INTRODUCTION

Flap necrosis is a physiologic consequence of prolonged ischemia in a surgical flap. Regardless of causes, partial or whole flap necrosis is regarded as a major complication and such events can result in wound dehiscence, infection, delayed healing, and fistula formation. These complications often prolong postoperative hospitalization and may require additional surgical management with less-than-optimal outcomes.

Background: Alprostadil and sildenafil are known vasodilators used independently to improve flap survival in animal models. In this study, we investigate whether these agents act synergistically to decrease flap necrosis in rat models.

Methods: After acclimation period, 4 groups of 10 male white rats were given a modified McFarlane skin flap. The postoperative treatment included saline control (Group A), sildenafil citrate-only (Group B), alprostadil-only (Group C), and both sildenafil and alprostadil (Group D). The flaps were observed on postoperative days 1, 3, 5 and 7. The animals were euthenized on postoperative day 7, and the flaps were evaluated for inflammation and neovascularization.

Results: At each observation, the mean necrotic index was significantly lower for all three treatment groups (Groups A, B, C) and was the lowest for the combined treatment group. On histologic evaluations, combined treatment was associated with decreased inflammation and increased capillary vessel formation, when compared with control group.

Conclusion: Both sildenafil-only and alprostadil treatments were independently associated with increased flap survival rate. Sildenafil citrate and alprostadil had a synergistic effect in increasing flap survival rate.

Keywords: Alprostadil / Sildenafil / Rats, Sprague-Dawley / Surgical flaps / Survival rate
in treating erectile dysfunction.

This animal study was designed to compare flap survivability between sildenafil citrate and PGE1 and to evaluate whether these effects are synergistic between the two agents.

**METHODS**

**Flap creation and postoperative management of rats**

Upon approved from the institutional animal experimentation ethics review committee, Sprague-Dawley rats were obtained (n=40). The rats were 12 weeks old, weighed between 300 to 400 g, and given a week of adaptation with ad lib water and feed.

On the day of operation, each rat was intramuscularly injected with 0.1 mL/100 g of tiletamine and zolazepam mixture (Virbac Korea Co., Seoul, Korea). Once under full anesthesia, the back was prepared with an electric shaver, and the dorsal surgical site was treated with povidone and alcohol. A modified 2×8 cm McFarlane skin flap was designed over the dorsum between the neck and iliac crest. The flap area was injected with 2% lidocaine without epinephrine. The skin was incised with a No. 15 knife, and the flap was elevated along the plane just deep to the panniculus carnosus. The flap was then sutured back into the original position using 4-0 nylon. Cefazedone sodium (Pazeron, Hanlim Pharm. Co., Seoul, Korea) was intramuscularly injected for three days postoperatively (5 mg/kg/day).

The rats were randomly assigned into four groups of 10 animals each. Group A served as control and was administered with oral an intravenous saline solution. Group B was administered with oral saline solution and intravenous alprostadil (Eglandin, Mitsubishi Tanabe Pharma Co., Ltd., Seoul, Korea). Group C was administered with oral sildenafil citrate (Viagra, Pfizer Australia Pty Ltd., West Ryde, Australia) and intravenous saline solution. Group D was administered with oral sildenafil citrate and intravenous alprostadil.

For Groups C and D, oral sildenafil table (100 mg/tab) was ground with pestle and mortar, dissolved in 10 mL saline solution, and fed through a flexible gastric tube (20 mg/kg/day) for 7 days. For Groups B and D, alprostadil (10 µg/2 mL ampoule) was diluted to a concentration of 1 µg/mL. After warming up the tail with infrared lamp to 30°C–35°C, the solution was injected into the dilated vein through a 24-gauge needle (1 µg/kg/day) for 7 days.

**Photographic flap necrosis evaluation**

The flaps were photographed on postoperative days 1, 3, 5, and 7. The photographs were loaded into Image J software (National Institute of Health) and evaluated by three observers who were blinded to the treatment regimen. Each flap was evaluated for the extent of necrosis as a percentage of total flap area (necrotic index, NI). The values were averaged between the observers and compared across the observations (Fig. 1).

**Tissue specimen preparation and histological observation**

In each group, a randomly selected animal was sacrificed on postoperative days 1, 3, and 5 for histologic evaluation. All of the remaining rats were euthanized on postoperative day 7. The flaps were harvested and fixed in a 10% neutral formalin solution for two days. After paraffin fixation, 2-cm sections were stained with H&E stain. The specimens were examined at 10, 20, and 200 magnifications and were evaluated for epidermal exfoliation due to necrosis, acute and chronic inflammation, granulation tissue formation, and fibrosis. Also, neovascularization was quantified by the number of new capillaries present within a 200× field.

**Statistical analysis**

The SPSS ver. 22.0 package was used for statistical analysis of the NI. Results were expressed in means and standard deviations. The values were compared across the four groups using one-way analysis of variance (ANOVA). Post-hoc analyses were used for multi-comparison through the least significance difference method. Statistical significance was accepted for p<0.05.
RESULTS

Photographic evaluation

All of the animals survived through the study duration, except for those sacrificed for early histologic evaluation. The mean final necrotic areas were 795.1, 558.8, 644.4, and 417.6 mm² for Groups A to D, respectively, and these represented 56%, 42%, 45%, 31% of the total respective flap areas (Table 1, Figs. 1, 2).

To investigate initial pace of necrosis, we compared the extent of necrosis in each group on days 1, 3, 5, and 7 to the final necrotic dimension. On postoperative day 1, the mean necrotic areas were 444.0 mm² (NI=31.8%), 268.3 mm² (NI=20.0%), 326.4 mm² (NI=24.4%), and 223.8 mm² (NI=17.1%) for Groups A to D. On postoperative day 3, the mean necrotic areas were 579.5 mm² (NI=41.3%), 366.1 mm² (NI=26.8%), 519.5 mm² (NI=33.1%), and 326.4 mm² (NI=23.7%). On postoperative day 5, the mean necrotic areas were 717.4 mm² (NI=51.0%), 439.8 mm² (NI=32.7%), and 550.2 mm² (NI=39.2%), and 399.3 mm² (NI=29.2%). On postoperative day 7, the mean necrotic areas were 795.1 mm² (NI=56.8%), 558.5 mm² (NI=42.0%), 644.4 mm² (NI=45.3%), and 417.6 mm² (NI=31.0%).

Overall, the control group of rats showed the highest average areas of flap necrosis at each evaluation. There were no significant patterns in maximum and minimum necrosis areas with respect to the administration of sildenafil or alprostadil (Table 1).

Histological findings

Epidermal exfoliation and flap detachment was greater in the

Table 1. Comparison of flap viability

| Group | Minimum necrotic area (mm²) | Maximum necrotic area (mm²) | Mean necrotic area (mm²) | Mean necrosis rate (ni) (%) |
|-------|-----------------------------|-----------------------------|--------------------------|-----------------------------|
| Day 1 |                             |                             |                          |                             |
| A     | 330.7                       | 513.5                       | 444.0                    | 31.8                        |
| B     | 18.4                        | 468.8                       | 268.3                    | 20.0                        |
| C     | 179.1                       | 456.6                       | 326.4                    | 24.4                        |
| D     | 56.4                        | 437.4                       | 223.8                    | 17.1                        |
| Day 3 |                             |                             |                          |                             |
| A     | 378.0                       | 657.8                       | 579.5                    | 41.3                        |
| B     | 210.0                       | 501.2                       | 366.1                    | 26.8                        |
| C     | 330.0                       | 681.6                       | 519.5                    | 33.1                        |
| D     | 75.0                        | 547.8                       | 326.4                    | 23.7                        |
| Day 5 |                             |                             |                          |                             |
| A     | 455.9                       | 904.1                       | 717.4                    | 51.0                        |
| B     | 258.2                       | 702.9                       | 439.8                    | 32.7                        |
| C     | 476.4                       | 684.2                       | 550.2                    | 39.2                        |
| D     | 138.3                       | 646.9                       | 399.3                    | 29.2                        |
| Day 7 |                             |                             |                          |                             |
| A     | 316.1                       | 975.5                       | 796.1                    | 56.8                        |
| B     | 368.4                       | 850.8                       | 558.5                    | 42.0                        |
| C     | 474.6                       | 798.0                       | 644.4                    | 45.3                        |
| D     | 141.1                       | 715.5                       | 417.6                    | 31.0                        |

Fig. 1. Comparison of flap viability on postoperative day 7. Throughout the entire study duration, flap necrosis was the highest in the control group (Group A).

Fig. 2. Mean necrosis area on postoperative day 7 in each group.
control group of rats (Group A) than in any other group. This group of rats demonstrated the greatest extent of neutrophil infiltration, inflammation, and hypodermic fibrosis.

In Group B, epidermal exfoliation was less than in Group A but greater than Group D. Flaps from Group B rats was found to have moderate neutrophil infiltration. This group had less fibrosis, more granulation tissue formation, and higher density of new capillary formation than Group A.

In Group C, epidermal exfoliation was less than in Group A and greater than in Group D. While more favorable than in Group A, the neutrophilic infiltration and inflammatory responses in this alprostadil-only group were more intense in this group when compared to sildenafil-only or sildenafil-and-alprostadil groups (B and D).

In Group D, epidermal exfoliation was less than in any other groups. The co-administration of sildenafil and alprostadil was associated with relatively lower levels of neutrophilic and lymphocytic infiltration. The highest density of capillary vessel formation was observed in this group (Table 2, Fig. 3).

Table 2. Relative degree of histological findings in each group

| Group | Neutrophil infiltration (acute inflammation) | Lymphocyte infiltration (chronic inflammation) | Fibrosis | Necrosis | Granulation tissue | New capillary blood vessel (count) |
|-------|---------------------------------------------|-----------------------------------------------|----------|----------|--------------------|----------------------------------|
| A     | ++                                          | ++                                            | +++      | +++      | ++                  | + (0–4)                          |
| B     | +                                            | ++                                            | ++       | ++       | +++ (18–22)        |                                  |
| C     | ++                                          | ++                                            | +++      | +        | ++                  | [12–16]                          |
| D     | +                                            | ++                                            | +        | ++       | +++ (31–38)        |                                  |

The degree of lymphocyte infiltration and fibrosis in each group showed no significant differences.

+, few; ++, moderately; +++: much.

Fig. 3. Histologic appearance on postoperative day 7. Control group (H&E): (A) ×10, (B) ×200. Alprostadil-only group (H&E): (C) ×10 (D) ×200. Sildenafil-only group (H&E): (E) ×10, (F) ×200. Alprostadil and Sildenafil group (H&E): (G) ×10, (H) ×200. Red line, skin detachment area; green line, skin maintenance area.
Statistical analysis of flap necrosis area

The mean NI was compared across the four groups using one-way ANOVA. The mean flap necrosis area rate was lowest in combined treatment (Group D, 31.0%) and was highest in control animals (Group A, 56.7%) throughout the experiment (p-value<0.05). Comparing single-agent treatments, the mean NI was higher for alprostadil-only administration (Group C) than for sildenafil-only administration (Group B), though this was not statistically significant. Aprostadil-only administration was associated with significantly higher mean NI, compared to combined treatment (p-value <0.05). Sildenafil-only administration was also associated with higher mean NI compared to combined treatment, though this was not statistically significant observation (Table 3).

DISCUSSION

Skin flaps are used widely in the reconstruction of surface defects in various parts of the body. Regardless of flap classification, all flaps require a minimal amount of oxygen and nutrient delivery in the initial period of grafting, which is dependent on adequate blood flow [2]. Tissue necrosis is not an uncommon complication following flap reconstruction. This is almost always due to insufficient blood flow (whether from insufficient arterial supply or from increased hydrostatic pressure secondary to venous congestion).

However, flap ischemia and necrosis are influenced by a complex set of pathophysiological events. Both local and systemic factors are associated with nutrition and oxygen delivery to flap tissue, precursor metabolites and markers of necrosis, and with wound healing and flap survival [3]. In understanding these factors, various methods have been devised to improve blood flow into flaps and to facilitate angiogenesis. Despite this, flap necrosis remains an unsolved important challenge for plastic surgeons [4].

In the management of patients undergoing flap reconstruction, modifiable factors include smoking cessation, nutrition optimization, minimal use of electrocautery, delicate handling of flap and vascular pedicles, and avoidance of pedicle twist. In addition, various pharmaceutical agents have been used to improve flap blood flow ever since such method was first reported by Adamson et al. [5]. For instance, Leite et al. [6] observed that dimethyl sulfoxide, a strong antioxidant, was useful in ameliorating the deleterious effect of nicotine on flap survival. Pentoxyfilline, a nonselective PDE inhibitor, reduces blood viscosity and improves flap survival, and anthrocyanins are found to have an anti-inflammatory action against reperfusion injuries in a rat model [7].

PGE1 has also been effective in improving flap survivability in experimental and clinical studies. Prostaglandins were first discovered by Euler, and are long chain polyunsaturated fatty acids generated from arachidonic acid through cyclooxygenation. These eicosanoid complexes are classified according to the number of intra-structural double bonds. Of the various prostaglandins, prostaglandin A and prostaglandin E are known to induce vasodilation, with prostaglandin F promoting vasoconstriction.

PGE1 specifically relaxes smooth muscles of medium-sized blood vessels, independent of β-receptors of sympathetic nervous system. PGE1-induced vasodilation reduces systemic vascular resistance and enhances blood flow rate. In addition to vascular smooth muscle tone, the prostaglandin inhibits secretion of platelet aggregation factors such as thrombin, adenosine diphosphate, intravascular collagen, and serotonin. It also inhibits platelet aggregation by increasing the cyclic adenosine monophosphate (c-AMP) concentration within platelets, and increases arteriole blood flow by improving the ability to transform erythrocyte membrane and by enhancing blood fluidity. It is also known to prevent oxygen free radicals from being generated and to help flaps survive surgical flap procedure by weakening neutrophil adherence competence and by reducing vascular endothelial cell injuries. While endogenous PGE1 is rapidly metabolized and inac-

Table 3. Statistical analysis of necrotic index (one-way analysis of variance test)

|       | Group A | Group B | Group C | Group D | p-value |
|-------|---------|---------|---------|---------|---------|
| Day 1 | 31.8±3.7 | 20.0±8.9 | 24.4±6.6 | 17.1±10.4 | 0.001   |
| Day 3 | 41.3±7.7 | 26.8±8.2 | 33.1±8.1 | 23.7±11.2 | <0.001  |
| Day 5 | 51.0±9.1 | 32.7±10.8 | 39.2±5.0 | 29.2±13.8 | <0.001  |
| Day 7 | 56.8±15.6 | 42.0±11.2 | 45.3±6.6 | 31.0±14.1 | <0.001  |

Values are presented as means.
Group A (n=10), saline (control group); Group B (n=10), alprostadil; Group C (n=10), sildenafil; Group D (n=10), alprostadil and sildenafil.
A p<0.05 for multiple comparison of mean values across the groups by post-hoc analysis except group B and group D.
tivated in the lung, the principle active component of apostil is Lipo-PGE1, which uses micro-fat globules as carriers and has a longer half-life in the body.

Sildenafil citrate is a phosphodiesterase-5 inhibitor and prevents degradation of cyclic GMP within the nitrogen guanylate cyclase pathway, which subsequently prevents NO depletion [8]. Tsai et al. [9] demonstrated that intraperitoneal administration of 10 mg/Kg/day of sildenafil citrate was effective in improving the flap survival rate in rats. In a similar study using the rat model, Sarıfakıoğlu et al. [10] found a dose-dependent relationship between sildenafil citrate, with higher doses associated with improved flap survival.

The current study investigated whether the combination of alprostadil and sildenafil citrate was superior to either sildenafil-only or alprostadil-only treatment. Compared to the control group, administration of sildenafil, alprostadil, or a combination of both was associated with significant decreases in flap necrosis. Administration of either sildenafil alone or alprostadil alone was associated with lower NI through the postoperative week, although this difference was not statistically significant.

The pharmacological mechanism of the combined use of alprostadil and sildenafil is not clearly understood. The synergistic effect between alprostadil and sildenafil observed in this study may be due to increased blood flow from vasodilation, as well as increased neovascularization from fibroblast growth factor and vascular endothelial growth factor secretion via the phosphokinase G pathway affected by sildenafil.

This study has some limitations. Photographic evaluation of necrotic areas can vary across observers. Pharmacokinetic management appeared to be associated with decreased flap necrosis, but the study may not have been adequately powered to detect statistical relationship. The flap model in rat represents random skin flap, and the result cannot be generalized to axial or free transfer flaps.

While sildenafil citrate was found to be very effective in enhancing the flap survival rate in this study, continuous administration can result in undesirable symptoms. Such side effects include headache, hot flush, nasal congestion, and dyspepsia, all of which are primarily attributed to vasodilation characterized by sildenafil citrate [11]. It can also inhibit the functions of platelet, prevent platelet aggregation, and cause gastric ulcer. In addition, it can be hazard-ous for heart disease patients since it may weaken cyclic adenosine monophosphate-specific PDE isoform-3 involved in cardiac constriction, dilate blood vessels, and decrease blood pressure abruptly [12]. It is necessary to conduct further research to determine a dose and concentration of sildenafil citrate that can maximize the flap survival rate while preventing systemic adverse effects and to identify long-term effects and complications of sildenafil administration combined with alprostadil and other agents.

In this study, sildenafil citrate and alprostadil were used in single-agent and combination prophylaxis against flap necrosis in rats. Single administration of sildenafil citrate following surgical flap procedure dilated blood vessels, increased blood flow into flaps, prevented flap necrosis, and improved the flap survival rate as much as that observed with alprostadil. The combined use of sildenafil and alprostadil was associated with lowest mean necrosis ratio, but the difference was statistically not significant from alprostadil-only management. Future research should be directed in elucidating the synergistic mechanism between the two vaso-dilatory agents in patients undergoing flap reconstruction and in patients with vascular disease.

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