Progesterone receptor variation and risk of ovarian cancer is limited to the invasive endometrioid subtype: results from the ovarian cancer association consortium pooled analysis

There is evidence that progesterone plays a role in the aetiology of invasive epithelial ovarian cancer. Therefore, genes involved in pathways that regulate progesterone may be candidates for susceptibility to this disease. Previous studies have suggested that genetic variants in the progesterone receptor gene (PGR) may be associated with ovarian cancer risk, although results have been inconsistent. We have established an international consortium to pool resources and data from many ovarian cancer case–control studies in an effort to identify variants that influence risk. In this study, three PGR single nucleotide polymorphisms (SNPs), for which previous data have suggested they affect ovarian cancer risk, were examined. These were the 331 C/T variant (rs10895068), PROGINS (rs1042838), and a 725 cases; OR = 0.80, 95% CI 0.62–1.04, P = 0.100). These data suggest that while these three variants in the PGR are not associated with ovarian cancer overall, the PROGINS variant may play a modest role in risk of endometrioid ovarian cancer.

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Several lines of evidence support a role for progesterone in the aetiology of ovarian cancer (Risch, 1998). Epidemiological studies have consistently shown a significant protective effect of parity.
The protective effect increases steadily with each birth and pregnancy is associated with high progesterone levels (Hartge et al, 1989; Cooper et al, 1999; Titus-Ernstoff et al, 2001; Whitteman et al, 2003; Pike et al, 2004). In the third trimester, progesterone levels are some 10–15 times higher than in the luteal phase of the normal menstrual cycle. Oral contraceptives are also protective against ovarian cancer (Study, 1987; Rosenblatt et al, 1992, 1994; Ness et al, 2000; Royar et al, 2001; Schildkraut et al, 2002) and use of progestin-containing oral contraceptives increases average circulating progesterone levels to 9.2 ng·mL⁻¹ compared to ~3.5 ng·mL⁻¹ during the normal menstrual cycle (Norman and Litwack, 1997). The protective effect of oral contraceptives per month of use is less than the protection from births, in line with the contribution of progesterone. There is also some evidence that oral contraceptives with higher progesterin content afford more protection against ovarian cancer (Schildkraut et al, 2002).

Animal models and in vitro data also suggest that progesterone has a significant influence on the ovary and on ovarian cancer. Studies in macaques suggest an apoptotic effect of progestins on the surface of the ovary (Rodriguez et al, 1998). In vitro treatment of both benign and malignant ovarian tumour cells with progestins results in an antiproliferative response (Zhou et al, 2002).

Progesterone binds to the progesterone receptor (PR) to initiate signalling. Two progesterone receptor isoforms (PR-A, PR-B) are encoded by a single gene (PGR). Except for a 164 amino-acid sequence at the N-terminal end of PR-B that is absent from PR-A, the PR isoforms are identical but their actions are divergent (Kastner et al, 1990). PR-B acts as a transcription activator whereas PR-A inhibits PR-B (and other members of the nuclear receptor superfamily) (Vegeto et al, 1993).

The PGR has long been hypothesised as a candidate gene for ovarian cancer susceptibility and its variation has been widely studied. Originally, an ALU in intron 7 named PROGINS was identified and found to be associated with increased risk of ovarian cancer (McKenna et al, 1995; Rowe et al, 1995). Subsequent characterisation of the coding region of the gene identified a non-synonymous single nucleotide polymorphism (SNP) in exon 4 and a synonymous SNP in exon 5 that were in perfect linkage disequilibrium with the PROGINS (De Vivo et al, 2002). The PROGINS (or variants in which it is in perfect linkage disequilibrium) has been studied by many groups in relation to ovarian cancer risk. The results are, however, equivocal (McKenna et al, 1995; Manolitsas et al, 1997; Lancaster et al, 1998, 2003; Spurtle et al, 2001; Tong et al, 2001; Agoulnik et al, 2004; Pearce et al, 2005; Terry et al, 2005; Romano et al, 2006). Pearce et al (2005), suggested that a variant 3′ of the PGR (rs608995), in partial linkage disequilibrium with the PROGINS, might be a better marker of ovarian cancer risk, but this has not been confirmed by other investigators.

In addition, a putative functional SNP, +331C/T (sometimes denoted as +331G/A), in the promoter region of the PGR that may affect the relative transcription of the PR-A and PR-B isoforms has been found to be associated with a reduced risk of ovarian cancer in studies from North Carolina and Australia (Berkhuck et al, 2004). This association was particularly strong among clear cell/endometrioid subtypes. However, Risch et al (2006) observed an increased risk of ovarian cancer associated with this SNP.

The inconsistent results with the PROGINS and the +331C/T SNP are not surprising. Genetic association studies are plagued by conflicting results that can be explained by heterogeneity across study populations as well as false-positive and -negative results. Lohmueller et al (2003) demonstrated that approximately two-thirds of genetic associations do not hold up on meta-analysis. Large sample sizes and pooling of data are therefore critical to evaluate the association between a phenotype and genetic variation with confidence.

To clarify the association between variation at the PGR locus and ovarian cancer risk, including histological subtype associa-

**Materials and Methods**

**Approval and consent**

All study participants provided written informed consent prior to the collection of biological samples or interview/clinical data. Each group involved in the OCAC has Institutional Review Board/ethics approval for this analysis and the University of Southern California and Duke University have Institutional Review Board approval to serve as data coordinating centres for the OCAC.

**Study populations**

The OCAC comprises investigators who collaborate on promising genetic associations by combining data from their individual ovarian cancer case–control studies. The participating groups for this pooled study are the Australian Cancer Study, (Merritt et al, 2008) the Australian Ovarian Cancer Study (Merritt et al, 2008), the Connecticut Ovary Study (CONN) (Risch et al, 2006), the Family Registry for Ovarian Cancer Study (Auranen et al, 2005; Song et al, 2006), the Hormones and Ovarian Cancer Prediction Study, the Danish Malignant Ovarian Cancer Study (MALOVA) (Auranen et al, 2005; Song et al, 2006), the Mayo Clinic Ovarian Cancer Case–Control Study (Sellors et al, 2005), the North Carolina Ovarian Cancer Study (Berkhuck et al, 2004), the New England-based Case–Control Study (NECC) (Terry et al, 2005), the Polish Ovarian Cancer Study (POCS) (García-Clossas et al, 2007), the UK SEARCH Ovarian Cancer Study (SEARCH) (Auranen et al, 2005; Song et al, 2006) and the USC/Los Angeles County Case–Control Studies of Ovarian Cancer (USC) (Pearce et al, 2005). Details of these studies have been published previously (Gayther et al, 2007); Table 1 shows the basic information for each study. The cases analysed here are restricted to women diagnosed with invasive epithelial ovarian cancer.

**Genotyping and quality control**

The three SNPs genotyped in this study were the +331C/T (rs10895068), PROGINS (measured by the exon 4 SNP rs1042838) and rs608995 (a variant 3′ of the PGR). The allele designations are based on the forward strand as given in the University of California at Santa Cruz genome browser.

All groups used the 5′ nuclease Taqman allelic discrimination assay (Taqman; Applied Biosystems, Foster City, CA, USA) to genotype samples with the exception of the Australian Cancer Study and Australian Ovarian Cancer Study, which used the iPLEX Sequenom MassArray system (Sequenom Inc., San Diego, CA, USA), CONN that used dot blotting (Risch et al, 2006), and Mayo Clinic Ovarian Cancer Case–Control Study that used Pyrosequencing for PROGINS and rs608995.

To confirm that laboratory to laboratory quality control was adequate, five SNPs were genotyped in the HAPMAPP01 panel of CEPH-Utah trios-standard plate provided by Coriell (http://locus.umdnj.edu/nigms/nigms cgi/?id = 2&query = HAPMAPP01). This 96-well plate contains 90 different DNA samples, five duplicate samples, and a negative template control. Genotyping call rates and concordance between studies were compared. Call rates for these five SNPs ranged from 96 to 99% and the concordance of results across the laboratories was >99%.
Clinical Studies

Table 1  Characteristics of the 12 case–control studies used in this analysis

| Study*   | Ascertainment                                                                 | Cases                  | Controls                | Participation rates |
|----------|-------------------------------------------------------------------------------|------------------------|-------------------------|---------------------|
| ACS, Australia | Cancer registries of New South Wales and Victoria; cases diagnosed July 2002–June 2005. | 111 91.0 59.8 Randomly selected from Commonwealth electoral roll. Frequency matched for age and geographical region | 156 95.8 55.2 | 684 97.4 58.2 Cases: 68% Controls: 47% |
| AOCs, Australia | Diagnosed from 2002 onwards; recruited through surgical treatment centres throughout Australia and cancer registries of Queensland, southern Australia and western Australia cases diagnosed 2002–2006. | 502 95.4 59.7 Randomly selected from Commonwealth electoral roll. Frequency matched for age and geographical region | 533 88.6 53.1 Cases: 69% Controls: 61% |
| CONN, USA | Rapid case ascertainment of consecutive cases identified from 30 Connecticut hospitals and through the Connecticut Tumour Registry between 1998 and 2003. | 365 90.7 59.1 HCPF (CMS) plus random-digtal dial identification from study area, frequency matched to cases on age group | 424 86.8 48.4 Cases: 75% Controls: 91% |
| FROC, USA | Consecutive cases diagnosed from 1997–2002 in Greater Bay Area Cancer Registry, San Francisco. | 324 87.3 50.8 Random-digtal dial identification from study area. Frequency matched to cases for race/ethnicity and 5-year age group | 152 94.7 56.1 Cases: 69% Controls: 81% |
| HOPE, USA | Variable source including physician offices, cancer registries and pathology databases from counties of western Pennsylvania, eastern Ohio and western New York. Incident cases (35–79 years) diagnosed 1994–1999 from municipalities of Copenhagen and Frederiksberg and surrounding counties. | 444 100.0 59.9 | 1221 100 56.8 Cases: 79% Controls: 67% |
| MALOVA, Denmark | 1994–1999 from municipalities of Copenhagen and Frederiksberg and surrounding counties. | 278 97.6 61.4 | 389 97.7 60.3 Cases: 84% Controls: 65% |
| MAYO, USA | Cases attending Mayo Clinic diagnosed from 2000 onwards, identified in a six-state surrounding region. | 610 83.0 56.8 | 843 81.5 54.4 Cases: 70% Controls: 63% |
| NCOCS, USA | Cases from 1999 onwards, identified from 48 counties within the region by rapid-case ascertainment. | 667 96.0 53.6 | 1011 96.6 50.5 Cases: 72% Controls: 69% |
| NECC, USA | Cases identified through hospital tumour boards and state cancer registries in New Hampshire and Massachusetts from 1992 to 2003. | 264 100.0 56.3 | 625 100 56.1 Cases: 71% Controls: 67% |
| POCS, Poland | Cases collected from the cities of Warsaw and Lodz, 2001–2003, by rapid ascertainment at participating hospitals. | 643 99.3 55.8 Selected from the EPIC-Norfolk cohort of 25,000 individuals aged 45–74, based in the same geographical regions as the cases. | 852 99.7 52.7 Cases: 67% Controls: 84% |
| SEARCH, UK | Cases <70 years from East Anglian, West Midlands and Trent regions of England. Prevalent cases diagnosed 1991–1998; incident cases diagnosed 1998 onwards. | 523 71.0 54.9 | 724 75.4 52.7 Cases: 73% Controls: 73% |

*See methods for full study name.

Hardy–Weinberg Equilibrium (HWE) was checked among controls by the racial/ethnic group. Data from one study (CONN) for two of the SNPs (PROGINS and rs608995) were excluded for gross deviations \( (P<10^{-4}) \) from HWE. The genotyping calls for studies with minor deviations from HWE \( (0.01<P<0.05) \) were examined to monitor the quality of the genotyping. There were no obvious reasons for deviation from HWE (e.g., genotyping irregularities), and therefore the minor deviations were assumed most likely due to chance. In addition, results were unchanged when excluding those studies with HWE \( P \)-values between 0.01 and 0.05 (data not shown). Concordance between duplicate samples was 100% across all studies for the three variants for all data included in these analyses.

Results were available for 12 groups for the +331C/T variant. The NECC study did not genotype rs608995. Results for the PROGINS and rs608995 were excluded for CONN due to significant deviations from HWE. Therefore, results were available for 11 studies for the PROGINS and 10 studies for the rs608995 3’ variant.

Statistical analysis

The variables available for this analysis were study, race/ethnicity (White, Latina, African-American), age, stage of disease (FIGO), histology (serous, mucinous, clear cell, and endometrioid), and time from diagnosis to blood collection (cases only).

Unconditional logistic regression was used to model the association between each SNP and risk of ovarian cancer stratified on study, age, and race/ethnicity. All single SNP models were log additive. Goodness of fit \( P \)-values were calculated to evaluate heterogeneity across the study populations. Statistical analyses were carried out using both SAS (Version 9, Cary, NC, USA) and
RESULTS

A total of 4788 invasive epithelial ovarian cancer cases and 7614 controls were available for the current analysis (Table 1). Overall, 92.0% of cases and 93.0% of controls were White and the mean ages were 56.7 and 54.3 years respectively. Information on stage at diagnosis was available on 73.7% of cases, the majority of which were FIGO stage III/IV (63.0%) and 55.5% had a serous histology (Supplementary Table 1).

Across the studies, the minor allele frequencies in White controls ranged from 4.6 to 7.3% for $+331\text{C/T (rs10895068)}$, 9.2 to 19.0% for the PROGINS (rs1042838) and 20.0 to 26.6% for the $30\text{A/T (rs608995); Supplementary Table 2.}$ The study-specific and summary effect estimates are shown in Figure 1 for all cases and endometrioid subtype associations.

There was no association with the $+331\text{C/T (rs10895068)}$ variant among all cases (per allele OR $= 1.00; 95\% \text{ CI } 0.89 – 1.13; P = 1.0$; Table 2a). In cell type-specific subgroup analyses, a suggestive association was observed with carrying a T allele and risk of endometrioid invasive ovarian cancers (per allele OR $= 0.80; 95\% \text{ CI } 0.62 – 1.04; P = 0.100$; Figure 1). Risk of clear cell ovarian cancer with this variant was reduced to a similar degree (OR $= 0.83; Table 2b$). No associations were observed between serous or mucinous subtypes and this allele (Table 2b).

No overall association was observed with risk of ovarian cancer and the PROGINS allele (rs1042838; OR $= 1.04; 95\% \text{ CI } 0.96 – 1.12; P = 0.38$; Table 2a). However, risk was statistically significantly elevated among endometrioid ovarian cancer cases (OR $= 1.17, 95\% \text{ CI } 1.01 – 1.36, P = 0.036$; Table 2b).

In a joint effects analysis, risk of endometrioid ovarian cancer associated with the PROGINS was observed only among non-carriers of the $+331$ minor allele (OR $= 1.22, 95\% \text{ CI } 1.01 – 1.46, P = 0.037$). Although not statistically significant, the protective effect of the $+331$ minor allele persisted among non-carriers of the PROGINS (OR $= 0.76, 95\% \text{ CI } 0.55 – 1.06, P = 0.11$) and carriers of the PROGINS (OR $= 0.79, 95\% \text{ CI } 0.40 – 1.57, P = 0.50$).

Table 2a Summary odds ratios (per allele) and 95% CI for the three PGR SNPs for all invasive cases among OCAC studies

| SNP                | Controls | All cases |
|--------------------|----------|-----------|
|                    | N        | N         | OR* (95% CI) | P       |
| $+331\text{C/T (rs10895068)}$ | 7338 | 4551 | 1.00 (0.89–1.13) | 1.0 |
| PROGINS C/A (rs1042838) | 6794 | 4124 | 1.04 (0.96–1.12) | 0.38 |
| rs608995 A/T        | 5796 | 3510 | 1.05 (0.98–1.13) | 0.17 |

CI = confidence interval; OR = odds ratio; OCAC = Ovarian Cancer Association Consortium; PGR = progesterone receptor gene; SNP = single nucleotide polymorphism. *All analyses stratified on study, race, and age.

Figure 1 Each panel shows the study-specific and summary odds ratios (boxes) and 95% confidence intervals (lines) for all cases and endometrioid subtype specific results for the three PGR SNPs. The size each box is proportionate to the number of subjects genotyped. See methods for full study names.
with regard to their functional effect. The T allele of the +331 variant (rs608995) and risk of ovarian cancer when all cases were considered (Table 2a). In subtype analysis, a borderline statistically significant association was observed between endometrioid cases and the rs608995 variant (OR = 1.14, 95% CI 0.99 – 1.31, P = 0.076), however, this effect was limited to individuals also carrying at least one copy of the PROGINS (data not shown).

**DISCUSSION**

Since the publication of the first paper examining the relationship between the PROGINS and ovarian cancer risk more than 10 years ago, there has been substantial interest in the role of the PGR in risk of this disease. We have evaluated three SNPs, +331C/T (rs10895068), PROGINS (rs1042838), and a +331 variant (rs608995), in the PGR in a pooled ovarian cancer dataset from 12 groups around the world and have found no overall role for this gene in disease risk.

The pooled analysis does provide statistically significant evidence of an association between the PROGINS and risk of invasive endometrioid ovarian cancer. The restriction of an association to this subtype only provides an explanation for the equivocal nature of the published results on the PROGINS and ovarian cancer risk, given that the proportion of endometrioid ovarian cancer cases likely varied by published study and typically accounts for no more than 15 – 20% of cases.

We also found suggestive evidence of an association between endometrioid ovarian cancer and the +331C/T variant (OR = 0.80, 95% CI 0.62 – 1.04, P = 0.100). As suggested by Berchuck et al (2004), combining endometrioid and clear cell histologies in which the effect is similar, resulted in a borderline statistically significant association (n = 1088 cases, OR = 0.81, 95% CI 0.65 – 1.01, P = 0.058).

Pearce et al (2005) had previously suggested that rs608995 may explain the PROGINS-ovarian cancer association, however, in this larger dataset in which the effect was restricted to endometrioid cases, this was not supported. When examining the joint effects of the PROGINS and rs608995, the OR for endometrioid ovarian cancer associated with the rs608995 minor allele was 0.79 (95% CI 0.59 – 1.07, P = 0.12) in the absence of the PROGINS allele. This suggests that the PROGINS allele or a marker in linkage disequilibrium with the PROGINS is responsible for the association and not the rs608995 variant.

Both the +331 variant and the PROGINS have been studied with regard to their functional effect. The T allele of the +331 331 favours an increase in the transcription of PR-B relative to PR-A (De Vivo et al, 2002); PR-B acts as a classic steroid receptor whereas PR-A acts as a repressor of both PR-B and other steroid receptors. PR-A therefore may lessen overall progesterone responsiveness through its repressive effect. Any variation which increases PR-B relative to PR-A may reduce risk of ovarian cancer by increasing exposure to the beneficial effects of progesterone. In a small study of 107 ovarian cancer cases, decreased risk of death was observed among cases positive for PRB (labelling index > 10) relative to cases negative for PRB (P = 0.037). However, this finding was amongst all cell types (Akahira et al, 2000). There is also a suggestion that the PROGINS allele as defined by the V660L exon five variant (as examined in the present study) decreases overall response to progesterone which would be consistent with an increased risk of disease associated with this variant (Romano et al, 2007).

In this collaborative effort, there were 4788 ovarian cancer cases, of which 766 (16.0%) were endometrioid tumours. With the samples sizes available in this current OCAC study, we had 80% power to detect odds ratios of 0.83 and 1.12 for the +331, and PROGINS variants, respectively for all cases using a log additive genetic model and a two-sided z of 0.05. Among endometrioid subtypes, we had 80% power to detect odds ratios of 0.67 and 1.25 for the +331 and PROGINS variants, respectively using a log additive genetic model and a two-sided z of 0.05. Although the power in the current OCAC study is still quite limited, it underscores the importance of collaborative efforts, as the largest individual OCAC study had only 124 endometrioid ovarian cancers. Thus the power of subgroups analyses is clear and will be enhanced in the future with continued patient accrual to existing studies and additional investigators contributing to OCAC studies.

Alternatively, the findings of an association with the +331C/T and PROGINS, variants with the endometrioid histology may simply be due to chance. By assigning priors of 0.05, 0.10, and 0.15, the resulting false positive report probabilities (Wacholder et al, 2004) are approximately 0.78, 0.62, and 0.52 for the +331 and PROGINS variants, respectively using a log additive genetic model and a two-sided z of 0.05. Although the power in the current OCAC study is still quite limited, it underscores the importance of collaborative efforts, as the largest individual OCAC study had only 124 endometrioid ovarian cancers. Thus they may represent false positive findings.

Our analysis is the largest report describing the association between ovarian cancer risk and variants in the PGR. However, there remain several limitations to the study. For example, it is possible that environmental modifiers, such as oral contraceptive use, may be important in refining the PGR ovarian cancer risk associations and such analyses are planned in the future. There are also weaknesses of this study. Firstly, there are variable participation rates for cases between studies (Table 1). If any or all of the variants analysed is related to survival, then the low participation rates among cases might be expected to influence the results. Efforts to evaluate this include the analyses of data stratified by FIGO stage and time from diagnosis to blood collection. None of the results differed significantly when conducting these analyses. Second, as is the nature of collaborative projects, each study had different level of pathology review and random misclassification cannot be ruled out, which would bias results towards the null in histologic-specific analyses suggesting that our results may be attenuated. Lastly, while we evaluated the best PGR candidate variants suggested by the literature, it remains possible that other, as yet unidentified variants at the locus, influence ovarian cancer risk.

Also, we observed significant heterogeneity of effect for the PROGINS allele and risk of ovarian cancer overall. Evaluation of the heterogeneity by removing one study at a time revealed that the NECC study population had a significantly different odds ratio relative to the other studies (P = 0.037).
endometrioid subtype is warranted to provide insight into the mechanisms underlying disease aetiology.

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