The HER-2 as a Target Gene of Curcumin to Protect Hepatocytes Against the Arsenic-induced Carcinoma in Mice

Mahdi Ahadi¹, Vahid Naseh¹, Masoud Salehipour*²

1. Dept. of Biology, Faculty of Biological Sciences, Parand Branch, Islamic Azad University, Parand, Iran

KEYWORDS

HER-2
Hepatocellular Carcinoma
Curcumin
Arsenic

ABSTRACT

Background & objective: The HER-2 gene is an important oncprotein overexpressed in many types of cancers. The current study hypothesized that curcumin downregulates HER-2 and inhibits the signal transduction pathway of PI3K/Akt, MAPK, and activation of NFκB, which could be useful to treat overexpressed-HER-2 hepatocellular carcinoma (HCC).

Methods: In the current study, 40 male NMRI (Naval Medical Research Institute) mice were divided into 4 groups of 10 as follow: Group1 (control group) only received 5 mL/kg corn oil, group 2 (poisoned group) received 30 mg/L arsenic (As₂O₃) dissolved in water, group 3 (curcumin treated), and group 4 (curcumin and arsenic treated) received 10 to 20 mg/5 mL/kg for 60 days. Once experimental period was completed, liver samples were collected. The analysis of the gene expression was performed by real-time polymerase chain reaction (PCR) technique.

Results: Gene expression analysis showed that curcumin had significantly downregulated the activity of HER-2, in poisoned mice.

Conclusion: According to the current study results, it could be concluded that curcumin has the inhibitory potential toward HER-2-overexpressed HCC.

Introduction

Hepatocellular carcinoma (HCC) is defined as a primary liver malignancy, which principally occurs in patients affected by the chronic hepatic disease and cirrhosis (1, 2). Tumors include local expansion, intra hepatic spread, and distant metastases (2). In addition, arsenic in the environment is etiologically associated with tumor development in a variety of tissue, including skin, bladder, lung, liver, and prostate gland (3, 4). The most important source of environmental arsenic exposure in most populations is drinking water, in which inorganic forms of arsenic (trivalent arsenite and pentavalent arsenate) are predominant (5). Curcumin (diferuloylmethane) is naturally a yellow pigment obtained from the rhizome of the plant Curcuma longa L. The powdered rhizome of this plant, called turmeric, is usually utilized to prepare curries. Curcumin, as a polyphenol with a diarylheptanoid structure containing α, β-unsaturated ketones, is considered as the major active part of turmeric (6, 7). In addition to its broad spectrum of pharmacological activities, the anticancer properties of curcumin (8, 9), and its capability to downregulate epidermal growth factor receptor (EGFR) and HER-2 oncoproteins (9, 10) are reported in many studies. Also, curcumin can have an impact on several pathways such as PI3K/Akt and mitogen-activated protein kinases (MAPK) (11, 12). Additionally, curcumin is a potent anti-inflammatory compound (13). Recently, several studies indicated that the deregulated inflammatory pathways play a pivotal role in a large number of chronic diseases, including cancer (14). The leading mechanism by which...
Curcumin serves as an anti-chronic inflammatory compound, and drives cancer initiation and progression via increasing production of pro-inflammatory mediators such as cytokines, chemokines, reactive oxygen species (ROS), overexpression of cyclooxygenase (COX-2), matrix metalloproteinase (MMPs), oncogenes, intracellular signaling pathway mediators, transcription factors including signal transducer and activator of transcription 3 (STAT3), nuclear factor κB (NF-κB), activator protein 1 (AP1), and protein kinase B (AKT), that drive tumor cell proliferation, transformation, invasion, metastasis, and angiogenesis (15). Regarding the possible properties provided for curcumin in this introductory comment, it can be assumed as a beneficial agent in the treatment of HER-2 positive cancers (16). Thus, the current study aimed at exploring the potential application(s) of curcumin on the HER-2 expression in arsenic-poisoned mice.

**Material and Methods**

**Induction of HCC and curcumin therapy**

In the current study, 40 male NMRI (Naval Medical Research Institute) mice weighing 30 to 35 gr were recruited. After being matched according to the body weight, the mice were divided into 4 different groups of 10: Group 1, control mice received 5 mL/kg/day corn oil (Sigma-Aldrich) for 60 days; group 2, (poisoned mice) received 5 mL/kg/day corn oil and 30mg/L arsenic (As₂O₃) for 60 days; group 3, treated mice received 5 mL/kg/day corn oil and 10 to 20 mg/5 mL/kg/day curcumin (Sigma-Aldrich) for 60 days; group 4, (treated mice) received 5 mL/kg corn oil and 10 to 20 mg/5 mL/kg/day curcumin and arsenic for 60 days. Also, 5 mL of the above-mentioned components were given to the animals via gavage syringe. Subsequently, mice were housed in the cages with free access to water and standard food. All of the animal handling processes were performed according to the guidelines of Iranian Animal Ethics Society, School of Science, Islamic Azad University, Parand Branch. At the end of the 60-day treatments, liver samples were collected, and the expression of HER-2 was analyzed by real-time polymerase chain reaction (PCR).

**RNA isolation**

To isolate RNA from tissue, mice were sacrificed; the liver removed under aseptic situations and immediately frozen in liquid nitrogen. Liver tissue samples were homogenized in TRIzol™ reagent (Sigma-Aldrich) using Mixer 301, and subjected to RNA extraction. The purity extracted RNA was determined by electrophoresis on an ethidium bromide pretreated agarose gel along with measuring absorption at 260/280 nm using the spectrophotometric method.

**Synthesis of cDNA and quantitative real-time PCR**

Five micrograms of RNA were reversely transcribed using reverse transcriptase enzyme for 1 hour at 37°C to synthesize cDNA. Quantitative changes of mRNA were assessed by quantitative real-time PCR (Bioneer, Exicycler™ 96 Korea) using SYBR green detection system contained SYBR green PCR Master Mix (Thermo Scientific). The sequences of the used primer are demonstrated in Table 1. The β-actin was used as a housekeeping gene, and each sample was normalized on the basis of its β-actin content. The mRNA encoding target genes were analyzed by employing real-time PCR method and were normalized by β-actin mRNA (as the housekeeping gene), using the 2-ΔΔCt formula.

**Histological staining of tissue**

Liver tissues were collected and fixed in 10% formalin after sacrifice. They were cut transversely or longitudinally to obtain ventricular sections or 4-chamber cross sections, respectively; then, embedded in paraffin and cut into 4-μm thick slices for morphological and pathological evaluations. Tissue sections were stained with hematoxylin and eosin (H&E), examined using a light microscope equipped with a charge coupled device (CCD) camera (KE2500, Nikon, Tokyo, Japan), and diagnosed by a clinical pathologist.

**Statistical Analysis**

Results were presented as mean± standard deviation (SD). The results were statistically analyzed using a one-way analysis of variance.
The HER-2 as a Target Gene of Curcumin…

(ANOVA), followed by Tukey multiple comparisons for all tests by SPSS software version 18 (SPSS Inc., Chicago, IL, USA). P ≤0.05 was considered statistically significant.

Results

The current study enrolled 40 mice, and HCC was induced in them by arsenic treatment. The mRNA levels of HER-2 in the liver of normal control group were regarded as 100% expression, and the expression in the other groups were calculated accordingly. The current study results showed that the expression of HER-2 was 6.6±0.057 in group 2, (received only arsenic, P<0.001), 0.18±0.23 in group 3 (curcumin treated, P<0.01), and 0.56±0.13 in group 4 (curcumin and arsenic treated, P<0.05), while expression of HER-2 in the control and β-actin control groups were 1.06±0.94 and 1.09±0.94, respectively (Figure 1).

Results of the current study showed that curcumin effectively downregulated the HER-2 expression in comparison with curcumin and arsenic treated group. Sections of liver tissue were stained with H&E illustrated that livers of the mice treated with arsenic developed HCC, and treatment of this group with curcumin showed that curcumin alone had the property to cure HCC. Also in the group that received arsenic and curcumin, results suggested that curcumin had beneficial effects to return the cancer cells back to normal status (Figure 2).

Discussion

Primary liver cancer, principally hepatocellular carcinoma (HCC), remains the 8th largest cause of cancer in females and the 5th largest in males (17, 18). HCC is the 3rd cause of cancer-related deaths worldwide (19, 20). Also, HCC is a malignancy with acutely ominous prospects and a 5-year survival rate reported below 9% (21). In addition, the role of HER-2 was discussed in different studies. Similar to HER-1, the HER-2 molecule is a member of a larger family of receptor tyrosine kinase overexpressed in various human malignancies such as liver, breast, ovarian, endometrial, cervical, lung, stomach, colon, mesenchymal, esophagus, bladder, and kidney cancers (22-26).

As a result, curcumin is an intelligent compound suppressing the expression of HER-2 and might be a useful option for cancer therapy. The current study showed that curcumin can be considered as an effective agent that downregulates the HER-2 expression in HCC. Consistent with the current study results, Hung et al., showed that the expression of HER-2 significantly increased in HCC (27). Chan et al., found that turmeric (Curcuma longa) inhibits the secretion of tumor necrosis factor (TNFα) (28). In several other studies, the antioxidant, antitoxic, anti-inflammatory, and anticancer properties of curcumin are reported (29). Curcumin has powerful antioxidant effects to neutralize free radicals and the ability to collect ROS (30). Cao et al., reported that curcumin inhibited the growth of atypical glandular cells of undetermined significance (AGS cells) and induced apoptosis through the activation of Ras/ERK signaling pathway. They suggested that curcumin can be a potential agent to treat gastric carcinoma (31).
Fig 2. Photomicrographs of liver sections stained with hematoxylin and eosin (bar, 100 μm). (A) Control mice, showing normal hepatocytes architecture; (B) Liver of HCC mice treated with arsenic, showing HCC development; (C) Liver of HCC mice treated with curcumin, shows approximately normal architecture; (D) liver of HCC mice received arsenic and curcumin, pointer shows the helpful effect of curcumin on cancerous cells that are returning to normal state.

Hu et al., reported that curcumin, with antioxidant and anti-inflammatory effects, can be helpful in cancer therapy (32). In agreement with the results of the current study, Wen Lai et al., demonstrated that curcumin could decrease the expression of HER-2 in patients with breast cancer (33). Soo Chun et al., showed that curcumin prevented tumor progression via inhibition of ERK1/2, NFKB, and COX-2 (34). In parallel with the results of the current study, Reddy et al., showed that curcumin can reduce the progression of colon cancer via reducing the expression of EGFR, ErbB-2, ErbB-3, and / or insulin-like growth factor 1 receptor (IGF-1R) genes (35). Several scientific reports stated the carcinogenic effect of arsenic and its impact on HER-2. For instance, Chuan shu et al., reported that arsenic played a fundamental role to activate signaling mediators including miR-199/148 / ERBB2 / PKM2 / NF-kB to increase the expression of hypoxia-inducible factor 1 (HIF-1), interleukin (IL)-8, and finally increased angiogenesis and contributed to the processes of tumor progression and development (36). Simeonova et al., showed that arsenic can affect the activation of the MAPK/ERK pathway group of molecules, including nuclear transcription factor AP-1, c-fos, c-jun, and c-myc (37). Flora et al., claimed that arsenic can increase the production of ROS. ROS itself causes altered signaling pathway and regulates the expression of transcription factors. His study also pointed to the role of arsenic in the activation of MAPK/ERK pathway cascade through the activation of signaling and phosphorylation of EGFR/MEK, EGFR/Ras/MEK, Src/EGFR performed (38).
Findings of the current study indicated that arsenic could lead to increased expression of HER-2 and other oncoproteins; and curcumin, as a chemopreventive and therapeutic agent in HCC, has helpful properties including antioxidant and anti-inflammatory impacts, and the capability to regulate a variety of signaling mechanisms. In vitro and in vivo preclinical models indicate that curcumin, along with different curcuminoids, are potential curative agents for HCC.

To draw a conclusion, considering the results of the current study, it could be suggested that curcumin has the complimentary potency to be developed as an antitumor operant to treat HCC. Additionally, maybe curcumin suppresses these types of hepatic tumors by balancing (up/down-regulating) the expression of HER-2 in arsenic-poisoned mice. Maybe enrichment in other antioxidant and antitumor agents makes curcumin a possible compound to protect hepatic cells from arsenic-poisoned damage.

Acknowledgements
Authors wish to thank all of the people who warmly cooperated in this research.

Conflict of Interest
The authors had no conflict of interest.

References
1. Ma Y, Wang J, Liu L, Zhu H, Chen X, Pan S, et al. Genistein potentiates the effect of arsenic trioxide against human hepatocellular carcinoma: role of Akt and nuclear factor-xB. Cancer letters. 2011;301(1):75-84.

2. Alison MR. Liver stem cells. Stem cell reviews. 2005;1(3):253-60.

3. Liu J, Waalkes MP. Liver is a target of arsenic carcinogenesis. Toxicological Sciences. 2008;105(1):24-32.

4. Waalkes MP, Ward JM, Liu J, Diwan BA. Transplacental carcinogenicity of inorganic arsenic in the drinking water induction of hepatic, ovarian, pulmonary, and adrenal tumors in mice. Toxicology and applied pharmacology. 2003;186(1):7-17.

5. Smith AH, Lingas EO, Rahman M. Contamination of drinking-water by arsenic in Bangladesh: a public health emergency. Bulletin of the World Health Organization. 2000;78(9):1093-103.

6. Aggarwal BB, Kumar A, Bharti AC. Anticancer potential of curcumin: preclinical and clinical studies. Anticancer res. 2003;23(1A):363-98.

7. Sharma R, Gescher A, Steward W. Curcumin: the story so far. European journal of cancer. 2005;41(13):1955-68.

8. Chauhan D. Chemotherapeutic potential of curcumin for colorectal cancer. Current pharmaceutical design. 2002;8(19):1695-706.

9. Chen A, Xu J, Johnson A. Curcumin inhibits human colon cancer cell growth by suppressing gene expression of epidermal growth factor receptor through reducing the activity of the transcription factor Egr-1. Oncogene. 2006;25(2):278-87.

10. Hong R-L, Spohn WH, Hung M-C. Curcumin inhibits tyrosine kinase activity of p185neu and also depletes p185neu. Clinical Cancer Research. 1999;5(7):1884-91.

11. Hussain A, Al-Rasheed M, Manogaran P, Al-Hussein K, Platanias L, Al Kuraya K, et al. Curcumin induces apoptosis via inhibition of PI3′-kinase/AKT pathway in acute T cell leukemias. Apoptosis. 2006;11(2):245-54.

12. Camacho-Barquero L, Villegas I, Sánchez-Calvo JM, Talero E, Sánchez-Fidalgo S, Motilva V, et al. Curcumin, a Curcuma longa constituent, acts on MAPK p38 pathway modulating COX-2 and iNOS expression in chronic experimental colitis. International immunopharmacology. 2007;7(3):333-42.

13. Cho JA, Park E. Curcumin utilizes the anti-inflammatory response pathway to protect the intestine against bacterial invasion. Nutrition research and practice. 2015;9(2):117-22.

14. Sethi G, Shanmugam MK, Ramachandran L, Kumar AP, Tergaonkar V. Multifaceted link between cancer and...
inflammation. Bioscience reports. 2012;32(1):1-15.

15. Shanmugam MK, Rane G, Kanchi MM, Arfuso F, Chinnathambi A, Zayed M, et al. The multifaceted role of curcumin in cancer prevention and treatment. Molecules. 2015;20(2):2728-69.

16. Bar-Sela G, Epelbaum R, Schaffer M. Curcumin as an anti-cancer agent: review of the gap between basic and clinical applications. Current medicinal chemistry. 2010;17(3):190-7.

17. Garcia M, Jemal A, Ward E, Center M, Hao Y, Siegel R, et al. Global cancer facts & figures 2007. Atlanta, GA: American cancer society. 2007;1(3):52.

18. Jemal A, Center MM, DeSantis C, Ward EM. Global patterns of cancer incidence and mortality rates and trends. Cancer Epidemiology Biomarkers & Prevention. 2010;19(8):1893-907.

19. Parkin DM, Bray F, Ferlay J, Pisani P. Global cancer statistics, 2002. CA: a cancer journal for clinicians. 2005;55(2):74-108.

20. Schütte K, Bornschein J, Malfertheiner P. Hepatocellular carcinoma–epidemiological trends and risk factors. Digestive Diseases. 2009;27(2):80-92.

21. Sherman M, editor Hepatocellular carcinoma: epidemiology, risk factors, and screening. Seminars in liver disease; 2005: Copyright© 2005 by Thieme Medical Publishers, Inc., 333 Seventh Avenue, New York, NY 10001, USA.

22. Ochs A-M, Wong L, Kakani V, Neerukonda S, Gorske J, Rao A, et al. Expression of vascular endothelial growth factor and HER2/neu in stage II colon cancer and correlation with survival. Clinical colorectal cancer. 2004;4(4):262-7.

23. Romond E, Suman V, Jeong J, Sledge G, Geyer C, Martino S, et al. Abstract S5-5: Trastuzumab plus adjuvant chemotherapy for HER2-positive breast cancer: Final planned joint analysis of overall survival (OS) from NSABP B-31 and NCCTG N9831. Cancer Research. 2012;72(24 Supplement):S5--S.

24. Krüger S, Weitsch G, Böttner H, Matthiessen A, Böhmer T, Marquardt T, et al. HER2 overexpression in muscle-invasive urothelial carcinoma of the bladder: Prognostic implications. International journal of cancer. 2002;102(5):514-8.

25. Xian Z, Zhang S, Cong W, Wu W, Wu M. Overexpression/amplification of HER-2/neu is uncommon in hepatocellular carcinoma. Journal of clinical pathology. 2005;58(5):500-3.

26. Wang W-H, Chiang I, Ding K, Chung J-G, Lin W-J, Lin S-S, et al. Curcumin-Induced Apoptosis in Human Hepatocellular Carcinoma J5 Cells: Critical Role of C a. Evidence-Based Complementary and Alternative Medicine. 2012;2012.

27. Hung C-M, Huang W-C, Pan H-L, Chien P-H, Lin C-W, Chen L-C, et al. Hepatitis B virus X upregulates HuR protein level to stabilize HER2 expression in hepatocellular carcinoma cells. BioMed research international. 2014;2014.

28. Chan MM-Y. Inhibition of tumor necrosis factor by curcumin, a phytochemical. Biochemical pharmacology. 1995;49(11):1551-6.

29. Aggarwal BB. Prostate cancer and curcumin: add spice to your life. Cancer biology & therapy. 2008;7(9):1436-40.

30. Carvalho DdM, Takeuchi KP, Geraldine RM, Moura CJd, Torres MCL. Production, solubility and antioxidant activity of curcumin nanosuspension. Food Science and Technology (Campinas). 2015;35(1):115-9.

31. Cao A-L, Tang Q-F, Zhou W-C, Qiu Y-Y, Hu S-J, Yin P-H. Ras/ERK signaling pathway is involved in curcumin-induced cell cycle arrest and apoptosis in human gastric carcinoma AGS cells. Journal of Asian natural products research. 2015;17(1):56-63

32. Hu S, Maiti P, Ma Q, Zuo X, Jones MR, Cole GM, et al. Clinical development of curcumin in neurodegenerative disease. Expert review of neurotherapeutics. 2015;15(6):629-37.
33. Lai H-W, Chien S-Y, Kuo S-J, Tseng L-M, Lin H-Y, Chi C-W, et al. The potential utility of curcumin in the treatment of HER-2-overexpressed breast cancer: an in vitro and in vivo comparison study with herceptin. Evidence-Based Complementary and Alternative Medicine. 2011;2012.

34. Chun K-S, Keum Y-S, Han SS, Song Y-S, Kim S-H, Surh Y-J. Curcumin inhibits phorbol ester-induced expression of cyclooxygenase-2 in mouse skin through suppression of extracellular signal-regulated kinase activity and NF-κB activation. Carcinogenesis. 2003;24(9):1515-24.

35. Reddy S, Rishi AK, Xu H, Levi E, Sarkar FH, Majumdar AP. Mechanisms of curcumin-and EGF-receptor related protein (ERRP)-dependent growth inhibition of colon cancer cells. Nutrition and cancer. 2006;55(2):185-94.

36. Huang C, Ke Q, Costa M, Shi X. Molecular mechanisms of arsenic carcinogenesis. Molecular and cellular biochemistry. 2004;255(1-2):57-66.

37. Simeonova PP, Wang S, Hulderman T, Luster MI. c-Src-dependent activation of the epidermal growth factor receptor and mitogen-activated protein kinase pathway by arsenic role in carcinogenesis. Journal of Biological Chemistry. 2002;277(4):2945-50.

38. Flora S, Mittal M, Mehta A. Heavy metal induced oxidative stress & its possible reversal by chelation therapy. Indian Journal of Medical Research.2008;128(4):501

How to Cite This Article:
Ahadi M, Naseh V, Salehipour M. The HER-2 as a Target Gene of Curcumin to Protect Hepatocytes Against the Arsenic-induced Carcinoma in Mice. Iran J Pathol. 2017;12(2):158-164.