Beta diversity patterns of fish and conservation implications in the Luoxiao Mountains, China

Jiajun Qin¹*, Xiongjun Liu²³*, Yang Xu¹, Xaioping Wu¹²³, Shan Ouyang¹

¹ School of Life Sciences, Nanchang University, Nanchang 330031, China ² Key Laboratory of Poyang Lake Environment and Resource Utilization, Ministry of Education, School of Environmental and Chemical Engineering, Nanchang University, Nanchang 330031, China ³ School of Resource, Environment and Chemical Engineering, Nanchang University, Nanchang 330031, China

Corresponding author: Shan Ouyang (ouys1963@qq.com); Xaioping Wu (xpwu@ncu.edu.cn)

Academic editor: M.E. Bichuette | Received 27 August 2018 | Accepted 20 December 2018 | Published 15 January 2019

http://zoobank.org/9691CDA3-F24B-4CE6-BBE9-88195385A2E3

Citation: Qin J, Liu X, Xu Y, Wu X, Ouyang S (2019) Beta diversity patterns of fish and conservation implications in the Luoxiao Mountains, China. ZooKeys 817: 73–93. https://doi.org/10.3897/zookeys.817.29337

Abstract
The Luoxiao Mountains play an important role in maintaining and supplementing the fish diversity of the Yangtze River Basin, which is also a biodiversity hotspot in China. However, fish biodiversity has declined rapidly in this area as the result of human activities and the consequent environmental changes. Beta diversity was a key concept for understanding the ecosystem function and biodiversity conservation. Beta diversity patterns are evaluated and important information provided for protection and management of fish biodiversity in the Luoxiao Mountains. The results showed that the spatial turnover component was the main contributor to beta diversity of Hemiramphidae, Amblycipitidae, Catostomidae, Clariidae, Balitoridae and Percichthyidae in the Luoxiao Mountains, which indicated that a number of protected areas would be necessary to conserve fish biodiversity and that these families would need conservation measures. Most protected areas are currently limited to some regions; therefore, in order to protect fish diversity, conservation efforts must target an increase in the number of protected areas which should be spread across each of the regions.

Keywords
beta diversity, commercial fishes, Luoxiao Mountains, protected areas

*Contributed equally as the first authors.

Copyright Jiajun Qin et al. This is an open access article distributed under the terms of the Creative Commons Attribution License (CC BY 4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.
Introduction

Biodiversity patterns and their formation mechanisms have been one of the hot issues, and it is also an important foundation for conservation (Kennedy and Norman 2005; Sutherland et al. 2009). Biodiversity is important for the future sustainability of freshwater natural resources (Hiddink et al. 2008). While it is axiomatic that biodiversity is essential for sustainable productive fisheries there is surprisingly little supporting evidence (Dulvy et al. 2000; Hilborn et al. 2003). Freshwater fishes are among the most diverse assemblages on Earth, which provide important economic value (e.g., nutrition) and valuable ecosystem services (e.g., natural water filtration; Naylor et al. 2000; Cressey 2009; De Silva 2012). However, due to dam construction, overfishing (commercial fish fishing), pollution, deforestation, and other human activities, fish numbers have declined rapidly in global terms (Fu et al. 2003; Arthington et al. 2016; Liu et al. 2017) and they are thus one of the most threatened assemblages.

Beta diversity is an important tool for conservation planning (Anderson et al. 2006); knowledge on beta diversity patterns can aid the decision on the number of protected areas needed and their sizes (Margules and Pressey 2000; Wiersma and Urban 2005). Beta diversity can be decomposed into species turnover (species replacement) and nestedness (richness difference; Baselga 2010; Carvalho et al. 2012). The species turnover component (species replacement) is the replacement of some species by others leading to a low number of shared species among two communities where turnover is high (Baselga 2010). In addition, the nestedness component (richness difference components) represents the differences between two communities only in terms of species richness, with the poorer community as a subset of the richer one (Baselga 2010). According to the percentage of spatial turnover and nestedness components in total beta diversity, different conservation strategies can be selected. If species turnover is the main component of beta diversity, a larger number of protected areas would be necessary to conserve regional biodiversity (Baselga 2010; Carvalho et al. 2012). If the nestedness is the main component of beta diversity, one large protected area comprising a high species richness could be sufficient (Baselga 2010; Carvalho et al. 2012).

The Luoxiao Mountains range is located in the southeast of China’s mainland and has a long history and complex environmental factors (Liao et al. 2014; Wei et al. 2015). The northern part of the mountains is connected with the Yangtze River, and the southern part is connected with the Nanling Mountain (Gong et al. 2016). It is the most important ecotone and fragile zone in the third step of eastern China, and is an important channel for the migration and diffusion of terrestrial organisms in the Northern Hemisphere (Liao et al. 2014; Gong et al. 2016). In addition, the Luoxiao Mountains is also a biodiversity hotspot in China (Liao et al. 2014; Gong et al. 2016). At the same time, as being the watershed of the Poyang Lake Basin and the Dongting Lake Basin in the middle reaches of the Yangtze River, the Luoxiao Mountains are a refuge to many endemic and endangered fishes (Liao et al. 2014; Gong et al. 2016). Therefore, fish resources of the Luoxiao Mountains play an important role in maintaining and supplementing the aquatic biodiversity of the Yangtze River Basin. However, due to dam construction, overfishing, pollution,
deforestation, and other human activities, fish diversity declined rapidly in this region. Here, we aim to evaluate beta diversity patterns and to provide useful information for the protection and management of fish biodiversity in the Luoxiao Mountains.

**Material and methods**

**Study area**

The Luoxiao Mountains (25°32’–29°28’N, 113°09’–114°26’E) are a large system of mountain ranges, located in the southeast of China’s mainland with an overall north-south trend, stretching across Hubei, Hunan, and Jiangxi provinces. It consists of Mufu Mountain, Jiuling Mountain, Wugong Mountain, Zhuguang Mountain, and others. The total length of the Luoxiao Mountains is 400 km and altitude ranges are 82–2120 m. Lingfeng Peak (2122 m) is one of the highest mountains in the southeastern Eurasia. Its average precipitation range is 1341–1943 mm and forest coverage in the watershed reaches 90% (Table 1). The tributaries of the Ganjiang River from the eastern stream of the Luoxiao Mountains flow into Poyang Lake. The tributaries of Xiangjiang River from the western stream of it flow into the Dongting Lake. The Fushui River alone flows into the Yangtze River.

**Sampling methods**

Sampling sites were selected by considering habitats, variations, and anthropogenic activities in the Luoxiao Mountains. Fish samples were collected from April 2014 to 2017 in eleven streams of the Luoxiao Mountains. We selected eleven streams (42 sampling sites) (Figure 1), including the (1) Fu River (sampling code is FR; three sampling sites), Xiuhe River (sampling code is XH; six sampling sites), Jinjiang River (sampling code is JJ; three sampling sites), Yuanshui River (sampling code is YS; three sampling sites), Heshui River (sampling code is HS; three sampling sites), Shushui River (sampling code is SS; two sampling sites), Suichuan River (sampling code is SC; five sampling sites), Shangyou River (sampling code is SY; three sampling sites), Miluo River (sampling code is ML; four sampling sites), Liuyang River (sampling code is LY; four sampling sites), Mishui River (sampling code is MS; six sampling sites). We collected the fish catch from professional fishermen who captured fish using fully standardized five gillnet clusters, each consisting of six gillnets of 50–80 m in length 4–10 m in height (mesh size = 1.0–10.0 cm) in the Luoxiao Mountains rivers. In addition, we assumed similar capture efficiencies from gillnet samples at each site. At the same time, we surveyed and collected fish in the township markets along the river which enhanced the species checklists at each section. All fish specimens were identified according to Chen (1998), Chu et al. (1999), and Yue (2000), and the scientific name was corrected according to Fishbase (http://www.fishbase.org/search.php). The division of endangered categories of fish was decided according to Jiang et al. (2016) and IUCN (2017).
Table 1. Hydrology and environmental characteristics of the streams of Luoxiao Mountains. JJ: Jinjiang River; YS: Yuanshui River; HS: Heshui River; SS: Shushui River; SC: Suichuan River; SY: Shangyou River; MS: Mishui River; ML: Miluo River; FS: Fushui River; XH: Xiuhe River; LY: Liuyang River.

| Stream | Latitude          | Longitude          | Length (km) | Area (km²) | Average gradient (%) | Average precipitation (mm) | Average temperature (°C) | Annual average runoff (×10⁸ m³) | Average Altitude (m) |
|--------|-------------------|--------------------|-------------|------------|----------------------|---------------------------|--------------------------|---------------------------|---------------------|
| LY     | 28°24’–28°46’     | 112°99’–114°04’    | 222         | 4665       | 0.57                 | 1598                      | 17.3                     | 39.41                   | 252                 |
| MS     | 27°16’–26°25’     | 112°88’–113°99’    | 296         | 10305      | 1.01                 | 1483                      | 18.1                     | 76.03                   | 352                 |
| ML     | 28°36’–29°02’     | 112°93’–114°05’    | 253         | 5543       | 0.46                 | 1400                      | 17.6                     | 43.04                   | 250                 |
| FS     | 29°49’–29°86’     | 114°40’–115°45’    | 196         | 5250       | 0.79                 | 1275                      | 16.6                     | 43.5                    | 613                 |
| XH     | 28°31’–29°12’     | 114°14’–116°01’    | 419         | 14700      | 0.48                 | 1663                      | 16.7                     | 135.1                   | 676                 |
| SY     | 25°37’–25°49’     | 113°43’–114°49’    | 204         | 4647       | 0.70                 | 1570                      | 18.8                     | 33                      | 615                 |
| SC     | 26°11’–26°30’     | 113°56’–114°44’    | 176         | 2882       | 2.36                 | 1640                      | 16.9                     | 27.1                    | 971                 |
| SS     | 126°29’–26°47’    | 14°04’–114°50’     | 152         | 1301       | 2.14                 | 1630                      | 16.7                     | 11.3                    | 1610                |
| HS     | 27°04’–27°24’     | 114°01’–114°59’    | 256         | 9103       | 0.59                 | 1580                      | 17.8                     | 27.4                    | 747                 |
| YS     | 27°27’–28°04’     | 114°10’–115°29’    | 279         | 6262       | 0.34                 | 1678                      | 17.2                     | 29.6                    | 391                 |
| JJ     | 27°57’–28°25’     | 114°01’–115°49’    | 307         | 7886       | 0.26                 | 1679                      | 17.6                     | 70                      | 391                 |

Figure 1. Map showing the sampling location of the streams of the Luoxiao Mountains.
Data analysis

Beta diversity is represented by the difference in species composition between different communities, which was determined by species turnover (species replacement) and nestedness (richness difference; Baselga 2010; Carvalho et al. 2012). In order to quantify the effects of two processes, Baselga (2010) systematically proposed the beta diversity decomposition method (BAS frameworks) based on the Sørensen index ($\beta_{sor}$), which was decomposed into species spatial turnover components ($\beta_{sim}$) and nestedness components ($\beta_{sne}$). Podani and Schmera (2011) and Carvalho et al. (2012) proposed the beta diversity decomposition method (POD frameworks) based on the Jaccard index ($\beta_{jac}$), which was decomposed into species replacement components ($\beta_{-3}$) and richness difference components ($\beta_{rich}$). Here, we analyzed the fish biodiversity based on both the BAS and POD frameworks.

BAS frameworks (Sørensen index):

\[
\beta_{sor} = \frac{b + c}{2a + b + c}
\]
\[
\beta_{sim} = \frac{\min(b, c)}{a + \min(b, c)}
\]
\[
\beta_{sne} = \frac{|b - c|}{2a + b + c} \times \frac{a}{a + \min(b, c)}
\]

POD frameworks (Jaccard index):

\[
\beta_{jac} = \frac{b + c}{a + b + c}
\]
\[
\beta_{-3} = \frac{2\min(b, c)}{a + b + c}
\]
\[
\beta_{rich} = \frac{|b - c|}{a + b + c}
\]

where a was the number of shared species among two streams, and b and c were the number of species only present in the first and second stream, respectively. Sørensen and Jaccard indices ranged from 0 to 1, representing respectively no species and all species in common among the two streams (Appendix 1).

A principal component analysis (PCA) was performed separately based on Sørensen and Jaccard indices to visualize patterns of fish assemblages among the Luoxiao Mountains rivers (Legendre and Legendre 2012). PCA results were then analyzed using R 3.2.0 version (R Development Core Team 2014) and using the “ade4” package (Dray and Dufour 2007).

We performed Mantel tests and partial Mantel tests (Legendre and Legendre 2012) with 9999 permutations to assess the correlations (Spearman’s method) between eight pairwise similarity matrices and the matrices of geographical drivers (difference in area, length, average precipitation, annual average runoff, average altitude, average gradient and average temperature among streams; Table 1) to explore the po-
potential mechanisms that explained beta diversity patterns. The partial Mantel tests were used to remove the effect of covariation because an inter-correlation between matrices of difference in area, length, average precipitation, annual average runoff, average altitude, average gradient and average temperature was detected ($P < 0.05$). All the analyses were performed in R 3.2.0 (R Development Core Team 2014) using the packages BAT (Cardoso et al. 2015), BETAPART package (Baselga and Orme 2012) and VEGAN (Oksanen et al. 2015).

**Results**

**Fish species composition**

The fish specimens sampled and identified in the Luoxiao Mountains were categorized into 113 species and 17 families (Figure 2; Appendix 2). The number of Cypriniformes was the greatest, accounting for 68.1% of the total number of fish species, followed by Siluriformes and Perciformes, accounting for 14.2% each, and Beloniformes, accounting for 0.1% (Figure 2). In addition, according to the endangered categories of the Jiang et al. (2016), Least Concern fish species were the greatest, accounting for 77.9% of the catch (Appendix 3). Critically Endangered, Vulnerable, and Near Threatened fish species accounted for 7.1% (Appendix 3).

![Figure 2. Fish composition from the streams of the Luoxiao Mountains.](image-url)
Beta diversity patterns

The fish composition similarity in the Luoxiao Mountains had a mean value of 0.50 and 0.67, based on BAS and POD frameworks respectively (SD ± 0.06 and SD ± 0.05, respectively; Table 2). The spatial turnover and replacement components ($\beta_{sor}$ and $\beta_{ Jac}$, 0.41 ± 0.03 and 0.32 ± 0.03) were higher than the nestedness and richness difference components ($\beta_{sne}$ and $\beta_{ Rich}$, 0.13 ± 0.09 and 0.28 ± 0.16). FS and SC had a high $\beta_{sor}$ and $\beta_{ Jac}$ (0.55 ± 0.07 and 0.54 ± 0.11; 0.71 ± 0.05 and 0.69 ± 0.09), a high spatial turnover and replacement components (0.39 ± 0.07 and 0.45 ± 0.12) in LY and nestedness and richness difference components (0.22 ± 0.10 and 0.41 ± 0.17) in SC (Table 2).

At the same time, fish composition similarity ($\beta_{sne}$ and $\beta_{ Rich}$) for the entire fish fauna had a mean value of 0.66 and 0.76 (SD ± 0.24 and 0.21, Table 3). The spatial turnover and replacement components ($\beta_{sor}$ and $\beta_{ Jac}$, 0.41 ± 0.03 and 0.32 ± 0.03) were higher than the nestedness and richness difference components ($\beta_{sne}$ and $\beta_{ Rich}$, 0.25 ± 0.02 and 0.44 ± 0.02). The greatest $\beta_{sor}$ and $\beta_{ Jac}$ (0.93±0.16 and 0.96±0.13) and spatial turnover and replacement components (0.80 ± 0.04 and 0.49 ± 0.03) was in Hemiramphidae, followed by Amblycipitidae, and the lowest was in Syngnathidae. The greatest nestedness and richness difference components (0.53 ± 0.25 and 0.65 ± 0.26) was in Syngnathidae, followed by Siluridae, and the lowest was in Hemiramphidae (Table 3).

The PCA showed that fish composition similarity of LY, SS, SY, XH, and MS were similar based on BAS and POD frameworks; FS, JJ and SC were similar; HS and ML were similar; and YS was uniquely divided into other areas, respectively (Figure 3).

Table 2. Fish compositional similarity by BAS and POD frameworks in the streams of Luoxiao Mountains. JJ: Jinjiang River; YS: Yuanshui River; HS: Heshui River; SS: Shushui River; SC: Suichuan River; SY: Shangyou River; MS: Mishui River; ML: Miluo River; FS: Fushui River; XH: Xiuhe River; LY: Liuyang River; ES: Eastern stream of Luoxiao Mountain; WS: Western stream of Luoxiao Mountains.

| Stream | BAS | POD |
|--------|-----|-----|
|        | $\beta_{sor}$ | $\beta_{ Jac}$ | $\beta_{ Rich}$ | $\beta_{sne}$ | $\beta_{ Jac}$ | $\beta_{ Rich}$ |
| JJ     | 0.52±0.08 | 0.32±0.11 | 0.19±0.13 | 0.68±0.07 | 0.33±0.16 | 0.35±0.19 |
| YS     | 0.48±0.08 | 0.27±0.06 | 0.21±0.11 | 0.65±0.07 | 0.27±0.11 | 0.37±0.17 |
| HS     | 0.52±0.08 | 0.39±0.09 | 0.13±0.09 | 0.68±0.07 | 0.42±0.14 | 0.26±0.16 |
| SS     | 0.49±0.07 | 0.38±0.07 | 0.11±0.07 | 0.66±0.06 | 0.42±0.12 | 0.24±0.14 |
| SC     | 0.54±0.11 | 0.31±0.09 | 0.22±0.10 | 0.69±0.09 | 0.28±0.11 | 0.41±0.17 |
| SY     | 0.48±0.03 | 0.37±0.10 | 0.11±0.09 | 0.65±0.03 | 0.42±0.15 | 0.22±0.14 |
| MS     | 0.47±0.05 | 0.36±0.06 | 0.11±0.08 | 0.64±0.05 | 0.41±0.12 | 0.22±0.14 |
| ML     | 0.50±0.04 | 0.38±0.10 | 0.12±0.09 | 0.66±0.04 | 0.42±0.14 | 0.24±0.15 |
| FS     | 0.55±0.07 | 0.39±0.06 | 0.16±0.10 | 0.71±0.05 | 0.39±0.15 | 0.32±0.19 |
| XH     | 0.48±0.04 | 0.36±0.08 | 0.11±0.08 | 0.65±0.03 | 0.42±0.14 | 0.23±0.15 |
| LY     | 0.49±0.03 | 0.39±0.07 | 0.10±0.07 | 0.66±0.03 | 0.45±0.12 | 0.21±0.12 |
| ES     | 0.50±0.07 | 0.34±0.09 | 0.16±0.11 | 0.66±0.06 | 0.36±0.14 | 0.30±0.17 |
| WS     | 0.50±0.06 | 0.38±0.07 | 0.12±0.09 | 0.67±0.05 | 0.42±0.13 | 0.25±0.15 |
| Total  | 0.50±0.06 | 0.36±0.08 | 0.14±0.09 | 0.67±0.05 | 0.39±0.13 | 0.28±0.16 |
Table 3. BAS and POD frameworks based on all species and 17 families in the streams of Luoxiao Mountain. Values are mean ± standard deviation.

| Family          | BAS       | POD       |
|-----------------|-----------|-----------|
|                 | $\beta_{sys}$ | $\beta_{sim}$ | $\beta_{syn}$ | $\beta_{jac}$ | $\beta_{rich}$ |
| Catostomidae    | 0.70±0.32  | 0.39±0.05  | 0.31±0.03  | 0.77±0.30  | 0.25±0.03  | 0.51±0.03  |
| Cyprinidae      | 0.65±0.24  | 0.41±0.03  | 0.25±0.02  | 0.76±0.20  | 0.32±0.02  | 0.43±0.02  |
| Cobitidae       | 0.66±0.26  | 0.37±0.04  | 0.28±0.03  | 0.75±0.23  | 0.28±0.03  | 0.47±0.03  |
| Balitoridae     | 0.69±0.25  | 0.49±0.04  | 0.20±0.02  | 0.78±0.21  | 0.39±0.03  | 0.39±0.02  |
| Siluridae       | 0.64±0.30  | 0.26±0.03  | 0.38±0.03  | 0.73±0.29  | 0.17±0.02  | 0.56±0.03  |
| Clariidae       | 0.70±0.32  | 0.39±0.05  | 0.31±0.03  | 0.77±0.30  | 0.25±0.03  | 0.51±0.03  |
| Bagridida       | 0.67±0.24  | 0.41±0.03  | 0.27±0.02  | 0.77±0.21  | 0.32±0.03  | 0.45±0.03  |
| Amblycipitidae  | 0.78±0.23  | 0.58±0.04  | 0.21±0.03  | 0.85±0.19  | 0.40±0.03  | 0.45±0.03  |
| Sisoridae       | 0.56±0.22  | 0.34±0.03  | 0.22±0.02  | 0.69±0.18  | 0.31±0.03  | 0.38±0.02  |
| Hemiramphidae   | 0.93±0.16  | 0.80±0.04  | 0.14±0.03  | 0.96±0.13  | 0.49±0.03  | 0.47±0.03  |
| Syngnathidae    | 0.53±0.25  | 0            | 0.53±0.25  | 0.65±0.26  | 0            | 0.65±0.26  |
| Mastacembelidae | 0.64±0.22  | 0.43±0.03  | 0.21±0.02  | 0.75±0.18  | 0.38±0.03  | 0.37±0.02  |
| Percichthyidae  | 0.69±0.24  | 0.49±0.04  | 0.20±0.02  | 0.78±0.20  | 0.40±0.03  | 0.39±0.02  |
| Odontobutidae   | 0.65±0.25  | 0.49±0.04  | 0.16±0.02  | 0.76±0.20  | 0.33±0.16  | 0.35±0.19  |
| Gobiidae        | 0.66±0.25  | 0.42±0.04  | 0.24±0.02  | 0.76±0.22  | 0.32±0.03  | 0.44±0.02  |
| Belontiidae     | 0.53±0.23  | 0.31±0.03  | 0.23±0.02  | 0.67±0.20  | 0.28±0.03  | 0.39±0.02  |
| Channidae       | 0.58±0.22  | 0.35±0.03  | 0.23±0.02  | 0.71±0.19  | 0.30±0.02  | 0.41±0.02  |
| All species     | 0.66±0.24  | 0.41±0.03  | 0.25±0.02  | 0.76±0.21  | 0.32±0.03  | 0.44±0.02  |

Figure 3. Results of the principal component analysis (PCA) on the compositional similarity of fish species in the streams of the Luoxiao Mountains. JJ: Jinjiang River; YS: Yuanshui River; HS: Heshui River; SS: Shushui River; SC: Suichuan River; SY: Shangyou River; MS: Mishui River; ML: Miluo River; FS: Fushui River; XH: Xiuhe River; LY: Liuyang River.
We found almost no significant effects of geographical drivers on overall beta diversity for the Luoxiao Mountains (Table 4). The correlation between BAS and POD frameworks and difference in length, average precipitation, mean temperature, and average gradient were not significant in the Luoxiao Mountains. The correlation between $\beta_{sne}$ ($\beta_{rich}$) and differences in area and annual average runoff was significant. The correlation between $\beta_{sim}$ ($\beta_{-3}$) and difference in average altitude was also significant (Table 4).

**Discussion**

**Fish species composition**

Studies on fish composition and diversity in streams is the basis for the conservation and management of stream fishes (Liu et al. 2017; Zhang et al. 2018). In this study, the fish specimens sampled and classified in the stream of the Luoxiao Mountains were categorized into 113 species. Compared with species numbers of the Shiwanda Mountains (102 species; Zhao and Zhang 2001), Wuyi Mountains (117 species; Song et al. 2017), and the Tibetan Plateau (114 species; Wu and Tan 1991), the fish abundance in the Luoxiao Mountains was also higher.

**Beta diversity patterns**

Abiotic and biotic factors and their ecological processes in different stream sizes varies substantially (Zhang et al. 2018). At least in streams, local species richness of fishes, habitat diversity and complexity often increase in large streams (Roberts and Hitt 2010; Zhang et al. 2018). Comparing alpha diversity and beta diversity at local and landscape
scales is an important, yet little-understood, area of basic and applied ecological research (Kessler et al. 2009). However, most studies on fish diversity of streams have focused on alpha diversity, whereas fewer studies have investigated beta diversity (Tisseuil et al. 2013; Johnson and Angeler 2014). Knowledge of beta diversity patterns can go beyond the systematic conservation planning method that only considers the location of protected area in relation to natural physical and biological patterns (Margules and Pressey 2000; Wiersma and Urban 2005). The efficiency of protected areas not only relies on species richness, but also on how well the complementarity among sites increases biodiversity conservation (Howard et al. 1998; Bush et al. 2016; Socolar et al. 2016). In this study, as turnover brought the larger contribution to beta diversity, additional conservation efforts must target an increase in the number of protected areas, which should be spread across each one of the regions, to maximize the protection of species diversity.

Biogeographical processes

The modern freshwater fish fauna of Eurasia originated in the early Tertiary (Chen et al. 1986; Liu and Quan 1996; Zhang 2012). At the same time, the primitive species of the Danioninae and Barbinae became the main component of the fish fauna with the flattened land and the warming climate (Chen et al. 1986; Tang et al. 2001). During the dramatic changes of landscape and climate of the Eurasian continent in the late Oligocene and the end of the Pliocene, the primitive species component had been reduced rapidly (Chen et al. 1986; Tang et al. 2001). After the Quaternary ice age, only some offspring fishes of the old Tertiary Period remained (Chen et al. 1986; Tang et al. 2001). Moreover, Labeoninae, Gastromyzontidae, Balitoridae and Sisoridae were dominant during the uplift of the Tibetan Plateau (Chen et al. 1986; Yang et al. 1982; Tang et al. 2001). At the same time, a large area of alluvial plains appeared in eastern China, and special habitats were created under the influence of the East Asian monsoon (Chen et al. 1986; Zhang and Chen 1997). The cold-water fishes, such as Leuciscinae and Gobioninae became the endemic fishes of the river plain in East Asia (Hypophthalmichthyinae, Culterinae, Xenocyprininae, Acheilognathinae, Gobiobotinae) and the warm-water fishes the endemic fishes of Southeast Asia (Botiinae, Clariidae, Amblycipitidae, Belontiidae, Channidae, Mastacembelidae). Since then, these taxa have become the major faunal component in southern China (Chen et al. 1986). In this study, Culterinae, Gobioninae, and Acheilognathinae had a high species composition (Appendix 2). At the same time, the spatial turnover component is the main contributor of beta diversity in Hemiramphidae, Amblycipitidae, Catostomidae, Clariidae, Balitoridae, and Percichthyidae, indicating that it would be necessary to conserve habitats in the Luoxiao Mountains.

Threats to fish diversity

The headwater stream is a tributary of a larger river, which is often located in a mountainous area with high altitude. Compared with large rivers, it had relatively
simple habitat structure, poor nutrition, obvious hydrological change, and low species diversity (Vannote et al. 1980; Grossman et al. 1990; Zhang et al. 2018). Therefore, the ecosystem of the stream is more fragile, its resistance to external disturbance and resilience is lower, and it would be more difficult to recover once it is damaged by humans. Fish, as the apex consumers of the stream, are very important to the stability and functioning of the stream ecosystems (Nogueira et al. 2010; Yan et al., 2011; Arthington et al. 2016; Liu et al. 2017). During the long evolution process, fishes have gradually adjusted their corresponding morphological characteristics, phenological rhythms, and life history countermeasures so that they could adapt to the unique natural environment of the stream (Lytle and Poff 2004; Osorio et al. 2011; Ren et al. 2016). However, due to habitat loss, water pollution, alien-species invasions, forest overcutting, climate change, overfishing etc., the fish biodiversity of most streams in China have been seriously threatened (Dudgeon et al. 2006; Allan and Castillo 2007). For example, numerous small dams in mountain streams were established (Huang et al. 2008; Hu et al. 2009). Dam constructions modified these small fast-flowing streams, which led to the decline of fish species adapted to rapid streams (Hu et al. 2009). In addition, a large number of fishing methods such as traps, gill nets, and electro-fishing has led to overfishing which has also caused a dramatic decline in fish biodiversity (Huang and Gong 2007; Zhang et al. 2010). Heavy metal pollution has affected the aquatic ecosystem in the Luoxiao Mountains (He et al. 1998). The contents of heavy metals have greatly exceeded the recommended standards (Xu et al. 2016). In this study, critically endangered (Myxocyprinus asiaticus), vulnerable (Leptobotia elongata, Pseudobagrus pratti, Liobagrus marginatus, Siniperca roulei), and near threatened (Onychostoma barbatulum, Siniperca obscura, Siniperca undulata) fish species accounted for 7.1% of the species recovered. At the same time, the PCA results showed that the fish composition among the streams sampled in the Luoxiao Mountains were similar. As turnover brought the larger contribution to beta diversity, additional conservation efforts must target an increase in the number of protected areas, which should be spread across each of the regions, to maximize the protection of species diversity.

**Conservation implications**

Freshwater fishes were thought to be the world’s most threatened group of vertebrates after amphibians (Bruton 1995; Hiddink et al. 2008; Liu et al. 2017) and, without protection, 20% of the world’s freshwater fishes may become extinct in the next 50 years (Moyle and Leidy 1992; Fu et al. 2003). Although endangered fish have raised public awareness, conservation strategies of fish biodiversity in China are concentrated on endangered species and economic fish (Fu et al. 2003; Liu et al. 2017). In addition, protected areas mainly occur in terrestrial conservation strategies, but freshwater habitats are commonly protected only incidentally as part of their inclusion within terrestrial reserves (Huang et al. 2013). For example, conservation areas of plants, animals, and wetlands in Jiangxi Province have been established, but there are very few freshwater protected areas nor are there any fish passage facilities in the rivers (Huang et al. 2013). In this study,
species turnover component is the main pattern of beta diversity, implying that a larger number of protected areas would be necessary to conserve the regional biodiversity in the Luoxiao Mountains. Therefore, in order to protect fish biodiversity, the establishment of freshwater protected areas in the streams of the Luoxiao Mountains should be considered.

Acknowledgements

This work is supported by grants from the Key Project of Science-Technology Basic Condition Platform from The Ministry of Science and Technology of the People’s Republic of China (Grant No. 2005DKA21402), and the foundation project of the National Ministry of Science and Technology of China (2013FY111500). The authors report no conflict of interest. The authors alone are responsible for the content and writing of this article.

References

Allan JD, Castillo MM (2007) Stream ecology: Structure and function of running waters (2nd edition). Springer, Netherlands. https://doi.org/10.1007/978-1-4020-5583-6

Anderson MJ, Ellingsen KE, McArdle BH (2006) Multivariate dispersion as a measure of beta diversity. Ecology Letters 9: 683–693. https://doi.org/10.1111/j.1461-0248.2006.00926.x

Arthington AH, Dulvy NK, Gladstone W, Winfield IJ (2016) Fish conservation in freshwater and marine realms: status, threats and management. Aquatic Conservation Marine & Freshwater Ecosystems 26: 838–857. https://doi.org/10.1002/aqc.2712

Baselga A (2010) Partitioning the turnover and nestedness components of beta diversity. Global Ecology and Biogeography 19: 134–143. https://doi.org/10.1111/j.1466-8238.2009.00490.x

Baselga A (2012) The relationship between species replacement, dissimilarity derived from nestedness, and nestedness. Global Ecology and Biogeography 21: 1223–1232. https://doi.org/10.1111/j.1466-8238.2011.00756.x

Baselga A, Orme CDL (2012) betapart: an R package for the study of beta diversity. Methods in Ecology & Evolution 3: 808–812. https://doi.org/10.1111/j.2041-210X.2012.00224.x

Bruton MN (1995) Have fishes had their chips? The dilemma of threatened fishes. Environmental Biology of Fish 43: 1–27. https://doi.org/10.1007/BF00001812

Bush A, Harwood T, Hoskins AJ, Mokany K, Ferrier S, et al. (2016) Current uses of beta-diversity in biodiversity conservation: a response to Socolar. Trends in Ecology and Evolution 31: 337–338. https://doi.org/10.1016/j.tree.2016.02.020

Cardoso P, Rigal F, Carvalho JC (2015) BAT-Biodiversity Assessment Tools, an R package for the measurement and estimation of alpha and beta taxon, phylogenetic and functional diversity. Methods in Ecology and Evolution 6: 232–236. https://doi.org/10.1111/2041-210X.12310
Carvalho JC, Cardoso P, Gomes P (2012) Determining the relative roles of species replacement and species richness differences in generating beta-diversity patterns. Global Ecology and Biogeography 21: 760–771. https://doi.org/10.1111/j.1466-8238.2011.00694.x

Chen YY (1998) Fauna Sinica: Osteichthyes Cypriniformes II. Science Press, Beijing, China.

Chen YY, Cao WX, Zheng CY (1986) Ichthyofauna of the Zhujiang River with a discussion on zoogeographical divisions for freshwater fishes. Acta Hydrobiologica Sinica 10: 228–236.

Chu XL, Zheng BS, Dai DY (1999) Fauna sinica, Osteichthyes, Siluiformes. Science Press, Beijing, China.

Cressey D (2009) Aquaculture: futurefish. Nature 458: 398–400. https://doi.org/10.1038/458398a

De Silva SS (2012) Aquaculture: a newly emergent food production sector and perspectives of its impacts on biodiversity and conservation. Biodiversity and Conservation 21: 3187–3220. https://doi.org/10.1007/s10531-012-0360-9

Dray S, Dufour A (2007) The ade4 package: implementing the duality diagram for ecologists. Journal of Statistical Software 22: 1–20. https://doi.org/10.18637/jss.v022.i04

Dulvy NK, Metcalfe JD, Glanville J, Pawson MG, Reynolds JD (2000) Fishery stability, local extinctions and shifts in community structure in skates. Conservation Biology 14: 283–293. https://doi.org/10.1046/j.1523-1739.2000.98540.x

Dudgeon D, Arthington AH, Gessner MO, Kawabata Z, Knowler D, Leveque C, Naiman RJ, Prieur-Richard AH, Soto D, Stiassny MLJ (2006) Freshwater biodiversity: importance, threats, status, and conservation challenges. Biological Reviews 81: 163–182. https://doi.org/10.1017/S1464793105006950

Fu C, Wu J, Chen J, Wu Q, Lei G (2003) Freshwater fish biodiversity in the Yangtze River basin of china: patterns, threats and conservation. Biodiversity & Conservation 12: 1649–1685. https://doi.org/10.1023/A:1023697714517

Gong HL, Zhuang WY, Liao WB (2016) Comprehensive Scientific Survey of Biodiversity in Luoxiao Mountain. China Science and Technology Achievements 17(22): 9–10.

Grossman GD, Dowd JE, Crawford M (1990) Assemblage stability in stream fishes: A review. Environmental management 14(5): 661–671. https://doi.org/10.1007/BF02394716

He MC, Wang ZJ, Tang HX (1998) The chemical, toxicological and ecological studies in assessing the heavy metal pollution in Le An River, China. Water Research 2(2):510–518. https://doi.org/10.1016/S0043-1354(97)00229-7

Hiddink JG, Mackenzie BR, Rijnsdorp A et al. (2008) Importance of fish biodiversity for the management of fisheries and ecosystems. Fisheries Research 90(1): 6–8. https://doi.org/10.1016/j.fishres.2007.11.025

Hilborn R, Quinn TP, Schindler DE, Rogers DE (2003) Biocomplexity and fisheries sustainability. Ecological Monographs 75: 3–36.

Howard PC, Viskanic P, Davenport TRB, Kigenyi FW, Baltzer M, Dickinson CJ, Lwanga JS, Matthews RA, Balmford A (1998) Complementarity and the use of indicator groups for reserve selection in Uganda. Nature 394: 472–475. https://doi.org/10.1038/28843

Hu ML, Wu ZQ, Liu YL (2009) The fish fauna of mountain streams in the Guanshan National Nature Reserve, Jiangxi, China. Environmental Biology of Fishes 86: 23–27. https://doi.org/10.1007/s10641-009-9496-1
Huang LL, Wu ZQ, Hu ML, Li Q, Zong DS, Wan ZQ, Zhao WQ (2008) Fish diversity in Lushan Nature Reserve, Jiangxi, China. Journal of Nanchang University 32: 161–164.

Huang LL, Wu ZQ, Li JH (2013) Fish fauna, biogeography and conservation of freshwater fish in Poyang Lake basin, China. Environmental Biology of Fishes 96: 1229–1243. https://doi.org/10.1007/s10641-011-9806-2

Huang XP, Gong Y (2007) Fishery resources in Poyang Lake and its conservation. Jiangxi Fishery Science Technology 112(4): 2–6.

Hughes JM, Schmidt DJ, Finn DS (2009) Genes in streams: using DNA to understand the movement of freshwater fauna and their riverine habitat. Biology Science 59: 573–583. https://doi.org/10.1525/bio.2009.59.7.8

IUCN (2017) The IUCN Red List of Threatened Species. Version 2016-3. https://www.iucnredlist.org [Accessed on 22 January 2017]

Jiang ZG, Jiang JP, Wang YZ, Zhang E, Zhang YY et al. (2016) Red List of China's Vertebrates. Biodiversity Science 24: 500–551. https://doi.org/10.17520/biods.2016076

Johnson RK, Angeler DG (2014) Effects of agricultural land use on stream assemblages: Taxon-specific responses of alpha and beta diversity. Ecological Indicators 45: 386–393. https://doi.org/10.1016/j.ecolind.2014.04.028

Kennedy D, Norman C (2005) What don’t we know? Science 309: 75–75. https://doi.org/10.1126/science.309.5731.75

Kessler M, Abrahamczyk S, Bos M, Buchori D, Putra DD, Gradstein SR et al. (2009) Alpha and beta diversity of plants and animals along a tropical land-use gradient. Ecological Applications 19: 2142–2156. https://doi.org/10.1890/08-1074.1

Legendre P, Legendre L (2012) Numerical ecology, 3rd edition. Elsevier, Amsterdam.

Liao WB, Wang YY, Li Z, Peng SL, Chen CQ, Fan Q, Jia FL, Wang L, Liu WQ, Yi GS, Shi XG, Zhang DD (2014) Comprehensive Scientific Survey of Biodiversity in Jinggangshan Area, China. Science Press, Beijing.

Liu BP, Quan QQ (1996) Course of Historical Geology, 3rd edition edition. Geological Publishing House, Beijing.

Liu XJ, Hu XY, Ao XF, Wu XP, Ouyang S (2017) Community characteristics of aquatic organisms and management implications after construction of Shihutang Dam in the Gangjiang River, China. Lake and Reservoir Management, 1–16.

Lytle DA, Poff NL (2004) Adaptation to natural flow regimes. Trends in Ecology and Evolution 19: 94–100. https://doi.org/10.1016/j.tree.2003.10.002

Margules CR, Pressey RL (2000) Systematic conservation planning. Nature 405: 243–253. https://doi.org/10.1038/35012251

Moyle PB, Leidy RA (1992) Loss of biodiversity in ecosystems: evidence from fish faunas. In: Fiedler PL, Jain SK (Eds) Conservation biology: the theory and practice of nature conservation, preservation and management. Chapman and Hall, New York, 127–169. https://doi.org/10.1007/978-1-4684-6426-9_6

Naylor RL, Goldburg RJ, Primavera JH, Kauntzy N, Beveridge MCM, Clay J et al. (2000) Effect of aquaculture on world fish supplies. Nature 405: 1017–1024. https://doi.org/10.1038/35016500

Nogueira C, Buckup PA, Menezes NA, Oyakawa OT, Kasecker TP, Ramos Neto MB, et al. (2010) Restricted-range fishes and the conservation of Brazilian freshwaters. Plos One 5: e11390. https://doi.org/10.1371/journal.pone.0011390
Oksanen J, Blanchet FG, Kindt R, Legendre P, Minchin PR, O’Hara R, Simpson GL, Solymos P, Stevens MHH, Wagner H (2015) vegan: community ecology package. R package version 2.3-2. Available at: http://cran.r-project.org

Osorio D, Terborgh J, Alvarez A, Ortega H, Quispe R, Chipollini V, Davenport LC (2011) Lateral migration of fish between an oxbow lake and an Amazonian headwater river. Ecology of Freshwater Fish 20: 619–627. https://doi.org/10.1111/j.1600-0633.2011.00511.x

Podani J, Schmera D (2011) A new conceptual and methodological framework for exploring and explaining pattern in presence-absence data. Oikos 120: 1625–1638. https://doi.org/10.1111/j.1600-0706.2011.19451.x

R Development Core Team (2014) R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. Available at: http://Rproject.org [accessed 23 December 2016]

Ren P, He H, Song YQ, Cheng F, Xie SG (2016) The spatial pattern of larval fish assemblages in the lower reach of the Yangtze River: potential influences of river-lake connectivity and tidal intrusion. Hydrobiologia 766: 365–379. https://doi.org/10.1007/s10750-015-2471-2

Roberts JH, Hitt NP (2010) Longitudinal structure in temperate stream fish communities: Evaluating conceptual models with temporal data. American Fisheries Society Symposium 73: 281–299.

Socolar JB, Gilroy JJ, Kunin WE, Edwards DP (2016) How should beta-diversity inform biodiversity conservation? Trends in Ecology & Evolution 31: 67–80. https://doi.org/10.1016/j.tree.2015.11.005

Song X, Tang W, Zhang Y (2017) Freshwater fish fauna and zoogeographical divisions in the wuyi-xianxialing mountains of eastern china. Biodiversity Science 25(12): 1331–1338. https://doi.org/10.17520/biods.2017207

Sutherland WJ, Adams WM, Aronson RB et al. (2009) One hundred questions of importance to the conservation of global biological diversity. Conservation Biology 23: 557–567. https://doi.org/10.1111/j.1523-1739.2009.00121.x

Tang WQ, Chen YY, Wu HL (2001) Fish species diversity of Wuling Mountains region and its zoogeographic analyses. Journal of Shanghai Ocean University 10: 6–15.

Tisseuil C, Leprieur F, Grenouillet G, Vrac M, Lek S (2013) Projected impacts of climate change on spatio-temporal patterns of freshwater fish beta diversity: A deconstructing approach. Global Ecology and Biogeography 21: 1213–1222. https://doi.org/10.1111/j.1466-8238.2012.00773.x

Vannote RL, Minshall GW, Cummins KW et al. (1980) The river continuum concept. Canadian Journal of Fishery & Aquatic Science 37(2): 130–137. https://doi.org/10.1139/f80-017

Wei SG, Li L, Xu R, Huang ZL, Cao HL (2015) Spatial Pattern and Interspecific Relationship of Dominant Species in Plant Community in Jinggang Mountain. Journal of Tropical and Subtropical Botany 23(1): 74–80.

Wiersma YF, Urban DL (2005) Beta diversity and nature reserve system design in the Yukon, Canada. Conservation Biology 19: 1262–1272. https://doi.org/10.1111/j.1523-1739.2005.00099.x

Wu YF, Tan Q J (1991) Characteristics of the fish-fauna of the characteristics of Qinghai-Xizang plateau and its geological distribution and formation. Acta Zoologica Sinica 37(2): 135–152.
Xu YZ, Liu JJ, Cheng YX (2016) Characteristics and ecological risk assessment of heavy metals contamination in sediments of the Xiangjiang River. Environmental Chemistry 35(1): 189–198.

Yan YZ, Xiang XY, Chu L, Zhan YJ, Fu CZ (2011) Influences of local habitat and stream spatial position on fish assemblages in a dammed watershed, the Qingyi stream, China. Ecology of Freshwater Fish 20: 199–208. https://doi.org/10.1111/j.1600-0633.2010.00478.x

Yang YC, Li BY, Yin ZS, Zhang QS (1982) The formation and evolution of landforms in the Xizang Plateau. Acta Geographica Sinica 37: 76–87.

Ye ZH, Liu JQ, Yin GS, Yan ZB (2013) Landform Features and Formation Mechanism in Mt. Jinggangshan, Jiangxi, China. Journal of Mountain Science 31(2): 250–256.

Yue PQ (2000) Fauna sinica, osteichthyes, cypriniformes III. Beijing, China: Science Press.

Zhang E, Chen YY (1997) Fish fauna in northeastern Jiangxi Province with a discussion on the zoogeographical division of east china. Acta Zoologica Sinica 21(3): 254–261.

Zhang D, Wan FY, Chu L, YanYZ (2018) Longitudinal patterns in α and β diversity of the taxonomic and functional organizations of stream fish assemblages in the Qingyi River. Biodiversity Science 26(1): 1–13. https://doi.org/10.17520/biods.2017263

Zhang JM, Wu ZQ, Hu ML (2010) Resource status of four major Chinese carps in the Xiajiang reach of Ganjiang River. Journal of Hydrology 3(1): 34–37.

Zhang LS (2012) Ancient Geography of China – the Formation of China’s Natural Environment. Science Press, Beijing.

Zhao YH, Zhang CG (2001) Fish fauna and zoogeographical analysis of ShiWan Da Shan Mountains, Guangxi, China. Biodiversity Science 9(4): 336–344.

**Appendix 1**

Species occurrence in the streams of Luoxiao Mountains. 1 = presence of the species as native in the stream, 0 = the species is absent from the stream, 2 = the species is present in the stream, but non-native from this stream. JJ: Jinjiang River; YS: Yuanshui River; HS: Heshui River; SS: Shushui River; SC: Sui-chuan River; SY: Shangyou River; MS: Mishui River; ML: Miluo River; FS: Fushui River; XH: Xiuhe River; LY: Liuyang River.

| Species                  | JJ | YS | HS | SS | SC | SY | MS | ML | FS | XH | LY |
|--------------------------|----|----|----|----|----|----|----|----|----|----|----|
| Myxocyprinus asiaticus   | 0  | 0  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  |
| Zacco platypus           | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  |
| Opsariichthys bidens     | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  |
| Mylopharyngodon piceus   | 0  | 1  | 0  | 0  | 0  | 1  | 0  | 0  | 0  | 1  | 1  |
| Ctenopharyngodon idella  | 1  | 1  | 0  | 0  | 1  | 1  | 0  | 1  | 1  | 0  | 0  |
| Elopichthys bambusa      | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 1  | 1  |
| Squaliobarbus curriculus | 0  | 1  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 1  |
| Hemiculter leuciscus     | 1  | 1  | 1  | 1  | 1  | 0  | 0  | 0  | 0  | 1  | 1  |
| Hemiculter bleekeri      | 0  | 1  | 0  | 1  | 1  | 0  | 0  | 0  | 1  | 0  | 1  |
| Hemiculterella sauvagei  | 1  | 1  | 1  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  |
| Hemiculterella unii      | 0  | 0  | 0  | 0  | 1  | 1  | 0  | 1  | 0  | 1  | 0  |
| Pseudohemiculter dispar  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  |
| Species                          | JJ | YS | HS | SS | SC | SY | MS | ML | FS | XH | LY |
|---------------------------------|----|----|----|----|----|----|----|----|----|----|----|
| Pseudolaubuca sinensis          | 0  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  | 0  |
| Sinibrama macrops               | 1  | 1  | 0  | 0  | 1  | 1  | 1  | 1  | 0  | 1  | 0  |
| Chanodichthys erythropterus     | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 1  | 1  | 1  |
| Culter alburnus                 | 0  | 1  | 0  | 1  | 0  | 1  | 0  | 1  | 0  | 0  | 0  |
| Chanodichthys mongolicus        | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  | 1  | 0  |
| Chanodichthys dabryi            | 0  | 1  | 0  | 1  | 0  | 0  | 0  | 0  | 1  | 0  | 1  |
| Culter oxycephaloides           | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 1  | 1  |
| Parabramis pekinensis           | 0  | 0  | 0  | 0  | 1  | 1  | 1  | 1  | 0  | 0  | 0  |
| Megalobrama terminalis          | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 1  | 1  | 1  |
| Megalobrama amblycephala        | 0  | 0  | 0  | 1  | 1  | 0  | 0  | 0  | 1  | 0  | 0  |
| Xenocypris macrolepis           | 0  | 1  | 0  | 1  | 1  | 1  | 0  | 0  | 0  | 1  | 0  |
| Xenocypris davidi               | 1  | 1  | 1  | 1  | 0  | 0  | 0  | 0  | 0  | 1  | 1  |
| Plagiognathops microlepis       | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 1  |
| Distoechodon tumirostris        | 1  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 1  | 1  | 0  |
| Hypophthalmichthys molitrix     | 0  | 1  | 0  | 1  | 1  | 1  | 0  | 0  | 0  | 0  | 1  |
| Hypophthalmichthys nobilis      | 0  | 1  | 0  | 0  | 1  | 1  | 0  | 0  | 0  | 0  | 1  |
| Abbottina risularis             | 0  | 1  | 1  | 1  | 1  | 0  | 1  | 1  | 0  | 1  | 0  |
| Pseudorabona parva              | 1  | 1  | 0  | 1  | 0  | 0  | 0  | 1  | 1  | 0  | 0  |
| Pseudogobio taiyuan             | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  |
| Pseudogobio gulinensis          | 0  | 0  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  |
| Hemibrus labo                   | 0  | 1  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  |
| Hemibrus maculatus              | 0  | 1  | 0  | 0  | 1  | 1  | 0  | 1  | 0  | 0  | 1  |
| Huigobio cheniensis             | 0  | 1  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  |
| Sarcocheilichthys sinensis      | 1  | 1  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 1  | 1  |
| Sarcocheilichthys kiangsiensis  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 1  | 0  |
| Sarcocheilichthys nigripinnis   | 0  | 0  | 0  | 0  | 0  | 0  | 1  | 0  | 0  | 0  | 0  |
| Squalius argentatus             | 0  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 0  | 1  | 0  |
| Rhinogobio typus                | 1  | 0  | 1  | 0  | 0  | 0  | 1  | 0  | 0  | 1  | 1  |
| Platysmacheilus exiguus         | 0  | 0  | 0  | 1  | 1  | 0  | 0  | 0  | 0  | 0  | 0  |
| Saurogobio dabryi               | 1  | 1  | 1  | 0  | 1  | 1  | 1  | 1  | 0  | 1  | 1  |
| Saurogobio xiangiangensis       | 0  | 0  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 1  |
| Microphygobio kiatingensis      | 0  | 0  | 0  | 1  | 1  | 0  | 0  | 0  | 0  | 0  | 0  |
| Microphygobio fukiensis         | 0  | 0  | 0  | 1  | 0  | 0  | 1  | 0  | 0  | 0  | 0  |
| Gobiobotia filifer              | 0  | 0  | 0  | 0  | 1  | 1  | 0  | 0  | 0  | 0  | 0  |
| Gobiobotia longibarba           | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 1  | 0  | 0  | 0  |
| Acheilognathus macropterus      | 0  | 0  | 0  | 1  | 1  | 0  | 0  | 0  | 0  | 0  | 1  |
| Acheilognathus gracilis         | 0  | 1  | 0  | 1  | 1  | 0  | 0  | 0  | 0  | 0  | 1  |
| Acheilognathus chankaensis      | 0  | 0  | 0  | 1  | 0  | 1  | 0  | 1  | 0  | 0  | 0  |
| Acheilognathus tonkinensis      | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 1  | 1  | 1  | 0  |
| Acheilognathus barbatulus       | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  |
| Rhodesus ocellatus              | 0  | 1  | 0  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  |
| Rhodesus lightyi                | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 1  | 1  | 0  | 0  |
| Acroscocheilus fasciatus        | 0  | 1  | 1  | 1  | 0  | 1  | 0  | 0  | 1  | 0  | 0  |
| Acroscocheilus paradoxus        | 0  | 0  | 0  | 1  | 1  | 0  | 1  | 0  | 0  | 0  | 0  |
| Acroscocheilus hemispinus       | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 1  | 1  | 0  | 0  |
| Acroscocheilus parallels        | 0  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 0  | 1  | 1  |
| Spinibarbus hollandi            | 0  | 1  | 1  | 0  | 1  | 1  | 0  | 0  | 0  | 0  | 0  |
| Onychostoma barbatulum          | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  |
| Carassius auratus               | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  |
| Cyprinus carpio                 | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  |
| Garra orientalis                | 0  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 1  | 0  | 0  |
| Cobitis sinensis                | 0  | 1  | 0  | 0  | 1  | 0  | 0  | 1  | 1  | 1  | 0  |
| Species                        | JJ | YS | HS | SS | SC | SY | MS | ML | FS | XH | LY |
|-------------------------------|----|----|----|----|----|----|----|----|----|----|----|
| *Misgurnus anguillicaudatus*   | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  |
| *Paramisgurnus dabryanus*      | 0  | 1  | 0  | 1  | 1  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Schistura fasciata*           | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Schistura incerta*            | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Leptobotia elongata*          | 0  | 0  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Parabotia banarescii*         | 0  | 0  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Parabotia fasciata*           | 0  | 1  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Parabotia maculosa*           | 0  | 1  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Erromyzon sinensis*           | 0  | 0  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Lepturichthys finibiata*      | 0  | 0  | 0  | 1  | 1  | 0  | 1  | 0  | 0  | 0  | 0  |
| *Vanmanenia stenosoma*         | 0  | 0  | 0  | 1  | 1  | 0  | 0  | 0  | 0  | 1  | 0  |
| *Vanmanenia pingtowensis*      | 0  | 1  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Pseudogastromyzon changtingensis* | 0  | 0  | 1  | 1  | 0  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Silurus asotus*               | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  |
| *Silurus meridionalis*         | 0  | 0  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Pterocyptis cochinhhagensis*  | 0  | 0  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Clarias fuscus*               | 0  | 0  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Hemibagrus macropterus*       | 0  | 0  | 0  | 0  | 1  | 0  | 1  | 1  | 0  | 0  | 1  |
| *Pseudobagrus crassilabris*    | 0  | 0  | 0  | 0  | 1  | 0  | 1  | 0  | 0  | 0  | 0  |
| *Pseudobagrus tenuis*          | 0  | 1  | 0  | 1  | 1  | 0  | 1  | 1  | 0  | 1  | 0  |
| *Pseudobagrus ondon*           | 0  | 1  | 0  | 0  | 0  | 0  | 1  | 0  | 0  | 0  | 0  |
| *Pseudobagrus pratti*          | 0  | 0  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Pseudobagrus albomarginatus*  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 1  | 0  | 0  |
| *Tachysurus fulvidraco*        | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  |
| *Tachysurus nitidus*           | 0  | 0  | 0  | 0  | 0  | 0  | 1  | 0  | 0  | 1  | 0  |
| *Liobagrus anguillicauda*      | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Liobagrus marginatus*         | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Liobagrus nigricauda*         | 0  | 1  | 1  | 1  | 0  | 0  | 0  | 1  | 0  | 0  | 0  |
| *Glyphorynchus sinense*        | 0  | 1  | 0  | 1  | 1  | 0  | 0  | 1  | 0  | 0  | 0  |
| *Hyphorhamphus intermedius*    | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 1  | 0  | 0  |
| *Monopterus albus*             | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  |
| *Macrogobius aculeatus*        | 0  | 1  | 0  | 0  | 0  | 0  | 1  | 0  | 0  | 0  | 0  |
| *Sinobagrus sinensis*          | 0  | 1  | 0  | 0  | 1  | 0  | 0  | 1  | 0  | 1  | 0  |
| *Siniperca chuatsi*            | 0  | 1  | 0  | 0  | 1  | 0  | 1  | 0  | 0  | 1  | 0  |
| *Siniperca kneri*              | 0  | 1  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Siniperca obscura*            | 0  | 0  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Siniperca roylei*             | 1  | 0  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Siniperca scherzeri*          | 0  | 0  | 0  | 0  | 0  | 1  | 0  | 0  | 1  | 0  | 1  |
| *Siniperca undulata*           | 0  | 0  | 0  | 0  | 0  | 1  | 0  | 0  | 0  | 0  | 1  |
| *Odontobutis sinensis*         | 0  | 1  | 0  | 1  | 0  | 0  | 1  | 0  | 0  | 0  | 0  |
| *Rhinogobius cliftordpopei*    | 1  | 1  | 1  | 1  | 0  | 0  | 1  | 0  | 0  | 1  | 0  |
| *Rhinogobius duospilus*        | 0  | 1  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Rhinogobius giurinus*         | 0  | 1  | 0  | 1  | 1  | 1  | 1  | 0  | 1  | 0  | 0  |
| *Rhinogobius lindbergi*        | 0  | 0  | 0  | 1  | 1  | 0  | 0  | 0  | 0  | 0  | 0  |
| *Rhinogobius leavelli*         | 0  | 0  | 0  | 1  | 0  | 0  | 1  | 0  | 0  | 0  | 0  |
| *Macropodus opercularis*       | 0  | 1  | 0  | 1  | 1  | 0  | 1  | 0  | 0  | 0  | 0  |
| *Channa argus*                 | 1  | 1  | 0  | 0  | 1  | 1  | 0  | 1  | 0  | 0  | 0  |
| *Channa asiatica*             | 0  | 1  | 0  | 1  | 1  | 1  | 0  | 0  | 0  | 0  | 0  |
| *Channa maculata*              | 0  | 0  | 0  | 0  | 1  | 1  | 1  | 0  | 0  | 0  | 0  |
Appendix 2

The proportion of order, family, and subfamily of fish species.

| Order           | Species (proportion) | Family               | Species (proportion) | Subfamily            | Species (proportion) |
|-----------------|----------------------|----------------------|----------------------|----------------------|----------------------|
| Cypriniformes   | 77(68.1%)            | Catostomidae         | 1(0.9%)              | Danioninae           | 2(3.2%)              |
|                 |                      | Cyprinidae           | 62(54.9%)            | Leuciscinae          | 4(6.5%)              |
|                 |                      | Cobitidae            | 9(8.0%)              | Culterinae           | 15(24.2%)            |
|                 |                      | Balitoridae          | 5(4.4%)              | Xenocyprininae       | 4(6.5%)              |
| Siluriformes    | 16(14.2%)            | Siluridae            | 3(2.7%)              | Hypophthalmichthyinae| 2(3.2%)              |
|                 |                      | Claridae             | 1(0.9%)              | Gobioninae           | 17(27.4%)            |
|                 |                      | Bagridae             | 8(7.1%)              | Gobiobotinae         | 2(3.2%)              |
|                 |                      | Amblycipitidae       | 3(2.7%)              | Acheilognathinae     | 7(11.3%)             |
|                 |                      | Sisoridae            | 1(0.9%)              | Barbinai             | 6(9.7%)              |
| Beloniformes    | 10(0.9%)             | Hemiramphidae        | 1(0.9%)              | Cyprininae           | 2(3.2%)              |
| Syngnathiformes | 3(2.7%)              | Syngnathidae         | 1(0.9%)              | Labeoninae           | 1(1.6%)              |
| Mastacembelidae |                      |                      |                      |                      |                      |
| Perciformes     | 16(14.2%)            | Percichthyidae       | 6(5.3%)              |                      |                      |
|                 |                      | Odontobutidae        | 1(0.9%)              |                      |                      |
|                 |                      | Gobiidae             | 5(4.4%)              |                      |                      |
|                 |                      | Belontidae           | 1(0.9%)              |                      |                      |
|                 |                      | Channidae            | 3(2.7%)              |                      |                      |
| Total           | 113(100%)            |                      | 113(100%)            |                      | 62(100%)             |

Appendix 3

Endangered categories of fish species in the Luoxiao Mountains. Key: DD: Data Deficient; LC: Least Concern; NT: Near Threatened; VU: Vulnerable; EN: Endangered; CR: Critically Endangered.

| Species                        | Endangered categories |
|--------------------------------|-----------------------|
| *Myxocyprinus asiaticus*       | CR                    |
| *Zacco platypus*               | LC                    |
| *Opsariichthys bidens*         | LC                    |
| *Mylopharyngodon piceus*       | LC                    |
| *Ctenopharyngodon idella*      | LC                    |
| *Elomichthys bambusa*          | LC                    |
| *Squaliobarbus cucculus*       | LC                    |
| *Hemiculter leuciscus*         | LC                    |
| *Hemiculter bleekeri*          | LC                    |
| *Hemiculterella sauvegi*       | LC                    |
| *Hemiculterella wui*           | LC                    |
| *Pseudohemiculter dispar*      | LC                    |
| *Pseudolaubuca sinensis*       | LC                    |
| *Sinibrama macrops*            | LC                    |
| *Chandichthys erythropterus*   | LC                    |
| *Culter alburnus*              | LC                    |
| *Chandichthys mongolicus*      | LC                    |
| *Chandichthys dabryi*          | LC                    |
| *Culter oxycephaloides*        | LC                    |
| *Parabramis pekinensis*        | LC                    |

*Jiang et al. (2016) IUCN (2017)*
| Species                                | Jiang et al. (2016) | IUCN (2017) |
|----------------------------------------|---------------------|-------------|
| Megalobrama terminalis                 | LC                  | DD          |
| Megalobrama amblycephala               | LC                  | LC          |
| Xenocypris macrolepis                  | LC                  | LC          |
| Xenocypris davidii                     | LC                  | DD          |
| Plagiognathops microlepis              | LC                  | LC          |
| Distoechodon tumirostris               | LC                  | LC          |
| Hypophthalmichthys molitrix            | LC                  | NT          |
| Hypophthalmichthys nobilis             | LC                  | DD          |
| Abbottina rivularis                    | LC                  | DD          |
| Pseudorasbora parva                    | LC                  | LC          |
| Pseudogobio vaillanti                  | LC                  | LC          |
| Pseudogobio guilinensis                | LC                  | DD          |
| Hemibarbus labo                        | LC                  | DD          |
| Hemibarbus maculatus                   | LC                  | DD          |
| Huigobio chenhiienensis                | LC                  | LC          |
| Sarcocheilichthys sinensis             | LC                  | LC          |
| Sarcocheilichthys kiangsiensis         | LC                  | DD          |
| Sarcocheilichthys nigripinnis          | LC                  | DD          |
| Squatilus argentatus                   | LC                  | DD          |
| Rhinogobio typus                       | LC                  | DD          |
| Platysmacheilus exigus                 | LC                  | LC          |
| Saurogobio dabryi                      | LC                  | DD          |
| Saurogobio xiangjiangensis             | LC                  | DD          |
| Microphysogobio kiatingensis           | DD                  | LC          |
| Microphysogobio fukiensis              | DD                  | LC          |
| Gobiobota filifer                      | LC                  | DD          |
| Gobiobota longibarba                   | DD                  | DD          |
| Acheilognathus macropterus             | LC                  | DD          |
| Acheilognathus gracilis                | LC                  | DD          |
| Acheilognathus chankaensis             | LC                  | DD          |
| Acheilognathus tonkinensis             | LC                  | DD          |
| Acheilognathus barbatulus              | LC                  | LC          |
| Rhodeus ocellatus                      | LC                  | DD          |
| Rhodeus lighti                         | LC                  | LC          |
| Acrossocheilus fasciatus               | LC                  | DD          |
| Acrossocheilus paradoxus               | LC                  | DD          |
| Acrossocheilus hemipinus               | LC                  | LC          |
| Acrossocheilus parallens               | LC                  | LC          |
| Spinibarbus hollandi                   | LC                  | DD          |
| Onychostoma barbatulum                 | NT                  | DD          |
| Carasius auratus                       | LC                  | LC          |
| Cyprinus carpio                        | LC                  | VU          |
| Garra orientalis                       | LC                  | LC          |
| Cobitis sinensis                       | LC                  | LC          |
| Misgurnus anguillicaudatus             | LC                  | LC          |
| Paramisgurnus dabryanus                 | LC                  | DD          |
| Schistura fasciolata                   | DD                  | DD          |
| Schistura incerta                      | DD                  | DD          |
| Leptobotia elongata                    | VU                  | VU          |
| Parabotia banaresci                    | LC                  | DD          |
| Parabotia fasciata                     | LC                  | LC          |
| Parabotia maculosa                     | LC                  | LC          |
| Species                                      | Endangered categories | Jiang et al. (2016) | IUCN (2017) |
|----------------------------------------------|------------------------|---------------------|-------------|
| Erromyzon sinensis                          | DD                     | DD                  | DD          |
| Lepturichthys fimbriata                     | DD                     | DD                  | LC          |
| Vanmanenia stenoema                         | DD                     | DD                  | DD          |
| Vanmanenia pingchowensis                    | DD                     | DD                  | DD          |
| Pseudogastromyzon changtingensis            | DD                     | DD                  | DD          |
| Silurus asotus                               | LC                     | LC                  | LC          |
| Silurus meridionalis                         | LC                     | LC                  | LC          |
| Pterocryptis cochinchinensis                 | LC                     | LC                  | LC          |
| Clarias fuscus                               | LC                     | LC                  | LC          |
| Hemibagrus macropterus                       | LC                     | LC                  | LC          |
| Pseudobagrus crassilabris                    | LC                     | LC                  | DD          |
| Pseudobagrus temuis                          | DD                     | DD                  | DD          |
| Pseudobagrus ondon                           | DD                     | DD                  | LC          |
| Pseudobagrus pratti                          | VU                     | DD                  | DD          |
| Pseudobagrus alboarginatus                   | LC                     | LC                  | DD          |
| Tachysurus fulvidraco                        | LC                     | LC                  | LC          |
| Tachysurus nitidus                           | LC                     | LC                  | DD          |
| Liobagrus anguillicauda                      | DD                     | DD                  | DD          |
| Liobagrus marginatus                         | VU                     | DD                  | DD          |
| Liobagrus nigricauda                         | DD                     | EN                  | DD          |
| Glyptothorax sinense                         | LC                     | DD                  | DD          |
| Hyporhamphus intermedius                     | LC                     | LC                  | DD          |
| Monopterus albus                             | LC                     | LC                  | LC          |
| Macragnathus aculeatus                       | LC                     | LC                  | DD          |
| Sinobdella sinensis                          | DD                     | LC                  | LC          |
| Siniperca chuatsi                            | LC                     | DD                  | DD          |
| Siniperca knerii                             | LC                     | DD                  | DD          |
| Siniperca obscura                            | NT                     | LC                  | LC          |
| Siniperca roulei                             | VU                     | DD                  | DD          |
| Siniperca scherzeri                          | LC                     | DD                  | DD          |
| Siniperca undulata                           | NT                     | NT                  | NT          |
| Odontobutis sinensis                         | LC                     | LC                  | DD          |
| Rhinogobius cliffordpopei                    | LC                     | LC                  | DD          |
| Rhinogobius duospilus                         | DD                     | DD                  | DD          |
| Rhinogobius giurinus                         | LC                     | LC                  | LC          |
| Rhinogobius lindbergi                        | DD                     | DD                  | DD          |
| Rhinogobius laevelli                         | LC                     | LC                  | LC          |
| Macropodus opercularis                       | LC                     | LC                  | LC          |
| Channa argus                                 | LC                     | DD                  | DD          |
| Channa asiatica                              | LC                     | LC                  | LC          |
| Channa maculata                              | LC                     | LC                  | LC          |