Six months vitamin K treatment does not affect systemic arterial calcification or bone mineral density in diabetes mellitus 2

Jonas W. Bartstra1 · Fieke Draaisma1 · Sabine R. Zwakenberg2 · Nikolas Lessmann3 · Jelmer M. Wolterink4 · Yvonne T. van der Schouw2 · Pim A. de Jong1 · Joline W. J. Beulens5

Received: 30 June 2020 / Accepted: 6 October 2020 / Published online: 17 October 2020
© The Author(s) 2020

Abstract
Purpose Vitamin K-dependent proteins are involved in (patho)physiological calcification of the vasculature and the bones. Type 2 diabetes mellitus (DM2) is associated with increased arterial calcification and increased fractures. This study investigates the effect of 6 months vitamin K2 supplementation on systemic arterial calcification and bone mineral density (BMD) in DM2 patients with a history of cardiovascular disease (CVD).

Methods In this pre-specified, post hoc analysis of a double-blind, randomized, controlled clinical trial, patients with DM2 and CVD were randomized to a daily, oral dose of 360 µg vitamin K2 or placebo for 6 months. CT scans were made at baseline and follow-up. Arterial calcification mass was quantified in several large arterial beds and a total arterial calcification mass score was calculated. BMD was assessed in all non-fractured thoracic and lumbar vertebrae.

Results 68 participants were randomized, 35 to vitamin K2 (33 completed follow-up) and 33 to placebo (27 completed follow-up). The vitamin K group had higher arterial calcification mass at baseline [median (IQR): 1694 (812–3584) vs 1182 (235–2445)] for the total arterial calcification mass). Six months vitamin K supplementation did not reduce arterial calcification progression (β [95% CI]: −0.02 [−0.10; 0.06] for the total arterial calcification mass) or slow BMD decline (β [95% CI]: −2.06 [−11.26; 7.30] Hounsfield units for all vertebrae) when compared to placebo.

Conclusion Six months vitamin K supplementation did not halt progression of arterial calcification or decline of BMD in patients with DM2 and CVD. Future clinical trials may want to pre-select patients with very low vitamin K status and longer follow-up time might be warranted. This trial was registered at clinicaltrials.gov as NCT02839044

Keywords Randomized controlled clinical trial · Type 2 diabetes · Cardiovascular disease · Vitamin K · Arterial calcification · Bone mineral density

Introduction

Vitamin K is a fat-soluble vitamin that is involved in the carboxylation and activation of several vitamin K-dependent proteins. It occurs in two different forms, phylloquinone (vitamin K1) and menaquinone (vitamin K2), which differ in the length and degree of saturation of the side chain [1]. Phylloquinones are mainly derived from green vegetables, whereas menaquinones are mainly derived from animal products like cheese and meat [2]. Phylloquinones and menaquinones both have a methylated naphthoquinone ring structure, which is the functional group, but menaquinones have a longer half-life time and higher bioavailability [1, 2]. For this reason, vitamin K2 is more effective in carboxylating extrahepatic vitamin K-dependent proteins than vitamin K1 [3]. Besides their role in the coagulation cascade,
vitamin K-dependent proteins are involved in (patho)physiological calcification of the vasculature [4] and the bones [5, 6]. Matrix Gla Protein (MGP) is a vitamin K-dependent protein that inhibits vascular calcification and plays a regulatory role in the bones [7]. Osteocalcin is involved in bone metabolism [8]. Both MGP and osteocalcin require post-translational carboxylation to be fully functional and vitamin K is a co-factor for activation [7].

In the vasculature, calcifications are increasingly recognized as independent risk factors for cardiovascular disease. Current treatment of cardiovascular risk focuses on hypercholesterolemia, hypertension and thrombosis, but despite optimal treatment, the risk of new cardiovascular events remains high in patients with DM2 [9, 10]. Arterial calcifications can occur in both the intimal and medial layer of the vascular wall and these calcifications differ in cause and consequence. Intimal calcifications are generally considered as stabilized atherosclerotic plaques, whereas medial calcifications are thought to contribute to hypertension, arterial stiffness and cardiac hypertrophy [11]. Arterial calcifications, in any vascular bed, are associated with a three- to fourfold increase in the risk of cardiovascular events and progression of calcification is associated with more events [12, 13]. In addition, calcifications in different arterial beds have different clinical consequences. For example, calcification of the leg arteries is associated with peripheral arterial disease, intracranial calcifications are associated with stroke, and aortic calcifications are associated with arterial stiffness, stroke and decreased bone mineral density [14–16]. Reduction of arterial calcification progression in patients with cardiovascular diseases might, therefore, reduce the high residual cardiovascular risk. Several observational studies have shown an association of low vitamin K intake with vascular calcification and cardiovascular events, and MGP might form the link between these associations [17–19].

Osteocalcin is a marker for osteoblast activity and bone metabolism. MGP is also thought to be involved in bone metabolism, but its role is not fully understood. It is speculated to promote bone formation and inhibit osteoclast activity [20]. Despite inconsistent results on bone mineral density (BMD) [21, 22], vitamin K supplementation is shown to reduce clinical fractures [23] and this reduction might be caused by carboxylation and activation of osteocalcin and MGP [24].

Type 2 diabetes mellitus (DM2) is associated with increased arterial calcification and increased fractures [25, 26] and the low vitamin K status found in DM2 patients might contribute to this [27]. DM2 patients are prone to develop medial arterial calcifications. Since MGP is thought to inhibit the formation and progression of medial arterial calcifications, vitamin K supplementation might reduce calcification progression in these patients. Recently, we showed that 6 months vitamin K2 supplementation reduced circulating levels of inactive MGP, but did not halt arterial calcification as measured with 18F sodium-fluoride positron emission tomography (18F NaF PET) and computed tomography (CT) in the femoral arteries in patients with DM2 and a history of cardiovascular disease [28]. Different arterial beds, however, may be more or less prone to calcification and the role of MGP might differ between these different arteries [29]. This post hoc analysis aims to investigate the effect of 6 months vitamin K2 supplementation on systemic CT-measured arterial calcification and CT-measured BMD in the spine in patients with DM2 and cardiovascular disease.

**Methods**

**Study design and population**

This study is a pre-specified post hoc analysis of a double-blind, randomized, placebo-controlled clinical trial conducted at the University Medical Center Utrecht (UMCU) in the Netherlands. The methods have previously been published [28]. In short, patients were randomized to vitamin K2 or placebo. The primary end point of the original trial was change in active vascular calcification in the femoral artery as measured with 18F-NaF PET [28]. The sample size calculation was based on the primary outcome of the trial. A previous study into 18NaF PET/CT found a mean TBR of 1.96 [30]. Considering a power of 80%, a two-sided α of 5%, an SD of the difference in TBR between baseline and follow-up of 0.41 and a 15% dropout rate, 70 participants were required to detect a 15% difference in TBR. The outcomes of this post hoc analysis were change in arterial calcification mass scores and BMD as measured on conventional CT. Participants were recruited via the Diabetes Pearl String Initiative-cohort, a Julius Center database of patients interested in participating in studies and via the outpatient clinic of the UMCU and Diakonessenhuis Utrecht. Middle-aged men and women (> 40), diagnosed with DM2 and with known pre-existing arterial disease, were included. Pre-existing arterial disease was based on an ankle-brachial index (ABI) < 0.9 and/or was diagnosed by a physician. Exclusion criteria were vitamin K antagonist use, the use of vitamin K containing (multi)vitamins and unwillingness to stop before randomization, coagulation problems (e.g., deep venous thrombosis) and an estimated glomerular filtration rate (eGFR) below 30 ml/kg/min. Randomization was performed stratified by sex by the data management department. The code was kept at the data management and the pharmacy departments. The study was approved by the medical ethical review board of the UMC Utrecht and registered in clinical trial registries (NCT02839044/NTR5287). All participants gave written informed consent.
Intervention

Participants were randomized in a 1:1 ratio to 360 µg/day vitamin K or placebo treatment as previously described [28]. The vitamin K tablets contained 360 µg menaquinone-7. The placebo tablets contained the same raw material and were similar in taste and appearance. Menaquinone-7 (Vitamin K2) was used because of its higher bioavailability and longer half-life time compared to phylloquinones (vitamin K1) [3]. The dose used in this study is at the high end of what can be achieved through diet and has previously been shown to effectively decrease circulating inactive dephosphorylated-uncarboxylated (dp-uc)MGP levels without affecting coagulation [31]. Leftover vitamin K and placebo tablets were returned after the trial and the compliance was estimated as the number of tablets that were returned divided by the number of tablets that should have been taken × 100. The compliance was high in both the vitamin K group (β [95% CI]: 97.4% [92.3%; 99.1%]) and placebo arm (β [95% CI]: 97.8% [94.2%; 99.7%]) [28].

Dp-ucMGP level determination

Serum dp-ucMGP levels were determined with a sandwich ELISA using the IDS Automated Analyser IDS-iSYS InaKtif MGP assay (Maastricht University, The Netherlands).

Computed tomography (CT) acquisition and analysis

All participants underwent a low-dose, full-body CT scan at baseline and 6-month follow-up (Siemens, Healthcare, Erlangen, Germany). Arterial calcification mass was quantified in several larger arterial beds that can reliably be quantified on low-dose CT scans. The arteries were the intracranial and common carotid arteries, the coronary arteries, the aorta, the iliac arteries and the arteries of the legs (femoral and crural arteries combined). The findings in the femoral arteries were the primary outcome of the trial and were published elsewhere [28]. The calcification mass score was measured using an in-house developed program (iX Viewer, Image Science Institute, Utrecht, The Netherlands) as previously described [32, 33] The observer manually identified potential calcifications that were segmented using a threshold of 130 Hounsfield Units (HU) for calcification. The calcification mass score was computed as the product of the volume of the lesion (in ml) and the mean attenuation (in HU) of the lesion. Bone mineral density was assessed using an in-house developed software tool as described previously [34]. Thoracic and lumbar vertebrae were automatically segmented using an iterative segmentation approach. Fractures were visually identified and defined as > 20% height loss of one the pillars of the vertebral body. BMD of the trabecular core of the non-fractured vertebra was expressed as the mean HU in all thoracic and lumbar vertebrae.

Statistical analysis

Descriptive data are presented as categorical variables (n, %), normally distributed continuous (mean ± SD) or non-normally distributed (median, interquartile range). The difference in absolute and relative change in CT-based calcification mass score or BMD between the treatment and placebo group was analyzed with the Mann–Whitney U test. The relative change was calculated with the following formula: (x follow-up − x baseline)/x baseline × 100. In addition, to adjust for baseline differences, calcification mass was log + 1-transformed and linear regression models were built. These models included treatment (vitamin K or placebo, vitamin K as the reference) and log-transformed calcification mass of the individual arteries or BMD at baseline as the independent variables and log-transformed calcification mass or BMD after 6 months as the dependent variable (e.g., arterial calcification mass FU ~ treatment (vitamin K or placebo) + arterial calcification mass baseline). A p value <0.05 was regarded statistically significant. All analyses were performed in SPSS version 25.0, figures were made in Rstudio v1.1.456.

Results

Trial population

Sixty-eight eligible patients were randomized: 35 to the intervention and 33 to the placebo arm. Mean age was 69 years in both groups; 74% vs 79% were male in the intervention and the placebo groups, respectively. In the intervention group, two patients were lost to follow-up versus six patients in the placebo group. The loss to follow-up was unrelated to the intervention and was mostly due to discomfort with the PET/CT scan. Segmentation failed in 98 of the 2176 vertebrae (5%) and 14 (0.6%) of the segmented vertebrae were fractured and therefore excluded. Baseline characteristics of the intervention and placebo group are presented in Table 1.

Despite randomization, participants in the vitamin K arm had higher arterial calcification mass scores in all arterial beds compared to the placebo arm at baseline. These differences were statistically significant in the aorta [median (IQR): 742 (322–1337) vs 365 (39–1144), p = 0.05], iliac arteries [median (IQR): 633 (242–1148) vs 337 (66–764), p = 0.02] and the total arterial calcification mass score [median (IQR): 1694 (812–3584) vs 1182 (235–2445), p = 0.03] and was nearly significant in the
calcification mass score of the legs [median (IQR): 309 (93–851) vs 90 (11–627), \( p = 0.07 \)].

At baseline, no difference in plasma dp-ucMGP [613 (513–684) pmol/l vs. 615 (489–743) pmol/l, \( p = 0.96 \)] and BMD (all vertebrae: 151 ± 39 HU vs. 156 ± 43 HU, \( p = 0.62 \)) was found between the vitamin K and placebo arm, respectively (Table 1).

**The effect of vitamin K supplementation on arterial calcification mass and BMD**

No significant difference in arterial calcification progression between the vitamin K arm and the placebo arm was found for any arterial bed, although a trend toward lower progression in the placebo arm was found in the iliac arteries.

### Table 1 Baseline characteristics of the trial population

| Characteristic                                      | Vitamin K \( (n = 35) \) | Placebo \( (n = 33) \) |
|-----------------------------------------------------|---------------------------|------------------------|
| Age, years                                          | 69 ± 8                    | 69 ± 8                 |
| Male sex, \( n \) (%)                                | 26 (74)                   | 26 (79)                |
| Body mass index                                     | 31 ± 6                    | 31 ± 5                 |
| Systolic blood pressure, mmHg                       | 136 ± 21                  | 138 ± 14               |
| Diastolic blood pressure, mmHg                      | 70 ± 11                   | 74 ± 9                 |
| Current smokers, \( n \) (%)                         | 6 (17)                    | 4 (12)                 |
| Previous smokers, \( n \) (%)                       | 24 (67)                   | 18 (55)                |
| Never smokers, \( n \) (%)                          | 5 (15)                    | 11 (33)                |
| Medical history                                     |                           |                        |
| Cerebrovascular disease, \( n \) (%)                | 12 (34)                   | 11 (33)                |
| Coronary artery disease, \( n \) (%)                | 23 (66)                   | 18 (55)                |
| Peripheral arterial disease, \( n \) (%)            | 6 (17)                    | 10 (30)                |
| Abdominal aortic aneurysm, \( n \) (%)              | 2 (6)                     | 4 (12)                 |
| Medication                                          |                           |                        |
| Antihypertensive, \( n \) (%)                       | 30 (86)                   | 30 (90)                |
| Glucose lowering, \( n \) (%)                       | 30 (86)                   | 30 (90)                |
| Lipid lowering, \( n \) (%)                         | 16 (46)                   | 15 (46)                |
| Laboratory measurements                             |                           |                        |
| HbA1c, mmol/mol                                      | 57 ± 15                   | 60 ± 17                |
| Total cholesterol, mmol/l                           | 4.5 ± 1.3                 | 4.2 ± 1.2              |
| LDL cholesterol, mmol/l                             | 2.1 ± 0.9                 | 2.0 ± 0.9              |
| HDL cholesterol, mmol/l                             | 1.1 ± 0.3                 | 1.1 ± 0.3              |
| Triglycerides, mmol/l                               | 2.8 (1.8–3.4)             | 1.9 (1.5–2.7)          |
| dp-ucMGP, pmol/l                                    | 613 (513–684)             | 615 (489–743)          |
| Arterial calcification                              |                           |                        |
| Intracranial internal carotid artery, mass score    | 12 (3–25)                 | 3 (0–35)               |
| Common carotid artery, mass score                   | 3 (0–25)                  | 2 (0–10)               |
| Coronary arteries, mass score                       | 74 (13–165)               | 46 (1–148)             |
| Aorta, mass score                                   | 742 (322–1337)            | 365 (39–1144)          |
| Iliac arteries, mass score                          | 633 (242–1148)            | 337 (66–764)           |
| Leg arteries, mass score                            | 309 (93–851)              | 90 (11–627)            |
| Total arterial calcification, mass score            | 1694 (812–3584)           | 1182 (235–2445)        |
| Bone mineral density                                |                           |                        |
| Thoracic vertebrae, HU                              | 162 ± 44                  | 166 ± 45               |
| Lumbar vertebrae, HU                                | 126 ± 36                  | 133 ± 48               |
| All vertebrae, HU                                   | 151 ± 39                  | 156 ± 43               |

Values are \( n \) (%), mean ± SD or median (interquartile range)

\( HDL \) high density lipoprotein, \( LDL \) low density lipoprotein, \( dp-ucMGP \) = dephosphorylated uncarboxylated matrix glycoprotein, \( HU \) Hounsfield units
In addition, no difference in BMD decline was found between the groups (median (IQR): 3 (−2; 16) HU vs −1 (−5; 10) HU, p = 0.24) in the vitamin K and placebo arm, respectively (Table 2). When adjusted for baseline arterial calcification mass scores, 6 months of vitamin K treatment did not halt progression of arterial calcification in any arterial bed when compared to the placebo (β: −0.02; 95% CI: −0.10; 0.06 mg; p = 0.64 for the total arterial calcification mass). No effect of vitamin K treatment on BMD was found (β: −2.06; 95% CI: −11.26; 7.30 HU; p = 0.66 for all vertebrae). See Fig. 1 and Table 2 for individual arterial beds and vertebral and lumbar vertebrae.

### Discussion

This study shows that 6 months of vitamin K supplementation does not affect CT-measured arterial calcification or CT-measured BMD in patients with DM2 and a history of cardiovascular disease. Although vitamin K supplementation reduced circulating inactive MGP, as was shown in the original trial, our findings do not support that vitamin K supplementation has a role in the prevention of arterial calcification progression or osteoporosis in patients with DM2, but some limitations have to be acknowledged.

Several clinical trials investigating the effect of vitamin K supplementation on arterial calcification have been performed. Despite a consistent decrease in the circulating inactive form of MPG (dephosphorylated-uncarboxylated (dp-uc) MGP) after vitamin K supplementation [28, 31, 35, 36], these trials showed differing results on arterial calcification progression. One year treatment with vitamin K2 in hemodialysis patients did not affect aortic calcification progression [37], although the dose in this study was lower compared to our study. Another study with 3-year follow-up did not find an effect of multivitamin treatment with vitamin K1 when compared to multivitamin alone on the progression of coronary artery calcification, although a subgroup analysis showed a small effect in patients that were >85% adherent to the treatment [38]. A meta-analysis of clinical trials investigating the effect of vitamin K on calcification progression showed a positive effect of vitamin K treatment on arterial calcification progression [39]. This effect was mainly driven by a study into the effect of vitamin K on aortic valve calcification which showed that 12 months treatment with 2 mg vitamin K1 reduced the progression of aortic valve calcification (10% progression in vitamin K1 vs 22% in the placebo group) in 72 patients with aortic valve stenosis [40].

In our trial, there was a significant difference in arterial calcification mass scores in several arterial beds between the intervention and placebo groups despite randomization. Although we adjusted for these differences in the regression models, this might have affected the outcomes since—from the coronary arteries—it is known that baseline calcification mass is a strong predictor of calcification progression [41]. Another explanation might be that our patients did not have very severe vitamin K deficiency. Although serum dp-ucMGP levels in this population were relatively high when compared to the healthy population of comparable

### Table 2  Difference in arterial calcification mass score and bone mineral density after 6 months of vitamin K treatment or placebo

| Arterial calcification mass | Difference baseline and follow-up | Linear regression models adjusted for baseline measurements |
|----------------------------|----------------------------------|----------------------------------------------------------|
|                            | Vitamin K, N=33                  | Placebo, N=27                                            |
| Intracranial internal carotid artery | 0 (−1; 5)                        | 0 (−0; 2)                                                |
| Common carotid artery      | 0 (−0; 3)                        | 0 (−1; 0)                                                |
| Coronary arteries          | 5 (−5; 12)                       | 1 (−4; 11)                                               |
| Aorta                     | 40 (−30; 125)                    | 11 (0; 47)                                               |
| Iliac arteries            | 25 (6; 87)                       | 5 (−4; 30)                                               |
| Leg arteries              | 35 (−8; 99)                      | 7 (0; 47)                                                |
| Total arterial calcification score | 84 (−37; 206)                   | 36 (1; 129)                                              |
| Bone mineral density       |                                  |                                                          |
| Thoracic                  | 5 (−3; 15)                       | −2 (−5; 12)                                              |
| Lumbar                    | 3 (−5; 18)                       | 1 (−5; 6)                                                |
| Total BMD                 | 3 (−2; 16)                       | −1 (−5; 10)                                              |

Data are presented as median (IQR) for non-normally distributed variables. The difference in arterial calcification mass score progression and BMD decline between the vitamin K and placebo groups were estimated with the Mann–Whitney U test. In addition, to adjust for baseline differences, linear regression models were built with vitamin K and placebo treatment and log-transformed calcification mass or BMD at baseline as the determinants and log-transformed calcification mass or BMD after 6 months of follow-up as the outcome.
age (372–376 pmol/l) [27, 42], much higher levels up to 1100 pmol/l are reported in patients with chronic kidney disease [43, 44]. Selection of patients with severe vitamin K deficiency might be warranted for future trials.

Despite these differing results on calcification progression, several trials showed a reduction of arterial stiffness progression with vitamin K supplementation [36, 39, 45]. This challenges our understanding of the role of MGP in vascular physiology. Since calcification is shown to add to arterial stiffness, the positive effects in these trials are thought to be mediated by a reduction in arterial calcification [46]. Possibly, MGP is involved in other mechanisms that contribute to arterial stiffness, including reduced elastin degradation and collagen formation in the arterial wall, as well [47].

We could not confirm a positive effect of vitamin K supplementation on BMD in patients with type 2 diabetes, although the duration of treatment was relatively short and we used CT scans instead of DXA scans for the measurements. Some clinical trials have shown an increase of BMD after vitamin K treatment [21, 48], whereas others did not [22, 49]. A meta-analysis of 36 clinical trials that investigated the effect of vitamin K treatment on bone mineral density and fractures showed vitamin K treatment does not affect BMD in postmenopausal women, although it may reduce clinical fractures [23]. This suggests that vitamin K plays a role in bone architecture and quality, rather than in BMD. DM2 patients have an increased risk of fractures despite normal or even increased BMD [50]. Future studies might therefore include the effect of vitamin K on bone structure and fractures, rather than on BMD [51].

The strengths of this study include the randomized controlled design, excellent compliance and extensive measurements of systemic arterial calcification and that we used CT scans to quantify vertebral BMD. Although areal BMD (aBMD) as measured with DXA is currently the golden standard for the measurement of BMD, and CT scans provide the opportunity to exclude the cortical bone from the analysis. Both low aBMD as measured with DXA and volumetric BMD as measured with CT are shown to be associated with increased fractures, but vBMD of the lumbar spine

Fig. 1 No difference was found in absolute (a, c) and relative (b, d) differences in total body arterial calcification mass (a, b) and bone mineral density (c, d) after 6 months of vitamin K treatment (red) or placebo (blue).
has been shown to be a better predictor for spine fractures than areal BMD as measured with DXA scans [52].

However, several limitations should be acknowledged. The relatively high dropout rate, especially in the placebo arm, might have affected the statistical power. Since the sample size calculation of this study was based on active arterial calcification progression, as measured with $^{18}$NaF PET/CT, it might have been underpowered to detect changes in BMD and CT-measured calcifications in this relatively short follow-up period. The reasons for dropout were mainly due to discomfort with the $^{18}$NaF PET/CT. Future clinical trials might want to consider using conventional CT instead of $^{18}$NaF PET/CT, since this requires shorter scan time, no radiotracer needs to be infused and it has a lower radiation dose. Since the baseline characteristics of the participants that fulfilled the follow-up period were similar to the baseline characteristics of all participants that were randomized, we do not think that selection bias has affected our results. Since we included DM2 patients with cardiovascular diseases, and approximately 75% of our participants were male, our results might not be generalizable to broader populations. Since we quantified arterial calcification mass on whole body CT scans without ECG-gated cardiac CT, the coronary artery calcification mass scores might not be accurate and future research into the effect of vitamin K on coronary artery calcification progression should consider using triggered scans or more advanced motion and partial volume correction [53, 54].

In conclusion, 6 months of vitamin K supplementation did not halt progression of arterial calcification and did not affect BMD in patients with DM2 and cardiovascular disease. Given the limitations of our study future clinical trials may want to pre-select patients with very low vitamin K status and longer follow-up time might be warranted.

**Funding** JWJB and SRZ were supported by the Senior Dr. Dekker Grant (2013T120) from the Dutch Heart Foundation.

**Compliance with ethical standards**

**Conflict of interest** The authors declare no conflicts of interest.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article’s Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

**References**

1. Halder M, Petsophonsakul P, Akbulut AC, Pavlic A, Bohan F, Anderson E, Maresz K, Kramann R, Schurgers L (2019) Vitamin K: double bonds beyond coagulation insights into differences between vitamin K1 and K2 in health and disease. Int J Mol Sci. https://doi.org/10.3390/ijms20040896

2. Schurgers LJ, Vermeer C (2000) Determination of phyloquinone and menaquinones in food. Effect of food matrix on circulating vitamin K concentrations. Haemostasis 30(6):298–307. https://doi.org/10.1159/0000541417

3. Schurgers LJ, Teunissen KJ, Hamulyak K, Knape MH, Vink H, Vermeer C (2007) Vitamin K-containing dietary supplements: comparison of synthetic vitamin K1 and natto-derived menaquinone-7. Blood 109(8):3279–3283. https://doi.org/10.1182/blood-2006-08-040709

4. Vossen LM, Kroon AA, Schurgers LJ, de Leeuw PW (2019) Pharmacological and nutritional modulation of vascular calcification. Nutrients. https://doi.org/10.3390/nu12010100

5. Myneni VD, Mezey E (2017) Regulation of bone remodeling by vitamin K2. Oral Dis 23(8):1021–1028. https://doi.org/10.1111/odi.12624

6. Cozzolino M, Fusaro M, Ciceri P, Gasparroni L, Cianciolo G (2019) The role of vitamin K in vascular calcification. Adv Chronic Kidney Dis 26(6):437–444. https://doi.org/10.1053/j.ackd.2019.10.005

7. Schurgers LJ, Uitto J, Reutelingsperger CP (2013) Vitamin K-dependent carboxylation of matrix Gla-protein: a crucial switch to control ectopic mineralization. Trends Mol Med 19(4):217–226. https://doi.org/10.1016/j.molmed.2012.12.008

8. Zoch ML, Clemens TL, Riddle KC (2016) New insights into the biology of osteocalcin. Bone 82:42–49. https://doi.org/10.1016/j.bone.2015.05.046

9. Kaasenbrood L, Boekholdt SM, van der Graaf Y, Ray KK, Peters RJ, Kastelein JJ, de Borst GJ, Visseren FL (2016) Distribution of estimated 10-year risk of recurrent vascular events and residual risk in a secondary prevention population. Circulation 134(19):1419–1429. https://doi.org/10.1161/CIRCULATIONAHA.116.021314

10. Mazzone T, Chait A, Plutzky J (2008) Cardiovascular disease risk in type 2 diabetes mellitus: insights from mechanistic studies. Lancet 371(9626):1800–1809. https://doi.org/10.1016/S0140-6736(08)60768-0

11. Lanzer P, Boehm M, Sorribas V, Thiriet M, Janzen J, Zeller T, St Hilaire C, Shanahan C (2014) Medial vascular calcification revisited: review and perspectives. Eur Heart J 35(23):1515–1525. https://doi.org/10.1093/eurheartj/ehu163

12. Kramer CK, Zinnman B, Gross JL, Canani LH, Rodrigues TC, Azevedo MJ, Retnakaran R (2013) Coronary artery calcium score prediction of all cause mortality and cardiovascular events in people with type 2 diabetes: systematic review and meta-analysis. BMJ 346:f1654. https://doi.org/10.1136/bmj.f1654

13. Rennenberg RJ, Kessels AG, Schurgers LJ, van Engelsroven JM, de Leeuw PW, Kroon AA (2009) Vascular calcifications as a marker of increased cardiovascular risk: a meta-analysis. Vasc Health Risk Manag 5(1):185–197. https://doi.org/10.2147/vhrm.s4822

14. Ho CY, Shanahan CM (2016) Medial arterial calcification: an overlooked player in peripheral arterial disease. Arterioscler Thromb Vasc Biol 36(8):1475–1482. https://doi.org/10.1161/ATVBAHA.116.306717

15. Bos D, Portegies ML, van der Lugt A, Bos MJ, Koudstaal PJ, Hofman A, Krestin GP, Franco OH, Vemmooy MW, Ikram MA (2014) Intracranial carotid artery atherosclerosis and the risk of stroke in...
19. Chen HG, Sheng LT, Zhang YB, Cao AL, Lai YW, Kunutsor SK, Dalmeijer GW, van der Schouw YT, Magdeleyns EJ, Vermeer Pivin E, Ponte B, Pruijm M, Ackermann D, Guessous I, Ehret G, Bartstra JW, Mali WP, Spiering W, de Jong PA (2020) Abdominal aortic calcification: from ancient friend to modern foe. Eur J Prev Cardiol. https://doi.org/10.1177/2047487320199895

18. Pivin E, Ponte B, Prijlim D, Ackermann D, Guessous I, Ehret G, Liu YP, Drummen NE, Knapen MH, Pechere-Bertsch A, Paccaud F, Mohaupt M, Vermeer C, Staessen JA, Vogt B, Martin PY, Burnier M, Bochud M (2015) Inactive matrix GlA protein is associated with arterial stiffness in an adult population-based study. Hypertension 66(1):85–92. https://doi.org/10.1161/HYPERTENSIONAHA.115.05177

17. Dalmeijer GW, van der Schouw YT, Magdeleyns EJ, Vermeer C, Verschuren WM, Boer JM, Beulens JW (2013) Matrix GlA protein species and risk of cardiovascular events in type 2 diabetic patients. Diabetes Care 36(11):3766–3771. https://doi.org/10.2337/dc13-0056

16. Bartstra JW, Mali WP, Spiering W, de Jong PA (2020) Abdominal aortic calcification: from ancient friend to modern foe. Eur J Prev Cardiol. https://doi.org/10.1177/2047487320199895

15. Pivin E, Ponte B, Prijlim D, Ackermann D, Guessous I, Ehret G, Liu YP, Drummen NE, Knapen MH, Pechere-Bertsch A, Paccaud F, Mohaupt M, Vermeer C, Staessen JA, Vogt B, Martin PY, Burnier M, Bochud M (2015) Inactive matrix GlA protein is associated with arterial stiffness in an adult population-based study. Hypertension 66(1):85–92. https://doi.org/10.1161/HYPERTENSIONAHA.115.05177

13. Chen HG, Sheng LT, Zhang YB, Cao AL, Lai YW, Kunutsor SK, Dalmeijer GW, van der Schouw YT, Magdeleyns EJ, Vermeer C, Vermeer C, Staessen JA, Vogt B, Martin PY, Burnier M, Bochud M (2015) Inactive matrix GlA protein is associated with arterial stiffness in an adult population-based study. Hypertension 66(1):85–92. https://doi.org/10.1161/HYPERTENSIONAHA.115.05177

11. Chen HG, Sheng LT, Zhang YB, Cao AL, Lai YW, Kunutsor SK, Dalmeijer GW, van der Schouw YT, Magdeleyns EJ, Vermeer C, Vermeer C, Staessen JA, Vogt B, Martin PY, Burnier M, Bochud M (2015) Inactive matrix GlA protein is associated with arterial stiffness in an adult population-based study. Hypertension 66(1):85–92. https://doi.org/10.1161/HYPERTENSIONAHA.115.05177

9. Chen HG, Sheng LT, Zhang YB, Cao AL, Lai YW, Kunutsor SK, Dalmeijer GW, van der Schouw YT, Magdeleyns EJ, Vermeer C, Vermeer C, Staessen JA, Vogt B, Martin PY, Burnier M, Bochud M (2015) Inactive matrix GlA protein is associated with arterial stiffness in an adult population-based study. Hypertension 66(1):85–92. https://doi.org/10.1161/HYPERTENSIONAHA.115.05177

7. Chen HG, Sheng LT, Zhang YB, Cao AL, Lai YW, Kunutsor SK, Dalmeijer GW, van der Schouw YT, Magdeleyns EJ, Vermeer C, Vermeer C, Staessen JA, Vogt B, Martin PY, Burnier M, Bochud M (2015) Inactive matrix GlA protein is associated with arterial stiffness in an adult population-based study. Hypertension 66(1):85–92. https://doi.org/10.1161/HYPERTENSIONAHA.115.05177

5. Chen HG, Sheng LT, Zhang YB, Cao AL, Lai YW, Kunutsor SK, Dalmeijer GW, van der Schouw YT, Magdeleyns EJ, Vermeer C, Vermeer C, Staessen JA, Vogt B, Martin PY, Burnier M, Bochud M (2015) Inactive matrix GlA protein is associated with arterial stiffness in an adult population-based study. Hypertension 66(1):85–92. https://doi.org/10.1161/HYPERTENSIONAHA.115.05177

3. Chen HG, Sheng LT, Zhang YB, Cao AL, Lai YW, Kunutsor SK, Dalmeijer GW, van der Schouw YT, Magdeleyns EJ, Vermeer C, Vermeer C, Staessen JA, Vogt B, Martin PY, Burnier M, Bochud M (2015) Inactive matrix GlA protein is associated with arterial stiffness in an adult population-based study. Hypertension 66(1):85–92. https://doi.org/10.1161/HYPERTENSIONAHA.115.05177

1. Chen HG, Sheng LT, Zhang YB, Cao AL, Lai YW, Kunutsor SK, Dalmeijer GW, van der Schouw YT, Magdeleyns EJ, Vermeer C, Vermeer C, Staessen JA, Vogt B, Martin PY, Burnier M, Bochud M (2015) Inactive matrix GlA protein is associated with arterial stiffness in an adult population-based study. Hypertension 66(1):85–92. https://doi.org/10.1161/HYPERTENSIONAHA.115.05177

European Journal of Nutrition (2021) 60:1691–1699

https://doi.org/10.3389/fendo.2019.00891

https://doi.org/10.1148/radiol.2461070006

https://doi.org/10.1101/j.mediatea.2019.02.005

https://doi.org/10.1148/radiol.2461070006

https://doi.org/10.1101/j.mediatea.2019.02.005

https://doi.org/10.1148/radiol.2461070006

https://doi.org/10.1101/j.mediatea.2019.02.005

https://doi.org/10.1148/radiol.2461070006

https://doi.org/10.1101/j.mediatea.2019.02.005

https://doi.org/10.1148/radiol.2461070006

https://doi.org/10.1101/j.mediatea.2019.02.005

https://doi.org/10.1148/radiol.2461070006

https://doi.org/10.1101/j.mediatea.2019.02.005

https://doi.org/10.1148/radiol.2461070006

https://doi.org/10.1101/j.mediatea.2019.02.005
association with 15 biomarkers. JACC Cardiovasc Imaging 10(8):858–866. https://doi.org/10.1016/j.jcmg.2017.05.010
42. Machado-Fragua MD, Hoogendijk EO, Struijk EA, Rodriguez-Artalejo F, Lopez-Garcia E, Beulens JW, van Ballegooijen AJ (2020) High dephospho-uncarboxylated matrix Gla protein concentrations, a plasma biomarker of vitamin K, in relation to frailty: the longitudinal aging study Amsterdam. Eur J Nutr 59(3):1243–1251. https://doi.org/10.1007/s00394-019-01984-9
43. Schurgers LJ, Barreto DV, Barreto FC, Liabeuf S, Renard C, Magdeleyns EJ, Vermeer C, Choukroun G, Massy ZA (2010) The circulating inactive form of matrix gla protein is a surrogate marker for vascular calcification in chronic kidney disease: a preliminary report. Clin J Am Soc Nephrol 5(4):568–575. https://doi.org/10.2215/CJN.07081009
44. Karnatowska I, Grzelak P, Masajtis-Zagajewska A, Kaczmarska M, Stefanczyk L, Vermeer C, Maresz K, Nowicki M (2015) Effect of vitamin K2 on progression of atherosclerosis and vascular calcification in nondialyzed patients with chronic kidney disease stages 3–5. Pol Arch Med Wewn 125(9):631–640. https://doi.org/10.20452/pamw.3041
45. Braam LA, Hoeks AP, Brouns F, Hamulyak K, Gerichhausen MJ, Vermeer C (2004) Beneficial effects of vitamins D and K on the elastic properties of the vessel wall in postmenopausal women: a follow-up study. Thromb Haemost 91(2):373–380. https://doi.org/10.1160/TH03-07-0423
46. Tsao CW, Pencina KM, Massaro JM, Benjamin EJ, Levy D, Vasan RS, Hoffmann U, O’Donnell CJ, Mitchell GF (2014) Cross-sectional relations of arterial stiffness, pressure pulsatility, wave reflection, and arterial calcification. Arterioscler Thromb Vasc Biol 34(11):2495–2500. https://doi.org/10.1161/ATVBAHA.114.303916
47. Cocciolone AJ, Hawes JZ, Staiculescu MC, Johnson EO, Murshed M, Wageseil JE (2018) Elastin, arterial mechanics, and cardiovascular disease. Am J Physiol Heart Circ Physiol 315(2):H189–H205. https://doi.org/10.1152/ajpheart.00087.2018
48. Je SH, Joo NS, Choi BH, Kim KM, Kim BT, Park SB, Cho DY, Kim KN, Lee DJ (2011) Vitamin K supplement along with vitamin D and calcium reduced serum concentration of under-carboxylated osteocalcin while increasing bone mineral density in Korean postmenopausal women over sixty-years-old. J Korean Med Sci 26(8):1093–1098. https://doi.org/10.3346/jkms.2011.26.8.1093
49. Volpe SL, Leung MM, Giordano H (2008) Vitamin K supplementation does not significantly impact bone mineral density and biochemical markers of bone in pre- and perimenopausal women. Nutr Res 28(9):577–582. https://doi.org/10.1016/j.nutre.2008.06.006
50. Iwamoto J, Sato Y, Takeda T, Matsumoto H (2011) Bone quality and vitamin K2 in type 2 diabetes: review of preclinical and clinical studies. Nutr Rev 69(3):162–167. https://doi.org/10.1111/j.1753-4887.2011.00380.x
51. Chang G, Boone S, Martel D, Rajapakse CS, Hallyburton RS, Valko M, Honig S, Regatte RR (2017) MRI assessment of bone structure and microarchitecture. J Magn Reson Imaging 46(2):323–337. https://doi.org/10.1002/jmri.25647
52. Chalhoub D, Orwell ES, Cawthon PM, Ensrud KE, Boudreau R, Greenspan S, Newman AB, Bauer D, Cummings S, Cauley JA, Osteoporotic Fractures in Men Study Research G (2016) Areal and volumetric bone mineral density and risk of multiple types of fracture in older men. Bone 92:100–106. https://doi.org/10.1016/j.bone.2016.08.014
53. Sprem J, de Bos JD, Lessmann N, de Jong PA, Viergever MA, Isgum I (2018) Impact of automatically detected motion artifacts on coronary calcium scoring in chest computed tomography. J Med Imaging (Bellingham) 5(4):044007. https://doi.org/10.1117/1.JMI.5.4.044007
54. Sprem J, de Bos JD, Lessmann N, van Hamersvelt RW, Greuter MJW, de Jong PA, Leiner T, Viergever MA, Isgum I (2018) Coronary calcium scoring with partial volume correction in anthropomorphic thorax phantom and screening chest CT images. PLoS ONE 13(12):e0209318. https://doi.org/10.1371/journal.pone.0209318