Secondary findings in inherited heart conditions: a genotype-first feasibility study to assess phenotype, behavioural and psychosocial outcomes

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Abstract

Disclosing secondary findings (SF) from genome sequencing (GS) can alert carriers to disease risk. However, evidence around variant-disease association and consequences of disclosure for individuals and healthcare services is limited. We report on the feasibility of an approach to identification of SF in inherited cardiac conditions (ICC) genes in participants in a rare disease GS study, followed by targeted clinical evaluation. Qualitative methods were used to explore behavioural and psychosocial consequences of disclosure. ICC genes were analysed in genome sequence data from 7203 research participants; a two-stage approach was used to recruit genotype-blind variant carriers and matched controls. Cardiac-focused medical and family history collection and genetic counselling were followed by standard clinical tests, blinded to genotype. Pathogenic ICC variants were identified in 0.61% of individuals; 20 were eligible for the present study. Four variant carriers and seven non-carrier controls participated. One variant carrier had a family history of ICC and was clinically affected; a second was clinically unaffected and had no relevant family history. One variant, in two unrelated participants, was subsequently reclassified as being of uncertain significance. Analysis of qualitative data highlights participant satisfaction with approach, willingness to follow clinical recommendations, but variable outcomes of relatives’ engagement with healthcare services. In conclusion, when offered access to SF, many people choose not to pursue them. For others, disclosure of ICC SF in a specialist setting is valued and of likely clinical utility, and can be expected to identify individuals with, and without a phenotype.

Introduction

Genome sequencing (GS) has been widely adopted by healthcare systems to investigate genomic contributions to rare disease and cancer [1]. GS coincidentally creates large amounts of data of potential health significance (secondary findings, SF), that are or may be unrelated to the presenting condition. SF are perceived both as an opportunity to predict and prevent disease [2] and a challenge to implementation of genomic medicine [3].

Inherited cardiac conditions (ICC) have a population prevalence of up to one in 200; a proportion result from a single variant in any one of a number of genes. ICC—which include hypertrophic cardiomyopathy (HCM), dilated cardiomyopathy (DCM), arrhythmogenic right ventricular cardiomyopathy (ARVC), long QT syndrome and Brugada syndrome—are often asymptomatic, yet pose a risk of complications including sudden cardiac death. Disease manifestation and risk of complications, including
arrhythmogenic events, can be assessed using standard cardiological tests based on imaging and ECG. Evidence-based risk strategies to manage risk include medical and lifestyle management, and cardiac defibrillator implantation [4–6].

Policy around search and return of SF will be informed by clinical actionability, including the positive predictive value of SF in populations not ascertained for the associated health condition. This question is more amenable to study in ICC than some other disorders, since disease expression can be assessed in any adult variant carrier through detailed clinical assessment at the time of presentation. In the context of manifest disease, family-based studies demonstrate the incomplete and variable penetrance of ICC variants [7–9]. While genotype-first approaches overcome the ascertainment bias inherent to case series and family-based studies, phenotypic features described are often derived from retrospective electronic medical records, lack specific ICC-targeted investigations, and hence may underestimate the magnitude of any effect [10–14]. The few reports of ICC SF disclosure highlight challenges with interpreting variants and correlating them with clinical findings and family history, and with patient management [15–17]. Pathogenicity assessment remains a challenge, especially in heterozygous disorders caused by missense variants [18].

The majority of patients, public and healthcare professionals support the return of ‘actionable’ SF, however, appreciation of the complexity of genomic information may temper attitudes to the scope of findings [19]. Increasingly, GS is expanding to ‘healthy’ cohorts; while recent initiatives have sought consent to return SF in variable gene lists, earlier ones did not, posing dilemmas relating to ethical recall-by-genotype studies [20].

Here we report on an approach to disclosure and targeted clinical evaluation of ICC SF in an established rare disease study cohort: a recall-by-genotype double blind, case-control feasibility study which aimed to assess the penetrance of ICC gene variants in an unselected population, and explore the psychosocial and behavioural sequelae of clinical assessment. The study, Secondary Cardiac Findings Evaluation (SCARFE), piloted a two-stage recruitment approach, designed to disclose SF information in an expert setting, and to enhance participant autonomy [21] in participants who were not offered SF options at initial recruitment.

### Materials and methods

#### NIHR BioResource for rare disease (BRRD)

BRRD ([https://bioresource.nihr.ac.uk/rare-diseases/rare-diseases/](https://bioresource.nihr.ac.uk/rare-diseases/rare-diseases/)) enrolled affected participants and their relatives across several different rare disease areas including infection and immunity, neuroscience, rare cancers and cardiovascular disease (Research Ethics Committee (REC) reference 13/EE/0325). Consent to BRRD included recontact for additional ‘Stage 2’ studies. The SCARFE study was approved by London Fulham research ethics committee (REC reference 17/LO/1579).

#### Genome sequencing

GS of DNA from blood samples from BRRD participants was undertaken as reported elsewhere [22].

#### Genomic analysis and participant re-identification

Twenty ICC genes robustly associated with HCM (ACTC1, GLA, LAMP2, MYL2, MYBPC3, MYL3, MYH7, TNNI3, TNNT2, TPM1), DCM (LMNA, VCL, MYH7, TNNI3, TNNT2, TPM1), ARVC (DSC2, DSG2, DSP, PKP2) and long QT syndrome (KCNH2, KCNQ1, SCN5A) were evaluated (Table 1) in sequence data from BRRD participants not known to have an ICC. Previously reported variants classified pathogenic in the context of autosomal dominant ICC (Oxford Medical Genetics Laboratory, OMGL; Partners Laboratory of Molecular Medicine [23]; or ClinVar), and truncating variants in genes where this class of variant is considered pathogenic, were extracted. Variants assigned pathogenic or likely pathogenic status were confirmed by Sanger sequencing by the BRRD. The TTN gene was not included since at the time of analysis, understanding of variant-disease association was incomplete and variants not considered actionable.

Codes of all participants harbouring a likely/highly likely pathogenic variant, together with variant details, were made available to the BRRD, who re-identified participants and assessed for eligibility: age 18–80 years, not known to have a condition associated with the SF, or that might confound clinical test interpretation, and matched with non-carrier controls by age, gender and primary disease cohort. Inclusion of matched non-variant carrier controls allowed us to approach BRRD participants without disclosing variant carrier status, to perform and analyse targeted tests while blind to genotype and to study impacts of the double blind approach on all participants.

#### Participant approach

Eligible BRRD participants and controls were initially sent an ‘opt-out’ letter [21] (Supplementary Material). Participants (cases and controls) who did not opt out within 4 weeks were sent an invitation letter to SCARFE requesting reply to the BRRD team, using a code for the
specific ICC associated with the variant. Contact details of participants who responded positively were passed to the SCARFE study team. Case/control status was blind to participants and SCARFE study team.

| Disease associated | Gene | Reference sequence | c.     | p.              | No. occurrences | Variant classification |
|--------------------|------|--------------------|--------|-----------------|-----------------|------------------------|
| ARVC               | DSC2 | NM_024422.3        | c.1123C>T | p.(Arg375*)     | 1               | Pathogenic             |
| ARVC               | DSC2 | NM_024422.3        | c.2533G>T (variant not confirmed) | p.(Glu845*) | 1 | Likely pathogenic     |
| ARVC               | DSC2 | NM_024422.3        | c.379G>T | p.(Glu127*)     | 1               | Likely pathogenic      |
| ARVC               | DSC2 | NM_024422.3        | c.939_942delAGAG | p.(Arg313fs) | 1 | Likely pathogenic     |
| ARVC               | DSC2 | NM_024422.3        | c.959delA | p.(Gln320fs)    | 1               | Likely pathogenic      |
| ARVC               | DSG2 | NM_001943.4        | c.1880-2A>G | p.?             | 1               | Pathogenic             |
| ARVC               | DSG2 | NM_001943.4        | c.2894_2897delAGAG | p.(Glu965fs) | 1 | Likely pathogenic     |
| ARVC               | DSG2 | NM_001943.4        | c.355C>T  | p.(Arg119*)     | 1               | Likely pathogenic      |
| ARVC               | DSG2 | NM_001943.4        | c.375dupT | p.(Leu126fs)    | 2               | Pathogenic             |
| ARVC               | DSG2 | NM_001943.4        | c.603_604delAT | p.(Leu203fs) | 1 | Likely pathogenic     |
| ARVC               | PKP2 | NM_004572.3        | c.1035-1G>C | p.?             | 1               | Likely pathogenic      |
| ARVC               | PKP2 | NM_004572.3        | c.1237C>T | p.(Arg413*)     | 1               | Pathogenic             |
| ARVC               | PKP2 | NM_004572.3        | c.2146-1G>C | p.?             | 1               | Pathogenic             |
| ARVC               | PKP2 | NM_004572.3        | c.2194C>T | p.(Gln732*)     | 1               | Pathogenic             |
| ARVC               | PKP2 | NM_004572.3        | c.378_2379dupCC | p.(Ser794fs) | 1 | Pathogenic            |
| ARVC               | PKP2 | NM_004572.3        | c.378_2379dupCC | p.(Ser794fs) | 1 | Pathogenic            |
| DCM                | LMNA | NM_170707.3        | c.949G>A  | p.(Glu317Lys)   | 1               | Likely pathogenic      |
| HCM                | GLA  | NM_000169.2        | c.644A>G  | p.(Asn215Ser)   | 1               | Pathogenic             |
| HCM                | MYBPC3 | NM_000256.3       | c.1504C>T | p.(Arg502Trp)  | 3               | Pathogenic             |
| HCM                | MYBPC3 | NM_000256.3       | c.1624G>C | p.(Glu542Gln)  | 1               | Pathogenic             |
| HCM                | MYBPC3 | NM_000256.3       | c.2373dupG | p.(Trp792fs) | 1 | Pathogenic             |
| HCM                | MYBPC3 | NM_000256.3       | c.26-2A>G | p.?             | 2               | Likely pathogenic      |
| HCM                | MYBPC3 | NM_000256.3       | c.3192dupC | p.(Lys1065fs) | 1 | Pathogenic             |
| HCM                | MYBPC3 | NM_000256.3       | c.3592dupG | p.(Ala1198fs) | 1 | Pathogenic             |
| HCM                | MYBPC3 | NM_000256.3       | c.772G>A  | p.(Glu258Lys)  | 2               | Pathogenic             |
| HCM                | MYBPC3 | NM_000256.3       | c.994G>T  | p.(Glu332*)    | 1               | Pathogenic             |
| HCM                | MYL3  | NM_000258.2        | c.170C>G  | p.(Ala57Gly)   | 2               | Likely pathogenic*     |
| HCM                | MYH7  | NM_000257.2        | c.4066G>A | p.(Glu135Lys)  | 1               | Likely pathogenic      |
| HCM                | TNNI3 | NM_000363.4        | c.433C>T  | p.(Arg145Trp)  | 2               | Pathogenic             |
| HCM                | TNNI3 | NM_000363.4        | c.434G>A  | p.(Arg145Gln)  | 1               | Likely pathogenic      |
| HCM                | TNNI3 | NM_000363.4        | c.484C>T  | p.(Arg162Trp)  | 1               | Likely pathogenic*     |
| HCM                | TNNI3 | NM_000363.4        | c.485G>A  | p.(Arg162Gln)  | 1               | Likely pathogenic*     |
| LQTS               | KCNH2 | NM_000238.3        | c.1744C>T | p.(Arg582Cys)  | 1               | Likely pathogenic      |
| LQTS               | KCNQ1 | NM_000218.2        | c.1066C>T | p.(Gln356*)    | 2               | Pathogenic             |
| LQTS               | KCNQ1 | NM_000218.2        | c.1780C>T | p.(Arg594*)    | 1               | Pathogenic             |
| LQTS               | KCNQ1 | NM_000218.2        | c.1781G>A | p.(Arg594Gln)  | 1               | Likely pathogenic      |
| LQTS               | SCN5A | NM_001099404.1     | c.6017dupC | p.(Ser2007fs) | 1               | Likely pathogenic      |

Total number of participants 45

No. occurrences = number of times variant found in the BRRD cohort (n = 7203). Variants reviewed and classified January–March 2017.

**Study visit**

Study rationale, possible outcomes, clinical and lifestyle consequences of significant clinical findings and presence
of an ICC gene variant for the participant and relatives, were discussed in full with potential SCARFE participants before written informed consent was taken. A cardiac-focussed medical history and three-generation pedigree were recorded. Clinical evaluation was performed by a single experienced operator using the same equipment according to the ICC associated with the variant detected. Clinical tests were interpreted by a cardiologist specialised in ICC and a diagnostic opinion recorded, prior to unblinding of case/control status. All findings were disclosed to participants the same day, with letter to participant general practitioner and recommendations regarding clinical follow-up in a National Health Service ICC clinic, and family cascade testing as appropriate, supported by family cascade letters. Variants were re-confirmed in a fresh sample by OMGL prior to cascade testing.

Qualitative data collection and analysis

Semi-structured interviews were undertaken by the first author between 6 and 12 months post visit using a guide developed through clinical experience and literature review. Interviews aimed to explore participant perceptions of being approached and of the study visit, their reactions to the information received, whom they had discussed the visit with, and reasons for any lifestyle changes. Interviews were audio recorded and verbatim transcripts analysed thematically [24] by the first two authors.

Results

Genetic analysis

Pathogenic/likely pathogenic variants within ICC genes were detected and confirmed in 44/7203 (0.61%) BRRD participants: numbers of participants carrying a variant associated with each disease: 16 ARVC, 1 DCM, 21 HCM, 6 LQTS (Table 1). Sanger sequencing failed to confirm one variant in one BRRD participant, who was not invited to SCARFE.

Participants

Twelve of the 44 variant carriers were ineligible and a further 12 could not be contacted or were considered unsuitable by the referring clinician. Initial opt-out letters were sent to 20 variant carriers, and returned by two. Invitation letters were sent to the remaining 18 eligible variant carriers, of whom 14 actively declined or did not respond to invitations. Four (mean age = 60) eligible variant carriers attended a study visit and participated. Approach and numbers at each stage are shown in Fig. 1. Variants carried by all four were in genes associated with HCM.

Seven non-carrier controls (mean age = 55) participated; four were matched to a case who declined or did not respond, and for one case, two controls were invited but declined or did not respond. Ten participants who attended a study visit took part in a semi-structured interview; one did not respond to requests.

Clinical findings, family history and genotype

Genetic variants, demographic data and clinical findings of variant carriers who attended for assessment are shown in Table 2.

B3 is a 70-year-old asymptomatic woman. Study visit echocardiogram showed septum measurement at the upper limit of normal with normal left ventricular size and systolic function, no left ventricular outflow tract obstruction, no systolic anterior motion, normal atri and no voltage criteria for left ventricular hypertrophy (LVH) on ECG. B3 has no
Table 2 Characteristics and clinical findings of variant carriers participating in SCARFE.

| ID  | Age (gender) | Echo MWT (cm) | Echo LA dimension (cm) | ECG comments | BP (mm Hg) | Prior family history HCM/SCD | Clinically affected status (blind to genotype) | Variant ACMG classification [18] (reassessment after SCARFE participation) | Relatives at known risk after SCARFE participation (gene test) | Recommendations |
|-----|--------------|---------------|------------------------|--------------|------------|------------------------------|-----------------------------------------------|-----------------------------------------------|-------------------------------------------------|------------------|
| B 3 | 70 years old (F) | 1.2 | 3.0 | No hypertrophy, normal contractility. Slightly enlarged atria. | Small voltages; not suggestive of HCM | 138/70 | No | Unaffected | c.2373dupG; p.(Trp792fs) MYBPC3 NM_000256.3 | Pathogenic | Parent (tested); sibling (tested); child (tested) | 5-yearly follow-up; cascade |
| B 13 | 46 years old (M) | 1.5 | 3.5 | Slightly hypertrophied mid-septum, suspicious for HCM. Unremarkable contractility. | Sinus bradycardia 55 bpm; normal voltages with no criteria for LVH | 113/65 | Yes: sibling with HCM; no gene test | Affected | c.3592dupG; p.(Ala1198fs) MYBPC3 NM_000256.3 | Pathogenic | Siblings n = 2 (testing awaited); children n = 2 (tested) | HCM risk stratification, annual follow-up, cascade |
| B 20 | 66 years old (F) | 1.2 | 3.0 | Slight septal bulge, insufficient for diagnosis | Small Q waves in L3, otherwise normal | 155/72 | Yes: 2 FDRs died suddenly in 70s | Unaffected | c.170C>G p.(Ala57Gly) MYL3 NM_000258.2 | Likely pathogenic (VUS) | Child (testing awaited); second degree relatives with deceased intervening relative n = 3 (2 tested, 1 test awaited) | 5-yearly follow-up; cascade |
| B 17 | 60 years old (F) | 1.1 | 3.4 | Normal septal thickness and contractility | No voltage criteria for LVH | 122/72 | No | Unaffected | c.170C>G p.(Ala57Gly) MYL3 NM_000258.2 | Likely pathogenic (VUS) | Siblings n = 3 (1 tested, 2 not yet sought testing); children n = 2 (1 tested, 1 not yet sought testing) | 5-yearly follow-up; cascade |

Variants carried by all were associated with HCM. All participants were of European ancestry.

**MWT** maximum wall thickness, **LA** left atrial, **bpm** beats per minute, **FDR** first-degree relative, **LVH** left ventricular hypertrophy, **BP** blood pressure.
known family history of HCM or sudden cardiac death. Clinical verdict was that B3 was not affected with HCM. After unblinding, B3 was found to carry a well-established pathogenic HCM variant in MYBPC3 [25] and was referred for clinical care: clinical cardiac magnetic resonance (CMR) imaging confirmed absence of LVH. B3 has three living adult first-degree relatives who have subsequently been tested, of whom one has the variant and is clinically unaffected.

B13 is a 46-year-old asymptomatic man employed in a profession requiring medical certification. Genotype-blind echocardiogram showed mild hypertrophy in the mid-septum with normal left ventricular size and systolic function, no left ventricular outflow tract obstruction, no systolic anterior motion and normal atria; ECG showed no criteria for LVH. One sibling had a prior diagnosis of HCM; no clinical genetic testing had been performed. Following the sibling’s B13 was clinically screened in his 30s, considered unaffected and discharged. Clinical verdict was that B13 was affected with HCM. He was found to carry a novel variant in MYBPC3 predicted to result in premature termination of translation resulting in haploinsufficiency, a known disease mechanism in HCM. Subsequent clinical CMR confirmed asymmetric septal hypertrophy (MWT 1.8 cm) with large areas of patchy fibrosis with late gadolinium enhancement. Twenty-four hours ECG and exercise test were normal. Family testing is ongoing.

B20 is a 66-year-old asymptomatic woman with normal left ventricular size and systolic function, no left ventricular outflow tract obstruction, no systolic anterior motion, normal atria and normal ECG. Two first-degree relatives suffered sudden deaths both at age 76. Clinical verdict was that B20 was not affected with HCM. She was found to have a variant in MYL3 considered likely pathogenic for HCM at the time of SCARFE genomic analysis: this variant is reported as pathogenic on ClinVar due to segregation and functional work in vitro [26] and in vivo [27], but has a higher than expected allele frequency (0.0072%) in gnomAD (r.2.1.1). After the study visit we reviewed the variant and considered evidence for pathogenicity uncertain, with further cardiac investigations not warranted. Cause of death in relatives was certified as cardiac failure and hypertension (parent); acute myocardial infarction, coronary atherosclerosis and myocardial hypertrophy and hypercholesterolaemia (sibling). Cascade testing is ongoing.

B17 is a 61-year-old asymptomatic woman. She has normal left ventricular size with normal wall dimensions and function, no left ventricular outflow tract obstruction, no systolic anterior motion and normal atria, normal ECG. She has no family history of HCM or sudden death. Clinical verdict was that B17 was not affected with HCM. B17 carries the same MYL3 variant detected in B20. Cascade testing is ongoing.

Clinical data for the non-variant carriers are shown in Table 3. None was considered to have clinical features diagnostic of the ICC for which they had been enrolled as controls.

Qualitative data analysis

Understanding of genetic health risks

Participants talked at length about the primary health condition (for which participants were enrolled in the BRRD) in them/their child, and the multiple psychosocial and practical burdens it had imposed on the family. No participant had yet received a genetic diagnosis. Experience with the primary health condition had informed participants about concepts such as inherited disease and risk, variable penetrance and expressivity, and diseases for which treatment is available but no cure. Some participants considered that leading a healthy lifestyle could mitigate disease development in general, despite presumed genetic risk. One of the participants carrying a variant subsequently reclassified already had experience of a variant of unknown significance (VUS) found in relation to the primary condition; she explained her understanding of the meaning of uncertainty in genetic disease:

‘This unknown significant stuff, I’d had a lot of time to get my head around with [child] having something, which they had downplayed considerably. That’s quite different from this unknown variation. I thought it was a very binary answer. It was either, ‘Yes,’ or, ‘No,’ and because I knew that I hadn’t got any heart problems, because my ECG had been fine and everything else, then, I just assumed that I was a no. It never occurred to me’. (B17).

Motivation for and satisfaction with participation in SCARFE

Participants agreed to take part for altruistic reasons as well as personal and family reasons. Only two (of ten) participants recollected the two-stage SCARFE approach. Of interest given the low response rate, one participant had assumed that she was ‘one of many’, and that her participation was not critical. Although some participants reported anxiety about the tests and what they might show, all were satisfied with the study processes before, during and after the study visit; they valued the ‘personalised approach’, ‘empathy’ and ‘clarity’, at the study visit, and availability of the study team post visit. All participants had clear and accurate recollection of personal genetic and cardiac findings from their SCARFE visit.
Table 3 Characteristics and clinical findings of control participants in SCARFE.

| ID    | Age    | Matched case | Phenotype associated with variant carried by matched case | Echo MWT (cm) | Echo LA dimension (cm) | Echo comments | ECG comments | Blood pressure (mm Hg) | Clinically affected status |
|-------|--------|--------------|----------------------------------------------------------|---------------|------------------------|---------------|--------------|------------------------|---------------------------|
| B5    | 48 years old (F) | Declined | HCM                                                     | 1.0           | 3.3                    | Normal contractility, normal cavity size and wall thickness. Borderline increase in LA volume | ST-T wave changes in V2/V3 of uncertain significance; no voltage criteria for LVH | 122/79                  | Unaffected               |
| B2    | 42 years old (F) | Declined | DCM                                                     | 0.8           | 3.7                    | Normal EF; normal cavity dimension | Normal         | 101/76                  | Unaffected               |
| B16   | 46 years old (M) | B13       | HCM                                                     | 1.2           | 4.2                    | MWT upper limit of normal; large vessels in keeping with athletic training, normal diastolic function | T wave inversion in L3, AVF; slight QRS widening (120 ms); prominent U wave, PR interval short (88 ms). No obvious delta wave | 146/98                  | Unaffected               |
| B22   | 67 years old (F) | B20       | HCM                                                     | 1.2           | 3.6                    | Borderline localised proximal hypertrophy, slightly dilated aortic root in keeping with hypertension | No evidence of prior infarct. PR interval upper limit of normal; shallow T wave inversion in V2. No voltage criteria for LVH | 134/77                  | Unaffected               |
| B9    | 39 years old (F) | No response | HCM                                                     | 0.8           | 2.5                    | Normal | Normal | 130/75                  | Unaffected               |
| B3M   | 63 years old (F) | Declined | LQTS                                                    | 1.2           | 3.7                    | Normal | Normal ST segment and T wave. QTc 402 ms | 132/80                  | Unaffected               |
| B7    | 70 years old (F) | B3        | HCM                                                     | 1.2           | 3.8                    | MWT upper limit of normal; normal contractility | Normal; no voltage criteria for LVH | 154/84                  | Unaffected               |

All participants were of European ancestry.

*MWT* maximum wall thickness, *LA* left atrial, *bpm* beats per minute, *LVH* left ventricular hypertrophy, *EF* ejection fraction, *QTc* QT interval (corrected using Bazett formula).
Risk perception

With one exception, participants did not consider, prior to attending the study visit, that they would have an ICC. This low perception of risk was informed variously by lack of symptoms, fitness levels or regular well-person checks in primary care with no heart problems noted. One participant perceived his risk as high: he appreciated that he could be genetically affected but clinically unaffected (earlier tests showed no signs of HCM). Another participant stated in hindsight (during interview) that she had expected to ‘have it’ and this was in some way entwined in her thoughts about her deceased child, albeit whose condition was unrelated to ICC:

‘I thought I had it...I didn’t think, oh my God I’m going to die or anything like that. I just thought, oh this is linked, perhaps, to [child]. I know [child] was so complex, so it was just, oh I thought, oh well perhaps we can help that area as well’. (B3)

However, her very active lifestyle, wellness and parents’ longevity were, for her, in contradiction with the possibility that she might have an ICC.

Personal consequences of participation

No participant regretted participation in SCARFE. For the one participant who found to have clinical signs of ICC, the finding provoked anxiety about the risk posed to continuing his occupation—a source of satisfaction, financial security and fulfilment of ambition since childhood—and period of uncertainty, while he underwent further assessment mandated by his employer. Having cleared this process by the time of interview, his concerns were then about the risk of HCM to his children, already burdened by significant illness. The other case participant with a pathogenic variant (clinically unaffected) found it hard to conceptualise her risk:

‘Am I lucky to be here? How come something didn’t happen?..it’s made me wary that I mustn’t go over the limit...so yes, it’s quite difficult that side’. (B3)

In two control participants, there were unexpected clinical findings unrelated to the ICC. One participant was found to have a subtly abnormal ECG, which the study team reported to his primary care doctor; he described subsequent potential cardiac symptoms on exercise and further medical investigation and monitoring. A further control participant believed she had suffered a ‘heart attack’ decades earlier. At her study visit, she was reassured that her heart showed no evidence of damage, and this evoked several reactions:

‘Then to find out that all those years when I couldn’t do things there was nothing wrong with my heart. A bit angry as well, because it did affect my life a lot...I go from being relieved—well, I am relieved, I’m very pleased about my heart’. (B22)

Other participants also recalled relief at learning their hearts were healthy, but only one (a case) described making positive lifestyle changes as a result of the study, wishing to maintain her health. One participant had ‘good intentions’, others felt they already had a healthy lifestyle.

Family communication and outcomes

The four case participants were advised to inform immediate relatives and were provided with cascade letters. While all four had informed relatives, they had not always found it straightforward depending on individual relatives’ personalities, circumstances and family dynamics. One participant with an elderly parent described her reasons for not informing them; another had arranged genetic testing with the parent’s care home. One participant knew of aunts and uncles who, with their descendants, could be at risk, but had no contact with them.

Responses of health care system

Aside from SCARFE, participants in general reported personal experience of strains on primary care practice. Relatives were distributed around the country. In two families, at-risk relatives had accessed appropriate testing/screening if they had sought it, but in two, relatives had struggled to persuade primary care physicians to refer for evaluation:

‘[Relative’s primary care doctor] was not helpful at all, no. She rang them up and sorted that out for herself. [Another relative’s doctor] wouldn’t refer him. He said he would do a blood test for him, but if he found that he’d got anything, he still wouldn’t refer him’. (B20)

Other relatives were told they would have to wait many months for an appointment. One relative had been able to access cardiac screening but not genetic testing. Primary care doctors were perceived as having limited understanding of genetic risk; this, and delays to appointments, had caused frustration and sometimes anxiety for participants and, reportedly, for relatives.

Discussion

To our knowledge, this is the first study reporting on a systematic approach to disclosure of SF ICC, with targeted
but genotype-blinded clinical evaluation and detailed exploration of sequelae of disclosure. Using this approach we have been able to explore many elements of a policy to look for and report ICC SF, critical for informing policy around SF. The possibility that SF might cause high levels of anxiety has been a concern among healthcare professionals and publics [19]. Distress or anxiety following SF disclosure should be factored into the risk/benefit equation, particularly if SF prove to have relatively low penetrance in unselected populations. Several studies [28–31], including one involving LQTS findings [32], have shown that this concern may be unfounded; but case reports show how anxiety may lead to disengagement with healthcare services [16]. Failure to assimilate SF information for whatever reason might also impede realisation of clinical utility: ‘patients’ (SF recipients) need to make cognitive and emotional adjustments to health risk information to enable relevant decisions about their own healthcare and lifestyle, and communicate risk information successfully to relatives [33]. Participants in our study denied ongoing anxiety caused by the SF per se, and were able to act as recommended, attending for further clinical testing and informing relatives. However, we cannot discount the possibility that among those who declined participation were some who might have found adaptation to new risk information challenging. Participants also demonstrated clear understanding of genetic health risks and concepts such as variable penetrance, informed by their experiences with rare disease; they valued information provided as part of the study design and the availability of continued access to the study team after the study visit.

Maximising the clinical utility of SF requires that healthcare services are able to enact appropriate clinical follow-up and cascade testing [34]. Unless carriers and their relatives are able to access genetic testing, clinical evaluation and follow-up, a search and disclosure policy for SF is of limited utility. In the present study, we found variable primary care responses to relatives’ request for a referral even when supported by a cascade letter: long waiting times and for one participant, a cardiology assessment offered before a genetic appointment. While variability is also seen in ICC clinical practice in the UK, healthcare professional engagement challenges may be exacerbated in families in which no one is clinically affected by the condition associated with the SF. Greater education and resourcing of primary care physicians may be required for this aspect of preventative medicine.

In this study, risk perception was low for all participants who had no family history of ICC. Risk perception was informed by lack of symptoms and high current or former levels of activity, as we have observed in an earlier study of pre-symptomatic testing in ICC [35]. These factors, as well as general health checks in primary care mentioned by some, do not preclude clinical signs of ICC. Further research is needed to understand interactions between risk perception and outcomes of SF disclosure.

The study identified one variant carrier with HCM, who had been previously clinically assessed after a diagnosis of HCM in a first-degree relative, and discharged. Had genetic testing been used in the proband and in family evaluation, it is likely the variant would have been detected and the participant identified as a carrier warranting periodic reassessment. This example highlights the value of clinical genetic testing and cascading for identifying ICC risk in relatives, for their personal health and risk assessment of employees in regulated professions. Incomplete utilisation of clinical genetic testing in cardiomyopathy (DCM) was also found in a genetics-first EMR-based study [36]. Genetic testing is a Class I recommendation for most ICC in a proband with clinical suspicion of disease [37]. However, where genetic testing is either not available or not informative, a single clinical evaluation of relatives in young/mid adulthood is insufficient to rule out risk of ICC.

A second carrier of a pathogenic (HCM) variant was clinically unaffected; cascade testing has clarified that her surviving parent and that parent’s many descendants are at population risk. The variant in question, a frameshift in MYBPC3, is considered unambiguously pathogenic, occurring at high frequency in the Dutch HCM population [25] and OMGL. Clearly it is possible that SF disclosure will fail to find evidence of disease in a family, although we cannot establish whether the deceased parent’s relatives have or are at risk of HCM, due to loss of contact with their descendants.

The study highlights challenges with variant assessment: the MYL3 variant (c.170C > G p.Ala57Gly) detected in our study was considered a pathogenic SF by Natajaran et al. [12] prior to reclassification. Variant interpretation remains inexact and subject to change, presenting a particular challenge in the context of SF when the pretest probability of disease is low [38]; expert laboratory input and genetic counselling will be critical to ensure patient understanding and appropriate clinical follow-up. Conversely, a variant considered a VUS at analysis but subsequently reclassified as pathogenic would not be disclosed.

A dilemma with recall-by-genotype studies is how to approach variant carriers to participate without divulging significant but unexpected risk information or compromising their right not-to-know [20]. This is a concern for studies in which participants had not originally consented to return of SF. Our study design used two strategies to recall participants: a two-step approach as proposed by Beskow et al. [21], whereby an initial letter asked participants in the original study to opt out if they did not wish to learn health or genetic information additional to what they already knew about. Two BRRD participants opted out at this stage,
suggesting that this approach promoted autonomy for some, however, most who accepted participation were unable to recall this letter as distinct from the study invitation letter. Second, we used a case/control strategy, in which participants could learn more about the study and possible implications with specialist genetic counselling, before deciding whether to proceed with clinical screening or learning their genetic status. While this strategy achieved its aim, it may have contributed to low recruitment rate. Further research is necessary to understand optimal approach in recall-by-genotype studies.

**Study limitations**

The main limitation is the small number of eligible participants and low response rate. From a cohort of over 7000 participants, we are unable to draw conclusions about ICC penetrance in the general population; given the rarity of pathogenic variants in these genes, meaningful estimates of ICC SF penetrance will require a very large sequenced population together with detailed cardiac phenotyping. While we have little information about individual reasons for declining, from participant interviews we can infer factors that may have contributed to low participation rate: study participants were recruited from a study investigating rare disease; many have experienced a high burden of disease, requiring high levels of care and multiple hospital appointments. This may have influenced participation rate, psychosocial and behavioural responses compared with a ‘healthy’ population. In addition, the case-control approach, while chosen for optimal design as well as to protect participants, restricted promotion of the possible health benefits of participation. In this study, many people chose not to pursue SF, however, interest in taking part in a research study may not be comparable with uptake of SF in other contexts.

In conclusion, we have piloted a protocol for return and clinical evaluation of putatively pathogenic ICC variants as SF to participants in a study where participants had not been asked to consent for SF. We show that ICC SF can be returned and acted upon without undue anxiety in recipients, but that UK clinical healthcare services are not fully utilising genetic testing in the context of ICC diagnosis, and may need additional resourcing and education to enact family cascading following SF disclosure. The finding of an unambiguously pathogenic ICC SF in a healthy participant highlights that ICC SF cannot be considered deterministic.

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**Compliance with ethical standards**

**Conflict of interest** The authors declare that they have no conflict of interest.

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**References**

1. Stark Z, Dolman L, Manolio TA, Ozengerber B, Hill SL, Caulfield MJ, et al. Integrating genomics into healthcare: a global responsibility. Am J Hum Genet. 2019;104:13–20.
2. Green RC, Berg JS, Grody WW, Kalia SS, Korf BR, Martin CL, et al. ACMG recommendations for reporting of incidental findings in clinical exome and genome sequencing. Genet Med. 2013;15:565–74.
3. Wright CF, Fitzgerald TW, Jones WD, Clayton S, McRae JF, van Kogelenberg M, et al. Genetic diagnosis of developmental disorders in the DDD study: a scalable analysis of genome-wide research data. Lancet. 2015;385:1305–14.
4. Priori SG, Wilde AA, Horie M, Cho Y, Behr ER, Berul C, et al. HRS/EHRA/APHRS expert consensus statement on the diagnosis and management of patients with inherited primary arrhythmia syndromes: document endorsed by HRS, EHRA, and APHRS in May 2013 and by ACCF, AHA, PACES, and AEPC in June 2013. Heart Rhythm. 2013;10:1932–63.
5. Authors/Task Force members, Elliott PM, Anastassakis A, Borger MA, Borggrefe M, Cecchi F, et al. 2014 ESC Guidelines on diagnosis and management of hypertrophic cardiomyopathy: the Task Force for the Diagnosis and Management of Hypertrophic Cardiomyopathy of the European Society of Cardiology (ESC). Eur Heart J. 2014;35:2733–79.
6. Towbin JA, McKenna WJ, Abrams DJ, Ackerman MJ, Calkins H, Darrieux FCC, et al. HRS expert consensus statement on evaluation, risk stratification, and management of arrhythmogenic cardiomyopathy. Heart Rhythm. 2019;16:e373–e407.
7. Christiaens I, Bimie E, Bonsel GJ, Mannens MM, Michels M, Majoors-Krakauer D, et al. Manifest disease, risk factors for sudden cardiac death, and cardiac events in a large nationwide cohort of predictively tested hypertrophic cardiomyopathy mutation carriers: determining the best cardiological screening strategy. Eur Heart J. 2011;32:1161–70.

8. Goldbergen I, Hovr S, Moss AJ, Lopes CM, Barshehet A, McNitt S, et al. Risk for life-threatening cardiac events in patients with genotype-confirmed long-QT syndrome and normal-range corrected QT intervals. J Am Coll Cardiol. 2011;57:51–9.

9. Groeneweg JA, Bhonsale A, James CA, Te Riele AS, Dooyees D, Tichnell C, et al. Clinical presentation, long-term follow-up, and outcomes of 1001 arrhythmogenic right ventricular dysplasia/cardiomyopathy patients and family members. Circ Cardiovasc Genet. 2015;8:437–46.

10. Ng D, Johnston JJ, Teer JK, Singh LN, Peller LC, Wynter JS, et al. Interpreting secondary cardiac disease variants in an exome cohort. Circ Cardiovasc Genet. 2013;6:337–46.

11. Van Driest SL, Wells QS, Stallings S, Bush WS, Gordon A, Nickerson DA, et al. Association of arrhythmia-related genetic variants with phenotypes documented in electronic medical records. JAMA. 2016;315:45–57.

12. Natarajan P, Gold NB, Bick AG, McLaughlin H, Kraft P, Rehm HL, et al. Aggregate penetrance of genomic variants for actionable disorders in European and African Americans. Sci Transl Med. 2016;8:364ra151.

13. Haggerty CM, James CA, Calkins H, Tichnell C, Leader JB, Hartzel DN, et al. Electronic health record phenotype in subjects with genetic variants associated with arrhythmogenic right ventricular cardiomyopathy: a study of 30,716 subjects with exome sequencing. Genet Med. 2017;19:1245–52.

14. Haggerty CM, Damrauer SM, Levin MG, Birtwell D, Carey DJ, Golden AM, et al. Genomics-first evaluation of heart disease associated with titin-truncating variants. Circulation. 2019;140:42–54.

15. Mackley M, McGuire K, Taylor J, Watkins H, Ormondroyd E. From genotype to phenotype. Circ Genom Precis Med. 2018;11:e002316.

16. Haggerty CM, Murray B, Tichnell C, Judge JP, Tandri H, Schwartz M, et al. Managing secondary genomic findings associated with arrhythmogenic right ventricular cardiomyopathy: case studies and proposal for clinical surveillance. Circ Genom Precis Med. 2018;11:e002337.

17. Hylind RJ, Chandler SF, Beuzzleur Ladouceur V, Roberts AE, Bezenndes V, Christensen KD, et al. Phenotypic characterization of individuals with variants in cardiovascular genes in the absence of a primary cardiovascular indication for testing. Circ Genom Precis Med. 2019;12:e002463.

18. Kelly MA, Caleshu C, Morales A, Buchan J, Wolf Z, Harrison SM, et al. Adaptation and validation of the ACMG/AMP variant classification framework for MYH7-associated inherited cardiomyopathies: recommendations by ClinGen’s Inherited Cardiomyopathy Expert Panel. Genet Med. 2018;20:351–9.

19. Mackley MP, Flether B, Parker M, Watkins H, Ormondroyd E. Stakeholder views on secondary findings in whole-genome and whole-exome sequencing: a systematic review of quantitative and qualitative studies. Genet Med. 2017;19:283–93.

20. Beskow LM, Linney KN, Radtke RA, Heizen EL, Goldstein DB. Ethical challenges in genotype-driven research recruitment. Genome Res. 2010;20:705–9.

21. Beskow LM, Fullerton SM, Namey EE, Nelson DK, Davis AM, Wilford BS. Recommendations for ethical approaches to genotype-driven research recruitment. Hum Genet. 2012;131:1423–31.

22. Thomson KL, Ormondroyd E, Harper AR, Dent T, McGuire K, Baks J, et al. Analysis of 51 proposed hypertrophic cardiomyopathy genes from genome sequencing data in sarcomere negative cases has negligible diagnostic yield. Genet Med. 2019;21:1576–84.

23. Alfares AA, Kelly MA, McDermott G, Funke BH, Lebo MS, Baxter SB, et al. Results of clinical genetic testing of 2,912 probands with hypertrophic cardiomyopathy: expanded panels offer limited additional sensitivity. Genet Med. 2015;17:880–8.

24. Braun V, Clarke V. Using thematic analysis in psychology. Qual Res Psychol. 2006;3:77–101.

25. Alders M. The 2373insG mutation in the MYBPC3 gene is a founder mutation, which accounts for nearly one-fourth of the HCM cases in the Netherlands. Eur Heart J. 2003;24:1848–53.

26. Lossie J, Ushakov DS, Ferenczi MA, Werner S, Keller S, Haase H, et al. Mutations of ventricular essential myosin light chain disturb myosin binding and sarcomeric sorting. Cardiovasc Res. 2012;93:390–6.

27. Kazmierczak K, Paulino EC, Huang W, Muthu P, Liang J, Yuan C-C, et al. Discrete effects of A57G-myosin essential light chain mutation associated with familial hypertrophic cardiomyopathy. Am J Physiol Heart Circ Physiol. 2013;305:H575–89.

28. Hart MR, Biesecker BB, Blout CL, Christensen KD, Amendola LM, Bergstrom KL, et al. Secondary findings from clinical genomic sequencing: prevalence, patient perspectives, family history assessment, and health-care costs from a multisite study. Genet Med. 2019;21:1100–10.

29. Wynn J, Holland DT, Duong J, Ahimz P, Chung WK. Examining the psychosocial impact of genetic testing for cardiomyopathies. J Genet Couns. 2018;27:927–34.

30. Lewis KL, Hooker GW, Connors PD, Hymas TC, Wright MF, Caldwell S, et al. Participant use and communication of findings from exome sequencing: a mixed-methods study. Genet Med. 2016;18:577–83.

31. Sanderson SC, Linderman MD, Suckiel SA, Zinberg R, Wasserstein M, Kasarskis A, et al. Psychological and behavioural impact of returning personal results from whole-genome sequencing: the HealthSeq project. Eur J Hum Genet. 2017;25:280–92.

32. Haukkala A, Kujala E, Alha P, Salomaa V, Koskinen S, Swan H, et al. The return of unexpected research results in a Biobank study and referral to health care for Heritable Long QT syndrome. Public Health Genomics. 2013;16:241–50.

33. Marteau TM, Weinman J. Self-regulation and the behavioural response to DNA risk information: a theoretical analysis and framework for future research. Soc Sci Med. 2006;62:1360–8.

34. Peterson JF, Roden DM, Orlando LA, Ramirez AH, Mensah GA, Williams MS. Building evidence and measuring clinical outcomes of returning personal results from whole-genome sequencing: the HealthSeq project. Eur J Hum Genet. 2019;27:604–10.

35. Ormondroyd E, Oates S, Parker M, Blair E, Watkins H. Pre-symptomatic genetic testing for inherited cardiac conditions: a qualitative exploration of psychosocial and ethical implications. Eur J Hum Genet. 2014;22:88–93.

36. Park J, Levin MG, Haggerty CM, Hartzel DN, Judy R, Kember RL, et al. A genome-first approach to aggregating rare genetic variants in LMNA for association with electronic health record phenotypes. Genet Med. 2020;22:102–11.

37. Ackerman MJ, Priori SG, Willems S, Berul C, Brugada R, Calkins H, et al. HRS/EHRA expert consensus statement on the state of the art and recommendations for the management of patients with channelopathies and cardiac phenotypes. Heart Rhythm. 2011;8:1308–39.

38. Amendola LM, Dorschner MO, Robertson PD, Salama JS, Hart R, Shirts BH, et al. Actionable exomic incidental findings in 6503 participants: challenges of variant classification. Genome Res. 2015;25:305–15.