Case Report

Utility of plasma cell-free DNA in metastatic castration-resistant prostate cancer

Itosu Hamano,1 Shingo Hatakeyama,2 Tomoko Hamaya,1 Kyo Togashi,1 Teppei Okamoto,1 Hayato Yamamoto,1 Tohru Yoneyama,3 Takahiro Yoneyama,3 Yasuhiro Hashimoto1 and Chikara Ohyama1,2,3

Departments of 1Urology, 2Advanced Blood Puriﬁcation Therapy and 3Advanced Transplant and Regenerative Medicine, Hirosaki University Graduate School of Medicine, Hirosaki, Japan

Abbreviations & Acronyms
ALP = alkaline phosphatase
AR-amp = androgen receptor amplification
ARAT = androgen receptor-axis-targeted therapy
CAB = combined androgen blockade
cfDNA = cell-free DNA
CNV = copy number variation
CT = computed tomography
LDH = lactate dehydrogenase
mCRPC = metastatic castration-resistant prostate cancer
PCa = prostate cancer
PSA = prostate-specific antigen

Correspondence: Shingo Hatakeyama M.D., Ph.D., Department of Advanced Blood Puriﬁcation Therapy, Hirosaki University Graduate School of Medicine, 5 Zaifu-chou, Hirosaki 036-8562, Japan.
Email: shingoh@hirosaki-u.ac.jp

How to cite this article: Hamano I, Hatakeyama S, Hamaya T et al. Utility of plasma cell-free DNA in metastatic castration-resistant prostate cancer. IJU Case Rep. 2020; 3: 141–144.

Introduction: Cell-free DNA is suggested as a prognostic biomarker in metastatic castration-resistant prostate cancer. However, it remains unknown which parameter of cell-free DNA is correlated with the progression and prognosis of metastatic castration-resistant prostate cancer.

Case presentation: A 75-year-old man with newly diagnosed prostate cancer (serum prostate-speciﬁc antigen 4891 ng/mL, Gleason score 4 + 5 = 9, cT3bN1M1) was referred to our department. He ﬁrst received sequential hormonal therapies and was consequently diagnosed metastatic castration-resistant prostate cancer 64 months after initial treatment. He underwent serial examinations of plasma cell-free DNA, including concentration, androgen receptor ampliﬁcation, TP53 point mutation, and PTEN loss. Only the cell-free DNA concentration increased along with disease progression and declined after the administration of abiraterone and enzalutamide.

Conclusion: This case presented that cell-free DNA concentration was possibly correlated with response to castration-resistant prostate cancer treatment and disease progression. Cell-free DNA concentration was proposed as a potential prognostic biomarker of metastatic castration-resistant prostate cancer.

Key words: androgen receptor, biomarker, castration-resistant prostate cancer, cell-free DNA, concentration.

Keynote message

We present a case of a patient with mCRPC who underwent serial examinations of plasma cfDNA. His cfDNA concentration potentially indicated disease progression and response to treatment with abiraterone and enzalutamide. Further studies are necessary to identify predictive parameters of cfDNA for the progression and prognosis of mCRPC.

Introduction

mCRPC remains a major cause of male cancer deaths.1–5 Not all patients with mCRPC present remarkable increases in serum PSA level as the disease progresses, and presently there are no useful biomarkers that predict disease progression of mCRPC.6–10 Recently, the clinical implication of cfDNA in mCRPC has been studied.11,12 However, it remains unknown which parameter of cfDNA is correlated with the progression and prognosis of mCRPC.13 We have examined longitudinal changes in various parameters of cfDNA in patients with mCRPC at our hospital. We present a case suggesting that the plasma cfDNA concentration potentially indicated mCRPC progression and response to treatment with abiraterone and enzalutamide.

Case presentation

A 75-year-old man with PCa (serum PSA 4891 ng/mL, Gleason score 4 + 5 = 9, cT3bN1M1, Fig. 1a, upper left panel) was referred to our department. He was ﬁrst treated with CAB by leuprolelin and bicalutamide and later received sequential hormonal therapies. He was diagnosed with mCRPC with a PSA of 2.03 ng/mL 64 months after the initial CAB (Fig. 1a, lower panel). We started the monthly examination of cfDNA to monitor the longitudinal changes in plasma
cfDNA concentration (pg/mL), CNV of AR-amp, TP53 point mutation (p.R175H, %), and PTEN loss (p.R233*, %) as recently suggested mCRPC biomarkers in cfDNA. In addition, the reproducibility of cfDNA concentration was confirmed as the error range was within 10% by multiple (two or more) measurements.

Although we administered abiraterone acetate (1000 mg/day) followed by enzalutamide (160 mg/day), PSA continuously rose during treatment. We could not detect radiologic progression by CT or bone scintigraphy (Fig. 1a, upper right panels) until his death. His main symptom was fatigue, which did not progress until 1 year after the administration of abiraterone. After that term, his general condition became worse with the PSA rose until he died 94 months since the PCa diagnosis. The cause of death was considered PCa progression due to no other findings for the cause of death. Meanwhile, AR-amp was not correlated with the PSA rise (Figs 1b, 2a), and frequency of TP53 point mutation and PTEN loss appeared to decline at 9 months after the administration of abiraterone (Fig. 2b). cfDNA concentration and serum ALP and LDH demonstrated responses to the administration of abiraterone and enzalutamide, which was not detected by serum PSA value (Figs 1b, 2a, c).

**Discussion**

This case implied that plasma cfDNA concentration is a potential biomarker for both progression of mCRPC and response to ARAT. There has been an unmet need for a non-invasive procedure that can examine cancer cells other than tissue biopsies. Thus, cfDNA has been focused on as a potential biomarker that demonstrates gene information as well as
the total amount of apoptotic and/or necrotic cells in the entire body. However, it remains unknown which cfDNA parameters might precisely predict prognosis and response to treatment in mCRPC.

While the association between total cfDNA amount and cancer prognosis has been gradually recognized in colorectal, lung, and breast cancers, only a few studies have proposed the clinical implication of total cfDNA in PCa. Kienel et al. assessed cfDNA before taxane-based chemotherapy in 59 patients with CRPC and concluded that cfDNA concentration before therapy was an independent predictor of overall survival. Also, Mehra et al. investigated 571 patients with mCRPC and suggested that cfDNA concentration before taxane-based chemotherapy was a predictive factor for shorter radiologic progression-free survival and overall survival. However, former studies have not investigated the longitudinal change in cfDNA concentration along with CRPC treatment. In the present case, cfDNA concentration declined but PSA did not remarkably change after each administration of ARAT, which implied that cfDNA is potentially a more sensitive biomarker for tumor burden than PSA in mCRPC.

Because of the heterogeneity of mCRPC and the sequence of molecular events in disease progression and therapy resistance, it remains challenging to utilize specific DNA alterations in cfDNA as a biomarker for mCRPC. Among various DNA alterations, AR gene status in cfDNA has been reported as a factor associated with therapy resistance in CRPC. Conteduca et al. analyzed AR copy number and mutations in 73 chemotherapy-naïve and 98 post-docetaxel patient groups and identified a correlation between AR gain and worse OS in both groups. Also, Sumiyoshi et al. analyzed the AR

Fig. 2 Longitudinal changes in the absolute value of potential biomarkers. While plasma cDNA concentration implied a reaction to the use of abiraterone and enzalutamide, AR-amp in cfDNA did not show a remarkable association with ARAT treatment (a). Frequency of TP53 point mutation and PTEN loss appeared to decline at 9 months after the administration of abiraterone (b). ALP and LDH declined after administration of abiraterone and enzalutamide (c).
status of 102 patients with CRPC and suggested the AR aberrations, especially AR-amp, in pretreatment cfDNA were associated with poor response to abiraterone.19 However, AR aberrations in cfDNA are not always detected in patients with CRPC.11,18,19 The longitudinal observation in our case suggested inconsistent changes in AR-amp along with CRPC progression.

Although aberrations in TP53 and PTEN are potential biomarkers of CRPC,12 single-point mutation could not predict disease progression because of the wide variety of mutations in those oncogenes. Indeed, we only analyzed specific point mutations of TP53 and PTEN, which did not imply correlation with mCRPC progression. Meanwhile, the frequency of TP53 and PTEN aberrations appeared to decline after the use of ARAT in this case. As no previous studies have reported such phenomenon, this case may be one of the first studies that suggested the clinical implication of longitudinal change in TP53 and PTEN aberrations after CRPC treatment.

This case implied that cfDNA concentration is a potential important correlate in understanding whole-body conditions in patients with end-stage mCRPC. Interestingly, the patient’s symptom and performance status did not become worse during the cfDNA concentration declined after the administration of abiraterone. Less invasive liquid markers may be beneficial for predicting the prognosis of patients with CRPC in a terminal state. This case also presented a trend in cfDNA concentration similar to those seen in conventional serum markers such as LDH and ALP. Moreover, a recent study suggested that aberrant serum N-glycans are associated with disease progression and poor prognosis in patients with mCRPC.20 Further studies are warranted to clarify whether cfDNA concentration can serve as a biomarker in end-stage mCRPC.

In conclusion, our case report of a patient with mCRPC proposed that plasma cfDNA concentration was possibly correlated with response to ARAT and disease progression not precisely demonstrated by PSA, AR-amp, or imaging modalities. Future studies need to investigate cfDNA as a potential prognostic biomarker of mCRPC.

Acknowledgments

This work was supported by Japan Society for the Promotion of Science (JSPS) KAKENHI Grant Numbers of 15H02563, 17K11119, 18K09157, and 19H05556. We would like to thank Yuki Fujita, Yukie Nishizawa, Satomi Sakamoto, and Mihoko Sutoh Yoneyama for their help in terms of data collection.

Ethical consideration

The genetic tests for this research project have been approved by the Ethics Committee of Hirosaki University Hospital (Approval No. 2019-094). Written informed consent was obtained from the subject.

Conflict of interest

The authors declare no conflict of interest.

References

1 Akamatsu S, Inoue T, Ogawa O, Gleave ME. Clinical and molecular features of treatment-related neuroendocrine prostate cancer. Int. J. Urol. 2018; 25: 345–51.
2 Oikita K, Hatakeyama S, Narita S et al. The effect of treatment sequence on overall survival for men with metastatic castration-resistant prostate cancer: a multicenter retrospective study. Clin. Genitourin. Cancer 2019; pii: S1558-7673(19)30275-7.
3 Narita S, Hatakeyama S, Takahashi M et al. Clinical outcomes and prognostic factors in patients with newly diagnosed metastatic prostate cancer initially treated with androgen deprivation therapy: a retrospective multicenter study in Japan. Int. J. Clin. Oncol. 2020; 25: 912–20.
4 Okamoto T, Hatakeyama S, Narita S et al. Validation and development of the CHAARTED criteria in patients with hormone-naive metastatic prostate cancer: a multi-institutional retrospective study in Japan. Int. J. Urol. 2020; 27: 90–1.
5 Fujita N, Hatakeyama S, Momota M et al. Safety and feasibility of radiation therapy to the primary tumor in patients with metastatic castration-resistant prostate cancer. Clin. Genitourin. Cancer 2020; pii: S1558-7673(20)30058-6.
6 Hamano I, Hatakeyama S, Narita S et al. Impact of nadir PSA level and time to nadir during initial androgen deprivation therapy on prognosis in patients with metastatic castration-resistant prostate cancer. World J. Urol. 2019; 37: 2365–73.
7 Kodama H, Hatakeyama S, Narita S et al. Clinical characterization of low prostate-specific antigen on prognosis in patients with metastatic castration-naive prostate cancer. Clin. Genitourin. Cancer 2019; 17: e1091–e1098.
8 Kodama H, Koie T, Okawa M et al. Castration-resistant prostate cancer without metastasis at presentation may achieve cancer-specific survival in patients who underwent prior radical prostatectomy. Int. Urol. Nephrol. 2020; 52: 671–9.
9 Momota M, Hatakeyama S, Soma O et al. Geriatric 8 screening of frailty in patients with prostate cancer. Int. J. Urol. 2020. https://doi.org/10.1111/iju.14256.
10 Hatakeyama S, Narita S, Takahashi M et al. Association of tumor burden with the eligibility of upfront intensification therapy in metastatic castration-sensitive prostate cancer: A multicenter retrospective study. Int. J. Urol. 2020. https://doi.org/10.1111/iju.14258.
11 Beliran H, Romanel A, Conteduca V et al. Circulating tumor DNA profile recognizes transformation to castration-resistant neuroendocrine prostate cancer. J. Clin. Invest. 2020; pii: 131041.
12 Boerrigter E, Groen LN, Van Erp NP, Verhaegh GW, Schalken JA. Clinical utility of emerging biomarkers in prostate cancer liquid biopsies. Expert. Rev. Mol. Diagn. 2020; 20: 219–30.
13 Spindler KG, Boysen AK, Palligard N et al. Cell-free DNA in metastatic colorectal cancer: a systematic review and meta-analysis. Oncologist 2017; 22: 1049–55.
14 Yi Z, Liu B, Guan X, Ma F. Plasma cell-free DNA and survival in non-small-cell lung cancer: a meta-analysis. Mol. Clin. Oncol. 2017; 7: 167–72.
15 Yu D, Tong Y, Guo X et al. Diagnostic value of concentration of circulating cell-free DNA in breast cancer: a meta-analysis. Front. Oncol. 2019; 9: 95.
16 Kienel A, Porres D, Heidenreich A, Pfeifer D. cfDNA as a prognostic marker of response to taxane based chemotherapy in patients with prostate cancer. J. Urol. 2015; 194: 966–71.
17 Mehra N, Dolling D, Sumansuriya S et al. Plasma cell-free DNA concentration and outcomes from taxane therapy in metastatic castration-resistant prostate cancer from two phase III trials (FIRSTANA and PROSELICA). Eur. Urol. 2018; 74: 283–91.
18 Conteduca V, Witterskog D, Sharabiani MTA et al. Androgen receptor gene status in plasma DNA associates with worse outcome on enzalutamide or abiraterone for castration-resistant prostate cancer: a multi-institutional correlationative biomarker study. Ann. Oncol. 2017; 28: 1508–16.
19 Sumiyoshi T, Mizuno K, Yamasaki T et al. Clinical utility of androgen receptor gene aberrations in circulating cell-free DNA as a biomarker for treatment of castration-resistant prostate cancer. Sci. Rep. 2019; 9: 4030.
20 Matsumoto T, Hatakeyama S, Yoneyama T et al. Serum N-glycan profiling is a potential biomarker for castration-resistant prostate cancer. Sci. Rep. 2019; 9: 16761.