C-MYC Aberrations as Prognostic Factors in Diffuse Large B-cell Lymphoma: A Meta-Analysis of Epidemiological Studies

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Abstract

Objectives: Various studies have investigated the prognostic value of C-MYC aberrations in diffuse large B-cell lymphoma (DLBCL). However, the role of C-MYC as an independent prognostic factor in clinical practice remains controversial. A systematic review and meta-analysis were performed to clarify the clinical significance of C-MYC aberrations in DLBCL patients.

Methods: The pooled hazard ratios (HRs) for overall survival (OS) and event-free survival (EFS) were calculated as the main effect size estimates. The procedure was conducted according to the Cochrane handbook and PRISMA guidelines, including the use of a heterogeneity test, publication bias assessment, and meta-regression, as well as subgroup analyses.

Results: Twenty-four eligible studies enrolling 4662 patients were included in this meta-analysis. According to the nature of C-MYC aberrations (gene, protein, and mRNA), studies were divided into several subgroups. For DLBCL patients with C-MYC gene abnormalities, the combined HR was 2.22 (95% confidence interval, 1.89 to 2.61) for OS and 2.29 (95% confidence interval, 1.81 to 2.90) for EFS, compared to patients without C-MYC gene abnormalities. For DLBCL patients with overexpression of C-MYC protein and C-MYC mRNA, pooled HRs for OS were 2.13 and 1.62, respectively. C-MYC aberrations appeared to play an independent role among other well-known prognostic factors in DLBCL. Addition of rituximab could not overcome the inferior prognosis conferred by C-MYC.

Conclusion: The present systematic review and meta-analysis confirm the prognostic value of C-MYC aberrations. Screening of C-MYC should have definite prognostic meaning for DLBCL stratification, thus guaranteeing a more tailored therapy.

Introduction

Non-Hodgkin lymphoma (NHL) is the fifth most frequent cancer worldwide, in which diffuse large B-cell lymphoma (DLBCL) ranks the most common histologic subtype. DLBCL comprises a heterogeneous group with varied clinical and molecular features and different prognoses, despite uniform treatment. Recognition of the biological heterogeneity of DLBCL is thus of clinical importance [1]. The International Prognostic Index (IPI) is currently the most well-established index for risk stratification of DLBCL patients [2]. However, this prognostic index has limitations in reflecting the biologic or genetic features of DLBCL. Even within the same IPI risk group, substantial variability in clinical outcome has been observed. A significant improvement in overall survival of DLBCL has been achieved since rituximab (R) was used in combination with cyclophosphamide, doxorubicin, vincristine, and prednisone (CHOP) as the first-line chemotherapy regimen [3]. However, the prognosis in high-risk DLBCL patients remains dismal. Identification and validation of novel prognostic biomarkers may contribute to better stratification of DLBCL and guide optimal treatment.

To date, several prognostic biomarkers of lymphoma have been investigated, in which C-MYC is one of the most prominent factors [4]. The nature of the C-MYC aberrations included gene translocation, gene amplification, and C-MYC mRNA or C-MYC protein overexpression. C-MYC gene translocation is a hallmark of Burkitt lymphoma and can be detected in 5–17% of DLBCL patients [5]. Despite the advances achieved in the assessing prognostic significance of C-MYC in DLBCL, recent studies have implied the prognostic value of C-MYC was complicated by other factors and is far from straightforward. For example, C-MYC gene translocation was frequently found in DLBCL with concurrent translocations of BCL2 and/or BCL6, referred to as “double-hit” or “triple-hit” lymphomas, and has a dismal prognosis [6]. Although some studies have shown that the presence of C-MYC aberrations was significantly associated with shorter survival in DLBCL [7,8,9,10,11,12,13,14,15,16,17,18,19,20,21,22,23,24], other studies failed to show such an association between C-
MYC and worse prognosis [23,26,27,28,29,30]. Therefore, the role of C-MYC as independent prognostic factors needs to be further addressed in well-designed clinical trials. On the other hand, although numerous studies had been conducted to explore the prognostic values of C-MYC, no meta-analysis has assessed the predictive role of C-MYC aberrations in DLBCL. In the present study, we have conducted the first comprehensive systematic review and meta-analysis regarding the impact of C-MYC aberrations on DLBCL patients.

Materials and Methods

Selection criteria
A literature search was conducted in PUBMED, EMBASE, and COCHRANE databases. All the studies published before 31 January 2014 were included. Search criteria used synonyms of the following terms variably combined: C-MYC, prognosis and diffuse large B-cell lymphoma. The search was restricted to human studies with no language limitations.

According to the PRISMA guidelines [31], studies included in this meta-analysis should meet the following criteria: (i) C-MYC aberrations in adult patients with primary DLBCL had been examined by fluorescent in situ hybridization (FISH), immunohistochemistry (IHC) or other techniques; (ii) detailed survival information was available; and (iii) the median follow-up time exceeded one year. In the screening and eligibility stage, patients with evidence of an indolent lymphoma, human immunodeficiency virus infection, or primary central nervous system disease were excluded. Because compared to other DLBCL patients, patients with evidence of these factors mentioned above had some distinct features. Moreover, all studies were carefully evaluated to identify duplicate patient populations. Criteria used to determine duplicate populations included the study period, hospital, treatment information, and any additional inclusion items. However, subsequent reports containing new data on prognostic factors or survival were also incorporated into pooled analyses of the specific point.

Quality assessment
Two investigators (KGZ and DMX) independently evaluated the methodological quality of the studies twice, applying the Newcastle-Ottawa Quality Assessment scale for case-control and cohort studies [32]. According to the quality scales, if a study met a requirement, then it gained a score of 1; otherwise, it gained a score of 0. REMARK guidelines, which provide a useful start for assessing tumor prognostic markers, were used to help identify study bias [33]. When discrepancies between investigators occurred, a third investigator (MH) conducted an additional evaluation.

Data extraction
Baseline characteristics of the included studies, such as follow-up time, were independently recorded on a spreadsheet. In each

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Figure 1. PRISMA flow chart of study selection.
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| Study     | Year  | Region   | Number of patients | Detection rate | Detection method          | Main treatment       | Outcome   | Median follow-up months (minimum, maximum) | HR | Study quality |
|-----------|-------|----------|--------------------|----------------|---------------------------|----------------------|-----------|--------------------------------------------|----|---------------|
| Klapper   | 2008  | Germany  | 177                | 7.9%           | FISH                      | CHOP-like            | EFS, OS   | 29 (1, 71)                                 |    | 9             |
| Yoon      | 2008  | Korea    | 156                | 16.1%          | FISH                      | Mainly CHOP-like     | OS        | 42 (3, 76)                                 |    | 7             |
| Niitsu    | 2009  | Japan    | 252                | 11%            | Conventional G-banding technique | CycloOBEAP or CHOP-like | EFS, OS   | 64 (18–90)                               |    | 7             |
| Savage    | 2009  | Canada   | 135                | 8.8%           | FISH                      | R-CHOP               | EFS, OS   | 36 (0.8, 84)                               |    | 8             |
| Barrans   | 2010  | Britain  | 245                | 14%            | FISH                      | R-CHOP               | OS        | 24 (1, 41)                                 |    | 7             |
| Zhang     | 2011  | China    | 106                | 12.3%          | FISH                      | Mainly CHOP-like     | OS        | 35 (4, 104)                               |    | 6             |
| Hummel    | 2006  | Germany  | 146                | 16.6%          | FISH                      | Mix treatment        | OS        | 60 (0.1, 209)                             |    | 8             |
| Cuccuini  | 2012  | France   | 161                | 17%            | FISH                      | R-DHAP or R-ICE      | EFS, OS   | 30 (2, 76)                                 |    | 8             |
| Green     | 2012  | Denmark  | 193                | 11%            | FISH                      | R-CHOP               | OS        | 47 (1, 102)                               |    | 8             |
| Kramer    | 1998  | Netherlands | 151               | 7%             | Southern blot             | CHOP-like            | OS        | 40 (1, 96)                                 |    | 7             |
| Akasaka   | 2000  | Japan    | 203                | 11.8%          | LD-PCR                    | CHOP-like            | OS        | 49 (29, 118)                              |    | 8             |
| Akyurek   | 2012  | Turkey   | 239                | 6%             | FISH                      | R-CHOP               | OS        | 26 (2, 96)                                 |    | 7             |
| Kawasaki  | 2001  | Japan    | 137                | 10.2%          | Southern blot             | CHOP-like            | OS        | 25 (0.1, 99)                              |    | 8             |
| McClure   | 2005  | America  | 76                 | 5%             | FISH                      | Mix treatment        | OS        | 32 (1, 219)                               |    | 8             |
| Saez      | 2003  | Spain    | 48                 | 27%            | RT-PCR                    | CHOP                 | EFS, OS   | 62 (12, 110)                              |    | 7             |
| Rimsza    | 2008  | Mix      | 208                | NA             | qNPA                      | R-CHOP and CHOP analyzed separately | OS        | 26 (0.6, 70)                              |    | 8             |
| Kojima    | 2013  | Japan    | 100                | 10%            | FISH                      | R-CHOP               | OS        | 49 (2, 118)                               |    | 8             |
| Horn      | 2013  | Germany  | 442                | 31.8%/8.8%     | IHC/FISH                  | Mix treatment        | EFS, OS   | 29 (4, 64)                                 |    | 8             |
| Johnson   | 2012  | Mix      | 307                | 33%/NA         | IHC/Microarray            | R-CHOP               | OS        | 49 (6, 136)                               |    | 8             |
| Aukema    | 2013  | Germany  | 562                | 11.5%          | FISH                      | Mix treatment        | OS        | 63 (3, 120)                               |    | 8             |
| Perry     | 2014  | Mix      | 106                | 65%            | IHC                       | Mix treatment        | OS        | 54 (7, 145)                               |    | 8             |
| Tzankovet | 2013  | America  | 432                | 9%             | FISH                      | R-CHOP               | EFS       | 50 (10, 95)                               |    | 8             |
| Kluk      | 2012  | America  | 56                 | 17.8%          | IHC                       | R-CHOP               | OS        | 42 (2, 87)                                 |    | 9             |
| Gupta     | 2012  | America  | 24                 | 29%/10.4%      | IHC/FISH                  | R-CHOP and epratuzumab | EFS       | 24 (3, 55)                                |    | 7             |

Abbreviations: HR, hazard ratio; OS, overall survival; EFS, event-free survival; IHC, immunohistochemistry; FISH, interphase fluorescent in situ hybridization; RT-PCR, reverse transcription-polymerase chain reaction; qNPA, quantitative nucleic acid protection assay; LD-PCR, long-distance polymerase chain reaction; CHOP, cyclophosphamide, doxorubicin, vincristine, and prednisone; R-CHOP, rituximab, cyclophosphamide, doxorubicin, vincristine, and prednisone; R-DHAP, rituximab, dexamethasone, aracytine, and cisplatin; R-ICE, rituximab, ifosfamide, etoposide, and carboplatin; NA, not available.

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study, both overall survival (OS) and event-free survival (EFS) were considered endpoints for survival analysis. We assessed the prognostic impact of C-MYC using hazard ratios (HRs) and their 95% confidence intervals (95%CIs) as the main effect size estimate. For each study, HR was estimated by a method depending on the data provided in the publication. For those studies that reported the value of HR and its standard error straightforward, these data would be extracted directly. For those studies that did not report the HR but provided sufficient data on survival, the log HRs and variances were estimated based on the methodology published previously [34]. An HR greater than 1 implied a worse survival for those patients with C-MYC. Otherwise, if the value of HR was smaller than 1, a better prognosis was indicated for the group with C-MYC. Additionally, if a 95% CI for the hazard ratio included the null value of 1, then this estimate of HR was not statistically significant. If the value of HR could be obtained from both multivariate and univariate analyses, we extracted the HR from multivariate analyses.

The secondary effect size estimate was odds ratio (OR), which reflects the possible association between the previously well-known prognostic factors with C-MYC status in DLBCL, and \( P < 0.05 \) was considered statistically significant.

**Statistical analysis**

The procedure was conducted according to the Cochrane handbook, including a sensitivity test, heterogeneity test, publication bias test, and meta-regression, as well as subgroup analyses. Heterogeneity among studies was evaluated using the Cochran’s Q test as well as the \( I^2 \) index. If the heterogeneity was substantial \( (I^2 \geq 50\%) \), the random effects model was performed. Otherwise, the fixed effects model was used. Meta-regression analysis of multiple covariates was also performed to examine the sources of heterogeneity, particularly the differences in the length of follow-up time [35,36]. Begg’s and Egger’s test were used to reveal possible publication bias [37,38]. A sensitivity analysis was performed to assess the stability. The inverse of the estimates’ variance was used as study weight, such that larger studies tended to contribute more than smaller studies to the weighted average. The weighting factor was \( 1/\text{standard error}^2 \) in our study. All calculations were performed using Stata 12.0 (StataCorp, College Station, TX).

**Results**

**Characteristics of the included studies**

The selection procedure of eligible studies was shown as a flow chart (Figure 1). Twenty-four articles finally met the inclusion criteria, and their data were summarized in Table 1. The sample size in each study varied from 24 to 562, with a median follow-up time ranging from 24 to 64 months. Eight studies had reported the effect of C-MYC on EFS endpoints, and twenty-two studies on OS endpoints. Among these publications, the nature of the C-MYC aberrations included gene translocation, gene amplification, and C-MYC mRNA or C-MYC protein overexpression. The HRs and 95%CIs were therefore calculated according to the manifestation of C-MYC aberrations (gene, protein, and mRNA). In particular, Gupta et al. and Horn et al. evaluated the association of survival endpoints with both C-MYC gene translocation and C-MYC protein overexpression in DLBCL [11,26]. In Johnson et al. study, high levels of both C-MYC mRNA and C-MYC protein were both investigated [27]. Based on the nature of C-MYC aberrations, the prognostic values of C-MYC aberrations were analyzed in corresponding subgroups. The hazard ratios were carefully retrieved. Fifteen studies had the direct data for HR. For the other nine studies, the hazard ratios were extrapolated from the graphical survival distributions.

**Meta-analysis for the associations between previously well-known prognostic factors with C-MYC aberrations**

Six studies investigated the correlation of C-MYC with the cell-of-origin classified subgroups, including the germinal center B-cell-like (GCB) and non-GCB groups [39]. Meta-analysis revealed C-MYC were significantly more prevalent in the GCB type than in the non-GCB type \( (P = 0.0002) \). The correlations of the previously well-known prognostic factors with C-MYC aberrations are summarized in Table 2. Patients with C-MYC presented more frequently with poor performance status (PS), an elevated level of lactate dehydrogenase (LDH), a high Ki-67 proliferation index and poor IPI. However, the frequencies of extranodal lesion, Ann Arbor stages III–IV or bone marrow involvement were not significantly different between DLBCL patients with and without C-MYC aberrations \( (P > 0.05) \).

**Meta-analysis for the prognostic effects of C-MYC aberrations in DLBCL patients**

The forest plots of HRs and 95%CIs for OS or EFS endpoints in DLBCL patients are shown in Figure 2 and Figure 3. In particular, subgroup analyses were performed according to the subtypes of C-MYC aberrations (gene, protein, and mRNA) and treatment.

Initially, the prognostic value of C-MYC gene abnormalities was analyzed. Nineteen selected studies had investigated C-MYC gene abnormalities [7,8,9,10,11,12,13,15,16,17,20,21,22,23,25,26,28,29,30]. Among them, seventeen studies focused on only C-MYC gene translocation, whereas the other two studies examined...
both C-MYC gene amplification and translocation [9,30]. For C-MYC gene abnormalities, the overall HRs for OS were 2.22 (95% CI, 1.89 to 2.61) and for EFS were 2.29 (95% CI, 1.81 to 2.90) compared with the patients without C-MYC gene abnormalities. Among the studies that investigated the prognostic value of C-MYC gene translocation, five studies had further explored whether the addition of rituximab to CHOP could reduce the prognostic value of C-MYC [7,8,10,15,20]. Our study revealed that the overall HR for OS in the R-CHOP subgroup was 2.17 (95% CI, 1.62 to 2.91), similar with the overall HR for OS in the patients who were not treated with rituximab-containing regimen (detailed results were shown in Table 3), indicating that the inferior OS conferred by C-MYC gene translocation could not be overcome by rituximab.

Among the eligible studies, nine studies [8,9,13,15,17,20,21,26,30] that comprised 1774 patients altogether evaluated the prognostic effects of C-MYC among multivariate risk factors, such as age, IPI, and LDH. They used multivariable regression analysis to determine the independent prognostic value of C-MYC gene abnormalities. The conclusion of an ‘independent effect’ was based on the multivariate analysis. If the coefficient of a factor in a regression model did not change substantially after including other factors, it was defined as an independent factor. For this subgroup that used multivariable analysis, the pooled HR for OS was 2.51
(95% CI, 1.87 to 2.86) compared with that for the C-MYC negative patients.

Next, the prognostic value of C-MYC overexpression was analyzed. Five studies had analyzed the prognostic value of C-MYC protein overexpression [11,14,24,26,27], and three studies had investigated that of overexpression of C-MYC mRNA [18,19,27]. For C-MYC protein overexpression, the pooled HR for OS was 2.13 (95% CI, 1.55 to 2.91) and that for EFS was 2.21 (95% CI, 1.36 to 3.61). For C-MYC mRNA overexpression, the pooled HR for OS was 1.62 with a 95% CI from 1.30 to 2.02. Meta-analysis for EFS endpoints of C-MYC mRNA could not be performed because only one study was available. In the R-CHOP subgroup, C-MYC overexpression remained to a poor prognostic factor because the pooled HR for OS was 1.93 for protein overexpression [14,27] and 1.73 for mRNA overexpression [18,27].

Detailed results of subgroup analyses for C-MYC aberrations are listed in Table 3. An additional meta-regression analysis also confirmed the worse survival of C-MYC was not influenced by the difference in follow-up time (P > 0.05). Taken together, our study demonstrated that there was no significant heterogeneity for either EFS or OS (I² value < 50%). Publication bias tests showed no bias (P > 0.05). Exclusion of any single study did not alter the overall findings in the sensitivity test.

Systematic review of the prognostic role of C-MYC amplification and the isolated C-MYC aberrations (single-hit lymphoma) in DLBCL

Apart from chromosome translocation, gene amplification was another mechanism for C-MYC overexpression that had not been widely investigated. With respect to the role of C-MYC amplification, only a few studies with diverse designs were performed. Therefore, a qualitative systematic review, rather than a quantitative meta-analysis, was performed. Only three studies with a small sample size singled out the prognosis value of C-MYC amplification in DLBCL. In the two studies that evaluated the copy-number changes by FISH, C-MYC amplification appeared to be associated with a similarly poor prognosis as C-MYC translocation [9,30]. However, in another study that detected C-

Figure 3. Forest plots of hazard ratios (HRs) and 95% confidence intervals (95% CIs) for event-free survival endpoints in DLBCL patients with C-MYC gene abnormalities (A), overexpression of C-MYC mRNA (B) and C-MYC protein (C). Squares represent the HR of each study, and the area of each square was proportional to the weight of each study in the meta-analysis; Horizontal lines, 95% CIs; Closed diamond, pooled HRs with their 95% CIs.
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Table 3. Subgroup analyses for the prognostic values of C-MYC aberrations in DLBCL patients.

| Endpoints                              | Studies | Pooled HRs | 95% CI       | $I^2$ value | Publication bias |
|----------------------------------------|---------|------------|--------------|-------------|------------------|
| **C-MYC gene abnormalities**           |         |            |              |             |                  |
| OS (translocation)                     | 17      | 2.22       | [1.89, 2.61] | 0%          | $P_{Begg} = 0.56$; $P_{Egger} = 0.45$ |
| OS (translocation and amplification)   | 2       | 1.88       | [1.18, 3.00] | 0%          | $P_{Begg} = 0.32$; $P_{Egger} = 1.00$ |
| OS (R-CHOP)                            | 5       | 2.17       | [1.62, 2.91] | 0%          | $P_{Begg} = 0.09$; $P_{Egger} = 0.06$ |
| OS (adjusted)                          | 9       | 2.31       | [1.87, 2.86] | 4.3%        | $P_{Begg} = 0.25$; $P_{Egger} = 0.08$ |
| EFS (translocation)                    | 7       | 2.44       | [1.87, 3.18] | 8.7%        | $P_{Begg} = 0.07$; $P_{Egger} = 0.06$ |
| EFS (translocation and amplification)  | 1       | 1.80       | [1.08, 3.01] | NA          | NA               |
| EFS (R-CHOP)                           | 4       | 3.18       | [2.09, 4.84] | 0%          | $P_{Begg} = 0.63$; $P_{Egger} = 0.77$ |
| EFS (without R)                        | 2       | 2.63       | [1.65, 4.17] | 0%          | $P_{Begg} = 0.32$; $P_{Egger} = 1.00$ |
| **C-MYC mRNA overexpression**          |         |            |              |             |                  |
| OS                                     | 3       | 1.62       | [1.30, 2.02] | 0%          | $P_{Begg} = 0.09$; $P_{Egger} = 0.07$ |
| OS (R-CHOP)                            | 2       | 1.73       | [1.27, 2.36] | 0%          | $P_{Begg} = 0.32$; $P_{Egger} = 1.00$ |
| EFS                                    | 1       | 2.94       | [1.22, 7.07] | NA          | NA               |
| **C-MYC protein overexpression**       |         |            |              |             |                  |
| OS                                     | 4       | 2.13       | [1.55, 2.91] | 0%          | $P_{Begg} = 0.09$; $P_{Egger} = 0.24$ |
| OS (R-CHOP)                            | 2       | 1.93       | [1.23, 3.05] | 2.2%        | $P_{Begg} = 0.32$; $P_{Egger} = 1.00$ |
| EFS                                    | 2       | 2.21       | [1.36, 3.61] | 0%          | $P_{Begg} = 0.32$; $P_{Egger} = 1.00$ |

Abbreviations: HR, hazard ratio; OS, overall survival; EFS, event-free survival; CI, confidence interval; R-CHOP, rituximab, cyclophosphamide, doxorubicin, vincristine, and prednisone; without R, treatment without rituximab; NA, not available.

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MYC amplification by array-based comparative genomic hybridization, extra copies of C-MYC were associated with poor OS and progression-free survival only in the present of concomitant del(8p), but not in all DLBCL patients [40].

Similarly, to find whether presence of double or triple hits might influence the outcome, a qualitative systematic review was performed to investigate the prognostic effects of isolated C-MYC aberrations (single-hit lymphoma). Generally, two comparison methods were applied to demonstrate the role of isolated C-MYC aberrations in DLBCL. Six studies compared single-hit patients with complex-hit cases. Among them, one study showed the complex-hit group had a worse prognosis than the single-hit group [17]; however, other studies found no difference between the two subgroups [9,12,22,26,30]. Furthermore, another three studies directly compared single-hit patients with C-MYC-negative patients. Two of them found the presence of C-MYC alone retained unfavorable prognostic significance [12,20], but another study did not [41]. Comparisons among different studies were hampered by only a few studies available, with the diverse study designs as well as the small-size cases.

Discussion

Thus far, the present study is the first systematic review and meta-analysis about the prognostic value of C-MYC in DLBCL. The current meta-analysis integrated 4662 DLBCL cases and strongly confirmed the role of C-MYC as a prognostic factor in DLBCL. To increase accuracy, studies were divided into several subgroups according to the nature of C-MYC aberrations and treatment. Based on this meta-analysis, several critical issues had been addressed. First, DLBCL patients with C-MYC were proven to be associated with several adverse clinical features. Second, the addition of rituximab did not seem to overcome the inferior outcome conferred by C-MYC. Moreover, after pooling the HRs adjusted by multivariate Cox models from nine studies, C-MYC appeared to maintain its independent prognostic value, regardless of other well-established factors. In light of meta-regression and subgroup analysis, the pooled results appeared not to be influenced by the length of follow-up time. In addition, the median follow-up time in all the included studies exceeded 24 months. C-MYC aberrations represented the most reproducible biomarker with unfavorable prognosis regardless of the biological test used. Screening of C-MYC should have definite prognostic meaning for DLBCL stratification, thus guaranteeing more tailored therapy.

Some questions remain uncertain regarding the prognostic role of C-MYC aberrations in DLBCL. First, a qualitative systematic review was performed on the role of C-MYC amplification due to the rarity and heterogeneity of the selected studies. It had been postulated that overexpression of C-MYC by amplification had a similar effect to up-regulation by C-MYC translocation in DLBCL [42,43,44]. However, different studies with small-size cases appeared to be inconsistent. Similarly, the prognostic effects of isolated C-MYC aberrations (single-hit lymphoma) also remained controversial due to the limited studies with inconsistent conclusion. In the future, prospective studies with large sample size or a patient-level meta-analysis need to be conducted to address these issues.

Second, assessment of C-MYC aberrations in pathology specimens is becoming increasingly important in the routine clinical practice. Statistical analysis has demonstrated there is a significant correlation between deregulation of C-MYC protein and mRNA with C-MYC gene abnormalities [43,44,45]. Unfor-
References

1. Sehn LH (2012) Paramount prognostic factors that guide therapeutic strategies in diffuse large B-cell lymphoma. Hematology Am Soc Hematol Educ Program 2012: 402–409.

2. The International Non-Hodgkin's Lymphoma Prognostic Factors Project (1993) A predictive model for aggressive non-Hodgkin's lymphoma. N Engl J Med 329: 987–994.

3. Coufler B, Lepage E, Briere J, Herbrecht R, Tilly H, et al. (2002) CHOP chemotherapy plus rituximab compared with CHOP alone in elderly patients with diffuse large-B-cell lymphoma. N Engl J Med 346: 235–242.

4. Perry AM, Mitrovic Z, Chan WC (2012) Biological prognostic markers in diffuse large B-cell lymphoma. Cancer Control 19: 214–226.

5. Slack GW, Gascoyne RD (2011) MYC and aggressive B-cell lymphomas. Adv Anat Pathol 18: 219–226.

6. Pfreundschuh M (2012) Growing importance of MYC/BCL2 immunohistochemistry in diffuse large B-cell lymphomas. J Clin Oncol 30: 3433–3435.

7. Akyurek N, Uner A, Benekli M, Barista I (2012) Prognostic significance of MYC, BCL2, and BCL6 rearrangements in patients with diffuse large B-cell lymphoma treated with cyclophosphamide, doxorubicin, vincristine, and prednisone plus rituximab. Cancer 118: 4175–4183.

8. Barrans S, Crouch S, Smith A, Turner K, Owen R, et al. (2010) Rearrangement of MYC is associated with poor prognosis in patients with diffuse large B-cell lymphoma treated in the era of rituximab. J Clin Oncol 28: 3360–3365.

9. Cuccurini W, Briere J, Mouneir N, Voelker HU, Rosenwald A, et al. (2012) MYC+ diffuse large B-cell lymphoma is not salvaged by classical R-CHOP or R-DHAP followed by BEAM plus autologous stem cell transplantation. Blood 119: 4619–4624.

10. Green TM, Young KH, Visco C, Xu-Monette ZY, Orazi A, et al. (2012) Immunohistochemical double-hit score is a strong predictor of outcome in patients with diffuse large B-cell lymphoma treated with rituximab plus cyclophosphamide, doxorubicin, vincristine, and prednisone. J Clin Oncol 30: 3460–3467.

11. Gupta M, Maurer MJ, Wellik JK, Law ME, Han J, et al. (2012) Expression of Myc but not pSTAT3, is an adverse prognostic factor for diffuse large cell lymphoma treated with etoposide+R-CHOP. Blood 119: 4400–4406.

12. Hamiel M, Bentink S, Berger H, Klapper W, Wessendorf S, et al. (2006) A biologic definition of Burkitt’s lymphoma from transcriptional and genomic profiling. N Engl J Med 354: 2419–2430.

13. Klapper W, Stoecklein H, Zeynalova S, Ott G, Kosari F, et al. (2008) Structural aberrations affecting the MYC locus indicate a poor prognosis independent of clinical risk factors in diffuse large B-cell lymphomas treated within randomized trials of the German High-Grade Non-Hodgkin’s Lymphoma Study Group (DSHNHL). Leukemia 22: 2226–2229.

14. Kluk MJ, Chalupski B, Sinha P, Roy A, Dal Cin P, et al. (2012) Immunohistochemical detection of MYC-driven diffuse large B-cell lymphomas. Plasr Onc 7: e3813.

15. Kojima N, Nishikawa H, Takizawa J, Aoki S, Noguchi M, et al. (2013) MYC rearrangements are useful for predicting outcomes following rituximab and chemotherapy: multivariate analysis of Japanese patients with diffuse large B-cell lymphoma. Leuk Lymphoma 54: 2149–2154.

16. McClure RF, Remstein ED, Macon WR, Dewald GW, Habermann TM, et al. (2005) Adult B-cell lymphomas with Burkitt-like morphology are phenotypically...
31. Moher D, Liberati A, Tetzlaff J, Altman DG (2009) Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. PLoS Med 6: e1000097.
32. Stang A (2010) Critical evaluation of the Newcastle-Ottawa scale for the assessment of the quality of nonrandomized studies in meta-analyses. Eur J Epidemiol 25: 603–605.
33. McShane LM, Altman DG, Sauerbrei W, Taube SE, Gion M, et al. (2005) Reporting recommendations for tumor marker prognostic studies. J Clin Oncol 23: 9067–9072.
34. Parmar MK, Torri V, Stewart L (1998) Extracting summary statistics to perform meta-analyses of the published literature for survival endpoints. Stat Med 17: 2815–2834.
35. Zhang LQ, Wang J, Jiang F, Xu L, Liu FY, et al. (2012) Prognostic value of survivin in patients with non-small cell lung carcinoma: a systematic review with meta-analysis. PLoS One 7: e34100.
36. Moja L, Piatti A, Pecoraro V, Ricci G, Virgili G, et al. (2012) Timing matters in hip fracture surgery: patients operated within 48 hours have better outcomes. A meta-analysis and meta-regression of over 190,000 patients. PLoS One 7: e46175.
37. Begg CB, Mazumdar M (1994) Operating characteristics of a rank correlation test for publication bias. Biometrics 50: 1088–1101.
38. Egger M, Davey Smith G, Schneider M, Minder C (1997) Bias in meta-analysis detected by a simple, graphical test. BMJ 315: 629–634.
39. Hans CP, Weisenburger DD, Greiner TC, Gascoyne RD, Delahie J, et al. (2004) Confirmation of the molecular classification of diffuse large B-cell lymphoma by immunohistochemistry using a tissue microarray. Blood 103: 275–282.
40. Testoni M, Kwee I, Greiner TC, Montes-Moreno S, Vose J, et al. (2011) Gains of MYC locus and outcome in patients with diffuse large B-cell lymphoma treated with R-CHOP. Br J Haematol 155: 274–277.
41. Visco G, Tzankov A, Xu-Monette ZY, Miranda RN, Tai YC, et al. (2013) Patients with diffuse large B cell lymphoma of germinal center origin with BCL2 translocations have poor outcome, irrespective of MYC status: a report from an International DLBCL rituximab-CHOP Consortium Program Study. Haematologica 98:255–263.
42. Mosaffa H, Damotte D, Jenabian A, Delarue R, Vincenoneau A, et al. (2006) Non-Hodgkin’s lymphomas with Burkitt-like cells are associated with c-Myc amplification and poor prognosis. Leuk Lymphoma 47: 1893–1899.
43. Staik CJ, Nita H, Zhang W, Mosher CH, Cook JR, et al. (2010) Increased MYC gene copy number correlates with increased mRNA levels in diffuse large B-cell lymphoma. Haematologica 95: 597–603.
44. Valentino C, Kendrick S, Johnson N, Gascoyne R, Chan WC, et al. (2013) Colorimetric in situ hybridization identifies MYC gene signal clusters correlating with increased copy number, mRNA, and protein in diffuse large B-cell lymphoma. Am J Clin Pathol 139: 242–254.
45. Green TM, Nielsen O, de Stricker K, Xu-Monette ZY, Young KH, et al. (2012) High levels of nuclear MYC protein predict the presence of MYC rearrangement in diffuse large B-cell lymphoma. Am J Surg Pathol 36: 612–619.