Serum Daytime Melatonin Levels Reflect Cerebrospinal Fluid Melatonin Levels in Alzheimer’s Disease but Are Not Correlated with Cognitive Decline

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Abstract.
Background: Nocturnal cerebrospinal fluid (CSF) and blood melatonin levels are altered in Alzheimer’s disease (AD). However, literature remains inconclusive on daytime blood melatonin levels. A positive correlation between melatonin levels and Mini-Mental State Examination (MMSE) scores in AD subjects has been evidenced following cross-sectional analyses. Whereas a correlation between serum and spinal CSF melatonin has been shown in healthy volunteers, an equal investigation in AD patients still has to be undertaken.

Objective: 1) To evaluate whether serum melatonin levels correlate with spinal CSF melatonin levels in AD. 2) To compare daytime CSF and serum melatonin levels between patients with AD dementia, mild cognitive impairment due to AD, and healthy controls, and to evaluate whether melatonin can affect cognitive decline in AD.

Methods: Subjects with AD and healthy controls included in two existing cohorts, of whom a CSF and serum sample was available at the neurobiobank and had at least 6 months of neuropsychological follow-up, were included in the present study. Melatonin concentrations were measured with liquid chromatography-mass spectrometry.

Results: Daytime serum melatonin levels correlated with spinal CSF melatonin levels in AD (r = 0.751, p < 0.001). No significant differences regarding daytime melatonin levels were found between patients and controls. No correlations were observed between daytime melatonin levels and MMSE score changes.

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Conclusion: Daytime serum melatonin accurately reflects CSF melatonin levels in AD, raising the possibility to assess melatonin alterations by solely performing blood sampling if also confirmed for night-time values. However, daytime melatonin levels are not associated with changes of cognitive impairment.

Keywords: Alzheimer’s disease, cerebrospinal fluid, dementia, melatonin

INTRODUCTION

Melatonin is a neurohormone produced by the pineal gland under influence of the hypothalamic suprachiasmatic nucleus (SCN) and is part of our circadian rhythm, with low levels during the day (<5 pg/ml in plasma [1]) and high levels during the night in healthy adults (average 60 pg/ml in serum [2] [3–5]). Melatonin produced in the pineal gland reaches the third ventricle directly via the pineal recess and might furthermore be produced by glial cells such as astrocytes, leading to high melatonin levels in cerebrospinal fluid (CSF) (32.5 pg/ml for daytime CSF melatonin collected from lumbar cistern [6, 7]). On the other hand, melatonin enters the bloodstream through passive diffusion when the synthesis of melatonin increases at night [8].

Melatonin production is disrupted in Alzheimer’s disease (AD), the most common cause of dementia [9–16]. A loss of volume and cells of the SCN when comparing AD patients with age-matched controls has been described, as well as decreased functional pineal volumes [17–22]. Furthermore, a loss of melatonin rhythmicity in postmortem pineal glands with disappearance of the significant increase in melatonin levels in those AD patients who died at night versus those who died during the day, and a significant reduction in pineal night-time melatonin levels in AD patients as compared to controls has been shown [10, 23]. Ventricular CSF melatonin levels of clinically and pathologically confirmed AD patients were found to be one fifth of age-matched healthy controls [9]. Several studies have been performed regarding blood melatonin levels in AD, showing a loss in diurnal variations [12, 13, 24, 25], decreased total levels of melatonin [15], decreased nocturnal peaks [11, 12, 15, 24, 25], and earlier onset of the nocturnal melatonin rise [12] or wider scatter of the acrophase [11]. However, increased [12, 13] as well as decreased [14, 26, 27] or unaltered [11, 15, 28] daytime blood melatonin levels have been reported in patients with AD as compared to healthy controls. In addition, the effect of APOE ε4 carrier status on melatonin levels in CSF and blood of AD patients remains unclear [9, 28, 29].

Aberrant melatonin production might contribute to sleep disruptions in AD, including nightly restlessness and sundowning [4]. This might lead to disease progression due to decreased night-time functioning of the glymphatic system, which according to the glymphatic hypothesis [30, 31], is responsible for drainage of solutes like amyloid-β (Aβ), or by a relative increase in neuronal activity with increased production of Aβ as a consequence [32]. Melatonin has positive effects on sleep and cognition in mild cognitive impairment (MCI) patients, diagnosed according to the Petersen criteria [33, 34], but not in patients with dementia due to AD (ADD) where it only had positive effects on sleep quality [35]. Correlations between daytime as well as nocturnal blood melatonin levels and Mini-Mental State Examination (MMSE) scores or severity of dementia have been found [11, 14], but have not yet been studied in CSF, nor have they been correlated with changes in melatonin levels in a longitudinal way.

By direct secretion of melatonin in the third ventricle via the pineal recess, concentrations in (ventricular) CSF are higher than in the peripheral blood circulation [7]. A study in 13 healthy humans showed a correlation between spinal CSF melatonin levels and serum melatonin levels [6]. However, a more recent study showed that sleep deprivation in healthy subjects led to increased CSF and serum melatonin levels but with lack of correlation between both [36]. Whereas a very strong positive correlation between CSF melatonin levels and pineal melatonin levels in AD patients has been found [23], this has not yet been studied for blood and spinal CSF.

The aim of this retrospective study was to evaluate whether serum (daytime) melatonin levels accurately correlate with spinal CSF melatonin levels in the AD continuum. Furthermore, since current literature on daytime melatonin levels in AD produced conflicting evidence, we aimed to compare CSF and serum melatonin concentrations between patients with ADD, MCI due to AD (abbreviated as MCI as from here on), and healthy controls, in order to increase the usability of melatonin as blood-based biomarker. Lastly, we
aimed to investigate whether these daytime melatonin levels are associated with cognitive decline in AD.

METHODS

Subjects

Patients with dementia, patients with MCI, and healthy controls were included in a prospective, longitudinal observational study at the memory clinic of Hospital Network Antwerp (ZNA) Middelheim and Hoge Beuken. Patients were recruited between 1995 and 2013, cognitively healthy controls between 2014 and 2017 [37–39]. The protocol was approved by the local Ethics Committee (UZA/UAntwerp and ZNA), approval numbers 2805 and 2806. All participants and/or their legal representatives gave a written informed consent. Behavioral assessment and neuropsychological testing at baseline and during follow-up visits, as well as CSF and blood sampling for AD biomarker analysis and sample storage was done in some, but not all participants in the prospective study. For the current study, we retrospectively selected patients and controls from the Neurobiobank of the Institute Born-Bunge (NBB-IBB; FAGG registration n° 190113) according to following diagnostic criteria and selection procedures.

Diagnostic criteria

We selected patients with a diagnosis of MCI and ADD based on the National Institute on Ageing and Alzheimer’s disease (NIA-AA) criteria [40, 41], implying that diagnosis was supported by the core AD CSF biomarkers (Aβ1–42, total tau (T-tau) and phospho-tau (P-tau181)) for these patients. Healthy controls had normal CSF Aβ1–42 values.

Selection procedure

Patients

All MCI and ADD patients of whom CSF and blood samples were available in the NBB-IBB 1) and in whom baseline neuropsychological testing was done within 3 months of CSF and blood sampling (before or after); 2) and who had neuropsychological testing done during follow-up visit at least 6 months after the first neuropsychological testing; 3) were included in the current research protocol. This resulted in the inclusion of 13 patients with MCI and 23 ADD patients in the current study.

Cognitively healthy controls

The 5 oldest healthy controls, best age-matched for the patient cohort, of whom CSF and blood samples were still available in the NBB-IBB 1) and in whom baseline neuropsychological testing was done within 4 months of CSF and blood sampling (before or after); 2) and who had neuropsychological testing done during follow-up visit at least 6 months after the first neuropsychological testing; 3) were included in the current research protocol.

None of the participants took beta- or alpha-blocking therapy, nor did they take melatonin treatment. However, information about medication intake was missing in 3 healthy controls.

Procedures

Behavioral assessment

Behavioral assessment was performed using a battery of behavioral assessment scales (CMAI [Cohen-Mansfield Agitation Inventory], BEHAVE-AD [Behavioral pathology in Alzheimer’s disease], MFS [Middelheim Frontality Score], GDeprS [30 item Geriatric Depression Scale], CSDD [Cornell Scale for Depression and Dementia], GDS [Global Deterioration Scale]) at baseline and during follow-up visits in patients as described earlier [37–39]. For controls, no behavioral assessment was done. However, we knew that healthy controls did not have significant depressive complaints.

For the current study, we focused on the BEHAVE-AD E cluster which evaluates day/night rhythm disturbances, with a score ranging from 0 to 3 with 0 having no day/night disturbance and 3 meaning complete reversal of day/night rhythm (< 50% of former sleep cycle at night). Furthermore, we took into account the CSDD and GDeprS scores to evaluate the possible association of depression with melatonin levels. The CSDD is a 19-item depression scale. Item scores range from 0 (absent) to 2 (severe), with a maximum total score of 38 points. The items are clustered in five groups: (A) mood-related signs: anxiety, sadness, and lack of reactivity to pleasant events; (B) behavioral disturbances: agitation, retardation, multiple physical complaints, and loss of interest; (C) physical signs: appetite loss, weight loss, and lack of energy; (D) cyclic functions: diurnal variation of mood, difficulty falling asleep, multiple awakenings during sleep and early morning awakening; (E) ideational disturbances: suicide, poor self-esteem, pessimism, and mood-congruent delusions [37, 42]. The GDeprS is a 30-item questionnaire in which
participants are asked to respond by answering yes or no in reference to how they felt over the past week and is used as an instrument to detect and evaluate significant depressive symptoms [43]. Although the GDeprS was developed for cognitively healthy elderly, this scale remains valid in patients with MCI [44].

Neuropsychological testing

Neuropsychological testing was performed at baseline and during follow-up visits up to 3 years after baseline. Neuropsychological testing consisted of MMSE, and a battery of neuropsychological tests as routinely used for diagnosis and evaluation of dementia [37–39].

In the present study we focused on MMSE scores and MMSE score changes over a certain time period in order to evaluate cognitive decline. We assessed cognitive decline by evaluating [(last MMSE score measured – MMSE score baseline) / years between both] and defined this as “yearly MMSE score change”.

Lumbar puncture (LP) procedure and blood sampling and pre-analytical sample handling

At baseline, a subgroup of patients and controls in the prospective research protocol underwent LP and blood sampling at the same time during daytime (between 08:00 AM and 04:00 PM, 78% of samples were taken between 08:00AM and 10:00AM). LP and blood sampling was performed in normal hospital lighting conditions throughout all seasons of the year. Blood was taken at the same time of LP via peripheral venous puncture. CSF and blood were centrifuged (after coagulation in case of blood) and aliquoted. CSF and blood samples were immediately snap-frozen in liquid nitrogen and were transferred in liquid nitrogen to the IBB biobank facilities where they were kept frozen at ~80°C until subsequent analysis [39].

CSF analysis of AD biomarkers

CSF biomarkers analysis (P-tau181, T-tau, Aβ1–42) was performed at the UAntwerp BIODEM laboratory, using manual ELISA techniques (INNOTEST® β-Amyloid1–42, INNOTEST® hTau-Ag, and INNOTEST® Phospho-Tau181P, respectively; Fujirebio Europe, Ghent, Belgium) [39].

Melatonin analysis

Melatonin analysis in CSF and serum was performed by the Chrono@Work laboratory, Groningen, using a liquid chromatography mass spectrometry (LC-MS/MS) method on the Waters® Xevo™ TQ-MS triple quadrupole mass spectrometer with lower limit of quantification of 2.32 pg/ml, inter-assay CV for low levels (4.99 pg/ml): 9.5%, for high levels (69.68 pg/ml): 4.2%, intra-assay CV for low levels (4.76 pg/ml): 8.9%, for high levels (73.86 pg/ml): 3.5% [45].

Statistical analysis

Analyses were performed using the SPSS®27.0 software package. Values are reported as number, median with interquartile range (IQR), displayed as Q1 and Q3, or mean with standard deviation (SD). Since 13 CSF samples and 20 serum samples had melatonin levels below the limit of quantification (BLOQ) (< 2.3 pg/ml), we performed two separate analyses. In the first analysis we substituted the values BLOQ as follows: [limit of quantification/2] = 1.15 pg/ml, while for the second analysis we excluded those values BLOQ.

Based on our sample size calculation with alpha set at 0.05 and beta set at 0.2, 11 patients belonging to the AD continuum needed to be included to assess the correlation between serum and CSF melatonin values (sample-size.net). Based on the values reported by Sirin et al. [14], and based on our study population distribution, 24 (13/8/3 [ADD/MCI/healthy controls]) participants needed to be included to assess for differences in melatonin values between these different groups, as calculated with GPower® 3.1. Due to limited sample size and not normally distributed melatonin values as measured by Kolmogorov-Smirnov test, nonparametric tests were applied. To detect differences in categorical binary data between groups, the Fisher-Exact test was used (gender, BEHAVE-AD E score, medication intake, APOE status). Nonparametric tests (Mann-Whitney U test for pairwise comparisons and Kruskall Wallis test with Bonferroni correction for multiple groups) were applied to assess for differences of continuous data (age, MMSE scores, GDeprS, CSDD, BEHAVE-AD, MFS, CMAI, melatonin concentrations in CSF and serum). We used Quade’s ANCOVA to correct for age in the comparison of CSF and serum melatonin between ADD, MCI and healthy controls. Alpha was set at 0.05.
RESULTS

Patient characteristics

We included 23 patients with ADD, 13 patients with MCI, and 5 cognitively healthy controls. Demographic and clinical characteristics of participants can be found in Table 1. ADD, MCI, and healthy controls did not differ significantly regarding gender, age, or APOE e4 carrier status. However, AD continuum patients tended to be older than the healthy controls. MMSE scores at inclusion were significantly lower in the ADD patients as compared to the MCI patients and healthy controls (p<0.001) and in the MCI patients as compared to healthy controls (p=0.003).

ADD and MCI patients tended to have a larger yearly MMSE score change decrease than healthy controls, however the differences did not reach significance. The MMSE score change from baseline to year 2 was significantly higher in the ADD patients than in healthy controls (p=0.042). BEHAVE-AD scores, GDeprS, CSDD, MFS, CMAI, intake of anti-AD medication, intake of sleep medication, and intake of antidepressants could only be compared between patients with ADD and patients with MCI because we did not have this information for healthy controls. There were no significant differences between ADD and MCI patients regarding BEHAVE-AD E score. However, there tended to be more MCI patients than ADD patients with BEHAVE-AD E score of 0.

Table 1
Overview of patient characteristics. Data is reported as number or median with IQR

|                | ADD (n = 23) | MCI (n = 13) | Healthy controls (n = 5) | p       |
|----------------|-------------|-------------|--------------------------|---------|
| M/F            | 7/16        | 5/8         | 1/4                      | 0.801   |
| Age            | 77 [72.5;80.5] | 74 [68;77] | 70 [69;70]               | 0.038*  |
| MMSE score at baseline | 13 [9;16.5]  | 25 [25;26]  | 29 [29;30]               | <0.001b |
| Yearly MMSE score change | –2 [0;–4.5]  | –1 [0.4;–1.8] | 0 [0]                   | 0.202   |
| MMSE year 1 – MMSE baseline | –2 [0;–4] (n = 21) | 0 [–1.75;2] (n = 10) | 0 [–0.5;0.5] (n = 3) | 0.137   |
| MMSE year 2 - baseline | –5 [–8;–3.75] (n = 8) | –3 [–3.05] (n = 7) | 0 [0;0.5] (n = 3) | 0.020c  |
| BEHAVE-AD total score | 10 [6;17.5] | 3 [2.6]     | –                       | 0.002   |
| BEHAVE-AD A (21) | 0 [0.3]     | 0 [0.0]     | –                       | 0.011   |
| BEHAVE-AD B (15) | 0 [0.0]     | 0 [0.0]     | –                       | 0.117   |
| BEHAVE-AD C (9) | 3 [2.5;5]   | 0 [0.0]     | –                       | <0.001  |
| BEHAVE-AD D (9) | 3 [0.5;5.5] | 0 [0.1]     | –                       | 0.012   |
| BEHAVE-AD E (3) | 0 [0.1;5]   | 0 [0.0]     | –                       | 0.121   |
| BEHAVE-AD F (6) | 1 [0.2]     | 1 [0.2]     | –                       | 0.780   |
| BEHAVE-AD G (12) | 0 [0.0]     | 0 [0.2]     | –                       | 0.359   |
| Behave-AD E score | 0           | 12 [52%]    | 10 [77%]                 | 0.175   |
| 1              | 5 [22%]     | 2 [15%]     | –                       | 1.000   |
| 2              | 5 [22%]     | 1 [8%]      | –                       | 0.385   |
| 3              | 1 [4%]      | 0 [0%]      | –                       | 1.000   |
| Anti-AD medication | 5           | 0           | –                       | 0.136   |
| Sleep medication | 7           | 4           | –                       | 1.000   |
| Antidepressants | 7           | 1           | –                       | 0.213   |
| GDeprS         | 9 [4;11] (n = 21) | 6 [3;8] (n = 12) | –                       | 0.084   |
| CSDD           | 6 [4;8.5]   | 4 [1.6]     | –                       | 0.094   |
| MFS            | 4 [3;5]     | 1.5 [0.2]   | –                       | <0.001  |
| CMAI           | 42 [33.5;54.5] | 31 [29;35] | –                       | 0.002   |

*Yearly MMSE score changes is defined as “(last MMSE score measured – MMSE score baseline) / years between both”. Medication intake was missing in 3 healthy controls and displayed as not available (–) for all healthy controls since 60% of data for healthy controls was missing. BEHAVE-AD E, GDeprS and CSDD scores were not available (–) for healthy controls. GDeprS was furthermore missing in 2 ADD and 1 MCI patient. APOE e4 carrier status was missing in 1 ADD and 3 MCI patients. "MMSE year 1 – MSSE baseline" was missing in 2 ADD, 3 MCI, and 2 healthy controls; "MMSE year 2 – MMSE baseline" in 15 ADD, 6 MCI, and 2 healthy controls. aNo significant differences between groups after Bonferroni correction. bADD versus MCI: p<0.001; ADD versus healthy controls: p=0.001; MCI versus healthy controls: p=0.003. c) ADD versus healthy controls: p=0.042 after Bonferroni correction, no significant differences between other groups. Patients only took donepezil as anti-AD medication. Sleep medication included chloralhydrate, benzodiazepines, trazodone, and zolpidem. Antidepressants included venlafaxine, sertraline, paroxetine, and trazodone.
[77% versus 52%; p = 0.175] and more ADD patients than MCI patients with BEHAVE-AD E score of 1–3 [48% versus 23%; p = 0.175]. Furthermore, the ADD group tended to score higher on depression scales and tended to have larger intake of antidepressants than the MCI group.

**Serum versus CSF daytime melatonin in AD continuum: data BLOQ substituted as 1.15 pg/ml**

Spearman rank correlation coefficient showed a significant positive association between serum and CSF melatonin levels in the AD continuum \((r = 0.751; p < 0.001; n = 36)\), and in the ADD group \((r = 0.669; p < 0.001; n = 23)\) and in the MCI group separately \((r = 0.671; p = 0.012; n = 13)\). The scatterplot for CSF and serum melatonin in the AD continuum is shown in Fig. 1. In case of healthy controls, there is also a significant positive association between CSF and serum melatonin levels \((r = 0.894; p = 0.041; n = 5)\).

Since depression and altered day-night rhythms might influence daytime melatonin levels, we assessed whether the correlation coefficient changed after removal of those patients with significant depressive symptoms suggestive of major depression (CSDD or GDeprS > 11; \(n = 8\)) and after removing those with day/night disturbances (BEHAVE AD E > 1; \(n = 7\)). After removing patients with major depressive complaints, the association remained significant \((r = 0.715; p < 0.001; n = 28)\). The same was seen after removal of the patients with significant depressive symptoms and with intake of antidepressants \((n = 12)\) \((r = 0.657; p < 0.001; n = 24)\) or after removal of the patients with day/night disturbances \((r = 0.707; p < 0.001; n = 29)\). Even after removal of 1 participant with complete reversal of day-night rhythm (BEHAVE AD E = 3; \(n = 1)\), the correlation coefficient remained unchanged \((r = 0.749; p < 0.001; n = 35)\). The effects of sleep medication (chloralhydrate, benzodiazepines, trazodone, and zolpidem) and antidepressants (venlafaxine, sertraline, paroxetine, and trazodone) intake were also assessed separately: correlation did not increase after removal of patients with intake of sleep medication \((n = 11)\) or antidepressants \((n = 8)\) \((r = 0.622; p < 0.001; n = 25\) and \(r = 0.708; p < 0.001; n = 28\) respectively).

**Serum versus CSF daytime melatonin in the AD continuum: data BLOQ disregarded**

When disregarding melatonin levels BLOQ, the correlation coefficient was lower than after substitution of melatonin levels BLOQ but remained significant in the AD continuum \((r = 0.543; p = 0.024; n = 17)\). However, after removing patients with significant depressive complaints (CSDD or GDeprS > 11, \(n = 4)\) or patients with significant depressive symptoms and antidepressant intake \((n = 7)\), correlation coefficient dropped and did not remain significant \((r = 0.482; p = 0.095; n = 13\) and \(r = 0.249; p = 0.487\);

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**Fig. 1.** Scatterplot of CSF melatonin versus serum melatonin in the AD continuum \((n = 36)\) with BLOQ values substituted as 1.15 pg/ml.

**Fig. 2.** Scatterplot of CSF melatonin versus serum melatonin in the AD continuum \((n = 17)\) when BLOQ values were disregarded.
n = 10). There were no significant differences in CSF and serum melatonin values between depressed and non-depressed AD patients (p = 0.367; p = 0.271 respectively). On the other hand, after removal of patients with diurnal rhythm disturbances (BEHAVE AD E > 1; n = 5), the correlation coefficient increased (r = 0.647; p = 0.023; n = 12). After removal of 1 participant with complete reversal of day-night rhythm (BEHAVE AD E = 3), the correlation coefficient remained unchanged (r = 0.575; p = 0.020; n = 16). The scatterplot for CSF and serum melatonin in the AD continuum with values BLOQ disregarded is shown in Fig. 2.

**Melatonin levels and cognition**

Mean +/- SD CSF and serum melatonin values in the AD, MCI, and healthy control group are shown in Table 2 (BLOQ values substituted as 1.15 pg/ml) and Table 3 (BLOQ values disregarded). No statistical differences were found regarding CSF and serum melatonin between ADD, MCI and healthy controls. However, we knew that healthy controls did not have major depressive complaints, so we did a re-analysis for comparison of CSF and serum melatonin between ADD, MCI and healthy controls without depressive complaints (GDeprS or CSDD > 11). Differences remained insignificant (p = 0.767; p = 0.258 for CSF and serum melatonin respectively with substitution of values BLOQ; p = 0.171 and p = 0.371 without substitution). Furthermore, we evaluated the link between daytime melatonin levels in the AD continuum and cognition by correlating melatonin levels with MMSE scores at baseline and yearly MMSE score change. No significant correlations were found between CSF or serum melatonin levels and MMSE at baseline (r = –0.226; p = 0.185 and r = –0.100; p = 0.561 for CSF and serum with substitution of values BLOQ respectively; r = –0.155; p = 0.471; r = –0.023; p = 0.926 for CSF and serum respectively without substitution of values BLOQ), nor between CSF or serum melatonin and MMSE score change (r = –0.092; p = 0.596 and r = –0.013, p = 0.938 for CSF and serum with substitution of values BLOQ respectively; r = 0.002; p = 0.993 and r = 0.376; p = 0.112 for CSF and serum respectively without substitution of values BLOQ).

**Melatonin levels and Aβ1–42, P-tau181, and T-tau**

We also evaluated the association between melatonin levels and CSF Aβ1–42, P-tau181, and T-tau levels in the AD continuum but could not find any correlation (r = –0.059; p = 0.730, r = –0.237; p = 0.171, r = –0.181; p = 0.297 for CSF Aβ1–42, P-tau181, and T-tau and CSF melatonin with substitution of values BLOQ respectively; r = 0.123; p = 0.474, r = –0.124; p = 0.480, r = –0.204; p = 0.241 for CSF Aβ1–42, P-tau181, and T-tau and serum melatonin with substitution of values BLOQ respectively; r = 0.059; p = 0.786, r = –0.240; p = 0.271, r = –0.152; p = 0.489 for CSF Aβ1–42, P-tau181, and T-tau and CSF melatonin without substitution of values BLOQ respectively; r = 0.294; p = 0.222, r = –0.028; p = 0.914; r = –0.076; p = 0.758 for CSF Aβ1–42, P-tau181, and T-tau and serum melatonin without substitution of values BLOQ respectively).

**Table 2**

|                  | ADD (n = 23) | MCI (n = 13) | Healthy controls (n = 5) | p     |
|------------------|-------------|-------------|-------------------------|-------|
| CSF melatonin [pg/ml] | 4.09 ± 2.83 | 3.46 ± 3.45 | 3.25 ± 1.41             | 0.578 |
| Serum melatonin [pg/ml] | 3.57 ± 2.92 | 2.79 ± 4.05 | 2.83 ± 3.15             | 0.225 |

**Table 3**

|                  | ADD                  | MCI                  | Healthy controls       | p     |
|------------------|----------------------|----------------------|------------------------|-------|
| CSF melatonin [pg/ml] | 5.38 ± 2.44 (n = 16) | 4.90 ± 3.77 (n = 8) | 3.78 ± 0.90 (n = 4)    | 0.455 |
| Serum melatonin [pg/ml] | 5.13 ± 2.78 (n = 14) | 5.42 ± 5.92 (n = 5) | 5.35 ± 4.31 (n = 2)    | 0.461 |
Melatonin levels and APOE

As shown in Table 1, there were no statistical differences in e4 carrier status between ADD, MCI patients, and healthy controls. Furthermore, no statistical differences could be found in CSF and serum melatonin levels comparing patients belonging to the AD continuum carrying two APOE e4 alleles versus those who do not carry an e4 allele \((p = 0.960; p = 0.797 \text{ with substitution of values BLOQ; } p = 0.909 \text{ without substitution of values BLOQ})\), nor regarding those who carry one e4 allele versus those not carrying e4 alleles \((p = 0.590; p = 0.979 \text{ with substitution; } p = 0.649; p = 0.701 \text{ without substitution})\), nor when comparing those carrying two e4 alleles versus those carrying one e4 allele \((p = 0.925; p = 0.732 \text{ with substitution; } p = 0.610; p = 1.000 \text{ without substitution})\).

Melatonin levels and BEHAVE-AD E scores

We compared CSF and blood melatonin values between those AD continuum patients with BEHAVE-AD E score \(\leq 1\) and \(> 1\). We could not find any significant differences in CSF or blood melatonin levels when values BLOQ were substituted \((p = 0.528, p = 0.364 \text{ for CSF and blood respectively})\), nor when values BLOQ were not substituted \((p = 0.499, p = 0.853 \text{ for CSF and blood respectively})\). We could neither find a correlation between BEHAVE-AD E score and CSF/serum melatonin values when values BLOQ were substituted \((r = 0.128; p = 0.458 \text{ for CSF and } r = 0.134, p = 0.435 \text{ for serum})\) and when values BLOQ were not substituted \((r = 0.125; p = 0.561 \text{ for CSF and } r = 0.086; p = 0.727 \text{ for serum})\) in the AD continuum.

DISCUSSION

Melatonin disruptions have been widely described in AD patients. SCN cell number and volume as well as functional pineal gland volume decreases in AD, with disrupted pineal gland melatonin production and alterations in ventricular CSF and blood melatonin levels as a consequence [9, 11–13, 16, 17, 19, 25]. Ventricular CSF melatonin levels correlate with pineal gland melatonin in a pathologically confirmed AD cohort [23], whereas the correlation between serum and spinal CSF melatonin concentrations has only been described in 13 healthy volunteers [6]. In the present study, we found that daytime serum melatonin levels could accurately reflect daytime spinal CSF melatonin levels in the AD continuum as well, with a correlation coefficient similar to that found in healthy volunteers when melatonin levels BLOQ were substituted [6] \((r = 0.751 \text{ here versus } r = 0.72 \text{ in healthy volunteers})\), suggesting valid use of blood melatonin levels to study melatonin disruptions in AD during the day. This entails an advantage since venipuncture is less invasive than lumbar puncture. Furthermore, multiple blood samples can be taken with minimal stress, giving the potential to assess changes in melatonin levels during a 24 h period and venipuncture can easier be done in dark environment than lumbar puncture, making it easier to assess night-time melatonin levels although accurate correlation between the much higher night-time blood and CSF melatonin levels remains to be confirmed in a prospective AD study. Presence of depressive complaints, sleeping disturbances, and intake of antidepressants or sleep medication did not seem to negatively influence the positive correlation between CSF and serum melatonin when melatonin values BLOQ were substituted. However, when melatonin values BLOQ were disregarded, the correlation coefficient decreased after removal of depressive patients but increased after removal of patients with day/night disturbances. Since there were no significant differences in melatonin values between depressed AD patients versus non-depressed AD patients, we hypothesize that this drop might be due to the low sample size remaining after removal of depressed patients. Sleeping disturbances might have a negative effect on the correlation coefficient which has already been demonstrated in healthy persons [36]. However, even with inclusion of patients with sleeping disturbances in this study, the correlation remained significant. Overall, we conclude that daytime blood melatonin reflects daytime spinal CSF melatonin in the AD continuum.

Current literature suggests a significant decrease in nocturnal blood melatonin production in AD patients [11, 12, 15, 24, 25, 46, 47]. Furthermore, a decrease in total daytime production [15], advanced chronophases [12, 13], and wider scatters of the acrophase [11] have been described. However, regarding daytime melatonin levels, literature is inconclusive [16]. Increased daytime blood melatonin levels in patients with dementia and ADD as compared to age-matched controls have been reported [12, 13], whereas others describe decreased daytime blood melatonin levels as compared to healthy controls [14, 26, 27], with blood sampling
done between 08:00–09:00 AM [14], 07:00–08:00 AM [26], and at 12:00 AM [27] respectively. Soùêtre et al. [48] and Zverova et al. [28] could not find a statistically significant difference in daytime blood melatonin levels between demented patients and healthy elderly. As reported in the systematic review by Nous et al. [16], the paper by Zverova et al. [28] is the most recent paper regarding daytime blood melatonin levels. Our findings are in line with their findings. We could not find a statistically significant difference regarding daytime melatonin levels between patients with ADD, MCI, and healthy controls, not even when correcting for age, or, after removal of all patients with depressive symptoms. The fact that Sirin et al. [14] and Ozcankaya et al. [27] found decreased blood melatonin levels in ADD patients as compared to controls could possibly be explained by their early blood sampling: ADD patients might have advanced chronophases [12, 13], possibly contributing to an earlier drop in melatonin levels in the morning as compared to the healthy controls. However, sampling by Soùêtre et al. [48] and Zverova et al. [28] was also performed at 08:00 AM or between 07:00 and 08:00 AM, arguing against previously mentioned argument. Blood and CSF sampling in our study was done between 08:00 AM and 04:00 PM (78% of sampling was performed between 08:00 AM and 10:00 AM). In this timeframe, we know that plasma melatonin is not tended toward a large degree in variation/shift in healthy controls [5].

Sirin et al. found a correlation between MMSE scores and daytime melatonin levels [14], which we could not confirm. Neither could we find any correlation between daytime melatonin levels and longitudinal MMSE score changes. We hypothesize that daytime melatonin levels do not contribute enough to total melatonin levels. Therefore daytime melatonin levels might not entail positive effects on cognition in AD. On the other hand, we suggest that disruptions in night-time melatonin production, and as such probably also in the total melatonin production, might be more predictive of cognitive decline in AD. Ferrari et al. found a correlation between night-time melatonin levels and MMSE scores in demented patients [11]. During night-time, melatonin levels increase, making it a much larger contributor to total melatonin production in humans and possible beneficial effects on AD symptoms than daytime melatonin levels. The link between nocturnal melatonin production in the AD continuum and longitudinal MMSE score changes should therefore be further investigated.

Liu et al. found ventricular CSF melatonin levels to be significantly reduced in patients carrying two APOE e4 alleles when compared to patients carrying an e3 and e4 allele [9]. In contrast with this, Zverova et al. found that plasma melatonin levels were significantly higher in a group of AD patients who carried at least one e4 allele versus those who did not carry an e4 allele [28]. We could not find any differences in CSF or serum melatonin concentrations when comparing AD patients expressing one or two e4 alleles as compared to those who did not. The fact that we could not find any differences in APOE e4 carrier status between our AD patients and healthy controls might be explained by the accidentally high prevalence of APOE e4 carrier status in our healthy control group (80%), which is normally only present in 23.1% of a Caucasian population [49]. Only one healthy control carrying one e4 allele had subjective memory complaints, whereas all other healthy controls had no memory complaints.

Limitations of our study are the retrospective nature of the study, with availability of serum and CSF only taken during the daytime. The fact that sampling was performed during the day, led to melatonin values below the limit of quantification in part of the participants. We tried to address this issue by performing two separate analyses: one where values were substituted as [limit of quantification/2] and one analysis where they were disregarded. Values reported in our study are rather low in comparison with other studies (±9 pg/ml Sirin et al. [14], 170 pg/ml Ozcankaya et al. [27], 34 pg/ml Zverova et al. [28] for AD patients). However, according to the recent review by Kennaway, melatonin levels in plasma are expected to be less than 5 pg/ml during the daytime when high-quality assay procedures are used, which makes us conclude that measured melatonin values in our study are accurate [1]. Further limitations include the relatively small sample size that however meets our sample size calculation, and the fact that CSF and serum samples from AD patients were older than those from healthy controls. However, melatonin has been shown to remain stable over time in propylene glycol, glycofurol, and dimethyl sulfoxide at 25°C [50], as well as in aqueous solution at 4°C and –70°C [51]. Furthermore, our samples were stored at –80°C and had undergone maximally 3 freeze-thaw cycles and it has been shown before that plasma melatonin levels remained unchanged following three freeze-thaw cycles [52].

Our study also exhibits strengths. We used a good analytical method (LC-MS/MS) for melatonin anal-
analysis as mass spectrometry provides us with target values for daytime melatonin levels that immunoassays still need to achieve [3]. Furthermore, we used core AD CSF biomarkers for clinical diagnostic categorization (ADD, MCI, and healthy controls) of participants which has not been done before in AD melatonin studies. As mentioned before, blood and CSF sampling between 08:00 AM and 04:00 PM would not lead to a large degree in variation/shift in melatonin levels, as shown in healthy volunteers’ plasma before [5]. Melatonin has been measured in serum and plasma [14, 27, 28, 36]. In contrast to melatonin’s precursor serotonin, which is stored in platelets requiring serotonin analysis in whole blood or platelet-rich plasma [53], melatonin does not seem to be stored in platelets leading to accurate melatonin measurement in serum and platelet-poor plasma [54]. A high correlation between serum and plasma melatonin has been found in blood samples drawn from 20 Caucasian men ($r = 0.97$) [55].

Larger, prospective clinical studies are needed to detect possible differences in night-time melatonin levels between ADD patients, MCI patients, and healthy controls and to investigate the possible correlations with cognitive decline, as mentioned above, in order to establish further evidence for the use of melatonin as a future treatment option in AD. By using longitudinal neuropsychological follow-up, new prospective studies would enable us to study the correlation between night-time melatonin values and cognitive decline in AD. Prospective studies entail the availability to standardize sampling at the same time of day in different participants, and to correct for environmental influences such as light, caffeine intake, and tryptophan-enriched food intake in particular, since melatonin is formed out of serotonin.

**CONCLUSION**

Serum daytime melatonin levels accurately reflect daytime CSF melatonin levels in the AD continuum, expressing the possibility to use blood sampling when studying melatonin’s disruptions and effects in AD. However, the correlation between night-time blood and CSF melatonin values remains to be proven. The sole need of blood for melatonin measurement in AD would bring along several advantages, because venipuncture is less invasive and easier performed in dark environment than lumbar puncture. Furthermore, there is a possibility to collect multiple blood samples within a 24-h period. We conclude from our retrospective study that daytime melatonin levels are not associated with cognitive status (ADD versus MCI due to AD versus healthy controls), nor can these levels predict cognitive decline. We hypothesize that there might be a more important role for night-time melatonin levels in the reflection and prediction of cognitive decline in AD.

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