Complete Genome Sequence of *Vibrio mediterranei* 117-T6, a Potentially Pathogenic Bacterium Isolated from the Conchocelis of *Pyropia* spp.

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**ABSTRACT** *Vibrio mediterranei* is a Gram-negative bacterium of the family *Vibrionaceae*. *Vibrio mediterranei* strain 117-T6 was pathogenic to *Pyropia yezoensis*, a red seaweed cultivated in China, by causing death to its conchocelis. Here, we report the complete genome sequence of *Vibrio mediterranei* strain 117-T6.

The genus *Vibrio* is a major group of the family *Vibrionaceae*, and its members are widespread in marine environments, or associate with hosts in those environments, and can be pathogenic or nonpathogenic (1, 2). Since *Vibrio mediterranei* was first described by Pujalte and Garay in 1986 (3), the bacterium has been isolated from various marine organisms, such as plants, shellfish, other marine invertebrates, and fish from different geographic regions (4–6). *V. mediterranei* 117-T6 was isolated from the bleached shell-born conchocelis of *Pyropia yezoensis* which was suffering from the fatal disease known as yellow spot disease. The strain was then sent to the China General Microbiological Culture Collection Center under the collection number CGMCC 1.16311. The original organism was isolated using a previously described method (7). In our study, *V. mediterranei* 117-T6 was pathogenic to the conchocelis of several *Pyropia* species, including *P. yezoensis* (Y. He, Q. Liu, M. Xu, Z. Tao, and R. Yang, unpublished data). *Pyropia* species are popular edible red algae with great commercial importance and have a long history of cultivation in northeast Asia. In this paper, the complete genome sequence of *V. mediterranei* strain 117-T6 was determined to facilitate further research into its virulence factors.

*V. mediterranei* 117-T6 was cultured in tryptic soy broth (TSB) medium containing 1% NaCl at 28°C for 12 h with shaking at 120 rpm. The genomic DNA (gDNA) was extracted using the Ezup column bacteria gDNA purification kit (Sangon Biotech Co., Ltd., Shanghai, China) according to the manufacturer’s protocols. The gene library was constructed using a SMRTbell template prep kit 1.0 and was quality controlled using Qubit 3.0 (Life Technologies, CA, USA) and a 2100 bioanalyzer (Agilent, Santa Clara, CA, USA). After library preparation, the genome was sequenced by single-molecule real-time sequencing (SMRT) using the PacBio sequel system (Pacific Biosciences, Menlo Park, CA) (8). The generated subreads were assembled using Hierarchical Genome Assembly Process (HGAP) version 4 with default parameters (9). The genome structure was annotated using Glimmer version 3.02 (10). To obtain the protein function annotation, the predicted protein sequences were compared with those in the nonredundant protein (NR), Cluster of Orthologous Groups (COG), KEGG, Gene Ontology (GO), and Swiss-Prot databases in NCBI using BLASTP with an E value of 1e–5 (11).

A total of 178,807 filtered subreads with a total length of 1,332,991,254 bp were generated after sequencing (average subread length, 7,454 bp; subread N50, 9,729 bp; 233-fold coverage). The reads were assembled, and we found that the complete
genome sequence of *V. mediterranei* 117-T6 consists of two circular chromosomes (3,698,726 bp and 2,081,621 bp, respectively) and one plasmid (195,680 bp). The total length of *V. mediterranei* 117-T6 was 5,976,027 bp with a GC content of 44.04%, including 5,539 coding DNA sequences (CDS), 31 rRNAs, 115 tRNAs, and 264 noncoding RNAs (ncRNAs). The number of tandem repeat finder (TRF) repeats of chromosome1, chromosome2, and the plasmid were 48, 20, and 9, respectively, and the number of simple sequence repeats (SSRs) were 9, 3, and 2, respectively. The plasmid contained 2 clustered regularly interspaced short palindromic repeat (CRISPR) sequence cassettes.

**Data availability.** The complete genome sequence for *V. mediterranei* 117-T6 has been deposited in DDBJ/ENA/GenBank under the accession numbers CP033577 to CP033579 (BioProject number PRJNA498774). The raw data have been submitted to the Sequence Read Archive (SRA) under run number SRR8294759.

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**REFERENCES**

1. Takemura AF, Chien DM, Polz MF. 2014. Associations and dynamics of *Vibrionaceae* in the environment, from the genus to the population level. Front Microbiol 5:38. [https://doi.org/10.3389/fmicb.2014.00038](https://doi.org/10.3389/fmicb.2014.00038).
2. Pruzzo C, Huq A, Colwell RR, Donelli G. 2005. Pathogenic *Vibrio* species in the marine and estuarine environment, p 217–252. In Belkin S, Colwell RR. Oceans and health: pathogens in the marine environment. Springer, Boston, MA.
3. Pujalte MJ, Garay E. 1986. Proposal of *Vibrio mediterranei* sp. nov.: a new marine member of the genus *Vibrio*. Int J Syst Evol Microbiol 36:278–281. [https://doi.org/10.1099/00207713-36-2-278](https://doi.org/10.1099/00207713-36-2-278).
4. Chimetto LA, Brocchi M, Gondo M, Thompson CC, Gomez-Gil B, Thompson FL. 2009. Genomic diversity of vibrios associated with the Brazilian coral *Mussismilia hispida* and its sympatric zoanthids (*Polythoa caribaeorum*, *Polythoa variabilis* and *Zoanthus solandri*). J Appl Microbiol 106:1818–1826. [https://doi.org/10.1111/j.1365-2672.2009.04149.x](https://doi.org/10.1111/j.1365-2672.2009.04149.x).
5. De La Pena LD, Lavilla-Pitogo CR, Paner MG. 2001. Luminescent vibrios associated with mortality in pond-cultured shrimp *Penaeus monodon* in the Philippines: species composition. Fish Pathol 36:133–138. [https://doi.org/10.3147/fsp.36.133](https://doi.org/10.3147/fsp.36.133).
6. Montes M, Farto R, Pérez MJ, Nieto TP, Larsen JL, Christensen H. 2003. Characterization of *Vibrio* strains isolated from turbot (*Scophthalmus maximus*) culture by phenotypic analysis, ribotyping and 16S rRNA gene sequence comparison. J Appl Microbiol 95:693–703. [https://doi.org/10.1046/j.1365-2672.2003.02028.x](https://doi.org/10.1046/j.1365-2672.2003.02028.x).
7. Guan XY, Li JB, Zhang Z, Li FC, Yang R, Jiang P, Qin S. 2013. Characterizing the microbial culprit of white spot disease of the conchocelis stage of *Porphyra yezoensis* (Bangiales, Rhodophyta). J Appl Phycol 25:1341–1348. [https://doi.org/10.1007/s10811-013-9976-8](https://doi.org/10.1007/s10811-013-9976-8).
8. Rhoads A, Au KF. 2015. PacBio sequencing and its applications. Genomics Proteomics Bioinformatics 13:278–289. [https://doi.org/10.1016/j.gpb.2015.08.002](https://doi.org/10.1016/j.gpb.2015.08.002).
9. Chin CS, Alexander DH, Marks P, Klammer A, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korf J. 2013. Non-hybrid, finished microbial genome assemblies from long-read SMRT sequencing data. Nat Methods 10:563–569. [https://doi.org/10.1038/nmeth.2474](https://doi.org/10.1038/nmeth.2474).
10. Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. 1999. Improved microbial gene identification with Glimmer. Nucleic Acids Res 27:4636–4641. [https://doi.org/10.1093/nar/27.23.4636](https://doi.org/10.1093/nar/27.23.4636).
11. Jacob A, Lancaster J, Buhler J, Harris B, Chamberlain RD. 2008. Mercury BLASTP: accelerating protein sequence alignment. ACM Trans Reconfigurable Technol Syst 1:1–44. [https://doi.org/10.1145/1371579.1371581](https://doi.org/10.1145/1371579.1371581).