Pollination Efficiency of *Amegilla calens* (Hymenoptera: Apidae) on *Gossypium hirsutum* L. (Malvaceae) Variety L457 Flowers at Meskine (Maroua, Cameroon)

Issaya Kodji Issaya, Beaudelaine Stephanie Kengni*, Sidonie Fameni Tope, Fernand-Nestor Tchuenguem Fohouo

Department of Biological Sciences, Faculty of Science, University of Maroua, Maroua, Cameroon

Email: *beaudelaine@yahoo.fr

**Abstract**

Animal pollination is a key of reproduction of more plants species including *Gossypium hirsutum*. This Malvaceae is the best cultivated oilseed plant in tropical and subtropical regions in the world. This research evaluates the interaction between *Amegilla calens* and *Gossypium hirsutum* from 2018 to 2019 based on the activities of the solitary bee on the flowers of this plant and their productive impact in field. The experiment was focused on four repeated treatments during the flowering phase: the first two were characterised by the existence or absence of protection of the flowers regarding all insects and the other two made up of flowers designed for the exclusive visit of *A. calens* or open then protected again without a visit from an insect or any other organism. Daily rhythm of activity, behavioural ecology, the pollination efficiency, the fruiting rate, the number of seeds per capsule and the percentage of normal seeds were evaluated. Among twenty insects species recorded *A. calens* is the most abundant with 30.72% of 655 visits and the most efficient pollinator. *Amegilla calens* is active on *G. hirsutum* flowers from 6 a.m. to 5 p.m. with a maximum activity situated from 8 a.m. to 9 a.m. All visits of this solitary bee are able to fertilize the flowers of Malvaceae. The mean duration of a visit per flower for pollen harvest is also more important (23.57 ± 0.96 sec) than nectar collection (13.69 ± 0.72 sec). For the two years, through its pollination efficiency, *A. calens* increased the fruiting rate by 20.30%, as well as the percentage of normal seeds by 32.39%. Therefore, we concluded that *A. calens* is the important pollinator and increases the fecundity and number of normal seeds of *G. hirsutum.*
1. Introduction

The importance of pollinating insects in agricultural production, mainly with regard to that of domestic and wild bees, is no longer to be proven but remains generally unknown [1]. In the natural environment and in agroecosystems, flowering insects in general and Apoidea in particular have great ecological and economical importance because they have a positive influence on food production [2] [3]. Insect pollination in many crops is essential for fruit and seed productions [4].

*Gossypium hirsutum* (cotton), commonly called white gold, is the second most important oil-seed crop after *Glycine max* (soybean) [5].

To our knowledge, the data published after detailed studies on the interactions between insects and *G. hirsutum* are those from Sudan [6], Russia [7], Australia [8] [9] [10], USA [7] [11] [12] [13] and Cameroon [2] [14] [15] [16].

In all these investigations, the foraging behaviour and pollination activity were carried out in detail only on *Apis mellifera* and *Macronomia vulpina*. The flowering entomofauna and the impact of insects on pollination, fruit and seed yields of a plant species may vary with the species of insect, time and space [17]. Cameroon is classified among the countries with a severe level of hunger with an overall hunger index of 17.6 with 22% of the undernourished population [18]. In this country, the demand for cotton seed is high (250,000 tons/year) while its production is low (240,000 tons/year) [19]. Currently, the processing of seed cotton produces around 15 million litres of cotton seed oil per year for an estimated population of 25 million inhabitants [19]. Cotton yields can be increased in this country if its flowering insects are well known and exploited [16]. Therefore, it is important to investigate the possibilities of increasing the production of this valuable plant in Cameroon.

The general objective of this work is to contribute to the understanding of the relationships between *A. calens* and *G. hirsutum*, for their optimal management. Specific objectives are to: 1) determine the place of *A. calens* in the *G. hirsutum* floral entomofauna; 2) study the activity of this Apidae on flowers of the Malvaceae; 3) evaluate the impact of the flowering insects including *A. calens* on pollination, fruit and seed yields of *G. hirsutum*; 4) estimate the pollination efficiency of *A. calens* on this plant species.

2. Material and Methods

2.1. Study Site, Experimental Plot and Biological Material

The experiment was carried out from 13th September to 12th December 2018 and from 11th September to 1st December 2019 at Meskine within an experimental
fields (latitude: 10°32'26"N; longitude: 014°14'53"E; altitude: 424 m above sea level). This Region belongs to the ecological zone with three phytogeographical areas (Sudano-Sahelian, Sahelian and Sudanian altitude) periodically flooded, with unimodal rainfall [20]. The climate is characterized by two seasons: a dry season (November to May) and a rainy season (June to October); August is the wettest month of the year [21]. Annual rainfall varies from 400 to 1100 mm [21]. The annual average temperature varies between 29°C and 38°C; daily temperature range between 6°C and 7°C [21]. The experimental plot was a field of 437 m². The bees, *A. calens* of the experimental station were recruited among the arthropods naturally present in the environment. The vegetation was represented by wild and cultivated species. The plant material was represented by the seeds of *G. hirsutum* variety L457 provided by the Institute of Agricultural Research for Development of Maroua.

2.2. Sowing and Weeding

From June 15th to 21st 2018 and from July 20th to 24th 2019, the experimental plot was divided into eight subplots of 8 × 4.5 m² each. Four seeds were sown per hole on six lines per subplot. There were 16 holes per subplot. Holes were separated 50 cm from each other, while lines were 70 cm apart [14]. Weeding was performed manually as necessary to maintain plots weeds-free.

2.3. Estimation of the Frequency of *Amegilla calens* Visits on *Gossypium hirsutum* Flowers

The frequency of *A. calens* visits on *G. hirsutum* flowers was determined based on observations of flowers of treatments 1 and 5, every day, from 13th to 22nd September 2018 and from 11th September to 1st October 2019 according to six daily time frames: 6 - 7 a.m., 8 - 9 a.m., 10 - 11 a.m., 12 - 1 p.m., 2 - 3 p.m. and 4 - 5 p.m. In a slow walk along all labelled flowers of treatments 1 and 5, the identity of all insects that visited *G. hirsutum* flowers was recorded [22]. Specimens of all insect taxa were caught using insect net on unlabelled flowers and conserved in 70% ethanol, excluding butterflies that were preserved dry [23], for subsequent taxonomic identification. All insects encountered on flowers were registered and the cumulated results expressed as the number of visits to determine the relative frequency of *A. calens* in the anthophilous entomofauna of *G. hirsutum* [22]. Data obtained were used to determine the frequency of visits (*F*) of each insect species on *G. hirsutum* flowers. For each study period, \[ F_i = \left[ \frac{V_i}{V_t} \right] \times 100 \], were \( V_i \) is the number of visits of insect *i* on treatment with unprotected flowers and \( V_t \) the total number of insect visits of all recorded insect species on these flowers.

2.4. Study of the Activity of *Amegilla calens* on *Gossypium hirsutum* Flowers

2.4.1. Floral Product Harvested

In addition to the determination of the flower visiting insect frequency, direct
observation of the foraging activity of *A. calens* on flowers was made in the experimental field. The floral product (nectar or pollen) harvested by *A. calens* during each flower visit was registered based on its foraging behaviour. Nectar foragers were seen extending their proboscis between the base of the corolla and stamens, while pollen gatherers scratched the anthers using their mandibles and legs [23] [24]. During the same time the duration of *A. calens* visits on flowers was registered, the type of floral product harvested by this solitary bee was noted [24]. In the morning of each sampling day, the number of opened flowers was counted. Data obtained were used to determine the relationship between the number of visits of *A. calens* and the corresponding flowers [22].

### 2.4.2. Duration of Visits and Foraging Speed

During the same days as for the registration of the frequency of visits, the duration of individual flower visit was recorded (using stopwatch) according to six daily time frames: 7 - 8 a.m., 9 - 10 a.m., 11 - 12 a.m., 1 - 2 p.m., 3 - 4 p.m. and 5 - 6 p.m. Moreover, the number of visits during which the bee came into contact with the stigma [25], was registered. Regarding the foraging speed (*F*) which is the number of flowers visited by an individual bee per minute [25], data were registered during the same dates and according to the same time frames and date as for the duration of visits. The stopwatch, previously set to zero was switched on as soon as an individual landed on a flower and the number of visited flowers was concomitantly counted. The stopwatch was stopped as soon as the visitor was lost to sight or when it left *G. hirsutum* flower for another plant species. The foraging speed (*F*) was calculated using the following formula: 

\[
F = \frac{Nf}{d} \times 60, 
\]

where *d* is the duration (in sec) given by stopwatch and *Nf* the number of flowers visited during *d*. [22]. During the observation, when a forager returned to previously visited flower, counting was performed as on two different flowers [22].

### 2.4.3. Abundances per Flower and per 1000 Flowers

The abundance of foragers (highest numbers of individuals foraging simultaneously) [26] per flower and per 1000 flowers (*A*1000) were recorded on the same dates and daily time frames as that for the registration of the duration of visits. Abundance per flower was recorded as a result of direct counting. For determining the abundance per 1000 flowers, foragers were counted on a known number of opening flowers and *A*1000 was calculated using the following formula:

\[
A_{1000} = \left( \frac{Ax}{Fx} \right) \times 1000, 
\]

where *Fx* and *Ax* are respectively the number of flowers and the number of foragers effectively counted on these flowers at time *x* [26].

### 2.4.4. Foraging Ecology

The disruption of the activity of foragers by competitors or predators and the attractiveness exerted by other plant species on *A. calens* was assessed by direct observations. For the second parameter, the number of times that the solitary
bee left these Malvaceae flowers to another plant species and vice versa was noted through the investigation period.

During each observation date, temperature and relative humidity in the station were registered every 30 minutes using a mobile thermo-hygrometer installed in the shade [22], from 6 a.m. to 6 p.m.

2.4.5. Evaluation of the Impact of Flowering Insects Including *Amegilla calens* on *Gossypium hirsutum* Yields

Parallel to the constitution of treatments 1, 2, 5 and 6, 600 flowers at bud stage were labelled in 2018 and in 2019, to form two treatments:

- treatment 3 in 2018 or 7 in 2019: 400 flowers protected using gauze bag nets to prevent insect or any other organism visits and destined to be visited exclusively by *A. calens*; as soon as each flower of these treatments was opened, the gauze bag was removed and this flower was observed for up to 10 minutes; the flower visited once by *A. calens* was marked and then protected once more [27];

- treatment 4 in 2018 or 8 in 2019: 200 flowers protected using gauze bag nets and destined to be uncovered then rebagged without the visit of insects or any other organism; as soon as each flower of these treatments was opened, the gauze bag was removed and this flower was observed for up to 10 minutes, while avoiding insect or any other organism visits.

At the maturity, fruits were harvested and counted from each treatment. The fruiting rate, the percentage of seeds per fruit and the percentage of normal seed were then determined for each treatment [28].

Evaluation of the effect of insects including *A. calens* on *G. hirsutum* production was based on the impact of flowering insects on pollination, the impact of pollination on *G. hirsutum* fruiting and the comparison of the fruiting rate, the number of seeds per fruit and the percentage of normal (that is well developed) seed [29] of treatments 1, 2, 4, 5, 6 and 8. For each year, the fruiting rate due to the foraging insects including *A. calens* (*Fri*) was calculated using the following formula [29]: 

$$Fri = \frac{(FX - FZ)(FX + FY - FZ)}{FX + FY - FZ} \times 100,$$

where *FX*, *FY* and *FZ* are the fruiting rates in treatment *X* (flowers left in free pollination), treatment *Y* (flowers protected from all insect visits) and treatment *Z* (flowers bagged then uncovered and rebagged without insect or any other organism visit). The fruiting rate of a treatment (*F*) is 

$$F = \frac{(b/a) \times 100}{\text{number of fruits formed}} \times \frac{\text{number of viable flowers initially set}}{a},$$

where *b* is the number of fruits formed and *a* the number of viable flowers initially set [30]. The impact of flower visiting insects including *A. calens* on the percentage of seeds per fruit and the percentage of normal seeds was evaluated using the same method as mentioned above for the fruiting rate [30].

2.4.6. Assessment of the Pollination Efficiency of *Amegilla calens* on *Gossypium hirsutum*

The contribution of *A. calens* on the fruiting rate, the number of seeds per fruit and the percentage of normal seeds, was calculated using the data of treatments
For each observation year, the contribution of *A. calens* in the fruiting rate \((FrA)\) was calculated using the following formula:

\[
FrA = \left( \frac{(FA - FZ)}{FA} \right) \times 100
\]

[26], where \(FA\) is the fruiting rate in treatment \(A\) (flowers visited exclusively by *A. calens*).

The impact of *A. calens* on the fruiting rate, number of seed per fruit and the percentage of normal seeds were evaluated using the same method as mentioned above for the fruiting rate.

### 2.5. Data Analysis

The data collected and aggregates were subjected to the analysis of variance (ANOVA) considering the different treatments: effect of visits of all insects, influence of or without visit, effect of an exclusive visit of *A. calens* and protection effect and opening and reprotection without visit, based on a fixed linear model and to the joint analysis within each year (2018 and 2019). These analyses were carried out using R commander, version i386 3.2.0. The treatment means were compared two by two by student’s \(t\)-test at \(p < 0.05\), the relationship study between two variables by Pearson correlation coefficient \((r)\) at \(p < 0.05\) and the comparison of percentages by chi-square \((\chi^2)\) at \(p < 0.05\).

### 3. Results

#### 3.1. Place of *Amegilla calens* in *Gossypium hirsutum* Floral Entomofauna

Among 655 visits of 11 and 19 insect species recorded on *G. hirsutum* flowers in 2018 and 2019 respectively, *A. calens* ranked third with 32 visits (13.22%) after *Lasioglossum* sp. 1 (36.36%) and *Apis mellifera* (14.82%) in 2018 and first with 172 visits (41.64%) in 2019 (Table 1). The difference between the percentages of *A. calens* visit for two years is highly significant \((\chi^2 = 57.48; df = 1; P < 0.001)\).

For the two cumulated years *A. calens* ranked first with 204 visits (30.72%).

#### 3.2. Duration of Visit per Flower

The mean duration of *A. calens* visit per *G. hirsutum* flower varied according to floral product harvested. In 2018, the mean duration of a flower visit for nectar harvest was 16.29 ± 1.29 sec \((n = 34; s = 7.42)\) and that for pollen collection was 25.80 ± 1.49 sec \((n = 72; s = 12.55)\); the difference between these means is highly significant \((t = 4.82; df = 104; P < 0.001)\). In 2019, the corresponding figures were 11.08 ± 0.77 sec \((n = 37; s = 4.60)\) for nectar harvest and 21.33 ± 1.25 sec \((n = 76; s = 10.82)\) for pollen collection. The difference between these two means is highly significant \((t = 6.99; df = 111; P < 0.001)\).

For the two cumulated years, the mean duration of a flower visit was 13.69 ± 0.72 sec \((n = 71; s = 6.01)\) for nectar harvest and 23.57 ± 0.96 sec \((n = 148; s = 11.69)\) for pollen collection. The difference between these two later means is highly significant \((t = 8.22; df = 217; P < 0.001)\).
Table 1. Diversity of flowering insects on *Gossypium hirsutum* in 2018 and 2019, number and percentage of visits of different insects.

| Insects | 2018 | 2019 | Total |
|---------|------|------|-------|
| Order   | Family | Genus and species | \( n \) | \( p_1 (%) \) | \( n \) | \( p_2 (%) \) | \( n_t \) | \( p_t (%) \) |
| Diptera | Muscidae | *Musca domestica* (ne) | 3 | 1.23 | 4 | 0.96 | 7 | 1.07 |
|         | Calliphoridae | *Chrysomia chloropyga* (ne) | 7 | 2.06 | - | - | 7 | 2.52 |
| Coleoptera | Scarabaeidae | (sp. 1) (po) | 30 | 12.39 | 9 | 2.17 | 39 | 5.99 |
|         | (sp. 2) (po) | 4 | 1.65 | 2 | 0.48 | 6 | 0.92 |
|         | (sp. 3) (po) | - | - | 6 | 1.45 | 6 | 1.45 |
| Meloidae | *Mylabris* sp. (ne. po) | 10 | 4.13 | 10 | 2.42 | 20 | 3.07 |
| Apidae | *Apis mellifera* (ne. po) | 36 | 14.87 | 21 | 5.08 | 57 | 8.75 |
|         | *Amegilla calens* (ne. po) | 32 | 13.22 | 172 | 41.64 | 204 | 30.72 |
|         | *Amegilla* sp. 1 (ne. po) | - | - | 3 | 0.72 | 3 | 0.72 |
|         | *Amegilla* sp. 2 (ne. po) | - | - | 4 | 0.96 | 4 | 0.96 |
|         | *Amegilla* sp. 3 (ne. po) | - | - | 2 | 4.48 | 2 | 4.48 |
|         | *Amegilla* sp. 4 (ne. po) | - | - | 19 | 0.60 | 19 | 0.60 |
|         | *Amegilla* sp. 5 (ne. po) | 9 | 3.71 | - | - | 9 | 3.78 |
|         | *Tetralonia* sp. (ne. po) | - | - | 7 | 1.69 | 7 | 1.07 |
| Halictidae | *Lasiglossum* sp. 1 (ne. po) | 88 | 36.36 | 107 | 25.90 | 195 | 29.95 |
|         | *Lipotriches azarensis* (ne. po) | - | - | 1 | 0.24 | 1 | 0.24 |
| Vespidae | (sp. 1) (ne. po) | 5 | 2.06 | 34 | 8.23 | 39 | 5.99 |
|         | (sp. 2) (po) | - | - | 6 | 1.45 | 6 | 1.45 |
| Lepidoptera | *Acraea acerata* (ne) | 12 | 4.95 | 3 | 1.72 | 15 | 2.30 |
|         | *Eurema* sp. (ne) | 6 | 2.47 | 3 | 1.72 | 9 | 1.38 |
| Total   | 242 | 100 | 413 | 100 | 655 | 100 |

\( n_1 \) and \( n_2 \): number of visits on 120 flowers in 2018 and in 2019; ne: collection of nectar; po: collection of pollen; \( p_1 \) and \( p_2 \): percentages of visits; sp: undetermined species. \( p_1 = (n_1/242) \times 100 \); \( p_2 = (n_2/413) \times 100 \); \( p_t = (n_t/655) \times 100 \).

3.3. Activity of *Amegilla calens* on *Gossypium hirsutum* Flowers

3.3.1. Floral Products Harvested

Individuals of *A. calens* were seen collecting nectar (Figure 1(A)) and pollen (Figure 1(B)) on *G. hirsutum* flowers. Pollen collection was regular and intensive whereas nectar collection was less. For 106 visits recorded in 2018, 72 (67.92%) were devoted to pollen collection and 34 (32.08%) to nectar harvest; in 2019, for 113 visits registered, 76 (67.25%) were devoted to pollen collection and 37 (32.74%) to nectar harvest. For the two cumulated years on 219 visits recorded, 148 (67.57%) were devoted to pollen collection and 71 (32.42%) to nectar harvest. Nectar and pollen were harvested during all scheduled observation daily time frames. For harvesting nectar, the individual of *A. calens* lands either on the stamens or on the stigma, then moves towards the inside of the corolla and...
deploys its proboscis to collect nectar. For the pollen collection the bee comes into contact with the flower from above, the thorax being in contact with the stigma; subsequently, it uses the hind legs, mandibles and abdominal hair to collect pollen which is stored at the level of their hind legs.

3.3.2. Daily Rhythm of Visits
*Amegilla calens* was active on *G. hirsutum* flowers from 6 a.m to 5 p.m in 2018 and in 2019. The peak of activity was situated between 8 and 9 a.m in 2018 as well as in 2019 (Figure 2).

In 2018, the correlation was not significant between the number of *A. calens* visits and the temperature ($r = 0.11; df = 4; P > 0.05$) and between the number of visits and relative humidity ($r = 0.34; df = 4; P > 0.05$). In 2019, the correlation was equally not significant between the number of *A. calens* visits and the temperature ($r = −0.12; df = 4; P > 0.05$) and between the number of these visits and relative humidity ($r = 0.73; df = 4; P > 0.05$) (Figure 3).

In 2018 as well as in 2019 *A. calens* visits were apparently more numerous on *G. hirsutum* individual plant when their number of opened flowers was highest. But, the correlation between the number of opened flowers and the number of visit was not significant in 2018 ($r = 0.62; df = 8; P > 0.05$) as well as in 2019 ($r = 0.14; df = 7; P > 0.05$) (Figure 4).

3.3.3. Foraging Speed of *Amegilla calens* on *Gossypium hirsutum* Flowers
In the experimental field, *A. calens* visited between 1 and 6 flowers per minute in 2018 and between 2 and 13 flowers per minute in 2019 (Table 2). The mean foraging speed was $3.07 \pm 0.33$ flowers per minute ($n = 69; s = 2.74$) in 2018 and $2.69 \pm 0.33$ flowers per minute ($n = 60; s = 2.52$) in 2019. There is no difference between these two means ($t = 0.81, df = 127; P > 0.05$). For the two cumulated years, the mean foraging speed was $2.88 \pm 0.23$ flowers per minute ($n = 129; s = 2.64$).

3.3.4. Abundance of *Amegilla calens*
In 2018, the highest mean number of *A. calens* individuals simultaneously in ac-
tivity was $1 \pm 0.00$ per flower ($n = 47; s = 0$) and $533.80 \pm 55.68$ per 1000 flowers ($n = 36; s = 329.42$). In 2019, the corresponding figures were $1 \pm 0.00$ per flower ($n = 89; s = 0$), and $547.67 \pm 32.19$ per 1000 flowers ($n = 102; s = 325.06$). There is no difference between the mean number of foragers per 1000 flowers in 2018 and 2019 ($t = 0.22; df = 136; P > 0.05$). For the two cumulated years the mean number of foragers per 1000 flowers was $540.74 \pm 43.94$.

Table 2. Foraging speed of *Amegilla calens* on *Gossypium hirsutum* flowers in 2018 and 2019 at Meskine.

| Years | Number of flowers/minute | Comparison of means |
|-------|--------------------------|---------------------|
|       | $n$ | $m$ | $sd$ | mini | maxi | $t$ | $df$ | $P > 0.05$ |
| 2018  | 69  | 3.07 | 0.33 | 2.74 | 6    | 0.82 | 127  | NS          |
| 2019  | 60  | 2.69 | 0.33 | 2.51 | 13   | 0.82 | 127  | NS          |
| Total | 129 | 2.97 | 0.23 | 2.37 | 13   | 0.82 | 127  | NS          |

$n$: number of speeds registered; $m$: mean; $sd$: standard deviation; $mini$: minimum; $maxi$: maximum; NS: not significant difference.

Figure 2. Variations of the number of *Amegilla calens* visits on *Gossypium hirsutum* flowers according to the daily time frames in 2018 (A) and 2019 (B) at Meskine.
Figure 3. Variation of the temperature, the humidity and the number of *Amegilla calens* visits on *Gossypium hirsutum* flowers according to the daily frames time in 2018 (A) and 2019 (B) at Meskine.

### 3.3.5. Influence of Neighbouring Flora

During the observation period, flowers of many other plant species growing in the environment of *G. hirsutum* field were visited by *A. calens*, for their nectar (ne) and/or pollen (po). Amongst these plants were: *Solanum lycopersicum* (Solanaceae: po); *Cosmos sulphureus* (Asteraceae: ne and po), *Hibiscus sabdariffa* (Malvaceae: ne and po), *Abelmoschus esculentus* (Malvaceae: ne and po), and *Ceratotheca sesamoides* (Pedaliaceae: ne and po). During the two years of study,
we observed no passage of *A. calens* from *G. hirsutum* flowers to flowers of another plant species and vice versa. Hence during foraging trips on *G. hirsutum*, individuals of *A. calens* were faithful to this Malvaceae.

### 3.3.6. Influence of Fauna

Individuals of *A. calens* were disturbed in their foraging activity by other individuals of the same species or those from other species, that were the competitor

---

**Figure 4.** Seasonal variations of the number *Gossypium hirsutum* opened flowers and the number of *Amegilla calens* visits on these organs in 2018 (A) and 2019 (B) at Meskine.
for *G. hirsutum* nectar and/or pollen. In 2018, for 106 visits, 4 (3.77%) were interrupted by *Lasioglossum* sp. 1 and 2 (1.88%) by individuals of a Vespidae (sp. 1). In 2019, for 113 visits, 3 (2.65%) were interrupted by a Vespidae (sp. 1) and 1 (0.88%) by *Mylabris* sp.

### 3.4. Impact of Anthophilous Insects Including *Amegilla calens* on *Gossypium hirsutum* Yields

The fruiting rate, the mean number of seeds per fruit and the percentage of normal seeds in the different treatments of *G. hirsutum* are shown in **Table 3**.

This table shows that:

1) The fruiting rates were 89.16%, 52.5%, 88.70%, 85.36%, 80%, 66.66%, 86.16% and 56% in treatments 1 to 8 respectively. The differences between these eight percentages are globally highly significant ($\chi^2 = 114.27; df = 7; P < 0.001$). The two to two comparisons showed that the difference observed is highly significant between treatments 1 and 2 ($\chi^2 = 39.05; df = 1; P < 0.001$), 2 and 4 ($\chi^2 = 30.75; df = 1; P < 0.001$), 7 and 8 ($\chi^2 = 32.22; df = 1; P < 0.001$), 4 and 8 ($\chi^2 = 25.74; df = 1; P < 0.001$), significant between treatments 5 and 6 ($\chi^2 = 5.46; df = 1; P < 0.05$), and not significant between treatments 1 and 5 ($\chi^2 = 3.87; df = 1; P > 0.05$), 3 and 4 ($\chi^2 = 0.73; df = 1; P > 0.05$), 6 and 8 ($\chi^2 = 2.93; df = 1; P > 0.05$), 1 and 3 ($\chi^2 = 0.016; df = 1; P > 0.05$), then 5 and 7 ($\chi^2 = 1.89; df = 1; P > 0.05$). Consequently, in 2018 and 2019, the fruiting rate of unprotected flowers (treatments 1 and 5) was higher than that of protected flowers (treatments 2 and 6).

2) The mean numbers of seeds per fruit were $26.68 \pm 2.75$, $24.09 \pm 0.87$, $25.54 \pm 0.89$, $23.82 \pm 0.78$, $25.38 \pm 0.73$ and $23.68 \pm 0.70$ in treatments 1 to 8 respectively. The differences between these eight means are

| Years | Treatments | NF | NFF | FR (%) | Seeds/fruit | TNS | NS | %NS |
|-------|------------|----|-----|--------|-------------|-----|----|-----|
|       |            | n  |     |        | Mean        | sd  | n  |     |
| 2018  | 1 (Uf)     | 120| 107 | 89.16  | $26.68 \pm 2.75$ | 6.25| 95 | 2535| 2408| 94.99
|       | 2 (Pf)     | 120| 63  | 52.5   | $24.09 \pm 0.87$ | 6.38| 55 | 1325| 1162| 87.69
|       | 3 (Fpva)   | 177| 157 | 88.70  | $25.54 \pm 0.89$ | 6.08| 48 | 1226| 1131| 92.25
|       | 4 (Fpwv)   | 123| 105 | 85.36  | $23.82 \pm 0.78$ | 4.46| 34 | 810 | 710 | 87.65
| 2019  | 5 (Uf)     | 120| 96  | 80.00  | $31.25 \pm 0.69$ | 6.03| 78 | 2438| 2242| 91.96
|       | 6 (Pf)     | 120| 80  | 66.66  | $25.67 \pm 0.74$ | 6.26| 73 | 1874| 1583| 84.47
|       | 7 (Pva)    | 159| 137 | 86.16  | $25.38 \pm 0.73$ | 5.68| 62 | 1574| 1474| 93.64
|       | 8 (Fpwv)   | 125| 70  | 56.00  | $23.68 \pm 0.70$ | 4.94| 51 | 1208| 1052| 87.08

Uf: unprotected flowers; Pf: protected flowers; Fpva: flowers visited exclusively by *Amegilla calens*; Fpwv: flowers bagged then uncovered and rebagged without visit by insect or any other organism; NF: number of flowers; NFF: number of fruits formed; FR: fruiting rate; TNS: total number of seeds; NS: number of normal seeds; %NS: percentage of normal seeds.
globally highly significant ($F = 11.4; \ df_1 = 7; \ df_2 = 488; \ P < 0.001$). The two to two comparison showed that the difference is highly significant between treatments 5 and 7 ($t = 5.87; \ df = 138; \ P < 0.001$), 1 and 5 ($t = 4.85; \ df = 171; \ P < 0.001$), 5 and 6 ($t = 5.53; \ df = 149; \ P < 0.001$), so significant between treatments 1 and 2 ($t = 2.40; \ df = 148; \ P < 0.05$), not significant between treatments 7 and 8 ($t = 1.69; \ df = 111; \ P > 0.05$), 2 and 4 ($t = 0.23; \ df = 87; \ P > 0.05$), 4 and 8 ($t = 0.13; \ df = 83; \ P > 0.05$), 3 and 4 ($t = 1.46; \ df = 80; \ P > 0.05$), 1 and 3 ($t = 1.04; \ df = 141; \ P > 0.05$), then 6 and 8 ($t = 1.96; \ df = 122; \ P > 0.05$). Thus, in 2018 as well as in 2019, the mean number of seeds per fruit of unprotected flowers was higher than that of protected flowers.

3) The percentages of normal seeds were 94.99%, 87.69%, 92.25%, 87.65%, 91.96%, 84.47%, 93.64% and 87.08% in treatments 1 to 8 respectively. The differences between these eight percentages are globally highly significant ($\chi^2 = 204.18; \ df = 7; \ P < 0.001$). Pairwise comparisons showed that the difference observed is highly significant between treatments 1 and 2 ($\chi^2 = 66.59; \ df = 1; \ P < 0.001$), 5 and 6 ($\chi^2 = 59.31; \ df = 1; \ P < 0.001$), 7 and 8 ($\chi^2 = 35.21; \ df = 1; \ P < 0.001$), 1 and 5 ($\chi^2 = 11.90; \ df = 1; \ P < 0.001$), 1 and 3 ($\chi^2 = 11.16; \ df = 1; \ P < 0.001$), significant between treatments 5 and 7 ($\chi^2 = 3.98; \ df = 1; \ P < 0.05$), 6 and 8 ($\chi^2 = 4.05; \ df = 1; \ P < 0.05$), and not significant between treatments 2 and 4 ($\chi^2 = 0.001; \ df = 1; \ P > 0.05$), then 4 and 8 ($\chi^2 = 0.14; \ df = 1; \ P > 0.05$). Hence, in 2018 and 2019, the percentage of normal seeds of unprotected flowers was higher than that of flowers protected during their opening period.

In 2018, the numeric contribution of anthophilous insects in the fruiting rate, the mean number of seeds per fruit and the percentage of normal seeds of *G. hirsutum* were 6.76%, 10.61% and 7.72% respectively. In 2019, the corresponding figures were 26.47%, 23% and 5.46%. For the two cumulated years, the numeric contribution of anthophilous insects including *A. calens* was 16.62%, 16.81% and 6.59% for the fruiting rate, the mean number of seeds per fruit and the percentage of normal seeds of *G. hirsutum* respectively.

### 3.5. Pollination efficiency of *Amegilla calens* on *Gossypium hirsutum*

During a single flower visit of *A. calens* for nectar or pollen harvest on *G. hirsutum* flowers, this bee regularly came into contact with anthers and stigma (100%), increasing the possibility of the Malvaceae pollination.

The fruiting rates (*Table 3*) due to *A. calens* were 88.70% in 2018, 86.16% in 2019 and 87.43% for the two cumulated years. Therefore, in 2019, the fruiting rate of flowers protected and visited exclusively by *A. calens* was higher than that of flowers protected and rebagged without insect or any other organism visit. The mean numbers of seeds per fruit due to *A. calens* were 25.54 ± 0.89 in 2018, 25.38 ± 0.73 in 2019 and 25.46 ± 0.81 for the two cumulated years. Thus, in 2018 as well as in 2019 the numbers of seeds per fruit in protected flowers and visited
exclusively by *A. calens* was not higher than that of flowers protected and re-bagged insect or any other organism visit. The percentage of normal seeds due to *A. calens* were 92.25% in 2018, 93.64% in 2019 and 92.95% for the two cumulated years. For each of the two years, results pointed out that flowers visited by *A. calens* have the highest number of normal seeds compare to those protected then uncovered and rebagged without the visit of insect or any other organism.

In 2018, the contribution of *A. calens* on the percentage of normal seeds via on single flower visit was 4.99%. In 2019, the fruiting rate and the percentage of normal seeds were 35% and 28.63% respectively. For the two cumulate years, the numerique contribution of *A. calens* via a single flower visit on the fruiting rate and the percentage of normal seeds were 20.30%, and 32.39% respectively.

4. Discussion

4.1. Activity of *Amegilla calens* on *Gossypium hirsutum* Flowers

Results obtained from these experiments indicated that *A. calens* was the main floral insect visitor of *G. hirsutum* flowers. The high frequency of individual foragers of *A. calens* on the flowers could be explained by the large number of individuals of this bee inside the experimental field and the good attractiveness of the floral products of this Malvaceae towards this bee. The significant difference between the percentages of *A. calens* visit for the two years of study could be attributed to a combination of climatic factors and seasonal variation in floral resources availability. It is well known that the anthophilous insect fauna of a plant varies over time [3] [22]. Other researches have revealed *Apis mellifera* [3] [6] [7] [8] [14] and *M. vulpina* [16] as the main insect visitors on the flowers of this Malvaceae. This difference could be explained by the absence of the nests of these two bee species within and near the experimental site, and the presence of other plant species with flowers able to attract *Apis mellifera* and *M. vulpina* foragers.

The significant difference observed between the mean duration of a pollen harvest visit and that of nectar harvest visit could be explained by the importance and accessibility of each of these floral products. Pollen is produced by the anthers, which are on the top of the stamens, whereas nectar is between the base of the style and stamens [26]. *Amegilla calens* do not make honey. It harvests more pollen and less nectar to make bread which is the protein source of young larva [31].

The high abundance of individual bees per 1000 flowers and the attractiveness of *G. hirsutum* nectar and pollen for *A. calens* could be partially explained by the higher availability of these substances, their accessibility and the needs of *A. calens* during the flowering period of the Malvaceae.

The absence of the passage of *A. calens* from *G. hirsutum* flowers to flowers of another plant species and vice versa could be explained by the fidelity of this solitary bee to the flowers of this plant during foraging bouts. This phenomenon is called floral constancy [32]. It is explained by the fact that the solitary bees
in general are able to memorize the form, the color and the smell of the flowers visited during previous foraging trips [33]. These same observations have been made for honey bees on the flowers of cotton in Mayel-Ibe [3] (Maroua-Cameroon).

The disruption of visits by other insects in 2018 by *Lasioglossum* sp. 1 (3.77%) and Vespidae (sp. 1) (1.88%), then in 2019 by Vespidae (sp. 1) (2.65%) and *Mylabris* sp. (0.88%) reduced the duration of some *A. calens* visits. This obliged some individuals of this bee to visit more flowers during a foraging trip to maximize their pollen or nectar loads. Similar observations have been made on *Apis mellifera* workers foraging on the flowers of this Malvaceae in Maroua [3] and in Garoua [16].

### 4.2. Impact of Anthophilous Insects Including *Amegilla calens* on *Gossypium hirsutum* Yields

The numeric contribution of anthophilous insects to the fruiting rate, the number of seeds per fruit and the percentage of normal seeds of *G. hirsutum* was positive. During nectar and/or pollen harvest on *G. hirsutum*, foraging insect in general always shook flowers and regularly contacted anthers and stigma, increasing self-pollination and/or cross-pollination possibilities of this Malvaceae. Our results agreed with those obtained in: Mayel-Ibbe (Maroua) [3], Dang (Ngaoundere) [14] and Djamboutou (Garoua) [16] on this Malvaceae.

### 4.3. Pollination Efficiency of *Amegilla calens* on *Gossypium hirsutum*

During a single flower visit of *A. calens* for the collection of nectar and/or pollen on each flower, individuals of *A. calens* always come into contact with the stigma and anthers (100%) and thus increasing the possibilities of *G. hirsutum* pollination. They could thus enhance self-pollination by applying pollen of one flower on its own stigma. *Amegilla calens* could provide allogamous pollination through carrying of pollen with their hairs, legs, mouthparts, thorax and abdomen, which is then deposited on flowers belonging to a different plant of the same species (geitogamy) [30]. The intervention of *A. calens* on the pollination of *G. hirsutum* is especially probable since their abundance per 1000 flowers and their foraging speed were high. The positive and significant contribution of *A. calens* in the fruiting rate and the percentage of normal seeds of *G. hirsutum* is justified by the action of this bee on the pollination of visited flowers. This significant contribution of *A. calens* on the fruiting rate and the percentage of normal seeds of *G. hirsutum* is in agreement with similar findings for *Apis mellifera* [3] [14] [16] and *M. vulpina* [15] on the same Malvaceae.

### Acknowledgements

I could not finish this work without thanking the Institute of Agricultural Research for Development of Maroua for providing us with an experimental plot.
Funding

This research has not received external funding.

Institutional Review Board Statement and Informed Consent Statement

It is not applicable.

Data Availability Statement

There is no new data created or analyzed in this research.

Conflicts of Interest

We declare there is no conflict of interest.

References

[1] AREM (2011) Enjeux de la pollinisation pour la production agricole en Tarn-Et-Garonne. Ecole d’ingénieurs, France, 106 p.

[2] Tchuenguem, F.F.N., Messi, J. and Pauly, A. (2002) L’activité de butinage des Apoïdes sauvages (Hymenoptera: Apoidea) sur les fleurs de maïs à Yaoundé (Caméroun) et réflexions sur la pollinisation des graminées tropicales. Biotechnologie, Agronomie, Société et Environnement, 6, 87-98.

[3] Dounia and Tchuenguem, F.F.N. (2013) Foraging and Pollination Activity of Apis mellifera adansonii Latreille (Hymenoptera: Apidae) on Flowers of Gossypium hirsutum L. (Malvaceae) at Maroua, Cameroon. International Research Journal of Plant Science, 4, 33-44.

[4] Klein, A.M., Brittain, C., Hendrix, S.D., Thorp, R., Williams, N. and Kremen, C. (2012) Wild Pollination Services to California Almond Rely on Semi-Natural Habitat. Journal of Applied Ecology, 49, 723-732. https://doi.org/10.1111/j.1365-2664.2012.02144.x

[5] Karavina, C., Andumbu, R.M., Parwada, C. and Ungunyana, T.M. (2012) Variety and Planting Date Effects on the Incidence of Bollworms and Insect Sucking Pests of Cotton (Gossypium hirsutum L.). Research Journal of Agricultural Sciences, 3, 607-610.

[6] Ahmed, H.M.H., Siddig, M.A. and El-Sarrag M.S.A. (1989) Honeybee Pollination of Some Cultivated Crops in Sudan. Proceedings of the Fourth International Conference on Apicultural in Tropical Climates, Cairo, 6-10 November 1988, 100-108.

[7] McGregor, S.E. (1976) Insect Pollination of Cultivated Crop Plants. Agricultural Research Service. United States Department of Agriculture, Agriculture Handbook, United States 411, 496 p.

[8] Thomson, N.J. (1966) Cotton Variety Trials in the Ord Valley, North Western Australia: Natural Crossing of Cotton. Empire Cotton Growing Review, 43, 18-21.

[9] Mungomery, J.E. and Glassop, A.J. (1969) Natural Cross-Pollination of Cotton in Central Queensland. Queensland Journal of Agriculture and Animal Sciences, 26, 69-74.

[10] Richard, J.S., Stanley, J.N. and Gregg, P.C.O. (2005) Viability of Cotton and Canola Pollen on the Proboscis of Helicoverpa armigera. Implication for Spread of Tran-
genes and Pollination Ecology. *Ecological Entomology*, **30**, 327-333. https://doi.org/10.1111/j.0307-6946.2005.00694.x

[11] Umbeck, P., Johnson, G., Barton, K. and Swain, W. (1987) Genetically Transformed Cotton (*Gossypium hirsutum* L.). *Plant Biotechnology*, **5**, 263-267. https://doi.org/10.1038/nbt0387-263

[12] Van Deynze, A.E., Sundstrom, F.J. and Bradford, K.J. (2005) Pollen-Mediated Gene Flow in California Cotton Depends on Pollinator Activity. *Crop Science*, **45**, 1565-1570. https://doi.org/10.2135/cropsci2004.0463

[13] Llewellyn, D.J., Tyson, C., Constable, G.A., Duggan, B., Beale, S. and Steel, P. (2007) Containment of Regulated Genetically Modified Cotton in the Field. *Agriculture Ecosystems and Environment*, **121**, 419-429. https://doi.org/10.1016/j.agee.2006.11.019

[14] Mazi, S., Tchuenguem, F.F.N. and Brückner, D. (2013) Foraging and Pollination Behaviour of *Apis mellifera adansonii* Latreille (Hymenoptera: Apidae) on *Gossypium hirsutum* (Malvaceae) Flowers at Dang (Ngaoundéré, Cameroon). *Journal of Agricultural Science and Technology*, **3**, 267-280.

[15] Dounia and Tchuenguem, F.F.N. (2014) Foraging and Pollination Activity of *Macronomia vulpina* (Gerstaecker, 1857) (Hymenoptera: Halictidae) on *Gossypium hirsutum* L. (Malvaceae) Flowers at Maroua, Cameroon. *International Journal of Agronomy and Agricultural Research*, **4**, 1-10.

[16] Basga, E., Fameni, S.T., Aitbita, E.N.O. and Tchuenguem, F.F.N. (2019) Efficacité pollinisatrice de *Apis mellifera* Linné (Hymenoptera: Apidae) sur les fleurs de *Gossypium hirsutum* (Malvaceae) à Djamboutou (Garoua, Cameroun). *Journal of Applied Biosciences*, **138**, 14123-14136. https://doi.org/10.4314/jab.v138i1.11

[17] Gallai, N., Salles, J.M., Settele, J. and Vaissière, B.E. (2009) Economic Valuation of the Vulnerability of World Agriculture Confronted with Pollinator Decline. *Ecological Economics*, **68**, 810-821. https://doi.org/10.1016/j.ecolecon.2008.06.014

[18] IRAM/MINADER (2017) Evaluation des risques agricoles au Cameroun. Rapport Final, 138 p.

[19] MINADER (2010) Annuaire des Statistiques du Secteur Agricole, Campagnes 2007 & 2008. Direction des Enquêtes et Statistiques, AGRI-STAT Cameroun, 16: 98.

[20] Letouzey, R. (1985) Notice de la carte phytogéographique du Cameroun au 1: 500000. Institut de la Cartographie Internationale et de la Végétation, Toulouse, 142 p.

[21] Kuete, M., Melingui, A., Mounkam, J. and Nofiele, D. (1993) Nouvelle Géographie du Camroun. 207 p.

[22] Tchuenguem, F.F.N. (2005) Activité de butinage et de pollinisation de *Apis mellifera adansonii* Latreille (Hymenoptera: Apidae, Apinae) sur les fleurs de trois plantes à Ngaoundéré (Cameroun): *Callistemon rigidus* (Myrtaceae), *Syzygium guineense var. macrocarpum* (Myrtaceae) et *Voacanga africana* (Apocynaceae). Thèse de Doctorat d’Etat, Université de Yaoundé I, Yaoundé, 103 p.

[23] Borror, D.J. and White, R.E. (1991) Les insectes de l’Amérique du Nord (au nord du Mexique). Broquet (éd.), Laprairie, 408 p.

[24] Jean-Prost, P. (1987) Apiculture: Connaître l’abeille—conduire le rucher. 6ème Edition, Lavoisier (éd.), Paris, 159 p.

[25] Jacob-Remacle, A. (1989) Comportement de butinage de l’abeille domestique et des abeilles sauvages dans des vergers de pomiers en Belgique. *Apidologie*, **20**, 271-285. https://doi.org/10.1051/apido:19890401
[26] Tchuenguem, F.F.N., Messi, J., Brückner, D., Bouba, B., Mbofung, G. and Hentchoya, H.J. (2004) Foraging and Pollination Behaviour of the African Honey Bee (Apis mellifera adansonii) on Callistemon rigidus Flowers at Ngaoundere (Cameroon). Journal of the Cameroon Academy of Sciences, 4, 133-140.

[27] Tchuenguem, F.F.N. and Népide, N.C. (2018) Efficacité pollinisatrice de Apis mellifera L. (Hymenoptera: Apidae) sur Sesamum indicum (Pedaliaceae) var. Graine Blanche et Lisse à Dang (Ngaoundéré, Cameroun). International Journal of Biological and Chemical Sciences, 12, 446-461. https://doi.org/10.4314/ijbcs.v12i1.35

[28] Tchuenguem, F.F.N., Ngakou, A. and Kengni, B.S. (2009) Pollination and Yield Responses of Cowpea (Vigna unguiculata L. Walp.) to the Foraging Activity of Apis mellifera adansonii (Hymenoptera: Apidae) at Ngaoundéré (Cameroon). African Journal of Biotechnology, 8, 1988-1996.

[29] Diguir, B.B., Pando, J.B., Fameni, T.S. and Tchuenguem, F.F.N. (2020) Pollination Efficiency of Dactylurina staudingeri (Hymenoptera: Apidae) on Vernonia amygdalina (Asteraceae) Florets at Dang (Ngaoundéré, Cameroon). International Journal of Research Studies, 6, 22-31. https://doi.org/10.20431/2454-6224.0602004

[30] Tchuenguem, F.F.N., Messi, J. and Pauly, A. (2001) Activité de Meliponula erythra sur les fleurs de Dacryodes edulis et son impact sur la fructification. Fruits, 56, 179-188. https://doi.org/10.1051/fruits:2001121

[31] Bakiri, E. (2018) Abeilles sauvages et abeilles domestique: Impact sur la biodiversité et la prodcitivité. Algerie, 16 p.

[32] Waser, N.M. (1986) Flower Constancy: Definition, Cause, and Measurement. The American Naturalist, 127, 593-603. https://doi.org/10.1086/284507

[33] Amaya-Márquez, M. (2009) Floral Constancy in Bees: A Revision of Theories and a Comparison with Other Pollinators. Revista Colombiana de Entomología, 35, 206-216.