Hematologic and serum biochemical reference intervals of the Oriental white stork (Ciconia boyciana) and the application of an automatic hematologic analyzer

Jae-Ik Han1, Hye-Jin Jang2, Ki-Jeong Na3,*

1Laboratory of Wildlife Diseases, College of Veterinary Medicine, Chonbuk National University, Iksan 54596, Korea
2Wildlife Center of Chungbuk, Chungbuk National University, Cheongju 28644, Korea
3Laboratory of Veterinary Laboratory Medicine, Veterinary Medical Center, and College of Veterinary Medicine, Chungbuk National University, Cheongju 28644, Korea

This study was conducted to establish accurate baseline values of clinical laboratory data with regard to age-related changes in the Oriental white stork (Ciconia boyciana). In addition, the availability of an automated hematological cell counter was evaluated. A total of 94 clinically normal storks, including 64 young storks (<1 year old; 30 male and 34 female) and 30 adults (>1 year old; 17 male and 13 female) were included. Hematological assays were performed using manual and automated cell counters and serum biochemistry profiles were examined using an automated analyzer. There were no significant differences in any parameters between male and female storks, while 16 parameters were significantly different between young and adult storks. Of these 16 parameters, total protein, albumin, aspartate aminotransferase, alanine aminotransferase, creatinine, triglyceride, total bilirubin, potassium, white blood cell count, packed cell volume, mean cell volume and hemoglobin levels were higher in adult storks than in young storks, while the latter showed higher glucose, uric acid and alkaline phosphatase levels, as well as a higher sodium/potassium ratio. The results presented herein will aid researchers who work for the conservation and rehabilitation of this endangered species.

Keywords: Ciconia boyciana, hematology, reference interval, serum biochemistry, stork

Introduction

The Oriental white stork (Ciconia [C.] boyciana) belongs to the family Ciconiidae and the order Ciconiiformes. This bird can be found in Japan, China, Korea, and Russia, although it is no longer a permanent resident in South Korea. Currently, this stork mainly inhabits the Heilong River and Wusuli River basins along the border between Russia and China [27]. The International Union for Conservation of Nature considers C. boyciana an endangered species. The Korea Institute of Oriental white stork Rehabilitation Research at Korea National University of Education is attempting to reintroduce this species as a breeding bird in Korea.

Blood analyses are widely used to diagnose and monitor general health and disease along with physiological processes in wild and captive birds as part of efforts to increase captive populations [6,11,23,31]. In addition, wild birds tend to hide clinical signs of disease; therefore, observations by the keeper should be complemented by periodic blood analyses if sick or injured birds are to be diagnosed and treated in the early stages of disease [14,17]. While interpreting data pertaining to blood analyses for animals, age must be considered because of significant differences between young and adult individuals [2,9,18]. Previous studies have reported hematological and biochemical parameters for some species of storks [19,22,26]; however, to the best of our knowledge, baseline and age-related changes in these parameters for young and adult C. boyciana have not been published to date.

Therefore, this study was conducted to establish accurate baseline values of clinical laboratory data for C. boyciana with regard to age- and sex-related changes. In addition, the availability of an automated hematological cell counter was evaluated for rapid and accurate examination of the C. boyciana population [3,7,24]. These data will help researchers who work...
for the conservation and rehabilitation of this endangered species.

Materials and Methods

Sample collection

A total of 94 clinically normal storks, including 64 young (< 1 year old; 30 male and 34 female) and 30 adult (> 1-year old; 17 male and 13 female) birds, were evaluated in this study. All storks showed visually normal behavior and appetite, as determined by experienced keepers and veterinarians, and were presexed through phenotyping and DNA tests as described by Han et al. [12]. All young storks evaluated in this study were born and raised in the Korea Institute of Oriental white stork Rehabilitation Research. During the experiment, the storks were housed individually in a 7 × 7 m outdoor cage with chain-link fencing. Each cage was covered with netting, and a pond and roost were established on the ground of the cage.

Hematological and serum biochemistry profiles

Peripheral blood was collected from the caudal tibial vein of the storks using a 24-gauge needle. The collected blood was placed into heparin-treated tubes and serum separator tubes, and serum was obtained from the clotted blood in the serum separator tube after centrifugation at 1,500 × g for 5 min.

A manual hematological examination was performed to determine the packed cell volume (PCV), hemoglobin (Hb) level, red blood cell (RBC) and white blood cell (WBC) counts, and the WBC differential count [29]. The WBC and RBC counts were determined using a hemocytometer and Natt–Herick’s solution. Microhematocrit tubes coated with heparin were filled with blood and centrifuged at 1,500 × g for 5 min to determine the PCV. Hb was assayed by the cyanmethemoglobin method. The mean cell volume (MCV), mean cell Hb (MCH), and mean cell Hb concentration (MCHC) were calculated using the following formulae: MCV = (PCV/RBC) × 10; MCH = (Hb/RBC) × 10; and MCHC = (Hb/PCV%) × 100. An air-dried blood smear was stained with Diff-Quik stain (Sysmex, Kobe, Japan), and a manual 100 cell differential count was obtained.

Serum biochemistry profiles were obtained using a 7020 Automatic analyzer (Hitachi High-Technologies, Japan). The assay included total protein, albumin, aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP), gamma-glutamyl transpeptidase, blood urea nitrogen (BUN), creatinine, total cholesterol, triglyceride, glucose, total bilirubin, creatine phosphokinase (CPK), amylase, lactate dehydrogenase (LDH), uric acid, total calcium, phosphorus, magnesium, sodium, potassium, and chloride levels. The albumin/globulin (A/G) ratio and sodium/potassium (Na/K) ratio were calculated based on the measured values of each parameter.

Evaluation of the automatic hematologic analyzer

Samples from 16 randomly selected adult storks were analyzed using the Cell-Dyn 3700 (Abbott Laboratories, USA) to evaluate the potential for use of the machine as an automatic hematological analyzer. For this evaluation, the analyzer was adjusted with the veterinary package software. Because of the

Table 1. Difference in hematological parameters between young (n = 64) and adult (n = 30) Oriental white storks (Ciconia boyciana)

| Parameter (unit) | Age (yr) | Mean ± SD | p value  | Percentile (%) |
|------------------|----------|-----------|----------|----------------|
|                  |          |           |          | 2.5 | 50 | 97.5 |
| WBC (10³/μL)     | < 1      | 7.94 ± 3.59 | 0.021*   | 3.11 | 6.67 | 16.00 |
|                  | > 1      | 15.30 ± 4.84 | 0.077    | 7.56 | 14.50 | 23.80 |
| RBC (10⁶/μL)     | < 1      | 2.51 ± 0.51  | 0.001*   | 1.63 | 2.56 | 3.48 |
|                  | > 1      | 2.44 ± 0.41  |           | 1.67 | 2.4 | 3.36 |
| PCV (%)          | < 1      | 48 ± 4       | < 0.001* | 40  | 49  | 56 |
|                  | > 1      | 50 ± 8       |           | 30  | 50  | 60 |
| Hb (g/dL)        | < 1      | 13.5 ± 1.3   | < 0.001* | 10.7 | 13.4 | 16.6 |
|                  | > 1      | 14.1 ± 2.3   |           | 7.7 | 14.4 | 17.6 |
| MCV (fL)         | < 1      | 201.6 ± 51.3 | < 0.001* | 123.6 | 191.5 | 306.7 |
|                  | > 1      | 207.0 ± 23.9 |           | 160.0 | 203.5 | 250.0 |
| MCH (pg)         | < 1      | 55.7 ± 11.5  | 0.128    | 38.2 | 52.5 | 79.8 |
|                  | > 1      | 58.4 ± 9.1   |           | 46.1 | 56.3 | 80.0 |
| MCHC (g/dL)      | < 1      | 28.1 ± 2.8   | 0.058    | 22.9 | 28.2 | 33.6 |
|                  | > 1      | 28.1 ± 1.8   |           | 24.5 | 28.3 | 32.0 |

*Significantly different between young (< 1-year old) and adult (> 1-year old) groups. WBC, white blood cell; RBC, red blood cell; PCV, packed cell volume; Hb, hemoglobin; MCV, mean cell volume; MCH, mean cell Hb; MCHC, mean cell Hb concentration.
Table 2. Difference in parameters of serum biochemistry profiles between young (n = 64) and adult (n = 30) Oriental white storks (Ciconia boyciana)

| Parameter (unit)            | Age (yr) | Mean ± SD   | p value     | Percentile (%) |
|-----------------------------|----------|-------------|-------------|----------------|
|                             |          |             |             | 2.5 | 50 | 97.5 |
| Total protein (g/dL)        | < 1      | 3.6 ± 0.6   | 0.002*      | 2.8 | 3.5 | 5.0  |
|                             | > 1      | 4.5 ± 0.9   |             | 2.9 | 4.4 | 6.5  |
| Albumin (g/dL)              | < 1      | 1.2 ± 0.2   | 0.003*      | 0.9 | 1.1 | 1.7  |
|                             | > 1      | 1.5 ± 0.5   |             | 0.9 | 1.5 | 3.5  |
| Globulin (g/dL)             | < 1      | 2.5 ± 0.5   | 0.060       | 1.7 | 2.4 | 3.4  |
|                             | > 1      | 3.0 ± 0.6   |             | 2.0 | 2.9 | 4.7  |
| A/G ratio                   | < 1      | 0.5 ± 0.1   | 0.210       | 0.4 | 0.5 | 0.7  |
|                             | > 1      | 0.5 ± 0.2   |             | 0.4 | 0.5 | 1.5  |
| AST (IU/L)                  | < 1      | 354 ± 111   | 0.011*      | 234 | 326 | 610  |
|                             | > 1      | 415 ± 155   |             | 185 | 404 | 759  |
| ALT (IU/L)                  | < 1      | 47 ± 16     | 0.003*      | 22  | 47  | 86   |
|                             | > 1      | 58 ± 29     |             | 28  | 52  | 139  |
| BUN (mg/dL)                 | < 1      | 5 ± 2       | 0.916       | 2   | 5   | 8    |
|                             | > 1      | 5 ± 2       |             | 2   | 5   | 9    |
| Creatinine (mg/dL)          | < 1      | 0.4 ± 0.1   | < 0.001*    | 0.3 | 0.4 | 0.6  |
|                             | > 1      | 0.4 ± 0.2   |             | 0.2 | 0.4 | 0.7  |
| Total cholesterol (mg/dL)   | < 1      | 211 ± 54    | 0.456       | 134 | 201 | 357  |
|                             | > 1      | 224 ± 60    |             | 136 | 222 | 381  |
| Glucose (mg/dL)             | < 1      | 248 ± 31    | 0.030*      | 191 | 250 | 306  |
|                             | > 1      | 243 ± 50    |             | 110 | 240 | 345  |
| Triglyceride (mg/dL)        | < 1      | 52 ± 18     | < 0.001*    | 28  | 47  | 111  |
|                             | > 1      | 74 ± 29     |             | 30  | 68  | 121  |
| Total bilirubin (mg/dL)     | < 1      | 0.5 ± 0.2   | 0.010*      | 0.2 | 0.5 | 0.9  |
|                             | > 1      | 0.6 ± 0.2   |             | 0.2 | 0.7 | 1.0  |
| CPK (IU/L)                  | < 1      | 560 ± 336   | 0.169       | 150 | 478 | 1387 |
|                             | > 1      | 392 ± 259   |             | 108 | 329 | 1137 |
| Amylase (IU/L)              | < 1      | 746 ± 190   | 0.575       | 431 | 735 | 1234 |
|                             | > 1      | 903 ± 199   |             | 544 | 871 | 1328 |
| LDH (IU/L)                  | < 1      | 902 ± 487   | 0.363       | 310 | 777 | 2338 |
|                             | > 1      | 1142 ± 354  |             | 552 | 1108 | 1957 |
| Uric acid (mg/dL)           | < 1      | 8.1 ± 3.8   | 0.028*      | 2.1 | 8.3 | 16.7 |
|                             | > 1      | 6.1 ± 2.6   |             | 0.7 | 5.4 | 12.1 |
| ALP (IU/L)                  | < 1      | 824 ± 360   | 0.003*      | 96  | 812 | 1468 |
|                             | > 1      | 451 ± 196   |             | 137 | 457 | 941  |
| GGT (IU/L)                  | < 1      | 0.5 ± 1.3   | 0.823       | 0.0 | 0.0 | 5.0  |
|                             | > 1      | 0.6 ± 1.2   |             | 0.0 | 0.0 | 4.0  |
| Sodium (mmol/L)             | < 1      | 155 ± 5     | 0.583       | 147 | 155 | 162  |
|                             | > 1      | 152 ± 5     |             | 137 | 153 | 161  |
| Potassium (mmol/L)          | < 1      | 3.4 ± 0.8   | 0.001*      | 2.3 | 3.1 | 5.8  |
|                             | > 1      | 4.0 ± 1.4   |             | 2.1 | 3.8 | 7.0  |
| Chloride (mmol/L)           | < 1      | 118 ± 4     | 0.125       | 111 | 118 | 130  |
|                             | > 1      | 116 ± 6     |             | 103 | 117 | 133  |
| Calcium (mg/dL)             | < 1      | 10.4 ± 1.4  | 0.597       | 6.9 | 10.5 | 12.8 |
|                             | > 1      | 9.1 ± 1.6   |             | 6.8 | 9.1 | 15.7 |
| Phosphorus (mg/dL)          | < 1      | 4.3 ± 1.0   | 0.765       | 2.3 | 4.4 | 6.3  |
|                             | > 1      | 3.6 ± 1.2   |             | 1.0 | 3.6 | 6.8  |
Table 2. Continued

| Parameter (unit)          | Age (yr) | Mean ± SD | p value | Percentile (%) |   |
|---------------------------|----------|-----------|---------|----------------|---|
| Magnesium (mg/dL)         | < 1      | 2.4 ± 0.5 | 0.664   | 1.4            | 2.6 | 3.1 |
|                           | > 1      | 1.8 ± 0.7 |         | 0.6            | 1.6 | 3.3 |
| Na/K ratio                | < 1      | 48.0 ± 10.0 | 0.002*  | 27.0           | 49.0 | 66.5 |
|                           | > 1      | 41.1 ± 15.7 |         | 15.4           | 39.3 | 73.3 |

*Significantly different between young (< 1-year old) and adult (> 1-year old) groups. A/G, albumin/globulin; AST, aspartate aminotransferase; ALT, alanine aminotransferase; BUN, blood urea nitrogen; CPK, creatine phosphokinase; LDH, lactate dehydrogenase; ALP, alkaline phosphatase; GGT, gamma-glutamyl transpeptidase.

Table 3. Passing-Bablok agreement between manual method and Cell-Dyn 3700 for hematological parameters in the Oriental white stork (Ciconia boyciana)

| Hematological parameter | Manual method versus automatic analyzer |
|-------------------------|----------------------------------------|
|                         | Intercept estimate (95% CI)             | Slope estimate (95% CI) |
| WBC                     | -6.71 (-42.89, 2.13)                    | 1.33 (0.67, 3.80) |
| RBC                     | -0.08 (-1.94, 1.33)                     | 1.03 (0.48, 1.73) |
| Hb                      | 0.16 (-4.23, 4.13)                      | 0.78 (0.44, 1.02) |
| PCV                     | 10.47 (-1.06, 27.61)                    | 0.78 (0.44, 1.02) |

Statistical analysis

Differences in variables between the young and adult groups or the manual and automated methods were analyzed using an independent t-test. Differences in variables between the male and female storks in the young and adult groups were analyzed using one-way ANOVA. The Kolmogorov–Smirnov test (p < 0.05) was used to determine if the data had a Gaussian distribution [20]. When data were from a Gaussian distribution, reference intervals were defined by minimum and maximum values for groups of fewer than 40 samples and by central 95% percentiles (mean ± 2SD) for groups of more than 40 samples [28]. For results that did not significantly differ between groups, reference intervals were determined from values pooled from two groups of storks [5,20,28]. All statistical analyses were conducted using SPSS statistical software for Windows (ver. 18.0.0.0; SPSS, USA). A p value of < 0.05 was considered statistically significant.

Results

Hematology

No significant differences were observed in hematological parameters between male and female storks in both the young and adult groups (i.e., young male versus adult male and young female versus adult female, respectively). Upon evaluation of age-related differences, the WBC count, PCV, Hb level, and MCV were significantly higher in adult storks than in young storks (p < 0.001 for both parameters), while none of the hematological parameters were higher in young storks (Table 1). Hematological reference intervals for storks are given in Table 1.

Serum biochemistry profiles

Serum biochemistry profiles did not differ significantly between male and female storks in the young or adult groups (i.e., young male versus adult male and young female versus adult female, respectively). Evaluation of age-related differences revealed that total protein, albumin, AST, ALT, creatinine, triglyceride, total bilirubin, and potassium levels were significantly higher in adult storks, while young storks showed significantly higher glucose, uric acid, and ALP levels, and a higher Na/K ratio (Table 2). Reference intervals for the serum biochemistry profiles of this stork are given in Table 2.

Automatic hematologic analyzer

There was good agreement between the results obtained using the manual method and those obtained using the analyzer for all hematological parameters (Table 3; Fig. 1). These
findings were consistent with those of Bland–Altman plots, although the biases were small and the 95% confidence intervals were wide (Fig. 2).

Discussion

Knowledge of the physiological state and causes of illness and death can improve and facilitate the selection and application of proper management strategies for the conservation of endangered species [10,16,21]. Clinical hematology and serum biochemistry profiles are useful diagnostic tools in clinical practice that are particularly important for birds because they generally hide the clinical signs of disease [13]. The primary purpose of this study was to establish reference intervals for hematological and serum biochemical parameters in healthy young and adult captive storks. The results presented herein will be valuable to conservation and rehabilitation projects for the stork.

In the field of veterinary medicine, automatic hematological instruments are widely used in most specialized laboratories and hospitals to simplify hematological analysis [24,25,32]. Unlike instruments used for serum biochemical parameters [1], the application of hematological instruments to bird species is somewhat complicated because of the presence of nucleated RBCs and thrombocytes [4]. Naked nuclei from lysed RBCs and nucleated thrombocytes are similar in size to lymphocytes; therefore, there is a false-positive increase in the leukocyte number in automatic cell counters when the WIC method is used alone. In contrast, the Cell-Dyn 3700 we used in this study uses both the WIC and WOC methods, providing two sets of results with an error message if both methods represent significantly different results [30]. Therefore, we evaluated this machine for hematological analysis of the Oriental white stork. Overall, the results demonstrated acceptable precision for every evaluated hematological parameter.

Among hematological parameters, WBC, PCV, and Hb levels, as well as MCV were higher in adults storks than in young storks. The results of RBC-related parameters may be due to the immature and developmental physiological states of young storks, which are similar to other bird species [8,15,17]. Additionally, the results of the present study were similar to those of studies of other species of stork chicks (Ciconia...
Fig. 2. Bland-Altman plots showing the agreement between the hematological data obtained on manual count and Cell-Dyn 3700 for hematological parameters in the Oriental white stork (Ciconia boyciana). X-axes represent the average for both methods, and Y-axes represent the difference between assessments.

ciconia) [22,26]. However, the higher WBC count of the adult storks in this study was different from the results of other species of stork chicks. Rather, the age-related change in WBC count was similar to that of a study of the bearded vulture (Gypaetus barbatus), which showed higher WBC counts in adult groups [15].

Among serum biochemical parameters, eight (total protein, albumin, AST, ALT, creatinine, triglyceride, total bilirubin, and potassium) were higher in the adult storks than in young ones, which showed higher glucose, uric acid and ALP levels, as well as higher Na/K ratios. The difference in total protein and albumin levels may have been due to the immature and developmental physiologic states of young storks, which are similar to other bird species [8,15,17]. The higher uric acid, ALP, and glucose levels found in young storks may have been caused by increased protein synthesis, increased bone metabolism, or higher energy requirements [17], whereas increased activities of other parameters (AST, ALT, creatinine, triglyceride, total bilirubin, and potassium) may have resulted from increased basal metabolism and tissue turnover [17].

Notably, none of these parameters showed sex-related differences in either the young or adult groups. These findings are inconsistent with those for the black stork C. nigra, which showed greater levels of Hb, total protein, ALP, and triglyceride in females, while males showed higher levels of albumin [19], probably reflecting interspecies variability.

In conclusion, this study determined hematological and serum biochemistry profiles for evaluating the health status of the Oriental white stork C. boyciana. We also demonstrated that the Cell-Dyn 3700, an automated cell counter, can provide accurate hematological data in a short period of time.

Acknowledgments

This study was supported by research funds for newly appointed professors of Chonbuk National University in 2015.

Conflict of Interest

The authors declare that there is no conflict of interest.
References

1. Ammersbach M, Beaufrè H, Gionet Rollick A, Tully T. Laboratory blood analysis in Strigiformes—Part II: plasma biochemistry reference intervals and agreement between the Abaxis Vetscan V2 and the Roche Cobas c501. Vet Clin Pathol 2015, 44, 128-140.

2. Bailey TA, Wernery U, Howlett J, Naldo J, Samour JH. Age-related plasma chemistry changes in houbara and kori bustards in the United Arab Emirates. J Wildl Dis 1999, 35, 31-37.

3. Bauer NB, Nakagawa J, Dunker C, Failing K, Moritz A. Evaluation of the impedance analyzer PocH-100iV Diff for analysis of canine and feline blood. Vet Clin Pathol 2012, 41, 194-206.

4. Beaufrè H, Ammersbach M, Tully TN Jr. Complete blood cell count in psitaciformes by using high-throughput image cytometry: a pilot study. J Avian Med Surg 2013, 27, 211-217.

5. Chung C, Cheng C, Chin S, Lee A, Chi C. Morphologic and cytochemical characteristics of Asian yellow pond turtle (Ocadia sinensis) blood cells and their hematologic and plasma biochemical reference values. J Zool Wildl Med 2009, 40, 76-85.

6. Cooper JE. Minimally invasive health monitoring of wildlife. Anim Welt 1998, 7, 35-44.

7. Criswell KA, Bock JL, Wildeboer SE, Johnson K, Giovanelly RP. Validation of Sysmex XT-2000iV generated quantitative bone marrow differential counts in untreated Wistar rats. Vet Clin Pathol 2014, 43, 125-136.

8. Dujowich M, Mazet JK, Zuba JR. Hematologic and biochemical reference ranges for captive California condors (Gymnogyps californianus). J Zoo Wildl Med 2005, 36, 590-597.

9. Fudge AM. Clinical haematology and chemistry of ratites. In: Tully TN Jr, Shane SM (eds.). Ratite Management, Medicine and Surgery, 1st ed. pp. 214-245. Krieger Publishing, Malabar, 1996.

10. González LM, Margalida A, Mañosa S, Sánchez R, Oriol J, Molina JL, Caldera J, Aranda A, Prada L. Causes and spatio-temporal variations of non-natural mortality in the vulnerable Spanish imperial eagle Aquila adalberti during a recovery period. Oryx 2007, 41, 495-502.

11. Han JL, Cang HJ, Lee SJ, Kang HM, Kim S, Park SR, Na KJ. Bacterial flora of the intestine in normal captive Oriental white storks. J Vet Clin 2011, 28, 516-518.

12. Han JL, Kim JH, Kim S, Park SR, Na KJ. A simple and improved DNA test for avian sex determination. Auk 2009, 126, 779-783.

13. Harr KE. Clinical chemistry of companion avian species: a review. Vet Clin Pathol 2002, 31, 140-151.

14. Hawkey CM, Dennet TB, Peirce MA (eds.). Color Atlas of Comparative Veterinary Hematology. p 192, Wolfe Medical, London, 1989.

15. Hernández M, Margalida A. Hematology and blood chemistry reference values and age-related changes in wild bearded vultures (Gypaetus barbatus). J Wildl Dis 2010, 46, 390-400.

16. Hernández M, Margalida A. Pesticide abuse in Europe: effects on the cinereous vulture (Aegypius monachus) population in Spain. Ecotoxicology 2008, 17, 264-272.

17. Hochleithner M. Biochemistry. In: Ritchie BW, Harrison GI, Harrison LR (eds.). Avian Medicine: Principles, and Application. pp. 223-245, Lake Worth, 1994.

18. Howlet JC, Bailey TA, Naldo JL. Age-related hematologic changes in captive reared kori bustards (Ardeotis kori). Comp Haematol Int 1998, 8, 26-30.

19. Lanzarot MP, Barahona MV, San Andrés MI, Fernández-García M, Rodríguez C. Hematologic, protein electrophoresis, biochemistry, and cholinesterase values of free-living black stork nestlings (Ciconia nigra). J Wildl Dis 2005, 41, 379-386.

20. Lumsden JH, Mullen K. On establishing reference values. Can J Comp Med 1978, 42, 293-301.

21. Margalida A. Bearded vultures (Gypaetus barbatus) prefer fatty bones. Behav Ecol Sociobiol 2008, 63, 187-193.

22. Montesinos A, Sainz A, Pablos MV, Mazzucchelli F, Tesouro MA. Hematological and plasma biochemical reference intervals in young white storks. J Wildl Dis 1997, 33, 405-412.

23. Naidoo V, Diekmann M, Wolters K, Swan GE. Establishment of selected baseline blood chemistry and hematologic parameters in captive and wild-caught African white-backed vultures (Gyps africanus). J Wildl Dis 2008, 44, 649-654.

24. Perez-Eci a A, Gonzalez-De Cara CA, Aguilera-Aguilera R, Estepa JC, Rubio MD, Mendoza FJ. Comparison of donkey hemogram using the LaserCyte hematology analyzer, an impedance system, and a manual method. Vet Clin Pathol 2014, 43, 525-537.

25. Piviani M, Segura D, Monreal L, Bach-Raich E, Mesalles M, Pastor J. Neutrophilic myeloperoxidase index and mean light absorbance in neonatal septic and nonseptic foals. Vet Clin Pathol 2011, 40, 340-344.

26. Puerta ML, Munoz Pulido R, Hucenas V, Abeleenda M. Hematology and blood chemistry of chicks of white and black storks (Ciconia ciconia and Ciconia nigra). Comp Biochem Physiol A Comp Physiol 1989, 94, 201-204.

27. Smirenski SM. Oriental white stork action plan in the USSR. In: Coulter MC, Wang Q, Luthin CS (eds.). Biology and Conservation of the Oriental White Stork (Ciconia boyciana). pp. 165-177. Savannah River Ecology Laboratory, Aiken, 1991.

28. Solberg HE. Establishment and use of reference values. In: Burtis CA, Ashwood ER, Tietz NW (eds.). Tietz Textbook of Clinical Chemistry. 3rd ed. pp. 336-356, WB Saunders, Philadelphia, 1999.

29. Stockholm SL, Scott MA (eds.). Fundamentals of Veterinary Clinical Pathology. 2nd ed. pp. 62-124, Wiley-Blackwell, Ames, 2008.

30. Sulj ević F, Fazlić M, Corić J, Kiseljaković JC. Evaluation of a hematology analyzer Cell-Dyn 3700 SL. Bosn J Basic Med Sci 2003, 3, 35-41.

31. Tamukai K, Takami Y, Akabane Y, Kanazawa Y, Une Y. Plasma biochemical reference values in clinically healthy captive bearded dragons (Pogona vitticeps) and the effects of sex and season. Vet Clin Path 2011, 40, 368-373.

32. Weiss DJ, Moritz A. Equine immune-mediated hemolytic anemia associated with Clostridium perfringens infection. Vet Clin Pathol 2003, 32, 22-26.

www.vetsci.org