Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.
Review

Emerging respiratory agents: New viruses for old diseases?

T.P. Sloots\textsuperscript{a,b,c,*}, D.M. Whiley\textsuperscript{a,b}, S.B. Lambert\textsuperscript{a,b}, M.D. Nissen\textsuperscript{a,b,c}

\textsuperscript{a} Queensland Paediatric Infectious Diseases Laboratory, Sir Albert Sakzewski Virus Research Centre, Royal Children's Hospital and Health Service District, Queensland, Australia
\textsuperscript{b} Clinical Medical Virology Centre, University of Queensland, Queensland, Australia
\textsuperscript{c} Microbiology Division, Pathology Queensland Central Laboratory, Queensland, Australia

Received 22 February 2008; accepted 3 March 2008

Abstract

The recent advances in molecular technology have enabled the detection of several new viral agents in specimens collected from the human respiratory tract. Human metapneumovirus was first described in 2001, and is a significant respiratory pathogen, particularly of children. Following the identification of severe acute respiratory syndrome (SARS) associated coronavirus, two other newly detected coronaviruses, NL63 and HKU1, have been linked to respiratory disease in humans. However, identifying a new virus as the causative agent of a specific disease is difficult, and ideally would involve satisfying Koch's postulates. The recently described human bocavirus and polyomaviruses KI and WU have been detected in samples collected from humans with acute respiratory infection, but as yet, have not been conclusively proven to be agents of human disease.

We review the new viral agents that have been detected in respiratory samples since 2001, and examine their contribution as agents of human disease.

© 2008 Elsevier B.V. All rights reserved.

Keywords: Emerging virus; Acute respiratory infection; Respiratory viruses; Epidemiology; Molecular; Clinical; Epidemiology

Contents

1. Introduction ................................................................. 234
2. Human metapneumovirus ...................................................... 234
3. Emerging human coronaviruses ............................................. 236
4. New viruses identified in the respiratory tract of humans ................. 237
4.1. Human bocavirus ...................................................... 237
4.2. Novel human polyomaviruses KVI and WUV ......................... 238
5. Conclusion .............................................................. 239
References ............................................................................ 239

* Corresponding author at: Queensland Paediatric Infectious Diseases Laboratory, Sir Albert Sakzewski Virus Research Centre, Royal Children's Hospital and Health Service District, Herston Road, Herston, Queensland 4029, Australia. Tel.: +61 7 3636 8833; fax: +61 7 3636 1401.
E-mail address: t.sloots@uq.edu.au (T.P. Sloots).

1386-6532/$ – see front matter © 2008 Elsevier B.V. All rights reserved.
doi:10.1016/j.jcv.2008.03.002
1. Introduction

Acute respiratory tract infections (ARTI) are a major cause of morbidity and mortality worldwide, particularly in children, who may experience multiple infections per year until they are 10 years of age (Arnold et al., 2006). Viruses are responsible for the majority of ARTI, with rhinoviruses (HRV), respiratory syncytial virus (RSV), influenza virus (INF) and parainfluenzaviruses (PIV) considered the major pathogens. Traditionally, the diagnosis of these infections relied on the isolation and identification of the viral agent by cell culture or detection of viral antigens by direct immunofluorescent assays (DFA) (Arnold et al., 2006). However, using these methods a large proportion of respiratory infections remained undiagnosed.

Recent advances in molecular biology, particularly the introduction of the polymerase chain reaction (PCR) assay, have greatly improved the detection of viral respiratory pathogens. Yet, even with the most sensitive molecular techniques, only 40–60% of infections were able to be associated consistently with a causative microorganism (Louie et al., 2005). This suggested that additional respiratory pathogens were likely to exist (van den Hoogen et al., 2001). In fact, since 2001, seven previously undescribed viruses have been identified by analysis of clinical specimens from the human respiratory tract (Table 1). These are human metapneumovirus (HMPV) in 2001 (van den Hoogen et al., 2001), three new human coronaviruses (HCoV); the severe acute respiratory syndrome (SARS) associated coronavirus in 2003 (Ksiazek et al., 2003), coronavirus NL63 (NL63) in 2004 (van der Hoek et al., 2004), coronavirus HKU1 (HKU1) in 2005 (Woo et al., 2005a), as well as human bocavirus (HBoV) in 2005 (Allander et al., 2005) and the recently described human polyomaviruses KI (KIV) and WU (WUV) in 2007 (Allander et al., 2007a; Gaynor et al., 2007). These new viral agents were detected by novel molecular methods such as VIDISCA (van der Hoek et al., 2004), pan-viral DNA microarrays (Wang et al., 2003), and high throughput sequencing (Allander et al., 2005, 2007a; Gaynor et al., 2007) (Fig. 1). These methods were comprehensively reviewed by Ambrose and Clewley (2006). More broadly, the advent of these new technologies has greatly stimulated efforts to identify novel viruses in the respiratory tract and in other human disease states.

Once new viral agents are identified, it is important to establish their pathogenic potential. Traditionally, Koch’s postulates have provided the standard for establishing a causal link between a pathogen and disease, but these have been modified over the years to match new knowledge of viral detection and disease processes (Fredericks and Relman, 1996). These modified postulates now propose that a causal relationship between an organism and disease may be established if (i) the organism is consistently present in patients with the disease at a significantly greater prevalence than in matched control subjects, (ii) the disease may be replicated in an appropriate animal model after challenging with the viral pathogen, and subsequent isolation of the agent from the challenged animal, and (iii) a specific immune response to the virus by the host can be demonstrated.

The process of establishing a causal link between a new virus and disease may be aided by the availability of a well-characterized specimen bank. Such specimens collected from patients with a variety of illnesses, supported with comprehensive clinical and epidemiologic data, are a valuable resource for establishing an association between presence of a virus and disease. However, the conclusion about such an association is only valid if they are tested together with appropriately matched controls.

Of the viruses discovered over the last 7 years, HMPV and the newly emerging HCoV are recognized to have global presence, and are considered causative agents of respiratory disease. However, to date, SARS coronavirus has been restricted geographically and has only been associated with limited and sporadic outbreaks of human disease. Of the other emerging viruses, an association with the respiratory tract, or respiratory infection, has been postulated, but not proven. These are HBoV and the newly described KIV and WUV.

This review examines the state of knowledge of newly emerging viruses that have been detected in the human respiratory tract, and their clinical association with human disease.

2. Human metapneumovirus

HMPV is a new human respiratory tract pathogen first described in 2001. The virus was isolated by cell culture from respiratory secretions collected during a 20-year period from Dutch children with ARTI. HMPV showed slow replication in cell culture with characteristic syncytia formation followed by rapid internal disruption of the cells at 10–14 days after inoculation (van den Hoogen et al., 2001).

It was established that HMPV was a respiratory pathogen of primates, by demonstrating active replication of the virus in four juvenile cynomolgus macaques, resulting in mild upper respiratory tract signs in two of these animals. The HMPV genome was characterized using random arbitrarily primed-PCR, and the virus was subsequently classified within Paramyxoviridae based on genome structure and organization. HMPV is the first known mammalian metapneumovirus associated with disease.

Many studies have shown a broad spectrum of clinical signs associated with HMPV infection in patients of all age groups. Like other human respiratory viruses, HMPV may cause upper and lower respiratory tract infection in infants and young children (Williams et al., 2004, 2006). It is second only to RSV as a cause of bronchiolitis in early childhood (Boivin et al., 2002; Freymouth et al., 2003; van den Hoogen et al., 2003; Viazov et al., 2003; Mullins et al., 2004; Williams et al., 2004; Chano et al., 2005; Sasaki et al., 2005). Children younger than 5 years of age are most susceptible to HMPV infection (van den Hoogen et al., 2004), and those younger
than 2 years of age are most likely to be hospitalized with severe symptoms, including bronchiolitis, pneumonia, and bronchitis (Kahn, 2006; Heikkinen et al., 2008). Recently, Heikkinen et al. (2008) showed that acute otitis media developed in 61% of HMPV-infected children younger than 3 years of age. There are no clinical manifestations of HMPV infection that are unique, and infection in young children cannot be distinguished from infection with RSV or INF (Peret et al., 2002; Biacchesi et al., 2003; Esper et al., 2004; Larcher et al., 2005; Martino et al., 2006). In these subjects the virus can cause prolonged and serious bronchitis, cough, wheezing, pneumonia, acute otitis media, asthma (von Linstow et al., 2004; Williams et al., 2004; Vicente et al., 2007), and with exacerbations of both asthma and chronic obstructive pulmonary disease (COPD) in adults (Falsey and Walsh, 2006).

HMPV infection in adults usually presents with mild common cold-like respiratory symptoms such as rhinorrhea, cough and sore throat (Stockton et al., 2002; Falsey et al., 2003). However, disease may be more severe in patients with underlying medical conditions such as cardiopulmonary disease, the elderly and immunocompromized subjects (Boivin et al., 2002; Faux et al., 2003; Hamelin et al., 2005; Honda et al., 2006). In these subjects the virus can cause prolonged and serious infections, and several case studies have described the detection of HMPV in patients with severe lung disease including several fatalities (Pelletier et al., 2002; Cane et al., 2003; Esper et al., 2004; Larcher et al., 2005; Martino et al., 2005; Williams et al., 2005a). Interestingly, one study reported an immunocompromized child, in whom HMPV infection with two different strains of the virus were detected during a 10-month period (Williams et al., 2005a).

It is now recognized that HMPV has a global distribution (van den Hoogen et al., 2001; Boivin et al., 2002, 2004; Nissen et al., 2002; Cuevas et al., 2003; Esper et al., 2003; Freymouth et al., 2003; Maggi et al., 2003b; Peiris et al., 2003; Dollner et al., 2004; Ebihara et al., 2004; Al-Sonboli et al., 2005; Kim and Lee, 2005; Ludewick et al., 2005; Noyola et al., 2005; Robinson et al., 2005; Garcia-Garcia et al., 2006; Gray et al., 2006; Samransamruajkit et al., 2006) circulating predominantly in late winter and spring. Usually the peak of infection is coincident with or follows the peak of RSV infection (Boivin et al., 2003; Karron et al., 2003; Osterhaus and Fouchier, 2003; Peiris et al., 2003; Esper et al., 2004; Konig et al., 2004; Mullins et al., 2004; Bouscambert-Duchamp et al., 2005; Chano et al., 2005; Agapov et al., 2006; Kaida et al., 2006; Nicholson et al., 2006; Sarasini et al., 2006). Many respiratory viruses share seasonality and susceptible populations. As a result, co-detection of HMPV with other respiratory viruses has been reported at rates ranging from 1% to greater than 10% (Viazov et al., 2003; Esper et al., 2004; Nissen et al., 2002; Cuevas et al., 2003; Esper et al., 2004; Xepapadaki et al., 2004). One study reported that dual infection with HMPV and RSV is associated with severe bronchiolitis and significantly increased the risk of admission to a paediatric intensive-care unit (Semple et al., 2005). This is at odds with previous findings which failed to detect a predisposition to more severe disease in patients with multiple respiratory virus detections (Lina et al., 2005).

Two major genotypes of HMPV, designated A and B are recognized, each with two subtypes (A1 and A2; B1 and B2) (Peret et al., 2002; Biacchesi et al., 2003; Esper et al., 2004; Mackay et al., 2004; Williams et al., 2004; Chano et al., 2005;
Vicente et al., 2007). Circulating strains of HMPV differ in different populations at any single time, and the same strain may be present in geographically distinct locations in different years. In an Australian study, the predominant genotype of HMPV switched over 4 consecutive years from genotype A1 to A2 to B1 (Sloots et al., 2006a). Similar observations have been reported elsewhere (Gerna et al., 2005). So far there are no data to suggest that severity of illness may be associated with a specific genotype (Kahn, 2006).

Seroprevalence studies have shown that the presence of HMPV-specific antibodies was greater than 90% by age 5 years, and nearly 100% in adults (van den Hoogen et al., 2001; Leung et al., 2005). This suggests that primary infection with HMPV occurs early in childhood, typically between the ages of 1–5 years (van den Hoogen et al., 2001; Ebihara et al., 2003; Wolf et al., 2003; Leung et al., 2005). The reported seroprevalence in infants younger than 3 months of age was greater than 90%, decreasing to a minimum by age 13 months and increasing to 52% by age 2 years (Wolf et al., 2003). Although this appears to be indicative of maternally derived antibodies (Leung et al., 2005), the role of such antibodies in the protection or moderation of HMPV infection remains to be determined.

3. Emerging human coronaviruses

Coronaviruses (CoV) infect many species of animals, including humans. The first HCoV, of which 229E and OC43 are the representatives, were identified in the 1960s (Tyrrell and Bynoe, 1965; Hamre and Procknow, 1966; McIntosh et al., 1967). Coronaviruses are classified into three distinct groups, with 229E and NL63 included in Group 1 coronaviruses and OC43 and HKU1 in Group 2 (Lai and Cavanagh,
SARS–CoV, represents an early split from Group 2 coronaviruses (Eickmann et al., 2003; Marra et al., 2003; Rota et al., 2003; Snijder et al., 2003) and is believed to have originated from wild animals (Guan et al., 2003; Lau et al., 2005).

3.1. SARS coronavirus

First reports of an outbreak of a new respiratory illness involving “atypical pneumonia” were published in 2003 (Center for Disease Control and Prevention (CDC), 2003; Tsang et al., 2003). The novel virus present in these cases was isolated from patients’ lungs and sputa on Vero E6 cells (Drosten et al., 2003a,b; Peiris et al., 2003; Ksiazek et al., 2003). Sequencing of the viral isolate showed the presence of a new human coronavirus (Rota et al., 2003; Snijder et al., 2003). It was demonstrated that this virus was the aetiological agent for SARS by transmission of respiratory disease to nonhuman primates (Fouchier et al., 2003). The SARS epidemic was halted by a highly effective global public health response co-ordinated by the World Health Organization, and there is no evidence that SARS–CoV is currently circulating in humans. However, the SARS outbreak focussed renewed attention on coronaviruses generally, resulting in the identification of two more human coronaviruses, NL63 and HKU1.

3.2. Human coronaviruses NL63 and HKU1

HCoV–NL63 was first detected in 2004 in a child with bronchiolitis in The Netherlands (van der Hoek et al., 2004), whilst HCoV–HKU1 was detected in 2005 in an adult with chronic pulmonary disease in Hong Kong (Woo et al., 2005a). After the initial discovery, several groups have reported presence of these viruses in their populations illustrating global presence (Fouchier et al., 2004; Arden et al., 2005; Bastien et al., 2005; Chiu et al., 2005; Ebihara et al., 2005a; Kaiser et al., 2005; Moes et al., 2005; Choi et al., 2006; Esper et al., 2006; Gerna et al., 2006; Sloots et al., 2006b; Vabret et al., 2005, 2006). These reports showed that NL63 and HKU1 may be detected in 1 to 10% of patients with acute respiratory tract infections. Co-detection of these viruses with other respiratory viruses was commonly reported (van der Hoek et al., 2006a). Like SARS–CoV, NL63 has been successfully cultured in monkey epithelial cell lines (Fouchier et al., 2004; van der Hoek et al., 2004; Schildgen et al., 2006), but to date, it has not been possible to culture HCoV–HKU1, despite attempts to do so in many cell lines. This difficulty with isolating some newly identified viruses particularly explains why they have not previously been detected.

Initial studies described the presence of NL63 in young children admitted to hospital with severe lower respiratory tract infections (LRTIs) (Fouchier et al., 2004; van der Hoek et al., 2004), but the virus has also been detected in elderly patients with a fatal outcome, showing that the severity of the respiratory disease can be substantial (Bastien et al., 2005).

HCoV–NL63 was proven to be a respiratory pathogen and has been identified as an agent of laryngotracheitis (croup) (Forster et al., 2004; Konig et al., 2004; van der Hoek et al., 2006b). In a case-controlled study of young German children it was reported that the chance of croup is 6.6 times higher in NL63-positive children than in NL63-negative children (Forster et al., 2004; Konig et al., 2004). Also, a link between NL63 and Kawasaki disease has been suggested (Esper et al., 2005), but other groups recently questioned this association (Belay et al., 2005; Ebihara et al., 2005b; Shimizu et al., 2005; Baker et al., 2006; Chang et al., 2006).

The first described HKU1 cases were amongst elderly patients and children with major underlying disease (Woo et al., 2005b). Similar findings in the USA (Esper et al., 2006) and France (Vabret et al., 2006) suggested that HKU1 infection may aggravate the conditions of persons with an underlying disease, thereby resulting in hospitalization.

Common respiratory symptoms accompanying HKU1 infection are rhinorrhea, fever, coughing, wheezing, and disease manifestations include bronchiolitis and pneumonia (Lau et al., 2006; Sloots et al., 2006b). It has been suggested that HKU1 might also be involved in gastrointestinal disease (Vabret et al., 2006).

HKU1 and NL63 infections are generally not life threatening, certainly not in otherwise healthy persons. This suggests that these coronaviruses, like 229E and OC43, are predominantly common cold viruses that can cause more severe clinical symptoms in young children, elderly persons, and the immunocompromized (McIntosh et al., 1974; Holmes, 2001; van Elden et al., 2004).

Phylogenetic analysis has shown that NL63 may be classified into two genotypes (van der Hoek et al., 2004; Arden et al., 2005; Chiu et al., 2005; Vabret et al., 2005), and full-genome sequencing of HKU1 isolates has also revealed the presence of two genotypes (A and B) with the possibility of a third genotype (C) that is the resultant of recombination between genotypes A and B (Woo et al., 2006).

4. New viruses identified in the respiratory tract of humans

4.1. Human bocavirus

Human bocavirus was first described in 2005 (Allander et al., 2005). This virus was identified by a procedure based on DNase treatment of pooled respiratory samples, random amplification and cloning, followed by large-scale sequencing and bioinformatic analyses. In their original study, Allander et al. (2005) showed HBoV had a prevalence of 3.1% in Swedish children with LRTI. Soon thereafter, a second study from Australia reported a 5.2% prevalence of HBoV in respiratory samples collected from children with ARTI during winter (Sloots et al., 2006b). In a study of hospitalized patients, Arnold et al. (2006) detected HBoV in 5.6% of children, showing a peak prevalence of infection in
spring. Subsequently, presence of the virus has been reported by investigators in Europe, the United Kingdom, the United States, Canada, Asia, and Australia, with prevalence rates ranging from 1.5% to 11.3% in these populations (Allander et al., 2005; Arnold et al., 2006; Bastien et al., 2006, 2007; Choi et al., 2006; Foulongne et al., 2006; Kesebir et al., 2006; Ma et al., 2006; Manning et al., 2006; Sloots et al., 2006b; Fry et al., 2007; Maggi et al., 2007; Naghipour et al., 2007; Neske et al., 2007). However, the results from these studies, regarding seasonality, indicate that there is no obvious regular seasonal occurrence of HBoV. A common feature of these studies was that HBoV was most frequently detected in infants younger than 3 years of age.

Arnold et al. (2006) proposed an association between detection of HBoV in the upper respiratory tracts of children and LRTI. They demonstrated that presence of HBoV in the upper respiratory tract was positively associated with asthma, acute otitis media, and pneumonia. The most common signs observed in HBoV-infected children were respiratory distress and hypoxia associated with a diagnosis of bronchiolitis, supporting the concept that HBoV may directly infect the lower respiratory tract.

Recent studies reporting an association of HBoV with ARTI, differed from other similar studies by including control populations (Kesebir et al., 2006; Manning et al., 2006; Fry et al., 2007; Maggi et al., 2007). HBoV was detected at high frequency in subjects with overt disease, whereas the virus was either not found or was found very infrequently in control subjects. Although a positive association with ARTI was suggested, the results remain inconclusive, given the high rate at which other viruses were detected in HBoV-positive specimens. These findings add important corroborative support, but still do not prove causality of disease.

In a similar study using control subjects, Allander et al. (2007b) proposed HBoV as a cause of acute wheezing in children. However, differences in specimen type, age groups, and time of year tested for controls and test subjects were considerable and may have confounded detection rates in this study.

HBoV has only rarely been detected in immunocompetent adult subjects (Bastien et al., 2006) yet was frequently detected in immunosuppressed adults (Kupfer et al., 2006; Manning et al., 2006; Maggi et al., 2007). However, the presence of HBoV in these subjects may be the result of re-infection, viral persistence or reactivation (Allander, 2008). To answer these questions, prospective longitudinal examination of HBoV in immunosuppressed patients will be an important area for future studies.

The idea that HBoV infection may play a possible role in gastroenteritis has been suggested because of the frequent manifestation of gastrointestinal symptoms in HBoV-positive subjects. Several studies have examined an association of HBoV with gastrointestinal disease, and have reported HBoV presence in faeces with a prevalence rate ranging from 0.8% to 9.1% in these patients (Lau et al., 2007; Lee et al., 2007; Neske et al., 2007; Vicente et al., 2007). Also, Albuquerque et al. (2008) reported the detection of HBoV in Brazilian children with gastroenteritis in the absence of respiratory symptoms. Although these findings suggest that HBoV may play a role as an agent of acute gastroenteritis, it is possible that this is a clinical presentation of a systemic response to HBoV infection by the host. This data however, does add new knowledge about potential mechanisms of transmission for this virus.

So far, two slightly different genetic lineages of HBoV have been reported globally that share greater than 98% amino acid identity (Allander et al., 2005; Kesebir et al., 2006; Bastien et al., 2007; Chieochansin et al., 2007; Neske et al., 2007). Also, Lau et al. (2007) reported a high level of genomic conservation of HBoV detected in samples from different anatomical sites, showing that a single lineage of HBoV was found in both respiratory tract and enteric specimens. Similarly, HBoV detected in patients over a 21-month period showed little genetic variation within the NP-1 and NS1 genes (Arnold et al., 2006).

4.2. Novel human polyomaviruses KIV and WUV

Recently, two new human polyomaviruses, KIV and WUV were detected in respiratory tract specimens (Allander et al., 2007a; Gaynor et al., 2007). The viruses were identified following large-scale molecular screening using high throughput DNA sequencing of random clones.

KIV and WUV are related members of a new group (or possibly sub-family) within the Polyomaviridae, small DNA viruses with circular, covalently closed double-stranded genomes infecting a wide range of mammalian and avian species. Polyomaviruses frequently establish persistent, lifelong infections characterized by highly restricted or latent infection.

In their initial reports Allander et al. (2007a) detected KIV in 1% of nasopharyngeal aspirates and 0.5% of stool samples collected from a Swedish population and Gaynor et al. (2007) reported a prevalence of 3% and 0.6% for WUV in respiratory samples from Australia and the USA, respectively. Neither group was able to detect these viruses in urine samples.

Since these first reports, KIV and WUV DNA have been detected in respiratory specimens from a number of geographic locations, suggesting a global presence for these viruses. Studies from Australia and Scotland reported a prevalence for KIV of 2.5% and 1.4%, respectively (Bialasiewicz et al., 2008; Norja et al., 2007), and for WUV prevalence rates ranged from 1% in a Scottish population to 7% in Australian and South Korean populations (Bialasiewicz et al., 2008; Han et al., 2007; Norja et al., 2007).

One striking feature of early findings concerning KIV and WUV is their high rate of co-detection with other respiratory viruses. A co-detection rate of 74% has been observed for KIV (Bialasiewicz et al., 2008) and rates ranging from 68% to 79% for WUV (Bialasiewicz et al., 2008; Han et al., 2007; Le et al., 2007). A level of 10% co-detection of KIV with WUV in the absence of any other respiratory virus was reported by Bialasiewicz et al. (2008).
Although an aetiologic role in childhood respiratory disease has been proposed for KIV and WUV (Allander et al., 2007a; Bialasiewicz et al., 2008; Gaynor et al., 2007), initial investigations have been based on large-scale screening of respiratory samples referred for general diagnostic testing from patients with moderate to severe respiratory disease. It is however difficult to assess the pathogenic role of these viruses without examining samples collected from matched control populations. Three recent studies have sought to do this for WUV, reporting prevalence rates in symptomatic versus asymptomatic subjects of 7.0% versus 4.2% (Han et al., 2007), 2.5% versus 6.4% (Abed et al., 2007) and 1.0% versus 5.4% (Norja et al., 2007). However, these data must be interpreted with caution as each of these studies examined only small numbers of controls and some of these were not well matched to the symptomatic population tested. Interestingly, although Norja et al. (2007) reported a high prevalence of KIV (5.4%) and WUV (5.4%) in their control subjects, 5 of the 6 (83%) positive specimens came from subjects who were classified as immunosuppressed. If consistent, the presence of KIV and WUV in control populations may represent a latent or persistent infection with subsequent asymptomatic reactivation or prolonged shedding from a prior respiratory tract infection.

Persistent shedding of WUV over a 6–8 week period was amongst a number of interesting observations reported in two children (1.5 and 4 years old) from St. Louis, USA, and WUV was also the only virus detected in a 1-day-old infant with respiratory symptoms raising interesting questions about possible transmission mechanisms of this virus (Le et al., 2007).

To date, little information is available regarding the molecular diversity of KIV and WUV strains present in different populations. Early data from Australian, USA and Canadian WUV-positive samples showed limited or no genetic variations based on sequencing from the large TAg region and the VP2 region (Abed et al., 2007; Gaynor et al., 2007). This was substantiated by whole genome analysis of Australian and USA samples which confirmed limited sequence variation amongst KIV and WUV strains from those patient groups (Bialasiewicz et al., 2008).

Further studies will need to be completed before the role of KIV and WUV as respiratory pathogens can be confirmed. It remains possible that these viruses are not involved in respiratory disease and their presence in the respiratory tract simply reflects their mode of transmission. However, considering the oncogenic potential of polyomaviruses in mammals, linking KIV and WUV to a particular disease will be important and may have significant medical implications.

5. Conclusion

With the development of new molecular technology, our ability to detect and characterize new viral agents has greatly improved. As a result, genome sequences have been described for new viruses that are associated with the human respiratory tract, gastrointestinal tract as well as new blood-borne viruses. Some of these are recognized as significant human pathogens causing disease in certain population groups. Others can be found in clinical specimens without definitive evidence for their role as the causative agent of disease, and yet others, like TT-(torqueteno) virus (Maggi et al., 2003a; Pifferi et al., 2006) and mimivirus (Khan et al., 2007; Raoult et al., 2007) have been loosely associated with respiratory disorders in humans.

Many putative viral respiratory pathogens could not be isolated from patients with overt disease, and researchers have as yet been unable to satisfy the modified Koch’s postulates that will prove a causal relationship. It is important therefore, that further extensive clinical studies be continued in an attempt to define the role of these agents as human pathogens.

Still, for a significant proportion of clinical infectious disease of suspected viral origin, a pathogen cannot be identified. Although new molecular methods are increasingly used to investigate these unknown causes of disease, they remain technically challenging and prone to the amplification of non-viral related sequence artefacts. However, with continuing advances in molecular technology and the development of more reliable, robust and reproducible molecular techniques, it seems certain that new potential viral pathogens of humans will be discovered.

References

Abed Y, Wang D, Boivin G. WU polyomavirus in children. Canada. Emerg Infect Dis 2007;13:1936–8.
Agapov E, Sumino KC, Gaudreault-Keener M, Storch GA, Holtzman MJ. Genetic variability of human metapneumovirus infection: evidence of a shift in viral genotype without a change in illness. J Infect Dis 2006;193:396–403.
Albuquerque MCM, Rocha LN, Benati FI, Soares CC, Maranhao AG, Ramírez ML, et al. Human bocavirus infection in children with gastroenteritis. Brazil. Emerg Infect Dis 2008;13:1756–8.
Al-Sonboli N, Hart CA, Al-Aryani A, Banajeh SM, Al-Aghbari N, Dove W, et al. Respiratory syncytial virus and human metapneumovirus in children with acute respiratory infections in Yemen. Pediatr Infect Dis J 2005;24:734–6.
Allander T, Tammi MT, Eriksson M, Bjerkner A, Tiveljung-Lindell A, Andersson B. Cloning of a human parvovirus by molecular screening of respiratory tract samples. Proc Natl Acad Sci USA 2005;102:12891–6.
Allander T, Andreasson K, Gupta S, Bjerkner A, Bogdanovic G, Persson MA, et al. Identification of a third human polyomavirus. J Virol 2007a;81:4130–6.
Allander T, Jarth T, Gupta S, Niesters HG, Lehtinen P, Osterback R, et al. Human bocavirus and acute wheezing in children. Clin Infect Dis 2007b;44:904–10.
Allander T. Human bocavirus. J Clin Virol 2008;41:29–33.
Ambrose HE, Clewley PJ. Virus discovery by sequence-independent genome amplification. Rev Med Virol 2006;16:365–83.
Arden KE, Nissen MD, Sloots TP, Mackay IM. New human coronavirus, HCoV–NL63, associated with severe lower respiratory tract disease in Australia. J Med Virol 2005;75:455–62.
Arnold JC, Singh KK, Spector SA, Sawyer MH. Human bocavirus: prevalence and clinical spectrum at a children’s hospital. Clin Infect Dis 2006;43:283–8.
Baker SC, Shimizu C, Shike H, Garcia F, van der HL, Kuiper TW, et al. Human coronavirus-NL63 infection is not associated with acute Kawasaki disease. Adv Exp Med Biol 2006;581:523–6.

Bastien N, Ward D, Van CP, Brandt K, Lee SH, McNabb G, et al. Human metapneumovirus infection in the Canadian population. J Clin Microbiol 2003;41:4642–6.

Bastien N, Anderson K, Hart L, Van CP, Brandt K, Milley D, et al. Human coronavirus NL63 infection in Canada. J Infect Dis 2005;191:503–6.

Bastien N, Brandt K, Dust K, Ward D, Li Y. Human Bocavirus infection, Canada. Emerg Infect Dis 2006;12:848–50.

Bastien N, Chui N, Robinson JL, Lee BE, Dust K, Hart L, et al. Detection of human bocavirus in Canadian children in a 1-year study. J Clin Microbiol 2007;45:610–9.

Belay ED, Erdman DD, Anderson LJ, Peret TC, Schrag SJ, Fields BS, et al. Kawasaki disease and human coronavirus. J Infect Dis 2005;192:352–3.

Biachesi S, Skiadopoulos MH, Boivin G, Hanson CT, Murphy BR, Collins PL, et al. Genetic diversity between human metapneumovirus subgroups. Virology 2003;315:1–9.

Bialasiewicz S, Whiley DM, Lambert SB, Jacob K, Bletchly C, Wang D, et al. Presence of the newly discovered human polyomaviruses KI and WU in Australian patients with acute respiratory tract infection. J Clin Virol 2008;41:63–8.

Boivin G, Abed Y, Pelletier G, Ruel L, Moisan D, Cote S, et al. Virological features and clinical manifestations associated with human metapneumovirus: a new paramyxovirus responsible for acute respiratory tract infections in all age groups. J Infect Dis 2002;186:1330–40.

Boivin G, De SG, Cote S, Gilca R, Abed Y, Rochette L, et al. Human metapneumovirus infections in hospitalized children. Emerg Infect Dis 2003;9:634–40.

Boivin G, Mackay I, Sloots TP, Madhi S, Freymuth F, Wolf D, et al. Global genetic diversity of human metapneumovirus fusion gene. Emerg Infect Dis 2004;10:1154–7.

Boucsumbert-Duchamp M, Lina B, Trompette A, Moret H, Motte J, Andreoletti L. Detection of human metapneumovirus RNA sequences in nasopharyngeal aspirates of young French children with acute bronchiolitis by real-time reverse transcriptase PCR and phylogenetic analysis. J Clin Microbiol 2005;43:1411–4.

Cane PA, van den Hoogen BG, Chakrabarti S, Fegan CD, Osterhaus AD. Human metapneumovirus in a haematopoietic stem cell transplant recipient with fatal lower respiratory tract disease. Bone Marrow Transplant 2003;31:309–10.

Center for Disease Control Prevention (CDC). Outbreak of severe acute respiratory syndrome—worldwide. Morb Mortal Wkly Rep 2003;52(11):226–8.

Chang LY, Chiang BL, Kao CL, Wu MH, Chen PJ, Kao CL, Wu MH, et al. Identification of a novel human coronavirus NL63 associated with respiratory tract infections in young children under <5 years. J Infect Dis 2004;189:1388–96.

Chen PJ, Berkhout B, et al. Presence of the new human metapneumovirus in young children with bronchiolitis. Pediatrics 2003;111:1407–9.

Chen PJ, Berkhout B, Anderson LJ, Walsh EE. Human metapneumovirus infections in young and elderly adults. J Infect Dis 2003;187:785–90.

Chen PJ, Walsh EE. Viral pneumonia in older adults. Clin Infect Dis 2006;42:518–24.

Chikahisa T, Shuichiro K, Naoe M, Junichi K, Hidetoshi K, et al. Human coronavirus infection in norwegian children. Pediatr Infect Dis J 2003;22:92–4.

Choi EH, Lee HJ, Kim SJ, Eun BW, Kim NH, Lee JA, et al. The association of newly identified respiratory viruses with lower respiratory tract infections in Korean children, 2000–2005. Clin Infect Dis 2006;43:585–92.

Cuevas LE, Nasser AM, Dove W, Gurgel RQ, Greensill J, Hart CA. Human metapneumovirus and respiratory syncytial virus, Brazil. Emerg Infect Dis 2003;9:1626–8.
Gerna G, Campanini G, Rovida F, Sarasin A, Lillero D, Paolucci S, et al. Changing circulation rate of human metapneumovirus strains and types among hospitalized pediatric patients during three consecutive winter-spring seasons. Brief report. Arch Virol 2005;150:2365–75.

Gerna G, Campanini G, Rovida F, Percivalle E, Sarasin A, Marchi A, et al. Genetic variability of human coronavirus OC43-, 229E-, and NL63-like strains and their association with lower respiratory tract infections of hospitalized infants and immunocompromised patients. J Med Virol 2006;78:938–49.

Gray GC, Capuano AW, Settenquist SF, Sanchez JL, Neville JS, Olson J, et al. Human metapneumovirus Peru. Emerg Infect Dis 2006;12:547–50.

Guan Y, Zheng BJ, He YQ, Liu XL, Zhuang ZX, Cheung CL, et al. Isolation and characterization of viruses related to the SARS coronavirus from animals in southern China. Science 2003;302:276–8.

Hamelin ME, Cote S, Laforgue J, Lampron N, Bourbeau J, Weiss K, et al. Human metapneumovirus infection in adults with community-acquired pneumonia and exacerbation of chronic obstructive pulmonary disease. Clin Infect Dis 2005;41:498–502.

Hamre D, Procknow JJ. A new virus isolated from the human respiratory tract. Proc Soc Exp Biol Med 1966;121:190–3.

Han TH, Chung JY, Koo JW, Kim SW, Hwang E-S. WU polyomavirus infection in children with acute lower respiratory tract infections, South Korea. Emerg Infect Dis 2007;13:1766–8.

Heikkinen T, Osterback R, Peltola V, Jartti T, Vainionpaa R. Human metapneumovirus infections in children. Emerg Infect Dis 2008;14:101–6.

Holmes K. Coronaviruses. In: Knipe D, Howley P, editors. Fields Virology. Philadelphia: Lippincott Williams & Wilkins; 2001. p. 1187–203.

Honda H, Iwashashi J, Kashitogi T, Imanura Y, Hamada N, Anraku T, et al. Outbreak of human metapneumovirus infection in elderly infants in Japan. J Am Geriatr Soc 2006;54:177–80.

Kahn JS. Epidemiology of human metapneumovirus. Clin Microbiol Rev 2006;19:546–57.

Kaida A, Iritani N, Kubo H, Shiomi M, Kohdera U, Murakami T. Seasonal distribution and phylogenetic analysis of human metapneumovirus among children in Osaka City, Japan. J Clin Virol 2006;35:394–9.

Kaiser L, Regamey N, Roiha H, Deffernez C, Frey U. Human coronavirus NL63 associated with lower respiratory tract symptoms in early life. Pediatr Infect Dis J 2005;24:1111–2.

Karron RA, Belshe RB, Wright PF, Thumar B, Burns B, Newman F, et al. A live human parainfluenza type 3 virus vaccine is attenuated and immunogenic in young infants. Pediatr Infect Dis J 2005;22:394–405.

Keserib D, Vazquez M, Weibel C, Shapiro ED, Fergusson D, Landry ML, et al. Human bocavirus infection in young children in the United States: molecular epidemiological profile and clinical characteristics of a newly emerging respiratory virus. J Infect Dis 2006;194:1276–82.

Khan M, La SB, Lepidi H, Raoul D. Pneumonia in mice inoculated experimentally with Acanthamoeba polyphaga mimivirus. Microb Pathog 2007;42:56–61.

Kim YK, Lee HJ. Human metapneumovirus-associated lower respiratory tract infections in korean infants and young children. Pediatr Infect Dis J 2005;24:1015–7.

Kong B, Konig W, Arnold M, Reracht H, Iborst G, Forster J, Prospective study of human metapneumovirus infection in children less than 3 years of age. J Clin Microbiol 2004;42:4632–5.

Ksiaczek TG, Erdman D, Goldsmith CS, Zaki SR, Peret T, Emery S, et al. A novel coronavirus associated with severe acute respiratory syndrome. N Engl J Med 2003;348:1953–66.

Kupfer B, Vehreschild J, Cornely O, Kaiser R, Plum G, Viazov S, et al. Severe pneumonia and human bocavirus in adult. Emerg Infect Dis 2006;12:1614–6.

Lai MM, Cavanagh D. The molecular biology of coronaviruses. Adv Virus Res 1997;48:1–100.

Larcher C, Geltner C, Fischer H, Nachbaur D, Muller LC, Huemer HP. Human metapneumovirus infection in lung transplant recipients: clinical presentation and epidemiology. J Heart Lung Transplant 2005;24:1891–901.

Lau SK, Woo PC, Li KS, Huang Y, Tsai HW, Wong BH, et al. Severe acute respiratory syndrome coronavirus-like virus in Chinese horseshoe bats. Proc Natl Acad Sci USA 2005;102:14040–5.

Lau SK, Woo PC, Yip CC, Tse H, Tsai HW, Cheng VC, et al. Coronavirus HKU1 and other coronavirus infections in Hong Kong. J Clin Microbiol 2006;44:2063–71.

Lee JI, Chung JY, Han TH, Song MO, Hwang ES. Detection of human bocavirus in children hospitalized because of acute gastroenteritis. J Infect Dis 2007;196:994–7.

Leung J, Esper F, Weibel C, Kahn JS. Seroepidemiology of human metapneumovirus (hMPV) on the basis of a novel enzyme-linked immunosorbent assay utilizing hMPV fusion protein expressed in recombinant vesicular stomatitis virus. J Clin Microbiol 2005;43:1213–9.

Lina B, Valette M, Foray S, Luciani J, Stagnara J, DM, et al. Surveillance of community-acquired viral infections due to respiratory viruses in Rhone-Alpes (France) during winter 1994 to 1995. J Clin Microbiol 1996;34:3071–11.

Louie JK, Hacker JK, Gonzales R, Mark J, Maselli JH, Yagi S, et al. Characterization of viral agents causing acute respiratory infection in a San Francisco University Medical Center Clinic during the influenza season. Clin Infect Dis 2005;41:922–8.

Ludewig HP, Abed Y, van NN, Boivin G, Klugman KP, Madhi SA. Human metapneumovirus genetic variability, South Africa. Emerg Infect Dis 2005;11:1074–8.

Ma X, Endo R, Ishiguro N, Ebihara T, Ishiko H, Ariga T, et al. Detection of human bocavirus in Japanese children with lower respiratory tract infections. J Clin Microbiol 2006;44:1132–4.

Mackay IM, Bialasiewicz S, Walluzzaman Z, Chadlow GR, Foged DC, Laingam S, et al. Use of the P gene to genotype human metapneumovirus identifies 4 viral subtypes. J Infect Dis 2004;190:1913–8.

Magni F, Pifferi M, Fornai C, Andreoli E, Tempestini E, Vatteroni M, et al. TT virus in the nasal secretions of children with acute respiratory diseases: relations to viremia and disease severity. J Virol 2003a;77:2418–25.

Magni F, Pifferi M, Vatteroni M, Fornai C, Tempestini E, Anzilotti S, et al. Human metapneumovirus associated with respiratory tract infections in a 3-year study of nasal swabs from infants in Italy. J Clin Microbiol 2003b;41:2987–91.

Magni F, Andreoli E, Pifferi M, Meschi S, Rocchi J, Bendinelli M. Human bocavirus in Italian patients with respiratory diseases. J Clin Virol 2007;38:321–5.

Manning A, Russell V, Eastick K, Leadbetter GH, Hallam N, Templeton K, et al. Epidemiological profile and clinical associations of human bocavirus and other human pariviruses. J Infect Dis 2006;194:1283–90.

Marra MA, Jones SJ, Astell CR, Holt RA, Brooks-Wilson A, Butterfield YS, et al. The genome sequence of the SARS-associated coronavirus. Science 2003;300:1399–404.

Martino R, Porras RP, Rabella N, Williams JV, Ramila E, Margall N, et al. Prospective study of the incidence, clinical features, and outcomes of symptomatic upper and lower respiratory tract infections by respiratory viruses in adult recipients of hematopoietic stem cell transplants for hematologic malignancies. Biol Blood Marrow Transplant 2005;11:781–96.

McAdam AJ, Hasenbein ME, Feldman HA, Cole SE, Offermann JT, Riley AM, et al. Human metapneumovirus in children tested at a tertiary-care hospital. J Infect Dis 2004;190:20–6.

McIntosh K, Chao RK, Krause HE, Wasi R, Mocca HE, Mufson MA. Coronavirus infection in acute lower respiratory tract disease of infants. J Infect Dis 1974;130:502–7.
Sarasini A, Percivalle E, Rovida F, Campanini G, Genini E, Torsellini M, et al. The discovery and characterization of Mimivirus. J Med Virol 2007;79:539–43.

Neske F, Blessing K, Tollmann F, Schubert J, Rethwilm A, Kreth HW, et al. Real-time PCR for diagnosis of human bocavirus infections and phylogenetic analysis. J Clin Microbiol 2007;45:2116–22.

Nicholson KG, McNally T, Silverman M, Simons P, Stockton JD, Zambon MC. Rates of hospitalisation for influenza, respiratory syncytial virus and human metapneumovirus among infants and young children. Vaccine 2006;24:102–8.

Nissen MD, Siebert DJ, Mackay IM, Sloots TP, Withers SJ. Evidence of human metapneumovirus in Australian children. Med J Aust 2002;176:188.

Noria P, Ulibos I, Templeton K, Simmonds P. No evidence for an association between infections with WU and KI polioviruses and respiratory disease. J Clin Virol 2007;40:307–11.

Noyola DE, puche-Solis AG, Herrera-Diaz A, Soria-Guerra RE, Sanchez-Alvarado J, Lopez-Revilla R. Human metapneumovirus infections in Mexico: epidemiological and clinical characteristics. J Med Virol 2005;79:960–74.

Osterhaus A, Fouchier R. Human metapneumovirus in the community. Lancet 2003;361:890–1.

Peiris JS, Chu CM, Cheng VC, Chan KS, Hung IF, Poons LL, et al. Clinical progression and viral load in a community outbreak of coronavirus-associated SARS pneumonia: a prospective study. Lancet 2003;361:1767–72.

Pelletier G, Dery P, Abed Y, Boivin G. Respiratory tract reinfections by the new human Metapneumovirus in an immunocompromised child. Emerg Infect Dis 2002;8:976–8.

Peret TC, Boivin G, Li Y, Couillard M, Humphrey C, Osterhaus AD, et al. Characterization of human metapneumoviruses isolated from patients in North America. J Infect Dis 2002;185:1660–3.

Pifferi M, Maggi F, Caramella D, De ME, Andreoli E, Meschi S, et al. High torquevirus loads are correlated with bronchiectasis and peripheral airflow limitation in children. Pediatr Infect Dis J 2006;25:804–8.

Raoult D, La SB, Birtles R. The discovery and characterization of Mimivirus, the largest known virus and putative pneumonia agent. Clin Infect Dis 2007;45:95–102.

Robinson JL, Lee BE, Bastien N, Li Y. Seasonality and clinical features of human metapneumovirus infection in children in Northern Alberta. J Med Virol 2005;76:98–105.

Rota PA, Oberste MS, Monroe SS, Nix WA, Campagnoli R, Icenogle JP, et al. Characterization of a novel coronavirus associated with severe acute respiratory syndrome. Science 2003;300:1394–9.

Samransruriyakit R, Thanasugarn W, Prapphal N, Theamboonlers A, Raoult D, La SB, Birtles R. Human metapneumovirus in infants and young children in Thailand with lower respiratory tract infections: molecular characteristics and clinical presentations. J Infect Dis 2006;52:254–63.

Sarasini A, Percivalle E, Rovida F, Campanini G, Genini E, Torsellini M, et al. Detection and pathogenicity of human metapneumovirus respiratory infection in pediatric Italian patients during a winter–spring season. J Clin Virol 2006;35:59–68.

Sasaki A, Suzuki H, Saito R, Sato M, Sato I, Sano Y, et al. Prevalence of human metapneumovirus and influenza virus infections among Japanese children during two successive winters. Pediatr Infect Dis J 2005;24:905–8.

Schildgen O, Jubbink MF, de VM, Pyrc K, Dijkman R, Simon A, et al. Identification of cell lines permissive for human coronavirus NL63. J Virol Methods 2006;138:207–10.

Semple MG, Collwell A, Dove W, Greensill I, McNamara PS, Hallihide C, et al. Dual infection of infants by human metapneumovirus and human respiratory syncytial virus is strongly associated with severe bronchiolitis. J Infect Dis 2005;191:382–6.

Shimizu C, Shike H, Baker SC, Garcia F, van der HL, Kuipers TW, et al. Human coronavirus NL63 is not detected in the respiratory tracts of children with acute Kawasaki disease. J Infect Dis 2005;192:1767–71.

Sloots TP, Mackay IM, Bialasiewicz S, Jacob KC, McQueen E, Harnett GB, et al. Human metapneumovirus, Australia, 2001–2004. Emerg Infect Dis 2006a;12:1263–6.

Sloots TP, McErlain PE, Speicher DJ, Arden KE, Nissen MD, Mackay IM. Evidence of human coronavirus HKU1 and human bocavirus in Australian children. J Clin Virol 2006b;35:99–102.

Snijder EJ, Bredenbeek PJ, Dobbe JC, Thiel V, Ziebuhr J, Poon LL, et al. Unique and conserved features of genome and proteome of SARS-coronavirus, an early split-off from the coronavirus group 2 lineage. J Mol Biol 2003;331:991–1004.

Sang A, Korn K, Wildner O, Uberla K. Characterization of virus isolates by particle-associated nucleic acid PCR. J Clin Microbiol 2005;43:716–20.

Stockton J, Stephenson I, Fleming D, Zambon M. Human metapneumovirus as a cause of community-acquired respiratory illness. Emerg Infect Dis 2002;8:907–911.

Tsang KW, Ho PL, Ooi GC, Yee WK, Wang T, Chan-Yeung M, et al. A cluster of cases of severe acute respiratory syndrome in Hong Kong. N Engl J Med 2003;348:1977–85.

Tyllrell DA, Bynoe ML. Cultivation of a novel type of common-cold virus in organ cultures. Br Med J 1965;1:1467–70.

Vabret A, Mourez T, Dina J, van der HL, Gouarin S, Petitjean J, et al. Human metapneumovirus NL63, France. Emerg Infect Dis 2005;11:1225–9.

Vabret A, Dina J, Gouarin S, Petitjean J, Corbet S, Freymuth F. Detection of the new human coronavirus HKU1: a report of 6 cases. Clin Infect Dis 2006;42:634–9.

van den Hoogen BG, de Jong JC, Groen J, Kuiken T, de GR, Fouchier RA, et al. A newly discovered human coronavirus isolated from young children with respiratory tract disease. Nat Med 2001;7:719–24.

van den Hoogen BG, van Doornum GJ, Fockens JC, Cornelissen JJ, Beyer WE, de GR, et al. Prevalence and clinical symptoms of human metapneumovirus infection in hospitalized patients. J Infect Dis 2003;188:1571–7.

van den Hoogen BG, Osterhaus DM, Fouchier RA. Clinical impact and diagnosis of human metapneumovirus infection. Pediatr Infect Dis J 2004;23:S25–32.

van der Hoek L, Pyrc K, Jubbink MF, Vermeulen-Oost W, Berkhout B, Wolthers KC, et al. Identification of a new human coronavirus. Nat Med 2004;10:368–73.

van der Hoek L, Pyrc K, Berkhout B. Human coronavirus NL63, a new respiratory virus. FEMS Microbiol Rev 2006a;30:760–73.

van der Hoek L, Sure K, Ihorst G, Stang A, Pyrc K, Jubbink MF, et al. Human coronavirus NL63 infection is associated with croup. Adv Exp Med Biol 2006b;581:485–91.

van Elden LJ, van Loon AM, van AF, Hendriksen KA, Hoepelman AI, van Kraaij MG, et al. Frequent detection of human coronaviruses in clinical specimens from patients with respiratory tract infection by use of a novel real-time reverse-transcriptase polymerase chain reaction. J Infect Dis 2004;189:652–7.

Vazov S, Ratjen F, Scheidhauer R, Fiedler M, Roggendorf M. High prevalence of human metapneumovirus infection in young children and genetic heterogeneity of the viral isolates. J Clin Microbiol 2003;41:3043–5.

Vicente D, Cilla G, Montes M, Perez-Yarza EG, Perez-Trallero E. Human bocavirus, a respiratory and enteric virus. Emerg Infect Dis 2007;13:636–7.

von Linstow ML, Larsen HH, Eugen-Olsen J, Koch A, Nordmann WT, Meyer AM, et al. Human metapneumovirus and respiratory syncytial virus in hospitalized Danish children with acute respiratory tract infection. Scand J Infect Dis 2004;36:578–84.
Wang D, Urisman A, Liu YT, Springer M, Ksiazek TG, Erdman DD, et al. Viral discovery and sequence recovery using DNA microarrays. PLoS Biol 2003;1:257–60.

Williams JV, Harris PA, Tollefson SJ, Halburnt-Rush LL, Pingsterhaus JM, Edwards KM, et al. Human metapneumovirus and lower respiratory tract disease in otherwise healthy infants and children. N Engl J Med 2004;350:443–50.

Williams JV, Martino R, Rabella N, Otegui M, Parody R, Heck JM, et al. A prospective study comparing human metapneumovirus with other respiratory viruses in adults with hematologic malignancies and respiratory tract infections. J Infect Dis 2005a;192:1061–5.

Williams JV, Tollefson SJ, Heymann PW, Carper HT, Patrie J, Crowe JE. Human metapneumovirus infection in children hospitalized for wheezing. J Allergy Clin Immunol 2005b;115:1311–2.

Williams JV, Wang CK, Yang CF, Tollefson SJ, House FS, Heck JM, et al. The role of human metapneumovirus in upper respiratory tract infections in children: a 20-year experience. J Infect Dis 2006;193:387–95.

Wolf DG, Zakay-Rones Z, Fadeela A, Greenberg D, Dagan R. High seroprevalence of human metapneumovirus among young children in Israel. J Infect Dis 2003;188:1865–7.

Woo PC, Lau SK, Chu CM, Chan KH, Tsoi HW, Huang Y, et al. Characterization and complete genome sequence of a novel coronavirus, coronavirus HKU1, from patients with pneumonia. J Virol 2005a;79:884–95.

Woo PC, Lau SK, Tsoi HW, Huang Y, Poon RW, Chu CM, et al. Clinical and molecular epidemiological features of coronavirus HKU1-associated community-acquired pneumonia. J Infect Dis 2005b;192:1898–907.

Woo PC, Lau SK, Yip CC, Huang Y, Tsoi HW, Chan KH, et al. Comparative analysis of 22 coronavirus HKU1 genomes reveals a novel genotype and evidence of natural recombination in coronavirus HKU1. J Virol 2006;80:7136–45.

Xepapadaki P, Psarras S, Bossios A, Tsolia M, Gourggiotis D, Liapi-Adamidou G, et al. Human Metapneumovirus as a causative agent of acute bronchiolitis in infants. J Clin Virol 2004;30:267–70.