Pilot study: bone marrow stem cells as a treatment for dogs with chronic spinal cord injury

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Abstract

Background: Chronic Spinal Cord injury is a common, severe, and medically untreatable disease. Since the functional outcomes of acute and experimental chronic spinal cord injury have been shown to improve with stem cell therapy, a case study was conducted to test if the application of stem cell also regenerates chronic SCI dysfunction. Transplantation of foetal bone marrow stem cells was applied in seven dogs with chronic spinal cord injury. Magnetic resonance images and assessments of symptoms according to the Olby scale were used to diagnose the severity of injury.

Result: All dogs improved locomotor and sensory function when examined 90 days after surgery, and showed increased movement of the hind limbs, and were able to stand upright, as well as to take small steps. Tail tone was observed in seven dogs, pain reflexes and defecation return were observed in five dogs.

Conclusion: The transplantation of bone marrow stem may be a promising, reliable and safe treatment for chronic spinal cord injury.

Keywords: Cell therapy, Dysfunction, Spinal cord, Disease

Background

Spinal Cord injury (SCI) is a disease with devastating effect on dogs, including paresis or paralysis and/or urinary and faecal incontinence. Chronic SCI is regarded as a medically untreatable condition and there is no effective treatment [1-3]. Spinal cord lesions are commonly reported in veterinary medicine, especially in dogs, where they usually occur in association with traumas that are induced by prolapsed intervertebral discs or exogenous sources such as motor vehicle accidents [4,5,1]. In humans, between 3 and 5 people per every 100,000 are affected with SCI [6]. Clinical signs in dogs are similar to those encountered in human patients, and dogs that suffer from severe SCI have the same poor prognosis for neurological recovery as their human counterparts [7]. The severity of the neurological signs are graded as follows: grade I (only spinal hyperaesthesia), grade II (ambulatory paraparesis, ataxia, proprioceptive deficits), grade III (non-ambulatory paraparesis), grade IV (paraplegia with nociception), and grade V (paraplegia with loss of nociception) [8]. Likewise, a scale of 14 points is used to assess the severity of the disease [9].

Diagnostic imaging has become increasingly important for assessing the prognosis and determining treatment decisions with intervertebral disc extrusion [8,10-12]. For dogs, the temporal aspects of the onset and duration of clinical signs after intervertebral disc herniation have been used as prognostic indicators [1], suggesting that clinical assessment is essential after SCI [12].

Treatments with stem cells have been performed in animal and human SCI models, with promising results [3,13-16]. In particular, transplantation of stem cells has been shown to improve the functional outcome of experimental SCI in rats [17-19]. Pre-clinical reports indicate that the transplantation of bone marrow mesenchymal stromal cells is encouraging in acute, SCI-simulating models, including monkeys [20], humans [21] and dogs [22,23]. The aim of the current pilot project is to evaluate if transplants of foetal canine bone marrow derived stem cells into spinal cord lesions improved the regeneration.
and spinal cord function in dogs with severe, chronic spinal cord lesions.

**Results**

On the day following surgery, the animals did not present with any adverse symptoms. There was no formation of local oedema, seroma, or other complications. No feeding issues or associated loss of weight was noted during the investigative period. Sutures were removed after 10 days without problems.

MRI did not show major differences after the treatment (Figure 1). All animals that were considered for statistical analysis showed significant improvement of the Olby scales after stem cell transplantation (Table 1, Figure 2). They also showed increased movement of the hind limbs on both treadmills. Animals 3, 4, 6 and 7 were able to support their own weight and to make small, uncoordinated steps and returned tail tone, superficial and deep pain reflexes as well as defaecation (Table 1). In detail, Animal 1 suffered from a flaccid paralysis without significant joint contracture that became slightly better after surgery, including movements of the hind limbs and improved muscle contraction, indicating a reactivation of the nervous pathways. Animal 2 showed

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**Figure 1 MRI of dogs thoracolumbar segment.** In (A) the pre treatment the animals showed severe extradural compressive myelopathy ventrolateral between T11-T12 and disc protrusion at L3-L4, disc degeneration and spondylosis multifocal in thoracic and lumbar spine. In (B) after treatment the animal showed the same diagnosis at MRI without regression or progression of the previous reported. In (C) homogeneous hyperintensity maintained between T9-L2, extradural compressive myelopathy, mild to moderate between T12-T13, previous surgical procedure between T11-T13, with secondary muscle atrophy, disc dehydration multifocal in thoracolumbar spine were observed. In (D) after the treatment the same alterations were observed in the MRI. In (E) myelopathy extradural compressive between T9-T120 and T13-L1, with the presence of secondary siringohidromielia. In (F) after treatment probable thoracolumbar muscular atrophy with presence of siringohidromielia between T12-L1 and L3, and severe at T13-L1, multifocal disc dehydration in thoracolumbar spine.
sharp muscle spasm that compromised the joints of the hind limbs. After surgery, the muscles recovered, in addition to an increased range of motion (Table 1). Animal 3 had little muscle contracture before therapy, but showed improved movement of the hind limbs in the aquatic treadmill afterwards. The animal was able to hold its body weight and to protrude the tail voluntarily (Table 1). Animal 4 was the only one that was affected by trauma. Before treatment it suffered from urinary incontinence and could not stand its body. After treatment, it was able to take several steps and spontaneous voiding returned. Animal 5 had a bilateral patella dislocation, in

Table 1: Group of animals showing height of the lesion by MRI and data relating to assessments of physical therapy animals, according to Olby scale before and after the surgical procedure and injection of stem cells

| Animal | Age-years | Breed       | Diagnosis            | Lesion       | Moment       | Olby scale                                                                 |
|--------|-----------|-------------|----------------------|--------------|--------------|-----------------------------------------------------------------------------|
| 1      | 6         | Teckel      | Intervertebral disc extrusion | T13-L1       | Pre – treatment | 0 Absence of movement of the hind limbs without feeling deep pain.          |
|        |           |             |                      |              | Pos – treatment | Minimum protraction (movement of a joint) without dislocation of the pelvic limb weight. |
| 2      | 8         | Teckel      | Intervertebral disc extrusion | T13-L3       | Pre – treatment | 0 Absence of movement of the hind limbs without feeling deep pain.          |
|        |           |             |                      |              | Pos – treatment | Minimum protraction (movement of a joint) without dislocation of the pelvic limb weight. |
| 3      | 6         | Lhasa Apso  | Intervertebral disc extrusion | T11-12       | Pre – treatment | 4 Protracão hínld limb with a displacement of less than 10 weight% of the time. |
|        |           |             |                      |              | Pos – treatment | 9 Protracão hínld limb with a displacement of from 10 to 50 weight% of the time. |
| 4      | 2         | Mongrel     | Trauma               | T10-11       | Pre – treatment | 6 Protracão hínld limb with a displacement of less than 10 weight% of the time. |
|        |           |             |                      |              | Pos – treatment | 10 Protracão hínld limb with a displacement of from 10 to 50 weight% of the time. |
| 6      | 8         | Poodle      | Intervertebral disc extrusion | L1-L4        | Pre – treatment | 7 Protracão hínld limb with a displacement of from 10 to 50 weight% of the time. |
|        |           |             |                      |              | Pos – treatment | 10 Protracão hínld limb with a displacement of from 10 to 50 weight% of the time. |
| 7      | 10        | Teckel      | Intervertebral disc extrusion | T11-12       | Pre – treatment | 3 Minimum protraction (movement of a joint) without dislocation of the pelvic limb weight. |
|        |           |             |                      |              | Pos – treatment | 9 Protracão hínld limb with a displacement of from 10 to 50 weight% of the time. |

Figure 2: Evolution of the animals according to the scale of Olby [9].
addition to the spinal lesion, resulting in high muscle contraction that became much better after surgery. However, one knee was not stable and required further surgery during the investigated period, and a final assessment of the Olby score was not possible and the animal was not included within the statistical analysis. The animal 6 showed aggressive behavior and the postoperative evaluation of superficial and deep pain sensation and panniculus reflex were performed with the aid of a forceps. In this case and for Animal 7, a significant improvement in walking and the return of voluntary movement of the tail was observed (Table 1). Additional file 1: Videos S1 and S2 were provided to show the evolution of the one animal treated after 30, 60 and 90 days after treatment.

Discussion
We showed in the current case study that adult dogs with chronic SCI could be successfully treated with the transplantation of foetal canine bone marrow stem cells. Examined 90 days after surgery, all animals had increased locomotor and sensory functions according to the Olby scale, without health complications or decline in the neurologic status. The patients received stem cell treatment, but also underwent physical therapy, so it is hard to determine the exact cause of any improvements. It should be given thought, however, that many patients with chronic SCIs do not recover strength so many time post injury with physical therapy alone [24]. The data suggested that transplanted stem cells may lead to a functional recovery of SCI, similar to what was reported for acute lesions in dogs [22]. Also, it was reported in dogs, indicated by higher values of the Texas SCI scale after the transplantation of bone marrow stem cells [23]. In others studies recovery of motor and sensory functions 53 days after initial cell transplantation and the ability to stand after 79 days was reported [25]. The observed improvements in dogs are similar to reports on humans with acute SCI [21]. In addition to the applications for SCI regeneration, bone marrow stem cells have broad therapeutic significance. They have been used as donor cells for haematologic diseases in animal models [26], or to treat cardiovascular diseases and cancer in humans [27]. Also, transplanted bone marrow stem cells improve neurologic disorders in models of central nervous system injury by generating neural or myelin-producing cells [28]. Donor cells from male humans have been shown to rebuild neurons in the brains of female recipients [29]. In general, bone marrow stem cells differentiate into a variety of non-hematopoietic cell lineages, e.g. muscle, skin, liver, lung, cardiac myocytes, endothelial and neuronal cells [30,31]. For stem cell therapy, the use of bone marrow-derived stem cells cause minor problems in regard to immunological rejections, graft-versus-host reactions or carcinogenesis compared to other embryonic or adult stem cells [32,33].

Conclusion
In addition to the former therapies, our case study indicates that foetal bone marrow stem cells represent a promising treatment for chronic SCI in canine patients, however others stem cell sources can also be considered to use for treatment, suggesting that clinical trials with more animals are needed.

Methods
Selection of animals and preoperative procedures
Seven dogs with chronic SCI, from different private veterinary clinics in São Paulo, were used (Table 1). The selected animals presented with paralysis, a lack of conscious proprioception, the absence of deep pain perception with presence of the withdrawal reflex in the hind limbs, assessed according to the Olby scale (see below), and diagnosis of spinal cord compression of thoracolumbar intervertebral discs between segments T10-L3 after magnetic resonance imaging (MRI). The absence of deep pain perception for more than 60 days was used as a diagnostic tool for chronic injury. The animals did not show any positive response to former treatments such as medication, physiotherapy or acupuncture. Conventional examinations (blood count, biochemistry, urinalysis, stool analysis and electrocardiogram) were performed, indicating normal values. MRI (Vet-MR Grande-Esaot) was performed to determine the precise location of the lesion. The owners signed an informed consent, making them aware of all procedures and risks associated with the surgery and cell transplantation. The project was approved by the Ethical committee of the University of Sao Paulo (Nr. 2063/2010).

Surgery and cell transplantation
Prior to anesthesia, the animals were injected with 0.05 mg/kg acepram (acepromazine1) associated with 4 mg/kg dolosal (Meperidine2), both intramuscularly. After 15 minutes anaesthesia was induced with 5 mg/kg propofol (Propovan3 - Cristália) intravenously and maintained by inhalational anaesthesia with isoflurane (Isoforine4 - Cristália). During surgery the animals underwent dorsolateral hemilaminectomy in the injured spinal cord. The incision was made lateral to the midline and dorsal to the lesion. After incision of the skin and muscles, the fascia was incised lumbodorsal alongside the corresponding spinous processes of the vertebrae, and, with the aid of a periosteum elevator, the spinal processes were isolated. By exposing the dorsal surface of the articular process, a window was opened to provide access to the spinal canal. After which, the medullary mass was compressed and damaged tissue was removed. After durotomy, intramedullary injection of 1×10⁶ of stem cells derived from foetal canine bone marrow [31] was completed cranial to the injury, at the focal point of the lesion, and caudal to the lesion. The postoperative therapy consisted of 30 mg/kg

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Kefazol* (cefazolin sodium) 30 mg/kg, 0.1 mg/kg meloxicam (Generic Biosintética) and 2 mg/kg Tramal* (tramadol hydrochloride) for seven days.

Postoperative examinations
Seven days after surgery, the animals were trained on aquatic and dry treadmills. To observe possible improvement, each animal was filmed in different positions 30 and 60 days after surgery (data not shown) as well as 90 days. The dogs were scored using the Olby scale by 14 points by three veterinary physiotherapists not involved in the experiments. Dogs were considered to be able to support their own weight when they maintained their body for at least four footsteps. The reflexes of superficial and deep pain were evaluated with the help of tweezers. Statistical analyses were performed using the PRISM 4.0 statistical software package (GraphPad, Inc., San Diego, CA). Significance for all analyses was set at P-values of 0.05.

Additional file

Additional file 1: Videos S1 and S2. Progressive response of the animal during the treatment between the analysis performed.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
CAPS and MAM conceived and designed the study, and wrote the manuscript; MNR and RZB performed the experiments and data analysis; AMM participated in manuscript preparation. All authors read and approved the final version of the manuscript.

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References
1. Webb AA, Ngan S, Fowler JD: Spinal cord injury I: a synopsis of the basic science. Can Vet J 2010, 51:485–492.
2. Mathé AJ, Tator CH: Advances in stem cell therapy for spinal cord injury. J Clin Invest 2012, 122:3824–3834.
3. Antonic A, Sena ES, Lees JS, Willis TE, Skeers P, Batchelor PE, Macleod MR, Howells DW: Stem cell transplantation in traumatic spinal cord injury: a systematic review and meta-analysis of animal studies. PLoS Biol 2013, 11:e1001738. doi:10.1371/journal.pbio.1001738.
4. McKee WM: Spinal trauma in dogs and cats: a review of 51 cases. Vet Rec 1990, 126:285–289.
5. Bruce CW, Brisson BA, Gyselinck K: Spinal fracture and luxation in dogs and cats; a retrospective evaluation of 95 cases. Vet Comp Orthop Traumatol 2008, 21:280–289.
6. Olson L: Medicine: clearing a path for nerve growth. Nature 2002, 416:589–590.
7. Jeffery ND, Lekatos A, Franklin RM: Autologous offactory glial cell transplantation is reliable and safe in naturally occurring canine spinal cord injury. J Neurotrauma 2005, 22:1282–1293.
8. Penning V, Platt SR, Dennis R, Capello R, Adams V: Association of spinal cord compression seen on magnetic resonance imaging with clinical outcome in 67 dogs with thoracolumbar intervertebral disc extrusion. J Small Anim Pract 2006, 47:644–655.
9. Olby NJ, Risio LD, Muñuera KR, Worsar MA, Skees TM, Sharp NJH, Keene BW: Development of a functional scoring system in dogs with acute spinal cord injuries. Am J Vet Res 2001, 62:1624–1628.
10. Levine GJ, Levine JM, Budke CM, Kerwin SC, Au J, Vinayak A, Hettlich BF, Slater MR: Description and repeatability of a newly developed spinal cord injury scale for dogs. Vet Prev Med 2009, 89:121–127.
11. Srugo I, Arocch L, Christopher MM, Choi O, Goradiki L, Bidoab-Abram T, Shmir MH: Signs and outcome in acute nonambulatory thoracolumbar disc disease in dogs. J Vet Intern Med 2011, 25:846–855.
12. Henke D, Vandervele M, Doher MG, Stocki M, Forterre F: Correlations between severity of clinical signs and histopathological changes in 60 dogs with spinal cord injury associated with acute thoracolumbar intervertebral disc disease. Vet J 2013, 198:70–75.
13. Mackay-Sim A, Feron F, Cochrane J, Basingthwaigte L, Bayliss C, Davies W, Fronck P, Gray C, Kerr G, Licina P, Nowitzke A, Perry C, Silburn PAS, Ushrath S, Geragthy T: Autologous olfactory enseathing cell implantation in human paraplegia: a 3-year clinical trial. Brain 2008, 131:2376–2386.
14. Kumar AA, Kumar SR, Narayanan R, Anil K, Bakkash M: Autologous bone marrow derived mesenchymal cell therapy for spinal cord injury: a phase II clinical safety and primary efficacy data. Exp Clin Transplant 2009, 7:241–248.
15. Attar A, Ayten M, Ozdemir M, Ogzencli E, Bozkurt M, Kaptanoglu E, Beksac M, Kanpolat Y: An attempt to treat patients who have injured spinal cords with intradiscal implantation of concentrated autologous bone marrow cells. Cytotherapy, 2011, 13:354–60.
16. Park HC, Shim VS, Yoon SH, Park SR, Choi BH, Park HS: Treatment of complete spinal cord injury patients by autologous bone marrow cell transplantation and administration of granulocyte-macrophage colony stimulating factor, Tissue Eng 2005, 11:913–922.
17. Osaka M, Hommou O, Murakami T, Nonaka T, Houkin K, Hamada H, Kocis JD: Intravenous administration of mesenchymal stem cells derived from bone marrow after contusive spinal cord injury improves functional outcome. Brain Res, 2010, 1433:226–235.
18. Hawryluk GW, Mothe A, Wang J, Wang S, Tator C, Feihlings MG: An in vivo characterization of trophic factor production following neural precursor cell or bone marrow stromal cell transplantation for spinal cord injury. Stem Cells Dev, 2012, 21:2222–238.
19. Eun-Sun K, Kee-Yong H, Young-Hoon K: Fate of transplanted bone marrow derived mesenchymal stem cells following spinal cord injury in rats by transplantation routes. J Korean Med Sci 2012, 27:586–593.
20. Deng YB, Yuan QT, Liu XG, Liu XL, Liu Y, Liu ZG, Zhang C: Functional recovery after rhesus monkey spinal cord injury by transplantation of bone marrow mesenchymal-stem derived neurons. Chin Med J 2005, 118:1533–1541.
21. Pal R, Venkataramak NR, Bansal A, Balasaju S, Jan M, Chandra R, Dixit A, Rauthan A, Murgod U, Totey S: Ex vivo expanded autologous bone marrow-derived mesenchymal stromal cells in human spinal cord injury/paraplegia: a pilot clinical study. Cytotherapy 2009, 11:897–911.
22. Jung DI, Ha J, Kang BT, Kim JW, Quan FS, Lee JH, Woo EJ, Park HM: A comparison of autologous and allogeneic bone marrow derived mesenchymal stem cell transplantation in canine spinal cord injury. J Neurosci Soc 2009, 285:67–77.
23. Nishida H, Nakayama M, Tanaka H, Kitamura M, Hatoya S, Sugiuara K, Suzuki Y, Ide C, Inaba T: Evaluation of transplantation of autologous bone marrow stromal cells into the cerebrospinal fluid for treatment of chronic spinal cord injury in dogs. Am J Vet Res 2011, 72:1118–1123.
24. Fawcett JW, Curt A, Steeven JD: Guidelines for the conduct of clinical trials for spinal cord injury as developed by the ICPP panel: spontaneous recovery after spinal cord injury and statistical power needed for therapeutic clinical trials. Spinal Cord 2007, 45:190–205.
25. Williams JB, Prabakaran R, Ayyappan S, Pushtirnaghi H, Rao D, Thamarakannan P, Dedepsiya VD, Xuruba S, Yoshikawa H, Mori Y, Preethy S, Abraham SJK: Functional recovery of spinal cord injury following application of intrathecal bone marrow mononuclear cells embedded in polymer scaffold - two year follow-up in a canine. J Stem Cell Res Ther 2011, 1:110.
26. Kroger N, Zabelina T, Renges H, Kruger W, Kordes U, Rischevski J, Schum J, Horstmann M, Ayuk F, Erttman R, Kabich B, Zander AR: Long-term follow-up of allogeneic stem cell transplantation in patients with severe aplastic anemia after conditioning with cyclophosphamide plus antithymocyte globulin. Ann Hematol 2002, 81:627–631.

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27. Park JH, Kim DY, Sung IY, Choi GH, Jeon MH, Kim KK, Jeon SR: Long-term results of spinal cord injury therapy using mesenchymal stem cells derived from bone marrow in humans. Neurosurgery 2012, 70:1238–1247.

28. Deda H, Inci MC, Kürekçi AE, Kayihan K, Özgün E, Ustünsoy GE, Kocabay S: Treatment of chronic spinal cord injured patients with autologous bone marrow-derived hematopoietic stem cell transplantation: 1-year follow-up. Cytotherapy 2008, 10:565–574.

29. Mezey E, Key S, Vogelsang G, Szalayova I, Lange GD, Crain B: Transplanted bone marrow generates new neurons in human brains. Proc Natl Acad Sci USA 2003, 100:1364–1369.

30. Krause DS: Plasticity of marrow-derived stem cells. Gene Ther 2002, 9:754–758.

31. Wenceslau CV, Miglino MA, Martinis DS, Ambrosio CE, Lizer NF, Pignatari GC, Kerks I: Mesenchymal progenitor cells from canine fetal tissues: yolk sac, liver, and bone marrow. Tissue Eng 2011, 17:2165–2176.

32. Strom TB, Field LJ, Ruediger M: Allogenic stem cells, clinical transplantation and the origins of regenerative medicine. Curr Opin Immunol 2002, 14:601–605.

33. Erdö F, Buhrlé C, Blank J, Hoehn M, Xia Y, Fleischmann B, Focking M, Küstermann E, Kolossov E, Hescheler J, Hossmann KA, Trapp T: Host-dependent tumorigenesis of embryonic stem cell transplantation in experimental stroke. J Cereb Blood Flow Metab 2003, 23:780–785.

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