Effectiveness of esterified whey proteins fractions against Egyptian Lethal Avian Influenza A (H5N1)

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Abstract

Background: Avian influenza A (H5N1) virus is one of the most important public health concerns worldwide. The antiviral activity of native and esterified whey proteins fractions (α-lactalbumin, β-lactoglobulin, and lactoferrin) was evaluated against A/chicken/Egypt/086Q-NLQP/2008 HPAI (H5N1) strain of clade 2.2.1 (for multiplicity of infection (1 MOI) after 72 h of incubation at 37°C in the presence of 5% CO2) using MDCK cell lines.

Result: Both the native and esterified lactoferrin seem to be the most active antiviral protein among the tested samples, followed by β-lactoglobulin. α-Lactalbumin had less antiviral activity even after esterification.

Conclusion: Esterification of whey proteins fractions especially lactoferrin and β-lactoglobulin enhanced their antiviral activity against H5N1 in a concentration dependent manner.

Background

Avian influenza A (H5N1) virus is one of the most important public health concerns worldwide. It has been detected and identified in South East Asia since 2003. In 2005, the virus was already spread to many countries in Europe, Asia and Africa [1]. Later it interrupted international travel and negatively affected the world economy especially tourism.

In December 2005, the first case of H5N1 in Egypt was detected in a migrating bird in Damietta Governorate. In mid-February 2006, H5N1 infection was reported in Egypt among domestic poultry in more than 15 governorates, resulting in severe losses for the poultry industry. In March 2006, the first human case of H5N1 in Egypt was detected [2]. Egypt had the highest number of confirmed human avian influenza cases outside Asia. As of 4 March 2010, 104 human cases, including 30 fatalities, had been recorded in Egypt [3,4].

Whey proteins have been reported to have numerous therapeutic applications including effects on bone (stimulate proliferation and differentiation of osteoblastic cells as well as suppress bone resorption and increase femoral bone strength), muscles (enhancing muscle hypertrophy and strength due to its leucine content), blood (lower blood pressure and reduce the risk of hypertension), brain (increase brain serotonin levels), immune system (stimulate immunity and improving immune function), cancer (increase NK cell function and glutathione levels), wound healing (essential for post-surgical wound healing and protein depletion delays healing time), and aging (antiaging agent due to the whey content of glutathione/antioxidant component). Furthermore, they act as antioxidant, antihypertensive, antiviral, antimicrobial, chelating agent and prevent cardiovascular diseases and osteoporosis [5,6]. The biological value of whey proteins has also been reviewed for their antimicrobial and antiviral functions [7-11].

Modification of whey proteins to enhance or alter their biological and functional properties may increase its applications. Whey proteins modification can be accomplished by chemical, enzymatic, or physical techniques [12]. Additional negative charges on β-lactoglobulin (BLG), α-lactalbumin (ALA) and human serum albumin (HSA) endowed them a significant antiviral activities against human immunodeficiency viruses—HIV-1 and HIV-2 [13-17]. BLG modified with 3-hydroxyphthaloylacid (3HP) inhibited the infection with HIV-1, Herpes simplex virus types 1 and 2 and human cytomegalovirus [18,19]. In addition, many studies reported that increasing the net positive charge on whey proteins...
led to enhancement of its antiviral activity. Esterified whey proteins showed antiviral activity against poliovirus type-1, Coxsackie virus B6, human cytomegalovirus, Herpes simplex virus type 1 and human influenza virus A subtype H3N2 & subtype H1N1 [20-24].

Therefore, it was thought worthwhile to test the efficacy of native and modified whey proteins fractions against influenza virus subtype H5N1 which may be useful for the prophylaxis and treatment of influenza viruses and at the same time can be a potential and low cost alternative candidate for an anti-influenza agent.

**Materials and methods**

**Materials**

The HPAI H5N1 A/chicken/Egypt/086Q-NLQP/2008 (referred to as EGYvar/H5N1) virus (GenBank accession number: EU496398.1) was isolated from an H5 vaccinated commercial chicken farm in 2008 during the routine national surveillance conducted by the National Laboratory for Veterinary Quality Control on Poultry Production (NLQP), Giza, Egypt. The virus has been titrated using Hemagglutination test (HA) 256 HAU. The tissue culture lethal dose 100% (TCLD100%), tissue culture infective dose 50% (TCID50%) and Embryonated egg infected dose 50% (EID50%) were calculated using Read and Minch (10^6, 10^6.26, 10^6.64/ml respectively) in Specific Pathogen Free Embryonated Chicken Egg (SPF ECE) and Madin-Darby Canine Kidney Cells (MDCK) as described in WHO manual [25]. SPF ECE was obtained from Qom Oshime SPF farm Egypt, and MDCK was obtained from NAMRU3 unit, Egypt.

α-Lactalbumin (97.46% protein) and β-lactoglobulin (97.8% protein) were kindly obtained from Davisco Food International (USA) while lactoferrin (95% protein) was kindly obtained from Armor Proteins (France). All other chemicals used in this study were of analytical grade.

**Methods**

1- **Protein Esterification**

The procedure of [26] was used for esterification of whey proteins fractions.

2- **Esterification extent**

The extent of esterification of proteins was quantified by the color reaction with hydroxylamine hydrochloride as described by [27].

3- **The antiviral activity of native and esterified whey protein**

The antiviral activity of native and esterified whey proteins: α-lactalbumin (ALA), β-lactoglobulin (BLG) and lactoferrin (LF) was assayed against Egyptian highly pathogenic avian virus A/chicken/Egypt/086Q-NLQP/2008 HPAI (H5N1) strain of clade 2.2.1 at concentrations of 1.00 MOI (multiplicity of infection) per cell according to [22].

4- **Statistical analysis**

All experiments were performed in triplicates and the results were expressed by the mean plus the standard deviation.

**Results**

**Extent of esterification**

The whey proteins fractions ALA, BLG and LF were modified to the extent of 68%, 100% and 100% respectively which indicates less esterification susceptibility of ALA as compared to both BLG and LF. The observed extents of such esterification are in accordance with [26].

**Antiviral activity of whey proteins fractions**

Data shown in Figure 1 demonstrate the antiviral effect of native and esterified whey proteins fractions against H5N1 propagated in MDCK cells at 100% (1.00 MOI) level of viral infection. Native proteins have exhibited different levels of inhibitory effects against the virus. It ranged from 21.62 ± 2.1 to 26.40 ± 1.5% for ALA, from 32.87 ± 2.3 to 42.43 ± 1.3% for BLG and from 34.98 ± 5.5 to 70.92 ± 3.2% for LF in response to protein concentration increasing from 20 to 80 µg/ml. The difference in viral inhibitory effect of the three native proteins may be due to the difference in their structural nature. Native lactoferrin seems to be the most active antiviral protein among the tested samples, probably due to its more basic nature that enables its interference and interaction with the viral constituents affecting viral replication and activity. Esterification of whey proteins fractions has further enhanced their antiviral activity against H5N1 in a concentration dependent manner. Met-ALA exhibited antiviral effect ranging from 54.84 ± 0.1 to 79.57 ± 2.0%, Met-BLG from 64.88 ± 1.9 to 99.05 ± 0.4% and Met-LF from 69.28 ± 1.8 to 99.42 ± 0.6% in response to protein concentration going from 20 to 80 µg/ml. Met-ALA was the lowest active as antiviral protein even after esterification while both Met-BLG and Met-LF reached maximum antiviral influence when the protein concentration was increased to 80 µg/ml. This may confirm that esterification is a potent tool which introduces this antiviral activity into native proteins. It may also show that the original differences between BLG and lactoferrin disappeared completely after esterification.

The antiviral activity of the tested proteins may be due to its interaction with influenza nuclear proteins (PB1, PB2, PA and NP), which catalyze the transcription of viral RNA [28-30]. Since these proteins are normally associated with RNA and undergo systematic dissociation during replication, the tested positively charged proteins may interfere with this association-dissociation process or compete for the negative charges on the
exposed regions of RNA, disturbing the overall replication pathways.

Conclusion
From the presented data, it can be concluded that esterification of LF followed by BLG and lastly by ALA enhances its antiviral activities against H5N1 infected into MDCK cell lines which is dependent on the concentration of the tested proteins. Consequently, applying this technique is associated with a protective action on the cell lines subjected to the viral infection. Further studies are needed to improve the antiviral activity of both of α-lactalbumin and to a less extent β-lactoglobulin.

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Authors’ contributions
SHT carried out the study design, participated in data organization, wrote and revised the manuscript; MAM helped in performance of the experiments; MZS carried out the study design and revised the manuscript; AIA participated in the design of the study and revised the manuscript; MMA carried out most of the experiments, participated in data organization, wrote and revised the manuscript; WHE helped in performance of the experiments.

All authors read and approved the final manuscript.

Competing interests
The authors declare that they have no competing interests.

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References
1. Yingst SL, Saad MD: Qinghai-like A (H5N1) from domestic cats, northern Iraq. Emerging Infectious Diseases 2006, 12:1295-1297.
2. Saad MD, Ahmed LS, Gamal-Eldeen MA, Fouda MK, Khalil FM, Parker AM, Monteville M: Possible avian influenza (H5N1) from migratory birds, Egypt. Emerging Infectious Diseases 2007, 13:1120-1121.
3. Earhart KC, Elsayed NM, Saad MD, Gaberena LV, Naylor A, Deyde VM: Oseltamivir resistance mutation N294 S in Human influenza A (H5N1) virus in Egypt. Journal of Infect Public Health 2009, 2:74-80.
4. Fasina FO, Ifende VI, Ajibade AA: Avian influenza A (H5N1) in humans: lessons from Egypt. Eurosurveillance 2010, 15:19473.
5. Marshall K: Therapeutic applications of whey protein. Alternative Medicine Review 2004, 9:136-156.
6. Frostedt LJ, John LZ, Michael AK, Loren SW, Eric DB: A whey-protein supplement increases fat loss and spares lean muscle in obese subjects: a randomized human clinical study. Nutrition & Metabolism 2008, 5:1-8.
7. Van der Strate BW, Beljaars L, Molema G, Meijer DK: Antiviral activities of lactoferrin (A Review). Antiviral Research 2001, 152:225-239.
8. Flors R, Recio I, Benkhout B, Visscher S: Antibacterial and antiviral effects of milk proteins and derivatives thereof. Current Pharmaceutical Design 2003, 9:1257-1275.
9. Kvistgaard AS, Pallesen LT, Arias CF, Lopez S, Petersen TE, Heegaard CW, Rasmussen JT: Inhibitory Effects of Human and Bovine Milk Constituents on Rotavirus Infections. Journal of Dairy Science 2004, 87:4088-4096.
10. Pan Y, Lee A, Wan J, Coventry MJ, Michalak WP, Sheilic B, Roginski H: Antiviral properties of milk proteins and peptides (A Review). International Dairy Journal 2006, 16:1252-1261.
11. Bojesen A, Buesa J, Montava R, Kvistgaard AS, Kongsbak MB, Petersen TE, Heegaard CW, Rasmussen JT: Inhibitory activities of bovine macromolecular whey proteins on rotavirus infections in vitro and in vivo. Journal of Dairy Science 2007, 90:66-74.
12. Kananen A, Savolainen J, Mak kinen J, Perttila KU, Myllykoski L, Pihlanto-Leppa KA: Influence of chemical modification of whey protein conformation on hydrolysis with pepsin and trypsin. International Dairy Journal 2000, 10:691-697.
13. Jansen RW, Moller G, Pauwels R, Schols D, De Clerq E, Meijer DKF: Potent in vitro anti-human immunodeficiency virus-1 activity of modified human serum albumins. Molecular Pharmacology 1991, 39:818-823.

Figure 1 Antiviral activity of native and esterified whey proteins fractions against H5N1 virus infecting MDCK cells. A: α-lactalbumin, B: β-lactoglobulin and C: lactoferrin.
14. Takami M, Sone T, Mizumoto K, Kino K, Tsunoo H: Maleylated human serum albumin inhibits HIV-1 infection in vitro. Biochimica et Biophysica Acta 1992, 1180:180-186.

15. Swart PJ, Meijer DKF: Negatively charged albumins: A novel class of polyanionic proteins with a potent anti-HIV activity. Antiviral News 1994, 2:69-71.

16. Jiang S, Lin K, Strick N, Li YY, Neurath AR: Chemically modified bovine β-lactoglobulin blocks uptake of HIV-1 by colon- and cervix-derived epithelial cell lines. Journal of Acquired Immune Deficiency Syndromes 1996, 13:461-462.

17. Schoen P, Conver J, Meijer DKF, Wilschut J, Swart PJ: Inhibition of influenza virus fusion by polyanionic proteins. Biochemical Pharmacology 1997, 53:995-1003.

18. Neurath AR, Debnath AK, Strick N, Li YY, Lin K: 3-Hydroxyphthaloyl-β-lactoglobulin. 1. Optimization of production and comparison with other compounds considered for chemoprophylaxis of mucosally transmitted human immunodeficiency virus type 1. Antiviral Chemistry & Chemotherapy 1997, 8:131-139.

19. Neurath AR, Strick N, Li YY: 3-Hydroxyphthaloyl-β-lactoglobulin. III. Antiviral activity against herpes viruses. Antiviral Chemistry & Chemotherapy 1998, 9:177-184.

20. Chobert JM, Sitohy M, Billaudel S, Dalgalarrondo M, Haertlé T: Anticytomegaloviral Activity of Esterified Milk Proteins and L-Polylysines. Journal of Molecular Microbiology and Biotechnology 2007, 13:255-258.

21. Sitohy M, Billaudel S, Haertlé T, Chobert JM: Antiviral activity of esterified alpha-lactalbumin and beta-lactoglobulin against herpes simplex virus type 1. Comparison with the effect of acyclovir and L-polylysines. Journal of Agricultural Food Chemistry 2007, 55:10214-10220.

22. Sitohy M, Michele D, Marie N, Besse B, Billaudel S, Haertle T, Chobert JM: The effect of bovine whey proteins on the ability of poliovirus and Coxsackie virus to infect Vero cell cultures. International Dairy Journal 2008, 18:658-668.

23. Sitohy M, Besse B, Billaudel S, Haertle T, Chobert JM: Antiviral Action of Methylated β-Lactoglobulin on the Human Influenza Virus A Subtype H3N2, Probiotics and Antimicrobial Proteins 2010, 2:104-111.

24. Sitohy M, Scanu M, Besse B, Mollat C, Billaudel S, Haertle T, Chobert JM: Influenza virus A subtype N1H1 is inhibited by methylated β-lactoglobulin. J Dairy Research 2010, 77:411-418.

25. WHO: Manual on animal influenza diagnosis and surveillance 2002, WHO/CDs/CSR/NCS.5.

26. Sitohy M, Chobert JM, Haertlé T: Simplified short-time method for the esterification of milk proteins. Milchwissenschaft 2001, 56:127-131.

27. Bertrand-Harb C, Chobert JM, Dufour E, Haertlé T: Esterification of food proteins: Characterization of the derivatives by a colorimetric method and by electrophoresis. Sciences des Aliments 1991, 11:641-652.

28. Huang TS, Palese P, Krystal M: Determination of influenza virus proteins required for genomic replication. Journal of Virology 1990, 64:5669-5673.

29. Poole E, Elton D, Medcalf L, Digard P: Functional domains of influenza A virus PB2 protein: identification of NP and PB1-binding sites. Virology 2004, 321:120-133.

30. Boulou S, Akansu H, Ruigrok RW, Baudin F: Nuclear traffic of influenza virus proteins and ribonucleoprotein complexes. Virus Research 2007, 124:12-21, 7.

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