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Cerebrospinal Fluid, Plasma, and Saliva in the BioFIND Study: Relationships Among Biomarkers and Parkinson’s Disease Features

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ABSTRACT: Objective: Examine relationships among neurodegenerative biomarkers and PD motor and nonmotor symptoms.

Background: CSF alpha-synuclein is decreased in PD versus healthy controls, but whether plasma and saliva alpha-synuclein differentiate these groups is controversial. Correlations of alpha-synuclein among biofluids (CSF, plasma, saliva) or biomarkers (eg, beta-amyloid, tau [total, phosphorylated]) are not fully understood. The relationships of these biomarkers with PD clinical features remain unclear.

Methods: BioFIND, a cross-sectional, observational study, examines clinical and biomarker characteristics in moderate-advanced PD and matched healthy controls. We compared alpha-synuclein concentrations across diagnosis, biofluids, and CSF biomarkers. Correlations of CSF biomarkers and MDS-UPDRS, motor phenotype, MoCA, and rapid eye movement sleep behavior disorder questionnaire scores in PD were examined.

Results: CSF alpha-synuclein was lower in PD versus controls (P = .01), controlling for age, gender, and education. Plasma and saliva alpha-synuclein did not differ...
between PD and controls, and alpha-synuclein did not significantly correlate among biofluids. CSF beta-amyloid1-42 was lower in PD versus controls (P < 0.01), and correlated weakly with MoCA recall scores (r = 0.23, P = 0.02). CSF alpha-synuclein was lower in the postural instability/gait difficulty phenotype than other motor phenotypes (P < 0.01). No CSF biomarkers predicted or correlated with total motor or rapid eye movement sleep behavior disorder scores. CSF alpha-synuclein correlated with beta-amyloid1-42, total-tau, and phosphorylated-tau (r = 0.41, 0.81, 0.43, respectively; Ps < 0.001).

Biomarkers have the potential to play a role in establishing PD diagnosis, understanding disease progression or disease-related features, and monitoring therapeutic effects. In PD, candidate biofluids include cerebrospinal fluid (CSF), plasma/serum, saliva, and urine, among others.1-3 Alpha-synuclein (α-syn) holds promise as a biomarker because it is a major component of Lewy bodies, can be found in peripheral tissues and body fluids, is readily secreted into extracellular spaces, and in part can be detected in exosomes.4 Studies consistently demonstrate reduced CSF total α-syn levels in PD when compared with healthy controls (HC).1,5,6 Because of the invasive nature of acquiring CSF, more readily accessible biofluids, such as blood or saliva, are attractive alternatives. Red blood cells express high levels of α-syn, but reports of plasma/serum α-syn in PD when compared with HC have yielded conflicting results.4,7,8 Saliva is an interesting source of α-syn because Lewy pathology has been noted in salivary glands,9 but studies of salivary α-syn have produced contradictory results, with levels either increased or not different between PD and HC.8,10 There are no studies of which we are aware that investigate the relationships among CSF, plasma, and salivary α-syn in PD.

Several studies have examined CSF markers of neurodegeneration in PD (eg, α-syn, beta-amyloid [Aβ] 1-40 and 1-42, total tau [t-tau], and phosphorylated tau [p-tau] concentrations), but most studies focus on either de novo PD patients (eg, deprenyl and tocopherol antioxidative therapy of parkinsonism [DATA-TOP]) or Parkinson’s Progression Markers Initiative (PPMI)), or specifically on cognitive measures in PD cohorts.1,11-16 Of this work, the prevailing evidence is that CSF α-syn is reduced in PD and that subspecies of α-syn oligomers may distinguish PD from controls. Furthermore, Aβ and tau may provide useful insight into prognosticating cognitive decline in PD. To date, few studies have examined CSF α-syn, Aβ, t-tau, and p-tau obtained simultaneously in moderate-advanced “typical” PD cohorts or investigated how they relate to specific motor and nonmotor features of PD, such as motor subtype, sleep, and other nonmotor features.

Conclusion: Lower CSF alpha-synuclein is associated with diagnosis and motor phenotype in moderate-advanced PD. Plasma and saliva alpha-synuclein neither correlate with CSF alpha-synuclein, nor distinguish PD from controls. CSF beta-amyloid1-42 remains a potential biomarker for cognitive impairment in PD. © 2017 The Authors. Movement Disorders published by Wiley Periodicals, Inc. on behalf of International Parkinson and Movement Disorder Society.

Key Words: alpha-synuclein; amyloid; cerebrospinal fluid; postural instability gait difficulty; tau

BioFIND, a cross-sectional, observational study of moderate-advanced PD patients and matched HCs evaluated with standardized clinical and biospecimen acquisition protocols, provides a unique resource for examining biomarkers in a well-characterized, “typical” (including good response to levodopa) PD population.17 The aim of our study was 3-fold: (1) examine relationships among (a) CSF markers of neurodegeneration (ie, α-syn, Aβ1-42, t-tau, p-tau) in PD and HC in the BioFIND cohort and (b) in PD participants, the relationships among CSF, plasma, and saliva α-syn levels; (2) determine differences between PD and HC in CSF α-syn, Aβ1-42, tau, and p-tau and in plasma and saliva levels of α-syn; and (3) investigate the associations of CSF markers with specific motor and nonmotor features of PD.

Methods

BioFIND Study and Design

BioFIND includes moderate-advanced PD participants and HCs enrolled at 8 sites in the United States.17 PD patients had “typical” features, meeting United Kingdom PD Society Brain Bank clinical diagnostic criteria and having all 3 classic parkinsonian motor signs (ie, bradykinesia, rigidity, resting tremor) by history or examination; represented all Hoehn and Yahr stages; had disease durations ≥ 4 years and onset between ages 50 to 75 years; and demonstrated well-established responses to dopaminergic agents and/or amantadine. Patients were excluded if they had features of atypical or secondary parkinsonian syndromes; a history of DBS or ablative brain surgery; a history of cancer (except basal or squamous cell skin cancers) within 5 years preceding enrollment; autoimmune, liver, or hematological disorders; or conditions precluding lumbar puncture. HCs were group matched by age and sex to PD patients, were free of any known neurological disorders, and scored ≥ 26 on the Montreal Cognitive Assessment (MoCA).18 Other exclusion criteria for HCs were similar to those for PD patients. Controls were excluded if they had a first-degree family member with
Evaluations

Clinical data and biospecimen collection occurred on 2 visits (baseline, [V1] and follow-up within 2 weeks of baseline, [V2]). For PD patients, V1 was performed in the on state (1-3 hours after the last PD medication dose) and V2 was performed in the practically defined off state (early morning before PD medications and approximately 12 hours after the last dose the night before). V1 included collection of blood for DNA and plasma and assessments including the International Parkinson and Movement Disorder Society–Sponsored revision of the UPDRS (MDS-UPDRS) parts I (nonmotor experiences of daily living), II (motor experiences of daily living), III (motor examination), and IV (motor complications) for PD patients and part III only for HCs. V2 included collection of blood for RNA and plasma, CSF, saliva (added after study startup), and MDS-UPDRS part III in PD patients. All PD and HCs either fasted or had a low-fat diet on the morning of V2. Assessments included demographics, family history of PD, medical/neurological histories, medications, neurological exams, and the MDS-UPDRS,19 MoCA,18 and Rapid Eye Movement Behavior Sleep Disorder (RBD) Questionnaire20 and for PD patients, the Modified Schwab and England Activities of Daily Living Scale.21

Motor and Nonmotor Features

For PD patients, motor phenotype was classified as tremor dominant (TD), postural instability/gait difficulty (PIGD), and indeterminate,22,23 calculated from MDS-UPDRS part III motor scores (V2, off) plus pertinent history questions from MDS-UPDRS part II (2.10 tremor, 2.12 walking and balance, 2.13 freezing; V1 and acquired only once during the study).23 Other motor and nonmotor features were examined using the published factor structure of the MDS-UPDRS for parts I to IV.19 Cognitive function was measured by MoCA total and subdomain scores (eg, visuospatial/executive, naming, attention, language, abstraction, delayed recall, orientation). RBD was examined using the RBD questionnaire total score, with a cut-off of > 5 as indicative of RBD symptoms.

The study was approved by the institutional review boards for the University of Rochester Clinical Trials Coordination Center and study sites. Written, informed consent was obtained from the study participants.

Biospecimen Analyses

Concentrations of α-syn in plasma, CSF, and saliva samples were analyzed using ELISA assays (BioLegend, cat. 844101). The concentration of α-syn in each sample was determined by interpolation of values against the standard curve established by the reference standards (range 1500 pg/ml-6.1 pg/ml) using a 4-parameter regression. Recombinant α-syn from rPeptide was used as the standard in this assay. Each sample was analyzed in duplicate at appropriate dilutions (1/20 for CSF, 1/200 for plasma, 1/4 for saliva). Concentrations of hemoglobin in CSF, plasma, and saliva samples were analyzed using ELISA assays (Bethyl Laboratories, cat. E80-134). Reference standards used in the assay ranged from 7.5 ng/ml to 125 ng/ml. The concentration of hemoglobin in each sample was determined by interpolation of values against the standard curve established by the reference standards using a 4-parameter regression. The assay has been validated in multiple matrices including in CSF, plasma, and saliva (BioLegend manual November 11, 2016). Interassay precision and spike/recovery data for saliva are depicted (Supplementary Figure).

### TABLE 1. Demographic and clinical features of the BioFIND study cohort

|                      | Healthy controls, n = 88 | PD, n = 115 | P value |
|----------------------|--------------------------|-------------|---------|
| Age, y               | 65.64 (7.36)             | 68.24 (6.40) | <.01    |
| Gender, % male       | 51.10                    | 62.60       | .10     |
| Age at PD onset, y   | 59.90 (6.17)             | 59.90 (6.17)| <.001   |
| Duration of PD, y (SD)| 8.34 (3.09)             | 8.34 (3.09) | <.001   |
| Education, y         | 17.00 (3.06)             | 16.98 (2.97) | .97     |
| LEDD, mg/day         | 740.50 (385.76)          | 740.50 (385.76) | <.001   |

Data are presented as mean (standard deviation) unless otherwise noted. RBD, Rapid Eye Movement Behavior Sleep Disorder; TD, tremor dominant; PIGD, postural instability/gait difficulty.
Simultaneous analysis of \( \text{A}_4 \)-42, t-tau, and p-tau in CSF was performed using a highly standardized microbead based research-use-only immunoassay (INNOBIA Alz Bio3 kits; Fujirebio, Ghent, Belgium).\(^{11}\) CSF samples without dilution were measured in 6 analytical runs in 96-well format, with each set of analyze calibration standards and quality control samples. A biomarker result was defined as the average of the duplicate concentration values. If a biomarker result was defined as the average of the duplicate concentration values. If a biomarker result did not meet specified criteria for acceptance, the result was not included in the final dataset. Two \( \text{A}_4 \)-42 and t-tau results had microbead counts \(<\)50 and 1 p-tau result had duplicate precision (%CV) > 25%; therefore, these 9 results were invalidated.

**Statistical Analysis**

The analyses were conducted in IBM SPSS Statistics version 23 (IBM Corp., Armonk, New York). Demographic data, disease characteristics, and biomarker concentrations were compared between PD and HC groups using \( t \) tests. Gender and dichotomous variables (e.g., RBD scores) were compared between groups using chi-square tests. Between-subjects tests were used to examine MoCA total and subdomain scores controlling for age and gender and CSF biomarker levels in motor phenotypes controlling for age, gender, and duration of disease. Pearson and Spearman rho correlations were used to examine the relationships among biofluids, CSF biomarkers, and for CSF with motor and nonmotor variables, as appropriate and controlling for age, gender, and education. Regression models controlling for age, gender, and education were used to examine CSF biomarker predictors of PD diagnosis.

**Results**

**Participant Characteristics**

Clinical characteristics of BioFIND participants who provided at least one of the biomarker specimens are presented in Table 1. Gender and education years did not differ significantly between PD and HC groups. PD patients were older than HCs (\( P < .01 \)), with mean age of disease onset 59.9 years, disease duration 8.34 years, MDS-UPDRS part III total motor score (off) 39.13, and a median Hoehn and Yahr stage of 2. PD motor phenotypes included 51.3% TD, 36.5% PIGD, and 12.2% indeterminate. MoCA total scores were significantly lower in PD patients when compared with HCs (\( P < .001 \)) as expected because HCs with MoCA scores \(<\)26 were excluded. Of the PD participants, 45.2% had RBD scores > 5, compared to 6.8% of HCs (\( P < .001 \)).

**Comparisons of CSF, Plasma, and Saliva Biomarkers in the BioFIND Cohort**

In the complete BioFIND cohort (PD and HC), CSF \( \alpha \)-syn concentrations were most strongly correlated with t-tau (\( r = 0.81 \)) and modestly with p-tau (\( r = 0.43 \)) and \( \text{A}_4 \)-42 (\( r = 0.41 \); \( P < .001 \); Fig. 1). Age was weakly, but significantly, correlated with CSF measures including hemoglobin (\( r = 0.18 \), \( P = .017 \)), \( \alpha \)-syn (\( r = 0.15 \), \( P = .043 \)), and t-tau (\( r = 0.20 \), \( P < .01 \)). Among PD participants only, correlations between CSF \( \alpha \)-syn concentrations and \( \text{A}_4 \)-42, t-tau, and p-tau remained consistent with those found in the complete BioFIND cohort.

Hemoglobin and \( \alpha \)-syn levels were modestly correlated in saliva (\( r = 0.41 \), \( P = .04 \)), but not in plasma (\( r = -0.28 \), \( P = .16 \)) or CSF (\( r = -0.12 \), \( P = .57 \)) in the BioFIND cohort. Saliva samples with hemoglobin concentrations \( > 1200 \text{ ng/ml} \) were excluded to avoid potential impact of contaminant hemoglobin on \( \alpha \)-syn levels.\(^{11,24}\) We did not exclude participants based on CSF or plasma hemoglobin levels because there was no significant correlation between hemoglobin and \( \alpha \)-syn levels in plasma and CSF.

Comparing \( \alpha \)-syn across the different biofluids in the BioFIND cohort, we did not find significant correlations for \( \alpha \)-syn concentrations between CSF and plasma.
TABLE 2. Comparison of CSF biomarkers in the BioFIND study cohort

| Biomarkers      | Healthy controls | PD          | P value | 95% CI       |
|-----------------|------------------|-------------|---------|--------------|
| CSFa            |                  |             |         |              |
| α-syn, pg/mL    | 1713.59 (637.16) | 1466.25 (669.20) | .01     | 89.47-491.30 |
| Aβ1-42, pg/mL   | 334.75 (89.40)   | 296.31 (79.16)   | <.01    |              |
| t-tau, pg/mL    | 42.06 (23.32)    | 37.15 (16.42)    | .08     |              |
| p-tau0/1, pg/mL | 20.61 (17.23)    | 16.42 (11.10)    | .05     |              |
| t-tau/Aβ1-42    | 0.15 (0.19)      | 0.07 (0.12)      | .28     |              |
| p-tau0/1/Aβ1-42 | 0.06 (0.06)      | 0.07 (0.12)      | .28     |              |
| Hb, ng/ml      | 165.73 (290.50)  | 157.04 (296.66) | .85     |              |
| Plasmab         |                  |             |         |              |
| α-syn, pg/mL    | 115,920.30       | 110,603.05 (71,392.10) | .64 |              |
| SalivaC         |                  |             |         |              |
| α-syn, pg/mL    | 165.97 (272.25)  | 285.42 (400.13) | .23 |              |

Data are presented as mean (standard deviation) unless otherwise noted. α-syn, alpha-synuclein; Aβ, beta-amyloid; t-tau, total tau; p-tau, phosphorylated tau.

CSF sample size: healthy controls, n = 76-79; PD, n = 98-106 (range).
Plasma sample size: healthy controls, n = 88; PD, n = 118.
Saliva sample size: healthy controls, n = 26; PD, n = 22.

(r = −0.08, P = .70, n = 177), CSF and saliva (r = −0.13, P = .53, n = 41), or plasma and saliva (r = 0.15, P = .48, n = 48). Likewise, among PD patients, there were no significant correlations between α-syn concentrations in CSF and plasma (r = −0.15, P = .14, n = 98), CSF and saliva (r = −0.41, P = .87, n = 20), or plasma and saliva (r = −0.25, P = .26, n = 22).

Comparisons of Biomarkers Between PD and HC

CSF α-syn (P = .01) levels were lower in PD patients when compared with HCs (Table 2, Fig. 2). The α-syn levels in plasma (P = .64) and saliva (P = .23), however, did not differ between PD and HCs. CSF Aβ1-42 (P < .01) and p-tau (P = .05) levels were lower in PD patients when compared with HCs, but t-tau (P = .08) levels did not differ between PD and HCs (Table 2). In regression models adjusted for age, gender, and education years, the CSF biomarkers were significant predictors of PD diagnosis (Table 3).

TABLE 3. Regression coefficients of CSF biomarkers in PD

| β-coefficient | t value | P value | 95% CI |
|---------------|---------|---------|-------|
| α-syn         | 290.39  | 2.85    | .005  | 89.47-491.30 |
| Aβ1-42        | 36.67   | 2.84    | .005  | 11.21-62.13  |
| t-tau         | 6.13    | 2.07    | .01   | 2.99-9.16    |
| p-tau         | 4.54    | 2.05    | .04   | 0.16-8.71    |

CI, confidence interval; α-syn, alpha-synuclein; Aβ, beta-amyloid; t-tau, total tau; p-tau, phosphorylated tau.

Relationship of CSF Biomarkers With Motor and Nonmotor Features in PD

None of the CSF biomarkers correlated with, or predicted, MDS-UPDRS total motor scores. However, CSF α-syn levels differed among motor phenotypes (P < .01), with lower concentrations in participants with PIGD phenotypes (1215.40 [491.56] pg/ml) than indeterminate (1682.99 [806.14] pg/ml) and TD phenotypes (1594.85 [702.45] pg/ml). Post hoc comparisons revealed differences between PIGD and TD (P < .01; Fig. 2) and PIGD and indeterminate phenotypes (P = .03). The significant difference between motor subtypes was noted when covariates of age, gender, and disease duration were considered. CSF Aβ1-42, t-tau, and p-tau did not differ significantly among the PD motor phenotypes. None of the CSF biomarkers correlated with factors of the MDS-UPDRS parts I to IV.

The only CSF biomarker associated with cognition was Aβ1-42, which correlated weakly with MoCA remote recall (Spearman rho = 0.23, P = .02) and attention subscores (Spearman rho = −0.19, P = .05). There were no significant correlations between any of the CSF biomarkers and dichotomized RBD scores in the PD patients.

Discussion

This cross-sectional, observational study examined the levels of α-syn across multiple biofluid matrices (ie, CSF, saliva, and plasma) and multiple CSF neurodegenerative biomarkers (ie, α-syn, Aβ1-42, t-tau, and p-tau) in a large sample of “typical,” moderate-advanced PD
and matched healthy control participants. CSF α-syn strongly correlated with t-tau and modestly, but significantly, with Ab1-42 and p-tau. α-syn levels in different biofluids such as CSF, plasma, and saliva did not significantly correlate with each other. Overall, CSF α-syn, Ab1-42, and p-tau concentrations were lower in PD when compared with healthy control participants, whereas t-tau levels did not significantly differ between groups. Furthermore, the concentration of CSF α-syn, but not the other CSF neurodegenerative markers, was significantly lower in those PD patients with PIGD motor phenotypes than in TD and indeterminate phenotypes. Finally, CSF Ab1-42 levels significantly correlated with aspects of cognition on the MoCA examination.

Our findings provide insights into the α-syn levels across multiple biofluid matrices. Peripheral manifestation of Lewy type pathology has been explored as a diagnostic marker. One study noted a weak correlation (r = 0.197) of CSF and total plasma α-syn, but we did not find a significant correlation of α-syn levels in peripheral biofluids (plasma and saliva) with α-syn levels in CSF in our cohort, perhaps because of the differences in biofluid methodologies or sample characteristics. Plasma and saliva α-syn levels did not significantly differentiate PD from healthy control participants. Thus, CSF α-syn has greater diagnostic utility for PD than peripheral α-syn despite its more invasive nature for obtaining samples.

Consistent with published literature including a study of 63 de novo PD and 32 healthy control participants in PPMI, CSF α-syn levels in PD strongly correlated with t-tau, and to a lesser degree, p-tau. In BioFIND, we also identified a modest correlation of CSF α-syn and Ab1-42, which supports findings from a larger PPMI study of 660 participants (r = 0.35, P < .01). These findings suggest that CSF α-syn, Ab1-42, and tau may interact on cellular and pathophysiological levels in PD. Interactions of α-syn and tau proteins have been described in animal models and postmortem brain studies, including colocalization of tau-positive tangles and α-syn-positive Lewy bodies, and α-syn and Ab1-42 may have a synergistic relationship. A greater understanding of the molecular mechanisms of overlapping proteinopathies and their underlying pathology in PD, along with relevant biomarkers, will contribute to the development and validation of biomarkers used for diagnosis, prognosis, and therapeutic monitoring in PD.

In agreement with previous reports, CSF α-syn levels were lower in PD than in HCs and, in classification and regression models, distinguished these groups from each other. In the BioFIND cohort, CS t-tau levels did not differ significantly between groups, although p-tau levels demonstrated borderline significance (P = .05). Ab1-42 also differentiated PD patients when compared with healthy controls. We recognize, however, that many studies represent healthy controls as the ceiling of health, such as our enrollment criteria of controls with MoCA scores ≥ 26; these groupings may introduce potential bias by detecting greater differences between disease and healthy states. However, it is interesting to note that CSF Ab1-42 may relate to certain cognitive domains such as remote memory; this finding supports CSF Ab1-42 as an important potential biomarker for cognitive impairment in PD.

It is notable that CSF α-syn levels did not correlate with several disease-related characteristics, including MDS-UPDRS total motor scores (part III), motor complications (part IV), and motor experiences of daily living (part II). Furthermore, CSF α-syn did not correlate with nonmotor features including RBD, cognition as measured by the MoCA, or nonmotor experiences of daily living (MDS-UPDRS part I). The lack of a relationship between RBD and CSF α-syn in our cohort differs from a study in which oligomeric α-syn levels were elevated in the CSF and serum in PD patients with RBD when compared with PD patients without RBD and controls. The different methodologies used and the more advanced PD participants included in our study may contribute to different study findings. CSF α-syn levels may be useful for distinguishing the disease state, but may not be strong biomarkers for disease progression once already at moderate-advanced stages.

CSF α-syn levels differed among the PD motor phenotypes, with lower levels in the PIGD subtype. Although lower CSF α-syn levels may not predict motor scores, they may be useful in distinguishing motor phenotypes and for prognostication because PIGD deficits may portend a worsened motor and cognitive course of PD. The relationship of CSF α-syn, but not other CSF markers, to PIGD motor phenotype in PD also occurs in early PD as described in the PPMI cohort. Although suggestive of a potential prognostic value, longitudinal studies are needed for confirmation.

The strengths of our study include well-characterized and “typical” PD patients, rigorously collected and detailed clinical and biospecimen data, and simultaneous measurement of multiple CSF neurodegenerative markers and of α-syn across multiple biofluid sources. We probed relationships of CSF biomarkers with motor and nonmotor variables, such as RBD and MDS-UPDRS factors, which have received limited attention to date. We note several study limitations, including its cross-sectional design and lack of clinico-pathological correlations, which preclude our ability to determine relationships between biomarkers and disease progression or neuropathology. RBD assessment was based on self-report without confirmation by polysomnography, and cognition was measured with a screening questionnaire rather than a full cognitive battery. Despite this, our findings provide direction for design of more in-depth future studies. Lastly, although our CSF and plasma sample sizes are large, the number of salivary specimens was small because this assessment was added after the study had begun, and thus larger studies are needed.
The BioFIND cohort offers a unique opportunity to compare α-syn across different biofluids. In our cohort, α-syn levels in different biofluids such as CSF, plasma, and saliva did not significantly correlate with each other. In addition, the finding that CSF α-syn levels are associated with PIGD motor phenotype suggests that this marker may also be useful for providing prognostic information for PD patients.

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Supporting Data

Additional Supporting Information may be found in the online version of this article at the publisher’s website.