BOLDINE ATTENUATES CANCER CELL GROWTH IN AN EXPERIMENTAL MODEL OF GLIOMA IN VIVO

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**Abstract**  
Plants are important sources of biologically active natural products which differ in terms of structure and biological properties. Some of the isolated plant constituents such as flavonoids, alkaloids, and others are responsible for many biological activities, such as antiinflammatory activity, analgesic activity, antiallergenic activity, among others. One of the biological activities of plant compounds that has attracted great interest is the ability to exert anticancer activity in different types of cancer. Boldine, (S)-2,9-dihydroxy-1,10-dimethoxy-aporphine, is an alkaloid that occurs abundantly in the leaves and bark of *Peumus boldus* Mol. a widely distributed native tree of Chile [1,2]. Boldo infusions have been traditionally employed in folk medicine and used as a medicinal plant for the treatment of digestive and hepatobiliary disorders [2-4]. Furthermore, it has been recognized as an herbal remedy in several pharmacopeias in South America and Europe [2]. Chemically pure boldine has been shown to exhibit, amongst others, immuno-modulating [5], smooth muscle relaxing [6], antipyretic and antiinflammary [7,8] and neuroleptic-like [9] properties.

However, in recent years, boldine has attracted more attention in relation to its potent antioxidative and cytoprotective properties [10-12]. Low micromolar concentrations of boldine have been shown to prevent both enzymatic and non-enzymatic mediated damage to biological systems. In vitro, boldine inhibits the free-radical-mediated initiation and propagation of the peroxidative damage induced to various membrane types such as liver homogenates, hepatic microsomes and erythrocytes, and it blocks the free radical-dependent lysis of red blood cells and intact hepatocytes [10,11,13]. These boldine cytoprotective actions are associated with its potent antioxidative activity, once this molecule acts as an efficient hydroxyl radical scavenger in biological systems [11]. Apart from its antioxidative activity, boldine has also been demonstrated to have anticancer properties against bladder carcinoma and glioma cancer cells [14,15].

Malignant gliomas are the most common primary tumors in the human brain. Current treatment includes surgical resection, radiation therapy, and chemotherapy. However, these neoplasms are extremely resistant and frequent tumor recurrence results in poor prognosis with a mean survival time of 12–15 months for grade IV glioma [16]. Exploring new adjuvant therapies are, therefore, necessary for improving the outcome of glioma treatment.

In our early study, we demonstrated the ability of boldine in inhibit the in vitro growth of malignant brain cells (U-138MG and U-87MG human glioma, and C6 rat glioma). With regard to in vivo approaches, it is not know until now whether boldine is safe to use and whether it is really effective in treating glioma grown in an orthotopic microenvironment. Once this alkaloid already has demonstrated anticancer activity in vitro, we examine the effect of this compound in an orthotopic and immunocompetent rat model of glioma that faithfully mirrors the hallmarks of human gliomas and evaluate the in vivo toxicity of boldine.

**Materials and Methods**  
**Chemicals**  
Cell culture media and Fetal Bovine Serum (FBS) were obtained from Gibco-Invitrogen (Grand Island, NY, USA). Boldine and Propidium iodide (PI) were obtained from Sigma Chemical Co (St. Louis, MO, USA). All other reagents were purchased from Sigma Chemical Co. (St. Louis, MO, USA) or Merck (Darmstadt, Germany). All other chemicals and solvents used were of analytical or pharmaceutical grade.

**Keywords:** Boldine; Toxicity; Glioma; Antitumor

**Introduction**  
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Animal maintenance

Healthy male Wistar rats (180–240 g) were obtained from in-house breeding colonies at the "Departamento de Bioquímica", “Universidade Federal do Rio Grande do Sul” (UFRGS - Porto Alegre, Brazil). Animals were housed in cages under optimum light conditions (12:12 h light–dark cycle), temperature (22 ± 1°C), and humidity (50 to 60%), with food and water provided ad libitum. All procedures used in the present study followed the "Principles of Laboratory Animal Care" from NIH publication No. 85-23 and were approved by the local Ethics Committee on the Use of Animals. All efforts were made to minimize the number of animals and their suffering.

Toxicity evaluation

Healthy animals were separated into three groups (4 animals per group) as follows: untreated (control group); treated with drug-vehicle (50% ethanol/water); treated with 50 mg/kg/day of boldine solubilized in vehicle. These animals were treated intraperitoneally for 14 days and weighed every day. After that, animals were euthanized for toxicological experiments evaluation. Blood samples were collected by cardiac puncture. Blood smears were prepared from freshly blood, stained with May-Grünwald-Giemsa Staining and then a manual White Blood Cells (WBC) differential was performed (100 WBCs were found, counted, and categorized according to type). For enzymatic assays (alkaline phosphatase, gamma-glutamyl transpherase, alanine aminotransferase (ALT), aspartate aminotransferase (AST), blood samples were centrifuged at 2,500 x g at 4°C for 10 min. The blood serum was collected and frozen at -70°C until the assays were performed. The enzymes activities, which were used as markers of hepatic tissue toxicity, were measured according to the recommendations of the kit supplier (Labtest, MG, Brazil). Brain, liver, stomach, kidney, heart and lung were removed, weighted, sectioned and fixed with 10% paraformaldehyde for posterior pathological analysis. Paraffin embedded sections were stained with hematoxylin and eosin (H&E) analyzed under a microscope (NikonEclipse TE3000) and analyzed using Image Tool SoftwareTM. The total tumor volume (mm³) was computed by the multiplication of the slice sections and by summing the segmented areas [18].

Immunohistochemical staining

Paraffin embedded, 5 μM formalin fixed tissue sections were mounted on microscope slides. Tissue sections were then dried overnight at 60°C, dewaxed in xylene and rehydrated with distilled water. Endogenous peroxidase was inhibited with 5% H2O2 in methanol for 15 min. Incubation with the following antibodies was performed overnight at 4°C temperature: anti-Ki67 (1:200) (Dako, USA) and anti-VEGF (1:30) (Dako, USA), followed by incubation with secondary antibody and Streptavidin-Avidin-Biotin (Kit Lsb, Dako, USA). The peroxidase reaction was performed using 3,3′diaminobenzidine tetrahydrochloride (DAB), according to the manufacturer's specifications. Finally, sections were counterstained with Harris hematoxylin. Glioma cell proliferation was assessed by counting the percentage of Ki67 positive glioma cell nuclei in five independent high-magnification (×400) fields per animal. Sections of rat spleen were used as positive controls.

Statistical analysis

Data are expressed as means ± SD. The statistical significance among groups was assessed by one-way Analysis of Variance (ANOVA) followed by Tukey's test. Differences between mean values were considered significant when p<0.05.

Results

Macroscopic and histopathological analysis of healthy animals

The treatment with 50 mg/Kg/day i.p. of boldine or vehicle for 14 days did not cause mortality of healthy animals. Moreover, In the period of 14 days of treatment, animals of control group and vehicle group did not improve significantly their body weights, and the treatment with boldine did not cause significant loose of body weight in rats, neither improvement (Figure 1A). Macroscopic investigation of brain, liver, kidney, lungs and hearts did not reveal any disturbance between groups. Organs weights did not present any alteration (Figure 1B). Microscopic investigation of the same organs by H&E analysis demonstrated absence of tissue toxicity in boldine-treated rats. Representative histologies of the tissues of control and boldine-treated rats are shown in Figure 2A. Macroscopic investigation of stomach and intestine to evaluate a possible gastrointestinal damage reveals absence of hemorrhage points or ulcer in both groups (Representative pieces of intestine are seen in Figure 2B).

Treatment of animals

After ten days of glioma implantation, the animals were randomly separated into three groups as follows: (1) untreated (control group–6 animals); (2) treated with drug-vehicle (50% ethanol/water–6 animals); (3) treated with 50 mg/Kg/day of boldine solubilized in vehicle (8 animals). Drug and vehicle were administered intraperitoneally (i.p.) to the animals for 10 consecutive days. At the end of the treatment, the rats were decapitated and the entire brain was removed, sectioned and fixed with 10% paraformaldehyde.

Pathological analysis and tumor volume quantification

Hematoxylin and eosin (H&E) sections (paraffin embedded) of each tumor were analyzed by a pathologist, blinded for the experimental data. For tumor size quantification, images were captured using a digital camera connected to a microscope (NikonEclipse TE3000) and analyzed using Image Tool SoftwareTM. The total tumor volume (mm³) was computed by the multiplication of the slice sections and by summing the segmented areas [18].

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Hematological analysis

To evaluate possible interference of boldine in blood cell populations, we prepared blood smears stained with May-Grünwald-Giemsa staining. Analyzing blood samples microscopically, we found a tendency to increase the percentage of the population of neutrophils in the detriment of the other populations (lymphocytes, monocytes) in the animals treated with boldine (Table 1).

Enzymatic analysis

Boldine treatment for 14 days did not exert hepatic toxicity in healthy animals as evaluated from determination of gamma-glutamyltranspeptidase and ALT (liver damage marker), AST (liver and heart unspecific damage marker), alkaline phosphatase (pancreas, liver and bone unspecific damage markers) (Figure 3).

Tumor size reduction induced by treatment with boldine in vivo

To determine whether boldine can suppress tumor growth in vivo, adult Wistar rats were inoculated in the right striatum with C6 glioma cells and, after 10 days of tumor development, they were treated with boldine 50mg/Kg/day. After 10-day of initiate treatment, (20 days after implantation of tumor cells) boldine group rats showed significantly lower tumor volumes compared with control and vehicle groups. There was no statistical difference between vehicle group and control group. The results demonstrated that boldine was effective to inhibit tumor growth in rat C6 glioma implantation model. Tumor volume quantification and representative images of tumors in different experimental groups are shown in Figure 4.

Histopathological analysis of tumors

Haematoxylin and eosin (H&E) stained tumor sections examination showed C6 cells growing in the intracerebral, intraventricular and intraparenchymal spaces (data not shown). H&E examination also shows that the untreated tumor presented nuclear pleomorphism, foci of tumor necrosis, intratumoral hemorrhage, lymphocytic infiltration and vascular proliferation, which are characteristics of glioblastoma multiforme in humans (Table 2). These characteristics were less pronounced when rats were treated with boldine indicating a less
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Figure 3: Enzymatic activity in healthy animals treated with boldine. Data show that boldine did not interfere in (A) Gamma-GT, (B) ALT, (C) AST and (D) alkaline phosphatase activities. The values are represented as means ± S.D. of all animals used (n=4 per group).

Figure 4: Boldine inhibits glioma growth in vivo. Animals were treated, as described in Section 2. Tumor size was measured 20 days after implantation of C6 cells by three hematoxylin and eosin (H&E) sections of each tumor. For tumor size quantification, images were captured using a digital camera connected to a microscope and total volume (mm$^3$) was determined using Image Tool SoftwareTM. (A) Representative tumor images of the control, vehicle and boldine-treated groups, magnification 10x; (B) Tumor size quantification of implanted gliomas. The values are represented as means ± S.D (Control group – 6 animals; Vehicle group – 6 animals; boldine-treated group – 8 animals). Data were analyzed by ANOVA followed by Tukey’s test. *** p < 0.001.

Table 2: Histological characteristics of implanted gliomas.

|                      | Control (n=6) | Vehicle (n=6) | Boldine (n=8) |
|----------------------|---------------|---------------|---------------|
| Coagulative necrosis | 5/6 (83,3%)   | 6/6 (100%)    | 5/8 (62,5%)   |
| Intratumoral hemorrhage | 6/6 (100%) | 6/6 (100%)    | 6/8 (75%)     |
| Vascular proliferation | 6/6 (100%)  | 6/6 (100%)    | 7/8 (87,5%)   |
| Microabscess         | 3/6 (50%)     | 4/6 (66,6%)   | 2/8 (25%)     |
| Lymphocytic infiltration | 6/6 (100%) | 6/6 (100%)    | 6/6 (100%)    |
| Peripheral pseudopalisading | 3/6 (50%) | 3/6 (50%)     | 5/8 (62,5%)   |

The histological variables were regarded as present or absent. (n = number of animals per group)

Discussion

Many of our present medicines are directly or indirectly derived from higher plants. Clinical plant-based research has made particularly
rewarding progress in the important fields of anticancer therapy, such as curcumin [19,20], resveratrol [21], and others. In this study we propose a novel natural compound, the alkaloid boldine, for the development of a new anticancer agent, which could be used in conjunction with the established chemotherapies regimens. The precedent of the prolonged tradition of pharmaceutical use of boldine and boldine-containing boldo preparations suggesting that boldine exhibits low toxicity and the fact that this alkaloid is present in high concentrations in the bark of the Chilean boldo tree (Peumus boldus Mol., Monimiaceae), makes it an interesting candidate for development as a natural drug [2].

In the present study, we performed toxicological experiments in healthy Wistar rats after treatment with 50 mg/Kg of boldine administrated intraperitoneally for 14 days. This treatment regimen did not cause mortality in animals. In fact, there are studies demonstrating that high doses are needed to induce death in several mammalian species (e.g., 15 g of boldine, administered orally, were necessary to kill a 12 kg dog [2]). Studies by Kreitmair (1952) [3] reported that 500 and 1000 mg/kg (p.o.) were required to induce the death of mice and guinea pigs, respectively. Studies conducted later by L’evy-Appert-Collin and L. evy (1977) [22], estimated a LD50 of 250 mg/kg (i.p.) in mice. The necropsy of the rats at the end of boldine treatment in our study did not show any macroscopic changes in the observed organs, and their weights did not differ from those of the control group (Figure 1B). Microscopic investigation of these organs by H&E analysis demonstrated absence of toxicity (Figure 2A). Because boldine was demonstrated to be an inhibitor of prostaglandin biosynthesis [7], the occurrence of gastrointestinal damage was evaluated. No lesions were found in stomach or intestine after the treatment regimen employed.

To evaluate the possible liver damage in the boldine-treated healthy rats, we measured the levels of the enzymes gamma-glutamyl transpehase, alanine aminotransaminase, aspartate aminotransaminase and alkaline phosphatase in the rat blood serum. None of the treated animals presented significant alterations in the investigated enzymes, discarding hepatic damage (Figure 3). Studying the effects of long-term administration of boldine, Almeida and colleagues (2000) [23], did also not observed hepatotoxicity, assessed by blood transaminases or urea levels, in rats given boldine p.o. daily at 500 mg/kg for 30 and 60 days, but a low degree of toxicity was seen at 800 mg/kg.

Manual white blood cells differential was performed in blood smears stained with May-Grünwald-Giemsa. Analyzing the results, we found that the healthy animals treated with boldine had a tendency to increase the population of neutrophils. Earlier work by González-Cabello and colleagues (1994) [5] suggested that boldine present immunomodulating properties on natural killer cells and Philipov and colleagues (1998) [24] found that boldine inhibit the in vitro concanavalin A-induced proliferation of mouse splenocytes. Although these works suggest some possible immunomodulating properties of boldine, the actual potential of this aporphine to modify cellular immune functions require confirmation and further assessment.

Our findings, in conjunction with the overall results available on the toxicity of boldine in the literature, point to a relatively low toxicity of this compound. However, its actual innocuousness in humans still remains to be established in further investigations.

To determine whether the natural alkaloid boldine is able to avert brain tumor growth in an authentic microenvironment, we examined the effects of the compound in an orthotopic and immunocompetent rat model of glioma that mirrors the hallmarks of human gliomas [18]. This in vivo model is one of the most useful tools for a variety of studies, especially for investigations of glial tumor biology, as well as for experimental chemotherapy of brain malignancies [25]. We treated C6-implanted Wistar rats with 50 mg/kg/day boldine, ip, from the 10th to the 20th day after glioma implantation. Our experiments demonstrate that the treatment significantly diminished the growth of implanted gliomas in rats after twenty days of tumor induction (Figure 4). In addition, pathological analysis demonstrated that malignant characteristics appeared to be lower in boldine-treated rats (Table 2 and Figure 5). Among these, the reduction of the mitotic index was significant in treated rats, as observed by Ki67 immunostaining (Figure 6), indicating a tumor less proliferative. The vehicle-treated group did not present any significant difference from control group in any of the parameters analyzed; therefore we did not present the data from this group in all figures.

Angiogenesis is an extremely important process for sustained tumor growth. Thus, it constitutes an important point in the control of cancer progression and its inhibition may be a valuable new approach to cancer therapy. Studies have demonstrated that C6 cells secrete VEGF, a major angiogenic factor in glioma, which takes part of a critical signaling pathway for regulation of tumor angiogenesis [26,27]. In our study, we assessed VEGF expression by immunohistochemical staining. The results indicate no difference in VEGF staining between rat tumors treated with boldine and the control group (Figure 7). The absent of any difference in VEGF staining could be due to the dose used in this study. Maybe, it is possible that highest doses could interfere in VEGF secretion by glioma cells. Further studies are necessary to clarify this hypothesis.

Effective therapies for glioblastomas are scarce, and long term survival is a rarity [28]. The limited success of current treatment regimens is mainly caused by the ability of malignant glioma cells to diffusely infiltrate the surrounding healthy brain [29], a hallmark of glioblastoma that is recapitulated in the rat model of glioma [18]. Therefore, any new modality to replace or support current treatments for malignant gliomas would be helpful. In summary, this is the first report demonstrating therapeutic effects of boldine on glioma growth in immunocompetent mice. The effectiveness of boldine as an antitumor agent in our in vivo glioma model and its relatively safety observed in the experiments with healthy animals now justify the pursuit of studies aimed to explore its pharmacokinetics, major biotransformation pathways, its actual innocuousness in humans and further its actual therapeutic value in phase I clinical studies. As combination therapies are promising in glioblastoma, boldine might serve as an adjunct to established chemotherapeutic drugs and/or radiation therapy.
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