Geometric consequences of electron delocalization for adenine tautomers in aqueous solution

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Abstract Geometric consequences of electron delocalization were studied for all possible adenine tautomers in aqueous solution by means of ab initio methods (PCM(water)//DFT(B3LYP)/6-311+G(d,p)) and compared to those in the gas phase (DFT(B3LYP)/6-311+G(d,p)). To measure the consequences of any type of resonance conjugation (π-π, n-π, and σ-π), the geometry-based harmonic oscillator model of electron delocalization (HOMED) index, recently extended to the isolated (DFT) and hydrated (PCM//DFT) molecules, was applied to the molecular fragments (imidazole, pyrimidine, 4-aminopyrimidine, and purine) and also to the whole tautomeric system. For individual tautomers, the resonance conjugations and consequently the bond lengths strongly depend on the position of the labile protons. The HOMED indices are larger for tautomers (or their fragments) possessing the labile proton(s) at the N rather than C atom. Solvent interactions with adenine tautomers slightly increase the resonance conjugations. Consequently, they slightly shorten the single bonds and lengthen the double bonds. When going from the gas phase to water solution, the HOMED indices increase (by less than 0.15 units). There is a good relation between the HOMED indices estimated in water solution and those in the gas phase for the neutral and ionized forms of adenine. Subtle effects, being a consequence of intramolecular interactions between the neighboring groups, are so strongly reduced by solvent that the relation between the HOMED indices and the relative energies for the neutral adenine tautomers seems to be better in water solution than in the gas phase.

Keywords Adenine · Electron delocalization · HOMED/ΔE relation · PCM//DFT · Solvent effects · Tautomers

Introduction

Various natural products, including pyrimidine and purine bases of nucleic acids, display prototropic tautomerism, the phenomenon which strongly influences their structure and biochemical transformations [1–4]. Depending on the number of the labile protons and on the number of tautomeric groups, they possess two or more tautomeric forms. Very often experimental techniques, applied to tautomeric compounds, cannot give the complete information on the structure of all possible tautomers. For example, application of spectroscopic methods such as ultraviolet (UV), infrared (IR), Raman, microwave (MW), nuclear magnetic resonance (NMR), mass spectrometry (MS), etc., leads mainly to the detection of the major tautomers, signals of which have significant intensities [1–3]. The minor and rare tautomers are not identified, probably because their amounts are too small and their signals cannot be distinguished from the background. Many internal and external factors such as polarity, aromaticity, stability of functionalities, acid–base properties of conjugated tautomeric sites, substituent and solvent effects, intra- and/or intermolecular interactions, oxidizing and reducing agents, ions, electrons, UV, γ-, and X-ray, etc. influence tautomeric equilibria. Depending on the tautomeric system, one or the other factor dictates the tautomeric preferences. In the case of simple tautomeric systems, a relation between prototropy and electron delocalization (also called resonance or mesomerism) has been signaled more than 50 years ago by Pauling [5]. This
relation seems to be very simple, because prototropy by definition is associated with changes in electron delocalization [1–3]. However, electron delocalization (e.g., aromaticity) is not always the main factor that influences tautomeric equilibria and the relation between prototropy and electron delocalization is frequently perturbed by other factors [3].

In the case of adenine (Chart 1) – a well-known component part of nucleic acids, electron delocalization (aromaticity) seems to play a principal role for tautomeric conversions. To our knowledge, there are only a few reports discussing aromaticity for the canonical form [6–10] and for N-benzyl derivatives of three amine tautomers of adenine in the gas phase [11]. Lack of sufficient literature data encouraged us to study electron delocalization by means of geometric criterion for all possible prototropic tautomers of adenine in the gas phase using quantum-chemical methods [12]. A good relation between prototropy and electron delocalization, perturbed solely by some subtle internal effects, has been found. Since various processes for living organisms take place not only in nonpolar (lipids) but also in polar environments (enzymes, receptors, proteins, nucleoproteins, etc.) containing water molecules, it is interesting to study how electron delocalization varies in two extreme environments, that is, when proceeding from the gas phase (isolated molecules) to aqueous solution (hydrated molecules). For this reason, we extended our studies on electron delocalization to aqueous solution. We investigated the geometric consequences of electron delocalization for the hydrated adenine tautomers and compared them to those reported previously for isolated ones [12]. Such kind of studies gave the possibilities to estimate the extreme external effects. They should not be smaller than those for adenine included in DNA in living organisms systems.

To properly determine the distribution of π- and n-electrons for all individual tautomers of adenine, and to describe well the relation between prototropy and electron delocalization, the recently extended geometry-based harmonic oscillator model of electron delocalization (HOMED) procedure [13, 14] was applied to the geometries of the adenine isomers optimized in aqueous solutions at the PCM(water)//DFT(B3LYP)/6-311+G(d,p) level [15]. The HOMED index is based on the original harmonic oscillator model of aromaticity (HOMA) idea [16, 17]. An abbreviation HOMED was proposed for the modified index [13], but it may also be abbreviated as moHOMA (modified original HOMA) or simply HOMA. Since the same phenomenon of electron delocalization takes place for the neutral, ionic, and radical species, the same geometric criterion (HOMED) was applied for the neutral and ionized adenine isomers. It should be mentioned here that one can find in the literature many other indices of aromaticity which can be classified as energetic and magnetic ones [18–22]. For simple five-membered rings, good correlations occur between the geometry-based, energetic, and magnetic indices [21]. However, for polycyclic systems, the situation is more complex, because the magnetic indices are local descriptors and cannot be applied to the whole molecule, whereas the energetic indices are rather global ones, and cannot be used for the fragments. Solely the geometry-based indices can be applied for the whole molecule as well as for its fragments [18]. They can measure the geometric consequences of local and global electron delocalization for polycyclic systems.

The HOMA index, reformulated by Krygowski in 1993 (rHOMA) [23] and applied to heterocycles [18, 19], including the canonical structures of nucleobases [6], and the harmonic oscillator model of heterocyclic electron delocalization (HOMHED) index, proposed by Frizzo and Martins in 2012 [24] and based on the hypotheses of the HOMED index [14], were not applied here for the adenine tautomers. The reasons are as follows. In the rHOMA procedure, different measures of π-electron delocalization were employed for the reference CC, CX, and XY bonds [18, 19, 23], and the rHOMA index is inappropriate for the systems containing heteroatoms, e.g., purine [25]. It may be solely used for compounds possessing the same type of bonds, e.g., hydrocarbons [14, 26]. In the HOMHED procedure, the statistical reference bond lengths were applied [24]. The use of the statistical reference CC, CX, and XY bond lengths leads to some kind of statistical HOMHED values which do not describe well the real electron delocalization in heteroatomic systems.

Methods

Geometries of adenine isomers in their ground states were fully optimized without symmetry constraints at the DFT(B3LYP)/6-311+G(d,p) level [27–30], and next re-optimized at the PCM(water)//DFT(B3LYP)/6-311+G(d,p) level [31, 32] as previously described [15]. For all calculations, the Gaussian 03 program [33] was used. The HOMED indices [13, 14] for the hydrated structures were estimated on the basis of the theoretically derived bond lengths (Tables S1
and S2, Supplementary material) according to Eq. (1) [16–18, 23] as described previously for purine [25]. In this equation, $\alpha$ is a normalization constant, $R_o$ is the optimum bond length (assumed to be realized for fully delocalized system), $R_i$ are the running bond lengths in the system, and $n$ is the number of bonds taken into account. For the systems containing the even number of bonds (2i), the normalization constant $\alpha$ were calculated from Eq. (2), where $R_s$ and $R_d$ are the reference single and double bonds, respectively. This equation is similar to that proposed for the reformulated HOMA index [23]. In the case of the systems containing the odd number of bonds (2i+1), i.e., (i+1) single bonds and (i) double bonds, the normalization constant $\alpha$ were calculated according to Eq. (1) [16, 17].

\[
\frac{1}{n} \left\{ \sum R_s^2 - \sum R_d^2 \right\}^{1/2} \quad (2)
\]

\[
\alpha = 2 \cdot \left( \sum R_s^2 - \sum R_d^2 \right)^{1/2} \quad (3)
\]

To quantitatively measure the geometric consequences of electron delocalization for the major, minor, and rare tautomers of adenine (A1-A23) in aqueous solution, the HOMED indices were estimated for the molecular fragments: imidazole (five bonds − HOMED5), pyrimidine (six bonds − HOMED6), 4-aminopyrimidine (seven bonds − HOMED7), and purine (ten bonds − HOMED10). They were also estimated for the whole tautomeric system including the exo-NH$_2$/NH groups (11 bonds − HOMED11). For parametrization, the following $R_s$, $R_d$, and $R_o$ values (in Å), calculated at the PCM(water)//DFT(B3LYP)/6-311+G(d,p) level [25], were taken here: 1.5308 (ethane), 1.3305 (ethene), and 1.3965 (benzene) for the CC bonds, and 1.4691 (methylamine), 1.2698 (methylimine), and 1.3354 (1,3,5-triazine) for the CN bonds, respectively. The $R_o$ bond lengths, computed for benzene and 1,3,5-triazine (reference aromatic systems, HOMED=1), are in accord with the harmonic oscillator method of optimization [Eq. (4)] [16, 17], in which $\omega$ (close to 2 for the CC and CN bonds) is the ratio of the stretching force constants for the pure double and single bonds. For the hydrated adenine fragments containing the even number of bonds, i.e., pyrimidine ring and purine system, the following normalization constants, calculated according to Eq. (5), were used: 89.32 (CC) and 90.18 (CN). For those with the odd number of bonds, the normalization constants were calculated according to Eq. (3). They are as follows: 79.59 (CC) and 80.34 (CN) for the imidazole fragment, 82.15 (CC) and 82.92 (CN) for the 4-aminopyrimidine fragment, and 84.64 (CC) and 85.42 (CN) for the whole tautomer adenine system.

### Results and discussion

**Adenine tautomers**

The tautomeric mixture of adenine contains 23 tautomers (Table 1), nine amine forms (A1-A9), and 14 imine forms (A10-A23) with the exo amine−NH$_2$ and imine−NH group, respectively. When geometric isomerism of the imine forms is taken into account, 37 isomers are possible (Fig. S1, Supplementary material) [15]. The number of the prototropic tautomers for adenine is a consequence of the number of the labile protons (two) and conjugated tautomeric sites (ten). In the majority of theoretical documents, the amine-imine tautomeric conversions {−NH−C(R)=N−→−N=C(R)=NH} have been investigated, and a maximum number of eight tautomers (A1, A3, A7, A9, A13, A15, A18, and A20) have been considered [34–38]. Application of NMR techniques have led to identification of three major tautomers (A3, A7, and A9) [39]. The enamine-imine conversions {−C=C(R)=NH−→−NH−C(R)=N} have usually been omitted. Some exceptions are the anionic states of adenine for which the rare tautomers dominate [40, 41]. These variations of the tautomeric preferences and the hypothesis of rare tautomers, suggested by Watson and Crick [42] in 1953 for DNA mutations, encouraged us to study all 37 isomers for adenine [15].

Adenine contains eight $\pi$- and ten $n$-electrons (Chart 1) which participate in electron delocalization during tautomeric conversions. More than one satisfactory Lewis structure, called resonance structures, can be drawn for each isomer [15, 43]. A single structure is composed of alternating single and double bonds. Electron delocalization for the resonance hybrid containing two or more resonance structures reveals shortening of the single bonds and lengthening of the double bonds. Solely full symmetry of the molecule leads to equalization of the corresponding bond lengths. Generally, the bond length alternation, or its lack of, is a consequence of various types of resonance conjugation in the molecule such as $\sigma$–$\pi$ hyperconjugation, $\pi$–$\pi$ and $n$–$\pi$ conjugations. For the NH-NH tautomers of adenine, containing the labile protons at the N atoms, the $\pi$–$\pi$ and $n$–$\pi$ resonance conjugations are possible. For the NH-CH tautomers, possessing the labile protons at the N and C atoms, the $\sigma$–$\pi$ hyperconjugation should be additionally considered. These different types of resonance conjugation lead to different geometric consequences of electron delocalization for the adenine tautomers. The number of the resonance structures substantially depends on the positions of the labile protons and on the positions of the double bonds in the Lewis structures. The number of the resonance structures...
affects the bond lengths, the stability of the tautomeric form, and its contribution in the tautomeric mixture. Consequently, prototropy and electron delocalization influence the structure of adenine, its physicochemical, chemical, and biochemical properties.

In this paper, all possible 37 isomers were considered for neutral adenine at the PCM(water)//DFT(B3LYP)/6-311+G(d,p) level. For selected isomers, the positively and negatively ionized forms (radical cations and anions) were then theoretically derived by removing and adding one electron to the neutral isomer. These transformations refer to one-electron gain and one-electron loss. The highest one-electron gain affects the bond lengths, the stability of the tautomeric form, and its contribution in the tautomeric mixture. Consequently, prototropy and electron delocalization influence the structure of adenine, its physicochemical, chemical, and biochemical properties.

Table 1: All possible 23 prototropic tautomers of adenine [15]

| Tautomer | Position of H  | Tautomer | Position of H  | Tautomer | Position of H  |
|----------|----------------|----------|----------------|----------|----------------|
| A1       | N1N10          | A9       | N9N10          | A17      | N3C5           |
| A2       | C2N10          | A10      | N1C2           | A18      | N3N7           |
| A3       | N3N10          | A11      | N1C4           | A19      | N3C8           |
| A4       | C4N10          | A12      | N1C5           | A20      | N3N9           |
| A5       | C5N10          | A13      | N1N7           | A21      | C5N7           |
| A6       | C6N10          | A14      | N1C8           | A22      | C5N9           |
| A7       | N7N10          | A15      | N1N9           | A23      | C8N9           |
| A8       | C8N10          | A16      | N3C4           |          |                |

In the presence of oxidizing or reducing agents, adenine may lose or gain one electron, and the charged radicals can be formed. They may also be generated electrochemically, photochemically, or during positive or negative ionization in various types of mass spectrometers. One-electron loss and one-electron gain affect the stability of adenine tautomers and their electron delocalization in different ways. However, when proceeding from the gas phase to aqueous solution, the estimated partial and total HOMED values slightly increase (by 0.001-0.006 units) indicating an increase of electron delocalization. The lowest solvent effect (0.001) takes place for the pyrimidine fragment which is already well delocalized (HOMED5 close to unity), and the largest one (0.006) for the imidazole fragment which is less delocalized (HOMED5 close to 0.9).

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|----------|----------------|----------|----------------|----------|----------------|
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| A2       | C2N10          | A10      | N1C2           | A18      | N3N7           |
| A3       | N3N10          | A11      | N1C4           | A19      | N3C8           |
| A4       | C4N10          | A12      | N1C5           | A20      | N3N9           |
| A5       | C5N10          | A13      | N1N7           | A21      | C5N7           |
| A6       | C6N10          | A14      | N1C8           | A22      | C5N9           |
| A7       | N7N10          | A15      | N1N9           | A23      | C8N9           |
| A8       | C8N10          | A16      | N3C4           |          |                |

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The canonical tautomer is the favored form of positively ionized adenine [15]. In both environments (for isolated and hydrated molecule), the most important amount of the spin density exists on the N10 atom [15]. This means that one non-bonding electron is taken from the exo–NH₂ group of the 4-aminopyrimidine fragment, rather than from the endo NH group of the imidazole ring. Lack of one electron on the N10 atom reduces the n-π conjugation of the exo–NH₂ group
(a) neutral

\[
\begin{align*}
\text{gas phase} & \quad \text{water solution} \\
0.992 & \quad 0.887 \\
0.931 & \quad 0.889 \\
0.995 & \quad 0.906 \\
0.938 & \quad 0.942 \\
\end{align*}
\]

(b) positively ionized (oxidized)

\[
\begin{align*}
\text{gas phase} & \quad \text{water solution} \\
0.911 & \quad 0.916 \\
0.904 & \quad 0.926 \\
0.890 & \quad 0.904 \\
0.916 & \quad 0.928 \\
\end{align*}
\]

(c) negatively ionized (reduced)

\[
\begin{align*}
\text{gas phase} & \quad \text{water solution} \\
0.881 & \quad 0.906 \\
0.614 & \quad 0.342 \\
0.906 & \quad 0.628 \\
0.342 & \quad 0.669 \\
0.638 & \quad 0.669 \\
\end{align*}
\]

Chart 2 Variations of the partial and total HOMED indices for the favored tautomers of neutral (a), positively (b), and negatively (c) ionized adenine when proceeding from the gas phase to aqueous solution.

with the pyrimidine ring, and decreases the corresponding HOMED values for A9⁺⁺ in comparison to those for A9: HOMED6 (from 0.995 to 0.890 in the gas phase and from 0.996 to 0.900 in water solution) and HOMED7 (from 0.992 to 0.911 in the gas phase and from 0.994 to 0.916 in water solution). Consequently, the other HOMED values also decrease: HOMED10 for the purine fragment (from 0.931 to 0.904 in the gas phase and from 0.936 to 0.920 in water solution) and HOMED11 for the whole tautomeric adenine system (from 0.938 to 0.916 in the gas phase and from 0.942 to 0.928 in water solution). The decrease effects of the HOMED values in water solution (0.096, 0.078, 0.016, and 0.014 units, respectively) are only slightly lower than those observed in the gas phase (0.105, 0.081, 0.027, and 0.022 units, respectively [12]). One exception is the HOMED5 index, which value increases when going from A9 to A9⁺⁺ (from 0.883 to 0.926 in the gas phase and from 0.889 to 0.943 in water solution). The increase effects (0.043 and 0.054 units, respectively) confirm the fact that one non-bonding electron is not taken from the endo N atom of the imidazole fragment. The explanation is as follows. For the favored NH tautomer of isolated imidazole, for which one non-bonding electron may be taken from the endo NH group, positive ionization decreases the HOMED5 value for both the isolated (from 0.902 to 0.877) and hydrated tautomers (from 0.917 to 0.907) [67]. This effect is completely different than that for the adenine imidazole fragment.

Negative ionization reduces the stability of the canonical tautomer such that A9⁺⁺ has larger energy value than A8⁻—the NH-CH one, favored for reduced but considered as the rare form for neutral and oxidized adenine [15, 40, 41]. For A8⁻, the HOMED5, HOMED6, HOMED7, HOMED10, and HOMED11 values in water solution (0.365, 0.924, 0.925, 0.628, and 0.669, respectively, Chart 2c) are only slightly larger (by 0.014-0.044 units) than those found for A8⁺ in the gas phase (0.342, 0.906, 0.881, 0.614, and 0.638, respectively [12]). This form is non-aromatic due to the presence of the C8sp³ atom in the imidazole fragment. Solely the pyrimidine and 4-aminopyrimidine fragments in A8⁻ are well delocalized (HOMEDs close to 0.9). The change of the tautomeric preference for negatively ionized adenine suggests that in this case aromaticity is not the main factor that affects the composition of the tautomeric mixture. This seems to be independent of environment.

HOMED(water)/HOMED(gas) relation

First perusal of the total HOMED values estimated for the selected neutral and ionized forms of adenine in the gas phase and in water solution suggests that electron delocalization and consequently bond length alternation or its lack seem to be less sensitive to solvation than to electron transfer from and to adenine molecule (Table S3, Supplementary material). However, the estimated HOMED values are consistent with a general tendency of the resonance conjugation: more delocalized system, larger HOMED index value [14]. The HOMED indices are between zero and unity for the neutral and ionized isomers. Generally, the HOMED values increase for the adenine tautomers when proceeding from the gas phase to water solution. This increase may be explained by specific interactions of polar solvent with polar functional groups of adenine. Specific interactions (e.g., H-bonding) augment the geometry-based indices [3, 6]. For the neutral adenine forms, solvent effects seem to be larger for the NH-CH tautomers than for the NH-NH ones. The total HOMED indices increase by 0.011-0.136 and 0.004-0.097 units, respectively. Positive and negative ionization change the total HOMED indices in different way, in some cases even by more than 0.1 units. However, solvent effects seem to be similar to those for neutral tautomers: less delocalized tautomer larger solvent effect.
The parallelism of the geometric consequences of electron delocalization for the adenine tautomers in apolar (gas phase) and polar (water) environments leads to a good linear relation between the HOMED11 indices estimated for the whole tautomeric system (11 bonds) in the gas phase and in water solution (Fig. S2, Supplementary material). The HOMED11 values for the adenine tautomers in the gas phase were taken from ref. [12]. The HOMED(water)/HOMED(gas) relation seems to be independent of the oxidation state. It is common for the neutral and ionized isomers of adenine. Similarly, a good linear relation exists between the total HOMED10 values estimated in water solution and in the gas phase for the neutral and ionized tautomers of the parent system–purine (ten bonds). This relationship is also present in Fig. S2 for comparison. The HOMED10 values for the purine isomers were taken from ref. [25]. A similar tendency was found for correlations between the relative energies (\(\Delta E\)) in the gas phase and in water solution for the neutral adenine and purine tautomers (Fig. S3, Supplementary material). However, for the ionized forms the changes in the relative energies are not parallel to those for the neutral forms and any common linear relation can be proposed.

HOMED/\(\Delta E\) relation

Adenine is a nice example of natural product with a good relationship observed between prototropy and electron delocalization for the isolated and hydrated neutral tautomers. The relative energies (\(\Delta E\)), which measure the relative stabilities of individual tautomers, correlate well with the total HOMED11 indices, which measure the geometric consequences of electron delocalization for the whole tautomeric system–11 bonds (Fig. 1). The relative energies for the hydrated adenine isomers were taken from ref. [15]. The HOMED and \(\Delta E\) values for the isolated (gas phase) adenine isomers were taken from refs. [12, 15]. The good HOMED/\(\Delta E\) relationship confirms that electron delocalization, and particularly aromaticity, is one of the main factors that dictates the tautomeric preferences for neutral adenine in water solution and in the gas phase. Solvent reduces solely the internal effects between the functional groups and diminishes the deviations of points for particular isomers from the HOMED/\(\Delta E\) relation.

Direct comparison of the \(\Delta E\) values calculated for the hydrated tautomers of adenine with the partial HOMED values estimated for their fragments (five, six, seven, and ten bonds) shows additionally that the variations of the HOMED5 indices for the imidazole fragment do not follow well the variations of the \(\Delta E\) values, and a scatter plot is rather observed in water solution (Fig. S4a, Supplementary material). The main reason is a small variation of the position of the labile proton(s) for the imidazole fragment [15]. There are 23 tautomers of the bicyclic adenine possessing different stabilities (\(\Delta E\) values vary from 0 to 68 kcal mol\(^{-1}\) in water solution [15]), whereas these tautomers contain solely ten different imidazole fragments, for which the HOMED5 values vary in four ranges in water solution: 0.89-0.96, 0.56-0.67, 0.43-0.49, and 0.33-0.39, respectively. The fragments similar to the free imidazole tautomers possess the HOMED5 values very close to those estimated in water solution for the imidazole N1H/N3H (0.917), C2H (0.354), and C4H/C5H tautomers (0.390) [67]. The small variations of the HOMED5 values for the adenine tautomers affect the deviations of points from the linear HOMED/\(\Delta E\) relation for the purine fragment (Fig. S4d), and also for the whole adenine tautomeric system (Fig. 1). Similar tendency has been found in the gas phase between the HOMED11 and \(\Delta G\) values [12].

The HOMED6 (Fig. S4b) and HOMED7 (Fig. S4c) values estimated for the pyrimidine and 4-aminopyrimidine fragments of the adenine tautomers, seem to be almost parallel to the \(\Delta E\) values in water solution. The variations of the position of the labile proton(s) in these fragments are considerably greater than that for the imidazole one. The 23 adenine tautomers contain 19 different 4-aminopyrimidine fragments. The HOMED7 values progressively change from 0.68 to 0.99 for the 4-aminopyrimidine fragments with the labile proton(s) at the N atom(s) and from 0.33 to 0.73 for the 4-aminopyrimidine...
fragments containing at least one labile proton at the C atom. Some subtle internal effects of the exo-NH$_2$ group cause some deviations of points for particular isomers from the HOMED/ΔE relation. These subtle effects also influence the deviations of points from the HOMED/ΔE relation for the whole adenine tautomeric system (Fig. 1). This tendency observed in water solution is similar to that in the gas phase [12].

Effects of the exo functional group

Total effects of the exo-NH$_2$ group in water solution are different than those in the gas phase (Table S5, Supplementary material). In the gas phase [12], the total effects are a mixture of the inductive and resonance effects of the N10H$_2$ group and also of those resulting from specific intramolecular interactions of this group with the endo neighboring N1/N1H and N7/N7H groups (Fig. S1, Supplementary material). In water solution, the total effects of the N10H$_2$ group additionally contain specific interactions with solvent. These interactions change the partial N10H$_2$ effects possible in the gas phase, and change the total N10H$_2$ effects in water solution. To quantitatively measure the total N10H$_2$ effects for neutral adenine, the HOMED indices (for the five, six, and ten bonds) and the relative energies (ΔE), estimated for the amine tautomers of adenine (A1-A9) were compared with those for the parent system – purine (P1-P9 - analogous to A1-A9) [25], both calculated at the same levels of theory, in the gas phase and in water solution. The comparisons lead to the geometric {δHOMED10=HOMED10(A)−HOMED10(P)} and energetic {δE=ΔE(A)−ΔE(P)} total N10H$_2$ effects. Their variations in water solution and in the gas phase are illustrated in Fig. 2.

The electron-withdrawing inductive effect of the N10H$_2$ group is energetically unfavorable. In the gas phase, this effect is well manifested for A6, for which the n-π conjugation between the N10H$_2$ group and the purine system is not possible [15]. Moreover, the N10H$_2$ group cannot intramolecularly interact with the endo functional groups [15]. The electron-withdrawing inductive effect of the N10H$_2$ group strongly reduces the basicity of the C6 atom containing the N10H$_2$ group such that this atom is exceptionally rare and may take the labile proton. Consequently, the δE value is strongly positive in the gas phase (19.1 kcal mol$^{-1}$). It is slightly lower in water solution (17.9 kcal mol$^{-1}$) due to interactions of the N10H$_2$ group with the polar solvent. The solvent also diminishes the geometric N10H$_2$ effect. The absolute δHOMED10 value decreases when going from the gas phase to water solution (by 0.019 units).

The electron-donating resonance effect of the N10H$_2$ group possible for the other amine tautomers of adenine (A1-A5 and A7-A9) depends on the position of the labile proton [15]. It is less or more energetically favorable. On the other hand, the specific intramolecular interactions of the N10H$_2$ group with the N1/N1H and N7/N7H groups seem to be energetically favorable solely for A2-A5, A8, and A9 (Fig. S1, Supplementary material). For A1 and A7, these interactions are energetically unfavorable due to some repulsions of the N10H/N1H and N10H/N7H groups, respectively. These unfavorable effects lead to the large positive δE values for A1 (5.6 kcal mol$^{-1}$) and A7 (4.3 kcal mol$^{-1}$) in the gas phase. In water solution, these effects are reduced by energetically favorable interactions with the polar solvent, and the δE values decrease (to 1.9 and 1.8 kcal mol$^{-1}$, respectively). The energetically unfavorable intramolecular interactions of the N10H and N1H groups in A1, and the N10H and N7H groups in A7 do not affect very much electron delocalization for the pyrimidine and imidazole fragments of A1 and A7, respectively (Table S5, Supplementary material).

Interestingly, the geometric and energetic total effects of the exo-NH$_2$ group are strongly favorable for some NH-CH$_2$ tautomers: A4, A5, and A8. The δHOMED values are very large in the gas phase and also in water solution. The δE values are even more negative in water solution (−7.0, −13.2, and −9.1 kcal mol$^{-1}$) than in the gas phase (−4.5, −10.1, and −6.1 kcal mol$^{-1}$, respectively). The largest

![Fig. 2 Variations of the geometric (δHOMED10) and energetic (δE in kcal mol$^{-1}$) total effects of the exo NH$_2$ group for the neutral amine tautomers of adenine in the gas phase and in water solution. Numbers 1–9 correspond to the amine tautomers A1-A9, respectively.](image-url)
favorable geometric and energetic effects take place for A5, indicating that the exo=NH₂ group strongly favors the transfer of the proton to the C5 atom of adenine in comparison to purine. This atom is also favored for the labile proton in uric acid [68]. The particular stability of the C5H tautomer for uric acid may partially explain the formation of the unstable intermediate, 5-hydroxisourea with the OH group at the C5 atom [15, 69–72].

For the imine tautomers of adenine (A10–A23, Fig. S1 in Supplementary material), the imine H atom in the exo=NH group may take two different positions (a and b). This leads to various specific intramolecular interactions between the exo and endo functional groups which affect the energetic and geometric parameters of individual isomers (Table S6, Supplementary material). When intramolecular interactions are energetically favorable for one of the isomers and energetically unfavorable for the other one, e.g., for A10, A11, A12, A14, A15, A18, and A21, the difference between the relative energies of these isomers {δE=ΔE(b)−ΔE(a)} is exceptionally large in the gas phase (4–7 kcal mol⁻¹). Solvent diminishes intramolecular interactions and reduces the δE values (to 1–2 kcal mol⁻¹). When there are no significant differences between intramolecular interactions for the isomers a and b, the δE values are close to zero in water solution and not larger than 2 kcal mol⁻¹ in the gas phase. Unfortunately, the variations of the energetic parameters are not in line with those of the geometric ones (Fig. 3). However, the differences between the total HOMED indices estimated for the isomers a and b {δHOMED11=HOMED(b)−HOMED11(a)} are lower in water solution than in the gas phase, and thus the deviations of points for the hydrated imine isomers in the HOMED/ΔE plot are smaller than those for the isolated ones (Fig. 1).

Conclusions

Quantum-chemical calculations, performed for the favored and rare tautomers of neutral and ionized adenine in aqueous solution {PCM(water)//DFT(B3LYP)/6-311+G(d,p)} and compared to those in the gas phase {DFT(B3LYP)/6-311+G(d,p)}, indicate that solvation and ionization effects influence the geometric parameters less drastically than the energetic ones. The geometry-based HOMED indices estimated here for the neutral and ionized forms of adenine in aqueous solution are parallel to those reported previously for the isolated molecules (gas phase) [12]. Ionization effects on the HOMED indices seem to be independent of environment. Quite a different situation takes place for the relative energies. A good relation between the relative energies found in the gas phase and in aqueous solution exists solely for the neutral adenine tautomers. For the ionized forms, no common linear relation can be proposed.

Adenine is a good example of natural product for which a good relation between prototropy and electron delocalization for all possible tautomers of neutral adenine can be proposed. The relative energies, which measure the tautomeric conversions, correlate well with the HOMED indices, which measure the geometric consequences of electron delocalization for individual tautomers. The solvent diminishes the subtle internal effects between the functional groups making the deviations of some isomers from the linear relationship smaller in aqueous solution than those in the gas phase. Aromaticity is the main factor that dictates the tautomeric preferences for neutral adenine similar to its parent system – purine [25]. Predominance of the aromatic NH-NH tautomers in the tautomeric mixture of neutral adenine seems to be independent of environment. Positive ionization (one-electron oxidation) does not change this tendency, and resonance conjugations seem to play the principal role for the oxidized adenine tautomers. Negative ionization (one-electron reduction) increases the stability of the non-aromatic NH-CH tautomers (considered as the rare forms for neutral adenine) that some of them are favored in the tautomeric mixture. In this case, the electron affinity is a more important factor than aromaticity and influences the tautomeric preferences [15]. However, the variations of the energetic parameters for the negatively ionized adenine NH-NH and NH-CH tautomers do not significantly change the order of their electron delocalizations.
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