Human Amniotic Membrane Effect on Perianal Fistula Healing in Rabbits: An Experimental Study

Mohammad Rafati1, MD; Seyed Vahid Hosseini2, MD; Farid Moradian3, MD; Mozhe Zaman1, PhD; Hajar Khazraei1, PhD; Maral Mokhtari3, MD

1Colorectal Research Center, Shiraz University of Medical Sciences, Shiraz, Iran; 2Department of Surgery, Yasouj University of Medical Sciences, Yasouj, Iran; 3Department of Pathology, Shiraz University of Medical Sciences, Shiraz, Iran

Correspondence:
Seyed Vahid Hosseini, MD; Colorectal Research Center, Shiraz University of Medical Sciences, Shiraz, Iran
Tel: +98 71 32306972
Email: colorectal2@sums.ac.ir hoseiniv@sums.ac.ir
Received: 15 January 2017
Revised: 5 April 2017
Accepted: 16 April 2017

Abstract

Background: Perianal fistula is a complicated disorder and most difficult to manage. New treatment methods would help surgeons to achieve a better outcome in patients with perianal fistula. Human amniotic membrane (HAM) has positive effects on wound healing in several conditions. The present study aimed to further determine the effect of HAM on wound healing of perianal fistula in rabbits.

Methods: In a prospective experimental study, 14 male rabbits (aged 4-6 months and weighing 3-4 kg) were randomly divided into two groups. After 12 weeks, the high type perianal fistula was repaired with endorectal flap (ERF) and ERF plus HAM in the control and case groups, respectively. In all rabbits of the case group, a 1×1 cm width wrap of HAM was applied and fixed around the ERF site. Three weeks later, the repaired site of the perianal fistula was sent for pathologic wound healing scoring. The results were analyzed with the SPSS 21.0 software using Mann-Whitney test.

Results: Six rabbits of each group survived the study period. There was a statistically significant difference in wound healing between the case and control groups (P<0.001). Wound healing process in the case group occurred better and faster than the control group.

Conclusion: HAM has an effective role in enhancing the ERF procedure and considered appropriate. A combination of HAM with other methods is recommended.

Please cite this article as: Rafati M, Hosseini SV, Moradian F, Zamani M, Khazraei H, Mokhtari M. Human Amniotic Membrane Effect on Perianal Fistula Healing in Rabbits: An Experimental Study. Iran J Med Sci. 2018;43(5):473-478.

Keywords ● Rectal fistula ● Amniotic ● Surgical flaps

Introduction

An anorectal fistula is one of the inflammatory disorders that involves epithelium of the anal canal and perineum or perianal skin. Perianal fistula has significant implications on patients’ quality of life and its squeal range from minor pain and social hygienic discomfiture to frank sepsis. Anal canal has 10^{12} to 10^{14} bacteria/mg faeces and the incidence of an anal fistula that originate from an anal abscess is usually 26-38% and the mean age of onset is 40 years old (range 20-60). Anorectal abscess is the most common cause of an anorectal fistula. Other causes of anorectal fistulas include Crohn’s disease, lymphogranuloma venereum, radiation proctitis, and rectal foreign bodies.

The management of anal fistula is an important challenge in colorectal surgery. The surgery is based on therapy with the final
goal of draining the local infection, eradicating
the fistulous tract, and avoiding recurrence while
preserving native sphincter function.6,7 Biologic
dressing to enhance wound healing is usually used.
Human amniotic membrane (HAM) includes three
elements, namely an epithelial layer, a basement
membrane, and a stroma which makes the inner
layer of the placenta. HAM has key features,
including anti-inflammatory, antimicrobial,
antifibrosis, antiscarring, low immunogenicity,
and high potency of differentiation. HAM as a
biologic matter has been used for wound healing
of abdominal adhesiolysis, tenolysis, neurolysis,
and injuries of the dura-mater and vagina for
many years.8,9

The precise mechanism of HAM remains
unidentified. Various mechanisms of anti-
inflammatory effect of HAM such as decreasing
lipid peroxidation, inducing apoptosis,
and inhibiting the chemotactic activity of
polymorphonuclear neutrophil (PMN) have
been proposed.10 HAM has been investigated
and demonstrated significant effects on wound
healing of rectovaginal fistula, gastrointestinal
tract, colon, and duodenum.10,11 Endorectal flaps
are used to protect wound healing with 70-80%
success rate. In the present study, for the first
time, we aimed to examine the effect of HAM on
wound healing process in perianal fistula.

Materials and Methods

Animals and Ethical Considerations

In collaboration with the Animal Laboratory of
Shiraz University of Medical Sciences (Shiraz,
Iran), 14 rabbits were selected and initially
evaluated by a veterinarian surgeon for any
underlying problems. The age of the rabbits
was 4-6 months, weighing 3-4 kg, and kept in
standard cages at 20-24 °C, 55±5% humidity,
12 times per hour ventilation, and 12:12 hours
light dark cycle. The rabbits received standard
chow and water ad libitum. The Animal Laboratory
of Shiraz University of Medical Sciences
supervised the procedures, preoperative, and
postoperative cares.12 The HAM was provided
by Shiraz Burn Research Center, Shiraz Zeinab
Hospital (Shiraz, Iran) and primarily evaluated
for viral markers (HIV, HCV, and HBV). It was
then preserved in gluteraldehyde and frozen at
20 °C. The study was approved by the Research
Vice Chancellor and the Ethics Committee of
Shiraz University of Medical Sciences (number
7172), Shiraz, Iran.

Experimental Design

Step 1: The rabbits were randomly divided
into two groups, namely case group (n=7) and
control group (n=7). Following 10-20 minutes
intravenous injection of Ketamine-Xylazine
(50 mg/kg), the animals underwent surgical
high type fistula tract creation by clamp and
electrocutery. A short French tube (number 10)
was inserted into the fistula and the internal
and external orifice of the fistula tract was
marsupialized with Vicryl 4-0.

Step 2: Twelve weeks later, after intravenous
sedation by Ketamine-Xylazine (50 mg/kg
administered 10-20 minutes in advance) followed
by preparation and draping, ERF was made in the
control rabbits using prolene 4.0 and ERF plus
HAM was applied in the case group. A 1×1cm
width wrap of HAM was sutured around the ERF
site and fixed by Vicryl 4-0 with simple interrupted
sutures. Surgical diet was started after 24 hours
in both groups. The protocol of anesthesia and all
procedures, preoperative care, and postoperative
care were similar for all the rabbits.

Step 3 (follow-up analysis of anastomosis):
The surviving rabbits were sacrificed after
3 weeks and perianal site surgery specimens
were resected and fixed in formaldehyde 10% solution (formalin). All samples were labeled
blindly and sent to the Histopathology Department
of Faghihi Teaching Hospital, Shiraz, Iran.

Pathological Evaluation

Histopathological assessment was performed
after tissue processing and slide preparation
with hematoxylin and eosin staining by a single
pathologist in the department of histopathology.
A modified scoring system for surgical wound
healing13 was used to determine healing grade in
each sample. Scoring was performed according
to multiple factors such as epithelialization,
inflammation, neovascularization, necrosis, and
granulation tissue formation (table 1).

Statistical Analysis

The statistical SPSS software version 21.0
for Windows (SPSS, Chicago, IL) was used
to perform the analyses with Mann-Whitney U
test. Two-tailed P values less than 0.05 were
considered statistically significant.

Results

Macroscopic Evaluation

After 12 weeks, no surgical site infection was
found in all rabbits (case and control groups).
Internal orifice perianal fistula sites were
resected and prepared for pathologic evaluation.

Pathological Evaluation

Pathological evaluation revealed no
significant difference between the case and
Adjunctive effect of HAM on perianal fistula healing

Iran J Med Sci September 2018; Vol 43 No 5

control groups, possibly due to the small sample size. The mean scores for each histological item are presented in table 2.

The pathological evaluation revealed more collagenization, neovascularization, and epithelialization as well as less tissue necrosis, inflammation, and granulation tissue formation in the case group compared with the control group. As shown in figure 1, clinical wound healing was more rapid in the HAM group.

Table 1: Modified scoring system for wound healing

| Score | Epithelialization | Collagenization | Inflammation | Neovascularization | Necrosis | Granulation |
|-------|-------------------|-----------------|--------------|--------------------|----------|-------------|
| 1     | None              | None            | Severe       | None               | Extensive| None        |
| 2     | None              | None            | Moderate     | None               | Focal    | Immature    |
| 3     | Partial           | Partial         | Mild         | <5/HPF             | None     | Mild mature |
| 4     | Complete, immature| Complete, non-regular| None | 6-10/HPF           | None     | Moderately mature |
| 5     | Complete, mature  | Complete, regular| None         | >10/HPF            | None     | Fully mature |

*HPF: High power field

Table 2: The mean of wound healing score in the two groups

| Groups       | Case   | Control | P value |
|--------------|--------|---------|---------|
| Epithelialization | 3.57   | 3.00    | 0.35    |
| Collagenization  | 4.14   | 3.71    | 0.20    |
| Inflammation     | 1.86   | 2.28    | 0.09    |
| Neovascularization | 1.86   | 1.57    | 0.42    |
| Necrosis        | 0.28   | 0.71    | 0.24    |
| Granulation     | 3.28   | 3.71    | 0.41    |

Discussion

The present experimental study was performed as the first investigation to assay the effects of HAM on wound healing and ERF in an animal model. Better histopathological results were found in the HAM group. There was no incident of wound infection, one of the most serious complications, in the control and case groups. One rabbit of each group expired during the study. In the HAM group, no evidence of ERF rejection due to infection was detected.

Various treatment approaches are primary fistulotomy, fibrin sealant injection, draining setons to preserve the sphincter mechanism, endoanal advancement flaps, fibrin glue, fistula plugs, modified Hanley procedure, ligation of the intersphincteric fistula tract, or diversion. In a study by Uludag et al., HAM had a preventive effect on the colonic anastomosis leakage. In the present study, pathological evaluation revealed more collagenization, neovascularization, epithelialization, and less tissue necrosis and granulation tissue formation in the case group compared with the control group. A range of studies assessed the effect of HAM on wound healing. In a study by Ghahramani et al., dogs were a candidate for duodenorraphy and HAM patch was used for wound repair. In gross evaluation with/without HAM, no difference in duodenal diameter was reported after the operation. However, in pathological evaluations, they revealed less inflammation and a better wound healing with HAM. In another study, HAM as a bioprosthesis was used with TachoSil® for the repair of rectovaginal fistula in a rabbit model. After 4 weeks, the histopathological results showed differences in fistula healing between the rabbits. The most rectovaginal fistula repair was related to HAM and TachoSil® did not have additional healing effect (P>0.05).

In an animal study, Barlas et al. evaluated the effectiveness of HAM as an intestinal patch and neomucosal growth was mentioned. They reported that after two weeks, the neomucosa consisted of a thin layer of columnar epithelial cells that covered all the patches. In an experimental study on dogs, Najibpour et al. indicated that the use of HAM results in a better colonic anastomosis and wound healing outcome. The left colon of dogs was resected and end-to-end anastomosis was performed in a single layer. HAM patch was used around the anastomotic line and the result showed significantly higher healing score in the HAM group (P=0.01).

Roshanravan et al. mentioned better outcomes when HAM was used as a bioprosthesis to repair the rectovaginal fistula in dogs. After 6 weeks, similar to previous and current investigations, there was no difference in...
the healing of fistula with or without HAM as a gross evaluation. However, the histopathological evaluation revealed significantly higher healing score in the HAM group (P=0.029).

A study on rats revealed that the wound healing process was accelerated due to the HAM effect on duodenal wall structure. The rats were subjected to duodenotomy and HAM was used on duodenal wound in one group. No changes were observed regarding the formation of duodenal fistula or peritoneal adherences in the region of duodenal wall repair. Schmidt et al. reported that regeneration of the mucosa and smooth muscle layer was increased and the healing of duodenal wall with HAM was better.20

Kuriu et al. demonstrated that HAM had a curing effect on the intraperitoneal adhesion in rats. They observed severe adhesions between the cecum and surrounding organs after 1 week of surgical trauma without HAM, while it was significantly reduced by HAM treatment. Histological evaluation showed a layered structure that consisted of myofibroblasts attached to the surfaces of HAM grafts. HAM was well absorbed after 10 weeks and it served as a substrate for regenerating mesothelium.

Some studies showed that the use of HAM might play a significant role and a feasible technical approach to prevent post-irradiation colorectal anastomosis leakage. In a study, animals underwent resection of colorectal segment and end-to-end anastomosis four weeks after irradiation. Then, HAM was applied around the site of anastomosis. A segment of anastomotic sites was obtained 8 weeks later for pathological evaluation and significant epithelialization and neovascularization with HAM demonstrated. The healing score was higher with HAM (P<0.001). Moslemi et al. suggested that the use of HAM could be protective from anastomosis leakage in post irradiation.

In another animal study, from a total of 10 dogs, 5 received 10 Gy of external radiation preoperatively while the other 5 received no radiation. Then, end-to-end anastomosis was done on the bowel remnant and HAM was used around the anastomosis. Tahamtan et al. demonstrated the ulceration of the mucosa with infiltration and granulation tissue formation. They showed that fibrotic tissue was formed between the serosa and HAM graft and suggested that complications of radiotherapy and loop ileostomy could be resolved with the application of HAM and eliminate the need for ileostomy insertion.23

Uludag et al. showed that HAM reduced inflammation and increase wound healing process. Additionally, inflammatory cytokines suppressed, tissue growth factor and anti-inflammatory proteins expressed, and fibroblast activity and angiogenesis increased. In the present study, granulation tissue formation was less in the case than the control group. The healing process in rabbits, due to their different immune system, is faster than humans. After 12 weeks, only the marked inner orifice was found and ERF was performed. The external orifice was healed in most rabbits.

The limitation of the present study is related to the small size of the animals. It is recommended to apply this method to large animals and even human beings. It should be noted that risk factors such as diabetes mellitus, obesity, anemia, and smoking could affect the results of human perianal fistula. Evaluation of these risk factors is recommended in future research.

**Conclusion**

In the present study, the effect of HAM on the ERF procedure was investigated on rabbits. The pathological evaluation revealed no statistically significant difference between the case and control groups. However, HAM placement had an effective role in enhancing the ERF procedure and showed a better histological findings and surgical outcomes in the HAM group.

**Acknowledgment**

The authors would like to thank the Vice Chancellor of Shiraz University of Medical Sciences for supporting the research (grant number: 7172)

**Conflict of Interest:** None declared.

**References**

1. Williams JG, Farrands PA, Williams AB, Taylor BA, Lunniss PJ, Sagar PM, et al. The treatment of anal fistula: ACPGBI position statement. Colorectal Dis. 2007;9:18-50. doi: 10.1111/j.1463-1318.2007.01372.x. PubMed PMID: 17880382.
2. Gurer A, Ozlem N, Gokakin AK, Ozdogan M, Kulacoglu H, Aydin R. A novel material in seton treatment of fistula-in-ano. Am J Surg. 2007;193:794-6. doi: 10.1016/j.amjsurg.2006.06.048. PubMed PMID: 17512300.
3. Hasan RM. Incidence of fistula after management of perianal abscess. Journal of Coloproctology. 2016;36:216-9. doi: 10.1016/j.jcol.2016.05.002.
4. Niyogi A, Agarwal T, Broadhurst J, Abel RM. Management of perianal abscess and fistula-in-ano in children. Eur J Pediatr Surg. 2010;20:35-9. doi: 10.1055/s-0029-1241878. PubMed PMID: 19899037.

5. Safar B, Sands D. Perianal Crohn’s disease. Clin Colon Rectal Surg. 2007;20:282-93. doi: 10.1055/s-2007-991027. PubMed PMID: 20011424; PubMed Central PMCID: PMCPMC2780223.

6. Goldberg JE, Steele SR. Rectal foreign bodies. Surg Clin North Am. 2010;90:173-84. doi: 10.1016/j.suc.2009.10.004. PubMed PMID: 20109641.

7. Kurer MA, Davey C, Khan S, Chintapatla S. Colorectal foreign bodies: a systematic review. Colorectal Dis. 2010;12:851-61. doi: 10.1111/j.1463-1318.2009.02109.x. PubMed PMID: 19895597.

8. Uludag M, Citgez B, Ozkaya O, Yetkin G, Ozcan O, Polat N, et al. Effects of amniotic membrane on the healing of primary colonic anastomoses in the cecal ligation and puncture model of secondary peritonitis in rats. Int J Colorectal Dis. 2009;24:559-67. doi: 10.1007/s00384-009-0645-y. PubMed PMID: 19172282.

9. Ghahramani L, Jahromi AB, Dehghani MR, Ashraf MJ, Rahimikazerouni S, Rezaianzadeh A, et al. Evaluation of repair in duodenal perforation with human amniotic membrane: An animal model (dog). Adv Biomed Res. 2014;3:113. doi: 10.4103/2277-9175.131029. PubMed PMID: 24804187; PubMed Central PMCID: PMCPMC4009825.

10. Niknejad H, Peirovi H, Jorjani M, Ahmadian A, Ghanavi J, Seifalian AM. Properties of the amniotic membrane for potential use in tissue engineering. Eur Cell Mater. 2008;15:88-99. PubMed PMID: 18446690.

11. Hosseini SV, Aski MH, Al-Hurry AMAH, Hassan A-RK, Khazraei H, Zabangirfard Z, et al. Simultaneous application of human amniotic membrane and TachoSil® in the repair of recto-vaginal fistula in an animal model. Comp Clin Path. 2017;26:405-9. doi: 10.1007/s00058-016-2391-1.

12. Consensus Author Guidelines for Animal Use [Internet]. USA: International Association of Veterinary Editors. c2010. Available from: http://www.veteditors.org/consensus-author-guidelines-on-animal-ethics-and-welfare-for-editors

13. Zekavat O, Amanat A, Karami M, Paydar S, Gramizadeh B, Zareian-Jahromi M. Wound Healing Studies Using Punica granatum Peel: An Animal Experimental Study. Adv Skin Wound Care. 2016;29:217-25. doi:10.1097/01.ASW.0000481116.16998.55. PubMed PMID: 27089150.

14. Rizzo JA, Naig AL, Johnson EK. Anorectal abscess and fistula-in-ano: evidence-based management. Surg Clin North Am. 2010;90:45-68. doi: 10.1016/j.suc.2008.10.001. PubMed PMID: 20109632.

15. Whiteford MH, Kilkenny J, 3rd, Hyman N, Buie WD, Cohen J, Orsay C, et al. Practice parameters for the treatment of perianal abscess and fistula-in-ano (revised). Dis Colon Rectum. 2005;48:1337-42. doi: 10.1007/s10350-005-0055-3. PubMed PMID: 15933794.

16. van Koperen PJ, Wind J, Bemelman WA, Sors JF. Fibrin glue and transanal rectal advancement flap for high transsphincteric perianal fistulas; is there any advantage? Int J Colorectal Dis. 2008;23:697-701. doi: 10.1007/s00384-008-0460-x. PubMed PMID: 18322659; PubMed Central PMCID: PMCPMC236753.

17. Barlas M, Gokcora H, Erekul S, Dindar H, Yucesan S. Human amniotic membrane as an intestinal patch for neumocosal growth in the rabbit model. J Pediatr Surg. 1992;27:597-601. PubMed PMID: 1625130.

18. Najbpour N, Jahantab MB, Hosseinazedeh M, Roshanravan R, Moslemi S, Rahimikazerouni S, et al. The effects of human amniotic membrane on healing of colonic anastomosis in dogs. Annals of Colorectal Research. 2013;1:97-100. doi: 10.17795/acr-16139.

19. Roshanravan R, Ghahramani L, Hosseinazedeh M, Mohammadipour M, Moslemi S, Rezaianzadeh A, et al. A new method to repair recto-vaginal fistula: Use of human amniotic membrane in an animal model. Adv Biomed Res. 2014;3:114. doi: 10.4103/2277-9175.131033. PubMed PMID: 24804188; PubMed Central PMCID: PMCPMC4009751.

20. Schimidt LR, Cardoso EJ, Schimidt RR, Back LA, Schiazzawa MB, d’Acampora AJ, et al. The use of amniotic membrane in the repair of duodenal wounds in Wistar rats. Acta Cir Bras. 2010;25:18-23. PubMed PMID: 20126882.

21. Kuriu Y, Yamagishi H, Otsuji E, Nakashima S, Miyagawa K, Yoshikawa T, et al. Regeneration of peritoneum using amnioticmembranetopreventpostoperative adhesions. Hepatogastroenterology. 2009;56:1064-8. PubMed PMID: 19760942.
22. Moslemi SM, Joraghi SAM, Roshanravan RM, Ghahramani LM, Mohammadianpanah MM, Hosseinzadeh MM, et al. Effect of Human Amniotic Membrane on Prevention of Colorectal Anastomosis Leakage in Cases with Neoadjuvant Radiotherapy: An Experimental Animal Study. Iran J Med Sci. 2016;41:501-6. PubMed PMID: 27853330; PubMed Central PMCID: PMCPMC5106565.

23. Tahamtan M, Hosseini SV, Khazraei H, Forozesh M, Bananzadeh A, Mokhtari M, et al. Effect of human amniotic membrane on anastomosis leakage in dog model. Comp Clin Path. 2016;25:1121-6. doi: 10.1007/s00580-016-2313-2.