Light Effect on Water Viscosity: Implication for ATP Biosynthesis

Andrei P. Sommer, Mike Kh. Haddad & Hans-Jörg Fecht

Previous work assumed that ATP synthase, the smallest known rotary motor in nature, operates at 100% efficiency. Calculations which arrive to this result assume that the water viscosity inside mitochondria is constant and corresponds to that of bulk water. In our opinion this assumption is not satisfactory for two reasons: (1) There is evidence that the water in mitochondria prevails to 100% as interfacial water. (2) Laboratory experiments which explore the properties of interfacial water suggest viscosities which exceed those of bulk water, specifically at hydrophilic interfaces. Here, we wish to suggest a physicochemical mechanism which assumes intramitochondrial water viscosity gradients and consistently explains two cellular responses: The decrease and increase in ATP synthesis in response to reactive oxygen species and non-destructive levels of near-infrared (NIR) laser light, respectively. The mechanism is derived from the results of a new experimental method, which combines the technique of nanoindentation with the modulation of interfacial water layers by laser irradiation. Results, including the elucidation of the principle of light-induced ATP production, are expected to have broad implications in all fields of medicine.

Institute of Micro and Nanomaterials, University of Ulm, 89081 Ulm, Germany. Correspondence and requests for materials should be addressed to A.P.S. (email: andrei.sommer@uni-ulm.de)

Received: 11 February 2015
Accepted: 09 June 2015
Published: 08 July 2015
result would already be sufficient to challenge the utilization of the viscosity of bulk water in the models and simulations used to assess the efficiency of the mitochondrial nanomotor.

The principal function of the mitochondrial ATP synthase is the synthesis of ATP — the primary energy carrier in cells. The root cause of the synthesis is the translocation of protons, which are turning the rotor19,20. The protons are driven across the inner membrane of mitochondria into the mitochondrial matrix by the energy of the proton gradient, the transmembrane proton-motive force21. It is thus plausible to assume that processes with direct impact on the dynamics of the nanomotor will affect ATP production. For instance, reactive oxygen species (ROS) induced oxidative stress was shown to cause depletion of ATP levels in mitochondria22,23, apparently a mechanism, which works in both mammalian and plant cells24. The question arises: What is the intrinsic mechanism by which ATP is depleted by elevated levels of ROS? Coincidentally, both compounds ATP and ROS are produced in mitochondria. Therefore, from the premise that bursts of ROS will accentuate the hydrophilic nature of the intramitochondrial space (due to the oxygen), the most plausible answer is that ROS enhances hydrophilicity, and thereby the viscous friction between surfaces moving relative to each other. Hence, pathological conditions that are triggering prolonged bursts of ROS will contribute to a transient increase in the viscosity of the interfacial water layers bound to exposed intramitochondrial surfaces. An increase in interfacial viscosity, concomitant with an increase in viscous friction, can only manifest itself in a reduced performance of the rotary motor, which explains the drop in ATP production. Indeed, molecular dynamics simulations predicted that interfacial viscosity would increase with hydrophilicity25. Recently, the correlation between viscosity and hydrophilicity received further confirmation by atomic force microscopy: The measured interfacial viscous forces were larger for materials with smaller contact angles, i.e., more hydrophilic surfaces, and vice versa15.

If indeed, a raise in interfacial viscosity impacts the rotation of the mitochondrial nanomotor, for instance, under conditions of prolonged oxidative stress, which are inescapably present during in vitro experiments, it can be reasonably assumed that a reduction of potentially elevated viscosity levels will manifest itself in an increase in ATP production. Earlier, we showed that the structure of nanoscopic interfacial water layers (about 2–3 monolayers)26 can be modulated with 670 nm laser light applied at moderate intensities as low as 50 W · m−2. Modulation includes volume expansion28 and viscosity reduction, specifically on hydrophilic surfaces, which are known to promote high viscosities14. Importantly, modulation effects were not restricted to 670 nm light. Comparable modulation effects were realized with other laser wavelengths, for instance, 633 nm, applied at 400 W · m−226. The expansion effect was exploited in vitro to force cancer cells to uptake various cytostatic drugs29, and predicted to be instrumental in the release of drugs from permeable nanovesicles, a prediction that has recently been confirmed experimentally30. Summarizing, there is observational evidence for the prevalence of intramitochondrial water viscosity gradients, and for a decrease in increase in ATP synthesis in response to ROS and laser light31, respectively. If indeed the root cause of the variation in ATP production is a variability in intramitochondrial viscous friction, then the same light which stimulated ATP production in cells should also reduce the viscous friction in subnanometer gaps.

Results and Discussion

Probing nanoscopic interfacial water layers by nanoindentation and NIR laser light. Here, we report on laboratory experiments focusing on nanoscopic interfacial water layers which prevail on hydrophilic surfaces and are confined in subnanometer gaps, and their light tunability. Similar water layers are expected to determine the efficiency of the rotary nanomotor (Fig. 1A). For their exploration we used a nanoindenter. We recorded the force required to penetrate 1 μm into different model substrates. We probed both hydrophilic and hydrophobic substrate materials, in dark and during irradiation of the tip/substrate contact zone with 670 nm laser light. Figure 1B illustrates the principle of the experiment. Hydrophilic materials comprised aluminum, zinc, copper and gold, while polymers, for instance, polystyrene, served as hydrophobic materials. Figure 2 presents the mean value curves of 2 × 10 representative nanoindentation measurements performed on a hydrophilic substrate (single crystal gold), irradiated with 670 nm laser light, and without laser irradiation. In both cases the measurements were performed in a closed box at room temperature and constant relative humidity of 67%. As can be clearly seen in Fig. 2 the laser light contributed to a reduction in the load required to penetrate the hydrophilic sample (assumed to carry a viscous film of interfacial water which is tunable by 670 nm laser light via collective interaction of photons with transiently immobilized interfacial water molecules)26–28. The drop in the load with laser light was estimated to be around 72% compared to the non-irradiated sample. The results for all tested metals are summarized in Table 1. Furthermore, at a relative humidity threshold of 48%, the effect disappeared, indicating that the amount of water confined at the tip/cavity interface was too small to build up a substantial viscous layer, reflecting a correlation with environmental humidity. On the basis of the humidity dependence we can safely exclude that the cause of the observed effect was the heating of the diamond tip and/or substrate. Notably, when we probed the hydrophilic species, the laser light had practically no effect. The results are in agreement with the observational evidence that elevated interfacial viscosity levels are present on hydrophilic, but not on hydrophobic surfaces. If we consider the classical friction components (adhesion, ploughing and asperity deformation) as well as the localized plastic deformation which presumably occur during the penetration of the diamond tip into the substrates, it is obvious that none of these factors can be modified by the low intensity laser light.
Figure 1. Mitochondrial nanomotor (A). During ATP synthesis, the rotor turns about 9000 times per minute. Artist's view of the principle of light-tuned nanoindentation. Blue spheres stand for water molecules forming the nanoscopic water layers confined in the space between the diamond tip and nanoindentation imprint (B). Reprinted by permission from Macmillan Publishers Ltd: [NATURE]39, copyright (2004).

Figure 2. Nanoindentation loading curves (average curves, n = 10) for hydrophilic sample (single crystal gold), with and without 670 nm laser. The force required for the diamond tip to penetrate ~100 nm into the metal is less when the tip/substrate contact zone is irradiated. Insets display the corresponding measurements shown as individual curves. Representative curves for single crystal aluminum have been presented during the International Conference on Laser Applications in Life Sciences, 201440.
Thus, it seems reasonable to assume that the origin of the relatively low force required to penetrate the first 100 nanometers into the hydrophilic samples, as depicted in Fig. 2, is a reduction of the viscous friction in the tip/cavity interface by the laser light. Going back to mitochondria, it is now tempting to assume that the nanomotor efficiency (ATP productivity) can be tuned with biologically tolerated intensities of red to NIR laser light. This perspective receives justification from the experimental side: Previously, it was reported that red laser light (632.8 nm (power 15 mW, fluence 5 J·cm⁻²)) changed the energy metabolism in mitochondria irradiated in vitro, and caused an increase in ATP synthesis. It was suggested that the extra ATP synthesis is directly produced by a laser-induced extra proton-motive force³¹. Remarkably, comparable levels of 670 nm laser light (power 33 mW, fluence 1 J·cm⁻²) increased both the proliferation and ATP production of cells in vitro³². We believe that the cause which gives the extra ATP is a reduction of interfacial viscosity within and/or around the mitochondrial nanomotor. The high torque of the nanomotor is best illustrated by its capacity to rotate objects several hundred times as large as the motor itself against the viscous friction of water. Torque is transferred to the ATP-producing part by the unit γ (Fig. 1). It is instructive to compare the mitochondrial rotary motor with a rotating cylinder viscometer. The measuring principle of the latter is based on the proportionality between the viscosity (η) and torque (T) \( T = \eta \cdot C \cdot \omega \), where C is a constant specific to the instrument and \( \omega \) stands for the angular velocity of the rotating cylinder². A reduction in viscosity by the laser light will increase the efficiency of the nanomotor, reflected by an increase in ATP production³¹.

### NIR laser light upregulates ATP synthesis.

Interestingly, equal levels of red to near-infrared (NIR) light are used routinely in clinical praxis to accelerate the healing of complicated wounds³³, to treat pain and inflammatory processes³⁵. Coincidentally, the therapeutic use of NIR light plays an increasingly important role in aeronautic medicine. As the gravitational force increases or decreases, the cell function responds in a linear fashion. This poses significant health risks for astronauts in long-term spaceflight. The use of NASA LEDs (central wavelengths 670 nm) will significantly improve the medical care that is available to astronauts on long-term space missions³⁶. For the majority of biostimulatory effects of NIR light described in the literature², irradiation parameters were virtually identical to those found to reduce the viscosity of interfacial water layers. Apparently, irradiation with biologically tolerated levels of laser light shows pronounced effects in biological systems which are exposed to oxidative stress. Thus, we feel justified to assume that the irradiation upregulates ATP turnover by reducing the viscosity of the nanoscopic interfacial water layers which seem to control the efficiency of the mitochondrial nanomotor. The insight deduced from our laboratory experiments is expected to allow the improvement of the present theories and hypotheses of light-induced ATP synthesis, and promises to enhance the predictive capability of existing models. Explicitly, realistic models designed to explore the functioning of ATP synthase may have to consider interfacial viscosity gradients, within and around the nanoturbine. This aspect is of considerable biological interest and may lead to a shift in the paradigm of ATP synthesis.

### Methods

#### Nanoindentation measurements and data analysis.

A nanoindenter (Nanoindenter XP, Nano Instruments Inc. Oak Ridge, Tennessee, U.S.A.) equipped with a Berkovich tip (three-faceted diamond pyramid with a total included tip angle of 130.6° and tip radius <20 nm) was used to measure the load required for the tip to penetrate into the surface of different model substrates (hydrophilic and hydrophobic materials) to a depth of 1 μm through a complete load/unload cycle. Data analysis focused on penetration depths of a few 100 nm. Hydrophilic materials comprised single crystal aluminum, single crystal gold, zinc and copper; polymers, for instance, polystyrene, served as hydrophobic materials.

#### Laser irradiation.

Substrates were exposed and during irradiation of the tip/Substrate contact zone with 670 nm laser light (power 4.5 mW, intensity 4.0 kW·m⁻²). Details of the laser configuration were reported previously³⁴. The measurements were performed in a closed box at room temperature and constant relative humidity of 67%. At relative humidity levels below 48% the effect disappeared, in accordance with analogous measurements performed by atomic force acoustic microscopy³⁷.

### References

1. Yasuda, R., Noji, H., Kinosita Jr., K. & Yoshida, M. F₁-ATPase is a highly efficient molecular motor that rotates with discrete 120° steps. Cell 93, 1117–1124 (1998).
2. Saraste, M. Oxidative phosphorylation at the fin de siècle. Science 283, 1488–1493 (1999).
3. Kinosita Jr., K., Yasuda, R., Noji, H. & Adachi, K. A rotary molecular motor that can work at near 100% efficiency. Philos. Trans. R. Soc. Lond. B Biol. Sci. 355, 473–489 (2000).

| Metal  | Au | Al | Zn | Cu |
|-------|----|----|----|----|
| Drop in load [%] | 72 | 56 | 70 | 80 |

Table 1. Reduction of indentation load in response to 670 nm laser irradiation.
Bioenergetics: From mitochondria to met_BITS
H. J. Fecht

1. Toyabe, S., Watanabe-Nakayama, T., Okamoto, T., Kudo, S. & Muneysu, E. Thermodynamic efficiency and mechnochemical coupling of F1-ATPase. *Progr. Natl. Acad. Sci. U. S. A.* 108, 17951–17956 (2011).

2. Kinosi Jr., K. Real time imaging of rotating molecular machines. *FASEB J.* 13, Suppl. 2, S201–S208 (1999).

3. Xu, L. The coupled chemomechanics of the F1-ATPase molecular motor. *Biochim. Biophys. Acta.* 1777, 1422–1431 (2008).

4. Usukura, E. et al. Torque generation and utilization in motor enzyme F0F1-ATP synthase. *J. Biol. Chem.* 287, 1884–1891 (2012).

5. Zimmermann, E. & Seifert, U. Efficiencies of a molecular motor: A generic hybrid model applied to the F1-ATPase. *New J. Phys.* 14, 103023 (2012).

6. Junge, W. Bioenergetics in mitochondria, bacteria and chloroplasts: Half a century of molecular bioenergetics. *Biochem. Soc. Trans.* 41, 1207–1218 (2013).

7. Chiwata, R. et al. None of the rotor residues of F1-ATPase are essential for torque generation. *Biophys. J.* 106, 2166–2174 (2014).

8. Vogelsberg, C. S. & Garcia-Garibay, M. A. Crystalline molecular machines: function, phase order, dimensionality, and composition. *Chem. Soc. Rev.* 41, 1892–1910 (2012).

9. Pannman, M. R. et al. Water lubricates hydrogen-bonded molecular machines. *Nat. Chem.* 5, 929–934 (2013).

10. Huesch, K. B. & Frenken, J. W. Capillary condensation in atomic scale friction: How water acts like a glue. *Phys. Rev. Lett.* 96, 166103 (2006).

11. Goertz, M. P., Houston, J. E. & Zhu, X. Y. Hydrophilicity and the viscosity of interfacial water. *Langmuir* 23, 5491–5497 (2007).

12. Kikitsu, Y., Bechert, J. & Nagyvary, J. New insights into ATP synthesis. *Proc. Natl. Acad. Sci. U. S. A.* 110, 12729–12734 (2013).

13. Huesch, K. B. & Frenken, J. W. Capillary condensation in atomic scale friction: How water acts like a glue. *Phys. Rev. Lett.* 96, 166103 (2006).

14. Goertz, M. P., Houston, J. E. & Zhu, X. Y. Hydrophilicity and the viscosity of interfacial water. *Langmuir* 23, 5491–5497 (2007).

15. Ortiz-Young, D., Chiu, H. C., Kim, S., Voitkowsky, K. & Riedo, E. The interplay between apparent viscosity and wettableness in nanofluidic water. *Nat. Commun.* 4, 2482 (2013).

16. Hutcheon, M. L., Duncan, T. M., Ngai, H. & Cross, R. L. Energy-driven subunit rotation at the interface between subunit a and the c oligomer in the F0 sector of *Escherichia coli* ATP synthase. *Proc. Natl. Acad. Sci. U. S. A.* 98, 8519–8524 (2001).

17. Minauro-Sanmiguel, F., Wilkens, S. & García, I. J. Structure of dimeric mitochondrial ATP synthase: Novel F0 bridging features and the structural basis of mitochondrial cristae biogenesis. *Proc. Natl. Acad. Sci. U. S. A.* 102, 12356–12358 (2005).

18. Ford, R. C. et al. Inelastic coherent neutron scattering measurements of intact cells and tissues and detection of interfacial water. *J. Am. Chem. Soc.* 126, 4682–4688 (2004).

19. Nagyvary, J. & Bechert, J. New insights into ATP synthesis. *Biochem. Educ.* 27, 193–199 (1999).

20. Watt, I. N., Montgomery, M. G., Runswick, M. J., Leslie, A. G. & Walker, J. E. Bioenergetic cost of making an adenosine triphosphate molecule in animal mitochondria. *Proc. Natl. Acad. Sci. U. S. A.* 107, 16823–16827 (2010).

21. Mitchell, P. Coupling of phosphorylation to electron and hydrogen transfer by a chemiosmotic type of mechanism. *Nature* 191, 144–148 (1961).

22. Janero, D. R., Hreniuk, D. & Sharif, H. M. Hydroperoxide-induced oxidative stress impairs heart muscle cell carbohydrate metabolism. *Am. J. Physiol.* 266, C179–C188 (1994).

23. Chen, H. et al. NAD+-carrying mesoporous silica nanoparticles can prevent oxidative stress-induced energy failures of both rodent astrocytes and PC12 cells. *PLoS One* 8, e74100 (2013).

24. Tiwari, B. S., Belenghi, B. & Levine, A. Oxidative stress increased respiration and generation of reactive oxygen species, resulting in ATP depletion, opening of mitochondrial permeability transition, and programmed cell death. *Plant Physiol.* 128, 1271–1281 (2002).

25. Sendner, C., Horinek, D., Boscquet, L. & Netz, R. R. Interfacial water at hydrophobic and hydrophilic surfaces: Slip, viscosity, and diffusion. *Langmuir* 25, 10768–10781 (2009).

26. Sommer, A. P., Zhu, D., Försterling, H.-D., Scharnweber, T. & Welle, A. Crystalline water at room temperature – Under water and in air. *Cryst. Growth Des.* 8, 2620–2622 (2008).

27. Sommer, A. P., Caron, A. & Fecht, H.-J. Tuning nanoscopic water layers on hydrophobic and hydrophilic surfaces with laser light. *Langmuir* 24, 635–636 (2008).

28. Sommer, A. P. et al. Breathing volume in interfacial water with laser light. *J. Phys. Chem. Lett.* 2, 562–565 (2011).

29. Sommer, A. P., Zhu, D. & Scharnweber, T. Laser modulated transmembrane convection: Implementation in cancer chemotherapy. *J. Control Release* 148, 131–134 (2010).

30. Carter, K. A. et al. Porphyrin-phospholipid liposomes permeabilized by near-infrared light. *Nat. Commun.* 5, 3546 (2014).

31. Passarella, S. et al. Increase of proton electrochemical potential and ATP synthesis in rat liver mitochondria irradiated in vitro by helium-neon laser. *FEBS Lett.* 175, 95–99 (1984).

32. Sommer, A. P. et al. 670 nm laser light and EGCG complementarily reduce amyloid-β aggregates in human neuroblastoma cells: Basis for treatment of Alzheimer’s disease? *Photomed. Laser Surg.* 30, 54–60 (2012).

33. Viswanath, D. S., Ghosh, T. K., Prasad, D. H. L., Dutt, N. V. K. & Rani, K. Y. Viscosity of Liquids: Theory, Estimation, Experiment, and Data. *Springer* pp 61–63 (2007).

34. Mester, A. & Mester, A. F. & Mester, A. The biomedical effects of laser application. *Lasers Surg. Med.* 5, 31–39 (1985).

35. Mester, A. Laser biostimulation. *Photomed. Laser Surg.* 31, 237–239 (2013).

36. Whelan, H. T. The use of NASA light-emitting diode near-infrared technology for biostimulation. In *Proceedings of the 2nd Int. Conf. on NOA*, Johnson Space Center, May 2001, Houston/TX, USA; NASA Publication CP-2002-210786 (2002).

37. Sommer, A. P., Pinheiro, A. L. B., Mester, A. R., Franke, R. P. & Whelan, H. T. Biostimulatory windows in low-intensity laser activation: lasers, scanners, and NASA’s light-emitting diode array system. *J. Clin. Laser Med. Surg.* 19, 29–33 (2001).

38. Sommer, A. P. et al. Interfacial water an exceptional biolubricant. *Cryst. Growth Des.* 9, 3852–3854 (2009).

39. Cross, R. L. Turning the ATP motor. *Nature* 427, 407–408 (2004).

40. Sommer, A. P., Haddad, M. K. h. & Fecht, H. J. Tuning the wheel of life with light. In *Abstracts of the International Conference on Laser Applications in Life Sciences*, Ulm, Germany, June 29 – July 2 (2014).

**Acknowledgement**

The authors wish to acknowledge financial support from the Accelerated Metallurgy Project, which is co-funded by the European Commission in the 7th Framework Programme (contract NMP4-LA-2011-263206), by the European Space Agency and by the individual partner organisations.

**Author Contributions**

A.P.S. designed the experiment and wrote the manuscript. A.P.S. and M. Kh.H. performed the experiments. H.J.F. contributed to the theoretical explanation of the experimental observations and derived the model parameters for their interpretation. All authors discussed the results and constructively commented on the manuscript.

**Additional Information**

**Competing financial interests:** The authors declare no competing financial interests.
How to cite this article: Sommer, A. P. et al. Light Effect on Water Viscosity: Implication for ATP Biosynthesis. Sci. Rep. 5, 12029; doi: 10.1038/srep12029 (2015).

This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article’s Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/