Clinical and molecular characterization of 12 prenatal cases of Cri-du-chat syndrome

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Abstract

Background: This study aimed to define the molecular basis for 12 prenatal cases of Cri-du-chat syndrome (CdCS) and the potential genotyping-phenotyping association.

Methods: Karyotyping and single nucleotide polymorphism array analyses for copy number variants were performed.

Results: Nine cases had 5p terminal deletions and three had 5p interstitial deletions, and these cases had variable deletion sizes with partial overlapping. Phenotypically, besides intrauterine growth restriction (IUGR) and brain as well as heart abnormalities, hypospadias, and lung dysplasia were observed. Potential genetic causes for specific phenotypes in these cases were identified.

Conclusion: This study defined the molecular bases for the patients of CdCS, which is important for genetic counseling for these families. The findings of present study expand the clinical features of CdCS in the fetal period, and provided important information for further refining the genotypic-phenotypic correlations for this syndrome.

KEYWORDS
5p deletion, Cri-du-chat syndrome, prenatal diagnosis, single nucleotide polymorphism array
1 | INTRODUCTION

Disorder resulting from a deletion of the short (p) arm of chromosome 5 is termed Cri-du-chat syndrome (CdCS) (OMIM#123450) or 5p deletion syndrome. CdCS is one of the most common chromosomal defect syndromes. It is estimated that CdCS occurs in 1 in 15,000–50,000 live birth infants (Niebuhr, 1978). Clinically, CdCS is typically characterized by high-pitched cry (cat’s cry), microcephaly, low birth weight, and hypotonia in infancy (Kondoh et al., 2005; Mainardi et al., 2001). Other features, including intellectual disability, distinctive facial features, congenital heart defect (CHD), and behavioral problems have also been noticed in affected individuals (Elmakky et al., 2014; Nguyen et al., 2015).

Recent studies have associated some critical regions of 5p with the clinical features of this disorder including haploinsufficiency of 5p15.3 for cat-like cry and speech delay, and haploinsufficiency of 5p15.2 for facial dysmorphism, microcephaly, and severe intellectual disability (Correa, Feltes, & Riegel, 2019; Zhang et al., 2005). Moreover, haploinsufficient deletions of some candidate genes in 5p have been associated with the specific clinical features of this disorder (Kondoh et al., 2005; Nguyen et al., 2015). For instance, haploinsufficiency of catenin delta 1 (CTNND2; OMIM *604275) and semaphorin 5A (SEMA5A; OMIM *609297) in 5p have linked to severe intellectual disability in individual affected with CdCs (Correa et al., 2019; Medina, Marinescu, Overhauser, & Kosik, 2000; Nguyen et al., 2015). Despite the progress in the clinical and molecular delineation of CdCs, the genotyping-phenotyping association of CdCs remains largely unclear.

In this study, we defined the molecular bases for 12 prenatal cases of CdCS, and further identified that haploinsufficiency of iroquois homebox 4 (IRX4; OMIM *606199), NADH-ubiquinone oxidoreductase subunits 6 (NDUFS6; OMIM*603848), steroid 5 alpha-reductase 1 (SRD5A1; OMIM*184753), and/or ADAM metallopeptidase with thrombospondin type 1 motif 16 (ADAMTS16; OMIM*607510) are associated with some specific phenotypes in CdCs.

2 | MATERIALS AND METHODS

2.1 | Ethical compliance

The study was approved by the ethics committee of Hunan Provincial Maternity and Infant Care Hospital.

2.2 | Subjects

Twelve prenatal cases of CdCS that were collected in Prenatal Diagnosis Central of Hunan Provincial Maternity and Infant Care Hospital were included in this study. A brief introduction of the conditions of the cases and their parents are presented in Table 1. The reasons for being referred to our prenatal diagnosis central including ultrasound abnormalities in six cases, high risk of fetal aneuploidy identified by maternal serum screening test or noninvasive prenatal testing (NIPT) in four cases, positive family history for genetic diseases in one case, and the history of miscarriages in one case. Following written informed consents, chorionic villus (CVS), amniocentesis cells, and cord blood were collected for molecular analysis for the fetuses, and peripheral blood was collected from parents to determine inheritance patterns of any deletions identified.

2.3 | Karyotyping and single nucleotide polymorphism array analyses

Karyotyping via routine chromosome G-banded (320–400 bands) analyses were performed in cells in metaphase according to standard protocols (Peng et al., 2015). For single nucleotide polymorphism (SNP) array analysis, the genomic DNA was extracted from the CVS, amino fluid cells, cord blood, and peripheral blood lymphocytes by using DNA Isolation Kit for Cells and Tissues and QIAamp DNA Blood Mini Kit (QIAGEN), respectively. SNP array analysis was conducted using Affymetrix CytoScan®750 K Array (Affymetrix Inc) according to the manufacturer's instruction. Data from SNP array analysis was analyzed by using Chromosome Analysis Suite (ChAS; version 2.1). All genomic coordinates were taken from the human reference sequence. Genes and Online Mendelian Inheritance in Man (OMIM) references were from RefSeq and OMIM entries, respectively (Liu et al., 2016).

3 | RESULT

3.1 | Clinical findings

Fetal ultrasound revealed that seven cases (case 1–5, 7, and 10) had developmental anomalies, and two cases (fetus 9, 11) showed no apparent abnormality. Results of fetal ultrasound for case 6, 8, and 12 were not found. Among those with abnormal ultrasound findings, three cases (1, 3, and 5) showed retarded embryo growth or intrauterine growth restriction (IUGR), five cases (2, 4, 5, 7, and 10) displayed cerebellar hypoplasia, and two cases (4 and 7) exhibited CHD. In addition, fetus 3 also had lung tissue dysplasia, and both fetus 7 and 10 showed hypospadias. The detail information regarding the clinical findings is listed in Table 2 and Figure 1.

Although fetus 11 appeared normal at 22 weeks of gestation by ultrasound testing, medical examination demonstrated
that the father who carried the same 5p deletion had slight dysmorphic faces and mild intellectual disability. In addition, according to family history information, the first pregnancy of the same parents of fetus 11 was aborted because of the findings of CHD and diaphragmatic hernia at 7-months gestation, and the second pregnancy was miscarried in the early stage.

### 3.2 Results of karyotyping and SNP array analyses

Karyotyping by G-banding were performed for 11 fetuses (2–12), and revealed that 9 of them except for fetus 9 and 11 showed 5p deletions. In addition, in accompany with the 5p deletion, duplication of 14q31.3-q32.33 and 8p23.3-p12 were detected in fetus 7 and fetus 8, respectively. Fetus 7 and 8 inherited the 5p deletion from the unaffected mother who underwent a balanced chromosome translocation between chromosome 5 and other chromosome. The karyotyping results are listed in Table 2.

Single nucleotide polymorphism array analysis identified 5p deletions in all 12 fetuses, and mapped the breakpoints for the deletion fragments. Nine cases had terminal deletions starting at 113,576 base pairs (bp) (position of the first probe from the used BeadChip in 5p) with distinct end breakpoints between 2,035,54 bp and 39,068,326 bp, and three cases had interstitial deletions. The smallest deletion encompassed 1.9 Mb and the largest spanned 39.0 Mb. The breakpoints for the duplicated fragment in fetus 7 and 8 were also mapped. SNP array analysis for the parents of the fetuses revealed that the 5p deletions in seven fetuses (fetus 1–5, 10, 12) were de novo, and in another five fetuses were inherited. The results of SNP array analysis are listed in Table 2.

### 3.3 Haploinsufficiency of OMIM genes

Recent studies have linked the haploinsufficiency of tubulin polymerization promoting protein (TPPP; OMIM*608773), telomerase reverse transcriptase (TERT; OMIM*187270), IRX4, NDUF56, ADAMTS16, CTNND2, NOP2/Sun RNA methyltransferase 2 (NSUN2; OMIM*610916), SEMA5A, membrane associated ring-CH-type finger 6 (MARCH6; OMIM*613297), dynein axonemal heavy chain 5 (DNAH5; OMIM*603335), and natriuretic peptide receptor C (NPR3; OMIM*108962) among others to some specific feature of CdCS (Correa et al., 2019; Nelson, Jin, Downs, Kamp, & Lyons, 2014; Tan et al., 2016).

Based on the information of the breakpoints for each deletion and the results of in silico analysis by using the UCSC Genome Browser Database and OMIM, OMIM genes in each deletion fragments were analyzed. The OMIM genes deleted in these fetuses are indicated in Figure 2.
| Case | Ultrasound findings                                                                                     | Karyotype                          | Genomic coordinates (hg19) start–end                                      | size (Mb) | Type | Parental Origin | Note                                                                 |
|------|--------------------------------------------------------------------------------------------------------|------------------------------------|---------------------------------------------------------------------------|-----------|------|-----------------|----------------------------------------------------------------------|
| 1    | Embryo stop growing                                                                                  | Not available                      | 5p15.33p13.1 (113,576–39,068,326)                                         | 39.0      | del  | De novo         |                                                                     |
| 2    | Dysgenesis of the cerebellar, widened posterior fossa pool, and abnormality of lateral ventricle      | 46, XX, del (5) (p13)              | 5p15.33p13.3 (113,576–32,733,570)                                         | 32.6      | del  | De novo         |                                                                     |
| 3    | Intrauterine growth restriction, lung tissue dysplasia                                               | 46, XY, del (5) (p13)              | 5p15.33p13.3 (113,576–29,739,642)                                         | 29.6      | del  | De novo         |                                                                     |
| 4    | Ventricular septal defect, cerebellar hypoplasia                                                     | 46, XX, del (5) (p14)              | 5p15.33p14.3 (113,576–19,255,769)                                         | 19.1      | del  | De novo         |                                                                     |
| 5    | Intrauterine growth restriction, left cerebellar hemispheric dysplasia, and widened posterior fossa pool | 46, XX, del (5) (p15)              | 5p15.33p15.2 (113,576–14,738,108)                                         | 14.6      | del  | De novo         |                                                                     |
| 6    | Unavailable                                                                                           | 46, XY, del (5) (p15)              | 5p15.33p15.2 (113,576–11,321,779)                                         | 11.2      | del  | mat            | The mother had severe mental retardation                             |
| 7    | enlarged lateral ventricle, absence of inferior vena cava, neural tube defects, and hypospadias      | 46, XY, der(5) t(5;14)(p15.2; p32)mat | 5p15.33p15.2 (113,576–10,459,497) (14q31.3q32.33) (85,163,601–107,284,437) | 10.3      | del  | mat            | The mother was normal                                                |
| 8    | Unavailable                                                                                           | 46, XX, der (5)t(5;8)(p15.3; p12)mat | 5p15.33 (113,576–3,923,103)                                               | 3.8       | del  | mat            | The mother was normal                                                |
| 9    | Normal                                                                                               | 46XX                               | 5p15.33 (113,576–2,035,548)                                               | 1.9       | del  | mat            | The mother was normal                                                |
| 10   | Enlargement of the lateral ventricle, hypospadias                                                    | 46, XY, del (5) (p13)              | 5p15.32p13.3 (4,538,650–32,005,542)                                       | 27.5      | del  | De novo         |                                                                     |
| 11   | Normal                                                                                               | 46, XX                             | 5p15.32–p15.2 (5,262,861–10,532,502)                                       | 5.2       | del  | pat            | The father had mild intellectual disability                          |
| 12   | Unavailable                                                                                           | 46, XY, del (5) (p14)              | 5p15.31p14.2 (7,116,243–24,560,212)                                       | 17.4      | del  | De novo         |                                                                     |

Abbreviations: Mb, megabase pair; SNP, single-nucleotide polymorphism.
Cri-du-chat syndrome was first identified in three patients by Lejeune in 1963 (Lejeune et al., 1963). Followed studies defined the typical clinical features of CdCS in adolescent and adult patients, including high-pitched cry, microcephaly, speech delay, and intellectual disability, which are present with variable frequency (Overhauser et al., 1994; Zhang et al., 2005). To date, less than 50 prenatal cases of CdCS had been reported (Mak et al., 2019; Su et al., 2019). Furthermore, specific symptoms of CdCS during fetus period remain largely unknown. From July 2015 to December 2019, 12 cases of CdCS were identified in 28,564 pregnant women that were referred to our center to perform invasive prenatal diagnosis because of abnormal findings from fetal ultrasound testing and/or maternal serum screening, resulting in an incidence of 0.042% (12/28,564), which was similar to that

**FIGURE 1** Images of ultrasound test on fetuses. A. Images of ultrasound test on fetus 2 at 30 weeks’ gestation. (a) Ultrasonographic image showing measurement of transverse cerebellar diameter (TCD). The TCD of this fetus was 2.92 cm. The normal value at 30 weeks’ gestation is 3.86 ± 0.34. The value of posterior fossa pool is 0.93 cm that is higher than the normal range (<0.80 cm). (b) Ultrasonographic image showing measurement of the left lateral ventricle. The highest value is 1.06 cm that is higher than the normal range (<0.80 cm). B. Images of ultrasound test on fetus 3 at 36 weeks’ gestation. (a) Ultrasonographic image showing that the lung tissue was scarce, suggesting lung dysplasia. (b) Ultrasonographic image showing measurement of biparietal diameter (BPD) and head circumference (HC). The value of BPD (7.6 cm) and HC (27.70 cm) represent 30 weeks’ gestation, suggesting an intrauterine growth retardation of fetus 3. C. Images of ultrasound test on fetus 4 at 24 weeks’ gestation. (a) Ultrasonographic image showing measurement of TCD. The TCD of fetus 4 was 2.06 cm. Normal value at 24 weeks’ gestation is 2.85 ± 0.17 cm. (b) Ultrasonographic image showing transventricular septal blood flow signal, suggesting ventricular septal defect. The ventricular septal defect sized 0.3 cm. D. Images of ultrasound test on fetus 5 at 35 weeks’ gestation. (a) Ultrasonographic image showing the TCD of this fetus was 4.00 cm. The normal value at 35 weeks’ gestation is 4.29 ± 0.26. The value of posterior fossa pool was 1.55 cm that is higher than the normal range (<0.80 cm). (b) Ultrasonographic image showing measurement of left and right cerebellar hemisphere, suggesting left cerebellar hemispheric dysplasia. E. Images of ultrasound test on fetus 7 at 26 weeks’ gestation. (a) Ultrasonographic image showing measurement of the left lateral ventricle. (b) Ultrasonographic image showing absence of inferior vena cava. (c) Ultrasonographic image showing neural tube defects. (d) Ultrasonographic image showing hypospadias. F. Images of ultrasound test on fetus 10 at 32 weeks’ gestation. (a) Ultrasonographic image showing measurement of the left lateral ventricle. The result of measurement was 1.28–1.35 cm. (b) Ultrasound image showing the coronal view of the external genitalia, suggesting hypospadias.

4 | DISCUSSION

Cri-du-chat syndrome was first identified in three patients by Lejeune in 1963 (Lejeune et al., 1963). Followed studies defined the typical clinical features of CdCS in adolescent and adult patients, including high-pitched cry, microcephaly, speech delay, and intellectual disability, which are present with variable frequency (Overhauser et al., 1994; Zhang et al., 2005). To date, less than 50 prenatal cases of CdCS had been reported (Mak et al., 2019; Su et al., 2019). Furthermore, specific symptoms of CdCS during fetus period remain largely unknown. From July 2015 to December 2019, 12 cases of CdCS were identified in 28,564 pregnant women that were referred to our center to perform invasive prenatal diagnosis because of abnormal findings from fetal ultrasound testing and/or maternal serum screening, resulting in an incidence of 0.042% (12/28,564), which was similar to that
reported by Su et al. (2019). Among them, nine cases had terminal deletions and three had interstitial deletions at 5p. The deletions in seven cases were de novo, while was inherited in other cases. The ratio of de novo 5p deletion in present study (58.3%) is lower than the previously reported (around 80%) (Mainardi et al., 2001). Phenotypically, among the nine cases with prenatal ultrasound results, seven cases had developmental anomalies, and two fetuses appeared normal. Among those with abnormal ultrasound findings, three had retarded embryo growth or IUGR, five had cerebellar hypoplasia, and two showed CHD. Hypospadias were noticed in two cases, and lung tissue dysplasia was detected in one case.

Abnormal brain development and function are one typical feature of CdCS patients in postnatal (Honjo et al., 2018; Zhang et al., 2005). In 36 prenatal cases of CdCS previously reported, 16 showed cerebral abnormalities such as cerebellar hypoplasia and abnormality of the cerebral ventricles (Mak et al., 2019; Su et al., 2019). In the present study, five fetuses (2, 4, 5, 7, and 10) showed abnormal brain development (Figure 1A,C–F). Notably, besides the 5p deletion, case 7 also contained a 22.1 Mb duplication of 14q31.3-q32.33. In patients with 14q distal trisomy, phenotypes including growth retardation, hypertelorism, facial dysmorphia, and cardiovascular anomalies have been reported (Villa et al., 2016), but not neural tube defect and hypospadias. Therefore, neural tube defect and hypospadias in case 7 was most likely from the 5p deletion. Although fetus 6 and 11 showed no brain anomaly by fetal ultrasound, the parent who carried the same chromosomal abnormalities showed intellectual disability, suggesting haploinsufficiency of the two regions were potentially associated with brain development. The absence of brain anomaly in fetus 6 and 11 may arise from incomplete penetration. Of note, the products of TPPP, NSUN2, and SEMA5A that localize in 5p13.3 have been demonstrated to play major roles in brain development (Blanco et al., 2014; Lehotzky et al., 2010; Mosca-Boidron et al., 2016; Sardina, Walters, Singh, Owen, & Kimonis, 2014). CTNND2 that localize in 5p13.2 have also been linked with brain development (Hofmeister et al., 2015; van Rootselaar et al., 2017). Moreover, fetuses 2, 4, 5, 6, 7, 10, and 11 contained an overlapping deletion region of 5p15.33-p15.2 (position: 5,626,861–10,495,497) that contained NSUN2 and SEMA5A (Figure 2). Together, these finding suggest that haploinsufficiency of 5p15.33-p15.2 contributes critically to the abnormal brain development and mental retardation in patients of CdCS.

CHD has been reported in 15%–20% of patients of CdCS (Hills, Moller, Finkelstein, Lohr, & Schimmenti, 2006; Nagy et al., 2019; Zhu et al., 2016). The cardiac anomaly in CdCS includes patent ductus arteriosus, septal defects, Tetralogy of Fallot (TOF), and other structural malformations (Hills et al., 2006). In present study, fetus 4 that contained a deletion of 5p15.33-p14.3 (position: 113,576–19,255,769) displayed ventricular septal defect (Figure 1C). Fetus 7 that contained a deletion of 5p15.33p15.2 (position: 113,576–10,459,497) showed the absence of inferior vena cava (IVC) (Figure 1E). These two fetuses shared an overlapping region from position 113,576 to 10,459,497. Proteins encoded by IRX4, NDUFS6, and DNAH5 that

**FIGURE 2** A comparison of the deleted regions and OMIM genes among the 5p deletion cases. OMIM, Online Mendelian Inheritance in Man.
localized in this regions have been demonstrated to be critical for cardiovascular development (Nelson et al., 2016; Nothe-Menchen et al., 2019; Rouzier et al., 2019). The deletion region in fetus 4 contained these three region, and the deletion region in fetus 7 contained IRX4 and NDUF56, suggesting that haploinsufficiency of IRX4 and/or NDUF56 lead to CHD.

Hypospadias were rarely reported in patients of CdCS. Chen et al. reported a prenatal case with distal deletion involving 5p15.1→pter that displayed cerebellar hypoplasia, hypospadias, and facial dysmorphisms (Chen et al., 2013). In the present study, two fetuses (7 and 10) showed hypospadias (Figure 1E,F). Fetus 7 contained a deletion of 5p15.33-p15.2 (position: 113,576–10,459,497), and fetus 10 carried a deletion of 5p15.32-p13.3 (position: 4,538,650–32,005,542). Among the OMIM genes in their overlapping deleted region, haploinsufficiency of SRD5A1 has been implicated as a candidate genetic reason for hypospadias. Notably, another two studies provided evidence that mutations of SRD5A2, instead of SRD5A1, were present in some boys with isolated hypospadias (Sun, Zhou, & Liu, 2019; Yuan et al., 2017). SRD5A1 and SRD5A2 encode isoform 1 and 2 of 5α-reductase that catalyzes the conversion of testosterone into the more potent androgen, dihydrotestosterone, and their protein products shared 50% sequence identity, but whether hemizygous deficiency each of them have similar effect in the development of hypospadias awaits future functional analysis.

In the present study, we report a case of CdCS showing lung dysplasia of fetus 3 (Figure 1B). The fetus 3 contained a deletion of 5p15.33-p13.3 (position: 113,576–29,739,642), which contained two OMIM genes DNAH5 and ADAMTS16 were the deletion region. Li et al. demonstrated that the protein production of DNAH5 played an important role in the development of lung (Li et al., 2016). Emerging evidence also suggested that ADAMTS16 expressed at high levels in fetal lung, but its involvement in lung development remain unknown (Surridge et al., 2009).

5p deletions with no apparent phenotype have been reported (Nguyen et al., 2015). Similar 5p deletions were also reported in individuals with/without clinical phenotypes (Nguyen et al., 2015). In this study, fetus 11 containing a deletion of 5p15.32-p15.2 (position: 5,262,861−10,532,502) showed no developmental abnormality by ultrasound testing. As such, the parents decided to continue the pregnancy, and a girl was born at the 39 weeks’ gestation. Physical examination showed the newborn had birth weight of 3,200 g (75th–90th), head circumference of 31.5 cm (10th–25th), and length of 45 cm (~50th). An Apgar score was 9 and 10 at the 1st and 5th minute after birth. Postoperative follow-up, feeding, and physical examination showed that the child was normal. In addition, results of development assessment, including cognitive aspects, language, mobility, and dexterity were all normal. In addition, fetus 9 that inherited a 1.9 Mb terminal 5p deletion from a healthy mother showed no developmental abnormality by ultrasound testing at the gestation of 23+ weeks. The reasons for the absence of clinical phenotypes in case 11 and 9 remain largely unknown, but in completed penetration may be a potential reason.

5p deletions mostly arise de novo, and 80%–90% of which are paternal in origin potentially due to breakage of chromosome 5 during gamete formation in males (Campbell et al., 2014; Eyal et al., 2019). Around 15% of 5p deletions are from parental translocation between chromosome 5 and others. Less common mechanisms including mosaicism, inversions, or ring chromosomes have also been implicated (Cerruti Mainardi, 2006). The risk of recurrence can be negligible for the cases of a de novo 5p deletion, but attention should be paid to the possibility of gonadal mosaicism in the parents which increases the risk of recurrence (Cerruti Mainardi, 2006). The risk of producing a CdCS offspring in the families with translocation involving 5p deletion ranged from 8.7% to 18.8% (Cerruti Mainardi, 2006). In the present study, the 5p deletions in case 7 and 8 were from the mother with balanced translocation involved in 5p deletion. Thus, genetic counseling is critically important for their families.

In summary, this study clarified the genetic diagnosis for 12 prenatal cases of CdCS from different families, which was important for guidance of the future pregnancy. In addition, this study provides more material for the clinical manifestation of CdCS in the fetal period, which is helpful for further analysis of the genotype-phenotype correlation in CdCS. This study also provided evidence supporting the critical regions and candidate genes whose haploinsufficiency is causative for clinical features of CdCS.

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CONFLICT OF INTEREST
All authors declare that they have no commercial or other conflicting interests.

AUTHOR CONTRIBUTION
Y Peng and H Wang designed the study. Y Peng and C Tang prepared the manuscript. J Pang and N Ma performed the SNP array analysis. Y Peng, J Liu, and S Yang interpreted the data of SNP array analysis. J Hu, H Xi, and Z Jia analyzed the karyotype study. X Huang performed the prenatal ultrasound testing.

STATEMENT OF ETHICS
Written informed consent was obtained prior to investigation. The authors have no ethical conflicts to disclose.
DATA AVAILABILITY STATEMENT
The data that support the findings of this study are available from the corresponding author upon reasonable request.

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REFERENCES
Blanco, S., Dietmann, S., Flores, I. V., Hussain, S., Kutter, C., Humphreys, P., … Frye, M. (2014). Aberrant methylation of tRNAs links cellular stress to neuro-developmental disorders. EMBO Journal, 33(18), 2020–2039. https://doi.org/10.15252/embj.201489282

Campbell, I. M., Stewart, J. R., James, R. A., Lupski, J. R., Stankiewicz, P., Olofsson, P., & Shaw, C. A. (2014). Parent of origin, mosaicism, and recurrence risk: Probabilistic modeling explains the broken symmetry of transmission genetics. American Journal of Human Genetics, 95(4), 345–359. https://doi.org/10.1016/j.ajhg.2014.08.010

Cerruti Mainardi, P. (2006). Cri du Chat syndrome. Orphanet Journal of Rare Diseases, 1, 33. https://doi.org/10.1186/1750-1172-1-33

Chen, C.-P., Huang, M.-C., Chen, Y.-Y., Chern, S.-R., Wu, P.-S., Su, Ying Peng, from the corresponding author upon reasonable request.

Correa, T., Feltes, B. C., & Riegel, M. (2019). Integrated analysis of the critical region 5p15.3-p15.2 associated with cri-du-chat syndrome. Genetics and Molecular Biology, 42(1 suppl 1), 186–196. https://doi.org/10.1590/1678-4685-GMB-2018-0173

Eyal, O., Berkenstadt, M., Reznik-Wolf, H., Poran, H., Ziv-Baran, T., Greenbaum, L., … Pras, E. (2019). Prenatal diagnosis for de novo mutations: Experience from a tertiary center over a 10-year period. Molecular Genetics & Genomic Medicine, 7(4), e00573. https://doi.org/10.1016/j.mgg3.573

Hills, C., Moller, J. H., Finkelstein, M., Lohr, J., & Schimmenti, L. A. (2006). Cri du chat syndrome and congenital heart disease: A review of previously reported cases and presentation of an additional 21 cases from the Pediatric Cardiac Care Consortium. Pediatrics, 117(5), e924–e927. https://doi.org/10.1542/peds.2005-1012

Hofmeister, W., Nilsson, D., Topa, A., Anderlid, B. M., Darki, F., Matsson, H., … Lindstrand, A. (2015). CTNND2-a candidate gene for reading problems and mild intellectual disability. Journal of Medical Genetics, 52(2), 111–122. https://doi.org/10.1136/jmg2014-102757

Honjo, R. S., Mello, C. B., Pimenta, L. S. E., Nufes-Vaca, E. C., Benedetto, L. M., Khoury, R. B. F., … Kim, C. A. (2018). Cri du Chat syndrome: Characteristics of 73 Brazilian patients. Journal of Intellectual Disability Research, 62(6), 467–473. https://doi.org/10.1111/jir.12476

Kondoh, T., Shimokawa, O., Harada, N., Doi, T., Yun, C., Gohda, Y., … Moriiuchi, H. (2005). Genotype-phenotype correlation of 5p-syndrome: Pitfall of diagnosis. Journal of Human Genetics, 50(1), 26–29. https://doi.org/10.1007/s10038-004-0213-9

Lehotzky, A., Lau, P., Tokesi, N., Muja, N., Hudson, L. D., & Ovadi, J. (2010). Tubulin polymerization-promoting protein (TPPP/p25) is critical for oligodendrocyte differentiation. Glia, 58(2), 157–168. https://doi.org/10.1002/glia.20909

Lejeune, J., Lafourcade, J., Berger, R., Vialatte, J., Boeswillwald, M., Seringe, P., & Turpin, R. (1963). 3 Cases of Partial Deletion of the Short Arm of a 5 Chromosome. Comptes Rendus Hebdomadaires Des Seances De L’academie Des Sciences, 257, 3098–3102.

Li, F., Fang, Z., Zhang, J., Li, C., Liu, H., Xia, J., … Ji, H. (2016). Identification of TRA2B-DNAH5 fusion as a novel oncogenic driver in human lung squamous cell carcinoma. Cell Research, 26(10), 1149–1164. https://doi.org/10.1038/cr.2016.111

Liu, J., Hu, H., Ma, N., Jia, Z., Zhou, Y., Hu, J., & Wang, H. (2016). A de novo duplication of chromosome 9q34.13-qter in a fetus with Tetralogy of Fallot Syndrome. Molecular Cytogenetics, 9, 54. https://doi.org/10.1186/s13039-016-0267-3

Mainardi, P. C., Perfumo, C., Cali, A., Coucourde, G., Pastore, G., Cavani, S., … Bricarelli, F. D. (2001). Clinical and molecular characterisation of 80 patients with 5p deletion: Genotype-phenotype correlation. Journal of Medical Genetics, 38(3), 151–158. https://doi.org/10.1136/jmg.38.3.151

Mak, A. S. L., Ma, T. W. L., Chan, K. Y. K., Kan, A. S. Y., Tang, M. H. Y., & Leung, K. Y. (2019). Prenatal diagnosis of 5p deletion syndrome: Report of five cases. Journal of Obstetrics and Gynaecology Research, 45(4), 923–926. https://doi.org/10.1111/jog.13911

Medina, M., Marincescu, R. C., Overhauser, J., & Kosik, K. S. (2000). Hemizygosity of delta-catenin (CTNND2) is associated with severe mental retardation in cri-du-chat syndrome. Genomics, 63(2), 157–164. https://doi.org/10.1006/geno.1999.6090

Mosca-Boidron, A.-L., Gueneau, L., Huguet, G., Goldenberg, A., Henry, C., Gigot, N., … Bourgeron, T. (2016). A de novo microdeletion of SEMA5A in a boy with autism spectrum disorder and intellectual disability. European Journal of Human Genetics, 24(6), 838–843. https://doi.org/10.1038/ejhg.2015.211

Nagy, O., Szaksonsz, K., Biró, B. O., Mogyorósy, G., Nagy, D., Nagy, B., … Ujfalusi, A. (2019). Copy number variants detection by microarray and multiplex ligation-dependent probe amplification in congenital heart diseases. Journal of Biotechnology, 299, 86–95. https://doi.org/10.1016/j.jbiotec.2019.04.025

Nelson, D. O., Jin, D. X., Downs, K. M., Kamp, T. J., & Lyons, G. E. (2014). Irx4 identifies a chamber-specific cell population that contributes to ventricular myocardium development. Developmental Dynamics, 243(3), 381–392. https://doi.org/10.1002/dvdy.24078

Nelson, D. O., Lalit, P. A., Biermann, M., Markandeya, Y. S., Capes, D. L., Addesso, L., … Lyons, G. E. (2016). Irx4 marks a multipotent, ventricular-specific progenitor cell. Stem Cells, 34(12), 2875–2888. https://doi.org/10.1002/stem.2486

Nguyen, J. M., Qualmann, K. J., Okashah, R., Reilly, A., Alexeyev, M. F., & Campbell, D. J. (2015). 5p deletions: Current knowledge and future directions. American Journal of Medical Genetics Part C: Seminars in Medical Genetics, 169(3), 224–238. https://doi.org/10.1002/ajmg.c.31444

Niebuhr, E. (1978). The Cri du Chat syndrome: Epidemiology, cytogentic, and clinical features. Human Genetics, 44(3), 227–275. https://doi.org/10.1007/bf00394291

Nöthe-Menchen T., Wallmeier J., Pennekamp P., Höben I. M., Olbrich H., Loges N. T., … Omran H. (2019). Randomization of left-right asymmetry and congenital heart defects: The role of DNAH5 in humans and mice. Circulation: Genomic and Precision Medicine, 12(11), 513–525. https://doi.org/10.1161/CIRCGEN.119.002686
Overhauser, J., Huang, X., Gersh, M., Wilson, W., McMahon, J., Bengtsson, U., … Wasmuth, J. J. (1994). Molecular and phenotypic mapping of the short arm of chromosome 5: Sublocalization of the critical region for the cri-du-chat syndrome. Human Molecular Genetics, 3(2), 247–252. https://doi.org/10.1093/hmg/3.2.247

Peng, Y., Ma, R., Zhou, Y., Xia, Y., Wen, J., Zhang, Y., … Wu, L. (2015). De Novo ring chromosome 11 and non-reciprocal translocation of 11p15.3-pter to 21qter in a patient with congenital heart disease. Molecular Cytogenetics, 8, 88. https://doi.org/10.1186/s13039-015-0191-y

Rouzier, C., Chaussenot, A., Fragaki, K., Serre, V., Ait-El-Mkadem, S., Richelme, C., … Bannwarth, S. (2019). NDUFS6 related Leigh syndrome: A case report and review of the literature. Journal of Human Genetics, 64(7), 637–645. https://doi.org/10.1038/s10038-019-0594-4

Sardina, J. M., Walters, A. R., Singh, K. E., Owen, R. X., & Kimonis, V. E. (2014). Amelioration of the typical cognitive phenotype in a patient with the 5pter deletion associated with Cri-du-chat syndrome in addition to a partial duplication of CTNND2. American Journal of Medical Genetics. Part A, 164A(7), 1761–1764. https://doi.org/10.1002/ajmg.a.36494

Su, J., Fu, H., Xie, B., Lu, W., Li, W., Wei, Y., … Qin, Z. (2019). Prenatal diagnosis of cri-du-chat syndrome by SNP array: Report of twelve cases and review of the literature. Molecular Cytogenetics, 12, 49. https://doi.org/10.1186/s13039-019-0462-0

Sun, L., Zhou, M., & Liu, T. (2019). Association between SRD5A2 polymorphism and hypospadias: A meta-analysis. Die Pharmazie, 74(2), 125–128. https://doi.org/10.1691/ph.2019.8768

Surridge, A. K., Rodgers, U. R., Swingler, T. E., Davidson, R. K., Kevorkian, L., Norton, R., … Clark, I. M. (2009). Characterization and regulation of ADAMTS-16. Matrix Biology, 28(7), 416–424. https://doi.org/10.1016/j.matbio.2009.07.001

Tan, S. C., Gomes, R. S. M., Yeoh, K. K., Perbellini, F., Malandraki-Miller, S., Ambrose, L., … Carr, C. A. (2016). Preconditioning of Cardiosphere-Derived Cells With Hypoxia or Prolyl-4-Hydroxylase Inhibitors Increases Stemness and Decreases Reliance on Oxidative Metabolism. Cell Transplantation, 25(1), 35–53. https://doi.org/10.3727/096368915X687697

van Rootseelaar, A. F., Groffen, A. J., de Vries, B., Callenbach, P. M. C., Santen, G. W. E., Koelewijn, S., … van den Maagdenberg, A. (2017). delta-Catenin (CTNND2) missense mutation in familial cortical myoclonic tremor and epilepsy. Neurology, 89(23), 2341–2350. https://doi.org/10.1212/WNL.0000000000004709

Villa, N., Scatigna, A., Redaelli, S., Conconi, D., Cianci, P., Farina, C., … Selicorni, A. (2016). 14q32.3-qter trisomic segment: A case report and literature review. Molecular Cytogenetics, 9, 60. https://doi.org/10.1186/s13039-016-0265-5

Yuan, S., Meng, L., Zhang, Y., Tu, C., Du, J., Li, W., … Tan, Y.-Q. (2017). Genotype-phenotype correlation and identification of two novel SRD5A2 mutations in 33 Chinese patients with hypospadias. Steroids, 125, 61–66. https://doi.org/10.1016/j.steroids.2017.06.010

Zhang, X., Smijders, A., Segraves, R., Zhang, X., Niebuhr, A., Albertson, D., … Pinkel, D. (2005). High-resolution mapping of genotype-phenotype relationships in cri du chat syndrome using array comparative genomic hybridization. American Journal of Human Genetics, 76(2), 312–326. https://doi.org/10.1086/427762

Zhu, X., Li, J., Ru, T., Wang, Y., Xu, Y., Yang, Y., … Hu, Y. (2016). Identification of copy number variations associated with congenital heart disease by chromosomal microarray analysis and next-generation sequencing. Prenatal Diagnosis, 36(4), 321–327. https://doi.org/10.1002/pd.4782

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