Paclitaxel-resistant cancer cell-derived secretomes elicit ABCB1-associated docetaxel cross-resistance and escape from apoptosis through FOXO3a-driven glycolytic regulation

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Chemotherapy-induced cancer cell secretomes promote resistance due, in part, to a predominant glycolytic energy metabolism, which drives aggressive cancer cell proliferation. However, the characterization of these secretomes and the molecular events that associate them with acquired drug resistance remain poorly understood. In this study, we show that secretomes of cancer cells with high-level paclitaxel resistance stimulated cell proliferation and suppressed drug-induced apoptosis of drug-sensitive cells. We also found that drug (docetaxel)-stimulated induction of interferon-α (IFN-α), IFN-λ, and tumor necrosis factor-α (TNF-α) release in drug-sensitive cells was lowered by these secretomes. The promotion of cell proliferation by paclitaxel-resistant (PacR) cancer cell secretomes was associated, in part, with an increase in S phase of the cell cycle and downregulation of the cell death pathway that supports escape from apoptosis. In addition, we also found that the regulation of targeted glycolysis in PacR cancer cells alters the effects of the secretomes on cell growth, apoptosis, ATP generation and acquired drug resistance. Further study revealed that the deletion of FOXO3a transcription exacerbates glycolytic shift-induced apoptosis by rescuing TRAIL expression. By generating a docetaxel–cross-resistant PacR cancer cell line (PacR/DCT), we further clarified the role of FOXO3a in glycolysis-associated mediation of P-glycoprotein/ABCB1 hyperactivity that induces docetaxel cross-resistance. These findings suggest that suppression of the cellular energy supply by targeting glycolysis may inhibit the multiplicity of acquired chemotherapy resistance. Therefore, the therapeutic inhibition of FOXO3a might direct glycolysis to induce apoptosis and overcome multidrug resistance in cancer cells.

INTRODUCTION

Acquired chemoresistance induces aggressiveness and causes relapse in a variety of cancer types. This resistance limits the efficacy of targeted therapies after a majority of patients show disease stabilization, which hampers the success of clinical treatments and increases the risk of death.1–3 Paclitaxel is the main treatment, along with platinum therapy, for ovarian, lung, prostate and breast cancer. Paclitaxel interrupts the dynamic equilibrium of tubulins and stabilizes the microtubule structure. Although studies have uncovered the mechanisms of paclitaxel resistance (PacR) in several malignancies, PacR remains a complex and unsolved issue in the clinical setting. Various mechanisms have been implicated in PacR, including the following: increased P-glycoprotein (P-gp), which is encoded by ABCB1 (MDR1); associated drug efflux; blocked death signals; changed microtubulin dynamics; and altered stress responses, such as the activation of DNA repair and detoxification signals.4–7 However, clinical agents that regulate these mechanisms, such as P-gp inhibitors, are often ineffective or toxic at the doses required to induce efficacy.8 Therefore, current options for overcoming PacR are limited, necessitating the identification of more selective cancer therapies.

Altered energy metabolism (Warburg effect) has been recognized as one of the hallmarks of cancer. It has been demonstrated that the metabolic properties of drug-resistant cancer cells are different from those of drug-sensitive cancer cells, the same is true for cancer cells versus normal cells.9 Dysregulated cellular metabolism has been linked to...
development of drug-resistant phenotypes, increased autophagy levels and regulation of critical glycolysis-associated molecules. Targeting dysregulated glucose metabolism overcomes therapeutic resistance in a number of models involving different mechanisms.\textsuperscript{10} Targeting glycolysis has been assessed using different approaches, one of the most frequent techniques used is modifying the glucose content in cells. Under glucose deprivation (GD), cells conserve energy to ensure survival and other related functions.\textsuperscript{11} Prolonged GD induces cellular stress, which regulates glucose-regulated protein 78 (GRP78) and other related factors that confer protection from apoptosis.\textsuperscript{12} Intracellularly, this method has been used to study glucose metabolism, but whether GD can be used to clinically access tumor response remains intriguing.\textsuperscript{13} Meanwhile, there has been slow progress in understanding the role of glucose metabolism in the secretion of complex factors that support tumorigenesis and drug resistance.

FOXO3a has been closely implicated in multidrug resistance through the expression of ABCB1 and PIK3CA in a limited number of malignancies.\textsuperscript{14–16} Highly drug-resistant cancer cells are characterized by anti-apoptotic mechanisms. FOXO3a is phosphorylated by Akt, which inhibits the transactivation of target genes associated with apoptosis and cell proliferation, such as p27\textsuperscript{kip1}, cyclin D, Bim and Bcl.\textsuperscript{17,18} This inhibition changes the status of FOXO3a as a major target of inactivation by PI3K/Akt. However, little is known regarding how FOXO3a affects the development of resistance in drug-sensitive cells as mediated by therapy/drug-induced cancer cell secretomes. Thus, a better understanding of this process is warranted to improve therapeutic outcomes.

Here we report that FOXO3a deletion and targeted glycolysis block drug-sensitive cancer cell escape from apoptosis and in the development of docetaxel cross-resistance in PacR cells induced by PacR cancer cell-derived secretomes.

**MATERIALS AND METHODS**

**Parental and drug-resistant cancer cell lines**

Parental A549 human lung cancer and PC-3 human prostate cancer cell lines were obtained from American Type Culture Collection (Manassas, VA, USA) and were cultured in RPMI supplemented with 10% heat-inactivated fetal bovine serum, 100 U ml\textsuperscript{-1} penicillin, 100 μg ml\textsuperscript{-1} streptomycin and 250 ng ml\textsuperscript{-1} amphotericin B at 37 °C in a humidified atmosphere with 5% CO\textsubscript{2}. PacR and PacR/DCT cell lines were developed from their parental cells (both A549 and PC-3) using stepwise selection for resistance with increasing concentrations of paclitaxel and docetaxel, respectively. The PacR cells were developed over a period of 12 months and were maintained with 100 nM paclitaxel. The PacR/DCT cells were derived from the PacR cells and were further developed for 4 weeks to have transient cross-resistance to docetaxel; the cells were maintained with low concentrations of paclitaxel and docetaxel. PacR-derived cells were characterized as being >200-fold more resistant to paclitaxel and to overexpress P-gp/ABCB1.\textsuperscript{19–21}

**Glucose modification**

For glucose supplementation, adherent cells were maintained in RPMI 1640, which contained 25 mM glucose supplemented with 5–10% fetal calf serum. In the experimental culture, the cells were seeded in high-glucose medium, and the medium was replaced after 48 h with either fresh high-glucose medium (+25 mM glucose) or glucose-deprived medium, which contained no glucose (0 mM glucose; Life Technologies, Grand Island, NY, USA). After glucose modification, PacR cells were washed, the medium was replaced with fresh PacR-maintaining cell culture media and cells were further conditioned for 24 h. This protocol ensured that the collected conditioned media (CM) of PacR cancer cells are not glucose-modified media (glucose-enriched or glucose-deprived) but rather the soluble secretomes of PacR cancer cells. The unmodified CM of PacR cancer cells was considered control secretomes.

**RNA silencing and transfection**

For transient gene silencing of FOXO3a, the cells were transfected with stealth scramble (5’-UUUCUCCGAACGUUCGGUTT-3’) or FOXO3a (5’-AAUGUGACAUAGGUCCAUUAAA-3’) short interfering RNAs (siRNAs; Santa Cruz Biotec, Santa Cruz, CA, USA) using Lipofectamine 3000. The transfectants were subjected to drug treatment and/or assays 48 h after transfection.\textsuperscript{22}

**Cytokine profiling**

The interleukin-6 (IL-6), interferon-α (IFN-α), IFN-β, IFN-λ1, IFN-α2 and tumor necrosis factor-α (TNF-α) levels were quantified using cytokine-specific enzyme-linked immunosorbent assay kits with pre-coated capture antibodies as per the manufacturer’s instructions (Sigma-Aldrich, St Louis, MO, USA). IL-6 levels were preliminarily detected using a Q-Plex Human cytokine screen (16-plex; Quansys Biosciences, Logan, UT, USA). Cell supernatants were prepared, and cytokine release was analyzed as described previously.\textsuperscript{23}

**Cell growth, cell death, cell cycle and caspase-3/-9 activity assays**

Cell viability was assessed using an MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) assay as described previously.\textsuperscript{24} Apoptosis was assessed by subjecting the cells to sub-G\textsubscript{1} fraction analysis by fluorescence-activated cell sorting (FACS) using a BD FACSCalibur flow cytometer (BD Biosciences, San Jose, CA, USA), a terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL) assay (Promega, Madison, MA, USA) and live-cell monitoring coupled with propidium iodide (PI) incorporation into dead/dying cells.\textsuperscript{20,24,25} The analysis of cell cycle distribution was performed by sorting fluorescent cells and evaluating DNA content by FACS.\textsuperscript{26} Caspase-3 and caspase-9 activity was detected using the CaspACE Assay System (Promega) with alternative Caspase-Glo assays.

**Confocal microscopy**

P-gp localization and DNA fluorescence indicated by 4,6-diamidino-2-phenylindole were observed using a Zeiss LSM 780 microscope (Carl Zeiss, Jena, Germany). Cells were grown on coverglass-bottomed, 35-mm dishes pre-coated with 0.2–0.5% gelatin. The cells were then fixed with 4% paraformaldehyde for 15 min and blocked with 1% bovine serum albumin (BSA) for 1 h. The fixed cells were incubated with P-gp primary human antibody at 4 °C overnight, and then the cells were incubated with fluorescein isothiocyanate-conjugated secondary antibody. 4,6-Diamidino-2-phenylindole was used to counterstain the nuclei.

**Determination of ATP content**

Transfected cells were seeded in 96-well plates and were treated with dimethylsulphoxide or the indicated drugs the following day with the
indicated stimulus and duration. CellTiter-Glo reagent (Promega) was used to investigate intracellular ATP content according to the manufacturer’s instructions. The data acquisition was measured as described previously.\textsuperscript{27}

**Western blotting**

Proteins were extracted using RIPA (radioimmunoprecipitation assay) lysis buffer containing 20 mM Tris-HCl (pH 7.5), 1% Triton X-100, 150 mM sodium chloride, 10% glycerol, 1 mM sodium orthovanadate, 50 mM sodium fluoride, 100 mM phenylmethylsulfonyl fluoride and 1× protease inhibitor (Roche Applied Science, Manheim, Germany) at 4 °C. The protein concentration of lysates and supernatants was determined using the Bradford method. Western blot analysis was carried out as described previously.\textsuperscript{20} In brief, the cells were lysed by boiling 2× sample loading buffer (250 mM Tris-HCl (pH 6.8), 4% SDS, 10% glycerol, 0.006% bromophenol blue, 2% β-mercaptoethanol, 50 mM sodium fluoride and 5 mM sodium orthovanadate) and were incubated for 10 min at 100 °C. Proteins were subjected to SDS-polyacrylamide gel electrophoresis and were transferred onto membranes (Millipore, Bedford, MA, USA). The blots were detected on X-ray flms or with an LAS-4000 imaging system (Fuji Film Corp., Tokyo, Japan). The blots were blocked with 5% BSA or 5× boiling 2× sample loading buffer (250 mM Tris-HCl (pH 6.8), 4% SDS, 10% glycerol, 0.006% bromophenol blue, 2% β-mercaptoethanol, 50 mM sodium fluoride and 5 mM sodium orthovanadate) and were incubated for 2 h at room temperature with the corresponding secondary antibodies in 2.5% BSA or 5% skim milk overnight at 4 °C. The membranes were washed with TBST and were incubated for 2 h at room temperature with the corresponding secondary antibodies diluted in 2.5% BSA or 5% skim milk. The membranes were washed with TBST and were exposed to enhanced chemiluminescence solution (Intron, Daejeon, Korea). The blots were detected on X-ray film or with an LAS-4000 imaging system (Fuji Film Corp., Tokyo, Japan).

**Quantitative real-time RT-PCR**

Total cellular RNA was isolated, and complementary DNA was synthesized using an OmniScript cDNA synthesis kit (Qiagen, Valencia, CA, USA). The samples for quantitative reverse transcription PCR were prepared using GoTaq and SYBR Green (Promega). Threshold cycle (Ct) values were determined, and samples were run on a Real-Time PCR system (Bio-Rad, Hercules, CA, USA) using a thermal prole that we have previously described.\textsuperscript{28} The primer sequences used for analysis are listed in Table 1. Real-time PCR was performed as previously described.\textsuperscript{29}

**Statistical analysis**

Statistical significance (\( P < 0.05 \)) was assessed using Student’s t-test or one-way analysis of variance coupled with Dunnett’s t-test performed using OriginPro 8.0 (OriginLab Corp, Northampton, MA, USA). For all compounds, half-maximal inhibitory concentration values and non-linear regressions were calculated using TableCurve 2D v5.01 (Systat Software, San Jose, CA).

**RESULTS**

A549-PacR cancer cell-derived secretomes lowered drug-stimulated release of IFN-α, IFN-λ, and TNF-α in A549 cells

Cytokines contribute to the regulation of drug-sensitive cell protection afforded by drug-resistant secretomes in vivo.\textsuperscript{30} As IFNs and TNF-α accelerate apoptosis in several malignancies, these factors are actively being considered for the development of new treatment modalities.\textsuperscript{31} In this context, the effects of PacR cell secretomes on the release of these cytokines were determined in A549 cells. Initially, the A549-PacR cells were exposed to three different conditions with or without trypsin (stimulation), two conditions with known cytokine-release-inducing effects (that is, hypoxic or serum starvation conditions) and a control condition (that is, normal culture conditions). The levels of IL-6 were found to be three- to eightfold higher in the supernatants. Because IL-6 augments drug sensitivity in lung cancer cells,\textsuperscript{30} we first determined which conditions could induce relatively lower, but still traceable, levels of IL-6 release by A549-PacR cell secretomes. As a result, the levels of IL-6 were found to be three- to eightfold higher in both the hypoxic and serum starvation conditions compared with normal culture conditions when cells were stimulated with trypsin (Figure 1b). On the basis of these findings, the subsequent cytokine production was determined under normal culture conditions. The production levels of IFN-λ1, IFN-λ2, IFN-α, IFN-β and TNF-α were primarily characterized under unstimulated or trypsin-stimulated conditions and variations...
were found in the trypsin-stimulated concentrations at the 24, 48 and 72 h time points (Figure 1c–e). Further studies were performed to elucidate the regulation of drug (docetaxel)-induced cytokine release (Figure 1f). Drug (docetaxel)-stimulated induction of the levels of IFN-α (Figure 1g), IFN-λ (Figure 1h) and TNF-α (Figure 1i) release was lowered by PacR cell secretomes in A549 cells.

**Figure 1** PacR cancer cell-induced secretomes decreased drug-stimulated release of interferon-α (IFN-α), IFN-λ, and tumor necrosis factor-α (TNF-α) in parental cells. (a) Schematic work flow of preliminary stimulating conditions and preparation of conditioned media (CM) from A549-PacR cells for cytokine profiling used for assays in b–e. (b) Secretion concentration profile of interleukin-6 from the collected PacR CM after time-dependent harvest in the indicated conditions. Trypsin (1.2 μg ml⁻¹) was used for stimulation. Note that the normal (unstimulated) conditions are highlighted and were used as the control c, d and e. (c–e) Time-dependent release of IFN-λ1 (left) and IFN-λ2 (right) (c), IFN-α (left) and IFN-β (right) (d), and TNF-α (e) following trypsin-induced *in vitro* stimulation with unstimulated conditions as the control. The data are presented as the mean ± s.d. of the maximal observed cytokine concentration (pg ml⁻¹). (f) Scheme of A549-PacR CM collection, transfer to and stimulation in adherent A549 cells before cytokine profiling used for assays in g–i. (g) Influence of docetaxel-stimulation in PacR secretome-induced changes in the IFN-α (g), IFN-β (h) and TNF-α (i) release from parental cells. The data are representative of three independent experiments with two biological replicates. The data are presented as the percentage (mean ± s.d.) *P<0.05, **P<0.001 versus the PacR secretome control.

PacR cancer cell-derived secretomes elicited cell outgrowth and escape from TRAIL-induced apoptosis associated with regulation of Akt/FOXO3a signaling in drug-sensitive cancer cells

Our previous study showed that secretomes derived from PacR cells with multiple acquired drug (5-fluourouracil, cisplatin and docetaxel) cross-resistance promoted FOXO3a-directed cell
growth and drug sensitivity of lung and prostate PacR cancer cells. To shift the focus onto drug-sensitive cancer cells, we evaluated the effects of PacR cell secretomes on cell growth and apoptosis of drug-sensitive cells. Secreted soluble mediators from drug-resistant cancer cells foster cell growth and regulate the drug-induced cell death response in sensitive cancer cells that feed from this microenvironment, indicating that acquisition of acquired drug resistance from these secreted...
We then examined TRAIL expression in these cells after exposed to PacR cell secretomes (Supplementary Figure S2). Furthermore, LY294002 induced apoptosis of A549 cells concurrent inhibition of FOXO3a protein expression. LY294002 induced FOXO3a ubiquitination with the FOXO3a in the presence of the Akt inhibitor LY294002. The activation of FOXO3a through Akt regulates TRAIL-targeting signals to affect cell death in drug-resistant cancer cells. Accordingly, targeting strategies that use molecules to modulate FOXO3a are under active investigation. In the present study, we found that PacR-derived cell secretomes upregulated the gene expression of FOXO3a and Akt in both A549 and PC-3 cells (Figure 2h). In addition, to further determine whether Akt controls FOXO3a expression in these cells, we initially identified the ubiquitination status of FOXO3a in the presence of the Akt inhibitor LY294002. LY294002 induced FOXO3a ubiquitination with the concurrent inhibition of FOXO3a protein expression. Furthermore, LY294002 induced apoptosis of A549 cells exposed to PacR cell secretomes (Supplementary Figure S2). We then examined TRAIL expression in these cells after treatment with apoptosis-inducing concentrations of paclitaxel and docetaxel. Exposure to PacR cell secretomes led to decreased drug-induced TRAIL gene expression in both A549 and PC-3 cells (Figure 2i). These results suggest that protection from drug-induced apoptosis mediated by TRAIL correlates with the acquired resistance-promoting effects of the PacR secretomes on drug-sensitive parental cells, which may involve Akt/FOXO3a. These findings are consistent with the previously reported increased expression and nuclear accumulation of FOXO3a despite enhanced PI3K/Akt activity in drug-resistant K562 cells.

Glucose-deprived PacR cell-derived secretomes altered their cell growth-promoting activity via FOXO3a-mediated ATP depletion in drug-sensitive cancer cells
The acquisition of drug-resistant cancer cell phenotypes has been associated with increased dependence on glucose for survival and stress defense. Therefore, we investigated the underlying mechanisms of the protective effects of PacR cell secretomes against drug-induced apoptosis in A549 and PC-3 cells. To this end, glucose metabolism (withdrawal or supplementation) was modified in PacR cells before the CM harvest and exposure to parental cancer cells (Scheme in Figure 3a). We also employed FOXO3a RNA interference to observe the regulatory mechanism of FOXO3a in Akt-induced glycolysis. The selected stealth siRNA for FOXO3a effectively suppressed the expression of FOXO3a messenger RNA and protein levels (Figure 3b). Glucose-deprived PacR cell-derived secretomes substantially altered the effect of secretomes on growth promotion, as observed by the time-dependent inhibition of cell growth, which was similar when FOXO3a transcription was suppressed both in A549 and PC-3 cells (Figure 3c). The susceptibility of tumor cells to cell death is known to be regulated by glucose metabolism and ATP coordination. Glucose withdrawal can lead to ATP depletion, possibly leading to apoptosis. On the basis of this hypothesis, we first determined whether glucose-deprived PacR cell secretomes modified the ATP production in parental cells. Glucose-deprived PacR cell secretomes depleted ATP production in a time-dependent manner in both A549 and PC-3 cells when the...
levels were normalized with those of the glucose-supplemented cells (Figure 3d), whereas ATP levels were totally inhibited when normalized to those of control (unmodified) cells (data not shown). We also found that the transient silencing of FOXO3a caused further depletion of ATP in both A549 and PC-3 cells after exposure to glucose-deprived PacR secretomes (Figure 3e), suggesting that ATP depletion is highly correlated with cell growth inhibition or, possibly, apoptosis. The results of experiments with drug-sensitive parental cells that were exposed to unmodified control PacR secretomes are presented in Supplementary Figure S3.

**Glucose modification in PacR cancer cells promoted the secretion of soluble factors that direct cell death via FOXO3a in drug-sensitive cancer cells**

By modifying glucose utilization in cancer cells, several secreted factors have been reported to promote the metastatic niche and protection from cell death-inducing molecules. To identify whether glucose modification in PacR cells can regulate the secretion of factors that mediate cell death in drug-sensitive cancer cells and determine the involvement of FOXO3a, FOXO3a RNA interference was employed to detect the effect on cell viability. Exposure to glucose-deprived PacR secretomes led to an increase in the apoptotic sub-G1 phase fraction with the deletion of FOXO3a, indicating the likelihood of paclitaxel-induced apoptosis (Figure 4a). In addition, exposure to glucose-deprived PacR cell secretomes also led to increased caspase-3 and caspase-9 activity with FOXO3a deletion, exacerbating the incidence of apoptosis in drug-sensitive cancer cells (Figure 4b). This finding suggests the probable involvement of additional cell death signals regulating this effect. Glucose-deprived PacR cell secretomes led to minimally induced PARP cleavage in A549 cells, and FOXO3a suppression did not seem to affect this outcome, even..
with paclitaxel stimulation (Figure 4c). Moreover, FOXO3a regulated the induced downregulation of Akt, and activation of Bak and Bax in A549 cells upon exposure to glucose-deprived PacR cell secretomes (Figure 4d). This result provides evidence that the apoptosis-promoting effect of glucose-deprived PacR cell secretomes in drug-sensitive cells may be mediated, at least in part, by FOXO3a.

**Regulation of apoptosis in PacR cells with docetaxel cross-resistance was mediated by glucose-modified cell secretion factors via FOXO3a**

To examine the mediators of apoptosis in relation to the acquisition of cross-resistance to docetaxel in PacR cells, we employed our established PacR/DCT cell lines derived from PacR variants. PacR/DCT cells have transient cross-resistance to docetaxel yet retain substantial resistance to paclitaxel. We assessed the effects of glucose modifications in PacR cell secretomes on adherent parental cancer cells based on high P-gp/ABCB1 dependence of acquired cross-resistance. We also examined the role of FOXO3a in apoptosis in these resistant variants. Active utilization of glucose from the microenvironment by drug-resistant tumors has been previously reported. Moreover, side populations of tumor cells with acquired resistance are also described to have elevated glycolytic activity, indicating higher glucose uptake and lower oxygen consumption, which increases the intensity of...
drug resistance. This information prompted us to compare glucose-enriched (25 mM glucose+) and glucose-deprived PacR cell secretomes and their effects on P-gp/ABCB1-associated docetaxel cross-resistance and apoptosis (Scheme on upper panel Figure 5a). P-gp distribution in the membrane and cytoplasm of A549-PacR/DCT cells was subdued in response to exposure to glucose-deprived PacR cell secretomes, whereas cells exposed to glucose-enriched PacR cell secretomes retained their P-gp distribution (Figure 5a). Furthermore, the inhibition of P-gp was apparent in A549-PacR/DCT cells after they were

Figure 5 Targeted glycolysis regulates P-glycoprotein (P-gp)/ABCB1-associated acquired drug resistance and apoptosis in cancer cells grown under exposure to PacR cancer cell-induced secretomes. (a) P-gp (green) distribution in A549-PacR/DCT cells after exposure to glucose-modified PacR cell secretomes. P-gp localization was observed by confocal microscopy. The nuclei were stained with 4,6-diamidino-2-phenylindole (DAPI; blue). Images are representative of three independent experiments. Images were scaled at 40 μm. (b) P-gp protein expression of A549-PacR/DCT cells after exposure to glucose-modified PacR cell secretomes following subsequent RNA interference (RNAi) experiments and TRAIL (400 ng ml⁻¹) treatment for 24 h. β-Actin was used as the loading control. (c) Apoptotic live-cell monitoring coupled with PI incorporation into dead/dying cells after A549-PacR/DCT cells were exposed to glucose-modified PacR cell secretomes following subsequent RNAi experiments and TRAIL (300 ng ml⁻¹) treatment for 24 h. The percent of PI incorporation was monitored time-dependently after transfection and treatment procedures. Non-overlapping PI-stained cells were counted according to the indicated time intervals, and the total number of PI-positive nuclei was scored. All data were normalized to the first time point (t=0) for convenient comparisons. The data are presented as the percentage (mean ± s.d.). (d) Messenger RNA (mRNA) gene expression of ABCB1 (MDR1), NF-κB, survivin and XIAP in A549-PacR/DCT cells after exposure to glucose-modified PacR secretomes following subsequent RNAi experiments and TRAIL (400 ng ml⁻¹) treatment for 24 h. GAPDH mRNA levels were used for normalization. The data are represented as the mean ± s.e.m. siRNA, short interfering RNA.
exposed to glucose-deprived PacR cell secretomes with or without TRAIL treatment, whereas glucose enrichment led to a maintained expression except after RNA interference transfection. Transfection with FOXO3a siRNA intensified this suppression compared with that of control cells (Figure 5b).

To correlate this finding with apoptosis, we exposed both A549-PacR/DCT and PC-3-PacR/DCT cells to glucose-enriched or glucose-deprived PacR cell secretomes and

Figure 6 Targeted glycolysis regulates PacR cancer cell-induced secretome-mediated promotion of docetaxel cross-resistance in PacR/DCT cells through FOXO3a. (a) Sensitivity screen of docetaxel in parental, PacR and PacR/DCT phenotypes of A549 and PC-3 cells after exposure to the non (control)- or glucose-deprived PacR cell secretome. Docetaxel half-maximal inhibitory concentration (IC₅₀) was identified by testing five different concentrations of the drug in a 48 h treatment window. Cell viability was assessed by MTT assay. The data are presented as percentages (mean ± s.d.). (b) Relative resistance/cross-resistance of docetaxel in PacR and PacR/DCT variants of A549 and PC-3 cells were calculated by dividing docetaxel IC₅₀ values in PacR and PacR/DCT variants by parental cell values. (c) Protein expression of P-glycoprotein (P-gp), p-Akt, Akt, p-FOXO3a, FOXO3a, p27/Kip1, p-IκB and IκB in the PacR/DCT phenotype of A549 and PC-3 cells after exposure to the control or glucose-deprived PacR cell secretomes following subsequent RNA interference (RNAi) experiments. (d) Phospho-AMPK-α expression in A549-PacR/DCT and PC-3-PacR/DCT cells after exposure to PacR cell secretomes for 24 h following subsequent glucose modification (+, 25 mM glucose in medium; −, glucose-depleted medium), RNAi experiments and docetaxel (40 nM) treatment in cultures of adherent cells. (e) Intracellular ATP luminescence intensities of A549-PacR/DCT cells after exposure to the control or glucose-deprived PacR cell secretomes following subsequent RNAi experiments and docetaxel (60 nM) treatment for 24 h. The data are presented as the percentage (mean ± s.d.). The results are representative of two (a) and three (b-e) independent experiments. *P<0.05, **P<0.001 versus control. siRNA, short interfering RNA.
transfected the cells with FOX3a siRNA. PI can fluorescently label the nuclei of necrotic or late-stage apoptotic cells because PI is normally membrane impermeable. Therefore, it has been shown that the increase in the number of PI-stained nuclei over time can be used as a metric for cell death. Images of live cells were acquired every 0.5 h over a total of 12 h, and the total number of PI-positive nuclei was determined and normalized to 1 (time; t = 0). In A549-PacR/DCT and PC-3-PacR/DCT cells exposed to glucose-enriched PacR cell secretomes, minimal differences were observed between the control and the FOXO3a siRNA-transfected cells over time; this finding was also consistent with the TRAIL-stimulated group (upper panels of Figure 5c and Supplementary Figure S4). However, glucose-deprived PacR cell secretomes induced higher levels of apoptotic cells, and significant differences can be observed between the control and FOXO3a siRNA-transfected cells: TRAIL-stimulated, FOXO3a siRNA-transfected cells showed relatively higher levels of apoptosis than did TRAIL-stimulated control cells (lower panels of Figure 5c; Supplementary Figure S4). To confirm these observed differences in cell death, we measured the gene expression levels of cell death-related signals and ABCB1 (MDR1) under the same conditions in A549-PacR/DCT cells. Exposure to glucose-deprived PacR cell secretomes inhibited the gene expression levels of ABCB1, NF-κB, survivin and XIAP, and further regulated their TRAIL-mediated levels in A549-PacR/DCT cells compared with those observed after exposure to glucose-enriched PacR cell secretomes, the deletion of FOXO3a intensified this inhibitory effect (Figure 5d). These results indicated that the knockdown of FOXO3a effectively mediated the induction of cell death, even after exposure to both types of glucose-modified PacR cell secretomes.

Targeted glycolysis involved FOXO3a/Akt in the regulation of docetaxel cross-resistance in PacR cancer cells induced by their secretomes

Modifications in metabolic energy produced by glycolysis are essential contributors to the progression of multidrug resistance in cancer cells. The deregulation of signals responsive to GD in drug-resistant cancer cells has been reported previously and has been shown to counteract drug-induced toxicity. In addition, transcriptome analysis of these signals in drug-sensitive cells in response to treatment-induced tumor secretomes has also been described. Therefore, we next determined whether the cell secretomes derived from A549-PacR and PC-3-PacR variants mediate the sensitivity to docetaxel in both PacR and PacR/DCT cells. Interestingly, we found that glucose modification in these cell secretomes affected the sensitivity to docetaxel in both PacR and PacR/DCT cells. In addition, FOXO3a siRNA enhanced the sensitivity to docetaxel in the same experimental conditions (Figure 6a). Relative acquired resistance to docetaxel was calculated based on the half-maximal inhibitory concentration levels, which further confirmed the docetaxel sensitivity results obtained from cells exposed to the secretomes of PacR cells (Figure 6b). Similarly, in our FOXO3a-based characterization
of the cell growth response to PacR cell secretomes with modified glucose content (that is, enriched with 25 mM glucose or deprived), we found significant changes in the growth rates (48 h cell viability normalized to 24 h) of parental, PacR and PacR/DCT variants in response to PacR secretomes and subsequent FOXO3a siRNA experiments (Table 2). This result suggested that targeting glycolysis in PacR cells prevents the secretion of factors that promote cross-resistance in drug-resistant cancer cells, which effectively addresses how multiplicity in drug resistance is primarily acquired in drug-resistant cancer cells. To support this claim, we analyzed the protein expression levels of P-gp and FOXO3a/Akt signaling in PacR/DCT cells after exposure to either control (normal glucose) or glucose-deprived PacR cell secretomes. Deprivation of glucose in PacR cell secretomes downregulated the expression levels of P-gp, phospho-Akt, phospho-FOXO3a and IκBα, with minimal changes in p27Kip1. These effects were augmented by the suppression of FOXO3a transcription (Figure 6c). AMP-activated protein kinase (AMPK) has been implicated in the regulation of glycolysis in several cancer types.54 More specifically, the phosphorylation of AMPK-α is involved in glucose-dependent Akt activity, possibly through the suppression of AMPK and the activation of Akt by glucose,55 affecting ABCG2 expression;48 ABCG2 is a member of the ATP-binding cassette family known to mediate drug efflux pumps. We found differential expression of drug-induced phospho-AMPK-α in A549-PacR/DCT and PC-3-PacR/DCT cells after transient transfection with FOXO3a siRNA under glucose deprivation; the expression was inhibited in A549-PacR/DCT cells but upregulated in PC-3-PacR/DCT cells. This differential expression might be associated with the wild-type status of LKB1 in the parental A549 (LKB1−/−) and PC-3 (LKB1+/+) cells, which differentially exhibit homozygous inactivating mutations of endogenous LKB1, a regulator of AMPK activation and phosphorylation.36,37 Nevertheless, docetaxel induced phospho-AMPK-α activation in both A549-PacR/DCT and PC-3-PacR/DCT cells after silencing of FOXO3a and cell exposure to glucose-deprived PacR cell secretomes (Figure 6d). Moreover, we found enhancement of the docetaxel-induced depletion of ATP by silencing FOXO3a in A549-PacR/DCT cells after exposure to both control and glucose-deprived PacR cell secretomes (Figure 6e). These results are consistent with those of previous reports that shown defective ATP production upon glucose content modification in drug-resistant cancer cells.58,59

**DISCUSSION**

Paclitaxel-based therapies, alone or in combination with kinase inhibitors such as erlotinib, carboplatin and flavopiridol, have shown significant clinical efficacy in patients with advanced non-small cell lung cancer or prostate cancer.60–62 Although adverse events are sometimes observed, they do not occur as commonly as they do when paclitaxel is used alone. Nonetheless, taxane-based therapies have been shown to have acceptable tolerability and have led to improvements in the duration and quality of life for most of these patients.63,64 However, drug resistance inevitably develops with progressive disease, representing a major obstacle for improving overall survival. Paclitaxel-resistant cancer types have been reported to develop more rapid cross-resistance to a variety of unrelated drugs.65–67 However, the development of rapid multiplicity and reaction to changes in the microenvironment during tumor progression for these paclitaxel-resistant tumor populations in acquired cross-resistance are not yet well understood.

Previously, we reported that microtubule stability and ABCB1 could be regulated by FOXO3a to induce multiplicity in acquired cross-resistance to unrelated drugs such as 5-fluorouracil and cisplatin in paclitaxel-resistant lung and prostate cancer cells. This function may augment the release of drug resistance-promoting secreted factors.31 In a more physiologically relevant context, this function of FOXO3a has been well described with the specific role of therapy-induced cancer cell secretomes (in the form of CM) in promoting resistance and progression in cancer cells exposed to these secretomes both in vitro and in vivo.32,33 A better
understanding of how targeted glycolysis mediates this process could lead to the improved efficacy of current targeted therapies.

To model how secretomes derived from cancer cells with high-fold resistance to paclitaxel affect drug-sensitive cells, we employed the established PacR cancer cell lines (A549-PacR and PC-3-PacR), which were developed over the course of >12 months and were 200 times more resistant to paclitaxel. We collected the CM of these PacR cells for 48 h and cultured the drug-sensitive A549 and PC-3 cells with these cell secretomes under normal culture conditions. We found that the PacR cell-derived secretomes significantly contribute to cell growth and consistently limit the efficacy of drugs, such as paclitaxel, docetaxel and cisplatin. Mechanistic evaluation clearly demonstrates that FOXO3a/Akt signaling triggers the ability of these PacR cell secretomes to mediate their effects on drug-sensitive cells, indicating a tight link between FOXO3a and acquired drug resistance. This finding was further extended to glucose modification in PacR cells. We found that GD prevents the secretion of complex factors by PacR cells that promote cell proliferation, escape from apoptosis and drug resistance in drug-sensitive cancer cells. Consistent with these findings, the enrichment of glucose in PacR cells either maintained or enhanced the cell growth-promoting effects of the PacR cell secretomes, providing complementary evidence that targeting glycolysis in PacR cancer cells mediates the secretion of drug resistance-promoting factors.

FOXO3a has been observed to amplify PI3K/Akt signaling in drug-resistant cancer cells, which are then characterized with elevated FOXO3a expression and activity.15 FOXO3a is recruited to the ABCB1 promoter regulating ABCB1 expression in response to chemotherapy drugs.14 Indeed, we found that the suppression of FOXO3a inhibited P-gp/ABC1 expression in PacR cancer cells exposed to PacR cell secretomes, which underwent either glucose enrichment or GD. Under both circumstances, FOXO3a mediated the effects of the secretomes on docetaxel drug sensitivity and escape from apoptosis, as demonstrated by the regulated expression of cell death markers and TRAIL-induced regulation. FOXO3a-driven glycolysis has been closely associated with downstream signals of Akt through tuberous sclerosis complex 1 (TSC1).44 Hyperactivation of PI3K/Akt activity has been described as a consequence of chronic induction of FOXO3a in multidrug-resistant leukemic cells.15 However, the exact mechanisms by which FOXO3a might be able to direct Akt through TSC1 to regulate the response of drug-sensitive cancer cells to PacR secretomes remain unclear and must be further investigated.

To further determine whether the promotion of docetaxel cross-resistance in PacR cells is evident, we employed the established PacR/DT cell lines, which were developed to have transient cross-resistance to docetaxel and a retained high-fold resistance to paclitaxel. By depriving PacR cells of glucose, the secreted factors failed to augment docetaxel resistance, which was amplified by silencing FOXO3a. These findings highlight an element of complexity associated with the mechanisms that drive the development of cross-resistance in PacR cancer cells; nevertheless, these findings further confirm that drug-resistant cancer cell secretomes promote resistance and outgrowth in drug-sensitive cells and confirm that targeted glycolysis can amend the process.

Further studies were performed to reveal these secreted factors of PacR-derived cell secretomes that cause the permissive stimulation of outgrowth and support the survival of drug-sensitive cancer cells. The limited efficacy of some drugs has been attributed to specific cytokines that support cell growth and the acquisition of aggressive phenotypes that promote the survival of drug-sensitive cancer cells. We found that the drug-stimulated levels of IFN-α, IFN-γ and TNF-α were alleviated in cancer cells by treatment with PacR cell secretomes, suggesting that PacR cell secretomes stimulate cell growth and resistance in drug-sensitive cancer cells (Figure 7).

In summary, the present findings reveal that PacR cancer cells secrete tumor growth-promoting factors that stimulate regulatory signals to shift apoptosis and develop acquired drug resistance in drug-sensitive cancer cells and cross-resistance in PacR cancer cells. It is especially notable that these findings support, at the basic level, how we can further regulate the effects of therapy-induced secretomes by targeting glycolysis, which appeared to be mediated by FOXO3a signaling. Considering that paclitaxel-based treatments are carefully being clinically evaluated, these findings reveal an approach to strategically improve the efficacy of therapies involving paclitaxel for successfully overcoming acquired drug resistance.

CONFLICT OF INTEREST
The authors declare no conflict of interest.

ACKNOWLEDGEMENTS

We thank Jayoung Song for the development and culture maintenance of the PC-3 and PC-3-PacR cell lines. This work was supported by a grant from procurement and development of biological resources funded by the Ministry of Education Science and Technology of the Korean government (2011-00499) and a National Research Foundation of Korea (NRF) grant funded by the Korean Government (MEST; no. 2009-008533).

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Supplementary Information accompanies the paper on Experimental & Molecular Medicine website (http://www.nature.com/emm)