Bioinspired pH and magnetic responsive catechol-functionalized chitosan hydrogels with tunable elastic properties

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We have developed pH- and magnetic-responsive hydrogels that are stabilized by both covalent bonding and catechol/Fe$^{3+}$ ligands. The viscoelastic properties of the gels are regulated by the complexation valence and can be used to tune drug release profiles. The stable incorporation of magnetic nanoparticles further expands control over the mechanical response and drug release, in addition to providing magnetic stimuli-responsivity to the gels.

The field of hydrogels has rapidly expanded in recent years, with bioinspired and biopolymer-based hydrogels representing an increasingly large fraction of these efforts. A growing theme is the development of hydrogels that are multi-functional, stimuli-responsive, and with tailorable physico-chemical and mechanical properties. Towards obtaining gels with tunable properties, metal coordination bonding has recently emerged as a versatile strategy, notably by exploiting catechol moiety/transition metal complexation. Such ligands are particularly useful to stabilize and modulate gels viscoelastic properties owing to: (i) their pH dependence that triggers either mono-, bi- or tri-dentate complexes, and (ii) the ability to further tune their properties by simple modification of the transition metal ions used to coordinate the catecholic moieties.

On the other hand, the conformational response of many hydrogels, which would be useful in many applications, remains slow. This limitation can be overcome by preparing magnetic hydrogels, composed of a discrete magnetic phase such as magnetic nanoparticles (MNPs) embedded within a polymeric matrix. Magnetically-controlled hydrogels provide unique functionalities in biomedical applications such as tissue engineering and drug delivery. For instance, they can be actuated in a controlled manner to prepare 3D tissue scaffolds during cell proliferation. Alternatively, the behavior of encapsulated cells can also be controlled in vitro by a remotely-activated mechanical stimulus, which is triggered by interactions between the MNPs and an external magnetic field. Using alternating magnetic fields, MNPs can also be remotely heated to induce: (i) conformational changes to the hydrogels, which can be exploited for the pulsatile release of drugs, or (ii) hydrophobic-to-hydrophilic phase transitions that can promote large dimensional changes. Finally, MNPs provide contrast in magnetic resonance imaging studies, enabling therapy and diagnostic (theranostic) related protocols. Thus, the introduction of magnetic responsiveness to the hydrogel increases its versatility to develop a truly multi-functional, multi-stimuli responsive material.

Chitosan, the deacetylated version of the polysaccharide chitin, represents a convenient source of biodegradable biopolymers for a wide range of soft materials, including hydrogels. It is highly abundant, notably in the waste from the seafood industry; it is soluble in weak acidic solutions; and it can readily be functionalized to expand its range of functional properties for biomedical applications such as gene delivery and tissue repair. The merging of the two approaches, namely catechol functionalization of chitosan, is quickly emerging as a method of choice to combine the specific advantages of catechol complexation and the ease of processing of chitosan, as exemplified in recent studies. Herein, we explore the potential of catechol-functionalized chitosan to engineer pH-sensitive and magnetic-responsive hydrogels by taking advantage of metal–catechol coordination. Hydrocaffeic acid (HCA) bearing the catechol moiety was used to functionalize chitosan, allowing the modulation of the viscoelastic response of HCA–chitosan gels via pH triggering and variation of the network mesh size, which in turn allowed the tuning of drug release profiles. In order to further expand the multi-functionality of these gels, we then incorporated iron oxide ($\gamma$Fe$_2$O$_3$) MNPs in their formulation and established that this strategy adds an additional level of multi-functionality.
First, it allows further modulation of the mechanical response; second it provides an additional control over drug release kinetics; and third, it confers magnetic responsivity on the gels, opening the door to their usage as stimuli-responsive, biocompatible hydrogels with tunability of drug release kinetics.

First, we functionalized high molecular weight chitosan ($M_w \approx 94$ KDa) with the HCA catecholic moiety, with a degree of substitution of $\sim 15\%$ as measured by $^1$H NMR (Fig. S1, ESI†). The preparation of hydrogels in the absence of MNPs was then investigated. Upon addition of Fe$^{3+}$ (HCA : Fe$^{3+} = 3 : 1$), increasing the pH of the HCA–chitosan solution to 6 and 12 led to the formation of bis- and tris-complexes, respectively. As a control, we also prepared covalently cross-linked hydrogels (hereafter referred to as “chemical hydrogels”) by increasing the pH of HCA–chitosan to 13 (Scheme 1).

![Scheme 1](Image)

**Scheme 1** Schematic of pH-tunable HCA–chitosan hydrogels stabilized by covalent bonding (“chemical gels”) and catechol/Fe$^{3+}$ complexation (“physical gels”). Gelation conditions used: (a) NaOH pH 13; (b) FeCl$_3$ (Fe$^{3+}$ : HCA = 1 : 3), pH 6; (c) FeCl$_3$ (Fe$^{3+}$ : HCA = 1 : 3), pH 12. Note the yellow–orange color of the covalently cross-linked hydrogel, and the blue–purple and red colors of the metal coordinated cross-linked hydrogels at pH 6 and 12, respectively.

Fig. 1 shows the UV-Vis spectra of the chemical and physical (stabilized by coordination complexation) hydrogels prepared with and without Fe$^{3+}$ (HCA : Fe$^{3+} = 3 : 1$) at different pHs. The presence of an absorbance peak at 280 nm for HCA–chitosan confirms the successful grafting of HCA to the chitosan backbone. Increasing the pH from acidic to basic, the color of HCA–chitosan solution changed from transparent to a yellowish/orange color due to the high molecular weight of chitosan (resulting in abundant free amino groups), the high degree of substitution (DS $\sim 15\%$), and the redox effect of Fe$^{3+}$. This is also reflected by the appearance of peaks at $\sim 260$ and 350 nm. It is worth mentioning that the mono-complex associated with a green color formation was observed when the pH of the HCA–chitosan solution was increased from $\sim 2$ to 3–4 in the presence of Fe$^{3+}$ (data not shown). The visco-elastic properties of the synthesized hydrogels were characterized two hours after gelation by rheology in the dynamic mode at 20 °C (Fig. 2). In the tested frequency range, HCA–chitosan (without Fe$^{3+}$) at pH 13 showed the characteristics of a soft hydrogel, with a constant storage modulus $G'_{\infty}$ of ca. 500 Pa. The damping factor ($\tan \delta = G'/G''$) was approximately 1 below 0.1 Hz, implying network properties at the boundary between fluid- and solid-like gel. However, above 0.1 Hz the gel was solid-like, with $G'/G'' > 1$. The addition of Fe$^{3+}$ greatly increased the storage modulus of the gels, which increased from 6 to ca. 20 KPa (at 1 Hz) as the pH of the HCA–chitosan solution was increased from 6 to 12. At pH 3 in the presence of Fe$^{3+}$, $G'$ values were considerably lower (< 1 Pa, Fig. S3, ESI†). This increase in stiffness can unambiguously be attributed to catechol/Fe$^{3+}$ complexation since no other bonds are introduced into the network. As previously reported, increasing the pH modifies the nature of the coordination complexes between the catechol side-chains and Fe$^{3+}$, with mono-complexes present below pH 3, bis-complexes at pH 6 and tris-complexes being the dominant species at pH 12. Using the classical relationship between storage modulus and mesh size ($G' \sim k_B T/\xi^3$), we determined that $\xi$ decreased from ca. 22 nm at pH 13 in the absence of Fe$^{3+}$ to 10 nm (pH 6, Fe$^{3+}$) and 5 nm (pH 12, Fe$^{3+}$), suggesting a direct correlation between the complexation state and the mesh size (Fig. 2B).

To assess the reversibility of the gelation process, the tris-complex hydrogel was treated with either HCl or EDTA overnight. The final pH of HCl treated hydrogel solution was found to be 5.5. As a control, the chemical hydrogel was also treated...
hydrogels maintained a dark red color, suggesting a faster kinetics to 26 kPa at 1 Hz (Fig. 3A and Fig. S4A, ESI).

The UV-Vis analysis supports this finding: a peak at 570 nm appeared after HCl treatment corresponding to the bis-complex at this lower pH. Treating the gel with EDTA showed no evidence for any absorbance peak above 450 nm, confirming the removal of all Fe³⁺ from the gel (Fig. 2D). The higher storage modulus of this hydrogel compared to that of the chemical hydrogel may be attributed to the formation of chemical crosslinks concomitantly taking place during metal chelation, leading to a gel with a higher covalent cross-link density. Collectively, these results indicate that the viscoelastic response of HCA–chitosan gels can be reversibly modulated by chelation or re-incorporation of metal ions.

We previously reported HCA–chitosan films reinforced with MNPs, in which the mechanical properties of the films were enhanced through load transfer from the functionalized-chitosan to MNPs via coordination interactions between catechol and Fe₂O₃. To confer dual pH and magnetic responsiveness, MNP particles were added to the hydrogels. To this end, HCA-chit were mixed with Fe₂O₃ MNPs (2.5 and 5 wt%) and gelled by increasing the pH in the absence or presence of Fe³⁺ (HCA:Fe³⁺ = 3 : 1), to yield both chemical and physical hybrid hydrogels. The chemical hydrogels featured a light-brown visual appearance consistent with the color of maghemite Fe₂O₃, whereas the physically cross-linked hydrogels maintained a dark red color, suggesting a faster kinetics of complexation for Fe³⁺/catechol than for Fe₂O₃/catechol. The addition of 2.5 and 5 wt% of MNPs to the physical gels resulted in a linear increase in the storage modulus with MNP content of up to 26 kPa at 1 Hz (Fig. 3A and Fig. S4A, ESI†). However, we note that the average mesh size ζ estimated from G’ ∼ k_B T ζ² decreased only moderately upon addition of MNPs (Fig. 3B), suggesting that catechols favour interactions with Fe³⁺ and that MNPs primarily act as fillers in the physical gels but do not specifically interact with MNPs. In contrast, the relative increase of G’ with MNP content for chemical gels was more distinct – though lower in absolute values than for the physical gels – and is reflected by the more pronounced relative decrease in mesh size upon addition of MNPs (Fig. 3B).

In the absence of Fe³⁺, HCA moieties in the chemical gels are free to coordinate the surface of Fe₂O₃, thus allowing load transfer from the soft chitosan–HCA matrix to the stiff MNPs. The increase in G’ may also be assessed in terms of decreased swelling upon addition of MNPs. The storage modulus of swollen networks is well-established to exhibit an inverse power law dependence on the swelling ratio Q (G’ ~ Q⁻ᵐ),²⁵ where m is in the range of 2.2 to 3 depending on the solvent. To verify this dependency, we plotted G’ vs. Q (Fig. S4B, ESI†) and obtained G’ ∼ Q⁻¹.²⁹ for chemical/MNPs gels, which is in good agreement with the power law predicted for theta solvents and similar to that measured for various PEG hybrid hydrogels.²⁶,²⁷ In contrast, in physical/MNPs hydrogels a weaker dependence of G’ on swelling was observed with G’ ∼ Q⁻¹.², which is consistent with a network where the catecholic moieties are saturated with Fe³⁺ coordination bonds and thus less available for water uptake. In summary, these data reveal a distinct dependency of G’ on the MNP content, which can be attributed to substantial differences in the swelling behaviour in the presence or absence of catechol/Fe³⁺ coordination bonds.

Vibrating Sample Magnetometer (VSM) measurements were conducted in order to confirm the magnetism of both chemical and physical hydrogels. Fig. 3C illustrates the magnetization versus magnetic field (M–H loop) for the chemical and physical hydrogels at various weight fractions of MNPs. All hydrogels displayed a low hysteresis behavior similar to native Fe₂O₃ (Fig. S2D, ESI†). The magnetic property values of the hydrogels are fully consistent with
the weight fraction of MNPs in the gels. No change was observed in the magnetic properties even after maintaining the gels in water for 10 days (data not shown). This indicates that the MNPs remained entrapped within the gel without further oxidation. For the same weight fraction of MNPs, physical gels exhibited slightly higher values (~0.18 vs. 0.16 and 0.09 vs. 0.07 emu g\(^{-1}\) at 5 and 2.5 wt%, respectively), which can be attributed to the smaller swelling ratio of physical gels compared to their chemical analogues (Fig. S5, ESI\(^{†}\)). A video illustrating the magnetically-induced shape morphing of the magnetic hydrogels is provided in the ESI\(^{†}\).

In addition to tuning the mechanical properties, MNP hydrogels can also generate additional stress and/or strain in DC magnetic fields, while an AC field could generate a temperature change, enabling thermal triggering. The quick magnetic responsivity could also generate additional stress and/or strain in DC magnetic fields, which might be used as implantable magnetic gels containing drugs or cells, enabling thermal triggering. The quick magnetic responsivity could be exploited to tailor the mechanical properties of the scaffold to suit optimal growth conditions, while the pH responsiveness could be used to trigger the release of growth factors at the appropriate times and/or spatial locations. Although the cytotoxicity of these stimuli-responsive gels remains to be assessed, recent studies have indicated a low cytotoxicity of metal coordination reinforced gels using vanadium ions as ligand centers, suggesting a promising alternative to Fe\(^{3+}\) cross-linked gels should the latter prove cytotoxic.

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