The mechanisms shaping CA2 pyramidal neuron action potential bursting induced by muscarinic acetylcholine receptor activation

Vincent Robert1, Ludivine Therreau1, M. Felicia Davatolhagh2, F. Javier Bernardo-Garcia3, Katie N. Clements4, Vivien Chevaleyre1, and Rebecca A. Piskorowski1

Recent studies have revealed that hippocampal area CA2 plays an important role in hippocampal network function. Disruption of this region has been implicated in neuropsychiatric disorders. It is well appreciated that cholinergic input to the hippocampus plays an important role in learning and memory. While the effect of elevated cholinergic tone has been well studied in areas CA1 and CA3, it remains unclear how changes in cholinergic tone impact synaptic transmission and the intrinsic properties of neurons in area CA2. In this study, we applied the cholinergic agonist carbachol and performed on-cell, whole-cell, and extracellular recordings in area CA2. We observed that under conditions of high cholinergic tone, CA2 pyramidal neurons depolarized and rhythmically fired bursts of action potentials. This depolarization depended on the activation of M1 and M3 cholinergic receptors. Furthermore, we examined how the intrinsic properties and action-potential firing were altered in CA2 pyramidal neurons treated with 10 µM carbachol. While this intrinsic burst firing persisted in the absence of synaptic transmission, bursts were shaped by synaptic inputs in the intact network. We found that both excitatory and inhibitory synaptic transmission were reduced upon carbachol treatment. Finally, we examined the contribution of different channels to the cholinergic-induced changes in neuronal properties. We found that a conductance from Kv7 channels partially contributed to carbachol-induced changes in resting membrane potential and membrane resistance. We also found that D-type potassium currents contributed to controlling several properties of the bursts, including firing rate and burst kinetics. Furthermore, we determined that T-type calcium channels and small conductance calcium-activated potassium channels play a role in regulating bursting activity.

Introduction

While hippocampal area CA2 was first described by Lorente de Nó >70 yr ago (de Nó, 1934), relatively little is known about the cellular properties of the neurons in this region. Recent studies using in vivo recording methods have revealed that neurons in area CA2 play potentially important roles in hippocampal network activity, including initiation of sharp wave ripples (Oliva et al., 2016) and spatial encoding during immobility (Kay et al., 2016). Lesion studies have found that synaptic transmission from area CA2 is required for social recognition memory (Hitti and Siegelbaum, 2014; Stevenson and Caldwell, 2014). Furthermore, there is mounting evidence that this area undergoes detrimental changes during schizophrenia (Benet et al., 1998; Piskorowski et al., 2016), Alzheimer’s disease (Ransmayr et al., 1989), Parkinson’s disease (Liu et al., 2019), and other neuropsychiatric and neurodegenerative diseases (Chevaleyre and Piskorowski, 2016). To understand the cellular mechanisms underlying these phenomena, the physiological properties of CA2 pyramidal neurons (PNs) require further examination. There is mounting evidence indicating that these neurons have numerous physiological properties that are markedly different from neighboring CA1 and CA3. The calcium buffering and G protein–signaling cascades in these neurons have been shown to actively prevent the expression of stereotypical postsynaptic NMDA-mediated long-term potentiation (Simons et al., 2009; Vellano et al., 2011; Zhao et al., 2007). Additionally, CA2 PNs have been shown to have a very different composition of ion channels and dendritic excitability than neighboring regions (Palacio et al., 2017; Srinivas et al., 2017; Sun et al., 2014; Dudek et al., 2016). CA2 PNs have also been shown to be modulated in the hippocampus by several neuromodulators.

1Université Paris Descartes, Inserm UMR1266, Institute of Psychiatry and Neuroscience of Paris, Team Synaptic Plasticity and Neural Networks, Paris, France; 2Department of Neuroscience, Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA; 3Department of Biochemistry and Biophysics, University of California San Francisco, San Francisco, CA; 4Department of Biology, East Carolina University, Greenville, NC.

Correspondence to Rebecca A. Piskorowski: rebecca.piskorowski@inserm.fr.
including oxytocin (Tirko et al., 2018) and vasopressin (Pagani et al., 2015).

The septo-hippocampal cholinergic projection plays a critical role in hippocampal memory formation. Acetylcholine released from septal fibers acts as both a neurotransmitter and a modulator of cellular excitability and synaptic transmission in the hippocampus, thus having wide-ranging effects on hippocampal oscillatory dynamics and synaptic plasticity. In this study, we examined how intrinsic excitability and synaptic transmission are modulated in CA2 PNs under conditions of increased muscarinic tone. We found that under conditions that approximately correspond to awake exploration in vivo (McIntyre et al., 2003; Yamamura et al., 1974), CA2 PNs depolarize and rhythmically fire bursts of action potentials (APs). Activation of M1 and M3 muscarinic receptors is required for the depolarization. We found that excitatory and inhibitory synaptic inputs contributed to the shape and timing of these AP bursts. We observed that burst firing persisted when all synaptic transmission was blocked, indicating that activation of muscarinic acetylcholine receptors altered currents that underlie this phenomenon. To investigate this further, we identified the contribution of K$_v$7 potassium channels, T-type calcium channels, D-type potassium channels and SK channels to the depolarization and bursting activity of CA2 PNs.

### Materials and methods

All procedures involving animals were performed in accordance with institutional regulations.

#### Slice preparation

Transverse hippocampal slices were prepared from 6–9-wk-old Swiss mice. Animals were anaesthetized with ketamine (100 mg/kg), xylazine (7 mg/kg), and isofluorane and perfused transcardially with an NMDG-based cutting solution containing (in mM): KCl 2.5, NaH$_2$PO$_4$ 1.25, transcardially with an NMDG-based cutting solution containing (100 mg/kg), xylazine (7 mg/kg), and isofluorane and perfused Swiss mice. Animals were anaesthetized with ketamine

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#### Electrophysiological recordings

All on-cell and whole-cell recordings of CA2 PNs were performed blind. Area CA2 has extremely dense extracellular matrix, making visually guided recordings very challenging. A recording pipette with positive pressure was inserted deeply into the pyramidal cell layer parallel to the stratum so that the tip of the recording pipette was even with the end of the mossy fibers. The pipette was then stepped deeper into the slice while a voltage step was delivered through the pipette. The current was monitored and used to determine when the pipette was close to a neuron; at that point, positive pressure was released, and a gigahm seal formed. Cell-attached recordings of CA2 PNs were obtained with ACSF in the recording pipette that had resistances between 2 and 5 MΩ. Whole-cell recordings were performed with potassium- or cesium-based intracellular solutions containing the following (in mM): K- or Cs-methyl sulfonate 135, KCl 5, EGTA-KOH 0.1, HEPES 10, NaCl 2, MgATP 5, Na$_2$GTP 0.4, Na$_2$-phosphocreatine 10, and bicytin (4 mg/ml). Series resistance was <20 MΩ and was monitored throughout the recordings. Data were discarded if the series resistance changed >10% during the experiment. For synaptic response measurements in voltage-clamp mode, no compensation was applied. In current-clamp mode, the bridge balance was measured every 20 s, compensated with internal circuitry, and monitored throughout experiments as well as before and after gap-free recordings. The liquid junction potential was not corrected for. Data were obtained using a Multiclamp 700B amplifier and digitized using a Digidata 1550 ADDA board. Data were sampled at 10 kHz.

pClamp10 software was used for data acquisition.

On-cell and whole-cell current clamp recordings of spontaneous activity were acquired in gap-free mode in sweeps of 10 min. For measurements of intrinsic properties in current clamp mode, direct current (DC) was injected and monitored as necessary to maintain an initial membrane potential (V_m) of ~70 mV. AP firing properties were measured with a series of 1-s-long depolarizing current steps.

For voltage-clamp experiments examining evoked and spontaneous synaptic transmission, cesium internal solution was used. Stimulating pipettes filled with ACSF were placed in stratum radiatum of CA1 to antidromically excite CA3-CA2 synapses and in stratum lacunosum moleculare (SLM) to stimulate distal dendritic inputs in area CA2. Synaptic currents were evoked with a constant voltage stimulating unit (Digitimer) set at 0.1 ms at a voltage range of 5–30 V. For minimal stimulation experiments, synaptic responses were evoked by a patch pipette filled with ACSF placed at the same depth as the recording pipette in close proximity to the apical dendrite of the CA2 PN. The stimulus strength (2–10 V, 0.1 ms) was adjusted to elicit small-amplitude excitatory postsynaptic currents (EPSCs; <20 pA) with >10% failure. Extracellular field potentials were recorded with patch pipettes filled with 1 M NaCl placed in CA2 stratum radiatum and SLM for detecting fEPSPs, or in the CA2 pyramidal layer for monitoring spontaneous activity. Pharmacological agents were added to the ACSF at the following concentrations (in µM): 10 carbamoylcholine chloride (CCh) to activate cholinergic receptors; 10 2,3-dihydroxy-6-nitro-7-sulfamoylbenzo [f]quinoxaline (NBQX) and 50 8-bromo-5-phosphonovalerate (APV) to block α-amino-3-hydroxy-5-methyl-4-isoxazolopyridin-1-yl] acetamide to block the M current; 2 TTA-A2 ([R]-2-[4-cyclopropylphenyl]-N-[1-[5-[2,2,2-trifluoroethoxy] pyridin-2-yl]ethyl] acetamide) to block T-type voltage-gated...
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Calcium channels; 100 4-aminopyridine (4-AP) to block voltage-activated potassium currents; 0.1 α-dendrotoxin (α-DTX) to block D-type potassium currents; 0.1 amamin to block small-conductance calcium-activated potassium (SK) channels; 10 pirenzepine to block M1 muscarinic receptors; 10 AF-DX 116 to block M2 muscarinic receptors; 11-dimethyl-4-diphenylacetoxy piperidin iodide (4-DAP) to block M3 muscarinic receptors; 10 mecamylamine to block nicotinic acetylcholine receptors (nAChRs); and 0.1 tetrodotoxin (TTX) to block voltage-activated sodium channels. Voltage clamp recordings of miniature excitatory postsynaptic currents (mEPSCs) were performed with 100 nM TTX and 4 mM CaCl2 in ACSF.

Immunocytochemistry and cell identification
Post hoc confirmation of all cells was performed by reconstruction of neuronal morphology. Following overnight incubation in 4% paraformaldehyde in PBS, slices were permeabilized with 0.2% Triton X-100 in PBS and blocked for 48 h with 3% goat serum in PBS with 0.2% Triton X-100. Alexa Fluor 464–conjugated streptavidin (Life Technologies, dilution 1:300) incubation was performed in block solution for 4 h at room temperature. Slices were mounted in longiss sodium mounting medium, resulting in a partial clarification of tissue following incubation at room temperature for 24 h. Images were collected with a Zeiss 710 laser-scanning confocal microscope.

Data analysis
Electrophysiological recordings were analyzed using custom-written macros with IGORpro (Wavemetrics) and Axograph software. APs were detected automatically by threshold crossing with IGOR TaroTools, followed by visual inspection to define AP bursts. In all whole-cell recordings, bursts were defined by clear depolarization and repolarization of the Vm flanking AP firing. For analysis of cell-attached data, bursts were defined based on the shortest bursts with lowest interburst interval seen in whole-cell recordings. This was defined as groups of ≥2 APs occurring <1 s apart from one another and separated from other groups of APs by ≥1 s. While this definition is broad, we used it to avoid constraining the analysis of cell-attached data. Kinetics of Vm variations were measured by linear fits of the Vm trace as it changed during the initial depolarization following CCh application, and from interburst-Vm plus 5 mV to burst-Vm minus 5 mV. Membrane resistance (Rm) measurements were calculated from linear fits of the voltage responses to current step injections. AP firing properties were measured on a positive current step 1.5 times over rheobase causing similar Vm depolarization for each cell and condition, typically 200 pA over rheobase. The threshold was measured at the first AP at rheobase. The AP full-width at half-maximal amplitude (AP width) and afterhyperpolarization potential (AHP) amplitude were measured for the first AP at threshold. The latency of the first AP and number of APs was measured with increasing current steps over rheobase. The instantaneous firing frequency and amplitude of the first six APs were measured at current injection steps 1.5 times over rheobase. The amplitude of the afterdepolarization (ADP) was measured 125 ms after step termination of a current 1.5 times over rheobase. Sag potentials were measured with a protocol injecting current as necessary to maintain a Vm of ~70 mV with a 1-s-long negative current step resulting in a membrane hyperpolarization of ~100 mV. The sag was calculated as the difference between the peak and steady-state voltage during the 1-s hyperpolarization.

Statistics
Results are reported as means ± SEM. Statistical significance of linear variables was assessed using paired or unpaired Student’s t test, one-sample t test, Mann–Whitney U test, Wilcoxon signed-rank test, Kruskal–Wallis test, one-way ANOVA, repeated-measures ANOVA, or Friedman ANOVA where appropriate. Normality was tested with the Jarque–Bera test, and homoscedasticity was tested using Barlett’s test to choose between parametric and nonparametric statistical analysis. The marks *, **, and *** denote P values <0.05, <0.01, and <0.001, respectively.

Results
Carbachol induces depolarization and AP bursting in CA2 PNs
Modulation of hippocampal neuronal activity by cholinergic agonists has been extensively described for PNs in areas CA3 and CA1. However, little is known about how activity in area CA2 is affected by increased muscarinic tone. To answer this question, we performed on-cell electrophysiological recordings in CA2 PNs in acute hippocampal slices from adult mice and monitored how the activity of the cells changed upon the application of 10 µM carbachol (CCh). This compound is the carbamate analogue of acetylcholine, allowing it to resist hydrolysis by acetylcholinesterase, and is a nonselective agonist for muscarinic acetylcholine receptors and a weaker nonselective agonist for nAChRs in the brain. We found that before CCh application, only 1 of 15 cells spontaneously fired APs. Following application of 10 µM CCh, 8 of 15 cells displayed regularly repeating bursts of APs (Fig. 1, A1 and A2). Within each burst, the cells fired 14.6 ± 2.2 APs at a frequency of 14.3 ± 4.2 Hz. Each burst lasted 6.4 ± 2.1 s, with an interburst interval of 90 ± 38 s.

The spontaneous rhythmic firing of bursts of APs that we observed in area CA2 are similar to previous reports of CA3 PNs following application of CCh (MacVicar and Tse, 1989; Williams and Kauer, 1997; Cobb et al., 1999; Cobb and Davies, 2005). We postulated that high muscarinic tone may be causing a change in the resting Vm in CA2 PNs, as has previously been reported in area CA3 (Dodd and Kelly, 1981; Cobb et al., 1999; Cobb and Davies, 2005) and in CA2 pyramidal cells in slice cultures (Fischer, 2004). To answer this question, we performed whole-cell current clamp recordings before and after bath-application of 10 µM CCh. Upon CCh application, we observed a depolarization of CA2 PN Vm from rest at ~75 ± 1.1 mV to ~65 ± 1.0 mV (Fig. 1, B1–B3; n = 29; paired t test, P < 0.001). CCh application caused Vm depolarization at a rate of 6.0 ± 0.7 mV/min (n = 29) before reaching a steady-state level. This depolarization was followed by rhythmic bursts of AP firing in 25 of 29 cells, with properties similar to those we observed in cell-attached recordings. We restricted the analysis of our whole-cell data to 15 min following CCh application because the on-cell data from
Carbachol application induces $V_M$ depolarization and burst firing in CA2 PNs. (A1) Sample trace of AP bursts recorded in cell-attached configuration from a putative CA2 PN following application of 10 µM CCh in acute hippocampal slice. (A2) Expanded view of a single burst of APs. (B) Sample trace of bursts recorded in whole-cell current-clamp configuration from a CA2 PN upon application of 10 µM CCh in acute hippocampal slice (B1) and expanded views of the $V_M$ initial depolarization (B2) and a burst of APs (B3). (C–G) Summary graphs of burst properties recorded in cell-attached (gray, $n = 8$) and whole-cell current-clamp (black, $n = 25$) configuration from CA2 PNs with application of 10 µM CCh (individual cells shown as dots; population averages shown as thick lines; error bars represent SEM). (C) Bursting onset (cell-attached, 10.3 ± 1.9 min; whole-cell, 6.4 ± 0.7 min; t test, $P = 0.088$). (D) Interburst interval (cell-attached, 90 ± 38 s; whole-cell, 100 ± 14.7 s; t test, $P = 0.80$). (E) Burst duration (cell-attached, 6.4 ± 2.1 s; whole-cell, 3.8 ± 0.5 s; Mann–Whitney U test, $P = 0.48$). (F) Number of APs per burst (cell-attached, 14.6 ± 2.2; whole-cell, 22 ± 2.5; Mann–Whitney U test, $P = 0.17$). (G) Firing rate during burst (cell-attached, 14.3 ± 4.2 Hz; whole-cell, 12.6 ± 1.9 Hz; Mann–Whitney U test, $P = 0.71$).
bursting cells showed that cells would burst during this time window. Furthermore, we wanted to avoid variation of cellular properties due to dialysis of the cytoplasm from whole-cell recording conditions. The fraction of bursting cells recorded in cell-attached \( n = 8/15 \) and whole-cell \( n = 25/29 \) configurations were not statistically different (chi-square test, \( P = 0.127 \)). Bursting occurred 10.3 ± 1.9 min \( ( n = 8 ) \) after CCh application in cell-attached mode and after 6.4 ± 0.7 min \( ( n = 28 ) \) in whole-cell recordings (Fig. 1 C). Bursts occurred with an interburst interval of 100 ± 14.7 s (Fig. 1 D) and lasted 3.8 ± 0.5 s (Fig. 1 E). During these bursts, CA2 PNs fired 22 ± 2.5 APs paced at 12.6 ± 1.9 Hz (Fig. 1, F–G). Importantly, none of the measurements reported above differed between cell-attached and whole-cell conditions, thus validating our approach. Following each burst of AP-firing, the \( V_M \) returned to an interburst potential that was significantly more depolarized than before CCh application of −62.3 ± 0.8 mV (paired \( t \) test, \( P < 0.001 \)).

We further studied the CCh-induced initial depolarization seen in whole-cell recordings before bursting. 10 \( \mu \)M CCh depolarized CA2 PN \( V_M \) within minutes of application (Fig. 2 A, \( n = 25 \) cells that subsequently fired bursts of APs; Fig. 2 B, \( n = 29 \) cells recorded in 10 \( \mu \)M CCh). This depolarization of the \( V_M \) following application of 10 \( \mu \)M CCh is likely acting via mAChRs, G-protein coupled metabotropic receptors that are expressed in the soma, dendrites, and axonal terminals of hippocampal PNs (Pitler and Alger, 1990; Levey et al., 1995). We found that with prior application of 10 \( \mu \)M pirenzepine, which will preferentially act on M1 receptors with partial antagonism of M3 receptors, CCh application resulted in a minor membrane depolarization of 1.6 ± 0.7 mV that was significantly lower than the 10.0 ± 0.9 mV depolarization observed in CCh alone (Fig. 2 B). Further, application of 10 \( \mu \)M AF-DX 116, an M2 receptor antagonist, had no significant effect on the CCh-induced depolarization (Fig. 2 B; 7.7 ± 1.1 mV). Application of 1 \( \mu \)M 4-DAMP, which should primarily act on M3 mAChRs but also a fraction of M1 mAChRs, also effectively prevented CCh-induced depolarization, resulting in a small depolarization (Fig. 2 B; 2.1 ± 0.5 mV). Consequently, CCh-induced AP bursting of CA2 PNs was blocked by pirenzepine and 4-DAMP but not by AF-DX 116 (Fig. 2 C). These results were consistent with the \( G_{opn} \)-coupled action of M1 and M3 receptors, which is known to inhibit potassium permeability, leading to membrane depolarization (Brown and Adams, 1980).

Next, we explored the dose-dependent effects of CCh on CA2 PNs and performed experiments with CCh concentrations ranging from 0.5 to 20 \( \mu \)M (Fig. 2, D1–D3). These experiments showed that, although application of 10 \( \mu \)M CCh robustly depolarized CA2 PNs, CCh concentrations as little as 0.5 \( \mu \)M significantly depolarized CA2 PNs (Fig. 2, E and F). However, AP bursts were observed consistently with CCh concentrations of ≥10 \( \mu \)M (Fig. 2 G). While burst firing was observed with lower concentrations, these occurred in only a fraction of experiments. Measurements of acetylcholine that have been performed in the hippocampus in vivo indicate that the concentration of acetylcholine during awake exploration is in the range of 0.5–1 \( \mu \)M (10–20 pmol acetylcholine/20 \( \mu \)l CSF; McIntyre et al., 2003). Because CCh affinity for mAChRs is 10-fold lower than that of acetylcholine (Yamamura et al., 1974), and because 10 \( \mu \)M CCh falls within this physiological range and consistently provokes bursts of AP firing, we used this concentration for the rest of the study.

To better understand how CA2 PN intrinsic properties and AP firing are altered by muscarinic tone, we performed whole-cell current clamp experiments. Current step injections before and after 10 \( \mu \)M CCh application revealed that many aspects of CA2 PN intrinsic properties and AP firing behavior were modified by CCh (Fig. 3, A1–A4). For these experiments, we took care to inject DC as necessary to maintain an initial \( V_M \) of −70 mV. First, we found that CCh application significantly increased CA2 PN \( R_M \) from 42 ± 2.8 to 90 ± 8.0 M\( \Omega \)m (Fig. 3 B). We examined several aspects of AP firing. We observed an increased AP threshold, from −47.0 ± 0.9 to −44.0 ± 0.8 mV (Fig. 3 C). Furthermore, we saw a decreased AP width, from 1.13 ± 0.04 to 1.02 ± 0.04 ms (Fig. 3 D), and an increased AHP from 6.2 ± 0.6 to 8.3 ± 0.4 mV (Fig. 3 E). In addition, the latency of first AP firing during a step was increased following CCh application, from 83 ± 11 to 156 ± 15 ms. The latency was compared at a current injection step 200 pA over rheobase (Fig. 3 F). At the same current injection, the number of APs per step also increased from 5.0 ± 0.5 to 11.9 ± 1.1 (Fig. 3 G). This increase is consistent with a decrease in AP duration and increase in AHP. CCh application also resulted in increased instantaneous AP frequency, which was measured from the first and second AP, from 15.9 ± 1.8 to 19.6 ± 2.1 Hz (Fig. 3 H). With this, we noticed that CCh prevented adaptation of AP firing frequency with increasing current injection steps. We measured a decreased AP amplitude, from 77.5 ± 1.4 to 71.1 ± 2.4 mV, following CCh application (Fig. 3 I). Following the termination of current steps eliciting AP firing, we observed an increased ADP of 7.0 ± 0.9 mV in CCh compared with 1.6 ± 0.3 mV in ACSF (Fig. 3, J and K). Finally, we observed that the sag potential measured following a hyperpolarizing step to −100 mV in CA2 PNs was increased by CCh application from 1.6 ± 0.1 mV to 4.5 ± 0.5 mV (Fig. 3, L and M). These results suggest that several conductances are modulated by CCh in CA2 PNs.

CA2 PN AP bursts are shaped by synaptic inputs

Muscarinic AChR activation has been shown to alter both synaptic transmission and ionic conductances that regulate intrinsic excitability in PNs and interneurons (Cobb and Davies, 2005; McQuiston, 2014). In area CA2, many of the properties of rhythmic burst-firing have been shown to be shaped by changes in the local inhibitory and recurrent excitatory network (Williams and Kauer, 1997; McMahon et al., 1998). For area CA2, the effect of CCh on both the local network activity and intrinsic properties has never been examined. To better understand the CCh-induced spontaneous activity in area CA2, we set out to evaluate the contribution of synaptic input to several properties of spontaneous AP bursting (Fig. 4 A). First, we assessed the contribution of excitatory transmission in CA2 PN bursting by blocking fast glutamatergic transmission with 10 \( \mu \)M NBQX and 50 \( \mu \)M APV. We saw that under these conditions, CA2 PNs continued to burst (Fig. 4, A2). We observed a significantly shorter interburst interval (Fig. 4 B and Table 1) and postulate that this is due to the significantly more depolarized \( V_M \) between bursts (Fig. 4 G and Table 1). Furthermore, with the block of
Figure 2. Depolarization of CA2 PN resting \( \text{V}_{\text{m}} \) by CCh. (A) Time course of the initial \( \text{V}_{\text{m}} \) depolarization leading to bursting onset following 10 µM CCh application \( (n = 25; \text{population average shown as black line; gray region represents SEM}) \). (B) Summary graph of \( \text{V}_{\text{m}} \) depolarization before and after application of 10 µM CCh in control conditions or with muscarinic acetylcholine receptors M1, M2, or M3 blocked by 10 µM pirenzepine, 10 µM AF-DX 116, or 1 µM 4-DAMP, respectively. The depolarization induced by CCh \( (n = 29) \) was significantly reduced by pirenzepine \( (p < 0.001, n = 12) \) and 4-DAMP \( (p < 0.001, n = 15) \) but not by AF-DX 116 \( (p = 0.185, n = 12; \text{individual cells shown as dots; population averages shown as thick lines; error bars represent SEM}) \). ***, \( p < 0.001 \). (C) Fraction of cells that burst following application of 10 µM CCh and in the presence of pirenzepine, AF-DX 116, or 4-DAMP \( (\chi^2 \text{-squared test, } p < 0.001) \). (D) Sample trace recorded in whole-cell current-clamp configuration from a CA2 PN \( (D1) \) and expanded view showing a small \( \text{V}_{\text{m}} \) depolarization without bursts during application of 1 µM CCh followed by further depolarization \( (D2) \) and AP bursts upon subsequent increased CCh concentration to 10 µM in acute hippocampal slice \( (D3) \). (E–G) Summary graphs of the initial \( \text{V}_{\text{m}} \) depolarization and subsequent bursting as a function of increasing CCh concentrations \( (0.5 \mu \text{M}, n = 9; 1 \mu \text{M}, n = 10; 2 \mu \text{M}, n = 5; 5 \mu \text{M}, n = 6; 10 \mu \text{M}, n = 29; 20 \mu \text{M}, n = 5; \text{individual cells shown as dots; population averages shown as thick lines; error bars represent SEM}) \). (E) Initial \( \text{V}_{\text{m}} \) depolarization induced by CCh \( (\text{one-sample } t \text{-tests: } 0.5 \mu \text{M}, P = 0.0079; 1 \mu \text{M}, P = 0.0022; 2 \mu \text{M}, P = 0.010; 5 \mu \text{M}, P = 0.0026; 10 \mu \text{M}, P < 0.001; 20 \mu \text{M}, P = 0.0056) \). (F) Initial \( \text{V}_{\text{m}} \) depolarization rate during CCh application. (G) Fraction of cells bursting during CCh application \( (\text{chi-square test, } P = 0.002) \).
excitatory transmission, we observed a significant decrease in the number of APs per burst (Fig. 4 D and Table 1), indicating that synaptic input from CA2 and CA3 PNs plays a significant role in the spontaneous activity. A possible consequence of blocking excitatory transmission is that less sustained AP firing during bursts would cause a decrease in activation of hyperpolarizing conductances, potentially leading to reduced repolarization of CA2 PN Vm following bursts.

Area CA2 has been shown to have a very high density of interneurons with unique morphologies and axonal projection patterns (Mercer et al., 2007, 2012; Botcher et al., 2014). To assess the contribution of the local inhibitory network to CCh-induced CA2 PN spontaneous AP bursting, we blocked all GABAergic and GABAB receptors by applying 1 µM SR95531 and 2 µM CGP55845A. We observed several notable changes following CCh application under these conditions. The burst duration was significantly shorter (Fig. 4 C and Table 1) resulting in a trend for less AP fired per burst (Fig. 4 D and Table 1), although the firing rate during a burst was significantly higher (Fig. 4 E and Table 1) than in control conditions with GABAergic transmission intact. Furthermore, the rate of depolarization preceding a burst, burst rise rate, was significantly faster (Fig. 4 H and Table 1) and the rate at which the Vm hyperpolarized following a burst, the burst decay rate, tended to be faster, although the difference was not significant (Fig. 4 I and Table 1). Our interpretation of these results is that local inhibition is acting to pace AP firing during bursts in CA2 PNs in the presence of CCh. Furthermore, the local field potential (LFP) recorded nearby in the pyramidal layer of area CA2 bore signatures of epileptiform-like events (Fig. 4 A3). These events consisted of brief (0.8 ± 0.3 s, n = 5) bursts of 3.3 ± 1.3 mV amplitude (before CCh, −47.0 ± 0.9 mV; after CCh, −44.0 ± 0.8 mV; Wilcoxon signed-rank test, P = 0.017, n = 34). (D) AP width at half-maximal amplitude (before CCh, 1.13 ± 0.04 ms; after CCh, 1.02 ± 0.04 ms; Wilcoxon signed-rank test, P = 0.005, n = 34). (E) AHP (before CCh, 6.2 ± 0.6 mV; after CCh, 8.3 ± 0.4 mV; Wilcoxon signed-rank test, P = 0.002, n = 29). (F) Latency to fire first AP per current step as a function of current injection over rheobase (200 pA over rheobase, before CCh, 15.9 ± 1.8 Hz; after CCh, 19.6 ± 2.1 Hz; Wilcoxon signed-rank test, P = 0.027, n = 31). (G) Number of APs fired per current step as a function of current injection over rheobase (200 pA over rheobase, before CCh, 5.0 ± 0.5; after CCh, 11.9 ± 1.1; Wilcoxon signed-rank test, P < 0.001, n = 34). (H) Instantaneous firing frequency of APs as a function of AP number during a current step of intensity chosen 1.5 times above rheobase (first couple of APs, before CCh, 15.9 ± 1.8 Hz; after CCh, 19.6 ± 2.1 Hz; Wilcoxon signed-rank test, P = 0.027, n = 31). (I) AP amplitude as a function of AP number during a current step of intensity chosen 1.5 times above rheobase (first AP, before CCh, 77.5 ± 1.4 mV; after CCh, 71.1 ± 2.4 mV; paired t test, P = 0.020, n = 29). (J) Sample traces of the ADP following a depolarizing current step injection in a CA2 PN recorded in whole-cell current-clamp configuration in control (gray) and 10 µM CCh (black). (K) Summary graph of CA2 PNs ADP in control (gray) and 10 µM CCh (black); before CCh, 1.6 ± 0.3 mV; after CCh, 7.0 ± 0.9 mV; paired t test, P < 0.001, n = 34; individual cells shown as thin lines; population averages shown as thick lines; error bars represent SEM). (L) Sample traces of voltage sag in response to hyperpolarizing current step injections recorded in whole-cell current-clamp configuration from a CA2 PN in control (A1, gray) and 10 µM CCh (A2, black), expanded view of the first AP in each condition (A3) and corresponding phase plane plot (A4). (B-I) Summary graphs of CA2 PNs AP firing properties in control (gray) and 10 µM CCh (black; individual cells shown as thin lines; population averages shown as thick lines, error bars represent SEM). (B) Rm (before CCh, 42 ± 2.8 MΩm; after CCh, 90 ± 8.0 MΩm; Wilcoxon signed-rank test, P < 0.001, n = 26). (C) AP threshold (before CCh, −100 ± 30 mV; after CCh, −70 ± 20 mV; Wilcoxon signed-rank test, P < 0.001, n = 34; individual cells shown as thin lines; population averages shown as thick lines; error bars represent SEM). * P < 0.05; ** P < 0.01; *** P < 0.001.

Figure 3. **Modification of CA2 PN intrinsic properties by CCh.** (A) Sample traces of AP firing in response to depolarizing current step injections recorded in whole-cell current-clamp configuration from a CA2 PN in control (A1, gray) and 10 µM CCh (A2, black), expanded view of the first AP in each condition (A3) and corresponding phase plane plot (A4). (B-I) Summary graphs of CA2 PNs AP firing properties in control (gray) and 10 µM CCh (black; individual cells shown as thin lines; population averages shown as thick lines, error bars represent SEM). (B) Rm (before CCh, 42 ± 2.8 MΩm; after CCh, 90 ± 8.0 MΩm; Wilcoxon signed-rank test, P < 0.001, n = 26). (C) AP threshold (before CCh, −47.0 ± 0.9 mV; after CCh, −44.0 ± 0.8 mV; Wilcoxon signed-rank test, P < 0.001, n = 26). (D) AP width at half-maximal amplitude (before CCh, 1.13 ± 0.04 ms; after CCh, 1.02 ± 0.04 ms; Wilcoxon signed-rank test, P = 0.005, n = 34). (E) AHP (before CCh, 6.2 ± 0.6 mV; after CCh, 8.3 ± 0.4 mV; Wilcoxon signed-rank test, P = 0.002, n = 29). (F) Latency to fire first AP per current step as a function of current injection over rheobase (200 pA over rheobase, before CCh, 15.9 ± 1.8 Hz; after CCh, 19.6 ± 2.1 Hz; Wilcoxon signed-rank test, P = 0.027, n = 31). (G) Number of APs fired per current step as a function of current injection over rheobase (200 pA over rheobase, before CCh, 5.0 ± 0.5; after CCh, 11.9 ± 1.1; Wilcoxon signed-rank test, P < 0.001, n = 34). (H) Instantaneous firing frequency of APs as a function of AP number during a current step of intensity chosen 1.5 times above rheobase (first couple of APs, before CCh, 15.9 ± 1.8 Hz; after CCh, 19.6 ± 2.1 Hz; Wilcoxon signed-rank test, P = 0.027, n = 31). (I) AP amplitude as a function of AP number during a current step of intensity chosen 1.5 times above rheobase (first AP, before CCh, 77.5 ± 1.4 mV; after CCh, 71.1 ± 2.4 mV; paired t test, P = 0.020, n = 29). (J) Sample traces of the ADP following a depolarizing current step injection in a CA2 PN recorded in whole-cell current-clamp configuration in control (gray) and 10 µM CCh (black). (K) Summary graph of CA2 PNs ADP in control (gray) and 10 µM CCh (black); before CCh, 1.6 ± 0.3 mV; after CCh, 7.0 ± 0.9 mV; paired t test, P < 0.001, n = 34; individual cells shown as thin lines; population averages shown as thick lines; error bars represent SEM). (L) Sample traces of voltage sag in response to hyperpolarizing current step injections recorded in whole-cell current-clamp configuration from a CA2 PN in control (gray) and 10 µM CCh (black). (M) Summary graph of CA2 PN voltage sag following hyperpolarization to −100 mV in control ACSF (gray) and 10 µM CCh (black; before CCh, 1.6 ± 0.1 mV; after CCh, 4.5 ± 0.5 mV; Wilcoxon signed-rank test, P < 0.001, n = 31; individual cells shown as thin lines; population averages shown as thick lines; error bars represent SEM). * P < 0.05; ** P < 0.01; *** P < 0.001.
synaptic potentials with superimposed population-spike-like signals. We detected these events in five of six CA2 PNs recorded in the presence of GABA blockers. In these PNs, epileptiform-like discharges accompanied 70 ± 8% of the AP bursts, which they tended to precede by 32 ± 12 ms. Our interpretation of this activity is that with inhibitory transmission blocked, PNs in the network undergo AP bursting that is not regulated by feed-forward and feedback inhibition. Hypersynchrony likely emerges in the network, leading to very short and sudden bursts of APs instead of paced activity.

When we blocked all excitatory and inhibitory synaptic transmission (Fig. 4, A4–I), we saw that following CCh application, the spontaneous rhythmic bursts of CA2 PNs were very similar to what we observed with NBQX and APV with only the
interburst-VM, interburst intervals and number of APs per burst different from control conditions (Fig. 4, B, D, and G; Table 1). Thus, while the CA2 PN AP bursting activity is shaped by both excitatory and inhibitory synaptic input, the intrinsic conductance of these neurons permits them to undergo rhythmic bursts of AP firing in the presence of 10 µM CCh.

CCh affects transmission onto CA2 PNs

High cholinergic tone has been shown to suppress synaptic transmission in the hippocampus in a laminar-selective way (Hasselmo and Schnell, 1994). These observations form a basis for computational models describing cholinergic modulation of hippocampal networks during learning. As this has never been explored in area CA2, and because we found that synaptic transmission shapes burst firing, we decided to examine how excitatory and inhibitory transmission is altered in this area by recording synaptic responses before and after application of 10 µM CCh.

Our first strategy was to use extracellular field recordings to examine evoked excitatory synaptic transmission in area CA2 at proximal inputs in stratum radiatum as well as distal inputs in (SLM in the presence of GABA receptor blockers (Fig. 5, A and B). We observed that application of 10 µM CCh reduced fEPSP amplitudes at both proximal (Fig. 5, A1–A3) and distal (Fig. 5, B1–B3) inputs. The reduction in fEPSP at stratum radiatum was larger than that measured in SLM, and this difference was statistically significant (P = 0.0079, two-way ANOVA, repeated measure). These results are very similar to what has been observed in areas CA1 (Hasselmo and Schnell, 1994) and CA3 (Kremin and Hasselmo, 2007).

We postulate that the application of 10 µM CCh is likely acting on CA2 PNs via activation of mAChRs. Our finding that application of the M1 and M3 receptor antagonists, pirenzepine and 4-DAMP, prevents the large membrane depolarization, and AP bursting supports this conclusion. However, in the hippocampus, nAChRs have been found to be located at both pre- and postsynaptic compartments and can modulate glutamatergic transmission (McGehee et al., 1995; Halff et al., 2014). To explore a possible contribution of nAChR participation in synaptic transmission in area CA2, we applied 10 µM of the nAChR blocker mecamylamine and measured the resulting CCh-induced decrease in synaptic transmission at stratum radiatum (Fig. 5 A3) and SLM (Fig. 5 B3) inputs. We found that blocking nAChRs had no impact on the CCh-induced reduction in synaptic transmission, indicating that these receptors are likely not contributing to the CCh-induced change in synaptic transmission as it is being measured here.

As shown in Fig. 2 B, application of M1/M3 mAChR antagonists prevented CCh-induced membrane depolarization. To determine

### Table 1. Effect of synaptic transmission on charbanchol-induced CA2 PN spontaneous activity

| Measurement                        | Control (n = 25) | Excitatory transmission blocked (n = 7) | Inhibitory transmission blocked (n = 6) | Excitatory and inhibitory transmission blocked (n = 6) | Statistics        |
|------------------------------------|-----------------|---------------------------------------|----------------------------------------|------------------------------------------------|------------------|
| ACSF-VM before CCh application (mV) | −75 ± 1.2       | −72 ± 3.1                              | −80 ± 1.9                              | −70 ± 4.1                                      | P = 0.054a       |
| CCh-VM (mV)                        | −64 ± 1.0       | −62 ± 2.1                              | −64 ± 1.5                              | −57 ± 2.9                                      | P = 0.036a       |
| ΔACSF-VM (mV)                      | 11 ± 1.0        | 9.5 ± 1.5                              | 16 ± 1.9                               | 13 ± 1.4                                       | P = 0.065a       |
| Depolarization rate (mV/min)       | 6.5 ± 0.8       | 9.2 ± 3.4                              | 9.4 ± 2.3                              | 8.6 ± 2.0                                       | P = 0.52a        |
| Burst onset (min)                  | 6.4 ± 0.7       | 4.6 ± 1.8                              | 4.7 ± 1.2                              | 3.5 ± 0.6                                       | P = 0.21a        |
| Interburst interval (s)            | 100 ± 14.7      | 36 ± 8.3b                              | 113 ± 41.9                             | 36 ± 5.6h                                      | P = 0.006a       |
| Burst-VM (mV)                      | −42 ± 1.0       | −36 ± 1.6                              | −42 ± 2.2                              | −40 ± 3.2                                       | P = 0.42a        |
| Interburst-VM (mV)                 | −62 ± 0.8       | −56 ± 1.4c                             | −63 ± 1.9                              | −56 ± 1.1l                                      | P < 0.001a       |
| Δburst-IBI (mV)                    | 21 ± 1.2        | 18 ± 2.6                                | 22 ± 3.7                               | 14 ± 2.3                                       | P = 0.11c        |
| Burst rise rate (mV/s)             | 3.5 ± 0.5       | 5.9 ± 1.1                               | 59 ± 26h                               | 3.6 ± 0.6                                       | P < 0.001f       |
| Burst decay rate (mV/s)            | −4.6 ± 0.8      | −4.3 ± 0.7                              | −13.7 ± 4.0                            | −4.8 ± 0.6                                       | P = 0.087h       |
| Burst duration (s)                 | 3.8 ± 0.5       | 2.6 ± 0.7                               | 11 ± 0.3h                              | 2.4 ± 0.6                                       | P = 0.037h       |
| Number of APs/burst                | 22 ± 2.5        | 11 ± 1.8h                               | 13 ± 2.3                               | 11 ± 2.5h                                      | P = 0.008f       |
| Firing frequency in burst (Hz)     | 13 ± 1.9        | 9.5 ± 1.4b                              | 36 ± 7.9h                              | 9.7 ± 4.5                                       | P = 0.012g       |

*10 µM CCh.
*10 µM CCh, 10 µM NBQX, and 50 µM APV.
*10 µM CCh, 1 µM SR95531, and 2 µM CGP5584A.
*10 µM CCh, 10 µM NBQX, 50 µM APV, 1 µM SR95531, and 2 µM CGP5584A.
One-way ANOVA.
P = 0.025 vs. CT; Tukey post hoc test.
*Kruskal–Wallis test.
P = 0.05 vs. CT; Dunn–Hollands–Wolfe post hoc test.
P = 0.001 vs. CT; P = 0.013 vs. SR; Tukey post hoc test.
P = 0.006 vs. CT; P = 0.033 vs. SR; Tukey post hoc test.

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CCh attenuates excitatory synaptic transmission onto CA2 PNs. (A) Diagram illustrating extracellular recording configuration with recording and stimulation electrodes in stratum radiatum. (A1) Sample traces of field responses to electrical stimulation of stratum radiatum input to area CA2 before (gray) and after (black) 10 µM CCh application in the presence of GABA receptor antagonists (1 µM SR95531 and 2 µM CGP5584A). (A2) Summary graph of the proportion of control response amplitude (Amp.) remaining in stratum radiatum after CCh with increasing stimulation intensities (n = 10). (A3) Summary graph of the proportion of fEPSP amplitude remaining with 20-V stimulation in stratum radiatum after application of 10 µM CCh alone (control, gray; n = 10, 57.2 ± 2.5%, one-sample t test, P = 2.8 × 10^−7) or in the presence of 10 µM mecamylamine, an nAChR blocker (turquoise; n = 10, 54.2 ± 3.6%, one-sample t test, P = 4.8 × 10^−7); 10 µM pirenzepine, an M1 muscarinic receptor blocker (red; n = 7, 92.6 ± 6.0%, one-sample t test, P = 0.26); 10 µM AF-DX 116, an M2 muscarinic receptor blocker (green; n = 6, 53.8 ± 3.3%, one-sample t test, P = 2.5 × 10^−5); and 10 µM 4-DAMP, an M3 muscarinic receptor blocker (blue; n = 6, 105 ± 3.7%, one-sample t test, P = 0.23). (B) Diagram illustrating extracellular recording configuration with recording and stimulation electrodes in SLM. (B1) Same as A1, except with SLM input stimulation. (B2) Summary graph of the proportion of control response amplitude remaining in SLM after CCh with increasing stimulation intensities (n = 10). (B3) Summary graph of the proportion of fEPSP amplitude remaining with 20-V stimulation in SLM after application of 10 µM CCh alone (control, gray; n = 10, 68.9 ± 2.2%, one-sample t test, P = 8.8 × 10^−7) or in the presence of 10 µM mecamylamine (turquoise; n = 10, 70.9 ± 3.8%, one-sample t test, P = 3.8 × 10^−5); 10 µM pirenzepine (red; n = 7, 97.7 ± 7.5%, one-sample t test, P = 0.766); 10 µM AF-DX 116 (green; n = 6, 63.46 ± 5.0%, one-sample t test, P = 7.9 × 10^−4); and 10 µM 4-DAMP (blue; n = 6, 98.2 ± 3%, one-sample t test, P = 0.567). (C) Diagram illustrating whole-cell recording configuration from CA2 PNs with stratum radiatum input stimulation in acute hippocampal slices. (C1) Sample traces of evoked EPSC recorded in a CA2 PN held at −70 mV with electrical stimulation of stratum radiatum input before (gray) and after (black) 10 µM CCh application in the presence of GABA receptor antagonists (1 µM SR95531 and 2 µM CGP5584A). (C2) Summary graph of the proportion of control response amplitude remaining after CCh with increasing stimulation intensities in stratum radiatum (n = 6; percentage remaining at 20-V stimulation = 57.3 ± 10.2%, one-sample t test, P = 0.008). (C3) Paired-pulse ratio at 20-V stimulation (n = 6; before CCh, 1.5 ± 0.1; after CCh, 1.8 ± 0.1; percentage increase = 19 ± 7%, paired t test, P = 0.041). (D) Same as C with SLM input stimulation. (D1) Same as C1 except with SLM input stimulation. (D2) Summary graph of the proportion of control response amplitude remaining after CCh with increasing stimulation intensities in SLM; n = 6; percentage remaining at 20-V stimulation = 49.1 ± 8.0%, one-sample t test, P = 0.031). (D3) Paired-pulse ratio at 20-V stimulation (n = 5; before CCh, 1.6 ± 0.3; after CCh, 1.8 ± 0.1; percentage increase = 13 ± 3%, paired t test, P = 0.024). * P < 0.05, ** P < 0.001.
if activation of these receptors is also a requirement for the attenuation of synaptic transmission, we performed extracellular recordings of synaptic transmission in the presence of pirenzepine (10 µM), AF-DX 116 (10 µM), or 4-DAMP (1 µM) mAChR blockers. We found that with either the M1 or M3 mAChRs blocked, CCh application had no effect on the amplitude of evoked EPSPs at synapses in both stratum radiatum and SLM (Fig. 5, A3 and B3). These data indicate that M1 and M3 receptors are primarily responsible for mediating the CCh-induced decrease in EPSP amplitude. With M2 receptors blocked with AF-DX 116, we observed that the EPSP amplitude remaining after CCh application was similar to control in both stratum radiatum and SLM.

To better examine the effect of CCh application on synaptic transmission independently from membrane depolarization effects, we performed whole-cell recordings of CA2 PNs with cesium intracellular solution. We voltage clamped the cells at −70 mV and evoked EPSCs from CA2 PNs with stimulating electrodes in stratum radiatum and SLM in the presence of...
GABA receptor blockers. Consistent with our field recordings, we observed a decrease in EPSC amplitude following CCh application in both stratum radiatum and SLM inputs (Fig. 5, C and D). However, in contrast to the field recordings, this reduction of EPSC amplitude did not significantly differ between stratum radiatum and SLM inputs ($t$ test between proportion of control response in stratum radiatum and SLM, $P = 0.47$). Thus, we conclude that CCh application decreases excitatory transmission in addition to depolarizing CA2 PNs.

To determine whether there is also a postsynaptic change in excitatory transmission, we monitored spontaneous EPSCs (sEPSCs). We found that the sEPSC frequency increased within 5 min following CCh application (Fig. 6, A–C). We hypothesize that this increase in frequency results from enhanced network activity, as connected synaptic inputs will be undergoing spontaneous bursts of AP firing. We did not observe a change in the amplitude of individual synaptic events (Fig. 6 D), consistent with the hypothesis that CCh did not act postsynaptically to reduce synaptic transmission. To further confirm the presynaptic action of CCh and decouple the network activity from synaptic transmission, we performed this experiment in the presence of 100 nM TTX and 4 mM extracellular CaCl₂, permitting us to record mEPSCs. We did not detect a change in mEPSC frequency before and after application of CCh (Fig. 6, E–G). Furthermore, we observed no change in mEPSC quantal
Figure 8. Contribution of the M-current to CCh effects on CA2 PN intrinsic properties. (A) Sample traces of AP firing in response to depolarizing current step injections recorded in whole-cell current-clamp configuration from a CA2 PN in control (A1, gray), 10 µM XE-991 (A2, purple), and 10 µM XE-991 and 10 µM CCh (A3, orange), expanded view of the first AP in each condition (A4), and corresponding phase plane plot (A5).

(B–I) Summary graphs of CA2 PNs AP firing properties in control (gray), 10 µM XE-991 (purple), and 10 µM XE-991 and 10 µM CCh (orange; n = 15; individual cells shown as thin lines; population averages shown as thick lines; error bars represent SEM).

(B) $R_m$ (before XE-991, 46 ± 2.6 MOhm; after XE-991, 53 ± 2.3 MOhm; after XE-991 and CCh, 79 ± 4.5 MOhm; Friedman ANOVA, P < 0.001).

(C) AP threshold (before XE-991, −66.3 ± 0.9 mV; after XE-991, −47.9 ± 1.4 mV; after XE-991 and CCh, −47.7 ± 1.2 mV; Friedman ANOVA, P = 0.37).

(D) AP width at half-maximal amplitude (before XE-991, 1.13 ± 0.03 ms; after XE-991, 1.06 ± 0.03 ms; after XE-991 and CCh, 1.17 ± 0.04 ms; repeated-measures ANOVA, P = 0.002).

(E) AHP (before XE-991, 6.7 ± 0.7 mV; after XE-991, 9.1 ± 0.4 mV; after XE-991 and CCh, 8.0 ± 0.5 mV, repeated-measures ANOVA, P < 0.001). (F) Latency to fire first AP per current step as a function of current injection over rheobase (200 pA over rheobase: before XE-991, 100 ± 7.6 ms; after XE-991, 132 ± 14.0 ms; after XE-991 and CCh, 147 ± 19.7 ms; Friedman ANOVA, P = 0.006).

(G) Number of APs fired per current step as a function of current injection over rheobase (200 pA over rheobase, before XE-991, 6 ± 1; after XE-991, 12 ± 2; after XE-991 and CCh, 15 ± 3; Friedman ANOVA, P = 0.27).

(H) Instantaneous firing frequency of APs as a function of AP number during a current step of intensity chosen 1.5 times above rheobase (first couple of APs frequency, before XE-991, 18 ± 1.8 Hz; after XE-991, 20 ± 2.3 Hz; after XE-991 and CCh, 20 ± 2.4 Hz; repeated-measures ANOVA, P = 0.55).

(I) AP amplitude as a function of AP number during a current step of intensity chosen 1.5 times above rheobase chosen 1.5 times above rheobase (first AP amplitude, before XE-991, 79 ± 2.0 mV; after XE-991, 84 ± 1.6 mV; after XE-991 and CCh, 78 ± 2.6 mV, repeated-measures ANOVA on first AP amplitude, P = 0.044).

(J) Sample traces of the ADP following a depolarizing current step injection in a CA2 PN recorded in whole-cell current-clamp configuration in control (gray), 10 µM XE-991 (purple), and 10 µM XE-991 and 10 µM CCh (orange).

(K) Summary graph of CA2 PNs ADP in control (gray), 10 µM XE-991 (purple), and 10 µM XE-991 and 10 µM CCh (orange; n = 12, preXE991, 2.1 ± 0.2 mV; after XE-991, 3.5 ± 0.3 mV; after XE-991 and CCh, 4.2 ± 0.5 mV; Friedman ANOVA, P = 0.001; individual cells shown as gray lines; population averages shown as thick lines; error bars represent SEM).

(L) Sample traces of voltage sag in response to hyperpolarizing current step injections recorded in whole-cell current-clamp configuration from a CA2 PN in control (gray), 10 µM XE-991 (purple), and 10 µM XE-991 and 10 µM CCh (orange).

(M) Summary graphs of CA2 PN voltage sag following hyperpolarization to −100 mV in control (gray), 10 µM XE-991 (purple), and 10 µM XE-991 and 10 µM CCh (orange; n = 12, preXE991, 2.1 ± 0.2 mV; after XE-991, 3.5 ± 0.3 mV; after XE-991 and CCh, 4.2 ± 0.5 mV; Friedman ANOVA, P = 0.001; individual cells shown as gray lines; population averages shown as thick lines; error bars represent SEM). * P < 0.05; ** P < 0.01; *** P < 0.001.
size (Fig. 6 H), consistent with a lack of postsynaptic effect of CCh. While these results support the hypothesis that CCh is acting presynaptically to reduce EPSC amplitude, the addition of TTX prevented >90% of spontaneous synaptic events, resulting in a very low mEPSC frequency. While this value is very similar to what has recently been reported by others for area CA2 (Modi et al., 2019), this low event frequency may be problematic in detecting changes in release probability. Furthermore, these spontaneous events are not input specific. Thus, we used a third method to investigate a potential presynaptic site of action of CCh, performing minimal stimulation of stratum radiatum inputs while recording CA2 PNs before and after CCh application (Fig. 6, I and J). We observed that the amplitude of synaptic events decreased when failures were included (Fig. 6 K) but was not changed when failures were excluded (Fig. 6 L). Consistently, when the amplitudes of the first and second stimulation were averaged with failures included, CCh application resulted in an increase in the paired pulse ratio (Fig. 6 M), indicating that release probability is decreased with CCh. Likewise, following CCh application, we observed an increase in failure rate (Fig. 6 N) as well as in the coefficient of variation (Fig. 6 O), consistent with a decrease in release probability.

We examined inhibitory transmission in area CA2 by using the same voltage clamp approach but held the CA2 PNs at +10 mV in the absence of GABA blockers. We observed that CCh application reduced inhibitory postsynaptic current (IPSC) amplitudes evoked by both stratum radiatum and SLM input stimulation (Fig. 7, A–C). This reduction of IPSC amplitude was largest in stratum radiatum compared with SLM (t test between proportion of control response in stratum radiatum and SLM, P = 0.028). Paired-pulse ratios of evoked IPSCs in stratum radiatum and SLM were also increased after CCh application.
Figure 10. Contribution of voltage-activated potassium currents to CCh-induced effects on CA2 PN intrinsic properties. (A) Sample traces of AP firing in response to depolarizing current step injections recorded in whole-cell current-clamp configuration from a CA2 PN in control ACSF (A1, gray), 100 nM α-DTX (A2, orange), and 100 nM α-DTX and 10 µM CCh (A3, green); expanded view of the first AP in each condition (A4); and corresponding phase plane plot (A5).

(B–I) Summary graphs of CA2 PNs AP firing properties in control (gray), 100 nM α-DTX (orange), and 100 nM α-DTX and 10 µM CCh (green; n = 12 for before and after α-DTX, n = 9 for after α-DTX and CCh; individual cells shown as thin lines; population averages shown as thick lines; error bars represent SEM).
presence of CCh. KV7 channels, which underlie the M-current, underlies the depolarization and spontaneous AP bursting in theaptic transmission, we examined which ionic conductance un-
dermination was not changed by CCh (Fig. 7 G), indicating that CCh
within 5 min (Fig. 7, D
Monitoring sIPSCs by holding CA2 PNs at a potential of +10 mV
after 20 min of bath application of XE-991. We confirmed that
reduction of inhibitory transmission by de-
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(200 pA over rheobase, before 4-AP, 4.0 ± 0.7; after 4-AP and CCh, 10.5 ± 1.6; Friedman ANOVA, P = 0.001). (Q) Instantaneous firing frequency of APs as a function of AP number during a current step of intensity chosen 1.5 times above rheobase (first couple of APs, before 4-AP, 11.3 ± 3.1Hz ; after 4-AP, 89 ± 2.4 mV; after 4-AP and CCh, 88 ± 2.2 mV; repeated-
measures ANOVA, P < 0.001). *(before –DTX, 70.9 ± 6.5 MOhm; after –DTX, 73.1 ± 6.6 MOhm; after α-DTX and CCh, 92.2 ± 9.4 MOhm; repeated-measures ANOVA, P = 0.0028). (C) AP threshold (before α-DTX, –42.9 ± 0.8 mV; after α-DTX, –44.7 ± 1.1 mV; after α-DTX and CCh, –48.5 ± 1.1 mV; repeated-measures ANOVA, P < 0.001). (D) AP width at half-maximal amplitude (before α-DTX, 1.36 ± 0.05 ms; after α-DTX, 1.34 ± 0.05 ms; after α-DTX and CCh, 1.38 ± 0.05 ms; repeated-measures ANOVA, P = 0.71). (E) AHP (before α-DTX, 7.0 ± 0.7 mV; after α-DTX, 6.9 ± 0.9 mV; after α-DTX and CCh, –4.3 ± 1.1 mV; repeated-measures ANOVA, P = 0.0023). (F) Latency to fire first AP per current step as a function of current injection over rheobase (200 pA over rheobase, before α-DTX, 180.5 ± 22.5 ms; after α-DTX, 138.1 ± 23.1 ms; after α-DTX and CCh, 91.0 ± 20.3 ms; repeated-measures ANOVA, P = 0.0016). (G) Number of APs fired per current step as a function of current injection over rheobase (200 pA over rheobase, before α-DTX, 5.5 ± 0.6; after α-DTX, 7.5 ± 1.0; after α-DTX and CCh, 15.1 ± 2.1; Friedman ANOVA, P < 0.001).

### Contribution of M-current to CCh-induced depolarization

Because CA2 PNs can fire bursts of APs in the absence of synap-
tic transmission, we examined which ionic conductance under-
lies the depolarization and spontaneous AP bursting in the presence of CCh. K\textsubscript{v7} channels, which underlie the M-current, have been modulated by acetylcholine and play a central role in controlling resting V\textsubscript{M} and repetitive and burst firing in CA3 and CA1 PNs (Cobb et al., 1999; Hönigsperger et al., 2015). We hypothesize that this current may play a similar role in CA2 PNs. To test this, we looked at the effect of blocking the M-current with 10 μM XE-991 on CA2 PN intrinsic properties (Fig. 8 A). For these experiments, care was taken to inject DC as needed to keep the initial V\textsubscript{M} at ~70 mV. Data were acquired after 20 min of bath application of XE-991. We confirmed that the drug was active by performing whole-cell recordings in area CA1 and reproducing previously published changes in EPSP summation and intrinsic properties (Hönigsperger et al., 2015). First, we examined the effects of XE-991 on CA2 PN intrinsic excitability. Consistent with previous observations that M-current channels are open at rest in CA2 PNs (Tirko et al., 2018), we observed an increase of R\textsubscript{M} in CA2 PNs following XE-
991 application, with subsequent application of CCh causing further R\textsubscript{M} increase (Fig. 8 B). Application of X-991 did not change AP threshold (Fig. 8 C) but did reduce AP width (Fig. 8 D) and increased AHP (Fig. 8 E). These effects were unchanged or attenuated by further application of CCh (Fig. 8, C–E). Application of XE-991 increased the latency of AP onset (Fig. 8 F) as well as the number of APs per step, albeit only at current injections well above threshold (Fig. 8 G), and occluded the effects of CCh application of these measurements. The milder effect of XE-991 alone compared with XE-991 plus CCh on AP number at low current injections indicates that CCh increases CA2 PN excitability by acting on additional conductances beside the M-current. Although the instantaneous firing frequency tended to be higher and to increase with AP number in XE-991 with CCh, no significant effects were seen in this dataset (Fig. 8 H). XE-991 application alone significantly increased AP amplitude, and addition of CCh reversed that effect, suggesting an M-current-independent effect of CCh in reducing spike amplitude (Fig. 8 I). The ADP following current step termination was reduced in XE-991 compared with ACSF, and subsequent application of CCh led to a further increase (Fig. 8, J and K). Application of XE-991 led to an increase in sag potential (Fig. 8 L). Our explanation of this finding is as follows. Block of the M-current results in a depolarization of the membrane, strongly supporting the conclusion that a fraction of these channels is open at resting V\textsubscript{M}. In normal ACSF, the sag current is a result of two

#### Figure 7 – B3–C3

We then gauged how inhibitory transmission was affected by CCh by recording spontaneous IPSCs (sIPSCs). Monitoring sIPSCs by holding CA2 PNs at a potential of ~10 mV showed that CCh application increased the sIPSC frequency within 5 min (Fig. 7, D–F). However, the sIPSC amplitude distribution was not changed by CCh (Fig. 7 G), indicating that CCh acts presynaptically to reduce inhibitory transmission by decreasing the probability of GABA release. Altogether, these results show a drastic influence of CCh on synaptic transmission in area CA2.
current at −100 mV, thus making the “sag” measurement larger. Further application of CCh in the presence of XE-991 resulted in similar levels of sag potential (Fig. 8 M). Altogether, these data show that the application of the Kv7 blocker XE-991 partially recapitulated and/or occluded many of the effects of CCh on CA2 PN intrinsic properties, including AHP amplitude, number of APs per current step, and sag current. This suggests a contribution of M-current closing to the effects of CCh on CA2 PN VM.

During these experiments, we also investigated the consequences of M-current block by XE-991 on CA2 PN VM by performing current-clamp recordings without injection of DC. We found that application of XE-991 led to a significant depolarization of CA2 PNs. This depolarization was insufficient to elicit AP bursting, and subsequent application of CCh brought further depolarization and AP bursting (Fig. 9 A). XE-991 has been found to be a voltage-dependent blocker of KV7.2 channels exogenously expressed in Chinese hamster ovary cells (Greene et al., 2017). Thus, we performed several experiments in which DC was injected to just below threshold to increase the probability of channel block. No difference was observed in the VM following this manipulation. When compared with CCh-induced initial VM depolarization of CA2 PNs, the XE-991 effect was smaller and did not bring VM to depolarized levels sufficient to AP bursting (Fig. 9, B–D). To confirm this, we analyzed the changes in VM before addition of XE-991, during and following the addition of CCh with XE-991 (Fig. 9, C–E). Summary graphs of burst characteristics with application of 10 µM CCh in control (black, n = 25; same CCh data shown in Fig. 12, C–E, as CCh in Fig. 4, H, I, and E, respectively), 100 nM α-DTX (green, n = 9), and 100 µM 4-AP (pink, n = 8; individual cells shown as dots; population averages shown as thick lines; error bars represent SEM). (C) Burst rise rate (Kruskal–Wallis test, P < 0.001). (D) Burst decay rate (Kruskal–Wallis test, P = 0.008). (E) Firing rate during burst (Kruskal–Wallis test, P = 0.027). *, P < 0.05; **, P < 0.01; ***, P < 0.001.

Figure 11. Voltage-gated potassium channels pace AP firing during CCh-induced bursts in CA2 PNs. (A1 and A2) Sample trace of bursts recorded in whole-cell current-clamp configuration from a CA2 PN (A1) and expanded view of a burst (A2) upon application of 10 µM CCh in acute hippocampal slice with D-type potassium channels blocked by application of 100 nM α-DTX. (A3 and A4) Sample trace of bursts (A3) and an expanded view (A4) recorded in the same configuration, except with the nonselective Kv channel blocker 4-AP applied at 100 µM with 10 µM CCh. (B1) VM levels at rest (gray) after application of 100 µM 4-AP (blue) and after further application of 10 µM CCh (pink; before 4-AP, −76.7 ± 1.6 mV; after 4-AP, −75.6 ± 1.5 mV; after 4-AP and CCh, −65.7 ± 1.5 mV; n = 10; repeated-measures ANOVA, P < 0.001). (B2) VM levels at rest (gray) and after application of α-DTX (orange) and further application of CCh (green; before α-DTX, −72.9 ± 0.8 mV; after α-DTX, −71.3 ± 1.1 mV; after α-DTX and CCh, −64.7 ± 0.7 mV; n = 10; repeated-measures ANOVA, P < 0.001).
M-current block alone is not sufficient to elicit bursts of AP firing in CA2 PNs in our experiments.

**CA2 PN AP firing and CCh-induced bursting are paced by D-type potassium currents**

D-type potassium currents have been shown to regulate AP timing and synchrony in hippocampal PNs (Storm, 1988; Cudmore et al., 2010). It may be possible that this current also participates in CCh-induced effect of AP bursting in CA2 pyramidal cells, as CCh application induces a delay in AP firing with current step injection. Furthermore, activation of M1 and M3 muscarinic receptors can result in depletion of phosphatidylinositol 4,5-bisphosphate (PIP2) in hippocampal PNs (Hackelberg et al., 2012). To examine this further, we blocked D-type potassium channels by applying 100 µM 4-AP. This blocks KV1.1, 1.2, and 1.6 voltage-gated ion channels. We then injected current steps to measure CA2 PN AP firing properties before and after the application of CCh to determine the contribution of this family of ion channels to mAChR-induced changes in intrinsic properties.

First, we assessed the role of D-type potassium currents on CA2 PN intrinsic properties and their contribution to CCh-induced changes by recording in whole-cell mode and injecting current steps before and after 100 nM α-DTX application alone followed by further application of 10 µM CCh (Fig. 10 A), similar to Fig. 8. We found that α-DTX application alone did not significantly change $R_M$, but subsequent addition of CCh led to a significant increase (Fig. 10 B). AP threshold, however, decreased following application of α-DTX and even further by subsequent CCh (Fig. 10 C). AP width was unaffected by α-DTX alone or combined with CCh (Fig. 10 D). Application of α-DTX had no effect on the AHP of the initial AP, and further application of CCh led to a significant decrease (Fig. 10 E). As a consequence of lowered threshold, AP firing occurred earlier during current step injection in the presence of α-DTX, leading to a reduced AP latency that prevented any further effect by the subsequent addition of CCh (Fig. 10 F). In addition, the number of APs per step was significantly increased by α-DTX, but subsequent addition of CCh resulted in a further increase, indicating the contribution of other channels in regulating the AP firing (Fig. 10 G). Further evidence of this was shown by the instantaneous AP firing frequency. Block of D-type channels by α-DTX resulted in a significant increase, but further application of CCh caused an even greater increase in firing frequency (Fig. 10 H). AP amplitude, however, was not altered by α-DTX alone or combined with CCh (Fig. 10 I). These data show a role for D-type potassium currents in regulating the firing pattern of CA2 PNs when $V_M$ reaches depolarized levels nearing threshold.

We also further explored the contribution of voltage-activated potassium channels by applying 100 µM 4-AP. This compound at this concentration will effectively block all D-type potassium currents as well as potentially a fraction of A-type potassium currents. Hippocampal area CA2 has an unusually dense extracellular matrix, and we recorded very deeply within the slice to have data from healthy neurons. Both of these factors

### Table 2. Effect of 4-AP and α-DTX on muscarinic-induced CA2 PN spontaneous activity

| Measurement                  | Control (n = 25) | Nonselective block of voltage-gated potassium channels (100 µM 4-AP; n = 8) | D-type potassium channels blocked (100 nM α-DTX; n = 9) | Statistics |
|------------------------------|-----------------|---------------------------------------------------------------------------|--------------------------------------------------------|------------|
| ACSF-VM (mV)                 | −75 ± 1.2       | −76 ± 1.5                                                                 | −71 ± 1.1                                              | P = 0.152* |
| CCh-VM (mV)                  | −64 ± 1.0       | −66 ± 1.5                                                                 | −65 ± 0.7                                              | P = 0.613  |
| ΔACSF,CCh-VM (mV)            | 11 ± 1.0        | 10 ± 1.5                                                                  | 7 ± 0.9                                                | P = 0.058  |
| Depolarization rate (mV/min) | 6.5 ± 0.8       | 5.4 ± 0.8                                                                 | 3.4 ± 0.6                                              | P = 0.091  |
| Burst onset (min)            | 6.4 ± 0.7       | 4.4 ± 1.0                                                                 | 6.1 ± 0.9                                              | P = 0.078* |
| Interburst interval (s)      | 100 ± 14.7      | 119 ± 26.2                                                                | 136 ± 33.0                                             | P = 0.512* |
| Burst-VM (mV)                | −42 ± 1.0       | −37 ± 3.2                                                                 | −44 ± 0.9                                              | P = 0.067  |
| Interburst-VM (mV)           | −62 ± 0.8       | −66 ± 1.5                                                                 | −64 ± 1.2                                              | P = 0.079* |
| ΔInterburst-III-VM (mV)      | 22 ± 1.3        | 29 ± 3.2*                                                                 | 20 ± 1.8                                               | P = 0.034* |
| Burst rise rate (mV/s)       | 3.5 ± 0.5       | 97.5 ± 38.0*                                                              | 65.6 ± 33.4*                                           | P < 0.001  |
| Burst decay rate (mV/s)      | −4.6 ± 0.8      | −21.6 ± 5.0*                                                              | −13.3 ± 6.5                                            | P = 0.008  |
| Burst duration (s)           | 3.8 ± 0.5       | 1.9 ± 0.7                                                                  | 6.2 ± 1.9                                              | P = 0.070  |
| Number of APs/burst          | 22 ± 2.5        | 25 ± 7.7                                                                  | 54 ± 19.9                                              | P = 0.266  |
| Firing frequency in burst (Hz)| 13 ± 1.9        | 30 ± 7.1*                                                                 | 16 ± 3.9                                               | P = 0.027  |

*One-way ANOVA.

**aP = 0.046 vs. CT; Tukey post hoc test.

**bP = 0.05 vs. CT; Dunn–Hollander–Wolfe post hoc test.**
Influence of T-type calcium channels on CA2 PN AP firing.

(A) Sample traces of AP firing in response to depolarizing current step injections recorded in whole-cell current-clamp configuration from a CA2 PN in control (A1, gray) and 2 µM TTA-A2 (A2, magenta); expanded view of the first projections recorded in whole-cell current-clamp configuration from a CA2 PN in control (A3); and corresponding phase plane plot (A4).

(B–I) Summary graphs of CA2 PNs AP firing properties in control (gray) and 2 µM TTA-A2 (magenta; n = 9; individual cells shown as thin lines; population averages shown as thick lines; error bars represent SEM). (B) Rm (before TTA-A2, 59.7 ± 9.8 MOhm; after TTA-A2, 79.8 ± 10.5 MOhm; paired t test, P = 0.028). (C) AP threshold (before TTA-A2, −43.1 ± 2.0 mV; after TTA-A2, −44.4 ± 2.0 mV; paired t test, P = 0.57). (D) AP width at half-maximal amplitude (before TTA-A2, 11.5 ± 1.2 mV; after TTA-A2, 10.5 ± 1.2 mV; paired t test, P = 0.54). (E) AHP (before TTA-A2, 11.5 ± 1.2 mV; after TTA-A2, 10.5 ± 1.2 mV; paired t test, P = 0.57). (F) Latency to first AP per current step as a function of current injection over rheobase (200 pA over rheobase: before TTA-A2, 276.8 ± 57.4 ms; after TTA-A2, 119.2 ± 33.8 ms; paired t test, P = 0.077). (G) Number of APs fired per current step as a function of current injection over rheobase (200 pA over rheobase: before TTA-A2, 4.0 ± 0.9 Hz; after TTA-A2, 3.4 ± 0.4; paired t test, P = 0.37). (H) Instantaneous firing frequency of APs as a function of AP number during a current step of intensity chosen 1.5 times above rheobase (first couple of APs, before TTA-A2, 5.6 ± 0.5 Hz; after TTA-A2, 4.0 ± 0.9 Hz; paired t test, P = 0.046). (I) AP amplitude as a function of AP number during a current step of intensity chosen 1.5 times above rheobase (first AP, before TTA-A2, 77.0 ± 3.1 mV; after TTA-A2, 85.9 ± 2.9 mV; Wilcoxon signed-rank test, P = 0.012). *, P < 0.05; **, P < 0.01.

T-type calcium currents control the duration of CA2 PN AP bursts

T-type calcium channels are activated at VM values near rest and cause transient calcium influx that depolarizes neurons. These channels are involved in setting the resting VM levels (Storm, 1988), application of α-DTX or 4-AP did not change the resting VM, while subsequent application of CCh caused an initial depolarization similar to control conditions (Fig. 11, B1 and B2). The main effect of blocking D-type potassium currents was an accelerated rise of VM at the onset of bursts seen with both α-DTX and 4-AP (Fig. 11 C and Table 2). Other effects with 4-AP potentially not restricted to D-type potassium currents were a faster burst decay rate (Fig. 4 D and Table 2) and increased firing rate during bursts (Fig. 4 E and Table 2). Other burst parameters were unchanged by α-DTX or 4-AP (Table 2). Altogether, these experiments show a key role of D-type potassium currents in controlling the early phase of burst firing induced by CCh in CA2 PNs.

Next, we assessed the contribution of voltage-activated potassium current to spontaneous burst firing by recording CA2 PNs in passive current-clamp mode and applying 100 nM α-DTX or 100 µM 4-AP followed by 10 µM CCh (Fig. 11 A). Consistent with D-type potassium channels being opened at only depolarized VM levels (Storm, 1988), application of α-DTX or 4-AP did not change the resting VM, while subsequent application of CCh caused an initial depolarization similar to control conditions (Fig. 11, B1 and B2). The main effect of blocking D-type potassium currents was an accelerated rise of VM at the onset of bursts seen with both α-DTX and 4-AP (Fig. 11 C and Table 2). Other effects with 4-AP potentially not restricted to D-type potassium currents were a faster burst decay rate (Fig. 4 D and Table 2) and increased firing rate during bursts (Fig. 4 E and Table 2). Other burst parameters were unchanged by α-DTX or 4-AP (Table 2). Altogether, these experiments show a key role of D-type potassium currents in controlling the early phase of burst firing induced by CCh in CA2 PNs.

Figure 12. Influence of T-type calcium channels on CA2 PN AP firing.

(A) Sample traces of AP firing in response to depolarizing current step injections recorded in whole-cell current-clamp configuration from a CA2 PN in control (A1, gray) and 2 µM TTA-A2 (A2, magenta); expanded view of the first projections recorded in whole-cell current-clamp configuration from a CA2 PN in control (A3); and corresponding phase plane plot (A4). (B–I) Summary graphs of CA2 PNs AP firing properties in control (gray) and 2 µM TTA-A2 (magenta; n = 9; individual cells shown as thin lines; population averages shown as thick lines; error bars represent SEM). (B) Rm (before TTA-A2, 59.7 ± 9.8 MOhm; after TTA-A2, 79.8 ± 10.5 MOhm; paired t test, P = 0.028). (C) AP threshold (before TTA-A2, −43.1 ± 2.0 mV; after TTA-A2, −44.4 ± 2.0 mV; paired t test, P = 0.57). (D) AP width at half-maximal amplitude (before TTA-A2, 11.5 ± 1.2 mV; after TTA-A2, 10.5 ± 1.2 mV; paired t test, P = 0.54). (E) AHP (before TTA-A2, 11.5 ± 1.2 mV; after TTA-A2, 10.5 ± 1.2 mV; paired t test, P = 0.57). (F) Latency to first AP per current step as a function of current injection over rheobase (200 pA over rheobase: before TTA-A2, 276.8 ± 57.4 ms; after TTA-A2, 119.2 ± 33.8 ms; paired t test, P = 0.077). (G) Number of APs fired per current step as a function of current injection over rheobase (200 pA over rheobase: before TTA-A2, 4.0 ± 0.9 Hz; after TTA-A2, 3.4 ± 0.4; paired t test, P = 0.37). (H) Instantaneous firing frequency of APs as a function of AP number during a current step of intensity chosen 1.5 times above rheobase (first couple of APs, before TTA-A2, 5.6 ± 0.5 Hz; after TTA-A2, 4.0 ± 0.9 Hz; paired t test, P = 0.046). (I) AP amplitude as a function of AP number during a current step of intensity chosen 1.5 times above rheobase (first AP, before TTA-A2, 77.0 ± 3.1 mV; after TTA-A2, 85.9 ± 2.9 mV; Wilcoxon signed-rank test, P = 0.012). *, P < 0.05; **, P < 0.01.
these channels had no effect on AP threshold (Fig. 12 C) but did result in a decrease in the AP width (Fig. 12 D), whereas there was no change in AHP (Fig. 12 E). Furthermore, we observed no change in AP latency or number of APs (Fig. 12, F and G).

After blocking T-type channels, we did observe a small but significant change in firing frequency and an increase in AP amplitude (Fig. 12, H and I). From these experiments, we can conclude that T-type calcium channels are expressed by CA2 PNs and contribute to several aspects of their intrinsic AP firing properties, including the timing of AP firing when the cell becomes depolarized.

Given these observations, we hypothesize that blocking T-type calcium channels may potentially alter the properties of CCh-induced rhythmic AP bursting. To test this, we first performed current clamp experiments in which we held the cells at −70 mV and progressively injected depolarizing steps of current in the presence of CCh alone or with application of CCh plus 2 µM TTA-A2 (Fig. 13 A). Blocking T-type channels significantly increased AP latency (Fig. 13 B) and effectively prevented the CCh-induced increase in AP firing (Fig. 13 C). Therefore, T-type calcium channels are likely involved in the modification of AP firing induced by CCh in CA2 PNs, opening with...
membrane depolarization and contributing to further depolarization and AP firing. Blocking these channels reveals this contribution. The decrease in the number of APs during the 1 s current injection step in the presence of CCh could potentially be due to the increased AP latency.

We next examined the effects of TTA-A2 on CCh-induced spontaneous AP bursts in CA2 PNs (Fig. 13 D). TTA-A2 block of T-type channels did not significantly change the resting VM level of CA2 PNs and did not affect their initial depolarization by CCh (Fig. 13 E). However, we found that application of TTA-A2 before CCh application resulted in a significant increase in the duration of AP bursts (Fig. 13 F and Table 3), while all other burst properties were unchanged (Table 3). Therefore, we conclude that T-type calcium channels are involved in controlling the duration of bursts, potentially by regulating calcium influx before and during AP burst firing that would eventually contribute to ending the burst by activating calcium-dependent potassium channels.

Small-conductance calcium-activated K⁺ channels contribute to CA2 PN VM repolarization between bursts

Sustained AP firing and VM depolarization as seen in CA2 PNs during CCh-induced AP bursting can lead to calcium influx and activation of calcium-activated potassium channels. In several types of neurons, SK channels frequently underlie a major component of the afterhyperpolarizing current following single or bursts of APs, acting to regulate burst firing (Stocker, 2004). In CA1 hippocampal pyramidal cells, these channels have been shown to be less important than Kv7 channels in regulating excitability (Chen et al., 2014). However, it has been shown that transient activation of M1 mAChRs does activate the store-release inositol triphosphate–mediated pathway that acts via SK channels to transiently control membrane excitability (Dasari et al., 2017). To examine this further, we first sought to establish the presence of SK channels in CA2 PNs and their contribution to AP firing by injecting current steps before and after application of 0.1 µM apamin (Fig. 14 A). While we found no change in Rm (Fig. 14 B), we did measure a small but significant decrease in AP threshold (Fig. 14 C), as well as a decrease in AP width (Fig. 14 D) and an increase in AHP amplitude (Fig. 14 E). This change in AHP is puzzling to us and merits further study.

Blocking SK channels had fairly minor or no effect on other aspects of CA2 PN firing (Fig. 14, F–I).

While apamin had minimal effects on CA2 PN AP firing, they are consistent with an increased RM that would be expected when blocking a potassium channel, thus warranting the study of SK channel contributions to the effects of CCh on CA2 PNs. To examine this further, we recorded pyramidal cells in area CA2 and injected depolarizing current steps in the presence of 10 µM CCh with and without 0.1 µM of apamin (Fig. 15 A). Block of SK channels by apamine resulted in increased AP latency in the presence of CCh (Fig. 15 B). Furthermore, blocking these channels appeared to prevent the CCh-induced increase in AP number with each current injection step (Fig. 15 C). The increased latency that we observed following CCh application when SK channels were blocked merits further study. Perhaps the block of SK allows increased influx of calcium that permits the activation of large-conductance calcium-activated potassium channels (BK channels), further hyperpolarizing the cells and delaying AP firing.

To determine how SK channels potentially contribute to CCh-induced spontaneous bursting of CA2 PNs, we recorded these cells in the presence of apamin followed by CCh application (Fig. 15 D). Block of SK channels by apamin alone did not

| Measurement                          | Control (n = 25) | T-type voltage-gated calcium channels blocked (2 µM TTA-A2; n = 9) | Statistics   |
|--------------------------------------|-----------------|------------------------------------------------------------------|-------------|
| ACSF-VM (mV)                         | −75 ± 1.2       | −72 ± 2.3                                                        | P = 0.28a   |
| CCh-VM (mV)                          | −64 ± 1.0       | −62 ± 1.8                                                        | P = 0.22a   |
| ΔACSF-CCh VM (mV)                    | 11 ± 1.0        | 11 ± 1.0                                                         | P = 0.89a   |
| Depolarization rate (mV/min)         | 6.5 ± 0.8       | 4.6 ± 0.9                                                        | P = 0.12a   |
| Burst onset (min)                    | 6.4 ± 0.7       | 6.0 ± 1.0                                                        | P = 0.78a   |
| Interburst interval (s)              | 100 ± 14.7      | 109 ± 18.4                                                       | P = 0.74a   |
| Burst-VM (mV)                        | −42 ± 1.0       | −42 ± 0.7                                                        | P = 0.74a   |
| Interburst-VM (mV)                   | −62 ± 0.8       | −60 ± 1.2                                                        | P = 0.17a   |
| Δburr−IBI VM (mV)                    | 21 ± 1.2        | 18 ± 0.9                                                         | P = 0.13b   |
| Burst rise rate (mV/s)               | 3.5 ± 0.5       | 2.8 ± 0.5                                                        | P = 0.69b   |
| Burst decay rate (mV/s)              | −4.6 ± 0.8      | −5.1 ± 1.1                                                       | P = 0.83b   |
| Burst duration (s)                   | 3.8 ± 0.5       | 8.9 ± 1.9                                                        | P = 0.006b  |
| Number of APs/burst                  | 22 ± 2.5        | 31 ± 6.4                                                         | P = 0.072b  |
| Firing frequency in burst (Hz)       | 13 ± 1.9        | 10 ± 1.9                                                         | P = 0.66b   |

* a t test.
* b Mann–Whitney U test.
Influence of SK channels on CA2 PN AP firing. (A) Sample traces of AP firing in response to depolarizing current step injections recorded in whole-cell current-clamp configuration from a CA2 PN in control (A1, gray) and 0.1 µM apamin (A2, blue); expanded view of the first AP in each condition (A3) and corresponding phase plane plot (A4). (B–I) Summary graphs of CA2 PNs AP firing properties in control (gray) and 0.1 µM apamin (blue; n = 10; individual cells shown as thin lines; population averages shown as thick lines; error bars represent SEM). (B) Rm (before apamin, 52.2 ± 5.0 MΩ; after apamin, 55.9 ± 3.8 MΩ; paired t test, P = 0.053). (C) AP threshold (before apamin, −41.7 ± 1.6 mV; after apamin, −44.7 ± 1.0 mV; paired t test, P = 0.010). (D) AP width at half-maximal amplitude (before apamin, 1.29 ± 0.05 ms; after apamin, 1.16 ± 0.03 ms; Wilcoxon signed-rank test, P = 0.006). (E) AHP (before apamin, 6.6 ± 1.1 mV; after apamin, 8.7 ± 0.7 mV; paired t test, P = 0.013). (F) Latency to fire first AP per current step as a function of current injection over rheobase (200 pA over rheobase, before apamin, 144.9 ± 18.5 ms; after apamin, 191.3 ± 25.6 ms; paired t test, P = 0.087). (G) Number of APs fired per current step as a function of current injection over rheobase (200 pA over rheobase, before apamin, 5.1 ± 0.9; after apamin, 5.4 ± 1.0; paired t test, P = 0.60). (H) Instantaneous firing frequency of APs as a function of AP number during a current step of intensity chosen 1.5 times above rheobase (first APs, before apamin, 13.2 ± 2.8 Hz; after apamin, 13.4 ± 2.6 Hz; paired t test, P = 0.74). (I) AP amplitude as a function of AP number during a current step of intensity chosen 1.5 times above rheobase (first AP, before apamin, 74.7 ± 3.0 mV; after apamin, 79.0 ± 2.1 mV; Wilcoxon signed-rank test on first AP amplitude, P = 0.13). * P < 0.05; **, P < 0.01.

Discussion

Using patch-clamp recordings in acute hippocampal slices from adult mice, we report several changes in hippocampal CA2 PN properties following mAChR activation. In summary, application of 10 µM CCh led to a depolarization of the VM and rhythmic bursts of AP firing. We found that the membrane depolarization would occur with 1 µM CCh, whereas the burst firing required higher concentrations. 10 µM CCh altered several intrinsic properties of CA2 PNs, including the Rm, AP threshold, AP width, AHP, latency, AP number, AP firing frequency, and sag current. While several aspects of the spontaneous AP bursting kinetics are shaped by excitatory and inhibitory synaptic transmission, CA2 PNs were found to be capable of spontaneously firing bursts of APs with all transmission blocked. Upon examination of synaptic transmission, we observed that CCh application attenuated both excitatory and inhibitory transmission. Lastly, we examined the contributions of the potassium M-current from KV7 ion channels, T-type calcium channels, D-type potassium channels, and SK channels to the intrinsic properties and AP bursting of CA2 PNs, finding that they play roles in regulating the excitability and AP firing of these cells.

The influence of the muscarinic tone on hippocampal physiology has long been recognized and extensively studied in area CA3 and CA1. Application of cholinergic agonists or stimulation of cholinergic inputs in acute hippocampal slices has been found to result in the depolarization of resting VM and an increase in Rm in CA3 and CA1 PNs (Dodd et al., 1981; Cole and Nicoll, 1983, 1984). In area CA2, we found that low concentrations of CCh resulted in VM depolarization. We also found that blocking the M1 and M3 muscarinic receptors effectively prevented depolarization and all spontaneous bursting activity. This is consistent with what has previously been described in CA3 PNs (Williams and Kauer, 1997). Furthermore, these findings are consistent with immunohistochemical evidence that M1 and M3 receptors are expressed in area CA2 (Levey et al., 1995). The pharmacological methods that we used in this paper support the premise that M1/M3 muscarinic acetylcholine receptors are expressed in CA2 PNs, as application of 1 µM 4-DAMP and 10 µM Pirenzepine prevents the large depolarization of the resting VM by CCh application and completely prevents spontaneous AP bursting. At 1 µM, a small fraction of M1 mAChRs would be expected to be blocked by 4-DAMP (Caulfield and Birdsal, 1998), and concrete discrimination between these two subtypes with our dataset cannot be done. Closer examination of RNA levels for different muscarinic receptor subtypes available on
the Allen Brain Atlas (http://mouse.brain-map.org/gene/show/12456) show that the relative levels of the M3 mAChR are lower in area CA2 compared with areas CA1 and CA3. This difference is intriguing, as different effects on PN depolarization and plasticity have previously been attributed to M3 and M1 mAChRs (Pitler and Alger, 1990; Auerbach and Segal, 1996). Furthermore, our results indicate that potential differences in downstream signaling cascades in area CA2 merit further investigation. The results demonstrate that CA2 PNs express muscarinic acetylcholine receptors that inhibit the M-current upon activation by CCh. We found that pharmacologically blocking the M-current led to a statistically significant depolarization of the $V_M$, albeit only a fraction of the depolarization observed with CCh. The extent of the depolarization that we observed following application of XE-991 is slightly lower than what had been reported in another study (3.2 ± 0.6 compared with 4.2 ± 0.5 mV; Tirko et al., 2018). In our experiments, we used a K-methanesulfonate–based solution while Tirko et al. (2018) used a K-gluconate–based intracellular solution. We hypothesize that this difference in intracellular solution may underlie the differences in cellular excitability that we observed.

Figure 15. **SK channels contribute to CA2 PN $V_M$ repolarization at the end of AP bursts.** (A) Sample traces of AP firing in response to depolarizing current step injections recorded in whole-cell current-clamp configuration from CA2 PNs in 10 µM CCh (A1, black) or 0.1 µM apamin and 10 µM CCh (A2, green). (B and C) Summary graphs of CA2 PN intrinsic properties in 10 µM CCh (CCh, black; ACSF, gray; $n = 34$; for reference, same CCh and control data as shown in Fig. 3, F and G), and 0.1 µM apamin and 10 µM CCh (green, $n = 10$; population averages shown as thick lines; error bars represent SEM). (B) Latency to fire first AP per current step by CA2 PNs as a function of current injection over rheobase (200 pA over rheobase, CCh, 156 ± 15.0 ms; CCh and apamin, 379 ± 64 ms; Mann–Whitney $U$ test, $P < 0.001$). (C) Number of APs fired per current step by CA2 PNs as a function of current injection over rheobase (200 pA over rheobase, CCh, 11.9 ± 1.1; CCh and apamin, 4.0 ± 0.7, Mann–Whitney $U$ test, $P < 0.001$). (D) Sample trace of bursts recorded in whole-cell current-clamp configuration from a CA2 PN (D1) and expanded view of a burst (D2) upon application of 10 µM CCh in acute hippocampal slice with SK channels blocked by application of 0.1 µM apamin. (E) $V_M$ levels at resting (gray), after application of 0.1 µM apamin (blue), and after further application of 10 µM CCh (green; before apamin $V_M$, −76.8 ± 2.2 mV; after apamin $V_M$, −75.1 ± 2.2 mV; after apamin and CCh $V_M$, −66.0 ± 2.3 mV, $n = 7$; repeated-measures ANOVA, $P < 0.001$). (F) Summary graph of CA2 PN $V_M$ level in between bursts with application of 10 µM CCh in control (black, $n = 25$; same CCh data shown in Fig. 14 F as CCh in Fig. 4 G) and 0.1 µM apamin (green, $n = 10$; t test, $P = 0.035$; individual cells shown as dots; population averages shown as thick lines; error bars represent SEM). *, $P < 0.05$; ***, $P < 0.001$. 

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specifically the M-current. We investigated how the M-current participates in synaptic plasticity in this area. It has been previously reported that activation of M1 mAChRs can activate a nonselective cation current, which has the potential to depolarize the membrane to allow spontaneous AP bursting. We observed that activation of M1 mAChRs resulted in an increase in depolarization and altered AP firing properties. This suggests that the M-current is an important player in regulating AP timing and network activity in CA2 PNs, similar to its role in CA1 and CA3 PNs.

As the calcium dynamics in area CA2 have previously been shown to prevent NMDA-mediated long-term potentiation (Half et al., 2014), we investigated how CCh affects AP firing in CA2 PNs. Our results indicate that in area CA2, this current can regulate AP firing and bursting, and that the role of M1 mAChRs in synaptic plasticity in area CA2 may be similar to that in CA1 and CA3.

In conclusion, our study reveals that activation of M1 mAChRs can regulate AP firing and bursting in CA2 PNs. This suggests that the role of M1 mAChRs in synaptic plasticity in area CA2 is similar to that in CA1 and CA3. Our findings highlight the importance of M1 mAChRs in regulating AP firing and bursting in CA2 PNs.
induction of persistent firing after current injection-evoked depolarization (Jochens and Yoshida, 2013; Knauer et al., 2013). Our experiments in CA2 PNs demonstrate that CCh application causes CA2 PNs to fire bursts of APs in a way that is very similar to CA3 PNs under similar conditions (Cobb et al., 1999). In this study, we demonstrated the role of T-type calcium channels in controlling the duration of these AP bursts in CA2 PNs. This finding is consistent with other cell types, as calcium currents have been known to be important for CA3 PN AP bursting (MacVicar and Tse, 1989). Furthermore, T-type calcium channels are partially opened near resting $V_M$ levels in cortical neurons (Hu and Bean, 2018), and their activation by muscarinic stimulation causes calcium entry in dentate gyrus granule cells (Martinello et al., 2015). In dopaminergic midbrain neurons, intrinsic bursting is regulated by coupled activity of T-type and SK channels (Wolfart and Roeper, 2002). In CA2 PNs, we show that SK channels are involved in the repolarization of $V_M$ following bursts. Thus, it may be possible that a similar mechanism is at play in these cells: that prolonged AP firing during bursts allows calcium influx that activates calcium-activated potassium channels, terminating the burst. Further work is required to better understand the calcium channel composition and contribution to excitability in CA2 pyramidal cells.

We did not investigate the potential role of large-conductance calcium-activated potassium channels in regulating the CCh-induced burst firing in this study. BK channels have been shown to be important for enabling high-frequency AP firing and early spike frequency adaptation in CA1 PNs (Gu et al., 2007), two properties that are nearly absent in CA2 PNs. That does not mean, however, that these channels are not contributing in a central way to the active properties of CA2 PN dendrites. In fact, given the unusual calcium buffering of CA2 PNs (Simons et al., 2009; Evans et al., 2018) and its implications for synaptic plasticity, the role of BK channels, and their possible interaction with SK channels in regulating AP firing in CA2, merits further investigation.

Of note, we saw that synaptic transmission played a role in shaping AP bursting frequency, duration, and $V_M$ between bursts. In our acute slice preparations, excitatory inputs to area CA2 originated from CA3 PNs, from recurrent connections from CA2 PNs, and from the entorhinal cortex. When these inputs are blocked, CA2 PNs fire fewer APs per burst and repolarize less after bursts. Excitatory transmission is likely important to sustain AP firing during bursts, which potentially contributes to allow calcium influx in CA2 PNs as well as promote subsequent repolarization of the $V_M$ to start a new burst. Removal of inhibitory transmission transformed bursts of APs into sudden brief AP discharges in which APs fired at higher frequency and rapidly hyperpolarized. Blocking excitatory transmission alone appeared to be indistinguishable from blocking all synaptic transmission. Clearly, feed-forward and feedback inhibition plays a central role in regulating overall excitability and AP firing in area CA2.

Interestingly, AP burst firing by CA2 PNs has recently been described to be induced by oxytocin (Tirko et al., 2018). These bursts of APs were mediated by PLC-mediated reduction of the M-current and activation of PKC. The oxytocin receptor–mediated AP bursts described by Tirko et al. (2018) lasted on the order of 2–3 s and had a firing frequency of ∼14 Hz, similar to the CCh-induced bursts that we report here. A question for future research will be to determine the downstream intracellular signaling pathways activated by M1 and M3 muscarinic receptors in CA2 PNs and how they are similar to those of other receptors. This question is very pertinent for CA2 PNs, as numerous G protein and growth factor receptors are either enriched or exclusively expressed in this region. These include the adenosine A1 receptor (Ochiishi et al., 1999), vasopressin 1b receptor (Young et al., 2006), and epidermal growth factor receptor (Tucker et al., 1993). Thus, having a better understanding of how the activity of CA2 PNs is altered in the presence and absence of high muscarinic tone is an important first step in fully understanding how this hippocampal area is affected by these signaling molecules.

The hippocampus receives a major cholinergic input from the septum (Amaral and Kurz, 1985); therefore, the hippocampal cholinergic tone is likely high when septal cholinergic neurons are active, during active exploration. It is well accepted that area CA2 plays a critical role in social memory formation (Hitti and Siegelbaum, 2014; Stevenson and Caldwell, 2014), and we predict that during social exploratory behaviors, cholinergic tone is likely to be elevated, influencing the cellular properties of CA2 PNs in a way similar to what we characterize in this paper. Major questions remain concerning how social information is encoded by the hippocampus and the precise role of area CA2 in this process. During social exploration, CA2 PNs likely receive theta-locked inputs from the entorhinal cortex, the septum, the supramammillary nucleus, and area CA3 (Robert et al., 2018). Our results indicate that changes in the $V_M$ resulting from M1 and M3 activation in high cholinergic tone suppress CA3 inputs more than cortical inputs, as has been observed in other CA regions (Kremin and Hasselmo, 2007; Hasselmo and Schnell, 1994). These diverse inputs would be summated, and the resulting CA2 output will then impact area CA1, CA3, and extrahippocampal structures. Our results will be a necessary tool for modeling and in vivo studies that consider area CA2 in the hippocampus.

Lastly, understanding the function and physiological mechanisms of CA2 PNs is important in the context of psychiatric and neurological disorders, as this region has been shown to undergo changes in human postmortem studies and animal models of schizophrenia (Benes et al., 1998; Piskorowski et al., 2016) and several other neuropsychiatric and neurodegenerative diseases (Chevaleyre and Piskorowski, 2016). Notably, the density of cholinergic input as well as acetylcholinesterase have been found to be decreased in patients with Alzheimer’s disease, most dramatically in area CA2 compared with other hippocampal areas (Ransmayr et al., 1989). Furthermore, it has recently been proposed that cholinergic dysfunction contributes to cognitive decline in Parkinson’s disease and is associated with increased Lewy pathology within the CA2 region (Liu et al., 2019).

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