Case Report

A case of non-immune hydrops fetalis with maternal mirror syndrome diagnosed by trio-based exome sequencing: An autopsy case report and literature review

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https://doi.org/10.1016/j.ymgmr.2022.100925
Received 29 July 2022; Received in revised form 6 October 2022; Accepted 8 October 2022
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1. Introduction

Non-immune hydrops fetalis (NIHF) affects approximately 1 in 2000–3000 pregnancies and is a high risk for preterm birth and neonatal complications or death [1]. Standard genetic testing using G-banded karyotyping or chromosomal microarray analysis detects the cause of only 25% of fetuses with NIHF [1]. Therefore, the cause of most fetal NIHF remains unknown. Not only for neonatal complications but also severe NIHF is known to be associated with mirror syndrome, a rare condition characterized by the combination of placental edema, hydrops fetalis, and maternal critical edema [2–4]. It is reported that maternal morbidity increases and fetal mortality are as high as 67% in mirror syndrome [4], but the pathogenesis and effective therapy for mirror syndrome remain unknown. The pregnancy should be terminated to resolve the maternal condition if the fetal hydrops is irreversible [3].

Recent advances in exome sequencing technology have improved diagnostic efficiency [5], especially in genetic disorders. Some genetic disorders, including RASopathies, were possible causes of NIHF [1,6]. One of the RASopathies, Noonan syndrome (NS; OMIM #163950), is an autosomal dominant congenital genetic disorder, and its prevalence is estimated to be 1 in 1000–2500 births [7]. It is a clinically and genetically heterogeneous condition characterized by distinctive facial features, cardiac defects, and other comorbidities [6–17]. Prenatally, NS indicates an increased risk for cystic hygroma or hydrops fetalis [7,9,10]. The diversity of phenotypical features is associated with various genotypes [6]. Several genes are associated with NS, and the genetic variants are identified in approximately 60–70% of patients with NS [7,16]. However, genetic variants relevant for severe fetal or maternal conditions in NIHF remained undetermined.

Here, we report a case of severe fetal hydrops and maternal mirror syndrome.
syndrome. The patient was diagnosed with NS and had a de novo RIT1 missense variant (NM_006912.6: c.246 T > G [p.F82L]; ClinVar ID: 181522) based on trio-based exome sequencing.

2. Case report

A 35-year-old G2P0 woman conceived after in vitro fertilization and embryo transfer; her partner was 34 years old. The couple was healthy and non-consanguineous. The singleton fetus presented polycystic

![Image of fetal ultrasound findings at 24+3 weeks of gestation.](image1)

The fetus had significant generalized skin edema, cystic hygroma, bilateral pleural effusion, and large amounts of ascites.

![Image of maternal blood test results at 24+3 weeks of gestation.](image2)

The left axis represents the level of total protein (TP) and albumin (Alb) [mg/dL], and the right axis represents the level of hematocrit (Hct) [%], respectively.

![Image of maternal chest X-ray at 24+3 weeks of gestation.](image3)

The enlarged cardiothoracic ratio and pulmonary congestion were significant.

![Image of placenta.](image4)

The left panel shows a macroscopic view of the placenta, showing that the placenta is easily collapsed. The upper right panel is a magnified image of the placenta, showing that the placenta is hydropic. The lower right panel shows the ultrasound finding of thickened placenta with a transverse diameter of 66.5 mm at 24+2 weeks of gestation.

![Image of newborns.](image5)

The female neonate had marked edema.

![Image of pathological findings of the autopsy.](image6)

Photomicrograph revealed the cyst wall was composed of fibrous connective tissue coated with one layer of the endothelium (D2–40, a lymphatic endothelial marker, arrows).

Fig. 1. Clinical findings.
A-C. Fetal ultrasound findings at 24+3 weeks of gestation.
The fetus had significant generalized skin edema, cystic hygroma, bilateral pleural effusion, and large amounts of ascites.
D. Time course of changes of the maternal blood test.
The left axis represents the level of total protein (TP) and albumin (Alb) [mg/dL], and the right axis represents the level of hematocrit (Hct) [%], respectively.
E. The results of the maternal blood test at 24+3 weeks of gestation.
WBC, white blood cell; Hb, hemoglobin; Hct, hematocrit; Plt, platelet count; TP, total protein; Alb, albumin; BUN, blood urea nitrogen; Cr, creatinine; UA, uric acid; AST, aspartate transaminase; ALT, alanine transaminase; LDH, lactate dehydrogenase; Na, sodium; Cl, chloride; K, potassium; BNP, brain natriuretic peptide.
F. Maternal chest X-ray at 24+3 weeks of gestation.
The enlarged cardiothoracic ratio and pulmonary congestion were significant.
G. Placenta
The left panel shows a macroscopic view of the placenta, showing that the placenta is easily collapsed. The upper right panel is a magnified image of the placenta, showing that the placenta is hydropic. The lower right panel shows the ultrasound finding of thickened placenta with a transverse diameter of 66.5 mm at 24+2 weeks of gestation.
H. Appearance of newborns.
The female neonate had marked edema.
I. Pathological findings of the autopsy (×20).
Photomicrograph revealed the cyst wall was composed of fibrous connective tissue coated with one layer of the endothelium (D2–40, a lymphatic endothelial marker, arrows).
hygroma, pleural effusion, and ascites at 16 weeks of gestation. Chromosome karyotype analysis showed a normal female karyotype (46, XX), and a fetal echocardiogram detected no anomaly. The cystic hygroma and hydrops worsened at 24 weeks of gestation, characterized by skin edema, bilateral pleural effusion, and large amounts of ascites (Fig. 1A-C). The placenta was thickened with a transverse diameter of 6.7 cm (Fig. 1G). The mother also had edema and palpitations that gradually worsened after 22 weeks of gestation, while her blood pressure was within the normal range (122/80 mmHg at 24-25 weeks of gestation). At 24-25 weeks of gestation, hemoglobin (Hb), total protein, and albumin decreased to 22.2%, 4.5 g/dL, and 2.3 g/dL, respectively (Fig. D-E). She had 1+ proteinuria on dipstick analysis. Chest X-ray was significant for the enlarged cardiothoracic ratio of 58.8% and pulmonary congestion with elevated serum brain natriuretic peptide (BNP) level of up to 102.3 pg/mL (Fig. 1E-F). From these findings, we diagnosed her with mirror syndrome and determined that further continuation of her pregnancy would be dangerous for her. We discussed with her and her partner and decided to perform a Cesarean section at 24-25 weeks of gestation due to her volume overload and oxygen demand.

The female newborn was found to have extreme hydrops, and her birth weight and height were 2356 g (>99th percentile) and 31.8 cm (73rd percentile), respectively. The umbilical artery blood pH was 7.40, and 1- and 5-min Apgar scores were 1 and 1, respectively. Neonatal resuscitation was immediately initiated; however, the baby died shortly after birth. The placenta was grossly hydropic and easily collapsed (Fig. 1G). The mother improved steadily after childbirth and was discharged without significant complications.

The autopsy of the newborn showed a large multifocal subcutaneous cyst with a diameter of >10 cm in the posterior neck. Her head and face were deformed by severe skin edema (Fig. 1H). The multifocal subcutaneous cyst contained approximately 70 mL of pale, bloody fluid, and the cyst wall was composed of fibrous connective tissue coated with one layer of endothelium. The endothelium was positive for D2-40 (arrows, Fig. 1I), suggesting fetal cystic lymphangioma. In the skin, dilation of lymphatic vessels was also observed from the dermis to the subcutaneous tissue, suggesting lymphatic congestion. The pleural effusion and ascites were 150 mL in total. Right and left lungs were 6 and 4 g, respectively, suggesting severe pulmonary hypoplasia due to compression of pleural effusion and ascites (17 g on average). The alveolar walls were lined with type II alveolar epithelium, but the air content in the alveolar spaces was poor. No other findings suggest apparent developmental abnormalities or dysfunction, including cardiac defects.

Trio-based exome sequencing showed a de novo heterozygous missense RIT1 variant at position 246 (c.246 T > G [p.F82L]) (Fig. 2A). Thus, this patient was diagnosed with Noonan syndrome.

3. Discussion

Here, we report a case of severe NS with hydrops and maternal mirror syndrome. According to the autopsy findings, the present neonate died mainly from circulatory failure associated with fetal cystic lymphangioma and fetal hydrops, combined with pleural effusion and respiratory failure due to the immaturity and severe hypoplasia of the lungs. In addition, her mother faced complications due to mirror syndrome. The exome sequence revealed a de novo heterozygous missense variant of RIT1 (c.246 T > G [p.F82L]), which suggested that NS was the cause of NIHF.

RIT1 (OMIM # 609591) variants, known causative variants of NS are responsible for approximately 4-5% of all patients with NS [7,9]. RIT1 has approximately 50% sequence identity to RAS [11] and activates multiple downstream signaling cascades [11,17]. Previously reported RIT1 variants were all missense variants, and most were gain-of-function variants [7,9,11,17]. The proteins encoded by the mutated RIT1 gene cause hyperactivation of RAS/MAPK or transactivation of the ELK pathways, participating in the pathogenesis of NS [11]. RIT1 is expressed ubiquitously in embryonic and adult tissues, and NS patients with RIT1 variants have a greater incidence of prenatal lymphatic malfunctions, one of the causes of cystic hygroma and hydrops fetalis, and hypertrophic cardiomyopathy (HCM) than in the overall population of patients with NS [6,9].

A total of 20 variants of RIT1 were identified in patients with NS and were located mainly in the switch II region (SW2) [6-20] (Fig. 2B), corresponding to RAS [6-8,15]. Table 1 shows the clinical phenotype of RIT1-associated NS, including the present and previously reported cases. Cystic hygroma has been detected in 7 patients with RIT1 variants, including the present case. Three of them had a p.F82L variant, and 2 had a p.M90I or p.M90V variant, which was the close site in the SW2 region. The present case was the second case of mirror syndrome with RIT1-associated NS. The previous one had the de novo p.M90V variant, which was also the close site in the SW2 region [20]. Our patient also had a de novo SW2 region variant, suggesting fetal factors might affect the progression to maternal mirror syndrome, while the evidence is limited. It is reported that fetal hydrops or cystic hygroma could affect the placental-derived factor production [21], possibly involved in the pathogenesis of mirror syndrome [22]. Further case accumulation is needed to investigate the association between the RIT1 domain and cystic hygroma or mirror syndrome. Although the pathologic association remains unclear, predicting mirror syndrome in earlier gestation would be very helpful for the mother and her partner.

In conclusion, exome sequencing was valuable in NIHF, where chromosome testing could not identify the cause. The present and previous cases speculated that exome sequencing, which can identify the variant type in detail, might help predict the phenotype and severity of NIHF and its association with mirror syndrome. However, further accumulation of cases is needed to confirm this speculation.
### Table 1
Clinical phenotype of RIT1-associated NS.

| RIT1 Mutation | Ref No. | Age | Sex | Inheritance | Mirror Syndrome | Prenatal findings | Cardiac anomalies |
|---------------|---------|-----|-----|-------------|-----------------|------------------|------------------|
| p.K23N        | 13      | 10m | M   | ND          |                 | CH   | NE | Others | HCM | ASD | VSD | Others |
| p.G31R        | 10      | 8y  | F   | Inherited  | ND              | –    | –  | –     | –   | –   | –   | –     |
| p.G31R        | 10      | 47y | F   | Not known  | ND              | ND   | ND | ND    | –   | –   | –   | –     |
| SW1           | p.S35T  | 10  | 1y  | De novo    | ND              | –    | –  | PH    | –   | –   | –   | PS    |
| p.S35T        | 10      | 5y  | F   | Not known  | ND              | –    | –  | PH    | +   | –   | –   | –     |
| p.S35T        | 10      | 11y | M   | Not known  | ND              | –    | –  | –     | –   | –   | –   | –     |
|               | 6       | 1y  | F   | ND          | ND              | –    | +  | –     | –   | +   | –   | PS    |
|               | 6       | 3y  | M   | ND          | ND              | –    | –  | +     | –   | +   | –   | PS, AS |
|               | 8       | 2y  | M   | ND          | ND              | –    | –  | –     | –   | –   | –   | PS    |
|               | 19      | 3m  | ND  | De novo    | ND              | –    | –  | PH    | +   | –   | –   | PS    |
|               | 15      | FD  | ND  | ND          | ND              | –    | +  | PE    | ND  | ND  | ND  | ND    |
| p.A57G        | 10      | 11y | F   | Not known  | ND              | ND   | ND | ND    | –   | –   | –   | PS    |
| p.A57G        | 6       | 15d | F   | ND          | ND              | –    | +  | PE    | +   | –   | –   | PDA   |
| p.A57G        | 6       | 5y  | F   | ND          | ND              | –    | –  | +     | –   | +   | –   | PDA   |
| p.A57G        | 6       | 11m | F   | ND          | ND              | –    | –  | PE    | +   | +   | –   | PDA   |
|               | 8       | 27y | F   | ND          | ND              | ND   | ND | ND    | +   | –   | –   | PS    |
| p.A57G        | 8       | 28y | M   | ND          | ND              | –    | –  | –     | –   | –   | –   | PS    |
| p.A57G        | 12      | 2y  | F   | ND          | ND              | ND   | ND | ND    | –   | –   | –   | PS    |
| p.A57G        | 11      | 15y | F   | ND          | ND              | –    | +  | +     | +   | +   | –   | PS    |
| p.A57G        | 18      | ND  | M   | ND          | ND              | ND   | ND | ND    | +   | –   | –   | PS    |
| p.A77T        | 10      | 3y  | F   | Not known  | ND              | –    | –  | PH    | –   | +   | –   | PS    |
| p.A77T        | 10      | 21y | M   | Not known  | ND              | ND   | ND | ND    | –   | –   | –   | PS, MVA |
| p.A77T        | 6       | 4y  | F   | ND          | ND              | –    | –  | PE    | –   | +   | +   | +     |
| p.A77T        | 6       | 1y  | F   | ND          | ND              | –    | –  | PH    | +   | –   | –   | PS    |
| p.A77T        | 19      | ND  | M   | ND          | ND              | ND   | ND | ND    | –   | –   | –   | PS    |
| p.A77T        | 12      | 22y | F   | ND          | ND              | ND   | ND | ND    | –   | –   | –   | PS    |
| p.A77S        | 6       | 3y  | F   | ND          | ND              | –    | –  | PE    | PH  | +   | +   | –     |
| p.A77S        | 14      | 14y | M   | ND          | ND              | –    | –  | –     | –   | –   | –   | –     |
| p.E81G        | 9       | ND  | ND  | ND          | ND              | ND   | ND | ND    | –   | –   | –   | +     |
| p.F82V        | 10      | 13y | F   | De novo    | ND              | –    | –  | PE    | PH  | –   | –   | –     |
| p.F82V        | 12      | 6y  | F   | ND          | ND              | –    | –  | +     | –   | +   | +   | PDA   |
| p.F82V        | 7       | 10y | F   | ND          | ND              | ND   | ND | ND    | –   | –   | –   | PDA   |
| p.F82S        | 10      | 29y | M   | Not known  | ND              | –    | –  | –     | –   | –   | –   | –     |
| Present case  | 0d      | F   | De novo | +   | +   | PE    | –    | –   | –     |
|               | 10      | 5y  | F   | De novo    | ND              | –    | +  | PE    | +   | –   | –   | MVA   |
|               | 10      | 38y | F   | Not known  | ND              | ND   | ND | ND    | –   | –   | –   | PS, MVA |
|               | 10      | 5y  | F   | Not known  | ND              | –    | –  | PH    | –   | –   | –   | PS    |
|               | 10      | 11y | M   | Not known  | ND              | –    | –  | PH    | +   | –   | –   | PS    |
|               | 10      | 13y | M   | Not known  | ND              | –    | –  | –     | –   | –   | –   | PS    |
|               | 10      | 1y  | F   | Not known  | ND              | –    | +  | PE    | +   | +   | –   | PDA   |
|               | 6       | 4y  | M   | ND          | ND              | –    | +  | PE    | –   | –   | –   | –     |
|               | 6       | 4y  | M   | ND          | ND              | –    | –  | PE    | +   | –   | –   | –     |
|               | 8       | 6d  | M   | ND          | ND              | +    | –  | –     | ND  | ND  | ND  | ND    |
|               | 18      | 10d | ND  | De novo    | ND              | +    | –  | –     | +   | –   | –   | PS    |
|               | 8       | 19y | M   | ND          | ND              | +    | –  | –     | +   | –   | –   | PS    |
| p.T83P        | 10      | 16y | F   | Not known  | ND              | –    | –  | –     | –   | –   | –   | –     |
| p.D87H        | 18      | ND  | M   | Not known  | ND              | –    | –  | –     | +   | –   | –   | PS    |
| p.Y89H        | 9       | ND  | ND  | Not known  | ND              | ND   | ND | ND    | +   | –   | –   | PS    |
| p.M90I        | 10      | 3y  | F   | Not known  | ND              | –    | –  | –     | –   | –   | –   | PS, MVA |
| p.M90I        | 6       | 5y  | M   | ND          | ND              | –    | –  | PH    | –   | –   | –   | PS    |
| p.M90I        | 7       | 5y  | F   | ND          | ND              | +    | –  | –     | +   | –   | –   | PS    |
| p.M90I        | 11      | 2y  | M   | ND          | ND              | –    | –  | PH    | +   | +   | +   | PDA   |
| p.M90V        | 20      | FD  | M   | De novo    | ND              | +    | –  | PE    | ND  | ND  | ND  | ND    |
| p.M90V        | 15      | FD  | ND  | ND          | ND              | +    | –  | PE    | ND  | ND  | ND  | ND    |
| p.G95A        | 10      | 8y  | M   | Inherited  | ND              | –    | –  | –     | –   | –   | –   | PS    |
| p.G95A        | 10      | 42y | F   | Not known  | ND              | –    | –  | –     | +   | –   | –   | –     |

(continued on next page)
## Table 1 (continued)

| RIT1 Mutation Region | Ref No. | Age | Sex | Inheritance | Mirror Syndrome | Prenatal findings | Cardiac anomalies |
|----------------------|---------|-----|-----|-------------|-----------------|-------------------|------------------|
|                      |         |     |     |             |                 | CH | NE | Others | HCM | ASD | VSD | Others |
| 0288                 | –       | 10  | 14y | F           | Inherited       | ND | –  | –      | –   | –   | –   | PS     |
| 10                   | 24y     | 10  | M   | Not known   | ND              | –  | +  | PH     | +   | –   | –   | –      |
| 10                   | 15y     | 10  | F   | Not known   | ND              | –  | +  | –      | –   | –   | –   | –      |
| 10                   | 11y     | 10  | M   | Not known   | ND              | –  | +  | –      | –   | –   | –   | –      |
| 6                    | 15y     | 6   | M   | ND          | ND              | –  | +  | –      | –   | –   | –   | –      |
| 6                    | 3mo     | 6   | F   | ND          | ND              | –  | –  | –      | –   | –   | –   | –      |
| 8                    | 16y     | 8   | F   | ND          | ND              | –  | –  | PH     | –   | –   | –   | –      |
| 8                    | 27y     | 8   | M   | ND          | ND              | –  | –  | –      | +   | –   | –   | –      |
| 12                   | 46y     | 12  | F   | ND          | ND              | –  | –  | ND     | –   | –   | –   | –      |
| 12                   | 18y     | 12  | M   | ND          | ND              | –  | +  | –      | –   | –   | –   | –      |
| 7                    | 17y     | 7   | F   | ND          | ND              | –  | –  | ND     | –   | –   | –   | –      |
| 7                    | 13y     | 7   | F   | ND          | ND              | –  | –  | ND     | –   | –   | –   | –      |
| 16                   | 11y     | 16  | F   | De novo     | ND              | –  | –  | ND     | –   | –   | –   | –      |

Bold letters indicate the present case. ND, no data; FD, intrauterine fetal demise; PH, polyhydramnios; CH, cystic hygroma; NE, fetal nuchal edema; PE, fetal preural effusion; FH, fetal hydrops; PS, pulmonary stenosis; HCM, hypertrophic cardiomyopathy; ASD, atrial septal defect; AS, aortial stenosis; VSD, ventricula septal defect; PDA, patent ductus arteriosus; MVA, mytral valve anomalies; LE, leg edema; CT cataract; ASC, ascites.

## Funding

This work was supported by the special coordination funds for Rare and Intractable Diseases Project of Japan from the Japan Agency for Medical Research and Development (T.O.).

## Author contributions

Conceptualization and planning: Sho Tano, Tomomi Kotani, and Tomoo Ogi; clinical data collection and interpretation: Masato Yoshihara, Seiko Matsuo, Takafumi Ushida, Kenji Imai, and Miharu Ito; bioinformatics analysis: Noriyuki Nakamura, Yasuyoshi Oka, Emi Sato, and Shin Hayashi; supervision: Tomoo Ogi and Hiroaki Kajiyama.

All authors discussed, read, and approved the manuscript in its final form.

## Ethics statement

This study was approved by the Ethical Committee for the Study of Human Gene Analysis at Nagoya University Graduate School of Medicine (approval number: 2018–0288). The patient’s parents gave written consent for the publication of the clinical information, including photographs.

## Declaration of Competing Interest

The authors have no conflicts of interest to declare.

## Data availability

Data will be made available on request.

## Acknowledgments

The authors would like to thank the parents of our patients for their willingness to share data to improve the diagnosis of rare disorders.

## References

[1] F.G. Sileo, et al., Non-immune fetal hydrops: etiology and outcome according to gestational age at diagnosis, Ultrasound Obstet. Gynecol. 56 (3) (2020) 416–421.

[4] A. Gledbuth, et al., Preeclampsia due to fetal non-immune hydrops: mirror syndrome and review of literature, Hypertens Pregnancy 30 (3) (2011) 322–330.

[5] S. Allaraki, et al., Characteristics and management of mirror syndrome: a systematic review (1956-2016), J. Perinat. Med. 45 (9) (2017) 1013–1021.

[8] D.R. Bertola, et al., Further evidence of the importance of RIT1 in Noonan syndrome, Am. J. Med. Genet. A 170 (7) (2016) 1874–1880.

[9] M. Nemcikova, et al., A novel heterozygous RIT1 mutation in a patient with Noonan syndrome, Proc. Natl. Acad. Sci. U. S. A. 111 (31) (2014) 11473–11478.

[10] K. Kouz, et al., Genotype and phenotype in patients with Noonan syndrome and a RIT1 mutation, Genet Med. 18 (12) (2016) 1326–1324.

[11] M. Koenighofer, et al., Mutations in RIT1 cause Noonan syndrome - additional functional evidence and expanding the clinical phenotype, Clin. Genet. 89 (3) (2016) 359–366.

[12] P.C. Chen, et al., Next-generation sequencing identifies rare variants associated with Noonan syndrome, Proc. Natl. Acad. Sci. U. S. A. 111 (2) (2014) 222–229.

[14] F. Ramond, et al., Expanding the cardiac spectrum of Noonan syndrome with RIT1 mutation, Genet Med. 18 (12) (2016) 1326–1324.

[16] A.L. Koh, et al., The spectrum of genetic variants and phenotypic features of southeast Asian patients with Noonan syndrome, Mol. Genet Genomic Med. 7 (4) (2019), e00581.

[17] X. Li, et al., Molecular and phenotypic spectrum of Noonan syndrome in Chinese patients, Clin. Genet. 96 (4) (2019) 290–299.

[18] G.K.C. Leung, et al., Integrating functional analysis in the next-generation sequencing diagnostic pipeline of RASopathies, Sci. Rep. 8 (1) (2018) 2421.

[19] M. Zhao, et al., Spectrum of mutations and genotype-phenotype analysis in Noonan syndrome patients, Clin. Genet. 96 (4) (2019) 290–299.

[20] G. Andelfinger, et al., Hypertrophic cardiomyopathy in Noonan syndrome treated by mek-inhibition, J. Am. Coll. Cardiol. 73 (17) (2019) 2237–2246.

[21] D. Milosavljevic, et al., Two cases of RIT1 associated Noonan syndrome: further delineation of the clinical phenotype and review of the literature, Eur. J. Pediatr. 175 (4) (2016) 587–592.

[22] G. Hirata, et al., Clinical characteristics of mirror syndrome: a systematic review (1956-2016), J. Perinat. Med. 45 (9) (2017) 1013–1021.

[23] F. Alvarez-Nava, et al., Elevated second-trimester maternal serum beta-human chorionic gonadotropin and amniotic fluid alpha-fetoprotein as indicators of adverse obstetric outcomes in fetal turner syndrome, J. Obstet. Gynaecol. Res. 41 (12) (2015) 1891–1898.