The recent trend in mycobacterial strain diversity among extra pulmonary lymph node tuberculosis and their association with drug resistance and the host immunological response in South India

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Abstract

Background: Tuberculosis (TB) though primarily affects the lungs it may also affect the other parts of the body and referred as extra pulmonary (EPTB). This study is focused on understanding the genetic diversity and molecular epidemiology of Mycobacterium tuberculosis (M.tb) among tuberculous lymphadenitis (TBL), a form of EPTB patients identified in Chennai, Tamil Nadu.

Methods: The genetic diversity was identified by performing spoligotyping on the M.tb clinical isolates that were recovered from lymph node samples. A total of 71 M.tb isolates were recovered from extra pulmonary lymph node samples and subjected to Drug susceptibility testing and spoligotyping was carried out. In addition, immunological characterization from blood of same individuals from whom M.tb was isolated was carried out between the two major lineages groups East African Indian 3 (EAI3) and non-EAI3 strains by ELISA. The results of spoligotyping patterns were compared with the world Spoligotyping Database of Institute Pasteur de Guadeloupe (SpolDB4).

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Results: We found 41 spoligotype patterns and their associated lineages. Out of 41 spoligotype pattern, only 22 patterns are available in the spoldB4 database with Spoligotype international Type (SIT) number and remaining patterns were orphan strains without SIT number. The most predominant spoligotype lineage that was found in lymph node sample in this region of India was EAI (36), followed by central Asian strain (CAS) (6), T1 (5), Beijing (3), Latin American & Mediterranean (LAM) (2), U (1), X2 (1) and orphan (22). In addition to EAI, CAS and Beijing, our study identified the presence of orphan and unique spoligotyping patterns in Chennai region. We observed six drug resistant isolates. Out of six drug resistant isolates, four were resistant to isoniazid drug and associated with EAI family. Moreover, we observed increased levels of type 2 and type 17 cytokine profiles between EAI3 and non-EAI family, infected individuals.

Conclusions: The study confirms that EAI lineage to be the most predominant lineages in EPTB patients with lymphadenitis and were found to have increased type 1 and type 17 proinflammatory cytokine profiles.

Keywords: Extra pulmonary tuberculosis, Mycobacterium tuberculosis, Spoligotyping, Th1 and Th17 cytokines, Lymph node, Mycobacterial lineage

Background

Tuberculosis (TB) occurs worldwide and it is caused by an infectious agent Mycobacterium tuberculosis (M.tb). According to World health organization (WHO), there were 1.3 million TB deaths was reported among HIV negative people and an additional 3, 00,000 deaths among HIV-positive people were reported in 2017. TB affected about 10 million people; among them 90% were adults, 9% were people living with HIV [1]. The two types of clinical manifestations of TB are pulmonary (PTB) and Extra-pulmonary (EPTB) tuberculosis. EPTB involves TB affects body parts other than the lungs, which include lymph nodes, skins, joints, or meninges. Among 6.4 million incident cases, 14% represents extra-pulmonary TB [1]. In India, about 15 to 20% of EPTB occurs in immune-competent patients and 50% of cases were reported in HIV-positive individuals [2]. Among them, lymph node TB (35%), pleural TB (20%), bone marrow TB (10%), genitourinary TB (9%), cerebrospinal, abdominal, skin, etc. accounts for 26% cases [2]. Though EPTB is not contagious, it has a huge impact on mortality and morbidity rate worldwide. Mycobacterium tuberculosis complex (MTBC) is the most common etiological agent in TB including EPTB and their diagnosis and therapeutics are highly challengeable [3].

Epidemiology of EPTB can be analyzed by genotyping of Mycobacterium tuberculosis, which will be helpful to detect unsuspected transmission, reinfection and relapse and to identify false-positive cultures [4]. The available genotyping methods are IS6110-RFLP, Polymorphic GC-rich repetitive sequences (PGRS)-RFLP, Mycobacterial interspersed repetitive unit (MIRU)-VNTR typing and Spoligotyping. Of the available genotyping tools, Spoligotyping and MIRU-VNTR is considered the most suitable tool in clinical settings [5].

Since Spoligotyping (Spacer Oligonucleotide typing) can simultaneous detect and type different lineages of M. tuberculosis bacterial complex (MTBC). The clinical data obtained by this method had a significant impact on immune surveillance and drug treatment regimens [6]. Previous studies demonstrated that M. tuberculosis strains have different epidemiology patterns. Evidence from a variety of sources suggests that MTBC lineages have different phenotypic properties. According to Centers for Disease Control and Prevention (CDC), MTBC genetic lineages are of two categories: Modern lineages and Ancestral lineages. The Modern lineages include East-Asian (Beijing), East-African-Indian (EAI) and Euro-American. The Ancestral lineages include Indo-Oceanic, M. bovis and M. africanum (West African 1 & West African 2) [7]. Therefore, spoligotypic information on clinical isolates can reveal the phylogenetic and evolutionary relationships among those strains. Previous studies have also documented that EAI (ancient strain) trigger significantly increased expression of certain (TNF-α, IL-1β, IL-12) cytokines and apoptosis than Beijing (modern lineage) and H37Rv (laboratory strain) up on in ex-vivo. Also, it has been shown that ancient strains were less lethal when compared to modern strains and they stimulate greater pro-inflammatory host response. In contrast, infection with Mtb Beijing strains might induce non-uniform immune responses [8–11]. In a given situation, characterization of those lineages in EPTB may provide better insights on strain pathogenicity and further will help us to document global tuberculosis epidemiology [12]. Therefore, in this study, we aimed to genotype 71 clinical isolates of M. tuberculosis from patients with extrapulmonary tuberculosis (EPTB) to identify the predominant spoligotyping pattern of strains in lymph node samples isolated from Southern India and studied strain specific immune responses among the infected individuals.
Methods
Clinical setting
The strains were received from an ongoing, clinical trial to study the efficacy and tolerability of a 4-month regimen containing ofloxacin compared to the standard 6-month regimen in the treatment of patients with superficial lymph node TB by National Institute for Research in Tuberculosis (NIRT), Chennai which is a Supranational Reference Laboratory and a WHO collaborating center for TB research and training. From this ongoing study, 71 mycobacterial isolates retrieved from suspected extra pulmonary cases and utilized for performing mycobacterial strain characterization (from the year 2013–2015). The Institutional Ethical Committee (IEC) of National Institute for Research in Tuberculosis (NIRT) approved the study and written informed consent was obtained from the patients before collecting the samples (NIRT IEC 2009 007).

Sample processing, biochemical characterization and identification
The lymph node samples were collected and transported in Kirchner’s medium to the laboratory. Lymph node sample processing: The biopsy sample (Lymph node) is cut into small pieces and grinded with sterile Teflon grinding rod, direct smear is made from the homogenate. The homogenate is treated with 1 ml 5% H2SO4 for 15 min and neutralized with sterile distilled water. The deposit is washed and inoculated onto two slopes each of plain Lowenstein Jensen (LJ) and LJ Sodium Pyruvate solid medium, remaining deposit is inoculated into two bottles of SK. LJ medium and SP medium are read for 8 weeks every week before declaring as negative. The SK medium is subculture on to LJ medium if positive. The drug susceptibility testing for First- and second-line anti-TB drugs were carried out for 71 isolates of M.tb. The minimum inhibitory concentration (MIC) was used to predict the sensitive and resistance patterns as per the guidelines recommended by WHO for First- and second-line anti-TB drugs (https://www.who.int/tb/publications/2018/WHO_technical_report_concentrations_TB_drug_susceptibility/en/). In Brief MIC method was performed using 4 mg moist weight per microlitre (ml) of culture suspension, one-third loopful of 2–3 wk. old culture on L-J medium was suspended in 1 ml of sterile distilled water and vortexed to obtain uniform suspension. The coarse particles or clumps in the suspension were allowed to settle at room temperature. For MIC method, 10 μl of the suspension was inoculated onto drug containing and drug free L-J medium. Results were read after 28 days of incubation at 37 °C for MIC methods. Isolates with ≥20 colony counts (1+ grading) were considered resistant to the particular drug concentration. In addition, MPT-64 Ag detection test were performed for these 71 isolates using SD Bioline TBAg MPT64 Rapid (Standard diagnostics Seoul, South Korea) Kit [13] to confirm M.tb.

DNA extraction
The DNA extraction from the confirmed M. tuberculosis positive patients was carried out as described [14]. The mycobacterial cultures were grown in LJ medium and scrapped in to 1 ml TE (Tris EDTA) buffer. At 80 °C, the cultures were heat killed for 30 min. To that, 50 μl of lysozyme (10 mg/ml) was added and the isolates were incubated at 37 °C. On the next day, 70 μl of 10% SDS and 6 μl of proteinase K (50 mg/ml) was added and further incubated at 65 °C for 15 min. Then, 100 μl of 5 M NaCl and 80 μl of CTAB were added, incubated again at 65 °C for 15 min. To the culture, 700 μl of chloroform/isomyl alcohol (24:1) was added and centrifugation was carried out for 20 min at 10,000 g at 25 °C. The supernatant was collected into a new Eppendorf tube. To precipitate the DNA, 600 μl of ice-cold isopropanol was added to the tubes and incubated at – 20 °C overnight. On the third day, Eppendorf tubes were subjected to centrifugation at 12,000 rpm for 15 min and the pellet was washed with 500 μl of 70% ethanol. The extracted genomic DNA was re-suspended in TE buffer and stored at 20 °C until use. To confirm the presence of DNA, the extracted DNA was run on 1% agarose gel electrophoresis and quantified using Nano drop.

Spoligotyping
Spoligotyping was performed as described previously [15]. DRa and DRb primers were used for amplifying the direct repeat (DR) region in the genome of MTB complex. M. tuberculosis H37Rv strain and Mycobacterium bovis BCG P3 chromosomal DNA were used as positive controls. The molecular biology grade water served as negative control. The commercial membrane pre-coated with spacer-oligos, which represents the spacer region of known sequences, was used for hybridizing the amplified product. The membrane was incubated with streptavidin-peroxide and ECL to visualize the presence/absence of spacer on X-ray film as black squares. Based on the combination of the presence/absence of theses spacers in a M.tb strain is distinguished as spoligotype. The available spoligotyping patterns in international spoligotype database (SpoIDB4) were used for comparing the spoligotyping patterns of lymph node samples obtained from this study and SIT number is assigned.

Elisa
In order to study the host immune response, plasma samples obtained from EPTB infected with respective strains were used to measure the systemic levels of different (IFNγ, TNFa, IL-2, IL-4, IL-5, IL-13, IL-12, IL-17A, IL-22, IL-10, IL-1α, IL-1β, IL-6 and GM-CSF)
cytokines and CC/CXC chemokines (CCL-1/1-309, CCL2/MCP-1, CCL3/MIP1α, CCL4/MIP-1β and CCL11/eotaxin, CXCL-1/Gro-α, CXCL2/Gro-β, CXCL9/MIG, CXCL10/IP-10 and CXCL11/ITAC 1) by bio-plex ELISA. The experiment was carried out according to the manufactures instruction provided (R & D systems).

Data analysis
The immunological parameters were analyzed using GraphPad PRISM (GraphPad Software, Inc.,San Diego, CA, USA). Geometric means (GM) were used for measurements of central tendency. Finally, Mann-Whitney U test was used to find the statistically significant differences among the cytokines and chemokines.

Results
We recovered 71 *Mycobacterium tuberculosis* strains from lymph node collected from EPTB patients recruited under this study. Majority of the patients recruited in this study were aged more than 18 years and other demographic and clinical details were reported in supplementary Table 1.

Drug resistance and EPTB recurrence
Out of the 71 isolates tested, 65 isolates were sensitive to all first line and second line anti-TB drugs. Four isolates were resistant to isoniazid; one isolate was mono resistant to rifampicin and one isolate shows resistant to ofloxacin (Table 1).

Spoligotyping
The isolated cultures from the lymph node were subjected to spoligotyping and it resulted in 41 spoligotype patterns as per the SpolDB4 database [16]. The resulted 41-spoligotype patterns and their associated lineages are shown in Fig. 1. The phylogenetic tree for the spoligotype pattern was generated using unweighted pair group method and arithmetic mean (Fig. 2). Out of 41 spoligotype pattern, only 22 patterns are available in the spoldB4 database with Spoligo international Type (SIT) number and remaining patterns were identified as orphan strains without SIT number. The most predominant spoligotype lineage that was found in lymph node sample in this region of India was EAI3 (21 Nos) followed by EA15 (9 Nos), EA11 (5 Nos) and EA16 (1 Nos). The sub-lineages identified from lymph node samples are summarized in Table 1. A total of 21 isolates belongs to EA13, comprising 20 isolates in EA13_IND and 1 isolate in EA13_IND 473. The next predominant lineage of EA1 family is EA15 (9 No) which are further classified in to different sub lineages as follows: EA15 126 (2 Nos), EA15 340 (1 No), EA15 355 (1 No), EA15 474 (1 No), EA15 517 (1 No), EA15 733 (1 No) AND EA15 938 (2 Nos). The third predominant lineage of EA1 family is EA11 (5 Nos) and was further identified to belong to four sub-lineages EA11_SOM 270 (1 No), EA11_SOM 48 (3 Nos) and EA11_SOM 734 (1 No) EA16 (1 No), Total of 18 new variants of EA1 were found without SIT numbers. The next prevalent lineage among 71 isolates of lymph node sample is CAS family (6 Nos). A total of 6 isolates belongs to CAS, comprising 3 isolates

| S. NO | SPOLOGOTYPES   | NO OF ISOLATES (NO OF RESISTANT STRAINS) |
|-------|----------------|------------------------------------------|
| 1.    | Beijing 1      | 3 (c-1)                                  |
| 2.    | CAS 1345       | 1                                         |
| 3.    | CAS 142        | 1                                         |
| 4.    | CAS 601        | 1                                         |
| 5.    | CAS1_DELHI 26  | 3                                         |
| 6.    | EAI 3 Orphan   | 9 (a-3)                                  |
| 7.    | EAI 5 Orphan   | 9                                         |
| 8.    | EAI1_SOM 270   | 1                                         |
| 9.    | EAI1_SOM 48    | 3 (d-2)                                  |
| 10.   | EAI1_SOM 734   | 1                                         |
| 11.   | EAI1_IND 11    | 20                                        |
| 12.   | EAI1_IND 473   | 1                                         |
| 13.   | EAI 126        | 2                                         |
| 14.   | EAI 340        | 1                                         |
| 15.   | EAI 355        | 1                                         |
| 16.   | EAI 474        | 1                                         |
| 17.   | EAI 517        | 1                                         |
| 18.   | EAI 733        | 1                                         |
| 19.   | EAI 938        | 2                                         |
| 20.   | EAI6_BGD1 591  | 1 (a-1)                                  |
| 21.   | Harleem1 Orphan| 3                                         |
| 22.   | LAM9 42        | 2                                         |
| 23.   | Orphan         | 22                                        |
| 24.   | S Orphan       | 1                                         |
| 25.   | T1 53          | 5 (b-1)                                  |
| 26.   | U 172          | 1                                         |
| 27.   | X2 137         | 1                                         |

*a*-Isoniazid resistant; *b*-Rifampicin resistant; *c*-Ofloxacin resistant; *d*-Recurrence

Sub-lineages identified from lymph node samples
Among the EA1 family, the most predominant lineage was EA13 (21 Nos) followed by EA15 (9 Nos), EA11 (5 Nos) and EA16 (1 Nos). The sub-lineages identified from lymph node samples are summarized in Table 1. Among the EA1 family, the most predominant lineage was EA13 (21 Nos) followed by EA15 (9 Nos), EA11 (5 Nos) and EA16 (1 Nos). The sub-lineages identified from lymph node samples are summarized in Table 1. Among the EA1 family, the most predominant lineage was EA13 (21 Nos) followed by EA15 (9 Nos), EA11 (5 Nos) and EA16 (1 Nos). The sub-lineages identified from lymph node samples are summarized in Table 1. Among the EA1 family, the most predominant lineage was EA13 (21 Nos) followed by EA15 (9 Nos), EA11 (5 Nos) and EA16 (1 Nos). The sub-lineages identified from lymph node samples are summarized in Table 1.
in CAS_DELHI 26 followed by CAS 1345 (1 No), CAS 142 (1 No) and CAS 601 (1 No). The existing spolDB4 database was compared and the patterns, which do not match the existing information, are called orphan strains. A total of 22 orphan strains were identified from this study.

**Spoligotyping pattern and their associated drug resistance**

Out of 71 isolates tested, 65 were sensitive to all the first- and second-line anti-TB drugs and belong to different lineages like EAI, CAS, LAM, U and X2. The spoligotype patterns and their associated drug resistance were analyzed in this study. We observed six drug resistant isolates. Out of six drug resistant isolates, four were resistant to isoniazid drug and associated with EAI family. Mostly, the sub-lineages of EAI3 orphan (3 Nos) and EA16_BGD1 (1 No) are associated with H mono resistance (Table 1). The remaining two strains were mono resistant to rifampicin and ofloxacin and belong to T1 and Beijing lineage respectively. In addition, we found recurrence of the disease in two patients out of 71 patients tested, they were primarily infected with EA11_ SOM sub-lineage, which is different from the most prevalent circulating EA13 strains in south India (Table 1).

**Differential immune response among patients infected with EA13 and non – EA1 3 strains**

To determine whether any significant immunological differences are associated with the individuals infected with either EA13 India or non-EA13 strains, we have measured the different (type 1, type 2, type 17, IL-1 family and pro-inflammatory) cytokines using the available plasma samples of respective EPTB individuals (Supplementary Fig. 1). Among them, the plasma levels of IL-2 and IL-1α were significantly higher in individuals infected with non-EA13 India strains upon comparison with EA13 India strains. In contrast, IL-13 cytokine level was significantly reduced in non-EA13 India strain infected individuals when compared to EA13 India strains (Fig. 3a). Similar to cytokines, we have also examined the systemic levels of different CC chemokines between the EA13 India and non-EA13 strains. As shown in Fig. 3b, the CC (CCL-3/ MIP-1α and CCL-4/ MIP-1β) chemokines were significantly different (diminished in EA13 India strain infected individuals) between the two study groups.

**Discussion**

A major public health concern, which still remains and contributes to the world epidemiology, is tuberculosis. For development of effective infection control and vaccines, the frequency of various *M.tbc* genotype and their transmission patterns need to be analyzed using molecular tools such as spoligotyping and MIRU-VNTR. To differentiate recurrence, re-infection and mixed infections in multiple strains of *M.tbc* these genotyping methods are widely used [17, 18]. Previous reports from our group have already reported the circulating lineages of south India from pulmonary TB samples [19]. However, this was the first report from our group, which focuses on
circulating lineage of extra pulmonary lymph node samples from ongoing clinical trials. In this study, we mainly focus on circulating lineages of extrapulmonary lymph node samples and their associated drug resistance pattern. Not many molecular epidemiological studies have been performed for lymph node extra pulmonary sample. In this context, this study provides a great insight and a better understanding of circulating lineages and their associated drug resistance patterns of extra pulmonary lymph node samples.

We performed Spoligotyping and Immunological assays for the extrapulmonary lymph node samples to understand the genetic diversity of different circulating strains that were recovered from ongoing clinical trial. Our present study identified 76.05% of EAI strains from Chennai, which was isolated from lymph nodes samples,
and found to be most predominant strains circulating in south India. This is in line with the previous studies, which identified 67.5 and 59.3% of EAI strains from pulmonary M. tb isolates [19, 20]. In comparison with North India, the high prevalence of EAI strains in pulmonary MTB isolates has already been reported in Kerala and Chennai [19–22]. The highest percentage of EAI strains (89%) were previously reported in Kerala [20]. Similarly, Joseph and co-workers also reported 64.3% of EAI strains in 2013 [21]. Various reports from North India mentioned 3.8% EAI strains from Delhi [18], 4.44% EAI strains from Maharashtra [23], 9.6% in UP and Varanasi [24] and 10% from the whole of north India [3]. Singh et al., 2007 reported that EAI lineages are more predominant in south India, whereas the CAS family is predominating the northern parts of India [25]. The origin, evolution and migration of two populations in India and the differences in lineages shows the presumption that Mycobacterium tuberculosis strains might have originated and evolved in a different way [3]. Similar study showed that Euro-American lineage were three times more likely to generate a secondary case during the 11-year study period than patients infected with any other strain. The data clearly shows that the immune response and genetic background plays a major role in deciding the infection with a particular lineage of Mycobacterium tuberculosis [26]. In the present study, we identified one EAI6_BGD1 from lymph node samples. A similar finding has been reported for the first time in Puducherry [27]. In the current study, drug susceptibility testing was carried out for all the isolates of lymph node samples. Majority of the strains were susceptible to first line drugs and six isolates were resistant to drugs like rifampicin, isoniazid and ofloxacin. Out of six isolates tested, all the four isolates are from EAI family and were resistant to Isoniazid. These resistant strains belong to EAI3 orphan strains (3 Isolates) and EAI6_BGD1 (1 Isolate).
The central and Middle Eastern Asia (CAS) family were the second largest Clade identified in our study; this is reported to be the predominant lineage in North India previously. Our study identifies 7.89% of CAS strain from lymph node samples, which accounts for the low prevalence of CAS family in Chennai. This is similar to the findings of previous research workers who also recorded the low prevalence of CAS family in south India, which accounts 1.5% in Trivandrum [20], 3.5% in Chennai [19] and 3.5% in Kerala [21]. Contrary to this low prevalence of CAS family in south India, North India recorded a very high prevalence of CAS family (57.2%) from extra pulmonary samples [3]. Similarly, Singh et al., 2007 reported 56.5%, 53.6 and 44.6% from Delhi, Pune and Lucknow, respectively [20]. This evidence clearly suggests that there are contrasting lineages circulating in north and south India. It is well-established historical fact that North Indians belongs to the Aryan descent whereas, South Indians belongs to Dravidian culture. This might be contributing for the demarcation of prevalent strains in different parts of India. In contrast, a study by Thomas et al., reported the higher prevalence of CAS family in Andhra Pradesh. This could be due to the migration of people to Hyderabad from Middle East countries and north India which contributes for the increased prevalence of CAS family.

The prevalence for Beijing strains in different parts of India ranges from 2.2% [19] to 11.5% [20] for pulmonary specimens. In the case of extra pulmonary specimens, the prevalence was slightly higher with the percentage of 10 and 18.8% [3, 28]. The Beijing strain was found to be more prevalent in multi drug extrapulmonary TB specimens and treatment failure cases with the percentage of 63.6% [23, 28]. In our study, Beijing strain was identified at a rate of 3.94% from lymph node. One of our lymph node samples were found to be resistant to Ofloxacin and associated with Beijing family. Previous studies from rural areas of Hyderabad and Mumbai have shown that Beijing strains were absent. However, in urban population of Andhra Pradesh it was prevalent at a rate of 6.8 and 10.4% among treatment failure cases [23, 29]. In addition to Beijing, the high rate of drug resistance is also associated with Haarlem lineages. This study reports the prevalence of Haarlem strain at a rate of 3.94% but none of these lineages contributed for the drug resistance genotypes. Interestingly, outside India the frequency of Beijing strains was more (56%) in TB meningitis patients which was reported by two consecutive studies from Bangkok and Thailand [30, 31].

In our study among the orphans, we found 22 unique patterns which were missing in SpolDB4, 9 M. tb Strains show features of EAI3 strains and 9 show similarity to EAI5 and three to Haarlem and one to S spoligotype. Previous reports by Desikan and his colleagues from Madhya Pradesh reported three EPTB isolates were found to be orphan [32]. In addition to this we also found very rare genotypes like LAM (2.63%), X2 (1.31%), U (1.31%) and S (1.31%) lineages in our study. This is in line with a study by Sankar et al. [3] who reports the rare isolates of LAM and X2 from extra pulmonary specimens. Furthermore, our study also identified Five T1 isolates from extra pulmonary specimens and for the first time one such report is also reported by Kandhakumari et al. [27] in India. In addition, drug susceptibility testing also found among six isolates tested; only one isolates belonging to the T1 lineage was found resistant to one of the first line drug rifampicin. We had four relapse in our study and genotyping results was available for two of them and both belong to EAII_SOM. Recurrent infections have been reported earlier by us in HIV infected TB patients by EAII_SOM M.tb strains [33]. Further studies need to carry out to explore association of EAII_SOM M.tb strains in recurrence.

Furthermore, exogenous reinfections were observed by many researchers in both immunocompetent and immunosuppressed individuals. Mostly, drug resistant strains and exogenous reinfections from drug sensitive organisms have been reported in so many lineages. Two patients in our study had a recurrent TB infection. Moreover, immunological characterization studies were performed between EAI and Non-EAI family in order to understand the differences in cytokine profiles which confers drug resistance to this particular lineage. Immune protection against tuberculosis are mediated by the induction of T cell responses specifically through type 1 and type 17 cytokines [34]. Besides, IL-1 family (IL-1α, IL-1β) and IL-18) and IL-12 cytokine are also necessary for protection against Mtb [35]. However, the relationship or the association of strain specific immune mediated responses in EPTB individuals has not been examined so far. Hence, for the first time we show there was significant immunological differences were observed between the individuals infected with EAI3 India and non-EAI strains. The major differences observed are for IL-2, IL-1α (reduced in EAI3 India strain) and IL-13 (increased in EAII3 strain). Decreased IL-2 levels in the EAII3 India strain infected individuals suggest that the former is associated with loss of immune protection and persistent chronic infection associated in EPTB individuals. The complete loss of IL-2 eventually leads to reduced immunity and expansion of Mtb growth and increased IL-2 provides protection along with substantial expansion of central memory T-cells [36, 37]. Like IL-2, IL-1 family cytokines are also equally important for immune protection and our data shows diminished levels of IL-α in EAII3 India strain infected individuals than the non-EAI. Previous studies also shown that deficiency in the IL-1 family cytokines are more susceptible to TB.
infection [35]. Finally, Type 2 cytokine (IL-13) were significantly enhanced in individuals infected with EAI3 India strain. Type 2 cytokines were harmful to host immune response to TB disease [34] and increased levels associated with chronic infection among the EAI3 India strain than non-EAI strain infected individuals. Therefore, individuals with EA13 infected strains might have suppressed host protective (Type 1) responses mediated by Type 2 cytokines. Apart from those, type 1 (IFNγ, TNFα), type 2 (IL-4, IL-5), type 17 (IL-17, IL-22), regulatory (IL-10) and pro-inflammatory (IL-6, IL-12 and GM-CSF) cytokines were not significantly different between the two strains (data were not shown).

In addition, we have also analyzed some of the major CC and CXC (CCL-1/1-309, CCL2/MCP-1, CCL3/MIP1α, CCL4/MIP-1β, and CCL11/eotaxin, CXCL-1/Gro-α, CXCL2/ Gro-β, CXCL9/MIG, CXCL10/IP-10 and CXCL11/ITAC 1) chemokines between the EA13 India and non-EAI strain infected individuals. Interestingly we found CCL3/MIP1α and CCL4/MIP-1β were higher in non-EAI strain infected individuals (Supplementary Fig. 1). Both chemokines have an important role in migration, homing, recruitment and stimulation of leukocytes to the sites of inflammation [38]. Hence the immune system fails to protect individuals infected with EA13 strains against EPTB infection. Further, we have also examined these cytokine and chemokine response to in drug sensitive (DS) versus Ofloxacin mono resistant (OMR) strains. Similar results were also observed between the two groups where increased Type 2 (IL-4 and IL-13) and Type 17 (IL-17 and IL-22) cytokines were observed in OMR strain infected individuals (Supplementary Fig. 1). It has been shown that Type 17 cytokine plays a crucial function in controlling Mtb through recruitment of immune cells, effective granuloma formation at early stages of infection. Meanwhile increased production might have the detrimental effects by promoting immunopathology [39]. Similar to EA13 and non-EAI groups, the other cytokines and chemokines do not possess any significant differences between the DS and OMR (data were not shown) strains infected individuals. These observations reinforce that cytokines or chemokines involved in the host protection are compromised or suppressed in the EA13 India strain or in OMR resistant strain infected individuals. Using only spoligotyping as the method for molecular epidemiology was limitation of our study, but we wanted to classify our strains into broader range, so that we can better study the immune response developed by the different spoligotypes. The other limitation is small sample number and drug resistance strains in EPTB samples analyzed.

**Conclusion**

The present study provides molecular epidemiological data of mycobacteria in lymph node samples isolated from extra-pulmonary patients from south India. The study confirms that EAI1 lineage to be the most predominant lineages in EPTB patients with lymphadenitis and were found to have increased type 1 and type 17 pro-inflammatory cytokine profiles. The presence of orphan and unique spoligotyping patterns in this study identifies the genetic diversity of M.tb strains in Chennai region and more molecular epidemiological studies are recommended in lymph node samples throughout India to elucidate their role in TB lymphadenitis.

**Supplementary information**

The online version contains supplementary material available at https://doi.org/10.1186/s12879-020-05597-0.

### Additional file 1: Supplementary Table 1.

**Demographic and clinical details of the study population**

### Additional file 2: Supplementary Fig. 1.

**Diminished plasma levels of type 2 and type 17 cytokines associated with individuals infected with drug sensitive strain.** The systemic levels of IL-4, IL-13, IL-17A and IL-22 were measured by ELISA between the drug sensitive (DS, n = 10) and orphan mono resistant (OMR, n = 3) strain infected individuals. The data were shown as bar graph and P values were calculated using the Mann–Whitney U test.

### Additional file 3: Supplementary fig. 2A.

**The Mtb spoligotypes distribution among male and female patients**

**The Mtb spoligotypes distribution among different age groups**

**Abbreviations**

Mtb: *Mycobacterium tuberculosis*; MTBC: *Mycobacterium tuberculosis* bacterial complex; EAI: East African Indian; CAS: Central Asian Strain; LAM: Latin America and Mediterranean; PGRS: Polymorphic GC-rich repetitive sequences; MIRU: Mycobacterial interspersed repetitive unit; SIT: Spoligotype international type; OMR: Ofloxacin mono resistant; DS: Drug sensitive; EPTB: Extra pulmonary tuberculosis; MIC: Minimum inhibitory concentration; IL: Interleukin; CCL: Chemokine (C-C motif) ligand; MCP: Monocyte chemoattractant protein; MMP: Macrophage inflammatory protein; CXCL: Chemokine (C-X-C motif) ligand; MIG: Monokine induced by gamma interferon; IP-10: IFNγ inducible protein 10; ITAC 1: Interferon-inducible T cell alpha chemoattractant 1

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**Authors’ contributions**

Conceived and designed the experiments: SS, YC, UDR. Performed the experiments: SS, YC, GRK. Analyzed the data: SS, YC, GRK. Contributed materials/reagents/analysis tools: SS, YC, GRK. Contributed writing: SS, YC, UDR. Wrote the paper: GRK, SS, YC, UDR. The authors have read and approved the final manuscript.

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**Availability of data and materials**

The data and materials supporting this article are included within the article.

**Ethics approval and consent to participate**

The Institutional Ethical Committee (IEC) of National Institute for Research in Tuberculosis (NIRT) approved the study and written informed consent was obtained from the patients before collecting the samples (NIRT IEC 2009 007).
Consent for publication
Not applicable.

Competing interests
None declared.

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