DNA photo-cross-linking using a pyranocarbazole-modified oligodeoxynucleotide with a D-threoninol linker†

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An alternative photo-cross-linker having a D-threoninol skeleton instead of the 2'-deoxyribose backbone in 3-cyanovinylcarbazole (CNVK) was investigated to improve the photoreactivity of photo-cross-linkers; the photo-cross-linking rate of 3-cyanovinylcarbazole with D-threoninol (CNVD) was found to be greater than that of CNVK. Therefore, in this study, a novel photo-cross-linker having pyranocarbazole (PCX) and D-threoninol instead of the 2'-deoxyribose backbone in PCX (PCXD) was developed. The PCXD in double-stranded DNA photo-cross-linked to a pyrimidine base at the 1 position of a complementary strand similar to PCX. Furthermore, the photoreactivity of PCXD was significantly higher than that of PCX. The introduction of D-threoninol improved the reactivity of pyranocarbazole to cytosine, the use of PCXD may extend the applicability of the photo-cross-linking reaction for DNA manipulation. In particular, this novel photo-cross-linker can contribute to the photochemical regulation of gene expression or biological events in a living cell.

Introduction

Photo-cross-linking reactions between biomolecules are used for various applications such as screening antigen interactions1 and improving detection sensitivity2 and the stability of biomolecular complexes.3 In particular, the use of DNA photo-cross-linking in the formation of a thymine dimer induced by UV-irradiation4 and interstrand photo-cross-linking with psoralen5 has been reported. In recent years, with the development of nucleic acid medicine and DNA nanotechnology, these have been used for the photochemical regulation of an antisense effect,6 improvement in stability of a DNA nanostructure,7 and other applications. Many photo-cross-linkers such as bromouracil8 and benzophenone9 have also been reported. Among these, DNA photo-cross-linking via [2 + 2] photocycloaddition by psoralens3 and coumarins10 is useful for detection, manipulation, and regulation of nucleic acids because photo-reversible manipulation is possible. Compared to the enzymatic method, photo-cross-linking can be used under a wide range of conditions without the addition of reagents. Although the intracellular usage of photo-cross-linking has already been reported, these require UV-irradiation, which poses a limitation for use in living cells because of phototoxicity. We report pyranocarbazole (PCX) as a photo-cross-linker that can photo-cross-link to pyrimidine in complementary DNA or RNA strand under visible light.11 It was anticipated that his photo-cross-linker would accelerate the intracellular application of nucleic acid photo-cross-linking such as photochemical regulation of gene expression12 and detection of RNA strand;13 however, photo-cross-linking using PCX to cytosine requires photoirradiation for 1 min, and it is necessary to speed up this process. Besides, the ribose backbone, D-threoninol backbone,14 and serinol backbone15 have been reported as the backbone of the artificial nucleic acid. It was determined that 3-cyanovinylcarbazole modified D-threoninol (CNVD)16 considerably accelerated the photo-cross-linking reaction with cytosine using the D-threoninol backbone (PCXD) (Fig. 1).

Fig. 1 Ultrafast photo-cross-linker (a) pyranocarbazole with D-threoninol (PCXD); (b) pyranocarbazole (PCX).
Results and discussion

PCXD was successfully synthesized according to the reaction scheme shown in Fig. 2. The PCXD was phosphoramidated using a general method \(^\text{17}\) after 5’ DMTr protection to synthesize the oligodeoxynucleotide (ODN) containing PCXD. The 9 mer ODN which has A, T, G, C having deoxyribose and PCXD having \(\nu\)-threoninol was synthesized with an automated DNA synthesis machine, and then, it was deprotected using 28% ammonium solution with a general method. After HPLC purification, it was analyzed using Matrix assisted Laser Desorption/Ionization (MALDI) analysis (Table 1).

First, we demonstrated the photo-cross-linking of PCXD in double-stranded DNA. The duplex containing PCXD was irradiated with 400 nm and analyzed using ultrahigh-performance liquid chromatography (UPLC). In the case of the pyrimidine base at the –1 position on the complementary ODN (cODN) strand, new peak identical to the photodimer appeared after photoradiation, although such new peak did not appear in the case of the purine base at the –1 position on the cODN strand. The new peak was identified to photoadduct of PCXD-ODN(C) and its complementary strand (found = 5591.2, calcd \([M + H]^+ = 5591.0\) by MALDI-TOF-MS analysis. This suggests that the photo-cross-linking reaction occurred only in the case of the pyrimidine base at the –1 position on the cODN strand.

To analyze the photoreactivity of PCXD, the time course of the photoreaction was monitored, and then, the reaction rate constant was analyzed with the assumption of first-order reaction kinetics. As shown in Fig. 3, the reaction rate constant of PCXD with cytosine is 4.3-fold larger than that of PCX, suggesting that the relatively flexible \(\nu\)-threoninol skeleton elevates the accessibility of the reactive double bond to the cytosine base on the complementary strand. The same effect was observed in the case of thymine as the target base of PCX; the reaction rate constant was 1.1-folds larger than that of PCX. The difference in the reactivity between the thymine and the cytosine bases decreased and the photoreactivity enhanced compared with PCX, which indicates that the use of PCXD extends the applicability of the photo-cross-linking reaction for DNA manipulation.

To evaluate the duplex stability of ODN(PCXD), the melting temperature \(T_m\) of the duplex consisting of ODN(A\(\text{PCXD}\)) and ODN(GT) was evaluated and compared with the duplex including PCX. The sigmoidal melting curve of ODN(PCXD)/ODN(T) duplex was observed, which suggests ODN(PCXD) and cODN(GT) formed a duplex structure. Then, we calculated the thermodynamic parameter from van’t Hoff plots.\(^\text{18}\) The \(\Delta S\) through ODN(PCXD)/ODN(T) hybridization was smaller than that of ODN(PCX)/ODN(T), suggesting that a relatively flexibility structure of PCXD inhibited the entropic loss via hybridization (Table 2). Therefore, PCXD has a higher flexibility than PCX, and it induces a higher photo-cross-linking ability in cytosine than PCX. In addition, PCX and PCXD containing pyranocarbazole moieties have lower \(T_m\) values compared to \(CNVK\) and \(CNVD\) containing 3-cyanovinylcarbazole moiety. When the polarity of pyranocarbazole (log \(P = 1.12\)) and 3-cyanovinylcarbazole (log \(P = 1.35\)) was examined,\(^\text{19}\) it was found that pyranocarbazole has

![Fig. 2](image-url)  

**Fig. 2** Synthesis of PCXD phosphoramidite.
a smaller log $P$ and is hydrophilic. Therefore, it is considered that the $T_m$ value of the duplex containing 3-cyanovinylcarbazole is high and the entropy loss is large because of the stacking between bases and hydrophobic interactions.

To evaluate the effects of the surrounding bases on the photo-cross-linking of ODNs containing PCXD, double strand and four di 

Table 2 Thermodynamic parameter of each photo-cross-linker

| Entry | $T_m$ [°C] | $\Delta G_{37}$ (kcal mol$^{-1}$) | $\Delta H$ (kcal mol$^{-1}$) | $\Delta S$ (cal mol$^{-1}$ K$^{-1}$) |
|-------|-----------|-------------------------------|-----------------------------|----------------------------------|
| PCXD  | 26        | -7.4                          | -16.7                       | -30.0                            |
| PCCX  | 33        | -0.58                         | -21.6                       | -67.7                            |
| CNXD  | 29        | -7.3                          | -33.8                       | -85.6                            |
| CNXK  | 34        | -7.5                          | -63.6                       | -181.1                           |

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Experimental

General

$^1$H NMR spectra were measured on a Bruker AVANCE III 400 system. Mass spectra were recorded on a Voyager PRO-SF, Applied Biosystems and HRMS were measured on a Solarix-JA (Bruker) HPLC was performed on a Chemcosorb 5-ODS-H column with JASCO PU-980, HG-980-31, DG-980-50 system equipped with a JASCO UV 970 detector at 260 nm. Reagents for synthesis for compound 1 to 5 were purchased from Aldrich, Wako, and TCI. Phosphoramidites of PCXD were synthesized by the procedure as follows. Other phosphoramidite reagents for the DNA synthesizer such as A, G, C, T-b-cyanoeethyl phosphoramidite, and CPG support were purchased from Glen research. Other oligonucleotides were purchased from Fasmac (Japan) and used without further purification. The synthesized ODN having deoxyribose backbone except PCXD.

Synthetic procedures

**Compound 2.** Pyranocarbazole (1) (628 mg, 2.67 mmol) and NaH (60% oil suspension, 320 mg, 13.4 mmol) were solved in dry DMF (10 mL) and stirred for 20 min on ice under N$_2$ atmosphere. Ethyl bromoacetate (0.59 mL, 5.34 mmol) and NaI (1.2 g, 8.0 mmol) was drop-wisely added over 30 min and stirred for 90 min at room temperature. Reaction mixture was poured onto water (200 mL) and extracted with chloroform and then dried over sodium sulfate. After removal of the solvent the residue was subjected to silica gel column chromatography (CHCl$_3$) to afford 2 (yellow solid, 1.4 g, 4.6 mmol, 32%).$^1$H NMR (400 MHz, CDCl$_3$) 8.06 (s, 1H), 8.01 (d, 1H, $J$ = 7.6 Hz), 7.8 (d, 1H, $J$ = 8.0 Hz), 7.44 (m, 1H), 7.25 (d, 1H, $J$ = 9.2 Hz), 7.24 (m, 1H), 7.13 (s, 1H), 6.26 (d, 1H, $J$ = 9.2 Hz), 4.91 (s, 2H), 4.17 (m, 1H), 1.19 (t, 3H, $J$ = 7.2 Hz).$^{13}$C NMR (100 MHz, CDCl$_3$) 167.8, 161.6, 153.5, 144.6, 143.1, 141.7, 126.9, 122.7, 121.1, 121.0, 120.6, 119.7, 112.9, 112.7, 108.9, 96.1, 62.2, 45.0, 14.3. ESI-FT-ICR MS(HRMS) calcd for [M + H]$^+$: 322.1073, found: 322.1073.

**Compound 3.** 2 (402 mg, 1.25 mmol) and NaOH (110 mg, 2.75 mmol) were solved in THF/MeOH/H$_2$O (3 : 2 : 1, 18 mL) and stirred for 90 min at room temperature. After the addition of 1 N HCl (150 mL), reaction mixture was extracted with EtOAc (150 mL) and washed with 1 N HCl. Organic layer was dried over MgSO$_4$ and evaporated to afford 3 (yellow solid, 1.2 g, 4.4 mmol, 96%).$^1$H NMR (400 MHz, DMSO-d$_6$) 13.17 (s, 1H), 8.50 (s, 1H), 8.19 (d, 2H, $J$ = 9.2 Hz), 7.65 (s, 1H), 7.59 (d, 1H, $J$ = 8.4 Hz), 7.50 (m, 1H), 7.30 (t, 1H, $J$ = 6.8 Hz), 6.36 (m, 1H), 5.30 (d, 2H).$^{13}$C NMR (100 MHz, DMSO-d$_6$) 169.8, 160.6, 152.8, 145.4, 134.0, 141.6, 126.5, 121.9, 120.4, 120.3, 120.1, 112.0, 119.7, 112.9, 112.7, 108.9, 96.1, 62.2, 45.0, 14.3. ESI-FT-ICR MS(HRMS) calcd for [M + H]$^+$: 306.0760, found: 306.0759.

**Compound 4.** 3 (156 mg, 0.53 mmol) and $\alpha$-threoninol (111 mg, 1.06 mmol) were dissolved in dry DMF (10 mL) containing 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride (203 mg, 1.06 mmol) and 1-hydroxybenzotriazole (143 mg, 1.06 mmol), and stirred for 11 h at room temperature. NaCl aq. soln was added and obtained precipitate was collected and dried under vacuum to afford 4 (yellow solid, 1.5 g, 4.1 mmol, 91%).$^1$H NMR (400 MHz, DMSO-d$_6$) 8.50 (s, 1H), 8.18~8.21 (m, 2H), 8.00 (d, 1H, $J$ = 8.8 Hz), 7.58~7.60 (m, 2H), 7.50 (t, 1H, $J$ = 7.2 Hz), 7.30 (t, 1H, $J$ = 7.2 Hz), 6.34 (d, 1H, $J$ = 9.2 Hz), 5.17 (d, 2H, $J$ = 3.6 Hz), 4.72 (d, 2H, $J$ = 4.7 Hz), 4.64 (t, 3H, $J$ = 4.2 Hz), 3.91 (m, 1H), 3.65 (br, 1H), 3.47~3.54 (m, 1H), 3.36~3.41 (m, 1H), 1.02 (d, 3H, $J$ = 6.4 Hz).$^{13}$C NMR (100 MHz, DMSO-d$_6$) 167.0, 160.6, 152.8, 145.4, 143.1, 141.8, 126.4, 121.8, 120.3, 120.1, 120.1,

Fig. 4 Effect of surrounding base of PCXD on its photoreactivity in dsDNA. 10 µM DNA in 50 mM cacodylate buffer (pH 7.4) containing 100 mM NaCl was 400 nm-irradiated for 10 s at 4 °C.
119.9, 111.9, 111.8, 109.8, 96.3, 63.9, 60.5, 55.9, 45.6, 20.3. ESI-FT-ICR MS(HRMS) calecd for [M + H]+: 381.1444, found: 381.1445. The molar extinction coefficient of PCX-D is 4400 M−1 cm−1.

**Compound 5.** 4 (97.6 mg, 0.26 mmol), 4,4′-dimethoxytritylchloride (105 mg, 0.31 mmol) and 4-dimethyl-aminopyridine (105 mg, 0.31 mmol) in dry pyridine (1 mL) were stirred at room temperature for 18 h. The reaction mixture was diluted with CHCl3, washed with H2O and organic layer was dried over Na2SO4. After the removal of the solvent, residue was subjected to silica gel column chromatography (CHCl3 with 0.1% TEA) to afford 5 (yellow solid, 0.63 g, 86%). 1H NMR (400 MHz, DMSO-d6) 8.47 (s, 1H) 8.17 (t, 3H, J = 8.8 Hz) 7.59 (d, 1H, J = 8.4 Hz), 7.57 (s, 1H), 7.44 (t, 1H, J = 8.8 Hz) 7.33–7.18 (m, 10H), 6.78 (t, 4H, J = 8.8 Hz), 6.31 (d, J = 1H, 9.2 Hz), 5.16 (s, 1H), 4.75 (s, 1H), 3.91 (d, 2H, 6.0 Hz) 3.70 (s, 6H), 3.11 (t, 1H, 4.6 Hz, 6.4 Hz), 2.93 (t, 1H, 2 Hz, 6.0 Hz). 13C NMR (100 MHz, DMSO-d6) 167.0, 160.5, 157.9, 157.8, 152.8, 145.4, 144.4, 144.9, 143.0, 141.8, 135.8, 135.7, 129.6, 129.6, 127.6, 126.4, 126.3, 121.8, 120.3, 120.1, 120.0, 113.0, 112.9, 111.9, 111.7, 109.9, 96.4, 85.2, 79.1, 64.8, 62.9, 54.9, 54.1, 45.7, 20.3. ESI-FT-ICR MS(HRMS) calecd for [M + H]+: 705.2571, found: 705.2568.

**Compound 6.** 5 (0187 mg, 0.28 mmol), 0.25 M solution of 5-benzylthio-1H-tetrazole (2.48 mL) in dry CH2Cl2 (10 mL) and 2-cyanoethyl N,N,N′,N′-tetraisopropylphosphorodiamidite (540 μL, 0.62 mmol) were mixed under N2 for 3 h. The crude mixture was dissolved in ethyl acetate. The solution containing 6 was washed with water, NaHCO3 aq. soln and brine. The organic layer was dried over MgSO4, and then filtered and the ethyl acetate was removed. The yellow foam (0.21 g, 0.24 mmol, 80%) was obtained and immediately used for DNA synthesis without further purification.

**Synthesis of oligodeoxynucleotide**

The oligonucleotides having PCX-D were synthesized by a 3400 DNA synthesizer (Applied Biosystems) and purified by a HPLC system (JASCO PU-980, HG-980-31, DG-980-50, UV-970) with an InertSustainTM C18 column (GL Science, 5 μm, 10 × 150 mm). Synthesized oligonucleotides was identified by MALDI-TOF-MS.

**Photoirradiation and UPLC analysis**

Photoirradiation was performed with an LED lamp (OminiCure, 400 nm, 8600 mW cm−2) on thermal cycler. The photoirradiated samples were analyzed with a UPLC system (Aquity, waters) equipped with BEH Shield RP18 column (1.7 μm, 2.1 × 50 mm), elution was with 0.05 M ammonium formate containing 1–10% CH3CN, linear gradient (10 min) at a flow rate of 0.4 mL min−1, 60 °C).

**Thermodynamic analysis of the hybridization**

Thermodynamic parameters were obtained according to a method in the literature: where TM is the melting temperature of duplex, ΔS° is entropy and ΔH is the entropy of duplex formation, respectively. R is the gas constant and CT is the total strand concentration. TM was measured at various concentrations of duplex in 50 mM Na-cacodylate buffer (pH 7.4) containing 100 mM NaCl by a spectrophotometer (V-630bio, Jasco) equipped with a temperature controller.

**Conclusions**

The PCX-D in double-stranded DNA photo-cross-linked to the pyrimidine base at the −1 position of the complementary strand was similar to that for PCX, and the photoreactivity of PCX-D was significantly higher than that of PCX. The photo-cross-linking rate of PCX-D having γ-threoinol and cytosine was faster than that of PCX and cytosine, the applicability of the photo-cross-linking reaction for DNA manipulation may be expanded by the use of PCX-D. We have already reported the importance of photo-cross-linking rate in photochemical antisense method in living cell and RNA FISH to E. coli 16S rRNA with a secondary structure. Therefore, this novel photo-cross-linker would contribute to the photoregulation of gene expression or biological events in living cells.

**Conflicts of interest**

There are no conflicts to declare.

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