Remus: System for remote deep brain interventions

Flexible manipulation of deep brain targets in task performing subjects

Taylor D. Webb, Matthew G. Wilson, Henrik Odéen, Jan Kubanek
taylor.webb@utah.edu (T.D.W.)
jan.kubanek@utah.edu (J.K.)

Highlights
Remus delivers ultrasound into deep brain targets of task-performing subjects

Targets are specified programmatically and at high spatial and temporal precision

Brief pulses delivered to deep brain regions modulate visual choice behavior

The system enables reproducible daily applications and continuous safety monitoring

Webb et al., iScience 25, 105251
November 18, 2022 © 2022
The Author(s).
https://doi.org/10.1016/j.isci.2022.105251
Remus: System for remote deep brain interventions

Taylor D. Webb,1,* Matthew G. Wilson,1 Henrik Odéen,2 and Jan Kubanek1,3,*

SUMMARY
Transcranial-focused ultrasound brings personalized medicine to the human brain. Ultrasound can modulate neural activity or release drugs in specific neural circuits but this personalized approach requires a system that delivers ultrasound into specified targets flexibly and on command. We developed a remote ultrasound system (Remus) that programmatically targets deep brain regions with high spatiotemporal precision and in a multi-focal manner. We validated these functions by modulating two deep brain nuclei—the left and right lateral geniculate nucleus—in a task-performing nonhuman primate. This flexible system will enable researchers and clinicians to diagnose and treat specific deep brain circuits in a noninvasive yet targeted manner, thus embodying the promise of personalized treatments of brain disorders.

INTRODUCTION
One in three patients with brain disorders is treatment-resistant (Bystritsky, 2006; Al-Harbi, 2012; Zesiewicz et al., 2010; Elias et al., 2016; Ferguson, 2001; Karceski, 2007; Lancet, 2017; Ahnsbrak et al., 2017). Neuromodulation provides these patients with new treatment options, but invasive, surgical approaches are risky and inflexible, and current noninvasive approaches do not have the necessary spatial resolution. Transcranial-focused ultrasound provides a new set of regimens that enable the manipulation of brain activity noninvasively and at high spatiotemporal precision. Depending on stimulus duration, ultrasound modulates neural activity (Fry, 1958; Kubanek et al., 2020; Deffieux et al., 2013; Tufail et al., 2010; Lee et al., 2015; Legon et al., 2014) or induces changes in functional connectivity (Verhagen et al., 2019; Folloni et al., 2019; Bongioanni et al., 2021). In addition, when combined with nanoparticles or microbubbles, ultrasound can be used to deliver drugs, genes, or stem cells selectively into the specified target/s. This can be achieved by releasing drugs from nanoparticle carriers, (Wang et al., 2018a; Rapoport et al., 2011; Lea-Banks et al., 2020) or using microbubbles to temporarily disrupt the blood-brain barrier (Abrahao et al., 2019) and so deliver large agents that would not pass otherwise. Thus, this form of energy provides treatment options for the large number of patients who are resistant to current forms of treatment.

Nonetheless, these applications are limited by technological challenges. A key requirement for the success of ultrasound-based therapies is the ability to target a specified site flexibly and reproducibly. This flexibility is critical given that the neural sources of many mental and neurological disorders are poorly understood and vary from individual to individual (Braun et al., 2018; Price and Drevets, 2012; Peirs and Seal, 2016; Insel and Cuthbert, 2015). Systems that could provide this functionality—phased arrays—are primarily designed for ablative treatments (Ghanouni et al., 2015) with only a few phased array systems available for other transcranial applications (Liu et al., 2014; Chaplin et al., 2018).

We developed Remus, a system that manipulates deep brain circuits remotely and programmatically, thus enabling personalized circuit manipulations in clinical and research settings. The platform enables an operator to specify one or more targets, together with the desired timing and ultrasonic waveform, in software. The design is MR-compatible, which enables clinicians and researchers to confirm precise targeting. The system’s imaging functionality ensures reproducible positioning of the device and evaluates the quality of the ultrasound coupling to a subject’s head. We validated these capabilities in a nonhuman primate (NHP). Remus can be used to validate existing and devise new ultrasonic protocols in large animals, NHPs, and humans. Ultimately, the system will be used to provide precision treatments to patients who are currently out of options.

RESULTS
Remus
Remus is a practical platform that delivers focused ultrasound into specified deep brain targets remotely, with the skull and skin intact (Figure 1, top). The targets of the ultrasound are specified in software. The

1Department of Biomedical Engineering, University of Utah, Salt Lake City, Utah, USA
2Department of Radiology and Imaging Sciences, University of Utah, Salt Lake City, Utah, USA
3Lead contact
*Correspondence:
taylor.webb@utah.edu
jan.kubanek@utah.edu
https://doi.org/10.1016/j.isci.2022.105251
This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).
software can flexibly target individual sites in sequence (Figure 1, bottom) or simultaneously. An animated
graphical abstract provides a visual representation of Remus and is available in the online version of this
article.

Remote modulation of choice behavior
We validated these features in an NHP. Specifically, we used the system to programatically deliver
brief pulses of focused ultrasound into the left and right LGN while quantifying the effects on behavior
(Figure 2A). The two LGNs were targeted electronically, without mechanical movement of the transducer.

We found that brief, 300 ms pulses of ultrasound directed into the left and right LGN transiently modulated
choice behavior in the NHP. The subject was asked to look at a left or a right target, whichever appeared
first. We quantified the effects of the ultrasonic neuromodulation in the controlled condition in which both
targets appeared at the same time. We assessed the proportion of choices of each target when the left
LGN was stimulated, and when the right LGN was stimulated and quantified the proportion of choices
The side of the stimulation was randomized. The animal was deciding whether to look at a left or a right target. The brief, low-intensity ultrasonic pulses influenced the choice behavior. Specifically, variation in the duty cycle of the ultrasound significantly changed the proportion of contralateral choices. Stimuli pulsed at 10% duty significantly decreased the proportion of contralateral choices relative to continuous stimuli. The decrease in contralateral choices is consistent with neuronal inhibition (vertical arrows). Data are represented as mean ± SEM. **: p < 0.01 as measured by a two-sided t-test.

(B) Repeated application of neuromodulatory ultrasound to deep brain circuits does not inhibit the subject from improving in the task over time. The break in the x axis marks an increase in the difficulty of the task. The positive slope of the linear fit is significant before the increase in difficulty (r = 0.27; F(28) = 10.1; p = 0.004).

Notably, in these analyses, we only compared effects for trials in which there always was an ultrasound stimulus: either the left or the right LGN was sonicated. This controls for potential generic artifacts that can be associated with ultrasound. Moreover, we found a dissociation in the effect size by the stimulus duty cycle (two-sided t-test, t(35) = −3.1; p = 0.004). Intriguingly, the stimulus that delivered 10 times less energy into the target (10% duty) produced an effect while the more potent 100% duty stimulus did not significantly alter the animal’s behavior (Figure 2A). This corroborates the notion that the duty cycle constitutes a critical variable in the neuromodulatory effects of ultrasound (Plaksin et al., 2016; Yu et al., 2021), and the growing consensus that the effects of low-intensity ultrasound are of mechanical, as opposed to thermal, nature (Tyler et al., 2018; Kubanek, 2018; Naor et al., 2016; Fomenko et al., 2020).

We delivered the ultrasound into the deep brain targets repeatedly, over 37 sessions. Figure 2 shows the subject’s accuracy across each session. The positive slope (r = 0.27; F(28) = 10.1; p = 0.004) demonstrates the subject’s improvement in task performance over time. Near the end of the study (day 96), we increased the difficulty of the task. Following this change the slope was insignificant (r = 0.27; F(5) = 1.57; p = 0.24). Thus, repeated daily sonication of the deep brain targets did not prevent the subject from improving their discrimination performance over time.

**Reproducible transducer positioning and validation of coupling**

Repeated delivery of ultrasound into the brain rests on reproducible positioning of the device and on the quality coupling of the transducer face to the subject’s skin. To address these critical aspects of ultrasound delivery, we equipped the platform with imaging functionality that ensures reproducible positioning with respect to the skull and high-quality coupling (Figure 3). Specifically, the device has a receive capability that also implements pulse-echo imaging. The ultrasound images acquired during each session are compared...
with a standard taken during an MRI imaging session, which validated the targeting. In addition, the magnitude of the received echoes (Figure 3, bottom) provides information on the coupling quality. The resulting ultrasound image allows the operator to confirm repeatable and accurate placement of the array relative to the skull in each session but the image should not be interpreted as a visualization of skull thickness or brain anatomy. The measured variation in distance averaged across all 256 elements and 36 sessions (the coupling check failed to run in one session) was 0.39 ± 0.77 mm. The variation in echo magnitude was 11 ± 5.5%.

Flexible deep brain targeting

Targets in the visual system, such as the LGN, will enable researchers to characterize the effects of ultrasound on neural tissue but future diagnostic and therapeutic protocols will require the ability to target systems involved in emotional regulation, such as the limbic system. Thus, we further validated our setup by demonstrating successful sonication of the LGN and amygdala in a second NHP subject (Figure 4).

DISCUSSION

We developed and validated a practical and MRI-compatible system that delivers ultrasound through the intact skull and skin into one or more neural targets in awake subjects. Remus unleashes the full potential of ultrasound: personalized, targeted, and noninvasive interventions deep in the brain (Figure 5).

Remus provides this intersection of features as a portable device. On the other hand, emerging systems based on single-element transducers are incapable of electronic targeting and have a limited axial resolution. Surgical systems (ex: ExAblate Neuro, Insightec (Ghanouni et al., 2015)) can deliver ultrasound into deep brain targets electronically, but the system was not designed for repeated, systematic applications. Moreover, in comparison with these systems, Remus provides imaging capabilities that validate the device’s position with respect to the head and test the quality of the ultrasound coupling.

The programmatic targeting enables clinicians to flexibly target multiple neural structures in a single session and the geometric design enables flexible mechanical placement of the array, resulting in a large treatable volume within the brain. This large treatment envelope, combined with the ease of targeting multiple nuclei in a single setting, provides the capacity to identify specific brain regions involved in an individual patient’s symptoms. For example, a physician can target multiple deep brain structures in a patient presenting with chronic pain. A significant change in the patient’s pain score would establish a causal
relationship between the targeted region and the patient’s pain, thus enabling the physician to replace systemic treatment (such as opioids) with a more effective and targeted therapy.

The dissociation between neuromodulatory effects and duty cycle (Figure 2A) provide additional evidence of ultrasonic modulation of neural structures (Naor et al., 2016; Tyler et al., 2018). In addition to these effects, ultrasound can elicit auditory or vestibular confounds (Guo et al., 2018; Sato et al., 2018). Remus’ fixed positioning and its capacity to programmatically deliver distinct ultrasound waveforms into distinct regions, with dissociated effects, enables scientists to control for such confounds.

Future applications

The manipulation of circuit function and dysfunction in the short term enables systematic and personalized diagnoses of the neural sources of brain disorders in the clinic. In the lab, transient manipulation of circuit function enables causal investigations of the function of specific deep brain regions in humans. Moreover, the system will enable researchers to deliver stimuli of durations necessary to induce circuit reset (Verhagen et al., 2019; Folloni et al., 2019; Bongioanni et al., 2021; Oh et al., 2019) into the malfunctioning regions.

Remus provides maximal pressure output over 5 MPa, well above the pressures needed to release drugs into a specified neural circuit during awake behavior. The awake setup enables investigators to quantify the effectiveness and duration of the neuromodulatory effects induced by specific neuromodulatory drugs, such as propofol (Wang et al., 2018b) or ketamine. In addition, the behavioral readout can be used to assess the safety of the release at the functional level (Figure 2B).

Analogous protocols can be used to assess the effectiveness and safety of cargo delivery across the blood-brain barrier (BBB) (Abrahao et al., 2019). To maximize therapeutic effects, agents, such as chemotherapy, need to be delivered across the BBB repeatedly over multiple sessions. This may raise safety concerns (Kovacs et al., 2017). Remus can be used for the systematic investigation of the effects of repeated ultrasound-based BBB opening.
Together, Remus affords multi-focal ultrasound-based brain intervention at high spatiotemporal resolution. The system’s flexible targeting and its validation of positioning and coupling open new possibilities for a large number of patients currently without options.

**Limitations of the study**

The study was designed to validate Remus’ capacity to reliably target deep brain structures during awake behavior. Behavioral modulation in one subject validates this capacity. With this validation, future studies will use Remus to measure the neuromodulatory effects of ultrasound in multiple subjects, including the effects of longer and higher intensity sonications.

---

**Figure 5. Applications of the platform for treatments of medication-resistant disorders**

Brief, millisecond-second ultrasound stimuli modulate neural activity in a transient fashion. This effect, when coupled with successive targeting of individual circuit candidates, provides a tool to dissect circuit function and dysfunction (top left). Ultrasound delivered into a target for minutes induces neuroplastic changes in the target. This could be used for a durable reset of the malfunctioning circuits (top right). To further increase the specificity of these effects, ultrasound can be combined with drug-carrying nanoparticles that release their cargo specifically at the ultrasound target (bottom left). If agents that do not naturally cross the blood-brain barrier (BBB) are to be delivered into a target, ultrasound can be combined with microbubbles that—upon sonication—transiently permeate the BBB (bottom right).

Together, Remus affords multi-focal ultrasound-based brain intervention at high spatiotemporal resolution. The system’s flexible targeting and its validation of positioning and coupling open new possibilities for a large number of patients currently without options.
In situ intensity estimates based on MR thermometry require several assumptions about brain tissue properties and do not account for spatial and temporal averaging inherent in MR thermometry measurements. Thus, there is considerable uncertainty in these estimates.

**STAR+METHODS**

Detailed methods are provided in the online version of this paper and include the following:

- KEY RESOURCES TABLE
- RESOURCE AVAILABILITY
  - Lead contact
  - Materials availability
  - Data and code availability
- EXPERIMENTAL MODEL AND SUBJECT DETAILS
  - Nonhuman primates
- METHOD DETAILS
  - System overview
  - MR validation
  - Targeting
  - Cost
  - NHP platform positioning and head fixation
  - MR imaging
  - Acoustic intensity at Target
  - Stimulation parameters
  - Task
  - Software
- QUANTIFICATION AND STATISTICAL ANALYSIS

**SUPPLEMENTAL INFORMATION**

Supplemental information can be found online at https://doi.org/10.1016/j.isci.2022.105251.

**ACKNOWLEDGMENTS**

This development of the platform and its application in NHPs was supported by the National Institute of Neurological Disorders and Stroke, Grants 1R00NS100986 and F32MH123019. We thank Tyler Davis, Eric Burdett, and Caroline Garrett for facilitating this work.

**AUTHOR CONTRIBUTIONS**

TDW and JK designed the system. TDW assembled and validated it. MGW and TDW trained the animal and collected the data. TDW analyzed the data. HO performed the MR imaging. TDW wrote a draft and all authors edited it.

**DECLARATION OF INTERESTS**

The authors declare no competing interests.

Received: December 20, 2021
Revised: May 17, 2022
Accepted: September 27, 2022
Published: November 18, 2022

**REFERENCES**

Abrahao, A., Meng, Y., Llinas, M., Huang, Y., Hamari, C., Manprize, T., Aubert, J., Heyn, C., Black, S.E., Hynynen, K., et al. (2019). First-in-human trial of blood–brain barrier opening in amyotrophic lateral sclerosis using MR-guided focused ultrasound. Nat. Commun. 10, 4373-4379.

Ahrensbrak, R., Bose, J., Hedden, S., Lipari, R., and Park-Lee, E. (2017). Key Substance Use and Mental Health Indicators in the United States: Results from the 2016 National Survey on Drug Use and Health (Center for Behavioral Health Statistics and Quality, Substance Abuse and Mental Health Services Administration).

Al-Harbi, K.S. (2012). Treatment-resistant depression: therapeutic trends, challenges, and future directions. Patient Prefer. Adherence 6, 369–388.

Bongioanni, A., Folloni, D., Verhagen, L., Sallet, J., Klein-Flugge, M.C., and Rushworth, M.F.S. (2021). Activation and disruption of a
neural mechanism for novel choice in monkeys. Nature 291, 270–274.

Braun, U., Schaefer, A., Betzel, R.F., Tost, H., Meyer-Lindenberg, A., and Bassett, D.S. (2018). From maps to multi-dimensional network mechanisms of mental disorders. Neurom 97, 14–31.

Bystritsky, A. (2006). Treatment-resistant anxiety disorders. Mol. Psychiatr. 11, 805–814.

Chaplin, V., Phipps, M.A., and Caskey, C.F. (2018). A random phased-array for MR-guided transcranial ultrasound neuromodulation in non-human primates. Phys. Med. Biol. 63, 105016.

Deffieux, T., Younan, Y., Wattiez, N., Tanter, M., Poquet, F., and Aubry, J.F. (2013). Low-intensity focused ultrasound modulates monkey visuomotor behavior. Curr. Biol. 23, 2430–2433.

 Elias, W.J., Lipsman, N., Ondo, W.G., Ghanouni, P., Kim, Y.G., Lee, W., Schwartz, M., Hynynen, K., Lozano, A.M., Shah, B.B., et al. (2016). A randomized trial of focused ultrasound thalamotomy for essential tremor. N. Engl. J. Med. 375, 730–739.

Ferguson, J.M. (2001). Ssri antidepressant medications: adverse effects and tolerability. Prim. Care Companion J. Clin. Psychi 3, 22–27.

Folloni, D., Verhagen, L., Marx, R.B., Fouragnan, E., Constats, C., Aubry, J.-F., Rushworth, M.F.S., and Sallet, J. (2019). Manipulation of subcortical and deep cortical activity in the primate brain using transcranial focused ultrasound stimulation. Neuron 101, 1109–1116.e5.

Fomenko, A., Chen, K.-H.S., Nankoo, J.-F., Saravanamuthu, J., Wang, Y., El-Baba, M., Xia, X., Seeral, S.S., Hynynen, K., Lozano, A.M., and Chen, R. (2020). Systematic examination of low-intensity ultrasound parameters on human motor cortex excitability and behavior. Elife 9, e54499.

Fry, W.J. (1958). Use of intense ultrasound in neurological research. Am. J. Physiol. Med. 37, 143–147.

Ghanouni, P., Paul, K.B., Elias, W.J., Henderson, J., Sheehan, J., Monteith, S., and Wintemark, M. (2015). Transcranial MRI-guided focused ultrasound: a review of the technologic and neurologic applications. AJR Am. J. Roentgenol. 205, 150–159.

Guo, H., Hamilton, M., Offutt, S.J., Gloeckner, C.D., Li, T., Kim, Y., Legon, W., Alford, J.K., and Lim, H.H. (2018). Ultrasound produces extensive brain activation via a cochlear pathway. Neuron 98, 1020–1030.e4.

Insel, T.R., and Cuthbert, B.N. (2015). Brain disorders? Precisely. Science 348, 499–500.

Kareski, S.C. (2007). Seizure medications and their side effects. Neurology 69, E27–E29.

Kovacs, Z.I., Kim, S., Jikaria, N., Qureshi, F., Milo, B., Lewis, B.K., Bresler, M., Burks, S.R., and Frank, J.A. (2017). Disrupting the blood-brain barrier by focused ultrasound induces sterile inflammation. Proc. Natl. Acad. Sci. USA 114, E75–E84.

Kubanek, J. (2018). Neuromodulation with transcranial focused ultrasound. Neurosurg. Focus 44, E14.

Kubanek, J., Brown, J., Ye, P., Pauly, K.B., Moore, T., and Newsome, W. (2020). Remote, brain region-specific control of choice behavior with ultrasonic waves. Sci. Adv. 6, eaaz4193.

Kubanek, J., Li, J.M., and Snyder, L.H. (2015). Motor role of parietal cortex in a monkey model of hemispatial neglect. Proc. Natl. Acad. Sci. USA 112, E2067–E2072.

Lancet (2017). Life, death, and disability in 2016. Lancet 390, 1083–1423.

Lee-Banks, H., O’Reilly, M.A., Hamani, C., and Hynynen, K. (2020). Localized anesthesia of a specific brain region using ultrasound-responsive barbiturate nanodroplets. Theranostics 10, 2849–2858.

Lee, W., Kim, H., Jung, Y., Song, I.-u., Chung, Y.A., and Yoo, S.-s. (2015). Image-guided transcranial focused ultrasound Stimulates human primary somatosensory cortex. Sci. Rep. 5, 6743.

Lee, W., Lee, S.D., Park, M.Y., Yang, J., and Yoo, S.-S. (2014). Evaluation of polyvinyl alcohol cryogel as an acoustic coupling medium for low-intensity ultrasound transcranial focal therapy. J. Clin. Neurosci. 21, 322–329.

Legon, W., Sato, T.F., Opitz, A., Mueller, J., Barbour, A., Williams, A., and Tyler, W.J. (2014). Transcranial focused ultrasound modulates the activity of primary somatosensory cortex in humans. Nat. Neurosci. 17, 322–329.

Liu, H.-L., Jan, C.-K., Chu, P.-C., Hong, J.-C., Lee, P.-Y., Huang, C.-C., Chen, P.-Y., and Wei, K.-C. (2014). Design and experimental evaluation of a 256-channel dual-frequency ultrasound phased-array system for transcranial blood-brain barrier opening and brain drug delivery. IEEE Trans. Biomed. Eng. 61, 1350–1360.

Naor, O., Krupa, S., and Shoham, S. (2016). Ultrasonic neuromodulation. J. Neuronal. Eng. 13, 031003.

Oh, S.-J., Lee, J.M., Kim, H.-B., Lee, J., Han, S., Bae, J.Y., Hong, G.-S., Koh, W., Kwon, J., Hwang, E.-S., et al. (2019). Ultrasonic neuromodulation via astrocytic trpa1. Curr. Biol. 29, 3386–3401.e8.

Peirs, C., and Seal, R.P. (2016). Neural circuits for pain: recent advances and current views. Science 354, 578–584.

Pennies, H.H. (1948). Analysis of tissue and arterial blood temperatures in the resting human forearm. J. Appl. Physiol. 1, 93–122.

Plaksin, M., Kimmel, E., and Shoham, S. (2016). Cell-Type-selective effects of intramembrane vesicle cavitation as a unifying theoretical framework for ultrasonic neuromodulation. eNeuro 3, ENEURO.0136, 15.2016.

Price, J.L., and Drevets, W.C. (2012). Neural circuits underlying the pathophysiology of mood disorders. Trends Cognit. Sci. 16, 61–71.

Rapport, N., Nam, K.H., Gupta, R., Gao, Z., Mohan, P., Payne, A., Todd, N., Liu, X., Kim, T., Shea, J., et al. (2011). Ultrasound-mediated tumor imaging and nanotherapy using drug loaded, block copolymer stabilized perfluorcarbon nanoemulsions. J. Contr. Release 153, 4–15.

Sato, T., Shapiro, M.G., and Tsao, D.Y. (2018). Ultrasonic neuromodulation Causes Widespread cortical activation via an indirect auditory mechanism. Neuron 98, 1031–1041.e5.

Szabo, T.L. (2004). Diagnostic Ultrasound Imaging: Inside Out (Academic Press).

Tufali, Y., Matyushov, A., Baldwin, N., Tauchmann, M.L., Georges, J., Yoshihiro, A., Timley, S.H., and Tyler, W.J. (2010). Transcranial pulsed ultrasound Stimulates intact brain circuits. Neuro 66, 681–694.

Tyler, W.J., Lani, S.W., and Hwang, G.M. (2018). Ultrasonic modulation of neural circuit activity. Curr. Opin. Neurobiol. 50, 222–231.

Verhagen, L., Gallego, C., Folloni, D., Constats, C., Jensen, D.E., Ahnine, H., Roumazieles, L., Santin, M., Ahmed, B., Lehericy, S., et al. (2019). Offline impact of transcranial focused ultrasound on cortical activation in primates. Elife 8, 40541–e40628.

Wang, J.B., Aryal, M., Zhong, Q., Vyas, D.B., and Aaran, R.D. (2018a). Noninvasive ultrasonic drug uncaging maps whole-brain functional networks. Neuro 100, 728–738.e7.

Wang, J.B., Aryal, M., Zhong, Q., Vyas, D.B., and Aaran, R.D. (2018b). Noninvasive ultrasonic drug uncaging maps whole-brain functional networks. Neuro 100, 728–738.e7.

Yu, K., Niu, X., Krook-Magnuson, E., and He, B. (2021). Intrinsic functional neuron-type selectivity of transcranial focused ultrasound neuromodulation. Nat. Commun. 12, 2519.

Zesiewicz, T.A., Chari, A., Jahan, I., Miller, A.M., and Sullivan, K.L. (2010). Overview of essential tremor. Neuropsychiatric. Dis. Treat. 6, 401–408.
STAR METHODS

KEY RESOURCES TABLE

| REAGENT or RESOURCE | SOURCE | IDENTIFIER |
|---------------------|--------|------------|
| Experimental models. Organisms/strains | Oregon Health and Science University | 35079, 35077 |
| Macaca Mulatta | Oregon Health and Science University | 35079, 35077 |
| Software and algorithms | Custom MATLAB scripts | 886115E |
| MR thermometry and targeting software | | |
| Other | |
| 256 Element Transducer | Doppler Electronic Technologies | |
| Vantage 256 driving hardware | Verasonics | Vantage 256 |
| Head Frame | Custom Solidworks Design | B32A6E4 |
| poly(vinyl alcohol) | Sigma-Aldrich | 363065 |

RESOURCE AVAILABILITY

Lead contact
Further information and requests for resources and reagents should be directed to and will be fulfilled by the lead contact, Jan Kubanek (jan.kubanek@utah.edu).

Materials availability
1. The transducer (HP-500K-256-BE) is available from Doppler Electronic Technologies (Guangzhou, China).
2. The Vantage 256 Driving system is available from Verasonics (Kirkland Washington).
3. The coupling material is made with 6% by weight poly(vinyl alcohol) in water (Lee et al., 2014). The poly(vinyl alcohol) is available at Sigma-Aldrich, product number 363065.
4. The titanium pins used to attach the frame to the subject’s head were custom made by Gray Matter Research (Bozeman, Mt).

Data and code availability
1. The software, which includes the element locations for the 256 element transducer, is available online at onetarget.us/software.
2. MR Imaging data and the subject’s behavioral data is available upon request to the lead contact (Jan Kubanek: jan.kubanek@utah.edu).
3. Any additional information required to reanalyze the data reported in this paper is available from the lead contact upon request.

EXPERIMENTAL MODEL AND SUBJECT DETAILS

Nonhuman primates
Two male macaque monkeys (macaca mulatta) participated in this study. One subject (age 6 years, weight 8 kg) participated in both MR based targeting validation and behavioral studies. A second subject (age 6 years, weight 12 kg) participated only in MR based target validation. The procedures were approved by the Institutional Animal Care and Use Committee of the University of Utah.

METHOD DETAILS

System overview
An overview of the system is given in Figure S1. Custom targeting software interfaces with the MR scanner for target validation and with the ultrasound driving system to deliver ultrasound to the desired location.
Thus, Remus delivers ultrasound into specific deep brain targets, enabling real-time modulation of awake behavior. Items in the green boxes are provided as part of this study at onetarget.us/software.

**MR validation**

A series of custom MATLAB functions allows the user to register the array to an MR image, select an acoustic focus within the image, and deliver a sonication to the desired anatomical location. The resulting thermometry images are overlaid on the anatomical image to verify accurate placement of the acoustic focus.

**Targeting**

Custom targeting software interfaces with the ultrasound driving system (Verasonics, Kirkland Washington) to measure the distance between the array and the skull, measure the quality of the acoustic coupling, and to provide the individual element phases necessary to place the acoustic focus at the desired location. Phases are determined using a time of flight algorithm (Szabo, 2004). The targeting software accepts user inputs through a serial connection to enable a user to specify target locations and to change those locations during individual sessions.

**Cost**

The cost of Remus is dictated by the costs of the driving system and the transducer array. Bulk manufacturing of the complete system is currently estimated to be around $50,000.

**NHP platform positioning and head fixation**

For the NHP experiments shown here, we developed an apparatus that ensures reproducible positioning of the platform with respect to the head from session to session, and at the same time allows us to head-fix NHPs and engage them in behavioral tasks. To achieve that, we developed a custom head frame that is attached to the skull via four titanium pins (Gray Matter Research, Bozeman, MT). Each pin is attached to the skull using three to four screws. The frame for the NHP experiments was designed in 3D cad software and produced with a 3D printer. A 3D drawing of the frame is available at onetarget.us/software. The frame is attached to a primate chair using two steel bars, mounted into the left and right side of the frame. After the frame is attached to the chair, the transducer is mounted to the frame via four screws (one on each corner). Coupling of the transducer to the skull is achieved using a 6% polyvinyl alcohol (PVA) gel (Lee et al., 2014) that is compressed between the subject’s head and the transducer. The subject’s hair is shaved prior to each session. Shaving may not be necessary in the ultimate human applications; the platform could detect the quality of the coupling and aberration due to the hair, and adjust the emitted ultrasound amplitude accordingly.

In each session, the system measures the accuracy of the device placement with respect to the head and the quality of the acoustic coupling using ultrasound imaging. Specifically, the system measures the distance between the transducer and the skull at six distinct locations. A pulse-echo measurement, performed using individual elements, provides an estimate of the location of the skull relative to the transducer. The distance is measured by detecting the front edge of the reflected ultrasound energy and assuming a speed of sound of 1500 m/s in the coupling gel. The average difference in position relative to the position measured during MR thermometry was 0.39 ± 0.77 mm. The total power returned to each element (Figure 3, bottom) was −11 ± 5.5% lower across sessions compared to the power measured during the MR thermometry session.

**MR imaging**

The system is fully MRI-compatible. Accurate targeting of specific deep brain regions can therefore be validated using MRI, such as MR thermometry or MR ARFI. We used MR thermometry. In this approach, a continuous, 5-second sonication is sufficient to increase temperature at the focal spot by about 2°C, enabling visualization of the focus without inducing long-term changes in the neural tissue.

All scans were performed using a 3T MRI scanner (Trio, Siemens Medical Solutions, Erlangen, Germany) with a 4-channel flex coil wrapped underneath the animal’s head. High resolution 3D T1-weighted images were used for anatomical imaging and target identification; Magnetization Prepared Rapid Acquisition Gradient Echo (MPRAGE), TR = 1900 ms, TE = 3.39 ms, TI-900 ms, BW = 180 Hz/pixel, flip angle 9°, FOV = 192 × 192 × 120 mm, resolution 1 × 1 × 1 mm, acquisition time 5:12 min. MR thermometry was performed with a 3D gradient recalled segmented echo planar imaging pulse sequence; TR = 24 ms,
TE = 11 ms, BW = 592 Hz/pixel, flip angle 12°, echo train length = 5 with monopolar readout, FOV = 144 × 117 × 36 mm, resolution 1.5 × 1.5 × 3.0 mm, acquisition time 4.6 s per dynamic image.

During thermometry, the subject was sedated and placed in a sphinx position. A custom 3D printed stand was mounted to the head frame by four screws located at the four corners of the frame. The stand held the subject’s head in position and provided space for the placement of the flex coil (3D drawings available at onetarget.us/software). After the subject was in place the transducer was mounted to the frame and coupled to the head using the same process used during awake procedures.

Registration of ultrasound and MR space was performed using three fiducial markers. The fiducials were placed in cavities with known locations that were designed into the printed frame. After registration, the average euclidean distance between the estimated and the actual location of the three fiducials (as measured by the 3-dimensional center of mass) was 2.5 mm. The transducer caused distortion along the subject’s inferior/superior axis resulting in approximately 4 mm of error in the transducers registration in that dimension. The registration of the transducer to the subject’s anatomy guided initial steering of the acoustic beam. Once the first thermometry signal was acquired, adjustments to the focal location were based primarily on the location of the thermometry signal, thus correcting any errors introduced by poor transducer localization.

It is well established that preferential absorption of ultrasound in the skull can lead to undesirable off-target heating. Indeed, clinical ablation trials rely on both a large transducer aperture and constant cooling of the scalp to mitigate this risk (Ghanouni et al., 2015). With a smaller transducer aperture and in the absence of active cooling investigators must assume that the temperature rise in the skull could be higher than at focus. Care should thus be taken to monitor the health of the skull and scalp between the transducer and the target when MR thermometry is used to validate acoustic targeting.

At the short durations and low frequencies used for neuromodulation in this study, heating of the skull was not a concern. The subject, who was awake, showed no signs of discomfort in response to the ultrasound.

**Acoustic intensity at Target**

While the energy delivered during MR thermometry is much greater than the energy used to achieve neuromodulation, we can scale the thermometry result to estimate the acoustic intensity at target during neuromodulation. First, using Pennes’ bioheat equation (Pennes, 1948), together with the measured temperature rise, we estimate the intensity during MR thermometry. We then scale this result by the voltage delivered by the neuromodulatory sonications to estimate the intensity delivered during neuromodulation. Assuming negligible conduction and perfusion, the acoustic intensity during MR thermometry, \( I_{\text{MRI}} \), is related to the temperature rise, \( \Delta T \), by

\[
I_{\text{MRI}} = \frac{\rho C \Delta T}{\Delta t_a}
\]

(Equation 1)

where \( \rho \), \( C \), and \( a \) are the density, specific heat capacity, and acoustic absorption of the tissue and \( \Delta t \) is the time in which the temperature increase occurs. To minimize the effects of conduction and perfusion \( \Delta T \) is chosen to be the maximum temperature measured in the first dynamic in which the ultrasound is on. The center of k-space is acquired 2.3 seconds into each acquisition. Thus, \( \Delta t \) was set to 2.3 s. We assumed a density of 1046 kg/m³, a specific heat capacity of 3630 J/K, and an acoustic absorption of 3.9 Np/m. The intensity during neuromodulation, \( I_n \), is then

\[
I_n = I_{\text{MRI}} \frac{V_n^2}{V_{\text{MRI}}^2}
\]

(Equation 2)

where \( V_n \) and \( V_{\text{MRI}} \) are the voltages applied to the transducer during neuromodulation and MR thermometry (15 Vp and 20 Vp).

Similarly, the MR thermometry can be used to estimate the maximum temperature rise during each neuromodulatory sonication. The temperature rise during the first 2.3 s of each thermometry pulse was 1.4 and 1.8 C in the left and right LGN respectively. The 10% duty cycle sonication delivered 0.73% of the energy delivered in the first 2.3 seconds of the thermometry sonication. Thus, we can estimate a temperature rise
of 0.010 and 0.013 °C in the left and right LGN. A continuous wave sonication delivers 10 times the energy, thus, the expected temperature rise for these sonications is 0.10 and 0.13 °C.

**Stimulation parameters**

Our custom, 256 element transducer (Doppler Electronic Technologies, Guangzhou, China) can produce focal pressures greater than 3 MPa at its 650 kHz center frequency and a usable frequency range of 480-850 kHz. The transducer geometry is semicircular, with a radius of curvature of 65 mm (see Figure 1). The elements are 4 × 4 mm square and the distance between elements is 4.2 mm. All stimuli used 650 kHz. The pulsed stimulus (10% duty) used a 500 Hz pulse repetition frequency. Both pulses used a square envelope. The peak focal pressure (estimated from the thermometry data) was 1.1 and 1.0 MPa in the right and left LGN respectively. The half power beam width measured by a hydrophone in a water tank for was 1, 3.75, and 3.75 mm in the left/right, anterior/posterior, and superior/inferior dimensions, respectively. The half power beam width measured by thermometry was 0.75 and 3.0 mm in the left/right dimension and 3.75 and 5.25 mm in the front/back dimension for the left and right LGN respectively. Using the same input voltage, the free field pressure—measured using a hydrophone—is 3 MPa at the location of the left and right LGN. Thus, the MR thermometry measurements suggest that about 30% of the pressure reaches the target through the individual layers. It is worth noting, nonetheless, that this estimate likely underestimates the actual pressure at target. In particular, the temperature measured by the MR is averaged spatially (across a voxel) and temporally (across the acquisition time); the actual peak temperature is thus higher than the measured average. In addition, a portion of the energy is distributed to the thermal conduction and vascular convection.

**Task**

We trained one NHP to perform a visual choice task. This task was used in many previous studies (e.g., (Kubanek et al., 2015, 2020)). Briefly, the subject was seated in a comfortable primate chair and was presented with visual stimuli shown on a monitor. The subject's eye movements were tracked and recorded using an infrared eye tracker (Eyelink, SR Research, Ottawa, Canada). In the task, the subject first fixates a central target. Following its offset, one target appears on the left and one target on the right side of the screen. There is a brief, controllable delay between the onset of the two targets, which can range from −60 to 60 ms. We varied the location of the targets within 2.5 visual degrees to circumvent adaptation. The sonication of the left and right LGN was randomly interspersed with trials in which no ultrasound was delivered. In an ultrasound trial, a 300 ms stimulus was delivered 150 ms before the fixation point offset. The subject receives a liquid reward if he looks at the target that appeared first within 2 s. In the key condition in which both targets appear at the same time and during which we quantify the effect of ultrasound, the subject is rewarded for either choice. The subject generally worked daily throughout the work week.

**Software**

We designed custom software to integrate ultrasound sonication with both the MR thermometry and the behavioral task. The software can register the transducer to an MR coordinate system, output the targeted focus in both transducer and MR coordinates, and overlay the temperature change (as measured by MR thermometry) on the anatomy image. The overlay of the MR thermometry and MR anatomy image is achieved by interpolating the thermometry image to the coordinates of the anatomy image. The coordinate systems are read from the image file headers (DICOM format) returned by the scanner. To protect the subject's scalp, skull, and neural tissue we used very small temperature increases to visualize the focal spot. Thus, to improve the visualization, we included a noise reduction algorithm in our visualization software. The algorithm assumes an exponential increase in temperature during the sonication and an exponential decay in temperature after the ultrasound turns off. Voxels in which the temperature does not follow such a trend are ignored.

Similarly, we developed software that integrates a behavioral task with flexible sonication of the targeted neural circuit. The ultrasound controller accepts target coordinates and intensities from the server running the behavioral task. Synchronization of the sonication with presentation of the visual stimulus is achieved with a TTL pulse. All of the ultrasound software was developed within the Verasonics framework and is available at onetarget.us/software.
QUANTIFICATION AND STATISTICAL ANALYSIS

Behavioral changes resulting from each set of stimuli were measured using a two-sided t-test (caption of Figure 2A, and results section; Figure 2A shows mean and standard error of the mean). Behavioral changes across the two stimuli were measured with a paired two-sided t-test (asterisks and caption of Figure 2A, and results section). The individual datapoints constituted the individual sessions, with 12 sessions collected for the 10% duty stimulation and 25 sessions for the 100% duty stimulation. Significance was defined as a p-value less than 0.05. P-values less than 0.05 were marked with one asterisk while p-values less than 0.01 were marked with two asterisks. A linear fit to the subject’s accuracy over time was characterized by the R-squared value and the F-statistic measured against a constant model (Caption to Figure 2B and results section). The analyses have included 27 sessions.