Effect of Physical and Chemical Mutagens on Morphological Characters in M1V2 Generation of Tuberose (Polianthes tuberosa L.)

K. Kayalvizhi*, M. Kannan and M. Ganga

Department of Floriculture and Landscaping, HC & RI, TNAU, Coimbatore-3, India

*Corresponding author

Abstract

An investigation was carried out at the Department of Floriculture and Landscaping, Horticultural College and Research Institute, Tamil Nadu Agricultural University, Coimbatore during 2014-2016 on the improvement of tuberose var. Prajwal through mutation breeding. The bulbs were treated with gamma rays, diethyl sulphate (DES) and ethyl methane sulphonate (EMS). The treatments consisted of 0.5, 1.0, 1.5, 2.0 and 2.5 kR of gamma rays, 15, 20, 25 and 30 mM of DES and 30, 45, 60 and 75 mM of EMS and control (untreated). Various morphological and floral characters were observed. In general, the treated population had manifested reduced expression than the control (untreated population) for most of the morphological and floral characters. Higher the dose of mutagens, lower was the expressivity of the traits. Expression of the morphological characters namely plant height, number of leaves, leaf length, leaf width and leaf thickness increased in the lower doses and decreased in the higher doses in M1V2 generation.

Keywords
Gamma rays, EMS, DES, Tuberose, Prajwal, Morphological variations and M1V2 generation

Introduction

Floriculture in India is estimated to cover an area of 2.55 lakh ha with a production of 17,54,000 MT of loose flowers. Nearly 77% of the area under floricultural crops is concentrated in seven states comprising Tamil Nadu, Karnataka, West Bengal, Maharashtra, Haryana, Uttar Pradesh and Delhi. Among different states, Tamil Nadu ranks first in area followed by Karnataka, West Bengal and Maharashtra. In Tamil Nadu 3,43,650 MT of loose flowers are produced in an area of about 55,000 hectares. (Anon., 2015).

Tuberose (Polianthes tuberosa L.) is one of the most important flowers used for both cut and loose flower purpose. It is extensively cultivated in many sub-tropical and tropical parts of the world including India. It is a native of Mexico and belongs to the family Amaryllidaceae. It is a bulbous perennial plant with tuberous roots producing long spikes, bearing waxy white fragrant flowers. It is a crop which flowers profusely throughout the year. Due to the longer keeping quality of flower spikes (Benschop, 1993), they are in great demand for making floral arrangements and bouquets in major cities of India. Three types of tuberose which are used in cultivation are single type with one row of corolla segments, semi-Double type with two to three rows of corolla segments and double type with more than three rows of corolla segments.
The spikes as a whole in double types can be used as cut flowers whereas the florets of single varieties are used for making garlands, veni, gajra, bangles, etc. and also for essential oil extraction. The flower yields a very valuable floral concrete (0.08 – 0.11 per cent) upon solvent extraction (Singh, 1995). The absolute of tuberose (essential oil) extracted from floral concrete is used in the preparation of various high value perfumes and cosmetics.

The main emphasis in flower breeding is to improve the varietal traits viz., colour, flower form, size, number of flowers, shelf life, vase life, year round production and growth habit.

Mutation breeding stands for the genetic improvement of crop plants for various economic characters by using physical and chemical mutagens. Mutation is the sudden heritable change that occurred in an organism. It may be caused by spontaneous or through artificial induction and the resulted mutant shows change in the gene or chromosomes. (De and Bhattacharjee, 2011).

Induced mutagenesis has been most successful in ornamental crops. Both physical and chemical mutagens have been used for improving the desired characters of many ornamental crops including amaryllis, asiatic hybrid lily, bougainvillea, chrysanthemum, dahlia, gladiolus, hibiscus, Lantana, marigold, rose, tuberose, gerbera, narcissus, etc. Induced mutations in ornamentals comprise traits, such as altered flower characters (colour, size, morphology, fragrance), leaf characters (form, size, pigmentation), growth habit (compact, climbing, branching) and physiological traits such as changes in photoperiodic response, early flowering, free flowering, keeping quality and tolerance to biotic and abiotic stresses. The main advantage of mutation breeding in vegetatively propagated crops is the ability to change one or a few characters of an otherwise outstanding variety without altering the unique part of the genotype (Datta, 2014).

In any mutation breeding programme, selection of an effective and efficient mutagen is very essential to produce high frequency desirable mutants. Several factors such as properties of mutagens and pH, duration of treatment, temperature etc. play an important role to produce a desirable mutant.

Mutations are induced by physical and chemical mutagen treatment in both seed and vegetative propagated crops. The mechanism of mutation induction is that the mutagen treatment will break the nuclear DNA and during the process of DNA repair mechanism, new mutations may occur randomly and are heritable. It is a simple, efficient, rapid and cheap option for obtaining desired genotypes from recalcitrant species. Induced mutation is one of the most widely used techniques for creating additional variability in desirable character. It can be done by physical and chemical mutagens.

In physical mutagens, atoms are the principle source material. Unstable atoms of same element having different weights giving energy or particles are called radioisotopes and electromagnetic waves associated with nuclear decay are called as radiation and the treatment of an organism or plant with radiation is known as irradiation. It is classified into two groups ionizing and non ionizing radiations. Alpha rays (α), Beta rays (β), X-rays, Gamma ray (γ) and Neutrons belongs to the group of ionizing radiation. Non ionizing radiation includes UV rays only. Ionizing radiations normally causes chromosomal rearrangements and deletions. (Bhat et al., 2007). Gamma rays are electromagnetic radiations having shorter wavelength than X rays with more energy and penetrating power. It is produced by a number of isotopes e.g. $^{14}$C, $^{60}$Co, $^{137}$Cs etc. for
chronic treatments requiring slow irradiation over long periods. (De and Bhattacharjee, 2011).

Mutation can also be induced chemically with alkylating agents such as Diethyl sulphate (DES) and Ethyl methane sulphonate (EMS) etc. The alkyl group of chemical mutagens reacts with DNA which may change the nucleotide sequence and cause a point mutation. (Broertijes and Harten, 1988). EMS alkylates are guanine bases and leads to mispairing-alkylated G pairs with T instead of C, resulting in primarily G/C to A/T transitions (Bhat et al., 2007).

Tuberose being a cross pollinated crop, there is need for high yielding variety with improved fragrance to overcome farmer’s predicament. Mutation breeding is the most effective and commonly employed tool to induce acceptable variations in the existing cultivars viz., high yielding and better quality of genotypes (Bhattacharjee, 2006). The present study was undertaken to induce desirable variations in tuberose using physical and chemical mutagens.

**Materials and Methods**

A study was conducted at the Department of Floriculture and Landscaping, Horticultural College and Research Institute, Tamil Nadu Agricultural University, Coimbatore during 2015-16 on the improvement of tuberose (*Polianthes tuberosa* L.) var. Prajwal through mutation breeding. The experimental site is geographically situated at an altitude of 426.72 metres above mean sea level (MSL) and between 11°02’” North Latitude and 76°57’” East Longitude. Bulbs of tuberose var. Prajwal (2-3 cm diameter) were subjected to gamma and chemical mutagens treatment.

Prajwal is a hybrid of tuberose (Shringar x Mexican Single) developed by IIHR, Bangalore. It is a single type with greenish white flower buds with dark pink tinge at the tips. It yields 18 t/ha/ year. It is ideal as loose flower and cut flower.

One physical mutagen (gamma ray) and two chemical mutagens viz., Diethyl Sulphate (DES) and Ethyl Methane Sulphonate (EMS) were used in the study.

For gamma irradiation of bulbs, the Gamma chamber - 1200 available at the Centre for Plant Breeding and Genetics of Tamil Nadu Agricultural University, Coimbatore, installed and maintained by the Board of Radiation and Isotope Technology (BRIT), DAE, Mumbai. Cobalt - $^{60}\text{(Co)}$ emitting 5000 rads per minute at the time of irradiation was used. The formula suggested by Kodym and Afza (2003) was used for the calculation of duration of exposure.

Diethyl Sulphate ($\text{C}_2\text{H}_2\text{O}_2\text{SO}_2$) with a molecular weight of 154.19, boiling point 208° C (lit.), density =1.777 g/ ml at 25°C (lit.) was obtained from M/s Sigma-Aldrich Company, U.S.A. Prior to use, it was removed from refrigerator and placed in a desiccator with calcium chloride until room temperature was reached.

Ethyl Methane Sulphonate ($\text{CH}_3\text{SO}_2\text{OC}_2\text{H}_5$) procured from M/s. Sigma-Aldrich Company, U.S.A was used. It has a Molecular weight-124.16, boiling point of 80/100 mm Hg and Density of $\text{D}_{4}^{25}$ = 1.203 g/ml). It was stored in dry air at 0°C to maintain its purity. Prior to use, it was removed from refrigerator and placed in a desiccator with calcium chloride until it reached room temperature. The treatment details are furnished below (Table 1).

Cultural practices were followed as per the Crop Production Manual, TNAU, 2012. The recommended fertilizer dose of 200:200:200 kg ha$^{-1}$ of NPK was applied. Half of the RDF was applied as basal and the remaining half
was applied in two splits at 30 and 45 days after planting respectively. Foliar spray of micronutrients (H$_3$BO$_3$ @ 0.1 % + ZnSO$_4$ @ 0.5 % + FeSO$_4$ @ 0.2 %) was given four times at 60, 120 and 180 days after planting (Ganesh, 2010). The observations recorded on growth, yield and quality parameters in M$_1$V$_2$ generation. For individual plants, the morphological variations were observed upto 200 days after planting.

**Results and Discussion**

Various morphological and floral characters were observed for the mutagen treated plants. In general, the treated population had manifested reduced expression than the control (untreated population) in most of the morphological and floral characters. Higher the dose of mutagens, there was reduction in the expressivity of the traits.

The expression of the morphological characters plant height, number of leaves, leaf length, leaf width and leaf thickness increased in the lower doses and decreased in the higher doses in the M$_1$V$_2$ generation. Similarly, days taken for spike emergence and first floret opening were observed earlier at lower doses. The floral and yield characters namely spike length, rachis length, floret length and diameter, number of spikes per plant, number of florets per spike, weight of single floret and flower yield per m$^2$ were observed maximum at lower doses and were minimum in the higher doses.

A total of 1617 plants were examined until 200 days to isolate the chlorophyll mutants. Chlorophyll mutants noticed in M$_1$V$_2$ generation are of four types viz. ‘albino’ ‘chlorina’ ‘striata’ and ‘xantha’ (Plate 1). The treatment T$_3$ (1.0 kR gamma ray) recorded maximum number of chlorophyll mutants and it was followed by T$_4$ (1.5 kR gamma ray) and T$_{12}$ (45 mM of EMS). Similar observations were made by Sambanthamurthi, (1983) and Kumar et al., (2013) in tuberose; Banerji and Datta (1998) and Sisodia and Singh (2014) in gladiolus.

**Table 1** Treatment details

| Treatment | Dose of physical and chemical mutagens |
|-----------|---------------------------------------|
| T$_1$     | Untreated control                      |
| T$_2$     | 0.5 kR gamma rays                      |
| T$_3$     | 1.0 kR gamma rays                      |
| T$_4$     | 1.50 kR gamma rays                     |
| T$_5$     | 2.00 kR gamma rays                     |
| T$_6$     | 2.50 kR gamma rays                     |
| T$_7$     | 15 mM DES                              |
| T$_8$     | 20 mM DES                              |
| T$_9$     | 25 mM DES                              |
| T$_{10}$  | 30 mM DES                              |
| T$_{11}$  | 30 mM EMS                              |
| T$_{12}$  | 45 mM EMS                              |
| T$_{13}$  | 60 mM EMS                              |
| T$_{14}$  | 75 mM EMS                              |
### Table 2: Morphological abnormalities

| Treatment | Variation / morphological abnormalities recorded in M₁V₂ generation |
|-----------|---------------------------------------------------------------------|
| T₃        | Absence of pink tinge on tip of the floret                         |
| T₄        | Leaf abnormalities such as merging of two leaves                   |
| T₅        | Leaf vein, single floret per node, fusion of two florets, uneven size of florets and broader stamen |
| T₆        | Two rows of florets in single type variety                         |
| T₇        | Sickle shaped leaf and tepal elongation                            |
| T₈        | Crinkled, lobed leaf, leaf vein, tepal serration, sharp and bent floret tip and pink colour of whole bud |
| T₉        | Leaf vein, bent floret tip                                         |
| T₁₀       | Leaf vein, spike abnormalities                                     |
| T₁₁       | Uneven size of florets                                            |

**Plate 1:** Chlorophyll mutants in M₁V₂ generation

- T₃ (1 kR gamma ray) - Striata
- T₃ (1 kR gamma ray) - Albino and Xantha
- T₄ (1.5 kR gamma ray) - Striata
- T₄ (1.5 kR gamma ray) – Broad leaf formation
Plate.2 Floret tepal variations in M₁V₂ generation

Broad leaf mutants were observed in 2.5 kR gamma ray with a leaf width of 2.53 cm and T₇ (15 mM DES) 2.87 cm. Branched leaf mutants were observed in 1.0 kR and 2.0 kR. These variations may be useful for landscape purpose twing to their enhanced aesthetic values. These findings are similar to those of Singh et al., (2013) in tuberose.

A non flowering spike was recorded in 25 mM DES. Similar result was reported by Sambanthamurthi (1983).

Floral mutants

Four tepal floral mutants were observed in all the treatments except T₁ (control) and T₂. Five tepal florets were also observed in all the treatments except T₁ (control), T₂, T₅ and T₆. Similarly, all the treatments except T₂ and T₃ produced seven tepal florets but in control plants only six tepal florets were produced. Likewise, eight tepal florets were observed in all the treatments except T₅. Nine tepal florets were observed in T₁₀ (30 mM of DES).
Eleven tepal floret was observed in T<sub>7</sub> (15 mM DES) (Plate 2). These findings are in accordance with those of Datta (1977), Van Harten (1998) and Datta (2000) in ornamentals and Anu <i>et al.</i>, (2003) and Kainthura and Srivastava (2015) in tuberose (Table 2).

These findings are in line with the morphological abnormalities in the foliage and florets observed by earlier workers in irradiated material of gladiolus Banerji and Datta (1998), Kumar <i>et al.</i>, (2013); Banerji <i>et al.</i>, (2000) and Singh <i>et al.</i>, 2013 gladiolus.

In conclusion, as chimerism and genetic variability play a key role in the variation observed in mutation treated population, there is a need to identify solid mutants in the future generations.

References

Anonymous, 2015. Present status and prospects of floriculture in Tamil Nadu. DFR technical Bulletin No. 20. ICAR – Directorate of floricultural research, Pune, Maharashtra, India.

Anu, K.G., Geetha, C.K., Rajeevan, P.K., Valsalakumari, P.K., Saifudeen, N. 2003. Induced mutation in tuberose (<i>Polianthes tuberosa</i> L.) by gamma rays. In: P.K. Rajeevan, P.K. Valsalakumari and R.L. Misra (eds.). One Hundred Research Papers in Floriculture. Indian Society of Ornamental Horticulture. pp. 255-259.

APEDA. 2016. http://apeda.gov.in/apedawebsite/six_head_product/floriculture.html.

Banerji, B.K. S.K. Datta. 1998. Improvement of gladiolus by induced mutations. Abst: National Symposium on Nuclear and Allied Techniques in Agriculture, Medicine and Environment Research. IARI, New Delhi. p. 11.

Banerji, B.K., A.K. Dwivedi and S.K. Datta. 2000. Gamma ray induced variability in gladiolus. Abst: National Conference on Gladiolus. NBRI, Lucknow. No. 8.

Benschop, M. 1993. Polianthes, In: De Hertogh A., Le Nard M., (Eds.), The physiology of flower bulbs, Elsevier, Amsterdam, The Netherlands, pp. 589-601.

Bhat, R., N. Upadhyaya, A. Chaudhury, A. Ragathan, C. Qiu, F. Wang, H. Wu, J. Mcnally, K. Leiung and B. TILL. 2007. Chemical and irradiation induced mutants and tillage. Rice functional genomics, p 148-180.

Bhattacharjee, S. K. 2006. Development and Research on Ornamental Horticulture In: Advances in Ornamental Horticulture, 1. Pointer Publishers, Jaipur. pp.1-33.

Broertjes, C. and A.V. Van Harten. 1978. Application of mutations breeding methods in the improvement of vegetatively propagated crops. Elsevier Scientific Publishing Company, New York.

Datta, S. 1977. Improvement of some vegetatively propagated plants by gamma radiation. Indian J. Hortic., 34(2): 169-174.

Datta, S. 2000. Role of classical mutation breeding in crop improvement. Daya Pub. House, Delhi. pp. 265-266.

Datta, S.K. 2014. Induced mutagenesis: basic knowledge for technological success. National Botanical Research Institute (NBRI-CSIR), Lucknow, Uttar Pradesh, India. 97-139.

De, L.C. and S.K. Bhattacharjee. 2011. Ornamental crop breeding. <i>Aavishkar Publishers, Distributors, Jaipur, India</i>. pp. 40-41.

Flowers Influence of 60Co gamma irradiation and cool storage. <i>Advances
Ganesh, S. 2010. Influence of growth regulators and micronutrients on growth, yield and quality of tuberose (*Polianthes tuberosa* L.) cv. Prajwal. M.Sc. (Hort.) Thesis. Tamil Nadu Agricultural University, Coimbatore.

Kainthura, P. and R. Srivastava. 2015. Induction of Genetic Variability and Isolation of Mutants in Tuberose (*Polianthes tuberosa* L.). Tropical Agricultural Research. 26 (4): 721 – 732.

Kodym, A. and R. Afza. 2003. Physical and chemical mutagenesis. Methods in molecular chrysanthemum biology, 236: 189-204.

Kumar, V., Chatterjee, S.R., Bhattacharjee, S.K. 2013. Shelf life of tuberose loose Sambandhamurthi, S. 1983. Studies on induced mutations in tuberose (*Polianthes tuberosa* L.). Ph.D. Thesis submitted to Tamil Nadu Agricultural University, Coimbatore.

Singh, A., K. Kumar and K.Anuj. 2013. Studies of gamma irradiation on morphological characters in gladiolus.

How to cite this article:

Kayalvizhi, K., M. Kannan and Ganga, M. 2017. Effect of Physical and Chemical Mutagens on Morphological Characters in M1V2 Generation of Tuberose (*Polianthes tuberosa* L.). *Int.J.Curr.Microbiol.App.Sci.* 6(4): 2492-2499. doi: [https://doi.org/10.20546/ijcmas.2017.604.290](https://doi.org/10.20546/ijcmas.2017.604.290)