A systematic review of association studies of common variants associated with idiopathic congenital talipes equinovarus (ICTEV) in humans in the past 30 years

Bi-Cheng Yong¹, Fu-Xing Xun¹, Lan-Juan Zhao², Hong-Wen Deng² and Hong-Wen Xu*¹

Abstract
The genetic cause of idiopathic congenital talipes equinovarus (ICTEV) is largely unknown. We performed a systematic review to describe the findings from 21 studies that have examined the genetic variants related to ICTEV, and to evaluate the quality of reporting. We found that ICTEV was positively associated with Hox family genes, collagen family genes, GLI3, N-acetylation genes, T-box family genes, apoptotic pathway genes, and muscle contractile family genes. Negative and controversial results were also discussed, and several genes associated with ICTEV were identified. Due to the limitation of the included studies, rare coding variants should be further investigated, sample size should be enlarged, and candidate genes should be replicated in larger ICTEV populations. Epigenetic study, pathways, chromosome capture, and detailed gene-environment interaction will also allow further elucidation of factors involved in ICTEV pathogenesis and may shed light on diagnosis and timely and accurate interventions.

Keywords: ICTEV, Etiology, Genetics

Background
Idiopathic congenital talipes equinovarus (ICTEV), also called clubfoot, is a common orthopedic birth defect found in 1 of 1000 infants (Wynne-Davies 1964). Males are more commonly affected than females by a ratio of 2 to 1 and the incidence of bilaterality is 50 %. The highest prevalence is found in Hawaiians and Maoris, and the lowest in Chinese (Chapman et al. 2000; Chung et al. 1969). The etiology of ICTEV is largely unknown, but it is universally acknowledged that gene-environment interaction plays a major role (Lochmiller et al. 1998). A genetic component to the etiology of clubfoot has been established in several studies (Bacino and Hecht 2014). In this paper, we systematically review and summarize studies performed on ICTEV probands and families regarding susceptible genes, pathways, and epigenetic changes.

Major findings in humans in the last 30 years are presented and ideas for further study are discussed.

Methods

Literature search strategy
This systematic review was conducted according to the guidelines of the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (Moher et al. 2009). The process began with the first author (YBC) performing a systematic electronic literature search of PubMed, Web of Science, China Wanfang Med Online, and Cochrane Library, for publications from Jan 1985 to Dec 2014. Queries to identify potentially relevant publications on the genetic study of patients with ICTEV were based on Boolean combinations of the following search terms: (“Talipes Equinovarus (MeSH) OR Club Foot (MeSH)” AND (Gene (MeSH) OR Genetics (MeSH))).

Study eligibility criteria
We limited this review to publications that were in English and Chinese, with full text available, concerning...
patients diagnosed with ICTEV. Family based, discordant sib pair association and case–control studies were included. Studies were excluded if they were case reports, dissertations, editorials, commentaries, or review articles.

Data extraction
The first author (YBC) screened the titles and abstracts of all the retrieved articles to determine whether they met the eligibility criteria, and appraised the methodological quality and evidence of each selected study. The second author (XFX) subsequently reviewed the accuracy and quality of the appraisal. Any disagreements were discussed between the first and the second authors until consensus was reached. The flow diagram describes the process used to select articles for this study, and the results of the literature search (Fig. 1). The following information was extracted from each study: (1) Year of publication, (2) Study objectives, (3) Study’s inclusion and exclusion criteria, (4) Sample size, (5) Method, (6) Gene examined, (7) Association or non-association study, (8) Results, and (9) Corresponding author. Due to the heterogeneity in the study design, in this review, meta-analysis was not performed for those observational gene(s) identified among the various reviewed studies. Two investigators independently assessed the quality of the reporting. Differences in the assessment were resolved by discussion.

Results

Literature search
We identified 902 relevant studies from various resources (Fig. 1); the included studies examined the genetic associations between relevant genes and ICTEV. After reading the titles, 715 irrelevant articles were excluded. A total of 67 abstracts addressing certain gene(s) associated with ICTEV were selected to be reviewed. After reviewing, 36 articles did not meet our criteria and were excluded, including case reports (n = 4), letters to the editor (n = 3), reviews (n = 16), animal only studies (n = 7), no specific genes studied (n = 5), and treatment related (n = 1). For the remaining 30 articles, full texts were screened. Nine more articles were excluded due to normal control being unavailable (n = 1), inconsistency between method and result (n = 1), and functional study and other reasons (n = 7). Twenty-one studies met the predetermined inclusion criteria (Fig. 1).

Description of the included studies (Table 1)
From the studies gathered, we identified 21 that investigated genes or pathways which may contribute to the occurrence of ICTEV. Among them, some genes are positively associated with ICTEV, while others have shown no evidence of association. The genes with positive results include: (1) Hox family genes (HoxA and HoxD), (2) collagen family genes (COL9A1 and COL1A1), (3) GLI3, (4) N-acetylation genes (NAT2), (5) T-box family genes (TBX3 and TBX4), (6) apoptotic pathway genes (Casp3, Casp8, Casp9, Casp10, Bid, Bcl-2, and Apaf1), and (7) muscle contractile family genes (TNNC2 and TPM1). The genes with negative or controversial results are CAND2, WNT7a, MYH, and DTDST. Our critical review of these genes affecting ICTEV is summarized as follows:

Positive gene results

Hox family genes
The HOX genes encode a highly conserved family of transcription factors that play fundamental roles in morphogenesis during embryonic development. This group of genes determines the segment identity and also helps pattern the developing embryo in the development of the axial skeleton and limbs. Hoxa13 and Hoxd10-Hoxd13 are expressed during specification of the hand/foot (autopod) (Favier and Dölle 1997). A variety of limb malformations including synpolydactyly and hand-foot-genital syndrome are known to be caused by specific mutations in HOXD13 and HOXA13, respectively (Muragaki et al. 1996; Mortlock and Innis 1997) In 2003, Wang identified 12 alleles at Hox4Ep-a microsatellite marker on HoxD gene; transmission of disequilibrium was found at the 12th allele, indicating that HoxD may be a potential gene for ICTEV (Wang et al. 2003). Wang and her colleagues, who identified the susceptibility of HoxD with ICTEV from the previous study group, further found SNP rs847154 located in 5′ flanking sequence of HoxD12 gene and SNP rs13392701 located in exon 1 of HoxD13 to be associated with ICTEV (Wang et al. 2005). In another study investigating HoxA in ICTEV patients, the authors found seven alleles at D7S516 microsatellite and the presence of transmission disequilibrium in Chinese populations (Wang et al. 2008). Variants in HoxA and HoxD clusters and altered transmission in multiplex and simplex families were validated in a larger Western population with ICTEV in 2009 (Ester et al. 2009).

Collagen family genes
COL9A1 encodes one of the three alpha chains of Type IX collagen, which is a minor (5–20 %) collagen component of hyaline cartilage. Lack of Type IX collagen is associated with early onset of osteoarthritis, epiphyseal dysplasia, and intervertebral disc degeneration (Czarny-Ratajczak et al. 2001; Alizadeh et al. 2005; Boyd et al. 2008). Liu et al. studied COL9A1 which maps to chromosome 6q12-13 and found that 84 nuclear pedigrees had transmission disequilibrium in SNPs rs592121 and
rs1135056, which are found in COL9A1 (Liu et al. 2007). Expression of COL9A1 mRNA is significantly higher in patients with ICTEV than in healthy human subjects. 

COL1A1 encodes the pro-alpha1 chains of Type I collagen, a fibril-forming collagen found in most connective tissues and abundant in bone, cornea, dermis, and tendon. Mutations in this gene are associated with osteogenesis imperfecta Types I–IV (Prockop et al. 1989; Takagi et al. 2015). Gene encoding collagen Type IV (COL1A1) was investigated in 2008 (Zhao et al. 2008). The results of this study show that expression of COL1A1 on mRNA levels is significantly higher in patients with ICTEV than in healthy patients. A $-161(T \rightarrow C)$ heterozygous mutation and a $+274(C \rightarrow G)$ homozygous mutation were also detected in the COL1A1 gene in patients with ICTEV, suggesting that COL1A1 mutations could cause ICTEV.
| Year | Study object | Study type | Inclusion criteria | Exclusion criteria | Race/Ethnicity | Sample size | Method | Gene examined | Association study | Results | Corresponding author |
|------|--------------|------------|-------------------|-------------------|----------------|-------------|---------|---------------|------------------|---------|---------------------|
| 2002 | To evaluate the relationship between R279W mutation in DTDST and occurrence of ICTEV | Family based | ICTEV patients and their families | Associate with other anomalies and syndromes | Hispanic and Nonhispanic White | 125 ICTEV probands and their parents | PCR, Genotyping | DTDST | N | Negative | Jacqueline T. Hecht |
| 2003 | To investigate possible association between ICTEV and HoxD gene | Family based | ICTEV patients and their families | Homozygous pedigree; Incomplete information | Chinese | 42 ICTEV probands and their parents | PCR, Genotyping, TDT | HoxD | Y | Positive | Shi-Jun Ji |
| 2004 | To investigate correlation between ICTEV and PAX5, PAX6 and TBX3 | Family based | ICTEV patients and their families | NOS | Chinese | 123 ICTEV probands in 41 nuclear family trios | PCR, Genotyping, TDT | PAX5 PAX6 TBX3 | Y | TBX3 Positive | Hong-wei Ma |
| 2005 | To study 2q31-33 SNP in ICTEV patients | Family based | ICTEV patients and their families | TEV from other causes | Hispanic and Nonhispanic White | 57 multiplex families and 83 simplex families | PCR, Genotyping | CASP8 CASP10 CFLAR | Y | CASP10 Positive | Jacqueline T. Hecht |
| 2005 | To study SNPs in HoxD10, HoxD12, HoxD13 and haplotypes distribution in ICTEV pedigree | Family based | ICTEV patients and their families | NOS | Chinese | 125 ICTEV probands | PCR, Genotyping, TDT | HoxD10 HoxD12 HoxD13 | Y | HoxD12 HoxD13 Positive | Chun-lian Jin |
| 2006 | To explore the association and mutation of GLI3 gene in ICTEV | Family based | ICTEV patients and their families | NOS | Chinese | 271 ICTEV probands and their parents 100 normal controls | PCR, Genotyping, TDT | GLI3 | Y | GLI3 Positive | Chun-lian Jin |
| Year | Study object | Study type | Inclusion criteria | Exclusion criteria | Race/Ethnicity | Sample size | Method | Gene examined | Association study | Results | Corresponding author |
|------|--------------|------------|-------------------|-------------------|----------------|-------------|--------|---------------|-----------------|---------|----------------------|
| 2006 | To study MTHFR C677T polymorphism, and maternal periconceptual folic acid supplement use, influenced risk of isolated clubfoot | Family based | ICTEV patients and their families | syndromic TEV | Not specified | 375 case-parent triads | PCR, Genotyping | MTHFR | N | MTHFR Positive | Linda Sharp |
| 2007 | To analyze SNPs within COL9A1 gene in ICTEV | Family based | ICTEV patients and their families | NOS | Chinese | 252 ICTEV probands in 41 nuclear family trios | PCR, Genotyping, ETDT | COL9A1 | Y | COL9A1 Positive | Chun-lian Jin |
| 2007 | To test the possible association between NAT2, NAT3 and ICTEV | Family based | ICTEV patients and their families | NOS | Hispanic and Caucasian | 56 extended multiplex families, 57 trios, and 157 Hispanic and 80 white non-Hispanic simplex trios | PCR, Genotyping, PDT, FBAT | NAT2, NAT3 | Y | NAT2 Positive | Jacqueline T. Hecht |
| 2007 | To study the association between Apoptotic genes and ICTEV | Family based | ICTEV patients and their families | NOS | Hispanic and Caucasian | 170 Caucasian families, 179 Hispanic families | PCR, Genotyping, FBAT, PDT | Casp3, Casp8, Casp9, Casp10, Bid, Bcl-2, Apaf1 | Y | Tested genes positive | Jacqueline T. Hecht |
| 2008 | To study possible association between ICTEV and HoxD | Family based | ICTEV patients and their families | Incomplete chart records | Chinese | 65 ICTEV patients 96 members from 32 families | PCR, Genotyping, TDT | HoxA | Y | HoxA Positive | Chun-lian Jin |
| 2008 | To detect the expressions of COL1A1 mRNA in 20 patients with ICTEV | Case-control | ICTEV patients | NOS | Chinese | 84 ICTEV probands and their parents 100 normal controls | PCR-DGGE, DNA sequencing | COL1A1 | N | COL1A1 Positive | Chun-lian Jin |
| Year | Study object | Study type | Inclusion criteria | Exclusion criteria | Race/Ethnicity | Sample size | Method | Gene examined | Association study | Results | Corresponding author |
|------|--------------|------------|--------------------|-------------------|-----------------|-------------|--------|--------------|------------------|--------|---------------------|
| 2009 | To test the hypothesis that CAND2 and WNT7a mutation associated with ICTEV | Case–control | ICTEV patients | Other syndromes including TEV | Not specified | 256 ICTEV patients and their parents 75 matched controls | PCR, DNA sequencing | CAND2, WNT7a | N | Negative | Jose A. Morcuende |
| 2009 | To detect the association between DTDST and ICTEV | Case–control | ICTEV patients | Associate with other abnormalities and syndromes | Chinese | 40 ICTEV patients 10 matched controls | RT-PCR, PCR-SSCP | DTDST | N | Positive | WU Xin-le |
| 2009 | To evaluate the expression level of CD-RAP | Case–control | ICTEV patients | Neuromuscular or syndromic TEV | Chinese | 25 ICTEV patients 5 controls | RT-PCR | CD-RAP | N | Positive | Chun-lian Jin |
| 2009 | To study HoxA, HoxD and IGFBP3 in patients with ICTEV | Family based | ICTEV patients | Chromosomal abnormality or syndrome | Hispanic and Non-Hispanic White | 179 extended families 331 simplex families 88 trios 144 families for validation | PCR, Genotyping, In Silico | HoxA, HoxD, IGFBP3 | Y | Tested genes positive Interactions with CASP3 | Jacqueline T. Hecht |
| 2010 | To study MYH genes in ICTEV patients | Case–control | ICTEV patients | Neuromuscular or other syndrome with TEV | Not specified | 200 patients 200 controls | PCR, DNA sequencing | MYH 1, MYH 2, MYH 3, MYH 8 | N | MYH genes not directly cause ICTEV | Jose A. Morcuende |
| 2012 | To assess whether variation in or around TBX4 is a common cause of non-syndromic clubfoot | Family based | ICTEV patients and their families | syndromic causes of clubfoot | Hispanic and Non-Hispanic White | 605 families | aCGH, PCR, Genotyping, DNA sequencing | TBX4 | Y | TBX4 variation is not a frequent cause | Jacqueline T. Hecht |
| 2012 | To interrogate muscle contractile complex genes in ICTEV | Family based and case–control | ICTEV patients and their families | NOS | Hispanic and Non-Hispanic White | 224 multiplex families 357 simplex families | PCR, Genotyping, DNA sequencing | Muscle contractile complex genes | Y | TNNC2 was identified in a validation group | Jacqueline T. Hecht |
Table 1 continued

| Year | Study object | Study type | Inclusion criteria | Exclusion criteria | Race/Ethnicity | Sample size | Method | Gene examined | Association study | Results | Corresponding author |
|------|--------------|------------|--------------------|------------------|-----------------|--------------|--------|---------------|-------------------|---------|----------------------|
| 2014 | To identify genetic risk factors associated with clubfoot | Case–control | ICTEV patients | Additional birth defects, known genetic, syndromes, developmental delay, mental retardation | Hispanic and non-Hispanic White | 396 ICTEV patients 1000 controls | Microarray genotyping, GWAS association study | Genome | Y | SNPs replication 12q24.31 FOKN3, SORCS1 MMP7/TMEM123 | Christina A Gurnett |
**GLI3 gene**
GLI3 encodes a protein which belongs to the C2H2-type zinc finger proteins subclass of the Gli family. The GLI3 protein localizes in the cytoplasm and activates patched Drosophila homolog (PTCH) gene expression. Mutations in the limb development related gene GLI3 have been associated with polydactyly (Volodarsky et al. 2014). SNP rs929387, located in exon 14 of the GLI3 gene, has transmission disequilibrium in 84 nuclear pedigrees, showing the association between the GLI3 gene and occurrence of ICTEV (Zha et al. 2006).

**N-Acetylation genes**
The NAT2 gene encodes an enzyme that functions to both activate and deactivate arylamine, hydrazine drugs and carcinogens. Polymorphisms in this gene are responsible for the N-acetylation polymorphism in which in human populations segregates into rapid, intermediate, and slow acetylator phenotypes. Since smoking is one of the known environmental risk factors for ICTEV and NAT2 metabolizes tobacco byproducts, Hecht et al. (2007) examined the variants of the NAT2 gene in 56 ICTEV multiplex families, 57 trios with a positive family history, and 160 simplex individuals. They reported a slight decrease in the expected number of homozygotes for the NAT2 normal allele in the Hispanic simplex trios. Significantly slow NAT2 acetylator phenotype was detected among the ICTEV patients, suggesting slow acetylation may be a risk factor for ICTEV.

**T-box family genes**
A possible association between TBX3 and ICTEV has been reported. The TBX3 gene is a member of a phylogenetically conserved family of genes that share a common DNA-binding domain, the T-box. T-box genes encode transcription factors involved in the regulation of developmental processes. The TBX3 protein is a transcriptional repressor and mutations in this gene affect limb development. The third allele short tandem repeat (D12S378) in the region of chromosome 12q24, where the TBX3 gene is located, was proven to have transmission disequilibrium in ICTEV patients, suggesting that TBX3 is a susceptible gene for ICTEV (Ren et al. 2004). TBX4 shares a similar structure with TBX3. Expression studies in mice and chickens show that TBX4 is expressed in developing hindlimb but not forelimb buds, suggesting a potential role for this gene in regulating limb development and specification of limb identity (Dai et al. 2014; Menke et al. 2008). TBX4 microdeletions/microduplications have been found in individuals with clubfoot (Alvarado et al. 2010). However, another study which examined the possible correlation between hindfoot specific gene TBX4 and ICTEV concluded that (1) there was minimal evidence indicating an association between TBX4 and clubfoot; (2) no pathogenic sequence variants were identified in the two known TBX4 hindlimb enhancer elements (Lu et al. 2012). However, the PITX-TBX4 pathway was studied and further investigated by the common disease-rare gene supporters (Gurnett et al. 2008). The study concludes that PITX1 or its pathways may be etiologically responsible for the increased incidence of ICTEV.

**Apoptotic pathway genes**
Cysteine-dependent aspartate-directed proteases (Caspases) are a family of cysteine proteases that play essential roles in apoptosis (programmed cell death), necrosis, and inflammation (Alnemri et al. 1996). The association between caspase genes and ICTEV was first studied in 2005 (Heck et al. 2005). The authors reported that the major allele of a variant in the CASP10 gene, a gene in the apoptotic pathway, is associated with ICTEV in simplex white and Hispanic trios. Further examination on the mitochondrial apoptotic related genes was performed to investigate their association with ICTEV (Ester et al. 2007). One SNP in each of the apoptotic genes (Casp3, Casp8, Casp9, Casp10, Bid, Bcl-2, and Apaf1) provided evidence implying correlation with ICTEV, suggesting the potential role of genetic variation in apoptotic genes in development of ICTEV (Gahlmann and Kedes 1990).

**Muscle contractile family genes**
Troponin (Tn), a key protein complex in the regulation of striated muscle contraction, is composed of three subunits (Tn-I, Tn-T, and Tn-C). The Tn-I subunit inhibits actomyosin ATPase. The Tn-T subunit binds tropomyosin and Tn-C. The Tn-C subunit binds calcium and overcomes the inhibitory action of the troponin complex on actin filaments. TNNC2 encodes Tn-C subunit and plays a key role in initiating muscle contraction in fast-twitching muscle fibers (Mckillop and Geeves 1993). In 2011, Weymouth et al. studied the association of the muscle contractile genes with ICTEV and identified two muscle contractile genes (TNNC2 and TPM1) associated with ICTEV (Weymouth et al. 2011).

TPM1 is a member of the tropomyosin family of highly conserved, widely distributed actin-binding proteins involved in the contractile system of striated and smooth muscles and the cytoskeleton of non-muscle cells. Tropomyosin functions in association with the troponin complex to regulate the calcium-dependent interaction of actin and myosin during muscle contraction. The associations of multiple SNPs in the TPM1 gene with ICTEV suggest a potential role of genes that encode contractile proteins of skeletal myofibers on the etiology of ICTEV (Shyy et al. 2010a, b).
Genome-wide association study
Besides the aforementioned candidate gene studies, a genome-wide association study was conducted in 396 isolated ICTEV patients and 1000 controls of European descent to identify novel genes for ICTEV (Zhang et al. 2014). The selected genetic variants from the genome-wide association study were further replicated with an independent cohort of 370 isolated ICTEV cases and 363 controls with the same ethnicity. The genome-wide association and replication study found an intergenic SNP on chromosome 12q24.31 between NCOR2 and ZNF664 that was significantly associated with ICTEV. However, Additional suggestive SNPs (Hox Genes, PITX1, TBX4, FOXN3, SORCS1 and MMP7/TMEM123) and identified pathways were not significant in the replication phase.

Negative or controversial results
Shyy et al. (2009) studied two candidate genes (CAND2 and WNT7a) and tested the hypothesis that mutations in these genes would be associated with the phenotype of ICTEV. After sequencing ICTEV patients, they found a polymorphism in each gene. However, the association results indicated that CAND2 and WNT7a are not the major genes that cause ICTEV. In a study exploring variation in MYH gene families, the authors sequenced the exons, splice sites, and predicted promoters of MYH genes in ICTEV patients (Shyy et al. 2010a, b). They found many SNPs, but none proved to be significantly associated with the phenotype of ICTEV. Bonafé et al. conducted research on diastrophic dysplasia sulphate transporter gene (DTDST) to test whether R279 W mutations are responsible for occurrence of ICTEV (Bonafé et al. 2002). Alterations in the coding region were not identified in 10 probands with ICTEV and a positive family history. The authors concluded that the R279 W mutation is no more frequent in this population of ICTEV probands than in controls. Contrary to this finding, another author reported in 2009 that DTDST gene mutations were detected in 27 children with ICTEV, but in only two normal children in the Chinese population, indicating the possible role of DTDST in ICTEV (He et al. 2009).

Other miscellaneous findings
In 2006, Sharp et al. found that children who carry the 677T variant of the MTHFR gene have a lower risk of ICTEV (Sharp et al. 2006). In 2009, Li and his colleagues compared the expression of CD-RAP (cartilage derived retinoic acid sensitive protein) in the abductor hallucis muscle from ICTEV and normal controls and found...
CD-RAP over-expressed in ICTEV patients, showing that CD-RAP might be a susceptibility gene of ICTEV (Li et al. 2009).

Discussion
In this systematic review, positive associations between genetic variants and ICTEV were established in several studies. Certain genetic variants were found to have significant association with ICTEV. However, it must be noted that conflicting and negative results were also identified which do not necessarily undermine their contribution to the occurrence of ICTEV.

ICTEV's genetic study history is described in Fig. 2. Simple major genes like X-linked genes, autosomal recessive genes and autosomal dominant genes used to be considered as possible genetic factors for ICTEV (Palmer 1964; Böök 1948; Wynne 1965). At the same time, another study concluded that multifactorial inheritance is significant in the etiology of ICTEV (Yamamoto 1979). Palmer et al. (1974) suggested simple major inheritance and multifactorial inheritance might be operating together to induce ICTEV. This theory was supported later by Wang et al. and Yang et al. who showed that a major gene component played a dominant role with additional minor contributions of multifactorial genes (Yang et al. 1987; Wang et al. 1988). In 1993, Rebbeck et al. (1993) rejected the non-Mendelian transmission pattern and concluded that the single Mendelian gene theory is adequate to explain the etiology of ICTEV. However, recent studies suggest that a polygenetic threshold model may explain its inheritance patterns. Contrary to the common disease-common variant hypothesis, one author introduced an alternative theory based on recent reports that rare genetic variants (with allele frequencies of <5 %) each confer a moderate risk with higher penetrance, which might be the genetic inheritance pattern of ICTEV. According to this theory, the PITX1-TBX4 transcriptional pathway directing early limb development is responsible for ICTEV (Dobbs and Christina 2012). However, further studies should be done to investigate other genes with low frequencies and how their variants affect ICTEV occurrence. Not only were gene mutations found in ICTEV, but chromosomal deletions and regulatory mutations were also reported.

Conclusion
Several genes were identified, though none of them could solely explain the occurrence of ICTEV. Because the sample size of most association studies was small, most of the studies included are largely underpowered. In those included studies, rare coding variants were rarely investigated. Candidate genes were not replicated in larger ICTEV populations. These limitations should be addressed in future studies. In the future, genetic research on ICTEV could be focused on at least five aspects. First, high-throughput sequencing instead of GWA studies might be used to detect replicable candidate genes. The sample size should be calculated and candidate genes must be replicated in other studies. Second, epigenetic sequencing examining regulatory mechanisms for RNA could be studied. Third, novel genes like FOXN3 and SORCS1 identified by the GWAS may be further investigated. Studying genes and their interactions could reveal common pathways which are responsible for the occurrence of ICTEV. Their functions and interactions are worthy of clarification. Fourth, recent advances in chromosome conformation capture may show more structural changes on a chromosomal level (Imakaev et al. 2012). Three-dimensional variants may shed light on ICTEV etiology and treatment. Fifth, in clinical practice, some patients do not have any recurrence although they are not completely compliant with the brace treatment, whereas other patients have a recurrence even though they are strictly compliant with the brace treatment (Zhao et al. 2014). It is conceivable that certain genes being activated at certain times results in the relapse of ICTEV. Therefore, integration of genomic risk assessment alongside other clinical investigations may help personalize the treatment of ICTEV and improve the prognosis in the era of precision medicine (Castaneda et al. 2015).

Authors' contributions
Dr. H-WX and Dr. B-CY designed the study. Dr. B-CY and F-XX searched articles and extracted the data. Dr. B-CY and Dr. L-JZ analyzed the data. Prof. H-WD and Dr. L-JZ revised the manuscript critically. All authors read and approved the final manuscript.

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Competing interests
The authors declare that they have no competing interests.
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