Smoking and COX-2 Functional Polymorphisms Interact to Increase the Risk of Gastric Cardia Adenocarcinoma in Chinese Population

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Abstract

Background: Over-expression and increased activity of cyclooxygenase (COX)-2 induced by smoking has been implicated in the development of cancer. This study aimed to explore the interaction between smoking and functional polymorphisms of COX-2 in modulation of gastric cardia adenocarcinoma (GCA) risk.

Methods and Findings: Three COX-2 polymorphisms, including –1195G>A (rs689466), –765G>C (rs20417), and 587Gly>Arg (rs3218625), were genotyped in 357 GCA patients and 985 controls. In the multivariate logistic regression analysis, we found that the –1195AA, –765GC, and 587Arg/Arg genotypes were associated with increased risk of GCA (OR = 1.50, 95% CI = 1.05–2.13; OR = 2.06, 95% CI = 1.29–3.29 and OR = 1.67, 95% CI = 1.04–2.66, respectively). Haplotype association analysis showed that compared with G–1195G–765G Gly587Arg, the A–1195C–765A Gly/Arg conferred an increased risk of GCA (OR = 2.49, 95% CI = 1.54–4.01). Moreover, significant multiplicative interactions were observed between smoking and these three polymorphisms of –1195G>A, –765G>C, and 587Gly>Arg, even after correction by false discovery rate (FDR) method for multiple comparisons (FDR-Pinteraction = 0.006, 5.239 x 10^-4 and 0.017, respectively). Similarly, haplotypes incorporating these three polymorphisms also showed significant interaction with smoking in the development of GCA (P for multiplicative interaction = 2.65 x 10^-6).

Conclusion: These findings indicated that the functional polymorphisms of COX-2, in interaction with smoking, may play a substantial role in the development of GCA.

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Introduction

Gastric cardia adenocarcinoma (GCA) is the second leading cause of cancer-related mortality in the world, with more than 700,000 deaths annually [1,2,3]. In China, GCA is also one of prevalent fatal malignancies. During the past two decades, epidemiological studies have shown a steady decline in incidence and mortality of GCA, thus emphasizing the importance of prevention strategy to GCA [4,5]. Extensive studies have revealed several environmental factors involved in the development of GCA, including cigarette smoking, alcohol consumption, inflammation, and diet. Most significantly, smoking has been established by considerable studies as a causal factor for GCA [6], which was supported by a recent meta-analysis including 33 studies that smokers had 1.8 fold increased risk of GCA than never-smokers [7]. However, the underlying mechanism how smoking promotes GCA development remains to be fully elucidated. Recently, cumulative evidence has shown that smoking contributed to carcinogenesis potentially through induction of COX-2 and its downstream metabolites [8].

Cyclooxygenase-2 (COX-2), a key enzyme converting arachidonic acid to prostaglandins, was absent from normal cells unless rapidly induced by various carcinogens. For instance, the tobacco specific carcinogen, nicotine, has been shown to up-regulate COX-2 expression and activity in vitro and in vivo [9]. Moreover, in nicotine treated hamsters, COX-2 was significantly increased in gastrointestinal cancer [10,11,12]. Interestingly, in the gastric cancer cells with nicotine-induced COX-2-derived PGE2 release and cell proliferation, the COX-2 inhibitor SC-236 caused G1 arrest and abrogated nicotine-induced cell proliferation [13]. It is therefore concluded that COX-2 played key role in smoke-
associated gastric cancer [8]. However, the expression level and activity of COX-2 induced by smoking may vary among individuals, and only small fraction of exposure individuals would develop to GCA during their life spans, suggesting genetic mechanism depending on COX-2 might be involved in susceptibility to smoke-associated GCA [14,15].

Intriguingly, by direct sequencing and biochemical assays, we have previously identified three functional single nucleotide polymorphisms (SNPs) in COX-2 gene, including –1195 G>A (rs689466), –765G>C (rs20417), and 587Gly>Arg (1759G>A, rs3218625) in coding region, of which, the G to A variant in –1195 locus created a c-myeloblastosis oncogene (c-MYB) binding site, resulting in higher transcriptional activity of COX-2 [16]. The –765C allele might attribute to heightened smoking-induced expression of COX-2 by creating a binding site for phosphorylated NPM (P-NPM), which acted as a specific transcriptional inhibitor and was driven to cytoplasm once with smoking stimulation [17]. In addition, the 587Gly>Arg variant was associated with the enhanced activity of COX-2 in vitro by causing the Gly to Aly amino acid substitution in codon 587 of exon10 [18]. Therefore, an alternative hypothesis was motivated by solid biological plausibility that these three functional SNPs might interact with smoking to modulate the GCA risk. To test this hypothesis, –1195 G>A (rs689466), –765G>C (rs20417), and 587Gly>Arg (rs3218625) were analyzed in a case-control study consisting of 357 GCA cases and 985 controls in a Chinese Han population, and the interaction between these three SNPs and smoking exposure was investigated in modulation of GCA risk.

Materials and Methods

Study subjects

This study consisted of 357 GCA patients and 985 controls. All subjects were unrelated Han Chinese from Beijing city and its surrounding region. Patients were recruited between July 1999 and July 2005 at the Peking Union Hospital and Cancer Hospital, Chinese Academy of Medical Sciences (Beijing). The inclusion criteria for patients included histopathologically confirmed GCA, without previous chemotherapy or radiotherapy, and no restriction in regards to sex, age, or disease stage. The controls were cancer-free individuals randomly selected from a pool of 3000 normal individuals in the Beijing area during the same period. The selection criteria for controls included cancer-free individuals and frequency matching to cases by sex and age (≤5 years). At recruitment, written informed consent was obtained from each subject, and the information on demographic characteristics, such as sex, age, and smoking status were collected via questionnaire. Subjects who had never smoked or smoked less than 1 cigarette per day and shorter than 1 year were defined as non-smokers; otherwise, they were considered as smokers (including current smokers and ex-smokers). This study was approved by the institutional review boards of the Chinese Academy of Medical Sciences Cancer Institute and Tongji Medical College of Huazhong University of Science and Technology.

COX-2 genotyping

Genomic DNA was extracted from whole blood samples of all subjects. Genotypes of three SNPs (including COX-2 –1195G>A, –765G>C, and 587Gly>Arg) were determined by polymerase chain reaction (PCR)-based restriction fragment length polymorphism (RFLP) methods as described previously [16]. Genotyping was performed without knowledge of the case or control status of the subjects. Genotypes determined by RFLP were further confirmed by direct sequencing in 30 random DNA samples, with the 100% concurrence rate of these two methods. Additionally, a 13% masked, random sample from cases and controls was tested twice by different investigators, and the results were concordant for all of the duplicate sets.

Statistical analysis

χ² test was used to compared the distribution of demographic characteristics between cases and controls. Hardy-Weinberg equilibrium for genotypes was tested by a goodness-of-fit χ² test in control group. Multivariate logistic regression was used to evaluate the associations between GCA risk and smoking, COX-2 genotypes or haplotypes. The interaction between smoking and COX-2 SNPs were estimated via multiplicative interaction term and the stratified analysis of the effect of SNPs on GCA by smoking status. A two-tailed P<0.05 was used as the criterion of statistical significance. The final P values were adjusted by the false discovery rate (FDR) correction for multiple comparisons [19]. All statistical analyses were conducted by SPSS v13.0 software. Linkage disequilibrium (LD) of these three SNPs was analyzed using Haploview v.4.0 [20]. Haplotypes composing these three SNPs were estimated using Phase v2.1 [21].

Results

Subjects characteristics

The demographic characteristics of all subjects are presented in Table 1. The cases and controls were matched well on sex and age distribution. There were 55.7% smokers among cases compared with 45% among controls. Significant difference in smoking status was observed between case and control groups (P=0.006). In the logistic regression model, smokers had an increased risk of GCA compared with non-smokers after adjusting for sex and age (OR = 1.41, 95% CI = 1.08–1.84).

COX-2 genotypes and GCA risk

The genotype distributions of the COX-2 –1195G>A, –765G>C, and 587Gly>Arg polymorphisms, no variant homozygotes were observed in this study population. Frequencies for the variant alleles of –1195A, –765C, and 587Arg were 0.56,

| Table 1. Distributions of select characteristics among cases and controls. |
|-----------------|-----------------|-----------------|-----------------|
| Variable        | Controls (n = 985) | Cases (n = 357) | P value         |
| Gender          |                 |                 |                 |
| Male            | 810 (82.2)      | 307 (86.0)      | 0.103           |
| Female          | 175 (17.8)      | 50 (14.0)       |                 |
| Age             |                 |                 | 0.453           |
| 45–65           |                 |                 |                 |
| 46–55           | 214 (21.7)      | 76 (21.3)       |                 |
| 56–65           | 380 (38.6)      | 134 (37.5)      |                 |
| >65             | 289 (29.3)      | 118 (33.1)      |                 |
| Smoking status  |                 |                 | 0.006           |
| Nonsmoker       | 542 (55.0)      | 166 (46.5)      |                 |
| Smoker          | 443 (45.0)      | 191 (53.5)      |                 |

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haplotypes both in cases and controls (48.7% and 50.6% for the A_1195G–C_765G–Gly587Arg haplotype and GCA risk were 0.09, and 0.08 in cases and 0.51, 0.05, 0.05 in controls, respectively.

In the logistic regression analysis, after adjusting for age, sex, and smoking status, individuals with the –1195AA genotype presented an elevated risk of GCA compared with those with the –1195GG genotype (OR = 1.50, 95% CI = 1.05–2.13). For the –765G/C SNP, individuals with the –765GC genotype had more than 2-fold increased risk of GCA (OR = 2.06, 95% CI = 1.29–3.29) compared with carriers of the –765GG genotype. Similarly, the 587Gly/Arg genotype also conferred 1.67 fold increased risk compared with the Gly/Gly genotype (95% CI = 1.04–2.66). Moreover, the effect of the –1195GG, –765GC, or 587Gly/Arg genotype achieved the significant level after correction for family-wise error rates (FDR-P<0.038, 0.009, and 0.033).

**COX-2 haplotypes and GCA risk**

LD analysis showed that these three SNPs are not in discernible linkage disequilibrium in this study population (Table S1). As shown in Table 3, five common haplotypes composing these three SNPs were observed in this study population, of which, the A_1195G–C_765G–Gly587Arg haplotype was the most prevalent haplotype both in cases and controls (48.7% and 50.6%, respectively). Treating the G_1195A–G_765C–Gly587Arg haplotype as the reference, the haplotype A_1195G–C_765G–Gly587Arg showed the significantly highest risk of GCA compared with other haplotypes (OR = 2.49, 95% CI = 1.54–4.01), and the G_1195G–C_765G–Arg variant was also significantly associated with the increased GCA risk (OR = 1.71, 95% CI = 1.01–2.86). Moreover, a significant allele-dose effect of haplotypes was observed in increasing risk of GCA (P = 1.000 x 10^-8).

Interaction of COX-2 genotypes and smoking

In this current study, we evaluated the interaction of these three SNPs and smoking by stratified analysis and the multiplicative interaction term (Table 4). In the stratified analysis, none of the –1195G>A, –765G>C or 587Gly>Arg variants were associated with risk of GCA among nonsmokers, whereas smokers with the 1195G>A, –765G/C or 587 Gly/Arg genotype all showed increased risk of GCA compared with non-smokers carrying the wild-type genotypes (OR = 1.64, 95% CI = 1.06–2.53; OR = 3.98, 95% CI = 2.06–7.69; and OR = 2.67, 95% CI = 1.36–5.23, respectively). Moreover, significantly multiplicative interaction were observed between these three COX-2 SNPs and smoking, even after correction by FDR for multiple comparisons (FDR-P_{interaction} = 0.006, 5.239 x 10^-4, 0.017).

Interaction of COX-2 haplotypes and smoking

We further explored the interaction of COX-2 haplotypes and smoking in GCA (Table 5). Defining nonsmokers with the A_1195G–C_765G–Gly587Arg haplotype as the reference group, no significant association was observed between any haplotype and GCA risk among nonsmokers, whereas smokers with the haplotype of A_1195G–C_765G–Gly587Arg, A_1195G–C_765G–Gly587Arg, A_1195G–C_765G–Gly587Arg, or A_1195G–C_765G–Arg all showed elevated risk for developing GCA (OR = 1.58, 95% CI = 1.05–1.82; OR = 1.62, 95% CI = 1.24–2.11; OR = 2.20, 95% CI = 1.04–4.63; and OR = 4.99, 95% CI = 2.54–9.81 and OR = 18.29, 95% CI = 2.11–158.24, respectively). Moreover, significantly multiplicative interactions were observed between COX-2 haplotypes and smoking (P_{interaction} = 2.65 x 10^-6).

**Discussion**

In this current study, we conducted a case-control study to investigate whether three functional polymorphisms in COX-2, including –1195G>A (rs689466), –765G>C (rs20417), and 587Gly>Arg (rs3218625), interacting with smoking, affect GCA risk in Chinese Han population. In stratified analysis, the effects of these three SNPs on GCA risk varied by smoking status.

**Table 2.** Genotype frequencies of COX-2 SNPs and their association with GCA risk.

| COX-2 genotypes | Cases/controls OR (95% CI) | FDR-P |
|-----------------|---------------------------|-------|
| –1195GG>A       |                           |       |
| GG              | 69/217                    | Reference |
| AG              | 175/527                   | 1.03 (0.75–1.42) | 0.847 |
| AA              | 113/241                   | 1.50 (1.05–2.13) | 0.038 |
| –765G>C         |                           |       |
| GG              | 324/940                   | Reference |
| GC              | 33/45                     | 2.06 (1.29–3.29) | 0.009 |
| 587Gly/Arg      |                           |       |
| Gly/Gly         | 327/933                   | Reference |
| Gly/Arg         | 30/52                     | 1.67 (1.04–2.66) | 0.033 |

**Table 3.** Distribution of COX-2 haplotypes and their association with GCA.

| Haplotype     | Controls n = 985 | Cases n = 357 | OR (95% CI) | P value |
|---------------|-----------------|---------------|-------------|---------|
| A_1195G–C_765G–Gly587Arg | 913 (46.3) | 290 (40.6) | Reference |
| A_1195G–C_765G–Gly587Arg | 960 (48.7) | 361 (50.6) | 1.19 (1.00–1.43) | 0.053 |
| G_1195G–C_765G–Gly587Arg | 44 (2.2) | 23 (3.2) | 1.71 (1.01–2.88) | 0.046 |
| A_1195G–C_765G–Gly587Arg | 41 (2.1) | 33 (4.6) | 2.49 (1.54–4.01) | 1.896 x 10^-4 |
| A_1195G–C_765G–Gly587Arg | 8 (0.4) | 7 (1.0) | 2.60 (0.93–7.23) | 0.068 |

1ORs and 95% CIs were calculated by unconditional logistic regression after adjusting for sex, age and smoking status.

P for trend = 1.000 x 10^-8.

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### Table 4. Stratify and interaction analysis between COX-2 genotypes and Smoking status associated with the risk of GCA.

| Smoking status | $-1195 \text{G} > A$ |          |          |          |          | FDR for multiplicative interaction |
|----------------|----------------------|----------|----------|----------|----------|-----------------------------------|
|                | GG                   | AG + AA  |          |          |          |                                   |
| non smoker     | 34/124               | Reference| 132/418  | 1.16 (0.75–1.77) | 0.006   |
| smoker         | 35/93                | 1.37 (0.78–2.38) | 156/350  | 1.64 (1.06–2.53) |          |
| Smoking status | $-765 \text{G} > C$ |          |          |          |          |                                   |
| non smoker     | 155/515              | Reference| 11/27    | 1.31 (0.63–2.71) | $5.239 \times 10^{-4}$ |
| smoker         | 169/425              | 1.32 (1.01–1.74) | 22/18    | 3.98 (2.06–7.69) |          |
| Smoking status | $587 \text{Gly/Arg}$ |          |          |          |          |                                   |
| Gly/Gly        | 153/511              | Reference| 13/31    | 1.43 (0.73–2.81) | 0.017   |
| Gly/Arg        | 174/422              | 1.38 (1.05–1.81) | 17/21    | 2.67 (1.36–5.23) |          |

1. ORs and 95% CIs were calculated by unconditional logistic regression after adjusting for sex and age.
2. $P$ values for multiplicative interaction were adjusted by the false discovery rate correction for multiple comparisons.

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Furthermore, significant multiplicative interactions were observed between these three SNPs and smoking in GCA development. This study firstly suggested that the interaction of smoking with the COX-2 −1195G>A, −765G>C, and 587Gly>Arg significantly contributed to GCA risk.

It was shown that COX-2 over-expression could increase proliferation, inhibit apoptosis, and enhance the invasiveness of cancer cells. Over-expression of COX-2 has been frequently seen in gastrointestinal malignancies, including pancreas, colon, non-cardia gastric cancer, and GCA [22,23,24,25]. Interestingly, the smoking specified carcinogen, nicotine might act through β-adrenoceptors while NNK through both β-adrenoceptors and α7-nAChR, and induce COX-2 and its derived PGE2 in gastric cancer cells [13]. Cigarette smoke condensate could also induce COX-2 expression by activating nuclear factor kappa B (NF-kB) [26]. Furthermore, nicotine has been shown to facilitate gastric tumor angiogenesis and invasion, suggesting heighten expression of COX-2 induced by smoking might contribute to gastric carcinogenesis. There was evidence that SNPs altering COX-2 expression and activity have been implicated in risk of smoking-associated cancers, including lung, pancreatic, and non-cardia gastric cancer. However, few have done to investigate whether SNPs in COX-2 affect GCA risk solely or in manner of interaction with smoking. Additionally, the effect of smoking on COX-2 enzyme was involved not only in the transcriptional expression but also in the enzymatic activity [13,27,28,29]. Therefore, we analyzed three functional SNPs, −1195G>A and −765G>C in promoter region and 587Gly>Arg in coding region, in this current study.

The most significant finding in this study was that the variants of three functional SNPs identified by our previously studies interacted with smoking to increase risk of GCA. Of these three functional SNPs, −1195A-containing or −765C-containing COX-2 promoter displays higher transcriptional activity, supported by examining the COX-2 expression level of these two variants by real-time PCR quantitation of COX-2 mRNA in individual esophageal tissues [16]. Moreover, smoking exposure can rapidly induce COX-2 expression; hence, the joint effect of the variant and smoking has been expected to increase COX-2 expression. In this current study, significantly association between the −1195AA genotype and increased risk of GCA was only seen among smokers, and multiplicative interaction was observed between this variant and smoking, suggesting the cooperation of −1195G>A and smoking in modulation of GCA risk. Similar result was also observed for the other functional SNP of −765G>C in the promoter region, which was consistent with our previously finding that this SNP interacted with smoking to intensify the risk of pancreatic cancer. Moreover, the biochemical evidence that the markedly higher expression of COX-2 drove by the −765G-containing promoter than the −765C-containing promoter was only seen in the cells treated with smoke condensate has strongly supported this current result [17]. Additionally, the variant of 587Gly>Arg in coding region was also found to interact with smoking exposure to intensified GCA risk, being consistent with our previous finding that heavy smokers with the 587 Gly/Arg genotype presented the highest risk for ESCC compared with non-smokers with the wild-type genotype. Furthermore, biochemical evidence has suggested that the substitution of Gly to Arg, might affect COX-2 activity which was examined in the MCF-7 cells by enzymatic activity assays [17]. Considering that cigarette smoking also influenced COX-2 enzymatic activity, our result for the interaction of the 587 Gly/Arg variant with smoking was biologically plausible. Analysis was also performed on the haplotypes composing these three functional SNPs in GCA. The findings that the variant haplotypes was only associated with smokers’ GCA risk and significant multiplicative interaction was observed between haplotypes and smoking further supported the hypothesis that the functional SNPs altering COX-2 expression and activity interacted with smoking to modulate GCA risk.

In summary, our study highlights the contribution of the interaction between smoking and functional SNPs in COX-2 to GCA susceptibility, raising the prospect of research in personalized prevention strategies to smoking-associated GCA.

### Supporting Information

#### Table S1

| Haplotype  | Nonsmoker | Smoker | P for multiplicative interaction |
|------------|-----------|--------|---------------------------------|
|            | Cases/controls | OR (95% CI) | P | Cases/controls | OR (95% CI) | P |  |
| −1195G>A, −765G>Gly587Arg | 136/502 | Reference | 154/411 | 1.38 (1.05–1.82); 0.021 | 2.650×10−6 |
| −1195G>A, −765G>Gly587Arg | 172/524 | 1.22 (0.95–1.58); 0.124 | 189/436 | 1.62 (1.24–2.11); 3.928×10−4 |
| −1195G>A, −765G>Gly587Arg | 11/24 | 1.78 (0.85–3.74); 0.127 | 12/20 | 2.20 (1.04–4.63); 0.038 |
| −1195G>A, −765G>Gly587Arg | 11/25 | 1.58 (0.76–3.29); 0.226 | 22/16 | 4.99 (2.54–9.81); 3.240×10−6 |

ORs and 95% CIs were calculated by unconditional logistic regression after adjusting for sex and age.

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Table 5. Stratify and interaction analysis between COX-2 haplotypes and Smoking associated with the risk of GCA.

Conceived and designed the experiments: X-PM X-MZ RZ LL. Performed the experiments: X-MZ RZ LL. Analyzed the data: RZ LL YW ZZ. Contributed reagents/materials/analysis tools: X-MZ RZ LL CS PW. Wrote the paper: X-MZ RZ LL XM. Collected the data: PW CS J-XY W-GS.
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