Review

The mechanistic pathways of oxidative stress in aortic stenosis and clinical implications

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Abstract

Despite the elucidation of the pathways behind the development of aortic stenosis (AS), there remains no effective medical treatment to slow or reverse its progress. Instead, the gold standard of care in severe or symptomatic AS is replacement of the aortic valve. Oxidative stress is implicated, both directly as well as indirectly, in lipid infiltration, inflammation and fibro-calcification, all of which are key processes underlying the pathophysiology of degenerative AS. This culminates in the breakdown of the extracellular matrix, differentiation of the valvular interstitial cells into an osteogenic phenotype, and finally, calcium deposition as well as thickening of the aortic valve. Oxidative stress is thus a promising and potential therapeutic target for the treatment of AS. Several studies focusing on the mitigation of oxidative stress in the context of AS have shown some success in animal and in vitro models, however similar benefits have yet to be seen in clinical trials. Statin therapy, once thought to be the key to the treatment of AS, has yielded disappointing results, however newer lipid lowering therapies may hold some promise. Other potential therapies, such as manipulation of microRNAs, blockade of the renin-angiotensin-aldosterone system and the use of dipeptidylpeptidase-4 inhibitors will also be reviewed.

Key words: Severe aortic stenosis; reactive oxidative species; oxidative stress

Introduction

Aortic stenosis (AS) is one of the most prevalent valvular heart diseases and constitutes a large portion of the burden on public health globally [1]. While degenerative disease is the most common aetiology of AS, other causes include congenital heart diseases, rheumatic heart disease. Moreover, systemic inflammatory and infiltrative disease can also contribute to AS development and progression [2]. Despite our understanding of this condition, there is currently no effective medical therapy to reduce the symptoms, mortality or valve progression of degenerative AS. The mainstay treatment of symptomatic AS is aortic valve replacement, either surgical (SAVR), or more recently, transcatheter aortic valve implantation (TAVI) in select patient populations [3]. SAVR and TAVI lead to better outcomes in survival and hospitalisations compared to medically managed patients with severe AS, with improved symptomology and ventricular function[3].

The aortic valve normally consists of three leaflets, each having a trilaminar matrix composing of ordered layers of collagen fibres, proteoglycans and elastic fibres respectively, bestowing compliance during systole, and the ability for leaflet apposition during diastole to prevent the backflow of blood [4]. The aortic valve is composed of three layers – the ventricularis, spongiosa and fibrosa. The ventricularis on the ventricular side of the leaflet contains fibres
rich in elastin, while the fibrosa on the aortic side of the leaflet comprises collagen fibres. These fibres give the aortic valve flexibility and strength to withstand decades of repetitive movement. The spongiosa consists of an extracellular matrix rich in glycosaminoglycans, and valvular interstitial cells (VICs, also called cardiac fibroblasts), are the predominant cell type in all layers [5].

The pathophysiology of degenerative AS is a complex interplay of inflammation, lipid infiltration and fibro-calcification [6]. Histological studies on calcified aortic valve tissues reveal inflammatory infiltrates characterised by macrophages and T-cells, while elevated levels of pro-inflammatory cytokines have been found in stenotic valves as well [7]. Coagulation also plays a part - the expression of coagulation factors such as factor VII and factor X have been found in stenotic aortic valves [8]. Thrombin and tissue factor is suspected to be involved in the calcification process of degenerative AS [9]. Under pro-inflammatory conditions, VICs will express these factors [5], which may trigger the coagulation cascade and leading to a build-up of fibrin within the valve. In fact, there is a positive correlation between the amount of fibrin in the aortic valve and the transvalvular pressure gradient [10].

These processes lead to progressive thickening of the valve leaflets, compromising leaflet motility and resulting in a narrowed valve orifice. In severe cases, this causes significant systolic blood flow obstruction, increased systemic afterload and ultimately, left ventricular remodelling, cardiac dysfunction, and heart failure [7].

The aetiology of degenerative AS shares many similarities to atherosclerosis. Risk factors such as age, smoking, obesity, hypertension, hypercholesterolaemia and diabetes are linked to the development of degenerative AS [8-15]. At the molecular level, stenotic aortic valves resemble aortic plaques and contain elevated levels of matrix metalloproteinases, oxidised phospholipids (oxPLs), and calcium [16-18].

Reactive oxidative species (ROS) are reactive molecules that are produced in the process of oxygen metabolism, and major sources include the mitochondrial electron transport chain as well as the oxidoreductase group of enzymes [15,19]. Oxidative stress, an imbalance between ROS such as superoxide and peroxynitrite, and antioxidant protective mechanisms, is one of the major upstream contributors to inflammation, lipid infiltration and calcification, with downstream effects leading to the development of degenerative AS. While oxidative stress affects both the extracellular matrix and the cells of the aortic valve, most of its influence revolves around the VICs (Central Illustration).

Oxidative stress is prevalent in older age, possibly due to increased ROS, lower levels of antioxidants, reduced repair/removal mechanisms as well as the presence of multiple comorbidities such as chronic kidney disease [20]. TAVI tends to be offered to this population on patients, compared to younger patients with fewer comorbidities in the SAVR cohort. The alleviation of AS with TAVI causes near-instantaneous decrease in levels of oxidative stress. Higher pre-TAVI baseline level of plasma superoxide has been associated with increased post-interventional inflammation and poorer clinical outcomes [21]; these ROS can act as signalling molecules in upregulating pro-osteogenic pathways that will be described later.

Hence, therapies targeting oxidative stress may potentially hold the key to slowing or reversing the progression of degenerative AS. A literature review was conducted on PubMed using the search terms ‘aortic stenosis’ and ‘oxidative stress’. This review will explore the mechanisms of oxidative stress in the development of degenerative AS, as well as discuss the clinical implications and possible medical therapies.

**Oxidative Stress and Lipid Infiltration**

The initiation of AS is hypothesized to be related to oscillatory shear stress, causing endothelial dysfunction [15]. In particular, bicuspid aortic valves are less efficient in dissipating this shear stress, resulting in accelerated endothelial damage in these phenotypes [22,23]. As such, patients with bicuspid aortic valves develop AS at an earlier age and display more rapid progression compared to those with tricuspid aortic valves [14]. Once the valve endothelium is damaged, there is infiltration of OxPL-containing lipoproteins such as lipoprotein(a) and oxidised low-density lipoprotein (oxLDL), similar to the pathogenesis of atherosclerosis [24,25]. These circulating lipids enter by initially adhering tightly to lysine-binding sites on the exposed valve surfaces [26]. Indeed, there is an association between elevated levels of oxPLs with the progression of AS and subsequent requirement for intervention [27]. OxPLs are also proapoptotic, and these apoptotic pathways may also be implicated in the pathophysiology of AS [15].

Oxidation of lipoproteins can be enzymatic or non-enzymatic; the latter is induced by ROS in a process called lipid peroxidation [19]. For example, myeloperoxidase (MPO) is an oxidoreductase enzyme that catalyses the formation of ROS and thus has been implicated in the oxidation of LDL [28]. High levels of oxLDLs interfere with the protective, antioxidant effects of high-density lipoprotein (HDL) cholesterol;
levels of MPO in valve tissue are found to increase along with the severity of AS, while the converse is true for HDL cholesterol [29]. Increasing concentrations of oxLDLs also lead to the uncoupling of endothelial nitric oxide synthase, resulting in a switch in the production from nitric oxide, which is a protective antioxidant, to the ROS superoxide [30-33].

Further lipid peroxidation and endothelial injury contribute to the inflammatory and calcification processes within the valve (Figure 1). Further studies demonstrating the association between lipid peroxidation and aortic stenosis are summarised in Table 1 [34-41].

### Table 1: Basic science and clinical studies demonstrating the association between oxidative stress and aortic stenosis.

| First Author [Ref] | Year | Study Type | Key Findings |
|-------------------|------|------------|--------------|
| Zheng KH [41]     | 2019 | Cohort study (n=145) | Patients with Lp(a) and OxPL levels in the top tertile had greater progression of valvular CT calcium score, faster haemodynamic progression on echocardiography, increased risk of aortic valve replacement and death |
| Zeng Q [40]       | 2014 | In vitro – human VICs incubated in an osteogenic medium | VICs incubated with oxLDLs had higher expression of the BMP-2 pathway and NOTCH1 signalling, with resultant increase in osteogenesis |
| Mohty D [38]      | 2015 | Case-control study (n=300) | PCSK9 levels were increased in human VICs incubated in an osteogenic medium, and a PCSK9 neutralising inhibitor decreases MPO and oxLDL deposition |
| Mohty D [38]      | 2014 | In vitro – human VICs incubated in an osteogenic medium | Prolonged incubation of the VICs with Lp(a) significantly increased calcium deposition |
| Yu B [45]         | 2017 | In vitro – human VICs incubated in an osteogenic medium containing Lp(a) and OxPLs | Calcium deposition was further augmented when VICs were incubated with both Lp(a) and OxPLs |
| Yu B [39]         | 2018 | In vitro – human VICs incubated in an osteogenic medium containing Lp(a) and OxPLs | Incubation of VICs with Lp(a) significantly increased ROS formation |
| Nsabia M [49]     | 2016 | Case-control study (n=300) | Lp(a) and oxPL levels were associated with higher autotaxin activity; patients with higher autotaxin and Lp(a) levels had an increased risk of AS |
| Nsabia M [49]     | 2008 | Ex vivo – human aortic valve tissue | NADPH oxidase 2 is significantly increased in human calcific aortic valves |
| Li F [37]         | 2015 | In vitro – porcine VICs | oxLDLs induce VIC osteogenesis via activation of the receptor for advanced glycation end products (RAGE) |
| Langsted A [77]   | 2016 | Cohort study (n=103,083) | PCSK9 loss-of-function mutation has lower levels of Lp(a) and reduced risk of AS |
| Hofmanis J [29]   | 2019 | Case control study (n=102) | AS severity is negatively correlated with levels of HDL cholesterol; higher MPO levels are negatively correlated with levels of HDL cholesterol as well MPO causes HDL cholesterol dysfunction via oxidation, reducing its protective effects |
| Kamstrup PR [36]  | 2013 | Cohort study (n=77,680) | Elevated Lp(a) levels and genotypes that increase plasma Lp(a) levels are associated with an increased risk of AS |
| Perrot N [78]     | 2015 | In vitro – human VICs incubated with serotonin | Superoxide and hydrogen peroxide levels were increased in calcified regions of the aortic valve Superoxide dismutase activity and expression were reduced in calcified regions of the aortic valve |
| mercier N [64]    | 2020 | In vitro – human VICs | SSAS levels were positively correlated with increasing calcification SSAS inhibition decreased VIC calcification |
| Miller JD [16]    | 2008 | In vitro – superoxide and superoxide dismutase levels were measured in human aortic valves | Superoxide levels were increased after incubation with serotonin Inhibitors of flavin-oxidases or monoamine oxidase prevented the serotonin-induced increase in superoxide levels |
| Mohy D [38]       | 2008 | Ex vivo – human aortic valve tissue | Valves with higher oxLDL content had higher levels of inflammatory cells, TNF-α and tissue remodelling Lp(a) and oxPL levels were associated with higher autotaxin activity; patients with higher autotaxin and Lp(a) and oxPL levels had an increased risk of AS |
| Peña-Silva RA [62]| 2009 | Ex vivo – human heart valves incubated with serotonin | Soluble ST2 disrupts mitochondrial fusion and oxidative phosphorylation capacity, as well as activates the osteogenic NF-kB pathway |
| Pena-Silva RA [62]| 2020 | Ex vivo – human heart valves | Soluble ST2 levels are positively correlated with oxidative stress and inflammation |
| Demir B [6]       | 2012 | Cohort study (n=64) | There is a positive correlation between serum uric acid levels and AS severity Uric acid accelerates the formation of oxLDLs and may decrease NO levels |
| Coté C [35]       | 2008 | Ex vivo – human aortic valve tissue | Increased levels of circulating oxLDLs are associated with worse fibrocalcific remodelling of valvular tissue in AS |
| Choi B [46]       | 2017 | In vitro – human VICs | NO depletion in human VICs activates the NF-κB pathway, which promotes DPP-4 expression and subsequently induces osteogenic differentiation via reducing IGF-1 signalling |
| Capoulade R [27]  | 2015 | Cohort study (n=220) | Elevated Lp(a) and OxPL levels are associated with faster AS progression and the need for aortic valve replacement |
| Bouchareb R [48]  | 2015 | In vitro – human VICs | Endothelial-derived NO signalling increases the expression of the NOTCH1 target gene |
| Bosse K [50]      | 2013 | In vitro – porcine VICs | Nitric oxide prevents spontaneous calcification of porcine VICs |
| First Author [Ref] | Year | Study Type | Key Findings |
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| **Key Findings**  |      |            |              |
| **AS** = aortic stenosis; **DPP-4** = dipeptidyl peptidase-4; **HDL** = high density lipoproteins; **IGF-1** = insulin-like growth factor 1; **Lp(a)** = lipoprotein(a); **MPO** = myeloperoxidase; **NADPH** = nicotinamide adenine dinucleotide phosphate; **NF-κB** = nuclear factor kappa light chain enhancer of activated B cells; **NO** = nitric oxide; **oxLDL** = oxidised low density lipoproteins; **OxPL** = oxidised phospholipids; **PCSK9** = proprotein convertase subtilisin/kexin type 9; **ROS** = reactive oxygen species; **SSAS** = semicarbazide-sensitive amine oxidase; **ST2** = interleukin 1 receptor-like 1; **TNF-α** = tumour necrosis factor-alpha; **VIC** = valvular interstitial cells. |
Oxidative Stress and Lipid Infiltration

Oxidised Phospholipids are linked to Valvular Inflammation and Calcification

The presence of oxPLs and continuing process of lipid peroxidation within valvular tissue leads to a cascade of inflammatory cell infiltrates characterised by macrophages and T-lymphocytes, and the production of pro-inflammatory cytokines such as tumour necrosis factor-alpha (TNF-α), transforming growth factor-beta 1 (TGF-β1) and interleukin-6 (IL-6) [26].

Oxidative stress also directly upregulates transcription factors leading to downstream expression of genes involved in the inflammatory process; subsequently there is positive feedback with the inflammation causing additional ROS generation that eventually promotes endothelial dysfunction and cell death, leading to a vicious circle [29].

The pro-inflammatory cytokines, coupled with a reduction in the availability of nitric oxide, disrupt the balance between the production and breakdown of the extracellular matrix in the valve [31,42]. This imbalance lays the foundations for dystrophic microcalcification [43], and the inflammatory stimuli induce VICs to express osteogenic genes such as those found in the bone morphogenetic protein 2 (BMP2) pathway [2]. This osteogenic process is not unlike the one observed in normal bone mineralisation. In fact, osteoblast-like cells have been found in calcified aortic valves [31]. In an *in vitro* study, treatment with L-Arginine, a precursor to nitric oxide, was found to be effective in preventing the osteogenic differentiation and reducing matrix calcification of VICs obtained from bovine valves [44].

OxPLs activate the toll-like receptors and NF-kB (nuclear factor kappa light chain enhancer of activated B cells) signalling pathway in VICs [2]. TNF-α contributes to this process, leading to downstream production of IL-6 (Figure 2). IL-6 then further promotes the expression of the BMP2 and receptor activator of NF-kB ligand superfamily member 11 (RANK)/RANK ligand (RANKL)/osteoprotegerin (OPG) pathways [2]. While the RANK/RANKL/OPG pathway promotes osteoclast activity in bones, the opposite is seen in valvular tissues, inducing an osteoblastic phenotype in VICs [14].

Furthermore, OxPLs upregulates alkaline phosphatase, which induces the differentiation of calcifying vascular cells; a similar process may be seen in valvular tissue [27]. Similarly, the incubation of aortic VICs with lipoprotein(a) was found to induce osteogenesis, resulting in increased apoptosis as well as a deposition in calcium [45].

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**Figure 1: Oxidative Stress and Lipid Infiltration**

- Particularly in bicuspid aortic valve
- Shear stress damages valve endothelium
- Infiltration of OxPLs

- OxPLs adhere tightly to lysine-binding sites on exposed valve surfaces
- Increased MPO catalyses formation of ROS, inducing LDL oxidation
- LDL oxidation interferes with HDL levels and uncoupling of endothelial NO synthase
The NF-κB pathway also promotes dipeptidyl-peptidase-4 (DPP-4) expression, which also induces osteogenesis in VICs by reducing insulin-like growth factor-1 (IGF-1) signalling [46]. High DPP-4 expression was found in calcified regions of aortic valves; conversely, levels were negligible in non-calcified regions. Sitagliptin, a DPP-4 inhibitor, was found to inhibit osteogenic changes in VICs in vitro, and also reduced aortic valve calcification as well as improved aortic valve indices such as aortic valve area, transaortic peak velocity as well as maximal and mean pressure gradients in animal models [46].

Autotaxin is an enzyme that is both transported in the blood plasma and secreted by several different types of cells, and it is involved in the production of lysophosphatidic acid, which is highly pro-inflammatory [47]. Autotaxin is transported into the aortic valve by lipoprotein(a), and is also secreted by VICs [48]. It contributes to the inflammatory process and subsequent promotion of osteogenesis via the NF-kB and BMP2 pathways; in fact, it can serve as an independent predictor of calcific AS [49].

**Oxidative Stress Causes Osteogenesis Via Cellular and Mitochondrial Pathways**

Besides lipid peroxidation, oxidative stress also directly promotes osteogenesis in valvular tissue via a variety of mechanisms. Endothelial-derived nitric oxide is the endogenous signalling molecule for NOTCH1, a transcriptional regulator that is involved in aortic valve development and inhibits osteogenesis [45,50]. The loss of nitric oxide reduces the activity of NOTCH1. There is then downstream activation of Lrp5 (low-density lipoprotein receptor-related protein 5), a protein involved in the osteogenic Wnt/β-catenin pathway [51]. Furthermore, ROS directly upregulates the expression of BMP2, which regulates osteogenic differentiation via Runx2 (runt-related transcription factor 2), SMAD1 (Mothers Against Decapentaplegic Homolog 1) and caspase-3 [52].

At the cellular level, oxidative stress causes damage to DNA, cellular proteins and lipids, as well as the activation of mitochondrial-driven apoptosis, and the resultant release of apoptotic bodies [15]. These apoptotic bodies facilitate the formation of hydroxyapatite crystals, which are nucleation sites for further calcium deposition [14,53]. This amorphous epitaxial mineral deposition is independent of osteogenesis and overrides antioxidant effects, causing an exponential increase in the rate of calcification [54].

As mentioned above, mitochondrial electron transport chain complexes I and III are major sources...
of superoxide degeneration; superoxide ions are converted to the more stable hydrogen peroxide by the superoxide dismutases, and crucially, levels of superoxide dismutases are significantly reduced in calcified regions of valve tissue, resulting in an increase in superoxide [15]. In vivo treatment of a rabbit model with lipolic acid, which aids the metabolism of hydrogen peroxide, abrogated aortic valve calcification [54]. Typically, levels of superoxide dismutases are increased in tissues that are exposed to elevated oxidative stress as an antioxidant defence mechanism, and its expression is found to be elevated in atherosclerotic plaques. Surprisingly however, its activity is reduced in calcified regions of the aortic valve [55].

NADPH (nicotinamide adenine dinucleotide phosphate) oxidase is a key mediator to increased oxidative stress in atherosclerosis, but its significance is debatable in AS. While an earlier landmark study found that its expression and activity was not increased in calcific aortic VICs [16], a more recent in vitro study found that specifically, levels of NADPH oxidase 2 are increased in calcific VICs [56]. In both in vitro and in vivo experiments, the same group showed that inhibition of NADPH oxidase 2 significantly reduced calcification in porcine VICs, and improved the rate of change in the aortic valve area, transaortic peak velocity as well as the maximal and mean pressure gradients in a rabbit model with calcific AS [56]. What is more universally agreed upon however is that the uncoupling of nitric oxide synthase drives a major production of ROS [57].

Asymmetric dimethylarginine is a mediator of endothelial dysfunction and a competitive inhibitor of nitric oxide synthase [58]. It induces oxidative stress via the upregulation of the renin-angiotensin system, with subsequent increases in ROS production [58]. In fact, plasma asymmetric dimethylarginine levels are correlated with severity of AS [59].

Increased levels of DRP1 (dynamin-related protein 1), a mitochondrial protein that regulates mitochondrial fission, have been found in calcified human cardiovascular cells [60]. DRP1 promotes osteogenic differentiation via oxidative stress, and its inhibition ameliorated calcification of VICs [60].

Other Mechanisms of Oxidative Stress

Oxidative stress may activate matrix metalloproteinases in the aortic valve, which in turn may contribute to the degradation of the extracellular matrix and predispose the valve to calcification [55].

Uric acid is produced by the metabolism of purines via xanthine oxidase, another enzyme involved in oxidative processes, and can exert both antioxidant or oxidative effects depending on the cellular environment [61]. In the context of degenerative AS, it contributes to endothelial dysfunction and accelerates the formation of oxLDLs; direct deposition of urate crystals in the aortic valve may accelerate the progression of degenerative AS too [6]. While levels of serum uric acid are determined by several factors such as diet, catabolism and renal function, there is a positive correlation between serum uric acid and the severity of AS indices [6].

Similarly, the metabolism of serotonin by monoamine oxidase generates ROS such as superoxide [62]. Serotonin, along with ROS, induces proliferation of VICs, leading to thickening and subsequent fibrosis and dysfunction of the valve; phenteramine, a monoamine oxidase inhibitor, in combination with fenfluramine, a serotonin reuptake inhibitor, can attenuate this effect [63].

Semicarbazide-sensitive amine oxidase (SSAO) is a mediator of oxidative stress and contributes to the development of atherosclerosis [64]. The SSAO enzyme is involved in the production of hydrogen peroxide, and is upregulated in calcified aortic valves compared to normal valves [64]. While serum SSAO levels are elevated in patients with traditional risk factors for atherosclerosis and degenerative AS – smoking, diabetes and obesity – SSAO levels within calcific valve tissue in degenerative AS are increased independent of these risk factors, and a positive correlation with the severity of AS is seen [64]. SSAO is significantly associated with PARP-1 (poly[ADP-ribose] polymerase 1) in calcified valve tissue; PARP-1 correlates with AS severity and may directly contribute to soft tissue calcification in response to inflammation and oxidative stress [65]. Immunohistochemical analysis of valve tissue found that SSAO is localised in proximity to calcified regions; with inhibition of SSAO, there was decreased calcification of valvular interstitial cells in vitro [64].

Soluble ST2 (interleukin 1 receptor-like 1) levels are positively correlated with the degree of oxidative stress and AS severity [66]. It interferes with mitofusin-1 activity, disrupting mitochondrial fusion and oxidative phosphorylation capacity in VICs [67]. This in turn may facilitate myocardial protein oxidation and inflammation. Furthermore, soluble ST2 can activate the osteogenic NF-kB pathway, and in vitro antioxidant treatment abolished these soluble ST2-mediated effects on oxidative stress and inflammation in cardiac fibroblasts [67].

Oxidative Stress in Bicuspid Aortic Valves

Bicuspid aortic valves are categorised into different subtypes based on their phenotypes [68]. Importantly, the mechanisms of progressive AS differ
based on their subtypes. In Type I – fusion of the right and left coronary cusps – oxidative stress predominates, while in Type II – fusion of the right and non-coronary cusps – endothelial dysfunction is the main contributor [69]. These however converge on the common pathway of inflammation, leading to subsequent valvular tissue remodelling and fibro-calcification.

As bicuspid aortic valves are less efficient at dissipating oscillatory shear stress compared to normal tricuspid aortic valves, accelerated endothelial dysfunction occurs. This stress induces ROS generation and lipid peroxidation [70], leading to an oxidative environment and further damaging the integrity of the extracellular matrix. Moreover, patients with bicuspid aortic valves have been demonstrated to have an abnormal antioxidant capacity, with an even greater lack of increased superoxide dismutase activity compared to patients with normal tricuspid valves [71].

Glutathione peroxidase-3 and sulfiredoxin-1 have also been found to be downregulated in endothelial cells in bicuspid aortic valves, compared to those in tricuspid aortic valves [72]. Glutathione peroxidase-3 and sulfiredoxin are involved in the reduction of hydrogen peroxide, and the absence of their protective effects contributes to increased oxidative stress, leading to DNA damage and increased apoptosis [72].

**Lipid Lowering Therapies – A Key Target?**

Given the key role of lipid peroxidation and infiltration, one of the most promising targets to combat degenerative AS was HMG-CoA reductase inhibitor therapy. One of the earlier animal studies involving rabbits with chronic hypercholesterolaemia demonstrated that atorvastatin inhibited bone mineralisation in the aortic valve by increasing the expression and activity of endothelial nitric oxide synthase [30]. A small open-label trial that followed in 2007 showed that treatment with rosuvastatin slowed the haemodynamic progression of AS [73]. However, large randomised controlled trials such as the SALTIRE [74], SEAS [75] and ASTRONOMER [76] failed to show the progress of AS with statin therapy. In fact, in the ASTRONOMER trial, lipoprotein(a) and oxPL levels were increased with rosuvastatin compared to placebo, which may have abrogated the positive effects of reduced LDL cholesterol levels. Consequently, the American Heart Association/American College of Cardiology guidelines recommend against the use of statin therapy to prevent the haemodynamic progression of AS [3].

There is some promising data regarding Proprotein Convertase Subtilisin/Kexin Type 9 (PCSK9) inhibitors in the setting of AS. PCSK9 is an enzyme that binds to and targets the LDL receptor for degradation, and a genetic association study found that a loss-of-function mutation of the PCSK9 gene was linked to lower levels of LDL cholesterol, lipoprotein(a) and a reduced incidence of AS [77]. These findings were echoed in a later genetic association study, which also found a higher level of PCSK9 expression in valvular tissues of patients with AS compared to those without [78]. Furthermore, PCSK9 inhibition in vitro reduced calcium accumulation in VICs when they were exposed to a pro-osteogenic medium [78]. In a secondary analysis of the FOURIER trial (an international, multicentre study which enrolled over 27,000 patients receiving statins for stable atherosclerotic disease and randomised them to receiving the PCSK9 inhibitor evolocumab versus a placebo), newly diagnosed or worsening AS, as well as aortic valve replacements, occurred in 63 patients. These events were associated with a higher level of lipoprotein(a), and there was a reduction in these events after the first year of treatment with evolocumab [79]. This data appears to indicate that treatment with PCSK9 inhibitors may reduce the risk of AS development or progression, however this was a post-hoc analysis involving a small number of events, and further validation requires dedicated large randomised controlled trials.

Cholesterol ester transfer protein (CETP) is a plasma protein that regulates the transfer of cholesteryl esters from HDL cholesterol to other lipoproteins; its actions lead to reduction in the plasma levels of HDL cholesterol and thus reduces its cardioprotective effects [80]. CETP is implicated in endothelial dysfunction, and induces ROS production in endothelial cells [80]. CETP activity is also found to be increased in states of high oxidative stress, such as in type 2 diabetes mellitus, and may partially explain the predisposition of diabetics to cardiovascular disease [81,82].

While CETP inhibition has been found to also significantly lower lipoprotein(a) levels, three large clinical trials investigating the use of CETP inhibitors in cardiovascular disease were terminated early due to toxicity or futility [83-89]. The only CETP inhibitor which showed a benefit was anacetrapib, which significantly reduced the incidence of major adverse coronary events compared to the placebo [90]. However, the use of CETP inhibitors in the context of slowing or reversing AS has not been studied.

Synthetic oligonucleotides are biologics used to inactivate genes involved in disease processes. There are two main approaches: using an antisense RNA
specific to the target disease-causing gene to disrupt its transcription, and using small interfering RNA fragments (siRNA) to cleave specific sequences in the mRNA transcript of the target disease-causing gene [91]. Both result in the silencing of the disease-causing gene.

Antisense oligonucleotides targeting both apolipoprotein A and apolipoprotein B have shown promise in lowering LDL and plasma lipoprotein levels. In clinical trials, investigators found that antisense oligonucleotides against apolipoprotein A significantly reduced levels of lipoprotein(a) in a dose-dependent fashion [92,93]. Antisense oligonucleotide therapy against apolipoprotein B resulted in significantly reduced lipoprotein(a) levels in multiple clinical trials involving patients with hypercholesterolaemia as well [94-100]. Their use to slow or reverse AS has not been studied. Importantly, use of antisense oligonucleotide therapy against apolipoprotein B is associated with hepatic steatosis, which will likely limit their clinical usage [101].

MicroRNAs – Targeting the Regulators of the Osteogenic Pathways

MicroRNAs (MiRNAs) are non-coding RNA sequences that are 21-23 nucleotides long, and regulate gene expression mainly via modulating mRNA translation during protein synthesis [52]. They are involved in many body pathways such as the BMP2 pathway mentioned earlier, which is implicated in osteogenesis. MiRNAs bind to the 3’ untranslated region of their target messenger RNA (mRNA) and mark them for destruction or by blocking its translation [52]. Hence, manipulation of miRNAs may be a potential avenue for therapeutics in the treatment and prevention of degenerative AS.

Like in many other disease processes, miRNAs are implicated in degenerative AS as well. Levels of miR-26a, miR-30b and miR-195 were found to be reduced in stenotic bicuspid aortic valve tissues compared to insufficient ones [102]. MiR-26a inhibits several calcification-related genes, while miR-30b reduces BMP2-induced osteogenic differentiation in VICs; the latter’s upregulation was found to reduce the risk of aortic valve calcification [103]. MiR-195 increases the expression of BMP2, RUNX2 and SMAD1, but also upregulates the expression of anti-calcification genes such as JAG2 (Jagged-2) and SMAD7 [52].

MiR-141, another inhibitor of BMP2, is also found to be under-expressed in bicuspid valves when compared to tricuspid valves, and levels are even further reduced in stenotic bicuspid valves [104].

Another major miRNA involved in the calcification of the aortic valve is miR-204, and it protects against calcification via the direct targeting of RUNX2 and SMAD4 [105,106]. Its expression was diminished in calcific aortic valve tissues compared to controls via the actions of TGF-β1 and BMP2, while its overexpression inhibited the osteogenic differentiation of VICs [106-108]. In contrast, miR-486 is upregulated in calcific aortic valves and targets a SMAD inhibitor known as SMURF2, thereby disinhibiting the SMAD pathway and resulting in the downregulation of miR-204 [109].

The effects of shear stress, which is the initiator of degenerative AS, are also modulated by miRNAs. Expression of miR-148-3p is increased in this abnormal haemodynamic environment, and upregulates the pro-osteogenic NF-kB pathway [110]. Shear stress also increases the levels of miR-214, which targets TWIST1 (twist-related protein 1), another inhibitor of osteogenic differentiation [111].

There are a number of circulating miRNAs whose levels are increased in the setting of degenerative AS. These may serve as biomarkers, however they appear to correlate more with the myocardial remodelling and dysfunction that are caused by degenerative AS [112]. Moreover, variance in the levels of valvular miRNA does not correlate directly with the changes of their plasma levels [112]. This may limit their use as potential biomarkers for the severity of AS before the onset of cardiac dysfunction.

MiRNAs have also been studied in relation to mitochondrial homeostasis, which is affected by oxidative stress and contributes to valvular calcification. An important protein in this process, DRP1, has been explored above, and it has been found that repression of miR-15a and miR-29a increases the levels of DRP1 in valvular tissues, leading to mitochondrial fission, subsequent apoptosis and contributing to calcification [113]. Incidentally, expression of miR-15a and miR-29a is reduced in stenotic valvular tissues [113].

Overexpression of protective miRNAs and inhibition of pro-osteogenic miRNAs may be potential strategies in the treatment of degenerative AS. MiRNA mimics and expression vectors can be used to increase the protective effects of select miRNAs, while sponge vectors and antisense oligonucleotides can be used to bind to and inhibit other miRNAs [114]. A recent study in a murine model of calcific AS found that delivery of a miR-34a antagonist significantly attenuated the calcification of the aortic valves [115], which hopefully can be replicated in human trials in the future.

Major challenges in the development of possible therapies centring on miRNA manipulation lie in the in vivo stability of these therapeutics, as well as their
A Note on Aortic Sclerosis

Aortic sclerosis, a precursor stage to degenerative AS, is characterised by the thickening of the aortic valve. There is remodelling of the extracellular matrix, with or without biominerization, and there is no alteration of the mechanical properties of the valve [128]. Even in the absence of calcification, VICs in sclerotic aortic valves display an osteogenic phenotype, with upregulation of pro-osteogenic proteins such as bone morphogenetic protein 4 (BMP4) and Runx2 [128]. Further osteogenic differentiation and subsequent calcification are driven by both BMP4 and tensile stretch forces, leading to a vicious circle. It can thus be thought that aortic sclerosis and degenerative AS are on the same spectrum.

There is scant literature on the role of oxidative stress in the pathogenesis of aortic sclerosis. Patients with aortic sclerosis are largely asymptomatic and challenging to identify; additionally; sclerotic human aortic valves are generally not available to investigators as they are usually only surgically replaced when symptomatic or severe AS occurs [129].

One of the studies available explores glutathione, an endogenous antioxidant that helps counteract the effects of ROS. The homeostasis between the oxidised and reduced forms of glutathione is an indicator of oxidative stress, when the balanced is tipped in favour of the oxidised form of glutathione [130]. ROS can react with the exposed cysteine residues of proteins in the aortic valve, which in turn can then be glutathionylated by reacting with the reduced form of glutathione [131]. The affected proteins can be altered in structure and function and contribute to the development of aortic sclerosis. The same study found that patients with both atherosclerosis and aortic sclerosis had an imbalance of their systemic glutathione homeostasis, with elevated levels of glutathionylated proteins in their valves [131].

Dietary Antioxidants

There is conflicting evidence regarding the benefits of dietary antioxidants such as vitamins in the prevention of other cardiovascular diseases such as ischaemic heart disease and atherosclerosis [132]; suggesting that their use in the context of AS may result in similar outcomes.

There are several challenges and limitations to antioxidant therapy in general. Antioxidant therapy should ideally be initiated before disease onset, and should be used over a long term to allow its beneficial effects to emerge [133]. Thus, their use in slowing the
progression of AS may be limited, as the main role that oxidative stress plays appears to lie in the initiation of AS rather than its propagation.

The apparent discrepancy between the success of antioxidant therapy in animal and in vitro models versus their conflicting outcomes in human studies could be due to several factors. Firstly, in vitro experiments are isolated models testing a single variable, and do not reflect the complex biochemical environment in humans. There could be off-target effects of the antioxidant in question leading to an abrogation of its desired outcomes when tested in clinical trials, or even resulting in adverse effects. For example, a Cochrane review found that vitamin A and β-carotene consumption, two types of dietary antioxidants, was associated with an increase in all-cause mortality [134].

### Table 2: Therapies targeting oxidative stress pathways in animal and in vitro models of aortic stenosis. BMP2 = bone morphogenetic protein 2; DPP-4 = dipeptidyl peptidase-4; DRP1 = dynamin-related protein 1; Lp(a) = lipoprotein(a); NADPH = nicotinamide adenine dinucleotide phosphate; NOS = nitric oxide synthase; oxLDL = oxidized low density lipoproteins; oxPL = oxidised phospholipids; VIC = valvular interstitial cells.

| First Author [Ref] | Year | Study Type | Key Findings |
|-------------------|------|------------|--------------|
| Bosse K [50]      | 2013 | *In vitro* – porcine VICs | Inhibition of NOS increases calcification |
| Choi B [46]       | 2017 | Mouse model – NOS -/- | DPP-4 inhibition reduced aortic valve calcification |
|                   |      | Rabbit model with cholesterol-enriched diet | Sitagliptin administration in a rabbit model with calcific aortic valve disease improved the rate of change in aortic valve area, transaortic peak velocity, and maximal and mean pressure gradients |
| Liberman M [54]   | 2008 | *In vitro* rabbit model with cholesterol-enriched diet | In vivo treatment with lipicolic acid, which aids the metabolism of hydrogen peroxide, decreases aortic valve calcification |
| Liu H [57]        | 2020 | *In vitro* – porcine VICs | Celastrol, which inhibits NADPH oxidase 2, significantly reduced calcification in porcine VICs |
| Nadlonek NA [123] | 2013 | *In vitro* – human VICs | oxLDLs increased the expression of phosphate inorganic transporter 1 and BMP2, with resultant calcium and phosphate deposition. Inhibition of phosphate inorganic transporter 1 with phosphonoformate hexahydrate acid prevented oxLDL-induced BMP2 expression. |
| Rajamannan NM [30] | 2005 | Rabbit model with chronic hypercholesterolaemia | Atorvastatin inhibits bone mineralisation in the aortic valve by increasing the protein expression and functional activity of endothelial NOSs |
| Ratazzi M [44]    | 2020 | *In vitro* – bovine VICs stimulated to acquire a pro-calcific phenotype with endotoxin | L-Arginine, a precursor of NO, prevents osteogenic differentiation and reduces calcification |
| Rogers MA [58]    | 2017 | *In vitro* – human VICs | DRP1 inhibition attenuates VIC calcification |
| Zheng KH [41]     | 2019 | *In vitro* – human VICs incubated in an osteogenic medium with Lp(a) and OxPL | Incubation with the E06 monoclonal antibody against OxPL reduced osteogenic differentiation of VICs |

### Table 3: Clinical studies of therapies targeting the oxidative stress pathways in the treatment of aortic stenosis. ACE = angiotensin-1 converting enzyme; ARB = angiotensin receptor blocker; AS = aortic stenosis; DPP-4 = dipeptidyl peptidase-4; Lp(a) = lipoprotein(a); PCSK9 = proprotein convertase subtilisin/kexin type 9.

| First Author [Ref] | Year | Study Type | Key Findings |
|-------------------|------|------------|--------------|
| Bergmark BA [79]  | 2020 | Randomised controlled trial; post-hoc analysis (n = 27,564) | Overall hazard ratio for AS events patients on evolocumab (PCSK9 inhibitor) was significantly lower after a year of treatment |
| Capoulade R [119] | 2013 | Observational study (n=338) | ARBs, but not ACE-inhibitors, were associated with slower AS progression in patients with concurrent hypertension and AS |
| Chan KL [76]      | 2010 | Randomised controlled trial (n=269) | Rosuvastatin 40mg once daily did not reduce the progression of AS in patients with known asymptomatic AS |
| Cowell SJ [74]    | 2005 | Randomised controlled trial (n = 155) | Atorvastatin 80mg once daily did not prevent the progression of AS nor induce regression in patients with known AS |
| Dicht W [127]     | 2008 | Randomised controlled trial (n=47) | Atorvastatin 20mg once daily did not prevent the progression of AS in patients with known asymptomatic AS |
| Lee S [122]       | 2020 | Retrospective analysis (n=212) | In diabetic patients with mild-moderate AS, use of linagliptin or gemigliptin was associated with a slower rate of progression of maximal transaortic velocity as compared to patients on alloglipin, sitagliptin or vildaglipin, as well as patients not on any DPP-4 inhibitors |
| Moura LM [73]     | 2007 | Prospective open label (n=121) | Rosuvastatin 20mg once daily slowed the haemodynamic progression of AS in patients with known asymptomatic AS |
| O’Brien KD [126]  | 2005 | Retrospective analysis (n=123) | Treatment with ACE-inhibitors slowed the rate of aortic valve calcium accumulation |
| Rosenhek R [125]  | 2004 | Retrospective analysis (n=211) | Treatment with statins, but not ACE-inhibitors, slowed the rate of AS progression in patients with known AS |
| Rossebo AB [75]   | 2008 | Randomised controlled trial (n=1873) | Simvastatin and ezetimibe did not reduce the composite outcome of death from cardiovascular causes, aortic-valve replacement, nonfatal myocardial infarction, hospitalization for unstable angina pectoris, heart failure, coronary-artery bypass grafting, percutaneous coronary intervention, and nonhemorrhagic stroke in patients with asymptomatic AS |
| Shavelle DM [124] | 2002 | Retrospective analysis (n=65) | Treatment with statins slowed progression of aortic valve calcium accumulation measured by electron-beam computed tomography |

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Secondly, lab feed that is provided to animals involved in experiments may be deficient in dietary antioxidants that are readily available in the normal human diet [135]. Thus, the apparent positive effects of antioxidant administration may simply be the correction of an artificial vitamin deficiency. Finally, antioxidants themselves may become oxidised in the body and lose efficacy, and there could be difficulties in achieving therapeutic concentrations of the antioxidant while avoiding the threshold of toxicity [136].

Patients with AS typically also have other comorbidities such as hypertension and hypercholesterolemia. It is possible that these patients are already benefiting from the antioxidant effects of their medications such as aspirin, statins, beta-blockers and ACE inhibitors/ARBs, and further antioxidant therapy may not have additional effects [136].

The innate intracellular activity of superoxide dismutase and other endogenous enzymes would outcompete most other antioxidant agents that are used intracellularly, hence intracellular scavenging of ROS as a therapeutic target is unlikely to have much benefit [135]. Extracellularly however, superoxide dismutase mimics have higher kinetic rate constants compared to non-enzymatic reactions, and may be of potential use as a ROS scavenger in the context of AS [135]. In fact, there is some early, exciting data of a recently approved superoxide dismutase mimic preventing aortic sclerosis in a murine model [137]. Glutathione peroxidase mimics are also currently undergoing clinical trials in a range of diseases such as bipolar disorder and Meniere’s disease [135]. Perhaps these may be translated to human research for AS in the near future.

**Conclusion**

Oxidative stress is a key denominator in the processes of inflammation, lipid infiltration and fibro-calcification in aortic stenosis, with distinct differentiation to the similar process of atherosclerosis. Statin therapy, while beneficial in the context of atherosclerosis, have demonstrated disappointing results in the slowing of AS progression. Other lipid lowering therapies such as PCSK9 inhibitors and CETP inhibitors may hold promise, however larger trials involving the use of these drugs are necessary. ARBs and DPP-4 inhibitors, which are commercially available prescription medications, also warrant further trials.

Manipulation of miRNAs involved in the pro-osteogenic pathways are also attractive targets, however this avenue is currently limited by the shortcomings in the stability and delivery of miRNA mimics, vectors and inhibitors, as well as their potential off-target effects.
Potential therapies that were studied for degenerative AS may have been commenced too late in the course of the disease to be effective. Hence, aortic sclerosis may provide an opportune target for pharmacological intervention. In fact, further osteogenic differentiation of VICs isolated from human aortic valves were amenable to reversal via antioxidant enzymes delivery [129].

As the pathways of oxidative stress in degenerative AS are complex, perhaps targeting a single component may not lead to clinically significant results. There is some evidence that targets outside the oxidative stress pathway, such as inhibition of cadherin 11, matrix metalloproteinases and even the use of direct oral anticoagulants (dabigatran, apixaban and rivaroxaban) may slow the progression of AS [138-139]. Although oxidative stress in itself forms a major part of the pathophysiology of AS, it is not the full picture. Hence, a multimodal approach addressing several targets both inside and outside the oxidative stress pathway may be the way forward.

It is of worth to note that while antioxidants do provide a defence against oxidative stress, excessive antioxidants can lead to a phenomenon known as reductive stress, which, over time, is linked to pathological myocardial remodelling, development of diastolic dysfunction, and subsequent heart failure [140].

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Competing Interests

The authors have declared that no competing interest exists.

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