Mitigation of particulate matter and airborne pathogens in swine barn emissions with filtration and UV-A photocatalysis

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Abstract: This study evaluated the use of filtration and UV-A photocatalysis for the reduction of particulate matter (PM) and airborne bacterial pathogens in swine barns. Two MERV filters (8 and 15) were used to mitigate PM concentrations measured at the PM 1, PM 2.5, respirable PM, and PM 10 ranges. Filtration was also used to generate different levels of airborne pathogens to be treated by UV-A. Results show that MERV 8 and 15 filters effectively reduced PM concentrations (96-98%) in air exhausted from a swine barn (p ranged from < 0.01 to 0.04). UV-A photocatalysis did not mitigate PM concentrations. UV-A photocatalysis treatment reduced measured colony-forming units (CFUs) by 15-95%. The CFU percent reduction was higher when airborne PM concentration was low. The numeric results suggested a real mitigation effect despite p-values that did not meet the usual statistical cut-off of <0.05 for significance due to the large variability of the CFU control samples. Normalization of measured airborne pathogen concentrations by smaller PM size range concentrations led to emerging significant treatment differences for CFUs. A significant decrease (~60% reduction; p < 0.03) in the concentration of viable airborne bacteria was shown for all PM below the 10-micron range.

Keywords: air pollution control; biosecurity; animal diseases; ultraviolet light; advanced oxidation; filtration; environmental technology;

1. Introduction

Bioaerosols composed of organic dust (proteins, complex carbohydrates), microorganisms (viruses, bacteria, and fungi) and endotoxins are believed to contribute to respiratory infections in workers and livestock [1,2,3]. Thus, human and animal health could benefit from improved air quality and reduced exposure to airborne pathogens inside barns. Air filtration can mitigate particulate matter (PM) and reduce airborne infectious pathogens, and some swine operations use high-efficiency particulate air (HEPA) filters to remove dust and infectious airborne from incoming ventilation air [4]. While inlet air filtration can reduce exposure to airborne pathogens and PM [5,6], it is expensive to install and maintain, e.g., the estimated maintenance cost for inlet air HEPA filters for a typical swine barn is > $80,000 per year [4].

Ultraviolet (UV) light can mitigate pathogens and gases, but the bactericidal effects depend on the wavelength, dose, and other factors that challenge the techno-economic considerations for farm-scale adoption [7]. The light within the UV portion of the spectrum (wavelengths 100-400 nm) exhibits widely different effects not only because of the
four-fold range of energy per photon but because of absorption probabilities that vary by orders of magnitude for different materials and wavelengths.

The 100-290 nm range (commonly referred to as "UV-C") is the most energetic and most universally absorbed, although even within that range, enormous differences in effect among wavelengths are observed because of variation in absorption. In fact, the range of 100-200 nm is often referred to as the Vacuum UV because these wavelengths are strongly absorbed by the ordinary components of air (and thus can only be transmitted in a vacuum), which means that for many applications, the practical UV-C range is roughly 200-290 nm. Nonetheless, even this range is considered bactericidal and most "bactericidal lights" are usually 254 nm lamps. Unfortunately, even the more practical 200-290 nm range sources (including the common 254 nm lamps) are more dangerous and toxic to humans and animals than longer wavelengths.

Alternatively, UV-A (320-400 nm) has the longest wavelength in the UV spectrum and is the least toxic. In broad terms, this is because it is absorbed by the fewest materials. However, that selectivity can be used to advantage in some cases because only certain things are affected. The 254 nm lamps are much more effective for use in cases where direct irradiation is desired and human or animal exposure can be avoided. However, coupling UV-A irradiation with a semiconductor photocatalyst that absorbs these wavelengths and produces surface-bound oxidative intermediates has been proven effective in mitigating air pollutants in animal agriculture. Costa et al. [8] and Guarino et al. [9] reported that UV-A photocatalysis in swine farrowing and nursery rooms reduced PM (17%), NH₃ (30%), and selected greenhouse gases (GHGs) (CH₄: ~27%, CO₂: ~15%) while improving feed conversion efficiency (12%). Follow-up research consistently reported the benefits of UV-A photocatalysis to mitigate livestock odor and gaseous emissions [10-20].

To date, there are no farm-scale data on the performance of UV-A photocatalysis in mitigating airborne pathogens in U.S.-based swine production facilities. Therefore, we conducted an on-farm study on mitigating airborne bacterial pathogens emitted from swine barn using PM filtration and UV-A photocatalysis. Two HEPA (high-efficiency particulate air) filters (MERV 8 and MERV 15 efficiency) were used to mitigate PM concentrations measured at the PM 1, PM 2.5, respirable PM, PM10, and total PM size ranges. To be clear, there was no expectation for photocatalysis or direct irradiation to directly affect the non-viable fraction of PM. The effect of UV-A photocatalysis on bacteria (sometimes referred to as 'viable PM') was measured using filtration to control PM concentration and generate specific concentrations of airborne bacterial pathogens, as measured in colony-forming units (CFU). The concept of the experimental design is shown in Figure 1.
2. Results and Discussion

2.1 Treatment of PM by HEPA filtration and UV-A photocatalysis

The effect of HEPA filtration and UV-A photocatalysis on PM was investigated under three conditions with increasingly higher PM concentrations. The 'best-case', 'midpoint', and 'worst-case' scenarios (Figure 1) were achieved by air filtration before the UV treatment (Tables 1, 2, and 3). Table 1 reports the results when MERV 8 and 15 filters removed 96-98% of incoming PM, and the treatment conditions reflect the low dust concentration, i.e., the 'best-case' scenario. Table 2 reports the results when the MERV 8 was used to remove 77-86% of incoming PM, i.e., the 'midpoint' scenario. Finally, Table 3 reports the results for unfiltered exhaust from the manure pit fan, i.e., the 'worst-case'. Notably, UV-A photocatalysis did not affect the PM concentration under any of the three scenarios.

Table 1. Particulate matter removal performance of the MERV 8 and 15 (Treatment I) and UV-A photocatalysis (Treatment II) in mitigating PM (the 'best-case' scenario). Bold signifies statistical significance.
Table 2. Particulate matter removal performance of the MERV 8 (Treatment I) and UV-A photocatalysis (Treatment II) in mitigating PM (the ‘midpoint’ scenario).

|                         | Total PM | PM 10 | Respirable PM | PM 2.5 | PM 1 |
|-------------------------|----------|-------|---------------|--------|------|
|                         | Conc (mg·m⁻³) | % R (p-value) | Conc (mg·m⁻³) | % R (p-value) | Conc (mg·m⁻³) | % R (p-value) | Conc (mg·m⁻³) | % R (p-value) |
| Control (before filtration, sampling port 1) | 0.267 ± 0.241 | - | 0.205 ± 0.185 | - | 0.153 ± 0.175 | - | 0.146 ± 0.174 | - |
| Treatment I (after MERV 8 and 15 filtration, sampling port 2) | 0.061 ± 0.028 | 77.1 (0.055) | 0.048 ± 0.018 | 76.6 (0.056) | 0.027 ± 0.009 | 82.6 (0.09) | 0.022 ± 0.007 | 84.7 (0.10) |
| Treatment II (after UV-A, sampling port 3) | 0.061 ± 0.016 | 0.7 (0.96) | 0.050 ± 0.011 | -3.2 (0.84) | 0.029 ± 0.005 | -7.0 (0.61) | 0.024 ± 0.005 | -7.1 (0.62) |

Table 3. Particulate matter removal performance of UV-A photocatalysis (Treatment II) in mitigating PM under no filtration condition (the ‘worst-case’ scenario).

|                         | Total PM | PM 10 | Respirable PM | PM 2.5 | PM 1 |
|-------------------------|----------|-------|---------------|--------|------|
|                         | Conc (mg·m⁻³) | % R (p-value) | Conc (mg·m⁻³) | % R (p-value) | Conc (mg·m⁻³) | % R (p-value) | Conc (mg·m⁻³) | % R (p-value) |
| Control (before filtration, sampling port 1) | 0.203 ± 0.169 | - | 0.122 ± 0.116 | - | 0.070 ± 0.075 | - | 0.063 ± 0.075 | - |
| Treatment I (after MERV 8 and 15 filtration, sampling port 2) | 0.201 ± 0.096 | 0.6 (0.98) | 0.124 ± 0.055 | -1.4 (0.97) | 0.064 ± 0.044 | 9.4 (0.85) | 0.057 ± 0.043 | 10.5 (0.85) |
| Treatment II (after UV-A, sampling port 3) | 0.201 ± 0.082 | 0.0 (0.99) | 0.139 ± 0.077 | -10.7 (0.72) | 0.081 ± 0.074 | -21.4 (0.64) | 0.073 ± 0.075 | -21.9 (0.67) |

Note: Conc = Concentration, % R = percent reduction, UV-A dose = 5.3 mJ·cm⁻²; Sampling port numbers and locations are illustrated in Methods.

MERV filtration was effective in removing airborne PM, regardless of the inherent variability in PM concentrations. As expected, it was verified that UV-A photocatalysis did not reduce PM. Variation in the real-time measured PM under the 'best-case', 'midpoint', and 'worst-case' scenarios for PM filtration prior to the UV treatment is illustrated in Figures S1-S3. The measured PM concentrations varied at ~4× the order of magnitude around the mean. Such variability is typical [21], i.e., the PM concentration in the manure pit headspace is instantaneously affected by barn ventilation, ambient air wind velocity, and wind direction. Also, the PM 'Control' concentrations, particulate size distribution, and composition vary with major diurnal and seasonal trends. Thus, variability in PM is a consideration in data analysis.
2.2 Treatment of airborne pathogens with UV-A photocatalysis

UV-A photocatalysis treatment reduced CFU by 15-95% (Table 4). The CFU percent reduction was higher when airborne PM concentration was low ('best-case' > 'worst-case'). To be specific, the 'best-case' PM filtering scenario resulted in the same-day reductions from 2,930 to 133 CFUs and 1,000 to 67 CFUs, i.e., 95 and 93% percent reduction. However, CFU counts were lowered such that measurements produced Control and UV treatment values outside of each other’s standard deviations, i.e., the variability meant that p-values did not meet the usual <0.05 test for statistical significance.

Table 4. CFU removal performance of UV-A photocatalysis (Treatment II) in mitigating airborne pathogens. Comparison of CFU before and after UV-A photocatalysis and resulting percent reduction.

| Scenario | Control (inlet to UV mobile lab, location #2, Figure 2) | UV treatment (outlet of UV mobile lab, location #3, Figure 2) | % Reduction | p-Value |
|----------|---------------------------------------------------------|-------------------------------------------------------------|--------------|---------|
|          | C1 (CFU) | C2 (CFU) | C3 (CFU) | Mean ± S.D. | T1 (CFU) | T2 (CFU) | T3 (CFU) | Mean ± S.D. |
| MERV 8&15 (Best) | 5.0×10³  | 2.0×10³  | 3.6×10³  | 2,930 ± 2,470 | 4.0×10¹  | 0  | 0  | 133 ± 231 | 95  | 0.17 |
|         | 6.0×10²  | 2.4×10³  | 0  | 1,000 ± 1,250 | 2.0×10²  | 0  | 0  | 67 ± 115 | 93  | 0.34 |
| MERV 8 (Midpoint) | 6.8×10³  | 7.2×10³  | 9.8×10³  | 7,930 ± 1,630 | 5.0×10¹  | 5.0×10³  | 5.8×10³  | 5,270 ± 462 | 34  | 0.06 |
| No filtration (Worst) | 1.3×10⁴  | 1.2×10⁴  | 1.1×10⁴  | 12,100 ± 1,410 | 9.6×10³  | 7.8×10³  | 1.3×10⁴  | 10,300 ± 2,860 | 15  | 0.52 |
|         | 6.4×10⁴  | 2.8×10⁴  | 1.6×10⁴  | 36,300 ± 24,700 | 2.4×10⁴  | 2.6×10⁴  | 1.6×10⁴  | 21,900 ± 5,500 | 40  | 0.38 |

Despite p-values that did not meet the usual statistical cut-off of <0.05 for significance, the large variability of the CFU control samples (Table 4), the numeric results suggest a real effect. A normalization procedure that accounts for the wide variability of the control data is discussed paragraphs below.

2.2.1 Normalization of the data

Unlike experimental conditions, variability in PM (including viable PM) concentrations and in laboratory-based CFU enumeration cannot be controlled in the field. Therefore, the CFU and PM data were normalized (see Materials and Methods Equations 2 and 3) prior to analysis. Statistical analysis of normalized data revealed that UV-A treatment had a significant effect on the partial filtration case (% R for CFU_D = 43%, p-value = 0.04, Table 5).

Table 5. CFU removal performance of UV-A photocatalysis (Treatment II) in mitigating airborne pathogens. Comparison of CFU_D (CFU∙m⁻³) normalized by the total PM concentration (µg∙m⁻³) before and after UV-A photocatalysis (CFU_pm, CFU∙µg⁻¹) and resulting percent reduction. Bold signifies statistical significance.

| Scenario       | Control (CFU_pm) | UV treatment (CFU_pm) | % Reduction | p-Value |
|----------------|------------------|-----------------------|--------------|---------|
| MERV 8&15 (Best) | 1,890 76 1,361 1,110 ± 933 | 148 0 0 49 ± 86 | 96  | 0.17 |
|               | 965 241 0 402 ± 502 | 57 0 0 19 ± 33 | 95  | 0.29 |

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Additional data analyses were performed to deconvolute the effect of UV treatment on airborne pathogens when normalized by measured PM concentrations at different sizes. Thus, the average of CFU\textsubscript{PM} values was normalized by different PM sizes’ concentrations under three different PM scenarios ('best-case', 'midpoint', and 'worst-case'; Table S1) and then compared the Control and Treatment (Table 6). The rationale was consistent with the fact that the PM size of airborne bacteria and viruses favors their overall contribution to the fine particulate (e.g., PM 2.5, PM 1). This had two practical implications; first, the averaging reduced some of the previously observed variability. Second, it removed the filtration effect altogether from the study (a filtration effect would affect both the Control and UV Treatment the same; thus, it cancels out when the difference is calculated).

Significant mitigation (60-61%) effect of UV-A photocatalysis on pathogen inactivation was reported in this study when considered in the context of the PM load and PM size ranges (Table 6), in particular for respirable, PM 2.5 and PM 1. Notably, normalization by smaller PM size range concentrations leads to emerging significant differences, while total and larger PM sizes (e.g., PM 10) did not indicate a statistically significant difference.

**Table 6. CFU removal performance of UV-A photocatalysis (Treatment II) in mitigating airborne pathogens. Comparison of CFU\textsubscript{D} (CFU m\textsuperscript{-3}) normalized by the different particle size (total, PM 10, respirable, PM 2.5, PM 1) concentrations (µg m\textsuperscript{-3}) before and after UV-A photocatalysis (CFU\textsubscript{PM}, CFU µg\textsuperscript{-1}). Bold signifies statistical significance.**

| Scale                     | Mean Control | Mean Treatment | Mean Difference | Test Statistic | % Reduction | p-value |
|---------------------------|--------------|----------------|-----------------|----------------|-------------|---------|
| Raw CFU                   | 12,000       | 7,387          | 4,612           | 1.80           | 56          | 0.147   |
| CFU\textsubscript{D} (CFU m\textsuperscript{-3}) normalized by Total PM | 514          | 135            | 379             | 2.15           | 64          | 0.098   |
| CFU\textsubscript{D} (CFU m\textsuperscript{-3}) normalized by PM 10 | 637          | 247            | 350             | 1.83           | 53          | 0.141   |
| CFU\textsubscript{D} (CFU m\textsuperscript{-3}) normalized by Respirable PM | 1,053        | 519            | 571             | 3.35           | 60          | 0.029   |
| CFU\textsubscript{D} (CFU m\textsuperscript{-3}) normalized by PM 2.5 | 1,236        | 608            | 666             | 4.24           | 61          | 0.013   |
| CFU\textsubscript{D} (CFU m\textsuperscript{-3}) normalized by PM 1 | 1,293        | 660            | 6,716           | 4.03           | 60          | 0.015   |

Note: The responses were averaged over the three replicates (C1, C2, C3 and T1, T2, T3 in Table 5), i.e., C\textsubscript{avg} and T\textsubscript{avg}, respectively. Then, the percent reduction (% R) was estimated as the ratio of (C\textsubscript{avg} – T\textsubscript{avg})/C\textsubscript{avg} *100%. Test Statistic was estimated as the ratio of Mean Difference and the standard deviation of the difference and was distributed as t-distribution. Mean Difference = Mean Control – Mean Treatment. Mean Control & Mean Treatment = averages of normalized CFU\textsubscript{D} in Control & Treatment, respectively.

| MERV 8 (Midpoint) | 302 | 320 | 435 | 352 ± 72 | 192 | 192 | 222 | 202 ± 18 | 43  | 0.04 |
|-------------------|-----|-----|-----|---------|-----|-----|-----|---------|-----|------|
| No filtration (Worst) | 393 | 358 | 311 | 354 ± 41 | 228 | 186 | 319 | 244 ± 68 | 31  | 0.20 |
|                   | 623 | 269 | 163 | 352 ± 241 | 174 | 189 | 117 | 160 ± 38 | 55  | 0.28 |
The results of this research support the anti-microbial effects of UV-A. UV-A is less bactericidal than UV-C [4,22]. However, a bactericidal effect has been reported for UV-A at sufficient dosage [23,24] under laboratory conditions. Further investigation of the potential UV-A treatment of airborne pathogens is warranted, especially for indoor barn applications.

Several improvements can be proposed to the PM sampling protocols, including selecting a fully mechanically ventilated barn and aiming for sampling periods of relative low variability in PM (e.g., nighttime, when the ‘Control’ is expected less variable). Further improvements could be made to optimizing the airborne pathogen sampling to target specific microorganisms besides the total load.

3. Materials and Methods
3.1 Viable particulate matter sampling system

The sampling system was designed to simultaneously collect replicate (n=3) airborne PM samples (Figure 2; Figures S4-S5). Three polytetrafluoroethylenes (PTFE) tubes were inserted into the treated air via the sampling ports (Figure S6), then connected to impingers (Biosampler, SKC Inc., Eighty Four, PA, USA). The impingers captured airborne PM using the culture medium (20 mL of brain heart infusion broth, BHI) and 0.1% (v/v) anti-foam A emulsion with a flow rate of 20 L·min\(^{-1}\) (Figure S7). Impingers were placed in a container filled with ice during sampling (Figure S7). Each impinger’s sampled airflow rate was adjusted by a rotameter (Dwyer Instruments Inc., Michigan City, IN, USA) (Figures S5, S8, S9). The manifold system was made for the vacuum distribution and simultaneous sample collection into three impingers. PVC adapter (ID = 89 mm, Lowes, Ames, IA, USA) and screw caps (ID = 89 mm, Lowes, Ames, IA, USA) were used to close both ends of manifolds (Figure S4). PVC primer and cement were used to seal the gaps between these parts. A gauge (Grainger, Des Moines, IA, USA) was installed in the manifold to monitor the vacuum. A constant airflow through the manifold was facilitated by a vacuum pump (Becker Pumps Corp., Cuyahoga Falls, OH, USA). The vacuum pump was turned on first, reached 20 L·min\(^{-1}\) airflow setpoint in each rotameter, and was maintained during each experiment.

![Figure 2. Schematic of an airborne pathogen sampling system. Red arrow: sampled air with the viable and non-viable particulate matter emitted from the swine barn ventilation fan and trapped in biosample impingers; Blue arrow: vacuum-induced airflow.](image)

3.2 Mobile lab setup in the swine farm
The mobile laboratory had an adjustable airflow system (0.28-1.25 m$^3$·s$^{-1}$) for mitigating emissions with UV-A photocatalysis [18-20]. The mobile lab was connected to the swine manure pit fan (Figure 3, Figure S10). The minimum treated airflow was used (facilitating 52 s UV-A treatment time from inlet to outlet in the mobile laboratory) in this experiment for maximizing the treatment time and the potential for mitigation effect. Also, the highest UV-A intensity (0.41 mW·cm$^{-2}$) was used (Figure S11) for the same purpose. Detailed information on the UV-A dose, light intensity, treatment time, airflow, the testing farm was published in Lee et al., 2021 [20].

The rationale for using the filtration upstream of UV treatment was to control the airborne PM concentrations and facilitate the separation of treatment effects between the UV and filtration (Figures S5, S12). A total of three PM concentration levels (conditions) were generated for testing the UV treatment in the mobile laboratory by using two HEPA filters (MERV 8 and 15, detailed information was shown in the previous research, Lee et al., 2021 [18]).

- The 'best-case' scenario – MERV 8 and 15 filters out most PM prior to UV treatment.
- The 'midpoint' scenario – partial filtration and a moderate PM load in the treated air. Only MERV 8 filter removes a fraction of airborne PM prior to the UV treatment.
- The 'worst-case' scenario – no PM filtration. Raw swine barn exhaust discharged from the manure pit is treated by UV.

**Figure 3.** Schematic of particulate matter and airborne pathogens sampling ports (yellow) in the UV mobile laboratory and filtration system. Three-dimensional ‘arrows’ represent free airflow. (1) Brown ‘arrow’: exhaust air from swine barn fan; (2) Red ‘arrow’: inlet air with reduced particle matter load after MERV filtration; (3) Blue ‘arrow’: treated air with UV-A photocatalysis.

### 3.3 UV-A dose

The maximum UV-A dose (5.3 mJ·cm$^{-2}$) provided by the mobile lab was then used to determine the mitigation effect of airborne pathogens. The UV-A dose was estimated as the average of the irradiated UV doses on all surfaces coated with TiO$_2$ (nanostructured TiO$_2$ anatase at 10 µg·cm$^{-2}$ from PureTi, Cincinnati, OH, USA) of the mobile lab. Each chamber inside the mobile laboratory was equipped with 5 UV-A LED lamps (0.04 mW·m$^{-2}$ intensity, 47.6 s treatment time). The treatment time represents a realistic residence times in mechanically ventilated barns. An additional 100 UV-A lamps (0.41 mW·m$^{-2}$, 9.5 s) were installed on a removable rack in chambers #2–#3 to maximize UV dose. The UV-A LED light used (T8 LED, Eildon Technology, Shenzhen, China) had 367 nm as the dominant
wavelength (Figure S11). Detailed information on the measurements of UV-A light intensity, treatment time, photocatalyst, UV lamps is published elsewhere [18].

3.4 Enumeration of total colony-forming unit

The procedure for total plate count was performed according to Laird et al., 2004 [25], with minor modifications. Briefly, air samples collected in 20 mL of brain heart infusion broth (BHI) (Remel INC, San Diego, CA, US) were centrifuged at 400G for 10 min, at room temperature. Then, 15 mL was pipetted out of the tube, and the remaining 5 mL were homogenized by vortexing. For CFU determination, serial ten-fold dilutions were prepared by pipetting 0.1 mL of the sample into 0.9 mL of phosphate buffer. Next, 0.1 mL of each dilution and the undiluted sample were inoculated onto the 5% sheep blood in tryptic soy agar (Hardy Diagnostics, Chicago, IL, US) plates. The liquid was spread into the agar with a sterile loop, immobilizing the cells on the surface of the agar and allowing the growth of distinct, non-overlapping colonies on the agar plates. The agar plates were incubated for 48 h at 37 °C, and colony-forming units (CFU) were counted at that dilution where 20-200 CFU were visible for counting (Figures S13, S14). Impingers, air sampling tubing, and connectors were autoclaved between trials (Figure S15).

3.5 Measurement of non-viable particulate matter size and concentrations

The PM concentration was measured using TSI DustTrak (Monitor 8533, Shoreview, MN, USA). Real-time airborne PM concentrations were measured at five size ranges (PM 1, PM 2.5, ‘respirable,’ PM 10, and total PM) for 1 h. The ‘respirable’ was defined as the ‘PM 4 to PM 10’ range (Figures S1-13). The airborne pathogens were sampled simultaneously with the PM real-time measurements for 1 h.

4.6 Data analysis

The rotameter’s (Figure 1) reading of the airflow rate for each impinger was corrected to the standard temperature and pressure using Eq 1 [26]:

$$ Q_{st} = Q_{ob} \times \frac{P_{st} \times T_{st}}{P_{ob} \times T_{ob}} $$

Where: $Q_{st}$ = standard flow corrected for pressure and temperature (normal temperature and pressure condition, 20 °C, and 14.5 psi, L·min$^{-1}$), $Q_{ob}$ = measured flowmeter reading from rotameter (L·min$^{-1}$), $P_{st}$ = standard pressure (1 atm, 14.7 psi), $P_{ob}$ = measured absolute pressure (psi): atmospheric pressure (14.7 psi) ± gauge pressure (psi), $T_{st}$ = standard temperature (293.15 K), $T_{ob}$ = measured temperature (273.15 + T °C)

The obtained CFU was divided by the airflow rate adjusted to normal temperature and pressure (NTP, defined as 20 °C and 1 atm), the number of CFU per sampled air volume for 1 h was estimated (Eq. 2).

$$ CFU_D = \frac{CFU}{V_{sampled \ air}} $$

Where: $CFU_D$ = density of sampled total CFU under NTP condition (CFU·m$^{-3}$), $CFU$ = measured total CFU per sample, $V_{sampled \ air}$ = total sampled air volume by each impinger for 1 h (m$^3$) at NTP.

Additional data analyses were performed to deconvolute the effect of UV treatment of airborne pathogens at different PM size ranges and when considered in the context of the PM load. The evidence of the PM load as a potential carrier of pathogens is illustrated in Figure S16. The value of CFU$D$ was divided by the measured PM concentration to calculate the CFU$PM$ (CFU per measured PM, Eq. 3). These additional analyses aimed to reduce the inherent variability in the real-time PM concentrations and associated with the CFU enumeration. The detailed process of calculating the quantified CFU$PM$ using Equations 1-3 was showed in Tables S2-S4.
\[ CFU_{PM} = \frac{CFU_D}{PM} \times 1,000 \]  

(3)

Where: \( CFU_{PM} \) (CFU µg⁻¹) = the ratio of \( CFU_D \) (CFU m⁻³) per PM concentration at a specific PM size range (mg m⁻³).

The mitigation effect was evaluated by the overall mean percent reduction (% R), Eq.4.

\[ \% R = \frac{Con - Treat}{Con} \times 100 \]  

(4)

Where: Con and Treat are \( CFU_{PM} \) (CFU µg⁻¹) and PM concentration in Control and Treatment, respectively.

3.7 Statistical analyses

The R Studio (version 3.6.2) was used to analyze the mitigation effect under UV-A photocatalysis treatment (reported in Table 1, 2, 3, 4, and 5). The control CFU value and treatment CFU value were statistically analyzed using one-way ANOVA. The statistical difference was confirmed by obtaining the \( p \)-value through the Paired Tukey test. A significant difference was defined for a \( p \)-value <0.05.

An additional statistical model in R Studio (version 4.0.6) was used to assess the effectiveness of UV treatment then the raw CFU data were normalized by PM size (reported in Table 6). A paired t-test was run over the averages of the three replicates.

4. Conclusions

This proof-of-the concept aimed to investigate whether particulate matter (PM) filtration (at MERV 8 and 15 range) and UV-A photocatalysis could reduce PM and airborne pathogens discharged from swine barn. The following general conclusions were made:

- MERV 8 and 15 effectively mitigated PM concentrations (96-98%, \( p \) ranged from < 0.01 to 0.04) in swine barn exhaust.
- UV-A photocatalysis does not affect PM concentrations in swine barn exhaust.
- UV-A photocatalysis treatment reduced CFU by 15-95%. The CFU percent reduction was higher when airborne PM concentration was low ('best-case' > 'worst-case'). UV-A photocatalysis reduced the concentration of airborne pathogens (43% reduction, \( p = 0.04 \)) in moderate PM concentration conditions.
- Despite \( p \)-values that did not meet the usual statistical cut-off of <0.05 for significance, the large variability of the CFU control samples, the numeric results suggested a real effect.
- Normalization of measured airborne pathogen concentrations by smaller PM size range concentrations led to emerging significant treatment differences for CFUs. Significant mitigation (60-61%, \( p \) ranged from 0.01 to 0.03) effect of UV-A photocatalysis on pathogen inactivation was observed when considered in the context of the PM load and PM size ranges in particular for the respirable, PM 2.5 and PM 1, i.e., below the 10-micron range.

Additional improvements to the experimental approach for farm-scale testing were proposed, and further investigation is warranted.

**Supplementary Materials:** Detailed PM and CFU data is presented in Figures S1-S3 and Tables S1-S4. Detailed information about the experimental setup and methods used is illustrated in Figures S4-S16.

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**References**

1. Seedorf, J. An emission inventory of livestock-related bioaerosols for Lower Saxony, Germany. *Atmospheric Environment*, 2004, 38(38), 6565-6581. doi:10.1016/j.atmosenv.2004.08.023.
2. Costa, A., Borgonovo, F., Leroy, T., Berckmans, D., & Guarino, M. Dust concentration variation in relation to animal activity in a pig barn. *Biosystems Engineering*, 2009, 104(1), 118-124. doi:10.1016/jbiosystemseng.2009.05.009.
3. Donham, K. J. (1991). Association of environmental air contaminants with disease and productivity in swine. *American Journal of Veterinary Research*, 1991, 52(10), 1723-1730.
4. Li, P.; Koziel, J.A.; Zimmerman, J.J.; Zhang, J.; Cheng, T.; Yim-Im, W.; Jenks, W.S.; Lee, M.; Chen, B.; Hoff, S.J. Mitigation of Airborne PRRSV Transmission with UV Light Treatment: Proof-of-concept. *Agriculture*, 2021, 11, 259, doi:10.3390/agriculture11030259.
5. Dee, S.; Batista, L.; Deen, J.; Pijoan, C. Evaluation of an air-filtration system for preventing aerosol transmission of porcine reproductive and respiratory syndrome virus. *Can. J. Vet. Res.*, 2005, 69, 293–298
6. Dee, S.; Otake, S.; Deen, J. Use of a production region model to assess the efficacy of various air filtration systems for preventing airborne transmission of porcine reproductive and respiratory syndrome virus and Mycoplasma hyopneumoniae: Results from a 2-year study. *Virus Res.*, 2010, 154, 177–184.
7. Holtkamp, D. J.; Johnson, C.; Koziel, J. A.; Li, P.; Murray, D.; Ruston, C. R.; Stephan, A.; Torremorell, M.; Wedel, K. Ultraviolet C (UVC) Standards and Best Practices for the Swine Industry. *Agricultural and Biosystems Engineering Technical Reports and White Papers*, 2020, 29.
8. Costa, A.; Chiarello, G.L.; Selli, E.; Guarino, M. Effects of TiO$_2$ based photocatalytic paint on concentrations and emissions of pollutants and on animal performance in a swine weaning unit. *J. Environ. Manag.*, 2012, 96, 86–90, doi:10.1016/j.jenvman.2011.08.025.
9. Guarino, M.; Costa, A.; Porro, M. Photocatalytic TiO$_2$ coating—to reduce ammonia and greenhouse gases concentration and emission from animal husbandries. *Bioresour. Technol.*, 2008, 99, 2650-2658, doi:10.1016/j.biortech.2007.04.025.
10. Lee, M.; Wi, J.; Koziel, J.A.; Ahn, H.; Li, P.; Chen, B.; Meiirkhanuly, Z.; Banik, C.; Jenks, W. Effects of UV-A light treatment on ammonia, hydrogen sulfide, greenhouse gases, and ozone in simulated poultry barn conditions. *Atmosphere*, 2020, 11, 283, doi:10.3390/atmos11030283.
11. Rockafellow, E.M.; Koziel, J.A.; Jenks, W.S. Laboratory-Scale Investigation of UV treatment of ammonia for livestock and poultry barn exhaust applications. *J. Environ. Qual.*, 2012, 41, 281–288, doi:10.2134/jeq2010.0536.
12. Koziel, J.A.; Yang, X.; van Leeuwen, J.; Jenks, W.S.; Laor, Y. Treatment of odorous VOCs with ultraviolet light. *Chem. Eng. Trans.* 2010, 23, 363, doi:10.3303/CET1023601.
13. Zhu, W.; Koziel, J.A.; Maurer, D.L. Mitigation of livestock odors using black light and a new titanium dioxide-based catalyst: Proof-of-concept. *Atmosphere*, 2017, 8, 103, doi:10.3390/atmos8060103.
14. Lee, M.; Li, P.; Koziel, J.A.; Ahn, H.; Wi, J.; Chen, B.; Meiirkhanuly, Z.; Banik, C.; Jenks, W.S. Pilot-scale testing of UV-A light treatment for mitigation of NH$_3$, H$_2$S, GHGs, VOCs, odor, and O$_3$ inside the poultry barn. *Front. Chem.*, 2020, 8, 613, doi:10.3389/fchem.2020.00613.
15. Maurer, D.L.; Koziel, J.A. On-farm pilot-scale testing of black ultraviolet light and photocatalytic coating for mitigation of odor, odorous VOCs, and greenhouse gases. *Chemosphere*, 2019, 221, 778–784, doi:10.1016/j.chemosphere.2019.01.086.
16. Koziel, J.A.; Yang, X.; Cutler, T.; Zhang, S.; Zimmerman, J.J.; Hoff, S.J.; Jenks, W.S.; Laor, Y.; Ravid, U.; Armon, R. Mitigation of odor and pathogens from CAFOs with UV/TiO₂: Exploring the cost effectiveness. In Proceedings of the Mitigating Air Emissions from Animal Feeding Operations, Des Moines, IA, USA, 19–21 May 2008; pp. 169–173.

17. Yang, X.; Koziel, J.A.; Laor, Y.; Zhu, W.; van Leeuwen, J.H.; Jenks, W.S.; Hoff, S.J.; Zimmerman, J.; Zhang, S.; Ravid, U. VOC removal from manure gaseous emissions with UV photolysis and UV-TiO₂ photocatalysis. Catalysts 2020, 10, 607, doi:10.3390/catal10060607.

18. Lee, M.; Koziel, J.A.; Murphy, W.; Jenks, W.S.; Fonken, B.; Storjohann, R.; Chen, B.; Li, P.; Banik, C.; Wahe, L. Design and testing of mobile laboratory for mitigation of gaseous emissions from livestock agriculture with photocatalysis. Int. J. Environ. Res. Public Health 2021, 18, 1523, doi:10.3390/ijerph18041523.

19. Lee, M.; Koziel, J.A.; Murphy, W.; Jenks, W.S.; Li, P.; Banik, C. Evaluation of TiO₂ based photocatalytic treatment of odor and gaseous emissions from swine manure with UV-A and UV-C. Animals, 2021, 11(5), 1289. doi: 10.3390/ani11051289.

20. Lee, M., Koziel, J.A., Murphy, W., Jenks, W.S., Chen, B., Li, P., and Banik, C. Mitigation of Odor and Gaseous Emissions from Swine Barn with UV-A and UV-C Photocatalysis. Atmosphere, 2021, 12(5), 585.

21. Heber, A. J.; Lim, T. T.; Ni, J. Q.; Tao, P. C.; Schmidt, A. M.; Koziel, J. A.; Hoff, S. J.; Jacobson, L. D.; Zhang, Y.; Baughman, G. B. Quality assured measurements of animal building emissions: particulate matter concentrations. Journal of the Air and Waste Management Association, 2006, 56, 1642-1648. doi:10.1080/10473289.2006.10464569

22. Le, T. T. N.; Nagata, H.; Takahashi, A.; Aihara, M.; Okamoto, T.; Shimohata, T.; Mawatari, K.; Akutagawa M.; Kinouchi, Y.; Haraguchi, M. Sterilization effect of UV light on Bacillus spores using TiO₂ films depends on wavelength. The Journal of Medical Investigation, 2012, 59(1,2), 53-58. doi:10.2152/jmi.59.53.

23. Ramesh, T.; Yaparatne, S.; Tripp, C. P.; Nayak, B.; Amirbahman, A. Ultraviolet light-assisted photocatalytic disinfection of Escherichia coli and its effects on the quality attributes of white grape juice. Food and Bioprocess Technology, 2018, 11(12), 2242-2252.

24. Kühn, K. P.; Chaberny, I. F.; Massholder, K.; Stickler, M.; Benz, V. W.; Sonntag, H. G.; Erdinger, L. Disinfection of surfaces by photocatalytic oxidation with titanium dioxide and UVA light. Chemosphere, 2003, 53(1), 71-77. doi:10.1016/S0045-6535(03)00562-X.

25. Laird, D.T.; Gambrel-Lenarz, S.A.; Scher, F.M.; Graham, T.E.; Reddy, R. Microbiological Count Methods, in HM Wehr and JF Frank (eds), Standard Methods for the Examination of Dairy Products, American Public Health Association, Washington, DC, 2004, pp. 153-186.

26. Li, P.; Koziel, J.; Zimmerman, J.; Hoff, S.; Zhang, J.; Cheng, T.; Yim-Im, W.; Lee, M.; Chen, B.; Jenks, W. Designing and Testing of a System for Aerosolization and Recovery of Viable Porcine Reproductive and Respiratory Syndrome Virus (PRRSV): Theoretical and Engineering Considerations. Frontiers in Bioengineering and Biotechnology, 2021, 9:659609. doi: 10.3389/fbioe.2021.659609.