Achyranthes bidentata is one of the most commonly used Chinese herbal medicines (CHM) that is currently considered for the treatment of osteoarthritis. The purpose of this study was to reveal the mechanism of Achyranthes bidentata in osteoarthritis treatment based on the network pharmacology.

Methods. The effective components of Achyranthes bidentata were firstly screened out from the TCMSP database with ADME property parameters. Then, osteoarthritis-related proteins targeted by the effective components were predicted based on the DrugBank and CTD databases. Subsequently, enrichment analysis and interaction network between targets of effective components and pathways were also studied. In addition, the differentially expressed genes (DEGs) of GSE55457 were used for validation of the osteoarthritis-related target proteins. Finally, the effective components–target molecular docking models were predicted.

Results. A total of 10 effective components were identified, of which kaempferol and quercetin had 1 and 29 targets, respectively. There were 26 target proteins of quercetin related to the osteoarthritis. These targets were mainly enriched in mitochondrial ATP synthesis coupled proton transport, cellular response to estradiol stimulus, and nitric oxide biosynthetic process. In addition, there were three common proteins, PIM1, CYP1B1, and HSPA2 based on the DEGs of GSE55457, which were considered as the key targeted proteins of the quercetin. Conclusion. The docking of PIM1–quercetin, CYP1B1–quercetin, and HSPA2–quercetin may play important roles during the treatment of osteoarthritis by Achyranthes bidentata.
by Traditional Chinese Medicine to treat the arthritis based on the kidney tonifying and blood activating method [8, 9]. *Achyranthes bidentata* contains many types of compounds, including polysaccharides, saponins, anthrones, flavonoids, peptides, organic acids, and various trace elements, which play an important role in the treatment of osteoarthritis [10].

Quercetin is a polyhydroxy flavonoid that exhibits high nutritional and medicinal properties due to its diverse biological activities. Quercetin is widely found in the flowers, leaves, and fruits of plants. It is known that more than 100 kinds of herbs contain quercetin, such as *Flos sophorae*, *Chrysanthemum*, *psyllium*, and *Eleutherococcus senticosus* [11]. Mollica et al. reported that *Juglans regia* leaves which contained quercetin could alleviate the symptoms of diabetic rats [12]. Besides, the extracts of two species of broccoli form north-central Italy which also contained quercetin decreased the biochemical parameters in diabetic rats [13].

Network pharmacology aims to reveal the mechanisms of natural medicines at the system and molecular levels. At present, network pharmacology plays a significant role in the study of the mechanisms of various natural medicines and TCM, such as *Liuwei Dihuang Pills* [14], *Qing-Luo-Yin* [15], and essential oil from *Aegle marmelos* leaves [16]. In this study, a comprehensive pharmacology network was constructed based on several databases and bioinformatics methods to understand the pharmacological mechanism of *Achyranthes bidentata* on osteoarthritis.

2. Methods

2.1. Screening of Effective Components of *Achyranthes bidentata*. The components and other information of *Achyranthes bidentata* were obtained from the Traditional Chinese Medicine Systems Pharmacology Database and Analysis Platform (TCMSP, http://lsp.nwu.edu.cn/tcmsp.php), such as molecule name, molecular weight (MW), lipid/ water partition coefficient (A log P), hydrogen-bond donors/acceptors (Hdon/Hacc), oral bioavailability (OB), intestinal epithelial permeability (Caco-2), blood-brain barrier (BBB), drug-likeness (DL), and drug half-life (HL). TCMSP contains a large number of herbal entries, and the drug-target networks and drug-disease networks obtained from the TCMSP will help revealing the mechanisms of action of Chinese herbs and developing new herb-oriented drugs [17]. Based on the absorption, distribution, metabolism, and excretion (ADME) property parameters with OB≥40% and DL≥0.2, effective components of *Achyranthes bidentata* were obtained [17–19].

2.2. Prediction of Proteins Targeted by Effective Components. The targeted proteins of effective molecules were predicted based on the TCMSP and DrugBank (https://www.drugbank.ca/) databases by matching the UniProt ID of the UniProt database (https://www.uniprot.org/) to the gene symbol. The drug information provided by the DrugBank database is experimentally proven, clinically tested, or marketed, and the results are accurate and reliable [20].

2.3. Identification of Osteoarthritis-Related Proteins. Comparative Toxicogenomics Database (CTD, update 2019, http://ctdbase.org/) published by Mount Desert Island Biological Laboratory provides information on chemical-gene/protein interactions, chemistry-disease, and gene-disease relationships [21]. In order to identify the key proteins related to osteoarthritis among the targeted proteins above, CTD database was used with the “osteoarthritis” as the key word. Then, the proteins of the intersection of two sets were selected for further study.

2.4. Enrichment Analysis of Targeted Proteins. Gene Ontology (GO) analysis was performed by the ClueGO and CluePedia plug-ins of the Cytoscape software with adjusted P<0.05 [22]. Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment was carried out based on the clusterProfiler of R package with the P<0.05. The kappa coefficient is used for consistency testing and can also be used to measure classification accuracy [23]. In the ClueGO plug-in, the kappa coefficient shows the relationship between GO terms based on overlapping genes. The GO functions were grouped based on kappa coefficients, and the higher the kappa coefficient, the stronger the term association strength.

2.5. Construction of Protein-Protein Interaction (PPI). PPI analysis based on the target proteins was performed by the STRING database (Version: 10.0, http://www.string-db.org/) with a Required Confidence (combined score) > 0.4 [24]. After downloading the tsv format files, the PPI was constructed by the Cytoscape software [25].

2.6. Construction of Pharmacological Network. In order to more clearly display the regulation mechanism of effective components of *Achyranthes bidentata* on the osteoarthritis-related proteins, pharmacological network was constructed by Cytoscape software.

2.7. Verification of the Target Protein by Osteoarthritis mRNA Expressed Profiles. Osteoarthritis-related mRNA profiles of GSE55457 were used to verify the targeted proteins of effective components. From the GSE55457 data set, 10 samples from osteoarthritis patients (OA group) and 10 control knee cartilage samples (control group) were selected for further analysis. Briefly, raw data file was downloaded and read by affy package of R language software (Version 1.50.0, http://www.bioconductor.org/packages/release/bioc/ html/affy.html). Then, data standardization preprocessing was performed by robust multi-array average (RMA) method, including background correction, standardization and normalization. Subsequently, the probes were annotated based on the platform annotation files. Unmatched probes were given up and the average value was considered as the expression level if the different probes matched the same gene. Finally, the differentially expressed genes (DEGs) between OA and control groups were identified by limma package (Version 3.26.9, http://bioconductor.org/packages/release/bioc/html/limma.html) with the P Value<0.05 and |log 2 Fold Change (FC)| > 0.585 [26].

2.8. Docking Prediction of Key Target Proteins and Effective Components. The intersection of target proteins of effective
components and DEGs between OA and control groups was screened out for further study. The docking prediction of key target protein and effective components was carried out by the Protein Data Bank (PDB) database [27]. Briefly, the 3D structure of key target proteins was shown through PyMOL software (Version 2.1.1, https://pymol.org/dokuwiki/?id=). Besides, the experimental dissociation/inhibition constant value (pKd/pKi) of docking model was predicted by systems-Dock online tool [28, 29]. The pKd/pKi represented binding affinity, and the higher the score, the stronger the affinity (1∼10). In addition, the schematic diagram of protein-ligand interactions was drawn.

3. Results

3.1. The Overall Design of This Study and the Main Results. As shown in Figure 1, the effective components of Achyranthes bidentata were firstly screened out from the TCMSP database with ADME property parameters. Then, proteins targeted by the effective components were predicted based on the TCMSP and DrugBank database. Keyword "osteoarthritis" was entered into the CTD database to search for the relevant genes. Based on the above results, there were 26 members in the intersection. Subsequently, the enrichment analysis and the interaction network of these target proteins were performed and constructed. After validation by GSE55457, PIM1, CY1B1, and HSPA2 were considered as the key proteins during the treatment of osteoarthritis by quercetin. Finally, the docking models of PIM-quercetin, CY1B1-quercetin, and HSPA2-quercetin were predicted.

3.2. Effective Components of Achyranthes bidentata and the Target Proteins. A total of 176 kinds of herbal ingredients were searched for in the TCMSP database with the key word "Achyranthes bidentata". With the OB≥40% and DL≥0.2, 10 effective components of Achyranthes bidentata were obtained (Table 1). The above 10 components were searched for one by one in the DrugBank database. The results showed that there were two components with target records, namely, quercetin and kaempferol, with 29 targets and 1 and target, respectively. The network of quercetin and kaempferol with their target proteins was shown in Figure 2(a). There were 31 nodes (quercetin and kaempferol and their target proteins) and 30 relationship pairs in this network.

3.3. Enrichment Analysis of 26 Targeted Proteins of Quercetin. Enrichment analysis of 26 targeted proteins of quercetin was performed by the clusterProfiler package. As shown in Figure 3(a) the 26 proteins are mainly enriched in several pathways, such as Estrogen signaling pathway (ESR1/ESR2/HSP90AA1/HSPA2), Adherens junction (ACTB/CSNK2A1/CSNK2B), and Measles (CSNK2A1/CSNK2B/HSPA2). The
Table 1: Effective components of *Achyranthes bidentata*.

| Mol ID    | Molecule Name                  | OB  | DL  |
|-----------|--------------------------------|-----|-----|
| MOL001006 | poriferasta-7,22E-dien-3beta-ol| 42.98 | 0.76 |
| MOL012537 | Spinoside A                    | 41.75 | 0.4  |
| MOL012542 | $\beta$-ecdysterone            | 44.23 | 0.82 |
| MOL002776 | Baicalin                       | 40.12 | 0.75 |
| MOL002897 | epiberberine                   | 43.09 | 0.78 |
| MOL000422 | kaempferol                     | 41.88 | 0.24 |
| MOL004355 | Spinasterol                    | 42.98 | 0.76 |
| MOL000785 | palmatine                      | 64.6  | 0.65 |
| MOL000098 | quercetin                      | 46.43 | 0.28 |

Table 2: The list of 26 osteoarthritis related proteins.

The intersection of 29 targeted proteins of quercetin and the osteoarthritis related proteins in the CTD database.

| Gene ID | Molecule Name |
|---------|---------------|
| UGT3A1  | ATP5A1        |
| HCK     | ATP5B         |
| PIK3CG  | ATP5C1        |
| HSP90AA1| SF3B3         |

GO function enrichment analysis showed that the targeted proteins are mainly related to 13 biological process items (Figure 3(b)). Based on the kappa value, the GO items were divided into three categories, nitric oxide biosynthetic process, cellular response to estradiol stimulus, and mitochondrial ATP synthesis coupled proton transport (Figure 3(c)). Figure 3(d) showed the GO function network; we found that a total of 9 target proteins which belonged to quercetin were significantly enriched in the 13 GO items. The most important three Go items were mitochondrial ATP synthesis coupled proton transport (the matching target proteins were ATP5C1, ATP5A1, and ATP5B), cellular response to estradiol stimulus (the matching target proteins were ESRI, ESR2, and RUVBL2), and nitric oxide biosynthetic process (the matching target proteins were ESRI, CYP1B1, and HSP90AA1).

3.4. PPI Network Based on the Target Proteins of Quercetin. Figure 4 displayed the PPI network of the target proteins of quercetin. The connectivity in the network represented the correlation of two molecules. Based on the connectivity of proteins in the PPI network, the top 5 members were HSP90AA1 (degree was 13), ACTB (degree was 7), ESRI (degree was 7), ATP5B (degree was 5), and CYP1B1 (degree was 5).
Figure 3: The enrichment analysis of 26 key target protein of quercetin. (a) The bubble map of enriched pathways. The vertical axis was the name of the pathway, the horizontal axis was the number of enriched genes, and the dot size was the ratio of the number of enriched genes to the total number of uploaded genes. The redder the dot color, the more significant the P values. (b), (c) Go function enrichment analysis. The vertical axis presented the names of the GO items; the abscissa represented the percentage of the enriched genes. (d) ClueGO Functional Network Diagram. The dots with red labels were target proteins and circles were the GO functions. The larger the P value, the larger the circle. The lines between two points represented the correlation between functions, and the larger the kappa coefficient, the thicker the line.
3.5. **Pharmacological Network.** Pharmacological network was constructed based on the effective components, target proteins, and pathways. As shown in Figure 5, there are 51 nodes in the network, with 113 relationship pairs, including 2 herbal component nodes, 29 target protein nodes, and 20 pathway nodes.

3.6. **Verification of the Target Protein by GSE55457.** A total of 557 DEGs between OA and control groups were screen out from the GSE55457. Compared with the target proteins of quercetin, there were three common members, protooncogene serine/threonine-protein kinase Pim-1 (PIM1), cytochrome P450 1B1 (CYP1B1), and heat shock-related 70 kDa protein 2 (HSPA2), which were considered as the key proteins during the treatment of osteoarthritis by quercetin.

3.7. **Prediction of Key Target Protein and Quercetin Docking.** The secondary and tertiary structure of quercetin were
Figure 6: Structure predictions of quercetin, PIM-quercetin, CYPIBI-quercetin, and HSPA2-quercetin. (a) The secondary and tertiary structure of quercetin. (b) The 3D structures of PIM1 (left), CYPIBI (middle), and HSPA2 (right). (c) The diagrams of protein-ligand interaction of PIM-quercetin (left), CYPIBI-quercetin (middle), and HSPA2-quercetin (right). The interactions were mediated by hydrogen bonding and hydrophobic contact. The dashed lines indicated hydrogen bonds; the arcs indicated hydrophobic contacts. Black dots represented carbon, red dots were oxygen, and blue dots were nitrogen. The black character labels were amino acids.

downloaded from the DrugBank database and shown in Figure 6(a). Besides, the 3D structures of PIM1, CYPIBI, and HSPA2 were constructed and shown in Figure 6(b). After prediction of potential docking target of quercetin, the PBD IDs of PIM1, CYPIBI, and HSPA2 were 1XWS, 3PM0, and 4FSV, respectively. In addition, the corresponding docking scores (pKd/pKi) were 6.526, 6.968, and 6.334. Figure 6(c) showed the diagram of protein-ligand interaction of PIM-quercetin, CYPIBI-quercetin, and HSPA2-quercetin.

4. Discussion

This study aimed to analyze the potential mechanism of treatment of osteoarthritis by *Achyranthes bidentata*. After comprehensive analysis of TCMSP, DrugBank, and CTD databases by the key words of "*Achyranthes bidentata*" and "osteoarthritis". A total of 26 target proteins of quercetin related to the osteoarthritis were obtained, mainly enriched in the mitochondrial ATP synthesis coupled proton transport,
cellular response to estradiol stimulus, and nitric oxide biosynthetic process. By validation of DEGs of GSE55457, three common proteins, PIM1, CYP1B1, and HSPA2, were considered as the key targeted proteins of the quercetin during the treatment of osteoarthritis by *Achyranthes bidentata*.

Quercetin is categorized as a flavonol, one of the six subclasses of flavonoid compounds [30]. Plenty studies have reported that quercetin showed many potential beneficial effects on human health, such as antioxidant activity and effects against cancer, cardiovascular disease, diabetes and diabetic complications, hypertension, immunity and infections, and arthritis [31]. Application of quercetin might reduce symptoms of osteoarthritis. Matsuno et al. reported that pain symptoms, daily activities, visual analogue scale, and the synovial fluid properties were significantly improved after application of quercetin glucoside for osteoarthritis patients [32]. Besides, quercetin could inhibit the activities of many kinases implicated in cancer cell biology, such as ABL1, Aurora-A, -B, -C, CLK1, FLT3, JAK3, and MET [33]. In our study, quercetin was the most important effective component during the treatment of osteoarthritis by *Achyranthes bidentata* based on searching several databases.

The normal nitric oxide biosynthetic process is important for human health. Increasing of nitric oxide production was detected in osteoarthritic joints indicating that nitric oxide was involved in the pathogenesis of osteoarthritis [34]. The nitric oxide production was regional differences in the knee meniscus in response to dynamic compression [35]. We found that the 26 target proteins of quercetin related to the osteoarthritis were mainly enriched in the nitric oxide biosynthetic process, indicating that nitric oxide related signal may play an important role in the treatment of osteoarthritis by *Achyranthes bidentata*.

PIM1 is a serine/threonine kinase encoded by the protooncogene Pim-1. PIM1 is mainly involved in cell cycle progression, apoptosis, and transcriptional activation, as well as more general signal transduction pathways [36]. Merkel et al. reported that PIM1 kinase had the potential as a target for cancer therapy [37]. Besides, many reports have shown that the activity of PIM1 could be inhibited by quercetin [38, 39]. In our study, Figure 5 showed that the quercetin regulated the PIM1, which was involved in the breast cancer. However, few literature works studied the correlation between PIM1 and osteoarthritis. CYP1B1 belongs to the cytochrome P450 superfamily of enzymes, which catalyze many reactions related to drug metabolism and synthesis of cholesterol, steroids, and other lipids [40]. We found that CYP1B1 was the target of quercetin, which was consistent with the previous reports. Choi et al. reported that quercetin acted as an antioxidant and downregulated CYP1B1 against 7,12-dimethylbenz(a)anthracene-induced oxidative stress in mice [41]. Quercetin showed strong and selective inhibition against CYP1B1, which required 2-3 double bonds on the C-ring [42]. HSPA2 is a 70-kilodalton heat shock protein, which plays a key role in regulating the recognition of sperm and egg [43]. Wang et al. reported that as the target of quercetin, HSPA2 might act as the ATP binding domain [44]. Although, there were few reports on the directed interaction of PIM1, CYP1B1, HSPA2, and osteoarthritis, the docking of PIM1-quercetin, CYP1B1-quercetin, and HSPA2-quercetin may play important roles during the treatment of osteoarthritis by *Achyranthes bidentata*. The expression levels and activities of PIM1, CYP1B1, and HSPA2 in the animal models and clinical samples have been arranged into our next research plan.

In conclusion, the docking of PIM1-quercetin, CYP1B1-quercetin, and HSPA2-quercetin may play important roles during the treatment of osteoarthritis by *Achyranthes bidentata*.

**Data Availability**

The data used to support the findings of this study are available from the corresponding author upon request.

**Disclosure**

Dujun Ma and Tian Yu should be regard as co-first authors.

**Conflicts of Interest**

The authors declare that they have no conflicts of interest.

**Acknowledgments**

This work was supported by National Natural Science Foundation of China (grant number 81804124) and National Science Foundation of Guangdong Province, China (grant number 2018A0303130138).

**References**

[1] M. B. Goldring and S. R. Goldring, "Osteoarthritis," *Journal of Cellular Physiology*, vol. 213, no. 3, pp. 626–634, 2007.

[2] J. W. J. Bijlsma, F. Berenbaum, and F. P. J. G. Lafeber, "Osteoarthritis: an update with relevance for clinical practice," *The Lancet*, vol. 377, no. 9783, pp. 2115–2126, 2011.

[3] L. Brosseau, G. A. Wells, G. P. Kenny et al., “The implementation of a community-based aerobic walking program for mild to moderate knee osteoarthritis (OA): a knowledge translation (KT) randomized controlled trial (RCT): part I: the Uptake of the Ottawa Panel clinical practice guidelines (CPGs),” *BMC Public Health*, vol. 12, article 871, 2012.

[4] J.-Y. Reginster, A. Neuprez, M.-P. Lecart, N. Sarlet, and O. Bruyere, "Role of glucosamine in the treatment for osteoarthritis," *Rheumatology International*, vol. 32, no. 10, pp. 2959–2967, 2012.

[5] F. P. Chen, C. M. Chang, S. J. Hwang, Y. C. Chen, and F. J. Chen, “Chinese herbal prescriptions for osteoarthritis in Taiwan: analysis of national health insurance dataset,” *BMC Complementary & Alternative Medicine*, vol. 14, no. 1, p. 91, 2014.

[6] S. Teekachunhatean, P. Kunanusorn, N. Rojanasthien et al., “Chinese herbal recipe versus diclofenac in symptomatic treatment of osteoarthritis of the knee: a randomized controlled trial [ISRCTN70292892],” *BMC Complementary and Alternative Medicine*, vol. 4, no. 1, article 19, 2004.

[7] L. Long, K. Soeken, and E. Ernst, “Herbal medicines for the treatment of osteoarthritis: a systematic review,” *Rheumatology*, vol. 40, no. 7, pp. 779–793, 2001.
Evidence-Based Complementary and Alternative Medicine

[8] S. Han, C. W. Lee, L. D. Yoon et al., “Prevention of arthritic inflammation using an oriental herbal combination BDX-1 isolated from Achyranthes bidentata and Atractylodes japonica,” Archives of Pharmacal Research, vol. 28, no. 8, pp. 902–908, 2005.

[9] X.-X. Xu, X.-H. Zhang, Y. Diao, and Y.-X. Huang, “Achyranthes bidentate saponins protect rat articular chondrocytes against interleukin-β-induced inflammation and apoptosis in vitro,” Kaohsiung Journal of Medical Sciences, vol. 33, no. 2, pp. 62–68, 2017.

[10] P. Lin, X. Weng, F. Liu, X. Liu, X. Li, and H. Ye, “Study on the mechanism of effective ingredients of Achyranthes bidentata in preventing and treating osteoarthritis,” Rheumatism and Arthritis, vol. 4, no. 2, pp. 56–59, 2015.

[11] M. Russo, C. Spagnuolo, I. Tedesco, S. Bilotto, and G. L. Russo, “The flavonoid quercetin in disease prevention and therapy: facts and fancies,” Biochemical Pharmacology, vol. 83, no. 1, pp. 6–15, 2012.

[12] A. Mollica, G. Zengin, M. Locatelli et al., “An assessment of the nutraceutical potential of Jugalans regia L. leaf powder in diabetic rats,” Food and Chemical Toxicology, vol. 107, pp. 554–564, 2017.

[13] A. Mollica, A. Stefanucci, G. Zengin et al., “Polyphenolic composition, enzyme inhibitory effects ex-vivo and in-vivo studies on two Brassicaceae of north-central Italy,” Biomedicine & Pharmacotherapy, vol. 107, pp. 129–138, 2018.

[14] X. Liang, H. Li, and S. Li, “A novel network pharmacology approach to analyse traditional herbal formulae: the Liu-Wei-Di-Huang pill as a case study,” Molecular Biosystems, vol. 10, no. 5, pp. 1014–1022, 2014.

[15] B. Zhang, X. Wang, and S. Li, “An integrative platform of TCM network pharmacology and its application on a herbal formula, Qing-Luo-Yin,” Evidence-Based Complementary and Alternative Medicine, vol. 2013, Article ID 456747, 12 pages, 2013.

[16] M. Fawzi Mahomoodyal, A. Mollica, A. Stefanucci, M. Zakariy-yah Aumeeruddy, R. Poorneeka, and G. Zengin, “Volatile components, pharmacological profile, and computational studies of essential oil from Aegle marmelos (Bael) leaves: A functional approach,” Industrial Crops and Products, vol. 126, pp. 13–21, 2018.

[17] J. Ru, P. Li, J. Wang et al., “TCMSP: a database of systems pharmacology for drug discovery from herbal medicines,” Journal of Cheminformatics, vol. 6, no. 1, article 13, 2014.

[18] J. C. Madden, In Silico Approaches for Predicting Adm Properties, 2010.

[19] P. Ertl, B. Rohde, and P. Selzer, “Fast calculation of molecular polar surface area as a sum of fragment-based contributions and its application to the prediction of drug transport properties,” Journal of Medicinal Chemistry, vol. 43, no. 20, pp. 3714–3717, 2000.

[20] D. S. Wishart, Y. D. Feunang, A. C. Guo et al., “DrugBank 5.0: A major update to the DrugBank database for 2018,” Nucleic Acids Research, vol. 46, no. 1, pp. DI074–DI082, 2018.

[21] A. P. Davis, C. G. Murphy, R. Johnson et al., “The comparative toxicogenomics database: update 2011,” Nucleic Acids Research, vol. 41, no. 1, pp. DI104–DI114, 2011.

[22] G. Bindea, B. Mlecnik, H. Hackl et al., “ChuGO: A Cytoscape plug-in to decipher functionally grouped gene ontology and pathway annotation networks,” Bioinformatics, vol. 25, no. 8, pp. 1091–1093, 2009.

[23] D. W. Huang, B. T. Sherman, Q. Tan et al., “The DAVID gene functional classification tool: a novel biological module-centric algorithm to functionally analyze large gene lists,” Genome Biology, vol. 8, no. 9, article R183, 2007.

[24] D. Szklarczyk, A. Franceschini, S. Wyder et al., “STRING v10: protein-protein interaction networks, integrated over the tree of life,” Nucleic Acids Research, vol. 43, pp. D447–D452, 2015.

[25] M. Kohl, S. Wiese, and B. Warscheid, “Cytoscape: software for visualization and analysis of biological networks,” Methods in Molecular Biology (Clifton, N.J.), vol. 696, pp. 291–303, 2011.

[26] G. K. Smyth, “Limma: linear models for microarray data,” Bioinformatics & Computational Biology Solutions Using R & Bioconductor, pp. 397–420, 2005.

[27] H. M. Berman, J. Westbrook, Z. Feng et al., “The protein data bank,” Nucleic Acids Research, vol. 28, no. 1, pp. 235–242, 2000.

[28] K. Hsin, Y. Matsuoka, Y. Asai et al., “systemsDock: a web server for network pharmacology-based prediction and analysis,” Nucleic Acids Research, vol. 44, no. W1, pp. W507–W513, 2016.

[29] K.-Y. Hsin, S. Ghosh, and H. Kitano, “Combining machine learning systems and multiple docking simulation packages to improve docking prediction reliability for network pharmacology,” PLoS ONE, vol. 8, no. 12, Article ID e83922, 2013.

[30] J. Æ. Ross and C. M. Kasum, “Dietary flavonoids: bioavailability, metabolic effects, and safety,” Annual Review of Nutrition, vol. 22, pp. 19–34, 2002.

[31] G. S. Kelly, “Quercetin,” Alternative Medicine Review, vol. 16, no. 2, pp. 172–195, 2011.

[32] H. Matsuno, H. Nakamura, K. Katayama et al., “Effects of an oral administration of glucosamine-chondroitin-quinacetin glucoside on the synovial fluid properties in patients with osteoarthritis and rheumatoid arthritis,” Journal of the Agricultural Chemical Society of Japan, vol. 73, no. 2, pp. 288–292, 2009.

[33] R. Boly, T. Gras, T. Lamkami et al., “Quercetin inhibits a large panel of kinases implicated in cancer cell biology,” International Journal of Oncology, vol. 38, no. 3, pp. 833–842, 2011.

[34] K. Vuoletennaho, T. Moilanen, R. G. Knowles, and E. Moilanen, “The role of nitric oxide in osteoarthritis,” Scandinavian Journal of Rheumatology, vol. 36, no. 4, pp. 247–258, 2007.

[35] A. Hennerbichler, B. Fennor, D. Hennerbichler, J. B. Weinberg, and F. Guilak, “Regional differences in prostaglandin E2 and nitric oxide production in the knee meniscus in response to dynamic compression,” Biochemical and Biophysical Research Communications, vol. 358, no. 4, pp. 1047–1053, 2007.

[36] M. Bachmann and T. Möröyi, “The serine/threonine kinase Pim-1,” The International Journal of Biochemistry & Cell Biology, vol. 37, no. 4, pp. 726–730, 2005.

[37] A. L. Merkel, E. Meggers, and M. Ocker, “PIM1 kinase as a target for cancer therapy,” Expert Opinion on Investigational Drugs, vol. 21, no. 4, pp. 425–436, 2012.

[38] B. Baron, M. Thirman, M. Giurcanu, and J. Baron, “Quercetin therapy for selected patients with PIM1 kinase-positive chronic lymphocytic leukemia/small lymphocytic lymphoma: a pilot study,” Acta Haematologica, vol. 139, no. 2, pp. 132–139, 2018.

[39] K. Tsuchanewaza, H. Watanabe, L. Parker et al., “A novel Pim-1 kinase inhibitor targeting residues that bind the substrate peptide,” Journal of Molecular Biology, vol. 417, no. 3, pp. 240–252, 2012.
[41] E. Choi, T. Kim, and G.-H. Kim, “Quercetin acts as an antioxidant and downregulates CYP1A1 and CYP1B1 against DMBA-induced oxidative stress in mice,” Oncology Reports, vol. 28, no. 1, pp. 291–296, 2012.

[42] H. Takemura, T. Itoh, K. Yamamoto, H. Sakakibara, and K. Shimoi, “Selective inhibition of methoxyflavonoids on human CYP1B1 activity,” Bioorganic & Medicinal Chemistry, vol. 18, no. 17, pp. 6310–6315, 2010.

[43] K. A. Redgrove, B. Nixon, M. A. Baker et al., “The molecular chaperone HSPA2 plays a key role in regulating the expression of sperm surface receptors that mediate sperm-egg recognition,” PLoS ONE, vol. 7, no. 11, p. e50851, 2012.

[44] R. E. Wang, C. R. Hunt, J. Chen, and J.-S. Taylor, “Biotinylated quercetin as an intrinsic photoaffinity proteomics probe for the identification of quercetin target proteins,” Bioorganic & Medicinal Chemistry, vol. 19, no. 16, pp. 4710–4720, 2011.