The extracellular matrix (ECM) is a complex structural and signaling extracellular entity that has multiple cell physiological and developmental functions [1,2]. These functions include:

A. Providing a structural scaffolding for the epithelium and other cellular tissues.

B. Providing a binding platform for growth factors (GFs) and other ligands that can affect cells that are in close proximity to the ECM or affect more distant cells once these GFs and ligands are released from the ECM.

C. Providing signaling motifs within each ECM component that trigger signaling pathways on those cells that are in contact with these motifs or these same peptide motifs that can be released from the ECM molecules via enzymatic reactions to subsequently affect cells.

D. ECM molecules are able to transfer signals to cells through specialized cell surface receptors, e.g. integrins, that specifically bind to motifs within the various ECM components so as to initiate complex signaling pathways for cell differentiation and cell function (ECM molecules also bind to non-integrin cell surface molecules such as syndecans and EGFR); and finally,

E. Matrix metalloproteases (MMPs) are functionally associated with the ECM in order that ECM components can be modified through hydrolysis. MMPs are involved in many ECM’s functions by acting directly on ECM components or by acting on ECM-associated molecules. These MMP-induced ECM modifications allow for altered matrix structure during morphogenesis and cellular/developmental processes [3]. MMPs also function to release ECM-associated molecules from the ECM to affect cell function.

Given the fundamental function of the ECM of metazoans, it is not surprising to find ECM components in ancient Taxa such as the Cnidaria that represents one of the earliest eukaryotic systems with a defined tissue organization [4]. Cnidarians are composed of an outer ectoderm and inner endoderm with an intervening ECM. Based on molecular studies focused on understanding the components and function of various macromolecules of the ECM in Hydra, we now have a basic understanding of the role of ECM in an early divergent metazoon that arose some 500 million years ago [5-7].

This review focuses on an important component of the ECM, namely laminin; the matrix molecule first cloned and characterized in the Cnidarian, Hydra. Its purpose is to show the highly conserved nature of ECM throughout the Metazoa by highlighting the structure/function relationships of an early laminin form. It does this by comparing this laminin to what we know of vertebrate laminins based on the full spectrum of Hydra laminins on current genomic and EST data bases.
Laminins of Hydra ECM: A Comparative Analysis of Hydra and Vertebrate Laminins
Based on Current Genomic and EST Databases

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Initial cloning and characterization of a Hydra laminin

From the standpoint of divergence of Taxa within the animal kingdom, Cnidarians are the earliest group of metazoans with a defined tissue organization (ectoderm and endoderm) and an ECM. A tree of Taxa divergence is shown in Figure 1. This tree of Taxa with a "Common Tool Kit" of ECM components indicates that Cnidaria, containing Hydra, arose some 500 million years ago, thereby representing one of the earliest metazoan life forms with an ECM [5].

The basic structure of Hydra's body wall is shown in Figure 2. As indicated, the entire body wall of Hydra (from the foot process, through the body column, and apically to the tip of head tentacles) is organized as an outer ectoderm and inner endoderm with an intervening ECM (Figure 2, left image). Based on molecular and cellular studies, Hydra’s ECM is itself organized as two basal lamina’s (adjacent to the two epithelial layers) with an intervening interstitial-like matrix composed of fibular collagens as shown in Figure 2, right image. Because of its simplified structure and high regenerative capacity, Hydra has been studied as a model developmental organism since the time of Trembley whose studies were published in 1744 [7].

Figure 1: A hieratical tree of metazoans with a "Common Tool Kit" of ECM components. Those Taxa with an asterisk have complete genome data bases available. Cnidaria divergence dates back approximately 500 million years ago.

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Laminins of Hydra ECM: A Comparative Analysis of Hydra and Vertebrate Laminins Based on Current Genomic and ETS DataBases

Hydra exists as a gastric tube with a mouth and several tentacles at the head pole and a peduncle and a basal disk at the foot pole (left diagram). The entire body wall of Hydra (from the tip of the tentacles to the basal disk) is organized as an epithelial bilayer with an intervening ECM along the entire longitudinal axis of the organism. Hydra's ECM is structured as two subepithelial zones (i.e. basal lamina matrix) with an intervening central fibrous zone (i.e. interstitial matrix). As shown in the composite diagram to the right that utilizes a transmission electron micrograph of Hydra ECM interposed between a drawing of the two cell layers (ectoderm and endoderm), Hydra laminin and Type IV Collagen are localized to the two subepithelial zones of the matrix while Hydra fibrillar collagens (e.g. Hcol-I) are localized to the central fibrous zone or interstitial matrix.

Using a combination of biochemical, cellular, and molecular approaches, studies were initiated in the 1980's to elucidate the structure and function of hydra ECM components [11]. The work involved:
A. Isolation of Hydra ECM and use of biochemical and immunological approaches to analyze the purified matrix preparation [12],
B. Use of the purified Hydra ECM preparation to biochemically analyze and to generate a battery of Hydra-specific polyclonal and monoclonal antibodies [13], and
C. Use of these Hydra specific antibodies as reagents to screen expression cDNA libraries and as probes to characterize the distribution of matrix components in Hydra ECM using morphological techniques [13]. In addition, Hydra-specific antibodies and isolated matrix component domains were also used as blocking reagents to study the role of cell-ECM interactions in Hydra using a number of regeneration bioassays [14]. These functional studies were later complimented with antisense RNA studies to selectively knockdown Hydra ECM components during regenerative processes [11].

While a number of Hydra ECM components were identified in this way, for the purposes of this review we will focus on those studies concerning Hydra laminin, the first molecule to be clearly identified and functionally characterized. Laminin chains were identified using antibodies generated to purified Hydra ECM. Initial antibody screening of expression cDNA libraries isolated clones matching the laminin α chain (likely α1) and β1 chain of vertebrates [13,15]. The complete ORF of the β1 chain was determined while only a partial sequence of the α chain (likely an α1 chain) was obtained. While Hydra laminin is localized to the two subepithelial zones (basal lamina) of Hydra ECM, it is synthesized exclusively by the endoderm, which means that the molecules have to diffuse through the mesoglea to reach the ectodermal layer [13,14]. The location at the basal region of both
cell layers (Subepithelial Zones of Hydra ECM as shown in Figure 2) suggests that laminin is required for proper cell function and differentiation. During Hydra ECM regeneration, laminin secretion from the endoderm precedes the secretion of Hydra collagen-I that arises from the ectoderm [16] and inhibition of laminin secretion through a RNA antisense technique will block collagen secretion [17]; indicating cross-talk between the two layers. Earlier studies have already established that antibodies to Hydra laminin will block Hydra morphogenesis [14] and other ECM-related processes such as cell migration [18]. mRNA expression studies found that laminin is up-regulated during regeneration and in situ expression studies found this up-regulation to be associated with the base of head and foot regeneration regions [17]. Potential cell surface binding domains were later identified in the laminin β1 chain along with cell binding proteins that had affinity for these binding domains [13,19]. As indicated, laminin expression was essential for Hydra morphogenesis and regeneration processes based on a number of functional studies [17].

These studies indicated that ECM components, such as laminin, formed early during the divergence of early metazoan groups (e.g. Cnidaria) and these molecules were essential for Hydra cell physiology and developmental/regenerative processes.

**Composition of vertebrate laminins**

Vertebrate laminins [1,2] are heterotrimeric cross-linked glycoproteins that bind to other matrix components in order to form the complete basement membrane network typically associated with epithelial cells. Laminins predominantly link with collagen IV, nidogens, agrin, and perlecan to form the ECM basement membrane (termed the basal laminin at the level of transmission electron microscopy) [20]. Laminins are composed of three chains, named the alpha (α), beta (β), and gamma (γ) chains, that are coded by separate genes. Genomic and biochemical studies with vertebrate systems have identified five α chains, three β chains, and three γ chains. These chains are assigned the symbols; LAMA1-5, LAMB1-3, and LAMC1-3. In vivo, we find 16 αβγ laminin trimers that exist in nature. The reduced number of laminin trimer chain combinations (as compared to the theoretical maximal number of combinations) is due to the biochemical properties of the chains and the distribution of laminins among the various tissues of the body. The most studied heterotrimeric laminin is composed of a α1, β1, and γ1 chain and is termed laminin-111. A diagram of laminin-111 is depicted in Figure 3.

As discussed, since the original identification of the first laminin in vertebrates, a total of 11 laminin chains have been identified and 6 of these chains have analogs in Hydra (see Section 3 and Table 1). The biochemistry of laminin chains has been well studied [1,2]. All laminin chains have common structural motifs that include:

A. Large globular laminin N-terminal domains (VI domains),
B. Rod-like stretches comprised of LE domains in all three types of chains,
C. Within the LE stretches are a number of globular domains (IV domains in Figure 3) that include; one in the β chain, one in the γ chain and two in the α chains (LiVa and LiVb). The β and γ chains end with a laminin coiled-coil (G) domain. The α chain contains five G domains (LG1-5). Functionally, the G domains are involved with the trimerization of the heterotrimer. During polymerization of ECM components, laminins interact with one another and also with other matrix components such as collagen IV. Cell surface integrins typically bind to laminin chains through their G domains.

Laminins have a critical role during many systemic processes such as embryogenesis [20], wound healing [21,22], and angiogenesis. In addition, laminins are central to many human diseases such as, muscular dystrophy type-1A [23], Pierson syndrome, epidermolysis bullousa [24-26], laryngo-onycho-cutaneous syndrome [27], and tumorigenesis [28,29].

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**Table 1:** Provides a list all identified Hydra laminins.

| Hydra Laminins Identified from Genomic and EST Analyses |
|--------------------------------------------------------|
| **Genomic Analyses**                                  | **EST Analyses**                        |
| Alpha chains                                          | Alpha chains                          |
| Laminin α1                                            | Laminin α 1                           |
| 100197510                                             | DN815114.2                            |
| Laminin α2                                            | Laminin α 1                           |
| 101235032                                             |                                   |
| Laminin α3                                            | Laminin α 1                           |
| 101241783                                             |                                   |
| Beta Chain                                            | Beta Chain                           |
| Laminin β1                                            | Laminin β1                           |
| 100202556                                             | D7613962.1                            |
| Laminin β2                                            | Laminin β1                           |
| 101236303                                             |                                   |
| Gamma chain                                           | Gamma chain                          |
| Laminin Y1                                            | Laminin Y1                           |
| 100198511                                             | D7613962.1                            |

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Composition of hydra laminins based on the most current NCBI data bases derived from various genomic and EST studies

Since the cloning and functional characterization of the Hydra laminin β1 chain and the Hydra laminin α chain (likely α1), no other articles have been published that focus on the detailed characterization of the other Hydra laminins. However, with the development of full genomic sequence data bases [8,9] (NCBI Hydra Genome Database: https://www.ncbi.nlm.nih.gov/genome/?term=Hydra) and EST data bases [10] (NCBI Hydra EST Database: https://www.ncbi.nlm.nih.gov/nucest), we now have a better understanding of the full spectrum of Hydra laminins. Based on current NCBI gene and EST data bases, Table 1 provides a list all identified Hydra laminins. Table 1 lists 6 Hydra laminin chains as analogs to vertebrate laminin chains. Some of these laminin chains were co-identified in both genomic studies and EST studies (laminin α1, laminin γ1). Other than Hydra laminin α1 and β1 [13,23], the functions for Hydra laminin α2, α3, β1, β3, and γ1 chains cannot be known without further cellular and molecular analyses.

Conclusion

From available studies and data bases, it is clear that laminins emerged early during the formation of metazoans and in tandem with the appearance of other ECM components. This likely reflects the critical function of ECM in metazoans. ECM functions encompass complex processes involving not only the structural integrity of tissues, but the dynamic nature of ECM/cell interactions as related to cell physiological processes, cell differentiation, morphogenesis, and development in general.

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