Non-Enzymatic Electrochemical Sensor Based on Sliver Nanoparticle-Decorated Carbon Nanotubes

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Abstract: The authors report a non-enzymatic electrochemical sensor based on a sliver nanoparticle-decorated carbon nanotube (AgNPs-MWCNT). Highly-dispersed AgNPs were loaded on the MWCNT surface through a simple and facile two-step method. The morphology, components, and the size of the AgNPs-MWCNT nanocomposite were characterized by transmission electron microscopy, X-ray diffraction, and ICP analysis. Benefitting from the synergistic effect between the AgNPs and MWCNT, the AgNPs-MWCNT nanocomposite exhibited high electrocatalytic activity for \( \text{H}_2\text{O}_2 \); the AgNPs-MWCNT electrochemical sensor was prepared by coating the AgNPs-MWCNT nanocomposite on a glassy carbon electrode, and it showed a fast and sensitive response to \( \text{H}_2\text{O}_2 \) with a linear range of 1 to 1000 \( \mu \text{M} \). The detection limit was 0.38 \( \mu \text{M} \) (S/N = 3). The sensor was applied to detect \( \text{H}_2\text{O}_2 \) in spiked human blood serum samples with satisfactory results.

Keywords: carbon nanotubes; silver nanoparticles; nonenzymatic; sensor; hydrogen peroxide

1. Introduction

Carbon nanotubes (CNTs) as perfect one-dimension nanomaterials are widely applied to various fields including energy storage, heterogeneous catalysis, sensors, biosensors, and DNA theranostic, due to their excellent physicochemical performance, unique electrochemical properties, and easy functionalization [1–6]. CNTs are easily functionalized through physical ultraphonic, chemical acidification and oxidation. The functionalized multi-wall carbon nanotubes (MWCNTs) have been used as carriers to immobilize various nanoparticles and biological molecules for various applications, such as biosensors and bioassays [7,8]. Silver is a low-cost noble metal and silver nanostructures have remarkable catalytic activity and good electron transfer efficiency [9–11]. Silver nanostructures, including nanoparticles and nanotubes, are used to construct electrochemical sensors for the ultrasensitive detection of hydrogen peroxide [12,13], proteins [14], lactose [15], glucose [16], and hydroquinone [17]. Due to the unique properties of CNT and silver nanoparticles, the preparation of CNT/AgNPs hybrid nanocomposites has received considerable interest [18–20]. Benefitting from the synergistic effect between the AgNPs and CNT, the AgNPs/CNT hybrid nanocomposite exhibits excellent performance on electron transfer and electrocatalytic activity. Various approaches including sputter deposition [21], silver mirror reaction [22], template self-volatilization [23], one-step hydrothermal method [24], electrostatic adsorption [25,26], and in-situ chemical reduction [27], were used to prepare the AgNPs-loaded CNT (AgNPs-CNT) nanocomposites. The AgNPs-CNT nanocomposite, prepared with the electrostatic adsorption method, has less AgNPs on the CNT surface, which influences its catalytic activity. The in-situ chemical reduction of \( \text{Ag}^+ \) ions in dispersed...
CNT solution would increase the amount of AgNPs on the CNT surface, but the AgNPs possess irregular morphology and size due to the uncontrolled nucleation and growth processes. Moreover, CNT aggregates in aqueous solutions, which affects its performance and applications. Although, many methods and techniques are used to prepare AgNPs-CNT nanocomposites, there is still a challenge to develop a simple and mature method for the controllable growth of AgNPs on carbon nanotubes substrate.

In this article, the highly-dispersed AgNPs with a diameter of 12 nm were loaded on MWCNTs through a simple and facile two-step procedure. The morphology and size of the AgNPs-MWCNT nanocomposite were characterized by transmission electron microscopy (TEM), X-ray diffraction, and ICP analysis. The AgNPs-MWCNT nanocomposite was used to develop a nonenzymatic electrochemical sensor for the detection of hydrogen peroxide (H$_2$O$_2$). H$_2$O$_2$ is widely used as an oxidizing agent in many fields, such as the clinical, pharmaceutical, and food industries. In addition, H$_2$O$_2$ is an important marker for oxidative stress and the side product of oxidase enzyme metabolism. Hence, the determination of H$_2$O$_2$ in trace levels in biological and various water samples are of great importance. Electrocatalytic activity of AgNPs-MWCNT nanocomposite to H$_2$O$_2$ was studied by cyclic voltammetry and amperometry was used to quantify the concentration of H$_2$O$_2$. The sensor was applied to detect H$_2$O$_2$ in spiked serum samples.

2. Results and Discussion

2.1. Characterization of AgNPs-MWCNT Nanocomposite

The AgNPs-MWCNT nanocomposite was synthesized through a two-step route. First, MWCNTs were treated with strong acids to shorten the length and introduce carboxylic groups on the CNT surface. Second, the mono-dispersed AgNPs were decorated on the MWCNTs’ surface by the in-situ reduction of AgNO$_3$ with sodium citrate. The size and morphology of the as-prepared AgNPs-MWCNT nanocomposite were characterized by transmission electron microscopy (TEM). Figure 1a,b are the TEM images of the shortened MWCNTs. One can see the length of the MWCNTs is about 200 nm (Figure 1a) and the diameter of the MWCNTs is around 20 nm (Figure 1b). Figure 1c presents the typical TEM image of the AgNPs-MWCNT nanocomposites. Large amounts of AgNPs were loaded on the MWCNTs’ surface and the distribution of AgNPs on the MWCNTs’ surface is uniform. The diameter of AgNPs ranges from 8 to 13 nm (Figure 1d). The components of the AgNPs-MWCNT were determined by inductively coupled plasma atomic emission spectrometer (ICP-AES) and the Ag content in the as-prepared AgNPs-MWCNT nanocomposite is 0.2356 wt%.

X-ray diffraction (XRD) was used to study the crystalline phases of the MWCNTs and AgNPs-MWCNT nanocomposite. As shown in Figure 2, the diffraction peaks of MWCNTs (black) at 25.8°, 43.5°, and 52.5° in the XRD pattern correspond to the lattice spacing of the carbon atom (002), (100), and (004), respectively [28]. The diffraction peaks of the AgNPs-MWCNT nanocomposite (red) at 38°, 44°, 64°, and 78° in XRD pattern ascribes to Ag (111), (200), (220), and (311), respectively. The above XRD pattern proves that the AgNPs-MWCNT nanocomposite was successfully synthesized [29].
Figure 1. Typical TEM images of the shortened MWCNTs (a and b) and the synthesized AgNPs-MWCNT nanocomposites (c and d).

Figure 2. XRD patterns of the shortened MWCNT (black) and AgNPs-MWCNT nanocomposite (red).

2.2. Electrocatalytic Activity of AgNPs-MWCNT Nanocomposite to H$_2$O$_2$

The AgNPs-MWCNT nanocomposite was used to prepare a nonenzymatic electrochemical sensor for the detection of H$_2$O$_2$. The electrocatalytic activity of the AgNPs-MWCNT nanocomposite to H$_2$O$_2$ was studied by cyclic voltammetry (CV). Figure 3a shows the typical cyclic voltammograms of the bare glassy carbon electrode (GCE), MWCNT modified GCE (MWCNT/GCE), and AgNPs-MWCNT modified GCE (AgNPs-MWCNT/GCE) in 0.1 M phosphate buffer saline (PBS, pH 7.4) containing 5 mM H$_2$O$_2$ at a potential scan rate of 100 mV/s. As shown in Figure 3a, there is no redox peak observed on the bare GCE electrode; an obvious reduction peak (peak potential: −0.55 V) was found with AgNPs-MWCNT/GCE, and the peak current was around −81.3 μA, which is much higher than that obtained with MWCNT/GCE. The above results show that the H$_2$O$_2$ were reduced at the AgNPs-MWCNT/GCE surface effectively, and the AgNPs-MWCNT has excellent electrocatalytic
activity to the reduction of H$_2$O$_2$. The electrocatalytic mechanism of AgNPs-MWCNT to H$_2$O$_2$ reduction can be illustrated as follows [30,31]:

$$
\text{H}_2\text{O}_2 + 2e^{\text{AgNP}(\text{pH 7.4})} \rightarrow 2\text{OH}^- \\
2\text{OH}^- \rightarrow \text{H}_2\text{O} + \frac{1}{2}\text{O}_2 + 2e
$$

Figure 3. (a) Cyclic voltammograms of GCE, MWCNT/GCE, and AgNPs-CNT/GCE in N$_2$-saturated 0.1M PBS solution containing 5 mM H$_2$O$_2$, potential scan rate: 100 mV/s; (b) cyclic voltammograms of AgNPs-MWCNT/GCE in N$_2$-saturated 0.1M PBS in the presence of H$_2$O$_2$ with different concentrations (1, 2, 3, 4, 5, 6, 7, 8, 9, and 10 mM), potential scan rate: 100 mV/s; (c) relationship between the reduction peak current and H$_2$O$_2$ concentrations. (d) Cyclic voltammograms of AgNPs-MWCNT/GCE in N$_2$-saturated 0.1M PBS containing 1 mM H$_2$O$_2$ at different scan rates (40, 60, 80, 100, 120, 140, and 160 mV/s); (e) relationship between the peak current vs. square root of potential scan rate.

The AgNPs on the MWCNT’s surface accelerate the reduction of H$_2$O$_2$ to generate hydroxide ions (OH$^-$). The reduction peak potential of H$_2$O$_2$ shifts from −0.85 V on MWCNT/GCE to −0.55 V on AgNPs-MWCNT/GCE. The enhanced reduction current and the shifts of peak potential indicate that the AgNPs-MWCNT nanocomposite has excellent electrocatalytic activities to H$_2$O$_2$ [24]. Figure 3b presents the CV curves of the AgNPs-MWCNT/GCE in the presence of different concentrations of H$_2$O$_2$ (0 to 10 mM). One can see the peak current at −0.55 V increases with the increase of H$_2$O$_2$ concentrations. There is a linear relationship (R = 0.9976) between the H$_2$O$_2$ concentration and the peak current (Figure 3c), indicating that the AgNPs-MWCNT/GCE can be used as a nonenzymatic electrochemical sensor for the detection of H$_2$O$_2$. The effect of the potential scan rate on the peak current was studied by varying the potential scan rate from 40 to 160 mV/s. Figure 3d presents the cyclic voltammograms of the AgNPs-MWCNT/GCE under the different potential scan rates. The reduction peak currents increase with the increase of potential scan rates. There is a good linear relationship between the square root of potential scan rate and the peak current (Figure 3e) and the correlation coefficient is 0.9989, indicating a classic diffusion-controlled process of H$_2$O$_2$ on AgNPs-MWCNT/GCE [32].

2.3. Amperometric Response of H$_2$O$_2$ on AgNPs-MWCNT/GCE

Amperometry was used to inspect the possibility of the AgNPs-MWCNT/GCE as a nonenzymatic electrochemical sensor for the detection of H$_2$O$_2$. In order to obtain the optimum working potential, the amperometric current of 1.0 mM H$_2$O$_2$ on AgNPs-MWCNT/GCE was measured at the working
potential ranging from −0.1 to −0.6 V with an interval of 0.1 V (Figure 4a). It can be seen that the amperometric response of \( \text{H}_2\text{O}_2 \) increased with the decrease of the potentials, and the highest response was obtained at −0.5 V. A further decrease of the working potential led to a decrease of the current. Therefore, a potential of −0.5 V was selected as the optimum potential for the amperometric detection of \( \text{H}_2\text{O}_2 \). We aimed to detect \( \text{H}_2\text{O}_2 \) in the blood or the physiological system with the AgNPs-MWCNT/GCE sensor, so we chose the pH value of 7.4 to match the physiological condition without further optimization. The AgNPs-MWCNT with the concentrations ranging from 1 mg mL\(^{-1}\) to 5 mg mL\(^{-1}\) were used to prepare the sensors. It was found the amperometric responses of the AgNPs-MWCNT/GCE increased with the increase of the amount of AgNPs-MWCNT on the GCE surface, a further concentration increase resulted in signal saturation. As a result, 4 mg mL\(^{-1}\) of AgNPs-MWCNT was used to prepare the sensor. Figure 4b shows the typical current-time (i–t) curve of AgNPs-MWCNT/GCE with the successive addition of \( \text{H}_2\text{O}_2 \) in 0.1 M PBS (pH 7.4) at room temperature under nitrogen atmosphere. One can see that the responsive current of AgNPs-MWCNT/GCE increases rapidly with the successive injection of \( \text{H}_2\text{O}_2 \) and reaches steady state within 3s. The sensitive and fast response of \( \text{H}_2\text{O}_2 \) on the AgNPs-MWCNT/GCE may be due to the large surface area of the AgNPs-MWCNT nanocomposite, which possesses a large amount of electrocatalytic active sites to improve the sensing sensitivity [33]. Figure 4c shows the relationship between the concentration of \( \text{H}_2\text{O}_2 \) and the step current. There are two linear ranges from 1 to 10 µM \( (I_r(\mu A) = -0.1806 C (\mu M) - 5.4262, R^2 = 0.9937) \), and from 10 to 1000 µM \( (I_r(\mu A) = -0.0125 C (\mu M) - 7.6935, R^2 = 0.9975) \). According to the linear regression equation, the detection limit of \( \text{H}_2\text{O}_2 \) on AgNPs-MWCNT/GCE is estimated to be 0.38 µM (S/N = 3). Moreover, the sensitivity of the detection was about 2556 µA cm\(^{-2}\) mM\(^{-1}\) at the low concentration range (1 to 10 µM). The above results indicate that the AgNPs-MWCNT nanocomposite possesses excellent electrocatalytic property to \( \text{H}_2\text{O}_2 \), which can be ascribed to the following reasons—first, both AgNPs and MWCNTs have excellent electron transfer efficiency, the hybrid nanocomposite (AgNPs-MWCNT) has higher electron transfer efficiency, which can enhance the reaction rate of hydrogen peroxide; second, the synthesized AgNPs possess a smaller size, well dispersion, and good uniformity deposited on MWCNT surface, which can enlarge the surface area and increase the electrocatalytic active sites of the nanocomposite; third, the synergistic effect was generated between AgNPs and CNTs, which can improve the sensitivity of the \( \text{H}_2\text{O}_2 \) detection [34].

Table 1 displays the detection limits and linear ranges of AgNPs–MWCNT/GCE and the other related sensors in literature [35–37]. The limit of detection (LOD) of the AgNPs–MWCNT sensor in this work is comparable or better than that of the other nonenzymatic \( \text{H}_2\text{O}_2 \) sensors.

| Modify Electrode       | Linear Range (µM) | LOD (µM) | Ref.   |
|------------------------|-------------------|----------|--------|
| PtNPs-MWCNTs           | 2–3800            | 0.7      | [35]   |
| AuNPs-MWCNT            | 20–300            | 0.4      | [36]   |
| AgNCs-GO               | 20–10000          | 3.0      | [13]   |
| Ag NPs-MWCNT           | 50–17000          | 0.5      | [37]   |
| AgNPs-MWCNT-rGO        | 100–100000        | 0.9      | [24]   |
| AgNPs-MWCNT            | 1–1000            | 0.38     | This work |
Figure 4. (a) The responsive currents of AgNPs-MWCNT/GCE in the presence of 1 mM H\textsubscript{2}O\textsubscript{2} at different working potential from −0.1 to −0.6 V with 0.1 V interval; (b) typical amperometric responses of AgNPs-MWCNT/GCE to the successive addition of H\textsubscript{2}O\textsubscript{2} in 0.1 M PBS at working potential of −0.5 V; (c) the dependence of the responses of electrodes on H\textsubscript{2}O\textsubscript{2} concentrations.

2.4. Selectivity, Reproducibility, and Stability of the AgNPs-MWCNT/GCE Sensor

Figure 5a shows the amperometric responses of 0.1 mM and 1.0 mM H\textsubscript{2}O\textsubscript{2} in the absence and presence of 0.5 M of potential interferences including ascorbic acid, NaCl, fructose, sucrose, and glucose (last injection). One can see that similar responsive currents were obtained in the absence and presence of interferences. In addition, there was no responsive current observed when injecting the interferences in the absence of H\textsubscript{2}O\textsubscript{2}. The above results indicate that the AgNPs-MWCNT/GCE sensor has excellent selectivity toward H\textsubscript{2}O\textsubscript{2} detection. The reproducibility of the AgNPs-MWCNT/GCE sensor was studied by measuring 0.1 mM H\textsubscript{2}O\textsubscript{2} with five electrodes fabricated at the same batch (Figure 5b). Similar responsive currents were obtained and the relative standard deviation (RSD) of the five measurements was 1.2%, indicating the excellent reproducibility of the sensor. The stability of AgNPs-MWCNT/GCE was tested by measuring the responsive current of 0.1 mM H\textsubscript{2}O\textsubscript{2} for 5 days, and the sensor was kept in room temperature. As shown in Figure 5c, no obvious current difference was observed in five days, and the RSD of the measured currents was 1.8%. The above experimental results indicate that the AgNPs-MWCNT/GCE has good stability.
3. Materials and Methods

3.1. Materials and Instruments

The AgNPs-MWCNT/GCE sensor was applied to detect $\text{H}_2\text{O}_2$ in human blood serum (HBS). The HBS was diluted 50 times by 0.1 M PBS (pH 7.4), and the $\text{H}_2\text{O}_2$ standards were added to the diluted HBS. The concentrations of $\text{H}_2\text{O}_2$ in the diluted HBS were fixed at 5.0, 50, and 500 µM, respectively. Each sample was detected three times at same condition. The data are displayed in Table 2. The recovery rates of $\text{H}_2\text{O}_2$ in the diluted HBS were 90.2, 92.8, and 96.8% respectively; and the RSD were 1.75, 0.47, and 0.61% respectively. The results indicated that the fabricated AgNPs-MWCNT/GC sensor has good ability for $\text{H}_2\text{O}_2$ detection in practical application.

| Samples | Added (µM) | Founded (µM) | Recovery (%) | RSD (%) |
|---------|------------|--------------|--------------|---------|
| 1       | 5.0        | 4.5          | 90.2         | 1.75    |
| 2       | 100        | 92.8         | 92.8         | 0.47    |
| 3       | 500        | 483.8        | 96.8         | 0.61    |

Table 2. Results of standard addition and recovery in human blood serum.

2.5. Detection of $\text{H}_2\text{O}_2$ in Human Blood Serum

The AgNPs-MWCNT/GCE sensor was applied to detect $\text{H}_2\text{O}_2$ in human blood serum (HBS). The HBS was diluted 50 times by 0.1 M PBS (pH 7.4), and the $\text{H}_2\text{O}_2$ standards were added to the diluted HBS. The concentrations of $\text{H}_2\text{O}_2$ in the diluted HBS were fixed at 5.0, 50, and 500 µM, respectively. Each sample was detected three times at same condition. The data are displayed in Table 2. The recovery rates of $\text{H}_2\text{O}_2$ in the diluted HBS were 90.2, 92.8, and 96.8% respectively; and the RSD were 1.75, 0.47, and 0.61% respectively. The results indicated that the fabricated AgNPs-MWCNT/GC sensor has good ability for $\text{H}_2\text{O}_2$ detection in practical application.

![Chronoamperometric curve](image-url)

**Figure 5.** (a) Chronoamperometric curve (black) of the AgNPs-MWCNT/GCE in response to the successive addition of 0.1 mM $\text{H}_2\text{O}_2$, 5 mM ascorbic acid, 5 mM NaCl, 5 mM fructose, 5 mM sucrose, 5 mM glucose and 0.1 mM $\text{H}_2\text{O}_2$ in 0.1 M PBS at a working potential of −0.5 V; chronoamperometric curve (red) of the AgNPs-MWCNT/GCE in response to the successive addition of 1 mM $\text{H}_2\text{O}_2$, 5 mM ascorbic acid, 5 mM NaCl, 5 mM fructose, 5 mM sucrose, 5 mM glucose and 1 mM $\text{H}_2\text{O}_2$ in PBS at a working potential of −0.5 V. (b) Current responses of five equally fabricated sensors to 0.1 mM $\text{H}_2\text{O}_2$. (c) Current responses of the AgNPs-MWCNT/GCE biosensor to 0.5 mM $\text{H}_2\text{O}_2$. Amperometric measurements were performed in 5 days using the same sensor.

Table 2. Results of standard addition and recovery in human blood serum.
3. Materials and Methods

3.1. Materials and Instruments

MWCNT was purchased from Nanjing XFNano Materials Technology Co., Ltd. (Nanjing, China). Silver nitrate (AgNO$_3$, 99.5%), sodium citrate, glucose, ascorbic acid, fructose, phosphate buffer saline (PBS, PH 7.4, 0.1 M), and lactose were purchased from Sigma (Shanghai, China). Sulfuric acid (H$_2$SO$_4$, 98%), nitric acid (HNO$_3$, 98%), and hydrogen peroxide (H$_2$O$_2$) were purchased from Sinopharm Chemical Reagent Co., Ltd. (Beijing, China). Human blood plasma was purchased from Senbeijia Biotechnology Co., Ltd. (Nanjing, China). All the reagents were of analytical grade and used as received without further purification. Ultrapure water (resistance >18 MΩ cm$^{-1}$, Milli-Q purification system) was used in all of the experiments.

Electrochemical studies were performed on a CHI 660D electrochemical system (Chenhua, Shanghai, China). The three-electrode system consists of a AgNPs-MWCNT modified glassy carbon working electrode, a Ag/AgCl/KCl (saturated) reference electrode, and a platinum wire counter electrode. The morphology and size of the CNT and AgNPs-MWCNT were characterized by TEM (JEOL, Kyoto, Japan, 2100F, 200KV). XRD measurements were performed on a Shimadzu XRD-6000 (Shimadzu, Kyoto, Japan) using Cu K$_\alpha$ radiation (1.5406 Å) and operated at 36 kV and 20 mA. All the experiments were carried out at room temperature. The content of Ag in the AgNPs-MWCNT nanocomposite was determined by ICP-AES (TJA IRIS Advantage ER/S, Thermo, Franklin, MA, USA).

3.2. Synthesis of AgNPs-MWCNT Nanocomposite

The MWCNTs were shortened and functionalized with the previous reported method [38]. Briefly, 40 mg of MWCNTs, 19.2 mL of concentrated sulfuric acid, and 6.4 mL of concentrated nitric acid were added to a glass flask with a volume of 50 mL. The mixture was sonicated for 6 h at room temperature and separated through centrifugation at 8,000 rpm for 10 min; then the MWCNT pellet was washed several times with deionized water and ethanol, respectively. The final product was dried for 24 h at 35 °C under nitrogen atmosphere.

The AgNPs-MWCNT nanocomposite was prepared by a reported method with a slight modification [39]. Three milligrams of the functionalized MWCNTs and 600 µL of 0.1 g mL$^{-1}$ sodium citrate solution were dispersed in 30 mL of deionized water, the mixture was heated at 100 °C for 5 h under vigorous stirring. Then, the mixture was cooled to 60 °C, and the 96.6 µL of 0.01 M silver nitrate solution was added to the mixture. The mixture solution continued to react for 5 h at 60 °C under vigorous stirring. After the system was cooled to room temperature, the resulting product (AgNPs-MWCNT) was separated through centrifugation, washed three times with deionized water and ethanol, and dried for 48 h at 35 °C under nitrogen atmosphere.

3.3. Preparation of the AgNPs-MWCNT/GCE

The GCE (Φ = 3 mm) was polished with 0.5 and 0.05 µm alumina slurries, respectively, and followed by sonication in deionized water and ethanol. The electrode was dried under nitrogen atmosphere. A 10 µL of AgNPs-MWCNT aqueous solution (4 mg mL$^{-1}$) was dropped on the cleaned GCE surface and dried in air, then a 10 µL of Nafion (0.05 wt%) was dropped again and dried at room temperature. The MWCNT-modified GCE was prepared in the same way.

3.4. Electrochemical Measurements

A three-electrode cell contains AgNPs-MWCNT/GCE as working electrode, platinum wire as the counter electrode, and Ag/AgCl as the reference electrode. Cyclic voltammograms (CV) were obtained at the potential range between 0.2 and −1.6 V in 0.1 M PBS at pH 7.4 at a scan rate of 100 mV s$^{-1}$. Amperometric experiments were carried out in 20.0 mL of 0.1 M PBS (pH 7.4) with the successive addition of H$_2$O$_2$ under mild stirring. The electrode potential was set at −0.5 V (vs. Ag/AgCl). Before the experiment, all the PBS solution was purged with high-purity nitrogen for at least 30 min.
4. Conclusions

The AgNPs-MWCNT nanocomposite were successfully prepared through a simple and facile two-step method. The advantage of this method was that the silver nanoparticles were highly dispersed on the functionalized carbon nanotubes. The AgNPs-MWCNT nanocomposite was used to prepare a nonenzymatic electrochemical sensor for the detection of H$_2$O$_2$ with wide linear range, low detection limit, and excellent reproducibility and stability. The sensor was applied to detect H$_2$O$_2$ in spiked human blood serum with satisfactory results.

Author Contributions: D.X., and B.H. did the experiments, analyzed the data, and wrote the paper. L.Q. and G.L. and X.Z. conceived, designed the experiment, and elaborated the paper, and all authors approved the final paper.

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**Sample Availability:** The AgNPs-MWCNT nanocomposite are available from the authors.

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