Rapid spread of OXA-244-producing Escherichia coli ST38 in Germany: insights from an integrated molecular surveillance approach; 2017 to January 2020

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Annually, increasing numbers of OXA-244-producing Escherichia coli in 13 German federal states prompted us to initiate an outbreak investigation. Whole genome sequencing revealed that among 148 isolates analysed, most belonged to sequence type 38 with 62 isolates forming a genetically distinct cluster. Although no epidemiological link could be identified between cases, ongoing investigations suggest non-healthcare associated transmission. A screening-PCR was developed facilitating early detection of ST38 cluster isolates to identify the source and transmission route.

A rise in detection of community-acquired infections with OXA-244-carbapenemase-producing Escherichia coli in at least nine European countries prompted a rapid risk assessment (RRA) by the European Centre for Disease Prevention and Control (ECDC) in the beginning of 2020 [1]. The rise in detection was deemed worrisome as despite of challenging detection of the chromosomally-encoded carbapenemase, a rapid and simultaneous increase was found. Genetically distinct clusters, predominantly of sequence type (ST) 38 were identified, thus a common source and a transmission via contaminated food products could not be ruled out. Germany reported the highest numbers of detected isolates among the affected countries.

Here we provide details on our integrated molecular surveillance (IMS) results and consecutive outbreak investigations in Germany that led to the RRA. Our IMS involved molecular investigation via whole genome sequencing (WGS) to identify relatedness of isolates, followed by epidemiological investigations of linked notified carbapenem-resistant E. coli cases to understand modes of transmission and to identify possible sources. A newly developed screening-PCR will facilitate rapid detection of the ST38 cluster isolates.

Molecular investigation of OXA-244-producing Escherichia coli
Between January 2017 and the end of February 2019, a total of 144 OXA-244-producing E. coli isolates were identified at the German National Reference Centre for multidrug-resistant Gram-negative bacteria (NRC) in Bochum (Figure 1), of which 86 isolates with information on hospital origin or diagnostic laboratory were subjected to Illumina (Illumina, San Diego, United States) WGS. As continued increase was noted and preliminary results pointed towards an ongoing nationwide spread [2], our analysis focused on most recent OXA-244-producing E. coli isolates from March 2019 onwards including those with only partially complete metadata.

Until the end of August 2019, 152 isolates were subjected to WGS with 148 isolates having sufficient sequence data for further analyses which included in silico multilocus sequence typing (MLST) and core genome (cg)MLST using the E. coli scheme (2,513 loci) as implemented in the SeqSphere+ software version 6.0.0 (Ridom, Muenster, Germany). Among the 148 isolates analysed, we identified 16 different sequence types (ST), with a dominance of ST38 (n=100). Results of cgMLST revealed close genetic relatedness of ST38 isolates, particularly among 62 isolates forming a distinct cluster with a maximum of 27 allelic differences.
among core genes, and a distance of >118 alleles to other ST38 isolates (Figure 2). By the end of January 2020, the NRC had identified a total of 243 OXA-244-producing *E. coli* isolates indicating a continuous spread in Germany (Figure 1).

**Epidemiological investigation**

To infer public health implications of closely related isolates by WGS, additional investigations for epidemiological association are required [3]. Our epidemiological investigation focused on the predominant cluster of 62 OXA-244-producing *E. coli* ST38 isolates. Isolates from cases were mapped based on the postal code of hospital of origin (where available) or diagnostic laboratory, revealing a supra-regional dissemination over 13 of the 16 German federal states (Figure 3) without a discernible pattern. Available information (specimen type, date of detection and patient age) for each isolate was extracted from the anonymized NRC database and used for linking with a second data set. In 2016 Germany has introduced a mandatory surveillance for colonisation or infection with carbapenem-resistant Enterobacterales and *Acinetobacter* spp.. We linked the available laboratory data with information from the national electronic reporting system for notifiable diseases. This enabled us to obtain additional case information and to identify the responsible health authority. Overall, 38 of 62 isolates could be linked (Figure 3). For these linked cases, additional personal data were available: sex, infection/colonisation ratio and age (age median: 56 years; age range: 0–82 years) were equally distributed (Table 1). All responsible public health authorities of affected German federal states were individually notified in August and October 2019. Together with the local public health authorities, we collected further case information via an exploratory questionnaire (Supplement 1) (response rate: 24/38, 63%), as summarised in Table 1. In the following all available information is summarised, the denominator varies as missing information was not counted. Of cases with available information, briefly, 27 of 36 cases were hospitalised at the time of detection of OXA-244-producing *E. coli*; 14 of 21 cases had a history of hospitalisation before their current hospital stay; eight of 18 cases had an underlying disease. Although nine of 14 cases had a stay abroad within 12 months before detection, no geographical areas with predominance were identified. Interestingly, half of the cases

**Figure 1**

Number of detected OXA-244-producing *Escherichia coli* isolates by month, Germany, January 2017–January 2020 (n = 243)
were detected via hospital admission screening or at outpatient treatment (12/24). These findings, and the absence of any common healthcare-related exposure or person-to-person transmission for closely related isolates, pointed towards transmissions outside healthcare settings.

Therefore, we further investigated community-related modes of transmission through a trawling questionnaire, interviewing the most recent notified cases to limit recall error by delay of interview [4] (Supplement 2). In six interviews, no common diet or common exposures could be identified among the cases, but in all households meat was processed and consumed. (Table 1).

**International alert**

Since the spread still continued, on 20 December 2019, our major findings were internationally shared via the European Early Warning and Response System (EWRS) and Epidemic Intelligence Information System (EPIS) of the ECDC. This led to the aforementioned ECDC RRA and identification of spread in further European countries [5].
Figure 3
Distribution of cases with OXA-244-producing *Escherichia coli* ST38 isolates by year and notification status, Germany, January 2017–January 2020 (n = 62)

Cases by hospital’s or submitting lab’s 3-digit post code
- 2017 (non-linked cases) (n = 2)
- 2017 (linked cases) (n = 2)
- 2018 (non-linked cases) (n = 10)
- 2018 (linked cases) (n = 9)
- 2019 (non-linked cases) (n = 12)
- 2019 (linked cases) (n = 27)

NRC: German National Reference Centre for multidrug-resistant Gram-negative bacteria. Year of isolate receipt at NRC are colour-coded in blue. Notified cases are labelled with diagonal stripes. Cases are mapped by 3-digit postcode of the sample collection locality (hospital) or alternatively of reporting laboratory. Background colour (orange) indicates population density.
**Table 1**

Case characteristics from notification and National Reference Centre for multidrug-resistant Gram-negative bacteria data as well as results of exploratory questionnaires and interviews, Germany, January 2017–January 2020 (n = 62)

| Characteristics                                      | Total cases (n = 62) | Non-linked cases (n = 24) | Linked cases (n = 38)* | Interviewed linked cases (n = 6) |
|------------------------------------------------------|----------------------|---------------------------|------------------------|---------------------------------|
| **Sex**                                              |                      |                           |                        |                                 |
| Male                                                 | 17                   | NA                        | 17                     | 3                               |
| Female                                               | 21                   | NA                        | 21                     | 3                               |
| Unknown                                              | 24                   | 24                        | 0                      | 0                               |
| **Median age (years) (range)**                       | 55 (0–86)            | 49.5 (0–86)               | 56 (0–82)              | 24 (0–62)                       |
| **Age groups (years)**                               |                      |                           |                        |                                 |
| < 3                                                  | 9                    | 3                         | 6                      | 1                               |
| 3–17                                                 | 6                    | 1                         | 5                      | 1                               |
| 18–39                                                | 10                   | 6                         | 4                      | 2                               |
| 40–64                                                | 16                   | 5                         | 11                     | 2                               |
| ≥ 65                                                 | 19                   | 7                         | 12                     | 0                               |
| Unknown                                              | 2                    | 2                         | 0                      | 0                               |
| **Material of isolate**                              |                      |                           |                        |                                 |
| Rectal                                               | 27                   | 10                        | 17                     | 2                               |
| Urine                                                | 20                   | 7                         | 13                     | 3                               |
| Blood                                                | 8                    | 4                         | 4                      | 0                               |
| Intraabdominal                                       | 1                    | 0                         | 1                      | 1                               |
| Respiratory                                          | 1                    | 0                         | 1                      | 0                               |
| Wound                                                | 1                    | 0                         | 1                      | 0                               |
| Other                                                | 4                    | 3                         | 1                      | 0                               |
| **Status of infection**                              |                      |                           |                        |                                 |
| Colonised                                            | Unknown b             | NA                        | 12                     | 4                               |
| Infected                                             | Unknown b             | NA                        | 11                     | 2                               |
| Unknown                                              | Unknown b             | NA                        | 15                     | 0                               |
| Hospitalisation at detection, Yes c                  | Unknown b             | NA                        | 27/36                  | 4                               |
| Previous Hospitalisation (<12 months), Yes c         | Unknown b             | NA                        | 14/21                  | 4                               |
| Detection via admission screening or outpatient treatment, Yes c | Unknown b   | NA                        | 12/24                  | 3                               |
| Underlying disease, Yes c                           | Unknown b             | NA                        | 8/18                   | 1                               |
| Travel history (<12 months), Yes c                   | Unknown b             | NA                        | 9/14                   | 4                               |
| Contacts to farm animals (<12 months), Yes c         | Unknown b             | NA                        | 1/13                   | 1                               |
| Vegetarian or vegan diet, Yes c                      | Unknown b             | NA                        | Unknown d              | 1                               |

NA: not available.

* Some of the information for linked cases is from the notification data (n = 38) with additional information coming from the exploratory questionnaire (n = 24).

b Information was only available from linked cases such that providing a total count for all cases was not possible.

c Denominator represents all cases with available information.

d Information on vegetarian or vegan diet was only obtained through trawling questionnaire and thus only available from interviewed linked cases.
Screening PCR for the identification of OXA-244 Escherichia coli ST38 cluster isolates

To accomplish early collection of patients’ epidemiological information for the identification of potential sources and routes of transmission, we developed a PCR assay specific for OXA-244-producing E. coli ST38 cluster isolates enabling nearly real-time screening. For this, we performed SNP analyses as implemented in Enterobase [5] to identify genetic differences of isolates. Using the assembly with the highest N50 value of a ST38 cluster isolate as reference genome, SNP analyses based on a total of 227,099 variable sites confirmed cgMLST results (Figure 4). Comparison of alignments revealed a ca 34 kb insertion that was carried only by ST38 cluster isolates (Figure 4) corresponding to an intact Mu-like bacteriophage, as annotated by rapid annotations using subsystems technology (RAST) and PHAge Search Tool Enhanced Release (PHASTER) analyses [6,7]. However, this segment was completely lacking in one isolate, whereas in two other isolates the prophage was not fully integrated but sequences upstream of the transposase gene were present (Figure 4). Based on the phage sequences found in 61 of the 62 ST38 cluster isolates, a specific PCR assay was designed to amplify a 514-bp-fragment partially encompassing the gene that encodes the repressor protein CI (Table 2). WGS results for 144 of the 148 isolates analysed indicated a high concordance rate (97.3%). However, two false positives (specificity 98.6%) as well as two false negatives (sensitivity 98.6%) were observed in our sample collection including the ST38 cluster isolate for which absence of the prophage was known. It has to be noted that the screening approach described here might only be valid for E. coli isolates known to carry the blaOXA-244, as ongoing analyses suggest that the prophage is also integrated in other ST38 contexts.

Ethics

A formal ethical review was not required for the outbreak investigation in accordance with Article 25, Section 1 of the German Infection Protection Act of 2001.

Discussion

Here, we used for the first time a systematic IMS approach for carbapenemase-producing Enterobacterales (CPE) in Germany by combining WGS data and epidemiological analyses on a national scale. Our results provide further evidence for the rapid and ongoing spread of blaOXA-244 [1,8]. In Germany, the spread was predominantly driven by genetically-clustered ST38 E. coli. Although instances of sporadic nosocomial infections cannot be excluded, it is unlikely that overall patterns reflect sustained healthcare-driven transmissions on a
supra-regional scale. Instead, available data suggest that infections are due to non-hospital-acquired transmissions as most cases were detected upon admission or during outpatient treatment. Notably in this regard, younger age groups were more affected in comparison to overall notifications of CRE in Germany: nearly half of cases were below 50 years compared with 18% of all CRE of which 91% were hospitalised in 2018 [9].

Although, worldwide, OXA-244-producing E. coli were detected in cases in Columbia [10] or cases were reported with epidemiological links to Indonesia [11], our investigations revealed no association with travel. Furthermore, no direct links to animals or certain food items were found. However, specific food products with a broad dissemination might be one possible vehicle to explain global spread. Carbapenemases were previously detected in various Enterobacterales species from food, livestock and companion animals in Germany and other European countries, although blaOXA-244 was not identified [12,13]. ST38 is known to be abundant in livestock, such as poultry, and has also been identified along the food production chain [14,15]. Although results of the trawling questionnaire remained ambiguous, food might be a possible vehicle. Ongoing molecular surveillance might reveal seasonal patterns in transmission risk thus enabling to narrow down potential sources of infection. Collaborating with food safety authorities, screening for clustered isolates should be considered and further interviews conducted accordingly. Multiple transmission hypotheses can be considered regarding the spread of OXA-244-producing E. coli isolates. The wide dissemination of these isolates in Europe [10,16,17] demonstrates the urgent need for additional investigations including additional data sources as well as enhanced screening capabilities [3].

Limitations

Isolates predominantly derived from hospitals and cases might not be representative for the underlying mode of transmission. Although this may hinder identification of community exposures, it strengthens the argument of a non-healthcare-related source in the absence of common links. Unknown stability and colonisation period of E. coli ST38 in the human gut, results in unknown incubation period, which limited the quality of the epidemiological investigations.

Although sensitivity as well as specificity of the PCR screening assay are high, blaOXA-244 has to be identified separately and is known be easily missed by phenotypic methods [17].

Conclusions

The ongoing spread of OXA-244-producing E. coli in the community is of great concern. Additional studies are needed to understand the driving forces responsible for the dissemination of blaOXA-244, which is not exclusively, but primarily driven by genetically-clustered E. coli ST38. In the absence of real-time WGS, the here described PCR screening assay may facilitate fast identification of ST38 cluster isolates and allow immediate epidemiological investigations of cases. Further cross-border data sharing and collaboration is needed to identify risk factors for OXA-244-producing E. coli colonisation, to investigate potential cross-border transmissions, to guide screenings of at-risk-population in healthcare settings and to guide control measures to stop dissemination in the community.

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Conflict of interest

None declared.

Authors’ contributions

KK, RK, SH, TE were part of the outbreak investigation team and conducted the epidemiological investigation. In addition, HS contributed to the investigations at the local level. GW and SG coordinated the typing investigations together with NP. JBH performed molecular work and analysed WGS data together with BN. KK and RK drafted the manuscript together with JBH. All authors contributed to the revision of the draft manuscript and approved the final version.

Table 2

| Primer sequences 5’ to 3’ | PCR reaction conditiona |
|--------------------------|-------------------------|
| Forward TGAACCAGAAAGATTGGTG | 95°C 5 min; 95°C 30 s, 58°C 30 s, 72°C 30 sec; 35 cycles |
| Reverse TCAATCCCATACGTTGA | 95°C 5 min; 95°C 30 s, 58°C 30 s, 72°C 30 sec; 35 cycles |

a In 25 μl containing: 1.8 mM MgCl₂, 200 μM dNTPs, 1 μM primers (each), 1.25 U AmpliTaq 360 Polymerase and 1 x AmpliTaq 360 Buffer (Thermo-Fisher Scientific, Waltham, Massachusetts, United States).
References

1. Vijayachari P, Sugunan AP, Shriram AN. Leptospirosis: European Centre for Disease Prevention and Control (ECDC). Rapid risk assessment: Increase in OXA-244-producing Escherichia coli in the European Union/European Economic Area and the UK since 2013. ECDC; Stockholm; 18 Feb 2020. Available from: https://www.ecdc.europa.eu/en/publications-data/rapid-risk-assessment-increase-oxa-244-producing-escherichia-coli-eu-eea

2. Hans JB, Kramer R, Kremer K, Pfennigwerth N, Haller S, Reuss A, et al. Ongoing outbreak of OXA-244 carbapenemase-producing Enterobacteriales in Germany is driven by clonal expansion of Escherichia coli ST38. In: 12th International Meeting on Microbial Epidemiological Markers; 2019 Sep 18-21; Dubrovnik, Croatia.

3. Besser JM, Carleton HA, Trees E, Stroika SG, Hise K, Wise M, et al. Interpretation of Whole-Genome Sequencing for Enteric Disease Surveillance and Outbreak Investigation. Foodborne Pathog Dis. 2019;16(7):504-12. https://doi.org/10.1089/fpd.2019.2650 PMID: 31246502

4. Gertler M, Czogielski I, Stark K, Wilking H. Assessment of recall error in self-reported food consumption histories among adults—Particularly delay of interviews decrease completeness of food histories-Germany, 2013. PLoS One. 2017;12(6):e0179121. https://doi.org/10.1371/journal.pone.0179121 PMID: 28640839

5. Zhou Z, Alikhan NF, Mohamed K, Fan Y, Achtmann M, Goorhuis A, Grobusch MP, et al. Prolonged carriage and IncK/blaCMY-2 resistance plasmids. Clin Microbiol Infect. 2017;23(6):407.e9-15. https://doi.org/10.1016/j.cmi.2016.12.035 PMID: 28082191

6. Arndt D, Grant JR, Marcu A, Sajed T, Pon A, Liang Y, et al. genome analyses of CMY-2-producing Escherichia coli isolates. Verrors A, et al. , RESET Study Group. Whole genome sequencing of Salmonella Typhimurium DT104 and Escherichia coli. Clin Microbiol Infect. 2016;22(5):E57-E65. https://doi.org/10.1016/j.cmi.2016.02.022 PMID: 27070438

7. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, et al. The RAST Server: rapid annotations using subsystems technology. BMC Genomics. 2008;9(1):75. https://doi.org/10.1186/1471-2164-9-75 PMID: 18261238

8. Masseron A, Poirel L, Falgenhauer L, Imirzalioglu C, Kessler J, Chakraborty T, et al. Ongoing dissemination of OXA-244 carbapenemase-producing Escherichia coli in Switzerland and their detection. Diagn Microbiol Infect Dis. 2020;97(3):115059. https://doi.org/10.1016/j.diagmicrobio.2020.115059 PMID: 32400363

9. Robert Koch Institut (RKI). Infektions epidemiologisches Jahrbuch für 2018. [Infectious Disease Epidemiology Annual Report – 2018] Berlin: RKI; 2019. Available from: https://www.rki.de/EN/Content/Infections/epidemiology/inf_dis_Germany/yearbook/yearbook_summer_2018.html

10. Abril D, Bustos Moya IG, Marquez-Ortiz RA, Josa Montero DF, Corredor Rozo ZL, Torres Molina I, et al. First Report and Comparative Genomics Analysis of a blaOXA-244-Harboring Escherichia coli isolate Recovered in the American Continent. Antibiotics (Basel). 2019;8(10):222. https://doi.org/10.3390/antibiotics8040222 PMID: 31766210

11. van Hattem JM, Arcilla MS, Bootsma MC, van Genderen PJ, Hogerhuis A, Grobusch MP, et al. Prolonged carriage and potential onward transmission of carbapenemase-producing Enterobacteriaceae in Dutch travelers. Future Microbiol. 2016;11(7):897-64. https://doi.org/10.2217/fmb.16.18 PMID: 27357522

12. Irgang A, Tenhagen B-A, Pauly N, Schmoger S, Kaesbohrer A, Hammer JA. Characterization of VIM-1-Producing E. coli isolated From a German Fattening Pig Farm by an Improved Isolation Procedure. Front Microbiol. 2019;10(2256):2256. https://doi.org/10.3389/fmicb.2019.02256 PMID: 31632372

13. Pulss S, Stolle I, Stamm I, Leidner U, Heydel C, Semmler T, et al. Multispecies and Clonal Dissemination of OXA-48 Carbapenemase in Enterobacteriaceae From Companion Animals in Germany, 2009-2016. Front Microbiol. 2018;9(1265):1265. https://doi.org/10.3389/fmicb.2018.01265 PMID: 29963026

14. Berg ES, Wester AL, Ahrenfeldt J, Mo SS, Slettemeås JS, Steinbak M, et al. Surveillance of OXA-244-producing Escherichia coli and epidemiologic investigation of cases, Denmark, January 2016 to August 2019. Euro Surveill. 2020;25(18):1900742. https://doi.org/10.2807/1560-7917.ES.2020.25.18.1900742 PMID: 32400363

15. Pietsch M, Irgang A, Roschanski N, Brenner Michael G, Hamprecht A, Rieber H, et al. , RESET Study Group. Whole genome analyses of CMY-2-producing Escherichia coli isolates from humans, animals and food in Germany. BMC Genomics. 2018;19(2):601. https://doi.org/10.1186/s12866-018-4976-3 PMID: 30092762

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