Mild Cognitive Impairment in novel SPG11 Mutation-Related Sporadic Hereditary Spastic Paraplegia with Thin Corpus Callosum

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Research article

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Abstract

**Background:** *SPG11* mutation-related autosomal recessive hereditary spastic paraplegia with thin corpus callosum (HSP-TCC) is the most common cause in complicated forms of HSP, usually presenting comprehensive mental retardation on early-onset stage preceding spastic paraplegias in childhood. However, there are still lots of sporadic late-onset HSP-TCC cases with negative family history, and potential mild cognitive deficits in multiple domains may be easily neglected and inaccurately described.

**Methods:** In this study, we performed next generation sequencing in four sporadic late-onset patients with spastic paraplegia and thin corpus callosum (TCC), and combined Mini-Mental State Examination (MMSE) and Montreal Cognitive Assessment (MoCA) to evaluate cognition of the patients.

**Results:** By evolutionary conservation and structural modeling analysis, we have revealed 4 novel pathogenic *SPG11* mutations, and firstly confirmed mild cognitive impairment (MCI) with normal MMSE scores (≥27) and decreased MoCA scores (<26) in these *SPG11* mutation-related HSP-TCC patients, predominantly presenting impairment of visuoexecutive function, delayed recall, abstraction and language correlated with prefrontal deficits.

**Conclusions:** The results expand the mutational spectrum of *SPG11*-associated HSP-TCC from sporadic cases, and confirm MCI characterized with dysfunction of prefrontal lobe in *SPG11*-related HSP-TCC, which should be paid more attention by neurologists.

**Background**

Hereditary spastic paraplegia (HSP) is a heterogeneous group of genetically-driven neurodegenerative disorder, inherited in autosomal dominant (AD), autosomal recessive (AR) or X-linked patterns with onset age varying from infancy to late adulthood, and traditionally classified into pure and complicated forms[1]. Among them, pure HSP is characterized by slowly progressive lower limb spasticity and weakness in isolation, while complicated HSP is characterized by spastic paraplegias associated with additional neurological features such as cognitive deficits, thinning corpus callosum (TCC), seizures, amyotrophy, ataxia, extrapyramidal disturbance, visual or auditory disturbances, orthopaedic abnormalities and peripheral neuropathy[2]. Up to now, at least 84 different loci and 67 genes have been shown to be associated with HSP. Among these genes, *SPG11* is the most common cause of complicated autosomal recessive hereditary spastic paraplegia with thin corpus callosum (HSP-TCC), which has been reported to be homozygous or compound heterozygous mutations[3].

General cognitive deficits in *SPG11*-related HSP-TCC preceding to spastic paraplegia is usually first noticed in childhood for learning difficulties and progresses insidiously to severe functional disability, diagnosed as mental retardation[4]. However, there are still lots of sporadic late-onset *SPG11*-related HSP-TCC cases without positive family history, and hardly diagnosed by only clinical manifestations and neuroimagings. Moreover, these patients may present spastic paraplegia without cognitive complaints,
and potential mild cognitive deficits in multiple domains may be easily neglected and inaccurately described by conventional Mini-Mental State Examination (MMSE) assessment [5–7].

In this study, we performed a combined approach of next generation sequencing, evolutionary conservation and structural modeling analysis to genetically assess 4 sporadic HSP-TCC patients without positive family history and prominent cognitive impairment. The results revealed 1 reported and 4 novel pathogenic $SPG11$ mutations. Furthermore, we combined MMSE and Montreal Cognitive Assessment (MoCA) to evaluate cognition of the patients, and firstly confirmed mild cognitive impairment (MCI) with normal MMSE scores (≥ 27) and decreased MoCA scores (< 26), predominantly presenting impairment of visuoeexecutive function, delayed recall, abstraction and language correlated with prefrontal deficits.

**Methods**

**Patients and clinical assessments**

Four cases with sporadic spastic paraplegias were recruited. The clinical assessments were approved by the Expert Committee (equal to the Institutional Review Board) of the Tangdu Hospital of Fourth Military Medical University (China), and we have obtained the written informed consents from all the patients and their family members. The patients and their relatives were all Chinese.

Except positive family history and prominent cognitive impairment in complaints, the characteristics in our patients were consistent with the clinical and radiological criteria for the complicated form of HSP-TCC reported in Japan[8], and Italy[9]. The diagnosis was determined by at least three experienced neurologists and radiologists. Because of negative family history, all known possible causes of spastic paraplegia were carefully excluded. All the patients were performed by Spastic Paraplegia Rating Scale (SPRS) for spastic paraplegia assessment[10], brain and spinal cord MRI scan, and electromyography (EMG) (including myoelectricity, nerve conduction velocity).

**Next Generation Sequencing**

The blood samples were collected for genetic analysis from 4 patients and all their relatives with informed consent. Extraction of genomic DNA from the peripheral blood leukocytes was obtained by using a standard protocol. Genomic DNA was isolated from peripheral leukocytes, fragmented into 150–200 bp length with the use of sonication. The DNA fragments were then processed by end-repairing, A-tailing and adaptor ligation, a 4-cycle pre-capture PCR amplification, and enriched by a custom-designed panel capturing the coding exons of 39 genes associated with spastic paraplegia, including $SPG11$.

Paired-end sequencing (150 bp) was performed on Illumina HiSeq X-ten platform to provide a mean sequence coverage of more than 100×, with more than 95% of the target bases having at least 20 × coverage.
Raw data was processed by the Illumina pipeline (version 1.3.4) for image analysis, error estimation, base calling and generating the primary sequence data. For the quality control, the Cutadapt (https://pypi.python.org/pypi/cutadapt) and FastQC (www.bioinformatics.babraham.ac.uk/projects/fastqc/) were used to remove 3′-/5′- adapters and low quality reads, respectively. The clean reads were mapped to the human reference genome (UCSC hg19) with the use of the BWA (version 0.7.10, http://bio-bwa.sourceforge.net)[11], duplicate sequence reads were removed by Picard (version 1.85; http://picard.sourceforge.net), and GATK (version 3.1, https://software.broadinstitute.org/gatk/)[12], was used to detect variants. Variants were annotated by ANNOVAR software (version 2015Dec14, http://www.openbioinformatics.org/annovar/)[13], which including function implication (gene region, functional effect, mRNA GenBank accession number, amino acid change, cytoband, etc.) and allele frequency in dbSNP138, 1000 Genomes (Phase3 - Variant Frequencies 5b)and ExAc (exac.broadinstitute.org/) [14], referring to transcriptNM_025137. Damaging missense mutations were predicted by SIFT (sift.bii.a-star.edu.sg/) and PolyPhen-2 (genetics.bwh.harvard.edu/pph2/). Interpretation of the variants according to the American College of Medical Genetics and Genomics (ACMG) recommended standards[15], and all the variants will be categorized into pathogenic, likely pathogenic, uncertain significance (VUS), likely benign and benign.

Sanger sequencing was performed to validate the putative pathogenic variants, allowing segregation analyses where possible. Genetic information of healthy Chinese obtained from local Chinese Millionome Database (CMDB) was identified as healthy controls.

**Structural And Functional Analysis**

*SPG11* protein sequence was obtained from the uniprot database (https://www.uniprot.org/uniprot/Q96JI7). Conserved domain database (CDD) version 3.18[16] was used to detect conserved structure domain in *SPG11* via RPS-blast with position-specific score matries (PSSMs), Expect Value threshold was set to 0.01.

Polyphen2 was used to predict possible structural or functional impact of amino acid substitution detected in human proteins using physical and evolutionary comparative algorithm, default setting was used. The prediction was against a precomputed database comprising ~ 150 million missense SNPs detected in all exons of UCSC human genome(hg19)[17].

The orthologous genes of *SPG11* were detected via blast, and the evolution tree was drawn by the gene orthology/paralogy predition method implemented in the ensemble database (http://asia.ensembl.org). Multiple sequence alignment was made using muscle 3.8[18], Green bars shows areas of conserved peptides in the sequence, white areas are gaps in the alignment. The multiple alignment sequence used to draw the seqlogo figure using the weblogo software[19] (http://weblogo.berkeley.edu/). The sequence logo consists of stacks of symbols for corresponding amino acids. The height of the stack indicates the conservation of amino acids. Chemical properties of amino acids were used to define the color system:
polar amino acids (G,S,T,Y,C,Q,N) are green, basic (K,R,H) blue, acidic (D,E) red and hydrophobic (A,V,L,I,P,W,F,M) amino acids are black.

De novo 3D structure modeling was performed using the I-TASSER algorithm[20] for both wild and mutant $SPG11$, it identified first 10 possible structure templates using a meta-server threading approach LOMETS[21] based on the highest significance Z-score of the threading alignments, and used SPICKER program to select the final simulation model based on pair-wise structure similarity using RMSD (TM-score). The confidence of the model is quantitatively measured by C-score. The first model with the best C-score is selected for further analysis.

Simulated protein 3D structures of wild and mutant $SPG11$ were aligned using superpose version 1.0[22]. Protein super positions were calculated using a quaternion approach. Rasmol version 2.7[23] was used to visualize the wild and mutant structures. Relative position of mutation site was determined using structure alignments.

**Neuropsychological Evaluation**

All the patients were performed by Neuropsychiatric Inventory (NPI), MMSE and MoCA for neuropsychological assessments. The NPI and MMSE were administered at the beginning, followed by MoCA on 7th hospital day to avoid effects of habituation. A cutoff of $\geq 27$ on the MMSE was chosen to indicate normal cognitive function and the accepted cutoff of $< 26$ on the MoCA was taken to indicate cognitive impairment[24, 25]. A cutoff of $\geq 26$ on the ADL was chosen to indicate functional disability.

MoCA is a 30-point test administered in 10 min consisting of seven subtests. Visuoexecutive functions are assessed using a clock-drawing task (3 points), a three-dimensional cube copy (1 point) and the Trail Making B task (1 point). Naming is assessed using a three-item confrontation naming task with low-familiarity animals (lion, camel, rhinoceros; 3 points). Attention is evaluated using a sustained attention task (target detection using tapping; 1 point), a serial subtraction task (3 points) and digits forward and backward (1 point each). Language is assessed using repetition of two syntactically complex sentences (2 points) and a phonemic fluency task (1 point). Abstraction is assessed using a two-item verbal abstraction task (2 points). The short-term memory recall task (5 points) involves two learning trials of five nouns and delayed recall after approximately 5 min. Finally, orientation to time and place is evaluated (6 points)[25].

**Results**

**Clinical features**

Four sporadic spastic paraplegias patients (2 male and 2 female) and their close relatives were studied. Ages at onset were based on information provided by the patients, the median age at onset for motor symptoms was 23 years (ranging from 18 to 27 years), and the median course of disease was 4 years
(ranging from 3 to 7 years). The symptoms at onset were all spasticity, and the spastic paraplegia was evaluated by Spastic Paraplegia Rating Scale (SPRS). Furthermore, no dysarthria, dysphagia, skeletal deformity, cerebellar signs, ophthalmoplegia, decreased vision, sphincter disturbance, amyotrophy, extrapyramidal signs, epilepsy, cataract, or optic atrophy was found in each patient. All patients accepted EMG examination, and both myoelectricity and nerve conduction velocity were normal. The clinical features of patients were listed in Table 1. Brain MRI showed thinning of whole corpus callosum in all patients, most pronounced in the genu and body parts, relatively spared in the splenium, without obvious periventricular white matter changes or cortical atrophy, while whole spinal cord MRI of each patient was normal (Fig. 1A-C).

**Genetic Findings**

In this study, we performed a custom-designed panel sequencing by next-generation sequencing in four sporadic patients with spastic paraplegia and TCC, and identified novel as well as reported *SPG11* mutations. Five mutations were recognized, and four of them, including 3 stop codons (c.4834C > T chr15:44881522/p.Q1612X, c.5137C > T chr15:44876741/p.Q1713X, c.5609T > A chr15:44876741/p.L1870X) and 1 missense mutation (c.316G > C chr15:44952756/p.A106P), were identified for the first time. (Table 2). The mutations were homozygous in 3 patients, and compound heterozygous in 1 patient. Results of Sanger sequencing indicated that these mutations segregated with the disease in patients while healthy parents and siblings of patients were all asymptomatic heterozygous carriers (Fig. 1D-G). Copy number variants (CNVs) of genes listed in the panel were also tested, and no associated CNVs were identified.

From patient 1, a novel homozygous stop coding mutation was detected in exon 28 (c.4834C > T) of *SPG11*, predicted to truncate the functional protein (PVS1). This mutation was never reported in public genomic databases (ExAC or gnomAD), nor detected in healthy controls (PM2). According to ACMG criteria, this mutation was classified to be likely pathogenic (PVS1 + PM2).

The *SPG11* gene of patient 2 displayed a novel homozygous missense c.316G > C (p.A106P) in exon 2. This missense was not identified in ExAC, gnomAD, or healthy controls (PM2). Some *in silico* algorithms, including SIFT and PolyPhen_2, performed the prediction of pathogenicity of this mutation. This mutation was graded as VUS based on the standard of ACMG. However, considering the absence of other variation or CNV in HSP related gene of this HSP-TCC patient, it is highly possible that this missense was the associated disease-causing loci.

The sequencing result indicated heterozygous mutations that c.5609T > A (p.L1870X) and c.5867-1G > T presented in two alleles of the *SPG11* gene respectively in patient 3. c.5609T > A was predicted to prematurely truncate the protein product (PVS1) and was neither reported in public genomic databases nor detected in healthy controls (PM2). It’s in *trans* mutation, mutation c.5867-1G > T, was previously published as Pathogenic (rs1060501168) (PM3)[26]. Thus mutation c.5609T > A was classified as Pathogenic (PVS1 + PM2 + PM3) according to ACMG criteria.
In patient 4, the novel homozygous nonsense was predicted to truncate the \textit{SPG11} in exon 30 (c.5137C > T) (PVS1). This mutation was never reported in ExAC or gnom AD and was not detected in controls (PM2). Two patients were identified in this family with genotypes co-segregated with the disease (PP1). Therefore, this mutation was classified as pathogenic (PVS1 + PM2 + PP1) through ACMG standards.

To accept the variants found via NGS, we have performed the Sanger sequencing test. All 4 mutations were validated and confirmed.

**Pathogenicity Of The Mutations**

Conservation of these 4 mutations were calculated based on multiple sequence alignment of 141 \textit{SPG11} homologous sequences. All these 4 residues were evolutionarily conserved in over 50\% orthologous. In patient 1, the mutation site (Q1612X) was located at the 2001 amino acid position where the main variants were Q and R. Q is the most conserved, suggesting the importance of Q. The stop gain mutation at this site may lead to the pathogenicity. In patient 2, the mutation site (A106P) was located at the 167 amino acid position in the below seqlogo, where A, T and V were common residues occurred in evolution across variants species. Alanine was the most conserved residue, however, Polin was rare. The Proline mutation was not presented in the evolution. In patient 3, the mutation site (L1870X) was located at the 2124 amino acid position where the main mutations was L, suggesting the importance of L at this position. The stop gain mutation may lead to the pathogenicity. In patient 4, the mutation site (Q1713X) was located at the 2124 amino acid position and the main variants were Q and N, and Q was the most conserved. The stop gain mutation may lead to the pathogenicity. (Fig. 2A-D)

Among them, A106P in patient 2 was predicted to be probably damaging with polyphen-2 score 0.98 (sensitivity 0.76, specificity 0.96), and the other 3 mutations in patient 1, 3 and 4 were all stop gains. To better understanding the role of these variants, structural modeling analysis was performed. The prediction with the Bepipred Linear Epitope Prediction 2.0 (http://www.cbs.dtu.dk/services/BepiPred/cite.php) showed that the mutation of A106P caused a sufficient surface exposure and orientation change of L104, and made the nearby peptide SRNSSTPTEPKL (92–104) to be a potential epitope. The other 3 variants were all stop gain mutations, causing the loss of the helix rich-c-terminal part of the SPG11, which could be important for structural stabilization. (Fig. 2E-H). Taking together, it seems that the c-terminal domain of SPG11 could be an important binding domain with other proteins and it is stabilized with a helix rich structure to the core region of SPG11. All these 3 variants (Q1612X, Q1713X, L1870X) introduced stop codon to the beginning position of the binding domain, thereby causing damages to both the SPG11 core structure and potential protein interaction function.

**Neuropsychological Findings**
All the patients have been educated for more than 12 years. NPI was performed in them for psychiatric assessment, and the results showed normal. MMSE was performed in these patients for preliminary cognitive assessment, and the score of each patient was normal ranging from 28 to 30, according to accepted normal MMSE cutoff value $\geq 27$, suggesting no general cognitive decline or dementia. Moreover, all the patients were reevaluated by MoCA in detail, and the score of each patient was decreased ranging from 16 to 22, according to accepted abnormal cutoff value $< 26$. Among the seven subtests of MoCA, the average score of “Visuoexecutive functions (5 points)” was 1.5 (ranging from 1 to 2); the average score of “Delayed recall (5 points)” was 1.5 (ranging from 1 to 2); the average score of “Abstraction (2 points)” was 0.25 (ranging from 0 to 1); the average score of “Language (3 points)” was 1 (ranging from 0 to 2); the average score of “Naming (3 points)” was 3; the average score of “Attention (6 points)” was 5.5 (ranging from 5 to 6); the average score of “Orientation (6 points)” was 5.75 (ranging from 5 to 6) (Table 1). The results of MoCA showed potential impairments in multiple cognitive domains that are not detected by the MMSE, including visuoexecutive function, delayed recall, abstraction and language, which have been reported to be associated with prefrontal lobe function[2]. Altogether, combination of MMSE and MoCA confirmed mild cognitive impairment (MCI) in these sporadic SPG11-related HSP-TCC cases, characterized with preferentially predominant dysfunction of prefrontal lobe.

**Discussion**

HSP is clinically divided into pure and complicated forms, and genetically into AD, AR and X-linked recessive forms. The number of families without genetic diagnosis after systematic testing ranged from 45–67% in the AD-HSP and 71–82% in the AR-HSP groups, and the percentage of sporadic cases was even higher to 85%[27]. Many retrospective studies have shown that SPG11 is the most frequent AR-HSP with homozygous or compound heterozygous types in different countries or regions, partly suggesting the candidates of disease-causing genes for sporadic HSP cases[28–30]. However, the SPG11 gene, comprising 40 exons and spanning 101 kilobases, have been discovered at least 127 mutations, distributing throughout the SPG11 gene without identified mutation hot spots[31]. Whole-exome capture by next generation sequencing provides a powerful and affordable mean to identify causative genes with less cost, more efficiency and accuracy[32].

SPG11 gene encodes endogenous expression of spatacsin, distributing in the neurons forming the corticospinal tract and corpus callosum, as well as in hippocampus, cerebellum, dentate nucleus and ponsin adult CNS[31]. Given the distribution of spatacsin relative to microtubules and vesicles, it is tempting to speculate that spatacsin was partially associated with cytoskeleton, endoplasmic reticulum and vesicles involved in protein trafficking. Loss of spatacsin function may affect axonal transport, contributing to the degeneration of the corticospinal tract and corpus callosum in HSP[33]. More recently, loss of spatacsin has also been demonstrated to be pivotal for autophagic lysosome reformation, and resulted in depletion of free lysosomes, accumulation of autolysosomes or lipids in lysosomes, and neuronal degeneration[34]. As shown in our study, we have reported four sporadic spastic paraplegias with TCC cases, and found novel SPG11 mutations in each patient by next generation sequencing. Moreover, systematical evaluation showed that all 4 variants occur at evolutionary conserved residues,
and present highly possible damages for either structural stabilization or potential protein binding ability of SPG11, suggesting pathogenicity of these mutations.

The corpus callosum is the main commissural pathway linking the hemispheres of the brain, and identifiable anatomically divided into four parts: rostrum, genu anteriorly, body centrally and splenium posteriorly. Thinning of the corpus callosum (TCC) is a key manifestation and often only imaging feature in early SPG11-related familial or sporadic HSP, and almost can be found in each patient, predominantly thinning in the genu and body, relatively sparing in the splenium of corpus callosum, which is also consistent with four sporadic patients reported in our study. The project fibers arising from prefrontal and motor cortex mainly pass through the genu and body respectively, and further research has also demonstrated that the prefrontal and motor portions of the corpus callosum are preferentially affected in SPG11-related HSP-TCC, suggesting specific impairments of multiple cognitive domains, such as visuoexecutive function, delayed recall, abstraction and language, associated with prefrontal lobe, in addition to spastic paraplegias[35–37]. Moreover, many retrospective studies have revealed that SPG11 is the most common disease-causing gene in HSP-TCC, followed by SPG15, while other rare mutations in SPG35, SPG46, SPG48, SPG54 and SPG56 have also been reported for HSP-TCC[38].

As TCC is the major hallmark of patients with complicated HSP and typically associated with cognitive impairment, early and widespread involvement of cognitive functions across all domains is a well-known phenomenon in SPG11-related HSP-TCC[37]. However, there are still lots of late-onset cases without cognitive complaints, and potential mild cognitive deficits in some specific domains of prefrontal functions may be easily neglected and inaccurately described by conventional MMSE, which is widely used for screening general cognitive decline or dementia, but inadequate for lack of prefrontal-related tasks[5–7]. Unlike MMSE, MoCA is designed to adding assessments or increasing difficulties of prefrontal functions such as visuoexecution, abstraction, delayed recall and language, and widely used to evaluate mild cognitive impairment (MCI) in degenerative diseases[25].

MCI is an intermediate clinical state between normal cognitive function and dementia with decline on objective cognitive tasks, and the aetiology of MCI is heterogeneous including different neurodegenerative diseases, ischaemia, trauma, metabolic disturbance, etc[39]. Neuropsychological testing can be helpful to distinguish MCI from normal or dementia cases, for dementia usually presenting clear impairment in functional activities and scores low on MMSE, while MCI showing particularly subtle deficits by more sensitive measures such as MoCA[40]. In fact, 75% of patients with MCI on neuropsychological testing had normal MMSE but abnormal MoCA, and sensitivity of MoCA for MCI is almost 90–100%[25]. In our study, the combination of MMSE and MoCA has confirmed MCI in SPG11-related HSP-TCC, characteristically presenting potential deficits in multiple cognitive domains associated with prefrontal functions that were not detected by the MMSE, including visuoexecutive function and abstraction (not tested by the MMSE) and delayed recall and language repetition (MMSE items too easy). These specific phenotypes of cognitive deficits in our patients were also consistent with preferentially affected prefrontal portions of the corpus callosum in SPG11-related HSP-TCC as described before[37].
Conclusions

Collectively, in present study, we have performed next generation sequencing in four sporadic HSP-TCC patients without cognitive complaints, and revealed 3 homozygous and 1 compound heterozygous novel SPG11 mutations. In addition, we systematically evaluated the pathogenenicity of these mutations via evolutionary conservation and structural modeling analysis. We found that all 4 variants occur at evolutionary conserved residues. And the mutations were all highly possible damaging for either structural stabilization or potential protein binding ability of SPG11. Our findings expand the mutational spectrum of SPG11-associated HSP-TCC from sporadic cases. Moreover, we firstly confirm MCI by combination of MMSE and MoCA in these patients, characteristically presenting potential deficits in multiple cognitive domains of prefrontal lobe in SPG11-related HSP-TCC, which should be paid more attention by neurologists.

Abbreviations

MCI: Mild cognitive impairment; HSP: Hereditary spastic paraplegias; AD: Autosomal dominant; AR: Autosomal recessive; MRI: Magnetic resonance imaging; TCC: Thincorpus callosum; MMSE: Mini-Mental State Examination; MoCA: Montreal Cognitive Assessment; SPRS: Spastic Paraplegia Rating Scale; NPI: Neuropsychiatric Inventory; CMDB: Chinese Millionome Database; ACMG: American College of Medical Genetics and Genomics; VUS: Variant of Uncertain Significance; CNVs: Copy Number Variations.

Declarations

Ethics approval and consent to participate

This study has been approved by the Ethical Committee of Tangdu Hospital, Fourth Military Medical University. We have obtained the written informed consents from all the patients and their family members.

Consent to publish

We obtained written informed consent from the patient for publication of this report and any accompanying images. A copy of the written consent is available for review by the Editor-in-Chief of this journal.

Availability of data and materials

The dataset analysed are available from the corresponding author on reasonable request.

Competing interests

The authors declare that they have no competing interests.
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Authors’ contributions

Conception and design of the study: WZ, YD. Acquisition of data: CL, QY, FD. Genetic analysis: QY, CZ, ZZ. Drafting of manuscript: CL, YD. Revision of the manuscript: WZ. All authors read and approved the final manuscript.

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**Tables**

Table 1. Clinical presentations of sporadic*SPG11*-associated HSP patients in this study
|                      | Patient 1 | Patient 2 | Patient 3 | Patient 4 |
|----------------------|-----------|-----------|-----------|-----------|
| **Inheritance**      | Sporadic  | Sporadic  | Sporadic  | Sporadic  |
| **Sex / Age at onset / Age at examination** | F/18/23   | F/27/32   | M/24/27   | M/23/26   |
| **Schooling years**  | 14        | 16        | 15        | 14        |
| **Initial symptoms** | Spasticity| Spasticity| Spasticity| Spasticity|
| **UL reflexes**      | ++        | ++        | ++        | ++        |
| **LL reflexes**      | ++++      | ++++      | +++       | +++       |
| **SPRS**             | 9         | 10        | 7         | 7         |
| **NPI**              | 0         | 0         | 0         | 0         |
| **MMSE**             | 30/30     | 30/30     | 29/30     | 28/30     |
| **MoCA**             | 22/30     | 18/30     | 16/30     | 17/30     |
| Visuoexecutive functions | 2/5      | 1/5      | 1/5      | 2/5      |
| Delayed recall       | 2/5       | 2/5       | 1/5       | 1/5       |
| Abstraction          | 1/2       | 0/2       | 0/2       | 0/2       |
| Language             | 2/3       | 1/3       | 0/3       | 0/3       |
| Naming               | 3/3       | 3/3       | 3/3       | 3/3       |
| Attention            | 6/6       | 5/6       | 5/6       | 6/6       |
| Orientation          | 6/6       | 6/6       | 6/6       | 5/6       |
| **MRI**              |           |           |           |           |
| TCC                  | +         | +         | +         | +         |
| PWM changes          | -         | -         | -         | -         |
| Cortical atrophy     | -         | -         | -         | -         |
| Ventricular Dilation | -         | -         | -         | -         |
| Cerebellar atrophy   | -         | -         | -         | -         |
| Spinal cord          | -         | -         | -         | -         |

M: male; F: female; UL: upper limbs; LL: lower limbs; SPRS: Spastic Paraplegia Rating Scale; NPI: Neuropsychiatric Inventory; MMSE: Mini Mental State Examination; MoCA: Montreal Cognitive
Assessment; nd: not done; MRI: magnetic resonance imaging; TCC: thin corpus callosum; PWM: periventricular white matter; +: presence;–: absence.

Table 2. Summary of the novel mutations of *SPG11* gene found in this study.

| Number | Inheritance | Location | Mutation | Consequence | Effect |
|--------|-------------|----------|----------|-------------|--------|
| Patient 1 | Homozygous (Non-consanguinity) | exon 28 | c.4834C>T chr15:44881522 | Stop codon | p.Q1612X |
| Patient 2 | Homozygous (Non-consanguinity) | exon 2 | c.316G>C chr15:44952756 | Missense mutation | p.A106P |
| Patient 3 | Compound heterozygous (Non-consanguinity) | exon 30 | c.5609T>A chr15:44876741 | Stop codon | p.L1870X |
| | | intro 31 | c.5867-1G>T chr15:44867240 | Frameshift mutation | |
| Patient 4 | Homozygous (Non-consanguinity) | exon 30 | c.5137C>T chr15:44876741 | Stop codon | p.Q1713X |

Figures
Figure 1

Presentations of corpus callosum in normal and SPG11-related HSP-TCC patients in midline sagittal brain MRI. (A) T1-weighted MR image shows normal corpus callosum anatomy in a 24-year-old female (1: rostrum, 2: genu, 3: body, 4: splenium). T1-weighted MR images of patient 1 (B) and patient 2 (C) with SPG11 homozygous mutation show thinning of whole corpus callosum most pronounced in the genu and body parts, relatively spared in the splenium. Pedigrees and mutation segregation of the families
with SPG11 mutations: patient 1 (D), patient 2 (E), patient 3 (F), and patient 4 (G). The squares indicate male; circles indicate female; filled shape, affected. M: mutation; +: wild type respectively. Chromatograms showing the mutations and respective wild type sequences are shown below the pedigrees.

Figure 2

Evolutionary and structural modeling analysis of 4 SPG11 mutants. (A) In patient 1, the mutation site(Q1612X) was located at the 2001 amino acid position, and Glutamine was the most conserved. (B) In patient 2, the mutation site(A106P). was located at the 167 amino acid position, Alanine was the most conserved residue, while Proline mutation was not presented in the evolution. (C) In patient 3, the mutation site (L1870X) was located at the 2331 amino acid position, where Leucine was highly conserved. (D) In patient 4, the mutation site(Q1713X) was located at the 2124 amino acid position and Glutamine was the most conserved. (E) Structural modeling analysis showed that the mutation of A106P in patient 2 caused a sufficient surface exposure and orientation change of L104, and made the nearby peptide SRNSSTPTEKPKL (92-104) to be a potential epitope. Structural modeling analysis showed that patient 1 (F), patient 3 (G) and patient 4 (H) were all stop gain mutations, causing the lost of the helix richc-terminal part of the SPG11, which could be important for structural stabilization.