A Splicing Variant in OCRL Gene Might Explain the Second Case of Lowe Syndrome in Iran

Massoud Houshmand1, Gholamreza Babamohammadi2, Hamidreza Moazzeni3, Ahmad Reza Salehi Chaleshtori 3 and Mohammad taghi Akbari 3,*

1Department of Medical Genetics. National Institute for Genetic Engineering and Biotechnology, Tehran, Iran
2Tehran Genetic Counseling Center, Tehran, Iran
3Tehran Medical Genetics Laboratory (TMGL), Tehran, Iran
*Corresponding author: Tehran Medical Genetics Laboratory, Postcode: 1581764111, Tehran, Iran. Email: mtakbari@gmail.com

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Abstract

Lowe syndrome is a condition that primarily affects eyes, brain, and kidneys. This disorder follows X-linked recessive mode of inheritance and it occurs in males mainly. Mutations in OCRL (located at Xq25) gene can cause accumulation of phosphatidylinositol bisphosphate and disturbed actin cytoskeleton remodeling. There are 268 mutations in OCRL gene causing Lowe syndrome or Dent disease 2 in HGMD database, however 10 - 20% of Lowe syndrome suspects remain undiagnosed at molecular level. Here we present a male case of Lowe syndrome with characteristic features. Comprehensive clinical examination and genetic counseling were performed. Sanger sequencing was employed to investigate the possible OCRL mutations and we identified a donor splice site variant (NM-000276: c.2469 + 1G > A) in hemizygous state. This is a pathogenic variant according to the ACMG standards and guidelines and might explain the clinical features of the patient. This result is in accordance with the clinical diagnosis of Lowe syndrome and it is absent from ExAC, 1000 G, Iranome, GME, gnomAD Genome databases of healthy controls. In-silico analysis of this splicing variant revealed that the position is highly conserved between species. Splicing prediction tools predicted some changes in splicing pattern of the OCRL transcript, elimination of some protein features, and malfunctioning the OCRL protein as a consequence of this variant. Accordingly, we proposed the c.2469 + 1G > A variant might explain the clinical features in studied patient and be employed for prenatal diagnosis of Lowe syndrome in the family.

Keywords: Splicing Variant, OCRL, Lowe Syndrome, Iran

1. Background

The oculocerebrorenal syndrome of Lowe (OCRL) [OMIM# 309000] is a rare multisystemic X-linked disorder, which affects 1 person in 500000 in general population (1). An spectrum of abnormalities in eye, kidney, skeletal, and neuronal systems are associated with Lowe syndrome (2). Clinical features are including but not limited to intellectual disability, failure to thrive, congenital cataract, glaucoma, joint hypermobility, scoliosis, genu valgum, sebaceous cysts, seizures, and kidney disease (2, 3).

Genetic basis of Lowe syndrome was first described through studying a girl with X-autosomal translocation (t(X;3) (q25;q27)). Hodgson et al suggested that underlying genetic defect might exist in a locus located at Xq25 band (4). Further details provided through studying two unrelated females with Lowe syndrome and the OCRL gene assigned as the most probable causative gene (5). The gene comprises 23 exons and stretches 52 kb on Xq25-26 chromosome. 6 All OCRL exons are coding region except the last one and alternative splicing of exon 18a has been demonstrated in the brain (6, 7). This gene encodes phosphatidylinositol 4,5-bisphosphate-5-phosphatase protein, a component of Golgi network that is involved in regulating membrane trafficking, and also play a role in primary cilium formation (8). This protein regulates traffic in the endosomal pathway by regulating the specific pool of phosphatidylinositol 4,5-bisphosphate that is associated with endosomes (9). The OCRL also acts as a regulator of phagocytosis, hydrolyzing phosphatidylinositol 4,5-bisphosphate (PtdIns(4,5)P2) to promote phagosome closure, through attenuation of PI3K signaling (10).

An array of mutations (268 mutations) in this gene cause Lowe syndrome or Dent Disease 2 except regulatory/repeat variations (HGMD Professional 2019.4), how-
ever in 10% - 20% of Lowe syndrome suspects genetic determinants remain undiagnosed molecularly (11). Cardinal features of the disease include dense congenital bilateral cataract at birth,1 involvement of central and the peripheral nervous systems (12), neonatal hypotonia,1 seizures (up to 50% of Lowe syndrome cases) (12), behavioral abnormalities in more than 80% of patients (13), neuropathological findings (14), proximal tubular dysfunction (15), progressive renal failure often leads to end-stage renal disease (ESRD) (16), and proteinuria which is observed in all patients. Some of these features were not observed in all patients of Lowe syndrome and Possible genotype-phenotype correlation and the spectrum of mutations are described elsewhere (2).

2. Objectives

Here we presenting complete clinical and molecular characterization of a Lowe syndrome patient. Since the disease is rare, this will help more accurate clinical/molecular diagnosis, improved genetic counseling, and offering appropriate reproductive options to such families.

3. Methods

3.1. Clinical Characterization

A 21 years old male patient was referred to Tehran Medical Genetic Laboratory (TMGL) in order to receive genetic counseling and possible molecular diagnosis. The patient was born to a consanguineous marriage, while there was a history of a deceased affected male sibling in the family with same clinical manifestations (Figure 1 and Figure 2A). Genetic counseling and clinical examination undertaken and patient’s consent form obtained from patient’s parents. Cardinal features of Lowe syndrome perfectly matched with the patient’s phenotype as listed in Table 1. Therefore, analysis of the OCRL gene was conducted to establish molecular diagnosis.

3.2. Molecular Characterization of the OCRL Gene

The patient’s peripheral blood was drained from ante-cubital vein and used to extract genomic DNA and molecular characterization. Genomic DNA was extracted using standard salting-out method and implemented in PCR reactions to amplify and analyze the entire coding region of the OCRL gene. Sanger sequencing of PCR products conducted on ABI 3130 genetic analyzer platform (Applied Biosystems, USA). DNA sequences were aligned to the GRCh37 assembly of the human genome and called variants were classified according to the ACMG standards and guidelines (17). We also studied the variant in all family members and performed segregation analysis for further evaluation of the variant.

3.3. Bioinformatic Analysis of the Variant

Databases of healthy control samples (gnomAD (https://gnomad.broadinstitute.org), 1000G (https://www.internationalgenome.org), GME (http://igm.ucsd.edu/gme), EVS (https://evs.gs.washington.edu/EVS), Iranome (http://www.iranome.ir), and our in-house database (TMGL db) (includes more than 200 exome sequencing projects)) were used to determine the minor allele frequency. The ClinVar database (https://www.ncbi.nlm.nih.gov/clinvar) and Mutationtaster (http://www.mutationtaster.org) were recruited to predict pathogenicity of the identified variant. To evaluate how this variant can affect splicing pattern of the transcript, we used NNSPLIC (https://www.fruitfly.org/seq_tools/splice.html), NetGene2 (http://www.cbs.dtu.dk/services/NetGene2), and Human Splicing Finder (Version 3.1) (http://umd.be/Redirect.html) online algorithms.

4. Results

The first set of questions aimed to be answered were about the clinical diagnosis of Lowe syndrome. It can be seen from the data in Table 1 that clinical presentations of the patient are in accordance with Lowe syndrome diagnosis. Cardinal features of the patient are illustrated in Figure 1 and all are consistent with the clinical diagnosis of Lowe syndrome. Through reviewing the literature, we extracted the prevalence of each finding in Lowe syndrome patients and compared with those findings observed in the patient (Table 1). The patient has shown many symptoms of the Lowe syndrome even those observed less frequent in such patients such as phosphaturia. Cognitive impairment in this patient was evaluated as mild-moderate and we were unable to examine his brain through MRI/CT-Scan since his parents refused our request. The clinical diagnosis is also supported by the pedigree (Figure 2A) at first glance. Since the disease is X-linked recessive and mutations of OCRL gene can cause Lowe syndrome, we analyzed the OCRL gene through Sanger sequencing method.

4.1. Molecular Characterization of the OCRL Gene

Thanks to Sanger sequencing and investigation of all OCRL coding regions, we identified a hemizygous variant
Table 1. Clinical Findings in the Patient

| Frequency in Other Studies | V1 | Clinical Findings in the Patient Lowe Syndrome |
|---------------------------|----|-----------------------------------------------|
|                           |    | **Growth**                                    |
|                           |    | Height                                        |
|                           | +  | Short height                                  |
|                           |    | Head circumference                            |
|                           | 12 cm | NA                |
|                           | +  | Head circumference 12 cm                     |
|                           |    | Weight                                        |
|                           | 59 kg | NA                |
|                           | +  | Failure to thrive                             |
|                           | + (33 cm) | NA          |
|                           |    | Eyes                                          |
|                           | +  | Congenital cataract                            |
|                           | +  | Glaucoma                                      |
|                           | +  | Decreased visual acuity                       |
|                           | +  | Corneal leek                                  |
|                           | -  | Freckles                                      |
|                           | +  | Strabismus                                    |
|                           | +  | Horizontal nystagmus                           |
|                           |    | Tooth                                         |
|                           | +  | Renal cyst                                    |
|                           | -  | Erupted hypoplasia                             |
|                           | +  | Loss of hair                                  |
|                           |    | Abdomen                                       |
|                           | -  | Constipation                                  |
|                           |    | Genitourinary                                 |
|                           | +  | Kidney                                        |
|                           | +  | Proximal renal tubular acidosis               |
|                           | -  | Renal Fanconi syndrome                        |
|                           | -  | Urinary incontinence                          |
|                           |    | Genitalia                                     |
|                           | -  | Cryptorchidism                                |
|                           |    | Skeletal                                      |
|                           | +  | Joint hypermobility                           |
|                           | +  | Osteomalacia                                  |
|                           | -  | Renal rickets                                 |
|                           | -  | Pathologic fractures                          |
|                           |    | Spine                                         |
|                           | +  | Kyphosis                                      |
|                           | -  | Platyspondyly                                 |
|                           | +  | Scoliosis                                     |
|                           | +  | Renal Fanconi syndrome                        |
|                           | -  | Renal rickets                                 |
|                           | -  | Pathologic fractures                          |
|                           |    | Pelvis                                        |
|                           | +  | Hip dislocation                               |
|                           | +  | Genitalia                                     |
|                           | +  | Gastrointestinal                              |
|                           | +  | Abdomen                                       |
|                           | -  | Skin, Nails, Hair                             |
|                           | +  | Dermal cysts                                  |
|                           | +  | Subcutaneous nodules                          |
|                           |    | Neurologic                                    |
|                           | +  | Sternum hypotonia                             |
|                           | +  | Atelecs                                       |
|                           | +  | Mental retardation                            |
|                           | +  | Intrinsic myopathy                            |
|                           |    | Central nervous system                        |
|                           |    | Increased signal intensity on T2-weighted scans in the pons, and cerebellar peduncle, NA |
|                           |    | Periventricular cysts                         |
|                           | +  | Bicarbonateuria                               |
|                           | +  | Aminoacidsura                                 |
|                           | +  | Protonin                                     |
|                           | +  | Phosphate                                    |
|                           | +  | Elevated serum acid phosphatase               |
|                           | +  | Abnormal serum protein electrophoresis        |
|                           | +  | Elevated total cholesterol                    |

in a donor splice site of the gene with the following coordinates; chrX: 128722991G > A; OCRL (NM-000276.4):c.2469 + 1G > A (Figure 2A and B), which is classified as a pathogenic variant (PVS1, PM2, PP3, PP5) according to the ACMG standards and guidelines (17). This variant was absent from our In-House database (TMGL db), ExAC, 1000 G, Iranome, EVS, GME, and gnomAD Genome databases, which indicates its rarity amongst healthy controls and its possible deleterious effects. Recently, the variant has been reported as a likely pathogenic variant in ClinVar database (variation ID:
835463). Through segregation analysis of the variant, we characterized patient’s mother and his two sisters as carriers of the OCRL splicing variant. In addition, his father carries the wild-type allele and he is healthy. Consequently, the OCRL variant and symptoms confers the X-linked pattern of inheritance consistent with the Lowe syndrome diagnosis (Figure 2A).

4.2. In-silico Analysis of the Splicing Variant

The mutation position is highly conserved between multiple species (Figure 2C) and our evaluations using NNSPLICE (https://www.fruitfly.org/seq_tools/splice.html),

Figure 1. Clinical presentations of the patient. A, Frontal view of the patient; B, lateral view of the patient; C, an example of subcutaneous nodules observed in this patient. D, Overcrowded teeth in the patient.
NetGene2 (http://www.cbs.dtu.dk/services/NetGene2/), and HSF (http://www.umd.be/HSF/) showed that c.2469 + 1G > A variant may critically affect the OCRL mature transcript due to changes in donor splice site (Table 2). This may cause elimination of some protein features in downstream of the altered splicing site. Other features of the variant and possible effects on OCRL protein structure are mentioned in Table 2 and Table 3. This splicing variant affects the Rho-GAP domain of OCRL protein and elimination of affected domains is a probable consequence of this pathogenic variant (Figure 2D).

Taken together, the c.2469 + 1G > A variation in OCRL gene might be a pathogenic variant with detrimental effects on protein structure and function.

5. Discussion

The oculocerebrorenal syndrome of Lowe is of interest because of its rarity and the allelic heterogeneity exhibited by the OCRL gene. In Iran, there is only one report of Lowe syndrome in the literature before this report (18). The patient we studied here showed most hallmarks of Lowe syndrome (Table 1 and Figure 1) and (2). The clinical consequences of c.2469 + 1G > A are in accordance with other OCRL mutations and manifestations of Lowe syndrome (https://www.omim.org/). This variant is absent from public databases of healthy controls as well as our in-house database (TMGL db) including more than 200 exome sequencing projects (Table 2). This splicing variant may cause elimination of six helices from the OCRL protein structure including the Rho-GAP domain (GTPase-activator protein for Rho-like GTPases) (Table 3 and Figure 2D). This family of protein domains transduce signals and control cell adhesion, motility and shape by actin cytoskeleton formation. Rho proteins act as molecular switches and need GTPase-activating proteins (GAPs) which stimulate the intrinsic GTPase activity of small G proteins (http://smart.embl-heidelberg.de/). Elimination of OCRL-Rho-GAP domain might alter the protein function through malfunctioning of signal transduction and cause development of the oculocerebrorenal phenotype. We have to bear in mind that c.2469 + 1G > A might influence on the last two coding exons of the OCRL gene, consequently attenuated clinical findings are expected due to non-coding inherent of exon 23 of the OCRL gene.

There is no significant genotype-phenotype correla-
Table 2. In Silico Evaluation of the Variant

| Gene (Exon/Intron) | Variants Coordinates | Pathogenicity Prediction | Minor Allele Frequency (MAF) |
|-------------------|----------------------|-------------------------|-----------------------------|
| OCRL (exon 22/intron 22) | NM_000276: c.2469 + 1G > A; ChrX (GRCh37): g.128722991G > A | MutationTaster: Disease Causing<br>ACMG: PVS1 Null Variant | ClinVar: Absent<br>ExAC: Absent<br>ACMG: PVS1 Null Variant<br>1000G: Absent<br>Iranome: Absent<br>EVs: Absent<br>NNSPLICE: Affecting donor splice site<br>NetGene2: Affecting donor splice site<br>gnomAD Genome: Absent<br>HSF: Alteration of the WT donor site<br>In-House (TMGL): Absent |

*aAt exon-intron boundary, protein features might be affected, splice site changes.*

Table 3. Protein Features may Lost as a Consequence of c.2469 + 1G > A Mutation in OCRL Gene

| Start (aa) | End (aa) | Feature Details | Consequences of Mutation |
|-----------|---------|-----------------|--------------------------|
| 721       | 901     | Rho-GAP DOMAIN  | might get lost (downstream of altered splice site) |
| 818       | 826     | HELIX           | might get lost (downstream of altered splice site) |
| 830       | 847     | HELIX           | might get lost (downstream of altered splice site) |
| 850       | 875     | HELIX           | might get lost (downstream of altered splice site) |
| 857       | 868     | HELIX           | might get lost (downstream of altered splice site) |
| 873       | 895     | HELIX           | might get lost (downstream of altered splice site) |

*Abbreviation: aa, amino acid.*

Management of this patient was conducted by early removal of cataracts with postoperative glasses regarding to his cataract. However, and despite surgical lens are not recommended for such patients (1), the patient V-1 has been benefited from intraocular lens implant. Generally, the OCRL phenotype is associated with variable degrees of intellectual disability (ID) (1). The patient V-1 shows mild-to-moderate ID, similar to 25% of OCRL male patients. Since, the patient presents with seizures, anticonvulsant therapy was conducted using lamotrigine (25 mg/day) (https://www.drugs.com/mtm/lamotrigine.html) and depakine chrono (500 mg/day) (https://www.drugs.com/international/depakine-chrono.html). Treatment of renal tubular dysfunction performed by oral supplements of sodium and potassium bicarbonate. The patient V-1 does not need to renal transplantation or dialysis. Other management strategies for Lowe syndrome patients are described elsewhere (21).
Correlation of OCRL mutations with the Lowe syndrome is well established in the literature while there is no clear genotype-phenotype correlation. Here we studied a characteristic case of Lowe syndrome comprehensively and this may help expansion of the clinical phenotype and mutation spectrum as well. We proposed the c.2469 + 1G > A pathogenic variant might affect the OCRL structure and function and may explain the clinical features in this patient.

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Footnotes

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References

1. Loi M. Lowe syndrome. Orphanet J Rare Dis. 2006;1:36. doi: 10.1186/1750-1172-1-36. [PubMed: 17022554]. [PubMed Central: PMC1526415].

2. Bokenkamp A, Ludwig M. The oculocerebrorenal syndrome of Lowe: an update. Pediatr Nephrol. 2016;31(12):2201-12. doi: 10.1007/s00467-016-3343-3. [PubMed: 27011217]. [PubMed Central: PMC5118406].

3. Streiff EB, Straub W, Golay L. [Ocular manifestations of Lowe syndrome]. Ophthalmologica. 1995;209(5-6):632-9. French. doi: 10.1159/000303367. [PubMed: 8599350].

4. Hodgson JV, Heckmann JZ, Hughes E, Crolla JA, Dubowitz V, Bobrow M. A balanced de novo X/autosome translocation in a girl with manifestations of Lowe syndrome. Am J Med Genet. 1986;23(3):837-47. doi: 10.1002/ajmg.1320230311. [PubMed: 3953680].

5. Attree O, Olivos IM, Okabe I, Bailey LC, Nelson DL, Lewis RA, et al. The Lowe’s oculocerebrorenal syndrome gene encodes a protein highly homologous to inositol polyphosphate-5-phosphatase. Nature. 1992;358(6383):239-42. doi: 10.1038/358239a0. [PubMed: 1321346].

6. Nussbaum RL, Orriss BM, Janne PA, Charnas L, Chinnault AC. Physical mapping and genomic structure of the Lowe syndrome gene OCRL. Hum Genet. 1997;99(2):245-50. doi: 10.1007/s004390050329. [PubMed: 9048491].

7. Choudhury R, Noakes CJ, McKenzie E, Kox C, Lowe M. Differential clathrin binding and subcellular localization of OCRL1 splice isoforms. J Biol Chem. 2009;284(15):9965-73. doi: 10.1074/jbc.M807442200. [PubMed: 19211563]. [PubMed Central: PMC2665120].

8. Suchy SF, Nussbaum RL. The deficiency of PIP2 5-phosphatase in Lowe syndrome affects actin polymerization. Am J Hum Genet. 2002;71(6):1420-7. doi: 10.1086/344557. [PubMed: 12428211]. [PubMed Central: PMC378584].

9. Vicinanza M, Di Campi A, Polichschuk E, Santoro M, Di Tullio G, Godi A, et al. OCRL controls trafficking through early endosomes via PtdIns4,5P(2)-dependent regulation of endosomal actin. EMBO J. 2010;29(4):4970-85. doi: 10.1038/emboj.2010.354. [PubMed: 29791085]. [PubMed Central: PMC342079].

10. Bohdanowicz M, Balkin DM, De Camilli P, Grinstein S. Recruitment of OCRL and Inppl5 to phagosomes by Rab5 and APPL1 depletes phosphoinositides and attenuates Akt signaling. Mol Biol Cell. 2012;23(1):76-87. doi: 10.1091/mbc.E10-04-0489. [PubMed: 22072788]. [PubMed Central: PMC348896].

11. Hichri H, Rennu J, Monnier N, Coutton C, Dorseuil O, Poussou RV, et al. From Lowe syndrome to Dent disease: correlations between mutations of the OCRL1 gene and clinical and biochemical phenotypes. Hum Mutat. 2011;32(4):379-88. doi: 10.1002/humu.21591. [PubMed: 2105165].

12. Nussbaum R, Suchy S. The oculocerebrorenal syndrome of Lowe (Lowe syndrome). The Online Metabolic and Molecular Bases of Inherited Disease. 2001. p. 6257-66.

13. Kenworthy L, Charnas L. Evidence for a discrete behavioral phenotype in the oculocerebrorenal syndrome of Lowe. Am J Med Genet. 1995;59(3):283-90. doi: 10.1002/ajmg.1320590304. [PubMed: 8599350].

14. Allmendinger AM, Desai NS, Burke AT, Viswanadhan N, Prabhu S. Neuroimaging and renal ultrasound manifestations of Oculocerebrorenal syndrome of Lowe. J Radiol Case Rep. 2014;8(10):3-7. doi: 10.1002/jrcr.1740. [PubMed: 25426299]. [PubMed Central: PMC424247].

15. Bockenhauer D, Bokenkamp A, van’t Hoff W, Levitschenko E, Kus-Van Holthe JE, Tasic V, et al. Renal phenotype in Lowe Syndrome: a selective proximal tubular dysfunction. Clin J Am Soc Nephrol. 2008;3(5):1340-6. doi: 10.2215/CJN.00520108. [PubMed: 18480301]. [PubMed Central: PMC2518783].

16. Charnas LR, Bernardini I, Rader D, Hoeg JM, Gahl WA. Clinical and laboratory findings in the oculocerebrorenal syndrome of Lowe, with special reference to growth and renal function. N Engl J Med. 1993;329(19):1318-25. doi: 10.1056/NEJM199311243291904. [PubMed: 7172228].

17. Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med. 2015;17(5):405-24. doi: 10.1038/gim.2015.30. [PubMed: 2574868]. [PubMed Central: PMC454475].

18. Amirhakimi G, Fallahzadeh M, Saneifard H. Lowe Syndrome: Report of a case and brief literature review. Iran J Pediatr. 2009;19(4):477-20.

19. Tosetto E, Addis M, Cardi G, Meloni C, Emma F, Vergine G, et al. Locus heterogeneity of Dent’s disease: OCRL1 and TMEM27 genes in patients with no CLCN5 mutations. Pediatr Nephrol. 2009;24(10):1967-73. doi: 10.1007/s00467-009-1228-4. [PubMed: 19582487].

20. Levin-Laina N, Dinour D. Renal disease with OCRL mutations: Dent-2 or Lowe syndrome? Journal of pediatric genetics. 2012;1(3).

21. Lewis RA, Nussbaum RL, Brewer ED. Lowe Syndrome. In: Adam MP, Ardinger HH, Pagon RA, Wallace SE, Bean LJH, Stephens K, et al., editors. GeneReviews(R). Seattle (WA); 1993.