Expression of CD34 and Myf5 Defines the Majority of Quiescent Adult Skeletal Muscle Satellite Cells

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Abstract. Skeletal muscle is one of a several adult post-mitotic tissues that retain the capacity to regenerate. This relies on a population of quiescent precursors, termed satellite cells. Here we describe two novel markers of quiescent satellite cells: CD34, an established marker of hematopoietic stem cells, and Myf5, the earliest marker of myogenic commitment. CD34⁺ve myoblasts can be detected in proliferating C2C12 cultures. In differentiating cultures, CD34⁺ve cells do not fuse into myotubes, nor express MyoD. Using isolated myofibers as a model of synchronous precursor cell activation, we show that quiescent satellite cells express CD34. An early feature of their activation is alternate splicing followed by complete transcriptional shutdown of CD34. This data implicates CD34 in the maintenance of satellite cell quiescence.  

In heterozygous Myf5

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Although not normally subject to rapid cell turnover, adult skeletal muscle also retains the ability to grow in response to increased work load and to repair and regenerate following damage. The mechanical functions of skeletal muscle are carried out by syncytial myofibers, each containing a highly specialized contractile apparatus maintained by large numbers of postmitotic myonuclei. The capacity to generate new myonuclei resides in a population of mononucleated precursors, termed satellite cells, which lie sequestered between the basal lamina and sarcolemma of each myofiber (reviewed in Bischoff, 1994). In immature muscle, many satellite cells are cycling and differentiating after a limited number of divisions to contribute myonuclei to growing myofibers (Moss and Leblond, 1971; Schultz, 1996). In mature muscle, satellite cells are mitotically quiescent (Schultz et al., 1978), but can be rapidly activated to provide myonuclei for nascent or pre-existing myofibers (Grounds and McGechie, 1987; Rantanen et al., 1995). Temporal studies of satellite cell proliferation suggest that those lost to differentiation are replaced, possibly by asymmetric division, from a distinct subpopulation.
defined in growing muscle by an extended cycle time (Schultz, 1996). The presence of populations of satellite cells with differing rates of division and proliferative capacities has been confirmed by in vitro clonal analyses (Schultz and Lipton, 1982; Molnar et al., 1996). Adult skeletal muscle also appears to contain rare multistep progenitors that can give rise to myonuclei and all hematopoietic lineages, but the identity and location of these cells have yet to be defined (Gussoni et al., 1999; Jackson et al., 1999). Although any direct lineage relationships between these putative subpopulations have yet to be established, the above observations suggest that the regenerative compartment of adult skeletal muscle may also conform to the hierarchical archetype of other self-renewing adult tissues.

Satellite cell subpopulations have thus far been defined only by behavioral criteria and there are no reports of differential expression of quiescent satellite cell protein markers such as M-cadherin (Irintchev et al., 1994), c-met (Cornelison and Wold, 1997), and myocyte nuclear factor (Garry et al., 1997). Here we describe two novel markers, CD34 and Myf5, that are expressed on most but, significantly, not all quiescent satellite cells.

The first of these markers, CD34, is an accepted, clinically exploited marker of adult hematopoietic stem cells (HSCs) and early blood-cell progenitors and has become the standard criterion for the isolation of such cells from both blood and bone marrow (Krause et al., 1996). Despite a pre-eminent status in transplantation biology, CD34 remains remarkably enigmatic. Structurally, CD34 is a highly O-glycosylated, transmembrane sialomucin, expressed by HSC and progenitors (Krause et al., 1996) and by small-vessel endothelium (Baumhueter et al., 1994). Two isoforms of CD34 are translated from alternatively spliced transcripts: the full-length protein (CD34full) has an intracellular domain with three potential phosphorylation sites that are lacking in the short cytoplasmic tail of truncated CD34 (CD34trunc) (Suda et al., 1992; Nakamura et al., 1993). The significance of the two isoforms is unclear since extracellular engagement of either promotes homotypic cytoadhesion via the same tyrosine kinase–mediated pathway (Tada et al., 1999). The function of CD34 is also obscure, although its expression on HSC has been implicated in the regulation of differentiation (Fackler et al., 1993). The significance of the two isoforms is unclear since extracellular engagement of either promotes homotypic cytoadhesion via the same tyrosine kinase–mediated pathway (Tada et al., 1999). The function of CD34 is also obscure, although its expression on HSC has been implicated in the regulation of differentiation (Fackler et al., 1993).

The second marker, Myf5, is one of a family of muscle-specific basic helix-loop-helix transcription factors. Myf5 is the earliest marker of myogenic commitment and, together with MyoD, is integral to the determination of skeletal muscle (reviewed in Tajbakhsh and Buckingham, 2000). Using the Myf5nlacZ mouse, which has a reporter gene encoding nuclear-localizing β-galactosidase (β-Gal) (nlacZ), targeted to the Myf5 locus (Tajbakhsh et al., 1996a), we show that the Myf5 locus is active in all CD34−ve quiescent satellite cells and that all Myf5−ve precursors express CD34.

We then determined the total number of satellite cells associated with individual isolated muscle fibers using the 3F-nlacZ:2E transgenic mouse carrying regulatory elements from the locus of the fast myosin light chain 1F/3F gene that drives a nlacZ reporter gene (Kelly et al., 1995). This transgene is expressed by all myonuclei in fast myofibers, but not by the associated satellite cells. We have found that the total number of satellite cells identified by a lack of 3F-nlacZ:2E transgene expression is significantly higher than the number of CD34−ve or Myf5−ve satellite cells. We therefore conclude that the satellite cell compartment consists of two populations: a majority expressing both CD34 and Myf5 and an as yet undefined minority that is negative for both markers.

The expression of CD34 on quiescent adult skeletal muscle satellite cells extends the role of CD34 in progenitor cell biology. Recently, the status of CD34 as a marker of stem cells has been called into question by the identification of CD34−/low HSCs (reviewed in Goodell, 1999) and the association of CD34 expression with activation and progress towards either self-renewal or differentiation (Sato et al., 1999). Coexpression with Myf5 and M-cadherin, both of which are restricted to the myogenic lineage, suggests that in adult skeletal muscle, CD34 does not mark stem cells but is expressed by precursors that are committed to a specific fate and have become arrested and held in reserve for subsequent activation. It seems likely therefore that CD34 plays a fundamental role in the regulation of lineage-primed progenitor compartments in a range of adult tissues, including blood and skeletal muscle.

**Materials and Methods**

**Primers and Probes**

CD34 primers were designed using the sequence of Brown et al. (1991). Amplification of both transcripts was carried out using a forward primer in exon 4 (5′-CCAGGTATCTGGCTGGAAC-3′) and a reverse primer in exon 5 (5′-GCTGAGGTTCTTGGGAGT-3′). Primers used to distinguish transcripts for CD34full and CD34trunc were 5′-AGCCAACAAAACTCCACAC-3′ in exons 5/6 and 5′-AGCCAACAAAACTCCACAC-3′ in exons 7/8, respectively. An M-cadherin cDNA fragment corresponding to 502 bp of exons 2–5 (Link et al., 1998) was produced using 5′-CTAACGTTCTGTGACAC-3′ and 5′-CTAACGTTCTGTGACAC-3′ for first-round amplification, before a second round of nested PCR using the primers designed to distinguish the two splice variants. External forward and reverse primers for MyoD cDNA were 5′-CGCTTCAAACTGTCATGGG-3′ and 5′-AAGAACGGGGCCACATCCC-3′, respectively, and the internal primers for nested amplification were 5′-CGGGCCAAGTGGCTAGCA-3′ and 5′-CGGGCCAAGTGGCTAGCA-3′; the nested primers lie in different exons separated by a 327-bp intron (Zingg et al., 1991). S16 primers were designed as described by Foley et al. (1993).

A CD34 cDNA fragment corresponding to 442 bp of exons 4–7 was generated by PCR amplification of cDNA reverse transcribed from total RNA extracted from embryonic day (E) 17.5 mouse embryos, using 5′-CAAGATTTCTGGCAATCGGGAAGTGAGT-3′ and 5′-ACTCTAAGAAGCCAGCTTTTCTCCTGTA-3′ as forward and reverse primers, respectively. Using cDNA from E14.5 muscle as a template, we used 5′-GACAAGTGGCAGTGGCTGGAAGT-3′ and 5′-GACAAGTGGCAGTGGCTGGAAGT-3′ as forward and reverse primers, respectively. A Mcadherin cDNA fragment corresponding to 502 bp of exons 2–5 (Link et al., 1998) was used to clone the sequence into pBluescript SK− (Stratagene).
Reverse Transcription-PCR and Northern Blotting

Total RNA was prepared according to the method of Chomczynski and Sacchi (1981). For reverse transcription (RT)-PCR analysis of cell lines, 2 μg of RNA were reverse transcribed in a 20-μl reaction, using Superscript RNase H⁻ reverse transcriptase (Life Technologies) primed with oligo-dT₃₁₋₁₅. 2-μl aliquots of cDNA were used as template for 24 cycles of PCR using Red Hot DNA polymerase (Advanced Biotechnologies) in the presence of 1 μCi of [γ³²P]dCTP (3,000 Ci/mmol; Amersham Pharmacia Biotech). PCR products were separated on polyacrylamide gels and analyzed using a Phosphor-Imager 445 SI equipped with ImageQuant software (Molecular Dynamics). For RT-PCR analysis of isolated fibers, individual muscle fibers were lysed in 10 mM Tris-HCl, pH 7.4, 0.15 M NaCl, 1 mM MgCl₂, 0.5% NP-40, and the entire lysate was reverse transcribed as above. 2-μl aliquots of cDNA were used as template for 35 cycles of PCR amplification, and 2 μl of product were then amplified through a further 35 cycles, using fully nested primers. For Northern blotting, total RNA was denatured at 50°C in 6.5% glyoxal, fractionated on a 1.5% agarose gel in 10 mM sodium phosphate, pH 7.0, and then transferred onto Hybond-N membrane (Amersham Pharmacia Biotech) and hybridized according to manufacturer’s instructions.

Myf5/nlacZ²⁺/⁺ Mice

The Myf5/nlacZ²⁺/⁺ mouse has nlacZ-SV40poly(A) RNApolyIII/Neo targeted to the first exon of the Myf5 gene such that β-Gal is produced as a fusion protein with the first 13 amino acids of Myf5. The Myf5 gene is also disrupted and a small deletion is introduced. Homozygous Myf5/nlacZ²⁺/⁺ animals die shortly after birth due to respiratory problems caused by abnormal rib development, whereas the heterozygous Myf5/nlacZ²⁺/⁻ mice used in the studies described here are viable (Tajbakhsh et al., 1996a).

3F-nlacZ-2E-transgenic Mice

The 3F-nlacZ-2E transgenic mouse contains seven copies of a construct consisting of 2 kb upstream of the myosin light chain (MLC)–3F transcriptional start site, nlacZ-SV40 poly(A) in frame in the second MLC3F-specific exon, 1 kb of MLC3F sequence 5' of nlacZ, and a 260-bp 3' MLC1F/3F enhancer (Kelly et al., 1995).

In Situ Hybridization

Hybridizations were carried out using (CBA × C57Bl/10) F₁ embryos. Noon of the day of the vaginal plug was designated E 0.5. Digoxygenin-labeled riboprobes were generated and hybridized to headless, eviscerated embryos as described (Zammit et al., 2000). The CD34 riboprobe was synthesized from the 502-bp cDNA fragment of exons 2–5 of cd34, and a 260-bp 3' MLC1F/3F enhancer (Kelly et al., 1995).

Whole Muscle Preparation

Mice were killed by cervical dislocation, muscles were removed complete with tendons, rinsed in PBS, and then fixed for 5 min in 20% normal goat serum for at least 30 min to block nonspecific antibody binding. Mouse anti-human MyoD1 (clone 5.8a; Dako) monoclonal antibody and rabbit anti-mouse Myf5 polyclonal antibody (Yablokna-Reuveni et al., 1999) were applied for 90 min at room temperature in 2% mouse serum in PBS. Anti-CD34 antibody was detected using biotin-conjugated goat anti-rat IgG polyclonal antibody (Sigma-Aldrich) followed by streptavidin conjugated to Alexa Fluor 594. Mouse monoclonal and rabbit polyclonal primary antibodies were detected using Alkfluor 488–conjugated goat anti-mouse or rabbit IgG polyclonal antibodies (fluorescent-labeled antibodies were from Molecular Probes). Isolated muscle fibers were stained with the anti-CD34 and –MyoD1 antibodies, rabbit anti-mouse M-cadherin polyclonal antibody (Irintchev et al., 1994), and mouse anti–Escherichia coli β-Gal monoclonal antibody (clone GALT13; Sigma-Aldrich). Fibers were fixed and treated as above except that permeabilization was carried out using 0.5% Triton X-100 and all primary antibodies were applied in 0.35% type IV lambda carrageenan (Sigma-Aldrich) in PBS for 16–40 h at 4°C. Primary antibodies were detected with appropriate species-specific Alexa Fluor–conjugated secondary antibodies, diluted in 0.35% carrageenan and applied for 2 h at room temperature. The same biotin-conjugated intermediate was used in conjunction with the anti-CD34 antibody.

Whole Muscle Preparation

Mice were killed by cervical dislocation, muscles were removed complete with tendons, rinsed in PBS, and then fixed for 5 min in freshly prepared 4% paraffinomdehyde in PBS. For cryostat sectioning, unfixed muscles were mounted in OCT (Raymond Lamb) compound and frozen in liquid nitrogen. Muscles were freeze or frozen within 10 min of the animal being killed.

Cell Culture and Single Fiber Preparation

Primary muscle cells were obtained by enzymatic disaggregation of leg muscle from 1-d-old C57Bl/10 mice and cultured as described previously (Beauchamp et al., 1999). The ICR/1An myogenic line was cloned from a primary culture prepared by enzymatic disaggregation of a crush-injured tibialis anterior (TA) muscle of a 6-wk-old ICR/1An mouse. Phosphorylase-deficient mice and was grown as a primary culture. Cultivation and maintenance of 128, C2C12, and H-2Kₘ-tsα58 cell lines have been described previously (Blau et al., 1983; Morgan et al., 1994; Irintchev et al., 1997). To induce myogenic differentiation, cultures were allowed to reach 70% confluence before transfer into differentiation medium consisting of DME, 4 mM l-glutamine, 100 U/ml penicillin, and 100 μg/ml streptomycin, supplemented with 10% (primary, ICR/1An cells and H-2Kₘ-tsα58 clones) or 2% (C2C12 and 128 cells) horse serum. All cultures were maintained on plastic pre-coated with 0.01% gelatin. s.End1 endothelial cells, a polymya virus-transformed line derived from a subcutaneous hemangioma in a 3-wk-old ICR mouse (Williams et al., 1988), were cultured in DME containing 4 mM l-glutamine, 100 U/ml penicillin, and 100 μg/ml streptomycin, supplemented with 10% fetal calf serum.

Results

Transcripts for both CD34<sup>all</sup> and CD34<sup>ran</sup> Are Expressed by Skeletal Muscle Precursor Cells In Vitro

To investigate the expression of CD34 in skeletal muscle precursor cells, total RNA from primary myogenic cultures and established myogenic cell lines was screened by RT-PCR. CD34 mRNA was present in all of the myogenic cultures examined, both in undifferentiated myoblasts and after differentiation into myotubes, although levels of expression varied considerably (Fig. 1 A). The highest levels were found in primary myogenic cultures prepared by enzymatic disaggregation of limb muscles dissected from newborn mice. Although primary muscle precursor cells are closest to their in vivo counterparts, there is an unde-
Expression of CD34 Is Associated with Skeletal Muscle Precursors that Do Not Differentiate In Vitro

Expression of CD34 protein in skeletal muscle cultures was investigated by immunofluorescent staining using an antibody raised against an extracellular epitope that recognizes both the full-length and truncated isoforms of CD34. In sEND.1 cultures, virtually all cells showed intense cell-surface expression (Fig. 2 A). In contrast, C2C12 myoblasts maintained in proliferation medium showed highly variable levels of expression, with most cells either negative or very faintly positive and only a small percentage (estimated as <5%) of strongly CD34+ve cells (Fig. 2 B). Heterogeneous expression of the myogenic regulatory factors Myf5 (Fig. 2 C) and MyoD (D) was also observed in proliferating C2C12 cultures, as reported previously in asynchronous cultures (Tapscott et al., 1988; Lindon et al., 1998). Although there was no apparent correlation between expression of CD34 and Myf5 (i.e., some CD34+ve myoblasts expressed Myf5, whereas others were Myf5−ve; Fig. 2 C), strongly CD34+ve myoblasts were invariably MyoD−ve (D).
In C2C12 cultures that had been allowed to differentiate for 5 d, strong expression of CD34 was maintained on a small number of mononucleated cells, usually closely associated with multinucleated myotubes (Fig. 2, E and F). In contrast to the variable levels of Myf5 expression observed among the CD34\(^{-\text{ve}}\) population in proliferating cultures, those that persisted after 5 d in differentiation medium were consistently Myf5\(^{+\text{ve}}\) (Fig. 2 E). Furthermore, the CD34\(^{-\text{ve}}\) cells did not express MyoD (Fig. 2 F), even though the majority of the culture had withdrawn from the cell cycle and entered terminal differentiation as defined by expression of p21 and myogenin (data not shown). Expression of CD34 therefore defines a subset of precursors that do not differentiate in culture, in contrast to the CD34\(^{+\text{ve}}\) majority.

**CD34 mRNA Is Absent during Primary Myogenesis, Expressed during Secondary Myogenesis, and Maintained after Birth**

In the mouse embryo, myogenesis occurs in distinct waves: an initial phase of primary muscle fiber formation is followed \(~2\) d later by the generation of secondary muscle fibers (Ontell and Kozeka, 1984). Distinct populations of muscle precursor cells are involved in each event: embryonic myoblasts fuse during primary myogenesis, whereas fetal or secondary myoblasts differentiate during secondary myogenesis. A further population of precursors, the satellite cells, persists in the postnatal animal and is involved in growth and regeneration (reviewed in Miller et al., 1999). Whole mount in situ hybridization of mouse embryos and dissected 3-d postnatal tissue was used to determine when CD34 transcript is expressed in skeletal muscle with respect to primary and secondary myogenesis. At E 11.5, staining for CD34 transcript was observed in the outflow tract of the heart and as a diffuse network throughout the embryo presumably reflecting expression in the developing circulatory system (Young et al., 1995) (Fig. 3 A). Although intersomitic blood vessels were clearly positive, no CD34 transcript was detected in the somites or in the limb buds. In marked contrast, transcript for M-cadherin, which marks all myogenic cells at this stage (Rose et al., 1994), was restricted to the myotome of the somites and the proximal region of the limb buds (Fig. 3 B) at E 11.5. The mutually exclusive patterns of expression at E 11.5 shows that CD34 is not present in skeletal muscle during primary myogenesis. However, at E 16.5, hybridization with either CD34 (Fig. 3 C) or M-cadherin (D) riboprobes produced strikingly similar lines of punctate staining, apparently in register with the underlying fibers. Previous developmental studies have shown that M-cadherin protein is uniformly expressed in the myotome at E 11.5, but becomes clustered at points of contact between primary and secondary fibers during secondary myogenesis between E 16 and 18 (Rose et al., 1994). In postnatal muscle,
M-cadherin protein is restricted to satellite cells and the adjacent region of the underlying fiber (Irintchev et al., 1994). Since both transcriptional activation and downregulation of M-cadherin occur before changes in protein (Moore and Walsh, 1993; Rose et al., 1994), it is likely that the discrete hybridization pattern obtained for M-cadherin (and by inference, that obtained for CD34) at E 16.5 reflects the distribution of satellite cells that continue to express M-cadherin protein after birth. Identical results were obtained with the CD34 probe on muscles from 3-d postnatal mice as on E 16.5 embryonic tissues (Fig. 3 E) and, when gently teased apart, at least some of the staining was clearly localized to putative satellite cells, closely applied to individual muscle fibers (F).

**CD34 Is Expressed on Adult Skeletal Muscle Satellite Cells Associated with Isolated Single Fibers**

The expression of CD34 protein on adult skeletal muscle satellite cells was confirmed using isolated single muscle fibers prepared by collagenase digestion and mechanical disruption of EDL muscles of 5–8-wk-old mice (Rosenblatt et al., 1995). This procedure produces viable single fibers together with their associated satellite cells, surrounded by an intact basal lamina but free of blood vessels, nerves, and connective tissue. Immunostaining of freshly isolated fibers that had been fixed within 2 h of the death of the animal revealed the presence of CD34+ve mononucleated cells attached to the sarcolemma of the muscle fibers. The same cells also expressed M-cadherin, a marker of quiescent myogenic precursors in adult skeletal muscle (Irintchev et al., 1994), thereby confirming their identity as satellite cells (Fig. 4, A–C).

Some of the CD34+ve cells associated with freshly isolated fibers were also found to express low levels of MyoD (Fig. 5, A–C). In preparations that had been maintained for 48 h in conditions that promote satellite cell activation and proliferation, all satellite cells showed strong coexpression of CD34 and MyoD, although the CD34 staining appeared more punctate compared with the uniform surface staining observed on freshly isolated cells (Fig. 5, D–F). CD34 was not detected in preparations maintained for longer than 48 h, when many satellite cells had migrated from the parent fiber and begun to proliferate on the surrounding dish (data not shown).

The expression of CD34 transcripts in adult satellite cells was investigated by fully nested PCR of cDNA prepared from individual muscle fibers. At the earliest time point after isolation, transcript for CD34trunc, but not CD34full, was detected in all preparations (Fig. 5 G). 3-h later, both isoforms were present, with different preparations containing transcript for either CD34full or CD34trunc, or both. After a further 3 h, all preparations contained transcript for CD34full, although a proportion continued to express low levels of transcript for the CD34trunc and by 24 h, no CD34 mRNA was detected in the isolated fiber preparations (data not shown). The same cDNA samples were also screened for the presence of MyoD transcript. Although the protein was clearly present in a small percentage of satellite cells associated with freshly isolated fibers albeit at low levels (Fig. 5 B), few samples were positive for MyoD mRNA either at the earliest time point or after 3 h in culture (Fig. 5 G). This suggests that either MyoD protein is highly stable in quiescent satellite cells or that the amount of transcript was below the level of detection. However, after 6 h, MyoD transcript was detected in almost all of the samples analyzed, attributable to satellite cell activation resulting in either the initiation or upregulation of MyoD transcription. Although the precise timings of the switch in CD34 transcripts and their subsequent disappearance varied between experiments, the transition from CD34trunc to CD34full always accompanied the initiation of satellite cell activation before transcriptional upregulation of satellite cell activation.
Myf5 Is Expressed on Adult Skeletal Muscle Satellite Cells Associated with Isolated Single Fibers

Myf5nlacZ/+ heterozygous mice were used to determine whether the CD34⁺ve satellite cells associated with freshly isolated fibers also express Myf5. The Myf5nlacZ/+ mouse has nlacZ targeted to the Myf5 locus such that activation of Myf5, and therefore expression of endogenous Myf5 from the untargeted allele, is reported by β-Gal activity (Tajbakhsh et al., 1996a). When freshly isolated fibers from Myf5nlacZ/+ mice were incubated in X-Gal solution, rare β-Gal⁺ve nuclei were observed, sometimes closely apposed to β-Gal⁻ve nuclei (Fig. 6, A and B). Since β-Gal can translocate from nuclei carrying nlacZ to other non-transgenic nuclei within a muscle fiber (Yang et al., 1997), the presence of adjacent β-Gal⁺ve nuclei suggests that each β-Gal⁺ve nucleus was contained within its own cytoplasm, isolated from the underlying syncytial fiber. This was confirmed by immunostaining of fibers isolated from heterozygous Myf5nlacZ/+ mice, which demonstrated that the β-Gal⁺ve nuclei were in satellite cells, as defined by surface expression of M-cadherin (Fig. 6, C and D) and CD34 (E and F). Furthermore, analysis of fibers prepared from mice of more than 6 mo of age, demonstrated that β-Gal is produced by satellite cells in mature muscle and is not a relic from earlier developmental stages (data not shown). Previous attempts to detect Myf5 protein in isolated fiber preparations were unsuccessful due to the high levels of nonspecific binding encountered with the available antibodies (Yablonka-Reuveni et al., 1999). However, fully nested RT-PCR analysis revealed the presence of transcript for Myf5 in all single-fiber preparations from C57Bl/10 mice analyzed immediately after isolation, or after 24 h in culture (data not shown), confirming the fidelity of the Myf5nlacZ/+ as a reporter of endogenous Myf5 transcription. These observations show that Myf5 is expressed by quiescent satellite cells associated with fibers obtained from normal adult skeletal muscle.

Quiescent Satellite Cells Express both CD34 and Myf5 In Vivo

Whole muscle preparations and cryosections were analyzed to confirm that CD34 and Myf5 are present on quiescent satellite cells in vivo and that expression on satellite cells associated with single muscle fibers was not due to activation during isolation. When intact muscles from 6-wk-old Myf5nlacZ/+ mice were incubated in X-Gal solution immediately after dissection, either with or without prior fixation, all showed a punctate pattern of β-Gal reaction product associated with myofibers (Fig. 7 A). That this pattern reflecting the distribution of satellite cells was confirmed when cryosections of TA muscle were immunostained or incubated in X-Gal (data not shown), both of which showed that the β-Gal activity was restricted to a few nuclei associated with the edges of muscle fibers (Fig. 7 B). Furthermore, when the same sections were immunostained with anti–CD34 antibody, the β-Gal⁺ve satellite cells were also clearly CD34⁺ve (Fig. 7, C–G).
derived from the CD34trunc and CD34 full alternately spliced transcripts are arrowed T (416 bp) and F (250 bp), respectively.

Figure 5. Expression of CD34 and MyoD on satellite cells associated with isolated single muscle fibers. (A–C) A freshly isolated EDL single fiber immunostained for CD34 (A, red), MyoD (B, green), and counterstained with DAPI (C). (Arrow) A single CD34\textsuperscript{+ve} satellite cell closely associated with an underlying myonucleus. (D–F) A single EDL fiber immunostained for CD34 (D, red), MyoD (E, green), and counterstained with DAPI (F) after 48 h in culture. Two CD34\textsuperscript{+ve}, MyoD\textsuperscript{+ve} satellite cells (arrows) are shown migrating off the parent fiber. The CD34 staining was punctate, in contrast to the continuous surface staining observed on quiescent cells. Bar, 30 \textmu m. (G) RT-nested PCR analysis of CD34 isoform expression in isolated single fibers during activation in vitro. Each pair of gels shows the PCR products obtained from 12 individual fibers using primers for CD34 (left) and MyoD (right). Groups of fibers were taken immediately after isolation (0 h) and after 3 or 6 h of culture in the presence of horse serum. PCR products derived from the CD34\textsuperscript{trunc} and CD34\textsuperscript{full} alternately spliced transcripts are arrowed T (416 bp) and F (250 bp), respectively.

β-Gal, confirming that the nuclei identified as DAPI fluorescent after incubation in X-Gal included those of the M-cadherin\textsuperscript{+ve}, CD34\textsuperscript{+ve} satellite-cell population.

Using the above approach, EDL fibers from 5- to 8-wk-old 3F-nlacZ-2E mice were found to contain an average of 5.6 \pm 0.3 satellite cells (\pm SEM, \(n = 125\) fibers from six mice) in which DAPI fluorescence remained unquenched. However, using fibers isolated from the same six muscles, significantly fewer satellite cells were identified by expression of CD34 (4.5 \pm 0.2 (\pm SEM, \(n = 114\) fibers)) or M-cadherin (4.4 \pm 0.2 (\pm SEM, \(n = 124\) fibers)) (Fig. 9). A value of 4.4 \pm 0.3 (\pm SEM, \(n = 122\) fibers from six animals) was also obtained by counting the number of β-Gal\textsuperscript{+ve} satellite cells associated with EDL fibers prepared from age-matched Myf5\textsuperscript{nlacZ/+} heterozygous mice. Therefore, there were no significant differences between satellite cell numbers obtained by counting CD34\textsuperscript{+ve}, M-cadherin\textsuperscript{+ve}, or Myf5\textsuperscript{+ve} cells, but all were significantly lower than the total number of satellite cells associated with the fibers. Furthermore, when fibers were immunostained for any two of the three markers (such as in Figs. 4, A–C, and 6, C–F), all satellite cells were found to express both markers. Together, this strongly suggests that the satellite cell population comprises a majority that is CD34\textsuperscript{+ve}, Myf5\textsuperscript{+ve}, and M-cadherin\textsuperscript{+ve} and an as yet undefined minority that is negative for all three markers.

Discussion

The renewal of several terminally differentiated adult tissues is sustained by populations of stem cells that both self-renew and generate a hierarchy of progressively lineage-restricted progenitors culminating in lineage-committed precursors fated to undergo terminal differentiation (Watt and Hogan, 2000; Weissman, 2000). In tissues with a high rate of turnover, such as blood, skin, and gut, the demands of replacement require a constant supply of precursors for terminal differentiation, such that progression from progenitor to functional, post-mitotic cell appears continuous. In adult skeletal muscle, however, new myonuclei are only required for growth and repair. Accordingly, the satellite cell compartment is normally quiescent and activated only in response to signals elicited by increased work load or damage (Seale and Rudnicki, 2000). Here we report two novel markers of the majority of quiescent satellite cells: CD34, a marker of stem cells and early progenitors in the hematopoietic system (Krause et al., 1996), and Myf5, the earliest marker of myogenic commitment (Tajbakhsh and Buckingham, 2000). This combination of markers suggests that most satellite cells become quiescent after committing to the skeletal muscle lineage and raises the possibility that CD34 may play a fundamental role in regulating progenitor cell differentiation.

Expression of CD34 transcript in several nonhematopoietic adult tissues has been attributed to the presence of small-vessel endothelium (Krause et al., 1996). Indeed, although CD34 transcript is present in extracts of whole skeletal muscle (Nakamura et al., 1993), CD34 protein has only been reported on capillaries, muscle spindle capsule, and axons (Baumhueter et al., 1994). Our results show that CD34 is also expressed by satellite cells in normal adult skeletal muscle, the identity of which was suggested by their morphology and distribution and confirmed by the
coexpression of M-cadherin (Irintchev et al., 1994). Furthermore, using heterozygous Myf5lacZ/+ mice, we were also able to show that CD34+/−ve satellite cells are clearly committed to the myogenic lineage by the criterion of transcriptional activation of Myf5. Developmental studies of Myf5lacZ/+ heterozygous mice have confirmed that the distribution of β-Gal faithfully reports endogenous expression of Myf5 transcript from the wild-type allele (Tajbakhsh et al., 1996a). Although Myf5 is active in nuclei within muscle fibers at birth (Tajbakhsh et al., 1996a), our results show that the gene is switched off during early postnatal development such that Myf5 is not expressed by myonuclei at 6 wk of age, but remains active in satellite cells. This is in contrast to a previous study that concluded that Myf5 is not expressed by quiescent satellite cells in sections of undamaged muscle (Cooper et al., 1999). However, we observed clear expression of Myf5 by satellite cells both in whole muscles and in sections of normal muscle that had been fixed or frozen only minutes after dissection. This discrepancy was probably due to differences in the sensitivity of the experimental procedures for β-Gal detection used in our studies compared with those of Cooper et al. (1999). We were also able to detect CD34 protein on satellite cells in sections of normal muscle and on freshly prepared single fibers, even when isolated in the presence of 100 μM cycloheximide (data not shown). These observations confirm that both CD34 and Myf5 are expressed by quiescent satellite cells and that their presence is not the result of de novo synthesis in response to experimentally induced activation.

A small percentage of CD34+/−ve, Myf5+/−ve satellite cells associated with freshly isolated fibers were also found to express MyoD protein, although apparently at low levels. In contrast, after 48 h of culture in the presence of serum, the vast majority of CD34+/−ve, Myf5+/−ve satellite cells were strongly MyoD+/−ve. This suggests that most adult skeletal muscle satellite cells are quiescent and express both CD34 and Myf5, but little or no MyoD, and become CD34+/−ve, Myf5+/−ve, MyoD+/−ve on activation. Although normal adult mouse muscle contains very few, if any, dividing satellite cells (Schultz et al., 1978; Irintchev et al., 1994), sporadic expression of MyoD has been reported and attributed to satellite cell activation, presumably in response to local stimuli (Grounds et al., 1992; Creuzet et al., 1998). It is therefore likely that expression of MyoD by a proportion of satellite cells associated with freshly isolated fibers defines those that had been activated in vivo before isolation. Our method of obtaining quiescent satellite cells associated with isolated single muscle fibers affords a unique in vitro system to study synchronous activation. Using fully nested RT-PCR, we detected a rapid increase in MyoD transcription that preceded the increase in MyoD protein expression observed by immunostaining, as described previously during satellite cell activation both in vivo (Cooper et al., 1999) and in vitro (Kitzmann et al., 1998). Although most quiescent satellite cells contained undetectable amounts of MyoD mRNA, a small percentage (5–10%) of satellite cells were strongly MyoD+/−ve. This discrepancy was probably due to differences in the sensitivity of the experimental procedures for β-Gal detection used in our studies compared with those of Cooper et al. (1999).
of MyoD mRNA, they did contain transcripts for Myf5 (data not shown) and CD34. Quiescent satellite cells expressed transcript for the truncated, but not the full-length isoform of CD34. Intriguingly, within hours of activation, the alternatively spliced transcript for CD34\textsuperscript{full} had become the predominant isoform. After 24 h in culture, neither CD34 transcript was present, although the protein could still be detected up to 48 h, presumably due to its long half life (Krause et al., 1996).

CD34 is also expressed by skeletal muscle precursors in culture. Using the C2C12 cell line, we found that at any given time <5\% of proliferating myoblasts expressed CD34 protein and, although the CD34\textsuperscript{+ve} cells showed variable levels of expression of Myf5, they were consistently MyoD\textsuperscript{-ve}. The variable expression of Myf5 and MyoD observed within our proliferating cultures is probably the result of asynchronicity, as both are cell-cycle regulated (Kitzmann et al., 1998). Using synchronous cultures, Kitzmann et al. (1998) showed that cells arrested in G0 are MyoD\textsuperscript{-ve}, Myf5\textsuperscript{+ve}, and that release into G1 is accompanied by a loss of Myf5 expression before upregulation of MyoD; it is therefore possible that expression of CD34 is associated with cells at the G0/G1 boundary. We also observed that, when induced to differentiate, the CD34\textsuperscript{+ve} cells remained morphologically undifferentiated, became uniformly Myf5\textsuperscript{-ve} and, in contrast to the CD34\textsuperscript{-ve} majority, showed no induction of MyoD or myogenin. These cells probably correspond to the slowly dividing, MyoD\textsuperscript{-ve}, Myf5\textsuperscript{-ve} ‘reserve’ cells described by Yoshida et al. (1998) that on return to appropriate conditions are able to resume proliferation and give rise to both MyoD\textsuperscript{+ve} differentiation-competent cells and more reserve cells. Behaviorally similar cells have been identified in primary cultures of human satellite cells (Baroffio et al., 1996). These ‘reserve’ or stem-like cells were described in cultures polarized by differentiation and so it is unclear whether they are a permanent subpopulation or a transitory phenotype in proliferating myogenic cultures. However, their presence in clonally derived cultures suggests plasticity of myoblast behavior, and that some of those that are CD34\textsuperscript{+ve}, MyoD\textsuperscript{-ve} (and probably at the G0/G1 boundary at the time of induction) become quiescent in response to differentiation cues.

The temporal appearance of CD34 during muscle development also suggests a potential role in establishing and/or maintaining the satellite cell compartment. We found that CD34 is not expressed in the somites during primary myogenesis, demonstrating that the earliest precursors committed to myogenesis are CD34\textsuperscript{-ve}. However, transcript was detected at E 16.5, coincident with the appearance of satellite cells (Cossu et al., 1983). Although satellite cells are generally assumed to be of somitic origin, recent evidence suggests that some may ultimately be derived from primordial endothelial cells that enter skeletal muscle either via the circulation or as perithelial cells associated with developing vessels, before becoming committed to the myogenic lineage through environmental influences (Bianco and Cossu, 1999; De Angelis et al., 1999).
Intriguingly, this implies a common ancestor with hematopoietic and endothelial cells, lineages with which CD34 is primarily associated. Whether satellite cells are a population sequestered within the somites or derived from elsewhere, their appearance coincides with the onset of CD34 expression in skeletal muscle.

Recent data suggest that in the adult hematopoietic system, expression of CD34 marks activated stem cells either about to self-renew and return to a state of CD34−ve quiescence, or to initiate commitment to differentiation, in which case CD34 expression is maintained (Sato et al., 1999). In skeletal muscle, we have also found that CD34 does not define lineage-negative stem cells; instead, the truncated isoform of CD34 is expressed by quiescent precurors that are committed to myogenesis by the criteria of Myf5 and M-cadherin expression. During development, Myf5 is unable to initiate muscle differentiation in the absence of MyoD, myogenin, and MRF4 (Renee Valdez et al., 2000) such that the expression of Myf5 should not instigate precocious myogenic differentiation. However, expression of Myf5 is required to prevent muscle cells from adopting alternative fates (Tajbakhsh et al., 1996b) and may act to restrict quiescent satellite cells to myogenesis. When satellite cells emerge from quiescence, alternate splicing and subsequent downregulation of CD34 means that MyoD is transiently coexpressed with CD34 full, whereas neither CD34 isoform is expressed once the cell has withdrawn from the cell cycle and entered terminal differentiation. The contrast between the quiescent CD34+ve satellite cell and the proliferating CD34+ve hematopoietic progenitor probably reflects the sporadic requirement for muscle repair and growth compared with the constant renewal of blood cells. However, in both cases, CD34 expression may play a role in preventing temporally or spatially premature differentiation.

Although our results show that the majority of quiescent satellite cells are CD34+ve, Myf5+ve, M-cadherin+ve, and MyoD−ve, a proportion do not appear to conform to this phenotype. When CD34, Myf5 and M-cadherin were used independently to count the numbers of satellite cells associated with single fibers isolated from the EDL muscles of adult mice, essentially the same values were obtained. This is consistent with the fact that in double-labeling experiments, all satellite cells expressed both markers. However, the CD34+ve, Myf5+ve, and M-cadherin+ve satellite cells account for only ~80% of the total number of satellite cells as determined using the 3F-nlacZ-2E transgenic mouse, strongly suggesting that the satellite cell compartment consists of at least two phenotypically distinct populations. In studies of muscle growth (Schultz, 1996) and regeneration (Rantanen et al., 1995; Heslop et al., 2000), the ability of precursors to repopulate host muscle after myoblast transplantation (Qu et al., 1998; Beauchamp et al., 1999) and in vitro clonal analyses (Schultz and Lipton, 1982; Baroffio et al., 1999).
maintenance and activation of quiescent, lineage-primed progenitors during adult tissue renewal and regeneration.

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Figure 9. Comparison of satellite cell numbers associated with isolated EDL muscle fibers determined by counting non–3F-nlacZ transgene-expressing nuclei, and M-cadherin–ve, CD34–ve, and Myf5–ve cells. Although the mean numbers of M-cadherin (4.5 ± 0.2, n = 114), CD34 (4.4 ± 0.2, n = 124), and Myf5–expressing cells (4.4 ± 0.3, n = 122) per EDL myofiber were not significantly different from each other (P < 0.05), the mean number of non–3F-nlacZ–transgenically expressing nuclei (5.6 ± 0.28) was significantly higher (P < 0.05). Values are expressed as population means ± SEM of the pooled data from six age-matched 3F-nlacZ–transgenic mice, and from six age-matched Myf5–nlacZ–transgenic mice.
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