Commentary

Paradoxical effects of osteoprotegerin on vascular function: inhibiting inflammation while promoting oxidative stress?

Nhat-Tu Le¹, Elizabeth A. Olmsted-Davis¹ and Jun-ichi Abe²

¹Academic Institute, Department of Cardiovascular Sciences, Center for Cardiovascular Sciences, Houston Methodist Research Institute, Weill Cornell Medical College, Houston, TX, U.S.A.; ²Department of Cardiology, The University of Texas MD Anderson Cancer Center, Houston, TX, U.S.A.

Correspondence: Jun-ichi Abe (jabe@mdanderson.org)

Osteoprotegerin (OPG), also known as osteoclastogenesis inhibitory factor or tumor necrosis factor receptor superfamily member 11B, is well known as a modulator of bone remodeling. The contribution of OPG to cardiovascular disease (CVD) has been suggested, but its molecular mechanism is complex and remains unclear. In the present study, Alves-Lopes et al. (Clin. Sci. (Lond.) (2021) 135(20); https://doi.org/10.1042/CS20210643) reported the critical role of syndecan-1 (SDC-1, also known as CD138), a surface protein part of the endothelial glycocalyx, in OPG-induced vascular dysfunction. The authors found that in endothelial cells (ECs), through SDC-1, OPG increased eNOS Thr495 phosphorylation, thereby inhibiting eNOS activity. Furthermore, the OPG–SDC-1 interaction increased reactive oxygen species (ROS) production through NOX1/4 activation. Both the reduced eNOS activity and induced ROS production inhibited NO production and impaired EC function. In vascular smooth muscle cells (VSMCs), the OPG–SDC-1 interaction increased ROS production through NOX1/4 activation, subsequently increased MLC phosphorylation-mediated Rho kinase-MYPT1 regulation, leading to increased vascular contraction. Utilizing wire myography and mechanistic studies, the authors nicely provide the evidence that SDC-1 plays a crucial role in OPG-induced vascular dysfunction. As we mentioned above, the molecular mechanism and roles of OPG in cardiovascular system are complex and somewhat confusing. In this commentary, we briefly summarize the OPG-mediated signaling pathways in cardiovascular system.

The OPG–RANKL interaction inhibits vascular inflammation

Osteoprotegerin (OPG), also known as osteoclastogenesis inhibitory factor or tumor necrosis factor receptor superfamily member 11B, is well known as a modulator of bone remodeling. The contribution of OPG to cardiovascular disease (CVD) has been suggested, but its molecular mechanism is complex and remains unclear. In the present study, Alves-Lopes et al. (Clin. Sci. (Lond.) (2021) 135(20); https://doi.org/10.1042/CS20210643) reported the critical role of syndecan-1 (SDC-1, also known as CD138), a surface protein part of the endothelial glycocalyx, in OPG-induced vascular dysfunction. The authors found that in endothelial cells (ECs), through SDC-1, OPG increased eNOS Thr495 phosphorylation, thereby inhibiting eNOS activity. Furthermore, the OPG–SDC-1 interaction increased reactive oxygen species (ROS) production through NOX1/4 activation. Both the reduced eNOS activity and induced ROS production inhibited NO production and impaired EC function. In vascular smooth muscle cells (VSMCs), the OPG–SDC-1 interaction increased ROS production through NOX1/4 activation, subsequently increased MLC phosphorylation-mediated Rho kinase-MYPT1 regulation, leading to increased vascular contraction. Utilizing wire myography and mechanistic studies, the authors nicely provide the evidence that SDC-1 plays a crucial role in OPG-induced vascular dysfunction. As we mentioned above, the molecular mechanism and roles of OPG in cardiovascular system are complex and somewhat confusing. In this commentary, we briefly summarize the OPG-mediated signaling pathways in cardiovascular system.
Figure 1. OPG-mediated signaling pathways

OPG inhibits RANKL and TRAIL binding to their receptors. In contrast, OPG associates with SDC-1, which promotes growth factor receptor-mediated signaling. RANK itself does not have any kinase activity, but rather signals through binding TRAFs, GRB-associated-binding protein 2 (GAB2), and Src kinase, which assist in initiating the signaling cascade leading to NF-κB, MAPK, and AP-1 activation inflammation, and vascular calcification [5,6]. TRAIL-R1/2 contains a death domain in its C-terminal, and when bound can produce apoptotic signals. However, in endothelial cells (ECs) and VSMCs, TRAIL-R1/2 can increase PI3-K/Akt signaling instead of Fas-associated protein with death domain (FADD)-mediated caspase 8 activation, to promote cellular survival and eNOS activation as described in the studies by Alves-Lopes et al. [7]. This phenomenon is also reported in cancer cells, and may be one of the mechanisms to explain the resistance to TRAIL-induced cancer cell apoptosis. However, how TRAIL-R1/2 can change its signaling from apoptosis to survival remains unclear [7]. Lastly, OPG associates with SDC-1 at a CAG side chain. CAG side chain contains heparan sulfate (HS), which forms a complex with growth receptor/ligands and acts as a co-receptor. In this manuscript, Alves-Lopes et al. [] showed that OPG can induce reactive oxygen species (ROS) production via SDC-1. Therefore, it is possible that OPG binds with HS, and promotes SDC-1 co-receptor activation as described in their studies. Made by Biorender.

suggesting that in the context of RANKL–RANK-mediated VSMC calcification via TRAF–NF-κB–BMP4 signaling, OPG exerts an inhibitory effect on RANKL-induced NF-κB activation. Indeed, the depletion of OPG in ApoE-/- mice exhibited the acceleration of not only vascular calcification, but also atherosclerosis formation [9]. Therefore, in this context, OPG functions as an anti-atherosclerotic factor probably by inhibiting vascular inflammation (Figure 1).

The OPG–TRAIL interaction inhibits vascular apoptosis

TRAIL-R1 and R2 are type 1 transmembrane receptors containing an intracellular death domain (DD) but the association of TRAIL with TRAIL-R1 and 2 does not always induce apoptosis. In contrast, TRAIL–TRAIL-R1/2 can induce survival by activating PI3-K-Akt and ERK1/2 signaling. Particularly in vascular cells such as ECs and VSMCs, the induction of pro-survival pathway of PI3-K-Akt and ERK1/2 activation induced by TRAIL is more prominent than pro-apoptotic signaling after the exposure of these cells to ligand (Figure 1). For example, it has been reported that TRAIL inhibits interferon-γ, TNF-α, and IL-1β-induced VSMC apoptosis, and enhances VSMC migration and proliferation [10]. In endothelial cells (ECs), TRAIL activates PI3-K/Akt pathway and subsequently increases eNOS activity [11,12]. In fact, systemic administration of TRAIL inhibits atherosclerotic plaque formation, and genetic deletion
of TRAIL accelerates it [13,14]. These data suggest that TRAIL possesses protective effects against vascular dysfunction and is atheroprotective. As stated above, since OPG inhibits TRAIL–TRAIL-R1/2 association, the detrimental effects of OPG on vascular function can also be explained by OPG-mediated inhibitory effects on TRAIL–TRAILR1/2 signaling.

The OPG–SDC-1 interaction induces oxidative stress

The binding of OPG with syndecan family has been reported [15]. Syndecans are transmembrane proteoglycans (PGs) with a highly conserved C-terminal cytoplasmic domain, a single-pass transmembrane domain, and a large N-terminal extracellular domain. The extracellular domain of SDC-1 possesses five glycosaminoglycan (CAG) side chains consisting of heparan sulfate (HS) and chondroitin sulfate (CS). HS chains play a major role for SDC-1 to associate with heparin-binding growth factors and their receptors such as FGFs, VEGFs, Wnt, and HGF [15,16]. HS associates with both the growth factor and its receptor, stabilizes the complex. Therefore, SDC-1 can act as ‘co-receptor’, and plays a crucial role in growth factor activation (Figure 1). For example, 2-O-sulfate is critical for FGF2 and HS binding, leading to increase in FGF2 and its receptor binding, and promote FGFR-mediated signaling [17]. The activation of NADPH oxidase induced by growth factor receptor activation has been well established [18–22]. Therefore, it is reasonable to speculate that OPG–SDC-1 association induces NOX1/4 activation via various growth factor receptor activation.

Conclusion

In the present study, Alves-Lopes et al. show that OPG can induce vascular dysfunction via inducing reactive oxygen species (ROS) production []; however, the atheroprotective role of OPG has been suggested in previous studies. At a glance, this seems to be contradictory, but it might be due to the different roles of inflammation and growth factor-induced ROS production during the process of particular cardiovascular events. It is well known that strong inflammatory events are instigated in significantly high cholesterol level-mediated atherosclerotic plaque formation in apoE−/− mice. In this case, the anti-inflammatory effects of OPG on RANKL–RANK signaling may play a key role in inhibiting atherosclerotic plaque formation. In contrast, without significant hypercholesterolemia condition, anti-inflammatory effects of OPG may play only a minor role. Instead OPG-induced NOX1/2 activation via SDC-1 may become more critical in regulating vascular contraction and accelerating vascular dysfunction. The data of OPG effects on vascular dysfunction may suggest an importance of the stage and severity of the vascular dysfunction mediated by both inflammation and ROS production. Inflammatory factors may not equally contribute to the process of vascular dysfunction and consequent atherosclerotic plaque formation. Therefore, the effects of inflammation and ROS on the vasculature may be very different. Future studies may need to focus on the similarities and differences between responses to inflammation and ROS. To study the effects of OPG during the process of atherosclerosis may provide a good model.

Data Availability

No new data are available in this manuscript.

Competing Interests

The authors declare that there are no competing interests associated with the manuscript.

Funding

This work was supported by the National Institutes of Health (NIH) [grant number HL-149303 (to Nhat-Tu Le and Jun-ichi Abe)].

Open Access

Open access for this article was enabled by the participation of University of Texas MD Anderson Cancer Center in an all-inclusive Read & Publish pilot with Portland Press and the Biochemical Society under a transformative agreement with EBSCO.

Abbreviations

eNOS, endothelial nitric oxide synthase; HS, heparan sulfate; NOX1/4, NADPH oxidase 1/4; OPG, osteoprotegerin; RANKL, receptor activator of nuclear factor-κB ligand; ROS, reactive oxygen species; SDC-1, syndecan-1; TNF, Tumor necrosis factor; TRAF, TNF receptor-associated factor; TRAIL, TNF-related apoptosis-inducing ligand.
References

1. Bernardi, S., Bossi, F., Toffoli, B. and Fabris, B. (2016) Roles and clinical applications of OPG and TRAIL as biomarkers in cardiovascular disease. *Biomed Res. Int.* **2016**, 1752854, https://doi.org/10.1155/2016/1752854

2. Rochette, L. et al. (2019) The role of osteoprotegerin and its ligands in vascular function. *Int. J. Mol. Sci.* **20**, https://doi.org/10.3390/ijms20030705

3. Montagnana, M., Lippi, G., Danese, E. and Guidi, G.C. (2013) The role of osteoprotegerin in cardiovascular disease. *Ann. Med.* **45**, 254–264, https://doi.org/10.3109/07853890.2012.727019

4. Simonet, W.S. et al. (1997) Osteoprotegerin: a novel secreted protein involved in the regulation of bone density. *Cell* **89**, 309–319, https://doi.org/10.1016/S0092-8674(00)80209-3

5. Ahern, E., Smyth, M.J., Dougall, W.C. and Teng, M.W.L. (2018) Roles of the RANKL–RANK axis in antitumour immunity - implications for therapy. *Nat. Rev. Clin. Oncol.* **15**, 676–693, https://doi.org/10.1038/s41571-018-0095-y

6. Dutka, M. et al. (2021) Osteoprotegerin and RANKL-RANK-OPG-TRAIL signalling axis in heart failure and other cardiovascular diseases. *Heart Fail. Rev.*, https://doi.org/10.1007/s10741-021-10153-2

7. Refaat, A., Abd-Rabou, A. and Reda, A. (2014) TRAIL combinations: the new ‘trail’ for cancer therapy (Review). *Oncol. Lett.* **7**, 1327–1332, https://doi.org/10.3892/ol.2014.1922

8. Panizo, S. et al. (2009) RANKL increases vascular smooth muscle cell calcification through a RANK-BMP4-dependent pathway. *Circ. Res.* **104**, 1041–1048, https://doi.org/10.1161/CIRCRESAHA.108.189001

9. Bennett, B.J. et al. (2006) Osteoprotegerin inactivation accelerates advanced atherosclerotic lesion progression and calcification in older ApoE-/- mice. *Arterioscler. Thromb. Vasc. Biol.* **26**, 2117–2124, https://doi.org/10.1161/01.ATV.0000236428.91125.e6

10. Secchiero, P. et al. (2006) Systemic tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) sequentially upregulates nitric oxide and prostanoid production in primary human endothelial cells. *Circ. Res.* **92**, 732–740, https://doi.org/10.1161/01.RES.000067928.83455.9C

11. Liu, M. et al. (2014) TRAIL protects against endothelium injury in diabetes via Akt-eNOS signaling. *Atherosclerosis* **237**, 716–724, https://doi.org/10.1016/j.atherosclerosis.2014.10.013

12. Fuchihata, T., Sekine, N., Inoue, M. and Kajita, T. (2016) TRAIL attenuates the development of atherosclerosis in apolipoprotein E deficient mice. *Atherosclerosis* **251**, 348–354, https://doi.org/10.1016/j.atherosclerosis.2016.11.010

13. Secchiero, P. et al. (2006) Systemic tumor necrosis factor-related apoptosis-inducing ligand delivery shows antiatherosclerotic activity in apolipoprotein E-null diabetic mice. *Circulation* **114**, 1522–1530, https://doi.org/10.1161/CIRCULATIONAHA.106.643841

14. Timmen, M. et al. (2020) The heparan sulfate proteoglycan Syndecan-1 influences local bone cell communication via the RANKL/OPG axis. *Sci. Rep.* **10**, 20510, https://doi.org/10.1038/s41598-020-77510-3

15. Rubin, J.S. et al. (2001) Dissociation of heparan sulfate and receptor binding domains of hepatocyte growth factor reveals that heparan sulfate-c-met interaction facilitates signaling. *J. Biol. Chem.* **276**, 32977–32983, https://doi.org/10.1074/jbc.M105486200

16. Yu, C., Griffiths, L.R. and Haupt, L.M. (2017) Exploiting heparan sulfate proteoglycans in human neurogenesis-controlling lineage specification and fate. *Front. Integr. Neurosci.* **11**, 28, https://doi.org/10.3389/fnint.2017.00028

17. Murillo, M.M. et al. (2007) Activation of NADPH oxidase by transforming growth factor-beta in hepatocytes mediates up-regulation of epidermal growth factor receptor ligands through a nuclear factor-kappaB-dependent mechanism. *Biochem. J.* **405**, 251–259, https://doi.org/10.1042/BJ20061846

18. Jagadeesha, D.K., Takapo, M., Banfi, B., Bhalla, R.C. and Miller, Jr, F.J. (2012) Nox1 transactivation of epidermal growth factor receptor promotes N-cadherin shedding and smooth muscle cell migration. *Cardiovasc. Res.* **93**, 406–413, https://doi.org/10.1093/cvr/cvr308

19. Fan, C.Y., Katsuyama, M. and Yabe-Nishimura, C. (2005) PKCdelta mediates up-regulation of NOX1, a catalytic subunit of NADPH oxidase, via transactivation of the EGF receptor: possible involvement of PKCdelta in vascular hypertrophy. Biochem. J. **390**, 761–767, https://doi.org/10.1042/BJ20050287

20. Boudreau, H.E., Casterline, B.W., Rada, B., Korzeniowska, A. and Loto, T.L. (2012) Nox4 involvement in TGF-beta and SMAD3-driven induction of the epithelial-to-mesenchymal transition and migration of breast epithelial cells. *Free Radic. Biol. Med.* **53**, 1489–1499, https://doi.org/10.1016/j.freeradbiomed.2012.06.016

21. Galan, M. et al. (2012) A novel role for epidermal growth factor receptor tyrosine kinase and its downstream endoplasmic reticulum stress in cardiac damage and microvascular dysfunction in type 1 diabetes mellitus. *Hypertension* **60**, 71–80, https://doi.org/10.1161/HYPERTENSIONAHA.112.192500

© 2022 The Author(s). This is an open access article published by Portland Press Limited on behalf of the Biochemical Society and distributed under the Creative Commons Attribution License 4.0 (CC BY).