Clinicians’ response to hyperoxia in ventilated patients in a Dutch ICU depends on the level of FiO₂

Abstract  

Purpose: Hyperoxia may induce pulmonary injury and may increase oxidative stress. In this retrospective database study we aimed to evaluate the response to hyperoxia by intensivists in a Dutch academic intensive care unit. 

Methods: All arterial blood gas (ABG) data from mechanically ventilated patients from 2005 until 2009 were extracted from an electronic storage database of a mixed 32-bed intensive care unit in a university hospital in Amsterdam. Mechanical ventilation settings at the time of the ABG tests were retrieved. 

Results: The results of 126,778 ABG tests from 5,498 mechanically ventilated patients were retrieved including corresponding ventilator settings. In 28,222 (22%) of the ABG tests the arterial oxygen tension (PaO₂) was >16 kPa (120 mmHg). In only 25% of the tests with PaO₂ >16 kPa (120 mmHg) was the fraction of inspired oxygen (FiO₂) decreased. Hyperoxia was accepted without adjustment in ventilator settings if FiO₂ was 0.4 or lower. 

Conclusion: Hyperoxia is frequently seen but in most cases does not lead to adjustment of ventilator settings if FiO₂ <0.4. Implementation of guidelines concerning oxygen therapy should be improved and further research is needed concerning the effects of frequently encountered hyperoxia. 

Keywords Mechanical ventilation · Hyperoxia · Guidelines · Lung injury · Oxygen

Introduction

Mechanical ventilation (MV) provides essential life support for patients with respiratory failure. Unfortunately, this life support may cause or aggravate lung injury in acute respiratory distress syndrome (ARDS). Ventilator induced lung injury caused by repeated shear stress and barotrauma can be reduced using protective lung ventilation with lower tidal volumes [1]. The ARDS network study demonstrated a compelling survival advantage when using low tidal volumes rather than conventional MV in patients with acute lung injury (ALI) or ARDS [2]. This led to the reappraisal of the goals of ventilatory support, allowing some hypercapnia, in order to limit inspiratory pressures and to lower tidal volumes. Not only barotrauma due to MV, but also hyperoxia can induce lung injury. Many experimental studies have demonstrated increased levels of reactive oxygen-derived free radicals, with an influx of inflammatory cells, increased pulmonary permeability and endothelial cell injury with the use of a high fraction of inspired oxygen (FiO₂) [3]. High inspiratory oxygen concentrations induce tissue hyperoxia and may mask pulmonary injury. Another detrimental effect of hyperoxia is the development of resorption atelectasis with increased shunting [4]. Accordingly, together with permissive hypercapnia, a certain extent of hypoxaemia is accepted. Generally a target arterial oxygen tension (PaO₂) of >8 kPa (60 mmHg) is suggested [5]. A Canadian questionnaire study, however, found considerable variation in the
attitudes, beliefs and practices of intensivists in the management of oxygen therapy. The responses suggested that most respondents believed that levels of FiO2 up to 0.4 are not harmful and that this is the ideal range of FiO2 when the PaO2 permits [6]. The actual response of intensivists to hyperoxia has never been studied. In this retrospective cohort study we aimed to study whether high PaO2 levels lead to adjustments in ventilator settings by intensivists in a Dutch academic intensive care unit (ICU).

Methods

Study centre

The Academic Medical Center (AMC) in Amsterdam is a teaching hospital where the ICU is a 32-bed “closed format” department in which medical and surgical patients (including cardiothoracic and neurosurgical patients) are under the direct care of the ICU team.

Patients and data collection

The study included all arterial blood gas (ABG) data from all mechanically ventilated patients from 2005 until 2009. Data were extracted automatically from the patient data management system (PDMS) storage database (Metavision, iMDsoft, Leiden, The Netherlands). Patients on noninvasive MV were excluded from the study. All patients received MV with an Evita4 ventilator (Dräger-Medical, Best, The Netherlands) or a Galileo ventilator (Hamilton Medical, Rhaïzüns, Switzerland). MV settings at the time of the ABG tests were retrieved. The following MV data were recorded: FiO2, measured positive end-expiratory pressure (PEEP) level, applied tidal volume (TV), inspiratory:expiratory (I:E) ratio and MV mode. Prone positioning, recruitment manoeuvres and other complementary measures that may improve oxygenation could not be explored in this database study. Every set of MV and ABG data was compared with the following set. The second set was then compared with the third, and so on. For every patient the last ABG test was excluded because no comparison was possible with a subsequent determination.

Demographic data were collected from the Dutch National Intensive Care Evaluation registry and included age, height, weight and acute physiological and chronic health evaluation (APACHE) II score. ABGs were determined routinely half an hour after every MV adjustment and also when blood glucose levels were measured. They were determined more frequently in patients without a good pulse oximetric saturation signal or on high pressure MV. All ABG test results were made available on the PDMS and were communicated to the clinician.

The predicted body weight (PBW) was calculated from the following formulas: in men, PBW (kg) = 50 + 0.91(height in centimetres – 152.4); in women, PBW (kg) = 45.5 + 0.91(height in centimetres – 152.4) [2]. In order to monitor the response to a certain extent of oxygenation we categorized the ABG test results into five PaO2 groups: <8 kPa (60 mmHg), 8.01–10 kPa (60–75 mmHg), 10.01–13 kPa (75–98 mmHg), 13.01–16 kPa (98–120 mmHg), and >16 kPa (120 mmHg). For the purpose of this study the last group with a PaO2 >16 kPa (120 mmHg) was regarded as hyperoxic.

MV protocol

In the AMC a written MV protocol is available for all ICU physicians and nurses. Briefly, this protocol advises pressure-controlled MV or pressure-support MV. In addition, adaptive support ventilation is used on the Hamilton Galileo ventilator in patients with an uncomplicated postoperative course in whom extubation is expected within 24 h. The MV protocol is based on an open lung concept [7]. MV is typically initiated with a FiO2 0.4–0.6, PEEP ≥5 cmH2O and I:E ratio 1:2, with a target PaO2 higher than 10 kPa (75 mmHg). PEEP and I:E ratio may be adjusted for lung recruitment if necessary. The FiO2 is targeted below 0.6. During procedures a higher FiO2 is accepted. Prone positioning is considered if the PEEP is >10 cmH2O and FiO2 is >0.6. The protocol suggests lowering FiO2 below 0.4 with a target PaO2 higher than 10 kPa (75 mmHg). In the weaning phase, patients are candidates for discontinuation of MV if FiO2 is ≤0.4, PEEP is ≤5 cmH2O and no signs of respiratory distress are present. Manual hyperinflation is performed as part of standard airway care, after disconnection of the ventilator and after adjustment of MV settings. Recommended tidal volumes are between 6 and 8 ml/kg PBW. All blood gas results are shown to the physicians. As part of the team, nurses can decrease the FiO2 if the PaO2 is higher than 10 kPa (75 mmHg), but all changes in FiO2 have to be reported to the physician. Nurses can make ventilator therapy recommendations, but unit policy mandates that all MV setting adjustments other than lowering of FiO2 are ordered by physicians. During the study period no changes in the MV protocol were made.

Statistical analysis

The statistical analysis was carried out with SPSS 16.01. An independent samples t-test, one-way ANOVA and the Kruskal-Wallis test were used as appropriate. Differences with a p value <0.05 were considered significant.
Results

A total of 148,136 ABG test results were retrieved from the database, of which 126,778 were obtained during MV. These ABG tests related to 5,498 mechanically ventilated patients, of whom 4,880 were first admissions. The demographic data are shown in Table 1. In 44% of the patients admission was after planned surgery. Trauma patients represented 4% of the study group. In the neurosurgery group 7% were elective patients and the others were admitted after urgent surgery for bleeding or trauma. Of the surgical patients almost 40% were ventilated for recovery from anaesthesia.

The baseline ventilator settings and adjustments together with subsequent oxygenation data for all ABG tests are shown in Table 2. Overall, approximately 40% of the ABG tests were performed during pressure-controlled MV with a median I:E ratio of 1:2. The mean tidal volume was 7.5 ± 2.2 ml/kg PBW. In 57% of the ABG tests the PaO2 was between 8 and 13 kPa (60–98 mmHg), in more than 40% the PaO2 was >10 kPa (75 mmHg), and in 22% the PaO2 was >16 kPa (120 mmHg). The median interval between two successive ABG tests was 166 min (interquartile range 100–258 min). This decreased significantly with a lower PaO2 (P<0.001). The PEEP and FiO2 were not changed in most patients in any of the PaO2 groups.

To study the response to hyperoxia, we subcategorized the highest PaO2 group (>16 kPa, >120 mmHg) based on the FiO2 (Table 3). In 57% of these ABG tests FiO2 was exactly 0.4 and 8% had FiO2 <0.4. In only one-quarter (24.9%) of the ABG tests was the FiO2 decreased. Of the total group with a PaO2 >16 kPa (120 mmHg), at time of the successive ABG test 11% had FiO2 <0.4, with a mean successive PaO2 of 18 ± 5.9 kPa (135 ± 44 mmHg). At the time of the successive ABG test 64% had a FiO2 of exactly 0.4. In the subsequent ABG test 33% had PEEP above 5 cmH2O. In 78% of cases with PaO2 >16 kPa (120 mmHg) and FiO2 <0.41 neither FiO2 nor PEEP was adjusted (Table 4). In contrast, in most cases with PaO2 >16 kPa (120 mmHg) and FiO2 >0.4, ventilator settings were adjusted.

Discussion

In this retrospective database study, we found that hyperoxia (defined as PaO2 >16 kPa, 120 mmHg) was frequently present in patients but in most cases did not lead to adjustment of ventilator settings. If the FiO2 was 0.4 or lower the hyperoxia was accepted in the majority of cases.

Oxygen therapy has been used in the care of critically ill patients since the early years of the last century. However, from the beginning of the 1970s increasing understanding emerged that oxygen therapy can cause pulmonary toxicity [8]. Baboons were found to have progressive alveolar septal injury after exposure to 60% oxygen for 14 days and hyperoxia (FiO2 0.5) exacerbated lung injury in a large tidal volume model in rabbits [9, 10]. The acute lung injury caused by hyperoxia is characterized by an influx of inflammatory cells representing an alteration in barrier properties of the alveolar capillary membrane [3]. The systemic effects of hyperoxia are less clear [11]. Some work has suggested that there is an important interaction between FiO2 and pulmonary free radicals potentially leading to an adverse effect on outcome in ARDS [12]. More recently, Li et al. [13] found in mice that the combined detrimental effect of oxygen and repetitive cyclic stretch results in the activation of specific independent intracellular signalling pathways, showing that hyperoxia-induced lung injury is more than free radical-mediated injury alone. Despite extensive experimental evidence in animals supporting a direct injurious effect of high inspired oxygen concentrations, only few human data exist on this topic. In 1992 a small study in 12 healthy nonsmoking adults exposed to a fractional inspired oxygen concentration of 50% (mean exposure 44 h) showed enhanced oxidative stress and stimulation of alveolar macrophages in bronchoalveolar lavage fluid [14]. Little is known about the dose range of oxygen toxicity. A recent study in Wistar rats found a dose-dependent hyperoxic lung injury [15]. However, to our knowledge no studies about the safety of an inspired oxygen concentration below 40% have been performed. Moreover, until now no studies concerning oxygen toxicity have been conducted in critically ill patients [16].

The fact that neither physicians nor nurses adjusted ventilator settings in the event of hyperoxia would be understandable if other clinical signs indicated

Table 1 Patient characteristics of the study population

| Characteristic                        | Value                  |
|---------------------------------------|------------------------|
| Age (years, mean ± SD)                | 61 ± 16                |
| Gender M/F (%)                        | 64/36                  |
| Height (cm, mean ± SD)                | 174 ± 9                |
| Actual body weight (kg, mean ± SD)    | 78 ± 6                 |
| Predicted body weight (kg, mean ± SD) | 67 ± 10                |
| APACHE II score (mean ± SD)           | 19 ± 8                 |
| Referring specialty (%)                |                        |
| Cardiothoracic surgery                | 37                     |
| General surgery                       | 20                     |
| Internal medicine                     | 12                     |
| Neurosurgery                          | 11                     |
| Cardiology                            | 9                      |
| Other                                 | 11                     |
| Type of admission (%)                 |                        |
| Medical                               | 37                     |
| Unplanned surgery                     | 19                     |
| Planned surgery                       | 44                     |

*Men = 50 + 0.91 height in centimetres − 152.4; women = 45.5 + 0.91 height in centimetres − 152.4*
deterioration of pulmonary function. In our study, however, this is a very unlikely explanation given the fact that mean PaO₂ in the subsequent ABG test was still 18 kPa (135 mmHg). Another potential but unlikely explanation would be that physicians and ICU nurses prefer some level of hyperoxia because monitoring of arterial oxy- genation is performed relatively infrequently. We found a median duration between ABG tests of 166 min, which is equal to almost nine ABG analyses per patient-day.

Significantly more ABG samples were taken in the group with a PaO₂ below 8 kPa (60 mmHg). This is more frequent than described in the past. In 1997, a study found that after the implementation of a practice guideline for ABG measurement, the number of tests decreased from 4.9 to 2.4 ABG tests per patient-day [17]. In a Swiss study the frequency decreased after a guideline from 8.2 to 4.8 tests per patient-day [18]. The high number of ABG tests in this study may be explained by the fact that blood glucose was also measured in the blood gas analyser. Nevertheless, in these cases ABG data were also pre- sented to the clinician. In addition to frequent ABG analysis, oxygen saturation is also continuously moni- tored transcutaneously in all our patients. Rice et al. [19] demonstrated in a medical patient population that pulse oximetric saturation/FiO₂ correlates with PaO₂/FiO₂ ratios, and thus may serve as a surrogate for the follow-up of mechanically ventilated critically ill patients. Thus, unnoticed hypoxaemia is unlikely to occur and there seems to be no reason for preferring some level of hyperoxia.

Table 2  Oxygenation data, ventilator settings and adjustments for all ABG tests

| PaO₂ category (kPa) | n (%) | PaO₂ (kPa, mean ± SD) | FiO₂ | PEEP (cmH₂O) | Time until next ABG test (min) | Delta FiO₂ | Delta PEEP (cmH₂O) | Delta PaO₂ (kPa, mean ± SD) | No decrease in FiO₂ or PEEP, n (%) |
|--------------------|-------|-----------------------|------|-------------|-------------------------------|-----------|-------------------|-------------------------------|---------------------------------|
| <8                 | 4,544 (3.6) | 7.3 ± 0.7          | 0.5 (0.4–0.61) | 10 (7–12) | 95 (56–159)                   | 0 (0–0.1) | 0 (0–2)            | 2.8 ± 4.6                      | 3.749 (83)                     |
| 8.01–10            | 21,335 (16.8) | 9.2 ± 0.6          | 0.45 (0.4–0.51) | 9 (5–12) | 143 (87–228)                  | 0 (0–0)   | 0 (0–0)            | 1.5 ± 3.2                      | 17.071 (80)                    |
| 10.01–13           | 45,545 (35.9) | 11.5 ± 0.8        | 0.4 (0.4–0.5)  | 8 (5–10) | 174 (108–264)                 | 0 (0–0)   | 0 (0–0)            | 0.6 ± 3.1                      | 32.973 (72)                    |
| 13.01–16           | 27,132 (21.4) | 14.4 ± 0.9        | 0.4 (0.4–0.42) | 5 (5–10) | 181 (111–276)                 | 0 (0–0)   | 0 (0–0)            | -0.4 ± 3.4                     | 18.297 (67)                    |
| >16.01             | 28,222 (22.3) | 20.7 ± 6.0        | 0.4 (0.4–0.45) | 5 (5–10) | 166 (99–261)                  | 0 (0–0)   | 0 (0–0)            | -2.6 ± 6.4                     | 17.527 (62)                    |

Data are presented as medians (interquartile range) unless otherwise stated.

Table 3  Oxygenation data, ventilator settings and adjustments in subcategory with PaO₂ >16 kPa

| FiO₂ | n (%) | PaO₂ (kPa, mean ± SD) | PEEP (cmH₂O) | Delta FiO₂ | Delta PEEP (cmH₂O) | Delta PaO₂ (kPa, mean ± SD) |
|------|-------|-----------------------|-------------|------------|--------------------|----------------------------|
| 0.21–0.4a | 18,287 (65) | 19.8 ± 3.6 | 5 (5–6) | 0 (0–0) | 0 (0–0) | -1.5 ± 4.5 |
| 0.41–0.6 | 8,451 (30) | 21.1 ± 5.7 | 8 (5–10) | -0.05 (–0.1 to 0) | 0 (0–0) | -3.8 ± 6.4 |
| 0.61–0.8 | 866 (3) | 23.8 ± 8.2 | 12 (10–15) | -0.15 (–0.2 to 0.1) | 0 (0–0) | -7.3 ± 8.6 |
| 0.81–1.0 | 618 (2) | 38.9 ± 18.9 | 10 (5–15) | -0.3 (–0.5 to −0.15) | 0 (0–0) | -17.4 ± 18 |

Data are presented as medians (interquartile range) unless otherwise stated.

| FiO₂ | Decreased FiO₂, n (%) | Decreased PEEP, n (%) | No decrease in FiO₂ or PEEP, n (%) |
|------|-----------------------|-----------------------|-----------------------------------|
| 0.21–0.4a | 998 (6) | 3,090 (17) | 14,289 (78) |
| 0.41–0.6 | 4,810 (57) | 1,798 (21) | 2,994 (35) |
| 0.61–0.8 | 715 (83) | 174 (20) | 130 (15) |
| 0.81–1.0 | 496 (81) | 105 (17) | 114 (18) |

Data are presented as medians (interquartile range) unless otherwise stated.

Notes:

- a In 88% of this group FiO₂ = 0.4
- b Delta difference between successive ABG tests

a In 88% of this group FiO₂ = 0.4

b Delta difference between successive ABG tests

Table 4  Ventilator adjustments in subcategory with PaO₂ >16 kPa

| FiO₂ | Decreased FiO₂, n (%) | Decreased PEEP, n (%) | No decrease in FiO₂ or PEEP, n (%) |
|------|-----------------------|-----------------------|-----------------------------------|
| 0.21–0.4a | 998 (6) | 3,090 (17) | 14,289 (78) |
| 0.41–0.6 | 4,810 (57) | 1,798 (21) | 2,994 (35) |
| 0.61–0.8 | 715 (83) | 174 (20) | 130 (15) |
| 0.81–1.0 | 496 (81) | 105 (17) | 114 (18) |

Data are presented as medians (interquartile range) unless otherwise stated.

- a In 88% of this group FiO₂ = 0.4
In our study, many patients were admitted after elective surgery, recovering from anaesthesia. These patients may have been disconnected from the ventilator when PaO₂ was high and FiO₂ was 0.4. However, as only ABG tests were included when patients were on mechanical ventilation, this is no explanation for our findings. Furthermore, some patients had a very high FiO₂ (0.81–1.0) as well as a high mean PaO₂ of 39 kPa (Table 3). This group only represents 0.5% of the total group of ABG tests. Explanations for unusually high PaO₂ values may include dysfunctioning pulse oximetric saturation measurements leading to high FiO₂, treatment for CO intoxication or hyperoxygenation during or after tracheal suction.

High PaO₂ values only led to adjustment of inspired oxygen concentration if FiO₂ was higher than 0.4. Our protocol mandates a target PaO₂ above 10 kPa (75 mmHg) and suggests decreasing the FiO₂ to 0.21 if the PaO₂ is above 10 kPa (75 mmHg). Furthermore, 33% of the group with PaO₂ >16 kPa (120 mmHg) still had PEEP above 5 cmH₂O in the subsequent ABG test. Previous reports from our institution show that feedback and education improve physician compliance in the use of lung-protective MV [20]. It seems very likely that feedback, education and adjusted protocols concerning oxygenation will improve the weaning to a minimal FiO₂ of 0.21 and minimal PEEP level. Apparently, physicians and nurses do not consider hyperoxia as potentially hazardous if FiO₂ is lower than 0.4 and do not adjust ventilator settings in these circumstances. Interestingly, in a questionnaire study from 1999, similar attitudes were reported among Canadian intensivists. Whereas high oxygen saturation measurements would urge intensivists to lower FiO₂ in almost all cases when FiO₂ was higher than 0.4, only 54% of respondents would alter oxygen administration if FiO₂ was lower than 0.4 [6].

Our study is the first to reveal adjustments of ventilator settings by intensivists and ICU nurses in the event of hyperoxia in intensive care patients. The main limitation of this study was the fact that it was a single-centre study. We do not know whether responses to hyperoxia are the same in other ICUs in the Netherlands or other countries. Another limitation of this study is that the clinical condition of the patient may influence decisions to change ventilator settings and that no information on the condition of the patients was available for this analysis. Also, we do not know whether hyperoxia is detrimental in critically ill patients, e.g. by increasing the severity of lung injury.

Conclusion

Hyperoxia was frequently present in ICU patients but in most cases did not lead to adjustment of ventilator settings. If FiO₂ was 0.4 or lower the hyperoxia was accepted in the majority of cases. Further research is needed concerning the effects of frequently encountered hyperoxia in ICU patients. For now and given the experimental evidence of hyperoxia-induced damage, it seems wise to emphasize in guidelines that hyperoxia should be limited to what is needed to ensure adequate tissue oxygenation.

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Conflict of interest The authors have no conflict of interest to declare.

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