Targeted inhibition of mitochondrial Hsp90 suppresses localised and metastatic prostate cancer growth in a genetic mouse model of disease

BACKGROUND: The molecular chaperone heat shock protein-90 (Hsp90) is a promising cancer drug target, but current Hsp90-based therapy has so far shown limited activity in the clinic.

METHODS: We tested the efficacy of a novel mitochondrial-targeted, small-molecule Hsp90 inhibitor, Gamitrinib (GA mitochondrial matrix inhibitor), in the Transgenic Adenocarcinoma of the Mouse Prostate (TRAMP) model. The TRAMP mice receiving 3-week or 5-week systemic treatment with Gamitrinib were evaluated for localised or metastatic prostate cancer; prostatic intraepithelial neoplasia (PIN) or localised inflammation using magnetic resonance imaging, histology and immunohistochemistry. Treatment safety was assessed histologically in organs collected at the end of treatment. The effect of Gamitrinib on mitochondrial dysfunction was studied in R1M1 cells isolated from TRAMP tumours.

RESULTS: Systemic administration of Gamitrinib to TRAMP mice inhibited the formation of localised prostate tumours of neuroendocrine or adenocarcinoma origin, as well as metastatic prostate cancer to abdominal lymph nodes and liver. The Gamitrinib treatment had no effect on PIN or prostatic inflammation, and caused no significant animal weight loss or organ toxicity. Mechanistically, Gamitrinib triggered acute mitochondrial dysfunction in R1M1 cells, with loss of organelle inner membrane potential and release of cytochrome-c in the cytosol.

CONCLUSIONS: The Gamitrinib has pre-clinical activity and favourable tolerability in a genetic model of localised and metastatic prostate cancer in immunocompetent mice. Selective targeting of mitochondrial Hsp90 could provide novel molecular therapy for patients with advanced prostate cancer.

Keywords: prostate cancer; metastasis; mitochondria; Hsp90; TRAMP

Despite advances in treating early-phase prostate cancer (Carter et al., 2006), advanced disease, characterised by castration resistance and bone metastases, poses significant therapeutic challenges (Taichman et al., 2007), with over 30,000 deaths in the United States alone. Cytotoxic or radiation has limited efficacy in these patients, and molecular therapies are still in early stages of evaluation (Vogiatzi et al., 2009). Because advanced prostate cancer is heterogeneous (Taylor et al., 2010), targeting so-called ‘nodal’ cancer genes (Lamb et al., 2006) overseeing multiple downstream pathways of tumour maintenance (Butcher, 2005) may offer concrete therapeutic prospects. In this context, the molecular chaperone heat shock protein-90 (Hsp90) is a nodal cancer gene (Whitesell and Lindquist, 2005), controlling the folding and/or maturation of client proteins involved in tumour cell proliferation, survival and adaptation (Trepel et al., 2010). The Hsp90 has been intensely pursued for cancer therapeutics, and several small-molecule antagonists of its ATPase pocket have been developed (Drysdale and Brough, 2008). However, Hsp90-based therapy (Kim et al., 2009) has shown so far modest activity in patients with epithelial malignancies, including prostate cancer (Heath et al., 2008), whether as single agent (Solit et al., 2008) or combined with cytotoxics (Tse et al., 2008) or molecular therapies (Modi et al., 2007).

A key feature of Hsp90 and Hsp90-like molecules is their localisation to multiple subcellular compartments (Trepel et al., 2010). Recently, a pool of Hsp90 has been found in mitochondria of tumour cells (Kang et al., 2007), where it promotes cell survival by antagonising cyclophilin D (CypD)-dependent organelle permeability transition and apoptosis (Green and Kroemer, 2004). Whether this pathway (Kang et al., 2007) influences the response to Hsp90-based therapy in the clinic (Trepel et al., 2010) is
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Currently unknown, but it is intriguing that none of the Hsp90 ATPase antagonists currently in preclinical development (Drysdale and Brough, 2008) has the ability to accumulate in mitochondria (Kang et al, 2009), thus leaving unscathed this general survival mechanism. To address this limitation, a new class of small-molecule Hsp90 inhibitors selectively targeted to mitochondria, that is, Gamitrinibs (GA mitochondrial matrix inhibitors) was recently generated (Kang et al, 2009), which exhibited potent anti-cancer activity in various xenograft tumour models, in vivo (Kang et al, 2009).

In this study, we evaluated the pre-clinical activity of Gamitrinib in the Transgenic Adenocarcinoma of the Mouse Prostate (TRAMP) model (Greenberg et al, 1995). Albeit with limitations (Pienta et al, 2008), prostate tumourigenesis in TRAMP mice recapitulates many aspects of the human disease on an immunocompetent background (Kaplan-Leffko et al, 2003), and is suitable for cancer drug discovery, in vivo (Zorn et al, 2007).

MATERIALS AND METHODS

Cells and reagents

RM1 cells derived from TRAMP mice have been described (Voeks et al, 2002). The chemical synthesis, HPLC profile and mass spectrometry of mitochondria-targeted small-molecule Hsp90 antagonists, Gamitrinibs, have been reported (Kang et al, 2009). This study utilised Gamitrinib–G4 (G–G4), which contains the Hsp90 ATPase inhibitory structure of 17-AAG (LC-Laboratories, Woburn, MA, USA) linked to four tandem repeats of guanidinium, used as a mitochondrial moiety (Kang et al, 2009).

The Gamitrinib treatment of TRAMP mice

All experiments involving animals were approved by an Institutional Animal Care and Use Committee. The TRAMP model has been described (Kaplan-Leffko et al, 2003), and involves expression of the SV40 large T and small t oncogene in the prostatic epithelium under the control of the minimal −426/+28 rat probasin promoter (Greenberg et al, 1995). Transgene expression is regulated by androgen, and results in a spectrum of lesions, including prostatic intraepithelial neoplasia (PIN), invasive adenocarcinoma, neuroendocrine tumours and metastases to loco-regional abdominal lymph nodes, liver and lungs (Greenberg et al, 1995; Kaplan-Leffko et al, 2003). Female TRAMP mice on a C57BL/6 background were bred with non-transgenic male pups to genotyped by PCR amplification of tail genomic DNA using transgene-specific primers. To test a potential anti-cancer activity of Gamitrinib (Kang et al, 2009) in this model, TRAMP mice were divided into two age groups to receive G–G4 monotherapy at 5 mg/kg in cremophor as i.p. injections, with the schedule 3 days on/2 days off. Mice in group 1 (short-term treatment) received G–G4 starting at 21.9 weeks of age for 3 weeks (24.9 weeks of age), with analysis of primary and metastatic prostate cancer as end point. Animals in group 2 (long-term treatment) were started on G–G4 at 14.7 weeks of age for 5 weeks (19.7 weeks of age) and assessed histologically for primary prostate cancer growth, PIN and localized inflammation.

Magnetic resonance imaging

The 2T magnetic resonance imaging (MRI) analysis of TRAMP mice was carried out on a Bruker/General Electric CSI-II 2.0 T/45 cm imaging spectrometer (Madison, WI, USA) equipped with a thermostatically controlled animal holder and gas anaesthesia, containing magnetic field gradients, RF phase, amplitude control and automatic shimming. For these experiments, 20-week-old non-castrated TRAMP mice were imaged by MRI using the following parameters: repetition time (TR) = 2000/600 ms; echo time (TE) = 25 ms; data acquisition field-of-view = 40 mm × 40 mm/30 mm × 30 mm; slice thickness (ST) = 1 mm; data acquisition matrix = 256 × 256; number of echo averages = 4; and display FO = 30 mm × 30 mm.

Analysis of mitochondrial dysfunction

RM1 cells isolated from TRAMP prostate tumours (Voeks et al, 2002) were incubated with 20 μM G–G4 or non-mitochondrially targeted Hsp90 inhibitor, 17-AAG, and analysed after 12 h for changes in mitochondrial membrane potential by JC-1 (200 μM) staining and multiparametric flow cytometry on a FACScalibur (Becton Dickinson, Franklin Lake, NJ, USA), as described (Kang et al, 2009). Alternatively, cytosolic extracts were isolated from treated RM1 cells using an ApoAlert Cell Fractionation Kit (Clontech, Otsu, Shiga, Japan), and analysed by western blotting.

Histology

The TRAMP mice in control or G–G4-treated groups were killed, and the entire genitourinary tract containing seminal vesicles, prostate (including dorsal, lateral, ventral and anterior lobes) and urethra (thus excluding the urinary bladder), was isolated, fixed and stained with hematoxylin/eosin. In some experiments, tissue sections were stained with an antibody to the proliferation-associated marker, Ki67, as described (Kang et al, 2009). At the end of the experiment, organs from control or G–G4-treated TRAMP mice were removed, paraffin embedded and analysed by H&E staining and light microscopy. The histological analysis in each case was performed by a veterinary pathologist (DSG), and the percentage of prostate gland involvement with PIN, adenocarcinoma or neuroendocrine tumours was assessed in individual prostatic lobes. The scoring system used to quantify inflammation or metastatic prostate cancer was as follows: 0, none; 1, mild; 2, moderate; and 3, marked.

Statistical analysis

Data were analysed using the unpaired t-test on a GraphPad software package (Prism 4.0, La Jolla, CA, USA) for Windows. All statistical tests were two sided. A P-value of 0.05 was considered to be statistically significant.

RESULTS

Prostate tumourigenesis in TRAMP mice

We began this study by quantifying prostate cancer growth in untreated TRAMP mice (Greenberg et al, 1995). Consistent with previous reports (Kaplan-Leffko et al, 2003), TRAMP mice at 22 weeks of age exhibited enlarged prostates, by MRI (Supplementary Figure 1). Prostatic lesions under these conditions included well-differentiated adenocarcinomas with low proliferative index as well as large neuroendocrine tumours, composed of sheets of small, undifferentiated cells that stained intensely for the proliferation-associated marker, Ki-67 (Supplementary Figure 1).

Gamitrinib inhibits localised prostate cancer growth in TRAMP mice

Consistent with these data, prostatic samples from untreated TRAMP mice harvested at 19.7 weeks of age (group 2) revealed the presence of neuroendocrine tumours, adenocarcinomas and PIN lesions, mixed with various degrees of local inflammation (Figure 1). Large neuroendocrine tumours occupying >50% of a prostatic lobe were observed in dorso-lateral and ventral prostatic samples (Figure 1A and B), whereas adenocarcinomas...
Figure 1C and D) were histologically well differentiated, of smaller size, that is, 5–25% of a prostatic lobe and equally distributed in dorso-lateral, ventral and anterior prostate. A complete histopathological analysis of control group 2 TRAMP mice is presented in Table 1.

Long-term treatment of group 2 TRAMP mice with G–G4 suppressed the growth of localised prostate cancer of both neuroendocrine and adenocarcinoma origin (Supplementary Figure 2 and Figure 2). Conversely, G–G4 treatment had no effect on localised prostatic inflammation in TRAMP mice, whereas it moderately but significantly increased the incidence and distribution of PIN lesions compared with age-matched control TRAMP mice (Supplementary Figure 2 and Figure 2). A complete histopathological characterisation of Gamitrinib-treated group 2 TRAMP mice is shown in Table 2.

Gamitrinib inhibits metastatic prostate cancer in TRAMP mice

Histological analysis of untreated group 1 TRAMP mice at 24.9 weeks of age revealed the presence of large neuroendocrine tumours in the dorso-lateral and ventral prostate, and well-differentiated adenocarcinomas in various prostatic lobes (Table 3). In all, 6 out of 10 of these animals also presented moderate prostate cancer dissemination to liver and loco-regional abdominal lymph nodes (Table 3 and Figure 2), consistent with previous observations (Hsieh et al., 2007). In contrast, none of the age-matched G–G4-treated animals in group 1 (0 out of 4 mice) had metastatic prostate cancer in liver or abdominal lymph nodes (Figure 2). Histological examination of brain, kidneys or lungs in these mice was also negative (not shown).

Safety of long-term Gamitrinib treatment in TRAMP mice

Both groups of TRAMP mice given Gamitrinib exhibited no significant weight loss throughout treatment (Supplementary Figure 3A). Similarly, organs harvested at the end of treatment from group 2 TRAMP mice were histologically unremarkable (Supplementary Figure 3B) compared with age-matched untreated mice (not shown).

'Mitochondriotoxic' activity of Gamitrinib

To begin elucidating the mechanism of anti-cancer activity of Gamitrinib in the TRAMP model, we next used RM1 cells that are derived from TRAMP tumours (Voeks et al., 2002). Treatment

| Mouse no. | Neuroendocrine (%) | Adenocarcinoma (%) | PIN (%) | Inflammation (Score) |
|-----------|--------------------|--------------------|---------|----------------------|
| 1137      | 100 (DLP, VP)      | 0                  | 10 (AP) | 0                    |
| 1145      | 100 (DLP, VP)      | 0                  | 10 (AP) | 0                    |
| 1150      | 75 (DLP)           | 0                  | 10 (AP) | 0                    |
| 1158      | 0                  | 25 (DLP)           | 50 (DLP); 50 (VP); 10 (AP) | +1 (DLP); +1 (VP) |
| 1208      | 0                  | 25 (DLP); 5 (VP)   | 50 (DLP); 75 (VP); 10 (AP) | +2 (DLP); +1 (VP) |
| 1211      | 0                  | 20 (DLP); 5 (AP)   | 70 (DLP); 100 (VP); 10 (AP) | +1 (VP); +1 (AP) |
| 1219      | 50 (VP)            | 5 (DLP)            | 75 (DLP); 50 (VP); 25 (AP) | +1 (DLP); +1 (AP) |
| 1224      | 0                  | 10 (DLP); 20 (VP)  | 60 (DLP); 80 (VP); 25 (AP) | +1 (VP) |

Abbreviations: AP = anterior prostate; DLP = dorso-lateral prostate; PIN = prostatic intraepithelial neoplasia; TRAMP = Transgenic Adenocarcinoma of the Mouse Prostate; VP = ventral prostate.
of inflammation (NS), PIN (score was determined, and expressed as arbitrary units (U). Quantification of prostatic lesions in representative matched samples of dorso-lateral prostate was carried out in TRAMP mice at 19.7 weeks of age (group 2). Quantification of metastasis to liver and loco-regional abdominal lymph nodes was determined in TRAMP mice at 24.9 weeks of age (group 1). Abbreviation: NS = not significant.

Table 2 Prostate histopathology of Gamitrinib–G4-treated TRAMP mice (group 2; 19.7 weeks)

| Mouse no. | Neuroendocrine (%) | Adenocarcinoma (%) | PIN (%) | Inflammation (Score) |
|-----------|--------------------|-------------------|---------|----------------------|
| 2999      | 0 (DLP)            | 99 (DLP)          | 0 (AP)  | +1 (VP)              |
| 4255      | 0 (DLP)            | 95 (DLP)          | 0 (AP)  | +2 (VP)              |
| 4260      | 0 (DLP)            | 100 (DLP)         | +1 (VP) |                      |
| 4464      | 0 (DLP)            | 95 (DLP)          | 25 (AP) | 0                    |
| 4473      | 0 (DLP)            | 100 (DLP)         | 0 (AP)  | 0                    |

Abbreviations: AP = anterior prostate; DLP = dorso-lateral prostate; Gamitrinib = GA mitochondrial matrix inhibitor; PIN = prostatic intraepithelial neoplasia; TRAMP = Transgenic Adenocarcinoma of the Mouse Prostate; VP = ventral prostate.

Table 3 Prostate histopathology of untreated TRAMP mice age matched to group 1 (24.9 weeks)

| Mouse no. | Neuroendocrine (%) | Adenocarcinoma | Metastasis |
|-----------|--------------------|----------------|------------|
| 1207      | 0                  | Well differentiated (DLP) | 0         |
| 1377      | 0                  | Well differentiated (DLP, AP) | + (Liver) |
| 1271      | 0                  | Well differentiated (DLP, AP) | + (Liver) |
| 1282      | 100 (DLP, VP)      | Well differentiated (AP) | + (Liver, lymph nodes) |
| 1295      | 100 (DLP, VP)      | 0               | + (Lymph nodes) |
| 1299      | 100                | 0               | + (Liver, lymph nodes) |
| 1300      | 0                  | Well differentiated (DLP) | 0         |
| 1265      | 0                  | Well differentiated (DLP) | 0         |
| 1381      | 0                  | Well differentiated (AP) | 0         |
| 1281      | 0                  | Well differentiated (AP) | 0         |

Abbreviations: AP = anterior prostate; DLP = dorso-lateral prostate; TRAMP = Transgenic Adenocarcinoma of the Mouse Prostate; VP = ventral prostate.

DISCUSSION

In this study, we have shown that systemic administration of Gamitrinib (Kang et al., 2009), a novel small molecule that targets exclusively the pool of Hsp90 in mitochondria (Kang et al., 2007), suppressed localised and metastatic prostate cancer growth in TRAMP mice (Greenberg et al., 1995), with no effect on PIN or local inflammation. Long-term systemic treatment of TRAMP mice with Gamitrinib was feasible, with no evidence of systemic or organ toxicity. Mechanistically, Gamitrinib functioned as a ‘mitochondriotoxic’ agent in the TRAMP model, triggering loss of organelle inner membrane potential and discharge of cytochrome-c in the cytosol.

Although still the backbone of cancer drug discovery, xenograft studies in immunocompromised mice have significant drawbacks (Kelland, 2004), as tumour growth in these settings does not recapitulate the complexity of clonal selection, cross-talk with the microenvironment, interplay of inflammatory responses and acquisition of metastatic traits. This has prompted renewed interest in exploiting genetically engineered mouse models for cancer drug discovery (Walrath et al., 2010), especially for prostate cancer, where cross-talk between the tumour cell population and its microenvironment has a critical role in progression to castration resistance and metastasis (Taichman et al., 2007). Despite its well-known limitations (Pienta et al., 2008), including the preponderance of neuroendocrine tumours compared with adenocarcinoma (Chiaverrotti et al., 2008), and the failure to metastasise to bones (Hsieh et al., 2007), prostatic tumourigenesis in TRAMP mice (Shappell et al., 2004) has provided a reliable genetic model for the human disease (Kaplan-Lefko et al., 2003), suitable for pre-clinical studies (Zorn et al., 2007).

Here, the anti-cancer activity of Gamitrinib in TRAMP mice extends recent studies in xenograft models (Kang et al., 2009), including prostate cancer, where systemic administration of Gamitrinib–TPP (Kang et al., 2009) suppressed the growth of subcutaneous or bone-localised PC3 prostate tumours in immunocompromised mice (Kang et al., 2010). In the TRAMP model, Gamitrinib–G4, which contains a structurally distinct mitochondrial-targeting moiety compared with Gamitrinib–TPP (Kang et al., 2009), was active across the spectrum of poorly differentiated, rapidly proliferating neuroendocrine tumours, as well as of differentiated adenocarcinoma. This is consistent with the abundant distribution of one of the targets of Gamitrinibs (Kang et al., 2009), the mitochondrial Hsp90 homologue TNF receptor-associated protein-1 (Trepel et al., 2010), in all Gleason grade localised and metastatic prostate cancer in humans, but not benign prostatic hyperplasia (Leav et al., 2010).

Consistent with our previous observations (Kang et al., 2009), the mechanism of action of Gamitrinib in the TRAMP model involved acute induction of mitochondrial dysfunction (Kang et al., 2010), with loss of organelle inner membrane potential and release of cytochrome-c in the cytosol (Green and Kroemer, 2004). This produces direct tumour cell killing by Gamitrinib, at variance with the mainly cytostatic activity of non-subcellularly targeted Hsp90 inhibitors (Kang et al., 2009). In prostate cancer, Gamitrinib-mediated killing indistinguishably affected androgen-dependent and -independent cell types (Kang et al., 2010; Leav et al., 2010), which may contribute to its activity against TRAMP tumours, often characterised by loss of androgen receptor (Huss et al., 2007) and androgen insensitivity (Kaplan-Lefko et al., 2003). With respect to the anti-metastatic activity of Gamitrinib in the TRAMP model,
it is possible that prostate cancer cells in the hypoxic environment of a metastatic niche, enriched in reactive oxygen species (Sung et al, 2008), may become especially ‘addicted’ to cytoprotection by mitochondrial Hsp90s (Kang et al, 2007). This model is consistent with an important role of CypD (Baines et al, 2005; Nakagawa et al, 2005) in mediating oxidative stress-induced mitochondrial permeability transition (Hua et al, 2007; Montesano Gesualdi et al, 2007), a cell death response antagonised by mitochondrial Hsp90s (Kang et al, 2007).

Long-term, continuous Gamitrinib treatment of TRAMP mice was feasible, devoid of systemic or organ side effects, in vivo. This tolerability likely reflects the low to undetectable expression of the targets of Gamitrinib, that is, mitochondrial Hsp90s, in most normal tissues, as opposed to tumours (Kang et al, 2007). This cytoprotective pathway may be also uniquely ‘wired’ in tumour cells, as suggested by the insensitivity of normal prostatic epithelium to Gamitrinib-mediated killing (Leav et al, 2010) and the lack of association between Hsp90s and CypD in mitochondria of normal tissues (Ghosh et al, 2010).

In sum, we have shown that one of the Gamitrinib variants, G–G4 (Kang et al, 2009), has activity in a pre-clinical genetic model of localised and metastatic prostate cancer in an immunocompetent background (Greenberg et al, 1995). Although additional work is required to define the drug-like properties of Gamitrinibs in anticipation of human testing, the data presented here suggest that selective suppression of mitochondrial Hsp90s may provide novel molecular therapy in prostate cancer, and improve the currently limited activity of Hsp90-based therapy in these patients (Trepel et al, 2010).

ACKNOWLEDGEMENTS

We thank Dr Christopher H Sotak (Worcester Polytechnic Institute) for MRI analysis. This work was supported by National Institutes of Health grants CA140043 (DCA, C-C H, LRL, JBL and GSS), CA78810, HL54131 and CA118005 (DCA).

Conflict of interest

The authors declare no conflict of interest.

Supplementary Information accompanies the paper on British Journal of Cancer website (http://www.nature.com/bjc)

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