Introduction

The treatment of high metal content liquid and solid residues has been gaining importance worldwide, as they represent a major threat to ecosystems and human health, due to its accumulative capacity [1], emerging then the imminent need of developing efficient methods to minimize or even eliminate metals prior to its discharge to the environment. Although there are treatments currently available to reduce the metal content from solid and liquid wastes, they consider the utilization of chemical compounds that generate more residues [2]. Hence, it is important to develop more eco-friendly methods to treat and/or recycle such materials. Thus, biotechnological approaches are an innovative and promising technology applicable for the removal of heavy metals from solid and liquid residues, that has been proved as an effective economically feasible technology [3,4], due to their low costs and higher efficiency at low metal concentrations, where physicochemical removal methods fail [5,6].

It has been previously reported that Bacillus megaterium possesses high levels of resistance to hostile conditions [7,8], including the exposition to metals like Hg and Ni [9,10], and some B. megaterium strains also present the demonstrated ability to remove metals from solid and liquid residues [10-12], establishing their potential to treat phosphogypsum waste [13], and spent catalysts [14,15]. Specifically, B. megaterium strain MNSH1-9K-1 presents the ability to resist up to 200 ppm of each Ni and V contained in a liquid mixture, being able to remove up to 15.38 ± 1.30 ppm of Ni contained in the former liquid medium [12].

Metals like Ni, Al, Mo, and V are commonly present in spent catalysts [14], being Ni and Al often found as water pollutants too, by reason of being used in numerous industrial processes. Furthermore, Ni is considered a highly valuable metal for modern industry, mainly due to its expanding need as a crucial component of stainless steel [16]. Although Ni has biological activity in bacteria at nanomolar concentrations [17-19], it is also considered as a highly toxic metal [20], by the aim of causing the elevated production of reactive oxygen species and DNA damage [21]. In the case of Al, its embryotoxic effects in animal models and its neurotoxic effects in humans have been demonstrated, suggesting that this metal exposition may cause specific encephalopathy with a dementia syndrome [22].

Thus, the current study analyzed the resistance of Bacillus megaterium strain MNSH1-9K-1 to Al, Ni, V, and Mo, metals that are commonly found in high metal content residues, and also to evaluate MNSH1-9K-1 Ni removal potential from liquid media at different concentrations. The results presented in this manuscript may contribute to the knowledge of B. megaterium potential application for high metal content residues biotreatment.
incubated for 24-48 h at 37 °C to perform colony counting. Followed by plating on LB solid medium. Finally, LB plates were prepared in phosphate-buffered saline (PBS) buffer [23], collected and was added to the cylindrical vial, and the supernatant was cooled 15 min. Afterwards, 20 ml of deionized water were included, as controls. Viability was assessed by taking a 100 μl aliquot from each culture, and serial 10-fold dilutions were prepared in phosphate-buffered saline (PBS) buffer [23], followed by plating on LB solid medium. Finally, LB plates were incubated for 24-48 h at 37 °C to perform colony counting.

**Materials and Methods**

**Bacterial strain and growth conditions**

*Bacillus megaterium* strain MNSH1–9K–1 (GenBank accession number KM654562.1) was used for this study, which isolation from a high metal content site has been previously described [15]. LB liquid medium was used for microbial growth at 200 rpm and 37°C. Inoculums were prepared overnight, and microbial density was adjusted to an Optical Density at a wavelength of 600 nanometers (O.D.600 nm) = 0.1 to begin experimentation.

**Metal resistance**

Cultures were grown for 48 hours in 125 ml Erlenmeyer flasks containing 10 ml of LB liquid medium and different concentrations of Al, Mo, V, or Ni, provided as AlCl₃, (NH₄)₆Mo₇O₂₄·4H₂O, NaVO₃, and Ni(NO₃)₂·6H₂O, respectively, as specified for each experiment. Also, samples without metals were included, as controls. Viability was assessed by taking a 100 μl aliquot from each culture, and serial 10-fold dilutions were prepared in phosphate-buffered saline (PBS) buffer [23], followed by plating on LB solid medium. Finally, LB plates were incubated for 24-48 h at 37 °C to perform colony counting.

**Ni removal from liquid medium**

Experimental sets were prepared in 125 ml Erlenmeyer flasks containing 10 ml of LB liquid medium supplemented with different Ni concentrations, to a 10 ml final volume. Also, samples without metals were included, as controls. After 24 hours of growth at 37°C and 200 rpm, samples were centrifuged at 5000 rpm for 10 min in order to separate cells from the supernatant, biomass was discarded and the liquid phase of each sample was filtered using a cellulose acetate syringe filter (Alltech, Deerfield, IL, USA) to remove any cell debris remaining in the samples. Subsequently, the samples were acid digested. For this purpose, 1 ml samples were placed in cylindrical silicon carbide vials, and 6 ml of concentrated HNO₃ and 2 ml of concentrated HCl were added, and samples were digested in a microwave reaction system (Multiwave PRO, Anton Paar), using an HF100 rotor. Digestion conditions were: 600 W for 6 vessels, 40 bar, 210-240°C, with pRate of 0.3 bar sec⁻¹, ramp 15 min, hold 15 min, and cooling 15 min. Afterwards, 20 ml of deionized water was added to the cylindrical vial, and the supernatant was collected and filled up to 100 ml with deionized water. Metal analysis was performed at 231.604 nm by Inductively Coupled Plasma Optical Emission Spectrometry (ICP−OES; Varian Model 710–ES). To determine Ni removal by *B. megaterium*, metal concentration remaining in liquid medium was calculated based on a calibration curve covering 0.1-10 mg kg⁻¹, using a commercial standard (High-Purity, cat. # ICP–200–7–6) [12].

**Statistical analyses**

Basic statistical parameters and analyses of variance (ANOVA) were performed using the commercial statistical software OriginPro 9.0. Differences with $P$ values of $\leq 0.05$ were considered statistically significant.

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**Results and Discussion**

It has been previously reported that members of the genus *Bacillus* present high heavy metal resistance properties [24–26]. Particularly, it was reported that *Bacillus megaterium* strain MNSH1–9K–1 possesses the ability to remove 149.5 mg/kg of Ni, and 127.5 mg/kg of V from a spent catalyst at a 16% (w/v) of pulp density [15]. Besides, MNSH1–9K–1 may tolerate up to 200 ppm of each Ni and V contained together in liquid medium [12], and removed up to 15.38 mg/kg of Ni from the latter system. Thus, in order to evaluate the ability of *B. megaterium* MNSH1–9K–1 to resist diverse concentrations of metals commonly encountered in solid and liquid high metal content residues, namely Al, Mo, Ni, and V, the strain was subjected to grow for 24 h in the presence of each metal at the different concentrations specified for each case, and cell viability was evaluated following the protocol described in Materials and Methods. As it can be observed in figure 1, *B. megaterium* MNSH1–9K–1 presented metal resistance following the ascendant order of Ni < Al < V < Mo, dictated by the respective median lethal doses of 87 ± 3, 248 ± 1, 1925 ± 247, and 10,400 ± 566 ppms calculated from figure 1A to 1D. Furthermore, Minimum Inhibitory Concentrations (MIC) were also calculated for each metal, and results are shown in table 1, where the metal resistance pattern Ni < Al < V < Mo was corroborated. In the case of Mo, it was not possible to obtain a specific value for Mo, because it was experimentally impossible to dissolve more (NH₄)₆Mo₇O₂₄·4H₂O to obtain higher concentrations and reach the MIC value.

**Table 1:** Minimum Inhibitory Concentrations (MIC) for the different elements tested.

| Metal | MIC (ppm) |
|-------|-----------|
| Ni    | 240       |
| V     | 35,000    |
| Al    | 2000      |
| Mo    | > 270,000 |

**Figure 1:** Resistance of *B. megaterium* MNSH1–9K–1 to Al (A), Mo (B), Ni (C), and V (D). The strain was grown in LB liquid medium in the presence of each metal at different concentrations, and the results were normalized to those from the control (without metal). Data are presented as averages ± standard deviations (n = 2).
The results are in accordance to previous studies, where it has been demonstrated the elevated toxicity of Ni above other metals [21, 27, 28], since this metal has been directly related to alterations in DNA repair mechanisms, epigenetic effects, and carcino genesis [29], besides the generation of reactive oxygen species (ROS), which has been broadly related to metal toxicity [30, 31]. It is interesting to note that an enhanced growth was observed when cells were subjected to 25, 75, and 200 ppm of Al. In this respect, it has been reported that many metals may have biological activities [32], participating in chemical processes, cell structures, and biological functions [33], so it may be the case that Al at these three concentrations is promoting B. megaterium growth, possibly by participating somehow in an up to date non specified metabolic function in this microorganism.

Due to Ni enhanced toxicity to living organisms, and hence the need of developing efficient approaches for its removal, B. megaterium MNSH1-9K-1 Ni resistance and removal abilities were studied in further detail. First, MNSH1-9K-1 growth kinetics were determined in the presence of five different Ni concentrations, comparing them with the growth behavior without the metal. To this end, the strain was subjected to grow for 24 hours in the presence of 50, 100, 150, 200, and 400 ppm of Ni at 37 °C and 200 rpm in 125 ml Erlenmeyer flasks containing LB liquid medium at a final volume of 10 mL, and the results are shown in figure 2. Microorganism growth was observed very similar to the control when 50 and 100 ppm of Ni were added to the medium. However, a lag phase of approximately 2 hours was observed when the strain was exposed to 150 ppm of the metal, although a subsequent growth recovery was achieved. Contrastingly, the strain was only able to tolerate 200 ppm of the metal, but not capable of growing under the stress generated by this Ni concentration, and a 50-fold decrease in cell viability was observed after 24 h of growth. Furthermore, B. megaterium MNSH1-9K-1 had an evident viability loss since the beginning of the experiment when exposed to 400 ppm of Ni, reaching up to a 1600-fold decrement in cell survival.

Previous studies show that Bacillus megaterium strain MNSH1-9K-1 possesses the ability to tolerate up to 200 ppm of Ni contained in a liquid Ni-V mixture [12]. The results presented here demonstrate the ability of MNSH1-9K-1 to grow in the presence of up to 150 ppm of Ni, and support the fact of this strain Ni tolerance at 200 ppm, evidencing also the elevated viability loss at 400 ppm of the metal.

It has been documented that the B. megaterium genome contains several open reading frames of genes involved in stress responses, including oxidative stress, osmotic stress, heat shock, and detoxification [34], which activations, at least in its most studied partner B. subtilis, overlap between general stress responses and more specific pathways, and confer the microorganism with efficient adaptation mechanisms [35]. In this regard, extracytoplasmic sigma (σ) factors regulate several overlapped stress responses. For example, σW regulon is induced by various cell wall antibiotics, alkaline shock, and other stresses affecting the cell envelope, and σE regulon activation may confer advantages to diverse stress factors, including high salinity, ethanol, heat, acid, phosphate starvation, superoxide stress, and exposure to cell wall antibiotics such as bacitracin, vancomycin, and cationic antimicrobial peptides [36]. In the case of B. megaterium, it has been reported that strain uyuni S29 presents a high capability to produce poly [(R)-3-hydroxybutyrate] (PHB) at excessive salinity conditions [37]. Hence, the differences in cell survival when B. megaterium is exposed to diverse Ni concentrations may be due to the diverse molecular mechanisms activated under the stress caused by each Ni concentration. Particularly, the strain was able to adapt at 150 ppm of Ni and resume growth, being evident that this adaptation ability is exceeded at 200 and 400 ppm of Ni under the conditions tested.

Once B. megaterium MNSH1-9K-1 Ni resistance was determined, analyses were performed to evaluate the ability of this microorganism to remove Ni from liquid medium at the same concentrations tested before (figure 3), observing that Ni removal was significantly obtained when the strain was subjected to grow in the presence of 200 ppm of the metal. This removal was evident since hour 2, and the highest Ni removal of up to 26 ppm was observed from hour 6 and beyond.
Reports have previously shown that *B. megaterium* possesses high metal sorption capacity [1], and also Ni specific response genes like *ncCA* and *smiAB* have been correlated to Ni removal by MNSH1-9K-1 [12]. Hence, as the uppermost Ni removal was accomplished at 200 ppm of Ni, even though cell population was 100–times less than in 50 to 150 ppm, the results strongly suggest that the removal of this metal may be accomplished by metabolism components, mainly activated by the stress generated by 200 ppm of Ni.

To date, little is known about the molecular mechanisms involved in the removal of Ni or other toxic metals from high content metal residues [12]. In consequence, transcriptome, proteome, and metabolome approaches are necessary perspectives to further understand the mechanisms involved in *B. megaterium* Ni removal capability, to enhance and further promote the use of this microorganism for the biotreatment of high Ni content residues.

**Conclusion**

The current study shows that *B. megaterium* MNSH1-9K-1 presents a metal resistance pattern of Ni < Al < V < Mo. Besides, the results strongly suggest that this microorganism presents the potential to be used for the biotreatment of high Ni content residues.

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