Impact of pretreatment dihydropyrimidine dehydrogenase genotype-guided fluoropyrimidine dosing on chemotherapy associated adverse events

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Abstract
Consensus guidelines exist for genotype-guided fluoropyrimidine dosing based on variation in the gene dihydropyrimidine dehydrogenase (DPYD). However, these guidelines have not been widely implemented in North America and most studies of pretreatment DPYD screening have been conducted in Europe. Given regional differences in treatment practices and rates of adverse events (AEs), we investigated the impact of pretreatment DPYD genotyping on AEs in a Canadian context. Patients referred for DPYD genotyping prior to fluoropyrimidine treatment were enrolled from December 2013 through November 2019 and followed until completion of fluoropyrimidine treatment. Patients were genotyped for DPYD c.1905+1G>A, c.2846A>T, c.1679T>G, and c.1236G>A. Genotype-guided dosing recommendations were informed by Clinical Pharmacogenetics Implementation Consortium guidelines. The primary outcome was the proportion of patients who experienced a severe fluoropyrimidine-related AE (grade ≥3, Common Terminology Criteria for Adverse Events version 5.0). Secondary outcomes included early severe AEs, severe AEs by toxicity category, discontinuation of fluoropyrimidine treatment due to AEs, and fluoropyrimidine-related death. Among 1394 patients, mean (SD) age was 64 (12) years, 764 (54.8%) were men, and 47 (3.4%) were DPYD variant carriers treated with dose reduction. Eleven variant carriers (23%) and 418 (31.0%) noncarriers experienced a severe fluoropyrimidine-related AE (p = 0.265). Six carriers (15%) and 284 noncarriers (21.1%) experienced early severe fluoropyrimidine-related AEs (p = 0.167). DPYD variant carriers treated with genotype-guided dosing did not experience an increased risk for severe AEs. Our data support a role for DPYD genotyping in the use of fluoropyrimidines in North America.

Study Highlights
WHAT IS THE CURRENT KNOWLEDGE ON THE TOPIC?
Dihydropyrimidine dehydrogenase (DPD) deficiency is associated with fluoropyrimidine-related adverse events (AEs), and screening for DPD deficiency can...
be carried out using *DPYD* genotype testing of clinically relevant variants, as noted in the Clinical Pharmacogenetics Implementation Consortium guidelines.

**WHAT QUESTION DID THIS STUDY ADDRESS?**

Given the paucity of data relating to pretreatment use of *DPYD* genotyping in North America, this Canadian study adds new insights to the clinical impact of *DPYD* testing.

**WHAT DOES THIS STUDY ADD TO OUR KNOWLEDGE?**

*DPYD* genotype-guided dosing can ameliorate fluoropyrimidine-related AE risk for patients treated with fluoropyrimidine dose and regimens prescribed in North America.

**HOW MIGHT THIS CHANGE CLINICAL PHARMACOLOGY OR TRANSLATIONAL SCIENCE?**

Although the European Medicines Agency supports DPD deficiency screening, this study suggests that similar efforts should be undertaken in North America.

**INTRODUCTION**

Five-fluouracil (5-FU) and capecitabine are fluoropyrimidines used in the treatment of solid tumours. Unfortunately, ~30% of patients experience severe fluoropyrimidine-related toxicity. Dihydropyrimidine dehydrogenase (DPD, gene name *DPYD*) is the rate-liming enzyme in fluoropyrimidine catabolism. DPD deficiency increases the risk of fluoropyrimidine-related toxicity, and there are heritable *DPYD* variants associated with decreased enzyme function and thereby DPD deficiency. Meta-analyses have narrowed the list of clinically relevant genetic variants allowing the implementation of genotype-guided dosing.

In 2013, the Clinical Pharmacogenetics Implementation Consortium (CPIC) published a consensus guideline detailing fluoropyrimidine dosing recommendations for 3 *DPYD* variants associated with reduced enzymatic activity: *DPYD* c.1905+1G>A (*2A, rs39818290), c.2846A>T (rs67376798), and c.1679T>G (*13, rs55886062). For heterozygous carriers of an individual variant, a 50% dose reduction was recommended, whereas avoidance of fluoropyrimidines was recommended for homozygous or compound heterozygous variant carriers. A fourth intronic variant, *DPYD* c.1129-5923C>G (rs75017182, in linkage disequilibrium with *DPYD* c.1236G>A [rs56038477]) was added to the guideline in 2017. These recommendations were also refined based on enzymatic activity scores (AS). The AS of each patient is the sum of the individual alleles where each allele is assigned a score of 0 to 1 based on functional characterization. The AS of clinically relevant alleles are 0 for *DPYD* c.1905+1G>A or c.1679T>G and 0.5 for *DPYD* c.2846A>T or c.1129-5923C>G. A 25% to 50% dose reduction was recommended for intermediate metabolizers with an AS of 1.5 and a 50% dose reduction was recommended for an AS of 1 and avoidance for an AS of 0 to 0.5. In 2018, results from Henricks et al. led to further updates of the CPIC guidelines to recommend a 50% dose reduction for AS of either 1 or 1.5. In addition, following a well-tolerated initial dose reduction, the CPIC encourages cautious dose escalation and with concurrent therapeutic drug monitoring if available. Of note, the CPIC guidelines provide reference for patients with available genotype data and do not comment on the necessity of preemptively determining the *DPYD* genotype.

In addition to the CPIC guidelines for response to known *DPYD* variants, Dutch and French initiatives have published guidelines that explicitly recommend DPD deficiency screening prior to fluoropyrimidine therapy. Despite this, adoption of pretreatment *DPYD* genotyping in Canada has been limited and currently is widely accessible only in Quebec. Given the abundance of data linking complete DPD deficiency to severe toxicity, a randomized controlled trial of pretreatment DPD deficiency screening was considered to be inappropriate for our center. The only two-arm comparative study of DPD deficiency screening was terminated prematurely due to the fluoropyrimidine-related death of a DPD-deficient patient in the control arm. However, two prospective *DPYD* single arm genotype-guided studies were completed in the Netherlands, the first examined the impact of one variant (c.1905+1G>A), and the second assessed four variants (c.1905+1G>A, c.2846A>T, c.1679T>G, and c.1236G>A). These studies demonstrated that genotype-guided dosing reduces the risk of adverse events (AEs) for *DPYD* variant carriers in a European population compared to the historical rate in *DPYD* variant carriers receiving the standard of care.

In contrast, the impact of pretreatment *DPYD* genotype-guided fluoropyrimidine dosing in North America is unpublished. There is an important distinction between results from a European population and the potential results in a North American population. Work by Haller et al. has identified regional variation in fluoropyrimidine-related AEs between the United States and Europe. Therefore, there is a need for regional data to support regional implementation. Here, we
conducted a study to determine the impact of pretreatment *DPYD* genotype-guided dosing on patient safety at a tertiary care center in London, Ontario, Canada. We hypothesized that *DPYD* variant carriers who received a genotype-guided dosing would have no greater risk of fluoropyrimidine-related AEs as compared with noncarriers.

**PATIENTS AND METHODS**

**Study Sample**

We conducted a single-center retrospective study of patients referred to the Personalized Medicine clinic at London Health Sciences Centre (LHSC), London, Ontario, Canada, for *DPYD* genotype testing between December 1, 2013, and November 30, 2019. The study was approved by the Institutional Review Board at Western University and all patients provided written informed consent. Of 1945 patients referred for testing, 1845 were tested prior to fluoropyrimidine treatment. At initiation, the study was based on the 2013 CPIC guidelines; however, in response to the 2017 update, the *DPYD* c.1236G>A variant was added to the testing panel in May 2018. Consequently, 41 c.1236G>A carriers identified retrospectively were removed from the study as they did not receive appropriate genotype-guided dosing. Two compound heterozygous carriers were identified among the genotype-guided patients and the treating oncologists were advised to select an alternative therapy. There were 1394 patients who initiated treatment through LHSC prior to December 1, 2019, that were included in the genotype-guided study (Figure 1, baseline characteristics are summarized in

![Flow diagram illustrating the study cohort. AE, adverse event; DPYD, dihydropyrimidine dehydrogenase](image-url)
Table 1). Prior chemotherapy, radiation therapy, concurrent antineoplastic therapies, and other concomitant medications were allowed. Baseline characteristics for patients lost to follow-up are shown in Table S1.

**DPYD genotype testing and dosing recommendations**

Whole blood samples were collected from each patient and DNA was extracted using the MagNA Pure Compact Instrument (Roche). DNA was assessed on a ViiA 7 real-time polymerase chain reaction system (ThermoFisher Scientific) using TaqMan allelic discrimination assays (ThermoFisher Scientific) for **DPYD** c.1905+1G>A (assay ID: C___30633851_20), c.2846A>T (assay ID: C__27530948_10), c.1679T>G (assay ID: C__11985548_10), and c.1236G>A (assay ID: C__25596099_30). Variant c.1236G>A is known to be in strong linkage disequilibrium with c.1129-5923C>G and was used as a proxy for genotyping, which is in alignment with the CPIC guidelines.

Results and dosing recommendations were provided to the referring physicians within the patients’ electronic health records (EHRs). Recommendations were as follows: for noncarriers, dose as per standard of care; for simple heterozygous carriers, apply a 50% initial dose reduction, and consider attempting dose escalation in subsequent cycles pending patient tolerance. A 25% to 50% initial dose reduction was recommended for heterozygous carriers of c.1236G>A upon its addition to the testing panel, with the same additional recommendation to attempt dose escalation based on patient tolerability. Avoidance of fluoropyrimidines in homozygous or compound heterozygous variant carriers was recommended throughout the study, recommendations are summarized in Table 2. Final treatment decisions were at the discretion of the treating oncologist.

**Data collection**

Treatment data, including regimen, dose, and radiation use, were collected from LHSC pharmacy records. Clinical variables and toxicity data were obtained by standardized review of the patients’ EHRs by trained study personnel, each record was reviewed independently by two study members. Toxicity data were recorded from clinic notes, admission records, discharge summaries, and emergency department reports. Severe AEs included grade greater than or equal to 3 toxicities according to the National Cancer Institutes’ Common Terminology Criteria for Adverse Events (CTCAE) version 5.0.22 Only those AEs determined to be possibly, probably, or definitely related to the fluoropyrimidine components were included in the outcome, following the standard definitions proposed in the National Institutes of Health protocol template for phase II/III trials.23 Based on these principles, the definitions require the AE to occur within 30 days of fluoropyrimidine administration, be pharmacologically plausible, and not be attributable to another component of the regimen. The effect of removing and reinstating the fluoropyrimidine were also considered when these challenges occurred. Based on the literature, the major toxicity categories considered were gastrointestinal (including primarily: diarrhea, oral mucositis, and nausea/vomiting), myelosuppression (primarily neutropenia/febrile neutropenia, as well as thrombocytopenia, and unexplained anemia), cardiac (sudden onset cardiac toxicity during fluoropyrimidine administration), and Palmar-Plantar Erythrodysesthesia (Hand-Foot Syndrome). The remaining AEs, in which fluoropyrimidines were likely contributors, were grouped under the other heading. The AEs reported by the initial reviewers and attribution of causality was reviewed by a medical oncology fellow under the supervision of a practicing medical oncologist. Conflicts in the records were reviewed by the initial coders and the reviewing medical oncologist. Patients were followed for their entire treatment period and until toxicity resolved.

**Outcomes**

The primary outcome was severe (grade ≥3, CTCAE version 5.0) fluoropyrimidine-related AEs. We included a secondary outcome of early fluoropyrimidine-related AEs during the first two cycles of treatment. Secondary outcomes further included fluoropyrimidine-related AEs by toxicity category, proportion of patients discontinuing fluoropyrimidines due to fluoropyrimidine-related AEs, and fluoropyrimidine-related deaths.

**Statistical methods**

The primary outcome was compared between **DPYD** variant carriers and noncarriers using a $\chi^2$ test. Other dichotomous outcomes were compared using $\chi^2$ or Fisher’s Exact test as appropriate. Fisher’s Exact tests were used when cell values in contingency tables were less than or equal to 5. A test for noninferiority between AEs in the variant carriers and noncarriers was performed using a two-one sided test of equivalence. The smallest effect size of interest (SESOI) was determined using the lower bound for the 95% confidence interval (CI) of the risk for the c.1236G>A variant carriers in the literature multiplied by the event rate in noncarriers in this study. The c.1236G>A demonstrated the lowest increased risk and using this value to set the SESOI was considered a conservative approach. Unadjusted relative risk was used to show the risk of grade greater than
or equal to 3 AE in our genotype-guided study and within the literature. Unadjusted relative risks are reported due to the low number of events among variant carriers, and for consistency with previous genotype-guided studies. A multivariable logistic regression determined the adjusted odds ratios and is available within the supplementary data. A Wilcoxon Mann-Whitney $U$ test was used to compare the number of cycles administered between variant carriers and noncarriers. Descriptive statistics are shown using number (percentage), mean (SD), and median (interquartile range).

| Characteristic | Genotype-guided cohort | Retrospective sample |
|----------------|------------------------|----------------------|
|                | Noncarrier ($N = 1347$) | Carrier ($N = 47$) | c.1236G>A carrier ($N = 41$) |
| Sex, $N$ (%)   |                        |                      |                                |
| Female         | 605 (44.9)             | 25 (53)             | 13 (31)                        |
| Male           | 742 (55.1)             | 22 (47)             | 28 (68)                        |
| Race, $N$ (%)  |                        |                      |                                |
| White          | 1267 (94)              | 45 (96)             | 40 (98)                        |
| Other$^a$      | 32 (2.4)               | 1 (2)               | 1 (2)                          |
| Unknown$^b$    | 48 (3.6)               | 1 (2)               | 0 (0)                          |
| Age, mean (SD), years | 64 (12) | 62 (13) | 66 (10.4) |
| Body surface area, mean (SD), m$^2$ | 1.9 (0.3) | 1.9 (0.2) | 1.94 (0.27) |
| Tumor site, $N$ (%) |                      |                      |                                |
| Colorectal     | 779 (57.8)             | 25 (53)             | 21 (51)                        |
| Gastric and esophagus | 189 (14.0) | 7 (15) | 5 (12) |
| Pancreas       | 106 (7.9)              | 6 (13)              | 2 (5)                          |
| Breast         | 89 (6.6)               | 3 (6)               | 1 (2)                          |
| Anus           | 48 (3.6)               | 1 (2)               | 3 (7)                          |
| Head and neck  | 27 (2.0)               | 2 (4)               | 3 (7)                          |
| Other$^c$      | 109 (8.1)              | 3 (6)               | 6 (15)                         |
| Regimen, $N$ (%) |                      |                      |                                |
| Capcitabine with radiation | 277 (20.6) | 11 (23) | 11 (27)                       |
| Capcitabine monotherapy$^d$ | 229 (17.0) | 7 (15) | 8 (20)                       |
| Capcitabine with oxaliplatin | 130 (9.7) | 2 (4) | 2 (5) |
| Capcitabine with other agents$^e$ | 68 (5.0) | 3 (6) | 1 (2) |
| FOLFOX$^d$     | 228 (16.9)             | 8 (17)              | 9 (22)                         |
| FOLFIRI/FOLFIRINOX | 135 (10.0) | 8 (17) | 0 (0) |
| 5-FU with cisplatin/oxaliplatin | 128 (9.5) | 4 (9) | 3 (7) |
| 5-FU with other agents$^f$ | 152 (11.3) | 4 (9) | 7 (17) |
| DPYD genotype, $N$ (%) |                      |                      |                                |
| Wild-type      | 1347 (100)             | 0 (0)               | 0 (0)                          |
| c.2846A>T heterozygous | 0 (0) | 19 (40) | 0 (0) |
| c.1905+1G>A heterozygous | 0 (0) | 9 (19) | 0 (0) |
| c.1679T>G heterozygous | 0 (0) | 1 (2) | 0 (0) |
| c.1236G>A heterozygous | 0 (0) | 18 (38) | 41 (100) |

Abbreviation: 5-FU, 5-fluorouracil.

$^a$Other includes Black, Asian, and Indigenous.

$^b$Due to self-declaration of race not all patients opted to provide this information and it remains unknown.

$^c$Other included appendix and small bowel, genitourinary, hepatobiliary, and primary site unknown.

$^d$Including with and without biologic agents.

$^e$Including gemcitabine, lapatinib, temozolomide, docetaxel, epirubicin, and mitomycin + radiation.

$^f$Including Degramount, FEC-D, and FLOT regimens, in addition to mitomycin + radiation.
range (IQR) as applicable. Reported p values are for two-sided tests, with p < 0.05 considered significant. All analyses were performed using R (version 4.0.2; R Foundation Inc.; http://cran.r-project.org/). In addition, the package “tidyverse” was used for data processing and both “epiR” and “TOSTER” were used for analysis, the script used for analysis is available in the supplementary materials.

**RESULTS**

**Study population**

Among the 1394 patients provided genotype-guided dosing, the mean (SD) age was 64 (12) years and 764 (54.8%) were men. The most common primary tumor site was colorectal (804, 57.7%). Overall fluoropyrimidine use was distributed between capecitabine (727, 52.2%) and 5-FU (667, 47.8%). Forty-seven patients (3.4%) were heterozygous carriers for one of $DPYD$ c.2846A>T (19, 1.4%), c.1236G>A (18, 1.3%), c.1905+1G>A (9, 0.6%), or c.1679T>G (1, <0.1%). The retrospectively identified c.1236G>A carriers did not appear to differ from the primary study, with the most common primary tumor site being colorectal (21, 51%), and an approximately equal use of capecitabine (23, 56%), and 5-FU (18, 44%). However, the retrospective sample contained more men (28/41, 68%) than women. The baseline characteristics are summarized in Table 1.

**Physician compliance with dose recommendations**

We confirmed that variant carriers were treated according to the dose recommendations provided to the treating oncologist. The mean initial dose intensity was 52% (18) of ideal for variant carriers and 87.4% (15.2) for noncarriers (Table 3). Variant carriers received a median (IQR) of 6 (2–7) cycles of fluoropyrimidine treatment, and noncarriers received a median of 4 (2–6) cycles. We also assessed the mean dose intensity throughout the treatment period and found that variant carriers received a mean dose intensity over the total treatment period of 55% (15), whereas mean intensity for noncarriers was 84.2% (14.7).

**Toxicity outcomes**

There were no significant differences in the primary or secondary toxicity outcomes between genotype-guided variant carriers and noncarriers. We observed that 23% (11/47) of variant carriers, and 31.0% (418/1347) of noncarriers
experienced severe fluoropyrimidine-related AEs during their total treatment periods \((p = 0.265; \text{Table } 3)\). We next examined severe fluoropyrimidine-related AEs that occurred during the early cycles \((1–2)\) of fluoropyrimidine treatment. We found that 13% \((6/47)\) of genotype-guided variant carriers compared to 21.4% \((284/1347)\) of genotype-guided noncarriers experienced an early fluoropyrimidine-related AE \((p = 0.167; \text{Table } 4)\). Secondary analyses of the major AE categories, proportion discontinuing fluoropyrimidines due to AEs and fluoropyrimidine-related deaths, during the total treatment period or the first two cycles, did not show any significant differences between genotype-guided variant carriers and noncarriers. Additionally, we performed noninferiority testing comparing the risk for global severe fluoropyrimidine-related AEs between carriers and noncarriers both during the total treatment and limited to the early cycles (Figure 2). In both early and total treatment periods, the CIs included no difference but did not cross the noninferiority margin. Therefore, we conclude that genotype-guided variant carriers do not experience increased risk of fluoropyrimidine-related AEs compared with noncarriers receiving the standard of care dosing practices. We determined the unadjusted relative risk \((\text{RR})\) of grade greater than or equal to 3 fluoropyrimidine-related AEs in our genotype-guided variant carriers to allow for comparison to literature values (Table 5)\(^{11,15}\). We report unadjusted RR values due to the small number of genotype-guided variant carriers in our cohort and the literature. We obtained historical values for RR of fluoropyrimidine-related AEs without genotype-guidance from a meta-analysis by Meulendijks et al.\(^{11}\). In our cohort, genotype-guided variant carriers were not at a significantly elevated risk for severe fluoropyrimidine-related AEs compared with noncarriers. Indeed, with the recommended 50% dose reduction the RR was 1.08 \((95\% \text{ CI: } 0.43–2.74)\) for c.1905+1G>A carriers, and 0.85 \((95\% \text{ CI: } 0.40–1.82)\) for c.2846A>T carriers. With the recommended 25% to 50% dose reduction recommendations, the RR for genotype-guided c.1236G>A carriers was 0.54 \((95\% \text{ CI: } 0.19–1.52)\). Finally, the single c.1679T>G carrier in our genotype-guided cohort was treated with a 50% dose reduction and did not suffer any fluoropyrimidine-related AEs during treatment.\(^{11}\) Additionally, we performed a secondary calculation of

| TABLE 3 Severe fluoropyrimidine-related adverse events during total treatment period |
|---------------------------------------------------------|
| **Genotype-guided cohort**                              |
| Noncarrier \((N = 1347)\) | Carrier \((N = 47)\) | Genotype | \(P\) value | c.1905+1G>A \((N = 9)\) | c.2846A>T \((N = 19)\) | c.1679T>G \((N = 1)\) | c.1236G>A \((N = 18)\) | Retrospective sample \((N = 41)\) |
|---------------------------------------------------------|
| Initial dose intensity, mean (SD)                       |
| 87.4 (15.2) | 52 (18) | NA | 47 (16) | 47 (21) | 43 (NA) | 59 (13) | 85 (17) |
| Dose intensity, mean (SD)                               |
| 84.2 (14.7) | 55 (13) | NA | 46 (8) | 55 (15) | 50 (NA) | 59 (12) | 85 (17) |
| Treatment cycles, median (IQR)                          |
| 4 (2–6) | 6 (2–7) | 0.201 | 6 (2–8) | 6 (4–8) | 6 (NA) | 4 (2–6) | 2 (2–4) |
| Total severe AEs\(^b\) (all cycles), \(N (\%)\)         |
| Global\(^c\) | 418 (31.0) | 11 (23) | 0.265 | 3 (33) | 5 (26) | 0 (0) | 3 (17) | 14 (34) |
| Gastrointestinal | 167 (12.4) | 6 (12) | 0.940 | 2 (22) | 2 (11) | 0 (0) | 2 (11) | 7 (17) |
| Myelosuppression | 157 (11.7) | 6 (12) | 0.816 | 2 (22) | 2 (11) | 0 (0) | 2 (11) | 2 (5) |
| Cardiac       | 33 (2.4) | 0 (0) | 0.625 | 0 (0) | 0 (0) | 0 (0) | 0 (0) |
| HFS           | 35 (2.6) | 1 (2) | >0.99 | 0 (0) | 1 (5) | 0 (0) | 0 (0) |
| Other\(^d\)   | 113 (8.4) | 2 (4) | 0.425 | 1 (11) | 1 (5) | 0 (0) | 0 (0) |
| AE-related death\(^e\)                                 |
| 10 (0.7) | 0 (0) | >0.99 | 0 (0) | 0 (0) | 0 (0) | 0 (0) |
| Discontinued treatment\(^f\)                           |
| 232 (17.2) | 10 (21) | 0.437 | 2 (22) | 3 (16) | 0 (0) | 5 (28) | 7 (17) |

Abbreviations: AEs, adverse events; HFS, hand-foot syndrome; IQR, interquartile range; NA, not applicable.

- \(^a\)\(P\) value for treatment cycles was calculated based on Wilcoxon-Mann-Whitney test. \(P\) values for fluoropyrimidine-related AEs calculated using the following tests: Global, Gastrointestinal, Myelosuppression, and Discontinued Treatment utilized \(\chi^2\) tests; Cardiac, HFS, Other, and AE-related Death utilized Fisher’s Exact Test.
- \(^b\)\(Grade \geq 3\) by Common Terminology Criteria for Adverse Events version 5.0.
- \(^c\)Global includes all fluoropyrimidine-related AEs grade \(\geq 3\) and fluoropyrimidine-related deaths. This does not include discontinuation.
- \(^d\)Other grade \(\geq 3\) AEs included: fatigue, infections, neurotoxicities, and laboratory abnormalities.
- \(^e\)At least one fluoropyrimidine-related AE contributed significantly to death.
- \(^f\)Patients discontinuing treatment with fluoropyrimidines due to a fluoropyrimidine-related AE of any grade.
multivariable logistic regression adjusting for age, sex, regimen, and initial intensity of therapy. We do not include this in the primary report due to the small sample size of variants and the potential for the introduction of bias during adjustment. However, we note that there were no significant differences from the unadjusted predictions and these results can be found in Table S2.

**Retrospectively identified DPYD c.1236G>A carriers**

*DPYD* c.1236G>A carriers identified retrospectively in May 2018 (*N* = 41) were removed from the genotype-guided cohort as they were treated as *DPYD* variant noncarriers. We predicted that these c.1236G>A carriers would experience an increased risk of fluoropyrimidine-related AEs given they were treated with standard dosing. However, c.1236G>A carriers treated with standard dosing did not experience an elevated toxicity profile (Table 3). In brief, 34% (14/41) of the retrospectively identified c.1236G>A carriers experienced a severe fluoropyrimidine-related AE during the total treatment period and 24% (10/41) experienced an early severe fluoropyrimidine-related AE (Table 4). In summary, compared to the genotype-guided cohort, the unadjusted relative risk of global severe fluoropyrimidine-related AEs was 1.09 (0.71–1.68).

**DISCUSSION**

We report the impact of pretreatment *DPYD* genotype-guided fluoropyrimidine dosing on AEs in a Canadian hospital assessed through retrospective follow-up of the Personalized Medicine Clinic. We show that when treated with genotype-guided dosing for *DPYD* c.1905+1G>A, c.1679T>G, c.2846A>T, or c.1236G>A, the proportion of variant carrying patients who experienced severe fluoropyrimidine-related AEs was not statistically different from noncarriers. We found that a 50% dose reduction for *DPYD* c.1905+1G>A and c.2846A>T carriers ameliorated the severe fluoropyrimidine-related AE risk compared to the historical RR for carriers treated with full dose (Table 5). Previously, Henricks et al. reported that a 25% initial dose reduction in carriers of *DPYD* c.2846A>T did not eliminate the elevated risk of severe fluoropyrimidine-related AEs.15 Together these findings suggest that an initial 50% dose reduction is an appropriate dosing strategy for carriers of *DPYD* c.1905+1G>A and c.2846A>T, consistent with the current CPIC guidelines.9

The Personalized Medicine Clinic attempted to provide *DPYD* genotype-guided dosing in alignment with the best available evidence. Indeed, the genotyping for *DPYD* c.1236G>A as a proxy for variant c.1129-5923C>G starting in 2018 reflects the latest CPIC guideline recommendations that note the association of c.1129-5923C>G with severe fluoropyrimidine-related AEs.11,24 In order to account for this

### Table 4 Early severe fluoropyrimidine-related AEs

| Genotype-guided cohort | Noncarrier (N = 1347) | Carrier (N = 47) | P value[^a] | c.1905+1G>A (N = 9) | c.2846A>T (N = 19) | c.1679T>G (N = 1) | c.1236G>A (N = 18) | Retrospective sample c.1236 G>A (N = 41) |
|-----------------------|----------------------|-----------------|------------|---------------------|-------------------|-------------------|-------------------|-----------------------------------------|
| Early severe AE[^b] (cycles 1–2), N (%) | N (%) | N (%) | N (%) | N (%) | N (%) | N (%) | N (%) | N (%) | N (%) |
| Global[^c] | 284 (21.1) | 6 (13) | 0.167 | 2 (22) | 3 (16) | 0 (0) | 1 (5) | 10 (24) |
| Gastrointestinal | 131 (9.7) | 3 (6) | 0.616 | 1 (11) | 1 (5) | 0 (0) | 1 (5) | 5 (12) |
| Myelosuppression | 102 (7.6) | 5 (11) | 0.401 | 2 (22) | 2 (11) | 0 (0) | 1 (5) | 1 (2) |
| Cardiac | 26 (1.9) | 0 (0) | >0.99 | 0 (0) | 0 (0) | 0 (0) | 0 (0) | 0 (0) |
| HFS | 13 (1.0) | 0 (0) | >0.99 | 0 (0) | 0 (0) | 0 (0) | 0 (0) | 2 (5) |
| Other[^d] | 68 (5.0) | 2 (4) | >0.99 | 1 (11) | 1 (5) | 0 (0) | 0 (0) | 3 (7) |
| AE-related death[^e] | 8 (0.6) | 0 (0) | >0.99 | 0 (0) | 0 (0) | 0 (0) | 0 (0) | 0 (0) |
| Discontinued treatment[^f] | 137 (10.2) | 5 (11) | 0.808 | 2 (22) | 1 (5) | 0 (0) | 2 (11) | 4 (10) |

Abbreviations: AEs, adverse events; HFS, hand-foot syndrome.

[^a]P values for fluoropyrimidine-related AEs calculated using the following tests: Global utilized χ² test; Gastrointestinal, Myelosuppression, Cardiac, HFS, Other, AE-related death, and Discontinued Treatment utilized Fisher’s Exact Test.

[^b]Grade ≥3 by Common Terminology Criteria for Adverse Events version 5.0.

[^c]Global includes all fluoropyrimidine-related AEs grade ≥3 and fluoropyrimidine-related deaths. This does not include discontinuation.

[^d]Other AEs included: fatigue, infections, neurotoxicities, and laboratory abnormalities.

[^e]At least one fluoropyrimidine-related AE contributed significantly to death.

[^f]Patients discontinuing treatment with fluoropyrimidines due to a fluoropyrimidine-related AE of any grade.
in this analysis, we carried out retrospective genotyping for 

DPYD c.1236G>A for patients who had been enrolled prior to 

inclusion of this variant as part of the DPYD test panel. We hypothesized that our patients who were DPYD c.1236G>A carriers treated with standard dosing would exhibit an increased risk of fluoropyrimidine-related AEs in alignment with previous meta-analysis data, as cited in the CPIC guidelines.\(^{11,14}\) However, the retrospectively identified DPYD c.1236G>A carriers in our study did not demonstrate an increased risk. In the meta-analysis by Meulendijks et al. that demonstrated an increased risk associated with DPYD c.1236G>A, however, the included studies consisted of only European populations (\(N = 4261\)).\(^{11}\) Subsequently to the meta-analysis publication, a large association study of American patients with colorectal cancer (\(N = 1953\)) demonstrated no significant association between DPYD c.1129-5923C>G and fluoropyrimidine-related

| DPYD variant | Genotype-guided dosing current cohort\(^a\) RR (95% CI)\(^d\) | Patients treated without genotype-guided dosing\(^b\) RR (95% CI)\(^d\) | Genotype-guided dosing literature cohort\(^c\) RR (95% CI)\(^d\) |
|--------------|-------------------------------------------------|-------------------------------------------------|-------------------------------------------------|
| c.1905+1G>A  | 1.08 (0.43–2.74)                                 | 2.87 (2.14–3.86)                                 | 1.31 (0.63–2.72)                                 |
| c.2846A>T    | 0.85 (0.40–1.82)                                 | 3.11 (2.25–4.28)                                 | 2.00 (1.19–3.34)                                 |
| c.1679T>G    | NA\(^e\)                                         | 4.30 (2.10–8.80)                                 | NA\(^e\)                                         |
| c.1236G>A    | 0.54 (0.19–1.52)                                 | 1.72 (1.22–2.42)                                 | 1.69 (1.18–2.42)                                 |

Abbreviations: CI, confidence interval; NA, not applicable; RR, relative risk.

\(^a\)Our genotype-guided cohort: 50% dose reduction recommended for carriers of c.1905+1G>A, c.2846A>T, and c.1679T>G; 25%–50% dose reduction for carriers of c.1236G>A.

\(^b\)Meulendijks et al. historical cohort derived from a meta-analysis\(^1\): standard of care dosing with adjustment due to tolerability resulting in the assumption that given no genotype was known the dose intensity was equivalent between DPYD variant carriers and noncarriers.

\(^c\)Henricks et al. genotype guided cohort\(^16\): 50% dose reduction recommended for carriers of c.1905+1G>A or c.1679T>G; 25% dose reduction for carriers of c.2856A>T or c.1236G>A. Followed by dose escalation pending patient tolerance.

\(^d\)Unadjusted RRs with 95% CIs are discussed due to small sample size of variant carriers in genotype-guided cohorts. Risks are calculated compared with noncarriers of the individual variant of interest.

\(^e\)Only one c.1679T>G carrier was detected in each genotype-guided cohort. In both cohorts, the carrier was treated with 50% dose reduction and did not suffer a fluoropyrimidine-related adverse event.
AEs (RR 1.27, 95% CI: 0.97–1.67) in their population. However, in the American study, they also confirmed that the proxy variant was in complete linkage disequilibrium with the causal variant. Lee et al. did demonstrate a trend toward an association and significance in a secondary outcome associating the c.1129-5923C>G variant with neutropenia. Given the difference between these previous findings and the known regional difference between the United States and European populations, we suggest the lack of significant association in our c.1236G>A carriers may reflect this difference. However, this difference was not proven and may be due to the limited sample size of retrospective c.1236G>A carriers in this study. The CPIC currently supports a 50% dose reduction for DPD c.1236G>A carriers, followed by dose escalation if the patient tolerates the reduced dose. More evidence is needed to elucidate the extent of the potential regional effect on carriers of this variant. In the meantime, we continue to support the CPIC recommendations for DPD c.1236G>A carriers. Given the uncertainty, therapeutic drug monitoring may be useful to limit AEs during dose escalations.

**Limitations**

The first major limitation of our study design is the experimental design. A robust two-arm comparative study directly comparing genotype-guided dosing to standard of care therapy would have provided stronger evidence to support these findings. However, a two-arm comparative study was deemed inappropriate given the body of evidence associating DPD variation and fluoropyrimidine-related AEs prior to initiating the program at the Personalized Medicine Clinic. The retrospective collection of AE outcomes also limits the design. However, as listed in the Methods sections, systems were in place to limit the bias of this data collection and the pragmatic nature was necessary given limitations of the clinic at the time of study initiation. As well, our study design lacks disease progression or survival outcomes. However, it has previously been shown that c.1905+1G>A carriers treated with a 50% starting dose reduction achieved the same fluoropyrimidine exposure as non-carriers with standard dosing. Additionally, a retrospective survival analysis showed no difference in survival outcomes between variant carriers receiving genotype-guided dosing and non-carriers receiving standard dosing. These data suggest that the DPD variant carriers treated with a dose reduction achieve the same systemic exposure and therapeutic outcomes. The four variants tested in this study have been validated in studies predominated by White people of European descent, as was our study population. Additional DPD variants may play an important role in other patient populations (e.g., DPD c.557A>G in people of African descent). Further research in other patient populations is needed to validate the utility of DPD genotype-guided dosing in more diverse populations.

Finally, this study used DPD genotype testing as a pretreatment screening method for DPD deficiency, however, we did not assess other methods of detecting DPD deficiency in this patient population.

**CONCLUSION**

Health Canada and the US Food and Drug Administration include warnings that DPD-deficient patients are at an increased risk of severe AEs on fluoropyrimidine product labels. However, to date, neither agency has recommended any pretreatment screening methods despite consensus guidelines from expert groups in Europe. In March of 2019, the French Medicines Agency triggered a formal review of preemptive DPD deficiency screening by the European Medicines Agency (EMA), and, in April 2020, the EMA issued a recommendation for DPD deficiency testing prior to initiation of fluoropyrimidines. Our data support equivalent efforts to study and implement DPD deficiency screening through DPD genotype testing be undertaken within North America.

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**CONFLICT OF INTEREST**

The authors declared no competing interests for this work.

**AUTHOR CONTRIBUTIONS**

T.J.W., U.T.S., and R.B.K. wrote the manuscript. W.A.T., U.I.S., and R.B.K. designed the research. T.J.W., B.L.P., W.A.T., R.M.L., J.L., S.N., V.P., D.K., J.M., V.S., S.S., Y-H.C., S.W., E.W., U.I.S., and R.B.K. performed the research. W.A.T., R.M.L., J.L., S.N., V.P., D.K., J.M., V.S., S.S., Y-H.C., S.W., E.W., U.I.S., and R.B.K. analyzed the data.

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