Nitric oxide (NO) is a short-lived free radical produced endogenously in biological tissues by nitric oxide synthases (NOSs)[1, 2]. Three NOS isoforms, namely NOS1 or neuronal NOS (nNOS), NOS2 or inducible NOS (iNOS), and NOS3 or endothelial NOS (eNOS) are present in most cell types, including cardiac myocytes and vascular endothelial cells. Vascular relaxation to mediators such as acetylcholine or increased blood flow depends on NO produced by the eNOS. The discovery of NO as the endothelium-derived relaxing factor (EDRF) and its crucial function as a signaling molecule in cardiovascular system was awarded the Nobel Prize in Physiology or Medicine in 1998[1].

Under normal circumstances the main function of eNOS is to produce NO. It catalyzes the conversion of L-arginine (L-Arg) to L-citrulline and NO via electron transfer from the reduced form of nicotinamide adenine dinucleotide phosphate (NADPH) through a flavin containing reductase domain to oxygen bound at the heme of an oxygenase domain containing tetrahydrobiopterin (BH$_4$) and L-Arg binding sites (Figure 1). The eNOS-derived NO activates the guanylate cyclase/cGMP/protein kinase G (PKG) pathway and modulates protein properties and function through nitrosylation of tyrosine and thiol-groups of cysteine in proteins, which are usually effective protection mechanisms against oxidative stress.

Under pathophysiological conditions such as hypertension, diabetes, septic shock and atherosclerosis, oxidative stress alters many functions of the endothelium and leads to endothelial dysfunction when the endothelium fails to serve its normal physiologic and protective mechanisms. A common feature of endothelial dysfunction is the reduced bioavailability of NO and increased production of superoxide (·O$_2$–) and other...
reactive oxygen species (ROS) in the vasculature. Multiple mechanisms may underlie the impaired NO availability. These include a reduction in the expression level of eNOS mRNA or protein, changes in subcellular compartmentalization of eNOS activity, and compromised availability of the substrates and/or enzymatic cofactors for eNOS. Depletion of the substrate L-arginine, accumulation of methylnitrogens, and oxidation of the cofactor BH₄ of eNOS can uncouple the electron transfer reactions and revert eNOS to function as an NADPH oxidase, thus producing ·O₂⁻ instead of NO (Figure 1). The rapid reaction of NO with ·O₂⁻ can form the most potent oxidant peroxynitrite anion (ONOO⁻) and causes cellular injury associated with many pathophysiologic conditions, such as hypertension, atherosclerosis, diabetes, myocardial hypertrophy, heart failure, and ischemia/reperfusion injury. The precise molecular mechanisms underlying the "switch" of the eNOS function from NO synthesis to ·O₂⁻ production under oxidative stress conditions, however, are still not fully understood.

Recently, Chen et al reported that S-glutathionylation of eNOS may be a unique mechanism for the redox regulation of eNOS. It has been demonstrated previously that cysteine residues are critical for the maintenance of normal eNOS function. Protein S-glutathionylation has been known as a specific post-translational modification of protein cysteine residues by adding the tripeptide glutathione through reversible thiol-disulfide exchange with oxidized glutathione (GSSG) or reaction of oxidant-induced protein thiyl radicals with reduced glutathione (GSH). Under oxidative stress, therefore, protein S-glutathionylation can serve to prevent irreversible oxidation of protein thiols. S-glutathionylation has now emerged as a potential mechanism for dynamic, post-translational regulation of a variety of regulatory, structural, and metabolic proteins. Increasing lines of evidence point to the important role of S-glutathionylation in the regulation of signaling and metabolic pathways in intact cellular systems. Indeed, Chen et al found that GSSG induced dose-dependent S-glutathionylation of human eNOS that was reversed by reducing agents β-mercaptoethanol or dithiothreitol. S-glutathionylation of eNOS reversibly decreases NO activity with a concomitant increase in ·O₂⁻ generation primarily from the reductase domain. Two highly conserved cysteine residues are identified as sites of S-glutathionylation and found to be critical for redox-regulation of eNOS function. They further demonstrated that S-glutathionylation of eNOS in endothelial cells turned off NO synthesis and turned on ·O₂⁻ generation (Figure 1). This conversion of eNOS function by S-glutathionylation is closely associated with impaired endothelium-dependent vasodilation. In hypertensive vessels, S-glutathionylation of eNOS is increased with impaired endothelium-dependent vasodilation. Thiophen Anthony et al reported that the switch of eNOS function is reversible and can be reverted by reducing agents.

In summary, S-glutathionylation has now emerged as a potential mechanism for dynamic, post-translational regulation of a variety of regulatory, structural, and metabolic proteins. Increasing lines of evidence point to the important role of S-glutathionylation in the regulation of signaling and metabolic pathways in intact cellular systems. Indeed, Chen et al found that GSSG induced dose-dependent S-glutathionylation of human eNOS that was reversed by reducing agents β-mercaptoethanol or dithiothreitol. S-glutathionylation of eNOS reversibly decreases NO activity with a concomitant increase in ·O₂⁻ generation primarily from the reductase domain. S-glutathionylation of eNOS reversibly decreases NO activity with a concomitant increase in ·O₂⁻ generation primarily from the reductase domain. Two highly conserved cysteine residues are identified as sites of S-glutathionylation and found to be critical for redox-regulation of eNOS function. They further demonstrated that S-glutathionylation of eNOS in endothelial cells turned off NO synthesis and turned on ·O₂⁻ generation (Figure 1). This conversion of eNOS function by S-glutathionylation is closely associated with impaired endothelium-dependent vasodilation. In hypertensive vessels, S-glutathionylation of eNOS is increased with impaired endothelium-dependent vasodilation. Thiophen Anthony et al reported that the switch of eNOS function is reversible and can be reverted by reducing agents.

Research Highlight

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