Review

Progesterone receptors – animal models and cell signaling in breast cancer

Progesterone’s role in mammary gland development and tumorigenesis as disclosed by experimental mouse genetics

Selma Soyal, Preeti M Ismail, Jie Li, Biserka Mulac-Jericevic, Orla M Conneely and John P Lydon

Department of Molecular and Cellular Biology, Baylor College of Medicine, Houston, Texas, USA

Correspondence: Dr John P Lydon, Department of Molecular and Cellular Biology, Baylor College of Medicine, One Baylor Plaza, Houston, TX 77030, USA. Tel: +1 713 798 3534; fax: +1 713 790 1275; e-mail: jlydon@bcm.tmc.edu

Introduction

Epidemiological studies have shown that early onset of menarche, delayed entry into menopause, cycle periodicity, nulliparity, and a late first pregnancy represent individual risk factors for breast cancer. However, early menopause and early first parity decrease this risk [1–3]. Because progesterone’s presence or absence directly influences the establishment of each of these reproductive endocrine states, assessing mammary gland development and tumorigenesis in relation to progesterone exposure is of paramount importance.

As for most epithelial cancers, the incidence of human breast cancer is age dependent, implicating a stochastic multistep progression in the development of this disease [4]. Although breast cancer incidence increases with age, the increase is not uniform but achieves its highest rate during the reproductive years of premenopause [3]. The data suggest that ovarian steroidal exposure of the mammary epithelial cell (MEC) during this reproductive period underlies the risk to breast cancer presented by many of the aforementioned reproductive endocrine states [3].

One hypothesis to account for the link between ovarian steroidal exposure and human breast cancer risk is that ovarian steroid-induced MEC proliferation provides a temporal window of opportunity for the progressive acquisition of genetic errors [3]. As a result of these errors, the transformed MEC is predicted to undergo unchecked clonal expansion to a mammary neoplasm.

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Abstract

The progesterone receptor knockout mouse demonstrated progesterone’s importance to parity-induced mammary tertiary branching and lobuloalveologenesis. Because early parity provides significant protection against breast cancer whereas prolonged exposure to premenopausal ovarian progesterone (or to postmenopausal supplemements thereof) has been linked to breast cancer risk, this steroid can be considered to exhibit contrasting roles in breast cancer etiology. This review describes the important mouse models that have contributed to our understanding of progesterone’s role in mammary gland development and neoplasia. We conclude by emphasising the urgent need to identify the molecular targets of the progesterone receptor, and to determine whether these targets are modulated differently by the progesterone receptor isoforms (A and B) during mammary morphogenesis and tumorigenesis.

Keywords: knockout, mammary gland, progesterone, progesterone receptor, tumorigenesis

MEC = mammary epithelial cell; PR = progesterone receptor; PRAKO = a specific knockout for the PR-A isoform; PRBKO = a specific knockout for the PR-B isoform; PRKO = progesterone receptor knockout.
Although estrogen’s proliferative effects on mammary gland development and tumorigenesis are well recognized, progesterone’s contribution to these processes has been mired in controversy [5–7]. Much of this controversy has been exacerbated in the past by: first, our inability to mechanistically dissect the individual roles of estrogen and progesterone in vivo; second, progesterone’s established inhibitory role on estrogen-induced luminal epithelial proliferation in the uterus, and the extrapolation of this inhibitory role to the mammary gland; and third, the increasing number of conflicting reports concerning the importance of synthetic progestins in mammary tumor progression in the rodent [8,9].

The acknowledged gaps in our understanding concerning progesterone’s involvement in human breast cancer has exposed the urgent need to better understand the role played by progesterone as an endocrine mammogen in normal mammary gland development.

In the present review, we provide an overview of how experimental mouse genetics has not only defined progesterone’s unique contribution to mammary gland morphogenesis and function, but has also aided in the formulation of new concepts concerning the role of this ovarian steroid in the normal and neoplastic development of this tissue.

**Postnatal mammary gland development**

*The mouse as an experimental tool of inquiry*

Because many of the fundamental aspects of mammary gland development and tumorigenesis are conserved between rodents and humans [10], the rodent model (in particular, the rat and the mouse) has historically served as the experimental system of choice for in vivo mammary gland research. With the recent advent of powerful genetic approaches to manipulate the murine genome, the mouse offers an unprecedented level of sophistication with which to query systemic or locally acting hormonal controls on mammogenesis, from the genetic, cellular, and molecular perspectives.

As with the human, murine postnatal mammary gland development consists of two distinct allometric growth stages. The first of these stages occurs at the onset of puberty, whereas the second manifests in response to pregnancy [11] (Fig. 1a). At puberty, in response to systemic estrogen and locally acting growth factors, cap cells of terminal end buds undergo extensive mitosis to drive ductal elongation and dichotomous branching to the limits of the fat pad. On reaching adulthood, the mammary gland is growth quiescent except for incipient side branching and alveolar budding (conspicuous in most strains, but not all), which develop over time as a result of the exposure of the tissue to cycling levels of ovarian steroids.

In response to the hormones of pregnancy, the epithelial compartment of the mammary gland undergoes proliferation and subsequent differentiation to generate alveoli that progressively fill the interductal spaces during late pregnancy, parturition, and lactation. Following weaning, removal of the suckling stimulus triggers collapse of the lobuloalveolar system through proteinase-mediated and apoptotic-mediated reductive remodeling processes, collectively termed involution. On completion of involution, the reinstatement of the mammary gland to a ductal architecture resembling the mammary phenotype of the prepregnant mouse completes the cycle of development (Fig. 1a).

Importantly, epidemiological and experimental investigations have provided irrefutable evidence that completion of this developmental cycle early in the reproductive life in rat, mouse and human species confers a significant protection against breast cancer in later life [1,5]. However, this protection is lost with a late first pregnancy. Given that the stage of mammary gland development at the time...
of carcinogen exposure greatly influences subsequent breast cancer initiation and progression, experimental mouse genetics has recently been applied to delineate progesterone’s role in the elaboration of each of these developmental stages.

The progesterone receptor knockout mouse

The progesterone receptor

Most of the physiological effects of progesterone are mediated by its intracellular receptor, the progesterone receptor (PR), which is a member of the nuclear receptor superfamily of transcription factors [12]. On binding progesterone ligand, the nuclear receptor ligand complex translocates to the nucleus to induce or to silence the expression of downstream target genes, which in turn manifest the physiological response of the target cell to the hormone, progesterone.

In most physiological systems, including the mammary gland, the PR is induced by estrogen, through its cognate nuclear receptor: the estrogen receptor. The PR comprises two isoforms, PR-A and PR-B, which are expressed from the same gene and are structurally identical except for a short amino acid extension contained in the N-terminal region of PR-B. Previous transient transfection experiments demonstrated that the two PR isoforms exhibit distinct transactivational functions, which are dependent on the cell of origin and on the target gene promoter context [13]. Specifically, in cell types in which PR-A is inactive, the PR-B isoform (in the absence of PR-A) is a strong transactivator of several PR-regulated promoters [13].

In a physiological setting, these results suggest that the PR-A and PR-B isoforms may regulate distinct sets of target genes and may exhibit different transactivational capabilities in a given progesterone-responsive target tissue. Moreover, in cell and promoter contexts in which PR-A lacks transactivational activity, the coexpression of PR-A and PR-B demonstrated that the PR-A isoform could act as a dominant repressor of PR-B activity. In a cellular context in which PR-A and PR-B are coexpressed, this observation suggests that PR-A has the ability to attenuate the general progesterone responsiveness of specific PR target genes in specific target tissues in vivo.

Characterization of the progesterone receptor knockout mouse

To directly examine the physiological significance of PR function in the murine mammary gland, a progesterone receptor knockout (PRKO) mouse model was generated in which both PR isoforms were simultaneously abrogated through gene targeting approaches [14]. Initial phenotypic characterization of the PRKO mouse revealed that removal of PR function resulted in a spectrum of reproductive abnormalities in the female, which included severe endocrine defects, an intrinsic impairment in ovulation, a dysfunctional uterus, and a loss in mating behaviors [14].

Unlike the estrogen receptor-α knockout mouse, in which the absence of estrogen signaling resulted in a block in mammary ductal outgrowth at puberty [15], the PRKO mouse mammary gland exhibited normal ductal elongation to generate a simple ductal architecture that was morphologically similar to the mammary gland of the young wild-type virgin [14]. Together, the estrogen receptor-α knockout and PRKO mouse models highlight the specific importance of ovarian estrogen rather than progesterone in epithelial ductal elongation in the pubescent mammary gland.

To address whether the progesterone signal is requisite for the observed increase in ductal side branching and lateral alveolar budding in the cycling nulliparous mouse, as well as for the observed morphological responses to the full spectrum of pregnancy hormones in the parous mouse, PRKO MECs were transplanted into the cleared mammary fat pad of a wild-type host [16]. The whole mammary gland transplantation approach was necessary to circumvent the inherent infertility defect of the PRKO mouse.

In the case of the cycling nulliparous host, the absence of ductal side branching and lateral alveolar budding in glands containing transplanted PRKO MECs (Fig. 1b) as compared with transplanted wild-type MECs (Fig. 1c) supports a role for progesterone in ovarian hormone-driven mammary epithelial cell proliferation. As indicated in the Introduction, ovarian steroid-induced mammary epithelial proliferation has been implicated as a basis for nulliparity-associated breast cancer risk [3]. Indeed, the recent utilization of the PRKO mouse in combination with the chemical carcinogen-induced mammary tumor model demonstrated the critical role that progesterone-induced mammary epithelial proliferation can play in the initiation and progression of carcinogen-induced mammary tumors [17]. As a corollary to the aforementioned PRKO mammary tumor studies, recent investigations have indicated that progesterone may also facilitate chromosomal instability (aneuploidy) in the subsequent stages of mammary tumor progression that follow the loss of p53 function [18].

In the parous host, despite exposure to the hormonal milieu of pregnancy, transplanted PRKO MECs (Fig. 1d) failed to elicit further ductal side-branching and lobuloalveologenesis, as observed with transplanted wild-type MECs (Fig. 1e). Because early parity provides significant protection against breast cancer in later life, understanding the cellular and molecular mechanisms by which progesterone exerts these pregnancy-associated morphological and functional changes in the mammary gland is now an important priority for contemporary mammary gland research. To address this issue, recent studies have provided convincing evidence that progesterone (with estrogen) exerts a pivotal role in the elaboration of persistent molecular changes (i.e. the activation of p53-signaling
pathways) in a subgroup of mammary epithelial cells that may contribute to parity-induced protection against mammary cancer [19]. Indeed, elegant studies recently reported by Wagner et al. [20] suggest that these parity-induced mammary epithelial populations may be identified and isolated in the near future.

**Future directions**

**A paracrine mode of action**

As in the human mammary gland [21], immunohistochemical studies on the murine mammary gland have demonstrated that PR expression is restricted to the luminal epithelial cell [22,23]. Importantly, recent immunofluorescence experiments have revealed that the majority of epithelial cells, which undergo proliferation in response to progesterone, are segregated from, but in close apposition to, PR-positive cells (Fig. 2a,b). These observations support a paracrine mode of action in which PR-positive cells (in response to progesterone) express and secrete a paracrine mediator(s) that impacts a neighboring PR-negative cell to proliferate (Fig. 2c). Interestingly, a paracrine mode of action for progesterone has also been shown to exist in the uterus [24]. Unlike the mammary gland, however, the progesterone-induced paracrine circuit emanates from the uterine stromal compartment to influence luminal epithelial proliferation.

The finding that the nonuniform cellular organization pattern for PR-positive cells and proliferating PR-negative cells is conserved between the human mammary gland and the rodent mammary gland [21,25] has provided strong support for an evolutionary conserved cellular mechanism of action by which PR-positive cells influence neighboring PR-negative cells to participate in ductal morphogenesis.

Although the reasons for such an evolutionary conserved nonuniform cellular organization for PR expression is uncertain, derailment of such an important cellular arrangement is predicted to adversely affect normal mammary gland development. Indeed, in the case of many steroid receptor positive human breast tumors, the majority of estrogen and progesterone receptor positive tumors also undergo proliferation, clearly contravening the paracrine signaling circuit that exists in the normal gland [21,25]. Although speculative, the above studies suggest that the genesis of these tumors may require an early switch in steroid-dependent regulation of proliferation from a paracrine to an autocrine mechanism.

Additional support for the importance of this cellular organization pattern for PR expression to normal mammary gland development comes from the recent analysis of the adult CCAAT/enhancer binding protein β knockout mammary gland, which does not respond to the proliferative effects of estrogen and progesterone [23]. In contrast to a nonuniform cellular distribution pattern for PR expression observed in the normal gland, the CCAAT/enhancer binding protein β knockout mammary gland exhibits a uniform cellular organization pattern for PR-positive cells with the attendant absence of PR-negative cells that proliferate in response to the progesterone paracrine signal.

Clearly, to gain a more meaningful mechanistic understanding of progesterone’s paracrine mode of action in the mammary gland, identification of the downstream genetic networks that relay the progesterone signal will be essential. Towards this end, recent studies have provided strong evidence that the wnt-4 signaling pathway may act as an important paracrine mediator of the progesterone-initiated proliferative signal in the mammary gland [26].

With the availability of the complete murine genome sequence, of gene discovery approaches, and of genetic strategies to conditionally turn on or turn off potential target genes in the murine mammary gland, we anticipate
that progesterone’s mammogenic effects will be readily exposed to molecular dissection in the near future.

**PR-A and PR-B**

Transgenic approaches have recently been employed to address whether the PR isoforms (PR-A and PR-B) mediate distinct physiological effects in the mammary gland [27,28]. Overexpression of PR-A resulted in increased ductal branching and eventual hyperplasia [27], whereas PR-B overexpression caused reduced ductal elongation and branching [28]. These *in vivo* investigations strongly support the concept that progesterone’s physiological signal is interpreted differently by its two receptor isoforms, and that misexpression of either one of these isoforms can result in aberrant mammary gland development.

Using gene targeting approaches, a specific knockout for the PR-A isoform (PRAKO) has recently been generated and characterized [29]. The PRAKO mouse, like the PRKO mouse, exhibited a infertility phenotype. Unlike the PRKO mouse, however, the PRAKO mammary gland developed normally in the absence of PR-A function [29]. Although PR-A is expressed in the mammary gland, these studies suggest that, unlike most other progesterone target tissues, the PR-B isoform may functionally compensate for PR-A in the murine mammary gland.

To determine whether PR-B is indispensable for mammary development or whether its actions can be compensated by PR-A, a specific knockout for the PR-B isoform (PRBKO) mouse model has recently been generated and is currently being characterized. Unlike the PRAKO mouse, the PRBKO female is fertile. Initial mammary gland studies, however, indicate that loss of PR-B function results in reduced pregnancy-associated ductal branching compared with wild-type mammary glands (Conneely OM, unpublished data, 2002). These data, although preliminary, suggest that most of the mammogenic effects of progesterone are mediated by the PR-B isoform in the mouse.

**Conclusions**

Experimental mouse genetics has highlighted progesterone’s indispensable role in pregnancy-induced morphological and functional changes in the mammary gland. If the aforementioned developmental changes occur early in reproductive life, a significant protection against breast cancer can be achieved. Because prolonged progesterone exposure, either through uninterrupted cyclical ovarian activity or by postmenopausal supplementation, has been linked to breast cancer risk, progesterone can be viewed as an endocrine mammogen with contrasting roles with respect to breast cancer etiology. With this view in mind, our concepts of breast cancer risk in relation to progesterone exposure will remain just that until the downstream genetic pathways through which progesterone affects normal and neoplastic mammary development are identified. With the availability of PRKO mouse models for PR-A, PR-B or both isoforms, in combination with DNA microarray approaches, we believe this goal will soon be realized.

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