A nomogram to predict the probability of axillary lymph node metastasis in female patients with breast cancer in China: A nationwide, multicenter, 10-year epidemiological study

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ABSTRACT

Axillary lymph node dissection (ALND) or sentinel lymph node biopsy (SLNB) alone may lead to postoperative complications. Among patients with positive ALN in the preoperative examination, approximately 40% patients do not have SLN metastasis. Herein, we aimed to develop a model to predict the probability of ALN metastasis as a preoperative tool to support clinical decision-making. We retrospectively analyzed the clinicopathological features of 4211 female patients with breast cancer who were diagnosed in seven breast cancer centers representing entire China, over 10 years (1999-2008). The patients were randomly categorized into a training cohort or validation cohort (3:1 ratio). Multivariate logistic regression analysis was performed for 1869 patients with complete information on the study variables. Age at diagnosis, tumor size, tumor quadrant, clinical nodal status, local invasion status, pathological type, and molecular subtypes were the independent predictors of ALN metastasis. The nomogram was then developed using the seven variables. Further, it was subsequently validated in 642 patients with complete data on variables in the validation cohort. Coefficient of determination (R²) and the area under the receiver-operating characteristic (ROC) curve (AUC) were calculated to be 0.979 and 0.7007, showing good calibration and discrimination of the model, respectively. The false-negative rates of the nomogram were 0 and 6.9% for the predicted risk cut-off values of 14.03% and 20%, respectively. Therefore, when the predicted risk is less than 20%, SLNB may be avoided. After further validation in various patient populations, this model may support increasingly limited axillary surgery in breast cancer.
INTRODUCTION

Breast cancer is the most common malignancy in women, accounting for 25% of all female cancer cases and 15% of all cancer-related deaths [1]. Recently, breast cancer incidence has plateaued [2]. The metastasis status of axillary lymph nodes (ALN) is an important factor affecting the prognosis of patients with breast cancer, a major component of breast cancer staging, and an important basis for designing treatment programs [3-5]. The sentinel lymph-node biopsy (SLNB) has been rapidly replacing ALN dissection (ALND) to become the standard surgical procedures for early breast cancer patients with clinical negative axillary lymph nodes [6-8].

Although SLNB is landmark progress in the field of surgery and can avoid unnecessary ALND for patients, a discussion of the disadvantages related to SLNB should not be omitted. Because of the need to assess the pathological state of SLN during surgery, this procedure is time consuming and expensive. Besides, although it causes less damage than ALND [9, 10], SLNB involves a certain degree of side injury, including upper limb edema, shoulder and back pain, arm numbness [11], weakened shoulder, and reduced arm strength [12]. Therefore, with either ALND or SLNB alone, postoperative associated complications may occur. In addition, studies have reported that among the patients with absence of ALN on preoperative clinical examination, more than 60% patients do not have SLN metastasis. Further, even in the patients with ALN on preoperative clinical examination, approximately 40% patients do not have SLN metastasis [13]. Therefore, it is important to screen patients with ALN and identify patients without SLN metastasis before surgery in order to avoid unnecessary SLNB. To this effect, researchers are attempting to determine methods to avoid unnecessary SLNB or ALND.

Medical Centers outside China have published models to predict the ALN status in patients. For example, the Memorial Sloan Kettering Cancer Center (MSKCC) developed a nomogram that was used for preoperative prediction of SLN status and prediction of non-SLN status when SLN was present. This model was verified in many medical centers and widely accepted, as it helped clinicians decide the surgical procedure required for regional lymph nodes before the surgery. However, there were several inconsistencies in the verification results between populations, which could have occurred due to differences in race, social and cultural background, level of economic development, level of medical care, and many other factors [14-19].

Prediction models are often built using clinical and pathological data of a specific population. Therefore, when used to predict disease in another group of people, their predictive value is limited. To our knowledge, the report on the establishment of prediction model of ALN metastasis in China was few at present, and these models included data from single-center studies that were not fully representative of the entire population of China.

Therefore, in this study, we aimed to (1) represent the entire population of China by retrospectively analyzing relevant medical records of female patients with breast cancer who were diagnosed over a period of 10 years, (2) determine the risk factors of ALN metastasis in breast cancer, and (3) build a prediction model of ALN metastasis in breast cancer in order to help clinicians in the decision-making process.

RESULTS

Clinicopathologic features and grouping of patients

Of the 4,211 patients, 3158 were included in the training cohort and 1053 were included in the validation cohort in a 3:1 ratio. The clinical and pathological data of the patients between the two groups did not differ significantly (p > 0.05), which was consistent with the randomization. Among patients who underwent SLNB and ALND, 48.74% (1426/2926) had ALN metastasis in the training cohort and 49.59% (483/973) had ALN metastasis in the validation cohort (Table 1).

Univariate logistic regression analysis of ALN metastasis in the training cohort

Univariate logistic regression analysis was used to explore ALN metastasis-related variables (Table 2) and showed that age, tumor size, primary tumor quadrant, clinical nodal status, local invasion status, pathological type, ER status, and molecular subtypes were related to breast cancer ALN metastasis (p < 0.05).

Processing of missing data and multivariate logistic regression analysis of ALN metastasis in the modeling group

Because of the longer duration of data collection, a large amount of data and collecting information, partial data were missing. We found no significant difference in the clinical and pathological features of patients with missing data between the two groups (p > 0.05; Table 3). Further, patients who did not undergo ALND or SLNB were excluded (n = 232 in the training cohort [7.35%] and n = 79 in the validation cohort [7.5%]). Considering that molecule subtype included ER status, the variable ER was excluded in multivariate regression analysis. Finally, 1869 and 642 patients with complete data on age, tumor size, primary tumor quadrant, clinical lymph nodes, local invasion status, pathological type, molecular
Figure 1: Geographic distribution of sites included in the study. The numbers in the map represent the following: 1: Cancer Institute/Hospital, Chinese Academy of Medical Sciences, 2: Liaoning Cancer Hospital, 3: Second Xiangya Hospital, Central South University, 4: Guangdong Sun Yat-Sen University Cancer Center, 5: Zhejiang Cancer Hospital, 6: First Affiliated Hospital of Xi’an Jiaotong University, 7: Sichuan Cancer Hospital [20].

Figure 2: Patients with complete information in the training cohort and the validation cohort. The 7 variables denote age at diagnosis, clinical tumor size, tumor location, clinical lymph node status, local invasion, pathological type, and molecular subtype.
**Table 1: Comparison of the descriptive characteristics between the training cohort and the validation cohort**

| Characteristics       | Training  | %    | Validation | %    | P value |
|-----------------------|-----------|------|------------|------|---------|
| Age at diagnosis (years) | N         | 3158 | 1053       | 0.947|         |
|                       |           | 48.69±10.45 | 48.66±10.52 |       |         |
| BMI (kg/m²)           | N         | 2476 | 805        | 0.256|         |
|                       |           | 23.32±3.24 | 23.47±3.38 |       |         |
| Tumor location        | N         | 2831 | 937        | 0.584|         |
|                       | UIQ       | 493  | 172        | 18.36|         |
|                       | UOQ       | 1359 | 459        | 48.99|         |
|                       | LIQ       | 168  | 60         | 6.4  |         |
|                       | LOQ       | 277  | 95         | 10.14|         |
|                       | central   | 173  | 49         | 5.23 |         |
|                       | others*   | 361  | 102        | 10.89|         |
| Clinical tumor size¹  | N         | 2668 | 898        | 0.592|         |
|                       | T1        | 783  | 277        | 30.85|         |
|                       | T2        | 1572 | 524        | 58.35|         |
|                       | T3        | 313  | 97         | 10.8 |         |
| Local invasion²       | N         | 2745 | 918        | 0.842|         |
|                       | yes       | 136  | 47         | 5.12 |         |
|                       | no        | 2609 | 871        | 94.88|         |
| Pathological type     | N         | 3001 | 1013       | 0.776|         |
|                       | DCIS-Mi   | 93   | 30         | 2.96 |         |
|                       | IDC       | 2585 | 873        | 86.18|         |
|                       | ILC       | 105  | 30         | 2.96 |         |
|                       | others**  | 218  | 80         | 7.9  |         |
| Clinical lymph node status³ | N        | 2803 | 907        | 0.080|         |
|                       | N0        | 1704 | 599        | 66.04|         |
|                       | N1-N3     | 1099 | 308        | 33.96|         |
| ER                    | N         | 2641 | 893        | 0.370|         |
|                       | Positive  | 1527 | 501        | 56.1 |         |
|                       | Negative  | 1114 | 392        | 43.9 |         |
| PR                    | N         | 2641 | 893        | 0.306|         |
|                       | Positive  | 1551 | 507        | 56.77|         |
|                       | Negative  | 1090 | 386        | 43.23|         |
| HR                    | N         | 2641 | 893        | 0.881|         |
|                       | Positive  | 1788 | 607        | 67.97|         |
|                       | Negative  | 853  | 286        | 32.03|         |
| HER-2 receptor status | N         | 2131 | 718        | 0.589|         |
|                       | Positive  | 556  | 180        | 25.07|         |
|                       | Negative  | 1575 | 538        | 74.93|         |
| Molecular subtype     | N         | 2447 | 830        | 0.604|         |
|                       | LM        | 1788 | 607        | 73.13|         |
|                       | HER2+     | 219  | 66         | 7.95 |         |
|                       | TN        | 440  | 157        | 18.92|         |
subtypes were included in the training cohort and validation cohort, respectively (Figure 2). Multivariate analysis confirmed that age, tumor size, primary tumor quadrant, clinical nodal status, invasion of the chest wall and skin, pathological type, and molecular subtype were independent predictors of ALN metastasis (Table 4).

Establishment of a prediction model for ALN metastasis

According to the results of multivariate analysis, the following seven variables were included in the prediction model of ALN metastasis: age, tumor size, primary tumor quadrant, clinical nodal status, local invasion status, pathological type, and molecular subtype. The weights of each variable in the model corresponded to different points (Figure 3). Points for the following factors were added to the total points, which corresponded to the linear predictors and risk predictors of ALN metastasis (Figure 3): size (T1, 0; T2, 7; T3, 23), location (LIQ, 0; UIQ, 17; UOQ, 31; LOQ, 41; central, 49; others, 27), invasion (no, 0; yes, 26), lymph node (no, 0; yes, 42), pathology (DCIS-Mi, 0; ILC, 98; IDC, 100; others, 72), subtype (TN, 0; HER2+, 5; LM, 11). According to the results of multivariate logistic regression analysis, the ALN metastasis risk of patients was expressed by the following equation:

$$\ln(p/(1-p)) = -0.014 \times a + 0.204 \times b2 + 0.663 \times b3 - 0.944 \times c1 - 0.529 \times c2 - 1.444 \times c3 - 0.237 \times c4 - 0.642 \\
\times c5 + 0.768 \times d + 1.235 \times e + 2.944 \times f1 + 2.884 \times f2 + 2.111 \times f3 + 0.322 \times g1 + 0.141 \times g2 - 2.483$$

where “p” represents the risk of ALN metastasis, “a” represents age at diagnosis, “b” represents tumor size (b2. T2; b3.T3), “c” represents tumor site (c1.UIQ; c2.UOQ; c3.LIQ; c4.LOQ; c5.others), “d” represents local invasion; “e” represents clinical lymph node status, “f” represents pathological type (f1.IDC; f2.ILC; f3.others), and “g” represents the molecular subtype (g1.LM; g2. HER2 +). This model was retrospectively utilized for patients in the training cohort (n = 1869), with an AUC value of 0.7157 (Figure 4), suggesting that it had a good predictive ability.

Prospective applications of the prediction model of ALN metastasis

This prediction model of ALN metastasis was prospectively used for patients in the validation cohort. It depicts the ROC curve, and the AUC value calculated was 0.7007 (Figure 5), indicating a good predictive ability. As seen in Figure 6, the curvilinear trend of predicted values and the real value was the same; there was no significant deviation, indicating that the predicted risk of ALN metastasis was consistent with the actual metastasis risk. The coefficient of determination represented the accuracy of model, and the R² value of the model was 0.979, suggesting good calibration. On further evaluation of the clinical value of the model using cutoffs, we found that when the cutoff values of 14.03% and 20% were considered, the false-negative rates of model were 0 and 6.9%, respectively (Table 5).

DISCUSSION

Breast cancer ALN status is a key factor in deciding the therapeutic options for patients and affects the prognosis of patients [3-5]. With more in-depth research on breast cancer, researchers consider ALND important for lymph node staging but has a small significance for treatment [28]. Therefore, if the ALN status of patients with breast cancer can be assessed in a noninvasive and accurate manner, clinicians can avoid
surgical trauma and associated complications for low-risk patients with lymph node metastasis. Clinical research in medical centers in China and abroad has focused on the prediction of ALN metastasis status. In 2003, the MSKCC in the US established two models. The models were validated in many medical centers, but the results were widely inconsistent among different populations [14-19]. Therefore, to improve the accuracy of prediction, some clinicians used preoperative breast ultrasound, mammography photography, and breast magnetic resonance imaging (MRI) to predict the risk of ALN metastasis. However, the false-negative rate of prediction of the ALN status by ultrasound was 16.7-22.9%. After combining with mammography photography, breast MRI, and positron emission tomography/computed tomography (PET/CT), the false-negative rate reduced to 14-16.9% [29, 30]. However, thus far, there is no international consensus on the preoperative routine use of MRI [31]. In China, some clinicians used ultrasound in combination with clinical data of patients to build a prediction model of ALN metastasis and obtained an AUC value of 0.864, indicating a good predictive value [32]. However, the number of patients included in that model (n = 322 for the modeling group and n = 234 for the validation group) was relatively small, and the patients were from a single medical center; therefore, they only represented a small

| Variables                                   | Coefficient | SE  | OR  | 95%CI Lower | 95%CI Upper | P value |
|---------------------------------------------|-------------|-----|-----|-------------|-------------|---------|
| Age at diagnosis (years)                    | 0.008       | 0.004 | 0.992 | 0.992       | 0.999       | 0.037   |
| BMI (kg/m2)                                 | 0.011       | 0.013 | 1.101 | 0.985       | 1.037       | 0.399   |
| Local invasion                              | 0.834       | 0.212 | 2.303 | 1.519       | 3.493       | 0.000   |
| ER                                          | 0.118       | 0.298 | 1.656 | 1.164       | 2.356       | 0.005   |
| PR                                          | 0.002       | 0.081 | 1.002 | 0.856       | 1.174       | 0.976   |
| HR                                          | 0.107       | 0.085 | 1.112 | 0.942       | 1.314       | 0.210   |
| HER-2                                       | 0.201       | 0.212 | 1.223 | 0.981       | 1.824       | 0.066   |
| Multifocality                               | 0.202       | 0.228 | 1.224 | 0.783       | 1.914       | 0.376   |
| Tumor location                              |             |     |     |             |             |         |
| UIQ versus Central                          | -1.004      | 0.190 | 0.366 | 0.253       | 0.531       | 0.000   |
| UOQ versus Central                          | -0.453      | 0.173 | 0.635 | 0.453       | 0.892       | 0.009   |
| LIQ versus Central                          | -0.991      | 0.232 | 0.371 | 0.236       | 0.584       | 0.000   |
| LOQ versus Central                          | -0.229      | 0.205 | 0.796 | 0.533       | 1.188       | 0.264   |
| Others versus Central                       | -0.538      | 0.197 | 0.584 | 0.397       | 0.860       | 0.006   |
| Molecular subtype                           |             |     |     |             |             |         |
| LM versus TN                                | 0.188       | 0.109 | 1.207 | 1.974       | 2.495       | 0.035   |
| HER-2+ versus TN                            | 0.121       | 0.169 | 1.129 | 0.811       | 1.571       | 0.473   |
| Clinical lymph node status                  | 1.436       | 0.088 | 4.203 | 3.534       | 4.997       | 0.000   |
| Clinical tumor size                         |             |     |     |             |             |         |
| T2 versus T1                                | 0.360       | 0.090 | 1.433 | 1.201       | 1.711       | 0.000   |
| T3 versus T1                                | 1.136       | 0.154 | 3.115 | 2.303       | 4.213       | 0.000   |
| Histological type                           |             |     |     |             |             |         |
| IDC versus DCIS-Mi                          | 2.219       | 0.376 | 9.194 | 4.403       | 19.197      | 0.000   |
| ILC versus DCIS-Mi                          | 1.940       | 0.423 | 6.961 | 3.039       | 15.942      | 0.000   |
| Others versus DCIS-Mi                       | 1.476       | 0.402 | 4.376 | 1.991       | 9.615       | 0.000   |

Abbreviations: UIQ, upper inner quadrant; UOQ, upper outer quadrant; LIQ, lower inner quadrant; LOQ, lower outer quadrant; BMI, body mass index; HR, hormone receptor; ER, estrogen receptor; PR, progesterone receptor; DCIS-Mi, ductal carcinoma in situ with micro-invasion; IDC, invasive ductal carcinoma; ILC, invasive lobular carcinoma; LM, luminal-like; TN, triple-negative; HER-2, human epidermal growth factor receptor-2; OR, odds ratio; CI, confidence interval; SE, standard error
Table 3: Comparison of the clinical and pathological features of patients with missing data between the two groups

| Variables                       | Training Percentage (%) | Validation Percentage (%) | P value |
|--------------------------------|-------------------------|---------------------------|---------|
| Age at diagnosis               | 0                       | 0                         | -       |
| Tumor location                 | 327                     | 10.35                     | 116     | 11.02 | 0.545 |
| Clinical tumor size            | 490                     | 15.52                     | 155     | 14.72 | 0.534 |
| Local invasion                 | 413                     | 13.08                     | 135     | 12.82 | 0.830 |
| Pathological type              | 157                     | 4.97                      | 40      | 3.8   | 0.119 |
| Clinical lymph node status     | 445                     | 14.09                     | 146     | 13.87 | 0.855 |
| Molecular subtype              | 711                     | 22.51                     | 223     | 21.18 | 0.366 |

Table 4: Multivariate logistic regression analysis for factors associated with axillary lymph node metastasis

| Variables                       | Coefficient | SE     | OR     | 95% CI Lower | 95% CI Upper | P value |
|--------------------------------|-------------|--------|--------|--------------|--------------|---------|
| Age at diagnosis (years)       | -0.014      | 0.005  | 0.986  | 0.976        | 0.995        | 0.004   |
| Clinical tumor size            |             |        |        |              |              |         |
| T2 versus T1                   | 0.204       | 0.112  | 1.226  | 0.985        | 1.528        | 0.069   |
| T3 versus T1                   | 0.663       | 0.210  | 1.940  | 1.286        | 2.927        | 0.002   |
| Local invasion                 |             |        |        |              |              |         |
| UIQ versus Central             | -0.944      | 0.252  | 0.389  | 0.237        | 0.638        | 0.000   |
| UOQ versus Central             | -0.529      | 0.230  | 0.589  | 0.375        | 0.926        | 0.022   |
| LIQ versus Central             | -1.444      | 0.321  | 0.236  | 0.126        | 0.443        | 0.000   |
| LOQ versus Central             | -0.237      | 0.267  | 0.789  | 0.467        | 1.332        | 0.375   |
| Others versus Central          | -0.642      | 0.261  | 0.526  | 0.315        | 0.878        | 0.014   |
| Clinical lymph node status     |             |        |        |              |              |         |
| Histological type              | 1.235       | 0.109  | 3.440  | 2.777        | 4.261        | 0.000   |
| IDC versus DCIS-Mi             | 2.944       | 0.624  | 18.998 | 5.595        | 64.509       | 0.000   |
| ILC versus DCIS-Mi             | 2.884       | 0.674  | 17.887 | 4.778        | 66.964       | 0.000   |
| Others versus DCIS-Mi          | 2.111       | 0.658  | 8.254  | 2.273        | 29.972       | 0.001   |
| Molecular subtype              |             |        |        |              |              |         |
| LM versus TN                   | 0.322       | 0.135  | 1.380  | 1.059        | 1.799        | 0.017   |
| HER-2+ versus TN               | 0.141       | 0.210  | 1.152  | 0.764        | 1.737        | 0.500   |

Abbreviations: UIQ, upper inner quadrant; UOQ, upper outer quadrant; LIQ, lower inner quadrant; LOQ, lower outer quadrant; DCIS-Mi, ductal carcinoma in situ with micro-invasion; IDC, invasive ductal carcinoma; ILC, invasive lobular carcinoma; LM, luminal-like; TN, triple-negative; HER-2, human epidermal growth factor receptor-2; OR, odds ratio; CI, confidence interval; SE, standard error
Table 5: Accuracy of the developed model in low-risk predictive patients in the validation cohort

| Predicted Risk (%) | No. of patients* (%) | Number of patients with ALN metastasis | Sensitivity (%) | Specificity (%) | Accuracy (%) | FNR (%) |
|--------------------|----------------------|--------------------------------------|----------------|----------------|--------------|---------|
| <10.00             | 14 (2.18)            | 0                                    | 100            | 0              | 100          | 0       |
| <14.03             | 21 (3.17)            | 0                                    | 99.56          | 4.86           | 93.1         | 6.9     |
| <20.00             | 29 (4.52)            | 1                                    | 99.56          | 4.96           | 91.2         | 8.8     |
| <22.01             | 49 (7.63)            | 2                                    | 98.45          | 8.06           | 89.8         | 10.2    |
| <25.02             | 78 (12.15)           | 12                                   | 96.12          | 14.36          | 84.62        | 15.38   |
| <31.27             | 104 (16.2)           | 19                                   | 92.12          | 25.00          | 81.37        | 18.27   |
| <35.00             | 159 (24.77)          | 37                                   | 87.35          | 34.30          | 76.73        | 23.27   |
| <40.00             | 244 (38.01)          | 68                                   | 76.58          | 51.55          | 72.13        | 27.87   |
| <41.00             | 261 (40.65)          | 75                                   | 74.36          | 54.96          | 71.26        | 28.74   |
| <41.50             | 265 (41.28)          | 77                                   | 73.03          | 56.51          | 70.94        | 29.06   |

A total of 642 patients had complete data in the validation group: 319 patients had actual positive axillary lymph node (49.69%) and 323 patients had actual negative axillary lymph node (50.31%)

*patients: the patients whose predicted risk is lower than the cutoff value
Abbreviations: FNR: false negative rate; ALN: axillary lymph nodes

Figure 3: Nomogram for predicting the probability of axillary lymph node metastasis. Age(years)- age at diagnosis in years; Size- clinical tumor size; Location- the location of tumor; Invasion- invasion of skin or chest wall; Lymph node- clinical lymph node status; Pathology- Pathological type; Subtype: molecular subtype There are a total of 11 rows in the nomogram. The behavioral variables are presented in rows 2 to 8, and points for each variable are correspond the scale in row 1. The points of the seven variables are added to the total points presented on the scale in row 9, which corresponds to the linear predictor and risk predictor of axillary lymph node metastasis in rows 10 and 11, respectively.
Figure 4: ROC curve of the predictive model for the training cohort (n = 1869) (ROC curve with an AUC value of 0.7157). ROC, receiver-operating characteristic ROC; AUC, area under the ROC curve.

Figure 5: ROC curve of the predictive model for the validation cohort (n = 642) (ROC curve with an AUC value of 0.7007). ROC, receiver-operating characteristic ROC; AUC, area under the ROC curve.

Figure 6: Calibration plot for the predictive model: The actual probability versus the predicted probability. The reference line represents perfect equality of the predicted probability and the actual incidence of ALN metastasis.
proportion of all patients in China. Thus, the development and application of this model in China were limited. In our current model, the seven breast cancer treatment centers included appropriately reflect the incidence, diagnosis, and treatment of breast cancer in women in China and represent the entire population of China [20].

In many risk predictions of cancer, the nomogram is considered an effective tool for quantitative assessment of risk factors to maximize the accuracy of prediction. It can reflect the contribution of predictive variables to the outcome visually and directly [33, 34]. In this study, we successfully established a nomogram model for prediction of breast cancer ALN metastasis that is suitable for Chinese people. Our results showed that the histopathological type played a crucial role in ALN metastasis, followed by tumor location, clinical lymph node status, age at diagnosis, invasion of the chest wall and skin, tumor size, and molecular subtype. Applying this model for patients in the training cohort and the validation cohort in this study, the performance of the nomogram in these two groups was similar (AUC = 0.7157 versus 0.7007), and the nomogram showed good predictive value in both. These results confirmed that our nomogram was useful in different populations.

Several studies demonstrated that age of patients with breast cancer at diagnosis, BMI, tumor size, primary tumor quadrant, presence of multiple tumors, clinical lymph node status, local invasion status, pathological type, ER/PR, HER2 status, molecular subtypes, and other factors were related to ALN metastasis status [35-37]. Our results showed that the significance of these variables was similar to that of the results previously reported in the literature, including age at diagnosis, tumor size, clinical lymph node status, local invasion status, and pathological type.

In our study, tumor in the central region of the breast was more prone to ALN metastasis than that in other quadrants, which is consistent with the findings of other studies, showing abundant lymphatic drainage in the central region of the breast [38]. Some studies reported that the tumor in the UIQ of the breast was the most difficult tumor location of axillary metastasis [38-40]. However, we found that compared to the other locations, the risk of ALN metastasis in the LIQ was the lowest. We speculate that this difference between our study and previous studies is related to the differences in tumor heterogeneity, ethnic differences, lifestyle factors, and so on. The exact reason of this phenomenon remains unclear. Further, some studies showed that molecular subtypes had no predictive value for ALN status [41], while others showed that the triple-negative breast cancer patients had the lowest incidence of ALN metastasis and the HER-2 subtype had the highest incidence of ALN metastasis [42, 43]. Contradictory to these findings, we observed that luminal-like breast cancer patients were associated with higher probability of ALN metastasis. One possible explanation for this finding could be that Luminal-like tumors had more lymphatic metastasis than triple-negative phenotype [32, 38, 44]. Nevertheless, these inconsistencies between our study and previous studies need to be investigated further.

To further assess the clinical application of the prediction model of ALN metastasis, we selected certain cutoff values for predicted risk for use in patients in the validation cohort. For patients with a metastasis risk below the cutoff value, we believed there was a low risk of ALN metastasis. Therefore, as per our model, they could be considered to be free from SLNB and ALND. According to the report of American Society of Clinical Oncology (ASCO), a false-negative rate of 0-29% was reported for SLNB, and the average false-negative rate was 8.4% [45]. In our model, the false-negative rate was only 6.9% when the cutoff value of 20%, which is less than the average false-negative rate of SLNB. Therefore, our nomogram should be acceptable in medical practice. SLNB may not be necessary when the predicted risk is less than 20%, especially for senile patients with other internal diseases and a lower surgical tolerance, who would have a low probability of ALN metastasis but might be more likely to suffer from postoperative complications [46, 47]. Thus, application of this model can reduce the surgical risk and postoperative complications in breast cancer patients with a low risk of ALN metastasis.

This study has several strengths that have been highlighted below: (1) To the best of our knowledge, this is the first nomogram prediction model of breast cancer ALN status that considered multi-center data and represents the entire Chinese population with breast cancer. (2) Our prediction model included data on seven variables, which can be obtained by conventional preoperative examination. This information will greatly improve the clinical application of the prediction model without additional examinations and costs. It has important implications for patients in developing countries and economically less-developed regions. (3) The AUC value obtained from the prospective data is 0.7007, suggesting a good predictive ability. Therefore, this model can help clinicians weigh the risks and benefits of SLNB before surgery in order to avoid unnecessary SLNB and ALND for patients.

Despite our important findings and strengths, our study had a few limitations that need to be addressed. First, the established model was based on clinical data of patients with breast cancer in multi-centers over a duration of 10 years. Owing to a long duration of data collection and differences in regional culture and educational and medical levels, there may be unavoidable biases introduced. For example, some data were missing in the study. Although 4211 patients were included in the analysis, only 2511 patients with complete data for variables were entered in the final model (1869 patients in the training cohort and 642 patients in the validation cohort). Second, in general, the predictive value of the model is considered good when the AUC value is 0.7-0.8.
and very good when the AUC value is 0.8-0.9. When we prospectively used the model for patients in the validation cohort \((n = 655)\), the obtained AUC value was 0.7007. The AUC value of our study is not perfect due to the large amount of data from multiple centers, it still needs more clinical central and a larger sample size to further evaluate and improve the predictive ability of model. In the future, we will validate the predictive ability of the model in larger clinical studies.

In conclusion, age of patients, tumor size, primary tumor quadrant, clinical nodal status, local invasion status, pathological type, and molecular subtypes were independent predictors of ALN metastasis. The nomogram model established in this study could provide an accurate and objective tool to predict the risk of breast cancer ALN metastasis by quantitative indicators. The developed model is easy to use and has a good predictive ability in the Chinese population. For low-risk patients with ALN metastasis, it can avoid the trauma and postoperative complications associated with axillary surgery, thereby improving the quality of life in patients.

**PATIENTS AND METHODS**

**Study design**

Data were obtained from the Nationwide Multicenter 10-year (1999-2008) Retrospective Clinical Epidemiological Study of Breast Cancer in China, led by Cancer Hospital/Institute, Chinese Academy of Medical Sciences (CICAMS) and jointly included seven Grade Three A hospitals nationwide.

**Selection of regions and hospitals**

To ensure that the samples were representative of the total population with breast cancer in China, we selected seven geographic regions across China, including North, North-East, Central, South, East, North-West, and South-West regions (Figure 1). These regions encompassed most of the country and represented different breast cancer burdens. A representative Grade Three A hospital was selected from each region based on the criteria used in our previous study [20]. Briefly, the criteria are listed below: (1) the city where the hospital is located must be an important city in the region; (2) participant hospitals must be leading public cancer hospitals and regional referral centers providing pathology diagnosis, surgery, radiotherapy, medical oncology, and routine follow-up care for patients with breast cancer; and (3) the source of the patient must be able to cover the corresponding research area in order to represent the region.

**Data collection and quality control**

Employees who uniformly received professional and systemic training in Beijing were responsible for recording patient’s information. The patient information was collected by standard case report forms (CRF) designed by the CICAMS and included data on general information, risk factors, diagnostic imaging tests, therapy models, and pathologic characteristics. The reliability and validity of the CRF were assessed by a preceding pilot study. The data were transmitted to Cancer Hospital/Institute, Chinese Academy of Medical Sciences and verified by EpiData (http://www.epidata.dk/). Specific details of this process are described in our previous studies [20].

In 1999-2008, each hospital randomly selected a month every year, and data of at least 50 female breast cancer patients were collected in this month (January and February were excluded from the random selection to eliminate any confounding effects of China’s largest annual holiday). If the number of patients included was less than 50 in the selected month, the patients from the immediately preceding month and the immediately following month were included until the total number of patients in that year reached 50. If, in the selected month, the number of patients exceeded 50, they were all included in the study. As such, a total of 4211 patients with breast cancer were included in the study.

**Patients and variables**

The 4211 patients included in this study were randomly categorized into a training cohort or validation cohort in a 3:1 ratio. Relevant clinical and pathological features were grouped according to international practice. Age at diagnosis and body mass index (BMI; weight [kg]/square of height [m²]) were considered continuous variables. The following features were considered categorical variables: clinical tumor size assessment by ultrasound (categorized as T1 ≤ 2 cm, 2 cm < T2 ≤ 5 cm, T3 > 5 cm); primary tumor quadrant (categorized into upper inner quadrant [UIQ], upper outer quadrant [UOQ], lower inner quadrant [LIQ], lower outer quadrant [LOQ], central, and others [occult breast cancer or tumor cannot be touched in the breast]); preoperative ALN assessment by palpation or imaging (categorized into non-clinical metastasis [N0] and clinical metastasis [N1-N3]; the patients whose regional lymph nodes could not be assessed [Nx] were excluded); invasion of the chest wall and skin (categorized as invasion and non-invasion); multifocal tumors were categorized as multifocal and unifocal; pathological types were categorized as ductal carcinoma in situ with micro-invasion (DCIS-Mi), invasive ductal carcinoma (IDC), invasive lobular carcinoma (ILC), and other types of invasive carcinoma (tubular carcinoma, mucinous carcinoma, medullary carcinoma); expression
of estrogen receptor (ER), progesterone receptor (PR), and human epidermal growth factor receptor-2 (HER-2) (categorized as positive and negative). The outcome variable—postoperative ALN status—was categorized as positive (presence of one or more ALN metastasis) and negative (no metastasis). Molecular subtypes were divided into three categories: luminal-like subtype (ER+ and/ or PR+, any HER2 status), HER-2+ subtype (ER-, PR-, HER2+), and triple-negative (ER-, PR-, HER2-) [21, 22]. All patients included in the study were female patients with breast cancer who were diagnosed by histopathology and underwent successful SLNB and ALND.

Pathologic processing

All nodes were examined postoperatively with serial section H&E staining. IHC staining was performed to determine whether micrometastasis (0.2-2 mm cancer foci) existed or not when no cancer cells were identified on H&E staining. ER, PR were considered positive if immunostaining was positive in more than 1% of tumour cells. HER-2 positivity was defined as a score of 3+ on IHC or amplification on FISH [23-25]. The histological subtype categorization was based on the 1981 and 2003 histological classification criteria of the World Health Organization [26]. Specific details are described in our previous studies [20, 27].

Statistical analysis

The mean, SD, median were calculated to describe continuous variables, and a constituent ratio was used to describe categorical variables. T-test was used for the comparison of continuous variables, and Chi-square test or Fisher’s exact test was used for comparison of categorical variables. Using the clinical and pathological data of the training cohort, univariate logistic regression analysis was performed to explore ALN metastasis-related variables. Subsequently, multivariate logistic regression analysis was used to determine the variables that were independent influence factors of ALN metastasis and establish the nomogram of the prediction model for breast cancer ALN metastasis. Receive-operating characteristic (ROC) curves, areas under the ROC curves (AUC), sensitivity, specificity were used to evaluate the predictive ability of model. The ROC curve was prepared by retrospectively using the data of the training cohort and calculating the AUC value. This prediction model was prospectively used for patients in the validation cohort by depicting the ROC curve and re-evaluating the accuracy of the prediction through the AUC value. To test the accuracy and stability of the model, the decile of predicted values of metastasis risk was segmented based on the data of the validation cohort, and the average metastasis risk was calculated in each segment. The predictive value was taken as the abscissa, and the average actual metastasis risk was taken as the ordinate to draw the calibration curve. To further evaluate the clinical value of the model, we considered certain cutoff values for prediction risk in patients in the validation cohort and calculated the corresponding accuracy and false-negative rate of the cutoff values in order to assess the screening indicators of low-risk patients with ALN metastasis.

All statistical analyses were performed using SPSS (version 19.0, Chicago, IL, USA), Stata (version 11.0, College Station, TX), and R software (version 3.1.0, Institute for Statistics and Mathematics, Vienna, Austria). A two-tailed p value < 0.05 was considered statistically significant. In this study, all data are reported in aggregates.

Ethics statement

The research was approved by Institutional Review Board of the Cancer Foundation of China. Because of the retrospective nature of the study, we were unable to contact all patients or their families. In addition, considering that it will not pose a risk to patients included in the study, informed consent was not obtained. All patient identifiers were removed, as per the approved procedures. De-identified data were maintained in a secure database, to which only research team members had access.

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CONFLICTS OF INTEREST

The authors declare that they have no conflict of interest.

REFERENCES

1. Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J and Jemal A. Global cancer statistics, 2012. CA: a cancer journal for clinicians. 2015; 65:87-108.
2. Siegel RL, Miller KD and Jemal A. Cancer statistics, 2016. CA: a cancer journal for clinicians. 2016; 66:7-30.
3. Carter CL, Allen C and Henson DE. Relation of tumor size, lymph node status, and survival in 24,740 breast cancer cases. Cancer. 1989; 63:181-187.
4. Rosen PP, Groshen S, Kinne DW and Hellman S. Pathological prognostic factors in stage I (T1N0M0) and stage II (T1N1M0) breast carcinoma: a study of 644 patients with median follow-up of 18 years. Journal of Clinical Oncology. 1989; 7:1239-1251.

5. Quiet CA, Ferguson DJ, Weichselbaum RR and Hellman S. Natural history of node-negative breast cancer: a study of 826 patients with long-term follow-up. Journal of clinical oncology. 1995; 13:1144-1151.

6. Krag DN, Anderson SJ, Julian TB, Brown AM, Harlow SP, Ashikaga T, Weaver DL, Miller BJ, Jalovec LM and Frazier TG. Technical outcomes of sentinel-lymph-node resection and conventional axillary-lymph-node dissection in patients with clinically node-negative breast cancer: results from the NSABP B-32 randomised phase III trial. The lancet oncology. 2007; 8:881-888.

7. Mansel RE, Fallowfield L, Kissin M, Goyal A, Newcombe RG, Dixon JM, Yangou C, Horgan K, Bundred N and Monypenny I. Randomized multicenter trial of sentinel node biopsy versus standard axillary treatment in operable breast cancer: the ALMANAC Trial. Journal of the National Cancer Institute. 2006; 98:599-609.

8. Purushotham AD, Upponi S, Klevessah MB, Bobrow L, Millar K, Myles JP and Duffy SW. Morbidity after sentinel lymph node biopsy in primary breast cancer: results from a randomized controlled trial. Journal of Clinical Oncology. 2005; 23:4312-4321.

9. Fleissig A, Fallowfield LJ, Langridge CI, Johnson L, Newcombe RG, Dixon JM, Kissin M and Mansel RE. Post-operative arm morbidity and quality of life. Results of the ALMANAC randomised trial comparing sentinel node biopsy with standard axillary treatment in the management of patients with early breast cancer. Breast cancer research and treatment. 2006; 95:279-293.

10. Del Bianco P, Zavagno G, Barelli P, Scalco G, Barutta L, Carraro P, Pietrarota P, Meneghini G, Morbin T and Tacchetti G. Morbidity comparison of sentinel lymph node biopsy versus conventional axillary lymph node dissection for breast cancer patients: results of the sentinella-GIVOM Italian randomised clinical trial. European Journal of Surgical Oncology (EJSO). 2008; 34:508-513.

11. Langer I, Guller U, Berclaz G, Koechli OR, Schaer G, Fehr MK, Hess T, Oertli D, Bronz L and Schnarwyler B. Morbidity of sentinel lymph node biopsy (SLN) alone versus SLN and completion axillary lymph node dissection after breast cancer surgery: a prospective Swiss multicenter study on 659 patients. Annals of surgery. 2007; 245:452-461.

12. Kootstra JJ, Hoekstra-Weebers JE, Rietman JS, de Vries J, Baas PC, Geertzen JH and Hoekstra HJ. A longitudinal comparison of arm morbidity in stage I-II breast cancer patients treated with sentinel lymph node biopsy, sentinel lymph node biopsy followed by completion lymph node dissection, or axillary lymph node dissection. Annals of surgical oncology. 2010; 17:2384-2394.

13. Nori J, Vanzi E, Bazzocchi M, Bufalini FN, Distante V, Branconi F and Susini T. Role of axillary ultrasound examination in the selection of breast cancer patients for sentinel node biopsy. The American journal of surgery. 2007; 193:16-20.

14. Lambert LA, Ayers GD, Hwang RF, Hunt KK, Ross MI, Kuerer HM, Singletary SE, Babiera GV, Ames FC and Feig B. Validation of a breast cancer nomogram for predicting nonsentinel lymph node metastases after a positive sentinel node biopsy. Annals of surgical oncology. 2006; 13:310-320.

15. Airan S, De Rycke Y, Fourchotte V, Charitansky H, Laki F, Falcou MC, Benamor M, Frenaux P, Salmon RJ and Sigal-Zafrani B. Validation and Limitations of Use of a Breast Cancer Nomogram Predicting the Likelihood of Non-Sentinel Node Involvement After Positive Sentinel Node Biopsy. Annals of Surgical Oncology. 2007; 14:2195-2201.

16. Smidt ML, Kuster DM, van der Wilt GJ, Thunnissen FB, Van Zee KJ and Strobbe LJ. Can the Memorial Sloan-Kettering Cancer Center nomogram predict the likelihood of nonsentinel lymph node metastases in breast cancer patients in the Netherlands? Annals of surgical oncology. 2005; 12:1066-1072.

17. Ponzone R, Maggiorotto F, Mariani L, Jacomuzzi ME, Magistris A, Mininni P, Biglia N and Sismondi P. Comparison of two models for the prediction of nonsentinel node metastases in breast cancer. The American journal of surgery. 2007; 193:686-692.

18. Zgajnar J, Perhavec A, Hocevar M, Podkrajsek M, Hertl K, Fkrovic-Grazio S, Pohar M and Besic N. Low performance of the MSKCC nomogram in preoperatively ultrasonically negative axillary lymph node in breast cancer patients. Journal of surgical oncology. 2007; 96:547-553.

19. Pal A, Provenzano E, Duffy S, Pinder S and Purushotham A. A model for predicting non-sentinel lymph node metastatic disease when the sentinel lymph node is positive. British Journal of Surgery. 2008; 95:302-309.

20. Li J, Zhang BN, Fan JH, Pang Y, Zhang P, Wang SL, Zheng S, Zhang B, Yang HJ and Xie XM. A nation-wide multicenter 10-year (1999-2008) retrospective clinical epidemiological study of female breast cancer in China. BMC cancer. 2011; 11:1.

21. Perou CM, Sorlie T, Eisen MB, van de Rijn M, Jeffrey SS, Rees CA, Pollack JR, Ross DT, Johnsen H and Akslen LA. Molecular portraits of human breast tumours. Nature. 2000; 406:747-752.

22. Sorlie T, Perou CM, Tibshirani R, Aas T, Geisler S, Johnsen H, Hastie T, Eisen MB, van de Rijn M and Jeffrey SS. Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. Proceedings of the National Academy of Sciences. 2001; 98:10869-10874.

23. Wolff AC, Hammond MEH, Schwartz JN, Hagerty KL, Allred DC, Cote RJ, Dowsett M, Fitzgibbons PL, Hanna WM and Langer A. American Society of Clinical Oncology/College of American Pathologists guideline
recommendations for human epidermal growth factor receptor 2 testing in breast cancer. Archives of pathology & laboratory medicine. 2007; 131:18-43.

24. Hammond MEH, Hayes DF, Dowsett M, Allred DC, Hagerty KL, Badve S, Fitzgibbons PL, Francis G, Goldstein NS and Hayes M. American Society of Clinical Oncology/College of American Pathologists guideline recommendations for immunohistochemical testing of estrogen and progesterone receptors in breast cancer (unabridged version). Archives of pathology & laboratory medicine. 2010; 134:e48-e72.

25. Perez EA, Dueck AC, McCullough AE, Reinholz MM, Tenner KS, Davidson NE, Gralow J, Harris LN, Kutteh LA and Hillman DW. Predictability of adjuvant trastuzumab benefit in N9831 patients using the ASCO/CAP HER2-postitivity criteria. Journal of the National Cancer Institute. 2012;104:159-62. doi: 10.1093/jnci/djr490.

26. Hartmann W, Ozzello L, Sobin L and Stalsberg H. Histological typing of breast tumors. World Health Organization, Geneva. 1981:15-25.

27. Zheng S, Bai JQ, Li J, Fan JH, Pang Y, Song QK, Huang R, Yang HJ, Xu F and Lu N. The pathologic characteristics of breast cancer in China and its shift during 1999&dash;2008: A national-wide multicenter cross-sectional image over 10 years. International Journal of Cancer. 2012; 131:2622-2631.

28. Fein DA, Fowlke BL, Hanlon AL, Hooks MA, Hoffman JP, Sigurdson ER, Jardines LA and Eisenberg BL. Identification of women with T1-T2 breast cancer at low risk of positive axillary nodes. Journal of surgical oncology. 1997; 65:34-39.

29. Hwang SO, Lee SW, Kim HJ, Kim WW, Park HY and Jung JH. The comparative study of ultrasonography, contrast-enhanced MRI, and 18F-FDG PET/CT for detecting axillary lymph node metastasis in T1 breast cancer. Journal of breast cancer. 2013; 16:315-321.

30. Valente SA, Levine GM, Silverstein MJ, Rayhanabad JA, Weng-Grumley JG, Ji L, Holmes DR, Spoto R and Sener SF. Accuracy of predicting axillary lymph node positivity by physical examination, mammography, ultrasonography, and magnetic resonance imaging. Annals of surgical oncology. 2012; 19:1825-1830.

31. Turnbull L, Brown S, Harvey I, Olivier C, Drew P, Napp V, Hanby A and Brown J. Comparative effectiveness of MRI in breast cancer (COMICE) trial: a randomised controlled trial. The Lancet. 2010; 375:563-571.

32. Qiu SQ, Zeng HC, Zhang F, Chen C, Huang WH, Pleijhuis RG, Wu JD, van Dam GM and Zhang GJ. A nomogram to predict the probability of axillary lymph node metastasis in early breast cancer patients with positive axillary ultrasound. Scientific reports. 2016; 6.

33. Karakiewicz PI, Briganti A, Chun FKH, Trinh QD, Perrotte P, Ficarra V, Cindolo L, De La Taille A, Tostain J and Mulders PF. Multi-institutional validation of a new renal cancer-specific survival nomogram. Journal of clinical oncology. 2007; 25:1316-1322.

34. Wierda WG, O’Brien S, Wang X, Faderl S, Ferrajoli A, Do KA, Cortes J, Thomas D, Garcia-Manero G and Koller C. Prognostic nomogram and index for overall survival in previously untreated patients with chronic lymphocytic leukemia. Blood. 2007; 109:4679-4685.

35. Xie F, Yang H, Wang S, Zhou B, Tong F, Yang D and Zhang J. A logistic regression model for predicting axillary lymph node metastases in early breast carcinoma patients. Sensors. 2012; 12:9936-9950.

36. Gann PH, Colilla SA, Gapstur SM, Winchester DJ and Winchester DP. Factors associated with axillary lymph node metastasis from breast carcinoma. Cancer. 1999; 86:1511-1519.

37. Viale G, Zurrida S, Maieronan E, Mazzarol G, Pruneri G, Paganelli G, Maisonneuve P and Veronesi U. Predicting the status of axillary sentinel lymph nodes in 4351 patients with invasive breast cancer treated in a single institution. Cancer. 2005; 103:492-500.

38. Bevilacqua JLB, Kattan MW, Fey JV, Cody HS, Borgen PI and Van Zee KJ. Doctor, what are my chances of having a positive sentinel node? A validated nomogram for risk estimation. Journal of Clinical Oncology. 2007; 25:3670-3679.

39. Bevilacqua J, Gucciardo G, Cody H, MacDonald K, Sacchini V, Borgen P and Van Zee K. A selection algorithm for internal mammary sentinel lymph node biopsy in breast cancer. European Journal of Surgical Oncology (EJSO). 2002; 28:603-614.

40. Estourgie SH, Tanis PJ, Nieweg OE, Olmos RAV, Emiel JT and Kroon BB. Should the hunt for internal mammary chain sentinel nodes begin? An evaluation of 150 breast cancer patients. Annals of surgical oncology. 2003; 10:935-941.

41. Jones T, Neboori H, Wu H, Yang Q, Haffty BG, Evans S, Higgins S and Moran MS. Are breast cancer subtypes prognostic for nodal involvement and associated with clinicopathologic features at presentation in early-stage breast cancer? Annals of surgical oncology. 2013; 20:2866-2872.

42. Crabb SJ, Cheang MC, Leung S, Immonen T, Nielsen TO, Huntsman DD, Bajdik CD and Chia SK. Basal breast cancer molecular subtype predicts for lower incidence of axillary lymph node metastases in primary breast cancer. Clinical breast cancer. 2008; 8:249-256.

43. Zhou W, He Z, Xue J, Wang M, Zha X, Ling L, Chen L, Wang S and Liu X. Molecular subtype classification is a determinant of non-sentinel lymph node metastasis in breast cancer patients with positive sentinel lymph nodes. PLoS One. 2012; 7:e35881.

44. Mattes MD, Bhatia JK, Metzger D, Ashamalla H and Katsoulakis E. Breast cancer subtype as a predictor of lymph node metastasis according to the SEER registry. Journal of breast cancer. 2015; 18:143-148.

45. Lyman GH, Giuliano AE, Somerfield MR, Benson AB,
Bodurka DC, Burstein HJ, Cochran AJ, Cody HS, Edge SB and Galper S. American Society of Clinical Oncology guideline recommendations for sentinel lymph node biopsy in early-stage breast cancer. Journal of Clinical Oncology. 2005; 23:7703-7720.

46. Sakorafas GH, Peros G, Cataliotti L and Vlastos G. Lymphedema following axillary lymph node dissection for breast cancer. Surgical oncology. 2006; 15:153-165.

47. della Rovere GQ, Bonomi R, Ashley S and Benson J. Axillary staging in women with small invasive breast tumours. European Journal of Surgical Oncology (EJSO). 2006; 32:733-737.