Abstract: Growing evidence showed that inflammation response plays an important role in cancer development and progression, and absolute lymphocyte count (ALC), absolute monocyte count (AMC), and lymphocyte to monocyte ratio (LMR) have been used as parameters of systemic inflammation in several tumors. In this study, we evaluated the prognostic significance of preoperative ALC, AMC and LMR in breast cancer and 2000 patients between January 2002 and December 2008 at Sun Yat-Sen University Cancer Center were enrolled. Patients were grouped by the cut-off value according to the receiver operating characteristics (ROC) curve analysis. Kaplan-Meier analysis showed that patients with elevated AMC levels (>0.48 × 10^9/L) had shorter overall survival (OS, P<0.001). In multivariate analysis, preoperative AMC was identified as an independent prognostic parameter for OS in breast cancer patients (hazard ratio = 1.374, 95% confidence interval: 1.045–1.807). Subgroup analyses revealed that AMC was an unfavorable prognostic factor in stage II–III breast cancer patients and Luminal B, human epithelial growth factor receptor-2 overexpressing subtype, and triple-negative breast cancer (all P<0.05). Additionally, the prognostic value of ALC and LMR could not be proven in the current study. Preoperative AMC may serve as an easily available and low-priced parameter to predict the outcomes of breast cancer.

Editor: Wen Zhou.
Received: October 16, 2015; revised: November 7, 2015; accepted: November 15, 2015.
From the Department of Breast Oncology, Sun Yat-Sen University Cancer Center, State Key Laboratory of Oncology in South China, Collaborative Innovation Center for Cancer Medicine, Guangzhou, Guangdong, P.R. China.
Correspondence: Xiaoming Xie, Department of Breast Oncology, State Key Laboratory of Oncology in South China, Sun Yat-Sen University Cancer Center, 651 East Dongfeng Road, Guangzhou 510060, P.R. China (email: xiem@sysucc.org.cn).
Supplemental Digital Content is available for this article.
This work was supported by funds from the National Natural Science Foundation of China (81472575, 81272513, and 81372133), the Key Programme of the National Natural Science Foundation of China (31300601), the Science and Technology Planning Project of Guangdong and Guangzhou (2013B060300009 and 20144100169).
The authors have no conflicts of interest to disclose.
Copyright © 2015 Wolters Kluwer Health, Inc. All rights reserved.
The work cannot be used commercially.
ISSN: 0025-7974
DOI: 10.1097/MD.0000000000002266

INTRODUCTION

Breast cancer is by far the most common type of cancer in women and becoming a health concern globally. There were approximately 232,670 new cases and 40,000 deaths from breast cancer in the United States in 2014.1 The improvement in detection and various treatments, such as surgical resection, adjuvant chemotherapy, hormonal therapy, targeted therapy, and radiation therapy, are responsible for the improved survival and reduced risk of recurrence.2,3 Classified according to the tumor node metastasis (TNM) staging system and their molecular features,4,5 breast cancer patients received various treatment options and experienced different prognoses. However, recurrence and metastasis still remain big problems for cure, especially for patients in advanced stages.6

Currently, besides the tumor-related factors, the host-related factors are considered as important factors in determining cancer recurrence and survival. Systemic inflammatory response has long been associated with tumor development.7–9 Cancer-associated inflammation is potentially implicated in the process of proliferation and metastasis, promoting angiogenesis, restraining antitumor immunity, and inducing subsequent poor prognosis.10–12 Furthermore, cytokines produced by tumor cells, such as granulocyte colony stimulating factor (G-CSF) and interleukin (IL)-6, could stimulate the proliferation of leukocytes and the latter in turn link to the progression of cancer.13 Lymphocytes and monocytes are essential inflammatory cells in the systemic inflammatory response.14,15 By inducing cytotoxic cell death and inhibiting tumor cell proliferation, lymphocyte could serve as tumor suppressors.9,16 Conversely, monocyte could differentiate into tumor-associated macrophages (TAMs) at tumor sites and promote the invasive and metastatic ability of tumor cells by constructing tumor microenvironment.17,18 A low pretreatment lymphocyte to monocyte ratio (LMR), indicating a lower lymphocyte count and a higher monocyte count, is found to link with poor prognosis in colon cancer, lung cancer, and hematological malignancies.19–21

However, the link between peripheral LMR and breast cancer is not completely defined. Therefore, we performed a retrospective cohort study on breast cancer patients underwent surgical treatment and investigated the prognostic value of preoperative peripheral lymphocyte, monocyte, and LMR for breast cancer.

MATERIALS AND METHODS

Study Population

Patient histologically confirmed as primary breast cancer between January 2002 and December 2008 in Sun Yat-Sen
University Cancer Center (SYSUCC) in Guangzhou, China were retrospectively reviewed. The inclusion criteria were as follows: received surgical treatment; female; and pathologic diagnosis as invasive ductal carcinoma or invasive lobular carcinoma. Exclusion criteria included: received neoadjuvant chemotherapy or radiotherapy before surgery; had surgical treatment before admission; with previous or coexisting cancers other than breast cancer; confirmed metastasis; current or potential inflammation: neutrophilic granulocyte percentage >70% or C-reactive protein >10 mg/L; and not enough data can be extracted. All patients were followed up to December 31, 2014 or date of deaths from any causes.

Clinical Data Collection
Baseline characteristics including age, menstrual status, pathological diagnosis, histologic grade, axillary lymph node status, hormonal receptor, and human epithelial growth factor receptor-2 (HER-2) status, date of last follow-up or death, and preoperative peripheral lymphocyte and monocyte count were collected. Blood sample was collected on an empty stomach and sent to the clinical laboratory. Complete blood count was performed as part of routine clinical evaluation prior to the surgery. The clinical stage was determined by the TNM staging system according to the American Joint Committee on Cancer. The intrinsic subtypes were as follow: Luminal A (estrogen receptor [ER]+, progesterone receptor [PR]+, HER-2−, and Ki-67 ≤14%), Luminal B (ER+ and HER-2+ or Ki-67 >14%), HER-2 overexpressing (ER−, PR−, HER-2+ and triple-negative breast cancer (ER−, PR−, HER-2−)), HER-2 positive was defined as “3+” in immunohistochemical test or “positive” in HER-2 fluorescence in situ hybridization test. The follow-up of patients was performed through out-patient medical records, telephone, or letters by Department of Follow-Up and Medical Record Management.

Statistical Analyses
Preoperative peripheral lymphocyte and monocyte count are expressed as means (± standard deviation), and categorical data were described using numbers and percentages. The LMR was defined as the absolute lymphocyte count (ALC) divided by the absolute monocyte count (AMC). Young breast cancer patient was defined as <35 years old. The endpoints assessed were overall survival (OS), calculated from the time of pathological diagnosis to the date of death from any causes or last follow-up. The relationship between patients’ characteristics and the ALC, AMC, and LMR were assessed by unpaired t test or 1-way analysis of variance (ANOVA), and difference between categories was examined using the Chi-squared test. The clinical significance and the optimal cut-off value of ALC, AMC and LMR were determined by the receiver operating characteristic (ROC) curve analysis, and patients were stratified into 2 ranges according to the cut-off value. Kaplan–Meier method was performed for survival analyses and compared by log-rank test. Univariate analyses and multivariate analyses (Cox proportional hazards model) were performed to determine the influence of potential confounding factors on OS. Hazard ratios (HRs) and the corresponding 95% confidence intervals (CIs) estimated from the Cox analysis were regarded as relative risks, and a 2-tailed P value <0.05 was considered significant. All statistical analyses were performed using SPSS 19.0 (SPSS, Inc., Chicago, IL) in the present study.

RESULTS
The 10-year OS rate was 77.6% for all 2000 patients, and the mean survival time was 123.6 months (95% CI: 121.6–125.5). In the univariate analysis, AMC and LMR were both

Association of ALC, AMC and LMR With OS
FIGURE 1. Flow chart of the patient selection.}

**Ethics Statement**
The study protocol was approved by the Independent Ethical Committee/Institutional Review Board of SYSUCC, and written informed consent was obtained from each participant prior to surgery. All patients were anonymous and de-identified prior to analysis.

**Patient Characteristics**
A total of 3156 consecutive patients with histopathologically confirmed breast cancer in SYSUCC were reviewed, and 2000 patients were finally enrolled after screening process (Figure 1) and no inflammatory breast cancer patient was included. Baseline clinicopathological characteristics are shown in Table 1. The median follow-up time was 75 months (range 3–144 months), and death occurred in 326 (16.3%) of the 2000 breast cancer patients. The median age of the enrolled patients was 49.4 years (range: 22–94 years), and 168 (8.4%) patients were under age 35. The mean ALC and AMC were 2.11 ± 0.62 (×10^9/L) and 0.47 ± 0.23 (×10^9/L), respectively, with a mean LMR of 5.10 ± 2.77.

**The Optimal Cut-Offs of ALC, AMC and LMR**
ROC curve analysis was performed to determine the optimal cut-off values for ALC, AMC and LMR (Figure 2). The cut-off values of ALC, AMC and LMR were 2.20 × 10^9/L, 0.48 × 10^9/L, and 3.80, respectively, with highest Youden’ s index. Enrolled patients were stratified into 2 levels (low- and high-) according to cut-off points. One thousand one hundred fifty (57.7%) patients were categorized as low-ALC group and 850 (42.5%) patients were categorized as high-ALC group. Similarly, 1038 (51.9%) patients were categorized as low-AMC group, and 590 (29.5%) patients were categorized as low-AMC group while the remaining 1092 (48.1%) patients as high-AMC group. The remaining 962 (48.1%) patients were categorized as high-AMC group, and 590 (29.5%) patients were categorized as low-LMR group while the remaining 1410 (70.5%) patients as high-LMR group. No correlation was identified between AMC level and age, menopausal status, tumor types, histologic grades, tumor sizes, lymph node status, ER/PR status, and HER-2 status (all P >0.05, Table 1). Patients who experienced poor outcome had significantly increased AMC compared with patients better prognosis (P <0.001).
significantly associated with OS in breast cancer patients with the HR was 1.565 and 0.776, respectively (both \( P < 0.05 \)). The HR for ALC was 1.166 (95% CI: 0.938–1.449, \( P = 0.167 \)) and no prognostic significance was proven. Other identified prognostic factors for OS included menstrual status, tumor size, lymph node status, ER/PR status, and HER-2 status (Table 2).

To identify independent prognostic factors for OS, multivariate analysis by the Cox proportional hazard model was performed and the AMC retained independent significance (HR = 1.374, 95% CI: 1.045–1.807, \( P = 0.023 \)). Survival analysis showed that breast cancer patients with higher monocyte count (>0.48 \( \times 10^9/\text{L} \)) had a significantly poorer survival than patients with lower monocyte count (116.0 vs 127.6 months, \( P < 0.001 \); Figure 3). No statistical significance of the prognostic effect of LMR was observed in the multivariate analysis (\( P = 0.236 \)). Other prognostic factors included menstrual status, tumor size, lymph node status, ER status, and HER-2 status.

In multivariate analysis stratified by clinical stages, elevated AMC was significantly related to worse OS in stage II–III breast cancer patients (HR = 1.450 and 1.693, respectively, both \( P < 0.05 \); Table 3), while no statistical significance was proven in the stage I patients (\( P = 0.231 \)). Meanwhile, the AMC was indicated as risk factor for breast cancer patients of Luminal B, HER-2 overexpressing subtype, and triple-negative breast cancer subtypes (all \( P < 0.05 \); Table 3). The clinical outcomes of Luminal A breast cancer patients with high monocyte counts were not significantly different with patients with low monocyte counts (\( P = 0.170 \)).

**DISCUSSION**

Growing evidence has suggested an important role of inflammation in cancer development, tumor angiogenesis, and metastasis. Chronic oxidative stress and the oxygen free radicals caused by the inflammatory response could lead to cancer initiation and promotion.22,23 As a marker of systemic inflammation, preoperative leukocyte levels are considered to be in association with prognosis in various carcinoma.24,25 The leukocytic cell ratios, including neutrophil to lymphocyte ratio,
platelets to lymphocyte ratio, and LMR, are routinely available markers of the systemic inflammatory response and identified as prognostic parameters in several solid tumors and leukemia.26–29

In the present study, we retrospectively analyzed 2008 consecutive breast cancer patients who received surgeries as primary treatments and found that: only AMC was an independent prognostic factor for patients with breast cancer, the prognostic value of ALC and LMR could not be confirmed \( (P = 0.241 \text{ and } P = 0.236) \); high AMC levels \( (>0.48 \times 10^9/L) \) were associated with poor clinical outcome in more advanced stages (II–III) and Luminal B, HER-2 overexpressing, and triple-negative breast cancer subtypes. Previous studies have shown a link between monocyte and cancer progression. The prognostic value of monocyte was reported not only in lymphomas, but also in nonhematologic malignancies.30,31 Increase of circulating monocyte was associated with poor progression-free survival and OS in nonsmall cell lung cancer, gynecological cancer, and gastrointestinal cancer.32–34

The possible mechanism of the independent association between circulating monocyte and cancer prognosis may be related to the activation of innate immunity during the tumor progression.34 Act as chemotactic factor, the monocyte chemotactic protein 1 is up regulated in tumors and can induce the increase of monocyte lineage cells in the tumor site directly, which usually correlate with a more advanced stage of cancer.35 Furthermore, the inflammatory monocytes recruited into tumor sites by the CCL2/CCR2 chemokine axis can differentiate into TAMs\(^{17}\) and become a major component of the tumor microenvironment. Positive correlation between the TAM count and

### TABLE 2. Univariate and Multivariate Analyses of AMC and LMR for OS in Breast Cancer

| Variable               | Univariate Analysis | Multivariate Analysis |
|------------------------|---------------------|-----------------------|
|                        | HR (95% CI)         | P Value               | HR (95% CI)         | P Value               |
| Age                    | 0.707 (0.497–1.003) | 0.052                 | 0.863 (0.598–1.248) | 0.434                 |
| Menstrual status       | 0.762 (0.604–0.962) | 0.022                 | 0.649 (0.508–0.830) | 0.001                 |
| Tumor type             | 0.796 (0.395–1.606) | 0.525                 | 0.723 (0.351–1.490) | 0.380                 |
| Histologic grade       |                     |                       |                      |                       |
| G1                     | 1 (reference)       | 1 (reference)         | 0.994 (0.363–2.727) | 0.991                 |
| G2                     | 1.807 (0.665–4.908) | 0.246                 | 1.329 (0.483–3.660) | 0.582                 |
| G3                     | 3.218 (1.188–8.731) | 0.022                 | 2.078 (1.412–3.059) | 0.001                 |
| Tumor size             |                     |                       |                      |                       |
| T1                     | 1 (reference)       | 1 (reference)         | 1.178 (0.920–1.509) | 0.194                 |
| T2                     | 1.533 (1.203–1.953) | 0.001                 | 2.078 (1.412–3.059) | 0.001                 |
| T3                     | 4.076 (2.826–5.879) | <0.001               | 2.078 (1.412–3.059) | 0.001                 |
| Lymph node status      |                     |                       |                      |                       |
| N0                     | 1 (reference)       | 1 (reference)         | 2.247 (1.623–3.111) | <0.001               |
| N1                     | 2.218 (1.607–3.060) | <0.001               | 3.886 (2.785–5.422) | <0.001               |
| N2                     | 3.984 (2.877–5.517) | <0.001               | 7.264 (5.244–10.062) | <0.001               |
| N3                     | 8.529 (6.244–11.649)| <0.001               | 0.674 (0.516–0.880) | 0.004                 |
| ER                     | 0.562 (0.452–0.698) | <0.001               | 0.945 (0.728–1.227) | 0.671                 |
| PR                     | 0.636 (0.510–0.793) | <0.001               | 1.446 (1.119–1.869) | 0.005                 |
| HER-2                  | 2.091 (1.665–2.626) | <0.001               | 1.167 (0.901–1.512) | 0.241                 |
| ALC                    | 1.166 (0.938–1.449) | 0.167                 | 1.167 (0.901–1.512) | 0.241                 |
| AMC                    | 1.565 (1.255–1.953) | 0.001                 | 1.374 (1.045–1.807) | 0.023                 |
| LMR                    | 0.776 (0.618–0.975) | 0.029                 | 0.840 (0.629–1.121) | 0.236                 |

**Note:** ALC = absolute lymphocyte count, AMC = absolute monocyte count, CI = confidence interval, ER = estrogen receptor, HER-2 = human epidermal growth factor receptor-2, HR = hazard ratio, LMR = lymphocyte to monocyte ratio, OS = overall survival, PR = progesterone receptor.

---

**FIGURE 2.** ROC curves assessing the cut-off of ALC, AMC, and LMR for predicting the overall survival in the cohort study. The AUCs for each parameter were 0.513 \((P = 0.461)\), 0.562 \((P = 0.001)\), and 0.459 \((P = 0.02)\), respectively. ALC = absolute lymphocyte count, AMC = absolute monocyte count, LMR = lymphocyte to monocyte ratio, OS = overall survival, ROC = receiver operating characteristic.
circulating monocytes is demonstrated in previous study. By producing a great diversity of cytokines, such as VEGFA, MMP-9 and PDGF, TAMs can promote the invasion and metastasis of cancer cells. Coculture of cancer cells with macrophages can enhance cancer cell matrix-degrading activity, promote the invasive potential of cancer cells, which correlate with poor prognosis in cancers.

In the subgroup analysis based on different clinical stages and intrinsic subtypes, the predictive value of AMC was significant in patients with stage II–III (both \( P < 0.05 \)), while the stage I patients showed no statistical significance (\( P = 0.231 \)). It is in line with the idea that the monocyte mainly plays roles in the cancer invasion and metastasis, not in the proliferation of the tumor cell. That may be the possible reason for no prognostic effect of AMC in the early stage breast cancer patients.

The prognostic value of AMC was statistically significant in Luminal B, HER-2 overexpressing subtype, and triple negative breast cancer subtypes, not in Luminal A subtype. Since the OS was distinctly different among various subtypes, and patients of Luminal A subtype experience best OS. HER-2 is considered as an oncogene of the MAP-Kinase and PI3K/AKT/mTOR signaling pathways leading to cell growth and proliferation. Previous studies indicate that patients with ER/PR-negative experienced higher risk of cancer relapse than those with ER/PR-positive. Thus monocyte may contact with tumor cells and assist the metastasis in the more invasive subtypes.

In our study cohort, the ALC was not identified as an independent favorable prognostic factor in breast cancer. The reason may be that different lymphocyte subsets manifest various effects in tumor development and metastasis. In breast cancer and lung cancer, CD8\(^+\) T lymphocytes were considered as antitumor lymphocyte and have favorable effect on cancer patients. Conversely, CD4\(^+\)CD25\(^+\)Foxp3\(^+\) regulatory T lymphocytes (Tregs) can inhibit cytotoxic responses and associate with poor clinical outcome. Thus the ALC could not accurately reflect the prognostic effect of lymphocytes in cancer patients.

AMC provides an easily available and low-priced biomarker for prognostic evaluation. However, several limitations existed in the present study. Firstly, because of the retrospective nature of the study, selection bias could not be excluded even though consecutive patients were enrolled. Secondly, although we excluded patients with potential inflammation by neutrophilic granulocyte percentage and CRP levels, potential confounding factors, such as autoimmune disease and hepatic dysfunction, still may still existed and affected the circulating monocyte counts. Finally, specific quality control analysis was performed by different quality inspectors since the white blood cell count was obtained as a routine clinical test before surgery.

In conclusion, our study suggested that high pretreatment count of circulating monocyte is independently associated with poor prognosis in breast cancer patients. And large, prospective studies are needed to validate the prognostic effect of circulating monocyte in the future.

### TABLE 3. Prognostic Impact of AMC in Various Clinical Stages and Intrinsic Subtypes

| Subgroup               | Univariate Analysis | Multivariate Analysis |
|------------------------|---------------------|-----------------------|
|                        | HR (95% CI)         | \( P \) Value         | HR (95% CI)         | \( P \) Value         |
| Clinical stages\(^a\) |                     |                       |                     |                       |
| I                      | 0.802 (0.352–1.829) | 0.600                 | 1.074 (0.418–2.759) | 0.231                 |
| II                     | 1.510 (1.054–2.162) | 0.024                 | 1.450 (1.011–2.080) | 0.043                 |
| III                    | 1.747 (1.296–2.355) | 0.001                 | 1.693 (1.254–2.286) | 0.001                 |
| Intrinsic subtypes\(^b\) |                    |                       |                     |                       |
| Luminal A              | 1.308 (0.853–2.004) | 0.218                 | 1.350 (0.879–2.075) | 0.170                 |
| Luminal B              | 1.545 (1.106–2.160) | 0.011                 | 1.531 (1.094–2.142) | 0.013                 |
| HER-2 overexpressing   | 1.860 (1.093–3.164) | 0.022                 | 1.772 (1.015–3.094) | 0.044                 |
| Triple-negative        | 1.923 (1.004–3.684) | 0.049                 | 2.183 (1.102–4.325) | 0.025                 |

AMC = absolute monocyte count, CI = confidence interval, HER-2 = human epithelial growth factor receptor-2, HR = hazard ratios.

\(^a\) Tumor size and lymph node status were not involved in the multivariate analyses.

\(^b\) Hormone receptor status and HER-2 status were not involved in the multivariate analyses.
REFERENCES

1. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2015. CA Cancer J Clin. 2015;65:5–29.

2. Fedele P, Orlando L, Schiavone P, et al. Recent advances in the treatment of hormone receptor positive HER2 negative metastatic breast cancer. Crit Rev Oncol Hematol. 2015;94:291–301.

3. Moran MS. Radiation therapy in the locoregional treatment of triple-negative breast cancer. Lancet Oncol. 2015;16:e113–e122.

4. Carey LA, Perou CM, Livasy CA, et al. Race, breast cancer subtypes, and survival in the Carolina Breast Cancer Study. JAMA. 2006;295:2492–2502.

5. Singletary SE, Allred C, Ashley P, et al. Revision of the American Joint Committee on Cancer staging system for breast cancer. J Clin Oncol. 2002;20:3626–3636.

6. Carlson RW, Allred DC, Anderson BO, et al. Metastatic breast cancer, version 1.2012: featured updates to the NCCN guidelines. J Natl Compr Canc Netw. 2012;10:821–829.

7. McMillan DC. Systemic inflammation, nutritional status and survival in patients with cancer. Curr Opin Clin Nutr Metab Care. 2009;12:223–226.

8. Roxburgh CS, McMillan DC. Role of systemic inflammatory response in predicting survival in patients with primary operable cancer. Future Oncol. 2010;6:149–163.

9. Coussens LM, Werb Z. Inflammation and cancer. Nature. 2002;420:860–867.

10. Iyengar NM, Hudis CA, Dannenberg AJ. Obesity and inflammation: new insights into breast cancer development and progression. Am Soc Clin Oncol Educ Book. 2013:46–51.

11. Colotta F, Allavena P, Sica A, et al. Cancer-related inflammation, the seventh hallmark of cancer: links to genetic instability. Carcinogenesis. 2009;30:1073–1081.

12. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell. 2011;144:646–674.

13. Kasuga I, Makino S, Kiyokawa H, et al. Tumor-related leukocytosis is linked with poor prognosis in patients with lung carcinoma. Cancer Am Cancer Soc. 2001;92:2399–2405.

14. Delogu G, Fanumero G, Tellan G, et al. Lymphocyte apoptosis, caspase activation and inflammatory response in septic shock. Infection. 2008;36:485–487.

15. Gillette DD, Trindandapani S, Butchar JP. Monocyte/macrophage inflammatory response pathways to combat Francisella infection: possible therapeutic targets? Front Cell Infect Microbiol. 2014;4:18.

16. Mantovani A, Allavena P, Sica A, et al. Cancer-related inflammation. Nature. 2008;454:436–444.

17. Qian BZ, Li J, Zhang H, et al. CCL2 recruits inflammatory monocytes to facilitate breast-tumour metastasis. Nature. 2011;475:222–225.

18. Chen JJ, Lin YC, Yao PL, et al. Tumor-associated macrophages: the double-edged sword in cancer progression. J Clin Oncol. 2005;23:953–964.

19. Stotz M, Pichler M, Absenger G, et al. The preoperative lymphocyte to monocyte ratio predicts clinical outcome in patients with stage III colon cancer. Br J Cancer. 2014;110:435–440.

20. Munoz E, Goldstein J, Benacquista T, et al. Diagnosis related group “all payer” hospital payment and medical diseases. Financial risk and hospital cost in medical noncomplicating condition-stratified diagnosis related groups. Arch Intern Med. 1989;149:417–420.

21. Wei X, Huang F, Wei Y, et al. Low lymphocyte-to-monocyte ratio predicts unfavorable prognosis in non-germinal center type diffuse large B-cell lymphoma. Leuk Res. 2014;38:694–696.

22. Salim AS. The permissive role of oxygen-derived free radicals in the development of colonic cancer in the rat. A new theory for carcinogenesis. Int J Cancer. 1993;53:1031–1035.

23. Hussain SP, Aguilar F, Amstad P, et al. Oxy-radical induced mutagenesis of hotspot codons 248 and 249 of the human p53 gene. Oncogene. 1994;9:2277–2281.

24. Tibaldi C, Vasile E, Bernardini I, et al. Baseline elevated leukocyte count in peripheral blood is associated with poor survival in patients with advanced non-small cell lung cancer: a prognostic model. J Cancer Res Clin Oncol. 2008;134:1143–1149.

25. Schmidt H, Baustolt L, Geertsen P, et al. Elevated neutrophil and monocyte counts in peripheral blood are associated with poor survival in patients with metastatic melanoma: a prognostic model. Br J Cancer. 2005;93:273–278.

26. Guthrie GJ, Charles KA, Roxburgh CS, et al. The systemic inflammation-based neutrophil-lymphocyte ratio: experience in patients with cancer. Crit Rev Oncol Hematol. 2013;88:218–230.

27. Cihan YB, Ozturk A, Mutlu H. Relationship between prognosis and neutrophil: lymphocyte and platelet: lymphocyte ratios in patients with malignant pleural mesotheliomas. Asian Pac J Cancer Prev. 2014;15:2061–2067.

28. Walsh SR, Cook EJ, Goulder F, et al. Neutrophil-lymphocyte ratio as a prognostic factor in colorectal cancer. J Surg Oncol. 2005;91:181–184.

29. Li ZM, Huang J, Xia Y, et al. Blood lymphocyte-to-monocyte ratio identifies high-risk patients in diffuse large B-cell lymphoma treated with R-CHOP. PLoS ONE. 2012;7:e14658.

30. Tsai YD, Wang CP, Chen CY, et al. Pretreatment circulating monocyte count associated with poor prognosis in patients with oral cavity cancer. Head Neck. 2014;36:947–953.

31. Roche NM, Kottschade LA, Grotz TE, et al. The prognostic role of the preoperative absolute lymphocyte count and absolute monocyte count in patients with resected advanced melanoma. Am J Clin Oncol. 2015;38:252–258.

32. Botta C, Barbieri V, Ciliberto D, et al. Systemic inflammatory status at baseline predicts bevacizumab benefit in advanced non-small cell lung cancer patients. Cancer Biol Ther. 2013;14:469–475.

33. Lee YY, Choi CH, Sung CO, et al. Prognostic value of pre-treatment circulating monocyte count in patients with cervical cancer: comparison with SCC-Ag level. Gynecol Oncol. 2012;124:92–97.

34. Bishara S, Griffin M, Cargill A, et al. Pre-treatment white blood cell subtypes as prognostic indicators in ovarian cancer. Eur J Obstet Gynecol Reprod Biol. 2008;138:71–75.

35. Marcus B, Arenberg D, Lee J, et al. Prognostic factors in oral cavity and oropharyngeal squamous cell carcinoma. Cancer Am Cancer Soc. 2004;101:2779–2787.

36. Koh YW, Kang HJ, Park C, et al. The ratio of the absolute lymphocyte count to the absolute monocyte count is associated with prognosis in Hodgkin’s lymphoma: correlation with tumor-associated macrophages. Onkolog. 2012;17:871–880.

37. Wang R, Zhang J, Chen S, et al. Tumor-associated macrophages provide a suitable microenvironment for non-small lung cancer invasion and progression. Lung Cancer. 2011;74:188–196.

38. Park KY, Li G, Platt MO. Monocyte-derived macrophage assisted breast cancer cell invasion as a personalized, predictive metric to score metastatic risk. Sci Rep. 2015;5:13855.

39. Yarden Y, Sliwkowski MX. Untangling the ErbB signalling network. Nature. 2001;2:127–137.

40. Yakes FM, Chinratanalab W, Ritter CA, et al. Herceptin-induced inhibition of phosphorylation/inositol-3 kinase and Akt is required for antibody-mediated effects on p27, cyclin D1, and antitumor action. Cancer Res. 2002;62:4132–4141.
41. Vaz-Luis I, Ottesen RA, Hughes ME, et al. Impact of hormone receptor status on patterns of recurrence and clinical outcomes among patients with human epidermal growth factor-2-positive breast cancer in the National Comprehensive Cancer Network: a prospective cohort study. Breast Cancer Res. 2012;14:R129.

42. Kawai O, Ishii G, Kubota K, et al. Predominant infiltration of macrophages and CD8(+) T cells in cancer nests is a significant predictor of survival in stage IV nonsmall cell lung cancer. Cancer Am Cancer Soc. 2008;113:1387–1395.

43. Mahmoud SM, Paish EC, Powe DG, et al. Tumor-infiltrating CD8+ lymphocytes predict clinical outcome in breast cancer. J Clin Oncol. 2011;29:1949–1955.

44. Tang Y, Xu X, Guo S, et al. An increased abundance of tumor-infiltrating regulatory T cells is correlated with the progression and prognosis of pancreatic ductal adenocarcinoma. PLoS ONE. 2014;9:e91551.