A Correlation Between Differentiation Phenotypes of Infused T Cells and Anti-Cancer Immunotherapy

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T-cell therapy, usually with ex-vivo expansion, is very promising to treat cancer. Differentiation status of infused T cells is a crucial parameter for their persistence and antitumor immunity. Key phenotypic molecules are effective and efficient to analyze differentiation status. Differentiation status is crucial for T cell exhaustion, in-vivo lifespan, antitumor immunity, and even antitumor pharmacological interventions. Strategies including cytokines, Akt, Wnt and Notch signaling, epigenetics, and metabolites have been developed to produce less differentiated T cells. Clinical trials have shown better clinical outcomes from infusion of T cells with less differentiated phenotypes. CD27+, CCR7+ and CD62L+ have been the most clinically relevant phenotypic molecules, while Tscm and Tcm the most clinically relevant subtypes. Currently, CD27+, CD62L+ and CCR7+ are recommended in the differentiation phenotype to evaluate strategies of enhancing stemness. Future studies may discover highly clinically relevant differentiation phenotypes for specific T-cell production methods or specific subtypes of cancer patients, with the advantages of precision medicine.

Keywords: memory T cells, phenotypic molecules, differentiation, clinical trials, anti-cancer immunotherapy

INTRODUCTION

T-cell therapy is a very promising anti-cancer therapy. It includes ex-vivo expansion and reinfusion of tumor-reactive T cells such as tumor-infiltrating lymphocytes (TILs) and, genetically-engineered T cells with conventional T-cell receptors (TCRs) or chimeric antigen receptors (CARs). CD19-specific CAR-T cell immunotherapy has achieved a complete response rate of more than 90% in B-cell leukemia and 30-50% in lymphoma (1). The efficacy of NY-ESO-1-specific TCR-T cell therapy in synovial cell sarcoma and melanoma has been up to 61% and 55%, respectively (1, 2). However, CAR-T cell therapy is not very effective in treating solid tumors. NY-ESO-1-specific TCR-T therapy is useful in just a few solid tumors. And even in these just a few solid tumors, the efficacy is still not satisfactory (1, 2). Therefore, tumor immunotherapy is still facing big challenges in the treatment of solid tumors (3).

Differentiation status of infused T cells is emerging as a crucial parameter for generating cell products with superior persistence and antitumor immunity (3, 4). The phenotype is a both effective and efficient indicator to analyze differentiation status of T cells. Quite a few clinical trials have shown that infused T-cell differentiation phenotypes are significantly relevant to clinical outcomes in anti-cancer therapies (5–11). However, the specific differentiation phenotypes indicating the
strongest T-cell therapeutic effects are still controversial. Here, we summarize the clinical investigations on differentiation phenotypes of infused T cells and their anti-cancer therapy, which may provide some insights to production of T cells with good quality for cancer immunotherapy.

**PHENOTYPIC MOLECULES OF T CELL DIFFERENTIATION**

The clinical trials revealed several T-cell differentiation phenotypes for anti-cancer therapies (5–11). The key molecules used to identify the phenotypes include CCR7, CD27, CD28, CD39, CD45RA, CD45RO, CD62L, CD69, CD95, IL7Rα.

**CCR7**

CC Chemokine Receptor 7 (CCR7), also known as CD197, is a kind of CCR receptors with specificity for chemokine CCL19 and chemokine CCL21. CCR7 regulates the homing of T cells to lymphoid organs. Controlling the migration of memory T cells by CCR7 modulation may determine their destiny by changing the location (12). In addition to controlling cell migration, CCR7-mediated signals affect T-cell homeostasis in lymph nodes and also influence T-cell activation and polarization (13).

Depending on the subtypes or functions of memory T cells, CCR7 may also determine the niche suitable for their survival. So CCR7 plays a crucial role in the activation of naive T cells (Tn) as well as the development and maintenance of memory T cell subsets (12).

**CD27**

CD27, also known as T-Cell Activation Antigen or TNFRSF7 Receptor (Tumor Necrosis Factor Receptor Superfamily, Member 7), is a member of the tumor necrosis factor receptor superfamily found on T-lymphocytes. It is a T cell co-stimulatory immune-checkpoint receptor (14), activated by the transient availability of its TNF-like ligand CD70 on lymphocytes and dendritic cells. This activation results in the increased proliferation of T cells. Key contributions of the CD27-CD70 system have been revealed on effector and memory T cell formation, which is probably based on improved cell survival (15). Co-stimulation of CD8+ T-cells through CD27 also enhances primary, secondary, memory and recall responses (14). An agonistic antibody has entered clinical trials for cancer immunotherapy (16).

**CD28**

CD28, also known as TP44 Receptor, is a kind of costimulatory T-lymphocyte receptors with specificity for CD80 antigen and CD86 antigen. Activation of T lymphocytes requires at least two signals, one delivered by TCR complex after antigen recognition, and one by co-stimulatory receptors, such as CD28 (17). CD28 also acts as a TCR-independent signaling unit to regulate the expression of pro-inflammatory cytokines (18). CD28 activation promotes T-cell proliferation, cytokine production, T-cell survival. Its stimulation also prevents clonal inactivation or anergy (19).

CD28 loss on T cells is the most consistent biological indicator of immune incompetence in aging. There is also mounting evidence for the high frequency of CD28- T cells among patients with inflammatory syndromes or with chronic infections disproportionate with their age (20), as well as in multiple solid and hematogenous tumors (21). They are derived from CD28+ precursors that have undergone repeated stimulation, indicating that CD28 silencing underlies the program of T-cell aging (20). CD28- T cells are very heterogeneous. Among them certain populations seem to promote allograft tolerance whereas others contribute to alloreactivity and costimulation blockade resistant rejection (22).

**CD39**

CD39, also known as ecto-apyrase, ectonucleoside triphosphate diphosphohydrolase 1, NTPDase-1, ectoADPase or ecto-ATP diphosphohydrolase, belongs to the GDA1/CD39 NTPase family. It is expressed primarily on activated lymphoid cells, and also in endothelial tissues. It hydrolyzes extracellular adenosine triphosphate (ATP), adenosine diphosphate (ADP) and adenosine monophosphate (AMP) into adenosine. CD39 modulates immunity (23). Its expression is induced on T cells and increased on B cells as a late activation antigen. It is a marker for terminally exhausted CD8+ T cells in patients with chronic hepatitis C virus and human immunodeficiency virus infections (24), and for exhaustion in tumor-infiltrating CD8+ T cells in melanoma and breast cancer (25). CD39+ neoantigen-specific TIL subsets are likely terminally differentiated TILs with poor proliferation, likely due to chronic antigenic stimulation.

As a cell surface marker, CD39 showed low expression in T-cell clusters associated with response in immunotherapy, and high expression in those associated with no response (26). Tumor control was enhanced when targeting CD39 in combination with anti-TIM3, PD-1 or PD-1/CTLA4 (27).

**CD45**

CD45, also known as T200, B220, 2H4 or leukocyte common antigen, is a kind of glycoproteins that constitute much of the cell surface of lymphocytes (28). Its cytoplasmic domain possesses protein tyrosine phosphatase activity (29). Its multiple isoforms result from alternative mRNA splicing of exons 4, 5 and 6, generating extracellular domains A, B, and C respectively (30). There are at least eight possible isoforms of CD45, but only RO (containing no alternatively spliced domains), RA, RB, RBC, and RABC polypeptides are found on human and murine lymphocytes (31).

Tn express the high molecular weight isoform CD45RA and that is lost after activation and replaced by the low molecular weight isoform CD45RO (32). Then T cells may also revert from CD45RO+ to CD45RA+ (33). Persistent viral infections, inflammatory syndromes and aging all induce the accumulation of highly differentiated CD45RA re-expressing memory T cells with decreased proliferative capacity, increased activation of senescence signaling pathways, greater susceptibility to apoptosis, as well as multiple effector functions (34).
**CD62L**

CD62L, also known as Mel-14, Leu-8, TQ1, L-Selectin, or Leukocyte Adhesion Molecule-1, is a homing receptor for Tn, stem central memory T cells (Tscm) and central memory T cells (Tcm) and to lymph node high endothelial venules (35). Tscm and Tcm, which lack immediate effector function but rapidly proliferate in secondary responses, express CD62L and are found in lymph nodes, whereas effector memory T cells with immediate effector function, lack CD62L and are found in blood, peripheral tissues and spleen but not in lymph nodes (36, 37).

**CD69**

CD69, also known as activation inducer molecule, early activation antigen EA1, Leu-23, C-type lectin domain family 2 member C, or p60, is a single-pass type II membrane protein. CD69 is one of the earliest markers up-regulated after T cell activation (38), and has been regarded as an activation marker (39). It is involved in lymphocyte proliferation and functions as a signal transmitting receptor in lymphocytes, natural killer cells, and platelets. CD69 is also critical for the generation and maintenance of professional memory Th lymphocytes, which can efficiently help humoral immunity in the late phase (40). CD69 functions as a homing receptor on CD4 T cells through S1P1, which regulates cell trafficking (41). CD69-overexpressing thymocytes accumulate in the thymus and do not enter the periphery (42, 43).

In contrast to CD69+ terminal effector T (TTE) cells, oligoclonal expansions are not evident within CD69+ TTE cells, which possess low perforin and granzyme expression and high inhibitory checkpoint expression and resemble T resident memory cells (44). The balance between CD69- and CD69+ cells within the bone marrow-TTE compartment may regulate immune responses in new diagnosed multiple myeloma and contribute to the clinical disease heterogeneity.

CD8+CD69+ T-cell prevalence may be an activity biomarker in Crohn’s disease (45). It is more abundant in active patients than in those in remission.

**CD95**

CD95, also known as APO-1, FAS, Fas Cell Surface Death Receptor, or TNFRSF6 (Tumor Necrosis Factor Receptor Superfamily, Member 6) Receptor, is a canonical death receptor found in a variety of tissues and on activated lymphocytes. It is engaged in the extrinsic pathway of cell death, leading to the formation of the death-inducing signaling complex (46). In T cells, CD95 plays a major role in activation-induced cell death (47). CD95 also facilitates T-cell differentiation (48). It is important in the control of T cell homeostasis and the contraction of immune responses (46).

**IL7Rα**

IL7Rα (Interleukin 7 Receptor alpha Subunit), also known as CD127, is a low affinity IL-7 receptor subunit. The high affinity receptor IL-7R is comprised of this unique α chain and CD132. IL-7 plays a key role in the development of both thymic and peripheral T cells (49). IL7Rα signals support the survival of peripheral mature T cells, not only for typical ζδ T cells, but also for γδ T cells, and even with a partial role for survival of regulatory T cells (Treg) (50). They maintain homeostasis of naive and memory T cells (51). They maintain Tn in an interphase whereas they induce occasional cell division of memory T cells (52). The interplay between TCR and IL-7R regulates IL-7R expression (50).

**T CELL DIFFERENTIATION STATUS AND ANTITUMOR RESPONSES**

Differentiation status is crucially related to T cell exhaustion, in vivo lifespan, and antitumor immunity. Differentiation status even impacts effects of antitumor pharmacological interventions.

**T Cell Exhaustion**

The transferred tumor-specific memory cells have become exhausted TILs with extensive heterogeneity in tamoxifen-induced autochthonous hepatocellular carcinoma (53) and fibrosarcoma (MCA205) (54) mice. Different cellular subsets show distinct cellular markers and functional capabilities including self-renewal, proliferation and immunoresponsiveness (Table 1). It implies differentiation status, which is the significant difference between these subsets, can play an important role in T cell lifespan and immunotherapy.

**Differentiation Status Regulates T Cell Lifespan**

After initial antigen encounter, naïve T cells are activated, rapidly proliferate and then produce cytokines and granules (55), which is a course of differentiation with progressive change in transcription factor expressions (56), epigenetics and metabolism (57). The less differentiated Tn, Tscm and Tcm cells have increased longevity and replicative capacity compared with effector memory T cells (Tem), effector T cells (Teff) and terminally differentiated Temra (58). Also, increased T cell differentiation has repressed telomere length (59) and transcription factors promoting self-renewal and cell longevity (56).

**Table 1**

| Exhausted T Cells | Cellular markers | Self-renewal and proliferation | Response to checkpoint inhibition |
|-------------------|-----------------|-------------------------------|---------------------------------|
|                   | PD1  | T-bet | TIM3 | CD38 | CD101 | CXCR5 | others |                   |                  |
| Memory-like       | mid  | hi    | low  | low  | low   | +     | −                  | High potential | Responsive       |
| Differentiated    | hi   | low   | hi   | hi   |       |       | −                  | Low potential  | Resistant        |

Ren et al. T-Cell Differentiation Phenotypes for Immunotherapy

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Less Differentiated T Cells Against Tumors

T-cell persistence is one of the key determinants for immunotherapy efficacy (58). Minimally differentiated T cells with enhanced self-renewal ability have mounted superior immune responses against established B16 melanoma in mice (60). Human-derived mesothelin-specific Tscm have shown more enhanced antitumor activity than Tcm and Tem in M108 mesothelioma mice (61). In melanoma patients, TILs exhibiting effective antitumor responses also have better persistence and survival in vivo (62). Less differentiated Tn, Tscm and Tcm cells have been more effective against established vascularized B16 melanoma in mice than Tem or Teff (63). Respectively, Tscm cells have shown more efficacious reconstitution in immunodeficient hosts (61). Tcm cells have stronger cytotoxicity (64), persisted longer in macaques (65) and humans (Clinicaltrials.gov ID: NCT03575806). Memory cell markers have also clinically been found closely related with telomere length and antitumor responses in metastatic melanoma patients (9, 66). Besides persistence, the superior antitumor efficacy of less differentiated T cells may also be because of their enhanced capacity to migrate to lymph nodes with homing receptor CCR7, primary immnosurveillance of peripheral tissues (3), and relatively weak inhibition by Tregs (67).

Differentiation Status Impacts Effectiveness of Pharmacological Interventions

Pharmacological intervention effects can also depend on the differentiation status of T cells. S-2-hydroxyglutarate (S-2HG) improves memory expansion and T-cell therapy in EG7-OVA tumor mice only when administered to naive T cells, and not when administered to the T cells with a more differentiated state (68). Not only the effect of pharmacological interventions, desirable therapeutic effects in one subset may also be prohibitive in another of different differentiation status. To sum up, low differentiation status of T cells is crucial for antitumor therapy.

STRATEGIES TO PRODUCE LESS DIFFERENTIATED T CELLS

Given the importance of the less differentiated T cells for antitumor therapy, strategies have been developed to produce them.

Cytokines

Cytokines IL-7 and IL-15 have been shown to promote the generation of Tcm-like cells (69). Each of them alone has also increased memory phenotypes (3). What is more, IL-7-expressing CAR-T cells have achieved complete regression of solid tumors (70). IL-7 receptor signaling has resulted in prosurvival in memory cells, while IL-15 positively affected mitochondrial (71) and catabolic metabolism (58).

Th9-culture condition using IL4 and TGF-β has enhanced proliferation, central memory phenotype and antitumor activity (72). 3-day IL-4 exposure during TCR stimulation has also induced long-lasting memory T cells (73). The mixture of IL-4, IL-7 and IL-21 has promoted central memory and stem cell memory phenotypes, and lowered inhibitory receptors PD-1, LAG-3 and TIM-3 in CAR19 T cells (74).

Akt, Wnt, and Notch Signaling

Controlling Akt/mTOR signaling has also driven a memory phenotype (75). Rapamycin has exerted an immunostimulatory effect to generate memory CD8+ T cells (76) including Tcm and Tem, and even impacted proliferation (75). Tacrolimus has also promoted Tcm cells. Akt inhibition has reduced T cell glycolysis, enhanced the expansion of both CD4+CD62L+ and CD8+CD62L+ memory TILs, promoted mitochondrial metabolism and antitumor properties (77). Fas signaling has also been implicated in Akt-driven T-cell differentiation (78). Impairment of TCR-driven Akt-mTOR phosphorylation by potassium has disrupted effector function and tumor clearance (79).

Glycogen-synthase-kinase-3β inhibitor TWS119 has modulated Wnt signaling, blocked T cell differentiation, enhanced metabolic fitness (80), promoted self-renewing memory stem cells with proliferative and antitumor capacities (60). Tscm can be generated from not only Tn, but also activated T cells. Coculturing with stromal cells expressing a Notch ligand has generated Tscm-like cells in vitro from activated mouse and human T cells (81).

Epigenetic Strategies

Conditional knockout of Dnmt3a in T cells has promoted the less-differentiated subsets with increased T-bet and TCF1 as well as decreased TIM3 and EOMES, thus enhanced persistence and cytokine production (82). Epigenetic strategies also include induced pluripotent stem cells (iPSCs). Transient expression of octamer-binding protein 4, SOX2, Krüppel-like factor 4 and MYC has reprogrammed T cells into iPSCs through stage-specific transcription factors, histone-modifying proteins, and chromatin-remodeling enzymes (83–87). Differentiated T cells may also be directly reprogrammed into less-differentiated ones (88), aiming at the silenced state of T cell-specific stemness and memory genes (89).

Bromodomain inhibitor JQ1 has inhibited effector differentiation by epigenetic modifying proteins, inhibited the histone acetylation reader BRD4 and histone deacetylase SIRT1, enhanced memory formation and persistence, and finally promoted antitumor activity (90). Metabolic by-product S-2HG has also targeted epigenetic proteins, inhibited effector differentiation, and promoted therapy (68). It has inhibited α-ketoglutarate-dependent proteins, such as the Jumonji family of histone demethylases and the TET family of DNA hydroxylases (91). C646 has inhibited the histone acetyltransferase p300, increased both CD8+CD45RA+CD62L+CCR7+ and CD8+CD45RA-CD62L+CCR7+ T cells, and eventually augmented antitumor effects (90).

Transcription factor c-Myb overexpression has enhanced CD8+ T cell memory formation, polyfunctionality and recall...
responses that promoted curative antitumor immunity in B16 tumor mice (92).

miR-155 has increased CD8+ T cell antitumor function by restraining T cell senescence and functional exhaustion through epigenetic silencing of drivers of terminal differentiation (93). miR-155 has promoted antitumor responses in melanoma mice through miR-155-Phf19-Polycomb repressor complex 2 axis.

Metabolites
Direct inhibition of hexokinase by 2-deoxyglucose has promoted CD62L+ memory cells and antitumor efficacy against established and vascularized melanoma (94). L-arginine has modulated T cell metabolism, promoted CD4+CCR7+ cells, antitumor activity and survival (95). Lactate dehydrogenase inhibition combined with IL-21 has promoted CD8+ T cell stemness and antitumor immunity in melanoma mice (96).

Others
Mitochondrial division inhibitor 1 has promoted fused mitochondrial structures and inhibited mitochondrial fission, increased memory cells, and improved antitumor activity (97).

RORγ agonists have also promoted durable memory and stemness of cells including Th17 and Tc17 (98). Central and tissue-resident memory populations have been conferred by P2RX7 (99), a purinergic receptor sensing extracellular ATP.

### INFUSION OF T CELLS WITH LESS DIFFERENTIATED PHENOTYPES HAS BETTER CLINICAL OUTCOMES IN ANTITUMOR THERAPY

9 clinical trials have analyzed the correlation between differentiation phenotypes of the infused T cells and their anti-cancer outcomes, with 2 in China and 7 in USA (Figure 1A). The first trial has been published in 2009, after which more and more clinical investigators have begun to study this correlation (Figure 1B). The characteristics of the trials have been summarized in Table 2. The associations of differentiation phenotypes with objective response have also been summarized (Table 3).

19 patients with neuroblastoma have been treated with CAR-T cells on 3 dose levels: 2×10^7 cells/m^2, 5×10^7 cells/m^2 or 1×10^8 cells/m^2. To produce CAR-T cells, peripheral blood mononuclear cells (PBMCs) have been activated with anti-CD3 mAb (OKT3) and recombinant human (rh)IL-2, transduced on day 3, expanded with 50 U/ml rhIL-2 added every 3 days, and frozen on day 12-18. 3 patients have shown complete response (CR), and 8 no evidence of disease (NED), 2 partial response (PR), 1 stable disease (SD), 4 progressive disease (PD), 2 tumor necrosis. The percentage of CD45RO+CD62L+ T cells in the infused product has been highly concordant with prolonged detection of transferred T cells within the peripheral blood (7). Each 1-unit increase in the percentage of this phenotype has been associated with a 6.1 increase in the log (duration) of CAR-expressing Epstein Barr-virus specific T cells (CAR-ATCs, p=0.055) and a 6.63 increase in the log (duration) of GD2-CAR expressing activated T cells (CAR-CTLs, p<0.0001) in the T-cell product. And this prolonged detection has been associated with prolongation of Time To Progression, which is the primary parameter for clinical outcomes. The persistence of even low levels of either CAR-ATCs or CAR-CTLs at or beyond 6 weeks has been associated with a significantly longer TTP (p=0.02) in patients with active disease. CD45RO+CD62L+ T cells mainly include the subtype of Tcm.

93 patients with measurable metastatic melanoma have been treated with the adoptive transfer of autologous TILs in conjunction with 720,000 IU/kg IL-2 following a lymphodepleting preparative regimen. TIL doses vary from less than 3×10^10 to more than 9×10^10. TILs have been grown from resected metastatic melanoma lesions in high-dose IL-2. 20 patients have achieved CR, and 32 PR, 41 NR (no response). The number of CD8+CD27+ cells infused has been significantly associated with objective response [CR vs. (PR+NR) p=0.001, (CR+PR) vs. NR p<0.001] (9). The CD8+CD27+ cell number in CR patients is (2.0 ± 0.3)×10^10, and (1.5 ± 0.2)×10^10 in PR, (0.8 ± 0.1)×10^10 in NR. CD8+CD27+ cells include the subtype of Tn, Tscm and Tcm.

19 patients with Epidermal growth factor receptor (EGFR)-positive advanced unresectable, relapsed/metastatic biliary tract cancers have received 1-3 cycles of CAR-T-EGFR cell infusion. The dose range is 0.8-4.1×10^7/kg. To produce CAR-T cells, PBMCs
| Publication year | Cancer type | No. of Patients | T-cell therapy type | Antigen Construct design | Transduction methods | Manufacture | Transfer dose | Objective response | Registration No. | Reference |
|------------------|-------------|----------------|---------------------|--------------------------|---------------------|-------------|--------------|-------------------|-----------------|-----------|
| 2009             | Melanoma    | 36             | TCR-T               | DMF5 DMF5 TCRα-T2A-DMF5 TCRβ | Retroviral vector | Stimulated with OKT3 (50 ng/mL) and IL2 (500 IU/mL) for 2 days, transduced and expanded with IL2 (6000 IU/mL) for 7-10 days, stimulated and expanded with OKT3, irradiated PBL and IL2 (6000 IU/mL) for 9-14 days | 1.5×10⁷ × 10⁹ | 1 CR, 8 PR, 27 NR | NCI-07-C-0174, NCI-07-C-0175 | (6) |
| 2011             | Neuroblastoma | 19             | CAR-CTL            | GD2 GD2-specific 14g2aζ CAR | Retroviral vector | Stimulated with OKT3 and IL2 (100 IU/mL) for 2 days, transduced, expanded with IL2 (50 U/mL) for 9-15 days | 2×10⁷, 6×10⁷ or 1×10⁸ cells/m² | 3 CR, 8 NED, 1 PR, 1 SD, 4 PD, 2 tumor necrosis | None | (7) |
| 2011             | Melanoma    | 93             | TIL                | N/A N/A N/A             | N/A             | Grown from resected metastatic melanoma lesions with IL-2 (6000 IU/mL) | Less than 3×10¹⁰ to more than 9×10¹⁰ | 20 CR, 32 PR, 41 NR | None | (9) |
| 2018             | Biliary tract cancer | 17             | CAR-T EGFR CD137-CD3ζ-CAR | EGFR-specific CD19 anti-EGFR scFv-4-1BB-CD3ζ CAR | Lentiviral vector | Stimulated with OKT3 (50 ng/mL) and IL2 (500 U/mL) for 2 days, transduced, expanded for 10 days | 0.2-7×10⁶/kg, 1-3 cycles within 6 months | 1 CR, 10 SD, 6 PD | NCT01869166 | (10) |
| 2018             | Chronic lymphocytic leukemia | 38             | CAR-T CD19 CD19-specific CAR with 4-1BB/CD3ζ domains | Lentiviral vector | Stimulated by anti-CD3/28 beads, transduced, expanded for 9-11 days | 0.8-4.1×10⁷/kg, 1-3 cycles within 6 months | 8 CR, 5 PR (3 PRIV), 25 NR | NCT01029366, NCT01747486, NCT02640209 | (6) |
| 2019             | Multiple myeloma | 25             | CAR-T BCMA anti-BCMA scFv-4-1BB-CD3ζ CAR | Lentiviral vector | Stimulated by anti-CD3/28 beads, transduced, expanded with IL2 for 10-12 days | 1-5×10⁷ or 1-5×10⁸ | 1 stringent CR, 1 CR, 5 very good PR, 5 PR, 5 minimal response, 6 SD, 2 PD | NCT02546167 | (8) |
| 2020             | Pancreatic carcinoma | 14             | CAR-T EGFR anti-EGFR scFv-CD137-CD3ζ CAR | Lentiviral vector | Stimulated with OKT3 (50 ng/mL) and IL2 (500 U/mL) for 2 days, transduced, expanded for 10 days | 1.31-8.9×10⁹ kg, 1-3 cycles within 6 months a single infusion of unselected 1.1×10⁹-1×10¹¹ | 4 PR, 8 SD, 2 PD | NCT01869166 | (11) |
| 2020             | Melanoma    | 38             | TIL                | N/A N/A N/A             | N/A             | Initially cultured from enzymatic tumor digests and tumor fragments, generated by rapid expansion with irradiated PBMC, anti-CD3 antibody (50 ng/mL), and IL-2 (5000 IU/mL) for 14 days | 17 CR, 21 NR | NCT00001832, NCT00513604, NCT01319565, NCT01468818, NCT01585415, NCT01993719 | None | (100) |
| 2020             | LBCL        | 24             | CAR-T CD19 scFv-CD28-CD3ζ CAR | Retroviral vector | Stimulated with anti-CD3 antibody and IL-2, transduced, and expanded | 2 × 10⁶ CAR-positive cells/kg | 9 CR, 1PR, 13 PD, 1 NE | None | (101) |
Ren et al.

T-Cell Differentiation Phenotypes for Immunotherapy

TABLE 3 | Association of differentiation phenotypes with response in clinical trials.

| Cancer                      | Differentiation phenotype | Corresponding subtype                        | Association with response | Reference |
|-----------------------------|---------------------------|----------------------------------------------|---------------------------|-----------|
| Melanoma                    | CD27+                      | Tscm and Tcm                                | Unsubstantial             | (5)       |
| Melanoma                    | CD28+                      | All                                          | Unsubstantial             | (5)       |
| Melanoma                    | CD45RA+                    | Tscm, Teff and Temra                        | Unsubstantial             | (5)       |
| Melanoma                    | CD45RO+                    | Tcm and Tcm                                 | Unsubstantial             | (5)       |
| Neuroblastoma               | CD45RO+/CD62L+             | Tcm                                          | Phenotype percentage - CAR-T persistence (p ≤ 0.055), CAR-T persistence – TPP (p = 0.02) | (7)       |
| Melanoma                    | CD8+/CD27+                 | Tn, Tscm and Tcm                            | CR vs. (PR+NR) p<0.001, (CR+PR) vs. NR p<0.001 | (9)       |
| Biliary tract cancer        | CD45RO+/CD62L+/CCR7+       | Tcm                                          | CR/SD vs. PD, p=0.0464    | (10)      |
| Chronic lymphocytic leukemia| CD8+/CD27+/CD45RO-        | Tscm                                         | CR/PRTD vs. PR/NR p=0.0009 | (5)       |
| Chronic lymphocytic leukemia| CD8+/CD45RO-/CCR7+/CD45RA+| CD8+ Tscm                                   | CR vs. NR p=0.00008, CAR/PRTD vs. PR/NR p=0.0055, CR vs. PR p=0.0121 | (5)       |
| Multiple myeloma            | CD8+/CD45RO-/CD27+        | ≥P R vs. <P R, p=0.0121                      | (8)                       |
| Pancreatic carcinoma        | CD45RO+/CD62L+/CCR7+      | Tcm                                          | PR/SD vs. PD, p=0.0468    | (11)      |
| Melanoma                    | CD8+/CD39-/CD69-           | Stem-like CD8 T cells                       | CR vs. NR, p<0.01         | (100)     |
| Melanoma                    | CD39-/CD69-                | Stem-like T cells                            | CR vs. NR, p=0.0096       | (100)     |
| LBCL                        | CD8+/CCR7+/CD27+          | CD8+ Tscm and Tcm                            | CR vs. PR/PD, p<2.2×10^-16 | (101)     |

The used phenotype is CD8+/CD45RO+/CD62L+/CCR7+.

38 patients have received CD19 CAR-T cell therapy for chronic lymphocytic leukemia. To produce CAR-T cells, PBMCs have been activated with anti-CD3 and anti-CD28 monoclonal-antibody-coated polystyrene beads, transduced with lentiviral vector, and expanded for 9-11 days. 8 patients have achieved CR, and 3 PR (partial response with transformed disease), 5 PR, 25 NR. Objective response (CR/PR vs. NR) has associated with the frequency of CD8+CD45RO+/CD62L+/CCR7+ (p=0.0009). Theoretically this phenotype involves Tn and Tscm. However, after substantial ex-vivo transduction and expansion, the likelihood of infusing gene-modified Tn cells is negligible. So it is mainly Tscm. Frequency of CD8+ Tscm from CR/PR patients at the time of leukapheresis has been significantly higher than that in PR/NR patients (p=0.0055) (5). And likewise, CR higher than NR (p=0.0008), CR higher than PR (p=0.0121). This phenotype is CD8+/CD45RO-/CCR7+/CD45RA+/CD62L+/CD27+/CD28+ IL7Rα+CD95+.

25 patients have received 1-5×10^7 or 1-5×10^8 anti-B cell maturation antigen (BCMA) CAR-T cells for relapsed/refractory multiple myeloma. To produce CAR-T cells, autologous T cells have been stimulated with paramagnetic polystyrene beads coated with anti-CD3 and anti-CD28 monoclonal antibodies, transduced with a lentiviral vector, and expanded for 10-12 days with rhIL-2. 1 patient has achieved stringent CR, and 1 CR, 5 very good PR, 5 PR, 5 minimal response, 6 SD, 2 PD. A higher proportion of CD8+CD45RO-/CD27+ T cell phenotype has been significantly associated with response (≥PR vs. <PR, p=0.0121) (8). This phenotype involves mainly Tscm.

16 patients with metastatic pancreatic carcinoma have received 1-3 cycles of CAR-T cell infusion. The dose range is 1.31-8.9×10^6/kg. To produce CAR-T cells, autologous PBMCs have been activated with OKT3 and cultured in GT-T551 medium with 0.5% autologous serum and 500 U/mL rhIL2, transduced by lentiviral particles after 2 days, and further expanded for 10 d in culture bags. 2 patients have lost follow-up. Of the 17 evaluable patients, 1 achieved CR for 22 months and 10 had SD for 2.5-15 months from the first cycle of treatment, the other 6 PD. Enrichment of Tcm in the infused cells has improved the clinical outcome (CR/SD vs. PD, p=0.0464) (10). The used phenotype is CD45RO+/CD62L+/CCR7+.

38 patients with metastatic melanoma have received a single infusion of autologous unselected TILs. The dose range is 1.1×10^8-1×10^11 cells. TILs have been initially cultured from enzymatic tumor digest and tumor fragments produced by sharp dissection. Infusion products have been generated by rapid expansion with irradiated PBMC, anti-CD3 antibody, and IL-2 in a 1 to 1 mixture of CM and AIM-V medium supplemented with 5% human AB serum, and harvested on day 14. 17 patients have achieved CR, and 21 NR. The percentages of stem-like T cells in CR infusion products have been significantly higher than those in NR ones (p=0.0096) (100). The used phenotype is CD39-/CD69-.

Among 24 patients with large B cell lymphoma (LBCL), there were 16 diffuse LBCL, 6 transformed follicular lymphoma, and 2 primary mediastinal B cell lymphoma. They have been treated with anti-CD19 CAR T cell. The dose is 2×10^6 CAR positive cells/kg. At 3-month follow-up, 9 have achieved CR (38%), 1 PR (4%), 13 PD (50%), and 1 not evaluable (NE) due to death from sepsis. Enrichment of CD8 memory T cells, including Tscm and Tcm, in the infused cells has improved the clinical outcome (101). The used phenotype is CD8+/CCR7+/CD27+.

However, there is a trial where analyzed phenotypes have shown no significant relevance to therapeutic outcomes. 36 patients with metastatic melanoma have received 1.5-107×10^6 TCR-T cells infused intravenously followed by 720,000 U/kg IL-2 every 8 hours to tolerance. To produce TCR-T cells, autologous peripheral blood lymphocytes have been stimulated for 2 days at 10^6/mL with 50 ng/mL OKT3 in complete AIMV media supplemented with 5% human serum and 300 IU IL-2, and have been stimulated by 50 ng/mL OKT3 and cultured in GT-T551 medium with 0.5% autologous serum and 500 U/mL rhIL2, transduced by lentiviral particles after 2 days, and further expanded for 10 d in culture bags. 2 patients have lost follow-up. Of the 17 evaluable patients, 1 achieved CR for 22 months and 10 had SD for 2.5-15 months from the first cycle of treatment, the other 6 PD. Enrichment of Tcm in the infused cells has improved the clinical outcome (CR/SD vs. PD, p=0.0464) (10). The used phenotype is CD45RO+/CD62L+/CCR7+.
transduced. On day 9-12, cells have been expanded for an additional 9-14 days in 6000 IU IL-2 with 50 ng/mL OKT-3 and irradiated allogeneic PBL feeder cells. 1 patient has achieved CR, and 8 PR, 27 NR. No substantial differences have been found in the cell phenotype between responding and nonresponding patients (6). The used phenotypic molecules include CD27, CD28, CD45RA and CD45RO. It may be because CD28+, CD45RA+ or CD45RO+ phenotypes contain the interference from more differentiated T cells such as Tem, Teff and Temra (58). This just from the opposite angle indicates the less differentiated phenotypes in better T-cell therapeutic outcomes. For CD27+ phenotype, it may be because the infused cell numbers have not been compared, but just the percentages have. Also, CD8 has not been included, different from the above-described trial using CD8+CD27+ phenotype (9). So, it may have been interfered by other subtypes such as Treg.

**FUTURE DIRECTIONS**

**The Most Clinically Relevant T-Cell Phenotypic Molecules**

Among the 11 conditions, CD27+, CCR7+, and CD62L+ have been demonstrated by the most clinical trials in better outcomes of T-cell therapy (Figure 2A). These 3 molecules involve all the least differentiated subtypes including Tn, Tscm and Tcm. Hence, they have been most frequently used in clinical research. It seems contradictory that both CD45RO+ and CD45RO- have been demonstrated in respectively 3 and 2 trials associated with better clinical outcomes (Figure 2A). They correspond to Tscm and Tcm respectively. Both subtypes have played important roles in T-cell therapy. So it is still reasonable these two opposite conditions have been implied in clinical trials. However, individual phenotypic molecules hardly distinguish T cell subsets and differentiation status. There should be firstly dynamic integration between these phenotypic molecules before applying them to assess T cell differentiation. Also, with inherent substantial differences with TILs and gene-modified T cells, it might not be easy to compare phenotypical properties among them. Besides, clinical trial amount is limited as well, so the clinically relevant analysis here is currently just an indication.

**The Most Clinically Relevant T Cell Subtypes**

The 2 least differentiated subtypes, Tscm and Tcm, have been demonstrated significantly associated with better clinical outcomes in various anti-cancer trials (Figure 2B). They are all important in T-cell therapy. Their relative importance may be due to different specific circumstances. However, taking into consideration the heterogenous nature of T cells from different trials and the limited number of the trials, the most clinically relevant T cell subtypes addressed here are just implications. Future development of a universal definition of T cell subsets may strongly facilitate comparisons and meta-analysis for more clinically relevant T cell subtype.

**Differentiation Phenotypes to Evaluate Strategies of Enhancing T-Cell Stemness**

Phenotypic analysis is effective and efficient to reveal T-cell products’ differentiation status, which is very important for therapeutic outcomes. Various strategies, including metabolites, epigenetic inhibitors, nutrient small molecules, and additional genetic engineering, have been shown to enhance T-cell stemness and anti-cancer therapy (3, 4, 58). But investigators still need to be very cautious in choosing phenotypic molecules. One cannot well develop strategies enhancing T-cell stemness without an efficient and stable differentiation phenotype. To the best of our knowledge, CD27+ CD62L+ and CCR7+ are currently recommended to be included in the differentiation phenotype. Single-cell high-throughput technologies, including single-cell transcriptomics, have discovered new subtypes such as slowly dividing T central memory precursors (CMPs) and rapidly dividing non-CMPs (102). The discovery of more clinically relevant T cell subtype or phenotype may strongly improve T-cell production and anti-cancer therapy.
Phenotypic Characteristics for Prognosis of T Cell Therapy

An efficient differentiation phenotype is important not only for research & development, but also for prognosis of T cell therapy. Before T cell infusion, the differentiation phenotype of the cell product can be analyzed to predict the potential clinical outcome. It may also be used to assess whether a cancer patient is suitable for the specific kind of T-cell therapy or T-cell production. Future studies may aim to discover highly clinically relevant differentiation phenotypes for specific T-cell production methods or specific subtypes of cancer patients, with the advantages of precision medicine.

In summary, differentiation status is crucial for T cell exhaustion, in-vivo lifespan, antitumor immunity, and even antitumor pharmacological interventions. Strategies including cytokines, gut microbiota, Akt signaling, epigenetics and metabolites have been developed to produce less differentiated T cells. Clinical trials have shown that infusion of T cells with less differentiated phenotypes achieves better clinical outcomes. Cancer-specific Tscm or Tcm cells, which express CD27, CCR7, and CD62L molecules, may be the most clinically relevant subtypes for successful T cell-based immunotherapy.

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