Arabis watsonii (P.H.Davis) F.K.Mey.: An overlooked cruciferous species from eastern Anatolia and its phylogenetic position

Barış Özüdoğru¹, Mehmet Fırat²

¹ Department of Biology, Faculty of Science, Hacettepe University, Ankara Turkey ² Department of Biology, Faculty of Education, Yüzüncü Yıl University, Van, Turkey

Corresponding author: Barış Özüdoğru (barisoz@hacettepe.edu.tr)

Academic editor: K. Marhold | Received 20 September 2016 | Accepted 19 November 2016 | Published 1 December 2016

Citation: Özüdoğru B, Fırat M (2016) Arabis watsonii (P.H.Davis) F.K.Mey.: An overlooked cruciferous species from eastern Anatolia and its phylogenetic position. PhytoKeys 75: 57–68. doi: 10.3897/phytokeys.75.10568

Abstract

Arabis watsonii (P.H.Davis) F.K.Mey. was initially reported as Thlaspi watsonii P.H.Davis in Flora of Turkey. Although F.K.Meyer transferred this species to Arabis L., this species has been overlooked and treated as Thlaspi L. in relevant literature for Flora of Turkey. In this study this species was evaluated using molecular (nuclear ITS and plastidic trnL-F sequences) and morphological data. Results clearly show that A. watsonii is sister to the A. hirsuta aggregate and its relatives. In conclusion, our results increased the number of known Arabis species in Turkey to 23. Furthermore, detailed description and distribution of the species are given and a new IUCN threat category for A. watsonii is proposed.

Keywords

Arabis watsonii, A. hirsuta aggregate, Arabideae, Brassicaceae, phylogeny

Introduction

The genus Arabis L. is represented by ca. 60 species and distributed in temperate regions of the northern hemisphere (Koch et al. 2010, Al-Shehbaz 2012). Although Al-Shehbaz (1988) earlier delimited the genus with ca. 180 species, molecular phylogenetical studies of Arabis and its relatives clearly show that Arabis s.l. is polyphyletic...
(Koch et al. 1999, 2000, 2001, 2010, Al-Shehbaz et al. 2011) and subsequently some of the highly supported phylogenetic lineages were described as separate genera, e.g. Scapiarabis M.A. Koch, R. Karl, D. German & Al-Shehbaz, Acistrostrum Y.Z. Zhao, and Sinoarabis R. Karl, D. German, M.A. Koch & Al-Shehbaz (Karl et al. 2012). Nevertheless, several highly supported Arabis clades (including main Arabis clade, A. alpina L. clade, A. auberchi Boiss. clade etc.) were also described (Karl and Koch 2013).

After the first revision of J. Cullen (1965) who reported 17 Arabis species in Flora of Turkey, five new species (A. lycia Parolly & P.Hein, A. alanyensis H.Duman, A. davisi H.Duman &A.Duran, A. erikii Mutlu and A. kaynakiae Daşkıın) and two new records for Turkey (A. allionii DC. and A. mollis Steven) were added (Davis et al. 1988, Parolly and Hein 2000, Duman 2001, Duman and Duran 2001, Mutlu and Dönmez 2003, Mutlu 2004, Daşkıın 2013). In addition, A. graellsiiiformis Hedge was treated as a subspecies of A. mollis Steven (Mutlu and Erik 2012) and Arabis turrita L. transferred to Pseudoturritis Al-Shehbaz (Al-Shehbaz 2005). Finally, the genus Arabis is currently represented by 22 species (24 taxa) in Turkey (Mutlu and Erik 2015).

Arabis watsonii (P.H.Davis) F.K.Mey. (Fig. 1) was initially published as Thlaspi watsonii P. H. Davis in Flora of Turkey by P. H. Davis. Davis argued that this taxon is closely related to the pink flowered Thlaspi lilacinum Boiss & Huet (Callothlaspi lilacinum (Boiss & Huet) F.K.Mey or Noccaea lilacina (Boiss. & A. Huet) Al-Shehbaz, depending on the authors and treatments). F.K. Meyer investigated type specimen of T. watsoni while working on his Callothlaspi account (Meyer 2006), and he transferred this species to Arabis since due to the presence of siliques (rather than silicles which are typical for Thlaspi) and leaves with bifid hairs. Ever since this taxon either has been overlooked (Al-Shehbaz et al. 2007, Özhatay et al. 2009, 2011, Mutlu and Erik 2015) or still treated as Thlaspi L. (Güner et al. 2014).

To verify Meyers’s treatment of Arabis watsonii, plant material was collected from type locality and nearby areas. Collected specimens were evaluated morphologically and molecularly to analyse 1) is A. watsonii a member of Arabis and 2), which main clade does it belong? We carried out morphological and molecular phylogenetic studies of A. watsonii plus representatives of Arabis and other Arabideae and combined this data with climatic and biogeographic data.

**Methods**

**Sampling**

The present study includes for the first time sequences of nuclear ribosomal ITS1, ITS2 and 5.8 S rRNA (hereafter ITS) and trnL(UAA) intron/trnL-trnF intergenic spacer sequence data (hereafter trnL-F) for Arabis watsonii (Voucher: M. Fırat 32513 at HUB). All other sequences of the 88 Arabis and other Arabideae species were taken from ITS and trnL-F data sets of Karl and Koch (2013). In addition, sequences of the related A. hirsuta aggregate (A. stelleri DC. and A. takesimana Nakai) were taken from
Arabis watsonii (P.H.Davis) F.K.Mey.: An overlooked cruciferous species...

Figure 1. Photographs of *Arabis watsonii*, **A** habit (in flower) **B** habit (in fruit), **C** basal leaves **D** stem leaves **E** trichomes on leaves margins **F** flowers **G** fruits **H** habitat of Bahcesaray population.

GenBank (http://www.ncbi.nlm.nih.gov/genbank) and incorporated into the above mentioned data sets. To determine the phylogenetic placement of *A. watsonii*, we reduced the above mentioned data as follows: All *Arabis* species were added along with
two species from *Aubrietia* Adans., two *Draba* L. species and one species each from other small genera including *Sinoarabis*, *Arcyosperma* O.E. Schulz, *Baimashania* Al-Shehbaz, *Acstrostrum*, *Botschantzevia* Nabiev, *Dendoarabis* (C.A. Mey.) D. German & Al-Shehbaz, *Pachyneurum* Bunge, *Pseudodraba* Al-Shehbaz, D. German & M. Koch, *Scapiarabis*, *Tomostima* Raf. *Pseudoturritis turrita* (L.) Al-Shehbaz was used as the out-group. Genbank accessions of species included in this study are at terminal nodes of phylogenetic trees (Figs 3, 4).

50 specimens belonging to five populations were used for extending description of *A. watsonii*. The vouchers were deposited at Hacettepe University Herbarium (HUB) and private herbarium of M. Fırat.

**DNA extraction, amplification and sequencing**

Total genomic DNA was isolated using DNeasy Plant Mini Kit (Qiagen, Hilden, Germany) following the manufacturer’s instructions. ITS and trnL-F regions were amplified using primers ITS1 and ITS 4 (White et al. 1990) and C and F of Taberlet et al. (1991), respectively. Amplification of ITS and trnL-F followed the protocol in Warwick et al. (2004) and Ansell et al. (2007), respectively. Purification and sequencing were performed by BIOEKSEN (İstanbul, Turkey).

**Data analysis**

Firstly, to determine whether *A. watsonii* belongs to tribe Arabideae, the phylogenetic tool in Brassibase (Koch et al. 2012; Kiefer et al. 2013) was used. ITS and trnL-F sequences data were edited with Codon Code Aligner (CodonCode Corporation) and directly incorporate into the alignment files of Karl and Koch (2013). Both data sets were analysed using a Bayesian approach as implemented in the software BEAST ver. 1.8 (Drummond et al. 2012).

Sequence evolution models were selected by the Akaike information criterion (AIC) implemented in MEGA v.6 (Tamura et al. 2013).

The GTR + G + I model was selected for ITS and GTR + I for trnL-F and a Yule process of speciation was used as the tree prior. Two independent Markov Chain Monte Carlo (MCMC) runs for each data set were conducted with 10 million generations and sampled every 1000 generations. Each run was checked using TRACER v1.6 (http://beast.bio.ed.ac.uk/Tracer) and then log and trees files were combined in LOGCOMBINER (Drummond et al. 2012).

A total of 20000 trees were obtained and 10% (2000) of these were discarded as burn-in. 18000 post-burn-in trees were used in the program TREEANNOTATER v.1.7.5 to obtain a single posterior probability and maximum clade credibility tree as visualized using FIGTREE v1.3.1.
Morphology, distribution and conservation

To redescribe morphological features of *Arabis watsonii* ca. 50 individuals from five populations were investigated. In order to evaluate the IUCN threat category of *Arabis watsonii*, occurrence data were obtained from both field and Yüzüncü Yıl University, Science faculty herbarium (VANF) and then total distribution area was calculated using DIVA-GIS (version 7.5.0, http://www.diva-gis.org). Some specimens that had no GPS coordinates were georeferenced using Google Earth 7.1. (http://www.google.com/earth) according to common names. To evaluate climatic requirements of *Arabis watsonii* and closely related taxa, bioclimatic data were taken from the WorldClim–Global Climate Database (http://www.worldclim.org) at a spatial resolution of 30 s.

Results

Morphology, distribution and conservation

*Arabis watsonii* (P.H.Davis) F.K.Mey (in Meyer 2006 p.187)

Basionym: *Thlaspi watsonii* P.H.Davis (in Davis et al. 1988 p. 235)

**Type.** Turkey B9 Van: Çuh pass, Halanduran Da. and Güzeldere Tepe, dry stony slopes, locally common, 2800 m, flowers white, vi 1966, Albury, Cheese & Watson 1438 (holo. K photo !).

Slender perennial herb. Stem erect, nearly glabrose, 13–30 cm high. Basal leaves up to 30 mm, oblong – obovate, petiolate, leaves on sterile shoots completely covered by branched trichomes, fertile shoot with branched trichomes on the leaves margin. Stem leaves narrowly oblong, very shortly auriculate, tapering to subacute apex, 5–32 × 2.5–10 mm with marginal trichomes. Sepals white-purplish, inner sepals saccate, 3–3.2 × 1–1.8 mm, outer sepals smooth, 2.7–3 × 1–1.5 mm. Petals white, 5.5–7 mm long, 1.5 mm broad above, tapering below into 1.5–2 mm claw. Stamens 6, long filaments 4, 3–4 mm, short filaments 2, 2.5 – 3 mm long, anther yellow, 0.8 – 1 × 0.3–0.4 mm. Pedicel up to 7–8 mm in fruit. Fruit ± constricted between seeds, 4–14 × 0.8–1.2 mm, with 6–8 seeds, style ca. 1 mm. Seeds brown, ovate-oblong, 0.9–1.1 × 0.5–0.7 mm.

Fl. and fr : 4–6. Alpine damp places, dry stony slopes, steppe. 1980–2800 m.

**Distribution:** Endemic. Irano – Turanian element (Fig. 2)

**Specimens examined. TURKEY. B9 Van:** Gürpınar district, from Güzeldere pass to Çuğ pass, damp places, 2495 m, 38°09'57"N, 43°57'47"E, 19 May 2015, M. Fırat 32513 (herb. M. Fırat); ibid M. Fırat 32572 (HUB, herb. M. Fırat) 11 June 2015; Bahçesaray district, from Ulubesli village to Hızan, damp steppe, 2265 m, 38°07'46"N, 42°40'53"E, 19 May 2014, M. Fırat 30870 (HUB, herb. M. Fırat); ibid M. Fırat 30989 (HUB, herb. M. Fırat) 21 June 2014; Bahçesaray, between Liçan Village and
Kavuşşahap Mountains (Deve Mountain), humid steppe, 1980 m, 04.04.1999. M. Fırat 1077 (herb. M. Fırat); Bahçesaray, Altındere Village Kavuşşahap Mountains, damp places, 2300 m, 17.05.2000, M. Fırat 2122 (herb. M. Fırat); Bahçesaray, between Cevizlibelen Village and Arnos Mountain, humid steppe, 2400 m, 23.06.2000, M. Fırat 2729 (herb. M. Fırat); between Güzelsu (Hoşap) and Başkale, Güzeldere pass, meadows, 2550–2650 m, 10.06.2001, M. Armağan 1073 (VANF); between Güzelsu (Hoşap) - Başkale, Güzeldere Pass, Güzeldere Hill, moist meadows, slopes, steppe, 2700–2800 m, 07.06.2002, M. Armağan 2423 (VANF); between Güzelsu (Hoşap) and Başkale, Güzeldere pass, from Güzeldere gendarmerie station to Başkale, slopes, steppe and moist meadows, 2600–2730 m, 19.05.2001, M. Armağan 1130 (VANF).

**Vernacular name.** In Van province, indigenous people use name ‘Nançûk’ for *Arabis* species (Fırat 2013).

Field observations and records taken from relevant herbaria indicate that *A. watsonii* has two distinct populations (Fig. 2). A large population growing around Bahçesaray district and a second population occurring in the Gürpinar district especially around the Güzeldere pass. Therefore total distribution areas for these populations were estimated separately. The area around Bahçesaray was calculated as 108.99 km² and the second area around Gürpinaras 2.69 km². In summary, to propose IUCN threat categories of *A. watsonii*, these two population groups and calculated areas were considered. The occupancy area (AOO) of *A. watsonii* was calculated as 111.68 km² in which about 1000 individuals in each population were estimated to occur. Overgrazing and reaping activities by the local people were observed in field studies. Therefore, in accordance with the
Figure 3. ITS-based phylogenetic backbone of Arabideae that focuses on the placement of *Arabis watsonii*. Shown is the Bayesian maximum clade credibility tree with posterior probability values > 0.5. Highlighted part of the tree, (magnified on the left), is the *Arabis hirsuta* aggregate and its relatives. Color codes: Green = *A. hirsuta* aggregate, red=the closest relatives of the *A. hirsuta* aggregate, blue = *A. watsonii*. Clade names follow Karl and Koch (2013).

criteria of the IUCN (2016), *A. watsonii* is assessed here as “Vulnerable” ([VU] (B2a, C2a(i))], because distribution area of the species is severely fragmented and the species is currently known from no more than ten localities occupying less than 2,000 km²,( although it was considered “Endangered” (EN) according to Ekim et al. (2000)).

The basic climatic requirements of *A. watsonii*, annual main temperature and annual precipitation were calculated as 5.7 °C and 583 ml respectively.

Phylogeny. The aligned ITS and trnL-F data matrices included 91 species. The ITS data set was 642 bp, of which 236 were variable and 168 parsimony informative, whereas the trnL-F data set incorporates 855 bp, of which 181 were variable and 108 parsimony informative.
Figure 4. trnL-F-based phylogenetic backbone of Arabideae that focuses on the placement of *Arabis watsonii*. Shown is the Bayesian maximum clade credibility tree with posterior probability values > 0.5. Highlighted part of the tree, (magnified on the left), is the *Arabis hirsuta* aggregate and its relatives. Color codes: Green = *A. hirsuta* aggregate, red=the closest relatives of the *A. hirsuta* aggregate, blue = *A. watsonii*. Clade names follow Karl and Koch (2013).

The query of ITS sequences of *A. watsonii* in BrassiBase (version 1.1) supported its phylogenetic placement within tribe Arabideae and clearly matching *Arabis*. The outcome of Bayesian phylogenetic analyses using ITS and trnL-F data sets (Figs 3, 4) were congruent with each other in regard to the placement of *A. watsonii*. In both analyses *A. watsonii* falls into the main *Arabis* clade as sister to *A. hirsuta* (L.) Scop. aggregate and its relatives outlined in Karl and Koch (2014). Whereas *A. watsonii* forms a monophyletic lineage with *A. cretica* (Bayesian posterior probability (pp) = 0.98) in
ITS analysis (Fig. 3), this sister relationship was not supported by trnL-F analysis (Fig. 4) and chloroplast data shows that *A. watsonii*, the *A. hirsuta* aggregate, its relatives plus non-European *Arabis* species are linked to this aggregate (including *A. georgiana* R.M.Harper, *A. pycnocarpa* M.Hopkins and *A. borealis* DC.) forming a monophyletic clade (pp=1.00).

**Discussion**

In this study, we used evidence from nuclear ITS and plastidic trnL-F sequences to determine the phylogenetic and taxonomic position of the overlooked Anatolian endemic *A. watsonii*. In addition, morphological and climatic data were used to better understand ecological and evolutionary relationships of *A. watsonii* with representatives of the well-defined *A. hirsuta* aggregate and its relatives.

The differences in the phylogenetic placement of *A. watsonii* in relation to its sister position to *A. cretica*, according to ITS and trnL-F analyses, indicates possible chromosome capture/ancient hybridization. These processes are well known in the *A. hirsuta* aggregate and its relatives (Koch et al. 2010, Karl and Koch 2014).

A recent study of *A. hirsuta* aggregate recognised eight European species including *A. hirsuta* (L.) Scop., *A. sagittata* (Bertol.) DC., *A. planisiliqua* (Pers.) Rchb., *A. nemorensis* (Wolf ex Hoffm.) W.D.J. Koch, *A. allionii* DC., *A. sudetica* Tausch, *A. sadina* (Samp.) Coutinho, and *A. juressi* Rothm. The historical definition and circumscription of such an aggregate depends on different authors (see Karl and Koch (2014)) and because relationships between the *A. hirsuta* aggregate and its European/non-European relatives have already been discussed in detail before, we will not repeat this discussion here.

As indicated above, the ITS phylogeny supports a clear monophyly between *A. watsonii* and Greece endemic *A. cretica* Boiss. & Heldr. Whereas this relationship does not supported by trnL-F, both species seems to be at a basal position for *A. hirsuta* aggregate and its relatives. This results is somewhat expected because the Western Irano-Turanian and the East Mediterranean regions have already been suggested as diversity centres for almost all Arabideae clades (Jordon-Thaden et al. 2010, Karl and Koch 2013).

Morphologically *A. watsonii* is similar to *A. hirsuta* and *A. sagittata*, therefore specimens were treated under these names in some herbaria. However *A. watsonii* differs from *A. hirsuta* and *A. sagittata* in having glabrous stems and relatively large petals. Meyer (2006) argued that *A. watsonii* is related to *A. abietina* Bornm. from the Ilgaz Mountain (Turkey) since this species is also characterized by glabrous stems and a similar petal length. The latter species has been treated as a synonym of *A. suedica* in Karl and Koch (2014) based on Jalas and Souminen (1994). Despite the relatively large geographic gap between *A. abietina* and *A. suedica*, ITS sequences of the taxa were identical and trnL-F sequences differ only in one single nucleotide position. Thus, *A. abietina* was not included in the current study, although it is treated as a valid species according to the Flora of Turkey (Cullen 1965) and the actual check-list (Güner et al. 2012). Apart from geographic isolation, branched trichomes on leaf margins and rela-
tively small fruits are the main diagnostic characters distinguishing *A. watsonii* from both *A. abietina* and *A. allionii*, which are also members of the *A. hirsuta* aggregate. In summary, more comprehensive studies are needed to clarify the validity of *A. abietina*. With the proper assignment of *A. watsonii* in the current study, the total number of the Turkish *Arabis* species increased from 22 to 23 (25 taxa).

Finally, distribution in the alpine zone and perennial life cycle of *A. watsonii* is concordant with general trends of the tribe Arabideae (Karl and Koch 2013). Estimated climatic conditions for *A. watsonii*, including annual mean temperature, annual precipitation and other bioclimatic variable (not provided here) reflect a continental climate also described for other members of the *A. hirsuta* aggregate including *A. sudeica, A. hirsuta* etc. In conclusion, all environmental parameters and life cycle strategies of *A. watsonii* are in agreement with the genetic affiliation to the *A. hirsuta* aggregate and its relatives within the main *Arabis* clade.

**Acknowledgement**

This study was supported by Hacettepe University Scientific Research Projects Coordination Unit (Projects no. 014D01601015-489). We thank Çağasan Karacaoğlu for helping us to use DIVAGIS and İsmail Kudret Sağlam for correcting style and grammar.

**References**

Al-Shehbaz IA (1988) The genera of Arabideae (Cruciferae; Brassicaceae) in the south eastern United States. Journal of the Arnold Arboretum 69: 85–166. doi: 10.5962/bhl.part.2391

Al-Shehbaz IA (2005) Nomenclatural notes on Eurasian *Arabis* (Brassicaceae). Novon 15: 519–524.

Al-Shehbaz IA, Mutlu B, Dönmez A (2007) The Brassicaceae of Turkey, updated. Turkish Journal of Botany 31: 327–336.

Al-Shehbaz IA, German DA, Karl R, Jordon-Thaden I, Koch MA (2011) Nomenclatural adjustments in the tribe Arabideae (Brassicaceae). Plant Diversity and Evolution 129(1): 71–76. doi: 10.1127/1869-6155/2011/0129-0044

Al-Shehbaz IA (2012) A generic and tribal synopsis of the Brassicaceae (Cruciferae). Taxon 61: 931–954.

Ansell SW, Schneider H, Pedersen N, Grundmann M, Russell SJ, Vogel JC (2007) Recombination diversifies chloroplast trnF pseudogenes in *Arabidopsis lyrata*. Journal of Evolutionary Biology 29: 2400–2411. doi: 10.1111/j.1420-9101.2007.01397.x

Cullen J (1965) *Arabis* L. In: Davis PH (Ed.) Flora of Turkey and the East Aegean Islands (Vol. 1). Edinburgh University Press, Edinburgh, 422–429.

Davis PH, Mill RR, Tan K (1988) *Arabis* L. In: Davis PH, Mill RR, Tan K (Eds) Flora of Turkey and the East Aegean Islands (Suppl. 1). Vol. 10, Edinburgh University Press, Edinburgh, 49.
Daşkin R (2013) Arabis kaynakiae (Brassicaceae), a new species from south Anatolia, Turkey. Phytotaxa 126: 43–48. doi: 10.11646/phytotaxa.126.1.5

Duman H (2001) A new species of Arabis L. (Brassicaceae) from south Anatolia. Botanical Journal of Linnean Society 137: 87–90. doi: 10.1111/j.1095-8339.2001.tb01109.x

Duman H, Duran A (2001) A new species of Arabis L. (Brassicaceae) from south Anatolia. Israel Journal of Plant Science 49: 237–240.

Drummond AJ, Suchard MA, Xie D, Rambaut A (2012) Bayesian phylogenetics with BEAUti and the BEAST 1.7. Molecular Biology and Evolution 29: 1969–1973. doi: 10.1093/molbev/mss075

Güner A, Aslan S, Ekim T, Vural M, Babaç MT (2012) Türkiye Bitkileri Listesi (Damarlı Bitkiler). Flora Araştırmaları Derneği ve Nezahat Gökyiğit Botanik Bahçesi Yayıncılığı, İstanbul.

Ekim T, Koyuncu M, Vural M, Duman H, Aytaç Z, Adigüzel N (2000) Red Data Book of Turkish Plants (Pteridophyta and Spermatophyta). Turkish Association for the Conservation of Nature and Natural Resources, Ankara.

Fırat M (2013) Ferhenga Navêni Riwêkên Bi Kurdî/Kürtçe Bitki Adları Sözlüğü/ Dictionary of Plant Names in Kurdish, Kalkan Offset, Ankara, 652 pp.

Fırat M (2016) Kemulariella tahirelcii (Asteraceae; Astereae), a new species from Şırnak, Turkey. Phytotaxa 253(1): 90–96. doi: 10.11646/phytotaxa.253.1.7

IUCN (2016) Standards and Petitions Subcommittee. Guidelines for Using the IUCN Red List Categories and Criteria. Version 12. Prepared by the Standards and Petitions Subcommittee.

Jalas J, Souminen J (1994) Atlas Florae europaeae, vol. 10. The Committee for Mapping the Flora of Europe & Societas Biologica Fennica Vanamo, Helsinki, Finland, 187–211.

Jordon-Thaden IE, Hase I, Al-Shehbaz IA, Koch M (2010) Molecular phylogeny and systematics of the genus Draba s.l (Brassicaceae) and identification of its closest related genera. Molecular Phylogenetics and Evolution 55: 524–540. doi: 10.1016/j.ympev.2010.02.012

Karl R, Kiefer C, Ansell S, Koch MA (2012) Systematics and evolution of arctic-alpine Arabis alpina L. (Brassicaceae) and its closest relatives in the eastern Mediterranean. American Journal of Botany 99: 778–794. doi: 10.3732/ajb.1100447

Karl R, Koch MA (2013) A world-wide perspective on crucifer speciation and evolution: Phylogenetics, biogeography and trait evolution in tribe Arabeae. Annals of Botany (Oxford) 112: 983–1001. doi: 10.1093/aob/mct165

Karl R, Koch MA (2014) Phylogenetic signatures of adaptation: the Arabis hirsuta species aggregate (Brassicaceae) revisited. Perspectives in Plant Ecology Evolution and Systematics 16: 247–264. doi: 10.1016/j.ppees.2014.06.001

Kiefer M, Schmickl R, German D, Lysak M, Al-Shehbaz IA, Franzke A, Mummenhoff K, Stamatakis A, Koch MA (2014) BrassiBase: introduction to a novel knowledge database on Brassicaceae evolution. Plant Cell Physiology 55: e3. doi: 10.1093/pcp/pct158

Koch M, Bishop J, Mitchell-Olds T (1999) Molecular systematics and evolution of Arabidopsis and Arabis. Plant Biology 1: 529–537. doi: 10.1111/j.1438-8677.1999.tb00779.x

Koch M, Haubold B, Mitchell-Olds T (2000) Comparative analysis of chalcone synthase and alcohol dehydrogenase loci in Arabidopsis, Arabis and related genera (Brassicaceae). Molecular Biology and Evolution 17: 1483–1498. doi: 10.1093/oxfordjournals.molbev.a026248
Koch M, Haubold B, Mitchell-Olds T (2001) Molecular systematics of the Cruciferae: Evidence from coding plastome matK and nuclear Chs sequences. American Journal of Botany 88: 534–544. doi: 10.2307/2657117

Koch MA, Karl R, Kiefer C, Al-Shehbaz IA (2010) Colonizing the American continent: systematics of the genus Arabis in North America (Brassicaceae). American Journal of Botany 97: 1040–1057. doi: 10.3732/ajb.0900366

Koch MA, Kiefer M, German D, Al-Shehbaz IA, Franzke A, Mummenhoff M, Schmickl R (2012) BrassiBase: tools and biological resources to study characters and traits in the Brassicaceae – version 1.1. Taxon 61: 1001–1009.

Meyer FK (2006) Kritische revision der „Thlaspi“-Arten Europas, Afrikas und Vorderasiens, Spezieller Tiel, VII. Callothlaspi F.K. Mey. Haussknechtia 11: 175–194.

Mutlu B (2004) A new species of Arabis L. (Brassicaceae) from inner Anatolia, Botanical Journal of Linnean Society 145: 251–256. doi: 10.1111/j.1095-8339.2004.00275.x

Mutlu B, Dönmez AA (2003) Arabis mollis Steven (Brassicaceae): A new record for Turkey. Turkish Journal of Botany 27: 235–238.

Mutlu B, Erik S (2012) The Taxonomical Position of Arabis graellsiiiformis Hedge (Brassicaceae) that it’s known as an endemic species of Turkey. Hacettepe Journal of Biology and Chemistry 40: 69–74.

Mutlu B, Erik S (2015) Distribution maps and New IUCN threat categories for the genus of Arabis, Pseudoturritis and Turritis (Brassicaceae) in Turkey. Hacettepe Journal of Biology and Chemistry 43(2): 133–143.

Özhatay N, Kültür Ş, Aslan S (2009) Check list of additional taxa to the supplement flora of Turkey 4. Turkish Journal of Botany 33: 191–226. doi: 10.3906/bot-1101-20

Özhatay N, Kültür Ş, Gürdal MB (2011) Check-list of additional taxa to the supplement Flora of Turkey 5. Turkish Journal of Botany 35: 589–624. doi: 10.3906/bot-1101-20

Parolly G, Hein P (2000) Arabis lycia (Cruciferae), a new chasmophyte from Taurus Mts, Turkey, and notes on related species. Willdenowia 30: 293–304. doi: 10.3372/wi.30.30208

Taberlet P, Gielly L, Pautou G, Bouvet J (1991) Universal primers for amplification of three non-coding regions of chloroplast DNA. Plant Molecular Biology 17: 1105–1109. doi: 10.1007/BF00037152

Tamura K, Stecher G, Peterson D, Filipski A, Kumar S (2013) MEGA6: Molecular evolutionary genetics analysis version 6.0. Molecular Biology and Evolution 30: 2725–2729. doi: 10.1093/molbev/mst197

Warwick SI, Al-Shehbaz IA, Sauder CA, Murray DF, Mummenhoff K (2004) Phylogeny of Smelowskia and related genera (Brassicaceae) based on nuclear ITS DNA and chloroplast trnL intron DNA sequences. Annals of Missouri Botanical Garden 91: 99–23.

White TJ, Bruns T, Lee S, Taylor J (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ (Eds) PCR protocols: A guide to methods and applications. Academic Press, New York, 315–322. doi: 10.1016/b978-0-12-372180-8.50042-1