Use of Probiotics to Control Aflatoxin Production in Peanut Grains

Juliana Fonseca Moreira da Silva,1,2 Joenes Mucci Peluzio,2 Guilherme Prado,3 Jovita Eugênia Gazzinelli Cruz Madeira,3 Marize Oliveira Silva,3 Paula Benevides de Morais,2 Carlos Augusto Rosa,1 Raphael Sanzio Pimenta,2 and Jacques Robert Nicoli1

1Departamento de Microbiologia, Instituto de Ciências Biológicas, Universidade Federal de Minas Gerais, Bloco J, 4ºAndar, Sala 171, Avenida Antônio Carlos 6627, CP 486, Pampulha, 31270-901 Belo Horizonte, MG, Brazil
2Laboratório de Microbiologia Ambiental e Biotecnologia, Universidade Federal do Tocantins, ALC No. 14 Campus Universitário, Avenida NS 15 Bloco II Sala 05, 77.001-090 Palmas, TO, Brazil
3Fundação Ezequiel Dias, Laboratório de Micologia e Micotoxinas, Rua Conde Pereira Carneiro 80, 30510010 Belo Horizonte, MG, Brazil

Correspondence should be addressed to Raphael Sanzio Pimenta; biorapha@yahoo.com.br

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1. Introduction

Peanut (Arachis hypogaea) is one of the major food crops cultivated throughout the tropics and subtropical regions and its annual world production is of approximately thirty-eight million tons [1]. However, this grain is very susceptible to mycotoxin contamination, especially by aflatoxins. Aflatoxins are secondary metabolites produced by three filamentous fungal species: Aspergillus flavus, A. parasiticus, and A. nomius [2, 3]. The aflatoxins can be classified as B1, B2, G1, and G2 according to their fluorescence under ultraviolet light and molecular weight and strains of A. parasiticus can produce all of them [4]. The B1 aflatoxin is recognized by the International Agency for Research on Cancer as a group 1 carcinogenic substance, whereas aflatoxins B2, G1, and G2 are classified as possible carcinogenic substances [5].

Aflatoxin contamination in peanuts is a great economical and public health concern [6] and many studies search for efficient methodologies capable of controlling filamentous fungal growth in pre− and postharvest. Use of GRAS...
(Generally Regarded as Safe) substances and antagonistic microorganisms has been tested with some success [7-9]. Antagonism is one of the most important phenomena observed in microecological relationships, and the principal mechanism responsible for the beneficial effect of probiotic yeasts and bacteria. Probiotics are defined by the Food and Agriculture Organization/World Health Organization [10] as “live microorganisms which when administered in adequate amounts confer a health benefit on the host.” An interesting possibility of using the antagonistic ability of probiotics would be to impair growth and/or aflatoxin production by phytopathogenic fungi during storage of grains.

Shetty and Jespersen [11] reported that *Saccharomyces cerevisiae* and lactic acid bacteria (LAB) can reduce the toxic effects of mycotoxins in foods by absorption on their cell walls. The LAB are known as predominant participants in many industrial food processes, especially in vegetables, meats, and dairy fermentations, where they can prevent contamination by harmful microorganisms. *Saccharomyces cerevisiae* is the most important microorganism responsible for production of alcoholic beverages and biofuel. Both LAB and *Saccharomyces* (particularly *S. cerevisiae* var. *boulardii*) have already been described for their use as probiotics [12-14].

*S. cerevisiae* (UFMG 905) reduced the translocation levels of pathogens, promoted the host immunomodulation, decreased the mortality and helped in the preservation of liver tissue and gut barrier integrity, and reached population decreased the mortality and helped in the preservation of liver tissue and gut barrier integrity, and reached population decreased the mortality and helped in the preservation of liver tissue and gut barrier integrity, and reached population decreased the mortality and helped in the preservation of liver tissue and gut barrier integrity, and reached population decreased the mortality and helped in the preservation of liver tissue and gut barrier integrity, and reached population decreased the mortality and helped in the preservation of liver tissue and gut barrier integrity, and reached population.

The objective of this work was to evaluate the capabilities of three probiotic microorganisms (two strains of *S. cerevisiae* and one of *L. delbrueckii*) to reduce the sporulation and aflatoxin production by *A. parasiticus* in peanuts.

2. Materials and Methods

2.1. Peanut Grains. Autoclaved peanuts grains, cultivar IAC Caiapó, available from Instituto Agronômico de Campinas were used. The grains had medium size, brownish color, high oil content, and no aflatoxin presence and were produced in the 2009/2010 crop.

2.2. Microorganisms. *Aspergillus parasiticus* IMI 242695 isolated from contaminated food products and aflatoxin producer (B₁, B₂, G₁, and G₂) was obtained from the International Mycological Institute (UK). *Saccharomyces boulardii* 17 was obtained from MERCK/SA (Rio de Janeiro, RJ, Brazil), *S. cerevisiae* UFMG 905 pertained to the Collection of Microorganisms and Cells of Federal University of Minas Gerais (UFMG), and *Lactobacillus delbrueckii* H2b20 was isolated from a healthy newborn and pertained to the culture collection of the Laboratório de Ecologia e Fisiologia de Microorganismos (Departamento de Microbiologia, Universidade Federal de Minas Gerais, Belo Horizonte, MG, Brazil).

2.3. Sample Preparation: Growth, Resuspension, and Dilution

2.3.1. *Aspergillus parasiticus* IMI 242695. Spores were obtained as described by Prado et al. [19]. Spore concentration was determined using a hemocytometer and adjusted to 1 × 10⁶ spores/mL in 50 mL of 0.1% Tween 80.

2.3.2. Yeasts. *Saccharomyces boulardii* and *S. cerevisiae* UFMG 905 cells were obtained by culture on YM agar medium (2% glucose, 0.5% peptone, 0.3% malt extract, 0.3% yeast extract, and 2% agar) incubated for 24 h at 37°C. Colonies were collected and suspended in 0.1% Tween 80 and the cell concentration was determined using a hemocytometer and the suspension was adjusted to 1 × 10⁸ cells/mL in a final volume of 50 mL.

2.3.3. *Lactobacillus delbrueckii*. Bacterial cells were obtained by culture on de Man, Rogosa and Sharpe (MRS) agar medium incubated for 24 h at 37°C in aerobic static conditions. Colonies were collected and suspended in 0.1% Tween 80 and the cell concentration was determined using a hemocytometer and the suspension was adjusted to 1 × 10⁸ cells/mL in a final volume of 50 mL.

2.4. Biological Control Assays. Autoclaved peanuts grains were used in the following eleven experimental groups (15 g in each): (1) first positive control: grains inoculated with 2.5 mL of *A. parasiticus* spore suspension and 2.5 mL of 0.1% Tween 80; (2, 3 and 4) negative controls: grains inoculated with 2.5 mL of each yeast or bacterial suspension plus 2.5 mL of 0.1% Tween 80; (5, 6 and 7) simple antagonistic tests: grains inoculated with 2.5 mL of each yeast or bacterial suspension plus 2.5 mL of 0.1% Tween 80; (8) second positive control: grains inoculated with 2.5 mL of *A. parasiticus* plus 5 mL of 0.1% Tween 80; (9, 10, and 11) combined antagonistic tests: grains inoculated with 2.5 mL of combined yeast or bacterial suspension in a factorial association for inoculation with two microorganisms (*L. delbrueckii* + *S. cerevisiae*; *S. boulardii* + *S. cerevisiae*; and *S. boulardii* + *L. delbrueckii*) plus 2.5 mL of *A. parasiticus* spore suspension. All the inoculations were performed by sprinkling of grains with microbial suspensions followed by incubation for seven days at 30°C.

2.5. Aflatoxin Quantification. The aflatoxin quantification was performed by fluorodensitometry at 365 nm using a fluorodensitometer (model CS9301, Shimadzu Corp., Kyoto, Japan). Concentration of aflatoxin B₁ was calculated by comparing the fluorescent intensity of sample spots with known standard amounts spotted on the same plate in the range from 0.8 to 0.88 ng [20].

2.6. Microbial Cell Viability on Peanut Grains. To determine the evolution of microbial cell viability on peanuts during storage, 15 g of peanut grains was placed in an plastic film covered Erlenmeyer flask (250 mL) and soaked with 2.5 mL of each yeast or bacterial suspension and the mixture was incubated at 25°C. At regular time intervals (weekly until 21
days and monthly until 330 days), viable cell counts were performed as follows. Three inoculated grains were introduced in three assay tubes containing 10 mL of sterile saline. The tubes were agitated for approximately 30 s, and an aliquot of the supernatant was submitted to serial decimal dilution up to 10^{-5} in saline sterile solution. Then, 50 µL of each dilution was plated onto MRS or YM agar media for Lactobacillus and yeasts counts, respectively, and incubated at 30 °C for 48 h. After incubation, viable cell number was determined and the results expressed as decimal logarithm of colony forming units per gram (log_{10} CFU/grain). Culture inoculation and counting were automatically performed by spiral plate and colonies counter (IUL Instruments, Barcelona, Spain).

2.7. In Vitro Antagonism Assay. To evaluate a possible in vitro antagonism of yeast and Lactobacillus probiotics against A. parasiticus, the agar double layer diffusion test was described by Tagg et al. [21] with some modifications. The antagonistic effect was evaluated using live and inactivated cells of each probiotic. In the assay using live cells (Assay 1) Petri dishes containing adequate medium (agar MRS or YM for bacteria or yeasts, resp.) were inoculated with 10 µL of yeast or bacterial suspension in the center and incubated at 37°C during 24 h. After incubation, 3.5 mL ofMEA medium (1.2% malt extract, 2% agar) supplemented with 10 µL of A. parasiticus spore suspension was spread onto the agar surface and the dishes were incubated at 25°C for seven days. In the assay using inactivated cells (Assay 2), after inoculation on the center of the Petri dish, the yeast or bacterial spot was killed by chloroform vapor exposition for 20 min. Then, Petri dishes were held open in a laminar flow hood until complete removal of residual chloroform and the dishes were treated as in Assay 1. After incubation, presence of growth inhibition zone and alterations in appearance of the phytopathogen were observed.

2.8. Determination of Number and Morphology of A. parasiticus Spores. This determination was performed with the same Petri dishes used for the in vitro antagonism assay described above. The spore count was done as follows: 10 mL of 0.1% Tween 80 and ten glass beads were added to each plate, and after shaking, the supernatant was transferred to a conical tube (50 mL). After homogenization, spore concentration in the supernatant was determined using a hemocytometer. The results were expressed as spores mL\(^{-1}\). To evaluate morphological alterations, 20 µL from each suspension was stained with cotton blue and visualized using an optical microscopy (Leica DM750), and the images were captured and analyzed with the Leica microsystems DFC 425 software.

2.9. Statistical Analysis. All the experiments followed a randomized design with five replicates each. Data were submitted to Kolmogorov-Smirnov test of normality and then compared by variance analysis and Tukey test at 5% of significance. Statistical analysis was performed using the Sisvar 5.3 software (UFLA, Lavras, MG, Brazil).

3. Results and Discussion

Saccharomyces boulardii, S. cerevisiae UFMG 905, and L. delbrueckii were able to reduce significantly A. parasiticus sporulation as shown in Table I. These reductions were observed with the utilization of both viable and inactivated yeasts and bacteria, but a higher reduction was obtained with live cells. Additionally, in these in vitro experiments with the use of viable and inactivated microorganisms, a color alteration in A. parasiticus appearance was observed when compared to the control group (olive green to yellow) (Figures 1(a)–1(g)). This alteration, due to the reduction in spore production, represents a considerable vantage for the use of the tested probiotics since this effect can reduce the pathogen dispersion in stored grains. On the other hand, there was no inhibition of the mycelium growth as well as no structural modifications of A. parasiticus spores which remained globose, rough (with spines), and hyaline.

In relation to the ability to reduce spore production, there was no statistical significant difference among the three viable microorganisms, despite the tendency to a better result for the Lactobacillus. Similar results were observed by Onilude et al. [22] using six lactic acid bacteria (LAB) (L. fermentum OYB, L. fermentum RS2, L. plantarum MW, L. plantarum YO, L. brevis WS3, and Lactococcus spp. RS3) to control the growth of two strains of A. parasiticus and four of A. flavus (all aflatoxigenic). L. plantarum YO strain significantly inhibited the vegetative growth and sporulation of all phytopathogens tested. These findings demonstrate the antifungal activity of some Lactobacillus, and the inhibition was possibly related to a pH reduction and/or a nutrient competition of the culture medium. Bueno et al. [23] evaluated the capacity of two Lactobacillus species (L. casei CRL 431 and L. rhamnosus CRL, 1224) to reduce the growth of three strains of A. flavus, and they observed a decreased growth of the phytopathogens when cultivated in association with the Lactobacillus strains (reduction of mycelium dry weight of 73% and 85% with CRL).
Figure 1: Positive control: *A. parasiticus* IMI 242695 (10 μL at 10^6 spores/mL) incubated for seven days at 30°C in semisolid malt agar (a). Group 1, live antagonistic cells (10 μL 10^8 cells/mL) incubated for seven days at 30°C. *L. delbrueckii* × overlapping culture (10 μL 10^6 spores/mL of *A. parasiticus* IMI 242695) (b); *S. boulardii* × overlapping culture (c); *S. cerevisiae* strain UFMG 905 × overlapping culture (d). Group 2, antagonistic cells inactivated by chloroform vapors (10 μL 10^6 cells/mL) incubated for seven days at 30°C. *L. delbrueckii* × overlapping culture (10 μL 10^6 spores/mL of *A. parasiticus* IMI 242695) (e); *S. boulardii* × overlapping culture (f); *S. cerevisiae* strain UFMG 905 × overlapping culture (g).

43 and CRL 1224 strain, resp.). Similar results were observed by Muñoz et al. [24], when three LAB and one *S. cerevisiae* strain were tested for antagonistic effects against *A. nomius* under different incubation conditions. After three days of coculture, 75%, 40%, 36%, and 20% of growth inhibition of *A. nomius* were observed for *L. rhamnosus* O236, *L. fermentum* ssp. *cellobiosus* 408, *L. fermentum* 27A, and *S. cerevisiae*, respectively. However, these antagonistic effects against
**Figure 2**: Viable cell count (CFU/Grain) of *S. boulardii* (a), *S. cerevisiae* UFMG 905 (b), and *L. delbrueckii* UFV H2b20 (c) during storage at 30°C for 330 days.

*A. nomius* growth were not accompanied by morphological alterations of the hyphae.

As shown above, better results for biological control of the phytopathogen were obtained when the yeasts and bacteria were used in viable form when compared to the inactivated form. As in Brazil, grains are generally stored during a mean period of about six months, and it was important to evaluate the evolution of viable population levels of the yeasts and bacteria after their inoculation onto the surface of peanuts grains during a simulated storage. Figure 2 shows that high levels of viable cells were maintained at least until 300 days after sprinkling of the three microorganisms.

Aflatoxin production by *A. parasiticus* was analyzed in the presence of each yeast and bacterium alone or in pair combinations. In the first situation, only the yeasts caused a significant reduction in aflatoxin production (Table 2), with statistically similar effects for *S. cerevisiae* UFMG 905 (72.8%) and *S. boulardii* (65.8%)

When inoculated in pairs, all combinations significantly reduced aflatoxin production by *A. parasiticus* (Table 3). Interestingly, the best result was obtained with the coinoculation of *S. boulardii* and *L. delbrueckii* (96.1%). This synergistic effect between yeast and the *Lactobacillus* was not observed in the combinations between *S. boulardii* and *S. cerevisiae* (71.1%) or *L. delbrueckii* and *S. cerevisiae* UFMG 905 (66.7%). Similar results were obtained by Prado et al. [6] when they used two yeasts pertaining to the *Saccharomycopsis* genus (*S. schoenii* and *S. crataegensis*) as biological control agents. These yeasts were able to reduce *A. parasiticus* production of aflatoxin B<sub>1</sub> and G<sub>1</sub> in peanuts. These results suggest that biological control with selected microorganisms could reduce not only the spore dispersion but also the production of mycotoxins by phytopathogens in stored grains.

Reddy et al. [25] used three bacterial species (*Rhodococcus erythropolis*, *Bacillus subtilis* and *Pseudomonas fluorescens*) for biological control of *A. flavus* and observed that *R. erythropolis* completely inhibited phytopathogen growth and, as a consequence, its aflatoxin B<sub>1</sub> production. The other microorganisms reduced mycelium growth in a range of 65 to 74% and aflatoxin production from 39% to 65%. The authors also noted that the inhibitory activity is likely due to a chemical antagonistic extracellular substance produced by the bacteria. Prado et al. [19] tested *S. cerevisiae* YEF 186 as an *A. parasiticus* antagonist in two peanut cultivars (IAC Runner and IAC Caiapó) with two different incubation times (seven and fourteen days) and two different inoculation sequences (yeast inoculated simultaneously or three hours before

### Table 2: Mean aflatoxin (µg/kg) production by *A. parasiticus* IMI 242695 incubated for seven days at 30°C in peanut grains cultivar IAC Caiapo and perceptual reduction of aflatoxin induced by *S. boulardii*, *S. cerevisiae* UFMG 905, and *L. delbrueckii* UFV H2b20. AP = *A. parasiticus*.

| Assays                | Average<sup>1</sup> S.D. | Reduction (%) |
|-----------------------|---------------------------|---------------|
| Control (AP)          | 36,695.7 ± 14,920.8       | —             |
| *S. boulardii* × AP   | 12,538.9 ± 9,731.9        | 65.8          |
| *S. cerevisiae* × AP  | 9,981.8 ± 2,244.4         | 72.8          |
| *L. delbrueckii* × AP | 34,265.6 ± 6,184.2        | 6.6           |

<sup>1</sup> Different letters indicate significant differences according to Tukey’s test (P < 0.05); S.D. = standard deviation.
Table 3: Mean aflatoxin (µg/kg) production by *A. parasiticus* IMI 242695 incubated for seven days at 30°C in peanut grains cultivar IACCaiapó and perceptual reduction of aflatoxin induced by combinations of *S. cerevisiae* UFMG 905, *S. boulardii*, and *L. delbrueckii* UFV H2b20. AP = *A. parasiticus*.

| Assays                              | Average ± deviation | Reduction (%) |
|-------------------------------------|---------------------|---------------|
| Control (AP)                        | 15,187.2 ± 3,957.0 | —             |
| *L. delbrueckii* + *S. cerevisiae* × AP | 5,062.1 ± 3,695.0 | 66.7          |
| *S. boulardii* + *S. cerevisiae* × AP | 4,389.4 ± 334.7   | 71.1          |
| *S. boulardii* + *L. delbrueckii* × AP | 589.4 ± 516.6    | 96.1          |

1Different letters indicate significant differences according to Tukey’s test (*P* < 0.05).

Figure 3: Erlenmeyer flasks with 15 g of autoclaved peanuts cultivar IAC caiapó inoculated with (a) negative control, only 5 mL of 0.1% Tween 80; (b) *S. boulardii* (2.5 mL at 10^8 cells/mL plus 2.5 mL of 0.1% Tween 80), (c) *S. cerevisiae* UFMG 905 (2.5 mL at 10^8 cells/mL plus 2.5 mL of 0.1% Tween 80), (d) *L. delbrueckii* UFV H2b20 (2.5 mL at 10^8 cells/mL plus 2.5 mL of 0.1% Tween 80) incubated at 30°C for 330 days.

The authors found that the best reduction of aflatoxin B₁ (74.4%) was obtained after seven days and when the yeast was inoculated before the pathogen. The authors suggested that this reduction was probably due to aflatoxin adhesion to the yeast cell wall or to aflatoxin degradation by the yeast, and this is probably a way of reduction observed in this study. Gerbaldo et al. [26] evaluated the antifungal activities and aflatoxin B₁ reduction promoted by two *Lactobacillus* species (*L. rhamnosus* L60 and *L. fermentum* L23) with known probiotic activities. The *Lactobacillus* strains were tested against ten aflatoxigenic *Aspergillus* strains (nine *A. flavus* strains and one *A. parasiticus*). They found that *Lactobacillus* L60 and L23 inhibited the mycelia growth of all *Aspergillus* strains tested and promoted a reduction in aflatoxin production from 73 to 99%. They also suggested three possible mechanisms to explain the effect: (1) aflatoxin degradation by enzymes from *Lactobacillus*, (2) competition for space or nutrients, or (3) absorption of aflatoxin onto the cell walls of the *Lactobacillus*.

Concluding, results of the present study showed that candidate microorganisms for biological control able to reduce the production of spores by aflatoxigenic fungi are more effective in a viable form than in an inactivated form. Additionally, the candidates tested here showed a high capacity to remain viable in high population levels even 300 days after their inoculation on the peanuts grains, which is an important property since in Brazil, the mean storage time for grains is of approximately six months. An interesting characteristic observed was that the tested microorganisms did not produce any apparent harm to the peanuts, and on the contrary, the microbial biofilm gave the grains an appearance similar to salted peanuts (Figures 3(a)–3(d)).
Another advantage of probiotics is that their commercial use in the food industry is not restricted by legislation as is the case with other microorganisms. Finally, the most important effect observed with the sprinkling of the tested microorganisms was the considerable reduction of aflatoxin production by A. parasiticus, especially when the combination of S. boulardii and L. delbrueckii H2b20 was used. Overall, these data suggest that the food industries could use the proposed method as an alternative treatment to control the dispersion and aflatoxin production by phytopathogen, remembering that beyond protection during storage, the method could provide an additional probiotic effect in the digestive tract of consumers after ingestion of the treated grains. However, more studies are needed to clarify the possible impact that lactic acid bacteria and yeasts added to reduce aflatoxins could have on the organoleptic characteristics of the products as well as the exact mechanisms responsible for the reduction of the aflatoxin contents.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

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