Interleukins in cancer: from biology to therapy

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Abstract | Interleukins and associated cytokines serve as the means of communication for innate and adaptive immune cells as well as non-immune cells and tissues. Thus, interleukins have a crucial role in cancer development, progression, and control. Interleukins can nurture an environment enabling and favouring cancer growth while simultaneously being essential for a productive tumour-directed immune response. These properties of interleukins can be exploited to improve immunotherapies to promote effectiveness as well as to limit side effects. This Review aims to unravel some of these complex interactions.

Advances in tumour biology during the past century have demonstrated the tight interplay between the immune system and healthy and malignant cells. These insights laid the foundation for the concept of immunosurveillance: the ability of the immune system to recognize and eliminate transformed cells. In contrast, immunoediting describes the reciprocal interaction and shaping of the immune system and cancer cells, eventually culminating in cancer development and progression1–5. The resistance to immune attack and the presence of protumoural inflammation are two of the major hallmarks of cancer6. Highlighting its importance, the immune milieu at the cancer site on presentation (immune contexture) can define the outcome of patients with colorectal cancer7,8.

Cytokines mediate key interactions between immune and non-immune cells in the tumour microenvironment (TME). It was recently demonstrated how the TME, for example, lung adenocarcinoma allows malignant cells to co-evolve with immune responses9. Among cytokines, several interleukins are particularly relevant in the development and progression of cancer. The multitude of cellular sources, receptors and signalling pathways and even dose-dependency define the pleiotropic role of interleukins in cancer. Along these lines, interleukin action can be cell specific and spans cancer initiation, tumour progression and tumour control10.

Delineating the exact mechanisms of tumour immune control and evasion enabled the development of novel, tailored and highly effective therapies. The therapeutic potential of interleukins has been of interest in both basic and translational cancer research in recent years. The increasing number of clinical trials currently under way highlights their value as a therapeutic agent and a target. Cytokines have been tested in clinical trials as singular therapeutic agents with limited success rates (extensively reviewed in11) and are now undergoing a revival in combinations with synthetic biology, gene and cellular therapies. In most cases, classification of cytokines relies on structural or receptor homology and gene proximity but not necessarily on their biological role in cancer. It may prove useful to readers in a broad range of disciplines.

Carcinogenesis

Chronic inflammation has long been established as one of the drivers for carcinogenesis in many cancer entities, such as lung, skin, oesophageal, gastric, colorectal and pancreatic cancer and hepatocellular carcinoma12. Some interleukins directly induce signalling in non-immune cells and sustain tissue homeostasis. However, after the oncogenic event, interleukin signalling in cancer cells can become a pathological mechanism of tumour growth, metastatic spread and cancer progression (Fig. 1).

IL-1 has long been implicated in inflammation-induced carcinogenesis13–16. IL-1α and IL-1β, which have more recently gained attention for their role in tumour biology, and their family members (Table 1) are produced by immune (for example, myeloid) and non-immune...
| Interleukin | Receptors | Function in cancer | Potential therapeutic strategy | Refs |
|------------|-----------|--------------------|-------------------------------|------|
| **IL-1 superfamily: IL-1 subfamily** | | | | |
| IL-1α | IL-1R1–IL-1R3 sIL-1R3 | Pleiotropic: promotes inflammatory carcinogenesis and antitumour immunity | Therapeutic neutralization to manage cachexia in clinical trials | 12,228 |
| IL-1β | IL-1R1–IL-1R3 IL-1R2–IL-1R3 sIL-1R2 sIL-1R3 | Pleiotropic: promotes inflammation-induced carcinogenesis, but recruits antineoplastic cells, may block metastatic outgrowth | Therapeutic neutralization to manage CRS in ACT, cancer prevention and treatment (CANTOS) | 12,229 |
| IL-33 | IL-1R3–IL-1R4 sIL-1R4 | Protumour: tumorigenic niche, Treg cell function, TH2 cell polarization | Preclinical neutralization | 15,124 |
| **IL-1 superfamily: IL-18 subfamily** | | | | |
| IL-18 | IL-1R5–IL-1R7 IL-18BP | Mostly antitumoural: activates lymphocytes to produce IFNγ, induces apoptosis | Preclinical engineered rIL-18 or in combination with ACT, hampered by IL-18BP | 97 |
| IL-37 | IL-1R8–IL-1R5 | NK cell function checkpoint inhibitor, but has some antitumour properties | Not explored | 230 |
| **IL-1 superfamily: IL-36 subfamily** | | | | |
| IL-36α, IL-36β and IL-36γ | IL-1R6–IL-1R3 | Antitumoural: promotes T<sub>H</sub>1-type inflammation, inhibited by IL-36Ra | Preclinical rIL-36γ as a tolerable alternative to IL-1 | 231 |
| IL-38 | IL-1R6–IL-1R9 | Has not been explored, evidently immunosuppressive | Not explored | 232,233 |
| **IL-2 (common γ-chain) family** | | | | |
| IL-2 | sIL-2Ra IL-2/IL-15Rβ–γc IL-2Rα–IL-2/IL-15Rβ–γc | Antitumoural: T cell and NK cell growth factor, but terminates T cell responses by the maintenance of T<sub>reg</sub> cells and induction of AICD | rIL-2 approved for monotherapy. Further engineered to avoid side effects and is used in ACT | 8,234 |
| IL-4 | IL-4Ra–γc | Protumoural: promotes T<sub>H</sub>2-type inflammation and T<sub>1</sub>,9 cell polarization. IL-4R, when overexpressed, promotes cancer growth | Targeting IL-4R-bearing cancer cells, blocking signalling. Producing antitumoural T<sub>1</sub>,9 cells for ACT | 235 |
| IL-7 | IL-7Ra–γc sIL-7Ra | Antitumoural: T cell and NK cell growth factor | rIL-7 or analogues in combination with interleukins or ACT | 236 |
| IL-9 | IL-9–γc | Pleiotropic: context dependent | Preclinical T<sub>1</sub>,9 cells in ACT | 114 |
| IL-15 | IL-15±IL15Ra + IL-2/IL-15Rβ–γc | Antitumoural: activates lymphocytes to produce IFNγ | rIL-15 or analogues in combination with interleukins or ACT | 8 |
| IL-21 | IL-21R–γc | Antitumoural: enhances cytotoxicity of CTLs | Combination therapies with rIL-21 in clinical trials | 8 |
| **IL-3 family** | | | | |
| IL-3 | IL-3Ra–βc | Haematopoietic factor, promotes haematological malignancies | Fused to toxins to target CD123-bearing cells | 103,106 |
| IL-5 | IL-5Ra–βc | Pleiotropic: via eosinophils and T<sub>1</sub>,2 cells | Preclinical neutralization | 101 |
| **IL-6 family** | | | | |
| IL-6 | IL-6Ra–gp130 (classic) sIL-6Ra–gp130 (trans) | Protumoural: activates carcinogenesis and tumour outgrowth, mediates CRS, promotes cachexia | Neutralization to manage CRS in ACT, cachexia | 36,37,44 |
| IL-11 | IL-11Ra–gp130 (classic) sIL-11Ra–gp130 (trans) | Protumoural: promotes inflammation-induced carcinogenesis and cancer progression | Preclinical neutralization and gp130 common receptor blockade | 38,39 |
| IL-31 | IL-31Ra–OSMRβ | T<sub>1</sub>,2-type cytokine, evidently tumorigenic | Not explored | 237 |
| **IL-10 family** | | | | |
| IL-10 | IL-10Ra–IL-10Rβ | Pleiotropic: promotes cytotoxicity, but inhibits antitumour responses | rIL-10 to increase cytotoxicity in trials | 102,105 |
| IL-19 | IL-20Ra–IL-20Rβ | Understudied, evidently protumoural | Not explored | 239 |
| IL-20 | IL-20Ra–IL-20Rβ IL-22Ra–IL-20Rβ | Protumoural: directly promotes carcinoma outgrowth, induces PD1 | Preclinical neutralization | 312,40 |
IL-10 family (cont.)

| Interleukin | Receptors | Function in cancer | Potential therapeutic strategy | Refs |
|------------|-----------|--------------------|-------------------------------|------|
| IL-22      | IL-22Ra1–IL-10Rβ IL-22Ra2 (also known as IL-22BP) | Promutual; promotes progression of carcinomas | Preclinical neutralization | 27,285 |
| IL-24      | IL-20Ra–IL-20Rβ IL-22Ra1–IL-20Rβ | Antimutual; induces apoptosis and autophagy of cancer cells | Preclinical rIL-24 combined with oncolytic virus | 241,242 |
| IL-26      | IL-20Ra–IL-10Rβ | Promutual via T<sub>1</sub>7 cells and neutrophils | Preclinical neutralization | 243,244 |

IL-12 family

| Interleukin | Receptors | Function in cancer | Potential therapeutic strategy | Refs |
|------------|-----------|--------------------|-------------------------------|------|
| IL-12      | IL-12Rβ1–IL-12Rβ2 | Antimutual; the main driver of T<sub>1</sub>7-type immunity, initiates and amplifies IFN<sub>γ</sub> production | rIL-12 has severe side effects, and thus is engineered or combined with other interleukins and ACT in trials | 4,245 |
| IL-23      | IL-23R–IL-12Rβ1 | Mainly promutual; direct and indirect effect via T<sub>1</sub>7 cells and T<sub>1</sub>722 cells | Neutralization in trials, enhances CAR T cell cytotoxicity | 212,227,245 |
| IL-27 and IL-30 (also known as IL-27 subunit p28) | IL-27Ra (also known as WSX1)–gp130 | Pleiotropic; induces NK cell and CTL cytoxicity, but enhances T<sub>reg</sub> cell activity and T cell exhaustion | Neutralization and engineered rIL-27 in trials | 120,121,246,247 |
| IL-35      | IL-12Rβ2–gp130 IL-12Rβ2–IL-12Rβ2 gp130–gp130 IL-27Ra–IL-12Rβ2 | Promutual; T<sub>reg</sub> cell-mediated suppression of T cell responses and exhaustion of T cells. Promotes metastatic colonization | Preclinical neutralization in combination with checkpoint inhibitors and other therapies | 83,120 |

IL-17 family

| Interleukin | Receptors | Function in cancer | Potential therapeutic strategy | Refs |
|------------|-----------|--------------------|-------------------------------|------|
| IL-17A/F   | IL-17RA–IL-17RC | Promutual; carcinogenesis, immunosuppression, EMT | Neutralization in clinical trials | 60,129 |
| IL-17B     | IL-17RB | Mostly promutual, but antimutual properties are also reported | Not explored, scarce preclinical evidence | 248 |
| IL-17C     | IL-17RA–IL-17RE | | Not explored, scarce preclinical evidence | 249 |
| IL-17D     | Unknown | | Not explored, scarce preclinical evidence | 250 |
| IL-25 (also known as IL-17E) | IL-17RA–IL-17RB | | | 63 |

IFNα family (type III interferons)

| Interleukin | Receptors | Function in cancer | Potential therapeutic strategy | Refs |
|------------|-----------|--------------------|-------------------------------|------|
| IL-28A and IL-28B | IL-28Ra (also known as IFNL1)–IL-10Rβ | Antimutual; induces apoptosis of malignant cells | Preclinical gene therapy using IL-28 and IL-29 | 102,103,251 |
| IL-29      | IL-28Ra–IL-10Rβ | | | |

Other interleukins

| Interleukin | Receptors | Function in cancer | Potential therapeutic strategy | Refs |
|------------|-----------|--------------------|-------------------------------|------|
| IL-8 (also known as CXCL8) | CXCR1, CXCR2 ACKR1/DARC | Promutual; attracts neutrophils, mediates suppressive environment | Therapeutic neutralization in clinical trials | 132 |
| IL-13      | IL-13Ra1–IL-4Ra IL-13Ra2 | Promutual; T<sub>2</sub>-type cytokine | Targeting or blocking IL-13R | 235 |
| IL-14a and IL-14β | IL-14R | B cell growth factor, also in lymphoma; not explored | Not explored | 252 |
| IL-16      | CD4 | Evidently promutual; lymphoma proliferation, chemoattractant | Not explored, scarce preclinical evidence | 253 |
| IL-32 (also known as NK4) | Unknown | Pleiotropic; depending on isoform (a to s) and cancer type | Preclinical antitumour effects in combination | 254 |
| IL-34      | CSF1R | Promutual; cancer progression, immune suppression and therapeutic resistance | Preclinical neutralization to alleviate promutual effects | 255,256 |

ACT, adoptive cell transfer; AICD, activation-induced cell death; CANTOS, Canakinumab Anti-inflammatory Thrombosis Outcomes Study; CAR, chimeric antigen receptor; CRS, cytokine release syndrome; CTL, cytotoxic T lymphocyte; CXCL8, CXC-chemokine ligand 8; CXCR, CXC-chemokine receptor; EMT, epithelial–mesenchymal transition; IFNγ, interferon-γ; IL-18BP, IL-18-binding protein; IL-1R, IL-1 receptor; IL-22BP, IL-22-binding protein; NK, natural killer; PD1, programmed cell death protein 1; rIL, recombinant interleukin; sIL, soluble interleukin; T<sub>reg</sub> cell, regulatory T cell.

(for example, epithelial) cells in the context of chronic inflammation. IL-1α and IL-1β are alarm cytokines (also known as alarmins) that both act through IL-1 receptor (IL-1R) to initiate and amplify local inflammation. The production of the IL-1β precursor pro-IL-1β occurs rapidly in response to danger-associated molecular patterns (DAMPs) and pathogen-associated molecular patterns (PAMPs) by pathogen-recognition receptors, such as Toll-like receptors, C-type lectin receptors or retinoic acid-inducible
Interleukins in carcinogenesis. Persistent inflammation in response to pathogen-associated molecular patterns (PAMPs) and danger-associated molecular patterns triggers activation of nuclear factor-κB (NF-κB), which primes pro-IL-1β production, and nucleotide-binding oligomerization domain (NOD)-like receptor (NLR) family pyrin domain-containing 3 (NLPR3) inflammasome activation, which causes the release of active IL-1β from fibroblasts, epithelial cells and myeloid cells such as dendritic cells (DCs), monocytes and macrophages (MΦ). In tumours, IL-33 derived from tumour-initiating cells recruits macrophages, which upon activator protein 1 (AP-1) signalling produce transforming growth factor-β (TGFβ) that suppresses the function of cytotoxic T lymphocytes (CTLs). IL-1β induces production of nitric oxide (NO) and reactive oxygen species (ROS) by epithelial cells, which may cause DNA damage, and promotes the production of IL-6 and IL-11 from epithelial and myeloid cells, and IL-22 from type 3 innate lymphoid cells (ILC3s) and γδ T cells. Under homeostatic conditions, IL-22 facilitates DNA repair caused by bacterial genotoxins, but in transformed cells, IL-6 and IL-11 together with IL-22 rapidly induce phosphorylation of signal transducer and activator of transcription 3 (STAT3). Activation of STAT3 signalling is observed in multiple types of cancer and induces proliferation, survival, stemness, epithelial–mesenchymal transition (EMT) and migration of transformed cells. IL-1β together with TGFβ induces differentiation of T helper 17 (T_{17}) cells, which upon IL-23 stimulation from DCs secrete IL-17A and IL-17F (IL-17A/F). IL-17, which typically mediates wound-healing signalling, and may exacerbate nascent tumour outgrowth.

Recent work demonstrated that IL-33 can create a self-amplifying tumorigenic niche that furthers the development of a nascent tumour. As shown in a model of squamous cell carcinoma, once cells have transformed, they acquire tumorigenic capacity (also known as tumour-initiating cells). Tumour-initiating cells can secrete IL-33, which in turn causes infiltration of tumour-associated macrophages and promotes transforming growth factor-β (TGFβ) signalling, which creates a tumorigenic niche.

In the context of chronic inflammation, IL-1α and IL-1β may directly promote the production of carcinogenic mediators, such as nitric oxide and reactive oxygen species. IL-1 initiates the downstream release of proinflammatory cytokines, such as IL-6, and mediates the recruitment of innate immune cells, triggering a cascade of inflammatory mechanisms. Here, the effects of IL-1 are mediated by phosphorylation of nuclear factor-κB (NF-κB), which in turn inactivates suppressor of cytokine signalling 3 (SOCS3) and potentiates phosphorylation of signal transducer and activator of transcription 3 (STAT3). Activation of STAT3 signalling is observed in multiple types of cancer and induces proliferation, survival, stemness, epithelial–mesenchymal transition (EMT) and migration of transformed cells. IL-1β together with TGFβ induces differentiation of T helper 17 (T_{17}) cells, which upon IL-23 stimulation from DCs secrete IL-17A and IL-17F (IL-17A/F). IL-17, which typically activates NF-κB to mediate wound-healing signalling, and may exacerbate nascent tumour outgrowth.
Similarly, in a mouse model of lung adenocarcinoma, myeloid differentiation primary response gene 88 protein (MyD88)-dependent IL-1 production from myeloid cells acted on γδ T cells to produce an inflammatory loop involving IL-17A and IL-22 secretion that led to the malignant transformation of cells bearing Kras and Trp53 mutations25.

IL-22, which is induced by IL-1 signalling in cancer settings25, has been associated with STAT3 activation and cancer-promoting properties26. IL-22 acts through IL-22R, exclusively expressed on non-haematopoietic cells, to promote wound healing and the production of microbicidal peptides27,28. Recent findings, however, highlight its stage-specific dual properties in carcinogenesis25. Under homeostatic conditions IL-22 is secreted mainly by group 3 innate lymphoid cells (ILC3s) and γδ T cells and can repair genotoxicity-induced DNA damage in the intestinal epithelium to prevent the malignant transformation of cells25. However, when its activity is not controlled by IL-22-binding protein (IL-22BP), its natural inhibitor, IL-22 has a protumorigenic effect in a mouse model of colitis-associated colon cancer29. IL-22-producing T cells were reported to accumulate in nascent lung and colon tumours in mouse and human samples30,31. Through STAT3 phosphorylation IL-22 provides proliferation and migration signals to transformed malignant cells and/or cells bearing oncogenic mutations, sustaining their stemness through the induction of SRY (sex-determining region Y)-box 2 (SOX-2) and NANOG as demonstrated in human colon tissues, and similarly in mouse models of lung, pancreatic and breast cancers32,33. IL-20 has been shown to act similarly to IL-22 in hepatocellular carcinoma and breast, prostate and oral cancer, and also induces immune inhibition through programmed cell death protein 1 (PDI) upregulation in pancreatic cancer34.

Tissue damage and release of alarm cytokines induce IL-6, IL-10, IL-11 and IL-23 expression by sentinel myeloid and tissue cells. Under homeostatic conditions, this results in a self-inhibitory loop to resolve inflammation and promote healing35. In turn, IL-6 and IL-11 are potent orchestrators of innate immune responses and inflammation35,36. Moreover, IL-6 is also a regulator of development and metabolic processes37. These effects are attributed to the ubiquitously expressed common transducing gp130 receptor subunit, which can dimerize with membrane IL-6Ra or IL-11Ra to initiate classical cis signalling or a soluble receptor form resulting in non-classical trans signalling38,39. gp130 is associated with activation of Janus kinase 1 (JAK1), JAK2 and non-receptor tyrosine-protein kinase 2 (TYK2)38,40, as well as tyrosine-protein phosphatase SH-PTP2 (SHP2) and SRC–Yes-associated protein (YAP)–Notch signalling, which activates proliferation and tissue regeneration36,39,41. Furthermore, activation of phosphoinositide 3-kinase (PI3K)–AKT–mechanistic target of rapamycin complex 1 (mTORC1) signalling integrates interleukin signalling and the metabolic cell programme35,36.

Classical IL-6 signalling is considered essential for homeostatic processes, whereas trans signalling was specifically demonstrated to amplify the inflammation and promote inflammation-induced carcinogenesis41,42. Excessive activation of STAT3 by the overabundance of IL-6 and IL-11 combined with oncogenic driver mutations licenses the development of malignant tumours such as colon and gastric40,42,43, pancreatic42,44 and lung45 cancers. IL-1 and IL-6 released by myeloid cells induce PI3K–AKT–mTORC signalling, which, via activation of hypoxia-inducible factor 1a (HIF1a), shifts their metabolism towards glycolysis and decreases oxidative phosphorylation leading to amplification of IL-1 and IL-6 production and exacerbated inflammation-induced carcinogenesis46,47. Of note, deregulated IL-6 and IL-1 signalling is also a contributor to cancer-induced cachexia (see BOX 1).

Importantly, IL-6 also induces angiogenesis and tumour vascularization mediated by vascular endothelial growth factor (VEGF)48. Further, classical IL-6 signalling via IL-6R on lymphocytes promotes proliferation of T cells and lineage commitment to T<sub>reg</sub> cells and T follicular helper cells49. IL-6 suppresses forkhead box P3 (FOXP3), limiting the ability of TGFβ to promote regulatory T cell (T<sub>reg</sub>) development, which enables T<sub>reg</sub> cell differentiation and amplifies proinflammatory response, as demonstrated in a mouse melanoma model of T<sub>reg</sub> cell adaptive cell transfer (ACT)50.

IL-23 is another interleukin produced in response to DAMPs at epithelial barriers51. Typically, it counteracts the antitumour action of IL-12, but it was also demonstrated to directly promote tumour incidence and growth. IL-23 triggers IL-17 production from ILC3s and committed T<sub>reg</sub> cells, synergizing with IL-6 in the amplification of inflammation, and prompting epithelial cells to acquire stemness and undergo malignant transformation52,53. IL-23 together with IL-1, IL-6 and IL-21 is capable of inducing IL-17 production independently of T cell receptor (TCR) signalling54,55.
During chronic inflammation, an abundance of microbial antigens may drive IL-17 and subsequent aberrant wound healing that results in tumorigenesis, as seen in murine models of skin and colon cancers. In skin stem cells, IL-17A signalling can recruit and transactivate epidermal growth factor receptor (EGFR), which induces expansion and migration of these cells. Hence, the inflammatory responses initiate cellular programmes in conditions of uncontrolled chronic activation and may provide a direct link to tumorigenesis.

**Cancer growth and progression**

Malignant tumours possess several characteristic traits termed by Hanahan and Weinberg 'hallmarks of cancer'. Importantly, several of these traits, namely sustained proliferation, inflammation, angiogenesis, active invasion and migration, are also the hallmarks of wound healing, and therefore may maliciously utilize cytokine signalling aimed at tissue repair (Fig. 2).

Thus, it was demonstrated that IL-1 not only promotes inflammation-induced carcinogenesis but also contributes to tumour invasiveness and angiogenesis. Although activation of, for example, the NLR family pyrin-domain containing 3 (NLRP3) inflammasome is reported ubiquitously in cancer, the consequences of NLRP3 activation and release of IL-1β and its family member IL-18 range from antitumour activity to cancer growth and metastasis. IL-18 was demonstrated to promote angiogenesis through the activation of NF-kB, which triggers the secretion of VEGF from cancer cells, induces their proliferation and invasion and...
Natural killer (NK) cells

Cytotoxic lymphocytes that can eliminate virus-infected cells or tumour cells by the recognition of non-self antigens presented on major histocompatibility complex class I molecules of the target cell.

Natural killer T cells (NKT cells).

A subset of T cells exhibiting antigen-specific immunity via a restricted T cell receptor recognizing CD1-presented lipid antigens, but also sharing natural killer cell functions in terms of their independence from major histocompatibility complex.

Antigen-presenting cells (APCs).

Dendritic cells, macrophages and B cells that present non-self peptide antigens to T cells via major histocompatibility complex class II molecules.

prevents apoptosis, and may also induce PD1-dependent immunosuppression of natural killer (NK) cells\(^{1,2}\).

In addition to its ability to amplify procarcinogenic chronic inflammation, IL-6 drives tumour-intrinsic mechanisms of cancer progression that recapitulate most of the hallmarks of cancer\(^1\). IL-6 induces PI3K–AKT, mitogen-activated protein kinase (MAPK)/extracellular signal-regulated kinase (ERK), NF-kB and STAT3 signalling. These pathways augment the expression of anti-apoptotic proteins (B cell lymphoma 2 (BCL-2), B-cell lymphoma, extra large (BCL-xl)), induced myeloid leukemia cell differentiation protein MCL1 and baculoviral inhibitor of apoptosis domain (IAP) repeat-containing protein 5 (BIRC5; also known as survivin)), evade growth control, activate metabolism and induce angiogenesis via VEGF production (reviewed in \(^4\)). STAT3 phosphorylation in cancer cells is also induced by other cytokines, such as IL-22, exacerbating cancer-promoting pathways\(^{22,27,29,57}\).

Furthermore, chronic STAT3 activation in cancer cells induces proliferation, expression of matrix metalloproteinases, and migration, thereby increasing the aggressiveness of carcinomas\(^{34,56,77}\). These effects are typically associated with epithelial-to-mesenchymal transition (EMT), which is orchestrated by EMT transcription factors, such as zinc-finger E-box-binding homeobox (ZEB), SNAI1, SNAI2, SLUG and TWIST, and enables cell detachment, metastatic spread and invasiveness\(^{18,49}\). EMT induced by the integrated network of IL-1β, IL-6, IL-17, IL-22 and IL-23 mediates the invasiveness of many cancer types. For example, STAT3 activation by IL-22 was found to upregulate EMT markers in mouse models of pancreatic ductal adenocarcinoma and breast carcinoma\(^{30,32}\). Moreover, chronic exposure to IL-1β induces MAPK (ERK and JUN amino-terminal kinase (JNK))–activating protein 1 (AP-1) engagement, which results in epigenetic modifications that cause long-lasting EMT phenotypes, including SLUG and ZEB2 transcription in lung cancer\(^{1,26}\). Similarly, EMT is promoted by IL-6-mediated and IL-11-mediated paracrine activation of STAT3 in invasive pancreatic ductal adenocarcinoma, IL-13-mediated activation of STAT6 in colorectal cancer and IL-35 in breast cancer\(^{49,53,54}\). IL-23 utilizes the Wnt–β-catenin pathway to induce EMT in oesophageal cancer\(^{36}\), whereas NF-kB activation induced by IL-17 directly promotes ZEB1 in lung cancer\(^{37}\). IL-8 induces EMT through PI3K–AKT and rat sarcoma (RAS)–rapidly accelerated fibrosarcoma (RAF)–mitogen-activated protein kinase kinase (MEK)–ERK in various cancers\(^{48}\). However, permanent activation of ZEB1 by IL-1β in breast cancer cells entraps them in a mesenchymal state and prevents metastatic colonization; hence, cancer cells require mesenchymal–epithelial transition for further progression\(^{38}\). Taken together, these findings provide a rationale for the therapeutic neutralization of these cytokines to reduce EMT-mediated cancer progression.

Cancer immunosurveillance

Innate immunity

Despite the plethora of oncogenic factors that constantly induce malignant transformation of cells, our immune system recognizes and eliminates most of these transformed cells through immunosurveillance\(^1\). Innate immune cells, namely NK cells, natural killer T cells (NKT cells) and γδ T cells, possess a wide array of the evolutionarily conserved ‘altered self’ and activation receptors that can recognize and eliminate transformed cells\(^1\). This process depends on production of perforin, granzyme B and interferon-γ (IFNγ) in these cells\(^{39}\). The last of these directly induces apoptosis in tumour cells and triggers a cascade of chemokines to initiate the recruitment of innate immune cells and induce the production of proinflammatory antitumoural cytokines\(^{40}\) (FIG. 5).

Proliferation, maturation and cytotoxic effector functions of innate immune cells are tightly controlled by IL-2 family cytokines, namely IL-2, IL-7, IL-15 and IL-21, which utilize heteromeric receptors and share a common γc receptor subunit\(^{41}\). IL-15 is the primary cytokine to regulate the biology of NK cells, which have particular importance for the control of haematological malignancies\(^{42}\). Unlike other IL-2 family members, IL-15 is typically presented in conjunction with the IL-15Ra subunit on the surface of antigen-presenting cells (APCs)\(^{43}\). To elicit signalling, the IL-15–IL-15Ra complex binds a heterodimeric IL-2Rβ–γc receptor complex shared with IL-2 (REFS\(^{44,45}\)). Upon binding to its receptor on NK cells, IL-15 triggers a cascade of reactions resulting in phosphorylation of the serine-threonine kinase AKT. Consequently, it causes accumulation and translocation of the transcription factor X-box-binding protein 1 (XBP1) into the nucleus, where T-box protein expressed (T-box) homology and FAS ligand (FASL) expression12. In addition to NK cells and cytotoxic T lymphocytes (CTLs)\(^{46,47}\), IL-15 secreted from APCs, in turn, induces transcription of granzyme B and genes responsible for proliferation, maturation and survival of NK cells\(^{48}\). Importantly, the activity of AKT downstream of IL-15 induces homeostatic priming in NK cells and cytotoxic T lymphocytes (CTLs)\(^{49}\).

NK cell-secreted IFNγ primes APCs, such as dendritic cells (DCs) and macrophages, to produce IL-12, which shares one of its subunits, p40, with IL-23 (REFS\(^{50,51}\)). Importantly, APCs serve as a bridge between the innate and adaptive immune responses, and their recruitment, maturation and activation are governed by cytokines\(^{52}\). IL-12 secreted from APCs, in turn, amplifies IFNγ production in NK cells (requires priming by IL-15 and/or IL-18 (REFS\(^{52,53}\))) and thereby antitumour responses\(^{54}\).

IL-18 acts through its receptor highly expressed in NK cells and triggers IFNγ production, cytotoxicity and FAS ligand (FASL) expression\(^{55}\). In addition to the reported ability to directly promote the proliferation and migration of cells in certain types of cancer, tumour-infiltrating NK cells and CD8 T cells express high levels of IL-18R\(^{56,57}\). However, IL-18 activity is moderated by IL-18-binding protein (IL-18BP), a soluble receptor that acts as a decoy for IL-18 and inhibits antitumoural and antiviral activities of NK cells and CTLs\(^{57,58}\). Moreover, mouse and human NK cells have high expression of IL-1R8, another member of the IL-1R family, which together with IL-1R5 (also known as IL-1R8a) binds IL-37, which is known for its anti-inflammatory properties\(^{1,29}\). IL-1R8 could inhibit IL-18-dependent mTOR and JNK signalling pathways, which are essential for NK cell metabolism, differentiation and activation, as
demonstrated in mouse models of liver carcinogenesis and haematogenous liver and lung metastasis\textsuperscript{99}.

Recently, eosinophils, a pleiotropic innate immune cell type primarily known for antihelminth and allergic responses, have received attention due to their distinct antitumour mechanisms in hepatocellular carcinoma and breast cancer\textsuperscript{100,101}. IL-5, which together with IL-3 and granulocyte–macrophage colony-stimulating factor (GM-CSF) constitutes a family of haematopoietic cytokines, is a key regulator of eosinophil differentiation, maturation, expansion and survival\textsuperscript{101}. Recruited by the tumour-released alarmin IL-33, eosinophils can directly exert cytotoxicity but also aid antitumoral TH1 cell responses\textsuperscript{100}.

IL-28A, IL-28B and IL-29 are cytokines distantly related to the IL-10 family, but are also interferons and therefore are also termed ‘type III interferons’ or ‘λ-interferons’. They typically mediate innate immune antiviral activity but also directly induce apoptosis of malignant cells\textsuperscript{102,103}.

Adaptive immunity

Upon immunogenic cancer cell death, antigens released by tumour cells are taken up by APCs, primarily DCs, and enter draining lymph nodes to initiate the development of antigen-specific adaptive immune responses. There, the cytokine milieu plays a fate-defining role for T cells\textsuperscript{2,104} (Fig. 3). Proliferation, survival, differentiation and termination of T cell responses are mainly governed by IL-2, IL-7 and IL-15 (REF.\textsuperscript{8}), whereas IL-3 is necessary for survival and proliferation of lymphocyte progenitors\textsuperscript{105,106}. IL-2 binds with intermediate affinity to γc–IL-2Rβ, inducing the clonal expansion of T cells, but also for termination of the T cell responses through...
Immunotherapy

Treatment of patients with factors shaping an immune response to enable the patient’s immune system to react to the disease.

T<sub>1</sub>-type responses

Polarized inflammatory responses inducing killing of intracellular pathogens and tumour cells mediated primarily by interferon-γ.

M2 macrophages

An inhibitory macrophage subtype associated with tissue remodelling and repair that secrete anti-inflammatory cytokines such as IL-10 and transforming growth factor-β (TGFβ).

Myeloid-derived suppressor cells (MDSCs)

A heterogeneous group of myeloid cells often recruited by tumour cells that lead to pronounced immunosuppression by inhibiting a wide range of inflammatory immune cells.

Group 2 innate lymphoid cells (ILC2)

Cells with characteristics of lymphoid cells but without rearranged antigen-specific surface receptors. Similarly to CD4<sup>+</sup> T cells, they produce type 2 cytokines and are also referred to as natural or innate helper cells.

T<sub>2</sub>-type responses

Immune responses against mainly extracellular antigens mediated by IL-4, IL-5 and IL-15.

FAS/FASL-mediated apoptosis of T cells<sup>2</sup>. IL-2Ra chain (CD25) expressed mainly in T<sub>reg</sub> cells increases the affinity of the IL-2R, mediates FOXP3 expression and is essential for self-tolerance<sup>3</sup>. CTLs and effector T<sub>H</sub>1 cells are the primary mediators of antitumoural adaptive immunity<sup>4</sup>. IL-12 derived from DCs provides an essential signal that drives expression of T-bet, and thus the differentiation of effector T<sub>H</sub>1 cells, and CTLs<sup>5,6,7</sup>. Successful anti-PDI immunotherapy relies on IL-12-dependent crosstalk between T cells and DCs to promote tumour cell killing<sup>8</sup>. Similarly to signalling in NK cells, IL-2, IL-15 and IL-18 synergize with IL-12 to trigger IFNγ production and direct cytotoxicity in CTLs and T<sub>H</sub>1 cells<sup>9,10</sup>. Thus, soluble IL-15–IL-15Ra complexes produced by myeloid cells were shown to be essential for establishing NK cell and CTL populations in tumours in mouse models of melanoma and colon cancer<sup>11</sup>. In addition, priming of T cells by IL-18 activates NF-κB, augments STAT4 activation by IL-12 and enables TCR-independent production of IFNγ<sup>12</sup>. Despite its ability to suppress IL-12 production and resolve inflammation, IL-10 also potently induces CTL cytotoxicity towards tumours with potential for clinical application<sup>13</sup>.

The plasticity of the interleukin-dependent T cell transcriptional profile is essential for tailoring immune responses and necessary effector mechanisms<sup>14</sup>. TH<sub>9</sub> cells and their prototypical cytokine IL-9, despite being closely related to the T<sub>H</sub>2 cell lineage, which typically antagonizes T<sub>1</sub>-type responses, can serve as persistent antitumour effectors that resist exhaustion and can be exploited in cellular therapy<sup>15,16</sup>. TH<sub>9</sub> cells are typically induced by IL-4 and TGFβ, and experiments have shown that IL-1β can replace TGFβ to promote the TH<sub>9</sub> cell lineage. TH<sub>9</sub> cells induced that way mounted a persistent response and possessed potent antitumoural properties, secreting IL-9, IL-21 and granulocyte B<sub>2</sub>-<sub>1</sub>-<sub>1</sub>-<sub>7</sub>, IL-21, besides its typical role in B cell responses, enhances cytokotoxicity and increases persistence of NK cells and CTLs in tumours as demonstrated in a melanoma mouse model of ACT<sup>18</sup>. Secretion of IL-21 can also be induced in T cells by IL-27, and limits differentiation of T<sub>reg</sub> cells, enhances proliferation of NK cells and CD8<sup>+</sup> T cells but also triggers the exhaustion programme in these cells as also recently investigated in a mouse model of melanoma<sup>19,20</sup>. TH<sub>2</sub> cells, typically seen as antagonists to antitumoural T<sub>H</sub>1 cell responses, can cause remodelling of tumour vasculature, thus leading to tumour starvation and regression<sup>12</sup>.

Cancer immune evasion

Despite the potent action of cancer immunosurveillance and immunoediting, malignant cells may evolve to evade antitumoural responses and exploit extrinsic immunoinhibitory mechanisms for progression<sup>21,22</sup>. In this way, they may secrete substances that enable domination of cell types that suppress antitumour immunity, such as T<sub>reg</sub> cells, ILC3s, T<sub>H</sub>17 cells, T<sub>eff</sub> cells, M2 macrophages and myeloid-derived suppressor cells (MDSCs), and induce anergy of antitumour cells through their metabolic reprogramming, tipping the balance towards cancer progression.

**T<sub>reg</sub> cells**

High-affinity IL-2 signalling together with TGFβ induces Foxp3. Foxp3 drives T<sub>reg</sub> cell differentiation and IL-10 production, thereby establishing an immunosuppressive TME<sup>23,24</sup>. Besides its role in creating a macrophage-dependent tumorigenic niche, IL-33 may directly promote TGFβ-elicited T<sub>reg</sub> cell differentiation, suppress IFNγ and promote T<sub>reg</sub> cell stability in the tumour<sup>24,25</sup>. The regulatory role of T<sub>reg</sub> cells in the TME is largely mediated by the secretion of IL-10, IL-35 and TGFβ<sup>26</sup>. IL-10 is anti-inflammatory and relies on its ability to suppress B lymphocyte-induced maturation protein 1 (BLIMP1)-dependent cytokine secretion and regulate cytotoxic effector function, whereas IL-35 induces inhibitory surface receptors (including PD1, lymphocyte-activation gene 3 protein (LAG3), T cell immunoglobulin and mucin domain-containing protein 3 (TIM3), T cell immunoreceptor with immunoglobulin and mucin domain-based inhibitory motif (ITIM) domains (TIGIT) and 2B4 (also known as CD244)) and limits the formation of T cell memory as demonstrated in mouse models of melanoma<sup>27</sup>. Mechanistically, IL-10 may also directly reduce TCR sensitivity upon increased cell membrane glycosylation<sup>28</sup>. T<sub>reg</sub> cells further limit the availability of IL-2 to CD4<sup>+</sup> T cells in the TME, causing deficiency of granzyme B expression in these cells and uncontrollable tumour outgrowth as demonstrated in a mouse sarcoma model<sup>29</sup>.

**T<sub>H</sub>17-type responses and myeloid suppressors**

In response to IL-1β stimulation, T<sub>H</sub>17 cells and γδ T cells produce IL-17, which recruits a large number of immunosuppressive granulocytes (reviewed in<sup>30,31</sup>), which was demonstrated to promote lung metastasis in a murine breast cancer model<sup>30</sup>. Moreover, in lung cancer, IL-23 converts group 1 innate lymphoid cells into IL-17-producing ILC3s that promote IL-17 mediated tumour cell proliferation<sup>31</sup>. Additionally, tumour-derived IL-8 (also known as CXCL8) provides a chemotactic signal for myeloid cells in patients with breast cancer and confers resistance to immunotherapy<sup>32,33</sup>. Similarly, IL-18 produced upon inflammation activates generation of MDSCs in multiple myeloma, conferring a mechanism of immunosuppression<sup>34</sup>.

**T<sub>eff</sub>2-type responses**

Tumour and stromal cells may secrete factors to promote T<sub>eff</sub>2 cell and M2 macrophage polarization, which suppress antitumoural T<sub>H</sub>1 cell polarization and responses<sup>35,36</sup>. In the same way, IL-33-induced group 2 innate lymphoid cells (ILC2) were demonstrated to antagonize NK cell function in a mouse model of melanoma<sup>37</sup>. In turn, IL-33, secreted by cancer-associated fibroblasts in breast cancer, is a potent enhancer of ILC2 and T<sub>eff</sub>2-type responses, and may induce TCR-independent secretion of IL-13 and recruitment of immunosuppressive granulocytes<sup>38,39</sup>. Moreover, IL-33 activates intrinsic signalling in T<sub>reg</sub> cells necessary for their immunosuppressive action in cancer<sup>34</sup>. 
Immune cell metabolic reprogramming

Cancer cells can induce metabolic reprogramming of immune cells in the TME and systemic changes in metabolism, which can induce the transition from proinflammatory to immunosuppressive responses. A glycolytic switch in effector T cells dependent on NF-κB-inducing kinase (NIK) signalling was recently demonstrated as pivotal for antitumour immunity in a mouse melanoma model. The accumulation of glycolysis-derived lactate in the TME can increase oxidative phosphorylation and together with IL-4 induces anti-inflammatory reprogramming (involving M2 polarization of macrophages and secretion of IL-10 and TGFβ). In addition, accumulation of lactate and depletion of amino acids and glucose in the TME in combination with TGFβ signalling can induce Treg cell polarization and amplify immunosuppression and T cell exhaustion, thereby rendering tumours resistant to immunotherapy (reviewed in [19]). In pancreatic ductal adenocarcinoma, it was shown that stromal cells released IL-6, which increased glycolysis and lactate efflux from the tumour cells, leading to M2 polarization of macrophages and reduction of the efficacy of anti-PD1 treatment [20].

Hence, the accumulation of tumour-derived substances together with chronic inflammatory responses renders antitumour immunity unable to restrain tumour growth and progression, resulting in an uncontrolled tumour outgrowth and distant metastasis and limiting the efficacy of targeted therapies.

Interleukins in cancer therapy

As some interleukins regulate immunosurveillance and thereby tumour control, clinicians have tried to utilize them for cancer therapy in the past few decades. However, cancer treatment by interleukins proved to be challenging as it has not lived up to its expectations in terms of effectiveness, while also encountering serious toxic effects in some cases. Because interleukins also nurture cancer progression, neutralization of their activity has emerged as a potential avenue.

There are reports suggesting several interleukins as biomarkers [21–24], but so far no interleukin has been implemented as a biomarker in the US Food and Drug Administration guidelines for oncologic therapeutics, and their potential overall value has been extensively reviewed in [25].

Here, we summarize recent advances in the translation of therapeutic exploitation of interleukins, covering therapeutic neutralization of interleukins as well as the use of their recombinant and engineered forms (Fig. 4).

Therapeutic neutralization

Interleukin-1. The Canakinumab Anti-inflammatory Thrombosis Outcomes Study (CANTOS) provided the strongest evidence for the potential of IL-1β-neutralization in cancer so far. The study enrolled patients with stable coronary artery disease receiving the standard of care, but with persistent elevation of the level of high-sensitivity C-reactive protein (hsCRP). Patients were treated with the anti-IL-1β antibody canakinumab and did not have a diagnosis of cancer at the time of enrolment. Canakinumab not only reduced hsCRP levels but also significantly lowered overall cancer mortality compared with the placebo group, especially concerning lung cancer [25]. This finding likely reflected the ability of IL-1β to impede invasion and metastasis of existing subclinical cancers rather than cancer initiation due to the median duration of the trial of 3.7 years [26]. In support of this, a substantial number of patients with diagnosed lung cancer already had circulating tumour DNA at the time of enrolment [27]. These findings spawned several clinical trials blocking IL-1β and other inflammatory interleukins in cancer with expected primary completion dates until 2024.

In 2018, a phase III trial of canakinumab as monotherapy compared with placebo started in patients with non-small-cell lung cancer (NCT03447769) [28]. Similarly, a phase II trial studying canakinumab monotherapy in myelodysplastic syndrome and chronic myelogenous leukaemia was initiated in 2020 (NCT04239157) [29]. Also, in 2018, a phase III trial using canakinumab in combination with pembrolizumab and chemotherapy in non-small-cell lung cancer (NCT03631199) [30], and in 2019 a phase III trial with canakinumab and chemotherapy alone (NCT03626545) were started [31]. Several additional studies were initiated between 2016 and 2020 (see Supplementary Table 1).

Anakinra is an IL-1R antagonist in clinical trials for cancer therapy. In a phase II trial, 47 patients with smouldering myeloma were treated with anakinra alone for 6 months. If a patient did not achieve a minor response or better, anakinra was combined with dexamethasone. Three of 47 patients achieved a minor response to anakinra alone, highlighting the need for a combinatorial approach. Upon addition of dexamethasone, five partial responses and four minor responses were achieved [32,33]. Anakinra is also under investigation in several other clinical trials in multiple cancer types as monotherapy and in combination therapy (see Supplementary Table 1).

Interleukin-6. While the anti-IL-6 antibody siltuximab reduced tumour growth in a xenografted cholangiocarcinoma model [34], it achieved only small effects in multiple myeloma as monotherapy [35], in combination with the proteasome inhibitor bortezomib [36] and with bortezomib, the cytostatic agent melphalan, and the anti-inflammatory glucocorticoid prednisone [37]. Siltuximab was also not effective in solid tumours as monotherapy. In prostate cancer, the best outcome achieved was stable disease [38], in renal cell carcinoma (RCC) the best outcome was a single partial response and no clinical efficacy was observed in a group of mixed solid tumours [39]. Furthermore, several clinical trials observed treatment-associated side effects, such as higher rates of infections upon siltuximab treatment [40–42], which overall does not support a benefit of siltuximab therapy.

IL-6 can protect cancer cells from therapy-induced DNA damage, oxidative stress and apoptosis and is known to induce immunosuppression and maintains highly therapy-resistant cancer stem cell populations [43]. Thus, IL-6 neutralization might reverse resistance to conventional cancer therapy. In combination with chemotherapy, the anti-IL-6R antibody tocilizumab
**a Recombinant and engineered interleukins**

**Increased persistence**

Inactive prodrug → Active form

PEGylated IL-2, IL-10, IL-12 or IL-15 → Clearance

**Targeted toxicity**

Toxin → IL-3 or IL-4 → Cell death

**Gene therapy**

Viral vector

**Targeted interleukin delivery**

IL-2, IL-12 or IL-15 Ab fusion → IL-2, IL-12 or IL-23 → Gene transcription

**b Complementing ACT**

**TRUCKs**

CAR → TIL

**Sensitizing CAR T cells to interleukins**

IL-7, IL-12 or IL-15 → IL-7R

**DC vaccine adjuvant**

DC → IL-2

**ACT adjuvant**

IL-2, IL-15

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**Immune checkpoint inhibitors (ICIs): Drugs used for therapeutic neutralization of signalling pathways that activate immune checkpoints, which normally lead to the inhibition of immune cell function to keep immune responses under control.**

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yielded first positive results. However, combining IL-6 neutralization with immune checkpoint inhibitors (ICIs) has engendered controversy. While IL-6 neutralization might enable patients to continue receiving ICIs by countering cytokine release syndrome (see BOX 2), IL-6 inhibition was shown to suppress the expression of PD1 and programmed death ligand 1 (PD-L1). This might decrease the effectiveness of ICIs. As preclinical studies mainly demonstrate asynergy between IL-6 neutralization and ICIs, several clinical trials have been initiated to test the combination in a clinical setting (see Supplementary Table 1).

Publications highlight the potential of blocking IL-6 trans signalling in hepatocellular carcinoma and colorectal cancer by comparing carcinogenesis in wild-type mice and mice transgenic for soluble gp130Fc, which consists of gp130 fused to a human IgG1 domain. These data may lead to clinical trials for omalizumab (also known as soluble gp130Fc) in patients with cancer in the future.

**Interleukin-8.** HuMax-IL-8 is an antibody neutralizing IL-8. While no objective responses were observed in monotherapy, no dose-limiting toxic effects were observed, and 73% of patients with solid tumours had stable disease. IL-8 induces resistance to several chemotherapeutic agents and ICIs, as summarized elsewhere. Therefore, several clinical trials are exploring the effect of HuMax-IL-8 in combination with nivolumab (NCT03400332), nivolumab and degralin (NCT03689699) and the anti-PD1 antibody nivolumab with stereotactic body radiotherapy (NCT04572451).

**Interleukin-17.** Blocking IL-17 in a murine model of gastric cancer led to a reduction of tumour growth and in combination with anti-PD1 therapy to tumour rejections in 80% of the mice. It also decreased the development of prostate cancer in a preclinical mouse model. IL-17 can accelerate the progression of multiple myeloma, and another study showed that IL-17 neutralization indeed can lead to a reduction in multiple myeloma growth. A clinical trial that started in 2017 (NCT03111992) is now exploring the efficacy of the anti-IL-17A antibody CJM112 in multiple myeloma.

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**Fig. 4 | Mechanisms of interleukin therapy.** a | Recombinant and engineered cytokines. Increased persistence: prolonging the half-life and controlling toxic effects by the progressive release of the active drug from conjugated polymers or Fc tags. Targeted toxicity: interleukin–toxin fusion proteins target the toxin to cells bearing the interleukin receptor, leading to cell death. Gene therapy: to avoid systemic toxic effects, the interleukin is expressed directly at the tumour site. Selective receptor affinity: interleukins can be engineered to alter their receptor affinity, thereby increasing efficacy or reducing side effects. Targeted interleukin delivery: by coupling of interleukins to tumour-targeting antibodies (Abs), they reach higher concentrations at the tumour site while decreasing side effects associated with high systemic concentrations. b | Complementing adoptive cell therapy (ACT). T cells redirected for antigen-unrestricted cytokine-initiated killing (TRUCKs); expression of interleukins by ACT cells leads to an accumulation of the interleukin at the tumour site, thereby avoiding systemic toxicity and mounting non-ACT immune responses by the activation of endogenous T cells (tumour-infiltrating T cells (TILs)) for targeted cancer cell killing, as well as macrophages, which can also mount an innate immune response against cancer cells that do not express the antigen. Sensitizing chimeric antigen receptor (CAR) T cells to interleukins: expression of interleukin receptors (for example, IL-7R) on ACT cells increases the likelihood of signalling at low interleukin concentrations. Dendritic cell (DC) vaccine adjuvant: DC vaccination can be accompanied by therapy with interleukins (for example, IL-2), enhancing the activation of DC-primed T cells. ACT adjuvant: increasing persistence, survival, proliferation and activation of ACT by interleukin therapy (for example, IL-2 or IL-15).
In addition to increasing its half-life, the polyethylene glycol (PEG) residues coupled to IL-2 block its binding site for the high-affinity IL-2Ra subunit. Over time, the PEG groups are lost and continually release the active form of bempegaldesleukin, thereby minimizing the risk of overdosing. Bempegaldesleukin was evaluated as monotherapy in phase I clinical trials in patients with metastatic solid tumours, predominantly consisting of patients with melanoma or RCC. The best overall responses observed were stable disease, which is disappointing in comparison with the 7% and 9% durable complete responses achieved by conventional IL-2 therapy in melanoma and RCC, respectively. Nevertheless, bempegaldesleukin showed a favourable safety profile, encouraging combination with ICIs. A phase I study of bempegaldesleukin in combination with nivolumab in metastatic solid tumours achieved a complete response rate of 19%. Several other clinical trials in combination with ICIs, including phase III clinical trials in patients with bladder cancer (NCT04209114), RCC (NCT03729245), or melanoma (NCT03635983), are currently ongoing. Other non-α IL-2 variants are being tested in clinical trials in combination with ICIs, such as ALKS 4230, a circularly permuted IL-2 and IL-2Ra designed to selectively activate the intermediate-affinity IL-2R. It is tested in four clinical trials in patients with advanced solid tumours or non-cutaneous squamous cell carcinoma of the head and neck alone and in combination with pembrolizumab (see Supplementary Table 1). THOR707 is an engineered non-α IL-2 being tested in patients with solid tumours alone and in combination with an unspecified ICI or with an anti-EGFR antibody (NCT04009681).

NARA1leukin is a fusion construct that consists of human IL-2 engineered into the anti-IL-2 antibody NARA1. NARA1 thereby permanently masks the CD25-binding site of IL-2 and thus abolishes the CD25-dependent IL-2Rα-mediated Treg cell development, while mediating potent antitumour responses in a murine melanoma model.

Another approach is the de novo design of Neo-2/15, a hyperstable interleukin that mimics only the binding site for IL-2Rβ-γc, but otherwise is structurally independent of IL-2 and IL-15. Therefore, it can mediate IL-2 and IL-15 signalling independently of IL-2Ra or IL-15Ra, and showed superior therapeutic activity compared with IL-2 in mouse models of melanoma and colon cancer.

Several attempts have been made to couple IL-2 to tumour-targeting antibodies to reduce side effects caused by high systemic concentrations while increasing the efficacy by concentrating IL-2 at the tumour. Some of those IL-2-antibody fusion constructs reached the stage of clinical trials. However, no breakthrough results have emerged so far (see Supplementary Table 1).

Interleukin-10. IL-10 has anti-inflammatory as well as CTL-stimulating properties, which led to its exploration as an anticancer therapeutic. The PEGylated IL-10 variant pegilodecakin can induce systemic immune activation, an increase of the levels of immunostimulatory cytokines and a reduction of TGFβ levels in patients
with solid cancer. Additionally, partial responses and prolonged stable disease were observed upon pegilodecakin treatment\(^{215}\). In a follow-up trial, in a mixed group of solid cancers, expansion and activation of intratumoural CTLs in response to treatment were observed\(^{216}\). The first clinical data on the combination of pegilodecakin with ICIs were promising as well\(^{217}\); however, two follow-up trials testing pegilodecakin in combination with nivolumab (NCT03382912 (REF.\(^{193}\)) and the anti-PD1 antibody pembrolizumab (NCT03382899 (REF.\(^{199}\)) were terminated early due to an unfavourable risk–benefit ratio.

CmAb-(IL-10), a bispecific fusion protein with one arm derived from the anti-EGFR antibody cetuximab and the other arm containing an IL-10 dimer, is an agent still in preclinical investigation. It has shown superior antitumour activity compared with IL-10 fused to a non-tumour-targeting antibody\(^{218}\).

**Interleukin-12.** IL-12 is an interleukin that demonstrates striking immune activation and tumour control but causes severe adverse effects with the doses necessary to induce an antitumour effect. However, targeted delivery strategies to achieve high concentrations at the tumour site while avoiding high systemic amounts led to a renaissance of IL-12 cancer therapy\(^{219}\). Gene therapy allows better control of the localization and amount of interleukins in a therapy regimen. Tavokinogene telseplasmid, an IL-12-encoding plasmid electroported into melanoma lesions, achieved an overall response rate of 36% and a complete response rate of 18% in a phase II trial. Of note, 46% of patients also showed regression in at least one lesion and 25% had a net regression in all lesions into which tavokinogene telseplasmid had not been injected\(^{220}\), suggesting the induction of systemic antitumour immunity. In combination with pembrolizumab, the overall response rate was increased to 41%, with a complete response rate of 36%\(^{199}\). Several follow-up trials were initiated between 2017 and 2020 in triple-negative breast cancer (NCT03566720 (REF.\(^{200}\))), head and neck squamous cell carcinoma (NCT03823131 (REF.\(^{201}\))) and melanoma (NCT04526730 (REF.\(^{202}\)) and NCT03132675 (REF.\(^{203}\))). Other therapeutic strategies using IL-12 have been extensively discussed elsewhere\(^{204,205}\), and recent clinical trials are listed in Supplementary Table 1.

CBD–IL-12 is a fusion protein consisting of IL-12 and a collagen-binding domain. It accumulates in the tumour due to exposed collagen in the disordered tumour vasculature and demonstrates antitumour effects superior to those of IL-12 by a broad activation of the immune system at the tumour site\(^{204}\). Li et al. designed multifunctional oncolytic nanoparticles with the intent to send several different signals triggering antitumour responses at once. They showed that the composition of nanoparticles containing self-replicating RNA can induce immunogenic cell death of tumour cells, as the contained RNA serves as a signal for pathogen-recognition receptors as well as coding for IL-12 to further enhance the induced immune response. When delivered intratumourally, control of large tumours is achieved in several tumour models\(^{205}\).

**Interleukin-15.** In contrast to IL-2, IL-15 does not lead to T\(_{reg}\) cell expansion, while having similar immunostimulatory properties. However, monotherapy with IL-15 and several engineered variants was ineffective, although there are several promising combination therapies in clinical trials\(^{216}\).

An example of this is ALT-803, consisting of recombinant IL-15 with an N72D mutation linked to the Sushi domain of IL-15Ra fused to an immunoglobulin G (IgG) Fc-fragment. While the N72D mutation increases binding to IL-15Rβ, the Sushi domain provides the IL-15Ra part and the Fc fragment increases its half-life. However, ALT-803 had only a limited effect as monotherapy in patients with solid tumours\(^{207}\) and showed responses in only 19% of patients with haematological malignancies\(^{208}\). Strikingly, in combination with intra-vascular bacillus Calmette–Guérin (BCG) therapy, complete responses were achieved in all nine patients with bladder cancer\(^{209}\), which led to the expansion of the trial to enrol nearly 600 patients (NCT02138734 (REF.\(^{210}\))).

Other IL-15-based therapeutic strategies include combination with ICIs and tumour-targeting antibodies (Supplementary Table 1); approaches for IL-15 in combination therapy for cancer have been extensively reviewed\(^{206,211}\).

**Interleukin-18.** IL-18 therapy has shown little preclinical efficacy, potentially due to IL-18BP, a high-affinity IL-18 decoy receptor. Using direct evolution, Zhou et al. developed decoy-resistant IL-18 (DR-18), which maintains signalling potential and unlike IL-18 exerted potent antitumour responses in mice, by supporting effector T cell development, reducing T cell exhaustion and enhancing activity and maturation of NK cells\(^{212}\). IL-36\(\gamma\) together with IL-23 can be successfully used in combination with OX40 ligand to promote acute inflammation for efficient tumour control\(^{213}\) and is currently under investigation in a clinical trial (NCT03739931 (REF.\(^{214}\))).

**Complementing ACT**

Interleukins play a key role in ACT, a breakthrough achievement in cancer therapy in the past decade. Interleukins such as IL-2, IL-7 and IL-15 have been widely used to improve in vitro expansion and differentiation of adoptive cells and to complement ACT therapy in humans by co-administration or by genetically engineering them into the transfused cells.

**Interleukin-2.** Treatment of patients with IL-2 to support ACT has been studied with variable success in several cancer types. For example, IL-2 was combined with tumour-infiltrating lymphocytes in melanoma\(^{214}\). While some of those trials yielded benefits, for example a correlation of chimeric antigen receptor (CAR) T cell engraftment with IL-2 concentrations\(^{215}\), this approach still faces toxicity problems similar to those of using interleukins alone.

A strategy to overcome the toxic effect of systemic IL-2 treatment while retaining its ability to support ACT therapy is orthogonal IL-2–IL-2R pairs. The designed
orthogonal IL-2 cannot bind to the wild-type IL-2R but transfers native IL-2 signalling when binding to the designed orthogonal IL-2Rβ, which was transduced into T cells. This creates a system where only the ACT product reacts to the orthogonal IL-2 treatment and systemic toxicity is avoided. Orthogonal IL-2 showed efficacy in a mouse melanoma model with the enhanced expansion of the orthogonal IL-2Rβ ACT product and negligible toxic effects. These data might indicate a potential advantage for clinical use in ACT.\(^{216}\)

**Interleukin-8.** A quite different approach is to use tumour-associated interleukins, such as IL-8, to improve infiltration of CAR T cells into the tumour by over-expressing their respective receptors. The expression of the IL-8 receptors CXCR1 or CXCR2 in CAR T cells markedly enhanced migration and persistence in the tumour and induced complete regression and long-lasting immunological memory in preclinical models of glioblastoma, ovarian cancer and pancreatic cancer.\(^{217}\)

**Interleukin-12.** Another strategy currently pursued in several trials is the expression of certain interleukins by cellular therapy products. In this manner, the interleukins should be expressed only locally at the tumour site, thereby avoiding systemic toxic effects and recruiting additional immune cells to establish immune responses against CAR antigen-negative tumour cell populations. In the case of T cells, this approach was named ‘T cells redirected for antigen-unrestricted cytokine-initiated killing’ (TRUCKs). The first interleukin used as a proof of concept for TRUCKs was IL-12 (REF.\(^{218}\)), and other interleukins, such as IL-7, IL-15 and IL-23, have since been used to engineer T cells as well.

In preclinical testing, IL-12 expression by CAR T cells led to the recruitment and activation of macrophages critical for the elimination of cancer cells with antigen loss that would normally escape from CAR-induced killing.\(^{219}\) The first in-human trial was a study expressing IL-12 under the control of a nuclear factor of activated T cells (NFAT)-inducible promoter in tumour-infiltrating lymphocytes from patients with metastatic melanoma. However, the study had to be terminated due to severe toxic effects likely caused by excessive IL-12 release.\(^{219}\) Recently, several clinical trials were started using IL-12-expressing CAR T cells, trying to avoid uncontrolled IL-12 release by the more restricted activation of CARs compared with polyclonal tumour-infiltrating lymphocytes and by using safety switches (see Supplementary Table 1).

Another approach is to induce only transient expression of IL-12 by electroporating IL-12 mRNA into ovalbumin-specific OT-I TCR transgenic CD8+ T cells. Injected intratumorally, but not intravenously, these cells led to complete rejections of lesions into which they had been injected and distant lesions in a B16-ovalbumin tumour model. The study authors claim that the transient expression was likely helpful in restricting IL-12-induced adverse events but do not show any data to prove this assumption.\(^{220}\)

A recent clinical trial using intratumourally injected activated DCs showed that successful DC therapy was associated with higher expression of IL-12 subunit p40 and IL-8 (REF.\(^{217}\)). This may indicate a favourable interplay between DC therapy and interleukin therapy.

**Interleukin-7.** Over time, the idea to express interleukins in the ACT product has spread from IL-12 to other interleukins. IL-7 and CC-chemokine ligand 19 (CCL19) are essential for the maintenance of T cell zones in lymphoid organs. In a preclinical setting, CAR T cells engineered to express IL-7 and CCL19 were superior in tumour killing in comparison with conventional CAR T cells. Histopathological analysis showed increased infiltration of DCs and CAR T cells as well as endogenous T cells into the tumour. The depletion of endogenous T cells dampened the therapeutic effect of IL-7 and CCL19-expressing CAR T cells. Together with the observation of abundance of memory CAR T cells and endogenous T cells in the spleen, this suggests co-expression of IL-7 and CCL19 by CAR T cells can initiate a CAR-independent immune response.\(^{222}\)

Four clinical trials evaluating the effect of CAR T cells overexpressing IL-7 and CCL19 were initiated between 2017 and 2020 (see Supplementary Table 1).

**Interleukin-15.** Preclinical studies showed that the expression of IL-15 in NKT cells led to a decreased expression of exhaustion markers, enhanced in vivo persistence, increased localization at the tumour site and improved tumour control.\(^{221}\) In CAR T cells, membrane-bound IL-15 led to an increase in the abundance of T memory stem cells with long persistence and potent rejection of CD19+ leukaemia.\(^{223}\) This led to a collection of ongoing clinical trials engineering IL-15 into CAR T cells and CAR NKT cells (Supplementary Table 1). An interim analysis showed that anti-GD2 CAR NKT cells expressing IL-15 did not cause any dose-limiting toxic effects in three heavily pretreated children with neuroblastoma. CAR NKT cells were shown to be present at the tumour sites, and one patient achieved regression of bone metastatic lesions.\(^{224}\)

Recently, a novel approach to reduce the toxicity of IL-15 was reported. Tang et al. designed a cell surface-conjugated nanogel that can be loaded with large quantities of protein drugs.\(^{225}\) The nanogel is designed to release its cargo upon an increase in T cell surface reduction potential, which is induced by T cell–antigen encounter. Thereby, the drug release is limited to places of TCR activation, predominantly the tumour site. The study authors also show that the system is translatable to CAR T cells. The potential to expand this method to other cytokines and protein drugs, maybe even combinations thereof, makes this system especially noteworthy, although the approach is inherently self-limiting as the nanogels will be depleted of their drugs over time.\(^{226}\)

**Interleukin-23.** IL-23 is a very recently reported alternative to potentially toxic interleukins such as IL-12 and IL-15. A preclinical study showed that engineering CAR T cells with IL-23 subunit p40 led to autocrine IL-23 signalling promoting the selective proliferation of activated T cells with increased antitumour capacity. Additionally,
Outlook

Limited understanding of the impact of interleukin therapy on carefully balanced systems in the TME, as well as the Janus-like functions of some interleukins, rendered the therapeutic exploitation of interleukins difficult to predict. This caused much disappointment in the field of interleukin therapy for cancer, as life-threatening side effects occurred and most therapeutic strategies did not live up to the expectations concerning effectiveness.

One example is the targeting of one of the first interleukins to be discovered (IL-1β), in which case neutralization could yield either disease exacerbation or reduction. Recent developments have pointed out that neutralization of this key pathway at a time point where the disease is already present might have therapeutic activity in patients with cancer. Also, IL-2 monotherapy in melanoma and RCC achieved durable complete responses, highlighting the potential of interleukin therapy.

However, in general, interleukin monotherapy still faces major limitations, and new strategies have emerged recently to tackle those problems. Thus, the trend is moving away from applying native forms of interleukins and is going towards sophisticated engineering approaches. Most strategies investigated in the past few years involved altering the receptor affinity for only the target selected cell populations or designing tumour-targeting fusion constructs to increase the efficacy and reduce systemic toxic effects.

Additionally, to improve the application of interleukin therapy, combination approaches are gaining more and more importance. New insights into how interleukins shape tissue repair, cancer progression and immune evasion will greatly help to identify how interleukins are involved in the resistance against established therapies such as chemotherapy or ICI treatment. Thereby, interleukin therapy could unleash the unused potential of other therapeutics and thus help otherwise therapy-resistant patients to benefit therefrom.

Another hypothesis on the low efficacy of interleukin monotherapy is the idea that delivering a single immunostimulatory signal is not sufficient to mount a durable immune response that can eliminate the tumour. Hence, strategies combining several immunostimulatory signals have been fast approaching. A strategy often pursued in this regard is the combination of interleukin therapy to support ACT, either by combining ACT with regular interleukin therapy or by the development of TRUCKs. ACT showed remarkable success in haematological malignancies but lacks efficiency in solid tumours due to poor infiltration and proliferation of the ACT product as well as a highly immunosuppressive TME impairing its function. Interleukins may support ACT by overcoming all of those hurdles, and the number of clinical trials in this area highlights the expectations set by the idea.

Conclusion

In conclusion, interleukins comprise key elements that orchestrate the TME and govern tumour–immune cell crossstalk as they are important players in large cytokine networks. Although interleukin or interleukin-targeted therapy still has significant hurdles to overcome on its path to the clinic, the intensive foundational research conducted on interleukins in cancer biology in the past decade will help us to better understand the mechanisms of interleukin therapy and thereby contribute to the development of new strategies. Also, there is a rich pipeline of exciting new preclinical and clinical studies on interleukin therapy for cancer that will yield interesting results in the coming years.

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