Title
Src family kinase-mediated vesicle trafficking is critical for neutrophil basement membrane penetration.

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Leukocyte recruitment into inflamed tissue is highly dependent on the activation and binding of integrins to their respective ligands, followed by the induction of various signaling events within the cell referred to as outside-in signaling. Src family kinases (SFK) are the central players in the outside-in signaling process, assigning them a critical role for proper immune cell function. Our study investigated the role of SFK on neutrophil recruitment in vivo using Hck−/− Fgr−/− Lyn−/− mice, which lack SFK expressed in neutrophils. We show that loss of SFK strongly reduces neutrophil adhesion and post-arrest modifications in a shear force dependent manner. Additionally, we found that in the absence of SFK, neutrophils display impaired Rab27a-dependent surface mobilization of neutrophil elastase, VLA3 and VLA6 containing vesicles. This results in a defect in neutrophil vascular basement membrane penetration and thus strongly impaired extravasation. Taken together, we demonstrate that SFK play a role in neutrophil post-arrest modifications and extravasation during acute inflammation. These findings may support the current efforts to use SFK-inhibitors in inflammatory diseases with unwanted neutrophil recruitment.

**ABSTRACT**

**Introduction**

Chronic inflammatory diseases are an increasing problem in western industrialized countries. A hallmark of these disorders is the recruitment of leukocytes from the circulation into affected tissues. This process follows a well-defined cascade of activation and adhesion events starting with the initial capture of leukocytes from the blood stream, followed by rolling along inflamed endothelium. Both steps are mediated by selectins interacting with glycosylated ligands on leukocytes.

Through binding of the leukocyte specific integrin LFA1 (αLβ2) to ICAM-1 on endothelial cells, leukocytes slow down their rolling velocity, and in combination with chemokine stimulation, firmly arrest on the endothelial surface. This is followed by post-arrest modifications, a process characterized by cell spreading, cytoskeleton rearrangements and crawling along the endothelium, that is critical for tight adhesion to the substrate and allows an appropriate spot for extravasation into tissue. Transmigration involves the crossing of the venular wall and the underlying vascular basement membrane (BM), two steps which are still incompletely understood. Several reports suggest a role for the integrins VLA5 (α5β1) and VLA6 (α6β1) along with neutrophil elastase (NE) in this process.

Recently, our group has shown that neutrophils translocate VLA5, VLA6 and NE from internally stored vesicles to the cell surface, to subsequently cross the vascular BM. The release of these vesicles is initiated by interactions of neutrophils with the inflamed endothelium in a PECAM-1/ICAM-1- and CXCL1-dependent manner. Vesicle transport is...
mainly regulated by Rab GTPases and their effector proteins with Rab27a being the main Rab molecule involved in the secretory machinery of neutrophils. Rab27a function is mediated by the two effector molecules synaptojanin-like protein 1 (JFC1, encoded by Syt11 in mice) and Munc13-4 (Unc13D). Although most of these processes rely on integrin signaling, integrins themselves lack enzymatic activity. Therefore, numerous proteins are recruited to their intracellular tails, among those Src family kinases (SFK). Their early recruitment during integrin activation assigns SFK a critical role in the so-called outside-in signaling process and thus regulation of central signaling pathways downstream of integrin-receptor ligation. Neutrophils express three members of this family, namely Hck, Fgr and Lyn. Their function has been intensely studied in SFK single, double (Hck-Fgr), and triple knockout (ko) (Hck-Fg-Lyn') mice, demonstrating a role for SFK in integrin activation and their downstream signaling. Neutrophils isolated from Hck-Fgr' mice showed poor spreading, indicating that integrin outside-in signaling is impaired. Additionally, neutrophil migration into the liver of Hck-Fgr' mice in an in vivo endotoxemia model is severely reduced. Furthermore, Kovács et al. recently showed that neutrophils exhibit reduced capability to create a microinflammatory environment, resulting in diminished neutrophil extravasation in a model of autoantibody-induced arthritis and inflammatory blistering skin disease.

Our study aimed to investigate the effect of SFK on leukocyte recruitment in an in vivo setting of acute inflammation, focusing on post-arrest modifications and the molecular mechanism of vascular BM penetration. We show that, in the genetic absence of SFK, these two steps are strongly impaired. Adherent Hck-Fgr-Lyn' neutrophils are unable to withstand shear forces and display diminished LFA1 clustering with reduced phosphorylation levels of Paxillin, Cortactin and Syk, suggesting that SFK depletion results in severely impaired adhesion strengthening. In addition, we show that SFK are critical for crossing the vascular BM during neutrophil extravasation by facilitating translocation of VLA3-, VLA6- and NE-containing vesicles to the cell surface. Therefore, SFK are not only important mediators of neutrophil post-arrest modifications, but are also required to breach the vascular BM.

Methods

Animals

Lyz2GFP, SFK-ko (Hck'Fgr'Lyn') and SFK-ko Lyz2GFP mice were generated as described earlier. C57BL/6 wildtype mice were purchased from Janvier Labs (Saint Berthevin, France). All animal experiments were approved by the Regierung von Oberbayern, Germany (AZ 55.2-1-54-2531-80-76/12 and 55.2-1-54-2532-102-2531-80-76/12). Animals were housed with a 12 h light/dark cycle and given ad libitum access to food and water.

Live cell imaging of in vitro laminin digestion

Transmigration of neutrophils was analyzed in μ-Slide membrane ibiPore flow chambers (Ibidix, Planegg, Germany) equipped with a 300-μm thick membrane with 5-μm pores, and a subjacent rat-tail collagen gel (1.5 mg/mL) containing 10 μM N-formylmethionyl-leucyl-phenylalanine (fMLP) as chemoattractant. The upper compartment was coated with Laminin (LN), PECAM-1 and ICAM-1, and, in addition, LN was visualized using an anti-LN antibody conjugated to Alexa Fluor-647 (novusbio, Littleton, CO, USA). Isolated neutrophils from wildtype and SFK-ko animals were labeled with CellTracker™ Green CMFDA Dye and CellTracker™ Red CMTX Dye (Thermo Fisher, Waltham, MA, USA), respectively, and distributed into the upper chamber compartment in a 1:1 ratio. Time-lapse microscopy with an interval of 50 seconds was performed using an upright spinning-disk confocal microscope ( Examiner; Zeiss) equipped with a confocal scanner unit CSU-X1 (Yokogawa Electric Corporation, Japan), an EMCCD camera (EvoFluo; Photometrics), and a x20/1.0 NA water immersion objective (Plan Apochromat; Zeiss). 3D images (70 z-stacks with a step size of 2μm) were acquired per time point and analyzed by generating maximum z-projections over time using Slidebook 6.0.8 software (Bitplane) and ImageJ. Interaction strength was analyzed with the MosaicIA interaction plugin of Fiji.

Functional in vitro and in vivo experiments, including intravital microscopy, peritonitis experiments, flow chamber experiments, vessel trafficking, flow cytometry, western blot analysis and statistics are described in detail in the Online Supplementary Methods.

Data sharing statement

Original data are available on request.

Results

Src family kinase depletion reduces neutrophil adhesion and extravasation in inflamed postcapillary venules in vivo

We first investigated how loss of neutrophil-expressed SFK influences neutrophil adhesion in TNFα-stimulated cremaster muscle postcapillary venules using intravital microscopy. Interestingly, the absolute number of adherent neutrophils was strongly increased compared to wildtype control mice (Online Supplementary Figure S1A). However, after adjusting the number of adherent neutrophils to the circulating neutrophil count, which was significantly higher in SFK-ko mice compared to wildtype mice (Online Supplementary Figure S1B and C), the neutrophil adhesion efficiency calculated as number of adherent cells/mm² divided by the systemic neutrophil count was significantly reduced in SFK-ko mice (0.48±0.04) compared to wildtype mice (1.04±0.08) (Figure 1A). This was not due to altered surface expression of rolling and adhesion relevant surface proteins including CD18, CD11a, CD11b, CD62L, PSGL1, CXCR2, and CD44 as their surface expression was equal between wildtype and SFK-ko neutrophils (Online Supplementary Figure S1D). We therefore conclude that SFK are critical for neutrophil adhesion in vivo.

Efficient neutrophil extravasation into tissue is required for proper host defense. Recently, Kovács et al. demonstrated in the K/BxN serum-transfer arthritis model that SFK deficiency resulted in lower levels of cytokine production and release leading to impaired leukocyte extravasation. To investigate a cell intrinsic adhesion defect in SFK-ko mice, we injected TNFα into the mouse scrotum. This approach circumvents possible effects of a reduced inflammatory environment and enables us to focus on neutrophil recruitment itself. Performing Giemsa staining of TNFα stimulated cremaster muscle whole mounts revealed strongly reduced numbers of extravasated neutrophils in SFK-ko mice compared to wildtype mice (306.5±43.7 vs. 758.6±72.9, respectively) (Figure 1B and Online Supplementary Figure S1E). Moreover, we calculated the extravasation efficiency as total number of extravasa-
ed leukocytes divided by the total number of adherent leukocytes to differentiate between reduced extravasation due to decreased adhesion versus a specific extravasation defect. Surprisingly, we observed a strongly reduced extravasation efficiency in SFK-ko mice (0.21 in SFK-ko mice vs. 0.77 in wildtype mice) (Figure 1C), indicating that SFK depletion does not only affect neutrophil adhesion, but also interferes with extravasation itself. Finally, we also compared neutrophil cell numbers in the peritoneal cavity 2 hours (h) after TNFα-induced peritonitis (Figure 1D). In SFK-ko animals, peritoneal neutrophil numbers were reduced to 50% compared to wildtype mice (0.42x10⁵ vs. 10⁶, respectively). Taken together, these experiments demonstrate an important role for SFK in neutrophil extravasation.

Adhesion strengthening is severely impaired in Src family kinase-knockout neutrophils in vitro

Our experiments suggest that SFK-deletion in neutrophils results in an extravasation defect in the presence of the proinflammatory cytokine TNFα. After firm adhesion to the inflamed endothelium, neutrophils start to polarize and crawl along the endothelium to find a suitable spot for extravasation. These changes are highly dependent on β2 integrins LFA1 (αιβ2) and Mac1 (αMβ2) and integrin outside-in signaling.20 We analyzed neutrophil crawling using time-lapse microscopy in TNFα-stimulated cremaster muscle venules of SFK-ko and wildtype mice. We observed no differences in crawling direction (Figure 2A), but, interestingly, crawling velocity was significantly increased in SFK-ko mice (10.7 µm/min) compared to wildtype mice (10.7 µm/min) (Figure 2B), suggesting defective adhesion strengthening. Subsequently, we performed ex vivo flow chamber assays using glass capillaries coated with E-Selectin/ICAM-1/CXCL1 to mimic the inflamed endothelium. Plots of single cell tracks displayed more SFK-ko neutrophils crawling in flow direction compared to wildtype neutrophils (Figure 2C). Crawling velocity was increased in SFK-ko neutrophils crawling in flow direction compared to wildtype neutrophils (Figure 2D), verifying our in vivo results. Interestingly, only 20% of SFK-ko neutrophils were able to crawl under flow (Figure 2E and Online Supplementary Mov1), compared to 80% of wildtype cells, suggesting that SFK-ko neutrophils are unable to maintain stable adhesion to the substrate. Together, these findings demonstrate a decreased capability of SFK-ko neutrophils to withstand shear forces, indicating that SFK are necessary for adhesion strengthening during post-arrest modifications. To further investigate a shear stress dependent adhesion defect in the absence of neutrophil SFK, we conducted detachment assays in the...
flow chamber using increasing shear forces. Adherent neutrophils were assessed for each shear stress level and related to the number of adherent neutrophils under low flow conditions (Figure 2F). Loss of SFK resulted in a decrease in shear stress resistant adhesion compared to wildtype cells. At wall shear stress levels of 140 dyn/cm², only 36% of SFK-ko leukocytes could adhere to the flow chamber, while 84% of wildtype leukocytes were still attached. To analyze if these findings also apply under in vivo conditions, we correlated in vivo shear rates to

**Figure 2. Neutrophil shear stress resistance is impaired in the genetic absence of Src family kinases (SFK).** In vivo crawling experiments were analyzed in TNFα-stimulated cremaster muscle tissue of 5 wildtype and 5 SFK-knockout (ko) mice. Time-lapse movies over 15 minutes were performed and leukocytes were tracked for 15 minutes. In vitro crawling experiments were analyzed using whole blood from 3 wildtype and 3 SFK-ko mice in flow chambers coated with E-Selectin/ICAM-1 and CXCL1. Time-lapse movies over 10 minutes were performed and leukocytes tracked for 200 time points. All data are presented as means ± standard error of the mean (SEM). *P < 0.05; **P < 0.005; ***P < 0.001; n.s.: not significant. (unpaired Student t-test). (A and C) Representative single cell migration tracks (upper panel) and rose plots (lower panel) for crawling in vivo (A) and in vitro (C) are displayed. Red lines indicate migration in, black lines migration against flow direction. Arrows indicate the direction of flow. In rose diagrams, the area of each sector is proportional to the frequency of the migration vectors of tracked wildtype or SFK-ko neutrophils. At least 50 cells were analyzed for each strain. (B) Intraluminal crawling velocity. (D) Crawling velocity under flow in vitro. (E) Relative number of crawling cells per Field of View (FOV) in vitro using wildtype and SFK-ko neutrophils. (F) Neutrophil detachment was analyzed using whole blood from 4 wildtype and 4 SFK-ko mice in flow chambers coated with E-Selectin/ICAM-1 and CXCL1. Flow was applied and increased every 30 seconds as indicated and numbers of attached cells per FOV counted. All data are presented as mean±SEM. *P < 0.05; **P < 0.005; ***P < 0.001 (two-way ANOVA, Sidak multiple comparison test). (G) In vivo correlation of shear rates and number of adherent cells in TNFα-stimulated cremaster muscle venules of 5 wildtype and 5 SFK-ko mice and 5 wildtype mice treated for 3 hours with the tyrosine kinase inhibitor Dasatinib (10 mg/kg).
absolute numbers of adherent cells/mm² in TNFα-stimulated post-capillary cremaster muscle venules of SFK-ko and wildtype mice (Figure 2G). We found a similar shear rate dependent decrease in the number of adherent cells/mm² in SFK-ko mice in vivo compared to wildtype mice. Additionally, we used a pharmacological approach and administered the broad-spectrum tyrosine kinase inhibitor Dasatinib before TNFα-stimulation. Dasatinib has been used in various studies as a SFK inhibitor and led to promising results as an immune modulator.21 Similar to the results in SFK-ko mice, we found a decrease in adherent cells with increasing shear rates. Importantly, SFK-ko mice receiving Dasatinib did not show any further decrease in adherent cells at high shear stress levels, indicating that Dasatinib exerts its observed effects on adhesion via inhibition of SFK (Online Supplementary Figure S1F). Taken together, these in vivo and in vitro findings indicate that loss of SFK in neutrophils leads to a shear force dependent inability to firmly adhere to the inflamed endothelium, a prerequisite for subsequent extravasation.

**Reduced phosphorylation of cytoskeleton-associated proteins in Src family kinase-deficient neutrophils**

To further elucidate the molecular mechanisms of SFK dependent neutrophil extravasation, we analyzed different steps of integrin outside-in signaling that occur after integrins bind to their receptor. This includes integrin clustering and cytoskeletal rearrangements.22 To analyze integrin clustering, we performed time-lapse confocal microscopy and studied LFA1 clustering in SFK-ko and wildtype neutrophils in flow chambers coated with E-selectin/ICAM-1/CXCL1 (Figure 3A and Online Supplementary Movie). We observed spreading and polarizing wild-type neutrophils which showed an accumulating LFA1 signal (clustering) over time at the neutrophil uropod (Figure 3B). In contrast, SFK-deficient neutrophils appeared round and unpolarized with no obvious LFA1 clustering. Several cytoskeleton-associated proteins and signaling molecules are known to be tyrosine phosphorylated upon integrin ligation.22 We investigated the phosphorylation and activation of the direct SFK target24,25 Syk in SFK-ko and wildtype neutrophils plated on ICAM-1 and then stimulated with CXCL1 or PMA. Western blot analysis of phospho-Syk (Tynt519/520) revealed a strong upregulation of phosphorylation following stimulation of wildtype cells. In contrast, SFK deficiency prevented the upregulation of phosphorylation following stimulation with CXCL1 or PMA (Figure 3C), indicating that Syk activation is defective in the absence of SFK. Moreover, we tested the phosphorylation of the adaptor protein Paxillin, one of the critical proteins for cell adhesion, migration and podosome formation, that is known to be tyrosine phosphorylated upon β1 integrin activation.26,27 Western blot analysis showed significant upregulation of Paxillin phosphorylation when stimulated with CXCL1 or PMA in wildtype cells, while no upregulation was detectable in SFK-ko neutrophils (Figure 3D). In addition, we also investigated the tyrosine phosphorylation of Cortactin, which regulates actin branching and therefore cell migration. Likewise, we observed no significant upregulation of Cortactin Tyr421 phosphorylation upon CXCL1 or PMA stimulation in SFK-ko cell, in contrast to wildtype neutrophils (Figure 3E). Because during neutrophil adhesion and migration SFK do not selectively signal via LFA1, but also via Mac1, we additionally analyzed Paxillin phosphorylation after plating the cells on Fibrinogen (Online Supplementary Figure S1G). Similar to ICAM1 we observed a stimulus dependent phosphorylation of Paxillin in wildtype neutrophils, which was absent in SFK-ko neutrophils. Taken together, these experiments show that β1-integrin clustering and subsequent outside-in signaling is severely impaired in SFK-ko neutrophils.

**Src family kinase are indispensable to pass the vascular basement membrane**

For successful extravasation into inflamed tissue, neutrophils need to cross the endothelial cell layer and the underlying vascular basement membrane. We aimed to investigate whether SFK are required to perform this last step of extravasation. First, we analyzed transmigration capacity of SFK-ko neutrophils in a transwell assay using CXCL1 as chemoattractant (Online Supplementary Figure S2A). In line with previous results,11 we observed no difference in transmigration between SFK-ko neutrophils and wildtype neutrophils. Both groups displayed a proportional increase in transmigration with rising CXCL1 concentrations. As a next step, we coated transwells with laminin-111 (LN1) or a combination of LN1 and PECAM-1/ICAM-1 and stimulated them with CXCL1 in order to mimic the situation at the vascular BM, as reported previously.2 No difference in transmigration through LN alone was detected between wildtype and SFK-ko neutrophils (Figure 4A). Coating with LN1/PECAM-1/ICAM-1 (LN/P/I) together with CXCL1 stimulation induced a strong increase in transmigration of wildtype cells. However, SFK-ko cells failed to cross the artificial BM. How exactly neutrophils cross the vascular BM is still a matter of debate, especially, whether the BM is permanently modified by extravasating neutrophils. To further address this, we conducted transwell assays as described above, coated with a combination of LN1 and PECAM-1/ICAM-1 and stimulated with CXCL1. We also used the same total cell number (2x10⁵), but this time mixed SFK-ko x Lyz₂GFP neutrophils and wildtype neutrophils in a 1:1 ratio (1x10⁵ wildtype and 1x10⁵ SFK-ko x Lyz₂GFP neutrophils). In this manner, transmigrated SFK-ko neutrophils were identified by their GFP signal. Interestingly, in combination with wildtype neutrophils, SFK-ko neutrophils were able to overcome the BM to the same extent as wildtype cells, suggesting that wildtype neutrophils facilitate BM penetration by providing exit points for SFK-ko neutrophils (Figure 4A). To confirm these findings in vivo, we performed whole mount staining of TNFα-stimulated cremaster muscles for LN (green) to visualize the BM, and MRP14 (red) to visualize neutrophils, then analyzed the localization of neutrophils by confocal microscopy. In wildtype tissue, we were able to detect local areas of increased extravasation, which is in accordance with published literature, describing transmigration hotspots28 (Figure 4B). In contrast, none of these spots could be found in cremaster muscle tissue of SFK-ko mice. MRP14-positive cells remained located within the vascular compartment and only few extravasated neutrophils could be detected, mainly remaining in close vicinity to the vessel (white arrows). Overall, SFK-ko neutrophils covered less distance away from the vessel wall when compared to wildtype neutrophils (8.0 μm vs. 17.3 μm, respectively) (Online Supplementary Figure S2B). These in vivo findings support our earlier in vitro findings that SFK are critical for neutrophils to cross the BM.
Figure 3. Defective LFA1 clustering and adhesion strengthening in Src family kinases (SFK)-knockout (ko) mice. Integrin downstream signaling was analyzed in isolated wildtype and SFK-knockout (ko) neutrophils. (A) In vitro integrin clustering experiments were performed using whole blood from wildtype and SFK-ko mice in flow chambers coated with E-Selectin/ICAM-1 and CXCL1. Blood samples were incubated with anti-LFA1-Alexa547 (2D7) to visualize LFA1 and crawling cells were analyzed for 15 minutes by confocal microscopy. LFA1 intensity is displayed at different time points. White arrows point to the uropod. Color code indicates signal intensity. Dashed lines are exemplary for drawn intensity profiles analyzed in (B) n=3 wildtype and 3 SFK-ko mice. Scale bar: 5 μm. (B) Mean signal intensities of an intensity profile of LFA1 signal, grouped into 3 regions: front, middle and rear of the neutrophil. (C-E) Western blot and respective quantitative analysis of Syk (Tyr519/520) (72kDa), Paxillin (Tyr118) (68kDa) and Cortactin (Tyr421) (80kDa) phosphorylation after CXCL1 and PMA stimulation in wildtype and SFK-ko neutrophils plated on ICAM1. Band intensity was normalized to respective total protein. All data are presented as mean± standard error of the mean (SEM). *P<0.05; ***P<0.001; n.s.: not significant (one-way ANOVA, Dunnett’s multiple comparisons test). N=3 wildtype and 3 SFK-ko mice.
Figure 4. Rab27a dependent vesicle transfer is impaired in Src family kinases (SFK)-knockout (ko) neutrophils, leading to an impaired basement membrane penetration. (A) Neutrophil transmigration in a transwell assay with or without CXCL1 through filters coated with laminin (LN), or LN, PECAM-1, and ICAM-1 (LN/P/I). All data are presented as mean ± standard error of the mean (SEM). *P<0.05; ***P<0.001; n.s. : not significant (two-way ANOVA, Sidak multiple comparison test). N=3 wildtype and 3 SFK-ko x Lyz2GFP mice, transwell assays were performed in duplicates. (B) Maximum projections of confocal microscopic images of venules in TNFα-stimulated cremaster muscle whole mounts from wildtype and SFK-ko mice. Basement membrane was visualized by anti-LN5 (green), neutrophils by anti-MRP14 (red). Surrounding muscle tissue was visualized by bright field microscopy. White arrows point at extravasated, but still attached neutrophils in SFK-ko tissue. Scale bar: 20 μm. (C) and (E) immunostaining of representative wildtype and SFK-ko neutrophils on BSA- or PECAM-1/ICAM-1/CXCL1–coated (P/I/CXCL1) coverslips for (C) VLA6 and (E) VLA3 analyzed by confocal microscopy. Scale bar: 10 μm. (D) and (F) Quantification of ring formation for VLA6 (D) and VLA3 (F). At least 80 cells from 3 wildtype and 3 SFK-ko mice were analyzed for each condition. (G) Immunostaining of representative wildtype and SFK-ko neutrophils on BSA- or PECAM-1/ICAM-1/CXCL1–coated (P/I/CXCL1) coverslips for Rab27a, Munc13-4 and JFC1. Scale bar is 5μm. (H) Quantification of ring formation for Rab27a, Munc13-4 and JFC1. At least 80 cells from 3 wildtype and 3 SFK-ko mice were analyzed for each condition. All data are presented as means±SEM. *P<0.05; **P=0.005; ***P<0.001; n.s. : not significant (two-way ANOVA, Sidak multiple comparison test).
Impaired vesicle transport in the absence of Src family kinase in neutrophils

We and others have previously shown that mobilization of vesicles containing VLA3 and VLA6 to the plasma membrane is an important process during vascular BM penetration. To investigate whether SFK have a role in this process, we analyzed the SFK-dependent translocation of VLA6 and VLA3 to the plasma membrane of SFK-ko and wildtype neutrophils. Bone marrow derived neutrophils of SFK-ko and wildtype mice were plated on BSA or, again, on a combination of PECAM-1/ICAM-1 and CXCL1 and subsequently stained for VLA6 or VLA3. Plasma membrane translocation was visualized by confocal microscopy as “ring formation” (Figure 4C and E). We observed fewer SFK-ko cells (57%) displaying a ring of VLA6, when plated on PECAM-1/ICAM-1/CXCL1 compared to wildtype (55%) (Figure 4D). This observation was also true for VLA3, where only 52% of SFK-deficient neutrophils showed vesicle translocation, compared to 82% of wildtype cells (Figure 4F). To test whether ring formation can also be observed in vivo, whole mount cremaster muscle stainings were conducted for VLA6, which revealed a similar trend as in vitro. In wildtype whole mounts, extravasated neutrophils displayed a ring-like staining for VLA6, while intravascular neutrophils showed only a diffuse signal. In SFK-ko whole mounts, intravascular and extravasated cells showed no clear peripheral staining (Online Supplementary Figure S2C). Next, we plated wildtype neutrophils on BSA or PECAM-1/ICAM-1/CXCL1 and stained for SFK. Confocal micrographs revealed diffuse intracellular SFK staining when cells were plated on BSA (Online Supplementary Figure S2D). Upon stimulation with PECAM-1/ICAM-1/CXCL1, we observed a clear SFK signal at the cell border. Again, we quantified SFK distribution and observed a pronounced SFK translocation to the cell periphery when neutrophils were plated on PECAM-1/ICAM-1/CXCL1 (Online Supplementary Figure S2E). These experiments support a critical role for SFK by strongly interfering in vesicle trafficking of neutrophils in vitro and in vivo.

Src family kinase regulate Rab27a-dependent vesicle translocation

Vesicle trafficking in neutrophils is mainly regulated by Rab27a GTPases and their corresponding effector proteins. In neutrophils, Rab27a works in collaboration with two of its known effectors, JFC1 and Munc13-4, on vesicle trafficking and exocytosis of secretory vesicles and azurophilic granules. We analyzed the translocation of Rab27a, JFC1 and Munc13-4, on vesicle trafficking and exocytosis of secretory vesicles and azurophilic granules. We analyzed the translocation of Rab27a, JFC1 and Munc13-4, on vesicle trafficking and exocytosis of secretory vesicles and azurophilic granules.1 We analyzed the translocation of Rab27a, JFC1 and Munc13-4, on vesicle trafficking and exocytosis of secretory vesicles and azurophilic granules. We analyzed the translocation of Rab27a, JFC1 and Munc13-4, on vesicle trafficking and exocytosis of secretory vesicles and azurophilic granules.1 We analyzed the translocation of Rab27a, JFC1 and Munc13-4, on vesicle trafficking and exocytosis of secretory vesicles and azurophilic granules.1 We analyzed the translocation of Rab27a, JFC1 and Munc13-4, on vesicle trafficking and exocytosis of secretory vesicles and azurophilic granules.1 We analyzed the translocation of Rab27a, JFC1 and Munc13-4, on vesicle trafficking and exocytosis of secretory vesicles and azurophilic granules.1 We analyzed the translocation of Rab27a, JFC1 and Munc13-4, on vesicle trafficking and exocytosis of secretory vesicles and azurophilic granules.1 We analyzed the translocation of Rab27a, JFC1 and Munc13-4, on vesicle trafficking and exocytosis of secretory vesicles and azurophilic granules.1 We analyzed the translocation of Rab27a, JFC1 and Munc13-4, on vesicle trafficking and exocytosis of secretory vesicles and azurophilic granules.1 We analyzed the translocation of Rab27a, JFC1 and Munc13-4, on vesicle trafficking and exocytosis of secretory vesicles and azurophilic granules.1

NE-dependent laminin degradation is defective in Src family kinase-ko neutrophils

Neutrophil extravasation is not only dependent on the translocation of LN binding β1 integrins, but also on the serine proteinase neutrophil elastase (NE). We first analyzed the localization of NE in neutrophils plated on either BSA or PECAM-1/ICAM-1/CXCL1 as described above (Figure 5A). Similar to VLA3 and VLA6, we observed an increase in ring formation from 21.9% in BSA stimulated wildtype neutrophils to 59.1% of wildtype neutrophils stimulated on PECAM-1/ICAM-1/CXCL1 (Figure 5B). This upregulation of vesicle translocation was absent in SFK-ko neutrophils (20.0% for BSA vs. 31.2% for PECAM-1/ICAM-1/CXCL1 coating). In neutrophils, NE is stored in azurophilic granules together with a variety of other proteinases and antimicrobial proteins such as myeloperoxidase (MPO). Neutrophils release MPO after stimulation with TNFα. Thus, we additionally quantified MPO release in the serum of SFK-ko and wildtype mice by ELISA 2 h after TNFα stimulation. While we did not observe upregulation of MPO in the serum of SFK-ko mice 2 h after TNFα stimulation (158.4 ng/mL before vs. 184.0 ng/mL after TNFα stimulation) (Figure 5C), MPO levels were markedly increased in the serum of TNFα stimulated wildtype mice (113.0 ng/mL before vs. 473.5 ng/mL after TNFα stimulation). This indicates that SFK are required for the release of azurophilic granule content including NE and MPO. Earlier studies revealed that NE is able to proteolytically cleave laminins. In addition, NE activates different matrix metalloproteinases (MMP) that are involved in matrix degradation. However, the relevance of these NE functions for neutrophil transmigration are still not completely understood. By applying a NE-fluorescent activatable substrate (NE680FAST) in postcapillary venules with or without TNFα stimulation we were able to investigate NE activity in wildtype and SFK-ko cremaster whole mounts. Compared to unstimulated controls, TNFα stimulation led to a robust NE activity signal in wildtype mice (Figure 5D). As expected, this activity was reduced in SFK-ko venules, indicating that there is reduced NE dependent substrate cleavage in the absence of SFK (Figure 5D).

Next, we investigated whether wildtype neutrophils are able to degrade BM constituents under in vitro conditions. To this end, we performed transwell experiments in a system that allows us to image neutrophil crawling and extravasation by spinning disc confocal microscopy. Neutrophils from wildtype and SFK-ko mice were isolated and stained with different cell trackers (CellTracker™ Red CMTPX Dye and CellTracker™ Green CMFDA Dye,) to image both neutrophil populations in one transmigration chamber. In addition, laminin was visualized with an Alexa-647 coupled antibody. We observed intensive degradation of LN by wildtype neutrophils only, which could be identified by the disappearance of fluorescently labeled LN over time (Figure 5E, Online Supplementary Figure S3A and Online Supplementary Movie3). When we used SFK-ko neutrophils only, no degradation of LN was observed (Online Supplementary Figure S3B). Interestingly we found that SFK-ko neutrophils, which were unable to digest the LN layer, migrated strongly towards wildtype neutrophils during the observation period (Figure 5F and G and Online Supplementary Movie4). We quantified this association of SFK-ko neutrophils around wildtype cells in a randomization approach. Here, the interaction strength is a measure of the degree of dependence of spatial distribution between wildtype and SFK-ko neutrophils. We observed an interaction strength of 5.1 after 1,000 seconds.
Figure 5. Src family kinases (SFK)-dependent neutrophil elastase (NE) translocation is critical for degradation of the basement membrane. (A) Immunostaining of representative wildtype and SFK-knockout (ko) neutrophils on BSA- or PECAM-1/ICAM-1/CXCL1–coated (P/I/CXCL1) coverslips for NE analyzed by confocal microscopy. Scale bar: 10 μm. N=3 wildtype and 3 SFK-ko mice. (B) Quantification of ring formation for NE. At least 80 cells from 3 wildtype and 3 SFK-ko mice were analyzed. All data are presented as mean± standard error of the mean (SEM). *P<0.05; ***P<0.001; n.s.: not significant (two-way ANOVA, Sidak multiple comparison test). (C) Quantitative analysis for myeloperoxidase (MPO) release in blood plasma samples from wildtype and SFK-ko mice 2 hours after TNF-α injection by quantitative ELISA assay; 3 wildtype and 3 SFK-ko mice were analyzed. All data are presented as mean±SEM. *P<0.05; **P<0.005; ***P<0.001; n.s.: not significant. (D) NE activity (white) within cremaster muscle venules without (upper image) or with TNF-α middle and lower image) stimulation from wildtype and SFK-ko mice imaged by confocal microscopy. Venules were visualized using a CD31 antibody (red) (n=3 mice per group). Scale bar: 40 μm. (E) Spinning disk confocal micrographs of LN degradation (indicated by black areas compared to white LN staining) by wildtype neutrophils at indicated time points (Lower panels). White lines outline the increase in degradation area over time (Upper panels). Scale bar: 10 μm. (F) Spinning disk confocal micrographs of wildtype (green) and SFK-ko (red) neutrophils plated on antibody-labeled LN (white) at indicated time points. Scale bar: 10 μm. (G) Randomization approach to test for wildtype/SFK-ko interaction strength during migration using the nearest-neighbor analysis. (H) Multi photon microscopy of TNF-α-stimulated SFK-ko x Lyz2GFP or SFK-ko x Lyz2GFP mice with 1x10^7 injected deep red labeled wildtype neutrophils. Venules were visualized using a CD31 antibody (turquoise). Dotted squares indicate areas of interest (see panel I) (n=4 mice per group). Scale bar: 30 μm. (I) Close-up of vessel wall of SFK-ko x Lyz2GFP or SFK-ko x Lyz2GFP mice with 1x10^7 injected deep red labeled wildtype neutrophils.
of observation (Online Supplementary Figure S3C and D), indicating that this accumulation is not random, but highly dependent. These experiments strongly support the concept of SFK-dependent BM degradation by extravasating wildtype neutrophils. In addition, our results indicate that LN degradation fragments might exert chemotactic activity.

To finally show that these in vitro findings are also relevant under in vivo conditions, we injected labeled (CellTracker™ Deep Red Dye) SFK-ko or wildtype neutrophils into Lyz2GFP or SFK-ko x Lyz2GFP mice, respectively, and observed neutrophil behavior using multi-photon microscopy of the mouse cremaster muscle vasculature. Compared to SFK-ko x Lyz2GFP mice, where neutrophils accumulated in the vessel or were stuck to the abluminal side, we observed strongly elevated numbers of extravasated neutrophils in the periphery, when wildtype neutrophils were present (Figure 5H and Online Supplementary Movies 5 and 6). Additionally, we were able to visualize SFK-ko x Lyz2GFP neutrophils accumulating around adherent wildtype cells, confirming the observed behavior of SFK-ko neutrophils in the in vitro setting (Figure 5I). Also, in the experiment in which we inverted the procedure, by injecting fluorescently labeled SFK-ko neutrophils into Lyz2GFP mice, we could observe extravasated SFK-ko neutrophils (Online Supplementary Figure S3E and Online Supplementary Movies 7 and 8). These experiments strengthen the concept of wildtype neutrophils creating a path through the BM which enables SFK-ko neutrophils to extravasate into inflamed tissue in vivo.

Discussion

Integrin outside-in signaling is critical for neutrophil firm adhesion to and extravasation through the vessel wall into inflamed tissue. The short intracellular integrin tail has no enzymatic activity and acts exclusively as a binding site for recruited proteins, which, in turn, activate a broad range of pathways, spanning from cytoskeletal rearrangements and integrin clustering to vesicle trafficking. It has been widely accepted that tyrosine phosphorylation of ITAM motives by members of the Src family kinases is one way of integrin outside-in signaling. Therefore, SFK triple knockout mice (Hck-/-Fgr-/-Lyn-/-), here named SFK-ko) have been used to study the function of these kinases in neutrophils in an in vivo model of acute inflammation. Interestingly, Hck/Fgr^-/- neutrophils displayed no adhesion defect in earlier static adhesion experiments on ICAM-1, while our findings show a dramatic decrease in neutrophil adhesion efficiency in inflamed cremaster muscle venules in vivo implying that SFK-dependent neutrophil adhesion is sensitive to shear stress. We also found that neutrophil extravasation in SFK-deficient mice was dramatically decreased when compared to wildtype mice, which was caused by an SFK-dependent intrinsic extravasation defect. This observation is in line with various reports in different inflammation models with SFK-ko mice where decreased numbers of neutrophils were observed. Lowell et al. linked low PMN numbers in the liver during endotoxemia to a failure of Hck/Fgr^-/- neutrophils to rearrange their actin cytoskeleton, while cytokine production was unchanged. We clearly demonstrate that in the absence of SFK, neutrophils are unable to strongly adhere to the substrate under flow and detach in vitro and in vivo, due to defective LFA1 clustering and poor adhesion strengthening. The application of the tyrosine inhibitor Dasatinib led to a comparable shear rate dependent decrease in adherent neutrophils in inflamed venules in vivo. This strengthens previous findings of an improved outcome during sepsis after Dasatinib application, where leukocyte infiltration into the inflamed peritoneum was reduced in a dose-dependent manner.

Interestingly, most in vitro studies were performed under steady state conditions, where integrin clustering and crawling were unaffected in SFK deficient leukocytes, neglecting the impact of shear stress on neutrophil adhesion. This phenomenon was previously observed in WASP- and mAbp1-deficient mice, where integrin clustering and migration defects only arose when neutrophils were analyzed under shear stress. Our study demonstrates that neutrophil adhesion strengthening under flow conditions is dependent on SFK which mediate clustering of integrins, the rearrangement of the actin cytoskeleton along with polarization of the cell. Several cytoskeleton associated proteins required for adhesion are reported phosphorylation targets of SFK (directly or indirectly via Syk) like Paxillin and Cortactin. Here we clearly show reduced phosphorylation of these adhesion-relevant proteins.

We also aimed to answer the question as to whether SFK are required for the extravasation process beyond the endothelial layer. It was suggested that leukocytes predominantly cross the vascular BM at LN low expression regions (LER), and recent publications, including those from our lab, showed that neutrophils need to translocate venules containing VLA3, VLA6 and NE to their surface in a MST1-Rab27a dependent manner in order to overcome the BM. Our results identify for the first time SFK to be an additional critical component of this process. We show that SFK-ko neutrophils fail to pass an artificial LN barrier, and even when stimulated with PECAM-1 and ICAM-1, SFK-ko neutrophils are unable to release aurophilic granules and translocate integrin containing secretory vesicles. Exocytosis and vesicle transfer in neutrophils is strongly regulated by the GTPase Rab27a and its two effectors JFC1 and Munc13-4. We demonstrate that PECAM-1/ICAM-1 and CXCL1 stimulation facilitates the translocation of all three proteins to the membrane of wildtype neutrophils, suggesting that SFK regulate vesicle transport in a Rab27a-dependent manner. How MST1 and SFK co-operate in this process is still unclear and needs further investigation. Of note, MPO release was not defective in MST1 deficient neutrophils (M Sperandio, 2019, unpublished data/personal communication) compared to SFK-ko neutrophils, where a marked impairment in MPO release could be observed, suggesting that SFK are involved in neutrophil vesicle trafficking independently of MST1.

The role of NE during neutrophil extravasation is still controversial, as is the role of BM degradation by elastases or MMPs. However, recent findings suggest that this might, at least in part, be related to the experimental model used. Reichel et al. could clearly demonstrate decreased extravasation into inflamed cremaster muscle tissue after blockage of gelatinases by the specific inhibitor MMP-2/-9 inhibitor III. Furthermore, Young et al. were able to show NE-dependent extravasation using the same model. It was also suggested that NE/MMP-
dependent LN degradation creates peptides that are chemoattractant for neutrophils. In line with these findings, we were able to show diminished LN degradation by SFK-ko neutrophils in *vivo* using spinning disk confocal microscopy. Surprisingly, we observed that SFK-ko neutrophils, which are unable to degrade the LN barrier themselves, arrange around wildtype neutrophils, strongly supporting the concept of neutrophil-mediated vascular BM digestion along this route.

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