Rethinking immune checkpoint blockade: ‘Beyond the T cell’

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ABSTRACT
The clinical success of immune checkpoint inhibitors has highlighted the central role of the immune system in cancer control. Immune checkpoint inhibitors can reinvigorate anti-cancer immunity and are now the standard of care in a number of malignancies. However, research on immune checkpoint blockade has largely been framed with the central dogma that checkpoint therapies intrinsically target the T cell, triggering the tumoricidal potential of the adaptive immune system. Although T cells undoubtedly remain a critical piece of the story, mounting evidence, reviewed herein, indicates that much of the efficacy of checkpoint therapies may be attributable to the innate immune system. Emerging research suggests that T cell-directed checkpoint antibodies such as anti-programmed cell death protein-1 (PD-1) or programmed death-ligand-1 (PD-L1) can impact innate immunity by both direct and indirect pathways, which may ultimately shape clinical efficacy. However, the mechanisms and impacts of these activities have yet to be fully elucidated, and checkpoint therapies have potentially beneficial and detrimental effects on innate antitumor immunity. Further research into the role of innate subsets during checkpoint blockade may be critical for developing combination therapies to help overcome checkpoint resistance. The potential of checkpoint therapies to amplify innate antitumor immunity represents a promising new field that can be translated into innovative immunotherapies for patients fighting refractory malignancies.

INTRODUCTION
Cancer research was transformed with the discovery that tumor-specific T cell dysfunction was reversible with immune checkpoint blockade.1 Antagonistic antibodies targeting cytotoxic T-lymphocyte-associated protein 4 (CTLA-4) and programmed cell death protein-1 (PD-1) or programmed death-ligand-1 (PD-L1) stimulate antitumor immunity and are now approved therapies in many cancer types, including metastatic melanoma and non-small cell lung cancer.2 These clinical successes highlight the immense potential for T cell-directed immunotherapy in cancer; however, we are just beginning to understand the full molecular activity of such agents. The remarkable achievements of these therapies in the clinic have elevated the T cell above all other immune lineages in the realm of antitumor immunity. Thus, the scope of research into immune checkpoint blockade may have been limited by ‘T cell centrism’. Growing evidence, reviewed below, highlights the emerging appreciation that innate immune cells mediate key aspects of checkpoint therapy biology. Despite numerous clinical successes, many patients do not respond to checkpoint therapies, and some cancer types are almost entirely resistant. An improved understanding of the mechanisms by which current checkpoint inhibitors function will enable clinicians to broaden the benefits of these treatments to greater numbers of patients.

Keeping T cells in check
PD-1 and its ligands are central to regulating inflammation and peripheral tolerance. PD-1-null mice spontaneously develop a lupus-like syndrome due in part to uncontrolled proliferation of autoreactive T cells.3 PD-1 restrains T cell activity when engaged by its ligands, PD-L1 and PD-L2.4 PD-L1 expression is inducible in a variety of cell types, including adaptive and innate immune cells, mesenchymal cells, and cancer cells.4 In contrast, expression of PD-L2 is limited to antigen-presenting cells (APCs) and a smaller subset of tumor cell types.4 PD-1/PD-L1 signaling profoundly modulates T cell cytokine secretion, dampens T cell receptor (TCR) signaling, and shortens synapse engagement between T cells and APCs, resulting in impaired antitumor immunity.5 PD-1/PD-L1 blockade partially reverses these negative effects, augmenting T cell proliferation, tumor infiltration, and cytotoxicity.4

CTLA-4 is another crucial T cell inhibitory receptor, which is upregulated in activated T cells and natively expressed by regulatory T cells (Tregs).6 In resting T cells, CTLA-4 is stored within cytosolic endosomes.6 After TCR engagement and costimulatory signaling via CD28, CTLA-4 molecules translocate to the cell surface, where they outcompete CD28 for its ligands, B7.1 and B7.2, expressed in APCs, restraining proliferation.

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and activation of T cells. CTLA-4 has a non-redundant immunosuppressive role; CTLA-4-deficient mice die at 1 month of age as a result of a lethal lymphoproliferative disorder. In multiple models, CTLA-4 blockade results in T cell-mediated tumor rejection. These findings spurred clinical trials that demonstrated the efficacy of anti-CTLA-4 in multiple cancers as a single agent or in combination with anti-PD-1.

The impact of checkpoint inhibitors on innate immune cells
Over the last two decades, research on checkpoint inhibitors has focused on the T cell as the principal therapeutic target; however, recent studies have highlighted the significant effects of checkpoint inhibitors on innate immune cells. Checkpoint blockade has both a direct and an indirect impact on innate immune lineages (figure 1). In the indirect pathway, anti-PD-1/PD-L1 or anti-CTLA-4 reinvigorates T cell immunity, which, in turn, shapes or phenotypically polarizes innate immune cell responses within the tumor microenvironment (TME). In the direct pathway, innate immune cells are direct targets of immune checkpoint blockade because subtypes of myeloid cells and innate lymphocytes express PD-1 and/or PD-L1. This highly nuanced interplay of cell types after checkpoint therapy testifies to the importance of investigating how checkpoint biology affects innate immune populations.

Tumor-associated macrophages and other myeloid cells in PD-1/PD-L1 checkpoint blockade
Indirect regulation
Macrophage function is orchestrated by activated T cells. T cell-associated cytokines such as interferon gamma (IFN-γ) stimulate macrophages to increase expression of major histocompatibility complex (MHC) molecules, costimulatory receptors, and the Th1-polarizing cytokine IL-12. Accordingly, checkpoint blockade-activated T cells dramatically alter phenotypes of tumor-associated macrophages (TAMs) and monocytes. Gubin and colleagues used single cell RNA-sequencing (scRNA-seq)
and mass cytometry to assess transcriptional and functional changes in tumor-infiltrating myeloid cells after treatment with anti-PD-1. PD-1 blockade resulted in a reduction in CD206+ TAMs and an increase in inducible nitric oxide synthase (iNOS) TAMs. This iNOS TAM cluster was enriched for genes involved in IFN-γ signaling, high glycolytic activity, and NF-κB activity, suggestive of antitumor potential. Moreover, on IFN-γ neutralization, anti-PD-1-mediated repolarization of TAMs/monocytes was significantly diminished. These results demonstrate the potential of checkpoint-activated T cells to secrete factors, such as IFN-γ, that remodel the TME toward a tumor hostile environment, rich in iNOS TAMs, which are associated with improved outcomes in many tumor models. Thus, anti-PD-1 can be added to a growing list of therapies aimed at ‘repolarizing’ TAMs away from a tumor permissive phenotype.

Direct regulation
Myeloid cells display detectable PD-1 levels. PD-1 expressing TAMs have been shown to promote tumor progression in several cancers including gastric cancer, colorectal cancers, and lung cancer. The appearance of PD-1 in myeloid progenitors is an early event in tumor progression, as the receptor may be induced by inflammatory conditions. Indeed, bone marrow–derived macrophages (BMDMs) rapidly upregulate PD-1 after Toll-like receptor (TLR)–2 engagement; correspondingly, PD-1 was upregulated by macrophages in a murine model of sepsis. The signaling downstream of PD-1 in macrophages is controversial and may be contextually dependent. PD-1-null BMDMs express more IL-6, and CCL2 (MCP-1) at 4-hour post TLR2 stimulation, suggesting an anti-inflammatory role of PD-1. In contrast, in vivo evidence found the opposite effect. Septic PD-1-null mice demonstrate decreased levels of peritoneal CCL2, tumor necrosis factor-alpha (TNF-α), and IL-1β, an observation that was abrogated on depletion of peritoneal macrophages. These in vivo data suggest that PD-1 expression on macrophages augments systemic inflammation. Despite the lack of a clear signaling pathway downstream of PD-1 in macrophages, there is a clear link between TAM expressed PD-1 and cancer-associated inflammation. In cancer models, synergistic antitumor effects were found with the combination of PD-1/PD-L1 blockade and neutralization of either IL-6 or IL-1β, indicating that targeting the inflammatory TME could amplify PD-1 blockade efficacy. This effect likely involves macrophage expressed PD-1, as the authors found that PD-1 agonism suppressed production of IL-6 by PD-1-bearing TAMs, whereas anti-PD-L1 enhanced IL-6 production. Thus, anti-inflammatory therapies may be necessary for maximizing the benefit of checkpoint blockade. These data suggest a complex role of myeloid intrinsic PD-1 signaling and highlight the need to delineate the mechanistic differences between PD-1 blockade and PD-1 deletion.

New model systems have permitted the investigation of the myeloid-specific effects of checkpoint blockade. Strauss et al generated a mouse model in which PD-1 was selectively deleted in myeloid cells. The authors employed these mice to dissect the relative contribution of myeloid versus T cell PD-1 signaling in colon cancer. Interestingly, myeloid-specific PD-1 deletion was as effective at limiting tumor growth as global PD-1 deletion, and more effective than selective ablation of PD-1 in T cells. One caveat to these studies is that genetic approaches to interrupt PD-1/PDL-1 signaling may not accurately model therapeutic antagonist therapies. However, the authors treated Recombination Activating Gene-2-null mice lacking T and B cells with anti-PD-1 and still observed a significant reduction in tumor growth, again emphasizing the critical importance of the innate immune system for checkpoint blockade.

PD-1 engagement on myeloid cells affects infiltration, differentiation, effector function, and cellular metabolism. Some of these pathways and outcomes are highlighted in figure 2. PD-1 engagement shifts activated human monocyte metabolism toward oxidative phosphorylation. PD-1/PD-L1 blockade is able to rescue glycolysis, which is correlated with enhanced antibody-dependent phagocytosis. PD-1-deficient myeloid cells exhibit altered development from common myeloid progenitors, with diminished accumulation of granulocyte/macrophage progenitors in the bone marrow and increased expansion of Ly6C+ monocytes and dendritic cells (DCs) within the tumor. These data suggest that PD-1 signaling in myeloid progenitors may direct myelopoiesis toward the granulocytic lineage, resulting in greater numbers of immunosuppressive granulocytic-MDSCs. These findings suggest that checkpoint therapies may benefit from drug combinations that limit tumor infiltration by myeloid subsets. Regarding effector function, PD-1 expressing TAMs demonstrate high levels of CD206, arginase 1 (ARG1), and IL-10, which dampen antitumor immune responses. In contrast, PD-1 deficiency in TAMs shifts their phenotype toward an antitumor profile, with higher levels of TNFα, iNOS, and MHCII. In multiple cancer models, TAM infiltration is skewed toward CD206+, ARG1-high macrophages; however, anti-PD-1 therapy reverses this trend, increasing the expression of iNOS, TNF-α, and IL-6, which may augment antitumor immunity. These findings corroborate the scRNA-seq results of Gubin et al and strikingly highlight that at a transcriptional level, checkpoint therapy has a concomitant, if not greater, impact on TAM phenotype than on T cell phenotype. Together, these data suggest that PD-1 blockade reprograms TAMs toward an antitumor phenotype.

Myeloid-specific effects of PD-L1 blockade
Canonically, PD-L1 interacts with its receptor PD-1 on tumor-specific T cells and limits their antitumor activity. Anti-PD-L1 therapy blocks this interaction, thereby reinvigorating T cell proliferation and effector functions such as IFN-γ secretion. However, like anti-PD-1, PD-L1 blockade can also directly and indirectly modulate myeloid cell function. Anti-PD-L1 has been shown to indirectly
repolarize TAMs toward a proinflammatory phenotype, in a T cell-dependent, IFN-γ-mediated process. These anti-PD-L1-treated TAMs exhibit decreased expression of ARG1 and enhanced iNOS, MHCII, and CD40 expression, indicative of an antitumor phenotype. In the direct pathway, TAMs can engage with activated T cells expressing PD-L1. T cell PD-L1 binds TAM-expressed PD-1 and induces a tolerogenic phenotype. These findings indicate that anti-PD-L1 may disrupt multiple axes of PD-1 engagement to restore the antitumor potential of TAMs.

The bulk of research on anti-PD-L1 therapy emphasizes the disrupted interaction between tumor-expressed PD-L1 and T cell-expressed PD-1. However, PD-L1 is widely inducible on immune subsets, tumor cells, and even endothelial cells, in an IFN-γ-dependent process. Thus, both tumor and host PD-L1 are intimately involved in checkpoint blockade. In some malignancies such as colon cancer, tumor immune infiltrates express PD-L1 at significantly greater levels than tumor cells. Thus, it is essential to study the role of tumor extrinsic PD-L1. Indeed, PD-L1 expressing lymph node resident APCs can inhibit T cell activation and prevent recruitment of primed T cells to the TME. PD-L1+ neutrophils have been shown to impair T cell immunity in hepatocellular carcinoma. In some models, PD-L1 expression by cancer cells is dispensable for anti-PD-L1 efficacy. PD-L1 blockade efficacy was retained in a PD-L1-deficient model of colon adenocarcinoma. However, efficacy was lost if the bone marrow from PD-L1-null mice was transplanted into tumor-bearing mice, indicating that a hematopoietic cell was responsible for the response. The efficacy was again lost after depleting CD11b+ PD-L1+ cells, presumably due to an absence of PD-L1-expressing APCs. These data support the essential role of PD-L1+ myeloid cells in anti-PD-L1 therapy. Other authors have disputed the dispensable nature of tumor expressed PD-L1 and have demonstrated that both tumor and APC expressed PD-L1 are involved in critical but distinct aspects of PD-1/PD-L1 blockade. These findings have broad implications for the use of PD-L1 as a biomarker for checkpoint efficacy. Patients may benefit from separate quantification of
tumor and tumor immune infiltrate PD-L1 expression; and differing cancer types may have divergent biology herein.

The portrait of myeloid PD-L1 engagement is further complicated by in vitro evidence that PD-L1 can have intrinsic signaling of its own. When engaged, PD-L1 can induce proliferation, costimulatory molecule expression, cytokine production, and mTOR signaling in macrophages. It is noteworthy that TAMs can express both PD-L1 and its ligands PD-1 and B7.1 (CD80), which have sufficient affinity to interact in cis. The evidence presented above suggests that PD-L1 inhibitors may have intrinsic effects on myeloid cells by either preventing direct engagement in trans or disrupting cis interactions. Further research is needed to elucidate the primary mode of myeloid specific anti-PD-L1 efficacy. Potential modalities include inhibition of PD-L1 engagement on myeloid cells, disruption of myeloid PD-L1 engagement with T cell PD-1 or inhibition of direct signaling through myeloid PD-L1. Regardless of the primary modality, it is increasingly appreciated that blocking PD-L1 may have a very different outcome compared with blocking PD-1. Indeed, in a model of pancreatic cancer, the combination of PD-1 and PD-L1 confer a synergistic benefit over either therapy alone. Based on the essential role of PD-1/PD-L1 in myeloid cells, it is likely that the success of anti-PD-L1 and anti-PD-1 combined therapy relies both on the disruption of immunosuppressive TAM-T cell crosstalk and on directly reshaping TAM phenotype toward an antitumor profile.

**Linking PD-1/PD-L1 blockade and DC function**

**Indirect regulation**

The efficacy of PD-1 blockade may depend on the indirect activation of tumor-infiltrating DCs. Anti-PD-1-activated T cells secrete IFN-γ, which in turn sets in motion a dramatic transcriptional shift in DC phenotype, as they express antigen presentation machinery, upregulating dramatic transcriptomic shift in DC phenotype, as they induce proliferation, costimulatory molecule expression, cytokine production, and mTOR signaling in macrophages. It is noteworthy that TAMs can express both PD-L1 and its ligands PD-1 and B7.1 (CD80), which have sufficient affinity to interact in cis. The evidence presented above suggests that PD-L1 inhibitors may have intrinsic effects on myeloid cells by either preventing direct engagement in trans or disrupting cis interactions. Further research is needed to elucidate the primary mode of myeloid specific anti-PD-L1 efficacy. Potential modalities include inhibition of PD-L1 engagement on myeloid cells, disruption of myeloid PD-L1 engagement with T cell PD-1 or inhibition of direct signaling through myeloid PD-L1. Regardless of the primary modality, it is increasingly appreciated that blocking PD-L1 may have a very different outcome compared with blocking PD-1. Indeed, in a model of pancreatic cancer, the combination of PD-1 and PD-L1 confer a synergistic benefit over either therapy alone. Based on the essential role of PD-1/PD-L1 in myeloid cells, it is likely that the success of anti-PD-L1 and anti-PD-1 combined therapy relies both on the disruption of immunosuppressive TAM-T cell crosstalk and on directly reshaping TAM phenotype toward an antitumor profile.

**DC-specific effects of PD-L1 blockade**

Anti-PD-L1 therapy is now employed to treat numerous solid tumor types; however, the mechanism of action still remains debated. One clue toward this mechanism is the recent observation that expression of PD-L1 by intratumor immune infiltrate is a better correlate to anti-PD-L1 clinical response than tumor expressed PD-L1. Interestingly, a DC-specific transcriptional signature, calculated from the expression levels of XCR1, BATF3, IRF8, and Flt3, stratified anti-PD-L1-treated renal cell carcinoma and lung cancer patients into long-term and short-term survivors, suggesting that DC-expressed PD-L1 may be a primary target of successful anti-PD-L1 therapy. PD-L1 is widely expressed in DC subsets, with increased expression in the setting of inflammation and cancer. DC-expressed PD-L1 acts as a homeostatic control of autoimmunity and directly restrains highly active T cell responses. In two recent publications, researchers sought to study the contribution of DCs to
anti-PD-L1 therapy by conditionally deleting PD-L1. In both cases, selective ablation of PD-L1 on DCs restricted tumor growth as effectively as systemic PD-L1 knockout mice, indicating that DCs are likely critical to the reinvigorated T cell response after anti-PD-L1 immunotherapy. At early timepoints, DC-specific PD-L1 deletion led to enhanced tumor infiltration of effector CD8+ T cells, but not proliferation, indicating there may be increased T cell recruitment. These findings highlight that checkpoint-responsive patients may be best identified by measuring DC expression of PD-L1 rather than tumor expression.

Aside from the classical PD-1/PD-L1 axis, APC-expressed PD-L1 can also bind the costimulatory molecule B7.1 (CD80) in cis with higher affinity than the canonical T cell costimulatory receptor CD28. Consequently, T cell immunity may be suppressed by both the PD-L1–B7.1 and the PD-L1–PD1 axes, either by directly dampening TCR signaling or by restricting a necessary costimulatory signal. In two recent publications, researchers reached differing conclusions as to whether blocking PD-L1’s cis interaction enhances CD28 signaling. Disrupting the PD-L1–B7.1 interaction may in fact accelerate tumor growth by permitting PD-L1/PD-1 engagement; however, this observation was accompanied by a significant increase in intratumoral T cell numbers, indicating that enhanced priming via CD28-B7.1 may have also occurred. Further research is needed to clarify the role of cis PD-L1–B7.1 binding on DCs, and whether DC-specific PD-L1 engages PD-1 expressed by other myeloid subsets such as TAMs. DCs are uniquely specialized APCs that efficiently prime naive T cell responses. The evidence above demonstrates that DCs are critical for anti-PD-L1 efficacy. Thus, PD-L1 blockade may offer a targeted approach to improve DC-mediated priming of antitumor T cell responses, by both releasing B7.1 and disinhbiting downstream TCR activation.

The complex role of MDSCs in PD-1/PD-L1 blockade
MDSCs are a heterogeneous group of relatively immature myeloid cells, which can play an immunosuppressive role in multiple cancers. The effects of PD-1/PD-L1 blockade on MDSCs are multifaceted and represent a double-edged sword because they may simultaneously induce antitumor immunity and promote tolerance.

Indirect regulation
While anti-PD-1/PD-L1 therapies have shown potential to induce antitumor myeloid activity, they can also have complex protumorigenic outcomes, such as recruiting immunosuppressive MDSCs. Circulating MDSCs are associated with poor survival in patients with melanoma who have received checkpoint therapy. The frequency of tumor-infiltrating MDSCs is significantly elevated in metastatic melanoma biopsies obtained from patients receiving anti-PD-1 therapy. This recruitment may be driven by anti-PD-1-activated T cells secreting IFN-γ, which partially triggers the tumor-intrinsic NLRP3 inflammasome. NLRP3 activity drives CXCL5-mediated recruitment of granulocytic MDSCs. Thus, anti-PD-1 therapy can lead to anti-PD-1 resistance, in an apparent negative feedback loop. In addition to IFN-γ, checkpoint-activated CD8+ T cells secrete more TNF-α, both of which lead to tumor production of CSF1. CSF1, in turn, induces the differentiation and survival of protumorigenic TAMs and MDSCs, thus magnifying checkpoint resistance. Based on these data, one clear strategy to improve checkpoint efficacy is to limit the infiltration of MDSCs. Indeed, in models of pancreatic cancer, colon cancer, and breast cancer, blockade of CXCR2 suppresses the recruitment of MDSCs, triggering antitumor immunity, increasing T cell numbers, and sensitizing tumors to anti-PD-1 therapy. The above studies clearly demonstrate that PD-1/PD-L1 checkpoint blockade may indirectly drive checkpoint resistance by expanding and recruiting MDSCs. Treatment of multiple solid tumors may benefit from combining checkpoint therapy with targeted strategies to limit MDSC chemotaxis.

Direct regulation
MDSCs may express both PD-1 and PD-L1, which can lead to a reversal of MDSC-related immunosuppression when targeted by checkpoint blockade. As with other myeloid lineages, MDSC expression of PD-1 and PD-L1 is inducible in inflammatory settings. Activated T cells may promote MDSC expression of PD-L1 through the IFN-γ–IFNGR1–STAT1–IRF1 axis. MDSCs with high expression of PD-1/PD-L1 demonstrate high rates of proliferation, leading to their robust expansion in the TME of many cancers. The joint expression of PD-1 and PD-L1 by MDSCs suggests that both anti-PD-1 and anti-PD-L1 should be evaluated for their MDSC remodeling potential. Indeed, in a model of multiple myeloma, joint therapy with anti-PD-1 and anti-PD-L1 prevented MDSC-mediated cancer promotion to a greater extent than either alone. In contrast to cancers such as melanoma, head and neck squamous cell carcinoma (HNSCC) demonstrates reduced granulocytic MDSC infiltration post-PD-1/PD-L1 blockade. It is unclear whether this model involves different mechanisms of MDSC recruitment or whether the direct blockade of PD-1 is inhibiting MDSC proliferation. Antagonism of PD-L1 in MDSCs has been shown to reduce the immunosuppressive polarization of T cells. In coculture experiments, treatment of MDSCs with anti-PD-L1 led to increased rates of T cell proliferation and IFN-γ production, which may be an outcome of reduced IL6 and IL10 production. The studies discussed above highlight the nuanced and complex effects of checkpoint therapy on MDSCs. PD-1/PD-L1 blockade can improve outcomes via a direct reversal of MDSC-related immunosuppression, while synchronously driving checkpoint resistance through MDSC recruitment.

Innate lymphocytes and PD-1/PD-L1 blockade
Recent studies have identified innate lymphocytes as novel targets of checkpoint inhibitors. Innate lymphocytes are divided into two large branches: natural killer (NK)
cells and innate lymphoid cells (ILCs). ILCs are further divided into ILC1s, ILC2s, ILC3s, and lymphoid tissue-inducer (LTi) cells. NK cells functionally mirror CD8+ T cells because they exhibit antitumor cytotoxic activity. In comparison, ILC subsets are tissue-resident populations with analogous roles to CD4+ T helper (Th) cell subsets. ILC1s produce IFN-γ to control intracellular pathogens, ILC2s produce IL-4, IL-5, and IL-13 to target parasites, and ILC3s secrete IL-17 and IL-22 to defend against extracellular bacteria and fungi. LTi cells are critical for the formation of secondary lymphoid tissues.

**NK cells**

Solid tumors, such as renal cell carcinoma, can be heavily infiltrated by NK cells. These innate lymphocytes are regulated by constitutively expressed activating and inhibitory receptors, which recognize stress-induced ligands and various conserved motifs on class I and non-canonical MHC molecules. There are conflicting reports as to whether NK cells express significant levels of PD-1. Some groups suggest that neither mouse nor human NK cells express PD-1, however, other groups have reported PD-1 expressing NK cells in several cancer types. It is likely that PD-1 is expressed by NK cells in certain inflammatory conditions. Tumor-educated NK cells upregulate PD-1, which, when engaged by PD-L1, dampens NK cell-mediated antitumor immunity. Emerging evidence suggests that PD-1/PD-L1 blockade increases both NK cell recruitment and cytotoxicity against multiple myeloma cells. Additionally, anti-PD-1 therapy is capable of triggering NK cell activation and production of IFN-γ. To further complicate matters, NK cells may express PD-L1, which induces enhanced antitumor functionality when agonized. In vivo studies suggest that the use of anti-PD-L1 not only blocks negative PD-1 engagement on NK cells, but also activates PD-L1+ NK cells, leading to enhanced tumor rejection. Although NK cells likely represent an additional target of PD-1/PD-L1 blockade, additional research is needed to determine whether NK cell-specific PD-1/PD-L1 blockade is clinically relevant in solid tumors, and whether a physiological role exists for PD-L1 signaling in these innate immune cells.

**Innate lymphoid cells**

Checkpoint inhibitors are also capable of reshaping ILC responses in pathological conditions such as cancer. While high PD-1 expression on ILC progenitors is lost on differentiation, PD-1 levels may be upregulated in response to tissue-specific cues. PD-1 is upregulated in tissue-resident ILC2s in the context of lung inflammation. The depletion of PD1<sup>high</sup> effector ILC2s reduces inflammation during influenza and allergen exposure. Expression of PD1 on ILCs is also relevant for antitumor immunity. Tumor-infiltrating ILC2s (TILC2s) are predictive of long-term survival in patients with pancreatic cancer. TILC2s express significant levels of PD-1, and TILC2 density increases post-anti-PD-1 therapy. In an orthotopic model of pancreatic cancer, TILC2s adoptively transferred into ILC2-deficient hosts were partially responsible for the reduction in tumor burden post-anti-PD-1 therapy. In addition to ILC2s, ILC3s are also affected by checkpoint therapies. ILC3s express PD-1 in primary and metastatic tumors and the PD-1/PD-L1 axis in ILC3s regulates cytokines secretion and immune tolerance. Although still in its early stages, research into the role of ILCs during checkpoint therapy represents a promising new field for cancer immunotherapy.

**The impact of CTLA-4 blockade on innate immune cells**

Although CTLA-4 expression is largely restricted to T cell lineages and some cancers, there are numerous indirect effects of anti-CTLA-4 therapy on innate immune subsets. Anti-CTLA-4 blockade reduces the numbers of tumor-infiltrating MDSCs and protumorigenic TAMs in a spontaneous model of HNSCC. In addition, patients with advanced melanoma treated with anti-CTLA-4 exhibit decreased intratumoral MDSC numbers with a reversal in their tolerogenic profiles. As mentioned previously, Gubin et al demonstrated the ability of checkpoint blockade, including anti-CTLA-4 therapy, to reshape the myeloid compartment. Anti-CTLA-4 therapy indirectly polarizes TAMs, in an IFN-γ-dependent process, toward an antitumor phenotype characterized by the increased expression of NF-κB-related genes. Of note, a recent publication demonstrated that the efficacy of human anti-CTLA-4 was partially attributable to the Fc portion of the antibody and it’s affinity to Fcγreceptors such as CD32a, expressed by multiple innate subsets. Treatment with anti-CTLA-4 resulted in depletion of CTLA-4 expressing Tregs, highlighting an additional role of innate subsets responsible for antibody-dependent-cellular cytotoxicity during checkpoint blockade. However, these effects may be highly dependent on the IgG class of the CTLA-4 targeting antibody. There is more limited evidence for the direct engagement of CTLA-4 on innate immune lineages. Subsets of tumor-infiltrating NK cells express CTLA-4 and CD28, and CTLA-4 blockade was found to inhibit IFN-γ release by NK cells on coculture with mature DCs. In summary, like anti-PD1/PD-L1 therapies, anti-CTLA-4 therapy is capable of inducing global shifts in tumor-infiltrating innate immune subsets. Reports of the expression of CTLA-4 and CD28 in tumor-infiltrating NK cells suggest that further efforts are needed to assess whether the NK cell-specific effects of anti-CTLA-4 blockade have been misattributed to T cells.

**Emerging checkpoints inhibitors and their impact on innate immune cells**

The clinical success of checkpoint therapies targeting PD-1/PD-L1 and CTLA-4 spurred intense interest in identifying additional coinhibitory and costimulatory receptors as potential therapeutic targets. After PD-1 and CTLA-4, lymphocyte-activation gene 3 (LAG3) was the third inhibitory receptor to be clinically targeted. In lymphoid lineages, LAG3 is highly expressed in activated T cells, Tregs, and NK cells. In myeloid subsets, LAG3...
is found in plasmacytoid DCs (pDCs), which are critical for mounting antiviral responses, and in TAMs in some cancer-specific conditions, such as B-cell lymphoma. Interestingly, LAG3-null mice have increased numbers of macrophages, granulocytes, and pDCs; thus, the receptor may play a role in regulating hematopoiesis. A population of Tregs coexpressing LAG3 and T-cell-immunoglobulin-and-mucin-domain-containing-molecule-3 (TIM3) suppress the proinflammatory activation of macrophages, suggesting that the blockade of LAG3 could synergize with the indirect effects of anti-PD-1/PD-L1 and/or anti-CTLA-4 therapies, which shift the TAM phenotype toward an antitumor state. Despite the known immunosuppressive role of LAG3 in T cells, research into the regulation of myeloid subsets by LAG3 is still limited.

Like LAG3, T-cell-immunoreceptor-with-Ig-and-ITIM-domains (TIGIT) is a coinhibitory receptor primarily expressed in T cells and NK cells. TIGIT interacts with a variety of nectin-like molecules expressed in APCs and various cancers, leading to impaired T and NK cell proliferation and IFN-γ production. Interestingly, MDSC-mediated suppression of NK cells may be partially TIGIT-dependent as an anti-TIGIT antibody was able to reverse NK cell dysfunction in MDSC coculture experiments. The mechanism underlying this finding may involve decreased Arg1 expression in MDSCs, as has been reported in the context of T cells. As with PD-1-null mice, TIGIT-null mice have reduced tumor growth in various cancer models, and NK cells isolated from these mice demonstrate enhanced IFN-γ secretion when activated by target cells. Further research is needed to evaluate the indirect effects of anti-TIGIT blockade on myeloid subsets.

Unlike TIGIT and LAG3, TIM3 is broadly expressed and directly regulates both innate and adaptive immune subsets. Specifically, TIM3 is found in T cells, NK cells, macrophages, TAMs, DCs, and mast cells. TIM3 agonism reduces inflammatory signaling in both adaptive and innate cell types, and as such, is an attractive target for future immunotherapies. In T cells, TIM3 blockade increases Th1 proliferation and IFN-γ production. Recent findings highlight that NK cell cytotoxicity is also attenuated by TIM3 expression, which may play a role in maternal-fetal tolerance. In macrophages, TIM3 signaling dampens inflammation and limits phagocytic capacity. Additionally, blockade of TIM3 reduces MDSC recruitment and slows tumor growth in a model of HNSCC. Within the DC lineage, TIM3 is primarily expressed by CD8+/CD103+ DCs, which are specialized for priming CD8+ T cell responses through cross-presentation. The outcome of TIM3 signaling in DCs is varied and can have both immunosuppressive and protumorigenic effects. Treating a DC cell line with lipopolysaccharide and the TIM3 ligand galectin-9 significantly increased TNF-α production. However, TIM3 has also been shown to dampen inflammation by interfering with the engagement of DNA-specific TLRs. TIM3 blockade improves antitumor DC-T cell crosstalk by enhancing cross-presentation by CD103+ DCs, and increasing DC expression of CXCL9, a T cell chemoattractant. Interestingly, in a fibrosarcoma model, DCs were required for anti-TIM3-enhanced chemotherapy efficacy. These results suggest that TIM3 may inhibit DC activation, antigen presentation, and DC crosstalk in the setting of chemotherapy-induced immunogenic cell death. Taken together, TIM3’s wide expression in innate and adaptive subsets, and its demonstrated role in the crosstalk between the two, highlight TIM3 as a promising immunotherapeutic target. The emergence of novel coinhibitory receptors will necessitate careful evaluation of which targets synergize with existing checkpoint therapies and broadly enhance antitumor immunity in both innate and adaptive compartments.

CONCLUSIONS

The studies highlighted in this review paint a complex picture of the relationship between checkpoint blockade and innate immunity. Myeloid cells and innate lymphocytes contribute to both checkpoint efficacy and resistance, through both direct and indirect mechanisms. The challenge for researchers and clinicians is to balance the varied, and sometimes opposing, effects of checkpoint therapies, so that they may intelligently employ and combine immunotherapies in the fight against cancer. Shifting the focus of checkpoint blockade from ‘T cell centricism’ to a more holistic view of the complex and interconnected TME may reveal new opportunities to broaden the benefits of checkpoint blockade to the many patients in need of prognosis-altering therapies.

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REFERENCES

1. Leach DR, Krummel MF, Allison JP. Enhancement of antitumor immunity by CTLA-4 blockade. Science 1996;271:1734–6.
2. Wilky BA. Immune checkpoint inhibitors: the linchpins of modern immunotherapy. Immunol Rev 2019;290:6–23.
3. Nishimura H, Nose M, Hiai H, et al. Development of lupus-like autoimmune diseases by disruption of the PD-1 gene encoding an ITIM motif-carrying immunoreceptor. Immunity 1999;11:141–51.
Henick BS, Herbst RS, Goldberg SB. The PD-1 pathway as a therapeutic target to overcome immune escape mechanisms in cancer. 

Expert Opin Ther Targets 2014;11:1–14.

Baumeister SH, Freeman GJ, Darnott G, et al. Coinhibitory pathways in immunotherapy for cancer. 

Ann Rev Immunol 2016;34:539–73.

Krummel MF, Allison JP. CTLA-4 engagement inhibits IL-2 accumulation and cell cycle progression upon activation of resting T cells. 

J Exp Med 1996;183:2533–40.

Tivol EA, Borrelli F, Schweitzer AN, et al. Loss of CTLA-4 leads to massive lymphoproliferation and fatal multiorgan tissue destruction, revealing a critical negative regulatory role of CTLA-4. 

Immunity 1995;3:541–7.

Mark Doherty T, Doherty TM. T-Cell regulation of macrophage function. 

Curr Opin Immunol 1995;7:400–4.

Haie C, Macatonia S, Tripp C, et al. Development of Th1 CD4+ T cells through IL-12 produced by Listeria-induced macrophages. 

Science 1993;260:547–9.

Gubin MM, Esaulova E, Ward JP, et al. High-dimensional analysis delineates myeloid and lymphoid compartment remodeling during successful immune-Checkpoint cancer therapy. 

Cell 2018;175:1443.

Rudd CE. A new perspective in cancer immunotherapy: PD-1 on myeloid cells takes center stage in orchestrating immune checkpoint blockade. 

Science Immunology 2020;5:eaa8128.

Wang F, Li B, Wei Y, et al. Tumor-Derived exosomes induce PD1+ macrophage population in human gastric cancer that promotes disease progression. 

Oncogenecsis 2018;7:41.

Gordon SR, Maute RL, Dulken BW, et al. PD-1 expression by tumour-associated macrophages inhibits phagocytosis and tumour immunity. 

Nature 2017;545:495–9.

Dhupkar P, Gordon N, Stewart J, et al. Anti-PD-1 therapy redirects macrophages from an M2 to an M1 phenotype inducing regression of os lung metastases. 

Cancer Med 2018;7:2654–64.

Strauss L, Mahmoud MAA, Weaver JD. Targeted deletion of PD-1 in myeloid cells induces antitumor immunity 2020/5.

Chen W, Wang J, Jia L, et al. Attenuation of the programmed cell death-1 pathway increases the M1 polarization of macrophages by inducing the IL-21 pathway that drives antitumor immunity. 

Cell death Dis 2016;7:e2115.

Huang X, Venet F, Wang YL, et al. PD-1 expression by macrophages plays a pathologic role in altering microbial clearance and the innate inflammatory response to sepsis. 

Proc Natl Acad Sci U S A 2009;106:6303–8.

Takakomo H, Fujioka K, Miyashita A, et al. Combined blockade of IL6 and PD-1/PD-L1 signaling abrogates mutual regulation of their immunosuppressive effects in the tumor microenvironment. 

Cancer Res 2018;78:5011–22.

Kaplanov I, Carmi Y, Kaplanov I, et al. Programmed cell death ligand 1 (PD-L1) on dendritic cells attenuates the polarization of macrophages/microglia to the M2 phenotype after induced macrophage/microglia accumulation and cell cycle progression upon activation of resting T cells. 

Sci Rep 2016;6:19434.

Diskin B, Adam S, Cassini MF, et al. PD-L1 engagement on T cells promotes self-tolerance and suppression of neighboring macrophages and effector T cells in cancer 2020;21:442–54.

Nguyen LT, Ohashi PS. Clinical blockade of PD1 and LAG3 potentiates mechanisms of action. 

Nat Rev Immunol 2015;15:45–56.

Kleinovink JW, Marijt KA, Schoonderwoerd MJA, et al. PD-L1 expression on malignant cells is no prerequisite for checkpoint therapy. 

Oncoimmunology 2017;6:e1294299.

Taube JM, Klein A, Brahter JR, et al. Association of PD-1, PD-1 ligands, and other members of the tumor immune microenvironment with response to Anti-PD-1 therapy. 

Clinical Cancer Research 2014;20:5064–74.

Liu X, et al. J Immunother Cancer. 2021;9:8001460. doi:10.1136/jitc-2020-001460
Steele CW, Karim SA, Leach JGD, et al. Cxcr2 inhibition profoundly suppresses metastases and augments immunotherapy in pancreatic ductal adenocarcinoma. Cancer Cell 2016;29:832–45.

Nam S, Lee A, Lim J, et al. Analysis of the expression and regulation of PD-1 protein on the surface of myeloid-derived suppressor cells (MDSCs). Biomed Ther 2019;27:63–70.

Lu C, Redd PS, Lee JR, et al. The expression profiles and regulation of PD-L1 in tumor-infiltrated myeloid-derived suppressor cells. Oncoimmunology 2016;5:e124713S.

Görgün G, Saad D, NKC, Cowens KB, et al. Lenalidomide enhances immune checkpoint Blockade-Induced immune response in multiple myeloma. Clinical Cancer Research 2015;21:4607–18.

GT Y, LL B, Huang CF, et al. Pd-1 blockade attenuates immunosuppressive myeloid cells due to inhibition of CD47/ SIRPAβ interaction in HPV negative head and neck squamous cell carcinoma. Oncotarget 2015;6:42067–80.

Noman MZ, Desantis G, Janji B, et al. Pd-L1 is a novel direct target of HIF-1α, and its blockade under hypoxia enhanced MDSC-mediated T cell activation. Journal of Experimental Medicine 2014;211:781–90.

Ballbach M, Dannert A, Singh A, et al. Expression of checkpoint molecules on myeloid-derived suppressor cells. Immunol Lett 2017;192:1–6.

Benson GM, Bajjan CE, Mishra A, et al. The PD-1/PD-L1 axis modulates the natural killer cell versus multiple myeloma effect: a therapeutic target for CT-011, a novel monoclonal anti-PD-1 antibody. Blood 2010;116:2286–94.

Tumino N, Martini S, Munari E, et al. Presence of innate lymphoid cells in pleural fluid of patients with pulmonary metastatic tumors: functional analysis and expression of PD-1 receptor 2019;145:1660–8.

Damele L, Ottonello S, Mingari MC, et al. Targeted Therapies: Friends or Foes for Patient’s NK Cell Immunotherapies. J Immunother Cancer 2020;8:12774.

Vivier E, Armitage K, Munna M, et al. Innate lymphoid cells: 10 years on. Cell 2018;174:1054–66.

Shirnfin N, Raulet DH, Ardlino M. NK cell self tolerance, responsiveness and missing self recognition. Semin Immunol 2014;26:138–44.

Judyce SJ, Duai C, Aguilar EG, et al. Minimal PD-1 expression in mouse and human NK cells under diverse conditions. J Clin Invest 2020;130:3051–68.

Pesce S, Greppi M, Tabellini G, et al. Identification of a subset of human natural killer cells expressing high levels of programmed death 1: A phenotypic and functional characterization. Journal of Allergy and Clinical Immunology 2017;139:335–46.

Hsu J, Hodkins JJ, Marathe M, et al. Contribution of NK cells to immunotherapy mediated by PD-1/PD-L1 blockade. Journal of Clinical Investigation 2018;128:6521–6.

Wiesmayr S, Webber SA, Macedo C, et al. Decreased Nkp46 and Nkg2D and elevated PD-1 are associated with altered NK-cell function in pediatric transplant patients with PTLD. Eur J Immunol 2012;42:541–50.

Dong W, Wu X, Ma S, et al. The mechanism of anti-PD-1 antibody efficacy against PD-L1-Negative tumors identifies NK cells expressing PD-L1 as a cytolytic effector 2019;9:1422–37.

Mariotti FR, Quattrini L, Munari E, et al. Innate lymphoid cells: expression of PD-1 and other checkpoints in normal and pathological conditions. Front Immunol 2019;10:910.

Yu Y, Tsang J, Wang C, et al. Single-Cell RNA-seq identifies a PD-1hi ILC progenitor and defines its development pathway. Nature 2016;539:102–6.

Moral JA, Leung J, Rojas LA, et al. Itc2S amplifies PD-1 blockade by activating tissue-specific cancer immunity. Nature 2020;578:130–5.

Vacca P, Pesce S, Greppi M, et al. Pd-1 is expressed by and regulates human group 3 innate lymphoid cells in human decidua. Mucosal Immunol 2019;12:624–31.