Arbuscular Mycorrhizal Fungi (AMF) Induced Defense Factors against the Damping-off Disease Pathogen, *Pythium aphanidermatum* in Chilli (*Capsicum annum*)

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**A B S T R A C T**

Arbuscular Mycorrhizal Fungi (AMF) from the small phylum *Glomeromycota* forms symbiotic association with the roots of higher plants. The AMF appears to benefit by improving the uptake of phosphate and other nutrients from soil and also increases the disease tolerance to the host plant. Mycorrhizal plants produce certain substances that have been indicated as copartners in resistance to the attack of root pathogens. The GC-MS analysis of root and leaf samples of AMF inoculated plants of chilli infected with the damping off disease pathogen, *Pythium aphanidermatum*, a tripartite interaction (chilli-AMF-pathogen) indicated the presence of compounds such as carboxy-1-methyl-2-azetidione, 1,2-cyclobutanedicarboxylic acid, tetra hydrogeranyl-2-methyl butyrate, 1, 2-benzenedicarboxylic acid and benzaldehyde that are reported as potential defense factors by various authors. Fatty acids with antimicrobial properties are also detected in both leaf and root samples of AMF inoculated chilli plants infected by damping-off pathogen. Dibutyl phthalate is a bioactive compound that was present at high level in leaf samples of AMF treated plants. Shahamin-B, a diterpenoid with anti-feedant properties was detected in chilli leaves due to AMF treatment. The results clearly reveal that the defense factors were induced by the mycorrhizal plants at higher level compared to non-mycorrhizal plants that in turn hinder the establishment of the pathogen. This strategy occurring in crop plants due to AMF colonization in roots is documented to prevent the incidence of pathogen attack, thereby enhancing the growth and yield of crop plants.

**Keywords**
Arbuscular mycorrhizal fungi, Chilli, Leaf and root, GC-MS analysis, Defense factors, Damping-off pathogen

**Introduction**

Arbuscular Mycorrhizal Fungi in symbiotic association with crop plants, improve not only the phosphorus nutrition of the host plant but also its growth and increases resistance to drought and diseases. AMF in its association with crop plants, receive greater attention for their application in sustainable agriculture (Min Chen *et al*. 2018). The Arbuscular Mycorrhizal Fungal functions range from stress alleviation to bioremediation in soils polluted with heavy metals. There is a great possibility of using mycorrhiza as a biological tool for sustainable agriculture in tropical countries (Orlando, 2003). AMF have been
used in the biological control of plant diseases, considering that they are widely distributed and establish long time relationships with the roots of most plant species. In several cases direct biocontrol potential has been demonstrated, especially for plant diseases caused by *Phytophthora*, *Rhizoctonia*, and *Fusarium* pathogens (Azcon and Barea, 1997). The colonization of common bean plants with AMF reduced the *Rhizoctonia* root rot disease incidence and severity due to accumulation of phenolic compounds and defense related enzymes (Abdel-Fattah et al., 2011). The mycorrhizal fungi are capable of producing morphological and physiological modifications in the root cells that increase plant’s tolerance to root diseases.

Mycorrhizal plants produce certain substances that have been indicated as copartners in resistance to the attack of root pathogens. Phenols, particularly flavonoids and isoflavonoids, are secondary metabolites involved in plant-microorganism interactions. The phytoalexins show defense role, which are important in plant protection mechanisms. Phormononetina and biocanina-A are the phenolic compounds with defense properties found in the roots of mycorrhizal plants (Lambais, 1996; Santos et al., 2017). Because of the defense mechanism by AMF in crop plants, this study is conducted to demonstrate the induction of defense factors in chilli plants due to the colonization of AMF in its rhizosphere.

**Materials and Methods**

**Analysis of defense factors by gas chromatography–mass spectrometry**

The analysis of the defense factors in the roots and leaves of AMF colonized chilli plants infected by damping-off pathogen was done by GC-MS (Seher et al., 2006).

**Preparation of plant extracts**

The root and leaf samples of *Pythium aphanidermatum* inoculated chilli plants that were both colonized and uncolonized by AMF were taken at the time of flowering for GC-MS analysis. The leaves and roots were dried at 30°C. The dried leaves and roots were crushed into fine particles and 10g of the powdered samples were soaked in 100ml of 100% methanol for 72 hours at room temperature with occasional mixing. The extract was then filtered using Whatman no.1 filter paper. The filtrates were concentrated in rotary vacuum evaporator and then the samples were filtered through membrane filter.

**GC-MS analysis**

The samples were analyzed using Clarus500 MS equipment. The column thickness of 30m x 0.25 micron was used. Helium was used as the carrier gas. Temperature program: 80-240°C at 8°C.min⁻¹, 240-300°C at 12°C.min⁻¹ and a 5 min hold at 300°C. The identification of components was accomplished using computer searches in commercial libraries.

**Results and Discussion**

The presence of defense factors in leaf and root samples of damping-off pathogen infected chilli plants that were AMF colonized and uncolonized was done by GC-MS and the compounds were identified as fatty acids and terpenoids.

**GC-MS pattern of chilli leaf analytes**

The leaf samples collected from AMF colonized and uncolonized chilli plants inoculated with *Pythium aphanidermatum* were used to identify the defense factors. The AMF treated and untreated chilli plant leaf samples showed the presence of various
compounds. AMF untreated chilli leaves revealed various biomolecules viz., pentanoic acid, 2- butyl ester, 1-2-bromododecanoic acid, 2-butene-1,4-diamine, 3-ethyl pentane, dibutyl phthalate, methyl palmitate, 1,2-propanedione, dodecane, 1-fluorodecyclic acid and ethyl-o-nitrobenzylidenentranilate. The AMF treated chilli plant leaf samples contained the defense factors viz., butanoic acid, tetra hydrogeranyl-2-methyl butyrate, propanoic acid, 1,2- benzenedicarboxylic acid, dibutyl phthalate and palmitic acid and these compounds were not found in AMF untreated chilli plants. Among the defense factors, the dibutyl phthalate, a diester of phthalic acid, is a bioactive defense factor that showed major peak in AMF treated plants, but the same compound was found in low level in AMF untreated plants. Shahamin-B, a diterpenoid with antifeedant properties is present only in the AMF treated leaves (Table 1).

GC-MS pattern of chilli root analytes

The defense factors present in root samples of AMF colonized chilli plants inoculated with Pythium aphanidermatum were carboxy-1-methyl-2-azetidinone, 1,2-cyclobutane dicarboxylic acid, benzaldehyde, didodecyl phthalate, butanoic acid and oleic acid. The carboxy-1-methyl-2-azetidinone is the compound present only in AMF treated root sample and in this compound the azetidinone is highly antimicrobial. 1, 2-cyclobutane dicarboxylic acid is a diester compound that exhibits defense role. Two fatty acids viz., butanoic acid and oleic acid with defense activity are present in the AMF colonized roots. Benzaldehyde is an aldehyde compound, which has antifungal and nematicidal activities. The didodecyl phthalate is a diester of phthalic acid, and it is a bioactive compound. Capric acid, a defense factor is also present in AMF colonized chilli plants.

Table 1 The defense factors detected by GC-MS in the leaves and roots of damping-off pathogen infected chilli plants colonized with AMF

| S.No | Defense factors in chilli leaves | Reference in which cited earlier. |
|------|---------------------------------|-----------------------------------|
| 1.   | Butanoic acid                   | Browing et al., 2005              |
| 2.   | Propanoic acid                  | Kitahara et al., 2004             |
| 3.   | 1,2-benzenedicarboxylic acid   | Sayed et al., 2008                |
| 4.   | Dibutyl phthalate               | Weiwei et al., 2008               |
| 5.   | Palmitic acid                   | Mishra and Sree, 2007             |
| 6.   | Tetra hydrogeranyl 2 methyl butyrate | Gokalp et al., 2003             |

| Defense factors in chilli roots | Reference in which cited earlier. |
|---------------------------------|----------------------------------|
| 1. Carboxy-1-methyl-2-azetidinone | Chavan and Nandini, 2007.    |
| 2. 1,2-cyclobutane dicarboxylic acid | Sayed et al., 2008.     |
| 3. Benzaldehyde                  | Shaukat et al., 2005.        |
| 4. Didodecyl phthalate           | Gokalp et al., 2003.         |
| 5. Butanoic acid                 | Browing et al., 2005.        |
| 6. Oleic acid                    | Kitahara et al., 2004.       |

The biomolecules present in AMF uncolonized roots were 5-hydroxy methyl-2-furanone, 1-methyl-2ethyl-pyrazolium bromide, 3-methyl-2-furan, 3-diazoundecan-2-one, decanoic acid, 2,5,6-trimethyl-4-ethyl pyrimidine, hexadecanoic acid, ethyl 5,5-
dimethyl cyclohex-2-en-1-one, 2,4-hexadienyl alpha-diazoacetate, capric acid, docosanedioic acid and ethyl phosphonic acid. But these compounds are absent in AMF colonized roots (Table 1). The results revealed comparatively more number and quantity of defense factors in the roots of AMF colonized chilli than the AMF uncolonized chilli plants.

Defense factors in the leaves

The GC-MS analysis of the AMF treated and untreated leaf samples of chilli plants inoculated with *Pythium aphanidermatum* showed the differences in the presence of various compounds. The AMF colonized chilli leaf samples contained compounds *viz.*, butanoic acid, tetrahydrogeranyl-2-methyl butyrate, propanoic acid, shahamin B, 1,2-benzenedicarboxylic acid, dibutyl phthalate and palmitic acid, whereas these compounds were not present in AMF uncolonized chilli. And these compounds are reported as potential defense factors by Kitahara *et al.* (2004) who studied the defense role of some fatty acids *viz.*, octanoic acid, decanoic acid, lauric acid, myristic acid, palmitic acid and stearic acid. Gonzalez (2007) studied the biological activities of diterpenoids and observed that shahamin-C is an antifeedant compound. Gokalp *et al.* (2003) studied the biological activity of some of the compounds *viz.*, 3-methyl-2-buten-1-ol, butyl-2- methyl butyrate, octyl acetate, decanol etc. Weiwei *et al.* (2008) identified that some of the volatile organic compounds *viz.*, dibutyl phthalate, acetic acid, o-xylene, 3-octanone, 2-decanne act as defense factors against fungi. Mishra and Sree (2007) identified the defense factor, palmitic acid in the leaves of mangrove plant by GC-MS analysis. Browing *et al.* (2006) found that butyric acid acts against soil borne fungal pathogen and nematode in strawberry. Shahamin-B is a diterpenoid that was present in the leaves of AMF colonized chilli and this compound possesses antifeedant properties. The presence of defense factors in AMF treated chilli leaves show the tolerance of the crops to disease causing pathogens.

Defense factors in the roots

The compounds present in AMF colonized chilli root samples are carboxy-1-methyl-2-azetidinone, 1,2-cyclobutanedicarboxylic acid, benzaldehyde, didodecyl phthalate, butanoic acid and oleic acid and these compounds also possess the defense role. Sayed *et al.* (2008) found that Phenazine-1-carboxylic acid acts against *Alternaria solani*. Benzaldehyde is found to be potential compound for the control of both fungal pathogens and phytoparasitic nematodes (Shaukat *et al.*, 2005). Walker *et al.* (2003) observed the defense factors *viz.*, butanoic acid, trans-cinnamic acid, O-coumaric acid, ferulic acid and vannilic acid in *Arabidopsis thaliana* by HPLC. Azetidinones are known to exhibit interesting biological activities (Chavan and Nandini, 2007). Capric acid is a fatty acid compound found in AMF colonized chilli plants. But the level of the capric or decanoic acid is less because the AMF colonized plants are induced to increase the synthesis of other defense factors.

Arbuscular mycorrhizal fungi are responsible for the alteration of genetic expression, mitochondrial and plastid proliferation of the host plants and increased production of terpenoids and jasmonic acid for defense against the pathogen (French, 2017). AMF have been used in the biological control of plant diseases, considering that they are widely distributed and establish long time relationships with the roots of most plant species. Mycorrhizal plants produce certain biomolecules that have been responsible for resistance to the attack of root pathogens. The presence of these defense factors in the leaf and root extracts of AMF colonized chilli
inoculated with *Pythium aphanidermatum* indicate that these compounds play a major role in the control of root pathogens. This implies that the host plants undergo certain important changes in the plant’s primary and secondary metabolism and regulation of the plant defense mechanisms (Fester and Hause, 2005; Sabin *et al*., 2012) altering the ability of the crops to sustain stresses. The transcriptomic and proteomic profiling of roots and leaves of wheat in symbiosis with *Xanthomonas translucens* confer stronger productivity and enhanced resistance to *X. translucens* (Fiorilli *et al*., 2018). During mycorrhiza establishment, induction of plant defense responses occurs, thus achieving an efficient activation of the plant immune responses leading to defense mechanisms to attack the pathogens (Jung *et al*., 2012). The protective effect of the plant – mycorrhizal symbiosis against pathogens leading to the crop protection is proved to be an altered metabolism of plants to overcome the stress caused by the disease causing pathogens.

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