Acquired EGFR C797G Mutation Detected by Liquid Biopsy as Resistance Mechanism After Treatment With Osimertinib: A Case Report

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Abstract. Background: Osimertinib is a third-generation EGFR-tyrosine kinase inhibitor approved for the treatment of T790M-positive non-small-cell lung cancer. More recently, osimertinib demonstrated improved disease control compared to other EGFR-TKIs. Multiple mechanisms of resistance have been described in T790M-positive patients who experienced treatment failure with osimertinib. Case Report: We report the case of a 78-year-old non-smoker woman with stage IV EGFR L858R-positive lung adenocarcinoma presented with T790M mutation after five years of treatment with gefitinib. The patient was started on osimertinib, but after two and a half years of treatment experienced disease progression. The analyses of circulating tumor DNA using next-generation sequencing showed, together with the pre-existing T790M and exon 21 L858R, the presence of the EGFR C797G resistance mutation. Conclusion: Our case report revealed a rare EGFR-dependent acquired resistance mutation to osimertinib in circulating tumor DNA. Liquid biopsy appears to be a promising resource to understand the biology of osimertinib resistance by clonal evolution monitoring and the identification of novel resistance mechanisms.

Osimertinib is a third-generation EGFR-tyrosine kinase inhibitor (TKI) with proven activity against T790M-positive EGFR mutant non-small-cell lung cancer (NSCLC) patients resistant to first- and second-generation EGFR-TKIs such as gefitinib, erlotinib, and afatinib (1).

More recently, based on the results of the FLAURA phase III trial, osimertinib demonstrated significantly longer progression-free survival and overall survival even when used as first-line treatment, compared to first-generation TKIs (2-3). The osimertinib safety profile was similar to that of the comparator EGFR-TKIs (2-3). However, disease progression eventually occurs during osimertinib therapy and resistance mechanisms may be identified through tissue or liquid biopsy (4). Amplification of MET, HER2, PIK3CA, secondary EGFR mutations (C797S and others), and histologic transformation are the most common resistance mechanisms detected (5, 6).

In the current report, we describe the case of a patient with an emerging rare EGFR C797G mutation during osimertinib treatment detected by liquid biopsy. Written informed consent was obtained from the patient for publication of this case report and any accompanying images.

Case Report

Acquisition of a rare mutation in EGFR Exon 20 (p. C797G, c. 2389T>G) was detected in a 78-year-old non-smoker female after the administration of third-generation EGFR-TKI osimertinib. In August 2007, the patient underwent left upper lobectomy with diagnosis of EGFR L858R-positive adenocarcinoma, pT2N1M0, followed by adjuvant chemotherapy with four cycles of cisplatin and vinorelbine. The radiological follow-up was performed with positron emission tomography–computed tomography (PET/CT).
Figure 1. Positron emission tomography scans of liver metastasis at different time points. (A) Liver progression on gefitinib treatment; (B) Liver localizations before osimertinib treatment: a liver biopsy revealed the presence of EGFR exon 21 L858R mutation and the acquisition of T790M mutation; (C) Partial response during osimertinib treatment; (D) Disease progression on osimertinib: a liquid biopsy showed an acquired EGFR C797G mutation together with the pre-existing T790M mutation and exon 21 L858R mutation; (E) Rapidly progressive disease few weeks before death.
because of hypersensitivity reaction to iodinated contrast medium. In July 2011, the patient presented with cough and dyspnea and a PET/CT demonstrated bilateral lung lesions with mediastinal involvement. Bone scan and brain magnetic resonance imaging (MRI) showed no abnormalities. A bronchoscopy with lung biopsy was performed, with diagnosis of EGFR L858R-positive adenocarcinoma (Exon 21 p.L858R, c.2573T>G). The patient received a first-line treatment with gefitinib, 250 mg per day, with partial remission for five years. In May 2016, a PET/CT revealed new lesions in the liver and right lung (Figure 1). Since the identification of disease progression after first-line gefitinib treatment, the patient was followed-up with liquid biopsy. Subsequent monitoring of circulating tumor DNA (ctDNA) was conducted using amplicon-based ultra deep-next-generation sequencing (NGS) on Illumina platform to detect a broader range of mutations.

A liquid biopsy did not reveal a T790M resistance mutation in ctDNA. Unfortunately, it was not possible to perform a tissue confirmatory biopsy because of patient’s refusal. Therefore, the patient received chemotherapy with carboplatin and pemetrexed for four cycles, until treatment discontinuation due to nephrotoxicity. In May 2017, the patient experienced shortness of breath and abdominal pain accompanied by radiological progression documented by PET imaging (Figure 1). A subsequent liver biopsy revealed the presence of exon 21 L858R mutation and the acquisition of Exon 20 T790M mutation (p.T790M, c.2369C>T), also confirmed by liquid biopsy. The patient was then put on osimertinib, 80 mg daily, until PET progression in November 2019 (Figure 1). A liquid biopsy showed an acquired EGFR C797G mutation at a mutant allele frequency (MAF) of 1.3%, together with the pre-existing T790M mutation (MAF: 1.3%) and exon 21 L858R mutation (MAF: 1.3%) (Figure 2). No activating mutations in other oncogene drivers (such as BRAF, MET, KRAS) were detected. Unfortunately, the patient never started another treatment due to a rapid general progressive failure that led to her death in February 2020.

Discussion

Approximately 50-60% of patients with EGFR-mutant lung cancer treated with first- and second-generation EGFR-TKIs will develop resistance via T790M mutation. Osimertinib is able to overcome this resistance restoring disease control with high response rates and a median progression-free survival of about 10 months, as observed in our clinical case (1). Unfortunately, when resistance to osimertinib occurs without acquired targetable mutations, chemotherapy represents the last therapeutic option for these patients. Common resistance mechanisms to osimertinib involve secondary EGFR-dependent mutations, which are found in about one-third of cases (4). The most frequent C797S variant is observed in up to 20-25% of patients, which occurs at the covalent binding site for osimertinib (7, 8). Other mutations may occur at G796/C797, L792, or L718/G719 residues (9). Additionally, EGFR-independent pathways may also be activated, with T790M loss and high degree of clonal heterogeneity, mainly involving MET, EGFR, KRAS amplification and activating mutations in PIK3CA and KRAS (10).

Most data on acquired resistance mechanisms are based on analyses in the second-line setting. Emerging data from both plasma genotyping and from tissue analyses reported differences in frequency of resistance mechanisms to osimertinib given in a front-line versus second-line setting. Patients receiving later-line osimertinib developed more frequently EGFR-dependent resistance compared to patients receiving initial osimertinib (11-13).

Our report describes a rare mutation located at the residue 797 of EGFR, C797G, as a mechanism of acquired resistance to osimertinib. Our patient did maintain the original mutations L858R and T790M, so finally we detected three mutations in the ctDNA. Few reports have identified this mutation from tissue biopsy of NSCLC patients who experienced disease progression on osimertinib, especially in the Asian population (14-16).

A secondary analysis of the AURA3 trial reported the ctDNA genomic profile of T790M-positive NSCLC patients who experienced disease progression during osimertinib treatment. Interestingly, in only one patient among the 73 included in the analysis an EGFR C797G resistance mutation was found by NGS analyzing ctDNA isolated from plasma samples (5).

Our case report underlined the pivotal role of liquid biopsy in establishing the molecular evolution of osimertinib resistance, similarly to other experiences (17, 18). Specifically, liquid biopsy, a non-invasive technique, is enabling physicians to monitor targeted treatment response, predicting clinical-radiological progression and the detection of heterogeneous resistance mutations, in order to administer precision-treatment regimens (19-21). Nevertheless, tissue biopsy should be kept into consideration in cases of suspected histologic transformation or non-informative liquid biopsy result.

Figure 2. Mutant allele frequency (MAF) % detected on EGFR exons 20 and 21 in the plasma ctDNA by ultra deep NGS. Media coverage of all analyzed amplicons is indicated as Total reads. MAF: Mutant allelic fractions; ctDNA: circulating tumor DNA; NGS: next-generation sequencing.
Further studies are warranted to investigate the heterogeneity of osimertinib resistance mechanisms, especially using osimertinib as upfront treatment, to guide therapeutic selection to overcome acquired resistance.

Conflicts of Interest

The Authors declare no conflicts of interest in relation to this study.

Authors’ Contributions

Alessandra Bearz, Elisa De Carlo, Giacomo Pelizzari and Monica Schiappacassi: Conception and preparation of the manuscript, data analysis and interpretation. Tania Baresic, Alessandro Del Conte, Brigida Stanzione and Valentina Da Ros: provision of study materials and data collection. Roberto Doliana and Gustavo Baldassare: provision of study materials, data analysis and interpretation. All Authors contributed and agreed with the content of the manuscript.

Acknowledgements

This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

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Received May 22, 2021
Revised June 17, 2021
Accepted June 18, 2021