Hand-made articulating spacers in two-stage revision for infected total knee arthroplasty

Good outcome in 30 patients

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Background and purpose   The most reliable results when treating an infected total knee arthroplasty have been obtained with a two-stage reimplantation protocol. We have used a simple technique for hand-made spacers and now report the outcome.

Patients and methods   30 patients with an infected total knee arthroplasty (TKA) were treated with a 2-stage reimplantation protocol. Spacers were built and customized to the type of defect using only 2 retractors and a high-speed tip burr. Partial weight bearing and discharge from the hospital were encouraged in the time between surgeries. 29 of 30 patients (97%) had successful reimplantations and they were followed for an average of 3 (2–5) years.

Results   Range of motion with the articulating spacer averaged 80° (55–100) and 21 of the 30 patients achieved motion greater than 75°. At the latest follow-up, there were no reinfections. According to the Knee Society score (KSS), the results were considered excellent or good in 25 patients, and fair or poor in 4 of the 29 patients with reimplantations. Motion after reimplantation averaged 107° (90–120).

Interpretation   One of the key factors related to a successful outcome with a two-stage reimplantation procedure is to keep the joint mobile and functional in the time between surgeries. Hand-made articulating spacers retain most of the advantages of more complex spacers in terms of mobility, pain, bone loss, success, or re-infection rate—with the major advantages of price and universal availability. Limitations related to this technique include potential knee instability, the need to walk with an orthosis, and the risk of subluxations.

Infection is one of the worst complications of a total knee arthroplasty procedure. It has been reported to affect between 2% and 3% of primary procedures, with double these rates in cases of revision arthroplasty (Mulvey and Thornhill 2001). Revision of an infected total knee arthroplasty is 3–4 times more expensive than the index procedure, and twice the cost of an aseptic revision (Hebert et al. 1996).

The goal of treatment of an infected knee arthroplasty is to eradicate the infection while preserving the function of the extremity. The surgeon can affect the final result at 3 fundamental steps: (1) the initial joint debridement, (2) the protocol and type of spacer used between operations, and (3) the surgical technique and selection of prosthesis at reimplantation (Whiteside 1994).

The most accepted and consistent results have been achieved with a two-stage reimplantation protocol including the removal of all the prosthetic components and the insertion of a temporary, antibiotic-loaded spacer between surgeries followed by a 4–6-week course of intravenous antibiotics, and then a delayed implantation of a new prosthesis fixed with antibiotic-loaded cement (Windsor...
Proposed advantages of the articulating spacers include maintenance of function and motion of the knee, minimization of soft tissue contractures, preservation of bone stock, and the possibility of delivering high concentrations of antibiotics. Spacers also reduce the need for extensile exposures at reimplantation and facilitate patient comfort and early hospital discharge (Calton et al. 1997, Haddad et al. 2000).

Although articulating spacers were introduced 20 years ago, surgeons have continuously described new modifications of spacers to adapt them to their specific technical and budgetary constraints. Most modifications have yielded good results (Insall et al. 1983, Cohen et al. 1988, Booth and Lotke 1989, Duncan et al. 1992, McPherson et al. 1995, Siebel et al. 2002, Durbhakula et al. 2004, Ha 2006, Mcavoy and Ries 2006). The simplest, cheapest, and universally available method for making articulating spacers is to build them during surgery (Villanueva et al. 2006).

We examined the outcome of a uniform technique of making hand-made articulating knee spacers. The procedure only requires three basic surgical tools, it does not involve autoclaving, and it does not require the use of trial plastic molds or of metallic components.

**Patients and methods**

30 patients (23 females), with an infected total knee arthroplasty were treated with a 2-stage revision protocol that included the use of a custom hand-made antibiotic-loaded articulating spacer built using a standardized technique. 26 patients had osteoarthritis and 4 patients had rheumatoid arthritis. All patients were reviewed retrospectively. We together started to perform this technique at the institution of the first author, and later on at other hospitals. The series include consecutive patients in which 2-stage reimplantation was considered the best option for treatment. Mean age at the time of treatment was 71 (64–82) years. 9 patients had had multiple previous operations on the infected knee. Time to our treatment, since the index arthroplasty, averaged 18 (1–144) months.

Before our procedure, 14 patients had had posterior stabilized designs (PS), 13 patients had had a cruciate retaining prosthesis (CR), and 4 patients had had revision designs including wedges and stems (2 with a PS model and 2 with a deep-dish ultracongruent polyethylene component). 2 patients had had a quadriceps snip at the time of a previous revision. None of the cases had been operated using minimum invasive surgery.

*Staphylococcus aureus* (SA) and coagulase-negative *Staphylococcus* (CNS) were each present in 12 of the patients, with 3 meticillin-resistant SA, 9 non-meticillin-resistant SA, 5 meticillin-resistant CNS, and 7 non-meticillin-resistant CNS. Enterobacteriae were present in 6 cases and anaerobes and streptococci were present in 3 other cases. In 3 cases, a polymicrobial infection was diagnosed.

At the first stage of surgery, all prosthesis components were removed and an extensive debridement of the knee was done. Samples were then taken for microbiological culture and pathological analysis, including intraoperative frozen sections to establish “definitive criteria of infection” (Atkins et al. 1998). The *articulating spacer*. 3 g of antibiotics (7.5%) per pack of cement was used in all but 4 cases. All spacers were constructed with CMW 1 with gentamycin (De Puy), to which 2 extra grams of gentamycin or 2 extra grams of vancomycin were added, or Simplex (Stryker-Howmedica) cement, to which 3 g of extra gentamycin or vancomycin per pack were added. The cement was modeled in its doughy phase to avoid excessive interdigitation in the remaining bone, and to facilitate its removal at reimplantation. The surgeon must know the time at which the cement cures and hardens with the antibiotics added. No pre-manufactured molds were used and no trial or previously autoclaved components were used to build the spacer. A Hohmann retractor, a curved osteotome, and a high-speed tip burr were all the surgical tools required. The components of the spacer were conformed manually following the basic guidelines that we have previously described (Villanueva et al. 2006) (Figures 1–3).

A continuous passive motion program was started within 24 h postoperatively with a Kinetec, progressing slowly in flexion and never surpassing 90° (Figure 4) to avoid potential subluxation and
Figure 1. The grooves, the central crest on the tibial component, and the trochlear groove on the femoral component can be seen. This shape of the spacer increases conformity and reduces instability.

Figure 2. Remodeling of posterior condyles with a curved osteotome.

Figure 3. Remodeling of the troclear groove with the high-speed burr.

Figure 4. Flexion over 90° in the same patient (A) and over 75° in another case (B).
locking of the spacer. Isometric exercises with and without bracing were encouraged, as well as active flexion and extension exercises with the heel resting on the bed or on the hand of a physiotherapist. Partial weight bearing with the aid of crutches or a cane was allowed as tolerated. An extension or hinge orthosis of the knee was worn during ambulation until soft tissue healing had occurred. If slight or mild laxity remained, the brace was worn until reimplantation.

Antibiotic therapy was instituted by the infectious diseases specialist and the patient entered a “hospital at home” program after discharge from the hospital. After discontinuation of antibiotics for at least 2 weeks, reimplantation was performed guided by intraoperative fresh-frozen samples, with more than 5 polymorphonuclear leukocytes (PMN) per high-power field being considered predictive of infection. In patients with inflammatory diseases, we based our decision on previous aspiration cultures. 2 cases had false positive histology and reimplantation was delayed for 15 days—when the cultures taken were negative (Fehring and McAllister 1994, Lonner et al. 1996). At reimplantation the same type of cement-loaded antibiotic was used, limiting the percentage of antibiotic to 2.5% of the final composite.

Results (Table)
Reimplantation was performed in 29 of the 30 patients (97%) at an average time of 14 (8–130) weeks. In a patient with rheumatoid arthritis, reimplantation was delayed 2.5 years due to the patient’s initial rejection of prosthetic knee reimplantation after 3 exchanges of the spacer because of persistent infection. One patient finally required an arthrodesis after several debridements and impairment of the extensor mechanism. At reimplantation, we used 21 semiconstrained implants (intercondylar stabilized), 6 posterior stabilized implants, and 2 rotating-hinge prostheses.

Range of motion (ROM) with the spacer averaged 80° (55–100). 21 patients achieved a motion greater than 75° with the articulating spacer.

At reimplantation, tibial tuberosity osteotomy was performed in the 2 patients who had the poorest motion between surgeries, and 5 “quadriceps snips” were performed. One patient required a proximal Insall’s realignment at the second stage of surgery. His final motion was 5–100°. Another patient required the realignment at the first stage, due to extensor mechanism and surgical wound dehiscence.

3 patients had a mild extension lag at the final follow-up. One of them had required an Insall’s realignment at reimplantation (ROM 5–110°) and the other 2 a tibial tuberosity osteotomy (ROM 5–110° and 10–100°, respectively).

According to the Knee Society score (KSS), the results following reimplantation were excellent (85–100 points) or good (70–84 points) in 25 patients and fair (60–69 points) or poor (less than 60 points) in 4 patients. Average motion after reimplantation was 107° (90–120°).

3 cases had complications directly related to the spacer. These included 2 subluxations, manually reduced, and a fracture of the posterior condyles of the femoral spacer. In 4 cases with segmentary defects, 1 component had to be rebuilt during the first surgery to obtain adequate shape and stability.

Discussion
In the treatment of infected total knee arthroplasties, the importance of every single factor affecting the final outcome (extensive debridement, time from clinical onset, the infecting microorganism, the dosage and combination of antibiotics, type of spacer, the technique, and prostheses selection at reimplantation) still remains to be defined as the series and techniques reported are not uniform—and hence difficult to compare. The use of articulating spacers as part of a 2-stage reimplantation protocol has, however, proven to be important in the eradication of infection and preservation of joint function (Haleem et al. 2004)—regardless of multiple modifications that include the use of specific molds and the use and resterilization of trial components or previous implants (Insall et al. 1983, Cohen et al. 1988, Booth and Lotke 1989, Duncan et al. 1992, McPherson et al. 1995, Goldstein et al. 2001, Siebel et al. 2002, Durbhakula et al. 2004, Ha 2006, MacAvoy and Ries 2006).
### Cases of infected knee arthroplasties

| Diagnosis, age, sex | Primary TKA | Microorganism / antibiotics | Time to infection | ROM with spacer | Knee exposure | Revision prosthesis | ROM d | KSS score follow-up | R |
|---------------------|-------------|------------------------------|-------------------|----------------|--------------|---------------------|-------|---------------------|----|
| OA 77 F             | Profix (S&N)| MRSA, Enterobacteriae/ rifampicin + cotrimoxazole | 6 weeks | 0–75° | 9 weeks Q snip | Profix (S&N) | 0–90° | 68/60 | 3 years | F |
| OA 82 M             | IB II (Zimmer) | Streptococcus viridans/ ceftriaxone + cotrimoxazole | 1.5 years | 0–85° | 10 weeks | IB-CCK (Zimmer) | 5–110° | IR | 95/85 | 5 years | E |
| RA 68 F             | IB II (Zimmer) | MSSA/ cefazolin + ciprofl oxacin | 1.5 years | 0–90° | 11 weeks | IB-CCK (Zimmer) | 0–135° | | 95/100 | 5 years | E |
| OA 76 F             | IB II (Zimmer) | Enterobacteriae, Clostridium/ ceftriaxone + teicoplanin + rifampicin | 3 years | 0–85° | 2 surgical procedures | Anthro d | 0° | Crutches | 3 years | P |
| OA 75 M             | Profix (S&N) | MSSA/ levofloxacin + rifampicin | 7 months | 0–55° | 11 weeks TTO | Profix PS (S&N) | 5–110° | IR | 86/75 | 3 years | G |
| OA 72 F             | Génesis I (S&N) | MSSSE/ ciprofl oxacin + rifampicin | 2 years | 0–90° | 8 weeks | IB-CCK (Zimmer) | 0–115° | IR | 92/85 | 4 years | E |
| OA 76 F             | Génesis I (S&N) | MRSA/ cotrimoxazole + rifampicin | 3 years | 0–80° | 8 weeks Q snip | IB-CCK (Zimmer) | 0–110° | IR | 95/80 | 3 years | G |
| OA 73 M             | IB II (Zimmer) | MSSA/ ciprofl oxacin + rifampicin | 14 months | 0–85° | 10 weeks | IB-CCK (Zimmer) | 0–110° | IR | 85/80 | 3 years | G |
| RA 70 F             | IB II (Zimmer) | MSSSE/ ciprofl oxacin + rifampicin | 22 months | 0–90° | 9 weeks | IB-CCK (Zimmer) | 0–115° | IR | 90/75 | 4 years | G |
| OA 71 M             | Profix (S&N) | MSSSE/ levofloxacin + rifampicin | 14 months | 0–80° | 8 weeks Q snip | Profix PS (S&N) | 0–100° | IR | 85/80 | 2 years | G |
| OA 74 F             | Profix (S&N) | MSSA/ ciprofl oxacin + rifampicin | 23 months | 0–80° | 10 weeks | Profix PS (S&N) | 0–105° | IR | 85/70 | 2 years | G |
| RA 65 F             | IB II (Zimmer) | MSSA/ ciprofl oxacin + rifampicin | 2 years | 0–85° | 11 weeks | IB-CCK (Zimmer) | 0–120° | IR | 91/80 | 4 years | E |
| OA 64 F             | Tricon- Profix revision (S&N) | Enterobacteriae/ teicoplanin + rifampicin | 3 months | 0–90° | 9 weeks Q snip | NG LCCK (Zimmer) | 0–105° | IR | 91/85 | 3 years | E |
| OA 68 F             | Profix (S&N) | MSSA/ ciprofl oxacin + rifampicin | 5 weeks | 0–75° | 11 weeks IR | NG LCCK (Zimmer) | 0–100° | IR | 90/80 | 3 years | G |
| RA 66 F             | Miller- Galante (Zimmer) | Streptococcus + MRSA/ amoxicillin-clavulanic 2 months and cotrimoxazole + rifampicin | 12 years | 0–85° | 2.5 years MR Hinge (Stryker) | | | 0–100° | IR | 65/55 | 2 years | P |
| OA 73 F             | Profix (S&N) | E. Coli ciprofl oxacin + rifampicin | 10 months | 0–90° | 10 weeks | NG LCCK (Zimmer) | 0–120° | IR | 94/90 | 2 years | E |
| OA 69 F             | Optetrak PS (ET) | MRSA/ linezolid + rifampicin | 9 months | 0–90° | 8 weeks | Optetrak semicons. (ET) | 0–95° | IR | 85/90 | 2.5 years | G |
| OA 69 F             | PFC (Johnson) | Proteus mirabilis/ ciprofl oxacin | 4 weeks | 0–70° | 12 weeks | Optetrak semicons. (ET) | 0–95° | | 81/70 | 2 years | G |
| OA 67 F             | Scorpio PS (Stryker) | MRSE/ cotrimoxazole + rifampicin | 1 year | 0–80° | 11 weeks | Optetrak semicons. (ET) | 0–105° | IR | 88/85 | 2 years | E |
### Table continued

| Diagnosis, age, sex | Primary TKA | Microorganism / antibiotics | Time to infection | ROM with spacer | Knee exposure | Revision prosthesis | ROM d | KSS score | follow-up |
|---------------------|-------------|------------------------------|-------------------|----------------|--------------|---------------------|-------|-----------|-----------|
| OA 68 F             | Scorpio PS (Stryker) | MSSE/levofloxacin + rifampicin | 8 months | 0–70° | 10 weeks | Optetrak revision (ET) | 0–100° | 82/70 | 2 years | G |
| OA 66 F             | Profix (S&N) | MRSE/linezolid + rifampicin | 5 months | 0–90° | 13 weeks | NG PS (Zimmer) | 0–100° | 85/80 | 2 years | G |
| OA 72 M             | CKS (Biomet) | MRSE/cotrimoxazole + rifampicin | 4 weeks | 0–85° | 11 weeks* | NGLCCK (Zimmer) | 0–115° | 90/85 | 4 years | E |
| OA 69 F             | NG PS (Zimmer) | MRSE/linezolid + rifampicin | 10 months | 0–60° | 24 weeks | Q-snip | 0–100° | 85/70 | 2 years | G |
| OA 70 M             | NG PS (Zimmer) | MSSE/levofloxacin + rifampicin | 6 months | 0–75° | 10 weeks | NG LCCK (Zimmer) | 0–120° | 87/85 | 2 years | E |
| OA 82 F             | NG revision PS (Zimmer) | Serratia/ciprofloxacin | 6 months | 0–100° | 10 weeks | MR Hinge (Stryker) | 0–100° | 80/55 | 2 years | P |
| OA 68 F             | Genesis I (S&N) | MSSA/levofloxacin + rifampicin | 4 weeks | 0–90° | 8 weeks | NG LCCK (Zimmer) | 0–115° | 93/80 | 2 years | G |
| OA 65 F             | Profix (S&N) | MRSE/cotrimoxazole + rifampicin | 8 months | 0–80° | 8 weeks | NG PS (Zimmer) | 0–100° | 65/70 | 4 years | G |
| OA 77 M             | NG PS (Zimmer) | MSSE + Streptococcus faecium/rifampicin + linezolid | 1.5 years | 0–75° | 16 weeks* | NG LCCK (Zimmer) | 0–110° | 95/90 | 2 years | E |
| OA 73 F             | Genesis I (S&N) | MSSE/ciprofloxacin + rifampicin | 5 weeks | 0–60° | 9 weeks | IB CCK (Zimmer) | 10–100° | 71/55 | 5 years | P |
| OA 72 F             | Tricon-Gen. I PS (S&N) | MSSA/ciprofloxacin + rifampicin | 4 years | 0–80° | 12 weeks | NG LCCK (Zimmer) | 0–110° | 82/80 | 4 years | G |

* OA: Osteoarthrosis, RA: Rheumatoid arthritis
* TKA: ET: Exactech
* IB: Insall-Burstein
* NG: Nex-Gen
* S&N: Smith and Nephew
* Microorganism:
  * MRSA: Methicillin resistant Staphylococcus aureus
  * MSSA: Methicillin sensitive Staphylococcus aureus
  * MRSE: Methicillin resistant Staphylococcus epidermidis
  * MSSE: Methicillin sensitive Staphylococcus epidermidis
* R: Insall’s realignment
* TTO: Tibial tubercle osteotomy
* Result:
  * R: Fresh-frozen samples +, negative cultures. Delayed reimplantation 15 days later.
use of these complex systems has not been general-
ized, however, and hand-made spacers are prob-
ably the most commonly used.

We have recently described a step-by-step tech-
nique to construct articulating spacers with just
3 basic surgical tools, thus representing an inex-
pensive method that is applicable in all operating
theaters (Villanueva et al. 2006). Our findings in
this consecutive series of 30 patients imply that our
hand-made spacers function as well as more com-
plex constructs.

29 of 30 patients were successfully reimplan-
tated without recurrence of infection at the most
recent follow-up, at a minimum of 2 years. 25 of
29 patients had excellent or good results and only 4
had fair or poor results according to the KS score.

Partial weight bearing and discharge from the
hospital were possible in the time between surger-
ies. Range of motion (ROM) with the spacer aver-
aged 80° and after reimplantation it was 107°.

2 patients required a tibial tubercle osteotomy
at reimplantation and 3 patients had an exten-
sion lag, but it could be partially related to lim-
ited ROM with the spacer in only 1 case. Other
advantages of spacers, such as minimizing the
retraction of the extensor mechanism, preventing
muscle atrophy and pain, preservation of bone
stock (Calton et al. 1997), and good knee motion
(Hoffman 2005) all appear to be achieved with
our hand-made spacers.

Complications included two subluxations,
reduced with a “Lachman test” maneuver, and a
spacer broken at the posterior condyle, which did
not affect knee mobility. The need to rebuild the
spacer in cases with considerable defects reflected
the learning curve of a manual procedure and did
not affect the final result, but it lengthened surgical
time. Specific problems when building the spacer
were excessive tissue tension, lack of posterior caps-
ule release that prevented extension of the poste-
rior part of the femoral spacer (leading to instabil-
ity of the femoral component), and elevation of the
posterior part of the tibial component—creating
an anterior slope or inclination that limited flexion
and caused anterior subluxation (Figure 5).

The possibility of adding high doses of antibi-
otic therapy alone (McLaren and Spooner 1996,
Jiranek et al. 2006), thus avoiding the development
of antibiotic resistance.

It has been shown that the maximum quantity
of antibiotic to be added that will not affect the
mechanical properties of the cement is around 5%
of the total weight of the cement (Haddad et al.
2001, Jiranek et al. 2006). However, most authors
use much higher doses of antibiotic in the tempo-
rary spacers used between surgeries. These systems
should not be used for longer than 8–12 weeks,
which is not enough for the expected increase in
fragility of the cement to have any clinical rel-
ence.

Hoffmann et al. (2005) reported 50 cases in
which they mixed tobramycin with the cement in
a 12% proportion to give the final composite. They
had a 12% reinfection rate, but the authors used
spacers including plastic and metal components.
MacAvoy and Ries (2006) used a 9% antibiotic-
cement composite with a ball and socket technique
not involving plastic or metal components. They
had a 30% reinfection rate at 18 months, probably
related to the co-morbidities of their patient popu-
lation. Using 10–12% antibiotic in the cement-anti-
biotic composite, Ha (2006) treated 12 cases with
a spacer made of cement alone, and there were
no recurrences of the infection. Durbhakula et al.
(2004) used silicone molds (ALACS) to build their
spacers without the inclusion of cement or plastic
elements. The authors used an antibiotic-loaded
cement spacer with a mean proportion of antibiotic

Figure 5. Subluxation of a spacer.
of 8%. 2 of 24 patients had persistent infection at the time of intended reimplantation.

Our spacers did not include plastic or metallic elements, and the cement proportion was 7.5%. This could be a real limitation of hand-made spacers because with proportions higher than 10%, the cement becomes stiffer and more difficult to model—so we were using theoretically suboptimal dosages (Jiranek et al. 2006). However, our high rate of eradication of infection compares to other reports using higher proportions, reflecting that this factor alone is only part of a successful protocol. 2 patients required repeated debridements: 1 had successful reimplantation and 1 was finally arthrodesed. There have been no recurrences of infection at a minimum 2 years follow-up.

We used semi-constrained designs at reimplantation in 21 patients. In a series of 13 patients with severe bone loss or collateral ligaments deficiencies, MacAvoy and Ries (2006) only used constrained designs in 5 cases. In their series of 50 patients, Hoffmann et al. (2005) reported 36 reimplantations performed with ultracongruent designs, with the same proportion of semi-constrained as in our series.

It may be expected that some residual laxity of the knees with the cemented spacers might contribute to secondary damage of the collateral ligaments, but this is not our experience. In cases of revision knee surgery, one frequently needs a higher level of constriction than in the prior surgery and 17 cases in our series already had a posterostabilized design at the time of onset of infection, some including stems, steering the preference of the surgeon at reimplantation to a semi-constrained design rather than to a posterostabilized prosthesis (Lampe and Hiller 2005). We used the hinge designs in 2 multi-operated patients with severe bone loss, soft tissue damage, and poor functional status.

We found a similar final result regardless of the time elapsed between the initial surgical debridement and final reimplantation. As long as the joint remains functional with controlled mobility and partial weight bearing, the final outcome seems not to be jeopardized.

In summary, hand-made articulating spacers appear to function as well as more complex, expensive spacers.

Contributions of authors
All authors have used the technique described, contributed cases to the series, and participated in writing of the manuscript.

No competing interests declared

A video illustrating this technique was given an award by the American Academy of Orthopaedic Surgeons (AAOS), San Diego 2007.
Link: http://www4.aaos.org/product/productpage.cfm?code=17102

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