A Kinetic Study of Acetylcholinesterase Inhibition by Fractions of *Oleo diox Roxb.* leaf and *Curculigo orchioides* Gaertn. Rhizome for the Treatment of Alzheimer’s Disease

G. K. Pratap and Manjula Shantaram

1Department of Studies and Research in Biochemistry, Jnana Kaveri Post Graduate Centre, Mangalore University, Chikka Aluvara, 571 232, Kodagu, Karnataka, India.

Authors’ contributions

This work was carried out in collaboration between both authors. Author PGK has designed the study, performed the statistical analysis, wrote the protocol and wrote the first draft of the manuscript. Author MS conceived the study and were in charge of overall direction, planning and manuscript corrections. Both authors read and approved the final manuscript.

ABSTRACT

Introduction and Aim: Acetylcholinesterase (AChE) has an important role in the biochemical studies indicating that AChE accelerates the formation of amyloid-β fibril and forms AChE-Aβ complexes in the brain. The medicinal plants of *Curculigo orchioides* Gaertn., and *Oleo diox Roxb.* collected from Western Ghat region of Kodagu, Karnataka, India have many medicinal values such as antioxidant, anti-cancer, anti-diabetic, neuroprotective activity and anti-inflammatory property. Aim of the study was to check whether *C. orchioides* Gaertn. and *Oleo diox Roxb.* fractions significantly inhibit acetylcholinesterase in different concentration-dependent manner (1-3 mg/mL).

Results: Plant fraction concentrations resulted in >60% AChE inhibition as compared with the standard Galantamine. The IC50 value was 0.15 mg/mL which was calculated from the equation of the percentage inhibition curve for the test or plant fraction. The Lineweaver-Burk plot indicated that
the fraction followed competitive and non-competitive inhibition kinetics. Finally, we conclude that the type of plot exhibited by the C. orchioides Gaertn. and Oleo dioxRoxb. fractions has a possible therapeutic application for improving memory and other cognitive functions. The medicinal plant-based drugs are known to have limitations due to their fewer side effects and problems associated with bioavailability, which necessitates the interest in finding better AChE inhibitors from plant sources.

**Keywords:** Acetylcholinesterase; amyloid-β fibril; C. Orchioides Gaertn; Oleo diox Roxb fraction; Lineweaver-Burk plot.

1. INTRODUCTION

Alzheimer’s disease (AD) is a progressive neurodegenerative disease distressing the brain. AD is the most frequent cause of dementia in an elderly population. Symptoms of AD are loss of memory, understanding, and speech, such as anxiety and dysphoria, unpleasant behavioral changes [1]. It has provided the intellectual structure for therapeutic intervention. It proposes that the development of deposition of the β-amyloid protein is the initial pathological event in AD, important to the creation of senile plaques and then to neurofibrillary tangles, neuronal cell death, and finally causes dementia. AD is globally recognized as the most common form of dementia. It will be increasing and by the year 2050, 115 million people will be affected throughout the world [2].

Acetylcholinesterase enzyme (AChE) is participating in cholinergic neurotransmission and stimulation of cholinergic receptors of acetylcholine released into the neuronal synaptic cleft by inhibiting acetylcholine hydrolysis by AChE through the use of acetylcholinesterase inhibitors [3]. Acetylcholinesterase enzyme is membrane-bound. The main role of acetylcholinesterase is the termination of nerve impulse transmission at the cholinergic synapses by fast hydrolysis of the neurotransmitter acetylcholine [4]. The AChEIs promote a boost in the concentration and duration of the act of synaptic acetylcholine [5].

Acetylcholinesterase has a significant role in the biochemical studies indicated that AChE accelerates the formation of amyloid-β fibril and form AChE-Aβ complexes in the brain. The neurotoxicity accelerates by AChE-Aβ complexes indicated that they prompt more neurodegeneration than those of the Aβ peptide [6].

Acetylcholinesterase activity is inhibited by many synthetic and natural compounds. There are two main types of AChEinhibitors which can be distinguished from drugs and toxins [5]. The inhibitors are different compounds with different structural motives as they can bind to the etherification part of the active site by etherification of serine hydroxyl and the aromatic gorge and the peripheral anionic site [7].

Alzheimer’s disease therapy is moderate and mainly based on acetylcholinesterase inhibitors (AChEIs) such as synthetic galantamine and donepezil. The plant-based drugs are known to have limitations due to their fewer side effects and problems associated with bioavailability, which necessitates the interest in finding better AChEIs from natural resources [8-14].

2. MATERIALS AND METHODS

2.1 Chemicals

Acetylthiocholine iodide (ATCI), Acetylcholinesterase (AChE) from electric eel, 5,5'-dithiobis [2-nitrobenzoicacid] (DTNB), Galantamine, purchased from Sigma. Ethanol and all other organic solvents (analytical grade) were purchased from Merck.

2.2 Collection of Sample

Rhizomes of C. orchioides Gaertn., and leaves of Oleo diox Roxb. were collected from the Western Ghat region of Kodagu, Karnataka, India.

2.3 Sample Preparation

Freshly collected rhizomes and leaves of the plants were washed 2-3 times with tap water and then with distilled water, followed by ethanol and then allowed to dry at room temperature for 23 days and finally mechanically powdered. 50 g of powder was Soxhlet extracted with ethanol and methanol, dried at room temperature and collected the sample in brown bottles until further analysis. Extracts of both plants were
reconstituted in ethanol and methanol and then subjected to column chromatography. Active fraction (C. orchioides Gaertn. F3 Fraction and oleo diox Roxb., F2 fraction) have exhibited positive results.

2.4 Estimation of the IC\textsubscript{50} Value

The different concentrations of the fraction inhibited 50\% of AChE activity (IC\textsubscript{50}). The estimation of IC\textsubscript{50} was carried out by the method of Kamal et al. [14]. It was performed by plotting % activity and % inhibition of acetylcholinesterase by the plant fraction (inhibitor) concentrations on the graph. The concentrations at the cross-section of these two curves was the IC\textsubscript{50} value. The assay mixture contained briefly 0.2 mL of 0.1 M Tris-HCl buffer (pH 8.0), 20 μL of fraction at different concentrations (1.2, and 3 mg/mL) and 20 μL AChE (3 U/mL) solutions were mixed and incubated for 15 min at room temperature and added 50 μl of 3 mM DTNB to this mixture. The reaction was then initiated by the addition of 50 μl of 15 mM AChI (acetylthiocholine iodide) substrate at different concentrations (0.5-4 mM). At this stage, yellow colour was formed. Galantamine was used as a positive control (1.5 mg/mL). The absorbance of the mixture was measured at 412 nm in a UV-Visible spectrophotometer (Beckman Coulter, USA). The final volume was adjusted to 2 mL with the 50 mM sodium phosphate buffer pH 8[11]. IC\textsubscript{50} values were obtained from (%) activity of the compound concentration graph by measuring the enzyme activity at different inhibition concentrations. In the mixture with inhibitor, the enzyme activity was measured at different concentrations of the substrate [4].

2.5 AChE Enzyme Kinetic Studies

2.5.1 Effects of various concentrations of the enzyme on the rate of reaction

The activity of acetylcholinesterase was assessed for the enzyme at different concentrations [12], ranging from 25-150 μl of AChE (3 U/mL) and 50 μl substrates were taken in all the test tubes. The assays were performed in triplicate.

2.5.2 Effects of varying concentrations of ATCI (acetylthiocholine iodide) substrate on the rate of reaction

The activity of acetylcholinesterase was assessed for acetylthiocholine iodide substrates at different concentrations [13], which ranged from 25-150 μl (ATCI, 15 mM) and uniform (AChE 25 μl, 3 U/mL) remains the same for all the test tubes. The assays were performed in triplicate.

2.5.3 Effects of various concentrations of the substrate on the rate of reaction

The activity of acetylcholinesterase was assessed for substrates at different concentrations. The substrate concentration ranged ATCI from 0.5-4 mM and the AChE enzyme concentration (3 U/mL) remains the same for all the tests [12]. The assays were performed in triplicate.

Fig. 1. Ellman’s reaction
Fig. 2. Inhibitor binds to the active site and directly blocks the active site

2.5.4 AChE enzyme saturation level

The saturation curves were constructed for the calculated acetylcholinesterase enzyme performance at various time(S) intervals [14,15]. Saturation curves were determined at room temperature. The 1mL of reaction mixture contains 50 μl of AChE enzyme (3 U/mL +500 μl DTNB (3 mM) +100 μl of substrate (15 mM, ATCI) 200 μl phosphate buffer ( pH 8.0). All assays were performed three times. The absorbance at 412 nm was determined by UV-spectro-photometer. The absorbance was recorded for every 30 seconds up to 4min. The assays were performed in triplicate.

2.6 Lineweaver-Burk (LB) plot and Michaelis-Menten Equation (Km and Vmax)

The kinetic inhibition of ethanolic and methanolic extracts of plant fraction was analyzed [16]. Ellman reaction mixture consisted 0.2 mL of 0.1 M Tris-HCl buffer (pH 8.0), 20 μL of fraction at different concentration (1.2, and 3 mg/mL) and 20 μl AChE (3 U/mL) solution which were mixed and incubated for 15 min at room temperature and added 50 μl of 3 mM DTNB. The reaction was then initiated by the addition of 50 μL of 15 mM AChI(acetylthiocholine iodide) substrate at different concentrations (0.5-4 mM) when yellow colour was formed. Galantamine was used as a positive control (1.5 mg/mL). The absorbance of the mixture was measured at 412 nm in a UV-VISIBLE spectrophotometer (Beckman Coulter, USA). Each assay was repeated three times. The final volume was adjusted to 2 mL with the 50mM sodium phosphate buffer with pH 8. The kinetic parameters Km and Vmax were determined from Linewear and Burk method [17]. Acetylcholinesterase enzyme concentration was kept constant with different concentrations of substrate (ATCI) ranging from 0.5-4 mM in the presence of AChE (3 U/mL) activity at the different concentration of inhibitor and measured the rate of reaction (1/V). At lower concentrations of substrate, the plot suggested less competitive reaction and at higher concentrations of substrate, the plot showed the competitive mode of production. The Km and Vmax were determined from Linewear and Burk double reciprocal plot analyzed over a different range of AChI concentrations (0.5 to 4 mM), AChE enzyme velocity is the dependent variable and AChI substrate is the independent variable. This experiment generates the Km and Vmax (Michaelis-Menten) plot.

3. RESULTS

3.1 Estimation of IC50 Value

Acetylcholinesterase is inhibited in a concentration-dependent manner. The IC50 values of rhizome of C. orchioides Gaertn., and Oleo diox Roxb fraction 1-3 mg/mL respectively. Galantamine was used to constant the standard curve. By increasing the concentration of plant fractions of C. orchioides Gaertn., and Oleo diox Roxb., the acetylcholinesterase inhibition rate was increased by >50% for 5 min incubation whereas that of rhizome of C. orchioides Gaertn., from>60% for 5 min incubation. The potency of C. orchioides Gaertn., and Oleo diox Roxb was compared to that of standard galantamine, the drug which is being used for the treatment of Alzheimer disease. Galantamine was found to have an IC50 value of 1.5 mg/mL O. diox Roxb., is at least 30 times more potent than galantamine, and C. orchioides Gaertn., is 50 times more potent than O.diox Roxb., and Galantamine. The AChE enzyme inhibition kinetics of C. orchioides Gaertn., and Oleo diox Roxb., fraction were determined from LB plots,
and both showed non-competitive and competitive inhibitions.

### 3.2 Effects of Various Concentrations of the Acetylcholinesterase Enzyme (AChE) and Acetylthiocholine Iodide (ATCI) Substrate on the Rate of Reaction

The varying AChE enzyme concentration with constant substrate concentration was taken. The effects of varying AChE enzyme concentrations on the rate of reaction, substrate concentrations remaining the same were being determined [Graph1]. The graph indicates that with an increase in AChE concentration using constant ATCI substrate concentration, the rate of reaction increased the formation of thiocholine and acetic acid. The AChE can take up to numerous catalytic reactions per second. To determine the maximum speed of an enzymatic reaction, the substrate concentration is the same until the increasing rate of reaction and product formation. This is the $V_{\text{max}}$ of the AChE enzyme, active sites of which are saturated with ATCI, Michaelis and Mentent [18-21].

### 3.3 Effects of Varying Concentrations of ATCI (Acetylthiocholine iodide) Substrate on the Rate of Reaction

The varying ATCI substrate concentrations and constant AChE concentration were taken and the effects were determined [Graph2]. The graph indicates that with an increase in substrate concentration, using AChE concentrations remaining the same, the rate of reaction increases with the formation of thiocholine and acetic acid. The substrate attentiveness is increased until a stable rate of product formation [18-21]. This is the $V_{\text{max}}$ of the enzyme. In this state, all AChE sites are active and saturated with the substrate (acetylthiocholine iodide). Since the substrate concentration at $V_{\text{max}}$ value exactly cannot be measured, AChE enzymes are categorized by the substrate concentration at the rate of reaction is half its maximum.

### 3.4 Pre-steady-state Kinetics

The AChE is mixed with the substrate, at zero seconds. There is no product formed and there are no intermediates exist. The next few seconds of the reaction mixture is called pre-steady-state

| Table 1. IC$_{50}$ of fraction of C. orchioides Gaertn and Oleo diox Roxb |
|-----------------------------------------------|
| **Inhibition IC50±SD (mg/mL)**                  |
| **Galanttamine**                              | **1 mg/mL** | **2 mg/mL** | **3 mg/mL** |
| 1.5 mg/mL                                      | 1.23±0.53    |             |             |
| Name of the plant                              | **1 mg/mL** | **2 mg/mL** | **3 mg/mL** |
| C. orchioides Gaertn                           | 2.29±0.73    | 3.59±0.80   | 5.36±1.11   |
| Oleo diox Roxb                                 | 2.21±0.43    | 3.25±1.08   | 2±0.81      |

Graph 1. Different concentrations of Enzyme Vs Substrate
kinetics, which is shown that the formation and consumption of AChE and ATCI substrate intermediates pending their steady-state kinetics are reached. Incase of AChE, an intermediate is formed by an attack on the active site of the substrate and the formation of the product [21,22]. In the first few seconds, the AChE enzyme forms the product. The final when steady state is reached the amount of product released is slowdown which indicates the saturation curves of AChE [Graph-3]. The amount of product formed is shown in the Y-axis and x-axis shows the steady-state time or saturation level time.

3.5 Effects of Varying Concentrations of ATCI Substrate in mM on the Rate of Reaction

The rate of a reaction concerning AChE also increases as the substrate concentration increases. The limited number of active sites are available in the enzyme. The available enzyme active sites are occupied with substrates. Consequently, increasing the substrate concentration it is observed that it will not alter the rate of diffusion [23]. Some maximum rate of reaction ($V_{\text{max}}$) when all the enzyme active sites are occupied by the substrate. The rate of reaction increases with increasing substrate concentration, but must asymptotically approach the saturation rate or level [Graph4]. $V_{\text{max}}$ is always directly proportional to the total enzyme concentration, and the catalytic constant of the enzyme [24].

3.6 Lineweaver-Burk (LB) plot and Michaelis-Menten equation ($K_m$ and $V_{\text{max}}$)

The substrate specificity of acetylcholinesterase (AChE) was evaluated by testing AChI as a substrate varying from 0.5 to 4 mM. AChE hydrolyzes acetylthiocholine. Substrate kinetic
parameters for AChI were calculated by Km and Vmax values by Lineweaver-Burk plot [5A & 6A] values [25,26]. The Lineweaver-Burk plot was used to calculate Vmax and K_m (Michaelis constant). The initial velocity of AChE was calculated for each substrate concentration individually. This K_m value is also called Michaelis-Menten constant. The enzyme-substrate complex is converted to a product. How quickly enzyme active sites become saturated can be described by the variable K_m, [Graphs 5B& 6B] the substrate concentration at which the reaction rate is V_max.

4. DISCUSSION

The medicinal plant extracts of O.diox Roxb. (F2 fraction) and Curculigo orchioides Gaertn. Rhizome (F3 fraction) caused significant inhibition of AChE activities in vitro studies. The highest inhibition in acetylcholinesterase activity was noticed in the Curculigo orchioides Gaertn. rhizome in vitro methods. Acetylcholinesterase is found in the cell membrane it degrades which the neurotransmitter acetylcholine into choline and acetate, is the important reaction for the regulation of synaptic activity in the nervous system [27].

The rate of product formation increases with increasing substrate concentration [Graphs 1,2&3]. However, in the presence of enzymes, the enzyme reaction has higher values and a much steeper slope. This implies enzymes greatly increase the rate of reaction. However, as the enzymes become saturated, the reaction rate levels remain constant [Graph3] [28]. The LB plot was used to study enzyme inhibition kinetics.

A total of 2 semi-purified plant fractions of 2 different plants were screened for AChE inhibition activity. The inhibition sample was initiate for each semi-purified fraction by plotting 1/[V] versus 1/[S] (LB plot). The following were the distribution of inhibition patterns: Two fractions at different concentrations had competitive inhibition, mixed inhibition, and non-competitive inhibition. Hence Lineweaver–Burk and Michaelis-Menten plots have been shown in Graphs 5A, 5B and 6A, 6B, showed mixed inhibition, uncompetitive inhibition, and competitive inhibition. The plots for galantamine, as a known AChE inhibitor, have been shown in graphs 5A, 5B and 6A,6B.

In this study, 2 plant fractions were assayed at one is O.dioxRoxb levels another one is Curculigo orchioides Gaertn. Rhizome extract at different concentrations (1-3 mg/mL), fraction have shown competitive inhibition in which inhibitor only binds to free enzyme and Vmax and Km increase. F2 fraction have shown mixed inhibition in which inhibitor can bind to either free enzyme or enzyme-substrate both and herein Vmax decreases while Km increases. Two extracts have shown non-competitive inhibition. Non-competitive inhibitors bind to enzyme and enzyme-substrate both with equal affinity and Km remains same but Vmax decreases. F2 and F3 Fraction have shown uncompetitive inhibition in which inhibitor only binds to enzyme-substrate complex. In uncompetitive pattern Vmax and Km both decrease [Table 2].

In this study, the best IC_{50} with 2mg/mL belongs to Curculigo orchioides Gaertn. (Rhizome: ethnoal) following by O. diox Roxb [Leaves; Methanol; Table 2].
Table 2. The plant name, part used, the solvent used, Km and Vmax values from Lineweaver-Burk plot, and inhibition pattern CI-competitive inhibition; MI-Mixed inhibition; NCI-Non-Competitive inhibition

| Sl. no | Plant name       | Part used  | Solvent used for extraction | $V_{max}$ 1.5 mg/mL | $K_{max}$ 1.5 mg/mL | Inhibition pattern |
|--------|------------------|------------|-----------------------------|----------------------|---------------------|--------------------|
| 1      | Galantamine      | -          | -                           | 5.291                | 0.645               | CI                 |
| 2      | Oleo diox Roxb.  | Leaf       | Methanol                    |                      |                     | I                  |
| 3      | Curculigo orchioides Gaertn. | Rhizome | Ethanol                     |                      |                     | NCI                |

$V_{max}$ and $K_{max}$ values are given in mg/mL.
At the lower concentrations of a plant fractions suggested uncompetitive inhibition and at higher concentrations (3 mg/mL) of fractions showed the non-competitive mode of inhibition [13].

Hence, overall from LB plot, it was a mixed type of inhibition, which is very common in medicinal plants due to the presence of a different type of compounds present in the fractions [29-32]. In *Oleo diox Roxb.*, F2 fraction and *Curculigo orchioides Gaertn.* Rhizome F3 fraction inhibited AChE in a concentration-dependent manner (1 to 3 mg/mL). The F3 fraction showed maximum inhibition of AChE enzyme at 3 mg/mL final concentration. The IC_{50} value obtained from the inhibition curve was 2 mg/mL and *Oleo diox*
5. CONCLUSION

Plant fractions (F2 and F3) contained certain bioactive compounds that exhibited the AChE inhibition. However, this might be due to the synergistic effect of many compounds in this plant fraction. Further purification and identification should be accomplished for understanding the exit mechanism of AChE inhibitory activity. The screening of AChE inhibition in O. diox Roxb., and Curculigo orchioides Gaertn., fractions was carried out using semi-purified fraction which might contain bioactive compounds such as the flavonoids, Phenolics and tannins. The type of bioactive compounds and the percentage of compounds which inhibit AChE in the fraction are yet to be explored. Generally, active compound percentage of any plant fraction is<1% and moreover, several types of bioactive compounds would effectively inhibit the AChE site and thus may display inhibition capacity. It is likely that the bioactive compound complexing with some other compounds would stop the reaction between bioactive compounds and AChE and thus decrease the inhibition activity probably due to change in solubility of bioactive compound to reduce its concentration in the final stock solution. Because of this reason, it is not fair to compare the inhibition activity of a pure compound with that of a plant fraction.

In the present study, we report that Oleo diox Roxb., F2 fraction and Curculigo orchioides Gaertn., rhizome F3 fraction showed significant AChE inhibitory activities which may contribute to the development of a more effective drug from these plants. The Curculigo orchioides Gaertn., rhizome fractions showed more specific and effective inhibitors against acetylcholinesterase enzyme which is useful in designing new drugs for the treatment of AD. Further examination of finding new bioactive compounds in Curculigo orchioides Gaertn., plant is going on.

CONSENT

It is not applicable.

ETHICAL APPROVAL

It is not applicable.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Kato G, Tan E, Yung J. Acetylcholinesterase kinetic studies on the mechanism of atropine inhibition. Journal of Biological Chemistry. 2019;247(10):3186-3190.
2. Lineweaver H, Burk D. The determination of enzyme dissociation constants. Journal of the American Chemical Society. 1934;56(3):658-666.
3. Ahmed M, Latif N, Khan R, Ahmad A, Rocha JBT, Mazzanti CM, et al. Enzymatic and biochemical characterization of Bungarussindanus snake venom acetylcholinesterase. Journal of Venomous Animals and Toxins including Tropical Diseases. 2012;18(2):236-243.
4. Sassa A, Beard WA, Shock DD, Wilson SH. Steady-state, pre-steady-state and single-turnover kinetic measurement for DNA glycosylase activity. Journal of Visualized Experiments. 2013;(78):50-95.
5. Ashraf M, Ahmad K, Ahmad I, Ahmad S, Arshad S, Shah SMS, et al. Acetylcholinesterase and NADH oxidase inhibitory activity of some medicinal plants. J. Med. Plants Res. 2011;5(10):2086-2089.
6. Bagewadi Z, Baligar P. Phytoconstituents investigation by lc-ms and evaluation of anti-microbial and anti-pyretic properties of cynodondactylon. IJPSR. 2014;5(7):2874-2889.
7. Tung BT, Thu DK, Thu NTK, Hai NT. Antioxidant and acetylcholinesterase inhibitory activities of ginger root (Zingiber Officinale Roscoe) extract. Journal of Complementary and Integrative Medicine. 2017;14(4):1-15.
8. Kumar S, Seal CJ, Okello EJ. Kinetics of acetylcholinesterase inhibition by an aqueous extract of Withania somnifera roots. International Journal of Pharmaceutical Sciences and Research. 2018;2:1-7.
9. James D, Harvey L, David B. Molecular cell biology, scientific American Books, W. H. Freeman and Co., New York, NY, Gamete Research. 2015;17(1):95-195.
10. Baldwin RL. Structure and mechanism in protein science. A guide to enzyme catalysis and protein folding. Protein Science. 2008;9(1):207-211.

11. Pratap GK, Ashwani S, Manjula S. Alzheimer’s disease: A challenge in managing with certain medicinal plants- A review. IJPSR, 2017;8(12):4960-4972.

12. De Ferrari GV, Canales MA, Shin I, Weiner LM, Silman I, Inestrosa NC. A structural motif of acetylcholinesterase that promotes Amyloid β-Peptide fibril formation†. Biochemistry. 2001;40(35):10447-10457.

13. Howes MJR, Houghton PJ. Plants used in Chinese and Indian traditional medicine for improvement of memory and cognitive function. Pharmacology Biochemistry and Behavior. 2003;75(3):513-527.

14. Kamal MA, Greig NH, Alhomida AS, Al-Jafari AA. Kinetics of human acetylcholinesterase inhibition by the novel experimental alzheimer therapeutic agent, tolserine. Biochemical Pharmacology. 2000;60(4):561-570.

15. Knapp MJ. A 30-week randomized controlled trial of high-dose tacrine in patients with Alzheimer’s disease. The Tacrine Study Group. JAMA: The Journal of the American Medical Association. 1994;271(13):985-991.

16. Lahiri DK, Farlow MR, Greig NH, Sambamurti K. Current drug targets for Alzheimer’s disease treatment. Drug Development Research. 2002;56(3):267-281.

17. Michaelis L, Menten M. The original Michaelis constant: translation of the 1913 Michaelis-Menten paper. Biochemistry. 2011;450(39):8264-9.

18. Pohanka M, Hrabinova M, Kuca K, Simonato JP. Assessment of Acetylcholinesterase Activity Using Indoxylacetate and Comparison with the Standard Ellman’s Method. International Journal of Molecular Sciences. 2011;12(4):2631-2640.

19. Saleem M, Jamous R, Yousef S. In-vitro screening of acetylcholinesterase inhibitory activity of extracts from Palestinian indigenous flora in relation to the treatment of Alzheimer’s disease. Functional Foods in Health and Disease. 2014;4(9):381-400.

20. Soni N, Singh DK, Singh VK. Inhibition kinetics of acetylcholinesterase and phosphatases by the active constituents of Terminalia arjuna and Tamarindus indica in the Cerebral Ganglion of Lymnaea acuminata. Pharmacognosy Journal. 2017;9(2):148-156.

21. Wetitay aklung P, Limmatvatiprat C, Phaechamud T, Keokitchai S. Kinetics of acetylcholinesterase inhibition of Quisqualis indica Linn. Flower extract. kinetics of acetylcholinesterase inhibition silpakorn U science and Tech J. 2007;1(2):20-28.

22. Pohanka M, Hrabinova M, Kuca K, Simonato JP. Assessment of acetylcholinesterase activity using indoxylacetate and comparison with the Standard Ellman’s Method. International Journal of Molecular Sciences. 2011;12(4):2631-2640.

23. Sobhani R, Pal AK, Bhattacharjee A, Mitra S, Aguan K. Screening indigenous medicinal plants of Northeast India for Their anti-alzheimer’s properties. Pharmacognosy Journal. 2016;9(1):46-54.

24. Croghan PC. Modeling dynamic phenomena in molecular and cellular biology. By L. E. Segel. Quarterly Journal of Experimental Physiology. 1985;70(4):642-656.

25. Chowdhury S, Kumar S. In vitro anti-acetylcholinesterase activity of an aqueous extract of Unicariatomentosa and in silico study of its active constituents. Bioinformation. 2016;12(3):112-118.

26. Niño J, Hernández JA, Correa YM, Mosquera OM. In vitro inhibition of acetylcholinesterase by crude plant extracts from Colombian flora. Memórias do Instituto Oswaldo Cruz. 2006;101(7):783-785.

27. Kumar S, Chowdhury S. Kinetics of acetylcholinesterase inhibition by an aqueous extract of Cuminum cyminum seeds. International Journal of Applied Sciences and Biotechnology. 2014;2(1):64-68.

28. Wszelaki N, Kuciun A, Kiss A. Screening of traditional European herbal medicines for acetylcholinesterase and butyrylcholinesterase inhibitory activity. Acta Pharmaceutica. 2010;60(1).

29. Zechel DL, Konermann L, Withers SG, Douglas DJ. Pre-steady state kinetic analysis of an enzymatic reaction monitored by time-resolved electrospray ionization mass spectrometry. Biochemistry. 1998;37(21):7664-7669.

30. Deveci E, Tel-Çayan G, Duru ME. Essential oil composition, antioxidant,
anticholinesterase and anti-tyrosinase activities of two Turkish plant species: *Ferula eleocharis* and *Sideritis stricta*. Natural Product Communications. 2018; 13(1):13-18.

31. Murata K, Tanaka K, Akiyama R, Noro I, Nishio A, Nakagawa S, et al. Anti-cholinesterase activity of crude drugs selected from the ingredients of incense sticks and heartwood of *Chamaecyparis obtusa*. Natural Product Communications. 2018;13(7):15-26.

32. Sichaem J, Tip-pyang S, Lugsanangarm K. Bioactive aporphine alkaloids from the Roots of *Artabotrys Spinosus*: Cholinesterase inhibitory activity and molecular docking studies. Natural Product Communications. 2018;13(10):1-8.

Peer-review history:
The peer review history for this paper can be accessed here:
http://www.sdiarticle4.com/review-history/53648