Use of coconut water (*Cocos nucifera L*) for the development of a symbiotic functional drink

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**ABSTRACT**

Many studies suggest that probiotic, prebiotic and symbiotic foods may be beneficial in the prevention and management of nutrition and health, the objective of this work was to develop a symbiotic drink based on coconut water. Fermentation was performed using lyophilized *Lactobacillus rhamnosus* SP1 and inulin as a source of soluble fiber. Different formulations were developed, determining the concentrations of fiber and probiotics. The growth of the probiotic in MRS broth was evaluated, using the plate counting technique in different periods of time. The fermentation time of the drink was 8 h and the shelf life in refrigeration was 14 days evaluated by pH and hedonic scale. The pH of the final drink was 3.48 and the probiotic content was $82 \times 10^8$ CFU/ml. It is concluded that coconut water can be processed by adding probiotic and prebiotic characteristics with sensory acceptance and adequate preservation characteristics.

1. Introduction

Coconut is the fruit of coconut palm (*Cocos nucifera*), also known as coconut, coconut palm, adiavan or Indian palm, and belongs to the Arecaceae family. It is a palm tree native to the eastern tropical regions, grown in Asia, America and Africa (Ramkhelawan and Paul, 2016).

In recent years the consumption of coconut products such as oil, dry coconut snacks, cosmetic products, bottled coconut water, coconut cream and milk has shown a much faster growth than the production of this tropical fruit (Granados and López, 2002; Prades et al., 2012). Coconut water is the liquid found in a young green coconut and should not be confused with coconut milk, since it refers to liquid products obtained by grating the solid endosperrm, with or without the addition of water (Yong et al., 2009).

Coconut water has been used successfully in various parts of the world for oral rehydration, treatment of childhood diarrhea, gastroenteritis and cholera (Ballit et al., 2018; Mujahid et al., 2019) and contains organic and inorganic compounds that play a vital role in helping the antioxidant system of the human body (Evans and Halliwell, 2001).

As it easily deteriorates once exposed to the air, most of the coconut water is consumed in its natural form in the areas where it is produced. Coconut water is commercially processed using ultra high temperature technology. However, it loses its delicate fresh flavor and some of its nutrients during heating (Awua et al., 2011; Kailaku et al., 2017), so would be desirable a non-thermal process to protect the fresh flavor and nutrient content of coconut water (Gautam et al., 2017). Until now, there is few studies in which alternative forms of conservation or transformation are proposed.

Consumers demand foods that, in addition to satisfying the taste, are healthy or contribute to their health, this demand can be covered by the so-called functional foods, hence the interest in the development of products through fermentation since these foods and its components have many possible health benefits (Melini et al., 2019). Fermented foods can be produced with simple economic ingredients and techniques, and can contribute significantly to the human diet, especially in rural homes and communities in villages around the world, so this research aims to develop a symbiotic functional drink using Coconut water as a base, *Lactobacillus rhamnosus* SP1 as probiotic and inulin as source of soluble fiber in quantity that allows to classify the product as functional and

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evaluate its probiotic characteristics through techniques that allow its standardization easily.

2. Materials and methods

A lyophilized strain of *Lactobacillus rhamnosus* SP1 (Clerici-Sacco, Italy), also known as *Lactobacillus rhamnosus* GG, was selected as it is one of the best studied and documented probiotic strains, with more than 300 clinical studies that have proven its efficacy and safety, is stable in acid and bile, has good adhesion to the mucous cells of the human intestine, and produces lactic acid. In experimental animals it has been shown to improve insulin sensitivity and reduce lipogenesis (Kim et al., 2013; Kun-Young et al., 2015), while in humans it has a positive immunomodulatory effect that could help reduce repeated rotavirus diarrhea episodes (Sindhu et al., 2014).

2.1. Preparation of the starter culture

To prepare the starter culture, the contents of the lyophilized strain sachet (10^{11}UFC) were poured into 1 L of sterile distilled water, homogenizing and subsequently cooling to 1.5 °C, the purity was verified via a Gram staining under a microscope, and short chains of gram positive bacilli were observed (Figure 1A). The strain was subcultured twice to obtain an active culture for the preparation of an inoculum by inoculating bacilli were observed (Figure 1A). The strain was subcultured twice to obtain an active culture for the preparation of an inoculum by inoculation in De Man Rogosa Sharpe broth (MRS, Merck, Germany) and incubated at 35 °C for 24 h. The culture was then stored at 1.5 °C and maintained on MRS broth until further use. In accordance with NOM-092-SSA1-1994, the purity was verified a literature review was conducted to determine fermentable carbohydrates by *L. rhamnosus*, selecting sucrose and lactose (to improve palatability), and it was decided to use a commercial coconut water, since the different treatments to which the fresh coconut water was subjected for pasteurization caused loss of aroma and development of slightly amber colors. The criteria for water coconut brand selection were that the only ingredient was coconut water (without additives or preservatives) and that would have been through inulin, for a total of 15 formulations (Table 1), 990 ml of each of these formulations was inoculated with 10 ml of the fermentation inoculum, then incubated at 35 °C and the pH was monitored until it decreased at least to pH < 5.0.

2.3. Sensory evaluation

A panel of 30 untrained homogenize (30 university students between 20 and 25 years of age) evaluated the sensory attributes of coconut water drinks for color, smell, taste and texture. The test was accomplished based on 5-point hedonic scale by panelists and scaled as 1 = dislike extremely, 2 = dislike moderately, 3 = neither like nor dislike, 4 = like moderately and 5 = like extremely. The samples, each of which received three digit code, served in plastic containers under normal light. The panelists received the random samples. They were asked to rinse their mouth with water between each simple tests.

2.4. Analytical determinations

Once the appropriate formulation was selected based on the sensory evaluation, the following analytical determinations were carried out, both for the fermented beverage and for the coconut water (control). The pH readings were made from the beginning of the incubation every hour to a pH < 5 and then every 3 days with a potentiometer (SM25CW Science Med, Finland), taking 20 ml aliquots of the drink (NMX-F-317-NORMEX-2013), until a minimum of pH 3 is reached, since at this pH all beverages were sensorially unacceptable. Humidity: by thermobalance (BL-MB45 Ohaus, New Jersey, USA). At a temperature of 70 °C. Ashes: by the calcination method, bringing the samples at a temperature of 500 °C for 8 h in a muffle (FB1300 Barnstead/Thermolyne, Iowa, USA). Proteins: by the method of Kjeldahl (KC24800 Labconco, Kansas, USA). Additionally, the dietary fiber to the fermented beverage was determined by the gravimetric-enzymatic method (AOAC 993.19).

2.5. Statistical analysis

The data obtained were analyzed using a unidirectional analysis of variance, and the Tukey test was used to evaluate the differences between treatments. All statistical analyzes were performed with the statistical package SPSS version 21 for Windows (IBM, Corp., Armonk, NY, USA).
Significant differences were considered if \( p < 0.05 \), and the results were expressed as mean values ± standard deviations.

2.6. Ethical aspects

The present study was presented to the Biochemistry Departmental Review Board, who after having made a series of questions concerning the safety of the study, informed participation and confidentiality of the information, decided to approve the study. All the tasters signed the consent form before performing sensory analyses.

3. Results and discussion

Currently, symbiotic foods (with mixtures of probiotics and prebiotics) are often used to take advantage of their synergistic effects in the application to food products (Al-Sheraji et al., 2013), and represent the new challenge for functional beverages, since prebiotics can improve the viability of probiotic bacteria and actively stimulate the beneficial microbiota in the human gastrointestinal tract (Corbo et al., 2014).

Once the viability and purity of the strain was verified, the different formulations shown in Table 1 were developed. The carbohydrates selected to add as carbon source were lactose and sucrose, both fermentable by Lactobacillus sp. (Camargo-Prado et al., 2015), however, in the case of lactose, it would have the disadvantage of not being suitable for the consumption of intolerant people. With regard to the prebiotic to be added, in addition to the multiple benefits reported in the literature for inulin, an important aspect is that the addition of 5% inulin increases the survival capacity of Lactobacillus spp. (Onal Darilmaz et al., 2019) with the additional advantage of further improving its activity against E. coli, so the initial percentage of inulin addition was 5% and it was decided to increase it to 15%, looking for the developed beverage to be prebiotic to show other possible functional effects in addition to those due to the probiotic (symbiotic). Maintaining in all formulations an inoculum of 1% starter culture, the incubation was carried out at 35 \(^\circ\)C and pH readings were made every 3 days until reaching a pH of 3, since from this value the sensory characteristics of the beverage was no longer properly preserved (Figure 2), which determines that the shelf life of the beverage in refrigeration once processed, is approximately 15 days.

In order to evaluate the repeatability of the formula and establish the time and temperature of incubation, 5 formulations in 200 ml bottles were prepared for one week, and allowed to incubate in the oven at 35 \(^\circ\)C until the desired pH was reached and after that they were refrigerated to 1.5 \(^\circ\)C, obtaining in each case a very similar drink in each processing batch (Figure 3) and establishing that the initial inoculum must be 1% of the starter culture and the incubation conditions 35 \(^\circ\)C for 8 h.

It was decided to employ untrained panelists because the objective was to determine the reaction of the potential consumer to the beverage and, according to Watts et al. (1989), with untrained panelists it is possible to obtain information about likes and dislikes, preferences and acceptability requirements. The results of the sensory evaluation are shown in Table 2. The color was statistically the same in all the formulations, however, in relation to other characteristics (smell, taste and texture), the lactose-containing formulations obtained the lowest scores, this finding was very favorable since it allowed to discard lactose as an
ingredient and develop a suitable drink for lactose intolerant people. Of the formulations with lactose, the only one that was not rejected was the 10% formulation but without inulin which would lead to a probiotic but not symbiotic drink. Significantly, the formulations K and M, both with sucrose, were the most accepted in terms of taste and general acceptance, without differences between them, so the formulation K was finally selected because it contained less sucrose and more fiber so it could be considered symbiotic.

One of the factors that most influence eating behavior, along with cost, safety and accessibility is the sensory aspect (Wiodarska et al., 2019), so the main criterion for the choice of the final formula was the sensory evaluation, also because the sensory effect derived from the inclusion of probiotics in products of plant origin is critical in the development of functional drinks (Ozcan et al., 2016).

This parameter has been explored by some authors, for example, salty, sour and scented flavors have been reported in fruit drinks with the addition of probiotics (Perricone et al., 2015). Luckow and Delahunty (2004) concluded that orange juices with probiotics (Lactobacillus rhamnosus GG, Lactobacillus casei strain 01, Lactobacillus paracasei NFBC 43338) presented unpleasant sensory profiles described as medicinal flavors and without dairy touches. However, it is suggested that exposure and familiarity with probiotic drinks helps improve consumer acceptance and taste for the sensory characteristics of fruit drinks with probiotics (Luckow et al., 2005).

Once the formulation was selected, the corresponding physicochemical analysis was performed. Table 3 shows the composition of the developed beverage. It can be seen that the proximal composition in the developed beverage is very similar to that reported for coconut water, however, according to different studies, it can be very variable, so it is difficult to establish comparisons (Prades et al., 2012; Yong et al., 2009).

Few processes have been reported for the transformation of coconut water, some of the works developed so far include a patent granted to Soccol (2009) and subsequently published (Camargo-Prado et al., 2015) of a fermented beverage with Lactobacillus plantarum using coconut water added with yeast extract, sugar and soy protein hydrolyzate; the coaddition of coconut water with Bacillus clausii and Saccharomyces boulardii (Rachana, 2017); the development of a hydration drink without fermentation based on serum and up to 30% coconut water (Murillo-Calderón, 2015); a drink fermented with Lactobacillus plantarum DW12 but of mature coconut water, which has different characteristics to those of immature coconut water, added with glutamate and fermented sugar cane or honey up to 20% (Kantachote et al., 2017), and a drink based on coconut water but fermented with Bacillus coagulans and oriented mainly to determine the probiotic potential of this bacterium (Gangwar et al., 2018), so these results allow us to point out that it is possible to develop a symbiotic drink with proximal characteristics quite similar to those of coconut water with the additional contribution of the reported beneficial effects on the health of L. rhamnosus and the content of dietary fiber that makes it possible to consider the developed beverage as symbiotic.

As a quick method to evaluate the growth of Lactobacillus rhamnosus, the use of the MacFarland scale and spectrophotometric readings were evaluated, however, due to the characteristics of the beverage, this method was not viable and the variation in pH with respect to the time compared to some commercial probiotic products was used as reference (Figure 4). Based on the pH, it could be concluded that the developed beverage is stable with respect to the pH compared to similar commercial products, the slow but continuous decrease of the pH of the developed beverage is probably due to the inulin content, since according to several researchers, it can be used by Lactobacillus spp, as carbon source (Onal Darilmaz et al., 2019; Balthazar et al., 2018; De Souza et al., 2012).

4. Conclusions

It was possible to develop a fermented coconut water formulation with L. rhamnosus stable in relation to physical and chemical
The viability of *L. rhamnosus* was above the minimum required to be considered probiotic. The fermented beverage developed has a good sensory acceptance and meets the criteria of functional feeding, since it has an amount of $82/\times10^8$ CFU/ml of *L. rhamnosus* and 15 g of inulin. The shelf life of the product developed in refrigeration was 15 days.

The critical characteristic for the shelf life of coconut water proved to be of a sensory nature, related to the appearance of flavors, odors and undesirable appearance, probably caused by residual enzymatic activity of the product. The critical parameter would be the decrease in consumer satisfaction, after day 15 of storage at $1.5^\circ C$.

**Declarations**

**Author contribution statement**

Orietta Segura-Badilla, Martín Lazcano-Hernández, Obdulia Vera-López, Joaquín Ramirez-Calixto: Performed the experiments; Wrote the paper.

Ashuin Kammar-García, Patricia Aguilar-Alonso: Performed the experiments; Analyzed and interpreted the data; Wrote the paper.

Addí Rhode Navarro-Cruz: Conceived and designed the experiments; Performed the experiments; Wrote the paper.

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**Competing interest statement**

The authors declare no conflict of interest.

**Additional information**

No additional information is available for this paper.

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Table 3. Percentage composition of the symbiotic drink developed and coconut water used compared to values reported in the literature.

|                       | Symbiotic drink | Coconut water used | Prades et al. (2012) | Yong et al. (2009) |
|-----------------------|-----------------|--------------------|-----------------------|--------------------|
| Dry material          | 25.07 ± 0.02    | 6.0 ± 0.08         | 4.5 ± 0.7             | 5.01               |
| Humidity              | 74.93 ± 0.02    | 94 ± 0.08          | NR                    | 94.99              |
| Ashes                 | 0.40 ± 0.01     | 0.45 ± 0.01        | 0.43 ± 0.04           | 0.39               |
| Protein               | 0.04 ± 0.002    | 0.19 ± 0.002       | 0.25 ± 0.26           | 0.72               |
| Fat                   | ND              | ND                 | 0.51 ± 0.33           | 0.20               |
| Dietary fiber         | 12.43 ± 0.19    | ND                 | NR                    | 1.1                |
| Nitrogen Free Extract | ND              | 5.2                | ND                    | 3.71               |

ND not detected, NR not reported.

Figure 4. pH of commercial probiotic drinks vs developed beverage.
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