Enhanced muscle pump during mild dynamic leg exercise inhibits sympathetic vasomotor outflow

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Abstract

Muscle sympathetic nerve activity (MSNA) is not increased during leg cycling at light and mild intensities, despite activation of central command and the exercise pressor reflex. We determined whether increasing central blood volume and loading the cardiopulmonary baroreceptors modulate sympathetic vasomotor outflow during leg cycling. To this end, we changed the pedaling frequency to enhance skeletal muscle pump. Subjects performed two leg cycle exercises at differential pedal rates of 60 and 80 rpm (60EX and 80EX trials) for two conditions (with and without MSNA measurement). In each trial, subjects completed leg cycling with a differential workload to maintain constant oxygen consumption (VO2). MSNA was recorded via microneurography at the right median nerve of the elbow. Without MSNA measurement, thoracic impedance, stroke volume (SV), and cardiac output (CO) were measured non-invasively using impedance cardiography. Heart rate and VO2 during exercise did not differ between the 60EX and 80EX trials. Changes in thoracic impedance, SV, and CO during the 80EX trial were greater than during the 60EX trial. MSNA during the 60EX trial was unchanged compared with that at rest (25.8 ± 3.1 [rest] to 28.3 ± 3.4 [exercise] bursts/min), whereas a significant decrease in MSNA was observed during the 80EX trial (25.8 ± 2.8 [rest] to 19.7 ± 2.0 [exercise] bursts/min). These results suggest that a muscle pump-induced increase in central blood volume, and thereby loading of cardiopulmonary baroreceptors, could inhibit sympathetic vasomotor outflow during mild dynamic leg exercise, despite activation of central command and the exercise pressor reflex.

Keywords
cardiopulmonary baroreceptors, dynamic leg exercise, sympathetic activity.

Introduction

Arterial blood pressure (BP) is regulated via sympathetic vasomotor outflow during dynamic exercise to maintain appropriate blood flow to active muscles. Several neural mechanisms, including the central command and the exercise pressor reflex (mechano- and metaboreflex within skeletal muscle), play major roles in this context, affecting the arterial baroreflex (Fadel and Raven 2012; Raven and Chapleau 2014). It is well-established that activations of these two physiological factors determine sympathetic vasomotor outflow (Saito et al. 1991; Ray and Mark 1993; Ettinger et al. 1996; Raven 2008). Indeed, muscle sympathetic nerve activity (MSNA) increases in proportion to exercise intensity when the exercise is static and performed using small muscle mass (i.e., a static handgrip) (Saito et al. 1986). However, when exercise is dynamic with large muscle mass (i.e., leg cycling), MSNA decreases or does not change from that at rest if the exercise is of light or mild intensity, and gradually rises thereafter in proportion to the increase in workload (Saito et al. 1993; Callister et al. 1994; Ichinose et al. 2008). These results suggest that changes in MSNA are not limited by activations of the central command and the exercise pressor reflex when dynamic leg exercise that is light or mild in intensity is performed.

This interesting phenomenon may be linked to loading of cardiopulmonary baroreceptors, attributable to muscle pump-induced increases in venous return or central...
blood volume (Mack et al. 1988; Ray et al. 1993; Ray and Saito 1999; Fadel and Raven 2012; Raven and Chapleau 2014). The cardiopulmonary baroreflex during dynamic exercise has been but seldom studied, and most representative work is performed by Ray et al. (1993). The cited authors showed, for the first time, that MSNA was affected by changes in central blood volume mediated by the cardiopulmonary baroreflex during dynamic exercise in the sitting and supine positions. Unfortunately, it remains unclear whether activation of the central command and the exercise pressor reflex was distinguished from the cardiopulmonary baroreflex in their study, because cardiopulmonary responses and the nature of limb muscle activation may differ when exercise are performed in the sitting and supine positions (Stenberg et al. 1967; Egana et al. 2010). Thus, the effect of the cardiopulmonary baroreflex on the modulation of MSNA remains unclear. Ogoh et al. (2007) showed that enhanced muscle pump-induced loading of the cardiopulmonary baroreceptors modified carotid-baroreflex function curve. This suggested that increases in central blood volume associated with the muscle pump might activate the cardiopulmonary baroreceptors and inhibit MSNA (Ray and Saito 1999; Fadel and Raven 2012). Therefore, it was necessary to clarify whether muscle pump-induced increase in central blood volume, thereby loading the cardiopulmonary baroreceptors, affected sympathetic vasomotor outflow, when central command, exercise pressor reflex, and the posture did not change during exercise.

The purpose of the present study was to clarify whether sympathetic vasomotor activity was suppressed by enhanced muscle pump during dynamic leg exercise. MSNA was recorded from the median nerve by using a microneurographic technique during leg cycling with an increase in central blood volume by increasing pedal frequency (e.g., increasing the frequency of muscle contraction) to enhance the skeletal muscle pump (Gotshall et al. 1996; Ogoh et al. 2007). In addition, we maintained constant oxygen consumption at the differential pedal cadence to keep central command and exercise pressor activation similar (Ogoh et al. 2007). We hypothesized that enhanced muscle pump during mild dynamic leg exercise would inhibit in sympathetic vasomotor outflow.

**Methods**

**Ethical approval**

The present study was approved by the human research committee of the Research Center of Health, Physical Fitness and Sports at Nagoya University.

**Subjects**

Eight healthy males were enrolled in the study; of whom six completed the study (means ± standard error [SE]: age = 23.7 ± 0.6 years, height = 176.0 ± 2.3 cm, body mass = 69.4 ± 3.3 kg). Most subjects participated in moderate-intensity exercise a couple of times per week, but none were engaged in high-intensity exercise training. Subjects were informed about the experimental procedures and potential risks involved, and written consent was obtained.

**Measurements**

**Respiratory variables**

Minute expired ventilation (VE), tidal volume (VT), breathing frequency (f), oxygen uptake (VO2), and carbon dioxide output (VCO2) were determined by an online system with a mixing chamber, as in our previous studies (Katayama et al. 2011, 2012, 2013). Subjects breathed through a leak-free nasal mask (5719; Hans Rudolph, Kansas City, MO). Expired gas volume was measured by a Fleisch pneumotachometer (PN-230; Arco System, Chiba, Japan). Sample gas was drawn through a sampling tube connected to the pneumotachometer to measure the expired gas fraction. Gas fractions were analyzed by a mass spectrometer (ARCO-1000; Arco System) that was calibrated and confirmed before each test. Breath-by-breath data were measured continuously using customized software on a computer (PC-9821Ra40; NEC, Tokyo, Japan).

**Cardiovascular variables**

An electrocardiogram (ECG) was measured using a three-lead electrocardiogram (AB-621; Nihon Koden, Tokyo, Japan), and HR was calculated from each R-R interval obtained from the ECG. Thoracic electrical impedance (the ΔZ waveform) and central hemodynamic variables (stroke volume [SV] and cardiac output [CO]) were estimated via non-invasively using the impedance cardiography (PhysioFlow PF-05 Lab1; Manatec Biomedical, Paris, France). The ΔZ value was outputted to be positive in this device, and therefore change in the ΔZ was presented negative numbers by convention. Thoracic impedance was used as an index of central blood volume (CBV) (Ebert et al. 1986; Cai et al. 2000). The reliability of SV and CO measurements obtained using this apparatus has been previously confirmed both at rest and during exercise (Charloux et al. 2000). ECG and thoracic impedance signals were sampled at a frequency of 200 Hz using an analog-to-digital converter (CBI-3133B; Interface, Hiroshima, Japan) and stored in a computer (CF-F8; Panasonic, Osaka, Japan). These variables were averaged over 1-min
both at rest and during exercise. SV was estimated not only by impedance cardiography, but also by beat-to-beat measurements of ascending aortic pulse waves. A Doppler echocardiographic technique employing an ultrasound system was used to this end (LOGIQ 5 PRO; GE-Yokogawa Medical Systems, Tokyo, Japan). The velocity curves were traced offline to obtain average velocity-time integral (VTI). Aortic cross-sectional areas were calculated from the aortic diameters (D values) at the sinotubular junction under resting conditions. SV was calculated as follows: \( SV = \pi \times (D / 2)^2 \times VTI. \) CO was the product of SV and HR. These variables were averaged over the last 30 s of every experimental minute. Arterial systolic and diastolic BP (SBP and DBP) were taken from the left arm using an automated BP unit (STBP-780; Colin Medical Instruments, San Antonio, TX). Automated values determined by the Colin STBP-780 were confirmed by manual readings taken by the experimenter listening through a headset (Katayama et al. 2011). Arterial BP was monitored at 1-min intervals throughout the tests. Mean arterial pressure (MBP) was calculated using the following equation: \( MBP = (SBP – DBP)/3 + DBP. \)

**Muscle sympathetic nerve activity**

Multiunit muscle sympathetic nerve discharges were recorded by the microneurographic technique using a recording system similar to that in our previous studies (Katayama et al. 2011, 2012, 2013). A tungsten microelectrode with a shaft diameter of 0.1 mm (impedance 1–5 MΩ) was inserted manually by an experimenter into the right median nerve at the cubital fossa. The right arm was fixed using equipment to prevent arm movement artifacts during the leg cycling exercise. After insertion, the electrode was adjusted until MSNA was recorded. Identification of MSNA was based on the following criteria: spontaneous burst discharge synchronized with heart rate and enhanced by the Valsalva maneuver or breath holding, but showing no change in response to sensory stimuli, such as a loud noise or cutaneous touch (Delius et al. 1972; Vallbo et al. 1979; Fagius and Wallin 1980; Saito et al. 1993). Moreover, subjects were asked to hold their breath to identify MSNA during exercise (at least 5 sec). The neurogram was fed to a differential amplifier and amplified 100,000 times through a band-pass filter (700–2000 Hz). The neurogram was full-wave rectified and integrated by a capacitance-integrated circuit with a time constant of 0.1 sec. The mean voltage neurogram was continuously digitized through an analog-to-digital converter with a sampling frequency of 200 Hz and stored electronically. MSNA bursts were identified from the mean voltage neurogram using a customized computer program-assisted inspection (Katayama et al. 2011, 2012, 2013), which accounted for the latency from the ECG-R wave to the sympathetic burst (Fagius and Wallin 1980). MSNA was quantified as burst frequency (BF, bursts/min) and incidence (BI, bursts/100 HR) (Saito et al. 1993; Saito et al., 1997; Katayama et al. 2011, 2012). MSNA burst amplitude and total activity could not be calculated because electromyographic, efferent and afferent nerve activities altered the baseline of the integrated neurogram during dynamic leg cycling in most recordings (Saito et al., 1997; Katayama et al. 2011, 2012, 2013).

**Experimental procedures**

All studies were performed at a constant temperature (22–24°C). On day 1, all subjects were familiarized with the equipment to be used in the experiment. Subjects were instructed how to laterally extend both arms and how to hold their arms during leg cycling using an electromechanically braked ergometer in a semirecumbent position (Aerobike 75XL III, Combi, Tokyo, Japan) (Saito et al. 1993; Katayama et al. 2011, 2012, 2013). On day 2, subjects carried out an incremental maximal exercise test using the ergometer. The exercise test began at an initial power output of 90 W, and the workload was increased 15 W per minute until exhaustion (Katayama et al. 2011, 2012, 2013). The pedaling rate was maintained at 60 rpm with the aid of a metronome. VO2 and HR were recorded during the test and were averaged every 30 sec afterward. The highest VO2 value obtained during the exercise protocol was used as peak VO2 (VO2peak). Workload at 40% VO2peak was calculated for submaximal exercise tests at a pedal cadence of 60 rpm. On day 3, two preliminary submaximal exercises were performed to determine workload at a pedal cadence of 80 rpm. The pedaling rate was maintained by means of a metronome. Subjects arrived at the laboratory and rested for 30 min. Subjects first performed submaximal exercise at 40% VO2peak at a pedaling rate of 60 rpm for 7 min, and we confirmed that VO2 attained the steady-state at a target VO2. After 15 min of cessation of the first exercise, subjects performed the second submaximal exercise at a pedal cadence of 80 rpm. The increase in pedal frequency (i.e., muscle contraction frequency) was used to enhance the effect of the muscle pump and increase central blood volume (Gotshall et al. 1996; Ogoh et al. 2007). The second exercise at 80 rpm began at 30% VO2peak at 60 rpm, and the workload at 80 rpm was adjusted to elicit the target VO2 that was obtained during the first submaximal exercise at 60 rpm. Thus, the workload at the same VO2 was lower \( (P < 0.05) \) at 80 rpm (58.5 ± 5.4 watts) than at 60 rpm (78.3 ± 4.9 watts), and these workloads were utilized in the submaximal exercise tests on days 4–6. During exercise at pedaling of 60 and 80 rpm, subjects were instructed again how to
hold their right arm. On day 4, all subjects performed two submaximal exercises during which we measured MSNA (MSNA test). Subjects arrived at the laboratory and rested for 30 min. Respiratory variables, HR, and MSNA were recorded throughout the experiment. The subjects rested for 3 min. Next, the subjects performed 7 min of exercise and rested for 1 min after cessation of exercise. The procedure was repeated twice, i.e., 60 rpm (control, 60EX trial) or 80 rpm (80EX trial) with a 15-min interval between trials. The order of exercise at 60 and 80 rpm was randomly assigned and counterbalanced. MSNA recordings during the 60EX and 80EX trials were successful in five of eight subjects. MSNA recording failed in the other three subjects, because the electrodes was displaced from the sympathetic nerve of the muscle or because of signal bursts from electromyographic, efferent, and afferent nerve activities. Such bursts concealed MSNA bursts attributable to arm or body movement. MSNA test was repeated 1 month later in these three subjects, and recording was successful in one. Consequently, six subjects from whom we obtained nerve recordings were used in the analysis. On day 5, two submaximal exercise tests were performed to measure thoracic impedance, SV, and CO (hemodynamic test; i.e., MSNA as not measured) in the six subjects for whom MSNA recordings were available. The procedure was identical to that of the MSNA test. The same procedure was repeated twice, i.e., the 60EX and 80 EX trials, with a 15-min interval between trials. Subjects were asked to provide their rates of perceived exertion (RPE values) (Borg 1982) during the last minute of exercise. The order of the 60EX and 80EX trials during hemodynamic test of each subject was identical to that during MSNA test. On day 6, five of six subjects performed two submaximal exercises during which SV and CO were measured using the Doppler echocardiographic technique (Doppler test; MSNA measurements were not made). The Doppler technique allowed us to confirm the differences in SV and CO measurements obtained via impedance cardiography during the 60EX and 80EX trials. The order of the two trials performed on day 6 by each subject was identical to that used in MSNA and hemodynamic tests.

**Statistical analysis**

Values are expressed as the means ± SE. Respiratory and cardiovascular variables and MSNA values during MSNA and hemodynamic tests were averaged every 1 min throughout the experiment. For all data, the assumption of normal distribution was verified using a Kolmogorov-Sminov test. Changes in variables during the experiment in each trial were analyzed using a Dunnett test (i.e., vs. the last minute of rest). Comparisons of parameters between the 60EX and 80EX trials were performed using a paired t-test (parametric test) if the distribution was regular. When the distribution was not regular, the Wilcoxon test (nonparametric test) was used. The SPSS (11.5; SPSS, Tokyo, Japan) statistical package was used only to execute the Kolmogorov-Sminov test; the StatView (5.0, SAS Institute, Tokyo, Japan) software was utilized for other statistical analyses. A P value P < 0.05 was considered to indicate statistical significance.

**Results**

**Maximal exercise test**

Cardiorespiratory parameters and workload at exhaustion during maximal exercise test were as follows: VO2 = 3.10 ± 0.08 l/min, 45.2 ± 2.4 mL/kg/min, HR = 184.7 ± 1.4 beats/min, and workload = 252.5 ± 4.3 watts.

**Submaximal exercise test**

**Baseline descriptive data**

There were no significant differences in any of the respiratory variables and HR at rest and during exercise between the MSNA and hemodynamic tests. Thus, we indicate the data for respiratory and cardiovascular variables in the hemodynamic test. Any differences in respiratory and cardiovascular variables of resting baseline were not found between 60EX and 80EX trials (Table 1). There was no significant difference in RPE during exercise between the 60EX and 80EX trials (60EX: 10.0 ± 0.7, 80EX: 10.2 ± 0.6).

**Respiratory variables**

All respiratory variables were increased significantly during exercise (Table 1). There were no significant differences in VE, VT, VO2, or VCO2 between the two trials throughout the experiment. The f value of the 80EX trial was slightly but significantly higher than that of the 60EX trial.

**Cardiovascular variables**

HR increased significantly during exercise in both trials, and this HR response to exercise was not different between the 60EX and 80EX trials (Fig. 1A). SBP and MBP increased, while DBP was unchanged during exercise (Table 1). There were no significant differences in BP variables between trials. Thoracic impedance (ΔZ) changed (from the resting values) in both exercise trials, and the change during the 80EX trial was greater (P < 0.05) than that during the 60EX trial (Fig. 2). The SV and CO levels...
(measured by impedance cardiography) during the 80EX trial were significantly higher than those of the 60EX trial (Fig. 1B and C). Similarly, the SV and CO values obtained via Doppler echocardiography were higher \( < P < 0.05 \) during the 80EX trial than those in the 60EX trial (Table 2).

**Muscle sympathetic nerve activity**

Representative MSNA recordings and changes in MSNA BF are presented in Fig. 3. MSNA BF did not change during exercise in the 60EX trial, while MSNA BF decreased significantly during exercise in the 80EX trial (Fig. 4). MSNA BF during exercise in the 80EX trial was lower \( < P < 0.05 \) than that in the 60EX trial. MSNA BI decreased significantly during exercise in both trials (Table 1), and MSNA BI during exercise in the 80EX trial was lower \( < P < 0.05 \) than that in the 60EX trial.

**Discussion**

The major finding of the present study was that MSNA BF fell during leg cycling at mild intensity when the pedaling frequency was increased (thus enhancing muscle pump). This result supports the hypothesis that no change or a decrease in sympathetic vasomotor outflow during dynamic leg exercise up to mild intensity may be due to muscle pump-induced increase in central blood volume, thereby loading the cardiopulmonary baroreceptors. The results from this study provide additional information concerning the mechanism of the MSNA response to dynamic leg exercise performed at mild intensity.

**Effect of the muscle pump-induced increase in central blood volume on MSNA**

Several previous studies have attempted to evaluate changes in MSNA during dynamic leg exercise, and
revealed that MSNA tended to decrease or was unchanged at light and mild exercise intensities compared with that at rest (Seals et al. 1988; Saito and Mano 1991; Ray et al. 1993; Saito et al. 1993; Callister et al. 1994; Ichinose et al. 2008; Katayama et al. 2011). Similarly, in the present study, MSNA BF during exercise in the 60EX trial showed little change from resting values, as shown in Fig. 4. Also, MSNA BI decreased during exercise in the 60EX trial (Table 1). One possible explanation for this observation is that dynamic leg exercise-induced muscle pump was accompanied by an increase in central blood volume. Ray et al. (1993) compared MSNA responses to dynamic a one-legged knee exercise performed in the sitting and supine positions. They compared MSNA response to dynamic exercise between supine and upright positions and noted that changes in central blood volume affected the MSNA response to dynamic exercise. In addition, Vollianitis and Secher (Vollianitis and Secher 2002) reported that arterial BP was reduced to a level below that achievable upon arm exercise alone when leg-cycling was added to an arm-cranking exercise. These results suggest that leg muscle pump strongly affects BP rather than the central command or the exercise pressor reflex. In addition, changing central blood volume affected peripheral vascular resistance (Donald and Shepherd 1978), and the extent of CO is negatively correlated with MSNA (Char-koudian et al. 2005). From these findings, it would be expected that loading of cardiopulmonary baroreceptors by a muscle pump-induced increase in central blood volume would suppress sympathetic vasomotor activity during dynamic leg exercise (Ray et al. 1993; Ray and Saito 1999; Fadel and Raven 2012; Raven and Chapleau 2014).

![Figure 2](image-url) Changes in thoracic impedance (Z values) from rest to exercise in the 60EX and 80EX trials. *P < 0.05 when comparing the 60EX and 80EX trials.

![Figure 4](image-url) Changes in MSNA BF in the 60EX and 80EX trials. *P < 0.05 vs. the last minute of rest in the 60EX trial. †P < 0.05 vs. the last minute of rest in the 80EX trial. §P < 0.05 when comparing the 60EX and 80EX trials.

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**Table 2.** Cardiac variables in the Doppler test.

| Trials | Rest  | Exercise | Recovery |
|--------|-------|----------|----------|
| HR (beats/min) | 60EX 70.0 ± 2.8 | 104.5 ± 4.1* | 78.0 ± 4.2* |
| 80EX 71.2 ± 1.4 | 105.5 ± 3.1† | 79.0 ± 3.7† |
| SV (mL) | 60EX 83.0 ± 4.1 | 106.6 ± 6.9* | 100.2 ± 5.9* |
| 80EX 81.5 ± 4.6 | 114.6 ± 7.4† | 100.8 ± 3.9† |
| CO (L/min) | 60EX 5.9 ± 0.2 | 11.5 ± 0.6* | 7.7 ± 0.3* |
| 80EX 5.8 ± 0.3 | 12.3 ± 0.6† | 7.9 ± 0.3† |

Values are mean ± SE (n = 5). Values are averaged over the last 30 sec of each session. HR, heart rate; SV, stroke volume; CO, cardiac output. SV and CO were obtained by the Doppler echo-cardiography. *P < 0.05 vs. at Rest in the 60EX trial. †P < 0.05 vs. at Rest in the 80EX trial. §P < 0.05 60EX trial vs. 80EX trial.

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**Figure 3.** Representative ECG and MSNA recordings made during the 60EX and 80EX trials.
In the present study, we utilized a traditional method (an increase in pedal frequency from 60 to 80 rpm) to further enhance skeletal muscle pump frequency (Gotshall et al. 1996; Rowland and Lisowski 2001; Ogoh et al. 2007). This manipulated central blood volume in a controlled manner; SV and CO increased significantly during leg cycling at 80 rpm (the 80EX trial) compared with the values recorded at 60 rpm (the 60EX trial) (Fig. 1). Since CO is equal to venous return under steady-state conditions (Badeer 1981; Young 2010), increased CO during the 80EX trial indicates enhanced central blood volume. Moreover, thoracic impedance, an index of central blood volume, was higher during the 80EX trial than the 60EX trial (Fig. 2). Consequently, MSNA BF decreased during the 80EX trial (Fig. 4), and MSNA BI (Table 1) during this trial was lower than during the 60EX trial, suggesting that enhanced muscle pump, which caused loading of cardiopulmonary baroreceptors, could inhibit sympathetic vasomotor outflow during dynamic leg exercise at mild intensity.

In an animal study, the BP response at the onset of exercise was not maintained when the cardiopulmonary receptors operated alone (without arterial baroreflex) (Walgenbach and Donald 1983). They concluded that cardiopulmonary receptors do not have a significant role in BP regulation. However, Ogoh et al. (2007) reported that loading of cardiopulmonary baroreceptors modified the arterial baroreflex evident during steady-state exercise. In addition, orthostatic stress increased peripheral vascular resistance via unloading of cardiopulmonary baroreceptors to maintain the required BP during mild-intensity exercise (Mack et al. 1988). These earlier findings indicated that the cardiopulmonary and arterial baroreflexes interacted to regulate BP during exercise. Similarly, in the present study, sympathetic vasoconstriction outflow was suppressed by loading of cardiopulmonary baroreceptors during the 80EX trial. Interestingly, BP during exercise in the 80EX trial did not differ from that in the 60EX trial (Table 1). Therefore, suppression of sympathetic vasoconstriction outflow by the cardiopulmonary reflex in the 80EX trial may prevent overshooting of BP during exercise. Taking these observations into consideration, cardiopulmonary receptors could play an important role for sympathetic vasomotor outflow to maintain adequate BP against changes in central blood volume during steady-state mild dynamic exercise.

We need to consider other possible mechanisms affecting MSNA during dynamic leg exercise. First, it is necessary to consider energy expenditure which is related to central command and the exercise pressor reflex. It is well known that an increase in pedaling cadence induces a rise in gross energy expenditure at a constant load because internal work is altered (Lollgen et al. 1980; Wells et al. 1986). Thus, increases in cycle pedaling cadence may produce greater activation of central command and the exercise pressor reflex when workload is kept constant. In order to apply the same central command and exercise pressor reflex, we used differential workloads to obtain the same energy expenditure (i.e., VO2) during the 60EX and 80EX trials (Table 1) (Ogoh et al. 2007). Similar to VO2, there were no significant differences in HR and RPE between the two trials. Therefore, we suppose that central command inputs were similar during two trials types; although, it is difficult to estimate the extent thereof.

In terms of the exercise pressor reflex, the metaboreflex and mechanoreflex within skeletal muscle contribute to changes in MSNA during exercise. In an animal study, it has been revealed that group IV muscle afferent was increased during low intensity dynamic exercise (Adreani et al. 1997). An increase in muscle contraction frequency may limit muscle blood flow, thereby triggering metabolite accumulation. However, Ferreira et al. (2006) found no difference in the extent of vastus lateralis deoxygenation (assessed using near-infrared spectroscopy) during cycling at 60 and 100 rpm, indicating that the increase in contraction frequency did not impair blood flow to the muscle. Further, the VCO2 value obtained during the 80EX trial did not differ from those of the 60EX trial (Table 1). Collectively, although metaboreflex during dynamic leg cycling at low intensity could enhance MSNA, it seems likely that the extent of metaboreflex did not differ between the 60EX and 80EX trials. As for the mechanoreflex, mechanoreceptor loading certainly increases as muscle contraction frequency rises, and group III muscle afferent was enhanced during dynamic exercise at low intensity (Adreani et al. 1997). Passive stretching of the hindlimb muscle significantly increased renal sympathetic nerve activity (Matsukawa et al. 1990). In addition, MSNA has been reported to increase during passive muscle stretching in humans (Cui et al. 2006). These findings suggest that an increase in mechanoreceptor loading during the 80EX trial could enhance, not inhibit, MSNA.

Finally, respiratory modulation of MSNA should be mentioned (Hagbarth and Vallbo 1968; Eckberg et al. 1985; Dempsey et al. 2002). In the present study, subjects adopted a high breathing frequency during the 80EX trial; however, this difference was small. Seals et al. (1990) showed that differences in depths and patterns of breathing influenced the within-breath MSNA modulation. However, St Croix et al. (2000) found that MSNA was unchanged when the breathing frequency increased threefold (via voluntary hyperpnea). Therefore, it is unlikely that the slight difference in breathing frequency between the 60EX and 80EX trials significantly affected MSNA; although, we cannot entirely rule out respiratory modulation of MSNA.
Technical considerations and limitations

Several technical considerations and limitations of the study should be noted. One limitation was the use of thoracic electrical impedance as an index of central blood volume; we could not obtain direct measurements of central venous pressure. Previous studies found that changes in thoracic impedance relates to central venous pressure during lower body positive or negative pressure (Ebert et al. 1986; Cai et al. 2000). As in the present study, Ogh et al. (2007) found that thoracic impedance decreased when the pedal cadence rose from 60 to 80 rpm (at the same VO2) during submaximal leg cycling at mild intensity. In addition to thoracic impedance, we measured the central hemodynamic variables SV and CO by both impedance cardiography and a Doppler echocardiographic technique. As CO is equal to the venous return under steady-state conditions (Badeer 1981; Young 2010), we presumed that changes in central blood volume were reflected in the CO values. Although the absolute values of SV and CO obtained by the Doppler technique were lower than those derived by the impedance method (in line with data of a previous study) (Christie et al. 1987), both SV and CO were significantly higher during the 80EX trial (Fig. 1 and Table 2). Taking these observations into consideration, it seems reasonable to suggest that central blood volume increased during the 80EX trial of the present study.

It is necessary to consider exercise intensity and duration. We utilized mild exercise intensity (40% VO2peak at 60 rpm), because the percentage of successful MSNA recordings is high when the exercise intensity is mild and movements of the arm and the body become greater during leg cycling with a high pedaling rate (Katayama et al. 2012, 2013). The extent of contribution of cardiopulmonary baroreceptor to sympathetic vasoconstriction outflow would alter with increasing exercise intensity, as well as other mechanisms (e.g., metaboreflex) (Ray et al. 1993; Fisher et al. 2005; Boushel 2010). As for the exercise duration, Saito et al. (1997) and Ray et al. (1993) demonstrated a progressive increase in MSNA during 30- and 40-min of dynamic leg exercise at mild intensity. Therefore, it may be that the influence of the cardiopulmonary receptors overwhelms when exercise intensity is high and exercise duration prolonged (Ray et al. 1993).

Another limitation of this study was the limited number and characteristics of subjects. Clinically, cardiopulmonary baroreceptor is particularly important to regulate arterial BP at rest and during exercise. It has been reported that resting MSNA in obesity and patients with heart failure and hypertension is higher than in healthy individuals (Yamada et al. 1989; Grassi et al. 2003; Witte et al. 2008). Furthermore, cardiopulmonary reflex is impaired in patients with heart failure (Dibner-Dunlap et al. 1996). Thus, it is assumed that the MSNA response to changes in central hemodynamics during dynamic leg exercise in patients differs from that in healthy subjects.

Conclusion

MSNA BF during leg cycling at mild intensity was reduced when central blood volume was increased by elevating pedaling frequency (e.g., muscle contraction frequency). These results suggest that a muscle pump-induced increase in central blood volume, and thereby loading of cardiopulmonary baroreceptors, could inhibit sympathetic vasomotor outflow during mild dynamic leg exercise, despite activation of central command and the exercise pressor reflex.

Conflict of Interest

No conflicts of interest, financial or otherwise, are declared by the authors.

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