Phytochemical screening and anti-oxidant activity of *Sargassum wightii* enhances the anti-bacterial activity against *Pseudomonas aeruginosa*

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**A B S T R A C T**

In this study, the phytochemical, phenolic, flavonoid and bioactive compounds were successfully screened from crude extract of *Sargassum wightii* by LC-MS analysis after NIST interpretation. Bacterial growth inhibition study result was shown with 24 mm zone inhibition at 200 μg/mL concentration against *P. aeruginosa*. The increased phenolic content was much closed to gallic acid and the range was observed at 250 μg/mL concentration. In addition, flavonoid contents of the algae extract was indicated more significant with rutin at 200 μg/mL. In result, both the phenolic and flavonoid contents of the extract were more correlated with gallic acid and rutin. Further, the total anti-oxidant and DPPH radical scavenging activities were shown increased activity at 200 μg/mL concentrations. Furthermore, the excellent anti-bacterial alteration result was observed at 200 μg/mL concentration by minimum inhibition concentration. Therefore, the result was revealed that the marine algae *Sargassum wightii* has excellent phytochemical and anti-oxidant activities, and it has improved anti-bacterial activity against *P. aeruginosa*.

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1. Introduction

The unpredictable marine environment is complex process and researcher’s great concern for marine biodiversity and identification of pharmaceutical products (Cai et al., 2020). It is most advantages environment due to the extraordinary nutrients and organic materials. It is produced excellent biodiversity, significant physical biodiversity and increased seafood activations to human (El-Din and El-Ahwany, 2016). It mainly contained several factors including pH, temperature, salinity, additional nutrients, high organic content, velocity and more carbon and nitrogen sources. The unpredictable environments increase the booster dose in pharmaceutical drugs and also marine organisms belonging to various levels of the ecological hierarchy from algae, bivalves, crustaceans and fishes (Mehdinezhad et al., 2016). Among the various sources, seaweeds are important algae providing enormous biological properties due to the rich polysaccharides, phytochemical derivatives and bioactive compounds (Angela et al., 2020). Sometimes, the alkaloid and flavonoid content are also available very rich in marine algae. All these rich content are exhibited with more anti-oxidant activity. All these properties are detected based on the algae nature (Cyril et al., 2017).
Among the algal family, *Sargassum wightii* is a brown macroalgae comes under the order of family of Sargassaceae and order of Fucales with more polysaccharides contents (Raman et al., 2014). It has variety of species, presenting throughout the world in the nearest of temporal and tropical region. The habitat of *Sargassum wightii* is living in shallow water and coral reefs. *S. wightii* is presenting in southeast Coast of Tamil Nadu, India and various other parts of Asia, it is highly reported for animal food, food ingredients and fertilizer usage (Dileepkumar et al., 2018). It has more flavonoid content leads to greater anti-oxidant activity suggesting ideal target for investigating presence of bio-molecules for various medical and industrial applications. In addition, the anti-oxidative properties of the O-heterocyclic analogues from *S. wightii* have been reported previously by Sastry and Rao (1995). It has the effective enzyme-ligand interactions to synthesis novel kind of pharmcophores and nutraceuticals to combat multi drug resistant bacteraial infections (Marudhupandi and Thangappan, 2013). Last 100 years, seaweed is a potential use in fertilizer, fodder and medicinal sources. In addition, these raw materials are used in industry by agar, align and carrageenan and also consumed largely as food in Asian countries. *S. wightii* is a large, most economically important and also ecologically dominant than other algae (Ramani et al., 2014).

Previously, the more polysaccharides compounds are frequently reported from *Sargassum wightii* with increased biological and chemical properties (v et al., 2016). Marine algae are three groups based on the pigment production, named as green, red and brown algae. These three algal groups are comes under the family of Chlorophyceae, Rhodophyceae and Phaeophyceae respectively (El-Din and El-Ahwany, 2016)). All these algae are structurally diverse characters including rich phytochemical, pharmaceutical, and biomedical properties. Previously, researchers are reported that the algae and its derivatives have excellent biological activities including anti-oxidant, anti-bacterial, anti-fungal and anti-cancer activities (Dileepkumar et al., 2018). Importantly, the notable compounds form seaweeds are very effective against some of the biological activities including Sesquiterpenes, acyclic diterpenes, Atlantic, two diterpenoids with a novel skeleton, dictyterpenoids A and B, tetraterpenes, Oleanane-type triterpene (Raman et al., 2014; Angela et al., 2020; Cyril et al., 2017; Putri et al., 2019). Based on this above facts, we have focused on phytochemical properties of *Sargassum wightii* extract and its anti-oxidant properties against *P. aeruginosa*.

2. Materials and methods

2.1. Collection and processing of seaweed

The fresh, undamaged brown algae *Sargassum wightii* was collected on the Sea shores of Mandapam Region, Ramanathapuram District, Southeast coast of Tamil Nadu, India. The collected samples were washed by tap water and stored in ice box for transferred to lab. The collected seaweeds were confirmed as *Sargassum wightii* after authenticated with Dr.N.Manoharan, Department of Marine Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India. The collected seaweed were soaked on double distilled water containing beaker and washed three times for removal of surface contaminations. Following alcohol treatment was done for removal of free floating microorganisms on the seaweed surface. Finally, the washed sample was kept in dry for 15 days in room temperature at dark condition. After complete dry, the seaweed samples were ground to get powder form. After receive the thin powder using sieving, the sample was tightly packed and covered by black cloth for protected from sunlight and stored in 4 °C for further use.

2.2. Crude extract preparation

The powdered sample of *Sargassum wightii* was used to prepare the crude extract for further identifications using ethanol as a solvent used for get a better yield (Sastry and Rao (1995)). The 20 g of powder sample was mixed in 1 L ethanol solution for extraction of crude extract using Soxhlet apparatus at the required temperature until the color of the solvent become clear or 12 h. The crude extract of the sample was received after complete solvent removal using the rotary vacuum evaporator. Then, the extracted sample was filled in 50 g volume of container. Finally, the yield of 25 g powder material was calculated by the bellowed formula,

\[
\text{Yield} = \frac{\text{Amount of product}}{\text{Amount of sample}} \times 100
\]

2.3. Phytochemical screening of crude extract

The complete profile of the crude extract including essential oils, phenol, alkaloids, flavonoids, tannins, terpenoids, alcohols, acids, esters, steroids, amino and organic compounds were qualitatively and quantitatively analyzed by LC-MS based on the previous report of Bibi and Mohamad Fawzi (2020). In LC-MS, 30 m × 200 μm × 0.30 μm of capillary column was fixed automatically, it made up of CP-Silicon 5 of chrome pack. In this process, 600C for 5 min was set as initially and extended up to 150 °C for 15 min. The injector coupled detector and constant temperature was maintained at up to 200–300 °C. Additionally, 1 mL/min flow rate with carrier gas of 30 cm/s with linier velocity were arranged. The flowed sample was used for detect the available chemical constituents of the tested seaweed of *Sargassum wightii* based on the peaks and their respective retention time and occupied percentages. The obtained peaks and their respective chemical derivatives were interpreted by NIST Wiley library of Bharathidasan University, Tiruchirappalli, Tamilnadu, India.

2.4. Antimicrobial activity of *Sargassum wightii* crude extract

The antimicrobial activity performance to identification of *Sargassum wightii* crude extract against bacteria was done by agar well diffusion experiment. This method was followed by earlier accepted procedure of Lin et al. (2019). Yesterday inoculated well grown *P. aeruginosa* culture was streaked on muller hinton agar plate with the help of sterile buds. Then 6 mm of 3 distance wells for first well of minimum concentration, second well of maximum of crude extract, third well of distilled water. Then, add 100 μL of crude extract properly in respective wells and kept in inside of the incubator one day. After, the zones of the wells were measured using measuring scale and compared with distilled water well for detect the anti-bacterial effect of *Sargassum wightii* crude extract.

2.5. Measurement of phenol and flavonoid properties

Based on the previous evidences, the phenol and flavonoid content of the purified crude compound of *Sargassum wightii* was calculated by folin-ciocalteu's method, and the result was taken in UV–vis spectroscopy (Rajivgandhi et al., 2020a, 2020b). The process of phenolic and flavonoid properties of the purified crude compound was done in a respective 50 mL test tubes including 1 mL of purified extract and 5 mL 10 fold dilution of folin–Ciocalteu’s reagent was reacted in one tube, and Na2 CO3 was mixed in the same tube after reaction. Next, all the sample in the same tube was shaken continuously with 10 min and held at half hour incubation. The reaction mixture content of the tube was measured at 540 nm by using UV-spectrometer. Additionally, the gallic acid with 50–250 μg/mL separately was used for negative control and same concentrations without extract of gallic acid was used for
positive control. The result of 1 μg gallic acid was converted to 1 mg equal content of tested sample after comparison of triplicate result of test and control O.D values. Further, the colorimetric analysis result was confirmed the availability of flavonoid content after following the reported procedure of Dussossoy et al. (2011). As same as, in same tube 200 μL of purified extract plus 75 μL of sodium nitrate was taken clearly. Aliquot: 1:1 concentration of 1 mg/mL mixed AlCl₃ solution plus 1% of potassium iodide solution was added gradually into the wall of the test tube. Then, the mixture solution was shaking vigorously and kept in room temperature 30 min. After incubation, 540 nm of colorimeter was taken for control rutin for comparison and test sample with blank. The test and control samples were used in triplicate and resulted flavonoid content was calculated by 1 μg gallic acid that equal to 1 mg of tested sample.

2.6. Measurement of Anti-oxidant activity of purified extract

Phosphor-molybdenum experiment based procedure was used in this study to detect the total anti-oxidant activity in crude extract of *Sargassum wightii* (Maneesh and Chakraborty (2018)). Each 5 mL solution of sodium phosphate and ammonium molybdate was taken in a separate test tube. Additionally, 5 mL of H₂SO₄ was also taken in a test tube. Each 2 mL of all the three solution taken together in a same test tube and maintained at water bath at 97 °C for 30 min with shaking facility. After cool, 1 mL of well mixed solution was taken in a test tube, followed by 10 μL of *Sargassum wightii* crude extract was added into the same tube at 37 °C for 10 min. Finally, the anti-oxidant production of prepared solution was taken 600 nm O.D using UV-spectrophotometer. The result was compared with control ascorbic acid O.D, and noted the interpreted anti-oxidant result. In the result, 1 μg of ascorbic acid is equal to tested *Sargassum wightii* crude extract.

2.7. DPPH free radical scavenging assay

The availability of radical scavenging activity in the crude extract of *Sargassum wightii* was calculated by DPPH assay (Putri et al., 2019). 150 μL DPPH and different concentration of crude extract of *Sargassum wightii* was taken together in the sterile test tube. Instead, the sterile test tube was filled with 5 μL of butylated hydroxyl toluene with DPPH was acted as an experiment control. Additionally, ethanol was used for control for this experiments. All the tubes were allowed to maintain at room temperature with 45 min. Finally, the formed anti-oxidant activity tubes were monitored under UV–vis spectrophotometer at O.D of 600 nm. The anti-oxidant result was noted after interpretation with control result using bellowed equation.

\[
\% \text{ of DPPH scavenged activity} = \frac{[\text{Control}_{OD} - \text{Test}_{OD}]}{\text{Control}_{OD}} \times 100.
\]

2.8. Minimum inhibition concentration

Overnight *P. aeruginosa* culture was used in this study with different purified extract concentration into the 96-well polystyrene plate wells by microtitre plate reader (Rajivgandhi et al., 2020a, 2020b). Firstly, the confluent culture of *P. aeruginosa* volume was 10 μL in all the wells of 96-well polystyrene plate and 25–200 μg/mL of purified extract of *Sargassum wightii* was added into the same plate and slightly shaken. Then, the plate was put in inside of the incubator and set 37 °C with one day. After one day, the high and low turbidity of the wells were observed on the naked eye and followed by taken O.D values using spectrophotometer with triplicate. The result was MIC was considered as which well was shown with very low turbidity in very low concentration. That concentration was fixed as a MIC. The following formula was used to analyze the percentage conversion of identified test and control results. MIC is referred as the lowest concentration of extract was inhibited the highest bacterial growth.

\[
\text{Inhibition (\%)} = 100 \left(\frac{\text{O.D. purified extract of } Sargassum wightii - \text{ O.D. only pathogen}}{\text{O.D. only pathogen}}\right) \times 100.
\]

3. Result

3.1. Phytochemical screening of crude extract of *Sargassum wightii*

The LC-MS peaks of the *Sargassum wightii* was showed with more peaks that including 30% of phytochemical derivatives, 20% of phenol, flavonoid, alkaloid, 40% bioactive compounds and other organic compounds of aromatic hydrocarbons. Among the all peaks and their respective chemical composition, the retention time, occupation area, occupation percentages are effectively identified. The retention time, occupied area and occupied percentages of *Sargassum wightii* extract was correlated with respective 25 peaks of previous reports with anti-oxidant and biological activities, and also confirmed by NIST data base of the Bharathidasan University Wiley library (Fig. 1). In the result, the important peaks of 2,6-Di-tert-buty, 1,4-benzoquinone, benzo, 1,1’-[oxbis(methy lene)bis, dodecanic acid, phenol Oleancic acid, 7,9-Di-tert-buty1-1-oxaspiro(4,5)dec-a-6,9-diene-2,8-dione, pyrrolo[1,2-alpyra zine, 1,4-dione, hexahydro-3, hexadecane, (1-hydroxypent-2,4-di en1-yl)oxy)anthracene-9,10-dione, trifluoroacetoxy hexadecane were observed based on the previously reported anti-microbial activities. This result was indicated that the chosen *Sargassum wightii* extract was suitable seaweed for inhibition of pathogens. Very recently, was reported that the seaweed synthesized chemical and aromatic compounds have extra ordinary anti-bacterial properties. This statement was agreed by Rajivgandhi et al. (2020a, 2020b), the increased biological activities of marine sources depends on unpredicted environmental sources like extreme temperature, carbon, nitrogen content, pH, salinity and other organic nutrients. Seaweed mediated phytochemical derivatives have the ability to inhibit in inside of the pathogen and it altered the virulence genes like quorum sensing and biofilm formation. The anti-bacterial activity effect was increased in the *Sargassum wightii* extract against multi drug resistant bacterial due to the influence of different environmental conditions (Raman et al., 2014; Xinjun et al., 2020). The LC-MS was used to detect the complete phytochemical derivatives, aromatic hydrocarbons, and other polysaccharide mediated compounds. Recent reports of our previous study was supported to present result and it confirmed that the algae *Turbinaria ornate* and *Cnulera taxifolia* have more phytochemical derivatives and it shown more anti-bacterial activity against multi drug resistant bacteria (Naiyf et al., 2020; Danjie et al., 2020).

3.2. Antimicrobial activity of *P. aeruginosa* by *Sargassum wightii* crude extract

The zones of maximum and minimum inhibitions of the wells were shown with 12 and 24 mm against *P. aeruginosa* containing plate. In this assay, the 50 μg/mL for minimum and 200 μg/mL for maximum concentration were used respectively (Fig. 2). The result was suggested that the crude extract of *Sargassum wightii* has antimicrobial activity. These zones and their respective concentrations are surprisingly very excellent compared with previously reported crude extract of *Sargassum wightii* (Hanjabam et al., 2019). This may be increased due to the under estimated environment of marine habitat including pH, temperature, organic
materials, surface light observation, NaCl, car bon and nitrogen sources (Kumar and Sahoo (2017)). This information as agreed by Antonisamy et al. (2012) and changed environmental factors were influenced the biological properties. Sometimes, the bioactive compounds, phytochemical derivatives were also very improved activities due to the influence of unpredictable environment (Anjana et al., 2014). Similar result was also reported by previous report of crude extract of Sargassum wightii against gram negative bacteria with higher biological activities. Sargassum wightii has more polysaccharides compounds compared with other algae, this polysaccharides have the ability to invade inside of the pathogens and damaged the nucleus effectively. After enter the nucleus, it affected entire bacteria and arrested the cell cycle growth that leads to cell death (Kumar et al., 2015). Finally, the result was suggested that the Sargassum wightii has increased antimicrobial activity against gram negative bacteria compared to other algal extract.

3.3. Detection of phenolic content of Sargassum wightii crude extract

Our result was shown with rich content of phenolic and flavonoid in the crude extract of Sargassum wightii after compared with respective control of gallic acid and rutin correlation. The available phenolic and flavonoid properties of the extract were given for both in 25–250 μg/mL concentration (Fig. 3). This concentration was very low for Sargassum wightii when compared with other seaweed extracts that reported previously. In addition the respective controls of the gallic acid and rutin was exhibited more phenols and flavonoids and it correlated highly for Sargassum wightii only. Additionally, the total sugar and protein level was too low in this study compared with phenol and flavonoid (Fig. 4). The polysaccharide rich seaweed extract has rich phenols and flavonoids contents as well as low level of proteins and sugars (Maneesh and Chakraborty, 2017a, Maneesh and Chakraborty, 2017b). Interestingly, Sivanandhan et al. (2015) stated that the phenol and flavonoid content of the seaweed extract has more anti-oxidant activity.

3.4. Antioxidant and DPPH scavenging activity

Anti-oxidant activity of Sargassum wightii crude extract was shown with excellent result, and it also more correlated with previous phenol and flavonoid content result. Because, the polysaccharides rich phenols and flavonoids seaweed extract exhibited more biological properties including anti-oxidant, anti-cancer and anti-microbial. In our study, the control result of gallic acid and rutin almost closely related to extract result at the concentration of 200 μg/mL (Fig. 5). Both the gallic acid and rutin was more correlated with extract each other. Therefore, our result was suggested that the anti-oxidant capacity was very high in crude extract of Sargassum wightii. Further, the supportive result of DPPH scavenging assay of Sargassum wightii crude extract was also very effective at 200 μg/mL concentration. Interestingly, the DPPH result was also much closed to control result and suggested the plant mediated anti-oxidant activity was useful for other biological studies (Fig. 6). In result, the extract may influence the scavenging activity at the same 200 μg/mL concentration by the activation of
phenol and flavonoid regulatory genes (Vijayanand et al., 2014). The similar result was reported recently by Immanuel et al. (2012) and seaweed based anti-oxidant activity was detected more compared to other plant and microbial extract (Maneesh and Chakraborty (2018)). Also, the phenol, flavonoid rich seaweed extract has more anti-oxidant activity and extended to intracellular damage and high level of ROS generation in infected pathogens (Matías et al., 2020).

Fig. 3. Available phenolic content of Sargassum wightii crude extract by folin-ciocalteu's method of invivo study.

Fig. 4. Presence of flavonoid content in the Sargassum wightii crude extract by colorimetric assay.

Fig. 5. Detection of anti-oxidant activity of Sargassum wightii crude extract by phosphor molybdenum method.
3.5. Minimum inhibition concentration

The turbidity based antimicrobial efficiency of Sargassum wightii crude extract was exhibited excellent minimum inhibition concentration at very lowest concentration of 200 μg/mL. In this concentration, the turbidity was looking very low and its originality was changed gradually at increasing concentration. Additionally, the inhibition level was very high at low concentration. Based on the triplicate calculation, the inhibition percentage was 84% at the 200 μg/mL wells and it confirmed that the Sargassum wightii crude extract was more sufficient against P. aeruginosa.

The result was suggested that the Sargassum wightii extract was may inhibit the P. aeruginosa culture at concentration dependent mode. Initially, the turbidity was more in 200 μg/mL wells compared with initial well (Fig. 7). Also, the control culture was shown with clear both color after 24 h also. Previously, MIC was one of the excellent method for detect the efficiency of plant and seaweed extract. Recently, Rajivgandhi et al., 2018 agreed our result and excellent method for detect the efficiency of plant and seaweed extract have more virulence deactivation activity. Based on the result, we have concluded that the crude extract of Sargassum wightii and Halimeda gracillii has excellent antibiofilm activity at decreased concentration (Suganya et al., 2019). In addition, the antioxidative and phytochemical compounds were reported from seaweed of Sargassum wightii (Maneesh and Chakraborty, 2017a, Maneesh and Chakraborty, 2017b; J. Marimuthu et al., 2012; Johnson et al., 2019).

4. Conclusion

Based on the result, we have concluded that the crude extract of Sargassum wightii has more phytochemical derivatives, bioactive compounds and some hydrocarbons. Also, it has very rich polysaccharides content, exhibited more phenol and flavonoid content with increasing concentration. Also, the anti-oxidant activity of Sargassum wightii was indicated that the extract has an excellent anti-oxidant activity at 200 μg/mL concentration. Further, the antimicrobial activity of Sargassum wightii extract against P. aeruginosa was confirmed at 200 μg/mL concentration. Finally, the result was confirmed that the Sargassum wightii as an excellent anti-oxidant and antimicrobial agent.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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