A Rapid In-Clinic Test Detects Acute Leptospirosis in Dogs with High Sensitivity and Specificity

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A rapid IgM-detection immunochromatographic test (WITNESS® Lepto, Zoetis) has recently become available to identify acute canine leptospirosis at the point of care. Diagnostic sensitivity and specificity of the test were evaluated by comparison with the microscopic agglutination assay (MAT), using a positive cut-off titer of ≥800. Banked serum samples from dogs exhibiting clinical signs and suspected leptospirosis were selected to form three groups based on MAT titer: (1) positive (n = 50); (2) borderline (n = 35); and (3) negative (n = 50). Using an analysis to weight group sizes to reflect French prevalence, the sensitivity and specificity were 98% and 93.5% (88.2% unweighted), respectively. This test rapidly identifies cases of acute canine leptospirosis with high levels of sensitivity and specificity with no interference from previous vaccination.

1. Introduction

Leptospirosis is a global bacterial zoonotic disease affecting humans and wild and domestic animals, including dogs. Clinical signs of leptospirosis in dogs are nonspecific, typically including renal and/or hepatic dysfunction, and can manifest as subclinical, chronic, or acute infections, sometimes with fatal outcomes. Infected animals excrete Leptospira in their urine and can pose a risk to humans who become infected through damaged skin or via the conjunctiva or mucosa, causing a potentially fatal disease [1].

The reference serological test for diagnosis is the microscopic agglutination test (MAT). Because the test is complex to perform and interpret, serum must be sent to a reference laboratory [1]. As such, MAT tends to be the last resort in first-opinion practice. An immunochromatographic test (WITNESS Lepto, Zoetis) has been developed, offering the advantage of a result in 10 min. The test uses whole cell antigen extracts of L. kirschneri serovar Grippotyphosa and L. interrogans serovar Bratislava to detect canine IgM made in response to infection. These serovars were selected as they are of clinical relevance and relative prevalence worldwide [2–8]. However, unpublished data from internal studies and the current study suggest broad serovar reactivity on the test, likely due to conserved antigens across pathogenic Leptospira. The objective of the present study was to evaluate the sensitivity (Se) and specificity (Sp) of this new test in client-owned French dogs as compared to MAT.

2. Materials and Methods

All samples in this study were taken from an established bank of clinical samples submitted by French veterinarians to the Laboratoire des Leptospires (VetAgro Sup, Marcy L’Etoile, France) for diagnostic testing because leptospirosis was suspected. A total of 135 canine serum samples were tested using WITNESS Lepto retrospectively compared to MAT. Based on previous MAT results, the samples were divided into three groups: (1) a positive group (n = 50) defined as having a MAT titer ≥800 for any of the 19 serovars used in the test and a diagnosis of leptospirosis; (2) a borderline group (n = 35) defined as having a MAT titer <800 for at least
Table 1: Weighted prevalence rates based on French data.

| Group                                      | n in present study | French prevalence* | Weighted n in present study |
|--------------------------------------------|-------------------|--------------------|-----------------------------|
| Positive (MAT titer ≥ 800)                 | 50                | 25%                | 33.75                       |
| Borderline (MAT titer < 800)               | 35                | 17%                | 22.95                       |
| Negative (MAT titer negative or vaccinal ≤ 400) | 50                | 58%                | 78.3                        |

MAT: microscopic agglutination test.
*Renaud et al., 2013 [7].

Table 2: MAT versus WITNESS Lepto: 2 × 2 table of weighted and unweighted results used for the calculation of sensitivity and specificity.

(a) Unweighted results

| WITNESS Lepto | MAT positive (n) | MAT negative (n) |
|---------------|-----------------|-----------------|
| Positive      | TP: 49 (positive group) | FP: 10 (borderline group) |
| Negative      | FN: 1 (positive group) | TN: 25 (borderline group), 50 (negative group) |

(b) Weighted data used to calculate Se and Sp

| WITNESS Lepto | MAT positive (n) | MAT negative (n) |
|---------------|-----------------|-----------------|
| Positive      | TP: 33.075      | FP: 6.55        |
| Negative      | FN: 0.675       | TN: 94.45       |

Sensitivity: 98% Specificity: 93.5%

MAT: microscopic agglutination test; Se: sensitivity; Sp: specificity; TP: true positive; TN: true negative; FP: false positive; FN: false negative.

3. Results and Discussion

Forty-nine of 50 (49/50) samples in the MAT positive group were positive, while 25/35 MAT borderline samples and all 50 MAT negativesamples were negative on WITNESS Lepto (Table 2(a)). Using the weighted distribution, Se was 98% (95% CI 88.7 to 99.9%) and Sp was 93.5% (95% CI 87.4 to 97.1%). With no weighting to the sample distribution, the Se remained at 98% and the Sp was 88.2%. The data is summarized in Table 2(b). Forty-two of the 50 MAT negative dogs had previously received a Leptospira vaccine with 38 of them being within a year of vaccination, suggesting that previous vaccination did not interfere with a correct test result.

The level of agreement between WITNESS Lepto and MAT was very high for samples that were either MAT positive or MAT negative. All but one of the samples where the WITNESS result differed with MAT came from the borderline group. The borderline samples were considered negative as their MAT titers were not ≥800. With this classification, some samples in the borderline group were still likely to be truly infected. An IgM immunoblot assay had 88% sensitivity in the first three days of human leptospirosis, compared to 2% sensitivity for the MAT [11]. Another human IgM ELISA detected infection in 29% of the cases before MAT had detectable titers [12]. Thus, WITNESS Lepto may have been correctly identifying positive samples in the borderline group before they reached a MAT titer of ≥800. This emphasizes the difficulty in the interpretation of a single MAT in practice. The sensitivity and specificity of MAT increase considerably when samples collected from the animal one or two weeks apart can be tested [13]. However, client compliance can
be difficult due to an extra trip to the clinic at extra cost. The WITNESS test provides a time-saving, easy to interpret, and economical solution in this regard. The test provides additional advantages to the MAT because the test is safe and does not require working with live organism, as is the case with the MAT. The convenience of testing in the clinic provides immediate identification of infected dogs, allowing for proper quarantine and handling protocols to be implemented which reduces the risk of transmission to humans and other animals. Whereas the MAT detects both IgM and IgG, WITNESS Lepto detects only IgM, simplifying result interpretation and negating cross-reactivity with IgG antibody from dogs with a previous *Leptospira* vaccination [14].

### 4. Conclusions

In conclusion, this study indicates that WITNESS Lepto is a reliable test with high levels of sensitivity and specificity, as compared to the MAT, for the diagnosis of acute leptospirosis in dogs that are showing compatible clinical signs. With its ease of use and immediate result, it can transform the veterinarian’s case management by enabling earlier diagnosis and treatment or ruling out of leptospirosis, all to the benefit of both the patient and the public health.

### Competing Interests

The authors Christophe Calleja, Michael Loenser, Dan Lin, and Joshua Lizer are employed by Zoetis and were responsible for the experimental design and analysis of the data. Angeli Kodjo conducted the testing and selected the individual samples to be used under a research agreement fully funded by Zoetis.

### References

[1] P. N. Levett, "Leptospirosis: a forgotten zoonosis?" *Clinical and Applied Immunology Reviews*, vol. 4, no. 6, pp. 435–448, 2004.

[2] R. D. Blazius, P. R. T. Romão, E. M. C. G. Blazius, and O. S. da Silva, "Occurrence of *Leptospira* spp. soropositive stray dogs in Itapema, Santa Catarina, Brazil," *Cadernos de Saúde pública*, vol. 21, no. 6, pp. 1952–1956, 2005.

[3] P. Boutilier, A. Carr, and R. L. Schulman, "Leptospirosis in dogs: a serologic survey and case series 1996 to 2001," *Veterinary Therapeutics*, vol. 4, no. 2, pp. 178–187, 2003.

[4] R. Gautam, C.-C. Wu, L. E. Guptill, A. Potter, and G. E. Moore, "Detection of antibodies against *Leptospira* serovars via microscopic agglutination tests in dogs in the United States, 2000–2007," *Journal of the American Veterinary Medical Association*, vol. 237, no. 3, pp. 293–298, 2010.

[5] V. Geisen, C. Stengel, S. Brem, W. Müller, C. Greene, and K. Hartmann, "Canine leptospirosis infections—clinical signs and outcome with different suspected *Leptospira* serogroups (42 cases)," *Journal of Small Animal Practice*, vol. 48, no. 6, pp. 324–328, 2007.

[6] M. Oliveira Lavinsky, R. A. Said, G. M. R. Strenzel, and H. Langoni, "Seroprevalence of anti-*Leptospira* spp. antibodies in dogs in Bahia, Brazil," *Preventive Veterinary Medicine*, vol. 106, no. 1, pp. 79–84, 2012.

[7] C. Renaud, S. Andrews, Z. Djeloudji et al., "Prevalence of the *Leptospira* serovars *bratislava*, *grippotyphosa*, *mozdok* and *pomona* in French dogs," *Veterinary Journal*, vol. 196, no. 1, pp. 126–127, 2013.

[8] J. E. Stokes, J. B. Kaneene, W. D. Schall et al., "Prevalence of serum antibodies against six *Leptospira* serovars in healthy dogs," *Journal of the American Veterinary Medical Association*, vol. 230, no. 11, pp. 1657–1664, 2007.

[9] H. Jeffreys, *Theory of Probability*, Oxford University Press, 1939.

[10] H. Jeffreys, "An invariant form for the prior probability in estimation problems," *Proceedings of the Royal Society of London, Series A: Mathematical and Physical Sciences*, vol. 186, no. 1007, pp. 453–461, 1946.

[11] J. E. Sykes, K. Hartmann, K. F. Lunn, G. E. Moore, R. A. Stoddard, and R. E. Goldstein, "2010 ACVIM small animal consensus statement on leptospirosis: diagnosis, epidemiology, treatment, and prevention," *Journal of Veterinary Internal Medicine*, vol. 25, no. 1, pp. 1–13, 2011.

[12] W. E. Winslow, D. J. Merry, M. L. Pirc, and P. L. Devine, "Evaluation of a commercial enzyme-linked immunosorbent assay for detection of immunoglobulin M antibody in diagnosis of human leptospiral infection," *Journal of Clinical Microbiology*, vol. 35, no. 8, pp. 1938–1942, 1997.

[13] S. Schuller, T. Francey, K. Hartmann et al., "European consensus statement on leptospirosis in dogs and cats," *Journal of Small Animal Practice*, vol. 56, no. 3, pp. 159–179, 2015.

[14] P. N. Levett, "Leptospirosis," *Clinical Microbiology Reviews*, vol. 14, no. 2, pp. 296–326, 2001.