REGIONAL LOCATION OF T CELL RECEPTOR GENE Tiα ON HUMAN CHROMOSOME 14

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The human T cell receptor is a molecular complex consisting of a clonotypic, disulfide-linked heterodimer, termed Ti, and three noncovalently associated monomorphic T3 components (1). The Ti molecule that presumably serves as the binding site for nominal antigen and the major histocompatibility complex (MHC) is composed of a 49–54 kilodalton (kD) α subunit and a 43 kD β subunit (1). In contrast, the 20–25 kD T3 proteins are likely involved in signal transduction after ligand binding to Ti (1). Sequence analysis at both the protein and DNA level indicates that the Ti α and β subunits are comprised of immunoglobulin-like domains and that each contains variable, joining, and constant (V, J, C) region encoding segments that rearrange during T lineage ontogeny (1).

Recent chromosomal mapping studies (2–4) localized the human T cell receptor β chain gene (Tiβ) to human chromosome 7, although its regional position remains unclear. Here, we demonstrate that human chromosome region 14pter>q21 contains the human Tiα gene. This is of particular cytogenetic interest, since chromosomal inversions and translocations involving this region have recently been observed in human T cell malignancies (5). Furthermore, the apparent lack of genetic linkage between the Tiα gene and the IgH cluster in humans can now be explained by the great physical distance between these loci on human chromosome 14.

Materials and Methods

Cell hybrid analysis (6) and blot hybridization experiments (3, 6) followed established methods except that the electro-eluted insert was labeled by the calf thymus primer protocol (7).

Isolation of Tiα REX cDNA Clones. Based on the previously determined amino acid sequence of the Tiα REX subunit, we have synthesized an oligonucleotide of 23 bases in length with a complexity of 256 (1). This oligonucleotide pool was used to screen the previously described REX lambda gt10 cDNA library (1). Positives were isolated and inserts subcloned in pBR322 (paREX) (8). DNA sequence analysis of paREX indicated that it encoded a portion of the V, J, and C regions of the REX human Tiα subunit (~400 nucleotides long).
FIGURE 1. Hybridization of paREX Tia-specific insert to nitrocellulose filters containing marker, genomic control, and mouse-human somatic cell hybrid DNAs. BamHI-cleaved genomic and cell hybrid DNAs were separated by gel electrophoresis, transferred to nitrocellulose filters, hybridized, and washed as reported (3, 6). (Lanes 1 and 12) λHindIII markers, (2 and 13) blanks, (3 and 14) mouse A9, (4 and 15) human placenta, (5) IIIMA9-6.1, (6) 5IIIMA9-6.1 SR3, (7) 55-87-c10, (8) 41pT2A, (9) WAV17, (10) BDA 14b25, (11) BDA 10a3, (16) FRY 4.A+ SEG, (17) AHA 16e, (18) AHA 16e6, (19) AHA 3d2-2, (20) AHA 3d2-3, (21) AHA 16e3. The size of marker DNA fragments is given to the left in kilobases.

Results and Discussion

Three major human Tia-specific fragments were detected at molecular weights of 5.5, 4.1, and 3.8 kilobases (kb) (Fig. 1, lanes 4 and 15). The mouse genomic signal (Fig. 1, lanes 3 and 14) showed bands of 10.0, 6.0, 3.1, and 2.65 kb. Each cell hybrid DNA contains mouse bands; however, the human signal segregates, with some hybrids lacking (lanes 5, 7, 9, 11, 19, 20), and some containing all three human bands. Co-segregation of the human bands indicated they originated from the same chromosome.

Table I shows the human chromosome complement of the hybrids as reported (3, 6). Each human chromosome except chromosome 14 is ruled out as the site of Tia by two or more hybrid DNAs. Hybrid 5IIIMA9-6.1-SR3, containing chromosomes 3, 4, 14, and X, contains the Tia signal, while hybrid IIIMA9-6.1 lacks human Tia and human chromosome 14 (Fig. 1), suggesting that human chromosome 14 contains Tia.

The human parent of BDA hybrids bears a translocation involving chromosomes 5 and 14 with the karyotype 46,XY,t(5;14)(5pter>5p14::5q21>5q14::14q21>14qter; 14pter>14q21::5p14>5pter). Markers NP at 14q13.1 (9), c-fos at 14q21>31 (6), and D14S1 at 14q32.1>32.2 (10), allowed identification of both translocation products (Fig. 2).

Cell hybrid BDA 10a3 contains D14S1 and c-fos, but not NP (3). Only the larger translocation product was present and this hybrid lacked Tia (Fig. 1, lane 11). Further, BDA 14b25 lacks c-fos, D14S1, and normal chromosome 14 (by cytogenetics), but expresses NP (6). Thus, normal chromosome 14 and the larger translocation product were absent but the shorter translocation product was
TABLE I
Human Chromosome Content of Somatic Cell Hybrids Used in Mapping the Human Tia Gene to Human Chromosomal Region 14pter>q21

| Cell hybrid         | Human chromosomes present* | Human-specific hybridization signal of cell hybrid DNA probed with pAREX insert: |
|---------------------|----------------------------|---------------------------------------------------------------------------------|
|                     |                            | 5.5 kb | 4.1 kb | 3.8 kb |
| IIIMA9-6.1          | 3, 4, X                    | +      | +      | +      |
| SIIMA9-6.1-SR3      | 3, 4, 14, X                | +      | +      | +      |
| 53-87-8c1.10        | 7                          | -      | -      | -      |
| 41pT22A             | 3, 6, 14, 15, 18, 19, 22, X| +      | +      | +      |
| BDA 14b25           | 1, 4, 7, 14, X             | +      | +      | +      |
| BDA 10a3            | 2, 5, 6, 8, 9, 10, 12, 13, 14, 16, 17, 20, 22, X | -      | -      | -      |
| FRY 4.A + SEG       | 1-4, 6-8, 10-12, 14, 15, 18, 21, 22, X | +      | +      | +      |
| AHA 16c             | 10-14, 17-21, X            | +      | +      | +      |
| AHA 16c6            | 1-3, 10, 11, 13, 14, 16, 19, 20, X | +      | +      | +      |
| AHA 3d2-2           | 1, 15                      | -      | -      | -      |
| AHA 3d2-3           | 3, 4, 8, 11, 12, 18, 19    | -      | -      | -      |
| AHA 16e-3           | 1, 3, 10-14, 19, 21, X     | +      | +      | +      |

* Cell hybrids were analyzed for human chromosome content by cytogenetic, isozyme, and DNA markers as reported (6).
  1 BDA 14b25 expressed the NP marker but lacked normal human chromosome 14 by cytogenetic analysis.
  2 BDA 10a3 did not express NP but contained distal human chromosome 14 DNA markers D14S1 and c-fos (6).

FIGURE 2. Idiogram of human chromosome 14 showing chromosomal location of markers used and the region found to contain the human Tia gene. NP, nucleoside phosphorylase isozyme marker (23); c-fos, human DNA sequence with homology to the transforming gene of Finkel-Biskis-Jinkins (FBJ)-murine sarcoma virus (6); D14S1, the pAW101 restriction fragment length polymorphic DNA marker (10); IgH, the immunoglobulin heavy chain cluster (11).

The human Tia gene is mapped to the region indicated (dotted line). The gene order for these and other previously mapped human chromosome 14 genes (9) is: [rDNA-centromere-NP]-(Tia)]-(c-fos,a-1-antitrypsin)-(D14S1, IgH)].

This hybrid contained human Tia (Fig. 1, lane 10). Tia segregated in BDA hybrids with the shorter translocation product so we conclude that the gene lies in region 14pter>q21.

Since Tiiβ is encoded by a chromosome 7 gene (2-4), the two Tii subunits are encoded by genes on different human chromosomes. Human IgH lies in region
14q32.3 (11) at great physical distance from the Tia gene at 14pter>q21. Given this great intergenic distance, it is not surprising that IgH and Tia are not genetically linked. Nevertheless, the presence of truncated Tia transcripts in Epstein Barr virus-transformed B lymphoblastoid lines and m transcripts in T cell lines (8) suggests that chromatin structure in these two regions of chromosome 14 may be altered in both differentiated cell types, permitting a common T and B recombinase to enter and interact with specific recognition sites.

Repeated observations of a chromosome translocation in human Burkitt lymphoma (12), followed by a subchromosomal mapping of c-myc (13) and IgH (14), led directly to the observation of underlying genetic events at the nucleotide level (15). Similarly, study of the chronic myelocytic leukemia (CML) translocation in the context of c-abl and Ig~ led to observation of molecular genetic events specific for CML (16).

With this in mind, we note that cytogenetically fragile regions of human chromosome 14 occur in normal individuals at 14q13 and 14q24.11 (17), and in malignant lymphocytes near 14q11 and 14q32 (5). The role of Tia in these conditions is unknown. Four patients with T11+ T cell acute lymphoblastic leukemia (ALL) have common disease-specific chromosome translocations with 14q13 breakpoints (18), within the region that we show contains Tia. Chromosome 14 aberrations in this region are also found in T cell lymphoma and T cell CLL (5). Based on our mapping results, these diseases are obvious subjects for study with regard to Tia.

Human B cell–related immunoglobulin loci have been mapped (11, 14); their map location in the mouse is also known (19, 20). Among T cell genes, the human T cell receptor β chain is clearly on human chromosome 7 (2–4), although its regional position is unclear. The gene encoding the Tα subunit is in human region 14pter>21. Analogous murine genes lie in proximal chromosome 6 (Tiβ) (2, 21) and 14 (Tia) (22).

The assignment of a locus in one species may predict its location in others for conserved linkage groups. Tia (human chromosome 14) predicts a mouse chromosome 14 assignment. In the mouse, the Tia-NP synteny parallels that in human chromosome 14. The human NP locus at 14q13.1 (23) is well within the region containing Tia, 14pter>q21. Thus, the syntenic region Tia-NP may be conserved between mice and humans. Further, human c-fos is at 14q21>31 (6) while mouse c-fos is on murine chromosome 12 (24), and thus syntetic with murine IgH (19). Interestingly, the Tia-NP syntenic region consists a block independent of the c-fos-IgH syntenic group. A possible manifestation of Tia-NP synteny may be the T, but not B, cell immunodeficiency in children with NP deficiency (25).

Summary

The chromosomal location of Tia was determined by hybridization of a radiolabeled cDNA for the α chain of human T cell receptor with 12 human × mouse cell hybrid DNAs cleaved with BamHI. Seven hybrids contained human Tia, while the remaining five lacked it. Only human chromosome 14 matched the distribution of human Tia signal across the mapping panel. Hybrids segregating a chromosome 14 translocation were used to demonstrate that Tia is in the region 14pter>14q21. Thus, the α and β chain genes that contribute structural components to the Ti moiety of the human T cell receptor lie on
different chromosomes. In humans, the immunoglobulin heavy chain locus and $T\kappa$ are in different regions of chromosome 14, with $T\kappa$ more proximal and the immunoglobulin heavy chain locus more distal.

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