Zfp57 inactivation illustrates the role of ICR methylation in imprinted gene expression during neural differentiation of mouse ESCs

Basilia Acurzio1,2,8, Ankit Verma1,2,8, Alessia Polito2,3,7, Carlo Giaccari1,2, Francesco Cecere1,2, Salvatore Fioriniello2, Floriana Della Ragione2, Annalisa Fico2, Flavia Cerrato1, Claudia Angelini4, Robert Feil5,6 & Andrea Riccio1,2*  

ZFP57 is required to maintain the germline-marked differential methylation at imprinting control regions (ICRs) in mouse embryonic stem cells (ESCs). Although DNA methylation has a key role in genomic imprinting, several imprinted genes are controlled by different mechanisms, and a comprehensive study of the relationship between DMR methylation and imprinted gene expression is lacking. To address the latter issue, we differentiated wild-type and Zfp57-/- hybrid mouse ESCs into neural precursor cells (NPCs) and evaluated allelic expression of imprinted genes. In mutant NPCs, we observed a reduction of allelic bias of all the 32 genes that were imprinted in wild-type cells, demonstrating that ZFP57-dependent methylation is required for maintaining or acquiring imprinted gene expression during differentiation. Analysis of expression levels showed that imprinted genes expressed from the non-methylated chromosome were generally up-regulated, and those expressed from the methylated chromosome were down-regulated in mutant cells. However, expression levels of several imprinted genes acquiring biallelic expression were not affected, suggesting the existence of compensatory mechanisms that control their RNA level. Since neural differentiation was partially impaired in Zfp57-mutant cells, this study also indicates that imprinted genes and/or non-imprinted ZFP57-target genes are required for proper neurogenesis in cultured ESCs.
DNA methylation. More recently, a non-canonical and H3K27me3-dependent form of imprinting has been demonstrated in the mouse.

The KRAB zinc finger protein ZFP57 has a critical role in maintaining DNA methylation at ICRs in both humans and mice. Homozygous mutations of ZFP57 have been reported in human individuals affected by type 1 transient neonatal diabetes mellitus (TNDM1), who present multi-locus imprinting disturbances. In the mouse, maternal-zygotic deletion of Zfp57 results in failure to maintain DNA methylation imprints, and causes mid-gestation embryonic lethality. ZFP57 recognizes the methylated hexameric motif “T/GGC CGC” within ICRs and recruits the corepressor KRAB-A-interacting protein (KAP1) together with DNA methyltransferases and histone H3K9 methyltransferases to these sites. In addition to ICRs, ZFP57 binds numerous non-imprinted genes and many transposable elements in mouse embryonic stem cells (ESCs). However, upon Zfp57 deletion, most or all ICRs lose their allelic DNA methylation and H3K9me3, while non-imprinted target loci lose H3K9me3 but maintain most of their DNA methylation, and repetitive elements are not apparently affected. Recently, the KRAB zinc finger protein ZNF445/ZFP445 was identified as a further important regulator of genomic imprinting. However, its role seems to be more relevant in humans, while in mice Zfp445 appears to be required for preserving methylation at only a subset of ICRs in the absence of Zfp57.

Loss of Zfp57 results in expression changes at only a few imprinted and non-imprinted genes in pluripotent ESCs. Allele-specific gene expression analysis in hybrid Zfp57−/− ESCs demonstrated that loss of Zfp57 causes derepression of the silenced allele of these imprinted loci. However, despite the fact that most ICRs displayed loss of methylation, only a few imprinted genes had their allele-specific expression status deregulated in Zfp57−/− ESCs. In fact, most imprinted genes have low RNA levels in pluripotent ESCs, but become activated in a lineage-specific manner during differentiation particularly during neurogenesis. In this study, we investigated the effect of Zfp57 inactivation upon differentiation of ESCs into neural precursor cells (NPCs). The results obtained highlight the complex relationship between ICR methylation maintained by ZFP57 and imprinted gene expression during neural differentiation.

**Results**

**Deregulation of ZFP57 target genes in Zfp57−/− ES cell-derived neural precursor cells.** It was previously shown that in vitro corticogenesis can recapitulate the imprinting identity of perinatal cortex, and yields the correct parent-of-origin-dependent expression and DNA methylation at imprinted gene loci. We reasoned, therefore, that this system could be a good model to test the consequences of Zfp57 inactivation on genomic imprinting. Neural differentiation was induced in pluripotent ESCs, following an established protocol with addition of the sonic hedgehog inhibitor cyclopoamine that generates cortical-identity neural progenitors (Fig. 1a). For this study, we used ESCs that were hybrid between the genetically divergent M. m. molossinus strain JF1 and C57BL6/J, so that at almost all imprinted genes the parental alleles could be told apart.

Wild-type (J1) ESCs and the previously obtained J1-derived Zfp57−/− ESCs were subjected to the differentiation protocol, and the expression of pluripotency and neural markers was investigated by quantitative RT-PCR and immunofluorescence (IF, Fig. 1b,c, Supplementary Fig. 1). Differentiation was stopped at day 12 to obtain neural precursor cells (NPCs), in which it has been reported that the expression of many imprinted genes is activated. The experiment was repeated several times, and down-regulation of the pluripotency markers Nanog and Pou5f1 was always observed in both wild-type and Zfp57−/− cells (Fig. 1b, Supplementary Fig. 1). In addition, the NPC-specific marker Nestin was increased from day 0 to day 12 in both wild-type and Zfp57−/− cells, although its expression was found lower in the mutant cells by quantitative RT-PCR (qRT-PCR, Fig. 1c). Furthermore, down-regulation of the pluripotency marker Pou5f1 in all the cells and activation of Nestin in most of them was demonstrated at day 12 in the Zfp57−/− mutant and wild-type strain by IF (Supplementary Fig. 1). Consistent with cell differentiation of both cell lines, there was also down-regulation of Zfp57, which was detected by qRT-PCR with the use of primers external to the deletion introduced during the knockout (Fig. 1b). Although these results demonstrate that after the 12 day-cyclopoamine treatment the Zfp57−/− mutant cells lost their pluripotency markers and acquired a NPC-specific marker, the more terminal marker Tubb3 was found activated only in the wild-type strain (Fig. 1c). Consistent with the differentiation protocol used (day 12), the glia-specific marker Gfap was not activated in either of the cell lines. Finally, some meso-endoderm-specific markers were activated with some differences in wild-type and mutant strains, suggesting that some of the cells also differentiated toward non-neural cell fates (Fig. 1d).

Concerning the imprinted genes, significant activation of Kcnq1ot1, H19 and Nnat was observed between day 0 and day 12 in both wild-type and mutant cells (Fig. 1e). A similar trend was observed for MEG3, Plagl1, Mest, Peg3 and Airn, although the changes of the individual genes did not reach statistical significance. Conversely, Cdkn1c, Dlk1, Igf2 and Zim1 were activated upon differentiation only in the wild-type cells (Fig. 1f). A further Zfp57 knockout clone, which was previously obtained in an inbred 129 ESC strain, was investigated by IF and qRT-PCR after the 12 day-cyclopaamine treatment. This cell strain demonstrated Pou5f1 down-regulation, Nestin activation and deregulation of imprinted gene expression similar to the Zfp57−/− J1 cells (Supplementary Fig. 2). In summary, these results indicate that the ESCs lacking Zfp57 are able to exit from pluripotency and partially differentiate into NPCs.

Two biological replicates of day 12-differentiated wild-type and Zfp57−/− IB1 cells were analyzed by RNA-seq. The results obtained were compared with the datasets from day 0-pluripotent wild-type and day 0-pluripotent Zfp57−/− IB1 cells (one replicate each) previously obtained in our laboratory, as well as with the datasets of day 0-pluripotent and day 12-differentiated wild-type IB1 and BJ1 cells (two replicates each) previously reported by Bouschet and collaborators (Fig. 2). Hierarchical clustering demonstrated that pluripotent and differentiated cells were separated, while wild-type and Zfp57−/− mutant cells grouped together, when either all genes or only imprinted genes were considered, indicating that gene expression of the wild-type and mutant strains can be compared (Fig. 2a,b). The analysis of the pluripotency and differentiation markers by RNA-seq confirmed the
results obtained by RT-PCR and IF (Fig. 2c). In particular, while ESC markers were generally down-regulated in both wild-type and mutant day 12-JB1 cells, Nestin but no other neural markers were activated in the Zfp57⁻/⁻ cells at a level similar to that of their control cells, or to previously reported hybrid NPCs. Nevertheless, activation of the endo-mesoderm markers Foxa2, Gata4 and Gata6 was similar in wild-type and mutant strains.
**Zfp57**

**Non-Imprinted**

Deregulated genes (padj<0.05)

Up

Down

Non-deregulated (padj>0.05)

All non-deregulated

**WT**

**day 12**

**JB1 log2**

(normalized read counts)

**Cluster Dendrogram**

**All genes**

**Zfp57**

**-/- cell lines**

**MARKERS**

**Z-score of normalized counts**

**ESCs/Pluripotency**

Markers

**Neural/Cortex**

**Endo/Mesoderm**

**Zfp57 expression**

**-log10(adj p-value)**

**Cluster Dendrogram**

**All genes**

**Imprinted genes**

**day 0**

**day 12**

**Distance from ZFP57/KAP1 peaks (bp)**

**Deregulated genes (padj<0.05)**

Up

Down

Non-Imprinted

Imprinted

Non-deregulated (padj>0.05)

All non-deregulated

**Nervous system development**

Neurogenesis

Generation of neurons

Neuron differentiation

Neuron development

Neural system development

Neuronal system development

Movement of cell or subcellular component

Anatomical structure development

Synapse organization

System development

Neuron projection development

Organic system organization

Multicellular system development

Cellular component organization

Asyn guidance

Neuron projection guidance

Cell morphogenesis involved in neuron differentiation

Anatomical structure morphogenesis

Cellular component organization or biogenesis
obtained from NPCs in an allele-specific manner. The mouse ESC lines used for neural differentiation derived **Gatm** genes with low-counts and genes located on the X chromosome (the JB1 ESCs are male), allele-specific expression with previous reports, imprinted ICR methylation was maintained in the wild-type NPCs26. Conversely, the ICR ing at the **Snrpn** differentiation was demonstrated by bisulfite conversion followed by pyrosequencing or direct Sanger sequenc-

**Zfp57**-mutant cells after inactivation24. Now, we investigated ICR methylation levels in wild-type and **Zfp57**-/- day 12-JB1 cells. Deregulated genes are indicated by red dots if downregulated (non-imprinted genes by light-red; imprinted genes by dark-red) and by blue dots if upregulated (non-imprinted genes by light-blue; imprinted genes by dark-blue). (e) Cumulative distribution of the distances (bp) of deregulated genes (imprinted genes: green; non-imprinted genes: orange) in **Zfp57**+/− day 12-JB1 cells from the ZFP57 target sites determined in wild-type day 0-JB1 cells24. The difference between the two curves is statistically significant according to the two-sample Kolmogorov–Smirnov test (p-value < 1e-10). (f,g) Gene ontology analysis of differentially expressed genes in **Zfp57**−/− day 12-JB1 cells (f, upregulated; g, downregulated). Bars represent GO:BP terms sorted by high to low -log10 (adjusted p-value). (h) Gene ontology analysis of down-regulated genes located < 100 Kb from ZFP57-KAP1 binding sites.

By using the DESeq2 software29, we identified 4912 differentially expressed genes in the **Zfp57**−/− day 12-JB1 cells with respect to the day 12-JB1 wild-type cells. These included both up-regulated and down-regulated genes in similar proportions (Fig. 2d, Supplementary Table 2). 50 of these genes were reported to be imprinted3, 4, indicating that the differentially expressed genes were enriched for imprinted genes (p-value < 1e-04).

We previously showed that loss of **Zfp57** causes profound changes in DNA methylation and histone marks at its target sites in pluripotent ESCs24. To determine if the inheritance of these epigenetic modifications may have directly affected gene expression in NPCs, we calculated the distance of deregulated genes from previously mapped ZFP57 binding sites in ESCs24. We found that deregulated imprinted genes were more frequently within 1000 kb from ZFP57 targets than deregulated non-imprinted genes (Fig. 2e). While imprinted gene expression was mostly altered by defective ZFP57 binding to ICRs at the imprinted loci themselves, many deregulated non-imprinted genes were located far from ZFP57 sites and were likely indirectly affected.

We then examined the identity and functional classification of the deregulated genes by gene ontology analysis. While up-regulated genes were mostly linked to intracellular functions and metabolic processes, down-regulated genes were prevalently involved in neurogenesis and neuron development pathways (Fig. 2g). Down-regulated genes involved in neuronal and developmental functions included 6 imprinted genes and 68 non-imprinted genes that were located within 100 kb from the ZFP57 binding sites (Fig. 2h, Table 1 and Supplementary Table 3). Significantly, 24 of these non-imprinted genes were overlapping ZFP57 binding sites. This suggests that in addition to disrupting the expression of important imprinted genes, loss of ZFP57 binding may directly alter the expression of a group of non-imprinted genes that controls neuronal differentiation of ESCs in culture.

Overall, these results demonstrate that when exposed to the differentiation conditions, **Zfp57**−/− ESCs exit from the pluripotency state and up-regulate some differentiation markers, but fail to fully activate neural mark-

**Massive deregulation of allele-specific expression of imprinted genes in **Zfp57**−/− cells after differentiation.** We previously demonstrated that pluripotent ESCs lose ICR methylation upon **Zfp57** inactivation27. Now, we investigated IRC methylation levels in wild-type and **Zfp57**−/− day 12 cells, to see if any change had occurred during differentiation of pluripotent ESCs. Methylation of 10 ICRs (7 maternally methyl-

**Zfp57**−/− day 12-JB1 versus wild-type day 0-JB1 cells (Fig. 2h, Table 1 and Supplementary Table 3). Significantly, 24 of these non-imprinted genes were overlapping ZFP57 binding sites. This suggests that in addition to disrupting the expression of important imprinted genes, loss of ZFP57 binding may directly alter the expression of a group of non-imprinted genes that controls neuronal differentiation of ESCs in culture.

To determine the consequences of **Zfp57** loss on imprinted gene expression, we analysed the RNA-Seq data obtained from NPCCs in an allele-specific manner. The mouse ESC lines used for neural differentiation derived from inter-sub-species (JF1 × C57BL/6)F1 hybrid blastocysts. The genetic divergence between the C57BL/6 and JF1 inbred mouse strains (> 4 SNPs/kb) allowed us to distinguish the parental origin of most imprinted genes. Only for a few protein-coding genes, including **Gatm**, **H13**, **Mcts2**, **Zim2**, and for the **SnoI115-6** cluster and many miRNAs, allele-specific expression could not be measured because of absence of suitable SNPs. After removal of genes with low-counts and genes located on the X chromosome (the J1 ESCs are male), allele-specific expression
| Gene* | Log2 fold change (Zfp57−/- vs WT) | Distance from ZFP57 peak (bp) | Functional categoryb |
|-------|---------------------------------|-------------------------------|----------------------|
| Ptpn5 | −5.650198389                   | 24,245                        | 1,2,3,4,5,6,8,9,10,14 |
| Hex5  | −5.178415842                   | 9979                          | 1,2,3,4,5,6,8,9,10,14,15,19 |
| Otx1  | −5.163770188                   | 3925                          | 3,6,8,10,14,15,19    |
| Pclh  | −3.996723886                   | 77,959                        | 6,8,10,14            |
| Dner  | −3.84008981                    | 0                             | 1,3,6,8,10,14,15     |
| Tron67 | −3.77993524                   | 8591                          | 1,2,3,4,5,6,8,9,10,14 |
| Gm2   | −3.332414676                   | 0                             | 6,8,10,14            |
| Lama2  | −3.187941857                   | 44,022                        | 1,2,3,4,5,6,8,9,10,11,12,13,14,16,17,18 |
| Solp  | −3.001779515                   | 23,142                        | 6,8,10,14            |
| Megf11 | −2.98280188                    | 0                             | 6,8,10,14            |
| Tub6  | −2.961888079                   | 4595                          | 1,2,3,4,5,6,8,9,10,11,12,13,14,16,17,18 |
| Stx1b | −2.953214561                   | 43,156                        | 1,2,3,4,5,6,8,9,10,14 |
| Unc5a | −2.861952413                   | 7113                          | 1,2,3,4,5,6,8,9,10,11,12,13,14,16,17,18 |
| Igf2  | −2.800585832                   | 68,640                        | 6,8,10,14            |
| Sprr1 | −2.622871614                   | 16,805                        | 6,7,14              |
| Cacna2d2 | −2.593159777                  | 60,968                        | 6,7,14              |
| Mapk8p2 | −2.581498233                  | 78,444                        | 1,2,3,4,5,6,8,9,10,13,14,17 |
| Tubb3 | −2.563740007                   | 0                             | 6,8,10,14            |
| Tubb2a | −2.557939166                   | 32,318                        | 1,2,3,4,5,6,7,8,9,10,13,14,15,16,17,18,19 |
| Akt3  | −2.447396122                   | 39,078                        | 3,6,8,10,14,15,19  |
| Prkcb | −2.447307746                   | 0                             | 6,8,10,14            |
| Tubb2b | −2.447307746                   | 88,398                        | 1,2,3,4,5,6,8,10,14 |
| Slit1  | −2.447307746                   | 0                             | 6,8,10,14            |
| Neff  | −2.447307746                   | 32,318                        | 1,2,3,4,5,6,8,9,10,11,12,13,14,16,17,18,19 |
| Ank3  | −2.377539166                   | 0                             | 1,2,3,4,5,6,8,9,10,11,12,13,14,16,17,18,19 |
| Adgrv1 | −2.367340007                   | 17,027                        | 3,6,8,10,14         |
| Lama4  | −2.171209218                   | 30,669                        | 6,8,10,14            |
| Fzd9  | −2.10585832                    | 0                             | 6,8,10,14            |
| Fzd9  | −2.10585832                    | 0                             | 6,8,10,14            |
| Bbs1  | −2.10585832                    | 0                             | 6,8,10,14            |
| Fgd4  | −2.10585832                    | 0                             | 6,8,10,14            |
| Hnrnph3 | −2.10585832                    | 0                             | 6,8,10,14            |
| Prkca | −2.10585832                    | 0                             | 6,8,10,14            |
| Plagl1 | −2.10585832                    | 0                             | 6,8,10,14            |
| Meis3 | −2.10585832                    | 0                             | 6,8,10,14            |
| Apbb1 | −2.10585832                    | 0                             | 6,8,10,14            |
| Shank3 | −2.10585832                    | 0                             | 6,8,10,14            |
| H113  | −2.10585832                    | 0                             | 6,8,10,14            |

Continued
Table 4. By following this approach, 15 imprinted genes demonstrated significant parent-of-origin-dependent
allelic expression in the wild-type day 0-JB1 cells and 32 in the day 12-JB1 cells (Fig. 3 and Supplementary
Table 5). Among the NPC imprinted genes, 9 were predominantly expressed from the maternal allele, and 23
were imprinted in the NPCs included genes that were either not imprinted in ESCs (e.g. Peg13, Peg3,
Plagl1, Meg3, Zfp57, and Mirg) maintained a significant allelic bias in the mutant ESCs. The
genes, which were not imprinted in wild-type and mutant ESCs, acquired imprinting in the wild-type NPCs, but not
in the mutant NPLCs.

| Gene  | Log2 fold change (Zfp57−/− vs WT) | Distance from ZFP57 peak (bp) | Functional categorya |
|-------|---------------------------------|-----------------------------|---------------------|
| Lmx1l | −1.003120469 | 97,408                     | 1,2,3,4,5,6,8,9,10,13,14,16,17,18 |
| Pam   | −0.9827988076 | 53,726                    | 3,6,8,10,14         |
| Naglu | −0.9714420126 | 95,071                     | 1,2,3,4,5,6,8,10,14,15,19 |
| Arhgef7 | −0.9214359692 | 0                         | 1,3,6,7,8,10,14     |
| Fam171a1 | −0.882925793 | 0                         | 6,14               |
| Art3  | −0.865781179 | 15,058                     | 1,2,3,4,5,6,8,10,14 |
| Usp5  | −0.8508211011 | 17,582                     | 6,8,10,14          |
| Ank   | −0.8178055036 | 0                         | 6,8,10,14          |
| Kdm1b | −0.8026958823 | 32,233                     | 6,10,14            |
| Rats6a | −0.7545602154 | 0                         | 6,8,10,14          |
| Bopp5f | −0.7516626695 | 0                         | 1,2,3,4,5,6,8,9,10,14,16 |
| Exoc5 | −0.7432131534 | 60,611                     | 6,14               |
| Phc2  | −0.6955736462 | 68,910                     | 6,10,14            |
| Pdez3n | −0.6821863965 | 0                         | 7                  |
| Alg10b | −0.6296632078 | 22,645                     | 6,10,14            |
| Gpi1  | −0.6184329932 | 22,645                     | 6,10,14            |
| Tri6  | −0.5863515137 | 0                         | 1,2,3,4,5,6,8,9,10,11,12,13,14,15,16,17,18 |
| Ptpg  | −0.5802367554 | 1086                      | 1,2,3,4,5,6,8,9,10,14,15,19 |

Table 1. Zfp57−/− day 12-JB1 cells down-regulated genes located < 100 kb from ZFP57/KAP1 binding
sites and involved in neuronal and developmental functions. *Imprinted genes are in bold. **GO
Category are as follows: 1. GO:0022008(neurogenesis), 2. GO:0048666(neuron development), 3.
GO:0007399(nervous system development), 4. GO:0048699(generation of neurons), 5. GO:0030182(neuron
differentiation), 6. GO:0048856(anatomical structure development), 7. GO:0050808(synapse
organization), 8. GO:0048731(system development), 9. GO:0031175(neuron projection development),
10. GO:0007275(multicellular organism development), 11. GO:0007409(axonogenesis), 12.
GO:0097485(neuron projection guidance), 13. GO:0048667(cell morphogenesis involved in neuron
differentiation), 14. GO:0032502(developmental process), 15. GO:0007417(cenral nervous system
development), 16. GO:0061564(axon development), 17. GO:0048812(neuron projection morphogenesis), 18.
GO:0007409(axonogenesis), 19. GO:0007420(brain development).

was determined by calculating the mat/(mat + pat) allelic ratio for each gene in the RNA-seq datasets. The genes with an average allelic ratio within the interval 0–0.33 were arbitrarily considered paternally expressed, and the genes with an average allelic ratio in the interval 0.67–1 were considered maternally expressed. In our previous study on pluripotent ESCs, we identified only a few genes with imprinted expression4. We have now re-evaluated the RNA-seq dataset of pluripotent ESCs reported in ref.24 together with the new NPC dataset by using SNPspilt,
an algorithm which efficiently splits the reads between genomes with known SNP genotypes (Supplementary Table 4)30. By following this approach, 15 imprinted genes demonstrated significant parent-of-origin-dependent allelic expression in the wild-type day 0-JB1 cells and 32 in the day 12-JB1 cells (Fig. 3 and Supplementary Table 5). Among the NPC imprinted genes, 9 were predominantly expressed from the maternal allele, and 23 from the paternal allele, consistent with what was previously demonstrated in mouse tissues4. The genes that were imprinted in the NPCs included genes that were either not imprinted in ESCs (e.g. Zfp57, Plac9, Peg13 and Impact) or whose expression in ESCs was too weak to assess allele-specificity (Supplementary Table 5). With a few exceptions, these results are consistent with those described in the study of Bouschet and collaborators48.

We next analyzed the allele-specific expression of the imprinted genes in the Zfp57−/− mutant JB1 cells at day 0 and day 12 of cyclopamine treatment. We found that all the genes (but Nespa in ESCs) that were imprinted in wild-type cells, reduced their allelic expression bias in the Zfp57−/− cells (Fig. 3, Supplementary Fig. 4 and Supplementary Table 5). However, the shift toward equivalent expression of the maternal and paternal loci was more dramatic in the mutant cells at day 12 than at day 0. Notably, several genes (e.g. Nespa, Peg3, Plag1, Meg3, Rian and Mirg) maintained a significant allelic bias in the mutant ESCs. The Zfp57−/−, Peg13 and Impact genes, which were not imprinted in wild-type and mutant ESCs, acquired imprinting in the wild-type NPCs, but not in the mutant NPLCs.

Loss of allelic bias is associated with different expression patterns of imprinted genes in Zfp57−/− NPCs. To better understand the consequence of ZFP57 loss on genomic imprinting in neural differentiation, we integrated the results of gene expression levels with the allelic ratios determined in the Zfp57−/− day 12-JB1 cells. An overall complex relationship between imprinting status and gene expression level was observed (Table 2). For several genes, the lack of biased allelic expression corresponded to global up-regulation of their RNA level, for others loss of imprinting was associated with down-regulation of their RNA levels. For the remaining ones, the loss of allele-specificity was not accompanied by a corresponding change in global expression levels.
The effect of ZFP57 deficiency on imprinted gene expression is better understood if each affected imprinted domain is described separately (Fig. 4 and Supplementary Fig. 5). On chromosome 1, the imprinted domain including the DBF-type zinc finger-containing protein 2 gene (*Zdbf2*) is controlled by a maternal germline DMR that is located within the *Gpr1* gene. This DMR regulates the paternal-specific expression of long isoforms of *Zdbf2* (*Liz*), which in turn activates the canonical *Zdbf2* promoter on the paternal chromosome during embryonic development. ZFP57 binds the *Gpr1* DMR on its methylated maternal allele, which loses its methylation in *Zfp57*-/- ESCs. Paternal-specific expression of *Zdbf2* was demonstrated in the wild-type day 12-cells, while this gene was activated twofold and imprinted expression was not acquired in the *Zfp57*-/- cells cultured in the same conditions (Supplementary Fig. 5a). Another gene, *Adam23*, reported to be preferentially expressed from its paternal allele in mouse brain, is located 100 kb from *Zdbf2*, in the same transcription orientation. We found *Adam23* biallelically expressed in both the wild-type and *Zfp57*-/- cells, but observed a twofold activation of the

**Figure 3.** Allele-specific expression of imprinted genes in the wild-type and *Zfp57*-/- day 0 and day 12-JB1 cells. Dot plot representing the allelic expression of imprinted genes in wild-type and *Zfp57*-/- day 0- (green and red circles, respectively) and wild-type and *Zfp57*-/- day 12- (green and red triangles, respectively) JB1 cells. The blue-shaded area corresponds to paternal (B6)-specific expression (allelic ratio interval: 0–0.33), the pink-shaded area corresponds to maternal (JF1)-specific expression (allelic ratio interval: 0.67–1), the white area corresponds to biallelically expressed genes (allelic ratio > 0.33 or < 0.67). The allelic ratio is the average of two biological duplicates (for details, see Supplementary Table 5). The data of wild-type and *Zfp57*-/- ESCs are derived from a previous study.

| Imprinted Genes | WT day 0 JB1 | WT day 12 JB1 | Zfp57/- day 0 JB1 | Zfp57/- day 12 JB1 |
|----------------|-------------|-------------|-----------------|-----------------|
|                 | Green circle | Red circle  | Green triangle  | Red triangle    |

The effect of ZFP57 deficiency on imprinted gene expression is better understood if each affected imprinted domain is described separately (Fig. 4 and Supplementary Fig. 5). On chromosome 1, the imprinted domain including the DBF-type zinc finger-containing protein 2 gene (*Zdbf2*) is controlled by a maternal germline DMR that is located within the *Gpr1* gene. This DMR regulates the paternal-specific expression of long isoforms of *Zdbf2* (*Liz*), which in turn activates the canonical *Zdbf2* promoter on the paternal chromosome during embryonic development. ZFP57 binds the *Gpr1* DMR on its methylated maternal allele, which loses its methylation in *Zfp57*-/- ESCs. Paternal-specific expression of *Zdbf2* was demonstrated in the wild-type day 12-cells, while this gene was activated twofold and imprinted expression was not acquired in the *Zfp57*-/- cells cultured in the same conditions (Supplementary Fig. 5a). Another gene, *Adam23*, reported to be preferentially expressed from its paternal allele in mouse brain, is located 100 kb from *Zdbf2*, in the same transcription orientation. We found *Adam23* biallelically expressed in both the wild-type and *Zfp57*-/- cells, but observed a twofold activation of the
global expression of this gene in the mutant cells, suggesting a coordinated regulation of this imprinted domain (Supplementary Fig. 5a).

In the Blcap/Nnat domain, a germline DMR overlaps the Nnat promoter that in turn is located in a Blcap intron. ZFP57 interacts with this region in mouse ESCs, confirming its identity as ICR. We found paternal-specific expression of Nnat in the wild-type but imprinting was not acquired by this gene in the Zfp57-mutant day 12-cells, demonstrating a direct role of ZFP57-dependent methylation in the imprinting control of this locus (Supplementary Fig. 5b). However, differently from Zdbf2, the global mRNA expression level of Nnat was unchanged in the Zfp57-mutant cells. Blcap appears slightly maternally expressed in the wild-type day 12-cells, but its allelic ratio does not fall in the fixed intervals of imprinted expression, possibly because of the presence of non-imprinted isoforms. However, the almost twofold down-regulation of this gene in the Zfp57-mutant day

| Domain | Chromosome | ICR methylation | Gene | Allelic expression wild-type NPCs | Allelic expression Zfp57−/− NPCs | Global expression change |
|--------|------------|-----------------|------|----------------------------------|----------------------------------|-------------------------|
| Gpr1   | chr1       | Maternal        | Zdbf2| Paternal                         | Biallelic                        | Up-regulated            |
| Nnat   | chr2       | Maternal        | Nnat | Paternal                         | Biallelic                        | Up-regulated            |
| Gnas   | chr2       | Maternal        | Nespas| Paternal                         | Biallelic                        | Up-regulated            |
| Peg10  | chr6       | Maternal        | Sge  | Paternal                         | Biallelic                        | Up-regulated            |
| Peg3   | chr7       | Maternal        | Zim1 | Maternal                         | All. bias reduced                | Down-regulated          |
| Surpr  | chr7       | Maternal        | Subg14| Paternal                         | Biallelic                        | Unchanged               |
| H19    | chr7       | Paternal        | H19  | Paternal                         | Biallelic                        | Up-regulated            |
| Kcnq1ot1| chr7      | Maternal        | Th   | Maternal                         | Biallelic                        | Down-regulated          |
| Plag1  | chr10      | Maternal        | Plag1| Paternal                         | Biallelic                        | Down-regulated          |
| Grb10  | chr11      | Maternal        | Grb10| Paternal                         | Biallelic                        | Down-regulated          |
| Commd1 | chr11      | Maternal        | Zrsr1| Paternal                         | Biallelic                        | Up-regulated            |
| Meg3   | chr12      | Paternal        | Dlk1 | Paternal                         | Biallelic                        | Down-regulated          |
| Trappc9| chr15      | Maternal        | Kcn9 | Pref. maternal                  | Biallelic                        | Down-regulated          |
| Ig2r   | chr17      | Maternal        | Ig2r | Pref. maternal                  | All. bias inverted               | Down-regulated          |
| Impact | chr18      | Maternal        | Impact| Paternal                        | Biallelic                        | Up-regulated            |

Table 2. Integration of global gene expression with allele-specific expression of imprinted genes in Zfp57−/− day 12-JB1 cells. *Not statistically significant.
Figure 4. Coordinated change of allelic and levels of expression at imprinted gene clusters in the Zfp57-/- day 12 cells. Panels (a–h) represent 8 different imprinted gene domains. A schematic of the gene cluster with indication of the imprinting status in wild-type and Zfp57-/- day 12-JB1 cells, and location of the ZFP57 binding site (ZFP57 BS) is shown at the top of each panel. In each panel, the box-plot in the lower left shows the allelic expression ratio of the genes of the domain in the wild type (green) and Zfp57-/- (red) day 12-JB1 cells. The dot-plot in the lower right shows the bulk gene expression changes detected in Zfp57-/- day 12-JB1 cells (Zfp57-/- day 12 / (wild-type day 12 + Zfp57-/- day 12)). All values correspond to the average of two biological duplicates (for details, see Supplementary Tables 2, 4 and 5).
12-cells confirms the hypothesis that Blec1 imprinting is co-regulated with Nnat by transcriptional interference (Supplementary Fig. 5b).

Imprinting of the Gnas domain is controlled by the paternally expressed lncRNA Nespas35, Nespas downregulates its sense counterpart Nesp, which in turn controls the reciprocal gamete of origin-specific expression of Gnas and GnasXL. The Nespas promoter overlaps a germline DMR that is maternally methylated and bound by ZFp57 in mouse ESCs36. We found Nespas expressed from its paternal allele in the wild-type but activated on both parental alleles with an overall twofold increase of its mRNA level in the Zfp57-mutant day 12-cells, consistent with ZFp57-dependent imprinting regulation (Supplementary Fig. 5c). Unfortunately, Nesp expression was too low to determine its allelism in NPCs, and assess the regulatory role of ZFp57-dependent methylation on this gene.

A ZFp57-marked maternally methylated germline DMR overlaps the promoters of Sgce and Peg10, on proximal chromosome 636. Consistent with Zp57-dependent imprinting control of these two genes, we observed their paternal-specific expression in wild-type and bi-allelic expression in Zfp57-mutant day 12-cells (Fig. 4a). The change in global mRNA levels of these two genes, however, was not consistent: Peg10 was up-regulated twofold, while Sgce displayed similar levels in wild-type and mutant cells. Another imprinted gene, Asb4, lies 700 kb distal from Sgce/Peg10. We found that Asb4 was preferentially expressed from the maternal allele in the wild-type and lost its allelic bias in the mutant day 12-cells, although its global mRNA level was unchanged (Fig. 4a). Since the Sgce/Peg10 DMR is the closest ZFp57 binding site in mouse ESCs36, this finding is consistent with the regulation of Asb4 by this ICR. Of note, two further imprinted genes of this cluster that are highly expressed in neural cells, namely Casd1 and Ppp1r9a37, were bi-allelically expressed in wild-type but down-regulated in the Zfp57-mutant day 12-cells. This suggests co-regulation of these genes with the rest of the cluster and is consistent with the incomplete neural differentiation of the Zfp57-/- cells.

A further maternally methylated germline DMR is present on chromosome 6 and overlaps the promoter of Peg3 and Usp29 genes (Fig. 4b)34. Consistent with their imprinting being regulated by this DMR, Peg3 and Usp29 were both paternally expressed in the wild-type but bi-allelically expressed in the Zfp57-mutant day 12-cells (Supplementary Fig. 5d). We also observed an almost twofold up-regulation of Peg1/Mest in mutant cells. Although we did not detect any allelic bias for Cog2, this gene was down-regulated in Zfp57-mutant cells, consistent with the hypothesis that its expression is controlled by the overlapping anti-sense MestXL38. A further chromosome 6 gene, Ctnm3, reported as imprinted in placenta but not in brain where it is highly expressed39, was bi-allelically expressed in the wild-type but strongly down-regulated in the Zfp57-mutant day 12-cells. This gene is not located close to an imprinted domain, but a ZFp57 binding site is located 1 Mb from its transcription binding site34. Its repression may be caused by loss of ZFp57 binding at this site, although it could also be a consequence of the incomplete neural differentiation of the Zfp57-/- cells.

On proximal chromosome 7, a ZFp57-bound DMR overlaps the promoters of the divergently transcribed Asb4 and Sgce genes (Fig. 4b)34. Consistent with their imprinting being regulated by this DMR, Peg3 and Usp29 were both paternally expressed in the wild-type but bi-allelically expressed in the Zfp57-mutant day 12-cells. However, at the global level, Peg3 was activated twofold while Usp29 expression remained unchanged. Within the same domain, the maternally expressed Zim1 also reduced its allelic bias, but its expression was strongly down-regulated in the mutant day 12-cells, consistent with the hypothesis that Peg3 is a repressor of Zim140.

In the central part of chromosome 7, another ZFp57-bound DMR overlaps the promoter of Snurp/Snrpn. This gene encodes large alternatively spliced transcripts, including the 460 kb IncRNA Snlgl14, which were paternally expressed in the wild-type but were bi-allelic in the Zfp57-mutant day 12-cells (Fig. 4c). This cluster includes several other paternally expressed genes (A230057D06Rik, Ndn, Mage2, Mkn3 and Peg12) that all lost or reduced their allelic bias in Zfp57-mutant cells, indicating that they are coordinately controlled by methylation of the above mentioned DMR. Concerning their global RNA level, only Snrpn was up-regulated almost twofold, while the others were either unchanged or down-regulated.

On distal chromosome 7, two ZFp57-bound DMRs mark two adjacent imprinted domains. The more centromeric domain includes the H19 and Igf2 genes. These both displayed imprinted expression in the wild-type and lost or reduced, respectively, their allelic bias in the Zfp57-mutant day 12-cells (Fig. 4d). However, the paternally expressed H19 was activated and Igf2 repressed, consistent with the model of the paternally methylated insulator corresponding with the ZFp57-bound DMR41. Interestingly, the Igf2 antisense transcript Igf2os was down-regulated similar to Igf2, indicating that it is controlled through a similar mechanism. The more distal ZFp57-bound DMR overlaps the promoter of the antisense noncoding gene Kcnq1ot1, which was found paternally expressed in the wild-type and bi-allelically expressed in the mutant day 12-cells, confirming its dependence from DMR methylation (Fig. 4e). The global level of the Kcnq1ot1 RNA was unchanged. Consistent with the model predicting the regulation of this domain through the cis-repressive function of Kcnq1ot1, the closely located Cdkn1c gene was down-regulated and its maternal expression bias reduced. Igh was also repressed but its allelic bias was borderline (adj. p-value = 0.0493). Another gene, Anol1, which lies 1 Mb from the Kcnq1ot1 DMR and was reported to be maternally expressed in placenta42, was found down-regulated in the Zfp57-mutant day 12-cells and therefore possibly coordinately regulated with the Kcnq1ot1 domain, although no allelic expression bias was detected in the wild-type NPCs.

A further ZFp57-bound DMR overlaps with the Plagl1 promoter on chromosome 1043. Consistent with regulation by this maternally methylated DMR, the Plagl1 gene was found paternally expressed in the wild-type and bi-allelically expressed in the Zfp57-mutant day 12-cells (Fig. 4f). At the global level, the Plagl1 mRNA was down-regulated in the mutant cells. The overlapping noncoding RNA Hymai is not currently mapped by UCSC, but according to the enrichment of the reads corresponding to its location (Plagl1 intron 1), it appears to have the same behavior as Plagl1. The imprinting status of the close-by Phactr2 gene is controversial44,45. However, we observed that Phactr2 was preferentially expressed from the maternal allele in the wild-type cells, and its allelic bias was abolished in the mutant cells, suggesting that its imprinting is co-regulated with Plagl1, although its global expression level was similar in the wild-type and mutant cells.
Chromosome 11 harbours two maternally methylated ZFp57-bound DMRs. The proximal one overlaps a CTCF binding site and an alternative promoter of the Grb10 gene. It has been proposed that DNA methylation interferes with the enhancer-blocking function of this DMR thereby allowing activation of the main Grb10 promoter on the maternal chromosome. Consistent with this hypothesis, we found Grb10 expressed from the maternal allele in the wild-type and from both parental alleles in the Zfp57-/- mutant day 12-cells (Supplementary Fig. 5f). However, loss of imprinting was accompanied by down-regulation of its global mRNA level. The Ddc gene is located downstream of Grb10. Ddc showed a paternal expression bias in the wild-type day 12-cells, which is consistent with previous reports, but its bias was not strong enough to be classified as imprinted under our stringent criteria. However, its allelic expression was inverted and its global expression activated in the Zfp57-/- day 12-cells, indicating that this gene may also be regulated by the Grb10 methylation-sensitive insulator. The distal chromosome 11 DMR overlaps the promoter of the Zrsr1 gene. Zrsr1 was expressed from its paternal allele in the wild-type and up-regulated twofold on both parental alleles in the Zfp57-mutant day 12-cells, consistent with its regulation from its maternally methylated DMR (Supplementary Fig. 5e). We did not find any allelic expression bias of the overlapping Commd1 gene in NPCs.

In the large cluster of imprinted genes on chromosome 12, Zfp57 binds a paternally methylated DMR that is located between the Dlk1 and Meg3 genes. We found Dlk1 paternally expressed and Meg3 maternally expressed in the wild-type but these genes were expressed from both parental alleles in the Zfp57-mutant day 12-cells, consistent with the model predicting imprinting dependent on the intergenic DMR methylation (Fig. 4g). In addition, Dlk1 was down-regulated, consistent with the repressor function of the IncRNA Meg3. A number of other genes (including Mirt and Rian) that are co-transcribed with Meg3 within a polycistronic RNA drastically reduced their allelic bias (Supplementary Fig. 6a). Although the global RNA level of Meg3 was only slightly changed, those of the small nucleolar RNA-containing gene Rian and the micro RNA-containing gene Mirt were up-regulated two- and fourfold, respectively, in the Zfp57-/- mutant day 12-cells.

On chromosome 15, a ZFp57-bound DMR overlaps the promoter of the IncRNA gene Peg13. This gene was paternally expressed in the wild-type and biallelically expressed in the Zfp57-mutant day 12-cells, consistent with being regulated by maternal methylation of this DMR, although its global expression was unchanged (Fig. 4h). The allelic bias of Trappc9, which overlaps Peg13, and that of Kenk9 located at its 3’ were not statistically significant in the wild-type but both of these genes were down-regulated in the Zfp57-mutant day 12-cells, consistent with their co-regulation with Peg13.

On chromosome 17, the promoter of the IncRNA Airn overlaps a maternally methylated and ZFp57-bound DMR. Consistent with the model predicting transcriptional interference between Airn and Igf2r, the former gene was activated and the latter repressed in the Zfp57-mutant day 12-cells (Supplementary Fig. 5g). Igf2r appears slightly paternally expressed in the wild-type NPCs but its allelic ratio does not fall in the fixed intervals of imprinted expression, while Airn appears paternally expressed, but its counts are too low to perform a statistical test. However, the symmetrical changes of their allelic ratios observed in the Zfp57-mutant day 12-cells is consistent with their reciprocal imprinting being regulated by Zfp57-dependent DMR methylation.

Finally, on chromosome 18, ZFp57 marks the maternally methylated DMR overlapping the promoter of the Impact gene. The imprinting mechanism controlling Impact is poorly investigated. We found that this gene is paternally expressed in the wild-type and that the allelic bias is reduced in the mutant day 12-cells, with a global upregulation of about fourfold. These findings suggest that its imprinting is controlled by the ZFp57-bound DMR (Supplementary Fig. 5h).

In conclusion, all the imprinted genes detected in wild-type NPCs lost or reduced their allelic expression bias in the Zfp57-/- NPCs. However, those expressed from the non-methylated chromosome were generally up-regulated, and those expressed from the methylated chromosome were generally down-regulated. In addition, several genes (such as Sgc, Asb4, Usp29, Snh1g14, Ndn, Magel2, Kcnq1ot1, Phactr2, Meg3 and Peg13), whose expression was expected to be increased upon loss of ICR methylation, acquired biallelic expression but overall were not significantly activated.

Discussion

Differential DNA methylation of ICRs is the best characterized mechanism by which imprinted gene expression is maintained in somatic cells. However, the relationship between ICR methylation and gene expression is not well understood for all imprinted genes. Zfp57-/- ESCs display a specific DNA methylation defect at the 20 ICRs that are bound by ZFp57 in mouse pluripotent cells, and represent an ideal model to study the role of methylation in the control of imprinted gene expression during differentiation. By employing an in vitro system of neural differentiation of hybrid ESCs, this study highlights the importance of ZFp57-dependent methylation in genomic imprinting, but also illustrates that imprinted gene control is more complex than what previously thought.

An unexpected finding of this study was that although the Zfp57-/- cells were able to leave the pluripotency state they only partially differentiated into NPCs if compared to their derivative wild-type cells, under the conditions used in this study. We cannot exclude that the deregulated expression of some imprinted genes is secondary to the differentiation differences observed between the wild-type and mutant strains. Nevertheless, the general loss of allelic expression and the coordinated expression changes of the genes located in the same domain suggest that most of the imprinting loss is directly caused by lack of ZFp57-dependent ICR methylation.

Despite the presence of differential methylation at ICRs, allele-specific expression was well established at only a few genes in the undifferentiated ESCs. Some genes acquired imprint expression after differentiation, others increased their allelic bias in NPCs. For several other genes, imprinting could be assessed only in NPCs, because their expression in ESCs was too low. Nevertheless, all the imprinted genes of wild-type NPCs, overall belonging to 15 of the 20 known imprinted gene clusters, lacked or reduced their gamete of origin-specific expression in Zfp57-mutant NPCs. This demonstrates that Zfp57-dependent methylation is generally required.
for both acquisition and maintenance of imprinted gene expression in somatic cells. Interestingly, some genes, such as Nespas, Peg3, Plagl1, Meg3, Rian and Mิง, partially retained their allelic bias in the Zfp57-mutant day 0-cells, but completely lost it in the Zfp57-mutant day 12-cells. Since both the Zfp57-mutant day 0 and Zfp57-mutant day 12-cells completely lack ICR methylation, this suggests that Zfp57-dependent methylation is the major determinant of imprinted gene expression in differentiated cells, but additional epigenetic mechanisms contribute to imprinting maintenance in pluripotent cells.

Lack of Zfp57 leads to a more limited ICR methylation loss in mouse embryos than in cultured cells. It is likely because the ESCs and the derived early differentiating cells undergo many more cell divisions than the equivalent cells in the early embryo, and there can thus be accumulation of small losses each cell cycle, eventually leading to fixation of the unmethylated state. So, an advantage of our ESC system is that it allows to score regions at which ZFP57 contributes only partly to the maintenance of methylation. The consequence of Zfp57-knockout on imprinted gene expression was investigated in mouse embryos in a very recent study.\textsuperscript{47} The data of this study are consistent with ours, but are limited by an incomplete loss of ICR methylation of Zfp57-null embryos. Differently from our neural cell-based system, this in vivo study analyzed whole E13.5 embryos, which did therefore not allow detection of allelic expression of several tissue-specific imprinted genes. It is important to note, however, that because the Zfp57 gene was inactivated prior to differentiation, none of these studies addresses the possible role of this gene in differentiated cells. In humans, ZFP57 inactivation results in a less severe phenotype than in mice, possibly because ZFPS7 is not expressed in human oocytes and is activated only post-implantation, and another zinc-finger protein gene, ZNF445, appears to play a major role instead.\textsuperscript{15,16}

Among the imprinted genes that were deregulated in the Zfp57-mutant NPLCs, some displayed up-regulation of the silenced allele, others down-regulation of the expressed allele, with respect to the wild-type cells. This behavior is consistent with the relationship between DMR methylation and allele-specific expression of each imprinted gene. In general, the gene in cis with DMR methylation was silenced in the parental parental allele, in mutant cells. Thus, maternally expressed genes were repressed and paternally expressed genes activated in the domains with maternal ICR methylation. Conversely, maternally expressed genes were activated and paternally expressed genes repressed in the domains with paternal ICR methylation.

Intriguingly, several imprinted genes that were expected to be activated twofold—because now transcribed from both the parental alleles—did not significantly change their global expression level in Zfp57-mutant cells (Table 2). This suggests that compensatory mechanisms exist for many imprinted genes that finely control their RNA level. Most of these genes, interestingly, are expressed at a lower level in wild-type NPCs. Since their overall expression levels were not affected by expressing two alleles versus one, it may also be speculated that these genes are not the ones that have driven the evolution of imprinted gene expression. In the context of possible dosage compensation mechanisms, it is interesting to note that multiple regulatory links exist between different imprinted domains.\textsuperscript{48} For instance, the maternally expressed miRNAs (miR-379/410) encoded by the Dlk1-Meg3 imprinting domain paternally expressed transcripts from other imprinted domains, including Plagl1, Peg3, Igf2 and Mrkrn3.\textsuperscript{35,50} The unexpected reduction in Plagl1 and Mrkrn3 RNA could be explained by up-regulation of these miRNAs in the Zfp57-mutant day 12-cells (Supplementary Fig. 6a–c). This could also be responsible of the indirect down-regulation of several neuronal non-imprinted miRNA-target genes in the Zfp57\textsuperscript{−−} NPLCs (Supplementary Fig. 3d).

Zfp57-mutant cells only partially differentiated towards neural precursors, suggesting that genes controlled by ZFP57 are necessary for neural differentiation of ESCs under the conditions used in this study. This finding contrasts with the absence of major problems of neurogenesis reported in Zfp57-null embryos.\textsuperscript{37} However, the Zfp57\textsuperscript{−−} ESCs show more methylation defects at the ZFP57-binding sites and are therefore expected to have more gene expression changes and severe phenotype than the mutant embryos.\textsuperscript{51} Consistent with this hypothesis, the Dnmt3l knockout that prevents establishment of methylation at all maternally imprinted DMRs is associated with exencephaly and other neural tube defects in the mouse.\textsuperscript{51} A possible explanation of the partial differentiation capacity of the Zfp57\textsuperscript{−−} cells toward the neural fate is the deregulation of imprinted genes that have a role in the control of neural cell proliferation, differentiation, apoptosis and migration, such as Cadk1, Dkk1, Igf2, Peg3 and Plagl1.\textsuperscript{52,56} Alternatively, the expression of non-imprinted genes involved in neurogenesis that are located close to the ZFP57 binding sites may be altered.

In conclusion, this study provides an extensive analysis of the relationship between DMR methylation and imprinted gene expression, and suggests that a number of ZFP57-target genes have a role in neural differentiation of mouse ESCs in vitro.

Materials and methods
Cell lines and culture conditions. The wild-type hybrid ESC line JB1, which is (JF1 × C57BL/6) F1, and the JB1-derived Zfp57\textsuperscript{−−} ESC line were described previously.\textsuperscript{24,57} ESCs were cultured under serum-free conditions on gelatinized tissue culture dishes and maintained in ESGRO Complete Plus Serum-free Clonal Grade 1i Medium (Merck-Millipore) in the presence of the Gsk3 inhibitor CHIR99021 (at 3 μM). The wild-type inbred 129 ESC line E14 and its derived Zfp57\textsuperscript{−−} ESC line were cultured under standard feeder-free conditions as previously described.\textsuperscript{24} The wild-type and Zfp57\textsuperscript{−−} JB1 and E14 ESCs were differentiated toward NPCs following the protocol described by Gaspard and collaborators (Fig. 1a)\textsuperscript{49,50}. In vitro corticogenesis of ESCs was started with plating at the optimal density of 7.500 ES cells/cm\textsuperscript{2} and culturing in chemically defined default medium (DDM).

From day 2 to 10 of culture, a Sonic hedgehog inhibitor (cyclopamine) was added at 1 μM concentration. From day 10 to 12, the medium was replaced with DDM only. The cells were cultured at 37 °C under an atmosphere of 5% CO\textsubscript{2}.
**DNA methylation analysis.** Genomic DNA from cultured cells was isolated using the Wizard Genomic DNA Purification Kit (Promega) following the manufacturer’s protocols. ~1 μg genomic DNA was subjected to bisulfite conversion using EpiTect Bisulfite Kit (Qiagen). The bisulfite sequencing was obtained by three methods: sequencing of individual clones, pyrosequencing and direct sequencing of the bisulfite-treated and PCR-amplified genomic DNA. PCR amplification was obtained with AmpliTaq DNA polymerase Kit (Thermo Fisher Scientific). For the Plcg1: TSS-DMR, Peg3: TSS-DMR and Igf2r: TSS-DMR, PCR products were cloned into the TOPO TA cloning vector (Invitrogen) and individual clones were isolated and sequenced (Eurofins Genomics). For the H19/Igf2:IG-DMR, Mest:TSS-DMR and Rasperf1:TSS-DMR, DNA methylation was assessed by Sanger sequencing (Eurofins Genomics) of the bisulfite-treated and PCR-amplified genomic DNA (AmpliTaq DNA polymerase Kit, Thermo Fisher Scientific). For the Kcnq1ot1:TSS-DMR, Meg3/Dk1:IG-DMR, Peg3:TSS-DMR, and Srrpn:TSS-DMR DNA methylation was analysed by pyrosequencing using PyroMark Q48 Autoprep sequencing machine (Qiagen). For the H19/Igf2:IG-DMR, Mest:TSS-DMR and Rasperf1:TSS-DMR, DNA methylation was assessed after direct sequencing of the bisulfite-treated and PCR-amplified genomic DNA. All the primers are listed in Supplementary Table 6.

**RNA analysis.** RNA was isolated using TRI reagent (Sigma-Aldrich) according to the manufacturer’s instructions. Approximately 900 ng of total RNA was retro-transcribed to cDNA by using QuantiTect Reverse Transcription Kit (Qiagen) according to the manufacturer protocol. For locus-specific gene expression analysis, cdNA was amplified by real-time PCR using SYBR Green PCR Master Mix (Bio-Rad) on a CFX Connect Real-Time PCR Detection System. For locus-specific allele-specific expression analysis, target genes were amplified, purified using PCR purification Kit (Qiagen), and sequenced using Sanger sequencing (Eurofins Genomics). PCR and RT-qPCR primers are reported in Supplementary Table 6.

For RNA-seq, ~1 μg of high quality RNA (260/280 > 1.80) samples in biological duplicates were subjected to library preparation using TruSeq Stranded Total RNA Ribo-Zero Gold Kit and sequencing with 2 × 125 bp paired end (PE) mode on Illumina HiSeq 2500 System (Institute of Applied Genomics: IGA, Italy).

**Bioinformatic analyses.** The reads obtained from RNA-seq were assessed for quality using FastQC tool and trimmed for adapter sequences using Trimomatic v0.36 (in PE mode, MINLEN:36, HEAD-CROP if required and other parameters as default)63. Only good quality reads were aligned to the mouse reference genome (gencode vM20) using STAR v2.5.4a (-outFilterMultimapNmax 1,-outFilterMismatchNmax 3 and other parameters set as default)63. Reads were assigned to genes (annotation gencode vM20) using the featureCounts function (-p option) from Subread package v.1.6.061. In the annotation file, two transcripts Gm49394 and Mirk483, which overlapped Igf2 coordinates and present in the gencode vM20 gtf annotation file, were removed. We arbitrarily set a cutoff of >8 reads (wild-type + mutant samples) to remove the genes with very low expression level. We used DESeq2 v.1.24.039 for normalization and differential expression between wildtype-NPCs and Zfp57−/− NPCs. Genes with adjusted p-value < 0.05 were considered statistically significant and were used for downstream analysis. To determine the differentiation potential of our NPCs we compared our RNA-Seq data with publicly available RNA-Seq data of ESCs and NPCs66. The raw data were processed as before and the read count matrix was normalized by the “upper quartile” method. Subsequently, we adjusted the expression of all genes by dividing by beta-actin expression of the respective samples. After adjustment, we computed the Z-scores for each sample and plotted for a set of lineage-specific markers of pluripotency, neural/cortex, endoderm and mesoderm. The functional enrichment analysis of deregulated genes was performed using gprofiler2 R package64. Biological process terms (GO:BP) from gprofiler2 were considered significantly enriched if adjusted p-value < 0.05 and only top 10–15 candidates were depicted on a barplot. The allele-specific RNA-Seq analysis was performed by utilizing the SNPs present between B6 and JF1 genome. SNPs were derived from Takada and collaborators63 and converted to mm10 using CrossMap v-0.2.564. These SNPs were overlapped with the JF1 SNPs downloaded from the Sanger Institute website (ftp://ftp-mouse.sanger.ac.uk/REL-1807-SNPs_Indels/mgg.v6.merged.norm.snp.indels.sillered.vcf.gz), which revealed ~11 millions SNP positions. These SNPs were used for creating N-masked genomes from mm10 reference genome using custom Python script by incorporating ‘Ns’ at the SNP positions. The alignment to N-masked genome was performed using STAR v2.5.4a with specific parameters (-alignEndsTypeEndToEnd, -outSAMattributes All,-outFilterMismatchNmax 1 -outFilterMismatchNmax 3 and other parameters as default). The aligned reads were assigned to maternal (JF1) or paternal (B6) allele using SNPSplit v0.3.258, accompanied with a file containing SNPs information. For visualization purposes, bigwig files were generated and uploaded to UCSC genome browser (mouse genome version GRChm38/mm10). The allele-specific reads were assigned to the genes using featureCounts61 and imported to DESeq2 for further normalization. As both parental alleles (viz. B6 and JF1) were originated from the same sample, the allele-specific counts were normalised by size factor obtained from bulk RNA-Seq using DESeq2. We filtered out the genes with <10 normalized counts (sum of mat and pat alleles) in both wild-type replicates. We confined our analysis only to imprinted genes, the list of which was collected from geneimprint database (www.geneimprint.com) and literature search65. The allele-specificity was estimated from the ratio of maternal allele normalized counts to the sum of maternal and paternal allele normalized counts (JF1/B6 + JF1) and only the genes with allelic ratio ≤0.33 or ≥0.67 were prioritized as monoallelic and plotted using ggplot2 R package. RNA-Seq raw and processed data is available in GEO (GSE164669).

ChIP-seq data for ZFP57 and KAP1 were generated previously20 and available in GEO repository (GSE74757). The raw reads were downloaded and aligned to mm10 using bowtie v1.2.1.1 in single-end mode. Only the uniquely mapped reads were obtained (bowtie option -m 1) and PCR duplicates were removed using custom shell script. Peak were called by using macs14 (MACS 1.4.2)60, which revealed 544 ZFP57 and 6986 KAP1 peaks (Supplementary Table 7). To increase the confidence over identified peaks, both ZFP57 and KAP1 peaks were
intersected using bedtools, which revealed 457 overlapping peaks. The distance of deregulated genes in the Zfp57−/− day 12 cells from the ZFP57/KAP1 peaks was calculated using bedtools (−closest function) and plotted using ggplot2 R package. The genes overlapping with ZFP57/KAP1 peaks were set at distance 0.

**Statistical analysis.** Statistical significance for the relative expression of the lineage-specific markers and imprinted genes was conducted by unpaired Student’s t-test. Enrichment of differentially expressed imprinted genes over all differentially expressed genes was assessed using the Hypergeometric test. The difference between the two empirical cumulative functions of the distances between genes and ZFP57/KAP1 peaks was evaluated using the two-sample Kolmogorov–Smirnov test. The allele-specific expression of genes in wild-type cells was assessed using a Proportion test and multiplicity correction was performed using the “Benjamini-Hochberg” method.

**Immunofluorescence.** For Nestin and Pou5f1 (also known as Oct3-4) immunofluorescence, the cells were fixed in 4% PFA and permeabilized with 0.1% Triton X-100/PBS. Cells were then, blocked with 10% normal goat serum (NGS)/1% BSA (bovine serum albumin)/PBS. Later, the cells were incubated with the primary antibodies (i.e., anti-Nestin, anti-Pou5f1, conjugated respectively with Alexa Fluor 594, sc-33677 AF594 and Alexa Fluor 488, sc-5279 AF488). After 3 washes with 0.1% Triton X-100/PBS cells were incubated with DAPI for 5 min. Images were captured with a fluorescence microscope (DMI6000B, Leica) run with the LAS AF 2.6 software (Leica).

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Author contributions
Conceptualization, A.R. and B.A.; methodology, B.A., A.V., A.P., R.E., C.A., S.F., F.D.R. and A.F.; validation, C.G. and Fr. C.; formal analysis, B.A., A.V. and A.P.; data curation, B. A. and A.V.; writing, review and editing, B.A., A.V., F.C., R.F. and A.R.; supervision, C. A. and A.R.; funding acquisition, A.R. All authors have read and agreed to the published version of the manuscript.

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Competing interests
The authors declare no competing interests.

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Correspondence and requests for materials should be addressed to A.R.

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