The androgen receptor cytosine-adenine-guanine repeat length contributes to the development of epithelial ovarian cancer

Xiangrui Meng¹, Peng Lu²,*, Zhi Chu³, Qingxia Fan¹

¹Department of Oncology, The First Affiliated Hospital of Zhengzhou University, Zheng Zhou, People’s Republic of China
²Department of Gastrointestinal Surgery, People’s Hospital of Zhengzhou, Zheng Zhou, People’s Republic of China
³General Hospital, Jinan Military Command, Jinan, Shandong Province, People’s Republic of China

*Co-first author

Correspondence to:
Qingxia Fan, e-mail: fan_qingxia@126.com

Keywords: ovarian cancer, AR, CAG, repeat, polymorphism

Received: June 17, 2015 Accepted: October 08, 2015 Published: October 20, 2015

ABSTRACT

Ovarian cancer is the main cause of death among women with gynecological malignancies. Androgen and its receptors play an important role in ovarian cancer pathogenesis. Here, we aim to evaluate the relationship between AR CAG and GGN repeat length polymorphisms and Epithelial Ovarian Cancer (EOC) risk in a two-stage, case-control study among Chinese women. The repeat length was analyzed as a categorical variable for CAG_A and GGN_A (average allele), CAG_S and GGN_S (shorter allele), CAG_L and GGN_L (longer allele), respectively. The median value of the repeat length among the controls was used as the cutoff point. Women with longer AR CAG repeats had a decreased risk of developing EOC. The results were replicated in an independent sample. Compared to those with shorter (<22) CAG_A repeat length, women with longer (≥22) CAG_A repeats length had a 31% decreased EOC risk (OR = 0.69, 95% CI: 0.62–0.77, \( P = 5.06 \times 10^{-11}\)). For CAG_S and CAG_L, the results remain consistent. However, we didn’t detect any significant associations for GGN_A, GGN_S, and GGN_L. This should be the first study to examine the association between AR repeat length polymorphisms and ovarian cancer risk in a relatively large group of Asian women.

INTRODUCTION

According to American Cancer Society’s report, ovarian cancer ranks 5th overall for cancer death in women, accounting for 5% of all cancer deaths in women [1]. Although previous studies have demonstrated that some etiologic factors, including early age at menarche, late age at menopause, obesity, use of estrogen and hormone-replacement therapy, and inherited susceptibility, are positively associated with ovarian cancer, however, the common genetic variants explain less than 3.1% of the excess familial risk of EOC so additional susceptibility loci are likely to exist [2–5].

Epidemiologic and biological data suggest a role for androgens and androgen receptor (AR) in ovarian cancer development [6, 7]. The AR is a nuclear transcription factor that mediates the actions of testosterone and dihydrotestosterone [8]. Since the early studies with tritiated DHT exchange assays carried out by the McGuire’s laboratory in the seventies, the presence of AR protein in cancer specimens has been detected [9]. In particular, two microsatellite polymorphic variants (CAG)n and (GGN)n repeats were found in exon 1 of the AR gene [8, 10]. Together, these polymorphisms make 90% of women heteroygotic for the AR gene. The CAG trinucleotide repeat codes for a polyglutamine tract which normally ranges from 6 to 39 repeats [11]. Molecular analyses have shown that the transactivation capacity of the AR decreases with increasing number of glutamines encoded by the CAG repeat tract [12]. The biological effects of changing the GGN repeat length have not been as widely studied as those of CAG. Studies found that AR protein levels were inversely affected by the GGN repeat length due to the reason of the GGN repeat forming a hairpin structure in AR mRNA [13].

There are several published studies that addressed the association between AR repeat polymorphisms and ovarian cancer [6, 12, 14–17]. However, none was...
conducted in non-Asians. Considering the important role of AR repeat polymorphisms in the carcinogenic process, we carried out a two-stage case-control study in a Chinese population to investigate the possible relationship between these two AR repeat polymorphisms and the risk of Epithelial ovarian cancer (EOC) in the Chinese population.

RESULTS

Table 1 presents the characteristics of the study population included in this study. In total, there were 2,795 cases and 2,800 controls included in this two-stage study. No significant differences were observed between cases and controls in distribution of age, education, smoking, and body mass index (BMI) in both stages, indicating satisfactory matching for the case-control studies. We found that there were significant differences in usage of hormone replacement therapy between cases and controls (Stage1: \( P = 0.028 \); Stage1: \( P = 0.0009 \)), which indicated that usage of hormone replacement therapy might be a potential risk factor for EOC risk. The most histological subtype was serous (71% in Stage 1 and 73% in Stage 2). Figures 1–6 showed the distribution of CAG and GGN repeat sequences between the control group and EOC patients, respectively. Range of allele length have been given for both the shorter and longer allele.

Table 2 presents the associations between AR CAG and GGN repeat length and EOC risk in stage 1. For CAG polymorphism, women in the category of longer (≥22) CAG_A repeats had a significant 34% decreased EOC risk (OR = 0.66, 95% CI: 0.57–0.75, \( P = 1.32 \times 10^{-9} \)), when compared to those with the shorter (<22) CAG_A repeat length. The results were consistent and also statistically significant for the CAG_L and CAG_S alleles. For GGN polymorphism, we didn’t detected any significant associations for GGN_A, GGN_S, and GGN_L. Then

| Category | Stage 1 | 
|----------|---------|
|          | Cases (\( N = 1,925 \)) | Controls (\( N = 1,900 \)) | \( P \) Value |
| Age (yr) | Range | 35–74 | 34–72 | - |
|          | Mean ± SD | 51.0 ± 7.5 | 50.9 ± 7.1 | 0.672 |
| Education | less than middle school | 764 (39.7%) | 709 (37.3%) | 0.132 |
|          | middle school and above | 1,161 (60.3%) | 1,191 (62.7%) | |
| Ever smoker | Yes | 279 (14.5%) | 268 (14.1%) | 0.731 |
|          | No | 1,646 (85.5%) | 1,632 (85.9%) | |
| Use of hormone replacement therapy | Yes | 81 (4.2%) | 55 (2.9%) | 0.028 |
|          | No | 1,844 (95.8%) | 1,845 (97.1%) | |
| Body mass index (kg/m²) | 24.0 ± 3.7 | 23.8 ± 3.0 | 0.211 |
| Histopathology | Serous | 1,367 (71%) | 635 (73%) |
|          | Mucinous | 154 (8%) | 78 (9%) |
|          | Clear cell | 116 (6%) | 44 (5%) |
|          | Endometrioid | 192 (10%) | 113 (13%) |
|          | Others | 96 (5%) | 0 (0%) |
The association of AR CAG repeat length with EOC risk was validated in an independent stage 2 samples. As shown in Table 3, the significant associations for CAG_A, CAG_L and CAG_S were replicated. When combined together, women with longer (≥22) CAG_A repeats had a significant 31% decreased EOC risk (OR = 0.69, 95% CI: 0.62–0.77, $P = 5.06 \times 10^{-11}$), when compared to those with the shorter (<22).

**DISCUSSION**

In this two-stage, case-control study, we examined the relationship between AR CAG and GGN repeat polymorphisms and EOC risk in Chinese women. We identified that women with longer AR CAG repeats had a decreased risk of developing EOC. To our knowledge, this is the first study to examine the association between AR repeat length polymorphisms and ovarian cancer risk in a relatively large group of Asian women.

The AR is codified by the AR gene which is located on the X chromosome (q11.2–q12). Two length polymorphisms were located at the exon 1: a 9–39 CAG repeat (polyglutamine, polyQ) and a 14–27 GGN repeat (polyglycine, polyG). Together, these polymorphisms make 90% of women heterozygotic for the AR gene [13]. First, a role in cancer predisposition for AR repeat polymorphisms was suggested by inverted associations between prostate cancer risk and CAG repeat length within the AR transactivation domain in 1995 [18]. Then, several studies aimed to address the associations between AR repeat polymorphism and ovarian cancer risk previously, although the results were inconsistent [6, 12, 14,
Spurdle1 et al [20] first showed no evidence for an association between ovarian cancer risk and the genotype defined by the CAG polymorphism, although they cannot exclude small effects, or threshold effects in a small subgroup. Then, Dagan et al [19] explore the association of AR CAG repeat length with risk of ovarian cancer in Jewish Israeli women who are BRCA1/2 mutation carriers, and null results were concluded due to the small sample size. Since, several studies published recently found shorter CAG repeat length could increase risk of ovarian cancer [12, 14, 24, 25], while two studies published previously showed inversed results [6, 23]. One possible reason is the ethnically diverse populations, for which potential lifestyle and cultural factors, as well as other genetic factors, that may modify risk. Another reason is the big difference of the sample sizes, which may result in false results. Also, observed associations in some studies were of marginal significance. Therefore, these results could potentially be due to chance. In current study, we identified that women with longer AR CAG repeats had a decreased risk of developing EOC, using a large sample size, two-stage design. This results was also supported by another study by Li et al [26], who reported that short androgen receptor allele length is a poor prognostic factor in epithelial ovarian carcinoma.

Strength of this study includes large sample size and the two-stage study design, which is more efficient on discovering potential risk factors. Limitations of our study, including potential biases and population generalizability, should also be taken attentions. Because the study population is primarily Chinese women and our analyses are restricted to Chinese, our results are not generalizable.
to other ethnicities. However, the homogeneity of our population is an advantage. Due to our case-control design, we must also consider the possibility of selection bias.

Our findings suggest that shorter CAG repeat length, modulates the actions of androgens in ovarian neoplasia and tumor growth. These data add to the growing body of evidence linking androgenicity to the pathogenesis and tumor biology of ovarian cancer. Functional studies characterizing hormone activity and response pathways may reveal specific mechanisms in which androgens function in ovarian cancer biology.

**MATERIALS AND METHODS**

**Subjects**

There were 1,925 EOC cases and 1,900 cancer-free controls included in Stage 1, and Stage 2 included an additional EOC 870 cases and 900 controls. All subjects were genetically unrelated ethnic Han Chinese. Controls were frequency-matched to the cases by age (the range of five years), and residential area. All cases were newly diagnosed according to International Classification of Diseases for Oncology (ICD-O) codes. All the controls were genetically unrelated with cases. Subjects with histories of cancer were excluded from the control group. We also obtained clinical information for all the cases. After informed consent was obtained, a questionnaire about lifestyle factors was completed by all the subjects through face-to-face interviews. After the interview, each subject provided 3–5 mL of venous blood. Approval for this study was obtained from the institutional review boards. Written informed consent was obtained from all participants or from the patients’ representatives.

**Genotyping**

DNA from blood of patients and control specimens was extracted from white blood cell fractions using the
Qiagen Blood Kit (Qiagen, Valencia, CA). Then DNA was PCR-amplified using fluorescently labeled primers (F: 5′-TCCAGAATCTGTTCCAGAGCGTGC-3′; R: 5′-GCTGTGAAGGTTGCTGTTCCTCAT-3′) for the AR CAG repeat polymorphism, while using the primers (F: 5′-CGGTTCTGGGTCACCCTC A-3′; R: 5′-TCACCATGCCGCCAGGTA-3′) for AR GGN repeat polymorphism, using the conditions as follows: with an initial denaturation step at 96°C followed by 35 cycles of 1 min at 96°C, 45 s at 61°C, and 2 min at 72°C, and a final extension step for 5 min at 72°C. Fragments were then run on denaturing polyacrylamide gels on the Applied Biosystems Prism 3700XL and analyzed by Applied Biosystems Prism Genescan automated fluorescence detection (Applied Biosystems, Foster City, CA).

**Statistical analyses**

We used the Chi-square test to examine the differences in the frequency distributions of categorical variables between groups. The Shapiro-Wilk test was used to verify the normality of distribution, and Levene’s test for equality of variances. When both of these assumptions were met, t-test was used to test the differences for the AR repeat lengths between case patients and control subjects. Otherwise, the Mann-Whitney U test was used. Odds ratios (ORs) and their 95% CI were calculated from unconditional logistic regression analyses. The repeat length was analyzed as a categorical variable for CAG_A and GGN_A (average allele), CAG_S and GGN_S (shorter allele), CAG_L and GGN_L (longer allele), respectively. The median value of the repeat length among the controls was used as the cutoff point. If some females have two alleles for the same repeat number, average allele, shorter allele, and longer allele share the same number without identifying which is longer. All statistical tests were 2-sided and a P value significance threshold of 0.05 was set. All analyses were conducted using SPSS 19.0.

### Table 2: Association of AR repeat length with EOC risk in Stage 1

|                | Cases, n (%) | Controls, n (%) | OR (95% CI)* | P     |
|----------------|--------------|-----------------|--------------|-------|
| **CAG repeat** |              |                 |              |       |
| CAG_A          |              |                 |              |       |
| <22            | 728 (37.8%)  | 543 (28.6%)     | Referent     |       |
| ≥22            | 1197 (62.2%) | 1357 (71.4%)    | 0.66 (0.57–0.75) | 1.32 × 10⁻⁹ |
| CAG_S          |              |                 |              |       |
| <19            | 1061 (55.1%) | 891 (46.9%)     | Referent     |       |
| ≥19            | 864 (44.9%)  | 1009 (53.1%)    | 0.72 (0.63–0.82) | 3.66 × 10⁻⁷ |
| CAG_L          |              |                 |              |       |
| <23            | 1132 (58.8%) | 935 (49.2%)     | Referent     |       |
| ≥23            | 793 (41.2%)  | 965 (50.8%)     | 0.68 (0.60–0.77) | 2.63 × 10⁻⁹ |
| **GGN repeat** |              |                 |              |       |
| GGN_A          |              |                 |              |       |
| ≤23            | 584 (67.1%)  | 578 (64.2%)     | Referent     |       |
| >23            | 286 (32.9%)  | 322 (35.8%)     | 0.87 (0.72–1.07) | 0.198  |
| GGN_S          |              |                 |              |       |
| ≤23            | 595 (68.4%)  | 602 (66.9%)     | Referent     |       |
| >23            | 275 (31.6%)  | 298 (33.1%)     | 0.93 (0.77–1.14) | 0.500  |
| GGN_L          |              |                 |              |       |
| ≤23            | 573 (65.9%)  | 554 (61.5%)     | Referent     |       |
| >23            | 297 (34.1%)  | 346 (38.5%)     | 0.83 (0.68–1.01) | 0.060  |

*adjusted for age, education and Use of hormone replacement therapy
Table 3: Association of AR repeat length with EOC risk in Stage 2 and combined results

| CAG repeat | Cases, n (%) | Controls, n (%) | OR (95% CI)* | P       |
|------------|--------------|-----------------|--------------|---------|
|             |              |                 |              |         |
| Stage 2     |              |                 |              |         |
| CAG_A       |              |                 |              |         |
| <22         | 320 (36.8%)  | 275 (30.6%)     | Referent     |         |
| ≥22         | 550 (63.2%)  | 625 (69.4%)     | 0.76 (0.62–0.92) | 0.006   |
| CAG_S       |              |                 |              |         |
| <19         | 470 (54.0%)  | 441 (49.0%)     | Referent     |         |
| ≥19         | 400 (46.0%)  | 459 (51.0%)     | 0.82 (0.68–0.99) | 0.03    |
| CAG_L       |              |                 |              |         |
| <23         | 480 (55.2%)  | 445 (49.4%)     | Referent     |         |
| ≥23         | 390 (44.8%)  | 455 (50.6%)     | 0.79 (0.66–0.96) | 0.01    |
| Combined    |              |                 |              |         |
| CAG_A       |              |                 |              |         |
| <22         | 1048 (37%)   | 818 (29%)       | Referent     |         |
| ≥22         | 1747 (63%)   | 1982 (71%)      | 0.69 (0.62–0.77) | 5.06 × 10⁻¹¹ |
| CAG_S       |              |                 |              |         |
| <19         | 1531 (55%)   | 1332 (48%)      | Referent     |         |
| ≥19         | 1264 (45%)   | 1468 (52%)      | 0.75 (0.67–0.83) | 7.02 × 10⁻⁸  |
| CAG_L       |              |                 |              |         |
| <23         | 1612 (58%)   | 1380 (49%)      | Referent     |         |
| ≥23         | 1183 (42%)   | 1420 (51%)      | 0.71 (0.64–0.79) | 3.18 × 10⁻¹⁰ |

*adjusted for age, education and Use of hormone replacement therapy

ACKNOWLEDGMENTS AND FUNDING

We thank all subjects who were involved in the study.

CONFLICTS OF INTEREST

The authors declare that they have no conflict of interest.

Authors’ contributions

FQ, MX, and CZ participated in the design, interpretation of the studies and analysis of the data and review of the manuscript; MX, LP and CZ conducted the survey and experiments, wrote the manuscript.

REFERENCES

1. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2015. CA Cancer J Clin. 2015; 65:5–29.
2. Kuchenbaecker KB, Ramus SJ, Tyrer J, Lee A, Shen HC, Beesley J, Lawrenson K, McGuffog L, Healey S, Lee JM, Spindler TJ, Lin YG, Pejovic T, Bean Y, Li Q, Coetzee S, et al. Identification of six new susceptibility loci for invasive epithelial ovarian cancer. Nat Genet. 2015; 47:164–171.
3. Barger CJ, Zhang W, Hillman J, Stablewski AB, Higgins MJ, Vanderhyden BC, Odunsi K, Karpf AR. Genetic determinants of FOXM1 overexpression in epithelial ovarian cancer and functional contribution to cell cycle progression. Oncotarget. 2015; 6:27613–27. doi: 10.18632/oncotarget.4546.
4. Lam F, Abbas AY, Shao H, Teo T, Adams J, Li P, Bradshaw TD, Fischer PM, Walsby E, Pepper C, Chen Y, Ding J, Wang S. Targeting RNA transcription and translation in ovarian cancer cells with pharmacological inhibitor CDKI-73. Oncotarget. 2014; 5:7691–7704. doi: 10.18632/oncotarget.2296.
5. Al-Wahab Z, Tebbe C, Chhina J, Dar SA, Morris RT, Ali-Fehmi R, Giri S, Munkarah AR, Rattan R. Dietary...
energy balance modulates ovarian cancer progression and metastasis. Oncotarget. 2014; 5:6063–6075. doi: 10.18632/oncotarget.2168.

6. Terry KL, De Vivo I, Titus-Ernstoff L, Shih MC, Cramer DW. Androgen receptor cytosine, adenine, guanine repeats, and haplotypes in relation to ovarian cancer risk. Cancer Res. 2005; 65:5974–5981.

7. Sun NK, Huang SL, Chang PY, Lu HP, Chao CC. Transcriptomic profiling of taxol-resistant ovarian cancer cells identifies FKBP5 and the androgen receptor as critical markers of chemotherapeutic response. Oncotarget. 2014; 5:11939–11956. doi 10.18632/oncotarget.2654.

8. Fujimoto J, Hirose R, Sakaguchi H, Tamaya T. Expression of size-polymorphic androgen receptor (AR) gene in ovarian endometriosis according to the number of cytosine, adenine, and guanine (CAG) repeats in AR alleles. Steroids. 1999; 64:526–529.

9. Allegra JC, Lippman ME, Thompson EB, Simon R, Barlock A, Green L, Huff KK, Do HM, Atién SC. Distribution, frequency, and quantitative analysis of estrogen, progesterone, androgen, and glucocorticoid receptors in human breast cancer. Cancer Res. 1979; 39:1447–1454.

10. Sleddens HF, Oostra BA, Brinkmann AO, Trapman J. Trinucleotide (GGN) repeat polymorphism in the human androgen receptor (AR) gene. Hum Mol Genet. 1993; 2:493.

11. Buchanan G, Yang M, Cheong A, Harris JM, Irvine RA, Lambert PF, Moore NL, Raynor M, Neufing PJ, Coetzee GA, Tilley WD. Structural and functional consequences of glutamine tract variation in the androgen receptor. Hum Mol Genet. 2004; 13:1677–1692.

12. Schildkraut JM, Murphy SK, Palmieri RT, Iversen E, Moorman PG, Huang Z, Halabi S, Calingaert B, Gusberg A, Marks JR, Berchuck A. Trinucleotide repeat polymorphisms in the androgen receptor gene and risk of ovarian cancer. Cancer Epidemiol Biomarkers Prev. 2007; 16:473–480.

13. Nicolas Diaz-Chico B, German Rodriguez F, Gonzalez A, Ramirez R, Bilbao C, Cabrera de Leon A, Aguirre Jaime A, Chirino R, Navarro D, Diaz-Chico JC. Androgens and androgen receptors in breast cancer. J Steroid Biochem Mol Biol. 2007; 105:1–15.

14. Ludwig AH, Murawska M, Panek G, Timorek A, Kupryjanczyk J. Androgen, progesterone, and FSH receptor polymorphisms in ovarian cancer risk and outcome. Endocr Relat Cancer. 2009; 16:1005–1016.

15. Terry KL, De Vivo I, Titus-Ernstoff L, Sluss PM, Cramer DW. Genetic variation in the progesterone receptor gene and ovarian cancer risk. Am J Epidemiol. 2005; 161:442–451.

16. Zhu T, Yuan J, Xie Y, Li H, Wang Y. Association of androgen receptor CAG repeat polymorphism and risk of epithelial ovarian cancer. Gene. 2015.

17. Kim SC, Ju W, Mahavni V, Geisler JP, Buller RE. CAG repeat length in exon 1 of the androgen receptor gene is related to age of diagnosis but not germ line BRCA1 mutation status in ovarian cancer. Int J Gynecol Cancer. 2006; 16:190–194.

18. Irvine RA, Yu MC, Ross RK, Coetzee GA. The CAG and GGC microsatellites of the androgen receptor gene are in linkage disequilibrium in men with prostate cancer. Cancer Res. 1995; 55:1937–1940.

19. Dagan E, Friedman E, Paperna T, Carmi N, Gershoni-Baruch R. Androgen receptor CAG repeat length in Jewish Israeli women who are BRCA1/2 mutation carriers: association with breast/ovarian cancer phenotype. Eur J Hum Genet. 2002; 10:724–728.

20. Spurdle AB, Webb PM, Chen X, Martin NG, Giles GG, Hopper JL, Chenexiv-Trench G. Androgen receptor exon 1 CAG repeat length and risk of ovarian cancer. Int J Cancer. 2000; 87:637–643.

21. Levine DA, Boyd J. The androgen receptor and genetic susceptibility to ovarian cancer: results from a case series. Cancer Res. 2001; 61:908–911.

22. Menin C, Banna GL, De Salvo G, Lazzarotto V, De Nicolò A, Agata S, Montagna M, Sordi G, Nicoletto O, Chieco-Bianchi L, D’Andrea E. Lack of association between androgen receptor CAG polymorphism and familial breast/ovarian cancer. Cancer Lett. 2001; 168:31–36.

23. Santarosa M, Bidoli E, Gallo A, Steffan A, Boiocchi M, Viel A. Polymorphic CAG repeat length within the androgen receptor gene: identification of a subgroup of patients with increased risk of ovarian cancer. Oncol Rep. 2002; 9:639–644.

24. Kassim S, Zoheiry NM, Hamed WM, Going JJ, Craft JA. Androgen receptor gene methylation and exon one CAG repeat length in ovarian cancer: differences from breast cancer. JUBMB Life. 2004; 56:417–426.

25. Li AJ, Elmore RG, Pavelka JC, Karlan BY. Hyperandrogenism, mediated by obesity and receptor polymorphisms, promotes aggressive epithelial ovarian cancer biology. Gynecol Oncol. 2007; 107:420–423.

26. Li AJ, Baldwin RL, Karlan BY. Short androgen receptor allele length is a poor prognostic factor in epithelial ovarian carcinoma. Clin Cancer Res. 2003; 9:3667–3673.