Research Paper

Superoxide Anion Radical Scavenging Activities of Herbs and Pastures in Northern Japan Determined Using Electron Spin Resonance Spectrometry

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Received: 2007.07.04; Accepted: 2007.07.26; Published: 2007.07.30

Free radicals are not only destructive to the living cells but also reduce the quality of animal products through oxidation. As a result the superoxide anion radical (O$_2^-\cdot$), one of the most destructive reactive oxygen species, is a matter of concern for the animal scientists as well as feed manufacturers to ensure the quality of product to reach consumers demand. The superoxide anion radical scavenging activities (SOSA) of water and MeOH extracts of 2 herbs and 9 pasture samples collected from lowland and highland swards were determined against a 5,5-dimethyl-1-pyroline-N-oxide-O$_2^-\cdot$ spin adduct based on a hypoxanthine-xanthine oxidase reaction using electron spin resonance spectrometry. Both the water and MeOH extracted SOSA differed among the herbs and pastures. Species and altitudinal variations were observed between extraction methods. The herbs were higher in both water and MeOH extracted SOSA than the pastures except for water extracts of one pasture, white clover (Trifolium repens L.). Among the pastures, quackgrass (Agropyron repens L.) showed higher SOSA in both the MeOH and water extracts, and timothy (Phleum pretense L.) showed higher MeOH extracted SOSA. It is apparent that the kind and amount of antioxidants differ among herbs and pastures. Animal health and quality of animal products could be improved by adequate selection and combining of herbs and pastures having higher SOSA.

Key words: Herbs and pastures, superoxide anion radical, scavenging activity, ESR, HPLC, altitudinal locations

1. Introduction

A part of the oxygen taken into living cells is changed to several harmful reactive oxygen species and free radicals. Once formed, free radicals can start a chain reaction leading to formation of more free radicals. Superoxide anion radical (O$_2^-\cdot$) is one of the strongest reactive oxygen species among the free radicals that are generated first after oxygen is taken into living cells. O$_2^-\cdot$ changes to other harmful reactive oxygen species and free radicals such as hydrogen peroxide and hydroxyl radical. Although detailed relationships between antioxidants and animal health have not yet been reported, antioxidants supplied through diets improve the antioxidative activity of animals, hence leading to improved animal health. Natural antioxidants supplied through forage grasses and legumes can also prolong the shelf life of meat and milk and protect meat from losing its desirable red color [1,2]. The freshness, fat content, flavor and color of meat are key factors used to judge the quality of meat by consumers, and diet has a potent influence on these factors [3-5]. The rate of loss of the red color is related to the degree of unsaturation of lipids and the presence of antioxidants supplied through the diet [6]. Natural antioxidants from plants also protect unsaturated fatty acids such as n-3 fatty acids from oxidation [7]. Conjugated linoleic acid (CLA) was reportedly much higher for pasture-fed cattle than for concentrate-fed cattle [8], and pasture contains a high amount of the precursor of CLA, i.e., polyunsaturated C18:2, linoleic acid [9] which could be protected from oxidation by antioxidants from feed. It is generally considered that fresh pastures are the principal supplier of antioxidants to animals [2]. Ensuring the supply of antioxidants through pastures is essential to produce good quality products from animals. As a result, natural herbs and pastures having high antioxidative activities have attracted considerable interest from scientists as well as feed manufacturers [10]. However, antioxidant properties vary from species to species and within species due to
environmental effects. For example, Collomb et al. [11] have reported that the composition related to antioxidative activities of milk and cheese differ depending on different altitudinal locations. The content of acteoside, an antioxidant, in forage herb plantain varies due to air temperature, light intensity, fertilizer application and harvesting season [12,13]. Plantain has a higher amount of superoxide anion radical scavenging activity (SOSA) than the principal temperate pastures used in northern Japan [14]. Despite the importance of natural herbs and pastures as a source of antioxidants, little attention has been paid to the determination of antioxidative properties of herbs and pastures. Few experiments have been carried out; including those of Lee et al. [15] on the antioxidative activities of alfalfa and timothy varieties using the DPPH method, and by Tamura and Masumizu [16] and Al-Mamun et al. [14] on SOSA measured using an ESR-spin trapping method for s limited number of pasture and herb species. Therefore, the present study was undertaken to find out the variation in the SOSA among a wide range of herbs and pastures, including grasses and legumes, collected from different altitudes in northern Japan.

2. Materials and Methods

Plant materials and preparation

Samples of herbs and pastures were collected at 3 cm above ground level from two swards differing in altitude, namely the Yamagami sward (lowlands, N 36°5′ E141°00′ and 177 m above sea level) and the Hayasaka sward (highlands, N 39°8′1″ E141°47″ and 920 m above sea level), which are usually used for harvesting hay and grazing by Japanese shorthorn cattle during the summer. From the lowlands, we collected one herb, plantain (PL, Plantago lanceolata L.), and nine dominant pasture species; perennial ryegrass (PR, Lolium perenne L.), orchardgrass (OR, Dactylis glomerata L.), timothy (TI, Phleum pretense L.), white clover (WC, Trifolium repens L.), quackgrass (QG, Agropyron repens L.), reed canarygrass (RC, Phalaris arundinacea L.), Kentucky bluegrass (KB, Poa pratensis L.), Japanese lawngrass (JL, Zoysia japonica L.) and alfalfa (AL, Medicago sativa L.) in the vegetative stage on 26 May, 2006. From the highlands we also collected one herb, dandelion (DD, Taraxacum officinale Weber) and five dominant pasture species; PR, OR, TI, WC and QG, at the vegetative stage on 26 May, 2006. The samples from the highlands were collected 3 weeks later than those of the lowlands in order to compare the same growth stage because the daily mean air temperature was higher at the latter (Fig. 1).

The samples from the two swards were then freeze-dried in a FRD-50M freeze-drier (Iwaki Glass Co., Ltd., Japan) and ground to a fine powder using a vibrating sample mill (Heko Co. Ltd., Japan). Each of the 1 mg of ground samples was extracted either into 10 mL of water (pH adjusted to 7.4 with phosphate buffer) or into 10 mL of pure MeOH, with linear shaking for 1 hour at a speed of 130 revolutions hour⁻¹ at room temperature. Finally, the samples were centrifuged at 3500 rpm for 10 min and the eluents used for the determination of SOSA by an ESR-spin trapping method. The experiment was conducted with three replicates.

Figure 1. The daily mean air temperature (°C) during sampling of the experimental areas. a, b; date of sampling at lowlands and highlands, respectively.

Chemicals used

Superoxide dismutase (SOD) from bovine erythrocytes (Nacalai Tesque Inc., Kyoto, Japan); trapping agent 5,5-dimethyl-1-pyroline-N-oxide (DMPO) (Labotec Co. Ltd., Tokyo, Japan); hypoxanthine (HPX) (Sigma Chemical Co., Tokyo, Japan); xanthine oxidase (XOD) for active oxygen generation (Roche Chemical Co., Tokyo, Japan); phosphate buffered solution pH 7.4 (1/15 mol L⁻¹) (Kanto Chemical, Tokyo, Japan); methanol (Wako Chemical, Osaka, Japan); catechin (Extrasyphes Inc. Genay, France); and Folin-Denis reagent (Sigma Chemical Co. MO, USA) were used.

Determination of SOSA

The SOSA was determined through a spin trapping method using an ESR spectrometer (JES-FA200, JEOL Ltd., Tokyo, Japan). Manganese oxide was used as an internal standard, which provided a constant signal intensity to which all peaks were compared. ESR spectrometer measurement conditions used to estimate O₂⁻ were as follows: magnetic field: 335±5 mT; power: 4 mW, 9.42 GHz; sweep time: 2 min; modulation frequency: 100 kHz, 0.08 mT; amplitude: 2.5 × 100; time constant: 0.1 sec.

An HPX-XOD reaction system was used to determine SOSA. The chemicals were mixed in the following order: 15 μL of 9.2 mol L⁻¹ DMPO, 50 μL of 2 mmol L⁻¹ HPX, 35 μL of water (pH 7.4, adjusted using PB), 50 μL of samples, and finally, 50 μL of 0.4 U mL⁻¹ XOD from cow’s milk. The mixture was transferred to an ESR spectrometry flat cell and the DMPO-O₂⁻ spin adduct was quantified for 1 min after the addition of XOD. Standard curves were made for both water and MeOH extracted samples and the SOSA was expressed as SOD equivalent units per milligram dry matter (DM) [17].

Determination of crude phenolics

Total polyphenol measurement of freeze-dried herb and pastures from the highlands was performed according to the Folin-Denis colorimetric method [18].
Each of the 0.03-0.04 g of freeze-dried samples was first mixed with 5 volumes of water or MeOH, and kept for 2 hours at room temperature before centrifugation at 10,000 rpm for 10 min. Then, 0.1 mL of the supernatant was mixed with 0.1 mL of Folin-Ciocalteau’s reagent. After 10 min, 0.1 mL of 10% saturated sodium bicarbonate was added and mixed well. After 1 hour the absorbance of the mixed samples at 655 nm was measured by a spectrophotometer. Catechin was used as the standard to which the samples were compared.

**Determination of vitamin C**

Freeze-dried samples from the highlands were used to determine vitamin C content using high performance liquid chromatography (HPLC) (LC 10AS, Shimadzu Co., Ltd., Kyoto, Japan). Quantitative analyses were performed at 35 °C using a 4.6 mm x 10 cm column (Senshupak silica-1100-N). The mobile phase was an acetyl ether, n-hexane, acetic acid and water (60:40:5:0.05) solution. The flow rate was 1.5 mL min⁻¹. Photodiode array detection was performed at 495 nm. First, 0.5 g of freeze-dried sample was extracted into 50 mL of 5% metalinic acid, homogenized and then centrifuged at 3000 rpm for 10 min. Then 1 mL of the eluent was mixed with 1 mL of 5% metallic acid, 100 μL of 0.2% 2,6-dichlorophenol-indophenol sodium salt dehydrate and 2 mL of 2% thiourea-5% metallic acid solution were added into the test tube. After that 0.5 mL of 2% of 2,4-dinitrophenol hydrazine-4.5 mol L⁻¹ H₂SO₄ was added and kept at 38-42 °C for 16 hours. Finally 3 mL of acetyl ether was added and the mixture was shaken for 1 hour, and then used for determination of ascorbic acid by HPLC.

**Statistical analysis**

All data were expressed as mean values with standard error of the mean (SEM). Statistical analysis was performed using the General Linear Model procedure of SAS with Tukey’s Studentized range (HSD) test of the SAS system [19]. In the case of individual species we used Student’s t-test to compare the water and MeOH extracted SOSA and polyphenol content and the SOSA values of highlands and lowlands.

3. **Results and Discussion**

**Differences in SOSA among herbs and pastures of lowlands**

The SOSA of herb and pastures varied largely among species both in the water and MeOH extracts (Fig. 2). For the water extracted SOSA, WC showed the highest and PL was the second highest of the pastures while PR, TI, KB, JL and AL grouped significantly to the lowest. OR, RC and QG were in between PL and the lowest group.

On the other hand, for MeOH extracted SOSA, PL showed an extremely high SOSA that was significantly higher than other grasses and PR, RC, KB and JL grouped significantly to the lowest SOSA while OR was in between. In the case of the leguminous species, WC showed very low SOSA, and no detectable SOSA was found in AL.

The most interesting observation was species variations between extraction methods. For instance, SOSA of WC was the highest in water extracts but the lowest in MeOH extracts. On the other hand, in PL, the MeOH extracted SOSA was extremely high, but that of water was relatively low, about half of the SOSA of WC. TI was the lowest for water extracted SOSA, but was highest for the MeOH extracted SOSA among the pastures. In PR, KB and JL, both the water and MeOH extracted SOSA were very low.

The present results indicate that the amount and kind of antioxidants accumulating differ largely between herbs and pastures and among pastures. Therefore, selection of pastures having higher antioxidative activity would be very important to improve animal health and the antioxidative status of meat and milk, and to prolong their shelf life. The effect of the water and MeOH extracted SOSA on animal health and on meat and milk quality is not clarified yet, but it could be considered that MeOH extracted SOSA is more important than water extracted SOSA, since MeOH extracted SOSA represents lipid soluble antioxidants. Bioactive compounds in each species having antioxidative activity are still not known in detail except for acteoside, a phenylethanoid compound that has high antioxidative activity [20]. The higher MeOH extracted SOSA of PL might be due to the presence of acteoside,
and the higher water extracted SOSA of WC might be due to higher vitamin C content than in the other pastures as reported by Tamura and Masumizu [16], and higher water soluble total polyphenol content. It is necessary to clarify the type of antioxidants and their mode of action in each species.

**Differences of SOSA among herbs and pastures of highlands**

The SOSA of herb and pastures also varied largely among species both for water and MeOH extracts (Fig. 3). For water extracted SOSA, WC showed the highest SOSA, and QG and DD also had significantly higher SOSA than PR, OR and TI. On the other hand, for the MeOH extracted SOSA, DD was significantly the highest while that of WC was the lowest. The PR, OR, TI and QG were in between DD and WC and no significant difference was found among them. The most interesting observations here were almost the same as found for the lowlands. For instance, WC showed the highest water extracted SOSA while that of MeOH extracts was the lowest, and among the pastures, TI had the highest MeOH extracted SOSA while that of water extracts was very low. Furthermore, we found that DD was very high both for the water and MeOH extracted SOSA.

**Comparison of SOSA between lowlands and highlands**

In the present experiment, only five species, PR, OR, TI, WC and QG, were compared, as they were available in both lowlands and highlands. The water extracted SOSA of WC was higher in lowlands than in highlands, whereas other pastures remained comparable between locations of different altitude (Fig. 4). The reason why the water extracted SOSA of WC in lowlands was so much higher than that of highlands is unknown. But it could be considered that the residual effects of a cold winter in the highlands negatively affected the spring growth of WC, which is less winter hardy than the graminaceae species.
attributable to higher amounts of flavonoids and phenolic compounds due to higher UV radiation.

**Polyphenol and vitamin C contents and their relationship to SOSA**

All of the water and MeOH extracted polyphenol and vitamin C contents differed significantly among herbs and pastures (Fig. 5, 6). For example, the water extracted polyphenol and vitamin C contents were significantly highest in WC and lowest in DD. Among other species, vitamin C content was significantly higher in PR, TI and QG than in OR, but no significant difference was observed in water extracted polyphenol. MeOH extracted polyphenol was significantly highest in TI and higher in QG and DD than other pastures, while OR was the lowest. The highest water extracted SOSA being found in WC might be due to its having the highest contents of both polyphenol and vitamin C [16]. On the other hand, the highest MeOH extracted SOSA in TI might be due to its having the highest MeOH extracted polyphenol. The same prediction is also applicable for DD and QG. Though the SOSA obtained in the present study may be related to the concentrations of polyphenol and vitamin C, the relationships are not that clear and the contribution coefficients of the polyphenol and vitamin C to SOSA were relatively low (Fig. 7). Other antioxidants presumably also affect the level of SOSA. DD was considered the most typical example, because although its water extracted polyphenol and vitamin C contents were the lowest, the water extracted SOSA was relatively high, indicating the presence of unknown water soluble antioxidants. Therefore, the determination of only polyphenol and and vitamin C is not enough to represent the antioxidative properties of an herb or pasture. The determination of total antioxidative properties by ESR-spin trapping or other established methods is essential.

![Graph 1](image1.png)

**Figure 5.** Water and MeOH extracted polyphenols of herbs and pastures of highlands. Values with different superscripts differed significantly ($P<0.05$).

![Graph 2](image2.png)

**Figure 6.** Vitamin C content of herbs and pastures of highlands. Values with different superscripts differed significantly ($P<0.05$).
4. Conclusions

From the present experiments it could be concluded that the antioxidant properties represented by both water and MeOH-extractable SOSA vary largely among species of the herbs and pastures available in northern Japan, and also vary depending on the altitudinal location. As a pioneer experiment, the results are promising and some of the species, especially PL, WC, DD, QG and TI, could be used as natural sources of antioxidants against superoxide radicals to protect animals from adverse effects of \( O_2 \cdot \) \(-\). However, based on the present experiments, further research should be performed to clarify specific antioxidants and their mode of actions in order to proper selection and combination of the herbs and pastures to improve the animal health ensuring the quality of animal products.

Acknowledgements

The authors thank to Professor emeritus Sansei Nishibe, Health Science University of Hokkaido for helpful suggestions, and Dr. Kanno and Mr. Deguchi, National Agricultural Research Center for Tohoku Region for providing air temperature data.

Conflict of interest

The authors have declared that no conflict of interest exists.

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