Herpes Simplex Virus Type 1 Infection Disturbs the Mitochondrial Network, Leading to Type I Interferon Production through the RNA Polymerase III/RIG-I Pathway

Noémie Berry,a,b Rodolphe Suspène,a Vincent Caval,a Pierre Khalfi,a,b Guillaume Beauclair,c§ Stéphane Rigaud,d Hervé Blanc,e Marco Vignuzzi,e Simon Wain-Hobson,a Jean-Pierre Vartanian*

aMolecular Retrovirology Unit, Institut Pasteur, Paris, France
bSorbonne Université, Complexité du Vivant, Paris, France
cViral Genomics and Vaccination, Institut Pasteur, Paris, France
dHub d’Analyse d’Images, Institut Pasteur, Paris, France
eViral Populations and Pathogenesis Unit, Institut Pasteur, CNRS UMR 3569, Paris, France

ABSTRACT Viruses have evolved a plethora of mechanisms to impair host innate immune responses. Herpes simplex virus type 1 (HSV-1), a double-stranded linear DNA virus, impairs the mitochondrial network and dynamics predominantly through the UL12.5 gene. We demonstrated that HSV-1 infection induced a remodeling of mitochondrial shape, resulting in a fragmentation of the mitochondria associated with a decrease in their volume and an increase in their sphericity. This damage leads to the release of mitochondrial DNA (mtDNA) to the cytosol. By generating a stable THP-1 cell line expressing the DNase I-mCherry fusion protein and a THP-1 cell line specifically depleted of mtDNA upon ethidium bromide treatment, we showed that cytosolic mtDNA contributes to type I interferon and APOBEC3A upregulation. This was confirmed by using an HSV-1 strain (KOS37 UL98-SPA) with a deletion of the UL12.5 gene that impaired its ability to induce mtDNA stress. Furthermore, by using an inhibitor of RNA polymerase III, we demonstrated that upon HSV-1 infection, cytosolic mtDNA enhanced type I interferon induction through the RNA polymerase III/RIG-I pathway. APOBEC3A was in turn induced by interferon. Deep sequencing analyses of cytosolic mtDNA mutations revealed an APOBEC3A signature predominantly in the 5’TpCpG context. These data demonstrate that upon HSV-1 infection, the mitochondrial network is disrupted, leading to the release of mtDNA and ultimately to its catabolism through APOBEC3-induced mutations.

IMPORTANCE Herpes simplex virus 1 (HSV-1) impairs the mitochondrial network through the viral protein UL12.5. This leads to the fusion of mitochondria and simultaneous release of mitochondrial DNA (mtDNA) in a mouse model. We have shown that released mtDNA is recognized as a danger signal, capable of stimulating signaling pathways and inducing the production of proinflammatory cytokines. The expression of the human cytidine deaminase APOBEC3A is highly upregulated by interferon responses. This enzyme catalyzes the deamination of cytidine to uridine in single-stranded DNA substrates, resulting in the catabolism of edited DNA. Using human cell lines deprived of mtDNA and viral strains deficient in UL12, we demonstrated the implication of mtDNA in the production of interferon and APOBEC3A expression during viral infection. We have shown that HSV-1 induces mitochondrial network fragmentation in a human model and confirmed the implication of RNA polymerase III/RIG-I signaling in the capture of cytosolic mtDNA.

KEYWORDS HSV-1, mitochondria, innate immunity, cytidine deaminase, APOBEC3A, herpes simplex virus
Mitochondria participate in a broad range of innate immune pathways, functioning as a signaling platform and contributing to effector responses. Not surprisingly, they are the target of many different viral strategies to circumvent them (1). For example, Epstein-Barr virus encodes latent membrane protein 2A (LMP2A), leading to an increase of mitochondrial fission (2), and hepatitis C virus induces oxidative stress that damages mitochondria (3), while the hepatitis B virus X protein (HBx) induces apoptosis by causing a perinuclear clustering of mitochondria as well as the loss of the mitochondrial membrane potential (4). Recently, it was demonstrated that human influenza A virus encodes PB1-F2, which targets mitochondria, altering their morphology (5).

Herpes simplex virus type 1 (HSV-1) does not escape the rule. Indeed, HSV-1 infection is known to disorganize mitochondrial dynamics and significantly reduce ATP production within 12 h postinfection (6). This perturbation is predominantly executed by UL12 (7, 8), which encodes two distinct yet similar proteins, UL12 and UL12.5. UL12 plays a crucial role in viral genome replication and processing, while the N-terminally truncated form, UL12.5, has nuclease and strand exchange activities and localizes predominantly to the mitochondria (9), where it triggers massive catabolism of mitochondrial DNA (mtDNA) during early HSV-1 replication (8). HSV-1 infection of mice results in the release of mtDNA to the cytosol and IFN induction through the cGAS/STING/IRF3 pathway (10), albeit without elimination of induction by other pathways. Infection of mice with an HSV-1 UL12 mutant strain, deficient in mitochondrial targeting, failed to stimulate innate immunity, emphasizing the role of mtDNA in signaling (10).

The fact that cytosolic double-stranded DNA (dsDNA) is read as a danger signal is reflected in the large number of sensor molecules, such as RNA polymerase III, which transcribes DNA, leading to dsRNA intermediates that are subsequently captured by RIG-I, ultimately leading to type I interferon (IFN) production (11–13). Cytoplasmic DNases such as TRED1 and TRED2 degrade dsDNA into small single-stranded DNA (ssDNA) molecules of around 60 nucleotides (14). In placental mammals, these can be further degraded by APOBEC3 cytidine deamination, deauracilation by uracil N-glycosylase (UNG), and ssDNA cleavage by apyrimidine endonucleases (APE1 and APE2), which together result in catabolism of ssDNA. For humans, chromosome 22 harbors a cluster of 7 APOBEC3 genes (APOBEC3A, A3B, A3C, A3DE, A3F, A3G, and A3H), encoding cytidine deaminases (15). All A3 enzymes, except A3DE, catalyze the deamination of cytidine to uracil on single-stranded DNA, leading to GC-to-AT editing (16–19). A3A, A3C, and A3H possess a nucleocytoplasmic localization, whereas A3D, A3F, and A3G are found exclusively in the cytoplasm. As an exception, A3B is strictly nuclear. The remaining A3DE protein is catalytically inactive but can modulate A3F and A3G by heterodimer formation (20). Many of the corresponding genes are interferon-stimulated genes (ISGs), most notably A3A, which can be upregulated 104-fold in primary leukocytes (13, 21–24).

A3A is so active that it can generate double-stranded-DNA breaks (DSBs) in an experimental setting and even edit mRNA at low levels (25–28). A3B can also edit nuclear DNA, albeit much less efficiently (25, 29). The role of A3A and A3B in cancer is highlighted by the fact that many cancer genomes encode tens of thousands of C-to-T transitions frequently in the 5’TpC dinucleotide context, a hallmark of A3A and A3B cytidine deamination (30–32). However, recent studies using high-throughput assays suggested that the preferred nucleotide motif for A3A is 5’TpCpG (33, 34).

We demonstrated here that HSV-1 infection in human acute monocytic leukemia cells triggers mitochondrial fragmentation and shapes the mitochondrial network by significantly reducing mitochondria volume. This leads to the release of mtDNA in the cytosol, inducing type I IFN through the RNA polymerase III/RIG-I pathway. A3A is in turn induced by IFN in a paracrine and autocrine manner to edit and shut down cytosolic mtDNA.

RESULTS

HSV-1 infection upregulates type I interferon and leads to APOBEC3A expression.

To correlate the impact of viral stress on the induction of ISGs, we used the HSV-1
strain SC16 to infect THP-1 cells. This cell line was used because it is an IFN-inducible human acute monocytic leukemia cell line (13). At 18 h postinfection, infected cells were lysed, and total RNA was extracted and quantitated by TaqMan reverse transcription-PCR (RT-PCR). Different multiplicities of infection (MOI) of HSV-1 generated a strong IFN-β response in a dose-dependent manner, ~160- to 2,000-fold greater than that in noninfected (NI) cells or THP-1 cells infected with heat-inactivated (HI) virus, but failed to induce potent induction of IFN-α expression (Fig. 1A).

Since some A3 genes can be induced by type I IFN, a transcription study of A3 genes was performed and showed that HSV-1 MOI increase was correlated with an increase in A3A expression at 18 h, in a dose-dependent manner, of ~60- to 440-fold (Fig. 1B). Compared to the other A3 genes, A3A upregulation was by far the most sensitive to HSV-1 infection, being upregulated by almost 2 orders of magnitude (Fig. S1A).

To further optimize our conditions of infection, THP-1 cells were infected for 18 h by HSV-1 at MOI from 0.05 to 0.5. As can be seen in Fig. 1C, only an MOI of 0.5 of HSV-1 generated DSBs in THP-1 cells ~10-fold over background (NI cells), which was correlated with induction of apoptosis as assessed using annexin V staining (Fig. 1D). Thus, infections were performed over 18 h without exceeding an MOI of 0.5. To validate the effect of HSV-1 under more physiological conditions, primary monocytes isolated from healthy patient peripheral blood mononuclear cells (PBMCs) were infected with HSV-1 strain SC16 with a wider range of MOI, from 0.05 to 3. At 18 h postinfection, we observed a common overexpression of both IFN-α and IFN-β (Fig. S1B), leading to a dose-dependent upregulation of A3A expression (Fig. S1C).

**Cytosolic DNA is the source of IFN-β production leading to APOBEC3A upregulation.** To ensure that cytosolic DNA was the main inducer of IFN and A3A upregulation, a stable THP-1 cell line expressing DNase I-mCherry fusion protein was established by lentiviral transduction using cell sorting to select for mCherry expression. DNase I cleaves single- or double-stranded DNA, considerably reducing cytosolic...
DNA. Expression of DNase I-mCherry was checked by Western blotting, and the cellular localization was assessed by confocal microscopy (Fig. 2A and B). Early apoptosis (annexin V-positive 7-aminoactinomycin D [7-AAD]-negative cells) and late apoptosis/necrosis (annexin V7-AAD+ cells) were analyzed and demonstrated that DNase I overexpression was not toxic for the cells (Fig. 2C).

The impact of the DNase I-mCherry construct was verified by transfecting dsDNA. When normalized to wild-type THP-1 cells, transfection of 250 ng of dsDNA generated an ~2-fold decrease in A3A as well as IFN-β responses (Fig. 2D; Fig. S1D). THP-1 cells expressing DNase I were then infected with HSV-1 SC16 and compared to infected wild-type THP-1 cells. At 18 h postinfection, expression of A3A as well as IFN-β responses in DNase I-expressing THP-1 cells was ~2-fold lower than in the wild-type cell line (Fig. 2D; Fig. S1D). To prove that the DNase I expressed in THP-1 was not involved in the degradation of viral DNA, reverse transcription-quantitative PCR (RT-qPCR) was performed on the HSV-1 UL42 early gene (encoding the viral DNA polymerase processivity factor) and on two late genes, UL44 (encoding the envelope glycoprotein C [gC]) and US6 (encoding the envelope glycoprotein D [gD]). As observed in Fig. 2E and Fig. S2A, identical UL42, UL44, and US6 mRNA expression was detected in

FIG 2

DNase I overexpression reduced A3A upregulation upon HSV-1 infection. Stable DNase I-overexpressing THP-1 cell lines were generated. (A) Western blot analysis of the DNase I protein expression in THP-1 cells, transfected or not with the DNase I-mCherry plasmid. β-Actin was used as a loading control. (B) Cellular localization of the DNase I protein by immunofluorescence after transfection of constructs in HeLa cells. Nuclei were stained using DAPI (blue), and mCherry constructions are in red. (C) Flow cytometry analysis of early apoptosis (annexin V+ 7-AAD− cells) and late apoptosis/necrosis (annexin V+ 7-AAD+ cells) at 48 h posttransfection in HeLa cells. The error bars represent standard deviations (SD) from three independent experiments using a two-way analysis of variance (ANOVA). (D) APOBEC3A profiling by RT-qPCR at 24 h posttransfection of 250 ng of DNA and at 18 h after HSV-1 infection in THP-1 cells expressing DNase I or not and a negative control with a heat-inactivated (HI) virus. Data were normalized to the RPL13A housekeeping gene. Mean values and SEM were calculated for three independent experiments in duplicate (n = 6). (E) HSV-1 UL42 expression by RT-qPCR at 18 h after HSV-1 infection (MOI of 0.2). Mean values and SEM were calculated for three independent experiments in duplicate (n = 6). ns, not statistically significant; ***, P < 0.005 (unpaired two-sided Student’s t test).
the DNase I-expressing THP-1 cell line as well as in wild-type THP-1 cells, suggesting that viral replication was not influenced by DNase I.

Together, these results proved that cytosolic DNA played a crucial role in the priming of the innate response. These findings highlighted a mechanism by which cytosolic DNA induces IFN-β, which in turn induces A3A expression.

**HSV-1 infection disturbs the mitochondrial network.** Loss of mitochondrial volume homeostasis and the accompanying mitochondrial swelling are two of the earliest and most striking signs of cell injury. Hence, in order to analyze different factors that are related to mitochondrial behavior following HSV-1 infection, THP-1 cells were infected with HSV-1 at an MOI of 0.2, and HI virus was used as a negative control. At 18 h postinfection, mitochondria and HSV-1 particles were stained with anti-TOMM20 (a mitochondrial import receptor subunit) and anti-ICP8 antibodies (Fig. 3A). Compared to the HI-virus-infected cells, the mitochondrial network of HSV-1-infected cells appeared to be disorganized and less aggregated around the nucleus. To statistically validate these observations, images acquired by confocal microscopy were deconvoluted and parameters related to the mitochondrial fraction count (MFC), total mitochondrial volume (MV) for the cell, and mitochondrial shape were analyzed. Twenty-eight infected cells and 64 noninfected randomly selected cells were imaged to calculate the respective MFCs. As observed in Fig. 3B, HSV-1-infected THP-1 cells demonstrated a more fragmented profile (mean MCF ≈ 7) than HI-virus-infected THP-1 cells (mean MCF ≈ 2.5), showing that HSV-1 infection led to ~3 times more mitochondrial network fragmentation than in HI-

**FIG 3** HSV-1 disrupted mitochondrial network. (A) Immunofluorescence of HSV-1-infected cells at 18 h postinfection by confocal microscopy. Nuclei were stained with DAPI (blue), HSV-1 particles with an anti-ICP8 monoclonal antibody (green), and mitochondria with an anti-TOMM20 monoclonal antibody (red). (B) Quantification of the mitochondrial fractional count (MFC). (C) Total mitochondrial volume (MV) normalized to the nucleus volume from confocal images. Box plots were statistically designed for 64 and 28 cells for HI (red) and HSV-1 at an MOI of 0.2 (blue), respectively. The P values in panels B and C are 10^-13 and 10^-6, respectively (Welch’s t test). (D) Evaluation of the sphericity of each mitochondrion; values describe an object’s closeness to a sphere when close to 1 and its opposite when close to 0. The quantity of mitochondria per sphericity value is represented by the density and was calculated for 3,736 and 2,624 mitochondria in cells infected with HI virus (red) and HSV-1 at an MOI of 0.2 (blue), respectively.
virus-infected cells. Moreover, when calculating the mean of the total mitochondrial volume for infected and HI-virus-infected cells, we observed a significant decline in the average mitochondrial volume that was more pronounced in infected cells (0.47 versus 0.33) (Fig. 3C).

The density distribution of mitochondrial sphericity was analyzed for 3,736 and 2,624 mitochondria in HI-HSV-1-infected cells (Fig. 3D, red curve) and HSV-1-infected cells (Fig. 3D, blue curve), respectively. Values close to 0 correspond to considerably deformed mitochondria, with an abstract shape, whereas values close to 1 correspond to spherical mitochondria. As observed in Fig. 3D, the comparison of mitochondria in HI-virus (red curve) versus infected cells (blue curve) highlighted a more pronounced sphericity index in HSV-1-infected cells. All together, these data demonstrated that HSV-1 seemed to fragment and shape the mitochondrial network by reducing their volume, leading to more spherical mitochondria.

**HSV-1 UL12.5 promotes the release of mitochondrial DNA to the cytosol.** Having demonstrated that HSV-1 induced mitochondrial fragmentation, we next examined the involvement of cytosolic DNA sensors in stress signaling pathways. Hence, THP-1 cells were infected with HSV-1, and released mtDNA was quantified. At 12 h postinfection, THP-1 cells were subjected to digitonin fractionation, and whole-cell extracts (WCE) or cytosolic extracts (CE) were blotted using the indicated antibodies. As visualized in Fig. 4A, compared to \( \beta \)-actin and GAPDH, TFAM (mitochondrial transcription factor A) and histone H2AX, specific markers for mitochondria and nuclei, respectively,
were missing from the cytosol extraction, demonstrating that the purification of the cytosol was not contaminated by remaining mitochondrial or nuclear proteins.

Quantification of mtDNA was performed by qPCR using the MT-COI (mitochondrial cytochrome c oxidase subunit I) gene present in the cytosolic fraction and normalized to the quantification of the nuclear gene β2M of the total fraction. Following HSV-1 infection, mtDNA in cytosolic extracts was increased in a dose-dependent manner by up to 2-fold (Fig. 4B), indicating a release of mtDNA. As the UL12.5 protein encoded by HSV-1 localizes to mitochondria and promotes rapid mtDNA depletion (8, 35), we investigated its impact in promoting IFN-β and A3A expression in uninfected cells. UL12.5 and UL98 (encoding a viral alkaline exonuclease homolog of UL12.5, in strain KOS37 UL98-SPA) were cloned in a pFLAP-GFP lentivirus-expressing vector, transfected into HEK-293T cells in order to produce lentiviral particles, and transduced in THP-1 cells. At 18 h posttransduction, Western blot analysis showed that UL12.5 and UL98 were produced similarly in HEK-293T and THP-1 cells (Fig. S3A). Confocal microscopy demonstrated a strong and significant colocalization of only UL12.5 with TOMM20 (Fig. S3B). Compared to one another, pFLAP-GFP (negative control)-, pFLAP-UL98-, and pFLAP-UL12.5-transduced THP-1 cells exhibited no significant increase of IFN-β (Fig. S3C), suggesting that UL12.5 probably needs viral and/or cellular cofactors to induce IFN-β and A3A expressions.

To explore the effect of UL12.5 in an infectious context, we used a recombinant UL12-deficient HSV-1 strain which was complemented by HCMV UL98, which impaired its ability to induce mtDNA stress but was still able to replicate similarly to the parental strain (KOS37 SPA) (36). THP-1 cells were infected with either KOS37 SPA or KOS37 UL98-SPA at an MOI of 0.02. The difference in MOI compared to the HSV-1 SC16 strain is due to the fact that the HSV-1 KOS37 strain has a more pronounced lytic effect in our experimental setup (Fig. 4B versus Fig. 4C and D). At 12 h postinfection, THP-1 cells were subjected to digitonin fractionation. WCE and CE were blotted and confirmed the absence of mitochondrial or nuclear contamination in the cytosol for both viral conditions (KOS37 SPA and KOS37 UL98-SPA) (Fig. S4A). Quantification of mtDNA demonstrated an ~2-fold increase of cytosolic mtDNA with KOS37 SPA but failed to induce mtDNA release with KOS37 UL98-SPA (Fig. 4C and D). Infection with KOS37 UL98-SPA resulted in ~1.7-fold attenuation of A3A and IFN-β expression (Fig. 4E; Fig. S4B), despite comparable UL12, UL12.5, and UL98 HSV-1 protein expression (Fig. S4C) and UL42, UL44, and US6 mRNA expression (Fig. S2A and S4D) and consistent with impaired antiviral innate immunity. All together, these data demonstrated that HSV-1 KOS37 SPA, expressing a functional UL12.5, was involved in the release of mtDNA in the cytosol that elicits a robust innate antiviral immune response.

Cytosolic mtDNA is involved in the innate immune signaling pathways. To explore the consequences of mtDNA release to the cytosol, THP-1 cells were specifically depleted of mtDNA by ethidium bromide (EtBr) treatment (37–39). As can be seen in Fig. 5A, 50 ng/mL EtBr treatment caused a strong decrease of the mtDNA content that was proportional to the duration of treatment. Treatment of THP-1 cells with 50 ng/mL EtBr for 4 weeks resulted in ~50% reduction of total mtDNA without loss of global mitochondrial mass, quantified by flow cytometry using TOMM20 staining (Fig. 5B; Fig. S2B). Furthermore, DNase I-expressing cells degrade ~60% of the cytosolic mtDNA (Fig. S2B). There is evidence of residual mtDNA persisting following EtBr treatment or in DNase I-expressing cells.

Mitochondrial DNA and mitochondria were simultaneously stained with anti-TFAM and anti-TOMM20 antibodies, respectively. TFAM proteins are involved in the packaging of the mtDNA to form nucleoids, leading to a positive correlation between the proportion of TFAM proteins and the pool of mtDNA (40), whereas TOMM20 proteins belong to the outer membrane of the mitochondria. As observed in Fig. 5B and C, a strong reduction of the TFAM signal in the EtBr-treated cells was evident, while no difference was observed for the mitochondrial mass represented by the TOMM20 signal.
As a control, protein samples extracted from HSV-1 infection in EtBr-treated and non-treated THP-1 cells were analyzed by Western blot and confirmed a massive depletion of mtDNA in EtBr-treated cells (Fig. 5D).

To prove that innate immune signaling pathways were not impaired in EtBr-treated THP-1 cells, IFN responsiveness was tested by quantifying A3A expression. As observed in Fig. 5E, EtBr-treated or nontreated cells incubated with IFN-supplemented medium for 24 h strongly expressed A3A ~100× over background in the same manner, demonstrating that the innate immune signaling pathways were not impacted by the EtBr treatment. EtBr-treated and nontreated THP-1 cells were then infected with HSV-1 SC16. At 18 h postinfection, IFN-β and A3A gene expression was quantified. As can be seen in Fig. 5F and G, IFN-β and A3A levels were ~2 times reduced in EtBr-treated THP-1 cells, demonstrating the involvement of mtDNA in triggering the innate immune signaling pathways upon HSV-1 infection. To prove that virus replication and gene

**FIG 5** EtBr-treated THP-1 cells are depleted of mtDNA and underexpress the A3A gene. THP-1 cells were treated with 50 ng/mL of EtBr for 4 weeks. (A) Quantification of the mtDNA content upon EtBr treatment by qPCR using primers mt3319 and mt3212 (59). Data were normalized to the quantification of the nuclear gene β2M. Mean values and SEM were calculated for three independent cell lines treated with EtBr. Differences relative to nontreated cells were estimated using an unpaired two-sided Student’s t test (**, P < 0.05). (B) Flow cytometry analysis of the TOMM20-positive cell population as a marker of the mitochondria mass in THP-1 cell lines, after 4 weeks of EtBr treatment, ns, not significant. (C) Immunofluorescence of EtBr-treated and nontreated THP-1 cell lines. Depletion of mtDNA was visualized by confocal microscopy. Nuclei were stained with DAPI (blue), mtDNA with an anti-TFAM monoclonal antibody (green), and mitochondria with an anti-TOMM20 monoclonal antibody (red). (D) Control of mtDNA depletion by immunoblotting with anti-TOMM20 and anti-TFAM antibodies against mitochondria and mtDNA, respectively. β-Actin was used as loading control. (E) Relative expression of A3A following IFN stimulation of THP-1 cell lines upon EtBr treatment or not, normalized to RPL13A housekeeping gene expression. (F and G) IFN-β and A3A profiling in EtBr-treated and nontreated THP-1 cells upon HSV-1 infection at an MOI 0.2 at 18 h postinfection. Data were normalized to HPRT1 housekeeping gene expression. Mean values and SEM were calculated for three independent experiments in duplicate (unpaired two-sided Student’s t test: **, P < 0.05; ***. P < 0.005; ns, not statistically significant). (H) Viral UL42 quantification by RT-qPCR at 18 h after HSV-1 infection (0.2 MOI). Mean values and SEM were calculated for three independent experiments in duplicate (n = 6) (unpaired two-sided Student’s t test: ns, not statistically significant).
expression were not inhibited in EtBr-treated cells relative to nontreated THP-1 cells, HSV-1 UL42, UL44, and US6 mRNAs were amplified by RT-qPCR. As observed in Fig. 5H and Fig. S2A, UL42, UL44, and US6 viral genes were similarly expressed in EtBr-treated and nontreated THP-1 cells, demonstrating that EtBr treatment does not impair viral replication.

**Cytosolic dsDNA is transcribed by RNA polymerase III and impacts RIG-1.** To identify the sensing molecules involved, THP-1 cells were infected at an MOI of 0.2. At 18 h postinfection, Western blot analysis showed increased levels of the phosphorylated forms of TBK1 (TBK1-P), IRF3 (IRF3-P), and IKKe (IKKe-P) as well as MOI-dependent increases in RIG-I, MDA5, and MAVS (Fig. 6A). Steady-state levels of STING and IKKe were unchanged. As observed, dose-dependent increases of TBK1-P and IRF3-P, key regulators of IFN production, suggest a signaling pathway via STING and/or MAVS. RT-qPCR performed on total extracted RNA from HSV-1-infected THP-1 cells demonstrated an upregulation of RIG-I and MDA5 expression ~22 times and 10 times over background levels (Fig. 6B and C). As the RNA polymerase III and cGAS DNA sensor molecules are upstream of STING, we investigated in detail these two pathways.

![FIG 6](image-url)
Expression of the DNA sensors RNA polymerase III and cGAS, as well as STING, was not affected by HSV-1 infection (Fig. 5A to C). To discriminate between the two DNA sensors, THP-1 cells were infected with HSV-1 at an MOI of 0.2 along with ML-60218, an inhibitor of RNA polymerase III or RU.521, a cGAS pathway inhibitor (41). When cells were incubated with 25 \( \mu \) M and 50 \( \mu \) M ML-60218, a strong extinction of TBK1 and IRF3 phosphorylated forms was observed (Fig. SSD), leading to a dose-dependent decrease in A3A as well as in IFN-\( \beta \) expression (Fig. 6D; Fig. S5E). To prove that ML-60218 treatment does not impair viral replication in THP-1 cells, early (annexin V\(^+\) 7-AAD\(^-\) cells) and late (annexin V\(^+\) 7-AAD\(^+\) ) apoptosis/necrosis as well as HSV-1 UL42, UL44, and US6 mRNA expression were analyzed. As visualized in Fig. S2A and F and S6A, THP-1 cells treated with ML-60218 did not induce apoptosis or impair viral replication. In contrast, when RU.521 was used, a less pronounced inhibition of A3A expression, \( \sim \)50\%, was detected (Fig. 6E). To prove that the cGAS pathway was functional, THP-1 cells were incubated with 500 ng of G3-YSD, an agonist of the cGAS pathway, in the presence of 5 \( \mu \) M or 10 \( \mu \) M RU.521. As observed in Fig. S6B, addition of G3-YSD led to a strong expression of A3A, which is inhibited in the presence of RU.521, demonstrating that the cGAS pathway is functional.

To show that cytosolic RNA synthesis by DNA-dependent RNA polymerase III binds to RIG-I, HSV-1-infected cells were lysed and incubated with anti-RIG-I or anti-MDA5 monoclonal antibodies along with an anti-IgG isotype used as a negative control to immunoprecipitate bound RNA. Total RNA was extracted and treated with DNase, and SYBR RT-qPCR was performed on NADH-ubiquinone oxidoreductase chain 4 (ND4), cytochrome c (MT-CYTC) and cytochrome b (MT-CYTB) mitochondrial genes along with UL42, UL44, and US6 viral genes. As observed in Fig. 6F and Fig. S2C, \( \sim \)3 to 4 times more ND4-, MT-CYTC-, and MT-CYTB-specific RT-PCR products were recovered only from the anti-RIG-I immunoprecipitates, which demonstrates that RIG-I remained the major sensing molecule. In comparison, RT-PCR products obtained from anti-MDA5 immunoprecipitates (Fig. 6G) or THP-1 cells treated with 25 \( \mu \) M and 50 \( \mu \) M ML-60218 (Fig. 6H) were not significantly different from the background. In addition, RIG-I bound HSV-1 viral RNA, compared to IgG, contained \( \sim \)2 times more UL42, UL44, and US6 specific RT-PCR products (Fig. S2C). Together, these data demonstrated that upon HSV-1 infection, IFN was induced by cGAS and RNA polymerase III/RIG-I pathways and both mitochondrial and viral RNAs could bind to RIG-I, leading to IFN pathway induction, with, however, mtRNA producing a higher impact on IFN production.

**Analyses of cytosolic mtDNA mutations reveal an APOBEC3A signature.** To analyze the mutational state of cytosolic mtDNA, MT-CO1 amplicons were generated by PCR from NI, HI-virus-infected, or HSV-1-infected cells at MOIs of 0.02 or 0.2 and were analyzed by next-generation sequencing (NGS). For each sample, reads obtained by Illumina sequencing were checked for quality and assessed using FastQC. Reads were analyzed using BLAST and a homemade Python script. The base substitution mutational signatures were described using a classification based on the six classes of single-base mutations: C to A, C to G, C to T, T to A, T to C, and T to G. To provide greater depth of insight into the operative mutational processes, the sequence context in which mutations occurred was incorporated by considering the bases immediately 5’ and 3’ of each mutated base.

All trinucleotide motifs (based on the reference sequences) as well as nucleotide substitutions contained in all reads were counted for each sample. Mutation content was obtained by dividing the number of each substitution in a given context by the count of the related trinucleotides sequence (i.e., number of 5’XpGpN3’-to-5’XpTpN3’ mutations divided by 5’XpCpN3’ motif count, with X and N being random nucleotides). The four data sets (NI, HI virus infected, HSV-1 infected at an MOI of 0.02, and HSV-1 infected at an MOI of 0.2) contained between 796,846 and 1,033,932 reads. Approximately 68.5% of the reads did not contain any mutation. The proportion of mutations to trinucleotide motifs is \( \sim \)0.41\% for each sample.

Mutation content was analyzed for each sample based on substitutions with their respective contexts. To estimate the implication of HSV-1 infection in modulating the
proportion of specific substitutions, the difference in mutation content observed between HI-virus-infected samples, samples infected with HSV-1 at an MOI of 0.02, samples infected with HSV-1 at an MOI of 0.2, and NI samples was investigated (Fig. 7). By comparing the differences between HSV-1-infected THP-1 cells, HI-virus-infected cells, and NI cells, we observed that C-to-T substitutions were predominant (Fig. 7). The most significant difference was observed for the 5’TpCpG-to-5’TpTpG substitution (the underlined nucleotide represents the mutated base) and slightly in the context 5’GpCpT to 5’GpTpT. The other substitutions fluctuated very slightly and had a difference lower than the greatest difference observed between NI and HI-virus-infected samples. We note that 5’TpCpG-to-5’TpTpG substitutions were not observed in a control sample generated from amplification of a plasmid following the identical protocol. Although 5’YpTpCpW has been described as a signature for A3A (42, 43), the preferred context we obtained was not without precedent; indeed, this specific signature is consistent with the mutagenic properties of A3A cytidine deaminase, which also preferentially targets 5’TpCpG motifs (33, 34).

**DISCUSSION**

These data demonstrated that in a human cellular model, HSV-1 infection disrupts the mitochondrial network with a remodeling of mitochondrial shape, resulting in a fragmentation of the mitochondria associated with a decrease in their volume and an increase in their sphericity. These data reinforce the notion that mitochondrial morphology in infected HSV-1 cells is not a static event, as HSV-1 infection leads to a loss of mitochondrial motility by fragmentation of the mitochondria (44). Therefore, since HSV-1 infects many cell types, it would not be surprising if it induces differential modulations of mitochondrial morphology in order to regulate cellular functions related to mitochondrial function, such as apoptosis or innate immunity signaling (45, 46).

**FIG 7** Pattern of mutations and contexts analyzed in mtDNA. The difference in mutation proportion between HSV-1 at MOI of 0.02 and 0.2 and the HI-virus-infected sample is shown.
been demonstrated that infection of HaCaT immortalized keratinocytes with HSV-1 resulted in an intracytoplasmic relocation of the Drp1 protein to the outer membrane of the mitochondria, which is involved in fission mechanisms, leading to fragmentation of the mitochondrial network (47).

Mitochondrial morphological changes can be associated with mtDNA release in the cytosol, potentially involving the action of the HSV-1 UL12.5 viral exonuclease, which targets mitochondria through its mitochondrial localization sequence (8, 35). However, in contrast to the mouse model (10), UL12.5 expression alone in THP-1 cells was not able and sufficient on its own to stimulate IFN-β (Fig. S3C), suggesting that other cofactors are probably involved in the release of mtDNA, leading to the induction of IFN-β and A3A. Indeed, it has been shown that ICP8 could interact with UL12.5, probably interfering with its function (9).

It is now well documented that cytosolic DNA, whatever its origins, plays an essential role in the IFN response, triggering the innate immune signaling pathway. Indeed, infection of a THP-1 lineage stably overexpressing cytoplasmic DNase I showed that cytosolic DNA was the trigger for IFN signaling following viral infection (Fig. 2D; Fig. S1D). Although we cannot exclude direct sensing of HSV-1 DNA, we demonstrated the contribution of cytosolic mtDNA in triggering IFN signaling response. In the same way, we showed that HSV-1-infected THP-1 cells lacking mtDNA following EtBr treatment (Fig. 5) demonstrated ~2-times-reduced expression of IFN-β and A3A, proving the involvement of mtDNA in triggering of the innate immune signaling pathway (Fig. 5F and G).

Which DNA-sensing pathway is involved during HSV-1 infection? A large number of cellular DNA sensors have been identified and characterized, which greatly broadened our knowledge of host-virus interaction. However, it has been shown that several DNA sensors exert distinct functions in different cell types or in response to different viruses. Among these cellular DNA sensors, cGAS can sense HSV-1 DNA and trigger host immune responses upon infection (48, 49). We demonstrated here that mtDNA released in the cytosol following HSV-1 infection was detected by RNA polymerase III (Fig. 6E). This is not without precedent; indeed, other viruses, such as adenovirus and Epstein-Barr virus, induce IFN-β expression in an RNA polymerase III-dependent manner through activation of RIG-I (11, 12). Moreover, it was recently demonstrated that HSV-1 infection leads to the relocalization and exposure of the cellular 5S rRNA pseudogene 141 transcript, allowing its recognition by RIG-I (50).

A specific inhibitor used against RNA polymerase III (ML-60218) significantly reduced IFN-β responses and A3A expression (Fig. 6D; Fig. S5E). Interestingly, we detected a more pronounced inhibition of IFN-β/A3A production in infected HSV-1 cells in the presence of ML-60218 (Fig. 6D; Fig. S5E) than in KOS37 UL98-SPA-infected cells (Fig. 4E; Fig. S4B) or with infected cells treated by EtBr (Fig. 5F and G). Concentrations of ML-60218 used in the present work were not toxic for the cells (Fig. S5F) but probably have a stronger inhibitory effect on IFN-β/A3A production than KOS37 UL98-SPA or cells treated with EtBr. Obviously, other unidentified parameters could influence the release of mtDNA that leads to IFN-β/A3A production.

As already documented, we also proved by using specific inhibitors that cGAS is also involved in the induction of IFN-β/A3A (Fig. 6E). Transcription of cytosolic mtDNA by RNA polymerase III allowed the synthesis of cytosolic and viral RNA that binds RIG-I (Fig. 6F; Fig. S2C), leading to the activation of the IFN signaling pathway. It is also conceivable that depending on HSV-1 tropism, signaling pathways leading to the production of IFN could be different.

Interestingly, the absence of a functional RNA polymerase III has been documented in children with severe varicella-zoster virus (VZV) infection (51), a virus also belonging to the family Herpesviridae. This worsening of the clinical picture has been associated with a defect in the IFN response, related to a genetic deficiency of RNA polymerase III due to heterozygous mutations (51). This highlights the essential role of RNA polymerase III in the antiviral response to VZV infection.
To increase the chance of survival, some viruses appear to have adopted the strategy of damaging the host cell mitochondrial DNA. Since mitochondria function as a source of energy and play an important role in antiviral immunity, it is possible that damage to mitochondrial DNA may help in evading mitochondrial antiviral immune responses (1). Some viruses have found different ways to degrade cytosolic mtDNA. Indeed, dengue fever virus induces mitochondrial elongation through the viral protein NS4B, and mitochondrial hyperfusion appears to have a proviral role in reducing IFN response (52). In the same vein, influenza virus-encoded NS1 protein, which interacts with the cytosolic mtDNA, leads to a reduction in the induction of IFN (5). As A3 DNA mutators are downstream of IFN induction, another viral strategy could be to exclude A3 from replication and transcription sites. Recent studies reported the relocation of A3A from the nuclear to cytoplasmic compartment, as is the case during HSV-1 infection (53, 54). This was observed for a variety of viral proteins, in particular with HSV-1 and ICP6, EBV and BORF-2, and KSHV and ORF61 (53, 54).

As a consequence, A3A relocation in the cytoplasm will lead to an increase of C-to-T editing in cytosolic DNA. Indeed, by performing deep sequencing of cytosolic mtDNA, we observed an increase of C-to-T editing occurring mainly in a specific context, 5’TpCpG (Fig. 7). As A3B is exclusively located in the nucleus, this excludes mtDNA editing by this enzyme (55, 56). These characteristics are consistent with the mutagenic properties of A3A cytidine deaminase, which was described to target 5’TpCpG motifs in single-stranded DNA (33). On the other hand, the role of A3A and A3B in cancer was highlighted by the fact that many cancer genomes encode tens of thousands of C-to-T transitions frequently in the 5’TpCpA and 5’TpCt dinucleotide context (30–32) and in 5’TpCpG, if the genomic underrepresentation of this motif is accounted for (33, 34). The difference between the signatures of nuclear and mitochondrial DNA could be linked to the affinity of A3A enzyme for the ssDNA substrate, which could be a function of substrate nucleotide composition, length, secondary structure, and pH variation (34).

The innate immune system is sensitive to dsDNA, with cellular sources of immune stimulatory self-DNA including viral DNA as well as nuclear DNA, cytosolic mtDNA, and DNA from phagosomal compartments. This suggests that foreign DNA, whatever its origin, present in the cytosol constitutes a danger signal and should be shut down by enzymatic processes, in order to reduce innate immune activation. In this sense, IFN activated A3A gene expression, which led to increased catabolism of ssDNA generated by cytoplasmic exonucleases (13, 24, 56). We also previously noticed that up to 17% of primary CD4+ T cells showed signs of A3-edited cytosolic mtDNA, demonstrating a very dynamic image of the mitochondrial network (55).

Together, these results demonstrate that viral stress induced by HSV-1 results in mitochondrial fragmentation. These changes in term of shape and sphericity help the release of mitochondrial DNA. Accumulation of mtDNA in the cytosol that probably escaped autophagic clearance will induce RNA polymerase III/RIG and cGAS pathways, leading to the expression of type I IFN and A3A cytidine deaminase. A3A could then probably lead to the catabolism of cytosolic mtDNA to decrease the danger signal. Finally, an HSV-1 infection, and probably any pathways involved in the induction of IFN, will inevitably lead to the presence of somatic mutations in cytosolic DNA and, to a lesser extent, mutations in nuclear DNA, assuming that the cell survives. As chronic inflammation underlies many cancers, while we live in an antigenic world, APOBEC3 mutator enzyme-induced somatic mutations are an inevitable part of human biology.

MATERIALS AND METHODS

Reagents. RNA polymerase III inhibitor (ML-60218) was from Merck Millipore, and cGAS inhibitor (ihn-ru521) and cGAS agonist (G3-15SD; hrh-ydna) were from InvivoGen. MDA5 (D74E4; rabbit monoclonal antibody [MAB] S321), STING (antibody 3337), phospho-IRF-3 (Ser396) (4D4G; rabbit MAB 4947), IRF-3 (D64C; XP rabbit MAB 11904), RIG-I (D14G6; rabbit MAB 3743), MAVS (antibody 3993), TBK1/NAK (D1B4; rabbit MAB 3504), phospho-TBK1/NAK (Ser 172) (D52C2; rabbit MAB 5483), IKKe (D20G4; rabbit MAB 2905), phospho-IKKe (Ser172) (D1B7; rabbit MAB 8766), β-actin horseradish peroxidase (HRP) conjugate (13E5; rabbit MAB G8795), HRP-linked anti-rabbit IgG (antibody 7074), and histone H2AX (antibody 2595) antibodies were from Cell Signaling Technology. GAPDH (mouse MAB G8795) was from Sigma-Aldrich.
Anti-mouse IgG, HRP-linked (antibody NA931V) was from GE Healthcare. Monoclonal anti-
β-actin (polyclonal, antibody A9874) and actinomycin D (D1411) were from Sigma-Aldrich. Monoclonal anti-
mouse TFAM (18G102B2E11) and Turbo DNase (2 U/µL; AM2238) were from Invitrogen. GlycoBlue
coprecipitant (AM9515) was from Applied Biosystems. TOM20 (EPR15581-39; rabbit Mab ab186734),
ICP8 (11E2; mouse MAb ab20194), and mCherry (1C51; rabbit Mab ab125096) were from Abcam. RIG-I
(D33H10; rabbit Mab 4200) and mouse anti-ribb IgG (conformation specific; L27A9; MAB 3678), cell
lysis buffer (98003), protein A magnetic beads (73778), glycine (7005), and phenylmethylsulfonyl fluoride
(PMSF) (8553) were from Cell Signaling. Recombinant RNasin RNase inhibitor (N2515) was from
Promega. Secondary Alexa Fluor 488 goat anti-mouse immunoglobulin (A11029), Alexa Fluor 488 goat
anti-rabbit immunoglobulin (A11034), Alexa Fluor 633 goat anti-mouse immunoglobulin (A21053), Alexa
Fluor 647 goat anti-rabbit immunoglobulin (A21244), Fluoromount-G with DAPI (4',6-diamidino-2-phenyl-
indole; 00-4599-52), and SlowFade Diamond antifade mountant without DAPI (S36972) were from
Invitrogen. DAPI solution 1.0 mg/ml (S96907) was from BD Pharmingen. Monoclonal Alexa Fluor 647
mouse anti-H2AX (g5139) (S60447) was from BD Pharmingen. The annexin V apoptosis detection Kit
(B88007) with annex in V eFluor 450 (48-8006-69) and the fixable viability dye eFluor 780 (65-0865-14)
were from eBioscience. CD14 Microbeads (human; 130-050-201) were from Miltenyi Biotec. JetPrime
(114-07) was from Polyplus Transfection, and Lipofectamine RNAiMAX (13778075) was from Invitrogen.
Sodium pyruvate (100 mM; 11360070) and HEPES buffer (1 M; 15630) were from Gibco. Uridine (U3003),
glucose solution (100 g/L; G8644), and ethidium bromide solution (10 mg/ml in H2O; E1510) were from
Sigma. NucleoSpin gel and the PCR clean-up Kit (740609) were from Macherey-Nagel. The Quant-IT
dsDNA assay Kit (high sensitivity; Q33120) was from Invitrogen, and the NEBNext DNA library prepara-
tion Kit (E7645) and NEBNext multiplex oligonucleotides for Illumina (E7600) were from New England Biolabs.

Cell lines. THP-1 cells (ATCC TIB-202) were maintained in Roswell Park Memorial Institute medium
(RPMI 1640; Gibco), supplemented with heat-inactivated fetal calf serum (10%), penicillin (50 U/ml),
streptomycin (50 µg/ml) and β-mercaptoethanol (0.05 mM) were grown in 150-cm2 cell culture flasks.
Human HeLa cells, Vero cells, and HEK-293T cells were maintained in Dulbecco’s modified Eagle’s
medium (DMEM; Gibco), supplemented with heat-inactivated fetal calf serum (10%), penicillin (50 U/ml),
and streptomycin (50 µg/ml) and were grown in 75-cm2 cell culture flasks in a humidified atmosphere
containing 5% CO2.

Construction of THP-1 DNase wild-type cell line. From the lab collection, a plasmid harboring the
sequence of the cDNA of the wild-type DNase I gene, previously designed by GeneCust, was cloned in
pcDNA3.1/V5 His-TOPO under the control of the CMV promoter, and the mCherry plasmid was cloned in
sequence of the cDNA of the wild-type DNase I gene, previously designed by GeneCust, was cloned in
pCR2.1 vector. Both were digested and ligated together. Ligated plasmid pcDNA 3.1 DNase (WT)-mCherry
was used to create the sequence of interest inside the lentiviral vector pTRIP harboring the promoter EF1a.
HEK-293T cells were then used to produce lentiviral vectors upon Fugene HD (Promega) transfection of DNase-
encoding viral proteins C-terminally tagged with a sequential peptide af-

Construction of THP-1 cell lines expressing UL98 and UL12.5. UL98-SPA and UL12.5-SPA sequences
were encoded via the genomic DNA (gDNA) of THP-1 cells. Cells were infected with several dilutions
(10-2 to 10-5) of HSV-1 in medium without serum. After 1 h, the inoculum was removed to avoid secondary infection and wells were covered with a solution of Avicel (2.4%)-complete DMEM. After 72 h at 37°C, the solution of Avicel was removed, and cells were

Preparation of mtDNA-depleted cell lines. THP-1 cells were treated with 50 ng/ml of ethidium bromide
(Sigma) or 4.5 g/L (Sigma). PBMC purification. Blood from healthy patients was provided by the Office of Biological Products of the Institut Pasteur. PBMC purification based on the Ficoll method (Ficoll-Paque Plus; GE Healthcare) was performed according to the guidelines of MACS Miltenyi Biotec. Primary monocytes were isolated using CD14 microbeads (Miltenyi Biotec) and maintained in supplemented RPMI for 3 days.

Virus strain and plaque assay. The wild-type HSV-1 strain SC16 was kindly provided by Marc
Labetoulle (I2BC, Gif-sur-Yvette, France). RIG-I (D33H10; rabbit Mab 4200) and mouse anti-ribb IgG (conformation specific; L27A9; MAB 3678), cell lysis buffer (98003), protein A magnetic beads (73778), glycine (7005), and phenylmethylsulfonyl fluoride (PMSF) (8553) were from Cell Signaling. Recombinant RNasin RNase inhibitor (N2515) was from
Promega. Secondary Alexa Fluor 488 goat anti-mouse immunoglobulin (A11029), Alexa Fluor 488 goat
anti-rabbit immunoglobulin (A11034), Alexa Fluor 633 goat anti-mouse immunoglobulin (A21053), Alexa
Fluor 647 goat anti-rabbit immunoglobulin (A21244), Fluoromount-G with DAPI (4',6-diamidino-2-phenyl-
indole; 00-4599-52), and SlowFade Diamond antifade mountant without DAPI (S36972) were from
Invitrogen. DAPI solution 1.0 mg/ml (S96907) was from BD Pharmingen. Monoclonal Alexa Fluor 647
mouse anti-H2AX (g5139) (S60447) was from BD Pharmingen. The annexin V apoptosis detection Kit
(B88007) with annex in V eFluor 450 (48-8006-69) and the fixable viability dye eFluor 780 (65-0865-14)
were from eBioscience. CD14 Microbeads (human; 130-050-201) were from Miltenyi Biotec. JetPrime
(114-07) was from Polyplus Transfection, and Lipofectamine RNAiMAX (13778075) was from Invitrogen.
Sodium pyruvate (100 mM; 11360070) and HEPES buffer (1 M; 15630) were from Gibco. Uridine (U3003),
glucose solution (100 g/L; G8644), and ethidium bromide solution (10 mg/ml in H2O; E1510) were from
Sigma. NucleoSpin gel and the PCR clean-up Kit (740609) were from Macherey-Nagel. The Quant-IT
dsDNA assay Kit (high sensitivity; Q33120) was from Invitrogen, and the NEBNext DNA library prepara-
tion Kit (E7645) and NEBNext multiplex oligonucleotides for Illumina (E7600) were from New England Biolabs.
fixed with 4% paraformaldehyde after washing. Plaques were stained with crystal violet (2.3%) and counted manually.

**HSV-1 infection.** THP-1 cells or primary monocytes were infected with HSV-1 at a multiplicity of infection (MOI) of 0.1, 0.2, 0.5, 1, or 3 for 18 h. As a negative control, cells were infected at an MOI of 0.5 with HSV-1 that had been inactivated at 70°C for 30 min. For both infections, viruses were absorbed in serum-free RPMI for a defined period at 37°C before dilution with complete RPMI. To study the role of the RNA polymerase III or cGAS, 2 h postinfection, inhibitors were added in the culture medium at final concentrations of 25 and 50 μM for ML-60218 (Merck Millipore) or at final concentrations of 1, 5, and 10 μM for RU.521 (InvivoGen).

**THP-1 stimulation.** DNase cells were assayed for their ability to degrade DNA, compared to wild-type THP-1 cells, by transfection of 250 ng of DNA with JetPRIME (Polyplus Transfection). EtBr-treated and nontreated THP-1 cell lines were tested for their ability to respond to IFN stimulation by addition of IFN-supplemented supernatant to the culture medium for 24 h.

The ability of RU.521 (InvivoGen) to inhibit the cGAS pathway, as well as the expression of IFN and APOBEC3A, was tested by incubating THP-1 cells with the inhibitor at final concentrations of 5 μM and 10 μM for 1 h before the transfection of 500 ng of G3-YSD (InvivoGen) using Lipofectamine RNAiMAX (Invitrogen).

**Subcellular fractionation.** Ten million THP-1 cells were infected with HSV-1. At 12 h postinfection, cells were recovered and separated into equal samples. One sample was considered the whole-cell extract (WCE). The second sample was resuspended in digitonin buffer (150 mM NaCl, 50 mM HEPES [pH 7.4], and 25 μg/mL of digitonin) and other aliquots were kept for future RNA or protein analysis. After 10 min of incubation with gentle agitation, lysate was centrifuged for 3 min at 980 × g to collect nuclei and unbroken cells. Centrifugation was repeated three times. Supernatant was centrifuged at 10,000 × g for 10 min to collect mitochondria. Finally, supernatant was centrifuged at 20,000 × g for 30 min to pellet other organelles. The clear cytosolic fraction was collected in a fresh tube.

**Coimmunoprecipitation.** HSV-1-infected cells were incubated at a final concentration of 2% of formaldehyde for 1 h under gentle agitation at room temperature. The cross-linking step was stopped by addition of glycine at a final concentration of 0.2 M for 5 min at room temperature. Cells were centrifuged at 1,800 rpm for 5 min at 4°C and lysed in cell lysis buffer (Cell Signaling), supplemented with RNasin (40 U/mL; Promega) and PMSF (1 mM; Cell Signaling) on ice for 5 min. Cell lysates were sonicated three times on ice and centrifuged for 10 min. The clear cell lysate, already preclarified with protein A magnetic beads (Cell Signaling) according to the manufacturer’s instructions, was incubated with the antibody of interest overnight, i.e., IgG (conformation specific, L27A9), RIG-I (D33H10), or MDA5 (D74E4; Cell Signaling) diluted 1/500 or 1/100. Protein A magnetic beads were added to the lysate-antibody mix for 20 min at room temperature under agitation. After incubation, beads were harvested and washed three times. To avoid DNA contamination, beads were treated with Turbo DNase (Invitrogen) according to the manufacturer’s instructions and washed twice before elution. Elution was performed by addition of 0.2 M glycine at three times the volume of beads under agitation for 10 min. Using the magnetic separation rack, supernatant with proteins of interest was separated from beads and was neutralized by the addition of an equivalent volume of Tris-HCl (pH 8). This elution step was repeated twice.

**Western blotting.** Cells were treated with lysis buffer (0.5% Nonidet P-40, 20 mM Tris-HCl [pH 7.4], 120 mM NaCl, and 1 mM EDTA and EDTA-free protease inhibitors [Roche Applied Science]). Total protein samples were loaded on NuPAGE 4 to 12% bis-Tris gel (Invitrogen) and transferred to a nitrocellulose membrane (Invitrogen) according to the manufacturer’s instructions. Specific primary antibodies were incubated overnight, followed by HRP-linked secondary antibodies for 1 h according to standard protocols. Detection was performed with a chemiluminescence assay (Pierce). β-Actin HRP antibody (Sigma) diluted 1/5,000,000 was used to detect β-actin protein as a loading control. At 18 h postinfection, HSV-1-infected cells were tested for the proteins of the IFN pathway with antibodies from the RIG-I pathway antibody sampler kit (Cell Signaling) using dilutions recommended by the manufacturer. The expression of DNase-mCherry protein from plasmids was verified with anti-mCherry antibody (Abcam) used at 1/500. SPA tag detection of HEK-293T transfected cells or stable THP1 UL98-SPA- or UL12.5 SPA-expressing cells was performed using anti-Flag HRP-coupled antibody (Sigma) diluted 1/5,000. Depletion of the mtDNA in EtBr-treated and nontreated THP-1 cell lines was controlled using anti-TOMM20 (Abcam), as a marker of mitochondria, and anti-TFAM (Invitrogen) antibodies, as a marker of mitochondrial DNA, at 1/5,000 and 1/1,000, respectively. Purity of the whole-cell extract and the cytosol from the subcellular fractionation was controlled using anti-GAPDH (1/10,000; Sigma) as a marker of cytosol, anti-H2AX (1/1,000; Cell Signaling) as a marker of nuclei, and anti-TFAM (1/1,000) antibodies. Protein expression of UL12, UL12.5, and UL98 upon KOS37 SPA or KOS37 UL98-SPA infection at 12 h postinfection was verified with anti-Flag HRP-coupled antibody (1/5,000; Sigma).

**DNA extraction and quantitative PCR.** Total DNA from infected cells, from samples produced by the subcellular fractionation and from EtBr cell lines were extracted with the MasterPure purification kit (Epicentre Illumina Company). mtDNA was quantified using quantitative PCR based on TaqMan (Applied Biosystems) or based on SYBR green (Applied Biosystems). Conditions were 10 min at 95°C followed by 40 cycles of 15 s at 95°C, 15 s at 55°C, and 1 min at 68°C and were 2 min at 50°C and 10 min at 95°C followed by 40 cycles of 30 s at 95°C and 1 min at 60°C with a melting curve step, respectively. Data were normalized to the expression levels of the nuclear reference gene β2M. Primers are described in Table S1.

**Real-time PCR.** All RNA was extracted from infected cells using the RNeasy Plus minikit (Qiagen), and RNA from coimmunoprecipitated eluted samples was harvested using the standard protocol of RNA
precipitation with 3 M sodium acetate, 100% ethanol, and 1 μL of GlycoBlue (Applied Biosystems). cDNA synthesis was performed using QuantiTect reverse transcription kit (Qiagen 205311) from 50 ng to 1 μg of RNA. Expression of APOBEC3s, IFN-α, and IFN-β genes was assayed by real-time PCR based on TaqMan (Applied Biosystems) along with RPL13A and HPRT1 as reference genes. Primers for the amplification of A3 as well as for the quantification of mtDNA were previously described (24, 59). The other primers and probes are presented in Table S1.

Deep sequencing. For the analysis of A3-induced mutations in mitochondrial DNA, cells were infected with HSV-1 for 18 h. Total DNA from cytosolic fractions was extracted, and the MT-COI gene was amplified by standard PCR using the primers dHCoxI fwd and dHCoxI rev (Table S1). For the MT-COI gene, conditions were 5 min at 95°C, then 35 cycles of 30 s at 95°C, 30 s at 60°C, and 1 min at 72°C, followed by 10 min at 72°C. PCR products were purified with the NucleoSpin gel and PCR clean-up kit (Macherey-Nagel) and quantified using the Quant-iT dsDNA assay kit (high sensitivity; Invitrogen) according to the manufacturer’s instructions. The preparation of the DNA library was performed with the NEBNext DNA library preparation kit and NEBNext multiplex oligonucleotides for Illumina (New England Biolabs) for the fragmentation and multiplex-adapter ligation steps. Deep sequencing was performed with illumina cBot and GAIIx technology.

Sequenced reads were cleaned from adapter sequences and trimmed using Trim Galore! 0.4.3 with a quality Phred score cutoff of 28 and a minimum length of 20 (www.bioinformatics.babraham.ac.uk/projects/trim_galore), and quality was assessed using FastQC (www.bioinformatics.babraham.ac.uk/projects/fastqc). Sequencing reads were subjected to BLAST search using BLASTn (60, 61) with the following parameters: -task dc-megablast -perc_identity 20 -max_target_seqs 1 -outfmt "b qseqid qstart qend sstart send qseq sseq eval bitscore score length mismatch gaps qframe sframe." Output was analyzed using a homemade python script (available on request). Further analyses were performed with R version 3.6.0 and R studio, as well as the Bioconductor packages ggplot2 and reshape2.

Immunofluorescence. THP-1 cells were plated and infected in 4-well chamber poly-l-lysine glass slides (Lab-Tek, Thermo Scientific). Cells were washed with phosphate-buffered saline (PBS), fixed in 4% paraformaldehyde, and permeabilized by incubation in 50/50 acetone-methanol (Sigma). Mouse monoclonal anti-ICP8 antibody (Life Technologies) and rabbit monoclonal anti-TOMM20 antibody (Abcam) were incubated at 1/750 and 1/250, respectively, in PBS-bovine serum albumin (BSA; 0.5%) for 1 h at room temperature. Secondary antibodies Alexa Fluor 488 goat anti-mouse and Alexa Fluor 647 goat anti-rabbit (Invitrogen) were both used at 1/750 for 45 min at room temperature, protected from the light. SPA tag detection in UL98-SPA- and UL12.5-SPA-expressing THP-1 cell lines was performed using Alexa Fluor 488 goat anti-mouse and Alexa Fluor 633 goat anti-rabbit (Invitrogen) were both used at 1/750 for 45 min at room temperature. Slides were mounted with Fluoromount-G medium, with DAPI (Invitrogen). Imaging was performed using a Leica SP8 confocal microscope. Both EtBr-treated and nontreated THP-1 cell lines were fixed and permeabilized in 4-well chamber poly-l-lysine glass slides as previously described (37–39). Rabbit monoclonal anti-TOMM20 antibody (Abcam) was used at 1/250 to stain mitochondria, and mouse monoclonal anti-TFAM antibody (Invitrogen) was used to stain mtDNA at 1/50 for 1 h. Secondary antibodies Alexa Fluor 488 goat anti-rabbit and Alexa Fluor 633 goat anti-mouse immunoglobulin (Invitrogen) were both used at 1/750 for 45 min at room temperature. Slides were mounted with Fluoromount-G medium, with DAPI (Invitrogen). Imaging was performed using a Leica SP8 confocal microscope. Transfected HeLa cells with pcDNA 3.1 DNase-mCherry were washed with PBS, fixed in 4% paraformaldehyde, and directly mounted with Fluoromount-G medium, with DAPI (Invitrogen). Imaging was performed using a Leica DM IR8 microscope. All images were analyzed using Fiji software (Imagej) version 1.52p.

Mitochondrial behavior analysis. All images were deconvolved with Huygens Professional version 19.04 (Scientific Volume Imaging, The Netherlands), using the CMLE algorithm, with stop criteria when quality was 0.05 or signal-to-noise ratio (SNR) was 20 or when a maximum of 40 iterations was reached. From the deconvoluted images, the nucleus and mitochondrion signals were processed separately to segment the different structures present. The nucleus channel was first smoothed using a 3- by 3- by 3-pixel median filter in order to remove the deconvolution artifacts and then thresholded using the Otsu algorithm (62). Finally, a morphological opening operation with a sphere element with a radius of 1 was used to remove possible small remaining salient structures from the binary image. The mitochondrion channel was thresholded using the Otsu algorithm. The different mitochondria were identified using a connected-component object labeling algorithm, which detects connected regions in the binary volume. For more precision, all structures under 10 pixels were removed for the analysis. Each region was then considered mitochondria, and several quantifications were computed from them.

The statistical analysis of the impact of HSV-1 infection on the mitochondrial network was performed using three characteristics: (i) the mitochondrial fraction count (MFC) (63), (ii) the total mitochondrial volume (MV) for the cell, normalized to the nucleus volume, and (iii) the mitochondrial shape through the computation of sphericity, which indicates that an object’s shape is close to a sphere when close to 1 and the opposite when close to 0. Both MFC and MV were computed per cell, allowing comparison using category plots. Sphericity was observed per mitochondrion directly. As we may be dealing with small objects that do not apply any noise filtering on the image, we were able to observe a large number of objects with a sphericity of $>1$. This phenomenon is due to the discretization of the values used to compute the sphericity, mainly the surface calculation when the particle size is drastically low. The particles were removed from the analysis and considered either noise or debris.

Flow cytometry analysis. For DNA double-strand break analysis, at 18 h after HSV-1 infection, cells were washed with PBS, fixed in 2 to 4% ice-cold paraformaldehyde (Electron Microscopy Sciences) for 10 min, and permeabilized in 90% ice-cold methanol (Sigma) for 30 min. After PBS washing, cells were...
incubated with 1:100 diluted Alexa Fluor 647 mouse anti-γH2A.X (pS139) antibody (BD Pharmingen) in PBS–5% BSA for 1 h. Mitochondrial quantification was performed by staining EtBr-treated and non-treated THP-1 cells with the rabbit monoclonal anti-TOMM20 antibody (Abcam) diluted 1/40 in PBS–5% BSA for 1 h. After PBS washings, cells were incubated with 1/100-diluted Alexa Fluor 488 goat anti-rabbit in PBS–5% BSA for 45 min. For apoptosis analysis, HeLa and THP-1 cells were collected and washed with PBS, stained with 7-AAD viability staining solution (1/40; eBioscience), then washed with annexin V binding buffer (eBioscience), and stained by anti-annexin V eFluor 450 (1/20; eBioscience). Treatment with 100 μM etoposide in dimethyl sulfoxide was used as positive control. All stained samples were acquired on a MACSQuant analyzer (Miltenyi Biotech), and data were analyzed with FlowJo software (Tree Star, Inc.; version 8.7.1).

**SUPPLEMENTAL MATERIAL**

Supplemental material is available online only.

**FIG S1**, TIF file, 0.2 MB.
**FIG S2**, TIF file, 0.2 MB.
**FIG S3**, TIF file, 0.5 MB.
**FIG S4**, TIF file, 0.3 MB.
**FIG S5**, TIF file, 0.3 MB.
**FIG S6**, TIF file, 0.1 MB.

**TABLE S1**, DOCX file, 0.02 MB.

**ACKNOWLEDGMENTS**

We thank Emmanuelle Pitré, Jérémy Dufloo, Mariaconetta Sicurella, Audrey Salles, Jean-Yves Tinevez, Thomas Vallet, Emma Partiot, Raphaël Gaudin, and Anastasia Komarova for helpful discussions and advice. The wild-type HSV-1 strain SC16 was kindly provided by Marc Labeltouille (I2BC, Gif-sur-Yvette, France). James Smiley (University of Alberta, Canada) kindly sent the KOS37 SPA and KOS37 UL95-SPA strains; the lentiviral vector was kindly provided by Pierre Charneau.

This work was supported by grants from the Institut Pasteur. N.B. and P.K. were supported by the Allocation de Recherche du Ministère de la Recherche et Ligue contre le Cancer, respectively. The funders had no role in study design, data collection and interpretation, or the decision to submit the work for publication.

N.B. and J.-P.V. designed the research. N.B., R.S., V.C., P.K., and S.R. performed the experiments. H.B. and G.B. performed NGS and analysis. N.B., R.S., V.C., P.K., G.B., S.R., H.B., M.V., S.W.-H., and J.P.-V. analyzed the data. N.B. and J.P.-V. wrote the paper. All authors reviewed and approved the manuscript.

No potential conflicts of interest were disclosed.

**REFERENCES**

1. Mills EL, Kelly B, O’Neill LAJ. 2017. Mitochondria are the powerhouses of immunity. Nat Immunol 18:488–498. https://doi.org/10.1038/ni.3704.
2. Young LS, Rickinson AB. 2004. Epstein-Barr virus: 40 years on. Nat Rev Cancer 4:757–768. https://doi.org/10.1038/nrc1452.
3. Ming-Ju H, Yih-Shou H, Tzy-Yen C, Hui-Ling C. 2011. Hepatitis C virus E2 protein induce reactive oxygen species (ROS)-related fibrogenesis in the HSC-T6 hepatic stellate cell line. J Cell Biochem 112:233–243. https://doi.org/10.1002/jcb.22926.
4. Takada S, Shirakata Y, Kaneniwa N, Koike K. 1999. Association of hepatitis B virus X protein with mitochondria causes mitochondrial aggregation at the nuclear periphery, leading to cell death. Oncogene 18:6965–6973. https://doi.org/10.1038/sj.onc.1203188.
5. Moriyama M, Koshiba T, Ichinohe T. 2011. Hepatitis C virus E2 protein induce reactive oxygen species (ROS)-related fibrogenesis in the HSC-T6 hepatic stellate cell line. J Cell Biochem 112:233–243. https://doi.org/10.1002/jcb.22926.
6. Saffran HA, Pare JM, Corcoran JA, Weller SK, Smiley JR. 2007. Herpes simplex virus eliminates host mitochondrial DNA. EMBO Rep 8:188–193. https://doi.org/10.1038/sjembr.7400878.
7. Reuven NB, Antoku S, Weller SK. 2004. The UL12.5 gene product of herpes simplex virus type 1 exhibits nuclease and strand exchange activities but does not localize to the nucleus. J Virol 78:4599–4608. https://doi.org/10.1128/jvi.78.9.4599-4608.2004.
8. West AP, Khoury-Hanold W, Staron M, Tal MC, Pineda CM, Lang SM, Bestwick M, Duguay BA, Raimundo N, MacDuff DA, Kaech SM, Smiley JR, Means RE, Iwasaki A, Shadel GS. 2015. Mitochondrial DNA stress primes the antiviral innate immune response. Nature 520:553–557. https://doi.org/10.1038/nature14156.
9. Hornung V, Ablasser A, Charrel-Dennis M, Bauernfeind F, Horvath G, Caffrey DR, Latz E, Fitzgerald KA. 2009. AIM2 recognizes cytosolic dsDNA and forms a caspase-1-activating inflammasome with ASC. Nature 458: 514–518. https://doi.org/10.1038/nature07725.
10. Chiu YH, Macmillan JB, Chen ZJ. 2009. RNA polymerase III detects cytosolic DNA and induces type I interferons through the RIG-I pathway. Cell 138:576–591. https://doi.org/10.1016/j.cell.2009.06.015.
11. Suspène R, Mussil B, Laude H, Galv C, Berry N, Bouzidi MS, Thiers V, Wain-Hobson S, Vartanian JP. 2017. Self-cytoplasmic DNA upregulates the mutator enzyme APOBEC3A leading to chromosomal DNA damage. Nucleic Acids Res 45:3231–3241. https://doi.org/10.1093/nar/gkx001.
1 and type 2 disrupt mitochondrial dynamics in human keratinocytes. Arch Virol 163:2663–2673. https://doi.org/10.1007/s00705-018-3890-y.

48. Orzalli MH, DeLuca NA, Knipe DM. 2012. Nuclear IFI16 induction of IRF-3 signaling during herpesviral infection and degradation of IFI16 by the viral ICP0 protein. Proc Natl Acad Sci U S A 109:E3008–E3017. https://doi.org/10.1073/pnas.1211302109.

49. Sun L, Wu J, Du F, Chen X, Chen ZJ. 2013. Cyclic GMP-AMP synthase is a cytosolic DNA sensor that activates the type I interferon pathway. Science 339:786–791. https://doi.org/10.1126/science.1232458.

50. Chiang JJ, Sparrer KMJ, van Gent M, Lassig C, Huang T, Osterrieder N, Hopfner KP, Gack MU. 2018. Viral unmasking of cellular 5S rRNA pseudogene transcripts induces RIG-I-mediated immunity. Nat Immunol 19:53–62. https://doi.org/10.1038/s41590-017-0005-y.

51. Ogunjimi B, Zhang SY, Sorensen KB, Skipper KA, Carter-Timofte M, Kerner G, Luecke S, Prabakaran T, Cai Y, Meester J, Osterrieder N, Hopfner KP, Gack MU. 2017. Inborn errors in RNA polymerase III underlie severe varicella zoster virus infections. J Clin Invest 127:3543–3556. https://doi.org/10.1172/JCI92280.

52. Chatel-Chaix L, Cortese M, Romero-Brey I, Bender S, Neufeldt CJ, Fischl W, Scaturro P, Schieber N, Schwab Y, Fischer B, Ruggieri A, Bartenschlager R. 2016. Dengue virus perturbs mitochondrial morphodynamics to dampen innate immune responses. Cell Host Microbe 20:342–356. https://doi.org/10.1016/j.chom.2016.07.008.

53. Cheng AZ, Moraes SN, Attarian C, Yockteng-Melgar J, Jarvis MC, Biollati M, Galitska G, Dell’Oste V, Frappier L, Bierle CJ, Rice SA, Harris RS. 2019. A conserved mechanism of APOBEC3 relocalization by herpesviral ribonucleotide reductase large subunits. J Virol 93:e01539-19. https://doi.org/10.1128/JVI.01539-19.

54. Stewart JA, Holland TC, Bhagwat AS. 2019. Human herpes simplex virus-1 depletes APOBEC3A from nuclei. Virology 537:104–109. https://doi.org/10.1016/j.virol.2019.08.012.

55. Mussil B, Suspene R, Caval V, Durandy A, Wain-Hobson S, Vartanian JP. 2016. Mitochondrial damage upon HSV-1 infection. Arch Virol 161:2463–2473. https://doi.org/10.1007/s00705-016-2890-y.

56. Suspene R, Aynaud M, Guetard D, Henry M, Eckhoff G, Marchio A, Pineau P, Dejean A, Vartanian JP, Wain-Hobson S. 2011. Somatic hypermutation of human mitochondrial and nuclear DNA by APOBEC3 cytidine deaminases, a pathway for DNA catabolism. Proc Natl Acad Sci U S A 108:4858–4863. https://doi.org/10.1073/pnas.1009687108.

57. Duguay BA, Smiley JR. 2013. Mitochondrial nucleases ENDOG and EXOG participate in mitochondrial DNA depletion initiated by herpes simplex virus 1 UL12.5. J Virol 87:11787–11797. https://doi.org/10.1128/JVI.02306-13.

58. Marconi P, Manservigi R. 2014. Herpes simplex virus growth, preparation, and assay. Methods Mol Biol 1144:19–29. https://doi.org/10.1007/978-1-4939-0428-0_2.

59. Bai RK, Perng CL, Hsu CH, Wong LJ. 2004. Quantitative PCR analysis of mitochondrial DNA content in patients with mitochondrial disease. Ann N Y Acad Sci 1011:304–309. https://doi.org/10.1111/j.1749-6632.2004.00968.x.

60. Mighelini BA, Smiley JR. 2013. Mitochondrial Damage upon HSV-1 Infection. November/December 2021 Volume 12 Issue 6 e02557-21 mbio.asm.org