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Evolutionary characterization of the emerging porcine epidemic diarrhea virus worldwide and 2014 epidemic in Taiwan

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Abstract

Since 2010, a new variant of PEDV belonging to Genogroup 2 has been transmitting in China and further spreading to the United States and other Asian countries including Taiwan. In order to characterize in detail the temporal and geographic relationships among PEDV strains, the present study systematically evaluated the evolutionary patterns and phylogenetic resolution in each gene of the whole PEDV genome in order to determine which regions provided the maximal interpretative power. The result was further applied to identify the origin of PEDV that caused the 2014 epidemic in Taiwan. Thirty-four full genome sequences were downloaded from GenBank and divided into three non-mutually exclusive groups, namely, worldwide, Genogroup 2 and China, to cover different ranges of secular and spatial trends. Each dataset was then divided into different alignments by different genes for likelihood mapping and phylogenetic analysis. Our study suggested that both nsp3 and S genes contained the highest phylogenetic signal with substitution rate and phylogenetic topology similar to those obtained from the complete genome. Furthermore, the proportion of nodes with high posterior support (posterior probability > 0.8) was similar between nsp3 and S genes. The nsp3 gene sequences from three clinical samples of swine with PEDV infections were aligned with other strains available from GenBank and the results suggested that the virus responsible for the 2014 PEDV outbreak in Taiwan clustered together with Clade I from the US within Genogroup 2. In conclusion, the current study identified the nsp3 gene as an alternative marker for a rapid and unequivocal classification of the circulating PEDV strains which provides complementary information to the S gene in identifying the emergence of epidemic strain resulting from recombination.

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1. Introduction

Porcine epidemic diarrhea virus (PEDV) causes an acute and highly contagious enteric tract infection, characterized by severe villus atrophy and congestion of the thin-walled intestines, which usually lead to high morbidity and mortality, especially in piglets (Song and Park, 2012). The disease was first observed among fattening pigs from United Kingdom in 1971 and the etiological agent was identified in Belgium as a new coronavirus, which was designated as PEDV prototype strain CV777 (Jung and Saif, 2015). Within the next two decades, PEDV was reported in several other European countries including Hungary, Italy, Germany, France, Switzerland and the Czech Republic, causing only isolated outbreaks in Europe (Hanke et al., 2015; Jung and Saif, 2015). In Asia, PEDV was first identified in 1982 and is now considered an endemic, causing devastating enteric diseases and substantial economic losses to the pork industry in many Asian countries such as China, South Korea, Japan, Thailand, and Taiwan (Lin et al., 2014; Park et al., 2007, 2011; Song and Park, 2012; Temeeyasen et al., 2014; Zhang et al., 2013).

Comparisons of full-length genomes showed that different PEDV strains are more closely related to alphacoronaviruses in a bat than other known alphacoronaviruses, suggesting that interspecies transmission of coronavirus may have occurred decades ago between bats and pigs or through other intermediate hosts (Huang et al., 2013; Tang et al., 2006; Woo et al., 2012). PEDV, belonging to the genus Alphacoronavirus within the family Coronaviridae, is an enveloped, single-stranded RNA genome with a 5′ cap and a 3′ polyadenylated tail (Jung and Saif, 2015). The size of its genome is approximately 28 Kb with two thirds of the 5′ genome containing two large open reading frames (ORFs), 1a and 1b, that encode two nonstructural polyproteins, pp1a and pp1b, which are involved in genome replication and transcription (Brian and Baric, 2005). The remaining PEDV genome contains ORFs encoding four structural and one nonstructural proteins in the following order: spike (S), ORF3, envelope (E), membrane (M)
and nucleoprotein (N). Similar to S proteins from other coronaviruses (CoVs), the PEDV S protein is a glycoprotein on the viral surface and has a pivotal function in regulating interactions with specific host cell receptor glycoproteins to mediate viral entry (Li, 2015). Thus, the S glycoprotein is often used as an important viral component to understand the genetic relationships of different PEDV strains and the epidemiological status of PEDV in the field (Chen et al., 2013b; Tian et al., 2013). Nevertheless, the high variability of other genes including ORF3, E, M or N protein has been previously utilized for phylogenetic inference (Chen et al., 2013a; Ge et al., 2013; Kubota et al., 1999; Pan et al., 2012; Temeeyasen et al., 2014; Yang et al., 2013). In order to get better phylogenetic resolution, several studies used PEDV full genome sequences for analysis (Chen et al., 2014; Pan et al., 2012; Sun et al., 2015; X. Wang et al., 2013); however, sequencing the full genome is expensive from a laboratory perspective and also limits the rapid characterization of novel PEDV strains. Furthermore, for many computationally intensive analyses, utilizing the full genome is unfeasible. It would be beneficial to use only those genomic regions that contain the highest phylogenetic signal to reduce cost without losing valuable information.

Since 2010, massive PED outbreaks were reported in China characterized by 80 to 100% illness among infected swine herds and a 50 to 90% mortality rate among infected suckling piglets (Zhang et al., 2013). The emerging strains in Asia are distinct from previous endemic PEDV strains characterized by multiple amino acid insertions/deletions or mutations on the S protein (J. Wang et al., 2013). The mutations on the S protein have been speculated to be associated with escape from neutralizing antibodies due to the use of a bivalent vaccine against transmissible gastroenteritis virus (TGEV) and PEDV (Chen et al., 2013a). This novel PEDV strain has continued to spread throughout the United States and Taiwan where no vaccination program has been implemented (Chen et al., 2014; Deng et al., 2014). Therefore, tracing the transmission and evolutionary changes of PEDV is important for future public health intervention. The objectives of the present study are (1) to systematically evaluate the evolutionary patterns and phylogenetic resolution in each gene in order to determine which regions provided the maximum interpretative power to infer temporal and geographic relationships among PEDV strains by using 34 complete genomic sequences from worldwide collection of PEDV field strains; and (2) to identify the origin of PEDV that caused the epidemic in Taiwan in 2014.

2. Materials and methods

2.1. Sequence data

We downloaded all full genome sequences (34) available in GenBank as of February 26, 2014 (http://www.ncbi.nlm.nih.gov/) for which the sampling year and country of collection was recorded (Supplementary Table 1). Since Genogroup 1 contained only 8 full-length sequences and no PEDV in the US was found until 2013, dataset was divided into three non-mutually exclusive groups: worldwide (34 sequences), Genogroup 2 (24 sequences) including all the recent emerging strains, and China (20 sequences) including all strains obtained from China, to cover different ranges of secular and spatial trends. Each of those datasets was then divided into different alignments by genes: structural (S, E, M, N) and non-structural (NSP1, NSP2, NSP3, NSP4, 3C-like protease, NSP6, NSP7, NSP8, NSP9, NSP10, RNA-dependent RNA polymerase, helicase, exoribonuclease, uridylyltransferase, specific endoribonuclease, and uridylyl-transferase) based on the sequence prediction from the whole genome of the strain CV777 (GenBank accession no. NC_003436). We created two additional concatenated datasets with length >2000 nucleotides for comparison: (1) NSP1 and NSP2 (denoted as 5′-nsp1-2); (2) ORF3, E, M, and N (denoted as 3′-OEMN). Sequences were aligned using ClustalW2 available from the European Molecular Biology Laboratory (EMBL, http://www.ebi.ac.uk/Tools/msa/clustalw2/) and manually edited.

Diarrhea among the piglets from the swine farms in Taiwan began in December 2013 and the etiologic agent of this epidemic was confirmed to be PEDV (Lin et al., 2014). The total nucleic acids from three representative clinical samples were extracted as previously described (Deng et al., 2014) and the cDNA was produced using the SuperScript III Reverse Transcription kit with random hexamers (Invitrogen, Carlsbad, CA, USA). The complete sequence of the NSP3 gene in each clinical sample was determined using the primers summarized in Table 1 and are available under GeneBank accession number KR632490-2.

2.2. Comparative analysis

To evaluate the phylogenetic resolution of each gene from each dataset, two bioinformatics approaches were employed: likelihood mapping (LM) and phylogenetic analysis involving the determination of the model of evolution and Bayesian Markov chain Monte-Carlo (MCMC) phylogenetic analyses.

2.2.1. LM analysis

To investigate the phylogenetic signal of each gene from all three datasets, LM was performed using TREE-PUZZLE program by analyzing 10,000 random quartets (Schmidt et al., 2002). LM assesses if a dataset is suitable for phylogenetic reconstruction by analyzing four randomly chosen groups of sequences, termed quartets. Each quartet was evaluated using maximum likelihood and three possible unrooted tree topologies were weighted for each quartet. The posterior weights were then plotted onto a triangular surface. Based on this method, the fully resolved tree topologies were plotted in the three corners, indicating the presence of a tree-like phylogenetic signal, and the unresolved quartets were shown in the central region of the triangle. The phylogenetic noise was computed probabilistically and a star-like signal was shown when more than 30% of the dots fell within the central area representing unresolved phylogenies (Strimmer and von Haeseler, 1997).

2.2.2. Phylogenetic analysis

Before proceeding to the phylogenetic analysis, each gene and each of the four datasets were tested against the best fitting nucleotide substitution model as specified. The best-fit model of nucleotide substitution was selected under the Akaike information criterion (AIC) and Bayesian information criterion (BIC) as implemented in jModelTest (Posada, 2008).

To estimate the genealogy and the evolutionary timescale of NSP3, S and E sequence alignments using the worldwide, Genogroup 2 and China datasets, the Bayesian framework implemented in BEAST software package version 2.1.3 (Bouckaert et al., 2014) under the designated molecular clock model and the nucleotide substitution model pre-determined by jModelTest was used. The BEAST program was run with two different rate categories for codon position 1 + 2 and position 3, allowing gamma-distributed rate variation when model selection results were suggested. The published year of isolation was assigned to each sequence (tip) to calibrate the rate estimate and the expected time to coalescence was modeled under constant population sizes. The MCMC process was run for 100 million iterations until convergence

Table 1

| Primer ID | Sequence (5′ to 3′) | Position | Used to amplify fragment |
|-----------|---------------------|----------|-------------------------|
| Nsp3-F2   | TCCACCGATGATAATGTG  | 2646–2665 | PCR forward primer      |
| Nsp3-R2   | TGAAGACGACAAAAACGAGAAG | 8088–8110 | PCR reverse primer      |
| Nsp3-F2-1 | TGGGTTGATGTGTCGGCTTG | 3706–3725 | Sequencing primer       |
| Nsp3-R2-1 | GCCCTCTAGGACTTACGAC  | 6960–6981 | Sequencing primer       |
| Nsp3-F2-2 | AGGACGATGACAGACAGTTC | 4700–4721 | Sequencing primer       |
| Nsp3-R2-2 | ACACCTCAATTTAATTACGAC | 6176–6197 | Sequencing primer       |

* Positions correspond to the PEDV CV777 strain (GenBank accession no. NC_003436).
with sampling every 10,000th generation (Drummond et al., 2012). The results were visualized using the Tracer program v.1.6 (http://beast.bio.ed.ac.uk/Tracer), and convergence of the Markov chain was assessed by calculating the effective sampling size (ESS) for each parameter. All the MCMC runs were repeated at least three times to confirm the convergence. All ESS values >200 indicated sufficient sampling. For each dataset, the maximum clade credibility (MCC) tree, which is the tree with the largest product of posterior clade probabilities, was selected from the posterior tree distribution after 10% burn-in using the program TreeAnnotator version 2.1.2 and displayed by FigTree v.1.4.2.

2.2.3. Evaluation of the molecular clock hypothesis

Molecular clock analyses were performed based on the NSP3, S and E genes from all three worldwide, Genogroup 2 and China datasets. The strict molecular clock assumes the same evolutionary rates along all branches in the tree, while the relaxed molecular clock allows different rates along different tree branches. These two models were compared by calculating the Bayes Factor (BF), which is the ratio of the marginal likelihoods (marginal with respect to the prior) between the two models being compared (Suchard et al., 2001; Vexler et al., 2013). The approximate marginal likelihood for each coalescent model was calculated through 1000 bootstrap samplings using the harmonic mean of the sampled likelihoods and the natural log difference of marginal likelihood between any two models is the log of the Bayes Factor, log(BF). Evidence against the null model (i.e., the one with lower marginal likelihood) is indicated by 2 log(BF). It is moderately rejected when the value is greater than 3 and strongly rejected when it is greater than 10. The calculations were performed with BEAST version 2.1.3 and Tracer v.1.5.

2.3. Statistical analysis

The proportion of constant sites between different datasets was analyzed by the non-parametric Mann-Whitney U test. The linear relationship between nucleotide length and phylogenetic noise was examined using Pearson correlation test.

3. Results

3.1. Phylogenetic signal using LM

LM analysis was used to calculate the phylogenetic noise for each gene in each dataset and it has been shown that datasets with less than 30% noise are usually reliable for phylogenetic inference based on previous simulation studies (Strimmer and von Haeseler, 1997). In our study, the phylogenetic noise ranged from 55.9% (ns7) to 3% (ns3) in the worldwide dataset (Table 2) and only four individual genes (ns1, 7, 9 and 10) contained noise greater than 30%. Similar results were found when only the China dataset was used, among which four genes (ns1, 7, 10 and E) contained noise greater than 30%. However, when the Genogroup 2 dataset was examined, 10 genes contained noise greater than 30%, including those genes detected in the worldwide and China datasets (Supplementary Tables 2 and 3). Overall, in the worldwide dataset, the triangle of LM analysis showed that ns3 and spike genes contained only 3.2% and 4.1% of the probabilities in the center region representing the star-like topologies (unresolved phylogenies), respectively. Similar results were also found in the Genogroup 2 and China datasets.

Next, we further examined the constant sites and alpha values among different genes since the sequence numbers were equivalent among the three datasets. The results suggested that Genogroup 2 dataset consistently contained higher constant sites than the worldwide and China datasets, particularly in 3C-like protease, ns6, 8, uridylate-specific endoribonuclease, M and N genes, with statistical significance (p < 0.05). Furthermore, a broad range of alpha values of the gamma-distribution was observed in all three datasets. Although different gamma-distributions might affect the levels of phylogenetic noise, no correlation was found. However, when the relationship between nucleotide length and phylogenetic signal was plotted, a linear correlation was observed with statistical significance (worldwide: R² = 0.52, p < 0.001; Genogroup 2: R² = 0.50, p < 0.001; China: R² = 0.51, p < 0.001) (Fig. 1). The results were further supported by the concatenated alignment of multiple genes. Both 5’ns1p-2 and 3’OMEIN reduced the noise to 10%, which was significantly lower compared to the noises obtained from individual gene analysis (Table 2). In summary, the LM analysis results suggested that while length is significantly correlated with phylogenetic signal, the ns3 and S genes contained the greatest phylogenetic signal, and had similar constant sites but different gamma-distributions under different temporal and spatial contents. We therefore focused on these two genes for further phylogenetic analysis.

3.2. Evolutionary model and rate estimation

The rate matrix, alpha value in the gamma distribution and the proportion of the invariable site were calculated for the complete, ns3, S and E genes in all three datasets (Supplementary Table 4). In the worldwide dataset, the estimated rate matrix was similar and the same evolutionary model (GTR + I + G) was selected for all three different gene sets except for the E gene. However, when the genogroup reduced or geographically reduced datasets were used, different evolutionary models for each gene were obtained. The small sample size of sequences in this study does not permit for further investigation.

The coalescent-based Bayesian MCMC approach was further used to estimate the evolutionary rate of the ns3, S and E genes from all three worldwide, Genogroup 2 and China datasets. The strict (SC) and relaxed molecular clock (RC) hypotheses were compared by using the BEAST program that incorporated the year of sample collection. Analyses were conducted using a constant coalescent prior and the specified substitution models based on the results were shown in Supplementary Table 4. For all three datasets, the SC hypothesis was strongly rejected (Bayes Factors > 70) when compared to RC model for the ns3 and S genes.
genes, which was consistent with that obtained using complete genome sequences, suggesting rate heterogeneity among branches (Table 3). For the worldwide dataset, although the 95% high posterior density intervals (95% HPDs) were overlapping, the mean evolutionary rate estimated under the RC assumption was slightly faster for S ($2.22 \times 10^{-2}$) than for nsp3 ($1.63 \times 10^{-2}$) gene or complete genome ($1.37 \times 10^{-2}$). Similar findings were also found in the Genogroup 2 and China datasets. On the contrary, the E gene didn’t show rate heterogeneity among branches since neither the RC model nor the SC model favored the result based on the Bayes Factor calculation. Furthermore, the E gene showed the highest evolutionary rate based on SC model estimation from all three different datasets.

Although different evolutionary rates were estimated for all three different genes, the TMRCA was similar for all three genes. For worldwide dataset, about 36 to 38 years before present were estimated from the RC model when complete genomes, nsp3 and S genes were

![Fig. 1. Correlation between nucleotide length and phylogenetic noise.](image)

### Table 3
Bayesian estimation of the molecular clock, evolutionary rate, and time to most recent common ancestor (TMRCA).

| Dataset        | Gene | Clock model | Bayes factor | Evolutionary rate (substitutions/site/year) | Lower HPD  | Upper HPD  | TMRCA (year) | Lower HPD  | Upper HPD  |
|----------------|------|-------------|--------------|---------------------------------------------|------------|------------|---------------|------------|------------|
| **Worldwide (N = 34)** |      |             |              |                                             |            |            |               |            |            |
| Complete       |      | Strict      |              | 1.31E-02                                          | 1.14E-02   | 1.48E-02   | 119.8         | 102.2      | 134.5      |
| Complete       |      | Relaxed     |              | 494.9                                          | 1.37E-02   | 9.77E-03   | 102.2         | 104.5      | 106.5      |
|                            |      | Strict      |              | 2.16E-02                                          | 1.81E-02   | 2.50E-02   | 60.4          | 53.5       | 67.6       |
|                            |      | Relaxed     |              | 146.2                                          | 1.09E-02   | 2.19E-02   | 37            | 35         | 41.2       |
| S               |      | Strict      |              | 2.24E-02                                          | 1.89E-02   | 2.63E-02   | 91.5          | 77.7       | 106.9      |
|                            |      | Relaxed     |              | 288.1                                          | 2.28E-02   | 3.10E-02   | 36.1          | 31.5       | 38.7       |
| E               |      | Strict      |              | 5.81E-02                                          | 4.30E-02   | 7.42E-02   | 35.7          | 35         | 37         |
|                            |      | Relaxed     |              | 39.9                                          | 2.90E-02   | 4.14E-02   | 36            | 35         | 38.3       |
| **Genogroup2 (N = 24)** |      |             |              |                                             |            |            |               |            |            |
| Complete       |      | Strict      |              | 3.93E-02                                          | 3.14E-02   | 4.78E-02   | 8.8           | 7.5        | 10.1       |
|                            |      | Relaxed     |              | 357                                          | 4.59E-02   | 6.71E-02   | 32            | 2.3        | 4.5        |
| S               |      | Strict      |              | 5.81E-02                                          | 4.42E-02   | 7.38E-02   | 4.6           | 3.8        | 5.3        |
|                            |      | Relaxed     |              | 108.9                                          | 4.50E-02   | 6.51E-02   | 2.9           | 2.2        | 3.8        |
| E               |      | Strict      |              | 6.07E-02                                          | 4.63E-02   | 7.78E-02   | 6.3           | 5.2        | 7.5        |
|                            |      | Relaxed     |              | 103.8                                          | 5.67E-02   | 8.46E-02   | 3.2           | 2.3        | 4.4        |
| **China (N = 20)**           |      | Resistant   |              | 1.29E-01                                          | 7.89E-02   | 1.85E-01   | 2.8           | 2.2        | 3.4        |
| Complete       |      | Relaxed     |              | 15.3                                          | 8.27E-02   | 4.01E-02   | 1.32E-01      | 2.6        | 2.1        | 3.2        |
| S               |      | Strict      |              | 3.35E-02                                          | 2.95E-02   | 3.78E-02   | 29.6          | 27.8       | 31.3       |
|                            |      | Relaxed     |              | 464                                          | 1.37E-02   | 9.83E-03   | 26.9          | 26         | 29         |
| E               |      | Strict      |              | 3.50E-02                                          | 2.82E-02   | 4.22E-02   | 26            | 26         | 27.8       |
|                            |      | Relaxed     |              | 97.3                                          | 2.31E-03   | 1.95E-03   | 26.2          | 26         | 26.9       |
| **Complete**     |      | Strict      |              | 5.15E-02                                          | 4.01E-02   | 6.35E-02   | 26.5          | 26         | 27.5       |
|                            |      | Relaxed     |              | 159.3                                          | 3.05E-02   | 4.43E-02   | 27.2          | 26         | 29.8       |
| S               |      | Strict      |              | 6.90E-02                                          | 4.97E-02   | 9.03E-02   | 26.5          | 26         | 27.5       |
| E               |      | Relaxed     |              | 40.1                                          | 2.63E-02   | 1.40E-02   | 26.8          | 26         | 28.9       |
used. Similar results were obtained when the E gene was used. For Genogroup 2 dataset, the TMRCA was estimated to be about 2 years before present, which was consistent among all genes. For the China dataset, about 26 years before present was estimated, which was also consistent among all genes.

3.3. Genealogy among different genes

In order to determine how the topology and support for the genealogies differed between genes with a strong phylogenetic signal (nsp3 and S genes) and weak signal (E gene), we examined both the topology and posterior probability of the MCC tree for the nsp3, S and E genes in all three different datasets under the relaxed molecular clock and the constant size coalescent prior. The example of the MCC trees based on three different genes from the worldwide dataset is shown in Fig. 2. Each gene is comprised of the same number of taxa (n = 34), and each tree is a bifurcating rooted tree, so the number of internal nodes (including the root) will be the same. To compare the degree of posterior support for the MCC tree inferred from each gene, we calculated the sum of the posterior support for all internal nodes of the MCC tree, which represented the estimate of the total probability of the given tree topology. The results suggested that the nsp3 and S genes showed a similar sum of the posterior probability in all three datasets and the E gene had the lowest results (Table 4). In order to compare the particular aspects of the topologies of each genealogy, clades with relatively high posterior support (posterior probability > 0.8) were counted and the proportion of nodes with high support was compared among three different genes from all three datasets. The result with the highest support was obtained from nsp3 and S genes from all three datasets, despite that a slightly lower proportion (0.52) of high support for nsp3 gene from the Genogroup 2 dataset was noticed than that of S gene (0.61) (Table 4). Similar to the low sum of posterior probability of the E gene, the proportion of high support was lowest for the E gene among all three datasets.

3.4. Phylogenetic analysis

Since the nsp3 and S genes showed similar evolutionary rate and phylogenetic topology, the nsp3 genes from three clinical samples of swine with PEDV infection were obtained and aligned with 34 nsp3 genes available in GenBank. The results suggested that the viruses from the 2014 PEDV epidemic in Taiwan were highly clustered with the viruses from the US within Genogroup 2 (Fig. 3A). While preparing the manuscript, comprehensive PEDV full-genome sequences collected from the US were obtained and the phylogenetic tree was further constructed based on the nsp3 gene including 74 and 3 PEDV sequences from the US and Taiwan, respectively. The results suggested that the virus responsible for the 2014 PEDV outbreak in Taiwan clustered closely together with Clade I from the US (Fig. 3B).

Table 4

| Dataset          | Gene | Sum of posterior probabilities | Proportion of nodes with posterior probability >0.8 |
|------------------|------|--------------------------------|-----------------------------------------------|
| Worldwide (N = 34) | NSP3 | 27.48                          | 0.76 (25/33)                                   |
|                  | S    | 26.79                          | 0.73 (24/33)                                   |
|                  | E    | 17.85                          | 0.45 (15/33)                                   |
| Genogroup 2 (N = 24) | NSP3 | 17.5                           | 0.52 (12/23)                                   |
|                  | S    | 17.32                          | 0.61 (14/23)                                   |
|                  | E    | 6.75                           | 0.13 (3/23)                                    |
| China (N = 20)   | NSP3 | 17.82                          | 0.84 (16/19)                                   |
|                  | S    | 18.55                          | 0.84 (16/19)                                   |
|                  | E    | 13.48                          | 0.63 (12/19)                                   |

Fig. 2. Bayesian maximum clade credibility phylogenetic trees based on the spike (S, left), nsp3 (middle), and E (right) genes for the worldwide dataset according to the indicated substitution models and molecular clock suggested by the ModelTest and Bayes factors. Branch lengths are scaled in years. Branch with posterior support >0.8 are labeled inside the node. Sequences colored in red clustered within Genogroup 1 and black within Genogroup 2 based on the full-genome phylogenetic analysis (Huang et al., 2013). Each strain is labeled based on the accession number on GenBank followed by the host, country and year, which could refer to supplementary Table S1 for details.

Fig. 3. Phylogenetic trees based on the spike (S, left), nsp3 (middle), and E (right) genes from the worldwide, Genogroup 2 and China databases. Branch lengths are scaled in years. Branch with posterior support >0.8 are labeled inside the node. Sequences colored in red clustered within Genogroup 1 and black within Genogroup 2 based on the full-genome phylogenetic analysis (Huang et al., 2013). Each strain is labeled based on the accession number on GenBank followed by the host, country and year, which could refer to supplementary Table S1 for details.
4. Discussion

Phylogeographic inference of the routes and rates of the dissemination are important to understand in order to plan effective intervention, particularly for new and potentially much more deadly infectious diseases. Until now, only a few studies that used full-genome sequences were able to define a meaningful geographic structure in Asia (Huang et al., 2013; Sun et al., 2015). However, because of the relatively high cost of sequencing the full genome and the enormous computational time required for long sequences, comprehensive phylodynamic analyses have been unfeasible. Although several investigators have reported that the S gene is appropriate to study the genetic relatedness of PEDV (Chen et al., 2014; Lee et al., 2010), a comprehensive analysis has not been performed to determine which genes are the most appropriate for such phylogenetic studies. In the current study, the phylogenetic noises have been evaluated by likelihood mapping method and validated by the phylogenetic method which compares the substitution model, evolution rate, tree topology and node support proportion. Our study suggested that both nsp3 and S genes are suitable for phylogenetic tree construction and phylodynamic analysis.

RNA synthesis during coronavirus infection is carried out by a replicase–transcriptase composed of 16 nonstructural protein (nsp) subunits. Among these, nsp3 is the largest and the first to be inserted into the endoplasmic reticulum (Hagemeijer et al., 2014). Nsp3 comprises multiple structural domains including two papain-like proteases (PLPs), which are predicted to be conserved among all CoVs. Other than polyprotein processing of the cleavage between nsp1–nsp2 and nsp2–nsp3, PLPs also possess a deubiquitinase activity, which acts to counter host innate immunity (Mielech et al., 2014). Also, the N-terminal region of the nsp3 is highly conserved, containing a ubiquitin-like globular fold followed by a flexible, extended acidic-domain rich in glutamic acid and catalytically active ADP-ribose-100-phosphatase (ADRP, app-100-pase) domain, which are thought to play a role during the synthesis of viral subgenomic RNAs (Báez-Santos et al., 2015; Oostra et al., 2008). Interestingly, comparative genome and proteome analyses of two bovine CoV (BCoV) isolates showed a predominant clustering of mutations within the nsp3 multidomain (Chouljenko et al., 2001). Despite that the detailed function of nsp3 during PEDV infection is currently unknown, the multifunctionality of the nsp3, the frequency of point mutations observed in nsp3 domains, and the involvement of nsp3 in structural arrangements of the replicase complex together with double-membrane vesicles may render pleiotropic effects, not only in pathogenicity, but also on future emerging coronaviruses (Snijder et al., 2003). A recent study identified four hypervariable regions suitable for phylogenetic analysis from whole genome analysis including regions covering the nsp3 and S genes (Sun et al., 2015). Our study confirmed the results and further suggested that differential evolutionary patterns were responsible for both genes under different genotypes or spatial transmission patterns.

The 3′ end of PEDV genome containing four structural proteins (S, E, M and N) and one nonstructural protein (ORF3) plays an important role during RNA binding after phosphorylation, virus assembly, immunogenic activity and virulence (de Haan et al., 1998). The E protein, a small transmembrane protein of 76 to 109 amino acids in length, plays a
In conclusion, we have shown that the S and nsp3 genes contained the lowest phylogenetic noise, which are both suitable for phylogenetic analysis and molecular characterization. Although differential evolutionary patterns were responsible for the evolution of both genes under different geographic or genogroup-reduced databases, the constructed phylogenetic trees utilizing the S and nsp3 genes could complement each other to identify the emergence of epidemic strain resulting from recombination. The robustness of the results between the S and nsp3 genes leads us to recommend the nsp3 genomic region as an alternative marker for a rapid and unequivocal classification of the circulating PEDV strains and in providing complementary information to the understanding of PEDV epidemiology.

Appendix A. Supplementary data

Supplementary data to this article can be found online at http://dx.doi.org/10.1016/j.meegid.2015.09.011.

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