Research Article

Enhancing and Complementary Mechanisms of Synergistic Action of Acori Tatarinowii Rhizoma and Codonopsis Radix for Alzheimer’s Disease Based on Systems Pharmacology

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Background and Aim. Alzheimer’s disease (AD) is a common neurological disorder worldwide. In traditional Chinese medicine (TCM), Acori Tatarinowii Rhizoma (ATR) and Codonopsis Radix (CR) are common herbs used to treat AD. However, due to the many active ingredients and targets in these herbs, it is difficult to clarify the synergistic mechanism of ATR and CR. To reveal the multicomponent synergistic mechanism of ATR and CR in Alzheimer’s disease, we analyzed important components, drug targets, and crucial pathways using a systems pharmacology strategy.

Materials and Methods. In this study, a systems pharmacology-based strategy was used to elucidate the synergistic mechanism of Acori Tatarinowii Rhizoma and Codonopsis Radix for the treatment of AD. This novel systems pharmacology model consisted of component information, pharmacokinetic analysis, and pharmacological data. Additionally, the related pathways were compressed using the Kyoto Encyclopedia of Genes and Genomes (KEGG) database, and the organ distributions were determined in the BioGPS bank.

Results. Sixty-eight active ingredients with suitable pharmacokinetic profiles and biological activities were selected through ADME screening in silico. Based on 62 AD-related targets, such as APP, CHRM1, and PTGS1, systematic analysis showed that these two herbs were mainly involved in the PI3K-Akt signaling pathway, MAPK signaling pathway, neuroactive ligand-receptor interaction, and fluid shear stress and atherosclerosis, indicating that they had a synergistic effect on AD. However, ATR acted on the KDR gene, while CR acted on IGF1R, MET, IL1B, and CHUK, showing that they also had complementary effects on AD. The ingredient contribution score involved 29 ingredients contributing 90.14% of the total contribution score of this formula for AD treatment, which emphasized that the effective therapeutic effects of these herbs for AD were derived from both ATR and CR, not a single herb. Organ distribution showed that the targets of the active ingredients were mainly located in the whole blood, the brain, and the muscle, which are associated with AD.

Conclusions. In sum, our findings suggest that the systems pharmacology methods successfully revealed the synergistic and complementary mechanisms of ATR and CR for the treatment of AD.

1. Introduction

More than 25 million people suffer from dementia worldwide, and Alzheimer’s disease (AD) accounts for approximately 70% of patients with dementia [1–3]. AD is a progressive neurodegenerative disorder caused by extensive synapse loss, the formation of intracellular neurofibrillary tangles, extracellular deposits of β-amyloid peptides (Aβ), inflammation, and oxidative stress in neurons [4–6]. Clinical data also showed that cerebral vascular diseases, especially atherosclerosis, are established risk factors for dementia, particularly AD [7, 8]. All this evidence suggests that the
clinical features of AD are diverse and that the pathological factors are complicated. Therefore, current pharmacology focusing on acetylcholine esterase inhibitors and an N-methyl-D-aspartic acid receptor (NMDAR) modulator is unsatisfactory for ameliorating the symptoms of AD [9]. For example, donepezil, the most effective drug for the treatment of AD, can only temporarily relieve the symptoms and cannot inhibit the progression of AD [10]. Development of LY450139, which has been investigated in a phase III trial of anti-Aβ drug candidates, has been halted due to its inhibition of the NOTCH protein, even though it can significantly reduce Aβ levels in patients [5]. Currently approved drugs for the treatment of AD, such as rivastigmine and memantine, result in various side effects, such as bradycardia or hepatotoxicity [11–13]. Hence, it is critical to explore more systemic, effective, and harmless anti-AD agents.

Traditional Chinese medicine (TCM) has a long history in the treatment of dementia [3, 14]. For example, Kai-Xin San, the representative prescription written by Sun Simiao in Bei Ji Qian Jin Yaofang during the Tang Dynasty, is a TCM formula with antiedementia effects [15]. Modern pharmacological research also verified that active ingredients extracted from Chinese herbs, such as apigenin, β-asarone, and huperzine A, exert curative effects against AD in vivo and in vitro; indeed, some of them have been used in clinical trials [15]. Notably, Acorus Tatarinowii Rhizoma (ATR, the rhizome of A. calamus, also named Acorus calamus var. angustatus, Shi Chang Pu in Chinese) functions by traditional biological experiments is expensive discovery [39]. However, identifying the pharmacokinetic mechanisms of action of TCM. In recent years, based on chemical, pharmacokinetic, and pharmacological data, the methodology of systems pharmacology has helped us to interpret the details of the synergistic mechanisms in Chinese formulas [27–29]. Systems pharmacology integrates pharmacokinetics synthesis data, target screening, pathway interaction, and network analysis to elucidate the therapeutic mechanisms of drugs from the molecular and cellular levels to the tissue and organism levels [26, 30]. Following a systems pharmacology-based strategy, many reports explain the polypharmacological and synergistic mechanisms and predict the principal ingredients and signaling pathways of herb pairs for various diseases [31–34]. Using this model, Lu et al. revealed that Huangqi and Huanglilan exert synergistic therapeutic effects on diabetes mellitus based on five pivotal ingredients [35]. Zhi-Zhu-Wan has efficacious therapeutic effects on functional dyspepsia due to 29 major active components [16]. Based on systems pharmacology, the ingredients from the herb Cistanche tubulosa show synergistic effects on neuroinflammation [36], and the compounds rutin and amentoflavone exert synergistic effects on relieving depression [37].

Consequently, we constructed a systems pharmacology model to explore the therapeutic mechanism of ATR and CR for treating AD. First, the active ingredients selected from the database were screened by two ADME parameters to ensure comprehensiveness. Second, a large-scale analysis of targets was performed by target identification to establish the network. Finally, network construction, including compound-target (C-T), target-pathway (T-P), and compound-target-organ (C-T-O) networks, was applied to reveal the underlying mechanism of the combination of ATR and CR on AD in our study (Figure 1).

2. Materials and Methods

2.1. Chemical Ingredients Database Construction. All of the constituent data of ATR and CR were retrieved from the TCM Systems Pharmacology Database and Analysis Platform (TCMSP, http://lsp.nwu.edu.cn/) [38]. Subsequently, the initial structures of all ingredients were depicted in TIF format using ChemDraw (version 16.0). Additionally, the pharmacology-related properties, including oral bioavailability (OB), drug-likeness (DL), molecular weight (MW), octanol-water partition coefficient predicted by ACD/PhysChem Suite (AlogP), number of acceptor atoms for H-bonds (nHAcc), and number of donor atoms for H-bonds (nHDon), were obtained from TCMSP.

2.2. Active Ingredient Screening by ADME. An early pharmacokinetic evaluation is important due to the failures caused by limited pharmacokinetic profiles in modern drug discovery [39]. However, identifying the pharmacokinetic properties by traditional biological experiments is expensive
and time-consuming, and in silico methods are efficient strategies for subsequent analysis [36, 40]. In our current study, to identify the active ingredients, we used OB and DL as criteria to select components of ATR and CR.

The OB is the key factor for evaluating the ability of drugs to deliver the compound to the systemic circulation by oral administration. Based on 805 structurally defined Western drugs and drug-like molecules, a novel in silico system, OBioavail 1.1, was used to calculate the OB values of all ingredients [41], and those ingredients with OB ≥ 30% were preserved for further analysis.

The values of the DL evaluation approach based on the Tanimoto coefficient were calculated using the following equation: $T(A, B) = \frac{(|A| \times |B|)}{(|A|^2 + |B|^2 - |A \times B|)}$. In this equation, $A$ represents the molecular descriptors of herbal compounds, and $B$ displays the average molecular properties of all compounds in DrugBank [42]. Although DL ≥ 0.18 was a common criterion for ADME screening [43], the number of ingredients of ATR with DL ≥ 0.18 and OB ≥ 30% was only four. Thus, to obtain more details and information about the two herbs, those ingredients with suitable DL ≥ 0.10 were selected as candidates for active ingredients as in a previous study [21, 44]. Additionally, some ingredients with a low OB or DL were supplemented because of their high bioactivity and enrichment through a text-mining method.

2.3. Target Collection. Based on the performance of several approaches integrated with chemometrics methods, message collection, and data mining, the related targets of the active ingredients of ATR and CR were searched. First, the targets of active ingredients were obtained from TCMSP. Second, the active ingredients were submitted to various servers, viz., BindingDB database (http://www.bindingdb.org/bind/index.jsp) [45] and STITCH (http://stitch.embl.de/) [46]. Finally, the targets obtained from the databases were input to UniProt (http://www.uniprot.org/) to standardize the names of the targets.

Then, for further elucidation of the role of ATR and CR in AD treatment, targets of AD were obtained from various servers, viz., Therapeutic Target Database (TTD, http://bidd.nus.edu.sg/group/cjttd/) [47], Comparative Toxicogenomics Database (CTD, http://ctdbase.org/) [48], Online Mendelian Inheritance in Man database (OMIM, http://www.omim.org/), PharmGKB (http://www.pharmgkb.org) [49], and DrugBank (https://www.drugbank.ca/) [50]. Finally, the common targets of ATR and AD or CR and AD were selected for further analysis. Importantly, all targets kept for further analysis were only derived from Homo sapiens.

2.4. Gene Ontology (GO) Analysis and Network Construction. To better decipher the function of the identified targets, we performed GO enrichment analysis for the 62 target proteins by Clue GO, a widely used Cytoscape plugin [51]. The $P$ values were corrected with a Bonferroni step-down method.

Then, for elucidation of the underlying molecular mechanism of action of ATR and CR, three corresponding networks were constructed: (1) A compound-target network (C-T network) was constructed to identify the key targets based on the active ingredients of ATR and CR and their corresponding targets. (2) A target-pathway network (T-P network) was constructed to identify the key targets based on the active ingredients of ATR and CR and their corresponding targets. (3) A compound-associated pathway network (C-T-O network) was constructed to identify the key pathways associated with the active ingredients of ATR and CR and their corresponding targets.
network) was built by determining the relationship between targets and pathways that were extracted from KEGG (Kyoto Encyclopedia of Genes and Genomes, http://www.kegg.jp). P values were set at 0.05 as the cut-off criterion. (3) The related targets and their tissue types were used in the compound-target-organ network (C-T-O network). The target organ location was determined in the BioGPS bank (http://biogps.org) [52]. All visualized networks were constructed by the open software Cytoscape 3.7.0 (http://www.cytoscape.org/) [53].

2.5. Contribution Score Calculation. For further elucidation of the role of each active ingredient of ATR and CR on AD treatment, a contribution score based on the degree listed by Cytoscape 3.7.0 was calculated by the following equation [16]:

\[
A_{ij} = \omega_{ei} + \frac{C_{Ai} + C_{Bi}}{C_{Ai} - C_{Bi}},
\]

\[
\omega_{ei} = \frac{\omega_{ei}}{C_{edge}},
\]

\[
CS (i) = \sum_{ij} C_{ij} \times [A_{ij} \times p_{ij}],
\]

where \(i\) and \(j\) represent the number of components and proteins, respectively. \(C\) is the degree of each component, and \(p\) is the degree of each protein. \(C_{Ai}\) represents the degree of each component only in the CR C-T network, and \(C_{Bi}\) represents the degree of each ingredient only in the ATR C-T network. \(\omega_{ei}\) is the ratio of the degree of each ingredient to all ingredients. \(A_{ij}\) is the index of affinity determined from the \(\omega_{ei}\) value. All degrees in this equation are calculated by Cytoscape 3.7.0. The contribution score (CS) represents the network contribution of one ingredient and its effectiveness in AD.

3. Results

3.1. Differences between ATR and CR. A total of 239 ingredients were retrieved in ATR (105) and CR (134) based on searching in a series of public databases. Lignans and volatile oil were the major components in ATR, whereas polysaccharides and volatile oil were the major components in CR. More details about these ingredients are listed in Table S1.

To explore the molecular differences between ATR and CR, we compared six parameters of these components: MW, AlogP, nHDon, nHAcc, OB, and DL. As shown in Figure 2, the values of these components mainly followed Lipinski’s rule of five [54]. (1) For MW, the average value of the components in ATR (227.44) was significantly lower than that in CR (316.17) \((P = 3.58E - 06)\). (2) The AlogP values of ATR and CR were 2.99 and 3.96, respectively, demonstrating that both components of these two herbs were hydrotropic. (3) For nHDon and nHAcc, the ATR values (0.99 and 2.42) were all significantly lower than those in CR (1.75 and 3.84) \((P = 6.16E - 03, 4.10E - 03)\). (4) For OB, unlike the values mentioned above, ATR possessed a higher average OB value (37.97) than CR (27.98) \((P = 4.35E - 05)\), indicating that ATR had better pharmacokinetic properties. (5) For DL, the DL value of ATR (0.289) \((P = 1.80E - 05)\) was significantly lower than that of CR (0.147) \((P = 5.01 by two tailed t-test (vs. CR).

In summary, these data showed a variation between the ingredients of ATR and CR because of their distinct chemophysical properties. Our results suggested that ATR showed better pharmacokinetic properties, but the ingredients in CR had a better DL. These two herbs contained different main ingredients, which may explain why ATR and CR could produce synergistic and complementary effects.

3.2. Identification of Active Ingredients in ATR and CR. The TCM formula usually contains multiple ingredients, but several of them possess unsatisfactory pharmacodynamic and pharmacokinetic properties, influencing the therapeutic responses [55]. Therefore, it is essential to identify the components with favorable properties, and ADME screening is a useful way to select suitable ingredients from herbs or TCM formulas that have many components. In our current work, we employed two major ADME parameters to filter the active ingredients from ATR and CR. As a result, a total of 68 active ingredients from the 239 ingredients were chosen for further analysis (Table 1).

3.3. Active Ingredients from ATR. Thirty-three active ingredients were selected from ATR through strict ADME screening rules, and most of them showed ideal biological activities. For instance, kaempferol (OB = 41.88, DL = 0.2) attenuates oxidative stress by regulating Bcl-2 in neuronal cells [56]. Marmesin (OB = 50.28, DL = 0.18) forms hydrogen bonds with paraoxonase to reduce nitric oxide levels to protect rats against myocardial infarction [57]. Cycloartenol (OB = 38.69, DL = 0.78) can reduce oxidase activity [58]. Majudin (OB = 42.21, DL = 0.13) inhibits cholinesterase to
| ID   | Molecule name                  | Structure | OB (%) | DL |
|------|--------------------------------|-----------|--------|----|
| CR1  | Poriferasta-7,22E-dien-3beta-ol | ![Structure Image](image1.png) | 42.98  | 0.76 |
| CR2  | 2-methoxyfuranodiene           | ![Structure Image](image2.png) | 53.58  | 0.13 |
| CR5  | EIC                            | ![Structure Image](image3.png) | 41.9   | 0.14 |
| CR14 | Methyl linoleate               | ![Structure Image](image4.png) | 41.93  | 0.17 |
| CR16 | (+/−)-Isoborneol               | ![Structure Image](image5.png) | 86.98  | 0.05 |
| CR21 | Perlolyrine                    | ![Structure Image](image6.png) | 65.95  | 0.27 |
| CR28 | DIOP                           | ![Structure Image](image7.png) | 43.59  | 0.39 |
| ID   | Molecule name                  | Structure | OB (%) | DL  |
|------|-------------------------------|-----------|--------|-----|
| CR32 | ZINC03978781                  | ![Structure](image1) | 43.83  | 0.76|
| CR37 | Stigmasterol                  | ![Structure](image2) | 43.83  | 0.76|
| CR38 | Syringin                      | ![Structure](image3) | 14.64  | 0.32|
| CR43 | Tectorigenin                  | ![Structure](image4) | 28.41  | 0.27|
| CR45 | 7-Methoxy-2-methyl isoflavone | ![Structure](image5) | 42.56  | 0.2 |
| CR48 | Spinasterol                   | ![Structure](image6) | 42.98  | 0.76|
| CR49 | Atractylenolide II           | ![Structure](image7) | 47.5   | 0.15|
| CR52 | Atractylenolide III          | ![Structure](image8) | 68.11  | 0.17|
| ID  | Molecule name                      | Structure | OB (%) | DL  |
|-----|-----------------------------------|-----------|--------|-----|
| CR61| Frutinone A                       |           | 65.9   | 0.34|
| CR63| Luteolin                          |           | 36.16  | 0.25|
| CR67| Taraxerol                         |           | 38.4   | 0.77|
| CR69| Stigmast-7-enol                   |           | 37.42  | 0.75|
| CR70| Norharman                         |           | 18.88  | 0.08|
| CR73| 3-Beta-hydroxymethylenetanshiquinone |         | 32.16  | 0.41|
| CR75| HMF                               |           | 45.07  | 0.02|
| CR76| Methyl icosa-11,14-dienoate       |           | 39.67  | 0.23|
| ID   | Molecule name                     | Structure | OB (%) | DL   |
|------|-----------------------------------|-----------|--------|------|
| CR77 | Apigenin                          | ![Apigenin Structure](image1.png) | 23.06  | 0.21 |
| CR81 | (1R)-2,3,4,9-Tetrahydro-1H-pyrido[3,4-b]indol-2-ium-1-carboxylate | ![Tetrahydro Structure](image2.png) | 52.9   | 0.13 |
| CR97 | 5-Alpha-stigmastan-3,6-dione      | ![5-Alpha-stigmastan Structure](image3.png) | 33.12  | 0.79 |
| CR99 | 7-(Beta-xylosyl)cephalomannine_qt | ![7-(Beta-xylosyl)cephalomannine Structure](image4.png) | 38.33  | 0.29 |
| CR101| Codonopsine                       | ![Codonopsine Structure](image5.png) | 45.83  | 0.13 |
| CR103| Daturilin                         | ![Daturilin Structure](image6.png) | 50.37  | 0.77 |
| CR106| Glycitein                         | ![Glycitein Structure](image7.png) | 50.48  | 0.24 |
| ID   | Molecule name                                                                 | Structure | OB (%) | DL  |
|------|-------------------------------------------------------------------------------|-----------|--------|-----|
| CR112 | Spinoside A                                                                   | ![Spinoside A](image) | 39.97  | 0.4 |
| CR113 | (8S,9S,10R,13R,14S,17R)-17-[(E,2R,5S)-5-ethyl-6-methylhept-3-en-2-yl]-10,13-dimethyl-1,2,4,7,8,9,11,12,14,15,16,17-dodecahydrocyclopenta[a]phenanthren-3-one | ![Structure](image) | 45.4   | 0.76|
| CR117 | 11-Hydroxyrainikindine                                                         | ![11-Hydroxyrainikindine](image) | 40     | 0.66|
| CR123 | Ethyl-β-D-fructofuranoside                                                     | ![Ethyl-β-D-fructofuranoside](image) | 33.84  | 0.15|
| CR130 | Furanodiene                                                                    | ![Furanodiene](image) | 45.11  | 0.1 |
| CR132 | (+)-Beta-pinene                                                                | ![(+)-Beta-pinene](image) | 44.77  | 0.05|
| ID   | Molecule name                      | Structure | OB (%) | DL |
|------|------------------------------------|-----------|--------|----|
| ATR3 | Vanillic acid                      | ![Structure](structure1.png) | 35.47  | 0.04 |
| ATR6 | (-)-Alloaromadendrene              | ![Structure](structure2.png) | 54.04  | 0.1  |
| ATR12| Calarene                           | ![Structure](structure3.png) | 52.16  | 0.11 |
| ATR15| p-MCA                              | ![Structure](structure4.png) | 31     | 0.05 |
| ATR19| Marmesin                           | ![Structure](structure5.png) | 50.28  | 0.18 |
| ATR20| Majudin                            | ![Structure](structure6.png) | 42.21  | 0.13 |
| ATR23| (-)-Caryophyllene oxide            | ![Structure](structure7.png) | 32.67  | 0.13 |
| ID   | Molecule name                  | Structure | OB (%) | DL |
|------|--------------------------------|-----------|--------|----|
| ATR28| beta-Asarone                  | ![Structure](image) | 35.61  | 0.06 |
| ATR30| beta-Gurjunene                | ![Structure](image) | 51.36  | 0.1 |
| ATR35| beta-Cubebe none              | ![Structure](image) | 32.81  | 0.11|
| ATR40| 2′-0.1Methylisoliquiritigenin | ![Structure](image) | 75.86  | 0.17|
| ATR50| (+)-Ledene                    | ![Structure](image) | 51.84  | 0.1 |
| ATR53| (+)-alpha-Longipinene         | ![Structure](image) | 57.47  | 0.12|
Table 1: Continued.

| ID   | Molecule name          | Structure | OB (%) | DL |
|------|------------------------|-----------|--------|----|
| ATR54| 8-Isopentenyl-kaempferol | ![Structure](image1.png) | 38.04  | 0.39 |
| ATR55| Aminacrine             | ![Structure](image2.png) | 35     | 0.12 |
| ATR57| Aristolene             | ![Structure](image3.png) | 52.2   | 0.11 |
| ATR58| Aristolone             | ![Structure](image4.png) | 45.31  | 0.13 |
| ATR59| Azaron                 | ![Structure](image5.png) | 38.39  | 0.06 |
| ATR63| Bisasarcin             | ![Structure](image6.png) | 18.55  | 0.5  |
| ATR65| Calamendiol            | ![Structure](image7.png) | 61.13  | 0.11 |
| ID   | Molecule name                  | Structure | OB (%) | DL  |
|------|--------------------------------|-----------|--------|-----|
| ATR73| Isocalamendiol                 | ![Structure](image) | 57.63  | 0.11 |
| ATR78| Longicyclene                   | ![Structure](image) | 46.07  | 0.15 |
| ATR79| Murolan-3,9(11)-dien-10-peroxy | ![Structure](image) | 36.72  | 0.11 |
| ATR81| Patchoulene                    | ![Structure](image) | 49.06  | 0.11 |
| ATR84| Spathulenol                    | ![Structure](image) | 81.61  | 0.12 |
| ATR87| $\alpha$-Gurjunene             | ![Structure](image) | 52.57  | 0.1  |
| ATR88| $\alpha$-Panasinsene           | ![Structure](image) | 56.77  | 0.12 |
increase acetylcholine and butyrylcholine levels [59]. Surprisingly, asarones, accounting for more than 90% of ATR oil, have low DL values [60]. However, β-asarone (OB = 35.61, DL = 0.06) and azaron (OB = 38.89, DL = 0.06) could promote neuronal differentiation and reduce intracellular reactive oxygen species accumulation [18, 60]. P-coumaric acid (OB = 43.29, DL = 0.04) can relieve the neuroinflammatory responses induced by amyloid-beta peptide [61]. Vanillic acid exhibits marked antioxidant properties [62]. P-MCA (p-methoxycinnamic acid, OB = 31, DL = 0.05) exerts antihyperglycemic [63] and neuroprotective effects [64]. Bisasaricin (OB = 28.94, DL = 0.5) exhibits antibacterial and anti-inflammatory activity [65]. Above all, a total of 33 active ingredients, including some mentioned above, were preserved for the active ingredients of ATR.

### 3.4. Active Ingredients from CR

Among the 134 ingredients in CR, 36 ingredients were selected by ADME screening. EIC (linoleic acid, OB = 41.90, DL = 0.14) could reduce the development of atherosclerosis, having a strong association with AD and vascular dementia due to its antioxidant and anti-inflammatory effects [66, 67]. Stigmasterol (OB = 43.83, DL = 0.76) has beneficial effects on vascular function by improving the lipoprotein profile [68]. DIOP (OB = 43.59, DL = 0.39) exhibits anti-inflammatory activity based on its cholinesterase inhibitory activity [51]. Evidence proves that syringin (OB = 14.64, DL = 0.32) is an efficient treatment for AD [69]. Luteolin (OB = 36.16, DL = 0.25) depresses the proliferation and migration of vascular smooth muscle cells to prevent atherosclerosis [70]. Hydroxymethylfurfural (HMF; OB = 45.07, DL = 0.02) and apigenin (OB = 23.06, O

| ID   | Molecule name               | Structure | OB (%) | DL |
|------|-----------------------------|-----------|--------|----|
| ATR89| (1R,3aS,4R,6aS)-1,4-bis(3,4-dimethoxyphenyl)-1,3,3a,4,6,6a-hexahydrofuro[4,3-c]furan | ![Structure](image1.png) | 52.35  | 0.62 |
| ATR91| Cycloartenol                | ![Structure](image2.png) | 38.69  | 0.78 |
| ATR93| Kaempferol                  | ![Structure](image3.png) | 41.88  | 0.24 |
| ATR98| (-)-alpha-Cedrene           | ![Structure](image4.png) | 55.56  | 0.1  |
| ATR102| p-Coumaric acid            | ![Structure](image5.png) | 43.29  | 0.04 |
| Gene      | Target name                                                                 | UniProt ID |
|-----------|------------------------------------------------------------------------------|------------|
| ABCA1     | ATP-binding cassette subfamily A member 1                                      | O95477     |
| ABCB1     | Multidrug resistance protein 1                                                | P08183     |
| ACHE      | Acetylcholinesterase                                                          | P22303     |
| ADH1C     | Alcohol dehydrogenase 1C                                                      | P00326     |
| ADRA2C    | Alpha-2C adrenergic receptor                                                  | P18825     |
| ADRB2     | Beta-2 adrenergic receptor                                                    | P07550     |
| AHR       | Aryl hydrocarbon receptor                                                     | P35869     |
| ALOX5     | Arachidonate 5-lipoxygenase                                                   | P09917     |
| APP       | Amyloid beta A4 protein                                                       | P05067     |
| BACE1     | Beta-secretase 1                                                              | P56817     |
| BAX       | Apoptosis regulator BAX                                                       | Q07812     |
| BCHE      | Cholinesterase                                                               | P06276     |
| BCL2      | Apoptosis regulator Bcl-2                                                     | P10415     |
| CALM1     | Calmodulin                                                                   | P0DP23     |
| CASP3     | Caspase-3                                                                    | P42574     |
| CASP7     | Caspase-7                                                                    | P55210     |
| CDK5      | Cyclin-dependent kinase 5 (CDK5)                                              | Q00535     |
| CHRM1     | Muscarinic acetylcholine receptor M1                                           | P11229     |
| CHRM2     | Muscarinic acetylcholine receptor M2                                           | P08172     |
| CHRM3     | Muscarinic acetylcholine receptor M3                                           | P20309     |
| CHRNA2    | Neuronal acetylcholine receptor subunit alpha-2                               | Q15822     |
| CHRNA7    | Neuronal acetylcholine receptor protein, alpha-7 chain                         | P36544     |
| CHUK      | Inhibitor of NF-kappa-B kinase (IKK)                                          | O15111     |
| CYPIA2    | Cytochrome P450 1A2                                                           | P05177     |
| CYP2A6    | Cytochrome P450 2A6                                                           | P11509     |
| CYP2D6    | Cytochrome P450 2D6 (2D6)                                                     | P10635     |
| CYP3A4    | Cytochrome P450 3A4                                                           | P08684     |
| DIO1      | Type I iodothyronine deiodinase                                                | P49895     |
| ESR1      | Estrogen receptor                                                             | P03372     |
| F2        | Thrombin                                                                      | P00734     |
| F7        | Coagulation factor VII                                                        | P08709     |
| FLT3      | Tyrosine-protein kinase receptor FLT3                                         | P36888     |
| GABRA1    | Gamma-aminobutyric acid receptor subunit alpha-1                             | P14867     |
| GABRA2    | Gamma-aminobutyric-acid receptor alpha-2 subunit                             | P47869     |
| GABRA5    | Gamma-aminobutyric-acid receptor alpha-5 subunit                             | P31644     |
| GSK3B     | Glycogen synthase kinase-3 beta                                               | P49841     |
| HMOX1     | Heme oxygenase 1                                                              | P09601     |
| HTR2A     | 5-Hydroxytryptamine 2A receptor                                               | P28223     |
| IGF1R     | Insulin-like growth factor 1 receptor                                         | P08069     |
| IKBKBP    | Inhibitor of nuclear factor kappa-B kinase subunit beta                       | O14920     |
| IL10      | Interleukin-10                                                                | P22301     |
| IL1B      | Interleukin-1 beta                                                            | P01584     |
| INS       | Insulin                                                                       | P01308     |
| INSR      | Insulin receptor                                                              | P06213     |
| KDR       | Vascular endothelial growth factor receptor 2                                 | P35968     |
| MAOA       | Amine oxidase [flavin-containing] A                                            | P21397     |
| MAOB       | Amine oxidase [flavin-containing] B                                            | P27338     |
| MAPK10     | Mitogen-activated protein kinase 10                                            | P53779     |
| MET        | Hepatocyte growth factor receptor                                              | P08581     |
| NOS3       | Nitric-oxide synthase, endothelial                                            | P29474     |
| NR1H2      | Nuclear receptor subfamily 1 group 1 member 2                                 | O75469     |
| PLAU       | Urokinase-type plasminogen activator                                          | P00749     |
| PPARG      | Peroxisome proliferator activated receptor gamma                              | P37231     |
| PTGES      | Prostaglandin E synthase                                                      | O14684     |
| PGTG1      | Prostaglandin G/H synthase 1                                                  | P23219     |
| PGTG2      | Prostaglandin G/H synthase 2                                                  | P35354     |
| RXRA       | Retinoic acid receptor RXR-alpha                                              | P19793     |
| RXRG       | Retinoic acid receptor RXR-gamma                                              | P48443     |
| SLC2A4     | Solute carrier family 2, facilitated glucose transporter member 4             | P14672     |
| SOAT1      | Acyl-cholesterol acyltransferase 1                                            | P36510     |
| TNF        | Tumor necrosis factor                                                        | P01375     |
| VEGFA      | Vascular endothelial growth factor A                                          | P15692     |
were shared by two herbs. HMF was selected because of its therapeutic effect on AD, and it provided effective protection against cognitive impairment via the activation of NMDA receptor signaling [71]. Oral treatment with apigenin plays an important role in the neuroprotective effects by relieving Aβ deposition and improving antioxidative activity [72]. All these components are listed as potential active ingredients for CR, and the details of the 68 ingredients of the two herbs are shown in Table 1.

3.5. Target Proteins of ATR and CR. Identifying the targets of candidate ingredients based on experimental approaches is complex and time-consuming. Recently, an integrated in silico approach was used to identify the corresponding targets for the active ingredients of ATR and CR. In our study, predictive models, including TCMSP, SEA, and STITCH, were used to search for the targets of ATR and CR, and 218 and 229 proteins were identified as targets of ATR and CR, respectively (Figure S1). Next, these targets were sent to TTD, CTD, OMIM, DrugBank, and PharmGKB to determine whether they were related to AD. In total, for ATR and CR, by target fishing, 53 out of 68 active ingredients (15 active ingredients have no AD-related targets) were valid for binding with 62 AD-related target proteins, and the details of 62 AD-related targets are listed in Table 2.

In ATR, 52 AD-related target proteins were validated to bind with 30 active ingredients. For example, the majority of ingredients in ATR, such as ATR6, ATR19, ATR30, ATR54, and ATR93, showed strong activation of the neurotransmitter receptors CHRM1, CHRM2, CHRM3, GABRA1, and GABRA2, modulating neurotransmission dysfunction in the AD brain [73, 74]. Intriguingly, ATR93 also interacted with CYP1A2 and CYP3A, which are major drug-metabolizing enzymes and can prevent cholinergic symptoms such as nausea and vomiting in patients with AD [75, 76]. Furthermore, three active ingredients, ATR3, ATR15, and ATR102, were shown to interact with MAOA, MAOB, and NOS3, which are related to inflammation and play a role in the pathogenesis and symptoms of mental disorders [77, 78].

In CR, 52 AD-related target proteins were validated to bind with 25 active ingredients, including ACHE, APP, BCHE, BCL2, BAX, CASP3, and CASP7, and are implicated in cell apoptosis and vascular and nervous system diseases. For instance, ACHE and BCHE are the major therapeutic targets of AD that improve β-amyloid plaques and cholinergic function in AD patients [79, 80]. Remarkably, luteolin (CR63) and glycitin (CR106) interact with APP, the central pathological target of AD [81]. Indeed, luteolin can prevent the formation of β-amyloid protein by inhibiting ppGalNAc-T activity [82]. Glycitin can also suppress β-amyloid deposition and has antioxidative activity [83]. Additionally, our results found that the apoptosis-related genes BCL2, BAX, CASP3, and CASP7, associated with AD, were potential targets of CR63 and CR77 [84, 85]. All these findings may explain why CR can treat AD with ATR.

3.6. Synergistic Mechanisms of ATR and CR

3.6.1. GO Enrichment Analysis for Targets. To clarify the synergistic mechanism of ATR and CR, we performed GO enrichment analysis on ClueGo. In Figure 3, the top 10 significantly enriched terms are ranked in the biological process (BP), molecular function (MF), and cellular component (CC) categories (P < 0.05; P values are corrected using the Benjamini–Hochberg procedure), including regulation of neurotransmitter levels, reactive oxygen species biosynthesis, and regulation of blood vessel diameter in the pre- and prosynaptic membrane, nuclear envelope lumen, and plasma membrane, to exert anti-AD potential. Particularly, the neurotransmitter imbalance caused by dysfunction of cholinergic neurons contributed to the cognitive deficits associated with AD [86, 87]. Studies have demonstrated that the degeneration of cholinergic neurons gives rise to memory loss in AD patients [88]. In the early stage of AD, the generation of reactive oxygen species is the prominent feature [89]. Oxidative stress is reported to be involved in the disruption of Aβ clearance in the brain, which contributes to the accumulation of Aβ [90]. Abnormal blood vessel diameters reduce blood flow and lead to an inadequate blood supply in the cortex [91, 92]. Undoubtedly, these BP terms are all closely associated with AD. For instance, synaptic dysfunction caused by disruption of neurotransmitter transmission and increased formation of reactive oxygen species in neural cells are the main pathogenic mechanisms of AD [93, 94]. Additionally, the lower cerebral blood flow caused by blood vessel size is a major risk factor for AD [95]. The results of GO enrichment analysis indicated that the combination of ATR and CR mainly by neuroprotective, antioxidative, and antiatherosclerotic effects.

3.6.2. Compound-Target Network Analysis. In Figure 4, 53 active ingredients and 62 AD-related targets were used to construct the C-T network. There were 373 component-target associations between 53 active ingredients and 62 AD-related targets. The average number of targets per ingredient was 6.0, and the mean degree of ingredients per target was 7.0, indicating complicated and complementary relationships between ingredients and targets. In total, more than half of the targets (67.7%), such as ACHE, BAX, BCL2, GABRA1, NOS3, and PTGS1, were synergistically regulated by various components of ATR and CR. However, several targets (32.3%), such as APP, CYP1A2, IL10, MAPK10, and VEGFA, were regulated by each ingredient of ATR and CR. However, this Chinese formula focused on the targets associated with AD, such as neurotransmitter release and interaction between neuroactive ligands and receptors (ADRA2C, APP, CASP3, CHRM1, CHRNA7, GABRA1, and CYP2D6). It also modulated other targets involved in the pathogenesis of AD, including oxidative stress, atherosclerosis, and inflammation (BCL2, IKKB, MAPK10, NOS3, TNF, PTGS2, and GSK3B). As shown in Figure 4, kaempferol (degree = 33) has the highest degree, followed by apigenin (degree = 30), stigmasterol (degree = 16), 7-
methoxy-2-methyl isoflavone (degree = 16), luteolin (degree = 15), and 8-isopentenyl-kaempferol (degree = 12), indicating that these ingredients play crucial roles in the treatment of AD. For instance, kaempferol is reported to promote the level of antioxidants and alleviate neuroinflammation in the hippocampus to protect against cognitive deficits in AD [96]. Apigenin can suppress the expression of cytokines and the production of nitric oxide, indicating a potential drug for AD [97]. Stigmasterol could decrease β-secretase activity and BACE1 internalization, which is related to APP β-secretase cleavage [98]. Luteolin also has several biological functions, including anti-neuroinflammatory and antioxidant activities, which are beneficial for the prevention of AD [99].
3.7. Target-Pathway Network Analysis. All of the targets interacting with the active ingredients were mapped onto the 67 KEGG pathways, and the T-P network was generated. As shown in Figure 5, the PI3K-Akt signaling pathway shows the highest number of target connections (degree = 15), followed by the MAPK signaling pathway (degree = 13), neuroactive ligand-receptor interaction (degree = 12), fluid shear stress and atherosclerosis (degree = 11), nonalcoholic fatty liver disease (NAFLD) (degree = 11), and Ras signaling pathway (degree = 11). These pathways were related to neurofibromatosis, antiatherosclerosis, antioxidation, and anti-inflammation. The PI3K-Akt signaling pathway is involved in atherosclerosis by blocking blood flow and inhibiting the migration of fibroblasts [9] and can suppress neuronal cell death and improve tau hyperphosphorylation by BCL2, GSK3B, and IGFr1 [100–102]. MAPK can directly affect AD, contributing to neuroinflammation and acting in some processes, such as excitotoxicity, synaptic plasticity, and tau phosphorylation [103, 104]. The neuroactive ligand-receptor interaction pathway has been applied in the analysis of neurodegenerative disorders [105]. As expected, our results showed that the combined herbs modulated neuroactive receptors such as CHRM1, CHRM2, GABRA1, GABRA2, etc., which are involved in this pathway and provide therapeutic benefits for AD patients. The fluid shear stress and atherosclerosis pathway can regulate the size of cerebral blood vessels [14] as well as plaque formation, which will lead to vascular dementia [106].

The compressed pathway was constructed to further explore the details of the synergetic mechanism of ATR and CR in the treatment of AD. As shown in Figure 6, ATR could act on the KDR gene, which is one of the key molecules related to angiogenesis and a strong risk factor for atherosclerosis [107], while CR can act on its ligand VEGFA. This result indicated that ATR and CR could treat AD through complementary effects on atherosclerosis, which is a pathogenic factor of AD. Additionally, CR can act on the genes of the upstream pathway, such as IL1B and TNF, which are proinflammatory mediators [17, 26], while ATR can act on downstream genes, such as CALM1 and NOS3, which are associated with learning and memory and anti-inflammatory processes. Consequently, these results showed that ATR and CR have a synergetic effect on antiatherosclerosis processes, anti-inflammatory processes, learning, and memory in a comprehensive pathway.

3.8. Contribution Score Analysis. To further explore the synergetic mechanism of ATR and CR, we performed CS analysis following a previously described method [16]. The CS value of each active ingredient is listed in Figure 7 and Table S2. The top 6 ingredients were kaempferol (SCP93), apigenin (DS77), 7-methoxy-2-methyl isoflavone (DS45), stigmasterol (DS37), syringin (DS38), and HMF (DS75) with a CS sum of 45.20%; these were active ingredients from both ATR and CR, especially apigenin and HMF, the shared ingredients. For instance, syringin exhibits anti-inflammatory activity and enhances the synthesis of acetylcholine in the hippocampus [69, 108]. HMF can mitigate the impairment of cognition and memory function induced by Aβ, inhibit beta-secretase activity, and increase antioxidative enzyme activities [109]. Moreover, 29 ingredients could contribute to the effects of this formula on AD with a CS sum of 90.14%, and ATR and CR accounted for 43.13% and 56.87%, respectively (Figure S2). Our data fully explained why the combination of ATR and CR could generate synergistic and combinatorial effects on AD.

3.9. Compound-Target-Organ Network Analysis. In TCM, the human body is considered an organic whole, and the organs are complementary to each other. The curative effect of diseases is usually exerted by multorgan cooperation. Therefore, to better understand the synergistic mechanism of ATR and CR treatment for AD at the organ level, we generated a compound-target-organ network. The mRNA expression results were obtained from the BioGPS database, and the target was positioned in the organization where it had the highest expression patterns. As shown in Figure 8, the targets were widely distributed on the brain, heart, kidney, liver, lung, whole blood, and other tissues. Particularly, the top three distributed organs of targets are whole blood, brain, and muscle. Undoubtedly, the key pathogenic mechanism of AD is the degradation of the brain, and the main targets located in the brain are neurotransmitter receptors such as CHRM3, GABRA1, and GABRA2. As a pivot between each tissue, whole blood improves the coordination of organs to provide a positive effect on AD. Additionally, the gene expression pattern in blood indicates that biological pathways associated with oxidative stress, inflammation, apoptosis, and immune activation are involved in AD [110, 111]. The dysfunction of smooth muscle, which is responsible for regulating blood vessels, could lead to AD complications, such as atherosclerosis and other vascular diseases [22]. In total, these data indicated that the combination of ATR and CR effectively prevents AD not only in the brain but also in other organs. This evidence indicated that ATR and CR have synergistic effects at the organ level as well.

4. Discussion

AD is a neurodegenerative disease characterized by synapse loss [112], neurofibrillary tangles in the brain [113], and extracellular deposits of β-amyloid peptides (Aβ) [114]. The pathogenesis of AD also involves inflammatory and oxidative stress in neuronal cells [4, 6], coronary heart disease [115, 116], and cardiovascular disease [66]. Our results demonstrated that the combination of ATR and CR can effectively relieve inflammation, oxidative stress, atherosclerosis development, and nervous system damage.

First, 68 active ingredients were chosen following ADME screening of 235 ingredients. In particular, apigenin and HMF were both shared ingredients of the two herbs that had not passed the screening criteria, but they were preserved because of their neuroprotective effects [71, 72]. Moreover, the volatile oils of ATR, such as β-asarone and γ-asarone, could improve learning and memory, mitigate the deposits
of Aβ, and prevent oxidative stress-induced apoptosis [17, 56]. The sterols of CR, such as stigmasterol and spinasterol, could attenuate the risk of coronary heart disease and inhibit the development of atherosclerosis [117, 118]. Consequently, these results suggest that ATR and CR could produce a combinatorial effect on AD.

Second, according to the CS calculation of each ingredient, ATR and CR can contribute to the effects for AD with a sum of 43.13% and 56.87%, respectively, and 29 ingredients of the two herbs can contribute a sum of 90.51%. This result suggested that the therapeutic effect of this formula was derived from ATR and CR, not a single herb nor a handful of ingredients. More remarkably, the top ingredients of the CS in ATR, such as kaempferol, not only could inhibit inflammation [119] but also attenuate glutamate-induced oxidative stress [56]. However, for CR, the top ingredients, such as stigmasterol and syringin, showed strong efficacy and treatment of atherosclerosis [68, 120]. These results suggested that ATR provided neuroprotective effects such as antioxidation and anti-inflammation, and subsequently, CR affected AD by inhibiting the development of atherosclerosis. These data proved evidence that the combination of ATR and CR had synergistic effects on AD.

Third, the GO enrichment analysis showed that the top ten BP terms were all closely related to neurotransmitter transduction, antioxidant effects, and vasodilatation, which are associated with AD in the synaptic membrane and plasma membrane. These data implied that these two herbs displayed an excellent ability to treat AD and its accompanying symptoms. The results of C-T network analysis showed that CR could act on AD-related genes, including APP [120], IL10 [56], CASP7 [68], and SOAT1 [121], and ATR could act on AD-related genes, including CYP2D6 [122], CHRNA2 [123], and ALOX5 [124], indicating that these two herbs displayed a complementary ability to treat AD. Several AD-related genes, such as ACHE [125], PTGS2 [126], IKBKB [127], GABAR1 [128], CHRM2/3 [129], etc., were synergistically regulated by various ingredients of ATR and CR, demonstrating that their combination enhances the ability to treat AD. Furthermore, based on the T-P network analysis, ATR and CR treatment of AD mainly depends on the PI3K signaling pathway, MAPK signaling pathway, neuroactive ligand–receptor interaction, and fluid shear stress and atherosclerosis, which regulate the nervous and vascular systems. Additionally, this combination activates nonalcoholic fatty liver disease, the Ras signaling pathway,

Figure 5: Target-pathway networks of ATR and CR. The green nodes are the targets of ATR and CR, while the orange nodes represent the pathways.
and human cytomegalovirus infection to relieve the AD-associated symptoms such as liver inflammation and cytomegalovirus infection to benefit patients with AD [6, 68].

Finally, the integrated pathway analysis showed that ATR and CR could act on genes that were involved in various signaling pathways. In particular, KDR was a unique
target of ATR, while CR could act on its ligand VEGFA, suggesting a complementary effect. Subsequently, CR could act on IGF1R, MET, IL1B, and CHUK alone. These results provided powerful evidence that each of them could act on genes to activate the complementary signaling pathway for treating AD. The C-T-O network analysis also showed that the targets of ATR and CR had a wide distribution, but the top organs were whole blood, brain, and muscle, which are strongly related to antioxidant, anti-inflammatory, anti-atherosclerotic, and neuroprotective activities. Consequently, the synergistic effect of ATR and CR involved not only the molecular and pathway levels but also the organ level.

In summary, this novel systems pharmacology model provided a powerful method to explore the therapeutic mechanism of TCM, which has multiple ingredients and targets. A combination of compatible remedies for nervous system diseases can relieve the symptoms of cardiovascular diseases and oxidative stress, which are the pathogenic mechanisms of AD at different levels. Given the above factors, a mixture of ATR and CR possesses synergism on AD and this study provides a solid theoretical basis for the effective prevention and treatment of the pain caused by AD. However, this conclusion mainly depends on model prediction and literature analysis, and more experiments should be performed to verify our conclusion.

4.1. Contribution to the Field Statements. AD is a progressive neurodegenerative disorder caused by extensive synapse loss, the formation of intracellular neurofibrillary tangles, extracellular deposits of β-amyloid peptides (Aβ), and inflammatory and oxidative stress in neurons. It has been reported that more than 35 million people suffer from AD. In this work, we evaluated the systems pharmacology-based strategy to explore the pathogenic mechanism of AD at the molecular pathway and organ levels. We first clarified the synergistic and complementary mechanisms of the Chinese
herbs ATR and CR in the treatment of AD without laborious and time-consuming experiments. In view of the rich natural compounds in Chinese herbs, this combination will provide an effective treatment for AD.

**Data Availability**

The data used to support the findings of this study are available from the corresponding author upon request.

**Conflicts of Interest**

The authors declare that they have no conflicts of interest regarding the publication of this paper.

**Authors’ Contributions**

ZS provided the concept, and SL designed the study and performed the experiments. YL, WM, and HL participated in data analysis, and SL and CH contributed to writing, revising, and proofreading the manuscript. All authors read and approved the final manuscript.

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**Supplementary Materials**

*Figure S1.* The number of targets of ATR (red), CR (blue), and Alzheimer’s disease (green) obtained from databases. *Figure S2.* CS and accumulative CS of the active ingredients in ATR or CR. *Table S1.* Detailed information on the ingredients in ATR and CR. *Table S2.* The value of active ingredients in ATR and CR. (Supplementary Materials)

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