Capsaicin Reverses the Inhibitory Effect of Licochalcone A/β-Arbutin on Tyrosinase Expression in B16 Mouse Melanoma Cells

Jun-Hui Hong, Huo-Ji Chen, Shi-Jian Xiang, Si-Wei Cao, Bai-Chao An, Shi-Fa Ruan, Bin Zhang, Jun-Xia Zhu, Qiang Liu

Department of Chinese medicine preparation, School of Traditional Chinese Medicine, Department of Pharmacy, The affiliated hospital of Qingdao University, 266071, Department of Pediatrics, Hospital of Integrated Chinese and Western Medicine, Southern Medical University, 510315, P.R. China

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ABSTRACT

Introduction: Melanin is synthesized by melanocytes, which are located in the basal layer of the skin. After synthesis, melanin is further deposited on the surface of the skin to form black spots or chloasma. Tyrosinase is a rate-limiting enzyme that plays an important role in melanogenesis. Currently, there are many drugs that inhibit tyrosinase expression to further reduce melanogenesis. Nevertheless, some of these could reverse the pharmacological effect of other drugs, when used simultaneously.

Materials and Methods: B16 mouse melanoma cells were treated with the tyrosinase inhibitors licochalcone A and β-arbutin, alone or in combination with capsaicin, an alkaloid found in peppers. Cytotoxicity, melanin content, and tyrosinase activity and expression were determined.

Results: Licochalcone A/β-arbutin inhibited tyrosinase expression and further hindered melanin synthesis when applied individually to B16 mouse melanoma cells. However, licochalcone A/β-arbutin combined with 50 µmol/L capsaicin enhanced the expression of tyrosinase in these cells and further increased melanin content. Conclusion: Our data implied that capsaicin could reverse the inhibitory effect of licochalcone A/β-arbutin on tyrosinase expression in B16 mouse melanoma cells.

Key words: Capsaicin, licochalcone A, melanin, melanocyte, tyrosinase, β-arbutin

SUMMARY

B16 mouse melanoma cells were treated with the tyrosinase inhibitors licochalcone A and β-arbutin, alone or in combination with capsaicin, an alkaloid found in peppers. Cytotoxicity, melanin content, and tyrosinase activity and expression were determined. Licochalcone A/β-arbutin inhibited tyrosinase expression and further hindered melanin synthesis when applied individually to B16 mouse melanoma cells. However, licochalcone A/β-arbutin combined with 50 µmol/L capsaicin enhanced the expression of tyrosinase in these cells and further increased melanin content. Our research implied that capsaicin could reverse the inhibitory effect of licochalcone A/β-arbutin on tyrosinase expression in B16 mouse melanoma cells.

INTRODUCTION

Melanin is a protein derivative that can be divided into two categories, specifically, pitchy pigment, which is called eumelanin, and reddish-brown pigment, which is called phaeomelanin.[1,2] Both the biological pigments are synthesized by melanocytes, and the formation of melanin is dependent on tyrosine, tyrosinase, and the concentration of oxygen. Tyrosinase, a rate-limiting enzyme that plays an important role in melanogenesis,[3] catalyzes the hydroxylation of tyrosine to 3, 4-L-dihydroxyphenylalanine (L-DOPA) and the further oxidation of L-DOPA to dopaquinone.[4]
Excess production of melanin can lead to freckles, melasma, and even malignant melanoma. To address these problems, many tyrosinase inhibitors have been applied to medications and cosmetics, such as arbutin, ellagic acid, licochalcone A, and kojic acid, among others. Licochalcone A is a flavonoid compound isolated from licorice [12] that has significant pharmacological applications such as anti-inflammatory, [8-10] anti-bacterial, [11-13] anti-angiogenic, [14] and weight loss effects, [15] among others. Additionally, Furusawa et al. [16] found that licochalcone A inhibits tyrosinase activity, and thus it has potential for use in skin-whitening cosmetics. Arbutin is one whitening ingredient that is commonly used in cosmetics. [17-20] According to different chemical structures, this compound can be categorized as α-arbutin and β-arbutin. [21] Currently, β-arbutin is widely used in cosmetics. Previous studies [20-22] have found that arbutin could inhibit the expression of tyrosinase and further reduce melanin content and the deposition of melanin in the skin. Moreover, it also greatly affects the intestinal microflora [23] and possesses anti-inflammatory properties. [24]

Capsaicin is a vanillin amine alkaloid found in chili pepper [24,25] of the Solanaceae family and mainly exists in the placenta and flesh of the pepper fruit at a concentration of 0.2%~1.0%. Capsaicin has been shown to have various biological functions. For example, it can induce nociceptor neuron desensitization and possesses anti-inflammatory and analgesic properties. [26] It also inhibits platelet aggregation [27] and it plays an important role in lowering blood pressure and preventing atherosclerosis. [28] Peppin et al. [29] found that a transdermal patch containing 8% capsaicin could be used for the treatment of peripheral neuralgia and its effect was obvious and without the side effects of opioid analgesics. A capsaicin transdermal patch is a prescription drug used for relieving neuralgia, and it has been included in the United States Pharmacopeia. Recently, studies have also found that capsaicin has many pharmacological effects such as anticancer, [30] carcinogenic, [31,32] anti-obesity, [33,34] anti-bacterial, [35] antioxidant, [36] and liver protective, [37] among others.

At first, since capsaicin is the main compound responsible for the spicy, pungent taste of chili peppers, [38] we hypothesized that it could be a potential permeation enhancer. To verify this, we performed preliminary experiments. We found that β-arbutin inhibited the expression of tyrosinase and further reduced melanin contents. However, after treating with capsaicin and β-arbutin simultaneously, the melanin content of B16 mouse melanoma cells increased. The results of this preliminary experiment were not consistent with that of the previous hypothesis. Therefore, we aspired to ascertain the reason as to why melanin content decreased after treatment with β-arbutin alone, but increased with a combination of β-arbutin and capsaicin.

The present study aimed to investigate whether capsaicin could reverse the inhibitory effects of licochalcone A/β-arbutin on tyrosinase expression in B16 mouse melanoma cells.

**MATERIALS AND METHODS**

**Materials**

B16 mouse melanoma cells were purchased from the Laboratory Animal Center of Sun Yat-sen University (China), Roswell Park Memorial Institute (RPMI)-1640 medium, fetal bovine serum (FBS), penicillin-streptomycin, and trypsin-EDTA were purchased from Thermo Fisher Scientific (Gibco, USA). In addition, 60-mm tissue culture dishes and 96-well plates were purchased from Corning (USA). Capsaicin, licochalcone A, and β-arbutin were purchased from Chroma Biotechnology (China). Dimethyl sulfoxide (DMSO), TritonX-100, sodium dodecyl sulfate, sodium salt (SDS)-polyacrylamide gels, polyvinylidene fluoride (PVDF) membranes, mouse anti-β-actin antibody (A1978), and rabbit anti-goat antibody conjugated with horseradish peroxidase were purchased from Sigma-Aldrich (USA). Phosphate-buffered saline (PBS), NaOH, L-DOPA, and normal saline were purchased from Koyos Biotechnology (China). (3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2-H-tetrazolium bromide [MTT]) and the BCA protein quantification kit were purchased from Beyotime Biotechnology (China). Cell lysis solution (1 mL radio-immunoprecipitation assay [RIPA] with 10 μL phenylmethylsulfonyl fluoride or phenylmethylsulfonyl fluoride [PMSF]) was purchased from Fdbio Science (China). The mouse anti-tyrosinase antibody (ab180753) was purchased from Abcam (England). An anti-mouse antibody, conjugated with horseradish peroxidase, was purchased from Cell Signaling Technology (USA). Multiskan® was purchased from Thermo Fisher Scientific (USA).

**Cell culture**

B16 mouse melanoma cells were cultured in RPMI-1640 medium containing 10% FBS and 1% penicillin-streptomycin in a 37°C thermostat incubator with 5% CO2. The cells were seeded in 60-mm tissue culture dishes to be used for subsequent experiments.

**Experimental groups**

The experimental groups were divided into control group (culture medium only), drug control group (culture medium containing the highest concentration of the agent), negative control group (culture medium with cells and vehicle control), and experimental group (culture medium, cells, and different concentrations of the agent, specifically, 800, 400, 200, 100, and 50 μmol/L). Capsaicin and licochalcone A were dissolved in DMSO, and β-arbutin was dissolved in normal saline.

**Cytotoxicity assays**

The cytotoxicity against B16 mouse melanoma cells was determined by MTT-assays. [39] First, cells were digested with a 0.25% trypsin-EDTA solution and then seeded in 96-well plates at a density of 5 × 104 cells per 100 μL of cell suspension. Subsequently, the cells were incubated in a 37°C thermostat incubator with 5% CO2. For all experimental groups, 100 μL of capsaicin, licochalcone A, or β-arbutin, at different defined concentrations, were added after 24 h, and cells were cultured for an additional 48 h. At this time, 30 μL of MTT solution at a concentration of 5 mg/mL was added to each well, and samples were incubated for 4 h. After this, 100 μL of DMSO was added to each well. The rate of cell proliferation was calculated based on measurements at 490 nm [39] using a Multiskan®.

**Measurement of relative melanin contents**

The relative content of melanin in B16 mouse melanoma cells was measured by performing an NaOH assay. [40] For the first experiment, after 24 h, 100 μL capsaicin, licochalcone A, or β-arbutin at different concentrations were individually added to the cells; in another experiment, 50 μL of 50 μmol/L capsaicin and 100 μL of capsaicin, licochalcone A, or β-arbutin at different concentrations in combination were added to the cells, which were cultured for an additional 48 h. After this, the cells were washed with PBS. Subsequently, 100 μL of 1 mol/L NaOH solution was added to each well, and samples were incubated for 1 h at 37°C. Finally, the samples were measured at 460 nm using a Multiskan®, and relative contents of melanin were calculated.

**Measurement of the tyrosinase activity**

Tyrosinase activity in B16 mouse melanoma cells was determined by performing an L-DOPA oxidation assay. [41] After 24 h, 100 μL capsaicin, licochalcone A, or β-arbutin at different concentrations were added to the first experiment; 50 μL of 50 μmol/L capsaicin and 100 μL of different concentrations of licochalcone A or β-arbutin were added for a different
experiment and cells were cultured for 48 h. After treatment, cells were washed with PBS and 100 μL of 1% Triton X-100 was added to each well; samples were agitated for 5 min and transferred to freeze at -20°C for 1 h. After the cells were thawed at room temperature for complete lysis, 100 μL of 0.1% L-DOPA was added to each well and cells were cultured for 2 h. Finally, samples were measured at 460 nm using a Multiskan, and tyrosinase activity was calculated.

Western blotting

Cells were divided into six groups, which were different from those of previous experiments; cells were seeded at a density of 5 × 10⁵ cells/well of a 60-mm dish. After 24 h, 3 mL of drug and 1 mL of 10% RPMI-1640 medium were added to each culture dish, except for the blank control group, and cells were cultured for 48 h. After treatment, cells were washed with PBS and 400 μL of cell lysis solution (1 mL RIPA and 10 μL PMSF) was added to each 60-mm dish to lyse the cells; samples were then centrifuged at 12,000 rpm for 5 min. Protein lysates were quantified using a BCA protein quantification kit. Subsequently, 30 μL of each sample was separated by 8.0% SDS-polyacrylamide gel electrophoresis and transferred onto a PVDF membrane. The membrane was soaked in blocking reagent for 1 h, and the membrane was incubated overnight with an anti-tyrosinase antibody (1:1000) at 4°C, and washed with PBS with Tween-20 (PBST) four times. The next day, the membrane was incubated with anti-goat antibody conjugated with horseradish peroxidase (1:30,000) for 1 h at 37°C, and washed with PBST five times. Finally, the membrane was developed using enhanced chemiluminescence.

Statistical analysis

Student’s t-test was applied to compare differences between groups, with cutoff P < 0.05, using SPSS statistical software 22.0 (IBM). Data are shown as mean ± standard deviation.

RESULTS

Cytotoxic effects of drugs on B16 mouse melanoma cells

To study the cytotoxic effects of capsaicin, licochalcone A, and β-arbutin, cells were treated with different drugs at different concentrations, and cell proliferation rates are shown in Table 1. The results showed that the cell proliferation rates were above 85% after treatment with 0–800 μmol/L licochalcone A/β-arbutin, and as such, neither licochalcone A nor β-arbutin was cytotoxic for the cells. When cells were treated with 100–800 μmol/L capsaicin, cell proliferation rates were below 70%, but upon treatment with 50 μmol/L capsaicin, the cell proliferation rate was 86.9%. This indicated that this compound lacked cytotoxicity only at 50 μmol/L. To verify this, cells were simultaneously treated with 50 μmol/L capsaicin and different concentrations of β-arbutin. The results showed that 50 μmol/L capsaicin is not cytotoxic for B16 mouse melanoma cells.

Effects of drugs on melanogenesis in B16 mouse melanoma cells

Cells from the first experiment were treated with licochalcone A or β-arbutin at concentrations of 800, 400, 200, 100, and 50 μmol/L. Cells from the second experiment were treated with 50 μmol/L capsaicin and licochalcone A/β-arbutin simultaneously. The melanin content of B16 mouse melanoma cells is shown in Table 2 and Figure 1. The results indicated that with increasing concentrations of licochalcone A/β-arbutin, the melanin content decreased. However, in contrast, when cells were treated with 50 μmol/L capsaicin and licochalcone A/β-arbutin at different concentrations, the melanin content increased.

Effects of drugs on tyrosinase activity in B16 mouse melanoma cells

Cells were treated as stated previously. Tyrosinase activity in B16 mouse melanoma cells, after treatment with drugs at different concentrations, is shown in Table 3 and Figure 2. Variations in tyrosinase activity were consistent with melanin content. Thus, regarding melanogenesis and tyrosinase activity, results of the aforementioned experiments indicated that capsaicin and licochalcone A/β-arbutin probably have antagonistic effects.

Effects of drugs on tyrosinase expression in B16 mouse melanoma cells

As is known, the expression of tyrosinase in B16 mouse melanoma cells can be determined by Western blot analysis. The results [Figure 3] implied that tyrosinase protein expression was significantly inhibited after treatment with β-arbutin, licochalcone A, or capsaicin, individually. Moreover, the expression of tyrosinase protein was augmented by treatment with capsaicin and licochalcone A/β-arbutin simultaneously.

Table 1: Cell proliferation rate of B16 mouse melanoma cells with different treatments (mean±standard deviation)

| Concentration (μmol/L) | Licochalcone A (%) | β-arbutin (%) | Capsaicin (%) | β-arbutin and 50 μmol/L capsaicin (%) |
|-----------------------|-------------------|--------------|--------------|--------------------------------------|
| 0                     | 100±0.01          | 100±0.09     | 100±0.02     | 100±0.03                             |
| 50                    | 91.2±2.56         | 95.9±1.86    | 86.9±1.25    | 97.8±2.94                            |
| 100                   | 90.3±1.36         | 96.6±1.07    | 69.7±0.94    | 101.0±1.81                           |
| 200                   | 88.8±0.73         | 97.9±0.22    | 32.7±0.17    | 101.0±1.94                           |
| 400                   | 91.3±1.61         | 99.2±1.61    | 3.05±1.47    | 94.7±1.26                            |
| 800                   | 87.4±0.85         | 92.1±1.02    | 0.54±1.12    | 93.8±0.66                            |

Table 2: Melanin content of B16 mouse melanoma cells with different treatments (mean±standard deviation)

| Concentration (μmol/L) | Licochalcone A (%) | Licochalcone A and 50 μmol/L capsaicin (%) | β-arbutin (%) | β-arbutin and 50 μmol/L capsaicin (%) |
|-----------------------|-------------------|------------------------------------------|--------------|--------------------------------------|
| 0                     | 100±0.00          | 100±0.00                                 | 100±0.00     | 100±0.00                             |
| 50                    | 94.2±10.60        | 92.1±11.60                               | 90.9±5.24    | 125.6±3.39                           |
| 100                   | 89.4±2.88         | 95.4±6.93                                | 79.5±1.17    | 141.9±2.53                           |
| 200                   | 76.0±11.25        | 91.8±13.00                               | 70.5±8.64    | 125.6±3.65                           |
| 400                   | 65.3±2.70         | 93.2±7.34                                | 56.8±3.94    | 93.0±3.58                            |
| 800                   | 32.7±8.81         | 85.9±2.45                                | 43.2±2.76    | 83.7±3.08                            |
DISCUSSION

Currently, more and more consumers, especially women around the world, desire to brighten or whiten their skin. Consequently, with respect to dermal pigmentation, many active ingredients such as β-arbutin have been studied and applied to cosmetics. If the effects of cosmetics were inferior, many people would question their efficacy; however, undoubtedly, diet also probably has an important role. Interestingly, in an experiment in which B16 mouse melanoma cells were treated with β-Arbutin, the tyrosinase activity was obviously inhibited; however, the opposite result was observed when capsaicin and β-arbutin were used simultaneously.

Table 3: Tyrosinase activity in B16 mouse melanoma cells with different treatments (mean±standard deviation)

| Concentration (μmol/L) | Licochalcone A (%) | Licochalcone A and 50 μmol/L capsaicin (%) | β-arbutin (%) | β-arbutin and 50 μmol/L capsaicin (%) |
|-------------------------|--------------------|------------------------------------------|---------------|--------------------------------------|
| 0                       | 100.0±0.00         | 100.0±0.00                               | 100.0±0.00    | 100.0±0.00                           |
| 50                      | 88.7±2.08          | 119.3±3.85                               | 102.6±2.13    | 120.8±4.52                           |
| 100                     | 77.4±3.92          | 102.8±2.52                               | 68.8±2.26     | 96.5±1.64                            |
| 200                     | 69.6±4.48          | 94.6±4.07                                | 53.3±2.99     | 79.6±1.55                            |
| 400                     | 53.0±2.94          | 77.7±1.31                                | 49.4±2.40     | 74.6±3.54                            |
| 800                     | 41.2±1.27          | 61.6±1.03                                | 36.4±4.00     | 65.0±1.78                            |

Figure 1: Effects of licochalcone A (a) or β-arbutin (b) on melanogenesis in B16 mouse melanoma cells simultaneously treated with or without 50 μmol/L capsaicin. Melanin production was assessed by performing a NaOH assay, and cells were treated for 48 h. All data are shown as mean ± standard deviation (n = 3). *P < 0.05, **P < 0.001, compared to the control group.

Figure 2: Effects of licochalcone A (a) or β-arbutin (b) on tyrosinase activity in B16 mouse melanoma cells simultaneously treated with or without 50 μmol/L capsaicin. Tyrosinase activity was assessed by performing an L-DOPA-oxidation-assay 48 h after treatment. All data are shown as mean ± SD (n = 3); *P < 0.05, **P < 0.001, compared to the control group.
In this study, we discovered that when licochalcone A, 50 μmol/L capsaicin, and β-arbutin were used individually, the expression of tyrosinase was inhibited, and melanogenesis was reduced. However, capsaicin and licochalcone A/β-arbutin together enhanced the expression of tyrosinase when used simultaneously, which indicates that capsaicin can reverse the function of licochalcone A/β-arbutin with respect to inhibiting the expression of tyrosinase. Since it is an interesting phenomenon, which nobody reported it, our work is currently ongoing to identify the mechanism of this reversal effect.

**SUMMARY**

The results of our study serve as a reminder to pay attention to dietary factors, such as food that contain capsaicin, when using whitening cosmetics, as this compound potentially inhibits the whitening effect. It also provides theoretical evidence for drug incompatibility between whitening cosmetics and capsaicin that could be further explored in the future.

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**Conflicts of interest**

There are no conflicts of interest.

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