Minireview

Preeclampsia As Modulator of Offspring Health

Violeta Stojanovska, Sicco A. Scherjon, and Torsten Plösch

Department of Obstetrics and Gynecology, University of Groningen, University Medical Center Groningen, The Netherlands

ABSTRACT

A balanced intrauterine homeostasis during pregnancy is crucial for optimal growth and development of the fetus. The intrauterine environment is extremely vulnerable to multisystem pregnancy disorders such as preeclampsia, which can be triggered by various pathophysiological factors, such as angiogenic imbalance, immune responses, and inflammation. The fetus adapts to these conditions by a mechanism known as developmental programming that can lead to increased risk of chronic noncommunicable diseases in later life. This is shown in a substantial number of epidemiological studies that associate preeclampsia with increased onset of cardiovascular and metabolic diseases in the later life of the offspring. Furthermore, animal models based predominantly on one of the pathophysiological mechanism of preeclampsia, for example, angiogenic imbalance, immune response, or inflammation, do address the susceptibility of the preeclamptic offspring to increased maternal blood pressure and disrupted metabolic homeostasis. Accordingly, we extensively reviewed the latest research on the role of preeclampsia on the offspring’s metabolism and cardiovascular phenotype. We conclude that future research on the pathophysiological changes during preeclampsia and methods to intervene in the harsh intrauterine environment will be essential for effective therapies.

early development, epigenetics, metabolism, preeclampsia

INTRODUCTION

The global prevalence of chronic cardiometabolic noncommunicable diseases (NCDs) diseases such as hypertension, cardiovascular disease, diabetes mellitus type 2, and metabolic syndrome has markedly increased during the past decades [1]. A number of genes and behavioral changes have been identified as initiators and mediators of these complex cardiometabolic diseases [2–4]. However, the increasing prevalence of NCDs cannot be accounted for by only these determinants. Biological factors already present during early development can lead to immediate cardiometabolic fetal responses that might have long-term effects.

The developmental origins of health and disease, or the Barker hypothesis, attempts to explain the high incidence of chronic NCDs by unfavorable in utero conditions. Depending on the severity of the insult during specific critical windows of fetal development, permanent tissue adjustments can occur, leading to long-term changes in organ function [5]. During pregnancy, the key regulatory organ of the intrauterine environment is the placenta, which serves as a metabolic, immune, and endocrine organ. It enables and regulates transport of gases, nutrients, hormones, immunoglobulins, and waste products between the mother and the fetus in order to maintain a favorable developmental homeostasis [6]. Hostile environmental factors present during early life, when rapid growth and differentiation is expected, can have a powerful impact on physiological health for a lifetime.

Preeclampsia is a pregnancy-associated syndrome, characterized by hypertension and proteinuria, affecting 2%–8% of the population worldwide [7]. It remains a major obstetric problem due to the high prevalence of maternal and fetal mortality and morbidity. Although the etiology is puzzling, several pathophysiologica mechanisms combined have proven to be involved at least in the clinical course of preeclampsia. Antiangiogenic imbalance, excessive inflammation, hypoxia, and/or autoantibodies targeting the renin-angiotensin system make up the harsh intrauterine environment during preeclampsia [8, 9]. All these factors may interact with the genome of the mother and the fetus in terms of gene expression modulation, ultimately affecting the expressed phenotype.

In this review, first we address epidemiological and human studies that show a contribution of preeclampsia to cardiometabolic alterations in the offspring. Further, we focus on animal studies in this research area, approaching three different mechanistic scenarios of preeclampsia. Finally, we discuss possible mechanisms that may explain relevance of preeclampsia in developmental programming of metabolic and cardiovascular diseases in the offspring.

EVIDENCE FROM HUMAN STUDIES: OFFSPRING STATUS AFTER PREECLAMPSIA

Birth weight screening is still an important assessment of optimal in utero nutrition and development. During preeclampsia, 13%–60% of the pregnancies are complicated by decreased birth weight depending on the region, maternal age, and the severity of the disease [10, 11]. Therefore, preeclampsia is one of the leading factors of fetal growth restriction [12, 13]. Low birth weight per se is already an established risk factor for
cardiovascular and metabolic diseases in later life, although the causal mechanisms are still speculative [14, 15].

Preeclampsia is characterized by new-onset hypertension during pregnancy (≥140/90 mmHg) along with proteinuria. However, little is known about neonatal blood pressure after this complication of pregnancy. An early report indicated that term neonates from preeclamptic mothers have a transient hypertension [16]. A more recent study showed that premature neonates from preeclamptic mothers, compared to controls, have early neonatal hypotension [17]. As indicated, blood pressure levels are also altered in these children, which appears to be associated with the gestational age. Furthermore, observation of blood pressure in school-age children previously exposed to preeclampsia showed higher systolic and diastolic blood pressure already at 8 yr of age [18–23]. Additionally, it was reported that these children have smaller hearts, increased heart rate, and features of cardiac diastolic dysfunction [24] as well as an increased risk of congenital heart defects, namely septal defects [25, 26]. However, in a cohort study, a 65-yr follow-up of preeclamptic offspring did not show an increased risk of coronary heart disease, but increased stroke incidence was reported [27].

Evaluation of endothelial functionality with noninvasive assessment can provide considerable insight into blood pressure risk stratification. School-age children previously exposed to preeclampsia showed increased vascular stiffness in the pulmonary and peripheral vascular system [24, 28]. Moreover, intact endothelial morphology is a potent vascular tone regulator. Analysis of endothelial cord cells showed a decreased number of endothelial colony-forming cells in contrast to increased senescent progenitor cells [29, 30]. This is indicative for at least advanced endothelial cell aging in the preeclamptic neonates.

The body mass index (BMI), plasma glucose, and lipid concentrations serve as strong indicators of optimal metabolic functioning and, when increased, are risk factors for cardiovascular and metabolic diseases. Preeclampsia shares many features with the metabolic syndrome, including increased maternal concentrations of proinflammatory cytokines, insulin, leptin, triglycerides, free fatty acid, and low-density cholesterol, usually in absence of diabetes [31]. Children from preeclamptic mothers show an increased risk of hospitalizations for endocrine and metabolic diseases in the first 5 yr of life [32]. In adolescence, premature-born preeclamptic males have an increased BMI in comparison to premature males born from normotensive pregnancies [33]. Cord blood samples from preeclamptic children show altered lipid profiles and increased tumor necrosis factor alpha (TNF-α) when studied for metabolic and inflammatory parameters [31, 34, 35], but in adolescence, these changes in glucose and lipid profiles are not prominent anymore [19, 21]. These effects may be influenced to some extent by maternal metabolic blood parameters and placental insufficiency. However, in school-age children previously exposed to preeclampsia, the metabolic phenotype shows changes only after subclustering of this group. The quantitative insulin sensitivity check index (QUICKI) serves as a predictive marker for diabetes onset based on fasting plasma glucose and insulin levels, and low values correspond to increased insulin resistance. Subdivision of groups based on QUICKI did show increased leptin and triglycerides levels in preeclampsia-exposed children that had independently low QUICKI values [36]. This suggests that insulin resistance independently, superimposed on earlier preeclampsia exposure, can serve as a strong predictor of the metabolic syndrome. These clinical observations reflect a transiently affected neonatal metabolism, which is not continuous through adolescence, but possibly can lead to increased susceptibility to the metabolic syndrome after a second environmental stressor, such as a metabolic stress.

**INTRAUTERINE ADVERSE ENVIRONMENT DURING PREECLAMPSIA AND OFFSPRING OUTCOME: ANIMAL STUDIES**

Animal models of preeclampsia can provide a unique possibility for understanding the causal relationship and the molecular networks of preeclampsia-induced offspring pathophysiology. Unfortunately, there is currently no perfect animal model of preeclampsia due to the complex and poorly understood pathophysiology of this disease (see Table 1 for an overview). Most of the presented models are based only on one pathophysiological feature, failing to reproduce the whole spectrum of preeclampsia characteristics. It is important to unravel whether all these experimental pathophysiological changes, which appear during preeclampsia, contribute to partial or complete cardiovascular and metabolic changes in the offspring. The use of several animal models of preeclampsia could help to distinguish the independent and/or dependent contribution of each of these factors to the developmental programming of offspring health. Below, we will discuss the animal studies that involve offspring follow-up after induction of major pathophysiological conditions of preeclampsia, excluding the genetic or surgically induced animal models of preeclampsia.

**Angiogenic Disparity**

Angiogenic dysbalance is a well-known feature of preeclampsia. The antiangiogenic factors soluble fms-like tyrosine kinase-1 (sFlt-1) and soluble endoglin (sEng) are increased in preeclamptic patients, after the 30th week of pregnancy [37]. Both sFlt-1 and sEng promote vascular dysfunction and capillary permeability, liver dysfunction, and neurological abnormalities via antagonization of proangiogenic factors such as VEGF and PGF or TGFβ signaling [38–40].

Adenoviral overexpression of sFlt-1 mimics the clinical course of preeclampsia in rodents [38]. Fetuses in this model show restricted growth that can be maintained until adulthood or can show catch-up growth until the age of 6 mo (Table 1) [41, 42]. Solely, sFlt-1 exposure during pregnancy imposes sex-specific glucose and/or insulin responses (to a glucose bolus) in the offspring, suggesting sex-specific differences in developmental programming of glucose metabolism. In addition, hypertension was observed only in the male offspring [41–43]. Sex-specific offspring outcomes are poorly understood, but one possible reason can be sexually dimorphic adaptations of the placenta [44].

When another environmental stressor, such as maternal obesity, is introduced during sFlt-1-induced preeclampsia, the offspring’s birth weight is not compromised. On the contrary, several biochemical parameters such as blood glucose, cholesterol, triglycerides, and leptin are increased in combination with increased fat tissue depositions and aberrant carotid vascular reactivity in both sexes [43, 45]. This may indicate that a single antiangiogenic intrauterine insult can influence sexual dimorphic changes in placenta by priming the males towards hypertensive phenotype, but this is not sufficient for profound metabolic alterations without additional trigger factors.

Unfortunately, the effects of increased sEng on offspring health are still largely unknown. In vivo studies in mice have
shown that direct administration of sEng increases the vascular resistance and subsequently the blood pressure [46]. In patients with diabetes and hypertension, sEng is positively correlated with the basal glucose levels, suggesting a potential role in glucose metabolism [47]. In accordance with previous findings and the known synergistic effect of sEng and sFlt-1 on preeclampsia outcome, we can speculate on the effects on offspring health in a similar or superimposed manner.

**Angiotensin II Type I Receptor Antibodies**

Angiotensin II type I receptor autoantibodies (AT1 AA) are found in 70%–95% of women diagnosed with preeclampsia, compared to 30% of healthy controls. A higher antibody titer is proportionally correlated to the severity of the disease [48, 49]. In addition, AT1 AA display an agonistic effect on the AT1 receptor, promoting vasoconstriction and aldosterone secretion, in a manner similar to angiotensin II [50–52].

Passive immunization with AT1 AA in rodents is associated with the development of proteinuria and hypertension at the end of pregnancy [53, 54]. The fetuses show growth restriction and remodeling in several organs, such as the liver, heart, and kidney. At the histopathological level, glomerular loss, myocardial apoptosis, and immature cell liver infiltration are observed in the offspring, suggesting an adaptive decline in fetal growth and organogenesis, possibly due to maternal-fetal transfer of AT1 AA. Irani et al. [53] reported unaffected functionality of these transported antibodies, and successful activation of fetal AT1 receptors may contribute to systemic vasoconstriction and hypoxia that can predispose the offspring to organ maladaptation.

Zhang et al. [54] did long-term follow-up on offspring derived from dams actively immunized against AT1 receptor antigen. Middle-age checkup at 10 mo of age showed elevated fasting insulin levels and an increased homeostasis model assessment index, suggesting the development of insulin resistance (Table 1). This was expected, especially because AT1 receptors are involved in insulin signaling of beta cells [55]. An additional 2 mo feeding with a high sugar diet of these adult offspring leads to even more pronounced metabolic alterations such as increased triglycerides, decreased high-density cholesterol, impaired glucose tolerance, and enlarged visceral fat depositions [54]. All these alterations are contributors to the progression of the metabolic syndrome. Surprisingly, blood pressure was normal in these animals, although they had been exposed to the AT1 antibodies in utero and in the weaning period via the maternal milk. One possible interpretation is that intrarenal angiotensin II, contrary to plasma angiotensin II, may be positively involved in blood pressure regulation. Another important comment is that vascular endothelium has relatively large regeneration capacities, and if there is no constant provocation with AT1 antibodies, no endothelial-related rise in blood pressure will occur.

**TABLE 1. Spectrum of cardiometabolic alterations in offspring from pre-eclamptic mothers (animal models).**

| Model | Species/strain | Offspring outcome | Offspring age | Reference |
|-------|----------------|-------------------|---------------|-----------|
| sFlt-1 overexpression | CD-1 mice | IPGTT variations | 24 wk | [42] |
| sFlt-1 overexpression | CD-1 mice | Hypertension in males | 9 wk | [41] |
| sFlt-1 overexpression | CD-1 mice | Metabolic changes: hypercholesterolemia, hyperleptinemia in females, and hypertriglyceridemia in males | 24 wk | [45] |
| Prepregnancy obesity and sFlt-1 overexpression | CD-1 mice | Fasting glucose increased in males | 12 wk | [43] |
| AT1 AA immunization | Wistar rats | Insulin resistance | 40 wk | [54] |
| Second impact: high sugar diet 20% sucrose | Wistar rats | Proteinuria | 3 wk | [143] |
| AT1 AA passive immunization | C67Bl/6J mice | Abnormal kidney and liver development | GD 18 | [53] |
| LPS injections | Sprague Dawley rats | Increased BW and fat deposits, hyperleptinemia, hypertension | 24 wk | [60] |
| LPS injections | Sprague Dawley rats | Hypertension | 25 wk | [61] |
| LPS injections | CD-1 mice | Decreased body weight | 32 wk | [62] |
| LPS injections | Sprague Dawley rats | Decreased glomeruli | 35 wk | [64] |
| LPS injections | Sprague Dawley rats | Impaired spermatogenesis | 12 wk | [65] |
| LPS injections + HF diet during pregnancy until 3 mo of age | Sprague Dawley rats | Insulin resistance | No data | [66] |
| LPS injections | ICR mice | Metabolic phenotype altered only due to the HF diet | No data | [66] |

*IPGTT, intraperitoneal glucose tolerance test; BW, body weight; AT1 AA, angiotensin II type I receptor autoantibodies; LPS, lipopolysaccharide; HF, high fat; GD, gestational day.
In sum, AT1 antibody exposure does not affect the fetal blood pressure but can have detrimental effects on organ formation and insulin resistance, which can be potentiated with an unhealthy diet. Nevertheless, more studies are needed in order to elucidate the underlying mechanisms of AT1 AA-induced fetal metabolic programming.

Inflammatory Milieu

Mild inflammation is generally considered a normal feature of pregnancy, whereas more exaggerated systemic inflammatory responses are characteristic of preeclampsia [9]. In accordance, proinflammatory cytokine concentrations are increased (TNFa, IL-6, IL-1b) in preeclamptic patients [56–58]. The association between inflammation and preeclampsia served as the basis for an experimental animal model of preeclampsia by low-dose intravenous infusion of bacterial endotoxin [59]. Nowadays, most of the developmental studies that involve exposure to lipopolysaccharide (LPS) during pregnancy are focused on the immunological consequences without concentrating on the possible preeclamptic symptoms in the dam.

Midgestational LPS exposure is characterized by a large range of cardiovascular events such as increased blood pressure, aortal vascular impairment, left ventricular hypertrophy, diastolic dysfunction, and myocardial apoptosis in adult offspring, without specific sex differences [60–64]. This implies striking endothelial and cardiac sensitivity of the fetus for inflammation that is maintained until adulthood, programing the offspring’s health toward cardiovascular functional decline. This, in part, can be explained by upregulation of the NF-kB signaling pathway, an increase of reactive oxygen species (ROS), and downregulation of the renal dopaminergic system leading to hypertension and vascular instability [64].

Combined effects of LPS and high-fat diet exposure during pregnancy have differential effects on offspring’s glucose and lipid metabolism (Table 1). It was shown that midgestation exposure to LPS and high-fat feeding until 3 mo of age can lead to impaired liver function and insulin resistance [65]. On the contrary, exposure to LPS in late gestation with additional high-fat diet stress after the lactation period did not result in an impaired metabolic phenotype in the offspring [66]. This suggests that timing of LPS exposure is crucial for fetal metabolic programming and in part can be explained by changes in maturational properties of the placenta, which in the last term of pregnancy are fully developed, possibly resulting in placental impermeability for the intermediate metabolic effectors of LPS [67]. Another important observation is that LPS and high-fat diet combined have a beneficial effect on blood pressure and the inflammatory response in the offspring, but not on the insulin resistance progression and liver dysfunction. Midgestation exposure to LPS seems to attenuate the offspring sensitivity to high-fat diet-induced inflammation [65]. In contrast, an aberrant inflammatory response on its own is not sufficient for a systemic breakdown in the regulation of insulin resistance.

Altogether, the data indicate that the developmental programming of offspring health via preeclampsia is caused by a two-hit combination of, first, systemic immunomodulatory and antiangiogenic signals during mid to late gestation and, second, a later host susceptibility marked by unhealthy lifestyle (e.g., a Western diet). These animal data have important translational consequences because the first hit is needed to affect the offspring’s development, and the presence of the second hit explains why only a minority of human fetuses exposed to preeclampsia develop detrimental cardiovascular and metabolic diseases later.

UNDERLYING MECHANISMS OF DEVELOPMENTAL PROGRAMMING

In order to interpret the developmental programming of cardiometabolic health via preeclampsia, we underline below the conserved mechanisms of chronic disease development, their interaction with the preeclamptic environment, and their effects on embryonic growth and epigenetic status (Fig. 1). Understanding the specific mechanisms by which preeclampsia impacts offspring welfare is crucial for developing appropriate strategies to improve the negative effects of the harsh intrauterine environment.

PLACENTAL PERMEABILITY: THE INITIATOR

The placental blood barrier serves as a protector and nutrient sensor between the mother and the child. In preeclampsia, placental morphology is perturbed showing superficial trophoblast invasion and insufficient remodeling of spiral arteries in the myometrium [68, 69]. Thus, with this defective placenta, two separate factors can influence its permeability: the placental composition and the exchange surface area.

Several factors influence placental composition, including an intact syncytiotrophoblast layer and cellular junctional assembly. The syncytiotrophoblast, a continuous membrane layer of the placenta, serves as a checkpoint for placental transport [70–72]. During preeclampsia, this layer is highly apoptotic [73], suggesting a dysfunctional adaptation of the placenta to increased fetal nutrient demand or an effect of the increased proinflammatory cytokines during preeclampsia. Consistent with this idea, tight junctions integral membrane proteins, important for paracellular transport of water and
nutrients, are extremely susceptible to TGFβ and IL-1β destruction, which can be reversed in vitro by specific cytokine inhibitors [74, 75]. Given that discontinuous placental membranes are accompanied with increased porosity, it is probable that loss of syncytiotrophoblast integrity underlines the defective nutrient transport of preeclamptic placenta.

An altered placental surface area in preeclampsia has been reported along with decreased placental weight, changes in the shape, and increased thickness, probably due to compensatory mechanisms [76]. The growth of the placenta was reported to be compromised only on the minor axis and corresponded to the severity of the preeclampsia. This axis is speculated to coincide with mediolateral development of the fetus, suggesting that this area is not spared during preeclampsia. In accordance, fetal length is less compromised in offspring in comparison with a severely affected abdominal circumference [34, 77].

A central question is whether these structural changes in the preeclamptic placenta are determinants for the transport of pathological signaling molecules to the fetus. It is known that inflammatory cytokines and AT1 AA can cross the placenta, but no data on the transfer of antiangiogenic molecules to the fetus is available [53, 78]. These molecules can act as potent signaling modifiers of glucose and lipid metabolism, but a definitive description of their mechanisms of action is lacking. Currently, we rely solely on animal data, for example, being challenged with an inflammatory cytokine such as TNFα induces insulin receptor downregulation in the liver that promotes the development of liver insulin resistance [79, 80]. An IL-6 challenge in rodents showed increased mobilization of acyl-CoA, a metabolic active form of fatty acid, in the skeletal muscles that has been strongly associated with lipid accumulation in muscles and peripheral insulin resistance [79–81]. AT1 AA exposure has detrimental effects on function of the liver, by NF-κB and NADPH oxidase dependent release of ROS [82, 83]. Ubiquitous exposure to sFlt-1 leads to hypovascularization in several organs, including pancreas and adipose tissue, that in turn can affect the beta cell mass and energy expenditure of the adipose tissue [84, 85]. In addition, prolonged exposure to sFlt-1 is involved in the development of diastolic dysfunction and heart failure [86]. Taken together, increased exposure of the fetal organism to these molecules may have a detrimental effect on proper metabolic and cardiovascular functioning.

ROS: THE MEDIATORS

Multiple lines of evidence suggest that oxidative damage is one of the underlying mechanisms of many chronic diseases such as type 2 diabetes, obesity, hypertension, atherosclerosis, and metabolic syndrome [87, 88]. Oxidative stress occurs as soon as the production and consumption of ROS are imbalanced.

During preeclampsia, inflammatory cytokines and AT1 AA promote increased ROS production by up to 40% when compared to control placentas [89–92]. Whether oxidative distress is a cause or consequence of placental dysfunction and/or fetal nutrient demand is a matter of ongoing debate, and most studies simply describe an association of ROS overflow with metabolic consequences rather than mechanistic connections.

Normal fetal development is dependent on tightly controlled oxidative stress exposure for optimal cellular signaling, differentiation, and proliferation [93]. However, during preeclampsia the functionality of the placenta is reduced and the antioxidant capacity is diminished, suggesting increased oxidative stress transfer to the fetus. Cord blood analyzed from preeclamptic mothers showed either decreased antioxidant activity [94] or increased oxidative stress markers [95–98], but not for all [99], suggesting a possibility of lipid peroxidation and protein inactivation in the fetus [100]. Several tissues are extremely susceptible to oxidative damage, including beta cells and vascular endothelium mainly due to low cytoprotective mechanisms [101, 102]. Furthermore, treatment of hypoxic dams with antioxidants during gestation ameliorates the vascular dysfunction in the offspring, indicating that antioxidant treatment may indeed be an interventional treatment [103]. On the contrary, clinical trials that involved routine antioxidant supplementation during pregnancy contradict the idea of preventive effect towards preeclampsia [104, 105]. Another trial that included only high-risk preeclampsia patients reported protective effect of antioxidant vitamins in combination with L-arginine [106], suggesting that exclusive antioxidant treatment is not sufficient to combat preeclampsia.

LEPTIN SIGNALING AS MEDIATOR

Leptin is a satiety hormone, and acting via JAK2/STAT3 and PI3K-Akt signaling pathways, leptin has a major impact on energy homeostasis, body composition, and appetite in early fetal and later adult life [107]. Moreover, leptin expression is responsive to the intrauterine and fetal environment, showing overexpression in monochorionic twin placenta only on the side of the growth restricted fetus [108]. The extent to which leptin signaling is implicated in overall fetal metabolism is unknown, but there is evidence that it stimulates fatty acid oxidation in muscles, increases the glucose turnover in brain, heart, and brown adipose tissue, and inhibits global lipid accumulation [109]. By contrast, reduction of leptin concentrations and the state of leptin resistance share similar effects on metabolism, promoting hyperinsulinemia and hyperglycemia. During preeclampsia, maternal leptin concentrations show a 2-fold increase in comparison to control subjects, irrespective of the BMI [110–114], which is usually combined with hyperinsulinemia, altered lipid profile, and decreased 2-methoxyestradiol, which serves as an important vasoprotector and vasodilator [115, 116]. There is a dichotomy, therefore, between the protective metabolic effects of leptin and apparently deleterious effects of hyperleptinemia on maternal health. In part, this can be explained by a development of leptin resistance, mainly due to inactivation of STAT3 intracellular activity that is also decreased in preeclamptic pregnancies [117].

Fetal cord leptin concentrations are increased [110, 118] in preeclamptic pregnancies, possibly due to increased nutrient demand of the fetus and/or increased placental permeability and consequent leptin flow to the fetus. Hyperleptinemia in utero can alter the adrenal responsiveness [56, 119] in the fetus and together with increased inflammatory markers can develop a defense mechanism of leptin resistance, which ultimately can lead to deleterious effects on cardiovascular and metabolic health.

CELLULAR ADAPTATIONS AS EFFECTORS

Optimal organ functioning is dependent on the quantity, morphology, and functionality of relevant cell types due to appropriate differentiation of pluripotent embryonic stem cells. Although the mechanisms of embryonic cell fate decisions are obscure, the presence of low energy levels and prominent signaling networks are strongly correlated with disturbed metabolic stem cell fate [120–122]. Importantly, all these
adverse conditions are also present in utero during preeclampsia.

Moreover, several reports showed changes in the number of nephrons, beta cells, and/or cardiomyocytes in offspring exposed to a harsh intrauterine environment [123–126]. A decreased number of nephrons contributes to low rates of renal ultrafiltration that affects blood circulating volume, which ultimately can lead to increased blood pressure [126]. Decreased beta-cell mass adaptation due to early life stressors, such as undernutrition and placental insufficiency, possibly can have an influence on later disease development, for example, diabetes mellitus [127]. Initial heart size has an influence on the end diastolic volume and serves as a predictive index for myocardial disease [24].

These (mal)adaptive changes are observed mainly in organs constructed from long-lived postmitotic cells [123, 128]. Because these cells are not—or rarely—dividing cells, their development during intrauterine life is extremely important in order to prepare them for long-term functionality. Tightly controlled processes regulate cell number while their functioning is dependent on specific signaling molecules and energy sources. During preeclampsia, the increased concentration of inflammatory markers and improper vascular signaling molecules might perturb these regulatory processes essential in organ formation [129]. Combined with poor nutrient supply via the placenta, this can lead to detrimental effects on offspring health.

**EPIGENETIC CHANGES AS EFFECTORS**

Exposure to different environmental stimuli, especially during critical windows of development, results in the formation of adaptive epigenetic marks as part of the adaptive stress response [5]. The epigenetic marking system includes changes in DNA methylation, histone modifications, and noncoding RNA (ncRNA) expression. Usually, they are established early in development and act as regulators of developmental, tissue, and sex-specific gene expression [130–133].

DNA methylation is a unique form of gene regulation because it involves direct covalent modification within the genome and can provide long-term stability in a heritable transgenerational way [134]. Methylation of important regulatory sites, for example, gene promoters or enhancers, is mostly connected to gene repression, resulting in gene expression downregulation [135].

DNA methylation analysis of cord blood cells is a valuable target for studying the early epigenetic consequences of preeclampsia on the fetus. Several studies analyzed DNA methylation of genes involved in fetal growth and development that are also highly sensitive to environmental perturbations. Hypomethylation has been observed in the promoter region of the 11β-hydroxysteroid dehydrogenase type 2 (HSD11B2) in cord blood samples from neonates exposed to preeclampsia [136]. Decreased methylation was also reported for insulin-like growth factor 2 (IGF2) in the differentially methylated regions, important for gene regulation of imprinted genes [137]. By contrast, in preeclamptic placentas, HSD11B2 and IGF2 gene expression levels are decreased [138, 139]. Therefore, there is a discrepancy between the reported hypomethylated status and the observed downregulated activity of these genes in other studies. It is tempting to speculate that this is a compensatory change in methylation to ensure favorable offspring functioning, but on the other side, it can be an atypical decrease in gene expression that can lead to metabolic maladaptation.

A recent study used a genomewide methylation analysis in which neonatal cord blood DNA from mothers diagnosed with early onset preeclampsia showed promoter hypo- or hyper-methylation for different subsets of genes. Prominent DNA modifications were primarily discovered in genes involved in lipid metabolism and inflammation, pointing out that early epigenetic disruptions can be seen in preeclamptic children [140]. Altogether, these findings support an effect of preeclampsia on the methylation status of the neonates cord blood, but it is unclear whether this is a protective or maladaptive effect. Although this does not prove any causal relationship with long-term health effects, it can be used as an initial proof of principle for conductions of new cohort studies.

To our knowledge, there are no data concerning histone modifications and/or ncRNAs in offspring from preeclamptic mothers. Communication between DNA methylation and chromatin modifiers or promoter regions of ncRNAs has been established [141, 142], and abnormal methylation either solely or via other epigenetic marks can be an important mediator of fetal metabolism. It is becoming clear that these molecules are implicated in several diseases, and successful unclosing of their role in developmental programming can lead to possible biological biomarkers or targets for therapy.

**CONCLUDING REMARKS**

Taking into consideration the great amount of evidence, it is reasonable to suggest that preeclampsia constrains the cardiometabolic health of the offspring. Still, it remains difficult to estimate the degree of involvement of preeclampsia into the cardiovascular and metabolic health programming of the offspring. The major obstacle is the presence of multiple pathophysiological pathways implicated in the development and the clinical course of preeclampsia that may influence each other or act independently all at once or in series. Depending on which mechanism is dominantly involved, and which secondary environmental stressors are present, different aspects of the metabolism and the cardiovascular system can be affected. The role of the placenta as a central initiator of long-term preeclamptic consequences in the offspring is just the beginning of what needs to be explored (Fig. 1). However, understanding of all mechanisms by which preeclampsia alters fetal growth and development and later on programs it toward chronic disorders is crucial for identification of individuals at risk and for development of future clinical interventions or prevention strategies.

**REFERENCES**

1. World Health Organization. WHO Noncommunicable Diseases Country Profiles. Geneva, Switzerland: WHO Press; 2014: 2014.
2. Pollex RL, Hegele RA. Genetic determinants of the metabolic syndrome. Nat Clin Pract Cardiovasc Med 2006; 3:482–489.
3. Mitchell BD, Imumorin IG. Genetic determinants of diabetes and atherosclerosis. Curr Atheroscler Rep 2002; 4:193–198.
4. Norman RE, Carpenter DO, Scott J, Brunе MN, Sły P.D. Environmental exposures: an unrecognized contribution to noncommunicable diseases. Rev Environ Health 2013; 28:59–65.
5. Jiménez-Chillarón JC, Díaz R, Martínez D, Pentinat T, Ramón-Krauel M, Ribó S, Plösch T. The role of nutrition on epigenetic modifications and their implications on health. Biochimie 2012; 94:2242–2263.
6. Carter AM. Evolution of placental function in mammals: the molecular basis of gas and nutrient transfer, hormone secretion, and immune responses. Physiol Rev 2012; 92:1543–1576.
7. Duley L. The global impact of pre-eclampsia and eclampsia. Semin Perinatol 2009; 33:130–137.
8. Larea-goiti-Servitje E, Gómez-Lopez N. The pathophysiology of preeclampsia involves altered levels of angiogenic factors promoted by...
hypoxya and autoantibody-mediated mechanisms. Biol Reprod 2012; 87: 36–36.

9. Redman CWG, Sargent IL. Immunology of pre-eclampsia. Am J Reprod Immunol 2010; 63:534–543.

10. Srinivas SK, Eddow AG, Neff PM, Samuel MD, Andrea CM, Elowitz MA. Rethinking IUGR in preeclampsia: dependent or independent of maternal hypertension? J Perinatol 2009; 29:680–684.

11. Weiler J, Tong S, Palmer KR. Is fetal growth restriction associated with a more severe maternal phenotype in the setting of early onset pre-eclampsia? A retrospective study. PLoS One 2011; 6:e26937.

12. Xiao R, Sorensen TK, Williams MA, Luthy DA. Influence of preeclampsia on fetal growth. J Matern Fetal Neonatal Med 2003; 13:157–162.

13. Ødega rd RA, Vatten LJ, Nilsen ST, Salvesen KÅ, Austgulen R. Preeclampsia and fetal growth. Obstet Gynecol 2000; 96:950–955.

14. Eriksson JG, Forse T, Tuomilehto J, Osmond C. Early growth, adult income, and risk of stroke. Stroke 2000; 31:869–875.

15. Jong M, Lafeber HN, Cranendonk A, van Weissenbruch MM. The association between maternal and offspring cardiovascular disease and their risk factors. Paediatr Perinat Epidemiol 2007; 21:127–122.

16. Miller FC, Read JA, Cabal L, Siassi B. Heart rate and blood pressure in neonates born to diabetic mothers. Horm Res Pediatr 2014; 81:43–49.

17. Miller FC, Read JA, Cabal L, Siassi B. Heart rate and blood pressure in infants of pre-eclamptic mothers during the first hour of life. Crit Care Med 1983; 11:532–535.

18. Teng RJ, Wu TJ, Sharma R, Garrison RD, Hudak ML. Early neonatal hypotension in premature infants born to preeclamptic mothers. J Perinatol 2006; 26:471–475.

19. Dowling P, Newton-Swuowski AJ, Lzadzam M, Kelly BA, Kyraku T, Leeson P. Preeclampsia and offspring cardiovascular health: mechanistic insights from experimental studies. Clin Sci 2012; 123: 53–72.

20. Tenhola S, Rahila E, Martikainen A, Halonen P, Voutilainen R. Blood pressure, serum lipids, fasting insulin, and adrenal hormones in 12-year-old children born with maternal preeclampsia. J Clin Endocrinol Metab 2004; 89:1217–1222.

21. Miettola S, Hartikainen AL, Voutilainen R, Sattar N, Ramsay JE, Greer IA. Fetal cardiac function in offspring 5-8 years after pregnancy complicated by preeclampsia. Early Hum Dev 2011; 87:531–535.

22. Brodwall K, Leirgul E, Greve G, Vollset SE, Holmstrøm H, Tell GS, Øyen N. Possible common aetiology behind maternal preeclampsia and congenital heart defects in the child: a cardiovascular diseases in Norway project study. Paediatr Perinat Epidemiol 2015; 30:76–85.

23. Auger N, Fraser WD, Healy-Proftis J, Arbour L. Association between preeclampsia and congenital heart defects. JAMA 2015; 314:1588.

24. Kajantie E, Eriksson JG, Osmond C, Thornburg K, Barker DJP. Preeclampsia is associated with an increased pro-inflammatory profile in newborns. J Reprod Immunol 2015; 112:111–114.

25. Seppä S, Voutilainen R, Tenhola S. Markers of insulin sensitivity in 12-year-old children born from preeclamptic pregnancies. J Pediatr 2015; 167:125–130.

26. Powers BW, Jeyabaln A, Clifton RG, van Dorsten P, Hauth JC, Klebanoff MA, Lindheimer MD, Sibai B, Landon M, Miodovnik M. Solute fructose-1,6-bisphosphate in preeclampsia among high-risk pregnancies. PLoS One 2010; 5:e13263.

27. Maynard SE, Min J, Merchant J, Lim K, Li J, Mondal S, Libermann TA, Morgan JP, Sellek FW, Stillman IE, Epstein FH, Sukhatme VP, et al. Excess placental solute fructose-1,6-bisphosphate in preeclampsia. J Clin Invest 2003; 111:649–658.

28. Chaiworapongsa T, Romero R, Espinoza J, Bujold E, Mee Kim Y, Goncalves LF, Gomez R, Edwin S. Evidence supporting a role for blockade of the vascular endothelial growth factor system in the pathophysiology of pre-eclampsia: Young Investigator Award. Am J Obstet Gynecol 2004; 190:1541–1550.

29. Levin RJ, Liao C, Qin J, Yu KP, Maynard SE, Sachs BP, Sibai BM, Epstein FH, Romero R, Thadhani R, Karumanchi SA. Solute fructose and other circulating angiogenic factors in preeclampsia. N Engl J Med 2006; 355:992–1005.

30. Lu F, Bytyaetine E, Tamayo E, Gamble P, Anderson GD, Hankins GDV, Longo M, Saade GR. Gender-specific effect of overexpression of sFlt-1 in pregnant mice on fetal programming of blood pressure in the offspring later in life. Am J Obstet Gynecol 2007; 197:1–5.

31. McDonnell M, Tamayo E, Kechichian T, Gamble P, Longo M, Hankins GDV, Saade GR, Costantine MM. The effect of prenatal pravastatin treatment on altered fetal programming of postnatal growth and metabolic function in a preeclampsia-like murine model. Am J Obstet Gynecol 2014; 210:542.

32. Byers BD, Betancourt A, Lu F, Hankins GDV, Longo M, Saade GR, Bytyaetine E. The effect of prepregnancy obesity and sFlt-1-induced preeclampsia-like syndrome on fetal programming of blood pressure in a mouse model. Am J Obstet Gynecol 2009; 200:432.

33. Pruis MG, Gellhaus A, Kühnel E, Lendvai A, Blows VW, Groen AK, Pötsch T. Sex-specific placental differences as a contributor to sex-specific metabolic programming? Acta Physiol 2015; 215:127–129.

34. Bytyaetine E, Tamayo E, Kechichian T, Derez N, Gamble P, Hankins GDV, Saade GR. Prepregnancy obesity and sFlt-1-induced preeclampsia in mice: developmental programming model of metabolic syndrome. Am J Obstet Gynecol 2011; 204:398.

35. Venkatesha S, Toporisan M, Lam C, Hanai J, Mammoth T, Kim YM, Bdlolah Y, Lim K-H, Yuan H-T, Libermann TA, Stillman IE, Roberts D, et al. Soluble endoglin contributes to the pathogenesis of preeclampsia. Nat Med 2006; 12:642–649.

36. Blázquez-Medella AM, García-Ortiz L, Gómez-Marcos MA, Recio-Rodriguez JJ, Sánchez-Rodriguez A, Martín-Martínez S, Martínez-Sánchez S, et al. Increased placental soluble endoglin levels as an indicator of cardiovascular alterations in hypertensive and diabetic patients. BMC Med 2010; 8:86.

37. Siddiqui AH, Irani RA, Blackwell SC, Ramín SM, Kellems RE, Xia Y. Angiotensin receptor agonist autoantibody is highly prevalent in preeclampsia: correlation with disease severity. Hypertension 2010; 55:386–393.

38. Herse F, Lamarca B. Angiotensin II type 1 receptor autoantibody (AT1-A)-mediated pregnancy hypertension. Am J Reprod Immunol 2013; 69: 413–418.

39. Wallukat G, Homuth V, Fischer T, Lindschau C, Horstkamp B, Jäppner A, Baur E, Nissen E, Vetter K, Neichel D, Dudenhausen JW, Haller H, et al.
Patients with preeclampsia develop怔istic autoantibodies against the angiotensin AT1 receptor. J Clin Invest 1999; 103:945–952.

51. Wenzel K, Rajakumar A, Haase H, Geusens N, Hubner N, Schulz H, Brewer J, Roberts L, Hubel CA, Herre F, Herling L, Qadri F, et al. Angiotensin Ⅱ type 1 receptor antibodies and increased angiotensin Ⅱ sensitivity in pregnant rats. Hypertension 2011; 58:77–84.

52. Hubel CA, Wallakat G, Wolf M, Herre F, Rajakumar A, Roberts JM, Markovic N, Thadhani R, Luft FC, Dechend R. Angonic angiotensin III type 1 receptor autoantibodies in postpartum women with a history of preeclampsia. Hypertension 2007; 49:612–617.

53. Irani RA, Zhang Y, Blackwell SC, Zhou CC, Ramin SM, Kellemes RE, Xia Y. The detrimental role of autoantibody angiogenic receptors in uterine growth restriction seen in preeclampsia. J Exp Med 2009; 206:2809–2822.

54. Zhang S, Zhang X, Yang L, Yan Z, Yan L, Tian J, Li X, Song L, Wang L, Yang X, Zheng R, Lau WB, et al. Increased susceptibility to metabolic syndrome in adult offspring of angiotensin type 1 receptor autoantibody-positive rats. Antioxid Redox Signal 2012; 17:733–743.

55. Chu KY, Tung L, Carlson PO, Leung PS, Angiotensin Ⅱ. Type 1 receptor blockade improves β-cell function and glucose tolerance in a mouse model of type 2 diabetes. Diabetes 2006; 55:367–374.

56. Lockwood CJ, Yen C-F, Basar M, Kayisli UA, Martel M, Buhimschi I, Buhimschi C, Huang SJ, Krikun G, Schatz F. Preeclampsia-related inflammatory cytokines regulate interleukin-6 expression in human decidual cells. Am J Pathol 2008; 172:1571–1579.

57. Sharma A, Satyam A, Sharma JB. Leptin, IL-10 and inflammatory mediators (TNF-α, IL-6 and IL-8) in pre-eclamptic, normotensive pregnant and healthy non-pregnant women. Am J Reprod Immunol 2007; 58:21–30.

58. Lau SY, Guild SJ, Barrett CJ, Chen Q, McCowan L, Jordan V, Chamley LW. Tumor necrosis factor-α, interleukin-6, and interleukin-10 levels are altered in preeclampsia: a systematic review and meta-analysis. Am J Reprod Immunol 2013; 70:412–427.

59. Faas M, Schuiling G, Baller J, Visscher C, Bakker W. A new model for human placentation: ultra-low-dose endotoxin infusion in pregnant rats. Am J Obstet Gynecol 1994; 171:158–164.

60. Wei Y, Li X, Zhou J. Prenatal exposure to lipopolysaccharide results in myocardial remodelling in adult murine offspring. J Inflamm 2013; 10:35.

61. Hao X-Q, Zhang H-G, Yuan Z-B, Yang D-L, Hao L-Y, Li X-H. Prenatal exposure to lipopolysaccharide alters the intrarenal renin-angiotensin system and renal damage in offspring rats. Hypertens Res 2010; 33:76–82.

62. Wei Y, Du W, Xiong X, He X, Yi P, Deng Y, Chen D. Prenatal exposure to lipopolysaccharide results in increases in blood pressure and body weight in rats. Acta Pharmacol Sin 2007; 28:651–656.

63. Hao X-Q, Zhang H-G, Yuan Z-B, Yang D-L, Hao L-Y, Li X-H. Prenatal exposure to lipopolysaccharide alters the intrarenal renin-angiotensin system and renal damage in offspring rats. Hypertens Res 2010; 33:76–82.

64. Wei Y, Du W, Xiong X, He X, Yi P, Deng Y, Chen D. Prenatal exposure to lipopolysaccharide results in myocardial remodelling in adult murine offspring. J Inflamm 2013; 10:35.

65. Zhao S, Zhang H, Cao D, Liu Y, Li X, Song L, Wang L, Yang X, Zheng R, Lau WB, et al. Increased susceptibility to metabolic syndrome in adult offspring of angiotensin type 1 receptor autoantibody-positive rats. Antioxid Redox Signal 2012; 17:733–743.

66. Liu X, Wang B, Zhao M, Zhang C, Chen Y. Effects of maternal LPS exposure during pregnancy on metabolic phenotypes in female offspring. PLoS One 2014; 9:e88127.

67. Liu X, Wang B, Zhao M, Zhang C, Chen Y. Effects of maternal LPS exposure during pregnancy on metabolic phenotypes in female offspring. PLoS One 2014; 9:e114780.

68. Malassine A, Frendo JL, Evain-Brion D. A comparison of placental development and endocrine functions between the human and mouse model. Hum Reprod Update 2003; 9:531–539.

69. Notis M, Perez-Casasnovas G. Mechanisms of disease: pre-eclampsia. Nat Clin Pract Nephrol 2005; 1:98–114.

70. Lyall F, Robson SC, Bulmer JN. Spiral artery remodelling and trophoblast invasion in preeclampsia and fetal growth restriction relationship to clinical outcome. Hypertension 2013; 62:1046–1054.

71. Sibley CP, Brownhill P, Dilworth M, Glazier JD. Review: adaptation in placental nutrient supply to meet fetal growth demand: Implications for programming. Placenta 2010; 31:570–574.

72. Gude NM, Roberts JT, Kallioni M, King RG. Growth and function of the normal human placenta. Thromb Res 2004; 114:397–407.

73. Powell TL, Jansson T, Illsley PN, Wennergren M, Korotkova M, Strandvik B. Composition and permeability of syncytiotrophoblast plasma membranes in pregnancies complicated by intrauterine growth restriction. Biochim Biophys Acta Biomembr 1999; 1420:86–94.

74. Ishihara N, Matsuo H, Murakoshi H, Laag-Fernandez JB, Samoto T, Maruo T. Increased apoptosis in the syncytiotrophoblast in human term placentas complicated by either preeclampsia or intrauterine growth retardation. Am J Obstet Gynecol 2002; 186:158–166.

75. Liévano S, Alarcón L, Chávez-Munguía B, González-Mariscal L. Endothelia of term human placentae display diminished expression of α tight junction proteins during preeclampsia. Cell Tissue Res 2006; 324:433–448.

76. Tossetta G, Paolinielli F, Avellini C, Salvolini E, Ciaramella M, Torri M, Totti P, Giulianini L, Cadeddu M, Cossu F, et al. IL-1β and TGF-β2 weaken the placental barrier through destruction of tight junctions: an in vivo and in vitro study. Placenta 2014; 35:509–516.

77. Kajantie E, Thornburg KL, Eriksson JG, Osmond C, Barker DJP. In preeclampsia, the placenta grows slowly along its minor axis. Int J Dev Biol 2010; 54:469–473.

78. Rasmussen S, Irgens LM. Fetal growth and body proportion in preeclampsia. Obstet Gynecol 2003; 101:575–583.

79. Dahlgren L, Samuelsson AM, Jansson T, Holmgård A. Increased susceptibility to metabolic syndrome in adult offspring of angiotensin type 1 receptor autoantibody-positive rats. Antioxid Redox Signal 2012; 17:733–743.

80. Cheung AT, Ree D, Kolls JK, Fuselier J, Coy DH, Bryer-Ash M. An in vivo model for elucidation of the mechanism of tumor necrosis factor-alpha (TNF-α)-induced inflammation: resistance: for differential regulation of inflammation signaling by TNF-α. Endocrinology 1998; 139:4928–4935.

81. Kim HJ, Higashimori T, Park SY, Choi H, Dong JY, Kim YJ, Noh HL, Cho YR, Cline G, Kim YB, Kim JK. Differential effects of interleukin-6 and -10 on skeletal muscle and liver insulin action in vivo. Diabetes 2004; 53:1060–1067.

82. Dechend R, Gieffers J, Dietz R, Joerres A, Rupp J, Luft FC, Maass M. Angiotensin type 1 receptor antibodies and increased angiotensin II sensitivity in diabetes: implication for the development of diabetes mellitus type 1. Exp Med 2009; 206:2809–2822.

83. Houstis N, Rosen ED, Lander ES. Reactive oxygen species have a causal role in multiple forms of insulin resistance. Nature 2006; 440:944–948.

84. D’Hoker J, De Leu N, Heremans A, Roosen A, Moutquin J, Leblanc S. Potential biomarkers of early gestation in pregnancies complicated by intrauterine growth retardation. Am J Obstet Gynecol 2002; 186:158–166.
Mistry HD, Wilson V, Ramsay MM, Symonds ME, Pipkin FB. Reduced selenium concentrations and glutathione peroxidase activity in preeclamptic pregnancies. Hypertension 2008; 52:881–888.

Tsukahara H, Ohita N, Sato S, Hiroaka M, Shukunami K-I, Uchiyama M, Kawakami H, Sekine K, Mayumi M. Concentrations of pentosidine, an advanced glycation end-product, in umbilical cord blood. Free Radic Res 2004; 38:691–695.

Torrance HL, Krediet TG, Vreman HJ, Visser GH, van Bel F. Oxidative stress and proinflammatory cytokine levels are increased in premature neonates of preeclamptic mothers with HELLP syndrome. Neonatology 2008; 94:138–142.

Hillal N, Kocyigit A, Demir M, Camuzcuoglu A, Incebiyik A, Camuzcuoglu H, Vural M, Taskin A. DNA damage and oxidative stress in patients with mild preeclampsia and offspring. Eur J Obstet Gynecol Reprod Biol 2013; 170:377–380.

Eredm M, Harma M, Harma IM, Arikani I, Barut A. Comparative study of oxidative stress in maternal blood with that of cord blood and maternal milk. Arch Gynecol Obstet 2012; 285:371–375.

Braekke K, Harsen NK, Staff AC. Oxidative stress and antioxidant status in fetal circulation in preeclampsia. Pediatr Res 2006; 60:560–564.

Yaacob N, Ohel G, Hochman A. Reactive oxygen species in the process of labor. Arch Gynecol Obstet 1999; 263:23–24.

Lenzen S, Drinkmier J. Low antioxidant enzyme gene expression in patients with HELLP syndrome. Free Radic Biol Med 1996; 20:463–466.

Kajiya M, Hirota M, Inai Y, Kiyosuka T, Morimoto T, Iwasaki T, Endo K. Maternal plasma leptin is increased in patients with mild preeclampsia and offspring. Eur J Obstet Gynecol Reprod Biol 2011; 204:327.

Vijekotte TG, Kruziener N, Hutton BA, Vollebregt KC, Van Eijden M, Twickler MB. Maternal lipid profile during early pregnancy and pregnancy complications and outcomes: the ABCD study. J Clin Endocrinol Metab 2012; 97:3917–3925.

Zhang Z, Yang X, Zhang L, Duan Z, Jia L, Wang P, Shi Y, Li Y, Gao J. Decreased expression and activation of Stat3 in severe preeclampsia. J Mol Histol 2015; 46:205–219.

Ogedagj RA, Vatten LJ, Nilseren ST, Salvesen KÅ, Austuglen R. Umbilical cord plasma leptin is increased in preeclampsia. Am J Obstet Gynecol 2002; 186:427–432.

Yuen BSJ, Owens PC, Symonds ME, Keisler DH, McFargane J, Kauter KG, McMullen IC. Effects of leptin on fetal plasma adrenocorticotrophic hormone and cortisol concentrations and the timing of parturition in the sheep. Biol Reprod 2004; 70:1650–1657.

Kobayashi CI, Suda T. Regulation of reactive oxygen species in stem cells and cancer stem cells. J Cell Physiol 2012; 227:421–430.

Bauer S. Cytokine control of adult neural stem cells. Ann N Y Acad Sci 2009; 1153:48–56.

Chaudhari P, Ye Z, Jang Y-Y. Roles of reactive oxygen species in the fate of stem cells. Antioxid Redox Signal 2014; 20:1881–1890.

Pallada DP, Khosla K, Kliot M, Baurer J, Baum M, Bates OA. Link between reduced nephron number and hypertension: studies in a mouse model. Pediatr Res 2006; 59:489–493.

Corstius HB, Zimaniy MA, Maka N, Herath T, Thomas W, Van Der Laarse A, Wreford NG, Black MJ. Effect of intrauterine growth restriction on the number of cardiomyocytes in rat hearts. Pediatr Res 2005; 57:796–800.

Hinchcliffe SA, Lynch MR, Sargen Ph, Howard CV, Van Velzen D. The effect of intrauterine growth retardation on the development of renal nephrons. Br J Obstet Gynaecol 1992; 99:296–301.

Cebran C, Asai N, D’Agati V, Costantini F. The number of fetal nephron progenitor cells limits ureteric branching and adult nephron endowment. Cell Rep 2014; 7:127–137.

Stanger BZ, Tanaka AJ, Melton DA. Organ size is limited by the number of embryonic progenitor cells in the pancreas but not the liver. Nature 2013; 498:455–461.

Terman A, Kurz T, Navratil M, Arriaga EA, Brunck UT. Mitochondrial turnover and aging of long-lived postmitotic cells: the mitochondrial-lysosomal axis theory of aging. Antioxid Redox Signal 2010; 12:503–535.

Rafalski VA, Mancini E, Brunet A. Energy metabolism and energy-sensing pathways in mammalian embryonic and adult stem cell fate. J Cell Sci 2012; 125:550–557.

Jaenisch R, Young R. Stem cells, the molecular circuitry of pluripotency and nuclear reprogramming. Cell 2008; 132:567–582.

Ellis HL, Shioida K, Rosenthal NF, Coser KR, Shioida T. Masculine epigenetic sex marks of the CYP19A1/aromatase promoter in genetically male chicken embryonic gonads are resistant to estrogen-induced phenotypic sex conversion. Biol Reprod 2012; 87:23.

Marcho C, Bevilacqua A, Tremblay KD, Mager T. Tissue-specific regulation of Igf2/Ins imprinting during gastrulation. Epigenetics Chromatin 2015; 8:10.

Cantone I, Fisher AG. Epigenetic programming and reprogramming during development. Nat Struct Mol Biol 2013; 20:282–9.

Orozco LD, Rubbi L, Martin LJ, Fang F, Hormozdari F, Che N, Smith AD, Lusis AJ, Pellegrini M. Intergenerational genomic DNA methylation patterns in mouse hybrid strains. Genome Biol 2014; 15:R68.

Hensenmann J, Bar-Nur O, Ben-David E, Benvenisty N. Global indiscriminate methylation in cell-specific gene promoters following reprogramming into human induced pluripotent stem cells. Stem Cell Reports 2013; 1:509–517.

Hu W, Weng X, Dong M, Liu Y, Li W, Huang H. Alteration in methylation level at 11β-hydroxysteroid dehydrogenase type 2 gene promoter in infants born to preeclamptic women. BMC Genet 2014; 15:26.

He J, Zhang A, Fang M, Fang G, Ge J, Jiang Y, Zhang H, Han C, Ye X, Yu D, Huang H, Liu Y, et al. Methylation levels at Igf2 and Gnas DMRs in infants born to preeclamptic pregnancies. BMC Genomics 2013; 14:472.
138. Aufdenblatten M, Baumann M, Raio L, Dick B, Frey BM, Schneider H, Surbek D, Hocher B, Mohaupt MG. Prematurity is related to high placental cortisol in preeclampsia. Pediatr Res 2009; 65:198–202.

139. Bourque DK, Avila L, Peñaherrera M, von Dadelszen P, Robinson WP. Decreased placental methylation at the H19/IGF2 imprinting control region is associated with normotensive intrauterine growth restriction but not preeclampsia. Placenta 2010; 31:197–202.

140. Ching T, Ha J, Song M-A, Tiirikainen M, Molnar J, Berry MJ, Towner D, Garmire LX. Genome-scale hypomethylation in the cord blood DNAs associated with early onset preeclampsia. Clin Epigenetics 2015; 7.

141. Reddington JP, Perricone SM, Nestor CE, Reichmann J, Youngson NA, Suzuki M, Reinhardt D, Dunican DS, Prendergast JG, Mjoseng H, Ramsahoye BH, Whitelaw E, et al. Redistribution of H3K27me3 upon DNA hypomethylation results in de-repression of Polycomb target genes. Genome Biol 2013; 14:R25.

142. Li Y, Zhang Y, Li S, Lu J, Chen J, Zhao Z, Bai J, Xu J, Li X. Genome-wide DNA methylome analysis reveals novel epigenetically dysregulated non-coding RNAs in human breast cancer. Sci Rep 2014; 1–12.

143. Jin Z, Zhang W, Yang H, Wang X, Zheng Y, Zhang Q, Zhi J. Maternal treatment with agonistic autoantibodies against type-1 angiotensin ii receptor in late pregnancy increases apoptosis of myocardial cells and myocardial susceptibility to ischemia-reperfusion injury in offspring rats. PLoS One 2013; 8:e80709.