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Reference

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In vivo absolute quantification of striatal and extrastriatal D$_{2/3}$ receptors with $[^{123}I]$epidepride SPECT

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Abstract

**Background:** $[^{123}I]$epidepride is a high-affinity radiotracer used in single-photon emission computed tomography (SPECT) imaging of the D$_{2/3}$ receptors. It binds with high affinity to striatal and extrastriatal receptors. Nevertheless, its slow kinetics in the striatum impedes quantification in this region. Thus, an approach that would allow a simultaneous quantification of both striatal and extrastriatal D$_{2/3}$ receptors would be of interest for preclinical and clinical SPECT neuroimaging. We describe a partial saturation protocol that allows us to produce an in vivo Scatchard plot and thus estimate $B_{\text{avail}}$ and $K_{\text{app}}$ separately in both striatal and extrastriatal regions, through a single dynamic SPECT session. To validate this approach, a multi-injection protocol is used for the full kinetic modeling of $[^{123}I]$epidepride using a two-tissue compartment, 5-parameter model (2T-5k).

**Methods:** Eighteen male rats were used. Binding parameters were estimated using the multi-injection protocol. Various simulations were performed to estimate the optimal conditions for the partial saturation protocol, which was applied at the region and voxel level. The results of the partial saturation study were compared to those obtained with the 2T-5k model. To illustrate the interest of the partial saturation approach, we performed a preliminary study of the effect of a chronic, subcutaneous administration of haloperidol (1 mg/kg/day), a D$_2$ receptor antagonist, on the $B_{\text{avail}}$ of $[^{123}I]$epidepride in the rat striatum.

**Results:** A series of simulations demonstrated that a mass of 3 ug/kg of unlabeled epidepride allows the formation of an in vivo Scatchard plot. The partial saturation study led to robust estimations of $B_{\text{avail}}$ in all brain regions that highly correlated ($r = 0.99$) with the corresponding values from the multi-injection study. A chronic haloperidol treatment resulted in a 17.9% increase in the $B_{\text{avail}}$ values in the left Caudate Putamen nucleus (CP) ($p = 0.07$) and a 13.8% increase in the right CP ($p = 0.12$).

**Conclusion:** A partial saturation method allowed the robust quantification of D$_{2/3}$ receptors in striatal and extrastriatal D$_{2/3}$ receptors with a single-scan approach. This approach may be applied in the mapping of the D$_{2/3}$ receptor in translational biological studies and potentially, in clinical SPECT imaging.

**Keywords:** $[^{123}I]$epidepride, D$_{2/3}$ receptor, Dopamine, Molecular imaging, SPECT, Rat

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**Background**

Molecular imaging of the dopaminergic system with positron emission tomography (PET) and single-photon emission tomography (SPECT) is a powerful tool for the non-invasive study of the living brain in clinical and translational settings. Molecular imaging of the D2/3 receptor in particular has provided insight into the pathophysiology of a wide spectrum of neuropsychiatric disorders, ranging from mood and psychotic disorders to addiction and neurodegeneration [1, 2]. Imaging of the D2/3 receptor provides information on the receptor density and, after appropriate experimental manipulation, such as the administration of amphetamine or the performance of a neurocognitive test, its interaction with the endogenous ligand, thus allowing an indirect measure of the endogenous dopamine availability [3].

Several D2/3-binding radiotracers have been developed and routinely used in research settings. Among them, [11C]raclopride and [123I]IBZM are perhaps the most widely employed in PET and SPECT imaging of the dopaminergic system, respectively. These radioligands have the disadvantage of a relatively low affinity for the D2/3 receptor, limiting their use to imaging of the striatal receptors with a relatively low signal-to-noise binding ratio [4, 5]. For this reason, high-affinity radiotracers have been developed, allowing extrastriatal receptor imaging [5]. Nevertheless, the high affinity of the radiotracer is a serious impediment to the quantification of striatal D2/3 receptors. In this region, the kinetics of these radiotracers is particularly slow, meaning that a quantitative approach with standard pharmacokinetic modeling would require an impractical scan duration, both for preclinical and, especially, for clinical imaging [6].

One prominent example of such radiotracers is [123I]epidepride [7, 8]. Because of its very high affinity for the D2/3 receptor, [123I]epidepride is only used for imaging of extrastriatal receptors in human SPECT imaging [9–18]. However, a quantification approach allowing the use of [123I]epidepride SPECT for whole-brain imaging would be highly advantageous: on the one hand, its high affinity and thus the high signal-to-noise ratio allow a better sensitivity for the quantification of biological changes compared to lower-affinity radiotracers. On the other hand, compared to PET imaging, SPECT has the advantage of being more accessible to clinical or experimental use, given that it requires no in-house cyclotron (as is the case for PET imaging). 123I has a considerably longer half-life (~13 h) than positron-emitting radioisotopes, which is a crucial feature for these high-affinity radiotracers, for which imaging over sufficiently long periods is required to accurately quantify receptor binding. In addition, the development of novel SPECT cameras, in the preclinical and very recently in the clinical level, has led to SPECT imaging with a high spatial resolution, comparable to that PET [19]. Interestingly, the use of 124I, a positron-emitting radioisotope has been employed to label epidepride, allowing the use of this radiotracer in PET and providing another argument for the development of epidepride imaging for both SPECT and PET [20].

In this context, a methodology that would allow to quantify [123/124]Iepidepride in both the striatum and the extrastriatal regions would be of great interest. Given that the main obstacle in [123I]epidepride imaging is its very slow kinetics in the striatum, we proposed to employ the partial saturation approach, which, in principle, implies an alteration of the radiotracer’s kinetics. The partial saturation approach has been originally developed for [11C]flumazenil imaging of GABA_A receptors [21, 22]. In this approach, a dose of an unlabeled ligand is co-injected with the labeled radiotracer. The resulting kinetics of the radiotracer allow an in vivo Scatchard plot to be formed and the B_{avail} and appK_d to be separately estimated. This separate estimation of B_{avail} and appK_d provides an important advantage: the dissociation of the quantification of the concentration of the receptor from the estimation of appK_d potentially removes a confounding effect. Indeed, appK_d is directly related to the affinity of the radiotracer for the receptor (1/appK_d) and may vary as a result of variations in the endogenous ligand of the receptor (i.e., dopamine). This protocol has been recently employed by our group [23] and others [24, 25] for D2/3 imaging using [123I]IBZM and [11C]raclopride in small animals. Overall, this approach has been used to quantify radiotracers with rapid kinetics and this work presents that first application for a radiotracer with slow kinetics, such as [123I]epidepride.

In this paper, we firstly perform a full quantification of [123I]epidepride kinetics using a multi-injection imaging protocol [23, 26–28], which separately identifies B_{avail} and appK_d. The results of the multi-injection protocol serve as a “gold-standard” for the validation of the partial saturation approach, which is applied at the region- and at the voxel-level.

**Methods**

**Animals and general SPECT scan protocol**

18 male Mdr1a (P-glycoprotein) knock-out (KO) rats [29], weighing between 380 and 500 g, were employed in the study. Using this rat strain allowed us to study the kinetics of [123I]epidepride independently of the potentially confounding effect of the P-glycoprotein [30–34]. Of these, 3 rats were employed in an in vivo multi-injection SPECT imaging protocol for absolute D2/3 receptor quantification (SPECT-MI in Table 1). Four rats were employed in an arterial plasma analysis for the study of plasma kinetics of the radiotracer and the estimation of the free parent radiotracer fraction (TLC in
Table 1) [23]. Four rats were employed in a SPECT experiment with a partial D2/3 receptor saturation design for the determination of Bavail and appKd parameters from an in vivo Scatchard plot (SPECT-PSA in Table 1) [23]. Finally, 7 rats were employed in a preliminary study of the effect of a chronic haloperidol treatment on the Bavail and appKd of the D2/3 receptors in rats (SPECT-PSA_HAL for haloperidol-treated and SPECT-PSA_CON for control rats in Table 1, see Additional file 1).

Table 1 Numerical values of the SPECT and metabolite protocol parameters corresponding to the 18 experiments

| Rats         | Modality/tracer       | Duration (min) | Injection 1 (T = 0 min) | Injection 2 | Injection 3 | Time (min) | J1 (μg) | J2 (μg) | Time (min) | J3 (μg) |
|--------------|-----------------------|----------------|-------------------------|-------------|-------------|------------|---------|---------|------------|---------|
| 1            | TLC/[123I]epidepride  | 180            | > 1000                  | 89.6/< 0.05 | –           | –          | –       | –       | –          | –       |
| 2            | TLC/[123I]epidepride  | 180            | > 1000                  | 98.2/< 0.05 | –           | –          | –       | –       | –          | –       |
| 3            | TLC/[123I]epidepride  | 180            | > 1000                  | 96.9/< 0.05 | –           | –          | –       | –       | –          | –       |
| 4            | TLC/[123I]epidepride  | 180            | > 1000                  | 77.4/< 0.05 | –           | –          | –       | –       | –          | –       |
| 5            | SPECT-MI/[123I]epidepride | 360       | 1065.86                | 95.4/0.043  | 180         | 103.4/0.047 | 1.07    | 240     | 214.00     |
| 6            | SPECT-MI/[123I]epidepride | 360       | 967.71                 | 87.4/0.043  | 180         | 93.5/0.046  | 2.35    | 240     | 234.50     |
| 7            | SPECT-MI/[123I]epidepride | 360       | 955.45                 | 87.6/0.044  | 180         | 97.5/0.049  | 1.10    | 240     | 221.00     |
| 8            | SPECT-PSA/[123I]epidepride | 180       | 19.50                  | 52.7/1.30   | –           | –          | –       | –       | –          | –       |
| 9            | SPECT-PSA/[123I]epidepride | 180       | 23.74                  | 58.6/1.19   | –           | –          | –       | –       | –          | –       |
| 10           | SPECT-PSA/[123I]epidepride | 180       | 40.67                  | 113.06/1.34 | –           | –          | –       | –       | –          | –       |
| 11           | SPECT-PSA/[123I]epidepride | 180       | 29.51                  | 75.24/1.23  | –           | –          | –       | –       | –          | –       |
| 12           | SPECT-PSA_CON/[123I]epidepride | 180       | 18.85                  | 49.35/1.26  | –           | –          | –       | –       | –          | –       |
| 13           | SPECT-PSA_CON/[123I]epidepride | 180       | 16.98                  | 43.48/1.23  | –           | –          | –       | –       | –          | –       |
| 14           | SPECT-PSA_CON/[123I]epidepride | 180       | 22.74                  | 45.65/0.97  | –           | –          | –       | –       | –          | –       |
| 15           | SPECT-PSA_CON/[123I]epidepride | 180       | 17.10                  | 41.15/1.16  | –           | –          | –       | –       | –          | –       |
| 16           | SPECT-PSA_HAL/[123I]epidepride | 180       | 18.81                  | 46.09/1.18  | –           | –          | –       | –       | –          | –       |
| 17           | SPECT-PSA_HAL/[123I]epidepride | 180       | 16.72                  | 40.7/1.17   | –           | –          | –       | –       | –          | –       |
| 18           | SPECT-PSA_HAL/[123I]epidepride | 180       | 16.95                  | 38.47/1.09  | –           | –          | –       | –       | –          | –       |

Spect scans were performed with a U-SPECT-II camera (MiLabs, Utrecht, Netherlands). In rats that underwent SPECT scans with the multi-injection protocol, two polyethylene catheters (i.d. = 0.58 mm, o.d. = 0.96 mm) were inserted in the left femoral vein and artery for radiotracer administration and blood sampling, respectively. On the other hand, in rats that underwent SPECT scans for the partial saturation protocol, radiotracer injection was performed via a tail vein catheter. SPECT scans were performed under isoflurane anesthesia (3% for induction and 1–2% for maintenance). Body temperature was monitored during the scans and maintained at 37 ± 1 °C by means of a thermostatically controlled heating blanket.

SPECT image reconstruction was performed using a pixel-ordered subset expectation maximization (POSEM, 0.4-mm voxel size, 4 iterations, 6 subsets) algorithm using MiLabs image reconstruction software. Radioactive decay correction was performed, while correction for attenuation or scatter was not. Following reconstruction, dynamic images from the partial saturation experiment were denoised with factor analysis (FA) using Pixies software (Apteryx, Issyles-Moulineaux, France) as previously described [23, 27, 35]. FA allows the decomposition of a dynamic signal into a few elementary components, termed factors [27, 28, 36]. In this study, 3 factors were retained and the rest of the signal was discarded as noise [23, 27]. SPECT images were processed with PMOD software v3.7 (PMOD Technologies Ltd, Zurich, Switzerland). Averaged images corresponding to the ten first frames of the acquisition were co-registered to a magnetic resonance imaging (MRI) template integrated in PMOD [37]. Transformation matrices were then applied to dynamic images. Tissue-activity curves (TACs) from the following regions were extracted: caudate-putamen (CP), nucleus accumbens (NAc), ventral tegmental area (VTA), frontal cortex (FC), amygdala (Amy), hypothalamus (Hyp), superior colliculus (SupC), inferior colliculus (InfC), and cerebellum (Cer).

All experimental procedures were approved by the Ethical Committee on Animal Experimentation of the
Canton of Geneva, Switzerland. All experimental data, without exception, is available upon request to any of the authors of the manuscript.

Radiotracer preparation

[^123I]epidepride was prepared as previously described [7], using a commercially available precursor (ABX, Radenberg, Germany). Radiochemical purity was > 99%. The specific activities in the experiments are shown in Table 1.

SPECT multi-injection imaging protocol and quantification, arterial plasma analysis, and free parent radiotracer fraction estimation

A multi-injection protocol for full kinetic modeling of [^123I]epidepride was employed [23, 28, 38]. The scan protocol began with a first injection of the radiotracer at a high-specific activity, followed by a second co-injection of [^123I]epidepride and the unlabeled compound at 180 min and a third injection of the unlabeled compound alone at 240 min. The overall scan protocol included 360 1-min frames (Fig. 1). The specific activities and radioactive doses of each experiment are presented in Table 1.

The whole multi-injection study TAC data was fitted in with a two-tissue compartment five-parameter model (2T-5k), to estimate $K_{1}$, $k_{2}$, $k_{off}$, $k_{on}$ and $B_{avail}$ as previously described [38–41]. The free, non-metabolized radiotracer fraction in the plasma was used as the input function [42]. During dynamic SPECT acquisitions, 40 arterial blood samples (of 25 μl each) were withdrawn after each radiotracer injection at regular time intervals. Radioactivity was measured in a gamma counting system and expressed in kBq/ml after calibration. To estimate the plasma input function in in vivo SPECT experiments, only whole-blood radioactivity was measured individually and the free, non-metabolized radiotracer fraction was fitted using a model estimated in an independent experiment. Indeed, metabolite correction and plasma protein binding analysis for the estimation of radiotracer plasma input function was performed in an independent group of 4 rats, as previously described [27, 28, 43–46] (the radioactive doses employed in this experiment are presented in Table 1). The analysis was performed in MATLAB software R2019 (Mathworks, USA).

Simulations and in vivo partial saturation study

The partial saturation protocol

A partial saturation protocol was employed to estimate $B_{avail}$ and $appK_{d}$ as recently described for another D₂/₃ radiotracer [23] based on the original description by Delforge et al. [21, 22], as optimized by Wimberley et al. [24, 25]. Briefly, co-injection of a dose of the unlabeled compound at doses that occupy 50–70% of the receptors allows to form a Scatchard plot when the specifically bound fraction of radiotracer ($C_{S}$) is plotted against the ratio of $C_{S}$ to the non-displaceable binding at the same region ($C_{S}/C_{ND}$). The intercept to the $C_{S}$ axis provides the $B_{avail}$ and the inverse of the slope the $appK_{d}$.

Simulation study 1: estimation of the dose of the unlabeled epidepride for the partial saturation study

A first simulation study was performed to estimate the dose of the unlabeled compound that is needed to produce an in vivo Scatchard plot. This simulation study was based on the results of the full quantification from one of the multi-injection experiments. This full-quantification approach allows the complete discrimination of the different components of the radiotracer’s kinetics, namely the free and the specific binding in each VOI. As a consequence, in simulation study 1, single injection experiments using variable doses of unlabeled epidepride were simulated and in vivo Scatchard plots were delineated using the free and the specific binding from the CP (a high-binding) and VTA (a low-binding VOI). This simulation study is particularly important. Indeed, given the potential differences in the in vivo affinity of [^123I]epidepride for striatal and extrastriatal D₂/₃ receptors (detailed description of the $appK_{d}$ here [47]), as described for other high-affinity D₂/₃ radiotracers [48], the dose-occupancy curves could be different for these two sites. To be able to apply the partial saturation protocol in both striatal and extrastriatal brain region simultaneously, a dose of [^123I]epidepride which occupies an optimal percentage of the D₂/₃ receptors in both sites to produce an in vivo Scatchard plot is necessary. For this reason, $B_{avail}$ and $appK_{d}$ were estimated and compared to the results of the full quantification using experiments.

![Fig. 1 TACs extracted from the CP of one multi-injection dynamic SPECT scan, along with 2T-5k model fit and the decomposition of the radioactive signal into the specific binding and the free radioligand](image)
the multi-injection study, to estimate the percent bias in the parameter values as a function of the dose of unlabeled epidepride. The dose of the unlabeled epidepride that allows an optimal estimation of $B_{\text{avail}}$ and $\text{app}K_d$ using the partial saturation method in both striatal and extrastriatal regions was thus determined.

**Estimation of the $r$ parameter for the correction of the cerebellar TAC**

In the partial saturation approach, the cerebellar TAC is employed as an approximation of the non-displaceable binding in the target brain VOIs. Given the differences in the concentration of this binding between the Cer and the target VOIs, as well as the presence of a non-negligible specific binding in the Cer with $[^{123}]$epidepride, the cerebellar TAC has to be corrected. As originally proposed by Delforge et al. [21, 22] and more recently by Wimberley et al. [24], the concentration of the non-displaceable binding in a given brain VOI ($C_{\text{ND}}$) is approximated by the corrected concentration of the total (measured) cerebellar binding ($C_{\text{cer}}$) using an estimated $r$ parameter, so that $C_{\text{ND}} = r \times C_{\text{cer}}$.

The estimation of $r$ was based on simulation study 1 using the optimal concentration of the unlabeled compound estimated in that study. Radioactivity data corresponding to the time points between the 30th and the 180th minutes from the simulated TACs of the non-displaceable binding in (1) the CP ($C_{\text{ND}-\text{CP}}$), representing the high-binding, striatal VOIs; (2) the VTA ($C_{\text{ND}-\text{VTA}}$), representing the low-binding, extrastriatal VOIs; and (3) the total binding in the $C_{\text{cer}}$, the reference region were used. The $r$ parameter was determined using the ratio $r = C_{\text{ND}-\text{CP}}/C_{\text{cer}}$ for the striatal VOI and $r = C_{\text{ND}-\text{VTA}}/C_{\text{cer}}$ for extrastriatal VOIs.

**Simulation study 2: evaluation of the bias induced by the presence of specific binding in the Cer**

$[^{123}]$epidepride has a non-negligible specific binding in the Cer, a region which is used as an approximation of the non-displaceable binding in the whole brain (reference region). The presence of specific binding may induce a bias in the estimation of the quantitative parameters. To evaluate this phenomenon, we employed the results of the full quantification from one of the multi-injection experiments, as in simulation study 1. Various levels of specific binding in the Cer were simulated. A partial saturation experiment, using the optimal dose of unlabeled epidepride as determined in simulation study 1, was simulated. The results of the quantification of $B_{\text{avail}}$ and $\text{app}K_d$ were compared to the results of the full quantification of the multi-injection study and the percent bias in these parameters was estimated as a function of the specific binding in the Cer.

**In vivo partial saturation study**

In vivo partial saturation SPECT scans and image reconstruction were performed in the same conditions as described for the SPECT multi-injection experiments protocol. A single radiotracer injection (containing a dose of unlabeled epidepride determined in the simulation study) was followed by a scan composed of 180 frames of 1 min. No arterial blood sampling took place and the Cer was employed as the reference region. The radioactive doses of each experiment are presented in Table 1. TACs were processed in PMOD (PKIN module), in which the model for the analysis of data from partial saturation experiments, as optimized by Wimberley et al. [24, 25], is implemented.

**In vivo partial saturation study with a fixed $\text{app}K_d$ in extrastriatal regions**

Given the low binding of $[^{123}]$epidepride in extrastriatal regions, the estimation of both $B_{\text{avail}}$ and $\text{app}K_d$ may be suboptimal, suffering from high inter-subject variability. For this reason, we performed the same in vivo partial saturation protocol as described in the previous paragraph (“In vivo partial saturation study”) by fixing the $\text{app}K_d$ value in [22] and thus only fitting $B_{\text{avail}}$. This fixed $\text{app}K_d$ value was determined as the average $\text{app}K_d$ across the extrastriatal VOIs of the three multi-injection experiments.

**Simulation study 3: impact of fixing the $\text{app}K_d$ value on the estimation of $B_{\text{avail}}$**

The $\text{app}K_d$ parameter may be altered across experimental conditions, notably with respect to the concentration of the endogenous ligand in the vicinity of the receptor under study. As a consequence, if the fixed $\text{app}K_d$ value in the partial saturation experiments (as in the paragraph “In vivo partial saturation study with a fixed $\text{app}K_d$ in extrastriatal regions”) differs from the real $\text{app}K_d$ value, a bias in the estimation of $B_{\text{avail}}$ may be induced. To study this phenomenon, we performed a simulation study based on simulation study 1. Using the optimal dose of the unlabeled compound, as determined in that same study, a series of partial saturation experiments was performed, in which varying $\text{app}K_d$ values were simulated. In vivo Scatchard plots were delineated and $B_{\text{avail}}$ was estimated using the same fixed $\text{app}K_d$ value as determined in “In vivo partial saturation study with a fixed $\text{app}K_d$ in extrastriatal regions.” The percent bias in the $B_{\text{avail}}$ estimates, compared to the simulated $B_{\text{avail}}$ values, was calculated.

**Parametric images of $B_{\text{avail}}$ using $[^{123}]$epidepride**

$B_{\text{avail}}$ values may also be estimated with the partial saturation approach at the voxel level to produce parametric images [23]. Nevertheless, the partial saturation
quantification method that we employed in this study, included in the PKIN module for VOI-wise estimations, has not been included in the PXMOD module of the PMOD software yet, in which parametric estimations are performed. For this reason, to produce parametric images of B\text{avail}, we applied the partial saturation protocol on a coronal section including the CP and the NAc and another one including the VTA. TACs were extracted from each pixel and processed in PKIN, exactly as described in “In vivo partial saturation study with a fixed appK\text{d} in extrastriatal regions”. The resulting pixel-wise B\text{avail} values were used to create parametric images in MATLAB.

**Statistical Analysis**

B\text{avail} and appK\text{d} values resulting from fitting the data of the whole duration of the multi-injection protocol were used as the “gold standard” for comparison of estimations with the partial saturation approach by means of regression analysis. Comparisons of average B\text{avail} and appK\text{d} values from the multi-injection study and the partial saturation experiments were also performed by means of a two-sample t test.

**Results**

**Quantitative parameters from the multi-injection study**

As described in the “Methods” section describing the multi-injection study protocol, in each dynamic brain SPECT study, only whole-arterial blood radioactivity was measured. The free, non-metabolized radiotracer fraction was estimated by fitting average data from an independent group of four rats in an experiment that was independent from the SPECT scan experiments. The parameters of the bi-exponential function describing the kinetics of the mean percent non-metabolized plasma radiotracer were A1 = 0.61, B1 = 0.20, A2 = 0.27, B2 = 0.006. Average percentage of radiotracer bound to plasma proteins was 22% (f1 = 0.22 ± 0.06). The 2T-5k model provided satisfactory fits to the TACs of the multi-injection study, as shown in Fig. 1. Parameter estimates are provided in Table 2. A non-negligible amount of displaceable binding in Cer is observed (data not shown).

**Simulation studies**

A *simulation study to determine the dose of unlabeled epidepride that is required for the partial saturation protocol*

As shown in Fig. 2, a dose of unlabeled epidepride of 3 \(\mu\)g/kg or higher produces a Scatchard plot in both high- and low-binding VOI (Fig. 2a, b; Supplemental Fig. 1a, b). We chose to employ this dose of 3 \(\mu\)g/kg, as it produces a Scatchard plot with the highest range of C\text{S}/C\text{ND} values. In this case, the impact of noise in the in vivo experiments should be the lowest.

A *simulation study to determine the r value for the correction of the cerebellar TAC*

In addition, according to the simulation study (Fig. 3), the r value for the correction of the cerebellar TAC was found to be equal to 1 for CP and NAc and equal to 0.4 for the extrastriatal VOI and this value presents an adequate temporal stability in time points beyond 70 min after the injection of the radiotracer, i.e., the time points that are employed in the Scatchard plot. In addition, small variations in the mass of the unlabeled epidepride do not remarkably modify the r value.

A *simulation study to assess the impact of variations in the specific binding in the Cer on B\text{avail} and appK\text{d} values*

The results of the simulation study, in which the impact of varying specific binding levels in the Cer on B\text{avail} and appK\text{d} are evaluated, are presented in Fig. 4. The B\text{avail} and appK\text{d} are in pmol/ml, K1 in mL.cm\(^{-3}\).min\(^{-1}\), k2, k3, k off, in min\(^{-1}\).

| Multi-injection study | Partial saturation |
|-----------------------|-------------------|
| VOI | Multi-injection | Partial saturation |
| | B\text{avail} ± SD | appK\text{d} ± SD | k1 ± SD | k2 ± SD | k3 ± SD | k\text{on} ± SD | k\text{off} ± SD | B\text{avail} ± SD | appK\text{d} ± SD |
| NAc | 10.18 ± 1.72 | 0.15 ± 0.04 | 0.53 ± 0.14 | 0.10 ± 0.02 | 0.26 ± 0.06 | 0.04 ± 0.00 | 11.75 ± 2.84 | 0.27 ± 0.02 |
| CP | 24.22 ± 2.62 | 0.22 ± 0.06 | 0.69 ± 0.16 | 0.10 ± 0.04 | 0.21 ± 0.06 | 0.04 ± 0.01 | 21.25 ± 1.25 | 0.08 ± 0.03 |
| VTA | 3.77 ± 1.86 | 0.34 ± 0.21 | 0.57 ± 0.07 | 0.14 ± 0.04 | 0.28 ± 0.36 | 0.04 ± 0.02 | 4.01 ± 1.40 | – |
| FC | 20.67 ± 22.60 | 4.50 ± 5.25 | 0.40 ± 0.15 | 0.09 ± 0.03 | 0.01 ± 0.01 | 0.03 ± 0.01 | 5.15 ± 2.05 | – |
| Amy | 2.71 ± 1.53 | 0.36 ± 0.12 | 0.38 ± 0.11 | 0.16 ± 0.10 | 0.09 ± 0.02 | 0.03 ± 0.00 | 2.77 ± 0.72 | – |
| Hyp | 3.64 ± 1.10 | 0.37 ± 0.09 | 0.50 ± 0.21 | 0.13 ± 0.03 | 0.15 ± 0.03 | 0.05 ± 0.01 | 3.85 ± 0.90 | – |
| SupC | 3.86 ± 1.65 | 0.28 ± 0.18 | 0.70 ± 0.19 | 0.16 ± 0.04 | 0.26 ± 0.20 | 0.05 ± 0.02 | 3.97 ± 0.93 | – |
| InfC | 3.03 ± 0.40 | 0.18 ± 0.02 | 0.80 ± 0.26 | 0.17 ± 0.03 | 0.23 ± 0.02 | 0.04 ± 0.00 | 4.46 ± 0.45 | – |
| Cer | 2.02 ± 1.09 | 0.65 ± 0.27 | 0.66 ± 0.18 | 0.20 ± 0.02 | 0.04 ± 0.03 | 0.02 ± 0.01 | – | – |

B\text{avail} and appK\text{d} are in pmol/ml, K1 in mL.cm\(^{-3}\).min\(^{-1}\), k2, k3, k off, in min\(^{-1}\).

NAc nucleus accumbens, CP caudate-putamen, VTA ventral tegmental area, FC frontal cortex, Amy amygdala, Hyp hypothalamus, SupC superior colliculus, InfC inferior colliculus, Cer cerebellum.
values estimated with the partial saturation method from high-binding VOI (the CP) are minimally influenced by this variation in cerebellar specific binding. However, the $B_{\text{avail}}$ values from the low-binding VOI (such as the VTA in this simulation) and the $appK_d$ values from both high- and low-binding VOI may be biased, depending on the variation in the specific binding in Cer. Indeed, a variation of $\pm30\%$ in the specific binding in the Cer will lead to a $\pm20\%$ bias in the $appK_d$ in the CP, a $\pm18\%$ bias in the $B_{\text{avail}}$ in the VTA and a $\pm7\%$ bias in the $appK_d$ in the VTA.

**Assessing the impact of fixing $appK_d$ in extrastriatal $B_{\text{avail}}$ value estimations**

In the last simulation study, the impact of fixing $appK_d$ values in extrastriatal $B_{\text{avail}}$ value estimations was
evaluated. A fixed appK_d led to a percent bias in the estimation of the B_{av} value in the VTA that is almost linear as a function of the percent difference between the “true”-simulated and the fixed appK_d (Fig. 5). In detail, a fixed appK_d which is higher than the simulated value by 60% leads to an underestimation of the B_{av} value by 16%. On the contrary, a fixed appK_d which is lower than the simulated value by 60% leads to an overestimation of the B_{av} by 25%.

Quantitative parameters from the in vivo partial saturation study at the VOI and pixel level

B_{av} and appK_d values estimated from the in vivo partial saturation experiments are also provided in Table 2. These values are comparable and highly correlated to the corresponding values from the multi-injection experiments, as the linear regression analysis demonstrates (r = 0.99, p < 0.01 for B_{av} Fig. 6). In addition, no significant difference in the average B_{av} or appK_d values from the multi-injection and the partial saturation studies was found (p > 0.05). The parametric images of B_{av} in the section including the CP and VTA and in the section including the VTA are shown in Fig. 7.

Preliminary study of the effect of chronic haloperidol treatment on the striatal D_{2/3} B_{av} values

A chronic (28 days) haloperidol treatment (1 mg/kg/day), as described in Additional file 1, resulted in an increase in the B_{av} values (Fig. 8): 25.67 ± 2.30 pmol/ml in the haloperidol-treated group vs 21.76 ± 2.20 pmol/ml in the vehicle-treated group in the left CP (p = 0.07 using two-samples t test) and 25.33 ± 2.52 pmol/ml vs 22.25 ± 3.40 pmol/ml in the right CP (p = 0.12).

Discussion

Validation of B_{av} and appK_d values

In this study, we applied a partial saturation approach in nuclear neuroimaging for the separate quantification of D_{2/3} B_{av} and appK_d. The multi-injection approach, the “gold-standard” for the validation of the results of the partial saturation method was also applied in a group of rats. The results of the multi-injection experiment are in accordance with previous studies of absolute D_{2/3} receptor quantification from our group [23], using the same protocol: using [123I]IBZM and a multi-injection protocol, B_{av} values in the CP were found in the range between 19 and 20 pmol/ml. At the level of NAc, the B_{av} values were found in the range between 13 and 15 pmol/ml, slightly higher than those found in the present study. Considering the extrastriatal VOI, B_{av} values were roughly between 2 and 4 pmol/ml. In a previous study from our group using [18F]fallypride, B_{av} values were found in the range between 2 and 9 pmol/ml [28]. This difference may be explained by the different delineation of the VOI in the two studies. In the [18F]fallypride study, VOI were spherical structures placed in the middle of the corresponding anatomical structure of the brain, whereas in the present study the Schiffer atlas [37] was employed to delineate VOI that correspond to the entire size of the anatomical structure.

For the partial saturation study, we first estimated the r parameter that is necessary for the correction of the cerebellar TAC and its use as an index of the non-
specific binding of a given VOI in the partial saturation studies. The $r$ parameter was estimated using a simulation study in which the total PET signal was decomposed and the non-displaceable binding in striatal and extrastriatal VOI was extracted. The ratio of this non-displaceable binding of the target VOI to the total binding in the Cer gave the $r$ parameter. We did not use a pre-saturation in vivo study as described previously from our group [23] and others [24, 25]. This was impossible, because a pre-saturation study would not be able to take the specific binding in the Cer into account in the estimation of $r$. This is because in that case, the specific binding in this region would have been displaced. In addition, we did not estimate the $r$ parameter directly by the ratio of the non-displaceable distribution volume ratio the target VOI to the cerebellar VOI ($V_{ND\text{-target}}/V_{ND\text{-CER}}$) because, as shown in Fig. 3, this ratio varies in the first frames of the SPECT scan and it remains relatively stable at later time-points, i.e., those employed in the quantification using the Scatchard plot. As a result, a direct $r = V_{ND\text{-target}}/V_{ND\text{-CER}}$ estimate would potentially be biased by the first time-frames, which are not taken into account for the quantification of the $B_{\text{avail}}$ and app$K_d$. Taking the specific binding in the reference region into account has already been described for the application of the partial saturation approach with $[^{11}C]$flumazenil [22]. In this way, the correction of the TAC of the reference region more accurately represents the ratio of the non-specific binding in the target VOI.

Another finding considering the use of the Cer as a reference region in $[^{123}I]$epidepride imaging is illustrated in simulation study 2. The results of this study suggest that should the specific binding in the Cer vary, the $B_{\text{avail}}$ values in the low-binding, extrastriatal regions will vary accordingly. $B_{\text{avail}}$ values in high-binding regions remain virtually unbiased even with large variations of the specific binding in the Cer. Similarly, app$K_d$ estimations in both striatal and extrastriatal regions will vary considerably as a function of the variation in the specific binding in the Cer in a given experimental context (e.g., a brain pathology). $[^{123}I]$epidepride has been employed in multiple studies (discussed in more detail in the following section) of different psychiatric conditions, in which the cerebellar specific binding of the radiotracer is considered unchanged. As demonstrated by Pinborg et al. [49], this issue may be particularly problematic when extra-striatal occupancy of $D_{2/3}$ receptors by pharmaceutical agents is estimated. Possible variations in the specific binding in the Cer have to be taken into account in the design of biological studies using $[^{123}I]$epidepride, and the stability of this specific binding across the experimental conditions has to be verified in order to obtain unbiased results.

The partial saturation approach using $[^{123}I]$epidepride gave highly similar values with previous studies of the $D_{2/3}$ receptor using the same method [23, 24] in the striatum. All $B_{\text{avail}}$ values obtained with the partial saturation approach highly correlated with the corresponding $B_{\text{avail}}$ values from the multi-injection experiment. However, the app$K_d$ values obtained using the partial saturation method in the striatal VOI differ considerably from the corresponding values obtained using the multi-injection approach (Table 2). This is could possibly be
due to inter-individual variations in the specific binding in the reference region and in the overall correction of the cerebellar TAC with the $r$ parameter. As predicted in the simulation study (Fig. 4), a variation in the specific binding in the Cer induces a considerable bias in the appKd in such high-binding VOI, whereas the $B_{\text{avail}}$ values are minimally impacted and this is observed in the estimated parameter values here. Regarding the extrastriatal VOI, the appKd values were fixed in a value obtained from the multi-injection experiment, as attempting to fit both $B_{\text{avail}}$ and appKd led to highly variable estimations (data not shown). Fixing appKd in these VOI allowed the estimation of $B_{\text{avail}}$ with acceptable variability (Table 2). In this case, as the simulation study suggests (Fig. 5), if the “true” appKd varies, the $B_{\text{avail}}$ estimations vary accordingly. This means that, by fixing the appKd parameter value, the $B_{\text{avail}}$ becomes a composite parameter which integrates information on both the absolute concentration of the target protein (i.e., the $B_{\text{avail}}$ per se) and the affinity of the radiotracer for the target protein (i.e., $1$/appKd). In this case, in a molecular neuroimaging study involving [123I]epidepride, a variation in the $B_{\text{avail}}$ value in the extrastriatal VOI in a given experimental condition may be interpreted as either a true elevation of the quantity of the D2/3 Receptor or a modification of the affinity of the radiotracer for the receptor, which could be linked to alterations in the concentration of dopamine in the synapse. This is essentially similar to the information provided by the binding potential ($BP = B_{\text{avail}}$/appKd), a composite measure that is most widely employed in molecular neuroimaging [50].

**Potential applications of [123I]epidepride imaging using the partial saturation approach**

The method proposed here is the first approach that allows the quantification of the striatal binding of [123I]epidepride. The high affinity of this radiotracer allows to obtain excellent quality images. Figure 9 illustrates the superiority of the quality of a [123I]epidepride SPECT image compared to an image obtained with another SPECT D2/3 radiotracer, the low-affinity [123I]IBZM. The binding of [123I]epidepride is considerably higher than the binding of [123I]IBZM (note the difference in the range of radioactive concentrations, 0–10 KBq/ml and 0–2.5 KBq/ml, respectively). The high binding of [123I]epidepride also allows for an adequate anatomical delineation of the striatal substructures and notably the visual distinction of the NAc from the CP, which is not possible with [123I]IBZM.

The quality of the radioactive signal and the robustness of the quantitative parameters that are obtained with the partial saturation approach are supported by the low-standard deviations and the percent coefficients of variation (CV) of the $B_{\text{avail}}$ values (Table 2): indeed, the CV of these values obtained with [123I]epidepride in the CP is 5.87% and in the NAc is 24.14%. In the extrastriatal regions, CV values range from 10 to 35%. In comparison, the same approach with [123I]IBZM yielded
B\textsubscript{avai}l values with a CV of 25.43% in the CP and 58% in the NAc in rats [23]. Using the same approach and the low-affinity PET radiotracer \[^{11}\text{C}]\text{raclopride}, Wimberley et al. found B\textsubscript{avai}l values with a CV around 40% in mice (as roughly depicted in Fig. 8 of the aforementioned paper) [24]. In rat PET imaging with \[^{18}\text{F}]\text{fallypride} [28], CV of BP values estimated with the Logan graphical analysis approach (38.31% in CP and 30% in the NAc and from 38 to 58% in the extrastriatal VOI) were also higher than the CV of B\textsubscript{avai}l values presented here. Overall, the robustness of the quantitative parameters obtained with the partial saturation approach in \[^{123}\text{I}]\text{epidepride} SPECT suggests that biological studies using this radiotracer may provide high statistical power compared to other available PET and SPECT imaging modalities of the dopaminergic system.

To illustrate the potential usefulness of the partial saturation approach in biological studies of the dopaminergic system, we performed a preliminary study of the effect of a chronic, high-dose haloperidol treatment [51] on the absolute quantity of D\textsubscript{2/3} receptors in the striatum. We found an upregulation of the D\textsubscript{2/3} receptor in the CP, as suggested by previous studies in the literature [52]. Chronic haloperidol treatment led to an increase in the B\textsubscript{avai}l of \[^{123}\text{I}]\text{epidepride} of 17.97% in the left CP and of 13.84% in the right CP. Associated p values with respect to a comparison by means of a two-sample t test were 0.07 and 0.12, respectively, i.e., the differences are not statistically significant using a two-tailed threshold of p = 0.05. Indeed, the small sample size (4 rats in the control and 3 in the haloperidol treatment group) suggests that this study was underpowered. However, if a one-tailed significance threshold is considered, the difference in B\textsubscript{avai}l values in the left CP achieves statistical significance. Given the purely illustrative purpose of this experiment, we chose to maintain its preliminary character in the context of the present study. It is for the same reason that we only studied the effect of haloperidol in high-binding VOIs, in which the haloperidol-induced D\textsubscript{2/3} receptor upregulation has been established in the literature [51, 52], and not in extrastriatal VOIs. Another prominent experimental paradigm where \[^{123}\text{I}]\text{epidepride} imaging with the partial saturation protocol is the study of endogenous dopamine alterations in physiological and pathological conditions. It has been experimentally and theoretically established [53] that \[^{123}\text{I}]\text{epidepride} binding is not sensitive to "rapid" changes in endogenous dopamine binding in the D\textsubscript{2/3} receptor, as in the case of amphetamine-induced dopamine release. Nevertheless, using the partial saturation protocol described here, one may gain access to enduring alterations of baseline receptor occupancy by endogenous dopamine in experimental models and pathological conditions, such as in schizophrenia [54, 55].

The high affinity of the binding of \[^{123}\text{I}]\text{epidepride} allowed it to be employed in clinical studies of the extrastriatal D\textsubscript{2/3} receptors in various psychiatric conditions, such as in major depressive disorder [56] and schizophrenia [12, 57, 58]. Using the partial saturation approach presented here, striatal and extrastriatal D\textsubscript{2/3} receptor binding can be studied simultaneously, which was possible because of the pharmacological properties of \[^{123}\text{I}]\text{epidepride}, which allows an optimal percentage of receptors to be saturated in both striatal and extrastriatal regions at the same time, as opposed to other high-affinity D\textsubscript{2/3} radiotracers (e.g., \[^{11}\text{C}]\text{FLB457} [48]). The interest of dissociating B\textsubscript{avai}l and appK\textsubscript{d} estimations in PET studies has been discussed extensively in a previous study from our group [23]. The application of the partial saturation protocol in translational \[^{123}\text{I}]\text{epidepride} imaging may further potentiate the preclinical studies of the dopaminergic system, well beyond the mesostriatal circuit. The partial saturation protocol described here for preclinical imaging is potentially applicable in clinical PET studies of the dopaminergic system. Indeed, occupying 50–70% of striatal D\textsubscript{2/3} receptors for the application of the partial saturation approach induces little, if any, adverse effects. Indeed, this level of the blockade is identical to the one induced by antipsychotic agents at therapeutic doses and it is accepted that adverse events appear when the blockade is higher than 80–85% [59]. After the safety of administering pharmacological doses of epidepride is confirmed, the application of this protocol in clinical imaging will be feasible and its impact in the understanding of brain function and disease can be invaluable.

**Conclusions**

In conclusion, we propose here, for the first time, a method that allows a simultaneous quantification of striatal and extrastriatal D\textsubscript{2/3} receptors using \[^{123}\text{I}]\text{epidepride}. A cartography of the D\textsubscript{2/3} receptor in various brain regions is now feasible using this radiotracer. The robustness of the radioactive signal (especially compared to low-affinity D\textsubscript{2/3} radiotracers) and the stability of the resulting quantitative parameters, which highlight the potential of conducting studies with high statistical power, have been demonstrated. In addition, this approach is relatively simple to apply and already integrated into a commercial image-analysis software. Overall, this study highlights an innovative tool of preclinical brain SPECT imaging in the study of dopamine neurochemistry in physiology and animal models of disease with an approach that is validated for rat SPECT studies but is potentially applicable in clinical brain SPECT.
Supplementary information

Supplementary information accompanies this paper at https://doi.org/10.1186/s13550-020-00650-0.

Additional file 1. Supplemental Materials and Methods

Abbreviations

[^123I]IBZM: [123I]-Iodobenzamide; 2T-5k: Two-tissue compartment, 5 parameter model; Amy: Amygdala; appKd: Dissociation constant; Basal: Absolute concentration of the receptor which is accessible for binding of the radioligand in the tissue; BP: Binding potential; Ccer: Cerebellum; CNP: Non-displaceable binding; CV: Coefficient of variation; CP: Caudate-Putamen; CS: Specific binding; D2/3: Dopamine receptors 2 and 3; FA: Factor analysis; FC: Frontal cortex; GABAα: Gamma aminobutyric acid receptor A; HYP: Hypothalamus; InfC: Inferior colliculus; KO: Knock-out; MRI: Magnetic resonance imaging; NAc: Nucleus accumbens; PET: Positron emission tomography; P-OSEM: Pixel-based ordered-subset expectation maximization; SPECT: Single-photon emission computed tomography; SPECT- Mi: SPECT multi-injection protocol; SPECT-PSA: SPECT partial saturation approach; SPECT-PSA: CON: SPECT partial saturation approach, control group; SPECT-PSA_HAL: SPECT partial saturation approach, haloperidol-treated; SupC: Superior colliculus; TACs: Tissue-activity curves; VNR: Non-displaceable volume of distribution; VOI: Volume-of-interest; VTA: Ventral tegmental area

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Ethical approval and consent to participate

All experimental procedures were approved by the Ethical Committee on Animal Experimentation of the Canton of Geneva, Switzerland.

Authors’ contributions

PM conceptualized and supervised the study. ST, BBT, and PM designed it. ST, BBT, and PM performed the experiments and analyzed the data. ST, BBT, and PM prepared the manuscript. The authors read and approved the manuscript.

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Availability of data and materials

All experimental data, without exception, is available upon request to any of the authors of the manuscript.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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