Inhibition of Active Substances of Vibrio Parahaemolyticus and its Application Research

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Abstract. Inhibition of vibrio parahaemolyticus by polyphenol compounds from carageen by plate growth inhibition tested to observe its bacteriostasis effect on V. parahaemolyticus. The test results show that within a certain range, polyphenols from carageen have obvious antibacterial activity against V. parahaemolyticus, the size of antibacterial activity is closely related to polyphenol concentration and molecular mass. The higher the content of polyphenols, the stronger the bacteriostasis it has. It indicates that carageen polyphenol compounds can inhibit V. parahaemolyticus.

1 Introduction

Vibrio parahaemolyticus is a gram-negative bacterium. It's a naturally pathogen bacteria, also known as V. parahaemolyticus. According to the classification of V. parahaemolyticus, it can be divided into pathogenic and non pathogenic. The pathogenic V. parahaemolyticus can cause gastroenteritis or diarrhea through the intestinal tract, or a septicemia caused by a wound infection. At present, there are more than 10 types of vibrio diseases reported by aquaculture animals, among them, vibrio alginolyticus, V. parahaemolyticus and vibrio vulnificus are the most common diseases[1]. Vibrio disease is the major disease of marine cultured animals. It has a wide spread, high morbidity and high mortality characteristic. There have been reports of numerous fish deaths caused by vibrio in Fujian and Zhejiang province[2]. Because of the abuse of antibiotics and the protruding bacterial drug resistance, how to inhibit vibrio parahaemolyticus activity has become an important topic. Seaweed polyphenols have strong antibacterial activity and are toxic to other algae, attached organisms and planktonic larvae[3]. In recent years, relevant studies at home and abroad have also found that polyphenol compounds in sargassum have strong antimicrobial activity, antibacterial activity, chemical defense effect[4] and deodorizing activity, etc[5]. Fabin Dang[6]etc found that polyphenols in carageen have broad antimicrobial spectrum against vibrio marine, but there are not reports about bacteriostatic activity of V. parahaemolyticus that have yet been reported. We use carageen as the experimental material, V. parahaemolyticus as the tested bacteria, the extraction of polyphenols from carageen by plate growth inhibition method, and we study the inhibitory effect of polyphenols from carageen on V. parahaemolyticus. It is used to provide new ideas and methods for the prevention and treatment of common vibrio diseases in aquaculture.

2 Experiment

2.1 Experimental materials

2.1.1 Carageen

Carageen bought from the Tumen Road vegetable market in Yangpu District, Shanghai. After washing, remove the impurities such as sand on the surface of seaweed. Naturally dry, then freeze and preserve. You need thaw it before you use.

2.1.2 Bacteria

V. parahaemolyticus was purchased from Shanghai Lu Wei Technology Co., Ltd.

2.2 Preparation of reagents

Folin-Denis: Adding 100g Na2WO4·2H2O, 20g phosphomolybdic acid hydrate, 50ml H3PO4 to 750ml water, backflowing 2h, cooling it then dilute to 1L.

Preparation of standard liquid for phloroglucinol: 5g phloroglucinol is dissolved in 100ml 95% ethanol.

2.3 Test instrument

Electrothermal constant temperature dryers (DHG-9053A), Electronic balance (FA2004N), Universal high speed crusher (DE-100g), Heat collector magnetic stirrer with constant temperature (DF-101S),
Biochemical incubator, etc.

2.4 Extraction and identification of polyphenols from carageen

2.4.1 Extraction of polyphenols from carageen

First we remove the first surface material, then wash with distilled water to remove second layers of epiphytic organisms, put them in the Electrothermal constant temperature dryers, temperature is set to 100 degree. Wait until the carageen becomes dry matter completely, cryopreserved it. We thaw before using it, dry mass of carageen 10g, use a Universal high speed crusher to crush it to complete, extracting 2h by adding 85% ethanol to avoid light, the amount of carageen and ethanol is about 1: 2 (g/ml), repeated extraction 3 times, mixing the extract, finally we will obtain the crude extract of carageen polyphenols. And we reduce pressure distillation (removal of ethanol), dissolve the finished product in water, then wash 2 times with 1/2 volume of ether and chloroform respectively, then reduce pressure distillation, the finished product dissolves in water, finally we will obtain the essence of the extract[7,8,9], put in the freezer for cryopreservation.

2.4.2 Determination of the content of polyphenols extract from carageen

Brown algae polyphenols are a kind of phenolic compounds with basic structure units of phloroglucinol. We use AOAC standard analysis method, using Folin-Denis[10] reagent to form a blue complex with polyphenols under alkaline conditions, colorimetric analysis at λ=709nm, standard curve drawing with phloroglucinol[11] as standard, and then the relative content of carageen polyphenols was calculated. Because polyphenols extract contained mannitol[12], mannitol and NaCl were used as controls.

X is absorbance and Y is concentration, data analysis by using Matlab, the relationship between the content of polyphenols and the absorbance is that: y=61.8201x+20.2077, the results show that the content of polyphenols in this experiment is 411.0289μg/ml.

2.4.3 Preparation of culture medium

Nutrient agar medium: peptone 10g/L, NaCl 5g/L, agar powder 15g/L, beef powder 3g/L, weighing the medium 33g, adding distilled water 1000ml, heat dissolving and put into the flask, sterilizing and standby by 121 degree 15min, pH7.6~7.8[9].

2.4.4 Preparation of polyphenols from carageen

The polyphenol extract was melted at room temperature before the experiment and we number the EP tube, we separately configure the concentration of 50μg/ml, 100μg/ml, 200μg/ml, 400μg/ml, 800μg/ml, 1200μg/ml, 1600μg/ml, 2000μg/ml.

2.4.5 Preparation of bacterial plate

Take the activated bacteria, make a certain density of bacteria suspension. Take 0.2 ml of the bacterial suspension and pour into the aseptic plate, and then immediately pour into the medium 15ml with high temperature sterilization and cooling to 45 degree, let the bacterial liquid mix well with the medium and ready for use.

2.5 Determination method of bacteriostasis activity

Use aseptic glass pipette to punch 2.5mm hole in bacteria plate, add a sample of 10μl per hole, each sample is added to a hole, and about 15mm from the side of the culture plate, 5% NaCl and 3%mannitol were used as the control group. The remaining 4 holes were put into different concentrations of carageen polyphenols extract, the concentrations were 50μg/ml, 100μg/ml, 200μg/ml, 400μg/ml, and so on. Then the culture plate was put into 36 degree incubator to cultivate 24h, measure the diameter of the bacteriostasis circle (deducted the diameter of the aperture), mean value of bacteriostasis.

3 Results and discussion

3.1 Morphology of VP

Electron microscopy revealed that it is round, curved, rod-shaped, filamentous, and has no spores, as shown in Fig. 2. A size of about 210 × 180 nm, as shown in Fig. 3.

3.2 Bacteriostatic effect and analysis of different concentrations of polyphenols

![Fig. 1. Standard curve of carageen polyphenols](image1)

![Fig. 2. and Fig. 3. Electron microscopy of VP](image2)
Table 1. Polyphenols content of carageen and diameter of bacteriostasis

| Polyphenols content (μg/ml) | 5% NaCl | 3% mannitol | 50  | 100  |
|-----------------------------|---------|-------------|-----|------|
| Diameter of bacteriostasis circle (mm) | 0 | 0 | 0 | 0.5  |
|                            | 0 | 0 | 0 | 0.5  |
|                            | 0 | 0 | 0 | 0.5  |
|                            | 0 | 0 | 0 | 0.7  |
|                            | 0 | 0 | 0 | 0.9  |
| Mean value (mm)            | 0 | 0 | 0 | 0.62 |

200  400  800  1200  1600  2000  
0.6  1.0  1.5  1.7  1.9  2.4  
0.6  1.5  2.0  2.2  2.2  2.5  
0.7  1.0  1.5  1.7  2.0  2.3  
1.0  1.5  1.6  1.7  2.1  2.5  
1.4  1.9  2.1  2.4  2.6  3.0  
0.86 1.38 1.74 1.94 2.16 2.54  

Fig. 4. Inhibitory effect of carageen polyphenols on V.parahaemolyticus
1. 5%NaCl 2. 3%Mannitol 3. 50μg/ml 4. 100μg/ml 5. 200μg/ml 6. 400μg/ml

Fig. 5. Inhibitory effect of carageen polyphenols on V.parahaemolyticus
1. 5%NaCl 2. 3%Mannitol 3. 800μg/ml 4. 1200μg/ml 5. 1600μg/ml 6. 2000μg/ml

The mean single factor analysis of the results of V.parahaemolyticus from the same source of carageen polyphenols, the results obtained (after removal of glass pipette) are converted to data, analysis of variance using SPSS 19. The results show that the antibacterial effect of polyphenol from the same source on V.parahaemolyticus was significantly different. Carageen polyphenols: F_{12,21}=6.059, P=0.000. Because P<0.05, the antibacterial ability is related to polyphenol content, as shown in Table 1, the higher the polyphenol content, the stronger the bacteriostatic ability.

3.3 Estimated marginal mean

The polyphenol content test showed that when the polyphenol content was low, carageen polyphenols had little difference in bacteriostatic ability of V.parahaemolyticus, but with the increase of polyphenol concentration, the difference became more significant. The results of simple effect test, as shown in Fig. 6.

4 Conclusion

On the whole, the results in this study demonstrated that the inhibitory effect of carageen on V.parahaemolyticus was determined by plate method. It was found that the inhibition of carageen polyphenols is closely related to its concentration, when the concentration of polyphenol decreased to a certain concentration, the inhibitory effect disappeared. Bacterial disease is one of the most common and serious diseases in aquaculture. Once the outbreak of infection can occur in a short time, a large number of fish and shrimp died, resulting in huge losses to the aquaculture industry. The results showed that the polyphenol of carageen has potential application value in the prevention and treatment of V.parahaemolyticus.

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