Relationship between free fatty acids, insulin resistance markers, and oxidized lipoproteins in myocardial infarction and acute left ventricular failure

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Background: The most common cause of myocardial infarction (MI) is stenotic atherosclerotic lesions in subepicardial coronary arteries. Artery disease progression induces clinical signs and symptoms, among which MI is the leader in mortality and morbidity. Recent studies have been trying to find new biochemical markers that could predict the evolution of clinical complications; among those markers, free fatty acids (FFA) and oxidative modification of low-density lipoproteins (oxidized LDL) have a special place.

Materials and methods: Seventy-nine ST-elevation MI patients were enrolled. The first group included MI patients without the signs of acute heart failure (Killip class I) while MI patients with Killip classes II–IV made up the second group. Thirty-three individuals with no cardiovascular disease were the controls. The lipid profile, serum oxidized LDL, and their antibodies, C-peptide and insulin were measured at days 1 and 12. The level of insulin resistance was assessed with the quantitative insulin sensitivity check index (QUICKI).

Results: MI patients had atherogenic dyslipidemia; however, the Killip II–IV group had the most pronounced and prolonged increase in FFA, oxidized LDL, and their antibodies. Additionally, positive correlations between FFA levels and creatine kinase activity (12 days, $R = 0.301; P = 0.001$) and negative correlations between the QUICKI index and FFA levels ($R = -0.46; P = 0.0013$ and $R = -0.5; P = 0.01$) were observed in the both groups.

Conclusion: The development of MI complications is accompanied by a significant increase in FFA levels, which not only demonstrate myocardial injury, but also take part in development of insulin resistance. Measuring FFA levels can have a great prognostic potential for risk stratification of both acute and recurrent coronary events and choice of treatment strategy.

Keywords: myocardial infarction, free fatty acids, insulin resistance

Introduction

Nowadays, ischemic heart disease (IHD) remains one of the most relevant and unaddressed issues of contemporary medicine. Moreover, in all its manifestations, IHD is the first leading cause of mortality worldwide. Coronary atherosclerosis underlying IHD usually occurs and progresses silently, so CHD symptoms occur in the later stages of the disease when full recovery cannot be achieved. Recent studies have been trying to detect new biochemical markers that could show the evolution of clinical atherosclerosis complications; among them free fatty acids (FFA) have a special place.
Cell FFA carry out some key functions, including adenosine triphosphate (ATP) production. They act as cell-signal mediators (activation of various protein kinase C isoforms, apoptosis initiation), ligand transcription factors, and basic components of biological membranes.\(^2\) FFA oxidation provides up to 70% of ATP to the heart; the other energy needs are satisfied by glucose oxidation. The intensity of FFA uptake by myocardial cells is defined, first of all, by their plasma concentrations.

Excess products and metabolites of fatty acid oxidation (acetyl-coenzyme A, reduced NADPH, FAD\(_2\)) are natural inhibitors of pyruvate dehydrogenase complex enzymes, responsible for aerobic glucose oxidation, which leads to a decreased glucose myocardial utilization.\(^3\) In ischemia, the main metabolic pathway providing energy to cardiomyocytes is anaerobic glycolysis because fatty acid oxidation requires more oxygen and altered myocardial utilization of fatty acids due to myocardial ischemia and necrosis results in fatty acid build-up in the blood.\(^4\) Some researchers think that increased FFA concentrations are the earliest ischemia predictor, and more sensitive to the degree of ischemia than electrocardiography.\(^5\) The prospective trial results of the Québec Cardiovascular Study show a strong relationship between increased plasma FFA concentrations and the risk of IHD development.\(^1\) Moreover, according to Opie and Knuuti, FFA can be regarded as a potential biochemical marker of postinfarct myocardial remodeling.\(^6\)

Increased blood FFA concentrations are known to be strongly related to insulin resistance (IR), which is a metabolic risk factor for development of arterial diseases.\(^7\) Recent publication highlighted the association between IR, atherosclerotic signs, and cardiovascular risk both in women and in men.\(^8\) One of the suggested pathophysiologic mechanisms of such a phenomenon is the extensive production of activated oxygen metabolites in endothelial cells due to high levels of FFA and IR, which are able to induce oxidative modification of low-density lipoproteins (oxidized LDL) and synthesis of their antibodies, inducing atherosclerotic changes in the vessel wall, which eventually causes myocardial ischemia.\(^9,10\)

A number of researchers suggested that FFA play an important role in atherosclerosis manifestation. Monitoring blood FFA levels in acute coronary events can play an important role in diagnostics and has prognostic value for choosing a strategy from a perspective of risk assessment in this category of patients.

This study was aimed at examining the dynamics of FFA levels and their relationship with IR markers and oxidized LDL and their antibodies in patients with acute myocardial infarction (MI).

**Material and methods**

**Study subjects and design**

Seventy-nine patients (50 males and 29 females, aged 63.8 ± 1.69 years) with ST elevation myocardial infarction (STEMI) were examined. The study design complied with the tenets of the Declaration of Helsinki and was approved by the Institutional Review Board of the Research Institute for Complex Issues of Cardiovascular Diseases, and all the patients signed the informed consent form before taking part in the study. MI was diagnosed according to the 2007 guidelines of the Russian National Cardiology Society. The inclusion criteria were chest pain (with no effect of nitroglycerin), signs of ischemia, ST segment elevation on electrocardiography and increased levels of cardiac enzymes: serum creatine kinase MB fraction (CK-MB) and troponin T. The patients were divided into two groups. Group I included 47 MI patients with no clinical signs of acute heart failure (Killip I) and group II enrolled 32 MI patients with acute heart failure (Killip II–IV) developed at day 1 from the onset of the disease. Coronary angiography showed that 58 patients (84.1%) had coronary lesions with >50% stenosis. The exclusion criteria were type 2 diabetes mellitus and laboratory signs of inflammation. The end diastolic (EDV, mL) and end systolic volumes (ESV, mL) reflecting structural and functional myocardial remodeling, which developed during MI, were 179.96 ± 11.04 mL and 157.19 ± 3.91 mL in Group I; and 78.52 ± 3.45 and 106.57 ± 10.46 in Group II, respectively. The echocardiography parameters were measured with the Aloka 5500 (SSD-5500 ProSound PHD; Hitachi Aloka Medical Ltd, Tokyo, Japan) in one- and two-dimensional modes with an ultrasound frequency of 3.5 MHz, according to a standard technique. Thirty-three age and sex-matched subjects without any cardiovascular disease were the control group.

**Assays**

Blood serum and plasma were tested. The tests were done at day 1 from MI and acute heart failure onset and at day 12 after the condition had been stabilized. Serum FFA, total cholesterol (TC), triglycerides (TG), LDL, very-low-density lipoproteins (VLDL), apolipoprotein B (apo B) as well as high-density lipoproteins (HDL) and apolipoprotein A (apo A) concentrations were measured with the standard Thermo Fisher Scientific test systems (Thermo Fisher Scientific Oy, Vantaa, Finland) on a Konelab 30i biochemistry analyzer (Thermo Fisher Scientific Oy). Serum oxidized LDL, their antibodies,
C-peptide and insulin concentrations were measured by ELISA with BioMedica (Waterloo, NSW, Australia) and Diagnostic Systems Laboratories (Webster, TX, USA) lab kits respectively. A structured mathematical model was used to assess IR level based on fasting insulin, glucose values and calculating the quantitative insulin sensitivity check index (QUICKI) values:

\[
\text{QUICKI} = \frac{1}{\log(I_h) + \log(G_w)}
\]

where \( I_h \) was the baseline glycemia (mg/dL) and \( G_w \) was the baseline insulinemia (mMU/mL). According to Katz et al, the mean QUICKI index of 0.382 ± 0.007 correlates with normal tissue sensitivity to insulin while the QUICKI indexes of 0.331 ± 0.010 and 0.304 ± 0.007 correlate with moderate and intensive IR.  

Statistical analysis
The results were statistically processed using nonparametric criteria; differences were considered to be statistically significant if \( P < 0.05 \). To determine the association between variables, Spearman’s correlation coefficient was calculated.

Results
Both groups had atherogenic dyslipidemia with higher TC, TG, LDL, VLDL, and apoB concentrations, greater apoB/apoA coefficient, and lower antiatherogenic HDL and apoA in healthy subjects than in the control group. (Table 1). There were no statistically significant differences in the lipid profiles of Killip I and II–IV patients.

FFA concentrations significantly differed in MI patients and healthy subjects as well as at different stages of the disease (Table 2). Indeed, at day 1 MI patients both with and without acute heart failure had, on average, sevenfold increased FFA levels than those in the control group. By day 12, FFA levels decreased, but were still 3.0 and 4.5 times (complicated and noncomplicated MI) higher than those in healthy subjects (Table 2).

Both groups had a significant rise in glucose, insulin, and C-peptide concentrations in the acute stage of the disease compared to the control group. Meanwhile, Killip I MI patients still had a tendency towards increased concentrations of the above-mentioned parameters up to day 12. On the contrary, Killip II–IV patients had much more different concentrations of these parameters, but by day 12 insulin and C-peptide levels decreased significantly, even lower than in healthy subjects (Table 3), with glucose levels being consistently high. The QUICKI index in both groups significantly differed from that in the controls; in the Killip I group it correlated with moderate IR (according to Katz et al\textsuperscript{11}) and in the Killip II–IV group it correlated with intensive IR (Table 3). When the patients’ condition stabilized, the parameters under study did not change significantly.

The correlation analysis showed a positive correlation between FFA and CK-MB activity at day 12, which demonstrated the size of myocardial necrosis (\( R = 0.301; P = 0.001 \)) (Figure 1). Besides, at day 1, the Killip II–IV group was found to have positive correlations between FFA levels and EDV (\( R = 0.34; P = 0.01 \)) (Figure 2), which proved a strong association between increased FFA and postinfarct myocardial remodeling. A negative correlation between the QUICKI index and FFA levels (\( R = −0.31; P = 0.0067 \)) (Figures 3 and 4) was found in both groups.

Oxidized LDL concentrations in MI patients changed according to the Killip class (Table 4). Both groups had higher oxidized LDL levels compared to those in the control group. Moreover, Killip II–IV patients had significantly higher oxidized LDL levels already at day 1 compared to those in Killip I patients. At day 12, both groups had a tendency toward increased oxidized LDL concentrations.

Both MI patients with and without acute heart failure were found to have positive correlations between oxidized LDL and FFA concentrations (\( R = 0.49; P = 0.0009 \)) (Figure 5), as well as negative correlations between the QUICKI index and oxidized LDL concentrations (\( R = −0.64; P = 0.0002 \)) Killip II–IV (Figure 6). There was an increase in oxidized LDL antibodies in both groups compared to healthy subjects.

### Table 1 Variables of blood lipid-transport function in patients with myocardial infarction on the first day of the disease

| Variables       | Control (n = 33) | Killip I (n = 47) | Killip II–IV (n = 32) |
|-----------------|-----------------|------------------|-----------------------|
| TC, mmol/L      | 5.02 (4.8–5.52) | 5.95 (4.7–6.9)\( ^a \) | 6.1 (6.0–7.4)\( ^a \) |
| TG, mmol/L      | 1.01 (0.9–1.2)  | 1.62 (1.027–2.35)\( ^a \) | 1.99 (1.28–3.2)\( ^a \) |
| HDL-C, mmol/L   | 1.1 (0.9–1.4)   | 1.04 (0.95–1.23)\( ^a \) | 1.005 (0.8–1.1)\( ^a \) |
| LDL-C, mmol/L   | 2.5 (2.1–3.0)   | 3.17 (2.32–3.8)\( ^a \) | 3.5 (3.02–3.77)\( ^a \) |
| VLDL-C, mmol/L  | 0.5 (0.35–0.61) | 0.73 (0.57–1.06)\( ^a \) | 0.9 (0.58–1.45)\( ^a \) |
| Apo B, g/L      | 0.9 (0.81–1.23) | 1.24 (0.87–1.35)\( ^a \) | 1.285 (1.12–1.5)\( ^a \) |
| Apo A1, g/L     | 1.2 (1.1–1.36)  | 1.25 (1.23–1.46)\( ^a \) | 1.18 (1.15–1.37) |
| Apo B/apo A1    | 0.6 (0.55–0.69) | 0.84 (0.64–0.99)\( ^a \) | 1.15 (0.89–1.188)\( ^a \) |

Note: \( ^a \) Compared with control group (\( P < 0.05 \)).

Abbreviations: TC, total cholesterol; TG, triglycerides; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; VLDL-C, very-low-density lipoprotein cholesterol; Apo B, apolipoprotein B; Apo A1, apolipoprotein A1.
| Variables       | Control (n = 33) | Killip I (n = 47) | Killip II–IV (n = 32) |
|-----------------|------------------|-------------------|-----------------------|
| Glucose, mmol/L | 5.1 (4.8–5.5)    | 6.9 (6.1–8.4)     | 7.6 (6.2–10.5)        |
| Insulin, mU/mL  | 12.4 (11.4–14.3) | 13.85 (8.3–18.3)  | 14.5 (6.1–19.5)       |
| C-peptide, ng/mL| 1.01 (0.89–1.23) | 0.93 (0.52–1.01)  | 0.35 (3.1–4.54)       |
| QUICKI          | 0.37 (0.34–0.4)  | 0.3 (0.28–0.33)   | 0.29 (0.28–0.32)      |

Notes: *Compared with the control group (P < 0.05); †reliable differences in indicators on day 1 and day 12 (P < 0.05); ‡reliable differences in indicators for Killip I and Killip II–IV groups (P < 0.05).

Abbreviation: QUICKI, quantitative insulin sensitivity check index.
ions, activating phospholipase A₂ with further membrane injury by lipid peroxidation products. If there is no early reperfusion, at first reversible metabolic changes inevitably become irreversible and result in cell death. Obviously, if MI or its complications develop, FFA increase does not only reflect myocardial ischemia, but may also act as a marker of the extent of myocardial necrosis. The results of the correlation analysis conducted in this study support this suggestion. Indeed, observed positive correlations between FFA levels at day 1 from the MI onset and CK-MB activity, showing the size of myocardial necrosis, as well as between FFA levels, ESV and EDV evidence show strong association between FFA and postinfarct myocardial remodeling.

It is suggested that FFA build-up in the blood during MI development can also be the cause of IR, which leads to a range of metabolic changes, contributing to develop-
ment of MI complications. Hepatocytes intensively produce atherogenic lipoproteins slowing down antiatherogenic HDL synthesis. Besides, hepatocytes collect lipid metabolism by-products (ceramides, diacylglycerol), changing the transport of glucose into the cells and, consequently, increasing its concentrations in the blood. Hepatic IR leads to decrease in glycogen production, glycogenolysis and gluconeogenesis activation, which together worsen hyperglycemia being, according to clinical studies, an independent risk factor for inhospital complications, adverse outcome, and mortality.7

We found general common patterns in the dynamics of IR and some differences in their changes depending on the presence or absence of MI complications (Table 3). Indeed, this study demonstrated a significant increase in glucose, insulin,
and C-peptide concentrations in both groups in the acute stage of the disease compared to those in the control group. The presence of IR in patients enrolled in the study was further confirmed by a low QUICKI index. An increase in the QUICKI index on day 12 in the complicated MI patients seems to be due to the decreased blood insulin. Apparently, FFA reduction results in a better insulin clearance, reduced hyperinsulinemia, and increase in QUICKI index.\textsuperscript{14}

Experimental IR, due to excess FFA, alters normal function of mitochondrial enzymes engaged in tissue respiration, which can potentially lead to reactive oxygen species production, result in lipoprotein oxidative modification, induce inflammation in vascular endothelium, and promote atherosclerotic plaque and ischemia progression.\textsuperscript{14} Our study showed that oxidized lipoprotein levels increased both in the acute stage of the disease and during the whole hospital stay. In order to confirm the association between IR and atherogenic oxidized LDL presence in the blood, a correlation analysis was conducted. It demonstrated positive correlations between oxidized LDL, FFA concentrations as well as a negative correlation between the QUICKI index and oxidized LDL levels, which confirmed the presence of a pathogenetic relationship between the parameters under study.

LDL oxidation is not only accompanied by changes in their chemical properties, but also leads to immunocompetent cell activation, producing antibodies to a ‘hostile’ agent, and oxidized LDL elimination from the blood flow.

Antibodies to oxidized LDLs can be also found in healthy subjects; however, their concentrations are significantly increased in patients with other cardiovascular diseases.\textsuperscript{15} This can be explained by the potential pathogenicity of these antibodies; increased levels of oxidized LDL and their antibodies form immune complexes and, binding with vascular intima, have additional damaging effects on the endothelium.\textsuperscript{16} In this study, we found increased

\begin{figure}[h]
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\includegraphics[width=\textwidth]{figure5.png}
\caption{Correlations between oxidized LDL and FFA concentrations at day 12 for the Killip II–IV group.\textbf{Abbreviations:} FFA, free fatty acids; LDL, low-density lipoprotein.}
\end{figure}

\begin{table}[h]
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\begin{tabular}{lccccc}
\hline
\textbf{Variables} & \textbf{Control (n = 33)} & \textbf{Killip I (n = 47)} & \textbf{Killip II–IV (n = 32)} \\
\hline
\multicolumn{2}{c}{\textbf{Day 1}} & \textbf{Day 12} & \textbf{Day 1} & \textbf{Day 12} \\
\hline
Oxidized LDL, ng/mL & 345.6 (299.2–405.5)\textsuperscript{a} & 470.04 (420–563.1)\textsuperscript{a} & 629.2 (589.6–690.9)\textsuperscript{b} & 712.71 (658.2–801.1)\textsuperscript{b} \\
oLABIgG, mU/mL & 201.82 (150.2–251.3) & 308.87 (255.1–372.1)\textsuperscript{a} & 324.94 (290.9–380.1) & 345.99 (301.9–402.1) \\
\hline
\end{tabular}
\textbf{Notes:} \textsuperscript{a}Compared with the control group (P < 0.05); \textsuperscript{b}reliable differences in indicators on day 1 and day 12 (P < 0.05); \textsuperscript{c}reliable differences in indicators for Killip I and Killip II–IV groups (P < 0.05).
\textbf{Abbreviations:} Oxidized LDL, oxidative modification of low-density lipoproteins; oLABIgG, oxidative modification of antibodies of low-density lipoproteins.
levels of oxidized LDL antibodies; moreover, patients with complicated MI had higher levels of these antibodies during the whole hospital stay.

The results obtained in this study correlate with the data of previous research, which demonstrated that increased levels of oxidized LDL antibodies did not only reflect the presence of IHD and were a marker of plaque instability, but also acted as a precursor of acute MI. Besides, based on the obtained data, measuring oxidized LDL antibody levels can help predict MI complications.

**Conclusion**

The results of this study show that MI and its clinical complications are accompanied by a significant increase in the level of FFA, which do not only reflect myocardial injury, but also take part in IR development and are a confounding factor for oxidized LDL. Furthermore, their antibodies build up in the blood flow, which has a powerful atherogenic potential.

Measurement of FFA levels can have a great prognostic value for risk stratification of both acute and recurrent coronary events and further choice of treatment strategy. High levels of FFA in MI, which still persist when the patients' condition has been stabilized, may theoretically be considered a prerequisite for the use of this marker as one of the criteria to assess the risk of complications.

**Disclosure**

The authors report no conflicts of interest in this work.

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