Relationship between plasma asymmetric dimethylarginine and nitric oxide levels affects aerobic exercise training-induced reduction of arterial stiffness in middle-aged and older adults

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INTRODUCTION

Alterations in arterial structure and function occur during aging in healthy individuals. An aging-induced decrease in endothelial function leads to deterioration of arterial stiffness. This functional deterioration impairs the conduit and buffering functions of arteries, leading to several pathological conditions including hypertension, atherosclerosis, congestive heart failure, stroke, and aortic root regeneration. Several studies have shown that arterial stiffness is lower in physically active individuals than in sedentary individuals. Furthermore, aerobic exercise training (AT) reduces arterial stiffness occurring with advancing age. Thus, AT prevents or improves arterial stiffness.

Nitric oxide (NO) is produced from l-arginine by endothelial NO synthase (eNOS). NO causes vasodilation and inhibits the development of arteriosclerosis and atherosclerosis. Asymmetric dimethylarginine (ADMA), an endogenous inhibitor of NOS, is associated with impaired endothelial function in humans. ADMA impairs NO bioavailability in older adults, which may result in enhanced arterial stiffness.

AT ameliorates endothelial dysfunction in older adults, and our previous studies reported that endurance exercise training increased arterial eNOS and phosphorylation, eNOS mRNA, and nitrite/nitrate (NOx) levels with a concomitant improvement of endothelial function in aged rodent models. Moreover, in middle-aged and older male and female adults, moderate AT elevated plasma NOx levels and reduced arterial stiffness. Similarly, AT led to decreases in circulating ADMA levels in patients with metabolic syndrome, type 1 diabetes mellitus, coronary and peripheral arterial diseases, obesity, and postmenopausal women. Additionally, the AT-induced decrease in circulating ADMA levels is
correlated with an increase in carotid arterial compliance in postmenopausal women. Therefore, the decrease in circulating ADMA levels by AT might contribute to the improvement of arterial stiffness in postmenopausal women. However, whether the AT-induced decrease in circulating ADMA levels is related to changes in circulating NOx levels remains unclear. The ratio of plasma NOx levels to ADMA levels decreases with advancing age, and the NOx/ADMA ratio is positively correlated with popliteal FMD in healthy men. Therefore, changes in the NOx/ADMA ratio by AT may be associated with a decrease in arterial stiffness in healthy middle-aged and older male and female adults.

Herein, we aimed to identify whether the relationship between plasma ADMA and NOx levels affected the AT-induced reduction in arterial stiffness. For this, we measured plasma ADMA levels, NOx levels, and arterial stiffness in middle-aged and older male and female adults. Additionally, we examined whether the effect of AT on circulating ADMA levels differed according to sex.

Methods

Subjects

Thirty-one healthy middle-aged and older subjects (total, n = 31, 66.4 ± 1.3 years [range, 53–79 years]; male, n = 13, 69.8 ± 1.6 years [range, 61–79 years]; female, n = 18, 63.9 ± 1.7 years [range, 53–79 years]) volunteered to participate in this study. All volunteers provided written informed consent before participating in the study, which was approved by the Ethics Committee of Ritsumeikan University and was conducted in accordance with the Declaration of Helsinki. This study was registered in the University Hospital Medical Information Network Clinical Trials Registry (UMIN-CTR, UMIN000035520). All subjects were free from overt signs and symptoms of chronic disease and did not smoke. Subjects taking medications such as anti-hyperlipidemics, anti-hypertensives, or anti-hyperglycemics as well as those with a history of stroke, diabetes, hypertension, hyperlipidemia, cardiac disease, chronic renal failure, and mental disorders were excluded from the study. None of the subjects regularly performed resistance and aerobic exercise. The subjects in this study drank very little alcohol. Since estrogen hormones influence arterial function, we recruited postmenopausal women. Subjects were randomly divided into 2 groups: a training group (n = 16 [male = 6/female = 10], 64.8 ± 2.0 years) and a control group (n = 15 [male = 7/female = 8], 68.1 ± 1.6 years).

Experimental design

For all subjects, peak oxygen uptake (VO2peak), body weight, body fat, height, resting systolic blood pressure (SBP), resting diastolic blood pressure (DBP), resting heart rate (HR), resting plasma ADMA concentration, resting plasma NOx concentration, as well as serum concentrations of total cholesterol, high-density lipoprotein (HDL) cholesterol, and triglycerides were measured at the beginning and end of the experiment. Carotid β-stiffness was examined as an index of arterial stiffness. Before the subjects were tested, they sat quietly for 30 min. Resting brachial SBP, DBP, and HR were measured in duplicate in the supine position using a vascular testing device (OMRON COLIN Co., Tokyo, Japan). At the beginning and end of the study period, fasting blood samples were drawn following at least 48 h of rest after the last exercise training session. All subjects were instructed not to eat or drink fluids other than water for at least 12 h prior to blood sampling. In addition, we confirmed that participants did not consume any dietary sources of NOx over the 24 h prior to testing in either group since NOx can be affected by diet. Thus, both acute effects from the most recent bout of exercise and oral sources of NOx other than NO should be avoided. Serum and plasma samples were immediately centrifuged (1500 × g, 15 min, 4°C). Blood samples were stored at −80°C until use. Room temperature was maintained at 24°C throughout the experiments.

Exercise intervention

Subjects attended the AT program, which consisted of cycling on a leg ergometer at 60%–70% VO2peak for 45 min/day, 3 days/week for 8 weeks. For the first two periods of the exercise program, subjects performed at 50% VO2peak for 40 min. The warm-up and cool-down periods consisted of 5 min of cycling at a 40% VO2peak. Exercise compliance was monitored carefully under direct supervision. Additionally, sedentary control subjects were encouraged to rest during the experimental period.

Measurement of VO2peak

The VO2peak was measured using an incremental cycle exercise test on a cycle ergometer (MINATO, AE-310SRD, Osaka, Japan). The incremental cycle exercise began at a work rate of 60 W (30–90 W) for men and 30 W (0–60 W) for women. Power output was increased by 15 W/min until the subjects could not maintain a fixed pedaling frequency of 60 rpm. The subjects were encouraged to exercise to maximum intensity during the ergometer test. HR and rating of perceived exertion (RPE) were monitored minute by minute during the exercise. RPE was obtained using a modified Borg scale. VO2 was monitored during the last 30 s of each period of the increased work rate. The highest value of VO2 during the exercise test was designated as VO2peak if three out of four of the following criteria were met: (I) a plateau in VO2 with an increase in external work, (II) maximal respiratory exchange ratio ≥ 1.1, (III) maximal HR ≥ 90% of the age-predicted maximum (208 − 0.7 × age), and (IV) RPE ≥ 18.

Measurement of the carotid β-stiffness index

Carotid β-stiffness was evaluated as an indicator of arterial stiffness. A combination of ultrasound imaging of the pulsatile common carotid artery and simultaneous application of tonometrically obtained arterial pressure from the contralateral carotid artery allowed noninvasive determination of arterial compliance. The carotid artery diameter was measured from images obtained using an ultrasound system equipped with a high-resolution linear array transducer. A
longitudinal image of the cephalic portion of the common carotid artery was acquired 1–2 cm proximal to the carotid bulb. All image analyses were performed by the same investigator.

Pressure waveforms and amplitudes were obtained from the common carotid artery using a pencil-shaped probe with a high-fidelity strain gauge transducer (SPT-301; Millar Instruments, Houston, TX, USA). Because baseline blood pressure levels are subjected to hold-down forces, the pressure signal obtained via tonometry was calibrated by equating the carotid mean arterial blood pressure and DBP to brachial artery values. The carotid \( \beta \)-stiffness index was calculated using the equation \( \ln(P1/P0)\left/\left[(D1 - D0)/D0\right] \right. \), where \( D1 \) and \( D0 \) are the maximum (systolic) and minimum (diastolic) diameters and \( P1 \) and \( P0 \) are the highest (systolic) and lowest (diastolic) blood pressures, respectively.

### Measurement of plasma NOx levels

NOx levels in the plasma were measured using the Griess assay (R&D Systems, Minneapolis, MN, USA) according to the manufacturer’s protocol. All samples were assayed in duplicate. The optical density at 540 nm was measured using a microplate reader (xMark microplate spectrophotometer; Bio-Rad Laboratories, Hercules, CA, USA). The day-to-day coefficient of variation of plasma NOx levels was 1.0 ± 0.3%.

### Measurement of plasma ADMA levels

ADMA levels in the plasma were measured using an enzyme-linked immunosorbent assay (ELISA; Immundiagnostik AG, Bensheim, Germany) according to the manufacturer’s protocol. All samples were assayed in duplicate. The optical density at 540 nm was quantified using a microplate reader (xMark microplate spectrophotometer; Bio-Rad Laboratories, Hercules, CA, USA). The day-to-day coefficient of variation of plasma ADMA levels was 0.6 ± 0.2%.

### Measurements of serum cholesterol and triglyceride levels

Fasting serum levels of total cholesterol, high-density lipoprotein (HDL) cholesterol, and triglycerides were determined using standard enzymatic techniques.

### Statistical analysis

Values are expressed as mean ± standard errors (SE). The differences between the groups and the two time points were assessed by a two-way repeated-measure analysis of variance followed by Fisher’s post hoc test, which was applied when a measurement was significantly different. The relationships between AT-induced changes in plasma ADMA levels and plasma NOx levels or carotid \( \beta \)-stiffness as well as changes in the NOx/ADMA ratio and carotid \( \beta \)-stiffness were determined using the Pearson correlation coefficient. Sex differences in the changes in these parameters before and after the AT intervention were compared using a paired Student’s t-test. Statistical significance was set at \( P < 0.05 \). All statistical analyses were performed using StatView (5.0, SAS Institute, Tokyo, Japan).

### RESULTS

#### Comparison of baseline and endpoint characteristics between the training and control groups

Before AT, there were no significant differences in VO_{2peak} between the training and control groups. However, there was a significant interaction between the group with intervention on VO_{2peak} (\( P < 0.05 \), Table 1). Specifically, in the training group, VO_{2peak} significantly increased after AT intervention (\( P < 0.05 \), Table 1). Although there was no significant difference in the carotid \( \beta \)-stiffness between the training and control groups before the AT, there was a significant interaction of the group with intervention on carotid \( \beta \)-stiffness (\( P < 0.01 \), Fig 1-A). Carotid \( \beta \)-stiffness significantly decreased after AT intervention (\( P < 0.01 \), Fig. 1-A). However, no significant differences between the groups or

### Table 1. Comparison of characteristics between training and control groups

|                     | Control | Post | Training | Group | Time | Interaction |
|---------------------|---------|------|----------|-------|------|-------------|
| Age, years          | 68.1±6.6 | 64.8±2.0  |          |       |      |             |
| Height, cm          | 160.6±2.5 | 159.0±2.3  | 159.1±2.2 | 0.548  | 0.978 | 0.940       |
| Body weight, kg     | 70.0±3.3  | 62.2±3.0  | 62.2±3.0 | 0.102  | 0.986 | 0.981       |
| BMI, kg/m²          | 22.0±1.0  | 24.6±1.1  | 24.6±1.1 | 0.016  | 0.999 | 0.949       |
| HR, bpm             | 60.0±2.0  | 59.6±2.2  | 59.6±2.2 | 0.077  | 0.975 | 0.590       |
| SBP, mmHg           | 87.7±3.6  | 118.8±3.8 | 118.8±3.8 | 0.047  | 0.952 | 0.493       |
| DBP, mmHg           | 79.6±2.0  | 73.9±2.9  | 73.9±2.9 | 0.637  | 0.237 | 0.730       |
| Total cholesterol, mg/dl | 219.6±7.7 | 219.0±8.5 | 0.333  | 0.765 | 0.367       |
| HDL cholesterol, mg/dl | 79.6±4.5  | 68.9±5.5  | 0.067  | 0.901 | 0.927       |
| Triglycerides, mg/dl | 111.7±25.3 | 123.1±18.0 | 0.417  | 0.727 | 0.707       |
| VO_{2peak}, ml/kg/min | 26.7±1.7 | 24.0±1.3  | 29.9±1.5 | 0.791  | 0.065 | 0.043       |

BMI, body mass index; HR, heart rate; SBP, systolic blood pressure; DBP, diastolic blood pressure; HDL, high-density lipoprotein; VO_{2peak}, peak oxygen uptake

The statistical analysis was used by 2-way ANOVA. The P values of Group and Time were the main effect while the P values of Interaction were Group × Time interaction. Values are mean and SE. * \( P < 0.05 \) vs. Training-Pre.
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Comparison of plasma ADMA and NOx levels and NOx/ADMA ratio between the training and control groups

Before AT, there were no significant differences in plasma ADMA or NOx levels and NOx/ADMA ratio between the training and control groups. There was significant interaction between plasma NOx levels in the intervention group (Fig. 1-B, P < 0.01). After AT, plasma NOx levels were significantly higher in the training group than in the control group (Fig. 1-B, P < 0.01). The interaction between groups and time points was significant for plasma ADMA levels (Fig. 1-C, P < 0.05). After AT, plasma ADMA concentrations were significantly lower in the training group than in the control group (Fig. 1-C, P < 0.01). Notably, there was a significant interaction between the intervention group and the NOx/ADMA ratio (Fig. 1-D, P < 0.01). After AT, the NOx/ADMA ratio was significantly higher in the training group than in the control group (Fig. 1-D, P < 0.01).

Relationships among plasma ADMA and NOx levels, NOx/ADMA ratio, and carotid β-stiffness

Changes in plasma ADMA levels were negatively correlated with changes in plasma NOx levels (Fig. 2-A, r = -0.414, P < 0.05). Moreover, there was a negative correlation between changes in plasma NOx levels and carotid β-stiffness (Fig. 2-B, r = -0.493, P < 0.01). Furthermore, changes in the NOx/ADMA ratio were negatively correlated with changes in carotid β-stiffness (Fig. 2-C, r = -0.514, P < 0.01).

Sex differences among changes in carotid β-stiffness, plasma ADMA and NOx levels, and NOx/ADMA ratio

There was no sex difference with regard to changes in carotid β-stiffness between male and female subjects (Fig. 3-A, male: -2.400 ± 0.411 vs. female: -2.390 ± 0.254). In addition, no sex differences were observed in the changes in plasma ADMA and NOx levels or in NOx/ADMA ratio.
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Figure 3. Comparisons of sex differences in changes in carotid β-stiffness, plasma nitrite/nitrate (NOx) levels, asymmetric dimethylarginine (ADMA) levels, and NOx/ADMA ratio.

Comparison of sex differences in changes in carotid β-stiffness (A), plasma nitrite/nitrate (NOx: B) levels, asymmetric dimethylarginine (ADMA: C) levels, and NOx/ADMA ratio (D) in middle-aged and older adults before and after 8 weeks of aerobic exercise (training group, n = 16) or sedentary (control group, n = 15). Data are expressed as the mean ± SE.

DISCUSSION

The present study investigated the effects of AT on circulating ADMA levels in middle-aged and older male and female adults before and after an 8-week AT. After AT intervention, plasma ADMA levels and arterial stiffness decreased, and concomitantly, plasma NOx levels and NOx/ADMA ratio increased. Additionally, we did not observe a sex difference in AT-induced changes in circulating ADMA levels and NOx/ADMA ratio between male and female subjects (Fig. 3-B, NOx: male: 24.412 ± 7.997 vs. female: 30.203 ± 5.810, Fig. 3-C, ADMA: male: -0.138 ± 0.024 vs. female: -0.142 ± 0.029, Fig. 3-D, NOx/ADMA ratio: male: 67.831 ± 16.707 vs. female: 87.964 ± 17.026).

circulating ADMA levels in postmenopausal women as well as in middle-aged and older male adults. Additionally, the amount of change in circulating ADMA levels by AT did not differ between the sexes. Thus, the data indicate that there is no sex difference in the effect of AT on circulating ADMA levels. Moreover, low circulating ADMA levels have been independently associated with increased FMD in subjects with low cardiovascular risk33 or decreased carotid-femoral pulse wave velocities in prediabetic individuals32. Accordingly, the decreased circulating ADMA levels by AT may be a novel biomarker, regardless of sex, for the prevention or treatment of arterial stiffness associated with impairments in NO production.

We demonstrated that an 8-week AT consisting of cycling on a leg ergometer at 60%–70% VO2peak for 45 min, 3 days/week, decreased plasma ADMA levels in middle-aged and older male and female adults. Similarly, 12 weeks of AT consisting of cycling at 65%–80% of maximal HR 40–60 min/day, 3–6 days/week, decreases circulating ADMA levels in postmenopausal women29. In addition, in patients with coronary artery disease, 12 weeks of endurance training consisting of running or walking for 30–60 min, 3–5 days/week, decreases circulating ADMA levels23. Moreover, in patients with type 1 diabetes mellitus, circulating ADMA levels were decreased after a 4-month-long aerobic exercise program of cycling on a leg ergometer at 60%–70% maximum HR for 40 min, 3 days/week22. Thus, AT may be effective in reducing plasma ADMA levels and arterial stiffness in both healthy and at-risk subjects.

After AT intervention, plasma ADMA levels decreased, and concomitantly, plasma NOx levels and NOx/ADMA ratio were increased in middle-aged and older adults. The mechanism underlying the effects of AT on plasma ADMA levels is unclear. ADMA is selectively degraded by dimethylarginine dimethylaminohydrolase (DDAH) enzymes33. DDAH exists in two isoforms. In an in vitro study using siRNA, individual silencing of either DDAH-1 or DDAH-2 reduced endothelial NO production by 31% and 48%, respectively33. These results indicate that both DDAH-1 and DDAH-2 play important roles in regulating endothelial NO production. Additionally, intracellular reactive oxygen species (ROS) can stimulate ADMA production or inhibit ADMA degradation, thus resulting in the accumulation of ADMA in endothelial cells34. Furthermore, ROS regulate the ADMA/DDAH-2/eNOS/NO pathway in endothelial cells35. AT increases eNOS phosphorylation and NOx levels in the aorta of aged rodent models17,18. Therefore, we speculate that AT could be associated with upregulated activity and protein expression of DDAH accompanied by a decrease in ADMA concentrations and an increase in...
NO metabolites, thereby reducing arterial stiffness. Further studies are needed to examine the effects of AT on DDAH activity and protein expression in the arterial wall to clarify the mechanistic association between AT-induced changes in ADMA levels and improvement in arterial stiffness.

In conclusion, we investigated the effects of AT on ADMA levels in middle-aged and older male and female adults before and after 8 weeks of AT. The training effect on plasma ADMA levels was related to that of plasma NOx levels, while the training effect on NOx/ADMA ratio was related to that of arterial stiffness. Thus, AT-induced reduction of plasma ADMA levels, regardless of sex, was related to an AT-induced increase in plasma NOx levels. The balance between ADMA and NO production may be associated with an underlying mechanism of AT-induced improvement in arterial stiffness in middle-aged and older men and women.

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