Predicting age and clinical risk from the neonatal connectome

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Abstract

The development of perinatal brain connectivity underpins motor, cognitive and behavioural abilities in later life. With the rise of advanced imaging methods such as diffusion MRI, the study of brain connectivity has emerged as an important tool to understand subtle alterations associated with neurodevelopmental conditions. Brain connectivity derived from diffusion MRI is complex, multi-dimensional and noisy, and hence it can be challenging to interpret on an individual basis. Machine learning methods have proven to be a powerful tool to uncover hidden patterns in such data, thus opening an opportunity for early identification of atypical development and potentially more efficient treatment.

In this work, we used Deep Neural Networks and Random Forests to predict neurodevelopmental characteristics from neonatal structural connectomes, in a large sample of neonates (N = 524) derived from the developing Human Connectome Project. We achieved a highly accurate prediction of post menstrual age (PMA) at scan on term-born infants (Mean absolute error (MAE) = 0.72 weeks, $r = 0.83$, $p<0.001$). We also achieved good accuracy when predicting gestational age at birth on a cohort of term and preterm babies scanned at term equivalent age (MAE = 2.21 weeks, $r = 0.82$, $p<0.001$). From our models of PMA at scan for infants born at term, we computed the brain maturation index (i.e. predicted minus actual age) of individual preterm neonates and found significant correlation of this index with motor outcome at 18 months corrected age. Our results suggest that the neural substrate for later neurological functioning is detectable within a few weeks after birth in the structural connectome.
1. INTRODUCTION

Magnetic Resonance Imaging (MRI) has provided a broad range of in vivo insights about the structure and function of the human brain. Diffusion MRI in particular, enables the characterization of microstructural changes in orientation and structure of white matter, opening the possibility to study brain structural connectivity and allowing the systematic description of whole-brain structural networks: the human connectome (Honey et al., 2010; Sporns et al., 2005).

During the perinatal period, the brain undergoes significant changes and consolidation of its structural connectivity, which underpins the expansion of motor, cognitive and behavioural abilities (Johnson, 2001). Since the inception of connectomics (Hagmann 2005; Sporns, Tononi, and Kötter 2005) several studies have tried to characterise early development of the structural connectome (Fan et al. 2011; Hagmann et al. 2010). Subtle alterations in the development of brain connectivity have been suggested to underlie atypical neurodevelopmental outcome in populations with perinatal risk factors, such as children born preterm (Battelle et al., 2018). This is of clear significance as preterm birth comprises approximately 11% of all births, and is the main global cause of death and disability in children under 5 years of age (Blencowe et al., 2012), as well as representing one of the most pervasive perinatal risk factors for atypical neurodevelopment (Wood et al., 2000). It has been associated with an increased risk of developing neurodevelopmental conditions such as motor, visuospatial and sensorimotor delay (Marlow et al., 2007), inattention, anxiety and social difficulties (Johnson and Marlow, 2014), autism spectrum (Johnson et al., 2010), cerebral palsy (Marlow et al., 2005) or psychiatric disorders in adulthood such as depression and bipolarity (Nosarti et al., 2012).

With the increasing interest in detailed study of neonatal brain connectivity, projects such as the developing Human Connectome Project (dHCP) have arisen which allow the development of bespoke methods to study the brain during this crucial period. The dHCP is an open science project which provides a large normative sample of neonatal structural, diffusion and functional MRI data with high spatial, angular and temporal resolutions. Features of this project include: advances in hardware (Hughes et al., 2017) and protocols for neonatal diffusion MRI acquisition (Hutter et al. 2018); the use of multiband techniques to accelerate acquisition time combined with approaches to correct motion (Cordero-Grande et al. 2016; Cordero-Grande et al. 2019); and the development of state-of-the-art preprocessing pipelines for neonatal MRI data (Bozek et al., 2018; Christiaens et al. 2018; Bastiani et al. 2019; Fitzgibbon et al. 2020; Makropoulos et al., 2018). These have together significantly improved neonatal MRI acquisition methods and data quality.

Despite this progress, studying the neonatal connectome remains challenging. Indeed, many methodological issues hamper the interpretation of the connectome (Sporns, 2013) including the difficulty of detecting origins and termination of connections (Jbabdi and Johansen-Berg, 2011) or a high number of false positive streamlines (Maier-Hein et al., 2017). Furthermore, the high dimensionality of the data and low number of scans usually available in neonates make underlying patterns very difficult to uncover.
Recent progress in machine learning offers a powerful alternative to study the structural connectome. Specifically, deep neural networks, which are known for their ability to model complex non-linear relationships, can help uncover hidden patterns in the connectome, and potentially detect abnormal patterns of connectivity in individual subjects. In adult brain connectivity research, a number of studies have used machine learning and deep learning to study the structural connectome (see Brown and Hamarneh, 2016 for a review). Several studies also used similar approaches in neonates: Kawahara and colleagues have developed BrainNetCNN, a Convolutional Neural Network composed of edge to edge, edge to node, and node to graph convolution filters on structural connectivity to predict post menstrual age (PMA) at scan and cognitive performance from the structural connectome (Kawahara et al., 2017). A recent study by Girault and colleagues similarly focused on using the structural connectome at birth to predict cognitive abilities (Mullen score) at age 2 with dense neural networks (Girault et al., 2019). However, little is known about the predictive power of the connectome in a large normative neonatal population such as that of the dHCP.

A promising method in adult and neonatal neuroscience is the study of the “brain maturation index” (also known as “brain delta” or “predicted age difference”) corresponding to the apparent age of the subject as compared to the norm (Dosenbach et al. 2010; Cao et al. 2015; Jonsson et al. 2019; Liem et al. 2017; Smith et al. 2019). By training regression models to fit the age of subjects from large normative imaging datasets, we can predict the age of individual subjects and compute the difference between the prediction and subject’s true age. This difference gives information about brain maturation and its divergence from the population norm. As such, in adults, a positive (predicted age > true age) difference is interpreted as demonstrating accelerated ageing, and is associated with disorders such as cognitive impairment (Liem et al., 2017), schizophrenia (Koutsouleris et al., 2014) or diabetes (Franke et al., 2013). In neurodevelopment, studying the brain maturation index is therefore extremely relevant for preterm born infants where neurodevelopmental delays and psychiatric disorders often occur (Brown et al., 2017; Galdi et al., 2020; Rasmussen et al., 2017).

In this work, we propose the use of two different machine learning algorithms - Random Forests (RF) and Deep Neural Networks (DNN) - to predict PMA at scan (i.e. brain maturation), and gestational age (GA) at birth (i.e. degree of prematurity), from the neonatal structural connectome in a large sample of neonates scanned at term equivalent age. Using models obtained for the term-born cohort, brain maturation index was computed for our preterm-born cohort allowing us to assess whether it predicted neurodevelopmental outcome at 18 months.
2. Methods and Materials

2.1 Participants

All participants were part of the dHCP, approved by the National Research Ethics Service West London committee (14/LO/1169).

524 infants (240 female and 284 male), born between 23^+0 weeks and 42^+2 week of gestation, underwent MRI between 37^+1 weeks and 45^+1 weeks. The participant gestational age at birth (GA) and postmenstrual age at scan (PMA) distribution is presented in Figure 1A-B. Full participant clinical information is presented in Table 1.

The Bayley III Scales of Infant and Toddler Development (BSID-III) (Bayley 2006) were collected at 18 months corrected age and available for 314 infants including 50 preterm-born infants. We used scores of motor (fine and gross), communication (expressive and receptive) and cognitive (raw) score. Assessments were carried out by experienced paediatricians or psychologists. Detailed assessment distributions are presented in Table 1.

![Figure 1](https://example.com/figure1.png)

Figure 1. Distribution of (A) GA at birth and (B) PMA at scan of full cohort (N=524). (C) GA at birth of cohort used for predicting GA at birth.

2.2 MRI acquisition

All scans were collected in the Evelina Newborn Imaging Centre based on the Neonatal Intensive Care Unit, St Thomas hospital London using a Philips Achieva 3T scanner (Best, NL). All scans were acquired using the dHCP neonatal brain imaging system which includes a 32 channel receive neonatal head coil (Rapid Biomedical GmbH, Rimpar, DE) (Hughes et al., 2017). Informed written parental consent was obtained prior to imaging. Positioning of all infants was done with a lightweight protective “shell”, which was positioned on an MRI safe trolley to ease transportation. Immobilization of the infants in the shell was done using bead...
filled inflatable pads (Pearltect, Zurich, CH). In addition to the pads, acoustic protection included earplugs moulded from a silicone-based putty (President Putty, Coltene Whaledent, Mahwah, NJ, USA) placed in the external auditory meatus and neonatal earmuffs (MiniMuffs, Natus Medical Inc, San Carlos, CA, USA). To avoid sudden sound changes which might wake up the infant, the MRI software was modified in order to gradually increase the noise from 0 to the average operating point (Hughes et al., 2017). All scans were supervised by a paediatrician or neonatal nurse experienced in MRI procedures; vital signs including pulse oximetry, temperature and electrocardiography data were monitored throughout data acquisition. All infants were scanned during natural unsedated sleep following feeding.

T2-weighted images were acquired using a Turbo spin echo sequence with parameters TR = 12s and TE = 156ms, SENSE factor 2.11 (axial) and 2.54 (sagittal) with overlapping slices (resolution = 0.8 x 0.8 x 1.6 mm³). Super-resolution methods (Kuklisova-Murgasova et al., 2012) as well as motion correction methods (Cordero-Grande et al., 2018) were combined to maximise precision and resolution of T2-weighted images (resolved to 0.8 x 0.8 x 0.8 mm³). Diffusion weighted imaging was acquired in 300 directions with parameters TR = 3.8s, TE = 90ms, SENSE factor 1.2, multiband factor 4, resolution = 1.5 x 1.5 x 3mm³ (with 1.5mm slice overlap), diffusion gradient encoding: b=0 s/mm² (n=20), b=400 s/mm² (n=64), b=1000 s/mm² (n=88), b=2600 s/mm² (n=128), and using interleaved phase encoding (Hutter et al., 2018b).

2.3 Pre-processing and connectome generation

Tissue segmentation of T2-weighted volumes was performed using a neonatal specific segmentation pipeline (Makropoulos et al., 2014) and template (Schuh et al. 2018). Parcellation of 90 cortical and subcortical regions (Shi et al., 2011) adapted to the dHCP weekly age-dependant high-resolution bespoke template (Schuh et al. 2018) was propagated to each subject’s T2w native space through non-linear registration based in a diffeomorphic symmetric image normalization method (SyN) available in ANTS software (Avants et al., 2011), using T2w contrast and tissue segmentation as input channels. Tissue maps and atlas parcellation were propagated from each T2w native space to each subject’s diffusion native space with a rigid registration using b=0 volumes as target. All rigid registrations were performed with IRTK software (Schnabel et al. 2001). Details of the 90 cortical and subcortical regions are presented in Supplementary Table 1.

Diffusion MRI was reconstructed at an effective resolution of 1.5mm isotropic and denoised using a patch-based estimation of the diffusion signal based on random matrix theory (Veraart et al., 2016). Gibbs ringing was suppressed (Kellner et al., 2016) and B0 field map estimated from b=0 volumes in order to correct magnetic susceptibility-induced distortion using FSL Topup (Andersson et al., 2003). Data was corrected for slice-level motion and distortion in a data-driven q-space representation using a bespoke spherical harmonics and radial decomposition (SHARD) basis of rank 89 corresponding to spherical harmonics of order lmax=0,4,6,8 for each respective shell, with registration operating at a reduced rank of 22 (Christiaens et al., 2018). DWI intensity inhomogeneity field correction was performed using the ANTs N4 algorithm (Tustison et al., 2010). Tools and pipelines implemented in MRtrix3 (Tournier et al., 2019) were used for quantitative analysis of the diffusion MRI data. Developing neonatal brain tissue undergoes rapid changes in cellular properties and water content that can be to a first approximation captured by a non-negative linear combination of anisotropic signal from relatively mature WM and from isotropic free fluid (Pietsch et al., 2019). We use data from 20 healthy full term control babies from our sample to extract a set
of two representative WM (Tournier et al., 2013) and fluid-like (Dhollander et al., 2016, Dhollander et al., 2018) signal fingerprint (response functions) that are used to deconvolve each subject’s diffusion signal into a fibre orientation distribution (FOD) image, capturing WM-GM-like signal, and scalar fluid density image using the multi-tissue multi-shell constrained spherical deconvolution technique (Jeurissen et al. 2014). Residual intensity inhomogeneity were corrected and component densities calibrated using a multi-tissue log-domain intensity normalisation (Raffelt, et al. 2017). Resulting normalised WM-GM-like FODs were used to generate 10 million streamlines with an anatomically constrained probabilistic tractography (ACT) (Smith et al., 2012) with biologically accurate weights (SIFT2) (Smith, et al. 2015). The fibre density SIFT2 proportionality coefficient (μ) for each subject was obtained to achieve inter-subject connection density normalisation. The structural connectivity network of each infant was then constructed by calculating the μ \times SIFT2-weighted sum of streamlines connecting each pair of regions (thus built as a symmetric adjacency matrix of size 90x90).

In addition, we used 73 structural connectivity matrices obtained from an independent dataset (Batalle et al., 2017) to test the design of the initial hyperparameters and architecture for predictive algorithms presented in sections 2.4.3 and 2.4.4.

2.4 Prediction of age at scan and age at birth

All analyses on this section were performed using Python 3.7. The machine learning library Scikit Learn (Pedregosa et al., 2011) was used for training the RF algorithm. The deep learning framework Keras (version 2.0.3) (Chollet et al., 2015) was used to train the deep learning models.

2.4.1 Feature set

As the structural connectome is presented as a symmetric adjacency matrix (in our case of size 90x90, with 90 brain regions) the lower triangle of the matrix contains all information. We thus extracted and reshaped the lower triangle of each subject’s structural connectome \( S_i \) as a 1D vector \( X_i \) with number of connectivity elements \( n = 4005 \), thus leading to the ensemble \( X \) of connectivity vectors across \( N \) subjects:

\[
X = \begin{bmatrix}
X_1 \\
X_2 \\
\vdots \\
X_N 
\end{bmatrix}, \text{with } X_i = [x_{i,1}, x_{i,2}, \ldots, x_{i,n}]; \ x_{i,j} \in \mathbb{R}^+ \text{ and } n = 4005
\]

We normalized each data point across the training sets, and normalized the testing set with the training normalization values using a min-max normalization:

\[
\tilde{x}_{i,j} = \frac{x_{i,j} - \text{Min}(X)}{\text{Max}(X) - \text{Min}(X)}, \text{with } \text{Min}(X) \text{ and } \text{Max}(X) \in \mathbb{R}^+
\]

Thus, assuming that testing data also falls between previous ranges, our training and testing data has the following form:
\[ \tilde{X} = \begin{bmatrix} \tilde{x}_1 \\ \tilde{x}_2 \\ \vdots \\ \tilde{x}_N \end{bmatrix}, \text{with } \tilde{x}_i = [\tilde{x}_{i,1}, \tilde{x}_{i,2}, ..., \tilde{x}_{i,n}] ; \tilde{x}_{i,j} \in [0, 1] \text{ and } n = 4005 \]

### 2.4.2 Prediction models

We carried a set of predictions of demographic information on different population samples using different regression algorithms. In each case, we fitted a regression model \( f \) to predict a variable \( Y \) representing demographic information (e.g. GA at birth or PMA at scan) for subjects in our dataset. We thus had prediction \( Y' \) as follows:

\[ Y' = f(\tilde{X}); \text{with } Y' = [y_1', y_2', ..., y_N']^T \]

We computed the regressor \( f \) that minimizes \( |Y' - Y|^2 \). We used two different supervised machine learning regression algorithms to do this: RF and DNN.

### 2.4.3 Random Forests regression

RF are an ensemble learning method for classification and regression based on constructing a multitude of decision trees (weak learners) which are individually trained through the technique of “bagging”. RF makes predictions by averaging the prediction of each individual tree, hence acting as a strong learner (Breiman L., 2001). For optimal performance, two main hyperparameters should be tuned: the number of trees (estimators) in the forest and the maximum depth of each tree. The number of trees determines the smoothness of the decision boundary and the depth corresponds to the maximum number of levels allowed for each tree. RF regressors’ performance often depends on finding the optimal value for these to ensure that there is no overfitting or underfitting.

Here we use the RF regressor from the Scikit Learn RandomForestRegressor implementation (Pedregosa et al., 2011). The RF were trained using mean squared error (MSE) as loss function. Hyperparameters were tuned separately for the PMA at scan and GA at birth prediction by performing a grid search on a set of 73 structural connectomes from an independent dataset (Batalle et al., 2017). This allowed us to choose optimal parameters without overfitting our model to the studied data. Hyperparameters used are presented in sections 2.4.6 and 2.4.7.

### 2.4.4 Deep Neural Networks regression

Deep (Fully Connected) Neural Networks (DNN) are universal function approximators whose parameters can be trained to model complex nonlinear relationships between features and labels via backpropagation (Rumelhart et al., 1986). However, the performance of a DNN also depends on the hyperparameters: design choices are mainly related to the architecture of the network (the layer types, number of layers and number of nodes per layers, activation functions at each layer), the loss function, and the training method (the number of epochs, the optimization function and its parameters).

The DNN in this work were implemented using the deep learning library Keras (version 2.0.3) (Chollet et al., 2015). As performing a grid search to find the best model hyperparameters is computationally expensive when training DNN, we started with a basic architecture built from
previous work and common DNN knowledge (Smith 2018), and subsequently optimized these via manual refinement architecture search. To avoid overfitting the model to the data used in this paper, this was done on the same set of 73 structural connectomes from an independent sample as was used for the RF training, independently for the GA at birth and PMA at scan prediction. For both prediction tasks, the models were trained using MSE as a loss function and the Adam optimizer (Kingma and Ba, 2015), albeit with different learning rates. Further details on the network architecture are included in sections 2.4.6 and 2.4.7.

2.4.5 Training and evaluation of the models

To assess the performance of the prediction models, the evaluation metric was calculated on test data excluded from training and hyperparameter tuning. We split the dataset into k groups (folds) and fit the model k times. Each time, one group is used to evaluate performance, while the rest of the groups are used for training and validation. The evaluation scheme is presented in Figure 2B.

We split the data into k=5 groups (folds), with 20% of data used for testing at each fold. The remaining 80% of the data were further split for training (65%) to fit the models and validation (15%) to tune the hyperparameters. Min-max normalisation presented in section 2.4.1 is fitted on the training/validation set, where normalization parameters are saved and applied to the test set.

We added a bias correction as previously described (Smith et al. 2019; Peng et al. 2019) to correct age dependency of the training residuals. Briefly, we used a linear model $Y' = f(X) = \alpha Y + \beta$ to obtain an unbiased estimate of $Y'$ as $\hat{Y} = \frac{Y'-\beta}{\alpha}$, where the parameters $\alpha$ and $\beta$ are estimated during training (on both the combination of training and validation set) and are thus applied directly to the test set. We obtained our final corrected prediction $\hat{Y}_i$ for each structural connectome as follows:

$$\hat{Y}_i = \frac{f(\tilde{X}_i) - \beta}{\alpha} = \frac{Y'_i - \beta}{\alpha} = F(\tilde{X}_i); \quad \alpha, \beta \in \mathbb{R}$$

The final performance is calculated by averaging test-set performance over the 5 folds. We used mean absolute error (MAE) as our evaluation metric, calculated on each test set $k$ as follows:

$$MAE_k = \frac{1}{N_k} \sum_{i \in S_k} |Y_i - \hat{Y}_i|$$

Where $N_k$ is the number of subjects belonging to test set $k (S_k)$ and $Y_i$ and $\hat{Y}_i$ are actual and predicted outcome of subject $i$. In addition, we also evaluate MSE and $R^2$ scores for each test set $k$, which are calculated as follows:

$$MSE_k = \frac{1}{N_k} \sum_{i \in S_k} (Y_i - \hat{Y}_i)^2$$
\[ R_k^2 = 1 - \frac{\sum_{i \in S_k} (Y_i - \hat{Y}_i)^2}{\sum_{i \in S_k} (Y_i - \bar{Y})^2} \]

Where $\bar{Y}$ is the mean actual age of test set $k$. We also calculated Pearson’s Correlation ($r_k$) and p-value ($p_k$) between actual ($Y$) and predicted output ($\hat{Y}$) for each test set. As we obtain a prediction for every subject (albeit with different models) we can compute the $MAE_{tot}$, $MSE_{tot}$, $R^2_{tot}$, $r_{tot}$ and $p_{tot}$ by considering the predictions of the 5 test sets (see Figure 2A). Finally, we assessed the presence of heteroscedasticity in our predictions by comparing the variance $\sigma^2_{err}$ of the absolute error – a lower variance signifies more homoscedastic predictions.

### 2.4.6 Prediction of PMA at scan in term-born infants

To build a model of typical development of connectivity we used the full cohort of 418 term-born babies (GA at birth >= 37) with PMA at scan between 37 and 45 weeks.

We first predicted PMA at scan from the vectorised and normalized structural connectome $\tilde{X}$ using RF regressor model. Optimal parameters of the model (max depth = 250, number of estimators = 30) were found by performing a grid search in an independent dataset (see section 2.4.3). We trained each fold on $N \approx 335$ samples (80%) including a validation set. We then tested the model on the remaining set ($N \approx 83$, 20%) in each fold, thus being able to predict age at scan on all 418 structural connectomes of term infants (see Figure 2A).

In a similar fashion, we also trained a regression DNN to predict PMA at scan from the vectorised and normalized structural connectome $\tilde{X}$. This DNN comprises one input layer with 4005 input nodes, 7 hidden layers, 6 activation layers (ReLu), 5 batch normalisation layers and one output layer with one node. Training was done for 50 epochs with learning rate of 0.007 and remaining parameters with default value. Detailed structure of the architecture of this DNN is provided in Figure 2D.

We applied the previously described bias correction method on both DNN and RF, by fitting $\alpha$ and $\beta$ for each model $f_k$ using both the training and validation set; thus reaching 5 distinct models $F_1$, $F_2$, ..., $F_5$ for both the DNN and RF methods.

### 2.4.7 Prediction of GA at birth

To assess the effect of preterm birth on structural connectivity we trained a prediction model for GA at birth from $\tilde{X}$ with both DNN and RF in a similar fashion as previously described for prediction of PMA at scan.

Since the dHCP cohort has significantly more term-born than preterm-born infants, there is a “class imbalance” in the GA distribution that may skew the model prediction. We therefore randomly selected a sub-sample of term subjects that had, on average, equal density of subjects on each GA at birth weekly bin. Our 106 preterm infants were distributed in 15 different GA at birth bins (22w-23w; 23w-24w ... 36w-37w), thus providing an average of 7 infants per age category. We kept all 106 preterm infants and randomly sampled 7 infants...
from each of the term age categories (37w-38w, 38w-39w, ..., 41w-42w) and the 4 subjects born 42w-43w (as only 4 were born between 42w and 43w GA), for a total of 39 term-born infants. This resulted in a total of 145 infants with a balanced distribution (see Figure 1C).

We first attempted prediction of GA at birth from the vectorised and normalized structural connectome $\tilde{X}$ using RF with optimal parameters max depth = 300, number of estimators = 50. For each fold, we trained using $N=116$ samples (80%) including a validation set. We then tested the model on the remaining set ($N=29$, 20%) in each fold, predicting GA at birth for all 145 structural connectomes considered.

In a similar fashion, we also trained a DNN to predict GA at birth. This DNN consists of one input layer, 6 hidden layers, 6 activation layers (ReLu), one dropout layer, and one output layer. 120 epochs were used for training, with learning rate 0.003 and remaining parameters at default value. Detailed information on the architecture is provided in Figure 2E.

We applied the bias correction method on both DNN and RF by fitting $\alpha$ and $\beta$ for each model $h_i$ from the validation set; thus reaching 5 distinct models $H_1, H_2, ..., H_5$ for both the DNN and RF methods.

2.4.8 Brain maturation index

We defined brain maturation index $\delta$ (also called brain age or predicted age difference in the literature) as the difference between the predicted age $\hat{Y}$ and true age $Y$ of a subject $n$ (Dosenbach et al., 2010): $\delta_i = \hat{Y}_i - Y_i$

We developed a model of typical brain development by training 5 models to predict PMA at scan on term-born infants only (section 2.4.6). Prediction of PMA at scan for each preterm subject was computed by taking the mean of the predictions from each of the 5 DNN trained models $F_k$ from each cross-validation partition:

$$\hat{Y}_i = \frac{1}{5} \sum_{k=1}^{5} F_k(\tilde{X}_i) = G(\tilde{X}_i)$$

Following this, we computed the brain maturation index $\delta_i$ of each preterm subject:

$$\delta_i = \hat{Y}_i - Y_i = G(\tilde{X}_i) - Y_i$$
Figure 2. (A) Pipeline for age prediction from MRI. (B) Cross Validation protocol. (C) Legend for DNN architecture components. (D) PMA at scan DNN architecture. (E) GA at birth DNN architecture.
2.5 Statistical methods

Differences between term and preterm cohorts on all relevant characteristics were assessed by computing a two tailed independent t-test or chi-square test as appropriate. The association between brain maturation index δ_i and neurodevelopmental outcomes was assessed with Pearson’s Correlation coefficient for all preterm infants having both δ_i and BSID-III developmental outcomes at age 18 months corrected age. All outcomes were corrected for socioeconomic status, captured by the English Index of Multiple Deprivation (IMD). IMD factor summarizes information from 38 different factors such as income, employment, education, crime rates and health situation for all neighbourhoods in England (Index of Multiple Deprivation, 2015). Lower IMD relates to lower level of deprivation. All p-values presented are uncorrected for multiple comparisons.

2.6 Data availability

The imaging and collateral data from the dHCP can be downloaded by registering at https://data.developingconnectome.org/

Structural connectivity networks and code used to predict age at birth and age at scan are available in https://github.com/CoDe-Neuro/Predicting-age-and-clinical-risk-from-the-neonatal-connectome
3. Results

3.1 Sample characteristics

There were no significant differences in PMA at scan and male/female proportion between term and preterm neonates in this study. For the subjects for which 18 months BSID-III follow-up neurodevelopmental assessment was available, there were no significant differences in outcomes between term and preterm infants. However, a significant group difference (p<<0.001) was found in IMD scores, with term infants showing significantly higher deprivation than preterm infants. Detailed cohort characteristics are provided in Table 1 and Figure 1. Neurodevelopmental outcome details are provided in Table 1.

Table 1. Detailed sample and outcome characteristics.

|                                   | Term Born N = 418 | Preterm born N = 106 | p-value* |
|-----------------------------------|-------------------|----------------------|----------|
| Gestational age at birth [weeks +days] | Median = 40⁺¹ IQR = 39⁺⁰ – 40⁺⁶ | Median = 32⁺² IQR = 28⁺⁵ – 34⁺⁴ | <<0.001 |
| Postmenstrual age at scan [weeks +days] | Median = 41⁺⁰ IQR = 39⁺⁴ – 42⁺² | Median = 41⁺⁰ IQR = 39⁺⁴ – 42⁺² | 0.771    |
| Sex, no. of female (%)            | 195 (47%)         | 45 (42%)             | 0.401    |
| BSID-III, no. (% of total population) | 264 (63%)       | 50 (47%)             | 0.002    |
| Gestational age at birth (weeks +days) of subjects with BSID-III data available | Median = 40⁺¹ IQR = 39⁺¹ – 40⁺⁶ | Median = 31⁺⁵ IQR = 28⁺² – 34⁺⁵ | << 0.001 |
| Postmenstrual age at scan (weeks +days) of subjects with BSID-III data available | Median = 40⁺⁶ IQR = 39⁺⁵ – 42⁺⁰ | Median = 41⁺² IQR = 39⁺⁵ – 42⁺⁶ | 0.158    |
| IMD Score of subjects with BSID-III data available | Mean = 26.57 STD = 12.33 | Mean = 18.74 STD = 10.68 | <<0.001 |
| Fine Motor                        | Mean = 11.364 STD = 2.33 | Mean = 10.80 STD = 2.74 | 0.130    |
| Gross Motor                       | Mean = 9.049 STD = 1.77 | Mean = 8.74 STD = 2.24 | 0.282    |
| Cognitive Score                   | Mean = 9.98 STD = 2.17 | Mean = 9.74 STD = 2.82 | 0.503    |
| Expressive communication           | Mean = 8.82 STD = 2.61 | Mean = 8.82 STD = 2.73 | 0.996    |
| Receptive Communication           | Mean = 9.90 STD = 3.21 | Mean = 10.22 STD = 3.31 | 0.524    |

*p-values computed with two tailed independent t-test or chi-square test as appropriately.
3.2 Prediction of PMA at scan

3.2.1 Prediction of PMA at scan with RF

We trained a RF regressor to fit for PMA at scan from vectorised and normalized structural connectome $\tilde{X}$ on term infants only. We obtained $MAE_{tot} = 0.84$ weeks, $MSE_{tot} = 1.10$, $R^2_{tot} = 0.61$, $\sigma^2_{err} = 1.10$ with correlation between true and predicted $r_{tot} = 0.79$ ($p_{tot} \ll 0.001$). Figure 3A shows true PMA vs predicted PMA on each of the 5 cross-validation folds. Detailed results of each fold are presented in Figure 3C.

3.2.2 PMA at scan prediction with DNN

Similarly, we trained a DNN regressor to fit for PMA at scan from vectorised and normalized structural connectome $\tilde{X}$ on term infants only. We obtained $MAE_{tot} = 0.72$ weeks, $MSE_{tot} = 0.94$, $R^2_{tot} = 0.67$, $\sigma^2_{err} = 0.94$ with correlation between true and predicted $r_{tot} = 0.83$ ($p_{tot} \ll 0.001$). Figure 3B shows true PMA vs predicted PMA on each of the 5 cross-validation folds. Detailed results of each fold are presented in Figure 3D.

3.3 Prediction of GA at birth

3.3.1 Prediction of GA at birth with RF

We trained a RF regressor to fit GA at birth from vectorised and normalized structural connectome $\tilde{X}$ on balanced data (145 infants). We obtained $MAE_{tot} = 2.76$ weeks, $MSE_{tot} = 12.95$, $R^2_{tot} = 0.43$, $\sigma^2_{err} = 12.93$ with correlation between true and predicted $r_{tot} = 0.67$ ($p_{tot} \ll 0.001$). Figure 4A shows true GA at birth vs predicted GA at birth on each of the 5 folds fold. Detailed results of each fold are presented in Figure 4B.

3.3.2 Prediction of GA at birth with DNN

Similarly, we trained a DNN from vectorised and normalized structural connectome $\tilde{X}$ on balanced data. We obtained $MAE_{tot} = 2.21$ weeks, $MSE_{tot} = 8.90$, $R^2_{tot} = 0.61$, $\sigma^2_{err} = 8.86$ with correlation between true and predicted $r_{tot} = 0.82$ ($p_{tot} \ll 0.001$). Figure 4B shows true GA vs predicted GA on each of the 5 folds. Detailed results of each fold are presented in Figure 4D.
Figure 3. Detailed results of prediction of PMA at scan on term cohort. True vs Predicted with (A) RF, (B) DNN. Fold by fold result with (C) RF, (D) DNN.
Figure 4. Detailed results of prediction of GA at birth. True vs Predicted with (A) RF, (B) DNN. Fold by fold result with (C)RF, (D)DNN.
3.3 Brain Maturation Index

We computed the predicted PMA at scan of each preterm-born infant scanned at term-equivalent age by averaging out the 5 predictions from the DNN term trained models. We obtained MAE of 1.16 weeks on prediction of 106 preterm infants ($MSE_{tot} = 2.24, R^2_{tot} = 0.42, \sigma^2_{err} = 1.45$), with correlation between true and predicted age between $r = 0.79$, ($p \ll 0.001$). True vs predicted age is presented in Figure 5A.

We computed the brain maturation index $\delta_i$ from each prediction. Brain maturation index ($\delta_i$) was significantly correlated with BSID-III gross motor scale at 18 months corrected age ( $r = 0.4590, p = 0.0008$, Figure 5BC).

| BSID-III outcomes       | Sample size | Correlation with prediction residuals | Correlation | p-value |
|-------------------------|-------------|---------------------------------------|-------------|---------|
| Fine Motor              | 50          | 0.0825                                | 0.5688      |         |
| Gross Motor             | 50          | 0.4590                                | 0.0008      |         |
| Receptive Communication | 50          | 0.1278                                | 0.3766      |         |
| Expressive Communication| 50          | 0.2578                                | 0.0707      |         |
| Cognitive scaled score  | 50          | 0.1986                                | 0.1667      |         |

Figure 5. Association of Brain Maturation Index with BSID-III outcomes in preterm-born infants. (A) True vs predicted PMA at scan for preterm infants. (B) Brain maturation index $\delta$ vs BSID-III Gross motor outcome corrected for IMD. (C) Detailed correlation and p-values of preterm brain maturation index and BSID-III outcomes – statistically significant results highlighted in bold red.
4. Discussion

This work demonstrates that machine learning can uncover connectivity patterns associated with typical and atypical development of structural brain connectivity, despite the high dimensionality of the data. We obtained accurate prediction of PMA at scan in term-born infants in a large normative sample of high-quality neonatal MRI data from the dHCP and predicted GA at birth in preterm infants from scans at term equivalent age, showing the impact of preterm birth on the structural connectome. Furthermore, we have also shown significant correlation between brain maturation index estimated from brain connectivity at birth and BSID-III gross motor outcome at 18 months of corrected age in preterm babies. Overall, our results show that machine learning approaches can extract relevant information on brain development from the neonatal structural connectome.

Prediction of age at scan

We achieved high accuracy in our prediction of PMA at scan on the term cohort, reaching a low MAE of 0.72 weeks ($MSE_{\text{tot}} = 0.94$, $R^2_{\text{tot}} = 0.67$) and a high correlation between true and predicted age ($r_{\text{tot}} = 0.83$; $p_{\text{tot}} < 0.001$). While the structural connectome presents several important challenges to study brain connectivity (Campbell and Pike, 2014) including high numbers of false positive streamlines (Maier-Hein et al., 2017), our results suggest that there is reliable information present to capture the subtle changes associated with weekly development. There have been several studies evaluating white matter microstructural and connectivity changes during the first days after birth. Indeed, it has been found that the postnatal period is marked by further dendritic arborization, refinement of existing intracortical connections, and an increase in synaptogenesis which results in an abundance of connections (for a review see (Keunen et al., 2017)). Some important changes have also been found in the structural connectome in the early postnatal period, mainly an increase in integration (the ease with which different brain regions communicate) and segregation (presence of clusters, i.e., capacity for specialised processing) (Batalle et al., 2017). These changes are likely captured by DNN and underlie the accurate prediction of PMA at scan.

Predicting PMA at scan from the structural connectome has, to our knowledge, only previously been done by Kawahara et al., where a MAE of 2.17 weeks and a correlation between true and predicted age of 0.87 were achieved in a cohort of 115 preterm infants (between 24 and 32 weeks PMA) (Kawahara et al., 2017). Although we achieved comparable correlation between true and predicted age, we achieved an improved MAE, which is likely due to the larger high-quality dataset and use of normative population.

Prediction of age at birth

Several studies have assessed the effects of prematurity in brain structure. Volumetric changes in the cerebellum (Limperopoulos et al., 2010), cortical and subcortical grey matter (Padilla et al., 2015) and altered shape of hippocampus (Thompson et al., 2009) have all been associated with preterm birth. The preterm brain white matter is also affected - studies have demonstrated significant alterations of white matter microstructure in the preterm infant without visible focal lesions on clinical MRI (Anjari et al., 2007; Hüppi et al., 1998), as well as more overt white matter injuries such as periventricular leukomalacia (Counsell et al., 2003;
Volpe, 2003). Other studies have focused on the impact of preterm birth on brain connectivity. Interestingly, several key components of structural connectivity appear to be unaltered by prematurity including rich club organization (Ball et al., 2014) and core connections (Batalle et al., 2017). However, whilst these core features are preserved, various other components are significantly impacted by the degree of prematurity, including decreased local connectivity in various brain regions (such as cerebellum and superior frontal lobe) (Batalle et al., 2017), or reduced fractional anisotropy in the corticospinal tracts and corpus callosum (Ball et al., 2012).

These alterations likely underlie the ability of our DNN to clearly decipher the level of prematurity of infants from the structural connectome and led to a relatively good prediction of GA at birth (DNN performance of \( \text{MAE}_{\text{tot}} = 2.21 \) weeks; \( \text{MSE}_{\text{tot}} = 8.90; \ R^2_{\text{tot}} = 0.61; \ r_{\text{tot}} = 0.82; \ p_{\text{tot}} \ll 0.001 \)) from the structural connectome of preterm-born infants scanned at term equivalent age. This shows that the structural connectome contains significant markers of cerebral white matter abnormalities, which are characteristic of prematurity level. This result is in keeping with an earlier study by Brown and colleagues which achieved prediction with precision of 1.6 weeks from the structural connectome of preterm infants only (77 scans, GA at birth between 24 and 32 weeks) using RF (Brown et al., 2017). This high performance is likely due to the reduced age range of the cohort compared to our study. Smyser and colleagues also attempted to predict preterm-birth from brain networks obtained from functional MRI, using support vector machines with a cohort of 100 babies (50 preterm) scanned at term equivalent age (Smyser et al., 2016), although their classification was only dichotomic. They achieved accuracy of 84% in classifying term- vs preterm-birth, suggesting that alterations in brain connectivity are also present in functional connectivity networks.

**Brain maturation index**

Brain maturation indices have been suggested as a powerful tool to capture alterations in the maturational trajectories of brain connectivity (Cao et al., 2015). Association with neurodevelopmental outcomes such as BSID-III is then important to evaluate the lasting impact of this predicted delay. Our bespoke brain maturation index \( \delta \) was thus computed for all preterm infants in which BSID-III outcome were available (50 infants). Contrary to adulthood where a positive \( \delta \) (predicted age > true age) is associated with emergence of various disorders such as cognitive decline (Jonsson et al., 2019), a negative \( \delta \) (predicted age < true age) can be associated with developmental delay in neonates. This hypothesis was verified as we found a positive correlation between the brain maturation index and BSID-III gross motor outcome in our cohort of preterm infants at 18 months of corrected age. This is in accordance with previous findings which have linked preterm birth to motor delay in later development (Foulder-Hughes and Cooke, 2007). This suggests that the brain maturation index may be a useful tool to capture potential delays and disorders in structural connectivity which may have a lasting impact on later neurological outcomes. Recent studies have shown the potential of normative modelling to find individual alterations in preterm babies (Dimitrova et al., 2020; O’Muircheartaigh et al., 2020), which are characterised by heterogeneous brain changes. In a similar way, we suggest that for an individual subject, a
high deviation from the population norm, translating to age predictions significantly lower than true age (negative brain maturation index) can be a marker of potential developmental delay so that these subjects should undergo further tests and may need follow up. Therefore, this marker may provide an opportunity for early preventive intervention, as other similar studies suggest (Cao et al., 2015).

**Methodological considerations**

We have assessed the performance of RF and DNN for prediction of key developmental characteristics in a large sample of neonates. Although both RF and DNN have been used extensively in the literature for brain imaging analysis, to our knowledge this is the first application in a large cohort of neonates. Previous studies have used convolutional neural networks (Kawahara et al., 2017) to extract information from the structural connectome which are particularly useful for data known to have local correlation, such is the case with segmenting brain MR images (Lecun et al., 1998). However, since the spatial distribution of adjacency matrices are not reflective of brain region locality and connectivity characteristics, we instead chose to use DNN in our investigation. Using this approach, we achieved better performance compared to RF on age prediction from the neonatal structural connectome. Although prediction of PMA at scan was also highly accurate with RF ($MAE_{tot} = 0.84$; $MSE_{tot} = 1.10$; $R^2_{tot} = 0.61$; $\sigma^2_{err} = 1.10$; $r = 0.79$) DNN achieved better performance ($MAE_{tot} = 0.72$; $MSE_{tot} = 0.94$; $R^2_{tot} = 0.67$; $\sigma^2_{err} = 0.94$; $r = 0.83$) with a more homoscedastic distribution of predictions over each of the 5 cross-validation folds (Figure 3). The improved performance of DNN over RF was more evident for prediction of GA at birth, with a better performance and more homoscedastic distribution of predictions on each fold with DNN ($MAE_{tot} = 2.21$; $MSE_{tot} = 8.90$; $R^2_{tot} = 0.61$; $\sigma^2_{tot} = 8.86$; $r = 0.82$) over RF ($MAE_{tot} = 2.76$; $MSE_{tot} = 12.95$; $R^2_{tot} = 0.43$; $\sigma^2_{tot} = 12.93$; $r = 0.67$) (Figure 4A-D). Our choice of undersampling the term cohort to achieve balance between age categories, although diminishing the sample size, was necessary to avoid a class imbalance problem, which could have caused a systematic positive bias for preterm infants (predicted GA > true GA). The relatively good performance of our model suggests that the impact of preterm birth on brain connectivity development is important and clearly apparent on the neonatal structural connectome.

We achieved high performance in the prediction of PMA at scan in our preterm cohort by averaging the predictions from all 5 term trained DNN models ($MAE_{tot} = 1.16$; $MSE_{tot} = 2.24$; $R^2_{tot} = 0.42$; $\sigma^2_{tot} = 1.45$; $r = 0.79$; $p \ll 0.001$). Although there was high accuracy in prediction and correlation between true and predicted age for preterm infants, predictions were on average inferior to those obtained for PMA at scan for term infants (Figure 5A). This is expected as preterm neonates are known to have specific differences in structural connectivity when compared with their term counterparts which may have reduced the generalizability of the predictive model (Ball et al., 2012; Batalle et al., 2017; Smyser et al., 2010). The difference between true and predicted PMA (brain maturation index) varies greatly across subjects, which we hypothesise is representative of the severity of the delay.
Limitations

There are several limitations to this work. Firstly, although the dHCP data set is the largest of its kind, a key next step will be to see whether the findings generalise to other populations as the regression algorithms were built and trained with the specific MRI acquisition protocol, brain parcellation and connectome generation methods developed for that project. To enable this, our predictive algorithms have been made publicly available, so other researchers can evaluate their performance in data sets with different acquisition and processing protocols. As the data was normalized prior to training, given a similar parcellation, we hypothesize that significant correlation between true and predicted GA at birth or PMA at scan should be obtained if tested on different data. There is increasing interest in predicting outcomes at age 18-24 months directly from neonatal brain connectivity, as done in (Girault et al., 2019). We have implemented our own version of their method and tested on the sub-set of our data set with available BSID-III at 18 months. However, no significant prediction capacity was reached with our data (data not shown). This might be due to different developmental outcome (Mullen scale instead of BSID-III), as well as differences in the pre-processing pipeline, or differences in the sample size and characteristics.

The recent progress in geometric deep learning, a new type of deep neural models specifically designed for data in non-Euclidean space (such as graphs), could be of great potential to improve the results of dense neural networks (Bronstein et al., 2017).

It remains difficult to deploy this type of study to clinical settings, as the highly nonlinear and multidimensional inner working of the algorithms are difficult to interpret by humans. Recent progress in deep learning explainability is of great potential to help on that matter (for a review, see (Xie et al., 2020)). In this context, identifying which specific edges influenced the decision of the network for a specific prediction could help clinicians to diagnose specific neurodevelopmental disorders and may have implications for targeted intervention.

5. Conclusion

In this work, we have used DNN to uncover important demographic and clinical information from the neonatal structural connectome, for the first time in a large sample of normative neonates. We achieved a MAE of 0.72 weeks in predicting PMA at scan, demonstrating that the neonatal structural connectome contains key developmental information. Furthermore, our prediction of GA at birth, with MAE of 2.21 weeks, shows that the patterns characteristic of prematurity are clearly present in the neonatal connectome, and can be uncovered with machine learning approaches. Finally, our brain maturation index computation on the preterm cohort was significantly correlated to BSID-III motor outcome at corrected age of 18 months. Brain maturation index thus appears to be a promising biomarker for prediction of neurodevelopmental disorders and delays, opening a potential path for early diagnosis and prevention of disorders in preterm born neonates.
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References

Andersson, J. L., Skare, S. and Ashburner, J. (2003), ‘How to correct susceptibility distortions in spin-echo echo-planar images: application to diffusion tensor imaging’, *Neuroimage* **20**(2), 870–888.

Anjari, M., Srinivasan, L., Allsop, J. M., Hajnal, J. V., Rutherford, M. A., Edwards, A. D. and Counsell, S. J. (2007), ‘Diffusion tensor imaging with tract-based spatial statistics reveals local white matter abnormalities in preterm infants’, *Neuroimage* **35**(3), 1021–1027.

Arbabshirani, M. R., Kiehl, K., Pearlson, G. and Calhoun, V. D. (2013), ‘Classification of schizophrenia patients based on resting-state functional network connectivity’, *Frontiers in neuroscience* **7**, 133.

Avants, B. B., Tustison, N. J., Song, G., Cook, P. A., Klein, A. and Gee, J. C. (2011), ‘A reproducible evaluation of ants similarity metric performance in brain image registration’, *Neuroimage* **54**(3), 2033–2044.

Ball, G., Aljabar, P., Zebari, S., Tusor, N., Arichi, T., Merchant, N., Robinson, E. C., Ogundipe, E., Rueckert, D., Edwards, A. D. et al. (2014), ‘Rich-club organization of the newborn human brain’, *Proceedings of the National Academy of Sciences* **111**(20), 7456–7461.

Ball, G., Boardman, J. P., Rueckert, D., Aljabar, P., Arichi, T., Merchant, N., Gousias, I. S., Edwards, A. D. and Counsell, S. J. (2012), ‘The effect of preterm birth on thalamic and cortical development’, *Cerebral cortex* **22**(5), 1016–1024.

Bastiani, M., Andersson, J. L., Cordero-Grande, L., Murgasova, M., Hutter, J., Price, A. N., Makropoulos, A., Fitzgibbon, S. P., Hughes, E., Rueckert, D. et al. (2019), ‘Automated processing pipeline for neonatal diffusion mri in the developing human connectome project’, *Neuroimage* **185**, 750–763.

Batalle, D., Edwards, A. D. and O’Muircheartaigh, J. (2018), ‘Annual research review: not just a small adult brain: understanding later neurodevelopment through imaging the neonatal brain’, *Journal of Child Psychology and Psychiatry* **59**(4), 350–371.

Batalle, D., Hughes, E. J., Zhang, H., Tournier, J.-D., Tusor, N., Aljabar, P., Wali, L., Alexander, D. C., Hajnal, J. V., Nosarti, C. et al. (2017), ‘Early development of structural networks and the impact of prematurity on brain connectivity’, *Neuroimage* **149**, 379–392.

Bayley, N. (2006), *Bayley scales of infant and toddler development*, PsychCorp, Pearson.

Blencowe, H., Cousens, S., Oestergaard, M. Z., Chou, D., Moller, A.-B., Narwal, R., Adler, A., Garcia, C. V., Rohde, S., Say, L. et al. (2012), ‘National, regional, and worldwide estimates of preterm birth rates in the year 2010 with time trends since 1990 for selected countries: a systematic analysis and implications’, *The lancet* **379**(9832), 2162–2172.

Bozek, J., Makropoulos, A., Schuh, A., Fitzgibbon, S., Wright, R., Glasser, M. F., Coalson, T. S., O’Muircheartaigh, J., Hutter, J., Price, A. N. et al. (2018), ‘Construction of a neonatal cortical surface atlas using multimodal surface matching in the developing human connectome project’, *Neuroimage* **179**, 11–29.

Breiman, L. (2001), ‘Random forests’, *Machine learning* **45**(1), 5–32.

Bronstein, M. M., Bruna, J., LeCun, Y., Szlam, A. and Vandergheynst, P. (2017), ‘Geometric deep learning: going beyond euclidean data’, *IEEE Signal Processing Magazine* **34**(4), 18–42.
Brown, C. J. and Hamarneh, G. (2016), ‘Machine learning on human connectome data from mri’, arXiv preprint arXiv:1611.08699.

Brown, C. J., Moriarty, K. P., Miller, S. P., Booth, B. G., Zwicker, J. G., Grunau, R. E., Synnes, A. R., Chau, V. and Hamarneh, G. (2017), Prediction of brain network age and factors of delayed maturation in very preterm infants, in ‘International Conference on Medical Image Computing and Computer-Assisted Intervention’, Springer, pp. 84–91.

Campbell, J. S. and Pike, G. B. (2014), ‘Potential and limitations of diffusion mri tractography for the study of language’, Brain and language 131, 65–73.

Cao, B., Mwangi, B., Hasan, K. M., Selvaraj, S., Zeni, C. P., Zunta-Soares, G. B. and Soares, J. C. (2015), ‘Development and validation of a brain maturation index using longitudinal neuroanatomical scans’, Neuroimage 117, 311–318.

Chollet, F. et al. (2015), ‘Keras’, https://github.com/fchollet/keras.

Christiaens, D., Cordero-Grande, L., Hutter, J., Price, A. N., Deprez, M., Hajnal, J. V. and Tournier, J.-D. (2018), ‘Learning compact q-space representations for multi-shell diffusion-weighted mri’, IEEE transactions on medical imaging 38(3), 834–843.

Christiaens, D., Cordero-Grande, L., Pietsch, M., Hutter, J., Edwards, A. D., Deprez, M., Hajnal, V. and Tournier, J. (2018), ‘Multi-shell shard reconstruction from scattered slice diffusion mri data in the neonatal brain’, ISMRM (Paris).

Cordero-Grande, L., Christiaens, D., Hutter, J., Price, A. N. and Hajnal, J. V. (2019), ‘Complex diffusion-weighted image estimation via matrix recovery under general noise models’, NeuroImage 200, 391–404.

Cordero-Grande, L., Hughes, E. J., Hutter, J., Price, A. N. and Hajnal, J. V. (2018), ‘Three-dimensional motion corrected sensitivity encoding reconstruction for multi-shot multi-slice mri: application to neonatal brain imaging’, Magnetic resonance in medicine 79(3), 1365–1376.

Cordero-Grande, L., Teixeira, R. P. A., Hughes, E. J., Hutter, J., Price, A. N. and Hajnal, J. V. (2016), ‘Sensitivity encoding for aligned multishot magnetic resonance reconstruction’, IEEE Transactions on Computational Imaging 2(3), 266–280.

Counsell, S., Rutherford, M., Cowan, F. and Edwards, A. (2003), ‘Magnetic resonance imaging of preterm brain injury’, Archives of Disease in Childhood-Fetal and Neonatal Edition 88(4), F269–F274.

Dhollander, T., Raffelt, D. and Connelly, A. (2016), Unsupervised 3-tissue response function estimation from single-shell or multi-shell diffusion mri data without a co-registered t1 image, in ‘ISMRM Workshop on Breaking the Barriers of Diffusion MRI’, Vol. 5, p. 5.

Dhollander, T., Raffelt, D. and Connelly, A. (2018), Accuracy of response function estimation algorithms for 3-tissue spherical deconvolution of diverse quality diffusion mri data, in ‘Proceedings of the Joint Meeting of the European Society for Magnetic Resonance in Medicine and Biology and the International Society of Magnetic Resonance in Medicine, Paris, France’, Vol. 1569.

Dimitrova, R., Pietsch, M., Christiaens, D., Ciarrustja, J., Wolfers, T., Batalle, D., Hughes, E., Hutter, J., Cordero-Grande, L., Price, A. N. et al. (2020), ‘Heterogeneity in brain microstructural development following preterm birth’, Cerebral Cortex 30(9), 4800–4810.
Dosenbach, N. U., Nardos, B., Cohen, A. L., Fair, D. A., Power, J. D., Church, J. A., Nelson, S. M., Wig, G. S., Vogel, A. C., Lessov-Slaggar, C. N. et al. (2010), ‘Prediction of individual brain maturity using fmri’, *Science* **329**(5997), 1358–1361.

Fan, Y., Shi, F., Smith, J. K., Lin, W., Gilmore, J. H. and Shen, D. (2011), ‘Brain anatomical networks in early human brain development’, *Neuroimage* **54**(3), 1862–1871.

Fitzgibbon, S. P., Harrison, S. J., Jenkinson, M., Baxter, L., Robinson, E. C., Bastiani, M., Bozek, J., Karolis, V., Grande, L. C., Price, A. N. et al. (2020), ‘The developing human connectome project (dhcp) automated resting-state functional processing framework for newborn infants.’, *BioRxiv* p. 766030.

Foulder-Hughes, L. and Cooke, R. (2003), ‘Motor, cognitive, and behavioural disorders in children born very preterm’, *Developmental Medicine & Child Neurology* **45**(2), 97–103.

Franke, K., Gaser, C., Manor, B. and Novak, V. (2013), ‘Advanced brainage in older adults with type 2 diabetes mellitus’, *Frontiers in aging neuroscience* **5**, 90.

Galdi, P., Blesa, M., Stoye, D. Q., Sullivan, G., Lamb, G. J., Quigley, A. J., Thrippleton, M. J., Basti, M. E. and Boardman, J. P. (2020), ‘Neonatal morphometric similarity mapping for predicting brain age and characterizing neuroanatomic variation associated with preterm birth’, *NeuroImage: Clinical* **25**, 102195.

Girault, J. B., Munsell, B. C., Puechmaille, D., Goldman, B. D., Prieto, J. C., Styner, M. and Gilmore, J. H. (2019), ‘White matter connectomes at birth accurately predict cognitive abilities at age 2’, *NeuroImage* **192**, 145–155.

Hagmann, P. (2005), ‘From diffusion mri to brain connectomics’, Technical report, EPFL.

Hagmann, P., Sporns, O., Madan, N., Cammoun, L., Pienaar, R., Wedeen, V. J., Meuli, R., Thiran, J.-P. and Grant, P. (2010), ‘White matter maturation reshapes structural connectivity in the late developing human brain’, *Proceedings of the National Academy of Sciences* **107**(44), 19067–19072.

Honey, C. J., Thivierge, J.-P. and Sporns, O. (2010), ‘Can structure predict function in the human brain?’, *NeuroImage* **52**(3), 766–776.

Hughes, E. J., Winchman, T., Padorno, F., Teixeira, R., Wurie, J., Sharma, M., Fox, M., Hutter, J., Cordero-Grande, L., Price, A. N. et al. (2017), ‘A dedicated neonatal brain imaging system’, *Magnetic resonance in medicine* **78**(2), 794–804.

Hüppi, P. S., Maier, S. E., Peled, S., Zientara, G. P., Barnes, P. D., Jolesz, F. A. and Volpe, J. J. (1998), ‘Microstructural development of human newborn cerebral white matter assessed in vivo by diffusion tensor magnetic resonance imaging’, *Pediatric research* **44**(4), 584–590.

Hutter, J., Christiaens, D. J., Schneider, T., Cordero-Grande, L., Slator, P. J., Deprez, M., Price, A. N., Tournier, J.-D., Rutherford, M. and Hajnal, J. V. (2018), ‘Slice-level diffusion encoding for motion and distortion correction’, *Medical image analysis* **48**, 214–229.

Hutter, J., Tournier, J. D., Price, A. N., Cordero-Grande, L., Hughes, E. J., Malik, S., Steinweg, J., Bastiani, M., Sotiropoulos, S. N., Jbabdi, S. et al. (2018), ‘Time-efficient and flexible design of optimized multishell hardi diffusion’, *Magnetic resonance in medicine* **79**(3), 1276–1292.
Jbabdi, S. and Johansen-Berg, H. (2011), ‘Tractography: where do we go from here?’, Brain connectivity 1(3), 169–183.

Jeurissen, B., Tournier, J.-D., Dholander, T., Connelly, A. and Sijbers, J. (2014), ‘Multi-tissue constrained spherical deconvolution for improved analysis of multi-shell diffusion mri data’, NeuroImage 103, 411–426.

Johnson, M. H. (2001), ‘Functional brain development in humans’, Nature Reviews Neuroscience 2(7), 475–483.

Johnson, S., Hollis, C., Kochhar, P., Hennessy, E., Wolke, D. and Marlow, N. (2010), ‘Autism spectrum disorders in extremely preterm children’, The Journal of Pediatrics 156(4), 525–531.

Johnson, S. and Marlow, N. (2014), Growing up after extremely preterm birth: lifespan mental health outcomes, in ‘Seminars in Fetal and Neonatal Medicine’, Vol. 19, Elsevier, pp. 97–104.

Jonsson, B. A., Bjornsdottir, G., Thorgeirsson, T., Ellingsen, L. M., Walters, G. B., Gudbjartsson, D., Stefansson, H., Stefansson, K. and Ulfarsson, M. (2019), ‘Brain age prediction using deep learning uncovers associated sequence variants’, Nature communications 10(1), 1–10.

Kawahara, J., Brown, C. J., Miller, S. P., Booth, B. G., Chau, V., Grunau, R. E., Zwicker, J. G. and Hamarneh, G. (2017), ‘BrainNetCNN: Convolutional neural networks for brain networks; towards predicting neurodevelopment’, NeuroImage 146, 1038–1049.

Kellner, E., Dhital, B., Kiselev, V. G. and Reisert, M. (2016), ‘Gibbs-ringing artifact removal based on local subvoxel-shifts’, Magnetic resonance in medicine 76(5), 1574–1581.

Keunen, K., Counsell, S. J. and Benders, M. J. (2017), ‘The emergence of functional architecture during early brain development’, Neuroimage 160, 2–14.

Kingma, D. P. and Ba, J. (2014), ‘Adam: A method for stochastic optimization’, arXiv preprint arXiv:1412.6980.

Koutsouleris, N., Davatzikos, C., Borgwardt, S., Gaser, C., Bottlender, R., Frodl, T., Falkai, P., Riecher-Rössler, A., Möller, H.-J., Reiser, M. et al. (2014), ‘Accelerated brain aging in schizophrenia and beyond: a neuroanatomical marker of psychiatric disorders’, Schizophrenia bulletin 40(5), 1140–1153.

Kuklisova-Murgasova, M., Quaghebeur, G., Rutherford, M. A., Hajnal, J. V. and Schnabel, J. A. (2012), ‘Reconstruction of fetal brain mri with intensity matching and complete outlier removal’, Medical image analysis 16(8), 1550–1564.

LeCun, Y., Bottou, L., Bengio, Y. and Haffner, P. (1998), ‘Gradient-based learning applied to document recognition’, Proceedings of the IEEE 86(11), 2278–2324.

Liem, F., Varoquaux, G., Kynast, J., Beyer, F., Masouleh, S. K., Huntenburg, J. M., Lampe, L., Rahim, M., Abraham, A., Craddock, R. C. et al. (2017), ‘Predicting brain-age from multimodal imaging data captures cognitive impairment’, Neuroimage 148, 179–188.

Limperopoulos, C., Chilingaryan, G., Guizard, N., Robertson, R. L. and Du Plessis, A. J. (2010), ‘Cerebellar injury in the premature infant is associated with impaired growth of specific cerebral regions’, Pediatric research 68(2), 145–150.
Maier-Hein, K. H., Neher, P. F., Houde, J.-C., Côté, M.-A., Garyfallidis, E., Zhong, J., Chamberland, M., Yeh, F.-C., Lin, Y.-C., Ji, Q. et al. (2017), ‘The challenge of mapping the human connectome based on diffusion tractography’, Nature communications 8(1), 1–13.

Makropoulos, A., Gousias, I. S., Ledig, C., Aljabar, P., Serag, A., Hajnal, J. V., Edwards, A. D., Counsell, S. J. and Rueckert, D. (2014), ‘Automatic whole brain mri segmentation of the developing neonatal brain’, IEEE transactions on medical imaging 33(9), 1818–1831.

Makropoulos, A., Robinson, E. C., Schuh, A., Wright, R., Fitzgibbon, S., Bozek, J., Counsell, S. J., Steinweg, J., Vecchiato, K., Passerat-Palmbach, J. et al. (2018), ‘The developing human connectome project: A minimal processing pipeline for neonatal cortical surface reconstruction’, Neuroimage 173, 88–112.

Marlow, N., Hennessy, E. M., Bracewell, M. A., Wolke, D. et al. (2007), ‘Motor and executive function at 6 years of age after extremely preterm birth’, Pediatrics 120(4), 793–804.

Marlow, N., Wolke, D., Bracewell, M. A. and Samara, M. (2005), ‘Neurologic and developmental disability at six years of age after extremely preterm birth’, New England journal of medicine 352(1), 9–19.

Mitra, J., Shen, K.-k., Ghose, S., Bourgeat, P., Fripp, J., Salvado, O., Pannek, K., Taylor, D. J., Mathias, J. L. and Rose, S. (2016), ‘Statistical machine learning to identify traumatic brain injury (tbi) from structural disconnections of white matter networks’, Neuroimage 129, 247–259.

Nosarti, C., Reichenberg, A., Murray, R. M., Cnattingius, S., Lambe, M. P., Yin, L., MacCabe, J., Rifkin, L. and Hultman, C. M. (2012), ‘Preterm birth and psychiatric disorders in young adult life’, Archives of general psychiatry 69(6), 610–617.

O’Muircheartaigh, J., Robinson, E. C., Pietsch, M., Wolfers, T., Aljabar, P., Grande, L. C., Teixeira, R. P., Bozek, J., Schuh, A., Makropoulos, A. et al. (2020), ‘Modelling brain development to detect white matter injury in term and preterm born neonates’, Brain 143(2), 467–479.

Padilla, N., Alexandrou, G., Blennow, M., Lagercrantz, H. and Ådén, U. (2015), ‘Brain growth gains and losses in extremely preterm infants at term’, Cerebral Cortex 25(7), 1897–1905.

Pedregosa, F., Varoquaux, G., Gramfort, A., Michel, V., Thirion, B., Grisel, O., Blondel, M., Prettenhofer, P., Weiss, R., Dubourg, V. et al. (2011), ‘Scikit-learn: Machine learning in python’, the Journal of machine Learning research 12, 2825–2830.

Peng, H., Gong, W., Beckmann, C. F., Vedaldi, A. and Smith, S. M. (2019), ‘Accurate brain age prediction with lightweight deep neural networks’, BioRxiv .

Pietsch, M., Christiaens, D., Hutter, J., Cordero-Grande, L., Price, A. N., Hughes, E., Edwards, A. D., Hajnal, J. V., Counsell, S. J. and Tournier, J.-D. (2019), ‘A framework for multi-component analysis of diffusion mri data over the neonatal period’, Neuroimage 186, 321–337.

Raffelt, D., Dholander, T., Tournier, J.-D., Tabbara, R., Smith, R. E., Pierre, E. and Connelly, A. (2017), Bias field correction and intensity normalisation for quantitative analysis of apparent fibre density, in ‘Proc. Intl. Soc. Mag. Reson. Med’, Vol. 25, p. 3541.

Rasmussen, J. M., Kruggel, F., Gilmore, J. H., Stynner, M., Entringer, S., Consing, K. N., Potkin, S. G., Wadhwa, P. D. and Buss, C. (2017), ‘A novel maturation index based on neonatal diffusion tensor imaging reflects typical perinatal white matter development in humans’, International Journal of Developmental Neuroscience 56, 42–51.
Rumelhart, D. E., Hinton, G. E. and Williams, R. J. (1986), ‘Learning representations by back-propagating errors’, *nature* **323**(6088), 533–536.

Schnabel, J. A., Rueckert, D., Quist, M., Blackall, J. M., Castellano-Smith, A. D., Hartkens, T., Penney, G. P., Hall, W. A., Liu, H., Truwit, C. L. et al. (2001), A generic framework for non-rigid registration based on non-uniform multi-level free-form deformations, in ‘International Conference on Medical Image Computing and Computer-Assisted Intervention’, Springer, pp. 573–581.

Schuh, A., Makropoulos, A., Robinson, E. C., Cordero-Grande, L., Hughes, E., Hutter, J., Price, A. N., Murgasova, M., Teixeira, R. P. A., Tusor, N. et al. (2018), ‘Unbiased construction of a temporally consistent morphological atlas of neonatal brain development’, *bioRxiv* p. 251512.

Shi, F., Yap, P.-T., Wu, G., Jia, H., Gilmore, J. H., Lin, W. and Shen, D. (2011), ‘Infant brain atlases from neonates to 1-and 2-year-olds’, *PloS one* **6**(4), e18746.

Smith, Leslie N. ‘A disciplined approach to neural network hyper-parameters: Part 1--learning rate, batch size, momentum, and weight decay.’ arXiv preprint arXiv:1803.09820 (2018).

Smith, R. E., Tournier, J.-D., Calamante, F. and Connelly, A. (2012), ‘Anatomically-constrained tractography: improved diffusion mri streamlines tractography through effective use of anatomical information’, *Neuroimage* **62**(3), 1924–1938.

Smith, R. E., Tournier, J.-D., Calamante, F. and Connelly, A. (2015), ‘Sift2: Enabling dense quantitative assessment of brain white matter connectivity using streamlines tractography’, *Neuroimage* **119**, 338–351.

Smith, S. M., Vidaurre, D., Alfaro-Almagro, F., Nichols, T. E. and Miller, K. L. (2019), ‘Estimation of brain age delta from brain imaging’, *Neuroimage* **200**, 528–539.

Smith, T., Noble, M., Noble, S., Wright, G., McLennan, D., Plunkett, E. et al. (2015), ‘The english indices of deprivation 2015’, *London: Department for Communities and Local Government*.

Smyser, C. D., Dosenbach, N. U., Smyser, T. A., Snyder, A. Z., Rogers, C. E., Inder, T. E., Schlaggar, B. L. and Neil, J. J. (2016), ‘Prediction of brain maturity in infants using machine-learning algorithms’, *NeuroImage* **136**, 1–9.

Smyser, C. D., Inder, T. E., Shimony, J. S., Hill, J. E., Degnan, A. J., Snyder, A. Z. and Neil, J. J. (2010), ‘Longitudinal analysis of neural network development in preterm infants’, *Cerebral cortex* **20**(12), 2852–2862.

Sporns, O. (2013), ‘The human connectome: origins and challenges’, *Neuroimage* **80**, 53–61.

Sporns, O., Tononi, G. and Kötter, R. (2005), ‘The human connectome: a structural description of the human brain’, *PLoS Comput Biol* **1**(4), e42.

Thompson, D. K., Wood, S. J., Doyle, L. W., Warfield, S. K., Egan, G. F. and Inder, T. E. (2009), ‘Mr-determined hippocampal asymmetry in fullterm and preterm neonates’, *Hippocampus* **19**(2), 118–123.

Tournier, J.-D., Calamante, F. and Connelly, A. (2013), ‘Determination of the appropriate b value and number of gradient directions for high-angular-resolution diffusion-weighted imaging’, *NMR in Biomedicine* **26**(12), 1775–1786.
Tournier, J.-D., Smith, R., Raffelt, D., Tabbara, R., Dhollander, T., Pietsch, M., Christiaens, D., Jeurissen, B., Yeh, C.-H. and Connelly, A. (2019), ‘Mrtrix3: A fast, flexible and open software framework for medical image processing and visualisation’, NeuroImage 202, 116137.

Tustison, N. J., Avants, B. B., Cook, P. A., Zheng, Y., Egan, A., Yushkevich, P. A. and Gee, J. C. (2010), ‘N4itk: improved n3 bias correction’, IEEE transactions on medical imaging 29(6), 1310–1320.

Veraart, J., Novikov, D. S., Christiaens, D., Ades-Aron, B., Sijbers, J. and Fieremans, E. (2016), ‘Denoising of diffusion mri using random matrix theory’, Neuroimage 142, 394–406.

Volpe, J. J. (2003), ‘Cerebral white matter injury of the premature infant—more common than you think’, Pediatrics 112(1), 176–180.

Wood, N. S., Marlow, N., Costelo, K., Gibson, A. T. and Wilkinson, A. R. (2000), ‘Neurologic and developmental disability after extremely preterm birth’, New England Journal of Medicine 343(6), 378–384.

Xie, N., Ras, G., van Gerven, M. and Doran, D. (2020), ‘Explainable deep learning: A field guide for the uninitiated’, arXiv preprint arXiv:2004.14545