Long-term outcomes of sentinel node identification using indocyanine green in patients with lung cancer

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Keywords
Indocyanine green; lung cancer; sentinel node.

Abstract
Background: Sentinel node (SN) biopsy is used in the management of numerous cancers to avoid unnecessary lymphadenectomy. This was a clinical exploration/feasibility study of a novel identification technique for SN biopsy using indocyanine green (ICG) fluorescence imaging during lung cancer surgery.

Methods: SN biopsy using ICG was performed on 22 patients who had cT1 or T2N0M0 lung cancer. ICG was injected just around the primary tumor. The fluorescence imaging system enabled visualization of the lymphatic vessels draining from the primary tumor toward the lymph nodes. Fluorescently labeled nodes were dissected, and patients were followed-up for prognosis and recurrence to confirm the pattern of lymph node metastasis after surgery.

Results: SNs were successfully identified in 16 (72.7%) of 22 patients. A total of 13 of 16 patients had pathological N0 and three had SN metastasis. The median follow-up time was 92.7 months. Only one patient had no SN metastasis at the postoperative pathological examination and lymph node metastasis during the follow-up period. The accuracy rate was 93.8% (15/16) and the false-negative rate was 7.7% (1/13).

Conclusions: SNs were identified by ICG fluorescence imaging, and this technique during lung cancer surgery had good identification and accuracy rates throughout the follow-up period.

Key points
Significant findings of the study: We attempted to identify sentinel lymph nodes by indocyanine green in lung cancer surgery. The identification rate was 72.7%. The accuracy rate was 100% immediately after surgery, and 93.8% after follow-up.

What this study adds: Sentinel node biopsy by indocyanine green may be useful for lymph node dissection during lung cancer surgery to avoid unnecessary lymphadenectomy.

Introduction
Due to the widespread use of thin-slice computed tomography (CT), small-sized and/or ground-glass peripheral lung cancers are being increasingly found. In such cases, radical resection can be performed by segmentectomy instead of lobectomy. Indications for segmentectomy require there to be no regional lymph node metastases. Thoracic surgeons have to confirm that there are no lymph node metastases by making an intraoperative rapid pathological diagnosis during the operation. However, it is difficult to determine which lymph nodes to sample for intraoperative rapid pathological diagnosis. Sentinel node (SN) biopsy has been used in the management of numerous cancers to...
determine the lymph vessel flow and avoid unnecessary lymphadenectomy.

The concept of the SN is an old but specific technique first reported by Morton et al. They identified lymph vessels and SNs for melanoma using dye. Krag et al. reported that they used radioisotopes (RI) to identify SNs in breast cancer. SN biopsy techniques have been established for melanoma and breast cancer because they were performed first, but they have yet to be established for lung cancer. Little et al. reported that the identification rate of SN in NSCLC was only 47% by the dye method. Several previous studies reported that it was 6.3% to 50.0%, which is poor. Liptay et al. reported the identification rate of SN in NSCLC to be 82% by the RI method. In other studies, the identification rate of SN biopsy by RI was 62.9%–100%. Although a phase II trial of the RI method was carried out by multiple institutions, the identification rate of SN was 61.5% and the accuracy rate was 51.2%, which were lower than expected.

In recent years, indocyanine green (ICG) fluorescence has become widely applied in surgery. ICG is a popular diagnostic reagent clinically approved for the examination of hepatic and circulatory function. ICG is less toxic, has no radiation exposure, is cost-effective, is excreted quickly, and rarely causes allergic reactions. Several studies have previously investigated SN identification using an ICG fluorescence method in lung cancer, however, they had a short follow-up period after surgery. In this study, we tried to identify the SN by the ICG fluorescence method and followed-up all patients for a long time after surgery to confirm the precise pattern of lymph node metastasis.

**Methods**

**Patients**

The protocol was approved by the Human Ethics Review Committee of Tokushima University Hospital on 21 February 2005 (No. 306) and registered in the Japanese clinical trial registry (UMIN000026155). All patients provided their written informed consent. Eligible patients had clinical T1 or T2 N0M0 NSCLC. Other inclusion criteria were Eastern Cooperative Oncology Group performance status of 0 to 2, absence of previous chemotherapy or radiotherapy, and satisfactory hematological, hepatic, and renal function.

All patients were enrolled and treated at our hospital between June 2007 and March 2009. They underwent chest X-ray, bronchoscopy, chest and abdominal CT, brain magnetic resonance imaging, and/or positron emission tomography-CT preoperatively. SN biopsy using ICG was performed on 22 patients. The characteristics of the patients are shown in Table 1. The mean age was 67.2 years old. A total of 15 were male (68.2%), 12 were smokers (54.5%) and the mean Brinkman index was 691. The tumor size was 27.1 ± 9.7 mm (mean ± standard deviation). All patients had lobectomy or segmentectomy with lymphadenectomy. Our criteria for the operation procedure were as follows: we performed segmentectomy when the tumor was located in the lower lobe or left upper lobe and was less than 2 cm in diameter, and there was no evidence of SN metastasis. Lobectomy was performed in all other cases.

### ICG fluorescence imaging system

Fluorescence images were obtained using the ICG fluorescence imaging system (Olympus, Tokyo, Japan). The light source was a xenon lamp equipped with an excitation filter that permeated light at a wavelength of 690–790 nm, and the detector was a charge-coupled device (CCD) camera equipped to filter out light with a wavelength below 800 nm. The xenon lamp and excitation filter were aligned on the light source equipment, and the CCD camera was placed at the head for thoracoscopy. The fluorescence signals were sent to a digital video processor. The scope had a diameter of 10 mm. This system can be used in video-assisted thoracoscopic surgery (VATS), and can switch between normal white light and infrared light for ICG fluorescence.

### SN identification using ICG

ICG (Diagnogreen; Daiichi Pharm. Co., Tokyo, Japan) at a volume of 0.5–1 mL (5 mg/mL) was injected at 1–4 sites

| Table 1 Patient characteristics |
|--------------------------------|
| **Age (mean)** | 67.2 (53–80) |
| **Gender** |
| Male | 15 (68.2%) |
| Female | 7 (31.8%) |
| **Non-smoker** | 10 (45.5%) |
| **Smoker** | 12 (54.5%) |
| **Brinkman index (mean)** | 691 |
| **Histology** |
| Adenocarcinoma | 16 (72.7%) |
| Squamous cell carcinoma | 5 (22.7%) |
| Large cell carcinoma | 1 (4.5%) |
| **Tumor location** |
| RUL | 7 |
| ML | 1 |
| RLL | 7 |
| LUL | 3 |
| LLL | 4 |
| **Tumor size (mean)** | 27.1 mm |
| **Operation** |
| Segmentectomy | 7 (31.8%) |
| **Lobectomy** | 15 (68.2%) |

LLL, left lower lobe; LUL, left upper lobe; ML, middle lobe; RUL, right upper lobe; RLL, right lower lobe.
around the primary tumor (Fig 1a). As a result, the ICG dose was 2.5–10 mg. ICG was injected into the collapsed lung for thoracoscopic surgery. The injection site was massaged to spread the ICG for approximately five minutes. The ICG fluorescence imaging system clearly visualized the ICG-injected sites (arrow) around the tumor and the spread of the ICG as bright fluorescence. (c) The hilar lymph node #10; and (d) the lymph node were visualized by fluorescence imaging.

Figure 1 (a) The tumor was resected using VATS to confirm lung cancer, as there was no definitive diagnosis preoperatively. During the operative rapid pathological diagnosis, indocyanine green (ICG) (0.5 mL) was injected at two points (arrow) to the proximal part of the stapler line (arrowhead). (b) The ICG fluorescence imaging system clearly visualized the ICG-injected sites (arrow) around the tumor and the spread of the ICG as bright fluorescence. (c) The hilar lymph node #10; and (d) the lymph node were visualized by fluorescence imaging.

Pathological examination

After SN dissection, SNs were examined by standard histology with hematoxylin-eosin using conventional methods. They were also examined by conventional intraoperative rapid pathological diagnosis using frozen tissue sections during the operation.

Statistical analysis

To examine the association between SN identification and patient characteristics (age, gender, Brinkman index, tumor location [right vs. left, upper and middle lobes vs. lower lobe], tumor size, pathology [adenocarcinoma vs. squamous cell carcinoma] and operation [lobectomy vs. segmentectomy]), we used the unpaired t-test and chi-square test.

Results

SN identification rate

SNs were successfully identified in 16 of 22 patients (identification rate: 72.7%) (Table 2). The lymphatic flow was clearly observed in all 16 patients. Among the six patients in whom the SNs were not identified, some did not show lymphatic flow and the SN was not identified, even though the injected ICG was visible in others. In 13 of 16 patients, there was no evidence of metastasis in SNs or non-SNs. There was SN metastasis detected in three patients (Cases 6, 12 and 14).

Representative patients with SN metastasis

In Case 14, the primary tumor was 16 mm in diameter and located in the left S10. It was resected using VATS and the operative rapid pathological diagnosis demonstrated adenocarcinoma. Immediately after intraoperative ICG injection, the fluorescence imaging system enabled visualization of the hilar lymph node (#10) (Fig 1c,d). The ICG flowed from the primary tumor via #10 to the subcarinal lymph node (#7). We defined #10 as the SN. The intraoperative rapid pathological diagnosis revealed metastasis in #10 and #7. We decided to convert from segmentectomy to lobectomy, and performed aortic and subcarinal lymph node dissection. Figure 2 shows the resected lymph node #10 ex vivo. The surrounding tissue often fluoresce strongly, not the lymph nodes themselves.
There was metastasis in SN (#10), the inferior interlobar lymph node (#11), and #7 LN. The subaortic lymph node (#5), lower lobar lymph node (#12), and segmental lymph node (#13) had no evidence of metastasis. The patient is alive 134 months after the operation and has had no recurrence.

In Case 6, the primary tumor was 20 mm in diameter and located in the left S3. The ICG flowed from the primary tumor via the upper lobar lymph node (#12) to the subaortic lymph node (#5). We defined #12 as the SN. The operative rapid pathological diagnosis revealed metastasis in #12 LN. However, lobectomy was not possible due to poor respiratory function, and segmentectomy was performed and only #5 that fluoresced by ICG was dissected. Following pathological examination, there was metastasis detected in #12u and the segmental lymph node (#13), whereas no evidence of metastasis in #5. The patient is alive at 148 months after the operation and has had no recurrence.

| No | B.I. | His. | Location | Tumor size | Operation | SN Dissected | mediastinal lymph node stations | pN | Follow-up period (months) | Recurrence (months after surgery) | Prognosis | Pathological findings of LN |
|----|------|------|----------|------------|-----------|--------------|--------------------------------|----|--------------------------|-----------------------------------|-----------|---------------------------|
| 1  | 0    | ad   | lt.S10   | 31 mm      | lob       | #10          | #4L, 5, 6, 7, 8, 9              | 0  | 62.9                     | None                             | Alive     | N0 (0/20)                 |
| 2  | 4000 | sq   | rt.S9 + 10 | 48 mm      | lob       | N.D.         | #4R, 7, 8, 9                      | 0  | 107.9                    | None                             | Alive     | N0 (0/32)                 |
| 3  | 800  | ad   | rt.S2    | 28 mm      | lob       | #11s         | #4R, 7                           | 0  | 61.3                     | None                             | Alive     | N0 (0/13)                 |
| 4  | 0    | ad   | rt.S8    | 15 mm      | seg       | N.D.         | #7, 8                           | 0  | 74.1                     | None                             | Alive     | N0 (0/6)                  |
| 5  | 0    | ad   | rt.S2    | 22 mm      | lob       | #4R          | #4R                             | 0  | 54.9                     | None                             | Alive     | N0 (0/26)                 |
| 6  | 1400 | ad   | lt.S3    | 20 mm      | seg       | #12          | #5                              | 0  | 148.0                    | None                             | Alive     | #5(0/6), #12 (1/1), #13(3/10) |
| 7  | 740  | ad   | lt.S1 + 2 | 31 mm      | lob       | #10          | #5, 6, 7                         | 0  | 103.2                    | Lung (37.9)                       | Dead      | N0 (0/18)                 |
| 8  | 1280 | ad   | lt.S9    | 19 mm      | seg       | #12          | #8, 9                           | 0  | 94.2                     | None                             | Alive     | N0 (0/15)                 |
| 9  | 0    | ad   | rt.S8    | 17 mm      | seg       | #7           | #7                              | 0  | 146.0                    | Lung (33.3)                       | Alive     | N0 (0/15)                 |
| 10 | 1620 | sq   | lt.S8    | 47 mm      | lob       | N.D.         | #7, 8, 9                         | 0  | 17.4                     | Bone (5.2)                       | Dead      | N0 (0/9)                  |
| 11 | 660  | ad   | rt.S6    | 15 mm      | seg       | #7           | #7, 8, 9                         | 0  | 17.5                     | Adrenal gland (17.0)             | Dead      | N0 (0/19)                 |
| 12 | 0    | ad   | rt.S4    | 33 mm      | lob       | #11s         | #4R, 7                          | 0  | 144.7                    | Pleura (73.2)                      | Alive     | #4R(0/15), #7 (0/10), #11s (1/6), #12 (0/1), #13(0/2) |
| 13 | 680  | ad   | lt.S1 + 2 | 18 mm      | seg       | N.D.         | #5, 6                           | 0  | 120.2                    | None                             | Alive     | N0 (0/10)                 |
| 14 | 720  | ad   | lt.S10   | 16 mm      | lob       | #10          | #5, 7                           | 2  | 134.4                    | None                             | Alive     | #5(0/10), #7 (1/2), #10 (1/1), #11 (1/4), #12 (0/10), #13(0/1) |
| 15 | 0    | sq   | rt.S3    | 37 mm      | lob       | #4R          | #4R, 7                          | 0  | 50.9                     | Adrenal gland (6.3)               | Dead (other disease)              | N0 (0/38) |
| 16 | 0    | ad   | rt.S2    | 17 mm      | lob       | #12          | #4R, 7                          | 0  | 85.0                     | None                             | Alive     | N0 (0/17)                 |
| 17 | 0    | ad   | rt.S2    | 26 mm      | lob       | #4R          | #4R                            | 0  | 59.7                     | None                             | Alive     | N0 (0/10)                 |
| 18 | 0    | ad   | rt.S10   | 25 mm      | lob       | N.D.         | #4R, 7                          | 0  | 102.5                    | None                             | Alive     | N0 (0/19)                 |
| 19 | 0    | ad   | rt.S6    | 28 mm      | seg       | #10          | #7                             | 0  | 132.5                    | None                             | Alive     | N0 (0/20)                 |
| 20 | 600  | sq   | rt.S2    | 40 mm      | lob       | N.D.         | #4R, 7                          | 0  | 111.8                    | None                             | Alive     | N0 (0/11)                 |
| 21 | 1500 | sq   | rt.S8    | 30 mm      | lob       | #10          | #4R, 7                          | 0  | 129.0                    | None                             | Alive     | N0 (0/22)                 |
| 22 | 1200 | la   | rt.S1    | 34 mm      | lob       | #11s         | #4R, 7                          | 0  | 81.1                     | None                             | Alive     | N0 (0/12)                 |

B.I., Brinkmann Index; His., histology; SN, sentinel node; lob, lobectomy; seg, segmentectomy; N.D., not detected; ad, adenocarcinoma; sq., squamous cell carcinoma; la, large cell carcinoma.

Table 2 Case details

There was metastasis in SN (#10), the inferior interlobar lymph node (#11), and #7 LN. The subaortic lymph node (#5), lower lobar lymph node (#12), and segmental lymph node (#13) had no evidence of metastasis. The patient is alive 134 months after the operation and has had no recurrence.

In Case 6, the primary tumor was 20 mm in diameter and located in the left S3. The ICG flowed from the primary tumor via the upper lobar lymph node (#12) to the subaortic lymph node (#5). We defined #12 as the SN. The operative rapid pathological diagnosis revealed metastasis in #12 LN. However, lobectomy was not possible due to poor respiratory function, and segmentectomy was performed and only #5 that fluoresced by ICG was dissected. Following pathological examination, there was metastasis detected in #12u and the segmental lymph node (#13), whereas no evidence of metastasis in #5. The patient is alive at 148 months after the operation and has had no recurrence.
nodes did. The interior portion of the lymph nodes did not emit bright fluorescence, but the external portion or the circuit of the lymph nodes did.

In Case 12, the primary tumor was 33 mm in diameter and located in the right S2. The ICG flowed from the primary tumor via the superior interlobar lymph node (#11s) to the right lower paratracheal lymph node (#4R). We defined #11s as the SN. We performed right middle lobectomy with lymph node dissection. There was only metastasis detected in the SN (#11s), and #4R and #7 had no metastasis. Disseminated lesions recurred 73 months after the operation, but the patient is alive at 144 months due to epidermal growth factor receptor-tyrosine kinase inhibitor (EGFR-TKI) treatment. Before follow-up, the false-positive rate was 0% (0/16) and the accuracy rate was 100% (16/16) in this study.

Follow-up of prognosis and recurrence

We followed-up all patients with regard to prognosis and recurrence to confirm the pattern of lymph node metastasis after surgery. The median follow-up time was 92.7 months (17.4–148.0 months). Seven (31.8%) of 22 patients had recurrence. Hematogenous metastasis was noted in five (lung metastasis; two, adrenal metastasis; two, bone metastasis; one), dissemination was noted in one, and lymph node metastasis was noted in one. Four of seven patients with recurrence died from lung cancer.

Of the 16 patients in whom the SN was identified, 15, including three with metastasis to the SN, had no postoperative lymph node recurrence. Only one patient (Case 5) had lymph node metastasis during follow-up. In this case, the primary tumor was located in the right S2. We defined #4R as the SN and performed lobectomy and superior mediastinal lymph node dissection. The postoperative pathological examination confirmed no metastasis in all resected lymph nodes. Recurrence in the right upper paratracheal lymph node (#2R) was found at one year and nine months after surgery. Four months later, it had spread to the left supraclavicular lymph node. In 15 (93.8%) of 16 patients with SNs, their metastatic patterns predicted the metastatic pattern of all lymph nodes. After follow-up was completed, the false-negative rate was 7.7% (1/13) and the accuracy rate was 93.8% (15/16) in this study.

Identification of the SN and clinical findings

We examined the association between SN identification and patient characteristics (age, gender, Brinkman index, tumor location [right vs. left, upper and middle lobe vs. lower lobe], tumor size, pathology [adenocarcinoma vs. squamous cell carcinoma] and operation [lobectomy vs. segmentectomy]). There was no significant association between SN identification and patient characteristics.

Discussion

SN biopsy has been used in the management of numerous cancers to avoid unnecessary lymphadenectomy. In recent years, there have been reports of lobe-specific selective lymph node dissection in lung cancer. The results in some studies have been reported to be similar to those of systematic lymph node dissection, but others reported more lymph node recurrence and a poorer prognosis than systematic dissection. Thus, as there is no consensus, SN identification for lymph vessel flow may be necessary to reduce the extent of lymph node dissection in each case.

We tried to identify the SN using an ICG fluorescence method. The results of SN identification by the ICG fluorescence method in NSCLC in four previous studies are shown in Table 3. The SN identification rate in our study was 72.7%, similar to that reported from other institutions (80.3%–83.7%). Although the results by Gilmore et al. were low (45.5%), they conducted an optimized ICG dose escalation trial for SN identification and reported that SN identification increased according to the ICG dose, with a success rate of 89% at 1000 mg or greater. The use of a sufficient amount of ICG can detect SNs in 70%–80%. The metastatic rate of SNs in this study was 3/15 (19%). A wide-ranging metastatic rate of SNs (4%–40%) has been previously reported. As the study by Gilmore et al. included large tumors (4, 6 and 6.5 cm), the metastatic rate of SNs may have been higher (40%). On the other hand, other studies included stage I cancer and the metastatic rate was relatively low (4.0%–9.7%).

In postoperative diagnosis, the false-positive rate was 0% and the accuracy rate was 100%, similar to results reported from other institutions. In other studies, the false-positive rate was 0%–2.3% and the accuracy rate was 97.3%–100%. The “true positive rate” should not change by following-up...
the patients, whereas the “true false-negative rate” and “true accuracy rate” may change during follow-up. Previous studies had a shorter period of follow-up after surgery. We followed-up all patients for 92.7 months (mean) after surgery to confirm the precise pattern of lymph node metastasis. A total of 12 patients had no SN metastasis at the intraoperative rapid or postoperative pathological diagnosis and no lymph node metastasis during the follow-up period. Only one patient exhibited no SN metastasis at the postoperative pathological diagnosis and lymph node metastasis during the follow-up period. Recurrence in #2R was noted at one year and nine months after surgery. In our study, the “true false-negative rate” was 7.7% (1/13) and the “true accuracy rate” was 93.8% (15/16). Three patients showed SN metastasis at the intraoperative rapid or postoperative pathological diagnosis, but had no lymph node metastasis during the follow-up period.

There was no significant association between SN identification and patient characteristics (age, gender, Brinkman index, tumor location, tumor size, pathology, and operation method) in our study, consistent with the study by Nomori et al.17

The limitations of the present study were the small number of patients and single-institution design. Many patients via a multi-institutional study may lead to more accurate results. We used the conventional pathological method for the intraoperative rapid or postoperative pathological diagnosis. In future, we may be able to precisely detect SN metastasis using reverse transcription-polymerase chain reaction (RT-PCR) or immunohistochemical analysis of CK-19 and CK-7.

We also tried another SN identification method using a CT contrast agent. Lymphography with transbronchial injection of a water-soluble extracellular CT contrast agent, iopamidol, was performed by bronchoscopy before surgery.22 The identification rate of the SN was 92.3% (12/13 patients). The metastatic rate was 16.7% (2/12 patients). The accuracy rate was 100% (12/12 patients) and the false negative rate was 0% (0/10 patients). We therefore consider it useful as a preoperative simulation tool for SN dissection. Compared with CT lymphography, the intraoperative ICG fluorescence method is advantageous in that bronchoscopy is unnecessary.

In conclusion, our identification technique for SN using ICG fluorescence imaging during lung cancer surgery had good identification and accuracy rates throughout the follow-up period. This technique may be useful for lymph node dissection during lung cancer surgery to perform segmentectomy and avoid unnecessary lymphadenectomy.

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Disclosure
The authors report no conflict of interest.

References
1. Morton DL, Wen DR, Wong JH et al. Technical details of intraoperative lymphatic mapping for early stage melanoma. Arch Surg 1992; 127: 392–9.
2. Krag DN, Weaver DL, Alex JC, Fairbank JT. Surgical resection and radiolocalization of the sentinel lymph node in breast cancer using a gamma probe. Surg Oncol 1993; 2: 335–9.
3. Little AG, DeHoyos A, Kirgan DM, Arcomano TR, Murray KD. Intraoperative lymphatic mapping for non-small cell lung cancer: The sentinel node technique. J Thorac Cardiovasc Surg 1999; 117: 220–4.
4. Sugi K, Fukuda M, Nakamura H, Kaneda Y. Comparison of three tracers for detecting sentinel lymph nodes in patients with clinical N0 lung cancer. Lung Cancer 2003; 39: 37–40.
5. Rzyman W, Hagen OM, Dziadziuszko R et al. Blue dye intraoperative sentinel lymph node mapping in early non-small cell lung cancer. Eur J Surg Oncol 2006; 32: 462–5.
6. Liptay MJ, Masters GA, Winchester DJ et al. Intraoperative radioisotope sentinel lymph node mapping in non-small cell lung cancer. Ann Thorac Surg 2000; 70: 384–90.
7. Liptay MJ, Grondin SC, Fry WA et al. Intraoperative sentinel lymph node mapping in non-small-cell lung cancer improves identification of micrometastases. J Clin Oncol 2002; 20: 1984–8.
8. Schmidt FE, Woltering EA, Webb WR, Garcia OM, Cohen JE, Rozans MH. Sentinel nodal assessment in patients with carcinoma of the lung. Ann Thorac Surg 2002; 74: 870–4.
9. Rzyman W, Hagen OM, Dziadziuszko R et al. Intraoperative, radio-guided sentinel lymph node mapping in 110 non-small cell lung cancer. Ann Thorac Surg 2006; 82: 237–42.
10. Nomori H, Horio H, Naruke T, Orikasa H, Yamazaki K, Suemasu K. Use of technetium-99m tin colloid for sentinel lymph node identification in non-small cell lung cancer. J Thorac Cardiovasc Surg 2002; 124: 486–92.
11. Sugi K, Kaneda Y, Sudoh M, Sakano H, Hamano K. Effect of radioisotope sentinel node mapping in patients with cT1 N0 M0 lung cancer. J Thorac Cardiovasc Surg 2003; 126: 568–73.
12. Nomori H, Ikeda K, Mori T et al. Sentinel node navigation segmentectomy for clinical stage IA non–small cell lung cancer. J Thorac Cardiovasc Surg 2007; 133: 780–5.
13. Liptay MJ, D’amico TA, Nwogu C et al. Intraoperative sentinel node mapping with technetium-99 in lung cancer: Results of CALGB 140203 multicenter phase II trial. J Thorac Oncol 2009; 4: 198–202.
14. Yamashita S, Tokuiishi K, Anami K et al. Video-assisted thoracoscopic indocyanine green fluorescence imaging system shows sentinel lymph nodes in non-small-cell lung cancer. J Thorac Cardiovasc Surg 2011; 141: 141–4.
15. Yamashita S, Tokuiishi K, Miyawaki M et al. Sentinel node navigation surgery by thoracoscopic fluorescence imaging system and molecular examination in non-small cell lung cancer. Ann Surg Oncol 2012; 19: 728–33.
16. Gilmore DM, Khullar OV, Jaklitsch MT, Chirieac LR, Frangioni JV, Colson YL. Identification of metastatic nodal disease in a phase I dose escalation trial of intraoperative sentinel lymph node mapping in NSCLC using near-infrared imaging. J Thorac Cardiovasc Surg 2013; 146: 562–70.
17. Nomori H, Cong Y, Sugimura H. Utility and pitfalls of sentinel node identification using indocyanine green during segmentectomy for cT1N0M0 non-small cell lung cancer. Surg Today 2016; 46: 908–13.
18. Hishida T, Miyaoaka E, Yokoi K et al. Lobe-specific nodal dissection for clinical stage I and II NSCLC: Japanese multi-institutional retrospective study using a propensity score analysis. J Thorac Oncol 2016; 11: 1529–37.
19. Meng D, Zhou Z, Wang Y, Wang L, Lv W, Hu J. Lymphadenectomy for clinical early-stage non-small-cell lung cancer: A systematic review and meta-analysis. Eur J Cardiothorac Surg 2016; 50: 597–604.
20. Maniwa T, Okumura T, Isaka M, Nakagawa K, Ohde Y, Kondo H. Recurrence of mediastinal node cancer after lobe-specific systematic nodal dissection for non-small-cell lung cancer. Eur J Cardiothorac Surg 2013; 44: 59–64.
21. Riquet M, Rivera C, Pricopi C et al. Is the lymphatic drainage of lung cancer lobe-specific? A surgical appraisal. Eur J Cardiothorac Surg 2015; 47: 543–9.
22. Takizawa H, Kondo K, Toba H et al. Computed tomography lymphography by transbronchial injection of iopamidol to identify sentinel nodes in preoperative patients with non-small cell lung cancer. J Thorac Cardiovasc Surg 2012; 144: 94–9.