Association of increased serum glycated albumin levels with low coronary collateralization in type 2 diabetic patients with stable angina and chronic total occlusion

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Abstract

Background: We investigated whether serum glycated albumin (GA) levels are related to coronary collateralization in type 2 diabetic patients with chronic total occlusion.

Methods: Blood levels of GA and glycosylated hemoglobin (HbA1c) were determined in 317 diabetic and 117 non-diabetic patients with stable angina and angiographic total occlusion of at least one major coronary artery. The degree of collaterals supplying the distal aspect of a total occlusion from the contra-lateral vessel was graded as low (Rentrop score of 0 or 1) or high collateralization (Rentrop score of 2 or 3).

Results: For diabetic patients, GA (21.2 ± 6.5% vs. 18.7 ± 5.6%, P < 0.001) but not HbA1c levels (7.0 ± 1.1% vs. 6.8 ± 1.3%, P = 0.27) was significantly elevated in low collateralization than in high collateralization group, and correlated inversely with Rentrop score (Spearmen’s r = −0.28, P < 0.001; Spearmen’s r = −0.10, P = 0.09, respectively). There was a trend towards a larger area under the curve of GA compared with that of HbA1c for detecting the presence of low collateralization (0.64 vs. 0.58, P = 0.15). In non-diabetic patients, both GA and HbA1c levels did not significantly differ regardless the status of coronary collateralization. In multivariable analysis, female gender, age > 65 years, smoke, non-hypertension, duration of diabetes > 10 years, metabolic syndrome, eGFR < 90 ml/min/1.73 m², and GA > 18.3% were independently determinants for low collateralization in diabetic patients.

Conclusions: Increased GA levels in serum are associated with impaired collateral growth in type 2 diabetic patients with stable angina and chronic total occlusion.

Keywords: Glycated albumin, Coronary collateralization, Diabetes

Background

Coronary collateralization is a physiological adaptive response to transient or permanent occlusion of major coronary arteries [1,2]. Well-developed coronary collaterals contribute to a reduction of infarct size, preservation of left ventricular function, and an improvement of survival in patients with coronary artery disease [3-5]. However, such a physiological response is significantly impaired in patients with type 2 diabetes, metabolic syndrome, and severe atherosclerosis [6,7]. In a diabetic setting, an excessive formation of advanced glycation endproducts (AGEs) and reactive oxygen species (ROS) frequently causes a reduction of endothelium-mediated dilation, while blockade of AGEs restores ischemia-induced angiogenesis [8,9].

Glycated albumin (GA) - a predominant early Amadori-type glycation protein in serum - serves as an alternative measure of dysglycemia over approximately 2–3 weeks, and is associated with the occurrence and severity of atherosclerosis in diabetes [10-14]. In cellular experiment, GA promotes inflammation via activation of NF-κB pathway in endothelial cells, induces adhesion of monocytes

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to endothelial cells through enhanced transcription of the cell surface adhesion molecules, and stimulates vascular smooth muscle cell proliferation [10]. It is also a prognostic marker for diabetic patients undergoing coronary artery stent implantation [14,15], and a risk factor of mortality and morbidity for those receiving hemodialysis [16,17]. GA could be also formed in a non-diabetic milieu triggered by inflammatory reaction [7,10]. Based upon these findings, it is reasonable to hypothesize that GA might exert a deleterious effect on coronary collateralization in diabetic patients. To verify this hypothesis, we analyzed serum levels of GA in diabetic and non-diabetic patients with stable angina and chronic coronary total occlusion. This angiographic criterion of inclusion was used because a severe coronary artery obstruction was a prerequisite for spontaneous collateral recruitment [18]. We assessed the degree of coronary collateralization according to the Rentrop grading system [19] as this method is easy to incorporate into the routine clinical practice. To the best of our knowledge, there is no previous report referring to the relation of Amadori-modified proteins and collateral formation in such a unique cohort.

Methods

Study population

A total of 537 consecutive patients with stable angina and chronic total occlusion (> 3 months) of at least one major epicardial coronary artery between January 2009 and July 2013 from the database of Shanghai Rui Jin Hospital Percutaneous Coronary Intervention (PCI) Outcomes Program were screened. Stable angina was diagnosed according to the criteria recommended by the American College of Cardiology/ American Heart Association [20]. The duration of coronary artery occlusion was estimated from the date of occurrence of myocardial infarction in the area of myocardium supplied by the occluded vessel, from an abrupt worsening of existing angina pectoris, or from information obtained from a previous angiogram. The diagnosis of type 2 diabetes mellitus and dyslipidemia were made according to the criteria of the American Diabetes Association and Third Report of The National Cholesterol Education Program (NCEP) [21,22]. Metabolic syndrome was defined by the updated 2005 National Cholesterol Education Program Adult Treatment Panel III (NCEP ATP III) [23]. We used body mass index (BMI) ≥ 25 kg/m² as the cut point of obesity according to the criteria of Asia-Oceania [24]. For the purpose of research, patients who received PCI within the prior 3 months (n = 41) or had a history of coronary artery bypass grafting (n = 38) were excluded. We also excluded those with chronic heart failure, pulmonary heart disease, malignant tumor or immune system disorders (n = 19). Patients with type 1 diabetes were excluded by measurement of peptide C level (n = 5). The remaining 434 patients (317 diabetics and 117 non-diabetics) were enrolled in this cross-sectional study.

The protocol was approved by the Hospital Ethics Committee and written informed consent was obtained from all patients. Since the study did not involve any intervention, it was not prospectively registered.

Coronary angiography

Coronary angiography was performed through the femoral or radial approach. All angiograms were reviewed by two experienced interventional cardiologists, according to lesion classification scheme of the American College of Cardiology/American Heart Association [25]. They were blinded to study protocol and biochemical measurements, and any difference in interpretation was resolved by a third reviewer. The presence and degree of collaterals supplying the distal aspect of a total coronary occlusion from the contra-lateral vessel were graded on a 4-point scale from 0 to 3 according to the Rentrop scoring system [18]: zero = no collateral vessels; 1 = thread-like, poorly opacified collaterals with faint visualization of the distal vessel; 2 = moderately opacified collateral channels; 3 = large, brightly filled collateral channels with immediate visualization of the entire distal vessel > 10 mm. Patients were then classified as low (Rentrop score of 0 and 1) and high (Rentrop score of 2 and 3) coronary collateralization, as in previous studies [26,27]. For those with more than one total coronary occlusion, the vessel with the highest collateral grade was chosen for analysis.

Biochemical measurement

Blood samples were collected at the day of angiography in patients after an overnight fasting. All samples were stored at −80 °C until analysis. Serum glucose, glycosylated hemoglobin (HbA1c), blood urea nitrogen, creatinine, uric acid, and lipid profiles were measured with standard laboratory techniques on a Hitachi 912 Analyzer (Roche Diagnostics, Germany). Modified estimated glomerular filtration rate (eGFR) was calculated, as described previously [28]. Serum GA levels were determined with an improved bromocresolpurple method using Lucia™ glycated albumin-L assay kit (Asahi Kasei Pharma, Japan). The linear range for this assay was 3.2-68.1% and the maximum inter-assay coefficient of variation (CV) was < 3.0% [11,12,14].

Statistical analysis

Continuous variables are presented as mean and standard deviation (SD), and categorical data are summarized as frequencies or percentages. For categorical clinical variables, differences between groups were evaluated with the chi-square test followed by Bonferroni’s correction to account for multiple comparisons. For continuous variables, the existence of a normal distribution was evaluated
with the Kolmogorov–Smirnov test, and logarithmic or square-root transformations were performed on the continuous variables of non-normal distribution. Differences among groups were analyzed by one-way analysis of variance (ANOVA) or the Kruskal–Wallis analysis followed by post-hoc analysis. Correlation between variables was determined by the Spearman’s rho tests as appropriate. Receiver operating characteristic (ROC) curve was plotted to assess the power of GA and HbA1c for detecting low collateralization, and the area under the curve was compared using the DeLong method. The independent determinants for low collateralization were assessed by multivariate logistic regression analysis, and the covariates chosen to enter the multivariate analysis model included age, gender, BMI, risk factors for coronary artery disease, duration of diabetes, metabolic syndrome,

| Table 1 Baseline characteristics and biochemical assessment in patients with chronic total occlusion |
|-------------------------------------------------|-------------------------------------------------|---------------------------------|---------------------------------|
| **Collateralization** | **Non-diabetes** | **Diabetes** | **P value** | **Non-diabetes** | **Diabetes** | **P value** |
| Number of patients | n = 22 | n = 95 | < 0.001 | n = 118 | n = 199 | < 0.001 |
| Female, n (%) | 12 (54.5) | 12 (12.6) | | 41 (34.7) | 27 (13.6) | < 0.001 |
| Age, y | 68 ± 11.1 | 62.4 ± 10.4 | 0.03 | 66.5 ± 10.1 | 64.6 ± 11.1 | 0.12 |
| Age > 65y | 16 (72.7) | 33 (34.7) | 0.001 | 76 (64.4) | 80 (40.2) | < 0.001 |
| Body mass index, Kg/m² | 25.3 ± 2.8 | 24.8 ± 3.0 | 0.53 | 25.8 ± 3.4 | 25.1 ± 3.4 | 0.07 |
| Smoke, n (%) | 8 (36.4) | 47 (49.5) | 0.27 | 8 (39.0) | 71 (35.7) | 0.56 |
| Hypertension, n (%) | 10 (45.5) | 69 (72.6) | 0.01 | 77 (65.3) | 157 (78.9) | 0.008 |
| Dyslipidemia, n (%) | 15 (68.2) | 37 (38.9) | 0.01 | 67 (56.8) | 81 (40.7) | 0.006 |
| Systolic blood pressure, mm Hg | 142.3 ± 18.3 | 139.6 ± 17.0 | 0.51 | 140.5 ± 18.7 | 139.9 ± 17.1 | 0.76 |
| Diastolic blood pressure, mm Hg | 86.4 ± 13.6 | 84.4 ± 8.1 | 0.53 | 84.1 ± 10.4 | 84.3 ± 9.1 | 0.86 |
| Duration of diabetes, y | / | / | / | 8.8 ± 4.6 | 5.7 ± 3.4 | < 0.001 |
| Fasting blood glucose, mmol/L | 5.4 ± 0.7 | 5.0 ± 0.7 | 0.03 | 5.9 ± 1.8 | 5.5 ± 1.5 | 0.049 |
| HbA1c,% | 6.3 ± 0.3 | 6.3 ± 0.5 | 0.86 | 7.0 ± 1.1 | 6.8 ± 1.3 | 0.27 |
| Glycated albumin,% | 19.1 ± 4.6 | 18.4 ± 5.2 | 0.54 | 21.2 ± 6.5 | 18.7 ± 5.6 | < 0.001 |
| Triglyceride, mmol/L | 1.8 ± 0.8 | 1.7 ± 0.8 | 0.49 | 1.9 ± 1.1 | 1.8 ± 1.2 | 0.36 |
| Total cholesterol, mmol/L | 3.9 ± 1.1 | 4.4 ± 1.1 | 0.08 | 4.5 ± 1.5 | 4.1 ± 1.2 | 0.009 |
| HDL-C, mmol/L | 0.9 ± 0.2 | 1.0 ± 0.3 | 0.10 | 1.0 ± 0.3 | 1.1 ± 0.3 | 0.10 |
| LDL-C, mmol/L | 2.4 ± 1.0 | 2.7 ± 0.9 | 0.15 | 2.8 ± 1.2 | 2.5 ± 1.0 | 0.045 |
| Metabolic syndrome, n (%) | 12 (54.5) | 42 (44.2) | 0.38 | 106 (89.8) | 148 (74.4) | 0.001 |
| Blood urea nitrogen, mmol/L | 5.1 ± 1.7 | 5.0 ± 2.0 | 0.82 | 5.3 ± 2.3 | 5.1 ± 1.6 | 0.40 |
| Uric acid, mmol/L | 354.5 ± 59.4 | 337.0 ± 83.7 | 0.36 | 353.3 ± 84.6 | 329.7 ± 77.4 | 0.01 |
| Creatinine, µmol/L | 85.2 ± 17.1 | 76.4 ± 22.3 | 0.08 | 89.7 ± 22.5 | 76.8 ± 23.7 | < 0.001 |
| eGFR, mL/min/1.73 m² | 809 ± 24.3 | 1043 ± 26.3 | < 0.001 | 805.6 ± 21.5 | 1024 ± 24.1 | < 0.001 |
| Number of significant CAD | 0.25 | | | 0.30 | | |
| 1-vessel | 8 (36.4) | 19 (20.0) | 30 (25.4) | 38 (19.1) | | |
| 2-vessel | 9 (40.9) | 46 (48.4) | 40 (33.9) | 82 (41.2) | | |
| 3-vessel | 5 (22.7) | 30 (31.6) | 48 (40.7) | 79 (39.7) | | |
| Medication, n (%) | | | | | | |
| ACE inhibitors | 7 (31.8) | 33 (34.7) | 0.80 | 63 (53.4) | 106 (53.3) | 0.98 |
| β-blockers | 11 (50.0) | 28 (29.5) | 0.07 | 44 (37.3) | 78 (39.2) | 0.74 |
| Calcium channel blockers | 7 (31.8) | 20 (21.1) | 0.28 | 32 (27.1) | 51 (25.6) | 0.77 |
| Nitrates | 5 (22.7) | 35 (36.8) | 0.21 | 69 (58.5) | 127 (63.8) | 0.34 |
| Statins* | 18 (81.2) | 82 (86.3) | 0.89 | 98 (83.7) | 166 (83.3) | 0.09 |
| Anti-diabetic treatment | 0 (0) | 0 (0) | / | 92 (78.0) | 170 (85.4) | 0.09 |

Data are mean ± SD or number (%). Abbreviations: ACE, angiotensin-converting enzyme; eGFR, estimated glomerular filtration rate; HbA1c, glycated hemoglobin A1c; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol *Statins: mainly simvastatin, pravastatin and atorvastatin.
renal impairment, multivessel disease, HbA1c, and GA measurements. All analyses used 2-sided tests with an overall significance level of alpha = 0.05, and were performed with the SPSS 15.0 for Windows (SPSS, Inc., Chicago, IL, USA).

Results
Baseline characteristics and bio-measurements
Baseline demographic and clinical characteristics and biochemical measurements are listed in Table 1. Both diabetic and non-diabetic patients with low collateralization were older and less hypertensive, but were females in higher percentage and had higher fasting glucose and more dyslipidemia, and renal impairment than those with high collateralization (for all comparison, \( P < 0.05 \)). Metabolic syndrome was more common, and serum levels of total cholesterol, low-density lipoprotein cholesterol, and uric acid were significantly elevated in diabetic patients with low collateralization.

Serum GA levels and coronary collateralization
GA levels (21.2 ± 6.5% vs. 18.7 ± 5.6%, \( P < 0.001 \)) but not HbA1c (7.0 ± 1.1% vs. 6.8 ± 1.3%, \( P = 0.27 \)) were consistently higher in diabetic patients with low collateralization as compared with those who had high collateralization (Table 1). This finding was persistent even after having the group age-matched (\( P = 0.004 \) and \( P = 0.11 \), respectively). Further statistics showed that subgroups stratified by Rentrop score exhibited significant difference in GA levels for diabetic patients (Figure 1), and GA but not HbA1c correlated inversely with the degree of collateralization categorized by Rentrop score after adjusting for age, gender, BMI, risk factors for coronary artery disease (including smoke, hypertension and dyslipidemia), multivessel disease, and eGFR (Spearman’s \( r = -0.28, P < 0.001; \) Spearmen’s \( r = -0.10, P = 0.09, \) respectively). In contrast, for non-diabetic counterparts, both GA and HbA1c levels did not significantly differ regardless the status of coronary collateralization and after having the group age-matched (\( P = 0.96 \) and \( P = 0.80 \), respectively).

ROC curve analysis showed that there was a trend towards a larger area under the curve of GA compared with that of HbA1c (0.64 vs. 0.58, \( P = 0.15 \)), and the cutoff of GA > 18.3% was more sensitive than HbA1c > 7.0% for detecting the presence of low collateralization in diabetic patients (67% vs. 47%, Figure 2).

Multivariable analysis
Multivariate logistic regression analysis revealed that in non-diabetic patients, female gender (\( P = 0.01 \)), age > 65 years (\( P = 0.04 \)), non-hypertension (\( P = 0.02 \)), dyslipidemia (\( P = 0.02 \)), and eGFR < 90 mL/min/1.73 m² (\( P = 0.04 \)) were independently associated with low collateralization. In diabetic patients, after adjusting for possible confounding factors including age, gender, BMI, risk factors for coronary artery disease, duration of diabetes, metabolic syndrome, impaired renal function, and multivessel disease, GA > 18.3% but not HbA1c > 7% remained an independent determinant for low collateralization (Figure 3).

Discussion
Coronary collateral circulation is an important prognostic marker of ischemic heart disease [5]. This study is the first to demonstrate that increased serum GA levels are inversely associated with the degree of coronary collateral formation in diabetic patients with stable angina and

![Figure 1](image1.png)

**Figure 1** Relationship between serum levels of glycated albumin and collateral flow grade in diabetic patients.

![Figure 2](image2.png)

**Figure 2** ROC curve for glycated albumin (GA) and glycated hemoglobin (HbA1c) in detecting the presence of low collateralization in diabetic patients.
chronic total occlusion, adding novel information on pathophysiology of impaired collateral growth in diabetes. Accumulating evidence suggests that diabetes may exert a deleterious effect on several components necessary for collateral development, including pro-angiogenic growth factors, endothelial function, redox state of the coronary circulation, intracellular signaling, leukocytes and bone marrow-derived progenitor cells [6]. Previous studies have shown that GA induces endothelial dysfunction, and produces pro-inflammatory effects in macrophages through ROS augmentation [6,29-32]. Moreover, this glycated protein reflecting poor glycemic control impairs angiogenic function and elicits apoptosis of endothelial progenitor cells (EPCs) [33-37]. Thus, these observations support a notion that GA may be either a risk factor or a marker for poor collateral growth in diabetes.

In the present study, at least mildly impaired renal function (eGFR < 90 ml/min/1.73 m²) was found to be a strong independent risk factor for low collateralization in patients with stable angina and chronic total occlusion, particularly in those with diabetes. This finding may be partly related to a high occurrence rate of chronic kidney disease in diabetic and non-diabetic patients with coronary artery disease, which is supported by prior reports [26,38,39]. The direct and indirect effects of uremic toxins cause ROS-mediated endothelial dysfunction and inhibit proliferation of EPCs [40], combined with pathophysiology of diabetes including the effect of GA, eventually lead to reduced collateral growth in patients with chronic kidney disease.

Notably, the present study showed that GA but not HbA1c levels were higher in diabetic patients with low coronary collateralization, and that GA levels were more sensitive for detecting low coronary collateralization as compared with HbA1c in diabetic patients. These results are consistent with previous observations including ours, indicating that GA is superior to HbA1c in evaluating vasculopathies in diabetic patients with certain specific disorders such as coronary artery disease and renal dysfunction [41].

**Limitations**

We recognize that there are several limitations in our study. First, the study is cross-sectional for the point of coronary collateral investigation, thereby allowing us to detect association, not to predict outcome. Second, we evaluated the presence and degree of collaterals according to the Rentrop scoring system. Coronary collaterals may be more accurately assessed by collateral flow index with simultaneous measurement of aortic pressure and the distal pressure within the occluded segment of the culprit coronary artery [1]. Nevertheless, angiographic assessment of coronary collaterals is easy to incorporate into the routine clinical practice [19,26]. Third, elevated serum GA levels and low coronary collateralization may be also caused by the presence of metabolic syndrome and not diabetes alone. Finally, further large-scale studies with molecular experiments are required to clarify the mechanisms of GA effect on coronary collateralization.

**Conclusions**

The present study has demonstrated an association between increased GA levels in serum and reduced coronary collaterals in diabetic patients with stable angina and chronic total occlusion. These findings might offer a rationale in an effort to improve clinical outcomes in this unique diabetic cohort with aggressive intervention of AGEs and inflammatory process.

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Abbreviations
AGES: Advanced glycation endproducts; ANOVA: Analysis of variance; CV: Coefficient of variation; eGFR: estimated glomerular filtration rate; EPCs: Endothelial progenitor cells; GA: Glycated albumin; HbA1c: Glycosylated hemoglobin; PCI: Percutaneous coronary intervention; ROC: Receiver operating characteristic; ROS: Reactive oxygen species; SD: Standard deviation.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
YS performed the study. WF designed the studies and drafted the manuscript. LL and ZS collected the data. FHD did statistical analysis. RYJ, QZ, ZKY, JH performed angiography and imaging assessment. QJC made biochemical analysis. All authors have read and approved the final manuscript.

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