Commentary

The Role of Cyclooxygenase Inhibition in the Antineoplastic Effects of Nonsteroidal Antiinflammatory Drugs (NSAIDs)

By Steven J. Shiff and Basil Rigas

From The Rockefeller University, New York, New York 10021

According to WHO statistics, in 1996 875,000 new cases of colorectal cancer were diagnosed and 495,000 deaths were attributed to it worldwide (1). It is the third most common cause of cancer deaths and its incidence and mortality rates continue to rise.

Most colorectal cancers develop via a characteristic series of pathological steps (Fig. 1). Colorectal epithelial cells acquire abnormal growth and morphological characteristics and form an adenoma, a tumor mass often protruding into the lumen of the colon or rectum. In time, these lesions enlarge and a subset of cells can acquire additional abnormal growth behaviors, which allows them to invade into the bowel wall and metastasize. At this point, the tumors are classified as adenocarcinomas and can be lethal. In the course of this typical multistep process, somatic mutations develop in key genes in the cells that comprise these lesions, such as in: the adenomatous polyposis coli (APC) and p53 tumor suppressor genes; the Ki-ras oncogene; and various genes that mediate DNA mismatch repair (2). The mutational events presumably drive the evolution of the pathological lesions and their neoplastic and malignant behavior.

Considerable evidence supports the view that aspirin (ASA) and other nonsteroidal antiinflammatory drugs (NSAIDs) prevent colorectal cancer (3). A combination of epidemiological, animal, and basic studies make a compelling case that regular use of these compounds lowers the risk for the development of colorectal cancer, as well as adenomas. Recently, sulindac was also shown to eliminate aberrant crypt foci in the colorectum of patients who have had adenomatous polyps. These lesions are similar to adenomas but are not yet visible to the naked eye, and probably grow to become adenomas (4). Overall, the weight of the evidence indicates that NSAIDs are the preeminent colorectal cancer chemopreventive agents. It is of paramount importance, then, to identify their mechanism of action in the chemoprevention of colorectal cancer.

NSAIDs inhibit the catalytic activity of cyclooxygenase (COX, more properly called prostaglandin H synthase, or PGHS), and this is thought to be the predominant mechanism by which they act as analgesic, antipyretic, and antiinflammatory agents (Fig. 2). Reports indicating that ASA and other NSAIDs inhibit colon carcinogenesis provided the impetus for a series of studies showing that the levels of both PGE₂ and COX-2 are augmented in human colon cancer (for review see reference 3). As a result, it generally has been assumed that the antineoplastic effects of NSAIDs were dependent upon inhibition of COX activity and PG synthesis. Yet evidence is mounting that NSAIDs produce some of their clinical and experimental cancer chemopreventive effects via mechanisms that are independent of COX inhibition.

In this issue of The Journal of Experimental Medicine, Zhang et al. provide additional compelling evidence for COX-independent chemopreventive effects of NSAIDs (5). They accomplish this by demonstrating that NSAID-induced inhibition of cell transformation, inhibition of cell proliferation, and initiation of apoptosis are not dependent on the expression of COX isozymes. By genetic manipulation of mouse embryos, they developed fibroblasts in which COX-1 or COX-2 or both COX-1 and COX-2 mRNAs and proteins were absent. The resulting three types of cell lines were equally capable of undergoing oncogenic transformation induced by ras and SV-40 oncogenes. In addition, the proliferation of transformed versions of these three cell types was inhibited and apoptosis was stimulated equally by various NSAIDs. Both non-COX-selective NSAIDs (i.e., sulindac, ibuprofen, piroxicam, and indomethacin) and the selective COX-2 inhibitor, NS-398 produced these effects in all three types of cell lines. Thus, targeted disruption of COX genes does not affect these NSAID-induced events related to transformation and cell turnover kinetics.

A few years ago, we demonstrated that NSAIDs inhibited the proliferation and induced apoptosis in human colon cancer cell lines (6, 7). Since this effect was obtained in medium supplemented with serum rich in PGs, we pursued this observation further to assess its dependence upon PGs and COX. In due course, a colon cancer cell line, HCT-15, which was intrinsically deficient in COX-1 and COX-2 expression and failed to produce any prostanoids even upon stimulation with arachidonic acid, meilitin, and the calcium ionophore A23187 was identified (8). NSAIDs had effects on proliferation and apoptosis in these cells similar to those in cells that express both COX enzymes and produce several PGs, such as HT-29. These observations, reinforced by subsequent studies demonstrating that compounds with meager COX inhibitory activity (e.g., sulindac sulfone, and salicylic acid [SA]) also inhibit cell proliferation and induce apoptosis in colonocytes, led to the hypothesis that COX-independent mechanisms account for at least some of the chemopreventive actions of...
Commentary

NSAIDs (3). A trivial explanation for our results and those of Zhang et al. is that there exist additional undiscovered COX isoforms that compensate for the loss of the known cyclooxygenases. However, this is practically untenable, because COX-1 and COX-2 null fibroblasts and HCT-15 cells do not produce PGs, a defining characteristic of COX enzymes; as well, HCT-15 cells do not produce PGs even upon stimulation. Regardless of this, the evidence should and already has aroused suspicion that NSAIDs may induce their chemopreventive effects by targeting other proteins or pathways, beyond the cyclooxygenases. Because of the probable importance of these proteins or pathways in preventing neoplasia, many laboratories are now actively working to identify them.

Several selected observations regarding the antineoplastic actions of NSAIDs are briefly reviewed below. They are broadly classified into three major categories according to their dependence on COX inhibition: (i) COX-dependent or COX-related; (ii) COX-independent; and (iii) uncertain, i.e., those that do not appear to involve COX, although their COX independence is not yet proven (Table I).

**COX-dependent Effects**

For the antineoplastic effects of NSAIDs to be exerted through COX inhibition, COX and PGs must contribute substantially to colorectal carcinogenesis. Evidence that eicosanoids and COX isozymes are important in colorectal cancer development includes: (a) PGE$_2$ levels are elevated in colorectal tumors; (b) eicosanoids, including PGs, stimulate proliferation and reduce apoptosis in colonocytes; (c) COX-2 is upregulated in colorectal tumors, more frequently in cancers than adenomas; and (d) COX-2 contributes to colorectal tumorigenesis in APC knockout mice (9) where APC$^{D716(1/2)}$COX-2$^{2/2}$ mice develop fewer intestinal tumors than do APC$^{D716(1/2)}$COX-2$^{1/1}$ mice, which in turn bear fewer tumors than do APC$^{D716(1/2)}$COX-2$^{2/2}$ mice (for review see reference 3). Aside from producing several eicosanoids, COX may promote carcinogenesis by activating carcinogens via its peroxidase activity, which can operate on substrates other than PGG$_2$; or by producing either malondialdehyde, a direct-acting mutagen, or peroxyl radicals (see Fig. 2).

Figure 2. Arachidonic acid metabolism by COX isozymes. Phospholipase A$_2$ (PLA$_2$) releases arachidonic acid (AA) from membrane phospholipids, which is in turn converted by either COX-1 or COX-2 to PGG$_2$ (C, cyclooxygenase catalytic activity of COX) and then to PGH$_2$ (P, peroxidase catalytic activity of COX). PGH$_2$ is converted to either PGs (e.g., E$_2$, F$_{2\alpha}$, I$_2$, D$_2$); thromboxane A$_2$ (TxA$_2$); or malondialdehyde (MDA). MDA is a direct-acting mutagen and carcinogen and can be produced without COX by direct lipid peroxidation. AA can be converted directly to 15-(R)-HETE by both COX isozymes. COX-1 is constitutively expressed in most tissues, whereas COX-2 is induced by cytokines, growth factors, tumor promoters, or other agents after the initiation of specific physiological events. Compounds other than PGG$_2$, e.g., procarcinogenic hydroperoxides, can serve as substrates for the peroxidase activity of both COX enzymes. Inactive carcinogens serving as electron acceptors can also become activated by this activity. The COX isozymes are also involved in the formation of peroxyl radicals that can activate procarcinogens. Figure is adapted from reference 3.

Angiogenesis, which is important in carcinogenesis, is related to PGs and COX. PGE$_2$ and COX-2 induce angiogenesis (10) and NSAIDs inhibit it (11). Clearly, inhibition...
of angiogenesis could explain the ability of NSAIDs to regress adenomas in familial adenomatous polyposis (FAP) patients and to inhibit colorectal adenocarcinoma formation. However, like most if not all of the mechanisms discussed here, it has yet to be formally demonstrated in vivo that angiogenesis mediates the chemopreventive effects of NSAIDs in colorectal cancer. It is noteworthy that sulindac sulfone also inhibits angiogenesis (12). This raises the possibility of PG- or COX-independent mechanisms of angiogenesis as well.

Whether COX or eicosanoids are necessary or essential for colorectal carcinogenesis has not been fully assessed. Clearly, some colorectal cancers develop without overexpressing COX or producing high levels of PGs. However, this still does not rule out a central role for COX enzymes, at least for most colorectal cancers.

COX-independent Effects

Cell Turnover. A potentially important means by which NSAIDs prevent colorectal neoplasia is to affect cell turnover in the colorectal epithelium. Cell death and renewal are critical for the regulation of the structural integrity of all tissues. The growth rate of a tissue or a tumor is determined by the rate of proliferation and counterbalanced by the rates of cell loss by apoptosis or necrosis of the cells that comprise them. We and others have shown that several NSAIDs, including SA, ASA, sulindac (and its metabolites sulindac sulfide and sulindac sulfone), indomethacin, piroxicam, naproxen, as well as selective COX-2 inhibitors, retard the proliferation and induce apoptosis in colon cancer cells (3, 13).

Human studies assessing the effects of NSAIDs on colonocytic proliferation have generated conflicting results, with sulindac reported either to show no effect upon proliferation or to reduce it (3). Apoptosis is suppressed in sporadic colorectal adenomas and carcinomas and in the flat mucosa or adenomas of patients with FAP. In FAP patients, sulindac normalizes apoptosis in normal rectal mucosal colonocytes while reducing the size and number of their adenomas (for review see reference 3). Thus, it is possible, but not yet fully substantiated, that cell kinetic effects play a major role in the antineoplastic effects of these compounds.

As we have shown previously, and as is now described in additional detail by Zhang et al., these events in vitro are clearly not dependent upon the expression of COX isozymes (5, 8). NSAIDs inhibit cell proliferation by inducing cell cycle quiescence in colonocytes, in part by reducing the levels of several key molecules that catalyze transitions through the various phases of the cell division cycle (3, 6, 7). However, the detailed molecular pathways that induce quiescence have yet to be fully elucidated.

Several groups contend that they have identified mechanisms by which NSAIDs induce PG- or COX-independent apoptosis. Preliminary reports claim that sulindac sulfone induces this form of cell death by inhibiting cGMP-dependent phosphodiesterase 5 (14). Others have shown that NSAID treatment of colon cancer cells generates the proapoptotic lipid, ceramide (15). By blocking the biosynthesis of prostanoids, NSAIDs increase the intracellular levels of arachidonic acid, which activates neutral sphingomyelinase, which in turn converts sphingomyelin to ceramide. Strictly speaking, this is a PG-independent mechanism, but, as presented, this model of ceramide formation is dependent upon COX inhibition and therefore may not explain the apoptosis induced by sulindac sulfone or SA, nor that induced in COX null cells, like HCT-15 or those generated by Zhang et al.

Cell Transformation. Aside from Zhang et al., other groups have also reported that NSAIDs inhibit cell transformation. Dong et al. showed that salicylates inhibited phorbol ester (TPA)-induced transformation of mouse epidermal JB6 cells (16). Hermann et al. demonstrated that sulindac sulfide inhibited transformation of primary rat embryo fibroblasts by activated H-ras and SV40 T antigen or other transformation-inducing stimuli (17). In both cases, transformation was inhibited at drug concentrations below those required to inhibit cell proliferation or cell viability. Dong et al. proposed that AP-1 activation is important for

| COX-dependent | COX-independent |
|---------------|-----------------|
| Cell turnover  | Cell turnover    | COX dependence unclear |
| proliferation/apoptosis | proliferation/apoptosis |
| Carcinogen formation | Cell transformation |
| Angiogenesis    | DNA repair       |
|               | Angiogenesis     |
|               | Ras signal transduction |
|               | MAP kinase activation |
|               | NF-kB activation |

The table shows the cellular or molecular mechanisms influenced by NSAIDs that may contribute to their antineoplastic effects, classified in relation to their dependence on inhibition of cyclooxygenase catalytic activity.
this effect of salicylates. This process appeared to be independent of PG inhibition, as indomethacin could not inhibit TPA-induced transformation, nor AP-1 activity. Mitogen-activated protein (MAP) kinase was shown not to be involved in this process. Hermann et al. proposed that sulindac sulfide inhibits oncogenic cell transformation by directly inhibiting R as signaling. They showed that sulindac sulfide binds noncovalently to R as and inhibits R as-dependent R af binding and R af activation without affecting its GT Pase or GTP binding activity.

Apart from its role in cell transformation, R as influences many pathways potentially important for the chemopreventive activity of NSAIDs, and thus it may be a pivotal target molecule integrating several disparate pathways influenced by NSAIDs. Inhibition of R as signaling may also explain the effects of NSAIDs on proliferation and apoptosis at high drug concentrations. R as inhibition may link the effect of NSAIDs to NF-kB and MAP kinase activity (see below) (18). Modulation of R as activity by NSAIDs may also relate to the eicosanoid pathway, as arachidonic acid and PGs regulate R as regulatory proteins such as p120GAP and N F-1GAP (19).

DNA Repair. Both ASA and non-salicylate NSAIDs reduce microsatellite instability in colon cancer cell lines deficient in DNA mismatch repair (i.e., HCT-116, HCT-15, SW-48, and LoVo cells) (20). Mismatch repair-deficient cells with mutations in hMLH1, hMSH2, or hMSH6, but not with a hPM2 S2 mutation, die selectively by apoptosis in response to NSAID treatment. Given that HCT-15 cells were used in this study, which we found to be deficient in COX expression (8), this effect is also likely to be COX independent. However, Ruschoff et al. (20) did not directly confirm the absence of COX isozymes or PG production in the cells they used. Patients with hereditary nonpolyposis colorectal cancer have germline mutations in DNA mismatch repair genes. Therefore, it was speculated that NSAIDs could prevent colon cancer in most of these patients. However, the clinical efficacy of NSAIDs in this particular high-risk group remains unknown.

MAP Kinases. SA activates p38 MAP kinase and induces apoptosis in FS-4 fibroblasts, and both are inhibited by the p38 MAP kinase inhibitor SB-203580. p38 MAP kinase activation may also be important for inhibition of nuclear factor (N F-)K B by SA (21).

N F-1 K B. ASA and SA, but not indomethacin, inhibit N F-1 K B activation (21, 22). There is evidence that these compounds bind to and inactivate I K B kinase β, which in turn prevents the degradation of I K B and the subsequent translocation of N F-1 K B to the nucleus, where it activates the transcription of a variety of genes (22). Depending on the cell type and the circumstances, N F-1 K B augments or inhibits apoptosis. Inhibition of the proliferation of ras-transformed rat fibroblasts by ASA may also be related to inhibition of N F-1 K B activation (18).

To show that activation of MAP kinases or inhibition of N F-1 K B activation are COX-independent processes, Yin et al. demonstrated that each is influenced exclusively by salicylates, and not by indomethacin (22). However, indomethacin and other non-salicylate NSAIDs prevent colon neoplasias and regress adenomas in FAP patients. As a result, it is unlikely that these two processes, as delineated in these experiments, play a major role in the chemopreventive effects of NSAIDs.

Role of COX Unclear

The observations that follow relate to processes that do not appear related to PGs or COX inhibition. However, since they have not been examined in detail in cells that lack COX enzymes or PG production, their relationship to COX and/or arachidonic acid metabolism remains uncertain.

Myc and N F-cOx 1 N c o p r o t e i n s . Lu et al. found that, in addition to apoptosis, Myc was markedly induced in serum-starved chicken embryo fibroblasts after activation of pp60-c-src and NSAID treatment. Apoptosis was markedly inhibited by transfection of antisense myc (23). Similar results were found in other cancer cell lines.

 Peroxisomal Proliferator-activated Receptors. Peroxisomal proliferator-activated receptors (PPARs) are a group of nuclear hormone receptor protein transcription factors that, when stimulated, induce differentiation of fibroblasts to adipocytes. PPAR-γ receptors are expressed in the colon and to an even greater degree in colon tumors (24). Activation of PPARs in colon cancer cells reduces their growth and induces differentiation in vitro (25). Interestingly, indomethacin and selected, but not all, NSAIDs bind to and activate PPAR-γ receptors (26). However, piroxicam, which regresses colorectal polyps in humans and animals, does not bind PPAR-γ as effectively as other NSAIDs (26) and PPAR-γ activators, such as troglitazone, increase colon tumors in APCmin mice (27, 28). These latter data suggest it is unlikely that PPAR activation is important for the chemoprevention of colorectal cancer by NSAIDs.

Tumor Immunity. There is ample indirect evidence for a role of NSAIDs, and by extension of COX enzymes, in immune phenomena related to various cancers. A clearly studied case concerns their role in the expression of HLAs, which is altered in many cancers and frequently downregulated in colon cancer (3). Such abnormalities may adversely affect the clinical course of cancer and the outcome of T cell–based immunotherapy.

NSAIDs may boost mechanisms of tumor immune surveillance: tumors are hypothesized to escape from immune-mediated destruction by thwarting mechanisms that detect tumor-associated antigens. Class I and II HLA antigens, participating in antigen presentation, may be critical to this process. The role of NSAIDs in these processes has been assessed in animal models of colon cancer and in cell culture systems. Piroxicam upregulates the expression of MHC genes in the colon mucosa of rats treated with a carcinogen (3). PGE2 reduces the transcription of HLA class II molecules and NSAIDs can increase it (29). That such effects occur in the presence of PG-rich serum in cell culture systems indicates, along with additional evidence (Ri-
Ent inconsistencies are not merely of theoretical interest, but are of presumptive chemopreventive effects of NSAIDs. This apparent inconsistency is not merely of theoretical interest, but is also of great significance for the rational design of strategies for colon chemoprevention and for assessing the relative significance of each mechanism in carcinogenesis. In all likelihood this is not a contradiction at all; rather, NSAIDs bring about their chemopreventive effects in the colon through both COX-dependent and -independent mechanisms. Indeed, we have proposed a model that assumes both mechanisms operate to produce the clinical antineoplastic effects of NSAIDs, in which COX-dependent and -independent pathways modulate different stages of the multistep process of colon carcinogenesis or different events regulating each stage (30). The multiplicity of action of NSAIDs, if confirmed, could in fact explain their high degree of effectiveness in colon cancer prevention in humans.

The great challenge will be to determine which of these or other yet unknown mechanisms produce the remarkable antitumor effects of NSAIDs, as well as the relative contribution of each. If and when these key questions are worked out, then a great deal will have been learned about colorectal carcinogenesis. In addition, new tools will have been collected to identify new compounds that hold promise to prevent colorectal cancer more safely and effectively than the conventional NSAIDs.

As this brief overview demonstrates, multiple processes with their attendant molecular pathways have been proposed to mediate the chemopreventive effects of NSAIDs (Table I). The NSAIDs, highly protein-bound molecules, probably interact with and inhibit the function of many proteins and, perhaps, other macromolecules. To complicate matters, it is not clear whether the key in vitro effects of NSAIDs that were discussed earlier, such as cell transformation, cell growth, or angiogenesis, are at all relevant to colorectal cancer chemoprevention.

Some of the data reviewed here indicate that COX inhibition by NSAIDs is indeed required for their chemopreventive effect. However, another body of data, including that of Zhang et al. in this issue (5), is consistent with this, if not required for certain presumed chemopreventive effects of NSAIDs. This apparent inconsistency is not merely of theoretical interest, but has important implications for the rational design of strategies for colon chemoprevention and for assessing the relative significance of each mechanism in carcinogenesis. In all likelihood this is not a contradiction at all; rather, NSAIDs bring about their chemopreventive effects in the colon through both COX-dependent and -independent mechanisms. Indeed, we have proposed a model that assumes both mechanisms operate to produce the clinical antineoplastic effects of NSAIDs, in which COX-dependent and -independent pathways modulate different stages of the multistep process of colon carcinogenesis or different events regulating each stage (30). The multiplicity of action of NSAIDs, if confirmed, could in fact explain their high degree of effectiveness in colon cancer prevention in humans.

The great challenge will be to determine which of these or other yet unknown mechanisms produce the remarkable antitumor effects of NSAIDs, as well as the relative contribution of each. If and when these key questions are worked out, then a great deal will have been learned about colorectal carcinogenesis. In addition, new tools will have been collected to identify new compounds that hold promise to prevent colorectal cancer more safely and effectively than the conventional NSAIDs.

We thank the Arthur and Rochelle Belfer Foundation and the National Institutes of Health (grant CA73298) for funding.

Address correspondence to Steven J. Shiff, The Rockefeller University, 1230 York Ave., Box 330, New York, NY 10021-6399. Phone: 212-327-7458; Fax: 212-327-7453; E-mail: shiffs@rockvax.rockefeller.edu

Submitted: 6 July 1999 Accepted: 13 July 1999

References

1. WHO. 1997. The World Health Report. World Health Organization, Geneva, Switzerland.

2. Kinzler, K.W., and B. Vogelstein. 1996. Lessons from hereditary colorectal cancer. Curr. Opin. Genet. Dev. 6:159–170.

3. Shiff, S.J., and B. Rigas. 1997. Nonsteroidal anti-inflammatory drugs and colorectal cancer: evolving concepts of their chemopreventive actions. Gastroenterol. 113:1992–1998.

4. Takayama, T., S. Katsuki, Y. Takahashi, M. Ohi, S. Nojiri, S. Sakamaki, J. Kato, K. Kobayashi, H. Miyake, and Y. Niiitsu. 1998. Aberrant crypt foci of the colon as precursors of adenoma and cancer. N. Engl. J. Med. 339:1277–1284.

5. Zhang, X., S.G. Morham, R. Langenbach, and D.A. Young. 1999. Malignant transformation and aneuploidy actions of nonsteroidal antiinflammatory drugs (NSAIDs) on cyclooxygenase-null embryo fibroblasts. J. Exp. Med. 190: 451–459.

6. Shiff, S.J., L. Qiao, L.-L. Tsai, and B. Rigas. 1995. Sulindac sulfide, a aspirin-like compound, inhibits proliferation, causes cell cycle quiescence, and induces apoptosis in HT-29 colon adenocarcinoma cells. J. Clin. Invest. 96:491–503.

7. Shiff, S.J., M.I. Koutsos, L. Qiao, and B. Rigas. 1996. Nonsteroidal antiinflammatory drugs inhibit the proliferation of colon adenocarcinoma cells: effects on cell cycle and apoptosis. Exp. Cell Res. 222:179–188.

8. Hanif, R., A. Pittas, Y. Feng, M.I. Koutsos, L. Staiano-Coico, S.J. Shiff, and B. Rigas. 1996. Effects of nonsteroidal anti-inflammatory drugs on proliferation and on induction of apoptosis in colon cancer cells by a prostaglandin-independent pathway. Biodem. Pharmaol. 52:237–245.

9. Oshima, M., J.E. Dinchuk, S.L. Kargman, H. Oshima, B. Hancock, E. Kwong, J.M. Trzaskos, J.F. Evans, and M.M. Taketo. 1996. Suppression of intestinal polyposis in Apc<sup>min</sup> knockout mice by inhibition of cyclooxygenase 2 (COX-2). Curr. Opin. Genet. Dev. 8:803–809.

10. Tsujii, M., S. Kawano, S. Tsuji, H. Sawaoka, M. Hori, and R.N. DuBois. 1998. Cyclooxygenase regulates angiogenesis in colon adenocarcinoma cells. Anticancer Res. 18:93–100.

11. Peterson, H.I. 1986. Tumor angiogenesis inhibition by prostaglandin synthetase inhibitors. Anticancer Res. 6:251–253.

12. Skopinska-Rózewska, E., G.A. Piazza, E. Sommer, R. Pamukcu, E. Barcz, M. Fillewska, W. Kupis, R. Caban, P. Rudzinski, J. Bogdan, et al. 1998. Inhibition of angiogene-
sis by sulindac and its sulfone metabolite (FGN-1): a potential mechanism for their antineoplastic properties. Int. J. Tis-
tissue Res. 20:85–89.
13. Elder, D.J., D.E. Halton, A. Hague, and C. Paraskeva. 1997. Induction of apoptotic cell death in human colorectal
carcinoma cell lines by a cyclooxygenase-2 (COX-2)-selective
nonsteroidal anti-inflammatory drug: independence
from COX-2 protein expression. Clin. Cancer Res. 3:1679–
1683.
14. Piazza, G.A., H. Li, L. Liu, G. Sperl, R. Pamukcu, W.J.
Thompson, and D.J. Ahnen. 1999. Cyclic GMP (CG)
phosphodiesterase (PDE) inhibition: a novel mechanism
for the antineoplastic properties of exisulind. Gastroenterology.
116:A485.
15. Chan, T.A., P.J. Morin, B. Vogelstein, and K.W. Kinzler.
1998. Mechanisms underlying nonsteroidal antiinflamma-
tory drug-mediated apoptosis. Proc. Natl. Acad. Sci. USA.
95:681–686.
16. Dong, Z., C. Huang, R.E. Brown, and W.Y. Ma. 1997.
Inhibition of activator protein 1 activity and neoplastic
transformation by aspirin. J. Biol. Chem. 272:9962–9970.
17. Herrmann, C., C. Block, C. Geisen, K. Haas, C. Weber,
G. Windle, T. Möroy, and O. Müller. 1998. Sulindac sul-
fide inhibits Ras signaling. Oncogene. 17:1769–1776.
18. Ljungdahl, S., M.C. Shoshan, and S. Linder. 1997. Inhibi-
tion of the growth of 12V-ras-transformed rat fibroblasts by
acyetylsalicylic acid correlates with inhibition of NF-κB.
Anticancer Drugs. 8:62–66.
19. Han, J.-W., F. McCormick, and I.G. Macara. 1991. Regu-
lation of ras-GAP and the neurofibromatosis-1 gene prod-
uct by eicosanoids. Science. 252:576–579.
20. Rüschoff, J., S. Wallinger, W. Dietmaier, T. Bocker,
G. Brockhoff, F. Hofstadter, and R. Fishel. 1998. Aspirin sup-
presses the mutator phenotype associated with hereditary
nonpolyposis colorectal cancer by genetic selection. Proc.
Natl. Acad. Sci. USA. 95:11301–11306.
21. Schwenger, P., D. Alpert, E.Y. Skolnik, and J. Vilcek.
1998. Activation of p38 mitogen-activated protein kinase
by sodium salicylate leads to inhibition of tumor necrosis
factor-induced iκB α phosphorylation and degradation.
Mol. Cell. Biol. 18:78–84.
22. Yin, M.J., Y. Yamamoto, and R.B. Gaynor. 1998. The
anti-inflammatory agents aspirin and salicylate inhibit the
activity of iκB kinase-β. Nature. 396:77–80.
23. Lu, X., D.W. Fairbarn, W.S. Bradshaw, K.L. O’Neill,
D.L. Ewert, and D.L. Simmons. 1997. NSAID-induced
apoptosis in Rous sarcoma virus-transformed chicken em-
broyo fibroblasts is dependent on v-src and c-myc and is
inhibited by bcl-2. Prostaglandins. 54:549–568.
24. DuBois, R.N., R. Gupta, J. Brockman, B.S. Reddy, S.L.
Krakow, and M.A. Lazar. 1998. The nuclear eicosanoid re-
ceptor, PPAR-γ, is aberrantly expressed in colonic cancers.
Carcinogenesis. 19:49–53.
25. Sarraf, P., E. Mueller, D. Jones, F.J. King, D.J. DeAngelo,
J.B. Partridge, S.A. Holden, L.B. Chen, S. Singer, C.
Fletcher, and B.M. Spiegelman. 1998. Differentiation and
reversal of malignant changes in colon cancer through
PPAR-γ. Nat. Med. 4:1046–1052.
26. Lehmann, J.M., J.M. Lenhard, B.B. Oliver, G.M. Ringold,
and S.A. Kliewer. 1997. Peroxisome proliferator-activated
receptors alpha and gamma are activated by indomethacin
and other non-steroidal anti-inflammatory drugs. J. Biol.
Chem. 272:3406–3410.
27. Saez, E., P. Tontonoz, M.C. Nelson, J.G. Alvarez, U.T.
Ming, S.M. Baird, V.A. Thomazy, and R.M. Evans. 1998.
Activators of the nuclear receptor PPAR-γamma enhance
colon polyp formation. Nat. Med. 4:1053–1061.
28. Lefebvre, A.-M., I. Chen, P. Desreumaux, J. Najib, J.-C.
Fruchart, K. Geboes, M. Briggs, R. Heyman, and J. Auw-
erx. 1998. Activation of the peroxisome proliferator-activated
receptor gamma promotes the development of colon
tumors in C57BL/6J-APC−/− mice. Nat. Med. 4:1053–
1057.
29. Arvind, P., E.D. Papavassiliou, G.J. Tsioulias, L. Qiao,
C.I.P. Lovelace, and B. Rigas. 1995. PGE2 down-regulates
the expression of HLA-DR antigen in human colon ade-
nocarcinoma cell lines. Biodrochemistry. 34:5604–5609.
30. Rigas, B., and S.J. Shiff. 1999. Is inhibition of cyclooxyge-
nase required for the chemopreventive effect of NSAIDs in
colon cancer? A model reconciling the current contradic-
tion. Med. Hypotheses. In press.