Recent Advances in Molecular Research on Hydrogen Sulfide (H$_2$S) Role in Diabetes Mellitus (DM)—A Systematic Review

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Abstract: Abundant experimental data suggest that hydrogen sulfide (H$_2$S) is related to the pathophysiology of Diabetes Mellitus (DM). Multiple molecular mechanisms, including receptors, membrane ion channels, signaling molecules, enzymes, and transcription factors, are known to be responsible for the H$_2$S biological actions; however, H$_2$S is not fully documented as a gaseous signaling molecule interfering with DM and vascular-linked pathology. In recent decades, multiple approaches regarding therapeutic exploitation of H$_2$S have been identified, either based on H$_2$S exogenous apport or on its modulated endogenous biosynthesis. This paper aims to synthesize and systematize, as comprehensively as possible, the recent literature-related data regarding the therapeutic/rehabilitative role of H$_2$S in DM. This review was conducted following the ‘Preferred reporting items for systematics reviews and meta-analyses’ (PRISMA) methodology, interrogating five international medically renowned databases by specific keyword combinations/”syntaxes” used contextually, over the last five years (2017–2021). The respective search/filtered and selection methodology we applied has identified, in the first step, 212 articles. After deploying the next specific quest steps, 51 unique published papers qualified for minute analysis resulted. To these bibliographic resources obtained through the PRISMA methodology, in order to have the best available information coverage, we added 86 papers that were freely found by a direct internet search. Finally, we selected for a connected meta-analysis eight relevant reports that included 1237 human subjects elicited from clinical trial registration platforms. Numerous H$_2$S releasing/stimulating compounds have been produced, some being used in experimental models. However, very few of them were further advanced in clinical studies, indicating that the development of H$_2$S as a therapeutic agent is still at the beginning.

Keywords: hydrogen sulfide (H$_2$S); Diabetes Mellitus (DM); DM vascular-linked pathology; systematic review; oxidative phosphorylation; ROS (Reactive Oxygen Species)

1. Introduction

Diabetes Mellitus (DM) is a non-communicable chronic metabolic disease [1] characterized by prolonged hyperglycemia. Type 1 DM is a chronic condition in which the body’s pancreatic $\beta$ cells, determined by different causes, reduce insulin production. Instead, type

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2 DM is caused mainly by lifestyle factors and is characterized by insulin resistance, thus inefficiency on target cells, also with complex pathophysiologic links. The International Diabetes Federation mentions more than 450 million people globally with DM at present, 90% representing Type 2 DM, and estimates an increase to over 600 million in the next 25 years [2]. Therefore, the diabetic “pandemic” seems to be a global public health problem that needs particular attention, considering the deaths and disabilities caused by this disease’s complications [3].

Severe and more frequently associated complications of DM may primarily involve the nervous system [4], kidneys [5], and/or eyes [6] due to damage of the microcirculation [7], respectively; and the cardiovascular systems [8], due to macroangiopathy [9]. Cardiomyopathy is responsible for more than 50% of deaths in diabetic patients [10]. Diabetic nephropathy [11] generates kidney injuries and tissue lesions and eventually leads to chronic kidney disease.

In DM, patients with insufficient oxygen supplies due to impaired angiogenesis/neovascularization [12], refractory wounds and critical ischemic sufferance in limbs are major vascular hazards [11], connected with a malfunction of endothelial cells (ECs) [13].

\( \text{H}_2\text{S} \), recognized by its smell of rotten eggs, is the simplest thiol (R-SH), a sulfur analog of alcohol, with a high redox potential [14]. However, disentangling \( \text{H}_2\text{S} \) chemistry and biochemistry is much more complicated; \( \text{H}_2\text{S} \) biological actions are dependent on its chemical (reductive and nucleophilic) attributes. \( \text{H}_2\text{S} \) is fast-dissolving in aqueous solutions (due to its fine acid quality), splitting to generate two anionic parts: sulfide (\( S_2^- \)) and hydrosulfide (\( \text{HS}^- \)). About 81.5% of the total \( \text{H}_2\text{S} \) exists as \( \text{HS}^- \) and \( S_2^- \), and only 18.5% as an undissociated acid at a specific physiological potential of hydrogen (pH) of 7.4 in an aqueous solution, and thus has biological functions similar to other ionic species [3]. As a result, \( \text{HS}^- \) is an excellent substrate for the Anion Exchanger 1 (AE1) [15]. The acid \( \text{H}_2\text{S}/\text{HS}^- \) balance follows the same principle as \( \text{CO}_2/\text{HCO}_3^- \) in the Jacobs–Stewart cycle [16]. \( \text{H}_2\text{S} \) is highly lipophilic—a feature that allows it to freely penetrate all cell membranes without the facilitation of membrane channels, thereby exerting biological activities [17]. Additionally, it has a very high permeability coefficient in human erythrocytes. Related to this capability, it was established that in cases characterized by a lack of extracellular hydration and intracellular dehydration, the pH buffer system of the \( \text{Cl}^-/\text{HS}^-/\text{H}_2\text{S} \) is faster than the \( \text{Cl}^-/\text{HCO}_3^-/\text{H}_2\text{CO}_3 \) cycle [18]; however, in severe DM, such a pH buffer, it is much lower than that in healthy persons, and therefore, its protective effect is diminished [19].

\( \text{H}_2\text{S} \) serves as a gasotransmitter in regulating organ development and maintaining homeostasis. Therefore, its abnormal levels are linked with multiple human diseases, such as DM, neurodegenerative diseases, myocardial injuries [20], or ophthalmic pathology [21]. Plasmatic \( \text{H}_2\text{S} \) levels are remarkably lower in diabetic patients [3]. In addition, the levels of \( \text{H}_2\text{S} \) in plasma, urine, and heart tissues are prone to be lower in aging diabetic rodents [22].

The striking lack of uniform data concerning \( \text{H}_2\text{S} \) levels under physiological conditions may contribute to uncertainty about the precise mechanistic roles of \( \text{H}_2\text{S} \) in various physiological processes. Hence, it is essential to improve the specificity of detection and reduce the threshold limits of related techniques, which may be helpful in achieving more accurate information regarding the physiological distribution of \( \text{H}_2\text{S} \) in the blood, cells, and tissues, as well as more consistent knowledge on its typical tasks at an intimate level based on more comprehensive data on its oxidative phosphorylation activities [23] to satisfy cellular energy requirements, and perhaps other biological involvements [3].

2. Materials and Methods

This systematic literature review is based on the PRISMA methodology, by searching free full-text available papers written in English, which have appeared in the last five years, by specific keywords combinations (Table 1), in the well known international databases: National Center for Biotechnology Information (NCBI)/PubMed, PubMed Central (PMC), Elsevier, and Web of Science. Cochrane and PEDro databases returned no results.
The scientific impact of each article was established using a customized quantification formula to obtain a PEDro score. We considered eligible the works that received a score of at least 4 (“fair quality = PEDro score 4–5”).

To evaluate the impact of hydrogen sulfide therapeutic interventions in DM, we searched on https://clinicaltrials.gov, https://trialsearch.who.int/, and https://www.clinicaltrialsregister.eu, (accessed on 1 January 2022), for clinical trials using as items Diabetes Mellitus and H₂S or hydrogen sulfide. The inclusion criteria were fixed regarding patients with Diabetes, age: 18 to the elderly, of all genders. Exclusion criteria correspond to study dates: not before 2017. In addition, a meta-analysis was included to analyze the different pathologies associated with diabetes and the number of subjects.

3. Results

3.1. Search and Filtering Results

Databases interrogation provided, initially, 212 articles. Applying the PRISMA selection filters and scoring resulted in 51 unique published qualified studies. We added another 94 free papers found based on Google Search by a direct internet search (Figure 1), which were highly related to our research (Table 2).

![Flow Diagram](image-url)

**Figure 1.** Our adapted PRISMA-type of the flow diagram.
### Table 2. PRISMA resulting conceptual skeleton structure of the article’s organization approach.

| Physiological Properties of H\textsubscript{2}S | Authors | Ref. No. | Subject-Data |
|-----------------------------------------------|---------|----------|--------------|
|                                               | (Sun, 2021) | [3] | An Updated Insight Into Molecular Mechanism of H\textsubscript{2}S in Cardiomyopathy |
|                                               | (George, 2018) | [6] | Treating inflammation and oxidative stress with H\textsubscript{2}S during age-related macular degeneration |
|                                               | (Zou, 2017) | [10] | H\textsubscript{2}S ameliorates cognitive dysfunction in streptozotocin-induced diabetic rats |
|                                               | (Rey, 2021) | [11] | Mitochondrial metabolism as target of the neuroprotective role of erythropoietin in Parkinson’s disease. |
|                                               | (Testai, 2021) | [12] | Modulation of EndMT by H\textsubscript{2}S in the Prevention of Cardiovascular Fibrosis |
|                                               | (Ciccone, 2021) | [13] | Endothelium as a Source and Target of H\textsubscript{2}S to Improve Its Trophism and Function |
|                                               | (Wu, 2017) | [16] | Exogenous H\textsubscript{2}S facilitating ubiquitin aggregates clearance via autophagy |
|                                               | (Hu, 2017) | [20] | Chelerythrine Attenuates Renal Ischemia/Reperfusion-induced Myocardial Injury |
|                                               | (Kar, 2019) | [22] | H\textsubscript{2}S -mediated regulation of cell death signaling ameliorates adverse cardiac remodeling |
|                                               | (Jeong, 2020) | [24] | Protective effect of H\textsubscript{2}S on oxidative stress-induced neurodegenerative diseases |
|                                               | (Luo, 2019) | [25] | H\textsubscript{2}S upregulates renal AQP-2 protein expression and promotes urine concentration |
|                                               | (Yang, 2019) | [26] | Exogenous H\textsubscript{2}S mitigates myocardial fibrosis through suppression of Wnt pathway |
|                                               | (Liu, 2018) | [27] | H\textsubscript{2}S attenuates myocardial fibrosis through the JAK/STAT signaling pathway |
|                                               | (Sun, 2019) | [28] | Exogenous H\textsubscript{2}S reduces the acetylation levels of mitochondrial respiratory enzymes |
|                                               | (Roa-Coria, 2019) | [29] | Possible involvement of peripheral TRP channels in the H\textsubscript{2}S-induced hyperalgesia |
|                                               | (Yang, 2017) | [30] | Exogenous H\textsubscript{2}S regulates endoplasmic reticulum-mitochondria crosstalk to inhibit apoptosis |
|                                               | (Zhao, 2021) | [31] | H\textsubscript{2}S Plays an Important Role in Diabetic Cardiomyopathy |
|                                               | (Liu, 2017) | [32] | H\textsubscript{2}S modulating mitochondrial morphology to promote mitophagy in endothelial cells |
|                                               | (Qiu, 2018) | [33] | Alpha-lipoic acid regulates the autophagy of vascular smooth muscle cells elevating H\textsubscript{2}S level |
|                                               | (Li, 2017) | [34] | H\textsubscript{2}S reduced renal tissue fibrosis by regulating autophagy in diabetic rats |
|                                               | (Yu, 2020) | [35] | Exogenous H\textsubscript{2}S Induces Hrd1 S-sulfhydration and Prevents CD36 Translocation via VAMP3 |
|                                               | (Kar, 2019) | [36] | H\textsubscript{2}S Ameliorates Homocysteine-Induced Cardiac Remodeling and Dysfunction |
|                                               | (Dominic, 2021) | [37] | Decreased availability of nitric oxide and H\textsubscript{2}S is a hallmark of COVID-19 |
|                                               | (Loiselle, 2020) | [38] | H\textsubscript{2}S and hepatic lipid metabolism-a critical pairing for liver health |
|                                               | (Ma, 2017) | [39] | Exogenous H\textsubscript{2}S Ameliorates Diabetes-Associated Cognitive Decline |
|                                               | (Jiang, 2020) | [40] | H\textsubscript{2}S Ameliorates Lung Ischemia-Reperfusion Injury Through SIRT1 Signaling Pathway |
|                                               | (Wu, 2019) | [41] | H\textsubscript{2}S Inhibits High Glucose-Induced Neuronal Senescence by Improving Autophagic Flux |

| Pathophysiological Properties H\textsubscript{2}S | Authors | Ref. No. | Subject-Data |
|-------------------------------------------------|---------|----------|--------------|
|                                                 | (Citi, 2021) | [7] | Role of H\textsubscript{2}S in endothelial dysfunction: Pathophysiology and therapeutic approaches |
|                                                 | (Kang, 2020) | [14] | H\textsubscript{2}S as a Potential Alternative for the Treatment of Myocardial Fibrosis |
|                                                 | (Sun, 2019) | [42] | H\textsubscript{2}S and Subsequent Liver Injury |
|                                                 | (Szabo, 2017) | [43] | Pharmacological Modulation of H\textsubscript{2}S Levels |
Our search on clinical trial registration platforms showed that despite the vast number of trials (25,969) on DM, only 46 included H$_2$S/hydrogen sulfide. In addition, 12 were duplicates, and 26 were performed/ended before 2017. Therefore, we selected/filtred for our meta-analysis eight relevant reports that included 1237 subjects.

### 3.2. Physiological Properties of H$_2$S

H$_2$S influences many cellular processes (Figure 2) through a broad spectrum of signaling molecules, reacting with superoxide anions, hypochlorite, hydrogen peroxide, peroxynitrite, metals, thiol derivatives, and NO [60]. Moreover, with its aforementioned high rate of the anionic chemical state in aqueous solution, there are reported antioxidant properties of H$_2$S that can mitigate oxidative stress-induced dysfunctions. It also acts through the potassium (KATP/K$^+$) and calcium (Ca$^{2+}$) ion channels to increase (antioxidant) glutathione (GSH) levels. GSH, Gpx (glutathione peroxidase), and superoxide dismutase (SOD) neutralize H$_2$O$_2$-induced oxidative damage in mitochondria. To be specified that ROS (Reactive Oxygen Species) are formed within the oxidative phosphorylation process.
(and in excessive quantities in such a process’ inefficiency, leading to oxidative stress and affecting mitochondrial metabolism) [11], and attenuation of mitochondrial ROS release results in completely preserved insulin sensitivity despite a high-fat diet [24].

H2S influences many cellular processes (Figure 2) through a broad spectrum of signaling molecules, reacting with superoxide anions, hypochlorite, hydrogen peroxide, peroxynitrite, metals, thiol derivatives, and NO [60]. Moreover, with its aforementioned high properties of H2S that can mitigate oxidative stress-induced dysfunctions. It also acts through blue arrows. Finally, the biosynthesis pathways are stated, represented by cystathionine-β-synthase (CBS), cystathionine-γ-lyase (CSE), and 3-mercaptopyruvate sulfurtransferase (MST), the latter connected with cysteine aminotransferase (CAT).

H2S is also endogenously produced, like nitric oxide (NO) and carbon monoxide (CO), which are similar gasotransmitters. H2S has been experimentally shown to be involved in the bio-molecular regulation of vital physiological processes such as the inflammatory response, apoptosis, oxidative stress, and angiogenesis. The brain, liver, kidney, and other organs produce H2S [7]. The cellular biogenesis of H2S is based on the desulfitation of cysteine or homocysteine, a process involving mainly three enzymes: cystathionin-β-synthase (CBS), cystathionine-γ-lyase (CSE), and 3-mercaptopyruvate sulfurtransferase (MST) [61]. H2S biogenesis at the mitochondria level implies cysteine aminotransferase (CAT) that catalyzes L-cysteine and glutamate to 3-mercaptopyruvate and α-ketoglutarate. Furthermore, 3-mercaptopyruvate is metabolized to pyruvate and H2S via 3-mercaptopyruvate sulfurtransferase (3-MST) [24].

MicroRNAs are factors involved in the upregulation of CSE expression. It was also found that some currently used drugs, including angiotensin-converting enzyme (ACE) inhibitors [62], statins [14], calcium channel antagonists, aspirin, and metformin vitamin D3 [42], and many others, may increase the biogenesis of H2S. From this list, statins, for example, can increase H2S synthesis via Akt-mediated control of CSE or suppress H2S degradation by decreasing coenzyme Q level, a sulfide quinone reductase cofactor [57].

Several routes could eliminate the H2S. Firstly, H2S can be transformed into thiosulfate by mitochondrial oxidative modification, or further converted into sulfate and sulfite. Next, cytosolic methylation is another pathway used to transform H2S to dimethylsulfide by
thiol S-methyltransferase. Finally, the excessive H₂S could be scavenged by Metallo- or disulfide-containing molecules or glutathione disulfide and could also be released by the lungs [63].

Exogenously supplied or endogenously generated, H₂S can be stored at the cellular level as bound sulfane, a reductant labile sulfur (e.g., persulfide, polysulfide, and protein-associated sulfur, among others) [14]. Human erythrocytes are about ~5 billion per mL of blood, and each has over 270 million hemoglobin molecules that can uptake H₂S, effectively controlling its clearance. This distribution ensures the maintenance of the physiological plasma and tissue concentration of free H₂S in the range of 15 to 150 nM. In addition, the high lipid and water solubility of H₂S allow quick passage through the alveolar membrane, which assures an equilibrium between blood and the alveolar air level of H₂S [18].

The potential of H₂S metabolite products as biomarkers is appreciated since the plasmatic and urinary levels of H₂S may reflect renal disease severity, such as chronic kidney disease [25]. Therefore, excessive exposure to H₂S can lead to cellular toxicity, orchestrate pathological processes, and increase the risk of various diseases [64]. H₂S is one of the most toxic poisons, and is even more harmful than cyanide on a mole-to-mole basis. A solution of dissolved H₂S diminishes the activity of mitochondrial cytochrome c oxidase at a concentration ranging from 10 to 30 µM. In vivo studies have shown that in rodents and large mammals, severe depression of the medullary respiratory neurons and/or cardiac contractility by infusion or inhaling H₂S at concentrations yield plasma concentrations of gaseous H₂S between 2 and 5 µM.

H₂S can attenuate matrix deposition and myocardial fibrosis [26] and improve MMP/TIMP disorder. The mechanism of H₂S protection against diabetic myocardial fibrosis depends on the down-regulation of JAK/STAT and TGF-β1 (transforming growth factor) signaling [27].

Many physiological and pathophysiological properties regarding antioxidation, apoptosis, or inflammation of H₂S are mediated through transcription factors such as Nrf2 (nuclear factor-E2-related factor), FoxO3 (Forkhead box O), and NF-kB (Nuclear factor kappa-light-chain-enhancer of activated B cells) [22]. The epigenetic role of H₂S is unveiled by Brg1 (Brahma-related gene 1) expression modulation at the promoter region, decreasing the ATP-dependent chromatin remodeling complex’s transcriptional level, which inhibits vascular smooth muscle cell proliferation. Moreover, H₂S may reduce the lysine acetylation of enzymes involved in fatty acid β-oxidation and glucose oxidation in diabetic statuses [28], exerting a beneficial effect on cardiac energy substrate utilization [65].

H₂S is known to regulate various physiological functions, such as decreasing blood pressure, acting on various targets, including ion channels, such as ATP-sensitive potassium channels (KATP) [66], voltage-gated potassium channels (Kv7) [67], transient receptor potential channels (TRPV) [29], or L/T-type Ca²⁺ channels [68], mitoKATP/Kv7 channels [69]. By activating ATP-sensitive K⁺ channels, H₂S lowers blood pressure, protects the heart from ischemia and reperfusion injury, inhibits insulin secretion in pancreatic β cells, and exerts anti-apoptotic, anti-inflammatory, and anti-nociceptive effects [70]. KATP channels also play a crucial role in insulin secretion in pancreatic cells, where the opening of the channels by H₂S decreases insulin secretion. Both endogenous and exogenous H₂S inhibits insulin secretion from cells by activating KATP channels and inhibiting L-type voltage-dependent calcium channels. In addition, by inhibiting glucose transporter-4 (GLUT-4), H₂S inhibits insulin-stimulated glucose uptake in adipocytes, indicating that H₂S decreases the insulin sensitivity of adipocytes [71].

During hyperglycemia, elevated levels of H₂S can open the KATP channels in the islets cell membrane, which can cause high hyperpolarization and lower insulin secretion. This effect is caused by several biochemical processes that inhibit insulin secretion [69].

The endoplasmic reticulum (ER) is the cytoplasmic location where proteins are synthesized. It maintains Ca²⁺ homeostasis and participates in protein folding [72]. The molecular markers of stress include C/EBP homologous protein, cleaved caspase-12, and the glucose-controlled protein 78 (GRP78). It has been observed that chronic ER stress can
trigger DM, Alzheimer’s disease, and other neurodegenerative disorders [73], engaging ER stress-induced apoptosis [10].

Studies on diabetic cardiomyopathy have shown that the effects of H₂S on the endoplasmic reticulum stress are related to its reduction in levels of mitochondria apoptotic proteins [30,31]. The endoplasmic reticulum’s interaction with mitochondria is regulated by the ROS pathway [74]. Mitofusin-2 is a critical protein that can bridge the endoplasmic reticulum and mitochondria. It plays a role in the fusion and fission of mitochondria. It is believed that Mfn-2 is involved in the cardiac system’s mitochondria function and is triggered by oxidative stress [30]. The high levels of Mfn-2 can also induce cardiomyocyte apoptosis [32].

The mitochondria control energy homeostasis and regulate ROS production [75]. Mitochondria play a significant role in the mechanism of fatty acids β-oxidation. On the other hand, mitochondrial dysregulations occur in insulin resistance. The number of mitochondria in hepatocytes decreases in CSE-deficit cells. Hyperglycaemia leads to the generation of mitochondria superoxide, which causes the synthesis of oxidants and endothelial dysfunction. H₂S works as an electron donor to the respiratory chain and plays a therapeutic role in DM and associated vascular diseases.

Mitochondrial DNA (mtDNA) content levels are significantly reduced in CSE-gene knockout mice. This depletion can be reversed by exogenous H₂S gain [76]. H₂S can provoke mtDNA replication and mitochondrial biogenesis by suppressing mitochondrial transcription factor A (TFAM) methylation. In contrast, H₂S may stimulate cardiac mitochondrial biogenesis by activating AMPK (5′ AMP-activated protein kinase) [33] PGC1α (peroxisome proliferator-activated receptor gamma coactivator 1-alpha) pathway [77]. Sulphydrization of AMPK and PP2A (protein phosphatase 2A) [78], which leads to AMPK activation and PP2A inhibition, respectively, has been proposed as a mechanism that may be involved in H₂S-mediated stimulation of unstressed mitochondrial biogenesis [14].

H₂S is a gasotransmitter with discovered roles in cellular signaling, which can also be stored as bound endosulfan, known to play a variety of physiological functions [79]. Some of these include vasodilation [80], anti-apoptosis [81], anti-inflammatory [82], cell survival/death [15], cell proliferation/hypertrophy [83], endoplasmic reticulum stress [84], antioxidative stress [32], mitochondrial bioenergetics/biogenesis [50], blood pressure reduction [85], and cell differentiation [86]. H₂S ameliorates diabetic complications, including endothelial dysfunction [87], nephropathy [34], retinopathy [88], and cardiovascular diseases.

H₂S could also increase the apoptosis of islet cells and inhibit the programmed cell death of pancreatic cells by blocking the ERK (extracellular signal-regulated) protein kinase [33]. It has also been shown to inhibit the anti-inflammatory or antioxidant signaling pathways of pancreatic cells. Injecting H₂S into STZ-induced diabetic rats can improve the status of their diabetes by blocking the PKC/ERK₁₂ signaling pathway. The effects of blocking the JAK/STAT signaling pathway are also linked to the H₂S’ anti-apoptotic effects [58]. The myocardial expressions of pro-fibrotic factors, such as MMP-2 (matrix metalloprotease 2), TIMP-2 (tissue inhibitor of metalloproteinase 2), transforming growth factor (TGF)-β1/SMAD family member 3 (Smad3) signaling pathway, and collagens are strikingly changed in diabetic rats. Many studies have shown that suppressing the STAT3 pathway can improve the physiological effects of H₂S. It can also contribute to the cardioprotective effects of H₂S by reducing the levels of ROS in the body [89].

H₂S can also activate the soluble guanylyl cyclase (sGC) by directing its interaction with the cGMP phosphodiesterase (PDE). In addition, this molecule can trigger the activation of the cyclic GMP-protein kinase G pathway [90]. It can also trigger the re-translation of eIF2 (eukaryotic initiation factor 2) [91] by increasing the phosphorylation of protein phosphatase-1 [27].

H₂S can also convert the -SH group of cysteine into a -SSH group, which can alter the activities of various enzymes such as the F₁F₀-ATPase pump, KATP channels, and the phosphatase and tensin homolog (PTEN) [14]. This can lead to the disappearance of certain protein S-sulfates. S-sulfhydration is a post-translational process that produces a
hydropersulfide moiety or polysulfide in specific body regions. It is known to regulate the cellular functions of H$_2$S [92]. HMG-CoA reductase [35] is an enzyme involved in the ubiquitination of various substrate proteins, such as Hrd1. H$_2$S induces the degradation of VAMP3 (vesicle-associated membrane protein 3), which controls exocytosis by Hrd1 S-sulfhydration. It is also known to trigger the translocation of CD36, which can cause lipid toxicity in the body [58].

Cytokines are small molecules that help the cell produce pro-inflammatory signals. Increased production of cytokines in the serum and heart muscle is a common feature of cardiovascular disease involving cell death. The JAK/STAT signaling pathway is a vital pathway for cytokine signal transduction and a pleiotropic cascade involved in growth hormone receptors’ activity and regulates various physiological and pathological processes, including proliferation, differentiation, apoptosis, and cellular immunity, and inflammation. In addition, this pathway can also increase the expression of TGF and type III collagen [93].

The pro-inflammatory cytokine TNFα can also induce apoptosis and necrosis. The stimulation of TNFα in the liver and macrophages can increase the secretion of H$_2$S. It has been observed that the treatment with LPS leads to an increase in the production of both IL-6 and TNFα, an epigenetic regulation mechanism [94].

In addition, treating patients with H$_2$S can decrease the production of neutrophils in the myocardium and contribute to the development of anti-apoptotic signaling. Neutrophils are recruited into the myocardium to express IL-1β and TNFα. H$_2$S reduces these immunity cells, correlated with promoted Bcl-2 anti-apoptotic signaling, decreases cytokine release, and preserves cardiac function [95].

VEGF is a pro-angiogenic cytokine that promotes endothelial cell survival. In the case of acute coronary syndrome, the reduction of VEGF leads to the depletion of microvessels. On the other hand, H$_2$S can prevent coronary artery disease and improve the survival of endothelial cells [96].

TGF is a critical cytokine in the development of cardiac remodeling. Myofibroblasts can promote the growth and deposition of collagen in the body. However, the presence of H$_2$S can inhibit the signaling cascade in myofibroblasts and limit the proliferation and survival of these cells. The cytoprotective effects of H$_2$S on cell death appear to act through cell types other than cardiomyocytes, where it influences TGFβ expression and inhibits the signaling cascade in fibroblasts. This restricts the differentiation and proliferation of fibroblasts into myofibroblasts and prevents the deposition of collagen in the heart [22].

H$_2$S and NO are physiological and pathological factors that have been extensively studied lately. They have been linked to the development of diabetes and heart failure [92]. The exposure of mice to H$_2$S can stimulate the production of NO through the activation of the eNOS pathway [97]. This can result in the development of more severe cardiac dilatation. Treating patients with H$_2$S using CSE overexpression can also improve the function and structure of their hearts after undergoing transverse aortic constriction. This therapy activates the eNOS-NO-cGMP pathway [36] and can prevent hepatic and myocardial ischemia-reperfusion injury [98–100].

3.3. H$_2$S in Pharmacology and Pathophysiology

H$_2$S levels are decreased in several conditions (e.g., DM, ischemia, and aging), even in COVID-19 [37], and are elevated in other statuses (e.g., inflammation, critical biological disbalances, and cancer). In recent decades, multiple approaches to the therapeutic exploitation of H$_2$S have been identified, either based on H$_2$S exogenous apport or decreased H$_2$S biosynthesis [43]. Inhibition and stimulation of H$_2$S synthesis have been suggested as potential interventions in DM.

Treating patients with H$_2$S can improve the recovery of liver and myocardial ischemia-reperfusion injury. It can also restore the damaged endothelium-dependent relaxation caused by NO depletion [13]. H$_2$S is known to produce nitroxyl, a one-electron reduction of NO. It can also help restore the relaxation caused by the depletion of NO [101].
Various studies have also shown that H$_2$S can promote the development of new blood vessels. Most of these studies were focused on the effects of VEGF on the angiogenic response [11]. Silencing of CSE by siRNA can also decrease the impact of VEGF-induced angiogenesis. It can also stimulate the activity of various cellular signaling pathways, such as the eNOS-NO-c pathway and the K1A2T7P signal transducer. H$_2$S promotes angiogenesis by increasing the activity of endothelial nitric oxide synthesis (eNOS), phosphatidylinositol 3 (PI3)-kinase/protein kinase B (AKT), p38/MAPK, K1A2T7P, signal transducer and activator of transcription 3 (STAT3), and sirtuin 1 (SIRT1)/VEGF/cyclic guanosine 5′-monophosphate (cGMP) cascade [102].

Although diabetes has been known to impair the development of new blood vessels [13], the mechanism of this process is not yet precise. The absence of vascular perfusion leads to diabetes-induced angiogenesis. H$_2$S rescues the migration of HUVECs in mice with hyperglycemia-induced migration. The effects of this condition on the pro-angiogenic and bio-energetic properties were also studied. H$_2$S improves the revascularization of diabetic mice through increasing NO bioavailability and promotes the development of vascular progenitors [86].

Under pathological conditions, the levels of H$_2$S and its production enzymes are significantly altered [103]. This can lead to the development of various cardiac disorders [38]. In addition, H$_2$S increases the filtration rate and kidney blood flow [62] and generates an increase in the excretion of certain nutrients, such as K$^+$ and Na$^+$. The role of the Renal-Angiotensin system (RAS) [22] is well established in the pathogenesis of various diseases. It plays a central role in regulating physiological function and possesses neuronal control of the circulatory system. H$_2$S is also known to interact with the zinc metalloproteinase, a zinc metalloproteinase. In addition, studies show that this protein can reduce the activity of the angiotensin-converting enzyme (ACE) in human endothelial cells.

Myocyte stretching releases angiotensin II (ANG II) [3], increases p53 binding to the ANG II promoter and the AT1 (angiotensin II type 1) receptor, and results in a four- to seven-fold increase in apoptosis. Adding Zn$^{2+}$ to the diet lowered the ACE mRNA level and reduced ROS production. H$_2$S could also alter RAS signaling, interacting with the ACE, a zinc metalloproteinase [62]. A dose-dependent drop in ACE activity in human endothelial cells after treatment with H$_2$S was observed. Supplementation of H$_2$S in DM rats reversed RAS activation and reduced ROS production. H$_2$S could alter RAS signaling, reducing oxidative stress [22].

As a neuromodulator, H$_2$S can improve the effects of diabetes on the central nervous system (CNS). Due to the impact of diabetes on the CNS, it is considered a leading cause of cognitive decline [39]. H$_2$S can also reduce the risk of cognitive decline and microvascular complications [94]. An equilibrated balance of oxidative stress/antioxidants is essential for maintaining cellular function. When this is disturbed, the other molecules, such as deoxyribonucleic acid, lipid, and protein oxidize, imprinting a pathological condition, like diabetes [24]. Oxidative stress comes from the overproduction of reactive oxygen and nitrogen species. The main source of ROS is the mitochondria [99]. Oxidative stress in diabetic patients determines dysfunctions during insulin secretion in the nervous system, and thus, neurodegeneration, such as diabetic peripheral neuropathy (DPN), occurs [104].

The pancreatic β cell is the most essential metabolically active part of the body, where metabolites take place for energy synthesis at the high glucose concentration level. H$_2$S displays antioxidant effects by directly silencing reactive oxygen species (ROS) via a hydro-sulfide anion (HS$^-$), a powerful one-electron chemical reductant dissociated from H$_2$S in a physiological fluid. H$_2$S can improve the function of the mitochondria, which is a type of respiratory chain that produces oxygen [105]. Overproduction of reactive nitrogen and oxygen species can lead to oxidative stress. The free radicals produced by these species can be suppressed by antioxidant molecules [14]. H$_2$S can also decrease ROS production by suppressing the copper/zinc superoxide activation. In addition, it can also prevent the degradation of antioxidant enzymes and proteins [63].
Autophagy is emerging as a critical cellular stress response that is involved in a variety of disease states. Autophagy is a highly conserved self-feeding pathway that degrades macromolecules and damaged organelles to maintain intracellular homeostasis. It has been shown that H₂S is a regulator of autophagy. Generally, autophagy serves a dual purpose: it may play a cytoprotective or harmful role in the body, hanging on the type and severity of the lesion it causes [22]. A certain degree of autophagic activity is essential in promoting tissue homeostasis and cell survival. However, excessive autophagic activity can contribute to apoptosis on the other side. In addition, autophagy dysfunction is involved in diabetic cardiomyopathy [106].

Autophagy is a well-coordinated, multi-stage process regulated by autophagy-related genetic products and proteins, such as Beclin1 and P62. Exogenous H₂S facilitates the elimination of autophagosome contents, which improves autophagy. The promotional effects of exogenous H₂S on autophagy may be essential for decreased ROS production. In addition, there are studies that ubiquitin aggregate clearance is mainly dependent on autophagy, and disruption of autophagy results in the accumulation of ubiquitin aggregates in cells [33].

H₂S has its regulatory role in autophagy during the development and progression of numerous diseases, such as diabetes, heart failure, or Parkinson’s disease [16]. Exogenous H₂S reduces the ubiquitination level. Recent studies have found that Keap-1 is crucial in eliminating ubiquitin proteins [107]. Keap-1 can be a critical factor in the protective role of exogenous H₂S on ubiquitin aggregate clearance via autophagy [16]. Exogenous H₂S upregulates the expression of Keap-1. Reported data show that Keap-1 regulates the translocation of Nrf2, a negative regulator of ROS production. However, exogenous H₂S had no significant effects on the translocation of Nrf2 to the nucleus. A recent study demonstrated that H₂S suppressed diabetes-accelerated atherosclerosis via Nrf2 [108].

H₂S rectifies high glucose/palmitate-induced excessive autophagy in endothelial cells. The Nrf2-ROS signaling pathway can trigger this effect. However, exogenous H₂S inhibits mitochondrial apoptosis and promotes mitochondrial autophagy, thus protecting endothelial cells against apoptosis induced by high glucose and palmitate. Therefore, it has been hypothesized that H₂S can promote the normal development of the diabetic endothelial system by suppressing the excessive autophagy that occurs following stressful events [44]. The optimal window of autophagy is maintained in response to stressful events. However, if it is excessive, autophagy is maladaptive, leading to cell death [22].

Some studies showed that H₂S upregulates autophagy and others that H₂S inhibits autophagy. H₂S plays diverse roles in autophagy depending on the tissue and disease. For example, H₂S could downregulate LC3BII and Beclin-1 protein expression and upregulate p62 protein expression in VSMCs (vascular smooth muscle cells) under HG (high glucose) conditions, which could be reversed by rapamycin, an autophagy activator. Furthermore, NaHS decreased the autophagy induced by HG in VSMCs. Similarly, ALA (Alpha-lipoic acid) could also inhibit autophagy in VSMCs under HG conditions via the AMPK/mTOR signaling pathway. Autophagy is regulated by many signaling pathways, among them the AMPK/mTOR signal pathway being crucial. The activation of the AMPK/mTOR pathway in DM has been widely studied. Increased AMPK phosphorylation and decreased mTOR phosphorylation activate autophagy [33]. H₂S also downregulates autophagy via the AMPK/mTOR signaling pathway [33].

An essential regulator of inflammation associated with metabolic syndrome is the nucleotide-binding domain, leucine-rich-containing family, pyrin domain containing-3 (NLRP3) inflammasome, which activates caspase-1, after interacting with the adaptor protein apoptosis-associated speck-like protein containing a C-terminal caspase recruitment domain (ASC). Induction of phosphorylation of the p65 subunit of NF-κB resulting in NF-κB signaling activation is a prerequisite for transcriptional activation of NLRP3 [109]. Cleavage, processing, and secretion of pro-inflammatory cytokines IL-1β and IL-18 result from NF-κB-mediated activation of NLRP3 inflammasome and subsequent caspase-1 activation [45].
H₂S can exert anti-inflammatory effects against free fatty acid (FFA)-induced inflammation and apoptosis in macrophages by suppressing TLR4/NF-κB-stimulated NLRP3 inflammasome activation. H₂S can thus prevent FFA-overload-mediated insulin resistance and type 2 DM [110].

H₂S exerted an anti-inflammatory role in diabetic myocardia by downregulation of Thioredoxin-interacting protein (TXNIP)-mediated NLRP3 inflammasome activation. H₂S alleviated hyperglycemia-mediated myocardial inflammation in type 1 DM. The mechanism may involve inhibiting TXNIP-mediated NLRP3 inflammasome activation, which might serve as an efficient, targeted therapy in diabetic cardiomyocytes [46].

Pyroptosis is a type of cell death with several characteristics that make it different from other forms of cell death. This type of cell death relies on the canonical pathway, dependent on caspase-1 [47], and the non-canonical pathway, reliant on caspase-11 [111]. Since pyroptosis is known to trigger the inflammatory response that contributes to chronic inflammatory diseases, it has shifted its focus away from the body’s natural defenses. The downstream inflammatory markers of pyroptosis are associated with toxic shock, nephropathy, and pathogen defense. The NLRP3 (NOD-like receptor protein 3) inflammasome activates caspase-1 in the canonical pyroptosis pathway. NLRP3 also localizes to the mitochondria and supplies high ROS production, but it can be inhibited by H₂S [48].

The Mfn-2 protein is known to promote early apoptotic events in the mitochondria. The fragmentation of the mitochondria network can also lead to the development of these events. It has also been shown that Mfn-2 can prevent the transfer of Ca²⁺ from the endoplasmic reticulum to the adjacent mitochondria. High glucose levels can also promote the growth of H9C2 cells through the increased expression of Mfn-2- and siRNA-mediated Mfn-2 silencing [30,49].

Necrosis is a version of cell death that occurs following severe injury. It is programmed to utilize the TNF receptor and the RIPK1/RIPK3 necrosome. The effects of H₂S on necroptosis and necrosis are limited. However, recent studies have shown that treating cardiomyocytes with high glucose levels can inhibit these markers [112].

A non-canonical death pathway known as MPT (mitochondrial permeability transition pore) is also initiated by Ca²⁺ and ROS. These stressors open the nonspecific MPT pore in the mitochondrial inner membrane, dissipate inner membrane potential, and rupture both mitochondrial membranes through osmotic swelling. The absence of the outer membrane can prevent the formation of apoptotic bodies. Instead, cell death occurs through the accumulation of necrosis. Data show that H₂S protects against MPT-driven necrosis in the heart and brain [113].

3.4. H₂S and Insulin Secretion and Sensitivity

Insulin resistance and compromised insulin secretion lead to impaired glucose metabolism, which contributes to the development of Diabetes [17]. H₂S could be produced endogenously in the pancreatic island’s β cells, liver, fat, skeletal muscle, and hypothalamus and regulates local and systemic carbohydrates metabolism [51]. Specifically, H₂S is reported to suppress insulin secretion and promote or reduce islet β-cell apoptosis. It influences insulin sensitivity. H₂S also suppresses glucose uptake and glycogen storage and promotes or inhibits gluconeogenesis, mitochondrial bioenergetics [50], and mitochondrial biogenesis in the liver [52]. This gas also promotes glucose uptake into adipocytes in the fat tissue, while other studies have reported inhibiting this process. H₂S has been shown, as well, to increase adipogenesis, inhibit lipolysis, and regulate adiponectin and MCP-1 secretion in adipocytes [53]. H₂S increases glucose absorption in skeletal muscle, improves insulin sensitivity and modulates circadian clock genes in myocytes. The hypothalamic CB₈ (cystathionin-β-synthase)/H₂S pathway reduces obesity [54] and improves insulin sensitivity through brain–adipose interactions. Most studies have shown that plasmatic H₂S levels are lower in diabetic patients [50,55].
H$_2$S can influence insulin secretion and modulate circulating glucose levels. H$_2$S administration to β cell lines attenuates insulin secretion triggered by a high glucose concentration. High levels of H$_2$S can also decrease the secretion of insulin. It can also cause the membrane to become polarized and inhibit the KATP channel’s independent signaling. H$_2$S can also inhibit insulin secretion by affecting various biochemical processes: activation of KATP channels, inhibition of ATP synthesis, and inactivation of L-type voltage-dependent Ca$^{2+}$ channels [114].

The effects of hyperglycemia on insulin secretion can vary depending on the phase of diabetes development. During the early stages of the disease, increasing levels of H$_2$S can protect islet cells from further damage. During the development of diabetes, an increase in H$_2$S can inhibit the secretion of insulin and reduce the overload of islet cells. This can also trigger an increase in ER stress response [50].

Inhibitory effects of sodium hydrosulfide (NaSH, 10 µM–1 mM) and L-cysteine (0.1–10 mM) on glucose (10 mM)-induced insulin secretion has been observed in both isolated mouse islets and pancreatic β cell lines, an effect that was not observed at a low glucose concentration (3 mM) [115]. One of the mechanisms through which H$_2$S inhibits insulin secretion is through the opening of KATP channels, as the inhibitory effects of NaSH and L-cysteine on insulin secretion were reproduced after using tolbutamide (a KATP blocker), α-ketoisocaproate (a mitochondrial fuel), and high K$^+$ condition (30 mmol/L). Interactions between H$_2$S with KATP channels seems to be mediated through functional manipulation, probably by decreasing selective cysteine residues of the KATP channel protein, independent of cytosolic second messengers. It has been suggested that the S-sulfhydration of KATP channels is a mechanism by which H$_2$S could influence insulin secretion [56].

Hepatic insulin resistance reveals the failure of insulin to inhibit glycogenolysis and gluconeogenesis in the liver to maintain normal plasma glucose levels. The enzymes CSE, CBS, and 3-MST, responsible for endogenous H$_2$S, are found in the liver. The effects of diabetes mellitus and its related pathologies on the H$_2$S production system in the liver are controversial. Compared with nondiabetic rats, H$_2$S production and CSE and CBS mRNA levels in the liver were increased in STZ diabetic rats, while insulin treatment reversed these effects [114]. H$_2$S regulates glucose uptake, glycogen storage, and gluconeogenesis. H$_2$S is a key component of liver glucose metabolism [50].

3.5. H$_2$S and Neurological Dysfunctions as Diabetes Associated Diseases

Neurological research concerning diabetes patients with complications such as Alzheimer’s disease, Parkinson’s disease, and amyotrophic lateral sclerosis present changes in the central nervous system because of the high blood glucose levels (HbA1c) correlated with poor cognitive function. Oxidative stress plays a part in inhibiting insulin signaling, which is necessary for brain function [110]. The regulation of Schwann cells, aggregation of sorbitol during signaling of the polyol pathway, inactivation of Na$^+$/K$^+$ signaling, and hyperglycemia-induced oxidative stress are causative factors of neuropathy in the brain. About 50% of diabetic patients are affected by neurological disorders, the most common comorbidities of DM. In addition, some neurodegenerative diseases, like Alzheimer’s disease (AD), Parkinson’s disease (PD), and amyotrophic lateral sclerosis (ALS), coincide in the central nervous system (CNS) in diabetic patients because of oxidative stress [24].

H$_2$S inhibits Aβ-induced neuronal apoptosis by regulating mitochondrial function. In addition, H$_2$S could inhibit the expression of IL-23/IL-17 axis and mitochondrial apoptotic proteins to alleviate the cognitive decline caused by DM [32].

The repercussions of hyperglycemia-induced oxidative stress on neurons vary in T1DM and T2DM, and DPN patients with T2DM have a low capacity to control hyperglycemia. In the peripheral nerves of T2DM patients, oxidative stress is increased from proximal to distal parts, passes from DRG to the sciatic nerve, and decreases the metabolism under the glycolytic and tricarboxylic acid cycle [29]. In AD, cholinergic homeostasis is hampered by downregulation of insulin/insulin growth factor (IGF) resistance, leading to
downregulation of target genes. There is evidence that 40% of patients with DM develop PD, and glucose is impaired at an early stage. However, H$_2$S can mitigate the effects of oxidative stress on nerve cells.

3.6. H$_2$S and Cardio-Vascular Dysfunctions as Diabetes Associated Diseases

Cardiovascular complications frequently cause hospitalization and death among diabetic patients. The early-onset diastolic dysfunction is the main characteristic of diabetes cardiomyopathy (DCM), an independent complication of diabetes, secondary to myocardial fibrosis [30]. DCM is a type of cardiomyopathy of unknown etiology, responsible for 75% of idiopathic dilated cardiomyopathy cases among diabetic patients. DCM characteristics are represented by impaired myocardial insulin signaling, mitochondrial dysfunction, overstimulation of the sympathetic nervous system, oxidative stress, increased inflammation, coronary microcirculation dysfunction, and inadequate immune response. These pathophysiological changes lead to fibrosis, hypertrophy, cardiac diastolic and/or systolic dysfunction, and ultimately, heart failure [35].

One of the alarming structural characteristics of DCM is represented by the overproduction and deposition of myocardial interstitial collagen, which leads to cardiac interstitial fibrosis, myocardial rigidity, and cardiac dysfunction. Although the precise mechanism of these changes has not been fully elucidated, the available literature suggests the cellular implication of oxidative stress, cell apoptosis, autophagy, inflammation, and endoplasmic reticulum stress are the main triggers [16].

Homocysteine transsulfuration produces H$_2$S, a gaseous signaling molecule with a cardioprotective role that is capable of preventing cardiac remodeling, cell death, and pyroptosis. Research activity in vitro and on mouse models demonstrated that H$_2$S inhibits caspase-1 activity and IL-1 secretion, with important suppression activity on pyroptosis in ischemic cardiomyopathy [116].

H$_2$S has reducing hypertensive effects and has an important protective role in cardiomyopathy models. Moreover, H$_2$S is an essential signaling molecule of the cardiovascular system with physiological and pathological mechanisms in ensuring homeostasis [80].

Accelerated atherosclerosis is a common cardiovascular complication in diabetic patients [117]. With a much higher incidence than in non-diabetic patients, atherosclerosis has an earlier onset and a higher mortality rate. Unfortunately, there is no proven treatment capable of slowing down atherosclerosis in DM. On the other hand, H$_2$S has important effects on atherosclerotic plaque stabilization and on hyperglycemia-induced endothelial dysfunction, being capable of ischemia-reperfusion injury, myocardial infarction, and heart failure prevention [118].

Different pathological states of the venous and/or arterial system characterize the vascular dysfunction or vascular disease, a pathology capable of inducing adverse cardiovascular events. We mention atherosclerosis, arterial remodeling, thrombosis, and restenosis, among these pathologies. The main cardiovascular risk factors (diabetes, obesity, hypertension, aging) are responsible for vascular dysfunction through mechanisms such as oxidative stress, an essential target in therapeutic and preventive strategies. Cellular oxidation is a tightly regulated process involving both pro- and antioxidant systems from different cellular compartments in physiological conditions [87].

Results from current literature support the multiple beneficial roles of H$_2$S in diabetic cardiovascular complications. First, H$_2$S slows down the onset and improves the prognosis of diabetic cardiomyopathy. Second, H$_2$S treatment ameliorates high-fat diet (HFD)-induced cardiac dysfunction through sulfide levels restoration; activation of adiponectin-AMPK signaling and decrease in HFD-induced ER stress secondary to H$_2$S underlines its protective effects. Third, adiponectin’s essential cardiovascular protective role strengthens the correlation between low adiponectin levels and high cardiovascular risk. Fourth, adiponectin uses the AMPK to deliver its metabolic regulatory effects. AMPK increases the expression of GLUT4, which stimulates glucose transport and modulates fatty acid
oxidation and cardiac lipid accumulation through the phosphorylation and inhibition of acetyl-coenzyme [50].

In recent studies, H$_2$S was shown to be involved in the regulation of various vascular conditions, such as nephropathy, retinopathy, and neuropathy. H$_2$S-releasing agents could potentially be used as a treatment for diabetes-related endothelial dysfunction. They could help restore the function of the vascular endothelial cells [7].

Many authors also noted that the use of H$_2$S-releasing agents could be beneficial for treating diabetes by blocking the formation of advanced glycation end products (AGEs), which can lead to the development of vascular complications, and contribute to the degradation of the endothelium’s functionality. The rats that were treated with H$_2$S-releasing agents exhibited a decrease in their vascular oxidation stress levels. This beneficial effect was also partially explained by the compound’s ability to increase the NO level [7].

3.7. H$_2$S and Renal Dysfunctions as Diabetes Associated Diseases

In renal physiology, H$_2$S induces vasodilation and increases renal blood flow and glomerular filtration rate, resulting in an indirect increase in the urinary excretion of Na$^+$ and K$^+$. In addition, H$_2$S exhibits an inhibitory effect on specific Na$^+$ and K$^+$ kidney transporters, thus further increasing the excretion of such electrolytes into the urine. Furthermore, H$_2$S acts as an oxygen sensor in the renal system, especially in the medulla. Moreover, H$_2$S is found to inhibit renin release in rat models of renovascular hypertension. Hypertension-related nephropathy, a consequence of long-term hypertension, is the second leading cause of chronic kidney disease in the world. The blood pressure-lowering [119] actions of exogenous H$_2$S donors have been demonstrated in spontaneously hypertensive rats, angiotensin II-induced hypertension, N$^\text{ω}$-nitro-L-argininemethyl ester (L-NAME)-induced hypertension, and renovascular hypertension [120]. Furthermore, renal protective effects of H$_2$S are observed in hypertensive animal models [121].

In Diabetic Nephropathy (DN) [122], the increased expression of TGF-β1 has been shown to promote the accumulation of ECMs such as collagens and fibronectin, apoptosis, dedifferentiation of podocytes, and epithelial-mesenchymal transition of proximal tubules, all of which are considered to facilitate renal hypertrophy and dysfunction. ERK$_1^2$, a member of the MAPK family, may be expressed in mesangial cells in the condition of high glucose. ERK$_1^2$ may upregulate TGF-β1 expression. Dysregulation of matrix metalloproteinases, (MMPs) or tissue inhibitors (TIMPs), are involved in the mechanism of renal fibrosis. MMPs are responsible for extracellular matrix degradation. MMPs and TIMPs construct a time-and-space-dependent system [68]. Treatment with H$_2$S could attenuate the progression of renal dysfunction in diabetic rats. The protective effects of H$_2$S are correlated with TGF-β1 signaling through the ERK$_1^2$ pathway [47].

3.8. H$_2$S Exogenous Sources as Possible Therapeutic Interventions in Diabetes or Related Diseases

H$_2$S is, on the one hand, a therapeutic natural gas [17] that is found in mofettic joints with carbon dioxide [123], in sulfurous waters [124], with appraised medical effects in balneotherapy [4], and, on the other hand, an endogenous gaseous signal substance in the organisms. Therefore, experimental animals can be exposed to an H$_2$S-rich environment to observe this gas’s physiological effects or toxicity. Reports show that when mice were exposed to 80 ppm of H$_2$S for 6 h, their oxygen intake dropped by ~50%, and the metabolic rate and core body temperature were also seriously decreased into a suspended animation state. Notably, lowering metabolic demand could help reduce tissue/cellular damage caused by trauma. However, a later study of other larger species indicated that H$_2$S only exerted thermoregulatory effects. In diabetes, H$_2$S could promote glucose uptake by ameliorating insulin resistance and reducing renal injury [125].

Peloid or therapeutic mud is a maturated mud with healing properties, composed of a complex intermixture of fine-grained natural substances of geologic and/or biologic origins, water, and standard organic composites from biological metabolic activity. Sapropelic muds or sapropels are found at the bottom of salt waters, originating from the action
of microorganisms on flora and fauna of the water basin [17]. The gaseous phase of sapropelic mud results from the biochemical processes involved in the mud formation (peloidogenesis): H₂S, CO₂, NH₄, CH₄, O₂, and Rn. H₂S has been reported as an active molecule of the mud, which can be absorbed through the skin [126], exerting numerous pharmacological effects. Under the action of mud, there is a harmonic stimulation in all glands to increase the enzymatic and synthetic activity, while maintaining the specificity of each. Usually, mud therapy is contraindicated in diabetic patients without glycemic control. Future research is necessary to elucidate the implications of mud therapy on diabetes [127].

Although less rigorously described in the scientific literature, H₂S is commonly used in the context of balneotherapy, where H₂S inhalation occurs as humans are soaking in H₂S-containing sulfurous waters, with at least 1 mg/L of H₂S. Hydrogen sulfide delivery into the body probably occurs via inhalation and absorption through the skin [128] or, in specific cases, when patients are sitting in closed rooms with H₂S donors and H₂S fountains of H₂S-containing thermal water placed in the middle of the room, where a sensor/ventilation feedback system regulates the H₂S concentration in the air of the room [124]. Small-scale preclinical studies demonstrate the beneficial effects of H₂S delivery via sulfurous waters [129]. In addition, exploratory clinical studies suggest the anti-inflammatory effects of ultrasonic nebulization with sulfurous water in asthmatic patients. However, the potential therapeutic effect of these approaches has not been studied in appropriately powered, randomized clinical trials [130].

One of the potential problems with all forms of H₂S delivery, but especially with H₂S inhalation, relates to the issue of possible overdosing and consequent intoxication. Although the inhibitory effect of H₂S on mitochondrial Complex IV is reversible and therefore supporting therapy can result in patient recovery in some cases, there are currently no well-characterized pharmacological antidotes to H₂S intoxication [131].

Under physiological pH, H₂S is in a specific equilibrium with HS⁻ in aqueous solutions. The HS⁻ and H₂S are in an 81 to 19% report. Inorganic sulfide salts, such as sodium sulfide (Na₂S) and sodium hydrosulfide (NaHS), are frequently used as H₂S equivalents in many kinds of research [132]. These salts are fast H₂S donors, as they produce H₂S after being dissolved in aqueous solutions [21].

The rapid volatilization of H₂S can cause it to escape from the buffers. This phenomenon could explain the discrepancy between the physiological responses required to trigger physiological responses in tissues and blood [133].

Many studies have used NaHS as a standard H₂S donor. For example, it was shown that NaHS could alleviate amyloid beta-peptide (Ab)-induced neural lesion in an Alzheimer’s disease cellular model. Furthermore, in hypoxic skin damage, NaHS could exert anti-inflammatory effects through inhibition of reactive oxygen species (ROS)-activated NF-kB/cyclooxygenase (COX)-2 [134].

Allicin is commonly used as a sulfur-containing compound in garlic. It can be considered an active H₂S pool. In aqueous solutions, it can transform various sulfur-containing combinations into H₂S. In contrast, the diallyl disulfide (DADS) produces only a limited amount of H₂S after a slow reaction with GSH. This process can be initiated by forming a cyclic disulfide [2].

3.9. Synthetic Slow-Releasing H₂S Donors

The types of donor that can be considered controlled are those with various release mechanisms [135]. Since using H₂S gas or sulfide salts in studies has been deemed dangerous, researchers have focused on synthetic molecules releasing H₂S [21]. For example, GYY4137 is a Lawesson’s reagent that can be used as a slow and safe source of H₂S. However, it is not as effective as an aqueous solution and can only be administered on animals. Another commonly used method is using sulfur-containing dithiolethione [132].

Thiomolybdate salts are thiol transfer reagents in organic synthesis [21]. The four sulfur atoms they present in their structures make them excellent copper chelators. Ammonium tetrathiomolybate (TTM) can release H₂S under strongly acidic conditions. As
such, it is possible to use TTM as an inorganic complex-based H₂S donor. TTM is a slow H₂S releaser. It was discovered that acidic pH increase TTM’s H₂S release [43].

In the last years, several ROS-activated H₂S donors were designed. For instance, carbonyl sulfide can be released through a cyclic anhydrase reaction. This process can be sped up by carbonic anhydrase. The tandem reaction will remove carbonyl sulfide (COS), as well as quinone and amine byproducts [136].

Further studies reveal that donors can also release H₂S through the intervention of an endogenous H₂O₂ in their cells. This method is similar to the cyclic anhydrase reaction. COS can easily undergo hydrolysis to produce H₂S if carbonic anhydrase (CA) is presented. However, studies showed that CA is unnecessary for the donors’ H₂S release, as H₂O₂ can also trigger the rapid H₂S release from COS [21]. The effects of oxidizing stress on the cell viability of donated human tissue were studied [76]. It was revealed that these individuals exhibited the most effective outcomes of oxidizing stress on their cells [137].

Recently were also communicated a series of esterase-activated H₂S donors. Association of esterase-activated donors with NSAID can form hybrid anti-inflammatory and anti-oxidative combined drugs that reduce NSAID-induced gastric damage [21].

Researchers also discovered nitroreductase-activated donors that could be used to release H₂S [21]. A type of H₂S-producing material is the polyNTA. This substance contains N-thiocarboxyanhydrides and undergoes a ring-opening reaction to donate H₂S [21]. A PEG-ADT (5-4-hydroxyphenyl-3H-1,2-dithiole-3-thione-conjugated with polyethylene glycol) can also be used to generate H₂S. Cell imaging studies also showed that PEG-ADT could enter cells through the endolysosome and last in the cytoplasm [21,43].

3.10. H₂S-Stimulating Agents

Aside from H₂S donors, some compounds can also stimulate the production of H₂S in vivo. For instance, the amino group L-cysteine is an essential substrate for the enzymes that produce H₂S. When acetylated, the resulting product N-acetyl-L-cysteine can increase the production of H₂S [138]. Two other cysteine derivatives, S-allyl-L-cysteine and S-propargyl-L-cysteine, can be used as CSE substrates to generate H₂S [139].

Vitamin D [11] is known to promote the growth and remodeling of bones [21]. In addition, researchers discovered that vitamin D could increase the concentration of H₂S in the liver and kidney. Notably, it was found that cholecalciferol, known as VD3, could increase tissue H₂S concentration in mouse heart, brain, and kidney. Meanwhile, another report suggested that VD3 could upregulate glucose transporter type 4 (GLUT4) and decrease glycemia in diabetes through stimulation of CSE expression and H₂S generation [21,50].

3.11. Clinical Studies on H₂S Donors/Exogenous Sources in Diabetes or Related Diseases—Meta-Analysis

Presentation of clinical studies on H₂S donors/exogenous sources in diabetes or related diseases (Figure 3, Table 3)

- https://clinicaltrials.gov
  - 17,498 Studies found for: Diabetes Mellitus
  - 2 Studies found for: H₂S/Hydrogen sulfide | Diabetes Mellitus
  - 18 Studies found for: hydrogen sulfide
- https://trialsearch.who.int
  - 6430 trials found: Diabetes Mellitus
  - 0 Studies found for: H₂S/Hydrogen sulfide | Diabetes Mellitus
  - 25 trials found for: hydrogen sulfide
- https://www.clinicaltrialsregister.eu
  - 2041 trials found: Diabetes Mellitus
  - 0 Studies found for: H₂S/Hydrogen sulfide | Diabetes Mellitus
  - 1 trial found for: hydrogen sulfide
The clinical trials that satisfied all the previous filtering criteria/PRISMA stages selected for qualitative synthesis were included in our meta-analysis to determine the using frequency of the medicine, as a poisonous and occasionally lethal toxic gas [66] formed from the decomposition of various organic materials. Therefore, it might also represent an industrial safety hazard, too, as it is colorless. However, H2S is a toxic byproduct of microbial metabolism in the atmosphere, depending on its concentration. The human nose could detect H2S at a level of 0.1 ppm [3].

On the other hand, H2S has been previously considered, inclusive in occupational and respiratory/associated diseases [141]. A wide range of interventions can extend the lifespan and healthspan of H2S, including dietary restriction. This is done through the removal of certain nutrients, such as sulfur-containing amino acids, but only methionine and cysteine are incorporated into proteins. Taurine was first isolated about 150 years ago from ox (Taurus) bile. Although taurine can be produced in vivo from cysteine, with the enzymatic help of cysteine dioxygenase [24], it cannot be synthesized de novo from other amino acids. One of the most common molecular factors that can affect the longevity of people is the altered metabolism of certain amino acids, such as methionine and cysteine, and the increased production capacity of H2S [142]. It is also believed that the presence of S has been previously considered, inclusive in occupational and respiratory/associated diseases [141]. A wide range of interventions can extend the lifespan and healthspan of H2S, including dietary restriction. This is done through the removal of certain nutrients, such as sulfur-containing amino acids, but only methionine and cysteine are incorporated into proteins. Taurine was first isolated about 150 years ago from ox (Taurus) bile. Although taurine can be produced in vivo from cysteine, with the enzymatic help of cysteine dioxygenase [24], it cannot be synthesized de novo from other amino acids. One of the most common molecular factors that can affect the longevity of people is the altered metabolism of certain amino acids, such as methionine and cysteine, and the increased production capacity of H2S [142]. It is also believed that the presence of H2S is indicated as a therapeutic intervention in these two pathological conditions [141].

It must be additionally specified that the pathogeny of DM has a common ground with the ischemia that generates a stroke: in both pathologic states, there is a depletion of ATP, the primary energy provider of metabolic processes [140], and an increase in the oxidative stress at the cellular level. Interestingly, H2S is indicated as a therapeutic intervention in these two pathological conditions [141].

On the other hand, H2S has been previously considered, inclusive in occupational medicine, as a poisonous and occasionally lethal toxic gas [66] formed from the decompo-
sition of various organic materials. Therefore, it might also represent an industrial safety hazard, too, as it is colorless. However, H\textsubscript{2}S is a toxic byproduct of microbial metabolism in the atmosphere, depending on its concentration. The human nose could detect H\textsubscript{2}S at a level of 0.1 ppm [3].

H\textsubscript{2}S is a widely used reducing agent that has unique chemical properties. It can be availed to target in this purpose various cellular and molecular components due to its nucleophilic nature. In addition, it can rapidly lose its chemical identity under multiple conditions, such as a tissue bath. For instance, under aerobic conditions, the half-life of H\textsubscript{2}S is about 2.0 min in human hepatic cells, 2.8 min in kidney tissues, and 10.0 min in brain homogenates [27].

H\textsubscript{2}S is a metabolite of sulfur amino acids in mammals, aside from SO\textsubscript{2} (sulfur dioxide) and Taurine. Taurine methionine, cysteine, and homocysteine are the four most common sulfur-containing amino acids, but only methionine and cysteine are incorporated into proteins. Taurine was first isolated about 150 years ago from ox (Taurus) bile. Although taurine can be produced in vivo from cysteine, with the enzymatic help of cysteine dioxygenase, it is mainly acquired from dietary sources, such as meat, eggs, and seafood. The mention of Taurine in the discussion session is determined by the fact that the only two clinical trials found, within our meta-analysis, in the above-mentioned searching platforms, address as primary pathologic condition DM, and H\textsubscript{2}S as the intervention, and these studies indicate Taurine as a drug used.

A wide range of interventions can extend the lifespan and healthspan of H\textsubscript{2}S, including dietary restriction. This is done through the removal of certain nutrients, such as amino acids. One of the most common molecular factors that can affect the longevity of people is the altered metabolism of certain amino acids, such as methionine and cysteine, and the increased production capacity of H\textsubscript{2}S [142]. It is also believed that the presence of H\textsubscript{2}S can delay the onset of aging by blocking the activation of the silent information regulator of the transcription 1 protein (SIRT1). In some studies, it is suggested that the use of dietary restriction for a specific duration can increase the production of H\textsubscript{2}S in rats [143].

As pointed out in the meta-analysis, most patients presented associated cardiovascular diseases (637, representing 51.50% of the total number). It must be emphasized that the main interventions proposed in the eligible clinical trials include Taurine (400 patients representing 32.34% of the total number) and sodium thiosulfate (380 patients representing 30.72% of the total number). It must also be underlined that there was no drug used for 387 patients/subjects (representing 31.29% of the total number), but only the measurement of the H\textsubscript{2}S plasma level.

Various analytical methods have been used to determine the concentration of H\textsubscript{2}S in blood and other tissues, such as fluorescent tools, colorimetry, spectrophotometric analysis, headspace gas determination, polarography, and liquid chromatography-mass spectrometry. Yet, different analysis methods have obtained very diverse intervals of H\textsubscript{2}S concentrations. Furthermore, the levels of H\textsubscript{2}S within tissues and plasma are also significantly different, ranging from 15 nM [18]—for instance, in human plasma—to 300 µM in animal tissues [8] in vivo. These high discrepancies—including as regards to sensitivity and specificity items—can be challenging and, at the same time, require both cautiousness in integrating the related data and must be worthy of further minute research in this domain [18].

In very recent studies, it has been shown that H\textsubscript{2}S can provide therapeutic effects in COVID-19 too. In addition, it is well known that DM is one of the comorbidities which dramatically increases the risk for aggravation of SARS-CoV-2 infection. “Cytokine storm” is a dominant paradigm in explaining the pathogenesis of COVID-19, being involved in many signaling pathways H\textsubscript{2}S influences, including the immune system functioning. Therefore, linking DM to H\textsubscript{2}S and COVID-19 is an interesting and justified quest direction [37,144–147].

H\textsubscript{2}S: entrance in the organism, its plasma levels, signaling, metabolism, and their regulation, and also its pathogenic roles, represent topics that warrant an enhanced quest
focus. At the same time, being a constituent of sapropelic muds, sulfurous mineral waters, and solfatara—natural sanogenic resources used in balneology—another scientific and practical goal is to promote them based on current, thorough evidence acquired through adequate research activities. Although H$_2$S biology and medical usefulness have expanded over the last decades, many related issues/hurdles remain to be further explored, explained, and hopefully overcome [17].

5. Conclusions

The available data in this field have revealed interesting sulfur-related biological mechanisms, potentially impacting DM pathophysiology and treatment (as considered, too, empirically in older balneological approaches). Hopefully, future studies will clarify many still poorly known and/or debatable aspects of the subject we approached and pave the way to a better turn to good account, including from bench side to bedside, of the interesting and subtle biological properties of H$_2$S.

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References
1. Munteanu, C.; Păun, D.-L.; Șută, A.-M.; Floreescu, S.A.; Onose, G. Diabetes mellitus and COVID-19 in the post-acute phase patients—possible links with physical and rehabilitation medicine and balneotherapy. Balneo Res. J. 2020, 11, 350–367. [CrossRef]
2. Melino, S.; Leo, S.; Papajani, V.T. Natural Hydrogen Sulfide Donors from Allium sp. as a Nutraceutical Approach in Type 2 Diabetes Prevention and Therapy. Nutrients 2019, 11, 1581. [CrossRef] [PubMed]
3. Sun, H.-J.; Wu, Z.-Y.; Nie, X.-W.; Wang, X.-Y.; Bian, J.-S. An Updated Insight Into Molecular Mechanism of Hydrogen Sulfide in Cardiomyopathy and Myocardial Ischemia/Reperfusion Injury under Diabetes. Front. Pharmacol. 2021, 12, 651884. [CrossRef] [PubMed]
4. Munteanu, C.; Rotariu, M.; Dogaru, G.; Ionescu, E.V.; Ciobanu, V.; Onose, G. Mud therapy and rehabilitation—Scientific relevance in the last six years (2015–2020) Systematic literature review and meta-analysis based on the PRISMA paradigm. Balneo PRM Res. J. 2021, 12, 1–15. [CrossRef]
5. Sashi Papu, J.A.; Kundu, S.; Pushpakumar, S.; Amin, M.; Tyagi, S.C.; Sen, U. Hydrogen sulfide inhibits Ca$_{2+}$-induced mitochondrial permeability transition pore opening in type-1 diabetes. Am. J. Physiol.—Endocrinol. Metab. 2019, 317, E269–E283. [CrossRef] [PubMed]
6. George, A.K.; Singh, M.; Homme, R.P.; Majumder, A.; Sandhu, H.S.; Tyagi, S.C. A hypothesis for treating inflammation and oxidative stress with hydrogen sulfide during age-related macular degeneration. Int. J. Ophthalmol. 2018, 11, 881–887. [CrossRef]
7. Citi, V.; Martelli, A.; Gorica, E.; Brogi, S.; Testai, L.; Calderone, V. Role of hydrogen sulfide in endothelial dysfunction: Pathophysiology and therapeutic approaches. J. Adv. Res. 2021, 27, 99–113. [CrossRef]
8. Gheibi, S.; Samsonov, A.P.; Gheibi, S.; Vazquez, A.B.; Kashfi, K. Regulation of carbohydrate metabolism by nitric oxide and hydrogen sulfide: Implications in diabetes. Biochem. Pharmacol. 2020, 176, 113819. [CrossRef]
9. Stehouwer, C.D.; Lambert, J.; Donker, A.; Van Hinsbergh, V.W. Endothelial dysfunction and pathogenesis of diabetic angiopathy. *Cardiovasc. Res.* 1997, 34, 55–68. [CrossRef]

10. Zou, W.; Yuan, J.; Tang, Z.-J.; Wei, H.-J.; Zhu, W.-W.; Zhang, P.; Gu, H.-F.; Wang, C.-Y.; Tang, X.-Q. Hydrogen sulfide ameliorates cognitive dysfunction in streptozotocin-induced diabetic rats: Involving suppression in hippocampal endoplasmic reticulum stress. *Oncotarget* 2017, 8, 64203–64216. [CrossRef]

11. Rey, F.; Ottolenghi, S.; Galliongo, T.; Balsari, A.; Martinelli, C.; Rey, R.; Allevi, R.; Giulio, A.M.; Zuccotti, G.V.; Mazzucchelli, S.; et al. Mitochondrial metabolism as target of the neuroprotective role of erythropoietin in Parkinson’s disease. *Antioxidants* 2021, 10, 121. [CrossRef] [PubMed]

12. Testai, L.; Brancalone, V.; Fiori, L.; Montanaro, R.; Calderone, V. Modulation of EndMT by Hydrogen Sulfide in the Prevention of Cardiovascular Fibrosis. *Antioxidants* 2021, 10, 910. [CrossRef] [PubMed]

13. Ciccone, V.; Genah, S.; Morbidelli, L. Endothelium as a Source and Target of H2S to Improve Its Trophism and Function. *Antioxidants* 2021, 10, 486. [CrossRef] [PubMed]

14. Kang, S.C.; Sohn, E.-H.; Lee, S.R. Hydrogen Sulfide as a Potential Alternative for the Treatment of Myocardial Fibrosis. *Oxidative Med. Cell. Longev.* 2020, 2020, 4105382. [CrossRef]

15. Lee, Z.-W.; Teo, X.-Y.; Tay, E.Y.-W.; Tan, C.-H.; Hagen, T.; Moore, P.K.; Deng, L.-W. Utilizing hydrogen sulfide as a novel anti-cancer agent by targeting cancer glycolysis and pH imbalance. *J. Cereb. Blood Flow Metab.* 2014, 171, 4322–4336. [CrossRef]

16. Wu, J.; Tian, Z.; Sun, Y.; Lu, C.; Liu, N.; Gao, Z.; Zhang, L.; Dong, S.; Yang, F.; Zhong, X.; et al. Exogenous H2S facilitating ubiquitin aggregates clearance via autophagy attenuates type 2 diabetes-induced cardiomyopathy. *Cell Death Dis.* 2017, 8, e2992-11. [CrossRef]

17. Munteanu, C.; Munteanu, D.; Onose, G. Hydrogen sulfide (H2S)—Therapeutic relevance in rehabilitation and balneotherapy Systematic literature review and meta-analysis based on the PRISMA paradigm. *Balneo PRM Res. J.* 2021, 12, 176–195. [CrossRef]

18. Ghashghaeinia, M.; Mrowietz, U. Human erythrocytes, nuclear factor kappaB (NF-κB) and hydrogen sulfide (H2S)—From non-genomic to genomic research. *Cell Cycle* 2021, 20, 2091–2101. [CrossRef]

19. Marunaka, Y. Roles of interstitial fluid pH in diabetes mellitus: Glycolysis and mitochondrial function. *World J. Diabetes* 2015, 6, 125. [CrossRef]

20. Hu, B.; Xu, G.; Zheng, Y.; Tong, F.; Qian, P.; Pan, X.; Zhou, X.; Shen, R. Chelerythrine Attenuates Renal Ischemia/Reperfusion-induced Myocardial Injury by Activating CSE/H2S via PKC/NF-κB Pathway in Diabetic Rats. *Kidney Blood Press. Res.* 2017, 42, 379–388. [CrossRef]

21. Yang, C.-T.; Chen, L.; Xu, S.; Day, J.J.; Li, X.; Xian, M. Recent Development of Hydrogen Sulfide Releasing/Stimulating Reagents and Their Potential Applications in Cancer and Glycometabolic Disorders. *Front. Pharmacol.* 2017, 8, 664. [CrossRef] [PubMed]

22. Kar, S.; Kambis, T.N.; Mishra, P.K. Hydrogen sulfide-mediated regulation of cell death signaling ameliorates adverse cardiac remodeling and diabetic cardiomyopathy. *Am. J. Physiol. Circ. Physiol.* 2019, 316, H1237–H1252. [CrossRef] [PubMed]

23. Munteanu, C.; Dogaru, G.; Rotariu, M.; Onose, G. Therapeutic gases used in balneotherapy and rehabilitation medicine—Scientific relevance in the last ten years (2011–2020)—Synthetic literature review. *Balneo PRM Res. J.* 2021, 12, 111–122. [CrossRef]

24. Jeong, N.Y.; Jung, J.; Tabassum, R. Protective effect of hydrogen sulfide on oxidative stress-induced neurodegenerative diseases. *Neural Regen. Res.* 2020, 15, 232–241. [CrossRef]

25. Luo, R.; Hu, S.; Liu, Q.; Han, M.; Wang, F.; Qiu, M.; Li, S.; Li, X.; Yang, T.; Fu, X.; et al. Hydrogen sulfide upregulates renal AQP-2 protein expression and promotes urine concentration. *EASEB J.* 2018, 33, 469–483. [CrossRef]

26. Yang, R.; Jia, Q.; Ma, S.; Wang, Y.; Mehmood, S.; Chen, Y. Exogenous H2S mitigates myocardial fibrosis in diabetic rats through suppression of the canonical Wnt pathway. *Int. J. Mol. Med.* 2019, 44, 549–558. [CrossRef]

27. Liu, M.; Li, Y.; Liang, B.; Li, Z.; Jiang, Z.; Chu, C.; Yang, J. Hydrogen sulfide attenuates myocardial fibrosis in diabetic rats through the JAK/STAT signaling pathway. *Int. J. Mol. Med.* 2018, 41, 1867–1876. [CrossRef]

28. Sun, Y.; Teng, Z.; Sun, X.; Zhang, L.; Chen, J.; Wang, B.; Lu, F.; Liu, N.; Yu, M.; Peng, S.; et al. Exogenous H2S reduces the acetylation levels of mitochondrial respiratory enzymes via regulating the NAD+-SIRT3 pathway in cardiac tissues of db/db mice. *Am. J. Physiol. Metab.* 2019, 317, E284–E297. [CrossRef]

29. Roa-Coria, J.E.; Pineda-Farias, J.B.; Barragán-Iglesias, P.; Quíñonez-Bastidas, G.N.; Zuñiga-Romero, Á.; Huerta-Cruz, J.C.; Reyes-García, J.G.; Flores-Murrieta, F.J.; Granados-Soto, V.; Rocha-González, H.I. Possible involvement of peripheral TRP channels in the hydrogen sulfide-induced hyperalgesia in diabetic rats. *BMC Neurosci.* 2019, 20, 1. [CrossRef]

30. Yang, F.; Yu, X.; Li, T.; Wu, J.; Zhao, Y.; Liu, J.; Sun, A.; Dong, S.; Wu, J.; Zhong, X.; et al. Exogenous H2S regulates endoplasmic reticulum-mitochondria cross-talk to inhibit apoptotic pathways in STZ-induced type I diabetes. *Am. J. Physiol.—Endocrinol. Metab.* 2017, 312, E190–E203. [CrossRef]

31. Zhao, S.; Li, X.; Li, X.; Wei, X.; Wang, H. Hydrogen Sulfide Plays an Important Role in Diabetic Cardiomyopathy. *Front. Cell Dev. Biol.* 2021, 9, 627336. [CrossRef] [PubMed]

32. Liu, N.; Wu, J.; Zhang, L.; Gao, Z.; Sun, Y.; Yu, M.; Zhao, Y.; Dong, S.; Lu, F.; Zhang, W. Hydrogen Sulphide modulating mitochondrial morphology to promote mitophagy in endothelial cells under high-glucose and high-palmitate. *J. Cell. Mol. Med.* 2017, 21, 3190–3203. [CrossRef] [PubMed]

33. Qiu, X.; Liu, K.; Xiao, L.; Jin, S.; Dong, J.; Teng, X.; Guo, Q.; Chen, Y.; Wu, Y. Alpha-lipoic acid regulates the autophagy of vascular smooth muscle cells in diabetes by elevating hydrogen sulfide level. *Biochim. Biophys. Acta (BBA)—Mol. Basis Dis.* 2018, 1864, 3723–3738. [CrossRef] [PubMed]
34. Li, L.; Xiao, T.; Li, F.; Li, Y.; Zeng, O.; Liu, M.; Liang, B.; Li, Z.; Chu, C.; Yang, J. Hydrogen sulfide reduced renal tissue fibrosis by regulating autophagy in diabetic rats. *Mol. Med. Rep.* 2017, 16, 1715–1722. [CrossRef]
35. Yu, M.; Du, H.; Wang, B.; Chen, J.; Lu, F.; Peng, S.; Sun, Y.; Liu, N.; Sun, X.; Shiyun, D.; et al. Exogenous H$_2$S Induces Hrd1 S-sulfhydration and Prevents CD36 Translocation via VAMP3 Ubiquitlization in Diabetic Hearts. *Aging Dis.* 2020, 11, 286–300. [CrossRef]
36. Kar, S.; Shahshahan, H.R.; Kambis, T.N.; Yadav, S.K.; Li, Z.; Lefer, D.J.; Mishra, P.K. Hydrogen Sulfide Ameliorates Homocysteine-Induced Cardiac Remodeling and Dysfunction. *Front. Physiol.* 2019, 10, 988. [CrossRef]
37. Szabo, C.; Papapetropoulos, A. International Union of Basic and Clinical Pharmacology. CII: Pharmacological Modulation of H$_2$S Signaling Pathway in Type 2 Diabetic Rats. *Front. Physiol.* 2020, 11, 596. [CrossRef]
38. Wu, L.; Chen, Y.; Wang, C.-Y.; Tang, Y.-Y.; Huang, H.-L.; Kang, X.; Li, X.; Xie, Y.-R.; Tang, X.-Q. Hydrogen Sulfide Inhibits High Glucose-Induced Neuronal Senescence by Improving Autophagic Flux via up-regulation of SIRT1. *Mol. Neurosci.* 2019, 12, 194. [CrossRef] [PubMed]
39. Lu, S.; Wu, Y.; Chen, J.; Zhong, D.; Ma, S.; Yang, R. Hydrogen sulfide mitigates myocardial inflammation by inhibiting nucleotide-binding oligomerization domain-like receptor protein 3 inflammasome activation in diabetic rats. *Exp. Biol. Med.* 2020, 245, 221–230. [CrossRef]
40. Li, Y.; Li, L.; Zeng, O.; Liu, J.M.; Yang, J. H$_2$S improves renal fibrosis in STZ-induced diabetic rats by ameliorating TGF-β1 expression. *Ren. Fail.* 2016, 39, 265–272. [CrossRef]
41. Kar, S.; Shahshahan, H.R.; Hackfort, B.T.; Yadav, S.K.; Yadav, R.; Kambis, T.N.; Lefer, D.J.; Mishra, P.K. Exercise Training Promotes Cardiac Hydrogen Sulfide Biosynthesis and Mitigates Pyroptosis to Prevent High-Fat Diet-Induced Diabetic Cardiomyopathy. *Antioxidants* 2019, 11, 638. [CrossRef]
42. Li, J.; Yuan, Y.; Zhang, L.; Zhang, H.; Zhang, S.; Zhang, Y.; Xuan, X.-X.; Wang, M.-J.; Zhang, J.-Y. Exogenous hydrogen sulfide protects against high glucose-induced apoptosis and oxidative stress by inhibiting the STAT3/HIF-1α pathway in H9c2 cardiomyocytes. *Exp. Ther. Med.* 2019, 18, 3948–3958. [CrossRef]
43. Zhang, H.; Huang, Y.; Chen, S.; Tang, C.; Wang, G.; Du, J.; Jin, H. Hydrogen sulfide regulates insulin secretion and insulin resistance in diabetes mellitus, a new promising target for diabetes mellitus treatment? A review. *J. Adv. Res.* 2021, 27, 19–30. [CrossRef]
44. Chen, H.-J.; Ngowi, E.E.; Qian, L.; Li, T.; Qin, Y.-Z.; Zhou, J.-J.; Li, K.; Ji, X.-Y.; Wu, D.-D. Role of Hydrogen Sulfide in the Endocrine System. *Front. Endocrinol.* 2021, 12, 704620. [CrossRef] [PubMed]
45. Gheibi, S.; Jeddidi, S.; Kashfi, K.; Ghasemi, A. Effects of Hydrogen Sulfide on Carbohydrate Metabolism in Obese Type 2 Diabetic Rats. *Molecules* 2019, 24, 190. [CrossRef]
46. Luo, Z.-L.; Ren, J.-D.; Huang, Z.; Wang, T.; Xiang, K.; Cheng, L.; Tang, L.-J. The Role of Exogenous Hydrogen Sulfide in Free Fatty Acids Induced Inflammation in Macrophages. *Cell Physiol. Biochem.* 2017, 42, 1635–1644. [CrossRef] [PubMed]
47. Comas, F.; Moreno-Navarrete, J.M. The Impact of H$_2$S on Obesity-Associated Metabolic Disturbances. *Antioxidants* 2021, 10, 633. [CrossRef] [PubMed]
48. Suzuki, K.; Sagara, M.; Aoki, C.; Tanaka, S.; Aso, Y. Clinical Implication of Plasma Hydrogen Sulfide Levels in Japanese Patients with Type 2 Diabetes. *Intern. Med.* 2017, 56, 17–21. [CrossRef]
49. Zhou, Y.B.; Zhou, H.; Li, L.; Kang, Y.; Cao, X.; Wu, Z.Y.; Ding, L.; Sethi, G.; Bian, J.-S. Hydrogen sulfide prevents elastin loss and attenuates calcification induced by high glucose in smooth muscle cells through suppression of stat3/cathepsin s signaling pathway. *Int. J. Mol. Sci.* 2019, 20, 4202. [CrossRef]
50. John, A.M.S.P.; Kundu, S.; Pushpakumar, S.; Fordham, M.; Weber, G.; Mukhopadhyay, M.; Sen, U. GYY4137, a Hydrogen Sulfide Donor Modulates miR194-Dependent Collagen Realignment in Diabetic Kidney. *Sci. Rep.* 2017, 7, 879. [CrossRef]
51. Bitar, M.S.; Nader, J.; Al-Ali, W.; Al Madhoun, A.; Arefanian, H.; Al-Mulla, F. Hydrogen Sulfide Donor NaHS Improves Metabolism and Reduces Muscle Atrophy in Type 2 Diabetes: Implication for Understanding Sarcopenic Pathophysiology. *Oxidative Med. Cell. Longev.* 2018, 2018, 6825452. [CrossRef]
59. Ding, T.; Chen, W.; Li, J.; Ding, J.; Mei, X.; Hu, H. High Glucose Induces Mouse Mesangial Cell Overproliferation via Inhibition of Hydrogen Sulfide Synthesis in a TLR-4-Dependent Manner. Cell. Physiol. Biochem. 2017, 41, 1035–1043. [CrossRef]

60. Lia, Q.; Lancaster, J.R. Chemical Foundations of Hydrogen Sulfide Biology. Nitric Oxide. 2013. Available online: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3624763/pdf/nihms412728.pdf (accessed on 20 May 2022).

61. Cheng, Z.; Kishore, R. Potential role of hydrogen sulfide in diabetes-impaired angiogenesis and ischemic tissue repair. Redox Biol. 2020, 37, 101704. [CrossRef]

62. Roorda, M.; Miljkovic, J.L.; Van Goor, H.; Henning, R.H.; Bouma, H.R. Spatiotemporal regulation of hydrogen sulfide signaling in the kidney. Redox Biol. 2021, 43, 101961. [CrossRef]

63. Bulboaca, A.E.; Boarescu, P.-M.; Forfire, A.S.; Dogaru, G.; Barbalata, C.; Valeanu, M.; Munteanu, C.; Răjnoweanu, R.M.; Nicula, C.A.; Stanescu, I.C. The Effect of Nano-Epigallocatechin-Gallate on Oxidative Stress and Matrix Metalloproteinases in Experimental Diabetes Mellitus. Antioxidants 2020, 9, 172. [CrossRef] [PubMed]

64. Han, Y.; Shang, Q.; Yao, J.; Ji, Y. Hydrogen sulfide: A gaseous signaling molecule modulates tissue homeostasis: Implications in ophthalmic diseases. Cell Death Dis. 2019, 10, 293. [CrossRef] [PubMed]

65. Saito, J.; Bhavsar, P.; Zhang, Q.; Hui, C.; Menzies-Gow, A.; Fan Chung, K. Sputum and serum hydrogen sulfide (H2S) as novel biomarker of asthma. Clin. Transl. Allergy 2013, 3. Available online: http://www.ctajournal.com/content/3/1/P3 (accessed on 20 May 2022). [CrossRef]

66. Lu, M.; Zhao, F.-F.; Tang, J.-J.; Su, C.-J.; Fan, Y.; Ding, J.-H.; Bian, J.; Hu, G. The Neuroprotection of Hydrogen Sulfide Against MPTP-Induced Dopaminergic Neuron Degeneration Involves Uncoupling Protein 2 Rather Than ATP-Sensitive Potassium Channels. Antioxid. Redox Signal. 2017, 29, 849–859. [CrossRef]

67. Takahashi, H.; Asahina, R.; Fujioka, M.; Matsu, T.K.; Kato, S.; Mori, E.; Hioki, H.; Yamamoto, T.; Kobayashi, K.; Tsuboi, A. Ras-like Gem GT-Pase induced by Nps4 promotes activity-dependent neuronal tolerance for ischemic stroke. Proc. Natl. Acad. Sci. USA 2021, 118, e2018850118. [CrossRef]

68. Vela-Aneroa, A.; Hermida-Gómez, T.; Gato-Calvo, L.; Vaamonde-García, C.; Díaz-Prado, S.; Meijide-Falide, R.; Blanco, F.; Burguera, E. Long-term effects of hydrogen sulfide on the anabolic-catabolic balance of articular cartilage in vitro. Nitric Oxide 2017, 70, 42–50. [CrossRef]

69. Kumarsamy, A.; Kurian, G.A. Hydrogen Sulfide Promotes Proliferation of HT-29 Colon Cancer Cells in a Mitochondria-independent Pathway. Indian J. Pharm. Sci. 2019, 81, 456–463. [CrossRef]

70. Yang, G.; Tang, G.; Zhang, L.; Wu, L.; Wang, R. The Pathogenic Role of Cystathionine γ-Lyase/Hydrogen Sulfide in Streptozotocin-Induced Diabetes in Mice. Am. J. Pathol. 2011, 179, 869–879. [CrossRef]

71. Xue, R.; Hao, D.-D.; Sun, J.-P.; Li, W.-W.; Zhao, M.-M.; Li, X.-H.; Chen, Y.; Zhu, J.-H.; Liu, J.; et al. Hydrogen Sulfide Treatment Promotes Glucose Uptake by Increasing Insulin Receptor Sensitivity and Ameliorates Kidney Lesions in Type 2 Diabetics. Antioxid. Redox Signal. 2013, 19, 5–23. [CrossRef]

72. Ngowi, E.E.; Afzal, A.; Sarfraz, M.; Khattak, S.; Zaman, S.U.; Khan, N.H.; Li, T.; Jiang, Q.-Y.; Zhang, X.; Duan, S.-F.; et al. Role of hydrogen sulfide donors in cancer development and progression. Int. J. Biol. Sci. 2020, 16, 73–88. [CrossRef] [PubMed]

73. Potenza, M.A.; Sgarra, L.; Desantis, V.; Nacci, C.; Montagnani, M. Diabetes and alzheimer’s disease: Might mitochondrial dysfunction help deciphering the common path? Antioxidants 2021, 10, 1257. [CrossRef] [PubMed]

74. Szabo, C. Hydrogen Sulfide, an Endogenous Stimulator of Mitochondrial Function in Cancer Cells. Cells 2021, 10, 220. [CrossRef]

75. Monteiro, B.S.; Freire-Brito, L.; Carrageta, D.F.; Oliveira, P.F.; Alves, M.G. Mitochondrial Uncoupling Proteins (UCPs) as Key Modulators of ROS Homeostasis: A Crossstalk between Diabesity and Male Infertility? Antioxidants 2021, 10, 1746. [CrossRef] [PubMed]

76. Tabassum, R.; Jeong, N.Y. Potential for therapeutic use of hydrogen sulfide in oxidative stress-induced neurodegenerative diseases. Int. J. Med. Sci. 2019, 16, 1386–1396. [CrossRef]

77. Xu, X.; Yan, Q.; Liu, X.; Li, P.; Li, X.; Chen, Y.; Simoncini, T.; Liu, J.; ZHU, D.; Fu, X. 17β-Estradiol nongenomically induces vascular endothelial H2S release by promoting phosphorylation of cystathionine γ-lyase. J. Biol. Chem. 2019, 294, 15577–15592. [CrossRef]

78. Zhou, Y.; Wang, D.; Gao, X.; Lew, K.; Richards, A.M.; Wang, P. mTORC2 Phosphorylation of Akt1: A Possible Mechanism for Hydrogen Sulfide-Induced Cardioprotection. PLoS ONE 2014, 9, e99665. [CrossRef]

79. Lee, Z.-W.; Teo, X.-Y.; Song, Z.J.; Nin, D.S.; Novera, W.; Choo, B.A.; Dymock, B.W.; Moore, P.K.; Huang, R.Y.-J.; Deng, L.-W.; Intracellular Hyper-Acidification Potentiated by Hydrogen Sulfide Mediates Invasive and Therapy Resistant Cancer Cell Death. Front. Pharmacol. 2017, 8, 763. [CrossRef]

80. Chen, G.; Mao, Y.-G.; Chen, X.; Zhang, Y. Hydrogen sulfide therapy: A narrative overview of current research and possible therapeutic implications in future. Med. Gas Res. 2020, 10, 185–188. [CrossRef]

81. Chen, G.; Dou, Y.; Wang, Z. The role of hydrogen sulfide in stroke. Med. Gas Res. 2016, 6, 79–84. [CrossRef]

82. Huang, L. Molecular hydrogen: A therapeutic antioxidant and beyond. Med. Gas Res. 2016, 6, 219–222. [CrossRef] [PubMed]

83. Choi, K.-S.; Song, H.; Kim, E.-H.; Choi, J.-H.; Hong, H.; Han, Y.-M.; Hahn, K.-B. Inhibition of Hydrogen Sulfide-induced Angiogenesis and Inflammation in Vascular Endothelial Cells: Potential Mechanisms of Gastric Cancer Prevention by Korean Red Ginseng. J. Ginseng. Res. 2012, 36, 135–145. [CrossRef] [PubMed]

84. Paul, B.D.; Snyder, S.H. H2S signalling through protein sulphydrylation and beyond. Nat. Rev. Mol. Cell Biol. 2012, 13, 499–507. [CrossRef] [PubMed]
85. Stier-Jarmer, M.; Frisch, D.; Oberhauser, C.; Immich, G.; Kirschneck, M.; Schuh, A. Effects of single moor baths on physiological stress response and psychological state: A pilot study. Int. J. Biometeorol. 2017, 61, 1957–1964. [CrossRef]

86. Zhan, Y.; Li, M.Z.; Yang, L.; Feng, X.F.; Zhang, Q.X.; Zhang, N.; Zhao, Y-Y.; Zhao, H. An MRI study of neurovascular restorative after combination treatment with xiaoshuanenteric-coated capsule and enriched environment in rats after stroke. Front. Neurosci. 2019, 13, 701. [CrossRef]

87. Streeter, E.Y.; Badoer, E.; Woodman, O.L.; Hart, J.L. Effect of type 1 diabetes on the production and vasoactivity of hydrogen sulfide in rat middle cerebral arteries. Physiol. Rep. 2013, 1, e00111. [CrossRef]

88. Allen, C.L.; Wolanska, K.; Malhi, N.K.; Benest, A.V.; Wood, M.E.; Amaoku, W.; Torregrossa, R.; Whiteman, M.; Bates, D.O.; Whatmore, J.L. Hydrogen Sulfide Is a Novel Protector of the Retinal Glycocalyx and Endothelial Permeability Barrier. Front. Cell Dev. Biol. 2021, 9, 724905. [CrossRef]

89. Liu, Z.; Ran, Y.; Qie, S.; Gong, W.; Gao, F.; Ding, Z.; Xi, J. Melatonin protects against ischemic stroke by modulating microglia/macrophage polarization toward anti-inflammatory phenotype through STAT3 pathway. CNS Neurol. Ther. 2019, 25, 1353–1362. [CrossRef]

90. Predmore, B.L.; Lefer, D.J.; Gojon, G. Hydrogen Sulfide Biochemistry in and Medicine. Antioxid. Redox Signal. 2012, 17, 119–140. [CrossRef]

91. Sakuma, S.; Minamino, S.; Takase, M.; Ishiyama, Y.; Hosokura, H.; Kohda, T.; Ikeda, Y.; Fujimoto, Y. Hydrogen sulfide donor GYY4137 suppresses proliferation of human colorectal cancer Caco-2 cells by inducing both cell cycle arrest and cell death. Heliny 2019, 5, e02244. [CrossRef]

92. Narne, P.; Pandey, V.; Panthithi, P.B. Role of Nitric Oxide and Hydrogen Sulfide in Ischemic Stroke and the Emergent Epigenetic Underpinnings. Mol. Neurobiol. 2019, 56, 1749–1769. [CrossRef] [PubMed]

93. Dilek, N.; Papapetropoulos, A.; Toliver-Kinsky, T.; Szabo, C. Hydrogen sulfide: An endogenous regulator of the immune system. Pharmacol. Res. 2020, 161, 105119. [CrossRef] [PubMed]

94. Ortega, E.; Gález, I.; Hinchado, M.D.; Guerrero, J.; Martin-Cordero, L.; Torres-Piles, S. Anti-inflammatory effect as a mechanism of effectiveness underlying the clinical benefits of pelotherapy in osteoarthritics patients: Regulation of the altered inflammatory and stress feedback response. Int. J. Biometeorol. 2017, 61, 1777–1785. [CrossRef] [PubMed]

95. Zhang, Y.; Li, H.; Zhao, G.; Sun, A.; Zong, N.C.; Li, Z.; Zhu, H.; Zou, Y.; Yang, X.; Ge, J. Hydrogen Sulfide Attenuates the Recruitment of CD11b^Gr-1^ Myeloid Cells and Regulates Bax/Bcl-2 Signaling in Myocardial Ischemia Injury. Sci. Rep. 2014, 4, 4774. [CrossRef]

96. Merighi, S.; Gessi, S.; Varani, K.; Fozzi, D.; Borea, P.A. Hydrogen sulfide modulates the release of nitric oxide and VEGF in human keratinocytes. Pharmacol. Res. 2012, 66, 428–436. [CrossRef]

97. Wu, L.; Yang, W.; Jia, X.; Yang, G.; Durudanova, D.; Cao, K.; Wang, R. Pancreatic islet overproduction of H2S and suppressed insulin release in Zucker diabetic rats. Lab. Investig. 2009, 89, 59–67. [CrossRef]

98. Xue, X.; Ling, X.; Xi, W.; Wang, P.; Sun, J.; Yang, Q.; Xiao, J. Exogenous hydrogen sulfide reduces atrial remodeling and atrial fibrillation induced by diabetes mellitus via activation of the PI3K/Akt/eNOS pathway. Mol. Med. Rep. 2020, 22, 1759–1766. [CrossRef]

99. Malagrini, F.; Zuhra, K.; Mascolo, L.; Mastronica, D.; Vicente, J.B.; Forte, E.; Giuffrè, A. Hydrogen Sulfide Oxidation: Adaptive Changes in Mitochondria of SW480 Colorectal Cancer Cells upon Exposure to Hypoxia. Oxidative Med. Cell. Longev. 2019, 2019, 8102936. [CrossRef]

100. Giuffrè, A.; Vicente, J.B. Hydrogen Sulfide Biochemistry and Interplay with Other Gaseous Mediators in Mammalian Physiology. Oxidative Med. Cell. Longev. 2018, 2018, 6290931. [CrossRef]

101. Li, P.; Liu, H.; Shi, X.; Prokosch, V. Hydrogen sulfide: Novel endogenous and exogenous modulator of oxidative stress in retinal degeneration diseases. Molecules 2021, 26, 2411. [CrossRef]

102. Li, H.; Xu, F.; Gao, G.; Gao, X.; Wu, B.; Zheng, C.; Wang, P.; Li, Z.; Hua, H.; Li, D. Hydrogen sulfide and its donors: Novel antitumor and antimetastatic therapies for triple-negative breast cancer. Redox Biol. 2020, 34, 101564. [CrossRef] [PubMed]

103. Hellmich, M.R.; Szabo, C. Hydrogen Sulfide and Cancer. Handb. Exp. Pharmacol. 2015, 230, 233–241. [CrossRef] [PubMed]

104. Qian, L.; Shen, J.; Chuai, Y.; Cai, J. Hydrogen as a New Class of Radioprotective Agent. Int. J. Biol. Sci. 2013, 9, 887–894. [CrossRef] [PubMed]

105. Gao, S.-H.; Ho, J.Y.; Fan, L.; Richardson, D.J.; Yuan, Z.; Bond, P.L. Antimicrobial Effects of Free Nitrous Acid on Desulfovibrio vulgaris: Implications for Sulfide-Induced Corrosion of Concrete. Appl. Environ. Microbiol. 2016, 82, 5563–5575. [CrossRef]

106. Shan, H.; Qiu, J.; Chang, P.; Chu, Y.; Gao, C.; Wang, H.; Chen, G.; Luo, C.; Wang, T.; Chen, X.; et al. Exogenous Hydrogen Sulfide Offers Neuroprotection on Intracerebral Hemorrhage Injury Through Modulating Endogenous H2S Metabolism in Mice. Front. Cell. Neurosci. 2019, 13, 349. [CrossRef] [PubMed]

107. Scammahorn, J.J.; Nguyen, I.T.N.; Bos, E.M.; Van Goor, H.; Joles, J.A. Fighting Oxidative Stress with Sulfur: Hydrogen Sulfide in the Renal and Cardiovascular Systems. Antioxidants 2021, 10, 373. [CrossRef]

108. Xie, L.; Gu, Y.; Wen, M.; Zhao, S.; Wang, W.; Ma, Y.; Meng, G.; Han, Y.; Wang, Y.; Liu, G.; et al. Hydrogen Sulfide Induces Keap1-S-sulfhydration and Suppresses Diabetes-Accelerated Atherosclerosis via Nrfr2 Activation. Diabetes 2016, 65, 3171–3184. [CrossRef]
109. Wu, D.; Luo, N.; Wang, L.; Zhao, Z.; Bu, H.; Xu, G.; Yan, Y.; Che, X.; Jiao, Z.; Zhao, T.; et al. Hydrogen sulfide ameliorates chronic renal failure in rats by inhibiting apoptosis and inflammation through ROS/MAPK and NF-κB signaling pathways. *Sci. Rep.* 2017, 7, 455. [CrossRef]

110. Ji, J.; Xiang, P.; Li, T.; Lan, L.; Xu, X.; Lu, G.; Ji, H.; Zhang, Y.; Li, Y. NOSH-NBP, a Novel Nitric Oxide and Hydrogen Sulfide-Releasing Hybrid, Attenuates Ischemic Stroke-Induced Neuroinflammatory Injury by Modulating Microglia Polarization. *Front. Cell. Neurosci.* 2017, 11, 154. [CrossRef]

111. Li, H.; Zhang, C.; Sun, W.; Li, L.; Wu, B.; Bai, S.; Li, H.; Zhong, X.; Wang, R.; Wu, L.; et al. Exogenous hydrogen sulfide restores cardioprotection of ischemic post-conditioning via inhibition of mPTP opening in the aging cardiomycocytes. *Cell Bioi.* 2015, 5, 43. [CrossRef]

112. Xuan, A.; Long, D.; Li, J.; Ji, W.; Zhang, M.; Hong, L.; Liu, J. Hydrogen sulfide attenuates spatial memory impairment and hippocampal neuroinflammation in beta-amyloid rat model of Alzheimer’s disease. *J. Neuroinflamm.* 2012, 9, 202. [CrossRef] [PubMed]

113. Fan, Y.-Y.; Shen, Z.; He, P.; Jiang, L.; Hou, W.-W.; Shen, Y.; Zhang, X.-N.; Chen, Z. A Novel Neuroprotective Strategy for Ischemic Stroke: Transient Mild Acidosis Treatment by CO₂ Inhalation at Reperfusion. *J. Cereb. Blood Flow Metab.* 2014, 34, 275–283. [CrossRef] [PubMed]

114. Beltowski, J.; WoJCicka, G.; Jamroz-Wiśniewska, A. Hydrogen sulfide in the regulation of insulin secretion and insulin sensitivity: Implications for the pathogenesis and treatment of diabetes mellitus. *Biochem. Pharmacol.* 2018, 149, 60–76. [CrossRef] [PubMed]

115. Chandrasekarar, P.; Ravindran, S.; Boovaran, S.R.; Kurian, G.A. Hydrogen sulfide-mediated cardioprotection against ischemia reperfusion is linked to KATP channel for mitochondrial preservation but not for its distinct preference on interfibrillar mitochondria. *Bangladesh J. Pharm.* 2019, 14, 107–115. [CrossRef]

116. Kamat, P.K.; Kyles, P.; Kalani, A.; Tyagi, N. Hydrogen Sulfide Ameliorates Homocysteine-Induced Alzheimer’s Disease-Like Pathology, Blood-Brain Barrier Disruption, and Synaptic Disorder. *Mol. Neurobiol.* 2016, 53, 2451–2467. [CrossRef] [PubMed]

117. Borlongan, C.V.; Gonzales-Portillo, B.; Lippert, T.; Nguyen, H.; Lee, J.-Y. Hyperbaric oxygen therapy: A new look on treating stroke and traumatic brain injury. *Brain Circ.* 2019, 5, 101–105. [CrossRef]

118. Durante, W. Hydrogen Sulfide Therapy in Diabetes-Accelerated Atherosclerosis: A Whiff of Success. *Diabetes 2016, 65, 2832–2834. [CrossRef]

119. Bredthauer, A.; Lehle, K.; Scheuerle, A.; Schelzig, H.; McCook, O.; Radermacher, P.; Szabo, C.; Wepler, M.; Simon, F. Intravenous hydrogen sulfide does not induce neuroprotection after aortic balloon occlusion-induced spinal cord ischemia/reperfusion injury in a human-like porcine model of ubiquitous arteriosclerosis. *Intensive Care Med.* 2018, 46, 44. [CrossRef] [PubMed]

120. Cheng, Z.; Shen, X.; Jiang, X.; Shan, H.; Cimini, M.; Fang, P.; Ji, Y.; Park, J.Y.; Drosatos, K.; Yang, X.; et al. Hyperhomocysteinemia potentiates diabetes-impaired EDHF-induced vascular relaxation: Role of insufficient hydrogen sulfide. *Redox Biol.* 2018, 16, 215–225. [CrossRef]

121. Wang, R. Physiological Implications of Hydrogen Sulfide: A Whiff Exploration That Blossomed. *Physiol. Rev.* 2012, 92, 791–896. [CrossRef]

122. Lambooy, S.P.H.; Bidaddkosh, A.; Nakladal, D.; Van Buiten, A.; Girgis, R.A.T.; Van Der Graaf, A.C.; Wiedenmann, T.J.; Koster, R.A.; Vogelaar, P.; Biukema, H.; et al. The Novel Compound Sul-1 Preserves Endothelial Function and Inhibits Progression of Kidney Damage in Type 2 Diabetes Mellitus in Mice. *Sci. Rep.* 2017, 7, 11165. [CrossRef] [PubMed]

123. Dogaru, G.; Bulboaca, A.; Boarescu, P.M.; Ciumarnean, L.; Rus, V.; Sitar-Taut, A.-V.; Munteanu, C.; Bodisz, G.; Stanescu, I. The Effect of Mofettes on Oxidative Stress/Antioxidant Balance in Experimental Myocardial Ischemia. *In Vivo* 2019, 33, 1911–1920. [CrossRef] [PubMed]

124. Gambari, L.; Grigolo, B.; Filardo, G.; Grassi, F. Sulfurous thermal waters stimulate the osteogenic differentiation of human mesenchymal stromal cells—An in vitro study. *Biomed. Pharmacother.* 2020, 129, 110344. [CrossRef] [PubMed]

125. Li, L.; Liu, Y.; Wang, Q.; Wang, Z.; Cui, L.; Xu, Y.; Guan, K. Levels of nasal exhaled hydrogen sulfide in the general population and allergic rhinitis patients. *J. Clin. Lab. Anal.* 2021, 35, e23678. [CrossRef] [PubMed]

126. Carubbi, C.; Gobbi, G.; Bucci, G.; Gesi, M.; Vitale, M.; Mirandola, P. Skin, Inflammation and Sulfurous Waters: What is Known, What is Believed. *Eur. J. Inflamm.* 2013, 11, 591–599. [CrossRef]

127. Hoteteu, M.; Romanian Association of Balneology; Munteanu, C.; Ionescu, E.V.; Almăsian, R.E. Techirghiol Balnear and Rehabilitation Sanatorium Bioactive substances of the Techirghiol therapeutic mud. *Balneo Res. J.* 2018, 9, 5–10. [CrossRef]

128. Carbajo, J.M.; Maraver, F. Sulphurous Mineral Waters: New Applications for Health. Evidence-Based Complement. *Altern. Med. 2017, 2017, 68034084. [CrossRef]

129. Viegas, J.; Esteves, A.F.; Cardoso, E.M.; Arosa, F.A.; Vitale, M.; Taborda-Barata, L. Biological Effects of Thermal Water-Associated Hydrogen Sulfide on Human Airways and Associated Immune Cells: Implications for Respiratory Diseases. *Front. Public Health 2019, 7, 128. [CrossRef]

130. Kida, K.; Yamada, M.; Tokuda, K.; Marutani, E.; Kakinohana, M.; Kaneki, M.; Ichinohe, F. Inhaled Hydrogen Sulfide Prevents Neurodegeneration and Movement Disorder in a Mouse Model of Parkinson’s Disease. *Antioxid. Redox Signal.* 2015, 11, 343–352. [CrossRef]

131. Kloesch, B.; Liszt, M.; Krehan, D.; Broell, J.; Kiener, H.; Steiner, G. High concentrations of hydrogen sulphide elevate the expression of a series of pro-inflammatory genes in fibroblast-like synoviocytes derived from rheumatoid and osteoarthritis patients. *Immunol. Lett.* 2011, 141, 197–203. [CrossRef]
132. Zhang, L.; Wang, Y.; Li, Y.; Li, L.; Xu, S.; Feng, X.; Liu, S. Hydrogen Sulfide (H$_2$S)-Releasing Compounds: Therapeutic Potential in Cardiovascular Diseases. *Front. Pharmacol.* 2018, 9, 1066. [CrossRef] [PubMed]

133. Sanderson, T.H.; Wider, J.M.; Lee, I.; Reynolds, C.A.; Liu, J.; Lepore, B.; Tousignant, R.; Bukowski, M.J.; Johnston, H.; Fite, A.; et al. Inhibitory modulation of cytochrome c oxidase activity with specific near-infrared light wavelengths attenuates brain ischemia/reperfusion injury. *Sci. Rep.* 2018, 8, 3481. [CrossRef] [PubMed]

134. Mirandola, P.; Gobbi, G.; Micheloni, C.; Vaccarezza, M.; Di Marcantonio, D.; Ruscitti, F.; De Panfilis, G.; Vitale, M. Hydrogen sulfide inhibits IL-8 expression in human keratinocytes via MAP kinase signaling. *Lab. Investig.* 2011, 91, 1188–1194. [CrossRef] [PubMed]

135. Kashfi, K.; Chattopadhyay, M.; Kodela, R. NOSH-sulindac (AVT-18A) is a novel nitric oxide- and hydrogen sulfide-releasing hybrid that is gastrointestinal safe and has potent anti-inflammatory, analgesic, antipyretic, anti-platelet, and anti-cancer properties. *Redox Biol.* 2015, 6, 287–296. [CrossRef]

136. Agency for Toxic Substances and Disease Registry (ATSDR). Toxicological Profile for Hydrogen Sulfide and Carbonyl Sulfide; U.S. Department of Health and Human Services, Public Health Service: Atlanta, GA, USA, 2016. Available online: https://www.atsdr.cdc.gov/toxprofiles/tp114.pdf (accessed on 20 May 2022).

137. Jia, J.; Xiao, Y.; Wang, W.; Qing, L.; Xu, Y.; Song, H.; Zhen, X.; Ao, G.; Alkayed, N.J.; Cheng, J. Differential mechanisms underlying neuroprotection of hydrogen sulfide donors against oxidative stress. *Neurochem. Int.* 2013, 62, 1072–1078. [CrossRef]

138. Huang, G.; Diao, J.; Yi, H.; Xu, L.; Xu, J.; Xu, W. Signaling pathways involved in HSP32 induction by hyperbaric oxygen in rat spinal neurons. *Redox Biol.* 2016, 10, 108–118. [CrossRef]

139. Liu, M.; Wu, L.; Montaut, S.; Yang, G. Hydrogen Sulfide Signaling Axis as a Target for Prostate Cancer Therapeutics. *Prostate Cancer* 2016, 2016, 8108549. [CrossRef]

140. Stoica, S.I.; Bleotu, C.; Ciobanu, V.; Ionescu, A.M.; Albadi, I.; Onose, G.; Munteanu, C. Considerations about Hypoxic Changes in Neuraxis Tissue Injuries and Recovery. *Biomedicines* 2022, 10, 481. [CrossRef]

141. Onose, G.; Anghelescu, A.; Blendea, D.; Ciobanu, V.; Daia, C.; Firan, F.C.; Oprea, M.; Spinu, A.; Popescu, C.; Ionescu, A.; et al. Cellular and Molecular Targets for Non-Invasive, Non-Pharmacological Therapeutic/Rehabilitative Interventions in Acute Ischemic Stroke. *Int. J. Mol. Sci.* 2022, 23, 907. [CrossRef]

142. Bithi, N.; Link, C.; Henderson, Y.O.; Kim, S.; Yang, J.; Li, L.; Wang, R.; Willard, B.; Hine, C. Dietary restriction transforms the mammalian protein persulfidome in a tissue-specific and cystathionine γ-lyase-dependent manner. *Nat. Commun.* 2021, 12, 1745. [CrossRef]

143. Wang, W.-J.; Cai, G.-Y.; Ning, Y.-C.; Cui, J.; Hong, Q.; Bai, X.-Y.; Xu, X.-M.; Bu, R.; Sun, X.-F.; Chen, X.-M. Hydrogen sulfide mediates the protection of dietary restriction against renal senescence in aged F344 rats. *Sci. Rep.* 2016, 6, 30292. [CrossRef] [PubMed]

144. Renieris, G.; Katrini, K.; Damoulari, C.; Akinosoglou, K.; Psarrakis, C.; Kyriakopoulou, M.; Dimopoulos, G.; Lada, M.; Koufargyris, P.; Giamarellos-Bourboulis, E.J. Serum Hydrogen Sulfide and Outcome Association in Pneumonia by the SARS-CoV-2 Coronavirus. *Shock* 2020, 54, 633–763. [CrossRef] [PubMed]

145. Citi, V.; Martelli, A.; Brancalone, V.; Brogi, S.; Gojen, G.; Montanaro, R.; Morales, G.; Testai, L.; Calderone, V. Anti-inflammatory and antiviral roles of hydrogen sulfide: Rationale for considering H$_2$S donors in COVID-19 therapy. *Br. J. Pharmacol.* 2020, 177, 4931–4941. [CrossRef] [PubMed]

146. Evgen’ev, M.B.; Frenkel, A. Possible application of H2S-producing compounds in therapy of coronavirus (COVID-19) infection and pneumonia. *Cell Stress Chaperones* 2020, 25, 713–715. [CrossRef]

147. Datzmann, T.; Merz, T.; McCook, O.; Szabo, C.; Radermacher, P. H$_2$S as a Therapeutic Adjuvant Against COVID-19: Why and How? *Shock* 2021, 56, 865–867. [CrossRef]