3’,4’,5’,5,7-Pentamethoxyflavone Sensitizes Cisplatin-Resistant A549 Cells to Cisplatin by Inhibition of Nrf2 Pathway

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Nuclear factor erythroid 2-related factor 2 (Nrf2) is an important redox-sensitive transcription factor that regulates the expression of several cytoprotective genes. More recently, genetic analyses of human tumors have indicated that Nrf2 may cause resistance to chemotherapy. In this study, we found that the expression levels of Nrf2 and its target genes GCLC, HO-1, NQO1 were significantly higher in cisplatin-resistant A549 (A549/CDDP) cells than those in A549 cells, and this resistance was partially reversed by Nrf2 siRNA. 3’,4’,5’,5,7-Pentamethoxyflavone (PMF), a natural flavonoid extracted from Rutaceae plants, sensitized A549/CDDP to CDDP and substantially induced apoptosis compared with that of CDDP alone treated group, and this reversal effect decreased when Nrf2 was downregulated by siRNA. Mechanistically, PMF reduced Nrf2 expression leading to a reduction of Nrf2 downstream genes, and in contrast, this effect was decreased by blocking Nrf2 with siRNA. Taken together, these results demonstrated that PMF could be used as an effective adjuvant sensitizer to increase the efficacy of chemotherapeutic drugs by downregulating Nrf2 signaling pathway.

INTRODUCTION

Lung cancer has become one of the main causes of cancer-related deaths in many countries. Non-small cell lung cancers (NSCLC) account for approximately 80% of all lung cancer cases (D’Addario et al., 2009). Cisplatin is widely used in the therapy of NSCLC and has a significant antitumor activity (Langer et al., 2002). However, one of the main factors contributing to the failure of cisplatin-based chemotherapy in NSCLC is the development of drug resistance (Longley and Johnston, 2005). Thus, it is a challenge to develop novel drug sensitizer that can enhance the efficacy of cisplatin-based chemotherapy and overcome chemoresistance.

Nuclear factor erythroid-related factor 2 (Nrf2) binds to antioxidant-response elements (AREs) and regulates the expression of several genes, including heme oxygenase (HO-1), NAD(P)H dehydrogenase quinine 1 (NQO1), UDP-Glucuronosyltransferase (UGTs), cysteine ligase catalytic subunit (GCLC) and several ATP-dependent drug efflux pumps (Mrps) (Maher et al., 2008; Venugopal and Jaiswal, 1996). Therefore, Nrf2 plays an important role in protecting cells from oxidative stress and regulating cellular redox homeostasis (Jaiswal, 2004).

More recently, genetic analyses of human tumours have indicated that Nrf2-mediated defense responses may cause resistance to chemotherapy. (Hayashi et al., 2003; Maher et al., 2008; Wang et al., 2008). In human cancers such as lung and breast cancer, frequent mutations of Kelch-like ECH-associated protein 1 (Keap1), a repressor of Nrf2, have been identified and such mutations lead to constitutive activation of Nrf2 and its downstream genes (Hayes and McMahon, 2009). Highly expressed Nrf2 was found in doxorubicin-resistant MCF-7 cell line, and the chemoresistance could be reversed by blocking Nrf2 with siRNA (Shim et al., 2009). Therefore, discovery of compounds which can potently inhibit Nrf2-mediated responses is desirable and such small molecular inhibitors may be applied as an adjuvant to counteract chemoresistance.

Several natural products have been proved to be potent Nrf2 inhibitors, such as triptolide, EGCG, Brusato and procyandins (Chen et al., 2013; Kweon et al., 2006; Ohnuma et al., 2011; Ren et al., 2011). Triptolide could enhance drug sensitivity of doxorubicin-resistant leukemia cell line HL60/A and imatinib-resistant cell line KS62/G to doxorubicin and imatinib through downregulation of HIF-1α and Nrf2 (Chen et al., 2013). Cinna-momi Cortex extract reduced the expression of efflux transporters via Nrf2 suppression in A549 cells and LU199 cells and enhanced the cytotoxicity of doxorubicin and etoposide by increasing their intracellular concentrations (Ohnuma et al., 2011).

3’,4’,5’,5,7-Pentamethoxyflavone (PMF) (Fig. 1) is a polymethoxy-substituted flavonoid presented in plants of Rutaceae plant family such as Murraya paniculata and Neoraputia magnifica (Tomazela et al., 2000). Murraya paniculata is a widely used botanical drug with anti-inflammation, anti-obesity and antifungal activities (Lu et al., 2011). Neoraputia magnifica is proved to have anti-trypanosome activity (Soeiro and Castro, 2009). Pre-
Cells were seeded in 6-well plates at a density of 1 × 10⁵/well and exposed to different treatments. After incubation for 24 h, the cells were harvested and washed twice with ice-cold PBS. Afterwards the cells were incubated with Annexin V-FITC and PI successively for 15 min at room temperature in the dark according to the manufacturer’s instructions. Analyses were performed at FACSCalibur analyzer (Beckman Coulter EPICS XL, USA).

**Western blot assay**

Cytosplasmic and nuclear extracts were extracted using a Nuclear Extract kit (Active Motif, USA) according to the manufacturer’s recommendations. Equivalent amounts of protein were separated by 12% SDS-polyacrylamide gel electrophoresis and transferred to polyvinylidene difluoride membranes (Millipore Co., USA). Blocking was performed with 5% nonfat milk in TBST (TBS-1% Tween 20) for 1 h, and then incubated with primary antibodies overnight at 4°C. Subsequently, a secondary horseradish peroxidase-conjugated anti-rabbit was applied, and specific bands were visualized using ECL detection kit (Engreen Biosystem, China). The intensity of protein bands was analyzed using ImageJ software (National Institute of Health, USA).

**Results**

Nrf2-mediated defenses are upregulated in A549/DDP cells

To ensure the drug resistance phenotype of A549/DDP cells, survival rates of cells exposed to CDDP were detected by SRB assay. Figure 2A showed that the IC₅₀ of A549/DDP cells (1136.3 μM) was significantly higher than that of A549 cells (317.9 μM, P < 0.05), which indicated that A549/CDDP cells were CDDP-resistance.

Up-regulation of Nrf2 is reported to play an important role in drug resistance. To evaluate whether Nrf2-mediated signaling pathway expression was different between A549 and A549/DDP cells, western blot assay was performed. The expression of total and nuclear Nrf2 protein in A549 cells was significantly lower than those in A549/DDP cells as well as its downstream target genes HO-1, NQO1 and GCLC (Figs. 2B-2C).

**PMF sensitized A549/CDDP cells to CDDP**

To determine whether PMF can enhance the sensitivity of A549/DDP cells to CDDP, SRB assay was carried out and IC₅₀ was calculated. As shown in Fig. 3A, IC₅₀ of PMF+CDDP treated group was significantly lower than that of the CDDP alone treated group, indicating that PMF reduced the resistance of A549/CDDP cells to CDDP. The effect of PMF on the cytotoxicity of CDDP was also investigated in A549 cells, which expressed lower Nrf2 than A549/DDP cells. The results showed that the effect of PMF was diminished in A549 cells compared to A549/DDP cells.

**Materials and methods**

**Materials**

3',4',5',5,7-Pentamethoxyflavone (PMF) was purchased from BioSun Sci & Tech Co., Ltd (China). Cis-diamminedichloroplatinum (II) (cisplatin, CDDP) was bought from Tokyo Chemical Industry (Japan) and kiton Red S (SRB) was purchased from Alfa Aesar (China). Primary antibody of Nrf2 was purchased from Santa Cruz Biotechnology (USA). NQO1, HO-1, ERK, p-ERK, PARP1 primary antibodies were got from Sangon Biotech (China). The A549 cell line and its CDDP-resistant cell line A549/CDDP were purchased from cell resource center of Institute of Basic Medical Sciences of Chinese Academy of Medical Sciences. Cells were cultured in Dulbecco's modified Eagle's medium (DMEM, HyClone, USA) supplemented with 10% fetal bovine serum (FBS, HyClone, USA) and antibiotics (50 μg/mL of penicillin and 50 μg/mL streptomycin). The cells were grown in a humidified incubator with 5% CO₂ at 37°C.

**Cell culture**

The A549 cell line and its CDDP-resistant cell line A549/CDDP cells were purchased from cell resource center of Institute of Basic Medical Sciences of Chinese Academy of Medical Sciences. Cells were cultured in Dulbecco’s modified Eagle’s medium (DMEM, HyClone, USA) supplemented with 10% fetal bovine serum (FBS, HyClone, USA) and antibiotics (50 μg/mL of penicillin and 50 μg/mL streptomycin). The cells were grown in a humidified incubator with 5% CO₂ at 37°C.

**Cytotoxicity assay**

Exponentially growing A549 or A549/CDDP cells were seeded in 96-well plates at a density of 5 × 10⁴ cells/well. Cells and siRNA-transfected cells were treated with the compounds for 48 h. Cell viability was assessed using SRB assay following the manufacturer’s protocol. Data were averaged from at least three independent experiments and are expressed as means ± SD.

**Flow cytometric analysis of apoptotic cells**

Cells were seeded in 6-well plates at a density of 1 × 10⁶/well and exposed to different treatments. After incubation for 24 h, the cells were harvested and washed twice with ice-cold PBS. Afterwards the cells were incubated with Annexin V-FITC and PI successively for 15 min at room temperature in the dark according to the manufacturer’s instructions. Analyses were performed at FACSCalibur analyzer (Beckman Coulter EPICS XL, USA).

**Transient transfection with Nrf2 siRNA or plasmids**

The siRNA against human Nrf2 or negative control was synthesized by RiboBio (China) and the sequence of Nrf2 siRNA is 5'-GAGAAAGAAGAUUGCCUGUAAUdTdT-3' and 3'-dTdTCTCUCUUGCUUUCUUGAAGAAAGUUGCCUGUAA-5'. The plasmid pGL3-ARE was a kind gift from Dr. Athanassios Frangoulis (University Hospital Aachen) (Boesch-Saadatmandi et al., 2009). Cells were transfected with siRNA or plasmids using Lipofectamine 2000 (Invitrogen) according to the product specification. A dual-luciferase reporter assay (Promega, USA) was used to determine ARE-driven promoter activity.
with that in A549/CDDP cells, indicating that PMF might sensitize A549/CDDP cells to cisplatin through inhibition of Nrf2. To investigate whether combination of PMF and CDDP would promote apoptosis of cells, A549/CDDP cells were exposed to CDDP (25 μM) or PMF (25 μM) or in combination for 24 h. Apoptotic cells were determined by PI/Annexin V staining and flow cytometric analysis. Figures 3C-3D showed that the ratio of apoptotic cells in the combined treatment group was significantly increased compared with that in the CDDP alone treated group (P < 0.05). Furthermore, apoptosis-related proteins were assayed by Western blotting. We found that the cleaved PARP1 and caspase3 levels induced by CDDP were significantly enhanced by PMF treatment (Fig. 3E). Taken together, these results illustrated that PMF enhanced the sensitivity of A549/CDDP cells to CDDP and promoted apoptosis.

**PMF inhibited Nrf2 signaling pathway**

To investigate whether the expression of Nrf2 and its target genes was regulated by PMF, A549/CDDP cells were incubated with PMF (10, 25 and 50 μM) for 24 h. Western blot analysis revealed that PMF treatment reduced the protein levels of Nrf2 and its downstream target genes, including HO-1, NQO1 and GCLC in a dose-dependent manner(Figs. 4B-4C).

Activated Nrf2 binds to ARE sites and causes the up-regulation of its target genes. To further confirm the effect of PMF on Nrf2 transactivation, HEK293T cells were transiently transfected with ARE plasmid constructs. Treatment with PMF (10, 25 and 50 μM) produced a significant decrease in ARE transcriptional activity (Fig. 4A).

Keap1 is the major repressor of Nrf2 and facilitates the degradation of Nrf2 through the ubiquitin–proteasome pathway in cytosol (Itoh et al., 2010). The effect of PMF on the expression of Keap1 was also investigated. The expression of Keap1 was significantly increased by PMF in a dose dependent manner (Fig. 4D), which suggested that the inhibition of PMF on Nrf2 signaling pathway might be through Keap1.

In addition, Nrf2 activity has been implicated to be under the regulation of a number of protein kinases (Kim and Jang, 2014). To further explore the mechanisms underlying the inhibitory effect of PMF on Nrf2 activity, the phosphorylated Nrf2 and phosphatase ERK were investigated. The p-Nrf2 level and p-ERK were significantly decrease by PMF (Fig. 5).

**PMF sensitized A549/CDDP cells to CDDP by inhibition of Nrf2**

To confirm whether Nrf2 was involved in the chemoresistance of A549/CDDP cells, Nrf2 siRNA was transfected to knock down Nrf2 expression in A549/CDDP cells. The sensitivity of cells to CDDP was detected by SRB assay. Figure 6A showed that Nrf2 siRNA-transfected A549/CDDP cells were much more sensitive to CDDP than A549/CDDP cells.

To clarify whether Nrf2 signaling pathway was involved in PMF increasing sensitivity of A549/CDDP cells to CDDP, A549/CDDP cells were transfected with Nrf2 siRNA to knock down Nrf2 expression (Fig. 6B-6D). Western blot analysis confirmed the successful knockdown of Nrf2 in A549/CDDP cells. The expression of Nrf2-target genes including HO-1, NQO1 and GCLC were also decreased in siRNA-transfected A549/CDDP cells. Furthermore, PMF did not significantly affect levels of Nrf2 and its target genes in A549/CDDP cells transfected with Nrf2 siRNA.

To investigate whether the sensitization by PMF was Nrf2 dependent, the sensitivity of A549/CDDP cells to CDDP was tested with Nrf2 knockdown. The effect of PMF was diminished in Nrf2 siRNA-transfected A549/CDDP cells (Fig. 6E). These results illustrated that PMF enhanced the sensitivity of A549/CDDP cells to CDDP and promoted apoptosis.
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Fig. 4. PMF inhibited Nrf2 signalling pathway. (A) HEK 293T cells were transfected with PGL3-ARE and pRL-TK plasmids. Twenty four hours after the transfection, the cells were treated with PMF (10, 25 and 50 \( \mu \)M) or tBHQ (10 \( \mu \)M) for 24 h. (B-D) A549/CDDP cells were treated with PMF (0-50 \( \mu \)M) for 24 hours. After treatment, nuclear fractions and total proteins were analyzed by western blot assay. Data are presented as means ± SD of three independent experiments and significant differences are indicated as *P < 0.05.

Further demonstrated that PMF increased the sensitivity of A549/CDDP cells to CDDP by inhibition of Nrf2.

DISCUSSION

Drug resistance during chemotherapy is considered as one of the major obstacle to the successful treatment of many cancers (El-Sheikh et al., 2013; Signore et al., 2013). Several studies have reported that Nrf2 played an important role in chemoresistance. (El-Sheikh et al., 2013; Maher et al., 2008; Na and Surh, 2014). The expression of Nrf2 and its target genes were significantly higher in tumor tissues than in their corresponding non-tumor tissues (Gao et al., 2013b). The proliferation of several cell lines was inhibited and became sensitive to anticancer drugs when Nrf2 expression was knocked down with siRNA (Ji et al., 2013; Singh et al., 2008). Both in vivo and in vitro studies demonstrated that Nrf2 was a potential target to increase drug sensitivity. In general, these findings indicated that up-regulation of Nrf2 and its target genes would increase resistance to the cytotoxic effects of chemotherapy drug.

In this study, the IC50 of A549/CDDP cells to CDDP was found to be higher than that of A549 cells (Fig. 1A), which suggested that A549/CDDP cells were CDDP-resistant. And higher levels of Nrf2 and Nrf2-mediated antioxidant enzymes were found in A549/CDDP cells than A549 cells. Transfection of A549/CDDP cells with Nrf2 siRNA decreased the Nrf2-mediated pathway, resulting in the partial recovery of sensitivity to CDDP in A549/CDDP cells (Fig. 6A). It is reported that HO-1 could potentiate tumor aggressiveness, by increasing tumor growth, angiogenesis and metastasis (Was et al., 2006). NQO1 could metabolically activate carcinogenic heterocyclic amines which present in smoke and higher levels of NQO1 expression have been detected in the lung tissues (Kiyohara et al., 2005). And GCLC is a subunit of a key enzyme in glutathione synthesis which was reported to enhance CDDP-resistance in NSCLC xenografts (Fujimori et al., 2004). Thus, we deduced that the up-regulation of Nrf2 and Nrf2-mediated antioxidant enzymes was one of the reasons that lead to resistance to CDDP in A549 cells, which was consistent with previous studies (Tang et al., 2011). Our findings suggested that the activation of Nrf2-mediated signaling is responsible for, at least in part, CDDP resistance in A549 cells. In addition to these antioxidant enzymes, several transporters such as Mrps were also reported to be regulated by Nrf2, which could lead to chemoresistance. We will further investigate Nrf2-mediated transporters in the future work.

Several natural products have been reported to be inhibitors of Nrf2 and can reduce drug resistance (Gao et al., 2013b; Kweon et al., 2006; Tang et al., 2011; Zhong et al., 2013). For example, apigenin was reported to sensitize doxorubicin-resistance BEL-7402/ADM cells to doxorubicin by inhibiting Nrf2 pathway (Gao et al., 2013b; Kweon et al., 2006; Tang et al., 2011; Zhong et al., 2013). For example, apigenin was reported to sensitize doxorubicin-resistance BEL-7402/ADM cells to doxorubicin by inhibiting Nrf2 pathway (Gao et al., 2013b; Kweon et al., 2006; Tang et al., 2011; Zhong et al., 2013). Luteolin was discovered to be a potent small-molecule inhibitor, which downregulated cytotoxic enzymes and sensitized NSCLC cells and colorectal cancer cell Lines to oxalipatin, bleomycin and doxorubicin (Chian et al., 2014; Tang et al., 2011). Wogonin was shown to suppress Nrf2 activity and reduce HO-1 and NQO1 expression in MCF-7/DOX cells, reversing their drug resistance (Zhong et al., 2013).

PMF is a natural polymethoxy-substituted flavonoid with anti-cancer potential (Kinoshita and Firman, 1997). In this study, we identified PMF as a potent Nrf2 inhibitor. PMF sensitized A549/
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