Protective effects of carnosine on white matter damage induced by chronic cerebral hypoperfusion

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Graphical Abstract

Carnosine has extensive neuroprotective effects on white matter damage after chronic cerebral ischemia

Abstract

Carnosine is a dipeptide that scavenges free radicals, inhibits inflammation in the central nervous system, and protects against ischemic and hypoxic brain damage through its anti-oxidative and anti-apoptotic actions. Therefore, we hypothesized that carnosine would also protect against white matter damage caused by subcortical ischemic injury. White matter damage was induced by right unilateral common carotid artery occlusion in mice. The animals were treated with 200, 500 or 750 mg/kg carnosine by intraperitoneal injection 30 minutes before injury and every other day after injury. Then, 37 days later, Klüver-Barrera staining, toluidine blue staining and immunofluorescence staining were performed. Carnosine (200, 500 mg/kg) substantially reduced damage to the white matter in the corpus callosum, internal capsule and optic tract, and it rescued expression of myelin basic protein, and alleviated the loss of oligodendrocytes. However, carnosine at the higher dose of 750 mg/kg did not have the same effects as the 200 and 500 mg/kg doses. These findings show that carnosine, at a particular dose range, protects against white matter damage caused by chronic cerebral ischemia in mice, likely by reducing oligodendroglial cell loss.

Key Words: nerve regeneration; subcortical ischemic vascular dementia; carnosine; corpus callosum; neuron; internal capsule; oligodendrocyte; optic tract; white matter damage; neural regeneration

Introduction

Vascular dementia is the second most prevalent form of dementia (Kalaria et al., 2008). Vascular dementia is a clinicopathological entity that is distinct from Lewy body dementia, Alzheimer’s dementia and frontotemporal dementia. Subcortical ischemic vascular dementia (SIVD), induced by chronic cerebral hypoperfusion due to small-artery disease, is a common type of vascular dementia. It is often observed in patients with atherosclerosis or hypertension, as well as in the aging population (Areosa et al., 2005). Many factors, such as hypertension, hyperlipidemia, atrial fibrillation and diabetes, may increase the risk of the disease (Selnes and Vinters, 2006). However, the pathogenetic mechanisms of SIVD are unclear. The characteristic features of the disease include progressive demyelination, white matter damage and cognitive impairment. Computed tomography scans and magnetic resonance imaging frequently detect periventricular and subcortical white matter damage in SIVD patients (Hashimoto and Ikeda, 2015). This white matter damage has been found to be associated with the loss of myelin and the apoptotic death of oligodendrocytes (Johnson et al., 2013). The cerebral white matter contains oligodendrocytes and myelin sheaths formed...
by oligodendrocytes, which are sensitive to various adverse stimuli, including ischemia (Benarroch, 2009). Proteolipid protein (PLP) and myelin basic protein (MBP), which are found throughout the myelin sheath, have been demonstrated to be linked with vascular dementia (Barker et al., 2013).

Therapy for SIVD has received great attention (Olivo et al., 2012; Safouis et al., 2015). There are several compounds with different mechanisms of action that display mild efficacy in SIVD patients (Craig and Birks, 2006; Taguchi, 2011; Tomimoto, 2011). However, there are no drugs approved for halting the progression of SIVD (McGuinness et al., 2009). Vasodilators and acetylcholinesterase inhibitors are used for treating SIVD, but they exhibit limited efficacy (Craig and Birks, 2006). Therefore, it is imperative to develop more effective drugs for the prevention and treatment of SIVD.

Carnosine (β-alanyl-L-histidine) is a natural dipeptide found at high levels in the central nervous system (Usui et al., 2013). Carnosine has been assigned many putative functions, including free radical scavenger, anti-inflammatory agent and pH buffer (Kurata et al., 2006; Babizhayev and Yegorov, 2010). Carnosine protects mice and rats against ischemia-induced brain damage and hypoxia through its antioxidative actions and by attenuating apoptosis in transient cerebral ischemia (Jin et al., 2005; Shen et al., 2010). Therefore, we hypothesized that carnosine may protect against SIVD.

In the present study, experimental SIVD was induced by right unilateral common carotid artery occlusion, which better simulates the clinical cognitive impairment caused by white matter damage in SIVD (Yoshizaki et al., 2008; Ma et al., 2012). In addition, in our previous study (Ma et al., 2015), we showed that the pathological lesions after right unilateral common carotid artery occlusion share common features with SIVD. The aim of our study is to investigate the effect of carnosine on white matter damage in the SIVD model. Our novel model should help in the study of the mechanisms and therapy of SIVD.

Materials and Methods
Ethics statement and animals
The animal studies were approved by the committee for experimental animals of the Xinhua Hospital of Shanghai Jiao Tong University School of Medicine, and were performed in accordance with the National Institutes of Health Guide for the Care and Use of Laboratory Animals. Precautions were taken to minimize suffering and the number of animals used in each experiment. Fifty male C57BL/6 mice (specific-pathogen-free/viral-antibody-free), 8-weeks-old, weighing 22–30 g, were purchased from Charles River (license No. SCXK 2011-0011). Mice were housed under a 12-hour light/dark cycle, and were allowed free access to food and water. The mice were equally and randomly divided into five groups: sham, SIVD, SIVD + carnosine 200 mg/kg, SIVD + carnosine 500 mg/kg, SIVD + carnosine 750 mg/kg.

SIVD model establishment
After anesthesia with sodium pentobarbital (60 mg/kg, intraperitoneally), the right common carotid artery was isolated from the adjacent vagus nerve, and then double-ligated with 6-0 silk sutures to achieve right unilateral common carotid artery occlusion. Mice in the sham group were subjected to the same procedure, except for carotid ligation. The mortality rate in this model was 0% (Ma et al., 2012).

Carnosine administration
Carnosine (Sigma, St. Louis, MO, USA), dissolved in sterile saline, was administered by intraperitoneal injection. Adult male mice were administered saline (0.1 mL) in the sham and SIVD group, and carnosine in the SIVD + carnosine 200 mg/kg, SIVD + carnosine 500 mg/kg and SIVD + carnosine 750 mg/kg group, 30 minutes before surgery and every other day until the mice were sacrificed. Sodium pentobarbital (60 mg/kg) was used for anesthesia. The mice were sacrificed on day 37 after model establishment, and the brain tissues were removed for Klüver-Barrera and histochemical staining.

Klüver-Barrera staining
Mice were deeply anesthetized and perfused transcardially with 4% paraformaldehyde in 0.1 M phosphate-buffered saline (PBS, pH 7.4). The brains were removed quickly and stored in 4% paraformaldehyde at 4°C for 24 hours, and then in 30% sucrose for 3 days. Frozen brain sections (10-μm-thick) were made on a cryostat (SM2000R; LEICA, Wetzlar, Germany). The severity of white matter lesions was evaluated by examining fiber density by Klüver-Barrera staining, as in Waktia et al. (2008).

Toluidine blue staining
Toluidine blue staining was performed on day 37 after model establishment. Frozen brain sections (10-μm thick) were incubated three times with PBS, and stained in toluidine blue (Sigma, St. Louis, MO, USA) working solution for 2–3 minutes and washed in water. The sections were dipped in 0.5% hydrochloric acid/alcohol (75%) for 5 seconds. Finally, the sections were observed under a fluorescence microscope (BX51; Olympus, Tokyo, Japan).

Immunofluorescence staining
Individual brain sections were incubated with PBS containing 0.3% Triton X-100 and 3% normal donkey serum for 2 hours, and then with the appropriate primary antibodies overnight for 4°C as follows: rabbit anti-Oligo2 polyclonal antibody (1:500; Millipore, Billerica, MA, USA), rat anti-PLP polyclonal antibody (1:250; Millipore) and rat anti-PLP polyclonal antibody (1:250; Millipore). The sections were washed three times in PBS and incubated with Cy3-conjugated goat anti-rabbit IgG (1:400; Invitrogen, Carlsbad, CA, USA), Cy3-conjugated goat anti-rat IgG (1:400; Invitrogen) or fluorescein isothiocyanate (FITC)-conjugated goat anti-rat IgG (1:400; Invitrogen) for 2 hours at room temperature. Finally, the sections were observed under a fluorescence microscope.

Statistical analysis
Data are presented as the mean ± SEM. Statistical analyses were done with SPSS 11.5 for Windows (SPSS, Chicago, IL, USA). One-way analysis of variance followed by the least significant
Myelin damage in the corpus callosum was examined by Klüver-Barrera staining. Myelin density in the corpus callosum was greatly reduced on day 37 after model establishment, compared with the sham group (P < 0.01). Carnosine significantly elevated myelin density to 87.84% (200 mg/kg; P < 0.01, vs. SIVD group) and 93.39% (500 mg/kg; P < 0.001, vs. SIVD group) of that in the sham group, as assessed by Klüver-Barrera staining. However, carnosine at 750 mg/kg had no effect on myelin damage in the corpus callosum (P >

difference or Dunnett’s T3 *post-hoc* test (where equal variances were not assumed) was used for multiple comparisons.

Results

Effects of carnosine on myelin damage in the brain of SIVD model mice

Corpus callosum

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Optic tract

Myelin damage in the optic tract was examined by Klüver-Barrera staining. Myelin density in the optic tract was greatly lower at day 37 after model establishment compared with the sham group ($P < 0.01$). Carnosine at 500 mg/kg greatly increased the myelin density to the level in the sham group on day 37 after model establishment ($P < 0.01$, vs. SIVD group). However, carnosine at 200 and 750 mg/kg had no effect on myelin damage in the optic tract ($P > 0.05$, vs. SIVD group; Figure 1).

Internal capsule

Myelin damage in the internal capsule was examined by Klüver-Barrera staining. Compared with the sham group, myelin density in the internal capsule was reduced to 74.89% of that in the sham group on day 37 after model establishment ($P < 0.05$). Carnosine at 200 and 500 mg/kg markedly elevated myelin density to 87.17% and 87.83% ($P < 0.05$, vs. SIVD group; Figure 2).
SIVD group) of that in the sham group, respectively. However, carnosine at 750 mg/kg showed no such effect ($P > 0.05$, vs. SIVD group; Figure 3).

Effects of carnosine on MBP expression in the corpus callosum of SIVD model mice
MBP is a major component of myelin, and reduced expression of the protein is associated with demyelination (Barker et al., 2013). We analyzed MBP expression in the corpus callosum after model establishment by immunofluorescence analysis. MBP expression was greatly reduced 37 days after model establishment compared with the sham group ($P < 0.01$). Carnosine (200 and 500 mg/kg) significantly improved MBP expression after model establishment ($P < 0.01$ or 0.05, vs. SIVD group; Figure 4).

Effects of carnosine on PLP expression in the corpus callosum of SIVD model mice
PLP, a tetraspan membrane protein, is the most abundant component of central nervous system myelin (Barker et al., 2013). We analyzed PLP expression in the corpus callosum after model establishment using immunofluorescence staining. There was no obvious difference between the sham and SIVD groups in PLP fluorescence intensities ($P > 0.05$). Carnosine had no significant effect on PLP expression ($P > 0.05$, vs. SIVD group; Figure 5).

Effects of carnosine on the number of oligodendrocytes in the corpus callosum of SIVD model mice
We characterized the oligodendrocyte population on day 37 after model establishment, and we evaluated the effects of carnosine treatment. The number of oligodendrocytes (Oligo2$^+$ cells) was strikingly decreased on day 37 after model establishment ($P < 0.001$, vs. sham group). We found that the administration of carnosine (200 and 500 mg/kg) increased the number of oligodendrocytes in SIVD mice ($P < 0.05$ or 0.001, vs. SIVD group; Figure 6).

Lack of effect of carnosine on neuronal density in the brain of SIVD model mice
By toluidine blue staining, no obvious neuronal loss was observed in the corpus striatum (Figure 7) or hippocampus (Figure 8) after model establishment and carnosine treatment ($P > 0.05$, vs. sham group). We also examined neuronal density in the cortex, and obtained similar results (data not shown).

Discussion
In our previous studies, we have already found that after right unilateral common carotid arteries occlusion, carnosine significantly improved the learning and memory in the object recognition test, passive avoidance task and Morris water maze (Ma et al., 2012). In this study, we found that carnosine significantly ameliorated myelin damage in the corpus callosum, optic tract and internal capsule in the mouse model of SIVD. In addition, carnosine prevented the decrease in MBP expression, and it reduced oligodendrocyte loss, suggesting that it protects against white matter damage in SIVD. Here, we observed extensive white matter damage in SIVD induced by right unilateral common carotid artery occlusion, particularly in the corpus callosum, which accords with a previous report (Yoshizaki et al., 2008). The corpus callosum is a white matter structure that is highly susceptible to ischemic damage. The accentuated vulnerability of the corpus callosum in ischemia-related white matter damage has also been observed by others (Cheng et al., 2015; Park et al., 2015). Farkas et al. (2004) reported that the optic tract seems to be particularly vulnerable to ischemia induced by bilateral common carotid artery occlusion. The corpus callosum is supplied by arterial blood from the anterior cerebral arteries, while the optic nerves and tract receive arterial blood directly from branches of the internal carotid arteries, and the internal capsule receives arterial blood from the middle cerebral artery (Farkas et al., 2004). Because the compensatory redistribution in the circle of Willis after carotid artery occlusion is not immediate or complete, our results suggest that specific brain regions may suffer more ischemic damage in different ischemic models. Case studies demonstrate that single, strategically placed lesions may result in SIVD, and the internal capsule is a key site (Tatemichi et al., 1992; Chui, 2007). Moreover, significant white matter damage in the internal capsule was observed in chronic cerebral ischemic injury induced by right unilateral common carotid artery occlusion in this study. We found that carnosine significantly ameliorated white matter damage in the corpus callosum, optic tract and internal capsule, indicative of a robust neuroprotective effect of carnosine against white matter damage. Thus, our results further demonstrate that carnosine may have therapeutic potential for the treatment of SIVD.

At present, the pathogenesis of SIVD is not clear. Recently, magnetic resonance imaging in patients with SIVD showed that white matter damage is progressive, often appearing before obvious cognitive dysfunction (Cavallari et al., 2015). The clinical and pathological changes in vascular dementia are different from Alzheimer’s disease, Lewy body dementia and frontotemporal dementia. The most striking difference from Alzheimer’s disease is that damage is primarily to the white matter, instead of the grey matter (Hashimoto and Ikeda, 2015). In this study, by toluidine blue staining, no obvious neuronal loss or apparent morphological change was observed in the cortex, striatum or hippocampus after model establishment and carnosine treatment, which further demonstrates that pathological changes in vascular dementia are mainly localized to the white matter. However, our data do not exclude the occurrence of neuronal dysfunction in SIVD, which will require additional studies to clarify.

Autopsy findings show that the main pathological changes in SIVD include damage to nerve fibers and the myelin sheath, which is produced by oligodendrocytes (Alvarez-Sabin and Roman, 2011). Although the pathogenesis of the white matter damage is still unclear, it frequently coincides with myelin degradation. Oligodendrocytes, myelin-forming glial cells of the central nervous system, are known to be quite vulnerable to ischemic stress, resulting in the loss of myelin after ischemia or hypoxia (Benarroch, 2009; Ihara et
al., 2010). In the present study, we found that administration of carnosine (200 and 500 mg/kg) increased the number of oligodendrocytes after SIVD, which may contribute to its protective effect on the white matter, and was associated with more intense Klüver-Barrera staining and increased MBP expression. However, PLP expression was not changed on day 37 after model establishment, which suggests that PLP and MBP may play different roles in SIVD.

Although the neuroprotection by carnosine in SIVD has been investigated previously (Ma et al., 2012), the effects of carnosine at high doses had not been examined. Carnosine treatment has been reported to exhibit significant neuroprotection with wide therapeutic and clinically relevant time windows in both permanent and transient ischemic models (Bae et al., 2013; Bandyopadhyay et al., 2014; Dolu et al., 2014). Therefore, carnosine was considered well tolerated and without toxicity. However, we found that the neuroprotective effect of carnosine in SIVD is dose-dependent—the high dose of the dipeptide had no effect on white matter damage induced by right unilateral common carotid artery occlusion. Although the reason for this lack of effect is not clear, our study suggests that the administered dose is critical for neuroprotection. This finding should help in the development of treatment strategies involving carnosine.

In conclusion, carnosine displayed an extensive neuroprotective effect on white matter damage in the corpus callosum, optic tract and internal capsule induced by right unilateral common carotid artery occlusion, which may be mediated by its cytoprotection of oligodendrocytes. These findings suggest that carnosine may have therapeutic potential for the treatment of SIVD.

Author contributions: JM participated in literature search, study design, data collection, data analysis and data interpretation, and wrote the paper. SHB and XTL carried out data collection and analysis, and provided critical revision. AJX and JZ designed the research, supervised the entire research, performed experiments, analyzed data and prepared the paper. All authors approved the final version of the paper.

Conflicts of interest: None declared.

Plagiarism check: This paper was screened twice using Cross-Check to verify originality before publication.

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Figure 6 Effect of carnosine on oligodendrocyte damage in the corpus callosum.
(A) The total oligodendrocyte (Oligo2+) count after carnosine treatment was calculated as the percentage of total cells (labeled by DAPI) after right unilateral common carotid artery occlusion. Fluorescent indicator is Cy3 (red). Scale bar: 50 μm. (B) Quantitative analysis of Oligo2+ cells. The numbers of oligodendrocytes (Oligo2+ cells) was decreased on day 37 after artery occlusion, and was increased by administration of carnosine (200, 500 mg/kg). Data are expressed as the mean ± SEM. n = 8–10 mice in each group. ###P < 0.001, vs. sham group; *P < 0.05, ###P < 0.001, vs. SIVD group. I–V: Sham, SIVD, SIVD + carnosine 200 mg/kg, SIVD + carnosine 500 mg/kg and SIVD + carnosine 750 mg/kg groups, respectively. SIVD: Subcortical ischemic vascular dementia; DAPI: 4′,6-diamidino-2-phenylindole.

Figure 8 Effect of carnosine on neuronal density in the hippocampus in the mouse model of SIVD.
(A) Effect of carnosine on neuronal density in the hippocampus was evaluated after right unilateral common carotid artery occlusion. Scale bar: 50 μm. (B) Quantitative analysis of cell density. There was no difference in neuronal number among these groups. Data are expressed as the mean ± SEM. n = 8–10 mice in each group. I–V: Sham, SIVD, SIVD + carnosine 200 mg/kg, SIVD + carnosine 500 mg/kg and SIVD + carnosine 750 mg/kg groups, respectively. SIVD: Subcortical ischemic vascular dementia.

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