A Panel of CircRNAs in the Serum Serves as Biomarkers for *Mycobacterium tuberculosis* Infection

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Tuberculosis (TB), one of the ancient and deadliest diseases, is a chronic immune disorder caused by *Mycobacterium tuberculosis* (*Mtb*) infection. Due to the lack of ideal diagnostic and therapeutic markers, TB is still posing a major health, social, and economic burden worldwide. Circular RNA (circRNA), a newly discovered endogenous RNA, is abundant and stable in the cytoplasm and has tissue specificity. More and more studies suggested circRNA is involved in a variety of human pathological and physiological processes. Recently, several studies have confirmed circRNAs not only existed in the serum but also could serve as ideal biomarkers for detecting diseases since the circRNAs have continuous, stable, and covalently closed circular structures and are not easily degraded by nucleases. In this study, we screened the circRNA expression profiles in active TB serum samples and healthy volunteers serum samples by circRNA microarrays. Then, we performed qRT-PCR to verified the dysregulated circRNAs and ROC curve analysis to evaluate the value of circRNAs for TB diagnosis. The results showed circRNA_051239, circRNA_029965, and circRNA_404022 could serve as biomarkers for TB diagnosis.

Keywords: biomarker, circRNA, serum, tuberculosis, infection

INTRODUCTION

Tuberculosis (TB) is the ninth leading cause of death worldwide with 1.5 million deaths and 10 million new cases worldwide in 2018 (Reinhart et al., 2012). Despite advances in the effective treatment of TB in recent years, the number of annual deaths and infections remains almost unchanged (Reinhart et al., 2012). The prevention, diagnosis and treatment of tuberculosis are mostly important for TB control. Since there are millions of infected people around the world, timely and effective isolation patients and treatment is the key factors to reduce the transmission
of TB. This is based on the effectively simple and cheap early diagnosis of TB. Currently, the commonly used diagnosis technologies for TB are imaging inspection, bacteriological inspection, molecular biological detection, and immunological experimental inspection. However, all of these methods have some unavoidable limitation. For example, typical imaging inspection (such as X-ray) is useful for the diagnosis of TB, but the specificity is too low to distinguish the TB infection from the other lung disease, such as cancer, pneumonia (Bhalla et al., 2015). The bacteriological inspection is still the gold standard for TB diagnosis, but it requires 4–8 weeks for the growth of M. tuberculosis. Therefore, identifying novel appropriate biomarkers for early diagnosis of TB is an urgent.

Circular RNAs (circRNAs) are a novel class of RNAs that participate in almost all the physiological and pathological processes by acting as competing endogenous (ceRNAs) RNAs to block the functions of their target miRNAs (miRNAs) by binding target miRNAs to relieve the suppression of miRNAs for their target gene (Thomson and Dinger, 2016). CircRNAs are highly stable and resistant to debranching and exonuclease-mediated degradation since they could form a circular structure closed by covalent bonds. Recently, several studies have provided circRNAs could not only exist in the body fluids, such as plasma/serum, interstitial fluid, and saliva, but also could serve as molecular biomarkers in the diagnosis and prognosis of many diseases (Zhang et al., 2018). For example, Sonnenschein et al. (2019) found circDNAJC6, circTMEM56, and circMBOAT2 in the serum could distinguish between healthy and hypertrophic cardiomyopathy patients. Ye et al. (2019) identified hsa_circ_0082182, hsa_circ_0000370 were significantly upregulated in colorectal cancer plasma, and the hsa_circ_0035445 was down-regulated. For qRT-PCR, cDNA was synthesized with SuperScript First-Strand Synthesis System (Invitrogen, Carlsbad, CA, United States) from RNA extracted from serum. Subsequently, the qRT-PCR reactions were performed in a total volume of 10 μL, including 3 μL cDNA, 5 μL 2 × SYBR Green, 0.25 μL primer forward (10 μmol/L) and 0.25 μL primer reverse (10 μmol/L) at 95°C for 10 min, followed by 40 cycles of 95°C for 15 s and 60°C for 60 s. The synthetic Caenorhabditis elegans miRNA cel-miR-39 (5’-UCACCGGGUGUAAAUCAGCUUG-3’) (RiboBio, Guangzhou, China) was spiked into the serum as a normalization control.

**MATERIALS AND METHODS**

**Patients and Healthy Controls**

Patients (n = 131) with active pulmonary TB, patients (n = 50) with community acquired pneumonia (CAP), and healthy controls (n = 53) were collected from the Nanjing Drum Tower Hospital. Serum samples were collected at the patients’ first admission to the hospital. All the TB patients were clinically diagnosed as active pulmonary TB by positive AFB smear staining or sputum culture. For CAP patients, they shared similar symptoms similar to TB infection (such as fever and chills, loss of appetite, shortness of breath), but didn’t have pulmonary TB by sputum smear or/and TB culture. The age- and sex-matched healthy controls, who had no clinical symptoms and normal physical examination, and sputum smear or/and TB culture for pulmonary TB were negative, were collected from the physical examination center of Nanjing Drum Tower Hospital. Patients and controls were matched based on age and gender. All the participants provided their written informed consent to participate in the study and the protocol was approved by the Institutional Research Board of the Nanjing Drum Tower Hospital. Protocols were designed and performed according to the principles of the Helsinki Declaration. Patient characteristics are summarized in Table 1. In order to identify serum circulating circRNAs which could serve as novel biomarkers for active TB diagnosis, a multiphase, case-control study was conducted (Figure 1).

**CircRNA Microarray**

Total RNA was isolated from serum with TRIzol reagent (Invitrogen, Carlsbad, CA, United States) according to the manufacturer’s protocol. The concentration and quality of total RNA of each sample were determined with NanoDrop ND-1000 spectrophotometer (NanoDrop, Wilmington, DE, United States). CircRNA expression profiles of serum for three active pulmonary TB patients and three healthy controls were generated using the 074301 Arraystar Human CircRNA microarray V2. The data processing and quantile normalization were performed by R (Version 3.4.1) software.

**qRT-PCR**

For qRT-PCR, cDNA was synthesized with SuperScript First-Strand Synthesis System (Invitrogen, Carlsbad, CA, United States) from RNA extracted from serum. Subsequently, qRT-PCR reaction was performed by an ABI 7500 real-time PCR System according to the manufacturer’s protocol. Briefly, PCR reactions were performed in a total volume of 10 μL, including 3 μL cDNA, 5 μL 2 × SYBR Green, 0.25 μL primer forward (10 μmol/L) and 0.25 μL primer reverse (10 μmol/L) at 95°C for 10 min, followed by 40 cycles of 95°C for 15 s and 60°C for 60 s. The synthetic Caenorhabditis elegans miRNA cel-miR-39 (5’-UCACCGGGUGUAAAUCAGCUUG-3’) (RiboBio, Guangzhou, China) was spiked into the serum as a normalization control.

**TABLE 1** | The demographic and clinical characteristics of active TB patients, community acquired pneumonia (CAP) patients, and healthy individuals in training and validation sets.

| Variable | TB (n = 128) | CAP (n = 50) | HC (n = 50) |
|----------|-------------|-------------|-------------|
| Age, yearsa | 43.5 (19.84) | 45.3 (18.25) | 44.4 (18.43) |
| Sex, n | | | |
| Male | 87 | 31 | 29 |
| Female | 41 | 19 | 21 |
| History of TB treatment | | | |
| Yes | 32 | | |
| No | 96 | | |

aAge data are present as the mean (SD).
TABLE 2 | circRNAs selected in Biomarker-screening phase by Arraystar Human CircRNA microarray V2 of pooled samples (three active TB patients, three healthy controls) with fold change = 2, and \( p < 0.01 \).

| CircRNA     | Chrom  | Strand | txStart  | txEnd    | CircRNA_type | Best_transcript | Gene_symbol | FC (TB vs. HC) | \( p \)     |
|-------------|--------|--------|----------|----------|--------------|----------------|-------------|----------------|------------|
| hsa_circRNA_051239 | chr19  | –      | 41938372 | 41945481 | Exonic       | uc010xwb.2    | ATP5SL      | 5.358899153 | 0.007123   |
| hsa_circRNA_029965  | chr13  | +      | 33306237 | 33448999 | Exonic       | NM_015032     | POS5B       | 5.174550937 | 0.004386   |
| hsa_circRNA_404022  | chr8   | +      | 41469694 | 41469511 | Exonic       | NM_178819     | AGPAT6      | 3.985618567 | 0.004119   |
| hsa_circRNA_102116  | chr17  | –      | 47388673 | 47389404 | Exonic       | NM_014897     | ZNF652      | 3.609132132 | 0.00888    |
| hsa_circRNA_065793  | chr3   | +      | 50131152 | 50131308 | Exonic       | NM_005778     | RBM5        | 3.175970107 | 0.004198   |
| hsa_circRNA_104588  | chr8   | –      | 37727937 | 37735069 | Exonic       | NM_025151     | RAB11FIP1   | 2.993253915 | 0.004175   |
| hsa_circRNA_048148  | chr19  | +      | 1037623  | 1039064  | Exonic       | uc0020uq.3    | CNN2        | 2.98009157 | 0.008405   |
| hsa_circRNA_406174  | chr22  | –      | 28249824 | 28269830 | Sense overlapping | NM_012399 | PTPNB       | 2.934103883 | 0.002732   |
| hsa_circRNA_100623  | chr11  | –      | 58317258 | 58322413 | Exonic       | NM_004811     | LIPXN       | 2.286347985 | 0.005376   |
| hsa_circRNA_029301  | chr12  | –      | 124832370 | 124835283 | Exonic       | NM_006312     | NCO2        | 2.123749587 | 0.00554    |

Expression and Statistical Analyses

SPSS 18.0 software was used for statistical analyses, and GraphPad Prism 6.0 (GraphPad Software, San Diego, CA, United States) was used to generate graphs. To compare significant differences in serum circRNA expression, the Mann-Whitney \( U \) test was used. A \( P \)-value of < 0.05 was regarded as statistically significant.

RESULTS

Dysregulated CircRNAs Expression Profile in TB Patients

To explore whether serum circRNAs are potential biomarkers for TB infection, the landscape of expressed circular transcripts and their expression pattern in serum was investigated by performing the Agilent-069978 Arraystar Human CircRNA microarray V1 for six serum samples (three active TB patients and three healthy controls) in the biomarker-screening phase (Supplementary Table S1). As showed in Figures 2A,B, Hierarchical clustering and scatter plots indicated that the serum circRNA expression patterns had the potential possibility to distinguish TB patients from HCs. Only 10 circRNAs showed significantly upregulated in TB patients (fold change > 2; \( p < 0.01 \)) (Table 2). The 10 upregulated circRNAs were selected for further analysis by qRT-PCR in the serum of 56 active TB patients, 20 community-acquired pneumonia and 20 healthy controls during the Biomarker-selection phase. Compared with healthy controls and community-acquired pneumonia, the expression level of circRNA_051239, circRNA_029965, and circRNA_404022 was significantly upregulated in TB patients (\( P < 0.01 \)), while no significant difference in the other seven circRNAs was found (all \( P > 0.05 \), shown in Figure 3).

Biomarker-Validation Phase of the Serum CircRNAs for TB

Next, these three circRNAs (circRNA_051239, circRNA_029965, and circRNA_404022) was further validated in a larger cohort,
comprising 72 active TB patients, 30 community acquired pneumonia, 30 healthy controls by qRT-PCR (Figure 1). As the results in the Biomarker-selection phase, the concentrations of these three circRNAs (circRNA_051239, circRNA_029965, and circRNA_404022) in TB patients were significantly increased in the TB patients, compared to the community acquired pneumonia and healthy controls (Figures 4A–C).

ROC Curve Analysis of Dysregulated CircRNAs

To assess the diagnostic value of these three serum circRNAs for TB detection, the ROC curve analysis was subsequently performed. As showed in Figures 4D–F, the area under the curve (AUC) values of these circRNAs were 0.9738 (95% CI, 0.9582–0.9893) for circRNA_051239, 0.9443 (95% CI, 0.9165–0.9721) for circRNA_029965, and 0.9682 (95% CI, 0.9496–0.9868) for circRNA_404022. Then, the diagnostic value of the combination of these three circRNAs was evaluated by a logistic regression model and found that the AUC was 0.9920 with $p < 0.01$ (Figure 4G). These results suggested circRNA_051239, circRNA_029965, and circRNA_404022 in the serum could serve as ideal potential biomarkers for TB diagnosis.

**CircRNA_051239 Was Significantly Upregulated in Drug-Resistant TB Patients**

In the 128 TB patients, we found 20 patients were resistant to at least one drug, while 31 samples were pan-susceptible TB patients. We further analyzed the expression level of circRNA_051239, circRNA_029965, and circRNA_404022 in these drug-resistant TB patients and pan-susceptible TB patients. As the results showed in Figure 5A, circRNA_051239 was significantly increased in the drug-resistant group, while the expression level of the other two circRNAs has no difference. More interestingly, Cui et al. (2017) reported levels of miR-320a decreased in the drug-resistant group compared to the pan-susceptible TB patients. We also evaluated the miR-320a level in our samples. As for the results reported by Cui et al. (2017), miR-320a significantly decreased in the drug-resistant group, compared to the pan-susceptible TB patients (Figure 5B).

We speculated circRNA_051239 might act as a ceRNA for miR-320a, and play a vital role in the TB drug-resistant progress. As predicted by RNAhybrid, miR-320a have three binding sites with circRNA_051239 (Figure 5D).

DISCUSSION

As newly discovered endogenous non-coding RNAs with covalently closed loop structures are resistant to RNase digestion,
(Memczak et al., 2013), circRNAs have been proved could serve as ideal potential biomarkers for tumors, Alzheimer’s disease, cardiovascular disease, and other diseases (Abu and Jamal, 2016). Recently, Fu et al. (2019) found circRNA_103017, circRNA_059914, and circRNA_101128 were increased in the peripheral blood mononuclear cells (PBMCs) from active tuberculosis (TB) patients, while circRNA_062400 was decreased in TB samples. Huang et al. (2018a) also identified the dysregulated circRNAs in the PBMCs from TB patients, and confirmed hsa_circRNA_001937, hsa_circRNA_009024, and hsa_circRNA_005086 were significantly elevated and hsa_circRNA_102101, hsa_circRNA_104964, and hsa_circRNA_104296 were significantly reduced in PBMCs from TB patients as compared to healthy controls. Subsequently, they compared the expression level of circRNAs in the plasma from TB patients and healthy controls, the results showed the expression level of hsa_circ_0001204 and hsa_circ_0001747 were significantly decreased in the plasma of TB patients (Huang et al., 2018b), and hsa_circ_0001953 and hsa_circ_0009024 were remarkably increased in the plasma of TB patients (Huang et al., 2018c). Yi et al. (2018) reported hsa_circRNA_103571 exhibited significant decrease in the plasma of active TB patients and showed potential interaction with active TB-related miRNAs, such as miR-29a and miR-16. As for the RNA isolation, transportation and preservation of PBMCs and plasma is too complexity to do quality control, the circRNAs in the PBMC and plasma could not serve as ideal biomarkers for TB diagnosis in clinical application. Compare to PBMCs and plasma, serum is an ideal material for molecular diagnosis. In order to investigate the potential diagnosis value of circRNAs in the serum for TB diagnosis, we characterized the profile of differentially expressed circulating circRNAs in the serum of active TB patients, CAP patients and healthy controls by circRNA microarray and qRT-PCR. As the results in the PBMCs and plasma, the expression profile of circulating circRNAs in the serum of active TB patients showed significantly different from the healthy controls, and we found that circRNA_051239, circRNA_029965, and circRNA_404022 significantly increased in the serum of TB patients and could serve as ideal biomarkers for TB diagnosis with the AUC of all of the three circRNAs is larger than 0.9.

Drug-resistant TB is one of the biggest threats for global TB control and remains a major public health concern in many developing countries. In 2018, more than 500,000 new cases of multidrug-resistant TB and an additional 100,000 cases with rifampicin-resistant TB were identified (Reinhart et al., 2012). Until now, there is still no biomarker to identify whether the TB patients was drug-resistant. In 2017, Cui et al. (2017) reported miR-320a in the plasma significantly decreased in the
drug-resistant TB patients. We also evaluated the expression level of miR-320a in the serum and found miR-320a in the serum also significantly decreased in the drug-resistant TB patients. However, the expression level of miR-320a in the serum/plasma altered in kinds of diseases. Such as, plasma miR-320a showed significantly decreased in the Arrhythmogenic CardioMyopathy (ACM), compared to the healthy controls (Sommariva et al., 2017). The concentrations of plasma miR-320a were decreased in patients with colorectal cancer and benign lesions (polyps and adenoma) compared with healthy controls, but increased in inflammatory bowel disease (IBD) (Fang et al., 2015). In the present study, circRNA_051239 was significantly upregulated in serum derived from drug-resistant TB patients compared to drug-susceptible patients, implying circRNA_051239 as a potential marker for discriminating drug-resistant TB patients. Moreover, we found the expression levels of circRNA_051239 and miR-320a showed negative correlation (Figure 5C). Recent studies have confirmed circRNAs play essential roles in various physiological and pathological processes via acting as a ceRNA for miRNAs to relieve the repression of miRNAs for their target genes (Thomson and Dinger, 2016). Here, circRNA_051239 was speculated to sponge miR-320a to release the target genes of miR-320a, and play a vital role in the TB drug-resistant progress. However, it needs to be further confirmed.

In summary, our study firstly provided a profile of circulating circRNAs in the serum of TB patients and healthy controls. We found circRNA_051239, circRNA_029965, and circRNA_404022 significantly increased in the serum of TB patients and could serve as ideal biomarkers for TB diagnosis. Moreover, circRNA_051239 could distinguish drug-resistant TB patients from pan-susceptible TB patients.
DATA AVAILABILITY STATEMENT

All datasets generated for this study are included in the article/Supplementary Material. Raw and processed data are stored in the laboratory and are available upon request.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Medical Ethics Committee, Nanjing Drum Tower Hospital, The Affiliated Hospital of Nanjing University Medical School. The patients/participants provided their written informed consent to participate in this study.

REFERENCES

Abu, N., and Jamal, R. (2016). Circular RNAs as promising biomarkers: a mini-review. Front Physiol 7:355. doi: 10.3389/fphys.2016.00355

Bhalla, A. S., Goyal, A., Guleria, R., and Gupta, A. K. (2015). Chest tuberculosis: radiological review and imaging recommendations. Indian J. Radiol. Imaging 25, 213–225. doi: 10.4103/0971-3026.161431

Cui, J. Y., Liang, H. W., Pan, X. L., Li, D., Jiao, N., Liu, Y. H., et al. (2017). Characterization of a novel panel of plasma microRNAs that discriminates between Mycobacterium tuberculosis infection and healthy individuals. PLoS One 12:e0184113. doi: 10.1371/journal.pone.0184113

Fang, Z. X., Tang, J., Bai, Y. Y., Lin, H. Y., You, H. Y., Jin, H. W., et al. (2015). Plasma levels of microRNA-24, microRNA-320a, and microRNA-423-5p are potential biomarkers for colorectal carcinoma. J. Exp. Clin. Cancer Res. 34:86. doi: 10.1186/s13046-015-0198-6

Fu, Y. R., Wang, J. D., Qiao, J. J., and Yi, Z. J. (2019). Signature of circular RNAs in peripheral blood mononuclear cells from patients with active tuberculosis. J. Cell Mol. Med. 23, 1917–1925. doi: 10.1111/jcmm.14093

Huang, Z. K., Su, R., Qing, C., Peng, Y. P., Luo, Q., and Li, J. M. (2018a). Plasma circular RNAs hsa_circ_0001953 and hsa_circ_0009024 as diagnostic biomarkers for active tuberculosis. Front Microbiol 9:2010. doi: 10.3389/fmicb.2018.02010

Huang, Z. K., Su, R. G., Yao, F. Y., Peng, Y. P., Luo, Q., and Li, J. M. (2018b). Circulating circular RNAs hsa_circ_0001204 and hsa_circ_0001747 act as diagnostic biomarkers for active tuberculosis detection. Int. J. Clin. Exp. Pathol. 11, 586–594.

Huang, Z. K., Yao, F. Y., Xu, J. Q., Deng, Z., Su, R. G., Peng, Y. P., et al. (2018c). Microarray expression profile of circular RNAs in peripheral blood mononuclear cells from active tuberculosis patients. Cell Physiol. Biochem. 45, 1230–1240. doi: 10.1159/000487545

Kroh, E. M., Parkin, R. K., Mitchell, P. S., and Tewari, M. (2010). Analysis of circulating microRNA biomarkers in plasma and serum using quantitative reverse transcription-PCR (qRT-PCR). Methods 50, 298–301.

AUTHOR CONTRIBUTIONS

FH and JW designed the experiments. HL, GL, WW, XJ, SG, and XY performed the experiments and analyzed the results. JW helped to analyze the microarray of circRNA and made critical reading of the manuscript. XY wrote the manuscript.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmicb.2020.01215/full#supplementary-material

TABLE S1 | The circRNA expression profile in the serum of TB patients and healthy controls by circRNA Microarray.

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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