Identification of a novel mutation in the cornea specific keratin 12 gene causing Meesmann`s corneal dystrophy in a German family

Ina Clausen, Gernot I.W. Duncker, Claudia Grünauer-Kloevekorn

Department of Ophthalmology, Martin-Luther-University Halle-Wittenberg, Halle (Saale), Germany

Purpose: To report a novel missense mutation of the cornea specific keratin 12 (KRT12) gene in two generations of a German family diagnosed with Meesmann`s corneal dystrophy.

Methods: Ophthalmologic examination of the proband and sequencing of keratin 3 (KRT3) and KRT12 of the proband and three other family members were performed. Restriction enzyme analysis was used to confirm the detected mutation in affected individuals of the family.

Results: Slit-lamp biomicroscopy of the proband revealed multiple intraepithelial microcysts comparable to a Meesmann dystrophy phenotype. A novel heterozygous A→G transversion at the first nucleotide position of codon 129 (ATG>GTG, M129V) in exon 1 of KRT12 was detected in the proband, her two affected sons but not in her unaffected husband or 50 control individuals.

Conclusions: We have identified a novel missense mutation within the highly conserved helix-initiation motif of KRT12 causing Meesmann`s corneal dystrophy in a German family.

The cornea represents the anterior surface of the eye and must maintain its structural integrity as well as its transparency to retain good vision. The outermost layer of the cornea is presented by the corneal epithelium.

In 1939 the German ophthalmologists Meesmann and Wilke described a dominantly inherited dystrophy affecting the corneal epithelium in three families living in northern Germany [1]. The 34 affected family members showed ocular irritation, tearing, and glare. Slit lamp biomicroscopy revealed myriads of microcystic epithelial opacities extending to the corneal limbus. Twenty-six years after the initial report by Meesmann and Wilke, Behnke and Thiel [2] published an extensive study of the same three northern German families. Kuwabara and Ciccarelli [3] were the first who examined the ultrastructure of the cornea from patients with Meesmann`s corneal dystrophy (MECD; OMIM 122100) in 1964. They were followed by Fine et al. [4] in 1977 and Tremblay and Dube [5] in 1982. The authors described the corneal epithelial basement membrane as thickened with an intracellular “peculiar substance” that reacted with periodic acid and Schiff`s reagent. This substance appeared to be derived from the tonofilaments.

Keratins are a group of water-insoluble proteins that form intermediate filaments in epithelial cells. They can be divided into acidic and basic-neutral subfamilies according to their relative charges, immunoreactivity, and sequence homologies to type I and II wool keratins, respectively. In vivo, a type I keratin (acidic) pairs with a specific type II keratin (basic or neutral) to form a heterodimer [6]. The structural integrity of corneal epithelial cells derives mainly from its unique composition of the cytoskeletal intermediate filaments keratin 3 and keratin 12, whereby disruption of a single keratin polymer can compromise epithelial structure and function [7]. Among keratin proteins the helix-initiation and helix-termination motifs are highly conserved and crucial for intermediate filament heterodimerization.

A breakthrough approached in 1997 when McLean and coworkers [8,9] examined two families from Northern Ireland and German descendants of the family originally described by Meesmann and Wilke. They were able to relate MECD to the keratin cluster on chromosome 17 (17q12–21). In addition, they performed molecular genetic studies by sequencing the candidate genes keratin 3 (KRT3) and keratin 12 (KRT12) [8, 9]. Irvine et al. [8] postulated that dominant-negative mutations in these keratins might be the cause of MECD. Indeed, they found linkage of the disorder to the KRT12 locus in Meesmann`s original German kindred. Furthermore, in the two pedigrees from Northern Ireland, they found that the disorder cosegregated with KRT12 in one pedigree and KRT3 in the other.

To our best knowledge three mutations in KRT3 and 19 mutations in KRT12 in patients showing MECD have been identified so far (Table 1) [8-22]. In KRT3 all detected mutations are located in the helix-termination motif (Figure 1), whereas in KRT12 mutations are found within the helix-initiation and the helix-termination motif as well (Figure 2).
Table 1. Review of known mutations in the keratin 3 and keratin 12 genes causing MECD.

| Gene | Exon | Codon number | Codon change | Amino acid change | References |
|------|------|--------------|--------------|-------------------|------------|
| KRT3 | 7    | 498          | GAG-GTG      | Glu-Val           | [10]       |
|      | 7    | 503          | CGC-CGC      | Arg-Pro           | [11]       |
|      | 7    | 509          | GAG-AAG      | Glu-Lys           | [8]        |
| KRT12| 1    | 129          | AGT-GTG      | Met-Val           | Current Report |
|      | 1    | 129          | AGT-ACG      | Met-Thr           | [9]        |
|      | 1    | 130          | CAA-CCA      | Gln-Pro           | [12]       |
|      | 1    | 132          | CTT-CAT      | Leu-His           | [13]       |
|      | 1    | 133          | AAT-AAG      | Asn-Lys           | [14]       |
|      | 1    | 135          | AGA-ACA      | Arg-Thr           | [8]        |
|      | 1    | 135          | AGA-ATA      | Arg-Ile           | [15]       |
|      | 1    | 135          | AGA-AGC      | Arg-Ser           | [16]       |
|      | 1    | 135          | AGA-GGA      | Arg-Gly           | [15]       |
|      | 1    | 137          | GCT-CCT      | Ala-Pro           | [17]       |
|      | 1    | 140          | CTG-CGG      | Leu-Arg           | [15]       |
|      | 1    | 143          | GTG-CTG      | Val-Leu           | [8]        |
|      | 1    | 143          | GTG-TTG      | Val-Leu           | [18]       |
|      | 6    | 122ins27     | 400ins9      |                   | [16]       |
|      | 6    | 426          | ATT-AGT      | Ile-Ser           | [19]       |
|      | 6    | 426          | ATT-AGT      | Ile-Val           | [20]       |
|      | 6    | 429          | TAT-GAC      | Tyr-Asp           | [15]       |
|      | 6    | 429          | TAT-TGT      | Tyr-Cys           | [11]       |
|      | 6    | 430          | CGC-CCC      | Arg-Pro           | [21]       |
|      | 6    | 433          | CTG-CGG      | Leu-Arg           | [22]       |

Figure 1. Scheme of the KRT3 structure. The localizations of the known point mutations within the helix-termination motif are indicated.

Here we report a novel heterozygous mutation within the helix-initiation motif of KRT12 causing MECD in three members of a German family.

METHODS

Subjects: A two generation family with three affected individuals was studied. The index patient (I/2), an 82-year-old woman, was examined by slit-lamp biomicroscopy. Peripheral blood samples of patient I/2, her unaffected husband (I/1) and the two affected sons (II/1, II/2) were obtained. The ophthalmological history of the proband’s deceased parents was unknown. Fifty unrelated volunteers were recruited to serve as controls. All subjects were treated in accordance with the tenets of the Declaration of Helsinki and signed a consent form.

DNA collection, polymerase chain reaction amplification, and sequencing: Genomic DNA was isolated from peripheral blood leucocytes by standard procedures using the Invitro Spin Blood Maxi Kit (Invitek, Berlin, Germany). Blood samples (10 ml) treated with EDTA were processed following the manufacturers instructions. Exons 1–9 of KRT3 and exons 1–8 of KRT12 were amplified using the primers shown in Table 2. Each PCR reaction was performed in a volume of 50 µl containing 1 µM of forward and reverse primer, 0.5 U of AmpliTaq Gold DNA Polymerase (Applied Biosystems Inc., Foster City, CA), 1 µl dNTP mixture (Applied
Biosystems), 5 µl of 10× PCR buffer (Applied Biosystems) and 100 ng of human genomic DNA. Amplification reactions were performed under the following conditions: 5 min of denaturation at 96 °C followed by 35 cycles of denaturation at 96 °C for 30 s, annealing at 60–66 °C for 30 s, extension at 72 °C for 45 s, and a further extension step at 72 °C for 5 min.

For direct sequencing, amplified DNA was purified using the QIAquick PCR purification kit (Qiagen GmbH, Hilden, Germany), cycle sequenced with BigDye Terminator v3.1 Cycle Sequencing kit (Perkin Elmer, Waltham, MA) and directly sequenced using an automatic sequencer (ABI 3100 Genetic Analyzer C; Applied Biosystems).

**Mutation confirmation:** The mutation M129V generated a HpyCH4III (New England Biolabs GmbH, Frankfurt, Germany) restriction enzyme site in exon 1b of KRT12. Therefore, PCR products of all four family members (three affected and one unaffected individual) as well as 50 unrelated control subjects were submitted to restriction digestion under the following conditions. A reaction mixture containing 10 µl PCR product, 5 µl enzyme, 25 µl NEBuffer 4 (50 mM potassium-acetate, 20 mM Tris-acetate, 10 mM magnesium-

### Table 2. Primers for Polymerase Chain Reaction Direct Sequencing of KRT3 and KRT12.

| Exon | Forward primer sequence (5′→3′) | Reverse primer sequence (5′→3′) | Size (bp) |
|------|----------------------------------|----------------------------------|-----------|
| Keratin 3  |                                   |                                  |           |
| 1    | TGCACAGGTCTCTCATTTCCCATCC         | TCTCAACCTGGATATCTTCCCA           | 887       |
| 2    | AGTGTGGGCTGTAGTGGCTCTCCTCTCT     | ACCATGGCTTGAGAAGAAGGTGA          | 439       |
| 3    | ATGGAGGGAGGAAGAGAGCATGAAGTACT    | ATGTGCTCAAGGGCTGACACTTG          | 275       |
| 4    | GCTCTCTTCTGTCTGAGTGTGTTG         | GCACAGCCTCAAAATCTGGAAACA         | 238       |
| 5    | AGTGAAGAAGAAGCTCCTCTGAGTGTG     | TGAACACCTCCAGTGGACCTCCGAA       | 235       |
| 6    | AAGGTGTTGTTGGTTGATGGTGGAG      | ATGGTGGAGATAGCTGCTGGTG          | 345       |
| 7    | AATCAATGCTGATGAGGATAGGAGC       | TATCTGGCACTTGGCCCTATG          | 354       |
| 8    | TCTGTGTTGATGATGCTTTGTTGAGG      | AGGCCTACATCCCTCTCTCCTCCT        | 228       |
| 9    | ACAATAAACATAGCAGCTGGCGCTG      | AATACCTAGAGGGCCGAGTAGGAA        | 756       |

| Keratin 12 |                               |                                  |           |
| 1a   | GGATCCAATTTTGAGTGGAGA          | ACCTAGAGAGCCACCTCTCTGAGG         | 459       |
| 1b   | AGCCCCAGGAGGGCTCCTCTCTA        | AGTCAGCTAAATGTGGAAAT             | 369       |
| 2    | TAGGCCTCTTCTGAGGCTCAAATC       | AGGCAGAGAGTAGGACA                | 194       |
| 3    | GCAAGAATAAGGGCCTGGAGA          | TTTTGGGTCTGGGAGA                 | 297       |
| 4    | GCGCCAGAGGAGGAAAAGTA          | GCAGCGCTTTCTTGTAATG              | 290       |
| 5    | AGAAGCGCGCTCAGGA             | AAAAGAGGAGGGTAGCAACC             | 278       |
| 6    | GTCCCCTCCATAGGTATTCTC         | CTATTTCTGCTCCCTACCT              | 330       |
| 7    | TCCATATCCAAAGCCAGTGT          | GTCTTACAGGGTTTGCATT              | 198       |
| 8    | GCCTACATATCAAACACCAGT         | TGACCAAATAAGGTCTGGAGACT          | 370       |
acetate, 1 mM dithiothreitol; New England Biolabs), and 170 µl H2O was incubated overnight at 37 °C. In the presence of the heterozygous M129V mutation, HpyCH4III digestion resulted in fragments of 293 bp and 76 bp with the additional undigested 369 bp PCR product. The digested products were separated and visualized on a 2% agarose gel stained with ethidium bromide. Digestion analysis was additionally used to screen fifty healthy control subjects for the M129V mutation.

RESULTS

Clinical exam: Slit lamp examination of patient I/2 revealed multiple microcysts within the corneal epithelium of both eyes with highest density in the center of the cornea (Figure 3). The proband had recurrent corneal erosions, blurred vision, and tearing. Visual acuity was highly reduced (OD finger count, OS 5/20) predominantly due to high myopia with myopic macular degeneration. Additionally, the patient was pseudophacic with posterior chamber lenses after cataract surgery. Ocular tension was within normal range.

KRT3 sequence analysis: The sequence analysis of all nine exons of KRT3 revealed no mutations in the four family members.

KRT12 sequence analysis: Direct forward and backward sequencing of exon 1b of KRT12 disclosed the heterozygous 409A>G transversion at the first nucleotide position of codon 129 (ATG>GTG) in the proband I/2 (Figure 4) and her two affected sons (II/1, II/2). This is predicted to substitute a valine residue for methionine (M129V, NM_000223). This missense mutation was absent in the unaffected husband (I/1). Additionally, the proband and her sons harbored the single-nucleotide polymorphism (SNP) 43C>T (Figure 5), resulting in an amino acid change in codon 15 from proline to serine (P15S, NM_000223). Further sequence analysis of exons 2–8 did not show any alterations.

Mutation confirmation: Restriction enzyme analysis employing HpyCH4III resulted in digested fragments of 76 bp and 293 bp in the proband (I/2) and her two affected sons (II/1, II/2) as shown in Figure 6. Since the found mutation is heterozygous, the undigested wild type fragment of 369 bp is also seen. Exclusively undigested wild type DNA was seen in the unaffected husband (I/1) and in fifty healthy control individuals (data not shown).

DISCUSSION

All known mutations in the cornea specific genes KRT3 and KRT12 are responsible for the development of Meesmann’s corneal dystrophy, an autosomal dominant disorder of the corneal epithelium. Until the present study, three mutations have been reported in KRT3 and 19 in KRT12, including a 27-nucleotide long insertion in exon 6. All mutations are summarized in Table 1. It is noteworthy, that the majority of
the KRT12 mutations are localized within the region encoding the helix-initiation motif. In contrast, the known mutations of KRT3 are exclusively within the helix-termination motif. In keratin proteins, the helix-initiation and –termination motifs are highly conserved regions playing an important role in normal filament assembly [23].

With the predicted amino acid substitution M129V we revealed the third mutation of KRT12 in a German family. Previously, Irvine and coworkers [8] identified the point mutation R135T in descendants of the German kindred originally described by Meesmann and Wilke [1]. In 2000, Corden et al. [12] reported a Q130P substitution in a southern German family.

Another nucleotide exchange within the same codon (M129T) predicting the substitution of the methionine (ATG) residue by threonine (ACG) was described independently by
Cordon et al. [9] and Nichini et al. [19] in American families. It is unknown whether these individuals are related.

Our index patient, an 82-year-old female, had a rather severe case of MECD with photophobia, tearing, and recurrent painful corneal erosions since adulthood. Visual acuity was highly reduced due to myopic macular degeneration. Additionally, the proband and her affected sons harboured the SNP 67C>T predicting the amino acid substitution P15S. This SNP has been previously described by Nielsen et al. [18] in an asymptomatic Danish family. The allele frequency in healthy probands was calculated to be 29%. Since the affected family members described by Nielsen et al. [18] presented an asymptomatic form of MECD, it seems unlikely, that the additional SNP should be responsible for the severity of symptoms in our patient.

Still, it seems cryptic why certain mutations lead to a more or less severe phenotype. Severe cases of MECD have been reported due to mutations within the helix initiation motif [12,14] as well as the helix termination motif [21]. Furthermore, the quality of the amino acid change seems not to elucidate the genotype-phenotype correlation. The V143L substitution in an asymptomatic Danish family means the exchange of two neutral aliphatic amino acids [18]. However, in our patient the aliphatic neutral methionine is replaced by another aliphatic and neutral amino acid (valine) as well.

Moreover, mutations in KRT3 cause a MECD phenotype indistinguishable from KRT12 mutations. Kao et al. [7] demonstrated the importance of functional KRT3/KRT12 heterodimers for corneal stability in a KRT12 knockout mouse model. Any modification that results in an impairment of the heterodimerization process will lead to corneal fragility. This could explain why mutations in any member of the heterodimer lead to almost identical phenotypes. Therefore, other individual factors may contribute to the MECD phenotype.

The present study adds another novel mutation to the collection of mutations causing Meesmann’s corneal dystrophy.

ACKNOWLEDGMENTS
We thank all the family members for their participation. We are highly grateful to Elke Vetter for her unique expertise and excellent laboratory skills.

REFERENCES
1. Meesmann A, Wilke F. Klinische und anatomische Untersuchungen über eine bisher unbekannte, dominant...
vererbte Epitheldystrophie der Hornhaut. Klin Monatsbl Augenheilkd 1939; 103:361-91.

2. Behnke H, Thiel HJ. On hereditary epithelial dystrophy of the cornea (type Meesmann-Wilke) in Schleswig-Holstein. Klin Monatsbl Augenheilkd 1965; 147:662-72. [PMID: 5296628]

3. Kuwabara T, Ciccarelli EC. Meesmann's Corneal Dystrophy. A Pathological Study. Arch Ophthalmol 1964; 71:672-82. [PMID: 14120659]

4. Fine BS, Yanoff M, Pitts E, Slaughter FD. Meesmann's epithelial dystrophy of the cornea. Am J Ophthalmol 1977; 83:633-42. [PMID: 301357]

5. Tremblay M, Dube I. Meesmann's corneal dystrophy: ultrastructural features. Can J Ophthalmol 1982; 17:24-8. [PMID: 6979375]

6. Moll R, Franke WW, Schiller DL, Geiger B, Krepler R. The catalog of human cytokeratins: patterns of expression in normal epithelia, tumors and cultured cells. Cell 1982; 31:11-24. [PMID: 6186379]

7. Kao WW, Liu CY, Converse RL, Shiraishi A, Kao CW, Ishizaki M, Doetschman T, Duffy J. Keratin 12-deficient mice have fragile corneal epithelia. Invest Ophthalmol Vis Sci 1996; 37:2572-84. [PMID: 8977471]

8. Irvine AD, Corden LD, Swensson O, Swensson B, Moore JE, Frazer DG, Smith FJ, Knowlton RG, Christophers E, Rochels R, Uitto J, McLean WH. Mutations in cornea-specific keratin K3 or K12 genes cause Meesmann's corneal dystrophy. Nat Genet 1997; 16:184-7. [PMID: 9171831]

9. Corden LD, Swensson O, Swensson B, Smith FJ, Rochels R, Uitto J, McLean WH. Molecular genetics of Meesmann's corneal dystrophy: ancestral and novel mutations in keratin 12 (K12) and complete sequence of the human KRT12 gene. Exp Eye Res 2000; 70:41-9. [PMID: 10644419]

10. Szaflrik JP, Oldak M, Maksym RB, Kaminska A, Pollak A, Udziala M, Ploski R, Szaflrik J. Genetics of Meesmann corneal dystrophy: a novel mutation in the keratin 3 gene in an asymptomatic family suggests genotype-phenotype correlation. Mol Vis 2008; 14:1713-8. [PMID: 18806880]

11. Chen YT, Tseng SH, Chao SC. Novel mutations in the helix termination motif of keratin 3 and keratin 12 in a Taiwanese family with Meesmann corneal dystrophy. Cornea 2005; 24:928-32. [PMID: 16227835]

12. Corden LD, Swensson O, Swensson B, Rochels R, Wannke B, Thiel HJ, McLean WH. A novel keratin 12 mutation in a German kindred with Meesmann's corneal dystrophy. Br J Ophthalmol 2000; 84:527-30. [PMID: 10781519]

13. Wang LJ, Tian X, Zhang QS, Liu L. Analysis of mutation in KRT12 gene in a Chinese family with Meesmann's corneal dystrophy. Zhonghua yan ke za zhi 2007; 43:885-9. [PMID: 18201524]

14. Irvine AD, Coleman CM, Moore JE, Swensson O, Morgan SJ, McCarthy JH, Smith FJ, Black GC, McLean WH. A novel mutation in KRT12 associated with Meesmann's epithelial corneal dystrophy. Br J Ophthalmol 2002; 86:729-32. [PMID: 12084738]

15. Nishida K, Homna Y, Doto A, Kawasaki S, Adachi W, Nakamura T, Quantock AJ, Hosotani H, Yamamoto S, Okada M, Shimomura Y, Kinoshita S. Isolation and chromosomal localization of a cornea-specific human keratin 12 gene and detection of four mutations in Meesmann corneal epithelial dystrophy. Am J Hum Genet 1997; 61:1268-75. [PMID: 9399908]

16. Yoon MK, Warren JF, Holscelaw DS, Gritz DC, Margolis TP. A novel arginine substitution mutation in 1A domain and a novel 27 bp insertion mutation in 2B domain of keratin 12 gene associated with Meesmann's corneal dystrophy. Br J Ophthalmol 2004; 88:752-6. [PMID: 15148206]

17. Takahashi K, Takahashi K, Murakami A, Okisaka S, Kimura T, Kanai A. Heterozygous Ala137Pro mutation in keratin 12 gene found in Japanese with Meesmann's corneal dystrophy. Jpn J Ophthalmol 2002; 46:673-4. [PMID: 12543196]

18. Nielsen K, Orntoft T, Hjortdal J, Rasmussen T, Ehlers N. A novel mutation as the basis for asymptomatic Meesmann dystrophy in a Danish family. Cornea 2008; 27:100-2. [PMID: 18245975]

19. Nichini O, Manzi V, Munier FL, Schorderet DF. Meesmann corneal dystrophy (MECD): report of 2 families and a novel mutation in the cornea specific keratin 12 (KRT12) gene. Ophthalmic Genet 2005; 26:169-73. [PMID: 16352477]

20. Coleman CM, Hannush S, Covello SP, Smith FJ, Uitto J, McLean WH. A novel mutation in the helix termination motif of keratin K12 in a US family with Meesmann corneal dystrophy. Am J Ophthalmol 1999; 128:687-91. [PMID: 10612503]

21. Sullivan LS, Baylin EB, Font R, Daiger SP, Pepose JS, Clinch TE, Nakamura H, Zhao XC, Yee RW. A novel mutation of the Keratin 12 gene responsible for a severe phenotype of Meesmann's corneal dystrophy. Mol Vis 2007; 13:975-80. [PMID: 17653038]

22. Seto T, Fujiki K, Kishishita H, Fujimaki T, Murakami A, Kanai A. A novel mutation in the cornea-specific keratin 12 gene in Meesmann corneal dystrophy. Jpn J Ophthalmol 2008; 52:224-6. [PMID: 18661274]

23. Hatfield M, Weber K. The coiled coil of in vitro assembled keratin filaments is a heterodimer of type I and II keratins: use of site-specific mutagenesis and recombinant protein expression. J Cell Biol 1990; 110:1199-210. [PMID: 16911898]