Evolutionary game theory of growth factor production: implications for tumour heterogeneity and resistance to therapies

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Background: Tumour heterogeneity is documented for many characters, including the production of growth factors, one of the hallmarks of cancer. What maintains heterogeneity remains an open question that has implications for diagnosis and treatment, as drugs that target growth factors are susceptible to the evolution of resistance.

Methods: I use evolutionary game theory to model collective interactions between cancer cells, to analyse the dynamics of the production of growth factors and the effect of therapies that reduce their amount.

Results: Five types of dynamics are possible, including the coexistence of producer and non-producer cells, depending on the production cost of the growth factor, on its diffusion range and on the degree of synergy of the benefit it confers to the cells. Perturbations of the equilibrium mimicking therapies that target growth factors are effective in reducing the amount of growth factor in the long term only if the reduction is extremely efficient and immediate.

Conclusion: Collective interactions within the tumour can maintain heterogeneity for the production of growth factors and explain why therapies like anti-angiogenic drugs and RNA interference that reduce the amount of available growth factors are effective in the short term but often lead to relapse. Alternative strategies for evolutionarily stable treatments are discussed.

Self-sufficiency of growth factor production is one of the hallmarks of cancer (Hanahan and Weinberg, 2000) and, like for other characters, there is evidence of intra-tumour heterogeneity in the ability of cells to produce growth factors (Achilles et al, 2001; Marusyk and Polyak, 2010). Jouanneau et al (1994) have shown that tumour cells cooperate for the production of FGF, that is, cells that do not produce the growth factor can benefit from the products of their neighbours. As they are diffusible molecules, it stands to reason that this is the case for other growth factors as well (Axelrod et al, 2006). What maintains heterogeneity? Given that cancer progression is a process of clonal selection (Cairns, 1975; Nowell, 1976; Crespi and Summers, 2005; Merlo et al, 2006; Greaves and Maley, 2012) in which cells compete for resources, space and nutrients, how can more than one clone stably coexist in a neoplasm? Current explanations include the possibility that different clones are evolutionarily neutral (Iwasa and Michor, 2011), specialise on different niches (Nagy, 2004; Gatenby and Gillies, 2008) or are not in equilibrium (Gonzalez-Garcia et al, 2002); which, if any, of these mechanisms are at work in neoplasms remains an open question (Merlo et al, 2006).

Heterogeneity has also implications for disease progression (Maley et al, 2006), diagnosis and therapy (Dexter and Leith, 1986). As diagnostic biopsies sample only a small region of the tumour, treatments based upon such samples might not be effective against all tumour cells. Understanding the origin, extent and dynamics of tumour heterogeneity, therefore, is essential for the development of successful anticancer therapies. It has been suggested that treatments that attack growth factors may be less susceptible than traditional drugs to the evolution of resistance (Pepper, 2012; Akitips and Nesse, 2013). Current drugs that target growth factors, however, like the anti-angiogenic drug Avastin, extend overall survival by only a few months (Amit et al, 2013), as resistance often arises.

We develop a model of growth factor production in the framework of evolutionary game theory and show how the evolutionary dynamics of the system can explain the maintenance...
of stable heterogeneity, how this affects the development of resistance to anticancer therapies that target growth factors, and show the implications for the development of stable therapies.

A fundamental analytical method in evolutionary biology, game theory has often been mentioned (for example, Gatenby and Maini, 2003; Axelrod et al, 2006; Merlo et al, 2006; Basanta and Deutsch, 2008; Lambert et al, 2011) as a promising avenue for cancer research, but only a few papers actually develop game theoretical models of cancer. Tomlinson (1997) and Tomlinson and Bodmer (1997) developed a mathematical model of a population consisting of two types of cells, with one type producing a factor that confers a proliferative advantage to both cells. Subsequent papers have extended this model up to four types of cells (Dingli et al, 2009; Basanta et al, 2008a,b, 2011; 2012; Gerstung et al, 2011) and allowed stochastic and spatial effects (Bach et al, 2003). These models, however, describe interactions between pairs of cells, whereas interactions among cancer cells are not normally pairwise and should be described by public goods games (PGGs) with collective interactions. It is known that results from the theory of two-player games cannot be extended to multiplayer PGGs and that this can actually lead to fundamental misunderstandings (Archetti and Scheuring, 2012).

Models of PGGs in evolutionary game theory, on the other hand (reviewed by Archetti and Scheuring (2012)) assume that the benefit of the public good is a linear function of the number of contributors (the N-person Prisoner’s Dilemma: NPD) or a step function with a fixed threshold (Archetti, 2009a,b). As the effect of enzyme production is generally a saturating function of its concentration (Hemker and Hemker, 1969; Ricard and Noat, 1986; Mendes, 1997; Eungdamrong and Iyengar, 2004), however, the public good produced by growth factors is likely to be a sigmoid function of the number of producer cells. We must therefore resort to a model with sigmoid benefits, which has been so far beyond the reach of evolutionary game theory (Archetti and Scheuring, 2011, 2012). As the dynamics of the NPD and that of threshold PGGs is radically different, it is not clear a priori, what the dynamics of PGGs with sigmoid benefits could be.

The model we use here, therefore, goes beyond current game theory models of cancer in that it is a multiplayer, collective action (public goods) game, rather than a game with two players, and it goes beyond standard models in evolutionary biology in that it assumes nonlinear, sigmoid benefits, and investigates the effect of therapies that affect the dynamics. To make the analysis possible, we must assume that cells maintain a large and constant population size, which may describe cancer cell populations that have reached a carrying capacity, due, for example, to resource limitation or immune response. The model is relevant for insulin-like growth factor (IGF) and other growth factors that confer direct benefits by inducing a growth advantage or by protecting against apoptosis. Other growth factors like vascular endothelial growth factor (VEGF) whose benefits are less direct will have more complicated dynamics.

MATERIALS AND METHODS

We consider the case in which cells can be either producers of the growth factor (cooperators; C) or non-producers (defectors; D); producers pay a cost of production $c > 0$ and the benefit $b(j)$ for a cell is an increasing function of the number of contributors $j$ among the other cells in the group (of size $n$) defined by the diffusion range of the growth factor, that is, $\Delta b_j = b(j + 1) - b(j) > 0$ for $j = 0, \ldots, n - 1$. More specifically, we assume that the benefit function has a sigmoid shape (for a given $j^*$, $j < j^* \Rightarrow \Delta b_{j+1} \geq \Delta b_j, j \geq j^* \Rightarrow \Delta b_{j+1} \leq \Delta b_j$). We use the simplest and most natural sigmoid function, the logistic function $b(j) = 1/(1 + e^{\beta(j - \beta)})$, where $k$ is the inflection point at which the function has steepness $s$ (with $0 < k \leq n$ and $s > 0$). A normalised version of the logistic function, given by $b_n(j) = [b(j) - b(0)]/[b(n) - b(0)]$, delivers the case of a fixed threshold (a benefit is produced if and only if at least $k$ producers are present; $s \to \infty$) and of a linear function ($s \to 0$) as limit cases. In a large population with no assortment, we can approximate the analysis by assuming an infinite, well-mixed population, and the fitness of producers and non-producers is given as follows:

$$
\pi_C(x) = \sum_{j=0}^{n-1} \binom{n-1}{j} x^{(1-x)}^{n-j} \cdot b(j+1) - c
$$

$$
\pi_D(x) = \sum_{j=0}^{n-1} \binom{n-1}{j} x^{(1-x)}^{n-j} \cdot b(j)
$$

where $0 \leq x \leq 1$ is the fraction of producers in the population, as a producer pays a cost $c$ that a non-producer does not pay, but its group has one more contributor (itself).

In a clonal population, the replicator dynamics (Hofbauer and Sigmund, 1998) of this game is given by

$$
\dot{x} = x(1-x) \cdot [\beta(x) - c]
$$

where the fitness difference $\pi_C(x) - \pi_D(x)$ is written in the form $\beta(x) - c$, and

$$
\beta(x) = \sum_{j=0}^{n-1} \binom{n-1}{j} x^{(1-x)}^{n-j} \cdot \Delta b_j
$$

The dynamics (1) has two trivial rest points $x = 0$ and $x = 1$; further possible, interior rest points are given by the roots of the equations

$$
\beta(x) - c = 0
$$

For the logistic function (or any other sigmoid function), an exact analytical solution is not possible. As $\beta(x)$ is a polynomial in Bernstein form, we can resort to the properties of Bernstein polynomials (Lorentz, 1953; Philips, 2003) to characterise the dynamics and find an approximate solution.

The benefit for having a fraction $x$ of producer cells is $b(x) = 1/(1 + e^x(1-x))$, the extension of $b(n)$ to all $x \in [0,1]$, with $h = k/n$. As $\beta(x)$ is a Bernstein polynomial (Bernstein, 1912; Lorentz, 1953; Philips, 2003) of the coefficient $\Delta b = b(j + 1)/n - b(j/n)$, by Bernstein theorem (Bernstein, 1912) we know that $\beta(x)$ converges uniformly to $\Delta b$, in $[0,1]$. Furthermore, because $\Delta b$ is the forward difference of the benefit function with spacing $1/n$, for large enough $n$ we can approximate $\Delta b \approx (1/n)b'(j/n)$. For any $x$, $j/n$ converges in probability to $x$ as $n \to \infty$; therefore, by Bernstein theorem, $\beta(x)$ converges to $(1/n)b'(x)$, and equation (3) can be approximated by

$$
(1/n)b'(x) - c = 0
$$

We can obtain an analogous solution using equation (4) for the normalised benefit function $b_n(x)$, and actually for any sigmoid function.

Because of the variation-diminishing property of Bernstein polynomials, we also know that the number of internal equilibria of $\beta$ is less than the number of sign changes of $\Delta b$ by an even amount; because of our assumption of sigmoid benefits, we know that $\beta(x)$ has a unique maximiser $x^*$ in $(0,1)$; finally, because of the end point values property of Bernstein polynomials, we know that $\beta(0) = \Delta b_0$ and $\beta(1) = \Delta b_{n-1}$.
RESULTS

It follows that \( x = 0 \) is a stable rest point of (1) if and only if \( \Delta b_0 < c \), and that \( x = 1 \) is a stable rest point of (1) if and only if \( \Delta b_n \geq c \). In addition, any interior stable rest point \( x_s \) must satisfy equation (3) and \( \beta'(x_s) < 0 \). It follows that there is at most one interior stable rest point \( x_s \) and that, if such a rest point exists, it satisfies \( x^* < x_s < 1 \). These conclusions define, for any sigmoid benefit function, the following five types of dynamics (Figure 1), where we simplify notation by defining \( \beta^* = \beta(x^*) \):

- If \( c > \beta^* \), then \( \beta(x) < c \ \forall x \), and \( x = 0 \) is the only stable equilibrium.
- If \( c = \min(\Delta b_0, \Delta b_{n-1}) \), then \( \beta(x) > c \ \forall x \), and \( x = 1 \) is the only stable equilibrium.
- If \( \min(\Delta b_0, \Delta b_{n-1}) < c < \max(\Delta b_0, \Delta b_{n-1}) \), then \( \beta(x) > c \) for \( x > x_u \) and \( \beta(x) < c \) for \( x < x_u \); therefore, the

![Figure 1. The five types of dynamics of growth factor production.](image-url)
unique interior unstable equilibrium \(x_u\) divides the basin of attraction of the two stable equilibria \(x = 1\) and \(x = 0\).

- If \(\min(\Delta h_0, \Delta h_{n-1}) < c < \max(\Delta h_0, \Delta h_{n-1})\), and \(\Delta h_u > \Delta h_{n-1}\), then \(\beta(x) > c\) for \(x < x_u\) and \(\beta(x) < c\) for \(x > x_u\); therefore, the unique interior stable equilibrium \(x_u\) divides the basin of attraction of the two unstable equilibria \(x = 1\) and \(x = 0\).

- If \(\max(\Delta h_0, \Delta h_{n-1}) < c < \beta^*\), then \(\beta(x) > c\) for \(x < x_u\) and \(\beta(x) < c\) for \(x > x_u\); therefore, the interior unstable equilibrium \(x_u\) divides the basins of attraction of the two stable equilibria \(x = 0\) and \(x = x_u\).

The internal equilibria for the normalised logistic function are given by (from equation (4))

\[
x_{\pm} = h \cdot \frac{s}{n} \log \left[ \frac{1 \pm \sqrt{1 - 4 \epsilon s \cdot B}}{2 \epsilon s \cdot B} - 1 \right]
\]

with \(x_+ = x_{\pm}\) for the stable equilibrium and \(x_u = x_{\pm}\) for the unstable equilibrium, where \(B = b(1) - b(0)\).

The results described so far reveal that stable heterogeneity can arise simply as a consequence of the fact that growth factors are diffusible public goods. Stable heterogeneity is more likely to occur for low levels of \(h\) (that is, when few producer cells are enough to confer a benefit to the whole group; Figure 2).

To conclude the comparative statics of the system, we note that (Figure 3) the equilibrium frequency of producers is always a decreasing function of \(c\) and \(n\) (the number of cells within the diffusion range of the growth factor); the effect of \(s\) (the steepness of the benefit function) is more complex because the equilibrium can be a monotonic or a single-peaked function of \(s\) depending on the other parameters (it is clear, however, that stable heterogeneity is possible only if \(s\) is not too low); finally, the equilibrium frequency of producers is an increasing function of \(h\) (the inflection of the benefit function). This has implications for the dynamics of anticancer therapies that target growth factors and for the evolution of resistance against such therapies.

An anticancer drug that acts by impairing growth factors will increase the amount of growth factors that the cells must produce to achieve a certain benefit, that is, it will increase \(h\). If the threshold is high, producers can go to fixation, whereas they remain at a mixed equilibrium if the threshold is lower (compare, for example, Figures 1B and D). Specifically, an anticancer drug that acts by reducing the amount of circulating growth factor may lead to the opposite of the desired effect: although the immediate effect is, of course, a sudden reduction in tumour growth (because there are not enough circulating growth factors), the amount of growth factors that must be produced to achieve the same benefit...
Figure 3. Effect of the parameters on the equilibrium fraction of producers. The fraction of producers is plotted over time for different values of $c$ (the cost of production), $n$ (the number of cells within the diffusion range of the growth factor), $h$ (the inflection of the benefit function) and $s$ (the steepness of the benefit function), starting from the same initial frequencies.

Figure 4. Evolution of resistance to anticancer treatments that target growth factors. A population reaches an equilibrium (after about 200 generations) in which producers and non-producers of growth factors coexist. When, at generation 1000, a drug is introduced that increases the threshold $h$ from 0.1 to 0.98 (i.e., it impairs almost completely the growth factor produced by the tumour), fitness immediately declines and remains low for a few generations; if the transition to the new dynamics is fast enough (20 generations), producers go extinct. If the transition is slower, however, (50 generations) the frequency of producers increases, and after a few generations fitness reaches levels comparable to the pre-drug treatment, as the fraction of producers approaches a new equilibrium. $n = 50; c = 0.01; s = 0.2$. 

(h) is now increased, which leads to an increase in frequency of the C cells, which eventually leads to a new equilibrium (Figure 4). In case of bistability, the final outcome will depend on the initial frequency of producers.

Even if a drug induces an almost complete annihilation of available growth factors, the speed of the transition from the original to the new threshold is essential for the success of the therapy. Although a fast transition to the new threshold can lead to a successful, stable therapy, a slower delivery of the anti-growth factor drug, or inefficient RNA interference (RNAi) delivery systems that require multiple treatments over an extended period of time, can make the system evolve back to relapse, even if the treatment itself is extremely efficient at impairing the growth factor (Figure 5A). Even therapies that are almost immediately effective will fail if the reduction in circulating growth factor is not large enough (Figure 5B).

CONCLUSION

Analysing the production of growth factors as a nonlinear PGG reveals that tumour heterogeneity can be maintained by the frequency-dependent selection that arises as a natural consequence of the fact that growth factors are diffusible, and therefore public goods, with no need to invoke neutral mutations (Iwasa and Michor, 2011), niche specialisation (Nagy, 2004; Gatenby and Gillies, 2008) or non-equilibrium (Gonzalez-Garcia et al, 2002). Five types of dynamics are possible, including the coexistence of producers and non-producers at a stable equilibrium. This diversity of dynamics was not captured by previous game theory models based on pairwise interactions or by previous models of public goods in evolutionary game theory.

Tumour heterogeneity has important implications for diagnosis and treatment. The results help us understand anticancer therapies that attack growth factors, either directly (using drugs like avastin that target the growth factors) or indirectly (using RNAi). Although it was suggested that attacking growth factors may be less susceptible to the evolution of resistance (Pepper, 2012; Aktipis and Nesse, 2013), the results shown here suggest that the issue is not so simple. In fact, it is clear that, in spite of the initial enthusiasm, anti-angiogenic drugs, one of the most prominent examples of therapies based on the disruption of growth factors, have been disappointing. A drug like Avastin, which was the best selling drug for Roche (Basel, Switzerland) in the past few years, only extend overall survival of patients with certain types of cancer by a few months, which are followed by relapse (Amit et al, 2013).

The rationale of the analysis is that when one reduces the amount of a growth factor, the immediate result is a sudden reduction in tumour growth, because the threshold necessary to achieve the original benefit is not reached; as a consequence, the growth rate of the tumour immediately declines. At the same time, however, the amount of growth factors necessary for the population to grow increases (because part of them are disrupted by the therapy), which changes the dynamics of the system; unfortunately, it changes in the wrong direction: by increasing the threshold, one increases the frequency of producers at equilibrium, which explains relapse simply as the new equilibrium reached by the system under the new conditions. Analysing anti-growth factor drugs as methods that increase the threshold of a PGG reveals that the initial expectations of stability for these therapies were based on faulty evolutionary predictions. Although it is too early to evaluate the efficacy of RNAi treatments, it seems reasonable that even silencing the gene for a growth factor should incur a similar problem and be susceptible to the evolution of resistance. As pointed out by Pepper (2012), therapies that target growth factors are certainly a more evolutionary robust approach than conventional drugs that target cells directly. As we have shown, however, an anti-growth factor drug must be evolutionarily stable in order to be effective. Besides efficacy, latency was also shown here to be fundamental for the success of such therapies.

In order to have quantitative results that may help design proper therapies, the methods used here should be extended to take into account more realistic features of cancer development, including, for example, spatial structure, as cells will form clusters of producers and non-producers with their own dynamics. It is known, however, that assortment in spatially structured
populations does not change the dynamics qualitatively if benefits are sigmoid (which is our case; Archetti and Scheuring, 2012; Perc et al., 2013). Drugs that target growth factors may be used in combination with other therapies, such as chemotherapy. As chemotherapy is more effective against rapidly dividing cells, it will affect preferentially non-producing cells (which grow faster), and therefore confer a benefit to producer cells, which would not help the treatment. An evolutionarily stable therapy should modify the dynamics to lead to the extinction of the producer cells. As we have seen, extinction of the producer cells is possible, depending on the shape of the benefit function, if the cost/benefit ratio of producing the growth factor is high enough, group size is large enough and if the initial amount of non-producer cells is above a critical threshold. The cost/benefit ratio could be changed by varying the amount of exogenous growth factor; group size could be changed by varying the diffusion range of the factor; the critical amount of non-producer cells can be achieved by autologous cell therapy, using cancer cells collected from the patient, in which genes coding for growth factors have been knocked out. Such therapy would be using cancer cells collected from the patient, in which genes coding for growth factors have been knocked out. Such therapy would be

Arkerti M (2009b) The volunteer's dilemma and the optimal size of a social group. J Theor Biol 261: 475–480.

Arkerti M, Scheuring I (2011) Coexistence of cooperation and defection in public goods games. Evolution 65: 1140–1148.

Arkerti M, Scheuring I (2012) Review: game theory of public goods in one-shot social dilemmas without assortment. J Theor Biol 299: 9–20.

Axelrod R, Axelrod De, Pienta KJ (2006) Evolution of cooperation among tumor cells. Proc Natl Acad Sci USA 103: 13474–13479.

Bach LA, Sumpter DJT, Alnser J, Loeschke V (2003) Spatial evolutionary games of interaction among generic cancer cells. J Theor Med 5: 47–58.

Basanta D, Deutsch A (2008) A Game Theoretical Perspective On The Somatic Evolution Of Cancer. In: Bellomo N, Chaplain M, De Angelis E (eds), Selected Topics On Cancer Modelling: Genesis Evolution Immune Competition Therapy. Birkhauser: Boston, MA, USA.

Basanta D, Hataikiriou H, Deutsch A (2008a) Studying the emergence of invasiveness in tumors using game theory, Eur Phys J 63: 393–397.

Basanta D, Scott JG, Fishman MN, Ayala G, Hayward SW, Anderson AR (2012) Investigating prostate cancer tumour-stroma interactions: clinical and biological insights from an evolutionary game. Br J Cancer 106: 174–181.

Basanta D, Scott JG, Rockne R, Swanson KR, Anderson AR (2011) The role of IDH1 mutated tumour cells in secondary glioblastomas: an evolutionary game theoretical review. Phys Biol 8: 015016.

Basanta D, Simon M, Hatzikirou H, Deutsch A (2008b) Evolutionary game theory elucidates the role of glycolysis in glioma progression and invasion. Cell Pro lif 41: 980–987.

Bernstein S (1912) Démonstration du théorème de Weierstrass fondée sur le calcul des probabilités. Comm Soc Math Kharkov 13: 1–2.

Cairns J (1975) Mutation selection and the natural history of cancer. Nature 255: 197–200.

Crespi B, Summers K (2005) Evolutionary biology of cancer. Trends Ecol Evol 20: 545–552.

Dexter DL, Leith JT (1986) Tumor heterogeneity and drug resistance. J Clin Oncol 4: 244–257.

Dingli D, Chalub FA, Santos FC, Van Segbroeck S, Pacheco JM (2009) Cancer phenotype as the outcome of an evolutionary game between normal and malignant cells. Br J Cancer 101: 1130–1136.

Eungdamrong NJ, Iyengar R (2004) Modeling cell signalling networks. Biol Cell 96: 355–362.

Gatenby R, Gillies RJ (2008) A microenvironmental model of carcinogenesis. Nat Rev Cancer 8: 56–61.

Gatenby RA, Maini P (2003) Cancer summed up. Nature 421: 321.

Gerstung M, Nakhlou H, Beerenwinkel N (2011) Evolutionary games with affine fitness functions: applications to cancer. Dynanm Games Appl 1: 370–385.

Gonzalez-Garcia I, Sole RV, Costa J (2002) Metapopulation dynamics and spatial heterogeneity in cancer. Proc Natl Acad Sci USA 99: 13085–13089.

Greaves M, Maley CC (2012) Clonal evolution in cancer. Nature 481: 306–313.

Hanahan D, Weinberg RA (2000) The hallmarks of cancer. Cell 100: 57–70.

Hemker HC, Hemker PW (1969) General kinetics of enzyme cascades. Proc R Soc Lond Ser B 173: 411–420.

Hofbauer J, Sigmund K (1998) Evolutionary Games and Population Dynamics. Cambridge University Press: Cambridge, UK.

Iwasa Y, Michor F (2011) Evolutionary dynamics of intratumor heterogeneity. PLoS One 6: e17866.

Jouanneau J, Moens G, Bourgeois Y, Poupon MF, Thiery JP (1994) A minority of carcinoma cells producing acidic fibroblast growth factor induces a community effect for tumor progression. Proc Natl Acad Sci USA 91: 286–290.

Lambert C, Estévez-Salmerón L, Oh S, Liao D, Emerson BM, Tisty TD, Austin RH (2011) An analogy between the evolution of drug resistance in bacterial communities and malignant tissues. Nat Rev Cancer 11: 375–382.

Lorentz GG (1953) Bernoulli Polynomials. University of Toronto Press: Toronto, Canada.

Maley CC, Galipeau PC, Finley JC, Wongsurawat VJ, Li X, Sanchez CA, Paulson TG, Blount PL, Risques RA, Rabinovitch PS, Reid BJ (2006) Genetic clonal diversity predicts progression to esophageal adenocarcinoma. Nat Genet 38: 468–473.

Marusyk A, Polyak K (2010) Tumor heterogeneity: causes and consequences. Biochem Biophys Acta 1805: 105–117.

Mendes P (1997) Biochemistry by numbers: simulation of biochemical pathways with Gepasi 3. Trends Biochem Sci 22: 361–363.

Merlo LMF, Pepper JW, Reid BJ, Maley CC (2006) Cancer as an evolutionary and ecological process. Nat Rev Cancer 6: 924–935.

Nagy JD (2004) Competition and natural selection in a mathematical model of cancer. Bull Math Biol 66: 663–687.

Nowell PC (1976) The clonal evolution of tumour cells. Science 194: 23–28.

Pepper JW (2012) Drugs that target pathogen public goods are robust against evolved drug resistance. Evolut Appl 5(7): 757–761.

Perc M, Gómez-Gardenés J, Szolnoki A, Moreno Y (2013) Evolutionary dynamics of group interactions on structured populations: a review. J R Soc Interface 10: 20120997.

Phillips GM (2003) Interpolation and Approximation by Polynomials. Chapter 7. Springer: Berlin, Germany.

Ricard J, Noat G (1986) Catalytic efficiency, kinetic co-operativity of ribulose 1,5-bisphosphate carboxylase/oxygenase. Eur J Biochem 160: 47–58.

Ricard J, Noat G (1986) Catalytic efficiency, kinetic co-operativity of ribulose 1,5-bisphosphate carboxylase/oxygenase. Eur J Biochem 160: 47–58.

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