Experimental and Kinetic Studies on Acid Red 88 Dye (AR88) Adsorption by Azolla filiculoides

Davoud Balarak1 and Yousef Mahdavi2*

1Department of Environmental Health, Health Promotion Research Center, School of Public Health, Zahedan University of Medical Sciences, Zahedan, Iran
2MSc Student of Environmental Health engineering, Student Research Committee, Mazandaran University of Medical Sciences, Mazandaran, Iran

Abstract

The use of Azolla filiculoides (AF) for the removal of Acid Red 88 dye (AR88) from aqueous solutions at different contact times, temperatures, pH, adsorbent doses and initial dye concentration was investigated. The extent of dye removal decreased with increasing adsorbent dosage and AR88 concentration, also increased with increasing contact time and temperature. The adsorption isotherms are described by means of the Langmuir, Freundlich, Tempkin and Sips isotherms. It was found that the Langmuir equation fit better than the other equation. Maximum adsorption capacity (qmax) was calculated at different temperatures (20, 30, 40 and 50°C) 22.45, 23.95, 25.29 and 26.17 mg/g, respectively. The adsorption kinetics of Acid Red 88 could be described by the pseudo-second-order reaction model. The obtained results are: (1) high levels of color removal (>98%) were achieved with low contact times adsorbent/dye (less than 90 min contact); and (2) the whole Azolla filiculoides can be successfully used as adsorbent of AR88 in aqueous solutions. Azolla biomass, an inexpensive and easily available material, can be alternative for more costly adsorbents used for dye removal in wastewater treatment processes.

Keywords: Adsorption; Isotherm; Acid Red 88; Azolla filiculoides; Kinetics

Introduction

Recently, the societies have been involved to the serious environmental issues due to development of industries and use of various chemical in industries [1-3]. The dyes have been introduced as a major group of pollutants. Although the dyes are applied in various industries; however, the textile industries are the main sectors in consumption of dyes [4-6]. Annually, a large amount of dyes are generated in the world. They are classified into several groups. The reactive dyes are one of these groups which are commonly applied in coloring of strings, wool and polyamide fibers. It has been expressed that 50% of reactive dyes is lost during the dying process [7]. The studies has been illustrated that there is a variety of unpleasant and detrimental consequences for human and environment associated with use of dyes [8-13]. Therefore, the dye removal from industrial effluents is high priority subject. Among the various techniques, the adsorption process has been showed substantial potential for dye removal from industrial effluents [14-16]. Currently, the activated carbon has become most popular adsorbent; but this is not economical adsorbent due to its high operation and regeneration costs [6,17,18]. Therefore, there is a strong demand to find a cost-effective alternative for dye removal from effluents. A number of materials such as Trametes versicolor, Cupuassu shell, Stoechospermum marginatum, pine tree leaves, Loofa egyptiaca and etc. were employed as adsorbent for dye removal [1,19-22]. Since the aquatic plant are capable to grow in contaminated water therefore they has been used in several study to remove the pollutants [23]. Azolla filiculoides is one of this plants which it is commonly found in ditches, ponds and slow moving streams. Its high growth rate is considered as main feature of Azolla [24,25]. Recently, the Azolla has been used in several studies to remove the pollutants [26-28]. Thus, the present study was conducted for following purposes: a) to investigate the potential of dried azolla for AR88 dye removal, b) to assess the effect of several variables including contact time, pH, temperature, initial AR88 dye concentration and adsorbent does on dye removal efficiency, c) to study the adsorption isotherm and kinetic.

Materials and Method

Azolla filiculoides was collected from Anzali wetland, Iran. The biomass was oven-dried at 50°C for 12 h, then ground and screened through a sieve with 110 meshes to obtain particle sizes less than 0.135 mm. The crushed particles were then treated with 0.1 M HCl for 5 h, then washing with distilled water and then kept for shaded dry. The resultant biomass was subsequently used in sorption experiments [29].

The morphological features and surface characteristics of A. filiculoides before and after use were examined using an environmental scanning electron microscopy (ESEM) instrument (Philips XL30). The specific surface area of adsorbent was determined by the BET method using the Gemini 2357 of micrometrics Co.

Acid Red 88 (EC number=216-760-3; chemical formula: C19H13N2NaO4S; molecular weight=400.39 g/mol; maximum wavelength=503 nm) were obtained from Merck Co. and used without further purification. All AR88 solutions used in this study were prepared by weighing and dissolving the required amounts AR88 in distilled water. The chemical structures of AR88 are presented in Figure 1.

Batch adsorption studies

Batch adsorption experiments were performed using 100 ml glass bottles with addition of 0.4 mg purified AF and 50 ml of AR88 solution of increased initial concentrations (C0) from 25 to 200 mg/L. The glass bottles were sealed and placed within a temperature control box to maintain water temperature. The pH of the samples was adjusted by adding 0.1 M HCl or 0.1 M NaOH to each 200 ml of the prepared
solution to pH 7. The pH of solutions was measured with a pH meter. In the experiments on the effect of temperature, the temperature was held at 20, 30, 40 and 50 °C and the pH was fixed at 7. At the end of the equilibrium period, the suspensions were separated for later analysis of the dye concentration. The amount of AR88 adsorption at equilibrium was calculated from the following equation [30]:

$$ q_e = \frac{C_0 - C_e}{W} V $$

Where $C_0$ and $C_e$ (mg/L) are the liquid-phase concentrations of dye at initial and equilibrium, respectively, $V$ (L) the volume of the solution and $W$ (g) is the mass of adsorbent used. The concentration of AR88 after and before adsorption was determined using a spectrophotometer ($k_{max} = 503$ nm).

The dye removal percentage can be calculated as follows [31]:

$$ R = \frac{C_0 - C_e}{C_0} \times 100 $$

**Results and Discussion**

The Dried *Azolla filiculoides* was also examined before and after use using environmental scanning electron microscopy. Figure 2 (a) clearly shows the pore textural structure of dried AB before use. However, as shown in Figure 2 (b), clear pore textural structure is not observed on the surface of dried AB after use which could be due to either agglomeration on the surface or the incursion of AR88 into the pores of dried AB.

**Effect of contact time**

The adsorption of AR88 onto Azolla was studied as a function of contact time in order to decide whether the equilibrium was reached. For this, 50 mg L$^{-1}$ of AR88 solutions at pH 3 were contacted with 4 g L$^{-1}$ of Azolla suspensions. The samples were taken at different periods of time and analyzed for their AR88 concentration (Figure 3). Dye adsorption onto Azolla filiculoides increased from 42.1% to 98.9% when the contact time was increased from 10 to 75 min. The AR88 adsorption rate is high at the beginning of the experiment because initially the adsorption sites are more available and AR88 ions are easily adsorbed on these sites [32,33]. The equilibrium can be reached within 75 min, and thus, further adsorption experiments were carried out for a contact time of 75 min.

**Effect of pH**

pH is an important factor in controlling the removal of AR88 dye onto AF adsorbent. The removal of AR88 dye was studied at a temperature of 27 ± 2 °C and 50 mg/L (Figure 4). Concentration by varying the pH from 3.0 to 11.0, the solution was equilibrated for 75 min. The result indicates that AF biomass adsorbent showed good removal capacity in acidic medium than in basic medium. The percentage removal of AR88 dye of adsorption on AF biomass adsorbents progressively decreased on the pH of the solution increased from 3.00 to 11.0. At higher pH, the percentage removal was found to decrease because the surface area of the adsorbent was more protonated and competitive adsorption occurred between H$^+$ and free AR88 ions and their OH$^-$ towards the fixation sites [29]. Therefore, H$^+$ ions react with anionic functional groups on the surface of the adsorbent and results in restriction of the number of binding sites favorable for the removal of AR88. However, a favorable increase in percentage removal for AF biomass adsorbent was observed below pH 3.

**Effect of adsorbent dosage**

Adsorbent dosage is an important parameter because this factor determines the capacity of an adsorbent for a given initial concentration of the adsorbate. The effect of adsorbent dosage (adsorbent prepared in different batch) was studied on AR88 removal by keeping all other experimental conditions constant (Figure 5). The results show that as the adsorbent concentration increased, the percentage of adsorption also increased, but the amount adsorbed per unit mass of the adsorbent decreased considerably. The decrease in unit adsorption with increasing dose of adsorbent is basically due to adsorption sites remaining unsaturated during the adsorption reaction [34,35].

![Figure 1: The chemical structures of AR88 [29].](image1)

![Figure 2: SEM image of Azolla filiculoides before and after used.](image2)

![Figure 3: Effect of contact time for AR88 adsorption (Con = 50 mg/L, pH = 3, dose: 4 g/L and temp 27°C).](image3)

![Figure 4: Effect of pH on adsorption (Con = 50 mg/L, Contact time = 75 min, dose: 4 g/L and temp 27°C).](image4)
The equilibrium adsorption capacity of the adsorbent for AR88 increased with increasing initial dye concentration, as is also shown in Figure 6. The dye adsorption capacity is highly concentration dependent. The increase in loading capacity of the adsorbent with relation to dye ions is probably due to a high driving force for mass transfer [36,37]. In fact, the more concentrated the solution, the better the adsorption. At 27 °C, when the initial dye concentration was increased from 25 to 200 mg/L, the loading capacity of dried adsorbent increased from 6.19 to 34.95 mg of AR88 per gram of Azolla biomass.

### Adsorption isotherms

The adsorption isotherms provide information about the distribution of dye molecules within the adsorbent at equilibrium. In general, adsorption isotherm studies are carried out in order to correlate the adsorption capacity and the residual concentration of the adsorbate, present in the aqueous solution. Several isotherm equations are available in literature; four of them were selected for this study: Langmuir, Freundlich, Temkin and Sips equations.

The Langmuir equation [38]:

\[
\frac{C_e}{q_e} = \frac{1}{K_L q_m} + \frac{C_e}{q_m}
\]

The Freundlich equation [39]:

\[
\ln q_e = \ln K_F + \frac{1}{n} \ln C_e
\]

The Tempkin equation [40]:

\[
\ln q_e = A + B \ln C_e
\]

The Sips equation [41]:

\[
\frac{1}{q_e} = \frac{1}{q_{max}} + \frac{1}{q_{max} C_e}
\]

\[
K_L \text{ is the Langmuir constant (L mg}^{-1}\text{) related to the affinity of binding sites and the free energy of sorption; } Q_{max}(\text{mg g}^{-1}) \text{ represent the maximum adsorption capacity when the surface is fully covered with dye molecules that assists in the comparison of adsorption performance; } q_e \text{ is the dye concentration at equilibrium onto the adsorbent (mg g}^{-1}\text{); } C_e \text{ is the dye concentration at equilibrium isolation (mg L}^{-1}\text{); } K_F \text{ and } n \text{ are the Freundlich constants, } A \text{ and } B \text{ are the Temkin constant, } T \text{ is the absolute temperature in Kelvin } R \text{ is the universal gas constant. The } K_F(1/\text{mg}) \text{ and } q_{max}(\text{mg/g}) \text{ are the Sips equilibrium constant and maximum adsorption capacity values. The essential Characteristic of the isotherm could be expressed by a dimensionless constant called equilibrium parameter } R_e \text{ Defined by as follows [42]:}

\[
R_e = \frac{1}{1 + K_L C_e}
\]

The value of } R_e \text{ indicates the type of the isotherm; which is unfavorable } (R_e>1), \text{ linear } (R_e=1), \text{ favorable } (0<R_e<1) \text{ or irreversible } (R_e=0).

Table 1 lists the parameters of the isotherm models along with the regression coefficients (R²). The R² values in Table 1 reveal that the Langmuir isotherm fits the experimental results better than other model, implying that the adsorption of AR88 onto AF biomass follows the mechanism of monolayer adsorption (chemisorption) on a homogenous surface. Although the Langmuir and Sips equations have similar correlation coefficients, the Langmuir is a simpler equation and therefore easier to use. Consequently, it was selected to predict } q_e \text{ values in the present work.

The effect of temperature on the adsorption rate of AR88 on Azolla biomass was investigated at four different temperatures (20, 30, 40 and 50 °C) using initial concentration of 100 mg/L (Table 1). The major effect of temperature is influence by the diffusion rate of adsorbate molecules and internal pores of the adsorbent particle. The increase in temperature from 20 to 50 °C increased the Azolla biomass monolayer adsorption capacities from 22.45 to 26.17 mg/g (Table 1). This phenomenon indicates that the adsorption process is endothermic in nature. This may be due to the mobility of molecules which increases generally with a rise in temperature, thereby facilitating the formation of surface monolayers.

| Tem (°C) | Langmuir model | Freundlich model | Temkin model | Sips model |
|---------|----------------|------------------|--------------|------------|
|         | q₀              | K₀               | R²           | n          | K₀        | R² | q₀        | K₀ | n  | R² |
| 20      | 22.45           | 0.0014           | 0.994        | 2.65       | 6.14      | 0.812 | 31.71 | 7.41 | 0.804 | 14.4 | 0.041 | 1.34 | 0.914 |
| 30      | 23.95           | 0.0039           | 0.986        | 2.17       | 7.39      | 0.845 | 37.45 | 6.52 | 0.781 | 17.7 | 0.075 | 1.69 | 0.943 |
| 40      | 25.29           | 0.0063           | 0.997        | 3.09       | 7.85      | 0.791 | 41.12 | 4.36 | 0.768 | 21.2 | 0.095 | 1.93 | 0.904 |
| 50      | 26.17           | 0.0098           | 0.998        | 3.44       | 8.13      | 0.826 | 44.69 | 2.79 | 0.819 | 23.4 | 0.104 | 2.18 | 0.935 |

Table 1: The adsorption isotherms constants for the removal AR88 onto Azolla biomass.
Adsorption kinetics

In order to evaluate the rate-controlling step in the overall adsorption process, three kinetic models, pseudo-first-order, pseudo-second-order and intra-particle diffusion, are adopted to investigate the adsorption process.

The pseudo-first-order equation can be expressed as the following equation [43]:

$$
\log (q_e - q_t) = \log q_e - \frac{k_1}{2.303} t
$$

Where $q_e$ (mg/g) and $q_t$ (mg/g) are the AR88 adsorption capacity at equilibrium and at time $t$ (min), respectively, and $k_1$ (min$^{-1}$) is the rate constant of the pseudo-first-order.

The parameters $k_1$ and $q_e$ could be calculated from the slope and intercept of the plots of $\log (q_e - q_t)$ versus $t$ and are given in Table 2. The values of the correlation coefficient $R^2$ obtained at all the studied concentrations are low, in the range 0.689–0.784. Furthermore, the experimental values of $q_e$, $q_{e, \text{exp}}$ (mg/g) are far from the calculated $q_e$, cal (mg/g). This suggests that the pseudo-first-order kinetic model is not suitable to describe the adsorption process.

On the other hand, the pseudo-second-order kinetic model can be expressed in linear form as follows [44]:

$$
t = \frac{1}{q_e} + \frac{t}{k_2 q_e^2}
$$

Where $k_2$ (g/mg min) is the rate constant of the pseudo-second-order. The kinetic parameters of AR88 under different conditions were calculated and are given in Table 2. It is seen that the pseudo-second-order model well represented the experimental data ($R^2>0.995$). This suggests that adsorption AR88 onto Azolla biomass predominantly follows the pseudo-second-order kinetic model. Besides, the calculated data ($q_e$, cal) agree well with the experimental data ($q_e$, exp). The parameters are listed in Table 2. The same conclusion was found in the literature on the adsorption of AR88 by Lemna minor [29].

The two models above cannot identify the diffusion mechanism during the adsorption process, so the experimental data are tested by the intra-particle diffusion model, which can be expressed by following equation [45]:

$$
q_t = k t^{1/2} + C
$$

Where $q_t$ (mg/g) is the amount of AR88 adsorbed at time $t$ (min), $k$ (mg/g min$^{-1/2}$) is the intra-particle diffusion rate constant and $C$ is the intercept.

The adsorption of a solute from solution by porous adsorbents is essentially related to three consecutive steps. The first step is the external surface adsorption or the instantaneous adsorption. The second step is the gradual adsorption stage where intra-particle diffusion is rate-limiting. The third step is the final equilibrium stage where intra-particle diffusion started to slow down due to the extremely low adsorbate concentrations left in the solutions. One or more of these three steps control the adsorption rate. The parameters are listed in Table 2 and show a double straight line nature. Initially, it is postulated that AR88 is transported to the external surface of Azolla biomass through film diffusion within a very short time. The first linear part could be due to the entry of AR88 molecules into the Azolla biomass particle by intra-particle diffusion. The second linear part represents the final equilibrium stage. As given in Table 2, it is obvious that values of $k$ increase from 1.65 to 6.94 mg/g min$^{-1/2}$ when the initial AR88 concentrations increase from 25 to 200 mg/L.

Table 2: Adsorption kinetic constants AR88 adsorption onto Azolla biomass

| $C_e$ (mg/L) | $q_e$ exp (mg/g) | $q_e$ Pseudo-first order | $q_e$ Pseudo-second order | $q_e$ Intraparticle diffusion |
|-------------|-----------------|------------------------|--------------------------|------------------------------|
|             | $K_1$ | $q_e$ | $R^2$ | $K_2$ | $q_e$ | $R^2$ | $K$ | $C$ | $R^2$ |
| 25          | 6.81  | 0.014 | 4.76  | 0.739 | 0.0042 | 6.45  | 0.996 | 1.65 | 2.17 | 0.918 |
| 50          | 12.48 | 0.021 | 9.14  | 0.689 | 0.0079 | 12.14 | 0.997 | 3.22 | 3.85 | 0.894 |
| 100         | 23.94 | 0.024 | 16.45 | 0.718 | 0.0087 | 22.86 | 0.998 | 5.09 | 1.19 | 0.906 |
| 200         | 34.95 | 0.028 | 28.32 | 0.784 | 0.0035 | 35.69 | 0.999 | 6.94 | 5.36 | 0.932 |

Conclusion

The efficiency of AR88 dye removal from aqueous solution by Azolla biomass as a natural biosorbent was evaluated. The effect of parameters such as contact time, pH, AR88 concentration, adsorbent doses and temperature were analyzed. In general, the best, efficiency of AR88 removal by Azolla biomass was obtained at pH acidic, dose adsorbent of 4 g/L, initial concentration of AR88 of 25 and 50 mg/L and contact time 75 min. The adsorption isotherms are described by means of the Langmuir, Freundlich, Temkin and Sips isotherms. It was found that the Langmuir equation fit better than the other equation. The rate of adsorption was found to conform to pseudo-second-order kinetics with a good correlation. The increase in temperature increased the Azolla biomass monolayer adsorption capacities. Finally Azolla biomass, an inexpensive and easily available material, can be an alternative for more costly adsorbents used for dye removal in wastewater treatment processes.

References

1. Daneshvar E, Kousha M, Sohrabi MS, Khataee A, Converti A (2012) Biosorption of three acid dyes by the brown macroalga Stoechospermum marginatum: Isotherm, kinetic and thermodynamic studies. Chemical Engineering Journal 195–196: 297-306.
2. Kadirevu K, Kaviyipria M, Karthika C, Radhika M, Vennilamani N, et al. (2003) Utilization of various agricultural wastes for activated carbon preparation and application for the removal of dyes and metal ions from aqueous solutions. Biosource Technology 87: 129–132.
3. Anouzla A, Abrourki Y, Souabi S, Safi M, Rhbal H (2009) Colour and COD removal of disperse dye solution by a novel coagulant: Application of statistical design for the optimization and regression analysis. J Hazard Mater 166: 1302–1306.
4. Zazzouli MA, Balarak D, Mahdavi Y, Karimnejad F (2014) The application of Azolla filiculoides biomasses in acid blue 15 dye (AB15) removal from aqueous solutions. J Bas Res Med Sci 1: 29-37.
5. Salima A, Ounissa KS, Lynda M, Mohamed B (2012) Cationic dye (MB) removal using polymer inclusion membrane (PMIs). Procedia Engineering 33: 38-46.
6. Arica MY, Bayraram Aybu G (2007) Biosorption of Reactive Red-120 dye from aqueous solution by native and modified fungus biomass preparations of Lentinus sajor-caju. J Hazard Mater 149: 499-507.
7. Galán J, Rodríguez A, Gómez JM, Allen SJ, Walker GM (2013) Reactive dye
adsorption onto a novel mesoporous carbon. Chemical Engineering Journal 219: 62-68.
8. Mezli C, Boukerroul A (2012) Removal of a Basic Textile Dye from Aqueous Solution by Adsorption on Regenerated Clay. Procedia Engineering 33: 303-312.
9. Rodríguez Couto S (2009) Dye removal by immobilised fungi. Biotechnol Adv 27: 227-235.
10. Mousavi GR, Mahmoudi M (2009) Degradation and biodegradability improvement of the reactive red 198 azo dye using catalytic ozonation with MgO nanocrystals. Chemical Engineering Journal 152: 1–7.
11. Yang Y, Wei B, Zhao Y, Wang J (2013) Construction of an integrated enzyme system consisting azooreductase and glucose 1-dehydrogenase for dye removal. Bioresour Technol 130: 517-521.
12. Akar T, Özcan AS, Tunali S, Özcan A (2008) Biosorption of a textile dye (Acid Blue 40) by cone biomass of Thuja orientalis: Estimation of equilibrium, thermodynamic and kinetic parameters. Bioresour Technol 99: 3057-3065.
13. Arami M, Limea NY, Mahmoodi NM (2006) Evaluation of the adsorption kinetics and equilibrium for the potential removal of acid dyes using a biosorbent. Chemical Engineering Journal 139: 2-10.
14. Amin NK (2008) Removal of reactive dye from aqueous solutions by adsorption onto activated carbons prepared from sugarcane bagasse pith. Desalination 223: 152-161.
15. Gök Ö, Özcan AS, Özcan A (2010) Adsorption behavior of a textile dye of Reactive Blue 19 from aqueous solutions onto modified bentonite. Applied Surface Science 256: 5439-5443.
16. Toor M, Jin B (2012) Adsorption characteristics, isotherm, kinetics, and diffusion of modified natural bentonite for removing diazo dye. Chemical Engineering Journal 187: 79-88.
17. Tor A, Cengeloglu Y (2006) Removal of Congo red from aqueous solution by adsorption onto acid activated red mud. J Hazard Mater 138: 409-415.
18. Zazouli MA, Yousefi Z, Yazdani-Charati J, Mahdavi Y (2014) Application of Azolla filiculoides Biomass for Acid Black 15 Dye Adsorption from Aqueous Solution. Procedia Engineering 33: 303-312.
19. Deniz F, Karaman S (2011) Removal of Basic Red 46 dye from aqueous solution by pine tree leaves. Chemical Engineering Journal 170: 67-74.
20. El Ashothuiky el SZ (2009) Loofa egiptica as a novel adsorbent for removing direct blue dye from aqueous solution. J Environ Manage 90: 2755-2761.
21. Bayramoglu G, Yakup Arica M (2007) Biosorption of benzidine based textile dyes "Direct Blue 1 and Direct Red 128" using native and heat-treated biomass of Trametes versicolor. J Hazard Mater 143: 135-143.
22. Cardoso NF, Lima EC, Pinto IS, Amavisca CV, Royer B, et al. (2011) Application of cupuassu shell as biosorbent for the removal of textile dyes from aqueous solution. J Environ Manage 92: 1237-1247.
23. Oren Benaroya R, Tzin V, Tei-Or E, Zamski E (2004) Lead accumulation in the aquatic fern Azolla filiculoides: Estimation of equilibrium, thermodynamic and kinetic parameters. Bioresour Technol 99: 3057-3065.
24. Padmesh TVN, Vijayaraghavan K, Sekaran G, Velan M (2006) Application of Azolla rongpong on biosorption of acid red 88, acid green , acid orange 7 and acid blue 15 from synthetic solutions. Chemical Engineering Journal 122: 55-63.
25. Padmesh TVN, Vijayaraghavan K, Sekaran G, Velan M (2005) Batch and column studies on biosorption of acid dyes on fresh water macro alga Azolla filiculoides. J Hazard Mater 125: 121-129.
26. Zazouli MA, Balarak D, Mahdavi Y (2013) Application of Azolla filiculoides biomass for 2-Chlorophenol and 4-Chlorophenol Removal from aqueous solutions. Iranian Journal of Health Science 1: 43-55.
27. Rahman IA, Saad B (2003) Utilization of guava seeds as a source of activated carbon for removal of methylene blue from aqueous solution. Malays J Chem 5: 8–14.
28. Ozdemir FA, Demirata B, Apak R (2009) Adsorptive removal of methylene blue from simulated dyeing wastewater with melanin-formaldehyde–urea resin. J Appl Polym Sci 112: 3442–3448.
29. Balarak D, Pirzadeh F, Mahdavi Y (2015) Biosorption of Acid Red 88 dyes using dried Lemna minor biomass. Journal of Science, Technology & Environmental Informatics 1: 81–90.
30. Zazouli MA, Balarak D, Mahdavi Y, Karimnejad F (2014) The application of Azolla filiculoides biomass in acid blue 15 dye (AB15) removal from aqueous solutions. Journal of Basic Research in Medical Science 1: 29-37.
31. Zazouli MA, Balarak D, Mahdavi Y (2014) Application of Azolla for 6-Trichlorophenol (TCP) Removal from aqueous solutions. Hygiene sciences 2: 17-24.
32. Yan C, Wang C, Yao J, Zhang L, Liu X (2009) Adsorption of methylene blue on mesoporous carbons prepared using acid-treated and alkaline-treated zeolite X as the template. Colloids Surf 333: 115–119.
33. Hameed BH, Ahmad AL, Latiff KNA (2007) Adsorption of basic dye (methylene blue) onto activated carbon prepared from rattan sawdust. Dyes Pigments 75: 143–149.
34. Almeida CA, Debacher NA, Downs AJ, Cottet L, Mello CA (2009) Removal of methylene blue from colored effluents by adsorption on montmorillonite clay. J Colloid Interface Sci 332: 46-53.
35. Weng CH, Pan YF (2007) Adsorption of a cationic dye (methylene blue) onto spent activated clay. J Hazard Mater 144: 355-362.
36. Saeed BB, Rauf MA, AIAI SS (2007) Removal of methylene blue from aqueous solution by adsorption on sand. Dyes Pigments 74: 85–87.
37. Uddin MT, Islam MA, Mahmud S, Rukanuzzaman M (2009) Adsorptive removal of methylene blue by tea waste. J Hazard Mater 164: 53-60.
38. Zazouli MA, Balarak D, Mahdavi Y (2014) Application of Canola Residuals in Adsorption of Reactive Red 198 (RR198) Dye from Aqueous Solutions. Journal of Neyshabur University of Medical Sciences 2: 55-66.
39. Han R, Wang Y, Han P, Shi J, Yang J, et al. (2006) Removal of methylene blue from aqueous solution by chaff in batch mode. J Hazard Mater 137: 550-557.
40. Ozer D, Dursun G, Ozer A (2007) Methylene blue adsorption from aqueous solution by deactivated peanut hull. J Hazard Mater 144: 171-179.
41. Pavan FA, Lima EC, Gias SL, Mazzocatto AC (2008) Methylene blue biosorption from aqueous solutions by yellow passion fruit waste. J Hazard Mater 150: 703-712.
42. Senthilkumar S, Varadarajan PR, Porkodi K, Subbhrasani CV (2005) Adsorption of methylene blue onto jute fiber carbon: kinetics and equilibrium studies. J Colloid Interface Sci 284: 78–82.
43. Zazouli MA, Balarak D, Mahdavi Y (2013) Effect of Azolla filiculoides on removal of reactive red 198 in aqueous solution. J Adv Environ Health Res 1: 1-7.
44. Diyanati RA, Yousefi Z, Cherati JY, Balarak D (2014) Comparison of phenol adsorption rate by modified Canola and Azolla: An Adsorption Isotherm and Kinetics Study. Journal of Health & Development 3: 17-25.
45. Balarak D, Mahdavi Y, Gharibi F, Sadeghi Sh (2014) Removal of hexavalent chromium from aqueous solution using canola biomass: Isotherms and kinetics studies. J Adv Environ Health Res 2: 45-52.