Proximate Composition, Phytoconstituents and Mineral Contents of Soybean (Glycine Max) Flour Grown and Processed in Northern Nigeria

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Abstract: Soybean (Glycine max), is a species of legume widely grown for its edible bean which has numerous uses. Soybeans are an important source of not only proteins, but carbohydrates, fats, and many essential vitamins and minerals. So, they facilitate normal growth and development in young children. In this study, the phytoconstituents, proximate composition and mineral contents of soybean flour was evaluated using standard processing techniques. The preliminary phytochemical screening indicated the presence of phenols, cardiac glycosides, steroids, saponins, flavonoids in both the methanol and aqueous extracts. The quantitative phytochemical screening showed that saponin had the highest content (18.4%) followed by phenol (16.8%) and flavonoids (12.4%) while phytate was the least (0.07%). The proximate composition was in this order: protein > carbohydrate > crude fat > moisture > crude fibre > ash content which showed that the soy flour is relatively high in protein. The minerals analyzed for were: Ca (231.6), Fe (5.790), Mg (249.8), Zn (2.414) and Mn (0.651) which were within the FAO/WHO standards for metals in foods.

Keywords: Soybean, Phytoconstituents, Protein, Soy Flour, Saponin

1. Introduction

The soybean (Glycine max (L.) Merrill) belongs to the legume family, native to Eastern Asia and introduced into Nigeria in 1908. The main producers of soy are the United States (36%), Brazil (36%), Argentina (18%), China (5%) and India (4%). [1]. Soybean is a leguminous vegetable of the pea family that grows in tropical, subtropical, and temperate climates. It has long been recognized as a plant food that, when compared with other plants, is relatively high in protein. The plant is classed as an oilseed rather than a pulse by the UN Food and Agriculture Organization (FAO). It is an important inexpensive food crop that contains about 40% protein, 30% carbohydrates, excellent amounts of dietary fiber, vitamins, minerals and 20% oil which makes it second only to groundnuts in terms of oil content amongst food legumes [2]. Many leguminous crops provide some protein, but soybean is the only available crop that provides an inexpensive and high quality source of protein comparable to meat, poultry and eggs [3]. Historically it is nick-named "meat of the field" or "meat without bones" because it is the only vegetable with a complete protein [4]. The gold standard for measuring protein quality is the Protein Digestibility Corrected Amino Acid Score (PDCAAS) and by this criterion soy protein is the nutritional equivalent of meat, eggs, and casein for human growth and health [5].

Over the years, soybean consumption has been a matter of much debate. But recently researchers have taken a very close look at the protein content of soybeans and arrived at many fascinating conclusions. Along with this increasing interest in soy protein came the discovery of very unique proteins, typically referred to as "peptides" which include defensins, glycinins, conglycinins and lunasin, and all are now known to provide us with health benefits in the areas of
Phytate and oxalates but these anti-nutritional factors are bloating and nausea. It can also cause allergic reactions particularly those in low-income classes, are weaned on most populations. Children in most developing countries, many parts of Africa as animal protein is too expensive for farmers [10].

Where fertilizers are hardly available and are expensive for farmers 

Malnutrition, mostly protein deficiency, is prevalent in many parts of Africa as animal protein is too expensive for most populations. Children in most developing countries, particularly those in low-income classes, are weaned on cheap, readily available starchy foods such as pap in which soybean flour is mainly used to supplement this popular weaning and breakfast recipe [11].

The beans can be processed in a variety of ways. Common forms of soy include soy meal (the material remaining after solvent extraction of oil from soybean flakes, with a 50% soy protein content. The meal is ‘toasted’ and ground in a hammer mill. Soybean meal is an essential element of the American production method of growing farm animals, such as poultry and swine, and more recently the aquaculture of catfish); soy flour; soy milk; tofu; tempeh; miso; natto and soybean oil (a vegetable oil extracted from the seeds of the soybean) [12].

Soy flour is used as an ingredient in foods, beverages, and condiments to promote higher optimal health benefit. However, soy can cause some mild side effects such as bloating and nausea. It can also cause allergic reactions involving rash and itching in some people. There is concern that taking high doses might cause abnormal tissue growth in the uterus. So while soy in moderation can be beneficial to your health, too much of it may cause a few health problems. But alongside of the many controversies swirling around soybeans and health, it’s important not to lose sight of the strong nutritional value of this legume [13].

Soybean seed, like other legumes, is known to contain significant amounts of all the essential amino acids, minerals, phytate and oxalates but these anti-nutritional factors are usually removed or greatly reduced by soaking, fermentation and hydrothermal treatment during processing [14]. In recent times, concerns over the possible make-up of this wonderful legume due to its numerous health benefits made this research work absolutely imperative.

2. Materials and Methods

2.1. Sample Collection and Preparation

The soybean (Glycine max (L.) Merrill) seeds used for this work were purchased from Kubwa market in Bwari Area Council, Abuja, Nigeria. It was transferred to the laboratory, sieved to remove stones and debris, washed and soaked for 48 hours, oven dried at 40°C for 5 minutes and dehulled with the aid of hammer mill. The dehulled beans was ground finely enough with the use of a specialized Alpine Fine Impact Mill, model A90BD to pass through a 0.25mm screen [15]. The so made soy flour was packaged in moisture-proof, air-tight polyethylene containers and kept at 4°C prior to analyses.

2.2. Methanol Extract

The methanol extract of the soybean flour was prepared by soaking 20 g of the sample in 200 ml of methanol for 12 hours. The extract was then filtered through a funnel lined with Whatman No. 42 filter paper. The filtrate was concentrated and stored in an airtight container.

2.3. Aqueous Extract

The aqueous extract of the soybean flour was prepared by soaking 20 g of the sample in 400 ml of distilled water for 12 hours. The extract was then filtered using filter paper, concentrated and stored in an airtight container.

3. Qualitative Phytochemical Constituents

A small portion of the filtrate was used for the qualitative phytochemical screening using standard methods [16, 17].

3.1. Test for Tannins (Braymer’s Test)

0.5 g of extract was stirred with 10 mls of distilled water and then filtered. 2 ml of the filtrate was treated with 1% alcoholic ferric chloride solution and observed for formation of blue-black or greenish colour solution.

3.2. Test for Phenols (Ferric Chloride Test)

The extract (50 mg) was dissolved in 5 ml of distilled water. To this few drops of neutral 5% ferric chloride solution was added. A dark green colour indicates the presence of phenolic compound.

3.3. Test for Cardenolides

2 ml of benzene was added to 1 ml of the sample extract. The formation of a turbid brown colour indicated the presence of cardenolides.

3.4. Test for Cardiac Glycosides (Keller Kelliani’s Test)

5ml of each extract was treated with 2ml of glacial acetic acid in a test tube and a drop of ferric chloride solution was
added to it. This was carefully underlayed with 1ml concentrated sulphuric acid. A brown ring formation at the interface indicated the presence of deoxysugar characteristics of cardiac glycosides.

3.5. Test for Phlobatannins (Precipitate Test)

Deposition of a red precipitate when 2 mls of extract was boiled with 1ml of 1% aqueous hydrochloric acid was taken as evidence for the presence of phlobatannins.

3.6. Test for Steroids (Libermann-Burchard Test)

To 2 ml of the test solution, a few drops of chloroform, 3 - 4 drops of acetic anhydride and one drop of concentrate sulphuric acid were added. Appearance of purple colour, which changes to blue or green colour, showed the presence of steroid.

3.7. Test for Saponins (Foam Test)

To 2mls of extract was added 6ml of water in a test tube. The mixture was shaken vigorously and observed for the formation of persistent foam that confirmed the presence of saponins.

3.8. Test for Volatile Oil

For volatile oil estimation 50 mg of extract was taken and subjected to hydro-distillation. The distillate was collected in graduate tube of the assembly, wherein the aqueous portion automatically separated out from the volatile oil.

3.9. Test for Flavonoids (Alkaline Reagent Test)

2 ml of extracts was treated with few drops of 20% sodium hydroxide solution. Formation of intense yellow colour, which becomes colourless on addition of dilute hydrochloric acid, indicated the presence of flavonoids.

3.10. Test for Carbohydrates (Molisch’s Test)

Few drops of Molisch’s reagent were added to 2ml portion of the various extracts. This was followed by addition of 2ml of conc. H2SO4 down the side of the test tube. The mixture was then allowed to stand for two-three minutes. Formation of a red or dull violet colour at the interphase of the two layers was a positive test.

4. Quantitative Phytochemical Constituents

4.1. Test for Phenol

This was determined using spectrophotometric method. The sample was boiled with diethyl ether for 15 mins. 5 ml of the boiled sample was then pipette into 50 ml flask and 5 ml of distilled water was added. Then, 2 ml of NH4OH solution and 5 ml of conc. butanol was added to the mixture. The sample was made up to mark and left for 30 mins. to react for colour development. The absorbance was measured at 505 nm [18].

4.2. Test for Saponin

0.5 g of the sample was added to 20 ml of 1 N HCl and was boiled for 4 hours. This was filtered after cooling and 50 ml of petroleum ether was added to the filtrate for ether layer and evaporated to dryness. 5 ml of acetone-ethanol was added to the residue. 0.4 ml each was taken into 3 different test tubes, 6 ml of FeSO4 reagent was added into them followed by 2 ml of concentrated H2SO4 acid. It was thoroughly mixed after 10 mins. and the absorbance was taken at 490 nm [19, 20].

4.3. Test for Flavonoid

This was done by acid hydrolysis of spectrophotometric method. 0.5 g of the sample was mixed with 5 ml of dilute HCl acid and boiled for 10 mins. the boiled extract was allowed to cool and then filtered. 1 ml of the filtrate was added to 3 ml of ethyl acetate and 5 ml of 1% NH3. This was then scanned from 420 -520nm for the absorbance [21].

4.4. Taste for Phytate

This was determined by the photometric method of Latta and Esin [22]. 2 g of the test sample was extracted with 100 ml of 2.4% HCl acid by shaking vigorously in a vortex mixer for 1 hour and then filtered through Whatman No. 5 filter paper. 5 ml of the filtrate was mixed with 1 ml of 0.1M Na-EDTA, 0.75 M NaOH solution and then made up to 25 ml with distilled water before being placed on an ion-exchange (AGIX4, 100-200mesh) column. The column was washed with 15 ml of distilled water and then 15 ml of 0.1 m NaCl before being eluted with 15 ml of 0.7 M NaCl. The eluate was collected and wet digested in a Kjeldahl apparatus with 0.5 ml conc. H2SO4 and 3 ml HCl. The digest was cooled to room temperature, 10 ml of distilled water added and the mixture heated again on a water bath with 2 ml of 2.5% ammonium molybdate solution in 1 N H2SO4, 1 ml conc. H2SO4 and then made up to 50 ml in a volumetric flask. The solution was allowed to stand for 15 mins before taking the absorbance at 640 nm against a blank. This was done in triplicate [23].

4.5. Test for Oxalate

Deionized water was heated at 29°C and the sample was soaked for 1 minute. 1 ml of 5 M NaOH, 1 ml of 5% CaCl2 and 3 drops of phenolphthalein was added to 5 ml of extract and allowed to stand for 3 hours. This was centrifuged at 300 rpm for 15 mins. The supernatant was discarded and the precipitate was washed with hot water and allowed to centrifuge. 2 ml of 3 M H2SO4 was pipette into it and transferred into a conical flask. The precipitate was dissolved by warming in a water bath at 80°C. The content of each tube was then titrated with freshly prepared 0.01M K2MnO4. The titration was carried out at ordinary temperature until a first colour change was observed. It was allowed to stand until the changed to colourless. The solution was warmed at 70°C and
titration was carried out until a pink colour persists at least for 30 seconds [24].

5. Proximate Composition

The proximate analysis (moisture, ash, crude protein, crude fibre, crude fat and carbohydrate) of the sample was determined using the standard method of AOAC [25]. All determinations were done in triplicate and reported in percentage.

Table 1. Qualitative analysis of phytochemical constituents of soy flour.

| Phytochemical parameters | Methanol extract | Aqueous extract |
|--------------------------|------------------|-----------------|
| Tannins                  | -                | -               |
| Phenols                  | +                | +               |
| Cardenolides             | -                | -               |
| Cardiac glycosides       | +                | +               |
| Phlobatannins            | -                | -               |
| Steroids                 | +                | +               |
| Saponins                 | +                | +               |
| Volatile oils            | -                | -               |
| Flavonoids               | +                | +               |
| Carbohydrates            | -                | +               |

Key: (+) = present, (-) = absent

Table 2. Quantitative analysis of phytochemical constituents.

| Variables | Percentage (%) |
|-----------|----------------|
| Phenols   | 16.8           |
| Saponins  | 18.4           |
| Flavonoids| 12.4           |
| Phytates  | 0.07           |
| Oxalates  | 0.15           |

Table 3. Proximate Composition of Soybean flour.

| Parameters     | Values (%) (n=3) |
|----------------|------------------|
| Moisture       | 6.84             |
| Ash            | 4.92             |
| Crude Protein  | 35.6             |
| Crude Fibre    | 6.27             |
| Crude Fat      | 18.5             |
| Carbohydrate   | 27.87            |

Table 4. Mineral contents of Soybean flour (mg/kg).

| Parameters | Values | FAO/WHO guidelines |
|------------|--------|--------------------|
| Ca         | 231.6  |                    |
| Fe         | 5.790  | 48                 |
| Mg         | 249.8  | 300                |
| Zn         | 2.414  | 60                 |
| Mn         | 0.651  | 1.0                |

6. Results and Discussion

The results of the phytoconstituents of Glycine max (L.) Merrill as determined are recorded in tables 1 and 2 while the proximate compositions and the mineral contents are presented in tables 3 and 4. Of the fifteen phytochemicals screened for, five were found present in two solvent extracts. The preliminary phytochemical screening indicated the presence of phenols, cardiac glycosides, steroids, saponins, flavonoids in both the methanol and aqueous extracts. In addition, the aqueous extract also showed the presence of carbohydrate while it was absence in the methanol extract. From the active compounds indicated, it is obvious that Glycine max (L.) Merrill is a potential source of useful drugs. From table 2, saponin had the highest percentage content (18.4%) followed by phenol (16.8%) and flavonoids (12.4%) while phytate was the least (0.07%).

Saponins are a class of chemical compounds found in particular abundance in various plant species. More specifically, they are amphipathic glycosides grouped phenomenologically by the soap-like foaming they produce when shaken in aqueous solutions, and structurally by having one or more hydrophilic glycoside moieties combined with a lipophilic triterpene derivative [26]. In other words, saponins are sterols that are present naturally in a wide variety of food-plants, including vegetables, legumes and cereals. Legumes such as soybean and chickpeas are the major source of saponins in form of soyasapogenins (group A and group B) and soyasapogenols in the human diet [27]. Soybeans also contain the isoflavonones genistein and daidzein, types of phytoestrogen which are closely related to the antioxidant flavonoids. Genistein is a soy component that could easily be singled out for its antioxidant properties! Increased activity of antioxidant enzymes—including superoxide dismutase, glutathione peroxidase, catalase, and glutathione reductase—has now been linked to intake of genistein from soy [28]. Another group of antioxidant phytonutrients called phenolic acids has also been recently investigated in soybeans. When we enjoy this antioxidant-rich legume, we also benefit from its phenolic acids, including caffeic, coumaric, ferulic, and sinapic acid. Phytic acid in soybeans has many effects including acting as an antioxidant and a chelating agent. It helps in reducing cancer, minimizing diabetes and reducing inflammation. However, phytic acid is also criticized for reducing vital minerals due to its chelating effect, but this effect is greatly reduced during processing [29].

From table 3, crude protein had the highest value (35.6%) while moisture content (4.84%) was the lowest which showed hindrance to microorganism growth. Most soy protein is a relatively heat-stable storage protein. This heat stability enables soy food products requiring high temperature cooking to be made. Soybeans are considered by many agencies to be a source of complete protein. For this reason, soy is a good source of protein, amongst many others, for people who want to reduce the amount of meat they eat. According to the US Food and Drug Administration, soy protein products can replace animal-based foods—which also have complete proteins but tend to contain more fat, especially saturated fat—without requiring major adjustments elsewhere in the diet. Even soy-based infant formula (SBIF) is sometimes given to infants who are not being strictly breastfed because of its richness in protein; it can be useful for infants who are either allergic to pasteurized cow milk proteins or who are being fed a vegan diet. Diverse studies have concluded there are no adverse effects in human growth, development, or reproduction as a result of the consumption of soy-based infant formula. SBIFs provide
7. Conclusion

Soybeans are perhaps best known for their fantastic blend of protein and fiber. They are a very good source of manganese, phosphorus, and protein as well as a good source of iron, omega-3 fatty acids, dietary fiber, magnesium, and vitamin K. There are also a wide range of unique proteins, peptides, and phytonutrients contained in soy which include flavonoids, isoflavonoids, phenolic acids and saponins (soyasaponins from group A and group B, and soyasapogenols) which are very useful for optimal health. Soy is used for high cholesterol, high blood pressure, and cancer, prostate cancer, and thyroid cancer, as well as preventing weak bones (osteoporosis), and slowing the progression of kidney disease. In foods, soy is used as a milk substitute in infant feeding formulas, and as an alternative to cow's milk. Soybeans are eaten boiled or roasted. Soy flour is used as an ingredient in foods, beverages, and condiments. Other food sources contain these nutrients, but soybean is readily available and affordable. We recommend that persons wanting to use soy in treatment of any ailment need to pay attention to the form of the soy, the amount consumed, their personal health history, and in some cases, the advice of their healthcare provider.

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