Silibinin suppresses epithelial–mesenchymal transition in human non-small cell lung cancer cells by restraining RHBDD1

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Abstract

Background: Rhomboid domain containing 1 (RHBDD1) plays a crucial role in tumorigenesis. Silibinin, which is a natural extract from milk thistle, has shown anti-tumor effects against various tumors. Here, we investigate whether silibinin affects the function of RHBDD1 in non-small cell lung cancer (NSCLC) cell proliferation, migration and invasion.

Methods: The Oncomine database and an immunohistochemistry (IHC) assay were used to determine the RHBDD1 expression levels in lung cancer tissues. The associations between RHBDD1 and overall survival rate or clinicopathological parameters were respectively assessed using the Kaplan-Meier overall survival analysis or Chi-squared test. CCK-8 and Transwell assays were applied to analyze cell proliferation, migration and invasion. A549 cells were incubated with increasing concentrations of silibinin. RHBDD1 knockdown and overexpression were achieved via transfection with si-RHBDD1 or RHBDD1 overexpression plasmid, respectively. Western blotting was performed to measure the expressions of epithelial–mesenchymal transition (EMT) markers.

Results: We found that overexpression of RHBDD1 in lung cancer tissues correlates with a poor prognosis of survival. Clinical specimen analysis showed that upregulation of RHBDD1 correlates remarkably well with TNM stage and lymph node metastasis. Silibinin suppresses A549 cell proliferation, migration, invasion and EMT in a dose-dependent manner. Importantly, RHBDD1 was downregulated in silibinin-treated A549 cells. RHBDD1 overexpression reversed the suppressive effects of silibinin on A549 cell proliferation, migration, invasion and EMT expression, while its knockdown enhanced them.

Conclusions: These findings shown an anti-tumor impact of silibinin on NSCLC cells via repression of RHBDD1.

Keywords: NSCLC, Silibinin, RHBDD1, Epithelial–mesenchymal transition
Background

Non-small cell lung cancer (NSCLC) is the leading cause of tumor-related death worldwide [1, 2]. Accounting for approximately 85% of lung cancer cases, NSCLC mainly consists of adenocarcinoma, large cell carcinoma and squamous cell carcinoma [3]. Considerable progress has been reported in diagnostic and therapeutic methods, but the five-year survival rate still remains poor in NSCLC patients because of the high rate of metastasis [4, 5]. Therefore, it is essential to elucidate the mechanisms underlying tumor metastasis to help develop effective therapies for NSCLC patients.

Tumor metastasis is defined as the detachment of malignant cells from the primary tumor and their invasion of new tissues with the formation of a new lesion formation [6]. Epithelial–mesenchymal transition (EMT), as reflected by the downregulation of E-cadherin and upregulation of N-cadherin, vimentin and Slug [7, 8], is a phenotypic conversion that occurs in cancer cells and is involved in promoting their invasive potential [9, 10]. There have been notable breakthroughs in identifying some of the biomarkers targeting key effectors of EMT occurring in the context of carcinogenesis [8, 11]. Of note, rhomboid domain containing 1 (RHBDD1) is a mammalian member of rhomboid family proteins as highly conserved intramembrane serine protease [12].

Emerging evidence indicates that silencing RHBDD1 suppresses cell growth and proliferation in hepatocellular carcinoma [13], glioblastoma [14] and colorectal cancer [15]. In breast cancer [16, 17] and colorectal cancer cells [18], RHBDD1 has been shown to promote migration, invasion and EMT. However, there are few reports on the impact of RHBDD1 on the EMT of NSCLC cells and the associated mechanisms.

Silibinin, which is a natural extract from the fruit and seeds of milk thistle plants, is the most effective flavonoid against hepatic disease, with a wide range of pharmacological activities, including antioxidant, antineoplastic and detoxification effects, and it helps maintain the stability of liver cell membranes [19, 20]. Interestingly, it has been established that silibinin exerts pro-apoptotic effects in breast cancer [21] and renal cell carcinoma [22]. In addition, silibinin can attenuate TGF-β1-induced migration and invasion ability by inhibiting EMT in bladder transitional cell carcinoma [23]. Wu et al. also demonstrated that silibinin decreases bladder cancer metastasis by inhibiting EMT and stemness [24]. Here, we hypothesize that silibinin might inhibit NSCLC cell migration, invasion and EMT and that its inhibitory effects might be associated with RHBDD1 regulation.

To validate our hypothesis, we first analyzed the expression of RHBDD1 using the Oncomine database and immunohistochemistry (IHC). In addition, we assessed the clinical significance of RHBDD1 in lung cancer patients. In vitro experiments enabled further exploration of silibinin’s effects on NSCLC cell migration, invasion and EMT through modulation of RHBDD1.

Materials and methods

Online database analysis

The online microarray database Oncomine (www.oncomine.org) was searched to compare the expression levels of RHBDD1 between lung cancer and normal tissues. Briefly, the input was Gene Name: RHBDD1, Data Type: mRNA, and Analysis Type: Cancer vs. Normal Analysis. Four datasets, including Garber Lung [25], Hou Lung [26],
Okayama Lung [27] and Selamat Lung [28] were screened to extract the expression of RHBDD1 mRNA for each specimen. The extracted log-transformed and normalized expression values of RHBDD1 were further analyzed using GraphPad Prism 6 software.

Kaplan-Meier overall survival analysis
The Kaplan-Meier Plotter database (http://kmplot.com/analysis/), which integrates gene expression and clinical data, was used to study the correlation between RHBDD1 expression and overall survival in lung cancer. We focused our analysis on patient overall survival information and compared the survival rate for groups with higher and lower RHBDD1 expression levels using a Kaplan-Meier survival plot. We further calculated the hazard ratio with 95% confidence intervals and log rank p value.

Patients and specimens
We obtained formalin-fixed, paraffin-embedded (FFPE) cancerous and paired adjacent normal tissues from 56 NSCLC patients who had undergone curative surgery between January 2012 and December 2016 at the Central China Fuwai Hospital of Zhengzhou University. Some basic patient information is summarized in Table 1. None of the patients had received radiotherapy, chemotherapy or immunotherapy prior to surgery. All signed written informed consent. The tumor stage was determined following the criteria provided in the tumor node metastasis (TNM) staging classification system of the American Joint Committee on Cancer, 8th Edition [29]. In accordance with Declaration of Helsinki, this study obtained approval from the Medical Ethics Committee of the Central China Fuwai Hospital of Zhengzhou University.

| Clinical pathologic parameters | Total (n = 56) | Expression of RHBDD1 | p-value (chi-square test) |
|--------------------------------|---------------|----------------------|--------------------------|
| Sex                            |               |                      |                          |
| Male                           | 37            | 26                   | 11                       | 0.0937                   |
| Female                         | 19            | 9                    | 10                       |
| Age                            | 0.5343        |                      |                          |
| < 60                           | 27            | 18                   | 9                        |
| ≥ 60                           | 29            | 17                   | 12                       |
| Tumor size (cm)                | 0.2035        |                      |                          |
| < 4                            | 22            | 16                   | 6                        |
| ≥ 4                            | 34            | 19                   | 15                       |
| TNM stage                      | 0.0008        |                      |                          |
| I + II                         | 35            | 16                   | 19                       |
| III + IV                       | 21            | 19                   | 2                        |
| Lymph node metastasis          | 0.0006        |                      |                          |
| Yes                            | 37            | 29                   | 8                        |
| No                             | 19            | 6                    | 13                       |

Note: The bold values indicates the P values which are less than 0.05
Abbreviations: NSCLC Non-small cell lung cancer, TNM Tumor-node-metastasis classification system
IHC assay
An IHC assay was conducted to evaluate the expression of RHBDD1 in FFPE NSCLC tissues and matched adjacent tissues using a previously described standard streptavidin–biotin–peroxidase complex method [30]. The samples were cut into 4-μm thick sections, which were deparaffinized in xylene and rehydrated in graded alcohol. Then, they were immersed in 10 mM citrate buffer (pH = 6.0) at 95 °C for 15 min in a microwave oven for antigen retrieval, followed by endogenous peroxidase blocking with 3% hydrogen peroxide for 10 min. Incubation with the primary antibody against RHBDD1 was performed at 4 °C overnight. The sections were rinsed with phosphate-buffered saline (PBS) and incubated with HRP-conjugated secondary antibody at 37 °C for 30 min. After DAB staining and hematoxylin staining, two pathologists independently calculated the IHC staining score of each sample in a blinded manner based on the proportion of the positive-staining cells [31]. The staining intensity scale was scored: 0 (no staining), 1 (weak staining), 2 (moderate staining) or 3 (strong staining). The percentage of stained tumor cells was scored: 1 (1–25%), 2 (26–50%), 3 (51–75%) or 4 (76–100%). The NSCLC samples could then be classified into high RHBDD1 (score ≥ 6) and low RHBDD1 (< 6) groups with the optimal cutoff value being an IHC score of 6 [32].

Cell culture and treatment
Human NSCLC cell line A549 was purchased from the Chinese Academy of Sciences and grown in RPMI-1640 medium with 10% fetal bovine serum (FBS; Gibco) containing 5% CO2 at 37 °C. The cultures were incubated with DMSO (0.1%) and 10, 20 and 40 μM of silibinin (S0417; Sigma-Aldrich) for 24 h.

Cell transfection
The small interfering RNA targeting RHBDD1 (si-RHBDD1: 5′-CATGCAGACATCTG CATGACTGAGTTGT-3′) and negative control (si-NC: 5′-UAUGAUGUGGAUUU UCCUCUU-3′) were synthesized by RiboBio. The full-length RHBDD1 cDNA was sequentially amplified with primers (forward: 5′-ATCTGGCTGGGATTCTTGTTG-3′ and reverse: 5′-GGCTGGCTTGTAATGCTCTC-3′) and ligated into the pcDNA3.1 vector to generate RHBDD1 overexpression plasmid by Shanghai GenePharma. When the A549 cell confluence reached about 80%, si-RHBDD1 or si-NC was transfected into the cells. For the rescue experiments, RHBDD1 overexpression plasmid or NC was transfected into the A549 cells, followed by treatment with 40 μM silibinin. All transfections were conducted for 48 h with Lipofectamine 3000 reagent (Invitrogen).

CCK-8 assay
Briefly, A549 cells were seeded in a 96-well plate (3000 cells/well) and cultured overnight. At 24, 48 or 72 h, the cells in each well were incubated with 10 μl CCK-8 solution (Dojindo) for 2 h at 37 °C. Using a Microplate Reader (Bio-RadA), we measured the absorbance at a wavelength of 450 nm. Each experiment was performed at least three times.

Cell migration and invasion assay
We used Corning Costar Transwell chambers with 8 μm pores and coated with Matrigel (BD Biosciences) to assess the cell migration ability; or uncoated to assess the
invasion ability. Briefly, A549 cells from different groups suspended in serum-free medium were plated into the upper chamber and 500 μl of medium containing 10% FBS was added into the lower chamber. After 24 h incubation, migrated or invasive cells in the lower chamber were fixed for 30 min with 4% paraformaldehyde and stained for 15 min with 0.1% crystal violet at room temperature. Subsequently, stained cells were counted under a light microscope (magnification, ×200). Each experiment was performed at least three times.

**Western blotting**

Extraction of total protein was performed with RIPA buffer containing protease inhibitor cocktail (Sigma-Aldrich) and the concentration was determined using a BCA protein assay kit (Sigma-Aldrich). Protein samples (30 μg) were separated via 10% SDS-PAGE and then transferred onto PVDF membranes. After blocking with 5% non-fat milk in 1x TBST at 4 °C for 1 h, the membranes were incubated with primary antibodies against RHBDD1, E-cadherin, N-cadherin, vimentin and GAPDH overnight at 4 °C, followed by incubation with HRP-conjugated secondary antibodies at room temperature for 2 h. Protein bands were visualized using an enhanced chemiluminescence detection kit (Sigma-Aldrich) according to the manufacturer’s protocols.

**Statistical analysis**

All experiments were performed in triplicate. Statistical analyses were performed using GraphPad Prism 6.0 software. Results are expressed as means ± standard deviation (SD). The correlation of RHBDD1 expression and clinicopathological parameters was evaluated using Chi-squared test. Difference comparison between two groups was done with Student’s t test and among multiple groups with one-way analysis of variance followed by Tukey’s post hoc analysis. \( p < 0.05 \) was considered statistically significant.

**Results**

**Elevated RHBDD1 correlates with lower survival rates in lung cancer**

Our bioinformatics investigation used the Garber Lung, Hou Lung, Okayama Lung and Selamat Lung datasets. We showed that the expression of RHBDD1 is significantly higher in lung cancers, including squamous cell lung carcinoma, lung adenocarcinoma and large cell lung carcinoma, than in corresponding normal lung tissues (Fig. 1a). Using the Kaplan-Meier plotter online software based on a public database, we determined that the overall survival of lung cancer patients with low expression of RHBDD1 is better than that for patients with high expression (Fig. 1b; HR = 1.22, 95% CI = 1.03–1.43, \( p < 0.01 \)).

**Elevated RHBDD1 is associated with TNM stage and metastasis in NSCLC patients**

To confirm the results of the bioinformatic analysis, we performed IHC staining to determine RHBDD1 expression in samples from 56 NSCLC patients. Representative photomicrographs of different degrees of RHBDD1 expression intensity are shown in Fig. 2a. Statistical analyses (Fig. 2b) showed weaker staining of RHBDD1 protein in paraffin-embedded NSCLC tissues than in adjacent normal tissues. However, significantly elevated moderate and strong staining of RHBDD1 protein was observed in NSCLC tissues compared with adjacent normal tissues.
Based on the IHC score of RHBDD1 staining, all the NSCLC patients were divided into high ($n = 35$) and low ($n = 21$) RHBDD1 expression groups. We then analyzed the correlation between the expression of RHBDD1 and the clinicopathological parameters of NSCLC using Chi-squared test. RHBDD1 expression levels significantly correlate with TNM stage ($p = 0.0008$) and lymph node metastasis ($p = 0.0006$). These observations indicate that high RHBDD1 expression might have value as a prognostic factor for NSCLC.

**RHBDD1 is downregulated after silibinin treatment, inhibiting NSCLC cell proliferation, migration and invasion**

To explore whether silibinin exerts anti-tumor effects against NSCLC cells, different dosages (10, 20 and 40 $\mu$M) were used to treat A549 cells. The CCK-8 assay results show that silibinin significantly decreases cell proliferation in a dose-dependent manner (Fig. 3a). The Transwell migration assay results show that a marked decrease in the number of migrated cells, from 187.0 ± 8.5 in DMSO group to 144.0 ± 8.5, 114.7 ± 5.5 or 52.7 ± 9.1 after treatment with 10, 20 or 40 $\mu$M of silibinin, respectively (Fig. 3b).

![Fig. 1](image1.png)  
**Fig. 1** RHBDD1 is overexpressed in lung cancer tissues and this correlates with a poor survival rate. **a** RHBDD1 gene analysis based on the Garber Lung, Hou Lung, Okayama Lung and Selamat Lung datasets from the Oncomine database. **b** The RHBDD1 gene in lung cancer. Kaplan-Meier plots showing overall survival in lung cancer. In red: patients with RHBDD1 expressions above the median. In black: patients with expressions below the median. The corresponding $p$-value is listed. HR: hazard ratio; CI: confidence interval; $p$ value: log-rank test

![Fig. 2](image2.png)  
**Fig. 2** RHBDD1 is highly expressed in NSCLC tissues. **a** Representative images of different degrees of RHBDD1 immunohistochemistry staining in NSCLC and adjacent tissues (−, weak staining, + moderate staining, ++ strong staining). **b** A significantly higher percentage of NSCLC tissues than adjacent normal tissues showed high RHBDD1 expression.
Similarly, silibinin markedly decreased invasive cell numbers in a dose-dependent manner (Fig. 3b).

Western blotting showed that silibinin significantly downregulated the expression of RHBDL1, implying a negative correlation between them. The same inhibition was observed in EMT markers, as reflected by upregulated E-cadherin and downregulated N-cadherin and vimentin in A549 cells (Fig. 3c). These findings imply that RHBDL1 might play an important role in NSCLC cell behaviors.

Knockdown of RHBDL1 suppresses NSCLC cell proliferation, migration and invasion
Since RHBDL1 was overexpressed in NSCLC tissues, we then performed a loss-of-function assay by transfecting si-RHBDL1 into A549 cells to investigate the functional
role of RHBDD1. si-RHBDD1 transfection significantly suppressed cell proliferation compared with si-NC transfection (Fig. 4a). Consistent with silibinin, RHBDD1 knockdown obviously inhibited the migration and invasion ability of A549 cells (Fig. 4b). Furthermore, we observed that the expressions of RHBDD1, N-cadherin and vimentin were downregulated, while E-cadherin was upregulated in A549 cells following si-RHBDD1 transfection.

Silibinin attenuates NSCLC cell proliferation, migration and invasion by restraining RHBDD1
To further investigate whether RHBDD1 is a downstream regulator involved in silibinin mediation of cell proliferation, migration and invasion, RHBDD1 overexpression plasmid, si-RHBDD1 and the NC were transfected into A549 cells, followed by treatment with 40 μM silibinin. We discovered that the inhibitory activity of silibinin on cell proliferation (Fig. 5a), migration and invasion (Fig. 5b) is reversed by RHBDD1 overexpression, but significantly enhanced by RHBDD1 knockdown. Additionally, the expressions of RHBDD1, E-cadherin, N-cadherin and vimentin in silibinin-treated A549 cells is evidently abolished by RHBDD1 overexpression, but promoted by RHBDD1 knockdown (Fig. 5c). These findings collectively demonstrate that silibinin exhibits the anti-tumor activity by restraining RHBDD1 expression in A549 cells.
Discussion

Several publications have demonstrated that silibinin exhibits anti-tumor activity, e.g., in gastric cancer by enhancing paclitaxel toxicity \[33\] in glioblastoma by inducing apoptosis \[34\], and in ovarian cancer \[35\]. Here, we found that silibinin significantly restrains NSCLC cell proliferation, migration, invasion and EMT. We also observed that RHBDD1 expression decreases in silibinin-treated A549 cells and that silibinin
hinders A549 cell proliferation, migration, invasion and EMT, possibly by downregulating RHBDD1.

Consistent with our data, silibinin significantly inhibits the growth of lung cancer cells [36]. Hou et al. also showed that silibinin could restrain NSCLC cell migration in an EGFR/LOX-dependent manner [37]. We showed that silibinin treatment upregulated E-cadherin while downregulating N-cadherin and vimentin. Supporting these results, Raina et al. [38] reported that silibinin inhibits cell migration and invasion in human prostate cancer by upregulating E-cadherin. Deep et al. [39] explained that silibinin treatment suppresses signaling molecules related to EMT (E-cadherin and β-catenin) and cell survival (survivin and Akt) in PC-3 cells.

EMT is one of the most well-established theories for cancer cell metastasis. Targeting EMT has yielded notable results in basic research and clinical trials [40, 41]. E-cadherin, well-known for regulating cell-to-cell contact, cell shape and polarity, has been reported to be upregulated in states of decreased tumor metastasis [41, 42]. The anti-migratory and anti-invasive efficacy of silibinin also correlates with EMT suppression in prostate cancer [42, 43].

RHBDD1 is an important and newly identified member of the Rhomboid family, with abnormal overexpression in various cancers [18, 44]. Two important studies demonstrated that RHBDD1 acts as an oncogene in breast cancer cells via promotion of cell migration, invasion and EMT [16, 45]. Zhao et al. [17] corroborated that downregulation of RHBDD1 by miR-138-5p inhibits cell migration, invasion and EMT in breast cancer, indicating a novel oncogenic role in breast cancer cells.

Here, using bioinformatics analysis and related clinical sample detection, we confirmed RHBDD1 overexpression in lung cancer tissues and showed its association with poor survival rate in lung cancer patients. In addition, we discovered that RHBDD1 expression is restrained in silibinin-treated A549 cells, indicating that RHBDD1 might play a vital role in the development of NSCLC. To further confirm the hypothesis, we performed rescue experiments and found that RHBDD1 overexpression reverses the inhibitory effects of silibinin on cell proliferation, migration and EMT in A549 cells. Moreover, knockdown of RHBDD1 enhanced the inhibitory effects of silibinin on cell proliferation, migration, invasion and EMT in A549 cells.

**Conclusion**

Our findings show that silibinin can inhibit EMT in NSCLC cells through downregulation of RHBDD1, thereby restraining their proliferation, migration and invasion. We also propose a new molecular mechanism for silibinin that could prove useful in NSCLC therapy. The identification of RHBDD1 as a novel therapeutic candidate might provide a basis for further clinical research on silibinin.

**Abbreviations**

- NSCLC: Non-small cell lung cancer; RHBDD1: Rhomboid domain containing 1; EMT: Epithelial–mesenchymal transition; FBS: Fetal bovine serum; CCK-8: Cell Counting Kit-8

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**Authors’ contributions**

This study was conceived and designed by LGF. XSY and ZHY performed the experiments and gathered the data. WAF and MYC analyzed the data and wrote the paper. GY edited the paper. All the authors approved the final manuscript.
References

1. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2017. CA Cancer J Clin. 2017;67:7–30. https://doi.org/10.3322/caac.21387.

2. Watanabe T, Hosaka T, Ohmori-Matsuda K, Suzuki Y, Suzuki H, Yabuki H, et al. High preoperative plasma vasohibin-1 concentration predicts better prognosis in patients with non-small cell lung carcinoma. Health Sci Rep. 2018;1:e40. https://doi.org/10.1002/hsr2.240.

3. Lu G, Zhang Y. MicroRNA-140-5p suppresses non-small cell lung cancer cell growth and metastasis by targeting ZNF503. Cell Mol Biol Lett. 2019;24:34. https://doi.org/10.1186/s1658-019-0161-1.

4. Chtouq S, Brown X. Lung cancer staging: clinical and radiologic perspectives. Semin Interv Radiol. 2013;30:99–113. https://doi.org/10.1055/s-0033-1342950.

5. Inage T, Nakajima I, Yoshino I, Yasufuku K. Early lung cancer detection. Clin Chest Med. 2018;39:45–55. https://doi.org/10.1016/j.ccm.2017.10.003.

6. Fidler IJ. The pathogenesis of cancer metastasis: the ‘seed and soil’ hypothesis revisited. Nat Rev Cancer. 2003;3:453–8. https://doi.org/10.1038/nrc1098.

7. Wang H, Wu Q, Zhang Y, Zhang H-N, Wang Y-B, Wang W. TGF-β1-induced epithelial–mesenchymal transition in lung cancer cells involves upregulation of miR-9 and downregulation of its target, E-cadherin. Cell Mol Biol Lett. 2017;22:22.

8. Shi B, Ma C, Liu G, Guo Y. MiR-10a directly targets LIMK1 to inhibit proliferation and EMT of oral carcinoma cells. Cell Mol Biol Lett. 2019;24:1. https://doi.org/10.1186/s1658-018-0127-8.

9. Jeon HM, Lee J. MET: roles in epithelial-mesenchymal transition and cancer stemness. Ann Transl Med. 2017;5:5. https://doi.org/10.21037/atm.2016.12.67.

10. Pradella D, Naro C, Sette C, Ghigna C. EMT and stemness: flexible processes tuned by alternative splicing in development and cancer progression. Mol Cancer. 2017;16:8. https://doi.org/10.1186/s12943-016-0579-2.

11. Huang QJ, Wei FK, Xu XL, Ye SX, Song JW, Ding PK, et al. SOX9 drives the epithelial–mesenchymal transition in non-small-cell lung cancer through the Wnt/b-catenin pathway; 2019.

12. Freeman M. The rhomboid-like superfamily: molecular mechanisms and biological roles. Annu Rev Cell Dev Biol. 2014;30:235–54. https://doi.org/10.1146/annurev-cellbio-100913-012944.

13. Liu XN, Tang ZH, Zhang Y, Pan QC, Chen JH, Yu YS, et al. Lentivirus-mediated silencing of rhomboid domain containing 1 suppresses tumor growth and induces apoptosis in hepatoma HepG2 cells. Asian Pac J Cancer Prev. 2013;14:45–9. https://doi.org/10.7314/apjc.2013.14.15.

14. Wei X, Lv T, Chen D, Guan J. Lentiviral vector mediated delivery of RHBDD1 shRNA down regulated the proliferation of human glioblastoma cells. Technol Cancer Res Treat. 2012;11:377–81. https://doi.org/10.7785/tcrt.2012.500362.

15. Han J, Bai J, Yang Y, Yin H, Gao W, Lu A, et al. Lentiviruses-mediated knockdown of rhomboid domain containing 1 inhibits colorectal cancer cell growth. Mol Med Rep. 2015;12:377–81. https://doi.org/10.3892/mmr.2015.3365.

16. Huang C, Ji X, Peng Y, Wu M, Wu W, Luo Y, et al. Silencing of rhomboid domain containing 1 to inhibit the metastasis of human breast cancer cells in vitro. Iran J Basic Med Sci. 2018;21:1161–6. https://doi.org/10.22038/ijbms.2018.29788.7191.

17. Zhao C, Ling X, Li X, Hou X, Zhao D. MicroRNA-138-5p inhibits cell migration, invasion and EMT in breast cancer by directly targeting RHBDD1. Breast Cancer. 2019;26:60-817. https://doi.org/10.1007/s12282-019-00989-w.

18. Zhang Y, Shao L. Decreased microRNA-146a-5p contributes to respiratory syncytial virus disease through targeting toll-like receptor 4. Exp Ther Med. 2018;16:993–9. https://doi.org/10.3892/etm.2018.5672.

19. Muthumani M, Prabu SM. Silibinin potentially protects arsenic-induced oxidative hepatic dysfunction in rats. Toxicol Mech Methods. 2012;22:277–88. https://doi.org/10.3109/15376616.2011.647113.

20. Sozhen H, Celik O, Cetin ES, Yilmaz N, Aksozek A, Topal Y, et al. Evaluation of the protective effect of silibinin in rats with liver damage caused by itraconazole. Cell Biochem Biophys. 2015;71:1215–23. https://doi.org/10.1007/s10568-013-0531-8.

21. Bayram D, Es Ç, Kara M, Özgöçmen M, Candan IA. The apoptotic effects of silibinin on MDA-MB-231 and MCF-7 human breast carcinoma cells. Hum Exp Toxicol. 2016;35:690-699. https://doi.org/10.1177/0960327116658105.

22. Ma Z, Liu W, Zeng J, Zhou J, Guo P, Xie H, et al. Silibinin induces apoptosis through inhibition of the mTOR-Gli1-Bcl2 pathway in renal cell carcinoma. Oncol Rep. 2015;34:2461–8.

23. Li F, Sun Y, Jia J, Yang C, Tang X, Jin B, et al. Silibinin attenuates TGFβ/β induced migration and invasion via EMT suppression and is associated with COX2 downregulation in bladder transitional cell carcinoma. Oncol Rep. 2018;40:3543–50. https://doi.org/10.3892/or.2018.6728.
24. Kaijie W, Zhongyuan N, Jin Z, Jinhai F, Jiancheng Z, Tingting Z, et al. Silibinin inhibits β-catenin/ZEBl signaling and suppresses bladder cancer metastasis via dual-blocking epithelial-mesenchymal transition and stemness. Cell Signal. 2013;25:2625–33.

25. Garber ME, Troyanskaya OG, Schluens K, Petersen S, Thaesler Z, Pacyna-Gengelbach M, et al. Diversity of gene expression in adenocarcinoma of the lung. Proc Natl Acad Sci U S A. 2001;98:13784–9. https://doi.org/10.1073/pnas.241500798.

26. Hou J, Aerts J, den Hamer B, van Jicklen W, den Bakker M, Riegman P, et al. Gene expression-based classification of non-small cell lung carcinomas and survival prediction. PLoS One. 2010;5:e10312. https://doi.org/10.1371/journal.pone.0010312.

27. Okayama H, Kohno T, Ishii Y, Shimada Y, Shiraiishi K, Iwashita R, et al. Identification of genes upregulated in ALK-positive and EGFR/ALK-negative lung adenocarcinomas. Cancer Res. 2012;72:100–11. https://doi.org/10.1158/0008-5472.CAN-11-1403.

28. Selamat SA, Chung BS, Girard L, Zhang W, Zhang Y, Campan M, et al. Genome-scale analysis of DNA methylation in lung adenocarcinoma and integration with mRNA expression. Genome Res. 2012;22:1197–211. https://doi.org/10.1101/gr.132662.111.

29. Amin MB, Greene FL, Edge SB, Compton CC, Gershenwald JE, Brookland RK, et al. The eighth edition AJCC cancer staging manual: continuing to build a bridge from a population-based to a more “personalized” approach to cancer staging. CA Cancer J Clin. 2017;67:93–9. https://doi.org/10.3322/caac.21388.

30. Zeng J, Ma X, Wang J, Liu R, Shao Y, Hou Y, et al. Down-regulated HSDL2 expression suppresses cell proliferation and promotes apoptosis in papillary thyroid carcinoma. Biosci Rep. 2019;39. https://doi.org/10.1042/BSR20190425.

31. Jiang L, Liu JY, Shi Y, Tang B, He T, Liu JJ, et al. MTMR2 promotes invasion and metastasis of gastric cancer through simultaneous inhibition of mTOR and YAP. Biomed Res Int. 2018;2018:1

32. Camp RL, Dolled-Filhart M, Rimm DL. X-tile: a new bio-informatics tool for biomarker assessment and outcome-based cut-point optimization. Clin Cancer Res. 2004;10:7252–9. https://doi.org/10.1158/1078-0432.CCR-04-0713.

33. Zhang Y, Ge Y, Ping X, Yu M, Lou D, Shi W. Synergistic apoptotic effects of silibinin in enhancing paclitaxel toxicity in human prostate carcinoma PC3 cells via targeting integrin signaling. Mutat Res. 2014;768:35–46. https://doi.org/10.1016/j.mrfmmm.2014.05.002.

34. Bai ZL, Tay V, Guo SZ, Ren J, Shu MG. Silibinin induced human glioblastoma cell apoptosis concomitant with autophagy through simultaneous inhibition of mTOR and YAP. Biomed Res Int. 2018;2018:1–10.

35. Pashaei-Asl F, Pashaei-Asl R, Khodadadi K, Akbarzadeh A, Ebrahimie E, Pashaiasl M. Enhancement of anticancer activity by silibinin and paclitaxel combination on the ovarian cancer. Artif Cells Nanomed Biotechnol. 2018;46:1483–7. https://doi.org/10.1080/00313020701329914.

36. Mateen S, Raina K, Agarwal R. Chemopreventive and anti-cancer efficacy of silibinin against growth and progression of lung cancer. Nutr Cancer. 2013;65(Suppl 1):13–11. https://doi.org/10.1080/01635581.2013.738504.

37. Hou X, Du H, Quan X, Shi L, Zhang Q, Wu Y, et al. Silibinin inhibits NSCLC metastasis by targeting the EGF/LOX pathway. Front Pharmacol. 2018;9:21. https://doi.org/10.3389/fphar.2018.00021.

38. Raina K, RajamaniKam S, Singh RP, Deep G, Chitezath M, Agarwal R. Stage-specific inhibitory effects and associated mechanisms of silibinin on tumor progression and metastasis in transgenic adenocarcinoma of the mouse prostate model. Cancer Res. 2008;68:6822–30. https://doi.org/10.1158/0008-5472.CAN-08-1332.

39. Deep G, Kumar R, Jain AK, Agarwal C, Agarwal R. Silibinin inhibits fibronectin induced motility, invasiveness and survival in human prostate cancer PC3 cells via targeting integrin signaling. Mutat Res. 2014;768:35–46. https://doi.org/10.1016/j.mrfmmm.2014.05.002.

40. Christiansen JJ, Rajasekaran AK. Reassessing epithelial to mesenchymal transition as a prerequisite for carcinoma invasion and metastasis. Cancer Res. 2006;66:319–26. https://doi.org/10.1158/0008-5472.CAN-06-0410.

41. Guarino M, Rubino B, Ballabio G. The role of epithelial-mesenchymal transition in cancer pathology. Pathology. 2007;39:305. https://doi.org/10.1080/00313020701329914.

42. Deep G, Gangar SC, Agarwal C, Agarwal R. Role of E-cadherin in Antimigratory and Antiinvasive efficacy of Silibinin in prostate cancer cells. Cancer Prev Res. 2011;4:1222–32.

43. Wu K, Zeng J, Li L, Fan J, Zhang D, Xue Y, et al. Silibinin reverses epithelial-to-mesenchymal transition in metastatic prostate cancer cells by targeting transcription factors. Oncol Rep. 2010;23:1545–52.

44. Zhang Q, Weng Y, Jiang Y, Zhao S, Zhou D, Xu N. Overexpression of mI-140 Sp inhibits lipopolysaccharide-induced human intervertebral disc inflammation and degeneration by downregulating toll-like receptor 4. Oncol Rep. 2018;40:793–802. https://doi.org/10.3892/or.2018.6488.

45. Zhang X, Zhao Y, Wang C, Ju H, Liu W, Zhang X, et al. Rhomboid domain-containing protein 1 promotes breast cancer progression by regulating the p-Akt and CDK2 levels. Cell Commun Signal. 2018;16:655. https://doi.org/10.1186/s12967-018-0267-5.

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