SYNTHESIS OF SILICA NANOPARTICLES VIA GREEN APPROACH BY USING HOT AQUEOUS EXTRACT OF THUJA ORIENTALIS LEAF AND THEIR EFFECT ON BIOFILM FORMATION

M.Th.Azawi 1 S.M.Hadi 2 Ch.H.Mohammed 3
Researcher Asst. Prof Asst. Prof
Dept. of Physics1, 3 and Biology2 - Coll. of Science - Univ. of Baghdad

mthamir50@gmail.com, sabahtech2013@gmail.com, ghusun.hamed@yahoo.com

ABSTRACT:

There is great interest for NPs manufacturing by environmentally friendly and economic manner. The aqueous leaves extract of Thuja orientalis was used to synthesized silica nanoparticles (SiO2NPs) by using two green methods, the first one is the magnetic stirrer method and the second method by using the cold plasma. The XRD pattern for both samples annealed at T=600°C for 1h, while they showed the characteristic of Bragg peaks for poly phases at (001), (010), (10-1), (1-10), (0-12) and (002) for SiO2 triclinic (anorthic). The average crystalline size was calculated by using Scherer’s formula, which was 11.1868 nm in magnetic stirrer method, while the cold plasma method showed amorphous structure. The morphology analysis using atomic force microscopy showed that the grain size was 33.94 and 18.37 nm for magnetic stirrer and cold plasma methods respectively. Fourier Transform Infrared Spectroscopy (FTIR) analysis indicates hydrophilic functional groups in the capping matrix, which can improve the stability of silica NPs. The biofilm inhibition of silica NPs were investigated for two genus of bacteria Staphylococcus aureus and Escherichia coli, the green silica NPs that synthesized by using cold plasma method showed the highest inhibition effect on S. aureus and E. coli respectively.

Keywords: green synthesis, annealed, cold plasma, magnetic stirrer, bacteria.

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INTRODUCTION
Nanotechnology is the most growing, widespread scientific and technical achievement in the world. This science was emerged by applying of nanosize structures as a solution to the previous difficult problems. Different fluorescent silica NPs with similar composition and functionalization have emerged as a particularly fascinating fluorescent probe and attracted widespread interest in biology and medicine (41,50). Silica NPs have high thermal, chemical stability, high surface area and good biocompatibility, but nude silica NPs shows no detrimental effects on bacteria. Sometimes silica NPs may be used as a mediator to enhance delivery doses of antimicrobials such as copper, zinc and silver NPs at the target sites and therefor reduce the antibiotics doses intake (20). Most of the chemical methods that used for the synthesis of NPs are too expensive and also involve the use of toxic, hazardous chemicals that are responsible for various biological risks. This enhances the growing need to develop environmentally friendly processes through green synthesis and other biological approaches. Sometimes the synthesis of NPs using various plants extract can be more useful than the other biological methods, which involve very complex procedures of maintaining microbial cultures. On the other hand the using of plant extracts is the most adopted method because the plants are widely distributed, available and much safer to handle. In addition, the plants have various secondary metabolites compounds that consider as another source for natural reducing and capping agents (22). Thuja orientalis plant is an evergreen coniferous tree that widely distributed in East Asia (53), it belongs to Cupressaceae family and used in landscape. T. orientalis leaves have essential oils which are used in traditional medicine as antifungal, bactericide and other properties like eradicate parasitic worms. Normally the oil is toxic, mainly for the presence of α- thujone, the other leaves compounds are; rhodoxanthin, amentoflavone, hinokilflavone, quercetin, myricetin, carotene, xanthophylls and ascorbic acid (10,49). In this study, we have used the green method in synthesis of silica NPs using T. orientalis aqueous leaf extract and TEOS for silica precursor as a sustainable, cheap and available material for synthesis of silica NPs in an environmentally friendly manner.

Synthesis chemistry of silica
The magnetic stirrer and cold plasma methods means the synthesis of an inorganic network by a chemical reaction produced in the solution at low temperature. The major advantage of this method is that it offers the possibility to obtain hyaloids solids, which are very difficult to be obtained by conventional techniques of burning at high temperatures and offers the possibility to obtain materials with predetermined structure, depending on the conditions of the experiment. The development of silica synthesis process in materials science starts with a solution of silicon alkoxide compound Si(OR)n as a precursor that melted in an alcohol or other low-molecular weight organic solvent, were R is an alkyl group (C3H6n+1) (8,52). Compared with colloid chemistry, the alkoxide route can be more easily controlled by controlling hydrolysis 1 and condensation reactions (2a-water condensation, 2b-alcohol condensation)

\[
\begin{align*}
\text{Hydrolysis} & \quad \text{Si} - \text{OR} + \text{HOH} \leftrightarrow \text{Si} - \text{OH} + \text{ROH} & (1) \\
\text{Water} & \quad \text{Condensation} \\
\text{Hydrolysis} & \quad \text{Si} - \text{OH} + \text{Si} - \text{OH} \leftrightarrow \text{Si} - \text{O} - \text{Si} + \text{HOH} & (2a) \\
\text{Alcohol} & \quad \text{Condensation} \\
\text{Alcoholysis} & \quad \text{Si} - \text{OH} + \text{Si} - \text{OR} \leftrightarrow \text{Si} - \text{O} - \text{Si} + \text{ROH} & (2b) \\
\end{align*}
\]

\[R = \text{C}_3\text{H}_5\]
The hydrolysis reaction 1-4, consists in replacing of the alkoxide groups (–OR) with hydroxyl groups (–OH) and releasing of the corresponding ROH alcohol molecules. A complete hydrolysis 4 is obtained when the stoichiometric molar ratio water: Si(OR)₃ is 4. Any intermediate species [(OR)₂-Si-(OH)] or [(OR)₃-Si- (OH)] would be considered the result of partial hydrolysis 3a, 3b. A small amount of water leads to a slow hydrolysis due to the reduced reactant concentration. A large amount of water gives a slow hydrolysis due to the increased reactant dilution. Subsequent condensation reactions involve the silanol groups (Si-OH), produce siloxane bonds (Si-O-Si) and water and alcohol as by-products:

(OR)₃-Si-OH + HO-Si(OR)₃ → [(OR)₂-Si-O-Si(OR)₃] + H₂O-H

(3a)

(OR)₃-Si-OR + HO-Si(OR)₃ → [(OR)₂-Si-O-Si(OR)₃] + R-OH

(3b)

Si(OR)₃ + 4 H₂O → Si(OH)₄ + 4 R-OH

Si(OH)₄ → SiO₂ + 2H₂O

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Floating electrode dielectric barrier discharge (FE-DBD) plasma technique

The samples were prepared by using the floating electrode dielectric barrier discharge (FE-DBD) plasma technique, which was manufactured by Dr. Hamid H. Murbit in the College of Science for Women/University of Baghdad. The floating electrode dielectric barrier discharge (FE-DBD) system, constructed similarly to conventional dielectric barrier discharges (DBDs) is inherently offers better potential for diversity, since it works in room temperature, air (normal atmospheric pressure) and it has a cold touch (15,29). This system operates at power densities of 0.1-2 W/cm². FE-DBD operates under the conditions where one of the electrodes is a dielectric-protected powered electrode and the second active electrode is the organ, liquid surface, animal or human skin without second surface present discharge does not ignite (22). In the FE-DBD setup, the second electrode is not grounded and remains at a floating potential. Of note is the fact that FE-DBD is completely safe from the electrical perspective and non-damaging for application to animal or human skin delicate surfaces which are likely to be damaged by thermal (hot) plasma. Discharge ignites when the powered electrode approaches the surface to be estimated (treated) at a distance (discharge gap) less than about 3 mm, depending on the form, duration, and polarity of the driving voltage (26). Power deposited into plasma discharge gap was analyzed by measuring current passing through the discharge gap and the voltage drop in the gap (17).

MATERIALS AND METHODS

Tetraethyl orthosilicate (TEOS) was obtained from (Aldrich chemical, purity > 98%). All the glass-wares were washed with concentrated hydrochloric acid then deionized water (diH₂O). The Milli-Q ultrapure deionized water with electrical conductivity (E.c = 0.7 µs cm⁻¹) was used for the all purposes.

Preparation of T. orientalis aqueous leaf extract

The leaves of T. orientalis were collected from the University of Baghdad gardens. Plant parts were washed several times by using tap water then by distilled water to remove all dirt then dried at room temperature. The samples were grinded into powder by using electrical grinder; 10 gm of dried powder was added to 100 ml of diH₂O. After boiling for 10 min then homogenized on the magnetic stirrer for 4h then the mixture was filtered with filter paper Whatman No.1. The extract was centrifuged at 5000 rpm for 15 min and storied at 4 °C until use.

Preparation of green silica nanoparticles by using magnetic stirrer method

Silica NPs were synthesized by added 15 ml of TEOS (10⁻³M) as silica precursor in a flask placed in a water bath with continuous stirring until the temperature stabilized at 60°C then 5 ml of T. orientalis aqueous leaf extract was added. The reaction was lasted for 30 min with continuous stirring, as shown in Figure 1a.

Preparation of green silica nanoparticles by using cold plasma method

Silica NPs were synthesized by added 15 ml of TEOS (10⁻³M) with 5ml of T. orientalis leaf extract, the mixture was left under the influence of plasma electrode for 5 min. In this experiment, continuous wave plasma high voltage (4 kV) was applied to a quartz protected electrode that generates plasma between the quartz and the liquid surface as shown in figure (1b). Discharge ignites when the powered electrode approaches the surface to be estimated (treated) at a distance (discharge gap) less than about 2 mm. In this
way, we obtain a light yellow nanoparticle solution.

![Image](a)
![Image](b)

**Figure 1. Green synthesis of SiO$_2$ NPs.** a) Synthesis SiO$_2$ NPs by using magnetic stirrer method. b) Synthesis SiO$_2$ NPs by using cold plasma method.

**Determination of antimicrobial activity by agar well diffusion method.**

The antimicrobial activity of silica nanoparticles were determined by using agar diffusion method against the pathogenic bacteria isolates (S. aureus and E. coli). The Muller Hinton Agar plates were inoculated with $0.5 \times 10^8$ (CFU/ml) of S. aureus and E. coli strain by using cotton swabs under aseptic conditions.

**The effect of silica nanoparticles on biofilm formation.**

Biofilm formation assays were performed using 96-well microtiter plate, based on the protocol by Goh, S. et al (19), with slight modifications. E. coli and S. aureus bacteria were cultured in tryptone soya broth (TSB) for 18-24 hrs and the resulting culture was adjusted to 0.5 McFarland tube. All the wells of microtiter plate were loaded with 100µl of TS broth with 100µl of nanoparticles solution except the control filled only TS broth, then the plate was incubated at 37°C for 24 h. All the NPs samples were tested three times. The microbial planktonic was removed by upset the plate over a waste tray then 0.1% w/v crystal violet solution was added to each well and left for 10 min at room temperature. The dye was removed by submerging the plate in a water tray, then was inverted and left in air to dry. The wells were treated with 95% v/v ethanol for 10 min to solubilize the dye. Optical density (OD) was measured at 630 nm (1).

**RESULTS AND DISCUSSION**

**X-ray diffraction analysis.**

The diffraction pattern gives information on translational symmetry - size and shape of the unit cell from peak positions and information on electron density within the unit cell, namely, where the atoms are located from peak intensities. It also, gives information on the deviations of perfect particles, if the size is less than approximately 100-200 nm, extended defects and micro strain from peak shapes and widths (28). Figure 2 shows the XRD pattern for samples which are prepared by tow method magnetic stirrer and cold plasma using the T. orientalis leaves extract in green method, annealed to 600°C for 1h (5). All parameters value resulted from XRD is shown in Table 1. It can be observed that all diffraction peaks of poly phases (001), (010), (10-1), (1-10), (0-12) and (002) facets can be indexed as the typical triclinic structure for silica. The crystallite sizes of silica NPs calculated for all peaks by using Scherer’s formula. It was found that the value of average crystalline size 11.1868 nm because this the size that lies within the NPs as shown in Table 1, these results are agreed with Boisen M. B. et al. (6). While The XRD diffractogram for synthesis silica NPs by using cold plasma is shown in Figure 2, which released that the broad peak in the diffraction angle range of (2θ=15-30°) who is centered at 22° attributed to the amorphous structure for SiO$_2$ NPs, without any defined peaks due to the existing of crystalline structure (18,32). The production of amorphous silica nanoparticles contributed to the positive impact as it has broad applications in our daily life. This is due to the special characteristics of amorphous silica nanoparticles that are stable in a long time compared to the crystalline silica (3). Secondly, amorphous silica is more reactive than crystalline silica. This is because of the hydroxyl group in the amorphous region is more applicable to use in the reaction compared to their crystalline region (51). These results were agreed with Mohd N. K. et al. (36).
Figure 2. The XRD pattern of SiO$_2$ NPs green synthesis

Table 1. X-ray diffraction data for Silica NPs

| Synthesis methods       | 2θ (Deg.) | FWHM (Deg.) | d$_{hkl}$ Exp.(Å) | G.S. (nm) | hkl   | d$_{hkl}$ Std.(Å) | Phase | Card No.          |
|-------------------------|-----------|--------------|-------------------|-----------|-------|------------------|-------|------------------|
| Magnetic stirrer        | 14.1683   | 0.7593       | 6.2460            | 10.5      | (001) | 6.2332           | Tri. SiO$_2$ | 96-900-6299     |
|                         | 16.9059   | 0.7781       | 5.2402            | 10.3      | (010) | 5.2328           | Tri. SiO$_2$ | 96-900-6301     |
|                         | 18.5419   | 0.3981       | 4.7814            | 20.2      | (10-1) | 4.8181           | Tri. SiO$_2$ | 96-900-6301     |
|                         | 25.4311   | 0.9279       | 3.4996            | 8.8       | (1-10) | 3.4707           | Tri. SiO$_2$ | 96-900-6299     |
|                         | 28.1547   | 1.3549       | 3.1669            | 6.0       | (0-12) | 3.1729           | Tri. SiO$_2$ | 96-900-6299     |
|                         | 32.5      | 1.3010       | 2.7570            | 6.4       | (002) | 2.7228           | Tri. SiO$_3$ | 96-900-6301     |

Cold plasma

Amorphous

Atomic force microscopy (AFM) measurement

Two and three dimensional (2D and 3D) profile of AFM was used to give information of surface morphology to materials. Figure 3a shows the surface morphology of silica NPs prepared by hydrothermal method, which notes that the particles were spherical, while Figure 3b refers to the granular distribution of silica NPs. Figure 4a show 2D and 3D the atomic force microscopy images that synthesized by using cold plasma method, it was observed that the shape of the particles was spherical. While Figures 4b refer to the granular distribution of silica nanoparticles of the same method. It is found the size of the grains was 33.94 nm and 18.37 nm for the samples that prepared by using the magnetic stirrer and cold plasma method respectively. The smaller silica nanoparticles have many positive attributes, such as good chemical stability and biofilm, which would make them suitable for many practical applications.
AFM imaging of the nanoparticles was performed by drying on a clean glass slide substrates. The surface morphology has been investigated using images of AFM which produces topological images of surface at very high magnification. Images of AFM are widely known, providing a useful tool for unambiguously describing the size and distribution of nanoparticle size. The results of AFM measurement are shown in Table 2. The increase in grain size of SiO$_2$ NPs by using magnetic stirrer method by is due to the increasing temperature of the magnetic stirrer (37).

**Table 2. The grain size of SiO$_2$ NPs**

| Synthesis methods | Average grain size (nm) |
|-------------------|-------------------------|
| Magnetic stirrer   | 33.94                   |
| Cold plasma       | 18.37                   |

From the above table it can be noted the increasing that happened in grain size for the samples was prepared by green method that used the magnetic stirrer due to the use of temperature in preparation, where high temperature increases crystalline growth of the material, so there is an increase in grain size.

Fourier transform infrared spectroscopy (FTIR)

The compounds were characterized via FTIR in Shimadzu IR-Affinity-1 (Japan) Spectrometer. The samples were mixed with potassium bromide and examined directly in their powder state without further preparation. The FTIR measurements of biosynthesized silica NPs were carried out to identify the possible interaction between protein and silica NPs. Results of FTIR study showed sharp absorption peaks located at about (1626, 1606.7 and 1608.6) and (3423.65, 3404.36 and 3398.57) cm$^{-1}$ Figures 5. Absorption peak at (1626, 1606.7 and 1608.6) cm$^{-1}$ may be assigned to the amide I bond of proteins arising due to carbonyl stretch in proteins, and peaks at (3423.65, 3404.36 and 3398.57) cm$^{-1}$ are assigned to OH stretching in alcohols and phenolic compounds (25). The absorption peak at (1626, 1606.7 and 1608.6) cm$^{-1}$ is close to that reported for native proteins (33). This evidence suggests that proteins are interacting with biosynthesized nanoparticles and also their secondary structure was not affected after binding with NPs (14). Phenolic compounds belonging to the lignin’s group have been earlier reported to be capable of chelating with elements to form complexes (7). Thus, it can be concluded that hydroxyl and carboxyl groups present in phenolic compounds of the *T. orientalis* leaf extract. These results confirm the presence of phenols and proteins which may act as stabilizing agents for NPs (47). Broad peak was observed around 3466 cm$^{-1}$ proved the presence of O-H stretching vibration due to the vibration of the silanol group on the silica surfaces. This finding was consistent with the previous study reported on silica NPs (2). Both of the spectra of silica NPs synthesis do not show any significant difference to each other.

![Figure 4. (a) 3D & 2D AFM images of SiO$_2$ NPs by using cold plasma method. (b) Granularity distribution of SiO$_2$ NPs by using cold plasma method.](image)

![Figure 5. FTIR spectra for prepared SiO$_2$ NPs](image)
Table 3. FTIR data for SiO₂ NPs

| Bonds                                      | Aqueous leaf extract | Magnetic Stirrer | Cold plasma |
|--------------------------------------------|---------------------|-----------------|------------|
| Si-O-Si bending (35,42,24)                 | 451.34              | 478.35          |            |
| Si-H bond (21,31)                          | 617.22              | 615.29          |            |
| Shoulder (Si-O-Si) asymmetric stretch (36,42,21,31,40,39,46,27) | 1066.64             | 1070.49         | 1064.71    |
| C–N stretch (aromatic amines) (47)         | 1384.50             | 1377.17         | 1375.25    |
| C-H bending (46,11)                        | 1436.97             |                 |            |
| The amide I bond of proteins arising due to carbonyl stretch in proteins (11,12,13,28,47) | 1625.99             | 1606.70         | 1608.63    |
| –COO– groups (35)                         | 1732.08             |                 |            |
| CO vibration (30,38)                      | 2364.73             |                 |            |
| C=O vibrations (35)                       | 2927.94             | 2931.80         | 2918.30    |
| Silanol (Si-O) stretching and Absorb H₂O on surface or -OH stretching and phenolic compounds with strong H bond (36,47,2,25) | 3423.65             | 3404.36         | 3398.57    |

The effect of SiO₂ NPs on biofilm formation

The results showed that a difference found in biofilm formation depends on the grain size of nanoparticles and the genus of bacteria as shown in Table 4. Each of the samples of silica nanoparticles that synthesized by using magnetic stirrer and cold plasma methods with Thuja orientalis aqueous leaf extract presented high inhibition effect against the entire bacteria compared with the control. The silica nanoparticles that synthesized by using cold plasma method had displayed the highest inhibition effect on the formation of biofilm by S. aureus and E. coli compared with the control (0.146333, 0.143333, 0.2773 and 0.2), and then followed by aqueous leaf extract of Thuja orientalis that presented inhibition effect against the biofilm formation (0.12, 0.125, 0.2773 and 0.2) respectively on the bacteria.

Table 4. The effect of different SiO₂ NPs on biofilm growth on bacteria

| Synthesis methods          | S. aureus | E. coli |
|----------------------------|-----------|---------|
| Control                    | **0.2773  | *0.2    |
| Aqueous leaf extract       | 0.12      | 0.125   |
| Magnetic stirrer           | 0.146333  | 0.143333|
| Cold plasma                | 0.1035    | 0.07    |

The effect of SiO₂ NPs on biofilm formation

While the silica NPs that synthesized by using magnetic stirrer method had less effect on biofilm formation compared with the control (0.146333, 0.143333, 0.2773 and 0.2) respectively as shown in Figure 6.

Figure 6. The differences in biofilm formation for bacteria after treatment with different SiO₂ NPs

Accordingly, isolates were classified as follows: O.D < 0.12 = no biofilm producer or weak biofilm producer, 0.12 < O.D ≤ 0.24 = moderate biofilm producer and 0.24 < O.D = strong biofilm producer (34). All the samples of SiO₂ NPs showed high effect against S. aureus biofilm formation compared with the control as shown in Table 4, while silica NPs that synthesized by using cold plasma method offered highest inhibition effect for biofilm growth than the rest of the samples because of the less average grain size of silica and then the extract of Thuja orientalis as shown in Figure 6. Based on the differences in the structure of the bacteria cell wall, they are
classified as gram negative or positive. The structural differences lie in the organization of a key component of the membrane, peptidoglycan. These differences in the cell wall confer different properties to the cell, in particular responses to external stresses, including heat, UV radiation and antibiotics (44). The differences in biofilm formation between the gram negative bacteria and positive bacteria may due to the differences in cell wall composition, the structure of the cell wall play an important role in tolerance or susceptibility of bacteria in the presence of nanoparticles and its diffusion inside biofilm matrixes by altering surface from hydrophilic to a highly hydrophobic towards nanoparticles due to change expression of cell wall proteinase. Silica NPs have high surface area, and good biocompatibility, so they found that silica NPs inhibit bacterial adherence to oral biofilms to reduce adhesion, and therefore proliferation, of bacteria. Therefor it is used as a good option to deliver drugs such as antibiotics. Although not strictly having a toxic mechanism (9). Silica NPs possess a net positive charge on their surfaces, which promotes more interaction with the negatively charged surface of bacteria and shows efficient antimicrobial activity (23). The effect of silica nanoparticles and aqueous leaf extract on inhibition of the biofilm formation is may be due to that silica NPs subsequently may bind with DNA molecules and lead to disordering of the helical structure by cross-linking within and between the nucleic acid strands and also disrupt the biochemical processes and protein denaturation and cause cell death (48). Another proposed mechanism is, Colloidal SiO$_2$ NPs are usually negatively charged and hydrophilic (43). May be the SiO$_2$ NPs that may attach to the negatively charged bacterial cell wall and rupture it, thereby leading to protein denaturation and cause cell death. The different synthesis methods of SiO$_2$ NPs affect on the percentage (%) of biofilm inhibition for pathogenic bacteria, as shown in Table 5. SiO$_2$ NPs which synthesized by using cold plasma method acheived the highest biofilm inhibition percentage (62.68, 65%) for S. aureus and E. coli bacteria respectively.

### Table 5. The effect of different NPs and synthesis methods on biofilm inhibition percentage (%) for pathogenic bacteria

| Samples                  | Biofilm inhibition percentage (%) | S. aureus | E. coli |
|--------------------------|----------------------------------|-----------|---------|
| Control                  | 0.2773                           | 0.2       |         |
| Aqueous leaf extract     | 56.73                            | 37.5      |         |
| Magnetic stirrer         | 47.23                            | 28.33     |         |
| Cold plasma              | 62.68                            | 65        |         |

### Antibacterial Activity

All the types of silica NPs did not showed any inhibition zone on the microorganisms by using well diffusion method and this behavior is in agreement with the results of Santra S. (45). Plants are abundant sources for natural and sustainable compounds, which are useful for green synthesis of nanostructures silica NPs. In this regard, we have been synthesis silica NPs via green method using $T$. orientalis aqueous leaf extract and by using TEOS as silica precursor. $T$. orientalis leaf extract contains bioactive compounds which can act as reducing and capping agent for biosynthesis of silica NPs. Prepared particles were surrounded with natural compounds from $T$. orientalis leaf extract. This matrix has hydrophilic functional groups that can make the particles colloidally stable in aqueous environment without applying any harsh reaction condition. This synthesis condition is so interesting from the economical point of view for production of silica NPs in industrial scales.

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