We screened African wild dogs (*Lycaon pictus*) in Kruger National Park, South Africa, for *Mycobacterium bovis* infection using an interferon-gamma release assay. We detected *M. bovis* sensitization in 20 of 21 packs; overall apparent infection prevalence was 83%. These animals experience high infection pressure, which may affect long-term survival and conservation strategies.

The African wild dog (*Lycaon pictus*) is an endangered carnivore occurring in fragmented, small populations (in South Africa, <500 animals). These factors make them susceptible to adverse factors, such as infectious diseases, that may threaten their long-term survival (1,2). Of particular concern are diseases caused by multihost pathogens that are capable of persisting in reservoir host species, such as *Mycobacterium bovis*, the causative agent of bovine tuberculosis (bTB). This pathogen may pose a major threat to the conservation of endangered host populations (3).

Since 2012, sporadic cases of wild dogs with macroscopic and histological lesions consistent with tuberculosis (TB) have been recorded in South Africa, specifically in Kruger National Park (KNP; *n* = 8), uMkuze Game Reserve (n = 1), and Hluhluwe-iMfolozi Park (HiP; *n* = 2). *M. bovis* infection is endemic in these parks and occurs in multiple species that are preyed upon by wild dogs, such as warthogs, which have an estimated *M. bovis* seroprevalence up to 38% in KNP (4,5). In 2 cases from KNP, acid-fast bacilli were associated with granulomatous lymphadenitis, and spoligotype analysis of *M. bovis* isolates from lesions in affected wild dogs from KNP (strain type SB0121) and HiP (strain type SB0130) were the same as those found in local prey (6).

*M. bovis* is a novel pathogen of wild dogs; understanding the impact of bTB disease in wild dogs is imperative to making informed management decisions regarding these animals’ conservation. Estimation of prevalence would provide a starting point for this investigation but requires diagnostic tools for accurate detection of *M. bovis* infection. To estimate prevalence in the KNP wild dog population, we assessed sensitization to TB antigens ESAT-6 and CFP-10.

During July 2016–January 2018, we tested blood samples from 77 wild dogs from KNP using an interferon-gamma release assay (IGRA) developed by our group (7). We tested animals from 21 wild dog packs; 20 of these included ≥1 IGRA-positive animal, indicating widespread exposure to *M. bovis* throughout KNP (Figure). We observed no significant difference in IGRA results based on sex (p = 0.79 by 2-tailed Mann-Whitney test). Overall, the apparent prevalence of *M. bovis* infection was 82% (63/77; 95% CI 72%–89% by modified Wald test).

Few reports of active bTB disease and related deaths have been documented in wild dogs, so the high apparent prevalence in this study merits further investigation.
The prevalence estimate in wild dogs was based on detection of immune sensitization to *M. bovis*–specific antigens. Although the infection status of a small number of animals was confirmed by antemortem mycobacterial culture of oropharyngeal swabs (4 [7.0%] of 57 wild dogs tested), we could not confirm the infection status of all of the animals included in this study. Furthermore, during the time of the study, most animals included in this analysis appeared to be healthy. Therefore, further investigations will be required to clarify the progression of *M. bovis* infection and risk of bTB disease in this species.

These results have implications for managing the wild dog metapopulation in South Africa, which involves translocation of animals across the country to maintain genetic diversity and to achieve conservation goals (2). The risk of introducing *M. bovis* into previously uninfected areas by an infected wild dog is unknown, and studies on transmission will be crucial in assessing this risk.

Survival of a species is affected by a complex array of factors, of which disease is only one. Currently, the KNP wild dog population appears to be stable (2), despite the apparent high prevalence of *M. bovis* infection. Favorable conditions, such as abundance of prey, may support high reproductive rates. However, with environmental changes, such as prolonged drought, the vulnerability of host populations to infectious disease may be more pronounced (10).

In conclusion, this study shows widespread exposure of wild dog packs to *M. bovis* in KNP, with high infection pressure to individual dogs. Although the impact of disease on population numbers is unknown, wild dogs infected with *M. bovis*, even young animals, have been observed to have generalized disease leading to death. Further investigations into the progression of *M. bovis* infection, the risk for transmission, and the probability of developing progressive disease are needed to assess the threat of this emerging disease in African wild dog populations.

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**About the Author**

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Ceftriaxone has been used to treat gonorrhea in China and most other countries for >1 decade, but the level of decreased susceptibility or clinical resistance to ceftriaxone has increased (1). Moreover, the international spread of ceftriaxone-resistant clones has been recognized as a threat to effective control of gonorrhea (2). We describe an imported ceftriaxone-resistant \textit{N. gonorrhoeae} strain isolated in China in 2016.

The patient was a heterosexual man in his late twenties. He reported unprotected 1-night heterosexual sex in Beijing in July 2016. Urethral discharge with dysuria occurred 3 days after the sexual activity. He was prescribed oral cephalosporin when he visited a private clinic in July. Because the urethral discharge did not resolve, he visited the sexually transmitted diseases clinic in Beijing Ditan Hospital (Beijing, China) in August.

Laboratory analysis of a urethra swab sample found gram-negative diplococci within leukocytes. Culture and nucleic acid amplification test were positive for \textit{N. gonorrhoeae}. Screening for other sexually transmitted infections by nucleic acid amplification test was negative for \textit{Chlamydia trachomatis}, \textit{Ureaplasma urealyticum}, and \textit{Mycoplasma hominis}.

The patient was treated with a 1-g intravenous dose of ceftriaxone once per day for 3 days. His symptoms improved after 3 days, and a test-of-cure by culture showed