Primary del 17 chronic lymphocytic leukaemia lymphocytes are hypersensitive to dasatinib in vitro

Chronic lymphocytic leukaemia (CLL) is characterized by the accumulation of mature quiescent B-lymphocytes in the G0/G1 phase of the cell cycle. B-lymphocyte accumulation is likely to be a consequence of an undefined defect in the apoptotic machinery rather than an increased proliferation of leukaemic cells (Hamblin & Oscier, 1997). The prolonged survival of CLL lymphocytes has also been linked to deregulated expression and/or activity of related non receptor tyrosine kinases including members of the SRC family kinases (SFK) and ABL1. Inhibition of either c-abl (Aloyz et al, 2004) or SFK (Contri et al, 2005) results in CLL lymphocyte death in vitro. Previous results from our laboratory suggest that dasatinib cytotoxicity in CLL lymphocytes is associated with c-abl rather than SFK inhibition (Amrein et al, 2008a). Recent results of phase I–II clinical trials using dasatinib in CLL suggest that the drug might be beneficial in only a small subset (≤10%) of previously treated patients (Amrein et al, 2008b). In agreement with these clinical results, we have recently reported that dasatinib is cytotoxic to primary CLL lymphocytes in vitro but mainly at clinically unobtainable concentrations (Amrein et al, 2008a). Although dasatinib resistance was associated with the basal expression of c-abl, our study suggested that wild type TP53 is important in CLL lymphocyte homeostasis and/or survival in the presence of dasatinib (Amrein et al, 2008a). To test this hypothesis we assessed: (i) dasatinib cytotoxicity in p53 proficient (wild type) CLL lymphocytes treated with dasatinib in the presence or absence of pifithrin-α, a small molecule inhibitor of p53 transcriptional activity (Steele et al, 2008) and 2) dasatinib cytotoxicity in primary CLL lymphocytes with impaired TP53 signalling from patients diagnosed with del 17p13.1. In the eleven samples we verified the functionality of p53 signalling as described by examining changes in p53 and its downstream target p21Cip1 (p21) after treatment with chlorambucil (Willmore et al, 2008). Briefly, the lymphocytes were treated with equivalent cytotoxic concentrations of chlorambucil (50% inhibitory concentrations [IC50]) for 24 h and induced p53 and p21 protein levels were monitored by Western blotting. As expected p53 and p21 protein levels were induced by chlorambucil only in TP53 wild type CLL lymphocytes (data not shown).

As previously reported, dasatinib IC50 in vitro in CLL lymphocytes expressing wild type TP53 were not in the clinically attainable range (mean value of 30 μmol/l, Table I) (Amrein et al, 2008a). In contrast, the dasatinib IC50 in del 17p13.1 lymphocytes were significantly lower (up to 100 times) than in the wild type TP53 lymphocytes. Moreover, in agreement with previous reports demonstrating that del 17p13.1 is associated with chemoresistance in CLL lymphocytes, we found that del 17p13.1 lymphocytes were significantly more resistant to chlorambucil that TP53 wild type CLL lymphocytes [Table I, Fig 1A (Zenz et al, 2008)].

In available wild type TP53 CLL samples, pifithrin-α sensitized the lymphocytes of 2 out 3 patients (1-3 and 12-fold) (Table I). Importantly, pifithrin-α was not toxic to CLL lymphocytes when used alone. Treatment with dasatinib for twenty four hours resulted in a dose dependent reduction of p53 and p21 basal expression levels in the lymphocytes of

Table I. Clinical characteristic of the patients and IC50 concentrations of dasatinib and chlorambucil in primary CLL-lymphocytes in vitro.

| Patient | Deletion 17 | Deletion 11 | RAI stage | Previous treatment | Dasatinib (μmol/l) | Chlorambucil (μmol/l) |
|---------|-------------|-------------|-----------|--------------------|-------------------|---------------------|
| 1       | Negative    | I           | CLB       | 0.8                | 17.3              |
| 2       | Negative    | 0           | FLU       | 30.6 (2.4)         | 23.0              |
| 3       | N.D.        | III         | CLB       | 7.2                | 30.0              |
| 4       | Negative    | 0           | No        | 0.1                | 41.0              |
| 5       | Negative    | 0           | No        | 36.0 (27.0)        | 18.6              |
| 6       | N.D.        | III         | CLB       | 28.2               | 10.9              |
| 7       | Negative    | II          | CLB       | 40.0 (40.0)        | 24.9              |
| 8       | Del 17p13.1 (86%) | II   | CLB       | 0.27               | 36.7              |
|         | Del 11q22–23 (N.D.) |     |           |                    |                   |
| 9       | Del 17p13.1 (94%) | I     | CLB/FLU   | 0.01               | 44.7              |
|         | Del 11q22–23 (5%) |     | Rituximab  |                    |                   |
| 10      | Del 17p13.1 (79%) | I     | No        | 0.01               | 100               |
|         | Del 1q22–23 (76%) |     |           |                    |                   |
| 11      | Del 17p13.1 (33%) | II    | CLB       | 0.17               | 100               |
|         | Del 11q22–23 (10%) |     |           |                    |                   |

Primary CLL lymphocytes, isolated and plated as described above, were incubated in the presence of dasatinib or chlorambucil (CLB). The IC50 concentrations are expressed in μmol/l and determined after 72 h incubation in vitro using the MTT (3-(4,5-Dimethylthiazol-2-yl)-2,5-Diphenyltetrazolium Bromide) assay as described (Amrein et al, 2008a). The numbers in parentheses indicate the dasatinib IC50 in the presence of 25 μmol/l pifithrin-α. The percentage of lymphocytes with del 17 and del 11 in each samples are indicated. Samples displaying percentages above 4.8% and 6.6% for del 17 and del 11 respectively were considered positive.

Negative percentage lower than the cut-off point.

N.D. not determined; CLB, chlorambucil.
Dasatinib decreases p53 basal expression levels in primary CLL lymphocytes expressing wild type p53. Dasatinib and chlorambucil IC$_{50}$s were significantly different between CLL lymphocytes expressing wild type TP53 or del 17 (\(P = 0.012\)). The bars represent the median values and 95% confidence intervals (CI 95%). (A). The lymphocytes of 11 CLL lymphocyte patients were treated for 24 h with vehicle (dimethyl sulphoxide), dasatinib 100 nmol/l or the IC$_{50}$ concentration as shown in Table I. Protein extracts were obtained as described before and 50 µg of proteins for each sample were resolved by sodium dodecyl sulphate polyacrylamide gel electrophoresis. p53 and p21 protein levels were assessed by Western blot using specific antibodies (Amrein et al., 2008a). The signals obtained in TP53 wild type lymphocytes (B) were analysed using National Institutes of Health - Scion image and normalized to actin, p53 or p21 levels (y-axis) and are expressed as the percentage of vehicle treated lymphocytes (control) value (\(|\text{OD value/control OD value}| \times 100\)) vis à vis the treatments indicated in the x-axis; * and ** indicates significance \(P = 0.003\) and \(P = 0.004\) respectively (C). Dasatinib-induced changes in p53 levels and p21 signal were not detected in protein extracts from del 17p13-1 CLL lymphocytes (D). Dasatinib IC$_{50}$s correlate with the percentage of residual p53 protein levels (in respect to vehicle treated lymphocytes) after dasatinib treatment, \(r = 0.82, P = 0.02\) (E). Two-sided tests with \(a\)-value of 0.05 were used. Correlations between the data were assessed using the Spearman test. All tests were performed using SigmaStat software.
patients expressing wild type TP53 (Fig 1B, C). In contrast, p53 levels were not affected in del 17 lymphocytes. Importantly, p21 was not detected in del 17p13.1 lymphocytes suggesting that TP53 is not functional (Fig 1D). These results are in agreement with previous reports suggesting that in the majority of CLL patients with malignant lymphocytes displaying del 17p13.1, the remaining TP53 allele is mutated (Zenz et al, 2008).

In addition, we found that dasatinib resistance (higher IC50) in CLL lymphocytes expressing wild type TP53 correlated with residual p53 protein levels (in respect to control) after dasatinib treatment ($r = 0.8$, $P = 0.02$, Fig 1E). As ATM is a key regulator of p53 functionality, we assessed del 11q22-23 status in del 17p13.1 lymphocytes (Table I) (Pettitt et al, 2001). Two of the three del 17p13.1 samples tested were positive for del 11q22–23. Although del 17p13.1 lymphocytes were hypersensitive when compared to p53 proficient lymphocytes, we did not find a correlation between the percentage of del 17p13.1 or del 11q22–23 in the lymphocytes and the IC50 of dasatinib. Studies regarding the role of p53 signalling (and its regulators e.g. ATM) in dasatinib sensitivity in a larger cohort of CLL samples should be informative.

Taken together, our results suggest that p53 is important to maintain CLL lymphocyte homeostasis following exposure to dasatinib and suggest that dasatinib may be effective to treat del 17p13.1 CLL patients. The recent report of an excellent clinical response to dasatinib of a CLL patient with lymphocytes displaying del 17p13.1 supports this hypothesis (Pitini et al, 2009).

Acknowledgement

This work was supported by a CIHR grant to R. Aloyz.

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Keywords: chronic lymphocytic leukaemia, p53, del 17, dasatinib.

First published online 2 September 2009

doi:10.1111/j.1365-2141.2009.07814.x

MicroRNA expression in chronic lymphocytic leukaemia

B-cell chronic lymphocytic leukaemia (B-CLL) is the most common adult leukaemia in the Western world and is characterized by the accumulation of CD5+CD19+/CD23+ mature B cells. The molecular pathology of the disease however