Compensatory expression and substrate inducibility of \(\gamma\)-glutamyl transferase GGT2 isoform in \textit{Arabidopsis thaliana}

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Abstract

\(\gamma\)-Glutamyl transferases (GGT; EC 2.3.2.2) are glutathione-degrading enzymes that are represented in \textit{Arabidopsis thaliana} by a small gene family of four members. Two isoforms, GGT1 and GGT2, are apoplastic, sharing broad similarities in their amino acid sequences, but they are differently expressed in the tissues: GGT1 is expressed in roots, leaves, and siliques, while GGT2 was thought to be expressed only in siliques. It is demonstrated here that GGT2 is also expressed in wild-type roots, albeit in very small amounts. GGT2 expression is enhanced in \textit{ggt1} knockout mutants, suggesting a compensatory effect to restore GGT activity in the root apoplast. Supplementation with 100 \(\mu\)M glutathione (GSH) resulted in the up-regulation of GGT2 gene expression in wild-type and \textit{ggt1} knockout roots, and of GGT1 gene expression in wild-type roots. Glutathione recovery was hampered by the GGT inhibitor serine/borate, suggesting a major role for apoplastic GGTs in this process. These findings can explain the ability of \textit{ggt1} knockout mutants to retrieve exogenously added glutathione from the growth medium.

Key words: Apoplast, \(\gamma\)-glutamyl cycle, \(\gamma\)-glutamyl transferase, glutathione degradation, thiol metabolism.

Introduction

Given its central role in cell homeostasis, glutathione (GSH) has received much attention from the scientific community, documented by numerous papers focusing on different aspects of GSH metabolism relating to plant nutrition and assimilation, plant development and cell cycle regulation, plant defence, adaptation to the environment, and redox control (May \textit{et al.}, 1998; Noctor \textit{et al.}, 1998; Vernoux \textit{et al.}, 2000; Beemster \textit{et al.}, 2003; Tausz \textit{et al.}, 2004; Mullineaux and Rausch, 2005; Meyer, 2008; Szalai \textit{et al.}, 2009; Foyer and Noctor, 2009). Many aspects of its metabolism have been fully elucidated in plants, and the biochemical steps leading to its synthesis have been described (Foyer and Noctor, 2001). In leaf cells, cytosol and chloroplast are a major source of GSH, which is transported to sink organs by phloem translocation (Herschbach and Rennenberg, 1995; Herschbach \textit{et al.}, 2000). The metabolic steps leading to xenobiotic detoxification through glutathione conjugation have also been described (Marrs, 1996; Dixon \textit{et al.}, 1998; Edwards \textit{et al.}, 2000).

The metabolic route relating to GSH degradation, which has been shown in animal systems to be fed by \(\gamma\)-glutamyl transferases (GGTs), has only recently received some attention in plant science. The initial assumption was that of a substantial difference between animal and plant organisms in terms of the subcellular localization, substrate affinity, and functions of GGT, but the description of its biochemical properties in plant tissues (Martin and Slovin, 2000) later pointed to broad similarities in GGTs from different organisms. An alternative pathway mediated by carboxypeptidase has also been demonstrated in \textit{Arabidopsis}, which may account for greater GSH degradation (Okkama-Ohtsu \textit{et al.}, 2008); this pathway is localized in the cytosol, however, and any conclusions are applicable to that compartment alone.

The significance of GGTs in plants has recently been addressed in several studies, in which \textit{Arabidopsis thaliana} was used as a model (Storozhenko \textit{et al.}, 2002; Grzam \textit{et al.}, 2007; Martin \textit{et al.}, 2007; Okkama-Otsu \textit{et al.} 2007\textit{a, b}). The
**Arabidopsis** genome is known to carry four GGT isoforms, with specific subcellular and organ-level spatial expression.

GGT1 and GGT2 are apoplastic enzymes. Their genes are consecutive and they are very similar (96% identical), indicating a duplication event during evolution. Despite this marked similarity, GGT1 is bound to the cell wall and expressed in all organs (and most intensively at the conductive element level), whereas GGT2 is likely to be bound to the plasma membrane, and it is expressed mainly in the siliques, where it accounts for 50% of all GGT activity (Martin et al., 2007).

GGT3 is considered a pseudogene because it has an unusually truncated sequence. GGT4 is expressed in vacuoles, where it contributes to the detoxification of xenobiotics following their conjugation with GSH by glutathione sulphotransferases (Grzam et al., 2007).

GGT1 knockout mutant prooplasts of *A. thaliana* are unable to retrieve GSH from the growth medium (Ohkama-Otsu et al., 2007a), but *Arabidopsis* plants lacking the functional GGT1 isoform (which was believed to be the only apoplastic GGT expressed in roots) can survive when grown in a nutrient solution supplemented with GSH as the sole source of sulphur (Martin et al., 2007). The ability of ggt1 plants to grow in media containing GSH, combined with the fact that ggt1 mutants exhibit no striking phenotype, has led to GGT1 being assigned a minor role in the recovery of extracellular GSH. Given a presumed functional redundancy in GSH retrieval mechanisms, it was concluded that extracellular GGT is dispensable, and it was argued that other mechanisms unrelated to GGT activity may be responsible for GSH uptake in ggt1 mutants. Another study found, however, that barley seedlings retrieved GSH from an external medium, but this function was severely impaired when serine/borate was added to inhibit GGT activity (Ferretti et al., 2009), suggesting that GSH recovery from an external medium relies on GGT activity.

Investigations were therefore carried out to determine whether the apoplastic, membrane-bound GGT2 isoform, normally strongly expressed in *Arabidopsis* seeds, might also be expressed in the roots to compensate for the loss of GGT1 function. To test this hypothesis, gene expression and enzyme histochemical analyses, in vivo and in vitro GGT activity measurements, and GSH uptake timing measurements were conducted in wild-type and ggt1 knockout mutant plants.

### Materials and methods

**Plant material and growth conditions**

After 4 d of stratification at 4 °C in the dark, *A. thaliana* L. ecotype Columbia (Col-0) plants were grown hydroponically on rock wool, as described in Huttner and Bar-Zvi (2003), in controlled-environment chambers (10/14 h light/dark cycle at a temperature of 22/18 °C, with 130 μmol m⁻² s⁻¹ photosynthetically active radiation, 70% relative humidity). The nutrient solution used was a modified Gibeaut medium (Gibeaut et al., 1997) containing: 1.5 mM Ca(NO₃)₂; 1.25 mM KNO₃; 0.75 mM Mg(SO₄); 0.5 mM KH₂PO₄; 0.1 mM FeNaEDTA, fresh; 50 μM KCl; 10 μM MnSO₄; 1.5 μM CuSO₄; 2 μM ZnSO₄; 50 μM H₃BO₃; 0.075 mM (NH₄)₆Mo₇O₂₄·4H₂O.

Roots, fully grown leaves, and seeds from untreated wild-type and ggt1 knockout mutant plants were harvested for gene expression and enzyme histochemical analyses. Plants of both genotypes at the fully expanded rosette stage, ~1 week before flowering, were used to monitor the time course of GSH depletion from the incubation medium, and for in vitro and in vivo GGT activity measurements.

The Arabidopsis Genome Initiative locus identifiers for the four members of the GGT gene family are as follows: GGT1 (At4g39640), GGT2 (At4g39650), GGT3 (At1G69820), and GGT4 (At4g29210). The ggt1 knockout mutant line (ecotype Columbia) was found in the mutant collection (Alonso et al., 2003) established at the Salk Institute and available from the Nottingham Arabidopsis Stock Centre (http://nasc.nott.ac.uk; polymorphism SALK_080363). This line was used and characterized in a previous study, where it was named ggt1-2 (Martin et al., 2007). The homoygous plants containing the T-DNA insert located in the second exon of the GGT1 gene were screened by PCR using the gene-specific primers GGT1-F6 (5'-AATTTTGT-TTTCGGGAAAGTTCGAG-3') and GGT1-R6 (5'-CGCTACTGATAAGGCTCCTTTCTT-3'), and the T-DNA left border primer (5'-GGCAATCAGTGTTCGGCCGTCT-CACT-3'). T-DNA integration was confirmed by sequencing the amplification products of the T-DNA flanking region, according to Sanger et al. (1977) at the CRIBI, University of Padua, using the ABI PRISM Big Dye Terminator (Perkin Elmer) Kit.

**Preparation of total RNA and cDNA synthesis**

Total RNA was extracted from *Arabidopsis* plant tissues using the RNeasy Plant Mini Kit™ (Qiagen, Valencia, CA, USA), followed by a DNA removal step with the QIAGEN RNase-Free DNase Set™ according to the manufacturer’s instructions.

The first-strand cDNA was synthesized from 2 μg of total RNA using the SuperScript™ III First-Strand Synthesis System (Invitrogen Corporation, Carlsbad, CA, USA) with an oligo(dT) primer, according to the manufacturer’s protocol.

**cDNA cloning and sequencing**

To confirm its identity, the GGT3 amplification product obtained was subcloned in the pGEM-T Easy Vector (Promega, Milano, Italy), and the plasmids from six recombinant colonies were sequenced. Plasmids were prepared using the QIAprep Spin Miniprep Kit (Qiagen, Hilden, Germany) and sequenced according to Sanger (1977). Sequences were compared using the BLASTn computer program (NCBI, National Center for Biotechnology Information).

**Semi-quantitative RT-PCR analysis**

The semi-quantitative RT-PCR analysis was conducted using the primer sequences described in Table 1.

For all PCR amplification reactions, 1 μl of the cDNA obtained was used in a 20 μl final volume, using 1.0 U of Platinum® Taq DNA Polymerase (Invitrogen Corporation). Different numbers of cycles (from 24 to 40) were tested to choose those corresponding to 50% of all GGT activity (Martin et al., 2007).

| Gene   | Gene locus | Primer name | Primer sequence |
|--------|------------|-------------|-----------------|
| GGT1   | At4g39640  | GGT1F       | GGTCTCTGTTATCGGATGGCG |
|        |            | GGT1R       | TTGGTCGCCGAAATATTGTGG |
| GGT2   | At4g39650  | GGT2F       | CCGTAAACCAGAAATTGCGTAT |
|        |            | GGT2R       | TTGGTACCGGCAGTGATGACA |
| GGT3   | At1g69820  | GGT3F       | GGCTCTTCGCGGTTCTAGCATT |
|        |            | GGT3R       | TGCCGGTGTTGGCCAGACT |
| GGT4   | At4g29210  | GGT4F       | CCAAATGCTCAGGAAGTGCA |
|        |            | GGT4R       | ACCCCATATGCTGACCGATG |
| ACT1   | At3g12110  | ACT1F       | CTGGAGATGTGACCAGAAG |
|        |            | ACT1R       | CCTCATCACCACAGTAAG |

**Table 1. Oligonucleotide primer sequences used in RT-PCRs**
the exponential phase for each gene, then all semi-quantitative PCR analyses were performed for 35 cycles. Reactions were carried out with the Gene Amp PCR system 9700 (PE Biosystems, Branchburg, NJ, USA) starting with an initial denaturation step at 94 °C for 5 min, then cycling at 94 °C for 30 s, 59 °C for 30 s; these steps were followed by a final incubation at 72 °C for 7 min. The A. thaliana β-actin gene (At3g21120) was used as an endogenous internal standard to normalize the gene expression results obtained. A 20 μl aliquot of the PCR products was resolved by electrophoresis on gels stained with ethidium bromide, then scanned and quantified using the Image J64 software (National Institute of Health, Bethesda, MD, USA). To confirm the results of the expression analysis, PCRs were carried out on cDNAs obtained from two different RNA extractions from plant tissues in two independent experiments and repeated three times for each cDNA.

**Exogenous glutathione supplementation**

For the depletion study, hydropically grown wild-type and ggt1 Arabidopsis plants were placed in test tubes containing 5 ml of 100 μM GSH, with or without 10 mM serine/borate as the competitive GGT inhibitor. A 50 μl aliquot of the solution was then drawn off after 0, 1, 3, 6, and 8 h of incubation and stored at −80 °C for several days before thiol chromatographic analysis. For in vivo GGT activity, plants were incubated with 100 μM GSH and harvested after 3 h and 6 h of incubation; single plant roots were transferred to a 5 ml tube containing 4.4 ml of bioassay solution.

**HPLC and thiol measurement**

Following reduction with tributylphosphine, 50 μl of growth medium was derivatized with the SBD-F (ammonium 7-fluoro 2,1,3-benzoazadiazole-4-sulphonate) fluorophore; low molecular weight thiol-SBD derivatives were then separated by chromatography according to the method described in Masi et al. (2002). Samples were collected from at least five distinct biological replicates.

**Enzyme histochemical analysis of GGT activity**

Leaves, roots, and seeds from A. thaliana were embedded in optimal cutting temperature medium (OCT, Cellpath, Newtown, UK) with no prior chemical fixation, then placed in 15 mm cryomoulds, snap-frozen, and stored at −80 °C until use. Sections were collected on glass slides with an adhesive coating and air dried overnight. Following immersion in cold pure acetone for 15 s and rinsing for 30 s in 50% acetone solution, the slides were stained essentially according to the procedure described by Rutenburg et al. (1969). After washing in distilled water, GGT activity was identified by placing the tissues in a freshly prepared staining solution containing 0.46 mM γ-glutamyl-4-methoxy-2-naphthylamide as a substrate, and 6.3 mM glycyl-glycine (gly-gly) as a γ-glutamyl acceptor, in phosphate buffer pH 7.4. Diazotization was done with the diazonium salt Fast Garnet GBC (0.35 mg ml⁻¹ incubation medium), extending the reaction for 90 min at room temperature, with gentle shaking. A control stain was obtained by incubating the tissue sections in a medium containing 10 mM serine/borate (Tate and Meister, 1978).

Enzyme histochemical analysis was also performed in wild-type and ggt1 leaves after incubating them overnight in pure acetone to remove chlorophyll and wounding them with a razor blade to enable infiltration of the incubation medium.

All images were acquired with a Leica DM4000B digital microscope, equipped with an Leica DCF300F camera and Leica Image Manager 50 software (Leica Microsystems).

**Enzyme extraction and in vitro GGT activity assay**

The sequential extraction procedure adopted for this study was based on the one used in Martin et al. (2007), with some modifications. A 1 g aliquot of root tissue from both genotypes was frozen in liquid nitrogen, pulverized in an ice-cold mortar, and homogenized in 2 ml of extraction buffer (20 mM TRIS-HCl pH 8.0, protease inhibitor cocktail, 1% Triton X-100). The homogenate was kept at 4 °C for 60 min, with gentle shaking, then centrifuged at 15 000 g for 20 min at 4 °C. The supernatant containing the soluble and membrane-bound protein fraction was collected and assayed for GGT activity.

The pellet was rinsed twice with the same buffer, then incubated with 20 mM TRIS-HCl pH 8.0 buffer containing 3 M NaCl for 60 min to solubilize the proteins ionically bound to the cell wall. It was found that 3 M NaCl enabled an ~30% higher recovery than 1 M NaCl (data not shown). This fraction was then assayed for GGT activity. The fractions obtained by rinsing the pellets were also tested, but they generally revealed a negligible residual activity, if any (data not shown).

GGT activity was assayed according to Huseby and Strömme (1974), involving the spectrophotometric measurement of absorbance at 407 nm of p-nitroaniline released from the γ-glutamyl-p-nitroanilide (GPNA) substrate.

The reaction was started by adding 0.2 ml of enzyme extract to 1 ml of solution A (4.6 mM GPNA in 100 mM TRIS-HCl pH 8.0, 10 mM MgCl₂) and 0.1 ml of solution B (575 mM gly-gly; the pH was adjusted to 8.0 by adding NaOH). The change in absorbance at 40 nm in the assay medium was recorded for 10 min.

**In vivo GGT activity measurements**

GGT activity in roots was measured in vivo, as described elsewhere (Ferretti et al., 2009). The method is essentially an adaptation of the procedure used to measure in vitro activity (Huseby and Strömme, 1974), and is based on the observation that the root apoplastic space is readily accessed by the external solution.

Single plant roots were placed in a test tube containing 4 ml of solution A (4.6 mM GPNA in 100 mM NaH₂PO₄ pH 8.0) and 0.4 ml of solution B (575 mM gly-gly in 100 mM NaH₂PO₄ pH 8.0); the solution was cycled through home-made piping from the cuvette to the spectrophotometer cell using a peristaltic pump operated at a flow rate of 3 ml min⁻¹. The change in absorbance at 407 nm induced by releasing p-nitroaniline into the assay medium was recorded for 10 min.

**Statistical analysis**

Data were submitted to analysis of variance (ANOVA), and the Tukey honest significant difference (HSD) multiple comparisons procedure was used to discriminate between means. A significance level of P < 0.05 was adopted for all comparisons. The Statgraphics Plus rel. 5.0 Software package (Manugistics Inc., Rockville, MD, USA) was used for the statistical analysis.

**Results**

The comparison between the hydropically grown A. thaliana wild-type and ggt1 knockout mutants showed that the latter exhibited no clearly visible phenotype, but a progressive senescence, demonstrated by a shorter time to flowering, which took ~4 d less (Fig. 1).

**GGT gene expression in Arabidopsis thaliana**

The relative expression of the known GGT isoforms in A. thaliana was investigated by semi-quantitative PCR analysis in different tissues (roots, leaves, and seeds).

The expression data in Fig. 2 indicate that GGT1 is expressed in all tissues, albeit with a different intensity (leaves > roots > seeds). GGT3 is only detected in seeds. GGT4 is detected in all tissues, but considerably more in roots.
GGT2 transcripts give rise to a very faint band in roots at 30 PCR cycles, but a clear band could be seen after increasing to 40 cycles. GGT3 transcripts yielded a clear band in siliques and very faint bands in leaves and roots. The authenticity of the GGT3 transcripts was confirmed by sequencing the 396 bp amplification product, which was 100% identical to the nucleotide sequence of At1g69820. No GGT2 expression was ever seen in leaves, not even at 40 cycles.

**Enzyme histochemical localization of GGT activity**

Figure 3 shows the enzyme histochemical detection of GGT activity in different organs and tissues of *A. thaliana* wild-type plants. Red to brownish staining results from the diazotization reaction of the unstable diazonium salt Fast Garnet GBC with p-nitroaniline released from GPNA by GGT activity. Staining was seen at the root apex (Fig. 3A), in conductive tissues closely associated with vascular bundles (Fig. 3B), and in the epidermal cell layer (Fig. 3E); in seeds, intense staining was apparent in the endosperm (Fig. 3C, D). The specificity of staining was demonstrated by using 10 mM serine/borate to inhibit GGT activity (Fig. 3F).

Enzyme histochemical analysis was also performed to compare wild-type and *ggt1* knockout mutants. As shown in Fig. 4A and C, root sections only exhibited staining in the vascular tissue in wild-type plants. Whole leaves (Fig. 4B, D) were analysed by wounding the leaves with a razor blade to enable infiltration of the incubation medium. Red staining was seen around the wounded region in wild-type, but not *ggt1* knockout leaves.

**GSH depletion time course analysis**

The ability of wild-type and *ggt1* roots to retrieve exogenous GSH is illustrated in Fig. 5. The wild-type *Arabidopsis* roots depleted a solution containing 100 μM GSH more quickly than the *ggt1* knockout mutants; adding the serine/borate inhibitor almost completely prevented the uptake process (Fig. 5A). GSH depletion from the growth medium was paralleled by an increase in cysteinyl-glycine and cysteine (Fig. 5B, C, respectively). This increase was lower in *ggt1* mutants compared with the wild type, and almost completely abolished in the presence of the serine/borate inhibitor.

**In vivo measurements of apoplastic GGT activity**

To test the hypothesis that GGT activity is partially restored in *ggt1* knockout mutants, the roots of intact plants were incubated with a GPNA solution and the rate of p-nitroaniline release was measured after 3 h and 6 h of incubation; this procedure was described in a previous study on barley seedlings (Ferretti *et al.*, 2009).

Figure 6 shows the GGT activity induced in *ggt1* roots; it was hardly detectable in vivo under normal conditions, but it increased noticeably after 3 h and 6 h of incubation with GSH. GGT activity in the wild-type roots was unaffected after 3 h and 6 h of incubation (not shown).

GGT activity in roots from wild-type and *ggt1* knockout plants was also measured after 24 h of incubation with 100 μM GSH, adopting a sequential extraction and using Triton X-100 and 3 M NaCl in order to discriminate between membrane-bound and soluble GGT activity from cell wall-bound activity. In the Triton X-100 fraction, GGT activity was similar in the wild-type and *ggt1* root extracts,
but 24 h of treatment with 100 μM GSH prompted a 3- to 4-fold increase in GGT activity by comparison with untreated plants. In wild-type extracts, GGT activity was much higher in the fractions obtained by 3 M NaCl extraction than in those obtained using Triton X-100; it was almost undetectable in the ggt1 extracts.

Semi-quantitative gene expression analysis following GSH supplementation

Based on the finding that GGT activity was partially restored in ggt1 roots after incubation with GSH, a gene expression analysis of the four ggt genes was performed in wild-type and ggt1 roots and shoots after GSH supplementation in the growth medium (Fig. 7).

As expected, inserting a T-DNA sequence in the mutant genotype results in the early truncation of the reading frame and complete loss of GGT1 transcripts. Incubation with 100 μM GSH led to a transient GGT1 stimulation in wild-type roots, observable after 1 h. GGT2 was constitutively up-regulated in ggt1 roots, and was stimulated by GSH supplementation in the wild type. No GGT2 transcripts were ever detected in leaves. Some stimulation of GGT3 and GGT4 expression was generally observed in roots, but the differences were not statistically significant.
Discussion

Histochemical detection of GGT activity in A. thaliana

The histochemical detection of GGT activity is an effective tool for investigating GGTs in their active form and in
different organisms. Here, enzyme histochemical analyses were used to compare wild-type and ggt1 Arabidopsis tissues (Figs 3, 4) to complement the findings of published reports on gene expression analyses. Consistently with previous observations in barley (Ferretti et al., 2009) and maize organs and tissues (Masi et al., 2007), Arabidopsis wild-type leaves exhibited intense staining in the vascular tissue, epidermis, and stomata, while virtually no staining was apparent in the ggt1 leaves (Fig. 4), confirming that GGT1 is the isoform most expressed in leaves, where GGT2 is not expressed at all. Consistently with previous observations in maize (Masi et al., 2007) and barley (Ferretti et al., 2009) roots, enzyme histochemical detection revealed a strong GGT activity in Arabidopsis wild-type root tips, but very little in ggt1 knockout roots (Fig. 4A, C). Overall, the histochemical analysis demonstrated that GGT activity correlates with the spatial distribution of GGT transcripts, as previously reported (Martin et al., 2007), suggesting that GGT activity is indeed regulated at the gene expression level—although post-translational changes, which are known to be induced by GGTs in animal cells, cannot be ruled out.

Regulation of GGT2 expression in A. thaliana roots by GSH availability and lack of functional GGT1

T-DNA insertion results in the disruption of the functional GGT1 gene in Arabidopsis, but GSH uptake is still possible in ggt1 mutants, albeit at a slower rate than in wild-type plants. Adding serine/borate (a competitive GGT inhibitor) significantly hampers this process, however (Fig. 5A). The present findings indicate that another protein with GGT activity (other than GGT1) sustains GSH degradation, as shown by the parallel rise in cysteinyl-glycine content in ggt1 plants. GGT3 (which is considered a pseudogene) is unlikely to contribute any activity and GGT4 is restricted to the vacuole, where it assists in the degradation of GSH conjugates (Grzam et al., 2007). It is worth mentioning that GGT3 transcripts were detected in this work (and verified by sequencing), in contrast to the report from Ohkama-Otsu et al. (2007a), but in accordance with the findings of Martin et al. (2007), who observed β-glucuronidase (GUS) activity in lines carrying the GUS:GGT3 fusion construct. Whether a functional protein originates from GGT3 transcripts is not known, but it would seem unlikely because it lacks a sequence that is important in conferring catalytic activity (Okada et al., 2007).

Here, it is experimentally demonstrated that GGT2 is also expressed in roots, and it is up-regulated both when the GGT1 gene is disrupted and in the presence of extracellular GSH. A transient up-regulation of GGT1 expression (observable after 1 h of incubation; see Fig. 7C) in wild-type roots was also induced by GSH supplementation, suggesting that the two genes might be similarly regulated by this stimulus.

The extracellular location of GGT1 and GGT2 activity can be exploited to measure root GGT activity in vivo by monitoring the release of p-nitroaniline from its γ-glutamyl
precursor, GPNA (Ferretti et al., 2009). This approach is effective in demonstrating large differences in GGT activity in *A. thaliana* wild-type versus *ggt1* roots. Incubation with 100 μM GSH restored in vivo GGT to nearly 50% by comparison with the wild-type after 6 h (Fig. 6), and this finding is consistent with the gene expression data in Fig. 7 and the data in Table 2, indicating a greater GGT activity in the membrane-bound protein fraction after GSH supplementation.

These findings suggest that GGT2 expression at least partially compensates for the loss of GGT1, and that it contributes to the retrieval of extracellular GSH, which also acts as a positive signal for GGT gene expression. The inducibility of GGT genes by GSH is also supported by some data available in the literature: high levels of GGT1, GGT2, and GGT3 expression were observed in natural conditions in developing trichomes (Martin et al., 2007), where the GSH content is nearly 300 times higher than in the epidermal layer of the leaves (Gutiérrez-Alcalá et al., 2000).

Compensatory GGT2 up-regulation occurs in roots, but not in leaves, where GGT2 transcripts were never detected in any of the present cases (Fig. 7), a finding consistent with data available in the literature. The lack of the GGT2 isoform in leaves when GGT1 is also missing suggests that a γ-glutamyl cycle for the retrieval of extracellular GSH might be unnecessary in leaves, which can rely on their intracellular biosynthetic machinery to control GSH availability. The GGT2 expression observed in this work is also consistent with the weak GUS expression already reported in roots of *Arabidopsis* lines carrying the GUS:GGT2 construct (Martin et al., 2007).

The contribution of GGT2 (known to be strongly expressed in silique) to total GGT activity in roots may be low, but not negligible, and it may increase through substrate availability. GGT1 and GGT4 are therefore not the only functional GGTs in roots. It follows that, despite major similarities between GGT1 and GGT2 in terms of gene sequences and apoplastic location, the two GGT isoforms exhibit some significant differences. For a start, they are associated with a different cell structure, since GGT1 is ionically bound to the cell wall, whereas GGT2 seems to be associated with membranes. They exhibit a different spatial and temporal expression; and their expression is differently regulated. GGT2 being inducible in roots by GSH, its natural substrate, whilst GGT1 expression among tissues is rather constitutive (although some inducibility by GSH has been observed in this work) and linked to developmental stage. The higher GGT2 expression in *ggt1* roots under basal conditions might also indirectly result from the increase in GSH content in the extracellular space.

The possible contribution of GGT2 activity to the recovery of extracellular GSH in *ggt1* mutants might explain the ability of the latter (see Martin et al., 2007), or *ggt1*50Pase, or *ggt1*ggg4 double mutants and *ggt1*ggg4/50Pase triple mutants to grow in media containing GSH as the sole source of sulphur (Ohkama-Ohtsu et al., 2008).

Taken together, these observations suggest that GGT1 and GGT2 isoforms cooperate in GSH recovery from the extracellular space in *A. thaliana* roots. Although additional mechanisms for GSH recovery may be at work in different plant organs and at different developmental stages, in the present experimental conditions GGTs seem to have a major role in the retrieval of extracellular GSH.

This means that the existence of apoplastic GGTs in roots should be taken into due account in GSH feeding experiments. Previous studies (e.g. Lappartient et al., 1999; Bashandy et al., 2010) should therefore be reconsidered bearing in mind that external GSH can be effectively degraded extracellularly and resynthesized starting from the constituent amino acids after their recovery by means of amino acid transporters.

### GGT and GSH degradation pathways

The role of the apoplastic GGTs, GGT1 and GGT2, remains an open question. It is tempting to suggest that they could participate in GSH retrieval and inter-organ redistribution, as in animal organisms, but this function is harder to assess in plants. In leaves, where GGT2 is never expressed under any conditions or at any developmental stage, even GGT1 silencing does not result in any visible phenotype. The glutathione level in these organs can be maintained, however, by the cysteine ultimately provided by photosynthesis. The effects in leaves might therefore be minimized by an autonomous capacity for GSH biosynthesis and supply.

In organs other than leaves, such as siliques and roots, where both GGT1 and GGT2 are expressed, it is likewise

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### Table 2. GGT activity in wild-type and ggt1 knockout root extracts with and without treatment with 100 μM GSH

| Treatment | Extraction medium | GGT activity (nmol p-nitroaniline g⁻¹ FW min⁻¹) | Wild type | ggt1 |
|-----------|-------------------|---------------------------------------------|-----------|------|
| Control   | + Triton X-100    | 44.22±8.70                                 | 34.19±5.44|
| + 100 μM GSH | + Triton X-100   | 151.28±38.89                              | 121.69±29.86|
| Control   | + 3 M NaCl        | 297.69±77.77                               | 7.23±1.48 |
| + 100 μM GSH | + 3 M NaCl      | 335.65±41.76                              | 6.91±4.64 |

Roots were incubated in 100 μM GSH medium and harvested after 24 h. GGT activity was measured in vitro on different fractions obtained from wild-type and *ggt1* roots with and without GSH treatment using the following sequential extraction procedure: roots were homogenized with Triton X-100 containing TRIS buffer pH 8, enabling the extraction of both soluble and membrane-bound proteins. After centrifugation, the supernatant was collected and the pellet was incubated twice with the same buffer to eliminate any residual activity completely. Then the pellet was incubated with 3 M NaCl containing TRIS buffer to solubilize proteins bound to the cell wall. Activity was measured as the release of p-nitroaniline from the artificial γ-glutamyl-p-nitroanilide substrate by monitoring the change in absorbance at 407 nm of the assay solution at 10 min intervals. The values given in the table are the means of four independent experiments. See Materials and methods for additional details.
impossible for the time being to draw any definitive conclusions. When the major GGT1 isoform is lacking in roots, GGT2 can compensate for the loss and ensure some activity, which is probably sufficient to minimize the effects of the mutation. In siliques, both isoforms are equally expressed, and silencing of either isoform is compensated for by the other.

A conclusive analysis on the role of extracellular GGT might be possible on ggt1/ggt2 double mutants, but, because the two genes are consecutive, the chances of obtaining a double mutant by conventional cross-breeding are virtually nil. Different strategies should therefore be explored (e.g. multiple gene silencing by RNA interference) in order to strongly reduce the expression of both isoforms.

At this stage, it is impossible to establish whether apoplastic GGTs are just one of a number of systems, or the most important one that plant cells use to retrieve extracellular GSH. It may also be that GGTs have as yet unknown functions that need to be clarified. In the extracellular space, where oxidizing conditions dominate, a γ-glutamyl cycle may be functional to some kind of apoplastic redox control and/or to transfer redox signals from the outside world to the cytosol. The only visible phenotype observed in ggt1 plants (i.e. a shorter life cycle) may actually be interpreted as an overall difficulty in the plant’s adaptation to the environment.

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References
Alonso JM, Stepanova AN, Leisse TJ, et al. 2003. Genomewide insertional mutagenesis of Arabidopsis thaliana. Science 301, 653–657.

Bashandy T, Guilleminot J, Vernoux T, Caparros-Ruiz D, Ljung K, Meyer Y, Reichheld JP. 2010. Interplay between the NADP-linked thioredoxin and glutathione systems in Arabidopsis auxin signaling. The Plant Cell 22, 376–391.

Beemster G, Fiorani F, Inzé D. 2003. Cell cycle: the key to plant growth control? Trends in Plant Science 8, 154–158.

Dixon DP, Cummins I, Cole DJ, Edwards R. 1998. Glutathione-mediated detoxification systems in plants. Current Opinion in Plant Biology 1, 258–266.

Edwards R, Dixon DP, Walbot V. 2000. Plant glutathione S-transferases: enzymes with multiple functions in sickness and in health. Trends in Plant Science 5, 193–198.

Ferretti M, Destro T, Tosatto SC, La Rocca N, Rascio N, Masi A. 2009. Gamma-glutamyl transferase in the cell wall participates in extracellular glutathione salvage from the root apoplast. New Phytologist 181, 115–126.

Foyer CH, Noctor G. 2001. The molecular biology and metabolism of glutathione. In: Grill D, Tausz M, de Kok L, eds. Significance of glutathione in plant adaptation to the environment. Dordrecht: Kluwer, 27–56.

Foyer CH, Noctor G. 2009. Redox regulation in photosynthetic organisms: signaling, acclimation, and practical implications. Antioxidants and Redox Signaling 11, 861–905.

Gibeaut DM, Hulett J, Cramer GR, Seemann JR, Romero LC. 2000. Glutathione biosynthesis in Arabidopsis trichome cells. Proceedings of the National Academy of Sciences, USA 97, 11108–11113.

Herschbach C, Rennenberg H. 1995. Long-distance transport of 35S-sulfur in 3-year-old beech trees (Fagus sylvatica). Physiologia Plantarum 95, 379–386.

Herschbach C, van Der Zalm E, Schneider A, Jouanin L, De Kok LJ, Rennenberg H. 2000. Regulation of sulfur nutrition in wild-type and transgenic poplar over-expressing gamma-glutamylcysteine synthetase in the cytosol as affected by atmospheric H2S. Plant Physiology 124, 461–473.

Huseby NE, Strömme JH. 1974. Practical points regarding routine determination of gamma-glutamyl transferase (gamma-GT) in serum with a kinetic method at 37°C. Scandinavian Journal of Clinical Laboratory Investigation 34, 357–361.

Huttner D, Bar-Zvi D. 2003. An improved, simple, hydroponic method for growing Arabidopsis thaliana. Plant Molecular Biology Reporter 21, 59–63.

Lappartient AG, Vidmar JJ, Leustek T, Glass AD, Touraine B. 1999. Inter-organ signaling in plants: regulation of ATP sulfurylase and sulfate transporter genes expression in roots mediated by phloem-translocated compound. The Plant Journal 18, 89–95.

Marrs KA. 1996. The functions and regulation of glutathione S-transferases in plants. Annual Review of Plant Physiology and Plant Molecular Biology 47, 127–158.

Martin MN, Saladores PH, Lambert E, Hudson AO, Leustek T. 2007. Localization of members of the γ-glutamyl transpeptidase family identifies sites of glutathione and glutathione S-conjugate hydrolysis. Plant Physiology 144, 1715–1732.

Martin MN, Slovin JP. 2000. Purified γ-glutamyl transpeptidases from tomato exhibit high affinity for glutathione and glutathione S-conjugates. Plant Physiology 122, 1417–1426.

Masi A, Destro T, Turetta L, Varotto S, Caporale G, Ferretti M. 2007. Localization of gamma-glutamyl transferase activity and protein...
in Zea mays organs and tissues. *Journal of Plant Physiology* **164**, 1527–1535.

**Masi A, Ghisi R, Ferretti M.** 2002. Measuring low-molecular-weight thiols by detecting the fluorescence of their SBD derivatives: application to studies of diurnal and UV-B induced changes in *Zea mays* L. *Journal of Plant Physiology* **159**, 499–507.

**May MJ, Vernoux T, Leaver C, Van Montagu M, Inzé D.** 1998. Glutathione homeostasis in plants: implications for environmental sensing and plant development. *Journal of Experimental Botany* **49**, 649–667.

**Meyer A.** 2008. The integration of glutathione homeostasis and redox signaling. *Journal of Plant Physiology* **165**, 1390–1403.

**Mullineaux P, Rausch T.** 2005. Glutathione, photosynthesis and the redox regulation of stress-responsive gene expression. *Photosynthesis Research* **86**, 459–474.

**Noctor G, Arisi A-CM, Jouanin L, Kunert KJ, Rennenberg H, Foyer CH.** 1998. Glutathione: biosynthesis, metabolism and relationship to stress tolerance explored in transformed plants. *Journal of Experimental Botany* **49**, 623–647.

**Ohkama-Ohtsu N, Oikawa A, Zhao P, Xiang C, Saito K, Oliver DJ.** 2008. A γ-glutamyl transpeptidase-independent pathway of glutathione catabolism to glutamate via 5-oxoproline in *Arabidopsis*. *Plant Physiology* **148**, 1603–1613.

**Ohkama-Ohtsu N, Radwan S, Peterson A, Zhao P, Badr A, Xiang C, Oliver D.** 2007a. Characterization of the extracellular γ-glutamyl transpeptidases, GGT1 and GGT2 in *Arabidopsis*. *The Plant Journal* **49**, 865–877.

**Ohkama-Ohtsu N, Zhao P, Xiang C, Oliver DJ.** 2007b. Glutathione-conjugates in the vacuole are degraded by γ-glutamyl transpeptidase GGT3 in *Arabidopsis*. *The Plant Journal* **49**, 878–888.

**Okada T, Suzuki H, Wada K, Kumagai H, Fukuyama K.** 2007. Crystal structure of the gamma-glutamyltranspeptidase precursor protein from *Escherichia coli*. Structural changes upon autocatalytic processing and implications for the maturation mechanism. *Journal of Biological Chemistry* **282**, 2433–2439.

**Rutenburg AM, Kim H, Fischbein JW, Hanker JS, Wasserkrug HL, Seligman AM.** 1969. Histochemical and ultrastructural demonstration of gamma-glutamyl transpeptidase activity. *Journal of Histochemistry and Cytochemistry* **17**, 517–526.

**Sanger F, Nicklen S, Coulson AR.** 1977. DNA sequence with chain-terminating inhibitors. *Proceedings of the National Academy of Sciences, USA* **74**, 5463–5467.

**Storozhenko S, Belles-Boix E, Babiychuk E, Herouart D, Davey MW, Slooten L, Van Montagu M, Inzé D, Kushnir S.** 2002. γ-Glutamyl transpeptidase in transgenic tobacco plants: cellular localization, processing, and biochemical properties. *Plant Physiology* **128**, 1109–1119.

**Szalai G, Kelló T, Galiba G, Kocsy G.** 2009. Glutathione as an antioxidant and regulatory molecule in plants under abiotic stress conditions. *Journal of Plant Growth Regulation* **28**, 66–80.

**Tate SS, Meister A.** 1978. Serine–borate complex as a transition state inhibitor of γ-glutamyl transpeptidase. *Proceedings of the National Academy of Sciences, USA* **75**, 4806–4809.

**Tausz M, Sircelj H, Grill D.** 2004. The glutathione system as a stress marker in plant ecophysiology: is a stress-response concept valid? *Journal of Experimental Botany* **55**, 1955–1962.

**Vernoux T, Wilson RC, Seeley KA, et al.** 2000. The ROOT MERISTEMLESS1/Cadmium Sensitive2 gene defines a glutathione-dependent pathway involved in initiation and maintenance of cell division during postembryonic root development. *The Plant Cell* **12**, 97–109.