Immune biomarkers of response to immune-checkpoint inhibitors in head and neck squamous cell carcinoma

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Anti-programmed cell death protein 1 (PD-1) agents have become the standard of care for platinum-refractory recurrent/metastatic head and neck squamous cell carcinoma (HNSCC) and are currently being evaluated in various disease settings. However, despite the gain in overall survival seen in some of the clinical trials, the majority of patients display primary resistance and do not benefit from these agents. Taking into consideration the potentially severe immune-related toxicities and their high cost, the search for predictive biomarkers of response is crucial. Besides Programmed death ligand-1 (PD-L1) expression, other biomarkers such as immune infiltration, tumor mutational burden or immune-gene expression profiling have been explored, but none of them has been validated in this disease. Among these, the microbiota has recently garnered tremendous interest since it has proven to influence the efficacy of PD-1 blockade in some tumor types. With the accumulating evidence on the effect of the microbiota in HNSCC tumorigenesis and progression, the study of its potential role as a predictive immune biomarker is warranted. This review examines the available evidence on emerging immune predictive biomarkers of response to anti-PD-1/PD-L1 therapy in HNSCC, introducing the microbiota and its potential use as a predictive immune biomarker in this disease.

Key words: head and neck squamous cell carcinoma, immune checkpoint inhibitors, anti-PD-1/PD-L1, biomarkers, microbiota

Introduction

Immune-checkpoint inhibitors (ICI) targeting cytotoxic T-lymphocyte antigen 4 and programmed cell death protein-1 (PD-1) and its ligands, programmed death ligand-1 (PD-L1)/2, have shown a significant and consistent benefit in survival when compared with standard therapies in prospective randomized clinical trials, leading to their regulatory approval in multiple tumor types [1–5]. In head and neck squamous cell carcinoma (HNSCC), anti-PD-1 antibodies are the first immunotherapeutic agents to demonstrate evidence of response durability and survival benefit in platinum-pretreated recurrent and metastatic (R/M) disease [6–9]. However, despite the encouraging results which led to the approval of nivolumab and accelerated approval of pembrolizumab by the US Food and Drug Administration (FDA) for platinum-refractory R/M HNSCC, the overall response rates (ORRs) of these agents ranged from only ~13%–18% [9, 10].

Up to 60% of patients across different tumor types, including HNSCC, display primary resistance to anti-PD-1/PD-L1 agents [11]. Several mechanisms have been suggested such as poor tumor immunogenicity, limited intratumoral immune cell infiltration, coexpression of multiple inhibitory receptors, and induction of immunosuppressive pathways within the tumor microenvironment (TME) [12–14]. To overcome this resistance,
many ongoing clinical trials are evaluating combination strategies with other immunotherapies, targeted agents, chemotherapy and radiotherapy, not only in R/M HNSCC, but also in the locoregionally advanced setting (NCT02952586, NCT03040999) [15]. This is of particular relevance as a proportion of patients with R/M HNSCC might experience rapid progression and decreased survival when treated with single-agent anti-PD-1/PD-L1 [16].

However, the potential immune-related toxicities of ICI and their high cost have urged the search for prospectively validated predictive biomarkers of response including PD-L1 protein expression, intratumoral immune cell infiltration, immune-gene expression profiling, and tumor mutational burden (TMB) [13, 14, 17]. Specifically, in HNSCC, none of them have been validated and ongoing exploration continues [9, 18].

Recently, the immunomodulatory role of the gut microbiota, defined as the collective microorganisms inhabiting the gastrointestinal tract, has raised a special interest, since its composition has proven to influence anti-PD-1 efficacy in preclinical models and has been associated with treatment responsiveness in patients with melanoma and some epithelial-derived tumors [19–22]. Interestingly, many retrospective studies in HNSCC have suggested that the oral microbiota might also be crucial for tumor development and progression, treatment-related toxicity and disease recurrence [23–25].

This review examines the available evidence on emerging immune predictive biomarkers of response to ICI in HNSCC, introducing the microbiota and its potential use as a predictive immune biomarker in this disease (Table 1).

Overview of emerging immune biomarkers in HNSCC

Is PD-L1 expression a reliable biomarker of response in HNSCC?

PD-L1+ tumors in general tend to demonstrate improved response rates to anti-PD-1/PD-L1 therapies, in comparison to PD-L1− tumors [26]. This correlation has been consistent with different anti-PD-1/PD-L1 drugs across many tumor types [5, 27, 28]. Most clinical trials evaluating ICI in R/M HNSCC suggested a similar pattern [29–31], and data from phase III randomized trials investigating pembrolizumab in the R/M setting

| Immune biomarkers | Assay | Predictive value in HNSCC | Evidence available |
|-------------------|-------|--------------------------|-------------------|
| PD-L1 expression  | PD-L1 staining by immunohistochemistry in tumor cells/immune cells (different cut-offs) | Positive<sup>b</sup> | Positive<sup>b</sup> | Prospective randomized clinical trials (Table 2). |
| Smoking           | • Smokers versus nonsmokers  
|                   | • Smoking mutational signatures in tumor samples | Negative | Uncertain | Retrospective analysis of prospective trials [9]. |
| Tumor immune-cell infiltration | Presence of CD8<sup>+</sup> T cells  
|                   | PD-1<sup>+</sup> TIM-3<sup>+</sup> CD8<sup>+</sup> T cells  
|                   | PD-1<sup>+</sup> LAG-3<sup>+</sup> CD8<sup>+</sup> T cells | Positive | Negative | Retrospective analysis of noncontrolled cohorts [73]. |
| Circulating immune cells | PD-1<sup>+</sup> CD8<sup>+</sup> T cells  
|                   | FoxP3<sup>+</sup> Tregs | Negative | Negative | Prospective analysis in a randomized clinical trial [102]. |
| Tumor mutational burden | Number of somatic coding missense mutations.  
|                   | • Tumor samples  
|                   | • Blood samples | Positive | Uncertain | Retrospective analysis of prospective clinical trial [74, 75]. |
| T-cell-inflamed phenotype | Immune-related gene expression signatures | Positive | Positive | Retrospective analysis of prospective clinical trial [74, 75, 104]. |
| Microbiota        | 16S rRNA high throughput sequencing of saliva and stool | Oral microbiota: nonpredictive  
|                   | Intestinal microbiota: no data yet | Retrospective analysis of prospective randomized clinical trial [133]. |

<sup>a</sup>Predictive values in HPV− and HPV+ subgroups were defined positive or negative if a statistically significant correlation between response and the immune biomarker was described in the referenced studies; uncertain if no significant correlation was found; no data if no studies had evaluated the role of the biomarker in this setting at the time of this publication.

<sup>b</sup>The positive correlation between PD-L1 expression and treatment response was not consistent across the studies.
Table 2. Anti-PD-1/PD-L1 agents tested in R/M HNSCC [6–9, 29–31, 33, 34, 52, 134]

| Agents         | Target   | Phase/study | N   | PD-L1 expression Location | Cut-off | ORR (%) | OS (HR)* |
|----------------|----------|-------------|-----|----------------------------|---------|---------|----------|
|                |          |             |     |                            | Overall | PD-L1+     | PD-L1-     | Overall | PD-L1+     | PD-L1-     |
| Nivolumab      | PD-1     | III (CHECKMATE-141) | 240 | TCs only                   | >1%     | 13.3%   | 17%       | 11.8%   | 0.68       | 0.55       | 0.73       |
| Pembrolizumab  | PD-1     | I (KEYNOTE-012) | 132 | TCs+ICs                    | >1%     | 18%     | 22%       | 4%      | NA         | NA         | NA         |
|                |          | III (KEYNOTE-040) | 247 | TCs+ICs(CPS)               | CPS > 1%| 14.6%   | 17.3%     | 0%      | 0.80 (P 0.016) | 0.74 (P 0.0049) | Ø         |
|                |          | III (KEYNOTE-048) | 882 | TCs+ ICs(CPS)              | CPS > 0%| 26.6%   | 0%        | Ø       | 0.53 (P 0.0014) | Ø         | Ø         |
| Durvalumab     | PD-L1    | I (MED4736-1108) | 62  | TCs only                   | >25%    | 10%     | 18%       | 8%      | NA         | NA         | NA         |
|                |          | II (HAWK)   | 112 | TCs only                   | >25%    | NA      | 16.2%     | NA      | NA         | NA         | NA         |
|                |          | II (CONDOR) | 67  | TCs only                   | <25%    | NA      | NA 6%     | 9%      | 0.99 (P 0.089) | Ø         | Ø         |
| Atezolizumab   | PD-L1    | I (GO27831) | 32  | ICs (CPS)                  | CPS >20 | 23.3%   | 0%        | Ø       | 0.61 (P 0.0007) | Ø         | Ø         |

*HR for OS resulting from: nivolumab and pembrolizumab versus investigator’s choice of chemotherapy (Docetaxel, Methotrexate and Cetuximab) in the CHECKMATE-141 and KEYNOTE-040 studies, respectively; pembrolizumab monotherapy versus EXTREME regimen in the KEYNOTE-048 study; durvalumab versus tremelimumab plus durvalumab in the CONDOR study.

ORR, overall response rate; OS, overall survival; HR, hazard ratio; TCs, tumor cells; ICs, immune cells; CPS, number of PD-L1-positive cells (tumor cells, lymphocytes, macrophages) divided by total number of tumor cells x 100; TPS, percentage of tumor cells with membranous PD-L1 expression; NA, not applicable; Ø, no data available.

(KEYNOTE-040 and KEYNOTE-048) endorsed this trend by demonstrating significantly increased survival in PD-L1+ patients [8, 32, 33]. However, CHECKMATE-141 failed to show a significant correlation between PD-L1 expression and tumor response or survival when evaluating nivolumab in the platinum-refractory R/M setting [9, 34] (Table 2).

The discordance of the results across studies might be explained by several reasons. One of the most relevant is the lack of uniformity in the assays and the variability in the thresholds used to define PD-L1 positivity, which have led to the launch of harmonization projects on PD-L1 assays by the scientific community and regulatory agencies [28, 35, 36]. This inconsistency is evident in the development of anti-PD-1/PD-L1 agents investigated to date in R/M HNSCC, including pembrolizumab, nivolumab, atezolizumab, durvalumab and avelumab, thus impairing cross-study comparisons and undermining the value of PD-L1 as a biomarker [6, 9, 30–32, 37, 38]. Importantly, PD-L1 expression seems to be regulated by multiple signaling pathways, including MAPK, PI3K and Akt/PKB that are commonly altered in HNSCC [39–41]. As a consequence of these molecular crosstalks, PD-L1 is a dynamic biomarker that is subject to temporal variations and spatial heterogeneity. Its expression may change from the point of initial diagnosis to recurrence or progression, and may differ between primary and coexisting metastatic lesions [42–45].

Published reports on the intratumoral heterogeneity of PD-L1 expression in HNSCC demonstrate conflicting results [46, 47]. In HNSCC, PD-L1 is highly expressed not only by tumor cells, but also by immune cells present in the TME, including regulatory T cells (Tregs), natural killer (NK) cells and antigen presenting cells (APCs) [18, 48–51]. Across various cancer types, it remains unclear whether PD-L1 expression and thresholds should take into consideration all or only selected cell populations. Both pembrolizumab and atezolizumab used combined scores based on the ratio between tumor cells and immune cells expressing PD-L1 to define tumor PD-L1 positivity, and pembrolizumab did show a positive correlation with response and survival in the phase III KEYNOTE-040 study when using the combined positive score (CPS) [52]. Recently, the results from the phase III KEYNOTE-048 study in first line R/M HNSCC revealed that pembrolizumab monotherapy improved OS when compared with the EXTREME regimen in patients whose tumors had PD-L1 expression ≥1% and ≥20% by CPS [hazard ratio (HR) 0.78 (0.64–0.96), P = 0.0086 and HR 0.61 (0.45–0.83), P = 0.0007, respectively] [33]. However, in KEYNOTE-040, the correlation with clinical outcome was also strongly positive when using PD-L1 expression in tumor cells only (TPS ≥ 50%), congruent with the experience in non-small-cell lung cancer (NSCLC) in KEYNOTE-010 [53, 54]. In contrast, there was no correlation in the nivolumab CHECKMATE-141 study where PD-L1 expression was exclusively determined in tumor cells, although the thresholds used were different (>1%, 5% and 10%) [9]. These divergent results and the limited data available suggest no firm conclusion can be made in this regard, although CPS seems to be more predictive than TPS in HNSCC, and the required cut-off for the latter appears to be higher in the mentioned studies.

Nonetheless, it is noteworthy that, although relevant in a smaller percentage, PD-L1− tumors also benefit from ICI [9].
Therefore, additional factors beyond PD-L1 expression, such as human papillomavirus (HPV) status, tumor immune infiltration or TMB, might also contribute to treatment response.

Are HPV+ tumors more responsive to immunotherapy?

HPV+ oropharyngeal squamous cell carcinoma (OPSCC) is a biologically distinct disease with better prognosis and improved treatment responsiveness when compared with HPV− disease at the same or similar stage [55–57]. Virus-related tumor types are postulated to be more responsive to ICI due to intrinsic characteristics including baseline tumor immunogenicity, increased immune infiltration and increased PD-L1 expression [58, 59]. HPV+ OPSCC have been shown to have a less immunosuppressive TME when compared with HPV− HNSCC, as it harbors greater infiltration by tumor infiltrating lymphocytes (TILs), higher proportion of CD8+ T cells, increased levels of interferon gamma (IFN-γ), decreased CD4+/CD8+ ratio, and lower numbers of Tregs [60–64]. These findings can be explained by a preexisting adaptive host immune response against viral and tumor-specific antigens, which may in turn lead to PD-L1 expression in immune cells. Indeed, a recent retrospective study showed that not only CD8+ TILs (>30%) but also high PD-L1 expression in immune cells (>5%) were both favorable prognostic factors in HPV+ disease regardless of stage [65, 66].

Altogether these findings suggest a potentially higher sensitivity of HPV+ disease to immune-checkpoint blockade. This hypothesis was initially supported by the results from the HNSCC cohort of the multibasket phase I KEYNOTE-012 trial in which HPV+ tumors had increased ORR to pembrolizumab compared with those that were HPV− (25%–32% versus 14%) [6, 7]. However, these results were not reproduced in the phase III KEYNOTE-040 trial, and further studies investigating other anti-PD-1/PD-L1 agents have reported mixed results. For instance, increased response rates were observed among HPV+ patients treated with durvalumab while no differences were seen with atezolizumab [30, 31]. In the CHECKMATE-141 study, nivolumab did not yield significant differences in ORR or OS between HPV+ and HPV− patients [HR for OS 0.60 (0.37–0.97) versus 0.59 (0.38–0.92), respectively] [9, 32, 34].

The inconsistencies in the abovementioned trials might be explained by other coexisting factors beyond PD-L1 expression and immune infiltration. Smoking, mutational signatures and TMB are thought to influence response to ICI in HNSCC although their relevance differs between HPV+ and HPV− disease (Table 1).

Smoking seems to contribute to a more immunosuppressive TME and negatively impact on anti-PD-1/PD-L1 efficacy in HNSCC. In CHECKMATE-141 study, the subgroup analysis reported a trend toward decreased survival benefit from nivolumab among smokers when compared with nonsmokers [9]. Similarly, a retrospective analysis of 81 HNSCC patients treated with anti-PD-1/PD-L1 showed that former/current smokers were less responsive to these agents when compared with never smokers. However, this correlation only remained significant among HPV+ patients, suggesting the immunosuppressive effects of smoking may not be as significant in HPV+ tumors [67]. In support of this, a genomic analysis of 287 HNSCC tumor samples revealed that smoking history and tumors with high smoking mutational signatures were correlated with decreased immune infiltration and downregulation of immune-signaling pathways in HPV+ but not HPV− tumors [67].

Conversely, the presence of other mutational signatures unrelated to smoking such as APOBEC (apolipoprotein B mRNA editing enzyme, catalytic polypeptide-like) is of particular relevance in HPV+ disease. Reduced exposure to exogenous carcinogens such as tobacco seems to favor the emergence of tumors with APOBEC-driven mutations such as PIK3CA [68, 69]. Moreover, APOBEC activity is known to be crucial for innate and adaptive immune responses, and HPV infection is thought to enhance it in an attempted host immune response against the virus. In a study analyzing over 500 HNSCC tumor samples, APOBEC mutational signatures were associated with upregulation of immune-signaling pathways [69]. APOBEC-driven mutagenesis might alter tumor immunogenicity in HPV+ disease impacting on immune checkpoint efficacy. Parenthetically, the presence of APOBEC signatures has been associated with increased immune infiltration and PD-L1 expression in other tumor types [70–72].

Increased TMB and neoantigen load have been shown to correlate with response to ICI in HPV+ HNSCC, whereas most of the studies conducted to date have refuted their predictive value in HPV− patients [73–75]. TMB is a quantitative measure of the total number of coding mutations in the tumor genome. Theoretically, the higher the number of missense mutations, the higher expression of tumor neoantigens which can elicit the greatest antitumor immune response and increase sensitivity to ICI. A retrospective analysis from KEYNOTE-012 and -055 demonstrated a stronger correlation between response to pembrolizumab and high TMB and neoantigen load in the HPV+ subgroup than HPV− subgroup [75]. As a matter of fact, in virally induced tumors such as HPV+ tumors or Merkel-cell carcinoma, response rates to ICI are higher than expected when adjusted for TMB and compared with other tumors types, suggesting immune responses may also be triggered by virus-specific antigens rather than by tumor-neoantigens alone [39, 76–78]. In support of this, a retrospective study analyzing a cohort of 126 patients with R/M HNSCC treated with anti-PD-1/PD-L1 agents showed that HPV+ patients had, as expected, lower TMB (8.2 versus 4.7 mut/MB, P = 0.01) when compared with HPV+ disease, while the number of responses was similar (7 versus 10 responses, P = 0.54) [73]. More importantly, among HPV+ patients, responders had increased CD8+ TILs regardless of TMB.

Overall, with the current available data, it is not possible to determine whether HPV+ OPSCC have higher (or lower) sensitivity to ICI when compared with HPV− disease. HPV positivity alone does not seem to be a reliable biomarker of response to ICI and needs to be interpreted along with other companion clinical and molecular biomarkers.

Is there a role for tumor immune infiltration and T-cell-inflamed phenotypes?

Tumor immune infiltration implies initial recognition by the immune system and might indicate an antitumor immune response [79]. Multiple immune cells coexist within the TME, including TILs (CD8+ T cells and Tregs), NK cells, macrophages, APC and myeloid-derived suppressor cells. The composition of these
immune cells within TME, recently defined as immune contexture, has prognostic implications but can also be predictive of response to therapies [17, 61, 80]. For instance, CD8\textsuperscript{+} T-cell infiltration at baseline has been correlated with increased response to anti-PD-1/PD-L1 agents in melanoma [81, 82].

HNSCC tumors are highly immune-infiltrated but overall characterized by an immunosuppressive TME [48, 83]. Many retrospective studies have attempted to assess the prognostic and predictive value of tumor immune cell infiltration (supplementary Table S1, available at Annals of Oncology online) [18, 62, 63, 84–89]. Despite the heterogeneity of these studies, increased infiltration by CD8\textsuperscript{+} T cells is the only immune cell type in HNSCC consistently proven to be correlated with increased survival regardless of tumor location, stage and treatment [61, 65]. A retrospective evaluation of 126 patients diagnosed with R/M HNSCC treated with anti-PD-1/PD-L1 agents showed that increased tumoral infiltration by CD8\textsuperscript{+} T cells and an increased ratio CD8\textsuperscript{+} T cells/Tregs were positively correlated with treatment response, indicating their potential role as predictive biomarkers [73].

In addition, the relative proportion of the various immune cell subsets and their location within the TME may be of relevance in predicting response to ICI. The immunoscore (IS) is a tool quantifying the density of CD8\textsuperscript{+} T cells within the tumor center versus the invasive margin. Increased number of CD8\textsuperscript{+} T cells in the tumor center (high IS) is thought to indicate an effective antitumor immune response and has been proven to be an independent prognostic biomarker in early stage colorectal cancer, melanoma and NSCLC [80, 90–92]. In HNSCC, a high IS is associated with lower levels of Tregs, increased PD-L1 and MHC type I expressions in tumor cells [62, 93], suggesting its potential to identify a subset of tumors with increased sensitivity to anti-PD-1/PD-L1 therapy. However, the predictive role of IS in HNSCC has not been explored yet.

The coexpression of other inhibitory immune-checkpoint molecules such as TIM-3 (T-cell immunoglobulin and mucin domain-containing protein 3), lymphocyte-activating gene 3 (LAG-3) and T-cell immunoreceptor with Ig and ITIM domains (TIGIT) has also shown to impair immune T-cell-mediated responses, conferring resistance to anti-PD-1/PD-L1 agents in preclinical models and in patients across different tumor types such as melanoma and NSCLC [15, 94–99]. In HNSCC, a high IS is associated with lower levels of Tregs, increased PD-L1 and MHC type I expressions in tumor cells [62, 93], suggesting its potential to identify a subset of tumors with increased sensitivity to anti-PD-1/PD-L1 therapy. However, the relative proportion of the various immune cell subsets and their location within the TME may be of relevance in predicting response to ICI. The immunoscore (IS) is a tool quantifying the density of CD8\textsuperscript{+} T cells within the tumor center versus the invasive margin. Increased number of CD8\textsuperscript{+} T cells in the tumor center (high IS) is thought to indicate an effective antitumor immune response and has been proven to be an independent prognostic biomarker in early stage colorectal cancer, melanoma and NSCLC [80, 90–92]. In HNSCC, a high IS is associated with lower levels of Tregs, increased PD-L1 and MHC type I expressions in tumor cells [62, 93], suggesting its potential to identify a subset of tumors with increased sensitivity to anti-PD-1/PD-L1 therapy. However, the predictive role of IS in HNSCC has not been explored yet.

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Tumor mutational burden and HNSCC mutational landscape

TMB has been recently evaluated as a potential biomarker of response to immune checkpoint blockade in prospective clinical trials and across many tumor types [77, 106–109]. An initial retrospective analysis of 27 tumor types and subtypes among patients who received PD-1/PD-L1 inhibitors demonstrated a significant correlation between TMB and response rate to these agents [77]. In this study, TMB was reported as a median number of coding somatic mutations per megabase (N mut/MB). Melanoma and squamous cell carcinoma of the skin (15–50 mut/MB) followed by tobacco-related cancers including NSCLC, urothelial cell carcinoma and HNSCC (5–10 mut/MB) comprised malignancies with the highest TMB [77]. Retrospective subset analyses of clinical trials evaluating pembrolizumab, atezolizumab and nivolumab in metastatic melanoma, NSCLC, urothelial carcinoma and HNSCC have demonstrated not only increased ORR but also improved survival in patients with high TMB [75, 106–108, 110]. These results were consistent across the studies, tumor type and anti-PD-1/PD-L1 agents. However, the cut-off and measure used to define a high TMB differed between studies, thus precluding direct comparisons. These results were further supported by a retrospective analysis of 126 HNSCC patients treated with anti-PD-1/PD-L1 agents. TMB was found to be significantly higher among responders (21.3 versus 8.2 mut/MB, \(P < 0.01\)) and was correlated with increased median OS (20 months if TMB > 10 mut/MB versus 6 months if TMB < 5 mut/MB, \(P = 0.01\)) in HPV\textsuperscript{+} disease [73]. A combined biomarker analysis of multiple studies evaluating the correlation between TMB, T-cell-inflamed GEP, PD-L1 expression by CPS and response to pembrolizumab in HNSCC showed no
significant correlation between TMB and inflammatory biomarkers (i.e. GEP or PD-L1). While this analysis did not stratify by HPV status, it suggests TMB and inflammatory biomarkers have distinct and independent predictive values, and may be used orthogonally to identify responders to pembrolizumab [105].

In addition to TMB, the specific tumor mutational landscape might be of biological relevance. Tumors characterized by mutations affecting DNA damage response, such as those with microsatellite instability high (MSI-H) or mismatch repair deficiency (dMMR), have the highest mutational load [77, 111]. These tumors have been shown to be particularly sensitive to ICI in prospective clinical trials, leading to the FDA approval of pembrolizumab for patients with dMMR or MSI-H tumors, regardless of histology [112, 113]. The estimated incidence of MSI-H tumors among HNSCC has been reported to be about 8% [114]. However, a recent study identified a subgroup of HNSCC responders to anti-PD-1/PD-L1 whose tumors were enriched with somatic mutations derived from frameshift events in tumor suppression genes such as NOTCH and SMARCA4 [73]. These cases are similar to what has been described in tumors with dMMR, with baseline increased mutational burden and greater sensitivity to ICI. The authors suggested this finding might represent a novel mutational signature in HNSCC with potential predictive value, although further validation is warranted.

HNSCC genomic classification described by the TGCA might be considered as well [39]. Four subtypes have been defined on the basis of gene expression: atypical, mesenchymal, basal and classical. The mesenchymal subtype, e.g. characterized by alterations in genes related to innate immunity, downregulation of MHC type I expression and deficient antigen-presentation machinery, would unlikely respond to anti-PD-1/PD-L1 agents.

Overall, while the predictive role of the specific molecular subtypes is yet to be explored, TMB has shown promising results and might become a useful predictive biomarker of immune-checkpoint blockade efficacy in HNSCC. However, similar to what occurred with the PD-L1 assay, the lack of uniformity in the methods used to determine the mutational burden (e.g. measured in the tumor or in the blood) and the variability of the thresholds used across studies are hampering the interpretation and extrapolation of the results obtained. Thus, standardization should be pursued when designing biomarker-validating studies using TMB. Moreover, TMB has not shown to correlate with PD-L1 expression or GEP signatures [73, 75, 105], again indicating the interactions between the tumor, TME and the immune system are complex and dynamic.

**Introducing the microbiota as a potential immune biomarker for HNSCC**

**The microbiota in head and neck cancer**

The composition of the microbiota present in the orogastrointestinal tract has been associated with immune dysregulation and initiation and progression of many cancers [23, 115–118]. The precise mechanisms of these associations are not known, but compositional and functional changes in the microbiota can induce or exacerbate chronic inflammation, resulting in cell damage and alteration of local and systemic immune homeostasis, which may affect local and distant carcinogenesis, ultimately dampening or enhancing antitumor immune responses [116, 119]. HNSCC arise from an epithelium and mucosa located in the oral cavity and the pharynx; both sites are constantly exposed to environmental factors that can alter the oral microbiota [120, 121]. Retrospective cohort studies have shown different microbiota composition in the saliva of HNSCC patients compared with healthy controls, while the presence of specific bacteria has been associated with reduced risk of developing HNSCC [23, 122–124]. Moreover, differentially enriched microbiota found in HPV+ and HPV+ OPSCC and oral cavity SCC indicates the existence of specific microbiota according to tumor location and HPV status [24]. Nonetheless, some authors have underlined the challenge of distinguishing whether the changes observed in the oral microbiota from HNSCC patients are influenced by the TME and/or by local and systemic cancer therapies, since most of the studies to date have retrospectively evaluated small, heterogeneous and noncontrolled cohorts of patients comprising different tumor sites, variable disease stages, and treatment with multiple modalities [23]. In this regard, a study analyzing the oral microbiota present in the saliva of HNSCC patients before and after treatment [including surgery, chemoradiotherapy (CRT) and ICI] showed an association between specific oral bacteria composition (Fusobacterium and Lactobacillus), down-regulation of immune-signaling pathways and upregulation of oncogenic Wnt/Beta-catenin pathways [125]. Altogether these findings suggest that the oral microbiota might represent a promising prognostic and predictive biomarker in this disease (Figure 1).

**Exploiting the microbiota as a biomarker of response to immunotherapy**

Accumulating evidence has implicated that intestinal microbiota can modulate host anticancer immune responses and alter the efficacy of anticancer therapies, including immunotherapy [19, 126–131]. Two preclinical studies using mouse models of melanoma and lung cancer revealed a correlation between the presence of specific commensal intestinal bacteria (Bifidobacterium) and response to ICI [20, 132]. This was further supported by two recent publications evaluating the gut microbiome in patients with melanoma and epithelial-derived tumors, showing improved anti-PD-1/PD-L1 efficacy among patients harboring specific intestinal bacteria (the species of Akkermansia muciniphila and members of the Ruminococcaceae family) and higher microbial diversity [21, 22]. Remarkably, these microbiota were also correlated with enhanced local and systemic immune response, reduction in tumor growth and restoration of response to anti-PD-1/PD-L1 therapy in germ-free mice transplanted with fecal microbiota from responding patients. These latter findings indicate the potential modulation of the microbiota as a viable therapeutic target to increase response to ICI.

Whether the microbiota has a role in predicting response to immunotherapy in HNSCC is yet to be determined. Only one substudy from CHECKMATE-141 explored the role of the oral microbiota measured in the saliva as a predictive biomarker in patients with R/M HNSCC treated with nivolumab, showing no
significant correlation with treatment efficacy or survival [9, 133]. However, the study had several limitations, including the lack of uniformity in sample collection, the small number of responses for correlation and importantly, the omission of intestinal microbiota. The predictive role of the oral microbiota was also investigated in melanoma patients treated with anti-PD-1/PD-L1 therapy, again reporting no association with treatment outcome, in contrast to the positive correlation observed with the intestinal microbiota composition [22]. Differential bacterial composition between these anatomical sites suggests oral and intestinal microbiota likely represent distinct entities with specific disease associations.

Considering the immunomodulatory effects of the intestinal microbiota and the growing evidence of the oral microbiota impacting HNSCC tumorigenesis and progression, the study of their role as a predictive biomarker of response to ICI in this disease is warranted. Hence, our group is currently conducting a research study at the Princess Margaret Cancer Centre to prospectively evaluate the oral and intestinal microbiota in a homogeneous cohort of patients diagnosed with locoregionally advanced OPSCC treated with definitive chemoradiotherapy. The overarching goal of this project is to characterize and explore the correlation with both oral and intestinal microbiota measured in the saliva and stool, respectively, by using 16S rRNA sequencing, in order to obtain a deeper understanding of their relationship with treatment response. The results of this ongoing study will serve as a fundamental basis to evaluate oral and intestinal microbiota signatures and their role as predictors of response to ICI in patients treated within the CCTG HN.9 clinical trial, a multicenter phase II noncomparative randomized study evaluating ICI plus RT followed by maintenance ICI versus standard chemoradiotherapy in intermediate-risk, HPV+ locoregionally advanced OPSCC (NCT034106615).

**Discussion**

**Conclusion**

Anti-PD-1 agents have become the standard of care for the platinum-refractory R/M HNSCC. Results from clinical trials evaluating their role in additional disease settings are pending, but clearly such compounds are already an important therapeutic backbone in this malignancy. As such, appropriate selection of patients who will benefit from these therapies is crucial. To date, there are no validated predictive biomarkers of response that are applicable uniformly to all HNSCC patients, although many candidate biomarkers with promising results are undergoing investigations. A systematic computational analysis of all clinically
annotated biomarker data would be invaluable to further the knowledge in this field.

Most of the biomarkers in HNSCC have been explored retrospectively, often using baseline archival tumor samples at a single time point which may not reflect the impact of spatial and temporal intratumoral heterogeneity. Also, standalone evaluation of potential biomarkers without considering interactions with other factors is likely oversimplifying the complexity of immune response. The microbiota is a dynamic and complex ecosystem that interrelates the immune system and the TME, thus, potentially representing an ideal biomarker that reflects the interactions between these biological entities in totality. Both oral and intestinal microbiota may be important regulators of local and systemic immune responses induced by environmental factors, shaping the TME and ultimately modulating the efficacy of cancer therapies. Considering the emerging immunomodulatory effects of the microbiota, the study of its role as a predictive immune biomarker in HNSCC is of special interest and should be integrated into prospective clinical trials.

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