Effect of Temperature on Anthocyanin Synthesis and Ethylene Production in the Fruit of Early- and Medium-maturing Apple Cultivars during Ripening Stages

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Abstract. The objective of this study was to investigate the effects of temperature treatments on anthocyanin accumulation and ethylene production in the fruit of early- and medium-maturing cultivars that were harvested early during fruit ripening. We first investigated the effects of various temperature treatments on anthocyanin accumulation in detached apples of ‘Tsugaru’, ‘Tsugaru Hime’, ‘Akane’ and ‘Akibae’ using an incubator. Three years of experiments demonstrated that at harvest, the lower-temperature treatments induced anthocyanin accumulation in ‘Tsugaru’, ‘Tsugaru Hime’, and ‘Akibae’ fruits, whereas the increases in anthocyanin accumulation under the 25 °C treatment were similar to those under the 15 and 20 °C treatments in ‘Akane’ fruit. The rate of ethylene production did not increase substantially during the temperature treatments in any of the four cultivars, except after the treatments of ‘Tsugaru’ fruit at harvest. The inhibition of ethylene action by the application of 1-methylcyclopropene (1-MCP) to detached fruits at harvest suppressed anthocyanin development under 15 and 20 °C temperature treatments in ‘Tsugaru’, ‘Tsugaru Hime’, and ‘Akibae’, but not in ‘Akane’.

Therefore, optimal temperatures for fruit pigmentation have been reported for various apple cultivars using detached fruit (Arakawa, 1991; Curry, 1997; Faragher, 1983; Marais et al., 2001). However, because the experimental conditions differed among the respective reports, comparing their results is difficult.

Ethylene is a key plant hormone involved in fruit ripening in apples (Saure, 1990). Whale and Singh (2007) reported that ethylene appeared to be a key factor regulating anthocyanin development in ‘Pink Lady’ fruit because ethylene production was significantly correlated with anthocyanin biosynthesis, but not with the accumulation of other flavonoids. Exogenous ethylene application promotes anthocyanin synthesis in fruit skin during the ripening stage (Larrigaudiere et al., 1996; Li et al., 2002). Recently, Johnston et al. (2009) showed that antisense suppression of MdACO1, an ethylene biosynthetic gene, decreased the ethylene concentration in the core cavity of fruit and led to poor red pigmentation in transgenic ‘Royal Gala’ apples. This reveals that ethylene production is essential for full anthocyanin accumulation in the apple fruit skin during ripening. Like anthocyanin synthesis, ethylene production in apple fruit is also influenced by temperature conditions. Tromp (1997) demonstrated that low-temperature treatment (16 °C) during a 6-week period after bloom repressed ethylene production in ‘Elstar’ fruit until a later stage, and cold storage of ‘Tsugaru’ fruit after harvest suppressed ethylene production (Tatsuki et al., 2011).

Genetic and molecular studies indicate that ethylene receptors are negative regulators of ethylene-dependent responses (Hua
The compound 1-MCP is thought to inhibit the perception of ethylene by competing for ethylene receptors in fruit (Blankenship and Dole, 2003), and its application to fruit slows ethylene-dependent responses, including ethylene evolution (Tatsuki et al., 2011). Several studies have investigated the effects of the inhibition of ethylene action in apples through the pre-harvest application of 1-MCP, including those on shelf life, ethylene evolution, and red skin color. Elfving et al. (2007) showed that the application of 1-MCP to ‘Scarletspar Delicious’ and ‘Caméo’ apple trees at 1 to 4 weeks before harvest (WBH) had little effect on skin color development. Yuan and Li (2008) reported that the application of 1-MCP to a ‘Delicious’ apple tree at 15 and 7 d before harvest did not affect the red coloration of the fruit. It is possible that the roles of ethylene action in anthocyanin accumulation differ among cultivars.

Anthocyanin accumulation in the fruit skin of four cultivars (Tsugaru, Tsugaru Hime, Akane, and Akibae) is susceptible to the effect of hot climatic conditions because their fruits are harvested in early autumn. In this study, we investigated the effect of temperature on anthocyanin accumulation in four cultivars of detached apples during ripening. The first experiment conducted in 2008–10 involved placing detached apples of the cultivars Tsugaru, Tsugaru Hime, Akane, and Akibae in an incubator. We measured the rates of ethylene production and anthocyanin accumulation at harvest in fruits of these cultivars to which 1-MCP had been applied to assess the effect of ethylene inhibition on the strength of pigmentation. Second, we examined changes in anthocyanin accumulation and ethylene generation in the fruit of potted ‘Misuzu Tsugaru’, a red sport of ‘Tsugaru’, under different climatic conditions in a greenhouse.

Materials and Methods

Plant materials. Apple trees (Malus ×domestica Borkh.) were grown in an orchard at the NARO Institute of Fruit Tree Science, Morioka, Japan. Morioka, with an average annual temperature of 10.2 °C, is a suitable area for apple production, particularly in terms of the temperature during fruit pigmentation. Two experiments were conducted to investigate the effect of temperature on anthocyanin accumulation in apple fruit. The first experiment involved an incubator and ‘Tsugaru’, ‘Tsugaru Hime’, ‘Akane’, and ‘Akibae’ apples, harvested from 2008 to 2010. In 2008, 2009, and 2010, the full bloom date was 4–9, 16 May, respectively. The second experiment required the use of a greenhouse and involved ‘Misuzu Tsugaru’ apple trees. All cultivars were grafted onto JM7 rootstock (Soejima et al., 1998) and trained as free spindle. The parentage, tree age, harvest date, and fruit quality at commercial harvest for the cultivars are listed in Table 1.

Temperature treatment of detached apples. The temperature treatments in the first experiment were conducted essentially according to the methods of Arakawa (1991). Fruits were bagged in single-layer paper bags for 2 months before commercial harvest to partially shade them from sunlight. Bagged fruits were sampled 2 and 4 WBH and at harvest. After the bags were removed, the fruits were placed in incubators set at 15, 20, 25, and 30 °C, respectively, with white and ultraviolet-B light provided by fluorescent tubes and left for 48 h. The temperature treatment period in Arakawa’s report was 96 h, but we used a 48-h period in this study because the anthocyanin concentration in fruit skins after 48 h was proportional to those after 96 h in a number of cultivars (Bessho, unpublished data). A total of six to 10 fruits from each cultivar were used for each temperature treatment, and their anthocyanin concentrations were measured. Attached fruits that were not bagged under field conditions were sampled concurrently for anthocyanin analysis.

1-MCP treatment. Bagged fruits of each cultivar were harvested at commercial maturity in 2011. On the day of harvest, after the paper bags were removed, ≈36 fruits were placed in a 117-L plastic container containing 1 ppm 1-MCP (SmartFresh®; AgroFresh Inc., Springhouse, PA) for 1 or 4 d (Fig. 1A). Untreated fruits were placed at room temperature after harvest and a proportion was subjected to temperature treatment at 2 DAH. The rates of ethylene production in 1-MCP-untreated and -treated fruits were measured at 0, 2, and 4 DAH. An anthocyanin analysis was conducted at 2 and 4 DAH.

Temperature treatment of potted trees. In the second experiment, two greenhouses located at the NARO Tohoku Agricultural Research Center were used to create control (25 °C day/15 °C night) and hotter climatic conditions (29 °C day/19 °C night). We performed the preliminary experiment under the control (25 °C day/15 °C night) and hotter climatic conditions (30 °C day/20 °C night) in 2008 and confirmed that a 5 °C increase in temperature suppressed anthocyanin accumulation in the fruit skin of ‘Misuzu Tsugaru’ in the greenhouse (data not shown). Therefore, the temperature difference was set at 4 °C (25 °C day/15 °C night) as the control condition and 29 °C day/19 °C night as the hotter climatic condition) in 2009. We set the duration of daily temperature variation as 12 h; the daytime (0600 to 1800 hr) temperature was maintained at 25 °C and the nighttime (1800 to 0600 hr) temperature was maintained at 15 °C for the control condition. Six potted ‘Misuzu Tsugaru’ apple trees were grown in a field until 5 WBH. Among them, two single-tree replicates were performed in each greenhouse at 5 WBH and grown under natural light conditions until harvest. The remaining two potted trees were grown in the field until harvest. Attached fruit on the potted trees were not bagged during the experiment. Fruits were collected at four points in time (1, 3, and 5 WBH and at harvest) for anthocyanin and ethylene analyses.

Anthocyanin and ethylene analyses. For the anthocyanin analyses, four disks (0.64-cm²/disk) were cut from the upper portion of each fruit with a cork borer and soaked in 10-mL hydrochloric acid/methanol (1:99, v/v) at 4 °C overnight. The absorbance of each extract at 530 nm was measured with a spectrophotometer (ultraviolet-1600; Shimadzu, Kyoto, Japan), and anthocyanin concentrations were calculated using a molar extinction coefficient of 3.43 × 10⁶ (Chalmers et al., 1973). For the ethylene analyses, whole apples were placed in a 1.2-L glass container equipped with a septum at room temperature (≈22 °C) for 30 min, and 1 mL of head-space gas was sampled using a syringe. The ethylene concentration was measured by gas chromatography (GC-201; Shimadzu).

Results

Effect of temperature on anthocyanin accumulation in detached apples. Temperature treatments were conducted at three points in time during fruit ripening to measure the potential ability to synthesize anthocyanin in the fruit skins of the early- and medium-maturing cultivars Tsugaru, Tsugaru Hime, Akane, and Akibae. Because the pattern of anthocyanin development in each cultivar was similar in 2008, 2009, and 2010, the results are shown as the mean of the data from all 3 years (Fig. 1).

In the fruit of ‘Tsugaru’, a major early-maturing cultivar in Japan, anthocyanin accumulation proceeded under the 15 and 20 °C treatments at the respective time points, and the maximum pigmentation potential was 5.9 μg·cm⁻² in the 15 °C treatment at harvest (Fig. 1A). The pigmentation potential in the 25 °C treatment was not found at 4 and 2 WBH but at harvest. The fruit of ‘Tsugaru Hime’, a red sport of ‘Tsugaru’, showed a similar pattern (Fig. 1B), except that at harvest the pigmentation potential in the 15, 20, and 25 °C treatments at harvest was more than 2-fold higher than that of ‘Tsugaru’, and the maximum pigmentation potential was found in the 15 °C treatment at harvest (12.3 μg·cm⁻²). The attached fruit of ‘Tsugaru Hime’ accumulated 2-fold higher anthocyanin levels (76.4 μg·cm⁻²) at harvest than did ‘Tsugaru’ (37.6 μg·cm⁻²).

In ‘Akane’ fruit, the pigmentation potential under the 15, 20, and 25 °C conditions was observed during fruit ripening (Fig. 1C). At 2 WBH, the pigmentation potential in the 25 °C treatment reached the same level as that in the 15 °C treatment. At harvest, there was no significant difference among the pigmentation potential in the 15, 20, and 25 °C treatment groups. The maximum pigmentation potential of ‘Akane’ fruit was found in the 25 °C treatment at harvest (15.1 μg·cm⁻²). The final anthocyanin concentration in the attached fruit of ‘Akane’ was 125.5 μg·cm⁻².
In the fruit of ‘Akibae’, the pigmentation potential under the 15, 20, and 25 °C conditions continued to increase during fruit ripening (Tukey-Kramer test, \( P < 0.05 \); data not shown); the maximum pigmentation potential was found in the 15 °C treatment at harvest (24.4 mg cm\(^{-2}\)) (Fig. 1D). The final anthocyanin concentration in the attached fruit of ‘Akibae’ was 135.6 mg cm\(^{-2}\), which was the highest among the four cultivars examined.

The temperature dependency of anthocyanin accumulation in fruit skin differed among the cultivars, and the interaction between cultivar and time point was significant with respect to the temperature treatment over 3 years (\( P < 0.001 \)). However, in the detached fruit of each cultivar, 15 °C was generally the optimal temperature for anthocyanin development during fruit ripening, whereas the increase in anthocyanin accumulation was repressed under the 30 °C treatment at all times, indicating that 30 °C was too high for anthocyanin synthesis in these apple cultivars. Therefore, we concluded that comparisons of pigmentation potential in the 15 and 25 °C treatments were effective for estimating anthocyanin accumulation in fruit skin under hotter climatic conditions in an early-maturing cultivar and in a medium-maturing cultivar that was harvested early.

Ethylene production during temperature treatment in detached apples. The ethylene

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Table 1. Parentage, tree age, harvest date, and fruit qualities at commercial harvest of the four cultivars.

| Cultivar          | Parentage              | Tree age in 2008 | Harvest date in 2008 | Harvest date in 2009 | Harvest date in 2010 | Percent blush (%) | Fruit firmness (N) | Starch index (0–5) | Sugar content (%) | Titration acidity (%) |
|-------------------|------------------------|------------------|----------------------|----------------------|----------------------|-------------------|-------------------|-------------------|-------------------|---------------------|
| Tsugaruy          | Golden Delicious × Jonathan | 8                | 15 Sept.             | 16 Sept.             | 20 Sept.             | 80                | 53.2              | 2.1               | 13.7              | 0.23                |
| Tsugaru Hime      | A red sport of Tsugaru | 4                | 19 Sept.             | 16 Sept.             | 21 Sept.             | —                 | —                | —                 | —                 | —                   |
| Akane             | Jonathan × Worcester   | 8                | 19 Sept.             | 23 Sept.             | 27 Sept.             | 75                | 54.4              | 0.5               | 11.8              | 0.64                |
| Akibaey           | Senshu × Tsugaru       | 7                | 30 Sept.             | 30 Sept.             | 4 Oct.               | > 80              | 63.6              | 2.9               | 13.8              | 0.47                |
| Misuzu Tsugaru    | A red sport of Tsugaru | 5                | 30 Sept.             | 15 Sept.             | —                   | —                 | —                | —                 | —                 | —                   |

\( ^{a} \)The starch index ranges from 0 to 5. A lower number means less intense staining of starch by iodine.

\( ^{b} \)The fruit quality values are the means from 2008 to 2010.

\( ^{c} \)The fruit qualities of red sports of ‘Tsugaru’ were similar to those of ‘Tsugaru’.

\( ^{d} \)The values given for various fruit qualities, except percent blush, are the mean values for 2004 and 2005. The value given for percent blush is the mean of the values from 2010 and 2011.
production rates in the four cultivars were measured in 2009 before and after the 15 and 25 °C treatments at the same time points as in Figure 1. Ethylene production in the fruits of ‘Tsugaru’ and ‘Tsugaru Hime’ at 4 WBH was below the detectable level before and after temperature treatment (Fig. 2A and B). The fruit began to produce ethylene at 2 WBH [up to 1.4 and 0.5 nl g⁻¹ fresh weight (FW)/h] in ‘Tsugaru’ and ‘Tsugaru Hime’, respectively. ‘Tsugaru’ fruit at harvest produced a large amount of ethylene (89.6 nl g⁻¹ FW/h); this increased significantly to 162.5 and 216.1 nl g⁻¹ FW/h after the 15 and 25 °C treatments, respectively. ‘Tsugaru’ fruit at harvest produced ethylene at a rate of 0.5 nl g⁻¹ FW/h, whereas ethylene production was below the detectable level in ‘Akibae’ fruit. In ‘Akane’, the ethylene level did not vary substantially during temperature treatment at any time point. ‘Akibae’ fruit at 2 WBH produced ethylene at a rate of 1.8 nl g⁻¹ FW/h, and it did not change during temperature treatment. At harvest, the ethylene production rate in ‘Akibae’ fruit increased after 25 °C treatment, but not after treatment at 15 °C. The effect of bagging on ethylene production was not found in the four cultivars at any point (data not shown).

The fruits of ‘Akane’ and ‘Akibae’ produced considerably lower levels of ethylene compared with those of ‘Tsugaru’ and ‘Tsugaru Hime’ during the experimental period (Fig. 2C and D). The interaction between cultivar and time point on ethylene production in detached fruit before temperature treatment was significant (P < 0.001), indicating that the rate of ethylene production varied among the cultivars. At 4 WBH, ‘Akane’ fruit produced ethylene at a rate of 0.5 nl g⁻¹ FW/h, whereas ethylene production was below the detectable level in ‘Akibae’ fruit. In ‘Akane’, the ethylene level did not vary substantially during temperature treatment at any time point. ‘Akibae’ fruit at 2 WBH produced ethylene at a rate of 1.8 nl g⁻¹ FW/h, and it did not change during temperature treatment. At harvest, the ethylene production rate in ‘Akibae’ fruit increased after 25 °C treatment, but not after treatment at 15 °C. The effect of bagging on ethylene production was not found in the four cultivars at any point (data not shown).

Effect of the postharvest application of 1-MCP on ethylene production and anthocyanin accumulation in detached apples. The rate of ethylene production in fruit with or without 1-MCP application was measured after harvest using detached fruit of each cultivar in 2011 (Fig. 3A). The application of 1-MCP repressed ethylene production in ‘Tsugaru’ fruit, as reported by Tatsuki et al. (2011). The rate of ethylene production in the 1-MCP-treated fruit decreased from 144.5 to 14.0 nl g⁻¹ FW/h at 4 DAH. Suppression of ethylene production by 1-MCP application was also evident in the ‘Tsugaru Hime’ fruit. The rate in the 1-MCP-treated ‘Tsugaru Hime’ fruit decreased from 50.1 to 1.0 nl g⁻¹ FW/h at 4 DAH. In contrast, 1-MCP application did not affect ethylene production by ‘Akane’ or ‘Akibae’ apples. The rate of ethylene production by the 1-MCP-untreated ‘Akane’ and ‘Akibae’ fruits at harvest was 0.8 and 0.3 nl g⁻¹ FW/h, respectively; ethylene production by 1-MCP-untreated and -treated fruits of both cultivars remained unchanged until 4 DAH.

The temperature treatment of 1-MCP-untreated and -treated fruits began at 2 DAH, and the anthocyanin concentration in the skin of the fruit was measured at 4 DAH (Fig. 3B). In ‘Tsugaru’ and ‘Tsugaru Hime’, the increase in anthocyanin concentration in fruit skin after temperature treatment was repressed by 1-MCP application. However, after the 15 °C treatment, there was no significant difference in anthocyanin concentration between fruits treated with and without 1-MCP application in ‘Tsugaru’. In ‘Akane’, after both temperature treatments, there was no significant difference in anthocyanin concentration between fruits treated with and without 1-MCP. In ‘Akibae’, the anthocyanin concentration in the 1-MCP-treated fruits was significantly lower than that in the 1-MCP-untreated fruit after the 15 °C treatment. After the 25 °C treatment, the concentration of anthocyanin increased to 7.3 and 5.6 μg·cm⁻² in the 1-MCP-untreated and -treated fruits, respectively; however, this difference was not significant. Thus, the inhibition of ethylene action by 1-MCP application suppressed anthocyanin accumulation in the fruits of ‘Tsugaru’, ‘Tsugaru Hime’, and ‘Akibae’, but not in ‘Akane’ fruit.

Effect of temperature on anthocyanin accumulation in attached fruit from potted trees. Potted trees of ‘Misuzu Tsugaru’, another red sport of ‘Tsugaru’, were used to investigate the effects of different climatic conditions on anthocyanin accumulation in attached apple fruit. The change in temperature in the field from July to September in 2009 followed the normal course, although the minimum temperature from late August to mid-September was lower than normal (Fig. 4A). Two greenhouses were used to create control (25 °C 12 h/15 °C 12 h) and hotter climatic conditions (29 °C 12 h/19 °C 12 h). The changes in ambient temperature in the greenhouses over 1 week during the experimental period (8 to 15 Sept.) are shown in Figure 4B.
The anthocyanin concentrations in fruits from potted trees in the field and under control and hotter climatic conditions were measured at four time points during fruit ripening (Fig. 4C). In the field, anthocyanin accumulation in the fruit skin continued to increase throughout the experiment; the anthocyanin concentration at harvest was 45.2 mg·cm⁻². In the potted trees from the two greenhouses, anthocyanin development in the fruit skin began later; in the control treatment, anthocyanin accumulation began between 1 and 3 WBH, and under the hotter climatic condition, anthocyanin accumulation began during the final week of the experiment. The final anthocyanin concentrations in fruits from trees subjected to the control and hotter climatic conditions were 26.8 and 19.0 µg·cm⁻², respectively. The rates of increase in anthocyanin concentration in the field, control greenhouse, and hotter greenhouse during the final week of the study were 2.5, 2.1, and 1.9 µg·cm⁻²·d⁻¹, respectively.

Ethylene production in fruits from the potted trees varied markedly (Fig. 4D–F) and the rate under field conditions at harvest was considerably lower in contrast to ‘Tsugaru’ and ‘Tsugaru Hime’ fruits from trees planted in the ground (Fig. 2A and B). In fruits from potted trees in the field, ethylene production remained under the detectable level until 1 WBH, and several fruits produced ethylene at harvest (up to 1.0 nl·g⁻¹ FW/h). With respect to trees subjected to control and hotter climatic conditions, more
than half of the fruit began to produce ethylene at 1 WBH, and the rates of production peaked at 18.8 and 181.7 nl·g⁻¹·h⁻¹, respectively, at harvest. The quality of the fruit from the potted trees (excluding fruit firmness) was similar to those of ‘Tsu- garu’ at commercial harvest (Table 1). Fruit firmness in the potted trees grown under field, control, and hotter climatic conditions was found to be 76.4, 66.4, and 70.4 N, respectively. Because there were no differences in fruit firmness among the three conditions, the high fruit firmness was likely the result of growing the trees in a pot.

In addition, the anthocyanin concentration after the 15 and 25 °C treatments was measured in 2011 in ‘Misuzu Tsugaru’ fruit detached from trees planted in the ground using the same method as in Figure 1. The pigmentation potential under the 15 and 25 °C treatments was found at 2 WBH and at harvest (Fig. 4G); moreover, the pattern was similar to that in ‘Tsugaru’ and ‘Tsugaru Hime’ (Fig. 1A and B). After the 15 °C treatment, the anthocyanin concentration was higher than that after the 25 °C treatment at 1 WBH; in comparison, there was no significant difference between the treatments at harvest. Therefore, anthocyanin accumulation in the skin of ‘Misuzu Tsugaru’ fruit at
harvest proceeded at a relatively high temperature (25 °C).

**Discussion**

We investigated the pigmentation potential for anthocyanin synthesis in fruit skins during ripening using detached fruit of four cultivars and found that the temperature dependency of anthocyanin accumulation differed among the cultivars, particularly for the 25 °C treatment (Fig. 1). Because ‘Tsugaru Hime’ was a red sport of ‘Tsugaru’, they showed a similar pattern in which the pigmentation potential in the 25 °C treatment increased at harvest compared with that at 2 WBH. The potential of ‘Akane’ in the 25 °C treated fruit reached the same level as in the 15 °C treated fruit at 2 WBH. These results indicate that in these cultivars, anthocyanin accumulation proceeded under hotter conditions toward harvest.

Lower-temperature treatment induced greater anthocyanin accumulation in ‘Akibae’ fruit up to the time of harvest, similar to ‘Tsugaru’ and ‘Tsugaru Hime’. In ‘Akibae’, the pigmentation potential at harvest after the 25 °C treatment (15.3 μg·cm⁻²) was the highest among the four cultivars. This trait of ‘Akibae’ would give rise to high anthocyanin concentrations in the skin of attached fruit at harvest.

The effect of the inhibition of ethylene action by 1-MCP application on ethylene production and anthocyanin accumulation differed among the cultivars (Fig. 3). The 1-MCP-treated fruits of ‘Tsugaru’ and ‘Tsugaru Hime’ showed a similar response; ethylene production and anthocyanin accumulation were repressed after the application of 1-MCP. In contrast, the inhibition of ethylene action by 1-MCP application did not affect either ethylene production or anthocyanin production in ‘Akane’ fruit at harvest. In the 1-MCP-treated fruit of ‘Akibae’, ethylene production remained unchanged, but anthocyanin synthesis in the 15 °C treatment was repressed. Blankenship and Dole (2003) assumed that 1-MCP binds irreversibly to ethylene receptors present at the time of treatment and that any recovery of ethylene sensitivity is the result of the appearance of a new site. In apple, six ethylene receptor genes (MdETR1, MdETR1B, MdETR2, MdETR5, MdERI, and MdERS2) have been isolated (Dal Cin et al., 2005; Tatsuki and Endo, 2006; Wiersma et al., 2007), but the rate of turnover of the receptors and the duration of the inhibitory effect by 1-MCP-bound receptors remain unknown (Tatsuki et al., 2007). Therefore, variations in the response to 1-MCP application may reflect differences in ethylene receptor characteristics between cultivars.

Lin-Wang et al. (2011) and Palmer et al. (2012) compared changes in the anthocyanin concentration of ‘Royal Gala’ fruit from trees grown in a region with moderate summer temperatures in New Zealand and in a region with high summer temperatures in Spain. They demonstrated that anthocyanin accumulation in fruit from the trees grown in Spain was suppressed to one-fourth compared with that in fruit from New Zealand. They also conducted orchard-based apple heating experiments using ‘Royal Gala’ trees and demonstrated that the maintenance of fruit skin temperatures above 20 °C for a 7-d period, starting 11 d before harvest, inhibited increases in the anthocyanin concentration in fruit skin. Moreover, it has been reported that a decrease in orchard temperature improved the color of apple fruit (Iglesias et al., 2002, 2005), clearly indicating the effect of temperature on anthocyanin bio-synthesis in apple fruit skin. We obtained a similar result; the anthocyanin concentration in ‘Misuzu Tsugaru’ fruit from potted trees grown at a hotter climate for 5 weeks was lower than that in fruit grown under control conditions (Fig. 4C). Nevertheless, the rate of increase in anthocyanin concentration in the hotter climate was elevated from 0.3 to 1.9 μg·cm⁻²·d⁻¹ at 1 WBH. The increased rate during the last week in the hotter climate reached a level that was close to that under control conditions (2.1 μg·cm⁻²·d⁻¹). This result was supported by the finding that detached fruit of ‘Misuzu Tsugaru’ at harvest accumulated identical anthocyanin levels after the 25 and 15 °C treatments, unlike the findings at 2 WBH (Fig. 4G).

Although the rates of ethylene production varied, ‘Misuzu Tsugaru’ fruit sampled from potted trees after the hotter climate treatment produced ethylene at up to 181.7 nl·g⁻¹·h⁻¹, which was 9-fold higher than under the control condition at harvest (Fig. 4E and F). This indicates that ‘Misuzu Tsugaru’ fruit grown under hotter climatic conditions during ripening produced a larger amount of ethylene after harvest. The development of red color proceeded in ‘Misuzu Tsugaru’ attached fruit from the potted trees in the field, although ethylene production was below the detectable level (Fig. 4C and D), suggesting that ‘Misuzu Tsugaru’, the ethylene action required for pigmentation in the fruit skin is activated by a small amount of ethylene. Arakawa et al. (1985) demonstrated that the application of aminooxyacetic acid, an inhibitor of ethylene synthesis, decreased ethylene production but had little effect on anthocyanin development in ultraviolet-B-irradiated ‘Jonathan’ fruit. Our speculation appears to support not only their report, but also our results in Figures 1 and 2. The final anthocyanin concentrations in attached fruits of ‘Akane’ and ‘Akibae’ were higher than those in attached fruits of ‘Tsugaru’ and ‘Tsugaru Hime’, whereas the ethylene levels in the former cultivars were considerably lower than those in the latter cultivars at harvest (Figs. 1 and 2A). In the former cultivars, a relatively lower amount of ethylene may be enough for inducing ethylene action to synthesize anthocyanin in fruit skin at harvest. Moreover, it is possible that the increase in ethylene in detached ‘Tsugaru’ fruit at harvest (Fig. 2A) did not contribute to the effect of ethylene on anthocyanin accumulation under 15 or 25 °C treatment (Fig. 1A).

In conclusion, comparisons of pigmentation potential after treatment at 15 and 25 °C in detached fruit were effective for estimating anthocyanin accumulation in fruit skin under hotter climatic conditions in early- and medium-maturing cultivars that were harvested early because the response of anthocyanin development in attached fruit of potted trees of ‘Misuzu Tsugaru’ to hotter climatic conditions during ripening was similar to the results using detached fruit. Our experiment using potted trees of ‘Misuzu Tsugaru’ in a greenhouse indicates that exposure to the hotter condition during ripening increased ethylene production in the fruit after harvest.

**Literature Cited**

Arakawa, O. 1991. Effect of temperature on anthocyanin accumulation in apple fruit as affected by cultivar, stage of fruit ripening and bagging. J. Hortic. Sci. 66:763–768.

Arakawa, O., Y. Hori, and R. Ogata. 1985. Relative effectiveness and interaction of ultraviolet-B, red and blue light in anthocyanin synthesis of apple fruit. Physiol. Plant. 34:323–327.

Ban, Y., C. Honda, Y. Hanayama, M. Igarashi, H. Bessho, and T. Moriguchi. 2007. Isolation and functional analysis of a MYB transcription factor gene that is a key regulator for the development of red coloration in apple skin. Plant Cell Physiol. 48:958–970.

Blankenship, S.M. and J.M. Dole. 2003. 1-methylcyclopropene: A review. Postharvest Biol. Technol. 28:1–25.

Chalmers, D.J., J.D. Faragher, and J.W. Raff. 1973. Changes in anthocyanin synthesis as an index of maturity in red apple varieties. J. Hortic. Sci. 48:378–392.

Curry, E.A. 1997. Temperature for optimum anthocyanin accumulation in apple tissue. J. Hortic. Sci. 72:723–729.

Dal Cin, V., M. Danesin, A. Boshetti, A. Dorigoni, and A. Ramina. 2005. Ethylene biosynthesis and perception in apple fruitlet abscission [Malus domestica (L.) Borkh]. J. Expt. Bot. 56:3095–3105.

Elfwing, D.C., R.D. Stephen, A.N. Reed, and D.B. Visser. 2007. Preharvest application of sprayable 1-methylcyclopropene in the orchard for management of apple harvest and postharvest conditions. HortScience 42:1192–1199.

Faragher, J.D. 1983. Temperature regulation of anthocyanin accumulation in apple skin. J. Expt. Bot. 34:1291–1298.

Hua, L. and E.M. Meyerowitz. 1998. Ethylene responses are negatively regulated by a receptor gene family in Arabidopsis thaliana. Cell 94:261–271.

Iglesias, I. and G. Echeverria. 2009. Does strain affect fruit color development, anthocyanin content and fruit quality in ‘Gala’ apples? A comparative study over three seasons. J. Amer. Pomol. Soc. 63:168–180.

Iglesias, I., G. Echeverria, and M.L. Lopez. 2012. Fruit color development, anthocyanin content, standard quality, volatile compound emissions and consumer acceptability of several ‘Fuji’ apple strains. Sci. Hortic. 137:138–147.

Iglesias, I., J. Graell, G. Echeverria, and M. Vendrell. 1999a. Differences in fruit colour development, anthocyanin content, yield and quality of seven ‘Delicious’ apple strains. Fruit Var. J. 53:133–145.

Iglesias, I., J. Graell, D. Faro, C. Larrigaudiere, I. Recasens, G. Echeverria, and M. Vendrell. 1999b. Efecto del sistema de riego en la

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coloración de los frutos, contenido de antocianos y actividad de la fenilalanina amonioilasa (PAL), en la variedad de manzana ‘Starking Delicious’. Investigacion Agraria Producción y Proteccion Vegetal 14:157–172.

Iglesias, I., J. Salvia, L. Torguet, and C. Cabús. 2002. Orchard cooling with overtree microsprinkler irrigation to improve fruit colour and quality of ‘Topred Delicious’ apples. Sci. Hort. 93:39–51.

Iglesias, I., J. Salvia, L. Torguet, and R. Montserrat. 2005. The evaporative cooling effects of overtree microsprinkler irrigation on ‘Mondial Gala’ apples. Sci. Hort. 93:267–287.

Johnston, J.W., K. Gunaseelan, P. Pidakala, M. Wang, and R.J. Schaffer. 2009. Coordination of early and late ripening events in apples is regulated through differential sensitivities to ethylene. J. Expt. Bot. 60:2689–2699.

Li, Z., H. Gemma, and S. Iwahori. 2002. Simulation of ‘Fuji’ apple skin color by ethephon and phosphorus-calcium mixed compounds in relation to flavonoid synthesis. Sci. Hort. 94:193–199.

Lin-Wang, K., D. Micheletti, J. Palmer, R. Volz, L. Lozano, R. Espley, R.P. Hellens, D. Chagné, D.D. Rowan, M. Troggio, I. Iglesias, and A.C. Allan. 2011. High temperature reduces apple fruit colour via modulation of the anthocyanin regulatory complex. Plant Cell Environ. 34:1176–1190.

Marais, E., G. Jacobs, and D.M. Holcroft. 2001. Light and temperature affect postharvest colour development in ‘Cripps Pink’ apples. Acta Hort. 553:91–93.

Palmer, J., L. Lozano, D. Chagné, R. Volz, K. Lin-Wang, J. Bonany, D. Micheletti, M. Troggio, A. White, S. Kumar, A.C. Allan, and I. Iglesias. 2012. Physiological, molecular and genetic control of apple skin colouration under hot temperature environments. In: Litz RE (ed.). Proc. XXVIIIth IHC-IS on Genomics and Genetic Transformation of Horticultural Crops. Acta Hort. 929:81–87.

Saure, M.C. 1990. External control of anthocyanin formation in apple. J. Amer. Soc. Hort. Sci. 121:746–750.

Li, Z., H. Gemma, and S. Iwahori. 2002. Simulation of ‘Fuji’ apple skin color by ethephon and phosphorus-calcium mixed compounds in relation to flavonoid synthesis. Sci. Hort. 94:193–199.

Lin-Wang, K., D. Micheletti, J. Palmer, R. Volz, L. Lozano, R. Espley, R.P. Hellens, D. Chagné, D.D. Rowan, M. Troggio, I. Iglesias, and A.C. Allan. 2011. High temperature reduces apple fruit colour via modulation of the anthocyanin regulatory complex. Plant Cell Environ. 34:1176–1190.

Tatsuki, M., H. Hayama, H. Yoshioka, and Y. Nakamura. 2011. Cold pre-treatment is effective for 1-MCP efficacy in ‘Tsugaru’ apple fruit. Postharvest Biol. Technol. 62:282–287.

Tromp, J. 1997. Maturity of apple cv. Elstar as affected by temperature during a six week period following bloom. J. Hort. Sci. 72:811–819.

Ubi, B.E. 2004. External stimulation of anthocyanin biosynthesis in apple fruit. Food Agr. Environ. 2:65–70.

Ubi, B.E., C. Honda, H. Bessho, S. Kondo, M. Wada, S. Kobayashi, and T. Moriguchi. 2006. Expression of anthocyanin biosynthetic genes in apple skin: Effect of UV-B and temperature. Plant Sci. 170:571–578.

Whale, S.K. and Z. Singh. 2007. Endogenous ethylene and color development in the skin of ‘Pink Lady’ apple. J. Amer. Soc. Hort. Sci. 132:20–28.

Wiersma, P.A., H. Zhang, C. Lu, A. Quail, and P.M.A. Toivonen. 2007. Survey of the expression of genes for ethylene synthesis and perception during maturation and ripening of ‘Sunrise’ and ‘Golden Delicious’ apple fruit. Postharvest Biol. Technol. 44:204–211.

Yuan, R. and J. Li. 2008. Effect of sprayable 1-MCP, AVG, and NAA on ethylene biosynthesis, preharvest fruit drop, fruit maturity, and quality of ‘Delicious’ apples. HortScience 43:1454–1460.