A comparison of the genetic and clinical profile of men that respond and do not respond to the immediate antihypertensive effects of aerobic exercise

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Abstract: We compared the genetic and clinical profile of men who lower and do not lower blood pressure (BP) after acute aerobic exercise. Volunteers were 45 men (Mean ± SEM, 43.5 ± 1.5 yr) with high BP (145.7 ± 1.5/85.7 ± 1.1 mmHg). They completed three experiments: nonexercise control and two cycle exercise sessions at 40% and 60% peak oxygen consumption, and were then instrumented to an ambulatory BP monitor. Logistic regression determined the genetic and clinical profile of men who lowered BP after exercise (responders [ExR n = 36]); and those who did not (nonresponders [ExNR n = 9]). ExR had higher C-reactive protein (CRP), high-density lipoprotein (HDL), the metabolic syndrome, family history of hypertension, more renin-angiotensin system (RAS) common alleles, and α-adducin Trp460 and endothelial nitric oxide synthase (ENOS) C786 risk alleles. ExNR had lower CRP and HDL, did not have the metabolic syndrome and a family history of hypertension, had more RAS risk alleles, and had the α-adducin Gly460Gly and ENOS T786T genotypes. This model had a sensitivity of 97.1%, specificity of 75.0%, and accounted for 46.3%–74.4% of the BP response. These results suggest genetic and clinical information may eventually be used to characterize people who do and do not respond to exercise as antihypertensive therapy.

Keywords: blood pressure, genetics, hypertension, metabolic syndrome, physical activity

Introduction
Hypertension is a significant public health problem (AHA 2008; Ong et al 2007). Approximately two thirds of adults in the United States have above optimal blood pressure (BP) (systolic BP [SBP] ≥ 120 and/or diastolic BP [DBP] ≥ 80 mmHg) (Wang and Wang 2004); and nearly all Americans will acquire hypertension if they live into old age (Chobanian et al 2003). Hypertension is the most common primary diagnosis in America and a major risk factor for heart disease, stroke, congestive heart failure, chronic kidney disease, and peripheral arterial disease. The current costs attributed to hypertension are US $63.5 billion annually, with antihypertensive medications accounting for 38% of this amount (AHA 2008). These expenditures underscore the substantial social and economic strain hypertension places upon the health care system.

Aerobic exercise training decreases BP an average of 5–7 mm Hg (Pescatello et al 2004a). Physical activity is recommended as nonpharmacologic therapy to prevent, treat, and control hypertension (Pescatello et al 2004a). Some, if not all, of the BP lowering effects of aerobic exercise training programs are the result of the acute or immediate BP benefits that result from a single exercise session, termed postexercise hypotension (PEH) (Thompson et al 2001). In a systematic review, we reported that PEH does not occur in about 25% of the people with hypertension.
(Pescatello and Kulikowich 2001). The heterogeneity of the individual response to the antihypertensive effects of exercise is attributed to interactions among genetic and environmental factors that are poorly understood (Arnett et al 2007)

There is great promise for the use of exercise genomics in the practice of personalized medicine approaches to exercise prescription (Roth 2007; Turner et al 2007). Yet, work related to identifying genetic and clinical determinants of the antihypertensive effects of exercise has gone slowly. The most recent installment of the human gene map for performance and health-related fitness phenotypes indicates that there have been nine published reports of candidate gene association studies examining the BP response to acute exercise (Rankinen et al 2006a). Collectively, these studies show that: 1) there is a significant genetic component in the acute BP response to exercise with heritability estimates of 45%–55%; 2) baseline BP is a major determinant of the BP response to exercise; and 3) polymorphisms in the renin-angiotensin system (RAS), sympathetic nervous system, and the endothelial L-arginine-nitric oxide synthase pathway associate with the BP response to acute exercise. These reports have primarily involved people with normal BP studied while exercising. Baseline BP is a major determinant of the BP response to exercise. Thus, findings may differ among people with hypertension that are followed in the postexercise period with ambulatory BP monitoring.

We are one of the few research groups that have conducted PEH candidate gene association studies among people with pre- to stage 1 hypertension using ambulatory BP monitoring (Blanchard et al 2006; Pescatello et al 2007a, 2007b). Our findings from these studies show genetic variants in the major BP regulatory pathways explain some of the variability in the BP response to acute aerobic exercise. Furthermore, exercise intensity interacted with genotype to exert differential post exercise BP effects. Other than baseline BP, the genetic and clinical characteristics of people who do and do not lower BP following exercise are largely unknown. The purpose of this study was to compare the genetic and clinical profile of men who do and do not respond to exercise as antihypertensive therapy.

Methods

Subjects

The methods have been published and are described here in abbreviated fashion (Blanchard et al 2006; Pescatello et al 2004b, 2007a, 2007b). Subjects signed an informed consent approved by the Institutional Review Boards of the University of Connecticut and Hartford Hospital. Volunteers were 45 men from 18 to 55 yr with pre- to stage 1 hypertension. They were sedentary, had no physical limitations to perform exercise, and did not smoke. Subjects were not enrolled if they had a SBP ≥ 160 mmHg and/or DBP ≥ 100 mmHg, symptomatic atherosclerotic cardiovascular disease, diabetes mellitus, asthma, thyroid dysfunction, pancreatitis, acute illness, and/or were on medication for depression. Any medications potentially influencing the BP response to exercise including antihypertensives, antilipemics, aspirin, nonsteroidal antiinflammatories, nutritional supplements with the exception of a 1-a-day vitamin, cold medications, and herbal supplements were stopped at least 4 wk before testing occurred. When antihypertensives were withdrawn, investigators regularly monitored participants’ BP. Men in whom the cessation of antihypertensive medication resulted in resting SBP ≥ 160 and/or DBP ≥ 100 mmHg were excluded and resumed their medication.

Procedures

Subjects completed an orientation session to acquaint them with the study and determine if their BP met the inclusion criteria. BP was measured three times, 5 min apart in both arms by auscultation and averaged according to the methods of the Joint National Committee (Chobanian et al 2003). Waist circumference was measured at the height of the iliac crest with a Gulick tape measure (Sammons Preston, Chicago, IL), and height and weight were taken on a standard balance beam scale (Model 339, Detecto, Webb City, MO) to calculate body mass index (kg/m²). During the orientation session, subjects were asked to self disclose a family history of hypertension among first degree relatives as a yes or no. Participants were instructed not to ingest any caffeinated beverage the morning of the testing sessions, and throughout the study to drink caffeinated (≤480 ml [2 cups]) and alcoholic (≤2 drinks/d) beverages in moderation. Otherwise, subjects were asked to maintain their usual diet. Weight maintenance was defined as ± 2.25 kg (5.0 lb) of orientation weight and used as an indication that subjects were adhering to their normal dietary patterns during the study. Men were weighed prior to the exercise test and three experiments to ensure weight maintenance.

At the end of the orientation session an ambulatory BP monitor (Accutracker II automatic noninvasive ambulatory BP monitor, Suntech Medical Instruments Inc., Raleigh, NC) was connected to each subject to acquaint them with the equipment and ensure their BP met the study inclusion criteria. The same investigator attached the ambulatory BP monitor to all subjects. A calibration check was performed with a mercury sphygmomanometer using a
t-tubule according to the manufacturer’s guidelines. The calibration check consisted of three test runs to ensure that three successive ambulatory BP measurements were within 5 mmHg of the auscultatory BP determinations. The monitor was programmed to record BP about every 20 min. BP was taken three times per hour until 2300 hr. All subjects exited the laboratory with instructions to proceed with their usual activities, not to exercise for the remainder of the day, let their arm remain still while the monitor was recording, note any usual emotional or physical events in a diary, and return the monitor and diary the next day. The computerized ambulatory BP recordings were considered acceptable if at least 80% of the BP readings were obtained.

Participants then completed a peak graded cardiopulmonary exercise stress test that was used to determine the workload of the two experimental exercise sessions. The graded exercise stress test was performed on a cycle ergometer (Monark Ergomedic 818E, Stockholm, Sweden). The protocol consisted of 2 min incremental stages of continuous cycling at a constant cadence of 60 rev·min⁻¹ with the resistance increased each stage by 0.5 kp (30 W) until volitional exhaustion. VO₂ peak was determined with breath by breath analysis of expired gases (Sensormedics Vmax 29 Metabolic Chart, SensorMedics Corp., Yorba Linda, CA). HR was recorded continuously with a 12 lead electrocardiogram (ECG) system. The same investigator measured BP by auscultation 30s prior to the end of each 2 min stage. At the conclusion of the graded cardiopulmonary exercise stress test, subjects were again attached to an ambulatory BP monitor to further familiarize them with the unit.

Subjects completed three, 40 min experiments in random order on three separate days that were at least 48 hr apart to avoid acute exercise effects on BP. Each experiment began with a 20 min baseline period of seated rest during which BP and HR were recorded every 2 min. HR was obtained with a HR monitor (Model #: 1902750, Polar Electro Inc, Woodbury, NY), and BP by auscultation in the nondominant arm (ie, hand with which the subject did not write). The same study investigator made all BP determinations during every experiment for all subjects. The BP measurements taken during the baseline period were averaged and denoted as baseline BP. The BP recordings were considered acceptable if at least 80% of the baseline period were averaged and denoted as baseline BP. The investigator made all BP determinations during every experiment for all subjects. The BP readings were obtained.

Blood sampling and analysis
Blood samples were taken in advance of the experiments for the purposes of genotyping and fasting blood lipid-lipoprotein, glucose and insulin determinations. Blood samples were also obtained during the experiments for high sensitivity-CRP as follows: at the end of the baseline period, at 30 min of each 40 min experiment, and at the conclusion of the experiments while still in the laboratory but prior to attachment to the ambulatory BP monitor. Samples were drawn into anti-coagulated EDTA tubes and centrifuged at 2500 g and 23 °C for 6 min. Plasma samples were transferred to storage tubes and frozen at −80 °C until analysis. Blood lipids and lipoproteins were determined by oxidoassays using colorimetric enzymatic methods (Cobras® Integra™, Roche Diagnostics, Mannheim, Germany). The intra- and inter-assay CV for the triglyceride assays were 1.6% and 1.9%, respectively. Low density lipoprotein was calculated with the Friedewald equation (Friedewald et al 1972). Plasma concentrations of insulin were measured in duplicate by radioimmunoassay using a solid phase, single antibody assay (Coat-a-Count Insulin, TKN2, Diagnostic Products Corporation, Los Angeles, CA) with an intra-assay CV of 7.0% and inter-assay CV of 7.8%. A multi-level control (CON6 Multivalent Control Module, Diagnostic Procedures Corporation) was analyzed during each insulin test run. Plasma glucose concentrations were determined in duplicate by an automated glucose oxidase method (Yellow Springs Instruments, Model 2003, Yellow Springs, OH). The intra- and inter-assay CV for the plasma glucose determinations were 2.8% and 3.6%, respectively, at 4.7 mmol·L⁻¹. The homeostasis model assessment (HOMA) of insulin resistance was calculated with the formula: [fasting glucose (mg·dL⁻¹) × insulin (uU·mL⁻¹)]·22.5⁻¹ (Matthews et al 1985). High sensitivity CRP concentrations were measured via particle enhanced immunoturbidimetry on the COBAS INTEGRA analyzer (CRP latex cassette, Roche Diagnostic Corporation, Indianapolis, IN) with low-, normal- and high-quality control values. The inter-assay CRP CV was 3.22%, 3.85%, and 3.66%, respectively.

Genotype methods
Genomic DNA was isolated from EDTA containing vacutainer tubes using Puregene kits (Gentra Systems, Inc, MN). Polymerase chain reaction based restriction fragment length
an analysis was performed using separate primers described previously to determine mutations in the angiotensin-converting enzyme insertion/deletion polymorphism (ACE DIP) (rs4646994) (Shamugam et al 1993), angiotensin II type 1 receptor (AT,R) 1166 A > C single nucleotide polymorphism (SNP) (rs5186) (Berg and Berg 1998), intron-2 conversion of the aldosterone synthase gene (CYP11B2) (Nicod et al 2003), α-adducin Gly460Trp SNP (rs4961) (Pescatello et al 2007a), α-subunit of guanine nucleotide-binding system (GNAS) 393 T > C SNP (rs7121) (Yang et al 2004), and endothelial nitric oxide synthase (ENOS) 786 T > C SNP (rs20270744) (Metzer et al 2007). Negative and positive controls (known genotypes) and random duplicates were used as quality control. In the event of discordant results, genotyping was repeated. All polymorphisms were in Hardy-Weinberg equilibrium.

Statistical analyses

Descriptive statistics were obtained on all study parameters. The BP difference from baseline was averaged at hourly intervals after the experimental sessions over the course of 9 hr; the time period over which PEH was noted when the men were awake and ambulating (Pescatello et al 2004b). The statistical analyses for the BP data used a two-pronged approach. The first approach involved repeated measures ANOVA to test if BP differed over time among experimental conditions (non-exercise control, LIGHT, MODERATE) and EXERCISE, LIGHT, and MODERATE responders and nonresponders. Small reductions in BP yield significant public health benefits in terms of reducing risks from coronary artery disease and stroke (Whelton et al 2002). We used the cohort and data from our previous work (Blanchard et al 2006; Pescatello et al 2004b, 2007a, 2007b) to define ExR as men with an average BP change from baseline that was reduced ≥1 mmHg after LIGHT and/or MODERATE exercise compared to nonexercise control over 9 hr. Men with an average BP change from baseline that was reduced <1 mmHg after LIGHT and/or MODERATE compared to nonexercise control over 9 hr were defined as an ExNR. Similarly, LIGHT responders were defined as men with an average BP change from baseline that was reduced ≥1 mmHg after LIGHT compared to nonexercise control over 9 hr; and all else were classified as LIGHT nonresponders. MODERATE responders were defined as men with an average BP change from baseline that was reduced ≥1 mmHg after MODERATE compared to nonexercise control over 9 hr; and all else were classified as MODERATE nonresponders.

The second approach used the BP response to exercise as a dichotomous dependent variable (ExR and ExNR). Independent t-tests determined if there were differences in subject clinical characteristics among ExR and ExNR. Genetic and clinical predictors of EXERCISE, LIGHT and MODERATE responders and nonresponders were tested using binary logistic regression models. For RAS genetic variants (ie, ACE DIP, AT,R 1166 A > C SNP, and CYP11B2) a genotype combination score of “at risk” alleles was calculated as 0–2 and ≥3 RAS risk alleles (Blanchard et al 2006), and this score was the dichotomous variable used to reflect RAS epistatic effects in the logistic regression analyses. Due to the low prevalence of the α-adducin Trp460 risk allele in the general population, no man was found to have the α-adducin TrpTrp genotype so that two α-adducin genotypes (GlyGly and GlyTrp) were used in the logistic regression analyses (Pescatello et al 2007a). Clinical markers entered into these models included: VO2 peak; waist circumference; fasting blood lipid-lipoproteins, glucose and insulin; CRP, and family history. Pseudo R2 values from the full logistic regression models (models containing genotype and clinical markers) were compared to constrained models (models containing only clinical markers) to estimate the proportion of variance in the BP response to exercise that was attributed to genetic variation. The total sample of ExR and ExNR consisted of 45 men; however, the sample size of EXERCISE (n = 42), LIGHT (n = 43), and MODERATE (n = 44) responders and nonresponders differed slightly due to genetic and clinical markers not being obtained on all 45 men in each of these three statistical models.

The final models displayed in Tables 3–5 were selected because: 1) they provided the best goodness of fit of the estimates fit to the actual outcomes; 2) the likelihood ratio test was not significant indicating the logistic regression model was more effective than the null model; and 3) the Cox and Snell and Nagelkerke pseudo R2, two additional descriptive measures of goodness of fit that are similar to the r2 concept of linear regression, explained a large proportion of the variability in the BP response to exercise. All statistical analyses were performed with Statistical Package for the Social Sciences for Windows version 14.0 (SPSS Inc., Chicago, IL) with p < 0.05 established as the level of significance.

Results

Subjects

Subjects were 45 overweight, middle-aged Caucasian men who had pre- to stage 1 hypertension, borderline dyslipidemia (NCEP 2002), and low cardiovascular fitness (ACSM 2006) (Table 1). Of these men, 40.0% had the Msyn (NCEP 2002),
51.1% reported a family history of hypertension, and 80.0% were classified as ExR. Clinical characteristics were not different between ExR and ExNR (p ≥ 0.05).

**Blood pressure response**

Table 2 displays the average BP difference from baseline after exercise versus nonexercise control over 9 hr among the men classified as EXERCISE, LIGHT, and MODERATE responders and nonresponders. In addition, the average BP change from baseline after exercise and nonexercise control at hourly intervals over 9 hr among ExR and ExNR is displayed in Figures 1 and 2, respectively. Among EXERCISE (Figure 1), LIGHT, and MODERATE responders, BP was lower after LIGHT and MODERATE compared to nonexercise control (Table 2) (p < 0.01). Among ExNR (Figure 2), BP was higher after LIGHT and MODERATE versus nonexercise control (p < 0.05). Among LIGHT nonresponders, BP was higher after LIGHT (p < 0.01) but not after MODERATE than nonexercise control (p ≥ 0.05). Among MODERATE nonresponders, SBP was higher after MODERATE (p < 0.01) but not LIGHT (p ≥ 0.05); and DBP was higher after LIGHT and MODERATE compared to nonexercise control (p < 0.05). Under these conditions, the postexercise BP response differed in all instances between EXERCISE, LIGHT and MODERATE responders versus nonresponders (Table 2) (p < 0.05). In addition, 84.4% of the men classified as LIGHT responders were deemed MODERATE responders; and 69.2% of LIGHT nonresponders were classified as MODERATE nonresponders.

**Genetic and clinical cardiometabolic profile**

**Exercise responders and nonresponders**

The genetic and clinical features of the cardiometabolic profile that emerged from the logistic regression analyses of ExR (n = 34, 81.0%) and ExNR (n = 8, 19.0%) were obtained on 42 of the 45 men enrolled in this study and is shown in Table 3. Based upon the designation of ExR as the reference category and the β in Table 3, ExR had: higher CRP and high density lipoprotein (HDL); the Msyn; a self-disclosed family history.

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**Table 1** Mean (± SEM) fasting clinical characteristics of the total sample and among exercise responders and nonresponders*

| Clinical characteristic | Total sample (n = 45) | Exercise responders (n = 36) | Exercise nonresponders (n = 9) |
|------------------------|----------------------|-----------------------------|-----------------------------|
| Age (yr)               | 43.5 ± 1.5           | 44.0 ± 1.7                   | 41.4 ± 3.4                   |
| Peak oxygen consumption (ml.kg⁻¹.min⁻¹) | 30.8 ± 1.0          | 30.4 ± 1.1                   | 32.4 ± 2.4                   |
| Body mass index (kg/m²) | 29.7 ± 0.7           | 29.6 ± 0.9                   | 30.0 ± 0.8                   |
| Waist circumference (cm) | 102.6 ± 2.2         | 102.1 ± 2.5                  | 104.7 ± 4.0                  |
| Orientation awake systolic blood pressure (mmHg)* | 145.7 ± 15          | 145.0 ± 1.8                  | 148.8 ± 2.6                  |
| Orientation awake diastolic blood pressure (mmHg) | 85.7 ± 1.1          | 85.2 ± 1.3                   | 87.7 ± 1.8                   |
| C-reactive protein (mg.L⁻¹) | 1.6 ± 0.2           | 1.7 ± 0.2                    | 1.2 ± 0.2                    |
| Insulin (pmol.L⁻¹)      | 76.7 ± 8.0           | 74.6 ± 8.5                   | 85.3 ± 8.0                   |
| Glucose (mmol.L⁻¹)      | 4.5 ± 0.1            | 4.5 ± 0.1                    | 4.6 ± 0.2                    |
| Homeostasis model assessment | 2.2 ± 1.6         | 2.2 ± 1.6                    | 2.3 ± 0.6                    |
| Total cholesterol (TCHOL) (mmol.L⁻¹) | 4.9 ± 0.2           | 5.0 ± 0.2                    | 4.6 ± 0.3                    |
| High-density lipoprotein-cholesterol (mmol.L⁻¹) | 1.1 ± 0.0           | 1.1 ± 0.0                    | 1.0 ± 0.0                    |
| Low-density lipoprotein-cholesterol (mmol.L⁻¹) | 3.1 ± 0.2           | 3.1 ± 0.2                    | 2.8 ± 0.2                    |
| Total cholesterol/high-density lipoprotein ratio (U) | 4.7 ± 0.2           | 4.6 ± 0.2                    | 4.9 ± 0.2                    |
| Triglycerides (mmol.L⁻¹) | 1.6 ± 0.3           | 1.6 ± 0.2                    | 2.0 ± 0.4                    |

Notes: Exercise responders; the average change in blood pressure from baseline ≥ 1 mmHg less after LIGHT and/or MODERATE compared to nonexercise control over 9 hr; all else were classified as nonresponders; Orientation awake blood pressure; ambulatory blood pressure averaged over the time period following the orientation session when men were awake, ie, day time blood pressure.

Abbreviations: LIGHT, 40% VO₂ peak; MODERATE, 60% VO₂ peak.
of hypertension; and more RAS common alleles (0–2); and were carriers of the risk allele of the α-adducin Gly460Trp (GlyTrp genotype) and ENOS 786 T > C (TC/CC genotypes) SNPs compared to ExNR. In contrast, ExNR: had lower CRP and HDL; did not have the Msyn and a family history of hypertension; had more RAS risk alleles (3+); and were homozygous for the common alleles of the α-adducin Gly460Trp (GlyGly genotype) and ENOS 786 T/H113501 mmHg less after LIGHT and/or MODERATE compared to nonexercise control over 9 hr; all else were classified as nonresponders; LIGHT, 40% VO2 peak; MODERATE, 60% VO2 peak; p < 0.05, p < 0.01, p < 0.001 blood pressure difference exercise vs nonexercise control; |p| < 0.05, |p| < 0.01, |p| < 0.001 responders vs nonresponders.

**Table 2** Blood pressure difference (mean ± SEM (95% confidence interval)) from baseline after exercise versus nonexercise control over 9 hr among EXERCISE, LIGHT, and MODERATE responders and nonresponders*

|                  | SBP difference (mmHg) | DBP difference (mmHg) |
|------------------|-----------------------|-----------------------|
|                  | EXERCISE              | LIGHT MODERATE        | LIGHT MODERATE        |
|                  | RESPONDER             | NONRESPONDER          | RESPONDER             | NONRESPONDER          |
| (n = 43)         | (n = 30)              | (n = 44)              | (n = 30)              | (n = 43)              |
|                  | (n = 42)              | (n = 8)               | (n = 42)              | (n = 34)              |
|                  | (n = 34)              | (n = 13)              | (n = 34)              | (n = 27)              |
| LIGHT RESPONDER  | −5.2 ± 1.3*           | −7.8 ± 1.4**          | −3.9 ± 1.1**          | −3.6 ± 1.1*           |
| LIGHT NONRESPONDER | −7.9, −2.5           | −10.5, −5.0           | −6.0, −1.7           | −5.8, −1.5            |
| MODERATE RESPONDER | 11.3 ± 3.5*           | 5.1 ± 1.9*           | 10.0 ± 2.4*          | 5.1 ± 1.5*           |
| MODERATE NONRESPONDER | 3.1, 19.6           | 1.2, 10.0           | −4.3, 15.7           | (1.5, 8.6)            |

**Notes:** *Exercise responders; the average change in blood pressure from baseline ≥ 1 mmHg less after LIGHT and/or MODERATE compared to nonexercise control over 9 hr; all else were classified as nonresponders; LIGHT, 40% VO2 peak; MODERATE, 60% VO2 peak; p < 0.05, p < 0.01, p < 0.001 blood pressure difference exercise vs nonexercise control; |p| < 0.05, |p| < 0.01, |p| < 0.001 responders vs nonresponders.

Abbreviations: LIGHT, 40% VO2 peak; MODERATE, 60% VO2 peak.

**Figure 1** The average adjusted systolic blood pressure change from baseline after LIGHT and MODERATE compared to nonexercise control at hourly intervals over 9 hr among exercise responders.

**Notes:** *Exercise responders; the average change in blood pressure from baseline ≥ 1 mmHg less after LIGHT and/or MODERATE compared to nonexercise control over 9 hr; all else were classified as nonresponders.

**Abbreviations:** LIGHT, 40% VO2 peak; MODERATE, 60% VO2 peak.
compared to the pseudo R² of the constrained logistic regression model, the genetic component accounted for 40.5% to 43.0% of the variability in the BP response to exercise.

**LIGHT responders and nonresponders**

The genetic and clinical components of the cardiometabolic profile that emerged from the logistic regression analyses of LIGHT responders (n = 30, 69.8%) and nonresponders (n = 13, 30.2%) were obtained on 43 of the 45 men (Table 4). Based upon the designation of LIGHT responders as the reference category and the β in Table 4, LIGHT responders: had higher CRP and HDL; did not have the Msyn; had a family history of hypertension; had more RAS common alleles (0–2); and were carriers of the risk allele of the α-adducin Gly460Trp.

![Figure 2](image)

**Table 3 Logistic regression model of genetic and clinical correlates of exercise responders (n = 34) and nonresponders (n = 8) to the immediate antihypertensive effects of aerobic exercise**

| Determinant                  | β      | Standard error β | X²    | df | p    | eβ odds ratio |
|------------------------------|--------|------------------|-------|----|------|---------------|
| C-reactive protein           | −16.396| 8.721            | 3.535 | 1  | 0.060| 0.000         |
| High-density lipoprotein     | −0.532 | 0.273            | 3.798 | 1  | 0.051| 0.587         |
| Metabolic syndrome^b         | −2.612 | 2.369            | 1.215 | 1  | 0.270| 0.073         |
| Family history^c             | −6.987 | 3.564            | 3.842 | 1  | 0.050| 0.001         |
| RAS polymorphisms^d          | 0.910  | 2.022            | 0.203 | 1  | 0.653| 2.485         |
| α-adducin Gly460Trp          | −0.947 | 1.592            | 0.354 | 1  | 0.552| 0.388         |
| ENOS 786 T > C^f             | −2.569 | 2.056            | 1.562 | 1  | 0.211| 0.077         |
| Test                         |        |                  |       |    |      |               |
| Likelihood ratio             | 14.770 | 7                | 0.000 |    |      |               |
| Goodness-of-fit              | 0.397  | 8                | 1.000 |    |      |               |

Notes: ^aResponders (Referent Group); the average change in blood pressure from baseline ≥1 mmHg less after LIGHT and/or MODERATE compared to nonexercise control over 9 hr; all else were classified as nonresponders; LIGHT, 40% VO2 peak; MODERATE, 60% VO2 peak; ^bMetabolic Syndrome, absent (n = 26) or present (n = 16) by National Cholesterol Education Adult Treatment Panel III Criteria (NCEP 2002); ^cFamily History, self disclosed family history of hypertension among first degree relatives no (n = 21) or yes (n = 21); ^dRAS SNPs, 0–2 risk alleles (n = 20) or ≥3 risk alleles (n = 22) from three polymorphisms of the RAS. Polymorphisms were angiotensin-converting enzyme insertion deletion polymorphism, an adenine/cytosine substitution at position 1166 of the angiotensin II type 1 receptor gene, and a gene conversion in intron 2 of the aldosterone synthase gene; ^eα-adducin Gly460Trp, GlyGly (n = 32) or GlyTrp (n = 10); ^fENOS 786 T > C, a threonine/cysteine substitution at position 786 of the endothelial nitric oxide synthase gene; TT (n = 23) vs TC/CC (n = 19).
Table 4 Logistic regression model of genetic and clinical correlates of LIGHT responders (n = 30) and nonresponders (n = 13) to the immediate antihypertensive effects of aerobic exercise*  

| Determinant                        | β       | Standard error β | X²     | df | p   | e^β odds ratio |
|------------------------------------|---------|------------------|--------|----|-----|----------------|
| C-reactive protein                 | −21.543 | 10.413           | 4.280  | 1  | 0.039| 0.000          |
| High-density lipoprotein           | −0.090  | 0.074            | 1.496  | 1  | 0.221| 0.914          |
| Metabolic syndrome                | 1.540   | 1.588            | 0.940  | 1  | 0.332| 4.664          |
| Family history                     | −1.866  | 1.301            | 2.057  | 1  | 0.151| 0.155          |
| RAS polymorphisms                 | 0.729   | 1.080            | 0.456  | 1  | 0.499| 2.074          |
| α-adducin Gly460Trp              | −2.692  | 1.492            | 3.254  | 1  | 0.071| 0.068          |
| GNAS 393 T > C                  | 1.453   | 1.197            | 1.472  | 1  | 0.225| 4.274          |

Notes: *LIGHT, 40% VO2 peak; Responders (Referent Group), the average change in blood pressure from baseline ≥1 mmHg less after LIGHT compared to nonexercise control over 9 hr; all else were as nonresponders; Meta-determinant, absent (n = 26) or present (n = 17) by National Cholesterol Education Adult Treatment Panel III Criteria (NCEP 2002); cFamily history, self-disclosed family history of hypertension among first degree relatives; no (n = 22) or yes (n = 21); dRAS SNPs, 0–2 risk alleles (n = 20) or ≥3 risk alleles (n = 23) from three polymorphisms of the RAS. Polymorphisms were angiotensin-converting enzyme insertion deletion polymorphism, an adenine/cytosine substitution at position 1166 of the angiotensin II type 1 receptor gene, and a gene conversion in intron 2 of the aldosterone synthase gene; eα-adducin Gly460Trp, Gly/Gly (n = 33) or Gly/Trp (n = 10); GNAS 393 T > C, a threonine/cysteine substitution at position 393 of the alpha subunit of the guanine binding system protein gene; TT/TC (n = 33) vs CC (n = 10).

(GlyTrp genotype) and GNAS 393 T > C (TT/TC) SNPs compared to LIGHT nonresponders. In contrast, LIGHT nonresponders: had lower CRP and HDL; had the Msyn; did not have a family history of hypertension; had more RAS risk alleles (3+); were homozygous for the common alleles of the α-adducin Gly460Trp (GlyGly genotype) SNP and GNAS 393 T > C (CC genotype) SNPs compared to LIGHT responders. The logistic regression model displayed in Table 4: 1) correctly classified 28 out of 30 LIGHT responders for a sensitivity of 93.3%, and nine out of 12 LIGHT nonresponders for a specificity of 75.0%; and 2) explained 41.0% (Cox and Snell R²) to 58.0% (Nagelkerke R²) of the variability in the BP response to exercise. When the pseudo R² of the full logistic regression model was compared to the pseudo R² of the constrained logistic regression model, the genetic component accounted for 40.7% of the variability in the BP response to exercise.

MODERATE responders and nonresponders

The genetic and clinical components of the cardiometabolic profile that emerged from the logistic regression analyses of MODERATE responders (n = 31, 70.5%) and nonresponders (n = 13, 29.5%) were obtained on 44 of the 45 men (Table 5). Based upon the designation of MODERATE responders as the reference category and the β in Table 5, MODERATE responders: had higher baseline BP and HDL; the Msyn; a family history of hypertension; more RAS common alleles (0–2); and were homozygous for the common allele of the α-adducin Gly460Trp (GlyGly genotype) SNP; and carriers of the risk allele of the ENOS 786 T > C (TC/CC genotypes) SNP compared to MODERATE nonresponders. In contrast, MODERATE nonresponders: had lower baseline BP and HDL; did not have the Msyn and a family history of hypertension; had more RAS risk alleles (3+); were carriers of the risk allele of the α-adducin Gly460Trp (GlyTrp genotype) SNP; and were homozygous for the common allele of the ENOS 786 T > C (TT genotype) SNP compared to MODERATE responders. The logistic regression model displayed in Table 4: 1) correctly classified 27 out of 31 MODERATE responders for a sensitivity of 87.1%, and nine out of 13 MODERATE nonresponders for a specificity of 69.2%; and 2) explained 41.4% (Cox and Snell R²) to 58.9% (Nagelkerke R²) of the variability in the BP response to exercise. When the pseudo R² of the full logistic regression model was compared to the pseudo R² of the constrained logistic regression model, the genetic component accounted for 65.9% to 66.2% of the variability in the BP response to exercise.

Discussion

The purpose of this study was to compare the genetic and clinical profile of men with pre- to stage 1 hypertension who did and did not respond to aerobic exercise as antihypertensive therapy. We found ExR had: higher CRP and HDL; the Msyn; a self-disclosed family history of hypertension; and more RAS common alleles (0–2); and were carriers of the
Table 5 Logistic regression model of genetic and clinical correlates of MODERATE responders (n = 31) and nonresponders (n = 13) to the immediate antihypertensive effects of aerobic exercise*a

| Determinant                                      | β       | Standard error β | X²    | df | p   | e^β odds ratio |
|-------------------------------------------------|---------|------------------|-------|----|-----|----------------|
| Baseline blood pressure                         | -0.068  | 0.048            | 2.016 | 1  | 0.156| 0.935          |
| High-density lipoprotein                        | -0.269  | 0.113            | 5.709 | 1  | 0.017| 0.674          |
| Metabolic syndrome                              | -2.853  | 1.516            | 3.541 | 1  | 0.060| 0.058          |
| Family history of hypertension                  | -3.100  | 1.388            | 4.992 | 1  | 0.025| 0.045          |
| RAS polymorphisms                               | 1.662   | 1.159            | 2.057 | 1  | 0.152| 5.269          |
| α-adducin Gly460Trp                             | 1.491   | 1.281            | 1.355 | 1  | 0.244| 4.442          |
| ENOS 786 T > C                                 | -2.666  | 1.491            | 3.198 | 1  | 0.074| 0.069          |
| Test                                            |         |                  |       |    |     |                |
| Likelihood ratio                                | 29.903  |                  | 7     | 0.001|     |                |
| Goodness-of-fit                                 | 7.077   |                  | 8     | 0.528|     |                |

Notes: *MODERATE, 60% VO2 peak; Responders (Referent Group), the average change in blood pressure from baseline ≥1 mmHg less after MODERATE compared to nonexercise control over 9 hr; all else were classified as nonresponders; bMetabolic Syndrome, absent (n = 27) or present (n = 17) by National Cholesterol Education Adult Treatment Panel III Criteria (NCEP 2002); cFamily history; self-disclosed family history of hypertension among first degree relatives: no (n = 22) or yes (n = 22); dRAS SNPs, 0-2 risk alleles (n = 21) or ≥3 risk alleles (n = 23) from three polymorphisms of the RAS. Polymorphisms were angiotensin-converting enzyme insertion deletion polymorphism, an adenine/cytosine substitution at position 1166 of the angiotensin II type 1 receptor gene, and a gene conversion in intron 2 of the aldosterone synthase gene; eENOS 786 T > C, a threonine/cysteine substitution at position 786 of the endothelial nitric oxide synthase gene; fα-adducin Gly460Trp, Gly/Gly (n = 33) or Gly/Trp (n = 11); gSNPs (Table 3). The genetic component of this model accounted for nearly two thirds of the variability in the BP response, with over 40% of this variability due to genetic factors. In addition, the model had a high sensitivity (97.1% of ExNR correctly classified) and adequate specificity (75.0% of ExNR correctly classified). These findings indicate that genetic and clinical information may eventually be used to characterize people who do and do not respond to exercise as antihypertensive therapy. Such information will ultimately enable clinicians and sports medicine professionals to personalize exercise prescriptions to maximize the effectiveness of exercise as a therapeutic option to prevent, treat and control hypertension.

Most men classified as LIGHT responders were MODERATE responders (ie, 84.4%). Despite the adequate repeatability of the BP phenotype, the cardiometabolic profile of LIGHT and MODERATE responders differed. LIGHT responders: had higher CRP and HDL; did not have the Msyn; had a family history of hypertension; more RAS common alleles (0–2); the α-adducin GlyTrp genotype; and were carriers of the ENOS C786 risk allele (TT/TC genotypes) (Table 4). The genetic component of this model accounted for just over 40% of the variability in the BP response. MODERATE responders: had higher baseline BP and HDL; the Msyn; a family history of hypertension; more RAS common alleles (0–2); and the α-adducin GlyGly genotype; and were carriers of the ENOS C786 risk allele (TC/CC genotypes) (Table 5). The genetic component of this model accounted for nearly two thirds of the variability in the BP response. Thus, CRP, baseline BP, and the presence or absence of the Msyn are the clinical determinants that appeared to differentiate between men who responded to LIGHT and MODERATE as antihypertensive therapy. Furthermore, the genetic makeup of men who lowered BP after LIGHT differed from those who lowered BP after MODERATE, and genetic factors accounted for more of the variability in the BP response after MODERATE than LIGHT. These clinical and genetic differences in the cardiometabolic profile of LIGHT and MODERATE responders may ultimately offer explanations for the differential exercise intensity of BP-lowering effects we (Blanchard et al 2006; Pescatello et al 2004b, 2007a, 2007b) and others (Quinn 2000; Smelker et al 2004) have reported; and suggest the mechanisms for the BP-lowering effects of exercise may differ by the intensity or level of physical exertion of the exercise bout.

In this study we examined a limited number of polymorphisms in a small sample of Caucasian, middle-aged men raising the possibility of finding false positive results. We preselected the polymorphisms examined in this study based upon literature evidence of their associations with hypertension and the exercise-induced BP response and location in major BP and metabolic regulatory pathways reducing the likelihood of false positive statistical results (Rankinen
et al 2000, 2006a, 2006b; Rankinen 2007). We used simple clinical measures of overall adiposity and insulin sensitivity. It remains possible that more sensitive measures of adiposity than the body mass index and waist circumference and of insulin sensitivity than HOMA may have shown significant associations with the postexercise BP response which we did not find in this study.

Nonetheless, our study has important strengths that increased our likelihood of finding associations among genetic and clinical determinants of the postexercise BP response (Cardon and Palmer 2003; Arnett et al 2007). These strengths included: 1) a randomized control design; 2) stringent assessment of the BP phenotype with ambulatory BP monitoring to isolate variability in the BP measurement; 3) purposeful inclusion of middle-aged Caucasian men to minimize genetic admixture; 4) a priori selection of polymorphisms; and 5) a functional approach with two different doses of exercise to isolate complex gene-gene and gene-environment interactions with the BP response to exercise. It is also important to note that our results confirm reports of others that there is a significant genetic component to the acute BP response to exercise (Bouchard and Rankinen 2001); and polymorphisms from the RAS, sympathetic nervous system, and ENOS pathway associate with the BP response to acute aerobic exercise (Rankinen et al 2000, 2006a; Rankinen 2007). Furthermore, finding family history to be an important component of the cardiometabolic profile suggests that other genetic factors contribute to the antihypertensive effects of exercise that remain to be found.

In conclusion, we have constructed models that profile the genetic and clinical characteristics of men who did and did not respond to exercise as antihypertensive therapy. Due to the small sample size and limited number of polymorphisms examined, these findings should be regarded as preliminary and hypothesis generating rather than hypothesis testing. Nonetheless, they illustrate the promise of the use of exercise genomics in personalized medicine. Genetic and clinical information may eventually be used by clinicians to individualize exercise prescriptions for subgroups of people that will maximize the effectiveness of exercise as antihypertensive therapy. The eventual translation of this information into clinical practice will enable exercise to take on increased importance in the prevention, treatment, and control of hypertension.

Disclosure
The authors report no conflicts of interest in this work.

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