Renoprotective effect of the antioxidant curcumin: Recent findings

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Abstract

For years, there have been studies based on the use of natural compounds plant-derived as potential therapeutic agents for various diseases in humans. Curcumin is a phenolic compound extracted from Curcuma longa rhizome commonly used in Asia as a spice, pigment and additive. In traditional medicine of India and China, curcumin is considered as a therapeutic agent used in several foods. Numerous studies have shown that curcumin has broad biological functions particularly antioxidant and anti-inflammatory. In fact, it has been established that curcumin is a bifunctional antioxidant; it exerts antioxidant activity in a direct and an indirect way by scavenging reactive oxygen species and inducing an antioxidant response, respectively. The renoprotective effect of curcumin has been evaluated in several experimental models including diabetic nephropathy, chronic renal failure, ischemia and reperfusion and nephrotoxicity induced by compounds such as gentamicin, adriamycin, chloroquine, iron nitritolriacetate, sodium fluoride, hexavalent chromium and cisplatin. It has been shown recently in a model of chronic renal failure that curcumin exerts a therapeutic effect; in fact it reverts not only systemic alterations but also glomerular hemodynamic changes. Another recent finding shows that the renoprotective effect of curcumin is associated to preservation of function and redox balance of mitochondria. Taking together, these studies attribute the protective effect of curcumin in the kidney to the induction of the master regulator of antioxidant response nuclear factor erythroid-derived 2 (Nrf2), inhibition of mitochondrial dysfunction, attenuation of inflammatory response, preservation of antioxidant enzymes and prevention of oxidative stress. The information presented in this paper identifies curcumin as a promising renoprotective molecule against renal injury.

Keywords:
Antioxidant properties of curcumin
Curcumin: Traditional uses in folk medicine and biological properties
Curcumin as a food additive
Renal diseases epidemiology
Renoprotective effect of curcumin

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Introduction

History and cultivation of Curcuma longa

Curcuma longa (turmeric or curcuma) is a rhizomatous monocotyledonous perennial herbaceous plant member of the ginger family (Zingiberaceae), endemic and prevalent in tropical and subtropical regions including India, China and South East Asia. India is the most important producer, consumer and exporter of spices to form curry powder.

Curcuminoids and curcumin

Curcuma contains 60–70% carbohydrate, 8.6% protein, 5–10% fat, 2–7% fiber, 3–5% curcuminoids (50–70% curcumin) and up to 5% essential oils and resins. The curcuminoid content in turmeric may vary between 2 and 9%, depending on geographical conditions [29]. The composition of curcuminoids is approximately 70% curcumin (curcumin I), 17% demethoxycurcumin (curcumin II), 3% bis-demethoxycurcumin (curcumin III) and the rest (10%) is called cyclocurcumin (curcumin IV) [5,8,36] (Fig. 1). However, the last compound has been associated with poor or non-biological activity [61]. The most active component of turmeric is curcumin [67]; Vogel first isolated it in 1815 [29]. Curcumin is an orange-yellow crystalline powder practically insoluble in water. The structure of curcumin (Fig. 1) was first described in 1910 by Lampe and Milobedesa and proved to be diferuloylmethane [29]. Studies indicate that functional groups associated to curcumin chemical structure including bis-α,β-unsaturated β-diketone, two methoxy groups, two phenolic hydroxy groups and two double-conjugated

Fig. 1. Chemical structures and abundance of curcuminoids in turmeric that have therapeutic effects.
bonds might play an essential role in antiproliferative and anti-inflammatory activities assigned to curcumin [3]. Curcumin has keto-enol tautomers, of which keto form predominates in acid and neutral solutions and enol form in alkaline solutions.

**Curcumin as a food additive**

Turmeric is an ingredient of spice blends, mainly curry powder, which generally consists of turmeric, clove, paprika, ginger, cardamom, coriander, cumin, mace, pepper and cinnamon. Turmeric is commonly used as natural pigment (yellow 3) in the cosmetic and textile production but it is widely used in food industry. Indeed, turmeric is classified as an additive in E100 category and is used as food color additive in mustard, pastries, daily products and canned fish [40]. Furthermore, according to the Joint FAO/WHO Expert Committee on Food Additives (JECFA) the admissible daily intake (ADI) is 0–3 mg/kg body weight and it was established at the 61st JECFA in 2003 [29].

**Curcumin: Traditional uses in folk medicine and biological properties**

Curcuminoids have been consumed as therapeutic infusions over the centuries worldwide. In Ayurvedic medicine, curcumin is a well-documented treatment for various respiratory conditions such as, asthma, bronchial hyperactivity and allergy, as well as for liver disorders, anorexia, rheumatism, diabetic wounds, runny nose, cough and sinusitis [10]. In traditional Chinese medicine curcumin has been used to treat diseases associated with abdominal pain [35]. In ancient Hindu medicine, it was used to treat sprains and swelling [10]. In Oriental cultures, it has traditionally been used as good therapeutic alternative, particularly as an anti-inflammatory, antioxidant, anticarcinogenic and antimicrobial reagent. In fact, it has scientifically proven that curcumin is indeed antioxidant [11,31,26,17], anti-inflammatory [2,82,86], and antibacterial [52]. Moreover, it has also been used because of its hepatoprotective [56,32,86], thrombosisuppressive, neuroprotective [97,18,63], cardioprotective [37,23,73], antineoplastic [4,17,68], antiproliferative [12], hypoglycemic and antiarthritic effect [35]. Curcumin has also been used for the treatment of intestinal parasites and as a remedy for poisoning, snakebites and various other complaints [49]. In this review we are focused on the renoprotective effects of curcumin and the mechanisms involved in this effect.

**Antioxidant properties of curcumin**

Oxidative stress plays a major role in the pathogenesis of various diseases including myocardial ischemia, cerebral brain ischemia-reperfusion injury, hemorrhage and shock, neuronal cell injury, hypoxia and cancer. Curcuminoids exhibit a differential antioxidant activity in several in vitro and in vivo models, for example, preventing lipid peroxidation in a variety of cells such as erythrocytes, rat brain homogenates, rat liver microsomes, liposomes and macrophages, where peroxidation is induced by Fenton’s reagent, as well as for metals, hydrogen peroxide (H2O2) and 2,2′-azo-bis(2-amidino-propane) hydrochloride (AAPH) [62]. Furthermore, it has been reported that curcumin is a bifunctional antioxidant [26] because of its ability to react directly with reactive species and to induce an up-regulation of various cytoprotection and antioxidant proteins. Curcumin is able to scavenge superoxide anion (O2−) [6,76], hydroxyl radicals (‘OH) [13], H2O2, [6,13], singlet oxygen [25], nitric oxide [77,78], peroxynitrite [44] and peroxyl radicals (ROO·) [13]. Together, these mechanisms might explain, at least in part, some of the cytoprotective effects of this compound. Features as the presence of phenolic groups in the structure of curcumin (Fig. 1) explains its ability to react with reactive oxygen species (ROS) and reactive nitrogen species (RNS) and might probably be one of the mechanisms through which curcumin treatment protects the epithelial cells of renal tubules (LLC-PK1) from oxidative damage induced by H2O2 [22].

The indirect antioxidant capacity of curcumin is defined by its ability to induce the expression of cytoprotective proteins such as superoxide dismutase (SOD), catalase (CAT) [59], glutathione reductase (GR), glutathione peroxidase (GPx) [87], heme oxygenase 1 (HO-1) [42,63], glutathione-S-transferase (GST), NAD(P)H: quinine oxidoreductase 1 (NQO1) [88] and γ-glutamylcysteine ligase (γGCL) [65]. Furthermore, it has been reported that curcumin can increase the synthesis and concentration of reduced glutathione (GSH) in astrocytes and neurons by induction of γGCL [47]. The cytoprotective proteins induced by curcumin are regulated by the nuclear factor erythroid-derived 2 (Nrf2, [24,64]), which in turns is also activated by curcumin [17,28]. On the other hand, it is well known that encoding genes for cytoprotective proteins are induced coordinately by a common molecular mechanism in which the inducers highly modify reactive thiol groups of cysteine in the Kelch-Like ECH-Associated Protein 1 (Keap1) [27]. Keap1 protein is a zinc metalloprotein cysteine-rich bound to Nrf2 and normally associated with the protein complex cullin 3 (Cul3), which promotes the ubiquitination and subsequent proteosomal degradation, preventing Nrf2 translocation into the nucleus. Also, it was established that the gene expression of cytoprotective proteins is regulated by three cellular components: (i) Antioxidant response element (ARE) sequence, a specific sequence present in regulatory regions of the genes of cytoprotective proteins, (ii) Nrf2, a transcription factor consisting of a basic leucine zipper that regulates basal and inducible expression of cytoprotective genes, and (iii) Keap1, the chemical sensor for inducers. In general, the cyteine residues interaction of protein-Keap1 with some compounds induces conformational changes that abrogate the ability of Keap1 to repress Nrf2; this transcription factor migrates to the nucleus where it is combined with small Maf transcription factors. This complex binds to ARE facilitating the transcription of cytoprotective gene. By this reason, Nrf2 is considered a master regulator of the antioxidant response against oxidative stress.

**Renal diseases epidemiology**

The homeostasis of body extracellular electrolyte composition and fluid volume is essential for all animals and humans to survive. The kidney plays a fundamental role in maintaining precise body and/or extracellular electrolyte, fluid balance and blood pressure homeostasis primarily through the actions of its proximal and distal tubular segments of nephrons [38]. Under renal insufficiency conditions, deregulation of extracellular electrolytes or overall fluids volume may lead to disturbance of the circulation, including cardiac output and blood pressure [39]. Prevalence of chronic kidney disease is estimated to be 8–16% worldwide [43] and it is expected that the number of patients with chronic kidney disease increase at a fastest rate in the poorest parts of the world. According to the 2010 Global Burden of Disease Study, chronic kidney disease was ranked 27th in the list of causes of total number of global deaths in 1990, but rose to 18th in 2010 [48]. Patients with chronic kidney disease are at an increase risk of acute kidney injury (AKI). Acute kidney injury might occur with the use of several drugs, such as non-esteroidal anti-inflammatory drugs, antibiotics, antineoplastic drugs, and angiotensin-converting-enzyme inhibitors. A meta-analysis of 13 cohort studies confirmed that AKI is an important risk factor for chronic and end-stage renal disease [21]. Severe, long, and repeated episodes of acute kidney injury increase the risk of progression of chronic kidney disease. Acute and chronic renal failures are global public health issues with different features to take into account in different parts of the world, renal complications, which involve most organ systems, can be treated and prevented, by using different therapeutic strategies.
Renoprotective effect of curcumin

Renal injury induced by curcumin

Diabetic nephropathy (DN) is one of the main causes of end-stage renal disease. DN is characterized by the presence of hyperfiltration, glomerular hypertrophy, tubular albuminuria, mesangial matrix expansion, and increased expression of extracellular matrix proteins that involves several profibrotic factors such as transforming growth factor β (TGF-β) and connective tissue growth factor (CTGF). The effect of curcumin on diabetic nephropathy has been studied [75]. Sharma et al. [69] have been found that curcumin administration (15 and 30 mg/kg/day for two weeks) protects against streptozotocin-induced diabetic nephropathy and oxidative stress. Furthermore, Soetikno et al. [72] evaluated in a diabetic nephropathy model the effect of oral curcumin administration (100 mg/kg/day for 8 weeks) showing that curcumin prevents progression of renal disease.

Curcumin treatment attenuates proteinuria and improves creatinine clearance after 3 weeks of streptozotocin injection. In addition, it decreased oxidative stress by reducing levels of subunits of nicotinamide adenine dinucleotide phosphate (NADPH) oxidase Nox4 and p67phox, which catalyzes the synthesis of O2−. Moreover, it also increased the activity of the antioxidant enzyme GPx.

The renoprotective effect of curcumin was related to the downregulation of the profibrotic cytokines vascular endothelial growth factor (VEGF), TGF-β, CTGF and osteopontin as well as in extracellular matrix proteins fibronectin and collagen IV. Moreover, it was observed a reduction in the development of structural damage evidenced by lower glomerulosclerosis index (GS), tubulointerstitial (IT) fibrosis and arteriopathy. These effects might be in part mediated by inhibition of protein kinase C-β (PKC-β), kinase responsible for the phosphorylation of a wide diversity of proteins [72]. Cellular events such as inhibition of nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) [20,71] and decrease of macrophage infiltration [71], histone acetyltransferase p300 protein and oxidative stress [20], also have been involved in the mechanisms through curcumin protects against diabetic nephropathy. Furthermore, the protective effect of curcumin in diabetic nephropathy has also been associated to the prevention of kidney triglycerides buildup [74]. Interestingly, curcumin also mitigates cardiac [73] and cerebral [46] complications in streptozotocin-induced diabetes.

Curcumin derivatives have also been proved to be effective in ameliorating diabetic nephropathy. Pan et al. [58,57] studied the effect of curcumin B06 and C66 analogues in diabetic rats (0.2, 1 and 5 mg/kg/day for 6 weeks) [57,58]. B06 treatment reduced the inflammatory kidney response by the attenuation of (a) renal macrophage infiltration, (b) expression of the profibrotic cytokine TGF-β, inducible nitric oxide synthase (iNOS), and cyclooxygenase-2 (COX-2) and (c) the proinflammatory cytokines such as tumor necrosis factor-alpha (TNF-α) and monocyte chemoattractant protein-1 (MCP-1). The anti-inflammatory effect of B06 was associated with the inhibition of c-Jun N-terminal kinase (JNK)/NF-κB activation [58,57]. Similarly, the oral administration (80 mg/kg/day for 45 days) of tetrahydrocurcumin (THU), another curcumin derivative, attenuated the renal and hepatic dysfunction found in rats with diabetes induced by streptozotocin and nicotinamide [53].

Renal injury induced by 5/6 nephrectomy

The study of chronic progressive renal injury in rats as the 5/6 nephrectomy (5/6NX), which involves the removal of 5/6 of the renal mass, is useful in the evaluation of strategies to reduce renal injury. This model is characterized by proteinuria, hypertension, proliferation of smooth muscle cells of the glomerular arterioles (arteriopathy), IT inflammation and hemodynamic alterations in individual nephrons [79].

In 5/6NX renal injury model, Ghosh et al. [33] found that curcumin (75 mg/kg/day for 8 weeks) exerted a renoprotective effect associated with the attenuation of inflammation, macrophage infiltration and the high levels of TNF-α in plasma and kidney and also of renal NF-κB activation [33]. Further studies in this model by Tapia et al. [79] evaluated the renoprotective effect of curcumin (60 mg/kg for 37 days). Nephrectomized animals developed hypertension, proteinuria, increase in serum creatinine and blood urea nitrogen (BUN) and glomerular hemodynamic alterations including hyperfiltration (high single nephron glomerular filtration rate and single nephron glomerular plasma flow), glomerular hypertension (high glomerular capillary pressure) and decrease in the afferent and efferent resistances as well as renal injury characterized by GS and IT fibrosis and inflammation. Interestingly, after curcumin administration, all the above systemic and glomerular alterations were attenuated. This was the first work showing that curcumin is able to prevent glomerular hemodynamic alterations secondary to 5/6NX (Fig. 2). The protective effect of curcumin in this experimental model was associated to an increased activity of the antioxidant enzymes CAT, GPx, GR, GST and SOD and a decrease in oxidative stress. Additionally, Soetikno et al. [70] found that curcumin administration (75 mg/kg/day for 8 weeks) in 5/6NX rats could reduce proteinuria, systolic blood pressure, GS, IT damage and inflammatory markers as TGF-β, TNF-α, NF-κB and COX-2. They also found that curcumin attenuated malondialdehyde (MDA) levels (a lipid peroxidation marker) which was associated with lower expression of p67phox and p22phox, essential subunits of NADPH oxidase [70]. Both studies conducted by Tapia et al. [79,80] established that the protective mechanism achieved by curcumin in the kidney was mainly due to the nuclear translocation of Nrf2. Later, Tapia et al. [80] showed that curcumin posttreatment (120 mg/kg for 30 days), given 30 days after the 5/6NX induction, was able to revert renal damage and oxidative stress. These experiments show that curcumin is a therapeutic agent in experimental chronic renal failure. It is important to highlight that tissue injury throwback effect of curcumin turns this molecule into a promising therapeutic agent. In this regard, it is also remarkable that curcumin can protect not only against renal injury but also against adverse effects derived from 5/6NX. For example, Correa et al. [23] showed that curcumin...
(120 mg/kg/day for 67 days) protects against cardiovascular disorders and cardiac tissue remodeling associated to the development of chronic renal failure in rats with 5/6NX. The cardioprotective effect of curcumin in 5/6NX model was associated with reduction in ROS production and oxidative stress markers, an increased antioxidant response and preservation of mitochondria function. Finally, Ghosh et al. [34] reported in this 5/6NX model that curcumin (75 mg/kg/day for 8 weeks) blocks overexpression of inflammatory mediators such as TNF-α and interleukin 1β (IL-1β) through activation of phospholipase 2 (PLP2) and COX-2, both key regulators of inflammation and oxidative stress inducers.

Renal injury induced by ischemia and reperfusion (I/R) or by glomerulonephritis

The effect of curcumin on acute kidney injury induced by I/R also has been studied. This renal injury may be consequence of several factors as renal transplantation and involves vascular factors and tubular damage associated with high significant morbidity and mortality. Curcumin was administered orally to rats (200 mg/kg/day for 7 days) subjected to bilateral renal ischemia for 45 min followed by 24 h reperfusion [15]. Curcumin significantly attenuated the reduction of serum GPx and the levels of urea, cystatin C, and MDA in serum and the increase of the MDA concentration, nitric oxide and protein carbonyl content in kidney tissue of rats with I/R [15]. On the other hand, Jacob et al. [41] observed, in mice with deficient immune response and glomerulonephritis injury, that curcumin administration reduces GS and improves renal function (evaluated by BUN and albuminuria), which was associated to a decrease in inflammatory markers (TGF-β and MCP-1) and matrix proteins (fibronectin, laminin and collagen).

Shock-wave lithotripsy (SWL)

SWL is commonly used for treatment of renal stones and ROS are involved in the pathophysiology of renal injury due to SWL. The protective effect of curcumin (75 mg/kg/day for 35 days) against renal injury induced by SWL was studied in Sprague-Dawley rats [14]. Curcumin prevented interstitial, glomerular, tubular epithelial and endothelial cellular injuries by decreasing of iNOS and p65 (the active subunit of NF-κB) expression and serum nitric oxide levels. This protective effect was also associated with increased levels of GSH and attenuation of high levels of MDA in kidney [14].

Triiodothyronine (T3)-induced renal injury

One of the most important effects of thyroid hormones (T3 and T4) is the elevation of mitochondrial respiration, producing a hypermetabolic state with excess generation of free radicals. Thyroxine has been reported to induce renal hypertrophy with a rise in the DNA content. However, there is a paucity of information on T3-induced oxidative damage to mammalian kidney in general and with respect to antioxidant treatment in particular [66]. The effect of curcumin treatment (30 mg/kg/day for 15 days) was evaluated on renal damage and oxidative stress induced by T3 administration. It was found that curcumin was able to attenuate the mitochondrial lipid peroxidation, the enhanced SOD activity and the histopathological changes secondary to T3-administration [66].

Renal injury induced by drugs

Cisplatin and oxaliplatin

Cisplatin is an effective anticancer drug used against lung, ovarian cancer and some lymphomas, however renal damage has limited its use. It has been well established that oxidative stress is one of the mechanisms involved in cell damage induced by cisplatin. Indeed, a decrease of antioxidant defense is clearly observed in vivo and in vitro experimental models [19], Antunes et al. [9] reported curcumin administration (8 mg/kg before and after cisplatin injection) provided protection against cisplatin-induced neurotoxicity, ototoxicity and nephrotoxicity (evaluated by serum creatinine and creatinine clearance) and oxidative stress (evaluated by MDA and GSH levels) in rats. Moreover, Kuhad et al. [45] designed a two-day curcumin pretreatment and in parallel treatment of 15, 30 and 60 mg/kg of curcumin in a model of cisplatin-induced nephrotoxicity. The cisplatin-treated group that received 60 mg/kg of curcumin showed normal renal function (evaluated by measuring urea levels and creatinine clearance), which correlated with lipid peroxidation reduction. Interestingly, curcumin administration in cisplatin-treated animals attenuated, in a dose dependent manner, the cisplatin-induced decrease in GSH, SOD and CAT [45]. In addition, Ueki et al. [82] studied the effect of curcumin administration (100 mg/kg ip) on the inflammatory mechanisms involved in the pathogenesis of cisplatin-induced renal injury in mice. Curcumin prevented cisplatin-induced tubular necrosis, decreased renal dysfunction and the increase of pro-inflammatory markers including of TNF-α in serum, and TNF-α and MCP-1 in renal tissue, and a rising of intracellular adhesion molecule 1 (ICAM-1) mRNA in kidney. Oxaliplatin, another platinum-based chemotherapeutic agent can induce renal damage and oxidant stress. In vitro studies performed by Waly et al. [84] showed that oxaliplatin or cisplatin induced oxidative stress in human embryonic kidney cells (HEK 293). These cells also showed a decrease in total antioxidant capacity (TAC) and inhibition of the activity of SOD, CAT and GPx. Interestingly, curcumin added to these cell cultures significantly restored TAC and activity of the above mentioned antioxidant enzymes. Together, these studies clear up the ability of curcumin to decrease oxidative stress through modulation of those enzymes.

 Gentamicin

Gentamicin is an aminoglycoside used in the treatment of infections caused by Gram-negative bacteria that induces renal injury as a side effect. Curcumin treatment (200 mg/kg/day for 10 days) ameliorated the gentamicin-induced nephrotoxicity in rats [7]. Moreover, Manikanandan et al. [50] observed a renoprotective effect after curcumin administration (200 mg/kg/day for 7, 15 and 30 days) in gentamicin-treated animals. Nephrotoxicity was evidenced by increased serum creatinine and BUN. An increase in ROS and renal lipoperoxidation and a reduction in GSH and in the antioxidant enzymes GPx, GST, SOD and CAT were related to the impaired glomerular filtration. The degree of these alterations was diminished in animals treated with curcumin. Furthermore, curcumin modulated the inflammatory response in gentamicin-treated rats through NF-κB expression in a time-dependent manner [50].

Cyclosporin A (CsA)

CsA is widely used as immunosuppressant drug in organ transplantation to prevent rejection. This therapeutic agent induces renal damage as side effect. Tirkay et al. [81] showed that curcumin was effective by protecting CsA-induced damage, since animals that received the phenolic compound plus CsA did not show renal function deterioration or depletion of antioxidant enzymes. CsA increased the thiobarbituric acid reactive substances (TBARS), decreased renal endogenous antioxidant enzymes and deteriorated the renal function as assessed by increased serum creatinine, BUN and decreased creatinine and urea clearance. Curcumin reduced
elevated levels of TBARS, attenuated renal dysfunction, increased the levels of antioxidant enzymes and normalized the altered renal morphology in CsA treated rats.

Adriamycin (doxorubicin)

Adriamycin is a chemotherapeutic drug that may induce nephrotoxicity as a side effect. Treatment with curcumin (200 mg/kg/day in 1% gum acacia for 30 days) markedly protected against adriamycin-induced proteinuria, albuminuria, hypoalbuminaemia and hyperlipidemia. Furthermore, curcumin also reduced urinary levels of the enzyme N-acetyl-β-D-glucosaminidase (NAG), a marker of tubular damage [83].

Chloroquine

Chloroquine is a drug used in the malaria treatment and induces renal injury and oxidant stress as secondary effect. It was found that THU (80 mg/kg/day for 15 days) and curcumin treatment prevents chloroquine-induced nephrotoxicity and lipid peroxidation and also the decrease in the antioxidants vitamin C, vitamin E, SOD, CAT and GPx in kidneys of these rats [60].

Renal injury induced by chemicals

Sodium fluoride (NaF)

Renal damage induced by chemicals also has drawn attention. Nabavi et al. [54] reported the renoprotective effect of curcumin (10 and 20 mg/kg) in rats with nephrotoxicity induced by NaF. Renal damage in these rats relies on an impairment of renal function evidenced by increased serum creatinine and BUN and structural impairment due to interstitial edema, inflammation and fibrosis which was associated with increased nitric oxide, peroxides, ROS and MDA. On the other hand, curcumin treated animals showed an improvement in renal function and showed less histological damage associated to restoration of SOD and CAT activities and increased GSH levels.

Heavy metals

Studies evaluating the effect of curcumin in experimental models of nephrotoxicity by heavy metals such as chromium (Cr) [51], cadmium [30] and mercury (Hg) [1] have been conducted. Curcumin treatment attenuated renal dysfunction, oxidative stress and the decrease in antioxidant enzymes induced by metals. Interestingly, Molina-Jijón et al. [51] demonstrated that curcumin has a protective effect against nephrotoxicity induced by hexavalent chromium (Cr VI), and this property was related to the nuclear translocation of Nrf2, prevention of oxidant stress and preservation of the activity of antioxidant enzymes and of mitochondrial function in the kidney. In this study, a pretreatment of 10-day with 400 mg/kg of curcumin attenuated the structural and functional damage to the kidney which was associated with the prevention of mitochondrial oxidant stress and of the decrease in the following mitochondrial determinations: oxygen consumption (state 3), respiratory control, ATP content, calcium retention and membrane potential. Also curcumin prevented the decrease in the following enzymatic activities: aconitase, antioxidant enzymes and mitochondrial respiratory complexes I, II, II-III and V [51], (Fig. 3). This was the first demonstration that the prevention of renal injury was associated to the preservation of mitochondrial function.

Fig. 3. Curcumin is able to prevent mitochondrial dysfunction associated to renal injury. Curcumin is able to prevent lipid peroxidation and the decrease in the following mitochondrial determinations: activity of complexes I, II, II-III and V, activity of aconitase and antioxidant enzymes, GSH content, membrane potential, calcium retention and ATP content [51]. GSH (Glutathione), SOD (superoxide dismutase), CAT (catalase), GPx (glutathione peroxidase), GST (glutathione-S-transferase), GR (glutathione reductase), NADH (nicotinamide adenine dinucleotide), NADPH (nicotinamide adenine dinucleotide, reduced form), FAD+ (flavin adenine dinucleotide), FADH2 (flavin adenine dinucleotide, reduced form), ATP (adenosine triphosphate), ADP (adenosine diphosphate).

Fig. 4. Curcumin is able to prevent several mechanisms leading to renal injury. Curcumin renoprotective effects have been associated with the prevention of three main factors, first the reduction of oxidative stress by (a) preventing the generation of O2− and scavenging different reactive oxygen species, and (b) by preventing the Nrf2 degradation by ubiquitin proteasome pathway, thus an increase of many antioxidant enzymes. Curcumin has been shown also to be able to reduce inflammatory process by reducing the inflammatory transcription factors such as NF-κB and TNF-α. On the other hand the reduction of cytokines such as TGF-β or CTGF eventually prevents a fibrotic process. ROS (reactive oxygen species), O2− (superoxide);OH (hydroxyl radical), H2O2 (hydrogen peroxide), ONOO− (Peroxynitrite), O2• (Singlet oxygen), NO+ (Nitric Oxide), ROP+ (peroxyl radical), Nit2 (translocation of nuclear factor erythroid-derived 2), ARE (Antioxidant responsive elements), Keap1 (Kelch-like ECH-associated protein 1), Cul3 (culin 3), NedD8 (Neural precursor cell expressed developmentally down-regulated 8), u (ubiquitin), SOD (superoxide dismutase), CAT (catalase), GPx (glutathione peroxidase), GST (glutathione-S-transferase), GCL (glutathione-cystein-ligase), TX (thioredoxin), TGF-β (factor transforming growth beta), CTGF (Connective tissue growth factor), NF-κB (nuclear factor kappa-light-chain-enhancer of activated B cells), TNF-α (tumor necrosis factor), PKC (protein kinase C), P38 (phosphoinositol 3-kinase), PERK (protein kinase RNA-like endoplasmic reticulum kinase) MAPK (mitogen-activated protein kinase), CK2 (Casein kinase2).
Factors associated to renoprotection by curcumin.

| Studied targets | Effect of curcumin treatment | Renal injury models |
|-----------------|------------------------------|---------------------|
| Transcription factors | Nrf2 | Promotes the Nrf2 translocation to the nucleus, the major regulator of the antioxidant response | 5/6 NX, HM (Cr VI) |
| Pro-oxidant enzymes | NADPH oxidase subunits: Nox4, p67phox, p22phox | Attenuates oxidative stress by reducing levels of subunits of NADPH oxidase | Diabetic nephropathy |
| Antioxidants | GPx, CAT, GR, GST, SOD, NQO1 | Increases the activity of antioxidant enzymes | Diabetic nephropathy, 5/6 NX, U/R, SWL, T3, Cisplatin, Gentamicin, CsA, Chlr, NaF, HM (CrVI), FNT |
| Pro-inflammatory mediators | TNF-α, MCP-1, NF-κB, p65 (NF-κB subunit), JNK/NF-κB, COX-2, iNOS, IL-1β, IL-10, PLP2, TGF-β | Reduces the inflammatory response | Diabetic nephropathy, 5/6 NX, U/R, SWL, Cisplatin, Gentamicin |
| Extracellular matrix protein | Osteopontin, Fibronectin, Collagen IV, Laminin | Promotes a decrease in matrix proteins | U/R |
| Pro-fibrotic cytokines | VEGF, TGF-β | Attenuates the expression of the cytokines promoting a decrease in the inflammatory response | Diabetic nephropathy, I/R |
| Mitochondrial function markers | Oxygen consumption, ATP content, Calcium retention, Mitochondrial membrane potential, Activity of mitochondrial respiratory complexes | Prevents the decrease of mitochondrial parameters | HM (Cr VI) |

Ferric nitrilotriacetate

Ferric nitrilotriacetate is a carcinogen and strong inducer of renal oxidative stress. The effect of curcumin and THU on ferric nitrilotriacetate induced oxidative stress in male ddY mice was studied [55]. Animals were fed along four weeks with 0.5% curcumin or 0.5% THU triacetate induced oxidant stress in male ddY mice was studied [55].

Final remarks

According to epidemiological evidence, acute renal injury is a serious health and economical problem across the world with increasing cases since this disease can have its own etiology but also can be a complication from other diseases or can be a side effect from several medical treatments. The urgency to develop renoprotective strategies sets the eyes in compounds as curcumin, which has been used in the traditional medicine, specifically because its protective effects against renal damage. In this context, the experiments of Tapia et al. [80], in which curcumin was able to revert established renal injury and systemic alterations in rats with 5/6NX, are promising. At cellular and molecular levels, recent studies have demonstrated that this compound attenuates ROS generation and activates signaling pathways that involve the release of Nrf2 from Keap1, promoting transcription of genes that induce the expression of antioxidant system (GPx, GST, CAT, and SOD). Also, recent evidence shows that improvement of mitochondrial dysfunction induced during nephrotoxicity seems to be a key mechanism in curcumin protection and indirect reduction of ROS production through mechanisms including a decrease of O$_2^{$-}$ through the down regulation of expression of some NADPH oxidase subunits such as p67phox, p22phox and NOX4 essential for O$_2^{$-}$ generation, which is useful against inflammation, a common process during kidney injury. In this regard, curcumin also diminishes the expression of NF-κB and TNF-α, TGF-β, extracellular matrix proteins collagen type IV, fibronectin and growth factors such as CTGF. These proteins are closely involved not only in inflammation but also, in the remodeling tissue which leads to renal fibrosis. Fig. 4 and Table 1 summarize some cell protection mechanisms of curcumin in renal damage. The information presented in this paper identifies curcumin as a promising renoprotective molecule against renal injury.

References

[1] R. Aggarwal, S.K. Goel, J.R. Behari, Detoxification and antioxidant effects of curcumin in rats experimentally exposed to mercury, Journal of Applied Toxicology 30 (2010) 457–468.
[2] B.B. Aggarwal, K.B. Harikumar, Potential therapeutic effects of curcumin, the anti-inflammator agent, against neurodegenerative, cardiovascular, pulmonary, metabolic, autoimmune and neoplastic diseases, The International Journal of Biochemistry & Cell Biology 41 (2009) 40–59.
[3] B.B. Aggarwal, B. Sung, Pharmacological basis for the role of curcumin in chronic diseases: an age-old spice with modern targets, Trends in Pharmacological Sciences 30 (2009) 85–94.
[4] B.B. Aggarwal, A. Kumar, A.C. Bharti, Anticancer potential of curcumin: preclinical and clinical studies, Anticancer Research 23 (2003) 363–398.
[5] B.B. Aggarwal, C. Sundaram, N. Malani, H. Ichikawa, Curcumin: the Indian solid gold, Advances in Experimental Medicine and Biology 595 (2007) 1–75.
[6] T. Ak, I. Gülçin, Antioxidant and radical scavenging properties of curcumin, Chemico-Biological Interactions 174 (2008) 27–37.

[7] B.H. Ali, N. Al-Wabel, O. Mahmoud, H.M. Mousa, M. Hashad, Curcumin has a palliative action on gentamicin-induced nephrotoxicity in rats, Fundamental & Clinical Pharmacology 15 (2001) 473–477.

[8] P. Anand, S.G. Thomas, A.B. Kunnumakkara, C. Sundaram, K.B. Harikumar, S. Sung, S.T. Tharakan, K. Misra, I.K. Priyadarsini, K.N. Rajasekharan, B.B. Aggarwal, Biological activities of curcumin and its analogues (Curcuminoids) made by man and Mother Nature, Chemical Pharmacology & Therapeutics 76 (2000) 1506–1511.

[9] L.M. Antunes, J.D. Darin-Niê, L. Bianchi, Effects of the antioxidants curcumin or selenium on cisplatin-induced nephrotoxicity and lipid peroxidation in rats, Pharmacological Research 43 (2001) 145–150.

[10] C.C. Araújo, L.L. Leon, Biological activities of curcuminoids, “a chemical cocktail from molecule to biological function”, Angewandte Chemie International Edition 44 (2005) 129–155.

[11] A.L. Eggler, K.A. Gay, A.D. Mesecar, Molecular mechanisms of natural products and synthetic antioxidants: an updated overview, Free Radical Research 44 (2010) 1216–1262.

[12] B.E. Bachmeier, P. Killian, U. Pfeffer, A.G. Nerlich, Novel aspects for the application of curcumin in chemoprevention of various cancers, Frontiers in Bioscience (Scholar edition) 5 (2010) 697–717.

[13] A. Barzegar, A.A. Moosavi-Movahedi, Intracellular ROS protection efficiency and free radical-scavenging activity of curcumin, Plots One 6 (2011) e20012.

[14] M. Bas, V. Tugcu, E. Kemahli, O. Bayrak, E. Uz, R. Bayrak, F. Turgut, A.F. Atmaca, S. Sahin, M.E. Yildirim, A.I. Tasci, Curcumin as Curcumin: from kitchen to clinic, Biochemical Pharmacology 75 (2008) 787–809.

[15] A. Goel, B.R. Aggarwal, Curcumin, the golden spice from Indian saffron, is a chemosensitizer and radiosensitizer for tumors and chemoprotector and radioprotector for normal organs, Nutrition and Cancer 62 (2010) 919–930.

[16] A. González-Salazar, E. Molina-Jijón, F. Correa, G. Zarco-Márquez, M. Calderón-Olivar, E. Tapia, C. Zazueta, J. Pedraza-Chaverri, Curcumin protects from cardiac reperfusion damage by attenuation of oxidative stress and mitochondrial dysfunction, Cardiology 2013: Article ID 424692, 1–19, 19 pages.

[17] J.E. Hall, M.W. Brands, E.W. Shek, Central role of the kidney and abnormal fluid volume control in hypertension, Journal of Human Hypertension 10 (1996) 633–639.

[18] S. Himesh, P.S. Sharan, K. Mishra, N. Govind, A. Singhai, Qualitative and quantitative profile of curcumin from exotic extract of curcuma longa, International Research Journal of Pharmacy 2 (2011) 180–184.

[19] A. Jacob, L. Chaves, M.T. Eadon, A. Chang, R.J. Quigg, J.J. Alexander, Curcumin alleviates immune-complex mediated glomerulonephritis in factor H deficient mice, Journal of Immunology 180 (2008) 159–164.

[20] V. Eybl, D. Kotyzová, M. Bludovská, The effect of curcumin on cadmium-induced toxicity and free radical-scavenging activity of curcumin and related compounds, Toxicological In Vitro 24 (2010) 563–569.

[21] S.S. Ghosh, H.D. Massey, R. Krieg, Z.A. Fazelbhoy, S. Ghosh, D.A. Sica, I. Fakhry, T.W. Gehr, Curcumin ameliorates renal failure in 5/6 nephrectomized rats: role of inflammation, Renal Physiology–American Journal of Physiology 296 (2009) F1146–F1157.

[22] S.S. Ghosh, R. Krieg, H.D. Massey, D.A. Sica, I. Fakhry, S. Ghosh, T.W. Gehr, Curcumin and enalapril ameliorate renal failure by antagonizing inflammation in 5/6 nephrectomized rats: role of phospholipase and cyclooxygenase, Renal Physiology–American Journal of Physiology 302 (2012) F439–F454.

[23] A. Goel, A.R. Kummar, Curcumin as Curcumin: from kitchen to clinic, Biochemical Pharmacology 75 (2008) 787–809.
pathogenic changes in rat kidney and heart, The Journal of Nutritional Biochemistry 24 (2013) 146–155.

[58] Y. Pan, G. Zhu, Y. Wang, L. Cai, Y. Cai, J. Hu, Y. Li, Y. Yan, Z. Wang, X. Li, T. Wei, G. Liang, Inhibition of high glucose-induced inflammatory response and macrophage infiltration by a novel curcumin derivative prevents renal injury in diabetic rats, British Journal of Pharmacology 166 (2012) 1169–1182.

[59] H.D. Panchal, K. Vranizan, C.Y. Lee, J. Ho, J. Ngai, P.S. Timiras, Early anti-inflammatory and anti-proliferative curcumin effects on neuroglioma cells suggest therapeutic targets, Neurochemical Research 33 (2008) 1701–1710.

[60] L. Pari, P. Murugan, Tetrahydrocurcumin: effect on chloroquine-mediated oxidative damage in rat kidney, Basic & Clinical Pharmacology & Toxicology 96 (2006) 329–334.

[61] I. Rahman, S.K. Biswas, P.A. Kirkham, Regulation of in vivo redox signaling by dietary polyphenols, Biochemical Pharmacology 72 (2006) 1439–1452.

[62] S. Reddy, B.B. Aggarwal, Curcumin is a non-competitive and selective inhibitor of phosphorylase kinase, FEBS Letters 341 (1994) 19–22.

[63] L.M. Reyes-Fermín, S. González-Reyes, N.G. Tarco-Álvarez, M. Hernández-Nava, M. Orozco-Ibarra, J. Pedraza-Chaverri, Neuroprotective effect of α-mangostin and curcumin against isocitrate-depleted cell death, Nutritional Neuroscience 15 (2012) 34–41.

[64] A.I. Rojo, O.N. Medina-Campos, P. Rada, A. Zúñiga-Toalá, A. López-Gazcón, L.M. Reyes-Fermín, S. González-Reyes, N.G. Tarco-Álvarez, M. Hernández-Nava, M. Orozco-Ibarra, J. Pedraza-Chaverri, Neuroprotective effect of α-mangostin and curcumin against isocitrate-depleted cell death, Nutritional Neuroscience 15 (2012) 34–41.

[65] S.K. Sandur, M.K. Pandey, B. Sung, K.S. Ahn, A. Murakami, G. Sethi, P. Limtrakul, L. Samanta, J. Panigrahi, S. Bhanja, G.B. Chainy, Effect of turmeric and its active principle, The Journal of Nutritional Biochemistry 33 (2018) 1701–1710.

[66] V. Soetikno, F.R. Sari, V. Sukumaran, A.P. Lakshmanan, S. Mito, M. Harima, R.A. Thandavarayan, K. Suzuki, M. Nagata, R. Takagi, K. Watanabe, Curcumin prevents diabetic cardiomyopathy in streptozotocin-induced diabetic rats: possible involvement of PKC- MAPK signaling pathway, European Journal of Pharmaceutical Sciences 47 (2012) 604–614.

[67] V. Soetikno, F.R. Sari, V. Sukumaran, A.P. Lakshmanan, M. Harima, K. Suzuki, H. Kawachi, K. Watanabe, Curcumin decreases renal triglyceride accumulation through AMPK-SREBP signaling pathway in streptozotocin-induced type 1 diabetic rats, Journal of Nutritional Biochemistry 24 (2013) 796–802.

[68] V. Soetikno, K. Suzuki, P.T. Veeraveedu, S. Arumugam, A.P. Lakshmanan, H. Sone, K. Watanabe, Molecular understanding of curcumin in diabetic nephropathy, Drug Discovery Today 18 (2013) 756–763.

[69] N. Sreejayan, M.N. Rao, Free radical scavenging activity of curcuminoids, Arzneimittelforschung 46 (1996) 169–171.

[70] R.M. Sreejayan, Nitric oxide scavenging by curcuminoids, The Journal of Phamacy and Pharmacology 49 (1997) 105–107.

[71] Y. Sumanont, Y. Murakami, M. Tohda, O. Vajragupta, K. Matsumoto, H. Watanabe, Evaluation of the nitric oxide radical scavenging activity of manganese complexes of curcumin and its derivative, Biological & Pharmaceutical Bulletin 27 (2004) 170–173.

[72] E. Tapia, V. Soto, K.M. Ortiz-Vega, G. Zarco-Márquez, E. Molina-Jijón, M. Cristóbál-García, J. Santamaría, W.R. García-Niño, F. Correa, C. Zazueta, J. Pedraza-Chaverri, Curcumin induces Nrf2 nuclear translocation and prevents glomerular hypertension, hyperfiltration, oxidant stress, and the decrease in antioxidant enzymes in 5/6 nephrectomized rats, Oxidative Medicine and Cellular Longevity (2012) 269038.

[73] E. Tapia, Z.L. Zatarain-Barrón, R. Hernández-Pando, G. Zarco-Márquez, E. Molina-Jijón, M. Cristóbál-García, J. Santamaría, J. Pedraza-Chaverri, Curcumin reverses glomerular hemodynamic alterations and oxidant stress in 5 / 6 nephrectomized rats, Phytomedicine 20 (2013) 359–366.

[74] N. Tirkay, G. Kaur, G. Vij, K. Chopra, Curcuma, a diferuloylmethane, attenuates cyclosporine-induced renal dysfunction and oxidative stress in rat kidneys, BMC Pharmacology 5 (2005) 15.

[75] M. Ueki, M. Ueno, J. Morishita, N. Maekawa, Curcumin ameliorates cisplatin-induced nephrotoxicity by inhibiting renal inflammation in mice, Journal of Bioscience and Bioengineering 115 (2013) 547–551.

[76] N. Venkatesan, D. Punithavarthi, V. Arumugam, Curcumin prevents Adriamycin nephrotoxicity in rats, British Journal of Pharmacology 129 (2000) 231–234.

[77] M.I. Waly, M.S. Al Moundhri, B.H. Ali, Effect of curcumin on cisplatin- and oxaliplatin-induced oxidative stress in human embryonic kidney (HEK) 293 cells, Renal Failure 33 (2011) 518–523.

[78] H.M. Wang, Y.X. Zhao, S. Zhang, G.D. Liu, W.Y. Kang, H.D. Tang, J.Q. Ding, S.D. Chen, PPARgamma agonist curcumin reduces the amyloid-beta-stimulated inflammatory responses in primary astrocytes, Journal of Alzheimer’s Disease 20 (2010) 1189–1199.

[79] M.E. Wang, Y.C. Chen, I.S. Chen, S.C. Hsieh, S.S. Chen, C.H. Chiu, Curcumin protects against thioacetamide-induced hepatic fibrosis by attenuating the inflammatory response and inducing apoptosis of damaged hepatocytes, The Journal of Nutritional Biochemistry 23 (2012) 1352–1366.

[80] L.P. Yarru, R.S. Settivari, N.K. Gowda, E. Antoniou, D.R. Ledoux, G.E. Rottinghaus, Effects of turmeric (Curcuma longa) on the expression of hepatic genes associated with biotransformation, antioxidant, and immune system in broiler chicks fed aflatoxin, Poultry Science 88 (2009) 2620–2627.

[81] S.F. Ye, Q.Z. Hou, L.M. Zhong, Q.Q. Zhang, Effect of curcumin on the induction of glutathione S-transferases and NADPH:quinone oxidoreductase and its possible mechanism of action, Yao Xue Xue Bao 42 (2007) 376–380.