MYCELIAL INCOMPATIBILITY OF SCLEROTINIA SCLEROTIORUM ISOLATES FROM A SINGLE RAPESEED FIELD IN SLOVAKIA

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Abstract

Sclerotinia sclerotiorum (Lib.) de Bary (1884) is economically important pathogen of rapeseeds damaging up to 20% of plants in Slovakia. There is a suggestion that S. sclerotiorum reproduces predominantly asexually, thus genetically identical clonal populations are dominating in fields. Genetically distinct isolates can be distinguished by mycelial compatibility-incompatibility grouping system. The aim of the study was to determine genetic variability of S. sclerotiorum based on mycelial interaction within a single oilseed rape field in Slovakia.

S. sclerotiorum isolates were obtained from flower petals of commercial rapeseeds field in Hul, Slovakia (2018) by transferring a single hyphal tip to the plates with potato dextrose agar (PDA). Two sets of 20 isolates were paired in all possible combination and cultivated on PDA amended by bromophenol blue. Mycelial interactions were scored after 7 days. Each of the 20 isolates in 1st set was unique and belonged to the different mycelial compatibility groups (MCGs). In the 2nd set, 18 MCGs was determined with one isolate and only 1 MCG consisted of 2 isolates. No prevalent MCG was found in this study, suggesting that S. sclerotiorum population affecting the target rapeseed field consisted of a very diverse group of isolates. The high level of incompatible reactions found in this first study from Slovakia may indicate that S. sclerotiorum undergoes frequent outcrossing in rapeseed stands.

Keywords: mycelial compatibility groups, mycelial interactions, Sclerotinia sclerotiorum, rapeseeds, genetic diversity

INTRODUCTION

In 2019, rapeseed (Brassica napus L.) was grown on an area of about 147,000.00 ha (11% of arable land area) in Slovakia (ŠUSR, 2020). Sclerotinia sclerotiorum (Lib.) de Bary (1884) is one of its economically most important pathogens damaging up to 20% of plants in years with higher precipitation during flowering (Bokor and Bečka, 2017).

Patterns of pathogen’s genetic diversity are likely affected by modes of reproduction (Aldrich-Wolfe and Nelson, 2015). Previous studies have suggested that homothallic S. sclerotiorum reproduces predominantly asexually and in fields is dominated by clonal populations that are genetically identical (Kohl et al. 1992). Genetically distinct isolates can be easily distinguished by a quick and inexpensive macroscopic assay of the self-nonself recognition method followed by inclusion them into mycelial compatibility-incompatibility groups (Kohn et al., 1990). Mycelial compatibility is the ability of two strains of filamentous fungi to anastomose and form one confluent colony. Different mycelial compatibility groups (MCGs) represent genetically distinct isolates (Kohn et al., 1991).

Data of the Kohn et al. (1991) demonstrate that the mycelial incompatibility in S. sclerotiorum occurs even within a local field population. Since there is no information available on MCGs of the S. sclerotinia populations in rapeseeds fields in Slovakia, the aim of the study was to determine genetic variability of S. sclerotiorum based on mycelial interaction within a single oilseed rape field.

MATERIAL AND METHODS

Isolates of S. sclerotiorum were obtained from flower petals of commercial rapeseed field. Fresh flower petals were sampled randomly along the diagonal direction in the target field. Collected flower petals were transported to the laboratory in paper bags and immediately cultivated on potato dextrose agar (PDA) at room temperature for 7 days. Four flower petals without surface sterilization were placed to each plate (90 mm in diameter). A total 40 isolates were purified by transferring a mycelial plug (6 mm in diameter) followed by transferring of the single hyphal tip 3 times to the plates with PDA. Hyphal segments were cut using a small needle under a binocular microscope in interval of 24 – 48 hours. Identification of S. sclerotiorum isolates was based on morphology of the mycelial mat and sclerotia formation traits. All isolates collected in 2018 originated from the same crop located in Hul, Nitra Region, Slovakia.

Mycelial compatibility grouping was determined according to the Schafer and Kohn (2006) with modification. Isolates were grown on PDA for 5 days in dark at the temperature of 20 ± 2°C. Mycelial plugs of two strains (6 mm in diameter), obtained at least 10 mm inside from the growing edge of the colony, were placed in Petri dish (90 mm in diameter) 30 mm apart. Mycelial plugs were cultivated on PDA amended by bromophenol blue 50 mg L−1. Bromophenol blue was added to enhance the visibility of incompatible reaction. Pairing plates were incubated in dark at the temperature of 20 ± 2°C. Compatible and incompatible reactions were scored after 7 days. Compatibility occurred when no reaction line occurred in interaction zone, and the pairings formed one confluent colony. Incompatible pairings formed a visible reaction in interaction zone such a yellow to green line visible on the colony reverse, or a band of sparse aerial mycelium on colony surface.

Two sets of isolates were paired in all possible combination. Each set consisted of 20 isolates and 190 combinations. Each isolate was paired by itself as a control of compatibility. Experiment was repeated three times. In case of inconsistent results, the assay was repeated once more.

RESULTS

Flower petal isolates of S. sclerotiorum obtained from a single rapeseed field were compared with each other in two sets. Each of the 20 isolates in 1st set was unique and belonged to the different MCGs (Table 1). In the 2nd set, 18 MCGs was determined with one isolate and only 1 MCG consisted of 2 isolates (Table 2). No prevalent MCG was found in this study, suggesting that S. sclerotiorum population affecting the target rapeseed field consisted of a very diverse group of isolates.

Of the 190 combinations, the most pairings (136 in 1st set and 151 in 2nd set) showed incompatible reaction with dark mycelium and/or green-yellow line in
interaction zone on the colony reverse (Figure 3 – 4). Only isolate 295 (2nd set) gave this single type of incompatible reaction in combination with all pairings. Rest of the combinations showed modest form of incompatible reaction – a band of sparse aerial mycelium on colony surface (Figure 2); and as mentioned above, 2 isolates (287 × 336) in 2nd set were compatible. All self-pairings gave a compatible reaction (Table 1 – 2, Figure 1).

**Table 1** First set of Sclerotinia sclerotiorum flower petal isolates obtained from a single rapeseed field in Hul, Slovakia in 2018

| 340 | 342 | 347 | 355 | 358 | 362 | 373 | 377 | 382 | 383 | 386 | 388 | 391 | 394 | 396 | 400 | 403 | 404 |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| □  | ●   | ●   | ○   | ●   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ○   | ●   | ●   | ●   | ●   | ●   |
| □  | ●   | ●   | ○   | ●   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ○   | ●   | ●   | ●   | ●   | ●   |
| □  | ○   | ●   | ○   | ●   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ○   | ●   | ●   | ●   | ●   | ●   |
| □  | ●   | ●   | ○   | ●   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ○   | ●   | ●   | ●   | ●   | ●   |
| □  | ○   | ●   | ●   | ○   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ○   | ●   | ●   | ●   | ●   | ●   |
| □  | ●   | ●   | ○   | ●   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ○   | ●   | ●   | ●   | ●   | ●   |
| □  | ○   | ●   | ○   | ●   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ○   | ●   | ●   | ●   | ●   | ●   |
| □  | ○   | ●   | ●   | ○   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ○   | ●   | ●   | ●   | ●   | ●   |
| □  | ○   | ●   | ●   | ○   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ○   | ●   | ●   | ●   | ●   | ●   |
| □  | ○   | ●   | ●   | ○   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ○   | ●   | ●   | ●   | ●   | ●   |
| □  | ○   | ●   | ●   | ○   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ○   | ●   | ●   | ●   | ●   | ●   |

**Table 2** Second set of Sclerotinia sclerotiorum flower petal isolates obtained from a single rapeseed field in Hul, Slovakia in 2018

| 269 | 272 | 277 | 280 | 283 | 287 | 290 | 291 | 295 | 297 | 299 | 300 | 303 | 308 | 309 | 318 | 320 | 323 | 329 | 333 | 336 |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| □  | ●   | ●   | ○   | ●   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   |
| □  | ○   | ●   | ●   | ○   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   |
| □  | ○   | ●   | ●   | ○   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   |
| □  | ●   | ●   | ○   | ●   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   |
| □  | ○   | ●   | ●   | ○   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   |
| □  | ○   | ●   | ●   | ○   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   |
| □  | ●   | ●   | ○   | ●   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   |
| □  | ○   | ●   | ●   | ○   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   |
| □  | ○   | ●   | ●   | ○   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   |
| □  | ○   | ●   | ●   | ○   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   |
| □  | ○   | ●   | ●   | ○   | ●   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   | ○   | ●   |

**Legend:**
- □ compatible reaction
- ○ incompatible reaction with a band of sparse aerial mycelium in the interaction zone
- ● incompatible reaction with dark mycelium and/or green-yellow line in interaction zone

**DISCUSSION**

In this study we assumed a presence of a clonal population with at least one prevalent MCG from the single rapeseed field. The reason of this assumption was that homothallic fungi as *S. sclerotiorum* are reproducing predominantly by haploid selfing (Kohn, 1995), what is functionally equivalent to clonal reproduction (Attanayake et al., 2014). Our assumptions were not confirmed, as the target rapeseed field consisted of a very diverse group of isolates mostly incompatible with each other. Since classification of isolates into MCGs is routinely used as a quick marker for genotyping of *S. sclerotiorum* (Schafer and Kohn, 2006) and a sexual recombination is the main source of genotypic variations (Attanayake et al., 2014), our results may indicate the occurrence of highly recombinant population with limited presence of clonality in the single field during the sampled year. Mechanisms of the widespread genetic recombination in homothallic species are still obscure (Attanayake et al., 2019). One of the possibilities is the forming of sclerotia from hyphae of different genotype resulting in recombinant ascospores (Sexton et al., 2006). Single sclerotium from Brazilian common bean comprised of 1 – 5 distinct haplotypes (Lehner et al., 2015), while single sclerotium from Australian sunflower comprised of only one genotype, despite of infections containing multiple genotypes (Ekins et al., 2010).

Our isolates were obtained from flower petals of rapeseeds contaminated or infected by ascospores. Ascospores released during flowering are considered as the main source of inoculum for rapeseeds infections. In Australian rapeseeds field was not found significant population subdivision between the ascospore and stem lesion populations, suggesting that there are no genetically defined subgroup of isolates causing stem infections (Sexton et al., 2006).

Many of our pairings showed incompatible reaction with dark mycelium in interaction zone. Fungal colonies of different isolates often display melanization due to hyphal killing or lysis in the zone between them (Henson et al., 1999). Karimi et al. (2012) in their study distinguished 3 levels of incompatible reactions – partial compatibility, intermediate incompatibility, and complete incompatibility. Kohn et al. (1990) scored pairings into the 5-type of incompatible reactions. The strong incompatible reactions given by the presence of dark melanised mycelium in interaction zone were not mentioned in any of these studies.
CONCLUSION

No prevalent mycelial compatibility group was found in this study, suggesting that S. sclerotiorum population affecting the target rapeseed field consisted of a very diverse group of isolates. The high level of incompatible reactions found in this first preliminary study from Slovakia should be in line with findings that S. sclerotiorum, in addition to haploid selfing and clonal sclerotial production, undergo frequent outcrossing in nature (Attanayake et al., 2014). However, to support this finding, there is a need for more extent research from larger geographical area and different host plant.

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SUPPLEMENTARY FIGURES

Figure 1 Compatible reaction in pairings between the same flower petal isolates of Sclerotinia sclerotiorum (isolates 391 × 391)

Figure 2 Incompatible reaction with thin a band of sparse aerial mycelium in the interaction zone in pairings between different flower petal isolates of Sclerotinia sclerotiorum (isolates 391 × 394)
Figure 3 Incompatible reaction with dark mycelium in interaction zone in pairings between different flower petal isolates of Sclerotinia sclerotiorum (isolates 382 × 396)

Figure 4 Incompatible reaction with green-yellow line in interaction zone in pairings between different flower petal isolates of Sclerotinia sclerotiorum (isolates 377 × 403)