Original Article

Synthesis of N-arylidene-2-(2-Phenoxyphenyl) Acetohydrazides as Anti-Inflammatory Agents

Maral Shekarchi\textsuperscript{a}, Latifeh Navidpour\textsuperscript{b}, Afshin Rajabi Khorami\textsuperscript{c}, Mahtab Shekarchi\textsuperscript{d}, Alireza Partoazar\textsuperscript{e}, Hamed Shafaroodi\textsuperscript{f}, Narges Rahmani\textsuperscript{g}, Abbas Shafiee\textsuperscript{h} and Maryam Shekarchi\textsuperscript{i}

\textsuperscript{a} Department of Chemistry, School of science, Karaj Branch, Islamic Azad University, Karaj, Iran. \textsuperscript{b}Department of Medicinal Chemistry, Faculty of Pharmacy and Pharmaceutical Sciences Research Center, Tehran University of Medical Sciences, Tehran 14176, Iran. \textsuperscript{c}Department of Pharmacology, Faculty of Medicine, Tehran University of Medical Sciences, Tehran 14174, Iran. \textsuperscript{d}Department of Pharmacology, Tehran medical unit, Islamic azad University, Tehran, Iran. \textsuperscript{e}Department of Research and Development, Food and Drug Laboratory Research Center, Tehran, Iran.

Abstract

Diclofenac sodium has been used for its anti-inflammatory actions for about 28 years, but since all the non-steroidal anti-inflammatory drugs (NSAIDs) suffer from the lethal gastrointestinal (GI) toxicities, diclofenac sodium is not an exception. The free –COOH group is thought to be responsible for the GI toxicity associated with all traditional NSAIDs. In the present research, the main motto was to develop new chemical entities as potential anti-inflammatory agents with no GI toxicities. A new type of 2-(2-Phenoxyphenyl) acetohydrazide possessing N-arylidene substituents, was synthesized for evaluation as anti-inflammatory agents. The starting material 2-(2-Phenoxyphenyl) acetohydrazide was synthesized from 2-phenoxybenzoic acid in several steps according to the previous published method. Various substituted arylidene-2-phenoxyacetic acid hydrazide derivatives were synthesized by the reaction of hydrazide 17 with selected aldehydes and screened for their potential anti-inflammatory activity. The structure of synthesized compounds was confirmed by different nuclear magnetic resonance technique, Fourier transform infrared spectroscopy (FTIR) and Mass-spectrometry data format. Qualitative structure-activity relationship data, acquired using the carrageenan-induced rat paw edema assay, showed that this group of arylidene-2-phenoxybenzoic acid hydrazides exhibit anti-inflammatory activity with significant reduction of rat paw edema (17-58% reduction in inflammation at different time intervals) in comparison with control group and a moderate to good activity range in comparison with diclofenac as the reference drug. Compounds 9a, 9d and 9e exhibited the most prominent and consistent anti-inflammatory activity. The compound, N-(4-Chlorobenzylidene)-2-(2-phenoxyphenyl) acetohydrazide (9d), exhibited the most in-vivo activity (32-58% reduction in inflammation) compared to the reference drug diclofenac (35-74% reduction in inflammation) in a carrageenan induced rat paw-edema assay.

Keywords: N-arylidene-2-(2-phenoxyphenyl) acetohydrazide derivatives; Anti-inflammatory activity; N-Acylhydrazones; Non-steroidal anti-inflammatory drugs.

* Corresponding author: Dr. M. Shekarchi
E-mail: Shekarchim@yahoo.com
Introduction

Non-steroidal anti-inflammatory drugs (NSAIDs) are the most prescribed drugs in the treatment of pain and inflammation, particularly for different types of arthritis (1-3). The pharmacological activity of NSAIDs exerts their anti-inflammatory activity. The inhibition of cyclooxygenase-derived (COX) prostaglandin synthesis is also responsible for the gastrointestinal (GI), renal, and hepatic side effects observed in patients undergoing long term treatment (4-9). It was discovered that cyclooxygenase exists in two isoforms, COX-1 and COX-2, which are regulated and expressed differently. COX-1 provides cytoprotection in the gastrointestinal tract (GIT), whereas inducible COX-2 selectively mediates inflammatory signals (10-11). Since most of the currently available NSAIDs in the market show greater selectivity for COX-1 compared to COX-2, chronic use of NSAIDs, including diclofenac, may elicit appreciable GI irritation, bleeding and ulceration (12). The GI damage from NSAID is generally attributed to two factors such as local irritation by the direct contact of carboxylic acid (–COOH) moiety of NSAID with GI mucosal cells (topical effect) and decreased tissue prostaglandin production in tissues which undermines the physiological role of prostaglandins in maintaining GI health and homoeostasis (13). Therefore, the discovery of new safer anti-inflammatory drugs represents a challenging goal for such a research area. Synthetic approaches based upon the chemical modification of NSAIDs have been taken with the aim of improving safety profile and, in turn, the therapeutic window of these NSAIDs.

Diclofenac, Mefenamic acid and Indomethacin are three traditional NSAIDs belonging to the class of fenamates (Figure 1). This class includes derivatives of N-phenyl (or heteroarylyl) anthranilic acid that are employed for their analgesic, anti-inflammatory and antipyretic properties. Unlike most of the NSAIDs, the fenamates also appear to compete with prostaglandins for binding at the prostaglandin receptor site and, thus, potentially antagonizing the physiopathological effects of prostaglandins that have already been formed. Fenamates are still endowed with most of the adverse effects induced by NSAIDs, particularly gastrointestinal (GI) bleeding, ulceration, and perforation. Several studies have described the derivatization of the carboxylate function of representative NSAIDs with amide or N-acylarylhydrazone having less acidic amide hydrogen resulted in an increased anti-inflammatory activity with reduced ulcerogenicity (14-17) (Figure 2). Some evidences suggest that the hydrazine moiety possesses a pharmacophoric character for the inhibition of cyclooxygenase (15). N-Acylhydrazones (NAH) have been widely described as potent anti-inflammatory, anti-nociceptive, and anti-platelet compounds, due to their ability to mimic the bis-allylic moiety of unsaturated fatty acids and amides, for example, arachidonic acid (AA), precursor of the eicosanoid biosynthesis, involved in the endocannabinoid system. This can be rationalized through the relative acidity in amide hydrogen of NAH group as well as its capacity of stabilizing the free radicals (14). Considering the above results and as a part of our ongoing program to design analgesic and anti-inflammatory agents (17), herein, we describe the design, synthesis and biological evaluation of a novel diverse group in N-acylarylhydrazone derivatives of 2-phenoxybenzoic acid 9a-g with different substituents on the terminal phenyl ring (Figure 2).

Experimental

Chemistry

Melting points were determined with a Reichert-Jung hot-stage microscope (Reichert-Jung, Germany) and are uncorrected. Infrared spectra were recorded on a Nicolet Magna 550-FT spectrometer (Nicolet, Madison, WI, USA). 1H-NMR (500 MHz) spectra were measured through a Bruker FT-500 MHz spectrometer (Bruker Bio Sciences, USA) in CDCl3 or DMSO-d6 with tetramethylsilane (TMS) as the internal standard, where J-values (coupling constant) are estimated in Hertz. Spin multiples are given as s (singlet), d (doublet), dd (doublet of doublets), t (triplet), q (quartet), m (multiplet), and b (broad). Mass spectra were obtained, using
a Finnigan Mat triple-stage quadrupole (TSQ 70) spectrometer (Thermo-Electron, Germany). All solvents and reagents were purchased from the Fluka (Aldrich, UK), or Merck Chemical Company (Merck, Germany). Male Wistar rats used in the anti-inflammatory screens were purchased from Pasteur Institute (Karaj, Iran) and experiments were carried out using protocols approved by the ethics committee of Tehran University of Medical Sciences. Compounds 10-16 have been synthesized according to the published methods in literatures.

2-(2-Phenoxyphenyl) acetohydrazide (17)

A solution of compound 16 (3g, 12 mmol) and hydrazine hydrate (3g, 60 mmol) in 20 mL methanol was stirred at room temperature for 5 h. The reaction mixture poured immediately into a mixture of crushed ice water, gave a crude precipitate collected by filtration and recrystallized in ethanol/water to give 2.6 g (94%) of hydrazide derivative 17, as a light yellow amorphous solid, m.p.: 62-64 °C; 1H-NMR (500 MHz, CDCl3) 7.21-7.24 (m, 1H, aromatic), 7.1 (m, 2H, aromatic), 6.94 (d, J = 7.5 Hz, 2H, aromatic), 6.87 (d, J = 7.5 Hz, 1H, aromatic), 3.76 (bs, 2H, NH2), 3.57 (s, 2H, CH 2); IR (KBr): Cm -1 3308 (N-H), 3037 (C-H, aromatic), 1654 (C=O), 1236 (C-O), 1096 (C-N); MS: m/z (%) 242 (M+, 42), 210 (68), 183 (100), 165 (32), 155 (54), 77 (45), 51 (36).

General procedure for preparing N-arylidene-2-(2-phenoxyphenyl) acetoxydrazides (9a-g)

The synthesis of final hydrazones 9a-g was carried out by the reaction of hydrazide 17 and the corresponding aldehydes in ethanol. A mixture of hydrazide 17 (1.5 mmol) and the corresponding aldehyde (1.5 mmol) in ethanol (10 mL) was stirred at room temperature for 1 to 24 h in the presence of hydrochloric acid (2 drops) as the catalyst. The completion of the reaction was monitored by thin layer chromatography (TLC). The reaction mixture was concentrated under reduced pressure and was neutralized with a 10% aqueous solution of sodium bicarbonate. The resulting precipitate was filtered, washed with water, and crystallized using a suitable solvent to give 9.

N-Benzylidene-2-(2-phenoxyphenyl) acetohydrazide (9a)

M.p.: 169-171°C (Ethanol/Water); as a mixture of diastereomers in ratio of 2.5:9:1; 1H-NMR (500 MHz, CDCl3): δ 9.01 (bs, 0.27H, NH), 8.77 (bs, 0.28H, NH), 7.80 (s, 0.28H, =CH), 7.72-7.63 (m, 1.28H, =CH2), 7.62-7.55 (m, 1.44H, Hδ), 7.49 (d, J = 7.25 Hz, 0.28H, Hδ), 7.45-7.32 (m, 4.28H, Hδ, Hδ, Hδ), 7.27-7.18 (m, 2.44H, Hβ, Hβ), 7.17-7.08 (m, 1.44H, Hβ, Hβ), 7.05-6.88 (m, 3.56H, Hα, Hα, Hα), 4.17 (s, 1.44H, CH3), 3.74 (s, 0.56H, CH2); MS: m/z (%) 330 (M +, 16), 240 (10), 227 (5), 210 (8), 183 (10), 165 (28), 155 (38), 149 (12), 134 (100), 119 (23), 104 (10), 89 (23), 77 (41), 68 (32), 55 (22).

N-(4-Methyl-benzylidene)-2-(2-phenoxyphenyl) acetoxydrazide (9b)

M.p.: 196-198°C (Ethanol/Water); as a mixture of diastereomers in ratio of 2.03:1; 1H-NMR (500 MHz, CDCl3): δ 9.40 (bs, 0.68H,
N-(4-Methoxybenzylidene)-2-(2-phenoxyphenyl) acetohydrazide (9c)

M.p.: 200-202°C (Ethanol/Water); as a mixture of diastereomers in ratio of 2.03:1; 1H-NMR (500 MHz, CDCl3); δ 9.43 (bs, 0.7H, NH), 8.74 (bs, 0.33H, NH), 7.71 (s, 0.33H, =CH), 7.63 (s, 0.67H, =CH), 7.60 (d, J = 8.0 Hz, 0.66H, Hg), 7.53 (d, J = 8.0 Hz, 1.34H, Hf), 7.48 (d, J = 7.10 Hz, 0.33H, Hh), 7.40 (d, J = 7.10 Hz, 0.67H, Hg), 7.35 (t, J = 7.15 Hz, 0.66H, Hf), 7.28-7.19 (m, 2.34H, Hg, Hf), 7.17-7.07 (m, 1.34H, Hf), 7.05-6.85 (m, 5.66H, Hg, Hf, Hf, Hg, Hf), 4.16 (s, 1.34H, CH2), 3.84 (s, 2.01H, OCH3), 3.82 (s, 0.99H, OCH3), 3.72 (s, 0.66H, CH2); IR (KBr): Cm⁻¹ 3262 (N-H), 3083 (C-H, aromatic), 1685 (C=O), 1242 (C-O); MS: m/z (%) 344 (M⁺, 5), 279 (10), 237 (3), 228 (3), 196 (3), 181 (50), 165 (20), 149 (70), 134 (65), 127 (12), 118 (20), 104 (30), 91 (32), 77 (60), 69 (62), 57 (97), 41 (100).

N-(4-Chlorobenzylidene)-2-(2-phenoxyphenyl) acetohydrazide (9d)

M.p.: 215-218°C (Ethanol/Water); as a mixture of diastereomers in ratio of 2.33:1; 1H-NMR (500 MHz, CDCl3); δ 9.34 (bs, 0.7H, NH), 8.82 (bs, 0.3H, NH), 7.79 (s, 0.3H, =CH), 7.63 (s, 0.7H, =CH), 7.59 (d, J = 8.1 Hz, 0.6H, Hg), 7.51 (d, J = 8.1 Hz, 1.4H, Hf), 7.48 (d, J = 7.25 Hz, 0.3H, Hg), 7.39 (d, J = 7.25 Hz, 0.7H, Hf), 7.38-7.30 (m, 2.3H, Hg, Hf), 7.29-7.20 (m, 2.4H, Hg, Hf), 7.19-7.07 (m, 1.4H, Hg, Hf), 7.05-6.96 (m, 1.3H, Hg, Hf, Hg, Hf), 6.95-6.85 (m, 2.3H, Hg, Hf), 4.15 (s, 1.4H, CH2), 3.73 (s, 0.6H, CH2); IR (KBr): Cm⁻¹ 3246 (N-H), 3085 (C-H, aromatic), 1681 (C=O), 1246 (C-O); MS: m/z (%) 366 (M⁺, 10), 330 (3), 282 (20), 257 (5), 218 (20), 194 (10), 183 (55), 165 (21), 149 (84), 134 (100), 121 (14), 105 (16), 97 (30), 81 (41), 69 (89), 57 (84), 43 (88).

N-(4-Chlorobenzylidene)-2-(2-phenoxyphenyl) acetohydrazide (9d)

M.p.: 215-218°C (Ethanol/Water); as a mixture of diastereomers in ratio of 2.33:1; 1H-NMR (500 MHz, CDCl3); δ 9.34 (bs, 0.7H, NH), 8.82 (bs, 0.3H, NH), 7.79 (s, 0.3H, =CH), 7.63 (s, 0.7H, =CH), 7.59 (d, J = 8.1 Hz, 0.6H, Hg), 7.51 (d, J = 8.1 Hz, 1.4H, Hf), 7.48 (d, J = 7.25 Hz, 0.3H, Hg), 7.39 (d, J = 7.25 Hz, 0.7H, Hf), 7.38-7.30 (m, 2.3H, Hg, Hf), 7.29-7.20 (m, 2.4H, Hg, Hf), 7.19-7.07 (m, 1.4H, Hg, Hf), 7.05-6.96 (m, 1.3H, Hg, Hf, Hg, Hf), 6.95-6.85 (m, 2.3H, Hg, Hf), 4.15 (s, 1.4H, CH2), 3.73 (s, 0.6H, CH2); IR (KBr): Cm⁻¹ 3246 (N-H), 3085 (C-H, aromatic), 1681 (C=O), 1246 (C-O); MS: m/z (%) 366 (M⁺, 10), 330 (3), 282 (20), 257 (5), 218 (20), 194 (10), 183 (55), 165 (21), 149 (84), 134 (100), 121 (14), 105 (16), 97 (30), 81 (41), 69 (89), 57 (84), 43 (88).

N-(4-Chlorobenzylidene)-2-(2-phenoxyphenyl) acetohydrazide (9d)

M.p.: 215-218°C (Ethanol/Water); as a mixture of diastereomers in ratio of 2.33:1; 1H-NMR (500 MHz, CDCl3); δ 9.34 (bs, 0.7H, NH), 8.82 (bs, 0.3H, NH), 7.79 (s, 0.3H, =CH), 7.63 (s, 0.7H, =CH), 7.59 (d, J = 8.1 Hz, 0.6H, Hg), 7.51 (d, J = 8.1 Hz, 1.4H, Hf), 7.48 (d, J = 7.25 Hz, 0.3H, Hg), 7.39 (d, J = 7.25 Hz, 0.7H, Hf), 7.38-7.30 (m, 2.3H, Hg, Hf), 7.29-7.20 (m, 2.4H, Hg, Hf), 7.19-7.07 (m, 1.4H, Hg, Hf), 7.05-6.96 (m, 1.3H, Hg, Hf, Hg, Hf), 6.95-6.85 (m, 2.3H, Hg, Hf), 4.15 (s, 1.4H, CH2), 3.73 (s, 0.6H, CH2); IR (KBr): Cm⁻¹ 3246 (N-H), 3085 (C-H, aromatic), 1681 (C=O), 1246 (C-O); MS: m/z (%) 366 (M⁺, 10), 330 (3), 282 (20), 257 (5), 218 (20), 194 (10), 183 (55), 165 (21), 149 (84), 134 (100), 121 (14), 105 (16), 97 (30), 81 (41), 69 (89), 57 (84), 43 (88).
### Figure 3. Reagents and conditions

(a) thionyl chloride, abs. EtOH, reflux, 4 h; 
(b) LiAlH₄, THF, stir, 24 h; 
(c) thionyl chloride, benzene/ pyridine, reflux, 2 h; 
(d) NaCN, DMSO, Stir, 2 h; 
(e) KOH, n- butanol, reflux, 2 h; 
(f) thionyl chloride, abs. EtOH, reflux, 4 h; 
(g) N₃, H₂O, abs. MeOH, Stir, 3 h; 
(h) aldehydes, EtOH, HCl (2 drops), stir, 1-24 h.

---

**N-(4-Fluorobenzylidene)-2-(2-phenoxyphenyl) acetohydrazide (9e)**

M.p.: 210-215°C (Ethanol/Water); as a mixture of diastereomers in ratio of 2.46:1; 
'H-NMR (500 MHz, CDCl₃): δ 9.24 (bs, 0.29H, NH), 8.80 (bs, 0.71H, NH), 7.80 (s, 0.28H, =CH), 7.70-7.60 (m, 1.29H, =CH, Hₖ), 7.57 (dd, J = 5.5, 7.5 Hz, 1.72H, Hₗ), 7.48 (d, J = 7.0 Hz, 0.29H, Hₖ), 7.40 (d, J = 7.0 Hz, 0.71H, Hₗ), 7.36 (t, J = 7.15 Hz, 0.58H, Hₖ), 7.26-7.18 (m, 2.42H, Hₗ, Hₖ), 7.15-7.09 (m, 1.42H, Hₗ, Hₖ), 7.08-6.97 (m, 3.71H, Hₗ, Hₖ, Hₗ, Hₖ), 6.96-6.85 (m, 2.29H, Hₗ, Hₖ), 4.16 (s, 1.42H, CH₂), 3.73 (s, 0.58H, CH₂); IR (KBr): Cm⁻¹ 3458 (N-H), 3222 (C-H, aromatic), 1677 (C=O), 1237 (C-O), 1096 (C-N); MS: m/z (%) 348 (M⁺, 14), 256 (7), 227 (5), 210 (5), 183 (100), 165 (31), 155 (42), 149 (70), 134 (65), 134 (80), 122 (10), 107 (14), 95 (17), 77 (28), 69 (40).

**N-(2,4-Dimethoxybenzylidene)-2-(2-phenoxyphenyl) acetohydrazide (9f)**

M.p.: 250-252°C (Ethanol/Water); as a mixture of diastereomers in ratio of 1.4:1; 
'H-NMR (500 MHz, CDCl₃): δ 8.00, 7.99 (bs, 1H, NH), 7.75, 7.73 (s, 1H, =CH), 7.46 (d, J = 7.5 Hz, 0.4H, Hₙ), 7.39 (d, J = 7.5 Hz, 0.6H, Hₚ), 7.33 (t, J = 7.8 Hz, 0.8H, Hₗ), 7.27-7.18 (m, 2.2H, Hₙ, Hₚ), 7.16-7.07 (m, 1.2H, Hₗ, Hₖ), 7.05-6.95 (m, 3.8H, Hₗ, Hₖ, Hₙ, Hₚ), 6.92-6.87 (m, 1H, Hₖ), 6.60-6.52 (m, 0.4H, Hₙ), 6.48 (dd, J = 2.1, 8.7 Hz, Hₖ), 6.46-6.44 (d, J = 2.1 Hz, 0.4H, Hₙ), 6.42 (d, J = 2.1 Hz, 0.6H, Hₖ), 4.14 (s, 1.2H, CH₃), 3.87, 3.88, 3.84, 3.83, 3.82, 3.81 (s, 6H, OCH₃); IR (KBr): Cm⁻¹ 1670 (C=O), 1207 (C-O), 1031 (C-N); MS: m/z (%) 390 (M⁺, 10), 359 (7), 227 (5), 210 (5), 183 (100), 164 (31), 150 (70), 137 (65), 122 (10), 106 (14), 95 (17), 77 (28), 69 (40).
N-(3,4,5-Trimethoxybenzylidene)-2-(2-phenoxophenyl) acetohydrazide (9g)

M.p.: 258-260°C (Ethanol/Water); as a mixture of diastereomers in ratio of 1.69:1; 1H-NMR (500 MHz, CDCl3): δ 9.55 (bs, 0.63H, NH), 8.75 (bs, 0.37H, =CH), 7.61 (s, 0.63H, =CH), 7.47 (d, J = 7.5 Hz, 0.74H, Hg), 7.40 (d, J = 7.5 Hz, 1.26H, Hg), 7.36 (t, J = 7.9 Hz, 0.74H, Hg), 7.29-7.20 (m, 2.26H, Hg, Hg), 7.15-7.08 (m, 1.26H, Hg, Hg), 7.04-6.95 (m, 1.37H, Ha, Hg, Hg), 6.95-6.87 (m, Hg, Hg), 6.88 (s, 0.74H, Hg), 6.82 (s, 1.26H, Hg), 4.15 (s, 1.26H, CHg), 3.87, 3.86 (s, 5H, OCHg), 3.85 (s, 4H, OCHg), 3.72 (s, 0.74H, CHg); IR (KBr): Cm⁻¹ 3243 (N-H), 3222 (C-H, aromatic), 1667 (C=O), 1237 (C-O), 1065 (C-N); MS: m/z (%) 420 (M+, 20%), 389 (10%), 226 (5%), 212 (5%), 184 (100), 170 (31%), 152 (42%), 149 (70%), 136 (65%), 121 (10%), 107 (14%), 90 (17%), 77 (28%), 69 (40%)

Anti-inflammatory activity

The anti-inflammatory activity was determined in-vivo using the carrageenan-induced rat paw edema test. Edema was induced in the right hind paw of all rats by subcutaneous injection of 0.1 mL of 1% (w/v) carrageenan (Sigma-Aldrich, Dorset, UK) in saline into their footpads 0.5 h after the intra-peritoneal (IP) administration of compounds (18). The paw thickness was measured from the ventral to the dorsal surfaces using a dial caliper immediately before and then two and four h after the carrageenan injection. The edema was calculated as the thickness variation between thickness of paw before and after carrageenan injection. The anti-inflammatory activity was expressed similar to the inhibition percentage of edema when compared with the control group and was calculated using the following formula:

\[ \text{Inhibition\%} = \left(1 - \frac{T_t}{T_c}\right) \times 100 \]

In this formula, \( T_t \) and \( T_c \) are defined as the thickness variation of test group and control group, respectively.

Statistics

The results are expressed as mean ± SEM of \( n \) animals per group. The data were statistically analyzed by one-way analysis of variance (ANOVA) followed by Tukey multi comparison test.

Differences with \( p < 0.05 \) among the experimental groups were considered statistically significant.
Results and Discussion

Chemistry

The synthetic reactions leading to the substituted arylidene-2-(2-phenoxyphenyl)-aceto-hydrazides 9a-g are outlined in Figure 3. 2-(2-Phenoxyphenyl) aceto-hydrazide (17) is the key intermediate for the production of the title compounds 9a-g. Esterification of compound 10 using thionyl chloride in ethanol afforded the ethyl esters 11 in 99% yield (19, 20). Next, hydroxyl compound 12 was prepared, in 95% yield by the reduction of compound 11 with lithium aluminum hydride in Tetrahydrofuran/Tetrahydrofolic acid/Tetrahydrofolate (THF) (20). The substitution of hydroxyl to chloride compound 13 was obtained in 97% yield by the reaction of compound 12 with thionyl chloride in benzene/pyridine (21). The treatment of compound 13 with sodium cyanide and subsequent hydrolysis of the synthesized compound 14, gave 2-(2-phenoxyphenyl) acetic acid (15) in 87 and 98% respectively. Reaction of 2-(2-phenoxyphenyl) acetic acid (15) with thionyl chloride in ethanol, furnished the ethyl 2-(2-phenoxyphenyl) acetate (16) in high yield after treatment with hydrazine hydrate in absolute methanol, it resulted in the formation of the required hydrazide 17, the key intermediate of this synthesis route, in 94% yield (20, 22). Finally, the title hydrazone compounds (9a-g) were prepared in great yields by the condensation of hydrazide 17 with different substituted aromatic aldehydes in ethanol, using hydrochloric acid as catalyst (22-25). The structures of various synthesized compounds were assigned on the basis of different chromatographic and spectral studies. The physical data, Fourier transform infrared spectroscopy (FTIR), 1H-NMR and Mass spectral data for synthesized compounds.

Table 2. Anti-inflammatory activities of compound 9a-g.

| Compound | R | Dose (mg/Kg) | Inhibition (%) | at 1 h | at 2 h | at 3 h |
|----------|---|-------------|---------------|-------|-------|-------|
| Control  | - | 10          |               | 35.6 ± 7.1 | 64.6 ± 7.4 | 70.38 ± 4.7 |
| Diclofenac| - | 20          |               | 62 ± 19.1 | 66.3 ± 5.7 | 74.1 ± 3.1 |
| 9a       | H | 10          |               | 47.9 ± 5.3 | 42.6 ± 3.9 | 41.8 ± 8  |
| 9b       | 4-CH₃ | 10   |               | 47.9 ± 5.3 | 45.6 ± 3.1 | 33 ± 4.3 |
| 9c       | 4-OCH₃ | 10  |               | 16.6 ± 4.6 | 26.7 ± 5.1 | 24.2 ± 2.8 |
| 9d       | 4-Cl | 10          |               | 31.2 ± 5.4 | 45.6 ± 3.1 | 43.7 ± 10.6 |
| 9e       | 4-F | 10          |               | 28.1 ± 5.3 | 37.6 ± 5.1 | 26.6 ± 9.4 |
| 9f       | 2,4-(OCH₃)₂ | 10 |               | 34.2 ± 7.2 | 37.3 ± 4.7 | 34 ± 3.3 |
| 9g       | 3,4,5-(OCH₃)₃ | 10 |               | 43 ± 7.4 | 42.5 ± 2.8 | 22.5 ± 38 |

ND: Not determined
are reported in experimental protocols.

The FTIR spectra of Schiff bases exhibited very similar features and showed the expected bands for the characteristic groups which were present in the compounds such as C–H and the C=O stretching vibrations and another specific band for Ar–C–N vibrations. Compounds of 9a-g have C=O stretching bands in the range of 1677-1685. In the proton 1H-NMR spectral data, all protons were seen according to the expected chemical shift and integral values. The aromatic protons appeared as multiplet peaks within the range of 6.9-7.7 ppm and the singlet signals derived from hydrazide 17 (–NH–NH₂) structure appeared at 3.76 ppm. Methylene protons resonated as singlet at 4.1-4.2 ppm. The 1H-NMR spectra of compounds 9a-g displayed singlet due to –NH– groups around 8.1, 9.01 and 9.4 ppm (probably due to their ability to get exchanged with D₂O) while each signal showed integration for one proton. For the compounds 9a-g, the signals which belonged to benzyldiene group were observed at aromatic region, while the signals of –NH₁NH₂ disappeared indicating the functionalization of hydrazide to hydrazone with substituted aromatic aldehydes.

The next step of this work was to determine the relative configuration of the imine double bond of arylidine benzoic acid derivatives 9a-g, in order to assure the stereoisomeric ratio essential for the complete understanding of the biological effect. The careful analysis of the 1H-NMR for 9a-g, enabled us to detect the presence of two singlet signals related to the imine hydrogen, which was attributed to respective (E) and (Z)-diastereomers. The existent ratio between the two diastereomers could be defined from the relative integration of imine-attached hydrogen in the corresponding 1H- NMR spectra (Table 1).

Anti-inflammatory activity

In-vivo pharmacological evaluation of 9a-g was carried out to assess their potential anti-inflammatory activity. Qualitative structure-activity relationship data acquired using the carrageenan-induced rat paw edema assay (18), showed that this group of arylidine-2-phenoxynicotinic acid hydrazides exhibit the anti-inflammatory activity with significant reduction of rat paw edema (17-58% reduction in inflammation at different time intervals) in comparison with the control group and a moderate to good activity range in comparison with diclofenac as the reference drug (Table 2). As shown in Table 2, the presence of electron-withdrawing substituents (9d-e) produced compounds with comparable activity to unsubstituted derivative (9a), while electron-donating (9b-c) substituted compounds (9b-c) showed to be less active in reduction of inflammation. Interestingly, most of the compounds showed better activity in 1 or 2 h rather than 3 h showing the faster onset of action compared to diclofenac which was possibly resulted from their more lipophilic structure. Poly-substitution in 9f-g resulted to a noticeable reduction in activity. Finally, 4-(4-chlorobenzylidene)-2-(2-phenoxypyphenyl) acetohydrazide (9d), was the most potent anti-inflammatory agent in this series, producing 32-58% reduction in inflammation 1 to 3 h prior to the administration of the drug.

Conclusion

Various substituted arylidine-2-phenoxynicotinic acid hydrazide derivatives were synthesized and screened for their potential anti-inflammatory activity. Most compounds exhibited moderate to good anti-inflammatory activities in comparison with diclofenac. Compounds 9a, 9d and 9e exhibited the most prominent and consistent anti-inflammatory activity.

References

(1) Lipsy P. The role of cyclooxygenase-2-specific inhibitors in clinical practice. Am. J. Med. (2001) 110: 15-58.
(2) Palomer A, Cabré F, Pascual J, Campos J, Trujillio MA, Entrena A, Gallo MA, Garcia L, Mauleon D, Espinosa A. Identification of novel cyclooxygenase-2 selective inhibitors using pharmacophore models. J. Med. Chem. (2002) 45: 1402-1411.
(3) Talley JI, Brown DL, Carter JS, Graneto MJ, Koboldt CM, Masferrer JL, Perkins WE, Rogers RS, Shaffer AF, Zhang YY, Zweifel BS, Seibert K. 4-[5-Methyl-3-phenylisoxazol-4-y1]-benzenesulfonamide, valdecoxib: a potent and selective inhibitor of COX-2. J. Med. Chem. (2000) 43: 775-777.
(4) Cryer B. NSAIDs and Risks. NSAID-associated deaths: the rise and fall of NSAID- associated GI mortality.
Am. J. Gastroenterol. (2005) 100: 1685-1693.

Go MF. Drug injury in the upper gastrointestinal tract: non-steroidal anti-inflammatory drugs. Gastrointest. Endosc. Clin. N. Am. (2006) 16: 83-97.

James MW and Hawkey JH. Assessment of non-steroidal anti-inflammatory drug (NSAID) damage in the human gastrointestinal tract. J. Clin. Pharmacol. (2003) 56: 146-155.

Lazzaroni M and Porro GB. Gastrointestinal side-effects of traditional non-steroidal anti-inflammatory drugs and new formulations. Aliment. Pharmacol. Ther. (2004) 20: 48-58.

Schneider V, Lévesque LE, Zhang B, Hutchinson T, Brophy JM. Association of selective and conventional nonsteroidal anti-inflammatories with acute renal failure: a population-based, nested case-control analysis. Am. J. Epidemiol. (2006) 164(9):881-9.

Adebayo D and Bjarnason I. Is non-steroidal anti-inflammatory drug (NSAID) enteropathy clinically more important than NSAID gastropathy? Postgrad. Med. J. (2006) 82: 186-191.

Chiroli V, Benedini F, Ongini E and Del Soldato P. Nitric oxide-donating non-steroidal antiinflammatory drugs. Eur. J. Med. Chem. (2003) 38: 441-446.

Corazzi T, Leone M, Mauceri R, Corazzi Land Gresele P. Direct and irreversible inhibition of cyclooxygenase-1 by nitrosparin (NCX 4016). J. Pharmacol. Exp. Ther. (2005) 315: 1331-1337.

Velazquez CA, Praveen Rao PN, Citro ML, Keefer LK, Knaus EE. O^-Acetoxyethylmethyl-protected diazeniumdiolate-based NSAIDs (NONO-NSAIDs): Synthesis, nitric oxide release, and biological evaluation studies. Bioorg. Med. Chem. (2007) 15: 4767-4774.

Downing JEG, Madden JC, Ingram MJ and Rostron C. Gastric and thymic assay of acute oral treatment of rats with nitric oxide esters of ibuprofen or indomethacin. Biochem. Biophys. Res. Commun. (2005) 334: 646-653.

Duarte CD, Tributino JL, Lacerda D, Martins MV, Moreira AM, Dutra F, Bechara EJ, Paula FS, Goulart MO, Ferreira J, Calixto JB, Nunes MP, Bertho AL, Miranda ALP, Barreiro E and Fraga CA. Synthesis, pharmacological evaluation and biochemical studies of novel 6-nitro-3,4-methylenedioxyphenyl-N-acylhydrazone derivatives: discovery of LASSBio-881, a new ligand of cannabinoid receptors. Bioorg. Med. Chem. (2007) 15: 2421-2433.

Figueiredo JM, Camara CA, Amarante EG, Miranda ALP, Santos FM, Radrigues CR, Fraga CA and Barreiro EJ. Design and synthesis of novel potent antinociceptive agents: methyl-imidazolyl N-acylhydrazone derivatives. Bioorg. Med. Chem. (2000) 8: 2243-2248.

Almasirad A, Tajik M, Bakhtiari D, Shafee A, Abdollahi M, Zamani MJ, Khorasani R and Esmaili A. Synthesis and analgesic activity of N-Arylhydrazone derivatives of mfenamic acid. J. Pharm. Pharmac. Sci. (2005) 8: 419-425.

Almasirad A, Hosseini R, Jalali-Zaedeh H, Rahimi M, Moghaddam Z, Abarian N, Janafrooz M, Abbaspour M, Ziaeve A, Dalvand A and Shafee A. Synthesis and analgesic activity of 2-phenoxybenzoic acid and N-phenylthranilic acid hydrazides. Biol. Pharm. Bull. (2006) 29: 1180-1185.

Al-Haboubi HA and Zeitlin J. Re-appraisal of the role of histamine in carrageenan-induced paw oedema. Eur. J. Pharmacol. (1983) 88: 169-176.

Perez MM, Docampo RM, Puigmarti Codina JM, Repolles-Moliner J and Solà JS. (1986) U.S. Pat. Appl. US 4623648.

Pirkle WH and Beure SD. Optically active solvents in nuclear magnetic resonance spectroscopy. IX. Direct determinations of optical purities and correlation of absolute configuration of a-amino acids. J. Am. Chem. Soc. (1969) 91: 5150-5155.

Firoozi F, Iavidnia F, Kamar M, Fooladi M, Forumadi A, Shafee A. Synthesis of substituted 1-methyl-2-(1,3,4-thiadiazol-2-yl)-4-nitropyroles. J. Heterocycl. Chem. (1995) 32: 123-128.

Kamal A, Khan MN, Reddy KS and Rohini K. Synthesis of a new class of 2-anilino substituted nicotinyl arylsulfonhydrazides as potential anticancer and antibacterial agents. Bioorg. Med. Chem. (2007) 15: 1004-1013.

Ribeiro IG, Silva KCM, Parrini SC, Miranda ALP, Fragal CAM and Barreiro E. Synthesis and antinociceptive properties of new structurally planned imidazo[1,2-a] pyridine 3-acylhydrazone derivatives. Eur. J. Med. Chem. (1998) 33: 225-235.

Figueiredo JM, Ca’ mara CA, Amarante EG, Miranda AL, Santos FM, Rodrigues CR, Fraga CA and Barreiro EJ. Design and synthesis of novel potent antinociceptive agents: methyl-imidazolyl N-acylhydrazone derivatives. Bioorg. Med. Chem. (2000) 8: 2243-2248.

Cunha AC, Figueiredo JM, Tributino JL, Miranda AL, Castro HC, Zingali RB, Fraga CA, Souza MC, Ferreira VF and Barreiro EJ. Antiplatelet properties of novel N-substituted-phenyl-1,2,3-triazole-4-acylhydrazone. Bioorg. Med. Chem. (2003) 11: 2051-205.