Toxins produced in cyanobacterial water blooms – toxicity and risks

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ABSTRACT
Cyanobacterial blooms in freshwaters represent a major ecological and human health problem worldwide. This paper briefly summarizes information on major cyanobacterial toxins (hepatotoxins, neurotoxins etc.) with special attention to microcystins - cyclic heptapeptides with high acute and chronic toxicities. Besides discussion of human health risks, microcystin ecotoxicology and consequent ecological risks are also highlighted. Although significant research attention has been paid to microcystins, cyanobacteria produce a wide range of currently unknown toxins, which will require research attention. Further research should also address possible additive, synergistic or antagonistic effects among different classes of cyanobacterial metabolites, as well as interactions with other toxic stressors such as metals or persistent organic pollutants.

KEY WORDS: microcystin; tumor promotion; peptide toxins; cylindrospermopsin; ecotoxicology

Toxic cyanobacterial water blooms
Massive proliferations of cyanobacteria in freshwater, brackish and coastal marine ecosystems have become a worldwide environmental problem. Anthropogenic eutrophication (i.e., increased input of nutrients, especially phosphorous but also nitrogen) of surface waters leads to accelerated growth of photoautotrophic organisms including cyanobacteria. In Europe, Asia and America, more than 40% of lakes and reservoirs are now eutrophic and offer favourable conditions for cyanobacterial mass development (Bartram et al., 1999). Furthermore, consequences of global climate changes (elevated temperature, increased atmospheric concentrations of carbon dioxide, elevated UV fluxes) have been discussed in connection with cyanobacterial ecology and growth (Beardall & Raven, 2004).

Cyanobacterial blooms formed by planktonic species or mats of benthic cyanobacteria have severe impacts on ecosystem functioning, e.g., disturbances of relationships among organisms, changes of biodiversity, light conditions or oxygen concentrations. The occurrence of cyanobacterial mass populations can create a significant water quality problem, especially as many cyanobacterial species are capable of synthesizing a wide range of odours, noxious compounds or potent toxins (Sivonen & Jones, 1999).

It has been estimated that 25 to 75% of cyanobacterial blooms are toxic (Chorus, 2001; Bláhová et al., 2007; 2008). Production of cyanobacterial toxins (cyanotoxins) includes human and animal health hazards, which can present risks of illness and mortality at environmentally relevant concentrations (Codd et al., 2005a). Thus, cyanotoxins represent important group of chemical compounds also from viewpoints of ecotoxicology, toxicology and environmental chemistry.

Health risks of cyanotoxins
Eutrophication but also other environmental factors enhance bloom formations such as low turbulence, stagnant water conditions, higher pH values and higher temperature. Under these circumstances, cyanotoxins can reach high concentrations in waters and might represent health and ecological risks (Codd et al., 2005b, Bláhová et al., 2008). Numerous incidents of animal and human poisonings (Table 1) associated with cyanobacterial blooms were reported. However, risks of cyanotoxins need not to be exclusively restricted to planktonic cyanobacteria and eutrophicated waters, because animal deaths linked to toxic populations of benthic cyanobacteria have been documented as well (Edwards et al., 1992) and lethal incidents occurred also in oligotrophic lakes (Mez et al., 1997).
Cyanobacterial toxins – cyanotoxins

Cyanobacteria have the ability to form a great variety of several secondary metabolites, which exhibit various types of biological or biochemical activities and some of them have been identified as potent toxins (cyanotoxins). The cyanotoxins are a diverse group of compounds, both from the chemical and the toxicological points of view. In terms of their toxicological target, cyanobacterial toxins are hepatotoxins, neurotoxins, cytotoxins, dermatotoxins and irritant toxins (Wiegand & Pflugmacher, 2005).

According to their chemical structures, cyanotoxins fall into several main groups: peptides, heterocyclic compounds (alkaloids) or lipidic compounds (Sivonen & Jones 1999). Cyanobacterial lipopolysaccharides, integral component of cell wall of all cyanobacteria, are usually classified as cyanotoxins (Sivonen & Jones, 1999; Codd et al., 2005a), because they possess some toxic effects. List of

| Year | Location (source) | Cyanobacteria | Toxin | Health outcomes |
|------|-------------------|---------------|-------|-----------------|
| 1931 | USA, Ohio river   | Microcystis   | ?     | gastroenteritis, abdominal pain, vomiting |
| 1960–1965 | Zimbabwe, Harare | Microcystis   | ?     | gastroenteritis |
| 1975 | USA, Pennsylvania | Schizothrix, Lyngbya, Phormidium | ? | gastroenteritis |
| 1979 | Australia, Palm Island | Cylindrospermopsis | CYN | gastroenteritis, liver, kidney and intestine damage |
| 1981 | Australia, Armidale | Microcystis | MC | liver damage |
| 1977–1996 | China | Microcystis | MC | colorectal cancer, deaths |
| 1972–1990 | China | Microcystis | MC | primary liver cancer, deaths |
| 1988 | Brazil, Itaparica dam | Microcystis, Anabaena | ? | gastroenteritis, diarrhoea, deaths |
| 1994 | Sweden, Malmö | Planktothrix | MC | gastroenteritis, fevers, abdominal and muscular pain |

| Year | Location (source) | Cyanobacteria | Toxin | Health outcomes |
|------|-------------------|---------------|-------|-----------------|
| 1959 | Canada, Saskatchewan | Microcystis, Anabaena circinalis | ? | headache, nausea, muscular pain, vomiting, diarrhoea, |
| 1980–1981 | USA, Pennsylvania and Nevada | Aphanizomenon, Anabaena | ? | eye and ear irritation, flu like symptoms |
| 1989 | UK, England, Stafordshire | Microcystis | MC | gastroenteritis, sore throat, blistered mouth, vomiting, abdominal pain, fever, pulmonary consolidation, diarrhoea |
| 1995 | Australia | Microcystis, Anabaena, Aphanizomenon, Nodularia | ? | gastroenteritis, flu like symptoms, blistered mouth, fever, eye and ear irritation, vomiting, diarrhoea |
| 1996 | UK | Planktothrix | MC | rashes, fever |
| 1996–1998 | Australia (coastal sea) | Lyngbya | ? | contact dermatitis, eye and ear irritation, respiratory irritation |
| 2002–2003 | Finland | Anabaena lemmermannii | STX | fever, eye irritation, abdominal pain, rashes |

| Year | Location (source) | Cyanobacteria | Toxin | Health outcomes |
|------|-------------------|---------------|-------|-----------------|
| 1974 | USA, Washington | present | LPS | fever, myalgia, chills, vomiting |
| 1996 | Brazil, Caruaru | present | MC, CYN | visual disturbance, tinnitus, nausea, vomiting, liver damage, deaths |
| 2001 | Brazil, Rio de Janeiro | Anabaena, Microcystis | MC | visual disturbance, tinnitus, nausea, vomiting, liver damage |

Abbreviations: MC - microcystin, CYN - cylindrospermopsin, STX - saxitoxin, LPS - lipopolysaccharides, “?” - toxin unknown. (compiled from WHO 1998b; Chorus and Bartram 1999; Duy et al. 2000; Codd et al. 2005a; Rapala et al. 2005; Falconer 2006).
the most important and investigated cyanotoxins is given in Table 2.

**Hepatotoxic heptapeptides – microcystins**

Microcystins are probably the most prevalent cyanotoxins in the environment and they are present in high amounts in cyanobacterial biomass (up to 1% of dry weight). In spite of their intensive research, the natural physiological or ecological function of microcystins is not well understood (Welker & von Dohren, 2006). Microcystins are family of monocyclic heptapeptides (more than 70 variants have been identified) with the characteristic feature, unusual β-amino acid, Adda (3-amino-9-methoxy-2,6,8-trimethyl-10-phenyldeca-4E, 6E-dienoic acid). Molecular weight of microcystins varies in the range of 909 to 1 115 (Figure 1). For microcystin-LR, World Health Organization (WHO) derived a value of tolerably daily intake (TDI) for human health risks assessment purposes. The TDI of 0.04 μg/kg bw/day (WHO, 1998a) was used for calculation of guidance value for the maximal acceptable concentration of microcystin-LR in drinking water, 1 μg/L (WHO, 1998a), and it was used also for human health risk assessment of microcystins resulting from other exposure routes, e.g., recreational exposure, consumption of contaminated food or blue-green algal food supplements (WHO, 1998b; Xie et al., 2005). This limit has been already implemented into Czech national legislation (National Drinking Water Decree No. 252/2004 Coll). However, microcystin-LR is not the only common structural variant of microcystin, and using of guideline value for total microcystin is preferable according to recent recommendation of experts (Chorus, 2005).

### Table 2. Principal groups of cyanobacterial toxins, their acute toxicities, structures and known producers.

| Toxins (LD50 - acute toxicityA) | Structure (number of variants) | Activity | Toxigenic genera |
|--------------------------------|-------------------------------|----------|-----------------|
| **Hepatotoxins**               |                               |          |                 |
| Microcystins (25 to ~1000)     | Cyclic heptapeptides (71)     | Hepatotoxic, protein phosphatase inhibition, membrane integrity and conductance disruption, tumour promoters | MicrocystisBCD, AnabaenaBCD, NostocBC, PlanktothrixBCD, Anabaenopsis B, HapalosiphonBC |
| Nodularins (30 to 50)          | Cyclic pentapeptides (9)      | Hepatotoxic, protein phosphatase inhibition, membrane integrity and conductance disruption, tumour promoters, carcinogenic | NodulariaBCD |
| Cylindrospermopsins (200 to 2100) | Guanidine alkaloids (3) | Necrotic injury to liver (also to kidneys, spleen, lungs, intestine), protein synthesis inhibitor, genotoxic | CylindrospermopsisBC, AphanizomenonBC, Anabaena2, RaphidiopsisBC, UmezakiaBC |
| **Neurotoxins**                |                               |          |                 |
| Anatoxin-a (250)               | Tropane-related alkaloids (5) | Postsynaptic, depolarising neuromuscular blockers | AphanizomenonBC, AnabaenaBCD, RaphidiopsisBC, OscillatoriaBC, PlanktothrixBC, CylindrospermumBC |
| Anatoxin-a(S) (40)             | Guanidine methyl phosphate ester (1) | Acetylcholinesterase inhibitor | AnabaenaBC |
| Saxitoxins (10 to 30)          | Carbamate alkaloids (20)      | Sodium channel blockers | AphanizomenonBC, AnabaenaBC, PlanktothrixBC, CylindrospermopsisBC, LyngbyaBC |
| **Dermatotoxins (irritants) and cytotoxins** | | | |
| Lyngbyatoxin-a                 | Alkaloid (1)                  | Inflammatory agent, protein kinase C activator | LyngbyaBC, SchizothrixB, OscillatoriaBC |
| Aplysioxin                     | Alkaloids (2)                 | Inflammatory agents, protein kinase C activators | LyngbyaBC, SchizothrixB, OscillatoriaBC |
| **Endotoxins (irritants)**     |                               |          |                 |
| Lipopolysaccharides            | Lipopoly-saccharides          | Inflammatory agents, gastrointestinal irritants | All cyanobacteria? |

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A acute toxicity in mouse bioassay (i.p. exposure, LD50 - μg/kg body weight); B toxin identified in natural population with dominant genera; C toxin identified in non-axenic monocyanobacterial culture (not bacteria free); D toxin identified in axenic monocyanobacterial culture (cyanobacteria free). (Compiled from Codd et al., 2005a; Codd et al., 2005b).
**Acute toxicity of microcystins**

Microcystins have been shown to be acutely (and also chronically toxic) to animals and humans (WHO, 1998a; Duy et al., 2000; Dietrich & Hoeger, 2005) with acute LD50s of the individual microcystin structural variants ranging between 50 (microcystin-LR) and 1000 ([(6Z)-Adda]microcystin-RR) μg/kg b.w. following i.p. injection in mice. The main mechanism of toxicity is the irreversible inhibition of protein phosphatases 1 and 2A, which are key regulatory enzymes in catalyzing dephosphorylation of serine/threonine residues in various phosphoproteins (structural proteins, enzymes, regulators). Inhibition of protein phosphatases is followed by loss of cytoskeletal integrity and subsequent cytolysis or apoptosis, primarily of hepatocytes (Dietrich & Hoeger, 2005). After acute i.p. exposure, severe liver damage is observed followed by haemodynamic shock, heart failure and death (Dawson, 1998). The oral LD50 in mice (5000 μg/kg b.w.) or in rats (>5000 μg/kg b.w.) is approximately 100 fold lower than the i.p. LD50 (Yoshida et al., 1997), may be due to slow gastrointestinal uptake of toxins in mice.

Oxidative stress seems to be another important biochemical mechanism of microcystin toxicity. Microcystins have been shown to induce formation of reactive oxygen species (ROS) that might cause serious cellular damage such as peroxidation of lipid membranes, genotoxicity, or modulation of apoptosis (Ding & Ong, 2003). The formation of ROS is the most likely the mechanism responsible for oxidative damage of DNA, genotoxic and clastogenic effects of microcystins (Humpage et al., 1999, 2000; Bouaicha et al., 2005). However, the exact mechanism of oxidative stress promoted by microcystins is still not known.

**Subchronic and chronic toxicity, tumour promotion**

Several experiments with mammals (rodents, pigs) showed significant subchronic and chronic toxicity of orally administered microcystins (Falconer 2006; Fawell et al., 1999), where harmful effects of microcystins such as increased mortality, liver injury (including histopathological changes, chronic inflammation, degeneration of hepatocytes, increased liver enzyme levels), renal damage or slightly higher number of tumours were observed.

Microcystins are considered to be tumour promotion factors. There has been evidence of tumor promotion properties of microcystins from several animal experiments (Humpage et al., 2000; Dietrich & Hoeger, 2005). These findings are supported by results of studies showing effects of microcystins on cell proliferation and cytokinesis, which might be associated with tumour promotion (Gehringer, 2004; Guzman et al., 2003; Fu et al., 2005). Moreover, in epidemiological studies in China, the incidence of liver or colorectal cancer was related to consumption of water originated from sources contaminated with microcystin or microcystin-producing cyanobacterial blooms (Yu, 1995; Zhou et al., 2002).

**Ecotoxicology of microcystins**

The majority of microcystin-related research has focused on mammalian toxicity. However, microcystin-producing blooms have been frequently involved in many incidents of fatal animal poisonings, including cattle, sheep, chickens,
pigs, horse s, dogs, poultry and wild birds, fish or even rhinoceroses (Duy et al., 2000; Briand et al., 2003).

Moreover, wide range of aquatic organisms is directly exposed to microcystins contained in their food (phytoplanktivorous fish, zooplankton etc.) and/or to microcystins dissolved in water, which may cause diverse effects. Therefore, more attention is also recently paid to investigations of microcystins ecotoxicity and their effects on aquatic biota. Although previous experiments concentrated mainly on fish and daphnids, there is an increasing number of studies with other species such as phytoplankton, submerged plants (macrophytes), various crustaceans and molluscs (Babica et al., 2006, 2007; Wiegand & Pfugmacher 2005; Zurawell et al., 2007). The ecological relevance of experiments (e.g., exposure duration and routes, experimental concentrations/doses) as well as evaluation of sublethal and chronic effects are particularly emphasized nowadays. As microcystins have been shown to accumulate in various organisms (plants, zooplankton, molluscs and fish), investigations of microcystin transfer through lake food webs are also highly needed (Adamovský et al., 2007). The bioaccumulation and possible effects on human health should be in the focus of future research.

Other cyanotoxins and bioactive compounds

In comparison with microcystins, substantially less attention has been paid to other cyanobacterial compounds, especially from the ecotoxicological point of view. Although there are several studies investigating effects of anatoxin-α or saxitoxins on aquatic organisms (see Wiegand & Pfugmacher, 2005 for review), only limited number of published papers concerned other cyanotoxins such as anatoxin-a(5), cylindrospermopsin or cyanobacterial lipo-polysaccharides. Since these “non-traditional” cyanotoxins are not routinely monitored, their importance in aquatic ecosystems can be underestimated (Bláhová et al., 2009).

Moreover, a number of studies reported toxic effects of cyanobacterial extracts and biomass, which could not be accounted for by those cyanobacterial metabolites currently termed “the cyanotoxins” (Oberemm et al., 2001). Besides cyanotoxins referred above, there have been identified many other substances of cyanobacterial origin which possess some kind of biological activity or toxicity (Wekerl & von Dohren, 2006). Toxicity of volatile compounds (e.g., geosmine) produced by cyanobacteria was also demonstrated (Watson, 2003). Interestingly, cyanobacterial fatty acids can also be toxic or modulate effects of other cyanotoxins (Ikawa et al., 1997; Reinikainen et al., 2001).

Conclusions

Degradation of aquatic ecosystems by nutrient pollution resulting in massive cyanobacterial water blooms is a global problem representing serious health and ecosystem risks. Although significant research attention has been paid to selected cyanotoxins (mostly microcystins), it is nowadays recognized that cyanobacteria may produce wide range of currently unknown toxins. The results of experiments, where have been observed toxic effects but no causative compound has been identified so far, are showing great ecotoxicological or toxicological significance of unidentified or as-yet-unknown substances. Further research is also needed on possible additive, synergistic or antagonistic effects to multiple classes of cyanobacterial bioactive metabolites, or studies of interactions between the cyanotoxins and other stressors, e.g., anthropogenic toxicants such as metals or persistent organic pollutants (Codd et al., 2005a).

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