Research Article

The Effect of Glycyrrhetinic Acid on Pharmacokinetics of Cortisone and Its Metabolite Cortisol in Rats

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The purpose of this paper is to study pharmacokinetics of cortisone (E) and its metabolite cortisol (F) in rats after administration of glycyrrhetinic acid (GA) and cortisone. Healthy male SD rats were randomized to be given 20 mg/kg E or E combined with 10 mg/kg GA. Blood samples were collected at 5, 10, 20, 40, 60, 90, 120, 150, 180, and 240 min after administration. The serum concentrations of E and F were determined by HPLC and pharmacokinetic parameters were calculated using DASver2.0 software. The parameters of AUC (0–t), AUC (0–∞), and Cmax for E in the group of E + GA were significantly higher than those in the group of E (P < 0.01); the half-time (t1/2β) was extended compared to E (P < 0.05) and CL/F was dropped obviously (P < 0.01). The rise in AUC (0–t), AUC (0–∞), and Cmax for cortisol in the group of E + GA was significantly compared to the group of E (P < 0.01). CL/F was lower than E (P < 0.01) and the half-time (t1/2β) was slightly extended. In this study, we find that GA restrains the metabolism of E and F and thus increases AUC, t1/2β, and Cmax of E and F, which may be related to its inhibition effect on 11β-hydroxysteroid dehydrogenase (11β-HSD).

1. Introduction

Licorice is one of the most commonly used clinical drugs [1]. When glycyrrhizin is administered orally, glycyrrhetinic acid is the major metabolite [2]. Glycyrrhetinic acid (GA) is an important bioactive substance of licorice with a variety of pharmacological effects, such as anti-inflammatory [3], antiviral [4], antiulcer [5], and adrenal cortical hormone kind function [6]. Clinical trials clearly show that glycyrrhizin has a good effect on all types of dermatitis [7], purulent scar disease [8], and hair follicle infection and can cure gingivitis, esophagus inflammatory disease. It plays a physiological function in reducing lipemia and antiatherosclerotic and preventing atherosclerosis [9, 10]. Many enzymes are inhibited by glycyrrhetinic acid in corticosteroids metabolic process, such as 17β-HSD, 11β-HSD, 5α-reductase, and 5β-reductase [11–13].

Two distinct isozymes of 11β-HSD catalyze the interconversion of hormonally active cortisol (F) and inactive cortisol (E), which are important human glucocorticoids [14–16]. This occurs via inhibition of the enzyme 11-β-hydroxysteroid dehydrogenase. Glucocorticoids have anti-inflammatory, immunosuppressive, antishock, and other pharmacological roles, with clinically important applications [17].

We develop a high performance liquid chromatography method for the simultaneous determination of cortisone and cortisol in rat serum. The pharmacokinetics of cortisone and cortisol in rats after the administration of glycyrrhettinic acid and cortisone were studied.

2. Experimental

2.1. Chemicals, Reagents, and Stock Solutions. Cortisone (lot no. 0001447641, purity >98.0%) and trifluoroacetic acid (TFA) (lot no. 064K3647, purity >98.0%) were purchased from Sigma-Aldrich Company (St. Louis, MO, USA).
Cortisol and pirfenidone (both purity >98.0%) were gifts from Rockefeller University (New York, USA). All other chemicals were analytical grade and used without further purification. Purified water was prepared in house with a Milli-Q water system from Millipore (Bedford, MA, USA).

The drugs were accurately weighed (±0.0001 g), on an analytical balance from METTLER TOLEDO AB204-A (Zurich, Switzerland), to prepare stock solutions of individual compounds at a concentration of 1 mg/mL in methanol. All of the standard solutions were stored in the dark at 277 K for a maximum of one month. Working solutions for calibration and controls were prepared from the stock solution by dilution using methanol. The internal standard (IS) working solution (100 μg/mL) was prepared by diluting its stock solution with methanol. Calibration curves were prepared using blank serum spiked at concentrations of 50, 100, 250, 500, 1000, 2000, 4000, and 6000 ng/mL. Low, medium, and high quality control (QC) samples at concentrations of 100, 1000, and 4000 ng/mL were prepared in the same way as the calibration standard.

2.2. Equipment and Conditions. HPLC system (Agilent 1100) was equipped with quaternary pump, on-line vacuum degasser, autosampler, column compartment, diode array detector, and Agilent Chem Station Rev A.10.02. Chromatographic separation was achieved on a 4.6 × 150 mm, 5 μm particle and an Agilent Zorbax SB-C18 column was kept constant during on the run: a source of temperature of 298 K using a one-step linear gradient. Mobile phases A (acetonitrile), B (0.1% TFA), and C (water) ratio changed as follows: 0–12 min, 24–34% A, 40% B, and 36–26% C. The detector operated at 246 nm for cortisone and cortisol. The injection volume was 20.0 μL, and the flow rate of mobile phase was 0.9 mL/min.

2.3. Calibration Standards and Quality Control Samples. Stock solutions of cortisone (1 mg/mL), cortisol (1 mg/mL), and IS (1 mg/mL) were separately prepared in 25 mL volumetric flasks with methanol and stored at 277 K. Working solutions for calibration and controls were prepared from the stock solution by dilution using methanol. The IS working solution (100 μg/mL) was prepared by diluting its stock solution with methanol. Calibration curves were prepared using blank serum spiked at concentrations of 50, 100, 250, 500, 1000, 2000, 4000, and 6000 ng/mL. Low, medium, and high quality control (QC) samples at concentrations of 100, 1000, and 4000 ng/mL were prepared in the same way as the calibration standard.

2.4. Animals and Treatment. Male SD rats, weighing 300 ± 20 g (Certificate no. 2007-0005) were purchased from Shanghai SLAC Laboratory animal Co., Ltd. Rats were divided into 2 groups, each group was administered with glycyrrhetinic acid 10 mg/kg or vehicle (DMSO versus water = 1:9 (V/V)) three times at 0, 12, and 24 h via intraperitoneal injection. Rats were anesthetized by 10% chloral hydrate before intraperitoneal injection of cortisone 20 mg/kg. Blood samples 800 μL were directly collected into a clean tube from the tail vein at 5, 10, 20, 40, 60, 90, 120, 150, 180, and 240 min. After centrifugation at 5000 rpm for 10 min, serum was separated and stored at 253 K until analysis.

2.5. Sample Preparation. The serum samples were prepared using liquid-liquid extraction technique. A 300 μL aliquot of the serum sample was taken in a 10 mL glass test tube, and on that 20 μL of IS (100 μg/mL) was spiked and 150 μL of sodium hydroxide solution (0.01 mol/L) was added. To this, 3 mL of extraction solvent (ethyl acetate) was added and vortex mixed on a multiple vortexer for 2.0 min, followed by centrifugation at 4000 rpm for 15 min. The supernatant was dried under nitrogen and dissolved in 200 μL of mobile phase. A 20 μL aliquot of the final supernatant was injected into the HPLC system for analysis.

2.6. Method Validation. The analytical method was validated to demonstrate the selectivity, recovery, accuracy, and precision of measurements.

Selectivity was established by the lack of interference peaks at the retention time for cortisone, cortisol, and IS. Recovery of the method was determined at three concentration levels (100, 1000, and 4000 ng/mL) by comparing the peak area obtained from the extracted serum samples with the peak area obtained by the direct injection of the corresponding concentration spiked standard solution in the extracted blank serum.

QC samples at three concentration levels (100, 1000, and 4000 ng/mL) were analyzed to assess the accuracy and precision of the method. Intraday accuracy and precision (n = 6) were evaluated by assays of QC samples at different times during the same day. Interday accuracy and precision were tested from the results of replicate assays of QC samples over three consecutive days. Accuracy of the method was estimated based on relative error (RE) and precision was estimated by the relative standard deviation (RSD).

2.7. Statistical Analysis. Data were expressed as X ± s. The significance of differences was assessed by unpaired t-test and one-way analysis of variance.

3. Results

3.1. Method Validation. In our study, the resolution of cortisone, cortisol, and their internal standard was satisfactory. No interference from internal materials in serum can be observed in the HPLC chromatograms. The retention time of cortisone, cortisol, and internal standard was 10.1 min, 9.3 min, and 7.6 min, respectively. The HPLC chromatograms are shown in Figure 1.

The regression equation of cortisone was \( Y_1 = 0.1991X_1 + 0.0041 \) (r = 0.9998) and its lower limit of quantitation (LLOQ) was 0.05 mg/L (n = 6; RSD = 3.70%); the regression equation of cortisol was \( Y_2 = 0.2262X_2 - 0.0033 \) (r = 0.9999) and its LLOQ was 0.05 mg/L (n = 6; RSD = 5.22%). The extractive recoveries for cortisone were (82.00 ± 0.64)%,(79.68 ± 2.04)% and (81.21 ± 0.98)% respectively at different serum standard concentrations of 0.1, 1.0, and
3.2. Pharmacokinetic Study of Cortisone and Cortisol. To investigate the pharmacokinetic parameters, we applied compartmental model. The mean concentration-time curves of cortisone in two groups were presented at Figure 2 and the pharmacokinetic parameters were presented at Table 1. The mean concentration-time curves of cortisol in two groups were presented in Figure 3 and the pharmacokinetic parameters were presented in Table 2.

4. Discussion

We studied the pharmacokinetic characteristics of E and F in rats combined using GA and E and then fitting their pharmacokinetic parameters. A two-compartment model was found meeting concentration-time curves of cortisone and cortisol in group E and E + GA, $T_{1/2}$, CL/F, $AUC_{(0–t)}$, $AUC_{(0–\infty)}$, and $C_{\text{max}}$ of cortisone and cortisol were affected.
Although radioimmunity is used to detect glucocorticoids usually for its high sensitivity; the result would be much higher than the actual concentration [18, 19]. Because target molecule, glucocorticoids, would cross-react with their isomers or metabolites [25]. Previous research had proved the result that conducted by ELISA and RIA assay it would have the same problem [26]. In this work, we employed an HPLC-DAD assay to detect E and F concentrations in serum. Compared to the methods mentioned above, the specificity of HPLC-DAD is much higher, so the result is much more reliable.

Authors’ Contribution

Dan Lin and Wei Sun contributed equally to this work.

Conflict of Interests

None of the authors has any other conflict of interests related to this paper.

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