DOG1-Positive Primary Mesenteric Leiomyosarcoma: Report of a Case and Review of the Literature

Nektarios Koufopoulos 1, Vasileia Damaskou 1, Vasiliki Siozopoulou 2, Panagiotis Kokoropoulos 3, Alin Roxani Gouloumis 4, Nikolaos Arkadopoulos 3, Ioannis G. Panayiotides 3

1. Pathology, Attikon University Hospital, National and Kapodistrian University of Athens, Athens, GRC. 2. Pathology, Antwerp University Hospital, Edegem, BEL. 3. Surgery, Attikon University Hospital, National and Kapodistrian University of Athens, Athens, GRC. 4. Pathology and Laboratory Medicine, Attikon University Hospital, National and Kapodistrian University of Athens, Athens, GRC.

Corresponding author: Nektarios Koufopoulos, koufonektar@yahoo.com

Abstract

The mesentery constitutes a common location for the metastatic spread of malignant gastrointestinal tumors. Primary mesenteric tumors, on the other hand, are very rare; lymphomas are the most common, followed by benign and malignant mesenchymal tumors. We present a case of a 43-year-old patient operated on for a primary mesenteric leiomyosarcoma with a positive immunostain for DOG1, despite having no KIT or PDGFRα mutations on molecular analysis. Moreover, we review the pertinent literature.

Categories: Pathology, General Surgery, Oncology
Keywords: dog1, immunohistochemistry, mesentery, leiomyosarcoma, gastrointestinal stromal tumor

Introduction

Leiomyosarcoma (LMS) is a malignant mesenchymal tumor originating from smooth muscle cells or from their precursor mesenchymal stem cells [1]. It usually arises in soft tissue, gastrointestinal tract, retroperitoneum, medium or large veins, and uterus [2,3]. Mesenteric LMS is a rare entity with a reported incidence of 1:350,000 [4], with only 20 immunohistochemically confirmed cases being reported. It is believed to arise from smooth muscle cells of the mesenteric blood vessel wall [2]. It is more often located in the small intestinal mesentery, followed by the transverse and sigmoid mesocolon or the gastrohepatic ligament [2]. The differential diagnosis primarily includes a mesenteric gastrointestinal stromal tumor (GIST). Since treatment and prognosis differ, CD117/KIT and DOG1 immunopositivity are the gold standards for a GIST diagnosis. Nevertheless, we report a DOG1 immunopositivity mesenteric LMS case, the first-ever reported to our best knowledge.

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Case Presentation

A 43-year-old patient was admitted in March 2020 due to a sizeable mesenteric mass discovered during a previous myomectomy for uterine leiomyomas (performed elsewhere) in October 2019. History was significant for cervical conization due to high-grade squamous intraepithelial lesion (HGSIL) fibroids. On laboratory investigation, inflammatory response markers and cancer tumor markers were within normal limits. The patient underwent a magnetic resonance imaging that revealed a mass with dimensions 4.7x4x3.3 cm that probably originated from the bowel wall or its mesentery with intermediate signal intensity in sequences T1 and T2 and intense inhomogeneous contrast enhancement. The patient was discussed in our local multidisciplinary tumor board (MDT), and the decision was made to proceed with surgical excision of the lesion. We performed an open excision of a mass that arose from the mesentery of the ileum. The tumor was excised with wide margins; it was of whitish color and firm texture. On microscopic examination, the tumor was seen to consist of fascicles of moderately pleomorphic, atypical spindle cells (Figure 1A) with up to eight mitoses per 10 high power fields (Figure 1B, 1C). Immunohistochemically, tumor cells stained positive for smooth muscle actin (SMA) (Figure 1D), desmin (Figure 1E), and DOG1 (Figure 1F), whereas they were negative for CD34 and S100 protein. The proliferation index assessed through immunostaining for Ki67 was estimated as 1%.

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FIGURE 1: The tumor consisted of atypical spindle cells arranged in fascicles with moderate pleomorphism 10x H&E (A). Up to eight mitoses per 10 high power fields (red arrows) were found, 40x H&E (B,C). Immunohistochemical study showed uniform positivity for SMA, 40x SMA (D), desmin, 40x desmin (E), and DOG1, 40x DOG1 (F)

H&E: hematoxylin and eosin stain; SMA: smooth muscle actin

Molecular analysis (performed at the Department of Pathology, Antwerp University Hospital, Edegem, Belgium) disclosed no mutations of KIT or PDGFRA genes. Based on the above findings, a diagnosis of malignant mesenchymal tumor consistent with low-grade leiomyosarcoma of the mesentery was rendered. A review of slides from the previous myomectomy specimens confirmed the diagnosis of leiomyomas.

A mass within the transversus abdominis muscle was found four months later during a postoperative check-up. Microscopic examination of the surgical specimen disclosed long, sweeping fascicles of uniform spindle cells with minimal cytologic atypia and low mitotic activity (one mitosis per 10 high-power field (HPF)) set within a collagenous stroma (Figure 2A, 2B). Tumor cells were diffusely positive for SMA (Figure 2C), as well as focally for muscle-specific actin (MSA) and desmin (Figure 2D); moreover, they showed diffuse cytoplasmic and focal nuclear β-catenin expression (Figure 2E), whereas they did not stain for either CD117 or DOG1. The proliferation index (assessed as the percentage of Ki67-positive cells) was estimated at around 1% (Figure 2F).
A diagnosis of abdominal fibromatosis/desmoid was therefore rendered. The MDT decided not to administer adjuvant therapy. At the two-year follow-up since surgical excision, the patient is free from any recurrence of either tumor or metastatic leiomyosarcoma.

Discussion
In contrast to metastases from gastrointestinal tumors, primary mesenteric tumors are rare, lymphomas being the most common, followed by usually benign mesenchymal tumors [2]. Mesenteric leiomyosarcoma is very rare, with only 20 cases reported [2-18]; six were male and 14 female. Age ranged from 13 to 86 years (median 54 years). Tumor size ranged from 5.3 to 36 cm (median 13.7 cm). Symptoms occur late in the disease course and are non-specific, usually a palpable mass, abdominal distention, and/or pain. Less commonly, abscess, acute intraperitoneal hemorrhage, or obstructive ileus may occur [2]. One patient presented with posterior reversible encephalopathy syndrome [15]. The liver was the most frequent location of metastatic disease. All cases were treated surgically for the primary and five for synchronous or metachronous metastatic lesions. Adjuvant chemotherapy was administered to eight patients.

In seventeen cases, follow-up was available, ranging from 3 to 59 months (median, 21 months). Eight patients died during the follow-up period, usually due to local recurrence or metastasis to the liver and lung. Two patients were alive with disease, and seven were alive with no evidence of disease. The demographic and clinical data are summarized in Table 1.
The management of LMS of the mesentery depends largely on preoperative imaging [2]. On CT, most solid lesions are neoplastic compared to cystic ones that are typically benign [19]. However, Dalal et al. have reported a case of entirely cystic LMS [2]. Radiologically, a helpful clue to differentiate GIST from sarcomas is the former’s less invasive appearance [20]. Percutaneous biopsy tends to be avoided since it may cause skin metastasis and peritoneal dissemination [6]. Thus, precise preoperative diagnosis based just on clinical and radiological findings is difficult [2]. The final and accurate diagnosis can be made only by the pathological examination of the surgical specimen.

In our case, we used an immunohistochemical panel consisting of SMA, desmin, CD117, and DOG1 to confirm the diagnosis. SMA and desmin staining are positive in almost all smooth muscle tumors compared to 31% and 4% of GISTs [21]. Both CD117 and DOG1 are considered relatively sensitive and specific markers for GIST diagnosis [22,23]. Focal DOG1 staining has been reported in almost 50% of the gastrointestinal ileum [22,23]. Therefore, SMA and desmin are useful for GIST diagnosis.

### TABLE 1: Demographic and clinical data of patients with mesenteric LMS.

| Case | Authors et al. | Year | Age | Gender | Symptoms | Size (cm) | Tumor location | Metastases | Chemo | Outcome (mo) |
|------|----------------|------|-----|--------|----------|-----------|---------------|------------|-------|--------------|
| 1    | Mettinen et al. [10] | 1999 | 41  | M      | N/M      | N/M       | Liver         | N/M        | N/M   | 12 AWD       |
| 2    | Mettinen et al. [10] | 1999 | 86  | M      | N/M      | N/M       | Liver         | N/M        | N/M   | 3 DOD        |
| 3    | Fukanaga et al. [11] | 2004 | 62  | F      | Mass     | 14        | Sigmoid colon | Liver      | Yes   | 10 DOD       |
| 4    | Simonovitch et al. [7] | 2006 | 82  | F      | Mass, pain | 11        | Small intestine | Liver      | No    | 24 Alive     |
| 5    | Iwasaki et al. [12] | 2010 | 13  | M      | Mass     | 10        | Small intestine | No         | No    | 3 ANED       |
| 6    | Koczowska et al. [8] | 2013 | 62  | F      | No       | 7.8       | Small intestine | Liver, LR  | Yes   | 24 DOD       |
| 7    | Koczowska et al. [8] | 2013 | 46  | M      | Mass     | N/M       | Sigmoid colon | Liver, LR  | Yes   | 58 DOD       |
| 8    | Mizobe et al. [13] | 2013 | 65  | M      | Mass, pain | 20        | Ascending colon | Liver, LR  | Yes   | 18 DOD       |
| 9    | Sidhic et al. [5] | 2015 | 33  | M      | Pain     | 15        | Small intestine | No         | N/M   | N/M          |
| 10   | Hamed et al. [9] | 2015 | 49  | F      | N/M      | N/M       | Small intestine | Liver      | N/M   | 59 AWD       |
| 11   | Hamed et al. [9] | 2015 | 46  | M      | N/M      | N/M       | Rectum        | Liver, lung | N/M   | 26 DOD       |
| 12   | Dasgupta et al. [2] | 2016 | 62  | F      | Fullness | 22        | Small intestine | No         | No    | 6 ANED       |
| 13   | Kato et al. [3] | 2016 | 76  | F      | Mass     | 14        | Descending colon | Liver      | No    | 40 ANED      |
| 14   | Varghese et al. [6] | 2016 | 40  | F      | Mass, Pain | 11        | Descending colon | Liver, lung | Yes   | 39 ANED      |
| 15   | Ilias et al. [4] | 2017 | 45  | F      | Mass     | 20        | Sigmoid colon | No         | No    | 9 ANED       |
| 16   | Dalal et al. [1] | 2017 | 50  | F      | Mass     | 15        | Small intestine | No         | No    | 6 ANED       |
| 17   | Schoucari et al. [14] | 2018 | 52  | F      | Pain, PRESS | 10        | Sigmoid colon | Liver, lung, LR | Yes   | 20 DOD       |
| 18   | Yoon et al. [15] | 2019 | 62  | F      | Mass, Pain | 25        | Descending colon | Omentum, peritoneum | Yes   | N/M          |
| 19   | Affas et al. [16] | 2020 | 68  | F      | Pain     | 5.7       | Ascending colon | LR         | N/M   | 12 DOD       |
| 20   | Deshmukh et al. [17] | 2021 | 31  | F      | Mass     | 36        | Descending colon | No         | Yes   | 15 ANED      |
| 21   | Present case | 2021 | 43  | F      | No       | 5.3       | ileum         | No         | No    | 24 ANED      |
stained. Uterine leiomyomas in the retroperitoneum and peritoneal leiomyomatosis were also focally DOG1 positive to a lesser degree (5/42 and 4/17 cases, respectively) [24]. Focal or diffuse DOG1 positivity has been previously reported in 27% of uterine LMS [25].

However, DOG1 expression has not been previously reported in a mesenteric LMS. In our case, uniform positive staining for SMA, desmin, and DOG1 was confusing. Despite the lack of CD117 immunopositivity, a GIST diagnosis could not be ruled out since DOG1 is a more sensitive marker than CD117 [26]. We, therefore, proceeded with molecular analysis, which showed no KIT or PDGFRα mutations, thus favoring the diagnosis of mesenteric LMS since only a small fraction (10-15%) of GISTs are devoid of these mutations [27].

Regarding prognosis, LMS of the mesentery has an aggressive clinical course with an unfavorable prognosis [2]. The overall five-year survival rate for abdominal LMS is only between 20% and 30% [19]. Earlier detection by ultrasonography and CT and complete surgical resection with tumor-free margins may improve long-term prognosis [6]. However, early detection is unusual due to the late appearance of symptoms. In several cases, LMS is detected after distant metastases occur [28].

Tumor recurrence can occur within five years in approximately 50% of patients despite adequate local control [29]. Careful postoperative follow-up is needed to detect a recurrence or metastasis for at least five years, with particular attention to the digestive tract, liver, and lung [6,7].

There is no clear precedent for treating mesenteric LMS, probably due to the small number of cases reported [6]. Treatment options depend upon the stage of presentation. For localized tumors, the treatment of choice is surgical excision with a wide margin of healthy tissue to achieve negative margins. Radiotherapy improves local control and preserves function, reduces local recurrence, but does not improve survival [1]. Metastatic disease is considered incurable since adjuvant chemotherapy or radiotherapy are ineffective [7,8].

Conclusions
Mesenteric LMS is an infrequent entity. Its diagnosis requires a combination of appropriate morphology and immunohistochemistry. As evidenced by the case we present, molecular analysis is necessary to make the correct diagnosis when morphological and immunohistochemical findings are inconclusive.

Additional Information
Disclosures
Human subjects: Consent was obtained or waived by all participants in this study. Conflicts of interest: In compliance with the ICMJE uniform disclosure form, all authors declare the following: Payment/services info: All authors have declared that no financial support was received from any organization for the submitted work. Financial relationships: All authors have declared that they have no financial relationships at present or within the previous three years with any organizations that might have an interest in the submitted work. Other relationships: All authors have declared that there are no other relationships or activities that could appear to have influenced the submitted work.

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