Tunisian Maturity-Onset Diabetes of the Young: A Short Review and a New Molecular and Clinical Investigation

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Introduction/Aims: Maturity-Onset Diabetes of the Young (MODY) is a monogenic non-autoimmune diabetes with 14 different genetic forms. MODY-related mutations are rarely found in the Tunisian population. Here, we explored MODY related genes sequences among seventeen unrelated Tunisian probands qualifying the MODY clinical criteria.

Materials and Methods: The GCK and HNF1A genes were systematically analyzed by direct sequencing in all probands. Then, clinical exome sequencing of 4,813 genes was performed on three unrelated patients. Among them, 130 genes have been reported to be involved in the regulation of glucose metabolism, β-cell development, differentiation and function. All identified variants were analyzed according to their frequencies in the GnomAD database and validated by direct sequencing.

Results: We identified the previously reported GCK mutation (rs1085307455) in one patient. The clinical features of the MODY2 proband were similar to previous reports. In this study, we revealed rare and novel alterations in GCK (rs780806456) and ABCC8 (rs201499958) genes with uncertain significance. We also found two likely benign alterations in HNF1A (rs1800574) and KLF11 (rs35927125) genes with minor allele frequencies similar to those depicted in public databases. No pathogenic variants have been identified through clinical exome analysis.

Conclusions: The most appropriate patients were selected, following a strict clinical screening approach, for genetic testing. However, the known MODY1-13 genes could not explain most of the Tunisian MODY cases, suggesting the involvement of unidentified genes in the majority of Tunisian affected families.

Keywords: MODY, genetic testing, Sanger sequencing, GCK, HNF1A, clinical exome sequencing
INTRODUCTION

Maturity-Onset Diabetes of the Young (MODY) is a dominantly inherited form of non-autoimmune monogenic diabetes. Mutations in one of the key genes related to β-cell function and pancreas development are the main factors of this disease (1). So far, the Online Mendelian Inheritance in Man (OMIM) database has summarized 14 genes responsible for the emergence of MODY subtypes (OMIM # 606391) (2). More than 75% of MODY cases are the outcome of mutations in four genes [Hepatocyte Nuclear Factor (HNF) 4 Alpha (HNF4A), HNF1A, HNF 1 Beta (HNF1B), and glucokinase (GCK)] but their frequency varies among tested populations (3, 4). On the other hand, the development of clinical molecular diagnostic technologies has facilitated the discovery of new and unclassified genes related to MODY (5, 6, 7). However, many young-onset diabetic cases exhibiting MODY-like phenotype remain genetically unexplained (MODY-X) (8, 9).

MODY gene mutations manifest clinically heterogeneous features in terms of age of onset, the pattern of hyperglycemia, and treatment modalities (9, 10). Therefore, an accurate molecular diagnosis of MODY is pivotal for early prognosis, treatment, and genetic counseling (1).

MODY represents less than 6% of entire diabetes cases (2, 11) and contributes to 1–6% of pediatric diabetes cases in European and North American populations (12). The factors such as undiagnosed cases, misdiagnosed patients as other types of diabetes (type 1 (T1D), type 2 (T2D), or gestational diabetes), and variable methods of clinical ascertainment (13, 14) result in inaccurate MODY prevalence reports globally (15).

In Tunisia, a few MODY cohorts have been screened but the accurate prevalence is not known. The clinical and familial inclusion criteria were lucid but specific for each investigation (Figure 1). The first study identified c.1357A>G mutation in the HNF4A gene of one patient, whereas 11 patients were negative for mutations in HNF4A, HNF1A, HNF1B, GCK, Pancreatic and Duodenal Homeobox 1 (PDX1), insulin (INS), and Neuronal Differentiation 1 (NEUROD1) genes (16). Index cases of six out of 11 families were also negative for mutations in paired box 4 (PAX4) gene (17). The second study reported P291fsinsC mutation in the HNFIA gene (18). Khelifa et al. (19) also studied 23 unrelated families and screened four MODY genes (INS, GCK, HNF1A, and HNF4A) for mutations. Two variants were found in GCK (c.-457C>T) and HNF4A (c.-169C>T) genes of three unrelated probands. Dallali et al. (20) studied 11 suspected MODY patients and identified five heterozygous pathogenic variants in four patients [GCK p.Arg191Trp, HNF1A c.710A>G, adenosine triphosphate (ATP)-binding cassette, sub-family, member 8 (ABCC8) [c.2376delC and c.4606+4A>G found at compound heterozygous state] and c.4606G>A]. Another study has also reported two pathogenic variants in HNF1A (c.476G>A) and HNF4A (c.763C>T) genes (21).

During this study, a cohort of seventeen unrelated Tunisian probands was selected based on strict clinical and familial diagnostic criteria common for all MODY subtypes. These probands were screened and analyzed for genotype-phenotype correlations.

MATERIALS AND METHODS

Participants and Clinical Data

Patients were recruited through the Departments of Endocrinology and Pediatric of Hedi Chaker University Hospital (Sfax, Tunisia). The study was approved by the Regional Committee of the Protection of Persons, Sfax, Tunisia (CPP SUD N°28/2019). Patients, their parents (if under the age of 18), and participating family members provided their written informed consent to participate in this study.

Details about the medical history of the patients and their extended family members (where available) were documented. Classification and diagnosis of diabetes were based on the latest
American Diabetes Association guidelines (22). Diabetes diagnostic criteria were based on thresholds of fasting plasma glucose, and/or 2-hour (2-h) plasma glucose after a 75-gram (75-g) oral glucose tolerance test (OGTT), and/or glycated hemoglobin (HbA1c) measures. The clinical and metabolic data included: date of birth, gender, nationality, native place, age at diagnosis, diagnostic approach, body mass index (BMI), fasting plasma glucose, HbA1c, diabetes autoantibodies [islet cell antibody (ICA), anti-glutamic acid decarboxylase (GAD), anti-insulin and antibody to protein tyrosine phosphatase (IA-2)], C-peptide, serum insulin, lipid profile [total cholesterol, triglycerides, high-density lipoprotein (HDL) and low-density lipoprotein (LDL)], liver function profile [alanine aminotransferase (ALAT), aspartate aminotransferase (ASAT) and C-reactive protein (CRP)], kidney function profile (urine ketones, urea, creatinine, and uric acid), patient’s medical history and initial/current medications.

Clinical Screening Strategy
Initially, the patients exhibiting (i) clinical symptoms of Maternally Inherited Diabetes Deafness syndrome (MIDD; OMIM # 520000), (ii) T1D confirmed by positive test for diabetes autoantibodies, or (iii) insulin resistance features (obesity, dyslipidemia, acanthosis nigricans, polycystic ovary syndrome...) were excluded.

The families included in this study fulfilled the following clinical MODY diagnostic criteria (1): a familial transmission of diabetes (two or more affected generations); (2) early-onset hyperglycemia (one or more members diagnosed under the age of 30); (3) absence of beta-cell antibodies; (4) personal history of unexplained beta-cell dysfunction (early repeated episodes of hypoglycemia or hyperglycemia); (5) non-insulin dependence (lack or low daily requirements of antidiabetic drugs for at least three years after the diagnosis); and (6) absence of obesity at diagnosis. Finally, a total of seventeen unrelated Tunisian pedigrees were recruited. Four families provided complete medical records with available family members for co-segregation analysis. The detailed clinical data of the probands from these four families are shown in Table 1.

A group of 181 people without clinical evidence of insulin resistance, abnormal glucose homeostasis, and/or autoimmune endocrine diseases served as a control in this study.

Genetic Screening Strategy
Genomic DNA was extracted from the peripheral blood leukocytes using the phenol-chloroform-based protocol (24). The DNA assessment was performed through NanoDrop 2000 (ThermoFisher Scientific) and Qubit® 2.0 fluorometer (Invitrogen, CA).

In the 17 unrelated participants, the entire exonic and consensus splice site regions of both GCK and HNF1A genes were screened for mutations by direct sequencing. Specific primers were designed using the Primer 3 program (25) and Refseq genes sequences. Primers and protocols of Polymerase Chain Reaction (PCR) are available on request.

Then, clinical exome enrichment was carried out using the TruSight One sequencing panel (Illumina, Inc. San Diego, CA, USA) targeting exonic regions of 4,813 genes associated with known clinical phenotypes. Only three patients with available family members’ medical records and DNA samples were investigated. All procedures were performed according to the manufacturer’s sample preparation protocol (Catalog No. FC-141-1007). Quantification and validation of the enriched libraries were performed using the Qubit dsDNA HS assay kit (Invitrogen) and 2100 Bioanalyzer Instruments (Agilent Technologies, Santa Clara, CA, USA). Generated libraries were paired-end sequenced (2 x 151 base pair) on a MiSeq instrument (Illumina) using the MiSeq Reagent kit V3 (300 cycles), according to the standard guidelines for preparing and loading samples on MiSeq.

Bioinformatics Data Analysis
Default parameters from Illumina’s MiSeq Reporter (MSR) v2.6.2.3 software were used to analyze the data of Illumina’s clinical sequencing assay. After demultiplexing and FASTQ files generation, the Burrows–Wheeler Aligner (BWA) enrichment packages (BaseSpace, Illumina) were used to read and map the raw sequence data against the GRCh37 (hg19) human reference assembly, while the Genome Analysis Toolkit (GATK) software (MSR, BaseSpace, Illumina) was used for variant calling. The sequencing quality parameters obtained from the MiSeq sequencing platform were noted as 140 K/mm² cluster density and 92.90% of clusters passing filter. In addition, 91.4% of reads showed a desirable quality score, at or above Q30. Data of all reads and variants were visually inspected through the Integrative Genomics Viewer tool (version 2.3) (26, 27), and almost all the genes had exons with enough coverage.

The bioinformatic analysis of the resulting variant call formats (VCFs) files was performed in the Illumina VariantStudio software (v3.0). This software provides access to specific tools and several resources including COSMIC (28), HGMD (29), ClinVar (30), and OMIM (31) that facilitate annotations at variant, gene, and transcript levels. The data of variants were gathered from different projects and databases including Single Nucleotide Polymorphism Database (dbSNP) (32), 1,000 genomes project (33), gnomAD database (V2.1.1) (34), and Exome Sequencing Project (ESP) (35). Several in silico prediction tools including SIFT (36), Polyphen-2 (37), and PROVEAN (38) were used to evaluate the effect of all missense variants reported in this study. Synonymous and splice site variants were evaluated in silico using splice site-altering scoring algorithm [Human Splice Finder (HSF) v3.1 (39)]. The Ensembl Variant Effect Predictor (VEP) (40) and VarSome database (41) were also used to access multiple databases, prediction tools, and publications at a single website. The guidelines of the American College of Medical Genetics and Genomics and the Association of Molecular Pathologists (ACMG/AMP) were applied to the newly identified variants (42).

Filtering Strategy
Initially, heterozygous variants identified by Sanger sequencing were analyzed according to their impact’s severity (missense, nonsense, splice sites, insertion–deletion (indel), in frame and frameshift) and their frequencies [minor allele frequency (MAF) below 1%] in multiple public databases. Two additional parameters were considered in our study in order to analyze
| Patient ID | Sex/Current age, years | Age at diagnosis, years | BMI, kg/m² | Fasting plasma glucose, mmol/L | HbA1c values (at diagnosis–last follow up), % | Treatment | Follow-up Complications | Probability of MODY, % | Gene (RefSeq) | Variants | Genotype | dbSNP ID | MAF in GnomAD | Pathogenicity according to ACMG, SIFT, Polyphen-2, Proveane, HSF | References |
|------------|------------------------|-------------------------|------------|--------------------------------|-----------------------------------------------|----------|------------------------|----------------------|--------------|-----------|----------|----------|-----------|--------------------------|-------------|
| F1-IV.2†   | M/8.3                  | 4.3                     | 15.1       | 6.61                           | 6.2–6.5                                      | OHA for two years then shifted to diet. Metformin | No                      | 75.5                 | GCK        | NM_000162.5 | c.571C>T, p.Arg191Trp | CT         | rs1085307455 | 7.96e−06 Pathogenic c,d | ClinVar, VarSome, LitVar‡ |
| F2-III.11  | F/63                   | 40                      | n.a.       | n.a.                           | n.a.–9                                       | No                                  | n.a.                  | GCK        | NM_000162.5 | c.774C>T, p.(Gly258Gly) | Exon 2:    | c.185A>G, p.Gln62Arg | CT         | rs78006456 | 1.77e−05 Uncertain significance b, affect splicing f | Present study |
|            |                        |                         |            |                                |                                              |                                     |                       |                       | KNLF11     | NM_003597.5 | c.185A>G, p.Gln62Arg | Exon 1:    | c.293C>T, p.Ala98Val | CT         | rs1800574  | 2.82e−02 Benign c,d | ClinVar, VarSome, LitVar‡ |
| F2-IV.16   | F/40                   | 33                      | n.a.       | n.a.–8                         | No                                           | Metformin                          | No                      | n.a.                 | GCK        | NM_000162.5 | c.774C>T, p.(Gly258Gly) | Exon 2:    | c.185A>G, p.Gln62Arg | TT         | rs78006456 | 1.77e−05 Uncertain significance b, affect splicing f | Present study |
|           |                        |                         |            |                                |                                              |                                     |                       |                       | KNLF11     | NM_003597.5 | c.774C>T, p.(Gly258Gly) | Exon 2:    | c.185A>G, p.Gln62Arg | TT         | rs35927125 | 9.13e−02 Benign c,d | ClinicVar, LitVar† |
| F3-II.10   | M/81                   | 40                      | n.a.       | n.a.–8.5                       | No                                           | Metformin and glimepiride           | No                      | n.a.                 | HNF1A      | NM_000545.8 | c.293C>T, p.Ala98Val | Exon 1:    | c.293C>T, p.Ala98Val | CT         | rs1800574  | 2.82e−02 Benign c,d | ClinicVar, VarSome, LitVar‡ |
| F3-III.1†  | M/30                   | 22                      | 25.59      | 8.5                            | 8–9                                          | Metformin and glimepiride           | No                      | 75.5                 | GCK        | NM_000162.5 | c.774C>T, p.(Gly258Gly) | Exon 2:    | c.185A>G, p.Gln62Arg | TT         | rs78006456 | 1.77e−05 Uncertain significance b, affect splicing f | Present study |
| F4-III.2   | F/65                   | 38                      | n.a.       | n.a.–9                         | No                                           | Metformin                          | No                      | 75.5                 | ABCD8      | NM_001287174.2 | c.293C>T, p.Ala98Val | Exon 25:   | c.293C>T, p.Ala98Val | GA         | rs201499958 | 3.59e−05 Tolerated c,d, probably damaging b, damaging a | Present study |
| F4-IV.2†   | F/37                   | 24                      | 19.5       | 8.5–11                         | No                                           | Metformin                          | No                      | 75.5                 | GCK        | NM_000162.5 | c.774C>T, p.(Gly258Gly) | Exon 2:    | c.185A>G, p.Gln62Arg | TT         | rs78006456 | 1.77e−05 Uncertain significance b, affect splicing f | Present study |
| F4-IV.8†   | M/33                   | 19                      | 19.5       | 23                              | Ketosis                                      | No Diabetic nephropathy and retinopathy | No                      | 58                   | GCK        | NM_000162.5 | c.774C>T, p.(Gly258Gly) | Exon 2:    | c.185A>G, p.Gln62Arg | AA         |                      |                      |                      |                      |                      |                      |                      |

BMI, Body Mass Index; HbA1c, glycated hemoglobin; n.a., Not available; OHA, oral hypoglycemic agents. *BMI and fasting plasma glucose values at first admission with/without the acute metabolic disorder. The probability of MODY was calculated only for the young-onset diabetes patients using MODY probability calculator. The prediction of the deleterious effect of variants using different tools. The estimated minor allele frequency for HNF1A p.A98V variant was around 3%; while the GCK p.(Gly258Gly) variant was absent in 50 healthy subjects. The restriction enzymes HaeIII and HindIII were used to digest the PCR products covering the rs1800574 and rs780806456, respectively. †Index case. ‡Variant found in LitVar (23).
the totality of MODY genes, including (i) the uncommon synonymous variants predicted to affect the mRNA splicing but not previously described in the HGMD Professional, and (ii) the autosomal recessive mode of inheritance (homozygous and compound heterozygous variants).

Subsequently, we established a list of 133 genes previously known to be involved in MODY transmission (MODY1-13 genetic subtypes), and/or monogenic (isolated or syndromic) diabetic forms or associated with T2D and glucose metabolism (Table S1). We particularly focused on variants previously associated with T2D or detected in the 133 genes. Finally, the retained variants within all enriched genes were analyzed according to their impact’s severity and frequencies (MAF below 1%) in GnomAD.

**Sanger Sequencing**

Sanger sequencing of available family members was carried out to confirm the familial co-segregation of retained variants. Primer sequences and amplification protocols are available on request. PCR products were purified using the PureLink Quick Gel Extraction Kit (Thermo Fisher Scientific) and then sequenced on an ABI 3100-4 automated sequencer (Applied Biosystems Inc, Foster City, CA, USA) according to the manufacturer’s instructions. Data of Sanger sequencing were analyzed using Bioedit software v7.1.3.

**RESULTS AND DISCUSSION**

Our genomic analysis included seventeen clinically diagnosed MODY patients. Their age of diagnosis ranged from 4 to 30 years. Most patients had a normal BMI at diagnosis; except one proband found to be overweight (Table 1). Blood glucose control was not ideal for some patients as the median HbA1c was 9.7% (6.2–13.2%). Diabetes treatment at diagnosis was heterogeneous. Two participants (2/17; 12%) were controlled with diet, fourteen (14/17; 82%) were treated with oral hypoglycemic agents (OHA) and 6% (1/17) with insulin. During the follow-up, we noticed that the treatment of only one patient, among the 14 participants previously treated with OHA, switched to diet. Additionally, we changed the treatment of the two patients initially controlled with diet into a combination of insulin and oral agents (Table 1). The long-term micro and macro-vascular complications were rarely found (11% of the probands).

The patients’ collected data were used to calculate the probability of MODY based on an online formula available at www.diabetesgenes.org (8). It is noteworthy that the probability of MODY occurrence increases among negative pancreatic autoantibodies patients. The mean probability of MODY was greater than 62% in our patients with young-onset diabetes (Table 1).

**Screening of GCK Gene and Clinical Phenotype Analysis**

We screened the promoter region, each of the ten exons and their flanking intron sequences specific for the pancreatic isoform of the GCK gene (NM_000162.5) in seventeen unrelated index cases. We identified two genetic variants (pathogenic and of uncertain significance) in two unrelated index cases. Table 1 shows the clinical, laboratory and genetic characteristics of patients with GCK gene variants.

The first substitution was the c.571C>T mutation (exon 5), and leads to the replacement of arginine to tryptophan in codon 191 (p.Arg191Trp; rs1085307455; Figure 2B). This mutation was observed in gnomAD with a MAF of 7.96e-26, but it remained absent in the large population cohorts (NHLBI-ESP and the 1000 Genomes Project). Bioinformatics analysis predicted that the GCK p.Arg191Trp mutation could affect the protein structure and function (43). Other computational studies have shown that the Arg191 residue contributes to the transformation of the enzyme’s spatial conformation (43, 44). Nevertheless, in vivo analysis have revealed a decreased thermostability of p.Arg191Trp mutant protein that ultimately leads to full kinetic inactivation of glucokinase (45).

The 191Trp residue was primarily found in Caucasian gestational diabetic subjects who did not meet MODY diagnostic criteria (46). Indeed, the c.571C>T mutation has been previously identified in individuals (pregnant women, children and adolescent) with MODY2 phenotype of British (46), Italian (47), Korean (48), Belgian (49), Norwegian (50), French (51), Japanese (52), Brazilian (53), Chinese (45) and Tunisian (20) descent. HGMD Professional has also reported three other amino acid substitutions on the same residue (p.Arg191Leu/Gln/Glu) related to MODY2 (47, 54, 55).

We identified c.571C>T mutation (p.Arg191Trp) in the F1-IV.2 proband. This proband was referred to the pediatric endocrinology clinic for mild fasting hyperglycemia of 6.81 mmol/L. The fasting blood glucose test was done as the proband exhibited persistent diabetes-specific symptoms (polyuria and polydipsia) at the age of 4.3. At admission, his weight, height and BMI were recorded as 17 kg (~0.5 SDS), 106 cm (~1 SDS), and 15.1 kg/m², respectively. His HbA1c was mildly elevated (6.2%) (Table 1) whereas the value of fasting C-peptide level was lower (0.07 pmol/ml) than the reference range (RR: 0.37–1.47). A slight increase in the glucose level (5.76 to 8.78 mmol/L) during a 2-h OGTT confirmed the impaired glucose tolerance diagnosis. The biochemical and metabolic parameters were found to be within the physiological limits without clinical characteristics of insulin resistance. The strong paternal family history of T2D is displayed in Figure 2A. The proband’s father (F1-III.6) was diagnosed with T2D at the age of 30. His fasting blood glucose measurements have always been below 8.5 mmol/L and he is currently on OHA therapy. The proband’s paternal aunt (F1-III.7) was diagnosed with T2D at the age of 40 and started OHA as well but she is currently on insulin therapy. However, cascade genetic testing confirmed the absence of this mutation in the proband’s mother (F1-III.5; Figure 2B). The proband’s father refused biochemical assessment, genetic testing or counseling. Genetic testing was useful in this case as it improves the clinical course and guided the management of our proband. The OHA therapy of this boy was immediately ceased (Table 1). Lifestyle counselling was...
provided for healthy eating and physical activity that stabilized his health. He was examined every 6 months at the outpatient clinic, and his HbA1c always remained stable at 6.5% (Table 1).

The clinical phenotype of our MODY2 subject (F1-IV.2) was similar to that described in previous reports (56). We recommended fasting blood glucose measurement, 2-h OGTT, and genetic testing of this mutation for each at-risk or affected family members. Hyperglycemia in F1-III.6 and F1-III.7 probands must be pharmacologically managed according to their genotypes. Furthermore, the c.571C>T mutation has already been reported in another unrelated Tunisian pedigree (20). The main reason for this mutation has never been explained whether it is due to the gene founder effect or mutational hotspot in Tunisia. Indeed, the GCK small domain has been specifically considered a mutation hotspot of Southern Italy (57). Of note, it has been observed that some subjects suffering from GCK-MODY were treated with low daily doses of OHA and/or insulin (50, 56). Therefore, the identification of this mutation among a large cohort of Tunisian patients with mild non-progressive early-onset hyperglycemia or gestational diabetes can facilitate early diagnosis of this disease and allow for a personalized treatment. It is noteworthy that according to previous studies, the treatment for GCK-MODY outside of pregnancy is not required (58, 59).

We also found, in the F2-IV.12 proband, a novel c.774C>T variant at exon 7 of the same gene resulting in an exonic silent substitution [p.(Gly258Gly); rs780806456; Figure 3B]. This variant has been reported in gnomAD (MAF = 1.77e−05), but not observed in healthy control subjects (n = 50). It was classified according to ACMG/AMP as a variant of uncertain significance (Table 1). Indeed, no data are available about its co-inheritance or association with any medical condition (mainly disorders of glucose metabolism).

The F2-IV.12 proband came to the emergency service, at 27-year-old, suffering from acute hyperglycemia (29.5 mmol/L). During clinical examination, his height, weight, BMI and blood...
pressure were recorded as 178 cm, 55 kg, 17.35 kg/m², and 110/70 mmHg, respectively. Initial HbA1c was elevated to 13.2% with an acute metabolic complication of diabetes (ketoacidosis). Mild osmotic symptoms (polyuria, polydipsia) were noticed at the age of 16 but there was no medical history of permanent neonatal diabetes. The uncontrolled hyperglycemia and recurrent infections might have caused the ketoacidosis condition. Biochemical analysis revealed 1.02 mmol/L of triglycerides (RR: 1.5–2.8), 1.84 mmol/L of total cholesterol (RR: 4–6), 0.92 mmol/L of HDL, 0.46 mmol/L of LDL, 55 U/L of ASAT (RR: 10–40), 31 U/L (RR: 10–50) of ALAT, 81.80 mg/L of CRP (RR: <6), 384 µmol/L of creatinine (RR: 80–115), 581 µmol/L of uric acid (RR: 220–420), and 10.76 mg/day of microalbuminuria. The thyroid function tests were normal. Ultrasonography revealed the normal shape of kidneys, pancreas, and liver. Initially, continuous insulin therapy was prescribed, to normalize the blood glucose, followed by a gradual dose reduction. The HbA1c of this proband decreased to 5.2% at the 6-month follow-up and remained stable for 24 months. His renal function was also stable during this period. Biochemical analysis revealed triglycerides of 0.44 mmol/L, total cholesterol of 2.62 mmol/L, HDL of 1.22 mmol/L, LDL of 1.2 mmol/L, and creatinine of 67.47 µmol/L. After examining these results, the treatment of proband was switched from very low-dose insulin (0.2 units (IU)/kg/day) to OHA for a few months. Indeed, the fasting blood glucose control was not satisfactory as the HbA1c
values were noted above 9%. Currently, the F2-IV.12 proband receives insulin at a dose of 0.3 IU/Kg/day (Table 1). During the last medical checkup, his HbA1c was noted as 12.2% because he failed to properly follow the hypoglycemic regimen. Diabetic retinopathy or retinitis pigmentosa was not observed during the fundus examination. Three siblings of the mother (F2-III.4, F2-III.6, F2-III.9) and their offspring (F2-IV.5, F2-IV.7) suffered from T2D as presented in Figure 3A. All the affected family members are on OHA therapy since the diagnosis of T2D and there are no micro- and macro-vascular diabetic complications in all affected proband’s family.

Cascade genetic testing revealed rs780806456 in F2-IV.12 and F2-IV.16 patients in the homozygous state (Figure 3B). The cosegregation analysis of rs780806456 showed that there is no correlation with diabetes in affected family members whose DNA was available for genetic testing (Figure 3B). Indeed, this variant was inherited from consanguineous parents (F2-III.11 and F2-III.12) who were heterozygous carriers and was absent in the maternal aunt of the proband (F2-III.4). Therefore, we could not confirm whether this variant is responsible for MODY.

The c.774C>T variant was predicted in silico to affect the mRNA splicing process through the creation of a new cryptic donor splice site or an exonic splicing silencer (ESS) site. Therefore, further functional investigation is required to evaluate its role in the phenotype of carrier patients. A recent in vitro study has supported the pathogenic effect of novel or rare synonymous GCK gene variants on splicing (60).

The clinical presentation of the F2-IV.12 proband did not meet classic MODY2 diagnostic criteria. Generally, patients suffering from GCK-MODY are asymptomatic with mild non-progressive hyperglycemia that is incidentally diagnosed (4). Their slightly higher HbA1c levels typically range from 5.6 to 7.3% at ≤40 years, and 5.9 to 7.6% at >40 years (61). GCK-MODY patients are usually not prone to long-term micro- or macro-vascular diabetic complications and are well responsive to diet without any pharmacological intervention (9, 62).

Screening of HNF1A Gene and Clinical Phenotype Analysis

HNF1A gene mutation analysis was also conducted in all index cases. The clinical, laboratory, and genetic characteristics of the patients with HNF1A relevant variants, are summarized in Table 1.

The heterozygous NM_000545.8: c.293C>T variant causes missense change in the protein from alanine to valine in residue 98 (p.Ala98Val; rs1800574). This variant co-segregates with diabetes in affected members (F3-II.10 and F3-III.1; Figure 4B). The c.293C>T variant is localized in the DNA-binding domain. It was first reported to be associated with a decrease in serum C-peptide and insulin responses to an oral glucose challenge (63). Then, several reports have demonstrated its mild impact on HNF1α function (64). The in silico analysis based on SIFT (score = 0.09) and Polyphen-2 (score = 0.229) predicted that this substitution is benign. We considered p.Ala98Val as a non-pathogenic variant due to its high frequency in the general population (MAF = 0.0282 according to GnomAD) and Tunisian control group (n = 181 subjects; MAF = 0.03; Table 1). rs1800574 frequency in diabetic patients and control groups has been widely studied in different Asian and European populations (65–67). Locke et al. (68) have reported that p.Ala98Val is not associated with the age during the diabetes diagnosis of HNF1A-MODY individuals. Previous studies have reported that there is no relationship between this variant and gestational diabetes in Danish (69), Polish (70), and Turkish (71) women. The role of HNF1A p.Ala98Val variant in early and late-onset of diabetes and MODY 3 differs between populations (67, 72, 73).

The c.293C>T variant was identified in a 22-year-old man (F3-III.1) with a 12-month history of polydipsia, polyuria and had lost 6 kg weight. At admission, the patient’s BMI was recorded as 25.59 kg/m² (82 kg weight and 179 cm height) and physical examination depicted normal vital signs. Laboratory investigations revealed 8.5 mmol/L fasting blood glucose and 8% HbA1c value (Table 1) with no acute metabolic disorder (no ketosis). Biochemical analysis showed normal levels of triglycerides, total cholesterol, HDL, LDL, ASAT, ALAT, CRP, and creatinine. He received OHA (metformin and glimepiride) for eight years. During the last medical checkups, his plasma glucose level was not satisfactory (HbA1c around 9%) revealing that he was not adhering to the hypoglycemic regimen. Diabetic retinopathy or retinitis pigmentosa were not noticed in fundus examination. The strong paternal family history of T2D is displayed in Figure 4A. All affected family members had been on diet control followed by, or not, OHA therapy since the diagnosis of T2D. Micro- and macro-vascular diabetic complications were not observed in the affected proband’s family. They have no clinical evidence of insulin resistance syndrome, cardiovascular or renal disorders. The proband’s father (F3-II.10) was diagnosed with diabetes at the age of 40 and treated with OHA for 21 years (Table 1). The proband’s mother (F3-II.9) and siblings (F3-III.2; F3-III.3) had normal blood glucose responses two hours after the administration of 75-g of glucose solution, and normal HbA1c levels (Figure 4A). The proband’s siblings were negative for all diabetes autoantibodies. The MODY phenotype was suspected for this family based on the clinical features such as the early onset of diabetes, and management of diabetes with long-term OHA therapy.

Clinical expression of MODY3 considerably varies depending upon the genetic predisposition (HNF1A gene SNPs) and environmental factors (physical activity and diet) (74, 75). Therefore, some MODY3 patients may not meet all classical diagnostic criteria. Most of the patients are very sensitive to low doses of sulfonylureas (4) which is usually effective in children and young adults for decades, until the production of beta-cell insulin is severely decreased (64). MODY3 patients are prone to cardiovascular and microvascular diabetes complications (64, 76).

The F3 family did not fulfill the classical diagnostic criteria of MODY3, and therefore it cannot be concluded that the p.Ala98Val variant is responsible for the described clinical phenotype.

Clinical Exome Sequencing and Variants Detection

By screening GCK and HNF1A genes, only one family (family F1) was classified as MODY2. Subsequently, clinical exome analysis was performed on three unrelated index cases (F2-IV.12, F3-III.1, F3-III.3).
F4-IV.8) selected based on the available family members’ medical records and DNA samples. Therefore, two genetic variants have been identified in two unrelated patients.

The known variant c.185A>G (p.Gln62Arg; rs35927125) was detected in the Kruppel-like factor 11 (KLF11) gene (NM_003597.5). This variant was identified in the F2-IV.12 index case, his affected mother, normoglycemic father (with heterozygous genotype AG), affected sister, and normoglycemic brother (with homozgygous genotype GG) (Figure 3C). rs35927125 was already present in public SNP databases as a common missense variant with a 9.13e−02 frequency of the G allele according to gnomAD. It was predicted as benign by multiple bioinformatics tools. However, Neve et al. (77) showed that the p.Gln62Arg variant impairs the activation of insulin promoter and reduces insulin expression levels in pancreatic beta cells. They also demonstrated a decrease in plasma insulin after an oral glucose challenge in the subjects carrying the 62Arg allele. rs35927125 was found to be significantly associated with T2D in North European populations (77). However, several association studies have reported contrasting results. Three studies of 8,676 Caucasian Northern-European individuals (78), 1,818 Japanese participants (79), and 1,337 Pima Indians (80) including case-control and/or family-based samples failed to replicate the association between KLF11 p.Gln62Arg with T2D or other insulin-related quantitative traits (78). Previous studies disputed the KLF11 gene association with MODY making the genetic diagnosis more challenging (12, 81).

For F4-IV.8 proband, clinical exome sequencing allowed the identification of a novel homozygous missense variant (c.2978G>A) in ABCC8 gene (NM_001287174.2) at exon 25. This substitution leads to the replacement of arginine to histidine [p.(Arg993His)] (Table 1). The elder sister (F4-IV.2) was diagnosed with T2D at the age of 24 and her BMI was 19 kg/m² (Table 1). She is currently being treated with insulin (0.7 IU/kg/day) and metformin. The proband refused to use the insulin, that worsened the glycemic control and led to early diabetic microvascular complications (diabetic nephropathy and retinopathy) at the age of 25. During the last medical checkup, an increased in HbA1c (13%) was noted because of irregular insulin usage. The laboratory examination also showed abnormal hepatic and renal functions. This patient was not followed up after the age of 30. His mother (F4-III.2) was diagnosed with T2D at the age of 38. Metformin was prescribed to manage plasma glucose levels and an HbA1c of 9% was noted at the last follow-up (Table 1). The elder sister (F4-IV.2) was diagnosed with T2D at the age of 24 and her BMI was 19 kg/m² (Table 1). She suffered from diabetic nephropathy and retinopathy at the age of 32. His 35 years old sister (F4-IV.3) was diagnosed with gestational diabetes at the age of 27 and insulin therapy was recommended (Figure 5B). Unfortunately, her laboratory and clinical follow-up were not available. Interestingly, diabetes autoantibodies were negative for both sisters of the proband.

Cascade genetic screening identified the same mutation in the proband’s mother in heterozygote state, but the sister (F4-IV.2) and her children (F4-V.1, F4-V.2) were negative (Figure 5B). The ABCC8 p.(Arg993His) missense variant was either de novo or inherited from the father. Unfortunately, proband’s father (F4-III.3) and affected sister (F4-IV.3) were not available for genetic testing. Further investigation needs to be conducted.
about the role of \textit{ABCC8} p.(Arg993His) variant in hyperglycemia of this family. It has been reported that heterozygous carriers of \textit{ABCC8} p.L171F mutation developed adult-onset diabetes, while homozygous carrier developed hyperinsulinemic hypoglycemia at neonatal period and diabetes later in life (83, 84).

This study revealed the absence of previously described or novel pathogenic variants within all enriched genes. This study can be further expanded in the future by (i) recruiting more MODY families to estimate MODY2 prevalence in Tunisia, (ii) conducting functional characterization of variants of uncertain significance to validate their effects on hyperglycemia, and (iii) complete testing of other affected/unaffected family members to track variants co-segregation in each family.

The clinical inclusion criteria for patients in this study were rigorous compared to other Tunisian studies (Figure 1). In recent years, the recruitment criteria of MODY patients have been modified than originally defined by Tattersall and Fajans (85). Non-traditional MODY characteristics including the age of onset, obesity, the presence of diabetes autoantibodies and ketonuria, and treatment modalities have been evaluated (10). This study included patients with decompensated diabetes at study inclusion. Indeed, the absence of ketonuria was believed to be a definite criterion to differentiate T1D from MODY patients. However, the presence of ketonuria has been reported among patients with \textit{HNF1A} (86, 87), \textit{PDX1} (88), \textit{NEUROD1} (89), and \textit{INS} (90) mutations. The patients involved in this study presented a less severe metabolic profile (BMI <26 kg/m²). But, we exclude unintentionally carriers of pathogenic/likely pathogenic variants within the most common MODY genes (\textit{HNF4A} and \textit{HNF1A}) (91, 92). Recently, Vaxillaire et al. (21) analyzed a cohort of Mediterranean patients suspected with monogenic diabetes and observed that carriers of pathogenic variants had the lowest BMI. This is not the first study in Tunisia that describes the genetic etiology of MODY/monogenic diabetes in patients with early-

![FIGURE 5](image-url)
onset diabetes. Overall, 89 unrelated MODY suspected Tunisian participants included in this study have been investigated using targeted gene panel sequencing, direct/indirect sequencing, or MLPA testing. Only eleven heterozygous pathogenic variants were found in sixteen unrelated patients (~18%) (16, 18, 20, 21). The results of this study support previous observations of Vaxillaire et al. (21) such as (i) the severity of the clinical presentation of non-European cases (including those of Tunisian ancestry) with early-onset diabetes (similar to F2-IV.12 and F4-IV.8 index cases), and (ii) low mutation rates within suspected MODY patients in Tunisia. Only one patient from the 17 studied index case (5.9%) presented a pathogenic variant; and showed improvement in his long-term monitoring.

Moreover, our findings emphasized the importance of exploring the role of variants of uncertain significance in glucose metabolism. Our study proved the efficiency of the whole exome sequencing for MODY diagnosis since more than one major gene might be involved in the emergence of MODY among the Tunisian population.

**CONCLUSIONS**

In this study, we report detailed genetic investigation results within suspected MODY patients in Tunisia. Only one patient from the 17 studied index case (5.9%) presented a pathogenic variant; and showed improvement in his long-term monitoring. Moreover, our findings emphasized the importance of exploring the role of variants of uncertain significance in glucose metabolism. Our study proved the efficiency of the whole exome sequencing for MODY diagnosis since more than one major gene might be involved in the emergence of MODY among the Tunisian population.

**DATA AVAILABILITY STATEMENT**

The datasets used and/or analyzed in the present study are available from the corresponding author on reasonable request.

**REFERENCES**

1. Urakami T. Maturity-Onset Diabetes of the Young (MODY): Current Perspectives on Diagnosis and Treatment. *Diabetes Metab Syndr Obes* (2019) 12:1047–56. doi: 10.2147/DMSO.S179793

2. Nkonge KM, Nkonge DK, Nkonge TN. The Epidemiology, Molecular Pathogenesis, Diagnosis, and Treatment of Maturity-Onset Diabetes of the Young (MODY). *Clin Diabetes Endocrinol* (2020) 6(1):20. doi: 10.1186/s40842-020-00112-5

3. Vaxillaire M, Froguet P, Bonnefond A. How Recent Advances in Genomics Improve Precision Diagnosis and Personalized Care of Maturity-Onset Diabetes of the Young. *Curr Diabetes Rep* (2019) 19(9):79. doi: 10.1007/s11892-019-1202-x

4. Peixoto-Barbosa R, Reis AF, Giuffrida FMA. Update on Clinical Screening of Maturity-Onset Diabetes of the Young (MODY). *Diabetol Metab Syndr* (2020) 12:50. doi: 10.1186/s13098-020-00557-9

5. Patel KA, Kettunen J, Laakso M, Stančáková A, Laver TW, Colclough K, et al. Heterozygous RFX6 Protein Truncating Variants are Associated With MODY With Reduced Penetrance. *Nat Commun* (2017) 8(1):888. doi: 10.1038/s41467-017-08959-9

6. Mohan V, Radha V, Nguyen TT, Stavisski EW, Pahuja KB, Goldstein LD, et al. Comprehensive Genomic Analysis Identifies Pathogenic Variants in Maturity-Onset Diabetes of the Young (MODY) Patients in South India. *BMC Med Genet* (2018) 19(1):22. doi: 10.1186/s12881-018-0528-6

7. Li J, Sun S, Wang X, Li Y, Zhu H, Zhang H, et al. A Missense Mutation in IRS1 is Associated With the Development of Early-Onset Type 2 Diabetes. *Int J Endocrinol* (2020) 2020:9569126. doi: 10.1155/2020/9569126

8. Shields BM, McDonald TJ, Ellard S, Campbell MJ, Hyde C, Hattersley AT. The Development and Validation of a Clinical Prediction Model to Determine the Probability of MODY in Patients With Young-Onset Diabetes. *Diabetologia* (2012) 55(5):1265–72. doi: 10.1007/s00125-011-2418-8

9. Vaxillaire M, Froguet P. Monogenic Diabetes: Implementation of Translational Genomic Research Towards Precision Medicine. *J Diabetes* (2016) 8(6):782–95. doi: 10.1111/1753-0407.12446

10. Delvecchio M, Pastore C, Giordano P. Treatment Options for MODY Patients: A Systematic Review of Literature. *Diabetes Ther* (2020) 11(8):1667–85. doi: 10.1007/s13300-020-00864-4

11. Thanabalasingham G, Pal A, Selwood MP, Dudley C, Fisher K, Bingley PJ, et al. Systematic Assessment of Etiology in Adults With a Clinical Diagnosis of Young-Onset Type 2 Diabetes is a Successful Strategy for Identifying Maturity-Onset Diabetes of the Young. *Diabetes Care* (2012) 35(6):1206–12. doi: 10.2337/dc11-1243

12. Hattersley AT, Gleeley SAW, Polak M, Rubio-Cabezas O, Njelstad PR, Mlynarski W, et al. ISPAD Clinical Practice Consensus Guidelines 2018: The Diagnosis and Management of Monogenic Diabetes in Children and Adolescents. *Pediatr Diabetes* (2018) 19(Suppl 27):47–63. doi: 10.1111/pedi.12772

**ETHICS STATEMENT**

The studies involving human participants were reviewed and approved by The Regional Committee of the Protection of Persons, Sfax, Tunisia (CPP SUD N°28/2019). Written informed consent to participate in this study was provided by the participants’ legal guardian/next of kin.

**AUTHOR CONTRIBUTIONS**

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

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**SUPPLEMENTARY MATERIAL**

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fendo.2021.684018/full#supplementary-material
null
85. Tattersall RB, Fajans SS. A Difference Between the Inheritance of Classical Juvenile-Onset and Maturity-Onset Type Diabetes of Young People. *Diabetes* (1975) 24(1):44–53. doi: 10.2337/diab.24.1.44

86. Pruhova S, Dusatkova P, Neumann D, Hollay E, Cinek O, Lebl J, et al. Two Cases of Diabetic Ketoadidosis in HNF1A-MODY Linked to Severe Dehydration: Is it Time to Change the Diagnostic Criteria for MODY? *Diabetes Care* (2013) 36(9):2573–4. doi: 10.2337/dc13-0058

87. Egan AM, Cunningham A, Jafar-Mohammadi B, Dunne FP. Diabetic Ketoadidosis in the Setting of HNF1A-Maturity Onset Diabetes of the Young. *BMJ Case Rep* (2015) 2015:1–3. doi: 10.1136/bcr-2014-209163

88. Deng M, Xiao X, Zhou L, Wang T. First Case Report of Maturity-Onset Diabetes of the Young Type 4 Pedigree in a Chinese Family. *Front Endocrinol (Lausanne)* (2019) 10:406. doi: 10.3389/fendo.2019.00406

89. Horikawa Y, Enya M. Genetic Dissection and Clinical Features of MODY6 (NEUROD1-MODY). *Curr Diabetes Rep* (2019) 19(3):12. doi: 10.1007/s11892-019-1130-9

90. Johnson SR, McGown I, Oppermann U, Conwell LS, Harris M, Duncan EL. A Novel INS Mutation in a Family With Maturity-Onset Diabetes of the Young: Variable Insulin Secretion and Putative Mechanisms. *Pediatr Diabetes* (2018) 19(5):905–9. doi: 10.1111/pedi.12679

91. Kleinberger JW, Copeland KC, Gandica RG, Haymond MW, Levitsky LL, Linder B, et al. Monogenic Diabetes in Overweight and Obese Youth Diagnosed With Type 2 Diabetes: The TODAY Clinical Trial. *Genet Med* (2018) 20(6):583–90. doi: 10.1038/gim.2017.150

92. Kyithar MP, Bacon S, Pannu KK, Rizvi SR, Colclough K, Ellard S, et al. Identification of HNF1A-MODY and HNF4A-MODY in Irish Families: Phenotypic Characteristics and Therapeutic Implications. *Diabetes Metab* (2011) 37(6):512–9. doi: 10.1016/j.diabet.2011.04.002

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