Impact of iron deposit on the accuracy of quantifying liver fat fraction using multi-material decomposition algorithm in dual-energy spectral computed tomography

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Abstract

Objectives: To investigate the accuracy of using multi-material decomposition (MMD) algorithm in dual-energy spectral computed tomography (CT) for quantifying fat fraction (FF) in the presence of iron.

Materials: Nine tubes with various proportions of fat and iron were prepared. FF were divided into three levels (10%, 20%, and 30%), recorded as references (FFref). Iron concentrations (in mg/100 g) were divided into three ranges (25.25–25.97, 50.38–51.55 and 75.57–77.72). The nine-tube phantom underwent dual-energy CT and MR. CT attenuation was measured and FF were determined using MMD in CT (FFCT) and Iterative Decomposition of water and fat with Echo Asymmetry and Least squares estimation (IDEAL-IQ) in MR (FFMR) for each tube. Statistical analyses used were: Spearman rank correlation for correlations between FFref and CT attenuation, FFCT, and FFMR; one-way ANOVA, and one-sample t-test for the differences between FFCT and FFref and between FFMR and FFref. A multivariate linear regression model was established to analyze the differences between the corresponding values with different iron concentrations under the same FFref.

Results: Fat fraction on CT (FFCT) and FFMR were positively correlated with FFref (all p < 0.001), while the CT attenuation was negatively correlated with FFref in the three iron concentration ranges. For a given FFref, FFCT decreased and FFMR increased as the iron concentration increased. The mean difference between FFCT and FFref over the nine tube measurements was 0.25 ± 2.45%, 5.7% lower than the 5.98 ± 3.33% value between FFMR and FFref (F = 310.017, p < 0.01).

Conclusion: The phantom results indicate that MMD in dual-energy CT can directly quantify volumetric FF and is less affected by iron concentration than MR IDEAL-IQ method.

Keywords: dual-energy CT, fat quantification, IDEAL-IQ, iron deposit, MRI, multi-material decomposition
Nonalcoholic fatty liver disease (NAFLD) is becoming the most common chronic liver pathology both in Chinese and Western countries. About 30% of the population is at risk, fatty liver is often associated with iron metabolism disorders, of which up to 30% of patients have hyperseroferritinemia and liver iron deposition, which is called dysmetabolic iron overload syndrome. Therefore, it is important to evaluate the severity of hepatic steatosis with iron overload. Liver biopsy has remained the “gold standard” mainly for assessment of liver fat content. However, the liver biopsy sample only accounts for 0.002% of the entire liver volume, so there is sampling errors in this method, and liver biopsy is an invasive surgery with complication rates of about 0.6%–18%.

Noninvasive quantitative methods for expressing liver fat concentration, therefore, have attracted more and more clinical attention, including those related to ultrasound (US), magnetic resonance (MR), and computed tomography (CT). US is widely used for the assessment of hepatic steatosis; however, it is relatively insensitive and nonspecific for the detection of individuals with mild steatosis. To our knowledge, there is no established quantitative US for assessing fatty infiltration of the liver. At present, there are two kinds of MR sequences used for fat quantification: magnetic resonance spectroscopy (MRS) and Iterative Decomposition of water and fat with Echo Asymmetry and Least Squares estimation (IDEAL-IQ), which have been verified in fat–water phantoms and animal models. However, iron deposition in the liver can change the uniformity of magnetic field, resulting in inaccurate measurement.

In the past, attenuation values derived from the single-energy CT is used to estimate density and is semi-quantitative for measuring liver fat content. However, due to the scanning parameters and the presence of iron, glycogen, drugs, or other substances, the measured fat content varies from patient to patient. In addition, the beam-hardening artifact has been pointed out as a cause for the attenuation value to drift, which further affects the semi-quantitative results. And researchers believe that the CT attenuation values are not sensitive to mild hepatic steatosis. On the other hand, the monochromatic images in dual-energy spectral CT can correct the beam-hardening effect, making the attenuation value more accurate and less affected by the shapes of the objects and the measurement locations in the objects than the polychromatic images. Multi-material decomposition (MMD) is a new technology in dual-energy spectral CT imaging that can distinguish two or more different substances at the same time and quantitatively and intuitively evaluate liver fat content in volume to generate volume fat fraction (FF).

The purpose of our study was to validate the accuracy of using MMD algorithm in dual-energy spectral CT for measuring FF using a fatty liver phantom with different FFs and iron concentrations, and to study the effect of iron on the measured FF accuracy, and to compare them with MR IDEAL-IQ measurements against the ground truth.

1 | MATERIALS

1.1 | Liver phantom preparation

Fresh swine liver was purchased from market and MR IDEAL-IQ scan was performed to confirm that the fat concentration in the liver was less than 2%. The blood vessels and bile duct tissue were first removed from the swine liver before it was grinded and filtered. The liver was then deaerated with a vacuum deaerator for 30 min at room temperature. Nine mixtures that contained different proportions (Table 1) of the swine liver, pure lard, iron (Iron sucrose Injection, Nanjing Hengsheng Pharmaceutical Co., Ltd; 100 mg of iron per 5 ml ampule), and distilled water were prepared and again deaerated until air bubbles were

| Tube no. | Swine liver (ml) | Lard (ml) | Irona (ml) | Distilled water (ml) | FFref (%) | Iron content (mg/100 g) |
|----------|-----------------|----------|------------|----------------------|-----------|------------------------|
| 1        | 30              | 4        | 0.5        | 5.5                  | 10        | 25.25                  |
| 2        | 30              | 4        | 1.0        | 5.0                  | 10        | 50.38                  |
| 3        | 30              | 4        | 1.5        | 4.5                  | 10        | 75.57                  |
| 4        | 26              | 8        | 0.5        | 5.5                  | 20        | 25.58                  |
| 5        | 26              | 8        | 1.0        | 5.0                  | 20        | 51.81                  |
| 6        | 26              | 8        | 1.5        | 4.5                  | 20        | 78.74                  |
| 7        | 22              | 12       | 0.5        | 5.5                  | 30        | 25.97                  |
| 8        | 22              | 12       | 1.0        | 5.0                  | 30        | 51.55                  |
| 9        | 22              | 12       | 1.5        | 4.5                  | 30        | 77.72                  |

Abbreviation: FF, fat fraction.

Iron sucrose injection (20 mg of iron per milliliter).
undetectable for 30 min. These mixtures were put in nine tubes and placed into a polypolypropylene phantom of 20 cm diameter (QSP-1; Fuyo Corporation). Based on the literatures, the nonalcoholic liver steatosis is graded in the following way: grade 0, 0%–6.4% (normal); grade 1, 6.5%–17.4% (mild); grade 2, 17.5%–22.1% (medium); and grade 3, 22.2% or higher (severe). Therefore, in our study the nine tubes were prepared to represent the three diseased levels at 10%, 20% and 30% and recorded as reference levels for the fat fraction (FFref). In each of the three tubes with same FF level, 0.5, 1.0, and 1.5 ml of iron sucrose were added by injection. The nine mixtures were weighed, and the iron concentration was calculated to be at roughly three levels (25.25–25.97, 50.38–51.81, and 75.57–77.72 mg/100 g). Iron content was reported to be less than 40 mg per 100 g of liver (wet weight) in healthy individuals. In our experiment, the molecular formula of the soluble sucrose iron added was C6H8FeO8 (Fe+2), simulating the relatively normal and high level of iron content in the liver. See Table 1.

1.2 CT acquisition and data analysis

The QSP-1 phantom that contained the nine tubes was scanned on a 256-section multidetector CT scanner (Revolution CT; GE Healthcare) using the dual-energy CT imaging mode. The scan parameters were as follows: tube voltage 80/140 kVp instantaneous switching; Tube current: 200 mA; Noise index: 6; gantry rotation speed: 0.5 r/s. Scan was repeated three times and the CT dose index value for each scan was 2.99 mGy. Images were reconstructed at layer thickness/spacing of 5.0 mm/1.25 mm using a new version adaptive statistical iterative reconstruction (ASIR-V) with 40% strength. The 120 kVp-like CT images were generated to measure CT attenuation in Hounsfield units (HU). The images were transmitted to an advanced workstation (AW Volumeshare 7; GE Healthcare) which was equipped with a currently not-yet-commercially available MMD software (GE Healthcare) to measure the FFCT (fat fraction on CT MMD image; Figure 1a). The MMD image is obtained from two sets of measurements associated with low and high energies from dual-energy spectral CT measurements. This method assumes three basis materials at the most within each pixel, and the material types alter among the pixels. It uses volume conservation to obtain three materials for reconstruction and decomposition to obtain live fat images for quantifying fat fraction.

Immediately after the CT scan, MR IDEAL-IQ imaging were performed on the nine tubes placed on the plate by using a 3.0 T MR clinical imager (Discovery 750 W; GE Healthcare) with a 24-channel head–neck combined coil. The IDEAL-IQ sequence had the following acquisition parameters: scan plane: Axial, TR: 7.4 ms, TE:1–5.3 ms, number of echoes 3, display field-of-view: 40 cm, matrix size: 160 mm×160 mm; flip angle 4°; Slice Thickness: 3 mm; space between slices 0 mm. Scan was repeated three times. The MR fat fraction (FFMR) maps were generated by post-processing software provided by the manufacturer (Figure 1b).

Two abdominal radiologists (more than 5 years of abdominal CT and MRI diagnosis experience) performed the measurement independently. CT and MR measurements were performed by placing a region of interest (ROI, 140 mm², 3 mm deviation) in the center of each tube over three consecutive image slices. The mean and SD of the measured values of each tube were calculated by averaging measurements over the three image slices and by the two radiologists (see Table 2).

**FIGURE 1** The exact values in nine ROI tubes by CT and MR fat fraction images. (a) Fat fraction image from MMD; (b) Fat fraction image from IDEAL-IQ. The numbers correspond to the nine tubes shown in Tables 1 and 2. IDEAL-IQ, Iterative Decomposition of water and fat with Echo Asymmetry and Least squares estimation; MMD, multi-material decomposition; MR, magnetic resonance; ROI, region of interest.
1.3 Statistical analysis

The original data were verified and analyzed with SPSS 17.0 statistical software. Spearman rank correlation was used to analyze the correlation between FF\text{ref} and corresponding CT measurements (dual-energy CT FF\text{CT} and CT value) and MR measurements (IDEAL-IQ FF\text{MR}). One-way ANOVA and one-sample t-test were used to compare the measurement errors from the gold standard (FF\text{ref}) between the dual-energy CT FF\text{CT} and IDEAL-IQ FF\text{MR}. A multivariate linear regression model was established to analyze the differences between the corresponding values with different iron concentrations under the same FF; setting test Level $\alpha = 0.05$.

2 RESULTS

Figure 2a shows the change of CT attenuation value (on a 120 kVp-like image) as a function of FF\text{ref}, affected by the three levels of iron concentration (25.25–25.97,
50.38–51.55, and 75.57–77.72 mg/100 g). The line charts showed that CT attenuation value was negatively correlated with FFref. Figure 2b–d show the scatterplots of the measured values of FFCT on MMD and FFMR on MR IDEAL-IQ as function of FFref, affected by the three levels of iron concentration. The three groups all showed good correlations: FFref positively correlated with dual-energy CT FFCT (\(\rho = 0.943\), \(\rho = 0.906\) and \(\rho = 0.943\)) and MR IDEAL-IQ FFMR (\(\rho = 0.906\), \(\rho = 0.943\) and \(\rho = 0.921\)) for the three iron concentration levels, respectively (all \(p < 0.001\)). Figure 3 shows the changes of FFCT and FFMR measurement values under different iron concentrations indicated that FFCT decreased and FFMR increased with the increase of iron concentration under the same FFref value.

The mean difference over the nine tube measurements between FFCT and FFref (named group A) was 0.25 ± 2.45%, and the difference between the FFMR and FFref (named group B) was 5.98 ± 3.33%. A one-way ANOVA was used to statistically evaluate the differences, and the results suggested that the difference between group A and group B was statistically significant (\(F = 310.017, p < 0.01\)). A one-sample t-test was performed on the overall sample: there was no statistical difference between group A and FFref with a mean of 0 (\(t = 1.313, p = 0.191\)), while group B was statistically different from FFref with a mean of 0 (\(t = 22.833, p < 0.01\)), indicating that overall the measured value from CT MMD was more consistent with FFref and the measured value from MR IDEAL-IQ had systematic positive bias compared with FFref.

Using the FF measurement results from CT MMD and MR IDEAL-IQ scan methods as the dependent variable, the actual FF, iron solution concentration, and scan method were used to fit the multiple linear regression model for the independent variables. The specific variable assignments are given in Table 3. An entrance tolerance of 0.05, and an exit tolerance of 0.10. The regression analysis indicated that the regression equation had a good fitting degree (\(F = 1251.640, p < 0.01\)) and multiple regression analysis with a determination coefficient of \(R^2 = 0.922\). The results are given in Table 3. According to the nonstandardized regression coefficients of the measurement methods, under the condition that other variables were fixed, the mean value of the MR IDEAL-IQ FF measurement was 5.7% higher than the mean value of the CT MMD measurement (\(p < 0.01\)). Specifically, there was underestimation of the FF by MMD measurement in dual-energy CT (FFCT) for the low FF setting, and overestimation by MRIDEAL-IQ (FFMR) at all three FF settings, which further illustrated the statistical difference between the FF measurement values of the two scanning methods (\(p < 0.01\)). The results also showed that there was no statistical difference in the effect of iron content on the measurement results of the two groups (\(p = 0.186\)).

3 | DISCUSSION

In principle, dual-energy CT can only accurately reconstruct two materials with different attenuation
coefficients when performing scanning measurements under high and low-energy fast switching conditions. In contrast-enhanced scans, liver fat quantification requires the differentiation of four materials: liver tissue, blood, fat, and contrast agent. This results in limited clinical use of current material decomposition techniques. MMD algorithm is a flexible model-based method developed for dual-energy CT, which can expand the core material discrimination ability of dual-energy CT and discriminate among two or more different materials at the same time. MMD algorithm has two main clinical applications for contrast-free dual-energy CT data or contrast-enhanced dual-energy CT data, the first is virtual un-enhancement, the second is liver-fat quantification. The algorithm used in our phantom study was for LFQ, which is performed with fat and healthy liver tissue in the material basis, and uses volume conservation to obtain three materials for reconstruction and decomposition for fat quantification, and obtains live fat images for quantifying FF. The second generation of dual-energy spectral CT (GE revolution CT) was used in our phantom study, which is superior to the previous generation (GE discovery CT). It adopts more advanced ASIR-V, and further optimizes the performance of multi-material qualification, quantification, and noise reduction.

In our study, the mixture of swine liver and fat was used to simulate the state of human fatty liver. In preparing the liver phantom, the blood vessels and bile duct tissue were first removed from the swine liver before it was ground and filtered, and then deaerated with a vacuum deaerator. As swine liver tissue contains more bile duct and blood vessel components compared with human liver tissue, the CT attenuation of swine liver is higher than that of human liver tissue and the removal of bile duct and blood vessel would make the CT attenuation in the phantom closer to that of human liver tissue. It is worth noting that gas will be mixed in the process of preparing the fatty liver phantoms, which will also affect the measurement values. Therefore, these preparations are the key steps to prepare a qualified fatty liver phantom, so as to make the measurement results more accurate.

At present, there are not many studies on the effect of liver iron deposit on the quantitative measurement of fat in nonalcoholic fatty liver. Eskreis-Winkler et al. verified the accuracy of IDEAL-IQ in 10 cancer patients with both elevated liver fat and elevated liver iron, and found that the measurement value of IDEAL-IQ fat content was more accurate than that of IOP (T1 in and out of phase). Furthermore, Catherine et al. verified the accuracy of MRS on fat-fraction quantitative in the mice model of steatosis and iron overload, and found that there was an excellent correlation between MRS fat-fraction and group based fat quantitative.

Our phantom results on the dual-energy CT MMD measurement value \( FF_{CT} \), 120kvp-like CT measurement value \( FF_{CT} \), and MRI IDEAL-IQ measurement value \( FF_{MR} \) demonstrated a strong correlation with \( FF_{ref} \) in the presence of different iron concentrations. However, for CT examination, the advantage of the MMD method over CT attenuation is the ability of the former to intuitively measure fat content in units of volume percentage. Our study results validated that the agreement in the measurement value of dual-energy CT MMD fat content was better than that of MRI IDEAL-IQ in the presence of iron. One previous phantom study by Tomoko Hyodo et al. assessed the influence of iron on fat measurements determined with Dual-energy CT (DECT) and MRS by using mixtures with iron concentration at three levels (0, 48.1–55.9, and 92.6–103.0 mg/100 g), and found that iron concentration had less influence on the measurement of FF by CT MMD, but led to the underestimation and overestimation of fat content measurements with MRS. Our phantom study also showed that the average value of the \( FF_{MR} \) was higher than \( FF_{CT} \), specifically in the presence of iron. \( FF_{CT} \) was underestimated in low fat concentrations and \( FF_{MR} \) was overestimated in low, medium, and high fat concentrations. Therefore, the MMD algorithm may produce fewer errors in patients with fatty liver and high iron content. For example, patients with chronic liver disease with excessive iron may get more accurate results with dual-energy CT. The range of iron concentration in our study was 25.25–77.72 mg/100 g, which cannot represent the concentration range of iron deposition for all diseases, such as genetic hemochromatosis (more than 100 mg iron per 100 g liver [wet weight]), etc, which needs further verification.

An unexpected result was that under the same \( FF_{ref} \) condition, with the increasing amount of iron the \( FF_{CT} \) measurement value gradually decreased, and the \( FF_{MR} \) measurement value gradually increased, indicating that the changes in iron concentration had a decreasing and increasing trend on CT and MR measurements. The results of multiple linear regression model analysis showed that iron concentration had similar impact of the measurement errors for CT MMD and MR IDEAL-IQ measurements regardless of the amount of fat content \( (p = 0.186) \). This indicated that both the MMD algorithm and IDEAL-IQ had good correction for the influence of iron.

There are a number of limitations in this study: (1) We did not make precise chemical extraction of the fat concentration and iron concentration in the fatty liver model; (2) The model did not cover the cases of clinically high-concentration liver iron; (3) A single DECT protocol was used in our study and the effect of altering CT acquisition and reconstruction parameters was not studied; (4) In order to avoid the interference of the phantom on the magnetic field stability during MR scanning, the tubes were placed in the air which may have some impact on the measurement results.
In summary, our phantom results demonstrate a strong correlation and consistency between the measurement results of MMD and the actual FF, and indicate that DECT based MMD method is promising for noninvasive quantification of hepatic steatosis with lower susceptibility to iron, which is of great importance for the diagnosis, characterization, and treatment of fatty liver disease, and has broad prospects for clinical application.

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AUTHORS’ CONTRIBUTIONS
D.D., X.W., J.W. and H.C. purchased experimental materials, prepared liver phantom and measured experimental data, and D.D. drafted the manuscript. J.S. scanned the phantom. B.L. participated in its design and coordination.

DATA AVAILABILITY STATEMENT
The datasets generated during and analyzed during the current study are available from the corresponding author on reasonable request.

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REFERENCES
1. Hentze MW, Muckenthaler MU, Galy B, Camaschella C. Two to Tango: regulation of mammalian iron metabolism. Cell. 2010;142(1):24-38.
2. Ratziu V, Bellentani S, Cortez-Pinto H, Day C, Marchesini G. A position statement on NAFLD/NASH based on the EASL 2009 special conference. J Hepatol. 2010;53(2):372-384.
3. Loomba R, Sanyal AJ. The global NAFLD epidemic. Nat Rev Gastroenterol Hepatol. 2013;10(11):686-690.
4. Datz C, Muller E, Aigner E. Iron overload and non-alcoholic fatty liver disease. Minerva Endocrinol. 2017;42(2):173-183.
5. Sanai FM, Keeffe EB. Liver biopsy for histological assessment: the case against. Saudi J Gastroenterol. 2010;16(2):124-132.
6. Guo Z, Blake GM, Li K, et al. Liver fat content measurement with quantitative CT validated against MRI proton density fat fraction: a prospective study of 400 healthy volunteers. Radiology. 2020;294(1):89-97.
7. Kramer H, Pickhardt PJ, Kliwer MA, et al. Accuracy of liver fat quantification with advanced CT, MRI, and ultrasound techniques: prospective comparison with MR spectroscopy. Am J Roentgenol. 2017;208(1):92-100.
8. Chiang HJ, Lin LH, Li CW, et al. Magnetic resonance fat quantification in living donor liver transplantation. Transplant Proc. 2014;46(3):666-668.
9. Idilman IS, Tuzun A, Savas B, et al. Quantification of liver, pancreas, kidney, and vertebral body MRI-PDFF in non-alcoholic fatty liver disease. Abdom Imaging. 2015;40(6):1512-1519.
10. Roberts NT, Hernando D, Holmes JH, Wiens CN, Reeder SB. Noise properties of proton density fat fraction estimated using chemical shift-encoded MRI. Magn Reson Med. 2018;80(2):685-695.
11. Jin KH, Jung CH, Bohyun K, et al. Accuracy and precision of proton density fat fraction measurement across field strengths and scan intervals: a phantom and human study. JMRI. 2019;50(1):305-314.
12. Hui SCN, So HK, Chan DFY, et al. Validation of water-fat MRI and proton MRS in assessment of hepatic fat and the heterogeneous distribution of hepatic fat and iron in subjects with non-alcoholic fatty liver disease. Eur J Radiol. 2018;107(107):7-13.
13. Qayyum A. MR spectroscopy of the liver: principles and clinical applications. Radiographics. 2009;29(6):1653-1664.
14. Pickhardt PJ, Park SH, Hahn L, Lee SG, Bae KT, Yu ES. Specificity of unenhanced CT for non-invasive diagnosis of hepatic steatosis: implications for the investigation of the natural history of incidental steatosis. Eur Radiol. 2012;22(5):1075-1082.
15. Matsuda I, Akahane M, Sato J, et al. Precision of the measurement of CT numbers: comparison of dual-energy CT spectral imaging with fast kVp switching and conventional CT with phantoms. Jpn J Radiol. 2012;30(1):34-39.
16. He T, Qian X, Zhai R, Yang Z. Computed tomography number measurement consistency under different beam hardening conditions. J Comput Assist Tomogr. 2015;39(6):981-985.
17. Niu T, Dong X, Petrongolo M, Zhu L. Iterative image-domain decomposition for dual-energy CT. Med Phys. 2014;41(4):041901.
18. Fischer MA, Reiner CS, Raptis D, et al. Quantification of liver iron content with CT-added value of dual-energy. Eur Radiol. 2011;21(8):1727-1732.
19. Tang A, Desai A, Hamilton G, et al. Accuracy of MR imaging-estimated proton density fat fraction for classification of dichotomized histologic steatosis grades in nonalcoholic fatty liver disease. Radiology. 2015;274(2):416-425.
20. Szczepaniak LS, Nurenberg P, Leonard D, et al. Magnetic resonance spectroscopy to measure hepatic triglyceride content: prevalence of hepatic steatosis in the general population. Am J Physiol Endocrinol Metab. 2005;288(2):E462-E468.
21. Brittenham GM, Farrell DE, Harris JW, et al. Magnetic-susceptibility measurement of human iron stores. N Engl J Med. 1982;307(27):1671-1675.
22. Buzzetti E, Petta S, Manuguerria R, et al. Evaluating the association of serum ferritin and hepatic iron with disease severity in non-alcoholic fatty liver disease. Liver Int. 2019;39(7):1325-1334.
23. Mendonca PR, Lamb P, Sahani DVA. Flexible method for multimaterial decomposition of dual-energy CT images. IEEE Trans Med Imaging. 2014;33(1):99-116.
24. Eskreis-Winkler S, Corrias G, Monti S, et al. IDEAL-IQ in an oncologic population: meeting the challenge of concomitant liver fat and liver iron. Cancer Imaging. 2018;18(1):51.
25. Hines CD, Agni R, Roen C, et al. Validation of MRI biomarkers of hepatic steatosis in the presence of iron overload in the ob/ob mouse. J Magn Reson Imaging. 2012;35(4):844-851.

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