Cell-specific histone modifications in atherosclerosis (Review)

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Abstract. Histone modifications are the key epigenetic mechanisms that have been identified to regulate gene expression in many human diseases. However, in the early developmental stages, such as in utero and the postnatal stages, histone modifications are essential for gene regulation and cell growth. Atherosclerosis represents a classical example of the involvement of different cell types, and their cumulative effects in the development of atheroma and the progression of the disease. Post translational modifications on proteins either induces their functional activity or renders them inactive. Post translational modifications such as methylation or acetylation on histones have been well characterized, and their role in enhancing or inhibiting specific gene expression was clearly elucidated. In the present review article, the critical roles of different histone modifications that occur in atherosclerosis have been summarized. Different histone proteins have been identified to serve a critical role in the development of atherosclerosis. Specifically, histone methylation and histone acetylation in monocytes, macrophages, vascular smooth muscle cells and in endothelial cells during the progression of atherosclerosis, have been well reported. In recent years, different target molecules and genes that regulate histone modifications have been examined for their effects in the treatment of atherosclerosis in animal models and in clinical trials. An increasing body of evidence suggests that these epigenetic changes resulting from DNA methylation and non-coding RNA may also be associated with histone modifications, thereby indicating that novel therapeutic strategies can be developed by targeting these post translational modifications, which may in turn aid in the treatment of atherosclerosis.

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1. Introduction

During the progression of atherosclerosis, oxidized low-density lipoprotein (oxLDL) initiates endothelial dysfunction and increases the expression of vascular cell adhesion molecule-1 (VCAM-1), P-selectin, and chemokine monocyte chemoattractant protein-I (MCP-I) that recruits monocytes, T-lymphocytes and platelets. Monocytes are stimulated by monocyte-colony stimulating factor (M-CSF), and pro-inflammatory chemokines and cytokines, which differentiate them into macrophages. Macrophages then engulf oxLDL by receptor-mediated phagocytosis and transform themselves into lipid-laden foam cells. Accumulated foam cells and immune cells in the vessel wall develop an identifiable pathological condition called, the ‘fatty streak’. This first symptom represents asymptomatic and non-stenotic plaque, and is the early sign for the development of atherosclerosis. Subsequently, the smooth muscle cells (SMCs) triggered by cytokines, chemokines, and growth factors, gain the ability to proliferate and migrate to develop a fibrous cap covering the atherosclerotic core. In this advanced pathological stage of atherosclerosis, matrix degradation, cytotoxic T-cells and cholesterol crystals accumulate to form a lipid-rich necrotic core. In the very late stage of atherosclerosis, apoptosis in the SMCs of the fibrous cap occurs which causes plaque instability and thrombosis. As a result, distal embolism occurs which leads to blockage in the cerebral arteries. Thus, atherosclerosis in coronary arteries and in carotid arteries results in coronary heart disease and ischemia in the brain which in severe conditions may cause significant morbidity and mortality (1). During the progression of atherosclerosis from ‘fatty streak’ to atheroma development, changes in the gene expression in different cell types have been reported. Below, we describe the de novo epigenetic changes, specifically histone modifications which were reported in
different cell types that are involved in the development of atherosclerosis.

2. Histone methylation

DNA normally remains coiled to the histones proteins (H1, H2A, H2B, H3 and H4) in eukaryotes systems. Tight control over gene expression occurs accurately and effectively via chromatin compaction or signaling, or by activating other signaling events such as phosphorylation, acetylation, ubiquitination and methylation on histones. These modifications on the histone proteins are collectively referred to as histone post-translational modifications (PTMs). Since these histone modifications have been identified to either induce or inhibit transcription, their role in regulating gene expression, which is specific to each type of modification on different histones, has been well studied (2,3).

Histone proteins are predominantly methylated on arginine and lysine residues. Multiple modifications can occur on these histone proteins such as mono-methylation (me1), di-methylation (me2) or tri-methylation (me3). The Su(var)3-9, Enhancer-of-zeste and Trithorax domain containing proteins (SET-domain-containing proteins), Disruptor of telomeric silencing 1-like (DOT1-like proteins), and Calmodulin-lysine N-methyltransferase are three families of proteins which transfer methyl groups from S-adenosyl methionine to histones. On the contrary, the two family of proteins which function as demethylases have also been identified which include amine oxidases and jumonji C (JmJc)-domain-containing proteins, and iron-dependent dioxygenases (4).

Histone methylation as a prognostic marker in atherosclerosis.

Methylation on H3K4 has been correlated with stage-specific progression of atherosclerosis mediated by GCN5L and MYST1, MLL2/4 proteins (3). In ApoE-/-mice, high levels of homocysteine (Hcy) induced EZH2 expression was detected which led to H3 at lysine 27 (H3K27) tri-methylation (5). The polycomb complex protein, BMI-1, is one of the proteins in PRC1-like complex, which is required to inhibit the expression of target genes through histone modification such as mono-ubiquitination on H2AK119. As part of PRC1 complex, BMI-1 promotes E3 ubiquitin-protein ligase activity (6). Global increase in trimethylation of H3K27 was observed in atherosclerotic plaques at late stage in the pathology. However, decreased trimethylation of H3K27 was not associated with either BMI-1, histone methyltransferase EZH2, or histone demethylase JMJD3, which targets H3K27 (7). Increased expression of H2AK119Ub and H2BK120Ub has been reported in diabetic cardiomyopathy, and treatment with Esculetin was identified to mitigate the renin-angiotensin system, oxidative stress (Keap1) and cell proliferation (Ki67) via H2A/H2B ubiquitination, which eventually attenuated metabolic alterations, hypertension, cardiomyocytes hypertrophy, and fibrosis in the hearts of type 2 diabetic rats (8). The S-Adenosyl homocysteine (SAH) level in the plasma holds a strong correlation with the size of atherosclerotic lesion and in the pathogenicity of hyperhomocysteinemia during atherosclerosis. The increased levels of SAH in plasma was reported to promote expression of glucose-regulated protein-78 and CEBP-homologous protein, which are indicators of endoplasmic reticulum stress resulting from the repression of trimethylation on histone H3 at lysine 9 (H3K9) (9). The histone-lysine N-methyltransferase (MLL5) activity, which serves an important role in cell cycle regulation, and SNP variations at this locus, have been attributed with coronary artery disease. The dominant genotype of rs12671368 and the recessive genotype of rs2192932 were found to provide protective effects in a study that was carried out on Chinese Han population (10).

Monocytes and macrophages. One of the primary functions of G9a is to mono-methylate and di-methylate H3K9 by forming G9a-GLP (G9a-like protein) complex, which plays a critical role in the regulation of gene expression in monocytes (11). In hyperhomocysteinemic ApoE-/-mice that were treated with methionine, decreased expression of G9a as well as H3K9 di-methylation was reported to promote apoptosis in the macrophages affecting the foam cell formation and also plaque stability (12). Inhibiting trimethylation of H3K4me3 and H3K27me3 was found tightly associated with increased expression of specific marker genes such as KLF4, IRF8, HOXA and FOXO which stimulate monocyte-phagocyte (MP) differentiation (13). In monocytes, reduced methylation on H3K9 and H3K27 was observed in inflammatory cells (14). Monocytes typically gets reprogrammed into pro-inflammatory phenotype by oxLDL, and this was found to increase during aerobic glycolysis. Analysis of the monocytes from patients with asymptomatic and symptomatic carotid plaques revealed that monocytes from patients with symptomatic carotid plaque expressed more pro-inflammatory cytokines after stimulating with LPS. Lower levels of H3K4me3 and H3K27me3 were found on the promotor region of TNF-α. At the same time, reduced glycolytic rate limiting enzymes hexokinase 2 and 6-phosphofructo-2-kinase/fructose-2,6-biphosphatase 3 expressions were detected in the carotid plaques of symptomatic compared to asymptomatic patients with carotid stenosis (15). The suppressor of variegation 3-9 homologuel (SUV39H1) is a histone methyltransferase which mediates trimethylation at H3K9, and acetylation at lysine residue 266 was found to decrease its activity. SIRT1 was reported to recruit and deacetylate SUV39H1, and this deacetylation of SUV39H1 contributes to the increased functional activity of SUV39H1 thereby, the levels of H3K9me3 will get elevated (16). In macrophages, high-glucose stimulation reduced trimethylation on H3K9 which was mediated through SUV39H1. This further led to increased expression of the inflammatory cytokines such as IL-6, IL-12, p40, MIP-1α, and MIP-1β (17). In peripheral blood monocytes (PBMC) of patients with type 2 diabetes mellitus (T2DM), Set7 induced mono-methylation at H3K4 was found increased at the promotor of NF-kB p65, leading to the activation of NF-kB-dependent oxidative and inflammatory signaling. This in turn elevated the expression levels of the downstream pro-inflammatory genes ICAM-1 and MCP-1. Set7 was also found associated with the oxidative marker 8-isoPGF2α and brachial artery flow-mediated dilation (FMD) (18). The oxLDL, TLR-2 and TLR-4 agonists can induce human monocyte differentiation into foam cells by stimulating trimethylation of H3K4 at the promotor regions of different
genes such as IL-6, IL-18, TNF-α, MCP-1, MMP2, MMP9, CD36, and SR-A (19). In Table 1, we summarize histone modifications occurring on different genes in monocytes and macrophages that are regulated during atherosclerosis.

**Endothelial cell.** TNF-α stimulates the expression of ICAM1 which causes leukocyte adhesion to endothelial cells via reducing methylation of H3K9 and H3K27 residues. Overexpression of G9α or inhibition of KDM4D can increase the expression of H3K9me2 and reduce adhesion of leukocytes to endothelial cells (20). In endothelial cells, SAH which is a potent inhibitor of S-adenosylmethionine-dependent methyltransferases, was found to induce endothelial cell activation by increasing the expression of adhesion molecules and cytokines, by stimulating NF-κB pathways. SAH reduced the expression of EZH2 and tri-methylation of H3K27 (21). In endothelial cells, TNF-α increased the expression of histone dimethylases KDM1B and KDM7A which further induced H3K9me2 di-methylation. KDM7A mediated upregulation of ICAM1 was found associated with decreased expression of the transcription factor EB which positively regulates the activity of lysosome formation (22). Arginine methyltransferase inhibitors, AMI-5 and AMI-1, inhibit migration of endothelial cells to prevent pathological angiogenesis by stimulating the formation of heterochromatin. This was reported to be mediated through downregulation of arginine and lysine histone methyltransferases (HMTs) (23). PHD finger protein 8 (PHF8) was shown to induce endothelial cell migration and formation of capillary-like structures by decreasing di-methylation of E2F4, suggesting its functions as a repressor (24). In human umbilical vein cell line EA.hy926, elevated di- and tri-methylation (H3K4me2/3) on MCP-1 promotor was found regulated by increased H3K4 histone methyltransferase MLL3, menin and SET7 along with a concomitant decrease in the demethylase LSD1 expression in high glucose conditions. Puerarin was also reported to decrease the expression of MCP-1 by reversing the expression of methyltransferases and demethylases (25). During the progression of atherosclerosis, a subset of dendritic cells also takes up the lipid molecules and this leads to the formation of foam cells. Elevated cholesterol levels in plasma indicates the onset of atherosclerotic phenotype in T-cells compared to anti-atherosclerotic phenotype in normal group (26). In human aortic endothelial cells (HAECs), Set7 was found to stimulate NF-κB-dependent oxidative and inflammatory signaling by mono-methylation of H3K4 at its promotor (18). In Endothelial cells, EZH2 was reported to be induced by LDL. This was observed to be mediated through the transcription activator, myocyte enhancing factor-2, which decreased the expression of KLF2. The decreased expression of KLF2 was found to affect the expression of different cellular regulators of atherosclerosis such as thrombomodulin, endothelial NO synthase, and plasminogen activator inhibitor-1, which promote accumulation of platelets on endothelial cells (27). ‘Hyperglycemic memory’ is a condition which induces deleterious effects in patients even after more than five years of the onset of glycemic control. In bovine aortic endothelial cells, hyperglycemia induced expression of NF-κB-p65 was found to upregulate the expression of H3K4m1, and this was associated with the reduced expression of di- and tri-methylation of histone H3 (H3K9m2 and H3K9m3). This reduction in histone methylation was identified as mediated through histone demethylase LSD1 (28,29). Hyperglycemia also stimulated the expression of p65 in aortic endothelial cells, which induced the expression of MCP-1 and VCAM-1. This suggests that by decreasing the levels of mitochondrial superoxide production or superoxide-induced α-oxoaldehydes, inflammation under hyperglycemic condition can be prevented (29). In Table II, we summarized the histone modifications occurring exclusively in the endothelial cells during atherosclerosis, affecting different target gene loci.

**Vascular SMCs.** Under in vitro conditions, high glucose increases the expression of KDM3a, causing reduction in H3K9 di-methylation at AGTR1 and ROCK2 loci. This indicates that the proliferation and migration of vascular SMCs (VSMCs) is enhanced through activation of Rho/ROCK and AngII/AGTR1 pathways (30). Increased histone acetylation at H3K9 and H3K27 led to a parallel decrease in the methylation at H3K9 and H3K27 which was found in the advanced atherosclerotic lesions (14). VSMCs upon treatment with Acrolein, which is an air pollutant and consists of cigarette smoke, was identified to induce histone acetylation at H3K9 and H3K56. Simultaneous decrease in the histone tri-methylation at H3K9 and H3K27me3 Decreased HOXA, FOXO, KLF4, IRF8 and TNF-α. KDM7A mediated upregulation of ICAM1 was found associated with decreased expression of the transcription factor EB which positively regulates the activity of lysosome formation (22). Arginine methyltransferase inhibitors, AMI-5 and AMI-1, inhibit migration of endothelial cells to prevent pathological angiogenesis by stimulating the formation of heterochromatin. This was reported to be mediated through downregulation of arginine and lysine histone methyltransferases (HMTs) (23). PHD finger protein 8 (PHF8) was shown to induce endothelial cell migration and formation of capillary-like structures by decreasing di-methylation of E2F4, suggesting its functions as a repressor (24). In human umbilical vein cell line EA.hy926, elevated di- and tri-methylation (H3K4me2/3) on MCP-1 promotor was found regulated by increased H3K4 histone methyltransferase MLL3, menin and SET7 along with a concomitant decrease in the demethylase LSD1 expression in high glucose conditions. Puerarin was also reported to decrease the expression of MCP-1 by reversing the expression of methyltransferases and demethylases (25). During the progression of atherosclerosis, a subset of dendritic cells also takes up the lipid molecules and this leads to the formation of foam cells. Elevated cholesterol levels in plasma indicates the onset of atherosclerotic phenotype in T-cells compared to anti-atherosclerotic phenotype in normal group (26). In human aortic endothelial cells (HAECs), Set7 was found to stimulate NF-κB-dependent oxidative and inflammatory signaling by mono-methylation of H3K4 at its promotor (18). In Endothelial cells, EZH2 was reported to be induced by LDL. This was observed to be mediated through the transcription activator, myocyte enhancing factor-2, which decreased the expression of KLF2. The decreased expression of KLF2 was found to affect the expression of different cellular regulators of atherosclerosis such as thrombomodulin, endothelial NO synthase, and plasminogen activator inhibitor-1, which promote accumulation of platelets on endothelial cells (27). ‘Hyperglycemic memory’ is a condition which induces deleterious effects in patients even after more than five years of the onset of glycemic control. In bovine aortic endothelial cells, hyperglycemia induced expression of NF-κB-p65 was found to upregulate the expression of H3K4m1, and this was associated with the reduced expression of di- and tri-methylation of histone H3 (H3K9m2 and H3K9m3). This reduction in histone methylation was identified as mediated through histone demethylase LSD1 (28,29). Hyperglycemia also stimulated the expression of p65 in aortic endothelial cells, which induced the expression of MCP-1 and VCAM-1. This suggests that by decreasing the levels of mitochondrial superoxide production or superoxide-induced α-oxoaldehydes, inflammation under hyperglycemic condition can be prevented (29). In Table II, we summarized the histone modifications occurring exclusively in the endothelial cells during atherosclerosis, affecting different target gene loci.

**Table I.** Histone methylation markers in monocytes and macrophages in atherosclerosis.

| Epigenetic mediator | Histone modification | Expression in atherosclerosis | Target genes |
|--------------------|----------------------|-------------------------------|--------------|
| G9a, SUV39H1       | H3K9                 | Decreased                     | IL-6, IL-12p40, MIP-1α, and MIP-1β |
| Set7               | H3K4me1              | Increased                     | NF-κB p65    |
|                    | H3K4me3              | Decreased                     | HOXA, FOXO, KLF4, IRF8, TNFα, IL-6, IL-18, MCP-1, MMP9, MMP2, CD36 and SR-A |
|                    | H3K27me3             | Decreased                     | HOXA, FOXO, KLF4, IRF8 and TNF-α |

SUV39H1, suppressor of variegation 3-9 homologue 1; H3K9, histone H3 at lysine 9; H3K4, histone H3 at lysine 4; H3K27, histone H3 at lysine 27; me 1/3, mono/tri-methylation; Set7, SET domain containing lysine methyltransferase 7; IL, interleukin; MIP, macrophage inflammatory protein; NF-κB, nuclear factor-κB; HOXA, homeobox A; FOXO, forkhead box O; KLF2, Kruppel like factor 2; IRF8, interferon regulatory factor 8; TNFα, tumor necrosis factor-α; MCP-1, monocyte chemoattractant protein 1; MMP, matrix metalloproteinase; CD, cluster of differentiation; SR-A, macrophage scavenger receptor 1.
was also observed. Co-culturing with N-acetyl cysteine has been shown to inhibit the toxicity of acrolein. In VSMCs this was correlated with significant decrease in histone acetylation at H3K9. At the same time, other histone modifications were also observed such as di-methylation at H3K4, and phosphorylation and di-methylation at H3S10 (31). In VSMCs, treatment with Roflumilast, which is a PDE4 inhibitor, was reported to affect TNF-α functions through VCAM-1 by promoting the expression of the cyclic AMP effector, Epac. Upregulation of Epac subsequently reduced histone di-methylation H3K4me2 at VCAM-1 and promoted its expression. This epigenetic action of Roflumilast on VCAM-1 can be reversed by treatment with an HDAC inhibitor (32). High-glucose conditions were also reported to increase JMJD2A expression in the VSMCs of the rat carotid arteries which enhanced cell proliferation. Tri-methylation of H3K9 was inversely correlated with VCAM-1 during atherosclerosis was summarized in Table III. In utero programming and postnatal histone methylation in atherosclerosis. Maternal hypercholesterolemia could have direct impact in inducing atherosclerosis in the offspring as reported in ApoE-/-Leiden mice (36). Compared with the offspring from wild-type mothers, the offspring from ApoE-/-mothers showed differences in histone tri-methylation in endothelial cells and VSMCs, which caused different responses to high cholesterol diet. This study highlights the importance of in-utero programming and postnatal histone modification in atherosclerosis which if addressed in early stages may help in treating the disease (37). In Fig. 1, we summarize the histone methylation in different cell types and their effects on regulating gene expression during atherosclerosis.

### 3. Histone acetylation

Histone acetylation is regulated by two key enzymes, histone acetyltransferases (HATs) and histone deacetylases (HDAC). By transferring an acetyl group to lysine side chains of histone proteins, HATs weaken the binding between histone and DNA. On the contrary, HDACs remove the acetyl group on the histones to increase the interaction between histone and DNA. HATs are further categorized into two groups, type-A and type-B (38). HDACs regulate transcription and cell cycle progression by deacetylating lysine residues located on the N-terminal of the core histone proteins (H2A, H2B, H3 and H4). HDACs are further classified into four Classes I, II, III and IV. Classes I, II and IV represent ‘classical’ HDACs which include 11 genes, and in Class III, 7 genes are included.

### Table II. Histone methylation markers in endothelial cells in atherosclerosis.

| Epigenetic mediator | Histone modification | Expression in atherosclerosis | Target genes |
|---------------------|----------------------|-------------------------------|--------------|
| G9a                 | H3K9m2               | Decreased                     | ICAM1        |
| KDM4D               | H3K9m2               | Increased                     | ICAM1        |
| KDM7A               | H3K9m2               | Increased                     | E2F4         |
| PHF8                | Demethylation        | Increased                     | MCP-1, p65   |
| MLL3                | H3K4m2, H3K4m3       | Increased                     | MCP-1, p65   |
| LDS1                | H3K4m2, H3K4m3       | Decreased                     | NF-κB        |
| SET7                | H3K4m1               | Increased                     | KLF2         |
| EZH2                |                      |                               |              |

H3K9, histone H3 at lysine 9; H3K4, histone H3 at lysine 4; MCP-1, monocyte chemoattractant protein 1; NF-κB, nuclear factor-κB; KLF2, Krüppel like factor 2; KDM 4D/7A, lysine demethylase 4D/7A; PHF8, PHD finger protein 8; MLL3, mixed-lineage leukemia protein 3; LDS1, Loey-Dietz syndrome 1; SET7, SET domain containing lysine methyltransferase 7; EZH2, enhancer of zeste homolog 2; ICAM1, intercellular adhesion molecule 1; E2F4, E2F transcription factor 4.

By stimulating A2b adenosine signaling pathway, PCAF/GEN5 which are histone H3K9 acetyltransferases, and WDR5 which consists of H3K4 methyltransferase complex, prevents their recruitment at the promoter region of CIITA transcription, thereby inhibiting the expression of CIITA in a STAT1-dependent manner (35). Specific methylation patterns in histone proteins that have been reported in the VSMCs during atherosclerosis was summarized in Table III.
which are named as sirtuins (39). Based on the type of modification occurring on the histones, gene expression is either favored or inhibited. This characteristic feature of the PTMs on histones and their role in transcriptional regulation is developmental and tissue specific. However, such histone modifications, to regulate gene expression, are frequently observed in many pathological conditions. Such modifications occurring in atherosclerosis are discussed below.

Histone acetylation as an indicative marker in atherosclerosis.

In the heart tissues of New Zealand white rabbits that were treated with high cholesterol diet and atorvastatin, the mRNA levels of ACE2 was reported to be elevated when compared with control high cholesterol diet. Notably, atorvastatin was identified to promote histone H3 acetylation (H3‑Ac) at the promoter region of ACE2 (40). High plasma levels of Hcy was identified not only to increase abnormal DNA methylation in the cells but also elevates the levels of N‑methyl‑d‑aspartate receptor‑1 (NMDAR1), DNA (cytosine‑5)‑methyltransferase‑1 (DNMT1) and MMP‑9, via acetylation on H3K9 and by inhibiting the expression of HDAC1. These regulatory proteins were identified as possible targets that have high potential to cause heart failure as reported in cardiomyocytes (41).

Monocytes and macrophages.

The ‘Silent mating type information regulator two homologue one’ (SIRT1), which belongs to class III HDAC, requires Nicotinamide adenine dinucleotide (NAD+) as a cofactor for its functional activity. The role of SIRT1 has been implicated in many cellular processes such as in cell cycle progression, inflammation, DNA damage, apoptosis, autophagy, aging as well as in metabolic disease (42). SIRT1 mediates transfer of the acetyl group from proteins to the targeting co‑substrate by removing the nicotinamide ribosyl bond of NAD+. In the peripheral blood mononuclear cells (PBMCs) of patients with T2DM, increased transcription of HDAC3 was reported. SIRT1, which was identified as a protective marker of cardiovascular diseases, was found reduced in PBMCS of these T2DM patients. In addition, the pro‑inflammatory markers TNF‑α, IL‑6, MCP‑1, IL1‑β, NFκB, TLR2, and TLR4 were also reported to be upregulated in the

| Table III. Histone methylation markers in vascular smooth muscle cells in atherosclerosis. |
|---------------------------------------------|-----------------------------|---------------------------------------------|-----------------------------|
| Epigenetic mediator | Histone modification | Expression in atherosclerosis | Target genes |
|------------------|-----------------|--------------------------|--------------|
| KDM3a | H3K9 | Increased | AGTR1, ROCR2 |
| Epac | H3K9m3 | Increased | ICAM1 |
| JMJD2A | H3K9 | Increased | MCP‑1, IL‑6 |
| EZH2 | H3K27 | Increased | - |
| SUV39H1 | H3K9m3 | Decreased | TNF‑α |

H3K9, histone H3 at lysine 9; H3K4, histone H3 at lysine 4; H3K27, histone H3 at lysine 27; ICAM1, intercellular adhesion molecule 1; IL‑, interleukin; MCP‑1, monocyte chemoattractant protein 1; TNF‑α, tumor necrosis factor‑α; AGTR1, angiotensin II receptor type 1; ROCR2, arginine utilization regulatory protein RocR; KDM3a, lysine demethylase 3A; Epac, exchange factor directly activated by cyclic adenosine monophosphate; JMJD2A, jumonji domain-containing protein 2A; EZH2, enhancer of zeste 2 polycomb repressive complex 2 subunit; SUV39H1, suppressor of variegation 3-9 homolog 1.

Figure 1. Histone methylation occurring during atherosclerosis. Methylation of different histone proteins and their effects on gene expression in various cell types during atherosclerosis is presented. Red arrows indicate upregulation and blue arrows indicate downregulation.
TABLE IV. Histone acetylation markers in monocytes and macrophages in atherosclerosis.

| Enzyme   | Expression in atherosclerosis | Target genes                        |
|----------|-------------------------------|-------------------------------------|
| HDAC7    | Increased                      | IL-6, MCP-1                         |
| P300     | Decreased                     | Transcription regulation            |
| HDAC9    | Increased                      | MMP12, ABCA1, ABCG1, PPAR-γ         |

HDAC, histone deacetylase; IL, interleukin; MCP-1, monocyte chemoattractant protein 1; ABCA1, adenosine triphosphate-binding cassette subfamily A member 1; adenosine triphosphate-binding cassette subfamily G member 1; PPAR-γ, peroxisome proliferator activated receptor-γ.

Figure 2. Histone acetylation occurring during atherosclerosis. Acetylation of different histone proteins and their effects on regulating gene expression in monocytes and macrophages during atherosclerosis is presented. Red arrows indicate upregulation and blue arrows indicate downregulation. HDAC, histone deacetylases; IL-6, interleukin 6; MCP-1, chemokine monocyte chemoattractant protein-1; MMP12, matrix metalloproteinase 12; ABCA1, adenosine triphosphate binding cassette subfamily A member 1; ABCG1, adenosine triphosphate binding cassette subfamily G member 1; PPAR-γ, peroxisome proliferator-activated receptor-γ.

plasma of the T2DM patients. Further, DBC1 (deleted in breast cancer 1), which represses HDAC3, was also reported to be decreased in the PBMCs of patients with T2DM (43). Myeloid HDAC3 deficiency was reported to have plaque stabilization effect by increasing the deposition of collagen, HDAC3 expression can potentially induce macrophage polarization, favoring the formation M2 phenotype. These M2 macrophages by secreting TGFβ1 were shown to initiate VSMCs to secrete excess collagen (44,45). Under hypercholesterolemic conditions, the orphan nuclear receptor NR4A1, which can effectively reduce IL-6 and MCP-1 expression, was found increased due to the recruitment of p300 acetyltransferase. As a result, HDAC7 (histone deacetylase 7) was found repressed, which contributes to the expression of NR4A1 in human monocytes, THP-1 and U937 (46). The histone deacetylase 9, HDAC9, was reported to be associated with MMP12 expression in the pro-inflammatory macrophages M2 and M4, in the advanced human plaques. However, the presence of both these types of macrophages could not be correlated with genes regulating plaque stabilization or thrombosis (47). HDAC9 deficient cells were identified to express increased levels of ABCA1, ABCG1, and PPAR-γ. This was identified to be mediated through upregulation of H3 and H4 acetylation, preventing the cholesterol efflux. Moreover, HDAC9 knockout macrophages were identified to be secreting lower levels of pro-inflammatory cytokines and showed prominent expression of M2 phenotype markers, and at the same time, it simultaneously decreased the expression of M1 markers (48,49). In Table IV, we summarized acetylation events on different residues of histones in the above monocytes and macrophages occurring during atherosclerosis. The same was represented in Fig. 2.

**Other cell types.** The protein ‘General Control Non-repressible 5’ (GCN5)-related N-acetyltransferase (GNAT), mediates transfer of the acyl group from acyl coenzyme A (acyl-CoA) to different proteins substrates which are involved in transcription, cell proliferation, antioxidant functions, antibiotic resistance and detoxification (50). The p300/CREB-binding protein (CBP) which is a HAT, acetylates non-histone proteins and controls transcription and DNA repair mechanisms in the cells (51). When compared between the cardiac mesenchymal stem cells from normoglycemic subjects, cardiac mesenchymal stem cells from type 2 diabetic patients had decreased levels of H3K9Ac and H3K14Ac acetylations, which are caused primarily due to the reduced activity of GCN5-related N-acetyltransferases (GNAT) p300/CBP-associated factor. Also its isoform, GCN5α, decreased proliferation and differentiation of MSCs. Moreover treatment with the HAT activator, pentadecylidenemalonolate 1b (SPV106) not only recovered the levels of H3K9Ac and H3K14Ac acetylation, but also decreased CpG hypermethylation in the genomic DNA, suggesting the involvement of two different epigenetic mechanisms regulating at the same time in these cells (52).

4. Targeting histone methylation and histone acetylation in the treatment of atherosclerosis

As cell specific histone modifications have been identified during atherosclerosis, targeting the enzymes which regulate these *de novo* changes can be a promising strategy for the treatment of atherosclerosis. This strategy is favored by two important features. Frist, like all epigenetic therapies, targeting histone modification alone do not affect the genetic component in the cells. Second, advances in the structural biology greatly assists the researchers to develop unique compounds that can specifically target the desired histone modification, such as the DNMT inhibitor (2’-deoxy-5-azacytidine, DAC) which targets H3K9 dimethylation, and the HDAC inhibitor TSA, which targets H3K27 trimethylation (53). Since any given histone modification is not unique for a specific disease and as they can be observed in different pathologies, drugs targeting a specific histone modification can be repurposed for other disease conditions. In addition, use of these specific histone targeting drugs as combination therapy with other therapeutic drugs appears to be more promising.

**Treatment strategies targeting histone methylation.** The selective inhibitor, 3-deazaneplanocin A (DZNep) prevents methylation at lysine 27 on histone H3 (H3K27me3) and lysine 20 on histone H4 (H4K20me3) is currently used in the treatment of cancer (54). Upregulation of Lysine methyltransferases (KMTs) plays an important role in the differentiation of monocytes into immature dendritic cells (iDCs). Two of the
know drugs that inhibit histone methylation, BIX-01294 which is KMT1c inhibitor, and DZNep, prevents global KMT activity were found to inhibit monocytes differentiating into iDCs (55).

**Treatment strategies targeting histone acetylation.** HDAC1 expression was identified to decrease acetylation of H3K9ac which induces accumulation of total cholesterol, free cholesterol, and triglycerides in foam cells, as seen in the aorta of ApoE−/−mice, which was abrogated by upregulation of the microRNA miR-34a (56). The compound 4-hydroperoxy-2-decenoc acid ethyl ester (4-HP-O-DAEE) isolated from the royal jelly was found to acetylate histone H3 and H4 at the proximal promoter region of EC-SOD (extracellular superoxide dismutase) in THP-1 cells, which is considered as a potential target for the treatment of atherosclerosis (57). Treatment with histone deacetylase inhibitor, valproic acid, was shown to promote the release of endogenous t-PA in males with vascular disease (58). Several HDAC inhibitors are used in the current standard of care. The known synthetic HDAC inhibitors which inhibit Class III HDACs include; dihydrocoumarin, naphthyroparanone, 2-hydroxynaphaldehydes and other sirtuin-inhibiting agents. Class I, II, and IV HDAC inhibitors were grouped as hydroxamates such as trichostatin A (TSA), vorinostat (suberoylanilide hydroxamic acid; SAHA), belinostat (PXD101), panobinostat (LBH589), LAQ824, peptide inhibitors such as cyclic tetrapeptides (trapoxin B) and depsipeptides, benzamides (entinostat (MS-275), mocetinostat (MGCD0103), C1994, aliphatic acid components (phenylbutyrate, valproic acid), and electrophilic ketones (51). Some of the plant polyphenols such as curcumin (diferuloylmethane) and resveratrol were identified as naturally available HDAC inhibitors.

**5. Future directions in targeting histone methylation and histone acetylation in the treatment of atherosclerosis**

Several histone modifications regulating the off-target drug effects have been reported which indicates their additional benefits. Both Hydralazine and Osalazine which are used to treat hypertension can be repurposed for treating neoplasia as their treatment can restore the functions of tumor suppressor genes. As a histone-deacetylase inhibitor, Romidepsin when used in endometriosis, efficiently induced apoptosis in the epithelial cells of endometrium. However, off-targets effects on histone modification can potentially exert negative effects in the cells. Similarly, the histone deacetylase inhibitor, valproic acid which is prescribed for epilepsy treatment, was identified as a causative factor for spina bifida in the growing embryos (59). Thus, for future considerations, specific mechanisms of histone modifications should be determined in patients for prognosis to avoid adverse drug effects.

**DNA methylation intercepts histone modifications.** During the early developmental stages of the embryo, unmethylated CpG islands were found to induce the expression of DNMT3A and DNMT3B. At the same time, DNMT3L interacts with the chromatin and targets H3K4. G9a was found to methylate H3K9 and recruit heterochromatin protein 1, DNMT3A and DNMT3B during stem cell production (60). Existence of such epigenetic cross connections may also lead to de novo gene regulations which can be critically involved in the disease progression.

**Non-coding RNA and histone modification.** Another key epigenetic mechanism that was found to intersect histone modifications is the expression of long non-coding RNAs. Under hypoxic conditions, upregulated expression of HDAC3 was reported to inhibit the expression of long non-coding RNA ‘lncRNA-LET’ (61). During the phenotype switch in VSMCs, the long non-coding RNA ‘taurine up-regulated gene-1’ (TUG1), was found to form a hetero-complex with EZH2 to methylate α-actin. As a result, the levels of α-actin were found decreased, and simultaneously F-actin was elevated. This imbalance was presumed to be a causative factor for the phenotype switch in the VSMCs which change from contractile to synthetic phenotype (62). In lung cancer, the long non-coding RNA, MALAT1, was reported to upregulate the expression of histone demethylase JMJDIA, which was known to promote cell migration and invasion (63).

**Hyperlipidemia and hypertension.** Several factors such as hyperlipidemia, hypertension, diabetes and also diet play an important role in the development and progression of atherosclerosis. The onset of de novo molecular mechanisms which are discussed above indicates epigenetic mechanisms to be critically involved in the pathogenesis. It was reported that high-fat diet decreases ATP citrate-lyase levels as well as acetyl-CoA and/or the ratio of acetyl-CoA:CoA in white adipose tissue and pancreas, leading to inhibition of H3K23 acetylation in mice fed with high-fat diet (64). Dyslipidemia can cause monocyte priming such as increase in the adhesion of monocytes. Chemotaxis in the monocytes was promoted due to reduction in the H3K27 acetylation in non-human primates (65). H3K4 methylation of lysine-specific demethylase-1 (LSD1), H3K6 methylation of dehydrogenase 2 (HSD11B2), K3K79 methylaion on epithelial sodium channel subunit α (SCNN1A) were also shown to be associated with arterial hypertension (66).

**Clinical significance of HDAC inhibitors for the treatment of atherosclerosis.** Panobinostat is a non-specific HDAC inhibitor that has been shown to have nanomolar potency, and can effectively inhibit Class I, II and IV HDACs. It induces apoptosis primarily by activating caspases and inducing PARP cleavage. In a phaseI/II clinical trial on 15 HIV positive patients, treatment with an HDAC inhibitor, panobinostat, was found to lower several inflammatory markers (67). Trichostatin A is a nonspecific inhibitor of Class I and Class II HDACs. Although it has been identified to prevent inflammatory markers such as IL-1β and IL-6, and have significant anti-tumor properties, it appears to have a more deleterious role in atherosclerosis. Treatment with Trichostatin A was found to promote proatherogenic markers such as TNF-α, SRA, CD36, eNOS and VCAM-1 (68). However, Trichostatin A was found to inhibit p21 mediated proliferation of VSMCs (69). These reports suggest that Trichostatin A may have a role in preventing neointimal hyperplasia or maintaining plaque stability. Trichostatin A treatment upregulates the expression...
of Arginase 2 enzyme which promotes eNOS expression, inflammation and cell proliferation during progression of atherosclerosis. Earlier, it was shown that HDAC2 directly binds to the Arginase 2 promoter and inhibits its expression. In the same study, treatment with the HDAC2 specific inhibitor ‘mocetinostat’, was shown to enhance oxLDL induced endothelial dysfunction (70).

Though certain HDAC inhibitors have shown promising results in treating cancer and other diseases, the same had a different role in atherosclerosis as identified above. However, there is no detailed information available yet on the clinical trials using the HDAC inhibitors for the treatment of atherosclerosis in humans. Additional studies emphasizing the specific role of each HDAC and their effects on different cell types during the progression of atherosclerosis are needed. The ultimate outcome of such studies should substantiate the need for extensive evaluation of HDAC inhibitors through clinical trials which are currently lacking.

Technological advancements and possible applications. Improvements in sequencing strategies, mass spectrometry, Chromatin immunoprecipitation, microarray and other novel techniques have contributed to substantial progress in health science research. Increasing applications of these technological advancements would have positive effects in the development of treatment strategies that target de novo epigenetic changes in different disease conditions including atherosclerosis. Time-resolved NMR spectroscopy was innovatively developed to characterize asymmetric histone PTMs (71). Chromatin immunoprecipitation in conjunction with sequencing methodologies are typically used for mapping histone modifications. Through inclusion of micrococcal nuclease (MNase) digestion and barcoding, disease specific signatures can be identified at the single-cell level. Moreover, single-cell DamID can also be performed for genome-wide analysis of histone modifications using a cell line expressing Escherichia coli deoxyadenosine methylase (Dam) and combining it with specific histone readers or modifiers (72).

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WJ wrote the manuscript and performed the literature review. CSB developed the format of the review and corrected the manuscript. DKA assisted with the scientific content of the review and edited the manuscript.

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Competing interests

The authors declare that they have no competing interests.

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