A Case of Late-onset Cerebrotendinous Xanthomatosis with a Novel Mutation in the CYP27A1 Gene

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Abstract:
Cerebrotendinous xanthomatosis (CTX) is a rare, autosomal recessive, inborn disruption in bile acid synthesis characterized by severe systemic xanthomas, cataracts and neurological injuries occurring before adolescence without elevation of the serum cholesterol or triglyceride levels. CTX is caused by a deficiency of the mitochondrial enzyme sterol 27-hydroxylase, which is encoded by the CYP27A1 gene. We herein report a 50-year-old Japanese woman with late-onset CTX who had no relevant symptoms before the development of bilateral Achilles tendon xanthomas in middle age. A genetic analysis revealed a compound heterozygous mutation in the CYP27A1 gene with a previously known missense mutation (NM_000784.3:c.1421 G>A) and a novel frame shift mutation of NM_000784.3:c.1342_1343insCACC.

Key words: cerebrotendinous xanthomatosis (CTX), sterol 27-hydroxylase (CYP27A1), cholestanol, norcholic acid, chenodeoxycholic acid (CDCA), cholesterol efflux

Introduction
Cerebrotendinous xanthomatosis (CTX) is an autosomal recessive lipid storage disease caused by disruption in bile acid synthesis induced by mutations in the CYP27A1 gene on chromosome 2q33-3qter (1). Fifty-four different mutations in CYP27A1 have been reported thus far (2). The first case of CTX was described in 1937 by Van Bogeaer (3). Since then, more than 300 cases have been reported in countries around the world. There is no sex predominance. CTX is an extremely rare disease; its prevalence was estimated as 1.9 per 100,000 people among Caucasians of European ancestry (2). The estimated incidence of CTX is higher in East Asia, at approximately 15.5 per 100,000 people (4). However, precise epidemiologic data from Japan are lacking.

The CYP27A1 gene encodes the mitochondrial enzyme sterol 27-hydroxylase, which hydroxylates the steroid side chain at C-27 (5). Deficiency of this enzyme leads to the accumulation of cholestanol and bile alcohols in connective tissues and neuronal cells and results in systemic symptoms, such as neurological disorders, atherosclerosis, tendon xanthomas and cataracts. Patients with CTX typically develop cataracts before their 10th birthday and develop systemic xanthomas in the second or third decade of life. Although most CTX patients have normal intelligence until puberty, intellectual functioning worsens with age, and over 50% of CTX patients show an intellectual decline in their 20s.
A slowly progressive neurologic symptoms, such as pyramidal tract signs, cerebellar ataxia and peripheral neuropathy, are typically observed in their 20s and 30s (2). There is significant clinical heterogeneity, even among siblings with the same mutation in the CYP27A1 gene; as such, the clinical phenotypes do not seem to be strictly correlated with the genotypes in CTX (2).

Chenodeoxycholic acid (CDCA) has been demonstrated to decrease the levels of serum cholestanol and urinary bile alcohols in CTX patients (6). Treatment with CDCA can stabilize or even reverse some of the symptoms in most patients with CTX (7). Starting CDCA during childhood or earlier is believed to be more beneficial than starting treatment in adulthood (8).

We herein report a Japanese woman diagnosed with CTX at age 50 who had no related symptoms of CTX before the development of bilateral Achilles tendon xanthomas in middle age.

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Case Report

Case presentation

The patient was a 50-year-old Japanese woman from non-consanguineous parents. She has no siblings but had a child. There was no apparent family history (parents, child and parent’s siblings) of neurological disorders, cardiovascular diseases or any lipid storage diseases inducing xanthomas, including CTX. She had no history of neonatal jaundice or chronic infantile diarrhea. She had been diagnosed with idiopathic epilepsy and medicated from 7 to 10 years of age. Thereafter, she had no remarkable medical developments until middle age. She insidiously developed painless tendon thickness of the bilateral Achilles tendons and right triceps muscle at 47 years of age. The gradual enlargement of the Achilles tendons caused gait disturbance when she was 49 years of age. T1- and T2-weighted magnetic resonance imaging (MRI) revealed low-intensity lesions in the thickened bilateral Achilles tendons (Fig. 1A) and right triceps muscle,
suggestive of xanthomas. She was admitted to Nagasaki University Hospital for further investigation at 50 years of age.

**Clinical, laboratory and imaging examinations**

The patient was lean (height 149.0 cm, weight 39.2 kg, BMI 17.7 kg/m²), and the findings of a physical examination were unremarkable except for confirmation of xanthomas on the bilateral Achilles tendons (Fig. 1B) and right triceps muscle. She had age-appropriate slight cataracts in the bilateral eyes and did not have dementia based on the Revised Hasegawa Dementia Scale (HSD-R; 26/30), Mini-Mental State Examination (MMSE; 27/30), and Frontal Assessment Battery (FAB; 16/18) scores. On a neurological examination, her gait appeared somewhat spastic, and all of her deep tendon reflexes were mildly enhanced bilaterally with positive Babinski and Chaddock reflexes, indicating impairment in the bilateral pyramidal tracts. There were no neurological findings suggesting cerebellar impairment.

Regarding the laboratory findings (Table 1), there was no elevation in the levels of serum cholesterol or triglycerides or deficiencies in apolipoproteins or thyroid hormones. As the clinical findings suggested CTX, we studied the bile acids and sterols in her serum and urine (Table 2). The primary bile acids were extremely low in the serum and urine when measured by both high-performance liquid chromatography (HPLC) and gas chromatography/mass spectrometer (GC/MS). The urinary levels of bile alcohols, especially 5β-cholestan-3α,7α,12α,24,25-pentol, were markedly elevated. Norcholic acid, which is mainly derived from 5β-cholestane-3α,7α,12α,23,25-pentol, was highly elevated and accounted for 78.4% of all bile acids in the urine. The serum level of cholestanol was extremely elevated, while those of sitosterol and campesterol were not elevated.

T2-weighted and fluid-attenuated inversion recovery (FLAIR) MRI of the brain showed bilateral low-intensity areas in the dentate nuclei and high-intensity areas in the peripheral white matter of the cerebellum (Fig. 1C). We also found bilateral high-intensity areas along the pyramidal tracts from the pons to the cerebral crus. Brain atrophy was not remarkable. Based on these clinical, laboratory and radiological findings, the patient was diagnosed with CTX.

Ultrasound of the carotid arteries showed a thickened intima-media complex at the left common carotid artery (maximum intima-media thickness, 1.2 mm). However, there was no evidence of cardiovascular disease on electrocardiogram, echocardiography and MR angiography of the brain and lower extremities.

Despite being premenopausal (Table 1), she had osteoporosis as determined by dual-energy X-ray absorptiometry, which showed a bone density of 61% and 75% the young-adult-mean in the lumbar vertebra and femoral neck, respectively.

**Genetic diagnosis**

Genomic DNA was isolated from peripheral blood leukocytes. All 9 exons of the CYP27A1 gene were amplified by polymerase chain reaction (PCR) and sequenced. Two muta-

### Table 1. Serum Lipids and Associated Laboratory Findings on Admission.

| Case | Reference | Case | Reference |
|------|-----------|------|-----------|
| Total cholesterol (mg/dL) | 142-248 | Total bilirubin (mg/dL) | 0.4-1.5 |
| Triglyceride (mg/dL) | 123 | Direct bilirubin (mg/dL) | 0.2 ≤ 0.4 |
| HDL-C (mg/dL) | 75 | AST (IU/L) | 16-30 |
| LDL-C (mg/dL) | 103 | ALT (IU/L) | 7-23 |
| Apo A-1 (mg/dL) | 167 | γ-GTP (IU/L) | 8-9.32 |
| Apo A-II (mg/dL) | 28.1 | BUN (mg/dL) | 9.0-8.20 |
| Apo B (mg/dL) | 80 | Creatinine (mg/dL) | 0.52-0.46-0.79 |
| Apo C-II (mg/dL) | 7.0 | Calcium (mg/dL) | 9.1-8.8-10.1 |
| Apo C-III (mg/dL) | 11.2 | Phosphate (mg/dL) | 3.0-2.7-4.6 |
| Apo E (mg/dL) | 6.3 | Free T4 (ng/dL) | 1.27-0.95-1.57 |
| Free cholesterol (mg/dL) | 56 | TSH (μIU/mL) | 0.43-0.48-5.08 |
| Free fatty acid (mEq/L) | 0.59 | Intact PTH (pg/mL) | 69.0-10.3-65.9 |
| Phospholipid (mg/dL) | 243 | 25(OH)D3 (ng/mL) | 14-30-60 |
| Lipoprotein(a) (mg/dL) | 11.4 ≤ 40 | 1,25(OH)2D3 (pg/mL) | 48-20-60 |
| MDA-LDL (U/L) | 119 | 46-105 | TRACP-5b (mU/L) | 385-120-420 |
| SAA-LDL (μg/dL) | 5.8 <20 | Intact P1NP (μg/L) | 66.2-17.1-64.7 |
| Glucose (mg/dL) | 80 | LH (IU/L) | 37.5-1.4-15 |
| Insulin (mIU/L) | 2.3 | FSH (IU/L) | 39.1-3.0-24.0 |
| HbA1c (%) | 5.3 | Estradiol (pg/mL) | 360.1-28.8-491.9 |

HDL-C: high-density lipoprotein cholesterol, LDL-C: low-density lipoprotein cholesterol, Apo: apoprotein, MDA-LDL: malondialdehyde-modified LDL, SAA-LDL: Serum amyloid A-LDL, AST: aspartate aminotransferase, ALT: alanine aminotransferase, γ-GTP: gamma-glutamyltransferase, BUN: blood urea nitrogen, TSH: thyrotropin, PTH: parathyroid hormone, TRACP-5b: tartrate resistant acid phosphatase-5b, P1NP: procollagen type 1 N-terminal propeptide, LH: luteinizing hormone, FSH: fertilizing hormone.
Table 2. Serum Levels of Bile Acids and Associated Biosynthetic Products before and 6 Months after Administration of CDCA.

| Before | After |
|--------|-------|
| Serum bile acid (HPLC; µmol/L) | Serum bile acid (GC/MS; µmol/L) |
| Glycoursoxycholeoycholic acid | N.D. | Total bile acid | 0.10 | 6.82 |
| Taurosoxycholeoycholic acid | N.D. | Cholic acid | N.D. | N.D. |
| Ursodeoxycholeoycholic acid | N.D. | Chenodeoxycholic acid | N.D. | 3.80 |
| Glycocholeoycholic acid | N.D. | Ursodeoxycholic acid | N.D. | 2.10 |
| Taurocholeoycholic acid | N.D. | Deoxycholeoycholic acid | N.D. | N.D. |
| Choloycholic acid | N.D. | Liihcholeoycholic acid | N.D. | N.D. |
| Glycocholeoycholic acid | N.D. | Norcholeoycholic acid | 0.10 | N.D. |
| Taurocholeoycholic acid | N.D. | Others | N.D. | 0.92 |
| Chenodeoxycholeoycholic acid | N.D. | | | |
| Glycocholeoycholic acid | N.D. | Urinary bile acid (GC/MS; mmol/mol-Cr) | | |
| Taurocholeoycholic acid | N.D. | Total bile acid | 0.50 | 1.09 |
| Deoxycholeoycholic acid | N.D. | Cholic acid | 0.03 | N.D. |
| Glycolithicholeoycholic acid | N.D. | Chenodeoxycholic acid | 0.01 | 0.12 |
| Tauroliihtioleoycholic acid | N.D. | Ursodeoxycholeoycholic acid | N.D. | 0.64 |
| Lithiitioleoycholic acid | N.D. | Deoxycholeoycholic acid | N.D. | N.D. |
| | | Liihcholeoycholic acid | N.D. | 0.06 |
| | | Norcholeoycholic acid | 0.39 | 0.04 |
| | | Others | 0.07 | 0.23 |

CDCA: chenodeoxycholeoycholic acid, HPLC: high performance liquid chromatography, GC: gas chromatography, GC/MS: gas chromatography mass spectrometry, N.D.: not detectable, Cr: creatinine

Figure 2. A direct nucleotide sequence analysis of the exon 8 of CYP27A1 gene. A: The patient was heterozygous for the missense mutation of NM_000784.3: c.1421 G>A, resulting in the amino acid substitution of arginine to glutamine at codon 474 (NP_000775.1: p.Arg474Gln), as previously reported (Ref. 9). B: The patient was also heterozygous for a frame-shift mutation of NM_000784.3: c.1342_1343insCACC (NP_000775.1: p.Arg448ProfsTer521), which has never been reported before.

...tions were identified. One was a known missense mutation of NM_000784.3:c.1421GA in exon 8 (Fig. 2A), resulting in an amino acid substitution of arginine to glutamine at codon 474 (R474Q), as reported previously (9, 10). The other was a novel frame-shift mutation of NM_000784.3:c.1342_1343insCACC in exon 8 (Fig. 2B). As we did not perform genetic analyses of these two mutations in the patient’s parents, we were unable to determine whether the patient was a compound heterozygote of two mutated alleles or had double mutations in one allele. Therefore, we assumed the patient to be a compound heterozygote of these two mutations based on the clinical and laboratory findings.

Cholesterol efflux assay

A cholesterol efflux assay was carried out as described previously (11). In brief, ATP-binding cassette transporter A1 (ABCA1)-transfected cell were incubated with [3H]-cholesterol. After incubation, the radioactive counts in media and cell lysates were measured by liquid scintillation counting. The ABCA1-mediated cholesterol efflux capacity, expressed as the percentage of labeled cholesterol moved from the cells to the medium, was 11.0% in the patient (reference...
Treatments

After CTX was diagnosed, the patient was treated with CDCA at 500 mg per day, divided into 2 doses. Alendronate at 35 mg per week and alfacalcidol at 0.5 μg per day were prescribed to treat her osteoporosis. These treatments are ongoing. She underwent bilateral Achilles-tendon plastic surgery, and the gait disturbance was improved dramatically after surgery. A pathological examination of the excised tissue revealed foamy cells admixed with inflammatory giant cells and cholesterol clefts surrounding tendinous tissues (Fig. 1D).

The serum levels of several bile acids were increased to detectable ranges, and the serum cholesterol and urinary norcholic acid levels were dramatically decreased six months after the administration of CDCA was started (Table 2). The serum cholesterol levels remained substantially low for at least 18 months after the initiation of CDCA treatment (4.5 and 5.4 μg/mL at 12 and 18 months, respectively). The patient has shown no regrowth of the tendon xanthomas or any other complications in the post-treatment period.

Discussion

We here present a case of late-onset CTX with compound heterozygous mutations with a known missense and a novel frame shift mutation in the CYP27A1 gene.

The pathogenesis of neuronal injury and other involvement in CTX has not been fully clarified. The leading hypothesis is that accumulation of cholesterol and cholestanol in the plasma membrane induces apoptosis of the neurons, and accumulation in other tissues causes xanthomas, juvenile-onset atherosclerosis and cataracts (12, 13). However, our patient was almost lacking in neurological and atherosclerotic findings other than the slight pyramidal-tract impairment and senile cataract. It is impossible to exclude that she had spinal cord involvements because we did not evaluate the spinal cord by MRI before starting CDCA treatment. The idiopathic epilepsy medicated in her childhood might be a symptom of CTX. It is unclear why she mainly developed xanthomas without other symptoms seen in CTX until middle age.

It was reported that sterol 27-hydroxylase expressed in macrophages enhances the cholesterol efflux capacity (14, 15). 27-hydroxycholesterol is a ligand of liver X receptor α (LXRα) (16) and plays a role in up-regulating the cholesterol efflux via the ABCA1 pathway (17). In this context, it would be of interest to know whether or not the cholesterol efflux is impaired for atherosclerotic formation in patients with sterol 27-hydroxylase deficiency, i.e. CTX. Although we performed an ABCA1-mediated cholesterol efflux assay in the present CTX patient, the efflux capacity was found to be relatively normal, which may partially account for the absence of apparent atherosclerotic disease in our case. More data are needed to clarify the relationship between sterol 27-hydroxylase and the cholesterol efflux in patients with CTX.

It has been reported that patients with CTX tend to develop osteoporosis (18). Our patient developed osteoporosis before menopause, and we found that her serum level of 25(OH)VitD was markedly decreased. Sterol 27-hydroxylase is one of several enzymes with vitamin D 25-hydroxylase activity (19), which may be a molecular mechanism underlying the development of osteoporosis in CTX. A previous report in a small group of CTX patients suggested an association of low serum 25(OH)VitD levels with osteoporosis (20), although other studies have reported relatively normal levels of 25(OH)VitD (18, 21). Therefore, the role of 25(OH)VitD in osteoporosis in CTX remains controversial (22).

Although a fundamental treatment for CTX has not yet been established, CDCA was shown to stabilize and even reverse some of the associated symptoms of CTX (2, 8). This is because the supplementation of CDCA, a primary bile acid, induces negative feedback in the catabolism of the various sterol intermediates (6). The abnormally high production of cholestanol thereby decreases, relieving its accumulation in the plasma membranes of neural cells. Indeed, it was reported that CDCA treatment improves the neurological manifestations and abnormal findings of brain MRI in CTX patients (8, 23). Our CTX patient mainly developed xanthoma of the tendons; as such, her neurological, neuropsychiatric, and cardiovascular manifestations might have been slowly progressive compared to other reported cases (8, 24, 25). As the serum levels of cholestanol decreased after starting CDCA, we remain hopeful that further complications of CTX can be prevented in this case.

Conclusion

We reported a middle-aged patient with late-onset CTX harboring a novel mutation in the CYP27A1 gene. It is important to begin CDCA treatment in patients with CTX before the onset of neurological dysfunction in order to prevent irreversible neuronal injury. Clinicians should consider CTX as a differential diagnosis when encountering patients with xanthomas of unknown etiology.

The authors state that they have no Conflict of Interest (COI).

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