High-Dose Testosterone Propionate Treatment Reverses the Effects of Endurance Training on Myocardial Antioxidant Defenses in Adolescent Male Rats

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Abstract This study was aimed at evaluation of changes in activities of selected antioxidant enzymes (superoxide dismutase, catalase, glutathione peroxidase, and glutathione reductase) and contents of key nonenzymatic antioxidants (glutathione, protein thiol groups, and \( \alpha \) - and \( \gamma \)-tocopherols) in the left heart ventricle of young male Wistar rats subjected to endurance training (treadmill running, 1 h daily, 5 days a week, for 6 weeks) or/and testosterone propionate treatment (8 or 80 mg/kg body weight, intramuscularly, once a week, for 6 weeks) during adolescence. The training alone increased the activities of key antioxidant enzymes, but lowered the pool of nonenzymatic antioxidants and enhanced myocardial oxidative stress as evidenced by elevation of the lipid peroxidation biomarker malondialdehyde. The lower-dose testosterone treatment showed mixed effects on the individual components of the antioxidant defense system, but markedly enhanced lipid peroxidation. The higher-dose testosterone treatment decreased the activities of the antioxidant enzymes, lowered the contents of the nonenzymatic antioxidants, except for that of \( \gamma \)-tocopherol, reversed the effect of endurance training on the antioxidant enzymes activities, and enhanced lipid peroxidation more than the lower-dose treatment. These data demonstrate the potential risk to cardiac health from exogenous androgen use, either alone or in combination with endurance training, in adolescents.

Keywords Adolescence · Antioxidant enzymes · Endurance training · Heart · Nonenzymatic antioxidants · Oxidative stress · Testosterone · Tocopherols

Introduction

The heart is characterized by one of the highest resting oxygen consumption rates among all body organs. High myocardial oxygen consumption, which is elevated in response to exercise [1], may result in enhanced production of reactive oxygen species (ROS) and impairment of the oxidant–antioxidant balance [2]. Indeed, increased generation of ROS is detectable in rat myocardium during acute exercise [3]. The excess ROS are disposed of quickly by the cellular antioxidant defense systems consisted of a number of antioxidant enzymes, including (but not limited to) superoxide dismutase (SOD; EC 1.15.1.1), catalase (CAT; EC 1.11.1.6), glutathione peroxidase (GPX; EC 1.11.1.9), and glutathione reductase (GR; EC 1.6.4.2), and nonenzymatic antioxidants, such as reduced glutathione (GSH) and \( \alpha \) - and \( \gamma \)-tocopherols. Electrically induced tachycardia, which imitates cardiac effects of exercise, acutely decreases the activities of the antioxidant enzymes in the left heart ventricle (LV) in the rat [4], whereas the reports on the effects of exercise training on enzymatic and
nonenzymatic antioxidant heart systems are equivocal [5–7]. Notably, there is no data on such effects in adolescents. Myocardium is a sex steroid-sensitive tissue in that it expresses receptors for both androgens and estrogens [8, 9]. Some studies suggest that abuse of androgenic steroids causes adverse cardiovascular effects (e.g. LV hypertrophy, hypertension, diastolic dysfunction, and cardiac failure) [8, 10, 11]. However, other reports suggest that testosterone improves myocardial lipid metabolism [12] and may act as a cardioprotectant and coronary vasodilator with effects comparable to those of conventional anti-ischemic drugs [13, 14]. It is also not clear whether androgens improve or worsen heart antioxidant status. For instance, androgens decrease activities of heart antioxidant enzymes in adult male rats [15]. On the other hand, orchidectomy markedly decreases LV activities of antioxidant enzymes in the rat [14, 16], while testosterone replacement reverses the deficit in tissue antioxidant capacity and substantially improves the recovery of cardiac work after ischemia/reperfusion in vitro [14].

Whereas exercise training is often combined with anabolic–androgenic steroid (AAS) use in real life (see [10, 11, 17–19]), attempts to dissect the response of the myocardial antioxidant system to such combination have not given clear-cut results. Moreover, there is no data on such effects in adolescents, in whom these phenomena may be affected by the ongoing somatic growth and the still-developing pattern of sex steroids’ production and secretion. Importantly, adolescence is the period that associates with increased interest of young people, particularly boys, in strength and/or endurance sports, or simply in bodybuilding. Their wish to succeed in such activities is often fulfilled by combined use of the most appropriate physical training and illegal use/abuse of AAS [10, 18, 19]. The aim of this study was to analyze the effects of endurance training and chronic androgen treatment of adolescent male Wistar rats on the capacity of both enzymatic and nonenzymatic antioxidant defense systems in the LV.

Materials and Methods

Animals

Adolescent (5–6 weeks old) male Wistar rats of 100–120 g body weight (BW), from the stock of Mossakowski Medical Research Center (Warsaw, Poland), were used for the study. The rats were housed four to five per cage, at 12-h light/12-h dark cycle (lights on at 7:00 a.m.), 22–24°C ambient temperature and 45–65% relative humidity, and were allowed standard pellet rat chow and tap water ad libitum. Before the start of the experiment, all rats were run-tested on a motorized rodent treadmill (3 × 5 min, with 15-min breaks) for three successive days to habituate them to the training environment and to eliminate rats unwilling to run. The selected rats were randomized between the following groups: (1) untrained controls (UTr, n = 8); (2) untrained rats receiving 8 mg/kgBW of testosterone propionate (TP) weekly (UTr+TP8, n = 8); (3) untrained rats receiving 80 mg/kgBW of TP weekly (UTr+TP80, n = 8); (4) endurance-trained TP-untreated rats (Tr, n = 9); (5) endurance-trained rats receiving 8 mg/kgBW of TP weekly (Tr+TP8, n = 8); and (6) endurance-trained animals receiving 80 mg/kgBW of TP weekly (Tr+TP80, n = 9).

Hormonal Treatment

Stock TP solution (Testosteronum propionicum; Jelfa, Poland) was diluted with sesame oil as needed and injected intramuscularly once a week for 6 weeks, alternatingly into the right and left hindlimb. TP-untreated rats were given the same volume of the oil according to the same schedule. Our choice of the lower TP dosage used was based on the following facts: (1) a single TP dose of 8 mg/kgBW is sufficient to restore physiological levels of serum testosterone for about 1 week in castrated sedentary young adult male Wistar rats [12, 16], while it almost doubles circulating blood testosterone levels in their intact counterparts (Langfort et al., unpublished data), and (2) it approximated well the AAS dosage typically used by athletes/bodybuilders [11, 17, 20]. The higher TP dosage employed (80 mg/kgBW/week) was chosen to roughly match the maximum reported AAS dosage [18]. The study protocol was approved by the First Warsaw Ethical Committee for Animal Experiments of the Polish Academy of Sciences, Warsaw, Poland (Certificate of approval No. 251).

Endurance Training

The rats scheduled for endurance training were exercised on a rodent treadmill (at 0° inclination) 5 days a week, for 6 weeks. The treadmill speed was 16 m/min during the first week, was increased by 4 m/min weekly over the next 3 weeks, and was kept at 28 m/min for the remaining training sessions. The session duration started each week at 40 min/day and was increased by 5 min daily during the first 4 weeks; during the last 2 weeks, the rats ran for 1 h daily. Two days after completion of the training, the rats were euthanized by decapitation. Trunk blood was collected, allowed to clot at room temperature, and spun to obtain serum for hormone assays. The chest was opened instantly, and the heart was perfused with the Krebs–Henseleit buffer supplemented with glucose (10 mM), immediately excised and weighed, and the LV was
separated, weighed, and cut into pieces that were snap-frozen in liquid nitrogen and stored at \(-80^\circ\text{C}\) until analysis.

### Analytical Procedures

LV samples were homogenized in ice-cold buffers according to the instructions provided by the manufacturer of diagnostic kits, or in ice-cold 0.1 M phosphate-buffered saline pH 7.0 (1:5 w/v), using an Ultra-Turrax T8 homogenizer (IKA Labortechnik, Staufen, Germany). The homogenates were either used immediately for the determination of reduced glutathione (GSH), \(\alpha\)-and \(\gamma\)-tocopherols and malondialdehyde (MDA) or spun at 4,000 \(x\) g (\(4^\circ\text{C}\)) for 15 min to obtain supernatants for quantification of enzyme activities and protein thiols (-SH). CAT activity and the contents of tocopherols and protein thiols were assessed as described previously [4, 16]. The activities of SOD, GPX and GR, and GSH content were measured using BIOXYTECH kits SOD-525, GPX-340, GR-340, and GSH-400 (OXIS International Inc., Portland, OR, USA) according to the manufacturer’s instructions. Serum total testosterone (TT) was determined with a radioimmunoassay kit (Testosterone RIA, DSL-4100, Diagnostic System Laboratories Inc., Webster, TX, USA), and serum-free testosterone (FT) was measured using a coated-tube radioimmunoassay (KIPI19000, DIASource Immunoassays S.A., Belgium). Serum 17\(\beta\)-estradiol (E2) level was quantified using Elecsys Estradiol electrochemiluminescence kit (Roche, Mannheim, Germany). Total protein content was determined by the 2,2'-bicinchoninic acid method using Sigma Protein Assay Kit BCA1.

### Statistics

All data are presented as means ± SD. A two-way ANOVA with endurance training and TP treatment as the main factors, followed, when appropriate, by the Tukey post hoc test, was used to test the significance of between-group differences. To assess possible associations between variables, Spearman’s rank order correlation (except when specified otherwise) coefficients were calculated. In all cases, a \(P < 0.05\) was considered significant. All statistical analyses were performed using Statistica 7.1 software (StatSoft, Tulsa, OK, USA).

### Results

#### Serum Sex Hormone Levels

TP treatment significantly and dose dependently elevated serum TT and FT levels both in sedentary (untrained) and in endurance-trained rats. Rats given the higher TP dose compared to the sedentary controls showed, irrespective of their training status, about 18-fold and 44-fold higher TT and FT levels, respectively. There was a strong positive correlation between TT and FT levels and a moderate correlation between TT and E2 levels. Two-way ANOVA revealed significant effects of TP treatment, no significant effect of endurance training, and significant training \(\times\) TP treatment interaction effects on TT and FT levels. E2 level was only marginally affected by the TP treatment and endurance training (Table 1).

#### Body and Heart Weights

Body weight was significantly lower in endurance-trained TP-untreated rats than in their sedentary counterparts and tended to be lower in animals receiving TP. Two-way ANOVA yielded significant effects of both endurance training and TP treatment and a significant interaction between these factors’ effects on BW. Body weight showed significant negative correlation with serum TT level, whereas the correlation between heart weight (HW) and serum TT level did not reach significance. HW was significantly increased by TP treatment but not by endurance training. Heart-to-body weight ratio (HW/BW) positively correlated with serum TT and reached the highest level in the endurance-trained rats given the high-dose TP treatment (Table 1).

#### Lipid Peroxidation

Endurance training only moderately elevated LV MDA content, while TP treatment markedly elevated it both in trained and in sedentary rats at both doses employed. There was no significant interaction between these factors’ effects on LV MDA content. Notably, there was a strong positive correlation between LV MDA content and serum TT level (see Table 1) and between LV MDA content and serum FT level \((R = 0.74, P < 10^{-6})\).

#### Left Ventricle Antioxidant Enzymes’ Activities and Nonenzymatic Antioxidants’ Contents

SOD and CAT activities were significantly higher in the endurance-trained TP-untreated rats than in their sedentary counterparts; no similar differences were found between the respective GPX and GR activities. TP-treated rats, particularly those receiving the higher TP dose, showed markedly lowered activities of all antioxidant enzymes studied (Table 2). There was a strong negative correlation between SOD and CAT activity or serum FT \((R = -0.77, P < 10^{-6})\) and \(R = -0.63, P < 10^{-5}\), respectively) and TT levels, whereas the correlations between GPX or GR activity and FT \((R = -0.63, P < 10^{-5})\), and \(R = -0.54, \ldots\)
Effects of testosterone propionate (TP) treatment and endurance training (Tr) on serum sex hormone levels, body (BW) and heart (HW) weights and heart left ventricle malondialdehyde (MDA) content in young male Wistar rats

| Variable                  | UTr   | UTr + TP8 | UTr + TP80 | Tr    | Tr + TP8 | Tr + TP80 | Two-way ANOVA results | Coefficient of correlation with serum total testosterone level<sup>a</sup> |
|---------------------------|-------|-----------|------------|-------|----------|-----------|------------------------|----------------------------------|
| Total testosterone        | 5.1   | 35.4***   | 93.5***††† | 17.8  | 35.0***  | 91.1***††† | Training: $F_{1,30} = 2.67$, $P = 0.11$  | $R$  | $P$     |
| [nmol/l]                  | (0.7) | (6.2)     | (3.5)      | (1.0) | (10.5)   | (3.9)     | TP treatment: $F_{2,30} = 582.7$, $P < 10^{-6}$ | -                | -      |
|                           |       |           |            |       |          |           | Interaction: $F_{2,30} = 5.68$, $P = 0.008$ |                  |        |
| Free testosterone         | 7.8   | 88.8**    | 337.9***†††| 44.0  | 54.1     | 343.0***†††| Training: $F_{1,32} = 0.059$, $P = 0.81$  | 0.96               | $<10^{-6}$ |
| [pmol/l]                  | (6.3) | (8.5)     | (41.7)     | (33.2)<sup>b</sup> | (26.8) | (18.1)     | TP treatment: $F_{2,32} = 490.1$, $P < 10^{-6}$ |                   |        |
|                           |       |           |            |       |          |           | Interaction: $F_{2,32} = 4.76$, $P = 0.016$ |                  |        |
| Estradiol-17β             | 77.9  | 76.5      | 112.9<sup>a</sup> | 93.7  | 108.4    | 105.7     | Training: $F_{1,34} = 4.98$, $P = 0.032$  | 0.48               | 0.003   |
| [pmol/l]                  | (15.9)| (8.5)     | (21.6)     | (17.9) | (14.2)   | (22.8)    | TP treatment: $F_{2,34} = 5.68$, $P = 0.008$ |                   |        |
|                           |       |           |            |       |          |           | Interaction: $F_{2,34} = 3.45$, $P = 0.043$ |                  |        |
| BW                        | 342.8 | 316.3     | 288.8**    | 293.9** | 311.4    | 273.6     | Training: $F_{1,44} = 9.81$, $P = 0.003$  | $-0.48$            | 0.0028  |
| [g]                       | (22.3)| (22.5)    | (9.5)      | (33.3) | (42.0)   | (9.2)     | TP treatment: $F_{2,44} = 10.3$, $P = 0.0002$ |                   |        |
|                           |       |           |            |       |          |           | Interaction: $F_{2,44} = 3.28$, $P = 0.047$ |                  |        |
| HW                        | 0.81  | 0.92      | 0.90       | 0.86  | 0.89     | 0.99<sup>a</sup> | Training: $F_{1,44} = 2.47$, $P = 0.12$  | 0.31               | 0.070   |
| [g]                       | (0.06)| (0.05)    | (0.06)     | (0.11) | (0.06)   | (0.10)    | TP treatment: $F_{2,44} = 8.74$, $P = 0.001$ |                   |        |
|                           |       |           |            |       |          |           | Interaction: $F_{2,44} = 2.40$, $P = 0.10$  |                  |        |
| HW/BW                     | 2.36  | 2.92<sup>+</sup> | 3.13** | 2.98<sup>a</sup> | 2.89 | 3.60<sup>*</sup>†† | Training: $F_{1,44} = 11.76$, $P = 0.0013$ | 0.59               | 0.0002  |
| [mg/g]                    | (0.17)| (0.21)    | (0.26)     | (0.62) | (0.38)   | (0.32)    | TP treatment: $F_{2,44} = 15.8$, $P = 10^{-5}$ |                   |        |
|                           |       |           |            |       |          |           | Interaction: $F_{2,44} = 3.43$, $P = 0.041$  |                  |        |
| MDA                       | 312.4 | 828.4***  | 883.2***   | 409.4 | 947.3*** | 1,108.1***†††| Training: $F_{1,42} = 28.9$, $P = 3 \times 10^{-6}$ | 0.79               | $<10^{-6}$ |
| [μmol/g wet wt]           | (51.8)| (119.7)   | (125.2)    | (187.1)| (77.0)   | (87.1)    | TP treatment: $F_{2,42} = 206.0$, $P < 10^{-6}$ |                   |        |
|                           |       |           |            |       |          |           | Interaction: $F_{2,42} = 2.09$, $P = 0.14$  |                  |        |

Data are means (SD). UTr untrained rats, Tr endurance-trained rats, TP8 8 mg/kgBW TP once a week, TP80 80 mg/kgBW TP once a week

<sup>a</sup> The correlation coefficients were calculated using Spearman rank correlation test, except that the correlation between free and total testosterone levels was calculated by linear regression analysis

<sup>b</sup> $P = 0.004$ versus the UTr group, Mann–Whitney U test

<sup>*</sup> $P < 0.05$, <sup>**</sup> $P < 0.01$, <sup>***</sup> $P < 0.001$ vs. the respective TP-untreated group; <sup>#</sup> $P < 0.05$, <sup>##</sup> $P < 0.01$, <sup>###</sup> $P < 0.001$ vs. the respective untrained group; <sup>†</sup> $P < 0.05$, <sup>‡</sup> $P < 0.01$, <sup>‡‡</sup> $P < 0.001$ vs. the respective low TP dose-treated group
Table 2: Effects of testosterone propionate (TP) and endurance training (Tr) on activities of superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPX), and glutathione reductase (GR) in the left ventricle of young male Wistar rats

| Enzyme   | UTr | UTr + TP8 | UTr + TP80 | Tr | Tr + TP8 | Tr + TP80 | Two-way ANOVA results                                                                 |
|----------|-----|-----------|------------|----|---------|----------|--------------------------------------------------------------------------------------|
| SOD [U/mg protein] | 3.93 | 3.30 | 1.57*+s+††† | 5.10### | 2.79*** | 0.68**+s+††† | Training: $F_{1,42} = 0.33, P = 0.57$  
TP treatment: $F_{2,42} = 215.4, P < 10^{-6}$  
Interaction: $F_{2,42} = 22.1, P < 10^{-6}$ |
| CAT [1/(s × g protein)] | 11.38 | 13.75 | 7.40†† | 20.92### | 11.73*** | 6.70###+† | Training: $F_{1,42} = 6.43, P = 0.015$  
TP treatment: $F_{2,42} = 35.1, P < 10^{-6}$  
Interaction: $F_{2,42} = 16.6, P = 5×10^{-6}$ |
| GPX [mU/mg protein] | 263.3 | 260.2 | 213.7* | 279.1 | 256.9 | 181.1###+s+††† | Training: $F_{1,42} = 0.54, P = 0.46$  
TP treatment: $F_{2,42} = 25.4, P < 10^{-6}$  
Interaction: $F_{2,42} = 2.4, P = 0.10$ |
| GR [mU/mg protein] | 5.09 | 4.39 | 3.84*** | 4.48 | 3.56* | 3.14*** | Training: $F_{1,42} = 21.5, P = 3×10^{-5}$  
TP treatment: $F_{2,42} = 24.7, P < 10^{-6}$  
Interaction: $F_{2,42} = 0.19, P = 0.83$ |

For group labels, see Table 1. Data are means (SD). * $P < 0.05$, *** $P < 0.001$ vs. the respective TP-untreated group value; # $P < 0.05$, ## $P < 0.01$, ### $P < 0.001$ vs. the respective untrained group value; † $P < 0.05$, †† $P < 0.01$, ††† $P < 0.001$ vs. the respective low TP dose-treated group value.
| Antioxidant | UTr | UTr + TP8 | UTr + TP80 | Tr | Tr + TP8 | Tr + TP80 | Two-way ANOVA results | Spearman rank correlation with serum total testosterone level |
|------------|-----|----------|-----------|----|---------|---------|------------------------|----------------------------------------------------------|
|            |     |          |           |    |         |         | Training: $F_{1,42} = 11.1, P = 0.002$ | $R$ | $P$ |
| GSH [μmol/g tissue] | 1.90 (0.21) | 1.76 (0.22) | 1.68 (0.19) | 1.54** (0.28) | 1.74 (0.76) | 1.49 | | $-0.005$ | $0.76$ |
| Protein thiols [μmol -SH/g protein] | 218.1 (10.5) | 256.3** (22.6) | 233.1 (9.0) | 200.7 (22.4) | 247.2** (25.2) | 205.6†† (28.7) | Training: $F_{1,42} = 8.75, P = 0.005$ | $0.23$ | $0.17$ |
| α-tocopherol [μmol/g tissue] | 0.0541 (0.0043) | 0.0358*** (0.0043) | 0.0438***† (0.0056) | 0.0480 (0.0051) | 0.0339*** (0.0030) | 0.0379*** (0.0030) | Training: $F_{1,42} = 12.7, P = 0.0009$ | $-0.49$ | $0.0025$ |
| γ-tocopherol [μmol/g tissue] | 0.00120 (0.00014) | 0.00149 (0.00033) | 0.00161 (0.00035) | 0.00088 (0.00019) | 0.00144** (0.00046) | 0.00129 (0.00068) | Training: $F_{1,42} = 6.97, P = 0.012$ | $0.35$ | $0.038$ |

For group labels, see Table 1. Data are means (SD). ** $P < 0.01$, *** $P < 0.001$ vs. the respective TP-untreated group value; ** $P < 0.01$ vs. the respective untrained group value; † $P < 0.05$, †† $P < 0.01$ vs. the respective low TP dose-treated group value.
phenomena related to apoptosis [24]. The lower TP dose activity of hormone-sensitive lipase and thereby stimulates that testosterone (serum levels of which positively correlated with LV MDA content in this study) elevates the decrease in major antioxidant enzyme activities and most nonenzymatic antioxidants studied. However, it is known that testosterone doses used in our study has been shown to attenuate this used in our study has been shown to attenuate this superoxide anions (O$_2^-$) due to a reversible reaction of O$_2^-$ radicals with native enzyme which involves a reduction of the ferric (Fe$^{3+}$) center to oxyferrous heme (Fe$^{2+}$)…O$_2$ [28]. Whereas consequences of increased oxidative stress in the model used in our study are not clear, it has been shown that such stress can induce apoptosis in rat ventricular myocytes in vitro [29]. Endurance training employing a protocol closely related to this used in our study has been shown to attenuate this effect by SOD- and Hsp70-mediated pathways in rats [30]. However, superoxide radical is not the only ROS that is involved in apoptosis, and different ROS may use distinct apoptotic pathways [29].

Our training protocol is known to shift anaerobic threshold to higher exercise intensity [31], and its

**Discussion**

The main findings of the present study are that (1) endurance training by itself increases the activity of two key components of heart enzymatic antioxidant system, namely SOD and CAT, but decreases the capacity of heart non-enzymatic antioxidant system and increases LV lipid peroxidation; (2) high-dose TP treatment decreases the capacity of both these antioxidant systems by itself, reverses the beneficial effect of endurance training on SOD and CAT activities, and potentiates myocardial lipid peroxidation. Whereas the effects of exercise training and androgens on heart antioxidant systems have been studied before, this study, to the best of our knowledge, is the first to investigate the effects of these factors in adolescent subjects. This issue is of considerable importance because of the well-known AAS abuse by adolescent bodybuilders and athletes and the emergence of the related spectrum of adverse side effects including a number of cardiac and cardiovascular complications leading eventually to death in some cases [10, 11, 17, 19–22].

While AAS can cause detrimental shifts in serum lipid profile, AAS abuse-related cardiac deaths are not necessarily related to coronary thrombosis or atherosclerosis [10, 22, 23], but may be caused also by a variety of pathological phenomena related to apoptosis [24]. The lower TP dose used in this study markedly elevated myocardial MDA in untrained rats, implying a substantial increase in oxidative stress. This may look surprising, as there was no concurrent decrease in major antioxidant enzyme activities and most nonenzymatic antioxidants studied. However, it is known that testosterone (serum levels of which positively correlated with LV MDA content in this study) elevates the activity of hormone-sensitive lipase and thereby stimulates lipolysis in rat cardiomyocytes [12]. This improves long-chain fatty acids availability for ATP synthesis, which in turn elevates oxygen utilization and thus enhances generation of ROS. The reported testosterone-induced increase in adipose tissue lipolysis [25] could have contributed also to the decreased BW of TP-treated rats in this study.

TP treatment substantially elevated both serum TT and FT in our rats, but the respective relative increases were much larger for FT (see Table 1). The traditional view holds that only FT represents biologically active testosterone. However, there was a high positive correlation between serum TT and FT levels in this study, and the correlation coefficients between the other indices measured and FT or TT level were, pair wise, remarkably similar (not shown), except that there was no significant correlation between serum FT and E2 levels (R = 0.22, P = 0.18). Thus, our data support the view that serum TT accurately reflects biologically active testosterone in adolescent rats [10, 26]. Interestingly, serum TT levels in the rats treated with the lower TP dose were, 9 days after the last dose, several times higher than those found during the first week following the injection of the same TP dose to naïve young adult male rats (Langfort et al., unpublished data). This shows extensive buildup of serum testosterone with weekly TP doses, which is in clear contrast with the need for administration of TP at 2- to 3-day intervals in humans with the aim of attaining reasonably stable TT levels.

Evidence from animal studies indicates that, compared with other aerobic tissues, the heart has moderate levels of antioxidant enzymes and thus rather limited ability to remove ROS [2]. Hence, it is possible that the LV antioxidant enzyme system was unable to inactivate the excessive ROS formation consequential to supraphysiological testosterone doses used in our study. However, significant decreases in the antioxidant enzymes’ activities could have contributed to increased oxidative stress in rats given the higher TP dose treatment. It is well documented that SOD is irreversibly inactivated (by up to 50%) by its product hydrogen peroxide in a concentration-dependent manner [27]. Besides, CAT, a heme enzyme, is rapidly inhibited by superoxide anions (O$_2^-$) center to oxyferrous heme (Fe$^{2+}$)…O$_2$ [28]. Whereas consequences of increased oxidative stress in the model used in our study are not clear, it has been shown that such stress can induce apoptosis in rat ventricular myocytes in vitro [29].

Endurance training employing a protocol closely related to this used in our study has been shown to attenuate this effect by SOD- and Hsp70-mediated pathways in rats [30]. However, superoxide radical is not the only ROS that is involved in apoptosis, and different ROS may use distinct apoptotic pathways [29].

Our training protocol is known to shift anaerobic threshold to higher exercise intensity [31], and its
effectiveness is additionally evidenced by the significant increase in HW/BW ratio in TP-untreated endurance-trained rats in this study. The shift in anaerobic threshold is associated with increased oxidative capacity and reliance of oxidative muscles on fat metabolism and presumably with enhanced ROS formation that may lead to enhanced lipid peroxidation [32].

The published reports on changes in lipid peroxidation induced by chronic exercise are equivocal. For instance, significant increases in MDA content in heart tissue of chronically exercised rats were reported by some researchers [33], whereas no changes were found in another study [3]. Similarly, the literature is ambiguous regarding which myocardial antioxidants are elevated after exercise training [7, 15, 34, 35]. Endurance training has been shown to increase the activities of antioxidant enzymes, particularly of SOD and CAT, in adult rats. The most robust responses to endurance training revealed in the present study were the increases in LV activities of SOD (the enzyme considered essential for protection against ROS) and CAT (the enzyme involved in removal of hydrogen peroxide produced by SOD) in TP-untreated rats. This apparent enhancement of enzymatic antioxidant defense was, however, associated with a significant negative impact of the training on all nonenzymatic antioxidants studied, and thus it could not prevent the enhancement of lipid peroxidation evidenced by the significant effect of the training on LV MDA content. Again, this observation is in agreement with the aforementioned moderate ability of exercise training on key components of the enzymatic antioxidant defense system. Conversely, it showed significant negative correlations with LV SOD, CAT and GR activities, and with LV $\alpha$-tocopherol level, with correlation coefficients ranging from $-0.32$ to $-0.45$. All these coefficients were considerably lower than those for serum TT level (see Table 2), which indicates that they merely reflected substrate–product relationship between testosterone and E2. These data are in agreement with earlier reports that showed no major effect of exogenous testosterone on circulating E2 level in male rodents [14] and suggest no cardioprotective effect from systemic testosterone aromatization.

Notably, there was no significant correlation between serum E2 level and LV $\gamma$-tocopherol content, whereas the latter correlated positively with serum TT level. ANOVA yielded significant decreasing effect on this antioxidant from endurance training and an opposing effect from TP treatment. The latter finding may be of importance in light of reports showing that $\gamma$-tocopherol has potent anti-inflammatory properties [38] and may be an effective cardioprotectant [39]. It is not clear whether the elevation in LV $\gamma$-tocopherol was driven by its mobilization from peripheral stores in response to oxidant attack, as previously reported for other localizations [40, 41], or resulted from the known inverse relationship between $\gamma$- and $\alpha$-tocopherol [42]. However, the substantial TP treatment-related elevation in LV MDA observed in the present study indicated that the increased $\gamma$-tocopherol content was insufficient to counteract the enhancement of myocardial oxidative stress induced by TP treatment.

To summarize, this study shows that in adolescent male rats: (1) all potentially beneficial effects of endurance training on the enzymatic components of the antioxidant defense system in the heart left ventricle are reversed by chronic high-dose testosterone propionate treatment and (2) none of the potentially detrimental effects of the training on the left ventricular antioxidant barrier are considerably counteracted by this treatment. These results imply that anabolic androgen use among adolescents, either alone or in combination with endurance training, may elevate oxidative stress-related risk to cardiac health.

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Conflict of interest The authors have no financial and/or personal relationships with other people or organizations that could influence this report.

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