied between 2.61 and 2.70 mg/L while in summer season, the range remained between 1.82 and 2.26 mg/L (Figure 6). Clawson et al. [39] reported the toxic levels of Mo as 5 ppm; current observations were considerably below this toxic limit. Current study values were lower than the standard values [22]. No imminent threat was found for studied livestock with respect to Mo concentration.
Correlation

Positive but non-significant correlation was observed for the three mediums under study (forage, soil, serum) to determine potential toxicity of Se revealing bioavailability and balance of flow. On the other hand, As had significantly positive correlation between serum and forage manifesting direct accumulation of As in animal bodies after forage consumption. Mo exhibited negative and non-significant correlation for forage-serum and soil-forage while non-significant but positive value was observed for serum and soil (Table 2) indicating probable deficiency and low cross-transmission.

### Table 2. Correlation among metal between Soil-Forage-Serum

| Metal | Soil-Forage | Forage-Serum | Soil-Serum |
|-------|-------------|--------------|------------|
| Se    | 0.339       | 0.145        | 0.407      |
| As    | 0.260       | 0.251        | 0.479(*)   |
| Mo    | -0.143      | 0.367        | -0.089     |

*Correlation is significant at the 0.05 level (2-tailed).

### Table 3. Bio concentration factor for metals from soil-forage-serum

| Metals | Sampling Seasons | Soil to Forage | Forage to Serun |
|--------|------------------|----------------|-----------------|
|        |                  | Lactating Buffalo | Non-Lactating Buffalo | Buffalo calves |
| Se     | Winter           | 2.1282          | 0.0237          | 0.0229         | 0.0139       |
|        | Summer           | 2.0338          | 0.0255          | 0.0127         | 0.0127       |
| As     | Winter           | 0.4722          | 0.0450          | 0.0228         | 0.0150       |
|        | Summer           | 0.4444          | 0.0470          | 0.0191         | 0.0147       |
| Mo     | Winter           | 68.197          | 0.0415          | 0.0433         | 0.0334       |
|        | Summer           | 65.641          | 0.0380          | 0.0313         | 0.0306       |

Bio-concentration factor (BCF)

BCF for Se (soil to forage) in winter (3.128) was higher than the summer season (2.603). Comparatively, BCF was quite low between forage and serum for all buffalo categories; BCF ranged from 0.013 to 0.023 in winter while in summer, its range was 0.012 to 0.025. Only lactating buffaloes displayed higher summer BCF (0.025) for Se. BCF for arsenic manifested low values between soil and forage in both winter (0.472) and summer (0.444). High transfer rate of metals from forage to non-lactating buffaloes was observed in winter (0.022) while in case of lactating buffaloes, high BCF was observed in summer (0.047). Interestingly, Mo exhibited unusually high BCF from soil to forage indicating high metal uptake in both winter (68.197) and summer (65.641).

However, BCF for Mo between forage and serum of buffaloes was almost similar to other metals. Winter BCF values for all buffalo categories were comparatively higher than summer season ranging between 0.0313 and 0.0433.

Over all high BCF values in winter season corresponded to efficient metal transfer and low resistance in forage varieties. Lower BCF could be due to limited absorption capacity of plants or animals [25]. Environmental factors, seasonal influence, age and type of plant, pH and type of soil could potentially affect metal uptake and transfer among different media.

### 4. Conclusions

Presence of hazardous minerals in three major components (soil, forage and buffalo serum) was evaluated for determining extent of metal contamination in Sargodha, Pakistan. Ten soil and forage samples for each season were taken from ten randomly selected sites. 30 grazing buffaloes of ‘niliravi’ breed (calves, non-lactating and lactating) were also sampled randomly from these sites for collection of serum. Heavy metal quantification via atomic absorption spectrophotometry was carried out for selenium, molybdenum and arsenic. Non-significant but positive correlation was found for selenium (Se) concentrations in soil, forage and serum and molybdenum (Mo) concentrations in soil and serum. Negative correlation (p>0.05) was obtained for Mo level between forage-serum and soil-forage which
was indicative of molybdenum deficiency in the soil. The results suggested that the heavy metals remained below maximum permissible levels and were non-toxic for the ruminants of the study area.

**Ethics:** All the study protocols were approved by the Institutional Animal Ethics Committee, University of Sargodha (Approval No.25-A18 IEC UOS). All the experiments performed complied with the rules of National Research Council [40] and all methods were performed according to relevant guidelines and regulations.

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