Comparative transcriptome analysis two genotypes of *Acer truncatum* Bunge seeds reveals candidate genes that influences seed VLCFAs accumulation

Rongkai Wang, Pei Liu, Jinsuan Fan & Lingli Li

The *Acer truncatum* Bunge is a particular widespread forest tree species in northern China. VLCFAs are important to eukaryotes survival and play diverse roles throughout the development. So far, there are reports that the *Acer truncatum* seeds fatty acid (FA) rich in VLCFAs, but little is known about the physiological mechanism responsible for the biosynthesis. A total of approximately 37.07 Gbp was generated, it was comprehensive enough to determine the majority of the regulation VLCFAs biosynthesis genes. The 97,053 different unigenes were assembled and identified, and large numbers of EST-SSRs were determined. The expression profiles of crucial genes (*KCS*, *KCR*, *HCD* and *ECR*) involved in VLCFAs elongation of fatty acids were also studied. To our knowledge, the present study provides the first comprehensive of the transcriptome of *Acer truncatum* seeds. This transcriptome dataset have been made publicly available NCBI, we believe that it may provide new resource for future high-throughput gene expression of *Acer truncatum* seeds growth and development and will provide theoretical basic information for improving the yield of VLCFAs, especially nervonic acid.

The *Acer truncatum* Bunge, a particular widespread forest tree species in northern China, Japan and Korea, and is also found in Europe and Northern America. The seeds fatty acid (FA) rich in nervonic acid (24:1; cis-tetracos-15-enolic acid) and it has been officially admitted as edible oil by the Ministry of Health of China. Nervonic acid is a very long chain fatty acids (VLCFAs). VLCFAs are fatty acids with an acyl chain of 20–30 carbons and longer. The chain length, the type of polar head, the degree of unsaturation and the associated lipids provide the structural and functional diversity of these fatty acids. VLCFAs are important to eukaryotes survival and play diverse roles throughout the growth and development. In addition, VLCFAs are important feedstocks for industrial, pharmaceutical and nutraceutical application. Vegetable oil, as neat fuel, has been to the main source of VLCFAs; therefore, the increase of VLCFA contents in seeds has become an important target for oilseed enhancement.

VLCFAs are synthesized by a microsomal fatty acid extension (FAE) system. Specifically, the C18 fatty acid is first synthesized using the *de novo* fatty acid synthesis pathway of plastids, and the C2 portion of malonyl-coenzyme A (CoA) is sequentially added to the previously synthesized C18 fatty acids. Analogous to FAS, the FAE is a membrane-bound fatty acid elongation complex, which involves 4 enzymatic reactions: condensation of C18-CoA with malonyl CoA to form a ketoacyl-CoA by ketoacyl-CoA synthase (KCS), reduction of ketoacyl-CoA to a hydroxyacyl-CoA by ketoacyl-CoA reductase (KCR), dehydration of hydroxyacyl-CoA to a enoyl-CoA by hydroxyacyl-CoA dehydratase (HCD), and reduction of enoyl-CoA by enoyl-CoA reductase (ECR). The KCS is thought to be the rate-limiting enzyme for the elongation of long-chain fatty acids, since it determines the substrate and tissue specificity of long-chain fatty acid elongation. In addition, regulating the expression of the KCS gene affects the final contents of VLCFAs. In contrast, the other 3 enzymes have no substrate specificity and tissue specificity for VLCFA biosynthesis.

Recently, RNA-Seq has become a very effective and powerful technology in generating comprehensive transcriptome dataset. Studies in the past have indicated the rapid identification and profiling of differentially
expressed genes by de novo transcriptome sequencing in some oil seeds, such as flax, castor bean, olive, peanut, sea buckthorn, tree peony and *Camellia oleifera*\(^{21-27}\). So far, there are reports that the *Acer truncatum* seeds FA rich in VLCFAs\(^3\), but little is known about the physiological mechanism responsible for VLCFAs biosynthesis.

In this study, we analyzed the transcriptome of *Acer truncatum* using high-throughput Illumina sequencing technology. In total, more than 37.07 Gbp was generated, it was comprehensive enough to determine the majority of the regulation VLCFAs biosynthesis genes. The 127,791 different transcripts and 97,053 unigenes were assembled and identified, and large numbers of EST-SSRs were determined. To our knowledge, this study provides the first comprehensive of the transcriptome of *Acer truncatum* seeds. This transcriptome dataset have been made publicly available NCBI, we believe that it may provide new resource for future research on bioengineering breeding and will provide theoretical basic information for improving the yield of VLCFAs.

**Results and Discussion**

**FA profiling in different genotypes of *Acer truncatum* seeds.** Based on the analysis of thirty-six genotypes *Acer truncatum* seeds FA, we found that different genotypes seeds showed different FA content and composition. There mainly were six kinds of VLCFAs in *Acer truncatum*, namely arachidic acid (C20:0), eicosenoic acid (C20:1), behenic acid (C22:0), erucic acid (C22:1), lignoceric acid (C24:0), and nervonic acid (C24:1). Although the accumulation of major VLCFAs varied in different genotypes seeds, the three dominant components of VLCFAs composed of eicosenoic acid (C20:1), erucic acid (C22:1) and nervonic acid (C24:1) (Tables S1 and S2).

The results showed that the highest content of VLCFAs genotype was H-11, and the lowest genotype was L-4. Additionally, there were higher content of Nervonic acid (C24:1) in H-11 seeds. However, no obvious differences were observed in several seed morphological traits including seed seed size and dry weight between seeds of H-11 and L-4 plants seeds (Fig. 1 and Table S2). To investigate the biological function on seed VLCFAs accumulation, we performed transcriptome sequencing of these two genotype seeds.

**Illumina sequencing and de novo assembly.** The total of the *Acer truncatum* seeds mRNA was isolated from a single plant (H-11 or L-4). The sequencing raw data through rigorous quality assessment and data filtering, about 6.23 Gb, 6.21 Gb and 6.08 Gb, as well as 6.13 Gb, 6.21 Gb and 6.21 Gb for H-11 and L-4, respectively. It is presented in Table 1. The high quality sequencing reads has been deposited in the NCBI. Using the Trinity software program, the high quality sequencing reads were de novo assembled\(^{28}\), which produced 127,791 transcripts with an N50 length of 1,122 bp and a mean length of 686.76 bp. The distribution of the transcripts are depicted in Fig. S1 and Table S3. These transcripts were further analyzed for cluster and assembly. We have obtained 97,053 unigenes with an N50 length of 938 bp and a mean length of 598.19 bp for further analysis. It is shown in Fig. 2 and Table 2.

**Functional annotation of all unigenes.** The assembled unigenes sequences were annotated based on the following databases: NR, SWISS-PROT, GO, COG and KEGG. An overview of functional annotation in Table 3. We used BLASTX to similarity analysis and compared against the NR database. In these assembled unigenes,
71,014 (73.57%) unigenes had significant matches, and about 30% unigenes were showed no significant matches. Previous studies have shown that sequencing of cDNA libraries does not significant hits sequences is about 25% to 35%. Among a wide range of plants with protein sequences, the Acer truncatum assembled unigenes had the highest number of hits against Citrus sinensis at 6,886 hits, followed by Citrus clementina at 4,687 hits, Theobroma cacao at 1,453 hits, Ricinus communis at 1,005 hits, Vitis vinifera at 755 hits, Manihot esculenta at 665 hits, Jatropha curcas at 656 hits and Ziziphus jujuba at 538 hits (Fig. 3). It depicts that the higher similarity of Acer
truncatum unigenes and Citrus sinensis genes suggests the possibility we can use Citrus sinensis transcriptomes and genomes as a reference for further analysis.

The SWISS-PROT database is the manually annotated and reviewed protein sequence database. It is a high-quality database of annotated and non-redundant protein sequences, and the results contain experimental results, computational features and scientific conclusions. Among the 97,053 unigenes, 26,076 (26.87%) were similar to the SWISS-PROT database (Table 3). Using the GO database enrichment analysis, the identified assembled unigenes were carried out to classify three independent sets (the cellular component, the molecular function, and the biological process). It depicts that the majority GO terms were assigned to the biological process is 59,615, the molecular function had 27,109 terms assigned, and the cellular component had 60,490 terms assigned (Fig. 4).

Furthermore, the assembled unigenes were searched against the COG database. A total of 30,605 unigenes have been assigned to the COG classification (Fig. 5). The highest group is the cluster for function prediction only (7,042, 23.01%), and followed by amino acid transport and metabolism (4,110, 13.43%); carbohydrate transport and metabolism (3,613, 11.81%); inorganic ion transport and metabolism (2,579, 8.43%); energy production and conversion (2,545, 8.32%); transcription (2,493, 8.15%); replication, recombination and repair (2,472, 8.08%); posttranslational modification, protein turnover, chaperones (2,304, 7.53%) and signal transduction mechanisms (1,860, 6.08%). However, only 11 and 10 annotations of unigenes are annotated to the nuclear structural and the extracellular structure. The KEGG database was used to analyze the active biological pathways. The 14,708 assembled unigenes were assigned to 120 biological pathways through this process (Table S4). Among them, the highest metabolic pathway assigned to the unigenes is Ribosome (ko03010, 698 unigenes), followed by Oxidative phosphorylation (ko00190, 566 unigenes), Purine metabolism (ko00230, 563 unigenes), Protein processing in endoplasmic reticulum (ko04141, 526 unigenes), Glycolysis/Gluconeogenesis (ko00010, 491 unigenes), RNA transport (ko03013, 423 unigenes), Spliceosome (ko03040,414 unigenes) and Pyrimidine metabolism (ko00240, 391 unigenes). These results indicate that the growth and development of Acer truncatum seeds is mainly dependent on a large number of substances and energy metabolism.
EST-SSR discovery. As containing highly informative molecular markers, SSR markers have become one of the most widely used molecular marker systems for genetics, evolution and breeding research. To explore EST-SSR markers in the assembled unigenes, the 4,039 sequences containing 5,774 EST-SSRs were produced from 15,837 unigenes. Di-nucleotide and tri-nucleotide motifs were the most plentiful with 31.24% (909) and 29.35% (854), respectively (Table 4). The most repeat was AG/CT (1,012), followed by AGC/CTT (400), AT/AT (328), and ACC/GGT (189) (Table S5). The large set of EST-SSR markers identified in this research will help future researchers to better understand the genome-wide adaptive pattern of this species.

Candidate enzymes involved in VLCFAs elongation of fatty acids in *Acer truncatum* seeds. VLCFAs are synthesized by a microsomal fatty acid extension (FAE) system. Specifically, the C18 fatty acid is first synthesized using the de novo fatty acid synthesis pathway of plastids, and the C2 portion of malonyl-coenzyme A (CoA) is sequentially added to the previously synthesized C18 fatty acids. Analogous to FAS, the elongase complex catalyzes the elongation of fatty acids. Four VLCFAs elongase complex catalyzes: KCS, KCR, HCD, and ECR.

Based on previous studies, 34 assembled unigenes related to 4 of the enzymes in the VLCFAs FAE were identified in the annotated *Acer truncatum* seeds transcriptome database (Fig. 6A). In the transcriptome database, KCS is considered to be the rate-limiting enzyme in VLCFAs biosynthesis. Therefore, the isolation and functional analyses of the genes of KCS genes have become the most important thing in the study of VLCFAs biosynthesis. Twenty unigene sequences were annotated as encoding KCS genes.

In order to identify the key genes regulating the VLCFAs biosynthesis, DEGs analysis was performed through comparing the expression levels. On the basis of the applied thresholds FDR (False Discovery Rate) < 0.01 and log2 (foldchange) ≥ 2, a total of 3,258 unigenes were identified as DEGs between these two samples (H-11 vs. L-4), which comprised 2,131 up-regulated genes and 1,127 down-regulated unigenes (Fig. S2). Among the 3,258 DEGs, the expression profile of 4 differentially expressed genes in VLCFAs elongation of fatty acids such as KCS, KCR, HCD, and ECR were DEGs analysis and confirm the results by RT-qPCR (Fig. 6B). The higher content of VLCFAs genotype group (H-28, H-7, H-14, H-26) and the lower content of VLCFAs genotype group (L-1, L-18, L-19) in 36 genotypes were also DEGs analysis and confirm the results by RT-qPCR (Fig. S3).

The results showed that 16 DEGs analysis exhibited the similar trends with the RT-qPCR transcript levels, among them, 10 DEGs were up-regulated, including c132128 (KCS-like), c121604 (KCS-like), c124374 (KCS-like), c117613 (KCS-like), c110367 (KCR-like), c117613 (KCS-like), c126208 (KCS-like), c123713 (KCS-like), c110367 (KCR-like), c70062

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**Table 4.** Length distribution of EST-SSRs based on the number of repeat units.

| Number of repeat units | Di- | Tri- | Tetra- | Penta- | Hexa- | Total | Percentage |
|------------------------|-----|------|--------|--------|-------|-------|------------|
| 5                      |     | 856  | 41     | 8      | 4     | 909   | 31.24%     |
| 6                      | 454 | 383  | 11     | 2      | 4     | 854   | 29.35%     |
| 7                      | 297 | 195  | 11     | 1      | 0     | 494   | 16.98%     |
| 8                      | 224 | 20   | 1      | 1      | 0     | 245   | 8.42%      |
| 9                      | 200 | 0    | 0      | 0      | 0     | 200   | 6.87%      |
| 10                     | 189 | 10   | 0      | 0      | 0     | 199   | 6.84%      |
| >10                    | 7   | 1    | 1      | 0      | 0     | 9     | 0.31%      |
(KCR-like), c127044 (HCD-like), c86364 (HCD-like) and 6 DEGs showed no difference between H-11 and L-4, such as c119348 (HCD-like), c125484 (ECR-like), c87572 (ECR-like), c128798 (ECR-like), c132697 (ECR-like) and c131607 (ECR-like) (Fig. 6B). These results again confirm that KCS is the rate-limiting enzyme for the elongation of long-chain fatty acids. In addition, regulating the expression of the KCS gene affects the final contents of VLCFAs. However, since VLCFAs biosynthesis are complex processes involving multiple parameters, we only clarifies part of the whole process through the transcriptome sequencing analysis, so it is difficult to determine a precise conclusion. Obviously, additional accurate molecular biology, genomics and proteomic analysis procedures studies are required to verify and validate and further build on our predictions.

Conclusions and Perspectives
To our knowledge, this study provides the first comprehensive of the transcriptome of Acer truncatum seeds. The coverage of the transcriptome, which includes 37.07 Gbp, was comprehensive enough to identify the majority of the regulation VLCFAs biosynthesis genes. A saturation curve for RNA-Seq was provided in Fig. S4. A total of 127,791 different transcripts and 97,053 unigenes were identified in this study. Additionally, we categorized the enzymes involved in the biosynthesis of VLCFAs, such as KCS, KCR, HCD and ECR. Additionally, large numbers of EST-SSRs were determined. This transcriptome dataset have been made publicly available NCBI Short Read Archive (Accession Number: SUB3838977), we believe that it may provide new resource for future high-throughput gene expression of Acer truncatum seeds growth and development as well as its breeding, especially involved in VLCFAs biosynthesis.

Materials and Methods

Plant materials. Thirty-six genotypes of Acer truncatum trees (10-year-old) seeds, were randomly collected from the Fufeng commercial planting base in Boji Country (N34°22′36.62″; E107°53′44.37″; with altitude 550–600m), Shannxi Province, China on the early September, 2015. The Acer truncatum trees seeds were fully mature in October. The reason for choosing this period is as follows: Our team previously found that VLCFA (especially 24:1 nervonic acid) are converted and accumulated in the middle and late stages of fatty acid synthesis, which is the first of the Acer truncatum seeds rich in VLCFA nervonic acid3.
Fatty acids extraction and composition analysis. The seeds of *Acer truncatum*, with three biological replicates, were used for extraction of fatty acids. Seeds FA were extracted and analyzed as reported previously in detail. Briefly, total FA was converted to FA methyl ester at 80 °C in a methanol solution containing 1 M HCl for 2 hours. After extraction, the fatty acid composition of the oil was analyzed by gas chromatography-mass spectrometry (Q GC-MS, Thermo Fisher). The FAME peaks were identified using the NIST 2014 database and their retention times compared to real standards. Prior to data analysis and statistics, all FAME peaks were quantified by area normalization with a threshold set at 0.1%.

The RNA-seq library construction for sequencing. The seeds were collected from *Acer truncatum* plants and dissected. After removal of pericarp, they were then immediately frozen and stored in liquid nitrogen prior to further analysis. We extracted the total mRNA using the Plant-RNA Kit (Aidlab -biotech, China). The spectrophotometer and the Agilent 2100 bioanalyzer were used to measure the quality and quantity of purified mRNA. The mRNA-seq library was constructed using the Illumina’s TruSeq RNA Sample Preparation Kit and the library quality was assessed on the Agilent Bioanalyzer 2100 system as previously reported in detail. The fragment (340 bp ± 25 bp) was purified by gel electrophoresis and then amplified by PCR as a sequencing template. Finally, the mRNA library was sequenced by the HiSeq4000 platform (Illumina Inc., USA).

Illumina sequencing data assembly and analysis. To obtain high quality clean read data for de novo assembly, all adapter sequences and low quality sequences were removed from the raw data. Using the Trinity program (k-mer = 25), the high-quality reads were de novo assembled (http://trinityrnaseq.sourceforge.net/). The contigs were clustered and the transcripts were further assembled according to the paired reads. The longest transcript in the cluster were defined as unigenes. We used EMBOSST Getorf Software to predict the coding area (http://emboss.bioinformatics.nl/cgi-bin/emboss/getorf).

Sequence clustering and functional categorization of unigenes. The assembled unigenes sequences were annotated based on the following databases: NR (NCBI non-redundant protein sequences); NT (nonredundant nucleotide sequence); the Swiss-Prot (Protein sequence database); GO (Gene Ontology); COG (Clusters of Orthologous Groups of proteins) and KEGG (Kyoto Encyclopedia of Genes and Genomes). The best alignment was selected from the matches with an E-value < 10^-5. According to the best BLAST comparison (highest score), we give a gene name for each assembly sequence. The ORFs were identified by the “GetORF” (EMBOSS software package). GO were assigned to the assembled unigene using the "Blast2GO". The KEGG pathways annotation by the KAAS (KEGG Automatic Annotation Server) (http://www.genome.jp/kegg/kaas/).

The EST-SSRs detection. Using the MISA software, the 12,845 unigenes (more than 1 kb) of *Acer truncatum* were used for the EST-SSRs detection (http://pgrc.ipk-gatersleben.de/misa/). The parameters were adjusted to identify perfect di-, tri-, tetra-, penta- and hexa-nucleotide motifs with a minimum of 6, 5, 4, 3 and 2 repeats respectively as previously described.

Quantitative real-time reverse transcription PCR (RT-qPCR). The RT-qPCRs were used to examine expression of potential candidate genes in the VLCFAs biosynthetic pathway in *Acer truncatum* seeds, such as KCS, KCR, HCD, and ECR. The expression of these potential candidate genes were calculated by relative quantification with the *actin* housekeeping gene as a reference. The specific primers are listed in Table S6 for the RT-qPCR reaction for the candidate genes. The RT-qPCR reactions were performed in a Step One Plus Real-Time PCR System (Applied Biosystems, USA) using a Super Real PreMix kit (SYBR Green) (Tiangen-biotech, China). The RNA relative expression of each gene was calculated as reported previously in detail.

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Author Contributions
R.K.W. and P.L. performed the experiments, analyzed the data, prepared figures and tables, reviewed drafts of the paper. J.S.F. reviewed drafts of the paper. L.L.L. conceived and designed the experiments, contributed reagents/materials/analysis tools, wrote the paper, reviewed drafts of the paper.

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