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Short Communication

Remdesivir–ivermectin combination displays synergistic interaction with improved in vitro activity against SARS-CoV-2

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A B S T R A C T

A key element for the prevention and management of coronavirus disease 2019 is the development of effective therapeutics. Drug combination strategies offer several advantages over monotherapies. They have the potential to achieve greater efficacy, to increase the therapeutic index of drugs and to reduce the emergence of drug resistance. We assessed the in vitro synergistic interaction between remdesivir and ivermectin, both approved by the US Food and Drug Administration, and demonstrated enhanced antiviral activity against severe acute respiratory syndrome coronavirus-2. Whilst the in vitro synergistic activity reported here does not support the clinical application of this combination treatment strategy due to insufficient exposure of ivermectin in vivo, the data do warrant further investigation. Efforts to define the mechanisms underpinning the observed synergistic action could lead to the development of novel treatment strategies.

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1. Introduction

At the time of writing, the World Health Organization (WHO) has reported more than 328 million cases of coronavirus disease 2019 (COVID-19) and more than 5.5 million deaths [1]. There remains a clear need for therapeutic strategies with activity against severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2). Potential therapeutic strategies may include the repurposing of existing drugs as well as the discovery of novel therapies. Thousands of clinical trials are currently underway, with therapeutic approaches involving direct-acting antivirals for the prevention of viral replication, and host-directed therapies aimed at mitigating against the disease pathology [2,3].

Combination therapies can offer several advantages over monotherapies. They have the potential to achieve greater efficacy, to increase the therapeutic index of drugs and to reduce the emergence of drug resistance. Strategies to identify effective combination therapies are emerging, with several laboratories reporting in vitro combination screens [4] and in vivo animal combination studies [5]. In a recent clinical trial, baricitinib administered in combination with remdesivir was found to be superior, and to elicit fewer adverse effects, compared with either drug in isolation [6]. Importantly, even in the absence of synergistic activity, an additive interaction between two drugs with separate mechanisms of action may profoundly reduce the speed at which drug resistance is established.

Both remdesivir and ivermectin have received attention for the treatment of COVID-19. Remdesivir is a prodrug C-adenosine nucleoside analogue that inhibits the viral RNA-dependent, RNA polymerase. Early in the pandemic, remdesivir was shown to display in vitro antiviral efficacy against SARS-CoV-2 [7]. In a double-blind, randomized, placebo-controlled trial, intravenous administration of

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remdesivir showed superiority relative to placebo in shortening the time to recovery in adults who were hospitalized with COVID-19 [8]. However, other studies indicated that its impact was negligible [9], and on 20 November 2020, WHO issued a conditional recommendation against the use of remdesivir in hospitalized patients (irrespective of disease severity) due to and absence of evidence supporting an improvement in survival or other outcomes in patients.

Ivermectin is an antiparasitic which is active against a wide range of parasites, including gastrointestinal roundworms, lungworms, mites, lice, hornflies and ticks [10]. Ivermectin is reported to exhibit broad-spectrum antiviral activity against a wide range of RNA and DNA viruses [11]. Recently, ivermectin was also shown to display antiviral activity against SARS-CoV-2 [12], but approved doses are not expected to be high enough to achieve in vitro-defined target exposures systemically [13]. Several clinical trials are now evaluating the potential of ivermectin for both prophylaxis and treatment of COVID-19, but low exposures make the anti-inflammatory and/or immunomodulatory mechanisms of action more plausible than direct antiviral activity of the monotherapy [14], particularly as studies with SARS-CoV-2 in Syrian golden hamsters showed an impact upon disease pathology in the absence of any effect on viral titres [15].

The authors found a synergistic interaction between remdesivir and ivermectin resulting in improved in vitro antiviral activity against SARS-CoV-2 using two distinct methodologies – determination of the fractional inhibitory concentration index (FICI) with isobologram analyses, and checkerboard combinations with SynergyFinder analyses. The data are discussed in the context of current therapeutic efforts against COVID-19.

2. Materials and methods

2.1. SARS-CoV-2 strain

SARS-CoV-2/Human/Liverpool/REMRQ0001/2020 was isolated from a nasopharyngeal swab from a patient in Liverpool and passed a further four times in Vero E6 cells. The mapped RNA sequence has been submitted to Genbank previously (Accession No. MW041156).

2.2. Vero E6 cell culture and plate preparation

Vero E6 cells were maintained in complete EMEM [EMEM supplemented with 10% heat-inactivated fetal bovine serum (Gibco; 10500-064) and 1% penicillin/streptomycin (Gibco; 15140-122)] in T175 flasks (Thermo Fisher Scientific, Waltham, MA, USA) at 37°C with 5% CO2. Cells were seeded in resting EMEM [EMEM supplemented with 10% heat-inactivated fetal bovine serum] at 1 × 105 cells/well in 96-well plates (Greiner Bio-one; 655090). Plates were incubated for 20 h at 37°C with 5% CO2 to allow the cells to reach 100% confluence. The resting minimal medium was removed, and the cells were used for downstream applications.

2.3. Concentration–response for remdesivir and ivermectin against SARS-CoV-2

Vero E6 cells were treated in triplicate with either drug in minimal medium at 25.00 µM, 8.33 µM, 2.78 µM, 0.93 µM, 0.31 µM, 0.10 µM and 0.03 µM (DMEM maintained at 0.25%) or control media, as appropriate. The plates were incubated at 37°C with 5% CO2 for 2 h. The minimal media containing the experimental compounds or the control media was then removed. Fifty microlitres of minimal media containing SARS-CoV-2 (MOI 0.05), 100 µL of 2× semi-solid media and 50 µL of minimal media containing experimental compounds and control media was added to each well, as appropriate. After 48 h, 4% v/v paraformaldehyde was added to each well and the plate was incubated for 1 h at room temperature. The medium was removed and cells were stained with crystal violet. Cells were washed three times with water, and cytopathic viral activity was determined by measuring the absorbance of each well at 590 nm using a Varioskan LUX microplate reader (Thermo Fisher Scientific).

Automated data quality control and data analyses were performed. For quality control, for the viral control, any well which had a log-transformed value that was 2 standard deviations above the mean of all log-transformed viral controls was excluded. Similarly, for the non-viral control, any well which had a log-transformed value that was 2 standard deviations below the mean of all log-transformed non-viral controls was excluded. If two or more wells were excluded on this basis for either control, the plate was voided and no further analysis was performed. Next, Z’ was calculated for each plate using the uninfected/untreated controls and infected/untreated controls according to Equation 1:

\[
Z' = 1 - \frac{3(\sigma_n + \sigma_v)}{|\bar{\mu}_n - \bar{\mu}_v|}
\]

where \(\sigma_n\) and \(\sigma_v\) represent the standard deviation of the non-viral and viral controls respectively, while \(\bar{\mu}_n\) and \(\bar{\mu}_v\) represent the corresponding means of these controls. Drug activity was expressed as a percentage of inhibition of viral growth relative to the uninfected/untreated control (100% inhibition of viral cytopathic activity) and the infected/untreated control (0% inhibition of viral cytopathic activity) on that plate. Half maximal effective concentration (EC50) and 90% maximal effective concentration (EC90) were calculated for each compound that generated a robust, converged four-parameter fit according to Equation 2:

\[
E = \frac{E_{\text{max}} \cdot C^4}{E_{50} \cdot C^6}
\]

where E is the drug effect at any given concentration (C), \(E_{\text{max}}\) is the maximal level of viral inhibition (0–100%), \(E_{50}\) is the concentration required to achieve half of this maximal inhibition, and \(C^6\) represents the hill slope which describes the steepness of the concentration–effect relationship.

Compounds that did not achieve ≥50% viral inhibition were deemed inactive without fitting. Concentrations that were deemed toxic, as evidenced by >20% (approximately two standard deviations of all data) drop in absorbance with concentration increase coupled with evidenced toxicity in drug controls, were excluded from fitting analysis.

2.4. FICI for remdesivir–ivermectin combinations against SARS-CoV-2

Following assessment of the inhibitory effect (EC50) of remdesivir and ivermectin monotherapy on the cytopathic viral activity of SARS-CoV-2, FICI was determined using the isobologram method developed by Berenbaum [16] using data from three independent biological replicates. Drug stocks were created in DMSO to provide a stock sufficient to produce a top concentration of 25 µM for each biological replicate. Drugs were combined to generate mixed ratios of 1:0, 0.8:0.2, 0.6:0.4, 0.4:0.6, 0.2:0.8 and 0.1:0. Fixed ratios were then diluted across a concentration range 1:2 (DMOS maintained at 1%) to generate concentration–response data for each ratio, as described previously. Ratio dilutions were performed in a single 2-ml deep-well plate, and added in parallel to three 96-well plates for each biological replicates. One additional plate which was not inoculated with virus was included to observe drug toxicity. Compound incubation and viral addition was performed as described above. Z’ was calculated and quality control was implemented as above. Interpretation of FICI (≤0.5 = synergistic; >4.0 = antago-
nistic; >0.5–4 = no interaction) was based on guidance provided by the Journal of Antimicrobial Chemotherapy [17].

2.5. Checkerboard combinations for remdesivir–ivermectin combinations against SARS-CoV-2

For robustness, a second method to assess pharmacodynamic drug combination interaction was utilized. Drug stocks were created by serial dilution. Compounds and controls were mixed 1:1 (DMSO maintained at 1%) to generate data for each combination alone and in combination. Remdesivir was studied at 10 μM, 5 μM, 2.5 μM, 1.25 μM and 0.63 μM and ivermectin was studied at 5 μM, 2.5 μM, 1.25 μM, 0.63 μM and 0.31 μM. These concentrations were selected as they were determined not to cause cell toxicity to Vero E6 cells. Ratio dilutions were performed in a single 2-ml deep-well plate, and added in parallel to three 96-well plates for each biological replicate. Compound incubation and viral addition was performed as described above. Z’ was calculated and quality control was implemented as described above. Data were analysed using SynergyFinder and a summary synergy score was generated (>10 synergistic, −10 to +10 additive, and <−10 antagonistic) [18].

3. Results

This study assessed the capacity of remdesivir and ivermectin combinations to inhibit the in vitro cytopathic activity of SARS-CoV-2. First, the activity of each compound in isolation was determined. For plates included in concentration–response analyses, the median signal to noise ratio was 29.3 and the median Z’ was 0.43 for concentration–response plates (Table 1). For each compound, a robust four-parameter fit was generated (Figure 1). EC50 was 2.4 ± 1.1 μM for ivermectin and 1.3 ± 2.1 μM for remdesivir (geometric mean ± geometric standard deviation).

Next, the combination interaction between remdesivir and ivermectin was determined by isobologram. The median signal to noise ratio was 26.4 and the median Z’ was 0.49 for isobologram plates (Table 1). The 0.2:0.8 [remdesivir:ivermectin (5 μM:20 μM)] ratio, 0.4:0.6 ratio (10 μM:15 μM) and 0.6:0.4 (15 μM:10 μM) ratio demonstrated synergy (FIC<0.5) across all three biological replicates (Figure 2). For the 0.8:0.2 (20 μM:5 μM) ratio, just one biological replicate met the defined threshold of synergy (Figure 2A). The other two biological replicates did, however, exceed the predicted effect assuming a purely additive relationship (Figure 2A).

The synergistic interaction was confirmed using interaction potency models using the SynergyFinder platform [18]. The median signal to noise ratio was 23.6 and the median Z’ was 0.62 for checkerboard plates (Table 1). All four integrated synergy models determined that interactions between remdesivir and ivermectin were synergistic with synergy scores that far exceeded the threshold for synergy (Table 2 and Figure 2B).

4. Discussion

This study found a synergistic interaction between remdesivir and ivermectin, both approved by the US Food and Drug Administration, resulting in enhanced in vitro antiviral activity against SARS-CoV-2. Although combination therapy offers a number of advantages compared with monotherapy, genuine descriptions of synergy are relatively infrequent [19]. Despite thousands of combination experiments having been performed, there have been very
few reports of validated synergistic interactions against SARS-CoV-2 [4,20].

At this stage, the mechanism underpinning the synergistic interaction between remdesivir and ivermectin is unclear; however, both drugs have previously been shown to inhibit SARS-CoV-2 replication [7,12]. Given that remdesivir is known to inhibit the RNA-dependent, RNA polymerase [21], it will be of interest to investigate whether ivermectin confers synergy by inhibiting an undefined alternative but complimentary role in RNA synthesis. Ivermectin has been shown to inhibit replication of HIV-1 and dengue through inhibition of importin-β-mediated nuclear transport [22]. In silico predictions suggest that ivermectin may interact with host-cell proteins such as importins, which are required for nuclear transport, as well as viral proteins, including Nsp13 helicase and Mpro protease, which facilitate replication and translation of SARS-CoV-2 [23]. Further mechanistic studies will be required to determine the validity of in silico predictions.

Special care was taken to assess in vitro activity across concentrations that likely cover the physiological exposure of remdesivir and ivermectin in human plasma and lung tissue. In humans, a single 225-mg dose of remdesivir has been shown to produce a plasma Cmax of approximately 4000 ng/mL [24], exceeding its in vitro EC50 (1.3 ± 2.1 μM). In humans, a high dose of 600 μg/kg/day of ivermectin has been shown to produce a plasma Cmax of 120 ng/mL [25], which is much less than its in vitro EC50 (2.4 ± 1.1 μM). The Cmax of remdesivir in lung epithelial lining fluid (ELF) has not been established, and it is likely that these concentrations are important in terms of clinical activity. Poor exposure in lung ELF may well explain the limited impact of remdesivir in the clinic [8]. Interestingly, concentrations of ivermectin are predicted to be some three-fold higher in the lung than in plasma [28]; however, even at these levels, ivermectin fails to meet its in vitro EC50 and no data are presented here, or elsewhere, that would support the clinical application of ivermectin for the treatment of SARS-CoV-2 infection. Given that 88–93.6% of remdesivir [27] and 93.2% of ivermectin [28] is protein-bound, the availability of unbound drug at target sites is predicted to be considerably less than the reported values based on total drug concentrations.

Data presented here demonstrate that remdesivir administered in combination with ivermectin enhances in vitro antiviral activity. As described above, with respect to ivermectin, due to insufficient exposure of unbound drug at the target site, this combination strategy does not represent a clinically tractable therapeutic strategy. In addition, the differing routes of administration would likely impact the ability to achieve therapeutic concentrations of both drugs simultaneously. Further investigations are now required to determine whether the observed synergistic interaction can be replicated in animal disease models and with drugs that share similar modes of action, such as, for example, the orally bioavailable polymerase inhibitors, favipiravir or molnupiravir. The underpinning mechanisms for this synergy warrant further investigation so that this pharmacodynamic phenomenon can be exploited for the development of optimal drug combinations.

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**Table 2**

| SynergyFinder synergy score summary table for remdesivir and ivermectin. |
|---------------------------|---------------------|---------------------|---------------------|
|                          | Mean synergy score  | Median (range)       |
| ZIP                      | 35.33 (28.01–40.84) |                     |
| HSA                      | 40.25 (36.02–44.34) |                     |
| Leowze                   | 26.34 (26.04–30.45) |                     |
| Bliss                    | 37.77 (27.61–41.69) |                     |

ZIP, zero interaction potency; HSA, highest single agent.
Competing interests

AO is a Director of Tandem Nano Ltd. AO has received research funding from ViIV, Merck and Janssen, and consultancy fees from Gilead. These associations had no influence on the content of the current manuscript. PON is currently engaged in a collaboration with Romark LLC, but this interaction had no influence on the content of the current manuscript. No other conflicts of interest are declared by the authors.

Ethical approval

Not required.

Supplementary materials

Supplementary material associated with this article can be found, in the online version, at doi:10.1016/j.jantimicrob.2022.106542.

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