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The genome of *Apis mellifera*: dialog between linkage mapping and sequence assembly

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Abstract

Two independent genome projects for the honey bee, a microsatellite linkage map and a genome sequence assembly, interactively produced an almost complete organization of the euchromatic genome. Assembly 4.0 now includes 626 scaffolds that were ordered and oriented into chromosomes according to the framework provided by the third-generation linkage map (AmelMap3). Each construct was used to control the quality of the other. The co-linearity of markers in the sequence and the map is almost perfect and argues in favor of the high quality of both.

Most eukaryotic genome sequencing projects are preceded by the construction of physical, genetic and/or cytological maps. For the honey bee genome project there was no physical map, and because of the low resolution of the cytogenetic map, the meiotic map was the only resource for organizing the sequence assembly on the chromosomes. The first generation map AmelMap1 comprised 541 markers on 24 linkage groups for 16 chromosomes [1,2]. Saturation was achieved by addition of 601 markers prepared from cDNAs [3] and bacterial artificial chromosomes (BACs) [4] sequences. AmelMap2 was not published, but was used by the Human Genome Sequencing Center at Baylor College for the first assembly of the *Apis mellifera* genome in January 2004. From that time a dialog was set up between the map and sequence projects that became interactive, each taking advantage of the progress of the other. The density of the third-generation map, AmelMap3, was doubled and contributed greatly to the ultimate assembly (version 4.0, March 2006) of the honey bee genome [5].

AmelMap3 comprises 2,008 microsatellite markers (see Additional data file 1) and is 4,000 cM long (M.S, F.M, D.V M.M and J-M.C, unpublished work). Improvements in the map between the second and third generation resulted exclusively from addition of markers designed from the sequence: 587 from previously placed scaffolds in assemblies 1.1 and 2.0 to reduce long genetic distances, orient scaffolds and homogenize the marker density along and among chromosomes and 436 in 379 large unplaced scaffolds (GroupUn) which efficiently increased the fraction of the sequence integrated in chromosomes in the later assemblies (Tables 1 and 2). Chromosomes were oriented by half-tetrad analysis [6]. This orientation was later confirmed by positioning telomeric regions [7] and cytogenetic analysis [5].

Great care was taken to eradicate errors in the final versions (AmelMap3, assembly 4.0). For single markers with uncertain chromosomal positions, new markers were designed; in three cases, the scaffold moved and in two cases the marker did not amplify the expected product. In three cases, two blocks of markers on the same scaffolds mapped to two different positions; adding
markers narrowed the region responsible for the chimerism in which the assembly had to be split. Most of the remaining discrepancies were local marker misordering, eradicated by correction of genotyping errors detected by double crossovers.

A few trivial differences persist between the latest versions of the map and the assembly. Sixteen small scaffolds were reversed and the order of eight groups of short scaffolds will also be revisited. This is attributable to the fact that the last map improvements occurred after the freeze of the version 4.0 assembly. Four unresolved discrepancies remain: the map positions of two short scaffolds (1.43 and 3.37), orientation of a long scaffold (10.30) and remnants in a false position of the break of scaffold 6.37. This generally excellent co-linearity pleads in favor of the quality of the two constructions. If some mistakes remain within scaffolds, they should be below the level of resolution of the map (average 93 kb).

This agreement could seem to be a circular argument as the map is the framework of the assembly. This is not the case. The genetic map and sequence scaffolds have been constructed independently. The maps were calculated with a version of the software Carthage-Gène [8] that does not use physical information and the assembly did not use the map to construct the scaffolds but only to organize them. The eradication of errors in the map, even if it used the sequence to detect them and helped their resolution, was based on genetic methods (controls or addition of genotypes).

Table 1

| Improvements between assembly versions 1.1 (January 2004) and 4.0 (March 2006) |
|---------------------------------|
| Map version | AmelMap2 | AmelMap3 |
| Number of markers | 1,050* | 2,013† |
| Assembly version | 1.1 | 4.0 |
| Total mapped sequence | 110 | 186 |
| Length (Mb) | 53% | 79% |
| Total unmapped sequence (GroupUn) | 96 | 49 |
| Percentage | 47% | 21% |
| Total scaffold length (Mb) | 206 | 235 |
| †After the freeze of assembly 4.0, some markers were added and others removed from the AmelMap3, which now comprises 2,008 markers.

To evaluate the final control of correctness, the scaffolds that contained at least three markers with two non-null genetic distances were selected. The number of markers flanking non-null distances was 1,319 (that is, two-thirds of the total) and they showed only four local and unresolved mistakes (0.3 %). In addition, the 387 markers that are at a null genetic distance within scaffolds are always clustered in the sequence. This accurate co-linearity within scaffolds may be considered indicative of that between scaffolds, which cannot be tested in this way. In the mouse, a very detailed genetic map existed before the sequence of the genome, but of the 12,000 markers, only 2,605 were considered as 'unambiguously' mapped and were used to assess the accuracy of the assembly [9]; most of the conflicts (1.8% of chromosomal misassignment and 0.7% of local misordering) were attributable to mapping errors. For the rat genome, the radiation hybrid map was consistent for 98% of markers with the genetic maps and for 96% with the genome sequence [10].

Among the 626 honey bee scaffolds, 320, representing a physical length of 152 Mb, are oriented (Table 3); the other half were too short to be oriented genetically; they represent only 18.4% of the physical length. Among them, 113 scaffolds forming 44 blocks are not ordered relative to one another (due to null genetic
distances). The unoriented scaffolds are nevertheless placed on chromosomes, but their orientation is random.

Missing sequences in the gaps are probably very short, as suggested by short interscaffold genetic distances. Manual superscaffolding of the five smallest chromosomes (12-16) [11], mainly achieved through relaxing matching criteria, conserved the general structure of the map, included 178 GroupUn scaffolds in the gaps and reduced the 139 scaffolds to 25 super-scaffolds by the addition of only 5.5% of the sequence length. For all chromosome arms, the telomeric regions are reached and the centromeric regions are close to being so [5,7]. Consequently, most of the euchromatic sequence of the chromosome arms is now organized and perhaps only 5% is not included in the assembly.

It may be asked if a genetic map alone provides sufficient information to organize an assembly. The large genetic length of the honey bee genome (about 4,000 cM) compared to its relatively small physical size (about 230 cM) was assuredly a great advantage because it suffices to genotype small families to observe recombination between markers at a short physical distance. The same resolution in organisms with shorter maps (that is, most organisms, if not all [12]), would require a larger genotyping effort in terms of the number of individuals, but it might be limited to a few markers within the largest scaffolds to get a reasonable picture of the genome organization.

Additional data files
Additional data file 1, a list of the primers used for mapping is available with this article online.

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