Evaluation of phytochemicals and activity index of some plant leaf extracts on typhoidal and non-typhoidal Salmonella isolates from selected hospitals in Bauchi, Nigeria

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Abstract
The phytochemicals and activity index evaluation was done for antibacterial activity of aqueous and methanol extracts of Cymbopogan citratus, Psidium guajava and Anacadium occidentale on clinical isolates of typhoidal and non typhoidal Salmonella. The typhoidal Salmonella isolates were S. typhi and S. paratyphi A, while non typhoidal was S. typhimurium. Well diffusion method was used. The activity of extracts was compared with that of orthodox drugs commonly used for the treatment of typhoid and non-typhoid salmonellosis thus; ciprofloxacin (10µg/Disc), amoxicillin (30µg/disc) and chloramphenicol (30µg/Disc) as standard. Phytochemical analysis of extracts was conducted using Trease and Evans methods of 2002. Both methanol and aqueous extracts had significant (p<0.05) in vitro activity Comparison of extract with antibiotics show activity index (AI) as follows: Chloramphenicol have AI≤1 on the salmonella serotypes of both solvent extracts implying that Chloramphenicol is still of use. Amoxacilin have AI>1 on the serotypes indicating that Amoxacillin is resistant to salmonella serotypes tested. Using Ciprofloxacin as a standard, AI varies with serotypes S. paratyphi A and S. typhimurium have AI≤1 and S. typhi have AI>1. Pythochemical screening of extract revealed the presence of some bioactive components like alkaloids saponin, tannins, anthraquinones steroid flavonoids glycosides. These properties determined the antimicrobial potential of the extracts. From the findings, it proved that plants crude extracts possess some potential as antibiotics and can be further studied to isolate the compound that is most active and formulated as drug against the disease.

Keywords: Phytochemicals; Activity Index (AI); Typhoida; antibiotics.

1. Introduction
Salmonellosis is a disease caused by salmonella species which are members of the family enterobacteriaceae. They are gram negative facultative anaerobic rods. Salmonella species are classified into serovars (serotypes) based on the lipopolysacharide (O) flagellar protein (H) and sometimes the capsular (VI) antigens. There are more than 2500 known serovars, within a serovar, there may be strains that differ in virulence [9] Salmonella is generally divided into two categories; non typhoidal and typhoidal. The non typhoidal salmonella is the most common form and is carried by both humans and animals. Most serotypes of salmonella such as Salmonella jariana and Salmonella enteritidis cause non typhoidal salmonellosis. Salmonella typhimurium is involving in the invasive non typhoidal. [9].

The serovars responsible for typhoid fever is restricted to human beings, which is transmitted through direct contact with the feacal matter of an infected person, that is to say it’s transmitted through feaco-oral route. Typhoid fever is endemic in developing and under developed world where unsanitary conditions are more likely to prevail and which...
can affect as many as 21.5 million people, each year. Recorded cases of typhoid fever in the developed world are mostly related to recent travel to areas where, salmonella Typhoid is endemic.

Typhoid fever symptoms appear between 8 – 14 days after eating contaminated food and last anywhere from 3 to 60 days. They include fever, weakness, lethargy abdominal pain, coughing, nose bleeding and delirium and enlarged organs. Typhoid fever is a serious illness that can result in death [4].

Herbal medicine is still the main stay of about 75-80% of the world population, particularly, in developing countries for primary health care [8]. This is primarily because of the general belief that herbal drugs are without any side effects. Besides, they are cheap and locally available. Before scientists made impact into the research for drugs curing human infections, the traditional means of treating diseases were done by means of concoctions from plants either in single form or mixtures without knowing that the agents were used against some pathogenic micro-organism [17]. Limited knowledge about the practices of use of plant medication that is herbal medicine and lack of scientific studies of plants have led to the neglect of novel bioactive components that may bring about remarkable result in treatment of infectious diseases with little or no side effect [15].

Medicinal plants are known to contain in one or more of its organ substances that can be used for therapeutic purpose or as precursors for synthesis of useful drugs [17]. Many of such plants known to be used primitively to alleviate symptoms of illness have been screened to have medicinal importance, some of which include Azadirachta indica (Dogonyaro), Zingiber officinale (Ginger), Piper guineese (Iyere) Allium sativiam (Garlic), Vernonia amydalina (Bitter leaf). These plants have been reportedly used in the traditional treatment of ailments such as stomach disorder, fever symptoms and cough [10].

Besides this efficacy, plants have little or no side effects in treatment of diseases because they act as food and as medicines. In the treatment of hypertension for instance herbs are used first to lower the blood pressure to clean the arteries, to slow and regulate the heart beat rate, to improve the circulation of blood and relax the mind, un like the fundamental conventional drugs that will dilate the arteries or the veins until they reach maximum elastic point which may suddenly burst and cause vascular accident, causing stroke or death [11]. Typhoidal salmonella which is the most resistant to antibiotics of the serovars, hence, finding the right antibiotic for it is a crucial matter; as a result, alternative means must be employed to find solutions to this problem such as use of herbs. This actually predates the introduction of antibiotics and modern drugs in Africa. The existence of human beings without the availability of plants would have been made very difficult. Different plants produce different compounds which vary in their antimicrobial action and organisms differ in their sensitivity to these compounds.

In the last three decades, the search for natural bioactive compounds that can serve as antimicrobial agents had increased tremendously due to the increasing resistance possessed by microorganisms to synthetic antibiotic [12].

The scope of this study includes isolation of Salmonella, extraction of African lemon grass (Cymbopogan Citratus) guava leaf (Psidium guajava) cashew leaf (anacardium Occidentale) and the testing of the efficacy of these extract on the isolate.

2. Material and methods

2.1. Ethical Clearance

The consent of the clinically suspected patients was sort, while the Ethics Committee of Abubakar Tafawa Balewa University teaching Hospital Bauchi gave approval for the study. Confidentiality of the subjects’ identities was duly maintained.

2.2. Study Area

The study area is Bauchi Local Government Area, where samples were collect from new General Hospital Bayara, Specialist Hospital and ATBU Teaching Hospital, Bauchi state is located on Lat. 10.6371 °N, long. 10.0807 °E the vegetation is that of savanna and sahel according to Koppen’s climat classification system. Bauchi LGA is located on Lat. 10.301 °N long. 9.8237 °E, it’s area is 3,687 km2 and has population of 493,810 according to 2006 population census.
2.3. Sample collection

2.3.1. Clinical samples
A total of 210 clinical samples were collected, blood and stool. Samples were collected from patient with febrile or diarrheal illness attending new General Hospital Bayara, Specialist Hospital and ATBU Teaching Hospital, Bauchi on weekly basis from May to August 2016. Blood sample were collected using syringe and needle while stool samples were collected using sterile bottles, patient where served with sterile bottles to collect a small quantity of their stools.

2.3.2. Plants collection and identification
Plant samples collected or used are African lemon grass (*Cymbopogan citratus*), Cashew leaf (*Anacardium Occidentale*) and guava leaf (*Psidium guajava*). These were collected at Mbak area of Dass town a local government in Bauchi state where they are found to be growing naturally or planted deliberately and transported to ATBU herbarium for authentication with the following voucher numbers; *Psidium guajava* ATBUHB 2489, *Anacardium Occidentale* ATBUHB 2490 and *Cymbopogan citratus* ATBUHB 2491.

2.4. Isolation, Characterization and Identification of Typhoidal and Non-Typhoidal Salmonella
Stool sample were inoculated in a non-selective broth, selenite F enrichment broth and incubated at 37 °C for 24hrs, the pre cultured stool sample were sub cultured on deoxycholate citrate agar (DCA), salmonella shigella agar (SSA) agar and brilliant green agar (BGA) and incubated at 37 °C for 18-24hrs where growth was detected based on their, colonial characteristic and morphological appearance on these media.

Blood samples were inoculated into Terathionate broth. A minimum of blood to broth ratio of 1 in 10mls, was maintained, and this was incubated at 37 °C and checked for signs of bacterial growth daily for up to seven days. Bottles that show signs of growth were subculture on to Brilliant Green Agar (BGA) *Salmonella Shigela Agar* (SSA), and Mac conkey Agar.

Blood culture broth with no bacterial growth after seven days was sub-cultured before being reported as negative result. Typical colonies of Salmonella appear as pink colonies with or without black centers. Many cultures of *Salmonella* produce colonies with large, glossy black centers or may appear as almost completely black colonies. [21].

Figure 1 Map of Bauchi State Indicating Bauchi LGA
2.5. Biochemical screening and serology

A range of biochemical tests were used to confirm the suspected colonies on the selective agar, these are Triple sugar iron agar (TSI), Urease, Indole and Simmons Citrate test [21].

Colonies considered to be *salmonella* ssp were further tested for somatic (O) and flagella (H) antigens polyvalent antisera (oxoid) [5]

| S/N | Test Substrate                  | Positive                  | Negative                  | Salmonella |
|-----|---------------------------------|---------------------------|---------------------------|------------|
| 1   | TSI (glucose)                   | Yellow Bult               | Red bult                  | +          |
| 2   | H2S (TS I lysine decarboxylase) | Blackening                | No blackening             | +          |
| 3   | Urease                          | Purple colour             | No colour change          | +          |
| 4   | Indole test                     | Violet colour at the surface | Yellow colour at the surface | -          |
| 5   | Simmons citrate                 | Growth blue colour        | No growth, No blue colour | V          |
| 6   | Polyvalent flagellate test      | Agglutination             | No agglutination          | +          |
| 7   | Polyvalent somatic test         | Agglutination             | No agglutination          | +          |

**Table 1** Biochemical and Serology test of Salmonella (carried out)

Key: V = Variable

2.5.1. Antibacterial Sensitivity of Orthodox Drug for Typhoid Treatment

The susceptibility testing was carried out by disc diffusion method using Mueller Hinton agar and it was tested in vitro for susceptibility to the following antibiotics (OXOID Ltd., UK) suggested by WHO, 2010, Ciprofloxacin (CPX, 10 µg), Amoxicillin (AMC, 30 µg) and Cloramphenicol (CH, 30 µg) [22].

2.5.2. Plant Sample Preparation

The leaves and grass were washed, drained and air dried at room temperature and grounded to powder using mortar and pestle. All the powdered samples were labeled and stored in a dark polythene bags at room temperature prior utilization.

2.5.3. Extraction Procedure of Plants Sample Prepared

100grams of each powder plant sample stored were weighed and extracted with methanol and aqueous (Water) for 3-6 hours using soxhlet extractor [3]. This was done by wrapping the powdered plant material in filter paper and tucked in to the soxhlet tube and the solvent in the soxhlet flask. The set was then placed on a hating mantle under electric current and allowed to run for the above mentioned hours until extraction is complete by comparing colour difference of the tube (plant material) with that of the flask (extracts).

2.5.4. Preparation of Inoculum

Macfarland standard; 0.5 macfarland standard was prepared by 0.5ml of 1% Barium chloride (BaCl₂) to 99.5mls of 1% Sulphurics acid (H₂SO₄). The turbidity of 0.5 macfarland stands was used for the estimation of the amount of salmonella. Colonies of Salmonella was suspended in sterilized 0.9% sodium chloride solution (normal saline) which was compared with 0.5 macfarland solution the microbial suspension (1 ml) in normal saline was added to 74 mls of sterile medium kept at 45 °C to give bacterial population density of 1.2×10⁷ Cells/ml [14].

2.5.5. Preparation of Stock Suspension of Extract

Stock preparation of plant extract was prepared by dissolving 0.4grams of extract in to 1ml of dymethyl sulfoxide (DMSO) for methanolic extracts and 1ml sterilized distilled water was used for aqueous extracts to make 400mg/ml and stored at room temperature pending usage.

2.5.6. Assays for Antibacterial Activity of Plant Extracts

The antibacterial activity was carried out by well diffusion method [6]. Preparation of inoculums was obtained using Macfarland standered to have inoculums density of 1.2×10⁷ CFU/mls, wells of 4mm in diameter were bored on already inoculated Mueller Hilton agar plates using a sterile well borer and 60µL of extracts was dispensed in the wells and
stand for 40 min., as pre-diffusion time, these were incubated at 37 °C for 18-24 hr. Diameters of zones of inhibition were determined by subtracting the diameter of the wells and recorded to the nearest millimeter.

Preliminary testing of this preparation was carried out using the stock concentration 400 mg/ml to test whether or not the plant is active. The antibacterial activity assay was done in duplicate at concentrations 400, 200, 100, 50 and 25 mg/ml were the means taken as the mean zones of inhibition (MZI).

To investigate the synergistic activity of the extracts equal volume of 0.5 ml from the stock preparation of both solvents, this combination was made in two and the three and diluted to make concentration of 400, 200, 100, 50 and 25 mg/ml.

2.5.7. Determination of the Susceptibility of the Isolates to Some commonly used Antibiotics

Susceptibility pattern of some commonly used antibiotic was carried to compare with that of the extract using disc diffusion method [22]. Out of the commercially available disc drug of choice for typhoid were cut out, these are ciprofloxacin (CPX) 10 µg, Amoxicillin (AM) 30 µg and chlorphenicol (CH) 30 µg. These were placed on inoculated Mueller Hillton agar plates; zones inhibitions were measured after 24 hr incubation at 37 °C. This was carried out in duplicates and then the mean zone of inhibitions was used to calculate the activity index (AI) = The mean zones of inhibition for the crude extracts test divided by the mean inhibition zones for the commercial antibiotics [1].

2.6. Phytochemical Analysis of Extract

Phytochemical screening of methanol and aqueous (water) extracts of *Cymbopogan citratus*, *Anacadium occidentale* and *Psidium guajava* was done using standard procedure to determine the bioactive agents in the extract [7, 16, 19].

2.6.1. Test for Alkaloids: Dragendorff’s Test

0.5 g portion of the leaves powder was stirred in 5 ml of 1% aq. HCl on a steam bath for about 5 minutes. The mixture was filtrated through a Whatman No. 1 filter paper. 2-4 drops of Dragendorff’s reagent was added to 1 ml of the filtrate. A change to orange color indicates the presence of alkaloids. Being alkaline, alkaloids are readily extracted by mildly acidic aqueous solvents like 1% aq. HCl. The test is valid only if the plant material is not colored.

2.6.2. Wagner’s Test

To the 1 ml of extract, add 2 ml of Wagner’s reagent. Reddish brown colored precipitate indicates the presence of alkaloids.

2.6.3. Test for Saponins

To about 5 ml distilled water in a test tube, 0.5 g of plant extract was added, shaken vigorously and observed, for frothing (small bubbles), the mixture was warmed by standing in a water bath (50 °C) for 10 minutes. Persistence of frothing on warming is an indication of the presence of saponins.

2.6.4. Test for Tannins

To 10 ml distilled water in a test tube, 5 g of the plant extract was added, stirred and then filtered through a Whatman No. 1 filter paper. 2-3 ml of ferric chloride solution was added gradually to the filtrate. A deep green color (olive-green) indicates the presence of tannins.

2.6.5. Test for Flavonoids

0.5 g portion of the extracts was added to 2 ml dilute NaOH solution in a test tube and shaken to dissolve. Then few drops of conc. H₂SO₄ were added. A colorless solution is an indication of flavonoids.

2.6.6. Test for Anthraquinones

To 10 ml dilute H₂SO₄ in test tube 5 g of extracts was added and boiled for a few minutes. Then filtered through Whatman No. 1 filter paper while the mixture is still hot. 5 ml of ether was added to the filtrate and shaken to mix. It was allowing standing until the ether and aqueous layers separate out, then, 2.5 ml of 10% ammonia solution was added. A pink or red or violet color in the aqueous layer indicates the presence of anthraquinones.
2.6.7. **Test for Steroids**

To 1mg of the extract in a test tube, 5ml chloroform added. Then, equal volume of conc. H₂SO₄ was added by sides. The turning of red in the upper layer and yellow with green fluorescence in the sulphuric acid layer indicates the presence of steroids.

2.6.8. **Test for Glycosides (most classes)**

To 10ml boiling distilled 5g of the extracts was added. Stirred and then filtered through Whatman No. 1 filter paper. A few drops of conc. HCl were added to 2ml portion of the filtrate. Boil for a few minutes to hydrolyze any glycosides present, a few drops of aqueous ammonia solution was added to make the mixture alkaline. Then five drops of the mixture to 2ml of Benedict’s reagent was added and boiled. A reddish brown precipitate indicates the presence of glycosides.

2.6.9. **Test for Phlobatannins**

Few drops of 1% HCl was added to 1ml of extract and boiled. A reddish precipitate indicates the presence of phlobatannins.

2.6.10. **Test for Terpenoids**

2ml of chloroform was added to 0.5g of the plant extract; follow by adding 3ml H₂SO₄. Reddish brown coloration at the interface which indicates the presence of terpenoids.

2.6.11. **Test for Resins**

To 2ml of extract plus equal volume of acetic anhydride solution was added and then drops of conc. H₂SO₄. Formation of colophony resins (violet coloration) indicates the presence of resins.

2.7. **Determination of Activity Index of the Plant Extracts**

Activity index was used to compare the inhibitory effect of the extract with that of the commercially available antibiotics. This was calculated as the mean zones of inhibition for the test extract divided by the mean inhibition zones for the commercial antibiotics [1, 2].

3. **Results and discussion**

3.1. **Physical Properties of Plant Extracts**

Some of the physical characteristics of plant extracts such a colour texture and their dry weight are presented on Table 2. The plant part used are the leaf or green part of the plants. The initial weight of the powdered plant material was 100 grams prior extraction, after the extraction, the colour of extract appeared green with methanol and brown to dark brown with aqueous. The of consistency varied, methanol extract of *Anacadium occidentale* was sticky and oily with dry weight 37.5g, *Psidium guajava* was also sticky with dry weight 26.82 and *Cymbopogan citratus* was creamy and soft to touch with dry weight 17.78g. The aqueous extracts have these varying properties *Anacadium occidentale* gummy and oily with dry weight 16.17g.

| Table 2 Physical Properties of Aqueous and Methanol Extracts of Plants |
|---------------------------------|
| **Plant Species** | **Part** | **Colour** | **Name of consistency** | **Initial weight in (grams)** | **Dry weight in (grams)** |
|-------------------|---------|-----------|------------------------|------------------------------|--------------------------|
| **Methanol Extract** |          |           |                        |                              |                          |
| *Anacadium occidentale* | leaf    | green     | Sticky/Oily            | 100                          | 37.5                     |
| *Psidium guajava*    | leaf    | green     | Sticky                 | 100                          | 26.82                    |
| *Cymbopogan citratus*| grass   | green     | Creamy/soft on touch   | 100                          | 17.78                    |
| **Aqueous Extract**  |          |           |                        |                              |                          |
| *Anacadium occidentale* | leaf    | Dark brown| Gummy/Oily            | 100                          | 16.17                    |
| *Psidium guajava*    | leaf    | brown     | Flakes                 | 100                          | 8.14                     |
| *Cymbopogan citratus*| grass   | brown     | Gummy                  | 100                          | 20.94                    |
3.2. Antibacterial Activity of Orthodox Antibiotics

The effect of most used orthodox antibiotics for the treatment of typhoid fever was used as standards. Ciprofloxacin (10µg), Amoxacilin (30µg) and chromplanicol (30µg) were tested on typhoidal and non typhoidal Salmonella isolates to compare their activities of various plant extracts against the isolates. This is as presented on Table 3. Chloramphenicol have a significant zone of inhibition on the three Salmonella serotypes at various levels. While ciprofloxacin (10µg) exhibited highest effect on S. paratyphi A with mean zone of inhibition 20.5mm followed by S. typhimurium 11mm and Amoxicilin (30µg) has highest activity on S. typhimurium with mean zone of inhibition 9mm.

Table 3 Sensitivity Tests of some Antibiotics Used for the Treatments of Typhoidal and Non Typhoidal Salmonellosis

| Antibiotics in (µg)      | Mean Zone of inhibition in (mm) |
|--------------------------|---------------------------------|
|                          | S. typhi | S. paratyphi | S. typhimurium |
| Ciprofloxacin (CPX) 10   | 1        | 20.5         | 11             |
| Amoxacilin (AM) 30       | 2        | 3            | 9              |
| Chloramphenicol (CH) 30  | 11.5     | 19           | 15             |

3.3. The Phytochemical Screening of Plants Extracts

Photochemical Screening of plant Cymbopogan citratus showed positive result for the ten compounds tested except for flavonoids with aqueous extract and plabotanins with methanol extract having negative results. Psidium Guajava showed positive result on the entire compound tested except for Resins with aqueous extract; Antraquinone and plabotanins of methanol extracts had negative results. Anacadium occidentale also showed positive result on all the compounds tested except for resins of aqueous extracts, antraquinones and plabotanins of methanol extract that gave negative results.

3.4. Phytochemical Analysis

The phytochemical present in the extracts of the three plants, Anacadium occidentale, Psidium guajava and Cymbopogan citratus with methanol and aqueous solvent shown on Table 3 revealed positive result for most of the of the bioactive compounds which may be the reason for the antibacterial activity as described by many researchers [20,6]. The bioactive compounds positive to the screening of both aqueous and methanolic extract were Alkaloids Saponins flavonoids steriods, glycosides and terpenoids with negative result on methanol extracts for Antraquinones and Phlabotanins also in aqueous extracts of Resins.

Table 4 Phyochemical Screening of plant extracts

| Bioactive Compound | Cymbopogan citratus | Psidium guajava | Anacadium occidentale |
|--------------------|----------------------|-----------------|-----------------------|
|                    | AE       | ME       | AE       | ME       | AE       | ME       |
| 1. Alkaloids       | +        | +        | +        | +        | +        | +        |
| 2. Saponins        | +        | +        | +        | +        | +        | +        |
| 3. Tanins          | +        | +        | +        | +        | +        | +        |
| 4. Flavonoids      | -        | +        | +        | +        | +        | +        |
| 5. Antraquinones   | +        | +        | -        | +        | -        | -        |
| 6. Steroids        | +        | +        | +        | +        | +        | +        |
| 7. Glycosides      | +        | +        | +        | +        | +        | +        |
| 8. Plabotanins     | +        | -        | +        | -        | +        | -        |
| 9. Terpenoids      | +        | +        | +        | +        | +        | +        |
| 10. Resins         | +        | +        | -        | +        | -        | +        |

Key: AE=Aqueous Extract ME=Methanol Extract

Bioactive compounds are known to possess antimicrobial properties [1]. Tennins have been found to form irreversible complexes with protein - rich compounds resulting in inhibition of cell protein synthesis. This is an important effect
for the treatment of inflamed or ulcerated tissue,[2] and can be used in ulcerated surfaces frequently experience by typhoid patient especially in perforated intestines and arthritis among the frequently occurring complication [18] while Tannins act by Coagulating the cell wall protein synthesis, Saponins are surface active agents which alter the permeability of the cells thus facilitates the flow of toxic materials or leakages of vital cell constituents, flavonoids being phenolic in nature are cytoplasmonic poisons, inhibiting the activity of enzyme [19] which are of benefit to the pathogenesis caused by Salmonella organism Tannins containing herbs are astringent in nature and are used for astringents intestinal disorders such as dysentery and diarrhea [13] which is a major sign and symptoms of typhoid and non-typhoid Salmonellosis.

3.5. Analysis of the Activity Indices of Extracts Using Standard Antibiotics

The activity index of extracts with the standard antibiotics is presented on Table 5.

Table 5 Activity Indices (AI) of Plant Extract at Concentration (400mg/ml) in Comparison with Antibiotics use for Treating Typhoid Fever

| Antibiotics                | Plant                      | Organism          | MZI of Antibiotics | Methanol | Aqueous |
|----------------------------|----------------------------|-------------------|--------------------|----------|---------|
| Ciprofloxacin (10 µg/disc) | Cymbopogan citratus        | S. typhi          | 1                  | 9        | 11      |
|                            |                            | S. paratyphi A    | 20.5               | 0.24     | 0.32    |
|                            |                            | S. Typhimurium    | 11                 | 1.27     | 1       |
|                            | Psidium guajava            | S. typhi          | 1                  | 17       | 16      |
|                            |                            | S. paratyphi A    | 20.5               | 0.82     | 0.90    |
|                            |                            | S. Typhimurium    | 11                 | 1        | 1.27    |
|                            | Anacadium Occidentale      | S. paratyphi A    | 20.5               | 0.95     | 0.82    |
|                            |                            | S. typhi          | 1                  | 19       | 13      |
| Amoxacillin (30 µg/disc)   | Cymbopogan citratus        | S. typhi          | 2                  | 4.5      | 5.5     |
|                            |                            | S. paratyphi A    | 3                  | 1.66     | 2.16    |
|                            |                            | S. typhimurium    | 9                  | 1.55     | 1.22    |
|                            | Psidium guajava            | S. typhi          | 2                  | 8.5      | 8       |
|                            |                            | S. paratyphi A    | 3                  | 5.66     | 6.16    |
|                            |                            | S. typhimurium    | 9                  | 1.22     | 1.55    |
|                            | Anacadium Occidentale      | S. paratyphi A    | 2                  | 9.5      | 6.5     |
|                            |                            | S. typhi          | 3                  | 6.5      | 5.66    |
|                            |                            | S. typhimurium    | 9                  | 1.88     | 1.88    |
| Chloramphenicol (30 µg/disc)| Cymbopogan citratus        | S. typhi          | 11.5               | 0.78     | 0.95    |
|                            |                            | S. paratyphi A    | 19                 | 0.45     | 0.34    |
|                            |                            | S. typhimurium    | 15                 | 0.93     | 0.73    |
|                            | Psidium guajava            | S. typhi          | 11.5               | 1.47     | 1.39    |
|                            |                            | S. paratyphi A.   | 19                 | 0.89     | 0.97    |
|                            |                            | S. typhimurium    | 15                 | 0.73     | 0.93    |
|                            | Anacadium Occidentale      | S. typhi          | 11.5               | 1.65     | 1.13    |
|                            |                            | S. paratyphi A    | 19                 | 1.02     | 0.89    |
|                            |                            | S. typhimurium    | 15                 | 1.13     | 1.13    |

Key: MZI = Mean Zone of Inhibition

Using ciprofloxacin (10µg) as standard, the indices of all the extracts were less than unity (AI<1) on S. paratyphi A. with both methanolic and aqueous extracts followed by S. typhimurium which is slightly greater than unity (AI>1) with both methanol and aqueous extracts while S. typhi is highly greater than unit (AI>1). With Amoxacillin (30µg) as a standard all extracts had indices greater than unity (AI>1) on all the three salmonella serotypes. Using
chloramphenicol (30µg) as a standard. S. paratyphi A showed index less than or equal to unity (AI ≤ 1) while S. typhi showed less than unity (AI<1) with cymbopogon citratus extracts indices was greater than unity (AI>1) with anacadium occidentale and psidium guajava extracts. S. typhimurium has indices less than unity (AI<1) with cymbopogon citratus and psidium guajava extracts indices was greater than unity (AI>1) with anacadium occidentale extracts indices was at unity (A=1).

3.6. Comparison of the Activity of the Plant Extracts with the Orthodox Antibiotics

The result of the investigation indicates that the extracts when compared with the standard antibiotics showed significant activity. The high activity indices above unity value in the crude forms of extracts is an indication of more promising therapeutic advantage than the likes of Amoxacillin and ciprofloxacin when refined to produce antibiotics especially in typhoid infection with serotype Salmonella typhi, the indication is that the extracts generally have a higher activity than the commercial antibiotics as a single crude drug with respect to S. typhi which had activity indices, above unity in comparison with the above mentioned antibiotics.

4. Conclusion

Extract showed significant in vitro activity as compared with the antibacterial activities of orthodox antibiotics, hence the pharmacological bases of use of these plants for treatment of typhoid fever. A base line data has been generated with respect to activity indices of these plants. The finding proved that chloramphenicol is still of value in the treatment of typhoid fever. Confirmed earlier results of other independent studies about activity of plants tested against the pathogen as promising. Plants prepared as drink can be packed into tea bags and used for treatment of typhoid and non-typhoid fevers.

Recommendation

Purification of photycheical compounds should be carried out to remove impurities and to identify the active compounds in the various plants. Pure compounds should be compared with the current antimicrobial agents or compounded together to yield better drugs and also take care of the problem of drug resistance that is presently becoming a problem globally.

Compliance with ethical standards

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Disclosure of conflict of interest

The authors whose names are mentioned above declare that they have no conflict of interest as far as this publication is concerned, and should there be any conflict that may arise, it can be resolved amicably. The research is solely sponsored by the corresponding author.

Statement of ethical approval

In compliance with the ethical standard the consent of the clinically suspected patients was sort, while the Ethics Committee of Abubakar Tafawa Balewa University teaching Hospital Bauchi gave approval for the research. Confidentiality of the subjects’ identities was duly maintained.

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