Case Report: Adolescent-Onset Isolated Nephronophthisis Caused by a Novel Homozygous Inversin Mutation

Zhengxia Zhong1*, Xiaoyong Yan1, Zhengying Fang2, Yijun Dong1, Jiaxing Tan3, Jingyuan Xie2, Linhong Hu1, Shibin Zhang4* and Wei Qin3

1Division of Nephrology, Department of Medicine, Affiliated Hospital of Zunyi Medical University, Zunyi, China, 2Department of Nephrology, Shanghai Ruijin Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China, 3Division of Nephrology, Department of Medicine, West China Hospital, Sichuan University, Chengdu, China, 4Laboratory Animal Centers, Zunyi Medical University, Zunyi, China

Objective: Nephronophthisis (NPHP) is a rare autosomal recessive inherited kidney disease that can cause cystic enlargement of the kidneys, and lead to end-stage renal disease (ESRD) before the age of 30 years. Herein we describe a case of adolescent-onset NPHP with a novel homozygous mutation in the inversin gene (INVS).

Methods: The patient was a 15-year-old Chinese boy who presented with ESRD. Genetic testing was performed via whole exome sequencing and validated via Sanger sequencing. A novel homozygous INVS mutation was identified (c. 1909C > T; p. Gln637Ter).

Results: The results of laboratory examinations included urinary protein 1.05 g/24 h, urine erythrocyte count 5/high-power field, serum creatinine 1,026.2 μmol/L, and estimated glomerular filtration rate 5.8 ml/min/1.73 mm². Extrarenal features included hypertension and moderate anemia, and his parents were consanguineous (first cousins). A homozygous 1-bp substitution resulting in a nonsense mutation (c. 1909C > T; p. Gln637Ter) in exon 15 of INVS was detected via whole exome sequencing, and validated via Sanger sequencing. According to the classification system of the American College of Medical Genetics and Genomics, the mutated gene in INVS is strongly pathogenic (PVS1+PM2+PP3+PP5). His parents and a younger brother were heterozygous carriers. Based on the above results he was diagnosed with juvenile type 2 NPHP. He underwent hemodialysis, and received a kidney transplant after 2 months. He is currently recovering well, with a serum creatinine level of 117 μmol/L and an estimated glomerular filtration rate of 79.6 ml/min/1.73 mm².

Conclusion: Here we have described an extremely rare case of adolescent-onset type 2 NPHP caused by a homozygous INVS mutation. The patient had progressed to ESRD by the age of 15 years. The current report will deepen our understanding of the clinical and genetic basis of this disease.

Keywords: adolescent-onset, homozygous mutation, INVs, NPHP2, ESRD
INTRODUCTION

Nephronophthisis (NPHP) is an extremely rare autosomal recessive kidney disease characterized by renal tubulointerstitial lesions, tubular basement membrane disruption, and renal cyst formation that progresses to end-stage renal disease (ESRD) in children (Otto et al., 2003). In addition to renal symptoms NPHP is often complicated with diverse extrarenal features, and approximately 20% of patients suffer from ciliopathy syndrome, including retinal degeneration, bone abnormalities, and liver fibrosis (Srivastava et al., 2017). Genetic studies have identified more than 20 genes that are causally associated with NPHP in humans. Juvenile type I NPHP (NPHP1) is most commonly caused by homozygous pathogenic variants in NPHP1, which encodes an SH3 domain protein (Hildebrandt et al., 1997a; Otto et al., 2008). Type 2 NPHP (NPHP2) has also been identified in patients with infantile NPHP, and is distinguished from other types of NPHP by an early age of onset and cystic enlargement of the kidneys. O’Toole et al. (2006) first reported NPHP2 mutation in a 2-year-old Arab boy with retinitis degeneration and kidney failure. Here in we report the novel INVS mutation p. Gln637Ter in an adolescent patient with NPHP2.

CASE

A 15-year-old Chinese boy who had been experiencing headache and vomiting for 1 week was admitted to the hospital (IV-2 in Figure 1). Ophthalmic examination did not reveal any obvious abnormalities, there were no neurological symptoms, and he did not exhibit short stature. His clinical symptoms were sudden and severe, and mainly manifested as hypertension and severe kidney failure. His blood pressure was 170/123 mmHg, his serum creatinine level was 1,026.2 μmol/L, and no prior symptoms were reported. His parents were consanguineous (first cousins), and his older brother had succumbed to an undiagnosed kidney disease at 8 years of age, without a kidney biopsy. The current patient was initially admitted to our medical center on 21 May 2021.

Ultrasonography depicted small kidneys and diffuse lesions (85–90 mm in diameter) in both kidneys, but no cystic enlargement of the kidneys. Laboratory tests revealed a hemoglobin level of 67 g/L and a urinary protein level of 1.05 g/24 h. Chest computed tomography was normal. Laboratory investigations on admission included screening for hepatitis B virus surface antigen, antinuclear antibodies, anti-double-stranded DNA antibodies, anti-glomerular basement membrane antibodies, and anti-neutrophil cytoplasmic antibodies, all of which were negative. The patient was started on hemodialysis. Fortunately he received a kidney transplant after 2 months of dialysis. He is currently recovering well, with a serum creatinine level of 117 μmol/L and an estimated glomerular filtration rate of 79.6 ml/min/1.73 mm². Whole exome sequencing was performed to further investigate the cause of his illness, and segregation was validated via Sanger sequencing in him and his family. Written informed consent was obtained from all family members.

METHODS

Blood samples were collected and DNA was extracted via a DNeasy Blood Kit (Qiagen, catalogue number 69504). DNA of the human exon region was highly enriched via an Agilent SureSelectHuman All ExonV6 Kit, and whole exome sequencing was performed using the Illumina platform. Sanger sequencing was then used to validate the exome sequencing results, and the loci of variation were analyzed in accordance with the standards and guidelines for the interpretation of genetic variants developed by the American College of Medical Genetics and Genomics in 2015 (Richards et al., 2015).

| Table 1 | Clinical characteristics of the patient and his family. |
| Parameter | Proband | Father | Mother | Brother |
|------------|---------|--------|--------|---------|
| Age (years) | 15 | 50 | 48 | 12 |
| Height (cm) | 173 | 166 | 155 | 140 |
| Hemoglobin (g/L) | 69 | 151 | 143 | 139 |
| Serum albumin (g/L) | 42.1 | 38.9 | 43.9 | 47 |
| Scr (μmol/L) | 1,026.2 | 79 | 69 | 57 |
| eGFR (ml/min/1.73 m²) | 5.8 | 99.5 | 89.8 | 148.6 |
| Uric acid (mmol/L) | 382 | 335 | 207 | 298 |
| Urine protein (g/24 h) | 1.05 | 0.10 | 0.09 | 0.09 |
| U-RBC (HPF) | 5 | 0 | 0 | 0 |

Scr, serum creatinine; eGFR, estimated glomerular filtration rate; U-RBC/HPF, red blood cell per high power field in urine.
RESULTS

Laboratory results included urinary protein 1.05 g/24 h, urine erythrocyte count 5/high-power field, serum creatinine 1.026.2 μmol/L, and an estimated glomerular filtration rate of 5.8 ml/min/1.73 mm². Extrarenal features mainly manifested as hypertension and moderate anemia. His parents (III-1 and III-2 in Figure 1) and younger brother (IV-3 in Figure 1) were in good health, and the results of their medical examinations were normal (Table 1). Mutational analysis of the inversin gene (INVS) identified a homozygous mutation in exon 15:c.1909C > T, with a predicted coding sequence change of p. Gln637Ter. A cytosine to thymidine substitution at nucleotide 1909 is predicted to result in glutamine at amino acid 637 being replaced by a stop codon, prematurely terminating the protein (Figure 2). According to the classification system of the American College of Medical Genetics and Genomics the INVS mutation identified is strongly pathogenic (PVS1+PM2+P3+P5). The presence of this mutation was then assessed in his immediate family members by sequencing the corresponding site via the Sanger method. The presence of the heterozygous pathogenic variant c.1909C > T in exons 15 of INVS was detected that is predicted to cause premature truncation of the inversin protein. Ultrasonography of the kidneys revealed hyperechogenic diffuse lesions, and the kidneys were shrunken. Clinically, the opportunity for renal pathological biopsy had been lost, so it was difficult to identify the etiology of his condition. Considering the consanguineous marriage of his parents, and that his older brother had died of ESRD, exome sequencing was performed in his entire immediate family. A novel and strongly pathogenic mutation was identified in the INVS gene, which may be associated with the ESRD in this patient. Inversin, the protein defective in NPHP2 patients, localizes to the primary cilia of renal epithelial cells and has been identified in patients with the infantile form of NPHP (Eley et al., 2004). Currently little is known about its specificity and potential interactions. Tory et al. (Tory et al., 2009) described five NPHP2 mutations that were recurrent in unrelated families, and the frequency of INVS mutations is reportedly up to 78% in patients who develop ESRD before 2 years of age. There are previous reports of patients with homozygous INVS mutations who developed renal failure before 5 years of age (Otto et al., 2008; O’Toole et al., 2006). Bellavia et al. (Bellavia et al., 2010) also described a patient with an INVS mutation who developed ESRD at the age of 11. Previous studies have demonstrated that NPHP2 mutations can cause ESRD, mainly the infantile-onset NPHP2 form. The age at which previously described patients progressed to ESRD differed from that of the current patient. The novel homozygous gene mutation identified in the current Chinese

DISCUSSION

NPHP is an uncommon autosomal recessive renal tubular interstitial disease known to cause renal ciliopathy. It is divided into infantile, juvenile, and adolescent forms based on respective median ages of onset and progression to ESRD of 1, 13, and 19 years. It is reportedly a common genetic condition that leads to ESRD in children and adolescents at the early stage of disease, characterized by polyuria, polydipsia, hypertension, and anemia (Hildebrandt et al., 2004). Currently little is known about its specificity and potential interactions. Tory et al. (Tory et al., 2009) described five NPHP2 mutations that were recurrent in unrelated families, and the frequency of INVS mutations is reportedly up to 78% in patients who develop ESRD before 2 years of age. There are previous reports of patients with homozygous INVS mutations who developed renal failure before 5 years of age (Otto et al., 2008; O’Toole et al., 2006). Bellavia et al. (Bellavia et al., 2010) also described a patient with an INVS mutation who developed ESRD at the age of 11. Previous studies have demonstrated that NPHP2 mutations can cause ESRD, mainly the infantile-onset NPHP2 form. The age at which previously described patients progressed to ESRD differed from that of the current patient. The novel homozygous gene mutation identified in the current Chinese
patient with the adolescent form of NPHP2 has not been previously reported, thus this report adds to the *NPHP2* variation spectrum.

There is currently a lack of effective therapy for NPHP2, so close long-term follow-up and supportive care strategies focused on attenuating the progression of renal impairment are generally utilized, sometimes in conjunction with alternative therapies. Kidney transplantation is preferred over replacement therapy when NPHP2 develops to ESRD. NPHP2 is a rare genetically heterogeneous disease accompanied by insidious clinical features and nonspecific manifestations, and it is easily misdiagnosed. Most patients have progressed to stage 5 chronic kidney disease by the time NPHP is diagnosed, and a kidney biopsy is not feasible due to the small size of the kidneys. It is thus very important to acquire family history, growth history, and other information for the diagnosis of NPHP. The possibility of undescribed pathogenic genes should also be fully considered in younger patients presenting with kidney disease.

The present case demonstrates that the clinical diagnosis of atypical NPHP2 is difficult. We have reported the first severe phenotype in a Chinese NPHP2 patient with

![FIGURE 3 | Sanger sequencing of the INVS gene. The patient was identified with 1-bp substitution; (C) C1909T, NM_001318382, Chr9:103059299 (A). His parents and younger brother are heterozygous carriers (B–D), and the arrows show the position of the mutation.](image-url)
adolescent-onset ESRD, and a novel INVS mutation was detected via whole exome sequencing. The case serves as a reminder to clinicians that precise medical genetic diagnostics are needed in cases of strongly suspected genetic diseases, especially in patients with a family history of consanguineous marriage.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/Supplementary Material, further inquiries can be directed to the corresponding authors.

REFERENCES

Bellavia, S., Dahan, K., Terryn, S., Cosyns, J.-P., Devuyst, O., and Pirson, Y. (2010). A Homozygous Mutation in INVS Causing Juvenile Nephronophthisis with Abnormal Reactivity of the Wnt/beta-Catenin Pathway. *Nephrol. Dial. Transpl.* 25, 4097–4102. doi:10.1093/ndt/gfq519

Eley, L., Turnpenny, L., Yates, L. M., Craighead, A. S., Morgan, D., Whistler, C., et al. (2004). A Perspective on Inversin. *Cell Biol Int* 28, 119–124. doi:10.1016/j.cbi.2003.11.009

Hildebrandt, F., Attanasio, M., and Otto, E. (2009). Nephronophthisis: Disease Mechanisms of a Ciliopathy. *JASN* 20, 23–35. doi:10.1681/asn.2008050456

Hildebrandt, F., Otto, E., Rensing, C., Nothwang, H. G., Vollmer, M., Adolphs, J., et al. (1997). A Novel Gene Encoding an SH3 Domain Protein Is Mutated in Nephronophthisis Type 1. *Nat. Genet.* 17, 149–153. doi:10.1038/ng1097-149

Hildebrandt, F., Strahm, B., Nothwang, H.-G., Gretz, N., Schnieders, B., Singh-Sawhney, L., et al. Members of the APN Study Group (1997). Molecular Genetic Identification of Families with Juvenile Nephronophthisis Type 1: Rate of Progression to Renal Failure. *Kidney Int.* 51, 261–269. doi:10.1038/ki.1997.31

König, J., Kranz, B., König, S., Schlingmann, K. P., Titierni, A., Tönshoff, B., et al. (2017). Gesellschaft für Pädiatrische Nephrologie (GPN). Phenotypic spectrum of children with nephronophthisis and related ciliopathies. *Clin. J. Am. Soc. Nephrol.* 12, 1974–1983.

Luo, F., and Tao, Y. H. (2018). Nephronophthisis: A Review of Genotype-Phenotype Correlation. *Nephrology* 23, 904–911. doi:10.1111/nep.13393

O’Toole, J. F., Otto, E. A., Frishberg, Y., and Hildebrandt, F. (2006). Retinitis Pigmentosa and Renal Failure in a Patient with Mutations in INVS. *Nephrol. Dial. Transpl.* 21, 1989–1991. doi:10.1039/n60088

Otto, E. A., Helou, J., Allen, S. J., O’Toole, J. F., Wise, E. L., Ashraf, S., et al. (2008). Mutation Analysis in Nephronophthisis Using a Combined Approach of Homozygosity Mapping, CEL I Endonuclease Cleavage, and Direct Sequencing. *Hum. Mutat.* 29, 418–426. doi:10.1002/humu.20669

Otto, E. A., Schermer, B., Obara, T., O’Toole, J. F., Hiller, K. S., Mueller, A. M., et al. (2003). Mutations in INVS Encoding Inversin Cause Nephronophthisis Type 2, Linking Renal Cystic Disease to the Function of Primary Cilia and Left-Right axis Determination. *Nat. Genet.* 34, 413–420. doi:10.1038/ng1217

Richards, S., Aziz, N., Bale, S., Bick, D., Das, S., Gastier-Foster, J., et al. (2015). Standards and Guidelines for the Interpretation of Sequence Variants: a Joint Consensus Recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet. Med.* 17, 405–424. doi:10.1038/gim.2015.30

Srivastava, S., Molinari, E., Raman, S., and Sayer, J. A. (2017). Many Genes-One Disease? Genetics of Nephronophthisis (NPHP) and NPHP-Associated Disorders. *Front. Pediatr.* 5, 287. doi:10.3389/fped.2017.00287

Srivastava, S., Molinari, E., Raman, S., and Sayer, J. A. (2018). Many Genes-One Disease? Genetics of Nephronophthisis (NPHP) and NPHP-Associated Disorders. *Front. Pediatr.* 5, 287. doi:10.3389/fped.2017.00287

Tory, K., Rousseau-Rouvière, C., Gubler, M.-C., Morinière, V., Pawtowski, A., Becker, C., et al. (2009). Mutations of NPHP2 and NPHP3 in Infantile Nephronophthisis. *Kidney Int.* 75, 839–847. doi:10.1038/ki.2008.662

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher’s Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Zhong, Yan, Fang, Dong, Tan, Xie, Hu, Zhang and Qin. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

ETHICS STATEMENT

Written informed consent was obtained from the individual(s) for the publication of any potentially identifiable images or data included in this article.

AUTHOR CONTRIBUTIONS

JX, XY and QW designed the study and critically revised the work. ZZ and SZ performed the experiments, collected the data, and wrote the manuscript. ZZ, JT, ZF and SZ performed the statistical analyses. YD and LH collected blood samples. All authors have read and approved the final manuscript.