RESEARCH ARTICLE

Accuracy of Presepsin in Sepsis Diagnosis: A Systematic Review and Meta-Analysis

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Abstract

Objective

It's difficult to differentiate sepsis from non-sepsis, especially non-infectious SIRS, because no good standard exists for proof of infection. Soluble CD14 subtype (sCD14-ST), recently re-named presepsin, was identified as a new marker for the diagnosis of sepsis in several reports. However, the findings were based on the results of individual clinical trials, rather than a comprehensive and overall estimation. Thus, we conducted this systematic review and meta-analysis to estimate the pooled accuracy of presepsin in patients with sepsis suspect.

Methods

A comprehensive electronic search was performed via internet retrieval system up to 15 December 2014. Methodological quality assessment was applied by using the QUADAS2 tool. The diagnostic value of presepsin in sepsis was evaluated by using the pooled estimate of sensitivity, specificity, likelihood ratio, and diagnostic odds ratio, as well as summary receiver operating characteristics curve.

Results

Nine studies with 10 trials and 2159 cases were included in the study. Only two trials had low concerns regarding applicability, whereas all trials were deemed to be at high risk of bias. Heterogeneity existed in the non-threshold effect, but not in the threshold effect. The pooled sensitivity of presepsin for sepsis was 0.78 (0.76–0.80), pooled specificity was 0.83 (0.80–0.85), pooled positive likelihood ratio was 4.63 (3.27–6.55), pooled negative likelihood ratio was 0.22 (0.16–0.30), and pooled diagnostic odds ratio was 21.73 (12.81–36.86). The area under curve of summary receiver operating characteristics curve was 0.89 (95%CI: 0.84 to 0.94) and Q* index was 0.82 (95%CI: 0.77 to 0.87).
Conclusion

This meta-analysis demonstrates that presepsin had some superiority in the management of patients, and may be a helpful and valuable biomarker in early diagnosis of sepsis. However, presepsin showed a moderate diagnostic accuracy in differentiating sepsis from non-sepsis which prevented it from being recommended as a definitive test for diagnosing sepsis in isolation, but the results should be interpreted cautiously.

Introduction

Sepsis is a type of systematic inflammatory response syndrome (SIRS) caused by the invasion of pathogens or conditional pathogenic bacteria into the blood circulation. It can develop into severe sepsis, septic shock, and multiple organ failure. Sepsis occurs in 1%–2% of all hospitalized patients and accounts for as much as 25% of intensive care unit (ICU) cases [1]. When accompanied by organ system dysfunction or cardiovascular shock, severe sepsis or septic shock occurs and causes millions of deaths worldwide each year [2, 3]. However, there is no good standard exists for proof of infection, no matter blood microbiological cultures which often lead to a late and imprecise report, or clinical symptoms which are non-specific and overlap with signs of SIRS without infection [4]. Delay of diagnosis and treatment with appropriate antimicrobial chemotherapy is the main reason for high morbidity and mortality associated with sepsis, thus looking for a reliable and timely biomarker for sepsis is of utmost importance [5]. At present, more than 178 markers have been found for sepsis, most of which are intermediate products of the inflammatory process and some are sepsis pro-inflammatory cytokines [6]. However, the most reliable biomarkers for precise diagnosis and prediction of the future process of patients suffering from severe sepsis or septic shock are still uncertain or are controversial [7].

As a glycoprotein expressed on monocytes and macrophages, cluster of differentiation 14 (CD14) serves as a receptor of the lipopolysaccharide (LPS)-lipopolysaccharide binding protein complexes and activates a series of signal transduction pathways and inflammatory cascades that finally lead to SIRS [8]. CD14 has two forms, namely, a membrane-bound CD14 (mCD14) and soluble CD14 (sCD14). sCD14 plays an important role in mediating the immune responses to LPS of CD14-negative cells, such as endothelial and epithelial cells. During inflammatory stress, sCD14 is cleaved in plasma, and the N-terminal fragment of 13 kDa has been identified as sCD14 subtype (sCD14-ST; also known as presepsin) [9].

In 2004, the value of presepsin in the diagnosis and evaluation of sepsis was discovered [10], and it has become an alternative biomarker to aid the diagnosis of sepsis. Since then, several studies have reported this compound as a new biomarker in the prediction of sepsis. However, there was a large variability regarding the results and sample sizes of these studies. For instance, the specificity in the report of Palmiere et al. [11] was only 0.44, whereas specificity was 0.98 in the study of Vodnik et al. [12]. Thus, the real value of presepsin in diagnosing sepsis is uncertain. Moreover, the findings of present reports were based on the results of individual clinical trials, and the literature lacks a pooled and robust appraisal of all the evidence for the diagnostic accuracy of presepsin testing. Systematic review and meta-analysis of the diagnostic efficiency are rigorous approaches for examining and synthesizing the evidence in the evaluation of the diagnostic and screening test [13]. Therefore, we conduct this systematic review and meta-analysis to evaluate the relationship between presepsin and sepsis to precisely estimate the diagnostic accuracy of the presepsin test.
Materials and Methods

Literature Search
A comprehensive electronic search of the PubMed, Embase, Medline, Cochrane Library, and China National Knowledge Infrastructure (CNKI) was performed via the Internet retrieval system. No language limitation was indicated, and the articles' inclusion period was until 15 December 2014. Search terms included (“presepsin,” or “sCD14-ST,” or “soluble CD14 subtype,”) and (“sepsis”) and (“diagnosis,” or “diagnostic value,” or “diagnostic biomarker”). In addition, content experts were contacted, and bibliographies of the relevant studies were reviewed to identify additional references. When multiple publications with the same or overlapping patient population from the same institution were identified, only the published report with the largest series was included. This meta-analysis was reported following the Preferred Reporting Items for Systematic Review and Meta-Analysis statement [14].

Study Selection and Data Extraction
A study was eligible for our meta-analysis if it satisfied the following requirements: (1) its purpose was to evaluate or explore the diagnostic value of presepsin as a single index for differentiation between critically ill patients with sepsis from those of non-infection, such as patients with SIRS without infection, with or without healthy people; (2) data were available for calculating the true positives, false positive, false negatives, and true negatives; (3) applying the clinical criteria of the American College of Chest Physicians and Society of Critical Care Medicine (ACCP/SCCM) [15–17] as reference standard for defining sepsis and SIRS; and (4) with a prospective controlled design. The studies considered ineligible for the meta-analysis were as follows: reviews, conference abstracts, editorials, or case reports; duplicate studies; and those with insufficient information to calculate accuracy estimates.

All data extractions were independently completed by two authors (WJY and HLR) and were checked by a third reviewer (HTP). Any disagreement was resolved through discussion. The following data were extracted from each eligible study: authors, years of publication, locations of the study, selection and characteristics of the sample population, diagnostic test performed, cut-off value, sensitivity, and specificity.

Quality Assessment
Each full-text article was reviewed independently by two authors (WJY and HLR) and scored with the Quality Assessment of Diagnostic Accuracy Studies 2 (QUADAS-2) tool, and all disagreements were resolved by consensus. QUADAS-2 tool consists of four domains: patient selection, index test, reference standard, and flow/timing. “Risk of bias” was evaluated for all the domains, whereas “concerns regarding applicability” was evaluated for the first three domains. QUADAS-2 does not utilize a comprehensive quality score, but rather an overall judgment of “low,” “high,” or “unclear” risk. For judging the “risk of bias,” the reviewers analyzed all articles by answering each signaling question with a “yes,” “no,” or “unclear” and then by recording the following risk scores for each domain: “L” for “low risk of bias,” “H” for “high risk,” and “U” for “unclear.” In QUADAS-2, “applicability” means whether certain aspects of an individual research match the review's question or not. The principle and method of judging the “concerns regarding applicability” are the same as those of the “risk of bias,” but without any signaling question. For overall judgment of "low risk of bias" or "low concern regarding applicability," a study must be ranked "low" on all relevant domains. If a study receives a "high" or "unclear" rating in one or more domains, then it may be judged as "at risk of bias" or having "concerns regarding applicability" [18].
Statistical Analysis

Data analysis was performed using the Meta-DiSc statistical software version 1.4 [19]. Sensitivity (Se), specificity (Sp), positive likelihood ratio (PLR), negative likelihood ratio (NLR), and diagnostic odds ratio (DOR) were computed for each study. DOR is a comprehensive measure for both Se and Sp or both PLR and NLR, and is regarded as an appropriate global measure for comparing the accuracy of different diagnostic tests, which is estimated by the following formula: (sensitivity/[1-sensitivity])/(1-specificity)/specificity) [20]. Pooled summary effect estimates were calculated using a random effects model (DerSimonian and Laird method) when high heterogeneity exists; otherwise, a fixed effect model (Mantel–Haenszel method) was used [21].

A summary receiver operating characteristics (SROC) curve was drawn to solve the inconsistency of various research results. Serving as global measures for the SROC curve, the area under curve (AUC), which could serve as a probability of correctly recognizing cases and non-cases by a diagnostic test, was also interpreted according to the following guidelines: low for 0.5–0.7, moderate for 0.7–0.9, and high for > 0.9 [22, 23]; as well as the Q² value, which is the point on the SROC curve where sensitivity equals specificity and is in the range 0–1 (1 indicates better test performance) [24, 25].

The heterogeneity of a diagnostic test is caused by threshold or non-threshold effects in general. In this study, the heterogeneity caused by threshold effect was evaluated by the appearance of a SROC curve (a “shoulder-like” distribution suggested the existence of a threshold effect) and the calculation of Spearman correlation coefficient (ρ) between sensitivity logarithm and (1-specificity) logarithms. Furthermore, heterogeneity caused by non-threshold effect was quantified by applying the χ² (for Se and Sp) and the Cochrane-Q test (for PLR, NLR, and DOR) and by determining the I² metric. Statistical significance was set at P < 0.05 or I² > 50% for the heterogeneity testing. A meta-regression was also performed to determine the factors for heterogeneity that have been influenced by the non-threshold effect. To eliminate the influence of the confounding factors, we also conducted a sensitivity analysis by calculating the pooled DOR and 95% CI after omitting the studies which including some possible confounding factors. Funnel plots, Begg’s rank correlation, and Egger’s linear regression method were also conducted to evaluate the potential publication bias through the STATA version 11.0 (STATA Corporation, College Station, TX, USA) [26, 27].

Results

Characteristics of included studies

Nine studies [7, 12, 28–34] met the inclusion criteria for our meta-analysis, one [34] of which included two trials. Therefore, a total of 10 trials with 2159 cases were included (1320 patients with sepsis, 512 with SIRS of non-infectious origin, and 327 healthy people), the sample sizes of which ranged from 104 to 959. These 10 trials were eligible for this meta-analysis. These studies principally originated from Western Europe (2 from Italy [29, 33], and 1 from Germany [7]) and Eastern Asia (3 from China [30–32], 1 from Korea [28], and 1 from Japan [34]). These studies along with one study from Serbia [12] were published between 2011 and 2014. Seven studies were written in English [7, 12, 28, 29, 32–34], and the other two were written in Chinese [30, 31]. All of the 9 studies were conducted in a prospective controlled design. Three studies were done in internal care unit (ICU) or critical care unit (CCU) [7, 29, 31], five studies in emergency department (ED) [12, 28, 31, 32, 33], while only one in CCU and ED [34]. Four studies clearly stated that the blinding method was used in presepsin determination [7, 28, 31, 33], and three studies included only patients with non-infectious SIRS as controls [31, 33, 34],
while the other six studies included both patients of SIRS without infection and healthy people in the control group [7, 12, 28–30, 32].

A summary of the characteristics of the 9 studies is outlined in Table 1, and the clinical natures of each trial are shown in Table 2. A flow diagram describes the details of the study selection progress (Fig 1).

### Results of quality assessment

The results of the quality assessment are listed in Table 3, and the details are presented in S1 Table. When the QUADAS-2 tool was used to assess the risk of bias in the 10 trials, 7 trials showed problems in patient selection, 5 showed problems in reference standard, 4 showed problems in study flow, and all 10 trials showed problems in the index test. Moreover, when

### Table 1. Characteristics of the included studies.

| Author (year) | Nation       | Language | Recruitment time | n  | Male/female | T/C | Testing method | Blind |
|---------------|--------------|----------|------------------|----|-------------|-----|----------------|-------|
| Behnes M (2014) [7] | Germany      | English  | Since 2011.10    | 176| 111/65      | 107/69| CLEIA          | Yes   |
| Kweon OJ (2014) [28] | Korea        | English  | 2012.9–2013.7    | 118| 59/59       | 73/45| CLEIA          | Yes   |
| Sargentini V (2014) [29] | Italy       | English  | 2013.3–2013.7    | 104| 47/57       | 60/44| CLEIA          | NR    |
| Su MH (2014) [30] | China        | Chinese  | 2012.11–2013.8   | 115| 65/50       | 72/43| CLEIA          | NR    |
| Yu J (2014) [31]  | China        | Chinese  | 2012.6–2012.12   | 176| 119/57      | 63/113| CLEIA         | Yes   |
| Liu B (2013) [32] | China        | English  | 2011.12–2012.10  | 959| 570/389     | 680/279| CLEIA        | NR    |
| Ulla M (2013) [33] | Italy        | English  | 2012.1–2013.1    | 189| 116/73      | 106/83| CLEIA         | Yes   |
| Vodnik T (2013) [12] | Serbia      | English  | NR               | 130| 71/59       | 30/100| CLEIA        | NR    |
| Shozushima T (2011) [34] | Japan      | English  | 2009.8–2010.7    | 192| 117/75      | 129/63| CLEIA        | NR    |

NR: none reported; T/C: test group/control group; CLEIA: chemiluminescent enzyme immunoassay.

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### Table 2. Clinical nature of the included studies.

| Author (year) | Setting       | Admission category | Severity                  | Controls                       | Sampling time          |
|---------------|---------------|--------------------|---------------------------|-------------------------------|------------------------|
| Behnes M (2014) [7] | ICU           | Medical            | Sepsis, severe sepsis, sepsis shock | SIRS without infection, health | At clinical onset      |
| Kweon OJ(2014) [28] | ED            | Medical            | Sepsis, severe sepsis, sepsis shock | SIRS without infection, health | At admission           |
| Sargentini V (2014) [29] | CCU          | Medical, traumatic | Sepsis, severe sepsis     | SIRS without infection, health | At admission           |
| Su MH (2014) [30] | ED            | Medical            | Sepsis, severe sepsis, sepsis shock | SIRS without infection, health | Before any treatment   |
| Yu J (2014) [31] | ICU           | Traumatic          | Sepsis                    | SIRS without infection         | At clinical onset      |
| Liu B (2013) [32] | ED            | Medical            | Sepsis, severe sepsis, sepsis shock | SIRS without infection, health | At admission           |
| Ulla M (2013) [33] | ED            | Medical, surgical, traumatic | Sepsis, severe sepsis, sepsis shock | SIRS without infection         | At first medical evaluation |
| Vodnik T (2013) [12] | ED            | Surgical           | Sepsis, severe sepsis, sepsis shock | SIRS without infection, health | At admission           |
| Shozushima T (2011) [34] | CCU, ED      | Medical, traumatic | Sepsis, severe sepsis     | SIRS without infection         | At admission           |

NR: none reported; ICU: internal care unit; ED: emergency department; CCU: critical care unit; SIRS: systemic inflammatory response syndrome; ACCP/SCCM: the American College of Chest Physicians/Society of Critical Care Medicine Consensus Conference

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the concerns of applicability were considered, 8 trials showed problems in patient selection, but the reference standard and index test were used appropriately in all trials.

In general, all trials were at risk of bias. Two trials had low concerns regarding applicability, whereas the other 8 trials showed problems regarding applicability.

Results of heterogeneity test

From the appearance of the SROC curve (Fig 2) and the estimation of the Spearman correlation coefficient ($\rho = -0.261, P = 0.467$), we could conclude that no threshold effect existed.

However, when non-threshold effect was investigated, significant heterogeneity was measured in the overall Se ($\chi^2 = 83.47, P < 0.001, I^2 = 89.2\%$), Sp ($\chi^2 = 56.40, P < 0.001, I^2 = 84.0\%$), PLR (Cochran-$Q = 46.13, P < 0.001, I^2 = 80.5\%$), NLR (Cochran-$Q = 45.49, P < 0.001, I^2 = 80.2\%$), and DOR (Cochran-$Q = 35.15, P = 0.0001, I^2 = 74.4\%$). Therefore, a random-effect model was applied for data synthesis. To determine the sources of heterogeneity, we used location, number of cases, blind, cutoff value, setting, admission category, and control’s component as variables in the meta-regression analysis. The results are shown in Table 4. Results showed that the heterogeneity could not be explained by meta regression analysis.
Results of meta-analysis

For the 9 studies that represented 10 trials, cutoff values were ranged from 317 pg/ml to 700 pg/ml in each trial. The reported sensitivity of presepsin diagnosing sepsis ranged from 0.71 to 1.00, the specificity ranged from 0.62 to 0.98, the positive likelihood ratio ranged from 1.71 to 39.75, the negative likelihood ratio ranged from 0.02 to 0.34, and the diagnostic odds ratio ranged from 6.09 to 2403.40.

Overall, the pooled Se of presepsin for sepsis was 0.78 (95%CI: 0.76 to 0.80) (Fig 3A), pooled Sp was 0.85 (95%CI: 0.80 to 0.85) (Fig 3B), pooled PLR was 4.63 (95%CI: 3.27 to 6.55) (Fig 3C), pooled NLR was 0.22 (95%CI: 0.16 to 0.30) (Fig 3D), and pooled DOR was 21.73 (95%CI: 12.81 to 36.86) (Fig 3E). In addition, the AUC of SROC curve was 0.89 (95%CI: 0.84 to 0.94), and Q index was 0.82 (95%CI: 0.77 to 0.87) (Fig 2).

The results of each trial, and overall outcomes were listed in Table 5.

Results of sensitivity analysis and publication bias

In the sensitivity analysis, the influence of each study on the pooled AUC was examined by omitting each study one at a time. Regardless which study was removed, the pooled AUC estimated by the remaining studies did not change significantly. This result indicated that no individual study dominated the results of the meta-analysis, thereby validating the credibility of the outcomes (Table 6). To compare the diagnostic accuracy of presepsin for sepsis in studies with or without healthy controls, we also conducted a sensitivity analysis by obtaining data from 10 trials (6 trials provided data for controls with healthy people, and 4 provided data for controls without healthy people). When healthy people were included in controls with SIRS of non-infectious origin, the diagnostic accuracy was similar to that of including only SIRS without infection in controls as evaluated by AUC (0.90 [95% CI: 0.88 to 0.92] vs. 0.85 [95%CI: 0.82 to 0.88]; not tested for significance).
Fig 2. Summary receiver operator characteristic plots with 95% CIs of sensitivity against (1-specificity) of presepsin testing for sepsis.
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Table 4. Possible sources of heterogeneity of meta-analysis (Results of meta-regression analysis).

| Variance                                      | Coefficient | Standard error | P value | RDOR (95% CI)   |
|-----------------------------------------------|-------------|----------------|---------|-----------------|
| Location (Europe vs. Asia)                    | 2.341       | 0.8083         | 0.0626  | 10.40 (0.79; 136.26) |
| No. of cases (> 150 vs. < 150)                | -0.601      | 0.8330         | 0.5227  | 0.55 (0.04; 7.77)   |
| Cut-off value (≥ 600 pg/ml vs. < 600 pg/ml)   | 0.628       | 1.0447         | 0.5698  | 1.87 (0.15; 24.15) |
| Blind (Blind vs. NR)                          | 0.651       | 0.8551         | 0.4756  | 1.92 (0.24; 15.53) |
| Setting (ICU/CCU vs. ED vs. CCU + ED)         | -0.533      | 0.2762         | 0.1494  | 0.59 (0.24; 1.41)   |
| Admission category (only medical vs. others)  | 0.983       | 0.9208         | 0.3642  | 2.67 (0.14; 50.04) |
| Controls (SIRS vs. SIRS + healthy)            | 1.555       | 1.0332         | 0.2294  | 4.73 (0.18; 126.84) |

RDOR: relative diagnostic odds ratio; CI: confidence interval; NR: none reported; ICU: internal care unit; ED: emergency department; CCU: critical care unit; SIRS: systemic inflammatory response syndrome.
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Publication bias was analyzed by funnel plot, Egger’s test, and Begg’s test, which provided negligible evidence of publication bias for the outcome of DOR ($P$ value for Egger’s test = 0.140; $P$ value for Begg’s test = 0.067). The funnel plot is shown in Fig 4.

**Discussion**

At present, various biomarkers (alone or in combination) are used in the diagnosis of sepsis, including procalcitonin (PCT), C-reactive protein (CRP), interleukin (IL), and soluble form of triggering receptor expressed on myeloid cells-1 (Strem-1). However, the clinical value of these biomarkers is still controversy. Moreover, blood culture is treated as the gold criteria for sepsis diagnosis, but it always takes 48–72 hours to obtain the outcome when this approach is used. Blood culture has a low positive rate, which results in diagnosis delay, and the best treatment time is missed. Therefore, finding a reliable biomarker for the early and rapid diagnosis of sepsis is critical.
A large number of studies have found that sCD14 has important pathophysiological implications in the occurrence and development of many human diseases. sCD14 exists in the blood and urine of humans, and comprises 99% of the total amount of CD14 in the human body.

### Table 5. The diagnostic parameters of presepsin for sepsis in the included trials, and overall outcome.

| Study                  | Cut-off (pg/ml) | TP   | FP   | FN   | TN | Se  | Sp   | PLR          | NLR          | DOR          |
|------------------------|----------------|------|------|------|----|-----|------|--------------|--------------|--------------|
| Behnes (2014) [7]      | 700            | 97   | 16   | 10   | 53 | 0.91 (0.83–0.95) | 0.77 (0.65–0.86) | 3.91 (2.53–6.03) | 0.12 (0.07–0.22) | 32.13 (13.62–75.79) |
| Kweon (2014) [28]      | 430            | 64   | 8    | 9    | 37 | 0.88 (0.78–0.94) | 0.82 (0.68–0.92) | 4.93 (2.62–9.30) | 0.15 (0.08–0.28) | 32.89 (11.69–92.57) |
| Sargentini (2014) [29] | 600            | 52   | 12   | 8    | 32 | 0.87 (0.75–0.94) | 0.73 (0.57–0.85) | 3.18 (1.94–5.20) | 0.18 (0.09–0.36) | 17.33 (6.40–46.98) |
| Su M (2014) [30]       | 407            | 71   | 4    | 1    | 39 | 0.99 (0.93–1.00) | 0.91 (0.78–0.97) | 10.60 (4.17–26.97) | 0.02 (0.00–0.11) | 32.13 (13.62–75.79) |
| Yu J (2014) [31]       | 540            | 46   | 15   | 17   | 98 | 0.73 (0.60–0.83) | 0.87 (0.79–0.92) | 5.50 (3.35–9.02) | 0.18 (0.09–0.36) | 17.68 (8.12–38.48) |
| Liu B (2013) [32]      | 317            | 481  | 40   | 199  | 239| 0.71 (0.67–0.74) | 0.86 (0.81–0.90) | 4.93 (3.69–6.60) | 0.34 (0.30–0.39) | 14.44 (9.94–20.98) |
| Ulla M (2013) [33]     | 600            | 84   | 32   | 22   | 51 | 0.79 (0.70–0.87) | 0.61 (0.50–0.72) | 2.06 (1.54–2.74) | 0.34 (0.22–0.51) | 6.09 (3.19–11.60) |
| Vodnik T (2013) [12]   | 630            | 30   | 2    | 0    | 98 | 1.00 (0.86–1.00) | 0.98 (0.93–1.00) | 39.75 (11.66–135.31) | 0.02 (0.00–0.26) | 2403.40 (112.31–51433.97) |
| Shozushima T (2011) a  | 399            | 104  | 14   | 25   | 49 | 0.81 (0.73–0.87) | 0.78 (0.66–0.87) | 3.63 (2.27–5.80) | 0.25 (0.17–0.36) | 14.56 (6.97–30.43) |
| Shozushima T (2011) b  | 415            | 103  | 12   | 26   | 51 | 0.80 (0.72–0.86) | 0.81 (0.69–0.90) | 4.19 (2.50–7.03) | 0.25 (0.17–0.36) | 16.84 (7.86–36.07) |
| Overall outcome        |                | 78   | 0    | 80   | 80 | 0.78 (0.76–0.80) | 0.83 (0.80–0.85) | 4.63 (3.27–6.55) | 0.22 (0.16–0.30) | 21.73 (12.81–36.86) |

*a Results of first of two trials in this article  
*b Results of second of two trials in this article  
TP: true positive; FP: false positive; FN: false negative; TN: true negative; Se: sensitivity; Sp: specificity; PLR: positive likelihood ratio; NLR: negative likelihood ratio; DOR: diagnostic odds ratio.

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### Table 6. The pooled AUC and 95% CI after omitting each trial in the meta-analysis (The results of sensitivity analysis).

| Study                  | AUC | 95% CI    |
|------------------------|-----|-----------|
| Behnes M (2014)        | 0.89| 0.83–0.95 |
| Kweon OJ (2014)        | 0.89| 0.83–0.95 |
| Sargentini V (2014)    | 0.90| 0.84–0.96 |
| Su MH (2014)           | 0.88| 0.84–0.92 |
| Yu J (2014)            | 0.89| 0.82–0.96 |
| Liu B (2013)           | 0.89| 0.81–0.97 |
| Ulla M (2013)          | 0.89| 0.87–0.91 |
| Vodnik T (2013)        | 0.88| 0.84–0.92 |
| Shozushima T (2011) a  | 0.90| 0.84–0.96 |
| Shozushima T (2011) b  | 0.89| 0.83–0.95 |

*a Results of first of two trials in this article  
*b Results of second of two trials in this article  
AUC: the area under the summary receiver operating characteristic curve; CI: confidence interval.

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with a normal concentration of 2 to 6 μg/ml in serum [35]. sCD14 concentration is closely related to the level of endotoxin and plays a critical role on mediating the inflammatory reaction of endothelial and epithelial cells. As a subtype of sCD14, presepsin can signal a much earlier and faster increase of sepsis when compared with PCT. Hence, presepsin may be a useful diagnostic biomarker for sepsis. However, the results of the current studies on presepsin showed considerable differences, such as in the SE range (0.73–1.00) and in the SP range (0.44–0.98), which may be due to the differences in the sample populations and study designs, differences in the selection of people for the control, and differences in cut-off values. Therefore, a meta-analysis, which is useful in integrating results from independent studies for a specified outcome, was conducted to determine the pooled outcomes.

In our meta-analysis, 9 studies that represented 10 trials were included, of which the sample populations were collected from ICU, ED, and CCU, or were typically seen in medical, surgical, and traumatic. The findings of this meta-analysis were therefore applicable to common clinical settings in which critically ill patients were managed. In these studies, the diagnostic value of presepsin was evaluated by testing the level in blood sample of septic patients compared with that of patients with non sepsis. Blood samples were drawn at admission, or clinical onset, or
before any treatment, and presepsin levels were measured by using a chemiluminescent enzyme immunoassay that allowed making automated measurements in a short time. The pooled results indicated that presepsin showed a moderate diagnostic value for distinguishing sepsis from non sepsis. However, systematic reviews of diagnostic accuracy studies are often characterized by a notable heterogeneity caused by the differences in the design and implementation of the studies. Thus, our results should be interpreted cautiously [36].

First, non-infectious SIRS patients were assessed as controls in each study, which is difficult to distinguish from sepsis because the clinical signs often overlap between them. However, healthy people also served as controls in some included studies, which may make the pooled outcomes much higher than the real results. Then we conducted a sensitivity analysis to make an exact evaluation, but the results had not changed much in the index of AUC compared with the overall AUC (0.89 [95%CI: 0.84 to 0.94] vs. 0.85 [95%CI: 0.82 to 0.88]), thus, the including of healthy people seemed to have little influence on the overall outcomes. But further study designed with better homogeneity is advisable.

Second, the cutoff values of presepsin among these included studies varied greatly, even though using the same test method. The difference may lie in the admission category, number of cases, and sample timing. Therefore, this research was limited in terms of qualitative analysis, but not in terms of quantitative analysis.

Third, all included studies declared that infection was confirmed by microbiologically or clinically according to an internationally recognized gold standard, but most failed to provide the information about whether previous antibacterial treatment was used. Under the impact of antibiotic drugs, bacteraemia occurs in only 30% of patients with sepsis [37, 38]. Moreover, the level of presepsin decreased rapidly after treatment [28]. Thus, the absence of such therapeutic details may increase interobserver variability, and add false judgment about the patient’s medical condition [4].

Fourth, extreme heterogeneity existed among these included studies. We detected substantial heterogeneity by bringing the study characteristics and clinical natures into meta-regression analysis, but none of them was responsible for the majority of heterogeneity. Thus, some unrecorded difference may contribute to the heterogeneity, and a study with better design and more homogenous population is needed to avoid the heterogeneity.

Fifth, LPS is a component of the Gram-negative bacterial cell wall. As a receptor of LPS, presepsin is easy to imagine that whether it only work for Gram-negative infections. Studies have shown that the Se of presepsin was not significantly different between Gram-positive and Gram-negative bacterial infections [17, 39, 40], while no disparity of presepsin concentration was found among the infection caused by Gram-positive or Gram-negative bacterial infections [30]. So it seems to be lack of evidence to prove that presepsin is not a predictor only for sepsis induced by Gram-negative cocci infection.

In previous clinical practices, many biomarkers were widely applied in diagnosis of sepsis, and PCT is the most widely used one. Several evidence-based researches have proved that PCT had a low diagnostic performance in differentiating sepsis from critically ill patients, and can not be recommended as the single definitive test for sepsis diagnosis [4, 41, 42]. When compared to PCT, presepsin also showed a similar diagnostic accuracy for sepsis with respect to Se (0.78 [95% CI: 0.76 to 0.80] vs. 0.77 [95% CI: 0.72 to 0.81]), Sp (0.83 [95% CI: 0.80 to 0.85] vs. 0.79 [95% CI: 0.74 to 0.84]), AUC (0.89 [95% CI: 0.84 to 0.94] vs. 0.85 [95% CI: 0.81 to 0.88]), according to a latest evidence about the accuracy of PCT for differentiating sepsis from SIRS without infection [4]. To avoid the bias of results raised by including healthy people in the research, we conducted a sensitivity analysis to exclude these studies. From the results of sensitivity analysis, we noted that, when focused on studies aimed at discriminating sepsis from non-infectious SIRS, the accuracy of presepsin was the same to that of PCT, with respect to
AUC (0.85 [95% CI: 0.82 to 0.88] vs. 0.85 [95% CI: 0.81 to 0.88]). Whether a marker can be used alone for the diagnosis of sepsis depends on its diagnostic performance. Therefore, presepsin did not show any advantage in diagnostic performance of sepsis compared to PCT, and it cannot be recommended as a single definitive test for sepsis diagnosis. However, presepsin showed some superiority in the management of patients, such as both presepsin and PCT are elevated in non-infectious SIRS and sepsis, but presepsin can signal a much earlier and faster increase in sepsis, and perform a unique capacity of distinguishing the severity of sepsis [7, 12, 33]. Furthermore, by using the PATHFAST analysis system, presepsin test only takes 17 min and can be conducted at bedside, thereby complying with the guidelines for the diagnosis and treatment of sepsis [1]. Consequently, although presepsin showed a moderate diagnostic accuracy which was similar to PCT, but it was still a helpful biomarker for early diagnosis of sepsis and severity evaluation.

In addition, other limitations are present. First, the search range was limited in published studies, which means we might have missed some unpublished, but valuable studies. Second, diagnostic tests were generally designed in case-control study or cross-sectional study, which belonged to the third level design in evidence-based medicine. Thus, many studies in this area were not designed with a blind method or did not clearly use the blind method to strengthen the argument, which may have contributed to the possibility of bias [43, 44]. Third, this systemic review did not investigate the prognostic value of presepsin in sepsis because not enough evidence is available.

In conclusion, presepsin had some superiority in the management of patients, and may be a helpful and valuable biomarker in early diagnosis of sepsis. However, presepsin showed a moderate diagnostic accuracy in differentiating sepsis from non-sepsis which prevented it from being recommended as a definitive test for diagnosing sepsis in isolation, but the results should be interpreted cautiously due to the heavy heterogeneity and different clinical natures in these included studies. Moreover, further randomized controlled researches are required in the context with unified clinical information.

Supporting Information

S1 Checklist. Prisma checklist.
(DOC)

S1 Table. Details of QUADAS-2 quality assessment for each study. This table presented the details of quality assessment for each study by the QUADAS-2 tool.
(DOC)

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Author Contributions

Conceived and designed the experiments: TH. Performed the experiments: JW FW. Analyzed the data: LH GZ. Contributed reagents/materials/analysis tools: LH. Wrote the paper: JW GZ.

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