Inferring Epidemiological Dynamics with Bayesian Coalescent Inference: The Merits of Deterministic and Stochastic Models

Alex Popinga,* Tim Vaughan,*† Tanja Stadler,§** and Alexei J. Drummond*†1

*Department of Computer Science, University of Auckland, Auckland, New Zealand 1010, †Allan Wilson Centre for Molecular Ecology and Evolution and ‡Massey University, Palmerston North, New Zealand 4442, §Department of Biosystems Science and Engineering, ETH Zürich, Basel, Switzerland 4058, and **Swiss Institute of Bioinformatics (SIB), Switzerland

ORCID ID: 0000-0003-4454-2576 (A.J.D.)

ABSTRACT Estimation of epidemiological and population parameters from molecular sequence data has become central to the understanding of infectious disease dynamics. Various models have been proposed to infer details of the dynamics that describe epidemic progression. These include inference approaches derived from Kingman’s coalescent theory. Here, we use recently described coalescent theory for epidemic dynamics to develop stochastic and deterministic coalescent susceptible–infected–removed (SIR) tree priors. We implement these in a Bayesian phylogenetic inference framework to permit joint estimation of SIR epidemic parameters and the sample genealogy. We assess the performance of the two coalescent models and also juxtapose results obtained with a recently published birth–death-sampling model for epidemic inference. Comparisons are made by analyzing sets of genealogies simulated under precisely known epidemiological parameters. Additionally, we analyze influenza A (H1N1) sequence data sampled in the Canterbury region of New Zealand and HIV-1 sequence data obtained from known United Kingdom infection clusters. We show that both coalescent SIR models are effective at estimating epidemiological parameters from data with large fundamental reproductive number $R_0$ and large population size $S_0$. Furthermore, we find that the stochastic variant generally outperforms its deterministic counterpart in terms of error, bias, and highest posterior density coverage, particularly for smaller $R_0$ and $S_0$. However, each of these inference models is shown to have undesirable properties in certain circumstances, especially for epidemic outbreaks with $R_0$ close to one or with small effective susceptible populations.

KEYWORDS Bayesian inference; phylodynamics; coalescent; epidemic; stochastic

Phylodynamics and the Coalescent

The epidemiological and evolutionary processes that underpin rapidly evolving species occur on a shared spatiotemporal frame of reference. Unified analyses that include both the dynamics of an epidemic and the reconstruction of the pathogen phylogeny can therefore uncover otherwise inaccessible information to aid in outbreak prevention. Such information includes the rates of pathogen transmission and host recovery, effective population sizes, and the “time of origin” representing the introduction of the first infected individual into a population of susceptible hosts.

The term phylodynamics was popularized by Grenfell et al. (2004) to describe the interlaced study of immunodynamics, epidemiology, and evolutionary mechanisms. Several phylodynamic models, both stochastic and deterministic in nature, have since been developed to characterize the phylogenetic history of the pathogen species and compartmentalizations of the host population throughout the epidemic. Such models grant the ability to infer key epidemiological parameters from genetic sequence data and include birth–death branching processes (Stadler et al. 2012, 2013; Gavryushkina et al. 2014; Kühnert et al. 2014), as well as coalescent approaches (Griffiths and Tavaré 1994; Pybus et al. 2001; Rasmussen et al. 2011, 2014; Koelle and Rasmussen 2012; Dearlove and Wilson 2013) derived from Kingman’s coalescent theory (Kingman 1982).

Significant steps toward the unification of epidemiology and statistical phylogenetics were made by Pybus et al. (2001),
Volz et al. (2009), and Dearlove and Wilson (2013), with the
formalization and application of Kingman’s n-coalescent to
pathogen population dynamics. These methods involved nu-
merical integration of a set of ordinary differential equations
(ODEs) to find deterministic approximations to the variation in
the number of sampled lineages through time. Volz (2012)
extended the tree density calculation from previous work (Volz
et al. 2009) to allow for serially sampled and spatially struc-
tured genetic sequence data. In this coalescent model, the birth
and death rates can vary in time and by the state of the host, so
that “the birth rate of a single gene copy is both time- and
state-dependent” (Volz 2012, p. 7).

In this article, we assess the ability of coalescent-based
phylodynamic models to infer, in a Bayesian setting, a range
of epidemiological parameters from simulated data. While
Dearlove and Wilson (2013) paved the way by implementing
a coalescent approach for deterministic susceptible–infected
(SI), susceptible–infected–susceptible (SIS), and suscep-
tible–infected–removed (SIR) models for Bayesian inference, we
implement and rigorously test both deterministic and stochas-
tic coalescent SIR models of epidemic dynamics extended for
heterochronously sampled data.

Stochastic and Deterministic Models

Stochasticity and determinism in population sizes each main-
tain dominant roles in particular stages of an epidemic. Once
the infected population has grown considerably large, on the
order of 1000–10,000 lineages, the probability densities of
stochastically expressed population size dynamics converge to-
ward the deterministic interpretation (Rouzine et al. 2001). How-
ever, during the early stages the population size of infected
individuals is small, and the dynamics of the epidemic are
therefore governed by stochastic processes due to the relative
significance of fluctuations in the demographic and rate pa-
rameters of the population model (Kühnert et al. 2014). Therefore,
approximating the prevalence of infection by a deterministic
function requires the number of infected hosts within the ef-
fective population to be assumed as very large throughout the
duration of the described epidemic, i.e., once the exponen-
tial growth phase has been reached (Rouzine et al. 2001).

Population size is critical to the epidemiological system
and, as with any parameter in a Bayesian setting, yields the
most accurate estimations when detailed prior information is
available and incorporated into the inference (Drummond
et al. 2006). In our extension and implementation of the
coalessent model for epidemics, both stochastic and deter-
ministic population size processes are used for the simul-
ation of trees and/or trajectories for subsequent inference.

Compartmental Population Models (SIR)

Host populations can be compartmentalized simply but
effectively in mathematical models that describe epidemic
progression. The specific division of the aggregate population
depends on the contagion, spanning a range of scenarios
where hosts may or may not recover from infection, may or
may not be reinfeeted, etc. Such examples include the SI, SIS,
and SIR models (Anderson and May 1991; Keeling and
Rohani 2008). Each of these compartments can be expressed
either (a) by a set of ODEs that describe the deterministic
time development of real-valued compartment occupations or
(b) in terms of integer-valued occupations governed by con-
tinuous-time Markov chains (CTMC) that allow for a degree
of uncertainty in the timing and number of events that occur
over the course of the epidemic.

In this article, we concentrate on the SIR model, which
describes epidemics that include infected individuals who
are at some point in time removed from the effective
population by way of immunity, death, behavioral changes,
or some other termination of infectiousness. The deter-
ministic variant of this model was introduced by Kermack
and Mckendrick (1932) and is given by the trio of coupled
ODEs,

\[ \frac{d}{dt} S(t) = -\beta I(t)S(t), \]

\[ \frac{d}{dt} I(t) = \beta I(t)S(t) - \gamma I(t), \]

\[ \frac{d}{dt} R(t) = \gamma I(t), \]

where \( \beta \) and \( \gamma \) respectively represent the transition rates
from susceptible \( S \) to infected \( I \) and infected \( I \) to removed
\( R \). The model fully defines the population dynamics with
initial conditions \( S(z_0) \), \( I(z_0) \), and \( R(z_0) \). It is worth recog-
nizing that, in the closed SIR model used here, there is no
demographic change in the host population. Therefore,
\( \frac{d}{dt} S(t) + \frac{d}{dt} I(t) + \frac{d}{dt} R(t) = 0 \) and \( S(t) + I(t) + \frac{d}{dt} R(t) = N \),
where \( N \) is the constant total population size.
Throughout this article we refer to the solutions to Equa-
tions 1–3 as deterministic SIR trajectories.

The comparable stochastic description is given in terms of
the probability of the epidemic state at time \( t \) given its initial
state and the rate parameters

\[ \pi(s, i, r; t) = \Pr(S(t) = s, I(t) = i, R(t) = r | S(0), I(0), R(0), \beta, \gamma), \]

which is governed by the following equation of motion:

\[ \frac{d}{dt} \pi(s, i, r; t) = \beta [(s + 1)(i - 1)\pi(s + 1, i - 1, r; t) - si\pi(s, i, r; t)] + \gamma [i + 1] \pi(s, i + 1, r - 1; t) - i\pi(s, i, r; t). \]

An explicit sampling process is incorporated by allowing
each removal event to coincide with a sampling event with
a fixed probability \( \psi/\psi + \mu \), where \( \psi \) and \( \mu \) are the overall
rates of sampled and unsampled removals, respectively, such
that \( \gamma = \psi + \mu \). We refer to epidemic histories sampled from
this model as stochastic SIR trajectories.
Both types of epidemic trajectories can be related to models of sampled transmission tree genealogies. In the deterministic case, this relationship is made via the coalescent distributions described in Volz (2012). We call this the deterministic coalescent SIR model. In the stochastic case, genealogies appear naturally from a branching process in which the branching events coincide with the transmission events in the CTMC and only those lineages ancestral to sampled removals are recorded. We call this the stochastic SIR model.

Another way of relating the stochastic SIR model to sampled transmission trees involves drawing a realization of a stochastic SIR epidemic and then using the coalescent distribution in Volz (2012) to produce a tree conditional on the particular piecewise constant infected compartment size corresponding to that realization. We call this approach the stochastic coalescent SIR model. Unlike BDSIR, the stochastic coalescent SIR model does not require the sampling process to be specified explicitly.

Both the transmission rate $\beta$ and the removal rate $\gamma$ can be estimated using each of the methods considered in this article from data ascribed to an SIR epidemic.

**Methods**

**Inference framework**

All phylodynamic inference discussed in this article is based on the joint posterior probability density

$$f(T, V, \eta, \theta | D) = \frac{\Pr(D | T, \theta) f(T | V, \eta) f(V | \eta) f(\eta | \theta) f(\theta)}{\Pr(D)},$$

where the sampled transmission tree $T$, the epidemic trajectory denoted $V = (S, I, R)$, the substitution parameters $\theta$, and the epidemiological parameters $\eta = \{\beta, \gamma, S_0, z_0\}$ are all estimated from the sequence data. The sampled transmission tree $T$ is assumed to be identical to the pathogen genealogy.

Here, $S$, $I$, and $R$ represent the host compartment sizes from the present time $t = 0$ back to the origin $z_0$, such that $S(t) = S(z_0 - t), I(t) = I(z_0 - t)$, and $R(t) = R(z_0 - t)$.

The various terms making up the right-hand side of Equation 6 are the tree likelihood $\Pr(D | T, \theta)$, the tree prior $f(T | V, \eta)$, the epidemic trajectory density $f(V | \eta)$, and the substitution and epidemiological parameter priors $f(\eta)$ and $f(\theta)$. The probability $\Pr(D)$ is merely a normalizing constant and can be ignored. It is the product of the tree prior and trajectory density $f(T | V, \eta) f(V | \eta)$ that distinguishes each of the models considered in this article.

For both the deterministic and stochastic coalescent SIR models, the tree prior $f(T | V, \eta)$ is calculated in the following way. First, consider the time span of a tree divided into segments bracketed by both sampling and coalescent events. By considering intervals ending in sampling events as well as coalescent-ending intervals, we follow previous work that extended coalescent approaches to time-stamped, serially sampled data (Rodrigo and Felsenstein 1999; Drummond et al. 2002). Interval $i$ is spanned by $k_i$ lineages and is the $i$th interval when ordered from the most recent tip to the root. The set of intervals $A$ ending in sample events and the set of intervals $Y$ ending in coalescent events together encompass all intervals, $V = A \cup Y$. Let the end time of an interval be $\tau_i$ (going back in time), with $\tau_0 = 0$ as the time of the most recent tip and with time increasing into the past. Then the probability density of a genealogy given an epidemic trajectory is

$$f(T | V, \eta) = \prod_{i \in Y} \lambda_k(\tau_i) \prod_{i \in V} \omega(\tau_i, k_i),$$

where $\lambda_k(\tau)$ is the instantaneous coalescent rate at $\tau$ prescribed by Volz (2012),

$$\lambda_k(\tau) = \left(\frac{k_i}{2}\right) \frac{2B S(\tau)}{I(\tau)},$$

and where $\omega(\tau_i, k_i)$ is the survival probability

$$\omega(\tau_i, k_i) = \exp\left(-\int_{\tau_{i-1}}^{\tau_i} \lambda_k(\tau) d\tau\right).$$

The deterministic coalescent SIR model assumes that the SIR epidemic trajectories are found by integrating the ODEs in Equations 1–3. Therefore, under this model each epidemic trajectory is a deterministic function of its parameters $V(\eta)$. This means that the trajectory density can be written as

$$f(V | \eta) = \delta(V - V(\eta)),$$

where $\delta(x)$ is the Dirac $\delta$-function and represents a point mass concentrated at $x = 0$.

In contrast, the stochastic coalescent SIR model assumes that the epidemic is generated by a jump process corresponding to the master equation given in Equation 5. In this case, the probability $f(V | \eta)$ is nonsingular and thus contributes to the uncertainty in the final inference result.

In the BDSIR model introduced by Kühnert et al. (2014), $f(V | \eta)$ is the same as for the stochastic coalescent SIR model, but $f(T | V, \eta)$ is defined differently. See Kühnert et al. (2014) for details.

**Markov chain Monte Carlo algorithm**

We use Markov chain Monte Carlo (MCMC) to sample from the joint posterior density given in Equation 6. Many of the specifics of the algorithm used have been discussed previously, in particular the method for calculating the tree likelihood (Felsenstein 1981, 2004) and the mechanism for exploring tree space (Drummond et al. 2002). However, the model-specific product $f(T | V, \eta) f(V | \eta)$ requires special attention.

As we are primarily interested in parametric inference rather than the epidemic trajectory itself, we can regard $V$ as a nuisance parameter to be marginalized over. This marginalization can be achieved implicitly by sampling it using MCMC and then ignoring this component of the sampled state, which is the strategy we use when reporting the BDSIR results. It can also be made an explicit part of the likelihood calculation, which is the approach we take with the deterministic
and stochastic coalescent SIR models. This marginalization
means that the product \( f(T|\mathcal{V}, \eta) f(\mathcal{V}|\eta) \) becomes
\[
  f(T|\eta) = \int f(T|\mathcal{V}, \eta) f(\mathcal{V}|\eta) d\mathcal{V},
\]
the probability density of the tree given the epidemiological parameters.

In the case of the deterministic coalescent SIR model, this density reduces to
\( f(T|\mathcal{V}(\eta), \eta) \), meaning that the density of the tree given epidemiological parameters \( \eta \) is obtained simply by substituting the numerical solution to Equations 1–3 for those parameters into Equation 7.

The stochastic coalescent SIR model is more complex, as in this case the trajectory density \( f(\mathcal{V}|\eta) \) is nonsingular, meaning that computing the integral in Equation 11 is nontrivial. We treat this here using the “pseudomarginal” approach (Beaumont 2003; Andrieu and Roberts 2009) in which, at each step in the MCMC chain, the marginalized tree density \( f(T|\eta) \) is replaced by the Monte Carlo estimate
\[
  \hat{f}(T|\eta) = \frac{1}{M} \sum_{r=1}^{M} f(T|\mathcal{V}_r, \eta),
\]
where each \( \mathcal{V}_r \) is a trajectory sampled independently from \( f(\mathcal{V}|\eta) \), using a stochastic simulation algorithm (Sehl et al. 2009). Perhaps counterintuitively within an MCMC framework, this stochastic likelihood converges to the true marginal posterior distribution regardless of the number \( M \) of realizations used in the estimate. However, the magnitude of \( M \) can significantly affect the rate at which the chain produces effectively independent samples from the posterior and must be tuned carefully.
Implementation and validation

We have implemented the schemes described above for performing inference under the deterministic and stochastic coalescent SIR models within the BEAST 2 phylodynamics package found at http://github.com/CompEvol/phylodynamics. This has a number of advantages over a stand-alone implementation. Foremost, we were able to avoid reimplementing components of the algorithm that are in common with other already-implemented phylogenetic and phylodynamic analyses, such as the MCMC proposal operators used to traverse the parameter space. Furthermore, this greatly increases the usefulness of the implementation, as it can be immediately used in conjunction with a wide variety of nucleotide and amino acid substitution models and parameter priors.

We have taken two steps to ensure our implementation is correct. First, we compared tree probability density \( f(T|V, \eta) \) values calculated using the main implementation of each of the two models with those calculated using completely independent implementations in R (R Core Team 2014).

Second, we used the implemented MCMC algorithms to sample transmission trees from the tree density given in Equation 11 for each model. We then compared the distributions of tree height, total edge length, and binary clade count summary statistics from these sampled ensembles with sample distributions obtained directly via stochastic simulation. As shown in Supporting Information, File S1, Figure S1, Figure S2, and Figure S3 (Sampling from the prior) and in the associated figures, the resulting pairs of distributions agree, providing strong support for our claim that the implementations of the methods described above are correct.

Instructions for downloading and using this package are also available on the project website located at http://github.com/CompEvol/phylodynamics.

Simulation study

To evaluate the implementation and extension of the coalescent models, we performed analyses on both sequence data and fixed trees simulated with known parameter values. The median estimated values produced by each model were then used to measure relative error and bias, along with the widths and coverage of 95% highest posterior density (HPD) intervals.

We used three methods for simulating the trees and trajectories, as shown below:

Inference model:
- Stoch. Coal. SIR
- Deter. Coal. SIR
- BDSIR

Simulation scheme:
- Stoch. Coal. SIR, Stoch. Coal. SIR, Stoch. Coal. SIR
- Deter. Coal. SIR, Deter. Coal. SIR, Deter. Coal. SIR
- Stochastic SIR, Stochastic SIR, Stochastic SIR.

The stochastic coalescent and deterministic coalescent simulation schemes were used to validate the coalescent SIR inference models. The stochastic SIR scheme, contrarily, is emphasized for its realistic properties.

Stochastic SIR trees and trajectories were generated using master equations in the simulation package MASTER (Vaughan and Drummond 2013). Deterministic coalescent trajectories were generated using a Runge–Kutta integrator (Runge 1895; Kutta 1901) with adaptive step sizes to solve a system of first order ODEs. Stochastic coalescent trajectories were generated using Sehl et al.’s (2009) SAL \( \tau \)-leaping algorithm (Sehl et al. 2009).

To simulate the stochastic coalescent SIR trees, we used the stochastic SIR trajectories, which could be converted to effective population size with the mathematical expression used to obtain Volz’s (2012) coalescent rate for the SIR model: \( N_e(\tau) = 1/\tau^2 = I(\tau)/(2\beta S(\tau)) \). The sampling times, generated by a sampling rate \( \psi \), for the stochastic
coalescent SIR trees were also taken from the MASTER output to allow for direct comparison between the sets of trees. In other words, the underlying epidemic function was the same for both stochastic SIR and stochastic coalescent SIR trees, the latter of which were then simulated under a piecewise constant population function.

Likewise, for the simulation of deterministic coalescent trees we used deterministic SIR trajectories to construct a population function and the relation $N_e = I/(2βS)$ to convert infected and susceptible host population sizes to effective population size. The sampling times were randomly generated from a probability distribution so that the density of samples taken through time was proportional to the number of infected individuals through time, as with the stochastic SIR trees.

We simulated stochastic SIR trees, using multiple combinations of parameter values. We were particularly interested in varying the basic reproductive ratio $R_0$ and the initial susceptible population size $S_0$, to observe the changes in relative error, bias, and uncertainty in stochastic and deterministic models. To alter the ratio $R_0 = βS_0/γ$ and still generate sensible trees with a consistent number of tips, one or more of the other parameters (birth rate β, removal rate γ, or $S_0$) must also change. Table 2, Table S6, Table S7, and Table S9 show the true values of the parameters for each set of simulations. (The birth rate β is not shown, as our implementation allows either β or $R_0$ to serve as a parameter in the inference, and $R_0$ is the parameter of interest. However, β can be calculated via the other three, using $β = R_0γ/S_0$. For example, when $R_0 = 1.0978$, $S_0 = 499$, and $γ = 0.25$, then $β = 5.50E+4$.)

**Heterochronous trees:** We generated 100 trees under each of the three (stochastic SIR, stochastic coalescent SIR, and deterministic coalescent SIR) models with parameters $S_0$, $β$, and $γ$. For heterochronously sampled trees, each removal generates a sample with probability $ψ/(ψ + μ)$, where $ψ$ is the overall rate of sampled removals and $μ$ is the rate of unsampled removals such that $γ = ψ + μ$.

The simulations ended once the number of infected individuals reached zero, i.e., when the last infected individual was removed. This ensured that the simulated trajectories spanned past the exponential growth phase of the epidemic and therefore included samples past the peak of infected individuals. This choice of procedure was motivated by (a) the suggestion of Stadler et al. (2014) that the behavior of the coalescent beyond the exponential phase could either inflate or reduce bias and (b) the observations of Dearlove and Wilson (2013) and Bošková et al. (2014) that deterministic coalescent SIR models might be properly fitted only once the epidemic has peaked. Figure 1 shows trajectories of susceptible, infected, and removed individuals underlying the simulation of stochastic SIR trees (Figure 2) generated in MASTER. An example XML for simulating these MASTER trees is provided in File S1.

We required that the trees had $n \geq 100$ leaves, filtering out those in which the epidemic died out in the early stages, i.e., when the initial infected individual was removed from the effective population too quickly to infect others. (Note that the inference procedures discussed in this article all implicitly condition on the number of leaves.) The probability that the first event in a given trajectory is the removal (by recovery, death, etc.) of patient zero is given by $δ/(βS_0 + δ) = 1/(1 + R_0)$. When $R_0 \approx 2.50$, this probability is $\approx 30\%$. In our case, $52/152$ (≈ 34%) trees were “empty” or containing only one node. The filtering process left us with a mean of $\approx 160$ leaves for the simulated trees.

**Homochronous trees:** A major concern in the comparison between Kühnert et al. (2014)’s birth–death-sampling SIR inference model, which includes explicit sampling, and our implementations of Volz (2012)’s coalescent SIR models, which do not include explicit sampling, is that the former is given extra information via the sampling process. Volz and Frost (2014) addressed this issue by providing a coalescent SIR model that does incorporate sampling explicitly.

That being said, results from Bošková et al. (2014) indicate that the poor performance of the deterministic coalescent SIR model in comparison with birth–death models was due to the lack of handling stochastic population size changes through time rather than the lack of information about the sampling proportion. Their results showed that
the coalescent is “very robust to changes in sampling schemes” (Boskova et al. 2014, p. 8).

Regardless, to ensure a fair comparison of BDSIR and the coalescent SIR models, we simulated an SIR epidemic with homochronous, or contemporaneous, sampling. This type of simulation affords no additional information about the population size for explicit-sampling models, as there is only a single time of sampling.

We selected a simulation time of \( t = 20 \) for the homochronously sampled trees, with the trajectories being sampled at high prevalence but also past the time of peak prevalence. This is important for distinguishing SIR from SI/SIS outbreaks, as it provides information about the removal parameter \( \gamma \). In this set of simulations, each lineage was sampled at \( t = 20 \) with probability 0.7 (the leaf count distribution for varied sampling probabilities is in File S1).

Simulated sequences: To assess the ability of each SIR model to infer epidemic parameters with the inclusion of phylogenetic uncertainty, we also simulated the evolution of 2000-bp sequences down each simulated tree. We time stamped the sequences with the tip dates of each corresponding tree and informed the inference with the true Hasegawa–Kishino–Yano (HKY) substitution model (Hasegawa et al. 1985), clock rate = SE-3, and \( \kappa = 5 \). These choices were made to reflect real data, specifically those of influenza (Vaughan et al. 2014).

Along with simulated sequence data, analyses were performed with the simulated trees fixed (results are in File S1), and the parameters \( R_0, S_0, \) and the origin of the tree \( z_0 \) were estimated with Bayesian prior distributions as listed in Table 4.

Deterministic coalescent SIR on higher \( R_0 \) and \( S_0 \): Finally, we had particular interest in the effects of varying the population size parameter \( S_0 \) on the deterministic coalescent SIR model, as comparisons from initial analyses with lower true \( R_0 \) (\( \approx 1.5 \) and \( \approx 1.1 \)) and \( S_0 \) (\( = 499 \)) showed higher error and bias and lower 95% HPD coverage. Also, it is often assumed that deterministic descriptions will perform well

Table 2 Simulation study results for fixed trees: \( R_0 = 2.50 \) and \( S_0 = 999 \). \( R_0 = 1.50 \) and \( S_0 = 499 \). and \( R_0 = 1.10 \) and \( S_0 = 499 \)

| \( \eta \) | Inference | Truth | Mean | Median | Error | Bias | Relative HPD width | 95% HPD accuracy (%) |
|---|---|---|---|---|---|---|---|---|
| \( R_0 \) | Stoch.Coal.SIR | 2.50 | 2.84 | 2.68 | 0.12 | 0.09 | 0.98 | 100.00 |
| | Deter.Coal.SIR | 2.50 | 2.68 | 2.49 | 0.13 | 0.04 | 0.81 | 98.00 |
| | BDSIR | 2.50 | 2.73 | 2.67 | 0.12 | 0.08 | 0.55 | 94.00 |
| \( \gamma \) | Stoch.Coal.SIR | 0.30 | 0.27 | 0.25 | 0.19 | \(-0.13\) | 1.14 | 99.00 |
| | Deter.Coal.SIR | 0.30 | 0.32 | 0.29 | 0.16 | 3.14E-3 | 1.27 | 99.00 |
| | BDSIR | 0.30 | 0.28 | 0.27 | 0.13 | \(-0.09\) | 0.62 | 95.00 |
| \( S_0 \) | Stoch.Coal.SIR | 999 | 1390 | 921 | 0.19 | \(-0.03\) | 3.85 | 100.00 |
| | Deter.Coal.SIR | 999 | 1807 | 1133 | 0.52 | 0.29 | 4.59 | 98.00 |
| | BDSIR | 999 | 1591 | 1142 | 0.39 | 0.24 | 3.42 | 99.00 |
| \( z_0 \) | Stoch.Coal.SIR | Varies | 41.81 | 40.35 | 0.03 | 0.01 | 0.20 | 99.00 |
| | Deter.Coal.SIR | Varies | 41.17 | 39.99 | 0.03 | 0.01 | 0.07 | 76.00 |
| | BDSIR | Varies | 40.89 | 39.72 | 0.65E-4 | \(-5.13E-4\) | 3.43E-3 | 97.00 |

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for higher $R_0$ and larger population sizes. Table S7 and Table S9 detail the parameter values we used to explore the behavior of the deterministic coalescent on varied $R_0$ and $S_0$ combinations.

**Interpretation of results**

We compared the coalescent SIR, as well as BDSIR, parameter estimations from the simulated data to the true values used to generate the SIR trajectories. Following Kühnert et al. (2014), the precision and accuracy of these methods were measured by relative error, bias, and HPD intervals. We used the posterior median value of the parameter value $\hat{\eta}$ compared with the true parameter $\eta \in \{R_0, \gamma, S_0, z_0\}$. Relative error and bias are then gauged by calculating the median value over medians from all 100 trees, such that

$$RE_\eta = \frac{\sum_{i=1}^{100} |\eta_i - \hat{\eta}|/\eta}{100}$$

and

$$RB_\eta = \frac{\sum_{i=1}^{100} |\eta_i - \hat{\eta}|/\eta}{100}.$$  

Measures of HPD interval widths are given by

\[
\text{95\% HPD upper bound} - \text{95\% HPD lower bound}.
\]

Table 1, Table 2, and Table 3 show these results, along with the percentages of posterior estimates that produced 95% HPD intervals containing the true values (i.e., 95% HPD coverage).

**H1N1 data analysis**

To test the efficacy of the coalescent SIR models on real data, epidemic parameters $R_0$, $\gamma$, $S_0$, and time of origin $z_0$ were estimated from 42 seasonal influenza A (H1N1) sequences sampled throughout the 2001 flu season in Canterbury, New Zealand.

Influenza infections are well known for their seasonal SIR behavior in nonequatorial populations, as each annual flu season begins with a supply of susceptible hosts and tapers off as the hosts recover with adaptive immunity (Iwasaki and Pillai 2014). Due partly to this seasonal pattern, the influenza virus is both a motivator for the development of specialized models and a prime subject for testing phylodynamic models (Koelle et al. 2006).

Sampling a particular region bypasses the necessity of specifying geographically structured populations, and New Zealand is an area of particular interest due to its geographic location and relative isolation from other regions with potentially varying dynamics. It is also assumed to play a key role in the global circulation of influenza strains (Rambaut and Holmes 2009; Bedford et al. 2010).

We used an HKY nucleotide substitution model, with a substitution rate of $5\times10^{-3}$ as estimated in Vaughan et al. (2014), and informed the models with dated sequences. Priors used for the Bayesian inference are shown in Table 4.

**HIV-1 data analysis**

In addition to our analysis of H1N1 sequence data, we selected HIV-1 subtype B nucleotide sequences collected from infected individuals located in the United Kingdom. The coalescent SIR results were collated with the results from the BDSIR data analysis performed by Kühnert et al. (2014), using the same sequences. More details of this analysis are provided in File S1.

**Results and Discussion**

**Simulation study**

Results for epidemic parameter inference from nucleotide sequences simulated from stochastic SIR trees are provided in Table 1 for $R_0 \approx 2.50$. Results for inference from fixed trees ($R_0 \approx 2.50$, $R_0 \approx 1.50$, $R_0 \approx 1.10$) are shown in Table 2, with 95% HPD coverage shown for each analysis in Figure 3. Inference results for analyses with true $R_0 = 1.0987$ and varying population size ($S_0 = 499, 999, 1999$) are described in Tables S1 and S2 in the supporting information, along with results from trees simulated under the stochastic and deterministic coalescent models for validation.

**Heterochronous trees:** For $R_0 \approx 2.50$, all three inference methods performed similarly for parameters $R_0$ and $\gamma$, with high 95% HPD coverage and low error and bias. The most weakly identifiable parameter $S_0$ yielded the largest HPD intervals for all three inference models. The deterministic coalescent returned higher error (0.52) and bias (0.29) than the stochastic coalescent SIR (0.19, −0.03) and BDSIR (0.39, 0.24) and recovered the origin parameter $z_0$ for only 76 of 100 simulated trees, while the stochastic coalescent and BDSIR respectively recovered $z_0$ for 99 and 97 of 100 simulations.

For $R_0 \approx 1.50$, the relative HPD widths (akin to variance) for three of the four estimated parameters ($R_0$, $\gamma$, and $z_0$) were smallest for BDSIR. For the parameter $S_0$, the relative HPD width is largest for BDSIR, although it also had slightly higher

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**Table 3 Epidemic parameter inference from H1N1 sequences in New Zealand**

| Inference model | $R_0$ | $\gamma$ | $S_0$ | Root of the tree (yr) | Origin $z_0$ of the epidemic (yr) |
|-----------------|-------|----------|-------|----------------------|-------------------------------|
| Stoch. Coal. SIR | 1.46 (1.04–2.14) | 27.08 (4.20–64.03) | 6.90E4 (175–2.86E5) | 0.53 (0.44–0.61) | 0.69 (0.45–1.03) |
| Deter. Coal. SIR | 1.35 (1.05–1.84) | 34.50 (3.86–82.16) | 1.20E5 (29–4.59E5) | 0.54 (0.45–0.62) | 0.73 (0.47–1.04) |
| BDSIR | 1.61 (1.09–2.29) | 27.72 (6.82–55.04) | 2.22E4 (259–9.38E4) | 0.49 (0.41–0.56) | 0.53 (0.43–0.65) |

Shown are mean estimates (and 95% HPD intervals) of each epidemic parameter inferred from seasonal influenza A (H1N1) sequence data collected in the Canterbury region of New Zealand throughout the 2001 flu season.
Table 4 Bayesian prior distributions

| Analysis       | $R_0$       | $\gamma$    | $S_{(0)}$ | $z_{(0)}$ | $\psi/(\phi + \mu)$ |
|----------------|-------------|-------------|-----------|-----------|---------------------|
| $R_0 \approx 2.5$, $S_0 = 999$ | LogN(1, 1)  | LogN(-1, 1) | LogN(7, 1) | Unif(0, 100) | Beta(1, 1)          |
| $R_0 \approx 1.5$, $S_0 = 499$ | LogN(0.5, 1) | LogN(-1, 1) | LogN(6, 1) | Unif(0, 500) | Beta(1, 1)          |
| $R_0 \approx 1.1$, $S_0 = 499$ | LogN(0.1, 1) | LogN(-1.5, 1) | LogN(6, 1) | Unif(0, 500) | Beta(1, 1)          |
| $R_0 \approx 1.1$, $S_0 = 999^a$ | LogN(0.1, 1) | LogN(-1.5, 1) | LogN(7, 1) | Unif(0, 500) | —                   |
| $R_0 \approx 1.1$, $S_0 = 1999^a$ | LogN(0.1, 1) | LogN(-1.5, 1) | LogN(7.5, 1) | Unif(0, 500) | —                   |
| $R_0 \approx 1.2$, $S_0 = 499^a$ | LogN(0.2, 1) | LogN(-1, 1) | LogN(6, 1) | Unif(0, 500) | —                   |
| H1N1          | Unif(0, 10) | LogN(3, 0.75) | LogN(13, 2) | Unif(0, 10) | Beta(1, 1)          |
| HIV-1         | LogN(1, 1)  | LogN(-1, 1) | LogN(7, 1) | Unif(0, 100) | Beta(1, 1)          |

Shown are prior distributions for the reestimation of SIR parameters—the reproductive ratio $R_0$, the rate of removal $\gamma$, the number of susceptible individuals at the start of the epidemic $S_{(0)}$, the time of origin $z_{(0)}$, and the sampling proportion $\psi/(\phi + \mu)$ for BDSIR—from the simulated trees, seasonal influenza A (H1N1), and human immunodeficiency virus (HIV-1) data analyses. LogN(M, S) is a log-normal distribution with mean $M$ and standard deviation $S$ in log space.

a Only applies to deterministic coalescent SIR; see details in File S1.

95% HPD coverage than deterministic coalescent SIR and the same as stochastic coalescent SIR. The deterministic coalescent SIR method recovered the truth for 85, 89, 91, and 88 of 100 trees for parameters $R_0$, $\gamma$, $S_0$, and $z_0$, while its stochastic analog recovered the truth for 100, 85, 100, and 99 of 100 trees for the same parameters. Finally, for stochastic coalescent SIR and BDSIR, error and (absolute) bias were relatively low for $R_0$, arguably the parameter of most interest to epidemiologists since it represents the number of individuals each infected individual will infect in a naive population. Deterministic coalescent SIR has a higher error (0.24) and bias (0.15) and also has significantly lower coverage for $R_0$ (85%).

For $R_0 \approx 1.10$, the two stochastic models again outperformed the deterministic coalescent in error, bias, and 95% HPD coverage. The stochastic coalescent most reliably recovered the truth for $R_0$ (99 of 100 simulations), while the deterministic coalescent had more than double the error and bias and still recovered the truth for only 25 of the 100 simulations. BDSIR had the lowest error and bias for $R_0$ under this scheme, although it recovered the truth for only 75 of 100 simulations. For removal parameter $\gamma$, BDSIR again yielded lower error and bias, in this case returning the truth for 100/100 trees (in contrast to 84 and 86 from the stochastic and deterministic coalescent, respectively).

In the stochastic models, there is a greater trade-off between parameters due to the impact the relationship between them has on the survival of trajectories at low $R_0$. A larger estimated removal rate tends to require a larger susceptible population for the epidemic to avoid dying out in the early stages. Likewise, a smaller susceptible population implies a smaller estimated $\gamma$.

**Deterministic coalescent SIR on higher $R_0$ and $S_0$:** As mentioned in the preceding subsection, the deterministic coalescent model yielded higher error and bias than both the stochastic coalescent and BDSIR for most parameters with $R_0 \approx 1.10$ and $S_0 = 499$.

To investigate the deterministic model’s sensitivity to population sizes, we also simulated a range of population sizes ($S_0 = 499, 999, and 1999$) for $R_0 = 1.0987$. Even with $S_0 = 1999$, the deterministic coalescent SIR model’s 95% HPD coverage was low. For parameters $R_0$, $\gamma$, $S_0$, and $z_0$, this coverage was respectively 40%, 64%, 66%, and 18%. Table S6 shows these results.

Additionally, we increased both $R_0$ (to 3.5 and 5) and $S_0$ (to 4999 and 9999). However, for parameters $R_0$, $\gamma$, and $S_0$, the deterministic coalescent SIR showed increased error, bias, and HPD widths, and the HPD coverage for $z_0$ did not improve. These results are shown in Table S9.

While each of these methods is an approximation, the deterministic coalescent particularly suffers from model misspecification since it does not account for the stochasticity that is always present in the early stages of epidemics, regardless of $S_0$.

**Homochronous trees:** Results for homochronously sampled trees are given in Table S3.

All three SIR inference models recover the truth for >95/100 trees within their respective 95% HPD widths for epidemic parameters $R_0$, $\gamma$, and $S_0$. The time of origin $z_0$ was recovered for 100/100 trees by BDSIR, 95/100 trees by stochastic coalescent SIR, and 73/100 trees by deterministic coalescent SIR. However, relative error and bias also increased consistently across all three models, along with the 95% HPD widths. The deterministic coalescent had the highest error, bias, and HPD width for $R_0$ and highest error and HPD width for $S_0$, which is consistent with the heterochronously sampled data.

Further consideration of the effects of sampling rate changes and sampling model misspecification are warranted for BDSIR and coalescent SIR, the latter of which has been facilitated by Volz and Frost (2014).

**Simulated sequences:** Relative error and bias were inflated across all three inference models with the addition of phylogenetic uncertainty, and in certain cases the 95% HPD coverage was lower than with fixed trees. The deterministic coalescent model recovered the truth within its 95% HPD intervals only for ≥90 of the 100 trees in the case of $S_0$. The true values for the parameters $R_0$, $\gamma$, and $z_0$ were covered by 95% HPD intervals for 87, 56, and 29 of the 100 trees, respectively. This is contrasted with the performance of the stochastic coalescent (100, 97, 47, and 37 for parameters $S_0$, $R_0$, $\gamma$, and $z_0$) and BDSIR (99, 100, 84, and 18 for $S_0$, $R_0$, $\gamma$, and $z_0$), as shown in Table 1.
Error, bias, and 95% HPD widths were higher with simulated sequences for all three inference models for parameters $g$, $S_0$, and $z_0$ than with fixed trees. This indicates the importance of calibrating epidemic parameters of interest. In our case, we emphasize the basic reproductive number $R_0$, often the parameter of most interest to epidemiologists. For $R_0$, stochastic coalescent SIR and BDSIR recovered the truth within their 95% HPD intervals for 97 and 100 of the 100 simulations, respectively. They also showed only slight changes in error and bias compared to inference performed on the fixed trees used to generate the sequences. The deterministic coalescent SIR model recovered $R_0$ for 87 of the 100 simulations (contrasted with 98/100 for the fixed trees) and with increased error.

**Priors and identifiability:** It is important to understand the impact of selected priors on inference results, as the priors are where the power of Bayesian inference lies. For example, we found relatively weak identifiability in the initial susceptible population parameter $S_0$, which must either be fixed or be estimated alongside the origin parameter $z_0$.

In addition to allowing each parameter to be either fixed or estimated, we have provided options for parameterization of our models, with either the transmission rate $\beta$ or $R_0$ acting as operable parameters in MCMC analysis. For the deterministic coalescent, there is also an option to use the intrinsic growth parameter described by Dearlove and Wilson (2013).

The choice of parameterization necessarily affects the prior that will be used in the inference and should be considered carefully. However, we found that once a parameterization has been selected, our inference models are robust to different prior distributions placed on each parameter. We also used broader prior distributions on the deterministic coalescent to test whether this would increase its lower 95% HPD coverage relative to the stochastic models. We found that doing so increased the error and bias of the results without increasing the accuracy (shown in Table S4).

**H1N1 data analysis**

Epidemic parameter estimates from serially sampled influenza A (H1N1) virus sequence data are shown in Table 3.

The estimated means of the basic reproductive number were $R_0 = 1.46$, 1.35, and 1.61 for the stochastic coalescent, the deterministic coalescent, and BDSIR, respectively. Estimates of $R_0$ from pandemic H1N1 in New Zealand range...
from ~1.2 to 1.5 (Paine et al. 2010; Opatowski et al. 2011; Roberts and Nishiura 2011; Roberts 2013; Biggerstaff et al. 2014), and estimates of $R_0$ for seasonal H1N1 from other countries also range from ~1.2 to 1.5 (Chowell et al. 2008). The 95% HPD intervals were very similar across each model, ranging from just over 1.0 to ~2.0.

The population of the Canterbury region in 2001 was reported to be ~481,431 by the Environment Canterbury Regional Council (Ecan 2001) and 521,832 by Statistics New Zealand (StatsNZ 2001). The mean estimates of $S_0$ were considerably lower using the stochastic coalescent ($S_0 = 69,000$), the deterministic coalescent ($S_0 = 120,000$), and BDSIR ($S_0 = 22,200$). However, the effective population of susceptibles is assumed to be much smaller, as the total population contains individuals of various susceptibility, e.g., those with partial immunity from vaccination and previous or secondary infections.

Most people recover from flu symptoms, the time they are likely to be most infectious, within a few days up to 2 weeks (CDC 2014; WHO 2014). This provides a range of probable true values for the removal parameter $\gamma$. The sequence data and molecular clock rate, and therefore the tree, are in units of years. Therefore, our $\gamma$ range would be from $365/14$ days to $365/2$ days or from $\gamma = 26.1$ to $\gamma = 182.5$. The stochastic coalescent, the deterministic coalescent, and BDSIR respectively inferred $\gamma$ means of 27.08, 34.50, and 27.72. These estimates are on the low side compared to epidemiological models for influenza that include explicit spatial and household effects (Ferguson et al. 2005), but a moderate misfit of the model is not unexpected when fitting a simple closed SIR model with no population substructure.

The root of the tree was very similar across all inference models, respectively 0.53, 0.54, and 0.49 for stochastic coalescent SIR, deterministic coalescent SIR, and BDSIR. The same was true for the origin $z_0$, with: 0.69, 0.73, and 0.53 for the stochastic coalescent, the deterministic coalescent, and BDSIR. All three inference models returned tree root and origin estimates that are consistent with previous estimates from single flu seasons. That is, the tree age is young and the root coincides with the start of the (winter) influenza season in the Southern Hemisphere. The time of introduction of influenza into the region, $z_0$, was 1 or 2 months before the root. This supports the notion that the sequences selected represent a single introduction of the strain into the Canterbury population (see File S1 for details of data selection and Figure S4 for representative trees inferred from an alternate data selection.).

The trees estimated by each of the three models are typical for influenza (see Figure 4 for representative trees from each posterior), with branches that are quick to coalesce moving backward in time from the most recently sampled tip.

**HIV-1 data analysis**

Results for inference from HIV-1 sequence data can be found in File S1. 95% HPD intervals are shown in Figure S5, Figure S6, Figure S7, and Table S8.

**Computational efficiency**

Finally, Table S5 shows comparisons of computation times under each inference model for each type of data.
analyzed. The deterministic coalescent SIR model is by far the fastest to sample and converge, with stochastic coalescent SIR and BDSIR varying, depending on the type of data.

Closing remarks

A key reason for the success of coalescent theory in population genetics is its mathematical simplicity and the computational efficiency of calculating the probability density of a sample genealogy. Our results show that a stochastic variant of coalescent theory can be successfully adapted to estimate epidemiological parameters in a true Bayesian inference context. This stochastic coalescent SIR model performs better than the deterministic analog for estimating epidemic parameters in some circumstances. Unfortunately, the stochastic model relies on a computationally demanding Monte Carlo estimate of the coalescent density via simulation of an ensemble of epidemic trajectories, neglecting one of the main advantages of coalescent theory. In fact, the current implementation is less computationally efficient than the implementation of the BDSIR model. However, an advantage of the stochastic coalescent over the explicit sampling model in BDSIR is its robustness to biased sampling schemes, as has been shown for the case of pure exponential growth dynamics (Bošková et al. 2014).

A more computationally efficient approach to computing the coalescent probability of the sample genealogy in the stochastic setting would be to use particle filtering (Andrieu and Roberts 2009; Andrieu et al. 2010; Rasmussen et al. 2011, 2014), but there are no theoretical barriers to applying particle MCMC to the exact model (Stadler et al. 2014). Therefore, an obvious extension of this work would be to apply particle MCMC algorithms to the exact stochastic SIR model that was used in simulations in this work. We anticipate that the exact model would outperform all the methods tested here, especially when \( R_0 \) is close to one.

In the meantime, the Bayesian coalescent inference methods developed here make it feasible to estimate epidemic parameters from time-stamped, serially sampled molecular sequence data, while accurately accounting for uncertainty in the topology and the divergence times of the phylogenetic tree.

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Literature Cited

Anderson, R. M., and R. M. May, 1991 Infectious Diseases of Humans: Dynamics and Control. Oxford University Press, Oxford.

Andrieu, C., and G. O. Roberts, 2009 The pseudo-marginal approach for efficient Monte Carlo computations. Ann. Stat. 37: 697.

Andrieu, C., A. Doucet, and R. Holenstein, 2010 Particle Markov chain Monte Carlo methods. J. R. Stat. Soc. Ser. B Stat. Methodol. 72: 269–342.

Beaumont, M. A., 2003 Estimation of population growth or decline in genetically monitored populations. Genetics 164: 1139–1160.

Bedford, T., S. Cobey, P. Beerli, and M. Pascual, 2010 Global migration dynamics underlie evolution and persistence of human influenza a (h3n2). PLoS Pathog. 6: e1000918.

Biggerstaff, M., S. Cauchemez, C. Reed, M. Gambhir, and L. Finelli, 2014 Estimates of the reproduction number for seasonal, pandemic, and zoonotic influenza: a systematic review of the literature. BMC Infect. Dis. 14: 480.

Bošková, V., S. Bonhoeffer, and T. Stadler, 2014 Inference of epidemiological dynamics based on simulated phylogenies using birth-death and coalescent models. PLoS Comput. Biol. 10: e1003913.

CDC, 2014 United States Centers for Disease Control and Prevention. Available at: http://www.cdc.gov/flu/. Accessed: November, 2014.

Chowell, G., M. Miller, and C. Viboud, 2008 Seasonal influenza in the United States, France, And Australia: transmission and prospects for control. Epidemiol. Infect. 6: 852–864.

Dearlove, B., and D. J. Wilson, 2013 Coalescent inference for infectious disease: meta-analysis of hepatitis C. Philos. Trans. R. Soc. Lond. B Biol. Sci. 368: 20120314.

Drummond, A. J., G. K. Nicholls, A. G. Rodrigo, and W. Solomon, 2002 Estimating mutation parameters, population history and genealogy simultaneously from temporally spaced sequence data. Genetics 161: 1307–1320.

Drummond, A. J., S. Y. W. Ho, M. J. Phillips, and A. Rambaut, 2006 Relaxed phylogenetics and dating with confidence. PLoS Biol. 4: e88.

ECAN, 2001 Environment Canterbury Regional Council. Available at: http://ecan.govt.nz/about-us/population/how-many/pages/census.aspx. Accessed: November, 2014.

Felsenstein, J., 1981 Evolutionary trees from DNA sequences: a maximum likelihood approach. J. Mol. Evol. 17: 368–376.

Felsenstein, J., 2004 Inferring Phylogenies. Sinauer Associates, Sunderland, MA.

Ferguson, N., D. Cummings, S. Cauchemez, C. Fraser, S. Riley et al., 2005 Strategies for containing an emerging influenza pandemic in southeast Asia. Nature 437: 209–214.

Gavryushkina, A., D. Welch, T. Stadler, and A. Drummond, 2014 Bayesian inference of sampled ancestor trees for epidemiology and fossil calibration. arXiv:1406.4573.

Grenfell, B. T., O. G. Pybus, J. R. Gog, J. L. N. Wood, J. M. Daly et al., 2004 Unifying the epidemiological and evolutionary dynamics of pathogens. Science 303: 327–332.

Griffiths, R. C., and S. Tavaré, 1994 Ancestral inference in population genetics. Stat. Sci. 9: 307–319.

Hasegawa, M., H. Kishino, and T. Yano, 1985 Dating of the human-ape splitting by a molecular clock of mitochondrial DNA. J. Mol. Evol. 22: 160–174.

Iwasaki, A., and P. S. Pillai, 2014 Innate immunity to influenza virus infection. Nat. Rev. Immunol. 14: 315–328.

Keeling, M. J., and P. Rohani, 2008 Modeling Infectious Diseases in Humans and Animals. Princeton University Press, Princeton.

Kermack, W., and A. McKendrick, 1932 Contributions to the mathematical theory of epidemics. ii. The problem of endemicity. Proc. R. Soc. A 138: 55–83.
Opatowski, L., C. Fraser, J. Grif
Kutta, M. W., 1901 Beitrag zur näherungsweisen integration totaler
differentialgleichungen. Zeitschrift für Mathematik und Physik

Kühnert, D., T. Stadler, T. G. Vaughan, and A. J. Drummond,
2012 Rates of coalescence for common epidemiological models at equilibriu

Koelle, K., S. Cobey, B. Grenfell, and M. Pascual, 2006 Epochal
evolution shapes the phylogenetics of interpandemic influen
za (h3n2) in humans. Science 314: 1898–1903.

Kühnert, D., T. Stadler, T. G. Vaughan, and A. J. Drummond,
2014 Simultaneous reconstruction of evolutionary history and
epidemiological dynamics from viral sequences with the
birth-death SIR model. J. R. Soc. Interface 11: 20131106.

Kutta, M. W., 1901 Beitrag zur näherungsweisen integration totaler
differentialgleichungen. Math. Ann. 46: 167–178.

Koelle, K., and D. A. Rasmussen, 2012 Rates of coalescence for
common epidemiological models at equilibrium. J. R. Soc. Inter-
fase 9: 997–1007.

Koelle, K., S. Cobey, B. Grenfell, and M. Pascual, 2006 Epochal
evolution shapes the phylogenetics of interpandemic influen
za (h3n2) in humans. Science 314: 1898–1903.

Kühnert, D., T. Stadler, T. G. Vaughan, and A. J. Drummond,
2014 Simultaneous reconstruction of evolutionary history and
epidemiological dynamics from viral sequences with the
birth-death SIR model. J. R. Soc. Interface 11: 20131106.

Kutta, M. W., 1901 Beitrag zur näherungsweisen integration totaler
differentialgleichungen. Math. Ann. 46: 167–178.

Koelle, K., and D. A. Rasmussen, 2012 Rates of coalescence for
common epidemiological models at equilibrium. J. R. Soc. Inter-
fase 9: 997–1007.

Koelle, K., S. Cobey, B. Grenfell, and M. Pascual, 2006 Epochal
evolution shapes the phylogenetics of interpandemic influen
za (h3n2) in humans. Science 314: 1898–1903.

Kühnert, D., T. Stadler, T. G. Vaughan, and A. J. Drummond,
2014 Simultaneous reconstruction of evolutionary history and
epidemiological dynamics from viral sequences with the
birth-death SIR model. J. R. Soc. Interface 11: 20131106.

Kutta, M. W., 1901 Beitrag zur näherungsweisen integration totaler
differentialgleichungen. Math. Ann. 46: 167–178.

Koelle, K., and D. A. Rasmussen, 2012 Rates of coalescence for
common epidemiological models at equilibrium. J. R. Soc. Inter-
fase 9: 997–1007.

Koelle, K., S. Cobey, B. Grenfell, and M. Pascual, 2006 Epochal
evolution shapes the phylogenetics of interpandemic influen
za (h3n2) in humans. Science 314: 1898–1903.

Kühnert, D., T. Stadler, T. G. Vaughan, and A. J. Drummond,
2014 Simultaneous reconstruction of evolutionary history and
epidemiological dynamics from viral sequences with the
birth-death SIR model. J. R. Soc. Interface 11: 20131106.

Kutta, M. W., 1901 Beitrag zur näherungsweisen integration totaler
differentialgleichungen. Math. Ann. 46: 167–178.

Koelle, K., and D. A. Rasmussen, 2012 Rates of coalescence for
common epidemiological models at equilibrium. J. R. Soc. Inter-
fase 9: 997–1007.

Koelle, K., S. Cobey, B. Grenfell, and M. Pascual, 2006 Epochal
evolution shapes the phylogenetics of interpandemic influen
za (h3n2) in humans. Science 314: 1898–1903.

Kühnert, D., T. Stadler, T. G. Vaughan, and A. J. Drummond,
2014 Simultaneous reconstruction of evolutionary history and
epidemiological dynamics from viral sequences with the
birth-death SIR model. J. R. Soc. Interface 11: 20131106.

Kutta, M. W., 1901 Beitrag zur näherungsweisen integration totaler
differentialgleichungen. Math. Ann. 46: 167–178.

Koelle, K., and D. A. Rasmussen, 2012 Rates of coalescence for
common epidemiological models at equilibrium. J. R. Soc. Inter-
fase 9: 997–1007.

Koelle, K., S. Cobey, B. Grenfell, and M. Pascual, 2006 Epochal
evolution shapes the phylogenetics of interpandemic influen
za (h3n2) in humans. Science 314: 1898–1903.

Kühnert, D., T. Stadler, T. G. Vaughan, and A. J. Drummond,
2014 Simultaneous reconstruction of evolutionary history and
epidemiological dynamics from viral sequences with the
birth-death SIR model. J. R. Soc. Interface 11: 20131106.

Kutta, M. W., 1901 Beitrag zur näherungsweisen integration totaler
differentialgleichungen. Math. Ann. 46: 167–178.

Koelle, K., and D. A. Rasmussen, 2012 Rates of coalescence for
common epidemiological models at equilibrium. J. R. Soc. Inter-
fase 9: 997–1007.

Koelle, K., S. Cobey, B. Grenfell, and M. Pascual, 2006 Epochal
evolution shapes the phylogenetics of interpandemic influen
za (h3n2) in humans. Science 314: 1898–1903.

Kühnert, D., T. Stadler, T. G. Vaughan, and A. J. Drummond,
2014 Simultaneous reconstruction of evolutionary history and
epidemiological dynamics from viral sequences with the
birth-death SIR model. J. R. Soc. Interface 11: 20131106.

Kutta, M. W., 1901 Beitrag zur näherungsweisen integration totaler
differentialgleichungen. Math. Ann. 46: 167–178.

Koelle, K., and D. A. Rasmussen, 2012 Rates of coalescence for
common epidemiological models at equilibrium. J. R. Soc. Inter-
fase 9: 997–1007.
Inferring Epidemiological Dynamics with Bayesian Coalescent Inference: The Merits of Deterministic and Stochastic Models

Alex Popinga, Tim Vaughan, Tanja Stadler, and Alexei J. Drummond
Supporting material for “Inferring epidemiological dynamics with Bayesian coalescent inference: The merits of deterministic and stochastic models”

File S1

1 Sampling from the prior

In order to assess the correctness of our implementation of the deterministic coalescent SIR and stochastic coalescent SIR models, for each model we used the MCMC algorithm to sample trees from the corresponding distribution $f(T|\eta)$, and compared these samples with coalescent trees simulated directly under the model.

The chosen $\eta$ included $\beta = 7.5 \times 10^{-4}$, $\gamma = 0.3$, $S_0 = 999$ and $z_0 = 30$. The comparisons were performed for trees generated from 20 leaves, sampled at integer times 0 through 19, inclusive.

For the deterministic coalescent SIR model, the direct simulation involved numerically solving the Eqs. (1)–(3) in the main text for $t \in [0,30]$ and using this solution in combination with Eq. (10) in the main text to determine the instantaneous coalescent rate $\lambda(\tau)$. This rate was used to simulate each of the coalescent trees in the usual fashion for heterochronous leaf times. In the case that the MRCA was not reached before the origin time of the epidemic, the tree was discarded and the simulation repeated.

The direct simulation proceeded in a similar way for the stochastic coalescent SIR model, the major difference being that the stochasticity of this model required each coalescent tree to be simulated under a distinct realization of the stochastic trajectory.

Comparisons between the direct simulation and MCMC results are shown in Figures S1 and S2 for three different summary statistics and show very close agreement.

2 Validation through simulated data analysis

As part of the validation of our implementation of the two coalescent SIR models, trees were simulated by their own methods (using stochastically- and deterministically-generated SIR trajectories, as discussed in the Methods section of the main paper), and relevant epidemiological parameters were inferred
using the stochastic and deterministic coalescent SIR models. Tables 1 and 2 show the results of these analyses, indicative of correct implementations.

Analyses for varying $R_0$ (and necessarily, slightly varied other parameters, such as the birth rate $\beta$) are provided in Tables S3 and S4. Results from tests of the influence of broader priors (with larger standard deviations in log space) are shown in Table S4. It appears that allowance of broader priors reduces 95% HPD coverage in some cases (e.g., for parameter $R_0$) when using the deterministic coalescent SIR inference model, as they increase error and bias.

Finally, it was noticed that even for the higher true parameter values of $R_0 = 2.50$ and $S_0 = 999$, under which deterministic coalescent SIR is expected to perform relatively well, there was an inability to accurately estimate the origin parameter $z_0$. Figure S3 provides some insight into this conundrum by examining the trajectories used for tree simulation and subsequent analysis.

2.1 H1N1 data selection

Initially, the H1N1 dataset contained 45 sequences. The ages of the inferred trees (Figure S4) using the original 45 sequences extended more than 1.5 years into the past for each of the SIR models, which is contrary to what we expect for a single, current strain of seasonal influenza. Three taxa (labelled 32197, 31893, and 31988) were hypothesized to belong to a unique strain, e.g., an additional seeding from outside the Canterbury region or a low-lying previous strain. Removing these three taxa caused the inferred trees to behave as expected, i.e., tree heights and epidemic origin $z_0$ less than a year old. It also raised the estimated $R_0$ values for all three SIR models (initially 1.24, 1.10, and 1.55 for stochastic coalescent SIR, deterministic coalescent SIR, and BDSIR, respectively), as well as those for $\gamma$ (initially 8.74, 12.65, and 11.33 for stochastic coalescent SIR, deterministic coalescent SIR, and BDSIR, respectively).

It will be interesting to further investigate the interplay between influenza strains and its contribution to the overall dynamics. For the closed SIR models discussed in this manuscript, however, this additional complexity leads to increased chance of model misspecification and misleading results. Therefore, we focused our attention on the analyses using 42 sequences.

2.2 HIV-1 data analysis

The original HIV-1 dataset (Hué et al. 2005) was agglomerated from both acute and chronic infections sampled in the United Kingdom (UK) and constitutes six phylogenetic clusters, from which the five used here (Clusters 1-4 and 6) were drawn. These particular clusters, with the omission of Cluster 5, were chosen simply for the purpose of direct comparison with Kühnert et al. (2014). Our extension to the models allowed us to imprint respective tip dates on the sequence data, sampled from 1999 to 2003, for inclusion in the likelihood computation.

For the selected five clusters, the nucleotide alignments contained 41, 62, 29, 26, and 35 sequences, respectively, each with 952 sites. The substitution
scheme chosen for phylogenetic analysis was the symmetric and independent general time reversible model (GTR), with gamma distributed rate variation and explicit proportion of invariable sites (GTR+G+I). Following Hué et al. (2005), the substitution rate was set to \( 2.55 \times 10^{-4} \) substitutions per site per year. All other parameters were estimated conjointly, and the Bayesian prior distributions are presented in Table 4: Bayesian prior distributions.

The pathophysiology of HIV is multifarious, and the patterns of its advancement within an infected host change throughout time. In addition to increased complexity potentially caused by recombination events, the transition between HIV’s acute and chronic phases alters the host’s infectivity (Guss 1994). The SIR compartmental model used for this particular phylodynamic analysis on the UK cluster data does not allow for independent infection rates for the acute and chronic phases (but see Volz et al. (2012) and Volz et al. (2013)). However, in this study we did not attempt to estimate the infection rate \( \beta \) and thus did not expect such a difference to significantly impact the estimation of the parameters of interest: the basic reproductive number \( R_0 \), removal rate \( \gamma \), size of the initial susceptible population \( S_0 \), and origin of the outbreak \( z_0 \).

### 2.2.1 HIV-1 inference results

In regard to parameter inference from the serially-sampled HIV-1 sequence data, the stochastic coalescent SIR, deterministic coalescent SIR, and BDSIR methods were most alike in light of the \( R_0 \) results. The medians and HPD intervals for all clusters pertaining to this parameter, (especially Clusters 1, 2, 3, and 6), were very close, and those of Cluster 4 were still congruent across the three analyses (Figure S5).

The coalescent SIR models and BDSIR disagreed with respect to the age of the most recent common ancestor and the origin \( z_0 \) (Figure S6). The coalescent SIR models also exhibited much larger 95% HPD intervals for \( z_0 \) in each of the clusters; while BDSIR encompassed an average of 16 years, the stochastic coalescent SIR and deterministic coalescent SIR models had averages of 49 and 37 years, respectively. Furthermore, the estimated age of the common ancestor of the tree was older under the coalescent SIR models than the estimates reported by either BDSIR or the original data analysis (Hué et al. 2005) for each cluster. This was also true for the time of origin for the epidemic, although for certain clusters the differences between the coalescent estimates of the origin \( z_0 \) and the birth-death estimates were much greater than others (e.g., Cluster 3).

The estimates of removal rate \( \gamma \) from Clusters 1 and 6 were very similar across the three methods (Figure S7). However, both coalescent SIR models estimated considerably higher \( \gamma \) values for Clusters 2-4 than BDSIR. This is reflective of the simulation study results, where the two coalescent models did not perform as well as BDSIR for the removal parameter.

Median estimates for the initial susceptible population \( S_0 \) were quite similar in all methods for Clusters 1-4, although BDSIR displayed much wider HPD intervals than stochastic coalescent SIR and deterministic coalescent SIR (Figure S8). In Cluster 6, the coalescent SIR models showed the smallest HPD intervals.
for their individual analyses on each cluster, while the opposite was true for BDSIR. There was also a disparity between the median estimates for the two coalescent approaches and that of BDSIR for Cluster 6. To this effect, it should be noted that the number of infections accrued throughout the duration of the epidemic was reported as \( N_e = 1,350 \) by Hué et al. This casts some suspicion on the low susceptible population estimates obtained by the stochastic coalescent SIR and deterministic coalescent SIR methods (median estimates of \( S_0 = 727 \) and \( S_0 = 693 \), respectively), since they appear lower than the estimated number of infected individuals from the original study.

There is disagreement in the literature in regard to the modelling of HIV-1 evolutionary dynamics under stochastic or deterministic processes (Nijhuis et al. 1998; Rouzine and Coffin 1999; Achaz et al. 2004; Shriner et al. 2004). The predicament dwells in the observation that the actual effective population size \( N_e \) for HIV-1 is often smaller than the total population size (Kouyos et al. 2006). While most of this debate has focused on within-host population dynamics, many of the arguments hold when considering the broader epidemic dynamics of host-to-host transmission. As previously mentioned, the appropriateness of these descriptions is hinged on the magnitude of the infected population, precisely, the effective infected population size. Consequently, even when the total infected population is quite large there may yet be significant stochastic effects in play.

Finally, as mentioned in the main article, the existence of two distinct infectious stages and the possibility of large effects due to recombination are reasons for any discrepancy produced by these SIR inference models.

### 2.2.2 Example XML

Below is an example XML for simulating 100 trees and trajectories in MASTER (Vaughan and Drummond 2013). This example is for \( R_0 = 2.4975 \) and \( S_0 = 999 \). The simulation ends when the infected \( I \) population returns to zero, i.e., when the last infected individual is removed.

```xml
<beast version='2.0' namespace='master.beast:beast.core.parameter:beast.evolution.tree.TreeHeightLogger'>
  <run spec='InheritanceEnsemble' nTraj='100'
       samplePopulationSizes='true'
       verbosity='1'>
    <model spec='InheritanceModel' id='model'>
      <population spec='Population' id='S' populationName='S'/>
      <population spec='Population' id='I' populationName='I'/>
      <population spec='Population' id='R' populationName='R'/>
      <population spec='Population' id='Rh' populationName='Rh'/>

      <!-- infection reaction -->
      <reaction spec='InheritanceReaction' reactionName='Infection' rate='0.00075'>
        S + I -> 2I
      </reaction>

    </model>
  </run>
</beast>
```
References

ACHAZ, G., S. PALMER, M. KEARNEY, F. MALDARELLI, J. W. MELLORS, et al., 2004 A robust measure of HIV-1 population turnover within chronically infected individuals. Mol Biol Evol 21: 1902-12.

GUSS, D. A., 1994 The acquired immune deficiency syndrome: An overview for the emergency physician, part 1. The Journal of Emergency Medicine 12: 375–384.

HUÉ, S., D. PILLAY, J. P. CLEWLEY, and O. G. PYBUS, 2005 Genetic analysis reveals the complex structure of hiv-1 transmission within defined risk groups. PNAS 102: 4425–4429.

KOUYOS, R. D., C. L. ALTHAUS, and S. BONHOEFFER, 2006 Stochastic or deterministic: what is the effective population size of HIV-1? Trends Microbiol 14: 507–11.

KÜHNERT, D., T. STADLER, T. G. VAUGHAN, and A. J. DRUMMOND, 2014 Simultaneous reconstruction of evolutionary history and epidemiological dynamics from viral sequences with the birth-death sir model. J R Soc Interface 11: 20131106.

NIJHUIS, M., C. A. BOUCHER, P. SCHIPPER, T. LEITNER, R. SCHUURMAN, et al., 1998 Stochastic processes strongly influence hiv-1 evolution during suboptimal protease-inhibitor therapy. Proc Natl Acad Sci U S A 95: 14441–6.

ROUZINE, I., and J. COFFIN, 1999 Linkage disequilibrium test implies a large effective population number for hiv in vivo. PNAS 96: 10758–10763.

SHRINER, D., R. SHANKARAPPA, M. A. JENSEN, D. C. NICKLE, J. E. MITTLER, et al., 2004 Influence of random genetic drift on human immunodeficiency virus type 1 env evolution during chronic infection. Genetics 166: 1155–64.

VAUGHAN, T. G., and A. J. DRUMMOND, 2013 A stochastic simulator of birth-death master equations with application to phylodynamics. Molecular Biology and Evolution .

VOLZ, E. M., 2012 Complex population dynamics and the coalescent under neutrality. Genetics 190: 187–201.

VOLZ, E. M., E. IONIDES, E. O. ROMERO-SEVERSON, M.-G. BRANDT, E. MOKOTOFF, et al., 2013 Hiv-1 transmission during early infection in men who have sex with men: a phylodynamic analysis. PLoS Med 10: e1001568; discussion e1001568.

VOLZ, E. M., J. S. KOOPMAN, M. J. WARD, A. L. BROWN, and S. D. W. FROST, 2012 Simple epidemiological dynamics explain phylogenetic clustering of hiv from patients with recent infection. PLoS Comput Biol 8: e1002552.
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Table S1: Simulation Study Results for Stochastic Coalescent Trees

| $\eta$ | Inference   | Truth | Mean | Median | Error | Bias | Relative HPD width | 95% HPD accuracy |
|--------|-------------|-------|------|--------|-------|------|---------------------|-----------------|
| $R_0$  | Stoch.Coal.SIR | 2.50  | 2.81 | 2.64   | 0.11  | 0.08 | 0.95                | 100.00%         |
|        | Deter.Coal.SIR | 2.50  | 2.73 | 2.65   | 0.14  | 0.06 | 0.85                | 96.00%          |
| $\gamma$ | Stoch.Coal.SIR | 0.30  | 0.28 | 0.26   | 0.16  | -0.11 | 1.17                | 99.00%          |
|        | Deter.Coal.SIR | 0.30  | 0.30 | 0.28   | 0.18  | -0.03 | 1.20                | 99.00%          |
| $S_{(0)}$ | Stoch.Coal.SIR | 999   | 1456 | 986    | 0.21  | 0.02 | 3.93                | 100.00%         |
|        | Deter.Coal.SIR | 999   | 1720 | 1057   | 0.48  | 0.24 | 4.28                | 99.00%          |
| $z_{(0)}$ | Stoch.Coal.SIR | (varies) | 42.36 | 40.43 | 0.03 | 0.02 | 0.20                | 98.00%          |
|        | Deter.Coal.SIR | (varies) | 41.25 | 39.77 | 0.03 | 0.01 | 0.07                | 64.00%          |
Table S2: Simulation Study Results for Deterministic Coalescent Trees

| $\eta$ | Inference    | Truth | Mean | Median | Error | Bias | Relative HPD width | 95% HPD accuracy |
|--------|--------------|-------|------|--------|-------|------|--------------------|------------------|
| $R_0$  | Stoch.Coal.SIR | 2.50  | 2.44 | 2.37   | 0.06  | -0.05| 0.67               | 100.00%          |
|        | Deter.Coal.SIR| 2.50  | 2.51 | 2.46   | 0.08  | -0.01| 0.59               | 99.00%           |
| $\gamma$ | Stoch.Coal.SIR | 0.30  | 0.33 | 0.31   | 0.07  | 0.05 | 1.00               | 100.00%          |
|        | Deter.Coal.SIR | 0.30  | 0.32 | 0.30   | 0.10  | 0.02 | 0.79               | 100.00%          |
| $S_{(0)}$ | Stoch.Coal.SIR | 999   | 1586 | 1142   | 0.26  | 0.20 | 3.83               | 100.00%          |
|        | Deter.Coal.SIR | 999   | 1426 | 1030   | 0.36  | 0.13 | 3.03               | 100.00%          |
| $z_{(0)}$ | Stoch.Coal.SIR | 44.12 | 45.52| 44.74  | 0.02  | 0.01 | 0.19               | 93.00%           |
|        | Deter.Coal.SIR | 44.12 | 44.34| 44.11  | 0.02  | 1.93e-3| 0.08             | 92.00%           |
Table S3: Results for Homochronous Sampling

| η       | Inference     | Truth | Mean | Median | Error | Bias | Relative HPD width | 95% HPD accuracy |
|---------|---------------|-------|------|--------|-------|------|--------------------|------------------|
| $R_0$   | Stoch.Coal.SIR| 2.50  | 3.04 | 2.74   | 0.13  | 0.11 | 1.32               | 100.00%          |
|         | Deter.Coal.SIR| 2.50  | 4.05 | 3.29   | 0.34  | 0.32 | 2.38               | 100.00%          |
|         | BDSIR         | 2.50  | 2.84 | 2.49   | 0.16  | 0.03 | 1.45               | 97.00%           |
| $\gamma$| Stoch.Coal.SIR| 0.30  | 0.26 | 0.23   | 0.25  | -0.21| 1.43               | 100.00%          |
|         | Deter.Coal.SIR| 0.30  | 0.26 | 0.19   | 0.36  | -0.31| 2.03               | 100.00%          |
|         | BDSIR         | 0.30  | 0.23 | 0.17   | 0.42  | -0.42| 2.04               | 100.00%          |
| $S_{(0)}$| Stoch.Coal.SIR| 999   | 1660 | 1065   | 0.18  | 0.09 | 4.75               | 100.00%          |
|         | Deter.Coal.SIR| 999   | 4127 | 679    | 0.78  | 0.09 | 10.24              | 100.00%          |
|         | BDSIR         | 999   | 1907 | 1320   | 0.41  | 0.41 | 4.86               | 100.00%          |
| $z_{(0)}$| Stoch.Coal.SIR| 20.0  | 20.17| 19.82  | 0.09  | -0.03| 0.43               | 95.00%           |
|         | Deter.Coal.SIR| 20.0  | 19.09| 19.21  | 0.09  | -0.05| 0.19               | 73.00%           |
|         | BDSIR         | 20.0  | 36.56| 29.38  | 0.55  | 0.54 | 4.24               | 100.00%          |
Table S4: Simulation Study Results: The Effect of Broader Priors on Deterministic Coalescent SIR

| \( \eta \) | St. Dev. | Truth | Mean | Median | Error | Bias | Relative HPD width | 95% HPD accuracy |
|---|---|---|---|---|---|---|---|---|
| \( R_0 \) | 2 | 1.50 | 2.06 | 1.75 | 0.40 | 0.35 | 0.86 | 79.00% |
| \( R_0 \) | 1 | 1.50 | 1.80 | 1.49 | 0.24 | 0.15 | 0.52 | 85.00% |
| \( R_0 \) | 2 | 2.50 | 3.31 | 2.85 | 0.34 | 0.24 | 1.43 | 95.00% |
| \( R_0 \) | 1 | 2.50 | 2.68 | 2.49 | 0.13 | 0.04 | 0.80 | 99.00% |
| \( \gamma \) | 2 | 0.30 | 0.31 | 0.23 | 0.37 | -0.12 | 1.59 | 96.00% |
| \( \gamma \) | 1 | 0.30 | 0.26 | 0.23 | 0.27 | -0.22 | 1.15 | 89.00% |
| \( \gamma \) | 2 | 0.30 | 0.31 | 0.25 | 0.33 | -0.09 | 1.59 | 95.00% |
| \( \gamma \) | 1 | 0.30 | 0.32 | 0.29 | 0.16 | 3.14E-3 | 1.27 | 99.00% |
| \( S_0(0) \) | 2 | 499 | 2041 | 249 | 1.40 | 0.49 | 7.75 | 85.00% |
| \( S_0(0) \) | 1 | 499 | 562 | 361 | 0.44 | -0.26 | 3.36 | 91.00% |
| \( S_0(0) \) | 2 | 999 | 3028 | 717 | 1.05 | 0.33 | 6.60 | 94.00% |
| \( S_0(0) \) | 1 | 499 | 553.38 | 337 | 0.42 | -0.26 | 3.08 | 92.00% |
| \( z(0) \) | 2 | (varies) | 65.10 | 62.01 | 0.04 | 0.03 | 0.25 | 86.00% |
| \( z(0) \) | 1 | (varies) | 91.03 | 72.51 | 0.39 | 0.38 | 0.42 | 88.00% |
| \( z(0) \) | 2 | (varies) | 40.97 | 39.85 | 0.03 | -6.78E-4 | 0.08 | 81.00% |
| \( z(0) \) | 1 | (varies) | 112.79 | 90.37 | 0.26 | 0.26 | 0.94 | 85.00% |
Table S5: Comparison of Computation Times for Bayesian Inference of Epidemic Parameters from Genetic Sequence Data using SIR Models

| Data Type               | Inference Model | Mean time per million samples (MCMC) |
|-------------------------|-----------------|--------------------------------------|
| Sim. Study ($R_0 \approx 2.50$) | Stoch.Coal.SIR  | 20m 41s                              |
|                         | Deter.Coal.SIR  | 3m 27s                               |
|                         | BDSIR           | 56m 27s                              |
| Sim. Study ($R_0 \approx 1.50$) | Stoch.Coal.SIR  | 1h 43m 30s                           |
|                         | Deter.Coal.SIR  | 3m 47s                               |
|                         | BDSIR           | 41m 35s                              |
| Sim. Study ($R_0 \approx 1.10$) | Stoch.Coal.SIR  | 1h 50m 41s                           |
|                         | Deter.Coal.SIR  | 6m 45s                               |
|                         | BDSIR           | 41m 21s                              |
| H1N1                    | Stoch.Coal.SIR  | 1h 20m 55s                           |
|                         | Deter.Coal.SIR  | 9m 44s                               |
|                         | BDSIR           | 47m 33s                              |
| HIV-1                   | Stoch.Coal.SIR  | 14h 37m 45s                          |
|                         | Deter.Coal.SIR  | 7m 56s                               |
|                         | BDSIR           | 1h 38m 54s                           |
Table S6: Deterministic Coalescent SIR Results for Simulated Sequences: $R_0 = 1.0987$ and $S_0 = 499$, $R_0 = 1.0989$ and $S_0 = 999$, $R_0 = 1.09945$ and $S_0 = 1999$

| $\eta$ | Truth | Mean | Median | Error | Bias | Relative HPD width | 95% HPD accuracy |
|--------|-------|------|--------|-------|------|-------------------|-----------------|
| $R_0$  | ≈1.10 | 1.89 | 1.28   | 0.62  | 0.63 | 0.40              | 52.00%          |
| $\gamma$ | 0.30 | 0.57 | 0.44   | 0.59  | 0.52 | 2.14              | 95.00%          |
| $S_{(0)}$ | 499  | 1830 | 1222   | 1.50  | 1.31 | 11.49             | 96.00%          |
| $z_{(0)}$ (varies) | 109.55 | 76.21 | 0.61   | 0.54  | 0.35 | 37.00%            |
| $R_0$  | ≈1.10 | 1.55 | 1.35   | 0.25  | 0.25 | 0.45              | 16.00%          |
| $\gamma$ | 0.30 | 0.27 | 0.24   | 0.20  | -0.12 | 1.23              | 61.00%          |
| $S_{(0)}$ | 999  | 1293 | 804    | 0.27  | -0.10 | 3.58              | 64.00%          |
| $z_{(0)}$ (varies) | 117.39 | 99.75 | 0.25   | 0.18  | 0.23 | 23.00%            |
| $R_0$  | ≈1.10 | 1.37 | 1.22   | 0.16  | 0.16 | 0.28              | 40.00%          |
| $\gamma$ | 0.30 | 0.26 | 0.24   | 0.18  | -0.14 | 1.13              | 64.00%          |
| $S_{(0)}$ | 1999 | 2292 | 1531   | 0.23  | -0.18 | 3.45              | 66.00%          |
| $z_{(0)}$ (varies) | 150.39 | 138.69 | 0.23   | 0.20  | 0.32 | 18.00%            |

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| Type of simulated data | Inference models used |
|------------------------|-----------------------|
| **1. Varying $R_0$ and $S_0$ (orig.)** | |
| (a) $R_0 \approx 1.1$, $S_0 = 499$, $\gamma = 0.25$, $\psi = 0.15$ | Deter.Coal.SIR, Stoch.Coal.SIR, BDSIR |
| (b) $R_0 \approx 1.2$, $S_0 = 499$, $\gamma = 0.30$, $\psi = 0.15$ | Deter.Coal.SIR |
| (c) $R_0 \approx 1.5$, $S_0 = 499$, $\gamma = 0.30$, $\psi = 0.15$ | Deter.Coal.SIR, Stoch.Coal.SIR, BDSIR |
| (d) $R_0 \approx 1.5$, $S_0 = 999$, $\gamma = 0.30$, $\psi = 0.20$ | Deter.Coal.SIR, Stoch.Coal.SIR, BDSIR |
| (e) $R_0 \approx 2.5$, $S_0 = 999$, $\gamma = 0.30$, $\psi = 0.05$ | Deter.Coal.SIR, Stoch.Coal.SIR, BDSIR |
| **2. Varying $S_0$ for fixed $R_0$** | |
| (a) $R_0 \approx 1.1$, $S_0 = 499$, $\gamma = 0.25$, $\psi = 0.15$ | Deter.Coal.SIR, Stoch.Coal.SIR, BDSIR |
| (f) $R_0 \approx 1.1$, $S_0 = 999$, $\gamma = 0.30$, $\psi = 0.20$ | Deter.Coal.SIR |
| (g) $R_0 \approx 1.1$, $S_0 = 1999$, $\gamma = 0.30$, $\psi = 0.09$ | Deter.Coal.SIR |
| **3. Contemporaneous sampling** | |
| (d) $R_0 \approx 1.5$, $S_0 = 999$, $\gamma = 0.30$, $\psi = 0.20$ | Deter.Coal.SIR, Stoch.Coal.SIR, BDSIR |
| (e) $R_0 \approx 2.5$, $S_0 = 999$, $\gamma = 0.30$, $\psi = 0.05$ | Deter.Coal.SIR, Stoch.Coal.SIR, BDSIR |
| **4. Phylogenetic uncertainty** | |
| (e) $R_0 \approx 2.5$, $S_0 = 999$, $\gamma = 0.30$, $\psi = 0.05$ | Deter.Coal.SIR, Stoch.Coal.SIR, BDSIR |
| **5. Reparameterization (growth rate)** | |
| (e) $R_0 \approx 2.5$, $S_0 = 999$, $\gamma = 0.30$, $\psi = 0.05$ | Deter.Coal.SIR |
Table S8: Epidemic Parameter Estimations from HIV-1 Subtype B Sequence Data

| Inference Model | HIV cluster | $R_0$     | $\gamma$     | $S_0$       | Root of the tree (yr) | Origin $z_0$ of the epidemic (yr) |
|-----------------|-------------|-----------|--------------|-------------|-----------------------|----------------------------------|
| **Stoch. Coal. SIR** |             |           |              |             |                       |                                  |
| Cluster 1       | 3.31        | 0.27      | 1165         | 1971        | (2.40 - 4.26)          | (1946 - 1987)                    |
| Cluster 2       | 2.42        | 0.32      | 976          | 1975        | (1.86 - 3.07)          | (1953 - 1988)                    |
| Cluster 3       | 2.10        | 0.41      | 1442         | 1979        | (1.07 - 3.73)          | (1959 - 1990)                    |
| Cluster 4       | 3.48        | 0.28      | 1757         | 1964        | (1.69 - 5.24)          | (1922 - 1990)                    |
| Cluster 6       | 3.09        | 0.19      | 727          | 1972        | (1.94 - 4.53)          | (1947 - 1988)                    |
| **Deter. Coal. SIR** |             |           |              |             |                       |                                  |
| Cluster 1       | 3.43        | 0.26      | 1158         | 1969        | (2.52 - 4.40)          | (1941 - 1987)                    |
| Cluster 2       | 2.34        | 0.38      | 1158         | 1979        | (1.92 - 2.81)          | (1967 - 1989)                    |
| Cluster 3       | 1.87        | 0.54      | 1298         | 1972        | (1.42 - 2.43)          | (1965 - 1989)                    |
| Cluster 4       | 3.35        | 0.34      | 1479         | 1971        | (2.02 - 4.86)          | (1948 - 1990)                    |
| Cluster 6       | 3.14        | 0.18      | 693          | 1969        | (1.98 - 4.64)          | (1943 - 1988)                    |
| **BDSIR**       |             |           |              |             |                       |                                  |
| Cluster 1       | 3.22        | 0.30      | 880          | 1986        | (2.18 - 4.27)          | (1983 - 1988)                    |
| Cluster 2       | 2.45        | 0.17      | 1745         | 1983        | (1.53 - 3.68)          | (1979 - 1986)                    |
| Cluster 3       | 1.90        | 0.20      | 1540         | 1985        | (1.22 - 2.78)          | (1981 - 1988)                    |
| Cluster 4       | 2.62        | 0.15      | 1921         | 1987        | (1.45 - 4.29)          | (1970 - 1988)                    |
| Cluster 6       | 3.17        | 0.15      | 2862         | 1986        | (1.73 - 5.43)          | (1975 - 1989)                    |
Table S9: Deterministic Coalescent SIR Results from Trees Simulated with Higher $S_0$ (with Fixed $R_0$) and Higher $R_0$ (with Fixed $S_0$)

| $\eta$ | Truth | Mean   | Median  | Error | Bias  | Relative HPD width | 95% HPD accuracy |
|-------|-------|--------|---------|-------|-------|-------------------|------------------|
| $R_0$ | 2.50  | 2.68   | 2.49    | 0.13  | 0.04  | 0.81              | 98.00%           |
| $\gamma$ | 0.30  | 0.32   | 0.29    | 0.16  | 3.14e-3 | 1.27              | 99.00%           |
| $S(0)$ | 999   | 1807   | 1133    | 0.52  | 0.29  | 4.59              | 98.00%           |
| $\xi(0)$ (varies) | 41.17 | 39.99  | 0.03    | 0.01  | 0.07  | 76.00%           |
| $R_0$ | 2.50  | 3.28   | 2.97    | 0.23  | 0.20  | 1.42              | 100.00%          |
| $\gamma$ | 0.35  | 0.30   | 0.28    | 0.24  | -0.20 | 1.28              | 99.00%           |
| $S(0)$ | 4999  | 7733   | 4838    | 0.34  | 0.03  | 4.18              | 100.00%          |
| $\xi(0)$ (varies) | 37.45 | 36.15  | 0.03    | 0.01  | 0.07  | 56.00%           |
| $R_0$ | 3.50  | 3.92   | 3.76    | 0.26  | 0.23  | 1.50              | 100.00%          |
| $\gamma$ | 0.40  | 0.33   | 0.31    | 0.26  | -0.22 | 1.22              | 100.00%          |
| $S(0)$ | 9999  | 12,609 | 7405    | 0.35  | -0.15 | 3.31              | 100.00%          |
| $\xi(0)$ (varies) | 34.99 | 34.28  | 0.04    | 0.01  | 0.07  | 43.00%           |
| $R_0$ | 5.00  | 6.13   | 5.53    | 0.20  | 0.12  | 1.39              | 100.00%          |
| $\gamma$ | 0.30  | 0.28   | 0.27    | 0.29  | -0.09 | 1.16              | 99.00%           |
| $S(0)$ | 9999  | 2144   | 1220    | 0.68  | 0.49  | 4.94              | 99.00%           |
| $\xi(0)$ (varies) | 26.28 | 25.26  | 0.03    | 0.01  | 0.03  | 52.00%           |
| $R_0$ | 5.00  | 7.20   | 6.37    | 0.27  | 0.23  | 2.53              | 100.00%          |
| $\gamma$ | 0.30  | 0.26   | 0.22    | 0.26  | -0.19 | 1.41              | 100.00%          |
| $S(0)$ | 9999  | 17,339 | 10,518  | 0.31  | 0.12  | 4.91              | 100.00%          |
| $\xi(0)$ (varies) | 28.20 | 26.93  | 0.03    | 0.01  | 1.05  | 36.00%           |