Lung cancer in never-smoker Asian females is driven by oncogenic mutations, most often involving EGFR

Ha, Sang Yun; Choi, So-Jung; Cho, Jong Ho; Choi, Hye Joo; Lee, Jinseon; Jung, Kyungsoo; Irwin, Darry; Liu, Xiaoli; Lira, Maruja E.; Mao, Mao; Kim, Hong Kwan; Choi, Yong Soo; Shim, Young Mog; Park, Woong Yang; Choi, Yoon-La; Kim, Jhingook

Published in: OncoTarget

DOI: 10.18632/oncotarget.2925

Publication date: 2015

Document version Publisher's PDF, also known as Version of record

Citation for published version (APA): Ha, S. Y., Choi, S-J., Cho, J. H., Choi, H. J., Lee, J., Jung, K., Irwin, D., Liu, X., Lira, M. E., Mao, M., Kim, H. K., Choi, Y. S., Shim, Y. M., Park, W. Y., Choi, Y-L., & Kim, J. (2015). Lung cancer in never-smoker Asian females is driven by oncogenic mutations, most often involving EGFR. OncoTarget, 6(7), 5465-5474. https://doi.org/10.18632/oncotarget.2925
Lung cancer in never-smoker Asian females is driven by oncogenic mutations, most often involving \textit{EGFR}

Sang Yun Ha$^{1,*}$, So-Jung Choi$^{2,*}$, Jong Ho Cho$^{3,*}$, Hye Joo Choi$^{2}$, Jinseon Lee$^{2}$, Kyungsoo Jung$^{4}$, Darry Irwin$^{5}$, Xiao Liu$^{6,7}$, Maruja E. Lira$^{8}$, Mao Mao$^{9}$, Hong Kwan Kim$^{3}$, Yong Soo Choi$^{3}$, Young Mog Shim$^{3}$, Woong Yang Park$^{10}$, Yoon-La Choi$^{1,4,10}$, Jhingook Kim$^{2,3}$

$^1$Department of Pathology and Translational Genomics, Samsung Medical Center, Sungkyunkwan University School of Medicine, Seoul, Korea
$^2$Samsung Biomedical Research Institute, Samsung Medical Center, Sungkyunkwan University School of Medicine, Seoul, Korea
$^3$Department of Thoracic and Cardiovascular Surgery, Samsung Medical Center, Samsung Biomedical Research Institute, Sungkyunkwan University School of Medicine, Seoul, Korea
$^4$Samsung Advanced Institute for Health Sciences & Technology, Sungkyunkwan University School of Medicine, Seoul, Korea
$^5$Agena Bioscience, Sequenom, San Diego, CA, USA
$^6$BGI-Shenzhen, Shenzhen, 518083, China
$^7$Department of Biology, University of Copenhagen, Copenhagen, Denmark
$^8$Oncology Research Unit, Pfizer Worldwide Research and Development, San Diego, CA, USA
$^9$WuXi AppTec, Shanghai, China
$^{10}$Samsung Genome Institute, Samsung Medical Center, Sungkyunkwan University School of Medicine, Seoul, Korea

*These authors contributed equally to this work

Correspondence to:
Jhingook Kim, e-mail: jhingookkim@gmail.com
Yoon-La Choi, e-mail: ylachoi@skku.edu

Keywords: non-small cell lung cancer, adenocarcinoma, never-smoker female, driver mutation, \textit{EGFR}

Received: August 07, 2014  Accepted: December 16, 2014  Published: February 28, 2015

ABSTRACT

The aim of this study was to determine the distribution of known oncogenic driver mutations in female never-smoker Asian patients with lung adenocarcinoma. We analyzed 214 mutations across 26 lung cancer-associated genes and three fusion genes using the MassARRAY® LungCarta Panel and the \textit{ALK}, \textit{ROS1}, and \textit{RET} fusion assays in 198 consecutively resected lung adenocarcinomas from never-smoker females at a single institution. \textit{EGFR} mutation, which was the most frequent driver gene mutation, was detected in 124 (63%) cases. Mutation of \textit{ALK}, \textit{KRAS}, \textit{PIK3CA}, \textit{ERBB2}, \textit{BRAF}, \textit{ROS1}, and \textit{RET} genes occurred in 7%, 4%, 2.5%, 1.5%, 1%, 1%, and 1% of cases, respectively. Thus, 79% of lung adenocarcinomas from never-smoker females harbored well-known oncogenic mutations. Mucinous adenocarcinomas tended to have a lower frequency of known driver gene mutations than other histologic subtypes. \textit{EGFR} mutation was associated with older age and a predominantly acinar pattern, while \textit{ALK} rearrangement was associated with younger age and a predominantly solid pattern. Lung cancer in never-smoker Asian females is a distinct entity, with the majority of these cancers developing from oncogenic mutations.

INTRODUCTION

Lung cancer is the leading cause of cancer-related mortality, with 1.38 million annual deaths worldwide [1]. Tobacco smoking is the main risk factor for lung cancer; however, approximately 25% of lung cancers worldwide occur in never-smokers [2, 3]. Moreover, the risk of lung cancer differs by race/ethnicity. In the United States, approximately 10% of patients with lung cancer are never-smokers [4], while in Asia, >30% of
patients with lung cancer are never-smokers and ≥50% of lung cancers occur in women who are never-smokers [5]. Never-smoker East Asian females have a tendency to develop adenocarcinoma, and these never-smokers exhibit higher treatment response rates to epidermal growth factor receptor tyrosine kinase inhibitors (EGFR-TKIs), such as gefitinib and erlotinib, than those with a history of tobacco smoking [6]. In several phase III studies, significantly better response rates and longer progression-free survival were observed in advanced non-small cell lung cancer (NSCLC) patients harboring activating EGFR mutations who were treated with first-line EGFR-TKIs than those patients treated with doublet platinum-based chemotherapy [7, 8]. In addition, a fusion protein of the N-terminal portion of the echinoderm microtubule-associated protein-like 4 (EML4) gene and the intracellular signaling portion of the anaplastic lymphoma kinase (ALK) tyrosine kinase receptor has been identified in a small subset of NSCLC patients [9]. Patients harboring the EML4-ALK fusion show unique clinicopathologic and physiological characteristics and respond positively to ALK inhibitors [10–12]. As for EGFR mutations, several reports have identified the EML4-ALK fusion protein predominantly in young female never-smokers with adenocarcinoma, although the presence of this fusion protein is mutually exclusive with EGFR mutation [9, 13]. More specifically, the EML4-ALK fusion gene was found in 23.7% of never-smoker female lung adenocarcinoma patients [14].

Lung adenocarcinoma in never-smoker females has been established as a distinct entity based on its particular epidemiologic, clinical, and biological characteristics. EGFR mutations and the EML4-ALK translocation are defined as driver mutations because these alterations are responsible for both initiation and maintenance of lung cancer. The discovery of driver oncogene variantic variants that are sensitive to molecular-targeted drugs is crucial for improvement of treatment strategies. Therefore, use of multi-mutational profiling in lung cancer studies is important for identification of driver gene alterations in order to validate the effectiveness of molecular-targeted therapies. In addition, the proportion of never-smokers with lung cancer is likely to increase as a result of smoking cessation and prevention programs. Thus, the aims of this study were to analyze the distribution of oncogenic driver mutations and to compare these mutations with clinicopathologic characteristics in female Asian never-smoker lung adenocarcinoma patients.

RESULTS

Patient population and histopathologic classification

The patient population is summarized in Table 1. All patients were women with no smoking history. Median age at diagnosis was 60 years (range, 29–81 years). Most patients (85.3%) had undergone lobectomy. Two patients had pleural metastasis at the time of surgery. Eleven (5.6%) patients had received neoadjuvant concurrent chemoradiation therapy. Among the 198 patients in this study, 104 (52.5%) patients had T1 stage tumors, 83 (41.9%) patients had T2 stage tumors, and 11 (5.6%) patients had T3 stage tumors. Nine patients did not undergo lymph node dissection and were excluded from the analysis of N stage. N0, N1, and N2 stage tumors were observed in 124 (65.6%), 25 (13.2%), and 40 (21.2%) patients, respectively. Two of the 198 cases were classified as minimally invasive adenocarcinomas with an invasive component of <5 mm. Invasive adenocarcinomas were classified as predominantly acinar (142 cases, 71.7%), predominantly papillary (18 cases, 9.1%), predominantly solid (17 cases, 8.6%), predominantly lepidic (9 cases, 4.5%), and predominantly micropapillary (1 case, 0.5%) patterns. Nine (4.5%) cases were classified as mucinous adenocarcinoma. Two minimally invasive adenocarcinomas were classified as predominantly lepidic for statistical analysis. In addition, all histologic patterns that were observed in >10% of the tumor area were recorded. An acinar pattern was observed in 169 (85.4%) cases. Papillary, lepidic, micropapillary, solid, and mucinous patterns were observed in 68 (34.3%), 36 (18.2%), 36 (18.2%), 33 (16.7%), and 17 (8.6%) cases, respectively.

Fusion gene and LungCarta analyses

The results of the fusion gene and LungCarta analyses are summarized in Figure 2 and Table 2, and a full list of mutations that were identified is shown in Supplemental Table 1. Among the 198 cases, driver mutations were detected in 157 (79%) cases. EGFR mutations were the most frequently found mutation in lung adenocarcinomas of female never-smokers (124 cases, 63%). EGFR mutations were detected in exon 19 in 47 (24%) cases, exon 20 in 16 (8%) cases, exon 21 in 58 (29%) cases, exons 18 and 21 in 2 (1%) cases, and exons 18 and 20 in 1 (1%) case. ALK rearrangement was observed in 14 (7%) of cases. The frequencies of KRAS, PIK3CA, TP53, ERBB2, and BRAF mutations were 4%, 2.5%, 2%, 1.5%, and 1%, respectively. ROS1 and RET gene rearrangements were each found in 2 (1%) cases. ALK, ROS1, and RET gene fusions were mutually exclusive with EGFR and KRAS mutations. The gene mutations were mutually exclusive with the exception of concurrent mutation of EGFR/PIK3CA (n = 3), EGFR/TP53 (n = 1), ALK/TP53 (n = 2), KRAS/PIK3CA (n = 1), and KRAS/ERBB2 (n = 1; Figure 3). The results of Sanger sequencing validation were consistent with the results of LungCarta analyses in all cases (Supplemental Table 2).
Relationship between driver mutation status and clinicopathologic characteristics

The relationship between the driver mutation status and histologic subclassification of adenocarcinoma is summarized in Table 2 and Figure 4A–4E. Tumors with EGFR mutation exhibited a high frequency of predominantly acinar patterns and an absence of a predominantly mucinous pattern. ALK rearrangements were frequently found in cases with a predominantly solid pattern. KRAS mutations were detected with high frequency in cases with predominantly mucinous tumors. BRAF mutations were detected only in predominantly papillary tumors. These four genes were significantly associated with the histologic subclassification. Mucinous adenocarcinoma tended to harbor a lower frequency of known driver gene mutations than other histologic subtypes (mutation rate, 44.4% vs. 80.4%; p = 0.090).

Table 1: Patient population

| Characteristics                      | Number of patients (%) |
|--------------------------------------|------------------------|
| Age, years                           | median 60 (range, 29–81) |
| Histologic classification            |                        |
| Minimally invasive adenocarcinoma    | 2 (1.0)                |
| Invasive adenocarcinoma              |                        |
| Lepidic predominant                  | 9 (4.5)                |
| Acinar predominant                   | 142 (71.7)             |
| Papillary predominant                | 18 (9.1)               |
| Micropapillary predominant           | 1 (0.5)                |
| Solid predominant                    | 17 (8.6)               |
| Mucinous adenocarcinoma              | 9 (4.5)                |
| T stage                              |                        |
| 1                                    | 104 (52.5)             |
| 2                                    | 83 (41.9)              |
| 3                                    | 11 (5.6)               |
| N stage\(^1\)                        |                        |
| 0                                    | 124 (65.6)             |
| 1                                    | 25 (13.2)              |
| 2                                    | 40 (21.2)              |
| M stage                              |                        |
| 0                                    | 196 (99.0)             |
| 1                                    | 2 (1.0)                |
| Operation                            |                        |
| Wedge resection                      | 16 (8.0)               |
| Lobectomy                            | 169 (85.3)             |
| Lobectomy +\(^\alpha\) \(^2\)       | 13 (6.6)               |
| Neoadjuvant CCRT\(^3\)               |                        |
| Yes                                  | 11 (5.6)               |
| No                                   | 187 (94.4)             |

\(^1\)Nine cases were excluded because lymph node dissection was not performed.  
\(^\alpha\) includes lobectomy with wedge resection of the other lobe, bilobectomy, or pneumonectomy.  
\(^3\)CCRT indicates concurrent chemoradiation therapy.
EGFR mutations were associated frequently with lepidic and acinar patterns and infrequently with mucinous patterns in cases where the histologic pattern predominated in >10% of the tumor area. ALK rearrangement was more frequently detected in tumors with solid and mucinous patterns, while KRAS mutations were also commonly found in tumors with a mucinous pattern. Other mutations were not associated with histologic subtypes (Supplemental Table 3).

Patients with tumors harboring EGFR and BRAF mutations were older than those with wild-type tumors, whereas ALK gene fusions and ERBB2 mutations were
Table 2: Relationship between driver mutation status and histologic subclassification of adenocarcinoma according to predominant type

|          | Total  | Lepidic | Acinar | Papillary | Micropapillary | Solid   | Mucinous | p-value   |
|----------|--------|---------|--------|-----------|----------------|---------|----------|-----------|
| n (%)    | 198 (100) | 11 (5.6) | 142 (71.7) | 18 (9.1) | 1 (0.5) | 17 (8.6) | 9 (4.5) |           |
| Wild-type| 41 (20.7) | 3 (7.3)  | 29 (70.7) | 2 (4.9)  | 0 (0)    | 3 (7.3)  | 4 (9.8) | 0.446     |
| Mutant type| 157 (79.3) | 8 (5.1)  | 113 (72.0) | 16 (10.2) | 1 (0.6) | 14 (8.9) | 5 (3.2) |           |
| EGFR     | 124 (62.6) | 7 (5.6)  | 102 (82.3) | 9 (7.3)  | 1 (0.8) | 5 (4.0)  | 0 (0)  | <0.001    |
| ALK      | 14 (7.1)  | 0 (0)    | 6 (42.9)  | 2 (14.3) | 0 (0)    | 5 (35.7) | 1 (7.1) | 0.012     |
| KRAS     | 8 (4.0)   | 1 (12.5) | 1 (12.5)  | 0 (0)    | 0 (0)    | 2 (25.0) | 4 (50.0) | <0.001    |
| PIK3CA   | 5 (2.5)   | 0 (0)    | 3 (60.0)  | 1 (20.0) | 0 (0)    | 0 (0)    | 1 (20.0) | 0.31      |
| TP53     | 4 (2.0)   | 0 (0)    | 4 (2.8)   | 0 (0)    | 0 (0)    | 0 (0)    | 0 (0)    | 1         |
| ERBB2    | 3 (1.5)   | 0 (0)    | 2 (66.7)  | 0 (0)    | 0 (0)    | 0 (0)    | 1 (33.3) | 0.272     |
| BRAF     | 2 (1.0)   | 0 (0)    | 0 (0)     | 2 (100)  | 0 (0)    | 0 (0)    | 0 (0)    | 0.043     |
| ROS1     | 2 (1.0)   | 0 (0)    | 0 (0)     | 1 (50.0) | 0 (0)    | 1 (50.0) | 0 (0)    | 0.086     |
| RET      | 2 (1.0)   | 0 (0)    | 1 (50.0)  | 1 (50.0) | 0 (0)    | 0 (0)    | 0 (0)    | 0.487     |
| NRAS     | 1 (0.5)   | 0 (0)    | 0 (0)     | 0 (0)    | 0 (0)    | 1 (100)  | 0 (0)    | 0.192     |

1 Two cases of minimally invasive adenocarcinoma were included in the lepidic predominant type.
2 Six cases with acinar pattern showed concurrent mutation of EGFR/PIK3CA (n = 3), EGFR/TP53 (n = 1), and ALK/TP53 (n = 2).
3 Two cases of mucinous adenocarcinoma showed concurrent mutation of KRAS/PIK3CA (n = 1) and KRAS/ERBB2 (n = 1).

Figure 3: Diagram demonstrating driver gene mutation status and clinicopathologic features in 198 adenocarcinomas from East Asian never-smoker females. Most mutations were mutually exclusive with the rare exception of concurrent mutation of EGFR/PIK3CA (n = 3), EGFR/TP53 (n = 1), ALK/TP53 (n = 2), KRAS/PIK3CA (n = 1), and KRAS/ERBB2 (n = 1). Patients who received concurrent chemoradiation therapy and those who did not undergo lymph node dissection were excluded in the analysis of TNM stage.

associated with younger age at diagnosis (Supplemental Table 4). Driver gene mutation status was not associated with tumor stage.

**DISCUSSION**

In this study, we evaluated the status of driver gene mutations in lung adenocarcinoma samples from 198 East Asian female never-smokers using the MassARRAY® LungCarta Panel and ALK, ROS1, and RET fusion assays. Approximately 79% of patients harbored driver gene mutations, and EGFR mutation (63%) was the most frequent driver mutation detected. Other genetic alterations occurred less frequently with ALK gene fusion, KRAS mutation, ERBB2 mutation, HER2 mutation, ROS1 rearrangement, and RET rearrangement occurring in 7%,
Figure 4: Frequency of driver gene mutations according to predominant histologic subtype. (A) Lepidic, (B) Acinar, (C) Papillary, (D) Solid, (E) Mucinous subtype. In acinar subtype, four cases with concurrent mutations of EGFR/PIK3CA ($n = 3$) and EGFR/TP53 ($n = 1$) are represented as an EGFR mutation, and 2 cases of ALK/TP53 mutation as an ALK mutation. In mucinous subtype, two cases with concurrent mutations of KRAS/PIK3CA and KRAS/ERBB2 are represented as a KRAS mutation.
4%, 1.5%, 1%, 1%, and 1% of cases, respectively. Most mutations were mutually exclusive.

 Ethnic background is a well-established factor in NSCLC. Asian female non-smokers were the specific epidemiologic subgroup in this study, and a few previous large-scale studies have shown that Asian ethnicity is a prognostic factor of overall survival for NSCLC patients [15–17]. The link between ethnicity and cancer survival rates is likely due to differences in genetic background among ethnicities. Compared to Caucasians, East Asians with NSCLC have higher rates of EGFR mutations and lower rates of KRAS and LKB1 mutations [18]. According to a recent meta-analysis of 94 studies, the rate of EGFR mutation in lung adenocarcinoma in Asians (47.9%) is higher than that in Westerners (19.2%), while KRAS (11.2%) and LKB1 (4.0%) mutation rates are lower in Asians than in Westerners (26.1% and 16.2%, respectively) [19]. Smoking status is also an important factor in the development of lung adenocarcinoma. Lung cancers in never-smokers are more frequently associated with adenocarcinoma with EGFR mutations and less frequently with KRAS mutations [20]. In a recent study of lung adenocarcinomas, mutation rates for EGFR (39%), KRAS (4%), ALK (15%), and HER2 (5%) in never-smoker groups differed from rates in current or former smoker groups (10%, 35%, 4%, and 1%, respectively) [21]. Notably, the proportion of female lung cancers in never-smokers is much higher in East Asia than in Europe or the United States (60% vs 15–20%) [20].

 Although many studies have investigated driver gene mutations in lung adenocarcinoma, only a few have focused specifically on Asian female never-smokers despite the demonstrated importance of this patient group for selection of targeted candidates for NSCLC therapy [22–24]. Zhang et al. analyzed EGFR, KRAS, ALK, HER2, and BRAF mutation in 349 Chinese never-smoker females with lung adenocarcinoma and discovered mutation rates of 76%, 5%, 4%, 2%, and 1% for EGFR, HER2, ALK, KRAS, and HER2, respectively, while only 12% of cases harbored no detected mutation [25]. In an analysis of mutations of 10 driver genes (EGFR, KRAS, NRAS, HRAS, HER2, BRAF, ALK, PIK3CA, TP53, and LKB1) in 52 lung adenocarcinomas from East Asian never-smokers including 41 women [26], Sun et al. found similar rates of genetic alterations: EGFR mutation in 79% of cases, EML4-ALK fusion in 6% of cases, HER2 mutation in 4% of cases, and KRAS mutation in 2% of cases. In addition, only 10% of patients did not harbor any detected mutation in these genes. Ren et al. reported EGFR mutation in 70% and ALK rearrangement in 9.6% in adenocarcinomas from never-smoker Chinese women (see Supplemental Table 5 for summary of these studies) [27]. The incidences of mutation detected in this study are consistent with those of previous studies.

 Several important features distinguish our study from previous studies. In our study, we analyzed a total of 26 known oncogenes and 3 fusion genes. We discovered that 1% of lung adenocarcinomas from East Asian never-smoker female patients harbored gene rearrangements of ROS1 and RET, which have recently been recognized as driver genes in lung adenocarcinoma [28–30] but had not been evaluated in East Asian female never-smokers. We used a multiplex method that can be used in routine clinical practice and our results demonstrated that this method was comparable to traditional mutational analysis using polymerase chain reaction (PCR) for each gene. The use of a multiplexed PCR-based assay to genotype NSCLCs was recently demonstrated to be clinically feasible [31]. Moreover, we confirmed well-known associations of driver gene mutations with histologic subtypes and clinical characteristics. EGFR mutation correlated positively with older age and a predominantly acinar pattern [25, 32, 33], while ALK rearrangement was associated with younger age and solid histology [34, 35]. KRAS mutation was detected more frequently in mucinous adenocarcinoma [25, 36]. Furthermore, most PIK3CA mutations coexisted with other mutations [26]. The fact that our findings are consistent with those of previous studies supports the value of the multiplex method in mutation analysis and suggests that clinicopathologic associations are useful for determination of the priority of driver mutation tests.

 The most outstanding result of our study was that driver gene mutations were detected in 79% of female never-smoker Asian patients. Targeted therapies for EGFR (erlotinib/gefitinib) and ALK (crizotinib) are currently available, and 70% of female never-smoker Asian patients with EGFR mutation (63%) or ALK rearrangement (7%) may benefit from such targeted therapies. An additional 10% of patients in our study may benefit from newly developed targeted drugs. A recent clinical trial described a partial response to the EGFR/HER2 inhibitor BIBW2922 in a HER2 mutant tumor [37], suggesting that this new drug is a promising treatment strategy. More comprehensive genomic analysis and deep sequencing may be necessary to identify genetic alterations in the remaining 20% of patients with no known mutations [26]. Notably, known driver gene mutations were found less frequently in mucinous adenocarcinoma in our study, suggesting the possibility of mutation in another driver gene in this histologic subtype.

 In conclusion, lung cancer in never-smoker Asian females is a distinct entity, with the majority of these lung cancers developing from oncogenic mutations.

 METHODS

 Study population

 Specimens were obtained from Samsung Medical Center (SMC) in Seoul, Korea with prior informed patient consent and approval by the Institutional Review Board of Samsung Medical Center. Between January 2008
and January 2013, 3796 consecutive patients underwent pulmonary resection with curative intent for primary lung cancer at our institute.

After excluding 1307 patients with non-adenocarcinoma (Figure 1A), the remaining 2489 patients consisted of 1302 men and 1187 women (Figure 1B). Of the 1187 female patients with lung adenocarcinoma, 422 patients were excluded due to a lack of smoking information. Of the remaining 765 female patients, 94.9% (726 women) were never-smokers (Figure 1C). Among the 726 never-smoker female pulmonary adenocarcinoma cases, 198 had sufficient tissue for genomic analysis and were included in this study.

Data collection and histologic classification of adenocarcinoma

Study data were abstracted from in-hospital charts and electronic medical records by trained experienced nurses from the Departments of Medical Oncology, Surgical Oncology, Laboratory Medicine, Pathology, and Nursing at Samsung Medical Center. Baseline clinical characteristics included gender, age at diagnosis, smoking history, alcohol consumption status, date of diagnosis of advanced lung cancer, tumor histology, tumor stage, and ambulatory status at diagnostic work-up. Tumor stage was defined according to the seventh edition of the American Joint Committee on Cancer [38]. Smoking history and alcohol consumption status were determined by self-reported answers on questionnaires. Never-smokers were defined as patients who smoked less than 100 cigarettes over their lifetime. Histologic classification was determined according to the International Association for the Study of Lung Cancer, American Thoracic Society, and European Respiratory Society classification of lung adenocarcinomas [39]. All histologic patterns that covered at least 10% of the tumor area were recorded, and the predominant pattern was defined as the pattern that covered the largest portion of the tumor area.

DNA and RNA extraction

Genomic DNA or RNA was extracted from lung tumors or distant histologically normal lung tissue using standard protocols (RNeasy Mini Kit and QIAamp DNA Mini Kit, Qiagen, Valencia, CA, USA).

ALK, ROS1, and RET fusion assays

The nCounter™ gene expression assays were custom-designed and synthesized by NanoString Technologies (Seattle, WA, USA). Hybridization, sample cleanup, and digital reporter counts were performed according to the manufacturer’s protocol. RNA was obtained from fresh-frozen tissues using the QIagen RNeasy Mini Kit (Qiagen, Valencia, CA, USA). RNA concentration was assessed by spectrophotometry using the Nanodrop 8000 (Thermo-Scientific, Wilmington, DE, USA).

Samples were processed according to the gene expression protocol of NanoString Technologies. Briefly, total RNA was hybridized to a multiplexed mixture of custom-designed nCounter™ capture and reporter probes complementary to ALK, ROS1, and RET target sequences (Supplemental Table 2) for at least 16 h at 65°C. The samples were cleaned up and processed using an automated nCounter™ Sample Prep Station (NanoString Technologies). Unhybridized probes were removed, and the hybridization complex was immobilized onto a cartridge and aligned. Fluorescently labeled, color-coded reporters were subsequently imaged on an nCounter™ Digital Analyzer (NanoString Technologies) set at 1155 fields of view. Raw reporter counts were collected using nSolver software v1.0 (NanoString Technologies).

LungCarta analysis

High-throughput multiplex mutation profiling was performed using the MassARRAY® LungCarta Panel Version 1.0 (Sequenom, San Diego, CA, USA). This panel permits screening of 214 mutations across 26 oncogenes and tumor suppressors with a limit of sensitivity of approximately 10% with the use of 480 ng DNA [17]. DNA was amplified using the OncoCarta PCR primer mix, unincorporated nucleotides were inactivated by shrimp alkaline phosphatase, and a single base extension reaction was performed using extension primers that hybridize adjacent to the mutation. Multiplexed reactions were spotted onto the SpectroChipII (Sequenom) using the MassARRAY Nanodispenser. Peaks with different mass were resolved by matrix-assisted laser desorption/ionization time-of-flight on the MassARRAY Compact Analyzer. A predefined ratio of expected normal allele to mutant allele at a specific nucleotide position allows mutations to be detected using primer extensions at that specific position. Because of the multiplexing capabilities of this assay, multiple mutations are detected simultaneously using one panel. Further details of the multiplex methodology can be found in the protocol provided by the manufacturer.

Sanger sequencing

To validate the LungCarta analysis, we performed Sanger sequencing of EGFR in 11 selected cases (4 cases of EGFR E746_A750del, 4 cases of EGFR L858R, and 3 cases of KRAS G12D) according to the method described previously by our group [40].

Statistical analysis

Pearson’s chi-squared, Fisher’s exact, and independent t tests were used as indicated. All tests were two sided, and a p-value of < 0.05 was considered statistically significant. Statistical analyses were performed using SPSS software (SPSS Inc., Chicago, IL, USA).
ACKNOWLEDGMENTS

This study was supported by the Converging Research Center Program (No. 2013K000278) funded by the Korean government (The Ministry of Science, Information and Communications Technology, and Future Planning) and by the R&D Program for the Society of the National Research Foundation (NRF) funded by the Ministry of Science, ICT, and Future Planning (NRF-2013M3C8A1078501) and by a grant of the Korea Health Technology R&D Project through the Korea Health Industry Development Institute(KHIDI), funded by the Ministry of Health & Welfare, Republic of Korea (grant number:HI13C2096).

REFERENCES

1. Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global cancer statistics. CA Cancer J Clin. 2011; 61:69–90.
2. Ferlay J, Shin HR, Bray F, Forman D, Mathers C, Parkin DM. Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. International journal of cancer Journal international du cancer. 2010; 127:2893–2917.
3. Siegel R, Ward E, Brawley O, Jemal A. Cancer statistics, the impact of eliminating socioeconomic and racial disparities on premature cancer deaths. CA Cancer J Clin. 2011; 61:212–236.
4. Scagliotti GV, Longo M, Novello S. Nonsmall cell lung cancer in never smokers. Current opinion in oncology. 2009; 21:99–104.
5. Toh CK, Wong EH, Lim WT, Leong SS, Fong KW, Wee J, Tan EH. The impact of smoking status on the behavior and survival outcome of patients with advanced non-small cell lung cancer: a retrospective analysis. Chest. 2004; 126:1750–1756.
6. Thatcher N, Chang A, Parikh P, Rodrigues Pereira J, Ciuleanu T, van Pawel J, Thongprasert S, Tan EH, Pemberton K, Archer V, Carroll K. Gefitinib plus best supportive care in previously treated patients with refractory advanced non-small-cell lung cancer: results from a randomised, placebo-controlled, multicentre study (Iressa Survival Evaluation in Lung Cancer). Lancet. 2005; 366:1527–1537.
7. Mok TS, Wu YL, Thongprasert S, Yang CH, Chu DT, Saijo N, Sunpaweravong P, Han B, Margono B, Ichinose Y, Nishiwaki Y, Ohe Y, Yang JJ, et al. Gefitinib or carboplatin-paclitaxel in pulmonary adenocarcinoma. The New England journal of medicine. 2009; 361:947–957.
8. Maemondo M, Inoue A, Kobayashi K, Sugawara S, Oizumi S, Isobe H, Gemma A, Harada M, Yoshizawa H, Kinoshita I, Fujita Y, Okinaga S, Hirano H, et al. Gefitinib or chemotherapy for non-small-cell lung cancer with mutated EGFR. The New England journal of medicine. 2010; 362:2380–2388.
9. Soda M, Choi YL, Enomoto M, Takada S, Yamashita Y, Ishikawa S, Fujiwara S, Watanabe H, Kurashina K, Hatanaka H, Bando M, Ohno S, Ishikawa Y, et al. Identification of the transforming EML4-ALK fusion gene in non-small-cell lung cancer. Nature. 2007; 448:561–566.
10. Takahashi T, Sonobe M, Kobayashi M, Yoshizawa A, Menju T, Nakayama E, Mino N, Iwakiri S, Sato K, Miyahara R, Okubo K, Manabe T, Date H. Clinicopathologic features of non-small-cell lung cancer with EML4-ALK fusion gene. Annals of surgical oncology. 2010; 17:889–897.
11. Solomon B, Varella-Garcia M, Camidge DR. ALK gene rearrangements: a new therapeutic target in a molecularly defined subset of non-small cell lung cancer. Journal of thoracic oncology: official publication of the International Association for the Study of Lung Cancer. 2009; 4:1450–1454.
12. Chen Z, Sasaki T, Tan X, Carretero J, Shimamura T, Li D, Xu C, Wang Y, Adelmant GO, Capelletti M, Lee HJ, Rodig SJ, Rogman C, et al. Inhibition of ALK, PI3K/MEK, and HSP90 in murine lung adenocarcinoma induced by EML4-ALK fusion oncogene. Cancer research. 2010; 70:9827–9836.
13. Zhang X, Zhang S, Yang X, Yang J, Zhou Q, Yin L, An S, Lin J, Chen S, Xie Z, Zhu M, Wu YL. Fusion of EML4 and ALK is associated with development of lung adenocarcinomas lacking EGFR and KRAS mutations and is correlated with ALK expression. Mol Cancer. 2010; 9:188.
14. Shaw AT, Yeap BY, Mino-Kenudson M, Digumarthy SR, Costa DB, Heist RS, Solomon B, Stubbs H, Admane S, McDermott U, Settleman J, Kobayashi S, Mark EJ, et al. Clinical features and outcome of patients with non-small-cell lung cancer who harbor EML4-ALK. Journal of clinical oncology: official journal of the American Society of Clinical Oncology. 2009; 27:4247–4253.
15. Ou SH, Ziozias A, Zell JA. Asian ethnicity is a favorable prognostic factor for overall survival in non-small cell lung cancer (NSCLC) and is independent of smoking status. J Thorac Oncol. 2009; 4:1083–1093.
16. Ahn MJ, Lee J, Park YH, Ahn JS, Ziozias A, Zell JA, Park K, Ou SH. Korean ethnicity as compared with white ethnicity is an independent favorable prognostic factor for overall survival in non-small cell lung cancer before and after the oral epidermal growth factor receptor tyrosine kinase inhibitor era. J Thorac Oncol. 2010; 5:1185–1196.
17. Kawaguchi T, Matsumura A, Fukai S, Tamura A, Saito R, Zell JA, Maruyama Y, Ziozias A, Kawahara M, Ignatius Ou SH. Japanese ethnicity compared with Caucasian ethnicity and never-smoking status are independent favorable prognostic factors for overall survival in non-small cell lung cancer: a collaborative epidemiologic study of the National Hospital OrganizationStudy Group for Lung Cancer (NIHSGLC) in Japan and a Southern California Regional Cancer Registry databases. J Thorac Oncol. 2010; 5:1001–1010.
29. RET gene fusion in lung adenocarcinoma revealed from
YT, Kim JI, Kang JH, Seo JS. A transforming KIF5B and
Ju YS, Lee WC, Shin YJ, Lee S, Bleazard T, Won JK, Kim
in lung adenocarcinoma. Nat Med. 2012; 18:375–377.
Iwakawa R, Ogiwara H, Oike T, et al. KIF5B-RET fusions
Nammo T, Sakamoto H, Tsuta K, Furuta K, Shimada Y,
Kohno T, Ichikawa H, Totoki Y, Yasuda K, Hiramoto M,
expression of DNA repair and synthesis genes in never-
geriatric mutant kinases. J Clin Oncol. 2010; 28:4616–4620.
Wang Y, Liu L, Hu Q, Zhang J, Tang L, Li X, et al. Clinical activity of afatinib (BIBW 2992) in patients with lung adenocarcinoma with mutations in the kinase domain
D, De Mey J, Everaert H, Umelo I, In’t Veld P, Schallier D.
De Greve J, Teugels E, Geers C, Decoster L, Galdermans
Rodig SJ, Mino-Kenudson M, Dacie S, Yeap BY, Shaw A,
Hunter IA, Stubbs H, Law K, Lindeman N, Mark E, Jane PA,
Lynch T, Johnson BE, et al. Unique clinicopathologic features characterize ALK-rearranged lung adenocarcinoma in the western population. Clin Cancer Res. 2009; 15:5216–5223.
Rekhtman N, Ang DC, Riely GJ, Ladanyi M, Moreira AL.
KRAS mutations are associated with solid growth pattern and tumor-infiltrating leukocytes in lung adenocarcinoma. Mod Pathol. 2013; 26:1307–1319.
De Greve J, Teugels C, Geers C, Decoster L, Gaedersmans D, De Mey J, Everaert H, Umelo I, In’t Veld P, Schallier D. Clinical activity of afatinib (BIBW 2992) in patients with lung adenocarcinoma with mutations in the kinase domain of HER2/neu. Lung Cancer. 2012; 76:123–127.
Tanoue LT, Detterbeck FC. New TNM classification for non-small-cell lung cancer. Expert Rev Anticancer Ther. 2009; 9:413–423.
Travis WD, Brambilla E, Noguchi M, Nicholson AG, Geisenger KR, Yatabe Y, Beer DG, Powell CA, Riely GJ, Van Schil PE, Gellin K, Austin JH, Asamura H, et al. International association for the study of lung cancer/american thoracic society/european respiratory society international multidisciplinary classification of lung adenocarcinoma. J Thorac Oncol. 2011; 6:249–245.
Huang SY, Han J, Lee JY, Kim YE, Choi Y-L, Kim HK. Mucoepidermoid Carcinoma of Tracheobronchial Tree: Clinicopathological Study of 31 Cases. The Korean Journal of Pathology. 2011; 45:175–181.