Tumor vascularity in ovarian cancer

T cells need breathing room

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In ovarian cancer, it has become increasingly clear that tumor-infiltrating lymphocytes (TILs) confer a survival benefit to patients.1 It is tempting to strive to further define the key TIL subsets driving antitumor immunity, as one can envision developing T cell-based therapy armed with this knowledge. Indeed, refinements in adoptive T-cell therapy seeking to identify antigen-specific populations are currently under intense investigation in melanoma studies. Given the immunosuppressive tumor environment, one must question how TILs perform their anticancer role under these hostile conditions. Generally, hypoxia is a common feature of tumors and low oxygen can attenuate immune responses due to the corresponding signaling events and metabolic changes initiated by T cells in hypoxic environments.2 In ovarian cancer, hypoxia has been observed to promote regulatory T cell (Treg) recruitment and alter CD4+ T helper type-17 (Th17) development.3,4 In response, hypoxic tumors increase the production of lactate that promotes the recruitment of myeloid-derived suppressor cells (MDSCs) which have a variety of immunosuppressive properties. For example, a high concentration of lactate has been observed to inhibit cytotoxic T-cell activation and causes a shift toward a Th17/23 cell phenotype.5 Our recent study has shown that T-cell infiltration and antitumor function is dependent upon the degree of tumor vascularization and corresponding oxygenation.6

Using 2 established markers of tissue vascularization and oxygenation, CD31, to mark blood vessel endothelial cells and vascular endothelial growth factor (VEGF), a hypoxia-inducible gene, we assessed the influence of hypoxia on TILs.7 Our initial analysis revealed that, compared with CD31 negative tumors (hypoxic), CD31 positive tumors (vascularized and normoxic) correlate with better disease-specific patient survival. Albeit counterintuitive, we posited that the increase in survival was related to lymphocyte infiltration into the tumor. When a series of tumor sections from the tissue array were histologically assessed for the presence of T cell markers (CD8, CD4, and FoxP3), we found that the expression of each marker had a strong positive correlation with CD31 expression. Further characterization using cytotoxicity markers (TIA-1 and granzyme B) showed that TILs within vascularized tumors were also activated and functional (Fig. 1).

We next performed a series of in vitro assays using murine OTI T cells to determine the direct effect of hypoxia on T-cell function.8 OVA-specific CD8+ T cells were isolated from OTI mice and stimulated under normoxic or hypoxic conditions with the cognate SIINFEKL peptide. As predicted, cells cultured under low oxygen produced lower levels of the cytotoxicity factors tumor necrosis factor α (TNFα) as well as interferon-γ (IFNγ). The decreased ability of hypoxia-exposed T cells to mount an effective cytotoxic response was confirmed using a cell lysis assay. Under hypoxia, T cells exhibited a lower capacity to kill target cells compared with those cultured in normoxia. In addition, the suppression of T-cell effector functions under hypoxic conditions, oxygen-starved T cells were found to activate autophagy as a survival mechanism. Although a common adaptation for tumor cells under hypoxia, autophagy had thus far been undocumented for hypoxic responses in T cells.

These results suggested that positive outcomes associated with TIL depend upon the oxygenation state of the tumor. Therefore, a series of Kaplan–Meier analyses were performed, comparing survival rates in groups of patients.
stratified according to the presence of each of the vascularity markers. These analyses showed that immune infiltrates led to longer disease-specific survival only when tumors were well vascularized and CD31⁺. Interestingly, in the case of the Treg marker FoxP3, there was a significant difference in these survival rates between patients harboring hypoxic vs. normoxic tumors, suggesting that the presence of T cells expressing FoxP3 are only beneficial to the patient when their tumor is highly vascularized.

Our study supports the concept that modulation of the tumor vasculature may be a viable avenue to improve anticancer immunity (Fig. 1). One such approach aims to stimulate T-cell adhesion to the epithelial cells within the tumor vasculature, and subsequent T-cell tumor infiltration. Considering that vascular growth factors such as VEGF can inhibit adhesion ligand expression in a state known as epithelial cell anergy (thus suppressing T-cell extravasation and tumor infiltration) increasing oxygenation to prevent such signals could be beneficial. A specific therapy where this approach is being pursued is the tumor vasculature targeting agent NGR-TNF in which the inflammatory cytokine TNFα is fused with a short peptide sequence targeting tumor-associated blood vessels. TNFα activates epithelial cells leading to adhesion molecule expression and immune cell infiltration. If this technique were applied in tumors undergoing angiogenesis, perhaps the infiltration suppression issue could be surpassed.

Because of the altered intratumoral T-cell activity, the hypoxic tumor environment may limit clinical strategies aiming to induce beneficial immune responses. This suggests that novel approaches altering the hypoxic response or the metabolism of T cells could counter the immunosuppressive effects of hypoxia. Overall, these findings demonstrate that the CD8⁺ T cell response is a crucial component of antitumor immunity and their presence within the malignant lesion requires adequate oxygenation in order for TILs to beneficially impact patient outcomes.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.
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