Anticonvulsant and anxiolytic evaluation of leaf extracts of *Ocimum gratissimum*, a culinary herb

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**ABSTRACT**

Anticonvulsant and anxiolytic activities of leaf extracts and fraction of *Ocimum gratissimum* L. (Lamiaceae) were studied using seizures induced by pentylentetrazol and open-field tests in mice. The results showed that the extracts and fraction increased the latency of tonic and tonic-clonic seizures and death and elicited 50% protection against mortality. In the open-field test, the extracts and fraction decreased the frequency of line crossing, center square entries, rearing against a wall and grooming, whereas grooming duration and freezing frequency and duration were increased. Acute toxicity test in mice gave an oral LD$_{50}$ greater than 5000 mg/kg for the methanol extract. These findings suggest that extracts of this plant possess anticonvulsant and anxiolytic-like properties.

**Key words:** Anticonvulsant, anxiolytic, *Ocimum gratissimum*, open field

**INTRODUCTION**

*Ocimum gratissimum* L. (Lamiaceae) is a small shrub commonly known as “scent leaf,” “tea bush” or “fever plant.” In Nigeria, it is variously called “Nchuanwu,” “Ahibi,” “Ahigbu” (Igbo), “Efirin” (Yoruba), “Ihiri eziza” (Bini), “Dai doya tagida” (Hausa) or “Ntion” (Efik) and is found in the wild or cultivated throughout the tropics and subtropics. In West Africa, *O. gratissimum* is commonly found around village huts and gardens and cultivated for medicinal and culinary purposes. The leaves have strong aromatic odor and are popularly used to flavor soup and spice meat, especially game. In southeastern part of Nigeria and beyond, the leaves popularly serve as indispensable flavoring agent in soups, especially “pepper soup,” and other such meals. In traditional medicine practice, it is used in the treatment of diarrhea, as a febrifuge and component of anti-malaria remedies, mosquito/insect repellent, stomachic and general tonic, antiseptic, in wound dressing, skin infections, conjunctivitis and bronchitis. An infusion of the leaves, called ‘Ocimum tea,’ is dispensed as a remedy for fever and diaphoresis. The roots are used as sedative for children. Extract of the crushed leaves is an excellent remedy for cough. In southeastern parts of Nigeria, in addition to serving culinary purposes, the leaves are also used for the treatment of convulsive disorders.

Experimental studies showed that extracts of this plant relaxed intestinal smooth muscle, exhibited antinociceptive effect, contracted the guinea pig ileum and rat colon, raised mean arterial pressure in rats and lowered blood glucose in diabetic rats. The volatile oil has been credited with antimicrobial, anthelminthic and insect-repellant properties, while the essential oil exhibited sedative and anxiolytic activities. The essential oil also protected mice against tonic seizures induced by maximum electroshock but was not effective against pentylentetrazol-induced seizures.

Several constituents have been identified in oil from the leaves of this plant. Eugenol, a monoterpen, has been severally identified as the dominant volatile constituent. Also present are the monoterpenes-1, 8-cineole, β-pinene, cis-Ocimene, trans-Ocimene, camphor and methyl eugenol; and the sesquiterpenes-trans-Caryophyllene and Germacrene-D, as well as thymol, xanthisones and lactones.

Although an earlier study has demonstrated the anti-convulsant, sedative and anxiolytic properties of essential oil of the leaves, these activities are not clearly attributable to any specific constituent of the oil. Also, the neuropharmacological effects of non-oil constituents of the leaves are not known. Consequently, in accordance with the need for additional information on the pharmacological properties of the leaves, anticonvulsant and anxiolytic evaluation of leaf extracts of *Ocimum gratissimum* L. was performed.
with its use in traditional medicine practice in southeastern Nigeria, we studied the effects of the leaf extracts and fraction on PTZ-induced seizures and paradigms of anxiety and depression in mice.

**MATERIALS AND METHODS**

**Drugs**
Diazepam (Valium™, Roche)

**Chemicals, solvents and reagents**
Ethanol (Fluka, Germany), methanol (Fluka, Germany), pentylenetetrazol (PTZ), petroleum ether (Fluka, Germany), Tween 80.

**Equipment**
Soxhlet (Staffordshire, ST 150BG; England), rotary evaporator (Staffordshire, ST 150BG; England), open-field apparatus (A plexiglass box measuring 72 × 72 cm with 36-cm high walls. The walls and the floor were painted white. Blue lines, drawn under the clear plexiglass floor with a marker, divided the floor into 16 squares (18 × 18 cm). A central square of equal size was drawn in the middle of the maze) and video camera.

**Animals**
Adult male Swiss albino mice (22-30 g) bred in the laboratory animal facility of the Department of Pharmacology and Toxicology, University of Nigeria, Nsukka, were used for the study. The animals were maintained freely on standard pellets and water and allowed 2 weeks acclimatization period before commencement of studies. All animal experiments were in compliance with the National Institute of Health Guide for Care and Use of Laboratory Animals (Pub. No. 85-23, revised 1985).

**Collection of plant material and preparation of extract**
Fresh leaves of O. gratissimum were collected in May 2007 from Orba, Enugu State, Nigeria. The plant material was identified and authenticated by Mr. A. Ozioko of the International Centre for Ethnomedicines and Drug Development (InterCEDD), Nsukka, Nigeria. The leaves were cut into smaller pieces, dried under shade for 5 days and pulverized to coarse powder using a manual blender. The leaf powder (250 g) was extracted with methanol by continuous extraction in a soxhlet. A fresh batch of leaf powder (750 g) was successively extracted with petroleum ether (40°C-60°C) and methanol in a soxhlet. The extracts were concentrated in a rotary evaporator at 40°C-50°C under reduced pressure to afford the methanol extract (ME: 19.5 g; 7.8% w/w), petroleum ether extract (PE: 9.48 g; 1.26% w/w) and methanol fraction (MF: 11.31 g; 1.51% w/w). The extracts and fraction were subjected to phytochemical analysis for constituent identification using standard procedures.[15,16]

**Acute toxicity test**
The acute toxicity and lethality of ME were determined in mice using the method described by Lorke.[17] Briefly, nine mice randomly divided into three groups (n = 3) were orally administered 10, 100 and 1000 mg/kg of ME, respectively, and observed for 24 hours for death. Since no death occurred, 1600, 2900 and 5000 mg/kg of ME was administered to a fresh batch of animals (n = 1) and the number of deaths in 24 hours recorded. The LD₅₀ was calculated as the geometric mean of the highest nonlethal dose and the lowest lethal dose.[17]

**Pentylenetetrazol-induced seizure test**
The anticonvulsant activity of ME, PE and MF was evaluated using the pentylenetetrazol-induced seizure in mice. Adult male albino mice (22-30 g) were randomly divided into eight groups (n = 6). Animals in groups I-VI received oral administration of ME, PE and MF (200 and 400 mg/kg), respectively, while groups VII and VIII received diazepam (1 mg/kg p.o.) and 10% Tween 80 (10 mL/kg p.o.), respectively. Thirty minutes later, pentylenetetrazol (PTZ) (70 mg/kg i.p.) was administered to each animal. The animals were observed for 60 minutes for seizures; an episode of clonic spasm that persisted for a minimum of 30 seconds was taken as a threshold convulsion. Animals devoid of threshold convulsion and without subsequent death during the 60 minutes of observation were considered protected.[18] The onset and duration of seizures, as well as quantal protection, were recorded for each group.

**Open field test**
The effects of ME, PE and MF on locomotor and exploratory activities, as well as grooming behavior in the open field,[19] were investigated. Adult male Swiss albino mice (19-30 g) were selected at random and divided into groups (n = 6). Each group received oral administration of one of ME, PE or MF (200 or 400 mg/kg) suspended in Tween 80 (10% v/v). The control groups received the vehicle (10 mL/kg) or diazepam (1 mg/kg). Thirty minutes later, each mouse was placed in the center square of the open field and observed for 5 minutes with the aid of video camera. The behavioral parameters recorded included line crossing, center square entries, rearing and grooming behaviors. The floor of the open field was cleaned with 70% ethanol and allowed to dry between tests.

**Statistical analysis**
Data obtained was analyzed using one-way ANOVA, and the results were expressed as mean ± SEM. Means were compared using LSD post hoc test and differences between treatment and control groups accepted as significant at P < 0.05.
RESULTS

Acute toxicity and lethality (LD₅₀) of methanol extract
The acute toxicity test showed that oral administration of ME caused no death in the two stages of the test. The oral LD₅₀ of ME in mice was therefore greater than 5000 mg/kg.

Phytochemical constituents of extracts and fraction
Phytochemical tests showed that ME tested positive to alkaloids, carbohydrates, flavonoids, glycosides, resins, saponins and tannins. Petroleum ether extract (PE) gave positive reactions for steroids and terpenoids, in addition to all the constituents of ME. The methanol fraction (MF) tested positive to alkaloids, carbohydrates and resins [Table 1].

Effect of extracts and fraction on pentyleneetetrazol-induced seizures
The extracts and fraction elicited a non-dose-related increase in the latency of tonic and tonic-clonic seizures and death. They also offered 50% protection of treated mice against seizure-induced mortality [Table 2].

Table 1: Phytochemical constituents of the extracts and fraction

| Phytochemical constituents | ME (7.8% w/w) | PE (1.26% w/w) | MF (1.51% w/w) |
|----------------------------|---------------|----------------|---------------|
| Alkaloids                | + + +         | + + +          | + +           |
| Carbohydrates            | + +           | + +            |               |
| Flavonoids               | + +           | + +            |               |
| Glycosides               | + +           | + +            |               |
| Reducing sugar           | -             | -              |               |
| Resins                   | + +           | + + +          | + +           |
| Saponins                 | + +           | + +            |               |
| Steroids                 | -             | + + +          | -             |
| Tannins                  | +             | + +            |               |
| Terpenoids               | -             | + +            |               |

Values in parentheses are extractive yields calculated relative to the weight of extracted plant material. ME = Methanol extract; PE = Petroleum ether extract; MF = Methanol fraction; + = Present; ++ = Conspicuously present; + + = Moderately present; ++ = Absent

Effect of extracts and fraction on locomotion and exploratory activities
The extracts and fraction significantly (P < 0.05) decreased the frequency of line crossing in a non-dose-related manner. The frequencies of center square entries, rearing against a wall and grooming were also reduced, whereas grooming duration and freezing frequency and duration were increased [Table 3].

DISCUSSION

Although the popularity of O. gratissimum is derived mainly from its culinary uses, the medicinal value has long been recognized. In this study, extracts of the leaves exhibited anticonvulsant activity by delaying the onset of PTZ-induced seizures and protecting treated mice from mortality induced by seizures. PTZ induces convulsion by antagonizing the γ-aminobutyric acid (GABA)₁ receptor chloride (Cl)-channel complex to attenuate GABA-dependent inhibition. Drugs protecting against tonic-clonic seizures induced by PTZ are considered useful in controlling myoclonic and absence seizures in humans. Thus, demonstration of activity in this seizure model suggests that the plant possesses anticonvulsant activity, which may underlie its traditional use in the treatment of convulsive disorders. Although the effect of the extracts and fraction on GABA was not evaluated in this study, antagonism of PTZ-induced seizures suggests possible enhancement of GABAergic transmission consistent with general depression of the central nervous system (CNS). An earlier study showed that the essential oil was not effective in protecting against PTZ seizures. In this study, however, the result has shown that constituents of the extracts and fraction protected treated mice against PTZ seizures. Thus, it is likely that constituents other than those of the essential oil are responsible for the anticonvulsant activity of leaves of this plant.

Evaluation of the effect of the extracts and fraction on paradigms of depression and anxiety in the open field showed decreased locomotor and exploratory activities

Table 2: Effect of extracts and fraction on pentyleneetetrazol-induced seizures

| Treatment | Dose (mg/kg) | Onset of tonic seizure (min) | Onset of tonic-clonic seizure (min) | Time of death (min) | Quantal protection | Protection (%) |
|-----------|-------------|-----------------------------|-----------------------------------|---------------------|-------------------|---------------|
| ME        | 200         | 3.68 ± 1.29                | 31.35 ± 10.83                    | 36.60 ± 8.86*       | 3/6               | 50            |
|           | 400         | 1.70 ± 0.55                | 6.13 ± 2.46*                     | 14.39 ± 5.05*       | 3/6               | 50            |
| PE        | 200         | 6.09 ± 1.11                | 32.70 ± 10.35*                   | 42.38 ± 7.68*       | 3/6               | 50            |
|           | 400         | 2.24 ± 0.44                | 2.81 ± 0.58                      | 33.68 ± 9.98*       | 0/6               | 0             |
| MF        | 200         | 1.76 ± 0.30                | 4.60 ± 0.32                      | 37.02 ± 8.69*       | 3/6               | 50            |
|           | 400         | 3.38 ± 0.59                | 18.79 ± 9.06                     | 38.64 ± 8.36*       | 3/6               | 50            |
| Control   | -           | 1.09 ± 0.25                | 1.41 ± 0.20                      | 9.44 ± 1.38         | 0/6               | 0             |
| Diazepam  | 1           | 46.0 ± 9.14*               | 60.00 ± 0.00                     | 60.00 ± 0.00*       | 6/6               | 100           |

n = 6; *P < 0.05 compared to control group (one-way ANOVA; LSD post hoc); ME = Methanol extract; PE = Petroleum ether extract; MF = Methanol fraction
and increased grooming and freezing behaviors. Reduction in locomotor and exploratory activities may derive from reduced excitability of the central nervous system due to interaction with sedatives/anxiolytics or anticonvulsants on concurrent use.

**CONCLUSION**

Findings from this study showed that leaves of *O. gratissimum* contain constituents which possess anticonvulsant and anxiolytic-like activities. Studies aimed at isolating the anticonvulsant constituents are ongoing.

**REFERENCES**

1. Iwu MM. Handbook of African medicinal plants. CRC Press Inc. Boca Raton: Florida; 1993.
2. Dalziel JM. Useful plants of West tropical Africa. Crown Agents for Overseas Governments: London; 1956.
3. Oliver B. Medicinal plants in Nigeria. Nigerian College of Arts, Science and Technology: Ibadan, Nigeria; 1960.
4. Di Stasi LC, Oliveira GP, Carvalhaes MA, Queiroz M Jr, Tien OS, Kakinami SH, et al. Medicinal plants popularly used in the Brazilian Tropical Atlantic Forest. Fitoterapia 2002;73:69-91.
5. Madeira SV, Matos FJ, Leal-Cardoso JH, Criddle DN. Relaxant effects of the essential oil of *Ocimum gratissimum* on isolated ileum of the guinea-pig. J Ethnopharmacol 2002;81:1-4.
6. Aziba PI, Bass D, Elegbe Y. Pharmacological investigation of *Ocimum gratissimum* in rodent for analgesic activity. Phytother Res 1999;13:427-9.
7. Rabelo M, Souza EP, Soares PM, Miranda AV, Matos FJ, Criddle DN. Antinociceptive properties of the essential oil of *Ocimum gratissimum* L. (Labiatae) in mice. Braz J Med Biol Res 2003;36:521-4.
8. Mohammed A, Tanko Y, Okasha MA, Magaji RA, Yaro AH. Effects of aqueous leaves extract of *Ocimum gratissimum* on blood glucose levels of streptozocin-induced diabetic Wistar rats. Afr J Biotech 2007;6:2087-90.
9. Freire CM, Marques MO, Costa M. Effects of seasonal variation on the central nervous system activity of *Ocimum gratissimum* L. essential oil. J Ethnopharmacol 2006;105:161-6.
10. Nakamura CV, Ueda-Nakamura T, Bando E, Melo AF, Cortez DA, Dias Filho BP. Antibacterial activity of *Ocimum gratissimum* L. essential oil. Mem Inst Oswaldo Cruz 1999;94:675-8.
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Shigellocidal properties of three Nigerian medicinal plants: Ocimum gratissimum, Terminalia avicennoides and Momordia balsamina. J Health Popul Nutr 2001;19:331-5.

12. Vieira RF, Grayer RJ, Paton A, Simon JE. Genetic diversity of Ocimum gratissimum L based on volatile oil constituents, flavonoids and RAPD markers. Biochem Syst Ecol 2001;29:287-04.

13. Matsayoh LG, Matsayoh JC, Wachira FN, Kinyua MG, Muigai AWT, Mukiama TK. Chemical composition and antimicrobial activity of the essential oil of Ocimum gratissimum L growing in Eastern Kenya. Afr J Biotech 2007;6:760-5.

14. Keita SM, Vincent C, Jean-Pierre S, Belanger A. Essential oil composition of Ocimum basilicum L, O. gratissimum L, and O. suave L in the Republic of Guinea. Flav Frag 2000;15:339-41.

15. Harborne JB. Phytochemical methods. Chapman and Hall: London; 1973.

16. Trease GE, Evans WC. Textbook of Pharmacognosy. 12th ed. Balliere Tindall: United Kingdom; 1983.

17. Lorke D. A new approach of practical acute toxicity testing. Arch Toxicol 1983;54:272-89.

18. Akah PA, Sampson A, Gamaniel K, Wambebe C. Effect of coconut water on the activity of some centrally acting drugs. Indian Drugs 1998;35:693-5.

19. Archer J. Tests for emotionality in rats and mice: A review. Anim Behav 1973;21:205-35.

20. Corda MG, Giorgi O, Longoni B, Orlandi M, Biggio G. Decrease in the function of γ-aminobutyric acid coupled chloride channel produced by repeated administration of pentylenetetrazole in rats. J Neurochem 1990;55:1216-21.

21. Nisar M, Khan I, Simjee SU, Gilani AH, Obaidullah, Perveen H. Anticonvulsant, analgesic and antipyretic activities of Taxus wallichiana Zucc. J Ethnopharmacol 2008;116:490-4.

22. Ozturk Y, Aydine S, Baser KH, Berberoglu H. Effects of Hypericum perforatum L. and Hypericum calycinum L. extracts on the central nervous system in mice. Phytomed 1996;3:139-46.

23. Perez RM, Perez JA, Garcia LM, Sossa H. Neuropharmacological activity of Solanum nigrum fruit. J Ethnopharmacol 1998;62:43-8.

24. Prut L, Belzung C. The open field as a paradigm to measure the effects of drugs on anxiety-like behaviors: A review. Eur J Pharmacol 2003;46:3-33.

25. Walsh RN, Cummins RA. The open-field test: A critical review. Psychol Bull 1976;83:482-504.

26. Choleris E, Thomas AW, Kavaliere M, Prato FS. A detailed ethological analysis of the mouse open field test: Effects of diazepam, chlordiazepoxide and an extremely low frequency pulsed magnetic field. Neurosci Biobehav Rev 2001;25:235-60.

27. Dallmeier K, Carlini EA. Anesthetic, hypotermic, myorelaxant and anticonvulsant effects of synthetic eugenol derivatives and natural analogues. Pharmacology 1981;22:113-27.

28. Elisabetsky E, Brum LF, Souza DO. Anticonvulsant properties of linalool in glutamate-related seizure models. Phytomedicine 1999;6:107-13.

29. Brum LF, Elisabetsky E, Souza D. Effects of linalool on MK801 and [(3)H] muscimol binding in mouse cortical membranes. Phytother Res 2001;15:422-5.

30. Viana GS, do Vale TG, Silva CM, Matos FJ. Anticonvulsant activity of essential oil and active principles from chemotypes of Lippia alba (Mill) NE Brown. Biol Pharm Bull 2000;23:1314-7.

31. do Vale TG, Furtado EC, Santos JG Jr, Viana GS. Central effects of citral, myrcene and limonene, constituents of essential oil chemotypes from Lippia alba (Mill) NE Brown. Phytomedicine 2002;9:709-14.

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