Functional Marker Assisted Improvement of Stable Cytoplasmic Male Sterile Lines of Rice for Bacterial Blight Resistance

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Bacterial blight (BB), caused by Xanthomonas oryzae pv. oryzae is one among the major diseases in rice, which in severe condition cause losses up to 60% in total yield. Marker assisted pyramiding of three broad spectrum BB resistance genes (xa5, xa13, and Xa21) in prominent rice varieties is the most economical and effective strategy for the management of the BB disease. We report here the pyramiding of three genes (xa5, xa13, and Xa21) in maintainer lines (CO 2B, CO 23B, and CO 24B) of three promising wild abortive cytoplasmic male sterile lines (CO 2A, CO 23A, and CO 24A) through functional markers assisted back cross breeding. IRBB60 with xa5, xa13, and Xa21 genes is used as a donor parent. BC2F1 and BC2F2 generations from a cross of CO 2B, CO 23B, and CO 24B with IRBB60 were evaluated for bacterial blight and non-fertility restoration. In BC2F1, plants with all three resistance genes (xa5, xa13, and Xa21) and high parent genome recovery was identified. In BC2F2, plants with all resistance genes and without fertility restorer (Rf3 and Rf4) were selected. Based on agronomic traits, BB resistance and maintenance of sterility, two plants each in CO 2B × IRBB60, CO 24B × IRBB60 and one plant in CO 23B × IRBB60 combinations were identified. The identified lines were crossed with respective male sterile lines for conversion of improved B line into CMS line through back-crossing, in addition to selfing. The plants with high recurrent genome and phenotypically similar to parental lines and sterile are being used for the hybrid rice development program. Currently, using these lines (improved CMS line), test crosses were made to develop new rice hybrids. Hybrids combinations viz., CO 23A × AD08009R and CO 24A × IET20898R were found to be stable at different locations with high yield. The R line used in this study has been introgressed with xa5, xa13, and Xa21 genes in a separate breeding program. These new hybrids with resistance against bacterial blight will increase the crop production at BB environment.

Keywords: bacterial blight resistance, functional markers, hybrid rice, cytoplasmic male sterility, marker-assisted backcross breeding, foreground selection, background selection, fertility restoration
INTRODUCTION

Rice (Oryza sativa L.) is one of the staple food crops and grown in a wide range of climatic conditions. Blight is an important bacterial disease in rice (Gnanamanickam et al., 1999), hence developing a BB resistant rice variety can give real time solution to the farmers. To develop resistant varieties, functional markers have a selective advantage over indirect selection using molecular markers (Andersen and Lübbertstedt, 2003; Varshney et al., 2005; Iyer-Pascuzzi and Mccouch, 2007; Ingvarsdson et al., 2008). Due to the lack of diverse parental lines with desirable traits and multiple resistances, hybrid rice production in India depends mostly on parental lines developed in China and IRRI. These outsourced lines are not well adapted to the Indian climate, resulting in poor grain quality. Therefore, we have decided to improve native CMS and maintainer line for biotic and abiotic stresses with higher production and productivity focusing on hybrid rice development. Currently, 10 CMS lines of native origin were developed at our University using Wild abortive (WA) cytoplasmic source. Quite a good number of stable hybrids were developed with these native CMS lines. Two hybrids, CORH 3 and CORH 4 were released recently from our center. CORH 4 is very popular among the Southern part of India. Low adaptability of earlier released hybrids is due to poor seed setting and grain quality. The hybrids developed using CO 23A overcomes the problem of low adaptability and poor grain setting shown by the earlier released hybrids, but the hybrids and their parental lines are more prone to bacterial blight disease.

Gene pyramid work has been successfully employed in several crops for agronomically important traits. Basavaraj et al. (2010) had successfully introgressed two bacterial blight resistance genes (xa13 and Xa21) into parental lines Pusa 6B and PRR 78 of Pusa RH10 hybrid. The elite restorer line of hybrid rice, Minghui 63, which become more susceptible to bacterial blight was improved for resistance through introgression of Xa21 gene within 3 generations of back-crossing (Chen and Ronald, 1999). A novel bacterial blight resistance gene Xa23, identified in wild rice species Oryza rufipogon was reported to confer resistance to 20 races of bacterial blight from China, India and Philippines right through all the growth stages of rice (Zhang et al., 2002; Zhang, 2009). The restorer lines Minghui 63, Y1671 and YR293 improved with the introgression of Xa23 gene revealed to have a wide range of resistance to bacterial blight (Zhou et al., 2011). Recently, more reliable Marker assisted selection is gaining significance in the field of disease resistance breeding for hybrid rice production. The elite restorer line RPHR 1005 was improved for BB and blast resistance with the aid of gene specific markers (Kumar et al., 2016).

Promising BB resistance genes viz., Xa21, xa13, Xa4, and xa5 with substantial resistance was introgressed into the background of restorer (KMR 3 and PRR 78) and maintainer lines (IR 58025B and Pusa 6B) through MAS (Shanti et al., 2010). Chen et al. (2000) successfully incorporated BB resistance genes by MAS.

Successful enhancement of resistance to bacterial blight in an elite restorer line Hau 1035 was accomplished by pyramiding Xa23, Xa22, Xa21, and Xa7 resistance genes by means of MAS (Huang et al., 2012). Pusa RH 1000, super fine rice hybrid and its parental lines- Pusa6B and PRR78 were improved for resistance to BB (xa13 and Xa21) along with agronomic, grain and cooking quality traits through MAS. Similarly, the maintainer line (DRR I7B) and restorer line (RPHR-1005) were improved for BB resistance (Xa21) and blast resistance (Pi54) through MAS (Balachiranjeevi et al., 2015; Kumar et al., 2016). Based on the earlier reports and our hypothesis, we selected two BB resistant recessive genes (xa5 and xa13) and one dominant gene (Xa21). Therefore, incorporation of three BB resistant genes (xa5, xa13, and Xa21) combination was taken up in the WA-CMS maintainer lines (CO 2B, CO 23B, and CO 24B) through back-crossing without fertility restorer genes (Rf3 and Rf4) of WA-CMS system using functional markers.

MATERIALS AND METHODS

Development of Plant Population

The experimental material consisted of CO 2A, CO 23A, CO 24A of WA cytoplasmic source and its maintainer, CO 2B, CO 23B, and CO 24B, as recurrent parents. IRBB 60 (BB donor line), IR 24 (susceptible check) and identified BB resistant restorer lines (AD 08009, AD 09525, IET 19863, IET 20885, IET 20897, and IET 20898) of WA cytoplasmic source were used as the genetic material for the present study. Pure seeds of rice lines were obtained from the Paddy Breeding Station, Centre for Plant Breeding and Genetics (CPBG), Coimbatore. The plant population was developed and maintained at the above mentioned center. Genomic DNA was extracted using modified CTAB method from freshly collected leaves (45 DAS) and its purity was assessed by Nanodrop™ 1,000 Spectrophotometer. PCR amplification of genes (xa5, xa13, and Xa21) for all the parents, F1, BC1F1, BC2F1, BC2F2, and BC2F3 generations from the cross of CO 2B, CO 23B, and CO 24B with IRBB60 respectively, and bacterial blight screening was carried out as described in our earlier study (Perumalsamy et al., 2010). PCR details have been provided in Datasheet S1. The amplified products were resolved in 2.5% agarose gel (Figure 1). The crossing layout of hybrid development is provided in Figure 2. Pollens were collected in randomly selected F2 plants from a cross of A × R line.

Pollens were collected in randomly selected F2 plants from a cross of A × R line. Pollen grains were squeezed out and placed on a glass slide for staining with (I- KI). Deep stained and round sized pollen was considered as fertile. Spikelet fertility was checked from the panicles of selected plants. Number of filled seeds to the number of spikelet in a panicle were taken into account to predict spikelet fertility.

Functional Marker Analysis

Functional marker for xa5 gene (Iyer-Pascuzzi and Mccouch, 2007), xa13 gene (Chu et al., 2006), and Xa21 gene (Perumalsamy et al., 2010) were used. For fertility restoration, markers linked to Rf3 and Rf4 were used. Three SSR markers, comprising one RM marker - RM 6100 (160 bp in non-restorer line and 175 bp for...
FIGURE 1 | Foreground selection in progenies. PCR amplification of BC2F2 plants of CO 23B × IRBB60 cross combination (1) xa5_1F, xa5_1R restricted with the enzyme BsrI to detect polymorphism (2) with xa13 (3) with Xa21 (4) DRRM-RF3-10 maker for Rf3 genes (5) RM6100 marker for Rf4. In figure, R, H, S indicates Resistant, heterozygote and susceptible respectively. CO2B, CO23B, CO24B, and CO24A were susceptible genotypes and IRBB60 was resistant source.

RESULTS

Introgression of BB Resistance Genes in Maintainer Line and CMS Line

Functional markers were used to incorporate both BB genes (xa5, xa13, and Xa21) and the two major Rf genes (Rf3 and Rf4) governing fertility restoration of cytoplasmic-geneic male sterility (CMS) in our targeted rice varieties. During the breeding procedure, foreground selection was practiced from the F1 generation to BC2F2 generation. At each stage, the plants having resistance alleles of the three target genes were selected and only progenies having resistance alleles of the three target genes were advanced to the next generation. A total of 20 (CO2B), 16 (CO23B), and 10 (CO24B) plants were identified to be true F1s and back crossed to respective recurrent parents.

In BC1F1, foreground selection with functional markers of target genes resulted in identification of heterozygous plants in

restorer line), two candidate gene markers DRCG-RF4-14 (885 bp in non-restorer line and 845 bp for restorer line) and DRCG-RF4-8 (1096 bp in non-restorer line and 760 bp for restorer line) for Rf4 locus and two SSR markers DRRM-RF3-5 (140 bp in non-restorer line and 160 bp for restorer line) and DRRM-RF3-10 (140 bp in non-restorer line and 150 bp for restorer line) for Rfβ reported by Suresh et al. (2012) were used in BC2F2 and BC2F3 generations. The details of markers sequences were given in Table 1. To examine parental polymorphism between donor and elite parents, microsatellite markers covering the whole rice genome were utilized. The primer information and chromosomal position were retrieved from the Gramene database (www.gramene.org).

The plants were scored as B (maintainer indicates allele), H (heterozygous state), and NR (non-fertility restorer genes). For fertility restoration genes (Rf3, Rf4), the amplified PCR product was separated in 3 per cent agarose gel.
all three crosses. A total of 13/60 (CO 2B), 2/58 (CO 23B), and 2/65 (CO 24 B) plants were found to be triple heterozygous for the target “R” gene functional markers. Background analysis was carried out with polymorphic SSR markers to assess the recovery of recurrent genome contribution in identified triple heterozygous plants in all three combinations. Resultant triple gene heterozygous (Xa21Xa21, Xa13xa13, and Xa5xa5) plants with maximum recovery of the recurrent genome in all three crosses, CO 2B (74.2%), CO 23B (73.8%), and CO 24B (73.5%) were further back crossed with their respective recurrent parents (Figure 3).

In BC2F1, a total of 12/96 (CO 2B), 9/101 (CO 23B), and 9/89 (CO 24B) plants were found to be triple heterozygous for Xa5, xa13, and Xa 21 genes. The heterozygous plants were subjected to back-ground selection using polymorphic SSR markers. Single triple heterozygous plant with maximum recovery of recurrent parent in CO 2B (87.1%), CO 23B (86.2%), and CO 24B (85.2%) were selfed and forwarded to F2 generation (Figure 3). In BC2F2 generation, a total of 190 (CO 2B), 160 (CO 23B), and 120 (CO 24 B) were subjected to PCR analysis using FMs (Figure 1). The segregation pattern for bacterial blight resistance in BC2F2 was tested using chi square (χ²) analysis. The segregation of all the three resistance genes was insignificant as the calculated χ² value is lower than the table values at 5% (5.99) and 1% (9.21) levels (Table 2). Plants with single, double, and triple gene combinations with BB resistance were identified. In CO 2B × IRBB60, forty one plant with Xa21 gene, 52 plants with xa13 and 43 plants with the recessive gene xa5 in homozygous condition were identified. Fifteen plants homozygous for two genes xa5 and xa13, 10 for xa5 and Xa21 and 10 plants for xa13 and Xa21 were identified. Twelve plants possessing all the three genes, xa5, xa13, and Xa21 in homozygous conditions were identified. Similarly, in CO 23B × IRBB60 and CO 24 B × IRBB 60 plants with different resistance genes combinations were obtained. Plants with triple homozygous for target genes in all three crosses, 12/190 (CO 2B), 9/160 (CO 23B), and 9/120 (CO 24 B) were found.

**Bacterial Blight Screening**

The selected backcross-derived 30 homozygous lines from three crosses along with recurrent parents, donor line and susceptible check were evaluated for their reactions to five predominantly available races of bacterial blight pathogen under greenhouse condition. A representative picture is given as Figure 4. The mean lesion lengths of plants with three resistance genes in BC2F3 population of three crosses were given in Table 3. As expected, all the 30 homozygous plants with three resistance genes (i.e., plants with genotype of xa5xa5, xa13xa13, and Xa21Xa21) showed higher levels of resistance with a mean lesion length of less than 5.0 cm for all the races with few exceptions. The susceptible check, IR 24 and recurrent parents, CO 2B, CO 23 B and CO 24 B showed more than 15.0 cm of lesion length and categorized as highly susceptible to bacterial blight pathogen. The donor parent, IRBB 60 showed high level of resistance with mean lesion length of 4.53 cm.

**Hybrid Development**

Presence/absence of both fertility restorer genes, Rf3 and Rf4 were verified in 30 homozygous. Two plants each in CO 2B × IRBB60 and CO 24B × IRBB60 combination, one plant in CO 23B × IRBB60 shows negative for both Rf3 and Rf4 fertility-restorer genes. Based on the agronomic traits, a single plant from CO 2B, CO 23B, and CO 24B cross combinations was selected and test crossed with respective CMS lines. The test crossed plants are intact for all three resistance genes and completely sterile for both pollen and spikelet fertility. Two more back crosses are needed to convert these lines into a male sterile source. Restorer lines, AD 08009 AD 09525, IET 19863, 20885, 20897 20898 of WA cytoplasmic source were crossed with improved lines of CO 2B, CO 23B, and CO 24B and hybrids were tested for their superiority for yield. Of several hybrid combinations, the hybrids combinations viz., CO 23A × AD08009R and CO 24A × IET20898R were found to be stable over locations with high yield (Table S1).

**DISCUSSION**

To overcome agrochemical pollution and time consuming conventional breeding methods, we should develop long lasting resistance varieties in the management of BB disease (Khush et al., 1989; Gnanamanickam et al., 1999). With the current advances in DNA marker based breeding and gene pyramiding, it becomes more comfortable in developing BB resistant rice varieties without compromising their yield and grain quality. Tagging of pyramided resistance genes and identifying the presence of multiple genes showing resistance to different races of Xoo pathogen through functional markers would provide a valid and reproducible selection strategy. Functional marker assisted selection played a key role in improving cytoplasmic male sterile lines by incorporating the three bacterial blight resistance genes (Xa21, xa13, and xa5) through back-crossing and selling. These

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**FIGURE 2** | Schematic representation of the introgression of xa5, xa13, and Xa21 into the maintainer and CMS line.
TABLE 1 | Details of markers used for bacterial blight resistance and fertility restoration.

| S.No | Marker | Primer Sequences 5′ → 3′ | Chromosomal location | Authors |
|------|--------|--------------------------|----------------------|---------|
| **BACTERIAL BLIGHT RESISTANCE** | | | | |
| 1. | xa 5-1F | F- ACGCTCGACGAGATGGTCTC | 5 | Iyer-Pascuzzi and McCouch, 2007 |
| | xa 5-1R | R- ATCACAAGCCCATATAGGAG | | |
| 2. | xa 13 | F- AGTCACAGCTCCAAATG | 8 | Chu et al., 2006 |
| | | R- GGCCATGGCTCAGTGTTTAT | | |
| 3. | Xa 21 | F- ATAGCTAGTTCATAGAGG | 7 | Perumalsamy et al., 2010 |
| | | R- ACATCCGTCACTCTTGCCA | | |
| **Rf3 LOCUS** | | | | |
| 4. | DRRM-RF3-5 | F- GATGGCACAGCTTCAGAACA | 1 | Suresh et al., 2012 |
| | | R- CTAATTCTGGGCGAGCAAAG | | |
| 5. | DRRM-RF3-10 | F- TCACCTCTTTCTGCTTCGAC | 1 | Suresh et al., 2012 |
| | | R- CTCCACCAGTGCAGGTTTT | | |
| **Rf4 LOCUS** | | | | |
| 6. | DRCG-RF4-14 | F- GCAATGCTTGTATTCAGCAAA | 10 | Suresh et al., 2012 |
| | | R- TCCAGCTGTAAATCCGTCAA | | |
| 7. | DRCG-RF4-8 | F- TTGCAACGCAAGGGTAATTT | 10 | Suresh et al., 2012 |
| | | R- TCACTGCGCATCTTTTTGAG | | |
| 8. | RM 6100 | F- TCCTCTACACAGTACCGACC | 10 | Sheeba et al., 2009 |
| | | R- GCTGGATCACAGATCATTGC | | |

FIGURE 3 | Graphical representation of background analysis. Y-axis represents the percentage of recurrent parent genome recovery using polymorphic SSR markers distributed all over the 12 chromosomes (RPGSSR) in BC1F1 generation (A) CO 2B × IRBB60, (B) CO 23B × IRBB60, and (C) CO 24B × IRBB60. While, X-axis indicates the number of agro-morphologically selected plants from foreground analysis subjected to RPGSSR analysis carrying xa5, xa13, and Xa21 gene combination.

Genes conferred a tough and wide spectrum resistance to Xoo strains distributed worldwide where rice is grown.

More than 38 major genes, named from Xa1 to Xa38 (Bhasin et al., 2011) have been identified from various environments and have conferred resistance to different strains of Xoo pathogens. The rice gene Xa21 has got its own significance in breeding practices for developing rice varieties with bacterial blight resistance due to its wide range of resistance against bacterial blight disease (Huang et al., 1997; Li et al., 2001; Pradhan et al., 2015; Luo et al., 2016).

xa5 is an important BB resistance gene because of its recessive nature and it does not belong to the emblematic structural classes of resistance genes. xa5 gene, which depends on the effector genes present in the pathogens, is broadly effective against the Xoo races (Jiang et al., 2006). The efficiency of xa5 in combating Xanthomonas oryzae pv. Oryzae depends mainly on TAL (Transcription Activation-like) effector genes, but regrettably futile in combating disease by strains in which S gene is expressed. The xa5 gene, though provides vertical resistance, but not associated with dominant resistance genes or clusters of resistance gene analogs (Ilag et al., 2000; Wang et al., 2001). The race specific xa13 mediated resistance is exclusively resistance to the most virulent Philippines Xoo race 6 which is mostly not conquered by other reported R genes. The fully recessive xa13 is insensitive to TAL effector PhxXo1 and therefore highly resistant to the PhxXo1 dependent virulence of pathogens (Khush and Angeles, 1999). Improved resistance was achieved by the suppression of either dominant or recessive allele, but also resulted in male sterility pointing that this gene plays a role in pollen development as well as in bacterial growth dependent modular (Chu et al., 2006).

The combination of these three genes (Xa21 + xa5 + xa13) can accomplish a tough and broad spectrum resistance in many BB prone rice growing areas in India. The order of gene combinations in conferring resistance xa5 + xa13 < xa5 + Xa21 < xa13 + Xa21 < xa5 + xa13 + Xa21 indicated the degree of severity of disease. Lines with Xa21 in combination with either xa5, xa13 or both have shown promise advocating the utility of Xa21 in achieving higher levels of resistance in rice as reported earlier (Sanchez et al., 2000; Singh et al., 2001). The present work was thus focused...
not only to pyramid both recessive and dominant genes for resistance, but to keep the maintaining ability of parental lines for sterility.

The genes of BB resistance should be incorporated into both the parental lines of the hybrid for the expression of desired level of resistance. To incorporate resistance genes into CMS lines, first, the genes must be transferred to the maintainer’s background. Thirty homozygous lines with high resistance to bacterial blight in the background of CO 2A, CO 23 A, and CO 25 A were developed. They were again screened for the presence of fertility restorer genes, Rf 3 and Rf 4 through molecular marker linked to these genes. SSR markers DRCG-RF4-14/DRCG-RF4-8/RM6100 for the Rf4 locus and DRRM-RF3-5/DRRM-RF3-10 for Rf3 linked to fertility restoration genes (Rf3, Rf4) were employed for detecting efficacious non-fertility restorer plants for WA-CMS system.

This helped us to identify the plants possessing resistance gene, which also had non restoring alleles in homozygous condition at BC2F2 generation. Sundaram et al. (2007) successfully pyramided the resistance genes (Xa21 + xa5 +xa13) into Samba Mashuri, an elite rice variety having high yield grain and cooking quality. Using rice microsatellite (RM) or simple sequence repeat (SSR) markers, Rf3 and Rf4 loci have been identified in different donors (He et al., 2002; Sattari et al., 2006; Bazrkar et al., 2008; Nematzadeh and Kiani, 2010). Marker combination of DRCG-RF4-14/DRCG-RF4-8/RM6100 for the Rf4 locus and DRRM-RF3-5/DRRM-RF3-10 for Rf3 showed the maximum efficiency to select the non-fertility restorer genes. The earlier attempts for MAS have used STS markers (Nas et al., 2003; Sattari et al., 2007) and either Rf3 or Rf4 markers (Nas et al., 2003; Sheeba et al., 2009; Ngangkham et al., 2010; Pranathi et al., 2016; Katara et al., 2017), only one study used two STS markers from Rf3 and Rf4 loci with selection accuracy of 100% in 13 R lines (Sattari et al., 2007). Selection accuracy of 94.9% in a set of 21 restorer lines with RM6100 from Rf4 was reported by Sheeba et al., (2009). Sattari et al., (2007) and Bazrkar et al., (2008) used the sequence tagged sites (STS) RG140/PvuII and S10019/BstUI in MAS for fertility restoration genes Rf3 on chromosome 1 and Rf4 on chromosome 10 in rice. This suggests that two dominant genes Rf3 and Rf4 seem to control the fertility restoration based on above studies. The effect of one of the two dominant genes (Rf3) in restoring fertility appears to be strong and as good as the two together (Rf3Rf4) while the other gene (Rf4) showed weak restoration in rice (Mahalingam and Saraswathi, 2016). These results show two dominant genes Rf3 and Rf4 to control the fertility restoration. If Rf3, Rf4 genes are present together, the effect of one of the two genes in restoring fertility appeared to be stronger than the other. This indicated that the recessive gene alone control the fertility restoration (Chu et al., 2006). From the above work, it was observed that in the presence of Rf3 and Rf4 genes plants shows complete fertility restoration. With the use of Rf3 (DRRM-RF3-5/DRRM-RF3-10) and Rf4 (DRCG-RF4-14/DRCG-RF4-8/RM6100) markers, the positive plants were ignored and the plants which are negative for the Rf3, Rf4 genes were selected which would never restore fertility and shows only sterile, also phenotypically similar to the recurrent parent. The above selected negative plants were confirmed as maintainer by crossing the improved lines with corresponding male sterile lines (CO 2A, CO 23A, and CO 24A).

A single plant in each cross was identified with all BB resistance and without fertility restorer genes- Rf3 and Rf4. These plants are highly resistance to all five races of Xoo pathogen and maintain 100% male sterility in the test crosses. Non-recovery

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**TABLE 2 | Segregating ratio of the marker genotypes in BC2F2 generation for BB.**

| S.No | Markers | 2B x IRBB60 | 23B x IRBB60 | 24B x IRBB60 |
|------|---------|-------------|-------------|-------------|
|      |         | Observed frequency | Observed frequency | Observed frequency |
|      |         | RR | Rr | Rr | Total | χ^2 | RR | r | r | Total | χ^2 | RR | r | r | Total | χ^2 |
| 1.   | Xa21 (F&F) | 41 | 104 | 45 | 190 | 1.873 | 32 | 77 | 51 | 160 | 4.237 | 28 | 59 | 33 | 120 | 0.449 |
| 2.   | xa13 (F&F) | 52 | 83 | 55 | 190 | 3.126 | 37 | 72 | 51 | 160 | 4.437 | 39 | 54 | 27 | 120 | 3.600 |
| 3.   | xa5 (F&F) | 43 | 97 | 50 | 190 | 0.599 | 29 | 78 | 53 | 160 | 4.950 | 32 | 54 | 34 | 120 | 1.266 |

RR, r: Homozygotes; Rr: Heterozygote.

**FIGURE 4 | Differential reactions of Xoo isolates on rice leaves at 14 dpi upon artificial clip inoculation.**
of recessive resistance gene xa5 with other genes was observed in earlier studies of Dokku et al. (2013). But in this study no such instance, was observed. Using the improved version of parental lines 2B, 23 B, and 24 B, number of hybrids were developed. Two hybrids, CO 23B × AD08009R and CO 24B × IET20898R, were found to be stable over locations with high yield. This new hybrid with resistance against bacterial blight will increase the rice productivity. Using functional markers, durable resistance cytoplasmic male sterile lines were developed in this study through MAS. It demonstrates the successful transfer of recessive and dominant genes with the intact recurrent genome.

CONCLUSION

We have introgressed three BB resistance genes (xa5, xa13, and xa21) which would be desirable to achieve durable and broad spectrum resistance in the maintainer lines of hybrid rice through functional markers assisted backcross breeding. Thirty homozygous plants, 12 in CO 2B × IRBB60, nine each in CO

### TABLE 3

| S.No | Pyramided lines | Screening against five Important pathotypes of bacterial blight pathogen |
|------|-----------------|-------------------------------------------------------------------------|
|      |                 | DX-002 | DX-020 | DX-027 | DX-148 | DX-321 |
| 2B × IRBB 60 |
| 1    |                 | 3.57 ± 0.30 | 7.07 ± 1.09 | 3.83 ± 0.38 | 4.02 ± 0.32 | 3.84 ± 0.19 |
| 2    |                 | 3.83 ± 0.26 | 5.95 ± 0.47 | 4.30 ± 0.26 | 4.17 ± 0.15 | 4.20 ± 0.35 |
| 3    |                 | 1.33 ± 0.12 | 5.97 ± 0.46 | 6.87 ± 1.28 | 4.03 ± 0.12 | 4.47 ± 0.20 |
| 4    |                 | 3.52 ± 0.29 | 6.03 ± 0.97 | 3.72 ± 0.55 | 4.13 ± 0.41 | 6.10 ± 0.40 |
| 5    |                 | 3.83 ± 0.44 | 3.72 ± 0.21 | 3.50 ± 0.29 | 6.43 ± 0.93 | 7.03 ± 1.28 |
| 6    |                 | 1.15 ± 0.15 | 3.92 ± 0.12 | 4.15 ± 0.23 | 3.33 ± 0.29 | 7.22 ± 1.14 |
| 7    |                 | 4.30 ± 0.21 | 3.64 ± 0.36 | 7.53 ± 1.28 | 4.28 ± 0.20 | 6.40 ± 0.76 |
| 8    |                 | 4.27 ± 0.27 | 7.57 ± 0.52 | 4.20 ± 0.21 | 3.53 ± 0.18 | 3.60 ± 0.15 |
| 9    |                 | 5.43 ± 0.44 | 4.17 ± 0.35 | 4.53 ± 0.50 | 4.17 ± 0.35 | 3.03 ± 0.50 |
| 10   |                 | 3.43 ± 0.26 | 5.03 ± 0.44 | 4.12 ± 0.29 | 6.33 ± 0.64 | 4.13 ± 0.38 |
| 11   |                 | 3.73 ± 0.33 | 4.03 ± 0.50 | 4.17 ± 0.35 | 7.22 ± 0.94 | 7.32 ± 1.09 |
| 12   |                 | 4.03 ± 0.38 | 7.03 ± 0.45 | 6.03 ± 0.88 | 3.67 ± 0.12 | 13.77 ± 0.69 |
| 23 B × IRBB 60 |
| 13   |                 | 5.57 ± 0.17 | 7.93 ± 0.97 | 4.20 ± 0.60 | 7.77 ± 0.72 | 8.77 ± 0.47 |
| 14   |                 | 3.70 ± 0.21 | 6.83 ± 0.36 | 3.73 ± 0.33 | 4.0 ± 0.70 | 3.78 ± 0.50 |
| 15   |                 | 6.80 ± 0.49 | 0.93 ± 0.35 | 4.10 ± 0.36 | 4.01 ± 0.23 | 4.00 ± 0.42 |
| 16   |                 | 8.10 ± 2.83 | 4.20 ± 0.06 | 5.07 ± 0.38 | 7.60 ± 0.47 | 7.33 ± 2.36 |
| 17   |                 | 3.23 ± 0.32 | 3.43 ± 0.30 | 3.70 ± 0.31 | 5.92 ± 0.91 | 3.57 ± 0.32 |
| 18   |                 | 3.55 ± 0.32 | 3.43 ± 0.32 | 3.98 ± 0.32 | 1.57 ± 0.22 | 7.40 ± 0.76 |
| 19   |                 | 3.17 ± 0.12 | 3.43 ± 0.26 | 3.60 ± 0.29 | 3.50 ± 0.35 | 1.00 ± 0.31 |
| 20   |                 | 3.83 ± 0.18 | 3.70 ± 0.15 | 3.57 ± 0.52 | 3.53 ± 0.30 | 3.83 ± 0.18 |
| 21   |                 | 3.21 ± 0.26 | 3.39 ± 0.36 | 3.32 ± 0.20 | 3.45 ± 0.24 | 1.42 ± 0.31 |
| 24 B × IRBB 60 |
| 22   |                 | 3.66 ± 0.32 | 3.73 ± 0.33 | 6.97 ± 0.97 | 3.70 ± 0.31 | 3.57 ± 0.32 |
| 23   |                 | 3.43 ± 0.32 | 0.97 ± 0.35 | 3.63 ± 0.32 | 0.57 ± 0.20 | 7.40 ± 0.76 |
| 24   |                 | 3.27 ± 0.11 | 3.48 ± 0.26 | 3.20 ± 0.28 | 3.59 ± 0.39 | 1.10 ± 0.31 |
| 25   |                 | 3.43 ± 0.18 | 3.70 ± 0.15 | 3.57 ± 0.52 | 3.53 ± 0.30 | 3.83 ± 0.18 |
| 26   |                 | 2.04 ± 0.22 | 1.55 ± 0.28 | 0.68 ± 0.30 | 2.67 ± 0.28 | 2.98 ± 0.15 |
| 27   |                 | 3.25 ± 0.32 | 3.29 ± 0.31 | 0.98 ± 0.42 | 1.97 ± 0.22 | 5.30 ± 0.76 |
| 28   |                 | 2.04 ± 0.22 | 1.65 ± 0.28 | 0.68 ± 0.30 | 2.67 ± 0.28 | 2.98 ± 0.15 |
| 29   |                 | 2.84 ± 0.32 | 2.57 ± 0.26 | 1.64 ± 0.35 | 2.36 ± 0.25 | 2.55 ± 0.25 |
| 30   |                 | 2.38 ± 0.25 | 1.55 ± 0.28 | 1.58 ± 0.33 | 2.60 ± 0.65 | 2.73 ± 0.18 |
| 31   | (Recurrent parent) | 24.47 ± 1.09 | 26.53 ± 1.21 | 26.27 ± 2.59 | 26.20 ± 2.43 | 9.30 ± 0.15 |
| 32   | (Recurrent parent) | 26.27 ± 1.85 | 27.03 ± 1.91 | 26.23 ± 2.46 | 13.07 ± 0.38 | 27.10 ± 2.26 |
| 33   | (Recurrent parent) | 28.50 ± 1.06 | 16.30 ± 0.47 | 12.47 ± 0.84 | 13.43 ± 0.60 | 27.13 ± 1.65 |
| 34   | (Susceptible check) | 23.89 ± 2.49 | 21.71 ± 1.39 | 19.40 ± 2.09 | 23.16 ± 3.01 | 26.54 ± 2.55 |
| 35   | (Donor parent) | 4.33 ± 0.29 | 4.43 ± 0.92 | 3.30 ± 0.17 | 0.83 ± 0.23 | 4.53 ± 0.92 |

The values are given in cm.
23B × IRBB60 and CO 24B × IRBB60 were identified and finally single line in each male sterile background without out fertility restorer genes, Rf3 and Rf4 was developed. The improved versions of male sterile and maintainer lines of stable CMS lines CO 2A, CO 2A, and CO 2A can form the base to develop new wide adoptable heterotic hybrids with resistance against the most destructive diseases on rice. Using these lines, two high yielding and stable hybrid combinations- CO 23B × AD08009R and CO 24B × IET20889R were identified in the current study.

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**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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