Phospho-Akt Immunoreactivity in Prostate Cancer: Relationship to Disease Severity and Outcome, Ki67 and Phosphorylated EGFR Expression

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Abstract

Background: In the present study, we have investigated the prognostic usefulness of phosphorylated Akt immunoreactivity (pAkt-IR) in prostate cancer using a well-characterised tissue microarray from men who had undergone transurethral resection due to lower urinary tract symptoms.

Methodology/Principal Findings: pAkt-IR in prostate epithelial and tumour cells was assessed using a monoclonal anti-pAkt (Ser⁴⁷³) antibody. Immunoreactive intensity was determined for 282 (tumour) and 240 (non-malignant tissue) cases. Tumour pAkt-IR scores correlated with Gleason score, tumour Ki67-IR (a marker of cell proliferation) and tumour phosphorylated epidermal growth factor receptor (pEGFR)-IR. For cases followed with expectancy, a high tumour pAkt-IR was associated with a poor disease-specific survival, and the prognostic information provided by this biomarker was additive to that provided by either (but not both) tumour pEGFR-IR or Ki67-IR. Upon division of the cases with respect to their Gleason scores, the prognostic value of pAkt-IR was seen for patients with Gleason score 8–10, but not for patients with Gleason score 6–7.

Conclusions/Significance: Tumour pAkt-IR is associated with both disease severity and disease-specific survival. However, its clinical use as a biomarker is limited, since it does not provide prognostic information in patients with Gleason scores 6–7.

Citation: Hammarsten P, Cipriano M, Josefsson A, Stattin P, Egevad L, et al. (2012) Phospho-Akt Immunoreactivity in Prostate Cancer: Relationship to Disease Severity and Outcome, Ki67 and Phosphorylated EGFR Expression. PLoS ONE 7(10): e47994. doi:10.1371/journal.pone.0047994

Editor: Kaustubh Datta, University of Nebraska Medical Center, United States of America

Received July 9, 2012; Accepted September 19, 2012; Published October 25, 2012

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Funding: The authors thank the Swedish Research Council (Grant no. 12158, medicine, C.J. Fowler); the Swedish Cancer Society (Grant no. CAN2010/457, C.J. Fowler); Lion’s Cancer Research Foundation, Umeå University (P. Hammarsten & C.J. Fowler); the Cancer Research Foundation in Northern Sweden (P. Hammarsten), and the Research Funds of the Medical Faculty, Umeå University (C.J. Fowler) for financial support. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

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Introduction

According to recent statistics, the global incidence of prostate cancer (Pca) in 2008 was approximately 900,000 cases [1]. The treatment of Pca varies according to tumour stage and histological grade. Options range from watchful waiting in elderly patients with low-grade disease to oncological treatment of generalised cancer. Radical prostatectomy and radiotherapy are associated with considerable morbidity [2] and a large proportion of the patients would have died of other causes than prostate cancer even without the curative treatment [3–5]. Therefore, robust biomarkers are greatly needed to aid treatment decisions.

The serine-threonine kinase Akt is an important regulator of cell proliferation and apoptosis. Downstream effects of Akt activation include the phosphorylation and thereby inactivation of the protein BAD, a pro-apoptotic protein [6] and a change in the transcriptional activity of androgen receptors [7]. Overexpression of a constitutively active form of Akt results in an increased growth of LNCaP prostate cancer cells in a xenograft model [8]. In man, immunohistochemical levels of phosphorylated Akt (pAkt) are higher in prostate tumour tissue and in bone metastases than in non-malignant prostate tissues [9,10] and are higher in Gleason score 8–10 tumours than lower Gleason scores [11]. Three independent studies have reported that high tumour pAkt immunoreactivity is associated with a poor clinical outcome (biochemical relapse assessed with serum prostate-specific antigen (PSA) [9,12], and survival [13]). In contrast, a fourth study found little prognostic value in pAkt IR [14]. The studies measuring survival, however used rather small cohort sizes (53 and 68 for [14] and [13], respectively). It is therefore important to assess whether or not pAkt immunoreactivity has prognostic significance upon disease-specific survival in a large cohort of well-characterised cases. In addition, it is important to assess whether pAkt immunoreactivity in benign tissue adjacent to tumours has prognostic significance upon disease-specific survival.
usefulness, or whether it is restricted to tumour expression alone.

Epidermal growth factor receptor (EGFR) is a cell surface receptor tyrosine kinase responsive to a number of different intracellular signalling pathways, in turn resulting in cell growth and survival [15]. Inhibition of EGFR has been shown to enhance castration-induced prostate involution [16]. In Pca, tumour pEGFR immunoreactivity is associated with biochemical recurrence rates [17] and with disease-specific survival [18]. One of the signalling pathways utilised by EGFR is the phosphoinositide 3-kinase/Akt pathway [19] which in Pca cell lines contributes to cell migration mediated by EGFR and to the transformation of these cells to give them characteristics reminiscent of epithelial-mesenchymal transition [20]. In Pca tumour tissues, pAkt is frequently co-expressed with EGFR [10], but it is not known whether or not cases with a high expression of pEGFR and pAkt may show different clinical outcomes to those with, for example, a high expression of pEGFR but a low expression of pAkt (or vice versa).

In the present study, we investigated pAkt immunoreactivity in a well-characterised Pca tissue array [3], and determined a) the relationship between pAkt immunoreactivity with disease severity and outcome (disease-specific survival) and b) the influence of pEGFR immunoreactivity upon this relationship.

Methods

Patients

The formalin-fixed, paraffin-embedded samples used in the present study were collected between 1975 and 1991 at the Central Hospital, Västerås, Sweden, from a total of 412 patients diagnosed with prostate cancer at transurethral resection for lower urinary tract symptoms [3].

The material was collected according to Swedish Regulations at a time when informed consent was not required. The research ethical committee at Umeå university hospital (Regional Ethical Review Board in Umeå, Sweden) approved of the study and waived the need for informed consent. In the database used for the analyses, the tissue samples were given a case number and year, and the patient names were not indicated in the database.

The presence of metastases was determined by bone scans shortly after the transurethral resection, and the patients were followed until death or until 2003. The Gleason scores, the percentage of the specimen that contained tumour and the disease stage were assessed in each sample. Cause of death was determined by evaluation of medical records. Tissue microarrays using cores with a diameter of 0.6 mm were constructed using a Beecher Instrument (Sun Prairie, WI, USA). Ki67 was analysed either as the number of positive stained cells crossing 11 horizontal lines across the core, or as the percentage of cells positive for this marker (reported in [21]). In the present paper, we have presented the data using the number rather than the percentage of positive stained cells. Unless otherwise stated, very similar results were seen with both methods of scoring, pEGFR (range of scores 0–5), and PDFR (range of scores 0–3) immunoreactive (IR) scores were available in the database, and have been reported elsewhere [18,22].

Analysis of Data

Tumour and non-malignant cores were scored for pAKT in prostate epithelial and tumour cells from digitally scanned images by two independent evaluators (MC & CF) who did not access the clinical data at the time of the evaluation. The scanned scores were analysed on the basis of intensity (0 = no staining, 4 = maximal staining) and distribution (0, 25, 50, 75 or 100% for each intensity).

The composite score for each core was then determined. Thus, for example, a core with 25% intensity 3 and 75% intensity 2 would be given a score of 0.25 × 3 + 0.75 × 2 = 2.25. Examples of staining intensities 1–4 are shown in Fig. 1. Not all cores are as well defined as those shown in Figure 1, and in some cases it is a judgement call as to whether or not a core should be scored (such as, for example, a case with mainly stroma and only one or two small areas of epithelial cells). In consequence, we only used cores that had been scored by both evaluators. For the 1648 cores scored independently by both evaluators, an intra-class correlation analysis using a mixed model and testing for consistency gave a Chronbach's alpha of 0.92. In fact 1219 of the 1648 cores (74%) were scored by the two investigators within 0.5 points of each other. Given that cores with staining somewhere between the units used are not uncommon, a difference in scores of up to 1 can be accepted. A total of 68 (4%) of the cores had differences in scores >1, and these were in consequence rescored, again independently and without accessing the previous scores, given that such large differences could be due to typographical errors or patterns of immunoreactivity that were difficult to score. Following the rescoring, 7 cores were discarded due to poor quality, 55 cores now had scores within 1 unit of each other, and only six cores with score differences >1 remained. These six cores were excluded from the analysis. After exclusion of data from one slide (24 cases) where there were no cores with scores >1, suggesting a technical failure, the median scores for each case were determined, and the medians from the two investigators were averaged and entered into the database. Subsequent investigation of discarded cases in the database indicated that 20 of the 24 cases had Gleason scores 4–5, and 4 had Gleason scores 8–10. The preponderance of Gleason score 4–5 means that their inclusion could bias the prognostic evaluation of pAkt if these immunoreactivities are artifactual. Thus, by excluding them, we are erring on the side of caution.

Statistics

With the exception of the intraclass correlation coefficient and Cox proportional-hazards regression analyses, which were conducted using SPSS software (SPSS Inc., Chicago, IL, USA), all statistical calculations were undertaken using the statistical package built into the GraphPad Prism 5 computer programme for the Macintosh (GraphPad Software Inc., San Diego, CA, USA). Partial correlation coefficients were calculated on an Excel spreadsheet from the Spearman correlation coefficients for matched samples (i.e. scored for all three parameters under investigation). For survival analyses, an event was defined as death
due to prostate cancer and entered into the database as “event=1”, thereby allowing us to determine disease-specific survival. Death from other causes was censored, as were cases where the patient was alive at the date of last follow-up. Three cases where the disease outcome was not known were excluded from the survival analyses.

**Results**

**Distribution of pAkt Immunoreactivity (pAkt-IR) in Non-malignant and Tumour Tissue**

Cores from 282 (tumour samples) and 240 (non-malignant tissue samples) cases were scored for pAkt-IR. Consistent with previous studies [9,10,12], pAkt-IR was associated with epithelial cells rather than the stroma (Fig. 1). There was a wide range of staining intensities (see e.g. Fig. 1 for examples and Fig. S1 for a distribution curve). In general, the pAkt-IR was greater in the tumour samples than in the non-malignant samples: the median scores were 2.75 and 2.0625 for the tumour and non-malignant samples, respectively. For the 189 cases shown as part of Fig. 1B where both tumour and non-malignant cores were scored, the pAkt-IR scores were significantly correlated (Spearman’s rho = 0.31, P<0.0001), and the scores for the tumour cores were significantly higher than those for the non-malignant cores (P<0.0001, Wilcoxon matched-pairs signed rank test).

**Correlation of pAkt-IR with Disease Severity at Diagnosis**

Correlation coefficients for the tumour and non-malignant pAkt-IR scores vs. clinical and biochemical parameters scored for the same samples are summarised in Table 1. The tumour pAkt-IR correlated significantly with the Gleason score, the percentage of the core that was tumour associated (%ca), the tumour stage, and the tumour Ki67-IR (a measure of cell proliferation). Thus, the tumour pAkt-IR is highly associated with disease severity at diagnosis. The non-malignant pAkt-IR was more weakly associated with these parameters.

An important question to be investigated is whether the associations between non-malignant pAkt-IR and the clinical markers of disease severity are true correlations, or simply reflect the correlation between tumour and non-malignant pAkt-IR. This can be assessed using the formula:

$$\Gamma_{ab,c} = (\Gamma_{ab} - \Gamma_{ac} \cdot \Gamma_{bc})/\sqrt{(1 - [\Gamma_{ac}]^2)(1 - [\Gamma_{bc}]^2))}.$$
where $\Gamma_{ab}$ is the Spearman correlation coefficient for the interaction between parameters a and b, and $\Gamma_{ab,c}$ refers to the first order partial coefficient between parameters a and b when parameter c is taken into consideration. The equation represents the general formula for partial correlation derived in [23]. Using this formula for the samples scored for both tumour and non-malignant tissue samples.

$pAkt-IR (T)$ and the Gleason score or the tumour proliferation marker Ki67-IR remain significant when either the influence of the tumour (T) $pAkt-IR$ is taken into account (Table S1). In contrast, the associations between $pAkt-IR$ (N) and the clinical variables are not significant when the influence of the tumour (T) $pAkt-IR$ is taken into account (Table S1), although the correlation between $pAkt-IR$ (T) and %ca remained significant when Ki67 was scored as % of stained cells positive for this marker rather than the number of positive stained cells crossing 11 horizontal lines across the core (data not shown).

Thus, it can be concluded that $pAkt-IR$ (T) is primarily associated with the Gleason score and the rate of tumour proliferation as assessed by Ki67-IR, whereas the $pAkt-IR$ (N) is not associated with disease severity.

### Table 1. Correlation coefficients for $pAkt-IR$ scores with clinical parameters and with the proliferation marker Ki67.

| Parameter | $Sp\rho$ | P | n | $Sp\rho$ | P | n |
|-----------|----------|---|---|----------|---|---|
| Age       | -0.03    | >0.5 | 282 | -0.12 | 0.065 | 240 |
| Gleason score | 0.39 | <0.0001 | 282 | 0.13 | 0.04 | 240 |
| % ca$^a$ | 0.27 | <0.0001 | 282 | 0.17 | 0.0069 | 240 |
| Tumour stage | 0.30 | <0.0001 | 280 | 0.14 | 0.03 | 238 |
| $K67-IR$ (T) | 0.38 | <0.0001 | 279 | 0.06 | >0.3 | 232 |
| $K67-IR$ (N) | 0.03 | >0.6 | 263 | 0.10 | >0.1 | 236 |

$Sp\rho$ refers to Spearman’s rho value for the non-parametric comparisons.

$^a$% of core that was tumour associated.

Abbreviations: T, tumour; N, non-malignant; Nl, non-malignant luminal, Nb; non-malignant basal; ep, epithelial; st, stroma. $Sp\rho$ refers to Spearman’s rho value for the non-parametric comparisons.

### Table 2. Correlation coefficients for $pAkt-IR$ scores with pEGFR-IR, total EGFR-IR and PDFRß-IR scores in the tumour and non-malignant tissue samples.

| Parameter | $Sp\rho$ | P | n | $Sp\rho$ | P | n |
|-----------|----------|---|---|----------|---|---|
| pEGFR-IR (T) | 0.27 | <0.0001 | 227 | 0.17 | 0.02 | 193 |
| pEGFR-IR (N) | 0.24 | 0.0004 | 219 | 0.31 | <0.0001 | 196 |
| pEGFR-IR (Nb) | 0.22 | 0.0008 | 219 | 0.30 | <0.0001 | 196 |
| PDFRß-IR (T, ep) | 0.04 | 0.95 | 230 | -0.001 | 1 | 190 |
| PDFRß-IR (N, ep) | 0.05 | 0.46 | 212 | 0.09 | 0.23 | 194 |
| PDFRß-IR (T, st) | 0.12 | 0.06 | 226 | -0.09 | 0.24 | 187 |
| PDFRß-IR (N, st) | 0.05 | 0.39 | 257 | 0.10 | 0.12 | 230 |

$Sp\rho$ refers to Spearman’s rho value for the non-parametric comparisons.

Abbreviations: T, tumour; N, non-malignant; Nb, non-malignant basal; ep, epithelial; st, stroma. $Sp\rho$ refers to Spearman’s rho value for the non-parametric comparisons.

### Table 3. Age, Gleason scores, incidence of metastases at diagnosis and tumour Ki67-IR at diagnosis for the cases divided on the basis of tumour $pAkt-IR$ and pEGFR-IR scores.

| pAkt-IR $<$2.75 | pAkt-IR $\geq$2.75 | pEGFR-IR $<$3.2 | pEGFR-IR $\geq$3.2 | pEGFR-IR $<$3.2 | pEGFR-IR $\geq$3.2 | P value |
|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|---------|
| Age in years: median (range) [n] | 75 (58–87) [30] | 73 (52–88) [30] | 76 (64–89) [37] | 74 (51–95) [90] | NS$^a$ |
| Number (%$^b$) of cases with: | | | | | |
| Gleason score | 4–5 | 19 (38%) | 11 (22%) | 2 (5%) | 5 (6%) | <0.0001$^c$ |
| 7 | 19 (38%) | 8 (16%) | 19 (51%) | 16 (18%) | |
| 8–10 | 7 (14%) | 14 (28%) | 9 (24%) | 18 (20%) | |
| pEGFR effect: | P<0.005$^a$ | P<0.0005$^a$ |
| Number (%$^b$) of cases with: | | | | | |
| Metastases | No | 37 (100%) | 35 (88%) | 23 (88%) | 57 (76%) | <0.01$^c$ |
| at diagnosis | Yes | 0 (0%) | 5 (13%) | 3 (12%) | 18 (24%) | |
| pEGFR effect: | P<0.05$^a$ | P<0.05$^a$ |
| K67-IR (T) median (range) [n] | 0.8 (0–14.4) [50] | 1.5 (0–40.8) [50] | 1.4 (0–65.1) [36] | 2.9 (0–78.5) [88] | <0.0001$^c$ |
| pEGFR effect: | NS$^d$ | NS$^d$ |

$^a$Kruskal-Wallis test.

$^b$The % value refers to the % of cases for the $pAkt-IR$/pEGFR-IR group in question (i.e. vertical numbers add up to 100%).

$^c$Dunn’s Multiple Comparison Test following significant Kruskal-Wallis test.

$^d$Not significant (P>0.05). "pEGFR effect" refers to the comparison between pEGFR-IR <3.2 and $\geq$3.2 for the given $pAkt-IR$ tranche. For comparisons between $pAkt-IR$ $<$2.75 and $\geq$2.75 for pEGFR <3.2 alone, the significance levels for Gleason score, metastases at diagnosis and K67-IR (T) were P<0.01$^d$, NS$^d$ and P<0.05$^d$, respectively. The corresponding significance levels for pEGFR-IR $\geq$3.2 alone were P<0.01$^d$, NS$^d$ and P<0.01$^d$, respectively.

Abbreviations: T, tumour; N, non-malignant; Nl, non-malignant luminal, Nb; non-malignant basal; ep, epithelial; st, stroma. $Sp\rho$ refers to Spearman’s rho value for the non-parametric comparisons.

### Table 4. Correlation coefficients for $pAkt-IR$ scores with pEGFR-IR, total EGFR-IR and PDFRß-IR scores in the tumour and non-malignant tissue samples.

| Parameter | $Sp\rho$ | P | n | $Sp\rho$ | P | n |
|-----------|----------|---|---|----------|---|---|
| pEGFR-IR (T) | 0.27 | <0.0001 | 227 | 0.17 | 0.02 | 193 |
| pEGFR-IR (N) | 0.24 | 0.0004 | 219 | 0.31 | <0.0001 | 196 |
| pEGFR-IR (Nb) | 0.22 | 0.0008 | 219 | 0.30 | <0.0001 | 196 |
| PDFRß-IR (T, ep) | 0.04 | 0.95 | 230 | -0.001 | 1 | 190 |
| PDFRß-IR (N, ep) | 0.05 | 0.46 | 212 | 0.09 | 0.23 | 194 |
| PDFRß-IR (T, st) | 0.12 | 0.06 | 226 | -0.09 | 0.24 | 187 |
| PDFRß-IR (N, st) | 0.05 | 0.39 | 257 | 0.10 | 0.12 | 230 |

$Sp\rho$ refers to Spearman’s rho value for the non-parametric comparisons.

Abbreviations: T, tumour; N, non-malignant; Nb, non-malignant basal; ep, epithelial; st, stroma. $Sp\rho$ refers to Spearman’s rho value for the non-parametric comparisons.
Correlation between pAkt-IR and pEGFR-IR

pAkt-IR (T) was significantly correlated to pEGFR-IR scores for both tumour and non-malignant samples (Table 2). In contrast, there were no significant correlations between the pAkt-IR scores and either the stromal or epithelial PDFRß-IR scores (Table 2). The correlation between pAkt-IR (T) and pEGFR-IR (T) remained significant when controlled for pAkt-IR (N) whereas the the correlation between pAkt-IR (T) and either pEGFR-IR in the luminal or basal non-malignant tissue was lost when controlled this variable. Conversely, the correlation between pEGFR-IR (T) and pAkt-IR (N) was lost when controlled for pAkt-IR (T) (Table S2). In other words, the association of pEGFR-IR and pAkt-IR is primarily within the same region of the tissue.

The samples were grouped into four groups on the basis of their tumour pAkt-IR and pEGFR-IR scores and their Gleason scores, incidence of metastases at diagnosis and tumour Ki67-IR scores were compared (Table 3). The distribution of Gleason scores was significantly different for the groups. At the extremes, only 10% of the cases with “low” pEGFR-IR (<3.2) and pAkt-IR (<2.75) scores were diagnosed with Gleason scores in the range 8–10, whereas the corresponding number for cases with “high” pEGFR-IR (≥3.2) and pAkt-IR (≥2.75) scores was 57%. For the Ki67-IR scores, there was no effect of pEGFR-IR at a given pAkt-IR,

Figure 2. Prognostic significance of tumour and non-malignant pAkt-IR for cases who were followed by expectancy. Panels A and B are for tumour pAkt-IR (n = 204), C and D for non-malignant pAkt-IR (n = 194). In Panels A and C, Exp(B) (±95% confidence intervals), obtained from Cox proportional-hazards regression analyses are shown for different cut-offs. Exp(B) is defined as the increase in risk for death due to prostate cancer for a score above the cut-off value relative to a score below the cut-off value. When both confidence limits are above unity (filled symbols in the figure), the cut-off value provides significant prognostic information. Values with a significance level 0.05 < P < 0.1 are shown as open triangles. The cut-off value with the highest significance is shown as a red filled symbol. The blue dotted line indicates the % of cases above the cut-off value. Thus, for example, for the symbol in Panel A at pAkt-IR cut-off value 2.5 (i.e sample divided as ≤2.5 and >2.5), 82 cases (40%) were below the cut-off value and 122 cases (60%) above the cut-off value. In Panels B and D, Kaplan-Meier plots are shown for the cut-offs showing the highest significances. \(^\text{1}P\text{ca}\) refers to the number of patients who died as a result of their prostate cancer during the follow-up period. The k \(^2\) values are for the log-rank (Mantel-Cox) tests, with the P values shown: ***P<0.001, *P<0.05.

doi:10.1371/journal.pone.0047994.g002
whereas the scores were higher for pAkt-IR $\geq 2.75$ compared with $<2.75$ for either pEGFR-IR group. The converse was seen for the incidence of metastases at diagnosis.

**Association of pAkt with Disease-specific Survival**

For the 282 cases where tumour pAkt-immunoreactivity (pAkt-IR) could be scored, 207 were followed by expectancy, as was the standard approach at the time in Sweden. The other patients received radiotherapy, hormonal treatment or radical prostatectomy. Three cases where outcome was not known were excluded, giving a total of 204 cases available for survival analysis. The corresponding number of cases for the non-malignant pAkt-IR was 194. These samples allow determination as to whether pAkt-IR has prognostic value. For the 204 tumour cases, 101, 20 and 83 cases were below, equal to and above the median value (2.75), respectively.

In the search of potential prognostic markers, the choice of cut-off is important. In order to investigate this, Cox proportional-hazards regression analyses were undertaken for both tumour and non-malignant pAkt-IR and over a wide range of cut-off values. The data are shown in Fig. 2. For tumour pAkt-IR, there is a reasonably broad band of cut-off values giving a significant increase in relative risk for values above the cut-off compared to those below the cut-off (Fig. 2A). In other words, dividing the data as, for example, $\leq 2.5$ and $>2.5$, or as a median split. We have chosen $2.6875$ and $2.6875$ [i.e. $<2.75$ and $\geq 2.75$, the median split (see above)] as the cut-off, since this gives the greatest significance (shown as the red symbol in Fig. 2A). Using this cut-off, patients with a pAkt-IR $\geq 2.75$ ("high") had a significantly poorer prognosis than those with a pAkt-IR $<2.75$ ("low") (Fig. 2B). The 15 year rate of disease-specific survival was $42\%$ and $72\%$ for high and low pAkt-IR scores, respectively.

In contrast to the situation for tumour pAkt-IR, non-malignant pAkt-IR scores had a narrow band where significant prognostic value was seen (Fig. 2C), and the Kaplan-Meier plot using the optimal cut-off showed a lower degree of significance than was seen for the tumour pAkt-IR (Fig. 2D). Furthermore, a bivariate COX regression analysis indicated that the significant effect of pAkt-IR scores had a narrow band where significant prognostic value. For the 204 tumour cases, 101, 20 and 83 cases were below, equal to and above the median value (2.75), respectively.

Kaplan-Meier plots of the influence of tumour pAkt-IR upon survival for the entire data set and subdivided on the basis of either the pEGFR-IR or the Ki67-IR cut-offs are shown in Fig. 3B and D. The general pattern seen in both cases was cases with the low scores on both markers had a good prognosis, those with high scores on both markers had a poor prognosis, with the other combinations clustering in the middle with rather similar prognoses. Pairwise comparisons indicated that at a low expression

### Table 4. COX proportional-hazards regression analyses for tumour and non-malignant pAkt-IR, tumour pEGFR-IR and Ki67-IR for patients followed by expectancy.

| Parameter       | Cut-off | n   | Exp(B) | Lower | Upper | P      |
|-----------------|---------|-----|--------|-------|-------|--------|
| **Univariate**  |         |     |        |       |       |        |
| pAkt-IR (T)     | $<2.75$ | 101 | 1      |       |       |        |
|                 | $\geq 2.75$ | 103 | 3.173 | 1.766 | 6.968 | 0.0001 |
| pAkt-IR (N)     | $<2.5$  | 137 | 1      |       |       |        |
|                 | $\geq 2.5$ | 57  | 1.963 | 1.072 | 3.596 | 0.029  |
| pEGFR-IR (T)    | $<3.2$  | 118 | 1      |       |       |        |
|                 | $\geq 3.2$ | 135 | 3.590 | 2.018 | 6.388 | $<0.0001$ |
| Ki67-IR (T)     | $<1.5$  | 162 | 1      |       |       |        |
|                 | $\geq 1.5$ | 57  | 3.515 | 1.853 | 6.668 | 0.0001 |
|                 | $\geq 3$ | 67  | 6.898 | 3.851 | 12.357| $<0.0001$ |
| **Bivariate**   |         |     |        |       |       |        |
| pAkt-IR (T)     | $<2.75$ | 76  | 1      |       |       |        |
|                 | $\geq 2.75$ | 73  | 2.412 | 1.163 | 5.003 | 0.018  |
| pAkt-IR (N)     | $<2.5$  | 106 | 1      |       |       |        |
|                 | $\geq 2.5$ | 43  | 1.776 | 0.878 | 3.591 | 0.11   |
| pAkt-IR (T)     | $<2.75$ | 85  | 1      |       |       |        |
|                 | $\geq 2.75$ | 100 | 2.772 | 1.485 | 5.175 | 0.001  |
| pEGFR-IR (T)    | $<3.2$  | 80  | 1      |       |       |        |
|                 | $\geq 3.2$ | 105 | 2.967 | 1.477 | 5.962 | 0.002  |
| pAkt-IR (T)     | $<2.75$ | 101 | 1      |       |       |        |
|                 | $\geq 2.75$ | 101 | 2.218 | 1.213 | 4.053 | 0.010  |
| Ki67-IR (T)     | $<1.5$  | 104 | 1      |       |       |        |
|                 | $\geq 1.5$ | 45  | 3.082 | 1.430 | 6.644 | 0.004  |
|                 | $\geq 3$ | 53  | 6.050 | 2.960 | 12.362| $<0.0001$ |
| pEGFR-IR (T)    | $<3.2$  | 116 | 1      |       |       |        |
|                 | $\geq 3.2$ | 133 | 2.891 | 1.593 | 5.245 | 0.0005 |
| Ki67-IR (T)     | $<1.5$  | 137 | 1      |       |       |        |
|                 | $\geq 1.5$ | 50  | 2.903 | 1.499 | 5.622 | 0.002  |
|                 | $\geq 3$ | 62  | 5.904 | 3.219 | 10.831| $<0.0001$ |
| **Trivariate**  |         |     |        |       |       |        |
| pAkt-IR (T)     | $<2.75$ | 85  | 1      |       |       |        |
|                 | $\geq 2.75$ | 98  | 1.897 | 0.988 | 3.642 | 0.054  |
| pEGFR-IR (T)    | $<3.2$  | 79  | 1      |       |       |        |
|                 | $\geq 3.2$ | 104 | 2.105 | 1.027 | 4.316 | 0.042  |
| Ki67-IR (T)     | $<1.5$  | 93  | 1      |       |       |        |
|                 | $\geq 1.5$ | 40  | 2.599 | 1.148 | 5.882 | 0.022  |
|                 | $\geq 3$ | 50  | 5.430 | 2.529 | 11.661| $<0.0001$ |

doi:10.1371/journal.pone.0047994.t004

Tumour pAkt-IR and Either Tumour pEGFR-IR or Ki67-IR Provide Additive Prognostic Information

Cox regression data using a wide range of cut-offs were also constructed for the tumour Ki67-IR and pEGFR-IR data available in the database [18,21]. For pEGFR-IR, the cut-off giving the highest significance corresponded to $<3.2$ and $\geq 3.2$ (Fig. 3A). In the case of Ki67-IR, there appeared to be two peaks of with approximately equal significance levels, and in consequence the highest points of significance were taken for both peaks, to give three tranches: $<1.5$, $1.5–2.9$ and $\geq 3$ (scores represent the number of tumour cells positive for this biomarker in crossing 11 horizontal lines across the core, Fig. 3C). Using these cut-off ranges, bivariate COX regression analyses indicated that pAkt-IR provided additive prognostic information to that provided either by pEGFR-IR or by Ki67-IR (Table 4). In a tri-variate analysis with all three parameters, the influence of pAkt-IR did not quite reach significance ($P = 0.054$, Table 4). A similar result was seen when Ki67-IR was expressed as % of cells positive for this marker, although there were fewer patients in the highest tranche (data not shown).
rate of pAkt-IR, the influence of pEGFR-IR does not quite reach significance (and vice versa), whereas at a high expression rate of pAkt-IR, the effect of pEGFR is highly significant (and vice versa) (Table 5).

Prognostic Usefulness of Tumour pAkt-IR at Different Gleason Scores

The curves shown in Fig. 3 did not take into account the Gleason scores of the samples. There were no cases of Gleason scores 4–5 that were scored for pAkt-IR who died as a result of their cancer, and so only the cases with Gleason scores 6–10 were investigated. A total of 153 cases followed by active expectancy and scored for tumour pAkt-IR had Gleason scores in this range. The Cox regressions at multiple cut-offs again showed an optimal cut-off at tumour pAkt-IR scores of \( <2.75 \) and \( \geq 2.75 \), although the range of significant Exp(B) values was very narrow, with only two cut-off points reaching significance (data not shown). For Gleason scores 6–7 (\( n = 102 \)), none of the cut-offs reached significance (Fig. 4A) and the Kaplan-Meier plot using the cut-off of \( <2.75 \) and \( \geq 2.75 \) (for illustrative purposes) showed no difference between the survival curves for the two populations. At Gleason scores 8–10 (\( n = 51 \)) only the cut-off at tumour pAkt-IR scores of \( <2.75 \) and \( \geq 2.75 \) was significant (Fig. 4C). From the Kaplan-Meier plot using this cut-off (Fig. 4D), 5 year disease-specific survival rates of 79±14% and 37±9% for pAkt-IR scores of \( <2.75 \) and \( \geq 2.75 \), respectively, were found. For comparative purposes, Exp(B) values were also determined for the Gleason groups 6–7 and 8–10 for both tumour pEGFR-IR and Ki67-IR. pEGFR-IR retained prognostic value for Gleason group 6–7 but not 8–10, whilst Ki67-IR retained prognostic value at both Gleason score groups (Figure S2). A bivariate Cox proportional-hazards regression analysis for the Gleason score 8–10 cases indicated that the tumour pAkt-IR score did not provide additive prognostic information to that provided by the tumour Ki67-IR score (Table S3). When Ki67-IR scores for the Gleason score 8–10 cases were analysed as % of cells positive for this marker rather than the number of positive stained cells crossing 11 horizontal lines across the core, only a single cut-off in the Cox proportional-hazards regression analyses gave a significant Exp(B) value, and the number of cases above this cut-off were so few (\( n = 5 \)) that bivariate Cox proportional-hazards regression analyses with pAkt-IR were not deemed to be meaningful.

Figure 3. Prognostic significance of tumour pEGFR-IR and Ki67-IR for cases who were followed by expectancy: relationship with tumour pAkt-IR. Panel A shows Exp(B) obtained from Cox proportional-hazards regression analyses are shown for different cut-offs of pEGFR-IR (\( n = 253 \)). The cut-off value with the highest significance is shown as a red filled symbol. Values with a significance level \( 0.05 < P < 0.1 \) are shown as open triangles. Panel B shows a Kaplan-Meier plot for the 185 cases scored for both tumour pAkt-IR and pEGFR-IR, divided up on the basis of their optimal cut-offs. In Panel C, Exp(B) values are shown for different cut-offs of Ki67-IR (\( n = 286 \)). The red and blue symbols indicate the highest significance levels for the lower range and for the higher range, respectively. Panel D shows a Kaplan-Meier plot for the 202 cases scored for both tumour pAkt-IR and Ki67-IR, divided up on the basis of their optimal cut-offs. \(^1\)Pca refers to the number of patients who died as a result of their prostate cancer during the follow-up period. The \( k \cdot \chi^2 \) values are for the log-rank (Mantel-Cox) tests, with the \( P \) values shown: \(* * * P < 0.001\).
**Table 5.** Pairwise comparisons of significance levels and 15 year rates of disease specific survival for the combinations of pEGFR-IR and pAkt-IR shown in Fig. 3B.

| Comparison          | n, Pca   | $\chi^2$, P* | 15 year rate of disease-specific survival |
|---------------------|----------|--------------|------------------------------------------|
| pAkt low/pEGFR low vs. | 46, 4    | 3.59, 0.06   | 87±6% vs.                                |
| pAkt low/pEGFR high  | 39, 11   | 62±10%       |                                          |
| pAkt low/pEGFR low vs. | 46, 4    | 2.88, 0.09   | 87±6% vs.                                |
| pAkt high/pEGFR low  | 34, 6    | 58±15%       |                                          |
| pAkt high/pEGFR low vs. | 34, 6    | 6.67, 0.01   | 58±15% vs.                                |
| pAkt high/pEGFR high | 66, 31   | 34±8%        |                                          |
| pAkt high/pEGFR high vs. | 39, 11   | 8.43, 0.005  | 62±10% vs.                                |
| pAkt high/pEGFR high  | 66, 31   | 34±8%        |                                          |

*low* and *high* refer to the cut-offs shown in Fig. 3B.
*Pca refers to the number of patients who died as a result of their prostate cancer during the follow-up period.

Discussion

In the present study, pAkt-IR was assessed in a well characterised Pca tissue microarray [3]. There are three main findings, that are discussed in turn below.

**Tumour pAkt-IR is Higher than Non-malignant pAkt-IR and is Correlated to Disease Severity at Diagnosis**

Previous studies have reported that pAkt-IR scores are higher in tumour tissue than in non-malignant tissue [9,10], and the present study has confirmed this finding. In our study, tumour pAkt-IR was correlated to both the Gleason score and the incidence of metastases at diagnosis, whereas the relationship of non-malignant pAkt-IR to disease severity was a reflection of its own correlation with tumour pAkt-IR. Tumour pAkt-IR was also highly correlated to Ki-67 IR, a marker of cell proliferation. Ayala et al. [9] reported in a large cohort that whilst tumour pAkt-IR was greater than the non-malignant pAkt-IR, the scores were correlated with tumour stage but not the Gleason score. In our hands, a significant correlation with tumour stage is also seen, but this is lost when the correlation with Gleason score is taken into account. In smaller cohorts, Le Page et al. [14] did not find any correlation between pAkt-IR and either Gleason score or tumour stage. In contrast, Malik et al. [11] reported that reported that 23/25 cases with Gleason scores 8–10 showed a strong staining intensity for pAkt, whereas the corresponding numbers for Gleason scores 5–6 and 7 were 4/25 and 5/14, respectively. This group also reported a significant correlation between the pAkt staining intensity and the tumour Ki67 labelling index [24]. Thus taken together, there is evidence that tumour pAkt-IR levels are associated with disease severity at diagnosis, but that the degree of association varies between studies. This may be due to differences in the composition of the cohorts. The large cohort described in [9] was primarily composed of cases with Gleason score 6–7 (488 (~86%) patients of a total 570 scored for pAkt-IR), whereas in our studies, these Gleason scores were less predominant (57 [20%], 121 [43%] and 104 [37%] of the cases had Gleason scores of 4–5, 6–7 and 8–10, respectively).

**Tumour pAkt-IR has Limited Prognostic Usefulness as a Marker for Disease-specific Survival in Pca**

In their large cohort, Ayala et al. [9] reported that the 41 cases with a very high pAkt-IR had a poorer biochemical recurrence-free survival (median 97 months) than the remaining 529 cases (median 133 months). The authors also reported that the 8 cases with this cut-off for non-malignant pAkt-IR also had a poorer biochemical recurrence-free survival than the remaining 556 cases. The prognostic value of the tumour pAkt-IR was retained when only cases with Gleason scores 6 or 7 were investigated [9]. In smaller cohorts, both significant and non-significant effects of pAkt-IR upon biochemical recurrence-free survival and/or survival have been reported [12–14]. The tissue microarrays of [9,12,14] consisted of samples obtained at radical prostatectomy, and whilst the patients had not been treated prior to surgery, it is important to note that the prognostic value of pAkt-IR in these studies has been assessed in this class of patients. In the study of [13], the samples were obtained at either TRUS-guided biopsy, and the patients were treated during the follow-up period. In contrast, in the present study, the prognostic value of pAkt-IR has been assessed in patients followed by expectancy alone, the long follow-up time allowing assessment of disease-specific survival.

We [16,25] have previously used receiver operating characteristic (ROC) curves to find optimal cut-off values. Although this method, originally developed to aid detection of radar signals, is widely used to find cut-offs of biochemical markers for disease processes, it may not be strictly valid in cases where the “diagnosis” in question is disease outcome over time rather than the disease itself. Ayala et al. [9] found their cut-off values (at very high pAkt-IR), following “an extensive search for the optimal cut-offs”. We have elected to run Cox proportional-hazards regression analyses over the whole gamut of scores, and pick the cut-off with the highest significance as our choice. This method, although tedious to perform, does take into account the chronological aspect of the endpoint, and additionally shows in a simple manner the range of cut-offs where significant prognostic information is provided. Further, the use of a ROC favours the choice of a single cut-off, which may not always be appropriate: in the present study, the Cox proportional-hazards regression analyses suggest that for tumour Ki67-IR, two cut-offs may be more appropriate.

For a useful prognostic marker, there should be a relatively wide range of cut-off values that provide a significant discrimination of cases with good/poor prognoses. This criteria is simply to allow for application of the marker in clinical praxis, and analysis of the previously published data for tumour pEGFR-IR and Ki67-IR scores [18,21] indicate that these markers pass this test. For the whole dataset, the tumour pAkt-IR also has a wide range of significant cut-offs, although we did not see any prognostic value at a very high pAkt-IR, in contrast to the study of Ayala et al. [9]. One possible explanation of this difference is in the nature of the samples in their study and in the present study: it may be that biochemical recurrence following radical prostatectomy requires a higher level of pAkt than disease-specific survival in untreated patients. As an aside, it was noted that for both tumour pEGFR-IR and tumour pAkt-IR, the optimal cut-offs measured using Exp[B] as a discriminant were the same as those found using the Youden (optimal) score in 15-year ROC analyses (data not shown and...
In the case of non-malignant pAkt-IR, however, the cut-off window is extremely small, and it does not provide additional prognostic value to that seen by tumour pAkt-IR. Although at first sight the wide window of significant prognostic information is promising, the clinical usefulness of tumour pAkt-IR is limited, since it provides no prognostic information at all at Gleason scores 6–7, i.e. those cases where treatment decisions are the most difficult. The finding, however, that pAkt-IR does provide prognostic information at Gleason scores 8–10 is mechanistically interesting, since it would suggest that activation of this survival pathway adds to the negative prognosis seen in poorly differentiated tumours. Cases with a Gleason score 8–10 in the database have a significantly higher median Ki-67 index than those with lower scores [21]. Although the sample size is small (only 50 cases followed by active expectancy with Gleason scores 8–10 were scored for both tumour pAkt-IR and Ki67-IR), the finding that pAkt-IR does not provide additional prognostic information when Ki67-IR (expressed as the number of positive stained cells crossing 11 horizontal lines across the core) is taken into account (Table S3) is consistent with the suggestion that the prognostic significance of pAkt-IR per se in Gleason score cases 8–10 is related to the proliferative capacity of the cells with high pAkt-IR expression.

Tumour pAkt-IR Correlates with pEGFR-IR and Provides Separate Prognostic Information

In the present study we found that there was a high correlation between tumour pAkt-IR and pEGFR-IR scores, confirming the study of Koumakpayi et al. [17], and consistent both with the finding of a high (54%) co-expression of EGFR and pAkt in prostate tumour cells [10] and the known signalling interrelation-ship between these two parameters [19]. We have additionally been able to show using partial multiple regression analyses that this correlation is not due to a “third-party” correlation with either the Gleason score or Ki67-IR. In Du145 prostate cancer cells, EGF produces pronounced cell migration and characteristics resembling epithelial-mesenchymal transition in a manner involving activation of Akt [20]. Epithelial-mesenchymal transition-like

![Figure 4. Prognostic significance of tumour and non-malignant pAkt-IR for cases who were followed by expectancy. Panel A and B are for cases with Gleason scores 6–7 (n = 102); Panels C and D are for cases with Gleason scores 8–10 (n = 51). The Exp(B) values obtained from Cox proportional-hazards regression analyses (Panels A and C) and the Kaplan-Meier plots (Panels B and D) were determined as described in the legend to Fig. 2. The \( \chi^2 \) values are for the log-rank (Mantel-Cox) tests, with the P values shown: *P < 0.05, NSP ≤ 0.8. doi:10.1371/journal.pone.0047994.g004]
states are believed to be an important factor in the ability of prostate cancer cells to metastasise [26]. Although the translation of data obtained in cultured cells to the clinical situation is far from easy, these data would predict that cases with high tumour pEGFR-IR and pAkt-IR scores would be more likely to have developed metastases at diagnosis and would be expected to have a poorer prognosis than cases where pEGFR-IR was high but pAkt-IR was low or where both parameters were low. The latter was found to be the case. When the Gleason score was taken into consideration, pEGFR-IR showed prognostic value for cases with Gleason scores 6–7 [10], in contrast to pAkt-IR. Taken together, our data are consistent with the hypothesis that the EGFR – Akt pathway is involved in the severity of the disease, but that additional EGFR pathways play an important role.

A final note concerns the lack of correlation between pAkt-IR and PDGFRβ in either the tumour or non-metastatic samples. PDGFRβ is a tyrosine kinase implicated in a number of cancers, including prostate cancer [22,27]. It has long been known that one of the pathways used by PDGFRβ is the Akt signalling pathway [28]. The lack of correlation between epithelial PDGFRβ-IR and pAkt-IR in the present study would suggest that this pathway is not a dominant pathway of PDGFRβ-mediated signalling in the prostate epithelial cells investigated here. It would of course be informative to investigate other receptors that affect growth of Pca cells and which involve Akt signalling in their actions. One such example is the insulin-like growth factor-1 receptor [29]. Similarly, given the finding that a low pERK-IR may compound the influence of pAkt-IR upon biochemical recurrence in Pca [12], it would be informative to investigate this marker in our cohort.

In conclusion, the present study has demonstrated that although tumour pAkt-IR is associated with both disease severity and disease-specific survival, it is of limited clinical usefulness as a biomarker, since it does not provide prognostic information in Gleason 6–7 cases. The association with tumour pEGFR-IR is consistent with the hypothesis that a high activity along the EGFR-Akt signalling pathway facilitates epithelial-mesenchymal transition and thereby an adverse clinical outcome.

**Supporting Information**

**Figure S1** Distribution of pAkt-IR scores in tumour (T) and non-malignant (N) samples. Shown are the scores for 282 (tumour) and 240 (non-malignant tissue) cases, using bin widths of 0.5 IR units to group the samples. The median (with 25% and 75% percentile in brackets) scores were: tumour 2.75 (2.25–3.125) and 2.0625 (1.578–2.609). For the 189 cases scored for both tumour and non-malignant pAkt-IR, the Spearman’s rho value was 0.3103 (P<0.0001), and the median values were significantly different (P<0.0001, Wilcoxon matched-pairs signed rank test).

(TIF)

**Figure S2** Exp(B) values for tumour pEGFR-IR (Panels A and B) and Ki67-IR (Panels C and D) for patients followed by expectancy; influence of Gleason score. Sample sizes are: pEGFR-IR, Gleason scores 6–7 (A), n = 141; pAkt-IR, Gleason scores 8–10 (B), n = 59; Ki67-IR, Gleason scores 6–7 (C), n = 140; Ki67-IR, Gleason scores 8–10 (D), n = 61. For explanation of the symbols, see legend to Fig. 3.

(TIF)

**Table S1** Non-parametric partial coefficients for pairwise comparisons of pAkt-IR vs. clinical parameters with a single controlling factor.

(DOCX)

**Table S2** Non-parametric partial coefficients for pairwise comparisons of pAkt-IR vs. clinical parameters with a single controlling factor.

(DOCX)

**Table S3** COX proportional-hazards regression analyses for tumour pAkt-IR and Ki-67-IR for patients with Gleason scores 8–10 at diagnosis and followed by expectancy.

(DOCX)

**Acknowledgments**

The authors are grateful to Pernilla Andersson for expert technical assistance.

**Author Contributions**

Conceived and designed the experiments: PH MC CJF. Performed the experiments: PH. Analyzed the data: MC CJF. Contributed reagents/materials/analysis tools: PH AJ PS LE TG. Wrote the paper: PH MC CJF.

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