Bat trypanosomatids (first report of *T. wauwau*) in Triângulo Mineiro, Brazil

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Abstract

In this study, trypanosomatids commonly found in bats, including *Trypanosoma cruzi marinkellei*, *T. dionisii*, and *Leishmania braziliensis*, were identified. Additionally, *T. wauwau* was identified in one specimen of *Anoura caudifer*, and represents the first report of this parasite from the Central West region of Brazil. *T. wauwau* was previously identified by other researchers in the North of the country, in only three species of bats in the genus *Pteronotus*: *P. parnellii* (Pará and Rondônia states), and *P. personatus* and *P. gymnnonotus* (Rondônia). The identification of *T. wauwau* indicates how different
trypanosomatids are able to adapt to new host species of bats. This is owing to bats’ high mobility, wide geographic distribution, social behavior, and ability to coexist in large colonies. These characteristics may facilitate the transmission of infectious agents in nature, which are responsible for outbreaks of some zoonoses. Therefore, health authorities should focus on both vertebrates and vectors associated with the environments where these bats are found.

Author summary

The prevalence of *Trypanosoma* in bats is high, with *T. cruzi*, *T. cruzi marinkellei*, and *T. dionisii* as the most prevalent infective species. This study reports for the first time the presence of *T. wauwau* in the southeast region of Brazil in the bat *Anoura caudifer*. Although this species of *Trypanosoma* has been found in bats of the genus *Pteronotus*, it was not detected in any other genus, including in the bats that share the same shelter with *Pteronotus*. The species *T. wauwau* was found infecting bats only in Brazil. Its occurrence was restricted to the northern region of the country, in the states of Pará, infecting the species *P. parnellii* and in Rondônia infecting *P. personatus*, *P. gymnonotus* as well as *P. parnellii*. Although its morphology is similar to that of *T. cruzi*, little is known about the development of *T. wauwau*, both in its vertebrate host and the existence of a plausible invertebrate vector. Its characteristics include its inability to develop in mammalian cells and its non-infectiousness in mice and triatomine insects. Further research, through molecular studies, may provide important and valuable data for understanding the origin, evolution, and global distribution of, and the association between the different species of *Trypanosoma* and their hosts.
Introduction

Until now, more than up to 30 species of trypanosomatids have been isolated from bats [1–4]. The most frequently reported species of *Trypanosoma* in bats are *T. cruzi*, *T. cruzi marinkellei*, *T. dionisii*, *T. hedricki*, *T. myoti*, *T. leonidasdeanei*, *T. desterrensis*, *T. pifanoi*, *T. pessoai*, *T. megaderma*-like, *T. theileri*, and *T. rangeli* [5–10]. An additional species, *T. wauwau*, was also recently described in Brazil [9–10]. Another group of bat-related trypanosomatids are those in the genus *Leishmania*, with *L. infantum chagasi* reported infecting bats in Venezuela [11], later, *L. mexicana* was found in the bats in southeastern Mexico [12] and the species *L. amazonensis*, *L. braziliensis*, and *L. infantum chagasi* were found in Brazil[3, 13–16]. Because of the detection of such medically important trypanosomatids in bats, epidemiological surveillance research is necessary in regions with endemic and non-endemic disease-causing species that may pose a risk of human infection.

Given the above observations and concerns, the objective of the present study was to evaluate the presence and identity of the possible trypanosomatid species (*Trypanosoma* spp. and *Leishmania* spp.) in bats of the Triângulo Mineiro Region of Brazil.

Material and methods

Area of study and capture of bats

This study was carried out in the city of Ituiutaba, Minas Gerais (MG), Brazil, located in the Triângulo Mineiro mesoregion (lat 18°58'08" S, lon 49°27'54" W; altitude: 544 m; area: 2595.2 km²), west of MG, Brazil.

Bats were captured from November 2014 to September 2015, at night between 18h00 and midnight (0h00) using mist nets, and during the day in shelters using manual
nets. The identification of bats species was done based on taxonomic keys to the family [17], genus, and species levels [18–19].

Blood collection

A total of 216 bats were collected, and 0.5 to 1.0 mL of blood was collected from each specimen by cardiac puncture. Of these samples, 25 µL was used to estimate microhematocrit, while the remaining blood was stored in EDTA V/V and guanidine solution (6 M Guanidine–HCl and 0.2 M disodium) at 4°C until further use.

Identification of trypanosomatid species

DNA extraction was performed using a GeneJET Genomic DNA Purification® kit from Thermo Scientific, according to the manufacturer's instructions.

For detecting Leishmania DNA, the primers HSP70F and HSP70R (5'CCGCCCATGCTCTGGGTACATC 3') were used, whose target is the HSP70 gene of Leshmania spp. [20]. For detecting Trypanosoma DNA, a Nested PCR was performed, in which the primers used in the first reaction were TRY927F (5'-GAAACAAGAAACACGGGAG-3') and TRY927R (5'-CTACTGGGCAGCTTGGA-3'), and those used in the second reaction were SSU561F (5'-TGGGATAAACAAGGGAGCA-3') and SSU561R (5'-CTGAGACTGTAAACCTCAAAGC- 3'), whose target was the 18S rDNA region [21]. Electrophoresis was performed on 6% polyacrylamide gel, stained with silver. Sequencing was performed at ACTGene Análises Moleculares (Brazil). Chromatograms were analyzed using ChromasPro 2.1.4, and consensus sequences were generated. The Phred threshold value was set at > 20. Sequences were aligned using SeaView 4.5.2, with the Muscle algorithm. Maximum Likelihood trees using Neighbor-Joining methods were generated using MEGA7: Molecular Evolutionary Genetics Analysis version 7.0 for
bigger datasets [22], with 1000 bootstrap iterations, and including reference sequences from trypanosomatids retrieved from GenBank.

**Statistical analyses**

The Chi-square test was used to assess the possible association between the dietary habits of bat hosts and the positivity of their PCR tests for both *Leishmania* and *Trypanosoma*, using the TIBCO® Statistica™ program [23], with a significance level of 5% (P < 0.05).

**Ethical considerations**

Bats were captured and manipulated according to the recommendations of the Brazilian Institute of Environment, with authorization for activities with scientific purpose number 45132-1 (IBAMA-SISBIO). The procedures used were approved by the Animal Research Committee of the Federal University of Triângulo Mineiro, as per protocol No. 51.

**Results**

**Identification of *Trypanosoma* species**

The captured animals (216) belonged to nine different bat species. Of these, 43 (19.90%) presented positive microhematocrit results for trypanosomatids. Among the blood samples analyzed, 18 (8.33%) were found positive for *Trypanosoma* spp. DNA, most (77.77%) of which came from nectarivorous bats (Table 1). However, according to the results of the Chi-square test, there was no significant statistical association between the dietary habits of hosts and infection with *Trypanosoma* spp. (p = 0.205) or *Leishmania* spp. (p = 0.635).
Table 1. Bat species captured in Minas Gerais, Brazil by family, feeding habits, sex and positivity

| Family       | Species             | Feeding habit | ♀/♂ | Total (%) | MIC | Leishmania sp. PCR | HSP70 | Trypanosoma sp. PCR TRY/SSU |
|--------------|---------------------|---------------|-----|-----------|-----|--------------------|-------|----------------------------|
| Molossidae   | Molossus molossus   | Insectivorous | 0/2 | 2 (0.93%) | 0   | 0                  | 0     | 0                          |
| Phyllostomidae | Anoura caudifer    | Nectarivorous | 36/18 | 54 (25.11%) | 4   | 3                  | 6     | 6                          |
|              | Artibeus lituratus  | Frugivorous   | 0/1  | 1 (0.46%)  | 1   | 0                  | 0     | 0                          |
|              | Carollia brevicauda | Frugivorous   | 2/3  | 5 (2.32%)  | 2   | 0                  | 0     | 0                          |
|              | Carollia perspicillata | Frugivorous    | 9/9  | 18 (8.37%) | 5   | 0                  | 0     | 0                          |
|              | Desmodus rotundus   | Hematophage   | 8/4  | 12 (5.58%) | 5   | 0                  | 3     | 8                          |
|              | Glossophaga soricina | Nectarivorous | 67/50 | 117 (54.41%) | 24  | 6                  | 8     | 18 (8,33%)                |
|              | Phyllostomus discolor | Omnivorous       | 1/0  | 1 (0.46%)  | 0   | 0                  | 1     | 1                          |
|              | Phyllostomus hastatus | Omnivorous     | 2/4  | 6 (2.79%)  | 2   | 0                  | 0     | 0                          |
| Total        |                     |               | 216  | 43 (19,90%) | 9   | 4 (4,46%)          | 18 (8,33%) |                       |

MIC – microhematocrit
♀/♂ – males/females
HSP70 – primer specific primer for Leishmania sp. research
TRY/SSU - specific primers for Trypanosoma sp. research

The species of bats with the highest infection rates were *Desmodus rotundus*, *Anoura caudifer*, and *Glossophaga soricina* (25.00%, 11.11%, and 6.83%, respectively).

The species of trypanosomes identified were *T. cruzi marinkellei* and *T. dionisii* in various bat species, and *Trypanosoma wauwau* in *Anoura caudifer* (Table 2, Figure 1).

Table 2. Trypanosomatids isolates, isolate code, bat hosts and primers used to DNA sequencing

| Trypanosomatids species | Quantity | Individual isolate code | Host                        | Primer used       |
|-------------------------|----------|--------------------------|-----------------------------|-------------------|
| *Leishmania braziliensis* | 2        | Mo205Bac/ Mo224Bac       | *Anoura caudifera*          | HSP70Leish        |
| *Leishmania braziliensis* | 1        | Mo110Bac                  | *Phyllostomus discolor*     | HSP70Leish        |
| *T. cruzi marinkellei*   | 4        | Mo220ID/ Mo254/ Mo256/ Mo258 | *Glossophaga soricina*     | TRY927/SSU561    |
| *T. cruzi marinkellei*   | 1        | Mo225                     | *Desmodus rotundus*         | TRY927/SSU561    |
| *T. cruzi marinkellei*   | 1        | Mo261                     | *Anoura caudifera*          | TRY927/SSU561    |
| *Trypanosoma dionisii*   | 2        | Mo176/ Mo218              | *Anoura caudifera*          | TRY927/SSU561    |
| *Trypanosoma wauwau*     | 1        | Mo184                     | *Anoura caudifera*          | TRY927/SSU561    |
The phylogenetic analysis was performed using the Maximum Likelihood method based on the Tamura-Nei model with 1000 bootstraps. A total of 1954 bases were analyzed together. All positions containing gap were eliminated. The phylogenetic analyses were performed in the MEGA7 program. The arrows indicate the species found in the sequencing of the present study.

**Identification of *Leishmania* species**

Of the samples analyzed, nine (4.46%) were positive for *Leishmania* spp. (Table 1). The only species of *Leishmania* identified was *L. braziliensis*, in the bats *Anoura caudifer* and *Phyllostomus discolor* (Table 2).
Discussion

Some species of trypanosomatids have already been found to naturally infect bats [4, 7–10, 24–27]. Many of these trypanosomatids are responsible for important zoonoses, but most of those found in bats remain poorly known. In this work, we investigated the presence of *Leishmania* spp. and *Trypanosoma* spp. in bats from the city of Ituiutaba, Triângulo Mineiro, Brazil, an area previously showing endemism for Chagas disease.

In this study, the use of the direct parasitological method of microhematocrit revealed frequent positive results for trypanosomatids, as has been demonstrated by other studies [4, 28]. It is worth highlighting that the microhematocrit method, in all the field studies carried out by our group, especially in small animals, has shown greater sensitivity or, in certain cases, similar behavior to that of traditional blood culture methods [29].

Regarding PCR positivity, the prevalence of trypanosomatids was similar to that observed by other studies in Brazil [3–4], as was the result that in terms of dietary habits most of these bats were nectarivores. It was also demonstrated in this study that the two most common species that circulate in bats in this region are *T. c. marinkellei* and *T. dionisii*, as has been previously demonstrated [4, 6, 30].

The only species of *Leishmania* found was *L. braziliensis*, in *Anoura caudifer* and *Phyllostomus discolor*. This is the first report of this species of *Leishmania* in these species of bats. In this region, the species *L. infantum*, *L. amazonensis*, and *L. braziliensis* were previously identified from the bat species *G. soricina* and *M. molossus* [3].

In this study, it was possible to identify *T. wauwau* in only one specimen of *Anoura caudifer* (a nectarivore of the family Phylostomatidae), with this being the first report of this parasite in the Central-West region of Brazil, and in a different species of bat from those previously reported to harbor it. This *Trypanosoma* was recently identified in Brazil in only three species of bats, all in the genus *Pteronotus*: *P. parnellii* in Pará and
Rondônia states, and *P. personatus* and *P. gymnonotus* in Rondônia only [9–10]. In none of our field studies in this region of the Triângulo Mineiro and Alto Paranaiba have we found bats of the genus *Pteronotus* spp. These bats are found in Brazil, in the Amazonian, *cerrado* and Atlantic forest biomes, living mainly near water sources, in caves, and under bridges. *Anoura caudifer*, in which *T. wauwau* was detected, is quite common in Brazil, and can be found in all biomes, particularly in humid forests and in areas with both primary and secondary vegetation.

*T. wauwau* is phylogenetically associated with Australian trypanosomes, possibly constituting one more piece of evidence that *T. cruzi* may have evolved from the recent dispersion of an ancestral bat host across several continents [9]. The finding of bats positive for a disease at a place where that disease is not endemic, rather is introduced from neighboring areas, demonstrates the considerable mobility of these animals, often involving migrations over long distances, including to urban areas, thereby acting as potential agents of zoonoses.

**Conflict of interest:** The authors declare that they have no conflicts of interest.

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