An ultraportable and versatile point-of-care DNA testing platform

Huan Xu1, Anyue Xia2, Dandan Wang3, Yiheng Zhang4, Shaoli Deng5, Weiping Li5, Jie Luo6, Qiu Zhong5, Fengling Zhang5, Lin Zhou1, Wenqing Zhang1, Yang Wang1, Cheng Yang1, Kai Chang1, Weiling Fu1, Jinhui Cui3*, Mingze Gan3, Dan Luo6,9*, Ming Chen1,10,11*

Point-of-care testing (POCT) has broad applications in resource-limited settings. Here, a POCT platform termed POCKET (point-of-care kit for the entire test) is demonstrated that is ultraportable and versatile for analyzing multiple types of DNA in different fields in a sample-to-answer manner. The POCKET is less than 100 g and smaller than 25 cm in length. The kit consists of an integrated chip (i-chip) and a foldable box (f-box). The i-chip integrates the sample preparation with a previously unidentified, triple signal amplification. The f-box uses a smartphone as a heater, a signal detector, and a result readout. We detected different types of DNA from clinics to environment to food to agriculture. The detection is sensitive (<10^9 copies/ml), specific (single-base differentiation), speedy (<2 hours), and stable (>10 weeks shelf life). This inexpensive, ultraportable POCKET platform may become a versatile sample-to-answer platform for clinical diagnostics, food safety, agricultural protection, and environmental monitoring.

INTRODUCTION

Laboratory-based detection methods such as polymerase chain reaction (PCR) are typically unavailable in resource-limited settings, where sophisticated infrastructures, reliable electricity supplies, and trained operators are lacking. Point-of-care testing (POCT), on the other hand, permits rapid on-site detections in these locations. Consequently, any form of molecular detection including DNA analyses requires POCT (1–2).

Over the last 5 years, POC DNA analysis research has progressed rapidly not only in clinical cares but also in agriculture, environmental protection, and food safety (3–6). Two keys to the success of practical applications are the portability and versatility of POCT. For the portability issue, one of the greatest challenges has been to ensure the portability throughout the entire process, from sample preparation to signal amplification to readouts without additional instruments. Many recent efforts have been dedicated to improving the portability of the signal amplification step (7–8). For example, paper-based DNA detection methods, such as lateral flow assays, offered the portability for DNA binding and fixing (onto thin strips of papers), but still requiring additional instruments such as a centrifuge for sample preparation and a thermal incubator or thermal cyclers for enzyme reactions. In addition, most instruments required an alternating current (AC) power supply. The need for instrument in the sample preparation stage and/or AC power supplies have severely limited the portability of the entire POCT. Although various methods without using an AC power such as using exothermic chemical reactions or positive temperature coefficient heaters had been developed to achieve isothermal amplification (9–12), those methods still weighted from hundreds to thousands of grams, too heavy to be ultraportable.

Versatility is another great challenge for POCT. The ideal DNA POCT system needs not only to be able to process diverse types of samples (such as saliva, urine, and blood) but also to be able to detect the multiple forms of DNA targets including pathogen-specific DNA sequences, gene mutations, single-nucleotide polymorphisms (SNPs), and allelic genes (13–14). So far, it is still difficult to detect pathogen-specific DNA directly from raw samples (e.g., saliva) in the fashion of "sample-in-result-out" (15). Considering the requirement of more accurate detection of other DNA target forms and compatibility with other biological samples (whole blood, etc.) containing more interferents, it is still quite challenging to develop a versatile POCT platform for the detection of multiple forms of DNA targets from diverse types of samples in the sample-to-answer manner.

In this study, we created a point-of-care kit for the entire test, the POCKET, as an ultraportable and versatile POCT platform. The POCKET platform was ultraportable: less than 100 g in weight and smaller than 25 cm in length, fitting into a quart-sized plastic bag or a regular-sized envelope (B5 size, 17 cm × 25 cm). The POCKET was ultraportable for the entire test process, from sample preparation to signal amplification to final readouts without any instrument. The POCKET was also versatile, detecting different forms of DNA from a variety of sample sources, from clinics (blood, buccal, and urine) to food (milk) to environment (river water) and to agriculture (plant leaves).

The POCKET was made of only two components: an integrated chip (i-chip), for both sample preparation and signal amplification, and a foldable detection box (f-box), using a smartphone as a heater, an incubator, a detector, and a result reader. The i-chip was smaller than fingertip and was fabricated using three-dimensional (3D) printing.
and microfabrication. The f-box was foldable for size reduction. The entire POCKET platform fitted within a quart-sized plastic bag or a regular-sized envelope (Fig. 1A). Furthermore, a previously unidentified heat-generating approach was created using only a smartphone; combined with the f-box, a smartphone-based isothermal incubator was realized without any conventional heaters, incubators, and AC power supplies (Fig. 1B, right). For the signal amplification, we developed a previously unidentified triple amplification method that combined an isothermal recombinase polymerase (iRP) amplification along with a further Au-silver complexes for a total of triple amplifications (termed iRPAS) (Fig. 1B, left top). The final amplified signals were colorimetric and were detected by the smartphone using the f-box (Fig. 1C). Thus, we achieved ultraportable from sample preparation to signal amplification to final readouts without any instrument.

Besides the ultraportability, the POCKET platform was versatile, capable of analyses of multiple types of DNA from a broad range of sources. First, the i-chip had Lego-type connectors 3D printed, rendering the POCKET platform readily expandable for the parallel detections of multiple samples. Second, the entire POCKET platform was designed to be generally usable for a variety types of DNA using samples from different sources. In other words, the POCKET was designed and constructed independent of what types of DNA were used. We successfully detected various types of DNA from clinical blood and buccal samples including gene mutations (long or short fragment deletions), SNPs (single-base substitution or insertion), and allele genes. We also successfully detected pathogen-specific DNA sequences from urine samples (Fig. 1D). The sensitivity and specificity of the detections were verified with the conventional clinical methods currently performed in a centralized laboratory.

In addition to real clinical samples, with the POCKET platform, we successfully detected the following specific DNA: bacteria in milk, river water, and on plant leaves, further demonstrating the versatility of the POCKET platform in sample sources.

RESULTS

Design strategy of the POCKET platform

The POCKET platform consisted of two major components: an i-chip and an f-box. The i-chip, only 4 g in weight, integrated two units: a 3D-printed sample preparation unit and a microfluidic signal amplification unit (Fig. 2A). For the 3D-printed sample preparation unit, we incorporated three modules: a basket, a rotary valve, and a block module. We designed an extraction disc at the bottom of the basket module for binding DNA from a broad range of sources (Fig. 2A, dotted box). The basket was connected to a polytetrafluoroethylene (PTFE) for injecting the reagents into the basket by a syringe. To streamline the delivery of reagents and to avoid using expensive pipetting tools, we developed a tandem reagent tube to preload and deliver reagents automatically. Each reagent was separated by an air gap such that multiple reagents were preloaded in a PTFE tube (fig. S1). The block module contained a wash chamber, which was fit with the basket module for DNA purification, and a reaction chamber, which was fit with the rotary valve for target DNA amplification through recombinase polymerase amplification (RPA). The rotary valve was designed as a hollow cylinder with a pore near the bottom side of the module for the connection with microfluidic signal amplification unit. There was another pore on the reaction chamber of the block module connecting with the microfluidic signal amplification unit through a PTFE tube. The open and close states of the microfluidic amplification unit connection were controlled by rotating the rotary valve.
The microfluidic amplification unit consisted of a microfluidic channel layer and an aldehyde-functionalized glass chip, designed for visual readouts via a smartphone (Fig. 2B). The glass chip was modified with multiple DNA capture probes, which were complementary to their targets. To fix capture probes on the glass chip, we first bound a polydimethylsiloxane (PDMS)–made probe channel layer that contained multiple channels onto the aldehyde-functionalized glass. Then, we infused each channel with one type of amino-functionalized capture probe. The probes were bond on the glass surface through aldimine condensation. Next, we peeled off the probe channel layer and assembled the microfluidic channel layer on top of the glass chip to complete the amplification unit (Fig. 2B, bottom, and fig. S2). The surface density of the capture probe was critical for the function of sensing layer (16). We optimized the capture probe concentrations and found that 6 μM was ideal to give a reliable result with approximately 10-fold signal increase (fig. S3A).

To amplify signals to such an extent that they could be visualized by naked eyes or a smartphone, we developed a previously unidentified iRPAS signal triple amplification method (Fig. 2C). Briefly, DNA was first amplified through RPA by tail-forward primers and biotinylated DNA was absorbed and extracted using the extraction disc at the bottom of the basket. Target DNA was purified using a wash buffer and amplified by RPA to obtain biotin-modified amplicons. The amplicons were bound to SA-AuNPs and captured by premodified capture probes to achieve a secondary signal amplification. The tertiary amplification was initiated by depositing silver ions onto the AuNPs. The POCKET platform was scalable for parallel sample detections by i-chips assembly (D). The f-box, which was designed to display the results. This included a foldable cradle, a smartphone, an optical unit with a macro-lens, and a LED lamp. Photo credit (D): Huan Xu, Army Medical University.
reverse primers for 20 min, resulting in biotin-modified, tailed amplicons. The reaction was transferred to the channel of microfluidic unit, and the biotin-modified amplicons were captured by the previously mentioned capture probes. The second amplification was through the affinity binding of biotin with the streptavidin-conjugated gold nanoparticles (SA-AuNPs), and the hybridization time was optimized to 15 min (fig. S3B). The third amplification was completed through a silver ion catalytic cascade that resulted in more and more silver ions deposited on the AuNPs. After this IRPAS signal triple amplifications, the signal was visualized by the naked eyes or semiquantified by a smartphone. The entire operation procedure for user was simple including five steps, and the operation time was short, less than 80 min (fig. S4).

Notably, the entire i-chip was designed with a Lego-like mortise and tenon connections for simple assembly and to allow multiple sample analyses in parallel (Fig. 2D). Thus, the POCKET platform was readily scalable for multiple DNA analyses.

Most enzyme-based DNA detection methods such as PCR and loop-mediated isothermal amplification (LAMP) require a bulky thermal cycler or an incubator. Not surprisingly, current POC systems still require an additional heat supply, such as an incubator or an exothermic chemical package (11, 17, 18). Here, to make our POCKET ultraportable without any additional instrument, we took the advantage of the heat generated by a smartphone and deliberately used that heat for maintaining an isothermal incubation at 37°C, the optimized temperature for RPA. In more detail, we programmed a smartphone application (called “heat-up” application) whose algorithms were called for intense central processing unit operations that resulted in a rapid temperature increase of the smartphone. By controlling both the number and timing of the algorithms running, we precisely controlled the temperature of the smartphone (fig. S5A). The heat transfer from the smartphone to the chip was almost instantaneously due to the extremely small nanoliter volume. To prevent heat dissipation in cold environments, we constructed a thermos packet with the aluminum foil to reduce heat loss in cold environments (4°C). We also incorporated a temperature display bar (30°C to 40°C with 2°C interval) that reported internal temperature in real time. Two additional temperature indication discs were also included to record whether the temperature was too cold (<37°C) or too hot (>42°C) (fig. S5B). To prevent smartphone from the biohazard, we put the smartphone into a disposable plastic bag to insulate from clinical samples, while the smartphone can still heat the chip outside the bag. After the detection, the bag can be recycled or disposed. We compared the smartphone-heating effects with and without the disposable plastic bag, and found that the enclosing bag had little impact on the smartphone heating after 20-min smartphone heating (fig. S6).

For signal readouts, we designed an f-box that integrated a smartphone, a smartphone cradle, and an optical unit with a macro-lens and a light-emitting diode (LED) (Fig. 2E). The smartphone cradle was designed to be foldable and fabricated from light-weight cardboard (fig. S7). The manufacturing cost of the disposable part of our platform is less than $4.1, and the reusable part is less than $2.5 (table S1). In comparison to the commercial devices such as GeneXpert IV ($17,000), our platform is very cost-effective.

Multiple types of DNA analyses
To develop the POCKET into a versatile platform, we tested different kinds of DNA including gene mutations (long and short fragment deletions), SNPs (single-base substitutions and insertions), allele genes, and pathogen-specific DNA sequences. To verify the false-negative result caused by poor DNA preparation, we added an inherent gene (human hemoglobin gene, hbb) in blood samples as an internal control. We can now distinguish the negative result between poor DNA preparation and no target: If both the internal control and the target are negative, then it indicates the poor DNA preparation. If the internal control is positive but the target is negative, then it indicates no target. We showed the results of positive, negative, and the poor DNA preparation in fig. S8. For the detection of long fragment deletions, we tested a 19-kb deletion of the Southeast Asia (SEA) α-thalassemia gene. Note that RPA was typical only for the amplification of sequences smaller than 500 base pairs, not for long fragments (19). Nevertheless, with the POCKET system and the designed paired primers (fig. S9A), we detected significant differences between the mutant sample and the healthy control; the results were further validated by DNA sequencing (Fig. 3A).

For the detection of other gene mutations, SNPs, and allele genes, we used terminal mismatch primers using a special kind of nucleic acids, the locked nucleic acids (LNAs). LNA is nucleic acid chemically modified by the addition of a 20-O, 40-C methylene bridge. LNA showed an increased specificity in the previously reported SNP detection (20). In addition, the introduction of mismatches in the primer, especially in the terminal ends, notably inhibited polymerase extension and thus discriminated the wild type and mutant types (fig. S9B) (21). We chose the following diseases for the gene mutations and SNP tests by the POCKET system: (i) β-thalassemia, a worldwide hereditary anemia that contained different types of gene mutations [we detected the CD41 mutation (four-base deletion) (Fig. 3B), the CD17 mutation (single-base substitution) (Fig. 3C), and the CD71 mutation (single-base insertion) (Fig. 3D)], and (i) allele genes (human ALDH2 gene), which correlated strongly with susceptibility to alcohol intoxication. In each case (Fig. 3, E to G), we obtained strong signals in the mutant samples and little to no signal in control samples. The results were further validated by DNA sequencing (Fig. 3, A to G, bottom).

Moreover, for the detection of the pathogen-specific DNA sequences, patient urine samples were successfully tested and diagnosed for two common bacterial urinary tract infections, Escherichia coli and Klebsiella pneumoniae (K. pn), via paired primers against conserved sequence regions (Fig. 3, H and I).

To summarize, the POCKET system showed versatility in detection of all types of DNA with single-base specificity including but not limited to gene mutations. Also, the entire test essentially remained the same among different DNA types; the only major alteration was to design different primers (table S2).

DNA detection in other fields beyond clinical medicine
In clinical medicine, the POCKET platform was very successful for detections of α-thalassemia and β-thalassemia in blood samples, ALDH2 in buccal swabs, and pathogens in urine samples. To further expand the versatility of the POCKET system in other areas, we explored its utility in food safety, agricultural protection, and environmental monitoring (Fig. 3, J to L).

For food safety applications, we mimicked food contamination by spiking 10⁶ copies of E. coli in 1 ml of sterile milk. The POCKET system successfully detected the contamination (Fig. 3) without much alterations for the entire test. For environmental protection, we chose river water spiked with E. coli as our model. Again, even with a vast amount of environmental impurity such as sand and microorganisms,
we successfully detected the $10^4$ copies of spiked *E. coli* in 1 ml of the river water sample (Fig. 3K). For agriculture protection, it had been notoriously difficult for plant sample preparation in POC device due to the presence of strong inhibitory compounds such as lignin, polysaccharides, and phenolics in the sample (22). To more efficiently prepare plant samples, we 3D-printed a miniaturized grinding pestle and mortar (fig. S10). We successfully detected *Pseudomonas syringae* pv. *actinidiae* (*P. sa*), a common fruit tree pathogen, in kiwifruit leaves using our POCKET system (Fig. 3L).

These results confirmed that the POCKET platform was versatile in sample resources, not only in clinical applications but also in food safety, agricultural protection, and environmental monitoring.

**Performance of the POCKET system in the field**

The performance of the POCKET system, from temperature control to comparison of gold standard methods, was evaluated. For the temperature control (e.g., in winter or in summer), we programmed and tested our heat-up application on a smartphone to reach the optimal 37°C in different environmental temperatures (4°, 15°, 25°, or 37°C). In each case, it took no longer than 20 min to reach and maintain 37°C (Fig. 4A). Strong signals were obtained under all tested temperatures (Fig. 4B). We also measured battery performance of the smartphone at each environmental temperature.

The performance parameters of POCKET were evaluated. For the limit of detection (LOD) of the system, we used a serial 10-fold dilution of plasmid DNA (from $1 \times 10^{10}$ to 10 copies/ml) as standard samples. As expected, the detection signal decreased along a sigmoidal function as the logarithm of the target concentration decreased (Fig. 4C). The LOD, defined as “blank using distilled water plus three times standard deviation” (23), was calculated to be 113 copies/ml (Fig. 4D).

![Fig. 3. DNA analysis using the POCKET platform.](http://advances.sciencemag.org/)
We also investigated the shelf life of i-chips without cold storage by keeping a batch of i-chips in a 25°C incubator up to 10 weeks. We then used them for detection once per week. No significant changes in results were observed after 10 weeks (P < 0.05), indicating that the i-chip was stable for at least 10 weeks without cold storage (fig. S11B).

For user friendliness, we compared the detection performance from a trained user group (previously already performed the POCKET at least three times) versus an untrained user group (the first time using the POCKET). No significant differences between the two groups were observed, suggesting that the POCKET system was very easy to perform and very user-friendly (fig. S11C). Furthermore, as a smartphone was the only nondisposable and expensive piece of instrument used in our system, we tested our POCKET performance using different brands of smartphones. The results showed no significant differences among any of three brands tested (fig. S11D).

The performance of the POCKET system was compared with conventional gold standard methods currently used in a centralized laboratory. Toward that end, we collected a total 172 samples including 63 peripheral venous blood samples, 55 DNA samples purified from participants’ blood, 20 buccal swabs, and 34 urine samples. To show the robustness of our DNA testing using the POCKET platform in clinical samples, we tested the clinical samples with common abnormalities. We summarized the abnormal characteristics of the clinical samples including some strong interferences...
performances. That our POCKET system detected multiple targets with excellent PPV was greater than 88.2% (Table 1). These results demonstrated and ALDH2*A. All other specificities were greater than 77.8%, and

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blood cells: 1 to 48,948 cells/ l, 8 samples with abnormal white blood cells: 1 to 47,577 cells/ l, 24 samples with abnormal white blood cells: 1 to 47,577 cells/ l, 8 samples with abnormal epithelial cells: 1 to 295 cells/ l (table S4). All samples were successfully analyzed using the POCKET system. They were also analyzed using standard methods including a thalassemia detection kit (a commercial clinical kit) for DNA and blood samples, DNA sequencing for buccal samples, and bacterial culture for urine samples (Fig. 4, E to G). The POCKET system showed successful detections. The total accuracy of our device for clinical samples is 97.1%, verified by laboratory standard methods: DNA sequencing for blood, buccal, and DNA samples, bacteria culture, and mass spectrometry for urine samples. (table S5).

Further, receiver-operating characteristic (ROC) curves were plotted to determine the area under the curve (AUC) (Fig. 4, H to J). High AUC value over 0.9 was considered excellent accuracy (24). Each AUC in our POCKET detections was more than 0.950. Here, the POCKET platform achieved a higher detection accuracy, comparable to the gold standard methods performed in centralized laboratories.

We established the value of the maximum sensitivity as the cut-off value, which led to a decrease in specificity and positive predictive value (PPV). The POCKET platform still achieved 100% specificity and PPV for the detection of SEA, CD71, E. coli, K. pn, and ALDH2*A. All other specificities were greater than 77.8%, and PPV was greater than 88.2% (Table 1). These results demonstrated that our POCKET system detected multiple targets with excellent performances.

**DISCUSSION**

Here, we established an ultraportable and versatile POCKET platform for DNA detection in nonlaboratory and resource-limited settings. To overcome the challenges from sample preparation to signal amplification to readouts in POCT workflow, we combined 3D printing, microfluidics, iRPAS signal amplification, and a smartphone. By 3D printing, miniaturized device prototypes can freely be fabricated with elaborate 3D structures, providing the high flexibilities in sample preparation and the fluid manipulation (25). Thus, a wide variety types of crude samples can be used in our POCKET platform. In many current POCT devices, including paper-based methods, clinical samples (e.g., blood) still rely on the laboratory-based pretreatments. It is difficult to perform direct, sample-to-answer detections in POC settings (26). We further adopted microfluidics and iRPAS signal amplification, enabling the sensitive, selective, and multiplexed testing on a chip. In addition to that, by producing the colorimetric signals, our POCKET eliminates the need of any optical filters, making the entire platform not only ultraportable and versatile but also more cost-effective, compared with fluorescence-based systems (17). We use a smartphone not only as a readout device for quantitative analyses but also as a heater, permitting the rapid and robust point-of-care DNA amplification even at the cold environment. Several heating methods exist in POCT using chemicals, peltier, and sunlight (9, 11, 17, 18). All of them require extra heating modules, resulting in more system weight and volume. Using a smartphone as a heater and readout device, we greatly reduce the weight and the volume of the whole platform, down to 60 g and the pocket size. As far as we know, the POCKET is the only platform that uses a smartphone itself as a heating supply. Because of our combinatory and integrated approach, the POCKET system was extremely light and small, rendering the POCKET system, to the best of our knowledge, to be one of the smallest and most portable POC DNA testing devices. Compared with other 3D-printed or microfluidic chips, our POCKET system was more versatile for a

| Target   | Resource       | Reference method | N  | Cutoff value | AUC  | 95% CI   | Sensitivity | Specificity | PPV   | NPV   |
|----------|----------------|------------------|----|--------------|------|----------|-------------|-------------|-------|-------|
| SEA      | Peripheral venous blood | Reverse dot blot | 18 | 4.053        | 1.000 | 1,000, 1,000 | 100.0%      | 100.0%      | 100.0% | 100.0% |
| CD41     | Purified DNA and blood | Reverse dot blot | 36 | 4.720        | 0.994 | 0.977, 1.000 | 100.0%      | 90.0%       | 88.9%  | 100.0% |
| CD17     | Purified DNA and blood | Reverse dot blot | 33 | 4.544        | 0.981 | 0.940, 1.000 | 100.0%      | 95.0%       | 92.9%  | 100.0% |
| CD71     | Purified DNA and blood | Reverse dot blot | 31 | 5.070        | 1.000 | 1,000, 1,000 | 100.0%      | 100.0%      | 100.0% | 100.0% |
| E. coli  | Urine          | Bacteria culture | 18 | 2.416        | 1.000 | 1,000, 1,000 | 100.0%      | 100.0%      | 100.0% | 100.0% |
| K. pn    | Urine          | Bacteria culture | 16 | 3.700        | 1.000 | 1,000, 1,000 | 100.0%      | 100.0%      | 100.0% | 100.0% |
| ALDH2*G  | Buccal         | DNA sequence    | 24 | 3.338        | 0.978 | 0.931, 1.000 | 100.0%      | 77.8%       | 88.2%  | 100.0% |
| ALDH2*A  | Buccal         | DNA sequence    | 16 | 3.700        | 1.000 | 1,000, 1,000 | 100.0%      | 100.0%      | 100.0% | 100.0% |
broader range of possible sample sources and achieved multiplex detections with reduced reagent costs (table S6) (6, 9, 11, 14, 17, 27–31).

Beside DNA testing, we are confident that our platform has the potential to detect more types of nucleic acids including but not limited to double-stranded DNA, single-stranded DNA, or RNA, considering that both DNA and RNA can be amplified through recombinase polymerase (14, 19, 32). Moreover, our POCKET platform is extendable to nucleic acids detections in other fields beyond clinical medicine including but not limited to food safety, agriculture protection, and environmental monitoring. The sample sources can be from blood, urine, buccal swabs, milk, river water, leaves, etc.

With two billion smartphones already existing (2016) and even more by 2020 (covering more than 70% of the world population), smartphones are among the most convenient, user-friendly, and powerful interfaces for POC detection in resource-limited settings (33, 34). Smartphones are also most suitable for the signal processing and readouts in a POC setting. On the basis of smartphones’ built-in wireless communication and geolocation capabilities, our POCKET platform makes it possible to real-time locate the distribution of the disease. Furthermore, the POCKET platform can serve as the Internet of Medical Things (IoMT) in DNA analysis to transmit on-site testing results to the centralized hospital or public health officials, providing spatiotemporal disease mapping for further investigation. In addition, end users including patients can receive fast feedbacks on disease control and prevention.

The POCT in this paper is our first version. In future improvements, we will expand the sample types to those that are known to be difficult to prepare, such as sputum and feces (35, 36). We will also further prevent cross-contamination from positive samples. The reagent loading method can also be better controlled using a power-free method (11, 37).

We envision that by prefabricating frequently emphasized targets on our chips, we will provide a series of comprehensive DNA detection panels for each separate application, from clinics to agriculture to environment to food and to drinking water. With further development, we firmly believe that the POCKET platform will become a widely used POCT device for a great number of real-world applications in various fields. We also hope that our POCKET system would become one of the gold standards for future POCT development.

### Construction of the f-box

We made the f-box with a foldable cradle, an optical unit with a white light LED (Supernatural Light, China) and a 60× macro-lens (Baise, China), and a smartphone (Lenovo ZUK Z2, China). The cradle was designed by SolidWorks software and made by a white cardboard (0.4 mm thick). Then, the cardboard was cut to the designed pattern by a laser engraving machine (Shenhui, China). Afterward, we folded the cardboard into the cradle, as the eight folding steps of the assembling process illustrated in fig. S7, and then fixed the cradle by the designed slot. On the upside wall of the cradle, there was an open window for the insertion of the detection unit into the cradle. A 0.3-W white light LED powered by two 1.5-V button batteries was mounted above the detection unit for illumination. From the top window of the cradle, the macro-lens was plug in and aligned with the detection unit. The macro-lens was tightly fixed on the rear camera of the smartphone by a clamp.

### Chip heating by smartphone

We controlled the chip heating by a heat-up application with functions of reading the internal temperature sensor and heating up the smartphone. The temperature was controlled by both the number and the time of the running algorithms. After the temperature reached 37°C, the i-chip was placed on the back of the smartphone for incubation. To avoid the heat loss and save the energy, the smartphone with the i-chip was placed in a thermos package with a temperature display bar (Jieyada, China). Then, the heating effects of the smartphone in the thermos package were tested in the incubator (Bluepard, China) at 4°C, 15°C, 25°C, and 37°C. When testing the performance of smartphone heating in the cold environment (4°C), the smartphone with the i-chip was put in the thermos package and sealed by clips before placing in the incubator. The temperature of i-chip was real-time monitored by an external temperature recorder (Elitech, China).

### Sample collection

The study was approved by the ethics committees of Daping Hospital and Southwest Hospital, Chongqing, China. Purified DNA samples,
buccal samples, and urine samples were collected from 55, 20, and 34 participants, respectively, from Daping Hospital, Chongqing, China. Peripheral venous blood samples from 63 participants were collected from Southwest Hospital, Chongqing, China. Spiked milk samples were mimicked by mixing 1 ml of sterile milk with E. coli [10^4 colony-forming units (CFU)/ml]. Spiked river water samples were prepared by mixing 1 ml of natural river water with 10^5 CFU/ml E. coli. P. sa–infected leaves and Colletotrichum gloeosporioides (C. gl)–infected leaves were obtained from X. Sun in Sichuan Agricultural University, Chengdu, China.

DNA sample preparation
We dipped the basket module into the sample liquid. About 2 μl of sample was absorbed by the extraction paper disc at the bottom. We prepared leaf samples by the extra crushing as following steps. Five milligrams of leaves with 100 μl of lysis buffer (Tiangen, China) was added to the 3D-printed mortar and grinded by the 3D-printed pestle for 5 min, and then 100 μl of neutralizing buffer (Tiangen, China) was added and mixed thoroughly. The extraction paper disc with sample liquid was dried at room temperature for 10 min. Thereafter, the basket module was inserted into the wash chamber (inner diameter, 5.0 mm; height, 6.5 mm) of the sample preparation unit and washed with 200 μl of nuclease-free water, 25 mM deoxynucleotide triphosphate, 6.6 μl of TE buffer (10 mM tris-HCl and 0.1 mM EDTA, pH 8.0) for 5 min. Once washed, the basket module with purified sample DNA on the paper disc was pulled out from the wash chamber and dried at room temperature for 15 min.

RPA amplification
We performed the RPA amplification following the manufacturer’s instructions (TwistDx, UK). For each reaction, 25 μl of TwistDx 2x reaction buffer was mixed with 5 μl of 10x basic E-mix, 3.6 μl of 25 mM deoxynucleotide triphosphate, 6.6 μl of nuclease-free water, and 2.5 μl of core mix. Next, 4.8 μl of primer mix, containing biotinylated reverse primer and tagged forward primer, was added to the RPA reaction reagent. Premixed RPA reaction reagent (47.5 μl) and 2.5 μl of magnesium acetate were successively infused into a PTFE tube (1.0 mm in inner diameter and 65 cm in length, Titan, China). Each reagent was separated by an air gap (over 5 cm in length) to form an isolated tandem reagent tube and stored at 4°C until required. At the start of amplification, the reagents in the isolated tandem reagent tube were injected into the reaction chamber (6.9 mm in inner diameter and 6.5 mm in depth) in the sample preparation unit by pushing air with a 1-ml syringe connecting to the tandem tube. The reaction was kept at 37°C for 20 min by the smartphone running with the heat-up application. All primers, amino-functionalized DNA probes, and LNA-modified probes (Zhanshan, China) were described in table S2. Plasmid (Tsingke, China) was described in fig. S12.

AuNP-silver amplification
After RPA amplification, we injected 50 μl of hybridization buffer (5× Denhardt’s solution, 3× saline sodium citrate, and 0.1% bovine serum albumin) containing 1% streptavidin-conjugated AuNPs (Nanoprobes, USA) into the reaction chamber to mix with the reacted RPA solution. Then, we opened rotary valve and injected the mixed solution into the microfluidic amplification unit through the connecting PTFE tube by pushing air with a 1-ml syringe. After that, the chip was incubated for 15 min by heating with the smartphone. The PDMS layer was then peeled off from the amplification unit, washed with water, and then stained with 20 μl of silver enhance buffer (Sigma, USA) for 7 min. Last, after iRPAS signal triple amplification, the microfluidic amplification unit was imaged with a smartphone installed with the POCKET software application for signal readout.

POCKET software
The smartphone application “POCKET version 1.0” was designed to image capture and image process the input of basic sample information including sample barcode, location, and the upload of results to the cloud database. Briefly, the image was captured and converted to the 8-bit grayscale format and calculated its grayscale value (black as 0 and white as 255) within the regions of interest for the sample, reference, and background (fig. S13). First, the mean grayscale value of sample and reference were obtained by subtracting the background value. Next, the relative fold change was obtained by dividing the mean grayscale value of the sample with the reference. Last, the relative fold changes of samples were recorded and compared with the threshold to determine the positive or negative result. Here, the threshold was designated as the cutoff value for each target.

Detection verification
All purified DNA samples and peripheral venous blood samples were initially diagnosed using a thalassemia diagnostic kit (Hybribio, China) by reverse dot blot method and further verified by DNA sequencing (Invitrogen, China). For the verification of allele genes, the ALDH2 gene from buccal swabs was detected by DNA sequencing (Invitrogen, China). Bacteria in urine samples were cultured on the blood agar medium plate (Pangtong, China) and then diagnosed by the matrix-assisted laser desorption ionization–time of flight mass spectrometry (bioMétrieux, France).

Data analysis and statistics
Statistical analysis was performed using GraphPad Prism 7.0 software (GraphPad Software, USA) and SPSS version 19.0 software (SPSS, USA). The data were normalized and expressed as mean ± SD. Independent sample two-tailed Student’s t tests were performed for comparisons between groups. Multiple-group statistical analyses were performed by one-way analysis of variance (ANOVA), with the least significant difference used for post hoc testing. P values <0.05 were regarded as statistically significant. Detection performance of the POCKET platform was calculated using area under ROC curves. As a screening test, cutoff values were selected on the basis of minimized false negatives, defined as the maximum sensitivity, taken at all points on the ROC curve.

SUPPLEMENTARY MATERIALS
Supplementary material for this article is available at http://advances.sciencemag.org/cgi/content/full/6/17/eaaz7445/DC1 View/request a protocol for this paper from Bio-protocol.

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