Screening and Isolation of Bacteria Producing Biosurfactants from Waste

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Abstract

Biosurfactant are surface active compounds (SAC), amphiphilic in nature consisting of hydrophobic (long chain fatty acids) and hydrophilic domain (amino acid, protein, carbohydrate). Classification of this compound is based on molecular weight, mode of action and physico-chemical structure. Biosurfactant are being utilized in laundry formulation, house hold cleaning products, as antimicrobial agents, in detergents, pesticides, food and pharmaceutical industries. Various type of microorganism has been studied capable of synthesizing wide range biosurfactant by utilizing hydrocarbon compounds as source of carbon and energy. Present study aims to isolate biosurfactant producing bacteria from different waste sources. In this study among twenty-one isolated microbial cultures from different waste samples were screened for biosurfactant production using methods like hemolytic activity, blue gar assay, hydrocarbon tolerance assay, oil spread test, Emulsification index (E24%).

Keywords. Biosurfactant, Oil Spreading Method, Emulsification index, Modified Drop Collapse, Tolerance against Hydrocarbon, Hemolytic activity, Blue agar plate, Agar well diffusion

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INTRODUCTION

Hydrocarbon compounds are widely used in vehicles and petroleum derived compound are major cause of contamination. Hydrocarbon molecule when released into environment cannot be removed easily, as they absorbs to the surfaces. Surfactant are chemical compounds, amphiphilic in nature, tend to accumulate at the interfaces between two liquid phases with different polarities (e.g., oil-water) and reduces surface tension. Most of surfactant are synthesized chemically, and are partially biodegradable, causing noxious effect on environment and human health. Some microorganism can degrade hydrocarbon by producing biosurfactant. Microbial surfactant is more effective and versatile than many synthetic chemical surfactant owing their potential action, are biodegradable, and stable at high temperature.

The application of biosurfactant ranges from agriculture to cosmetics; from food to petrochemical and from textiles to petroleum production, oil recovery; remediation of organic and metal contaminated sites.

MATERIALS AND METHODS

Sample collection

Four types of waste were collected. These were the water drained during washing of uncooked rice, water left after cooking of rice, water drained after boiling of corn and mixture of cooked rice water and boiled corn water. The samples were collected in glass bottles sterilized in hot air oven, at 170°C for 60 minutes. These samples were left at 30°C for 48 hours. After 48 hours, coconut oil parachute brand of Marico Limited, Mumbai, India) having saturated fatty acid 91 g/200ml (in mixture and rice water), homemade ghee from buffalo milk having 86.46g saturated fatty acid, (in corn water), petrol obtained from nearby petrol pump containing paraffin 30%, naphthenes 49%, asphatic 6%, aromatic compounds 15%, respectively were added. After adding different hydrocarbons in all sample bottles, these were further incubated at room temperature for 6 days (Fig. 1).
Characterization of biosurfactant produced in waste

Each sample after 6 days incubation were analyzed for production of biosurfactant by performing various tests as mentioned herewith.

**Oil Spreading Method (OSM)**

10 ml distilled water added to Petri dish. To this, paraffin oil containing 1 ml of 0.5% Sudan black dye was added. This was followed by addition of 1 drop incubated sample onto paraffin surface. The incubated sample containing biosurfactant separated the Sudan black dye containing paraffin layer and formed a clear zone. The same procedure was done with Vim liquid soap (Hindustan Unilever Limited, India) as a control. Vim liquid soap contains lauryl sulphate, disodium EDTA, lime juice, chlorine, water.

**Emulsification index (EI24%)**

Equal volume of supernatant from incubated wastes and engine oil in one test tube (1:1), and equal volume of supernatant from incubated wastes and paraffin oil (1:1) were taken. Contents of each tube were thoroughly mixed for 2 minutes using vortex mixer. The contents in the tubes were then left to stand for 24 hours at 37°C. The same test was performed with vim liquid soap as a control. After 24 h, the emulsification activity was calculated using following formula:

\[ E24 (\%) = \frac{\text{total height of the emulsified layer}}{\text{total height of the liquid layer}} \]

**Modified Drop Collapse (MDC) assay**

Screening of biosurfactant production was performed using the qualitative drop-collapse test described by Bodour et al. and Jain et al. This assay relies on the destabilization of liquid droplets by surfactants. Engine oil (80%base oil-1-10%, alkenyl succinic, 1-10% salicylic acid, base oil –petroleum, poly alpha olifins) was applied as a coating, on glass plate. A drop of incubated sample was placed over it to analyze drop collapse. Vim liquid soap was used as control.

**Isolation of biosurfactant producing bacteria from the incubated samples by streak plate method**

Incubated wastes samples that showed biosurfactant production were streaked on nutrient agar plate, followed by incubation of plate at 37°C for 24 hours.

**Spot inoculation of isolated colonies to another nutrient agar plate**

Isolated colonies from nutrient agar plates were spot inoculated on to a new nutrient agar plate. Which were then incubated at 37°C for 24 hours (Fig. 2).
Tolerance Against Hydrocarbon (TAH)

In order to analyze the ability of isolate to grow in presence of hydrocarbon, nutrient agar plate having isolated colonies were coated with seed agar followed by spreading of engine oil over surface. The plates were then incubated at 37°C for 24-48 hours. Colony surrounded by emulsified halos was considered positive for biosurfactant production. Positive isolates were cultured in sterile medium containing tap water and engine oil as sole source of carbon.

Hemolytic activity (HA)

Isolated bacterial culture which were inoculated in the nutrient broth from broth and incubated for 4 hours at 37°C was streaked on sterile 5% blood agar plate and incubated at 37°C for 24 - 48 hours. Hemolytic activity was detected as the presence of a definite clear zone around a

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**Table 1. Modified Oil Spread (MOS) test**

| Name of the test | Control (VIM liquid soap) | Broth 1 (Rice water + coconut oil) | Broth 2 (Rice water+Corn water+Coconut oil) | Broth 3 (Corn water + ghee) | Broth 4 (corn water + petrol) | Broth 5 (garage soil + water) |
|------------------|---------------------------|-----------------------------------|---------------------------------------------|-----------------------------|-------------------------------|-----------------------------|
| Oil spread test  |                           | Oil drop                          | Slight displacement                         | Oil drop                     | Negative                      | Slight displacement          |
|                  |                           | completely displaced at edge       |                                             | completely displaced at edge |                              |                              |

**Table 2. Emulsification index**

| Name of the test | Control (VIM) | Broth 1 (Rice water + coconut oil) | Broth 2 (Rice water+Corn water+Coconut oil) | Broth 3 (Corn water + ghee) | Broth 4 (corn water + petrol) | Broth 5 (garage soil + water) |
|------------------|----------------|-----------------------------------|---------------------------------------------|-----------------------------|-------------------------------|-----------------------------|
| Emulsification index | pending | 2.94% | 3.33% | 16.66% | 6.98% | 20% |
| E24 with paraffin oil | 42% | 46% | 57% | 48% | 51% | 40% |

**Table 3. Modified Drop Collapse (MDC) test**

| Name of the test | Control (VIM) | Broth 1 (Rice water + coconut oil) | Broth 2 (Rice water+Corn water+Coconut oil) | Broth 3 (Corn water + ghee) | Broth 4 (corn water + petrol) | Broth 5 (garage soil + water) |
|------------------|----------------|-----------------------------------|---------------------------------------------|-----------------------------|-------------------------------|-----------------------------|
| Drop collapse test (Result) | + | _ | + | +++ | ++ | + |
| Diameter of collapsed drop | 0.5cm | _ | 0.5cm | 2cm | 1.0cm | 0.9cm |

**Table 4. Colony characters of isolates on nutrient agar plate (rice water+coconut oil)**

| Colony characters | Sector 1 | Sector 2 | Sector 3 | Sector 4 | Sector 5 | Sector 6 |
|-------------------|----------|----------|----------|----------|----------|----------|
| Size              | Small    | Small    | Small    | Large    | Moderate | Large    |
| Shape             | Round    | Irregular| Round    | Irregular| Irregular| Regular  |
| Margin            | Entire   | Entire   | Entire   | Curly    | Entire   | Lobate   |
| Elevation         | Flat     | Flat     | Flat     | Flat     | Raised   | Flat     |
| Texture           | Smooth   | Smooth   | Smooth   | Rough    | Rough    | Rough    |
| Pigmentation      | Colorless| Yellowish| Colorless| Colorless| Dull white| Colorless|
| Opacity           | Opaque   | Opaque   | Opaque   | Opaque   | Opaque   | Opaque   |

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colony.12-14 Gram staining performed from isolated colony from blood agar plate.

Blue agar plate (BAP) method

Mineral salt agar medium supplemented with engine oil (2ml). Wells were made using sterile cup borer on methylene blue agar plate which were loaded with broth inoculated with isolated bacterial culture and incubated for 4 hours at 37°C. This was followed by incubation at 37°C for 24-48 hours. A dark blue halo zone around the wells with active culture was considered positive for anionic biosurfactant production.

Agar well diffusion (AWD) assay

The ionic property of cell bound biosurfactant producing strain was determine by using agar well diffusion method. Uniformly

Table 5. Colony characters of isolates on blue agar plate (rice water coconut oil+corn water)

| Colony characters | Observation                              |
|------------------|------------------------------------------|
| Size             | Small                                    |
| Shape            | Round                                    |
| Margin           | Entire                                   |
| Elevation        | Flat                                     |
| Texture          | Dry                                      |
| Color            | Dark blue (indicates positive result)    |
| Opacity          | Opaque                                   |

Table 6. Colony characters of isolates on nutrient agar plate (corn water +ghee)

| Colony characters | Sector 1 | Sector 2 | Sector 3 | Sector 4 | Sector 5 | Sector 6 |
|-------------------|----------|----------|----------|----------|----------|----------|
| Size              | Moderate | Moderate | Moderate | Large    | Moderate | Small    |
| Shape             | Irregular| Round    | Round    | Irregular| Round    | Irregular|
| Margin            | Lobate   | Entire   | Entire   | Lobate   | Entire   | Entire   |
| Elevation         | Flat     | Raised   | Flat     | Raised   | Flat     | Flat     |
| Texture           | Smooth   | Smooth   | Smooth   | Rough    | Smooth   | Rough    |
| Pigmentation      | Colorless| Yellowish| Colorless| Colorless| Dull white| Colorless|
| Opacity           | Opaque   | Opaque   | Opaque   | Opaque   | Opaque   | Opaque   |

Table 7. Colony characters of isolates on nutrient agar plate (corn water +petrol)

| Colony characters | Sector 1 | Sector 2 | Sector 3 | Sector 4 |
|-------------------|----------|----------|----------|----------|
| Size              | Small    | Large    | Large    | Small    |
| Shape             | Irregular| Irregular| Irregular| Round    |
| Margin            | Entire   | Filamentous| Lobate   | Entire   |
| Elevation         | Raised   | Flat     | Flat     | Flat     |
| Texture           | Smooth   | Smooth   | Smooth   | Smooth   |
| Pigmentation      | Yellowish| Off white| Colorless| Colorless|
| Opacity           | Opaque   | Opaque   | Opaque   | Opaque   |

Determination of anionic and cationic characteristics

Anionic or cationic property of biosurfactant can be analyzed by using methylene blue and methyl orange. Where cationic surfactant can be analyzed by using methyl orange dye and anionic biosurfactant can be analyzed by using methylene blue dye. To perform this test about 1 ml of sample was taken in test tube followed by adding methylene blue in one tube to determine anionic characteristics and addition of methyl orange for cationic characteristics. Extracted with chloroform left for overnight. Observed for blue or yellow colour.

RESULTS

The results of each sample after 6 days incubation which were analyzed for production and characterization of biosurfactant are discussed here.
**Oil Spreading Method (OSM)**

The incubated sample containing biosurfactant separated the Sudan black dye containing paraffin layer and formed a clear zone. Same result was observed with positive control (vim liquid soap). Results obtained with different samples are as shown in Table 1. Corn water along with ghee shows higher degree for oil displacement.

**Emulsification index**

Emulsification test of broth was performed with engine oil and paraffin oil. E24 obtained for each broth after 24 hour is using above formula:

\[ E24(\%) = \frac{\text{total height of the emulsified layer}}{\text{total height of the liquid layer}} \]

Higher emulsification index was observed from broth of corn water with petrol for both servo engine oil and paraffin oil.

**Modified Drop Collapse (MDC) test**

Drop got collapsed after 1min showing different diameter for each broth while water taken as control showed negative result. Results are shown in Table 3. Large diameter for oil drop collapse observed with broth containing corn water and petrol.

**Results of isolates producing biosurfactant**

Among 21 selected isolates obtained from different waste sources, 4 isolates showed positive result for hydrocarbon tolerance assay and 7 isolates exhibited hemolytic activity.

**Isolation by streak plate method**

Colonial morphology of all isolates was observed. 6 colonies observed on sector streaked with broth (corn water +ghee), 16 colonies observed on sector streaked with broth containing (corn water + petrol). 16 colonies observed on sector streaked with broth containing (Mix+ coconut oil). 15 colonies observed on sector streaked with broth containing (Rice water+ coconut oil).

**Spot inoculation of isolated colonies to another nutrient agar plate**

Colonial morphology of all isolates observed as shown in tables 4-7 and Fig. 2.

**Tolerance Against Hydrocarbon (TAH)**

Colony surrounded by emulsified halo was observed and selected for further work, inoculated in broth (distilled water and engine oil). Results for each broth are shown in Table 8. and Fig. 3.

**Hemolytic Activity**

Zone of hemolysis was observed with the various diameter. Hemolytic activity for each broth is along with colonial morphology and Gram staining Results of gram staining are illustrated in Table 9. Inoculated colonies utilized engine oil as carbon source, turbidity was observed over the surface of the each broth after 48 hours of incubation (Fig. 4).
Blue agar plate method
Isolates produced dark blue halo around the well which was considered positive activity for biosurfactant production. Results for each broth is shown in Table 10 (Fig. 5).

Agar well diffusion assay
The taste is based on diffusion of two compounds bearing charges of the same or opposite type in a weakly concentrated gel, revealed precipitation lines between two compounds, no precipitation lines observed in any corresponding loaded well as shown in Table 11.

Determination of anionic and cationic characteristics
Test performed from each broth was observed for blue or yellow color after extraction with chloroform. Blue color was observed indicating anionic characteristics of isolated biosurfactant (Table 12).

DISCUSSION
Biosurfactant has wide range of application in remediation of hydrocarbons as they can intensify the growth of bacteria producing biosurfactant on hydrophobic surface and allow it to take nutrients from hydrophobic surface. In this project detail study was done on isolate from each collected sample. Waste used here are from corn and cooked rice. Corn contains higher amount of starch acts as good substrate for bacterial growth. Rodrigo et al.\textsuperscript{16} reported that oil spread technique is sensitive method for detection of small amount of biosurfactant produced. Here in oil spread assay in which sample 1 showed complete displacement of oil in edge indicating its greater potential to displace paraffin oil compared to other collected samples. Similar results were observed for modified drop collapse test. Upon screening among twenty-one cultures five isolates were able to tolerate hydrocarbons, in which all the isolates are Gram positive \textit{Bacillus}. These isolates showed positive result for hydrocarbon tolerance while the same test was found to be negative for \textit{Bacillus} with the hydrocarbon toluene 6. Hemolytic activity is used as preliminary test to describe biosurfactant production. Biosurfactant can lyse erythrocytes if biosurfactant is produced by organism. In this study, all the isolates produces zone of hemolysis with the size more than 2.30 cm. Bacteria isolated from samples 1, 2 and 3 showed more hemolytic activity compared to hemolytic activity of \textit{Bacillus} culture that showed zone of hemolysis around 1.0 to 2.5 in the study performed by Thavasi et al\textsuperscript{17}. Blue agar plate method results were found similar to activity by \textit{Pseudomonas}.\textsuperscript{18} Potential activity for biosurfactant production by bacteria as can be determined by performing emulsification activity. Here emulsification activity was performed with three different hydrocarbons, servo engine oil, diesel and liquid paraffin. Highest emulsification was observed toward hydrocarbon servo engine oil which is high compared to the report of Nishanthi et al.\textsuperscript{19} The isolates obtained have shown promising results, so more studies
### Table 8. Hydrocarbon tolerance assay

| Name of the test | Broth 1 (Rice water + coconut oil) | Broth 2 (Rice water + corn water + coconut oil) | Broth 3 (Corn water + ghee) | Broth 4 (corn water + petrol) | Broth 5 (garage soil + water) |
|------------------|-----------------------------------|-----------------------------------------------|----------------------------|-------------------------------|-------------------------------|
| Hydrocarbon tolerance assay | Colony showing emulsified halo observed on sector 4 colony which is large in size, with irregular shape, undulate margin, and rough in texture. Flat elevation, colorless. | Colony showing emulsified halo observed on sector 2 colony which is medium in size, with round shape, entire margin and smooth texture. Raised elevation off white in color. | Colony showing emulsified halo observed on sector 2 colony which is medium in size, with round shape, entire margin and smooth texture. Raised elevation off white in color. | Colony showing emulsified halo observed on sector 4 colony which is large in size, with irregular shape, wrinkled margin, and rough in texture. Raised elevation off white in color. | Colony showing emulsified halo observed on coated agar is large in size, with irregular shape, and rough in texture. |

### Table 9. Hemolytic Activity (HA) and Gram staining of isolates

| Name of the test | Broth 1 (Rice water + coconut oil) | Broth 2 (Rice water + corn water + coconut oil) | Broth 3 (Corn water + ghee) | Broth 4 (corn water + petrol) | Broth 5 (garage soil + water) |
|------------------|-----------------------------------|-----------------------------------------------|----------------------------|-------------------------------|-------------------------------|
| Hemolytic activity | Zone of beta hemolysis is observed with diameter of 3.83CM. Colony morphology: large, irregular margin, smooth, opaque, no pigmentation. Gram positive, rods, chain, non-swelled spores centrally located. | Zone of beta hemolysis observed with diameter of 2.33CM. Colony morphology: medium, irregular in shape dry, opaque no pigmentation. Gram positive short rods with round ends arranged single and chain. | Zone of beta hemolysis is observed with diameter of 26CM. Colony morphology: Small, irregular in shape wrinkled, dry, opaque. Gram negative, rod. | Zone of beta hemolysis observed with the diameter of colonial morphology is Large, irregular Flowery, flat 3.66Dry, slight reddish, opaque Gram positive rod arranged in chain with, round ends. | Zone of beta hemolysis observed with the diameter of colonial morphology is Large, irregular Flowery, flat 3.66Dry, slight reddish, opaque Gram positive rod arranged in chain with, round ends. |
### Table 10. Blue agar plate method

| Name of the test | Broth 1  (Rice water + coconut oil) | Broth 2  (Rice water + corn water + coconut oil) | Broth 3  (Corn water + ghee) | Broth 4  (Corn water + petrol) | Broth 5  (Garage soil + water) |
|------------------|------------------------------------|-----------------------------------------------|----------------------------|-------------------------------|--------------------------------|
| Blue agar plate method | Positive. Well surrounded by dark blue color observed. | Positive. Well surrounded by dark blue color observed. | Positive. Well surrounded by dark blue color observed. | Positive. Colony surrounded by dark blue colour is observed. | Positive. Well surrounded by dark blue color observed. |

### Table 11. Agar well diffusion assay

| Name of the test | Broth 1  (Rice water + coconut oil) | Broth 2  (Rice water + corn water + coconut oil) | Broth 3  (Corn water + ghee) | Broth 4  (Corn water + petrol) | Broth 5  (Garage soil + water) |
|------------------|------------------------------------|-----------------------------------------------|----------------------------|-------------------------------|--------------------------------|
| Agar well diffusion assay | No precipitation line observed. | No precipitation line observed. | No precipitation line observed. | No precipitation line observed. | No precipitation line observed. |

### Table 12. Determination of anionic and cationic characteristics

| Determination of Anionic/Cationic characteristics | Sodium lauryl sulphate. (Anionic standard) | Cetyl pyridinium chloride. (Cationic standard) | Sample 1 | Sample 2 | Sample 3  (Rice water + coconut oil) | Sample 4 | Sample 5 |
|--------------------------------------------------|--------------------------------------------|----------------------------------------------|----------|----------|------------------------------------|----------|----------|
| Observation Result | Blue Anionic | Yellow Cationic | Blue Anionic | Blue Anionic | Blue Anionic | Blue Anionic | Blue Anionic |

**Fig. 5.** Blue agar plate method.
regarding the isolates and the biosurfactant produced would be taken up in future studies.

CONCLUSION
These tests provide rapid and easy screening of biosurfactant producing bacteria. Upon observation of results all isolates studied gives positive results for biosurfactant production. Among all studied bacterial cultures isolates from sample 1 shows highest potential for biosurfactant production followed by isolates of sample 3. Among five isolated cultures, four cultures are gram positive one is gram negative having potential to produce biosurfactant can be used for biodegradation of hydrocarbons, and other industrial processes.

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CONFLICT OF INTEREST
The authors declare that there is no conflict of interest.

AUTHORS’ CONTRIBUTION
All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

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DATA AVAILABILITY
All datasets generated or analyzed during this study are included in the manuscript.

ETHICS STATEMENT
This article does not contain any studies with human participants or animals performed by any of the authors.

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