Human Placentae in Pregnancy Induced Hypertension - A Histologic and Morphometric Analysis

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Abstract

Introduction: Placenta is the mirror of maternal and fetal status. It has been called a pregnancy diary that permits the clinician to study the intra uterine environment of the fetus and some of the fetal responses to disease. Reduced maternal utero placental blood flow leading indirectly to constriction of foetal stem arteries has been associated with the changes seen in the placentae of pre-eclamptic women. Maternal vasospasm leads to fetal hypoxia, causing fetal distress and death. Histopathological changes related to confined placental mosaicism may be associated within adequate placentation and retroplacental ischemia.

Aim: To study morphometry and histological changes in placentae of women with pregnancy induced hypertension (PIH) and to observe the perinatal outcome of such pregnancies.

Materials: Total number of 50 placentae were used for the present study, out of which 25 placentae were obtained from women with PIH and 25 placentae from women who were normal having uncomplicated full-term deliveries.

Results: The mean weight of the placenta was 470.04g in control group and 385.70g in PIH group. Histopathological examination of the placental PIH showed increase in number of syncytial knots and areas of fibrinoid necrosis.

Conclusion: PIH has remarkable influence on morphology and histology of the placenta, and thereby affects the growth of the fetus. The changes caused by placental insufficiency has an adverse effect on the neonatal birth weight due to compromised utero-placental blood flow.

Keywords: Feto-Placental Ratio; Perinatal Outcome; Vasospasm; Birth Weight; Histopathological.

Abbreviations: PIH: Pregnancy Induced Hypertension; H&E: Haematoxylin & Eosin; ISSHP: International Society for the Study of Hypertension in Pregnancy; DBP: Diastolic Blood Pressure; FP: Feto-Placental.

Introduction

The placenta is an organ that connects the developing fetus to the uterine wall to allow nutrient uptake, provide thermoregulation to the fetus, waste elimination, and gas exchange via the mother’s blood supply, fight against internal infection and produce hormones to support pregnancy. Placenta is a vital organ for fetal development and is a mirror of maternal and fetal status. Complications in pregnancy like gestational diabetes or hypertension are reflected in the placenta in a conspicuous way [1]. According to WHO systematic review, hypertensive disorders of pregnancy contribute to 16% of maternal mortality, which is more than other leading causes namely hemorrhage (13%), abortion (8%) and sepsis (2%) [2]. Hypertensive disorders are also the cause of fetal and neonatal mortality and morbidity. Hypertensive disorders complicate about 5-10% of all pregnancies. Researchers suggest that there is reduction in surface area of placenta in pregnancy induced hypertension (PIH). These changes have been attributed to constriction of the fetal stem arteries due to decreased utero-placental blood flow [1].

The placenta develops from 2 sources - the chorion frondosum (fetal component) and the decidua basalis (maternal component).
Placenta helps in exchange of gases, passage of nutrients and IgG antibodies from maternal to fetal side and passage of waste products from fetal to maternal side. It also produces hormones like human placental lactogen, human chorionic gonadotropin, estrogen, progesterone and relaxin. On gross examination, a normal full term placenta appears disc shaped. It weighs around 500-600g with a diameter of 15-20cm [3]. The placenta shows 2 surfaces-maternal surface with a rough cobblestone appearance due to presence of 15-20 cotyledons separated by grooves occupied by incomplete placental septa and a fetal surface which is smooth and transparent as it is covered by amnion. The umbilical cord is normally attached to the center of fetal surface [3].

**Aim**

1. To study the morphometry and histological changes in placentae of women with pregnancy induced hypertension (PIH).
2. To observe the perinatal outcome of such pregnancies, compare the results with those of normal pregnancies.

**Materials and Methods**

The study was conducted on 50 placentae obtained from the Department of Obstetrics and Gynecology after attaining due consent from patients and approval from the Institutional Ethics Committee (IEC743/2016).

**Inclusion Criteria**

25 placentae were obtained from women diagnosed with PIH and 25 placentae from women who were having normal uncomplicated full-term deliveries.

**Exclusion Criteria**

Patients suffering from chronic hypertension, renal failure, cardiac problems, gestational diabetes mellitus and multiple pregnancy were excluded.

The collected placentae were squeezed to evacuate the blood and washed under running tap water to remove clots. The membranes were trimmed and the site of insertion of the umbilical cord was noted. The diameter, the weight and the central thickness of placenta was measured. The diameter of the placentae were calculated by taking the mean value of the shortest & longest diameter of placenta. The placent all surface area was calculated using the formula 1.

\[ A = \pi d_1 d_2 /4 \]

Where:

- \( A \) = placental surface area
- \( d_1 \) = minimum diameter (cm)
- \( d_2 \) = maximum diameter (cm)

The number of cotyledons were counted from maternal side. The feto-placental ratio was calculated after obtaining weight of the baby. Data analysis was done by using Statistical Package for Social Sciences (SPSS) 19 version, and \( P<0.05 \) was considered statistically significant.

Two tissue sections were taken from each placenta-one from the center and one from the edge. The tissue sample was fixed in formalin fluid, processed to prepare paraffin embedded blocks and 5-micron thick sections were cut. The slides are stained with Haematoxylin & Eosin (H&E) stains. The stained sections were observed and photographed using Olympus BX43 microscope.

**Results**

In the present study the placental weight (380.70gm), placental diameter (16.04cm) and placental surface area (201.43cm²) was comparatively less in the PIH group in comparison to the control group. It was also observed that the birth weight was less in the PIH group (Table 1). The site of attachment of the umbilical cord was observed to be eccentric both in case of the PIH placentae as well as the control group (Table 2).

Histopathological examination of the placenta in PIH showed increase in number of syncytial knots, villous edema and areas of fibrinoid necrosis (Figure 1).

### Table 1. The mean and standard deviation of various parameters of the placentae measured in the control group and PIH group.

| Parameter         | Control (n=25) | PIH (n=25) | p-value |
|-------------------|----------------|------------|---------|
| Placental weight (gm) | 470.04 ± 53.07 | 380.70 ± 47.25 | 0.017   |
| Placental diameter (cm) | 18.67 ± 4.85  | 16.04 ± 0.14  | 0.031   |
| Placental surface area (cm²) | 267.53 ± 59.17 | 201.43 ± 42.61 | 0.298   |
| Placental central thickness (cm) | 1.90 ± 0.32  | 1.53 ± 0.15  | 0.354   |
| Placental cotyledons | 14.73 ± 2.53  | 14.23 ± 1.41  | 0.130   |
| Baby’s birth weight (gm) | 2684.47 ± 514.03 | 2138 ± 438.50 | 0.057   |
| Feto-Placental weight ratio | 5.87 ± 0.93   | 5.56 ± 0.42   | 0.207   |

### Table 2. The incidence of site of attachment of umbilical cord in the control group and PIH group.

| Parameters       | Control | PIH     | p-value |
|------------------|---------|---------|---------|
| Centric          | 8 (32%) | 10 (40%) | 7 (28%) |
| Eccentric        | 10 (40%) | 11 (44%) | 4 (16%) |
| Marginal         | 7 (28%) | 4 (16%) |         |
Discussion

Hypertensive disorders are seen in 5-10% of all pregnancies [2]. According to the International Society for the Study of Hypertension in Pregnancy (ISSHP), hypertension is defined as a diastolic blood pressure (DBP) > 90mm Hg or Systolic blood pressure (SBP) >140mm Hg on at least 2 occasions taken 6 hours apart. In normal pregnancy, the DBP begins of all in early pregnancy and continues of all in second trimester or each anadirat 22-24 weeks. Then it steadily rises to reach prepregnant levels by term. This fall is due to reduced vascular tone, which leads to peripheral vasodilation. Pregnancy may aggravate hypertension in women, who are already hypertensive and may also induce hypertension in those who are not motensive before they became pregnant. 3 grades of PIH are recognized: gestational hypertension, pre-eclampsia and eclampsia [1]. What ever the cause of hypertension, the potential complications are placental insufficiency, fetal growth restriction, preterm labour, abrupton placenta, renal changes and maternal convulsions [2, 4].

The mean weight of the placenta was 470.04gm in control group and 385.70g min PIH group (Table 5). According to Dutta and Dutta as grade of PIH increases, there is decrease in the weight of placenta [5]. Placental weight affects the fetal weight due to its relation with the villous surface area. Sirpurkar et al., observed that placental weight was quite low in PIH group than normal [13]. Placental weights show great variations which has been attributed to differences in genes regulating placental and fetal weight [6].

The mean placental diameter in control group and PIH group was 19.45 ± 6.53cm and 16.08 ± 4.63cm respectively (Table 1). The values in control group were 21.225 ± 1.385cm and 14.208 ± 1.2103cm in PIH group. Sultana et al., and Ashfaq et al., observed differences in placental weight and diameter between PIH group and controls, but these differences were not significant [8, 9]. The mean placental thickness was recorded as 1.96 cm and 1.57cm in control group and PIH group respectively. In preterm infants, thickness and weight of placenta vary directly with duration of pregnancy [1, 4].

The fetus derives its nutrition from the maternal blood. Fetal growth thus depends on the placental surface area through which diffusion of nutrients occurs from them aternal blood to the fetal circulation through the placental barrier. In PIH group, placental surface area was found to be 201.43 ± 42.61cm² which was less than the mean placental surface area in the normal placentae (Table 1). However the difference was not found to be significant. In the present study, the number of cotyledons in the placentae from PIH group were found to be less than that of the control group (Table 1), similar to her studies [8, 10] however the difference between controls and cases was not found to be significant, similar to results obtained by Manjunath et al., and Mandhana et al., [11, 12].

In our study the mean birth weight of babies was 2138 ± 438.50g min PIH group and 2684.47 ± 514.03g min control group (Table 3). According to one study placental weight and the placental decidus area have a definite influence on the fetal birth weight [14]. Some studies have shown that size and weight of placenta is directly proportional to the birth weight of babies [1]. This may be due to altered intra cotyledonous vasculature seen in hypertension [15]. The site of attachment of umbilical cord did not show any significant differences between PIH group and control group in our study (Table 2).

Maternal vasospasm in PIH leads to decreased utero-placental blood flow. Fetal complications in PIH include intra uterine death, intra uterine growth retardation (due to chronic placental insufficiency), asphyxia and prematurity. The fetal risk in PIH is related to severity of pre-eclampsia, duration of disease and degree of protein uria. Metabolic abnormalities such as impaired glycolytic pathway, poor utilization of glucose and decrease in oxygen consumption have been observed in placentae obtained from PIH pregnancies [1].

The feto-placental weight (F/P) ratio was 5.56 ± 0.42 and 5.87 ± 0.93 in PIH group and control group respectively (Table 4). Macpherson opined that F/P ratio can be used for evaluation of placental weight deviation [16]. According to some studies if fetus is small, the placenta will also show diminished growth a sitisa fetal organ [17].

Histopathological examination of the placenta in PIH showed increase in number of syncytial knots and areas of fibrinoid necrosis. Areas of calcification and hyalinized villi were observed along with a cute atherosis of decidual arteries. According to Foxetal, reduced utero-placental blood flow causes decrease in villous perfusion leading to excess syncytial knot formation and stromal fibrosis [18].

Studies have reported that hyalinization, endothelial proliferation of arteries and fibrinoid necrosis illustrate placental mosaicism and presumably the after math of PIH [19, 20]. Plausibly this may cause placental insufficiency leading to fetal complications encountered in PIH.
Table 3. Fetal birth weight (gm) comparison with previous studies.

| Studies | Fetal birth weight (gm) |
|---------|-------------------------|
| Present study | 2138 ± 562.50 |
| Keche et al., [2015] [1] | 2140.00 ± 439.60 |
| Londhe and Mane [2011] | 2.26 |
| Majumdar et al., [2005] [11] | 2.04 ± 0.48 |
| Udainia and Jain [2001] [20] | 2.2 |

Table 4. Feto-placental weight ratio comparison with other studies.

| Studies | F/P ratio | Mean ± SD |
|---------|-----------|-----------|
| Present study | 5.56 ± 0.42 |
| Nag U et al., [2013] [15] | 6.02 ± 0.36 |
| Sirpurkar et al., [2015] [13] | 5.70 |
| Usman et al., [2015] [4] | 5.21 ± 0.85 |
| Ranga, et al., [2017] [10] | 5.5 ± 1.9 |
| Keche et al., [2015] [1] | 5.51 ± 0.18 |

Table 5. Placental weight (gm) comparison with other studies.

| Studies | Placental weight (gm) |
|---------|-----------------------|
| Present study | 380.70 ± 47.25 |
| Ranga, et al., [2017] [10] | 455.7 ± 59.9 |
| Keche et al., [2015] [1] | 387.00 ± 72.54 |
| Nag U et al., [2015] [15] | 404 ± 42 |
| Sirpurkar et al., [2015] [13] | 398 ± 98 |
| Usman et al., [2015] [4] | 397.08 ± 89.32 |

Conclusions

The high incidence of pregnancy induced hypertension and its adverse effects on the placental morphology which directly affects the growth and nutrition of fetus in utero is of significant concern. Among the other morbidities of pregnancy such as hemorrhage and infection; PIH plays a crucial role. The complications as seen in this study not only affect macroscopically but also microscopically.

References

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