Research article

Early response genes induced in chondrocytes stimulated with the inflammatory cytokine interleukin-1β

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Abstract

Recent work has established that IL-1β plays a central role in the inflammation and connective tissue destruction observed in both rheumatoid arthritis and osteoarthritis. These processes result from the activity of this inflammatory cytokine to activate expression of genes for neutral proteases, such as the matrix metalloproteinases. While IL-1β activates matrix metalloproteinase genes within several hours, it also activates immediate early genes, which are required for the later expression of matrix metalloproteinases and other arthritis-perpetuating genes, are also activated. To identify putative immediate early genes involved in IL-1β-mediated arthritic disease, a chondrocytic cell line (SW1353) was stimulated with this cytokine for 2 hours, total RNA was isolated, and expressed genes were identified by microarray analysis. This analysis identified alterations in the expression of multiple transcription factors, cytokines, growth factors and their receptors, adhesion molecules, proteases, and signaling intermediates that may contribute to inflammation and cartilage destruction in arthritis. Interestingly, confirmation of the expression of activating protein-1 family members by reverse transcriptase polymerase chain reaction revealed a preferential increase in junB, a known transcriptional antagonist of c-jun. The failure to observe induction of early growth response gene-1, which was detected by reverse transcriptase polymerase chain reaction to be substantially and transiently induced by 1 hour of IL-1 treatment, may be explained by the known instability of the message after early induction. However, this analysis has identified numerous IL-1β-responsive genes that warrant further investigation as mediators of disease in arthritis.

Keywords: chondrocytes, interleukin-1, matrix metalloproteinases, signal transduction, transcription factors

Introduction

IL-1β is a proinflammatory cytokine that has myriad effects on cells, increasing proliferation, activating inflammatory responses, and inducing matrix remodeling through the production of neutral proteases. Since most cells can both express and respond to IL-1β, complex signaling programs are likely required to modulate the timing and magnitude of responses to this cytokine [1]. It is well documented that elevated expression of IL-1β in the joint results in the activation of inflammatory and degradative programs in synovial cells, contributing to the progression of rheumatoid arthritis [2]. Recent work by

AP-1 = activator protein-1; COL2A1 = procollagen 2 alpha 1; egr-1 = early growth response gene-1; ets = erythroblastosis gene twenty-six; IL-1β = interleukin-1β; LH = lactalbumin hydrolysate; LIF = leukemia inhibitory factor; MAPK = mitogen-activated protein kinase; MMP = matrix metalloproteinase; NF-xB = nuclear factor-xB; OA = osteoarthritis; PDGF = platelet-derived growth factor; RT-PCR = reverse transcriptase polymerase chain reaction; SMAD4 = mothers against dpp homolog 4; TGF-β = transforming growth factor-β.
Hasty and coworkers [3] has demonstrated that local expression of IL-1β by articular chondrocytes activates autocrine gene expression, which contributes to the pathogenesis of osteoarthritis (OA). Furthermore, IL-1-receptor antagonists are effective inhibitors of inflammation and connective tissue destruction in animal models of arthritis [2,4,5]. Together, these studies implicate IL-1β as a central mediator of pathogenesis in arthritis.

Activation of this pathologic process is complex, most likely involving the activation or repression of numerous signal/transduction pathways, with subsequent effects on many genes. Our previous work has focused on the pathways by which IL-1β induces several members of the family of matrix metalloproteinases (MMPs), enzymes that are active at neutral pH and that collectively degrade the various components of the extracellular matrix. Specifically, we have focused on MMP-1 and MMP-13, interstitial collagenases that have the unique ability to initiate cleavage of the triple helix comprising the stromal collagen types I, II, and III. Elevated expression of MMP-1 and MMP-13 by stromal cells and chondrocytes in response to IL-1β contributes to the pathogenesis of connective tissue disorders [6–11]. Studies from this laboratory and others showed that IL-1 activates MMP-1 and MMP-13 gene expression through common signaling pathways [11–15]. Specifically, nuclear translocation of NF-κB and mitogen-activated protein kinase (MAPK) stimulation of activator protein-1 (AP-1) are required for transcription of cytokine-induced MMP-1 and MMP-13. Furthermore, cycloheximide blocks mRNA expression of both collagenases [11,16], indicating that expression of immediate early genes is required for MMP-1 and MMP-13 gene expression. Thus, IL-1β activation of collagenase gene expression requires the increased expression of a panel of essential transcription factors and signaling intermediates, as well as the initiation of signaling cascades that activate these factors.

IL-1 also contributes to the pathogenesis of arthritis by increasing the proliferation of mesenchymal cells and by enhancing the expression of other inflammatory mediators [17–19]. This complex alteration in cellular phenotype probably involves the coordinated increase and decrease of genes for cytokines and growth factors, adhesion molecules, and signaling intermediates. The recent availability of microarray technology has made it possible to perform an extensive analysis of IL-1 modulation of genes in cells that are central players in arthritic disease.

We therefore carried out a microarray analysis of IL-1β-stimulated chondrocytic cells in order to identify immediate early genes that could contribute to phenotypic changes relevant to OA. In these studies, we used the human chondrosarcoma cell line SW1353, for three important reasons. First, we, and others, have found that when treated with IL-1β, SW1353 cells serve as an appropriate model for primary chondrocytes in OA [11,20–22]. Second, the amount of total RNA required for microarray analysis precludes the use of primary human chondrocytes for these studies. Third, the inherent genetic variation in primary human chondrocyte cultures could bias the gene expression profile and lead to erroneous conclusions. We found increased expression of a large cohort of transcription factors, cytokines, growth factors, and signaling intermediate genes that are potential regulators of metalloproteinase gene expression, proliferation, and sustained inflammation. Alternatively, this was accompanied by a decrease in a modest number of genes also belonging to these categories, suggesting that IL-1β substantially reprograms gene expression in chondrocytes. A subset of these IL-1β-stimulated genes was then confirmed by semiquantitative reverse transcriptase polymerase chain reaction (RT-PCR). This characterization of IL-1β-induced immediate early genes in chondrocytic cells identifies candidate mediators of the expression of collagenase genes in arthritis.

Materials and methods

Cell culture

SW1353 chondrosarcoma cells were grown to confluence in Dulbecco’s modified Eagle’s medium supplemented with 10% fetal bovine serum (HyClone, Logan, UT, USA), penicillin/streptomycin, and L-glutamine (Cellgro, Mediatech, Herndon, VA, USA). At the beginning of each experiment, cells were washed three times with Hank’s buffered sterile saline (Cellgro) to remove traces of serum and were placed in Dulbecco’s modified Eagle’s medium containing 0.2% lactalbumin hydrolysate (Gibco/BRL, Rockville, MD, USA). Then cells were either left unstimulated or treated with 10 ng/ml recombinant IL-1β (Promega, Madison, WI, USA). This concentration of IL-1 was found to be optimal for activation of MMP-13 gene expression in these cells (data not shown).

Harvest and microarray analysis of mRNA

For microarray analysis, two confluent 150-mm plates of SW1353 cells were cultured in lactalbumin hydrolysate, or lactalbumin hydrolysate plus 10 ng/ml IL-1β, for 2 hours. Cells were then washed with Hank’s buffered sterile saline solution and homogenized in 2 ml Triazol Reagent (Gibco/BRL), and RNA was purified in accordance with the manufacturer’s instructions. The RNA pellets were stored in 70% ethanol and shipped on dry ice to Clontech Laboratories (Palo Alto, CA, USA) via express courier. Clontech analyzed the RNA using their Atlas Human Cancer 1.2K array (1176 unique genes) and AtlasImage 1.0 software. We chose this data baseline because it represents a large number of genes that may be expressed during development and again during disease pathology. We further classified genes that increased or decreased by twofold or more, and sorted them using Microsoft Excel.
Reverse transcriptase polymerase chain reaction
A subset of genes whose expression was increased by IL-1β was then confirmed using a radioactive RT-PCR assay. Briefly, RNA was treated with DNase I and then 2 µg was reverse transcribed with Moloney murine leukemia virus reverse transcriptase and oligodT primers. One tenth of the reverse transcriptase reaction was amplified using gene-specific primers, Platinum Taq DNA polymerase and 33PdATP. Single-band products were resolved on native acrylamide gels and visualized by autoradiography. Table 1 shows the gene-specific primers that were used.

Results and discussion
Table 2 shows the results of an analysis using Clontech’s Human Cancer 1.2K Array and total RNA from SW1353 cells treated with IL-1β (10 ng/ml) for 2 hours. This analysis revealed substantial increases or decreases in the expression of several known IL-1β-responsive genes. These include transcription factors (panel A), cytokines, growth factors, and their receptors (panel B), proteases, matrix proteins, and adhesion molecules (panel C), as well as signaling intermediates and tumor suppressors (panel D).

We focused on genes for transcription factors because they have been reported as IL-1-responsive immediate early genes in chondrocytes [23]. Members of the NF-xB, AP-1, and ets families of transcription factors substantially increased in the Human Cancer 1.2K array (see Table 2, panel A). To confirm the effect of IL-1β on AP-1 family members, we assayed Jun family members by RT-PCR (Fig. 1). We found that the c-Jun and JunB genes increased transiently within 1 hour of IL-1 treatment, before the induction of the MMP-13 gene. The expression profiles for c-Jun and JunB were similar to the profile for egr-1, another immediate early gene in IL-1-stimulated chondrocytes [23]. In contrast, the JunD gene was constitutively expressed, and IL-1β increased this expression modestly. These data show that in IL-1β-stimulated chondrocytes, all three Jun family members are expressed and can contribute to MMP gene expression. Furthermore, RT-PCR analysis of the JunB gene (Fig. 1) validated the gene expression profile derived from the Human Cancer 1.2K array (Table 2, panel A).

Transcription factor genes that were modulated by IL-1β included members from the NF-xB family (relB, p100/p52, p105/p50, IkB-x, c-rel, and p65), the AP-1 family (fra-1 and junB), and the ETS family (ets-1). While it is well established that NF-xB proteins translocate to the nucleus upon IL-1 stimulation [24], we were surprised to find so many family members expressed simultaneously. This suggests that in IL-1β-stimulated chondrocytes, NF-xB family members may compete for dimerization partners and for binding sites in responsive genes.

Members of the AP-1 family of transcription factors are essential transcriptional activators for both the MMP-1 and MMP-13 genes [25–27], and these transcription factors are activated by extracellular stimuli through MAPK pathways [28]. JunB was identified as an IL-1-inducible gene in chondrocytes by the Human Cancer 1.2K array (Table 1) and this finding was confirmed by RT-PCR (Fig. 1). Levels of JunB mRNA peaked within 1 hour of IL-1 stimulation and began to decline after 2 hours. This was somewhat surprising, since MAPK pathways do not activate JunB [29] and in some systems JunB antagonizes transcriptional activation by c-Jun [30]. Since c-Jun, JunB,

Table 1
| PCR primers and conditions |
|---------------------------|
| **Gene** | **Primers** | **Annealing** | **Product** |
|   |           | **temperature** | **Cycles** | **length** |
| MMP-13 | CCAGTGGTGGTATGAGATG | Sense | 52°C | 25 | 143 |
|       | TTCTCGAGCTCTCAGTC | Antisense |   |   |   |
| c-Jun | GAAACGACCTCTATGAGATGGCTCA | Sense | 63°C | 22 | 316 |
|       | GAACCCCTCTGCTATCGTCAGTTCT | Antisense |   |   |   |
| JunD | GATGGTTCGATTCTGCCTCA | Sense | 53°C | 22 | 214 |
|       | TGTGGACTCGTAGCAAAACA | Antisense |   |   |   |
| JunB | CCAGTCCTCCACTCGACTGGTTGAAAG | Sense | 58°C | 22 | 257 |
|       | GACTAATGCGGTGGTTCCTCTCTCACCAGTAC | Antisense |   |   |   |
| egr-1 | TCCCGGCTCCTCGACCTAC | Sense | 60°C | 25 | 177 |
|       | AAGGGTGGCTAGTCATGCCTGAAAG | Antisense |   |   |   |

GAPDH was amplified using Clontech’s human GPDH amplimer set with an annealing temperature of 58°C and 25 cycles of amplification. MMP-13 = matrix metalloproteinase-13; egr-1 = early growth response gene-1. *Measured in base pairs.
### Table 2

**IL-1-responsive genes in chondrocytic cells**

| Fold ↑ | Fold ↓ | Gene description | Genbank# |
|--------|--------|------------------|----------|
| 18.0   |        | I-rel (RELB)     | M83221   |
| 17.0   |        | nuclear factor NF-kappa-B p100 subunit; nuclear factor NF-kappa-B p52 subunit; H2TF1; oncogene lyt-10 | X61498 |
| 15.7   |        | interferon regulatory factor 1 (IRF1) | X14454 |
| 13.7   |        | major histocompatibility complex enhancer-binding protein MAD3 | M69043 |
| 8.1    |        | nuclear factor kappa-B DNA binding subunit (NF-kappaB; NFKB) | M58603 |
| 5.5    |        | octamer-binding transcription factor 2 (oct-2; OTF2); lymphoid-restricted immunoglobulin octamer binding protein NF-A2; POU2F2 | M36542 |
| 4.7    |        | IkappaB epsilon (IKBE) | U91616 |
| 3.0    |        | c-rel proto-oncogene protein | X75042 |
| 2.8    |        | fos-related antigen (FRA1) | X16707 |
| 2.4    |        | STAT-induced STAT inhibitor 3 | ABO04904 |
| 2.4    |        | erythroblastosis virus oncogene homolog 1 (ETS-1); p54 | J04101 |
| 2.3    |        | jun-B | M29039 |
| 2.2    |        | ELAV-like neuronal protein 1 (HEL-N1) + HEL-N2 | U12431 + |
| 2.0    |        | myb proto-oncogene; c-myb | M15024 |
| 2.0    |        | NF-kappaB transcription factor p65 subunit; RELA; NFKB3 | L19067 |
| 2.0    |        | transcription factor erf-1; AP2 gamma transcription factor | U85658 |
| 2.0    |        | early growth response alpha (EGR alpha) | S81439 |
| 6.0    |        | homeobox protein HOX-D3; HOX-4A | D11117 |
| 2.0    |        | retinoblastoma-like protein 2 (RBL2; RB2); 130-kDa retinoblastoma-associated protein | X74594 |
| 2.0    |        | mothers against dpp homolog 4 (SMAD4); MADR4; pancreatic carcinoma gene 4 (DPC4) | U44378 |

**Panel B: Cytokines/Growth Factors/Receptors**

| Fold ↑ | Fold ↓ | Gene description | Genbank# |
|--------|--------|------------------|----------|
| 21     |        | leukemia inhibitory factor precursor (LIF); differentiation-stimulating factor (D factor); melanoma-derived LPL inhibitor (MLPLI); HILDA | X13967; |
| 8.7    |        | interleukin-11 (IL-11); adipogenesis inhibitory factor (AGIF) | M65420 |
| 6.1    |        | interferon-gamma (IFN-gamma) receptor beta subunit precursor; IFN-gamma accessory factor 1 (AF1); IFN-gamma transducer 1 (IFNGT1) | M57765 |
| 6.1    |        | interleukin-6 precursor (IL-6); B-cell stimulatory factor 2 (BSF2); interferon beta-2 (IFNB2); hybridoma growth factor | U05875 |
| 5.9    |        | heparin-binding EGF-like growth factor (HEBGF); diphtheria toxin receptor (DTR) | M14584 |
| 5.0    |        | interferon gamma-induced protein precursor (gamma-IP10) | M60278 |
| 4.3    |        | putative serine/threonine protein kinase PRK | X02530 |
| 4.3    |        | CD40 receptor-associated factor 1 (CRAF1) | U56998 |
| 3.6    |        | platelet-derived growth factor A subunit precursor (PDGFA; PDGF1) | U21092 |
| 3.0    |        | bone morphogenetic protein 3B precursor (BMP3B); growth differentiation factor 10 (GDF10); bone inducing protein (BIP) | X06374 |
| 2.5    |        | CD40 ligand (CD40-L); tumor necrosis factor (TNF)-related activation protein (TRAP); T-cell antigen GP39 | D49493 |
| 2.1    |        | interleukin-12 beta subunit precursor (IL-12B); cytotoxic lymphocyte maturation factor 40-kDa subunit (CLMF p40); NK cell stimulatory factor subunit 2 (NKSF2) | L07414 |
| 2.1    |        | cellular growth factor 2 precursor (GGF2PP2); neutregulin; heresulin-beta3 + neu differentiation factor + heregulin-alpha | M65290 |
| 2.0    |        | platelet-derived growth factor B subunit precursor (PDGFB; PDGF2); bacaplermin; c-sis | L12260; |
| 7.8    |        | bone morphogenetic protein 4 (BMP4) + bone morphogenetic protein 2B (BMP2B) | L12261; |
| 2.3    |        | platelet-derived growth factor receptor alpha subunit (PDGFRα); CD140A antigen | X02811; |
| 2.3    |        | platelet-derived growth factor receptor beta subunit (PDGFRβ); CD140B antigen | X02744; |

*Continued overleaf*
| Fold | Fold | Gene description | Genbank# |
|------|------|------------------|----------|
| 3.0  |      | matrix metalloproteinase 13 (MMP-13); collagenase 3 precursor | X75308   |
| 2.3  |      | matrix metalloproteinase 7 (MMP-7); matrilysin | X07819   |
| 2.1  |      | matrix metalloproteinase 12 (MMP-12); metalloelastase | L23808   |
| 2.1  |      | collagen 6 alpha 2 subunit (COL6A2) | M34570   |
| 2.1  |      | tissue-type plasminogen activator precursor (T-plasminogen activator; TPA) | M15518; X07393; M18182 |

**Panel C: Proteases/Matrix Proteins**

| Fold | Gene description | Genbank# |
|------|------------------|----------|
| 2.0  | urokinase-type plasminogen activator receptor GPI-anchored form precursor (U-PAR); monocyte activation antigen MO3; CD87 antigen | U08839; M83246; X51675 |
| 3.0  | chymotrypsin-like protease CTRL-1 precursor | X71874   |
| 2.0  | cysteine protease | D55696   |

**Adhesion Molecules**

| Fold | Gene description | Genbank# |
|------|------------------|----------|
| 3.5  | ninjurin-1        | U72661   |
| 3.0  | desmocollin 3A/3B precursor (DSC3) + desmocollin 4 (DSC4) | X83929 + D17427 |
| 2.8  | tenascin precursor (TN); hexabrachion (HXB); cytotactin; neuronectin; GMEM; miotendinous antigen; glioma-associated extracellular matrix antigen | X78565; M55618 |
| 2.2  | alpha-catenin related protein (CAPR); alpha 2 catenin (CTNNA2); alpha N-catenin | M94151   |

**Panel D: Signaling Intermediates**

| Fold | Gene description | Genbank# |
|------|------------------|----------|
| 7.0  | rho6 protein     | Y07923   |
| 2.9  | Gem; induced immediate early protein; ras family member (ras-like protein KIR) | U10550   |
| 2.6  | regulator of G protein signaling 3 (RGS3; RGp3) | U27655   |
| 2.5  | dual-specificity protein phosphatase 8; HVH5 | U27193   |
| 2.3  | death receptor 5 (DR5); cytotoxic TRAIL receptor 2 (TRICK2A) | AF016268 |
| 2.1  | protein kinase DYRK3 | Y12735   |
| 2.1  | jagged 1 (JH1)   | AF028593 |

**Tumor Suppressors**

| Fold | Gene description | Genbank# |
|------|------------------|----------|
| 11.0 | melanoma differentiaction-associated protein 7 precursor (MDA7) | U16261   |
| 2.3  | tumor suppressor protein DCC precursor; colorectal cancer suppressor | X76132   |
and JunD are all expressed in IL-1-treated SW1353 chondrocytic cells, it is not clear which family member is responsible for transcriptional activation of collagenase genes. Perhaps the functional transcriptional complex is determined by induced expression and activation of AP-1 accessory proteins, such as the Fos family member Fra-1, or the ets family member ets-1 [31]. This cohort of AP-1, Fos, and ets family members may also preferentially bind to different AP-1 sites in MMP promoters, with subtle effects on gene expression [32].

egr-1 is an IL-1-inducible immediate early gene in chondrocytes [23] and has been implicated in the elevated MMP-1 gene expression observed in rheumatoid synoviocytes [33,34]. Surprisingly, egr-1 expression was increased only 1.3-fold in this Human Cancer 1.2K array (data not shown). However, when this gene was analyzed by RT-PCR, we found a robust increase in egr-1 mRNA within 1 hour of IL-1 stimulation, which declined significantly by 2 hours (Fig. 1). Thus, the relatively short half-life of egr-1 mRNA resulted in an underestimation of IL-1 induction of this gene in the microarray. It is therefore probable that an array using RNA from cells treated for 1 hour with IL-1 would identify additional important genes.

Cytokines and growth factors and their receptors were a major group of IL-1-modulated genes in chondrocytes (Table 2, panel B). For instance, leukemia inhibitor factor precursor (LIF) was substantially induced by IL-1β in SW1353 cells, and this is consistent with previous reports of LIF as an IL-1-responsive gene in chondrocytes [38,39]. Recently, LIF was reported to induce MMP-13 gene expression [40], and this may contribute to the role of this cytokine in arthritis pathology [38]. IL-6 expression was increased in IL-1-treated cells, and while this may affect a variety of cellular functions in chondrocytes, this cytokine does not seem to be required for IL-1-induced suppression of proteoglycan synthesis in cartilage [41]. Interestingly, there was a concurrent increase in expression of platelet-derived growth factor (PDGF) subunits, and a decrease in PDGF receptor subunits. Perhaps this is a mechanism of ensuring paracrine, but not autocrine, effects of PDGF in cytokine-stimulated chondrocytes. The role of growth factors in IL-1-dependent effects is not clear cut, since IL-1 can both induce and inhibit proliferation of chondrocytes [17]. However, in the present model of SW1353 cells under serum-free conditions, we did not observe a mitogenic effect of IL-1 (data not shown).

Bone morphogenetic protein 4 (BMP4) is a member of the TGF-β superfamily of proteins and is required for chondrocyte differentiation and cartilage maintenance [42,43]. The observed suppression of its expression in chondrocytes by IL-1β (Table 2, panel B) is consistent with a previous report in osteoblastic cells [44] and suggests that IL-1β may enhance cartilage degradation by blocking an essential autocrine signal for chondrocyte maturation and function.

IL-1 induction of several MMP family members is seen, including collagenase-3 (MMP-13), matrilysin (MMP-7), and metalloelastase (MMP-12) (Table 2, panel C). The collagenase-1 (MMP-1) gene was not significantly induced in IL-1β repressed three transcription factor genes, including HOX-4A, the retinoblastoma-like protein 2, and mothers against dpp homologue 4 (SMAD4). SMAD4 contributes to transforming growth factor-β (TGF-β)-dependent transcription of the alpha 2(l)-collagen (COL1A2) [35] and aggrecan [36] genes. Furthermore, our microarray data demonstrated that IL-1 inhibited Col2A1 expression by 1.6-fold in SW1353 cells (data not shown). It has been reported that IL-1 inhibition of Col2A1 gene expression requires the p38 MAPK pathway [37]. Perhaps inhibition of SMAD4 synthesis is another component of IL-1-dependent Col2A1 gene repression. While IL-1 promotes cartilage degradation through MMP synthesis and inhibits matrix deposition by reducing Col2A1 gene expression, TGF-β has the opposite effect on chondrocytes [3]. Thus, it is tempting to speculate that IL-1 antagonizes the effects of TGF-β by blocking expression of essential signaling intermediates, such as SMAD4.
Tenascin, an adhesion molecule that is expressed during cartilage development [45], was induced by IL-1β in SW1353 cells (Table 2, panel C). Interestingly, the tenascin gene has been reported to be IL-1-inducible in synovial cells and chondrocytes, and its expression is enhanced in rheumatoid arthritis and OA [46,47]. A possible physiologic consequence of IL-1β-induced tenascin expression is that it may promote chondrocyte migration towards the sites of lesions in OA [47]. IL-1β coordinately reduced expression of integrin α1, a cell-attachment molecule that chondrocytes use to bind to type II collagen [48]. Thus, IL-1β treatment may modify adhesion molecule expression, so that matrix adhesion is compromised.

We found that IL-1β changed the expression levels for several signaling proteins and tumor suppressor genes (Table 2, panel D), but the significance of these changes is not clear. Proteins associated with G-protein signaling were both increased (rho6, Gem, and regulator of G protein signaling) and decreased (rho7, RAD1, and p160ROCK). Jagged, which is a transmembrane ligand for the Notch signaling pathway [49], is induced by IL-1β. Interestingly, jagged-1 is important for epithelial–mesenchymal cell interaction in development [50], and mutations in the jagged gene lead to Alagille syndrome, a congenital connective tissue disorder whose hallmarks include craniofacial and vascular deformities [51]. Zhao and colleagues [52] identified frizzled (denoted FZD2) as the human homologue of the Drosophila polarity-determining gene. Of potential importance is the fact that frizzled family members have recently been implicated by Carson and colleagues in rheumatoid arthritis [53]. A new frizzled family member that is expressed in chondrocytes and is involved in skeletal morphogenesis has been described [54]. Perhaps down-regulation of frizzled family members by IL-1β is deleterious to chondrocyte function and can contribute to OA.

Conclusions
Along with confirming changes in gene expression already known to be associated with IL-1β stimulation, chondrocyte biology, and MMP gene regulation, this microarray identified several other induced and repressed genes whose roles in chondrocyte biology are yet to be defined. While the significance of these findings in terms of understanding IL-1 effects on chondrocytes is still uncertain, the documentation of these changes in gene expression may provide the basis for future studies on the molecular effects of IL-1 on chondrocytes and on other cell types as well.

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