Genetic Variation for Nitrogen Use Efficiency Traits in Global Diversity Panel and Parents of Mapping Populations in Pearl Millet

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Nitrogen (N) is one of the primary macronutrients required for crop growth and yield. This nutrient is especially limiting in the dry and low fertility soils where pearl millet \([\textit{Pennisetum glaucum} (L.) R. Br]\) is typically grown. Globally, pearl millet is the sixth most important cereal grown by subsistence farmers in the arid and semi-arid regions of sub-Saharan Africa and the Indian subcontinent. Most of these agro-ecologies have low N in the root zone soil strata. Therefore, there is an immense need to identify lines that use nitrogen efficiently. A set of 380 diverse pearl millet lines consisting of a global diversity panel (345), parents of mapping populations (20), and standard checks (15) were evaluated in an alpha-lattice design with two replications, 25 blocks, a three-row plot for 11 nitrogen use efficiency (NUE) related traits across three growing seasons (Summer 2017, Rainy 2017, and Summer 2018) in an N-depleted precision field under three different N levels (0%-N\(_0\), 50%-N\(_{50}\), 100%-N\(_{100}\) of recommended N, i.e., 100 kg ha\(^{-1}\)). Analysis of variance revealed significant genetic variation for NUE-related traits across treatments and seasons. Nitrogen in limited condition (N\(_0\)) resulted in a 27.6 and 17.6% reduction in grain yield (GY) and dry stover yield (DSY) compared to N\(_{50}\). Higher reduction in GY and DSY traits by 24.6 and 23.6% were observed under N\(_0\) compared to N\(_{100}\). Among the assessed traits, GY exhibited significant positive correlations with nitrogen utilization efficiency (NUtE) and nitrogen harvest index (NHI). This indicated the pivotal role of N remobilization to the grain in enhancing yield levels. Top 25 N-insensitive (NIS-top grain yielders) and N-sensitive (NS-poor grain yielders) genotypes were identified under low N conditions. Out of 25 NIS lines, nine genotypes (IP 10820, IP 17720, ICMB 01222-P1, IP 10379, ICMB 89111-P2, IP 8069, ICMB 90111-P2, ICMV IS89305, and ICMV 221) were common with the top 25 lines for N\(_{100}\) level showing the genotype plasticity toward varying N levels. Low N tolerant genotypes identified from the current investigation may help in the identification of genomic regions responsible for NUE and its deployment in pearl millet breeding programs through marker-assisted selection (MAS).

Keywords: pearl millet, nitrogen use efficiency, genotypic variations, phenotyping, nitrogen-responsive
INTRODUCTION

Global nitrogen (N) demand, one of the most expensive farm inputs currently stands at about 117 million metric tons with a projected annual increase of ~1.5% in the future (FAO, 2019). Indian agriculture consumes over 17 million tons of N fertilizer per year. However, plants can utilize only 30–40% of the applied N for food production and the remaining (up to 60%) is lost to the environment by leaching, de-nitrification, and runoff. Surplus nitrogen pollutes freshwater streams and air which is hazardous to the majority of living species (Hickman et al., 2014; Russo et al., 2017).

Besides, excess usage of N fertilizer not only decreases the efficiency of nutrient use but also affects the rate of economic returns per unit of chemical fertilizer applied. The effect of negative environmental and economic impacts could be reduced through better agronomic practices and also by utilizing N efficient lines with improved nitrogen use efficiency (NUE) (Raun et al., 1999). Hence, improving the NUE of crop plants could help in reducing fertilizer input, increased productivity and profitability coupled with a reduced negative impact on the environment.

Globally, pearl millet (Pennisetum glaucum (L.) R. Br) widely grown for food and fodder is considered as the sixth most important cereal crop in terms of area. It is one of the oldest cultivated cereal crops, originated in Africa but later spread to many countries (D’Andrea and Casey, 2002; Manning et al., 2011). This millet crop is vital for food and nutritional security for the world’s poorest people living in different agro-ecological zones. Besides, pearl millet has a rich nutritional profile, wide genetic diversity, high photosynthetic rates being a C4 and also provides a healthy balanced diet, thus contributes to the economic security of poor farmers (Srivastava et al., 2019). In the Indian agriculture scenario, pearl millet is grown by poor and marginal farmers under low fertile and rainfall dependent areas often facing drought during different stages of crop growth. From the past few years, pearl millet demand has been increased for its nutritional characteristics and its adaptability to a wide range of climatic conditions (Tako et al., 2015). Since then farmers started applying a high dose of N fertilizer for maximizing grain yield (GY), but excessive usage of N fertilizer leads to low NUE of the crop along with various environmental hazards. In some parts of India, poor farmers lack knowledge about the ideal dose of fertilizer to harvest the real yield potential. Hence, yield improvement of pearl millet under low nitrogen input is indeed beneficial for economic and environmentally sustainable cultivation.

Nitrogen use efficiency is a complex trait, which is associated with various morphological, physiological, molecular, and biochemical changes in plants throughout the life cycle. For a clear understanding of this complex nature, studies of various physiological traits and their close correlation with one or more economically important traits like GY is foremost critical. Nevertheless, it will help in selecting low N tolerant/high yielding lines at different N conditions (Monostori et al., 2016). In general, plant function is always associated with chlorophyll content, which directly indicates the N status of the leaf (Yang et al., 2014). Leaf chlorophyll content and photosynthetic capacity are appropriate benchmarks for identifying high NUE (HNUE) genotypes under low N conditions during field trials (Vijayalakshmi et al., 2015; Kiran et al., 2016). Leaf N status was usually measured by using a hand-held optical chlorophyll meter to monitor the leaf nitrogen status and chlorophyll content. Moreover, leaf area (LA) is also one of the important physiological traits, plays a critical role in enhancing plant biomass (Gianquinto et al., 2004; Monostori et al., 2016).

Nitrogen use efficiency is defined as the ability to produce GY per unit N available in the soil. NUE mainly depends on the results of two main processes, such as N uptake efficiency (NupE), and nitrogen utilization efficiency (NutE) (Good et al., 2004; Hakeem et al., 2012; Vijayalakshmi et al., 2013). NupE is the ability of the plant to take up N from the soil and NutE is the ability to use N to produce gain yield (Ladha et al., 1998; Hirel et al., 2007). Studies on genotypic variations under low and recommended N for NUE traits at both seedling and maturity stage of plants under controlled and field conditions in various crops resulted in the identification of high NUE genotypes possessing high yield sustainability under low N condition (Muchow, 1998; Inthapanya et al., 2000; Le Gouis et al., 2000; Presterl et al., 2003; Anbessa et al., 2009; Namai et al., 2009; Vijayalakshmi et al., 2015). The high NUE/N-insensitive genotypes (NIS-top grain yielders) are defined as genotypes that give more or equal GY with minimal application of N fertilizer when compared to recommended or standard N fertilizer conditions (Hawkesford, 2017). The understanding of GY and its associated NUE traits performance is still largely lagging in the pearl millet diversity panel. However, few studies had explored genotypic variations for NUE at different N levels. In one such report, 20 diverse pearl millet genotypes and few high-yielding hybrids were screened under field conditions at two N levels (Alagarswamy and Bidinger, 1987). In another study, pearl millet hybrids were grown over 3 years in western and eastern Nebraska at four different N levels to determine the optimum N rate for cultivation and concluded that the NUE trait was less responsive than the N uptake with increased N levels (Maman et al., 2006).

Nitrogen use efficiency, which is dependent on soil or external N supply, is an output of available N uptake, its efficient utilization and remobilization to grain at end of the season. Several studies revealed that improved NUE might lead to enhanced NUE under low N conditions, particularly in cereals (Moll et al., 1982; Good et al., 2004). NUE is defined as the genotype ability to assimilate and remobilize N which ultimately convert into GY. It is an essential physiological parameter that unveiled the positive relationship with GY. Whereas in pearl millet comprehensive studies are required to pinpoint the critical factors underlying NUE under different N levels. GY is a complex trait controlled by a network of multiple traits and their associations. The GY in pearl millet is a result of many yield components, such as grain number, grain weight, tiller number, and panicle number (Rai et al., 2012; Basava et al., 2019). Hence, uncovering the genetic basis of GY, and other related NUE traits under low and high N conditions is a prerequisite to understand the mechanism and also to identify...
ideal NUE associated traits for selection. Thus, the current study was aimed to evaluate a diverse set of pearl millet lines for their response to GY and NUE related traits under different N levels and also to determine the suitable NUE traits for selection. This study provides useful information toward uncovering the physiological and genetic basis of GY and its related traits under low N which further facilitates the development of low N stress-resilient pearl millet cultivars.

MATERIALS AND METHODS

Experimental Details

The experiment was conducted at an N-depleted precision field of the International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), Patancheru, Telangana, India. The farm is geographically situated at an altitude of 545 m above mean sea level on 17.53° N latitude and 78.27° E longitude. The climate of the location is semi-arid with an average rainfall of 898 mm. The minimum and maximum temperatures ranged from 38 to 42°C which was observed during the experimental seasons. A set of 380 diverse pearl millet lines consisting of a world diversity panel, Pearl Millet Inbred Germplasm Association Panel-(PMiGAP) (Sehgal et al., 2015), mapping population parents and checks were screened during three seasons (summer 2017, rainy 2017, and summer 2018) in the same field with minimal fertility and moisture gradient. Across all the three seasons, field experiments were carried out in a split-plot alpha-lattice design with 2 m and three-row plots with two replications under three N treatments [N₀ (without additional N application), N₅₀ (@50 kg ha⁻¹), and N 100 (@100 kg ha⁻¹)]. Here N source used was urea (46.4% N). Field area with three different N treatments was considered as the main plot and it was divided into six blocks (two replications for each treatment). Each N level block was divided into 25 sub-blocks, and each sub-block consisted of 16 genotypes. Nitrogen depleted plot was developed and maintained at ICRISAT over past many seasons before the inception of the experiment by withholding N application. Before initiating the experiment, important physical and chemical properties of the soil were measured. Soil samples were collected at 15 cm depth from different places and four corners of the plot; mixed composite was used to determine the soil properties (Supplementary Table 1).

Crop Management

Crop management including land preparation, sowing/planting, management of water, nutrient, disease, and pest control were taken care of throughout the crop period. Seeds were sown on the raised furrows by using a four cone planter by maintaining sufficient spacing of 60 cm from row to row and 15 cm from plant to plant. The crop was fertilized by manual broadcasting as per the treatment. N fertilization was done as per the treatments using urea (46.5%) in two equal splits at 20–25 and 40–45 days after sowing. Besides, other nutrients such as single super phosphate @250 kg ha⁻¹ and muriate of potash (60% K₂O) @50 kg ha⁻¹ was also applied at the time of sowing. All the pearl millet genotypes were evaluated for a total of 11 traits across three consecutive growing seasons under different N regimes. The measured phenotypic traits were (1) SPAD chlorophyll content (soil plant analytical device), (2) leaf number (LN), (3) LA, (4) GY, (5) dry stover yield (DSY), (6) N percent in dry stover (NPS), (7) N uptake in grain (NUpG), (8) N uptake in stover (NUpS), (9) total nitrogen uptake (TNUp), (10) NUtE, and (11) nitrogen harvest index (NHI). The phenotypic data were averaged across three seasons in each genotype for the first five traits SPAD chlorophyll content, LN, LA, GY, and DSY. For the remaining six NUE related traits data was presented as a pooled mean of two summer seasons, due to lack of quality data in the rainy season.

Physiological Traits

SPAD Chlorophyll Content (Soil Plant Analytical Device)

Soil plant analytical device chlorophyll content was measured by using Minolta Corporation’s Chlorophyll SPAD-502 plus, United States. Plants from all the treatments were marked a day before with different color ribbons. Fully expanded uppermost leaves at 45 days’ stage (from the day of leaf emergence) were selected and an average of three reads was recorded from a total of three plants from the middle row.

Leaf Area and Leaf Number

In order to examine the LA and LN, leaf samples were collected approximately 80 days after emergence for all the treatments and replications. Healthy plant per plot was harvested from the middle row in the field in early morning and on the same day, specific LA and number were measured by passing in all the collected leaves of the plant through LA meter LI-3100C (LI-COR, United States).

Grain and Stover Yield

At the crop harvesting stage, GY was measured after threshing sundried panicles to remove the grains from their central rachis. Harvested plants were selected from the middle row and expressed in grams (g) per 2-meter area. Similarly, DSY was also measured after drying the stover samples of all the middle row plants without panicles and expressed in grams per 2-meter area.

Nitrogen Estimation and Nitrogen Indices

Nitrogen Estimation

Nitrogen content in grain and stover samples was estimated by using the sulfuric acid-selenium digestion method. Grain and stover were made into fine powder by using clone mixture (cyclone sample mill) and 0.250 gm was used for nitrogen estimation in the Charles Renard Analytical Laboratory at ICRISAT, Patancheru. Pre-weighed samples were digested with the sulfuric acid-selenium and then analyzed (Sahrawat et al., 2002) by using an auto-analyzer (Skalar SAN System, AA Breda, Netherlands). Nitrogen concentration is expressed as Nitrogen% (N%).
Different Nitrogen Use Efficiencies
Total N uptake was calculated from the sum of grain uptake and stover N uptake values. Then different N efficiencies were calculated as per the formulas given by Fageria et al. (2010).

Nitrogen utilization efficiency (NUE)
\[ \text{NUE} = \frac{\text{GY}}{\text{TNUp}} \]

Nitrogen harvest index (NHI)
\[ \text{NHI} = \frac{\text{NUpG}}{\text{TNUp}} \times 100 \]

Statistical Analysis
The phenotypic mean values over the replications of each genotype for all the traits were prepared and the best linear unbiased estimates (BLUE) for genotypes were estimated at each and across N levels using Genstat software, 20th edition. The minimum, maximum, and BLUE of each trait at each N level were calculated. ANOVA was performed for the individual and pooled seasons by modeling individual season’s residuals variance using Genstat software, 20th edition (VSN International, 2019). Fisher’s t-test was used to ascertain the significant difference among the genotypes, treatments and interactions. Pearson’s correlations coefficient analysis was performed by using pooled data of two summer seasons for all the 11 traits to identify the relationship between traits at each level of N treatments.

Cluster Analysis
Hierarchical cluster analysis was carried out using the Euclidean distance metric and UPGMA method (un-weighted paired group method and arithmetic averages)¹. By using the pooled data of three seasons treatment wise (N₀, N₅₀, and N₁₀₀) cluster analysis was performed for the five traits viz., SPAD chlorophyll content, LN, LA, GY, and DSY.

RESULTS
According to the fertilizer map, the available nitrogen (N) content of the soil sample was divided in to low, medium, and high. Therefore, low N conditions were observed when soil nitrogen content is below 280 kg/ha (critical limits of the soil maintained as per ICRISAT guide limits). The selected experimental field was not used for any cultivation from the past 5–6 years and before starting the trial, soil samples were collected across the field and undergone soil nutrient analysis. Nutrient analysis (Supplementary Table 1) revealed 0.5% organic carbon (OC), indicating a low available N in the soil and the basal N availability was common across all the N regimes before the initiation of the experiment. In addition, the soil micronutrient availability was in the medium range. The basal N availability in the soil is common for all the treatments but coming to the N₅₀ and N₁₀₀ regimes, an additional N source in the form of urea was applied as per the protocol mentioned in the methodology section. Overall, the N₀ regime was maintained strictly without any additional supply of nitrogen.

¹http://darwin.cirad.fr/darwin

A total of 380 pearl millet genotypes including the diversity panel, parents of mapping populations, and checks were screened for 11 physio-agronomic and NUE traits at three nitrogen levels (0, 50, and 100% of the recommended N doses) with two replications. Data recorded were averaged across the three seasons for each trait in each N level. Results revealed the native variation across the genotypes toward N response which had given the scope to identify nitrogen use efficient lines under low and high N conditions for the marginal and favorable ecologies, respectively. ANOVA results revealed significant variations among the genotypes, treatments and their interactions (season × treatment (N levels), season × genotype, treatment × genotypes, and season × treatment × genotype). The pooled means over three seasons in each genotype in each N level for five physio agronomic traits were provided in Supplementary Table 2. The pooled mean of over two seasons for six N related traits in each genotypes were provided in Supplementary Table 3.

Physiological Traits
Key physiological traits viz., SPAD chlorophyll content, LN, and LA were recorded for all three seasons. Significant variations across the genotypes under different N regimes were noticed. Under N₀, SPAD chlorophyll content was ranged from 28.18 to 49.11 with an average of 37.85. Whereas in N₅₀, it was ranged from 32.16 to 50.1 with an average of 40.59. In N₁₀₀, the values were ranged from 29.04 to 49.07 with an average of 41.77. In N₀, SPAD chlorophyll content values were reduced by 6.8 and 9.4% compared to N₅₀ and N₁₀₀ conditions, respectively. LN was decreased under N₀ conditions by 19.3 to 29.6% compared to N₅₀ and N₁₀₀ conditions, respectively. The range of LA was lower under N₀ compared to N₁₀₀ conditions. The mean of LA was reduced significantly under the N₀ condition compared to the N₁₀₀ condition. In addition, the analysis of variance (ANOVA) results revealed that the effect of genotype, treatments and their interactions were significant (P < 0.001) for all the three physiological traits studied. The data was averaged across the three seasons for each trait in each N level. Descriptive statistics over the three seasons, and ANOVA results for all the physiological traits were provided in Table 1.

Agronomic Traits
Grain Yield
In N₀, the average GY of the tested genotypes was 67.04 g and ranged from 9.64 g (IP13363) to 138.48 g (IP18621). Similarly, significant genotypic variations were observed in other treatments. In N₅₀, GY values varied from 20.72 g (Tift238 D1) to 168.85 g (IP16096) with a mean of 92.61 g. Whereas in N₁₀₀, GY ranged from 20.48 g (IP13363) to 164.87 g (ICMV-IS89305) with a mean of 88.69 g. Nitrogen in limited condition (N₀) resulted in a 27.6 and 24.6% reduction in GY compared to N₅₀, and N₁₀₀, respectively. The ANOVA results indicated that the effect of genotype, treatments were significant (P < 0.001) and their interactions were also significant between seasons × treatment (P < 0.001), seasons × genotypes (P < 0.001), treatment × genotype (P < 0.001), and season × treatments × genotype.
TABLE 1 | Summary statistics and ANOVA results from three seasons pooled data for physiological and agronomic traits under three N levels.

| Traits | SPAD chlorophyll content | LN | LA | GY (g/2m²) | DSY g/2m² |
|--------|--------------------------|----|----|------------|-----------|
| Range  | N₀            | 37.85 | 17.89 | 722.59 | 67.04 | 202.06 |
|        | N₀₀          | 40.59 | 22.18 | 1212.4 | 92.61 | 246.16 |
|        | N₁₀₀        | 41.77 | 25.40 | 1484.04 | 88.89 | 264.41 |
| Mean   | N₀            | 28.18 | 49.11 | 8.47–34.81 | 202.64–2220 | 9.64–159.71 | 58.47–806.09 |
|        | N₀₀          | 32.16 | 50.1 | 13.25–42.77 | 652.35–2967.8 | 20.72–236.43 | 62.42–740.78 |
|        | N₁₀₀        | 29.04–49.07 | 10.30–41.73 | 363.65–2742.06 | 20.48–242.12 | 73.92–678.87 |
| Percent reduction | N₀ compared with N₀₀ | 6.8 | 19.3 | 40.4 | 27.6 | 17.9 |
|        | N₀ compared with N₁₀₀ | 9.4 | 29.6 | 51.4 | 24.6 | 23.6 |
|        | N₀₀ compared with N₁₀₀ | 2.8 | 12.7 | 18.4 | -4.2 | 6.9 |
| F statistic | S               | 12.45*** | 345.29*** | 742.27*** | 580.24*** | 609.32*** |
|        | T              | 2.27**ns | 40.48*** | 227.09*** | 213.16*** | 55.08*** |
|        | S × T           | 0.67ns | 15.21*** | 67.86*** | 67.77*** | 94.49*** |
|        | G              | 5.94*** | 7.44*** | 8.48*** | 9.48*** | 14.97*** |
|        | S × G           | 4.65*** | 5.19*** | 7.77*** | 6.87*** | 7.54*** |
|        | T × G           | 2.49*** | 3.5*** | 3.6*** | 2.86*** | 4.01*** |
|        | S × T × G       | 2.17*** | 2.87*** | 4.3*** | 2.32*** | 2.73*** |

SPAD: Chlorophyll content; LN, leaf number; LA, Leaf area; GY, grain yield; DSY, dry stover yield; G, Genotype; S, Season; T, treatment. F statistic values with asterisk indicate significant at levels of 0.001 (***) and NS denote for non-significant.

Based on GY data, top 25 (N-insensitive [NIS-Top grain yielders]) and least 25 (N-sensitive [NS-Poor grain yielders]) genotypes were identified under N₀ conditions (Table 2). Out of 25 NIS lines, nine genotypes (IP10820, IP17720, ICMB01222-P1, IP10379, ICMB89111-P2, IP8069, ICMB90111-P2, ICMV-IS89305, and ICMV221) were common in the top 25 lines at the N₁₀₀ level which shows the genotype plasticity toward N₀ and N₁₀₀ conditions (Figure 1). Similarly, the top 25 high yielding genotypes were identified in N₁₀₀. The 25 least grain yielding genotypes [N-sensitive (NS)] in N₀ and N₁₀₀ conditions were presented in Figure 2.

Furthermore, agronomic data was provided for the top five lines viz., IP 18621, IP 10820, IP 17720, IPC804, and ICMB01222-P1, out of selected 25 NIS lines (Supplementary Table 4). Interestingly, days to 50% flowering (DF50%) and plant height (PH) was more in all the tested genotypes at N₀ indicating the genotype efficiency in terms of agronomic traits. These genotypes can be used as parents in the pearl millet breeding program to develop N efficient genotypes under low N input conditions.

Dry Stover Yield

In N₀, DSY ranged from 58.47 g (LGD-1-B-10) to 806.19 g (IP8863) with a mean of 256.3 g. Whereas in N₀₀, values ranged from 62.42 g (IP13363) to 740.78 g (IP11584) with a mean of 246.16 g. In N₁₀₀, the average DW of 380 genotypes was 264.41 g, and values varied from 73.92 g (Tift32D281-P1-P2) to 678.87 g (IP8786). Application of N significantly increased DSY by 12.6% in N₀ and 18.02% in N₁₀₀ compared to the N₀ condition. Significant differences were observed for dry stover weight among the genotypes (P < 0.001), and interactions were also significant between seasons × treatment (P < 0.00), seasons × genotypes (P < 0.001), treatment × genotype (P < 0.001), and season × treatments × genotype.

Interestingly, out of the top 25 high dry stover yielding lines, nine genotypes (IP14398, IP10579, IP15857, IP10394, IP17125, IP13608, IP6098, IP12020, and IP11584) were common in the top 25 lines at N₁₀₀ condition (Figure 3). At both N levels, the top 25 poor-performing dry stover genotypes were also identified (Figure 4).

TABLE 2 | Top 25 NIS and NS lines under low nitrogen (N₀) with contrasting NUE grain yield derived from the pooled mean over three seasons.

| S. No | Genotypes | NIS | Genotypes | NS |
|-------|------------|-----|------------|----|
| 1     | IP18621    | 138.42 | IP13363   | 9.64 |
| 2     | IP10820    | 131.19 | IP3110    | 11.54 |
| 3     | IP17720    | 130.81 | IP6282    | 13.56 |
| 4     | IPC804     | 127.12 | IP4378    | 15.27 |
| 5     | ICMB01222-P1 | 119.13 | IP8500    | 15.43 |
| 6     | IP10379    | 116.92 | IP15344   | 15.75 |
| 7     | IP3108     | 115.54 | IP14398   | 18.45 |
| 8     | IP11577    | 115.52 | IP14311   | 18.72 |
| 9     | IP16096    | 114.84 | IP7941    | 19.73 |
| 10    | IP5560     | 113.03 | IP5031    | 20.88 |
| 11    | ICMB89111-P2 | 110.84 | IP10456   | 21.15 |
| 12    | IP8069     | 109.29 | IP13608   | 21.84 |
| 13    | IP8665     | 109.15 | AMIP92901-S1-15-1-2-B-P03 | 22.09 |
| 14    | ICMV-IS92222 | 108.9 | IP6310    | 22.31 |
| 15    | ICMB90111-P2 | 108.34 | IP4979    | 24.2 |
| 16    | IP6099     | 107.22 | IP6869    | 24.92 |
| 17    | IP13016    | 104.61 | ICMB90111-P5 | 26.13 |
| 18    | IP17028    | 103.4 | IP19584   | 26.53 |
| 19    | Tift32D1-P158 | 103.31 | IP79300   | 27.56 |
| 20    | IP16403    | 102.3 | IP11593   | 29.33 |
| 21    | ICMV-IS89305 | 101.87 | IR8333/47-P1 | 29.81 |
| 22    | IP6109     | 101.32 | IP15070   | 29.94 |
| 23    | IP6060     | 100.37 | IP5438    | 30.23 |
| 24    | IP22424    | 100.3 | IP8000    | 31.19 |
| 25    | ICMV221 = ICMV88904 | 100.22 | Tift186   | 31.22 |
In addition, three high yielding dual purpose genotypes (IP5560, IP15857, and IP17125) were identified in the top 25 GY and DSY genotypes under N₀. Whereas in N₁₀₀, only one common high yielding genotype (IP3106) was identified. Descriptive statistics over the three seasons, and ANOVA results for the agronomic traits were provided in Table 1.

**N Content in Grain and Stover**

Nitrogen percent in grain (NPG) varied from 1.18% (IC804) to 2.43% (Tift 186), with an average of 1.61% in N₀ condition. Whereas in N₅₀, NPG ranged from 1.19% (9444) to 2.43% (IP4942) with a mean of 1.67%. Under N limiting conditions NPG was reduced by 3.6 and 11.4% as compared with N₅₀ and N₁₀₀ conditions, respectively. In N₅₀, an average mean of NPG decreased by 8.24% as compared to N₁₀₀. The response of genotypes in the N₁₀₀ condition varied significantly from 1.27% (IP10539) to 2.59% (IP4828) with a mean of 1.82%. Significant differences were observed in NPG among the genotypes (P < 0.001), treatments (P < 0.001), and interactions and are also significant.
FIGURE 3 | Top 25 high dry stover yielding (DSY) genotypes under N₀ and N₁₀₀ conditions. Total 9 genotypes (highlighted in red color) viz., IP 14398, IP 10579, IP 15857, IP 10394, IP 17125, IP 13608, IP 6098, IP 12020, and IP 11584 exhibited high dry stover yield (DSY) in both N₀ and N₁₀₀ conditions.

FIGURE 4 | Least performing genotypes for dry stover yield (DSY) under N₀ and N₁₀₀ conditions.
between seasons × treatment (P < 0.00), seasons × genotypes (P < 0.001), and season × treatments × genotype except for treatment × genotype (0.677).

Genotypic variations found in NPS in N₀ and the minimum and maximum value of NPS ranged from 0.42% (IP8786) to 1.08% (ICMB90111-P5) with a mean of 0.61%. The average NPS was more in N₀ compared to N₅₀ and N₁₀₀ conditions. Significant variations were observed in all the interactions except season × treatment (0.6771) and treatment × genotypes (0.0102).

**Nitrogen Uptake for Dry Stover**

Genotypic variations were observed for NUpS across the N levels. The mean of all genotypes was 1.65 kg ha⁻¹ ranged from 3.34 kg ha⁻¹ (ICMB9533-P5) to 8.03 kg ha⁻¹ (IP5560) and decreased by 9.8 and 14.45% in N₀ compared to N₅₀ and N₁₀₀, respectively. Whereas, in N₅₀, the values varied from 0.51 kg ha⁻¹ (AIMP92901-S1-15-1-B-P3) to 8.03 kg ha⁻¹ (IP10543) with a mean of 1.83 kg ha⁻¹ and decreased by 5.67% when compared with N₁₀₀. The minimum and maximum values of NUpS varied from 0.38 kg ha⁻¹ (IP155363) to 1.08 kg ha⁻¹ (IP11584) with a mean of 6.59 kg ha⁻¹ under N₁₀₀ condition.

**Total Nitrogen Uptake**

Application of N fertilizer resulted in an increase of TNUp and also revealed significant genotypic variations among the genotypes with in the treatments. Under N₀, the values ranged from 1.09 to 10.47 with an average of 3.03 kg ha⁻¹ and decreased by 14.4 and 21% as compared with N₅₀ and N₁₀₀, respectively. Whereas in N₅₀ and N₁₀₀, values varied from 1.08 (IP11229) to 9.57 (843B) and 0.58 (IP13363) to 8.97 (IP11584) with a mean of 3.54 and 3.83 kg ha⁻¹, respectively. The results of ANOVA for TNUp indicated significant effects for all the interactions except treatments.

**Nitrogen Utilization Efficiency**

Large variation was observed for NUtE of genotypes, ranged from 2.40 (IP31110) to 50.60 (IP3557) with an average of 30.81 under N₀ and the efficiency was reduced as compared with N₅₀ and N₁₀₀ by 7.3 and 5.95%, respectively. Whereas in N₅₀, values varied from 10.63 to 50.86 with a mean of 33.22, and a 7.57% reduction was observed as compared with the N₁₀₀ condition. The application of N fertilizer (N₁₀₀) resulted in a significant increase in NUtE. The minimum and maximum NUtE were observed in IP31110 (10.06) and IP10456 (56.25), respectively with an average of 32.76. Significant differences were observed among the genotypes (P < 0.001), treatments (P < 0.001), and seasons × treatments (P < 0.001), seasons × genotypes (P < 0.001), treatment × genotypes and interactions were also significant between season × treatments × genotype.

Nitrogen utilization efficiency under N₀ was compared with NUtE under N₁₀₀ to determine the genotypic efficiency and their responsiveness to N. Based on the NUtE, the genotypes were classified into four groups (Figure 5) viz., (1) N efficient non-responsive (NENR), (2) N efficient responsive (NER), (3) N responsive inefficient, and (4) N inefficient non-responsive (Rengel and Graham, 1995; Worku et al., 2007). In this study, NUtE data recorded in 374 genotypes across different N levels. The average NUtE of 374 genotypes under N₀ (30.81) and N₁₀₀
condition (32.76) were considered as cut off for the identification of genotypic efficiency and responsiveness for N use. Under low N, above-average genotypes were considered as efficient and below-average genotypes were considered as inefficient. Similarly, in N100, above-average genotypes were considered as responders, and below-average genotypes are considered as non-responders. Overall, efficient genotypes are higher in the utilization of absorbed N over inefficient genotypes.

**Nitrogen Harvest Index (NHI)**

The nitrogen harvest index varied from 5.46% (IP3110) to 74.52% (ICMB 95333-P5) with an average of 47.56% in N0 condition and a reduction of about 10.3 and 15.43% compared to N50 and N100 condition, respectively. Whereas in N50, NHI values ranged from 21.10% (IP10488) to 84.02% (IP5923) with a mean of 53.0%. The response of genotypes under the N100 condition varied significantly and ranged from 23.51% (IP12020) to 78.89% (IP6110) with a mean of 56.24%. Further, the NHI reduction under N0 was about 10.3 and 15.43% as compared with N50 and N100, respectively. In N50, a reduction of NHI about 5.46% was observed compared to the N100 condition. Significant differences were observed among the genotypes (P < 0.001), treatments (P < 0.001), and interactions were also significant between seasons × treatment (P < 0.001), seasons × genotypes (P < 0.001), and season × treatments × genotype except for the treatment × genotype (0.677) (Table 3).

**Phenotypic Correlation**

Correlation coefficient analysis was performed to identify the interrelation among the traits for all the N levels. The complete list of correlation coefficient values among the traits in each N level is provided in Figure 6. GY showed a significant positive correlation with TNUp, NUtE, and NHI. The results revealed that the GY of pearl millet was increased through the selection of higher NUtE and NHI lines. Furthermore, GY negatively correlated with DSY, NPS, NPG, and NUpS at both N50 and N100 conditions. The results suggested that the higher GY was, the lower DSY, NPS, NPG, and NUpS. Further DSY showed a negative significant correlation with all the measured traits except NUpS and TNUp across N levels. Interestingly NUtE showed a positive correlation with GY and NHI across N levels. SPAD chlorophyll content was found to be positively correlated with all traits except LA, DSY, and NUpS across all the treatments. Further LA was negatively correlated with GY, NUtE, and NHI across the treatments.

**Cluster Analysis**

Hierarchical cluster analysis of 380 pearl millet genotypes revealed large genotypic variation in five traits viz., SPAD chlorophyll content, LN LA, GY, and DSY. This analysis is useful for grouping genotypes for NUE. Therefore, clustering 380 genotypes based on five traits identified two to three major clusters. Based on this, genotypes were classified into high, medium, and low-performing genotypes. At both N levels, clusters I and II contained better-performing genotypes and also found the least dissimilarity among the genotypes. Interestingly few common genotypes viz., IP 15536, IP 6098, IP11961, IP8863, IP11378, IP 20679, and IP 12138 were identified in N0 and N100 levels in the first two clusters, implying that genotype plasticity toward N under N0 and N100 conditions (Figures 7, 8). Cluster III contained more number of medium performing genotypes that form a maximum number of sub clusters at both N levels. Finally, the least performing genotypes were present in Cluster IV. Overall, the maximum dissimilarity (from all these matrices) was observed between cluster I and IV.

### Table 3: ANOVA for N-related traits under three nitrogen levels during Summer 2017 and 2018.

| Traits          | Range (N0) | Range (N50) | Range (N100) | Mean (N0) | Mean (N50) | Mean (N100) | Percent reduction N0 compared with N50 | Percent reduction N0 compared with N100 | Percent reduction N50 compared with N100 | Percent reduction N0 compared with N100 | F statistic | NPG (%) | NPS (%) | NUpS (kg ha\(^{-1}\)) | TNUp (kg ha\(^{-1}\)) | NUtE (kg ha\(^{-1}\)) | NHI (%) |
|-----------------|------------|-------------|-------------|-----------|------------|------------|----------------------------------------|----------------------------------------|----------------------------------------|----------------------------------------|-------------|---------|---------|----------------|----------------|----------------|---------|
| Range           | N0         | 1.18–2.43   | 0.42–1.08   | 0.34–0.83 | 1.09–10.47 | 2.42–50.6  | 5.46–74.52                                      |                                       |                                        |                                        | 1808.86*** | 717.71*** | 7105.54*** | 12241.28*** | 129.2*** | 45.79 |
|                 | N50        | 1.19–2.43   | 0.4–0.94    | 0.51–6.27 | 1.08–9.57  | 10.63–56.86 | 21.07–84.02                                 |                                       |                                        |                                        | 39.22***   | 12.04*** | 1.75*** | 2.52*** | 43.13*** | 43.8*** |
|                 | N100       | 1.27–2.59   | 0.52–1.03   | 0.38–6.59 | 0.58–8.97  | 10.06–56.27 | 23.51–78.89                                 |                                       |                                        |                                        | 0.39**      | 5.39**    | 148.57*** | 212.2***  | 172.45*** | 16.36*** |
|                 | G          | 9.34***     | 3.48***     | 7.7***    | 6.66***    | 9.86***    | 8.33***                                   |                                       |                                        |                                        | 5.19***     | 2.67***   | 6.44*** | 4.58*** | 4.5***   | 4.72*** |
|                 | S × T      | 1.85***     | 1.15**      | 3.91***   | 3.07***    | 2.09***    | 2.47***                                  |                                       |                                        |                                        | 1.47***     | 1.16***   | 2.26*** | 1.85*** | 2.09*** | 2.31*** |
|                 | S × G      | 1.85***     | 1.16**      | 3.91***   | 3.07***    | 2.09***    | 2.47***                                  |                                       |                                        |                                        | 1.47***     | 1.16***   | 2.26*** | 1.85*** | 2.09*** | 2.31*** |
|                 | T × G      | 1.85***     | 1.16**      | 3.91***   | 3.07***    | 2.09***    | 2.47***                                  |                                       |                                        |                                        | 1.47***     | 1.16***   | 2.26*** | 1.85*** | 2.09*** | 2.31*** |
|                 | S × T × G  | 1.85***     | 1.16**      | 3.91***   | 3.07***    | 2.09***    | 2.47***                                  |                                       |                                        |                                        | 1.47***     | 1.16***   | 2.26*** | 1.85*** | 2.09*** | 2.31*** |

NPG: N percent in grain; NPS: N percent in dry stover; NUpS, N uptake in stover; TNUp, total nitrogen uptake; NUtE, nitrogen utilization efficiency; NHI, nitrogen harvest index; G, Genotype; S, Season; T, treatment.

F statistic values with asterisk indicate significance at the level of 0.01 (*) and 0.001 (***), and NS denotes no significance.
DISCUSSION

Nitrogen (N) is an essential macronutrient required for plant growth and is often a limiting factor for crop yield. From the past 50 years of agriculture, augmented food production was attained by the extensive application of N fertilizer in combination with N-responsive cultivars across the globe. Excessive application of N fertilizer is becoming expensive which accounts for the loss of economic profit to the farmers along with negative impacts on the environment (Raun et al., 2002; Hawkesford and Griffiths, 2019). Hence to overcome this problem, a clear understanding of genotype behavior, identification and development of genotypes (without compromising GY) with high NUE under low N conditions is a paramount need for improving NUE.

Therefore, the present study aimed to identify the N-Sensitive (NS) and N-insensitive (NIS) genotypes and also to determine the traits regulating low N tolerance. However, accurate and reliable phenotyping under low N input is challenging and influenced by the genotype (G), environment (E), and G × E interactions (Chen et al., 2014; Rao et al., 2018). Moreover, there are limited studies available in pearl millet toward the identification of nitrogen insensitive genotypes under low N conditions at field level. Based on the current knowledge, this study presents the first report of genetic variations for NUE in the global association panel, Pearl Millet inbred Germplasm Association Panel (PMiGAP) (PMiGAP collected from a large set of diverse germplasm and breeding lines) and from mapping population parents. Notably, very few pearl millet breeding programs are targeting the development of low N tolerance traits which are a must for sustainable agriculture with minimal negative impacts on the environment. Components of NUE were studied in pearl millet hybrids at two N levels found that the efficient genotype (Souna B) had a 32% of higher NUE value than the N inefficient Indian genotype (BJ104) (Alagarswamy and Bidinger, 1987).

Another study revealed that NUtE contributes more to genetic variation in NUE (Maman et al., 2006).

In this study, a total of 380 diverse pearl millet genotypes were characterized for SPAD chlorophyll content, LN, LA, GY, DSY, and NUE related traits at three different N levels under field conditions. Notably, diverse responses have been observed among the genotypes across N levels, despite a similar growth conditions and an equal amount of N fertilizer in a given N
These observed genotypic variations purely reveal the genotype plasticity toward traits. This set of lines used in this study which are collected from different regions across the globe and these lines were used for various abiotic stress studies. Previous studies in rice, maize, wheat, and oilseed rape, etc., have established significant genetic variation for NUE-related traits with large germplasm panels, hybrids, open-pollinated and recombinant inbred line.
populations (Chen et al., 2014; Li et al., 2015; Vijayalakshmi et al., 2015; Ertiro et al., 2017; He et al., 2017; Rao et al., 2018). Generally, the results of ANOVA explained statistical differences among the treatments and genotypes and their interactions. In the current study, ANOVA results revealed significant variations among the genotypes and treatments for all measured traits except a few traits (Tables 1, 3). These results were indicating that the environment under different N levels was a crucial factor.
in explaining the genotypic variance of GY and its related traits, which are strongly influenced by the relative contribution of G × E interactions under field evaluation. The results concur with other reported field experiments in rice (Srikanth et al., 2016) and wheat (Sial et al., 2005; Belete et al., 2018).

In the present study, SPAD chlorophyll content significantly increased with nitrogen application, which might be sufficient availability of N in the leaf. Similar results of increased SPAD chlorophyll content by increasing N application was reported by Kitajima and Hogan (2003), Pramanik and Bera (2013). The relation between the slow or controlled release of N fertilizers supporting more N absorption and related physiological mechanisms has been well studied in different crops (Li et al., 2003; Long et al., 2013). LA of all the pearl millet genotypes was significantly increased with N application which might be attributed to translocation of N to leaves, which brings variation in plant architecture and leaf internal structure (Singh et al., 2004; Mhaskar et al., 2005). Reduction in LN under N0 condition was observed, due to deprivation of N, thereby reduced source size and hindering the plant development.

Across the year on average, GY ranged from 9.64 g to 159.71 g (N0), 20.72 g to 236.43 g (N50), and 20.48 g to 242.12 g (N100), indicating the genotypic variability in GY at different N regimes. In general, GY increased was correlated with the N application rate, which might be due to sufficient nitrogen availability. Recently similar kind of results was reported in a diverse set of foxtail millet genotypes (Bandyopadhyay et al., 2020). In wheat, GY and quality traits were evaluated by Sarčevec et al. (2014) at low and normal N and found that GY was reduced by 10% under low N condition compared to normal condition. The current results also are in tune with the earlier reports as there was an increased GY with increase N levels. Hawkesford and Griffiths (2019) reported that split application of N is the best option to recover the maximum applied N in the form of harvested grain/increased GY and found a strong significant relationship between GY and physiological traits.

Remarkably, 25 high yielding genotypes were identified under the N0 condition and considered as N-insensitive (NIS). Out of 25, nine genotypes (IP14398, IP10579, IP10394, IP17125, IP13608, IP6098, IP12020, and IP11584) were identified and has unique features like dry season crop, production environments having low nitrogen, high photosynthetic efficiency, and high dry forage capacity (Govintharaj et al., 2018). Several reports depict the dry fodder demand and it would require approximately 568 million tonnes across the globe by the year 2030 (Govintharaj et al., 2018). To tackle this problem, high fodder yielding genotypes need to be identified. Generally, most of the genotypes have lower dry matter production under low N conditions, hence identification of genotypes with high dry matter production under low N conditions is important for sustainable fodder production. In this framework, the top and least performing 25 genotypes for DSY were identified under N0 and N100 conditions (Table 4).

These contrasting genotypes can be further exploited in the breeding program and molecular studies to develop varieties with high NUE for biomass. Irrespective of N, the common nine genotypes (IP14398, IP10579, IP10394, IP17125, IP13608, IP6098, IP12020, and IP11584) were identified and can produce more dry matter production under both N0 and N100 conditions. These genotypes need further physiological and molecular characterization under low nitrogen conditions to understand the molecular basis for low nitrogen tolerance and biomass production. Overall, observed genetic variability in pearl millet genotypes are suited well for improving livestock productivity. Studies on various crops to determine the relationship between nitrogen levels and biomass traits have shown that N fertilizer application increased the biomass related traits, including DSY (Chan-Navarrete et al., 2014). The present

| S. No | Genotypes | Top 25 DSY | Genotypes | Least 25 DSY |
|-------|------------|------------|------------|--------------|
| 1     | IP8863     | 806.09     | LGD-1-B-10 | 58.47        |
| 2     | IP5560     | 741.82     | ICMV85333-P5 | 59.72      |
| 3     | IP14398    | 529.71     | IP6103     | 60.13        |
| 4     | IP10579    | 523.27     | IP4542     | 74.33        |
| 5     | IP15857    | 481.52     | IP6584     | 83.17        |
| 6     | IP14418    | 475.12     | AIMP92901-S1-15-1-2-B-P03 | 85.84 |
| 7     | IP13840    | 443.79     | IP9347     | 86.42        |
| 8     | IP6310     | 428.9      | IP8275     | 87.84        |
| 9     | IP10394    | 419.14     | IP13863    | 88.54        |
| 10    | ICMV221    | 395.32     | W504-1-1   | 92.31        |
| 11    | IP14624    | 395.1      | IP22455    | 92.39        |
| 12    | IP17125    | 389.29     | IP8276     | 93.25        |
| 13    | IP18246    | 387.36     | IP9426     | 93.73        |
| 14    | IP13608    | 386.07     | IP2557     | 96.39        |
| 15    | IP15070    | 380.06     | GB8735     | 96.83        |
| 16    | IP18500    | 377.18     | AIMP92901  | 97.12        |
| 17    | IP8074     | 362.87     | IP9332     | 99.22        |
| 18    | IP14311    | 362.63     | Tl232D281-P1-P5 | 102.62 |
| 19    | IP6098     | 360.08     | Okashana1  | 102.92       |
| 20    | IP9981     | 359.06     | IP11310    | 103.67       |
| 21    | IP10811    | 356.19     | ICTP8203   | 104          |
| 22    | IP12020    | 353.03     | 863B-P2    | 104.48       |
| 23    | IP12116    | 351.76     | RIB335/74-P1 | 104.52     |
| 24    | IP11584    | 344.58     | ICM57703   | 105.77       |
| 25    | IP17493    | 337.88     | IP3201     | 105.86       |
results were tuned with these observations as DSY was more in N\textsubscript{100} condition than in N\textsubscript{0} condition; the increasing quantity was up to the significant levels. Another study revealed wide genetic variations for biomass traits and stover nitrogen in pearl millet germplasm (Gupta et al., 2015). Few studies revealed that significant genetic variation present in pearl millet for biomass traits in dual-purpose hybrids and open-pollinated varieties (OPVs), populations, and top cross hybrids (Bidinger et al., 2010; Blümmel et al., 2010; Rai et al., 2012). More importantly from this study, dual-purpose genotypes were identified and these can be used as a reference set for the developing high grain and stover yielding lines.

In the present study, a total of 11 traits were recorded for all three seasons. However, due to heavy rain, we are unable to generate quality data for the rainy season. Hence to avoid misinterpretation of results/findings, data for only six NUE traits were pooled from two summer seasons and presented. Moreover, misinterpretation of results/findings, data for only six NUE traits to generate quality data for the rainy season. Hence to avoid all three seasons. However, due to heavy rain, we are unable this study, dual-purpose genotypes were identified and these can be used in the utilization of absorbed nutrients than inefficient genotypes. Vast genotypic variations in NUE have been reported under field/pot screening in various wheat genotypes and other crops by several researchers (Bouchet et al., 2014; Ma et al., 2015).

The nitrogen harvest index indicates the level of efficiency of plants to use acquired total N for grain formation (He et al., 2017). In the present study, significant genotypic variation was observed for NHI at both N\textsubscript{0} and N\textsubscript{100} conditions. NHI was positively correlated with NUE and GY in all three N levels. These findings concur with previous studies in oilseed rape (Ulasc, 2013; Stahl et al., 2016) and wheat (Monostori et al., 2017).

In the current study, the Pearson correlation coefficient analysis for the GY and NUE related traits indicated that NUE trait has the largest contribution to the GY followed by NHI and TNUp across N levels, implying that GY improvement in pearl millet could be possible by selecting genotypes that attain a higher NUE and NHI across N levels. Similarly, a significant correlation between GY and NUE was reported in maize, wheat, and oilseed rape (He et al., 2017; Belete et al., 2018). Fageria et al. (2010) reported that strong associations between GY and NUE related traits could be a better option for GY improvements under limited N condition. Furthermore, GY was negatively correlated with NPG and NPS across N levels. Previous studies also reported a similar significant negative correlation between grain N concentration and GY (Sinebo et al., 2004). In the present study, a strong positive correlation of DSY with NUpS and TNUp across N levels was observed indicating that more DSY was with higher NUpS and TNUp, where it was negatively correlated with NPS, NUE, and NHI. The inverse relationship between NPS and DSY was reported in rice (Subudhi et al., 2020). Identified genetic stocks will be further utilized to carry an in-depth investigation to understand the genes and associated pathways related to NUE. Overall, important correlations identified from this study, will certainly help for the future pearl millet NUE breeding programs.

CONCLUSION

Improving NUE of pearl millet is pivotal for sustainable crop growth and yield especially under low nitrogen soils. Likewise, improving crop productivity using N fertilization is important for achieving climate resilience. Nevertheless, the genetic improvement of pearl millet NUE depends on the nature and extent of variation among the germplasm. As studies on pearl millet NUE are still nascent, this study was aimed to derive morphologic and agronomical traits associated with NUE in a
set of 380 diverse lines under different N levels. The first step in this research revealed extensive genetic variations across the diversity panel and mapping population parents under N0 and N100 conditions. Also, large environmental variations (including N inputs) for GY and its related NUE traits were observed. Nitrogen limitation resulted in the reduction of GY and DSY in N0, as compared with N20 and N100. Under low N, the best grain yielding genotypes were identified and considered as nitrogen-insensitive (NIS). Genotypes IP10820, IP17720, ICMB01222-P1, IP10379, ICMB89111-P2, IP8069, ICMB90111-P2, ICVM-I89305, and ICVM221 (ICMV88904) proved to be the most efficient genotypes in terms of GY at low and high N levels, and indeed shows their inherent genotypic plasticity toward N application. Furthermore, the use of NIS genotypes will help in breeding N-efficient genotypes for arid and marginal agro-ecologies. Overall results suggest that genotypes with more yield and high to moderate NUtilE can be used as parents for the breeding of N efficient genotypes. The lines identified from the present study, coupled with multi-omics technologies can help to identify candidate genes for NUE in pearl millet. The available genetic stocks will be useful to carry an in-depth dissection of the genes and pathways related to NUE. These contrasting genetic stocks may help to map the genome, transcriptome, proteome, and metabolome signatures of the traits leading to a better understanding of NUE in pearl millet. These current findings may also help the farmers for optimizing the use of fertilizer inputs for economic and environmentally sustainable food production.

**DATA AVAILABILITY STATEMENT**

The original contributions presented in the study are included in the article/Supplementary Material, further inquiries can be directed to the corresponding author/s.

**AUTHOR CONTRIBUTIONS**

RKS and RG planned and designed this research. VP, MP, SB, RR, and VT performed the field experiments, phenotyping, and analyzed the data. AR, RD, and GA carried out statistical analysis. RKS and RG interpreted the data. VP, MP, RKS, and RG wrote the manuscript. All the authors have made their contribution in editing the manuscript for publication.

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**SUPPLEMENTARY MATERIAL**

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fpls.2021.625915/full#supplementary-material

**REFERENCES**

Alagarswamy, G., and Bidinger, F. R. (1987). "Genotypic variation in biomass production and nitrogen use efficiency in pearl millet [Pennisetum americanum (L.) Leeke]," in Genetic Aspects of Plant Mineral Nutrition, eds W. H. Gabelman and B. C. Loughman (Dordrecht: Springer), 281–286. doi: 10.1007/978-94-009-3581-5_25

Anbessa, Y., Juskiw, P., Good, A., Nyachiro, J., and Helm, J. (2009). Genetic variability in nitrogen use efficiency of spring barley. *Crop Sci.* 49, 1259–1269. doi: 10.2135/cropsci2008.09.0586

Arduini, I., Masoni, A., Ercozi, L., and Mariotti, M. (2006). Grain yield, and dry matter and nitrogen accumulation and remobilization in durum wheat as affected by variety and seeding rate. *Eur. J. Agron.* 25, 309–318. doi: 10.1016/j.eja.2006.06.009

Bandyopadhyay, T., Swarbrick, S. M., Jaiswal, V., Gupta, R., Bentley, A. R., Griffiths, H., et al. (2020). Grain number and genotype drive nitrogen-dependent yield response in the C4 model *Setaria italica* (L.) P. Beauv. *bioRxiv* [Preprint]. doi: 10.1101/2020.03.23.003004

Basava, R. K., Hash, C. T., Mahendrakar, M. D., Kishor, P. B. K., Satyavathi, C. T., Kumar, S., et al. (2019). Discriming combining ability loci for divergent environments using chromosome segment substitution lines (CSSLs) in pearl millet. *Plos One* 14:e0218916. doi: 10.1371/journal.pone.0218916

Belette, F., Dechassa, N., Molla, A., and Tana, T. (2018). Effect of nitrogen fertilizer rates on grain yield and nitrogen uptake and use efficiency of bread wheat (*Triticum aestivum* L.) varieties on the Vertisols of central highlands of Ethiopia. *Agric. Food Secur.* 7, 1–12. doi: 10.1186/s40066-018-0231-z

Bidinger, F. R., Blumme, M., Hash, C. T., and Choudhary, S. (2010). Genetic enhancement for superior food-feed traits in a pearl millet (*Pennisetum glaucum* (L.) R. Br.) variety by recurrent selection. *Anim. Nutr. Feed Tech.* 10, 61–68.

Blumme, M., Khan, A. A., Vadez, V., Hash, C. T., and Rai, K. N. (2010). Variability in stover quality traits in commercial hybrids of pearl millet (*Pennisetum glaucum* (L.) R. Br.) and grain-stover trait relationships. *Anim. Nutr. Feed Tech.* 10, 29–38.

Bouchet, A. S., Nesi, N., Bissuel, C., Bregoon, M., Lariepe, A., Navier, H., et al. (2014). Genetic control of yield and yield components in winter oilseed rape...
(Brassica napus L.) grown under nitrogen limitation. *Euphytica* 199, 183–205. doi: 10.1007/s10681-014-1130-4

Chan-Naravate, R., Kawai, A., Dolstra, O., Van Bueren, E. T. L., and Van der Linden, C. G. (2014). Genetic diversity for nitrogen use efficiency in spinach (*Spinacia oleracea* L.) cultivars using the Ingestad model on hydroponics. *Euphytica* 199, 155–166. doi: 10.1007/s10681-014-1186-1

Chen, B., Xu, K., Li, J., Li, F., Qiao, J., Li, H., et al. (2014). Evaluation of yield and agronomic traits and their genetic variation in 488 global collections of *Brassica napus* L. *Genet. Resour. Crop Evol.* 61, 979–999. doi: 10.1007/s10722-014-0091-8

D’Andrea, A. C., and Casey, J. (2002). Pearl millet and Kintampo subsistence. *Afr. Archaeol. Rev.* 19, 147–173. doi: 10.1023/A:1016518919072

Ertiro, B. T., Beyene, Y., Das, B., Mugo, S., Olsen, M., Oikeh, S., et al. (2016). Combining ability and testcross performance of drought-tolerant maize inbred lines under stress and non-stress environments in Kenya. *Plant Breeding*. 136, 197–205. doi: 10.1111/pbr.12464

Fageria, N. K. (2014). Nitrogen harvest index and its association with crop yields. *J. Plant Nutr.* 37, 795–810. doi: 10.1080/01904167.2014.881855

Fageria, N. K., and Baligar, V. C. (2008). The role of nutrient efficient genotypes in *Brassica napus* L. *J. Plant Nutr.* 31, 1121–1157. doi: 10.1080/01904160802116068

Fageria, N. K., De Morais, O. P., and Dos Santos, A. B. (2010). Nitrogen use efficiency in upland rice genotypes. *J. Plant Nutr.* 33, 1696–1711. doi: 10.1080/019041610.2010.496892

FAO (2019). FAO World Fertilizer Trends and Outlook to 2020. Available online at: http://www.fao.org/3/a-i6895e.pdf (accessed June 11, 2019).

Gianquinto, G., Goffart, J. P., Olivier, M., Guarda, G., Colauzzi, M., Dalla Costa, L., et al. (2004). The use of hand-held chlorophyll meters as a tool to assess the nitrogen status and to guide nitrogen fertilization of potato crop. *Potato Res.* 47, 35–40. doi: 10.1007/bf02731970

Good, A. G., Shrawat, A. K., and Muench, D. G. (2004). Can less yield more? Is fertilizers on lowland rice yield and nitrogen use efficiency. *Clin. J. Soil Sci.* 35, 311–315.

Li, F., Chen, F., Cai, H., Liu, J., Pan, Q., Liu, Z., et al. (2015). A genetic relationship between nitrogen use efficiency and seedling root traits in maize as revealed by QTL analysis. *J. Exp. Bot.* 66, 3175–3188. doi: 10.1093/jxb/erv127

Long, J. R., Wan, Y. Z., Song, C. F., Jian, S. U. N., and Qin, R. J. (2013). Effects of nitrogen fertilizer level on chlorophyll fluorescence characteristics in flag leaf of super hybrid rice at late growth stage. *Rice Sci.* 20, 220–228. doi: 10.1007/s12010-013-6308(13)0138-9

Lopez-Bellido, R. J., Shepherd, C. E., and Barraclough, P. B. (2004). Predicting post-anthesis N requirements of bread wheat with a Minolta SPAD meter. *Eur. J. Agron.* 20, 313–320. doi: 10.1016/j.eja.2003.03.0025-X

Ma, B. L., Biswas, D. K., Herath, A. W., Whalen, J. K., Ruan, S. Q., Caldwell, C., et al. (2015). Growth, yield, and yield components of canola as affected by nitrogen, sulfur, and boron application. *J. Plant Nutr. Soil Sci.* 178, 658–670. doi: 10.1002/jpln.201400280

Maman, N., Mason, S. C., and Lyon, D. J. (2006). Nitrogen rate influence on pearl millet yield, nitrogen uptake, and nitrogen use efficiency in Nebraska. *Commun. Soil Sci. Plant Anal.* 37, 127–141. doi: 10.1080/00103530504061112

Manning, K., Pelling, R., Higham, T., Schwenniger, J. L., and Fuller, D. Q. (2011). 4500-Year old domesticated pearl millet (*Pennisetum glaucum*) from the Tilemsi Valley, Mali: new insights into an alternative cereal domestication pathway. *J. Archaeol. Sci.* 38, 312–322. doi: 10.1016/j.jars.2012.16

Mhaskar, N. V., Dongale, J. H., Dademal, A. A., and Khanvilkar, S. A. (2005). Performance of scented rice varieties under different nitrogen levels in lateritic soil of Konkan. *Oryza* 43, 233.

Moll, R. H., Kamprath, E. J., and Jackson, W. A. (1982). Analysis and interpretation of factors which contribute to efficiency of nitrogen utilization. *Field Crops Res.* 42, 3489–3499. doi: 10.1016/0378-5179(92)90012-3

Muchow, R. C. (1998). Nitrogen utilization efficiency in maize and grain sorghum. *Field Crops Res.* 56, 209–216. doi: 10.1016/S0378-4290(97)00132-9

Nami, S., Toriyama, K., and Fukuta, Y. (2009). Genetic variations in dry matter production and physiological nitrogen use efficiency in rice (*Oryza sativa* L.) varieties. *Breed. Sci.* 59, 269–276. doi: 10.1276/bbss.59.269

Pramanik, K., and Bera, A. K. (2013). Effect of seeding age and nitrogen fertilizer on growth, chlorophyll content, yield and economics of hybrid rice (*Oryza sativa* L.). *Int. J. Agron. Prod. Prod.* 4, 3489–3499.

Presterl, T., Seitz, G., Landbeck, M., Thiemt, E. M., Schmidt, W., and Geiger, H. H. (2003). Improving nitrogen-use efficiency in European maize. *Crop Sci.* 43, 1259–1265. doi: 10.2136/cropscis2003.1259

Rai, K. N., Blummel, M., Singh, A. K., and Rao, A. S. (2012). Variability and relationships among forage yield and quality traits in pearl millet. *Eur. J. Plant Sci. Biotechnol.* 6, 118–124.
Pujarula et al.  

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Raon, W. R., Johnson, G. V., and Westerman, R. L. (1999). Fertilizer nitrogen recovery in long-term continuous winter wheat. *Soil Sci. Soc. Am. J.*, 63, 645–650. doi: 10.2136/sssaj1999.036159950063000300x

Raun, W. R., Solie, J. B., Johnson, G. V., Stone, M. L., Mullen, R. W., Freeman, K., et al. (2002). Improving nitrogen use efficiency in cereal grain production with optical sensing and variable rate application. *J. Agron.* 94, 815–820. doi: 10.2134/agronj2002.8150

Rengel, Z., and Graham, R. D. (1995). Wheat genotypes differ in Zn efficiency when grown in chelate-buffered nutrient solution. *Plant Soil* 176, 307–316. doi: 10.1007/BF00011795

Sahrawat, K. L., Kumar, G. R., and Murthy, K. V. S. (2002). Sulfuric acid–selenium digestion for multi-element analysis in a single plant digest. *Commun. Soil Sci. Plant Anal.* 33, 3757–3765. doi: 10.1081/CSI-120015920

Sinha, R., Giri, M., and Prasad, A. (2017). Mapping QTL for agronomic traits in breeding populations. *Front. Genet.* 18, 208–222. doi: 10.3389/fgene.2017.00398

Stahl, A., Friedt, W., Wittkop, B., and Snowdon, R. J. (2016). Complementary diversity for nitrogen uptake and utilisation efficiency reveals broad potential for increased sustainability of oilseed rape production. *Plant Soil* 400, 245–262. doi: 10.1007/s11104-015-2726-8

Subudhi, H. N., Prasad, K. V. S. V., Ramakrishna, C., Rameswar, P. S., Pathak, H., Ravi, D., et al. (2020). Genetic variation for grain yield, straw yield and straw quality traits in 132 diverse rice varieties released for different ecologies such as upland, lowland, irrigated and salinity prone areas in India. *Field Crops Res.* 245:107626. doi: 10.1016/j.fcr.2019.107626

Tako, E., Reed, S. M., Budiman, J., Hart, J. J., and Glahn, R. P. (2015). Higher iron pearl millet (*Pennisetum glaucum* L.) 967 provides more absorbable iron that is limited by increased polyphenolic content. *Natr. J.* 14, 1–9. https://doi: 10.1186/1478-7981-14-11

Ulas, A., Behrens, T., Wiesler, F., and Horst, W. J. (2013). Does genotypic variation in nitrogen remobilisation efficiency contribute to nitrogen efficiency of winter oilseed-rape cultivars (*Brassica napus* L.). *Plant Soil* 371, 463–471. doi: 10.1007/s11104-013-1688-y

Vijayalakshmi, P., Kiran, T. V., Rao, Y. V., Srikanth, B., Rao, I. S., Sailaja, B., et al. (2013). Physiological approaches for increasing nitrogen use efficiency in rice. *Indian J. Plant Physiol.* 18, 208–222. doi: 10.1007/s40502-013-0042-y

Vijayalakshmi, P., Vishnuvirkiran, T., Kumari, B. R., Srikanth, B., Rao, I. S., Swamy, K. N., et al. (2015). Biochemical and physiological characterization for nitrogen use efficiency in aromatic rice genotypes. *Field Crops Res.* 179, 132–143. doi: 10.1016/j.fcr.2015.04.012

VSN International (2019). *Genstat for Windows*, 20th Edn. Hemel Hempstead: VSN International.

Worku, M., Bänziger, M., Erley, G. S. A. M., Friesen, D., Diallo, A. O., and Horst, W. J. (2007). Nitrogen uptake and utilization in contrasting nitrogen efficient tropical maize hybrids. *Crop Sci.* 47, 519–528. doi: 10.2135/cropsci2005.05.0070

Würschum, T. (2012). Mapping QTL for agronomic traits in breeding populations. *Theor. Appl. Genet.* 125, 201–210. doi: 10.1007/s00122-012-1887-6

Yang, H., Yang, J., Li, Y., and He, J. (2014). SPAD values and nitrogen nutrition index for the evaluation of rice nitrogen status. *Plant Pro. Sci.* 17, 81–92. doi: 10.1666/pps.17/81

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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