Sentinel Lymph Node Analysis in Colorectal Cancer Patients Using One-Step Nucleic Acid Amplification in Combination With Fluorescence and Indocyanine Green

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Purpose: Analysis of the sentinel lymph node (SLN) in colorectal cancer (CRC) patients was proposed for more accurate staging and tailored lymphadenectomy. The aim of this study was to assess the ability to predict lymph node (LN) involvement through analysis of the SLN with a one-step nucleic acid (OSNA) technique in combination with peritumoral injection of indocyanine green (ICG) and near-infrared (NIR) lymphangiography in CRC patients.

Methods: A total of 34 patients were enrolled. Overall, 51 LNs were analyzed with OSNA. LNs of 17 patients (50%) were examined simultaneously with hematoxylin and eosin (H&E) and OSNA.

Results: SLN analysis of 17 patients examined with H&E and OSNA revealed that OSNA had a higher sensitivity (1 vs. 0.55), higher negative predictive value (1 vs. 0.66) and higher accuracy (100% vs. 76.4%) in predicting LN involvement. Overall, OSNA showed a sensitivity of 0.69, specificity of 1, accuracy of 88.2%, and stage migration of 8.8%. Compared to those who were OSNA (−), OSNA (+) patients had a greater number of LN metastases (4.8 vs. 0.16, P = 0.04), higher G3 rate (44.4% vs. 4%, P = 0.01), more advanced stage of disease (stage III: 77.8% vs. 16%; P = 0.00) and were more rapidly subjected to adjuvant chemotherapy (39.1 days vs. 50.2 days, P = 0.01).

Conclusion: SLN analysis with OSNA in combination with ICG-NIR lymphangiography is feasible and can detect LN involvement in CRC patients. Furthermore, it allows for more accurate staging reducing the delay between surgery and adjuvant chemotherapy.

Keywords: Colorectal cancer; Sentinel lymph node analysis; One-step nucleic acid; Fluorescence; Indocyanine green

INTRODUCTION

Colorectal cancer (CRC) is the third most common cancer worldwide and lymph node (LN) status is the most powerful prognostic factor [1, 2]. After curative treatment, between 30% and 65% of patients with CRC will develop recurrent disease [3]. Under-staging for CRC patients is thought to occur in around 10%–20% of patients and the relevant literature shows that examination of only one LN slide using hematoxylin and eosin (H&E) staining leaves up to 33% of metastases unidentified [4].

Moreover, some studies have shown that extensive lymphadenectomy is associated with improved prognosis in patients with more advanced stage CRC, even though numerous postoperative complications related to this extensive surgery are described [5, 6]. On the other hand, organ-preserving surgery without lymphadenectomy has been proposed for early-CRC with encouraging results [7, 8].

Among colorectal surgeons and treatments, it is now accepted...
that “one size does not fit all,” and there is increasing agreement regarding the need for a more targeted surgery and a tailored lymphadenectomy [9].

The challenge is the careful selection of patients; early CRC disease could benefit from organ-preserving surgery while more advanced stage disease would require tailored lymphadenectomy taking into consideration LN status, the presence of aberrant lymphatic drainage and “skip” metastases.

The sentinel lymph node (SLN) is defined as the first LN in the lymphatic basin that drains the primary tumor, and it has recently been proposed as a strategy for more precise staging in gastrointestinal tumors [10]. Some groups have demonstrated that peritumoral injection of indocyanine green (ICG) with near-infrared (NIR) provides for visualization of LNs and aberrant lymphatic drainage during CRC resection [11]. Furthermore, a novel technique for SLN analysis has been developed: one-step nucleic acid amplification (OSNA; Sysmex Corp., Kobe, Japan) is a rapid mRNA assay that by measuring levels of cytokeratin 19 (CK19) can identify the presence of metastases and micro-metastases in the LNs within 30–40 minutes of their removal [12].

The aims of this pilot study were to assess the ability to predict LN involvement in patients with CRC using SLN analysis with OSNA in combination with ICG-NIR lymphangiography and to estimate the benefits of the routine use of this technique.

METHODS

Study design and population
This cohort study relies on data retrieved from a prospectively maintained database of consecutive patients undergoing elective colorectal resection for adenocarcinoma at St. Giuseppe Moscati Hospital of Avellino, Italy. The database was implemented in 2016 and included preoperative, operative, and postoperative data. Inclusion criteria were any adult patients diagnosed with CRC requiring surgical treatment. Exclusion criteria included participants with an allergy to any ICG. In total, 34 patients were included in this study.

The study was conducted according to the ethical principles of the Institution following the Declaration of Helsinki and was approved by the Institutional Review Board of the St. Giuseppe Moscati Hospital (approval number: 201801). Written informed consent was granted by study participants for publication of this article.

Surgical technique and histopathological evaluation
All patients underwent colonic preparation within 3 days prior to surgery. Immediately after access to the abdominal cavity by laparotomy or laparoscopy, a flexible endoscope was passed to the tumor site to allow a 1- to 2-mL submucosal injection of ICG (2.5–5 mg/mL) in four points around the tumor. The mesocolic or mesorectal tissue was then examined under NIR illumination (KARL STORZ SE & Co., Tuttlingen, Germany) and any identifiable LNs were excised and submitted immediately to the pathology department (Fig. 1). The retrieved LNs were staged according to the classification system of the Japanese Society for Cancer of the Colon and Rectum [13]. LNs harvested in this way were cleaned of fat, frozen in liquid nitrogen and stored at −80°C for processing. LNs weighing between 0.05 g and 0.6 g or with a cross diameter of less than 8 mm were exclusively processed using the OSNA technique. LNs weighing more than 0.6 g or more than 8 mm in diameter were dissected and analyzed using both the OSNA and H&E methods. In the latter case, LNs were divided at 2-mm intervals, and nonadjacent blocks were alternatively subjected to histopathological examination with H&E or OSNA as described by Yamamoto et al. [14]. For the OSNA assay, larger LNs or slices of smaller LNs were placed in homogenized lysis buffer (Ly-
### Table 1. Clinicopathologic characteristics of patients according to OSNA status

| Characteristic                                      | All patients (n = 34) | OSNA (−) (n = 25) | OSNA (+) (n = 9) | P-value |
|-----------------------------------------------------|-----------------------|-------------------|------------------|---------|
| Age (yr)                                            | 69.3 ± 11.8           | 71 ± 9.9          | 64.6 ± 15.8      | 0.82    |
| Male sex                                            | 21 (61.8)             | 16 (64.0)         | 5 (55.6)         | 0.70    |
| Body mass index (kg/m²)                             | 24.8 ± 2.7            | 25.1 ± 2.3        | 23.9 ± 3.4       | 0.33    |
| Tumor location                                      |                       |                   |                  |         |
| Right colon                                         | 10 (29.4)             | 6 (24.0)          | 4 (44.4)         | 0.39    |
| Left colon                                          | 11 (32.4)             | 8 (32.0)          | 3 (33.3)         | 1.00    |
| Rectum                                              | 13 (38.2)             | 11 (44.0)         | 2 (22.2)         | 0.42    |
| Neoadjuvant chemotherapy                            | 2 (5.9)               | 0 (0)             | 2 (22.2)         | 0.06    |
| Surgical approach                                   |                       |                   |                  |         |
| Open                                                | 26 (76.5)             | 17 (68.0)         | 9 (100)          | 0.07    |
| Laparoscopic                                        | 8 (23.5)              | 8 (32.0)          | 0 (0)            |         |
| Stadiation                                          |                       |                   |                  |         |
| pT stage                                            |                       |                   |                  |         |
| pT1–pT2                                             | 9 (26.5)              | 9 (36.0)          | 0 (0)            | 0.07    |
| pT3–pT4                                             | 25 (73.5)             | 16 (64.0)         | 9 (100)          | 0.07    |
| pN stage                                            |                       |                   |                  |         |
| Negative                                            | 21 (61.8)             | 21 (84.0)         | 0 (0)            | 0.00*   |
| Positive                                            | 13 (38.2)             | 4 (16.0)          | 9 (100)          |         |
| Sentinel lymph node number examined                  | 1.5 ± 0.6             | 1.5 ± 0.7         | 1.3 ± 0.5        | 0.31    |
| Total lymph node number                             | 19.9 ± 11.1           | 18.2 ± 10.2       | 24.7 ± 12.9      | 0.19    |
| Metastatic lymph node number                        | 1.38 ± 3.5            | 0.16 ± 0.3        | 4.8 ± 5.8        | 0.04*   |
| >3 Metastatic lymph nodes                           | 3 (8.8)               | 0 (0)             | 3 (33.3)         | 0.01*   |
| Lymph node level (JSCCR)                            |                       |                   |                  |         |
| 1                                                   | 12 (35.2)             | 7 (28.0)          | 5 (55.6)         | 0.22    |
| 2                                                   | 11 (32.4)             | 8 (32.0)          | 3 (33.3)         | 1.00    |
| 3                                                   | 11 (32.4)             | 10 (40.0)         | 1 (11.1)         | 0.21    |
| M1                                                  | 2 (5.9)               | 0 (0)             | 2 (22.2)         | 0.06    |
| Histological grade                                  |                       |                   |                  |         |
| G1 (well differentiated)                            | 1 (2.9)               | 1 (4.0)           | 0 (0)            | 1.00    |
| G2 (moderate differentiated)                        | 28 (82.4)             | 23 (92.0)         | 5 (55.6)         | 0.03*   |
| G3 (poorly differentiated)                          | 5 (14.7)              | 1 (4.0)           | 4 (44.4)         | 0.01*   |
| Stage                                               |                       |                   |                  |         |
| I                                                   | 7 (20.6)              | 7 (28.0)          | 0 (0)            | 0.15    |
| II                                                  | 14 (41.2)             | 14 (56.0)         | 0 (0)            | 0.00*   |
| III                                                 | 11 (32.4)             | 4 (16.0)          | 7 (77.8)         | 0.00*   |
| IV                                                  | 2 (5.9)               | 0 (0)             | 2 (22.2)         | 0.06    |
| Adjuvant chemotherapy                               | 12 (35.3)             | 4 (16.0)          | 8 (88.9)         | 0.00*   |
| Time to start adjuvant chemotherapy (day)           | 42.8 ± 6.1            | 50.2 ± 4.1        | 39.1± 1.9        | 0.01*   |

Values are presented as mean ± standard deviation or number (%).
OSNA, one-step nucleic acid; JSCCR, Japanese Society for Cancer of the Colon and Rectum.
*P < 0.05, significant difference.
norhag, Sysmex) and then centrifuged. Lysate was then extracted for CK19 mRNA RT-LAMP in the RD-100i system (Sysmex) using the Lynoamp (Sysmex) reagent kit [15]. On the basis of previous investigations, OSNA results with CK19 mRNA above 250 copies/L were designated as positive and those with fewer than 250 copies/L were considered negative [12]. The result of the SLN analysis was intraoperatively communicated to the surgeon but the surgical procedure was not modified. All patients underwent standard CRC resection in the conventional manner. A total or partial mesocolon or mesorectal excision was performed. Subsequently, records of each patient were discussed in a multidisciplinary conference with surgeons, pathologists and oncologists, taking into consideration the definitive examination of the surgical specimen and the result of the SLN analysis. The final histology of the surgical specimen and harvested LN were always concluded based on H&E analysis.

Statistical analysis
Statistical analyses were carried out by dividing the study population into 2 groups based on the results of the SLN analysis with OSNA. Continuous variables are presented as the mean ± standard deviations and were compared using Student t-test or the nonparametric Mann-Whitney test, as appropriate. Categorical variables are presented as numbers (percentages) and were compared across groups using the chi-square test or Fisher exact test, as appropriate. Furthermore, sensitivity, specificity, positive predictive value (PPV), negative predictive value (NPV), and test accuracy were calculated comparing the intraoperative result of OSNA with the definitive examination of all LNs detected in the surgical specimen, as routinely determined by the pathologist. All statistical analyses were performed with IBM SPSS Statistics ver. 23.0 (IBM Co., Armonk, NY, USA).

Table 2. Comparing definitive histology with OSNA analysis of retrieved fresh lymph nodes also evaluated with hematoxylin and eosin

| OSNA  | Histology (+) | Histology (−) | Total |
|-------|---------------|---------------|-------|
| (+)   | 9             | 0             | 9     |
| (−)   | 0             | 8             | 8     |
| Total | 9             | 8             | 17    |

OSNA, one-step nucleic acid.

Table 3. Comparing definitive histology with H&E analysis of retrieved fresh lymph nodes also evaluated with one-step nucleic acid

| H&E   | Histology (+) | Histology (−) | Total |
|-------|---------------|---------------|-------|
| (+)   | 5             | 0             | 5     |
| (−)   | 4             | 8             | 12    |
| Total | 9             | 8             | 17    |

H&E, hematoxylin and eosin.

RESULTS

Study population
The characteristics of all patients are shown in Table 1. Thirty-four patients with colorectal adenocarcinoma were included in this study. Intraoperative evaluation of the SLN with OSNA was positive (OSNA [+] in 9 patients (26.5%) and negative (OSNA [−]) in 25 patients (73.5%). No complications related to ICG injection occurred during surgery. Globally, 51 LNs were retrieved and examined with a median of 1 (1–3) LN harvested for each patient.

In 17 patients (50%), the SLN weighed more than 0.6 g or had a diameter greater than 8 mm, making LN evaluation possible with both OSNA and H&E as described. Comparing definitive histology with simultaneous analysis of retrieved fresh LN with OSNA and H&E, the OSNA technique had higher sensitivity (95% confidence interval [CI]), 1 (0.77–1.00) vs. 0.55 (0.30–0.78), a higher NPV (95% CI), 1 (0.77–1.00) vs. 0.66 (0.40–0.86), and higher accuracy, 100% vs. 76.4%. Both methods showed high specificity (95% CI), 1 (0.77–1.00) vs. 1 (0.77–1.00), and a high PPV (95% CI), 1 (0.77–1.00) vs. 1 (0.77–1.00) (Tables 2 and 3).

Overall, comparing the definitive histology with OSNA analysis of retrieved fresh SLN of all patients, OSNA had a sensitivity (95% CI) of 0.69 (0.51–0.81), specificity (95% CI) of 1 (0.87–1.00), and accuracy of 88.2%. Its PPV (95% CI) was 1 (0.87–1.00), and its NPV (95% CI) was 0.84 (0.66–0.93) (Table 4). A summary of the sensitivity, specificity, PPV, and NPV are shown in Table 5.

Study OSNA (+) vs. OSNA (−) population characteristics
OSNA (+) and OSNA (−) patients were statistically similar in age (P = 0.82), sex (P = 0.70), BMI (P = 0.33), primary disease (P = 0.39 for right colon, P = 1.00 for left colon and P = 0.42 for rectal cancer) and the number of patients undergoing neoadjuvant chemotherapy (22.2% vs. 0%, P = 0.06) (Table 1). No statistically significant difference was found between the 2 groups in surgical approach, but 100% of the procedures for patients in the OSNA (+) group were performed by laparotomy (P = 0.07). In 100% (vs. 16%) of patients in the OSNA (+) group, pN was positive on definitive histological examination (P = 0.00). Globally, 69.2% (9 of 13) of patients with pN positive on definitive histological examination were identified during surgery. Yet, a greater number of LNs metastases were found in OSNA (+) patients (4.8 ± 5.8 vs. 0.16 ± 0.3, P = 0.04). Moreover, 33.3% (vs. 0%) of patients in the

Table 4. Comparing definitive histology with OSNA analysis of all retrieved fresh lymph nodes

| OSNA  | Histology (+) | Histology (−) | Total |
|-------|---------------|---------------|-------|
| (+)   | 9             | 0             | 9     |
| (−)   | 4             | 21            | 25    |
| Total | 13            | 21            | 34    |

OSNA, one-step nucleic acid.
Table 5. Summary of sensitivity, specificity, positive predictive value, negative predictive value, and test accuracy

| Variable                                      | No.  | Sensitivity (95% CI) | Specificity (95% CI) | Accuracy (%) | PPV (95% CI) | NPV (95% CI) |
|-----------------------------------------------|------|----------------------|----------------------|--------------|--------------|--------------|
| Comparing definitive histology with OSNA analysis of SLN also evaluated with H&E | 17   | 1 (0.77–1.00)        | 1 (0.77–1.00)       | 100          | 1 (0.77–1.00) | 1 (0.77–1.00) |
| Comparing definitive histology with H&E analysis of SLN also evaluated with OSNA | 17   | 0.55 (0.30–0.78)     | 1 (0.77–1.00)       | 76.4         | 1 (0.77–1.00) | 0.66 (0.40–0.86) |
| Comparing definitive histology with OSNA analysis of all retrieved SLN | 34   | 0.69 (0.51–0.81)     | 1 (0.87–1.00)       | 88.2         | 1 (0.87–1.00) | 0.84 (0.66–0.93) |

OSNA (+) group had more than 3 metastatic LNs (P = 0.01). Regarding grading of the tumor, patients with OSNA (−) had a higher rate of G2 (moderately differentiated) (92% vs. 55.6%, P = 0.03), while patients who were OSNA (+) had a higher G3 (poorly differentiated) rate (44.4% vs. 4%, P = 0.01). A higher number of patients in the OSNA (+) group had distant metastases OSNA (+), though the difference did not reach statistically significance (22.2% vs. 0%, P = 0.06). Globally, OSNA (+) patients had more advanced stages of disease (stage III: 77.8% vs. 16%, P = 0.00; stage IV: 22.2% vs. 0%, P = 0.06). After a multidisciplinary discussion, 3 patients (8.8%) with SLN positive by the OSNA method were considered pN positive and subjected to adjuvant chemotherapy. Adjuvant chemotherapy was administered in 88.9% of patients in the OSNA (+) group (vs. 16%, P = 0.00), and the time to start chemotherapy was lower OSNA (+) (39.1 ± 19.1 days vs. 50.2 ± 41.1 days in the OSNA [−] group, P = 0.01).

DISCUSSION

In this study, we showed that intraoperative SLN analysis with OSNA in combination with ICG-NIR lymphangiography is safe, feasible and can predict the state of LN involvement in patients with CRC. Overall, OSNA showed a sensitivity of 69%, specificity of 100%, accuracy of 88.2%, PPV of 100%, negative predictive of 84%, false negative rate of 30.7% (4 of 13) and stage migration of 8.8% (3 of 34). Furthermore, 66.7% (8 of 12) of OSNA (+) patients received adjuvant chemotherapy more rapidly.

LN status plays a crucial role in oncologic therapeutic strategies, and despite the use of increasingly sophisticated imaging techniques, pre-operative metastatic LN identification in patients with CRC is unsatisfactory [16, 17].

Innovative organ preserving procedures such as transanal mininally invasive surgery allow an adequate resection of early tumors without resorting to extensive interventions [18]. Nevertheless, it is estimated that 10%–20% of patients in stage T1 will have LN metastasis, and such patients subjected to a localized resection are undertreated [19]. In Japan, on the other hand, unlike in western countries, a lateral LN dissection in association with conventional total mesorectal excision is performed for advanced rectal cancers (T3–4), and although it is associated with sexual and urinary dysfunction, the risk of intrapelvic recurrence is reduced by 50%, and survival increases [5, 20, 21].

Particularly in the latter 2 cases, SLN mapping with ICG-NIR lymphangiography may allow the identification of a smaller number of LNs representing the tumor status of the entire nodal basin, directing the surgeon to perform a targeted lymphadenectomy in place of a more radical or conservative treatment. Furthermore, exhaustive analysis of the SLN facilitates a more accurate staging of patients and can influence the decision for adjuvant treatment.

Two results of our study should be highlighted. First, in 3 patients (8.8%), the SLN analysis was strongly positive by OSNA but negative by H&E evaluation. After multidisciplinary assessment, these patients were considered pN positive and subjected to migration from stage II to III. This type of approach could lead to more accurate staging of the disease, allowing patients to access adjuvant chemotherapy. In fact, we know that 25% of CRC patients in stage II will die from recurrent disease [22]. One of the most important risk factors is the presence of unidentified LN micrometastasis. Rahbari et al. [23] concluded in a recent meta-analysis that the presence of LN micrometastasis is associated with poor overall survival and shorter disease-free survival in stage II CRC patients. SLN micro-metastasis can easily be missed by H&E evaluation [24, 25]. Only immunohistochemistry (IHC) or molecular biology techniques can detect micrometastatic dissemination without doubt, but they are laborious and time-consuming [25, 26]. OSNA allows detection of the specific mRNA of cytokeratins without classical nucleic acid extraction and purification. Moreover, the analysis of four LNs takes only 40 minutes. Yamamoto et al. [14] found a 95% concordance rate between OSNA and classical histological analysis with H&E and IHC. Recently, Miyake et al. [27] demonstrated that the ex vivo analysis of perirectal LNs by OSNA predicts LLN metastasis in advanced rectal cancer with a sensitivity of 100%, specificity of 86%, PPV of 57% and NPV of 100%. Other studies and a meta-analysis have also confirmed the ability of the OSNA to predict LN involvement [28]. In our study, OSNA was superior to H&E in identifying LN metastases, with a false negative rate of 0% vs. 44.4% and accuracy of 100% vs. 76.4%, respectively.

Another benefit we found was that OSNA (+) patients received adjuvant chemotherapy more rapidly, an advantage also found by other authors [29]. Indeed, although no definite cutoff has been established, the greater the delay in starting adjuvant chemother-
apy, the higher the risk of metastatic progression in patients with advanced CRC [30]. When we found LN metastasis during surgery, we planned the placement of a port-a-cath 2 weeks after surgery and started chemotherapy without necessarily waiting for the final pathologic reports.

The limitations of our study are the small number of patients and false-negative rate (30.7%). We attribute this latter result to the method of lymphangiography rather than OSNA. Emile et al. [31], in a recent meta-analysis that enrolled 12 articles and 248 patients showed a median sensitivity of 73.7% (range, 0%–100%), median specificity of 100% (range, 0%–100%) and accuracy rates of 75.7% (range, 0%–100%) for the detection of metastatic SLN in CRC patients with the use of ICG and NIR combined with H&E and IHC examination [31]. Except for the higher accuracy rate of our study (88.2% vs. 75.7%), the remaining results are similar to those reported in this meta-analysis [31]. The site of the ICG injection plays a vital role in determining the outcomes of the test. Emile et al. [31] in effect, have shown that combined subserosal and submucosal injection achieved 100% rates of sensitivity, specificity and accuracy. Thus far, we have only used submucosal ICG injection just before surgery and will try to modify our technique in order to improve sensitivity. A further limit for the routine use of OSNA is the price, which remains quite high.

In conclusion, SLN analysis with OSNA in combination with ICG-NIR lymphangiography is feasible and may allow intraoperative prediction of LN status in patients with CRC. Furthermore, its implementation allows more precise staging, reducing the delay between surgery and the onset of adjuvant chemotherapy. In our study, the results of SLN analysis did not influence surgical strategy, but OSNA and ICG-NIR lymphangiography could be used to develop customized surgery and tailored lymphadenectomy in patients with CRC.

CONFLICT OF INTEREST

No potential conflict of interest relevant to this article was reported.

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