Abstract

Introduction: C-reactive protein (CRP) is an acute phase protein and was shown to be a predictor for all-cause and cardiovascular death in end stage renal disease patients. C-reactive protein is a member of the pentraxin family and plays a key role in the innate immune response. Several single nucleotide polymorphisms (SNP) associated with plasma CRP levels have been identified. This study aimed to evaluate the frequency of the rs1800947 polymorphism in the human CRP gene in maintenance hemodialysis (HD) patients, and to determine whether it is related to nutritional status.

Material and methods: We investigated 99 patients maintained on chronic hemodialysis (HD). 109 apparently healthy volunteers served as a control group. HD subjects were followed for 12 months. Anthropometric measurements were performed, the malnutrition-inflammation scale (MIS) score was determined, and blood samples were obtained within 1 month of enrollment and at 3 and 12 months of follow-up. rs1800947 SNP in the CRP gene was genotyped using restriction fragment length polymorphism-polymerase chain reaction.

Results: There were no significant differences in the investigated genotype frequencies between HD patients and controls. Moreover, there was no influence of genotype on anthropometric and laboratory indices of nutritional status and MIS score.

Conclusions: The results of this study indicate that the rs1800947 single nucleotide polymorphism in the CRP gene does not seem to correlate with nutritional status in hemodialysis patients.

Key words: C-reactive protein, single nucleotide polymorphism, chronic kidney disease, hemodialysis, nutritional status.
to 1.1 million deaths worldwide each year [1, 2]. CKD patients treated with chronic hemodialysis (HD) have the highest mortality rate within this group [3]. There is a strong association between mortality in patients who are treated with HD and their nutritional status as assessed by simple indicators [4].

Persistent low-grade inflammation has been implicated in the pathogenesis of malnutrition in HD patients, and several investigators have found that in patients maintained on chronic HD there is an association between markers of inflammation and nutritional status, cardiovascular disease and overall morbidity and mortality [5, 6]. Up to 30% to 60% of HD patients have elevated CRP levels [7–9], which are independently inversely related to serum albumin levels and the risk of death [10]. However, causality has not been established for any specific inflammatory mediator [11].

Several cytokines have been implicated as factors that link malnutrition, accelerated atherosclerosis, and excessive morbidity and mortality in end-stage renal disease (ESRD) patients maintained on HD [12], also the polymorphism in several cytokine coding genes was associated with the risk of malnutrition [13, 14].

C-reactive protein (CRP) belongs to the pentraxin family. It is produced in the liver and is secreted in increased amounts within 6 h of an acute inflammatory stimulus. Interleukin-6 is the chief stimulator of the production of CRP [15]. CRP is encoded by a gene found on chromosome 1 close to the serum A amyloid gene (SAA). The locus coding for CRP is polymorphic, which may influence its expression, and subsequently plasma levels. In 2000 Cao and Hegele described CRP polymorphism based on substitution of the single nucleotide in the 1059 position of the 2nd exon (+1059G>C, rs1800947) [16]. Subsequently, several single nucleotide polymorphisms (SNP) associated with plasma CRP levels were identified [17].

Taking all this into account, we hypothesized that inflammation-related CRP gene polymorphism may contribute to nutritional status in HD patients, and in the present study we sought to determine the relative frequency of +1059 (G>C) single nucleotide polymorphism in the CRP gene in patients with ESRD treated with chronic HD, and to compare that to a control population. We also assessed the relationship between specific alleles and different indices of nutritional status in HD patients.

**Material and methods**

**Subjects**

This was a multicenter longitudinal study. The study cohort consisted of 99 Caucasian patients with ESRD, maintained on chronic HD, who were recruited from 3 hemodialysis centers. HD was performed three times weekly. Dialyzers with low-flux, modified cellulose membranes were used. No major changes were made in the dialysis treatment or schedules during the follow-up period. Table I presents baseline characteristics of the study cohort.

Most of the patients were on antihypertensive drugs and phosphate binders. Dietary recommendations during the study were as follows: protein 0.8–1.0 g/kg of ideal body mass (with at least half of the protein being of high biologic value), 30% of calories derived from fat and 62% of calories from carbohydrates.

The control group consisted of 109 apparently healthy volunteers, 46 males and 63 females, aged 46.9 ±17.9 years.

**Study protocol**

Patients were eligible for the study if they were between the ages of 18 and 90 years, had been receiving HD treatment for at least 3 months before entering the study, and agreed to participation. Exclusion criteria included: diabetes mellitus, lupus erythematosus, ulcerative colitis, Crohn’s disease, history of juvenile rheumatoid arthritis, recent acute inflammatory disease, history of neoplastic disease and treatment with steroids and immunosuppressive drugs.

| Parameter          | Value                  |
|--------------------|------------------------|
| Age [years]        | 61.6 ±13.5             |
| Gender, male, n (%)| 53 (53.5)              |
| Cause of CKD, n (%):|                       |
| Chronic glomerulonephritis | 28 (28.3)   |
| Unknown            | 19 (19.2)              |
| Chronic interstitial nephritis | 19 (19.2)   |
| Polycystic diseases | 9 (9.1)               |
| Vascular disease   | 8 (8.1)                |
| Amyloidosis        | 6 (6.1)                |
| Other              | 10 (10.1)              |
| Dialysis vintage [years] | 6.4 ±5.5       |
| BMI [kg/m²]        | 23.3 ±4.0              |
| Kt/V               | 1.31 ±0.24             |
| Serum albumin [g/l]| 4.0 ±0.74              |
| Cholesterol [mg/ml]| 177.8 ±51.6            |
Blood samples for analysis of CRP albumin, creatinine, urea, cholesterol and triglycerides were drawn before scheduled hemodialysis, as part of the routine protocol, within one month of enrollment and after 3 and 12 months. Serum was separated after centrifugation and stored at -70°C until analysis.

Nutritional status was assessed using modified SGA scale (MIS) proposed by Kalantar-Zadeh et al. [18]. Each MIS component has four levels of severity from 0 (normal) to 3 (very severe). The sum of all 10 MIS components ranges from 0 to 30, denoting the increasing degree of severity. Nutritional status was also evaluated by anthropometric measurements including: waist circumference, hip circumference, mid arm circumference (MAC), biceps skinfold thickness, triceps skinfold thickness (TSF), subscapular skinfold thickness and midarm muscle circumference (MAMC). MAMC was calculated as follows: MAMC = MAC – (3.1415 × TSF).

Subjects were weighed in light clothing. Weight was rounded to the nearest 0.5 kg and their height rounded to the nearest 0.5 cm. Skinfold thickness (SFT) was measured using a Harpenden skinfold caliper, by trained investigators with standard technique, at the triceps, biceps and subscapular sites using defined anatomical landmarks [19]. The mean of 3 measurements at each site was calculated. The waist and hip circumference were measured to the nearest 0.5 cm at the umbilicus and greater trochanter.

Kt/V (a measure of “dialysis dose”) was calculated according to the Daugirdas formula [20].

The local Ethics Committee approved the study protocol, and informed consent was obtained from all the patients and control subjects.

Laboratory methods

Routine laboratory parameters were measured in local hospital clinical laboratories. IL-6 concentration was determined with a Quantikine human IL-6 immunoassay kit (R&D systems, USA). Genomic DNA was isolated using the spin column method and a commercially available kit (QIAamp DNA Blood Mini Kit, Qiagen, Germany). Genotyping was performed after amplification of the target sequence (corresponding to nucleotides 514–1258, within 2nd exon) using specific primers (F primer corresponding to nucleotides 514–535, and reverse corresponding to nucleotides 1239–1258) [16]. After MaelII digestion reaction products were electrophoresed in polyacrylamide gel, silver stained and scanned. Digestion of the 1059G allele with MaelII produced three fragments with sizes of 312 bp, 233 bp and 200 bp, whereas digestion of the 1059C allele gave rise to two fragments with sizes of 312 bp and 433 bp.

Statistical analysis

Results are expressed as means ± standard deviations. Because baseline CRP and IL-6 concentrations were right skewed, they were log transformed before analysis.

Univariate statistical analysis was performed using the χ² test, Fisher’s exact test or Student’s t-test as appropriate. Associations between variables were analyzed using Pearson’s correlation coefficient or Spearman’s rank correlation as appropriate.

In order to estimate the effect of 1059(G>C) on variables of interest over the time of the study, we used the weighted generalized estimating equations (wGEE) approach [21] to account for individual patients with multiple exacerbations. Predictor variables in the model included gender, squared time of measurement, and age. Working correlation structure (autoregressive, exchangeable or unstructured) was selected using the quasi-AIC (QIC) method [22]. Results of GEE analysis are presented as estimated increase/decrease in the variable of interest from GEE multivariable analysis.

P-values ≤ 0.05 were considered statistically significant. The STATA software, version 9.2 for Macintosh (Stata Corporation, College Station, TX, USA) was used for most statistical computations. Weighted GEE analyses were performed using R version 3.6.1, wgeesel_1.5 package (https://CRAN.R-project.org/package=wgeesel) [23].

Results

The demographic and laboratory characteristics of the patients are provided in Table I.

Sixteen subjects died and 16 dropped out of the study because of renal transplantation (n = 5), consent withdrawal (n = 9), new-onset diabetes mellitus (n = 1) and transfer to another dialysis center (n = 1).

There were no significant differences in the GC/CC genotype frequencies between HD patients and controls (16.2% and 11.8%, respectively; p = 0.364). Fourteen patients who died had GG genotype and 2 had GC/CC genotypes (p = 0.736).

There was no difference, across the 12-month study interval, in any investigated biochemical, clinical or anthropometric parameters, between GG and GC/CC groups (Table II). “Dialysis dose” (as measured by Kt/V) was similar in both SNP groups. CRP 1059G>C SNP was not associated with serum log CRP concentration, although male GG carriers had a non-significantly higher log CRP level than GC/CC variant carriers (coefficient estimate log CRP = 0.222, robust SE = 0.1184; p = 0.06).

Male sex was associated with higher log CRP levels (coefficient estimate = 0.299 robust SE = 0.0748; p < 0.001) and log IL-6 (coefficient estimate 0.275, robust SE = 0.0966; p = 0.004) levels over...
Table II. Relationship between SNP CRP and nutritional variables in HD patients

| Variable                          | Genotype | Baseline | At 3 months | At 12 months | P-value |
|-----------------------------------|----------|----------|-------------|--------------|---------|
| Kt/V                              |          |          |             |              |         |
|                                   | GC/CC    | 1.38 ±0.20 | 1.32 ±0.21 | 1.38 ±0.15  | 0.335   |
|                                   | GG       | 1.30 ±0.24 | 1.37 ±0.22 | 1.38 ±0.22  |         |
| BMI [kg/m²]                       |          |          |             |              |         |
|                                   | GC/CC    | 22.4 ±3.0  | 22.3 ±2.4  | 23.1 ±2.8   | 0.664   |
|                                   | GG       | 23.5 ±4.1  | 23.5 ±4.3  | 22.7 ±6.3   |         |
| Waist circumference [cm]          |          |          |             |              |         |
|                                   | GC/CC    | 84.5 ±9.8  | 84.7 ±10.2 | 88.1 ±11.9  | 0.367   |
|                                   | GG       | 88.9 ±12.5 | 88.7 ±12.5 | 90.4 ±13.2  |         |
| Hip circumference [cm]            |          |          |             |              |         |
|                                   | GC/CC    | 91.9 ±5.4  | 92.9 ±4.5  | 94.5 ±6.2   | 0.750   |
|                                   | GG       | 93.4 ±7.9  | 93.7 ±8.2  | 92.3 ±14.6  |         |
| WHR                               |          |          |             |              |         |
|                                   | GC/CC    | 0.92 ±0.14  | 0.91 ±0.08 | 0.93 ±0.08  | 0.204   |
|                                   | GG       | 0.95 ±0.09  | 0.95 ±0.09 | 1.15 ±1.48  |         |
| Midarm circumference [cm]         |          |          |             |              |         |
|                                   | GC/CC    | 25.1 ±3.2  | 25.2 ±3.6  | 26.0 ±3.0   | 0.869   |
|                                   | GG       | 25.8 ±3.6  | 25.5 ±3.6  | 25.3 ±3.9   |         |
| Biceps skinfold [mm]              |          |          |             |              |         |
|                                   | GC/CC    | 8.6 ±4.2   | 9.1 ±4.3   | 10.1 ±5.6   | 0.133   |
|                                   | GG       | 10.1 ±4.8  | 9.6 ±4.4   | 11.1 ±6.6   |         |
| MAMC [cm]                         |          |          |             |              |         |
|                                   | GC/CC    | 21.5 ±3.0  | 21.2 ±2.69 | 21.6 ±2.65  | 0.391   |
|                                   | GG       | 21.3 ±1.2  | 21.3 ±2.95 | 21.0 ±3.18  |         |
| Triceps skinfold [mm]             |          |          |             |              |         |
|                                   | GC/CC    | 11.6 ±5.7  | 12.9 ±6.7  | 13.8 ±7.7   | 0.092   |
|                                   | GG       | 14.2 ±6.9  | 13.4 ±6.6  | 13.8 ±7.0   |         |
| Subscapular skinfold [mm]         |          |          |             |              |         |
|                                   | GC/CC    | 13.0 ±6.1  | 13.4 ±4.8  | 14.1 ±6.4   | 0.234   |
|                                   | GG       | 14.9 ±6.6  | 15.0 ±6.9  | 15.7 ±8.1   |         |
| Log CRP [ng/ml]                   |          |          |             |              |         |
|                                   | GC/CC    | 0.62 ±0.51 | 0.59 ±0.33 | 0.41 ±0.50  | 0.631   |
|                                   | GG       | 0.66 ±0.48 | 0.70 ±0.52 | 0.60 ±0.53  |         |
| Log IL-6 [pg/ml]                  |          |          |             |              |         |
|                                   | GC/CC    | 0.91 ±0.25 | 0.90 ±0.15 | 0.83 ±0.38  | 0.455   |
|                                   | GG       | 0.92 ±0.28 | 0.89 ±0.30 | 0.86 ±0.25  |         |
| Creatinine pre-HD [mg/dl]         |          |          |             |              |         |
|                                   | GC/CC    | 9.3 ±2.0   | 9.4 ±2.3   | 9.9 ±2.3    | 0.796   |
|                                   | GG       | 9.2 ±2.0   | 9.2 ±2.4   | 9.5 ±2.3    |         |
| Serum cholesterol [mg/dl]         |          |          |             |              |         |
|                                   | GC/CC    | 168.8 ±37.5 | 176.6 ±43.4 | 172.4 ±40.3 | 0.189   |
|                                   | GG       | 179.7 ±54.1 | 181.0 ±49.6 | 172.6 ±49.5 |         |
| Triglycerides [mg/dl]             |          |          |             |              |         |
|                                   | GC/CC    | 164.7 ±79.5 | 138.4 ±76.9 | 181.2 ±94.6 | 0.196   |
|                                   | GG       | 182.9 ±135.4 | 164.4 ±84.2 | 171.7 ±132.3|         |
| Serum protein [g/l]               |          |          |             |              |         |
|                                   | GC/CC    | 7.3 ±0.9   | 6.9 ±0.6   | 7.1 ±0.9    | 0.397   |
|                                   | GG       | 7.4 ±0.9   | 7.2 ±2.4   | 7.1 ±0.8    |         |
| Serum albumin [g/l]               |          |          |             |              |         |
|                                   | GC/CC    | 3.9 ±0.7   | 3.8 ±0.5   | 4.1 ±0.6    | 0.205   |
|                                   | GG       | 4.1 ±0.8   | 4.0 ±1.6   | 4.2 ±0.5    |         |
| TIBC [mg/dl]                      |          |          |             |              |         |
|                                   | GC/CC    | 248.2 ±63.9 | 244.2 ±67.0 | 265.1 ±50.7 | 0.885   |
|                                   | GG       | 255.3 ±59.6 | 224.1 ±64.0 | 261.3 ±83.8 |         |
| MIS (median 25–75 percentile)     |          |          |             |              |         |
|                                   | GC/CC    | 9 (5–12)   | 9 (7–12)   | 7.5 (4–10)  | 0.381   |
|                                   | GG       | 7.5 (5–10) | 9 (7–13)   | 8 (7–12)    |         |

Values derived from weighted GEE multivariable analysis for association over 12 months of variable of interest with 1059(G>C) SNP.

12 months of the study (Figure 1), and lower levels of triglycerides (coefficient estimate −50.740, robust SE = 20.7686; \(p = 0.015\)), and total cholesterol (coefficient estimate 44.565, robust SE = 8.2751; \(p < 0.001\)). Log IL-6 levels were associated with age (coefficient estimate 0.01349, robust SE = 0.0034; \(p < 0.001\)) over 12 months of the study (Figure 2).

At baseline of the study there was a positive correlation between log CRP and log IL-6 (\(r = 0.278\); \(p < 0.0001\)), and between both of the inflammatory markers and waist-hip ratio (WHR) (\(r = 0.231\); \(p = 0.0001\) and \(r = 0.191\); \(p = 0.002\) respectively). There was also an inverse correlation of baseline serum albumin and CRP and IL-6 (\(r = −0.279\); \(p < 0.0001\) and \(r = −0.169\); \(p = 0.007\) respectively).

**Discussion**

The main finding of our study is that apparently there was no difference in time course in any investigated biochemical, clinical or anthropometric parameters over 12 months, between GG and GC/CC groups.

As CRP SNPs are associated with serum CRP levels in the general population [24], which in turn is associated with nutritional status in CKD stage 5 patients, we were interested in whether long-term average concentration of CRP (as approximated by a 12-month period) would be affected by rs1800947 CRP SNP.
Frequency of the C allele (low producer) in our control group was similar to that reported by Cao and Hegele [16] for Caucasians, and did not differ significantly from the HD patient group. On the other hand, Zee and Ridker [25] found a somewhat lower frequency of that allele in a group of 700 healthy individuals in the USA. Furthermore, they additionally found that plasma CRP concentrations were significantly reduced among carriers of a 1059G>C polymorphism (GC/CC) as compared with non-carriers (GG) [25].

In our study CRP 1059G>C SNP was not significantly associated with serum CRP concentration. This is at variance with findings of others [25, 26]. It probably can be explained by the low number of subjects and confounding. We observed non-significantly higher levels of CRP in male carriers of GG genotype. Interestingly, Eklund et al. found that male carriers of the +1059 C allele had significantly lower CRP concentrations than GG homozygotes. No significant difference was found in women [27].

SNPs in the CRP gene may also influence the acute phase response, which is frequently observed in the HD patient population in addition to prolonged low-intensity reaction. In fact, this was confirmed for the rs1800947 SNP in patients with acute coronary ischemia [28], acute pancreatitis [29], ischemic stroke [30], inherited depression [31] and another CRP gene variant (+1444C/T), which influenced both basal and stimulated CRP level after strenuous physical exercise and perioperatively in coronary artery bypass grafting [32].

Differently to our findings (of no association between rs1800947 SNP and anthropometric and laboratory indices of nutritional status), subjects treated with chronic HD, with high producer genotypes for other proinflammatory cytokines such as IL-6 or TNF-α, had higher comorbidity and lower functional scores, and lower serum albumin levels [14], and the TNF-α-308 polymorphism was reportedly associated with the risk of malnutrition inflammation syndrome (MIS) [13].

The so-called ‘malnutrition–inflammation complex syndrome’ (MICS) [33] or ‘malnutrition–inflammation–atherosclerosis’ syndrome [34] is the simultaneous combination of malnutrition and inflammation. MICS appears to play a central role in poor clinical outcome including the high rate of mortality and hospitalization and diminished quality of life seen in dialysis patients. MICS is also believed to be the underlying condition of the phenomenon known as “reverse epidemiology” of cardiovascular risks in these patients, where a low, and not a high, BMI or serum cholesterol is associated with poor dialysis outcome [35].

Due to the relatively short duration of the study we have not attempted to correlate actual morbidity and mortality with CRP genotype. There are limitations of our study that merit consideration. First, and most important, is the small size of the study, which therefore may be regarded rather as hypothesis generating. Second, while our analysis focused on the impact of only one CRP polymorphism, a combination of different polymorphisms, and other genetic factors as well as environmental factors likely interact with respect to investigated parameters.

In conclusion, the investigated polymorphism does not seem to correlate with nutritional status in hemodialysis patients.

**Acknowledgments**

This study was supported by grant no. 3P05B02024 from the Committee for Scientific Research, Poland.

**Conflict of interest**

The authors declare no conflict of interest.
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