Phytochemical and biological investigation of Indian medicinal plant of *givotia rotterifromis griff*

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Abstract: A study on *givotia rotterifromis griff* shows that studies on a tremendous of pharmacological and physiological activities and other applications. Moreover, *givotia rotterifromis griff* has diverse physiological effects. This tempted us to use the bark of *givotia rotterifromis griff* for extraction of new compound from the extract. Attempt is made to develop an efficient method for isolation of compounds from the barks of *givotia rotterifromis griff*. Our scheme of work is to separate a portion by treatment with pure methanol (CH₃OH) using soxhlet apparatus at boiling temperature and the sample is crystallized to separate to the pure compound. The compound was identified further by chromatographic studies, melting point determination, and spectral analysis (IR, 1D and 2D NMR) etc. Furthermore, antimicrobial studies are tested various antibacterial and antifungal species and also molecular docking studies are checked with HIV protease with Schrodinger 9.5 software.

Keywords: *Givotia rotterifromis griff*, Soxhlet apparatus, 1D and 2D NMR, Molecular docking studies.

1. Introduction

A natural product is a chemical compound or substance produced by a living organism that is found in nature. In the broadest sense natural products include any substance produced by life [1]. Natural products can also be prepared by chemical synthesis (both semi synthesis and total synthesis) and have played a central role in the development of the field of organic chemistry by providing challenging synthetic targets [2]. The term natural product has also been extended for commercial purposes to refer to cosmetics, dietary supplements, and foods produced from natural sources without added artificial ingredients [3].

Within the field of organic chemistry, the definition of natural products is usually restricted to mean purified organic compounds isolated from natural sources that are produced by the pathways of primary or secondary metabolism [4,5]. Within the field of medicinal chemistry, the definition is often further restricted to secondary metabolites [6,7] Secondary metabolites are not essential for survival, but nevertheless provide organisms that produce them an evolutionary advantage [8,9]. Many secondary metabolites are cytotoxic and have been selected and optimized through evolution for use as "chemical warfare" agents against prey, predators, and competing organisms [10-15].

In the present investigation new compound has been extracted from the *givotia rotterifromis griff* and screened for its chemical and pharmacological activities.

2. Experimental methods

2.1 Plant Material

This plant material is available in Tamil Nadu, Karnataka, Andhra Pradesh and Srilanka. The bark is collected from Kanjimalai (Namakkal). The barks used for the study were cleaned manually to free them from stones and other undesirable matter and then ground in an electrically operated grinder to fine particle size.

2.2 Chemicals

All chemicals were analytical grade expect petroleum ether (60-80°C), Ethyl acetate, methanol, Silica gel for TLC, Silica gel for Column and ethanol was of laboratory grades. Except ethanol all other solvents were purchased from Merck chemicals.
A Soxhlet extractor is a piece of laboratory apparatus invented in 1879 by Franz von Soxhlet. It was originally designed for the extraction of a lipid from a solid material. However, a Soxhlet extractor is not limited to the extraction of lipids. Typically, a Soxhlet extraction is only required where the desired compound has a limited solubility in a solvent, and the impurity is insoluble in that solvent. If the desired compound has a significant solubility in a solvent then a simple filtration can be used to separate the compound from the insoluble substance. Normally a solid material containing some of the desired compound is placed inside a thimble made from thick filter paper, which is loaded into the main chamber of the Soxhlet extractor. The Soxhlet extractor is placed onto a flask containing the extraction solvent. The Soxhlet is then equipped with a condenser. The solvent is heated to reflux. The solvent vapour travels up a distillation, and flows into the chamber housing the thimble of solid. The condenser ensures that any solvent vapour cools, and drips back down into the chamber housing the solid material.

The chamber containing the solid material slowly fills with warm solvent. Some of the desired compound will then dissolve in the warm solvent. When the Soxhlet chamber is almost full, the chamber is automatically emptied by a siphon side arm, with the solvent running back down to the distillation flask. This cycle may be allowed to repeat many times, over hours or days. During each cycle, a portion of the non-volatile compound dissolves in the solvent. After many cycles the desired compound is concentrated in the distillation flask. The advantage of this system is that instead of many portions of warm solvent being passed through the sample, just one batch of solvent is recycled. After extraction the solvent is removed, typically by means of a rotary evaporator, yielding the extracted compound. The non-soluble portion of the extracted solid remains in the thimble, and is usually discarded. The concentrate extract was involved to column chromatographic technique.

2.3 Chemicals Test
2.3.1 Test for Nitrogen element

The pure compound was taken for the analysis for the analysis of elements by standard methods and it was found that Nitrogen is present as a special element. A small quantity of the sample is added with sodium metal is taken in a fusion tube. It is made alkaline by adding 2-3 drops of sodium hydroxide (NaOH) solution. 1 mL of freshly prepared solution of ferrous sulphate is added to this solution. The mixture of the two solutions is boiled and then acidified with dilute sulphuric acid. The Prussian blue coloration of the precipitate is obtained. It confirms the presence of nitrogen in the compound.

2.4 Tests for Unsaturation

The test of unsaturation by alkaline KMnO4 method showed the presence of unsaturation in the compound. A small amount of the compound is treated with small quantity of sodium carbonate solution and very dilute solution of KMnO4. The pink colour is decolourised. It confirms the presence of unsaturation in the compound.

2.5 Column Chromatographic Analysis

In column chromatography the powdered adsorbent (silica gel) is placed in a vertical glass tube. The extract is added to the top of the column. A solvent (methanol) is then allowed to flow down the tube separation of the components of the extract takes place as a result single adsorption. The compound is visible, coloured band appear down the column. The flow of the solvent is continued until all the compound is eluted from the bottom. The pure single compound is obtained and dried.

2.6 Thin-Layer Chromatographic Analysis

Transfer a small portion of the purified sample and dissolve them in a few drops of ethyl acetate. Prepare a silica gel thin-layer chromatography plate for the spotting of sample. With the aid of a capillary, place a sample of the crude oil remaining from the extraction procedure, a sample of the filtrate from the isolation procedure, the sample prepared from the purified chemical compound, and a sample of a chemical compound standard on separate points of the thin-layer plate. Develop the plate using 3:2 Petroleum ether & ethyl acetate. Visualize under in an iodine chamber. A single spot is appeared in TLC.

2.7 Other Characterizations

Melting points were recorded with melting point apparatus Macro Scientific Works found to be 164°C. The percentage of yield is 30%. Thin Layer Chromatography (TLC) was performed on glass plates coated with silica gel 60 (E. Merck, India Ltd.) and column chromatography technique was isolated for pure single compounds with silica gel. FT IR spectrum was recorded employing Bruker FT-IR spectrometer. The 1H NMR and 13C NMR spectra of the compound were recorded using the AMX 500MHz Bruker spectrometer with DMSO as the internal standard reference. The 1H-1H COSY and 13C-1H COSY spectrum of the compound were recorded using the AMX 500MHz Bruker spectrometer with DMSO as the internal standard reference. In vitro antimicrobial activity was examined for various alcoholic extract of plant of the study species against four bacterial and four fungal species. Molecular docking studies of the isolated compound was docked in to the active site of
3. Results and Discussion

3.1 Spectral analysis

The *Givotia rottleriiformis* griff (Venthal tree) bark extract contains several important olefinic compounds. The olefinic compounds are important in motional biology. In the present work we have attempted to bark extract an important component from by solvent extraction procedure. In our experiment we have selected solvent such as methanol throughout the extraction process. Extracted product was isolated, purified and identified by TLC, column, FT-IR, $^1$H NMR, $^{13}$C NMR and COSY ($^1$H-$^1$H, $^{13}$C-$^1$H) NMR spectra.

3.2 FT-IR spectrum of the compound

The FT-IR spectrum of the compound is shown figure 1. IR data reveals that the compound consist cyclic ring, double bond, and hydroxyl and amine functional group. In this spectrum to given some important idea of the found the structure. The absorption frequency of the olefinic in conjugation with some functional group is also lower by the 400 cm$^{-1}$=1610 cm$^{-1}$ the intensity of absorption is increased on conjugation of an olefinic double bond the C=C stretch is lowered and the intensity of absorption increased. Resonance result in partial $\pi$ bond character between the two double bonds therefore vibrate at lower frequency these do similar isolated double bond. In the region of 3385 cm$^{-1}$ and 3237 cm$^{-1}$, we have to find out the main point of O-H stretching and N-H stretching vibration respectively. The N-H stretching of hydrogen bonded primary amine coupled doublet in broad peak rang 3237 cm$^{-1}$.

3.3 $^1$H and $^{13}$C -NMR Spectrum of the isolated compound

The $^1$H NMR (δ ppm) spectrum was recorded in 500 MHz Brucker instrument. The NMR result again confirms the purity of the compound. It is also indicating 10 proton of olefinic regions of 3 to 6 ppm. The proton NMR spectrum of compound (Figures 2, 3 & 4) as only ABX system for the hetero cyclic ring proton (H3a, H2a and H2e) and (H4a, H5a and H5e). Since no coupling partner is available at C3-C4 proton. The olefinic methyl analog indicated the axial position for the proton and equatorial orientation for the methyl protons groups. The signle at 4.4ppm with $^3$J value of 12Hz J(H5a,H4e & H4e, H5e) and J5e,4e-3 Hz is assigned to the axial proton at HC=CH and it conformed the equatorial orientation of C2 and C5 groups. The presence of N-H proton and O-H proton at 2.93 ppm and 4.6 ppm respectively. A mass spectrum of this compound the presence of at M$^+$ ion at m/e 99 and the fragmentation pattern conform the structure of the isolated compound. In the $^{13}$C NMR (δ ppm) spectrum (Figure 5) the ranges of carbon atom appear between 72.3 & 75.7ppm, suggesting only an olefinic unsaturated carbon atoms are present.

3.4 2D NMR (COSY)spectrum of the compound

The structure of the compound also confirmed by correlation spectra $^1$H-$^1$H COSY and $^1$H-$^{13}$C COSY (Figures 6, 7 & 8). Tentative proposed structure of the isolated compound. DEPT – 135 spectrums also to find out odd and even carbon to help of find the structure (Figures 9 & 10).

*Figure 1. FT-IR spectra of the N- cyclopent-1-eny1-hydroxylamine*
Figure 2. $^1$H NMR spectra of the N-cyclopent-1-enyl-hydroxylamine

Figure 3. $^1$H NMR spectra of the N-cyclopent-1-enyl-hydroxylamine

Figure 4. $^1$H NMR [Ex] spectra of the N-cyclopent-1-enyl-hydroxylamine
Figure 5 $^{13}$C-NMR spectra of the N-cyclopeny-1-enyl-hydroxylamine

Figure 6 $^{13}$C-NMR[Ex] spectra of the N-cyclopent-1-enyl-hydroxylamine

Figure 7 $^1$H- $^{13}$C COSY NMR spectra of the N-cyclopent-1-enyl-hydroxylamine
Figure 8 ¹H-¹H COSY NMR [Ex] spectra of the N-cyclopent-1-enyl-hydroxylamine

Figure 9 DEPT spectra of the N-cyclopent-1-enyl-hydroxylamine

Figure 10 DEPT [Ex] spectra of the N-cyclopent-1-enyl-hydroxylamine
3.5 Anti microbial activities

The dried leaf powder of (50g) was successively extracted in soxhlet apparatus using hexane, ethyl acetate and methanol (250mL of each). The extracts were dried under reduced pressure using rotary vacuum evaporator to get the crude and were stored below 4°C until further use.

In vitro antimicrobial activity was examined for various alcoholic extract of bark of the study species against four bacterial species and four fungus which included Staphylococcus aureus, Bacillus subtilis, Escherichia coli and Pseudomonas aeruginosa and Staphylococcus aureus, Bacillus subtilis, Escherichia coli and Pseudomonas aeruginosa species respectively. All these microbial strains were obtained from the Department of Biotechnology, Kongunadu Arts and Science College. The bacterial stock cultures were maintained on nutrient agar slants and fungal stock culture maintained on Potato dextrose agar slant at 4°C.

The isolated compound were screened for their in-vitro growth inhibitory action against different strains of bacteria viz., Staphylococcus aureus, Bacillus subtilis, Escherichia coli, Pseudomonas aeruginosa, the results are presented in Table - 1 and Figure 11. The compound shows a significant antibacterial activity against Staphylococcus aureus, Bacillus subtilis, Escherichia coli, Pseudomonas aeruginosa. All extract has been moderate activity compared to standard drugs of Tetracycline.

3.6 Anti-fungal activity

The in vitro antifungal activity of the compound and of tetracycles as the reference drug on a panel of fungi such as Aspergillus niger, A. flavus, A. baumanii, and Fusarium oxysporum measured as diameter of inhibition is provided in Table 2. Against all the fungal strains, isolated compound, exhibited antifungal activity against Fusarium oxysporum diameter of inhibition of 9.18± 0.68 in petroleum ether extract and methanal extract values 11.75±0.28. Of isolated compound extract are found to be potent against Fusarium oxysporum, and Aspergillus niger showing of 9.23± 0.49 & 15.79±0.66 µg/ml respectively, rest of the isolated compound extract were no effect potent against all the tested strains.

3.7 Molecular Docking studies

The crystal structure of HIV integrase was obtained from protein databank (PDB ID: 1HXW) with a resolution factor Å. Before docking, the crystal structure of the protein was cleaned by removing the water molecule, added the hydrogen atom to the protein structure for tautomeric and ionization states of amino acid residues using OPLS-2005 forcew field. Finally the protein structure energy was minimized reached until the average root mean square deviation non bonded hydrogen atom 0.30Å. The prepared protein were input file for molecular docking.

The HIV protease (PDB ID: 1HXW) was retrieved from protein data bank further taken to CASTp for binding site prediction which revealed different binding site. The best binding residues from the CASTp result given below. R8, 23L, 25D, 26T, 27G, 28A, 29D, 30D, V32, 47I, 48G, 49G, 50I, R8, P9, L10, L23, D25, T26, G27, A28, D29, D30, P31, V32, I47, 48G, R87.. The isolated compound was drawn using chem draw version . The ligand structure was further carried out into ligprep for ligand preparation version , Ligprep convert 3D structure from 2D structure, include different tautomers and ionized form at a pH range 7.0± 2.0. the prepared ligands were input file for molecular docking studies.

The isolated compound was docked in to the active site of the HIV protease (PDB ID: 1HXW) using Glide from Schrodinger 9.5. Grid was prepared for the HIV protease with the exact same center and the size of the bounding box set on 20 Å. The Glide algorithm was based on a systematic search of positions, orientations, and conformations of the ligand in the receptor binding site using funnel type approach.
The search begins with a rough positioning and scoring phase that significantly limits the search space and reduces the number of poses to be selected for minimization on the pre-computed OPLS-2005 van der Waals and electrostatic grids for the protein. The glide score and glide energy was analyzed using XP visualize Table-3.

3.8 Binding mode of isolated compound into HIV Protease

Docking simulation of isolated compound within the active site of the HIV Protease has been analyzed. The Glide Score and Glide Energy value for compound was -3.767Kcal/mol and -23.176Kcal/mol. Upon the examination of docking features between isolated compound and HIV protease, it was found only three hydrogen bond interaction with bond distance (2.46Å, 1.92Å, 2.16Å).

Table - 1 Antibacterial activity of certain alcoholic bark extracts of the species, *Givotia rotterifomis* Griff

| Bacteria                | Standard (Tetracycline) | Solvent extracts |
|-------------------------|-------------------------|------------------|
|                         |                         | Petroleum ether | Methanol | Water        |
| *Staphylococcus aureus* | 8.13±0.57               | 15.19±0.38       | 14.16±0.56 | 12.83±0.45  |
| *Bacillus subtilis*     | 9.03 ± 0.45             | 14.17±0.33       | 15.23±0.71 | 11.34±0.23  |
| *Escherichia coli*      | 22.43±0.38              | 10.71±0.55       | 9.05±0.33  | 10.56±0.33  |
| *Pseudomonas aeruginosa*| 23.03 ± 0.54            | 9.07±0.65        | 10.11±0.45 | 9.41±0.66   |

Table - 2 Antifungal activity of certain alcoholic bark extracts of the species *Givotia rotterifomis* Griff

| Fungi                  | Standard (Tetracycline) | Solvent extracts |
|------------------------|-------------------------|------------------|
|                        |                         | Petroleum ether | Methanol | Water        |
| *Aspergillus niger*    | 33.22±0.58              | 9.23±0.49        | 15.79±0.66 | 14.58±0.80  |
| *A. flavus*            | 35.17±0.45              | 10.13±0.25       | 14.18±0.63 | 10.17±0.48  |
| *A. baumanii*          | 36.68±0.40              | 8.38±0.48        | 12.45±0.33 | 12.85±0.78  |
| *Fusarium oxysporum*   | 34.25±0.87              | 9.18±0.68        | 11.75±0.28 | 12.69±0.27  |

Experiments were performed in triplicates and represented as mean±standard deviation (SD).
The sidechain hydrogen atom of the negative charged residue of ASP 30 were strongly interacted with oxygen atom of the compound 4, compound 34 were strongly interacted with the negative charged residue of ASP 30. Furthermore ILE 47, ALA 28, ILE 50, LEU 76 a number of hydrophobic interactions was bound between compound 4 into IV Protease (Figure 12a-b).

![Figure 12 (a) 3-Dimensional Structure of Target Protease with protein HIV Protease (PDB ID: 1HXW) (b) Docked structure of target protein HIV with N-cyclopent-1-enyl-hydroxylamine](image)

**Table-3** Glide extra precision (XP) results isolated molecules by use Schrodinger-9.5

| Compound Name                        | Glide Score | Glide Energy | No of H bond Interactions | Interacting Residue | Distance (Å) |
|--------------------------------------|-------------|--------------|---------------------------|---------------------|--------------|
| Givotia rottleriformis griff         | -3.767      | -23.176      | 3                         | ILE 50              | 2.46 1.92    |
|                                      |             |              |                           | ILE 50              | 2.16         |

4. Conclusion

In conclusion, this study is a complementary survey to phytochemical and spectral studies carried out upon N-cyclopent-1-enyl-hydroxylamine. This project work involves bark extraction of the givotia rottleriformis griff with petroleum ether from which a pure component has been identified by crystallization with benzene. TLC report also gives the indication of a pure component. Melting point determination, IR, 1H-NMR, 13C-NMR and 2D-NMR spectral techniques identify the purity of the compound and also the spectral data’s indicate that the compound is tentatively N-cyclopent-1-enyl-hydroxylamine. Further pharmacological studies like Anti-microbial, and anti-fungal activities studies are required to find out the complete physiological and pharmacological action of the compound.

So that identified compound may be correctly predicted as N-cyclopent-1-enyl- hydroxylamine. Thus, our method, reported here is found to give moderate yield of a compound likely to be N-cyclopent-1-enyl-hydroxylamine of fairly pure quality. This is a very rapid, straight forward and new innovative method of extraction of the olefilic compounds, N-cyclopent-1-enyl- hydroxylamine from the bark of givotia rottleriformis griff. Docking simulation of isolated compound within the active site of the HIV Protease has been analyzed. The Glide Score and Glide Energy value for compound was -3.767Kcal/mol and -23.176Kcal/mol.
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Conflict of interest
None of the authors have any conflicts of interest to declare.

Author’s contribution
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