Use of Natural Clinoptilolite in the Preparation of an Efficient Adsorbent for Ciprofloxacin Removal from Aqueous Media

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Abstract: The adsorption of the antibiotic ciprofloxacin (CIP) from an aqueous solution by natural zeolite, the calcium-rich clinoptilolite (CLI), and magnetite-coated CLI (MAG-CLI) was investigated. Both CLI and MAG-CLI showed a high adsorption affinity towards CIP at 283, 288 and 293 K at a pH of 5. Adsorption kinetics studied for the initial concentrations of 15–75 mg CIP dm$^{-3}$ follow Lagergren’s pseudo-second order equation and the adsorption is best represented by the Langmuir model. The adsorption mechanism involves strong electrostatic interactions between negatively charged aluminosilicate lattice and the cationic form of CIP accompanied by an ion-exchange reaction. Magnetite coverage (approx. 12 wt.%) induces magnetism, which can facilitate the separation process. The coverage does not influence the adsorption activity of CLI. The leaching test showed that the MAG coating protects the adsorbent from CIP leaching. This is ascribed to interactions between the CIP carboxyl groups and magnetite nano-particles. Antibacterial tests showed strong antibacterial activity of the ciprofloxacin-containing adsorbents towards pathogenic E. coli and S. aureus.

Keywords: natural zeolite; magnetite; ciprofloxacin; adsorption; disinfection

1. Introduction

In the last few decades, the use of antibiotics has constantly increased in both human and veterinary medicine. Although some countries have established monitoring systems in order to restrict inappropriate use of antibiotics, water pollution by antibiotics is generally poorly regulated. Different types of antibiotics can be found in both surface waters and in drinking water [1]. Due to their structural complexity, antibiotic molecules are usually very stable, exhibiting a long-lived persistence in the environment and therefore pose a risk to the environment and to human health [2]. They are toxic towards algae and other aquatic organisms and their excessive use leads to bacterial resistance [3,4].

Ciprofloxacin (CIP) is one of the most widely applied fluoroquinolone-type antibiotics for treatment of human and animal bacterial infections because it shows excellent activity against both Gram-positive and Gram-negative bacteria. Its extensive use leads to the presence of fluoroquinolone in waters in a wide range of concentrations. Thus, the CIP concentration in wastewater effluents varies from ng to mg dm$^{-3}$. An extremely high concentration of up to 50 mg dm$^{-3}$ can be found near drug manufacturing plants [5].

The CIP molecule (Figure 1) has two ionizable groups, amine and carboxyl, with $pK_a = 5.90 \pm 0.15$ and $8.89 \pm 0.11$, respectively. Depending on the pH value, CIP can exist as a cation, zwitterion or anion [6,7].

The removal of CIP from wastewater by conventional wastewater treatments, including oxidation [8], ozonation [9], as well as several photocatalytic [10] and biological treatments [11,12], is usually complex, energy-inefficient, and incomplete [13].

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Adsorption is generally regarded as a simple method for the removal of different types of pollutants due to its relatively simple design, easy operation and cost effectiveness. Since the operating cost of a treatment based on the adsorption process is greatly dependent on the cost of the adsorbent material and its recyclability, a variety of low-cost adsorbents have been explored, showing adsorption to be superior in comparison to other methods [14–18]. Thus, carbon-based materials [19–22], clay minerals [23], nano-sized metal oxides [5,24], waste biomaterials [25], biochar [26], coal fly ash [27], as well as different zeolites [28–30] have been studied as adsorbents for the removal of antibiotics from wastewater. Removal efficiency of CIP on different adsorbents are listed in Table 1.

**Table 1.** Removal efficiency of ciprofloxacin on different types of adsorbent.

| Adsorbent                           | Experimental Conditions | CIP Removal Efficiency, % | Recyclability | Ref. |
|-------------------------------------|-------------------------|---------------------------|---------------|------|
| Nano-sized magnetite                | $C_0 = 33$ mg dm$^{-3}$  | 80                        | * n.r.        | [5]  |
|                                     | Adsorbent dose = 10 g dm$^{-3}$  |                           |               |      |
|                                     | pH = 5.97               |                           |               |      |
|                                     | $t = 24$ h              |                           |               |      |
| Graphene hydrogel                   | $C_0 = 50$ mg dm$^{-3}$  | 75                        | n.r.          | [19] |
|                                     | $T = 25^\circ$C         |                           |               |      |
|                                     | $t \approx 36$ h        |                           |               |      |
| Carbon from date palm leaflets      | $C_0 = 100$ mg dm$^{-3}$ | 56                        | n.r.          | [22] |
|                                     | Adsorbent dose = 2 g dm$^{-3}$  |                           |               |      |
|                                     | pH = 6                  |                           |               |      |
|                                     | $T = 45^\circ$C         |                           |               |      |
|                                     | $t = 48$ h              |                           |               |      |
| Halloysite nanotubes                | $C_0 = 30$ mg dm$^{-3}$  | 95                        | 95% after five cycles | [23] |
|                                     | Adsorbent dose = 1.7 g dm$^{-3}$  |                           |               |      |
|                                     | pH = 5–6                |                           |               |      |
|                                     | $T = 20^\circ$C         |                           |               |      |
|                                     | $t = 90$ min            |                           |               |      |
| $\gamma$-Al$_2$O$_3$ nanoparticles  | $C_0 = 20$ mg dm$^{-3}$  | 53                        | n.r.          | [24] |
|                                     | Adsorbent dose = 0.775 g dm$^{-3}$  |                           |               |      |
|                                     | pH = 7.5                |                           |               |      |
|                                     | $t = 46.25$ min         |                           |               |      |
| Biomaterials from banyan aerial roots | $C_0 = 60$ mg dm$^{-3}$  | 97                        | n.r.          | [25] |
|                                     | Adsorbent dose = 1.2 g dm$^{-3}$  |                           |               |      |
|                                     | pH = 8                  |                           |               |      |
|                                     | $T = 25^\circ$C         |                           |               |      |
|                                     | $t = 48$ h              |                           |               |      |
| Biochar-montmorillonite             | $C_0 = 25$ mg dm$^{-3}$  | 86                        | n.r.          | [26] |
|                                     | Adsorbent dose = 1 g dm$^{-3}$  |                           |               |      |
|                                     | pH = 5–6                |                           |               |      |
|                                     | $t = 400$ min           |                           |               |      |
Table 1. Cont.

| Adsorbent                        | Experimental Conditions          | CIP Removal Efficiency, % | Recyclability | Ref.   |
|----------------------------------|----------------------------------|---------------------------|---------------|--------|
| Coal fly ash                     | $C_0 = 160$ mg dm$^{-3}$          | 39                        | n.r.          | [27]   |
|                                  | Adsorbent dose = 40 g dm$^{-3}$   |                           |               |        |
|                                  | $T = 40 \, ^\circ C$             |                           |               |        |
|                                  | $t = 100$ min                    |                           |               |        |
| Clinoptilolite                   | $C_0 = 5$ mg dm$^{-3}$           | 99                        | n.r.          | [29]   |
|                                  | Adsorbent dose = 2 g dm$^{-3}$   |                           |               |        |
|                                  | pH = 6                           |                           |               |        |
|                                  | $T = 25 \, ^\circ C$            |                           |               |        |
| Synthesized zeolites (A, X, Y)   | $C_0 = 150$ mg dm$^{-3}$         | 27–61.4                   | n.r.          | [30]   |
|                                  | Adsorbent dose = 0.5 g dm$^{-3}$ |                           |               |        |
|                                  | pH = 3                           |                           |               |        |
|                                  | $T = 20 \, ^\circ C$            |                           |               |        |
|                                  | $t = 24$ h                       |                           |               |        |
| Commercial zeolites (A, X, Y)    | $C_0 = 150$ mg dm$^{-3}$         | 34–87                     | n.r.          | [30]   |
|                                  | Adsorbent dose = 0.5 g dm$^{-3}$ |                           |               |        |
|                                  | pH = 3                           |                           |               |        |
|                                  | $T = 20 \, ^\circ C$            |                           |               |        |
|                                  | $t = 24$ h                       |                           |               |        |

* n.r.—not reported.

It should be noted that the data shown in Table 1 are difficult to compare because they are obtained under different experimental conditions. Moreover, for most adsorbents CIP adsorption is an irreversible process.

Natural zeolites have been extensively studied because of their unique ion-exchange, molecular sieving, and adsorption properties. Their worldwide abundance recommends them as available, low-cost materials [31]. Clinoptilolite (CLI) is the most abundant natural zeolite. Its three-dimensional structure consists of two interconnected micropore channels occupied by exchangeable cations and water molecules. CLI can efficiently remove different pollutants from wastewaters, including inorganic toxic ions such as heavy metal cations [32,33], anions [34,35] as well as various organic species such as pesticides, dyes, or pharmaceuticals [36]. The removal efficacy of CLI can be ascribed to its unique structural features as well as to its propensity to be modified without structural changes. Impregnation of CLI with oxide nanoparticles or with biopolymers leads to an increase of its specific surface and adsorption performance [37–43].

Generally, the literature data on the adsorption of organic pollutants by zeolites are relatively scarce which can be ascribed to the fact that most zeolites are hydrophilic. Hydrophobicity of zeolites increases by the increase of the Si/Al ratio. However, it should be noted that CLI has been the most studied natural zeolite adsorbent not only for inorganic but also for organic pollutants [6]. Its adsorptive effectiveness towards specific pollutants varies because the Si/Al molar ratio and elemental composition of CLI from different regions differ. Among synthetic zeolites, the zeolite Y was reported to be an efficient antibiotic adsorbent [44]. However, the adsorption was found to be irreversible, so the reuse of the spent adsorbent is questionable.

The aim of this research was to study: (1) the adsorptive efficacy of natural clinoptilolite (CLI) for CIP removal at different temperatures and for different initial CIP concentrations, (2) adsorption kinetics and mechanisms, (3) impregnation of CLI with magnetite nanoparticles (MAG-CLI) to allow a facile adsorbent separation, and (4) antibacterial activity of the CIP-containing CLI.

CLI was used for the CIP removal because an extensive study conducted in the last decade showed that clinoptilolite-reach tuffs from Serbia can be perspective adsorbents for the removal of various toxic species [32,33].
2. Materials and Methods

2.1. Materials

The clinoptilolite-rich zeolitic tuff obtained from the Serbian deposit in Slanci (near the capital city of Belgrade) was used as a starting material. Rietveld analysis [45] showed that CLI was the major mineral phase (>80 wt.%), with quartz (<7.5 wt.%) and feldspar (<13 wt.%) the major satellite phases. The cation exchange capacity (CEC), measured by standard procedure [46], was 162.2 mmol M⁺/100 g.

Prior to the experiments, the sample was sieved, washed with deionized water to remove impurities, and then dried in an oven at 105 °C overnight to a constant mass. The particle size used in the experiments was in the range of 0.063–0.125 mm since previous studies showed the best adsorptive performance was in particles in that size range [32,33].

All chemicals used were of analytical grade. All the experiments were carried out under controlled conditions. The experiments were performed in a thermostated bath where the temperature was maintained constant to within ±0.1 °C; the CLI samples were weighed to a four-digit accuracy, and the solution concentrations were determined with a four-digit accuracy. In all experiments, deionized water was used.

2.2. Preparation of Magnetite

The magnetite nanoparticles were synthesized per the literature procedure [5,47]. Briefly, FeCl₃·6H₂O and FeCl₂·4H₂O were dissolved in degassed distilled water under N₂ in a molar ratio of 2:1. A solution of ammonia was then added dropwise to the prepared Fe(II) and Fe(III) solution, increasing the pH to 10. A black solid that precipitated was magnetically separated and washed with distilled water until a negative reaction to chloride ions.

2.3. Preparation of Magnetite-Coated Clinoptilolite (MAG-CLI)

Conversion of CLI to MAG-CLI was carried out following a slightly modified method as described by Mohseni-Bandpi [48]. Briefly, 10 g of CLI were suspended in 120 cm³ of deionized water and stirred under the N₂ atmosphere at 350 rpm for about 10 min. Then, a solution, which contained FeCl₃·6H₂O and FeSO₄·7H₂O in a molar ratio of 2:1, was added to the CLI suspension under stirring. The pH of the suspension was adjusted to pH ~10 by the dropwise addition of a water solution of NH₄OH (25 wt.%), followed by a vigorous stirring in a water bath at ~80 °C under N₂ for 1 h. The resulting black precipitate was separated from the suspension by centrifugation and washed several times with deionized water until a negative reaction to chloride ions. Finally, the obtained MAG-CLI was left to dry at 80 °C overnight until constant mass.

2.4. Characterization

2.4.1. Powder X-ray Diffraction Analysis (PXRD)

Analysis of the mineral phases present in the samples was done via the powder X-ray diffraction method (PXRD). PXRD patterns were obtained using a Rigaku SmartLab powder diffractometer (Rigaku, Tokyo, Japan), with CuKα (λ = 1.54178 Å) radiation at 40 kV and 30 mA and Bragg-Brentano focusing geometry. The samples were scanned in the 2θ range from 5° to 65°, with 0.01° step and a scan rate of 5° min⁻¹.

2.4.2. Energy Dispersive X-ray Spectroscopy (EDS)

Elemental compositions of CLI and MAG-CLI were obtained by a Carl Zeiss Supra™ 3VP (Zeiss, Jena, Germany) field-emission gun scanning electron microscope (FESEM) equipped with an EDS detector (Oxford Analytical) with the INCA Energy system for quantification of elements. Before analysis, the samples were coated with gold.

2.4.3. Thermal Analysis

Thermal behavior was examined by simultaneous thermogravimetric (TGA) and differential thermal analysis (DTA) using a SDT Q-600 instrument (TA Instruments, New
Castle, DE, USA). The samples were heated in standard alumina sample cups (90 µL) from room temperature to 800 °C at a heating rate of 10 °C min⁻¹ under synthetic air with a flow rate of 100 cm³ min⁻¹.

2.4.4. Textural Properties

The specific surface area and porosity characteristics were determined by the N₂ adsorption at −196 °C using a Micromeritics ASAP 2020 instrument (Micromeritics, Norcross, GA, USA). Prior to the measurements, the samples were degassed in a vacuum at 423 K for 10 h. The specific surface area of samples was calculated using the Brunauer-Emmett-Teller (BET) method from the linear part of the N₂ adsorption isotherm. The total pore volume \( V_{\text{tot}} \) was determined from the desorption isotherm at \( p/p_0 = 0.998 \), and the volume of mesopores was calculated by the Barrett-Joyner-Halenda (BJH) method from the desorption isotherms. The average maximum pore diameter \( D_{\text{max}} \) was determined from the BJH desorption analysis.

2.4.5. TEM Analysis

TEM analysis was carried out at 200 kV using a transmission electron microscope (JEM-2100, JEOL Ltd., Tokyo, Japan) equipped with an ultra-high-resolution objective lens pole piece with a point-to-point resolution of 0.19 nm. The samples were crushed in an agate mortar, dispersed in absolute ethanol, and a fine fraction of the suspension was deposited on Cu holey carbon-coated grids. To determine the average size of magnetite particles, their diameters were measured from the obtained TEM images. Due to the relatively small size of the magnetite nanoparticles, the selected area electron diffraction (SAED) was performed over multiple nanocrystals to obtain the characteristic diffraction rings with the structure-specific \( d \)-values.

2.4.6. FTIR Analysis

The Fourier transform infrared spectroscopy (FTIR) analysis was performed using a Digilab FTS-80 interferometer (Bio-Rad, Cambridge, MA, USA). The spectra were recorded in the range of 400 to 4000 cm⁻¹, at a resolution of 4 cm⁻¹ and 100 scans.

2.4.7. Zeta Potential Measurement

The zeta potentials were measured using a Zetasizer Nano ZS90 (Malvern Instruments Ltd., Malvern, UK) according to the method described by Gulicovski [49]. Briefly, the sample (0.1 g) was suspended in 50 cm³ of deionized water and left at room temperature overnight. The pH value of the suspension was then adjusted in the range of 2–12 using 0.1 mol dm⁻³ HCl or NaOH. Before measurement, the suspensions were ultrasonically treated for 20 s. The pH values were measured using a Mettler Toledo (Columbus, OH, USA) digital pH meter.

2.4.8. Magnetic Measurements

Magnetic properties were measured using a vibrating sample magnetometer LakeShore 7400 Series VSM (LakeShore, Westerville, OH, USA) at the room temperature and with an applied magnetic field strength of 1.53 T.

2.5. CIP Adsorption Experiments

CIP adsorption experiments were performed at 283, 288 and 293 K by the batch method using 0.2 g of CLI or MAG-CLI in 50 cm³ of the chosen CIP concentration (15, 25, 50 or 75 mg dm⁻³). The adsorption experiments were carried out at a pH of 5, which previous studies showed had the best CIP removal efficacy (vide infra). The pH adjustment was performed using HCl or NaOH solutions (0.1 mol dm⁻³). The suspensions were left shaking (100 rpm) in a thermostated water bath (Memmert, WBE 22) at the chosen temperature for 5 to 60 min. The solid was separated by centrifugation and then filtered through a 0.22 µm nylon filter. The CIP concentration was measured in the filtrate by
standard procedure using a UV-Vis spectrometer, Perkin Elmer Lambda 365 (Perkin Elmer Inc., Waltham, MA, USA) at 278 nm [50]. The amount of the adsorbed CIP per unit of the adsorbent after time \( t \) \( (q_t) \) was calculated using the following formula:

\[
q_e = (C_0 - C_e) \cdot V / m
\]  

(1)

2.6. Leaching Test

Zero point two grams of the spent CLI or MAG-CLI, containing about 21 mg CIP per gram of the adsorbent (CLI or MAG-CLI), were suspended in 50 cm\(^3\) of 0.1 mol dm\(^{-3}\) NaCl and left for 24 h in a thermostated water bath at 25 °C. After filtration, the CIP content in the filtrate was determined by UV-Vis. The desorbed amount of CIP was calculated as the ratio of the CIP amount in the solution and on the adsorbent.

The concentration of exchangeable cations (K\(^+\), Mg\(^{2+}\), and Ca\(^{2+}\)) in filtrates after the CIP adsorption was measured using an atomic absorption spectrophotometer, Varian SpectrAA 55B (SpectraLab Inc., Markham, ON, Canada).

2.7. Antibacterial Activity Test

The antibacterial activity of CIP and of the spent adsorbents (CLI-CIP and MAG-CLI-CIP) was tested towards the Gram-negative bacterium *Escherichia coli* strain DSM 498 and the Gram-positive bacterium *Staphylococcus aureus* strain ATCC 25923. Bacteria were pre-grown on a nutrient agar (NA, Torlak, Serbia) for 16 h at 37 ± 0.1 °C to obtain cultures in the log phase of growth. Prior to the experiments, the samples were sterilized by UV light for 30 min.

The disk diffusion method was used for a qualitative assessment of antibacterial activity. The biomass of each bacterial strain was separately suspended in a sterile physiological solution to achieve the bacterial concentration of about \( 10^9 \) CFU cm\(^{-3}\) (colony forming units) and then inoculated onto NA with a sterile swab. Antibacterial activity was tested as follows: 0.01 cm\(^3\) of suspensions containing 5, 10 or 20 mg of CIP, CLI-CIP, or MAG-CLI-CIP per cm\(^3\) in a sterile phosphate buffer solution (pH = 7.02) was dropped onto the bacteria inoculated NA and the plates were incubated for 24 h at 37 °C. The results were reported as the presence or absence of the inhibition zone.

3. Results and Discussion

3.1. Powder X-ray Diffraction (PXRD)

The PXRD pattern (Figure 2) showed that CLI was the main mineral phase, displaying diffractions at \( 2\theta = 9.79^\circ, 11.16^\circ, 13.06^\circ, 16.89^\circ, 19.00^\circ, 22.31^\circ, 25.01^\circ, 25.98^\circ, 29.90^\circ, 31.84^\circ \) and 32.68° [51]. Diffractions at \( 2\theta = 26.6^\circ \) and 27.1° corresponded to quartz and feldspar, respectively [38,51]. The PXRD pattern of MAG-CLI showed that the modification of CLI to MAG-CLI did not significantly affect the CLI crystallinity. New diffraction peaks that appeared in the pattern of MAG-CLI at \( 2\theta = 35.64^\circ, 43.23^\circ \) and 62.96° suggest the presence of magnetite Fe\(_3\)O\(_4\) [52,53]. According to the EDS analysis (vide infra), the content of magnetite is 12.2 wt.%. 

3.2. Energy Dispersive X-ray Spectroscopy (EDS)

The average chemical compositions of CLI and MAG-CLI obtained by the EDS analysis are listed in Table 2. It is evident that the zeolite phase consisted of the calcium-rich clinoptilolite with a Si/Al molar ratio of 5.03. The Si/Al molar ratio increased to 6.12 during modification to MAG-CLI, suggesting a partial dealumination. Moreover, the content of exchangeable cations (Ca and K) decreased in MAG-CLI, whereas the content of Fe increased. Since the content increase of Fe is significantly higher than the content decrease of exchangeable cations, it can be concluded that the modification of CLI to MAG-CLI proceeds not only through an ion exchange but also through precipitation of the Fe species on the CLI surface.
3.3. Thermogravimetric Analysis (TGA)

The thermograms of CLI and MAG-CLI (Figure 3) show that the TG curves of both samples display similar continual weight loss up to 300 °C. At higher temperatures, the weight loss of MAG-CLI differed from that of CLI. CLI lost weight continuously up to 800 °C, whereas the weight loss of MAG-CLI took place in two stages (300–500 °C and 500–800 °C). The total weight losses were similar (wt. %: 14.2—CLI and 13.1—MAG-CLI). The weight losses are accompanied by the following DTG events: (a) weak DTG maxima at 58, 160 and 600 °C on the DTG curve of CLI, suggesting a release of water molecules from different crystallographic sites in the aluminosilicate lattice, and (b) a strong maximum at 470 °C on the DTG curve of MAG-CLI, indicating a conversion of magnetite to maghemite (γ-Fe₂O₃) [54].

Table 2. The average chemical composition of the CLI phase of CLI and MAG–CLI obtained by EDS analysis.

| Sample   | Si   | Al  | O    | K  | Ca | Fe  | Si/Al |
|----------|------|-----|------|----|----|-----|-------|
| CLI      | 18.35| 3.65| 75.97| 0.44| 1.20| 0.40| 5.03  |
| MAG-CLI  | 18.39| 3.01| 72.15| 0.27| 0.18| 5.63| 6.12  |

Figure 2. PXRD patterns of CLI and MAG-CLI.
3.4. TEM Analysis

Figure 4 gives the TEM image of MAG–CLI. For comparison, the image of pure magnetite (MAG) is given in Figure 4a. The magnetite particles are in the range of 5–30 nm, with few particles exceeding 100 nm. The selected area electron diffraction (SAED) pattern displays rings typical for the spinel (magnetite) structure, with the most intense ring of the \{311\} reflections in randomly oriented nanocrystals. MAG shows characteristic octahedral morphology with many crystals connected by the \{111\} spinel-twin law that forms through self-assembly [55]. On the TEM image of MAG-CLI (Figure 4b) an abundant coverage of the CLI plates by rounded magnetite nanocrystals with highly degraded morphology is evident. The SAED pattern shows interrupted ring patterns that are indicative of nanocrystal ordering. The intensity peaks with a six-fold symmetry, which can be seen for the \{111\} and \{311\} magnetite reflections [56,57].

3.5. FTIR Analysis

Figure 5 shows the FTIR spectra of the synthesized samples. The MAG spectrum displays characteristic vibration bands of the Fe–O stretching at about 570 cm$^{-1}$ [47]. The bands overlap with the vibration of the (Al, Si)O$_4$ tetrahedral units of the aluminosilicate lattice [58]. On the spectra of CLI and MAG-CLI, the stretching vibrations of the OH groups of water molecules (1590–1670 and 3400–3700 cm$^{-1}$) and of the (Al, Si)O$_4$ tetrahedral units (1050–1250 cm$^{-1}$) are evident, confirming that the modification of CLI into MAG-CLI did not influence the zeolite aluminosilicate lattice.
3.6. Zeta Potential Measurements

The graphs of the zeta potential as a function of pH (Figure 6) show that the zeta potential of CLI changed by the modification to MAG-CLI, although both CLI and MAG-CLI have a negatively charged surface in a wide pH range from 2 to 12. For MAG-CLI, the point of zero charge (PZC) was at a pH of ~2, which is in accordance with the literature data [59]. The effect of pH on the CIP removal efficiency is presented in Figure 7. The removal efficiency decreased in alkaline media, which is in accord with the fact that both CIP and the surfaces of CLI and MAG-CLI are negatively charged. In acidic media, the CIP is positively charged (CIP\(^+\)) and the attractive forces between CIP\(^+\) and CLI/MAG-CLI favor adsorption. The highest removal efficiency is at pH = 5.

![Figure 5. FTIR spectra of MAG (a), CLI (b) and MAG-CLI (c).](image)

![Figure 6. Zeta potential measurements as a function of pH for CLI (square) and MAG-CLI (circle).](image)
Figure 7. The influence of pH in the CIP adsorption on CLI (squares) and MAG-CLI (circles); 
$C_0 = 50 \text{ mg dm}^{-3}$, adsorption time = 60 min, adsorbent dose = 4 g dm$^{-3}$ and $T = 293 \text{ K}$.

Similar results were found for metal oxides, clays, and a graphene oxide composite [5,24,60,61].

3.7. Textural Analysis

Table 3 presents the textural parameters of CLI and MAG-CLI, which show that textural properties changed during conversion. The specific surface area ($S_{BET}$) of MAG-CLI is two times larger than that of CLI and so is the total pore volume ($V_{tot}$). The pore size diameter ($D_{max}$) of MAG-CLI is smaller than in CLI, which could be explained by two phenomena: (1) the formation of a second layer at the CLI surface consisting of magnetite nanoparticles [39,40], and (2) a partial blockage of the CLI pore system caused by the formation of MAG particles.

Table 3. Textural properties of CLI and MAG–CLI.

| Sample   | $S_{BET}$ (m$^2$ g$^{-1}$) | $V_{tot}$ (cm$^3$ g$^{-1}$) | $D_{max}$ (nm) |
|----------|-----------------------------|-----------------------------|-----------------|
| CLI      | 23.57                       | 0.0988                      | 16.26           |
| MAG-CLI  | 45.17                       | 0.1531                      | 3.50            |

$S_{BET}$—specific surface area, m$^2$ g$^{-1}$; $V_{tot}$—total pore volume, cm$^3$ g$^{-1}$; $D_{max}$—average maximum pore diameter, nm.

3.8. Magnetic Measurements

The magnetic properties of MAG-CLI were also studied and compared with those of MAG. The magnetization curves are presented in Figure 8a. Both MAG and MAG-CLI showed similar magnetic behavior with the saturation magnetization values of 49.57 and 8.93 emu g$^{-1}$ within magnetic field lower than 1.5 T, respectively. The lower saturation magnetization value of MAG-CLI is the result of MAG precipitation on the CLI surface. However, this value indicates that MAG-CLI possesses magnetic properties since CLI itself does not exhibit magnetism (not shown). Furthermore, the magnetization loops showed an absence of hysteresis, which is a characteristic of the superparamagnetic behavior of some nanoparticles [53]. It is worth noting that MAG-CLI retains magnetic properties after the CIP adsorption (Figure 8b), suggesting that the magnetic separation of the spent MAG-CLI from water media is possible.
3.9. Adsorption Isotherm Study

Adsorption of CIP on both CLI and MAG-CLI was studied at 283, 288 and 293 K for the initial CIP concentrations of 15, 25, 50 and 75 mg dm$^{-3}$ and at a pH of 5. The solid to liquid ratio was 1:250, which the experiments had shown to be optimal (Figure 9).

The adsorption capacity of both CLI and MAG-CLI increased with the initial CIP concentration at all studied temperatures (Figure 10). At 293 K, the adsorption of CIP onto CLI increased from 4.39 mg CIP g$^{-1}$ (for $C_0 = 15$ mg CIP dm$^{-3}$) to 12.52 mg CIP g$^{-1}$ (for $C_0 = 75$ mg CIP dm$^{-3}$). Similar increases were observed for MAG-CLI: 4.26 mg CIP g$^{-1}$ (for $C_0 = 15$ mg CIP dm$^{-3}$) to 12.59 mg CIP g$^{-1}$ (for $C_0 = 75$ mg CIP dm$^{-3}$), suggesting that the MAG cover did not influence the CIP adsorption.
Figure 10. The adsorption isotherms for CIP on CLI (a) and MAG-CLI (b); \( q_e \) is the amount of the adsorbed CIP (mg per 1 g of CLI or MAG-CLI) and \( C_e \) is the CIP solution concentration at equilibrium.

The adsorption equilibrium data given in Figure 10 were analyzed by widely applied empirical adsorption isotherm models [17,62]. Among the two-parameter models, only the Langmuir and Freundlich isotherm models gave acceptable fits.

The Langmuir model [63] is represented as:

\[
q_e = \frac{q_{\text{max}} \cdot b_L \cdot C_e}{1 + b_L \cdot C_e} \tag{2}
\]

where \( C_e \) is the equilibrium concentration of CIP (mg dm\(^{-3} \)), \( q_e \) is the equilibrium concentration of the adsorbed CIP (mg g\(^{-1} \)), while \( q_{\text{max}} \) (mg g\(^{-1} \)) and \( b_L \) (dm\(^3\) mg\(^{-1} \)) are Langmuir constants (\( q_{\text{max}} \) corresponding to the maximum achievable uptake by the system, and \( b_L \) is related to the affinity between CIP and CLI or MAG-CLI).

The Freundlich isotherm model is described with the following equation:

\[
q_e = K_f \cdot C_e^{1/n} \tag{3}
\]

where \( C_e \) is the equilibrium concentration of CIP (mg dm\(^{-3} \)), \( q_e \) is the equilibrium concentration of the adsorbed CIP (mg g\(^{-1} \)), \( K_f \) is the Freundlich isotherm constant (mg g\(^{-1} \)) and \( n \) is a parameter related to the adsorption intensity.

The results are listed in Table 4. As can be seen from the obtained values of the linear regression correlation coefficients \( R^2 \), the Langmuir model gave a slightly better description for the CIP adsorption on both CLI and MAG-CLI. The maximum calculated capacities were 17.30 mg CIP g\(^{-1} \) (at 293 K) and 15.86 mg CIP g\(^{-1} \) (at 288 K) for CLI and MAG-CLI, respectively. Considering the assumptions of the Langmuir model [63], it could be concluded that one active site at the surface of MAG-CLI is occupied by only one CIP species and that the CIP removal from the water solution proceeds as a monolayer adsorption. Furthermore, the Langmuir separation factor \( R_L \) (results not shown) was in the range of 0–1, indicating that the adsorption is a favorable process.

Adsorption Mechanism

The obtained results indicate a strong affinity of both CLI and MAG-CLI towards CIP, which could be explained by electrostatic interactions between the positively charged CIP and negatively charged CLI and MAG-CLI surface. Moreover, the measurements of concentration of exchangeable cations in the filtrate after the CIP adsorption (Table 5) showed that K\(^+\), Mg\(^{2+}\), and Ca\(^{2+}\) are present in the filtrate, suggesting that ion exchange reactions occur during adsorption. Since dimensions of the CIP molecule (1.35 x 0.3 x 0.74 nm) [64] exceed the size of the pore openings in the CIP lattice (0.4 nm), the ion exchange likely
takes place only at the surface of the adsorbent. Ion exchange was also proposed as the main adsorption mechanism for the CIP adsorption by montmorillonite [30].

Table 4. The parameters obtained by the adsorption isotherm Equations (2) and (3) for the adsorption of CIP on CLI and MAG-CLI; $R^2$ is the correlation coefficient.

| $T$, K | CLI | Langmuir Isotherm | Freundlich Isotherm |
|--------|-----|-------------------|--------------------|
|        | $q_{\text{max}}$, mg g$^{-1}$ | $b_L$, dm$^3$ mg$^{-1}$ | $R^2$ | $K_f$, mg g$^{-1}$ (dm$^3$ mg$^{-1}$)$^{1/n}$ | $n$ | $R^2$ |
| 283    | 14.96 | 0.1028 | 0.9944 | 2.24 | 2.05 | 0.9924 |
| 288    | 16.31 | 0.0697 | 0.9877 | 1.73 | 1.80 | 0.9804 |
| 293    | 17.30 | 0.0893 | 0.9909 | 2.34 | 1.97 | 0.9993 |

| $T$, K | MAG-CLI | Langmuir Isotherm | Freundlich Isotherm |
|--------|----------|-------------------|--------------------|
|        | $q_{\text{max}}$, mg g$^{-1}$ | $b_L$, dm$^3$ mg$^{-1}$ | $R^2$ | $K_f$, mg g$^{-1}$ (dm$^3$ mg$^{-1}$)$^{1/n}$ | $n$ | $R^2$ |
| 283    | 13.27 | 0.2235 | 0.9834 | 3.34 | 2.62 | 0.9831 |
| 288    | 15.86 | 0.0914 | 0.9827 | 2.16 | 1.97 | 0.9809 |
| 293    | 14.25 | 0.2263 | 0.9870 | 3.85 | 2.81 | 0.9981 |

$q_{\text{max}}$—maximum adsorption capacity, mg g$^{-1}$; $b_L$—Langmuir constant, dm$^3$ mg$^{-1}$; $K_f$—Freundlich constant, mg g$^{-1}$ (dm$^3$ mg$^{-1}$)$^{1/n}$; $n$—adsorption intensity.

Table 5. The concentration of exchangeable cations in filtrates after the CIP adsorption on CLI and MAG-CLI.

| Sample | Cations | K$^+$ | Mg$^{2+}$ | Ca$^{2+}$ |
|--------|---------|-------|-----------|-----------|
| CLI    | G$^{+}$ | 0.9325 | 1.2513 | 4.2280 |
| MAG-CLI| G$^{+}$ | 0.6796 | 0.3823 | 2.1890 |

3.10. Kinetic Analysis

The CIP adsorption dependence on time was investigated at 283, 288 and 293 K for solutions with $C_0 = 15, 25, 50$ and 75 mg CIP dm$^{-3}$. Figures 11 and 12 show the CIP uptake from water solution for CLI and MAG-CLI, respectively.

For both CLI and MAG-CLI, the CIP uptake increases rather sharply in the first 10 min, as is evident for all studied temperatures and initial concentrations. Later, the adsorption occurs more slowly. More than 80% of the maximum adsorption capacity was achieved within the first 10 min, indicating fast adsorption kinetics.

The data from Figures 11 and 12 were analyzed using two reaction–based kinetic models and a diffusion-based model. The first model is given by the Lagergren’s first-order rate equation [63]:

$$\frac{dq_t}{dt} = k_1 (q_e - q_t)$$  \hspace{1cm} (4)

where $q_e$ (mg g$^{-1}$) is the adsorption capacity at equilibrium and $k_1$ (min$^{-1}$) is the first-order kinetic rate constant. By integrating the Equation (4) using the boundary conditions $t = 0$ to $t = t$ as well as $q_t = 0$ ($t = 0$) and $q_t = q_e$ ($t = t$), the following expression is obtained:

$$\log(q_e - q_t) = \log q_e - (k_1/2.303) t$$  \hspace{1cm} (5)

The plot of $\log(q_e - q_t)$ vs. $t$ is a straight line if the experimental data follow this kinetic model.

The second reaction–based model that was applied is described by the pseudo-second order rate equation [65]:

$$\frac{dq_t}{dt} = k_2 (q_e - q_t)^2$$  \hspace{1cm} (6)
Here $k_2$ (g mg$^{-1}$ min$^{-1}$) is the rate constant of the pseudo-second order adsorption. Integration between the same limits as above gives the following expression:

$$t/q_t = 1/(k_2 q_e^2) + (1/q_e) t$$

(7)

The plot of $t/q_t$ vs. $t$ is a straight line if the experimental data conform to this kinetic model and the values of $q_e$ and $k_2$ are obtained from the slope and intercept of the plot, respectively.

The influence of diffusion in the CIP adsorption was analyzed by the Weber–Morris mass transfer model [28,58]. The following equation defined this model:

$$q_t = K_{id} t^{1/2} + I$$

(8)

where $K_{id}$ (mg g$^{-1}$ min$^{-1/2}$) is the intra-particle diffusion rate constant and $I$ (mg g$^{-1}$) is the parameter related to the thickness of the boundary layer.

![Figure 11. Adsorption kinetics for CIP on CLI for different temperatures; $q_t$ is the amount of the adsorbed CIP (mg per 1 g of CLI) after time $t$.](image-url)
Application of the two reaction-based kinetic models on the experimental data showed that Lagergren’s first-order model gives a poor agreement (the results are not shown since the $R^2$ was lower than 0.80). The pseudo-second order model gives the best fit as judged by the $R^2$ values (Table 6). This suggests that chemisorption occurs during the CIP adsorption. In addition, the maximum calculated capacities obtained for the highest studied concentration, 13.27 mg CIP g$^{-1}$ (at 293 K) and 13.00 mg CIP g$^{-1}$ (at 283 K) for CLI and MAG-CLI, respectively, correspond to the experimentally obtained values. The Lagergren’s pseudo-second order rate constant ($k_2$) was in the range of 0.0346–0.4011 g mg$^{-1}$ min$^{-1}$ for both CLI and MAG-CLI. The $k_2$ values changed rather irregularly with temperature for all initial CIP concentrations. A similar phenomenon was reported for the CIP adsorption on clay minerals and on $\gamma$-Al$_2$O$_3$ [23,24], indicating complexity of the CIP adsorption mechanism.
Table 6. Rate constant for the two studied kinetic models for the adsorption of CIP on CLI and MAG-CLI ($R^2$ is the correlation coefficient of the linear regression).

| $C_0$, mg CIP dm$^{-3}$ | $T$, K | Weber-Morris Model Parameters | Lagergren's Pseudo-Second-Order Rate Parameters |
|-------------------------|--------|-------------------------------|-----------------------------------------------|
|                         |        | $K_{id}$, mg g$^{-1}$ min$^{-1/2}$ | $I$, mg g$^{-1}$ | $R^2$ | $k_2$, g mg$^{-1}$ min$^{-1}$ | $q_e$, mg g$^{-1}$ | $R^2$ |
| CLI                     |        |                               |                               |       |                               |                               |       |
|                         | 15     | 283                           | 0.0729                        | 3.54  | 0.9301                        | 0.3393                        | 4.09  | 0.9999 |
|                         |        | 288                           | 0.0768                        | 3.66  | 0.9638                        | 0.2490                        | 4.28  | 0.9998 |
|                         |        | 293                           | 0.0599                        | 4.29  | 0.9221                        | 0.2275                        | 4.82  | 0.9996 |
|                         | 25     | 283                           | 0.0592                        | 4.35  | 0.7707                        | 0.1896                        | 4.90  | 0.9988 |
|                         |        | 288                           | 0.0820                        | 4.50  | 0.8933                        | 0.2900                        | 5.13  | 0.9999 |
|                         |        | 293                           | 0.0377                        | 4.99  | 0.5450                        | 0.4011                        | 5.24  | 0.9995 |
|                         | 50     | 283                           | 0.3033                        | 7.15  | 0.9436                        | 0.0436                        | 9.78  | 0.9983 |
|                         |        | 288                           | 0.1946                        | 6.93  | 0.9386                        | 0.0703                        | 8.63  | 0.9991 |
|                         |        | 293                           | 0.1632                        | 7.44  | 0.8792                        | 0.0714                        | 8.94  | 0.9989 |
|                         | 75     | 283                           | 0.1442                        | 10.26 | 0.7653                        | 0.0868                        | 11.58 | 0.9997 |
|                         |        | 288                           | 0.1822                        | 9.77  | 0.7612                        | 0.0886                        | 11.28 | 0.9991 |
|                         |        | 293                           | 0.3307                        | 10.46 | 0.9285                        | 0.0443                        | 13.27 | 0.9982 |
| MAG-CLI                 |        |                               |                               |       |                               |                               |       |
|                         | 15     | 283                           | 0.0390                        | 3.07  | 0.7268                        | 0.3133                        | 3.43  | 0.9997 |
|                         |        | 288                           | 0.0877                        | 2.24  | 0.9268                        | 0.1361                        | 3.03  | 0.9964 |
|                         |        | 293                           | 0.0434                        | 3.88  | 0.7702                        | 0.3242                        | 4.26  | 0.9995 |
|                         | 25     | 283                           | 0.1337                        | 4.46  | 0.9771                        | 0.1159                        | 5.59  | 0.9995 |
|                         |        | 288                           | 0.0399                        | 4.88  | 0.4335                        | 0.1185                        | 5.08  | 0.9996 |
|                         |        | 293                           | 0.0885                        | 4.59  | 0.8507                        | 0.1257                        | 5.41  | 0.9980 |
|                         | 50     | 283                           | 0.1972                        | 7.29  | 0.9393                        | 0.1359                        | 8.60  | 0.9995 |
|                         |        | 288                           | 0.1708                        | 6.38  | 0.9613                        | 0.0861                        | 7.84  | 0.9993 |
|                         |        | 293                           | 0.1060                        | 8.21  | 0.5944                        | 0.0673                        | 8.85  | 0.9994 |
|                         | 75     | 283                           | 0.3887                        | 9.59  | 0.7299                        | 0.0346                        | 13.00 | 0.9930 |
|                         |        | 288                           | 0.2409                        | 10.12 | 0.8105                        | 0.0859                        | 12.00 | 0.9994 |
|                         |        | 293                           | 0.3047                        | 9.96  | 0.9832                        | 0.0558                        | 12.46 | 0.9997 |

$K_{id}$—intra-particle diffusion rate constant, mg g$^{-1}$ min$^{-1/2}$; $I$—parameter related to the thickness of the boundary layer, mg g$^{-1}$; $k_2$—pseudo-second-order rate constant, g mg$^{-1}$ min$^{-1}$; $q_e$—adsorption capacity, mg g$^{-1}$. 
Table 6 shows that the analysis of the adsorption data by the Weber-Morris mass transfer model gave parameter I values that were higher than one, suggesting that the intra-particle diffusion is not the rate-limiting step [32,65].

3.11. Leaching Test

Leaching of the adsorbed CIP from the spent adsorbents was studied by treating the spent adsorbents in NaCl solution over 24 h. It was found that 20.6% of the CIP is released from CLI (containing 20.2 mg CIP g\(^{-1}\)) and 4.40% from MAG-CLI (with 22.1 mg CIP g\(^{-1}\)). This suggests the following: (1) magnetite coverage prevents leaching of CIP from the adsorbent, and (2) the CIP adsorption is an irreversible process. The protection of the magnetite coverage could be explained by an interaction of the CIP carboxylic groups with magnetite particles. Namely, adsorptive ability of nano-magnetite was ascribed to the formation of an inner-sphere complex between CIP and magnetite, mediated by carboxylic acid groups [5]. Moreover, the fact that only 20% of the adsorbed CIP can be replaced by Na\(^+\) ions from CLI suggests a strong interaction of CIP and the clinoptilolite surface.

3.12. Regeneration of the Adsorbent

The unsuccessful removal of adsorbed CIP by ion exchange confirms that the adsorption is irreversible. A similar result was also reported for the CIP adsorption onto zeolite A, Y and X [44]. Adsorbent reusability is a very important issue for the operational cost of wastewater treatment based on the adsorption process. With this in mind, we explored a low-temperature atmospheric pressure plasma treatment for the recovery of the spent adsorbent. Plasma treatment is an effective method for the surface modification of various materials including zeolites. Plasma treatments of zeolites were found to increase the removal efficiency of organic species from water after a short treatment time [66,67]. This effect is attributed to an interaction between the reactive species produced in plasma and the zeolite surface [68].

A surface dielectric barrier discharge (SDBD) source with a segmented electrode operating in air was used for plasma treatment of the spent MAG-CLI. Plasma was generated using a 50 Hz sine wave high-voltage signal (V(amplitude) = 12.5 kV) within the \(d = 2\) mm gap between the upper (powered) and lower (grounded) electrode which also served as a sample holder. The duration of the treatment was \(t = 20\) min. The preliminary results of four regeneration cycles are shown in Figure 13.

![Figure 13. CIP removal efficiency on the MAG-CLI during four adsorption and plasma regeneration cycles \((C_0 = 50\) mg dm\(^{-3}\), adsorption time = 60 min, \(T = 298\) K and pH = 5).](image-url)

...
The results indicate that almost 80% of the adsorbent capacity can be recovered by the plasma treatment (Figure 13). This suggests that such a method is suitable for adsorbent recycling. Further investigation into the optimization of the plasma operational parameters is in progress and will be published.

3.13. Antibacterial Test

In order to make the management of the spent adsorbents cost-effective, antibacterial activity of the CIP-containing CLI and MAG-CLI was tested.

Previously published results showed that clinoptilolite itself does not exhibit antibacterial activity towards tested bacteria strains [69]. Figure 14 shows that CIP has a strong antibacterial effect towards both \textit{E. coli} and \textit{S. aureus}, which is in accordance with the literature data [70,71]. Moreover, according to the inhibition zone, it is evident that both CLI-CIP and MAG-CLI-CIP exhibit a strong antibacterial effect. The effect of the CIP, as well as that of the spent adsorbents, is stronger towards \textit{E. coli} than towards \textit{S. aureus}. This can be explained by differences in structure and composition of the cell wall of Gram-negative and Gram-positive bacteria [72,73]. It is important to note that the CIP concentration used in this assessment was 100 times higher than that on CLI-CIP and MAG-CLI-CIP. This suggests that the CIP immobilized on CLI could be used in water disinfection.

\textbf{Figure 14.} Antibacterial activity of CIP, CLI-CIP and MAG-CLI-CIP towards \textit{E. coli} (a) and \textit{S. aureus} (b). Initial number of bacteria ($t_0$): \textit{E. coli} = $1.4 \cdot 10^9$; \textit{S. aureus} = $1.2 \cdot 10^9$ CFU cm$^{-3}$.

4. Conclusions

From the obtained results, the following can be concluded:

- Calcium-rich natural clinoptilolite shows a high adsorptive activity towards antibiotic ciprofloxacin;
- Clinoptilolite strongly adsorbs ciprofloxacin at a pH of 5 via electrostatic interactions and ion exchange reaction occurring between the ciprofloxacin cations and clinoptilolite;
- The adsorption proceeds quickly following the Lagergren’s pseudo-second order rate equation. More than 80% of the maximum adsorption capacity was achieved within the first 10 min for the temperature range of 283 to 293 K;
- Impregnation of clinoptilolite by nano-magnetite particles does not influence the adsorption ability and capacity of clinoptilolite, but brings magnetism to the clinoptilolite-based adsorbent which allows for the easy removal of the spent adsorbent by magnetic separation;
- Magnetite coverage protects the spent adsorbent from the CIP leaching through an interaction of the carboxylic groups of the adsorbed CIP and magnetite particles;
- Preliminary studies indicate that atmospheric pressure plasma could be an efficient method for the regeneration of spent adsorbent;
• Ciprofloxacin-containing clinoptilolite shows strong antibacterial activity towards pathogens (E. coli and S. aureus), suggesting its possible use in a tertiary stage of water treatment.

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