Inflammation, Metalloproteinases, Chronic Venous Disease and Sulodexide

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Abstract

Inflammation and extracellular Matrix Metallo Proteinases (MMPs) have been recently considered as an important step in the pathogenesis of Chronic Venous Disease (CVD). To understand how inflammation and MMP may affect venous tissue, a review on these items and on the pathogenesis of CVD was performed. Prolonged or repeated venous hypertension due to well-known predisposing factors causes abnormal shear stress leading to shedding of endothelial glycocalyx and activation of endothelial cells. The latter expose adhesion molecules and release pro-inflammatory cytokines. Diapedesis follows, leading to leukocytes infiltration of Extra Cellular Matrix (ECM) in the sub endothelial space of vein walls and valves. Activated leukocytes, mainly monocytes-macrophages, release chemotactic cytokines that amplify the inflammatory response, they also produce Nitric Oxide molecules (NO) and proteases, including MMPs. High concentration of MMPs, especially MMP-9, is found in venous wall with CVD and in venous ulcers. The role of MMP-9 in CVD is also supported by experimental data. MMP-9 can degrade components of ECM as collagen, elastin, fibrinectin and laminin. MMP-9 and other proteolytic enzymes may disrupt ECM structure, damaging and debilitating venous wall that lead to varicose veins and venous ulcers. Sulodexide, a glycosaminoglycan, counteracts several of these inflammatory changes. In subjects with CVD administration of sulodexide improves venous hemodynamic changes and CVD symptoms and accelerates venous leg ulcers healing.

Keywords: Varicose veins; Inflammation; Metalloproteinases; MMP; Venous leg ulcers; Sulodexide; Chronic venous disease; Venous insufficiency.

Abbreviations: CGRP: Calcitonin Gene Related Protein; COX: Cyclooxygenase; CVD: Chronic Venous Disease; C2,C3,C4: Clinical stages 2, 3 and 4 of CEAP Classification for CVD; ECM: Extra Cellular Matrix; EGF: Epidermal Growth Factor; FG: Fibroblast Growth Factor; G-CSF: Granulocyte Colony Stimulating Factor; GM-CSF: Granulocyte-Monocyte Stimulating Factor; HSP: Heat Shock Protein; ICAM: Inter Cellular Adhesion Molecule; IL: Inter Leukin; LDL: Low Density Lipoprotein; LPS: Lipo Poly Saccharide; MCP-1: Monocyte Chemotactic Protein-1; MIP-1β: Macrophage Inflammatory Protein-1 beta; NFKB: Nuclear Factor kappa B; NO: Nitric Oxide; PDGF: Platelet Derived Growth Factor; PGE2: Prosta Glandin E2; ROS: Reactive Oxygen Species; SEM: Scanning Electron Microscopy; TGFβ: Transforming Growth Factor beta; TIMP: Tissue Inhibitor of Metallo Proteinase; TNFa, TNFβ: Tumor Necrosis Factor alfa, beta; VCA: Vascular Cell Adhesion Molecule; VEGF: Vascular Endothelial Growth Factor.

Introduction

In recent years, multiple evidences have emerged that strongly support the involvement of inflammation and extracellular Matrix Metallo Proteinases (MMP) production as a fundamental step in the pathophysiology of Chronic Venous Disease (CVD), in both its appearance and its progression to the most advanced stages, i.e., venous leg ulcers. However, the inflammatory process, from its origin to the production of MMP and the consequent deterioration of venous walls and their valves which leads to CVD, may be complicated and it frequently requires an integral explanation. The purpose of this review is to organize these concepts and facilitate the knowledge on the participation of the inflammatory process in the pathophysiology of CVD.

Inflammation General Concepts

Inflammation is a physiological response of the body to aggression. It can be caused by biological agents (e.g., an infection), chemical agents (e.g., oxidative substances such as oxygen reactive species), physical agents (e.g., heat) or mechanical agents (e.g., trauma or vessel shear stress). The inflammatory phenomenon is essential for survival, since it tends to protect the organ and repair the damage, ultimately its objective is the return to normal conditions. Aggression can be of exogenous or endogenous origin. Sometimes, the inflammatory response may cause damage to tissues and organs, as occurs in autoimmune diseases.

The main signs of inflammation were described by Celsus (Aulus Cornelius Celsus; ca.25 b. C. - 50 a. C.): heat, redness, tumor (volume increase) and pain; 200 years later, Galen of Pergamon added function loss or alteration (functionaela), which was confirmed by Virchow. The common substrate of inflammation is the leukocytic infiltrate which is nearly always preceded by vasodilation with an increase in vascular permeability that causes fluid exudation (edema).

Inflammation is a complex defense mechanism in which leukocytes migrate from inside the vessels towards the damaged tissue to destroy the agent causing tissue damage. Acute inflammation is a limited beneficial response, particularly during an infection, while chronic inflammation is a persistent phenomenon that may lead to tissue damage. One characteristic of acute inflammation is that initially, the leukocyte infiltrate consists of neutrophils, but after 24 to 48 hours monocytes predominate. In contrast, chronic inflammation is
associated with mononuclear cell infiltrate: monocytes-macrophages and lymphocytes. The leukocyte infiltrate is the main characteristic that allows the pathologist to recognize the presence of inflammation under the microscope [1,2].

**How is vasodilation produced?**

A noxious agent stimulates certain cells: mast cells, endothelial cells and macrophages, as well as free nervous endings that release different chemical mediators, which dilate the arterioles and venules and increase fluid and protein leakage. Mast cells thus activated produce histamine, eicosanoids (thromboxanes, prostaglandins, leukotrienes), tryptases and C-type natriuretic peptide (CGRP) (Calcitonin Gene Related Product), which cause arteriolar dilation. Endothelial cells produce Nitric Oxide (NO), PGI2 and convert kallikrein into bradykinin, which are substances that also cause arteriolar dilation. Activated macrophages cause arteriolar vasodilation and increased vascular permeability by the action of Cyclooxygenase (COX), which converts arachidonic acid into PGE2, they also produce NO. Free nervous endings release P substance and neurokinins, which stimulate mast cells and cause venular vasodilation and permeability increase. The greater vascular permeability enables the presence of a water and protein exudate and facilitates the leukocytic infiltrate [3].

**How is the leukocytic infiltrate produced?**

The noxious agent that prompts any cell alteration causes damaged cells of any kind to release heat shock proteins or HSP that act on tissue macrophages, smooth muscle cells and endothelial cells. Thus activated endothelial cells express adhesion molecules on their surface: ICAM – Inter Cellular Adhesion Molecule, VCAM - Vascular Cell Adhesion Molecule, E and P selectins. The three types of cells produce cytokines, especially Interleukins (IL) which regulate and amplify inflammation [3].

Cytokines are soluble proteins released by some cells and they act as a chemical messenger between the cells involved in immunity and inflammation. IL among them, produced mainly by different types of leukocytes, tumoral necrosis factor, interferons and various growth factors [4]. Endothelial cells and smooth muscle cells produce IL-6, macrophages produce IL-1, IL-6, IL-12 and IL-15 which modulate inflammation and the immune response, macrophages also produce TNF-α (Tumor Necrosis Factor alpha) and nitric oxide which is a vasodilator. IL-1α and TNF-α contribute to the expression of adhesion molecules in endothelial cells. IL-6 is pleiotropic, it has various pro-inflammatory actions, among which change from a neutrophil infiltrate to a monocyte-macrophage infiltrate in the extracellular matrix stands out, it participates in the regulation of the immune response and in the production of acute phase reactants [5,6]. The exposure of adhesion molecules on the surface of endothelial cells and also on leukocyte’s surface (L-selectin, integrins) causes these to circulate more slowly and finally adhere to endothelial cells, to later penetrate into the intercellular spaces until reaching the extracellular matrix (diapedesis).

The participation of the endothelial glycoalkal in inflammation is essential. An intact glycoalkal rejects leukocytes and prevents these from approaching adhesion molecules. Mechanical tensile, frictional or shear stresses on vascular walls, as well as hyperglycemia, oxidized LDL, some enzymes and tumor necrosis factor TNF-α may cause sufficient glycoalkal thinning for small adhesion molecules to become exposed to leukocytes, thus, these may adhere and migrate towards the ECM beyond the endothelium [7,8]. Activated leukocytes produce IL and chemokines, which attract more leukocytes and increase the inflammatory infiltrate in the extracellular matrix. Chemokines are small molecules (8-12 kD), there are around 50, they induce chemotaxis and they recruit leukocytes; at some point they also recruit fibroblasts that initiate lesion healing or tissue remodeling [4].

**Inflammation as a cause of extracellular matrix damage**

Besides production of different cytokines, both endothelial cells and activated leukocytes produce Reactive Oxygen Species (ROS) that may oxidize different exogenous substances (for example, phagocytized bacteria) but also components of the body itself. Leukocytes also produce different enzymes that break protein peptide bonds (proteinases), such as elastase, cathepsin G, proteases and extracellular matrix metalloproteinases. Elastase hydrolyzes elastin and fibronectin, it also detaches a leukocyte coating, thus exposing integrins, facilitating their adhesion and transmigration towards the ECM, which induces the production of ROS and more proteinases. Cathepsin G is produced by neutrophils and it is important for the digestion of phagocytized material. Proteases digest proteins and peptides of different kinds. MMPs are worthy of being considered separately. Proteinases and ROS may damage the extracellular matrix structure. Extracellular proteolysis appears to be a prerequisite for normal wound and injured tissue healing, but excessive or uncontrolled tissue destruction may be a pathogenic factor leading to tissue damage and remodeling [3]. The main aspects of inflammation development are summarized in (Figure 1).

**Extracellular Matrix Metalloproteinases (MMPs)**

Metalloproteinases are produced by several cells, especially leukocytes. Their physiological function is in the turnover, regulation and composition of the extracellular matrix components, therefore, they are present in all tissues. They are designated as metalloproteinases due to the presence of zinc at the active site.

There are more than 20 MMP, but they can be divided into 5 groups according to their structure and specificity [9].

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**Figure 1:** General diagram of inflammation. The two main histologic features are leukocyte infiltration and vasodilation. The first initiate with heat shock proteins inducing leukocyte adhesion and diapedesis. Leukocyte infiltration is enhanced by chemokines produced mainly by macrophages, leukocytes release MMPs and other proteinases, as well as reactive oxygen species, leading to the destruction of noxious agent. Vasodilation is produced through several chemical mediators such as histamine, eicosanoids, bradykinin, PGE2, nitric oxide released by endothelial cells, mast cells, macrophages and nerve endings. Vasodilation and increased vascular permeability causes edema.
• Collagenases, which degrade type I, II, III and X fibrillar collagen (MMP-1, 8 and 13).
• Gelatinases, which degrade denatured collagen, laminin, fibronectin, elastin and basement membrane type IV collagen (MMP 2 and 9).
• Stromelysins, which degrade proteoglycans, laminins, fibronectin and some types of collagen (MMP-3,10,11 and 12).
• Matrylsin, which degrade type IV collagen and proteoglycans (MMP-7).
• Membrane metalloproteinases, which degrade several components of ECM (MMP-14 to 17).

They are secreted aszymogens and must be activated (by plasmin, ROS and other enzymes) to exert their proteolytic action. They are inhibited by TIMP (Tissue Inhibitors of Metallo Proteinase). MMP production is kept in balance with its tissue inhibitors, but the balance is broken in the presence of inflammation (and of additional activated monocytes-macrophages). Pro-inflammatory cytokines: IL-1, IL-6, TGF-β (transforming growth factor β), TNF-a, EGF (epidermal growth factor), PDGF (platelet-derived growth factor) increase their synthesis, while corticosteroids, heparin and IL-4 slow down their synthesis [10].

MMPs have been considered important in vascular diseases for the following reasons:

a) A marked increase in MMP-9 plasma concentration was found in blood drawn from varicose veins after 30 minutes of stasis (in orthostatic posture) [11].

b) Greater amounts of different MMP with variations in distribution have been observed in varicose vein walls, compared to normal veins. MMP-9 has been found at higher concentrations in the middle layer of varicose veins [12,13].

c) In rat vena cava rings subjected to tension, it was demonstrated, by means of immune-histochemical staining, that the rise of tension markedly increases the presence of MMP-2 and MMP-9 in the middle layer. MMP-2 and MMP-9 concentration was higher with greater tensile strength. The greater the tensile strength (2 g vs. 0.5 g) and its duration (24 hours vs. 1 hour), the higher the MMP-2 and MMP-9 concentration (p<0.01). Moreover, MMPs increase coincides with a reduction in the contraction strength of the venous wall induced by phenylephrine, angiotensin II and potassium chloride, an effect prevented with TIMP, thus demonstrating that contractility reduction is totally or partially due to MMP increase. The direct effect of MMP-2 and MMP-9 on venous contraction capacity was proved by the incubation of vena cava segments with these MMPs, contractility is lower as incubation time elapses, this effect is also counteracted with TIMP [14].

d) The importance of MMPs in venous ulcers is supported by various studies that demonstrate the increase of MMP in the ulcerated tissue, mainly in chronic ulcers or ulcers of torpid evolution. MMPs concentration is higher in chronic ulcers than in acute ulcers [15,16].

e) Specifically, MMP-9 is found at higher concentrations in fluid from chronic ulcers than in fluid from acute ulcers. The high concentration of MMP-9 in ulcers is associated to an unfavorable evolution and it is reduced in ulcers during compressive treatment or in ulcers that are in healing process [17-20].

f) In diabetic foot ulcers, the initial MMP-9 concentration is also associated with the outcome: those with low MMP-9 concentrations heal rapidly, while those with high concentrations have a more chronic evolution [21].

g) Administration of MMP-9 to experimental skin ulcers in mice delays healing [22].

Pathophysiology of Chronic Venous Disease

There are two main groups of causative or predisposing factors in the appearance of CVD: genetic and environmental factors. Besides some genetic syndromes accompanied by venous disorders, which are sometimes very marked, the influence of inheritance is certain in “common” CVD cases and it has been demonstrated in multiple observations. If both parents have varices, the risk of developing CVD is very high (90%), while if one parent is affected the risk is 25% for men and 62% for women [23]. Maternal influence is greater than paternal influence, with the former involving a risk of around 50% and the latter a 25% risk [24]. In Mexico, it was observed that more than 60% of 1,013 patients with CVD had history of maternal varicose veins [25]. The mechanism of genetic predisposition is not clear enough, but it is likely to be related to venous wall or valve weakness, or both. Other predisposing factors, such as obesity, multiparity, prolonged standing, previous venous thrombosis, etc. [24-26] have a clear haemodynamical influence, since they delay blood flow return and they chronically elevate hydrostatic venous pressure in a repeated and prolonged way.

Venous hypertension increases mechanical tensile, frictional or shear stresses which act on the endothelial glycocalyx and endothelial cells. Glycocalyx becomes thinner and endothelial cells are activated [27]. It has been demonstrated that the increase in venous pressure causes an increase of endothelial adhesion molecules [28]. A recent review of the influence of blood flow alterations and shear stress on the endothelium reveals that these induce the expression of pro-oxidant, pro-inflammatory, pro-coagulant and pro-apoptotic genes in endothelial cells, as well as the expression of adhesion molecules, leukocyte adhesion and diapedesis and the increase of metalloproteinases and ROS [29].

Examination of the endothelial surface of surgical samples of saphenous veins with CVD by SEM (scanning electron microscopy), compared with control saphenous veins without venous disease show an irregular orientation, discontinued areas of the endothelium, plates and erythrocytes adhered to the surface and the presence of “microvilli” on endothelial cell surface. The abnormalities were of greater severity in more severe degrees (C4>C3>C2). In the same study, endothelial cells from veins (saphenous veins and tertiary branches R3) from healthy controls and patients with CVD show clear differences between the groups: the expression of surface molecules, adhesion molecules (ICAM-1), the proliferation, transcriptional activity (NFKB), pro-inflammatory cytokine release and VEGF capacity are greater in CVD grade C3 than in grade C2, and are greater in these than in control individuals. In short, during CVD, endothelial cells show a pro-inflammatory phenotype whose intensity depends on the degree of CVD [30].

The degradation of the glycocalyx and the inflammatory activation of the endothelium enable diapedesis and leukocyte infiltration of the vein walls. In this respect, more than 15 years ago, with the use of monoclonal antibodies, Ono et al. identified the presence of monocye/macrophage infiltrate in the venous walls and valves in all the ten saphenous veins of patients with venous disease and in no saphenous vein of four individuals without venous disease [31].
The inflammatory infiltrate originates the release of proteinases (mainly MMP) and other enzymes that degrade the ECM. Moreover, a pro-oxidative state has been demonstrated in CVD, since an increase of oxidative substances and a reduction in antioxidants has been found in plasma from patients with CVD compared to healthy volunteers [32]. The role of oxidative stress in venous wall damage is not clear, but it is probably a contributing factor.

The degradation of the extracellular matrix weakens the venous wall and allows vein dilation and valve impairment, which in turn increases reflux and worsens venous hypertension. An imbalance of the protein content of ECM of varicose vein walls has been proven, which appears even before the presence of valves insufficiency. Although inflammation also occurs in venous valves, deforming them or even damaging them until disappearance, some evidence suggests that venous dilation appears first, followed by valves incompetence [33,34]. Valves dysfunction worsens venous hypertension, and the pathophysiological circle increases.

The same changes that affect glycocalyx and endothelial cells occur in venules and capillaries, which explains edema, skin disturbances and venous ulcers. The increase of water permeability causes edema, whose consistency increases due to the passage of proteins, erythrocyte filtration causes oedema, debridement towards the skin and the subcutaneous tissue favors inflammation which, together with local ischemia due to alterations of capillary flow, lead to the emergence of skin ulcers [27]. The involvement of inflammation in venous disease and microcirculation in the pathophysiology of lipodermatosclerosis and skin ulceration has been recognized since many years ago and it has recently been reviewed incorporating the current concepts [35,36].

In venous ulcers, a high concentration of multiple pro-inflammatory cytokines, such as various interleukins, granulocyte and granulocyte-monocyte colony stimulating factors, monocyte chemotactic protein-1, interferons γ and α and other has been demonstrated; many of these decrease after 4 weeks of treatment. High concentrations of anti-inflammatory cytokines are also found in ulcers that later heal faster [37]. This inflammatory activity is consistent with high MMP-9 concentrations, especially in ulcers with unfavorable evolution and its reduction with treatment and healing, as previously mentioned. In keeping with the foregoing, the proteomic analysis of exudate collected from rapidly healing ulcers shows proteins that take part in tissue formation, while that of non-rapidly healing ulcers rather show inflammatory and tissue destruction proteins, such as MMP-9, elastase and protease 3 [20].

Inflammation and microcirculation in the pathophysiology of chronic venous disease, including skin disturbances, is summarized in (Figures 2 and 3).

**Pleiotropic anti-Inflammatory Therapeutics in CVD**

Although there are many anti-inflammatory drugs, most of them are cyclooxigenase inhibitors and they have not been shown to have favorable actions in CVD. In contrast, the glycosaminoglycan sulodexide has shown utility in CVD, since it has different actions in the pathophysiology of CVD [38-40] as follows:

- It restores impaired glycocalyx volume. For example: the glycocalyx dimension observed in the sublingual vessels of individuals with type 2 diabetes mellitus is lower than in healthy controls. After two months of treatment with sulodexide administered by oral route, the dimensions of the glycocalyx increased reaching normal values [41].
- It suppresses the inflammatory-type response of endothelial cells. Cultured endothelial cells subjected to high glucose concentrations or to ageing changes release free radicals (reactive oxygen species), monocyte chemotactic protein-1 and interleukin 6 and reduce their multiplication rate, measured as the time of recovery from a wound. These alterations are prevented when sulodexide is added to the culture medium [42,43].
- It reduces the release of interleukins, chemokines, and colony-stimulating factors by macrophages. In an *in vitro* experiment in cell culture, lipopolysaccharide (LPS)-stimulated macrophages significantly increase the production of cytokines, chemokines and colony-stimulating factors, when sulodexide is added at two different
concentrations to previously stimulated macrophages a dose-dependent inhibition of the release of various interleukins is observed: IL-1β, IL-12 and TNF-alpha, which are proinflammatory, IL-7 and IL-17, which are immune-modulatory, several cytokines with chemotactic activity, such as interleukin 8, monocyte chemotactic protein-1 (MCP-1) and macrophage inflammatory protein β (MIP 1β), as well as granulocyte colony-stimulating factor (G-CSF) and granulocyte-monocyte colony-stimulating factor (GM-CSF). In the same experiment, a dose-independent inhibition of other interleukins was observed: IL-2, IL-6, IL-10, IL-13 and interferon β [44].

- It reduces MMP-9 secretion by macrophages. The incubation of blood from healthy volunteers with increasing sulodexide concentrations caused a dose-dependent decrease in serum MMP-9 concentration. Incubation of serum alone (therefore, without blood cells) with sulodexide had no effect, thus demonstrating that its action is not on the existing MMP concentration, but on its release by white cells. The incubation of cultured myelo-monocytic cells also demonstrated that sulodexide causes a reduction of MMP-9 release [45].

The anti-inflammatory actions of sulodexide, most of them compiled by Mattana et al. [46], are summarized in (Table 1).

Different phlebodynamic measurements suggest that treatment with sulodexide improves venous wall conditions: it reduces vein distensibility, increases venous tone, reduces maximum incremental volume upon exposure to an external obstruction with 40 or 60 mmHg, compared to placebo, it reduces posterior tibial vein pressure compared to phlebotonics and reduces capillary filtration coefficient (Figure 3) [47-49].

The clinical translation of anti-inflammatory and phlebodynamic properties manifests as an improvement of the signs and symptoms of chronic venous disease at its different stages, including chronic venous leg ulcers (Figure 4) [25,50-53]. As can be seen in (Figure 5), sulodexide acts in the different stages of inflammation that lead to venous disease.

**Conclusions**

The long-term increase of blood pressure in the veins and the consequent alteration of shear stress cause endothelial glycocalyx shedding and activate endothelial cells, allowing diapedesis and inflammation of the extracellular matrix, production of Matrix Metallo Proteinases (MMP) and other proteinases, which disrupts the matrix and weakens venous walls and valves. This leads to varicose veins, greater venous hypertension, inflammatory alterations of the skin and eventually venous ulcers. Sulodexide counteracts several of the foregoing pathogenic steps, improves vein function and symptoms of CVD and accelerates healing of venous ulcers.

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**Table 1**: Actions of sulodexide in different steps of the inflammatory process and their implications [4-6, 41-43, 46-48, 54-60].

| Action | Experimental system | Probable implication |
|--------|---------------------|----------------------|
| **On the regulation of inflammatory response** | | |
| Reduction of monocyte chemotactant protein-1 (MCP-1) | In vitro | Decrease of leukocyte infiltration |
| Reduction of vascular endothelial growth factor (VEGF) | Mouse | Decrease of monocyte infiltrate, vascular permeability and vasodilation |
| Reduction of transforming growth factor-β1 (TGF-β1) | Human | Reduction of the inflammatory infiltrate |
| Reduction of tumor necrosis factor-α (TNF-α) | Mouse | Reduction of the inflammatory infiltrate |
| Decrease of IL-6, IL-8, IL-1b, IL-2, IL-10, IL-13, Interferon β and C reactive protein | Human, mouse, in vitro | Reduction of the inflammatory infiltrate, modulation of the inflammatory/immune response |
| Reduction of macrophage inflammatory protein-1β (MIP-1β) | In vitro | Reduction of the inflammatory infiltrate |
| Reduction of granulocyte colony-stimulating factor (G-CSF) and of granulocyte monocyte colony-stimulating factor (GM-CSF) | In vitro | Reduction of neutrophils and activated monocytes. Modulation of the inflammatory/immune response |
| **On oxidative stress** | | |
| Increase of superoxide dismutate (SOD) | Mouse | Reduction of the oxidative stress (↓ tissue damage) |
| Reduction of reactive oxygen species (ROS) | In vitro | Reduction of the oxidative stress (↓ tissue damage) |
| **On tissue damage** | | |
| Reduction of glomerular heparanase-1 | In vitro | Decreased degradation of the glycocalyx and the extracellular matrix |
| Decrease of MMP-9 | In vitro | Decreased degradation of the extracellular matrix |
| **On tissue repair** | | |
| Increase of heparan sulfate of cell surface | In vitro | Glycocalyx and extracellular matrix repair |
| Increase of hepatocyte growth factor (HGF) | Human | Tissue regeneration (endothelial cells and other), antioxidant and antiapoptotic |
| Increase of the mitogenic activity of fibroblast growth factor-1 and 2 (FGF-1 and FGF-2) | In vitro | Tissue re-epithelization, angiogenesis |
| Increase of endothelial glycocalyx thickness | Human | Restoration of damaged endothelial glycocalyx and its functions |
| Restore endothelial cells replication | In vitro | Increase ability of endothelial cells to heal after mechanical injury |

**Figure 4**: The steps that occur in the inflammatory process of chronic venous disease are shown: glycocalyx thinning, activation of endothelial cells, diapedesis, the release of various inflammatory cytokines by endothelial cells and macrophages, the release of Reactive Oxygen Species (ROS) and the production of enzymes that may damage the extracellular matrix. The anti-inflammatory actions of sulodexide marked with an “X” protect the glycocalyx and the endothelial cells and reduce the release of ROS, proinflammatory cytokines and proteolytic enzymes, especially MMP.
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Disclosure

Dr. Alberto Frati is at present Medical Director of Alfa Wasserman (Mexico) producer of sulodexide. Dr. Luis Fernando Flota has been speaker for Alfa Wasserman, Bayer, Bristol, UCB, Takeda and Goretex.

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