Screening of essential oils activity against a gram negative psychrophilic bacterium isolated from aquatic environment (Water & Biofilm)

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Abstract. Among the applications of medicinal plants, it is their use as antimicrobial agents. The objective of this study was to investigate the effect of some essential oils against an etiological pathogen Flavobacterium spp. responsible for several lost in rainbow trout, (Oncorhynchus mykiss) hatcheries, the strains used in this study were isolated from rearing tanks water and biofilm, identified as Flavobacterium spp. based on phenotypic, biochemical and enzymatic characterizations. A collection of eight essential oils were extracted, analyzed and tested for an inhibitory activity against the isolated strains, the effect on this bacterium has been demonstrated by the aromatogram method based on a screening of bacterial growth in a solid medium culture with disks containing essential oils. Our study's results show that the chemical composition of the extracted essential oils play a crucial role in their antibacterial activity, which varies from 6 mm up to 34 mm as maximal inhibitory diameter.

1 Introduction

A bacterial isolate from rearing tank water and biofilm in a fish farm in Morocco, was characterized for its physiological, enzymatic and biochemical features as Flavobacterium spp, in comparison with seven strains of Flavobacterium psychrophilum.

In general, Flavobacteria are aerobic non-fermentative bacteria, catalase and oxidase positive, gram negative, yellow rods frequently isolated from different ecosystems, and it is considered to be a bacterial group of special relevance for aquatic environments (marine and freshwater). The importance of Flavobacterium spp. as a fish pathogen and the increasing significance of the disease are given by the huge number of economic lost known in the salmonid hatcheries world wild, especially in early ages. Currently there is no vaccine and the only treatment existing is the antibiotic therapy. Antibiotics become more and more complex and chemically synthesized but despite that the new generations may not be able to inhibit and/or control the development, and growth of pathogenic microorganisms, due to their capacity of transmitting genetic material and acquiring new resistances against the active molecules of these antibiotics.

Nowadays, antibiotics residues are reported to be ineffective in controlling diseases in aquaculture due to the misuse or overuse of antibiotics by fish farmers. Furthermore, antibiotics residues are found as a threat to human health [1] and the environment [2].

This situation leads to the recurring public health problems related to bacterial resistance to antibiotics, this leads to think about new natural treatments which means a return to the traditional medicine, and more specifically the herbal medicine, on the basis of essential oils extracted from aromatic and medicinal plants; theses essential oils are a part of secondary metabolites, which the plant produces in some conditions for the majority of cases to protect themselves against pests.

These essential oils have several uses, they can be used in medicine, and power supplies are for aromatization or conservation, also in cosmetic [3]. The antimicrobial properties of essential oils have been described [4], as natural products from the aromatic and medicinal plants, they are known by their antiviral, antibacterial, antifungal and antioxidant power [5-6]. Currently there is interest in herbal medicine as an alternative of synthesized antibiotics [7].

Many studies have been made to reveal the antibacterial power of essential oils either on the bacteria Gram-positive or negative, which justifies the research that been made in traditional medicine for the characterization of this power, the more it is admitted that the essential oils can be active even on the multi resistant pathogenic microorganisms [8]. The plants and their components are sources potentially rich in antimicrobial substances. Many studies have been published confirming the presence of this activity against different types of microorganisms, [9-10-11].

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2 Material and Methods

2.1 Biological material

Water samples were collected from the rearing tanks in the fish farm, sampled during the four seasons in 2015. Water samples were collected from each tank using acid-rinsed and autoclaved polycarbonate bottles. Water samples were kept dark at 4°C before analysis. Biofilm samples were taken in the same time and in aseptic conditions, from the same fish rearing tanks and transported in sterile sampling pots under controlled conditions.

Processed samples were plated onto TYES agar. Plates were incubated at 16°C for 7 to 10 d and bacterial growth was recorded. Yellow pigmented flat or very thin colonies, spreading, with uneven, rhizoid, or filamentous margins were selected and sub-cultured for phenotypic, enzymatic and biochemical analysis.

To be identified as Flavobacterium spp, the isolates were tested for a number of key characteristics using standard procedures [12], such as cell size and morphology (phase-contrast microscopy), Gram stain, congo red absorption, KOH string test, cytochrome oxidase and catalase (3% H2O2). Furthermore, production of indole, growth on enriched medium with casein, tyrosine, starch, gelatin, chitinase, cellulase and pectinase activities were tested as described by [13]. The strains were also characterized by using the whole test spectrum of the identification systems API 20E and API 20NE at 16°C. The gleeding motility was tested using the hanging drop method.

2.2 Vegetal material

The essential oils (EO) are not simple bodies, but in general, they are assemblies of molecules each once have her specific properties. The importance of the knowledge of families, botanical genera and species is obvious, and also their provenance. The plants witch they are botanically identic can, indeed, give some essences whose the differences are more or less important.

The true lavender (lavandula vera or angustifolia or officinalis) is part of the Labiatae. It grows in Mediterranean Europe, on limestone land, 700 m up to 1 800 m of altitude. Its leaves are long, narrow and whitish; the whole plant has aromatic smell very pleasant [14].

Sage (Salvia officinalis, Labiatae) was seen among our ancestors (the Gauls), as well as among all the other peoples of Antiquity, as the plant salutary by excellence, so wonderful that they considered all able to cure all diseases [15].

The wild chamomile, Cladanthus mixtus, also called Chamomile commercially of Morocco, is an Asteraceae biennial chamephyte to numerous erect stems, this species is characterized by a fresh smell balsamic. It gives an essential oil of campfre smell sought in cosmetic, perfumery and Medicine [16].

Cloves (which is a male name, contrary to the current use) or gerofle or clove, is the floral button of a tree originating in the Moluccas Islands, which are the homeland of giroflier (Eugenia caryophyllata, or caryophyllus, Myrtaceae) [17].

The geranium the Pelargonium graveolens (family Geraniaceae), belongs to the category of perennial plants to fragrant foliage. Evergreen leaves are lobed and opposite, covered with glandular hairs microscopic organisms that release their scents to the touch or to the heat [18]. The plant is cultivated in many Mediterranean regions and subtropics [19].

Rosemary, rosmarinus officinalis Labiaceae, common plant in the wild, is, without doubt, one of the plants the most popular in Morocco This plant belongs to the family of Labiatae. It presents itself in the form of shrub, under shrub or herbaceous. The leaves are narrowly lanceolate linear, brittle and tough, the flowers of a pale blue, stained inwardly from the purple are arranged in short dense clusters thrive almost throughout the year [20].

The eucalyptus (Eucalyptus globulus - Myrtaceae) is originating in Australia and Tasmania and it is cultivated in some subtropical regions of the South of Europe, Africa, Asia, and America. The oil of Eucalyptus is obtained from the leaves of the plant [21-22].

The hydrodistillation of the wood of cedars (Cedrus atlantica), a bio-test performed with the essential oil gross and its fractions showed an antimicrobial activity observed predominantly in the fractions rich in terpenols. [23].

The harvested plants were manipulated carefully and all the essential oils tested have been extracted by steam drive of water using an experimental setup within « Les Aromes du Maroc » Domain -Tiddas, except the essential oil of Lavandula angustifolia that comes from a traditional distillery in Ben Smin-Azrou; and the essential oil of the Cedrus atlantica which comes from the society « Cèdre de l'Atlas-Azrou ». The chemical analysis of essential oils has been carried out by CPG-MS, in the laboratory of physico-chemical analysis in « Les Aromes du Maroc » Domain -Tiddas.

2.3 Aromatogram

The Purpose of the Aromatogram is to determinate (in vitro) which Essential Oils are to be considered as active or inactive against a bacterium. The choice of using certain Essential Oils will be determined by various criteria gathered from the Aromatogram - same as in the case of an Antibiogram.

The designation of an Essential Oil by the Aromatogram is an indication of its antibacterial properties in vitro against a considered germ. So it is a method which allows determining the inhibitory activity of essential oils, if it exists by the measurement of the diameter of inhibition around a disk of cellulose impregnated with essential oil.

It is a qualitative method which makes it possible to study on a solid culture medium the action of an agent antibacterial, and to give indications on its effectiveness.
in vitro. The antibacterial activity of essential oils was determined separately, using the disc diffusion method [24].

A suspension of each strain is prepared in physiological water and adjusted to 10^8 bacteria/ml. Each suspension (100 μl) is spread on a Petri dish of 90 mm diameter. The surface of the Petri dishes is dried under the hood laminar flow with slightly opened cover of the Petri dishes.

The sterile paper disks blotting of 6 mm in diameter are impregnated with essential oil and then tests were made three times. The reading of the inhibition diameters was done after 7 to 10 days of incubation in the oven at 15 °C. The results are expressed according to three levels of activity: resistant (d < 6 mm), intermediate (13 mm < d > 6 mm) and sensitive (D > 13 mm).

### 3 Results and Discussion

The identification of bacteria is respecting the classical protocols, the morphological and phenotypic characteristics of all the bacterial strains examined were in agreement with those described in previous published reports, and they are yellow pigmented colonies, gram negative bacillus and adopting a movement by gliding [25]. However, our strains present a homogeneity enzymatic pattern.

Despite, the biochemical analysis show that the studied strains are all Congo red absorbent, CIT positive, Indole negative, they grow up in a medium supplemented with 1% of NaCl and they can’t degrade glucans. But they respond differently on some tests; the found results are illustrated in the table (1).

**Table 1. Biochemical characterization of the studied strains**

| Bacterial strains | Biochemical tests | ONPG | ADH | LDC | ODC | H2S | URE | F.P.T |
|-------------------|-------------------|------|-----|-----|-----|-----|-----|-------|
| 404               |                   | N    | N   | N   | N   | N   | N   | P     |
| 405               |                   | P    | N   | N   | N   | N   | N   | N     |
| 406               |                   | P    | N   | P   | N   | N   | N   | N     |
| 409               |                   | N    | N   | N   | N   | N   | N   | N     |
| 901               |                   | N    | P   | N   | N   | N   | N   | N     |
| 903               |                   | N    | N   | N   | N   | N   | N   | P     |
| 907               |                   | N    | N   | N   | N   | N   | N   | N     |
| 911               |                   | N    | P   | N   | N   | N   | N   | N     |
| 111               |                   | N    | N   | N   | N   | N   | N   | N     |
| 114               |                   | N    | N   | P   | N   | N   | N   | N     |
| 117               |                   | N    | N   | N   | N   | N   | N   | N     |
| 118               |                   | N    | N   | N   | N   | N   | N   | P     |
| 442               |                   | N    | N   | N   | N   | P   | N   | N     |
| 445               |                   | P    | N   | P   | N   | N   | P   | N     |
| 817               |                   | P    | P   | P   | N   | P   | N   | N     |
| 733               |                   | N    | N   | N   | N   | N   | N   | N     |
| 738               |                   | P    | P   | P   | N   | N   | N   | N     |
| 607               |                   | N    | N   | N   | P   | N   | P   | N     |
| 760               |                   | N    | N   | N   | P   | N   | P   | N     |

N: negative result / P: positive result

### Table 2. Majority compounds of essential oils used in the study

| Essential oil | Majority compounds | Color | Density |
|---------------|--------------------|-------|---------|
| Lavender      | Cineol 1-8: 38.22% Linalol: 15.93% Camphor: 8.57% Linalyl acetate: 6.53% Borneol: 3.59% Pinene alpha: 1% Pinene beta: 1.70% | Pale yellow | 0.933 |
| Sage          | Pinene alpha: 7.61% Camphene: 4.89% Pinene beta: 4.06% Myrcene: 2.58% Limonene cineol 1-8: 19.83% Thujone alpha: 2.75% Thujone beta: 25.51% Camphor: 10.66% Humulene alpha: 4.26% | Orange-yellow | 0.779 |
| Chamomile     | Pinene alpha: 15.91% Pinocarveol trans: 2.31% Pinocarvone: 1.75% Pinene beta: 3.88% Limonene: 8.10% Elemene Delta: 3.05% Cinéol 1-8: 4.77% Caryophyllene beta: 1.07% Santolina alcohol: 7.56% Farnesene beta: 5.87% Germacrene d: 7.22% Nerolidol trans: 0.94% | Pale yellow | 0.952 |
| Clove         | Eugenol: 85.42% Caraphyllene beta: 9.95% | Pale yellow | 1.040 |
| Geranium      | Isomenthone: 14.45% Geraniol: 14.45% Linalol: 8.83% Citronellyl formate: 5.77% Geranyl formate: 5.23% Cinetroleol: 19.69% Butyrate Geranyl: 2.39% 10-epi-gamma-eudesmol: 9.46% | Pale yellow | 0.873 |
| Rosemary      | Pinene alpha: 11.95% Camphene: 3.25% Pinene beta: 6.63% Linalol: 8.75% Myrcene: 1.63% Térpénin-4-OL: 2.18% Limonene: 2.15% Caryophyllene beta: 2.34% Cinéol 1-8: 46.12% Borneol: 2.88% Terpinene gamma: 1.30% Verbenone: 2.15% | Colorless | 0.804 |
| Eucalyptus    | Cineol 1-8: 58% Alpha pinene 21, 97% Limonene: 7.95% Paracyrenme: 2.89% Globulo: 1.5% Trans pinocarveol: 2.63% Aomadendrene: 1.74% | Colorless | 0.781 |
| Cedar         | Himachaline alpha: 17.35% Himachaline beta: 46.97% Himachaline gamma: 12.39% Trans Atlantone alpha: 3.12% Cis Atlantone alpha: 1.46% Atlantone beta: 1.13% | Pale yellow-colorless | 0.968 |

The physico-chemical composition of the essential oils is presented in the table (2). To avoid overload in the text, only the compound having content greater than 1% will be mentioned. The identification is carried out through the comparison of retention times observed on the one hand on samples to characterize, and on the other hand on the heels available.
The density tells us about the chemical composition, so a density less than 0.9 indicates the presence of terpene and aliphatic compounds at high rates, while a density greater than 1 indicates a very varied composition of polycyclic terpene compounds [26].

However, the pharmacological properties of essential oils from a few Labiatae, have been demonstrated since 1973 by [27], but in 1974, [28], has described the antibacterial properties of the other essential oils, such as that obtained from the clove and which does not contain phenols but aromatic aldehydes.

The aromatogram realized with strains of Flavobacterium spp. shows that the strains are sensitive to the essential oils of eucalyptus, geranium, lavender, rosemary and the clove, with inhibition diameters up to 35 mm and they show a resistance to essential oils of cedar, chamomile and sage (Table 3).

**Table 3. Inhibition diameters of essential oils against Flavobacterium spp. Strains**

| Strains      | R. officinalis (10 µl) | S. officinalis (10 µl) | C. mixtus (10 µl) | L. angustifolia (10 µl) | C. alatunica (10 µl) | E. globulus (10 µl) | P. graveolens (10 µl) | S. aromaticum (10 µl) |
|--------------|------------------------|------------------------|-------------------|------------------------|---------------------|---------------------|----------------------|-----------------------|
| 404          | 0.8                    | 0.8                    | 0.8               | 2.0                    | 2.8                 | 2.0                 | 1.2                  |                       |
| 405          | -                      | -                      | 3.0               | 1.7                    | 0.8                 | -                   | 1.8                  |                       |
| 406          | -                      | -                      | 0.8               | 3.2                    | -                   | -                   | 1.3                  | -                     |
| 409          | -                      | -                      | -                 | -                      | 1.0                 | 1.0                 | 1.0                  |                       |
| 901          | -                      | -                      | -                 | 1.0                    | -                   | -                   | 1.8                  |                       |
| 903          | 0.8                    | -                      | 1.0               | 1.8                    | 2.8                 | 3.0                 | 0.8                  |                       |
| 907          | -                      | -                      | -                 | -                      | -                   | -                   | 1.0                  |                       |
| 911          | -                      | -                      | -                 | 1.1                    | 1.4                 | -                   | -                    |                       |
| 111          | -                      | -                      | -                 | -                      | -                   | -                   | -                    |                       |
| 114          | -                      | -                      | 2.8               | 1.2                    | -                   | 1.2                 | 2.0                  |                       |
| 117          | 0.8                    | 0.8                    | 1.0               | 3.4                    | 1.6                 | 3.5                 | 3.0                  | 0.8                   |
| 118          | 0.8                    | 0.8                    | 1.0               | 3.4                    | 1.6                 | 3.5                 | 3.0                  | 1.4                   |
| 442          | -                      | -                      | -                 | -                      | -                   | 3.0                 | 1.4                  | 1.8                   |
| 445          | -                      | -                      | -                 | 1.1                    | -                   | 2.6                 | -                    | 1.6                   |
| 817          | -                      | -                      | -                 | -                      | -                   | 1.5                 | -                    | -                     |
| 733          | -                      | -                      | 1.0               | -                      | -                   | 1.6                 | -                    | -                     |
| 738          | -                      | -                      | -                 | -                      | -                   | 3.2                 | -                    | -                     |
| 607          | -                      | -                      | 0.8               | -                      | -                   | 2.0                 | -                    | -                     |
| 760          | -                      | -                      | -                 | -                      | -                   | 2.4                 | -                    | -                     |

_E. globulus_ and _L. officinalis_ come in first position with highest antibacterial activity and inhibition diameters varying between 6 and 30.5 mm and 6 and 30.4 mm respectively.

As it has been demonstrated by [27], the essential oil of _E. globulus_ has a powerful antibacterial activity which is in accordance with our finding. As for the tested _L. officinalis_, the antimicrobial activity of this essential oil is mainly due to its richness in the following constituents: Cineol 1-8, camphor, borneol and esters. Indeed, all these compounds are known for their antimicrobial properties, this goes with the results demonstrated in [14].

As for _P. graveolens_ and _S. aromaticum_, their essential oils exhibit less activity and inhibition diameters varying between 6 and 30 mm and 6 and 20 mm respectively.

For the activity of _P. graveolens_ essential oil it has been approved by [28], and they propose the use of this natural substance in the pharmaceutical industry for the fight against contamination and biofilms composition. The _S. aromaticum_ activity is due principally to eugenol, its major compound which is known by its antibacterial, antifungal and antiviral activities [17].

The essential oils of _C. atlantica_ and _C. mixtus_ exhibit low activity with inhibition diameters ranging from 6 to 20 mm and 6 to 11 mm respectively.

Our essential oil of _C. mixtus_, have a weaken antibacterial activity compared with the results found by [21].

With inhibition diameters not exceeding 8 mm, the essential oils of _R. officinalis_ and _S. officinalis_ do not exhibit anti-flavobacteric activity. This agrees with the results of [29] who reported that the essential oil of _R. officinalis_ from Sardinia has moderate antibacterial activity.

From the found results it can be seen that the reference strains were present more resistance to the oils tested in comparison with our Moroccan isolates, only the essential oil of _E. globulus_ showed a strong activity translated by the diameters of inhibition measured.

Based on the disc diffusion method, this test makes it possible to assess the antibacterial activity of essential oils from the inhibition diameters they generate on a standardized bacterial inoculum. Among the essential oils screened, those with the highest activity were selected to study their mode of action on Flavobacterium spp.

These in vitro experiments have demonstrated effective antimicrobial efficacy of the essential oils of _P. graveolens, L. officinalis, E. globulus_ and _S. aromaticum_, compared to those of _C. atlantica_ and _C. mixtus_ except for the essential oil of _S. officinalis_ and _R. officinalis_ which seem to be ineffective on Flavobacterium spp.

**4 Conclusion**

These results show the importance of making the aromatogram in functions of the targeted germs, as a preliminary step in the selection of essential oils with antibacterial effect and those who do not possess, before starting the second part of the tests which concerns the determination of the CMI (minimum inhibitory concentration), and CMB (The minimal bactericidal concentration) if the power of essential oils is bactericidal.

These first results are encouraging new systematic studies of many essential oils on other samples of pathogenic bacteria with increased resistance against the
conventional antibiotics. The use of natural products in the place of some synthesis products is also envisaged in the field of animal health because the concept of bio and naturalness is in full progress; we can well imagine the use of these E.O.

New perspectives can be envisaged by a further study of the antibacterial activity, not only on the E.O used alone, but also in a mixture, thus allowing a possible synergy.

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