Variations in Alveolar Partial Pressure for Carbon Dioxide and Oxygen Have Additive Not Synergistic Acute Effects on Human Pulmonary Vasconstriction

Quentin P. P. Croft1, Federico Formenti1, Nick P. Talbot1, Daniel Lunn2, Peter A. Robbins1, Keith L. Dorrington1*

1 Department of Physiology, Anatomy and Genetics, University of Oxford, Oxford, United Kingdom, 2 Department of Statistics, University of Oxford, Oxford, United Kingdom

Abstract

The human pulmonary vasculature constricts in response to hypercapnia and hypoxia, with important consequences for homeostasis and adaptation. One function of these responses is to direct blood flow away from poorly-ventilated regions of the lung. In humans it is not known whether the stimuli of hypercapnia and hypoxia constrict the pulmonary blood vessels independently of each other or whether they act synergistically, such that the combination of hypercapnia and hypoxia is more effective than the sum of the responses to each stimulus on its own. We independently controlled the alveolar partial pressures of carbon dioxide (PACO₂) and oxygen (PAO₂) to examine their possible interaction on human pulmonary vasoconstriction. Nine volunteers each experienced sixteen possible combinations of four levels of PACO₂ (+6, +1, −4 and −9 mmHg, relative to baseline) with four levels of PAO₂ (175, 100, 75 and 50 mmHg). During each of these sixteen protocols Doppler echocardiography was used to evaluate cardiac output and systolic tricuspid pressure gradient, an index of pulmonary vasoconstriction. The degree of constriction varied linearly with both PACO₂ and the calculated haemoglobin oxygen desaturation (1-SO₂). Mixed effects modelling delivered coefficients defining the interdependence of cardiac output, systolic tricuspid pressure gradient, ventilation, PACO₂ and SO₂. No interaction was observed in the effects on pulmonary vasoconstriction of carbon dioxide and oxygen (p>0.64). Direct effects of the alveolar gases on systolic tricuspid pressure gradient greatly exceeded indirect effects arising from concurrent changes in cardiac output.

Introduction

The human pulmonary vasculature constricts in response to both hypercapnia and hypoxia [1–4]. Sometimes, variations in CO₂ and O₂ are such as to work in synchrony on the vasculature. For example, this occurs in a poorly ventilated region of the lung where they both act to direct blood flow away from the region to better ventilated lung tissue, thereby enhancing the efficiency of gas exchange [5]. At other times, variations in CO₂ and O₂ are such as to act in opposition on the vasculature. An example is human exposure to high altitude, where the whole lung is exposed to coexisting hypoxia and hypocapnia [6], and the potentially harmful pressor effect of the alveolar hypoxia is obviated by the dilatory effect of the alveolar hypocapnia. It is not known in what way a combination of the stimuli of hypercapnia and hypoxia affect the blood vessels in the human lung. It is unclear, therefore, whether the effects of the stimuli are additive or synergistic, that is to say, whether variations in O₂ could potentially enhance the response to CO₂ or vice-versa.

The question of whether there is a synergy between the effects CO₂ and O₂ in the sensing mechanisms of the pulmonary vasculature is of broader interest than in the context of this tissue alone. In relation to the mammalian carotid body a stimulus interaction in the responses of single afferent fibres to CO₂ and O₂ has been known since 1975 [7], and considerable attention has been directed at establishing at what cellular level of transduction this synergy might occur [8,9]. The important consequences of this stimulus interaction on the control of breathing in humans in a wide variety of conditions has been recognized for many years [10,11]. In comparison, responses of pulmonary vascular smooth muscle to the combined stimuli CO₂ and O₂ have received little attention, but are arguably of a similar importance for understanding the behaviour of the lung in health and disease [12,13]. Animal preparations have not provided a clear indication of what one might expect for the human lung. Most, but not all [14], preparations show vasomotor responses to both respiratory gases, with some degree of synergistic interaction between the effects of CO₂ and O₂ being common but variable [15–20]. Study of vasoconstrictor responses in the in vivo healthy human lung is made particularly difficult by the fact that changes in PACO₂ and PAO₂ induce changes in pulmonary artery pressure and pulmonary vascular resistance (PVR) that are a summation of a direct active effect of the gases on vascular smooth muscle and an indirect passive effect of concurrent changes in pulmonary blood flow and,
potentially, ventilation [21]. The indirect effect may be quite small, because pulmonary vessels tend to be quite distensible, and thus accommodate large changes in flow with little rise in perfusion pressure and with a fall in resistance. This nevertheless makes it misleading to measure either pulmonary artery pressure or PVR, as a sole index of pulmonary vascular smooth muscle constriction.

The luxury available in animal preparations of being able to impose a constant pulmonary flow, and using pulmonary artery pressure or PVR as the index of vasoconstriction, has not been achieved in humans [22]. We address this problem by using mixed effects modelling to extract coefficients in direct and indirect pathways linking $P_{A\text{CO}_2}$ and $P_{A\text{O}_2}$ with pulmonary artery pressure, and the relative contribution of each pathway. Direct effects of alveolar gases on pulmonary artery pressure are found to dominate. This approach also evaluates whether the gases have an additive or synergistic action; an additive action is observed, consistent with the approach adopted in an earlier model of feedback control of regional gas exchange in the human lung [13].

**Methods**

**Ethics Statement**

The study was approved by the Oxfordshire Research Ethics Committee and performed in accordance with the Declaration of Helsinki. Informed written consent was obtained from all volunteers.

**General approach to the measurement of pulmonary vasoconstriction**

The general approach adopted was to use non-invasive measurement of systolic pulmonary artery pressure as our index of pulmonary vasoconstriction, whilst at the same time taking into account the dependence of this pressure upon other variables: ventilation and cardiac output. This separation of direct and indirect influences of $P_{A\text{CO}_2}$ and $P_{A\text{O}_2}$ on systolic pulmonary artery pressure was achieved using mixed effects modelling.

**Volunteers**

Nine healthy volunteers (5 women and 4 men), aged 24±4 years and with BMI 22.5±2 kg/m² (mean ± S.D.), completed the study. Female volunteers were asked to participate only during the first 14 days of their menstrual cycle. Volunteers visited the laboratory before undergoing the experimental protocols in order to discuss the procedures and confirm that they were suitable for echocardiographic assessment of tricuspid regurgitation.

**Study design**

The pulmonary vascular response to four different levels of $P_{\text{ETCO}_2}$ was studied at each of four different levels of $P_{\text{ETO}_2}$. This led to 16 different combinations of $P_{\text{ETCO}_2}$ and $P_{\text{ETO}_2}$ overall, each called a protocol. Each protocol comprised a ten-minute exposure to the particular $P_{\text{ETCO}_2}/P_{\text{ETO}_2}$ combination which was preceded by 5 min of baseline conditions (see below). Cardiovascular and respiratory variables were measured throughout each protocol.

Each volunteer completed the sixteen protocols in one of four different orders, determined by block randomization based on date of first contact. Volunteers completed these protocols in two batches of eight in two afternoons. Each protocol was preceded by at least ten minutes of quiet rest. The sixteen protocols were the sixteen combinations of four levels each of end-tidal partial pressures of CO$_2$ ($P_{\text{ETCO}_2}$) and O$_2$ ($P_{\text{ETO}_2}$). These end-tidal values were assumed to be equivalent to alveolar partial pressures. The following four levels of $P_{\text{ETCO}_2}$ were chosen (relative to normal baseline): +6, +1, −4 and −9 mmHg. The levels of $P_{\text{ETO}_2}$ used were 175, 100, 75 and 50 mmHg. This provided an opportunity to span the range from relative hyperoxia to the hypoxia used in other studies [23–25], and so cover the likely regional values for these variables encountered within the healthy lung at sea level [13,26].

**Gas control**

$P_{\text{ETCO}_2}$ and $P_{\text{ETO}_2}$ were controlled using an end-tidal forcing system as previously described [27–29]. Volunteers lay in a semi-left lateral position and breathed through a mouthpiece with the nose occluded. Ventilatory volumes and flows were measured by turbine and pneumotachograph respectively. Gases were sampled by a catheter close to the mouth and analysed continuously by mass spectrometry.

Ventilation during the protocols conducted at $P_{\text{ETCO}_2}$ values of −9 and −4 mmHg was achieved by voluntary hyperventilation. Volunteers controlled the frequency of breathing through the use of an audible metronome, and the depth of breathing through feedback presented on an oscilloscope connected to the output of the turbine measuring ventilatory flows. Ventilation during the protocols conducted at $P_{\text{ETCO}_2}$ values of +6 and +1 mmHg was spontaneous. Each protocol consisted of 5 min of spontaneous ventilation, or voluntary hyperventilation, with end-tidal gases held constant at baseline values (100 mmHg $P_{\text{ETCO}_2}$ and the measured baseline $P_{\text{ETCO}_2}$) followed by ten minutes with these gases at the specified levels for the protocol. For protocols involving hypocapnia, a constant combination of breathing depth and frequency was used throughout.

**Echocardiography**

In approximately 70% of healthy volunteers it is possible to detect with Doppler ultrasound a regurgitant blood flow from the right ventricle to the right atrium during ventricular systole. Measurement of the peak velocity (v) of this regurgitant jet affords an opportunity to estimate the systolic pressure difference $\Delta P_{\text{max}}$ between the right ventricle (where the pressure is close to pulmonary artery systolic pressure) and right atrial pressure. This relationship is given by the Bernoulli equation: $\Delta P_{\text{max}} = \rho v^2/2$, where $\rho$ is blood density. The peak systolic tricuspid pressure gradient ($\Delta P_{\text{max}}$) and cardiac output were measured using a GE Vivid-i ultrasound machine with a S4 transducer (2–4 MHz). Assessment of $\Delta P_{\text{max}}$ used Doppler echocardiography, via a 4-chamber view of the heart, to measure the peak pressure difference between the right ventricle and the right atrium during systole. Since right atrial pressure changes little during hypocxia, changes in $\Delta P_{\text{max}}$ reflect changes in systolic pulmonary arterial pressure [30,31]. The utility of measuring $\Delta P_{\text{max}}$ as an index of pulmonary vascular constriction in healthy humans has been shown during hypoxia [24,25], hypercapnia and hypocapnia [2,13].

Cardiac output (Q) was measured using Doppler echocardiography to assess non-turbulent flow through the centre of the left ventricular outflow tract (LVOT). The cross-sectional area of the LVOT was obtained by measuring the diameter of the aortic valve using a parasternal long-axis view of the heart. Flow through the LVOT was imaged using an apical five-chamber view of the heart and measured using the velocity-time integral. Systolic flow was multiplied by the cross-sectional area of the LVOT to provide an estimate of stroke volume. Heart rate was recorded simultaneously. The stroke volume was multiplied by the heart rate to provide an estimate of cardiac output.

For both measurements, results depend to some extent upon the phase of the respiratory cycle, so end-expiration was chosen as the phase of that cycle giving minimal disturbance; images of the
spectral traces at or as near as possible to end-expiration were saved digitally for later analysis.

Data analysis

Ventilation (VE) and end-tidal gases were assessed using 30 s averages of the values calculated from each breath. For ΔPmax and Q, approximately five measurements of each variable were obtained each minute and then 2 min averages were calculated.

Baseline variables were the average of values recorded during the first five minutes of each protocol. Protocol variables were the average of the last six minutes of each protocol. The change in each variable was the difference between the protocol and baseline values.

Petco2 values were converted to an equivalent fractional oxyhaemoglobin saturation (SO2) using the equation provided by Severinghaus [32]. Although the major stimulus to pulmonary vascular constriction is the partial pressure of the sensed gases, the response to oxygen is known to be markedly non-linear and the purpose of this sigmoid transformation was to permit us to use a virtual saturation in place of PO2 in our analysis, and thereby assess the suggestion of previous authors [33] that hypoxic constriction tends to be a linear function of SO2 whilst being a markedly curvilinear function of PO2.

Modelling and statistical analysis

The experimental data were analysed using the following linear model:

\[ \Delta P_{\text{max}} \text{ protocol value} = a + \beta(B \Delta P_{\text{max}}) + \eta(B \text{SO}_2) + a(d \text{SO}_2) + b(B \text{PETCO}_2) + \mu(d \text{SO}_2^* d \text{PETCO}_2) + \gamma(B \ln V_E) + h(\Delta \ln V_E) \]

where \( B \Delta P_{\text{max}}, B \text{SO}_2, B \text{PETCO}_2, BQ \) and \( B \ln V_E \) refer to baseline values of the respective variables \( \Delta P_{\text{max}}, \text{SO}_2, \text{PETCO}_2, \text{Q} \) and \( \ln V_E \), whilst \( d \text{SO}_2, d \text{PETCO}_2, \Delta Q \) and \( \Delta \ln V_E \) refer to the differences between protocol and baseline values. \( \Delta \text{SO}_2^* \Delta \text{PETCO}_2 \) allows for possible interaction between the stimuli. The logarithm of \( V_E \) was required in the analysis instead of \( V_E \) itself so as to avoid giving undue dominance to a small number of high values of \( V_E \). The coefficients preceding each term were obtained by fitting the model to the experimental data.

Figure 1 shows the conceptual framework for our modelling approach. \( \Delta P_{\text{max}} \) is viewed as primarily a measure of pulmonary vasoconstriction dependent upon a direct effect of alveolar gases on vascular smooth muscle, whilst also being a weak function of Q and VE. These in turn are functions of alveolar gases, and provide an indirect route via which alveolar gases can change \( \Delta P_{\text{max}} \). The modelling described below delivers mean values plus confidence intervals, expressed as standard error of these means, to the nine coefficients displayed in Figure 1, as well as assessing the significance of the interactive term \( \Delta \text{SO}_2^* d \text{PETCO}_2 \) in Eq. 1.

The data were analysed with linear mixed effects modelling to account for correlation within individual volunteers and for variability between volunteers. A two-level multilevel model with an exchangeable correlation structure was fitted. This statistical technique can be used for analysing data that occur as repeated measurements on each of a number of participants in order to identify and quantify responses common to all participants, taking into account individual variability, with no two individuals being the same. Models similar to that in Eq. 1 were derived for Q and \( \ln(V_E) \).

Data were analysed using ‘R’, open-source computer software for statistical analyses. R uses a penalised likelihood method to fit the data to a given model iteratively until no improvement in the residual deviance is achieved. Data were initially fitted to a model in which all of the possible contributing factors in Eq. 1 were considered. The model was then adjusted to exclude the least significant factor until all remaining factors showed significance with \( p<0.05 \). This provided individual coefficients for each contributing factor that define the linear relationships. Each coefficient was then fitted as a random variable, with the mean and standard deviation estimated from the data, retaining adjustments that enhanced the explanatory power of the model. This was judged by two methods: first, if the random factor correlated well with another random factor then no additional

Figure 1. Diagram of the relationships involved in the study. ΔPmax is viewed as the primary measure of pulmonary vasoconstriction, influenced directly by alveolar gases (paths a and b), whilst also being a weak function of cardiac output and possibly ventilation (paths g and h). The latter two are also functions of alveolar gases (via the pathways c-f). Interactions are not represented.

doi:10.1371/journal.pone.0067886.g001
explanatory power was added, the variability being explicable by one of the two factors. The constant in the model (which provides the y-axis intercept on a graph of the function) was always modelled as a random factor, and if it correlated well with another random factor then it was acting as a surrogate for that factor and the factor could be subsumed by the intercept factor. Secondly, if the residual deviance was not decreased by a large amount then the explanatory power was not enhanced, and the addition of a random factor was not necessary.

Results

Protocols were conducted between August 2007 and June 2009. Figure 2 shows representative data from two protocols that illustrate spontaneous ventilation during hypercapnia and controlled ventilation to induce hypocapnia. The left panel shows a protocol involving hypoxia with hypercapnia, and the right panel shows a protocol involving hypocapnia with hyperoxia. The upper panels show the control of PETO₂ and PETCO₂ for the two protocols; gas control achieved a rapid (<1 min) step from euoxia and eucapnia to protocol values and little variation from target end-tidal values either side of the change. The middle panels show the ventilations and cardiac outputs achieved during the protocols and
SO2* and fit, suggesting that they had no significant role in determining sequentially, in that order, without significantly worsening the individual changes in DCO2, the errors in gas control are well below 0.1 mmHg, whilst for O2, the errors in gas control are around 1 mmHg. Table 2 gives the individual changes in ΔPmax for each of the sixteen protocols.

Results of statistical analysis

A major objective of this study was to investigate whether the stimuli of hypercapnia and hypoxia constrict the pulmonary blood vessels independently of each other, or whether they act synergistically; in other words, evidence of an interaction was sought.

The main analysis used the model given in Eq. 1. Of the included factors baseline Q˙, baseline PETCO2, baseline V˙E, baseline SO2 and ΔSO2*ΔPETCO2, were all removed from the model sequentially, in that order, without significantly worsening the fit, suggesting that they had no significant role in determining ΔPmax protocol value. The interactive term was insignificant at the level p>0.64.

To ensure the study had sufficient power to detect any interaction between the effects of hypoxia and hypercapnia, we calculated power as a function of the percentage change in the ΔPmax response attributable to the interaction term (ΔSO2*ΔPETCO2). At the 5% significance level, the study had a power of 80% for the detection of a 4% change in the ΔPmax response due to interaction; the power for detecting a 10% change in the response was close to 100%. Despite adequate power, no evidence of an interaction was identified.

The final model fitted the following equation:

$$\Delta P_{\text{max}} \text{ protocol value} = a + b(\Delta B A \text{P}_{\text{max}}) + a(\Delta S O_2) + b(\Delta P_{E \text{TCO}_2}) + g(\Delta Q)$$

where the coefficients are given in Table 3 as a value ± standard error. The model that best explains the experimental data delivers

### Table 1. Errors (mean and standard deviation) in control of end-tidal gases calculated as the measured end-tidal partial pressure minus the target end-tidal partial pressure for the four levels of CO₂ and four levels of O₂ used in the study.

| Target PCO₂ (mmHg) | -9 | -4 | 1 | 6 |
|--------------------|----|----|---|---|
| Error              | 0.023 | -0.008 | 0.018 | -0.051 |
| SD                 | 0.325 | 0.256 | 0.255 | 0.352 |

| Target PO₂ (mmHg) | 175 | 100 | 75 | 50 |
|-------------------|-----|-----|----|----|
| Error             | -1.315 | -1.201 | 0.610 | 0.813 |
| SD                | 1.792 | 0.766 | 1.100 | 1.557 |

### Table 2. Individual changes in systolic tricuspid pressure gradient (ΔPmax) in response to sixteen combinations of end-tidal gas composition.

| End-tidal PO₂ (mmHg) | 50 mmHg | 75 mmHg | 100 mmHg | 175 mmHg |
|----------------------|---------|---------|----------|----------|
| Change in end-tidal PCO₂ (mmHg) | +6 | +1 | -4 | -9 | +6 | +1 | -4 | -9 | +6 | +1 | -4 | -9 |
| Subject 1662         | 12.7 | 10.4 | 3.1 | 2.5 | 10.6 | 3.7 | 2.0 | 0.8 | 2.6 | 1.4 | 1.0 | -1.2 | 5.6 | 0.1 | 0.1 | 0.5 |
| Subject 1664         | 3.9 | 3.5 | 3.7 | 3.4 | 0.0 | 0.0 | 0.0 | 0.1 | 3.0 | 1.9 | 0.3 | -1.7 | 2.4 | 1.3 | -0.6 | -0.7 |
| Subject 1701         | 6.7 | 6.1 | 5.1 | 2.5 | 3.6 | 0.9 | 0.6 | 1.1 | 3.0 | 1.3 | -0.5 | -0.8 | 1.7 | -0.2 | -0.2 | 0.4 |
| Subject 1703         | 5.9 | 1.6 | 3.7 | 1.9 | 5.5 | 2.0 | 0.0 | 1.5 | 2.8 | 1.9 | 2.5 | 1.0 | 3.4 | 0.5 | 0.2 | 2.1 |
| Subject 1714         | 9.7 | 10.7 | 6.6 | 7.3 | 7.8 | 1.8 | 2.3 | -0.8 | 5.8 | 0.1 | -0.2 | 0.6 | 2.4 | -0.7 | -0.4 | 0.1 |
| Subject 1719         | 15.1 | 12.6 | 15.0 | 13.8 | 4.7 | 3.6 | 5.2 | -1.6 | 3.9 | 0.0 | -0.7 | 0.8 | 3.2 | 0.0 | 1.4 | -2.4 |
| Subject 1730         | 4.2 | 6.0 | 3.8 | 2.0 | 2.9 | 1.1 | -1.7 | 2.2 | 3.6 | 1.2 | 1.0 | 1.1 | 0.8 | -0.7 | -2.6 | -1.8 |
| Subject 1096         | 12.4 | 12.7 | 9.9 | 7.2 | 2.6 | 0.8 | -1.8 | -0.7 | 3.2 | -1.0 | -0.2 | -1.9 | 1.0 | -1.5 | -2.5 | -0.9 |
| Subject 1751         | 6.8 | 7.9 | 4.7 | 5.4 | 1.2 | -0.1 | 1.5 | 1.3 | -0.7 | 0.6 | 1.5 | -0.8 | 0.1 | -0.1 | -0.7 | -1.3 |
| Mean                 | 8.6 | 7.9 | 6.2 | 5.1 | 4.3 | 1.6 | 0.9 | 0.4 | 3.0 | 0.8 | 0.5 | -0.3 | 2.3 | -0.1 | -0.6 | -0.6 |
| SEM                  | 1.3 | 1.3 | 1.3 | 1.3 | 1.1 | 0.5 | 0.7 | 0.4 | 0.6 | 0.4 | 0.3 | 0.4 | 0.4 | 0.4 | 0.3 | 0.4 |

Volunteers were exposed to each combination of end-tidal PO₂ and PCO₂ for 10 min, preceded by 5 min baseline breathing with end-tidal gases held close to baseline values (100 mmHg end-tidal PO₂ and the measured baseline end-tidal PCO₂). The change in peak systolic tricuspid pressure gradient (ΔPmax) was calculated as the difference between the mean baseline ΔPmax and the mean ΔPmax during the last 6 minutes of each protocol. Gas control was achieved by means of end-tidal forcing.

doi:10.1371/journal.pone.0067886.t002
coefficients $\beta$ and $b$ as fixed coefficients with $\alpha$, $a$ and $g$ as coefficients that vary between individuals with normal distribution and standard deviations of 0.59 mmHg, 0.26 mmHg/%desaturation and 0.36 mmHg/l/min, respectively.

The usual linear regression assumptions of normality and constant variance are confirmed by plotting the residuals against the fitted values (Fig. 3A) and inspection of a normal residuals-quantile plot (Fig. 3B). The purpose of the former plot is to show whether variance changes throughout the range of data, which would appear as a trend for the residuals to deviate from 0 as a function of the fitted values. One or two outliers on a dataset of this size are to be expected and are not necessarily inconsistent with a good fit. The latter plot shows whether the data are approximately normal, an assumption which is violated to the extent that the plot deviates from being linear.

The independent effects of altered PETCO2 and SO2 on Q were modelled using the same approach. The analysis fitted the equation:

$$Q\text{ protocol value} = \gamma + \varepsilon(B\text{ }Q) + i(A\text{ }\ln V E) + \varepsilon_t(A\text{ }\Delta SO_2) + f(A\text{ }\Delta PETCO2)$$

(3)

where the coefficients are given in Table 3. The model delivered $\varepsilon$, $i$, $f$ and $f$ as fixed coefficients, whilst $\gamma$ was taken to be normally distributed with a standard deviation of 0.08 l/min.

A similar approach was used for lnVe. Data from protocols involving hypocapnia were excluded from this analysis because Ve was consciously controlled in these protocols in order to achieve hypocapnia. The final model for Ve derived the following equation:

$$\ln(V E)\text{ protocol value} = \lambda + \zeta(B\text{ }\ln(V E)) + c(A\text{ }\Delta SO_2) + d(A\text{ }\Delta PETCO2)$$

(4)

where the coefficients are given in Table 3. The model delivered $\zeta$, $c$ and $d$ as fixed coefficients, whilst $\lambda$ was taken to be normally distributed with a standard deviation of 0.22 l/min.

Figure 4 gives the results for the coefficients defined in Fig. 1, and summarizes direct and indirect pathways via which O2 and CO2 influence APmax. For both gases, the direct pathway dominates.

### Discussion

The main finding of this study is that the effects of CO2 and O2 on human pulmonary artery pressure are additive rather than synergistic. Specifically, the retention in the model for systolic pulmonary artery pressure of a term incorporating the product of oxyhaemoglobin saturation and carbon dioxide partial pressure could not improve the predictive power of the model. An additional finding is that the direct effects of alveolar gases on pulmonary artery pressure via vasoconstriction dominate the indirect effects that come about via changes in ventilation and cardiac output.

Methods for measuring pulmonary vasoconstriction in vivo are controversial. In reduced preparations, typically perfusions of non-human animal lungs or vessels in vitro, it is common to manipulate pulmonary flow to be constant and then use either the pressure drop across the pulmonary circulation or PVR as measures of vascular ‘tone’ or ‘constriction’ [34,35]. An alternative approach is to maintain perfusion pressure constant, and associate changes in vascular constriction with changes in blood flow [20,36]. In awake humans neither of these approaches has proved accessible, and measurements of pulmonary vasoconstriction are complicated by the fact that both pulmonary arterial pressure and pulmonary blood flow usually change in response to changes in alveolar gases. A common invasive strategy has been to measure PVR using a Swan-Ganz pulmonary artery catheter, whilst accepting that changes in PVR occur independently in response to changes in both cardiac output [37] and alveolar gas composition [38]. This study demonstrates that the non-invasive measurement of systolic pulmonary artery pressure using Doppler ultrasound is a useful tool to assess vasoconstriction in response to changes in alveolar gases, as long as account is taken, with catheter measurements, of the separate effect of cardiac output on this variable.

### Comparison of pulmonary vascular response with previous human studies

Fig. 4[B] suggests for this study that 10–15% of the effect of alveolar gases on APmax occurs via indirect pathways. Two such pathways have been identified here: changes in cardiac output induced by changes in ventilation alone, and changes in cardiac output induced by CO2 and O2 in the absence of changes in ventilation. Few data are available from the literature for comparison. A study focusing on longer durations of hypoxia

| Protocol value | Intercept | Baseline | $\Delta$SO2 | $\Delta$PETCO2 | $\Delta$Q | $\Delta$lnVe |
|----------------|-----------|----------|--------------|---------------|-------------|-------------|
| APmax          | $\alpha$  | $\beta$  | $a$          | $b$           | $g$         | $h$         |
| mmHg           | 3.4±1.5   | 0.89±0.06| 0.43±0.09    | 0.18±0.03     | 0.66±0.32   | 0           |
| Q              | $\gamma$ | $\varepsilon$ | $e$       | $f$           | $i$         |
| l/min          | 1.1±0.3  | 0.79±0.06| 0.06±0.01    | 0.02±0.01     | 0.33±0.14   |
| ln(VE)         | $\lambda$ | $\zeta$ | $c$          | $d$           |             |
| l/min/mmHg     | 1.2±0.2 | 0.52±0.08| 0.039±0.004  | 0.099±0.009   |

Roman alphabet coefficients are depicted in Fig. 1. Roman and Greek coefficients are defined in Eqs. 2, 3 and 4. Coefficients are given as a value ± standard error.

doi:10.1371/journal.pone.0067886.t003
(0.5–8 h) found that approximately 5% of the rise in $\Delta P_{\text{max}}$ with hypoxia could be attributed to indirect effects via cardiac output [39].

The sensitivity of $\Delta P_{\text{max}}$ to acute changes in $Q$ is defined by coefficient $g$ in Eq. 2 and Fig. 1. The contribution of $Q$ alone is defined by $g = 0.66 \text{ mmHg/l/min}$. A previous study [39] observed spontaneous concurrent changes in $\Delta P_{\text{max}}$ with changes in $Q$ during air breathing in the absence of changes in alveolar gas composition and found a value for $g$ of $0.60 \text{ mmHg/l/min}$, in good agreement with that found here.

Other coefficients accessible from previous studies on similar human volunteers permit estimates for $e$ (0.06 l/min/%desat from hypoxic exposures [13,40]; here identically 0.06 l/min/%desat) and $f$ (0.04 l/min/mmHg from hypocapnic exposures at constant ventilation, [13]; here 0.02 l/min/mmHg).

**Figure 3. Plots of residuals for $\Delta P_{\text{max}}$ associated with model in Eq. 1.** (A) Residuals for $\Delta P_{\text{max}}$ plotted against the values for $\Delta P_{\text{max}}$ fitted to the model in Eq. 1. A skewed plot would show that the assumption of constant variance had been violated. No such pattern is discernible in this plot. (B) Residuals for $\Delta P_{\text{max}}$ plotted against the standardized expected quantiles (units of standard deviation) fitted to the model in Eq. 1. The linear relationship demonstrates that the residual deviances map on to a Normal distribution.

doi:10.1371/journal.pone.0067886.g003

**Limitations of the study**

The study measured changes in cardiopulmonary variables between 4 and 10 min after induction of new values of alveolar gases. A maximum exposure of 10 min to the perturbation in alveolar gas composition was chosen in part because of the difficulty experienced by volunteers in tolerating longer exposure to extremes such as combined hypoxia ($P_{\text{ETO}} = 50 \text{ mmHg}$) and hypercapnia ($P_{\text{ETCO}} = +6 \text{ mmHg}$). Previous work has suggested that this is a sufficiently long period in which to capture the initial acute phase of human hypoxic pulmonary vasoconstriction and the hypoxic increase in cardiac output, in which the time constants of the responses are around 2 min [40,41]. Recent work has found, however, that the time courses of the acute human cardiopulmonary responses to eucapnic hypercapnia and hypocapnia have time constants in the range 4–10 min [13], suggesting that the present experiments have measured a substantial but partial component of the acute changes in $\Delta P_{\text{max}}$ and $Q$ to changes in $P_{\text{Aco}}$. It is consequently difficult to obtain reliable...
estimates from previous studies for comparison with coefficients b and f in Eqs. 2 & 3; this may be why it is these coefficients that agree least well with estimates from previous studies. With regard to the coefficients relating to the cardiopulmonary responses to oxygen, values for a, g, & e show fair agreement with published values obtained from well-defined steady-state measurements.

A second limitation of the study arises from the requirement to establish voluntarily controlled ventilation for half of all measurements made, in order to achieve hypocapnia. The resulting halving of the number of data pertaining to the coefficients linking to ventilation in Fig. 1 will have reduced the precision with which coefficient i in Eq. 3 could be estimated and reduced the probability of detecting a small but non-zero value of the coefficient h linking DPmax directly with VE.

Thirdly, this study did not seek to understand the cellular basis for any interaction between CO2 and O2 in the pulmonary circulation, but instead to understand the effects of alveolar gas composition at the integrative level in humans. For example, we did not address the question of whether changes in pulmonary vascular tone result directly from alterations in PCO2, or whether they are secondary to the associated change in pH. This question has been addressed in animal studies, some of which suggest an effect of hypercapnia per se in the pulmonary vasculature [42,43], but further studies would be needed to explore this issue in humans.

**Physiological significance of the findings**

An accurate appreciation of the way in which the stimuli CO2 and O2 work together on pulmonary vessels is of importance to the understanding of situations in which they act in synchrony or in opposition. The spontaneous matching of perfusion to ventilation in the lung is usually modelled as being achieved solely by the vasoconstrictor effects of hypoxia on small pulmonary arteries [44,45], but the local vasoconstrictor effect of hypercapnia has the potential to enhance this matching [1,36]. It remains a possibility that the effects of hypoxia and hypercapnia acting only within an isolated small region of lung tissue might display a different, possibly interactive, relationship from the global effects on all lung tissue studied here. One possible reason for this is that the experiments subjected volunteers to relatively stressful perturba-
tions in end-tidal gas composition that might lead to global autonomic effects on the pulmonary circulation that would not occur with perturbations limited to small regions of lung tissue. Even on the assumption of additive, rather than interactive, effects of the two stimuli recent calculations suggest that CO₂ may play a more substantial role than O₂ in matching ventilation in the healthy lung at sea level [13]. Under conditions of therapeutic artificial ventilation, clinicians recognize the potential adverse effect on oxygenation of the patient of a low P<sub>sh</sub>, in a hyperventilated hypoxic lung leading to inhibition or elimination of hypoxic vasoconstriction in that lung [46,47], but the relative contributions of the stimuli have remained unclear.

Pulmonary hypertension at high altitude is associated with global hypoxia with hypocapnia throughout the lung [10] and appears to be responsible for high altitude pulmonary edema in patients who have an exaggerated vasconstrictor response [48]. It remains uncertain to what extent in affected individuals a weak vasodilatatory effect of hypocapnia might inadequately ameliorate the pulmonary hypertension that results from a strong vasconstrictor effect of hypoxia, because these stimuli have not been examined separately in this setting [49]. The human lung shows considerable potential to dilate in response to sustained hypocapnia [2], and it would clearly be beneficial at altitude for there to be a balance between the vasodilatatory effects of hypocapnia and the constriction brought about by hypoxia. The present experiments have quantified the extent of this balance for very acute responses in the period 4–10 min following a step change of alveolar gases. Further work is required to find whether the considerably more intense responses to more sustained combinations of CO₂ and O₂ stimuli, such as those occurring over hours and days at high altitude, combine in a similar additive manner.

A novel finding from the study has been the possibility of obtaining a quantitative estimate of the effect of V<sub>E</sub> on Q that is independent of the effects of alveolar gases, namely the coefficient i. The value of i = 0.33 l/min/h/(l/min) suggests a 0.33 l/min rise in cardiac output attributable to a 2.72-fold rise in ventilation. Another interpretation, assuming linearity over a broad range of ventilation, is that a rise in ventilation from a resting value of about 4.5 l/min to a twenty-fold value of 90 l/min associated with very vigorous exercise might contribute a rise in cardiac output of ~1 litre/min from the direct effect of ventilation on the cardiovascular system alone. Interestingly, ventilation alone appears to have no direct effect upon ΔPmax (i.e. h = 0). Further studies are required to establish the magnitude of these interrelationships over wider ranges of physiological disturbance.

Acknowledgments

We thank Mr David O’Connor and Dr Marziah Fatemian for their technical assistance and the volunteers for their participation.

Author Contributions

Conceived and designed the experiments: QPPC NPT PAR KLD. Performed the experiments: QPPC FNT. Analyzed the data: QPPC DL PAR. Wrote the paper: QPPC PAR KLD.

References

1. Visvanathan R, Lodi ST, Subramanian S, Radha TG (1976) Pulmonary vascular response to ventilation hypcapnia in man. Respiration 35: 165-176.
2. Balanos GM, Talbot NP, Dorrington KL, Robbins PA (2003) Human pulmonary vascular response to 4 h of hypcapnia and hypoxia measured using Doppler echocardiography. J Appl Physiol 94: 1543–1551.
3. Molty H, Gournaud A, Werko L, Himmelstein A, Dresdale D (1947) The influence of short periods of acute anoxia upon pulmonary arterial pressure in man. Am J Physiol 150: 315–320.
4. Carlsson AJ, Bindslev L, Santesson J, Gottlieb I, Hedenstierna G (1985) Hypoxic pulmonary vasoconstriction in the human lung: the effect of prolonged unilateral hypoxic challenge during anaesthesia. Acta Anaesthesiol Scand 29: 346–351.
5. Naeije R, Brimouille S (2004) Physiology in medicine: importance of hypoxic pulmonary vasoconstriction in maintaining arterial oxygenation during acute respiratory failure. Crit Care 5: 67–71.
6. Dehurt C, Berger MM, Mairbaurl H, Barsch P (2007) High altitude pulmonary edema: a pressure-induced leak. Respir Physiol Neurobiol 156: 266–273.
7. Lahiri S, DeLaney RG (1975) Stimulus interaction in the responses of carotid body chemoreceptor single afferent fibers. Respir Physiol 24: 249–266.
8. Roy A, Rozanov C, Mokashi A, Lahiri S (2000) P(O₂)-P(CO₂) stimulus interaction in [Ca(2+i)] and CSN activity in the adult rat carotid body. Respir Physiol 122: 13–26.
9. Peers C (2004) Interactions of chemostimuli at the single cell level: studies in a model system. Exp Physiol 89: 60–65.
10. Millelde JS, West JB, Schorner RB (2007) High Altitude Medicine & Physiology London: Hodder Arnold.
11. Lloyd BD, Buls NCM, Cunningham DFC (1958) The relation between alveolar oxygen pressure and the respiratory response to carbon dioxide in man. Quarterly J Exp Physiol 43: 214–227.
12. Howell K, Ooi H, Preston R, McCloughlin P (2004) Structural basis of hypoxic pulmonary hypertension: the modifying effect of chronic hypoxia. Exp Physiol 89: 66–72.
13. Dorrington KL, Balanos GM, Talbot NP, Robbins PA (2010) Extent to which pulmonary vascular responses to PCO₂ and PO₂ play a functional role within the healthy human lung. J Appl Physiol 108: 1084–1096.
14. Geant BJ, Davies EE, Jones HA, Hughes JM (1976) Local regulation of pulmonary blood flow and ventilation-perfusion ratios in the continuous. J Appl Physiol 40: 216–228.
15. Brimouille S, Lejeune P, Vachieri JL, Lerman M, Melot C, et al. (1990) Effects of acidosis and alkalosis on hypoxic pulmonary vasconstriction in dogs. Am J Physiol 258: H347–353.
16. Benumof JL, Wahrenbrock EA (1976) Blunted hypoxic pulmonary vasoconstriction by increased lung vascular pressures. J Appl Physiol 38: 846–1450.
17. Viles PH, Shepard, JT (1968) Relationship between pH, PO₂, and PCO₂ on the pulmonary vascular bed of the cat. Am J Physiol 215: 1170–1176.
18. Von Euler US, Ljøstad G (1946) Observations on the pulmonary arterial pressure in the cat. Acta Physiol Scand 12: 311–320.
19. Shirai M, Sada K, Ninomiya I (1986) Effects of regional alveolar hypoxia and hypopcapnia on small pulmonary vessels in cats. J Appl Physiol 61: 440–448.
20. Sheehan DW, Farhi LE (1999) Local pulmonary blood flow: control and gas exchange. Respir Physiol 94: 91–107.
21. Dorrington KL, Talbot NP (2004) Human pulmonary vascular responses to hypoxia and hypercapnia. Physiologist 49: 1–15.
22. Sylvestor JT, Shimoada LA, Aaronson PL, Ward JP (2005) Hypoxic pulmonary vasconstriction. Physiol Rev 92: 367–520.
23. Liu C, Smith TG, Balanos GM, Brookes J, Crosby A, et al. (2007) Lack of involvement of the autonomic nervous system in early ventilatory and pulmonary vascular acclimatization to hypoxia in man. J Physiol 579: 215–225.
24. Smith TG, Balanos GM, Croft QP, Talbot NP, Dorrington KL, et al. (2008) The increase in pulmonary arterial pressure caused by hypoxia depends on iron status. J Physiol 586: 5999–6005.
25. Smith TG, Brookes JT, Balanos GM, Lappin TR, Layton DM, et al. (2006) Mutation of von Hippel-Lindau tumour suppressor and human cardiopulmonary physiology. PLoS Med 3: e290.
26. West JB (1990) Ventilation/Blood Flow and Gas Exchange. Oxford: Blackwell.
27. Robbins PA, Swanson GD, Howson MG (1982) Prediction-correction scheme for forced alveolar gases along certain time courses. J Appl Physiol 52: 1353–1357.
28. Robbins PA, Swanson GD, Michos WP, Schubert WP (1981) A fast computer-controlled binary gas-mixing system for breath-to-breath respiratory control studies. J Appl Physiol 52: 1358–1362.
29. Howson MG, Khamnei S, McIntyre ME, O’Connor DF, Robbins PA (1987) A rapid computer-controlled binary gas-mixing system for fresh breath-to-breath respiratory control studies. J Physiol 394: 7P.
30. Peacock AJ, Challenor V, Sutherland G (1990) Estimation of pulmonary artery pressure using Doppler echocardiography. J Appl Physiol 69: 1347–353.
31. Stevenson JG (1989) Comparison of several noninvasive methods for estimation of pulmonary artery pressure. J Am Soc Echocardiogr 2: 157–171.
32. Severinghaus JW (1979) Simple, accurate equations for human blood O₂ dissociation computations. J Appl Physiol 46: 599–602.
33. Marshall C, Marshall B (1983) Site and sensitivity of stimulation of hypoxic pulmonary vasoconstriction. J Appl Physiol 55: 711–716.
34. Kiss L, Schatte H, Mayer K, Grimm H, Padberg W, et al. (2006) Synthesis of arachidonic acid-derived lipoxigenase and cytochrome P450 products in the intact human lung vasculature. Am J Respir Crit Care Med 161: 1917–1923.
35. Weissmann N, Akkayagil E, Quanz K, Schermuly RT, Ghofrani HA, et al. (2004) Basic features of hypoxic pulmonary vasoconstriction in mice. Respir Physiol Neurobiol 139: 191–202.
36. Barer GR, Howard P, Shaw JW (1970) Stimulus-response curves for the pulmonary vascular bed to hypoxia and hypercapnia. J Physiol 211: 139–155.
37. Kovacs G, Obuchowski A, Bergfeld A, Obuchowski H (2012) Pulmonary vascular resistances during exercise in normal subjects: a systematic review. Eur Respir J 39: 319–328.
38. Groves BM, Reeves JT, Sutton JR, Wagner PD, Cymerman A, et al. (1987) Operation Everest II: Elevated high-altitude pulmonary resistance unresponsive to oxygen. J Appl Physiol 63: 521–530.
39. Balanos GM, Talbot NP, Robbins PA, Dorrington KL (2005) Separating the direct effect of hypoxia from the indirect effect of changes in cardiac output on the maximum pressure difference across the tricuspid valve in healthy humans. Pflügers Arch 450: 372–380.
40. Talbot NP, Balanos GM, Dorrington KL, Robbins PA (2005) Two temporal components within the human pulmonary vascular response to approximately 2 h of isocapnic hypoxia. J Appl Physiol 98: 1125–1139.
41. Morrell NW, Nijman KS, Biggs T, Seed WA (1995) Magnitude and time course of acute hypoxic pulmonary vasoconstriction in man. Respir Physiol 100: 271–281.
42. Viles PH, Shepherd JT (1968) Evidence for a dilator action of carbon dioxide on the pulmonary vessels of the cat. Circ Res 22: 325–332.
43. Ketabchi F, Egemnazarov B, Schermuly RT, Ghofrani HA, Seeger W, et al. (2009) Effects of hypercapnia with and without acidosis on hypoxic pulmonary vasoconstriction. Am J Physiol 297: L977–983.
44. Marshall BE, Hanson CW, Frisch F, Marshall C (1994) Role of hypoxic pulmonary vasoconstriction in pulmonary gas exchange and blood flow distribution. 2. Pathophysiology. Intensive Care Med 20: 379–389.
45. Bironville S, LeJeune P, Naeije R (1996) Effects of hypoxic pulmonary vasoconstriction on pulmonary gas exchange. J Appl Physiol 81: 1535–1543.
46. Bindels L, Jolin-Carlsson A, Santesson J, Gottlieb I (1983) Hypoxic pulmonary vasoconstriction in man: effects of hyperventilation. Acta Anaesthesiol Scand 29: 547–551.
47. Noble WH, Kay JC, Fisher JA (1981) The effect of PCO2 on hypoxic pulmonary vasoconstriction. Can Anaesth Soc J 28: 422–430.
48. Bartsch P, Mairbaur H, Maggiorini M, Swenson ER (2005) Physiological aspects of high-altitude pulmonary edema. J Appl Physiol 98: 1101–1110.
49. Grünig E, Meredès D, Hildebrad W, Swenson ER, Kuhler W, et al. (2000) Stress doppler echocardiography for identification of susceptibility to high altitude pulmonary edema. J Am Coll Cardiol 35: 960–967.