Aboveground and below-ground carbon allocation of summer rape under elevated CO₂ and air temperature

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In studies on plant responses to climate change more attention has been given to aboveground processes although carbon input by plants into the soil is a major flux in the global carbon cycle. The objective of study was to investigate the effects of elevated CO₂ and temperature on carbon allocation and partitioning in different parts of plant, soil, and microbial biomass. An experiment was conducted on summer rape (Brassica napus L.) under increased levels of air temperature and atmospheric CO₂ in controlled environment chambers. Results showed that the amount of leaf, stem and root carbon statistically significantly increased under elevated CO₂ and temperature conditions. Microbial biomass carbon significantly increased by 11.2% and 13.5% under elevated CO₂ and elevated CO₂ and temperature, respectively, although soil carbon under both treatments decreased. It is concluded that carbon allocation is controlled under different climate conditions; however, elevated CO₂ and temperature together will have a more significant effect for carbon allocation to different plant parts and microbial biomass carbon compared to elevated CO₂ alone.

Key words: carbon allocation, soil carbon, microbial biomass carbon, leaf area, summer rape

Introduction

Studies on plant responses to increased atmospheric CO₂ concentration and elevated temperatures have become abundant in the last 20 years (Franks et al. 2013). Allocation of carbon to different plant organs is central in this discussion and provides a mechanism by which plants can adapt to changes in the environment (Chaves et al. 2002). Carbon partitioning varies with plant development stage (reflecting changing priorities) but also depends on species-specific strategies (Weiner 2004), such as preferred allocation in below-ground storage compounds (Kuzyakov and Domanski 2000), responses to environmental conditions, e.g. drought or other stress factors (Sanaullah et al. 2012).

Although usually more attention is given to aboveground processes, carbon input by plants into the soil is a major flux in the global carbon cycle and is crucial not only for carbon sequestration, but also for maintenance of soil fertility, ecosystem stability, and functions (Pausch and Kuzyakov 2017).

Below-ground processes play a key role in global carbon cycle because they regulate the storage of large quantities of carbon and are potentially very sensitive to direct and indirect effects of elevated CO₂ and temperature (Pendall et al. 2004). Elevated CO₂ increases carbon supply to below-ground parts of plants, whereas warming is likely to increase respiration and decomposition rates, leading to speculations that these effects will moderate one another (Pendall et al. 2004). The variations in plant carbon allocation suggest there could be significant differences in plant carbon allocation across crop types, climatic zones, and soil types. The differences are certainly critical in the eventual deposition of plant carbon into soil carbon pools and can be used to select crop varieties with superior carbon sequestration potential (Mathew et al. 2017). Therefore, it is important to understand the carbon input to different parts (root vs. shoot) to strategize options intended to increase soil organic carbon (Rasse et al. 2005). Additionally, microbial biomass has been used as a sensitive indicator of alteration patterns in soil organic matter (Balota et al. 2003). If climate change alters soil microbial communities and this change determines plant species establishment and growth, then ecosystem responses will be dependent on the interactions between plants and soil communities (Clasen et al. 2015).

The objective of this study was to investigate the effects of elevated CO₂ and temperature on biomass accumulation and carbon content in different parts of Brassica napus, soil carbon, and microbial biomass, to investigate the physiological and morphological responses of the crop to different climatic treatments and to ascertain whether elevated CO₂ and elevated CO₂ and temperature, projected by the end of this century, would differentially affect carbon allocation to different plant parts.
Material and methods

Plant material and growth conditions

The experiment was conducted in the growth chambers in the Vytautas Magnus University. Summer rape (Brassica napus L.) was chosen because it is one of the major crops of the EU27 member states (Donatelli et al. 2015) and one of the most popular crops in Lithuania. Seeds of summer rape (Brassica napus L. var. ‘Fenja’) (15 seeds per container) were sown in 3 l plastic containers (height 10.6 cm) containing a growth substrate composed of a mixture of field soil (the soil was taken from ASU Training Farm, Kaunas District), perlite, and fine sand (5:3:2 by volume). In the control chamber plants were grown under conditions of current climate—an average day/night temperature of 21/14 °C and 400 µmol mol⁻¹ of CO₂ (CON). Elevated CO₂ and temperature (day/night temperature of 25/18 °C and 800 ppm of CO₂, ETC) and elevated CO₂ (day/night temperature of 21/14 °C and 800 ppm of CO₂, EC) treatments started when the seedlings of summer rape were germinated, and lasted for 4 weeks. The elevated temperature and CO₂ concentrations were increased to the level in accordance with the climate change scenario for Lithuania (Juknys et al. 2017). The pre-set values of the air temperature in the growth chambers were controlled manually at each of the growth chambers (Emerson Network Power S.r.l., Italy, model No. S06UC021V300020FX051260). The concentration of CO₂ was manipulated automatically by controlling the amount of injected CO₂ gas and chamber conditioner. The climate program was controlled by the IGSS 9-13175 software. The following stable conditions were maintained in all chambers: a photoperiod of 14 h, relative humidity of 50/60%, and 226 µmol m⁻² s⁻¹, on average, photon flux density of photosynthetically active radiation (PAR). A nutrient supply corresponding to 120 kg N ha⁻¹ was used after sowing the seeds. Additional fertilization with a complex nutrient (NPK 12-11-18 + microelements) solution, increasing the N level to 180 kg N ha⁻¹, was applied in the middle of treatments (2 weeks after the seedlings were germinated). The pots in the chambers were watered sufficiently and regularly. All treatments were run in three replicates (three pots per chamber). Avoiding edge effects, the pots were rotated every second day.

Photosynthesis rate and leaf area measurement

Photosynthetic rates (Pr, µmol CO₂ m⁻² s⁻¹) were measured with a portable photosynthesis system LI-6400 (LI-COR, USA) equipped with a 6 cm² leaf chamber. Photosynthetic rates were recorded automatically for approximately 5 minutes every 3 s when Pr reached the steady state level. The measurements were made on the most recent fully expanded leaves (i.e. one leaf per plant) of intact plants (randomly chosen plants were measured). Three plants per pot (the average of them was taken for statistical analysis) and three pots per treatment were measured (n = 3) under ambient, elevated CO₂, and elevated CO₂ and temperature climate treatments from 10:00 h to 15:00 h on the last day of the experiment. During the measurements, leaf chamber conditions were controlled at 400 or 800 µmol mol⁻¹ CO₂, and 21 or 25 °C (block temperature), according to the climate treatments. Airflow rate through the assimilation chamber was maintained at 500 µmol s⁻¹. The water vapour concentration of air entering the leaf chamber was not controlled and tracked ambient conditions. Relative humidity was 51 ± 0.9% in CON, 62 ± 3.5% in EC, and 39 ± 1.6% in ETC treatment (mean ± SE). PAR outside the leaf chamber was 226 ± 4.0 µmol m⁻² s⁻¹, on average, under the different climate conditions.

The measurements of leaf area were carried out on the last day of treatment. The leaf area per plant (three replicates per treatment) was measured with a scanner (CanoScan 4400F, Canon, USA) and then the leaf area was determined by GIMP 2.8 software. Leaf area ratio (LAR) was calculated as the ratio of leaf area and total plant weight.

Dry weight and carbon content measurements

Samples of plant leaves, stems, and roots were dried in an air-forced oven at 70 °C until a constant dry weight was obtained (at least 72 hours). Soil samples were also taken at the end of the experiment. The samples were air dried at room temperature and sieved through 2 mm mesh to remove roots and plant remains. The dried samples of shoots, roots, and soil were ground to a fine powder with a mill (Retsch HM400, Germany).

Organic carbon content (%) in dried plant and soil samples was analyzed with a Shimadzu TOC-V solid sample module SSM-5000A. The root carbon stock (Rcs) and shoot carbon stock (Scs) were defined as the total amount of carbon (g) in dry weight measured in the corresponding plant parts. Microbial biomass carbon was determined by a chloroform fumigation direct extraction method (Beck et al. 1997).
Data analysis

Statistical analyses were carried out using STATISTICA 8 software. Mean values of the parameters (plant photosynthetic rate (Pr), LAR, plant dry weight, carbon content of leaf, stem and root, the ratio of root carbon stock and shoot carbon stock (Rcs/Scs), microbial biomass carbon, and soil carbon) and their standard errors of mean (±SE) were calculated. The Mann-Whitney U-test was used to estimate the difference between parameters under different climatic conditions. Spearman rank correlation was used to determine the strength of relationships between variables.

Results

Changes in plant photosynthesis and dry weight

The analysis of photosynthesis and carbon allocation is useful for understanding how the plants will respond to climate change, including the impact on biomass production. Changes in total plant dry weight and dry weight of different parts of plant, LAR, and Pr are presented in Figure 1. Total plant dry weight was significantly higher under climate treatment conditions (elevated CO$_2$ [EC] and elevated CO$_2$ and temperature [ETC]) in comparison with the ambient climate (CON) ($p<0.05$) (Fig. 1.A). ETC significantly increased the dry mass of plants on average by 142% ($p<0.05$). Elevated CO$_2$ and temperature had the highest positive effect on leaf dry weight (Fig. 1.A): the amount of leaf dry weight increased by 142% ($p<0.05$) under ETC compared to CON conditions. Both EC and ETC significantly decreased LAR: by 30% and by 46%, respectively, compared to CON (Fig. 1.B). The photosynthetic rate was significantly higher under EC and ETC, compared to CON – 29% and 75%, respectively ($p<0.05$, Fig. 1.C).

Carbon content in different plant parts

The carbon content increased in all plant parts under ETC conditions, but there was almost no changed under EC. Carbon content in leaf and root was significantly higher by 3% ($p<0.05$) and by 6% ($p<0.05$), respectively, under ETC, compared to ambient climate conditions (Fig. 2.A). Also, elevated CO$_2$ and temperature have had the highest positive effect on stem carbon: the content of stem carbon increased by 19% ($p<0.05$) under ETC compared to CON conditions. However, EC had less impact on stem carbon changes—its amount increased only by 2% compared to ambient climate conditions ($p>0.05$). Correlation analysis showed a significant positive relationship between photosynthesis, plant biomass and carbon content in different plant parts ($p<0.05$). While photosynthesis increased, plant biomass ($r=0.88$, $p<0.05$), carbon content in plant leaf ($r=0.72$, $p<0.05$), stem ($r=0.85$, $p<0.05$), and root ($r=0.68$, $p<0.05$) also increased.
To assess carbon allocation to roots under different climate conditions, the ratio of root carbon stock (Rcs) to shoot carbon stock (Scs) was determined (Fig. 2B). Under EC climate conditions, Rcs/Scs ratio increased by 8% (p>0.05); while under ETC conditions, the Rcs/Scs ratio decreased by 13% (p>0.05) compared to ambient climate conditions. The Rcs/Scs ratio decreased under ETC conditions, because the increase in Scs was relatively much higher than the increase in Rcs under ETC conditions.

Microbial biomass carbon and soil carbon

Microbial biomass carbon increased by 11.2% (p<0.05) and by 13.5% (p<0.05) under EC and ETC conditions, respectively, although soil carbon decreased by 5% (p>0.05) and by 6% (p>0.05), respectively, compared to ambient climate conditions (Fig. 3). There was no significant correlation between microbial biomass carbon and soil carbon, but it was estimated that microbial biomass carbon significantly correlated with plant root biomass. While plant root biomass increased, microbial biomass carbon also was statistically significantly higher (r=0.71, p<0.05). Soil carbon was negatively correlated with the carbon content in plant stem, stem biomass, and leaf biomass. While plant stem carbon (r=−0.72, p<0.05), stem biomass (r=−0.7, p<0.05), leaf biomass (r=−0.74, p<0.05) increased, soil carbon significantly decreased.

Fig. 2. A. Change of carbon content in different plant parts (%) under changed climate conditions compared with ambient climate. (* - statistically significant difference, compared to ambient climate conditions at P< 0.05). B. The ratio of root carbon stock (Rcs) and shoot carbon stock (Scs) under different climate conditions. Different letters indicate significant differences among treatments (p<0.05) based on U-test.

CON = ambient temperature and CO₂, EC = elevated CO₂, ETC = elevated temperature and CO₂.

Fig. 3. Microbial biomass carbon and soil carbon under modified climate conditions (mean ± SE). Different letters indicate significant differences among treatments (p<0.05) based on U-test. Treatments: CON = ambient temperature and CO₂, EC = elevated CO₂, ETC = elevated temperature and CO₂.
Discussion

The results showed that the photosynthetic rate under elevated CO₂ was 29% (p<0.05) higher, and under elevated CO₂ and temperature, 75% (p<0.05) higher compared to ambient climate conditions. With the reference to Ainsworth and Long (2005), the elevated CO₂ stimulates photosynthetic carbon assimilation rate by an average of 31%, which is similar to the current experiment. This stimulation of photosynthesis in C₃ plants such as *Brassica napus* due to elevated CO₂ occurs because Rubisco is CO₂ substrate-limited at ambient CO₂ (Long et al. 2004, Tcherkez et al. 2006). Elevated CO₂ enhances photosynthesis and, in turn, dry matter accumulation increases (Lawlor and Mitchell 2000, Ainsworth and Long 2005, Taub and Wang 2013). This is in accordance with the current experiment results; elevated CO₂ increased the biomass of *Brassica napus* by 1.3 times, and elevated both CO₂ and temperature by 2.4 times compared to ambient climate conditions. Positive interaction between elevated CO₂ and increased air temperature on photosynthesis and biomass production of C₃ plants has also been reported in other studies (Vu 2005, Borjigida et al. 2006, Alonso et al. 2009, Yoon et al. 2009, Juknys et al. 2011, 2012, Kaciene et al. 2017). This positive interaction between elevated CO₂ and temperature is explained by increased optimal temperature for plant growth (Long and Drake 1991, McMurtrie and Wang 1993) and net photosynthesis (Bernacchi et al. 2006, Alonso et al. 2009) under elevated CO₂. According to Long and Drake (1991), the optimal temperature for many C₃ plants may increase by approximately 5 °C, as CO₂ increases by 300 μmol mol⁻¹, as was the case in the current experiment.

However, an increase of the photosynthetic rate, which often is the result of increased CO₂, is not necessarily directly linked to higher crop production and yield (French et al. 2011). As reported by French et al. (2011), only in one of four *Brassica napus* cultivar (‘Bolero’) the biomass was significantly increased under elevated CO₂. It should be realized that there is no 1:1 translation of a photosynthetic CO₂ response into a growth response, as is highlighted in the review by Körner (2006). The discrepancy between the almost uniform stimulation of leaf photosynthetic rates in proportion to a rise in CO₂ concentration and rather variable growth responses, from zero to a large positive effect, has confounded researchers for as long as this research has been conducted, and that puzzle has not been resolved (Nowak et al. 2004). In addition, it is well known that the initial stimulation of photosynthetic rate by elevated CO₂ for most C₃ plants is temporal, and slows with future exposure, particularly under relatively long-term impact of elevated CO₂, and stabilizes at a lower level, the phenomenon known as downregulation (Kant et al. 2012, Xu et al. 2015). Because photosynthetic downregulation may be both plant development and species-ecotype dependent (Li et al. 2008, Aranjuelo et al. 2009, Kaplan et al. 2012), the extent of the increase in photosynthesis under elevated CO₂ conditions varies greatly among the species and even different varieties and functional groups of plants (Long et al. 2004, Leakey et al. 2009). Global changes in photosynthetic uptake could lead to a rapid response from short-lived C pools (such as foliage, fine roots, and litter) or a prolonged response from long-lived C pools (such as woody biomass and soil C) with very different outcomes on ecosystem source-sink behavior (Bloom et al. 2016).

Short-term C assimilation is typically linked to growth, which contradicts evidence that show significant temporal lags between assimilation and leaf/stem growth (Zweifel et al. 2006, Gough et al. 2009, Richardson et al. 2013). Results showed that carbon content increased in all *Brassica napus* parts under elevated CO₂ and temperature conditions, but it almost was unchanged under elevated CO₂ compared to ambient climate conditions. Leaf carbon increased only by 3% (p<0.05) under elevated CO₂ and temperature conditions, which was probably due to rapid carbon loss by leaf respiration and carbon export to other pools (Hill et al. 2007, Wu et al. 2010). Elevated CO₂ and temperature had the highest positive effect on stem carbon—the amount of stem carbon increased by 19% (p<0.05) compared to ambient climate conditions. Also, the Rcs/Scs analysis showed the amount of carbon was increased in shoots under elevated temperature and CO₂ conditions. According to Mathew et al. (2017), generally, all plants allocate more carbon in the shoots, showing that roots are relatively weaker carbon sinks compared to shoots. Shoot carbon stocks are higher than root carbon stocks because carbon is only exported to other sinks when the supply exceeds local demand (Ludewig and Flügge 2013). Also, Pausch and Kuziyakov (2017) showed that the main part of assimilated C remains aboveground and is used for shoot respiration and for shoot biomass production or C storage. According to results of the current experiment, elevated CO₂ increased the carbon content in both roots and shoots, especially in ETC, compared to ambient climate conditions. However, the Rcs/Scs decreased in ETC due to higher relative increase in carbon in the shoots. The reason may be that CO₂ enrichment increases carbon partitioning to the rapidly cycling carbon pools (below-ground) and root turnover due to increased demand for below-ground resources (Hungate et al. 1997, Ge et al. 2012). Rhizodeposition consists of a continuous flow of carbon-containing compounds from the roots to the soil (Gougoulias et al. 2014). Increased atmospheric CO₂ stimulates photosynthesis (Dijkstra et al. 2005, Hungate et al. 2006) and the release of root exudates, which in turn, means more labile carbon available for microbial decomposition and respiration (Ainsworth and Long 2005, Heath et al. 2005, Rayner et al. 2005, Friedlingstein et al. 2006, Hungate et al. 2006).
Microbial biomass carbon increased under elevated CO$_2$ and elevated CO$_2$ and temperature conditions compared to the ambient climate in the current experiment, although soil carbon decreased under both climatic treatments compared to ambient climate. Jackson et al. (2017) indicated increases in carbon uptake by plants under elevated atmospheric CO$_2$ might be partially offset by the accelerated loss of soil carbon due to plant-induced stimulation of microbial decomposition. Also, the decline in soil carbon may be driven by changes in soil microbial composition and activity. Soils exposed to elevated CO$_2$ have higher relative abundances of fungi and higher activities of a soil carbon degrading enzyme, which led to more rapid rates of soil organic matter degradation than soils exposed to ambient CO$_2$ (Carney et al. 2007). Allison et al. (2010) concluded, that the soil carbon response to climate warming depends on the efficiency of soil microbes in using carbon, however according to Pausch and Kuzyakov (2017), total carbon allocated below-ground also depends on photosynthetic intensity.

Conclusion

The analysis of photosynthesis and carbon allocation is useful to ascertain how the plants will respond to climate change, including the impact on biomass production. The objective of this study was to investigate the effect of climate change for carbon allocation in different parts of the plant, soil carbon, and microbial biomass carbon and to investigate physiological and morphological responses of crop Brassica napus to different climatic conditions. Results show increased biomass allocation in all plant parts under elevated CO$_2$, in both temperature treatments, but effects of CO$_2$ on crop and soil carbon contents differed. Stem carbon of the crop was most positively affected (19%) by elevated CO$_2$ and temperature, while exposure to CO$_2$ alone had almost no effect on the amount of carbon in different plant parts. Soil carbon decreased under elevated CO$_2$ and elevated CO$_2$ and temperature conditions, and it was estimated that the decline in soil carbon was driven by changes in soil microbial composition and activity. Microbial biomass carbon increased statistically significantly by 11.2% and by 13.5% under elevated CO$_2$ and elevated CO$_2$ and temperature conditions, respectively. Physiological and morphological responses of crop Brassica napus to different climatic treatments showed stimulation of plant growth. Accelerated plant growth was shown by higher plant dry weight and increased photosynthetic rate.

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