Association of APOE Serum Levels and APOE $\varepsilon2$, $\varepsilon3$, and $\varepsilon4$ Alleles with Optic Neuritis

Liucija Momkute, Alvita Vilkeviciute, Greta Gedvilaite, Gabriele Dubinskaite, Loresa Kriauciuniene and Rasa Liutkeviciene

Abstract: Optical neuritis (ON), otherwise known as optical nerve damage, is a term used to describe various environmental and body conditions that lead to optic nerve dysfunction. Neurologists are well aware of conditions that cause optic neuropathy, such as trauma, infections, malnutrition, and various toxins. As optic neuritis is a multifactorial demyelinating or infectious process, genetic predisposition may also influence the progression of optic neuritis. This study aimed to evaluate the association of ON (with and without multiple sclerosis) with APOE alleles and APOE serum levels. We found that the APOE $\varepsilon3/\varepsilon3$ genotype was statistically less common in the ON group of males than in the control group ($p = 0.045$). Moreover, the APOE $\varepsilon3/\varepsilon3$ genotype had a 3.7-fold increase in the odds of ON development in males (OR = 3.698; CI: 1.503–9.095; $p = 0.004$). In contrast, the APOE $\varepsilon3/\varepsilon4$ genotype had a 4.1-fold decrease in the odds of ON development in males (OR = 0.242; CI: 0.083–0.704; $p = 0.009$). APOE serum levels were statistically significantly higher in the ON group than in the control group ($p = 0.042$). The APOE $\varepsilon3/\varepsilon3$ genotype may increase males’ risk of developing ON, while the $\varepsilon3/\varepsilon4$ genotype may reduce males’ risk of developing ON.

Keywords: optic neuritis (ON); APOE; $\varepsilon3/\varepsilon3$

1. Introduction

Optic neuritis (ON) is an acute inflammatory disorder demyelinating the optic nerve. The isolated optic neuritis characteristics include unilateral, acute, and painful vision loss without systemic or other neurological symptoms [1]. In most cases, the etiology of the disease may be unknown, so it is called idiopathic ON [1]. The mechanism responsible for acute ON has not been definitively identified, but it is believed that an autoimmune reaction damages the myelin sheath that encompasses the neurons within the optic nerve [2]. When a disturbed nerve impulse causes an inflammatory response, type IV hyperallergic reactions occur. Activated peripheral T lymphocytes begin to secrete cytokines (inflammatory mediators), which easily cross the blood–cerebral protective barrier and cause myelin rupture. ON disease occurs when inflammatory mediators with T lymphocytes reach the central nervous system [3]. When the initial ON episode occurs, the patient experiences decreased color contrast sensitivity (depending on the period, blue and yellow in acute cases, and red and green in non-acute cases), photopsia (“flashes of light”), or impaired depth sensation in moving objects (Pulfrich phenomenon) [4,5]. Further tests are then applied to confirm the diagnosis of the disease. For evaluation, routine blood and magnetic resonance imaging, cerebrospinal fluid testing, and visual evoked potential tests are performed [1].

There is a close association between optic neuritis and multiple sclerosis (MS). MS is a chronic inflammatory autoimmune demyelinating disease that affects the central nervous system. Its mechanism of action is linked to the immune system; it destroys myelin and
forms scar tissue, which ultimately results in a disruption of nerve impulse transmission [6]. Studies have shown that ON occurs in approximately 70% of patients with MS [7]. Swelling of the optic disc may be observed during ophthalmoscopy. The retina is usually unchanged, but the resulting peripheral lining may be associated with an increased risk of multiple sclerosis. Therefore, optic neuritis may also be considered one of the manifestations of multiple sclerosis [8].

Optic neuritis is considered to be a multifactorial demyelinating process. The causes of the disease have also been examined at the genetic level, but they are not exactly known yet. APOE consists of 299 amino acids encoded in chromosome 19q13. The gene is involved in transporting fat molecules associated with low-density lipoproteins (LDLs) and very low-density lipoproteins (VLDLS). Studies have shown that the APOE gene is essential in the healing process of various CNSs and that this function is impaired when the protein exists in the ε4 isoform (encoded by the ε4 allele). Thus, we aimed to investigate the impact of APOE on ON development. A total of three isoforms are known to exist in the APOE gene. APOE2, APOE3, and APOE4 are the products of three alleles (ε2, ε3, and ε4) at a single gene locus. The ε3 isoform is considered usual. Higher frequencies of specific alleles are observed in certain diseases, e.g., ε4 in Alzheimer’s disease [9–13]. APOE2 has cysteine in both positions, APOE3 has cysteine in 112 positions and arginine in 158 positions, and APOE4 has arginine in both positions. These are the products of three alleles (ε2, ε3, and ε4) in a single gene locus. There are six genotypes in the general population: three homozygous (APOE ε2/ε2, APOE ε3/ε3, and APOE ε4/ε4) and three heterozygous (APOE ε2/ε3, APOE ε2/ε4, and 3 A). The ε3 allele is the most common and is found in 77% of the general population. Meanwhile, the ε2 allele is found in 8% of the general population, and the ε4 allele is found in 15% of the general population. However, the frequency of APOE alleles varies between different ethnic groups [9,10].

In the periphery, APOE is synthesized by the liver, macrophages, and astrocytes, which are transmitted to neurons through LDL receptors. APOE2 binds to LDL less strongly than APOE3 and APOE4. Consequently, APOE2 carries lipids less efficiently, and ε2 is homozygous for the risk of type III hyperlipoproteinemia. APOE4 binds to lipoprotein particles and, therefore, is associated with an increased risk of hypercholesterolemia and atherosclerosis, a faster progression of HIV infection, and accelerated telomere shortening. The ε4 allele is a vital genetic risk factor for late-onset Alzheimer’s disease and neurodegenerative diseases, such as cerebral amyloid angiopathy, Lewy cell dementia, and MS. APOE2 has been associated with white matter loss in the brain, traumatic brain healing, and hemorrhagic and ischemic stroke [11–15].

The apolipoprotein E gene has also been linked to the development of another pathology, perpetual macular degeneration. Studies have confirmed that the ε4 allele provides protective care by reducing the risk of early-onset macular degeneration (AMD) and late AMD [16,17] in individuals with a family history of AMD. Individuals with the ε4 allele have a lower risk of developing the disorder than those with the ε2 allele, whose risk becomes higher for the development of AMD [18,19].

We aimed to find an association between optic neuritis with and without multiple sclerosis and the APOE allele and APOE serum levels.

2. Materials and Methods

The permission for this study was obtained from the Ethics Committee for Biomedical Research (No. BE–2–102). This study was carried out in the Department of Ophthalmology and at the Institute of Neurosciences, the Lithuanian University of Health Sciences (LUHS).

From 1 January 2012 to 1 February 2022, patients with acute ON from the Department of Ophthalmology were enrolled. The study participants were 87 subjects with optic neuritis and 583 healthy control subjects.

The criteria for the inclusion of patients with optic neuritis were as follows:

- The first occurrence of an acute ON episode;
- Patient’s consent to participate in the study;
• Consultation with a neurologist;
• Neurological examination if multiple sclerosis has been diagnosed according to McDonald’s (2005) criteria.

The exclusion criteria for study patients were as follows:
• Other diseases related to the optic nerve;
• Systemic diseases, such as diabetes, chronic infectious diseases, and oncological diseases after transplantation.

Other criteria for the inclusion of the population not suffering from an attack of optic neuritis were as follows:
• A positive diagnosis of other ocular diseases (after ophthalmological examination);
• Exclusion of other chronic/acute systemic diseases;
• Consent to participate in the study.

The exclusion criteria for healthy subjects were as follows:
• A negative diagnosis of ocular disease or visual impairment;
• Therapeutic disorders of other systems.

The average age of the control group and patients with optic neuritis was statistically significantly different ($p < 0.05$). Thus, a further analysis was performed and adjusted by age and gender. The demographic data of the subjects are shown in Table 1.

### Table 1. Demographic characteristics of patients with optic neuritis (ON) and the control group subjects.

| Characteristics | ON Group ($n = 87$) | Control Group ($n = 583$) | $p$-Value |
|-----------------|---------------------|--------------------------|-----------|
| Gender          |                     |                          |           |
| Females, $n$ (%)| 56 (64.4)           | 276 (47.3)               | 0.003 *   |
| Males, $n$ (%)  | 31 (35.6)           | 307 (35.6)               |           |
| Age, years      |                     |                          |           |
| (Median, IQR)   | 33 (16)             | 58 (34)                  | $<0.001$ *|
| Multiple sclerosis |                 |                          |           |
| With            | 30 (34.5)           |                          |           |
| Without         | 46 (52.9)           |                          |           |
| Not determined  | 11 (12.6)           |                          |           |

* Mann–Whitney U test.

DNA was isolated from peripheral blood leukocytes using silica gel columns from a Thermo Scientific GeneJET Genomic DNA Purification Kit. This kit allows for the rapid and efficient extraction of high-quality DNA and is suitable for further RT-PCR assays. Two single-nucleotide polymorphisms of the $APOE$ gene (rs429358 and rs7412) were evaluated using the genotype sets C___3084793_20 and C____904973_10 (Applied Biosystems, Foster City, CA, USA). The program started by heating for 10 min at 95 °C, then heating for 15 s to 95 °C for another 40 heating cycles, and finally raising the temperature to 60 °C for another 1 min. A Human Apo E (AD2) ELISA Kit was used to assess serum APOE levels.

Statistical analysis was performed using IBM SPSS Statistics 27.0. The qualitative characteristics of the subjects are expressed in absolute numbers and percentages in parentheses. The hypothesis about the normal distribution of the measured trait values was tested using the Kolmogorov–Smirnov and Shapiro–Wolf tests. The following descriptive statistical characteristics used for the study did not meet the criteria for a normal distribution: median and IQR. The Mann–Whitney U test was used to assess the difference between the two independent groups. The distributions of $APOE$ genotypes and alleles in the ON and control groups were assessed using $\chi^2$ and Fisher unilateral criteria calculations. Moreover, Hardy–Weinberg’s equilibrium (HWE) was performed in the control group. Subjects were divided into two groups ($\leq$35 years old and $>$35 years old) based on the median age of all subjects to evaluate the distributions of $APOE$ genotypes and alleles within younger and
older subjects. Binary logistic regression was used to assess the influence of genotypes on ON development, indicating an OD of 95% confidence interval (CI). The differences were considered statistically significant when \( p < 0.05 \).

3. Results

\( APOE \) rs429358 and rs7412 genotyping was performed to evaluate the differences between the ON and control groups. Unfortunately, no statistically significant differences were found. The results are presented in Table 2.

Table 2. \( APOE \) genotype and allele frequencies in patients with ON and control groups.

| Genotype, Allele | ON (n = 87) | Control (n = 583) | \( p \)-Value |
|------------------|------------|------------------|--------------|
| \( APOE \varepsilon2/\varepsilon2, n (%) \) | 0 (0)      | 3 (0.5)          | -            |
| \( APOE \varepsilon2/\varepsilon3, n (%) \) | 14 (16.1)  | 84 (14.4)        | 0.774        |
| \( APOE \varepsilon2/\varepsilon4, n (%) \) | 1 (1.1)    | 17 (2.9)         | 0.342        |
| \( APOE \varepsilon3/\varepsilon3, n (%) \) | 57 (65.5)  | 387 (66.4)       | 0.874        |
| \( APOE \varepsilon3/\varepsilon4, n (%) \) | 15 (17.2)  | 85 (14.6)        | 0.516        |
| \( APOE \varepsilon4/\varepsilon4, n (%) \) | 0 (0)      | 7 (1.2)          | -            |
| \( \varepsilon2 \) allele, n (%) | 15 (8.6)   | 107 (9.2)        | 0.812        |
| \( \varepsilon3 \) allele, n (%) | 143 (82.2) | 943 (80.9)       | 0.681        |
| \( \varepsilon4 \) allele, n (%) | 16 (9.2)   | 116 (9.9)        | 0.756        |

Binary logistic regression was performed to evaluate the impact of \( APOE \) on the development of ON. No statistically significant results were found (Table 3).

Table 3. \( APOE \) binary logistic regression in patients with ON and control groups.

| Genotype | OR (95 %, CI) | \( p \)-Value |
|----------|---------------|--------------|
| \( APOE \varepsilon2/\varepsilon2 \) | -             | -            |
| \( APOE \varepsilon2/\varepsilon3 \) | 0.852 (0.440–1.652) | 0.636        |
| \( APOE \varepsilon2/\varepsilon4 \) | 1.285 (0.156–10.553) | 0.815        |
| \( APOE \varepsilon3/\varepsilon3 \) | 1.162 (0.669–1.930) | 0.563        |
| \( APOE \varepsilon3/\varepsilon4 \) | 0.765 (0.400–1.461) | 0.416        |
| \( APOE \varepsilon4/\varepsilon4 \) | -             | -            |

The \( APOE \) rs429358 and rs7412 polymorphisms were evaluated in the ON and control groups according to the gender of the subjects. The \( APOE \varepsilon3/\varepsilon3 \) genotype was statistically significantly less common in the ON group of males than in the control group of males (48.4% vs. 66.4%, \( p = 0.045 \)). The results are shown in Table 4.

Table 4. \( APOE \) genotype and allele frequencies in patients with ON and controls grouped by gender.

| Genotype, Allele | Males | Females | \( p \)-Value | Males | Females | \( p \)-Value |
|------------------|-------|---------|--------------|-------|---------|--------------|
| \( APOE \varepsilon2/\varepsilon2, n (%) \) | 0 (0) | 0 (0) | -            | 1 (0.3) | 1 (0.3) | -            |
| \( APOE \varepsilon2/\varepsilon3, n (%) \) | 7 (22.6) | 48 (15.6) | 0.318 | 7 (12.5) | 48 (15.6) | 0.547 |
| \( APOE \varepsilon2/\varepsilon4, n (%) \) | 1 (3.2) | 8 (2.6) | 0.838 | 0 (0) | 8 (2.6) | -            |
| \( APOE \varepsilon3/\varepsilon3, n (%) \) | 15 (48.4) | 204 (66.4) | 0.045 | 42 (75.0) | 204 (66.4) | 0.208 |
| \( APOE \varepsilon3/\varepsilon4, n (%) \) | 8 (25.8) | 43 (14.0) | 0.080 | 7 (12.5) | 43 (14.0) | 0.764 |
Table 4. Cont.

| Genotype, Allele | Males | | Females | |
|------------------|-------|---|---------|---|
|                  | ON (n = 31) | Control (n = 307) | p-Value | ON (n = 56) | Control (n = 276) | p-Value |
| APOE ε4/ε4, n (%) | 0 (0) | 3 (1.0) | - | 0 (0) | 3 (1.0) | - |
| ε2 allele, n (%)  | 8 (12.9) | 58 (9.4) | 0.382 | 7 (6.25) | 58 (9.4) | 0.276 |
| ε3 allele, n (%)  | 45 (72.6) | 499 (81.3) | 0.100 | 98 (87.5) | 499 (81.3) | 0.113 |
| ε4 allele, n (%)  | 9 (14.5) | 57 (9.3) | 0.186 | 7 (3.25) | 57 (9.3) | 0.298 |

A binary logistic regression analysis was performed in the ON and control groups to assess the effects of APOE genotypes on the incidence of ON by gender. We found that the APOE ε3/ε3 genotype is associated with a 3.7-fold increase in the odds of ON development in males (OR = 3.698; CI: 1.503–9.095; p = 0.004). In contrast, the APOE ε3/ε4 genotype is associated with a 4.1-fold decrease in the odds of ON development in males (OR = 0.242; CI: 0.083–0.704; p = 0.009). The results are shown in Table 5.

Table 5. APOE binary logistic regression in patients with ON and controls grouped by gender.

| Group | Genotype | OR (95 %, CI) | p-Value |
|-------|----------|---------------|---------|
| Males | APOE ε2/ε2 | - | - |
|       | APOE ε2/ε3 | 0.540 (0.190–1.539) | 0.249 |
|       | APOE ε2/ε4 | 0.258 (0.027–2.428) | 0.236 |
|       | APOE ε3/ε3 | 3.698 (1.503–9.095) | 0.004 |
|       | APOE ε3/ε4 | 0.242 (0.083–0.704) | 0.009 |
|       | APOE ε4/ε4 | - | - |
| Females | APOE ε2/ε2 | - | - |
|         | APOE ε2/ε3 | 1.125 (0.463–2.733) | 0.794 |
|         | APOE ε2/ε4 | - | - |
|         | APOE ε3/ε3 | 0.644 (0.329–1.261) | 0.200 |
|         | APOE ε3/ε4 | 1.359 (0.564–3.276) | 0.495 |
|         | APOE ε4/ε4 | - | - |

This study aimed to compare the genotypes and allele frequencies of the APOE in different groups of ON: ON without multiple sclerosis vs. the control group and ON with multiple sclerosis vs. the control group. However, no statistically significant differences were found between the groups. The data of the subjects are shown in Table 6.

Table 6. APOE genotype and allele frequencies in patients with ON with MS and without MS.

| Genotype, Allele | ON without MS (n = 46) | Control Group (n = 583) | p-Value | ON with MS (n = 30) | Control Group (n = 583) | p-Value |
|------------------|------------------------|------------------------|---------|---------------------|------------------------|---------|
| APOE ε2/ε2, n (%) | 0 (0) | 3 (0.5) | - | 0 (0) | 3 (0.5) | - |
| APOE ε2/ε3, n (%) | 7 (15.2) | 84 (14.4) | 0.881 | 4 (13.3) | 84 (14.4) | 0.870 |
| APOE ε2/ε4, n (%) | 1 (2.2) | 17 (2.9) | 0.771 | 0 (0) | 17 (2.9) | - |
| APOE ε3/ε3, n (%) | 32 (69.6) | 387 (66.4) | 0.659 | 22 (73.3) | 387 (66.4) | 0.431 |
| APOE ε3/ε4, n (%) | 6 (13.0) | 85 (14.6) | 0.776 | 4 (13.3) | 85 (14.6) | 0.850 |
| APOE ε4/ε4, n (%) | 0 (0) | 7 (1.2) | - | 0 (0) | 7 (1.2) | - |
| ε2 allele, n (%) | 8 (8.7) | 107 (9.2) | 0.878 | 4 (7.1) | 107 (9.2) | 0.605 |
Table 6. Cont.

| Genotype, Allele | ON without MS ($n = 46$) | Control Group ($n = 583$) | $p$-Value | ON with MS ($n = 30$) | Control Group ($n = 583$) | $p$-Value |
|------------------|--------------------------|---------------------------|-----------|----------------------|---------------------------|-----------|
| $\varepsilon3$ allele, $n$ (%) | 77 (83.7) | 943 (80.9) | 0.176 | 48 (85.8) | 943 (80.9) | 0.533 |
| $\varepsilon4$ allele, $n$ (%) | 7 (7.6) | 116 (9.9) | 0.467 | 4 (7.1) | 116 (9.9) | 0.491 |

A binary logistic regression analysis was performed in patients with ON without MS and with MS. No statistically significant differences were found between patients with ON without MS and patients with ON with MS. The results are shown in Table 7.

Table 7. APOE binary logistic regression in patients with ON with MS and without MS.

| ON Groups | Genotypes | OR (95 %, CI) | $p$-Value |
|-----------|-----------|---------------|-----------|
| ON with MS | $\text{APOE} \varepsilon2/\varepsilon2$ | - | - |
|           | $\text{APOE} \varepsilon2/\varepsilon3$ | 1.065 (0.354–3.270) | 0.910 |
|           | $\text{APOE} \varepsilon2/\varepsilon4$ | - | - |
|           | $\text{APOE} \varepsilon3/\varepsilon3$ | 0.781 (0.335–1.823) | 0.568 |
|           | $\text{APOE} \varepsilon3/\varepsilon4$ | 1.055 (0.349–3.189) | 0.925 |
| ON without MS | $\text{APOE} \varepsilon2/\varepsilon2$ | - | - |
|           | $\text{APOE} \varepsilon2/\varepsilon3$ | 0.940 (0.392–2.255) | 0.890 |
|           | $\text{APOE} \varepsilon2/\varepsilon4$ | 0.636 (0.067–5.353) | 0.677 |
|           | $\text{APOE} \varepsilon3/\varepsilon3$ | 0.996 (0.505–2.067) | 0.992 |
|           | $\text{APOE} \varepsilon3/\varepsilon4$ | 0.977 (0.387–2.467) | 0.961 |
|           | $\text{APOE} \varepsilon4/\varepsilon4$ | - | - |

Serum APOE levels were measured in the ON ($n = 20$) and healthy subject ($n = 20$) groups. An analysis was performed using the Mann–Whitney U test. A statistically significant difference was found between the groups (median (IQR): 136.65 (75.71) vs. 107.89 (25.14), $p = 0.042$). APOE serum levels were statistically significantly higher in the ON group than in the control group. The results are shown in Figure 1.

Figure 1. APOE Serum levels in control and ON groups.
4. Discussion

This study examined single-nucleotide polymorphisms rs429358 and rs7412 in the APOE gene in 87 patients with ON and 583 healthy subjects (without other ophthalmological and systemic diseases). HWE was performed in the control group (n = 583). The APOE rs429358 and rs7412 genotype distributions were consistent with HWE (p = 0.828 and 0.233, respectively). The frequency of genotypes and alleles was analyzed, but there were no statistically significant differences between the ON and control groups. A further analysis of SNPs was performed to evaluate the impact of age on ON development; unfortunately, it did not reveal any statistically significant differences. In contrast, statistically significant differences were found when the analysis was based on gender (p < 0.05). The APOE ε3/ε3 genotype was statistically significantly less frequent in males with ON than in control group males (p = 0.045). Moreover, the APOE ε3/ε3 genotype was associated with a 3.7-fold increase in the odds of ON development in males (p = 0.004). In contrast, the APOE ε3/ε4 genotype was associated with a 4.1-fold decrease in the odds of ON development in males (p = 0.009).

Many studies have analyzed the association of APOE with MS, but the data are contradictory. Many studies support the notion that the APOE ε4 allele is not associated with a higher risk of developing MS [20–39]. However, other studies suggest the opposite, stating that patients carrying the APOE ε4 allele may have decreased neuronal regeneration, leading to a faster progression of MS [27,40–43]. Moreover, a study conducted by Oliveri and co-authors found that the APOE gene−491A/T polymorphism is associated with severe cognitive impairment in patients with MS with a homozygous AA genotype [34]. Other researchers have found that APOE ε4 carriers are more likely to develop a more severe form of MS, while APOE ε2 carriers are more likely to develop a milder form of MS [44,45]. Julian and co-authors conducted a study to elucidate the association of the APOE gene with depression, which often occurs in patients with MS. Their results concluded that the APOE ε2 allele shows a protective effect by reducing the incidence of depressive disorders [46].

Only one study conducted by Pinholt et al. analyzed the association of APOE gene single-nucleotide polymorphisms with optic neuritis and multiple sclerosis. The study concluded that the APOE gene does not affect the development of either ON or MS [31]. The study looked for associations between the ε4 allele of the APOE gene and ON or MS. No statistically significant difference in alleles was found between genders. Researchers concluded that Apo E genotypes do not influence the development of MS or ON, but the Apo epsilon4 allele seems to predispose carriers with MS to a faster progression of disease [31].

However, Lazo et al. (a Mexican study) (abstract) examined 58 patients with ON and found no association between APOE and ON development [47]. In contrast, we found that the APOE ε3/ε3 genotype may reduce males’ risk of developing ON. Moreover, we found that APOE serum levels were statistically significantly higher in the ON group than in the control group (median (IQR): 136.65 (75.71) vs. 107.89 (25.14), p = 0.042). In comparison, a study conducted by Tamam et al. revealed that male patients with MS were likely to have lower serum APOE levels and that the most common APOE genotype in patients with MS was ε3/ε3 (82.0%). Male patients with MS were significantly more likely to have ε4, and at baseline, the disease duration was shorter, the Expanded Disability Status Scale (EDSS) scores were higher, the serum total APOE levels were lower, and the PI was significantly higher. The MS onset age, clinical types, EDSS scores, and PI had no significant correlation with the ε4 allele. Visual onset and sensory onset are good prognostic factors. There were no patients with visual onset and only a few patients with sensory onset in the ε4-positive group [48]. No statistically significant associations were found between APOE and ON with or without MS, but further and larger scale studies are needed to evaluate the associations of APOE SNPs and APOE serum levels with ON (with and without MS). Moreover, the links between APOE gene polymorphisms, Alzheimer’s disease, and cardiovascular disease have been extensively studied. There is scientific evidence that the ε4 allele of this gene is involved in the pathogenesis mechanism and influences the
development of the mentioned diseases [9–15]. Kulminski et al. found that the APOE gene, along with five other single-nucleotide polymorphisms, influenced the pathogenesis of Alzheimer’s disease [49].

The strengths and limitations of our study are as follows: this is the first study to examine the relationship between APOE serum levels and APOE ε2, ε3, and ε4 alleles with ON and MS in the Baltic states, and this is the first study to divide patients into two groups, namely, patients that only had isolated optic neuritis and patients that had optic neuritis and multiple sclerosis.

We confirmed that the APOE ε3/ε3 genotype might increase ON development risk in males, while the ε3/ε4 genotype may reduce males’ risk of developing ON, as APOE serum levels were statistically significantly higher in the ON group than in the control group. However, because of a large number of SNPs, it is necessary to conduct replication studies in the future, particularly with a bigger sample size, to confirm the association between SNPs and serum levels and ON development.

5. Conclusions

The APOE ε3/ε3 genotype may increase ON development risk in males, while the ε3/ε4 genotype may reduce males’ risk of developing ON. APOE serum levels were statistically significantly higher in the ON group than in the control group.

**Author Contributions:** Conceptualization, L.M., G.D., A.V., G.G., L.K. and R.L.; methodology, A.V. and G.G.; software, A.V. and G.G.; validation, A.V. and G.G.; formal analysis, A.V. and G.G.; investigation, L.M., G.D., A.V., G.G., L.K. and R.L.; resources, R.L.; data curation, A.V. and G.G.; writing—original draft preparation, A.V. and G.G.; writing—review and editing, A.V., G.G. and R.L.; visualization, A.V., G.G. and R.L.; supervision, R.L.; project administration, A.V. and G.G. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research received no external funding.

**Institutional Review Board Statement:** The study was conducted according to the guidelines of the Declaration of Helsinki and approved by the Ethics Committee of the Lithuanian University of Health Sciences (No. BE–2–102).

**Informed Consent Statement:** Informed consent was obtained from all subjects involved in the study.

**Data Availability Statement:** Data will be provided if a request is made by editors, reviewers, or scientists.

**Conflicts of Interest:** The authors declare no conflict of interest.

**References**

1. Kale, N. Optic neuritis as an early sign of multiple sclerosis. *Eye Brain.* 2016, 8, 195–202. [CrossRef] [PubMed]
2. Guier, C.P.; Stokkermans, T.J. Optic Neuritis. In *StatPearls*; StatPearls Publishing: Treasure Island, FL, USA, 2021.
3. Banevicius, M.; Vilkeviciute, A.; Glebauskiene, B.; Kriauciuniene, L.; Liutkeviciene, R. Association of Optic Neuritis with CYP4F2 Gene Single Nucleotide Polymorphism and IL-17A Concentration. *J. Ophthalmol.* 2018, 2018, 1686297. [CrossRef] [PubMed]
4. Pau, D.; Al, Z.N.; Yalamanchili, S.; Plant, G.T.; Lee, A.G. Optic neuritis. *Eye* 2011, 25, 833–842. [CrossRef]
5. Weerasinghe, D.; Lueck, C. Mimics and chameleons of optic neuritis. *Pract. Neurol.* 2016, 16, 96–110. [CrossRef]
6. Nicholas, R.; Rashid, W. Multiple sclerosis. *Am. Fam. Phys.* 2013, 87, 712–714.
7. Bennett, J.L. Optic Neuritis. *Continuum* 2019, 25, 1236–1264. [CrossRef]
8. Theodoridou, A.; Settas, L. Demyelination in rheumatic diseases. *J. Neurol. Neurosurg. Psychiatry* 2006, 77, 290–295. [CrossRef] [PubMed]
9. Diptiranjan, S.; Harshitha, S.M.; Sibin, M.K.; Arati, S.; Chetan, G.K.; Bhat, D.I. Role of APOE and IL18RAP gene polymorphisms in cervical spondylotic myelopathy in Indian population. *J. Clin. Neurosci.* 2019, 66, 83–86. [CrossRef] [PubMed]
10. Sidaraite, A.; Vilkeviciute, A.; Glebauskiene, B.; Kriauciuniene, L.; Zaliuniene, D.; Liutkeviciene, R. Association of ApoE haplotype with clinical evidence of pituitary adenoma. *Gene* 2019, 706, 154–161. [CrossRef]
11. Ghebranious, N.; Ivacic, L.; Mallum, J.; Dokken, C. Detection of ApoE E2, E3 and E4 alleles using MALDI-TOF mass spectrometry and the homogeneous mass-extend technology. *Nucleic Acids Res.* 2005, 33, e149. [CrossRef]
12. Armstrong, R.A. Risk factors for Alzheimer’s disease. *Folia Neuropathol.* 2019, 57, 87–105. [CrossRef] [PubMed]
13. Iacono, D.; Felitis, G.C. Impact of Apolipoprotein E gene polymorphism during normal and pathological conditions of the brain across the lifespan. *Aging* **2019**, *11*, 787–816. [CrossRef] [PubMed]

14. Chapleau, R.R.; Martin, C.A.; Hughes, S.R.; Baldwin, J.C.; Sladky, J.; Sherman, P.M.; Grinkemeyer, M. Apolipoprotein E promoter genotypes are not associated with white matter hyperintensity development in high-altitude careers. *BMC Res. Notes* **2019**, *12*, 630. [CrossRef] [PubMed]

15. Zhong, L.; Xie, Y.Z.; Cao, T.T.; Wang, Z.; Wang, T.; Li, X.; Shen, R.C.; Xu, H.; Bu, G.; Chen, X.-F. A rapid and cost-effective method for genotyping apolipoprotein E gene polymorphism. *Mol. Neurodegener.* **2016**, *11*, 2. [CrossRef] [PubMed]

16. Klaver, C.C.; Kliken, M.; van Duijn, C.M.; Hofman, A.; Crutsch, M.; Grobbe, D.E.; van Broeckhoven, C.; de Jong, P.T.V.M. Genetic association of apolipoprotein E with age-related macular degeneration. *Am. J. Hum. Genet.* **1998**, *63*, 200–206. [CrossRef]

17. Souied, E.H.; Benliah, P.; Amouyel, P.; Feingold, J.; Lagarde, J.P.; Munnich, A.; Kaplan, J.; Coscas, G.; Soubbrane, G. The epsilon 4 allele of the apolipoprotein E gene as a potential protective factor for exudative age-related macular degeneration. *Am. J. Ophthalmol.* **1998**, *125*, 353–359. [CrossRef]

18. Lee, S.H.; Schloss, G.J.; Swain, J.L. Maintenance of vascular integrity in the embryo requires signaling through the fibroblast growth factor receptor. *J. Biol. Chem.* **2006**, *275*, 33679–33687. [CrossRef]

19. Davignon, J.; Cohn, J.S.; Mabile, L.; Bernier, L. Apolipoprotein E and atherosclerosis: Insight from animal and human studies. *Clin. Chim. Acta* **1999**, *28*, 115–143. [CrossRef]

20. Santos, M.; Costa, M.D.C.; Rio, M.E.; Sa’, M.J.; Monteiro, M.; Valenca, A.; Sá, M.J.; Monteiro, M.; Valença, A.; Sá, A.; et al. Genotypes at the Apo E and SCA2 loci do not predict the course of multiple sclerosis in patients of Portuguese origin. *Mult. Scler.* **2004**, *10*, 153–157. [CrossRef]

21. Kantarci, O.H.; Hebrink, D.D.; Achenbach, S.J.; Pittock, S.J.; Altintas, A.; Schaefer-Klein, J.L.; Atkinson, E.J.; de Andrade, M.; McMurray, C.T.; Rodriguez, M.; et al. Association of APOE polymorphisms with disease severity in MS is limited to women. *Neurology* **2004**, *62*, 811–814. [CrossRef]

22. Weatherby, S.J.; Mann Cl Fryer, A.A.; Strange, R.C.; Hawkins, C.P. No association between the APOE ε4 allele and outcome and susceptibility in progressive primary multiple sclerosis. *J. Neurol. Neurosurg. Psychiatry* **2000**, *68*, 532. [CrossRef]

23. Weatherby, S.J.; Mann, C.L.; Davies, M.B.; Carthy, D.; Fryer, A.A.; Boggild, M.D.; Young, C.; Strange, R.C.; Ollier, W.; Hawkins, C.P. Polymorphisms of apolipoprotein E; outcome and susceptibility in multiple sclerosis. *Mult. Scler.* **2000**, *6*, 32–36. [CrossRef] [PubMed]

24. Niino, M.; Kikuchi, S.; Fukazawa, T.; Yabe, I.; Tashiro, K. Polymorphisms of apolipoprotein E and Japanese patients with multiple sclerosis. *Mult. Scler.* **2003**, *9*, 382–386. [CrossRef] [PubMed]

25. Ballerini, C.; Campani, D.; Rombola, G.; Gran, B.; Nacmias, B.; Amato, M.P.; Siracusa, G.; Bartolozza, L.; Sorbi, S.; Massacesia, L. Association of apolipoprotein E polymorphism to clinical heterogeneity of multiple sclerosis. *Neurosci. Lett.* **2000**, *296*, 174–176. [CrossRef]

26. Carlin, C.; Murray, L.; Graham, D.; Doyle, D.; Nicoll, J. Involvement of Apolipoprotein E in multiple sclerosis: Absence of remyelination associated with possession of the APOE ε2 allele. *J. Neuropathol. Exp. Neurol.* **2000**, *59*, 361–367. [CrossRef]

27. Chapman, J.; Sylantiev, C.; Nisipeanu, P.; Korczyn, A.D. Preliminary observations on APOE ε4 allele and progression of disability in multiple sclerosis. *Arch. Neurol.* **1999**, *56*, 1484–1487. [CrossRef]

28. Chapman, J.; Vinokurov, S.; Achiron, A.; Karussis, D.M.; MitosekSzewczyk, K.; Birnbaum, M.; Michaelson, D.M.; Korczyn, A.D. APOE genotype is a major predictor of long-term progression of disability in MS. *Neurology* **2001**, *56*, 312–316. [CrossRef]

29. Enzinger, C.; Ropole, S.; Strasser-Fuchs, S.; Kapeller, P.; Schmidt, H.; Poltrum, B.; Schmidt, R.; Hartung, H.-P.; Fazekas, F. Lower levels of N-acetulaspartate in multiple sclerosis patients with the apolipoprotein E ε4 allele. *Acta Neurol. Scand.* **2003**, *60*, 65–70. [CrossRef]

30. Evangelou, N.; Jackson, M.; Beeson, D.; Palace, J. Association of the APOE ε4 allele with disease activity in multiple sclerosis. *J. Neurol. Neurosurg. Psychiatry* **1999**, *67*, 203–205. [CrossRef]

31. Pinholt, M.; Frederiksen, J.L.; Andersen, P.S.; Christiansen, M. Apo E in multiple sclerosis and optic neuritis: The apo E-epsilon4 allele is associated with progression of multiple sclerosis. *Mult. Scler.* **2005**, *11*, 511–515. [CrossRef]

32. Ferri, C.; Sciaccia, F.L.; Veglia, F.; Martinelli, F.; Comi, G.; Canal, N.; Grimaldi, L.M.E. APOE ε4–ε2 and/491 polymorphisms are not associated with MS. *Neurology* **1999**, *53*, 888–889. [CrossRef] [PubMed]

33. Gervais, A.; Gaillard, O.; Plassart, E.; Reubol, J.; Fontaine, B.; Schuller, E. Apolipoprotein E polymorphism in multiple sclerosis. *Ann. Clin. Biochem.* **1999**, *35*, 135–136. [CrossRef] [PubMed]

34. Oliveri, R.L.; Cittadella, R.; Sibilia, G.; Manna, I.; Valentino, P.; Gambardella, A.; Aguglia, U.; Zappia, M.; Romeo, N.; Andreoli, V.; et al. APOE and risk of cognitive impairment in multiple sclerosis. *Acta Neurol. Scand.* **1999**, *100*, 290–295. [CrossRef] [PubMed]

35. Pirttilä, T.; Haanpää, M.; Mehta, P.D.; Lehtimäki, T. Apolipoprotein E (APOE) phenotype and APOE concentrations in multiple sclerosis and acute herpetic zoster. *Acta Neurol. Scand.* **2000**, *102*, 94–98. [CrossRef]

36. Rubinsztein, D.C.; Hanlon, C.S.; Irving, R.M.; Goodburn, S.; Evans, D.G.R.; Kellar-Wood, H.; Xuereb, J.H.; Bandmann, O.; Harding, A.E. Apo E genotypes in multiple sclerosis, Parkinson’s disease, schwannomas and late-onset Alzheimer’s disease. *Mol. Cell Probes* **1994**, *8*, 519–525. [CrossRef]

37. Savettieri, G.; Andreoli, V.; Bonavita, S.; Cittadella, R.; Caltagirone, C.; Fazio, M.C.; Siracusa, G.; Le Pira, F.; Liguori, M.; Logroscino, G.; et al. Apolipoprotein E genotype does not influence the progression of multiple sclerosis. *J. Neurol.* **2003**, *250*, 1094–1098. [CrossRef]
38. Zakrzewska-Pniewska, B.; Styczynska, M.; Podlecka, A.; Samocka, R.; Peplonska, B.; Barcikowska, M.; Kwiecinski, H. Association of apolipoprotein E and myeloperoxidase genotypes to clinical course of familial and sporadic multiple sclerosis. Mult. Scler. 2004, 10, 266–271. [CrossRef]

39. Zwemmer, J.N.P.; Van Veen, T.; Van Wijnen, L.; Van Kamp, G.J.; Barkhof, F.; Polman, C.H.; Uitdehaag, B.M.J. No major association of Apo E genotype with disease characteristics and MRI findings in multiple sclerosis. Mult. Scler. 2004, 10, 272–277. [CrossRef]

40. Masterman, T.; Zhang, Z.; Hellgren, D.; Salter, H.; Anvret, M.; Lilius, L.; Lannfelt, L.; Hillert, J. APOE genotypes and disease severity in multiple sclerosis. Mult. Scler. 2002, 8, 98–103. [CrossRef]

41. Fazekas, F.; Strasser-Fuchs, S.; Schmidt, H.; Enzinger, C.; Ropele, S.; Lechner, A.; Flooh, E.; Schmidt, R.; Hartung, H.-P. Apolipoprotein E genotype related differences in brain lesions of multiple sclerosis. J. Neurol. Neurosurg. Psychiatry 2000, 69, 25–28. [CrossRef]

42. Hogh, P.; Oturai, A.; Schreiber, K.; Blinkenberg, M.; Jorgensen, O.S.; Ryder, L.; Paulson, O.B.; Suhrensen, P.S.; Knudsen, G.M. Apolipoprotein E and multiple sclerosis: Impact of the epsilon-4 allele on susceptibility, clinical type and progression rate. Mult. Scler. 2000, 6, 226–230. [CrossRef] [PubMed]

43. Zhang, H.L.; Wu, J.; Zhu, J. The immune-modulatory role of apolipoprotein E with emphasis on multiple sclerosis and experimental autoimmune encephalomyelitis. Clin. Dev. Immunol. 2010, 2010, 186813. [CrossRef] [PubMed]

44. Schmidt, S.; Barcellos, L.F.; Desombre, K.; Rimmler, J.B.; Lincoln, R.R.; Bucher, P.; Saunders, A.M.; Lai, E.; Martin, E.R.; Vance, J.M.; et al. Association of polymorphisms in the apolipoprotein E region with susceptibility to and progression of multiple sclerosis. Am. J. Hum. Genet. 2002, 70, 708–717. [CrossRef] [PubMed]

45. Fazekas, F.; Strasser-Fuchs, S.; Kollegger, H.; Berger, T.; Kristoferitsch, W.; Schmidt, H.; Enzinger, C.; Schiefermeier, M.; Schwarz, C.; Kornek, B.; et al. Apolipoprotein E epsilon 4 is associated with rapid progression of multiple sclerosis. Neurology 2001, 11, 853–857. [CrossRef]

46. Julian, L.J.; Vella, L.; Frankel, D.; Minden, S.L.; Oksenberg, J.R.; Mohr, D.C. APOE alleles, depression and positive affect in multiple sclerosis. Mult. Scler. 2009, 15, 311–315. [CrossRef]

47. Lazo, M.; Alonso, M.E.; Corona, T. Apo E genotype and idiopathic optic neuritis. Mult. Scler. 2003, 9, 241.

48. Tamam, Y.; Tasdemir, N.; Yalman, M.; Tamam, B. Association of apolipoprotein E genotypes with prognosis in multiple sclerosis. Eur. Rev. Med. Pharmacol. Sci. 2011, 15, 1122–1130.

49. Kulminski, A.M.; Shu, L.; Loika, Y.; He, L.; Nazarian, A.; Arbeev, K.; Ukrain'tseva, S.; Yashin, A.; Culminskaya, I. Genetic and regulatory architecture of Alzheimer’s disease in the APOE region. Alzheimers Dement 2020, 12, e12008. [CrossRef]