INVESTIGATION OF PHARMACOGNOSTICAL, PHYTOCHEMICAL, AND PHARMACOLOGICAL ACTIVITY OF AERIAL ROOTS OF *FICUS BENGHALENSIS* LINN

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ABSTRACT

Objective: *Ficus benghalensis* Linn. (Moraceae family) is commonly known as banyan tree in English, which is used traditionally in India. The literature survey showed that the aerial roots of this plant are yet to be explored. Our main interest is to evaluate its pharmacognostic and phytochemical character by the standard monograph and to explore its in vitro antioxidant and in vivo analgesic activity study with ethyl acetate extract.

Methods: Pharmacognostic evaluation and phytochemical screening have been done using standard monograph. An in vitro antioxidant activity using ethyl acetate extract has been done using four different methods. In vivo analgesic activity of the ethyl acetate extract has been evaluated by acetic acid-induced writhing test in mice and tail flick method.

Results: Aerial roots of *F. benghalensis* have been found the rich source of steroidal glycosides, cardiac glycosides, flavonoids, tri-terpenoids, and phenols. The presence of phellem, phellogen, xylem, and phloem has been found after microscopic investigation. All the pharmacognostic parameters proved its purity. Results showed the absence of heavy metals. The ethyl acetate extract has shown potent antioxidant activity at 100 µg/ml concentration and higher analgesic activity at the concentration of 400 mg/kg than 200 mg/kg.

Conclusion: Pharmacognostic characteristics and phytochemical properties revealed in this study could be used for the pharmacopoeial standard. Ethyl acetate extract showed potent antioxidant and analgesic activity.

Keywords: Antioxidant, Analgesic, Glycoside, Terpenoid, Pharmacognostic.

INTRODUCTION

Nature has been a source of medicinal agents for thousands of years. Even today, plant materials continue to play a major role in primary health care for therapeutic remedies in many developing countries. According to the World Health Organization (WHO), a major part of the traditional therapies involves the use of plant extracts or their active constituents. The world health organisation (WHO) estimated that about 80% of the populations living in the developing countries depended on traditional medicine for their primary health-care needs [1].

Our plant of interest *Ficus benghalensis* Linn. (Moraceae family) [2] is commonly known as banyan tree in English, “Bot” in Bengali, and “Bargad” in Hindi. It is a large deciduous tree with aerial roots distributed all over India from Sub-Himalayan region and in the forests of Deccan and South India. It is found throughout the year grows in evergreen [3,4].

Different parts of banyan tree used as traditional medicine to get remedies from several ailments. The bark is useful in burning sensation, hemorrhages, diarrhea, dysentery, diabetes, ulcer, and skin diseases. The leaves are good for ulcers, leprosy, skin allergies, etc. The buds are used in diarrhea and dysentery. The latex is useful in rheumatism, hemorrhoids, inflammation, and skin diseases [5].

The present studies were undertaken to establish the pharmacognostic characteristics, phytochemical nature, in vitro antioxidant activity, and in vivo analgesic activity of aerial roots of *F. benghalensis* Linn.

METHODS

Chemical used

Folin–Ciocalteu, sodium carbonate, ethylenediaminetetraacetic acid (EDTA), hydrogen peroxide, and trichloroacetic acid (TCA) were purchased from Merck Specialist Pvt., Ltd., Mumbai, India. 1, 1-diphenyl-2-picrylhydrazyl (DPPH) and deoxyribose were purchased from Hi-Media Laboratories Pvt., Ltd., Mumbai, India. Ascorbic acid, thioarbituric acid (TBA), and phenazine methosulphate (PMS) were procured from Loba Chemie Pvt., Ltd., Mumbai, India. Ferric chloride, potassium ferric cyanide and nitroblue tetrazolium (NBT), and butylated hydroxyanisole (BHA) were purchased from Sigma Chemical Co., Ltd., St. Louis, MO, USA. Diclofenac was from Macleods and pentazocine was purchased from Sun Pharmaceutical Industries Ltd. All other chemicals and reagents were of analytical grade.

Collection and authentication of plant material

The aerial roots of banyan tree were collected from Amdanga, North 24 Parganas, West Bengal, India, in October–November. The plant materials were authenticated and herbarium was deposited to (CNH/Tech II/2015/9/245) Central National Herbarium, Botanical Survey of India, Howrah – 711 103.

Macroscopic characteristics

Macroscopic characteristics identification of medicinal plant materials is based on its shape, size, surface, texture, fracture, color, and appearance of the cut surface. The collected cut pieces of aerial roots were 8-10 cm long and macroscopic characteristics were evaluated.

Microscopic characteristics

Fresh specimens of aerial roots of *F. benghalensis* were collected and washed thoroughly with the distilled water. Then, the samples were fixed in 2.5% glutaraldehyde. Freehand semi-thin transverse sections were done with the help of a razor. The sections were allowed to dehydrate by passing it from various gradations of ethanol starting from 30% to 100%. After the dehydration, sections were mounted over
a drop of DPX taken in a clean slide and directly observed under light microscope (LEICA DM750) under low magnification.

Drying
After collection, the aerial roots were washed with fresh water and shade dried at temperature 27°C–30°C for 5–7 days. Fresh dried aerial roots were cut into small pieces (2–3 cm), grounded in a mechanical mill, and screened through 0.6–0.8 mm sized particle mesh. This powder was stored in an airtight container and used for further extraction.

Physicochemical parameters
Evaluation of physicochemical parameters such as total ash, water-soluble ash, acid-insoluble ash, loss on drying, and presence of heavy metals were detected as per the guidelines of IP [6]. The intensity of the color produced with the test solution matched with that produced with the standard solution. Water, alcohol, chloroform, and petroleum ether-soluble extractive values [7] were also determined.

Extraction
The powdered plant materials (1000 g) were macerated using methanol as solvent and subjected to a rotary evaporator for evaporation of the solvent. The extract was further fractionated using hexane and ethyl acetate.

Phytochemical screening
The extracts were subjected to preliminary phytochemical analysis for detection of fats and oils, flavonoid, alkaloid, glycoside, steroid, triterpenoid, saponin, tannin, protein, and carbohydrate as per standard procedures [8].

Antioxidant activity study
Ascorbic acid was selected and used as a standard for the all in vitro antioxidant activities. All the solutions were freshly prepared.

Hydroxyl radical scavenging activity
The hydroxyl radical scavenging activity was measured by following the method of Halliwell et al. [9], and it was slightly modified. To the extract of different concentrations (10, 25, 50, and 100 µg/ml), 0.1 ml (104 µM) of EDTA, 0.01 ml (100 µM) of FeCl3, 0.1 ml (1 mM) of H2O2, and 0.36 ml (2.8 mM) of deoxyribose were mixed. The final volume was prepared 1 ml with phosphate buffer (20 mM, pH - 7.4). The mixture was then incubated at 37°C for 1 h. Then, 1.0 ml portion of the incubated mixture was mixed with 1.0 ml of 10% TCA. 1.0 ml of 0.5% TBA (in 0.025 M NaOH containing 0.25% BHA) and heated at 95°C on water bath for 15 min to develop the pink color, and the absorbance was measured at 532 nm against the blank sample using a UV-visible spectrophotometer [3]. The hydroxyl radical scavenging activity of the extract was determined using the following equation:

\[
\text{Percentage inhibition} = \frac{1 - \text{absorbance of test/absorbance of control}}{\text{absorbance of test/absorbance of control}} \times 100
\]

Reductive activity
To the 1 ml ethyl acetate extract of different concentrations (10, 25, 50, and 100 µg/ml), 2.5 ml of 100 mM phosphate buffer of pH 6.6 and 2.5 ml of 1% potassium ferricyanide solution were added. The mixture was incubated at 50°C for 20 min and 2.5 ml of 10% TCA was added. It was centrifuged at 3000 rpm for 10 min. 2.5 ml of the supernatant was mixed with 2.5 ml of distilled water and 0.5 ml of 0.1% ferric chloride. Then, the absorbance of the reaction mixture was measured spectrophotically at 700 nm against the blank sample using a UV-visible spectrophotometer [10]. Higher the absorbance of the test mixtures indicated higher reductive activity.

Superoxide radical scavenging activity
Superoxide radical scavenging activity was determined by mixing 1 ml of NBT solution (156 µM NBT in 100 mM phosphate buffer, pH 7.4), 1 ml NADH solution (468 µM in 100 mM phosphate buffer, pH 7.4), and 0.1 ml of sample solution of different concentrations of extract (10, 25, 50, and 100 µg/ml). To this mixture, 100 µl of PMS solution (60 µM PMS in 100 mM phosphate buffer, pH 7.4) was added. The reaction mixture was then incubated at 25°C for 5 min, and the absorbance was measured at 560 nm against the blank sample using a UV-visible spectrophotometer [11]. Lower the absorbance of the test mixtures indicated higher superoxide radical scavenging activity.

Superoxide radical scavenging effect (%) = (1–absorbance of test/absorbance of control)×100

DPPH radical scavenging activity
1 ml of 0.1 mM methanolic solution of DPPH was added to 3 ml of the extract of different concentrations (10, 25, 50, and 100 µg/ml). After continuous shaking of the mixture, it is allowed to stand for 30 min at room temperature. Then, the absorbance was measured at 517 nm against the blank using UV-visible spectrophotometer. Decreased the absorbance values of the reaction mixture were indicated increased free radical scavenging activity [12]. DPPH radical scavenging capability was calculated using the following equation:

\[
\text{DPPH scavenging effect} (\%) = \left(1 - \frac{\text{absorbance of test/absorbance of control}}{\text{absorbance of control}}\right) \times 100
\]

50% inhibitory concentration (IC50) value determination
IC50 value of the standard and extract from DPPH radical, hydroxyl radical, and superoxide radical scavenging activity was calculated by interpolation from the linear regression analysis.

Estimation of total phenolic content
The total phenolic content of ethyl acetate extract was estimated by following the Folin–Gioeal mean method of Sinkard and Singleton [13]. 0.1 ml (1000 µg) solution of the extract was taken in volumetric flask and diluted with distilled water up to 46 ml. 1 ml of Folin–Gioeal reagent was added to it and mixed thoroughly. After 3 min, 3 ml of 2% NaCO3 was added and the mixture was allowed to stand for 2 h with random shaking. The absorbance was measured at 760 nm. Pyrocatechol was taken as standard. The concentration of total phenolic contents within the extract estimated as equivalent to the microgram of pyrocatechol by the following equation obtained from standard pyrocatechol graph.

\[
\text{Absorbance} = \frac{0.001 \times \text{concentration of pyrocatechol (µg)}}{0.0033} [14].
\]

Analogic activity study
All the animals (albino mice) of 25–30 g wt were obtained from animal house of BCDA College of Pharmacy and Technology, Barasat. The animals were kept under standard laboratory conditions at room temperature with normal humidity. They were given standard diet and water ad libitum. The animals have fasted for 12 h with the availability of normal water before the experiment. The animals were divided into four groups (n=6), control, standard, test 1, and test 2 (two different concentrations of ethyl acetate extract). All the procedures for this in vivo analgesic activity study were approved by the institutional animal ethical committee with approval no BCDA/CPT/IAEC/02/2014–2015.

Acetic acid-induced writhing test
All the animals from the control group were treated with acetic acid 1%v/v solution. Second, third, and fourth groups are treated with standard drug solution diclofenac sodium 10 mg/kg, ethyl acetate extract 200 mg/kg, and ethyl acetate extract 400 mg/kg, respectively, and also administered acetic acid to all the animals of three groups 15 min after the standard and test treatment [15]. Note the onset of writhing and calculate the percentage inhibition of writhing response compared with control group.

\[
\% \text{ inhibition of writhing} = \left\{1 - \frac{\text{no of writhing of test/no of writhing of control}}{\text{no of writhing of control}}\right\} \times 100
\]

Tail flick method
Basal reaction time (tail withdrawal time) to radiant heat was noted by placing the tail end (2 cm) of mice to the radiant heat source using
analyse a microtome. A cutoff time of 15 s is maintained. The four groups were treated with carboxymethyl cellulose (CMC) 0.5% w/v, standard pentazocine 30 mg/kg, ethyl acetate extract 200 mg/kg, and ethyl acetate extract 400 mg/kg, respectively, and noted the reaction time at 30, 60, 90, and 120 min after the drug administration [16].

**Statistical analysis**

All the statistical analyses were done using Microsoft Office Excel 2007. Statistical significance was calculated by paired t-test.

**RESULTS**

**Pharmacognostic investigation**

The value of loss on drying is 3.62%, the alcohol-soluble extractive value is 7.44%, water-soluble extractive value is 12%, the chloroform-soluble extractive value is 11.1%, and the ether-soluble extractive value is 7.6%. Total ash value is 3.32%, acid-insoluble ash is 2.71%, and the water-soluble ash value is 2.92%. The limit test of heavy metals showed less intense color than standard solution, so it could be inferred that heavy metal is absent.

**Macroscopic character**

Cut pieces of aerial roots occurred in 8–10 cm long, 0.1–0.5 cm thick, cylindrical, unbranched, or branched. The external surface of the aerial root of *F. benghalensis* is gray, the cut surface is reddish-brown, rough due to longitudinal and transverse cracks and transverse rows of lenticels, fracture, and bark portion is fibrous, tough, and short in wood portion.

**Microscopic character**

Anatomy of the roots revealed a well-characterized secondary growth pattern. Outermost layer consists of bark followed by the phellem and phellogen. The structure of primary vascular bundle in this stage was revealed by the radially arranged exarch primary xylem located in the central region. The strands of secondary vascular tissues were arranged collaterally. Cambial activity causes the formation of medullary rays which runs between the xylem and the phloem through the cambium. Xylem vessel shows almost circular in outline. Overall, the anatomical structures revealed the picture of a typical dicot root showing a secondary growth. No other kind of special character, as well as an anomaly, was observed. The present anatomical study was predictable to conclude that in this case there is hardly any change in their anatomical structures and cellular pattern in relation to their metabolic and chemical profile.

**Phytochemical investigations**

The N-hexane extract of aerial roots of *F. benghalensis* Linn. has shown the presence of saponin, triterpenoid, fats, and oils. Ethyl acetate and methanol extract showed the presence of flavonoid, saponin, steroid, triterpenoids, glycosides, tannins, and phenols. The presence of alkaloid was observed only in the methanol extract. Carbohydrate and protein were absent.

**Total phenolic content**

1 mg of each concentration of the ethyl acetate extract contained 36.66±0.05 μg of pyrocatechol equivalents of phenols.

**Antioxidant activity**

**Hydroxyl radical scavenging activity**

The extracts have shown Hydroxyl radical scavenging activity in the dose-dependent manner which is expressed as the percentage of inhibition (Fig. 1). About 73.49±8.93% of the Hydroxyl radical scavenging activity was observed in case of the extracts at 100 μg/mL. The percentages of inhibition of hydroxyl scavenging activity of extract in different concentrations are lower than standard (98.83±5.85%) but are significant statistically.

**Reductive activity**

The extract showed the increased reducing power activity with the increment of dose. It has shown a significant and highest reducing power activity (optical density 0.304) at highest dose of 100 μg/ml which is lower than standard (0.35±0.009) (Fig. 2).

**Superoxide radical scavenging activity**

Ethyl acetate extract showed significant superoxide radical scavenging property at the highest concentration (100 μg/ml). It has shown about 54.83±0.73% of superoxide radical scavenging (Fig. 3) activity, whereas standard showed 71.65±1.43% of superoxide radical scavenging property. The extract showed the dose-dependent activity.

**DPPH radical scavenging activity**

The ethyl acetate extract showed highest antioxidant property in DPPH radical scavenging method. The extract showed a dose-dependent inhibition of DPPH radical. At dose of 100 μg/mL, extract showed nearly 83.90±2.6% of inhibition which significant and close to the activity shown by the standard drug (91.37±1.23) (Fig. 4).

All statistically analyzed data are expressed as mean ± standard deviation (n=6). Statistical significance (p) calculated by paired t-test. p<0.05 considered to be statistically significant.

**Fig. 1:** Hydroxyl radical scavenging activity of different concentrations of std. (standard ascorbic acid) and ethyl acetate extract. Values are expressed as mean ± standard deviation where n=6. All data are statistically significant at p<0.05

**Fig. 2:** Reductive activity of different concentrations of std. (standard ascorbic acid) and ethyl acetate extract. Each point represents mean ± standard deviation where n=6. All data are statistically significant at p<0.05
In hydroxyl radical scavenging ability method, the results showed that the IC$_{50}$ value of standard and ethyl acetate extract was 11.53 µg/ml and 45.25 µg/ml, respectively. In superoxide radical scavenging method, the results showed that the IC$_{50}$ value of standard and ethyl acetate extract was 39.18 µg/ml and 72.75 µg/ml, respectively. In DPPH radicals scavenging method, the results showed that the IC$_{50}$ value of standard and ethyl acetate extract was 10.57 µg/ml and 14.75 µg/ml, respectively.

Acetic acid-induced writhing test
Ethyl acetate extract showed 39.33±0.75% and 60±0.63% inhibition of writhing at 200 mg/ml and 400 mg/ml concentration, respectively, and diclofenac sodium showed 66.67±0.52% inhibition. Results showed that the inhibitory effect of diclofenac sodium was more than the extract, but all the data are statistically significant at p<0.001 when compared with control (Fig. 5).

Tail flick method
Results showed that normal CMC (0.5%)-treated group (control) did not show any significant difference in the reaction time on tail flick up to 120 min of observation. Pentazocine (30 mg/kg)-treated group (standard) showed maximum reaction time of 13.2±0.3 s at 90 min, but the response is lowered when observed at 120 min. Ethyl acetate extract of 200 mg/kg body weight and 400 mg/kg body weight showed maximum reaction time of 7.2±0.33 s and 10.03±0.5 s, respectively, and diclofenac sodium showed 66.67±0.52% inhibition. Results showed that the inhibitory effect of diclofenac sodium was more than the extract, but all the data are statistically significant at p<0.001 when compared with control (Fig. 5).
The DPPH radical scavenging activity

This model is widely used to evaluate free radical scavenging activity. DPPH is a stable free radical that accepts an electron or hydrogen atom to become a stable diamagnetic molecule [20]. The absorption maximum of a stable DPPH radical in methanol was 517 nm [21]. The decrease in absorbance of DPPH radical caused by antioxidants because of the reaction between antioxidant molecules and radical progressed, results in the scavenging of the radicals by hydrogen donation. It is visually noticeable that as a change in color from purple to yellow. Based on the data obtained from this study, ethyl acetate extract was effective free radical scavenger that reacts with free radicals and acts as a primary antioxidant. Fig. 4 shows a significant (p<0.05) DPPH radicals scavenging ability of the standard (91.36±1.24 % inhibition at 100 µg/ml concentration and IC<sub>50</sub> value is 10.57 µg/ml) and extract (83.96±2.95% inhibition at 100 µg/ml concentration and IC<sub>50</sub> value is 14.75 µg/ml).

Acetic acid-induced writhing reflex

This method was done to evaluate the peripheral analgesic effect of the extract. The pain sensation is due to the production of prostaglandin through cyclooxygenase pathway. Diclofenac sodium and the extract may be acted by inhibition of cyclooxygenase pathway [22].

Tail flick method

This method was done to evaluate central Analgesic activity of the extract. The results showed that there was a significant time-dependent initial increase (from 30 min to 90 min) and then gradual decrease (at 120 min) in the tail flick time of mice compared to control. This may be due to the agonist-antagonist action of pentoazone on opioid receptors [23]. The ethyl acetate extract showed its potent analgesic activity may be by acting on opioid receptors.

CONCLUSION

The present study proved that the ethyl acetate extract of aerial roots of *F. benghalensis* Linn. exhibited significant antioxidant potential with increased concentration and significant analgesic activity which is most effective at higher concentration. The ethyl acetate extract showed high total phenolic content and the phytochemicals which are present in the extract are important sources of antioxidants. Further studies will be carried out to explore the bioactive molecules responsible for the antioxidant and analgesic activity.

ACKNOWLEDGMENT

The authors are thankful to Mr. Sandeep Chakraborty, Asst. Prof. Department of Botany, Achhruram Memorial College, Purulia, for his help to the study.

AUTHOR’S CONTRIBUTION

Mrs. Kamalika Mazumder has performed all the experiments and prepared the manuscript. Dr. Himangshu S. Maji and Dr. Nirupranath Bala helped in choosing the experimental protocols and manuscript correction.

CONFLICTS OF INTEREST

All the authors declare that there are no conflicts of interest.

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Table 1: Preliminary pharmacognostic evaluation

| Parameters                  | Values (%w/w) |
|-----------------------------|---------------|
| Loss of drying              | 3.62          |
| Extractive value            |               |
| Alcohol-soluble extractive  | 7.4           |
| Water-soluble extractive    | 12            |
| Glucomannan-soluble extractive | 11.1     |
| Ether-soluble extractive    | 7.6           |
| Ash value                   |               |
| Total ash                   | 3.32          |
| Acid-insoluble ash          | 2.71          |
| Water-soluble ash           | 2.92          |
| Heavy metals                | Absent (No turbidity shown) |

Table 2: Phytochemical investigation

| Phytochemical extract | N-hexane extract | Ethyl acetate extract | Methanol extract |
|-----------------------|------------------|----------------------|------------------|
| Alkaloid              | –                | –                    | +                |
| Flavonoid             | –                | ++                   | +                |
| Saponin               | +                | +                    | +                |
| Steroid               | –                | ++                   | +                |
| Cardiac glycoside     | –                | +                    | +                |
| Fats and Oils         | ++               | –                    | +                |
| Tannin                | –                | +                    | +                |
| Phenol                | –                | +                    | ++               |
| Triterpenoid          | ++               | +                    | +                |
| Carbohydrate          | –                | –                    | –                |
| Proteins and amino acid | –             | –                    | –                |

increased concentrations proved the antioxidant property of extract and standard. Fig. 1 shows a significant (p<0.05) increase in hydroxyl radical scavenging ability with increased concentration of the standard (98.8±5.85% inhibition at 100 µg/ml concentration and IC<sub>50</sub> value is 11.53 µg/ml) and extract (73.49±8.93% inhibition at 100 µg/ml concentration and IC<sub>50</sub> value is 45.25 µg/ml).

Effect on reductive ability

The reducing power activity of the extract is a significant indicator of the antioxidant property. This property measured by the ability of the extracts to convert Fe³⁺ to Fe²⁺ and by the donation of an electron [18]. The reducing power of a compound is a significant indicator of its potential antioxidant activity. The reducing power of extracts increased with increased concentration of samples. Fig. 2 shows a significant (p<0.05) increase in reducing ability with increased concentration of the standard (optical density 0.351±0.009 at 100 µg/ml concentration) and extract (optical density 0.304±0.008 at 100 µg/ml concentration).

Effects on superoxide anion radical scavenging activity

Superoxide radical is a biological source of reactive oxygen species. It is responsible for the production of hydroxyl radicals and oxygen free radicals. These free radicals are causing oxidative stress [19]. In the PMS-NADH-NBT system, superoxide anion from dissolved oxygen from the coupling reaction of PMS-NADH reduces NBT. The decrease in absorbance at 560 nm with antioxidants indicates the consumption of superoxide anion in the reaction mixtures, Fig. 3 shows a significant (p<0.05) increase in superoxide radical scavenging ability with increased concentration of the standard (71.65±1.43% inhibition at 100 µg/ml concentration and IC<sub>50</sub> value is 39.18 µg/ml) and extract (54.83±0.73% inhibition at 100 µg/ml concentration and IC<sub>50</sub> value is 72.75 µg/ml).

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