Microvascular imaging and monitoring of hemodynamic changes in the skin during arterial-venous occlusion using multispectral raster-scanning optoacoustic mesoscopy

Amalina Binte Ebrahim Attia a,1,2, Mohesh Moothanchery a,1,2, Xiuting Li a,2, Yik Weng Yew b, Steven Tien Guan Thng c, U.S. Dinish a,2,x, Malini Olivo a,2,x

a Laboratory of Bio Optical Imaging, Singapore Bioimaging Consortium, Agency of Science, Technology and Research (A*STAR), Singapore
b National Skin Centre, Singapore

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A B S T R A C T

The ability to monitor oxygen delivery in microvasculature plays a vital role in measuring the viability of skin tissue and the probability of recovery. Using currently available clinical imaging tools, it is difficult to observe non-invasive hemodynamic regulation in the peripheral vessels. Here we propose the use of a novel multispectral raster-scanning optoacoustic mesoscopy (RSOM) system for noninvasive clinical monitoring of hemodynamic changes in the skin microvasculature’s oxy- (HbO2) and deoxy-hemoglobin (Hb), total hemoglobin (HbT) and oxygen saturation (rsO2). High resolution images of hemoglobin distribution in the skin microvasculature from six healthy volunteers during venous and arterial occlusion, simulating systemic vascular diseases are presented. During venous occlusion, Hb and HbO2 optoacoustic signals showed an increasing trend with time, followed by a drop in the values after cuff deflation. During arterial occlusion, an increase in Hb value and decrease in HbO2 values was observed, followed by a drop in Hb and jump in HbO2 values after the cuff deflation. A decrease in rsO2 values during both venous and arterial occlusion was observed with an increase in value after occlusion release. Using this proof of concept study, hereby we propose multispectral RSOM as a novel tool to measure high resolution hemodynamic changes in microvasculature for investigating systemic vascular diseases on peripheral tissues and also for monitoring inflammatory skin diseases, and its therapeutic interventions.

1. Introduction

Skin microcirculation consists of vessels with diameters < 150 μm, including arterioles, capillaries and venules within the superficial plexus at 400–500 μm depth and deep plexus at 1.9 mm below the skin surface [1,2]. Measuring skin microcirculation and oxidative metabolism is of interest in clinical settings as it provides information to understand multiple skin conditions, not limited to angiogenesis, neovascularization, inflammation, infection, etc [3,4] and systemic co-morbidities such as vascular diseases (e.g. cardiovascular disease) and metabolic syndromes (e.g. diabetes) [5]. Emerging evidence have shown that the cutaneous microcirculation can serve as a representative vascular bed to assess local function by examining the arterioles and venules within a defined region [6]. Systemic vascular dysfunction can be manifested in cutaneous microcirculation [7,8], reflecting the systemic dysfunction in the same extent and mechanism [9], making cutaneous microvasculature a useful surrogate model for studying vascular function and hemodynamic response. Therefore, having a non-invasive skin-specific technique to assess the local vascular function and response is desirable and potentially useful in the assessment and prognosis of dermatologic conditions and systemic vascular comorbidities.

Pulse oximetry is a widespread clinical monitoring tool that provides the measurement of oxygenated hemoglobin percentage in arterial blood by obtaining a ratio of signal in the pulsatile component of arterial blood in relation to that from tissue, venous blood, and non-pulsatile arterial blood. However, the accuracy of this technique decreases at saturation measurements below 70 %, and in hypoxic states experienced...
in vascular diseases such as hypotension, hypothermia and reduced perfusion states due to low-amplitude pulses [10]. This is disadvantageous as the low signal-to-noise ratio in such patients will prevent accurate readings to be obtained where they are needed most. Doppler ultrasound is another widely used tool in clinics to assess blood flow [11] albeit the low contrast to the flow in smaller vessels, makes it suboptimal to measure hemodynamic changes. Magnetic resonance imaging (MRI) and positron emission tomography (PET) are promising ultrasound is another widely used tool in clinics to assess blood flow and can only give an indirect measurement of microcirculations by relating blood flow with the recorded infrared images. SGP is a relatively simpler technique which has been used for many years for point perfusion measurements and thus unable to provide spatial images. As for tcpO2, it employs the principle of inducing local hypoxemia by heating while requiring repeated calibrations and a time-consuming process to set appropriate measurement conditions. Both laser Doppler flowmetry and NIRS have been used to study vascular diseases with relatively low spatial resolutions. NIRS in particular does not measure blood flow directly, complicating the interpretation of tissue oxygenation.

Optoacoustic (photoacoustic) imaging (OAI) on the other hand is a hybrid imaging modality based on the principle of thermoelastic expansion of tissue using pulsed laser excitation and ultrasound signal detection [22]. OAI has shown great potential in scalable deep tissue imaging with high spatio-temporal resolution while providing complementary functional, molecular and structural information of tissue [23]. Due to its distinct optical absorption contrast, it is increasingly playing a pivotal role in clinical and biological research with the ability to resolve main endogenous tissue absorbers i.e. hemoglobin, melanin and water. Optoacoustic signal from hemoglobin can provide vascular structural information and in vivo hemodynamic changes by utilizing the absorption contrast between oxygenated-hemoglobin (HbO2) and deoxygenated-hemoglobin (Hb) [24]. In one such configuration of OAI systems, optoacoustic tomography has been exploited to demonstrate hemodynamic changes in muscle tissue [25,26] and foot [27] with resolutions ≥150 μm. This configuration however is not enough to resolve skin microvasculature in high resolution. Another configuration of OAI system, optoacoustic microscopy could enable microvascular imaging and demonstrated vascular response during arterial occlusion by using two wavelengths [28] while another study reported using a single wavelength on the vascular hemodynamic during venous occlusion [29]. However, the reported optoacoustic microscopy did not manage to resolve the changes in Hb and HbO2 in individual vessels and its relation to venous and arterial occlusion.

Raster-scanning optoacoustic mesoscopy (RSOM) is a novel OAI system configuration that offers ultrawideband ultrasound detection to yield high resolution (10 and 40 μm, axial and lateral resolutions respectively) images up to a few millimeters deep with a single wavelength excitation. It has been successfully employed to clinically image cutaneous inflammatory skin diseases such as psoriasis [31] and atopic dermatitis [32] and nailbed microvasculature [33]. When fitted with a tunable laser, the RSOM system is capable of resolving the tissue absorbers, giving a spatial map and relative concentration of the absorbers [34]. By quantifying the levels of Hb and HbO2 signals, the oxygenation status (rSo2) of the bulk vascular structure and individual vessels can be determined. This multispectral RSOM setup yielded mean oxygen saturation values of 85 ± 4 % in superficial vessels and about 54 ± 7 % in a deeper vessel in the dermis of the forearm, similar to the bulk oxygen saturation values obtained from other optical measurements such as reflectance spectroscopy and NIRS in the same area [34]. Evidently, multispectral RSOM has demonstrated its potential in imaging microvasculature and its oxygenation status at different depths and at specific locations of the skin layers, unlike pulse oximetry. The quantification of the vascular features is also possible down to individual vessels in the 3D images while pulse oximetry gives a bulk saturation value. Because optoacoustic imaging is not dependent on pulsatile of blood vessels and rather on the absorbance of chromophores, their quantification of hypoxic states is still possible from the spatial Hb and HbO2 signals. The aim of this study is to demonstrate the capability of high-resolution multispectral RSOM technique for monitoring hemodynamic changes in the microvasculature of the forearm during cuff occlusion, simulating ischemic and thrombosis conditions.

2. Methods

2.1. Subjects and protocols

Healthy adult volunteers (n = 6) of Fitzpatrick skin types II to IV participated in this study which was approved by National Healthcare Group Domain Specific Review Board 2020/00079, Singapore. In this study, the subjects were in a sitting position on a chair with their arm outstretched and resting at heart level. The imaging head was placed by a flexible mechanical arm over the right forearm at mid-point between hand and elbow on the volar forearm. A blood pressure cuff was attached around the right upper arm. The forearm was upward and supported by a pillow. The cuff occlusion protocol for the subjects are detailed in Fig. 1. Briefly, the venous-arterial occlusion challenges comprised of seven stages. First, a baseline image was captured from the volunteer in the resting state before occlusion (i). Second, venous outflow was stopped using an occlusion cuff, by inflating the cuff up to 80 mm Hg pressures and an image was captured at 1 min venous occlusion (ii). Third, another image was captured at 3 min venous occlusion (iii). Fourth, the occlusion cuff was deflated rapidly and an image was captured at 2 min post-release (iv). Fifth, both venous outflow and arterial inflow was stopped by inflating the cuff and an image was captured at 1 min arterial occlusion (v). The arterial occlusion pressure used was 40 mm Hg more than that of systolic pressure. Sixth, another image was captured at 3 min arterial occlusion (vi). Finally, the occlusion cuff was deflated rapidly and an image was captured at 2 min post-release (vii). To minimize movements of the imaging area, the arm was rested on a flat surface while the volunteers sat on a chair without wheels comfortably.

2.2. Multispectral optoacoustic mesoscopic imaging system

An RSOM Explorer ms-CS50 system (iThera Medical GmbH, Munich, Germany) was used for optoacoustic imaging. Briefly, the skin was illuminated with a multispectral, nanosecond pulsed, Raman laser having four different wavelengths (532, 555, 579, 606 nm) and 2.5-3.2 ns pulse width. The time difference between successive pulses is in the range of 0.5-2 ms. The repetition rate of the laser can be varied between 325 Hz per wavelength to a maximum of 1.3 kHz for all wavelengths, depending on the field of view and number of wavelengths used. The laser light with a pulse energy ~25 μJ was delivered through a single customized fiber for optimum light illumination which combined with a broadband spherically focused transducer (50 MHz center frequency, 11–99 MHz bandwidth, 4 mm focal diameter, 4 mm active element distance, f-number: 1). The combined illumination-detector system was attached to a two axis motorized stage enabling raster scanning the two-dimensional (2D) field of view of up to 6 × 6 mm with a step size of 20 μm, which forms the scanning head. The scanning head was attached at the end of an articulated arm for optimal positioning. A water-filled unit with the bottom covered with an interchangeable polyethylene
membrane will connect the scan head to the skin surface by means of ultrasound gel. All the imaging parameters were within maximum permissible exposure (MPE) limits and the system was equipped with a key switch, a laser safety foot pedal and an emergency off button for safe operation of the laser. The data acquired were amplified using two low noise amplifiers of 30 dB (one pre and one post data detection).

2.3. Image processing

The recorded optoacoustic signals were subsequently reconstructed with a beam-forming algorithm by assuming that at the focal point of the transducer, a point detector is located, which accepts signals within a cone with its angle defined by the numerical aperture of the transducer. To facilitate the visualization of the skin layers and vascular structures, the skin surface was identified and then flattened with motion correction to reduce the motion effects from the subjects [35,36]. The flattened and motion corrected reconstructed images were unmixed by linear regression for different absorbers namely melanin, Hb, and HbO$_2$ using all four available wavelengths. Two frequency sub-bands are divided from optoacoustic signals for concurrent visualization of bigger and smaller vascular structures, in which low frequency band (LF) was colored in red from 11 to 33 MHz representing bigger structures, high frequency band (HF) was colored in green from 33 to 99 MHz representing smaller structures. The 3D volume images and maximum intensity projection (MIP) images along the depth direction ($z$-axis) were generated for further processing. By averaging all the pixels in the $x$-$y$ plane along the depth, the multispectral RSOM optoacoustic profile along the skin depth was obtained. The depth ranges of epidermis (E) and dermis (D) regions was segmented manually based on the optoacoustic MIP profiles along the depth direction.

2.4. Data analysis

The quantification of each absorber’s optoacoustic signal was calculated by averaging all the voxels in the 3D volume of multispectral RSOM images. The total hemoglobin (HbT) and oxygen saturation (rsO$_2$) were then calculated by the equations Hb + HbO$_2$ and HbO$_2$/ (Hb + HbO$_2$) respectively. The plots of absorber values for all the subjects at various time points over the period of occlusion challenges were presented. Data were presented as mean ± standard error of the mean (SEM) values. Two-sided paired t-test was used to compare variables with $p < 0.05$ regarded as statistically significant. The calculation was also applied on a small demarcated area with individual vessels of one subject to validate the sensitivity of multispectral RSOM, and on LF and HF bands of the same subject to explore the bigger and smaller vascular structures changes during the occlusion challenges.

3. Results and discussion

3.1. Multispectral optoacoustic images of the forearm during occlusion challenge

Fig. 1 shows the cuff occlusion challenge protocol and the various discrete time points at which multispectral RSOM imaging of the forearm was acquired during the occlusion challenge, labelled as (i) to (vii). The corresponding and representative multispectral RSOM MIP images are shown in Fig. 2 with each hemoglobin absorber shown separately and merged together with melanin absorber in the $xz$ plane. The blood oxygen saturation (rsO$_2$) maps of the MIPs at each time point are also shown in Fig. 2. The multispectral RSOM was capable of resolving the structural architecture of the skin with the images showing melanin in uppermost epidermis (green) layer with a present hair shaft followed by the dermis layer underneath. The epidermal layer is characterized mainly by strong absorption of melanin signals with minimal presence of hemoglobin absorption. On the other hand, the contribution from melanin in the dermis was not present, as expected. The vascular plexus network could be visualized in the dermis showing the spatial maps of optoacoustic signals of HbO$_2$ and Hb, from which the rsO$_2$ map could be derived. At baseline (i), the rsO$_2$ values for the representative case in Fig. 2 were in the range between 65 % and 92 % over the skin depth.
In general, the Hb optoacoustic signals increased in intensity (blue signals) during occlusion of venous or arterial flow and dropped close to baseline values 2 min post-release of occlusion (Fig. 2). The time point at which the Hb optoacoustic signals were the highest was at (iii) at which 3 min venous occlusion occurred compared to the arterial occlusion challenge. On the other hand, the HbO$_2$ optoacoustic signals peaked at time point (iii) with vessel dilation present for some individual vessels and was the lowest at time point (iv) (Fig. 2). In the rsO$_2$ spatial maps, the rsO$_2$ change during the venous and arterial occlusion challenges was only observed in the dermis as vascular supply is not present in the epidermis. The baseline (i) rsO$_2$ values in the dermis were above 50 % in the heat map while an increase of signals signifying less than 50 % rsO$_2$ was observed during both occlusions in the spatial maps. Notably, the 3 min arterial occlusion time point (vi) exhibited the highest number of signals signifying less than 50 % rsO$_2$.

Depth-dependent oxygen saturation can be observed from the spatial heat maps of rsO$_2$ values in Fig. 2. At time points (i) and (vii) i.e. the baseline and recovery from occlusion challenge respectively, the highest rsO$_2$ values were seen in the vessels of superficial plexus just underneath the epidermis. During arterial occlusion (time points (v) and (vi)), the highest rsO$_2$ values were observed in the deeper vessels instead. Atmospheric oxygen being taken up by the skin through diffusion is well-documented, reaching a depth of 0.25 to 0.40 mm from the skin surface [37] explaining why the superficial plexus vessels exhibited the higher oxygen saturation values at baseline and post-occlusion.

### 3.2. Venous occlusion challenge

Fig. 3 shows the mean optoacoustic signals of Hb, HbO$_2$, HbT and calculated rsO$_2$ values over the occlusion challenges for the subjects ($n = 6$). During the venous occlusion challenge (time points (i) to (iv)) whereby the venous outflow is impeded while the arterial supply is retained, an increase in Hb, HbO$_2$ and HbT optoacoustic signals was observed (Fig. 3A-C). The maximum Hb, HbO$_2$ and HbT optoacoustic signals occurred at time point (iii), i.e. the 3rd minute of venous occlusion (increase by 403 %, 41 % and 112 % respectively compared to baseline (i)). The increase in HbT was probably due to the increase in blood volume when the blood accumulates from the blocked venous outflow during the occlusion. Notably, the increase in Hb signals is more drastic than that of HbO$_2$ signals. At time point (iv) i.e. 2 min post cuff release, the Hb, HbO$_2$ and HbT optoacoustic signals decreased from their peak values at time point (iii) to 38.9 %, -17.9 % and -5.9 % of baseline (i) values respectively, indicating venous drainage had resumed.

The mean oxygen saturation level decreased from 74.2 % at baseline to 58.9 % at 1 min post occlusion before further decreasing to 50.2 % at 3 min post-occlusion (Fig. 3D). While both Hb and HbO$_2$ signals increased during venous occlusion due to the accumulation of blood, the drastic increase in Hb signals compared to that of HbO$_2$ signals resulted in a decrease in oxygen saturation levels during the occlusion period. This can be attributed to the disruption of transport of deoxygenated blood from the peripheral blood vessels back to the lungs for the exchange of carbon dioxide for oxygen while blood is accumulating during the occlusion. The increase in HbO$_2$ signals was slower probably due to the increasing intraluminal blood pressure with the increased volume of
blood which can gradually reduce the arterial inflow. At 2 min post cuff release (iv), the oxygen saturation level rebounded to a mean value of 66.2 % while still not recovering near to the baseline values \((p = 0.024)\) compared to baseline values. This can be attributed to the fact that HbO\(_2\) signals only recovered to below baseline values upon cuff release. As venous drainage resumes after cuff release, the blood volume will decrease, leading to the decrease of Hb and HbO\(_2\) signals. However, the strong drainage of the accumulated blood was probably propelled by the intraluminal blood pressure achieved during the venous occlusion challenge, thereby reducing the HbO\(_2\) signals before ultimately achieving baseline values.

### 3.3. Arterial occlusion challenge

During the arterial occlusion challenge (time points (iv) to (vii)) whereby both the venous outflow and arterial inflow are interrupted, an increase in Hb and a decrease in HbO\(_2\) optoacoustic signals were observed (Fig. 3A-B). At the 3\(^{rd}\) min of the occlusion challenge i.e. time point (vi), Hb signals increased to 64.4 % of that of time point (iv) while HbO\(_2\) signals decreased by 32.5 % of that of time point (iv). HbT optoacoustic signals on the other hand remained steady (within 2.5 % of that of time point (iv)) throughout the occlusion challenge period as no change in blood volume was expected with no outflow and inflow of blood occurring (Fig. 3C). The simultaneous increase in Hb and decrease in HbO\(_2\) signals during arterial occlusion is indicative of the blood oxygen consumption in the blood vessels with no supply of oxygenated blood being delivered i.e. towards tissue hypoxia. Upon release of the cuff, both Hb and HbO\(_2\) signals returned within 10 % of that of time point (iv). While it is expected to observe a rebound hyperemia effect in response to pro-vasodilatory molecules secreted to increase blood flow to meet the oxygen demand, this was not observed in this study. The RSOM ms-C50 system is limited to time-point measurements due to the raster-scanning of the imaging site during image acquisition and the hyperemia effect may have been only apparent immediately post-cuff release which was not captured during image acquisition.

Similar to the venous occlusion challenge, the mean oxygen saturation level decreased during the arterial occlusion challenge, from 66.2 % at time point (iv) to a minimum of 47.6 % at 3 min of arterial occlusion (time point (vi)) before recovering to 69.4 % at 2 min post cuff release (Fig. 3D). The decrease in oxygen saturation level is due to the cutaneous oxygen consumption in maintaining metabolism in the vessels with limited delivery of oxygenated blood. Notably, the mean oxygen saturation values during arterial occlusion is lower than that of venous occlusion probably due to the interrupted inflow of oxygenated blood in the former.

### 3.4. Sensitivity of multispectral RSOM imaging

While Fig. 3 was obtained from bulk calculations of Hb and HbO\(_2\) signals from the acquired multispectral RSOM images, we also investigated the sensitivity of the system on measuring the hemodynamic variations in single vessels in a small area. The Hb, HbO\(_2\) and HbT signals in a small area of the vascular plexus as demarcated in Fig. 2 in a white dashed box were calculated for all the time points in one representative subject (Fig. 4). Fig. 4A shows that in a small area of interrogation for one subject, the trends of Hb, HbO\(_2\) and HbT hemodynamic changes were consistent with that of Fig. 3, which was calculated from the bulk vasculature in the images and averaged among all the subjects. The Hb, HbO\(_2\) and HbT signals peaked at time point (iii) during the 3\(^{rd}\) min of venous occlusion in Fig. 4A. The Hb signals increased during arterial occlusion whereas the HbO\(_2\) signals decreased during the challenge instead compared to that of time point (iv). Similarly, the rsO\(_2\)
values of the vessels in the small area followed the same hemodynamic variation with that of the bulk vasculature. The rsO$_2$ values decreased from the baseline value during both occlusion challenges with the lowest being at time point (vi) i.e. 3$^{rd}$ min of arterial occlusion. Recovery of oxygen saturation after venous occlusion did not reach baseline values while recovery after arterial occlusion did, similar to that observed in the bulk quantification of oxygen saturation. Thus, the multispectral RSOM technique demonstrates promise and sensitivity for visualizing and quantifying the hemodynamic responses in single vessels in a small area of interrogation.

In Fig. 3D, a decrease of 26 % in the rsO$_2$ was observed between baseline and its minimum value at time point (vi). In comparison, a decrease of 45 % in the rsO$_2$ values was observed between baseline and that of time point (vi) in Fig. 4B. While the hemodynamic trends of the vessels in the small area were comparable to that of bulk vasculature, this difference in rsO$_2$ reductions between the vessels in the small area and bulk vasculature is significant. Firstly, this is probably due to the difference in the relative vasculature signal in relation to the background signals in the region-of-interest in both the small area and bulk volume. The defined small area was vasculature rich i.e. the density of the Hb and HbO$_2$ signals were higher compared to the non-vascular background. This will enhance the contrast between the vasculature and background hemoglobin signals. Thus, the reduction of rsO$_2$ during the occlusion challenge was more pronounced in the small area than that of the bulk volume. One way to address this limitation is by segmenting the vessels from the background in rsO$_2$ calculations. This requires a higher frequency transducer to effectively increase the resolution of the outline of the vascular structures for easier segmentation. Secondly, the selected small area in which the rsO$_2$ were quantified was near the skin surface compared to the overall bulk vasculature measurements in the Fig. 4.

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**Fig. 4.** Validating sensitivity of multispectral RSOM. Dotted white box in Fig. 2 showing the area with individual vessels for tracking the (A) Hb, HbO2 and HbT signals and (B) rsO$_2$ values in one small area in one representative subject over the time points. Time points (i) to (iv) corresponds to venous occlusion challenge with time point iv representing release of the occlusion. Time points (iv) to (vii) corresponds to arterial occlusion challenge with time point vii representing release of the occlusion.

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**Fig. 5.** The ultrawide detection bandwidth of multispectral RSOM divided into low (LF) and high frequency (HF) bands. (A) Representative cross-sectional multispectral RSOM images at various time points (i to vii) of venous and arterial occlusion challenges with concurrent visualization of LF band (11 – 33 MHz) in red and HF band (33 – 99 MHz) in green at single wavelength 532 nm. Bigger vascular structures are shown in LF band while smaller vascular structures in HF band. Plots of unmixed Hb, HbO$_2$ and HbT optoacoustic signals for (B) LF and (C) HF bands for the subjects (n = 6) over the occlusion challenges period (mean ± SEM). Scale bar; 500 μm.
3D volume. As the incident light reaching the dermis gets lower with the increasing depth, the changes in rsO2 values of the deeper dermis vessels captured from optical imaging would be less sensitive. Therefore, the rsO2 values in the selected small area will vary significantly during occlusion compared to the averaged values in the overall 3D bulk volume. Vogt et al. have similarly reported that the accuracy of oxygen saturation measurement is depth-dependent, which is attributed to fluence artifacts [38].

3.5. Low and high-frequency bands of multispectral RSOM images

With a broadband transducer fitted in the multispectral RSOM system, the detected ultrasound signals can be divided into two separate frequency bands for better visualization. The cross-sectional multispectral RSOM images for the same representative subject is shown in Fig. 5A in which the low (LF) and high frequency (HF) components acquired at single wavelength 532 nm are labelled in red and green respectively. The plots of the LF and HF components of the unmixed Hb, HbO2 and HbT signals are shown in Fig. 5B-C. Larger vascular structures are shown in LF band (~20 to 50 μm in size for 11–33 MHz) while smaller vascular structures are visualized in HF band (~10 to 20 μm in size for 33–90 MHz). Any overlap of both frequency bands will result in yellow signals. The multispectral RSOM images in Fig. 5A revealed a depth-dependent variation of blood vessel size. In the cross-sectional images, the HF band signals are present in the vessels immediately underneath the epidermis. In the depth axis direction, the frequency band signals turn to yellow indicating a presence of LF band signals followed by predominantly red LF band signals deeper to the hypodermis. This was especially apparent in time point (iii) of Fig. 5A. It is known that arterioles deeper in the dermis are larger in diameter than that near the dermal papillae.

During venous occlusion, the LF and HF signals at 532 nm (Fig. 5A) and of HbT absorber (Fig. 5B-C) increased in intensity compared to that of baseline with the signals most intense at time point (iii) (209 % and 167 % change from baseline, respectively for HbT). The signals decreased upon release of occlusion at time point (iv). However, during arterial occlusion, no discernible differences in the LF and HF signals at 532 nm were observed among time points (iv) to (vii) (Fig. 5A). While it was shown previously that accumulation of blood occurs during venous occlusion, it demonstrates herein that both small and big vessels exhibited the same hemodynamic response to the venous occlusion challenge. The increase in both LF and HF signals signify vessel dilation occurring upon obstruction of venous outflow. As both outflow and inflow of blood was impeded in arterial occlusion, no change in LF and HF signals were expected. Similarly, when the individual absorber signals were divided into LF and HF bands in Fig. 5B-C, both frequency bands show similar trend of hemodynamic response for Hb, HbO2 and HbT absorbers.

4. Conclusion

In this study, we present a novel high-resolution multispectral optoacoustic system and its application to image skin microvasculature and monitor hemodynamic changes. The system performance has been validated under physiological interventions such as venous and arterial cuff occlusion challenges to simulate systemic vascular conditions in the skin from six healthy volunteers. During both venous and arterial occlusion, Hb optoacoustic values showed an increasing trend with time compared to baseline, whereas HbO2 values showed an increasing trend during venous occlusion, and decreasing trend during arterial occlusion, compared to baseline. The rsO2 values decreased during both venous and arterial occlusions. The optoacoustic absorber signals and rsO2 values recovered near to baseline values upon release of occlusion. An accumulation of blood was observed during venous occlusion in both smaller and bigger vessels in the vascular plexus leading to their dilation during the occlusion challenge. The hemodynamic trends were also observed in single vessels in a small defined region of monitoring, corroborating multispectral RSOM’s sensitivity to detect absorber signals in single vessels. Thus, multispectral RSOM has demonstrated its potential as a novel clinical imaging tool to measure hemodynamic changes in peripheral tissue microvasculature. Understanding the hemodynamic response in the vessels of peripheral tissues using imaging techniques present a novel way of non-invasive investigation of systemic vascular diseases. Additionally, the ability to monitor microcirculation and changes in blood oxygenation at different depths below the skin surface at high resolution opens the window of opportunity for non-invasive evaluation of various inflammatory skin diseases and conditions in which vascular remodeling is commonly observed [39]. In this context, we envision that this study will open up new avenues in understanding inflammatory skin conditions such as eczema and psoriasis and its associated comorbidities.

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Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Steven Tien Guan Thng (Assoc Prof) graduated from National University of Singapore in 1992 and went on to pursue his interest in Humanitarian and Disaster medicine with the Singapore Armed Forces from 1992 to 2002. After completing his dermatology training in Singapore, Dr Steven Thng went on to Amsterdam Medical Centre, Netherlands, to continue his training in pigmentary disorder, under Professor Westerhof. Upon returning from his training, Dr Steven Thng started the pigment clinic in National Skin Centre as well as set up the melanocyte culture lab to start his work on cultured melanocyte grafting for vitiligo patients. Currently, Dr Thng is the head of the pigment clinic, in charge of managing all complex pigmentary disorders in National Skin Centre, and he also the principal surgeon for tissue grafting for vitiligo. In 2012, he has also started a research clinic for hyperpigmentary disorders exploring novel ways to manage difficult hyperpigmentary conditions. He took over as Executive Director of Skin Research Institute of Singapore from Jan 2017 to Feb 2020, and was responsible for the transformation of SRIS from a virtual institute to a dynamic institute, working on 4 main multi-institutional, interdisciplinary research programs of chronic wound, acne, skin microbiome and atopic dermatitis. His current research interest is on skin pigmentary disorder, novel drug delivery systems, as well as advanced imaging for skin cancer and skin diseases. He has translated much of his research to improve patient care in dermatology. He has been serving as the Chief Dermatologist of Skin Research Institute of Singapore from Feb 2020. For his dedication in research and patient care, he was awarded Excellence in Public Service Award in 2013, Healthcare Humanity Award in 2014, Singapore Clinician Investigator Award in 2015 and 2016.