Wiskott-Aldrich syndrome protein forms nuclear condensates and regulates alternative splicing

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The diverse functions of WASP, the deficiency of which causes Wiskott-Aldrich syndrome (WAS), remain poorly defined. We generated three isogenic WAS models using patient induced pluripotent stem cells and genome editing. These models recapitulated WAS phenotypes and revealed that WASP deficiency causes an upregulation of numerous RNA splicing factors and widespread altered splicing. Loss of WASP binding to splicing factor gene promoters frequently leads to aberrant epigenetic activation. WASP interacts with dozens of nuclear speckle constituents and constrains SRSF2 mobility. Using an optogenetic system, we showed that WASP forms phase-separated condensates that encompasses SRSF2, nascent RNA and active Pol II. The role of WASP in gene body condensates is corroborated by ChIPseq and RIPseq. Together our data reveal that WASP is a nexus regulator of RNA splicing that controls the transcription of splicing factors epigenetically and the dynamics of the splicing machinery through liquid-liquid phase separation.
Wiskott–Aldrich syndrome protein (WASP) is the founding member of a family of actin nucleation factors that also include N-WASP (neural WASP), SCAR/WAVE, WHAMM, and WASP12. WASP family proteins respond to various cellular signals to promote actin polymerization via the Arp2/3 complex3. Mutations in WASP cause Wiskott–Aldrich syndrome (WAS), an X-linked primary immunodeficiency that manifests in microthrombocytopenia, eczema, recurrent infections, autoimmunity and predisposition to malignancy7. Some but not all disease phenotypes can be linked to the actin nucleation function of WASP1,4. It has been revealed recently that WASP family proteins function in the nucleus. For example, Drosophila WASH regulates nuclear organization, especially in the heterochromatin compartment5. WASP also functions in transcriptional regulation during differentiation of T helper 1 (TH1) cells8,9, and homology-directed repair5. Despite these advances, the multifaceted functions of WASP remain poorly understood.

Precursor mRNA splicing, a process that removes introns from the primary transcript to generate the mature mRNA, contributes to the expression of almost 95% of human genes9. It is also a complex and fine-tuned process that involves a large number of splicing factors (SFs). Mutations or dysregulated expression of RNA SFs could result in altered RNA splicing that is associated with cancer10. Previous research found increased intron retention and decreased productive splicing of mRNA of IFNG and TBX21 in WASP-deficient T_1 cells, which correlated with R-loop accumulation at these loci11. However, it remains unclear if WASP directly participates in RNA splicing.

Cells perform many biochemical processes including transcription, chromatin organization, RNA splicing, and receptor activation and signaling in diverse membraneless cellular compartments, referred to as condensates, which assemble through liquid–liquid phase separation (LLPS)12. N-WASP (encoded by the WASL gene)—a ubiquitously expressed WASP family protein—forms two-dimensional micron-scale phase-separated signaling clusters on the plasma membrane through multivalent interactions with the adhesion receptor Nephrin and its cytoplasmic adaptor Nck13. The N-WASP-Nephrin-Nck condensates control actin assembly by the Arp2/3 complex14. Using an optogenetic tool15, a recent study showed that N-WASP can form phase-separated liquid droplets with RNA Pol II and nuclear actin16. WASP shares 50% homology with N-WASP. It remains unknown if WASP participates in LLPS and exerts its function in condensates, and, more importantly, what (if any) mechanistic links there are between condensates and the pathogenesis of WAS. In addition, LLPS is the organizing principle of many nuclear compartments17, it could provide a framework to study the nuclear functions of WASP, which remain underexplored.

Research in WAS faces several challenges. Murine models of WAS recapitulate some aspects of WAS, but they lack the presentation of microthrombocytopenia, eczema, and hematological malignancies, yet show consistent development of chronic colitis that is rarely seen in patients18. Primary patient cells for research are scarce, owing to the rarity of WAS. Cell lines can be used to study WAS pathogenesis, but the findings in transformed cell lines must be verified in a normal cellular context. Mutations in the WAS gene affect all hematopoietic lineages except for erythrocytes19. Patient-derived induced pluripotent stem cells (iPSCs) can differentiate into any cell type, therefore constituting an attractive way to model diseases20-22. Two WAS models based on patient-specific iPSCs have been reported23,24. However, these models lacked an isogenic wild-type control, nor did they provide new mechanistic insights into the disease. Similarly, isogenic pluripotent stem cell (PSC) models of inborn blood disorders can be generated de novo from wild-type cells using genomeediting technologies25, which afford greater flexibility in modeling a wide range of mutations and avoid confounding factors associated with patient cells (e.g., mutational burden in Fanconi anemia patient cells25).

Here, we generate two isogenic models of WAS based on either patient-specific iPSCs followed by precise gene correction of disease-causing mutations or on WAS knockout iPSCs generated de novo from a wild-type iPSC line by CRISPR-mediated gene deletion (Fig. 1a). Combined with efficient hematopoietic differentiation these models recapitulate WAS phenotypes and revealed roles of WASP in alternative splicing. Pathogenic mutations or deletion of the WAS gene disrupt this process. Briefly, we find that RNA SFs such as SRSF2 were overexpressed in WASP-deficient cells. Knocking down SRSF2 expression in WASP-deficient cells partially rescues altered RNA splicing. Mechanistically, genome-wide WASP binding is enriched at the promoter of RNA SFs. These SFs show a gain of the H3K27ac modification in WAS mutants, suggesting a mechanism of transcriptional regulation. We prove that WASP can form LLPS condensates in cells using the CRIS-2-based optoDroplet system. A subset of WASP condenses clusters with actively transcribing Pol II, SRSF2, and nascent RNA, suggesting a direct involvement of WASP in RNA splicing. Imaging and biochemical evidence show that WASP physically interacts with SFs and RNA. Fluorescence recovery after photobleaching (FRAP) experiments show that WASP normally acts to constrain the mobility of SRSF2. Taken together, our findings provide mechanistic insights into the role of WASP in regulating RNA splicing.

Results

Generation of isogenic WAS-iPSC models. We derived integration-free iPSCs from fibroblasts of two patients with WAS as previously described (Supplementary Fig. S1a-c). Patient 1 had a clinical score 4 on the Ochs scale28,29, and a nonsense mutation (c.107_108del, p.F36). Patient 2 (clinical score unknown) carried a previously unreported mutation (c.1271dupG, p.L425fs). The patient cells showed an expected reprogramming efficiency (Patient 1, 0.097%, Patient 2, 0.08%, and normal range, 0.023–0.2925,26). The established WAS-iPSC lines expressed markers of pluripotency, differentiated into all three germ layers (Fig. 1b and Supplementary Fig. S1d), retained the original WAS mutation (Supplementary Fig. 1e), and demonstrated a normal karyotype (Fig. 1f and Supplementary Fig. S1g).

To generate an isogenic wild-type control of WAS-iPSC, we performed targeted gene correction by homologous recombina-
tion using a helper-dependent adenoviral vector (HDAdV)—WAS-c-HDAdV30,33 (Fig. 1c). The efficiency of gene correction was 5–50% (Supplementary Fig. S1f). The corrected WAS-iPSCs (named cWAS-iPSCs) had a normal karyotype, retained pluripotency, and were free of random vector integration (Fig. 1f and Supplementary Fig. S1g, h).

The genotype–phenotype relationship in WAS is complex. There are over 200 mutations that lead to hypomorphic levels or complete loss of WASP, and it is impossible to predict clinical severity from the genotype6-34. To avoid phenotype ambiguity due to mutational background and ascertain the true loss-of-function phenotype, we deleted the entire WAS gene in a well-characterized wild-type iPSC line6,33 (using the CRISPR-CAS9 system (Fig. 1d, e). WAS knockout had no appreciable effect on pluripotency and karyotype (Fig. 1f and Supplementary Fig. S1g, i). These WAS wild-type iPSCs (WT-iPSCs) and isogenic knockout iPSCs (KO-iPSCs, three independent clones with two different breakpoints, see Fig. 1e) constitute an independent bona fide null WAS-iPSC model that complements the patient iPSC models.
Isogenic WAS-iPSC models recapitulate disease phenotypes. WAS-iPSCs expressed markers of the three germ layers upon spontaneous differentiation (Supplementary Fig. S2a). The expression of the WAS gene was reduced at the mRNA level in both patients, which was rescued by gene correction (Supplementary Fig. S2b, c). Both WAS-iPSCs and KO-iPSCs could be differentiated into CD34\(^+\) and CD43\(^+\) hematopoietic progenitor cells that form typical hematopoietic colonies (Supplementary Fig. S2d, e). We then differentiated KO-iPSCs into macrophages and WAS-iPSCs into macrophages and dendritic cells (DCs) to examine if they could recapitulate phenotypes observed in patients. Undifferentiated WAS-iPSCs and macrophages from both WAS-iPSCs or KO-iPSCs had no detectable WASP protein expression as compared to the isogenic controls (Fig. 1g). DCs
(WAS-iDCs) derived from WAS-iPSCs, and macrophages derived from WAS-iPSCs and KO-iPSCs (WAS-iMPs and KO-WAS-iMPs) expressed typical lineage markers, consistent with clinical observations (Supplementary Fig. S2f–h)\(^{19,35}\). WAS-iMPs and KO-iMPs had an abnormal elongated morphology that is consistent with known defects in substrate interaction and spreading, as seen in patient macrophages (Fig. 2a)\(^{35,36}\). Podosome defects of WAS\(^{36}\) were highly penetrant in both patient WAS-iMPs and all three clones of KO-iMPs (Fig. 2b). Defective CD43 expression observed in patients\(^{37–40}\) was recapitulated in WAS-iMPs and KO-iMPs (Supplementary Fig. S2i). In addition, chemotaxis of WAS macrophages is known to be impaired\(^{41,42}\), which was recapitulated in KO-iMPs and WAS-iMPs (Fig. 2c and Supplementary Fig. S2j, k). Phagocytosis of *E. coli* particles was significantly impaired but not completely abolished in WAS-iMPs and KO-iMPs (Fig. 2d and Supplementary Fig. S2l), which was documented in patient monocytes and macrophages\(^{43,44}\). Importantly, all these phenotypes were corrected or absent in cWAS-iPSC or WT-iPSC-derived cells (Fig. 2c and Supplementary Fig. S2a–l). WASP and N-WASP share the highest homology among the members in WAS family\(^{45}\). N-WASP expression was similar at both the mRNA and protein levels between the WT iMPs and three KO-iMPs (Supplementary Fig. S2m), suggesting that N-WASP cannot account for the above-mentioned phenotypes. Together, these data from five iPSC lines in three independent genetic backgrounds strongly show that isogenic WAS models faithfully recapitulate known disease phenotypes and are suitable for studying pathogenic mechanisms of WAS.

Splicing factors are overexpressed in WAS. Consistent with previous reports\(^{47}\), WASP localized to both the cytoplasm and the nucleus in major blood lineages except for erythrocytes (Supplementary Fig. S3a–c). Although the roles of WASP in transcription have been described in TH cells\(^{46,47}\), the nuclear localization and function of WASP in monocytes/macrophages have not been reported. Nuclear and cytoplasmic WASP staining in macrophages was verified with four different WASP antibodies (see “Methods”, Supplementary Fig. S3b). WASP puncta occupied interchromatin regions in blood cells (Supplementary Fig. S3b, c). WASP is known to co-localize with hyperphosphorylated RNA polymerase II (Pol II)\(^{47}\). However, whether WASP participates in other nuclear compartments remains unknown. We, therefore, examined the nuclear compartments using TEM analysis using WAS-iMPs (Fig. 3a) and WASP mutant B cells (Supplementary Fig. S4a–c). Mutant cells contained numerous prominent interchromatin granule clusters (IGCs) that correspond to nuclear speckles by negative staining in TEM\(^{46}\), whereas IGCs in gene-corrected or wild-type cells were inconspicuous (Fig. 3a and Supplementary Fig. S4c).

Nuclear speckles are dynamic structures enriched in SFs and involved in pre-mRNA splicing\(^{46,47}\). To confirm the nuclear speckle phenotype, we next performed immunofluorescence staining of nuclear speckle marker SRSF2. Consistent with the TEM data, patient iMPs, KO-iMPs, and WASP mutant B cells contained significantly more prominent and irregularly sized nuclear speckles compared to wild-type or gene-corrected cells (Fig. 3b and Supplementary Fig. S4d–f). The higher expression level of SRSF2 was confirmed by western blot (Supplementary Fig. S4g). Immunofluorescence staining of SF3B1, another nuclear speckle marker\(^{48}\), well recapitulated the nuclear speckle phenotype shown by SRSF2 immunofluorescence ( Supplementary Fig. S4h). Thus, these data suggest that WASP regulates the expression of SRSF2 and the organization of nuclear speckles.

Nuclear speckles play key roles in transcription-linked biological processes\(^{47,49}\). We therefore examined gene expression in WASP mutant cells. We first performed RNA-seq of WAS-iMPs (four biological replicates), cWAS-iMPs (four biological replicates), and macrophages derived from normal cord blood (CB-MPs). Two patient WAS-iMPs differed from CB-MPs in distinctive ways in principal component analysis (PCA), yet both resembled normal macrophages after gene correction (Fig. 3c). There were 652 genes upregulated and 480 downregulated (FDR < 0.05). Consistent with the aforementioned nuclear speckle phenotypes, we found 49 nuclear speckle-associated SFs to be overexpressed in both patient iMPs (Fig. 3d). The enrichment of upregulated SFs in the upregulated genes is statistically highly significant (P < 3.517 × 10\(^{-34}\), hypergeometric test).

WAS patients display dysfunction, clonal proliferation, and malignant transformation of the innate and adaptive immune lineages\(^4\), we, therefore, analyzed primary T and B cells (despite our attempts, none of the samples produced enough macrophages for analysis) in cryopreserved peripheral blood mononuclear cells of WAS patients using RNA-seq (Supplementary Fig. S5a, b). PCA and hierarchical clustering of differentially expressed genes showed that the two patients (P1 and P2) differ from the controls in a distinct fashion (Supplementary Fig. S5b, c). T and B cells of P1 (who had a worse prognosis) showed much greater differences and formed a separate cluster outside of the T- or B-cell clusters (Supplementary Fig. S5c). Similarly, 126 and 92 RNA SFs were upregulated in T cells and B cells of WAS patients, respectively (Supplementary Fig. S5d, e).

The overexpression of SFs in WAS patient macrophages, T cells, and B cells suggested a possible role of WASP in RNA splicing. To test this hypothesis, we next performed ultra-deep RNA-seq in the KO-iMP and WASP mutant B-cell models. The three WASP-KO-iMP samples were transcriptomically distinct from WT iMPs (Fig. 3e). Fifty-eight SFs were upregulated in KO-iMPs, which overlapped the upregulated SFs in patient iMPs by 53.06% (Fig. 3d, f). In contrast, only six SFs were downregulated. The enrichment of upregulated SFs in the upregulated genes is statistically highly significant (P < 3.607 × 10\(^{-22}\), hypergeometric test). A similar observation was made in mutant B-cell lines (Supplementary Fig. S5f). To explore if transcriptional upregulation corresponded to elevated protein levels, we quantitated...
protein expression using the data-independent acquisition mass spectrometry (DIA-MS). As expected, the DIA-MS data showed that WASP was the most significantly downregulated protein in KO-iMPs (Fig. 3g). Consistent with the RNA-seq data, 124 RNA SFs were overexpressed at the protein level in KO-iMPs (Supplementary Data 1), of which 46 overlapped with those detected by RNA-seq in either patient iMPs or KO-iMPs (Fig. 3h). In contrast, only 8 SFs were downregulated in the DIA-MS data. RNA splicing-related processes were the most enriched gene ontology (GO) terms in the upregulated proteins in KO-iMPs (Fig. 3i). Taken together, the results showed that WASP deficiency causes an upregulation of RNA SFs at the mRNA and protein levels.

Altered RNA splicing universally occurs in WASP deficiency cells. To investigate the effect of WASP deficiency on RNA
splicing, we examined alternative splicing (AS) by comparing gene expression at the isoform level (see “Methods”). We identified 303, 310, and 726 genes with differentially expressed isoforms in WAS-iMPs, patient T and B cells, respectively (Supplementary Fig. S6a). Most of these were also differentially expressed at the gene level. About 16% of the isoforms were shared by at least two cell types (Supplementary Fig. S6a), and were highly enriched in GO categories including the nucleoplasm and RNA binding (Supplementary Fig. S6b). We confirmed the AS differences using an alternative bioinformatics pipeline (Supplementary Fig. S6c). Patient cells had a strong preference for exon promotion (higher inclusion) in AS events, and the genes with promoted exons identified in both B and T cells were enriched in protein and nucleic acid turnover (Supplementary Fig. S6d–f).

To better quantify the different types of AS events, we analyzed AS events using the ultra-deep RNA-seq data of KO-iMPs and B-cell lines. The results showed that skipped exon (SE) was the predominant form of AS change in KO-iMPs, followed by retained intron (RI), mutually exclusive exon (MXE), and alternative 3’ or 5’ splice site (A3SS and A5SSS) (Fig. 4a). Similarly, alternative exon usage was the predominant form of AS change (SE and MXE in Fig. 4b, >75% on average) in mutant B cells (Fig. 4b). Many SE events were consistently detected in WAS-iMPs, KO-iMPs and WASP mutant B cells, including those in CTTN, SENP1, and PCBP2 (Fig. 5f and Supplementary Fig. S6g). We validated these shared altered AS events (AASEs) of CTTN, SENP1, TCF12, PCBP2, TPD52L2, and HMG20A using exon-specific qPCR probes that distinguish the isoforms with the included or skipped exons (Fig. 5g and Supplementary Figs. S6h and S9c). We also validated AASEs that were specific to WASP-deficient macrophages in the FN1, DKGZ, and HYAL2 genes, which are associated with immune functions (Supplementary Fig. S9a, b and Supplementary Data 2). Overall, the AASEs were enriched in biological processes such as organelle organization, chromosome organization, and cellular response to stress (Fig. 4c, d). The AASEs were present in both upregulated and downregulated genes (Fig. 4e). Motif analysis (see “Methods”) revealed that the cassette exons or downstream exons of AASEs contained multiple motifs that overlapped with binding sequences of human SFs that were upregulated in mutant cells (e.g., SRSF10, SRSF9, SRSF2, SFPQ, etc.) (Fig. 4f, g and Supplementary Fig. S6i, j). These findings suggest that aberrant overexpression of SFs (Fig. 3 and Supplementary Figs. S4 and S5) could be responsible for the AASEs in WASP-deficient cells.

WASP binds to splicing-factor gene promoters and regulates local chromatin states. To understand if WASP directly participated in transcriptional regulation of SFs, we performed chromatin immunoprecipitation followed by high-throughput sequencing (ChIPseq) of endogenous WASP using a previously validated antibody7 in a wild-type B-cell line. We identified 80,711 high-confidence peaks that were strongly enriched at the transcription start site (TSS) (Fig. 5a). Major categories of WASP binding included proximal promoters (≤1 kb upstream of TSS), distal promoters (between 1 and 10 kb upstream of TSS), distal intergenic DNA, introns, exons, 3’ UTR and 5’ UTR (Fig. 5b). WASP ChIP-seq peaks in the promoter and/or gene body of multiple genes were validated by ChIP-qPCR (Supplementary Fig. S8), and the genome-wide binding pattern of WASP was independently confirmed in WASnull B cells overexpressing a wild-type WASP (Supplementary Fig. S7a, b, d).

WASP ChIP-seq peaks localized near the promoter of the SF genes that are misregulated in WAS (Fig. 5c and Supplementary Fig. S7c, d). The overlap between WASP-binding sites and RNA-splicing genes was statistically significant (Fig. 5d). For instance, 79% (46 out of 58) of the SFs upregulated in KO-iMPs were bound by WASP (Figs. 3f and 5c, d and Supplementary Fig. S7c, d). To understand the effect of WASP binding, we analyzed the active chromatin mark histone H3 lysine 27 acetylation (H3K27ac) in four SFs bound by WASP (Fig. 5c, d) and consistently upregulated in WAS mutant cells (Fig. 3d, f, h and Supplementary Data 1). We showed that the upregulation of SFs was correlated with a gain of H3K27ac, which is a marker of active transcription30, in WASP-bound promoter regions in WASnull cells (Fig. 5e), suggesting that WASP regulates the local chromatin state of SF promoters. To explore the regulation role of WASP, we performed Homer de novo motif analysis using the promoter regions of upregulated SFs bound by WASP. Interestingly, three of the top-ranked motifs were best-matched to binding motifs of Kruppel-like zinc finger transcription factors (TFs) that contain the transcription repression KRAB domain (Supplementary Fig. S7f). The best-matched TFs included ZNF460 and ZNF189, which are associated with lymphoma and immunodeficiency, respectively. These data suggest that WASP might mediate transcriptional repression of SFs through KRAB-zinc finger TFs.

Rescue of WAS-relevant phenotypes by genetic downregulation of splicing factors. Specific AS isoforms have been shown to be enriched in cancer due to their ability to promote proliferation and metastasis31,32. The full-length isoform of cortactin (CTTNwt)33 is overexpressed in B-cell chronic lymphocytic leukemia but not normal B cells and correlates with poor prognosis34. RNA-seq and isoform-specific RT-PCR showed that CTTN-wt was specifically upregulated in WAS-iMPs, KO-iMPs and WASnull B cells, whereas cWAS-iMPs, WT iMPs, CB-iMP and WASP-rescued WASnull B cells predominantly expressed a shorter isoform.

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Fig. 2 Recapitulation of WAS phenotypes using isogenic WAS-iPSC models. a Representative phase-contrast microscopy images of macrophages and quantitative analysis of their circularity. PB-MP: macrophages derived from normal peripheral blood. Analyzed cell n = 36 (PB-MP), 36 (WT iMP r1), 50 (WAS-iMP1), 50 (cWAS-iMP1), 36 (WAS-iMP2), 36 (cWAS-iMP2), 30 (KO-C52 iMP), 47 (KO-C152 iMP), 30 (KO-C157 iMP), and 28 (WT iMP). Data are shown as mean ± SEM, Two-sided Student’s t test. ****P < 0.0001. Scale bar = 50 μm. b Representative immunofluorescence images of podosomes (yellow arrowheads) in macrophages. Podosomes are marked by F-actin and cortactin (encoded by the CTTN gene). DNA was stained with DAPI. DIC differential interference contrast. Scale bar = 10 μm. The experiment was repeated three times independently. c Transwell migration assay. Left: schematic of the principle of the transwell migration assay. Each upper compartment was seeded with the same number of WT iMPs and KO-iMPs (three clones, three replicates). The images show macrophages that migrated to the underside of the porous membrane and were stained by crystal violet. Rightmost: quantitation of cell number in the images. Biological replicate n = 3. Individual dots represent cell number per sample, and the bars show mean ± SD, ****P = 0.0001 (one-way ANOVA). Scale bar = 200 μm. d Quantitative measurement of phagocytosis. Left: schematic of the principle of the phagocytosis assay using the pH-sensitive fluorescence dye pHrodo Red-conjugated E. coli particles. Right: fluorescence images and the quantification of pHrodo Red fluorescence in the same number of WT iMPs and KO-iMPs (three clones, three replicates). Rightmost: quantitation of pHrodo Red fluorescence in each cell. Biological replicate n = 3, cell n = 2271 (WT iMP), 1653 (KO-C52 iMP), 1452 (KO-C152 iMP), 1180 (KO-C157 iMP). Data are shown as mean ± SD, ****P = 0.0001, ***P < 0.0001 (one-way ANOVA). Scale bar = 200 μm.
lacking exon 11 (CTTN-SV1)\textsuperscript{53} (Fig. 5f, g and Supplementary Fig. S9c). Moreover, overexpression of two WASP mutants resulted in a dramatic increase in CTTN-wt, suggesting that mutant WASP is responsible for the AS event (Supplementary Fig. S9c). Intriguingly, CTTN exon 11 contains a high density of the consensus exonic splicing enhancer motifs of SRSF2. The abundance of exon 11-containing transcript (CTTN-wt) correlated with the levels of SRSF2 (Supplementary Figs. S9c and S4g). Knocking down SRSF2 in KO-iMPs and WAS\textsuperscript{null} and WAS\textsuperscript{F36\*B} B cells significantly reduced the ratio of CTTN-wt to CTTN-SV1 (Fig. 5h and Supplementary Fig. S9e), suggesting the overexpression of SRSF2 in WASP mutant cells is responsible for this.
Fig. 3 WASP deficiency results in overexpression of splicing factors. a Representative TEM images of 70-nm sections of WAS-iMPs and cWAS-iMPs collected under identical conditions. Bar = 1 μm in the large panels; Bar = 0.5 μm in the insets. IGC: interchromatin granule cluster (nuclear speckles). Number of cells analyzed: n = 7 (WAS-iMP), n = 4 (cWAS-iMP). b Representative maximum projection confocal images and image-based quantitative analysis of the nuclear speckle marker SRSF2 in WAS-iMPs, cWAS-iMPs, WT iMPs and KO-iMPs (three clones). The mean fluorescence intensity (MFI) of SRSF2 signal per nucleus was measured in the 2D images in cWAS-iMPs (n = 15), WAS-iMPs (n = 22), WT iMPs (n = 49), CS2 KO-iMPs (n = 53), CS2 KO-iMPs (n = 59), and CS17 KO-iMPs (n = 54). For the 3D analysis of the MFI and volume of nuclear speckles, SRSF2 speckles were segmented and quantitated using the Avizo software (see “Methods”). Number of particles analyzed: n = 217 (cWAS iMPs), n = 379 (WAS iMPs), n = 129 (WT iMPs), n = 64 (CS2 KO-iMPs), n = 51 (CS17 KO-iMPs), n = 62 (CS17 KO-iMPs). Data are normalized by the corresponding wild-type and shown as mean ± SEM. Comparisons were performed with two-sided Student’s t test, or Mann-Whitney test if data are not normally distributed. *P = 0.0173, ***P = 0.001, ****P < 0.0001. Bar = 5 μm. Source data are provided as a Source Data file.

c PCA of RNA-seq data of WAS-iMPs, cWAS-iMPs derived from two patients, and wild-type CB-MP. Heatmap of samples shown in C based on upregulated SFs. d PCA of RNA-seq data of WT iMPs and WASP-KO-iMPs. e Heatmap of upregulated SFs in WASP-KO-iMPs shown in (c). f Volcano plot of mass spectrometry of WT and WASP-KO-iMP samples. Parametric test with Benjamini–Hochberg correction. g Heatmap of upregulated SFs in WASP-KO-iMPs shown in (g). h GO analysis of upregulated proteins in WASP-KO-iMPs.

AASE. Similarly, SRSF2 knockdown partially rescued the AASE events in the DGKZ gene (Fig. 5h), which is involved in carcinogenesis and immune response35,36, while knocking down three other upregulated SFs (SRSF1, SRSF3, and HNRNPD) had no effects (Supplementary Fig. S9d). These data suggest that overexpression of SRSF2 may play a larger role than that of other SFs in AASEs.

A study in the WAS knockout mouse model and WAS patient cells suggested that WASP-deficient macrophages upregulate the pro-inflammatory cytokine TNF, which was linked to exacerbated Th1/Th17-helper cell response and autoimmune inflammatory colitis57. This pro-inflammatory phenotype was confirmed in our WAS models at the mRNA and protein levels (Supplementary Fig. S9f). While KO-iMPs secreted significantly more TNF than WT iMPs after treatment with lipopolysaccharide (LPS), this TNF overproduction phenotype was partially rescued by SRSF2 knockdown in all three KO-iMP lines (Fig. 5i).

WASP physically interacts with splicing factors and regulates SRSF2 mobility. Surprisingly, WASP-binding sites were also significantly enriched in AASEs of WASP mutant cells (Fig. 6a), indicating that WASP could directly participate in RNA splicing. We further performed an integrated analysis of WASP ChIP-seq and AASEs and found that, besides the expected enrichment at promoters (Fig. 5a, b), WASP binding was also enriched around the cassette exon and its flanking exons, albeit to lower levels—especially for the downstream exon—than WASP binding at promoters (Fig. 6b). The exonic WASP binding was validated by ChIP-qPCR (Supplementary Fig. S8). These patterns of WASP binding suggest that WASP may work with SFs co-transcriptionally. To test this hypothesis, we first examined if WASP was present in the same complex as SFs. Only a few proteins are known to interact with WASP in the nucleoplasm217. To systematically extend the knowledge on how WASP participates in nuclear functions such as splicing, we performed MudPIT (multidimensional protein identification technology) analysis using co-immunoprecipitation (co-IP) of endogenous WASP, which achieved specific and sensitive detection of WASP interaction partners (see “Methods”). MudPIT analysis in whole cells and purified nuclei (Supplementary Fig. S10a) identified 78 and 299 high-confidence WASP-interacting polypeptides, respectively. These included known partners of WASP, including SMARCA5 (mean spectral count (SC) 6, mean sequence coverage (Seq Cov) 5.3%)6, WIPF1 (SC 17.5, Seq Cov 9.7%)4, and VIM (SC 5, Seq Cov 9.7%)38, thus validating our method (Supplementary Data 3). Analysis of proteomic datasets of nuclear subcompartments47,59,60 showed that the 99 proteins of WASP interactome overlapped with components of nuclear speckle (e.g., SRSF2, SF3B3, and CDC5L, Supplementary Data 3), which are enriched in RNA-splicing-related GO terms (Supplementary Fig. S10b). Similarly, WASP-interacting proteins were highly enriched in RNA-binding proteins (Supplementary Fig. S10c)61.

The association between WASP and nuclear speckle components indicated an unexpected role of WASP in the RNA-splicing process. We thus confirmed the physical interaction between WASP and SFs that are bona fide nuclear speckle components using co-immunoprecipitation assay (Fig. 6c). Proximity ligation assay (PLA, see “Methods”), which is a sensitive and specific method for in situ detection of endogenous protein interactions, further showed that the locations where WASP physically interacts with SFs were consistent with known subcellular distribution of these factors (i.e., SRSF2: mainly nuclear62, SF3B3 (also known as SAP130) and HNRNPA2B1: shuttling between the nucleus and cytoplasm63,64) (Fig. 6d). Furthermore, purified recombinant WASP and SRSF2 proteins showed direct binding in the microscale thermophoresis (MST) assay (Supplementary Fig. S10f, g).

SRSF2 is shown to move dynamically between nuclear speckles and their surroundings65. We hypothesized that this process could be regulated through its interaction with WASP. To test this hypothesis, we expressed a GFP-tagged SRSF2 construct65 that had a similar nuclear distribution as the endogenous SRSF2 (Supplementary Fig. S10h) in WT and KO-iMPs and performed FRAP analysis. The FRAP data showed that the absence of WASP resulted in a faster fluorescence recovery and a higher mobile fraction of SRSF2. The difference between WT and KO-iMPs was statistically significant, although some clonal variation was observed (Fig. 6e, f and Supplementary Fig. S10i, j). Interestingly, the dynamics of SRSF2 within the nuclear speckles (i.e., SRSF2 bright speckles) was not significantly different between WT and KO-iMPs (Supplementary Fig. S10k), suggesting that the increase of SRSF2 dynamics was in the non-speckled regions. Besides, we used a non-speckle nuclear protein HP1α-GFP to evaluate if nuclear mobility was generally higher in WASP-KO cells. The results indicated that HP1α was highly immobile in both WT and KO-iMPs (Supplementary Fig. S10l). We confirmed that these observations were not artifacts as the same HP1α-GFP construct showed FRAP dynamics in HEK293T cell as documented in the literature66 (Supplementary Fig. S10l). These data supported the idea that WASP specifically alters SRSF2 mobility in the nucleoplasm. Endogenous WASP puncta were often observed near SRSF2 bright nuclear speckles (Supplementary Fig. S11a). Consistently, most WASP-SRSF2 interactions were detected near or outside of nuclear speckles (Supplementary Fig. S11b). Therefore, WASP physically interacts with the components of nuclear speckles including SRSF2 and constrains the mobility of SRSF2.

Optogenetic WASP clusters with SRSF2, active Pol II, and newly transcribed RNA. Nuclear speckles are membraneless
Fig. 4 WASP deficiency results in altered RNA splicing in iMPs and B-cell lines. 

**a** Altered alternative splicing events (AASEs) in KO-iMPs (three clones). A3SS alternative 3' splicing site, A5SS alternative 5' splicing site, MXE multi-exon skipping, RI intron retention, SE exon skipping. WT iMP biological replicate \( n = 2 \), KO-iMP biological replicate \( n = 3 \). The WASP-null B-cell lines are derived from the same biological replicate as the WASP KO-iMPs. 

**b** AASEs in 3 WASP mutant B-cell lines. Biological replicate \( n = 2 \). 

**c** GO enrichment of AASEs in KO-iMPs. WT-iMP biological replicate \( n = 2 \), KO-iMP biological replicate \( n = 3 \). 

**d** GO enrichment of common AASEs in three WASP mutant B-cell lines. Biological replicate \( n = 2 \). Hypergeometric testing \( P \) values are corrected for multiple testing using a false discovery rate (FDR). 

**e** AASEs in both upregulated genes and downregulated genes in KO-iMPs. 

**f** Splicing-factor motif analysis of altered SE, RI, and MXE events in KO-iMPs. 

**g** Splicing-factor motif analysis of the downstream exon of altered SE in WASP-null B cells.
nuclear bodies with phase-separated, liquid-like properties that allow their constituents to shuttle between the nuclear body and nucleoplasm dynamically and transiently interact with nucleoplasmic factors. These phase-separated condensates are driven by intrinsically disordered proteins (IDPs) that contain intrinsically disordered regions (IDRs). Intriguingly, WSP partners showed a strong enrichment for bioinformatically predicted IDPs (Fig. 6g, Supplementary Fig. S10d, e, and Supplementary Data 3), which implies a potential role of WSP in condensates.
Fig. 5 WASP binds to promoters of splicing-factor genes to regulate their transcription. a Heatmap of endogenous WASP ChIP-seq peaks around the TSS. b Genomic annotation of WASP ChIP-seq peaks. c WASP ChIP-seq peaks located in SF genes visualized in Integrative Genomics Viewer (IGV). d Venn diagram shows overlap between WASP-binding genes, RNA-splicing genes (GO term 0008380), and upregulated SFs of KO-iMPS. The hypergeometric testing P value is 1.27 × 10⁻⁵⁶ for RNA-splicing term vs WASP ChIP-seq and 5.21 × 10⁻¹² for upregulated SF vs WASP ChIP-seq. e Representative ChIP-qPCR results of the level of H3K27ac in WASP binding sites in selected SF genes. Biological replicates n = 2 for gene SRSF7 and HNRNPF, and for gene SRSF2 and HNRNP2B1 n = 3. One of the independent biological replicates is shown, and other biological replicate data are provided in a Source Data file. Data are shown as mean ± SEM. f Isoform-level analysis of RNA-seq data of macrophages and B-cell lines. The graph below the gene diagram shows IGV tracks of RNA-seq reads mapped to the isoforms. g Isoform-specific qRT-PCR analysis of two CTTN variants in macrophages. Bottom: Semi-quantitative RT-PCR shows expression of two CTTN isoforms in WAS-iMPS. Rep1 and Rep2: two biological replicates. The numbers below the gel pictures are densitometry ratio of CTTN-wt and CTTN-SVL. ***P = 0.0196 (KO-C52 IMP), **P = 0.0131 (KO-C157 IMP), ***P = 0.0002 (KO-C152 IMP), ***P = 0.0002 (WAS-iMP), ***P = 0.0003 (WAS-iMP2). h qRT-PCR analysis of the expression of the CTTN and DGKZ isoforms upon knockdown of SRSF2 in KO-iMPS. Biological replicate n = 3. CTTN: **P = 0.0181 (KO-C152 IMP), **P = 0.0487 (KO-C157 IMP), ***P = 0.0001; DGKZ, VI: **P = 0.0256, ***P = 0.0017, ***P = 0.0007. i ELISA quantitation of TNF secretion by indicated macrophages upon stimulation with LPS. Biological replicate n = 3. mKO-C52 IMP), **P = 0.0116 (KO-C52 IMP), ***P = 0.0026 (KO-C157 IMP), ***P = 0.0006. All statistics in this figure were done using a two-sided Student’s t-test. Data in (g-i) are shown as mean ± SD.

morphology in wild-type cells (Fig. 3b and Supplementary Fig. S4d, e), we hypothesized that WASP could control the localization of SFs (e.g., SRSF2, SF3B1) through LLPS. Indeed, WASP was predicted to contain multiple IDRs (http://www.pondr.com, Fig. 7a). To test if WASP could phase separate in vivo, we cloned the WT WASP and the predicted N- and C-terminal IDRs of WASP to the N-terminus of the Cry2-based optogenetic droplet construct15. Optogenetic WT WASP (opto-WASP) readily formed LLPS droplets in cells, but neither IDR was sufficient for droplet formation (Fig. 7a and Supplementary Movies 1–3). Furthermore, we observed the growth and fusion of optoWASP droplets over time by live-cell imaging and the dynamic exchange of WASP molecules between the soluble and droplet phases by FRAP (Fig. 7a–c and Supplementary Movies 1 and 4), proving the liquid-like properties of optoWASP droplets.

WASP is a nucleation factor of actin polymerization. Using a GFP-tagged nuclear actin chromobody, we showed that nuclear filamentous actin (F-actin) colocalized with optoWASP droplets (Supplementary Fig. S10m), suggesting that optoWASP retained the extra actin-binding domain) remains to be elucidated54. It has been demonstrated that endogenous WASP participates in phase-separated condensates encompassing SFs and active Pol II. Consistently, we found that optoWASP droplets could also cluster nascent RNA (produced by serum-stimulated transcription) together with SRSF2 (Fig. 7d, e), which further supported the role of WASP in RNA splicing. All observations made with N-terminal WASP-Cry2 fusion construct were reproduced with a C-terminal Cry2-WASP fusion construct (Fig. 7d, e and Supplementary Fig. S11c, d).

To confirm the interaction of WASP and RNA, we performed RNA immunoprecipitation followed by high-throughput sequencing (RIPseq). WASP antibody pulled down specific RNA species in WT but not in WASPnull B cells in either the whole cell or nuclear fraction (Supplementary Data 4). We further validated the enrichment of select RNA in WASP immunoprecipitation in wild-type cells using RIP-qPCR (Supplementary Fig. S11f). Taken together, our data reveal that WASP is a phase-separated protein and it could regulate RNA processing via interaction with SRSF2, active Pol II, and the nascent RNA in gene condensates.

Discussion

WASP is a multifaceted protein important for diverse hematopoietic and immune processes1. The pathogenic mechanisms of WAS remain poorly defined, partly due to the lack of an experimental system of suitable for dissecting the function of WAS in appropriate cellular context in humans. Here, we take advantage of isogenic iPSC models to investigate the role of WASP in RNA splicing. This study revealed that WASP deficiency caused by diverse WAS genotypes results in overexpression of SFs and leads to altered splicing events. We found a significant enrichment of WAS-associated AASEs in the immunity-associated gene list in the Gene Set Enrichment Analysis (GSEA) database (P < 0.002, Supplementary Data 2). Some of these AASEs affect genes that are not only associated with immunity but also play essential roles in inflammation (e.g., FN1 and HYAL2) and cancer oncogenesis (e., CTTN, FN1, and DGKZ). The WAS-associated CTTN-wt isoform (Fig. 5g and Supplementary Fig. S9c) encodes an extra actin-binding domain compared to the CTTN-SVL isoform. CTTN-wt is found to be overexpressed in B-cell chronic lymphocytic leukemia, but the functional relevance of the exon (i.e., the extra actin-binding domain) remains to be elucidated54. It has
been reported that HYAL2 has two variants that contain either exon 1 or exon 2 at the 5′UTR. The splicing variant exons 1–3 is absent in cancer cell lines, but the variant exons 2–3 show a higher expression in cancer cell line73. In our model, exon 2 of HYAL2 exhibited a higher inclusion ratio in WASP-KO-iMPs, which might be related to the high cancer susceptibility in WAS patients. In addition, exon 2 of DGKZ is more frequently skipped in WASP-KO-iMPs, leading to a truncated protein with two different alternative N-terminal domains (DGKZ_V1 and DGKZ_V2) with the same internal deletion (Met1-Ala243del). In summary, these new analyses show that WAS AASEs could lead to altered protein sequences and may be involved in disease-relevant processes.

We uncovered two mechanisms by which WASP regulates RNA splicing. First, WASP represses the transcription of SF genes under normal circumstances by directly binding to their...
promoters and regulating the local chromatin state. Second, we demonstrate that WASP is a phase-separated protein that can directly participate in the process of RNA splicing during transcription through physical association with target genes and components of the gene-body condensates (e.g., active Pol II, SFs, and nascent RNA) (Supplementary Fig. S12).

The evidence of WASP binding to splicing-factor promoters provides a hint that WASP regulates the transcription of SF genes. WASP could regulate chromatin remodeling and/or histone modifications of these promoters, which is supported by our observation of the gain of H3K27ac in mutant cells. Increasing evidence has shown that nuclear actin or nuclear actin polymerization regulates gene transcript by RNA Pol II16,74–76. It will be interesting to test if the transcriptional regulation of SFs by WASP depends on its actin-polymerization function. The nuclear actin phase separates with Pol II during serum-induced transcription, as process that requires N-WASP16. Another possibility is that WASP plays a role in regulating transcription initiation. Besides SFs, the identified WASP-binding partners include transcription factors and epigenetic regulators (Supplementary Data 3), which could be recruited by WASP (whose binding is enriched at TSS) to the gene promoter condensate that contains the pre-initiation complex, co-activators, and nuclear actin17,77 to regulate transcription. As WASP also occupies the gene body (i.e., exons and introns), an intriguing possibility that WASP may facilitate interactions between the promoter and distal sequences also warrants future study.

Recently, a WASP family protein, N-WASP (or WASL), was shown to drive LLPS via multivalent interactions with signaling complexes on the cell membrane14. Bioinformatics tools such as PScore69 and PSPredictor8 predict that WASP has phase-separation propensity, but no experimental evidence existed prior to this study. We proved WASP is a phase-separated protein in the nucleus by using an optogenetic tool and by disrupting endogenous WASP condensates with 1,6-HD. WASP in the liquid-droplet phase carries out normal function in actin nucleation (Fig. 7a and Supplementary Fig. S10m). However, the N- and C-terminal IDRs of WASP are not sufficient for liquid–liquid phase separation (Fig. 7a), suggesting that multiple IDRs are involved in LLPS of WASP. Our data revealed extensive protein–protein interactions that WASP mediates in the nuclear compartment, including interactions with numerous SFs. We showed that WASP in normal situations constrains the mobility of SRSF2. Interestingly, the nuclear speckles are enlarged and irregularly sized in WASP-deficient cells (Fig. 3b). The nuclear speckle morphology can be disturbed by carcinogenesis, viral infection, and phosphorylation of serine/arginine-rich SFs and assumes an enlarged and irregular appearance.65,47,79. This is similar to the phenotype of WASP-deficient cells (Fig. 3a, b and Supplementary Fig. S4c–e, h). In addition, we showed WASP droplets encompass SRSF2, active Pol II, and the nascent RNA. It is thus conceivable that WASP may facilitate the function of the splicing machinery in the splicing condensate described recently17,77 through LLPS in the nucleus. Future work is needed to decipher the relative contribution of the two mechanisms and how they coordinate splicing regulation.

The cellular models here presented can be used for elucidating how the mechanisms and protein factors identified herein contribute to pathologies of WAS. Finally, the significant alterations in gene expression and AS described in this study may serve as diagnostic biomarkers for WAS patients and constitute the basis upon which to develop targeted therapies for WAS.

**Methods**

**cells used and culture conditions.** The use of human samples that were obtained with informed consent was approved by the Institutional Review Board (IRB) of the Salk Institute for Biological Studies and KAUST Institutional Biosafety and Bioethics Committee (IBEC). Human fibroblasts from a male Wiskott–Aldrich syndrome patient with the c.1271dupG mutation (in the WAS gene) were purchased from the Coriell Institute (Camden, NJ). Human fibroblasts from a male patient with the c.1078G > A mutation and from the mother of the patient (unaffected) were purchased from the Cell Line and DNA Bank from Patients Affected by Genetic Diseases (Genova, Italy). All human fibroblasts were cultured at 37 °C in DMEM containing GlutaMAX, nonessential amino acids, sodium pyruvate, and 15% fetal bovine serum (FBS). The B-lymphocyte cell lines ID00003 (WASm4, c.430G > A, E133K, clinical score 4–5) and GM11518 (wild type) were purchased from the Coriell Institute (Camden, NJ). All lymphoblastoid cell lines were cultured at 37 °C in RPMI-1640 containing 2mL-glutamine and 15% FBS. H1 and H9 human ESs were purchased from WiCell Research. Human ESs and generated iPSC lines were cultured on Matrigel and/or mitotically inactivated MEF cells at 37 °C in DMEM/F12 (Invitrogen) supplemented with 0.1 mM nonessential amino acids (Invitrogen), 1 mM GlutaMAX (Invitrogen), 20% Knockout Serum Replacement (Invitrogen), 55 mM beta-mercaptoethanol (Invitrogen) and 10 ng/mL bFGF (Joint Protein Central) as described. Peripheral blood mononuclear cells from two WAS patients were obtained with informed consent under approval by IRB at ICRCS Fondazione Policlinico San Matteo. One patient (P1, clinical score 4) manifested thrombocytopenia, eczema that was difficult to control, infections, humoral immune deficiency, but no autoimmunity or cancer. P1 was diagnosed with WAS over 25 years ago, and no genetic test was done. The other patient (P2, clinical score 2) manifested thrombocytopenia and humoral immune deficiency, but no autoimmunity or cancer. P2 carries the mutation p.P413fs, which impairs the c-terminal VCA domain of WASP. All cells of P1 and P2 were used to produce the data presented in the manuscript and therefore are no longer available. Human buffy coats were purchased from San Diego Blood Bank (San Diego, CA). Human umbilical cord blood were gifts from StemCyte (Covina, CA) and San Diego Blood Bank (San Diego, CA). The cord blood units were anonymized and donated to research by individuals who gave informed consent. Cord blood CD34+ cells were isolated by immune selection using human CD34 magnetic microbeads and MiniMACS separator (Miltenyi Biotec, San Diego, CA) according to the manufacturer’s instructions. The purity of the cells was checked by staining with anti-CD34-PE antibody on a BD LSRFortessa cytometer. Cryopreserved CB CD34+ cells were FACS sorted again before being used in experiments. The HEK293T cell

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**Fig. 6** WASP directly binds alternatively spliced genes and physically interacts with splicing factors. **a** Venn diagram of WASP-bound genes and AASEs in WASP mutant cells. Hypergeometric testing P values are 1.39 × 10−41 (KO-iMPs), <10−324 (Null), <10−324 (Null + F36), and <10−324 (Null + L425S) cells (shown in (a)). The vertical lines in the model gene diagram indicate the center of the elements. The distance from the center in kb is given by the scale bar. The y axes are drawn to the same scale. **b** co-IP validation of physical interaction between WASP and selected proteins. IP immunoprecipitation, IB immunoblotting. This experiment was done once for SF3B3, LMNB, and RNAPII, and twice for SRSF2. **d** Representative immunofluorescence images of PLA signals for WASP-SRSF2, WASP-SF3B3, and WASP-HNRNPA2B1 in the WT B-cell line. The pie chart below each image shows the percentage of PLA signals in different cellular compartments. Right: quantitative analysis of PLA signals. Cell number n = 50 for each setting. Data are shown as mean ± SEM. Mann–Whitney test, ***P < 0.0001. Nuclei were stained with DAPI. Bar = 5 μm. NE nuclear envelope, NO nucleolus, OIN other locations inside the nucleus. **e** Left: representative FRAP images of SRSF2-GFP expressing WT and WASP-KO-iMPs (KO-C152 is shown here, and other clones are shown in Supplementary Fig. S10d). Scale bar = 2 μm. Right: time-lapse FRAP curve of SRSF2-GFP fluorescence generated from representative iMP cells. **f** Statistic analysis for the half time of FRAP recovery and mobile fraction of SRSF2-GFP fluorescence in WT (n = 39) and WASP-KO-iMPs (n = 78). Data are shown as mean ± SD. Two-sided Student’s t test. **P = 0.004, ****P < 0.0001. Source data are provided as a Source Data file. **g** Percentage of predicted phase-separation proteins (PScore ≥ 4) in the human proteome (n = 18,476) and WASP partners among nuclear speckle proteins (n = 95). P value is calculated with Fisher’s exact test. Source data are provided as a Source Data file.
A iPSC line was purchased from ATCC and cultured in DMEM medium containing 10% FBS (heat-inactivated).

iPSCs generation. The episomal vector-based reprogramming was performed as in refs. 25, 26. Briefly, fibroblasts were nucleofected with the episomal vectors using a Nucleofector II (Lonza) with program U20. Five days after nucleofection, fibroblasts were reseeded onto mitotically inactivated mouse embryonic fibroblasts (MEFs). The reprogrammed colonies were allowed to grow until they were ready to be mechanically picked and transferred onto MEFs. Upon successful expansion of the iPSC lines on MEFs, they were then cultured on Matrigel (BD Biosciences) in mTeSR medium (StemCell Technologies). Analyses of pluripotency of iPSCs, including detection of transgene insertion, transgene expression, teratoma formation, immunostaining of pluripotency markers, and karyotyping were carried out as in refs. 25, 26. The genotype of the WAS-iPSCs was periodically verified during the course of this study. The identity of the iPSCs was checked by Sanger sequencing and DNA fingerprinting.

Construction and preparation of HDAdV. Construction and preparation of WAS-c-HDAdV for gene correction were generated using a BAC clone containing the human WAS locus (RP11-1148L6, BACPAC Resources)81. In brief, an FRT-
PGK-EM7-neo-bPA-FRT fragment was recombined into intron 2 of WAS in the BAC clone. A total of 20.4 kb of WAS homology sequence and FRT-PGK-EM7-neo-bPA-FRT cassette was subcloned into the HDADV plasmid p5C3RDhADCT8-4 (kindly provided by Dr. Kohnosuke Mitani) (Suzuki et al. [22]). The WAS-c-HDADV virus was generated according to a published method[22,23]. Briefly, the WAS-c-HDADV plasmid was linearized by Pci-Scl (New England Biolabs) and transfected into 116 cells (a kind gift from Dr. Philip Ng) in the presence of helper virus AdΔHPRG35 (a kind gift from Dr. Anders M. Åkerberg). Cells were serially amplified in 116 cells and the purified, g·al·transducing units (btu) were determined in 293 cells to define infectious vector titers.

Isolation of gene-corrected human iPSCs. For the generation of gene-corrected iPSCs, 2.5–3.0 × 10^6 WAS patient iPSCs were infected with WAS-c-HDADV at the multiplicity of infection (MOI) of 3–30 btu/cell. Two days after infection, G418 (50 μg/ml; Invitrogen) was added to the medium to start positive selection. After 9 days, 2 μg Ganciclovir (Ganciclovir, Invitrogen) in addition to G418 was added to the medium to start negative selection. After an additional 8 days, G418/GANC double-resistant clones were transferred to 96-well plates and expanded for further characterization. Gene-targeted clones were determined by PCR of genomic DNA from drug-resistant clones with the following primers (P1, 5‘-CATTCCGCCCATACCCATACCTCGCTC-3’; P2, 5‘-TCCATTGCTCA-3’; P3, 5‘-CTACCTGGCCATTGACGCAACAGGCAAACATC-3’; P4, 5‘-AAATTTCGCGGATCTCTTCCGGCAGGTACG-3’; see Fig. 1e) using PrimeSTAR GXL DNA Polymerase (TAKARA). To determine gene-corrected clones for exon 10, exon 10 of WAS was PCR-amplified with the following primers: 5‘-AAGAAACCCAGGCCCATGACTACTCCTTGCCACA-3’ and 5‘-AGGCAACATTCCGGGTGAGAAGAATACCCA-3’; 5‘-GGGATTTCACCCCCTAAGGGC-AGG-3’; 5‘-ACTGCTTCAGTCAGGAGTTG-3’; 5‘-GGAGGCTGACACAAGATTCAT-3’; and 5‘-ACTGCTTCAGTCAGGAGTTG-3’ with PrimeSTAR GXL DNA Polymerase. To determine gene-corrected clones for exon 10, exon 10 of WAS was PCR-amplified with the following primers: 5‘-ACTGCTTCAGTCAGGAGTTG-3’ and 5‘-GGGATTTCACCCCCTAAGGGC-AGG-3’ (downstream)—were synthesized by in vitro transcription using the T7 MEGAshortscript kit (Thermo Scientific) and subjected to Fluorescence in situ hybridization. The WAS-c-HDADV probe was custom synthesized by in vitro transcription using the T7 MEGAshortscript kit (Thermo Fisher Scientiﬁc) and labeled with 5-TAMRA by Empire Genomics (Buffalo, NY). 5-fluorescein-labeled complementary probe was synthesized from Empire Genomics. Cells were fixed in 4% formaldehyde for 10 min at room temperature, permeabilized with Triton X-100 and repeated freezing in liquid nitrogen, and deproteinized by treating with 0.1 M HCl. Hybridization was carried out at 37 °C overnight. Samples were counterstained with DAPI, mounted, and examined using a Zeiss LSM 780 Laser Scanning Confocal Microscope.

Hematopoietic differentiation. Hematopoietic differentiation efficiency and hematopoietic colony formation activity were assayed as described in ref. 83 with some modifications. Briefly, iPSCs and ESCs cultured on MEF feeders in human ESC medium were manually dissociated and plated on OP9 feeders to start differentiation. Cells were co-cultured with OP9 OP9 feeders for 2 weeks in differentiation medium (Alpha MEM containing 10% Hyclone defined FBS (GE Life Sciences catalog number SH30070.03), 100 μM monothioglycerol (Cat# M61145, Sigma-Aldrich) and 50 μg/ml ascorbic acid (Cat# A4544, Sigma-Aldrich). Differentiated cells were dissociated with collagenase and Accutase (Innovative Cell Technologies) and subjected to flow cytometry analyses, cell sorting, qPCR analyses and clonogenic progenitor cell assays. Macrophage and dendritic cell differentiation were performed as described[84]. Briefly, 2-week differentiated hiPSC were harvested and the non-adherent fraction (or FACS sorted CD34+CD43+) were cultured under myeloid progenitor expansion condition (IMDM 10% FBS, GM-CSF (100 ng/ml), Neomycin (100 μg/ml), and G418 (50 mg/ml) for 1 week. DC differentiation was initiated using DC differentiation medium (RPMI-1640 plus 10% FBS, GM-CSF (100 ng/ml), IL4 (10 ng/ml, Peprotech)). DCs were cultured in RPMI-1640 supplemented with 10% FBS, 10 ng/ml TNFa, 10 ng/ml IL4, and 2 μg/ml LPS (Sigma-Aldrich, Cat# L0242-SMG).

Characterization of iPS-derived cells. Characterization of the identity of iMPS and iDGs using lineage-specific cell surface markers were performed as described in a published protocol[85]. Briefly, 100 K cells were washed in FACS buffer (1x PBS without Ca2+ or Mg2+, 2 mM EDTA, 2% FBS) and pelleted at 400 g for 5 min at 4 °C. The pellet was resuspended in 100 μl FACS buffer in a 5-ml polystyrene test tube. Antibodies or isotype controls were added to the cell suspension and incubated for 30 min at 4 °C with agitation. Four milliliters of FACS buffer were added to wash the cells. The washed pellets were centrifuged at 400 g for 5 min at 4 °C. The pellet was resuspended in FACS buffer without FBS and analyzed on a flow cytometer. Macrophage morphology was quantified by using the circularity plugin of ImageJ. Macrophage phagocytosis assay was performed with pHrodo® Red E. coli BioParticles® Conjugate for Phagocytosis (Cat# P35361, Life Technologies, Carlsbad, CA). The pHrodo red particles were prepared according to the manual and opsonized Escherichia coli BioParticles® opsonizing Reagent (Life Technologies, Carlsbad, CA) as instructed. Adherent macrophages were incubated with pHrodo red E. coli particles for 1 h at 37 °C, dissociated, washed, and analyzed by FACS. Migration assay was performed using the CytoSelect 24-Well Cell Migration Assay, 5 μm kit (Cat# CBA-102, Cell Biologics, Inc., San Diego, CA), or using 24-well Transwell plates with 5-μm pore inserts (Corning, Inc., Main, USA) as follows: 5000 iMPS were resuspended in 100 μl X-Vivo-10 serum-free medium (Lonza, Cat#EBD4-380Q) with 30 μM R848 (Stem Cell Technologies) and seeded in the lower compartment; IMDM medium (Thermo Fisher Scientific) with 10% FBS and 5 ng/ml M-CSF was added in the lower compartment. The cells of the insert were stained by crystal violet (BD) after 12 h of incubation at 37 °C, 5% CO2, and the cells on the upper surface of the insert were removed by cotton swabs; the number of migratory cells was measured using the imageJ software (ver. 1.5.1i).
activated by a laser with a wavelength of 488 nm and imaged by a laser with a wavelength of 580 nm.

**Serum starvation and stimulation.** Before serum starvation, the HEK293 cells were transfected with the Cry2 fusion constructs using Lipofectamine 3000 and cultured in DMEM medium with 10% FBS for 24 h. Subsequently, cells were harvested and seeded with 2 million cells in eight-chamber slides in DMEM medium without FBS overnight. The medium was replaced by 20% FBS DMEM medium for one hour (for nascent RNA labeling, the 20% FBS medium was mixed with 1 mM 5-ethynyl uridine (Thermo Fisher, cat # C10329)). The cells were exposed under blue light for 10 min to induce optogenetic droplet formation and immediately fixed by 4% formaldehyde in PBS for 20 min at room temperature to be ready for the further staining experiment. We used a Click-it RNA imaging kit (Thermo Fisher, cat # C10329) to stain nascent RNA followed a standard protocol provided by the kit.

**Fluorescence recovery after photobleaching assay.** FRAP assay of optoWASP was performed on a Leica SP8 confocal equipped with a live-imaging chambered stage. The optoWASP positive cells were activated by 488 nm laser and monitored by 588 nm laser in a live mode every 5 s until the droplets reached proper sizes. The photobleaching applied 588 nm laser with 100% power for 20 iterations, for a circular region with ~1.0 µm in diameter containing a droplet inside. Images were acquired for five prebleach frames with 2 s interval time and 30 postbleach frames with 10-s interval time. For the FRAP assay of SRSF2-GFP and HP1α-GFP, 0.5 µg plasmid was delivered to 1 x 10^6 WT iPSCs or WASP KO-iPSCs using the Neon transfection system (Thermo Scientific). The electroporation parameters are as follows: pulse voltage 1900 V; pulse width 30 µs; pulse number 1. Cells were cultured for 24 h before imaging. The same amount of HP1α-GFP was transferred into HEK293T cells using Lipofectamine 3000. FRAP was performed on ZEISS LSM 710 inverted confocal microscope. Pixel dwell is 0.42 microseconds for SRSF2-GFP and 0.67 microseconds for HP1α-GFP. Bleaching was carried out for 50 iterations with 100% laser power using the 100x dry objective in the 100 μm field of view. The same amount of HP1α-GFP was delivered into the cells every 1 s for HP1α-GFP. After image acquisition, the data were analyzed using FRAP module installed in ZEISS software. The t-half and mobile fraction were used for statistical analysis and the time-lapse data were used for generating the FRAP curve.

**Immunofluorescence analysis.** Cells were fixed with 4% formaldehyde in PBS for 20 min at room temperature, and then permeabilized with 0.4% Triton X-100. After a blocking step with 10% normal donkey serum in PBS, cells were incubated with the primary antibody at 4 °C overnight, followed by incubation at room temperature with the corresponding secondary antibody for 1 h. Nuclei were counterstained with 4,6-diamidino-2-phenylindole (DAPI) and mounted with ProLong Gold Antifade Mountant (Thermo Fisher, cat # P36930).

**Transmission electron microscopy.** Cells grown on coverslips were fixed for 5 min at 37 °C and then transferred to 4 °C and brought to the Salk Biophotonics core facility for further processing. Media were removed and washed with warmed buffer before the fixative solution (2.5% glutaraldehyde + 2% paraformaldehyde + 2 mM CaCl₂ in 0.15 M cacodylate buffer). Cells were washed and fixed again (1% OsO₄, 0.1% potassium ferricyanide in 0.15 M cacodylate buffer) for 1 h at 4 °C before dehydration. After drying, cells were coated with 2% uranyl acetate for one hour on ice in the dark. The cells were then washed in distilled water and acetone or ethanol series. Embedding was done with Spurr’s embedding kit. After cleaning with methanol, the slides were coated with Teflon and let dry. The embedded samples were flattened using two glass slides in resin. The samples were imaged using a Carl Zeiss Libra 120 kV PLUS Energy Filtered Transmission Electron Microscope that is equipped with a 2 k x 2 k fiber optically coupled YAG CCD camera.

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**Immunoprecipitation and immunoblot analysis.** The WASP-GFP cells and WT B cells described above or WASP-GFP cells expressing FLAG-tagged wild-type WASP were lysed with a buffer comprising 50 mM Tris-HCl (pH 7.5), 150 mM NaCl, 0.5% NP-40, 100 µM aprotinin, 100 µM leupeptin (Roche), 400 µM Na3VO4, 400 µM EDTA, 10 mM NaF, and 10 mM sodium pyrophosphate. The lysates were centrifuged at 20,000xg for 10 min at 4 °C to remove debris, and the resulting supernatants were then incubated for 30 min at 4 °C with protein A-Sepharose 4 Fast Flow (Cat# P9424, Sigma) conjugated with antibodies as indicated. The resulting immunoprecipitates were washed three times with cold 1/10 binding buffer, fractionated by SDS-PAGE, and subjected to immunoblot analysis with primary antibodies and HRP-conjugated secondary antibodies to either mouse (Cat# 7076, Cell Signaling Technology), or rabbit (7074, Cell Signaling Technology) IgGs. Immune complexes were detected with an ECL system (GE Healthcare Cat# RPN2232).

**Proximity ligation assay and colocalization analysis.** Proximity ligation assay (PLA) is a method that allows in situ detection of protein interactions with high sensitivity and specificity. Briefly, two primary antibodies were used to label the target proteins (e.g., WASP and SRSF2). PLA probes, which are species-specific secondary antibodies conjugated with a short oligonucleotide sequence, bind to the primary antibodies. When the target proteins are in proximity (<30-40 nm), the oligonucleotides of the two different PLA probes can participate in rolling amplification. The amplified DNA can then be visualized by a fluorescent oligonucleotide probe, generating strong fluorescence signals (e.g., red spots in confocal images). These red PLA spots indicated the locations where WASP interacts with its partner proteins. PLA was carried out according to the protocol described in the online protocol for the Duolink® Kit. In situ Red Starter Kit Mouse/Rabbit (DUO92101-KT, Sigma-Aldrich). In all, 0.2 million WT B cells were spun onto slides (pre-coated with 1% FBS) using a Pro-Cyt Centrifuge. Then, WT B cells were fixed with 4% formaldehyde in PBS for 20 min at room temperature and further permeabilized with 0.4% Triton X-100 for 30 min. After blocking with the blocking buffer in the kit (DUO92004-30XN, Sigma-Aldrich), slides were treated with two different primary antibodies at 4 °C overnight. The primary antibodies, mouse monoclonal anti-WASP antibody (D-1) (Cat# SC-5300, Santa Cruz Biotechnology) and rabbit polyclonal antibody (Cat# ab37415, Abcam), were used as a negative control. The next day, Duolink® In Situ PLA® Probe Anti-Mouse MINUS and Duolink® In Situ PLA® Probe Anti-Rabbit PLUS was mixed together and applied to the samples. Each slide was incubated at 37 °C for 60 min. After washing, the ligation and amplification were performed following the Duolink® In Situ Detection Reagents Red kit instructions. If needed, an Alexa fluor 488 secondary antibody was applied to the sample to visualize the green immunofluorescence signal of target proteins. Finally, using Duolink® In Situ Mounting Medium with DAPI, slides were incubated to stain nuclei and to protect PLA signals. PLA signals were imaged using a Zeiss LSM 880 with Airyscan function. To quantify the PLA signal number, we randomly picked 50 cells from five different images using a x63 oil objective. PLA signals were checked carefully in the Z-stack mode in the ZEN software. For colocalization analysis, the color threshold function in the Fiji software was used. We define a yellow color (20-50) as colocalization, which is generated from the PLA signal in the red channel and SRSF2 or POI II signal in the green channel.
5 µg of antibody at 4 °C on a rotating platform. The chromatin was incubated with the antibody-coupled beads overnight at 4 °C on a rotating platform. On a magnetic stir plate, the beads were separated by centrifugation. To reverse cross-linking the immunoprecipitated chromatin and input control were incubated on a thermomixer at 65 °C for 2 h followed by 95 °C for 15 min in elution buffer and the addition of Proteinase K. DNA was extracted with the Qiagen DNA Clean and Concentrator Kit (DS205, Zymo Research) and resuspended in 40 µl of Elution buffer. For qPCR, 0.25 µl of DNA was added per reaction using the StepOnePlus™ Real-Time PCR System (Thermo Scientific) coupled with an Eittest™ 3000 UHPLC was used for DIA-MS analysis. A Sputa Moxia software against the Pan-Human library was applied for protein/peptide identification and quantification.

**Enzyme-linked immunosorbent assay (ELISA).** KO-iMPS were transduced with 30 pmol siRNA targeting SRSF2 per million cells by RNAiMAX (Thermo Fisher) and the same amount of non-targeting siRNA was transduced to both WT and KO-iMPS as control. After 24 h transfection, the cells were harvested and seeded 100k/well in 24-well with 1 µg/l P/LPS in the medium. The supernatant was collected after 8 h LPS treatment and centrifuged at 500xg for 5 min at 4 °C to remove any cell debris. The TNF concentration in the supernatant was measured by an ELISA kit (PeproTech, Cat # 900-TM25) followed a standard protocol provided by the kit.

**Microscale thermophoresis (MST).** His6-tagged SRSF2 were labeled with red–tris–NTA dye in PBS-T buffer (Nano-Temper-technology) with a ratio 1:2 dye to protein. The reaction was then incubated for 30 min at room temperature. The interaction of SRSF2 with WASP was measured using MSTR instrument monolight NT.115 in premium coated capillaries supplied by Nano temper. The recombinant WASP purchased from Abnova (Cat # H00007454-P01) is a full-length protein (1-502 a.a.) with a GST tag at the N-terminus that was expressed in an in vitro wheat germ expression system and purified by the Glutathione Sepharose 4 Fast Flow method. A series of 10 µl reaction of 16 different concentrations from 90 nm to 0.165 µM was prepared. The MST reaction was run in water with 1 µl of His6-tagged SRSF2 labeled with red-Tris-NTA to obtain a final concentration of 1 nM. All reactions were done in buffer (50 mM HEPES pH 7.5, 10 mM MgCl2, 100 mM KCl, 0.1 mg/ml BSA, 5% glycerol and 1 mM DTT). The fluorescent molecules were excited with a red laser (640 nm) to monitor the spatial distribution of the molecules in each capillary. Thermophoretic mobility was determined with the addition of a temperature gradient induced by an infrared (IR) laser with an emission wavelength of 1480 nm focused on a local defined sample volume. Upon heating, the protein molecules diffused away from the peak of the temperature and bound and unbound proteins respond differently. The change in the thermophoresis of fluorescent molecules due to binding was plotted and used to calculate the bound protein fraction and the dissociation constant was obtained by fitting the results to a binding isotherm.

**Expression and purification of recombinant human SRSF2.** Human full-length SRSF2 (accession no. Q01130) N-terminally 8x histidine- and SUMO-tagged and C-terminus Strep-tagged E. coli optimized sequence was cloned into pET-24a by GenScript. SRSF2 plasmid was transformed into E. coli strain BL21 (DE3) competent cells (Novagen) and colonies were selected on agar plates containing 50 µg/ml kanamycin. SRSF2 was over-produced by growing the transformed cells in 1 L of 2YT media (Teknova) supplemented with kanamycin. Cells were grown at 37 °C to OD600 of 0.8 and then induced with 0.2 µM isopropyl β-D-thiogalactosidase (IPTG) and incubated further for 19 h at 16 °C. Cells were collected by centrifugation (10,000× g, 10 min) and the clear supernatant was directly loaded onto HisTrap HP 5 ml affinity column (GE Healthcare) pre-equilibrated with buffer A [50 mM NaCl, 0.1 mM imidazole, 0.1% Triton X-100, 0.05% SDS, 10 mM imidazole, 5 mM β-mercaptoethanol, 0.2% NP-40, 1 mM PMSF, 5% glycerol and EDTA free protease inhibitor cocktail tablet/50 ml (Roche, UK)]. All further steps were performed at 4 °C. Cells were lysed enzymatically by adding 2 mg/ml lysozyme and 10 mg/ml protease inhibitors, 10 mM β-mercaptoethanol and 0.2 µg/ml RNaseA and incubated for 30 min on ice and the clear supernatant was centrifuged (22,000×g, 30 min) and the clear supernatant was diluted on to HisTrap HP 5 ml affinity column (GE Healthcare) pre-equilibrated with buffer A [50 mM Tris (pH 8), 700 mM NaCl, 40 mM imidazole, 5 mM β-mercaptoethanol and 5% glycerol]. The column was then washed with ten column volumes (CVs) of buffer A followed by elution gradient with ten CVs using buffer B [50 mM Tris (pH 8), 500 mM NaCl, 700 mM imidazole, 5 mM β-mercaptoethanol and 5% glycerol]. The peak fractions were pooled and dialyzed overnight in a dialysis buffer [50 mM Tris (pH 8), 300 mM NaCl, 5 mM β-mercaptoethanol, and 10% glycerol]. The dialyzed fractions were concentrated, flash frozen and stored at −80 °C.

**Sample preparation for RNA immunoprecipitation sequencing (RIP-seq).** WT and WASP KO-iMPS were harvested after 1-month differentiation with M-CSF and GM-CSF cytokines. About 2 x 109 macrophages were lysed in 500 µl RIPA buffer with protease inhibitor cocktail and were sonicated by sonication on ice (Qsonica XL-2000, Newtown, CT). The lysates were passed through 0.45 µm filter and centrifuged at 10,000× g for 10 min. The supernatant was desalted using Sep-Pak C18 cartridge (Waters). Approximately 1.5 µg of peptide mixture per sample was injected in size exclusion chromatography and the eluates were collected and mixed with equal volume of RIPA lysis buffer with freshly added 0.5 mM DTT, 1x protease inhibitor cocktail (PI), and 100 µM RNaseOUT. After 10 min of incubation at 4 °C, sonication was performed by 30 s on and 45 s off for total 10 min using Bioruptor Pico (Diagenode). The samples were centrifuged at 10,000×g for 10 min at 4 °C and mixed the supernatant with equal volume of RIPA buffer (150 mM KCl, 25 mM Tris-HCl (pH 7.5), 5 mM EDTA, 0.5% NP-40, 0.5 mM DTT, 1x PIC and 100 U RNaseOUT. The 1% sample was saved as input and the rest was incubated with 6 µg WASP antibody (B9) for 2 hours at 4 °C.

**Data-independence mass spectrometry (DIA-MS).** WT iMPS and WASP KO-iMPS were harvested after 1-month differentiation with M-CSF and GM-CSF cytokines. About 2 x 109 macrophages were lysed in 500 µl RIPA buffer with protease inhibitor cocktail and were sonicated by sonication on ice (Qsonica XL-2000, Newtown, CT). The lysates were passed through 0.45 µm filter and centrifuged at 10,000× g for 10 min. The supernatant was desalted using Sep-Pak C18 cartridge (Waters). Approximately 1.5 µg of peptide mixture per sample was injected in size exclusion chromatography and the eluates were collected and mixed with equal volume of RIPA lysis buffer with freshly added 0.5 mM DTT, 1x protease inhibitor cocktail (PI), and 100 µM RNaseOUT. After 10 min of incubation at 4 °C, sonication was performed by 30 s on and 45 s off for total 10 min using Bioruptor Pico (Diagenode). The samples were centrifuged at 10,000×g for 10 min at 4 °C and mixed the supernatant with equal volume of RIPA buffer (150 mM KCl, 25 mM Tris-HCl (pH 7.5), 5 mM EDTA, 0.5% NP-40, 0.5 mM DTT, 1x PIC and 100 U RNaseOUT. The 1% sample was saved as input and the rest was incubated with 6 µg WASP antibody (B9) for 2 hours at 4 °C.
and further incubated with 50 µl Dynabeads for 1 h at 4 °C. After washing the beads twice with RIPA buffer, the beads were resuspended in 56 µl RNase-free water and 33 at 1 x 75 bp using the Illumina HiSeq 2500, paired-end 75 bp using the Illumina NovaSeq(RNA OUtcome followed by incubation at 4 °C for 5 h. For the nuclear fractionation step, the nuclei were isolated using nuclear isolation buffer (1.28 M sucrose, 40 mM Tris-HCl (PH 7.5), 20 mM MgCl2, 4 Tn100, 1 µl PCR and 100 U/ml RNAseOUT) and resuspended in 1 ml RIPA buffer with 1 h incubation at 4 °C. Sonication was performed by 30 s on and 45 s off for total 10 min using Bioruptor®Pico (Diag.

After 13, 000 rpm centrifugation for 10 min, the supernatant was collected, of which 1% was saved as input, the rest incubated with 6 µg WASP antibody (B) overnight at 4 °C. The complexes were captured with Protein A/G-coated beads for 3 h at 4 °C. The beads were washed with RIPA buffer, low salt buffer (20 mM Tris-HCl (PH 7.4), 150 mM NaCl, 2 mM EDTA, 1% Triton, 0.1% SDS), high salt buffer (20 mM Tris-HCl (pH 7.4), 500 mM NaCl, 2 mM EDTA, 1% Triton, 0.1% SDS), low salt buffer, high salt buffer, and RIPA buffer. The beads from the final step of both whole-cell RIP and nuclear fraction RIP and input samples were resus-

sembled in 1 ml Trizol followed by RNA purification. The Qubit Fluorometer was used for RNA concentration measurement and Bioanalyzer was used to determine RNA integrity prior to library preparation.

Antibodies. Antibodies used in this study are listed in Supplementary Data 6.

High-throughput whole transcriptome sequencing. RNA was isolated by Qiagen RNeasy kit with in-column DNase digestion. The Agilent TapeStation or Bioanalyzer was used to determine RNA integrity (RIN) numbers prior to library preparation. Stranded mRNA-Seq libraries were prepared using the TruSeq Stranded mRNA Library Prep Kit (Cat# RS-122-2101, Illumina) or TruSeq RNA Access Library Prep Kit (Cat# RS-301-2001, Illumina) according to the manu-

facturer’s instructions. Briefly, RNA with polyA tail was isolated using poly-T oligos conjugated to magnetic beads. mRNA was then fragmented and reverse-

transcribed into cDNA using cDNA synthesis. The dUTP-incorporated second strand was not amplifi-

ed. cDNA synthesis was then performed in 1 ml TRIzol followed by RNA puri-

fication. The isolation of the nuclei were isolated using nuclear isolation buffer (1.28 M sucrose, 40 mM Tris-HCl (PH 7.4), 150 mM NaCl, 2 mM EDTA, 1% Triton, 0.1% SDS), high-salt buffer (20 mM Tris-HCl (pH 7.4), 500 mM NaCl, 2 mM EDTA, 1% Triton, 0.1% SDS), low salt buffer, high salt buffer, and RIPA buffer. The beads from the final step of both whole-cell RIP and nuclear fraction RIP and input samples were resus-

sembled in 1 ml Trizol followed by RNA purification. The Qubit Fluorometer was used for RNA concentration measurement and Bioanalyzer was used to determine RNA integrity prior to library preparation.

Bioinformatics analysis. Sequencing reads were aligned to the hg19 human genome using STAR (v2.5.3a) with default parameters. Cuffdiff (v2.2.1) (https://www.sabatierlab.org/cufflinks) was used to analyze the differentially expressed (DE) genes and isoforms (-b -u -library-type r-strand) on a pair-wise experiment design. A default cutoff of q-value < 0.05 was used to identify the DE genes. The spliced ratio was defined as the expression of one isoform over all isoforms of a specific gene under one condition. Isoforms with a difference in spliced ratio > 20% between two conditions and FPKM (Fragments Per Kilobase of transcript per Million mapped reads) > 5 in both conditions were identified as differentially spliced (DS) isoforms. Pathway analysis was performed using DAVID (v6.8) (1) where the list of DE genes/DS isoforms was used as input and the whole genome was used as back-

ground. As an alternative isoform expression analysis method, we used STAR v2.5.3a to build a genome index from ENSEMBL GRCh38 human primary ground. As an alternative isoform expression analysis method, we used STAR (v2.5.3a) with default parameters and the soft-clipping option off. The resulted skipped exon events with >10% difference in splicing ratio and FDR < 0.05 were identified as significantly alternatively spliced (AS) exons.

Motif enrichment tests on the exons were performed using Homer (v4.8; PMID: 20513432). The RBBPmap database containing motif information of 94 RNA binding proteins was used to scan the aberrant skipped exons of WASP-deficient cells, and their upstream or downstream exons, respectively. The motif lengths from 4 nt to 15 nt were tested for enrichment with respect to the background. Significantly enriched motifs were defined as motifs with hypergeometric P < 1e-2, greater than 5% enrichment in target sequences, and fold-enrichment > 1.5.

R (http://www.R-project.org/) was used to generate graphs, unless mentioned specifically. Venn diagrams were plotted using the R package VennDiagram (https://doi.org/10.1186/1471-2105-12-33). ChIPpeakAnno package (https://doi. org/10.1186/1471-2105-11-237) was used for the integrated analysis of the WASP ChIP-seq and AASEs.

Raw reads of RNA-seq data of B-cell lines were processed with FASTQC (https://www.bioinformatics.babraham.ac.uk/projects/fastqc/) and BBDuk (https://jgi.doe.gov/data-and-tools/bbtools/) in order to check the quality and remove low-quality bases or adapters. A minimum quality of 25 and a minimum read length of 35 nt were required. High-quality reads were then mapped against the Homo sapiens reference genome (NCBI GRCh38.p12) with STAR. FeatureCounts (https://www.bioinformatics.babraham.ac.uk/projects/featurecounts/) was used to perform read summarization at the gene level, only reads with quality higher than 30 were used. In addition, strand-specific and paired-end modes were included. Statistical analyses and plots were generated with R. Lowly expressed genes were filtered out with the HTSFilter (https://doi.org/10.1093/bioinformatics/bt350) package, then a differential expression analysis was performed using edgeR. Genes with an FDR less or equal than 0.05 were considered significantly differentially expressed. Differential Splicing Analysis was performed with the software RMATS using the trimmed reads as input for mapping. Only splicing events with an FDR < 0.05 were considered differential.

Data availability

The human GRCh38 genome (https://ftp.ncbi.nlm.nih.gov/pub-release/99/fasta/homo_sapiens/) was used as the reference genome. The RNA-seq data generated in this study including raw sequencing data and the processed gene expression tables have been deposited in the NCBI GEO database under the accession number GSE170963 and GSE165382, and in NCBI SRA database within the BioProject under the accession code PRJNA541823. Processed ChIP-seq dependency data generated in this study have been deposited in the NCBI GEO under the accession number GSE126483. The mass spectrometry proteomics data generated in this study have been deposited to the ProteomeXchange Consortium via the PRIDE® partner repository with the dataset identifier PXD003326. All data are available in the main text or the Supplementary Information. Source data are provided with this paper.

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Author contributions

M.L., Y.Z., K.S., and J.C.I.B. conceived the project, designed the experiments, and performed data analysis. M.L., B.Y., X.Z., K.S., G.R.M., M.W., M.T., L.V.C., J.J.M., S.D., R.H.B., T.H., N.Y.K., M.M.A., M.K., Y.T., J.Q., E.A., J.Q., G.L., Z.Y., F.L., C.B., Y.G., C.S., J.R., Y.L., X.D.F., C.R.E., and S.H. contributed to project design and conducted the experiments. L.H., C.B., and M.S. carried out bioinformatics work and interpreted data. P.C. and F.F. contributed to the design and experiments related to the primary patient samples. B.Y., X.Z., M.W., J.C.I.B., and M.L. wrote the paper.

Competing interests

The authors declare no competing interests.

Additional information

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