Spatio-Temporal Pattern of Dinoflagellates Along the Tropical Eastern Pacific Coast (Ecuador)

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Among marine phytoplankton, dinoflagellates are a key component in marine ecosystems as primary producers. Some species synthesize toxins, associated with human seafood poisoning, and mortality in marine organisms. Thus, there is a large necessity to understand the role of environmental variables in dinoflagellates spatial-temporal patterns in response to future climate scenarios. In that sense, a monthly four-year (2013–2017) monitoring was taken to evaluate dinoflagellates abundances and physical-chemical parameters in the water column at different depths. Sampling sites were established at 10 miles in four locations within the Ecuadorian coast. A total of 102 taxa were identified, corresponding to 8 orders, 22 families, and 31 genera. Eight potentially harmful genera were registered but no massive blooms were detected. The most frequent dinoflagellates were Gymnodinium sp. and Gyrodinium sp. Environmental variables showed different mixing layer thickness and a conspicuous and deepening thermocline/oxycline/halocline and nutricline depending on annual and seasonal oceanographic fluctuations. This study confirms that seasonal and spatial distribution of the environmental variables are linked to the main current systems on the Eastern Tropical Pacific, thus the warm Panama current lead to a less dinoflagellates abundance in the north of Ecuador (Esmeraldas), while the Equatorial Upwelling and the cold nutrient-rich Humboldt Current influence dinoflagellates abundance at the central (Manta, La Libertad) and South of Ecuador (Puerto Bolivar), respectively. Inter-annual variability of dinoflagellates abundance is associated with ENSO and upwelling conditions. Climate change scenarios predict an increase in water surface temperature and extreme events frequency in tropical areas, so it is crucial to involve policy-makers and stakeholders in the implementation of future laws involving long-term monitoring and sanitary programs, not covered at present.

Keywords: dinoflagellates, HABs, ENSO, tropical Eastern Pacific, nutrients, upwelling, humboldt current

INTRODUCTION

The Equatorial Pacific region is characterized by a unique complex oceanographic variability with a well-defined Equatorial Front where El Niño Southern Oscillation (ENSO) events eventually occur (Santos, 2006; Gierach et al., 2012). The Ecuadorian coast is divided in two biogeographical regions, a northern “Bay of Panama” ecoregion from Azuero Peninsula to Bahía de Caráquez and a
southern “Guayaquil” ecoregion from Bahía de Caráquez to the Illescas peninsula in Perú (Sullivan and Bustamante, 1999). The northern coast of Ecuador is influenced by the warm current of Panama and the current of El Niño, while the Humboldt current brings cold water to the southern Ecuadorian coast which contributes to outcrops formation in the area (Cucalon, 1989). The southern sector is the most productive area regarding fisheries and shrimp production (Pennington et al., 2006). Due to the variability found in oceanographic conditions over the Ecuadorian coast, there is a noticeable heterogeneity concerning physical, chemical, and biological parameters which leads to a high diversity in phytoplankton communities (Smyda, 2008) and the potential formation of harmful algae blooms (HABs).

The oceanographic conditions identified as drivers of HABs, are linked to those conditions associated to upwelling systems (cold nutrient-rich waters), high column water stratification (mostly associated to ENSO conditions), and long-term sea surface temperature (SST) increase due to climate change (Hallegraeff, 2010; McCabe et al., 2016; Kudela et al., 2017). Those studies have related the particular oceanographic conditions with the occurrence of HABs in the Pacific Ocean (Brown et al., 2003; Franco-Gordo et al., 2004; Moore et al., 2010), have shown that those factors module the phytoplankton community and dynamics. In general, the phytoplankton community in the Eastern Tropical Pacific (ETP) region is characterized by relatively low levels of biomass, with the dominance of small species and rare large bloom-forming diatoms (Brown et al., 2003). Diatoms and dinoflagellates are two major groups of phytoplankton that flourish in the oceans (Abate et al., 2017), mostly registered in the south-eastern Equatorial Front (Jimenez-Bonilla, 1980; Torres and Tapia, 2002), and upwelling systems (Shulman et al., 2012). Particularly, dinoflagellates exhibit environmentally induced adaptation and survival to changing environmental conditions (Kudela et al., 2010).

Among the dinoflagellate community, there are some toxin producer dinoflagellates which may impact negatively marine ecosystems causing massive marine organism mortality, cause economic damage in coastal locations and affect human health by seafood consumption (Li et al., 2014). In addition, ballast waters contribute to the transfer of phytoplankton in commercial and oil tankers resulting in the dispersion of non-native phytoplankton species, increasing the risk of phytoplankton proliferation (Hallegraeff, 2003). Moreover, physiological changes may affect phytoplankton communities in response to climate change (Poloczanska et al., 2016) with consequences in their distribution and abundances worldwide. In that sense, it is important to perform studies related to phytoplankton biodiversity characterization and its spatio-temporal variability to guarantee an optimal integrated coastal management, contributing to future decisions from policy-makers to optimize waters vigilance.

Some countries of Latin America, such as Chile or Mexico, have implemented HABs monitoring regarding seafood production safety (Daguer et al., 2018; León-Muñoz et al., 2018). However, in Ecuador there is a lack of a national regulation plan, related to phytoplankton observations. Nevertheless, the Oceanographic Institute of Ecuador (INOCAR) has performed periodic monitoring since 1989 along the Ecuadorian coast. Torres (2015) observed the presence of algal proliferations along the Ecuadorian coast, which were more frequent in the Gulf of Guayaquil, where most aquaculture activities are located. Therefore, through the analysis of the INOCAR data bases, this study aims to provide information related to dinoflagellates abundances dynamic across a broad latitudinal area from 2013 to 2017 that constitute a base-line for future investigations.

**MATERIALS AND METHODS**

**Study Area and Samples Collection**

The study area was located between 0°89 North and 03°036 South, in the ETP Ocean. Along the study area, four geographic points were settled as sampling stations: three of them located 10 miles from the coast: (1) Esmeraldas (Galera San Francisco), (2) Manta, (3) La Libertad (Santa Elena Peninsula), and (4) Puerto Bolívar (Figure 1). All stations have an approximate depth of 100 m. except for Puerto Bolívar station, which is located approximately 27 miles, inside the submarine platform in a pit that is 9 m deep, northeast of Santa Clara Island in the Gulf of Guayaquil.

At each sampling station, 200 mL water samples were collected in plastic bottles from surface, 10, 20, 30, 40, 50, and 75 m depth using a 3 Liter volume Vand Dorn bottle, then preserved in neutral Lugol’s solution (final concentration 0.4%). Samplings were performed monthly from February 2013 to December 2017, between 09h00 to 12h00 am.

**Phytoplankton Identification and Abundance Estimation**

Water samples were settled in 25 mL volume Utermöhl chambers (Utermöhl, 1958), and observed in a Leica (ML) inverted microscope after 24 h. Cell abundance estimation was performed in horizontal transects of the chamber at 400× magnification. Results are reported in cell. L⁻¹. The validity of names of the different taxa was checked on the World Register of Marine Species (Horton et al., 2018). Dinoflagellate species identification was based on Steidinger and Jangen (1997); Hoppenrath et al. (2009) Hoppenrath et al. (2014); Omura et al. (2012), and Lassus et al. (2016). The list of toxic species was checked on the Taxonomic Reference List of Harmful Micro Algae (Moestrup et al., 2009).

In Esmeraldas and Puerto Bolivar no sampling was performed during February to December 2016 due to economic issues.

**Environmental Variables**

Temperature and salinity vertical profiles were obtained using a SBE 19plus V2 SeaCat Profiler CTD, at each sampling site. Data was processed using the SEABIRD software. Water samples for dissolved oxygen and nutrient estimation were collected with Van Dorn bottles at same depths as for phytoplankton analysis. Determination of dissolved oxygen for each sample was carried out *in situ* following the protocol for Dissolved Oxygen Test PEE/LAB-DOQ/01-INOCAR based on Apha (2005). Water samples for nutrient analysis were filtered with 0.45 µm millipore
filters and kept at −20°C. Nitrate, silicate and total phosphate were analyzed by techniques described in Test PEE/LAB-DOQ/01-INOCAR based on Strickland and Parsons (1972). For phosphates, as values in Equatorial Pacific waters are below 3 μmol L−1, the analysis followed is detailed in Part II of the protocol cited above. In Esmeraldas and Puerto Bolivar there is no data for dissolved oxygen and nutrients from February to December 2016 due to economic issues. In this study N/P is the ratio between Nitrate and Phosphate.

Data Analysis
Before any formal statistical analysis, an exploratory data analysis (EDA) was conducted to both, environmental and biological databases in order to avoid further statistical problems (i.e., proper family distribution of the variable, homoscedasticity, independence of the values), as suggested by Zuur et al. (2010). Beside this, the environmental variables were inspected in order to avoid collinearity (i.e., correlation among explanatory variables), thus all variables who showed a Pearson correlation coefficient (rho) higher than 0.7 were dropped from the analysis (Supplementary Figure S1). Community analysis considered the calculation of the common Shannon-Weaver diversity index and its spatio-temporal variability. From an ecological perspective those species with a percentage of occurrence higher than 2% were related with the environmental variables using a canonical correspondence analysis (CCA) for each year (ter Braak and Verdonschot, 1995) through the “cca” function of the vegan package (Oksanen et al., 2018). The significance of the models were achieved through the function “anova.cca” (from the vegan package, op.cit) which performs an ANOVA like permutation test using 999 permutations (Borcard et al., 2018). All the statistical analysis were conducted under the R software (R Core Team, 2018; version 3.4.4) and supervised by OneMind-DataScience1.

RESULTS
Dinoflagellate Community
Dinoflagellate Occurrence
During the studied period, a total of 102 dinoflagellate taxa were identified, corresponding to 8 orders, 22 families and 31 genera (Table 1). The genera *Tripus* and *Protoperidinium* were the most diverse, comprising 22 and 16 species, respectively (Table 1).

The most dominating taxa were *Gymnodinium sp* (70% occurrence), *Gyrodinium sp* (53%), and *Gyrodinium acutum* (21%). Other representative genera were *Protoperidinium* spp (16%), *Oxytoxum* spp (14%), *Proorocentrum micans* (13%), *Scrispiella* spp (12%), *Gonyaulax* spp (10%), and *Oxytoxum scolopax* (9.5%). The rest of the taxa presented lower percentage of occurrence (Table 1).

With respect to the temporal variability of the species mentioned above (most representative in terms of occurrence), *Gymnodinium sp*, *Gyrodinium sp*, *Oxytoxum* spp, and *O. scolopax* were present during the entire study period (Table 1). *Gonyaulax* spp was present also for both seasons (wet and dry seasons) and all years except 2016, which was not observed (Table 1). *Gyrodinium acutum* and *Scrispiella* spp was found in both wet and dry seasons, but only for the 2015 to 2017 period (Table 1).

*Protoperidinium* spp was found also in both wet and dry seasons but for the 2013–2015 period (Table 1). Instead, for the years 2016 and 2017, *Protoperidinium* spp was only observed during the dry season. *P. micans* was only found in dry season for 2013 and for wet season in 2016; instead during the period 2015–2017, *P. micans* was present in both seasons (Table 1).

Spatio-Temporal Variability
The two most dominant species, *Gymnodinium sp* and *Gyrodinium sp*, were more abundant in Manta, with bloom formation up to $1.2 \times 10^6$ cell. L$^{-1}$ for *Gymnodinium sp* in April 2017 (Figure 2A) and $5.4 \times 10^4$ cell. L$^{-1}$ for *Gyrodinium sp* in February 2016 (Figure 2B).

Regarding cell abundance estimation, 2013 registered for the entire water column studied (0–75 m depth) the lowest abundances comparing with the rest of the years (Figure 3). For 2013, the maximum values were found at surface in La Libertad (mostly corresponding to March ($2.8 \times 10^5$ cell. L$^{-1}$) and at 10 m depth in Puerto Bolivar (mostly in February, $2.0 \times 10^5$ cell. L$^{-1}$). In 2014, abundances were higher in the first 10 m of the water column except for Esmeraldas where abundances were similar in the first 30 m, being lower comparing with the other sampling sites for the same year (Figure 3). The maximum abundances in 2014, were observed at 10 m in Manta and at surface in Puerto Bolivar (with the maximum value corresponding to May, with $2.9 \times 10^5$ cell. L$^{-1}$ and $3.8 \times 10^5$ cell. L$^{-1}$, respectively). In 2015, Esmeraldas registered again the lowest concentrations in the first layers, comparing with the other sampling sites for the same year. The same vertical pattern was observed, as in 2014, with similar abundances from surface until 30 m (Figure 3).

Maximum of the year occurred in La Libertad at surface, in March ($1.1 \times 10^6$ cell. L$^{-1}$) and at 20 m depth in Puerto Bolivar in July (maximum value of $4.5 \times 10^5$ cell. L$^{-1}$). No peaks were registered during the year 2016, but data is not available for Esmeraldas and Puerto Bolivar. Thus, the maximum concentrations registered were observed in the first 20 m for Manta and La Libertad sites (Figure 3). The maximum abundances were found at surface and at 75 m depth for Manta (in February with $1.7 \times 10^5$ cell. L$^{-1}$ and $1.4 \times 10^5$ cell. L$^{-1}$, respectively) and at surface for La Libertad site (with maximum value for March with $1.0 \times 10^5$ cell. L$^{-1}$). The highest concentrations of the sampling period were reported in 2017 (Figure 3), mostly between January and June. The maximum abundance was found in Manta, in April at surface, $1.5 \times 10^6$ cell. L$^{-1}$ and at 10 m depth in Manta with an abundance of $4.0 \times 10^5$ cell. L$^{-1}$. In February, at the first 10 m layer, Manta and Esmeraldas sites registered also high abundance values ($\sim3.5 \times 10^5$ cell. L$^{-1}$ and $\sim2.5 \times 10^5$ cell. L$^{-1}$, respectively). The pattern of decreasing dinoflagellate concentration with depth, (very significant from 20 m onward) observed in the vertical abundances’ distribution for the rest of the study period, in 2017 is not so marked (Figure 3). In 2017, high abundances are observed in more deep layers (30–40 m) respecting to the

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1https://onemind-datascience.com/
| Order                | Family             | Species                                                                 | Occurrence (%) | 2013 | 2014 | 2015 | 2016 | 2017 |
|---------------------|-------------------|-------------------------------------------------------------------------|----------------|------|------|------|------|------|
| Dinophysiales       | Amphisoleniaceae  | Dinophysis bidentata Schröder, 1900                                    | 0.21           | +    | +    | +    |      |      |
|                     | Dinophysiaceae    | Dinophysis acuminata Claparède and Lachmann, 1859                      | 0.62           | +    | +    | +    | +    | +    |
|                     |                   | Dinophysis caudata Saville-Kent, 1881                                  | 1.46           | +    | +    | +    | +    | +    |
|                     |                   | Dinophysis cf. rotondata Claparède and Lachmann, 1859                  | 1.45           | +    | +    | +    | +    | +    |
|                     |                   | Dinophysis ovum Schutt, 1895                                          | 1.45           | +    | +    | +    | +    | +    |
|                     |                   | Dinophysis spp. Ehrenberg, 1839                                        | 2.56           | +    | +    | +    | +    | +    |
|                     |                   | Omothocercus quadratus Schutt, 1900                                    | 0.14           |      |      |      | +    |      |
|                     |                   | Omothocercus steini Schutt, 1900                                       | 0.42           |      |      | +    | +    |      |
|                     |                   | Omothocercus thunii (Schmidt) Kofoid and Skogsberg, 1928               | 0.14           |      |      | +    |      |      |
| Oxyphysaceae        | Phalacroma       | Phalacroma doryphorum Stein, 1883                                      | 0.07           |      |      |      |      |      |
|                     |                   | Phalacroma rapa Jorgensen, 1923                                        | 0.07           |      |      |      |      | +    |
| Gymnurales          | Ceratiaceae       | Tripos boehmi (H.W.Graham and Bronikovsky) F.Gómez, 2013               | 0.07           |      |      |      |      |      |
|                     |                   | Tripos breve (Ostenfeld and Schmidt) F. Gómez, 2013                    | 0.28           | +    | +    | +    |      |      |
|                     |                   | Tripos candelabrus (Ehrenberg) F. Gómez, 2013                          | 0.35           | +    | +    | +    |      |      |
|                     |                   | Tripos declinatus (G.Karsten) F.Gómez, 2013                            | 0.83           | +    | +    | +    |      |      |
|                     |                   | Tripos deflexus (Kofoid) F.Gómez, 2013                                 | 0.48           | +    | +    | +    | +    | +    |
|                     |                   | Tripos dens (Ostenfeld and Johannes Schmidt) F.Gómez, 2013             | 0.97           | +    | +    | +    | +    | +    |
|                     |                   | Tripos extensus (Gourret) F.Gómez, 2013                                | 0.07           |      |      |      |      |      |
|                     |                   | Tripos falcatus (Kofoid) F.Gómez, 2013                                 | 0.21           | +    | +    | +    |      |      |
|                     |                   | Tripos falciformis (Jorgensen) F.Gómez, 2013                           | 0.07           |      |      |      |      | +    |
|                     |                   | Tripos furca (Ehrenberg) F. Gómez, 2013                                | 4.85           | +    | +    | +    | +    | +    |
|                     |                   | Tripos fusus (Ehrenberg) F.Gómez, 2013                                 | 3.88           | +    | +    | +    | +    | +    |
|                     |                   | Tripos hexacanthus (Gourret) F.Gómez, 2013                             | 0.07           |      |      |      |      | +    |
|                     |                   | Tripos lunula (Schimper ex Karsten) F.Gómez, 2013                      | 0.07           |      |      |      |      |      |
|                     |                   | Tripos massiliensis (Gourret) F.Gómez, 2013                            | 0.35           | +    | +    | +    |      |      |
|                     |                   | Tripos muelleri Bory de Saint-Vincent, 1824 . F. Gómez, 2013           | 1.11           | +    | +    | +    | +    | +    |
|                     |                   | Tripos pentagonus (Gourret) F.Gómez, 2013 var. tenerum Jorgensen, 1920| 3.11           | +    | +    | +    |      |      |
|                     |                   | Tripos pulchellus (Schröder) F.Gómez, 2013                             | 0.21           |      |      |      |      | +    |
|                     |                   | Tripos spp. Tripos Bory de Saint-Vincent, 1823                         | 0.28           |      |      |      |      |      |
|                     |                   | Tripos strictus (Okamura and Nishikawa) F.Gómez, 2013                 | 0.07           |      |      |      |      | +    |
|                     |                   | Tripos subrobustus (Jörgensen) F.Gómez, 2013                           | 1.45           | +    | +    | +    | +    | +    |
|                     |                   | Tripos teres (Kofoid) F.Gómez, 2013                                   | 0.28           | +    | +    | +    |      |      |
|                     |                   | Tripos trichoceros (Ehrenberg) Gómez, 2013                             | 1.39           | +    | +    | +    | +    | +    |

(Continued)
| Order          | Family            | Species                                                                 | Ocurrence (%) |
|----------------|-------------------|-------------------------------------------------------------------------|----------------|
|                |                   |                                                                         | 2013 | 2014 | 2015 | 2016 | 2017 |
|                |                   |                                                                         | R   | D   | R   | D   | R   | D   |
| Ceratocoryaceae| Ceratocorys horrida (Schütt) Kofoid, 1910 | 0.14 | +   |     |     |     |     |
|                 |                   |                                                                         |     |     |     |     |     |     |
| Goniodomataceae| Triadinium polyedricum (Pouchet) Dodge, 1981 | 0.07 |     |     |     |     |     |
|                 |                   |                                                                         |     |     |     |     |     |     |
| Gonyaulacaceae  | Gonyaulax polygramma Stein, 1883 | 3.05 | +   | +   | +   | +   | +   |
|                 |                   |                                                                         |     |     |     |     |     |     |
|                  | Gonyaulax spinifera (Claparède and Lachmann) Diesing, 1866 | 1.11 |     | +   | +   | +   |     |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Gonyaulax spp. Diesing, 1866 | 9.83 | +   | +   | +   | +   | +   |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Gonyaulax turbinal Murray and Whitting, 1899 | 0.28 | +   | +   |     |     |     |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Lingulodinium polyedra (F.Stein) J.D.Dodge, 1989 | 0.07 |     |     |     |     |     |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Lingulodinium spp. D.Wall, 1967 | 1.52 | +   | +   | +   | +   | +   |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Heterodiniaceae   | Heterodinium spp. Kofoid, 1906 | 0.42 | +   |     |     |     |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Ostreopsisidaceae | Ostreopsis Halim, 1960 | 0.55 | +   | +   | +   | +   |
|                  |                   |                                                                         |     |     |     |     |     |     |
| Pyrophacaceae    | Pyrophacus steinii (Schiller) Wall and Dale, 1971 | 0.62 | +   | +   | +   | +   |     |
|                  |                   |                                                                         |     |     |     |     |     |     |
| Gymnodiniales    | Gymnodiniaceae     | Amphidinium carterae Hulburt, 1957 | 0.07 | +   |     |     |     |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Amphidinium spp. Claparède and Lachmann, 1859 | 0.48 | +   | +   | +   | +   | +   |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Cochlodinium spp. Schütt, 1896 | 1.04 | +   | +   | +   | +   | +   |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Gymnodinium catenatum H.W.Graham, 1943 | 3.12 | +   | +   | +   | +   | +   |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Gymnodinium spp. Stein, 1878 | 69.94 | +   | +   | +   | +   | +   |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Gymnodinium spirale (Bergh) Kofoid and Swezy, 1921 | 3.39 |     | +   | +   | +   | +   |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Gymnodinium spp. Kofoid and Swezy, 1921 | 52.7 | +   | +   | +   | +   | +   |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Gymnodinium acutum (Schütt) Kofoid and Swezy, 1921 | 24.72 | +   | +   | +   | +   | +   |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Margalefidinium catenatum (Okamura, 1916) F.Gómez, Richlen and D.M.Anderson, 2017 | 6.23 | +   | +   | +   | +   | +   |
|                  |                   |                                                                         |     |     |     |     |     |     |
| Kareniaceae      | Karenia spp. G.Hansen and Moestrup, 2000 | 0.07 |     |     |     |     |     |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Karenia mikimotoi (Miyake and Kominami ex Oda) Gert Hansen and Ø.Moestrup, 2000 | 0.28 | +   |     |     |     |     |
|                  |                   |                                                                         |     |     |     |     |     |     |
| Polykrikaceae    | Polykrikos kofoidii Chatton, 1914 | 0.07 |     |     |     |     |     |
|                  |                   |                                                                         |     |     |     |     |     |     |
| Noctilucales     | Noctiluca scintillans (Macartney) Kofoid and Swezy, 1921 | 1.32 | +   | +   | +   | +   | +   |
|                  |                   |                                                                         |     |     |     |     |     |     |
| Protodiniferaceae| Pronoctiluca rostrata F.J.R.Taylor, 1976 | 0.48 |     |     |     |     |     |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Pronoctiluca spp Fabre-Domergue, 1889 | 0.07 | +   |     |     |     |     |
|                  |                   |                                                                         |     |     |     |     |     |     |
| Peridiniales     | Corythodinium tesselatum (Stein) Loeblich Jr. and Loeblich, Ill, 1966 | 0.35 |     |     |     |     |     |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Corythodinium sp 1 Loeblich Jr. and Loeblich Ill, 1966 | 0.07 | +   |     |     |     |     |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Corythodinium sp 2 Loeblich Jr. and Loeblich Ill, 1966 | 0.07 |     |     |     |     |     |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Oxytoxum mediterraneum Schiller | 0.14 |     |     |     |     |     |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Oxytoxum scolopax Stein, 1883 | 9.56 | +   | +   | +   | +   | +   |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Oxytoxum sceptrum (F.Stein) Schröder, 1906 | 0.14 | +   |     |     |     |     |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Oxytoxum turbo Kofoid, 1907 | 2.63 | +   | +   | +   | +   | +   |
|                  |                   |                                                                         |     |     |     |     |     |     |
|                  | Oxytoxum spp. Stein, 1883 | 14.4 | +   | +   | +   | +   | +   |

(Continued)
| Order          | Family                      | Species                                           | Ocurrence (%) | 2013 | 2014 | 2015 | 2016 | 2017 |
|---------------|-----------------------------|---------------------------------------------------|---------------|------|------|------|------|------|
|               |                             |                                                   |               | R    | D    | R    | D    | R    | D    |
| Peridiniaceae |                             |                                                   |               |      |      |      |      |      |      |
|               |                             | Scrippsiella Balech spp. ex A.R.Loeblch III, 1965 | 12,12         | +    | +    | +    | +    | +    | +    |
|               |                             | Blepharocystis splendor-maris (Ehrenberg) Ehrenberg, 1873 | 0,21          |      |      |      |      | +    |      |
|               | Podolampadaceae             |                                                   |               |      |      |      |      |      |      |
|               |                             | Podolampas bipes Stein, 1883                      | 0,21          |      |      |      |      |      |      |
|               |                             | Podolampas palmipes Stein, 1883                   | 1,66          |      |      |      |      |      |      |
|               |                             | Podolampas spinifera Okamura, 1912                | 0,42          |      |      |      |      |      |      |
|               | Protoperidiniaceae          |                                                   |               |      |      |      |      |      |      |
|               |                             | Preperidinium meunieri (Pavillard) Eibraechter, 1900 | 0,21          |      |      |      |      |      |      |
|               |                             | Protoperidinium bispinum (Schiller, 1937) Balech, 1974 | 0,14          |      |      |      |      |      |      |
|               |                             | Protoperidinium brochii (Kofold and Swezy, 1921) Balech, 1974 | 1,94         |      |      |      |      |      |      |
|               |                             | Protoperidinium cassum (Balech, 1971) Balech, 1974 | 0,9           |      |      |      |      |      |      |
|               |                             | Protoperidinium compressum (Abé, 1927) Balech, 1974 | 0,9           |      |      |      |      |      |      |
|               |                             | Protoperidinium depressum (Bailey, 1854) Balech, 1974 | 0,35          |      |      |      |      |      |      |
|               |                             | Protoperidinium elegans (Cleve, 1900) Balech, 1974 | 0,62          |      |      |      |      |      |      |
|               |                             | Protoperidinium latapinum (Mangin, 1926) Balech, 1974 | 0,48          |      |      |      |      |      |      |
|               |                             | Protoperidinium longipes Balech, 1974             | 0,14          |      |      |      |      |      |      |
|               |                             | Protoperidinium oceanicum (VanHöffen, 1897) Balech, 1974 | 0,42         |      |      |      |      |      |      |
|               |                             | Protoperidinium pentagonum (Gran, 1902) Balech, 1974 | 0,42          |      |      |      |      |      |      |
|               |                             | Protoperidinium pedunculatum (Schüt, 1895) Balech, 1974 | 2,42         |      |      |      |      |      |      |
|               |                             | Protoperidinium quamerense (B.Schröder, 1900) Balech, 1974 | 2,22         |      |      |      |      |      |      |
|               |                             | Protoperidinium simulum (Paulsen, 1931) Balech, 1974 | 5,12          |      |      |      |      |      |      |
|               |                             | Protoperidinium steini (Jorgensen, 1899) Balech, 1974 | 0,28          |      |      |      |      |      |      |
|               |                             | Protoperidinium spp. Bergh, 1882                  | 15,93         |      |      |      |      |      |      |
| Prorocentrales| Prorocentaceae              |                                                   |               |      |      |      |      |      |      |
|               |                             | Prorocentrum compressum (J.W.Bailey) Abé J.D.Dodge, 1975 | 2,98         |      |      |      |      |      |      |
|               |                             | Prorocentrum dentatum Stein, 1883                 | 3,74          |      |      |      |      |      |      |
|               |                             | Prorocentrum gracile Schüt, 1896                  | 2,49          |      |      |      |      |      |      |
|               |                             | Prorocentrum lima (Ehrenberg) F.Stein, 1878        | 1,73          |      |      |      |      |      |      |
|               |                             | Prorocentrum mexicanum Osorio-Tafall, 1942        | 0,55          |      |      |      |      |      |      |
|               |                             | Prorocentrum micans Ehrenberg, 1834               | 12,88         |      |      |      |      |      |      |
|               |                             | Prorocentrum scutellum Schröder, 1900             | 0,55          |      |      |      |      |      |      |
|               |                             | Prorocentrum spp. Ehrenberg, 1834                 | 1,8           |      |      |      |      |      |      |
|               | Pyrocystales                |                                                   |               |      |      |      |      |      |      |
|               |                             | Pyrocystis fusiformis C.W.Thomson, 1876           | 2,08          |      |      |      |      |      |      |
|               |                             | Pyrocystis hamulus var. semicircularis Schröder, 1900 | 0,14         |      |      |      |      |      |      |
|               |                             | Pyrocystis lunula (Schutt) Schüt, 1896           | 0,21          |      |      |      |      |      |      |
|               |                             | Pyrocystis pseudonoccula Wyville-Thompson, 1876    | 0,35          |      |      |      |      |      |      |
|               |                             | Pyrocystis spp. Murray ex Haeckel, 1890           | 0,07          |      |      |      |      |      |      |
|               | Thoracosphaerales           |                                                   |               |      |      |      |      |      |      |
|               |                             | Goniodoma sp. Stein, 1883                         | 1,52          |      |      |      |      |      |      |
|               |                             |                                                   |               |      |      |      |      |      |      |
|               |                             | TOTAL                                             |               | 36   | 31   | 28   | 43   | 41   | 63   |
|               |                             |                                                   |               | 40   | 33   | 53   | 70   |
FIGURE 1 | Sampling sites; Esmeraldas (00°55'16.0"N; 80°06'24.0"O); Manta (00°52'45.0"S; 80°50'05.2"O); La Libertad (02°03'55.0" S; 81°07'15.0"O); Puerto Bolivar (03°06'10.0"S; 80°29'47.0"O).

other years (Figure 3) (i.e., at 30 m in May in Puerto Bolivar with $3.8 \times 10^5$ cell. L$^{-1}$ and at 50 m in April in Manta with $3.9 \times 10^5$ cell. L$^{-1}$).

Species Richness
Species richness values ranged from zero to a maximum of 21 species recorded in Puerto Bolivar during 2015 (August). Higher values were found in 0 m with a negative trend with depth (Figure 4). At surface waters (0 m) Manta and Puerto Bolivar showed the highest average values (median value of 5 for both sites), followed by Esmeraldas and La Libertad (median value of 4 for both sites). At 10 m depth the pattern is the same, except Puerto Bolivar that showed lower values (median of 4 species). Respect to the temporal variability, there is a general increase pattern along the study period, visualized as a predominance of observations above the average value across the time span of the study. In addition, along time series there are several periods were the species richness values showed the highest values. Those periods were between July to September 2013; the period between April to June 2014; February and March of 2015 in La Libertad; April and May of 2015 in Puerto Bolivar and Esmeraldas; the period of August and October in Puerto Bolivar where its maximum value was recorded; beginning of 2016, particularly for Puerto Bolivar and Manta and finally, most notably a general increase in richness values during 2017 starting in February and ending during May for the northern sites (Esmeraldas and Manta) and ending in September for southern sites (La Libertad and Puerto Bolivar). A new increase at the end of the study period (November – December) was evident for all sites. maximum richness was found in August 2015 in Puerto Bolivar at 10 and 75 m depth (20 and 21 taxa, respectively). Richness above 10 taxa were also observed the same year at same depths in La Libertad (February and March) and in 2017 at surface and 10 m in Manta (April), Puerto Bolivar (April, May, July and August) and Esmeraldas (May) (Figure 4).

Dinoflagellate Diversity
Diversity (Shannon–Wiener index) values ranged from 0 to 2.35 with southern stations (Puerto Bolivar and La Libertad) showing the highest values (2.21 and 2.35, respectively). In addition, higher richness values were found in surface waters across sampling stations (probably influenced by the richness gradient, see previous section) and along the sampling period. However, in some occasions high richness values were found in deeper waters during February to March 2013 (mostly for Manta and Puerto Bolivar), during the period between February to May of 2014 (except for Manta), between February to May 2015, between August to October 2015 and since the year 2017 where a general increase in diversity index was found. This general increase for 2017 showed a maximum for all sites during April – May (Supplementary Figure S2).

Harmful Algae
A total of 8 potential harmful dinoflagellate species were observed in low abundances belonging to 4 orders and
6 families, *Alexandrium* sp, *Cochlodinium catenatum*, *Dinophysis acuminata*, *Dinophysis ovum*, *Gymnodinium cf. catenatum*, *Karenia* sp., *Prorocentrum lima* and *Prorocentrum mexicanum*. The taxa that showed the highest occurrence were *C. catenatum* and *G. cf. catenatum* (6 and 3%, respectively).

The highest cell abundance was recorded by *C. catenatum* in May and June 2014 ($7.5 \times 10^4$ and $9.6 \times 10^4$ cell. L$^{-1}$) in Puerto Bolivar. In general, the species was present in southern stations. Contrary, *G. cf. catenatum* was registered more times in northern stations, mainly in Esmeraldas, maximum abundances were observed in February 2013 ($4 \times 10^4$ cell. L$^{-1}$). *P. lima* had an occurrence of 1.5%, the highest abundance was recorded in February – March 2015 at Puerto Bolivar at surface ($9.1 \times 10^4$ cell. L$^{-1}$). *Alexandrium* spp, *Karenia* spp, *P. mexicanum*, *D.
acuminata, D. ovum, recorded the lowest occurrences (<1%) (Table 1).

Environmental Variables
Temperature and Salinity
In general, water temperature was higher in Esmeraldas compared to the rest of the stations during the studied period with a maximum in surface of 28.86°C in May 2017 (Figure 5A).

Concerning temporal variability, lowest temperatures (<22°C) registered in the first 10 m of the water column, occurred in 2013 from June to August in Puerto Bolivar, August and November in La Libertad, March and September in Manta and March in Esmeraldas (Figure 5A).

The next 2 years, 2014 and 2015, lowest temperatures were observed in February and March in Puerto Bolivar and La Libertad in surface and 10 m while in Esmeraldas and Manta there were also present at 20 m. In Puerto Bolivar and La Libertad
the lowest values occurred in September in all depths of the water column (Figure 5A).

In 2016, lowest temperatures were reported in March in Manta for the first 20 m and in April in Puerto Bolivar from 10 m (Figure 5A).

During 2017, lowest temperatures were registered in February and March for Esmeraldas, Manta and La Libertad, while in Puerto Bolivar there were also registered in April (Figure 5A).

Contrary, higher temperatures (>25°C) registered in 2013 occurred from April to December in the first 40 m in Esmeraldas and Manta, however, in La Libertad and Puerto Bolivar, warmer waters were limited to the first layers, 20 and 10 m, respectively (Figure 5A).

In 2014 and 2015, Esmeraldas and Manta registered warmer waters in the 50 m of the water column from April to December, evident in La Libertad and Puerto Bolivar (Figure 5A).

The thermocline was deeper in northern stations, Esmeraldas and Manta (between 30 and 50 m), while in La Libertad and Puerto Bolivar it was between 10 and 30 m (Figure 5A). Annual vertical profiles of monthly mean temperature are available in Supplementary Figure S3.

Lower salinity values (<32) were observed mainly in Esmeraldas in 2013, 2014, and 2017. In the other stations, values were between 33 and 34 during the studied period in the first 30–40 m of the water column. Saltier water layers (>35.00) were present in deeper layers (between 40 and 100 m) with the exception in 2015 and 2016 where in February, April, August and October, higher salinities were observed up to 20 m (Figure 5B). Annual vertical profiles of monthly mean salinity are available in Supplementary Figure S4.

Nutrients and Dissolved Oxygen

Regarding nutrients, highest nitrate concentrations (5 to 27.6 µg-at-L⁻¹) were registered from 20 to 50 m, mainly during the warm season (February and March) and dry season (From July to October) in all sampling sites (Figure 6A). Lowest nitrate concentrations (<1 µg-at-L⁻¹) were observed in surface, up to 30 m and there was not a seasonal trend among sampling sites. In general, Esmeraldas registered lowest values compared with the rest of sampling sites. Annual vertical profiles of monthly mean nitrate are available in Supplementary Figure S5.

Concerning phosphates, highest concentrations (>1 µg-at-L⁻¹) were registered in 2013 in Manta and Esmeraldas in the first 50 m of the water column in February and March and between 30 and 50 m from May to September. For the rest of the years, higher values decreased between 0.3 and 1.9 ug-at/l and no seasonal pattern was observed among stations (Figure 6B).

Lowest phosphate concentrations (<0.2 µg-at-L⁻¹) were mainly observed in Esmeraldas without a particular pattern. Manta also registered low values but less frequency. Annual vertical profiles of monthly mean phosphate are available in Supplementary Figure S6.

Moreover, high silicate concentrations (>5 µg-at-L⁻¹) were registered in 2013, 2014, and 2017, with a slightly variability among sampling sites (Figure 6C). Annual
Concerning dissolved oxygen, values in all sampling sites were between 4 and 5 mL L\(^{-1}\) in the first 30–40 m approximately in Esmeraldas and Manta, while in the southern sites, this layer was found in higher depth (between 20 and 30 m). More oxygenic waters (>5 mL L\(^{-1}\)) were found occasionally in surface and 10 m layer in February and March 2013 and 2017, between April and June 2016. Lower values of dissolved oxygen (<2.5 mL L\(^{-1}\)) were observed between 40 and 50 m (Figure 6D).

Annual vertical profiles of monthly mean silicates are available in Supplementary Figure S8.

### Relationship With Environmental Variables

For those species with a percentage of occurrence higher than 2.8% the association between the environmental variables and the structure of dinoflagellates was inspected through CCA per year. Results showed for year 2013 a significant proportion of total constrained inertia (TCI) explained by the environmental variables (TCI = 8%, \(p\)-value < 0.05) with the first two axis explaining more than 80% of this constrained inertia (Table 2). Permutation showed that salinity, phosphate and temperature significantly affected the community structure for this year (Table 3). The community composition (CCA analysis) showed that the species, *Protoperidinium simulum* and *Prorocentrum compressum*, were positively correlated with salinity and with N/P (this last being non-significant). The taxa *Oxytoxum sp* and *Protoperidinium sp*. were positively associated with high values of nitrite. The group formed by *Gymnodinium cf. catenatum*, *Oxytoxum turbo* and *P. micans* were positively associated with high values of nitrate. The group formed by *Gymnodinium cf. catenatum*, *Oxytoxum turbo* and *Gonyaulax polygramma* showed a negative association with this variable. Finally, the
group formed by the species *Gonyaulax sp*, *Gymnodinium sp*, *Tripos fusus* and *Tripos furca* were negatively associated with nitrite and phosphate, but showed an unimodal response with the rest of the variables (a central position in the CCA-Triplot, Figure 7A).

For year 2014 a lower than previous year, but still significant proportion of TCI explained by the environmental variables (TCI = 5.8%, \(p\)-value < 0.05) with the first two axes explaining 78.8% of this constrained inertia (Table 2). For this year (same as 2013) salinity, temperature and phosphate significantly affected the community structure (Table 3). Moreover, a clear positive correlation between phosphate with nitrite and N/P (both being non-significant) and those three variables showed a negatively association with temperature (Figure 7B). The association of species with environmental variables showed that *Dinophysis sp* was positively associated with N/P and nitrite – phosphate variables. With respect to salinity *Oxytoxum turbo* was positively associated whereas *T. fusus*, *O. scolopax* and *P. simulum* were negatively associated. *T. furca* and *C. catenatum* (in a lesser extent) showed a positive association with temperature. The rest of the species showed a central position in the CCA-Triplot representing unimodal response to the environmental variables (Figure 7B).

For year 2015 the proportion of TCI explained by the environmental variables was also significant (TCI = 5.8%, \(p\)-value < 0.05) with the first two axes explaining the highest amount of this constrained inertia along the study period (89.4%, Table 2). During 2015 the temperature, the N/P ratio and salinity were those variables which significantly explained the dinoflagellate's community structure (Table 3). A positive relationship was observed among salinity and nitrite and phosphate and a negative association between temperature and N/P (Figure 7C). From the species-specific relationship, the CCA analysis revealed a positive association between *Dinophysis sp* and *P. simulum* with N/P ratio and negatively with temperature. The species *Gymnodinium cf. catenatum* was associated with positive values of nitrite and phosphate and in a lesser extent, showing a non-linear response, with salinity. Finally, the species *P. micans*, *P. compressum* and *T. furca* were associated to low values of salinity (negative relationship) (Figure 7C). Interestingly, this year none species was associated with high, but low temperature values.

For year 2016 and 2017 a decrease in the TCI from 7.8% to 5.5 was evident. However, for both years the model was able to significantly explain this constrained inertia with the first two axes explaining 82.6% and 65.1% of this constrained inertia.
TABLE 2 | Summary table of canonical correspondence analysis (CCA) statistics per year.

| Year | n   | TMI | TCI (%) | F-statistic (p-value) | CI-1 (%) | CI-2 (%) | CI (%) |
|------|-----|-----|---------|-----------------------|----------|----------|--------|
| 2013 | 223 | 4.9 | 8.0     | 3.82 (<0.05)          | 52.5     | 27.9     | 80.4   |
| 2014 | 274 | 4.1 | 5.8     | 3.32 (<0.01)          | 58.2     | 20.6     | 78.8   |
| 2015 | 271 | 4.2 | 6.9     | 3.96 (<0.01)          | 64.5     | 24.9     | 89.4   |
| 2016 | 179 | 1.95| 7.8     | 2.93 (<0.01)          | 52.6     | 30.0     | 82.6   |
| 2017 | 309 | 1.82| 5.5     | 3.55 (<0.01)          | 45.1     | 20.0     | 65.1   |

The total number of observations (n), the total model inertia (TMI), the proportion of total constrained inertia explained by the environmental variables [TCI (%)], the model significance at $\alpha = 0.05$ (p-value), the proportion of constrained inertia explained by axis 1 [CI-1 (%)] and by axis 2 [CI-2 (%)], and the total proportion of constrained inertia explained by the first two axes [CI (%)] is showed for each year.

inertia, respectively (Table 2). Particularly during 2016 the environmental variables who were significantly associated with the structure of the dinoflagellate community were temperature, nitrite and phosphate (Table 3). Salinity and phosphate were positively correlated and temperature and nitrite were negatively correlated. The CCA analysis for this year showed that Gymnodinium cf. catenatum was correlated positively with salinity and negatively with nitrite. The species Gymnodinium acutum was associated with high values of phosphate with a non-linear response (center position of the species related with the environmental vector). The species C. catenatum, O. scolopax, Scripsiella sp, G. polygramma, T. furca, Protoperidinium sp and Gymnodinium sp. showed a negative association with phosphate and Oxyttoxum sp, Gymnodinium spirale, G. polygramma, Scripsiella sp, O. scolopax, and Gymnodinium sp a negatively association with nitrite (Figure 7D). During 2017 all the environmental variables were significant when related to the observed dinoflagellate community structure (Table 3). Among the environmental variables, a negative association was evident for salinity and phosphate (Figure 7E). From the community composition perspective, the species P. simulum, T. fusus, Gymnodinium cf. catenatum, and Prorocentrum dentatum showed a clear positive relationship with nitrite and negatively with salinity and with the N/P ratio. The species T. fusus, P. compressum, O. scolopax, and Gymnodinium sp were positively associated with phosphate and negatively with salinity. For this year, non-species were related positively with temperature, but some species showed a negative association with this environmental variable (e.g., Oxyttoxum turbo, Gymnodinium spirale, Oxyttoxum sp, and Gymnodinium sp).

Interestingly, the total model inertia (TMI), which represents the total variability n-dimensional inside the model, remained more or less constant for the first 3 years (2013–2015) ranging from 4.1 to 4.9. However, during years 2016 and 2017 this TMI was significant lower with values for 2016 of 1.95 and for 2017 a TMI of 1.82 (Table 2).

DISCUSSION

Dinoflagellate Community

In this study, there were reported a total of 97 taxa, corresponding to 8 orders, 22 families, and 31 genera during the sampling period from 2013 to 2017 in 4 stations in the coast of Ecuador at different depths. Phytoplankton dynamics across the ETP coast is more characterized in other countries and there is a lack of information in the central area corresponding to the Ecuadorian coast. Interestingly, recent studies regarding phytoplankton distribution in response to ENSO events have been published (Conde and Prado, 2018; Conde et al., 2018), but, unfortunately, species identification is not indicated as a result. Another recent study explored the oceanography of red tides using remote sensing data from 1997 to 2017, confirmed that potential HABs have been dominated by dinoflagellates during wet season mostly at the Gulf of Guayaquil (Borbor-Cordova et al., 2019). Furthermore, dinoflagellates abundance reported in the present study was not high and the number of proliferations was low, only Gymnodinium sp exceeded $10^6$ cell. L$^{-1}$ in April 2017. In the coast of Ecuador, INOCAR and other researchers have reported blooms of dinoflagellates, for example Gymnodinium sp was reported in 2003, 2004, 2005 reaching high concentrations ($1 \times 10^5$ cell. L$^{-1}$ – $1.1 \times 10^7$ cell. L$^{-1}$) in the Gulf of Guayaquil both in wet and dry season (Torres and Tapia, 2002; Torres et al., 2017).

It is important to mention that species identification resulted difficult to perform without specific equipment and adequate formation. In that sense, some of the taxa found in the area were not identified at species level. Dinoflagellates identification under inverted microscope based on morphological features constitutes a challenge since some species from several genera share the same plate pattern and overlap in size (e.g., the genus Ostreopsis – Carnicer et al., 2016). In the last decades, molecular techniques have allowed to perform accurate identifications in dinoflagellates using rDNA sequences (Litaker et al., 2007). Unfortunately, there is no molecular characterization of marine dinoflagellates in the Ecuadorian coasts, with the exception of Ostreopsis cf. ovata (Carnicer et al., 2016). It is important for Ecuadorian institutions to invest in the implementation of molecular techniques for future investigations, especially important to identify potentially toxic species.

Higher species richness in the water column was observed in surface layers, with a maximum of 21 taxa. These results are in agreement with a study performed in 2015 at the southern coast of the country, in El Oro province, where they reported species richness average between 28 and 23 dinoflagellates in surface samples (Prado et al., 2015; Conde et al., 2018). The time series length of the present study avoids us to conduct a formal time series
Analysis to statistically prove this increase pattern observed. In addition, some oscillatory behavior is evident and probably associated with the interannual variability pattern (Conde et al., 2018).

In 2015, associated with El Niño ENSO event, higher dinoflagellate species richness values were registered during the wet season in La Libertad, Puerto Bolivar and Esmeraldas, reaching a maximum of 28 species in dry season in Puerto Bolivar, that could be linked with upwelling and higher nutrients concentrations (Borbó-Cordova et al., 2019). In 2017, higher values are also observed in each station with the highest found in Puerto Bolivar in agreement with Prado et al. (2015), associating the event with the presence of the Humboldt current. The higher amount of species during El Niño ENSO events can be linked with the interannual variability pattern analysis to statistically prove this increase pattern observed.

Table 3

| Variable               | 2013 F-statistic | p-value | 2014 F-statistic | p-value | 2015 F-statistic | p-value | 2016 F-statistic | p-value | 2017 F-statistic | p-value |
|------------------------|------------------|---------|------------------|---------|------------------|---------|------------------|---------|------------------|---------|
| Temperature (Anomaly)  | 5.78             | <0.01   | 4.22             | <0.01   | 6.76             | <0.01   | 2.00             | <0.05   | 3.57             | <0.01   |
| Salinity (Anomaly)     | 3.80             | <0.05   | 5.47             | <0.01   | 5.66             | <0.05   | 1.25             | n.s.    | 4.19             | <0.01   |
| Phosphate              | 5.04             | <0.01   | 3.78             | <0.01   | 1.74             | n.s.    | 5.13             | <0.01   | 4.6              | <0.01   |
| Nitrite                | 0.91             | n.s.    | 0.82             | n.s.    | 0.31             | n.s.    | 2.40             | <0.05   | 4.61             | <0.01   |
| N/P                    | 1.44             | n.s.    | n.s.             | n.s.    | 3.60             | <0.05   | 1.99             | n.s.    | 3.67             | <0.05   |

Significant environmental variables (terms) are showed in bold and the value of the F statistic (p-value) are also given. n.s., means non-significant term.

Environmental Variables and Seasonality

Overall, in this study, SST and salinity anomalies, nutrients, as well as the N/P ratio were strongly associated to the spatio-temporal distribution of dinoflagellate community (Figure 7). Studies elsewhere have found that HAB have been related during positive anomalies of SST, stratification with a deeper thermocline, pulses of upwelling during summer or dry season and the biological interaction among phytoplankton community (Hallegaard, 2010; Diaz et al., 2016), some of these conditions were found in this analysis.

The environmental variables presented in this study confirm the main patterns of current systems on the ETP, which are driven by cold Humboldt Current in the central and south coast of Ecuador (La Libertad and Puerto Bolivar), associated with the equatorial upwelling (EU) system and the warm Panama current toward the north of Ecuador (Esmeraldas), these oceanographic settings drives the biogeochemical conditions that may promote extensive phytoplankton growth and potential HABs (Cucalan, 1989; Pennington et al., 2006). The north of the Ecuadorian coast is under the influence of the inter-tropical convergence zone (ITCZ), receiving higher levels of precipitation as well as runoff from Panama Gulf generating a low salinity and warmer temperatures which lead to stratified conditions, shallow halocline-nutricline (30–40 m), and low nutrients concentration (Pennington et al., 2006). Those conditions are reflected in the northern station of Esmeraldas, where minimum dinoflagellate abundances were reported (Figure 2 and Supplementary Figure S2) compared to other stations, highlighting the influence of the oceanographic characteristics and strong seasonality of the ETP (Chavez et al., 2002).

Oceanographic studies have established the seasonal pattern of the coastal EU at the center of Ecuador, and the upwelling from the Humboldt Current on the south of the Ecuadorian Coast (Pennington et al., 2006). Seasonality of those upwelling system are reflected in all stations (based on higher nutrient concentrations and lower temperatures) except at Esmeraldas, being deeper and high nutrient concentration during wet season (December to May) but also between August to September. It is expected that Manta is mostly influenced by the EU while at La Libertad and Puerto Bolivar it is the upwelling of Humboldt Current, considered one of the most productive of
FIGURE 7 | Canonical correspondence analysis triplot for (A) 2013, (B) 2014, (C) 2015, (D) 2016, and (E) 2017. Sampling stations are shown as triangles with color-code according to the sampling area (figure legend). The species (with occurrence >2%) present in the community CCA analysis follows: Otu.
In this study, highest concentrations of dinoflagellates were found in Manta (April 2017), La Libertad (March 2015), and Puerto Bolivar (May 2014 and May 2017), during wet season. During this period, there is a large amount of nutrients coming with the intense runoff from the agricultural Guayas Basin (Borbor-Cordova et al., 2006) and interestingly, during the period of this study it is reported higher levels of nutrients, mainly nitrate and phosphate (Supplementary Figures S5 in the present study), and interestingly, in the present study it is reported higher levels of nutrients, mainly nitrate and phosphate (Supplementary Figures S5, S6), coming from deeper water, suggesting advection from the coastal Humboldt Upwelling in dry season with peaks on August. Thus, data on this study suggests that a combination of extreme hydrodynamic conditions, climate variability associated to the warm phase of the Pacific Decadal Oscillation (PDO) and ENSO in the coast of Ecuador (2015–2016), influencing salinity and SST anomalies and nutrient limitation conditions, led to shifts in the phytoplankton community and maybe to HABs.

In this region phytoplankton productivity and growth are influenced by the upwelling dynamics which can be associated to the pycnocline-nutricline-oxygen depth, and their variability along the coast is driven by local specific conditions of coastal waves, seasonality and inter-annual variability of ENSO cycle (Montecino and Lange, 2009). The thermo-nutricline determines the supply of limiting nutrients (related to N/P ratio) to the euphotic zone and influencing the phytoplankton community assemblage during each ENSO event. This is evidenced in the present study with the inter-annual variability of dinoflagellates associated to ENSO events which is characterized by high positive anomaly of SST, changes on the timing of seasonality and strengthening (cold phase) or weakening (warm phase) the coastal upwelling (Pizarro and Montecinos, 2004; Pennington et al., 2006; Montecino and Lange, 2009). Similar dynamics have been reported in North and ETP during ENSO events (Chavez et al., 2002; Jacox et al., 2016). Previous ENSO studies pointed out that southwest winds weakness lead to a coastal upwelling decline which diminish phytoplankton blooms occurrence (Tam et al., 2008; Wang et al., 2008; Montecino and Lange, 2009). However, during warm phase ENSO event in 2015 occurred anomalous conditions related with sustained wind stress, intermittent coastal upwelling, that could be the reason why dinoflagellates were able to persist even in a nutrient limited environment, and warm stratified conditions (Smyady and Reynolds, 2001; Du et al., 2015; McCabe et al., 2016; McKibben et al., 2017; Conde and Prado, 2018).

During the period of this study (2013–2017), 2013 is considered a normal year with the lower abundances of dinoflagellates of the study, a weak ENSO warm phase in 2014, followed by a strong warm phase ENSO in 2015–2016 (with a different seasonality of the one in 1997–1998) beginning a transition to cold phase ENSO during the dry season 2016 is extended to 2017 characterized by abundance of dinoflagellates (see Figure 3). Each year of the study develop specific environmental conditions; however, SST anomalies, salinity and nutrients persistently drove the dinoflagellates community for all the study period (see Figure 7).

In 2013 a normal year with a slightly cold SST anomaly (see Supplementary Figure S9), lower salinities, and nutrients were the drivers for the dinoflagellates. Gymnodinium cf catenatum, Oxytoxum turbo and P. micans were associated to less saline, low nutrients and warm waters typical conditions during wet season in Esmeraldas influenced by the Panama Current. While P. simulum, P. compressum, Oxytoxum sp, and Protoperidinium sp are positively related to nutrient and salinity which are characteristics of upwelled waters, suggesting a preference of nutrient-rich water along the coast of Ecuador (Figure 7A).

In 2014, considered a weak ENSO year with ONI strong anomalies (>1.5) during wet season decreasing rapidly toward December 2014 (see Supplementary Figure S9). T. fusus, O. scolopax and P. simulum were associated to low salinity conditions mostly on Esmeraldas, Manta and La Libertad suggesting in/Out of freshwater runoff and precipitation. While Dinophysis sp was associated to nutrients from upwelled waters from EU system at the Esmeraldas station, the upwelling process is related with the nitrate distribution at 30–50 m (Supplementary Figure S5). T. furca and C. catenatum were associated to positive anomalies of temperature, suggesting that have adapted to the warm conditions on the Esmeraldas site.

Although the SST anomalies of ENSO events in 2015 were of similar magnitude to those in 1997–1998, the responses were diverse in relation to the oceanographic characteristic of the equatorial front (Conde et al., 2018). By April 2015, the thermocline deepened to 40–75 m until December 2015 generating stable thermal stratification, but nutrients were sustained by a weak upwelling and a pycnocline-nutricline between 30–50 m. Even though the nutrients were reduced at the surface layer (see Figures 6, 7), surprisingly the abundance levels of dinoflagellates in La Libertad and Puerto Bolivar where high between 10–30 m (see Figure 3). By August 2015, nutrients (nitrate, and phosphate) were upwelled between 30 and 50 m at La Libertad, and Puerto Bolivar. These conditions of thermal stratification, deepened thermocline, slightly upwelling, light for growth and ciliate prey (Mesodinium rubrum) appear to establish the optimal combination for the development of potential HABs such as Dinophysis sp and P. simulum (Reguera et al., 2012; Diaz et al., 2013). G. catenatum have affinity for the nitrate and phosphate and less with salinity, bloom of this specie has been
related to advected vegetative populations to the coasts during the relaxation of coastal upwelling (Bravo and Figueroa, 2014). *P. micans, P. compressum,* and *T. furca* were associated to less saline water characteristic of the Panama current or influenced by precipitation at the northern stations of Esmeraldas and Manta. Considered an extreme ENSO year and warm PDO, expecting a limited algal growth, surprisingly there was a relative high abundance of dinoflagellate and higher richness reflected in the diversity of species (21). If this increase in richness is a pattern of the ENSO need to be verified in other years.

Year 2016 shows the transition between a warm ENSO to cold and normal conditions, with ONI decreasing from 2.5 in January to – 0.7 in September in the dry season (see Supplementary Figure S9). In this year, *Gymnodinium cf. catenatum* appeared during dry season associated with salinity and inversely with nitrite at the Manta and La Libertad, unfortunately no data in Esmeraldas and Puerto Bolivar are available. *Gymnodinium cf. catenatum* has been recognized to cause paralytic shellfish poisoning across regions and as a survivor by using strategies based on its motile forms and cysts in plankton assemblages and in surface sediments (Yamamoto et al., 2004; Bravo and Figueroa, 2014). *Gymnodinium cf. catenatum* is also identified as one of ten most damaging potential domestic target species, considering the environmental and economic impact in aquaculture and also in human health (Hallegraeff, 2003). *C. catenatum* is another specie that seems to be associated to the upwelling at La Libertad in dry season.

During 2017, persistent negative SST anomalies associated to upwelling and water nutrients richness lead to an increase in the productivity of the region in all the ETP (Brown et al., 2015). Concentrations of dinoflagellates increased and reached their maximum in all the stations due to conditions of a persistent upwelling specially in Puerto Bolivar station. Some of the specie that were associated with upwelled nutrients were *P. simulum, T. furca, Gymnodinium cf. catenatum* and *P. dentatum*.

In the present study, the most abundant specie were *Gymnodinium sp* and *Gyrodiunum sp* which were present every year in both seasons of the studied period and registered up to 40 m depth. Those specie correspond to free living large cell dinoflagellates, ubiquitous in the Pacific Ocean, previously registered in México, California and Ecuador (Gomez, 2005; Meave-del Castillo et al., 2012; Torres, 2015). Both species appeared in all coastal stations during the seasonal transition in the weak ENSO in 2014 and strong ENSO in 2015 (Figure 2 and Table 1) indicating their capacity to adjust to warm and deeper thermo-nutricline.

Even though some microalgae exhibit a rapid adaptability to high temperatures in short-term experiments, the physiological plasticity and genetic response of many microalgae under future environmental conditions is unknown (Hallegraeff, 2010). Organism size and elemental composition of the phytoplankton community will influence processes at the level of individuals, populations, communities and ecosystems (Finkel et al., 2010). Some researchers have found that dinoflagellates are species with a smaller surface and have the ability to grow and sustain during ENSO, ocean heatwaves, stratified conditions, and nutrient poorer waters (Smayda and Reynolds, 2003; Glibert et al., 2005). However, there is also evidence that nutrient enriched waters can stimulate dinoflagellates blooms (Glibert et al., 2005), therefore, dinoflagellates are considered strategist and survivors either in extreme conditions.

The contrasting behavior of *Tripos* species in divergent lake types (with different climatic, morphometric, geological, hydrological, and trophic features) explains the existence of ecotypes of these species adapted to diverse environmental conditions and exhibiting high intra- and inter-population and morphological variability (Cavalante et al., 2016). *T. furca* is also known to perform active vertical migration, depending on nutrients and water temperature in both natural and laboratory conditions, which means that different species of dinoflagellates have different ecological abilities. These abilities are possibly linked with their ecological responses for surviving under local environmental conditions. *T. furca* has a competitive advantage because of physiological adaptations to low nutrient concentration waters (Baek et al., 2011). Therefore, dinoflagellates can alter their vertical position in the water column by swimming, which allows them to maintain an optimal depth in terms of light or nutrients. Consequently, the actual duration of high irradiance in the coastal bays is considered to regulate the maintenance of the bloom (Baek et al., 2011).

In general, under N or P limitation or some specific N/P ratio, some toxic dinoflagellates are probably outcompeted, and toxin production may be an adaption strategy to offset the ecological disadvantages of dinoflagellates with low nutrient affinity (Smayda, 1997). How these biochemical shifts shape phytoplankton community structure and the presence of potentially toxic species need to be explored in Ecuador.

Montecino and Lange (2009) proposed that during warm ENSO as 2015, there is microbial trophic web path mostly dominated by small-sized phytoplankton, in this study taxa such as *Gonyaulax polygrama, P. dentatum* and *Gymnodium sp.* were characterized to be trophic web dominated. Still it is not very clear which are the main ecological drivers to explain interannual variability of phytoplankton community structure, including toxin producer species. In addition, mixotrophy relationships need to be explored because many of the motile bloom formers are mixotrophic protists (Stoecker et al., 2017). Some argued that PDO is a good indicator to explain those shifts on the community structure and potential toxins generation (Du et al., 2015). Unfortunately, in this study only dinoflagellates community was considered, so it is not possible to confirm this ascertain. An effort in future works need to be done in order to encompass more taxa to better understand phytoplankton distribution in the area.

**CONCLUSION**

Dinoflagellates abundances reported in the present study were not elevated, and no HABs were observed in the studied period. The taxa presented in this study represents a baseline in which future work may referred in order to understand certain dinoflagellate dynamics. In synthesis, the environmental variables and the oceanographic ENSO index served to understand how the current system of the ETP, their thermocline
dynamics, and the coastal upwelling affect eutrophic zone nutrient supply and hence, dinoflagellates abundance and richness along the coast of Ecuador.

Moreover, toxic species were detected and, considering that in future decades it is predicted an increase in seawater temperature and extreme events frequency (IPCC, 2014), phytoplankton dynamics may be influenced. In order to be able to predict future scenarios of HABs, it is important to study their distribution in relation to environmental variables in areas where extreme events may affect their proliferations. Thus, it is crucial to involve policymakers and stakeholders in the implementation of environmental policies and climate change adaptation strategies involving long-term monitoring and sanitary programs, not covered at present.

AUTHOR CONTRIBUTIONS

GT, SR, RN, and EP made substantial contributions to conception and design of the work and conducted the sampling, nutrient, and dinoflagellates analysis. OC, GT, MB-C, PDLF, and AC made substantial contributions in data analysis and interpretation and participated in drafting the article. MB-C gave the final approval of the version to be submitted.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmars.2019.00145/full#supplementary-material
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**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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