Association of adiponectin levels and insulin demand in critically ill patients

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Purpose: Intensive care unit patients usually have a deregulated glucose homeostasis and present with hyperglycemia and hyperinsulinemia, suggesting overall insulin resistance. Adiponectin has significant anti-inflammatory and insulin-sensitizing effects and is diminished in morbidly obese and in critically ill patients. Reduced adiponectin could contribute to insulin resistance in these patients. We examined how far insulin demand in critically ill patients is correlated with patient adiponectin levels.

Patients and methods: Adiponectin, resistin, leptin, insulin demand, minimal and maximal blood sugar levels, epinephrine, and hydrocortisone demand were measured 1 day after diagnosis of severe sepsis or septic shock in 25 patients (8 female, 17 male; median age 65 years; range: 31 to 87 years).

Results: Insulin demand (range: 0–8 IU/h; median 3.5 IU) was positively correlated with serum adiponectin levels (median: 10.1 µg/mL; range: 2.9–47.6 µg/mL; r = +0.56, P < 0.01). There was no significant correlation between insulin demand and leptin serum levels (median: 18.1 ng/mL; range: 0.3–80.7 ng/mL; r = +0.29, P = 0.08) or resistin serum levels (median: 103.9 ng/mL; range: 14.7–352.3 ng/mL; r = +0.13, P = 0.27). Epinephrine demand (median: 0.08 µg/kg*min; range: 0.02–0.63 µg/kg*min) was negatively correlated with male adiponectin levels (r = -0.58; P < 0.01; females: r = -0.36; P = 0.19) and positively correlated with resistin levels (r = 0.43; P = 0.02). Patient body mass index (median 26 kg/m²; range: 18–37) was positively correlated with serum leptin (r = +0.60; P < 0.01) but was not correlated with insulin demand (r = 0.19; P = 0.19), or adiponectin (females: r = -0.37, P = 0.18; males: r = -0.16, P = 0.27), or resistin levels (r = +0.17; P = 0.21).

Conclusion: Adiponectin levels and insulin demand were positively correlated during sepsis. Adiponectin levels were negatively correlated with epinephrine demand in male patients and epinephrine demand was positively correlated with resistin levels, which might have increased insulin resistance. The relationship between adiponectin and insulin action in humans is more complex than often suggested.

Keywords: adipokines, sepsis, diabetes, resistin, leptin

Introduction

Patients who are critically ill often require prolonged intensive care and vital organ function support. These patients uniformly have a deregulated glucose homeostasis and present with hyperglycemia and hyperinsulinemia, suggesting overall insulin resistance. Maintaining normoglycemia with insulin therapy improves survival and reduces morbidity in surgical intensive care unit patients.1,2

Hyperglycemia in the presence of hyperinsulinemia also accompanies type 2 diabetes (T2DM), obesity, and the metabolic syndrome. These conditions are characterized by
Reduced inhibition of hepatic gluconeogenesis and impaired glucose uptake in insulin-sensitive tissues, such as skeletal muscle.

These metabolic processes are influenced by products of the adipose tissue, so-called adipokines, which play a crucial role in the pathogenesis of the metabolic syndrome. Adipokine levels are changed in the morbidly obese and these changed levels contribute to skeletal muscle and hepatic insulin resistance.

Adiponectin, the most abundant adipokine exclusively derived from adipocytes with higher average levels in females compared with males, has excited intense interest because of robust correlation of its circulating levels with insulin resistance and risk of T2DM. Further, adiponectin has a significant anti-inflammatory and insulin-sensitizing effect. We have already demonstrated that adipokines like adiponectin or leptin are shifted in critically ill patients in the same direction as in morbidly obese and patients with T2DM. Adiponectin has an unusual inverse relationship with body mass index (BMI) and thus is diminished in the obese, but also in the critically ill. Since reduced adiponectin levels seem to contribute to insulin resistance in the obese, reduced adiponectin levels could also contribute to insulin resistance in critically ill patients.

Since higher adiponectin levels are associated with a lower risk of T2DM across diverse populations, we could speculate that higher adiponectin levels in critically ill patients cause a lower insulin demand.

Therefore, in this study, we determined adiponectin, resistin, and leptin levels in critically ill patients and analyzed the insulin demand in correlation to patients’ adipokine levels.

Materials and methods

Patient characteristics

Twenty-five patients who fulfilled the clinical criteria for severe sepsis or septic shock were enrolled in this study. Eight patients were female and 17 male, with a median age of 65 years (range: 31–87 years; female median age: 81 years, range: 42–87 years; male median age: 65 years, range: 31–83 years). The criteria for severe sepsis and septic shock were in accordance with those defined by Bone. The Simplified Acute Physiology Score (SAPS II) and Sequential Organ Failure Assessment score (SOFA) without Glasgow Coma Scale were used to define the severity of disease and organ dysfunctions, respectively. Total adiponectin, resistin, and leptin were measured 1 day after diagnosis of severe sepsis or septic shock. Blood samples were collected at 5 AM in the fasting state. Body weight and height for calculation of BMI of septic patients were self-reported (or estimated if no self-report was possible). Insulin was administered via perfusor syringe. International Units of insulin demand, minimal and maximal blood sugar levels, as well as epinephrine and hydrocortisone demand were determined within the first 24 hours after diagnosis of severe sepsis or septic shock.

For all measurements, 10 mL venous blood was collected in a chilled syringe with EDTA and immediately centrifuged at 1600 × g for 15 minutes. Plasma was stored at −80°C until analysis.

The study was performed with the permission of the independent local ethics committee of the University of Ulm (approval number 82/2002 and 73/2009). A written informed consent was obtained. If the patient was not capable of making decisions because of sedation or altered mental function, the informed consent was obtained after recovery as advised by the ethics committee. Exclusion criteria were: age <18 years, pregnancy, rapidly progressing underlying disease, HIV/AIDS, cardiogenic shock as the primary underlying disease, underlying hematologic disease, or cytotoxic therapy given within the previous 6 months.

Adipokine measurement and reagents

Multiplex analysis kits for leptin, resistin, and total adiponectin were obtained from Millipore, Hamburg, Germany. In brief, the appropriate adipokine standards and samples, diluted in plasma dilution buffer, were added to wells of a filtered plate. The samples were incubated with 50 µL of the antibody-coupled microsphere set on a plate shaker in the dark at room temperature for 30 minutes. The samples were washed 3 times with 100 µL wash buffer. Freshly diluted secondary detection antibody (25 µL/well) was added to the wells and then incubated on a plate shaker in the dark at room temperature. Thereafter, samples were washed 3 times with 100 µL wash buffer. Fifty microliters of strepavidin-PE (16 µg/mL in assay buffer) was added to each well, and incubation continued on a plate shaker at room temperature for the first 10 minutes. Unbound analytes were filtered through the wells using the vacuum manifold. The bound beads were washed 3 times with 100 µL wash buffer. After the last wash step, 125 µL of assay buffer was added to each well and the plate was placed on a plate shaker set at 500 rpm for 1 minute and then for 3 minutes at a reduced speed of 300 rpm. Finally, samples were analyzed on the Luminex system in accordance with the manufacturers’ instructions. The system was sufficiently sensitive to analyze leptin values as low as 0.3 ng/mL.

Statistical analysis

All values were expressed as median with range. The Mann-Whitney U test was used to compare the data of septic patients. Analysis was performed with WinSTAT, Version 2009.1 (R.F. FitchSoftware).
The Spearman rank-order correlation coefficient was calculated for correlation analysis. $R$ indicates the correlation coefficient. A value of $P < 0.05$ was considered statistically significant. Correlation coefficient values between 0.3 and 0.7 ($-0.3$ and $-0.7$) indicate a moderate linear relationship, and values between 0.7 and 1.0 ($-0.7$ and $-1.0$) indicate a strong positive (negative) linear relationship. No adjustment for multiple testing has been made.

**Results**

The 25 septic patients (17 male and 8 female patients) had median SAPS II and SOFA scores of 37 (range: 21–58) and 9 (3–11), respectively. Median BMI was 26 kg/m² (range: 18–37). The underlying cause of sepsis with side diagnosis, gender, age, CRP and leukocyte levels, BMI, SAPS II (without Glasgow Coma Scale), and sepsis score according to the definition of Bone et al$^{10}$ are shown in Table 1.

### Table 1: Gender, age, underlying reason of sepsis, side diagnosis, C-reactive protein levels, white blood count, self-reported (or estimated) body mass index, and Simplified Acute Physiology Score (SAPS II) without Glasgow Coma Scale of the 25 septic patients

| Age/gender | Underlying cause of sepsis | CRP (mg/L) | White bloodcount (G/l) | BMI | Side diagnosis | SAPS II (without GCS) | SOFA (without GCS) | Sepsis score* |
|------------|---------------------------|-----------|-----------------------|-----|----------------|------------------------|---------------------|---------------|
| 62/m       | Acute necrotizing pancreatitis | 85        | 12.7                  | 27  | Spastic hemiparesis, nicotine and AA/D | 49         | 10          | 6             |
| 83/m       | Perforated gastric ulcer pancreatic fistula | 334       | 44.8                  | 25  | NIDDM, aHT, TAA | 50         | 10          | 6             |
| 67/m       | Postoperative pancreatic fistula | 259       | 46.2                  | 29  | NIDDM, aHT     | 21         | 7           | 6             |
| 42/f       | Necrotizing fasciitis        | 323       | 17.5                  | 28  | –              | 58         | 9           | 7             |
| 65/m       | Necrotizing fasciitis        | 283       | 24.3                  | 25  | DIDDM, aHT, TAA, RI | 29         | 11          | 6             |
| 66/m       | Ischemic colitis             | 289       | 18.2                  | 26  | DIDDM, aHT; RI, PAOD | 34         | 9           | 6             |
| 63/m       | Postoperative bile peritonitis | 266       | 12.9                  | 37  | CHD, adipositas, NIDDM, aHT | 31         | 9           | 6             |
| 64/m       | Postoperative paralytic ileus | 250       | 16.2                  | 30  | –              | 24         | 10          | 6             |
| 86/f       | Perforated duodenal ulcers   | 273       | 37.7                  | 31  | aHT, dementia  | 31         | 5           | 4             |
| 50/m       | Esophageal perforation       | 138       | 15.5                  | 26  | Nicotine and AA/D; liver cirrhosis | 40         | 11          | 6             |
| 69/m       | Acute necrotizing pancreatitis | 219       | 14.7                  | 23  | Nicotine and AA/D | 24         | 8           | 6             |
| 44/m       | Blunt chest trauma           | 135       | 9.7                   | 32  | –              | 29         | 9           | 6             |
| 67/m       | Colon anastomosis insufficiency | 319       | 28.9                  | 21  | Ulcerative colitis | 47         | 11          | 6             |
| 65/m       | Arterial bypass infection    | 259       | 17.5                  | 26  | PAOD, AA/D, COPD, HF | 37         | 9           | 6             |
| 58/f       | Gastric ulcers perforation   | 259       | 16.9                  | 32  | Nicotine and AA/D, liver cirrhosis, aHT, asthma | 53         | 10          | 6             |
| 31/m       | Pleural empyema              | 330       | 17.5                  | 28  | –              | 29         | 7           | 6             |
| 64/m       | Colon anastomosis insufficiency | 102       | 21.6                  | 23  | –              | 46         | 11          | 6             |
| 66/m       | Gastric ulcers perforation   | 200       | 26.7                  | 24  | CHD, PAOD, COPD, aHT, TAA, HF, PAOD, AA/D, COPD, HF | 48         | 9           | 4             |
| 87/f       | Gastric ulcers perforation   | 213       | 16.3                  | 22  | NIDDM, aHT     | 41         | 5           | 4             |
| 80/f       | Anaphylactic shock           | 134       | 14.3                  | 25  | NIDDM, aHT     | 54         | 5           | 5             |
| 81/f       | Postoperative abdominal infection | 274       | 37.3                  | 24  | COPD           | 46         | 6           | 6             |
| 87/f       | Aspiration pneumonia         | 235       | 18.9                  | 28  | COPD           | 54         | 9           | 6             |
| 50/f       | Retroperitoneal abscess      | 178       | 18.8                  | 26  | RI, aHT        | 31         | 3           | 4             |
| 44/m       | Necrotizing fasciitis        | 130       | 15.1                  | 31  | Nicotine and AA/D, RI | 28         | 6           | 6             |
| 80/m       | Empyema                     | 225       | 17.6                  | 18  | COPD, nicotine abuse, RI | 31         | 9           | 4             |

*Note: Sepsis score (Bone et al$^{10}$): 4, sepsis, 5, severe sepsis, 6, septic shock.

**Abbreviations:** AA/D, alcohol abuse/dependence; aHT, arterial hypertension; CHD, coronary heart disease; COPD, chronic obstructive pulmonary disease; CRP, C-reactive protein; GCS, Glasgow Coma Scale; G/l, Giga/liter; HF, heart failure; NIDDM, noninsulin dependent diabetes mellitus; PAOD, peripheral artery occlusive disease; RI, renal insufficiency; TAA, tachyarrhythmia absoluta.
Median resistin level was 103.9 ng/mL (range: 14.7–352.3 ng/mL), median leptin level was 18.1 ng/mL (range: 0.3–80.7 ng/mL). Median gender specific adiponectin serum levels were 14.1 µg/mL for female (range: 5.3–47.6 µg/mL) and 9.6 µg/mL (range: 2.9–32.6 µg/mL) for male patients. The 2 extreme values of adiponectin (47.6 µg/mL in female patients and 32.6 µg/mL in male patients) represent the 2 patients with the lowest BMI (22 kg/m² and 18 kg/m²) and the lowest leptin levels (0.9 ng/mL and 0.3 ng/mL, respectively), although BMI showed no significant negative correlation with adiponectin levels (females: r = -0.37, P = 0.18; males: r = -0.16, P = 0.27).

Minimal blood sugar levels ranged between 63 mg/dL and 119 mg/dL (median 91 mg/dL), and maximal levels ranged from 122 to 398 mg/dL (median 193 mg/dL).

Insulin demand ranged from 0 to 8 IU/h (median 3.5 IU/h). When sepsis was diagnosed, 23 out of 25 patients required insulin; 15 out of 25 patients required hydrocortisone (preponderant 8 mg/h; range: 0–10 mg/h), and all septic patients required epinephrine (median: 0.08 µg/kg*min; range: 0.02–0.63 µg/kg*min). There was no gender difference in resistin and leptin levels, insulin, hydrocortison, and adrenalin demand or in minimal and maximal blood sugar levels.

There was a positive correlation of insulin demand after diagnosis of sepsis and serum adiponectin levels (all patients: r = +0.57, P < 0.01; female patients: r = +0.61, P = 0.05; Figure 1A; male patients: r = +0.53, P = 0.01; Figure 1B). Multivariate regression analyses with adjustment for age and BMI showed a largely unchanged significant positive correlation between insulin demand after diagnosis of sepsis and serum adiponectin levels (P = 0.02; 95% confidence interval: 0.02–0.2). There was no significant correlation between insulin demand and leptin (r = +0.29, P = 0.08) or resistin (r = +0.13, P = 0.27; Table 2) serum levels.

Epinephrine demand was negatively correlated with male adiponectin levels (r = -0.58; P < 0.01; female: r = -0.36; P = 0.19) and positively correlated with resistin levels (r = 0.43; P = 0.02; Table 2). There was no correlation between epinephrine and insulin (r = 0.12; P = 0.28) and leptin (r = 0.24; P = 0.13).

Patients who received hydrocortisone (n = 15) had mean insulin demand of 5 IU/h (range: 1.0–8.0 IU/h), while patients who received no hydrocortisone (n = 10) had mean insulin demand of 2 IU/h (range: 0.0–5.0 IU/h; P = 0.01). The mean adiponectin levels were not significantly different regardless of whether patients received hydrocortisone (10.3 µg/mL vs 10.0 µg/mL; P = 0.20). Furthermore, groups showed no difference in SAPS II and SOFA scores (P = 0.46; P = 0.41).

There was no correlation between patient age and SAPS II and SOFA scores, insulin, hydrocortisone, and adrenalin demand, or any analyzed serum parameter (adiponectin, resistin, and leptin). Five patients had pre-existing T2DM. These 5 patients showed no difference in any analyzed parameter.

Patient BMI was positively correlated with serum leptin (r = 0.60; P < 0.01) but not with insulin demand (r = 0.19; P = 0.19), or adiponectin (female: r = -0.37, P = 0.18; male: r = -0.16, P = 0.27), or resistin (r = +0.17; P = 0.21) levels.

### Discussion

Adipose tissue is essentially involved in complex body hormone and cytokine production and cannot stand by passively in septic conditions. In septic conditions, adipokines are shifted in the same direction as in obese subjects and patients with T2DM, which implies lowered serum levels of adiponectin. Recent research indicated that adiponectin has anti-inflammatory, anti-atherogenic,
antidiabetic properties. The ability of adiponectin to reduce insulin resistance in conjunction with its anti-inflammatory properties makes adiponectin a potentially promising therapeutic target, and agents that enhance adiponectin secretion or action have potential to treat metabolic disease.14

Because lowered adiponectin levels are associated with insulin resistance,15 we hypothesized that patients with lower adiponectin serum levels in septic conditions have a higher insulin demand. Surprisingly, however, insulin demand showed a positive correlation to serum adiponectin levels – especially since direct testing of the effect of insulin infusion in healthy subjects has shown that insulin suppresses adiponectin levels in vivo.16

A possible rationale for this unexpected correlation may be that adiponectin levels are more influenced by the septic condition than adiponectin levels influence insulin demand, particularly since adiponectin levels correlate negatively with SAPS II and SOFA scores.7

Furthermore, insulin secretion is influenced by multiple hormones. Besides adipose tissue-derived hormones, a variety of other hormones has been discovered in recent years. Possible candidate hormones that can enhance insulin secretion are the so-called incretins, ie, gut peptide hormones, which are secreted in response to a meal.17 Referring to insulin demand, these hormones could be more involved compared with adiponectin through their insulin-secreting effect, although incretins seem to be responsible for higher insulin release after an oral glucose load compared with an intravenous glucose load.18

In addition to incretins, there are also a number of other hormones like ghrelin or peptide YY. Ghrelin has been shown to inhibit insulin secretion in humans19 and ghrelin levels seem to be elevated in fasting conditions.20 Similarly, recent studies have suggested more direct effects of peptide YY on insulin sensitivity,21 but the role of peptide YY independent of food intake still needs to be confirmed. Since all septic patients enrolled in this study were fed exclusively parenterally, the insulin-secreting stimulus of incretins and other gut hormones was expected to be reduced.22 No data exist to show the extent to which septic conditions influence gut hormone release.

It is difficult to say how patient insulin demand in this study was influenced by hydrocortisone supply. Insulin resistance in critically ill patients and patients with systemic inflammatory response syndrome was classically attributed to increased serum levels of cortisol, which promotes gluconeogenesis and inhibits peripheral glucose disposal in a stress-dependent manner.23 Ten out of 25 patients received low-dose hydrocortisone (8–10 mg/h). Notably, mean adiponectin levels of patients receiving hydrocortisone were almost identical with those of patients receiving no hydrocortisone (10.3 µg/mL vs 10.0 µg/mL). Also, both groups had similar SAPS II and SOFA scores.7

Resistin, a protein with proinflammatory properties secreted by adipocytes and monocytes, has recently been reported to participate in the inflammatory response.24 Secretion of resistin is stimulated by insulin and inflammatory processes, glucocorticoids, and lipopolysaccharides, whereas tumor necrosis factor α and β-adrenergic stimulation act as inhibitors. Resistin levels are generally more elevated in obese patients and those with T2DM.25 Elevated resistin levels also seem to play a crucial role in insulin resistance.26 Resistin levels were positively correlated with SAPS II score (r = 0.39, P = 0.03) and epinephrine demand (r = 0.43; P = 0.02) supporting participation of resistin in the inflammatory response process. On the other hand, gender specific adiponectin, a protein with anti-inflammatory or
Fritsche L, Weigert C, Häring HU, Lehmann R. How insulin receptor

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