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Characterization of a relaxase belonging to the MOB\(_T\) family, a widespread family in Firmicutes mediating the transfer of ICEs

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Abstract

Background: Conjugative spread of antibiotic resistance and virulence genes in bacteria constitutes an important threat to public health. Beyond the well-known conjugative plasmids, recent genome analyses have shown that integrative and conjugative elements (ICEs) are the most widespread conjugative elements, even if their transfer mechanism has been little studied until now. The initiator of conjugation is the relaxase, a protein catalyzing a site-specific nick on the origin of transfer (ori\(T\)) of the ICE. Besides canonical relaxases, recent studies revealed non-canonical ones, such as relaxases of the MOB\(_T\) family that are related to rolling-circle replication proteins of the Rep\(_\text{trans}\) family. MOB\(_T\) relaxases are encoded by ICEs of the ICE\(_{St3}\)/ICE\(_{Bs1}\)/Tn\(_{916}\) superfamily, a superfamily widespread in Firmicutes, and frequently conferring antibiotic resistance.

Results: Here, we present the first biochemical and structural characterization of a MOB\(_T\) relaxase: the Rel\(_{St3}\) relaxase encoded by ICE\(_{St3}\) from \(\text{Streptococcus thermophilus}\). We identified the ori\(T\) region of ICE\(_{St3}\) and demonstrated that Rel\(_{St3}\) is required for its conjugative transfer. The purified Rel\(_{St3}\) protein is a stable dimer that provides a Mn\(^{2+}\)-dependent single-stranded endonuclease activity. Sequence comparisons of MOB\(_T\) relaxases led to the identification of MOB\(_T\) conserved motifs. These motifs, together with the construction of a 3D model of the relaxase domain of Rel\(_{St3}\), allowed us to determine conserved residues of the Rel\(_{St3}\) active site. The involvement of these residues in DNA nicking activity was demonstrated by targeted mutagenesis.

Conclusions: All together, this work argues in favor of MOB\(_T\) being a full family of non-canonical relaxases. The biochemical and structural characterization of a MOB\(_T\) member provides new insights on the molecular mechanism of conjugative transfer mediated by ICEs in Gram-positive bacteria. This could be a first step towards conceiving rational strategies to control gene transfer in these bacteria.

Keywords: Integrative and conjugative elements, Relaxase, MOB\(_T\), Conjugation, Tn\(_{916}\), Firmicutes

Background

Horizontal gene transfer (HGT) drives the evolution of bacterial genomes and plays an important role in the adaptation of bacteria to different environments or the colonization of novel niches. Conjugative elements are major actors that trigger HGT\(^{1, 2}\). Besides conjugative plasmids, many integrative and conjugative elements (ICEs) in the host chromosome encode their own DNA transfer from donor to recipient cells by conjugation\(^{3–5}\). Genome analyses indicate that ICEs are widespread in most phyla of bacteria, and that many of them carry adaptive genes such as antibiotic resistance or virulence genes\(^{3, 4, 6, 7}\), hence contributing to the dissemination of these traits among bacterial populations. Thus, many Firmicutes have recently become resistant to antibiotics through ICE transfer, including various strains of Clostridia, Enterococci, Staphylococci and Streptococci, some of which are severe human or zoonotic pathogens\(^{8–14}\). Although Gram-positive ICEs

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Relaxases, also termed Mob (for mobilization) proteins, belong to different families of proteins. The canonical oriT organizes the origin of transfer (multi-protein complex called the relaxosome, that recognizes oriT on the excised ICE DNA to be transferred [17]. As demonstrated for conjugative plasmids, the main protein of the relaxosome is the relaxase, a trans-esterase enzyme encoded by ICEs. The relaxase introduces a single-stranded nick at the site, leading to a re-circularization of the ssDNA in the recipient cell [20].

Relaxases, also termed Mob (for mobilization) proteins, belong to different families of proteins. The canonical relaxases harbor a histidine triad with a ‘HxH’ signature that is known to coordinate a metallic cofactor [21]. They include the MOB\(_F\), MOB\(_M\), MOB\(_L\), MOB\(_Q\), MOB\(_V\) and MOB\(_B\) families [7, 18, 22, 23]. This ‘HxH’ motif is also found in rolling-circle replication initiators of numerous plasmids and viruses, usually termed Rep proteins [24]. Several other families of relaxases have been identified which do not harbor the ‘HxH’ signature and which are unrelated to canonical relaxases [23]. The best described are the MOB\(_H\) family related to HD-hydrolases [23, 25], and the MOB\(_C\) family of relaxases that has no typical relaxase motif, but surprisingly has been shown to be related to restriction endonucleases [26]. Additionally, recent observations indicate that several other protein families also function as relaxases, especially in Firmicutes. This is the case of the TcpM protein from the pCW3 conjugative plasmid (Clostridium perfringens) that is related to tyrosine recombinases [27]. Furthermore, rolling circle replication (RCR) initiators of the Rep\(_1\) family have been found to act as a relaxase, being responsible for the mobilization of plasmids by ICEs, in addition to their role in plasmid maintenance [28].

Prior to transfer, ICEs excise from the host chromosome as a circular form. As in conjugative plasmids, the initiation of conjugative transfer is likely performed by a multi-protein complex called the relaxosome, that recognizes the origin of transfer (oriT) on the excised ICE DNA to be transferred [17]. As demonstrated for conjugative plasmids, the main protein of the relaxosome is the relaxase, a trans-esterase enzyme encoded by ICEs. The relaxase introduces a single-stranded nick at the site, leading to a re-circularization of the ssDNA in the recipient cell [19]. The relaxase is then thought to join the transferred ssDNA at the nic site of oriT. Whereas the 3’-end of the nick is used to initiate rolling-circle replication of the element, the relaxase remains covalently bound to the 5’-end via a catalytic tyrosine residue [18]. A coupling protein is assumed to recruit the relaxase-DNA complex to a type-IV secretion system (T4SS) encoded by the ICE, providing energy to translocate this complex into the recipient cell [19].

Rolling Circle Replication (RCR) initiators encoded by the \textit{Rep\_trans} family which are involved in the maintenance of many small Firmicutes plasmids [34]. A structural and biochemical characterization of the active site of the \textit{Rep\_trans} RCR initiator protein encoded by the pSTK1 plasmid (\textit{Geobacillus stearothermophilus}) has been performed recently [34].

From a structural point of view, 3D structures are available for three canonical relaxase families: MOB\(_T\) [23, 35–37], MOB\(_Q\) [23, 38], and MOB\(_V\) [39]. To date, no 3D structure is known for the non-canonical relaxases (MOB\(_B\), MOB\(_C\), and MOB\(_B\)).

In this work, we characterized the MOB\(_T\) relaxase encoded by ICE\(S\)3/Tn916 superfamily of ICEs, which is widespread in Firmicutes, also encodes non-canonical relaxases. These relaxases were unclassified by Garcia-Barcia et al [23], but were grouped in a family called MOB\(_T\) in 2011 [7]. Interestingly, putative MOB\(_T\) relaxases have also been detected in a number of putative integrated and mobilizable elements (IMEs) in streptococcal genomes [29, 30]. Up to now, the activity of only two MOB\(_T\) relaxases has been demonstrated, namely those encoded by Tn916 (Orf20) and ICEBs1 (NicK). The Orf20 protein was reported to involve the Tn916 integrase for DNA sequence specificity [31]. However, this result needs to be confirmed because the Orf20 version used lacked the N-terminal helix-turn-helix (HTH) DNA-binding domain due to misidentification of the start codon [32]. The NicK relaxase has been shown to be essential for conjugation and the nic site within ICEBs1 oriT was identified [33]. Up to now, no MOB\(_T\) relaxase has been characterized biochemically or structurally.

MOB\(_T\) relaxases are non-canonical relaxases that harbor a PF02486 domain. This domain is also found within Rolling Circle Replication (RCR) initiator proteins of the \textit{Rep\_trans} family which are involved in the maintenance of many small Firmicutes plasmids [34]. A structural and biochemical characterization of the active site of the \textit{Rep\_trans} RCR initiator protein encoded by the pSTK1 plasmid (\textit{Geobacillus stearothermophilus}) has been performed recently [34].

The RELSt3 ICEBs1/Tn916 superfamily of ICEs, which is widespread in Firmicutes, also encodes non-canonical relaxases. These relaxases were unclassified by Garcia-Barcia et al [23], but were grouped in a family called MOB\(_T\) in 2011 [7]. Interestingly, putative MOB\(_T\) relaxases have also been detected in a number of putative integrated and mobilizable elements (IMEs) in streptococcal genomes [29, 30]. Up to now, the activity of only two MOB\(_T\) relaxases has been demonstrated, namely those encoded by Tn916 (Orf20) and ICEBs1 (NicK). The Orf20 protein was reported to involve the Tn916 integrase for DNA sequence specificity [31]. However, this result needs to be confirmed because the Orf20 version used lacked the N-terminal helix-turn-helix (HTH) DNA-binding domain due to misidentification of the start codon [32]. The NicK relaxase has been shown to be essential for conjugation and the nic site within ICEBs1 oriT was identified [33]. Up to now, no MOB\(_T\) relaxase has been characterized biochemically or structurally.

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To investigate further this relationship, a selected set of MOB_T relaxase sequences including RelSt3 was phylogenetically analyzed together with Rep_trans RCR initiator sequences from staphylococcal plasmids (Fig. 1a). The MOB_T sequences selected for this analysis include relaxases encoded by the characterized ICEs ICEBs1 (NicK), Tn916 (Orf20), ICE_S15_tRNA^lys, ICEcp1, ICE6013, Tn6202, Tn6098, and nisin-sucrose transposon [31, 33, 40–45], and putative relaxases encoded by several Tn916-related elements from different bacterial phyla, and by other distantly related ICEs belonging to the ICESt3/ICEBs1/Tn916 superfamily. Rep_trans sequences include RepSTK1 whose 3-D structure was recently solved [34], and several members of the Rep proteins belonging to the pT181 family [46]. As expected, RelSt3 groups with the MOB_T proteins (Fig. 1a). The topology of the tree clearly indicated that MOB_T and Rep_trans from the pT181 family are distantly related (Fig. 1a). Interestingly, the RepSTK1 sequence grouped with MOB_T sequences rather than with other

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**Fig. 1** Relationships between Rep_trans RCR initiators and MOB_T relaxases. 

- **a** Unrooted maximum likelihood tree of MOB_T and Rep_trans proteins. Fifty-four homologous positions were selected manually for phylogenetic analysis by PhyML. Bootstrap values (100 replicates) above 50% are indicated. The scale bar shows the number of substitutions per site. MOB_T proteins: RelSt3 encoded by ICESt3 from *S. thermophilus* [89], RelICE_S15 encoded by ICE_S15_tRNA^lys from *S. agalactiae* [40], Orf20-Tn916 encoded by ICE Tn916 from *Enterococcus faecalis* [32], NicK_ICEBs1 encoded by ICEBs1 from *B. subtilis* [33], Rel_Tn6202 encoded by Tn6202 from *E. faecalis*, conferring vancomycin resistance [41], Rel_ICE_SmuUA159 encoded by ICE_SmuUA159_tRNA^leu from *S. mutans*, Rel_Tn6098 encoded by Tn6098 from *Lactococcus lactis*, and 4 other Tn916 relaxases from strains of different phyla: Rel_Tn916B from *Bifidobacterium longum*, Rel_Tn916M from *Mycobacterium abscessus*, Rel_Tn916S from *Streptomyces alboniger*, and Rel_Tn916C from *Chlamydia trachomatis*. Rep_trans proteins: RepSTK1 corresponds to the Rep_trans RCR initiator encoded by the plasmid pSTK1 from *Gardiacillus stearothermophilus* [90], whose structure have been solved recently (PDB ID 4CIJ) [34]. Besides the well-characterized RepC (pT181) and RepD (pC221) proteins, Rep_trans proteins from 5 other members of the pT181 family of staphylococcal plasmids were included: RepE (pS194), RepJ (pUB112), RepI (pC223), RepN (pcW7) and the Rep from the pH6 plasmid [46]. 

- **b** MOB_T relaxases (red boxes) and Rep_trans proteins (blue boxes) are linked according to their global sequence identity (see the bottom panel). 

- **c** Weblogos of the six conserved motifs identified for MOB_T relaxase domain. 2016 sequences recovered through CDART analysis with RelSt3 sequence as a query were clustered with a sequence identity of 90%, and this WebLogo was obtained using a representative of each of the 261 clusters obtained. The Chemistry color scheme was chosen for representation of amino-acid residues (WebLogo 3 application): green for polar, purple for neutral, blue for basic, red for acidic, and black for hydrophobic residues. Each motif is numbered on the top, and the motif boundaries in the RelSt3 sequence are indicated on the bottom. See Material and Methods for details.
Rep_trans proteins, even if this topology was not clearly supported by bootstrap analysis. This relationship between RepSTK1 and MOB_T proteins was also supported by sequence-based searches for fold similarities using the Phyre2 [47] and iTasser [48] programs. Indeed, the use of any MOB_T sequence as query gave the RepSTK1 structure as first hit, with the RepDE structure that belongs to the pT181 family being the second hit [34]. A comparison of sequence identities also illustrates that RepSTK1 is closer to MOB_T relaxases than the pT181 cluster (Fig. 1b). As expected, relaxases encoded by Tn916 and Tn916-like elements clustered together among MOB_T proteins with a good bootstrap support. RelSt3 is more closely related to the relaxase encoded by ICE_S15_tRNA from S. agalactiae than to NicK encoded by ICEBs1 from B. subtilis. These relationships were also confirmed by multiple alignments of MOB_T and Rep_trans protein sequences (Additional files 1 and 2: Figure S1 and S2), which allowed the identification in RelSt3 and other MOB_T relaxases of the 5 conserved motifs described in Rep_trans proteins by Carr et al [34]. Taken together, these data argue in favor of RelSt3 being a bona fide MOB_T relaxase.

To identify conserved motifs among MOB_T proteins, we recovered 2016 sequences of proteins harboring the same architecture as RelSt3, i.e. with a N-terminal HTH domain and a PF02486 domain (see Materials and Methods for details). To our knowledge, this architecture is only found in proteins encoded by ICEs (and not by plasmids), suggesting that they correspond to relaxases involved in conjugation and not to RCR Rep_trans proteins. Multiple sequence alignment followed by WebLogo analysis allowed the identification of 6 conserved motifs (Fig. 1c), distinct from all other motifs found in the other MOB families [22, 23]. Among these motifs, 5 have already been identified in Rep_trans proteins, whereas an additional motif, called N′′, was found between motifs N′ and I (Fig. 1c). The 3 acidic residues involved in cation coordination in Rep_trans proteins [34] were also conserved in the same 3 motifs in MOB_T relaxases: motifs N′ (D72 in RelSt3 sequence), I (D177) and IV (D280). In motif III, the catalytic tyrosine identified in Rep_trans proteins [49] is also conserved (Y252 in RelSt3 sequence).

The RelSt3 gene (orfJ) is required for ICESt3 conjugation
If it encodes a functional relaxase, the orfJ gene is expected to be essential for ICESt3 conjugative transfer, as demonstrated for the related gene nicK from ICEBs1 [33]. We constructed a S. thermophilus strain carrying a derivative of ICESt3cat deleted for the orfJ gene (referred as LMG18311 ICESt3DeorfJcat, see Additional file 4, Supplementary Materials and methods). The ICESt3 conjugation/recombination modules are composed of 17 genes transcriptionally coupled, including orfJ at the fifth position downstream of the Pcr promoter [50]. Our strategy was to delete the orfJ CDS while keeping the predicted oriT region and other ICESt3cat genes intact, without introducing any foreign sequence. The mating frequency for the LMG18311 ICESt3cat [51] as donor strain was $1.0 \times 10^{-8} \pm 2.0 \times 10^{-5}$ transconjugants per donor cell. No transconjugant was observed when three independent clones of LMG18311 ICESt3DeorfJcat were used (with a detection level $\geq 10^{-9}$ transconjugants per donor cell), thus demonstrating that the orfJ gene is required for ICESt3 conjugation.

Determination of ICESt3 oriT
In order to determine the location of ICESt3 oriT, we constructed the pOr1180-oriT plasmid harboring the circa 200 bp sequence of the intergenic orfK/orfJ region from ICESt3, where the conserved nic site was previously predicted [52]. This plasmid carrying a spectinomycin resistance marker was tested for mobilization in trans by ICESt3, and was successfully transferred to recipient cells with a mobilization frequency of $2.5 \times 10^{-3}$. In contrast, no transconjugant was obtained with the plasmid lacking the orfK-orfJ intergenic region (below the $10^{-6}$ detection level) (Table 1). These results confirmed that the oriT sequence is located between orfK and orfJ genes in ICESt3 sequence, and indicated that RelSt3 is able to mobilize a replicon in trans.

RelSt3 is dimeric in solution
RelSt3 was over-expressed and purified from E. coli using affinity (nickel) chromatography (Fig. 2a) as described in Additional file 4, Supplementary Materials and Methods. RelSt3 was purified to homogeneity, with a yield of about 1.5 mg per g of cell pellet. As the final gel filtration step was performed with a calibrated Superdex 200 column (Additional file 3, Figure S3), the elution volume of the single elution peak (72.4 mL) allowed us to estimate the apparent molecular weight of the protein in solution to be about 88 kDa (Fig. 2b). As the predicted size of RelSt3 is 48.5 kDa, this finding suggested that RelSt3 is dimeric.

The RelSt3 protein was also submitted to chemical cross-linking experiments. To do so, RelSt3 was incubated with increasing concentrations of glutaraldehyde (0.001 to 0.1%, Fig. 2c), and a well-defined band appeared with 0.01% glutaraldehyde around 100 kDa, a molecular mass corresponding to twice the predicted mass of the monomer. At higher concentration of cross-linker, higher molecular mass species appeared.

We next studied the protein homogeneity by velocity sedimentation in order to determine the molecular weight and radius of RelSt3 in solution (Fig. 2d). Data were collected using 1 and 5 mM of protein, which both gave
similar results. Most of the RelSt3 absorbance corresponded to a single peak, indicating the absence of higher aggregates, and demonstrating that the oligomeric form of RelSt3 is homogeneous at these protein concentrations. The Sedfit program was used to analyze the c(s) distribution model. The unique peak was associated with a sedimentation coefficient (S20, w) of 5.2 S, a frictional ratio of 1.5 and a hydrodynamic radius of 4.4 nm. Taken together, these data indicate that the RelSt3 protein is an elongated dimer in solution.

Secondary structure content and stability of RelSt3
Using various computational methods, the secondary structure of RelSt3 protein was predicted to consist of 41.7% α–helixes, 14.8% β-strands and 43.6% random coils (Table 2). Circular dichroism (CD) was used to analyze RelSt3 folding experimentally. Far-UV spectra were recorded at 20 °C. The spectral data were converted into mean residue ellipticity (Fig. 3a) and showed a maximum at 195 nm, a characteristic feature of β-strand structures, and two minima at 208 and 222 nm, typical of α–helix content [53]. Deconvolution using

Table 1 ICESt3 mobilization in trans of an oriT-harboring plasmid

| Donor strain | Recipient strain | Plasmid ICESt3 | ICESt3 transconjugants frequency | Plasmid mobilization frequency |
|--------------|------------------|----------------|----------------------------------|-------------------------------|
| LMG18311 (ICESt3cat) | LMG18311 (pOri1180) | pOri1180 | 8.8 × 10^{-3} | < 10^{-6} |
| LMG18311 (ICESt3cat) | LMG18311 (pOri1180-oriT) | 2.0 × 10^{-4} | 2.5 × 10^{-3} |

Fig. 2 Purification of RelSt3 protein and determination of its oligomeric state. a Different steps of RelSt3 purification from E. coli (12% SDS-PAGE). Crude extract (lane 1) was submitted to a His-Trap affinity chromatography. Flow-through analysis (lane 2) indicated that most of the protein was retained. The pool of imidazole elution fractions (lane 3) was then digested by the TEV protease to remove His-tag (lane 4), and the sample was then loaded on a second His affinity chromatography. Without tag, the protein was not retained (lane 5), and was concentrated to be injected on a gel filtration column. Finally, the resulting RelSt3-containing fractions were concentrated (lane 6). Lane 7 corresponds to the Spectra Multicolor Broad Range protein ladder (Thermo Scientific). The gel was stained with Coomassie Brilliant Blue. b Gel filtration (Sephadex 200) analysis of RelSt3 protein. The elution profile recorded showed a large main peak with a maximum elution volume of 72.4 ml. c Glutaraldehyde cross-linking of RelSt3 purified protein. 5 µg of RelSt3 protein were incubated without (lane 1) or with 0.001% (lane 2), 0.01% (lane 3), 0.025% (lane 4), 0.05% (lane 5) or 0.1% (lane 6) of glutaraldehyde. The products were then analyzed by SDS-PAGE (10%), with a Coomassie Brilliant Blue staining. See Materials and Methods for details. Lane 7: Spectra Multicolor Broad Range protein ladder. d Analysis of the dispersion state of RelSt3 by analytical ultracentrifugation. Sedimentation velocity of RelSt3 was analyzed at 5 µM. The diagram represents the sedimentation coefficient distribution [c(s)].
different methods provided an average content of 39.8% α-helix, 19.4% β-strands, 17.0% turns, and 24.4% unordered (Table 2), indicating that this experimentally determined secondary structure content is in agreement with the in silico prediction. This mixed α/β proportion is similar to those of other CD-characterized relaxase proteins, including MobM from pMV158 and TraA from pIP501 [54, 55], as well as known structures of other relaxases [35, 36, 56, 57].

To estimate the stability of RelSt3 folding, thermal denaturation was monitored from 20 °C to 80 °C at 222 nm (typical minimum ellipticity of α-helix) and at 195 nm (typical maximum ellipticity of β-strands). Approximately 75% of the ellipticity at 195 nm and 55% of the ellipticity at 222 nm were lost between 20 and 80 °C, with a one-step cooperative transition midpoint at 55 °C (Fig. 3b). After cooling back to 20 °C, almost no ellipticity was recovered (data not shown), indicating that the thermal denaturation is irreversible in our conditions. In conclusion, RelSt3 is a well-folded protein, but is sensitive to denaturation, that is consistent with the observation of protein aggregates during RelSt3 purification trials in presence of ionic force lower than 500 mM NaCl.

Using DSC experiments, we next investigated the overall stability of RelSt3. RelSt3 denaturation occurred in a single step with an enthalpy variation of 144 kcal/mol and a determined Tm of 50.95 °C (Fig. 3c), which is in good accordance with our CD results.

### Table 2 Secondary structure content of RelSt3

| Prediction programs | α-helix | β-strand | Turns | Unordered |
|---------------------|---------|----------|-------|-----------|
| PSI-pred            | 43.6    | 10.7     | ND    | 45.7      |
| SABLE               | 47.3    | 12.7     | ND    | 40        |
| Jpred4              | 42.2    | 14.4     | ND    | 43.4      |
| PredictProtein      | 38.5    | 15.1     | ND    | 46.4      |
| APSSP2              | 36.3    | 18.8     | ND    | 44.9      |
| Phyre2              | 42      | 17       | ND    | 41        |
| **Average**         | 41.7 ± 3.9 | 14.8 ± 2.9 | 43.6 ± 2.6 |

Data and standard deviation are given in percentages. ND: not determined

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### RelSt3 structure model

To obtain further structural data of a MOB_T relaxase, we built a 3D model of the tertiary structure of the RelSt3 relaxase domain (PFO2486) by homology. As phylogenetic analysis indicated that RepSTK1 was the closest protein to RelSt3 for which an experimental structure has been solved (Fig. 1), the X-ray structure of RepSTK1 (PDB 4CII) was used as a template. Like the RepSTK1 structure, the RelSt3 model is crescent-shaped, with two lobes of similar secondary structure patterns, each involving 5 beta-strands: β-α-(α)-β-β-β-α-α-α (N-lobe) and β-(α)-β-β-β-α-α-β (C-lobe) (Fig. 4) [34]. The underlined elements are absent from the template structure RepSTK1. The two 5-strand beta-sheets are adjacent and form a continuous 10-strand sheet, whose structure and position in the sequence are highly conserved with Rep_trans proteins (despite RelSt3 and Rep_trans proteins being poorly conserved in sequence, Fig. 1b).

In our homology model of RelSt3 relaxase domain, the secondary and tertiary structures are very well conserved compared to the template, and stable over the dynamic simulation. The main differences are a shorter β1-α1 loop (20 vs 27 residues), a longer β9-β10 linker (23 vs 12 residues) including an additional helix (α8), and shorter β7 and β8 strands (3–4 vs 8–9 residues). The parts of our model that correspond to the active site in the template are located at the junction of the two 5-strands sheets, involving the adjacent strands β1 (N-lobe) and β6-β9-β10 (C-lobe), and a short helix downstream of β9 (α7, Figs. 4 and 7).

### In vitro relaxase activity of RelSt3

The relaxation activity of RelSt3 was assayed as previously described by Lorenzo-Diaz et al [54], using a pBR322 plasmid harbouring ICESt3 oriT (Additional file 4; Table S2). We were able to demonstrate a nicking activity on this supercoiled substrate (FI), leading to the formation of a relaxed form of the plasmid (FII) (Fig. 5a). To determine the optimal conditions for this relaxation activity, different ions (Fig. 5a), incubation times (Fig. 5b), and cations (Fig. 6a) were tested. As shown in Fig. 5a and b, a 20 min incubation at 30 °C of RelSt3 with supercoiled plasmid appeared to give a maximum amount of relaxed form FII. A small amount of linear plasmid was also observed (FIIL), probably as a by-product from already nicked DNA. A dose-response experiment was also carried out (Fig. 5c) indicating that the nicking activity is dependent on RelSt3 concentration. As shown for Rep_trans proteins [34, 58], no activity was observed without any cationic cofactor (Fig. 6a) indicating that RelSt3 activity depends on the presence of a cationic cofactor. Whereas Ca²⁺, Zn²⁺ and Ni²⁺ were not bona fide cofactors for RelSt3, the presence of Co²⁺, and especially Mg²⁺ or Mn²⁺ is needed for RelSt3 relaxation activity (Fig. 6a). The RelSt3 activity is dependent on Mn²⁺ concentration, and a maximal conversion of form I was observed above 1 mM Mn²⁺ (Fig. 6b).

### Characterization of RelSt3 variants affected in their nicking activity

As mentioned above, multiple alignment of protein sequences (Additional files 1 and 2; Figure S1 and S2) together
with our 3D homology model (Fig. 4) identified motifs in RelSt3 similar to conserved motifs proposed for Rep_trans proteins [34]. These motifs are also found in other MOB T relaxases (Fig. 1c). Carr et al (2016) demonstrated that 3 acidic residues located in 3 different motifs (N’, I and IV, shown by a star in Additional file 1: Figure S1) are involved in the coordination of the cationic cofactor in Rep_trans proteins. As these acidic residues are conserved in the RelSt3 sequence (D72, D177, E280, Fig. 1c and Additional file 1: Figure S1), each one was independently mutated into alanine in order to assess its involvement in RelSt3 activity. In motif III (Additional file 1: Figure S1), the RelSt3 tyrosine Y252 is conserved within Rep_trans proteins from plasmids of the pT181 family. The corresponding residue (Y188) of the RepD protein from the staphylococcal plasmid pC221 has already been demonstrated to be catalytic [49]. To confirm that this residue is also catalytic in a MOB T protein, a Y252A variant of RelSt3 was generated. Two other conserved residues were also mutared, K254 and E257, located in the vicinity of the putative catalytic Y252 residue in motif III. All variants were expressed using the same expression vector and with the same purification protocol as for the purification of the WT protein. The nicking activity of each variant was compared to the WT (Fig. 5 and Fig. 7a) with a standard dose-response experiment using Mn2+ as cofactor.

As shown in Fig. 7a, each RelSt3 single mutant D72A, D177A and E280A had completely lost the nicking activity, which is consistent with the results obtained for Rep_trans proteins modified at these conserved positions [34]. The Y252A variant also showed an absence of nicking activity, suggesting that this residue plays a role of catalytic tyrosine for RelSt3. Concerning the two other conserved residues in

![Secondary structure analysis of RelSt3 folding and stability.](image)
Fig. 4 Homology 3-D model of RelSt3 core-domain. a Schematic view of the secondary structure of RelSt3 core domain (PF02486). Beta-strands are represented as arrows and helices as cylinders, with lengths proportional to the numbers of residues. The N- and C-lobes, each including a 5-strand beta-sheet, are coloured in blue and pink, respectively. Loops are represented as grey lines, with white circles indicating their length (number of residues). The five motifs conserved both in MOB₂ and Rep_trans proteins are boxed in dotted rectangles. In addition, the motif N° conserved in MOB₂ proteins identified in this work is also boxed. The residues mutated in this work are indicated and coloured as follows: yellow for residues chelating the divalent cation, blue for the catalytic conserved tyrosine, green for other conserved residues. b Front view of the homology 3-D model of RelSt3 core-domain. Beta-strands are coloured as in Fig. 4a. Helices and loops are in green. Important residues of the active site are represented as sticks and coloured as in Fig. 4a. c Side view of the 3-D RelSt3 model, illustrating the crescent-shaped structure with the active site in the centre. The yellow ball indicates the probable location of the divalent cofactor, according to the model.

Fig. 5 RelSt3 harbours nicking activity. In all assays, 200 ng of pBR322-oriT plasmidic DNA were incubated with the indicated amount of RelSt3 protein. a RelSt3 nicking activity according to temperature. pBR322-oriT plasmid was incubated without or with 2 μg of RelSt3 protein for 20 min at 4, 10, 20, or 30 °C in the presence of 5 mM Mn²⁺. FI: supercoiled plasmid DNA. FII: open circular form. FIII: linear form. b Time course in the presence of 2 μg of RelSt3 and 5 mM Mn²⁺ at 30 °C. The nicking activity was established by band quantification monitoring the decrease of FI form. c Dose response of RelSt3 nicking activity. DNA was incubated for 20 min at 30 °C with 5 mM Mn²⁺ without (lane 1) or with 0.02 (lane 2), 0.05 (lane 3), 0.2 (lane 4), 0.5 (lane 5), 1 (lane 6), 2 (lane 7), 5 (lane 8), 7 (lane 9) or 10 (lane 10) μg of RelSt3 protein.
motif III, K254 appeared to be essential, whereas E257 was found to be important but not essential for DNA relaxation (Fig. 7a). The positions of these residues in the active site structure model of RelSt3 (Fig. 7b) suggest that E257 could be involved in maintaining the catalytic Tyr in the correct position, whereas K254 could be involved in the stabilization of the acidic residues assumed to coordinate the cation (especially D72 and E280, with Calpha-Calpha distances of 10.5 and 13.0 Å respectively, and maximal side-chain sizes of 6.4 Å, 3.8 Å and 4.8 Å for K254, D72 and E280, respectively).

Discussion
In this work, we undertook the biochemical characterization of the RelSt3 protein encoded by ICESt3 of *Streptococcus thermophilus*, a MOB$_T$ relaxase. MOB$_T$ relaxases are encoded by ICEs belonging to the ICESt3/ICEBs1/Tn916 superfamily found in many Firmicutes [15]. We highlighted here the relationship between the MOB$_T$ family of relaxases and the RCR initiator proteins of the Rep$_{trans}$ family, including Rep proteins encoded by the pT181 plasmid family and the structurally characterized RepSTK1 from *Geobacillus stearothermophilus* [34]. Interestingly, an activity of RCR initiation was discovered for several MOB$_T$ relaxases encoded by ICEs (Orf20 and Nick). Indeed, these relaxases were shown to be responsible for autonomous RCR initiation starting from the oriT [32, 59]. Reciprocally, it has already been demonstrated that RCR initiators involved in the maintenance of small plasmids from Firmicutes can also act as conjugative relaxases. Indeed, these plasmids are still mobilized in trans by ICEBs1 deleted from its relaxase gene [28]. However, this mobilization was only demonstrated for RCR plasmids encoding a Rep_1 initiator protein, and was not tested for a RCR plasmid encoding a Rep$_{trans}$ protein.

The purified RelSt3 protein was demonstrated to be homogeneously dimeric. This dimeric state is in agreement with the Rep$_{trans}$ 3D structures solved recently, where the corresponding proteins crystallized as dimers [34]. We demonstrated the relaxation activity of RelSt3 on a plasmidic DNA harboring ICESt3 oriT, confirming the functionality of this protein as a relaxase for ICESt3. As observed for Rep$_{trans}$ proteins, this single-strand endonuclease activity is dependent on the presence of a divalent metal cofactor that turned out to be preferentially Mg$^{2+}$ or Mn$^{2+}$. Although historically the activity of most relaxases was mainly tested with Mg$^{2+}$ [22, 60–62], when compared with Mn$^{2+}$ the latter was found (i) to be more efficient for nicking activity [54, 63], or (ii) to have a better affinity as measured by micro-calorimetry [54, 64], or (iii) to be the most probable cation according to structural and ICP-MS data [35]. We also noticed that Mn$^{2+}$ was identified in the RepSTK1 structure provided by Carr et al [34]. Accordingly, we observed a slightly higher efficiency in relaxation assays for RelSt3 with Mn$^{2+}$ compared to Mg$^{2+}$.

The secondary structure of the RelSt3 protein deduced from the circular dichroism analysis is very similar to the in silico predictions. A mix of $\alpha$–helices and $\beta$–strands is observed as for most, if not all, characterized RCR Rep and Mob proteins [21, 26, 34, 39]. The thermal stability of a few canonical relaxases has already been studied using circular dichroism, and the addition of a
cationic cofactor induced a stabilization of the protein at higher temperatures. Whereas the MobA_R1162 and TraA_pIP501 (MOBQ) relaxases present a unique and short transition with a midpoint around 42 °C, indicating a cooperative unfolding [55, 63], the MobM_pMV158 (MOBV) relaxase harbors a complex denaturation profile with two transitions (midpoints at 21 and 42 °C) spanning more than 30 °C [54]. In comparison, the MOB_T relaxase RelSt3 exhibits a one-step transition as shown by CD and DSC experiments, with midpoints at 55 and 51 °C, respectively. This is clearly higher than observed for other characterized relaxases, indicating that RelSt3 could be intrinsically more stable.

Phenotype analyses of RelSt3 point mutations confirmed the relationship between MOB_T and Rep_trans proteins. We showed that the catalytic tyrosine of RelSt3 is located in the conserved motif III as in Rep_trans proteins. This is the first characterization of catalytic motif for a MOB_T relaxase. We also demonstrated the involvement of the 3 conserved acidic residues D72, D177 and E280 (located in motifs N', I and IV, respectively) in relaxation activity, as demonstrated for RepD variants [34]. These results suggest that both MOB_T and Rep_trans families (i.e. the PF02486 domain) use 3 distant acidic residues in primary sequence to coordinate their cationic cofactor. In addition, we analyzed the involvement of two other conserved residues of motif III in RelSt3 activity, K254 and E257. Both residues are important for RelSt3 nicking activity, indicating that they could assume a structural role within the active site. Indeed, our 3D structural model of the PF02486 RelSt3 domain suggests that E257 could interact with the catalytic Tyr, and that K254 could interact with the acidic residues D72 and E280 that are assumed to coordinate the cation, maintaining them in the correct position.
The 3D structure model based on RepSTK1 crystal structure revealed a fold that resembles classical HxH endonucleases [21], as it presents a central 10-strand beta sheet (2 × 5 strands) decorated by several alpha-helices. Whereas HxH endonucleases harbour their 3 histidine residues that coordinate the cationic cofactor on two of the central beta-sheets, RelSt3 as well as Rep_trans structures instead use 3 acidic residues to do so, each located on a different strand of the central sheet. This correlates with the quite important distance between each of these residues in the primary sequence, lying respectively in motifs N’, I and IV, compared to the classical 3 His that are close to each other in the primary sequence. Another striking difference with HxH endonucleases is that RelSt3 and Rep_trans structures harbour their conserved catalytic Tyr residue on one of the central beta-strands (strand beta9 in RelSt3 model), whereas the HxH proteins have their catalytic Tyr residue(s) on an alpha-helix close to the cationic cofactor. These observations suggest that the catalytic assembly orchestrating a Tyr residue and a divalent cation to cleave a DNA substrate arose at least twice in evolution, giving rise to the HxH endonucleases on one hand, and the Rep_trans/MOB_T families (harbouring the PF02486 domain) on the other hand, as a remarkable example of evolutionary convergence.

Conclusions
Even if two MOB_T relaxases were identified circa 10 years ago (for Tn916 and ICEBs1 conjugative elements) [31, 33], their affiliation to a unique relaxase family was only pointed out later. This family was unclassified in 2009 [23], proposed in 2011 [7], but surprisingly, it was not mentioned recently [22, 39]. Taken together, our data commit us to propose the MOB_T family as a full non-canonical relaxase family alongside the two previously described non-canonical families (MOB_p and MOB_c) (Fig. 8).

In conclusion, we performed the first biochemical characterization of a member of the MOB_T family of relaxases, the RelSt3 protein. Its active site characterized here constitutes a potential target for the design of conjugation inhibitors. With this aim in mind, further work is needed, in particular to determine an experimental 3D structure of a MOB_T protein.

Methods
Phylogenetic analysis and identification of conserved motifs
BLAST searches were performed using the NCBI non-redundant (nr) database. The multiple alignment for the phylogenetic analysis was built with the MultAlin interface [65]. Only homologous positions (54 in total) were selected manually for phylogenetic analysis. A tree was constructed with PhyML 3.1 (downloaded from http://www.atgc-montpellier.fr/phyml/versions.php) [66]. The JTT model of amino acid substitution was chosen [67], and a gamma correction with four classes of sites was used. The alpha parameter and the proportion of invariable sites were estimated. The robustness of the tree was tested with PhyML by non-parametric bootstrap analysis (100 replicates).

To identify the conserved motifs in MOB_T relaxases, we first constructed a set of MOB_T sequences using NCBI’s CDART (Conserved Domain Architecture Retrieval Tool) application, with the RelSt3 sequence as a query [68]. All sequences harboring the RelSt3 architecture (N-terminal HTH domain and PF02486 domain) were aggregated together with the sequences harboring the NicK domain (PF02486 domain only, lacking the HTH domain). A total of 2016 sequences were then clustered using the CD-HIT suite [69], giving rise to 711, 383 and 261 clusters accordingly. One member of each cluster was selected for sequence alignment with Muscle [70] or ClustalW [71], and the resulting alignments were submitted to the WebLogo 3 interface [72]. The resulting files were very similar for all sequence identity cut-offs (used for CD-HIT clustering) and multiple alignment programs used.

Bacterial strains and plasmids
The bacterial strains and plasmids used in this study are listed in Additional file 4: Tables S1 and S2. Streptococcus thermophilus LMG18311 harboring ICESt3 labelled with a chloramphenicol resistance gene [51], or its derivative depleted of the orfJ gene, were used as donor strains during mating experiments. A strain that derives from S.
thermophilus LMG18311 and harbors a pMG36e plasmid carrying erythromycin resistance was used as recipient cell. E. coli EC101 was used as the host for cloning procedures with pG’host9 [73]. For heterologous protein production in E. coli, constructs were cloned into the DH5α strain, and over-expression was performed in BL21(DE3) cells.

Construction of ICESt3ΔorfJcat and of a mobilizable oriT-harboring plasmid

The deletion strategy used to construct the S. thermophilus ICESt3ΔorfJcat strain is derived from Biswas et al., 1993 [74], with several adaptations described in Additional file 4, Supplementary Materials and Methods. Three independent mutants were verified by sequencing and then used in conjugation assays.

The mobilizable oriT-harboring plasmid was constructed using the pOri23 backbone [75], where the Erm<sup>+</sup> cassette was replaced by a Spec<sup>+</sup> cassette, giving rise to the pOri1180 plasmid. In order to experimentally identify the oriT region of ICESt3, we constructed a plasmid encompassing the intergenic region between orfK and orfJ using EcoRI and ApaI restriction sites (according to primers described in Additional file 4, Table S3), giving rise to the pOri1180-oriT plasmid.

Mating experiments

Mating experiments were done as described in Dahmane et al, 2017 [76]. Briefly, donor and recipient strains were grown overnight with appropriate antibiotics. Cultures diluted to 1:100 were grown without any antibiotic until mid-exponential phase, and then mixed and concentrated 30 times in LM17 broth. For pOri1180 mobilization experiments, the donor strains were grown in the presence of spectinomycin (500 μg/mL) until mid-exponential phase and washed before the mating experiment. For each mating experiment, three 150 μL aliquots of cells were spread separately on 0.45 μm pore-size nitrocellulose filters (Millipore) deposited on soft agar (0.8%) LM17 plates and then incubated overnight at 42 °C. To select transconjugant cells, bacteria recovered from the filters were directly spread or concentrated 10 times before plating on chloramphenicol and erythromycin, or spectinomycin and erythromycin containing-LM17. After dilutions, the donor and the recipient cells were selected on LM17 plates containing the appropriate antibiotics. After a 24 h-incubation at 42 °C, CFUs counting of three independent biological replicates enabled us to quantify the mating frequency as the number of transconjugants per donor cell.

Cloning of orfJ gene for production in E. coli and mutagenesis

Mutagenesis was performed by overlap PCR with the oligonucleotides described in Additional file 4, Table S3.
were recorded with a scan speed of 60 nm min\(^{-1}\) and 1 nm spectral bandwidth. After an average of 3 scans and subtraction of buffer contribution, raw data were converted to mean residue ellipticity (deg.cm\(^2\).dmol\(^{-1}\) per residue) using Pro-Data Viewer software (Applied Photophysics, Ltd., UK), and was deconvolved using SELCON3, CONTINLL and CDSSTR algorithms available at the DichroWeb site [82, 83], with the reference data set #3. Data were also deconvolved with the CDNN program [84]. Thermal denaturation curves were followed at 195 and 222 nm from 20 to 80 °C (1 °C/min), and finally a 180–260 nm spectrum was recorded after cooling the sample back to 20 °C.

Differential scanning calorimetry (DSC) experiments were performed with a VP-DSC microcalorimeter (Microcal, Malvern Panalytical, Vénissieux, France). 500 μL of 6.6 μM RelSt3 was incubated with a heating rate of 60 °C/h in buffer A (20 mM Tris–HCl pH 7.5, 500 mM NaCl, 2 mM β-mercapto-ethanol). The experiments were done twice.

**Homology modeling**

Ten 3D models of the tertiary structure of the RelSt3 PF02486 domain (residues C61-D285) were built by homology with MODELLER [85], using as a template the X-ray structure of the RepSTK1 protein (PDB 4CJ), 30% sequence identity with the RelSt3 PF02486 domain. The model with the best Discrete Optimized Protein Energy (DOPE) score [86] was then refined by a molecular dynamics simulation of 100 ns in explicit solvent, and the most representative structure was selected by clustering the conformations selected from each ps of the simulation. Molecular dynamics simulation were performed with the NAMD software [87], using the charmm36 force field [88], and a box of explicit TIP3P water molecules. Counter-ions were added to neutralize the system.

**Supercoiled plasmidic DNA relaxation assays**

Relaxation assays were adapted from Lorenzo-Diaz et al, 2011 [54]. Plasmidic DNA (200 ng) of pBR322-oriT (Additional file 4, Table S2) was incubated 20 min at 30 °C with increasing amounts of RelSt3 protein (WT or variants, see figure legends), in R buffer [20 mM Tris–HCl pH 7.5, 200 mM NaCl, 5 mM MnCl\(_2\), 0.05 mM EDTA, 1% glycerol (w/v) and 1 mM β-mercapto-ethanol] with a final volume of 20 μL. Reactions were stopped by addition of final concentrations of 0.5% SDS and 100 μg/mL of proteinase K, followed by a second incubation at 30 °C for 20 min. Samples were electrophoresed on 1% agarose gel in Tris-Borate-EDTA-1X for 20 h at 2.5 V/cm and the gel was stained with ethidium bromide. Gel pictures were captured using a Bio-Rad XR+ system, and bands were quantified with Image Lab software (Bio-Rad Laboratories). Variations from these standard conditions are indicated in the figure legends.

**Additional files**

**Additional file 1:** Figure S1. Alignment of MOB\(_T\) relaxase representatives with Rel\(_{\text{trans}}\) RCR initiator representatives. This alignment with reduced number of sequences allows the identification of the different motifs cited in this study. See complete legend in Additional file 4. (RTF 1402 kb)

**Additional file 2:** Figure S2. Sequence alignment of MOB\(_T\) relaxase and Rel\(_{\text{trans}}\) RCR initiators. This larger alignment with more MOB\(_T\) and Rel\(_{\text{trans}}\) initiator sequences illustrates the conservation of the motifs through very distant proteins. See complete legend in Additional file 4. (RTF 40 kb)

**Additional file 3:** Figure S3. Calibration curve of the gel filtration column. A Sephadex G200 HiLoad 16/60 column (GE Healthcare) was used as a final step of purification of RelSt3 protein. This column was calibrated and allowed us to estimate the apparent molecular weight of RelSt3. See complete legend in Additional file 4. (PPTX 40 kb)

**Additional file 4:** Supplementary material text. This text includes supplementary Materials and Methods, complete Additional file legends, and tables for bacterial strains, plasmids and oligonucleotides used in this study. (DOC 20 kb)

**Abbreviations**

CD: Circular dichroism; DSC: Differential scanning calorimetry; HGT: Horizontal gene transfer; ICE: Integrative and conjugative elements; oriT: origin-of-transfer; RCR: Rolling-circle replication; T4SS: Type-IV secretion system

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**Availability of data and materials**

The datasets used during the current study are available from the corresponding author on reasonable request. The accession number of all protein sequences used for sequence alignments or phylogenetic analysis are listed in the Additional file 4, Table S4.

**Authors’ contributions**

NS, SP, MDD and NLB designed experiments. ER, ICB, PM, VL, BM and JS performed experiments. NS, ICB, VL, BM, GG, SP, MDD and NLB analyzed the data. NS wrote the original draft of the manuscript. ICB, VL, DWR, GG, SP, MDD and NLB reviewed the manuscript. This work was supervised by NS, DWR, MDD and NLB. All the authors approved the final manuscript.

**Ethics approval and consent to participate**

Not applicable.

**Consent for publication**

Not applicable.
Competing interests
The authors declare that they have no competing interests.

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