Review

Natural Products in Cardiovascular Diseases: The Potential of Plants from the Allioideae Subfamily (Ex-Alliaceae Family) and Their Sulphur-Containing Compounds

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Abstract: Cardiovascular diseases (CVDs) are the leading cause of mortality worldwide and, together with associated risk factors such as diabetes, hypertension, and dyslipidaemia, greatly impact patients’ quality of life and health care systems. This burden can be alleviated by fomenting lifestyle modifications and/or resorting to pharmacological approaches. However, due to several side effects, current therapies show low patient compliance, thus compromising their efficacy and enforcing the need to develop more amenable preventive/therapeutic strategies. In this scenario, medicinal and aromatic plants are a potential source of new effective agents. Specifically, plants from the Allioideae subfamily (formerly Alliaceae family), particularly those from the genus Allium and Tulbaghia, have been extensively used in traditional medicine for the management of several CVDs and associated risk factors, mainly due to the presence of sulphur-containing compounds. Bearing in mind this potential, the present review aims to gather information on traditional uses ascribed to these genera and provide an updated compilation of in vitro and in vivo studies validating these claims as well as clinical trials carried out in the context of CVDs. Furthermore, the effect of isolated sulphur-containing compounds is presented, and whenever possible, the relation between composition and activity and the mechanisms underlying the beneficial effects are pointed out.

Keywords: Allium; Tulbaghia; extracts; sulphur-containing compounds; antiplatelet aggregation; hypertension; diabetes; dyslipidaemia

1. Introduction

Cardiovascular diseases (CVDs) continue to lead mortality rates worldwide [1], accounting for nearly 18 million annual deaths, primarily due to coronary heart disease and stroke [2]. Unfortunately, these numbers tend to increase as several non-modifiable and modifiable risk factors associated with the onset and development of these disorders are also escalating. While non-modifiable risk factors such as aging, gender, genetic predisposition, family history of heart-related problems and ethnicity cannot be altered [3–5], modifiable risk factors are changeable. These include hypertension, dyslipidaemia, diabetes, obesity, smoking, alcohol misuse, unhealthy diet, sedentary lifestyle, and psychosocial factors [6] and are recognised as relevant targets to manage CVDs. For example, the INTERHEART case–control study pointed out that 90% of acute myocardial infarction cases are due to these risk factors and that controlling or eliminating them per se could lead to a drastic decrease in CVD mortality [7,8]. Indeed, due to their huge impact on CVDs, these risk factors are included in the World Health Organisation (WHO) target list that...
aims to reduce their prevalence by 2025 [9]. The negative impact of CVDs is further fuelled by the fact that 60% of patients fail to correctly adhere to the therapeutic regimen [10], mainly due to the cost of CVD therapies [11]. Therefore, new therapeutic interventions and/or preventive strategies with fewer side effects are mandatory, with aromatic and medicinal plants emerging as promising agents to manage both CVDs and associated risk factors. In fact, herbal medicines are relevant sources of bioactive molecules, used by ca. 80% of the world’s population in basic health care [12]. Moreover, many of these medicinal plants have already been used in the treatment of chronic and acute conditions including CVDs [13–16] and are part of the Mediterranean-style diet with proven beneficial effects on cardiovascular risk factors [17], as pointed out in several meta-analysis and critical reviews [18–23]. Interestingly, these effects are associated with the increased consumption of fruit, vegetables, spices, garlic, and onions [24]. Overall, the preventive/therapeutic potential of aromatic and medicinal plants is mainly attributed to the presence of secondary metabolites [25] including phenolic compounds, terpenes, alkaloids, and organosulfur compounds [26]. Organosulfur compounds are widely found in plants from the Allioideae subfamily (ex-Alliaceae family) and, together with extracts or raw bulbs from these plants, are widely reported for their medicinal properties [27]. Therefore, bearing in mind the bioactive potential of these plants, a systematised review gathering information on the effects of sulphur-containing extracts/compounds on major CVD risk factors, namely hypertension and dyslipidaemia/diabetes, is presented. Additionally, whenever reported, the mechanisms underlying the observed effects are referred to and the relation between composition and activity pointed out. To achieve this, a bibliographic search was conducted using Pubmed, Scopus and Google scholar databases, combining the keywords “Allium”, “Tulbaghia”, “Alliaceae” or “Allioideae” with “cardiovascular”, “diabetes”, “obesity”, “dyslipidaemia”, “hypertension” or “vasorelaxation”. Studies published over the last 20 years that had an available DOI were considered.

2. Importance of Allioideae Species in Cardiovascular Diseases

In the following sections, both the relevance and potential of plants from the Allioideae subfamily are described. First, the traditional uses ascribed to these plants in several ethnobotanical surveys is shown in order to highlight their importance in local health care systems. Then, studies validating some of these effects are systematised, considering pre-clinical approaches and clinical trials. The effect of isolated sulphur-containing compounds is also presented, and whenever possible, the relation between composition and activity is discussed, thus opening new avenues for further investigations in the field.

2.1. Traditional Uses of Allioideae

A plethora of traditional uses are ascribed to Allioideae plants or plant-based preparations, as summarised in Table 1. Plants’ scientific and common names are included as well as the region of use. In addition, the plant part or preparations used (with reference to the preparation method and posology, when known) and beneficial effects on the cardiovascular system are pointed out. Overall, the majority of the studies focus on the genus Allium, with only a few studies reporting the effects of two species from the Tulbaghia genus. In traditional preparations, the plant bulb is commonly used (7/11 total studies), with leaves (1/11), aerial parts (1/11) or whole plants (1/11) referred to in much less often. In addition, the use of a combination of plants is frequent and, therefore, this information is also provided. A list of abbreviations, used throughout the table, is provided at the end of the table.

2.2. Pre-Clinical Studies Validating the Cardioprotective Effects of Allioideae

Given the importance of Allioideae plants in the management of CVDs and associated risk factors in ethnopharmacological studies, we next compile several pre-clinical studies validating these effects. First, the effect of plants or their extracts is pointed out (Table 2) followed by the effect of isolated sulphur-containing compounds (Table 3) and then clinical trials.
Table 1. Traditional uses ascribed to plants from the Allioideae subfamily.

| Scientific Name (Common Name) | Region of Use                      | Plant Part or Preparation (Mode of Administration; Posology) | Cardiovascular Disease/Risk Factor (Observed Effect) | Ref. |
|-------------------------------|------------------------------------|---------------------------------------------------------------|-----------------------------------------------------|------|
| Allium ampeloprasum L. (wild leek) | Suva planina, Serbia               | Raw aerial parts (oral)                                        | Diabetes                                            | [28] |
|                               | Serra de Mariola, Spain            | Whole plant (oral)                                             | Hypertension                                        | [29] |
|                               | Beni Mellal, Morocco               | Raw bulb (oral)                                                | Diabetes                                            | [30] |
|                               | Gabon                              | Bulb (oral)                                                    | Diabetes, hypertension                              | [31] |
| Allium cepa L. (onion)        | Edo, Nigeria                       | Mined and blended bulb mixed with honey (oral; one tablespoonful twice a day) | Hypertension                                        | [32] |
|                               |                                    | Bulb maceration—soaked in water with Vernonia amygdalina and Zingiber officinale for 5 days (oral; one cup twice a day) |                                      |      |
|                               |                                    | Bulb decoction with snail water and Capsicum frutescens, add small salt and filter (oral; one small cup twice daily) | | |
|                               |                                    | Bulb concoction with Viscum album, Persea americana, Ocimum gratissimum added with Elaeis guineensis kernel oil and boiled for 10 min (oral; drink as a soup) | | |
|                               | Tamil Nadu, India                  | Bulb mixed with buttermilk (oral)                              | Cardiovascular disease                              |      |
|                               |                                    | Bulb boiled in milk, sugar from Borassus flabellifer added (oral; once a day in the evening) |                                      |      |
|                               |                                    | Raw bulb, (oral; daily before eating for 45 days)               | Dyslipidaemia                                       | [33] |
|                               |                                    | Raw bulb (oral; daily in the morning for 20 days)               | | |
|                               |                                    | Juice from bulbs (oral; 25 mL in the morning for two weeks)     | | |
|                               |                                    | Bulb decoction with Macrotyloma uniflorum, Zingiber officinale and honey (oral; daily in the morning for 30 days) | | |
| Scientific Name (Common Name) | Region of Use | Plant Part or Preparation (Mode of Administration; Posology) | Cardiovascular Disease/ Risk Factor (Observed Effect) | Ref. |
|-----------------------------|---------------|-----------------------------------------------------------|-----------------------------------------------------|-----|
| Allium jacquemontii Kunth.  | Dir, Pakistan | Raw bulb, (oral; for 3 weeks)                             | Hypertension                                         | [34]|
| Allium rotundum L.          | Aladaglar, Turkey | Raw bulb (oral)                                         | Hypertension (regulates blood pressure)              | [35]|
|                            |                |                                                           |                                                     |     |
| Eastern Cape, South Africa  |                | Not referred                                             | Diabetes                                            | [36]|
| Gabon                      |                | Bulb maceration                                          | Diabetes, dyslipidaemia                              | [31]|
| Suva planina, Serbia        |                | 3 peeled bulbs with 3 chopped lemons in 1 L of hot water for 12 h) (oral; 1 cup a day for 40 days) | Dyslipidaemia (decreases TG), hypertension (improves blood circulation) | [28]|
| Togo                       |                | Bulb decoction with *Khaya senegalensis* bark (oral)      | Diabetes                                            |     |
|                            |                | Bulb maceration with honey (oral)                        |                                                     |     |
|                            |                | Bulb maceration with *Parkia biglobosa* (oral)           |                                                     |     |
|                            |                | Bulb powder containing *Lippia multiflora*, *Stachytarpheta angustifolia* and *Persea americana* (oral) | Hypertension                                        | [37]|
| Edo, Nigeria               |                | Bulb maceration—soaked with guava, *Vernonia amygdalina* in water for 5 days (oral; half a cup daily) |                                                     |     |
|                            |                | Bulb decoction combined with *Allium cepa* and boiled in water (oral; half a cup twice a day) |                                                     |     |
|                            |                | Bulb decoction combined with *Allium cepa* and *Zingiber officinale* and boiled in water (oral; one cup twice a day) |                                                     |     |
|                            |                | Bulb decoction with *Cocos nucifera* boiled for 3 days (oral; half a cup twice a day) |                                                     |     |
|                            |                | Bulb infusion after pounding with *Carica papaya* (oral; one small cup 3 times a day) |                                                     |     |
| Scientific Name (Common Name) | Region of Use | Plant Part or Preparation (Mode of Administration; Posology) | Cardiovascular Disease/ Risk Factor (Observed Effect) | Ref. |
|-----------------------------|---------------|-------------------------------------------------------------|------------------------------------------------------|-----|
| Bulb decoction with *Musanga cecropoides*, *Talinum triangulare*, *Carica papaya* boiled in water (oral; half a cup twice a day) | Western Anti-Atlas, Morocco | Raw bulb (oral) | Diabetes | [38] |
| Bulb decoction with *Hunteria umbellata, Sida acuta* and potash in cold water (oral; one cup a day) | | | | |
| Boiled bulb with *Zingiber officinale* rhizome and added milk (oral; twice a day) | Tamil Nadu, India | Bulb cooked with *Foeniculum vulgare* in milk (oral; daily in the morning until cure) | Cardiovascular disease | |
| Bulb cooked with *Trachyspermum ammi* in milk (oral; daily in the morning until cure) | | Bulb boiled in water, and added milk (oral; daily in the evening) | Hypertension | |
| Bulb boiled in milk (oral; daily in the evening for a month) | | Bulb as a food supplement with *Moringa oleifera* (oral; twice a week at lunch) | Obesity | [33] |
| Bulb syrup with *Citrus limon* juice, *Zingiber officinale* rhizome, *Malus pumila* cider vinegar and honey (oral; 10 mL twice a day) | | Bulb syrup with *Citrus limon* juice, *Zingiber officinale* rhizome, *Malus pumila* cider vinegar and honey (oral; 15 mL twice a day after meals) | Cardiovascular disease, obesity | |
| Bulb paste with *Coriandrum sativum, Solanum torvum* and *Zingiber officinale*, consumed with honey (oral; once a day in the morning) | | Bulb combined with honey (oral; 5 mL once a day in the morning) | Cardiovascular disease | |
| | | | | |
| Scientific Name (Common Name) | Region of Use          | Plant Part or Preparation (Mode of Administration; Posology) | Cardiovascular Disease/Risk Factor (Observed Effect) | Ref. |
|-------------------------------|------------------------|----------------------------------------------------------------|------------------------------------------------------|------|
|                              |                        | Bulb combined with honey (oral; 5 mL twice a day for a month) | Dyslipidaemia                                         |      |
|                              |                        | Bulb combined with honey (oral; 10 mL twice a day)             | Hypertension                                          |      |
|                              |                        | Bulb jam with sugar from *Borassus flabellifer* and oil from *Sesamum indicum* (oral; 10 g twice a day for 45 days) | Dyslipidaemia                                         |      |
| Tamil Nadu, India             |                        | Bulb jam with sugar from *Borassus flabellifer* (oral; 20 g twice a day till cure) | Obesity                                              |      |
|                              |                        | Bulb jam with sugar from *Borassus flabellifer* and oil from *Sesamum indicum* (oral; 10 g twice a day) | Hypertension                                         |      |
|                              |                        | Bulb powder with *Cinnamomum verum*, *Piper cubeba* and *Vitis vinifera* (oral; 2–3 g once a day in the evening) | Dyslipidaemia                                         |      |
| Beni Mellal, Morocco          |                        | Bulb gravy with *Arachis hypogea*, *Cissus quadrangularis*, *Murraya koenigii*, *Tamarindus indica* and clarified butter (oral; twice a week until cure) | Dyslipidaemia                                         |      |
| *Allium ursinum* L. (wild garlic) | Suva planina, Serbia | Leaf tincture diluted in a small glass of water (oral; 10 drops 3 times a day before meals) | Hypertension, hypercholesterolemia (lowers blood cholesterol) | [28] |
| *Tulbaghia alliacea* L.       | Eastern Cape, South Africa |                                                |                                                      |      |
| *Tulbaghia violacea* Harv.    |                                                | Not referred                                                  | Diabetes                                              | [36] |

TG—triglycerides.
Table 2. Effects of plant parts/extracts from the Allioideae on the cardiovascular system.

| Plant Species          | Plant Part or Extract Used (Preparation; Concentration) | Study Model: Insult or Injury | Main Findings                                                                 | Ref. |
|------------------------|---------------------------------------------------------|------------------------------|--------------------------------------------------------------------------------|------|
| *Allium cepa*          | Aqueous extract (i.v. 30 min before injury; 0.1, 0.3 and 1 g/kg) | Rat: Brain ischaemia         | ↓ Brain edema; prevented ZO-1 and occludin disruption; ↑ Cat and GPx; ↓ MDA     | [39] |
|                        | Methanolic extract (0.01, 0.05 and 0.1 g/mL)            | Cardiomyoblasts (H9c2): Hypoxia | ↓ ROS production, mitochondrial membrane depolarisation, cytochrome c and caspase-3 release | [40] |
|                        | Methanolic extract (p.o. 14 days before injury; 0.1, 1 and 10 g/Kg) | Rat: Ischaemic injury        | ↓ Infarct area, apoptotic cell death and MDA                                  |      |
| *Allium macrostemon*   | Decoction with bulbs and *Trichosanthes kirilowii* (gavage for 4 weeks; 1.14, 2.27 and 4.53 g/Kg) | Rat: LAD ligation-induced infarction | ↓ HW/BW, LV/BW, systemic inflammation, myocardial fibrosis, and collagen I and III expressions; ↓ TGFβ1, TGFβ2 and Smad 2/3 expression; ↑ Smad7 expression | [41] |
| *Allium sativum*       | Aged garlic extract (p.o. for 3 weeks; 2 and 5 mL/Kg)   | Rat: ISO-induced myocardial infarction | ↑ Heart function, SOD and Cat; ↓ LDH, CK-MB and MDA | [42,43] |
|                        | Homogenate (p.o. for 30 days; 125, 250 and 500 mg/kg)   |                                             | ↑ SOD and cat; ↓ LDH, CK-MB and structural changes | [44] |
|                        | Raw homogenate (p.o. 30 days before injury; 125, 250 and 500 mg/kg) | Rat: I/R                        | ↓ MDA and structural changes; ↑ SOD, cat, GSH and GPx | [45] |
|                        | Black garlic extract (gavage for 4 weeks; 300 mg/kg)    |                                             | ↑ HO-1                                                                    | [47] |
|                        | Raw garlic extract (gavage for 4 weeks; 300 mg/kg)      | Rat: ISO-induced myocardial necrosis     | ↓ HW, LDH, CK-MB, cTnC and systemic inflammation; ↑ SOD and cat               | [48] |
| *Allium ursinum*       | Methanolic extract (p.o. 28 days before injury in drinking water; 125, 250, and 500 mg/kg) | Rat: I/R                        | ↑ Cardiac function and antioxidant system                                   | [49] |
| *Tulbaghia violacea*   | Methanolic extract (intragastric for 30 days before injury; 60 mg/kg) | Rat: ISO-induced myocardial infarction | ↓ CK, CK-MB, LDH and MDA; ↑ LV function, SOD and GSH | [50] |
| Plant Species               | Plant Part or Extract Used (Preparation; Concentration) | Study Model: Insult or Injury | Main Findings                                                                                   | Ref. |
|----------------------------|----------------------------------------------------------|-------------------------------|-------------------------------------------------------------------------------------------------|------|
| Allium cepa                | Aqueous extract (p.o. for 4 weeks; 0.5, 1.5 and 4.5 g/kg) | Rat: HFD-induced hyperlipidaemia | ↓ TC, LDL, MDA, lipid droplets in liver, foam cell accumulation and HMG-CoA; ↑ HDL, SOD and LDLR | [51] |
| Allium cepa var. destiny   | Raw onion (p.o. for 6 weeks; 16 and 40 g/kg)            |                               | ↓ TC, glucose, LDL, HDL, TG, erythrocyte number and haemoglobin; ↑ white blood cell number     | [52] |
| Allium elburzense          | Hydroalcoholic extract (intragastric for 7 days, 100, 200, and 400 mg/kg) | Rat: DEX-induced diabetes     | ↓ TG, TC, LDL, MDA and liver steatosis; ↑ HDL                                                | [53] |
| Allium eriophyllum         | Hydroalcoholic extract (gavage for 4 weeks; 30 and 100 mg/kg) | Rat: T2DM + Hypertension      | ↓ SBP, BG, CK-MB, infarct size and coronary resistance; ↑ SOD, GSR                             | [54] |
| Allium hirtifolium         | Ethyl acetate fraction from hydroalcoholic extract (gavage for 4 weeks; 5 mg/kg) | Rat: STZ-induced diabetes     | ↓ BW, BW gain, adipose tissue, TG, TC, LDL, AI, cardiac risk factor, LDH, AST and ALP        | [55] |
| Allium hookeri             | Powder (p.o. for 4 weeks; 3% and 5% in chow)           | Rat: HFD-induced obesity      | ↓ TG, TC and LDL                                                                              | [56] |
| Allium hookeri             | Powder (p.o. for 13 weeks; 0.2 g/Kg)                   | Hamster: HFD-induced obesity  | ↓ liver and adipose tissue weight, TG, TC, LDL, AI, AST and ALT; ↑ HDL                        | [57] |
| Allium hookeri             | Hydroalcoholic extract (p.o. for 4 weeks; 200 and 400 mg/kg in chow) | Mice: HFD-induced obesity     | ↓ liver and adipose tissue weight, TG, TC, LDL, AI, AST and ALT; ↑ HDL                        | [58] |
| Allium sativum             | Aged garlic extract (intra-abdominal injection every 12 h for one month; 125 mg/kg) | Rat: Metabolic syndrome       | ↓ TG, insulin, leptin, AGE, SBP and MDA; ↑ GSH, and GPx; restored vascular and cardiac function | [59] |
|                            | Aged garlic extract (1, 2.5 and 5 g/L)                 | HUVEC: oxidLDL                | ↓ LDH release and cell damage                                                                  | [60] |
|                            | Aged garlic extract (p.o. for 12 or 24 weeks; 3% in chow) | Mice: ApoE−/−                 | ↓ Atherosclerotic lesions, TC, TG and CD11b+ cells in spleen                                  | [61] |
| Plant Species                               | Plant Part or Extract Used (Preparation; Concentration) | Study Model: Insult or Injury                                                                 | Main Findings                                                                                                                                                                                                 | Ref. |
|--------------------------------------------|--------------------------------------------------------|---------------------------------------------------------------------------------------------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|------|
| Fresh homogenate                           | (intragastric for 41 days; 100 mg/kg)                 | Pregnant rat: High cholesterol diet                                                        | On mothers: ↓ systemic inflammation, disruption of mitochondrial network, infiltration of foam cells, TC, TG, LDL and CK  
On offspring: ↓ abnormalities and abortions                                                                 | [62] |
| Homogenate and raw garlic                  | (p.o. for 100 weeks; 0.5% in chow)                    | Rat: High cholesterol diet                                                                 | Homogenate: ↓ TC, LDL and TG  
Raw garlic: ↓ TC, LDL and TG; ↑ excretion of TG and TC                                                                                                          | [63] |
| Aged garlic extract                        | (p.o. for 56 days; 500 mg/kg)                          | Rat: STZ-induced diabetes                                                                  | ↓ Glucose, CK, LDH and AGER gene expression; ↑ Mn-SOD                                                                                                                     | [64] |
| Raw garlic                                 | (p.o. for 4 weeks; 250 mg/kg)                          | Rat: STZ-induced diabetes                                                                  | ↑ Cat, SOD, SIRT3 activity, TFAM and PGC-1α mRNA; ↓ ROS                                                                                                               | [65] |
| Black garlic extract                       | (p.o. for one month; 250 mg/kg)                        | Rat: High fat/sucrose diet                                                                 | ↓ Calory intake, BW, TG, LDL, insulin, leptin and leptin receptor, pro-inflammatory genes; induced vasorelaxation                                                           | [66] |
| Garlic oil                                 | (p.o. for 8 weeks; 1% in chow)                         | Hamster: High cholesterol                                                                  | ↓ Cardiac apoptosis and apoptotic markers; ↑ IGFR/PI3K/Akt pathway                                                                                                  | [67] |
| Garlic oil                                 | (gavage daily for 16 days; 100 mg/kg)                  | Rat: Diabetic cardiomyopathy                                                              | ↓ Cardiac apoptosis and apoptotic markers dependent of death receptor and mitochondria; ↑ IGFR/PI3K/Akt pathway                                           | [68] |
| Aqueous extract (5 mg/mL added to the blood collection tube) |                           | Human: Healthy individuals                                                                | ↓ TC and TG                                                                                                                                                                                                   | [69] |
| Aqueous extract (i.p. for 8 weeks; 100 mg/kg) |                           | Rat: STZ-induced diabetes                                                                  | ↓ STZ-induced vasoconstriction                                                                                                                                       | [70–72] |
| Aqueous extract (p.o. for 16 weeks; 100 mg/kg) |                           | Rat: STZ-induced diabetes                                                                  | ↓ Coronary arterioles thickening and BG; ↑ aortic/coronary blood flow                                                                                                    | [73] |
| Aqueous extract (gavage for 28 days; 2500 and 500 mg/kg) |                           | Rat: Obese and insulin resistant                                                         | ↓ Insulin, BG, and lipid levels; ↑ cardiac function and mitochondrial homeostasis                                                                                            | [74] |
| High pressure garlic extract               | (p.o. for 5 weeks; 2% in chow)                         | Rat: High-fat diet                                                                         | ↓ Plasma and hepatic LDL and TG; ↑ plasma HDL, hepatic mRNA ApoA1, ABCA1 and LCAT                                                                                       | [75] |
| Bulb powder                                | (gavage for 28 days; 200 mg/kg)                        | Rat: STZ/Nicotinamide-induced diabetes                                                     | ↓ Hyperglycaemia, dyslipidaemia, AI and MDA; ↑ Insulin production, GSH activity                                                                                   | [76] |
Table 2. Cont.

| Plant Species | Plant Part or Extract Used (Preparation; Concentration) | Study Model: Insult or Injury | Main Findings | Ref. |
|---------------|----------------------------------------------------------|-------------------------------|---------------|------|
| **Allium ursinum** | Powder (p.o. for 35 days; 300 mg/kg) | Rabbit: HC-induced atherogenesis | ↓ Neointima formation, cholesterol, TG, PL and collagen accumulation; ↓ TG, TC, PL blood levels; ↓ AI | [77] |
| | Powder (n/a) | Rat: In vivo Fe²⁺-induced LDL oxidation | ↓ LDL oxidation and oxLDL mobility; ↓ MDA hepatic, serum and heart levels | [78] |
| **Tulbaghia violacea** | Leaf lyophilizate (p.o. for 8 weeks; 2% in chow) | Rabbit: Hypercholesterolaemic | ↑ Heart function in vivo and ex vivo; ↑ HO-1; ↓ TC, TG, ApoB and atherosclerotic lesions | [79] |
| **Allium cepa** | Raw onion (p.o. for 3 weeks; 5% in chow) | Rat: L-NAME-induced hypertension | ↓ SBP and TBARS; ↑ NO metabolites excretion | [81] |
| **Allium fistulosum** | Raw or boiled juice (cumulative doses from $3 \times 10^{-5}$ to $4 \times 10^{-3}$ g/mL) | Aortic rings: NE precontracted | Raw juice: Induced relaxation; Boiled juice: ↑ EDCF | [82] |
| | Raw green part (p.o. for 4 weeks; 5% in chow) | Rat: HFD-induced hypertension | ↓ SBP, O₂⁻⁻ and NOX activity; ↑ NO levels | [83] |
| **Allium macrostemon** | Volatile extract (cumulative doses from 0.01% to 0.1%) | Pulmonary arteries: Phe contracted | Induced relaxation; ↑ NOS phosphorylation and Ca²⁺ influx to ECs | [84] |
| | Aged garlic extract (gavage for 12 weeks; 2 g/kg) | Rat: Dahl salt-sensitive hypertensive | ↓ LVEDP, pressure half-time, interstitial fibrosis, LV mass and SBP | [85] |
| **Allium sativum** | Aged garlic extract (cumulative doses from 0.001 to 1%) | Aortic rings: NE-contracted | Induced relaxation in a dose-dependent manner | [86] |
| | Fresh homogenate (p.o. for 8 weeks; 250 mg/kg) | Rat: High fructose | ↓ LVH, NF-κB and oxidative stress; ↑ cat, GSH, GPx, and Nrf2 | [87] |
| | Homogenate (p.o. for 3 weeks; 125 and 250 mg/kg) | | ↓ SBP, HR, TC, TG, glucose, LDH, CK-MB; ↑ SOD, cat and heart function | [88] |
| Plant Species      | Plant Part or Extract Used (Preparation; Concentration) | Study Model: Insult or Injury | Main Findings                                                                 | Ref.  |
|-------------------|----------------------------------------------------------|-------------------------------|-------------------------------------------------------------------------------|-------|
| Raw garlic        | (p.o. 1 day or 3 weeks before MCT injection + 3 weeks; 1% in chow) | Rat: MCT-induced PH           | ↓ RVSP, RVH, vasoconstriction in CEC; induced relaxation                      | [89]  |
| Garlic juice (cumulative doses from 1 to 50 µg/mL) | Aortic rings: Phe contracted | Induced relaxation in a dose-dependent manner |                                                                             | [90]  |
| Aqueous extract   | (p.o. for 4 weeks; 50 mg/kg)                             | Rat: 2-kindey-1-clip hypertension | ↓ SBP and ACE activity                                                        | [91]  |
| 100% methanol fraction from a methanolic extract (cumulative doses from 30 to 750 µg/mL) | Aortic rings | Precontracted with KCl or Phe: Induced relaxation | Pre-treatment with the fraction: Prevented contraction evoked by KCl or Phe | [92]  |
| Aqueous extract (0.045 mg/mL) | Aortic rings: NE-contracted | Prolonged relaxation induced by GSNO; Inhibited chloride channels |                                                                     | [93]  |
| Aqueous extract (cumulative doses from 3 to 500 µg/mL) | Pulmonary arteries | Normoxia: Induced dose-dependent relaxation | Hypoxia: Inhibited the transient relaxation and sustained contraction elicited by hypoxia; ↓ ET-1 induced contractions | [94]  |
| Aqueous and 5% ethanol extracts (cumulative doses from 1 to 500 µg/mL) | Atria: Spontaneously or EPI-induced contraction | ↓ Phe-induced contractions; ↑ ACh-induced relaxation |                                                                   | [95]  |
| Aqueous and ethanol extract (non-cumulative doses from 0.1 to 3 mg/L) | | | Negative inotropic and chronotropic effect | [96]  |
| Allium ursinum    | Leaf lyophilizate (p.o. for 8 weeks; 2% in chow)         | Rat: MCT-induced PH           | ↑ RV function and PDE5 activity; ↓ Medial thickness of PA                    | [97]  |
| Tulbaghia violacea| Methanolic extract (i.p. for 7 weeks; 50 mg/kg)          | Rat: Dahl salt-sensitive hypertensive | ↓ SBP; ↑ [Na] in urine and AT-1a receptor levels                         | [98]  |
| Allium cepa       | Raw juice (intragastric intubation for 14 days; 1 mL)   | Rat: DOX-induced cardiotoxicity | ↓ Apoptotic cells; ↓ CK, CK-MB, LDH, cTn1 and MDA levels; ↑ SOD, GSH, GPx | [99]  |
| Plant Species | Plant Part or Extract Used (Preparation; Concentration) | Study Model: Insult or Injury | Main Findings | Ref. |
|---------------|--------------------------------------------------------|-----------------------------|---------------|------|
| **Allium sativum** | Raw juice (intragastric intubation for 14 days; 1 mL) | Rat: DOX-induced endothelial dysfunction | ↓ Apoptotic cells; ↓ MDA levels; ↑ GSH | [100] |
| | Raw juice (intragastric intubation for 8 weeks; 1 mL/100 g BW) | Rat: Cd-induced cardiotoxicity | ↓ Apoptotic cells; ↓ CK, CK-MB, LDH, cTnT and MDA levels; ↑ SOD, GSH, GPx | [101] |
| | Aged garlic extract (1000 µg) | Rat: DOX-induced cardiomyocyte apoptosis | ↓ p53 activation, and caspase-3 activity; ↑ 8-isoprostane levels | [103] |
| | Aged garlic extract (p.o. for 6 days before DOX; 2860 mg/kg) | Mice: DOX-induced cardiotoxicity | ↑ survivability, and tumour uptake of DOX | [103] |
| | Aged garlic extract (p.o. for 28 days; 250 mg/kg) | | ↓ LDH, CK and MDA | [104] |
| | Homogenate (p.o. days; 250 and 500 mg/kg) | Rat: Adriamycin-induced cardiotoxicity | ↑ SOD, GPx, and cat; ↓ MDA, TNF-α accumulation | [105] |
| | Aqueous extract (p.o. for 3 weeks; 250 mg/kg) | Rat: Gentamycin-induced renal failure | ↑ Renal function, BW, HW/BW, cardiac Na⁺/K⁺-ATPase activity, and antioxidant capacity; ↓ BP, LDH, CK-MB, MDA | [106] |
| **Allium ursinum** | Water and methanolic extracts (4 h pre-treatment; 50 µg/mL) | Cardiomyoblasts (H9c2): DOX-induced toxicity | Water: ↓ intracellular and mitochondrial ROS and cell death induced by DOX Methanolic: ↓ intracellular and mitochondrial ROS | [107] |
| **Allium ampeloprasum** | Raw juice (n/a) | Human: Platelet aggregation in whole blood | ↓ Platelet aggregation (IC₅₀ = 114.9 and 117.3 mg/mL) | [108] |
| **Allium ascalonicum** | Heated extract (n/a) | Human: Platelet-rich plasma | ↓ Platelet aggregation, which is lost with higher heating times or microwave heating | [109] |
| **Allium cepa** | Peel aqueous extract (50, 100 and 500 µg/mL) | Rat: Collagen-induced platelet aggregation | ↓ Platelet aggregation, [Ca²⁺]i, TXA₂; ↑ cAMP | [110] |
| | Methanolic extract and methanolic fractions (0.5, 1, 3 and 5 mg/mL) | | ↓ Platelet aggregation | [111] |
| Plant Species | Plant Part or Extract Used (Preparation; Concentration) | Study Model: Insult or Injury | Main Findings | Ref. |
|---------------|--------------------------------------------------------|------------------------------|---------------|-----|
| Allium fistulosum | Raw juice (n/a) | Human: Platelet aggregation in whole blood | ↓ Platelet aggregation (IC$_{50}$ = 46.7 and 116.7 mg/mL) | [108] |
| Allium fistulosum | Raw juice (i.v. after CFR induction; 0.09 ± 0.01 mL/kg) | Dog: Chronic platelet-mediated thrombosis | ↓ Platelet aggregation | [112] |
| Allium fistulosum | Raw homogenate (intragastric after CFR induction; 2 g/kg) | Human and dog: In vitro platelet aggregation | ↓ Platelet aggregation in both blood type, stronger effect on dog | |
| Allium fistulosum | Raw juice (1, 10 and 100 mL/L) | | | |
| Allium fistulosum | Raw juice (n/a) | Human: Platelet aggregation in whole blood | ↓ Platelet aggregation (IC$_{50}$ = 113.8 and 113.2 mg/mL) | [108] |
| Allium fistulosum | Raw juice (p.o. for 4 weeks; 2 g/Kg) | Rat | ↓ SBP, platelet adhesion to fibrinogen, platelet aggregation and thromboxane release; ↑ bleeding time, cAMP and 6-keto prostacyclin F$_{1\alpha}$ | [113] |
| Allium fistulosum | Raw or boiled juice (0–4 mg/mL) | Human: ADP-induced platelet aggregation | Raw juice: ↓ [Ca$^{2+}$]$_i$ and thromboxane production; ↑ cAMP levels Boiled juice: ↑ [Ca$^{2+}$]$_i$ and thromboxane production; induced morphological changes | [114] |
| Allium fistulosum | Aged garlic extract (3.12 to 12.5%) | Human: Fibrinogen- and ADP-induced platelet aggregation | ↓ Platelet adhesion to fibrinogen; Prevented platelet conformational changes induced by ADP; ↑ cAMP | [115] |
| Allium fistulosum | Aged garlic extract (0.78-25%) | Human: ADP-induced platelet aggregation | ↓ Platelet aggregation and [Ca$^{2+}$]$_i$. | [116,117] |
| Allium sativum | Aged garlic extract (p.o. for 7 or 14 days; 1, 2 or 5 g/kg) | Rat: Healthy fed AGE | ↓ Platelet aggregation after 14 days without prolonging bleeding time; ↑ extracellular ATP, TXB$_2$ and ↓ phosphorylation of ERK, p38 and JNK after collagen treatment | [118] |
| Allium sativum | Aged garlic extract (p.o. for 13 weeks; 5 mL) | Human: ADP-induced platelet aggregation (13 days pre-treatment) | ↓ % of aggregated platelets and the initial rate of aggregation | [119] |
| Plant Species          | Plant Part or Extract Used (Preparation; Concentration) | Study Model: Insult or Injury | Main Findings                                                                 | Ref.  |
|------------------------|----------------------------------------------------------|-------------------------------|-------------------------------------------------------------------------------|-------|
|                        |                                                          |                               | ↓ Platelet aggregation; ↑ cGMP and cAMP which were inhibited by ODQ and SQ22536 | [120] |
| Aged garlic extract (0.19–6.25%) | Human: ADP-induced platelet aggregation |                               |                                                                               |       |
| Garlic juice (n/a)     | Human: Platelet aggregation in whole blood                |                               | ↓ Platelet aggregation (IC₅₀ = 3.2 and 4.0 mg/mL)                             | [108] |
| Aqueous and alcoholic extract (n/a) | Human: Platelet-rich plasma |                               | Aqueous: ↓ ADP-induced aggregation                                             | [121] |
| Odourless powder (p.o. for 2 weeks; 1 g/kg in chow) | Rat: In situ loop |                               | ↓ Thrombus formation                                                          | [122] |
| Allium schoenoprasum   | Raw juice (n/a)                                          | Human: Platelet aggregation in whole blood | ↓ Platelet aggregation (IC₅₀ = 45.4 and 50.1 mg/mL)                           | [108] |
| Allium ursinum         | Aqueous extract (n/a)                                    | Rat: Platelet aggregation     | ↓ ADP-, collagen-, AA- and EPI-induced aggregation                             | [121] |
|                        |                                                          |                               | Other activities                                                              |       |
| Allium ampeloprasum    | Aqueous extract (0.045 mg/mL)                            |                               | Induced NO release                                                             | [93]  |
| Allium ascalonicum     | Ethyl acetate fraction from a hydroethanolic extract (500 and 800 ng/mL) | HUVEC: Angiogenesis          | Promoted angiogenesis                                                          | [123] |
| Allium cepa            | Aqueous extract (0.045 mg/mL)                            |                               | Induced NO release                                                             | [93]  |
| Allium sativum         | Aged garlic extract (1–4 mg/mL)                          | HUVEC                         | ↑ HO-1, GCLM and Nrf2 activation                                               | [124] |
|                        | Aged garlic extract (p.o. for 6 weeks; 4% in chow)       | Rat: Folate-deficient diet    | ↓ Homocysteine total, protein-bound and free levels                            | [125] |
|                        | Chloroform extract of aged black garlic (30 min before treatment; 30 µg/mL) | HUVEC: TNF-α                  | ↓ ROS, NF-κB activation, VCAM-1 mRNA and protein expression and THP-1 adhesion to HUVEC | [126] |
|                        | Aqueous extract (0.045 mg/mL)                            |                               | Induced NO release                                                             | [93]  |
Table 2. Cont.

| Plant Species                        | Plant Part or Extract Used (Preparation; concentration) | Study Model: Insult or Injury | Main Findings                                                                 | Ref. |
|--------------------------------------|---------------------------------------------------------|-------------------------------|-------------------------------------------------------------------------------|------|
| Aqueous extract (0.2–1.0%)           | Macrophages/HUVEC: LPS- and IFNγ stimulated             | Macrophages: ↓ iNOS expression | HUVEC: ↑ eNOS activity and cGMP levels                                        | [127]|
| Garlic skin or flesh extract (1, 2.5 and 5 µL/mL) | Cardiomyocyte: NE-induced hypertrophy                    | ↓ Cell hypertrophy, cell death, apoptosis, and oxidative stress |                                                                 | [128]|
| Aqueous extract (p.o. for 5 weeks; 800 µg/Kg) | Rabbit: Vascular restenosis                             | ↓ Myointimal hyperplasia      |                                                                 | [129]|
| Hydroalcoholic extract (1, 10, 50, and 100 µg/mL) | Mice: LPS-stimulated heart                              | ↓ PGE2 and 8-iso-PGF2α levels; ↓ COX2, IL-6 and NF-κB mRNA |                                                                 | [130]|
| Aqueous powder (4 days before treatment; 0.25–4.0 mg/mL) | CAEC: IL-1α                                            | ↓ ICAM-1, VCAM-1 and monocyte adhesion to ECs |                                                                 | [131]|

↑—Increase; ↓—Decrease; +dp/dt—Ratio of pressure change in the ventricular cavity during the isovolumetric contraction period; 8-iso-PGF2α—8-iso-Prostaglandin F2α; AA—Arachidonic acid; ABCA1—ATP-binding cassette transporter; ACE—Angiotensin-converting enzyme; Ach—Acetylcholine; ADP—Adenosine diphosphate; AGE—Advanced glycation end products; AGER—Advanced glycation end products receptor; AI—Atherogenic index; Akt—Akt Murine thymoma viral oncogene/Protein kinase-B; ALP—Alkaline phosphatase; ALT—Alanine aminotransferase; ApoA1—Apolipoprotein A1; ApoB—Apolipoprotein B; ApoE—Apolipoprotein E; AST—Aspartate aminotransferase; AT-1a—Type 1A angiotensin II receptor; ATP—Adenosine triphosphate; BG—Blood glucose; BW—Body weight; Ca²⁺—Calcium; CAEC—Coronary artery endothelial cells; CAMP—Cyclic adenosine monophosphate; Cat—Catalase; Cd—Cadmium; CEC—Coronary endothelial cells; cGMP—Cyclic guanosine monophosphate; CK—Creatine kinase; CK-MB—Creatine kinase—muscle/brain isoform; COX2—Cyclooxygenase-2; CRF—Cyclic flow reduction cTnI—Cardiac troponin T; Cu²⁺—Copper (II); DEX—Dexamethasone; DOX—Doxorubicin; EC—Endothelial cell; EDCF—Endothelium-derived contracting factor; eNOS—Endothelial nitric oxide synthase; EPI—Epinephrine; ERK—Extracellular signal-regulated kinases; ET-1—Endothelin-1; Fe²⁺—Iron (II); GCLM—Glutamate-cysteine ligase modifier subunit; GSH—Glutathione; GSNO—S-Nitrosoglutathione; GSR—Glutathione reductase; γ-δ—High cholesterol; HDL—High-density lipoprotein; HFD—High-fat diet; HMG-CoA—β-Hydroxy β-methylglutaryl-Coenzyme A; HO-1—Hemeoxygenase-1; HR—Heart rate; HUVEC—Human umbilical cord vein endothelial cell; HW/BW—Heart weight/body weight ratio; I/R—Ischaemia/Reperfusion; IC₅₀—Concentration needed to inhibit 50% of the enzyme activity; ICAM-1—Intercellular adhesion molecule 1; IFNγ—Interferon gamma; IGFR—Insulin-like growth factor 1 receptor; i.p.—Intraperitoneal injection; IL-1α—Interleukin-1 α; IL-6—Interleukin-6; iNOS—Inducible nitric oxide synthase; IS—Isoproterenol; JNK—c-Jun N-terminal kinase; KCl—Potassium chloride; LAD—Left anterior descending; LCAT—Lecithin-cholesterol acyltransferase; LDL—Lactate dehydrogenase; LDLR—Low-density lipoprotein receptor; L-NAME—Nω-Nitro-L-arginine methyl ester; LPS—Lipopolysaccharide; LV—Left ventricle; LV/BW—Left ventricle weight/body weight ratio; LVDP—Left ventricle diastolic pressure; 8-iso-PGF2α—8-iso-Prostaglandin F2α; MCT—Monocrotaline; MDA—Malondialdehyde; MCT—Monocrotaline; MDA—Malondialdehyde; Mn-SOD—Manganese superoxide dismutase; mRNA—Messenger RNA; n/a—Not available; Na—Sodium; Nε—Nε,Nε′-Diallyl-lysine; NADPH—Nicotinamide adenine dinucleotide phosphate oxidase; Nrf2—Nuclear factor erythroid 2-related factor 2; O₂—Superoxide; ODQ—[1H-[1,2,4]oxadiazolo-[4,3-a]quinazolin-1-one]; oxLDL—Oxidised low-density lipoprotein; p38—p38 Mitogen-activated protein kinases; PA—Pulmonary artery; PDES—Phosphodiesterase type 5; PGC-1α—Peroxisome proliferator-activated receptor-gamma coactivator-1α; PGF₂α—Prostaglandin E2; PH—Pulmonary hypertension; Phe—Phenylephrine; PFK—Phosphofructokinase 3-kinase; PL—Phospholipid; p.o.—Per os (orally); PPR—Platelet-rich plasma; ROS—Reactive oxygen species; RPP—Rate pressure product; RV—Right ventricle; RVH—Right ventricle hypertrophy; RVSP—Right ventricle systolic pressure; SBP—Systolic blood pressure; SIRT3—Sirtuin 3; SOD—Superoxide dismutase; SQ22536—Inhibitor of adenylyl cyclase; STZ—Streptozotocin; T2DM—Type 2 diabetes mellitus; TBARS—Thiobarbituric acid reactive substances; TC—Total cholesterol; TFAM—Mitochondrial transcription factor A; TG—Triglycerides; TGF—Transforming growth factor; THP-1—Spontaneously immortalised monocyte-like cell line; TNF-α—Tumour necrosis factor alpha; TXA₂—Thromboxane A2; TXB₂—Thromboxane B2; VCAM-1—Vascular cell adhesion protein 1; VLDL—Very low-density lipoprotein; ZO-1—Zonula occludens-1.
2.2.1. The Effect of Plant Parts or Extracts

In Table 2, studies reporting the beneficial effects of plant parts or extracts is presented with reference to the species name, the plant part/extract used (with reference to the preparation method and concentration), the study model and the main findings regarding the effect observed in the cardiovascular system. Unless stated, a daily administration was used. Studies are grouped considering the cardiovascular disease and/or risk factor assessed with plants organised in alphabetical order of their scientific name. A list of abbreviations, used throughout the table, is provided at the end of the table.

Plants from this subfamily are rich in cysteine sulfoxide derivatives, such as alliin [132], which, by the action of alliinase, are converted into thiosulfimates, e.g., allicin, which in turn are instable and change into organosulfur compounds like ajoene [133]. Thiosulfimates are considered to be the main class of compounds responsible for the biological activities reported for plants from Allioideae subfamily [134]. Accordingly, several studies have assessed the role of alliinase activity on the effect of the extracts. Indeed, the antihypertensive effect of onions (Allium cepa) is lost or is much weaker upon boiling [81]. Similarly, the antiplatelet aggregation potential of these bulbs is also compromised, since longer heating times in either a conventional oven or microwave led to a pro-aggregatory effect rather than the expected anti-aggregatory potential [109]. Additionally, with the loss of alliinase activity, the vasorelaxant properties of Allium sativum, were abolished in aortic rings pre-contracted with phenylephrine [95]. Similar dependency on alliinase activity was reported for the hypolipidemic activity of A. sativum where long heating times or microwave heating compromised this effect [135].

On the other hand, the antidysslipidaemic and antidiabetic effects of A. sativum seem to depend on the PI3K/Akt/Nrf2 [87] or IGFIR/PI3K/Akt [67,68] pathways since, upon treatment, activation of these pathways is observed.

In order to better disclose the putative factors underlying the hypolipidemic effect of A. hookeri, a metabolomic analysis on the serum of hamsters consuming a high-fat diet and administered A. hookeri powder orally was carried out. The authors found 25 putative markers which could explain the lipid-lowering effect of this species, with phosphatidylcholines, lysophosphatidylcholines and lysophosphatidylethanolamines the most common targets. Furthermore, the metabolism for glycerophospholipids was increased in the treated group [57].

2.2.2. The Effect of Isolated Sulphur-Containing Compounds

In this section, the effect of isolated sulphur-containing compounds found in the Allioideae subfamily is presented. Then, a composition–activity relation is discussed in order to bring attention to potential active extracts. Table 3 systematises the main studies performed in these compounds, with the compound name, chemical structure, study model used, and the main findings of the study pointed out. Additionally, whenever reported, the route of administration and concentration used is highlighted. A list of abbreviations, used throughout the table, is provided at the end of the table.

### Table 3. Effects of sulphur-containing compounds on the cardiovascular system.

| Compound | Study Model: Insult or Injury (Route of Administration; Concentration) | Main Findings | Ref. |
|----------|------------------------------------------------------------------------|---------------|------|
| Ajoene   | Smooth muscle cells (1–50 μM)                                         | ↓ Proliferation, cholesterol biosynthesis | [136] |
| Allicin  | Mice: ApoE-deficient and LDLR-deficient (p.o.; 9 mg/kg)               | ↓ Atherosclerotic plaque, uptake and degradation of oxLDL by macrophages | [137] |
|          | HUVEC: oxLDL-induced damage (10, 30, 100 μM)                          | ↓ Apoptosis   | [138] |
### Table 3. Cont.

| Compound | Study Model: Insult or Injury (Route of Administration; Concentration) | Main Findings | Ref. |
|----------|-------------------------------------------------|----------------|------|
| **Alliin** | Rat: High fructose (p.o. for 3 weeks; 0.111 and 0.222 mg/kg) | ↑ Heart function; ↓ SBP and TG | [139] |
| **Diallyl disulphide** | Rat: ISO-induced myocardial infarction (gastric intubation for 35 days; 40 and 80 mg/kg) | ↓ CK, CK-MB, LDH, ALT, AST, TC, LDL, VLDL, TG, FFA, PL, MDA levels, HMGR activity; ↑ HDL levels, LCAT activity | [140] |
| **Diallyl trisulphide** | HEK293 cells: Whole cell patch clamp (n/a) | ↓ IKr and hERG channel trafficking | [146] |

Note: The table continues with similar entries for various compounds and study models, including changes in concentrations and related biological effects.
Although the compounds presented in Table 3 are commonly found in plants from the Allioideae subfamily, there are others that, despite being found in lower amounts, have been studied for their potential bioactivity. These compounds are shown to exert various effects, including antioxidative, anti-inflammatory, and anti-apoptotic properties, in cell culture and animal models. The mechanism of action underlying these effects is complex and involves various pathways and targets, as detailed in the table below.

Despite the interest in these compounds and their potential, only one study from those listed in Table 3 focused on the mechanisms of action underlying the observed effects. Indeed, it was shown that allicin reduced oxidised low-density lipoprotein-induced damage by inhibiting apoptosis and decreasing oxidative stress [138].

Although the compounds presented in Table 3 are commonly found in plants from the Allioideae subfamily, there are others that, despite being found in lower amounts, have...
been assessed for their cardioprotective effect. For example, the antidyslipidaemic effects reported for garlic (*A. sativum*) seem to be due to the capacity of S-allyl cysteine, N-acetyl-S-allyl cysteine, alliin, allixin, and allylmercaptocysteine to suppress low-density lipoprotein oxidation since all these compounds were able to reduce LDL oxidation induced by copper (II) [60]. Additionally, S-methylcysteine sulfoxide in high cholesterol-fed rats, was able to reduce the levels of total cholesterol, triglycerides and phospholipids. Furthermore, this compound reduced the activity of lipoprotein lipase without affecting the activity of other lipogenic proteins, while decreasing the levels of free fatty acids. In addition, the excretion of bile acids and sterols was enhanced in the treated group [154].

Furthermore, the antiplatelet activity of aged garlic extract was related to the presence of S-ethylcysteine, S-methyl-L-cysteine, S-propionyl-L-cysteine, since the remaining constituents of the extract (alliin, cycloalliin, S-allyl-L-cysteine, S-allylmercapto-L-cysteine, and fructosyl-arginine) failed to significantly inhibit platelet aggregation [116]. Moreover, two compounds, sodium n-propyl thiosulfate and sodium 2-propenyl thiosulfate decreased adenosine diphosphate-induced platelet aggregation in both dogs and human blood [155].

Regarding the vascular protective effect of garlic, it seems that allithiamine (vitamin B analogue found in garlic) might play a relevant role. Indeed, the presence of this compound in HUVEC growing in high glucose conditions showed a lower level of advanced glycation end products as well as a lower inflammatory profile when compared to high glucose-only treated cells. In addition, this compound also showed a very potent antioxidant potential [156]. Moreover, 2-vinyl-4H-1,3-dithin, an organosulfur compound found in macerated garlic oil or in stir-fried garlic, decreased spontaneously hypertensive rat’s vascular smooth muscle cells proliferation and cell migration and arrested cell cycle at G2 phase. Furthermore, it decreased reactive oxygen species production induced by angiotensin II [157]. Also, diallyl disulphide and diallyl trisulphide have been reported for their capacity to induce neovascularogenesis via PI3K/Akt pathway activation [68,141]. In addition, reduction of cell death dependent on death receptor and mitochondria is also reported for both compounds [68]. Furthermore, for diallyl trisulphide, the promotion of neovascularogenesis is also attributed to a decrease in the microRNA 221 [68,141]. This compound also activated Nrf2 via the PI3K/Akt pathway [147] and induced the release of hydrogen sulphide by cystathionine-γ-lyase [148] using in vitro conditions mimicking diabetes. The reported effects for ajoene might be due to its capacity to inhibit protein prenylation, particularly that dependent on protein farnesyltransferase and protein geranylgeranyltransferase type I [136].

Some studies also assessed the activity of synthetic derivatives of naturally occurring sulphur-containing compounds. A study compared the antihypercholesterolaemic properties of diallyl disulphide analogues and showed that all the tested analogues lowered serum and hepatic levels of several lipids, including low-density lipoprotein while increasing those of high-density lipoprotein. The authors suggested that this lipid-lowering effect is due to the modulation of the 3-hydroxy-3-methylglutaryl-CoA reductase activity since a decrease in mRNA levels with a concomitant inactivation of sterol regulatory element-binding protein-2 and cyclic adenosine monophosphate response element-binding protein is observed [158]. Another study assessed the antihypertensive and vasorelaxant properties of five synthetic derivatives of diallyl disulphide. The results showed that all analogues were able to decrease systolic blood pressure in the Nω-nitro-L-arginine methyl ester-induced hypertensive animal model. Similarly, all compounds restored the antioxidant defences as observed by an increase in the activity of glutathione peroxidase, glutathione and superoxide dismutase with concomitant decrease in malondialdehyde and protein carbonyl levels. Furthermore, nitric oxide metabolites and cyclic guanosine monophosphate levels were restored by all the analogues, while the activity of angiotensin-convertase enzyme was decreased [159].

The effect of sulphur-containing compounds on the pharmacodynamic and pharmacokinetics of other drugs was also assessed. Indeed, it was reported that the oral co-consumption of diallyl trisulphide and nifedipine led to a higher maximum concen-
tration and area under the curve, thus suggesting that the compound might affect the gastrointestinal metabolism of nifedipine, since no effect on the pharmacokinetics was observed when nifedipine was given intravenously [160].

2.2.3. Clinical Trials

The importance of plants from the Allioideae subfamily is also validated by a small number of clinical trials. However, some contradictory results have been reported that may be related to the different doses used, duration of the treatment and/or association with other compounds. For example, in a small placebo-controlled and double-blind trial, firefighters were given four tablets containing 300 mg/table of aged garlic extract and 30 mg/table of coenzyme Q10 for up to 1 year. The results showed that the consumption improved their vascular elasticity and endothelial function [161]. In another study, the consumption of 1200 mg of this extract daily for 4 weeks followed by 4 weeks of washout had no effect on several parameters assessed such as glycated haemoglobin A1c, blood pressure, total cholesterol, triglycerides and high-density lipoprotein, and did not prevent endothelial dysfunction, oxidative stress or inflammation in patients with type 2 diabetes with high cardiovascular risk [162]. Furthermore, the administration of aged garlic extract (250 mg) supplemented with vitamins B12 and B6, folic acid and L-arginine daily for a 12-month period increased the ratio between brown and white epicardial adipose tissues with concomitant increase in the temperature-rebound index, while decreasing homocysteine levels and preventing the progression of coronary artery calcification [163]. In patients with coronary artery calcification and increased cardiovascular disease risk, the consumption of 2400 mg of aged garlic extract daily for 1 year inhibited the progression of the calcification. Regarding secondary outcomes, the extract decreased interleukin-6 levels as well as the glucose levels and blood pressure [164]. The same concentration increased cutaneous microcirculation in diabetic patients, thus suggesting that aged garlic extract might promote wound healing in these patients [165]. Overall, it seems that longer treatment durations (up to 1 year) have better outcomes.

Regarding other extracts, the consumption of 125 mL of red wine extract of onion twice daily for 10 weeks by healthy individuals showed hypocholesterolaemic, antioxidant and anti-inflammatory effects [166]. Additionally, the consumption of 300 mg of *A. sativum* standardised powder for 8 weeks by patients undergoing haemodialysis decreased the absolute values for oxidised low-density lipoprotein and homocysteine. In addition, the powder significantly ameliorated the values of calcium, triglycerides, oxidised low-density lipoprotein and homocysteine [167]. The consumption of quercetin-rich *A. cepa* extract daily for 6 weeks decreased systolic blood pressure in hypertensive individuals when compared to the placebo group [168]. The consumption of *A. cepa* peel extract twice daily for 12 weeks improved the flow-mediated dilation as well as the number of circulating endothelial progenitor cells in healthy overweight and obese patients. Indeed, the rate of patients with endothelial dysfunction decreased from 26% to 9% after extract administration [169].

Concerning CVD risk factors, some studies have been performed, such as the Tehran Lipid and Glucose study that assessed the effect of dietary consumption of *A. sativum* and *A. cepa* in cardiometabolic risk factors (body mass index, waist circumference, systolic blood pressure, diastolic blood pressure, fasting plasma glucose, triglycerides-to-high-density lipoprotein ratio, insulin, creatinine, estimated glomerular filtration rate and creatinine clearance) for 6 years. The results showed that high consumption of these vegetables led to a 64% reduction in CVD outcomes, as well as a lower incidence of chronic kidney disease and hypertension while no association was made with type 2 diabetes. Furthermore, it improved TG levels and creatine clearance [170].

Although the majority of the reported clinical trials are conducted using a small cohort of patients and are usually single-centre studies, they highlight the potential of plants from the Allioideae subfamily in the management of CVDs and associated risk factors. Nevertheless, these effects should be validated in more complete clinical trials with access.
to bigger multicentre cohorts to account for the genetic polymorphisms which impact the activity of drug metabolising enzymes leading to altered pharmacokinetics [171].

Drug interactions between conventional drugs or between these and herbal medicines are common, with both beneficial and detrimental effects reported. For example, the consumption of capsules containing 0.5 g of *A. macrostemon* bulb extract powder (three times a day) for eight weeks by patients undergoing baseline therapy for unstable angina, led to lower oxidised low-density lipoprotein and plasminogen activator inhibitor-1 level, while increasing plasminogen activity [172]. Moreover, in patients undergoing simvastatin therapy, supplementation with fenugreek and garlic for 8 weeks significantly reduced total cholesterol, triglycerides, non-high-density lipoprotein and low-density lipoprotein levels and increased those of high-density lipoprotein [173]. On the other hand, care must be taken with antiplatelet drugs, particularly warfarin and aspirin, as a simultaneous consumption of garlic or onion with these drugs can increase the risk of bleeding [174,175]. This interaction is attributed to their capacity to decrease platelet adhesion and aggregation, by inhibiting plasminogen activating factor and fibrinogen receptors and by decreasing thromboxane X

3. Final Remarks

The present review sheds light on the potential of plants from the Allioideae subfamily in the management of CVDs and associated risk factors. Traditional uses of some of these species are widely recognised, with garlic (*Allium sativum*) and onions (*Allium cepa*) being the most common. Additionally, pre-clinical studies and clinical trials validating their beneficial potential are frequent, thus confirming their importance. Nevertheless, other species such as *A. jacquermontii*, *A. rotundum* and *Tulbaghia alliacea*, despite being used in traditional remedies in some regions, lack scientific validation while other plants have undergone clinical trials but with no beneficial effects on the cardiovascular system.

Regarding CVD risk factors, plants from the Allioideae subfamily showed promising antiplatelet aggregation, antidiabetic, and dyslipidaemic effects, and were able to exert protection against atherosclerotic events.

Overall, we gathered information on both the tapped and untapped potential of plants belonging to the Allioideae subfamily, by highlighting scientific gaps as well as well-validated effects that pave the way for the development of new preventive/therapeutic approaches for CVDs.

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52. Ostrowska, E.; Gabler, N.K.; Sterling, S.J.; Tatham, B.G.; Jones, R.B.; Eagling, D.R.; Jois, M.; Dunshea, F.R. Consumption of Brown Onions (Allium Cepa Var. Cavalier and Var. Destiny) Moderately Modulates Blood Lipids, Haematological and Haemostatic Variables in Healthy Pigs. *Br. J. Nutr.* **2004**, *91*, 211–218. [CrossRef]

53. Safaeian, L.; Zolfaghari, B.; Karimi, S.; Talebi, A.; Ghazvini, M. The Effects of Hydroalcoholic Extract of Allium Elburzense Wendelbo Bulb on Dexamethasone-Induced Dyslipidemia, Hyperglycemia, and Oxidative Stress in Rats. *Res. Pharm. Sci.* **2018**, *13*, 22. [CrossRef]

54. Janahmadi, Z.; Nekoeoeian, A.A.; Mozafari, M. Hydroalcoholic Extract of Allium Hookeri in Rats Fed with a High Fat Diet. *Food Chem.* **2019**, *280*, 165–175. [CrossRef]

55. Khaleghi, S.K. Ethyl Acetate Fraction of Allium Hirtifolium Improves Functional Parameters of Isolated Hearts of Diabetic Rats. *Korean J. Community Living Sci.* **2017**, *11*, 452. [CrossRef] [PubMed]

56. Jang, G.-J.; Sung, M.J.; Hur, H.J.; Yoo, M.; Choi, J.H.; Hwang, I.K.; Lee, S. Metabolomics Analysis of the Lipid-Regulating Effect of Allium hookeri leaf extract in high-fat-diet-fed mice. *J. Nutr.* **2015**, *145*, 1363-1371. [CrossRef] [PubMed]

57. Lee, S.; Joo, H.; Kim, I.-H.; Kim, Y. High Hydrostatic Pressure Extract of Garlic Increases the HDL Cholesterol Level via Up-Regulation of Apolipoprotein A-I Gene Expression in Rats Fed a High-Fat Diet. *Lipids Health Dis.* **2012**, *11*, 1–7. [CrossRef] [PubMed]

58. Madkor, H.R.; Mansour, S.W.; Ramadan, G. Modulatory Effects of Garlic, Ginger, Turmeric and Their Mixture on Hyperglycaemia, Dyslipidaemia and Oxidative Stress in Streptozotocin–Nicotinamide Diabetic Rats. *Br. J. Nutr.* **2011**, *105*, 1210–1217. [CrossRef]
Plants 2022, 11, 1920

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77. Sobenin, I.A.; Andrianova, I.V.; Lakunin, K.Y.; Karagodin, V.P.; Bobryshev, Y.V.; Orekhov, A.N. Anti-Atherosclerotic Effects of Garlic Preparation in Freeze Injury Model of Atherosclerosis in Cholesterol-Fed Rabbits. *Phytomedicine* 2016, 23, 1235–1239. [CrossRef]

78. Mukthamba, P.; Srinivasan, K. Protective Effect of Dietary Fenugreek (*Trigonella foenum-graecum*) Seeds and Garlic (*Allium sativum*) on Induced Oxidation of Low-Density Lipoprotein in Rats. *J. Basic Clin. Physiol. Pharmacol.* 2016, 27, 39–47. [CrossRef] [PubMed]

79. Bombicz, M.; Prikzsz, D.; Varga, B.; Gesztesy, R.; Kertesz, A.; Lengyel, P.; Balogh, P.; Csupor, D.; Hohmann, J.; Bhattoo, H.; et al. Anti-Atherogenic Properties of Allium Ursinum Lipolyhalisate: Impact on Lipoprotein Homeostasis and Cardiac Biomarkers in Hypercholesterolemic Rabbits. *Int. J. Mol. Sci.* 2016, 17, 1284. [CrossRef] [PubMed]

80. Olorunnisola, O.S.; Bradley, G.; Afolayan, A.J. Protective Effect of Tulbaghia Violacea Harv. on Aortical Pathology, Tissue Antioxidant Enzymes and Liver Damage in Diet-Induced Atherosclerotic Rats. *Int. J. Mol. Sci.* 2017, 13, 12747–12760. [CrossRef]

81. Kawamoto, E.; Sakai, Y.; Okamura, Y.; Yamamoto, Y. Effects of Boiling on the Antihypertensive and Antioxidant Activities of Onion. *J. Nutr. Sci. Vitamino.* 2004, 50, 171–176. [CrossRef]

82. Chen, J.-H.; Tsai, S.-J.; Chen, H. Welsh Onion (*Allium fistulosum* L.) Extracts Alter Vascular Responses in Rat Aortae. *J. Cardiovasc. Pharmacol.* 1999, 33, 515–520. [CrossRef]

83. Yamamoto, Y.; Aoyama, S.; Hamaguchi, N.; Rhi, G.-S. Antioxidative and Antihypertensive Effects of Welsh Onion on Rats Fed with a High-Fat High-Sucrose Diet. *Biosci. Biotechnol.Biochem.* 2005, 69, 1311–1317. [CrossRef] [PubMed]

84. Han, C.; Qi, J.; Gao, S.; Li, C.; Ma, Y.; Wang, J.; Bai, Y.; Zheng, X. Vasodilation Effect of Volatile Oil from Allium Macrostemon Bunge Are Mediated by PKA/NO Pathway and Its Constituent Dimethyl Disulfide in Isolated Rat Pulmonary Arterial. *Fito terapia 2017*, 120, 52–57. [CrossRef] [PubMed]

85. Hara, Y.; Noda, A.; Miyata, S.; Minoshima, M.; Sugiiura, M.; Kojima, J.; Otake, M.; Furukawa, M.; Cheng, X.W.; Nagata, K.; et al. Effects of Aged Garlic Extract on Left Ventricular Diastolic Function and Fibrosis in a Rat Hypertension Model. *Exp. Anim.* 2013, 62, 305–310. [CrossRef] [PubMed]

86. Takashima, M.; Kanamori, Y.; Kodera, Y.; Morihara, N.; Tamura, K. Aged Garlic Extract Exerts Endothelium-Dependent Vasorelaxant Effect on Rat Aorta by Increasing Nitric Oxide Production. *Phytomedicine* 2017, 24, 56–61. [CrossRef] [PubMed]

87. Padiya, R.; Chowdhury, D.; Borkar, R.; Srinivas, R.; Pal Bhadra, M.; Banerjee, S.K. Garlic Attenuates Cardiac Oxidative Stress via Activation of PI3K/AKT/Nrf2-Keap1 Pathway in Fructose-Fed Diabetic Rat. *PLoS ONE* 2014, 9, e94228. [CrossRef]

88. Asdaq, S.M.; Inamdar, M.N. Potential of Garlic and Its Active Constituent, S-Allyl Cysteine, as Antihypertensive and Cardio protective in Presence of Captopril. *Phytomedicine 2010*, 17, 1016–1026. [CrossRef] [PubMed]

89. Sun, X.; Ku, D.D. Allium in Garlic Protects against Coronary Endothelial Dysfunction and Right Heart Hypertrophy in Pulmonary Hypertensive Rats. *Am. J. Physiol.-Heart Circ. Physiol.* 2006, 291, H2431–H2438. [CrossRef] [PubMed]

90. Ashraf, M.Z.; Hussain, M.E.; Fahim, M. Endothelium Mediated Vasorelaxant Response of Garlic in Isolated Rat Aorta: Role of Nitric Oxide. *J. Ethnopharmacol.* 2004, 90, 5–9. [CrossRef] [PubMed]

91. Sharifi, A.M.; Darabi, R.; Akbarloo, N. Investigation of Antihypertensive Mechanism of Garlic in 2K1C Hypertensive Rat. *J. Ethnopharmacol.* 2003, 86, 219–224. [CrossRef]

92. Ganado, P.; Sanz, M.; Padilla, E.; Tejerina, T. An In Vitro Study of Different Extracts and Fractions of *Allium Sativum* (Garlic): Vascular Reactivity. *J. Pharmacol. Sci.* 2004, 94, 434–442. [CrossRef]

93. Grman, M.; Misak, A.; Cacanyiova, S.; Kristek, F.; Tomaskova, Z.; Bertova, A.; Ondrias, K. The Aqueous Garlic, Onion and Leek Extracts Release Nitric Oxide from S-Nitrosoglutathione and Prolong Relaxation of Aortic Rings. *Gen. Physiol. Biophys.* 2012, 30, 396–402. [CrossRef]

94. Kim-Park, S.; Ku, D.D. Garlic Elicits A Nitric Oxide-Dependent Relaxation And Inhibits Hypoxic Pulmonary Vasoconstriction In Rats. *Clin. Exp. Pharmacol. Physiol.* 2000, 27, 780–786. [CrossRef]

95. Ku, D.D.; Abdel-Razeq, T.T.; Dai, J.; Kim-Park, S.; Fallon, M.B.; Abrams, G.A. Garlic And Its Active Metabolite Allicin Produce Endothelium- And Nitric Oxide-Dependent Relaxation In Rat Pulmonary Arteries. *Clin. Exp. Pharmacol. Physiol.* 2002, 29, 84–91. [CrossRef]

96. Radenkovic, M.; Brankovic, S.; Kitic, D.; Veljkovic, S.; Ivetic, V.; Nesić, M.; Miladinovic, B. Inhibitory Effect of Aqueous and Ethanolic Garlic (*Allium Sativum L.*, *Liliaceae*) Extracts on the Rat Atria. *Clin. Exp. Hypertens.* 2010, 32, 251–255. [CrossRef] [PubMed]

97. Bombicz, M.; Prikzsz, D.; Varga, B.; Kurucz, A.; Kertész, A.; Takacs, A.; Posa, A.; Kiss, R.; Szilvassy, Z.; Juhasz, B. A Novel Therapeutic Approach in the Treatment of Pulmonary Arterial Hypertension: Allium Ursinum Lipolyphate Alleviates Symptoms Comparably to Sildenafil. *Int. J. Mol. Sci.* 2017, 18, 1436. [CrossRef]

98. Moodley, K.; Mackraj, I.; Naidoo, Y. Cardiovascular Effects of Tulbaghia Violacea Harv. (Alliaceae) Root Methanolic Extract in Dahl Salt-Sensitive (DSS) Rats. *J. Nutr. Sci. Vitaminol.* 2013, 59, 364–369. [CrossRef] [PubMed]

99. Alpsoy, S.; Aktas, C.; Uygun, R.; Topcu, B.; Kanter, M.; Erboga, M.; Karakaya, O.; Gedikbasi, A. Antioxidant and Anti-apoptotic Effects of Onion (*Allium cepa*) Extract on Doxorubicin-Induced Cardiotoxicity in Rats. *J. Appl. Toxicol.* 2013, 33, 202–208. [CrossRef] [PubMed]

100. Alpsoy, S.; Uygun, R.; Aktas, C.; Topcu, B.; Kanter, M.; Erboga, M.; Karakaya, O.; Gedikbasi, A. The Effects of Onion (*Allium cepa*) Extract on Doxorubicin-Induced Apoptosis in Aortic Endothelial Cells. *J. Appl. Toxicol.* 2013, 33, 364–369. [CrossRef] [PubMed]
26 of 29

Plants 2022, 11, 1920

102. Ige, S.F.; Akhigbe, R.E. Common Onion (Allium cepa) Extract Reverses Cadmium-Induced Organ Toxicity and Dyslipidaemia via Redox Alteration in Rats. Pathophysiology 2013, 20, 269–274. [CrossRef]

103. Alkreathy, H.M.; Damanhoury, Z.A.; Ahmed, N.; Slevin, M.; Osman, A.-M.M. Mechanisms of Cardioprotective Effect of Aged Garlic Extract Against Doxorubicin-Induced Cardiotoxicity. Int. J. Cancer Ther. 2012, 11, 364–370. [CrossRef]

104. Alkreathy, H.; Damanhoury, Z.A.; Ahmed, N.; Slevin, M.; Ali, S.S.; Osman, A.-M.M. Aged Garlic Extract Protects against Doxorubicin-Induced Cardiotoxicity in Rats. Food Chem. Toxicol. 2010, 48, 951–956. [CrossRef]

105. Mukherjee, S.; Banerjee, S.K.; Maulik, M.; Dinda, A.K.; Talwar, K.K.; Maulik, S.K. Protection against Acute Adriamycin-Induced Cardiotoxicity by Garlic: Role of Endogenous Antioxidants and Inhibition of TNF-α-Alpha Expression. BMC Pharm. 2003, 3, 15. [CrossRef]

106. Gomaa, A.M.S.; Abdelhafez, A.T.; Aamer, H.A. Garlic (Allium sativum) Exhibits a Cardioprotective Effect in Experimental Chronic Renal Failure Rat Model by Reducing Oxidative Stress and Controlling Cardiac Na+/K+-ATPase Activity and Ca²⁺ Levels. Cell Stress Chaperones 2018, 23, 913–920. [CrossRef]

107. Pop, R.M.; Bocsan, I.C.; Buzoianu, A.D.; Chedea, V.S.; Socaci, S.A.; Pecoraro, M.; Popolo, A. Evaluation of the Antioxidant Activity of Nigella Sativa L. and Allium Ursinum Extracts in a Cellular Model of Doxorubicin-Induced Cardiotoxicity. Molecules 2020, 25, 5259. [CrossRef]

108. Beretta, H.V.; Bannoud, F.; Insani, M.; Berli, F.; Hirschegger, P.; Galmarini, C.R.; Cavagnaro, P.F. Relationships Between Bioactive Compound Content and the Antiplatelet and Antioxidant Activities of Six Allium Vegetable Species. Food Technol. Biotechnol. 2017, 55, 266–273. [CrossRef] [PubMed]

109. Cavagnaro, P.F.; Galmarini, C.R. Effect of Processing and Cooking Conditions on Onion (Allium cepa L.) Extracts. J. Agric. Food Chem. 2012, 60, 8731–8737. [CrossRef] [PubMed]

110. Ro, J.-Y.; Ryu, J.-H.; Park, H.-J.; Cho, H.-J. Onion (Allium cepa L.) Peel Extract Has Anti-Platelet Effects in Rat Platelets. Springerplus 2015, 4, 1–8. [CrossRef] [PubMed]

111. Ko, E.Y.; Nile, S.H.; Jung, Y.-S.; Keum, Y.S. Antioxidant and Antiplatelet Potential of Different Methanol Fractions and Flavoronols Extracted from Onion (Allium cepa L.). 3 Biotech 2018, 8, 1–10. [CrossRef] [PubMed]

112. Briggs, W.H.; Folts, J.D.; Osman, H.E.; Goldman, I.L. Administration of Raw Onion Inhibits Platelet-Mediated Thrombosis in Dogs. J. Nutr. 2001, 131, 2619–2622. [CrossRef]

113. Chen, J.-H.; Chen, H.; Tsai, S.-J.; Jen, C.J. Chronic Consumption of Raw But Not Boiled Welsh Onion Juice Inhibits Rat Platelet Function. J. Nutr. 2000, 130, 34–37. [CrossRef]

114. Chen, J.-H.; Chen, H.; Wang, J.-S.; Tsai, S.-J.; Jen, C.J. Effects of Welsh Onion Extracts on Human Platelet Function in Vitro. Life Sci. 2000, 66, 1571–1579. [CrossRef]

115. Allison, G.L.; Lowe, G.M.; Rahman, K. Aged Garlic Extract Inhibits Platelet Activation by Increasing Intracellular CAMP and Reducing the Interaction of GP IIb/IIIa Receptor with Fibrinogen. Life Sci. 2012, 91, 1275–1280. [CrossRef]

116. Allison, G.L.; Lowe, G.M.; Rahman, K. Aged Garlic Extract and Its Constituents Inhibit Platelet Aggregation through Multiple Mechanisms. J. Nutr. 2006, 136, 782S–788S. [CrossRef]

117. Allison, G.L.; Lowe, G.M.; Rahman, K. Aged Garlic Extract May Inhibit Aggregation in Human Platelets by Suppressing Calcium Mobilization. J. Nutr. 2006, 136, 799S–792S. [CrossRef]

118. Morihara, N.; Hino, A. Aged Garlic Extract Suppresses Platelet Aggregation by Changing the Functional Property of Platelets. J. Nutr. Met. 2017, 71, 249–256. [CrossRef] [PubMed]

119. Rahman, K.; Billington, D. Dietary Supplementation with Aged Garlic Extract Inhibits ADP-Induced Platelet Aggregation in Humans. J. Nutr. 2000, 130, 2662–2665. [CrossRef] [PubMed]

120. Rahman, K.; Lowe, G.M.; Smith, S. Aged Garlic Extract Inhibits Human Platelet Aggregation by Altering Intracellular Signaling and Platelet Shape Change. J. Nutr. 2016, 146, 410S–415S. [CrossRef] [PubMed]

121. Hiyasat, B.; Sabha, D.; Grötzing, K.; Kempfert, J.; Rauwald, J.-W.; Mohr, F.-W.; Dhein, S. Antiplatelet Activity of Allium Ursinum and Allium Sativum. Pharmacology 2009, 83, 197–204. [CrossRef] [PubMed]

122. Fukao, H.; Yoshida, H.; Tazawa, Y.; Hada, T. Antithrombotic Effects of Odorless Garlic Powder Both in Vitro and in Vivo. Biosci. Biotechnol. Biochem. 2007, 71, 84–90. [CrossRef] [PubMed]

123. Seyfi, P.; Mostafaei, A.; Mansouri, K.; Arshadi, D.; Mohammad-Motlagh, H.-R.; Kiani, A. In Vitro and in Vivo Anti-Angiogenesis Effect of Shallot (Allium ascalonicum): A Heat-Stable and Flavonoid-Rich Fraction of Shallot Extract Potently Inhibits Angiogenesis. Toxicol. In Vitro 2010, 24, 1655–1661. [CrossRef]

124. Hiramatsu, K.; Tsuneyoshi, T.; Ogawa, T.; Morihara, N. Aged Garlic Extract Enhances Heme Oxygenase-1 and Glutamate-Cysteine Ligase Modifier Subunit Expression via the Nuclear Factor Erythroid 2-Related Factor 2–Antioxidant Response Element Signaling Pathway in Human Endothelial Cells. Nutr. Res. 2016, 36, 143–149. [CrossRef]

125. Yeh, Y.-Y.; Yeh, S. Homocysteine-Lowering Action Is Another Potential Cardiovascular Protective Factor of Aged Garlic Extract. J. Nutr. 2006, 136, 745S–749S. [CrossRef]

126. Lee, E.N.; Choi, Y.W.; Kim, H.K.; Park, J.K.; Kim, H.J.; Kim, M.J.; Lee, H.W.; Kim, K.-H.; Bae, S.S.; Kim, B.S.; et al. Chloroform Extract of Aged Black Garlic Attenuates TNF-α-Induced ROS Generation, VCAM-1 Expression, NF-κB Activation and Adhesiveness for Monocytes in Human Umbilical Vein Endothelial Cells. Phytother. Res. 2011, 25, 92–100. [CrossRef]
Plants 2022, 11, 1920

127. Kim, K.-M.; Chun, S.-B.; Koo, M.-S.; Choi, W.-J.; Kim, T.-W.; Kwon, Y.-G.; Chung, H.-T.; Billiar, T.R.; Kim, Y.-M. Differential Regulation of NO Availability from Macrophages and Endothelial Cells by the Garlic Component S-Allyl Cysteine. Free Radic. Biol. Med. 2001, 30, 747–756. [CrossRef]

128. Louis, X.L.; Murphy, R.; Thandapilly, S.J.; Yu, L.; Netticadan, T. Garlic Extracts Prevent Oxidative Stress, Hypertrophy and Apoptosis in Cardiomyocytes: A Role for Nitric Oxide and Hydrogen Sulfide. BMC Complement. Altern. Med. 2012, 12, 140. [CrossRef] [PubMed]

129. Rassoul, F.; Salvetter, J.; Reissig, D.; Schneider, W.; Thiery, J.; Richter, V. The Influence of Garlic (Allium sativum) Extract on Interleukin 1α-Induced Expression of Endothelial Intercellular Adhesion Molecule-1 and Vascular Cell Adhesion Molecule-1. Phytomedicine 2006, 13, 230–235. [CrossRef]

130. Rose, P.; Whiteman, M.; Moore, P.K.; Zhu, Y.Z. Bioactive S-Alk(En)Yl Cysteine Sulfoxide Metabolites in the Genus Allium: The Chemistry of Potential Therapeutic Agents. Nat. Prod. Rep. 2005, 22, 351–368. [CrossRef]

131. Gonen, A.; Harats, D.; Rabinkov, A.; Miron, T.; Mirelman, D.; Wilchek, M.; Weiner, L.; Ulman, E.; Levkovitz, H.; Ben-Shushan, D.; et al. The Antiatherogenic Effect of Allicin: Possible Mode of Action. Pathobiology 2005, 72, 325–334. [CrossRef]

132. Rose, P.; Whiteman, M.; Moore, P.K.; Zhu, Y.Z. Bioactive S-Alk(En)Yl Cysteine Sulfoxide Metabolites in the Genus Allium: The Chemistry of Potential Therapeutic Agents. Nat. Prod. Rep. 2005, 22, 351–368. [CrossRef]

133. Goel, V.K.; Caruso, G. Chapter 5—Onion. In Nutritional Composition and Antioxidant Properties of Fruits and Vegetables; Jaiswal, A.K., Ed.; Academic Press: San Diego, CA, USA, 2000; pp. 73–87, ISBN 978-0-12-812780-3.

134. Golubkina, N.; Caruso, G. Chapter 5—Onion. In Nutritional Composition and Antioxidant Properties of Fruits and Vegetables; Jaiswal, A.K., Ed.; Academic Press: San Diego, CA, USA, 2000; pp. 73–87, ISBN 978-0-12-812780-3.

135. Najman, K.; Sadowska, A.; Buczak, K.; Leontowicz, H.; Leontowicz, M. Effect of Heat-Treated Garlic (Allium sativum L.) Leaves on Interleukin 1α-Induced Expression of Endothelial Intercellular Adhesion Molecule-1 and Vascular Cell Adhesion Molecule-1. Phytomedicine 2006, 13, 230–235. [CrossRef]

136. Ferri, N.; Yokoyama, K.; Chiu, S.-C.; Pai, M.-H.; Wang, F.-Y.; Kuo, Y.-H.; Tang, F.-Y. Organosulfur Garlic Compounds Induce Multiple Apoptosis and Oxidative Stress Pathway. BMC Complement. Altern. Med. 2016, 16, 133. [CrossRef] [PubMed]

137. Louis, X.L.; Murphy, R.; Thandapilly, S.J.; Yu, L.; Netticadan, T. Garlic Extracts Prevent Oxidative Stress, Hypertrophy and Apoptosis in Cardiomyocytes: A Role for Nitric Oxide and Hydrogen Sulfide. BMC Complement. Altern. Med. 2012, 12, 140. [CrossRef] [PubMed]

138. Li, G.; Cheng, G.; Wu, J.; Ma, S.; Zhang, A.; Han, W.; Sun, C. Allitridin Reduces IKr Current by Disrupting the Trafficking of the Human Ether-α-Go-Related Gene Channels. Cardiology 2014, 128, 1–8. [CrossRef]

139. Tsai, C.-Y.; Wang, C.-C.; Lai, T.-Y.; Tsu, H.-N.; Wang, C.-H.; Liang, H.-Y.; Kuo, W.-W. Antioxidant Effects of Diallyl Trisulfide on High Glucose-Induced Apoptosis Are Mediated by the PI3K/Akt-Dependent Activation of Nrf2 in Cardiomyocytes. Int. J. Cardiol. 2013, 168, 1286–1297. [CrossRef]

140. Tsai, C.-Y.; Wang, C.-C.; Shibue, M.A.; Yang, Y.-C.; Peng, H.; Wang, B.; Wei, Y.-M.; Chang, H.-Y.; Lee, C.-Y.; Huang, C.-Y.; et al. Diallyl Trisulfide Protects against High Glucose-Induced Cardiac Apoptosis by Stimulating the Production of Cystathionine Gamma-Lyase-Derived Hydrogen Sulfide. Int. J. Cardiol. 2015, 195, 300–310. [CrossRef]

141. Yang, H.-B.; Liu, H.-M.; Yan, J.-C.; Lu, Z.-Y. Effect of Diallyl Trisulfide on Ischemic Tissue Injury and Revascularization in a Diabetic Mouse Model. J. Cardiovasc. Pharmacol. 2018, 71, 367–374. [CrossRef] [PubMed]

142. Xiao, D.; Li, M.; Herman-Antosiewicz, A.; Antosiewicz, J.; Xiao, H.; Lew, K.L.; Zeng, Y.; Marynowski, S.W.; Singh, S.V. Diallyl Trisulfide Inhibits Angiogenic Features of Human Umbilical Vein Endothelial Cells by Causing Akt Inactivation and Down-Regulation of VEGF and VEGF-R2. Nutr. Cancer 2006, 55, 94–107. [CrossRef]
151. Chuah, S.C.; Moore, P.K.; Zhu, Y.Z. S-Allylcysteine Mediates Cardioprotection in an Acute Myocardial Infarction Rat Model via a Hydrogen Sulfide-Mediated Pathway. *Am. J. Physiol.-Heart Circ. Physiol.* 2007, 293, H2693–H2701. [CrossRef] [PubMed]

152. Ho, S.E.; Ide, N.; Lau, B.H.S. S-Allyl Cysteine Reduces Oxidant Load in Cells Involved in the Atherogenic Process. *Phytomedicine* 2001, 8, 39–46. [CrossRef]

153. Padmanabhan, M.; Prince, P.S.M. Preventive Effect of S-Allylcysteine on Lipid Peroxides and Antioxidants in Normal and Isoproterenol-Induced Cardiotoxicity in Rats: A Histopathological Study. *Toxicology* 2006, 224, 128–137. [CrossRef] [PubMed]

154. Kumar, K.; Augusti, K.T. Lipid Lowering Effect of S-Methyl Cysteine Sulfoxide from Allium Cepa Linn in High Cholesterol Diet Fed Rats. *J. Ethnopharmacol.* 2007, 109, 367–371. [CrossRef]

155. Chang, H.S.; Yamato, O.; Sakai, Y.; Yamasaki, M.; Maede, Y. Acceleration of Superoxide Generation in Polymorphonuclear Leukocytes and Inhibition of Platelet Aggregation by Alk(En)Yl Thiosulfates Derived from Onion and Garlic in Dogs and Humans. *Prostaglandins Leukot. Essent. Fat. Acids* 2004, 70, 77–83. [CrossRef]

156. Biró, Á.; Markovics, A.; Fazekas, M.E.; Fidler, G.; Szalóki, G.; Paholcsek, M.; Lukács, J.; Stündl, L.; Remenyik, J. Alliliphane Alleviates Hyperglycaemia-Induced Endothelial Dysfunction. *Nutrients* 2020, 12, 1690. [CrossRef]

157. Torres-Palazzolo, C.; de Paola, M.; Quesada, I.; Camargo, A.; Castro, C. 2-Vinyl-4H-1,3-Dithiin, a Bioavailable Compound from Allium Sativum. *Phytother. Res.* 2020, 34, 1131–1135. [CrossRef] [PubMed]

158. Rai, S.K.; Sharma, M.; Manral, A.; Saini, V.; Singh, A.; Srivivasan, B.P.; Tiwari, M. Novel Diallyldisulfide Analogs Ameliorate HMG-CoA Reductase Expression in Healthy Hypercholesterolemic Subjects. *Phytomedicine* 2016, 23, 723–727. [CrossRef]

159. Kumari, K.; Augusti, K.T. Lipid Lowering Effect of S-Methyl Cysteine Sulfoxide from Allium Cepa Linn in High Cholesterol Diet Fed Rats. *J. Ethnopharmacol.* 2020, 255, 320–327. [CrossRef]

160. Wang, Y.; Zou, M.-J.; Zhao, N.; Ren, J.-G.; Zhou, H.; Cheng, G. Effect of Diallyl Trisulfide on the Pharmacokinetics of Nifedipine in Rats. *J. Ethnopharmacol.* 2016, 177, 132–137. [CrossRef]

161. Larijani, V.N.; Ahmadi, N.; Zeb, I.; Khan, F.; Flores, F.; Budoff, M. Beneficial Effects of Aged Garlic Extract and Coenzyme Q10 in Rats with L-NAME-Induced Hypertension. *Eur. J. Pharmacol.* 2012, 691, 198–208. [CrossRef] [PubMed]

162. Atkin, M.; Laight, D.; Cummings, M.H. The Effects of Garlic Extract upon Endothelial Function, Vascular Inflammation, Oxidative Stress and Insulin Resistance in Adults with Type 2 Diabetes at High Cardiovascular Risk. A Pilot Double Blind Randomized Placebo Controlled Trial. *J. Diabetes Its Complicat.* 2016, 30, 723–727. [CrossRef]

163. Ahmadi, N.; Nabavi, V.; Hajasadeghi, F.; Zeb, I.; Flores, F.; Ebrahimi, R.; Budoff, M. Aged Garlic Extract with Supplement Is Associated with Increase in Brown Adipose, Decrease in White Adipose Tissue and Predict Lack of Progression in Coronary Atherosclerosis. *Int. J. Cardiol.* 2013, 168, 2310–2314. [CrossRef] [PubMed]

164. Wlosinska, M.; Nilsson, A.-C.; Hlebowicz, J.; Hauggaard, A.; Kjellin, M.; Fakhro, M.; Lindstedt, S. The Effect of Aged Garlic Extract on the Atherosclerotic Process—A Randomized Double-Blind Placebo-Controlled Trial. *BM Complement. Med. Ther.* 2020, 20, 132. [CrossRef]

165. Wlosinska, M.; Nilsson, A.; Hlebowicz, J.; Malmsjö, M.; Fakhro, M.; Lindstedt, S. Aged Garlic Extract Preserves Cutaneous Microcirculation in Patients with Increased Risk for Cardiovascular Diseases: A Double-blinded placebo-controlled Study. *Int. Wound J.* 2019, 16, 1487–1493. [CrossRef]

166. Chiu, H.-F.; Shen, Y.-C.; Huang, T.-Y.; Venkatakrishnan, K.; Wang, C.-K. Cardioprotective Efficacy of Red Wine Extract of Onion in Rats. *J. Food Sci.* 2011, 76, T30–T34. [CrossRef] [PubMed]

167. Larijani, V.N.; Ahmadi, N.; Zeb, I.; Khan, F.; Flores, F.; Budoff, M. Beneficial Effects of Aged Garlic Extract and Coenzyme Q10 on Vascular Elasticity and Endothelial Function: The FAITH Randomized Clinical Trial. *Nutrition* 2013, 29, 71–75. [CrossRef] [PubMed]

168. Atkin, M.; Laight, D.; Cummings, M.H. The Effects of Garlic Extract upon Endothelial Function, Vascular Inflammation, Oxidative Stress and Insulin Resistance in Adults with Type 2 Diabetes at High Cardiovascular Risk. A Pilot Double Blind Randomized Placebo Controlled Trial. *J. Diabetes Its Complicat.* 2016, 30, 723–727. [CrossRef]

169. Ahmadi, N.; Nabavi, V.; Hajasadeghi, F.; Zeb, I.; Flores, F.; Ebrahimi, R.; Budoff, M. Aged Garlic Extract with Supplement Is Associated with Increase in Brown Adipose, Decrease in White Adipose Tissue and Predict Lack of Progression in Coronary Atherosclerosis. *Int. J. Cardiol.* 2013, 168, 2310–2314. [CrossRef] [PubMed]

170. Choi, E.-Y.; Lee, H.; Woo, J.S.; Jang, H.H.; Hwang, S.J.; Kim, H.S.; Kim, W.-S.; Kim, Y.-S.; Choue, R.; Cha, Y.-J.; et al. Effect of Onion Peel Extract on Endothelial Function and Endothelial Progenitor Cells in Overweight and Obese Individuals. *Nutrition* 2015, 31, 1131–1135. [CrossRef] [PubMed]

171. Bahadoran, Z.; Mirmiran, P.; Momenan, A.A.; Azizi, F. Allium Vegetable Intakes and the Incidence of Cardiovascular Disease, Hypertension, Chronic Kidney Disease, and Type 2 Diabetes in Adults. *J. Hypertens.* 2017, 35, 1909–1916. [CrossRef] [PubMed]

172. Yoon, S.; Shiga, T.; Lam, P.; Edwards, R.J. Genetic Polymorphisms and Cardiovascular Drug Metabolism. In *Cardiovascular Pharmacogenetics*. Handbook of Experimental Pharmacology; Springer: Berlin/Heidelberg, Germany, 2004; Volume 160, pp. 39–77.

173. Liu, Y.; Zhang, L.; Liu, Y.-F.; Yan, F.-F.; Zhao, Y.-X. Effects of Bulbus Allii Macrostemi on Clinical Outcomes and Oxidized Low-Density Lipoprotein and Plasminogen in Unstable Angina/Non-ST-Segment Elevation Myocardial Infarction Patients. *Phytot. Res.* 2008, 22, 1539–1543. [CrossRef]

174. Alobaidi, A. Effect of Nigella Sativa and Allium Sativum Coadministered with Simvastatin in Dyslipidemia Patients: A Prospective, Randomized, Double-Blind Trial. *Anti-Inflamm. Anti-Allergy Agents Med. Chem.* 2014, 13, 68–74. [CrossRef] [PubMed]
174. Capasso, F.; Gaginella, T.S.; Grandolini, G.; Izzo, A.A. *Phytotherapy: A Quick Reference to Herbal Medicine*; Springer: Berlin/Heidelberg, Germany, 2003; ISBN 3540000526.

175. Leite, P.M.; Martins, M.A.P.; Castilho, R.O. Review on Mechanisms and Interactions in Concomitant Use of Herbs and Warfarin Therapy. *Biomed. Pharmacother.* **2016**, *83*, 14–21. [CrossRef] [PubMed]

176. Leite, P.M.; Martins, M.A.P.; Carvalho, M.D.G.; Castilho, R.O. Mechanisms and Interactions in Concomitant Use of Herbs and Warfarin Therapy: An Updated Review. *Biomed. Pharmacother.* **2021**, *143*, 112103. [CrossRef] [PubMed]