Changes in Antioxidant/Pro-Oxidant Ratio of Brain Tissue Homogenates in Response to Cocos nucifera Treatment

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Authors’ contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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ABSTRACT

Objective: This study revealed the effect of Cocos nucifera juice on antioxidant/pro-oxidant ratio in brain tissue homogenates of wistar rats.

Materials and Methods: 40 male wistar rats were collected and randomly selected into 4 groups. Treatment protocols were; i-normal saline, ii-3ml C. nucifera juice, iii-6ml C. nucifera juice, iv-9ml C. nucifera juice. The study period lasted for 42 days. Brain tissue homogenate was prepared and biochemical analysis for antioxidant enzymes superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPx); pro-oxidant biomarkers hydrogen peroxide (H$_2$O$_2$), thiobarbituric acid reactive substances (TBARS) and nitric oxide (NO) were assayed for.

Results: The result showed that there was an increase in antioxidants concentration and a simultaneous decrease in pro-oxidant species generation as the dose of treatment was increased. The ratio of antioxidants to pro-oxidants showed a direct proportionality to the treatment dose.

Conclusion: C. nucifera juice has the ability to improve the level of production or synthesis of the assayed antioxidant enzymes and also has the efficacy to suppress pro-oxidant species generation.

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1. INTRODUCTION

Oftentimes, excessive free radical generation occurs in the body and the oxidant system cannot cope with such phenomenon [1]. Under normal conditions, there is a balance between the level of pro-oxidants and antioxidants in living systems [2]. A loss in this balance towards pro-oxidants is a condition called oxidative stress. Oxidative stress occurs when the generation of free radicals and active intermediates in a system exceeds the system's ability to neutralize and eliminate them [1]. To cope with the oxidative stress elicited by aerobic metabolism, animal and human cells have developed ubiquitous antioxidant defense system, which consists of superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx) and glutathione reductase (GR) together with a number of low molecular-weight (LMW) antioxidants such as ascorbate, α-tocopherol, glutathione, cysteine, thioredoxin and vitamins [1][3]. However, these antioxidant defense systems may be overwhelmed by various pathological or environmental factors so that a fraction of reactive oxygen species may escape destruction and form the far more reactive hydroxyl radicals. Oxidative stress has been implicated in several neurodegenerative diseases like Parkinsonism [1][4] and Alzheimer's disease [1][5]. Recent studies also suggest that free radical generation is positively correlated with learning, memory and motor impairment [1]. Coconut, botanically known as Cocos nucifera [6], belongs to the family of Arecaceae (Palmae), an important member of monocotyledons [7]. Indonesia, Philippines, India and Brazil are the world's largest coconut producers, accounting for over 25% of world production [6][8]. C. nucifera (Coconut) juice acts as a natural energy or sports drink, as it is rich in mineral content especially in potassium levels. C. nucifera juice has a high demand among consumers for its zero fat content and low contents of carbohydrates and sodium. C. nucifera juice serves as a potential healthy drink for adults and oldpersons as it has promising health utilities. C. nucifera is considered to be sterile unless the fruit is damaged from an external source [8][9]. Coconut water is rich in mineral content with high potassium and antioxidant contents which has various medical utilities. C. nucifera also contains cytokinin [10] which is one of themeney beneficial components it is composed of. Around the world, C. nucifera juice has been used in popular medicine for the management of various diseases, such as arthritis, diarrhea, liver and kidney diseases [11]. It is believed to be anti-gingivitic, febrifugal, antibronchitis and antibleenorragic. This study revealed further effect of C. nucifera juice on antioxidant/pro-oxidant ratio in brain tissue homogenate.

2. MATERIALS AND METHODS

2.1 Plant Material

C. nucifera juice was collected daily from fresh fruits into sterilized bottles. The fruits were purchased during the dry season dated between 16th November and 31st December, 2018.

2.2 Animal Collection

40 male wistar rats aged 80 days with weight between 150 to 170 grams were collected within the matrices of Experimental Animals Unit, Department of Human Physiology, Madonna University. The animals were confirmed to be healthy by a veterinarian in same institution.

2.3 Study Design

This study included 4 groups sampled randomly from a population of 40 wistar rats. The treatments were; i- normal saline and served as control, ii-3ml C. nucifera juice, iii-6ml C. nucifera juice, iv-9ml C. nucifera juice. The animals were exposed to an almost constant temperature of 25±3 ºC in normal day/night rhythm and pelleted feed was provided ad libitum. The duration of study was 42 days; 14 days for acclimatization, 28 days for treatment protocols.

2.4 Sacrifice and Sample Collection

Sacrifice and brain sample collection was done on day 42 using methods previously described [1].

2.5 Biochemical Analysis

Brain tissue homogenate was prepared following methods previously described. Antioxidants assayed for include the enzymatic antioxidants;
superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPx), and pro-oxidants; hydrogen peroxide, thiobarbituric acid reactive substances (TBARS) and the nitrosative agent, nitric oxide. Biochemical assay for each marker was performed following standard protocols [1].

2.6 Statistical Analysis

IBM® SPSS version 20.0 was the statistical tool used for this study. Data was expressed as Mean ± standard error of mean (SEM). One way analyses of variance (ANOVA) was calculated using recommended bio-statistical guidelines [11].

3. RESULTS

The following results were obtained from this study:

3.1 Effect of *C. nucifera* Treatment on Antioxidants

*C. nucifera* juice treatment caused an increase in antioxidant enzymes and this increase was directly proportional to the dose administered. As the dose of treatment was increased, there was a significant increase in the brain tissue level of SOD, CAT and GPx compared to control. From this result, the juice has a dose-dependent positive effect on brain tissue antioxidant enzyme markers.

3.2 Effect of *C. nucifera* Treatment on Pro-Oxidants

*C. nucifera* juice treatment caused a decrease in level of brain tissue pro-oxidants, H$_2$O$_2$, TBARS and NO. This decrease was dose-dependent or negatively correlated with the dose of treatment. The juice may have enhanced the level of antioxidant thereby indirectly reducing the level of pro-oxidants in brain tissue.

3.3 Effect of *C. nucifera* on Antioxidant/Pro-Oxidant Ratio

The ratio of antioxidant to pro-oxidant revealed CAT to be a predominant antioxidant enzyme against the pro-oxidant H$_2$O$_2$, when treatment dose of 3ml was administered. SOD showed highest level of prevalence against NO at treatment doses 6ml and 9ml.

4. DISCUSSION

Oxidative stress is a serious burden since it has been implicated in several diseases like diabetes, hypertension and neurodegenerative

| Treatments    | SOD (μg/l) | CAT (μg/l) | GPx (μg/l) |
|---------------|------------|------------|------------|
| Normal saline | 221.1±0.4  | 102.0±0.1  | 98.2±0.4   |
| 3ml           | 221.3±0.3  | 227.2±2.1* | 111.4±0.2* |
| 6ml           | 322.1±0.3* | 233.2±1.3* | 117.0±1.3* |
| 9ml           | 446.4±1.2* | 241.1±0.1* | 242.0±0.3* |

Key; *=values statistically significant at 95% confidence interval compared to control

| Treatments    | H$_2$O$_2$ (μg/l) | TBARS (μg/l) | NO (μg/l) |
|---------------|-------------------|--------------|-----------|
| Normal saline | 43.4±0.2          | 37.3±1.0     | 22.1±0.2  |
| 3ml           | 42.2±0.3          | 22.1±0.4*    | 21.3±1.7  |
| 6ml           | 20.3±1.4*         | 17.2±1.1*    | 24.0±0.1* |
| 9ml           | 17.2±1.3*         | 12.4±0.2*    | 24.3±1.1* |

Key; *=values statistically significant at 95% confidence interval compared to control

| Treatment | Antioxidant-Pro-oxidant Ratio |
|-----------|------------------------------|
| Normal saline | SOD[2] CAT[1] GPx[1] H$_2$O$_2$[2] TBARS[2] NO[1] |
| 3ml | CAT[2] SOD[2] GPx[1] H$_2$O$_2$[2] TBARS[1] NO[1] |
| 6ml | SOD[2] CAT[2] GPx[1] NO[1] H$_2$O$_2$[1] TBARS[1] |
| 9ml | SOD[2] GPx[1] CAT[1] NO[2] H$_2$O$_2$[1] TBARS[1] |
disorders [1]. Biomedical studies have been conducted extensively to determine possible prevention and management of oxidative burden [1] [12]. This study revealed further effect of C. nucifera juice on antioxidant enzymes and pro-oxidant generation in brain tissue homogenate of animal model. The phytochemical composition of C. nucifera juice has already been evaluated [11] [12]. From the outcome of this study, the juice significantly increased the level of antioxidants in all sample and this increase was directly proportional to the dose administered. As the dose of treatment was increased, there was a corresponding increase in all antioxidant enzymes. There was a corresponding dose-dependent progressive decrease in pro-oxidants in the brain tissue homogenates. This decrease in pro-oxidant occurred simultaneously with the increase in level of antioxidants. It can be deduced from this study that as the dose of C. nucifera juice treatment is increased, there will be a progressive increase in antioxidant/pro-oxidant ratio. The reason behind these physiologic manifestations may be because of some phytochemicals like flavonoids and other phenolic compounds which have the capacity to boost antioxidant level and suppress pro-oxidant generation in a system [11]. An increase in endogenous antioxidant generation may cause a progressive or proportionate decrease in pro-oxidant species generation in a living system [13]. This can be applied in the management of certain diseases with an oxidative causality [14]. The effect of the juice on antioxidant and pro-oxidant level was predominant when the highest dose in this study was administered, this may infer that at the dose of treatment, the concentration of the active agents in the juice were either just enough or more than enough to cause marked significant changes in the tested biomarkers.

5. CONCLUSION

From the outcome of this study, C. nucifera juice has the ability to enhance antioxidant/pro-oxidant ratio in neural tissues and may have the therapeutic potential in prevention and management of oxidative stress related changes.

DISCLAIMER

The products used for this research are commonly and predominantly use products in our area of research and country. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

ETHICAL APPROVAL

As per international standard or university standard written ethical approval has been collected and preserved by the author(s).

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Ilochi Ogadinma, Arthur Nwafor Chuemere, Ekwemlechewwu, Bassey Samuel. Neuroprotective and antitoxic potential of hydromethanolic extract of Allium cepa in Experimental Rats. European Journal of Pharmaceutical and Medical Research. EJPMR. 2018;5(9):138-143. ISSN 2394-3211 EJPMR
2. Yaep SK, Beh BK, Ali NM, Yusof HM. Antistress and antioxidant effects of virgin coconut oil in vivo. Exp Ther Med. 2015; 9:39.
3. Mariani E, Polidori MC, Cherubini A, Meccoci P. Oxidative stress in brain aging, neurodegenerative and vascular diseases: an overview. J Chromatogr B Analyst Technol Biomed Life Sci. 2005; 827:65-75.
4. Paul CA, Arthur NC, Ogadinma I. Impairment of cardiovascular function indices in male rats induced by aluminium-tainted water: Atherogenic indices and predictor ratio assessment. Discovery. 2018;54(275):442-446
5. Shaw PJ. Molecular and cellular pathways of neurodegeneration in motor neuron disease.J NeurolNeurosurg Psychiatry. 2005;76:1046_57.
6. Khan M, Nasiruddin MU, Rehman, Khurram KW. A study of chemical composition of Cocosnucifera L. (Coconut) water and its usefulness as rehydration fluid. Pakistan Journal of Botany. 2003; 35(5):925-930
7. Joshi SS, Gaikwad VJ, Pati ISS, Makandar AY. Comparative Study of Production Of Wine From Milk And Water Of Cocosnucifera L. International Journal of
Advanced Biotechnology and Research. 2013;4(2):274-279.

8. Intahphuak S, Khonsung P, Panthong A. Anti-inflammatory antipyretic activities of virgin coconut oil. Pharm Biol. 2010; 48:151-7.

9. Shivashankar Sanganamoni, Mallesh S, Vandana K, SrinivasaRao P. Thermal Treatment of Tender Coconut Water-Enzyme Inactivation and Biochemical Characterization International Journal of Current Microbiology and Applied Sciences. 2017; 6(5):2919-2931
Available:https://doi.org/10.20546/ijcmas.2017.6.05.331
ISSN: 2319-7706.

10. Ilochi Ogadinma, Arthur Nwafor Chuemere, Vurey Vitalis (2018) Evaluation of Aromatherapeutic Potential of Allium cepa in Carbon Monoxide-induced Respiratory Tissue Toxicity in Wistar Rats. Asian Journal of Research in Medical and Pharmaceutical Sciences. 2018;4(4):1-7. Article no.AJRIMPS.44256
ISSN: 2457-0745.

11. Ilochi Ogadinma, Kolawole Tolunigba Abisola, Arthur Nwafor Chuemere, Bassey Samuel, Paul Moundipa. Hepatorenoprotective Potential of Cocos nucifera (coconut) Juice in Normal Male Humans. Saudi Journal of Medical and Pharmaceutical Sciences. DOI: 10.21276/sjmps.2018.4.12.4.

12. Sandhya VG, Rajamohan T. Beneficial effects of coconut water feeding on lipid metabolism in cholesterol-fed rats. Journal of Medicinal Food. 2006;9(3):400-407.

13. Ilochi Ogadinma, Daniel Yaro Onoja, Chuemere Arthur Nwafor. Neurodynamic Influence of Hydrocortisone on Rhinencephalic and Telencephalic Brain Areas, International Neuropsychiatric Disease Journal. 2019;13(2):1-7. Article no.INDJ.52924
ISSN: 2321-7235
NLM ID: 101632319.

14. Ekwem Ikechukwu, Ilochi Ogadinma, Arthur Nwafor Chuemere. Epidemiological investigation, monitoring and surveillance; strategies in environmental health sustenance. Submitted to Asian Journal of Advanced Research and Reports. 2019; 5(2):1-9. Article no: AJARR.45207.(2019)