Thymidylate synthase polymorphisms in genomic DNA as clinical outcome predictors in a European population of advanced non-small cell lung cancer patients receiving pemetrexed

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Abstract

Background: We studied whether thymidylate synthase (TS) genotype has an independent prognostic/predictive impact on a European population of advanced non-small cell lung cancer (NSCLC) patients receiving pemetrexed.

Methods: Twenty-five patients treated with pemetrexed-based regimens were included. Genomic DNA was isolated prior to treatment. The variable number of tandem repeat (VNTR) polymorphisms, the G > C single nucleotide polymorphisms (SNP) and the TS 6-bp insertion/deletion (6/6) in the 3′ untranslated region (UTR) polymorphisms were analyzed and correlated with overall response rate (ORR), progression-free survival (PFS), overall-survival (OS) and toxicity.

Results: The genotype +6/+6 predicted a higher ORR among active/former smokers compared to +6/-6 genotype (100% vs. 50%; p = 0.085). Overall, the 3R/3R genotype predicted a higher ORR (100%) over the rest VNTR polymorphisms (p = 0.055). The presence of 3R/3R genotype significantly correlated with a superior ORR in patients without EGFR activating mutations (100%) compared to 2R/2R, 2R/3R and 3R/4R genotype (77.8%, 33.3% and 0% respectively; p = 0.017). After a median follow-up of 21 months, a trend towards a better PFS, although not significant, was found among subjects showing 3R/3R polymorphisms (p = 0.089). A significantly superior OS was found in patients showing 3R/3R genotype rather than other VNTR polymorphisms (p = 0.019). No significant correlation with the toxicity was observed.

Conclusion: In our series, 3R/3R polymorphism correlated with a superior OS. Also, this polymorphism, when associated to wild type EGFR, was related to a higher ORR to pemetrexed. Toxicity was not significantly correlated with a specific TS genotype.

Keywords: Thymidylate synthase, Polymorphisms, Epidermal growth factor receptor, Predictive factors, Prognostic factors, Non-small cell lung cancer
Background

Lung cancer represents the most frequent cause of cancer deaths. More than 225,000 new cases were diagnosed during 2012 only in the United States of America, accounting for approximately 160,000 annual deaths [1,2]. More than 50% of the patients diagnosed with non-small cell lung cancer (NSCLC) present advanced disease (stage III and IV) at onset. The most common histology is adenocarcinoma representing approximately 80% of all cases [3].

Pemetrexed, is a multitargeted antifolate drug and one of the latest active drugs against NSCLC [4] approved for the first-line [5] (in combination with cisplatin) [6] and second-line treatment (monotherapy) of patients with non-squamous histology [7]. More recently, pemetrexed gained approval for its use as a single-agent maintenance therapy [8] after response/stabilization to four cycles of a platinum doublet with or without pemetrexed.

Thymidylate synthase (TS) is the main biological target of antifolate drugs such as pemetrexed or 5-fluorouracil. Different studies have evaluated the correlation between tumor TS expression and TS genotype and the prognosis of patients with different cancer types treated with antifolates [9-11]. In NSCLC, constitutive expression of TS is lower in tumors with adenocarcinoma histology than among those with squamous differentiation [12]. This finding could possibly explain the higher efficacy of the drug among non-squamous histology patients. The potential predictive role of TS polymorphisms in NSCLC has never been studied in a European population. In addition, how differential TS genotypes may impact on the outcome of patients depending on their smoking status or with Epidermal Growth Factor Receptor (EGFR) activating mutations tumors is to be determined.

Finally, although the toxicity profile described in most patients receiving pemetrexed in combination or as a single agent is usually favorable, there are several reported cases of fundamentally dermatological, hematological and potentially serious renal toxicities, even when the recommended vitamin prophylaxis guidelines have been followed [13-15]. Nonetheless, the tumor TS levels or its polymorphisms in each patient could explain these cases of severe toxicity, as it has been suggested in other neoplasms treated with other antifolate drugs [9]. The potential association between different TS genotypes and the toxicity experienced by a European population of patients with NSCLC receiving pemetrexed is also to be studied.

Three different types of polymorphisms have been described in the TS gene. In the gene promoter, there is a variable number of tandem repeats (VNTR) of 28 pb in the 5′-region. Thus, cases of two or three repetitions of this tandem TS gene promoter enhancer region (TSER) have been described [16]. A second type of polymorphism consists in a change in a single nucleotide (G → C), in one of the sequences of the repetition comprising a single nucleotide polymorphism (SNP) [16]. A third modality of polymorphisms consists in the deletion or insertion of 6 pair of bases (bp) in the 3′-UTR region (untranslated region) [16].

In summary, the potential usefulness of TS genotype as an independent prognostic factor or predictor of response to pemetrexed-based regimens in a European NSCLC population has not been studied. Similarly, no clear evidence is available about the potential correlation between the different TS genotypes and the toxicity experienced by those patients.

Therefore we decided to investigate the three known polymorphisms of TS gene, and their correlation with objective response rate (ORR), progression-free survival (PFS) and overall survival (OS), as well as toxicity in European patients with advanced NSCLC treated with pemetrexed-based regimens.

Methods

Patients and samples

Overall, 25 consecutive stage III-IV NSCLC patients treated at a single institution, from which peripheral blood samples were available, were analyzed. All of them received pemetrexed-based regimens in the first, second or third line settings, according to our institutional therapeutic protocols.

After the informed consent was obtained from all patients, 10 ml of peripheral blood samples were collected before the administration of the first cycle of a pemetrexed-containing regimen. Blood samples were stored at the Biobank of the University of Navarra and were processed following standard operating procedures approved by the Ethical and Scientific Committees.

Tumor ORR to the treatment was assessed using computerized tomography (CT) scans every two pemetrexed-based chemotherapy cycles and categorized according to the Response Evaluation Criteria In Solid Tumors (RECIST) v1.1, as per institutional protocol.

The toxicities recorded during pemetrexed-based treatment were graded according to the Common Terminology Criteria for Adverse Events (CTCAE) version 4.0.

TS enhancer region genotyping analysis

The genomic DNA was extracted from the peripheral leukocytes. The genotypes of the TSER (VNTR) and SNP were determined by polymerase chain reaction (PCR). The variable number tandem repeat (VNTR) of 28 bp polymorphism and the G → C SNP in the first and second repeat were analyzed. A DNA fragment was amplified using previously described PCR conditions and primers [17], and directly sequenced using an ABI PRISM 3100 Genetic Analyzer (Applied Biosystems, Foster City, CA, USA). The forward primer 5′-CGTGCTCCTGCGTTTCC-3′ and...
the reverse primer 3′-GAGCCGGCCACAGGCAT-5′ were used. A modification of conventional conditions was necessary. PCR was performed in a reaction mixture with dNTP: 0.35 μl, Buffer: 0.25 μl, MgCl₂: 17.5 μl, Tap polymerase: 0.5 μl, H₂O: 18 μl, primers 0.1 + 0.1 μl, DMSO: 1.25 μl and DNA: 2 μl. The cycling conditions were denaturation 95°C for 10 minutes, and 30 cycles at 95°C for one minute, then at 64°C for one minute, and 72°C one minute, and finally, seven minutes at 72°C. Aliquots of amplified fragments were separated on a 3% agarose gel and the TS VNTR genotype was determined, staining 2R (210 base pairs; bp) and 3R (238 bp) alleles. After that we performed a PCR-restriction fragment length polymorphisms (RFLP), by Hae III digestion. The mixture was PCR product: 10 μl, H₂O: 7 μl, Buffer 2 μl and Hae III: 1 μl. After that we incubated the mixture at 37°C overnight. Aliquots of digested fragments were separated on 12% acrylamide gel and the SNP genotype was determined. The digestion of fragments showed the different genotypes 2RGC: 66, 47, 46, 44 and 7 bp, 2RCC: 113, 46, 44 and 7 bp, 3RGGCC (3RG): 66, 47, 46, 44 and 7 bp, 2R (210 base pairs; bp) and 3R (238 bp) alleles. After that we performed a PCR-restriction fragment length polymorphisms (RFLP), by Hae III digestion. The mixture was PCR product: 10 μl, H₂O: 7 μl, Buffer 2 μl and Hae III: 1 μl. After that we incubated the mixture at 37°C overnight. Aliquots of digested fragments were separated on 12% acrylamide gel and the SNP genotype was determined. The digestion of fragments showed the different genotypes 2RGC: 66, 47, 46, 44 and 7 bp, 2RCC: 113, 46, 44 and 7 bp, 3RGGCC (3RG): 66, 47, 46, 44 and 7 bp, 2R (210 base pairs; bp) and 3R (238 bp) alleles. After that we performed a PCR-restriction fragment length polymorphisms (RFLP), by Hae III digestion. The mixture was PCR product: 10 μl, H₂O: 7 μl, Buffer 2 μl and Hae III: 1 μl. After that we incubated the mixture at 37°C overnight. Aliquots of digested fragments were separated on 12% acrylamide gel and the SNP genotype was determined. The digestion of fragments showed the different genotypes 2RGC: 66, 47, 46, 44 and 7 bp, 2RCC: 113, 46, 44 and 7 bp, 3RGGCC (3RG): 66, 47, 46, 44 and 7 bp.

**TS 3′UTR region genotyping analysis**

The 3′UTR polymorphisms were analyzed by Restriction Fragment Length Polymorphism (RFLP). A fragment containing the 6 bp deletion/insertion was amplified using the reverse primer 5′-CAGATAAGTGGCAGTA-CAGA-3′ and the forward primer 3′-CAAATCTGAGGGAGCTGAGT-5′ in 10 μl of reaction mixture with dNTP: 4 μl, Buffer: 5 μl, MgCl₂: 4 μl, Tap polymerase: 0.5 μl, H₂O: 26.5 μl, primers 4 + 4 μl and DNA: 2 μl. The cycling conditions were denaturation 95°C for 10 minutes, and 35 cycles at 95°C for 30 minutes, then at 57°C for 30 minutes, and 72°C one minute, and finally, seven minutes at 72°C. The fragments were amplified on 2% agarose gel. Afterwards the products were digested with Dra I and the mixture of PCR product: 20 μl, BSA 10%: 0.5 μl, Buffer: 5 μl, H₂O: 23.5 μl and Dra I: 1 μl. Posteriorly, the product was incubated one hour at 37°C. The final digested product was separated in a 3% agarose gel. The different genotypes were deletion 6 bp/insertion 6 bp, insertion 6 bp/insertion 6 bp and deletion 6 bp/deletion 6 bp. The expected fragment sizes by genotyping were deletion 6 bp/insertion 6 bp: 148, 142, 88 and 60 bp, insertion 6 bp/insertion 6 bp: 88 and 60 bp, and deletion 6 bp/deletion 6 bp: 142 bp. We repeated the PCR three times to ensure final results.

**EGFR mutations analysis**

As per institutional protocol, all patients with advanced NSCLC were tested for *EGFR* activating mutations before treatment initiation. In brief, after having the samples fixed in alcohol and stained by Papanicolaou stain, DNA was extracted and amplified by PCR technique, using *EGFR* gene exons 18, 19, 20 and 21 specific primers. ABI PRISM® 310 Genetic Analyzer equipment was used for the analysis of the sequencing reactions with both forward and reverse primers.

**Statistical analysis**

Fisher’s exact test was used to investigate the correlation between each genotype and the response to the treatment and the toxicity presented. Kaplan-Meier curves and log-rank test or Tarone-Ware test, when indicated, were calculated to correlate each genotype with the survival outcomes (PFS and OS). For the subgroup analysis, *EGFR* mutation status and smoking history were considered in order to analyze potential differences in clinical outcome measures (ORR, PFS and OS).

The SPSS 15.0 software (SPSS, Inc., Chicago, IL) was employed to perform the statistical analysis.

**Results**

**Patients’ characteristics and treatment**

The clinical and pathological characteristics of the patients included are summarized in Table 1. In brief, our cohort was mainly composed by males with a median age of 59 years and a past smoking history showing good performance status. Most of the patients showed adenocarcinoma histology (88%) and showed distant metastasis (M1) at onset (72%). Most of the patients received a pemetrexed-based regimen in first line (84%). After a median follow up of 21 months, 80% of patients have already progressed and 52% of them have died due to disease progression (Table 1).

In addition, in 8 out of the 18 subjects showing multiple brain metastases at onset, conventional whole-brain radiotherapy (300 cGy) was administered between first and second chemotherapy cycles, following our institutional treatment guidelines.

Finally, 4 out of the 7 patients showing no distant metastases at onset responded to the pemetrexed-based induction chemotherapy. As per institutional protocol, all four subjects underwent a 3-D conformal radiotherapy program with concurrent chemotherapy, as previously published [18].

**Correlation between ORR to the treatment and polymorphisms**

We studied the potential correlation between the different polymorphisms observed and the response to the treatment obtained (Table 2). For this purpose, any kind of radiological response (complete or partial response), was compared to no response to the treatment (disease stabilization or progression). The presence of 3R/3R polymorphism seemed to predict a higher ORR (100%), compared to the rest of the genotypes with a trend toward statistical significance (p = 0.055). In the subgroup
analysis, a significantly higher ORR to pemetrexed for wild-type EGFR patients showing a 3R/3R genotype (100%) compared to the 2R/2R (77.8%), 2R/3R (33.3%) and 3R/4R (0%) was observed (p = 0.017).

No statistically significant differences were observed comparing the presence and the absence of a SNP G > C as shown in Table 2.

Overall, a non-significant correlation between the different 3′-UTR polymorphisms and the ORR was observed. However, the genotype +6/+6 seemed to predict a higher ORR among active/former smokers (A/FS) compared to +6/-6 (100% vs. 50%; p = 0.085).

Correlation between PFS and polymorphisms

Regarding TSER polymorphisms we found a trend toward statistical significance (p = 0.089) in the differences in PFS observed among the different genotypes in favor of the 3R/3R genotype, (Figure 1A).

In the case of the absence or presence of a SNP at the third repetition (3R allele), we observed a non-significant increased PFS in the subgroup of patients showing an absence of SNP (Figure 1B).

Finally, no significant correlations regarding the 3′UTR genotypes and PFS were observed, (Figure 1C).

Correlation between OS and polymorphisms

In this cohort, we found a significant correlation between TSER polymorphisms and OS (Figure 2A). The median OS was not reached for 3R/3R genotype patients, whereas 2R/3R genotype subjects showed a 70 m
OS followed by 3R/4R and 2R/2R genotypes with a median OS of 15 m and 13 m, respectively (p = 0.019), (Figure 2A).

No significant differences in OS were observed with regards to the presence/absence of SNP (Figure 2B) or regarding the 3′-UTR polymorphisms (Figure 2C).

**Correlation between toxicity and polymorphisms**

The most frequent toxicity was grade (G)1 anemia (28%) and nausea (20%) and G2 leucopenia (40%). The most common G3-4 toxicities were leucopenia (16%), asthenia (8%), anemia (4%), neutropenia (4%) and dyspnea (4%). Overall, we found no significant correlations between the toxicity profiles experienced by the patients and the different TS genotypes (Table 3).

**Discussion**

Pemetrexed, a multitargeted antifolate drug, is essential for the first and second-line as well as maintenance treatment of NSCLC patients with non-squamous histology [6]. TS is the main biological target of pemetrexed. Some studies have suggested that TS expression could be a predictive factor of response in NSCLC [19]. Moreover, some VNTR genotypes have been associated with TS expression and activity in other tumor types such as colorectal cancer [17].

In NSCLC patients, a correlation between different genotypes and the TS protein expression has been shown [20]. Shintani et al. [20] also confirmed that the TS mRNA levels were significantly higher in lung cancer tissues with the 3R/3R genotype as compared to those with the 2R/2R genotype.

Nonetheless, definitive studies addressing the correlation of the different genotypes of TS in circulating genomic DNA with response to the treatment, PFS or OS in pemetrexed-treated NSCLC European patients are lacking. The potential influence of the EGFR status on those polymorphisms and their correlation with clinical outcome after pemetrexed-based treatment is also unexplored.

A recent study by Hu et al. [21] investigated the different TS polymorphisms in genomic DNA of 90 Asian NSCLC patients. In contrast with our findings, no specific genotype regarding the TSER or 3′-UTR polymorphisms studied seemed to correlate with a significant difference in ORR, PFS or OS. This could be explained by substantial clinical differences between both populations. Our cohort was constituted by Caucasian patients compared to the Asian population studied by Hu et al. In addition, our patients were mostly current or former smokers (72%) compared to the Asian population that showed 62% of never smokers. Also in our cohort, the subjects mainly received pemetrexed-based chemotherapy as a first line regimen (92%), whereas the cohort studied by Hu et al. [21] was treated with pemetrexed as a second or further line in 62.2% of the cases. These remarkable differences in basic clinical characteristics, and in particular the ethnicity, between both cohorts are probably also explaining the differences observed in the 3′-UTR genotype frequency between our population and the one studied by Hu et al. In our cohort, +6 bp/+6 bp, +6 bp/-6 bp and -6 bp/-6 bp genotypes were found in 48%, 44% and 8% of the cases, respectively. In contrast, 7.8%, 47.8% and 44.4% were respectively found in the population studied by Hu et al. [21]. In a previous analysis performed on another Caucasian NSCLC population evaluated at the M.D. Anderson Cancer Center [22], a similar proportion of 3′-UTR genotypes according to our findings was observed (49.2% of +6 bp/+6 bp, 42.4% of +6 bp/-6 bp and 8.4% of -6 bp/-6 bp). Additionally, the low prevalence of the +6 bp/+6 bp genotype in an Asian population compared to our cohort may be confirmed by a recent study in which from 106 Asian NSCLC patients investigated, none of them showed a +6 bp/+6 bp genotype in genomic circulating DNA [23]. Nonetheless, in this latter study [23] a significantly higher ORR was observed among patients showing a -6 bp/-6 bp 3′-UTR genotype compared to the ORR reported for patients presenting a -6 bp/+6 bp
polymorphism (32.2% vs 12.7%; p = 0.008). Accordingly, in our cohort, a higher ORR in patients showing a −6 bp/−6 bp genotype compared to those presenting a −6 bp/+6 bp polymorphism was also observed (100% vs. 54.5%). However, the statistical significance was not reached probably due to the relatively low number of patients included in our analysis. Interestingly enough, in the subgroup analysis of our data, the +6 bp/+6 bp genotype seemed to predict a higher ORR only among active/former smokers compared to +6 bp/−6 bp (100% vs. 50%; p = 0.085). This novel observation, if validated in future studies, could be relevant for selecting specific drugs for each patient in a second or third line setting.

With regards to the TSER polymorphisms, the presence of a 3R/3R polymorphism seemed to predict a higher ORR with a clear trend toward statistical significance (p = 0.055). Moreover, that difference was even greater and statistically significant benefiting the subpopulation of wild-type EGFR patients. To our knowledge, this is the first time that such observation has been made. An interesting preclinical study by Giovannetti et al. [24] investigated the activity profile of a combination therapy against NSCLC cell lines with different genotypes with Erlotinib and pemetrexed. Remarkably, pemetrexed increased EGFR phosphorylation and reduced Akt phosphorylation. Additionally, erlotinib significantly reduced TS expression and activity. Thus, when erlotinib and pemetrexed were combined, a strong synergism in all NSCLC cells, regardless of their genetic signature, was observed. This potential crosstalk between the EGFR signaling pathway and the TS expression and activity could in part explain our novel findings showing a significantly higher ORR to pemetrexed in those wild-type EGFR patients harboring a 3R/3R polymorphism. However, none of the previous studies have described the EGFR status of the patients analyzed and how that status impacted on the ORR to pemetrexed for certain TS polymorphisms.

In terms of survival, in the present series after a median follow-up of 21 months, PFS was superior for those
Table 3 Correlation between grades of toxicity and different genotypes

| Global distribution polymorphisms (Pol) | No toxicity | Grade 1-2 | Grade 3-4 | p value |
|----------------------------------------|-------------|-----------|-----------|---------|
| VNTR polymorphisms                     |             |           |           |         |
| 2R/2R                                  | 2 (22.2)    | 4 (44.2)  | 3 (33.4)  | 0.545   |
| 3R/3R                                  | 2 (25)      | 5 (75)    | 0 (0)     |         |
| 2R/3R                                  | 2 (25)      | 4 (50)    | 2 (25)    |         |
| 3R/4R                                  | 1 (100)     | 0 (0)     | 0 (0)     |         |
| SNP polymorphisms                      |             |           |           |         |
| Absence                                | 3 (42.9)    | 4 (57.1)  | 0 (0)     | 0.3     |
| Presence                               | 4 (22.2)    | 9 (50)    | 5 (27.8)  |         |
| 3’-UTR polymorphisms                   |             |           |           |         |
| +6/+6                                  | 3 (25)      | 6 (50)    | 3 (25)    | > 0.05  |
| +6/-6                                  | 3 (27.3)    | 6 (54.5)  | 2 (18.2)  |         |
| −6/-6                                  | 1 (50)      | 1 (50)    | 0 (0)     |         |
patients showing a 3R/3R genotype with a trend toward statistical significance, as expected considering the higher ORR observed for patients with the same TSER genotype. The relatively low statistical power of our clinical cohort may be accounting for the lack of a full statistical significance observed.

Regarding OS, the advantage in PFS observed in patients showing the 3R/3R genotype translated into a significantly higher median OS in patients with the same polymorphism compared to the rest. Conversely, in the study by Hu et al. [21] no specific genotype was significantly correlated with a superior PFS or OS. As aforementioned, the dramatic differences in the population's characteristics between both series might possibly explain this discordance. In our study, conversely to the observations made by Wang et al. [23] a significantly superior PFS and OS in patients with the −6 bp/−6 bp 3′-UTR genotype has not been confirmed. Most probably, this is also due to the differences in the genotype distribution among populations with markedly different ethnicity and epidemiological characteristics.

Finally, in accordance with Wang et al. [23], the toxicity profile was not significantly correlated with any TS genotype in our series. As aforementioned, this study has some limitations due to its retrospective nature and the low number of patients investigated. Both could be responsible for a low statistical power that may impair our ability to find significant differences between subgroups of patients.

Conclusions
For the first time in a European population of NSCLC patients receiving pemetrexed, the presence of the TSER 3R/3R polymorphism significantly correlated with a superior OS. Moreover, this same polymorphism, when associated to wild-type EGFR, was correlated with a higher ORR to pemetrexed. The presence of the +6/-6 bp 3′-UTR genotype among active or former smokers was correlated to a higher ORR showing a trend toward statistical significance. Finally, pemetrexed-induced toxicity was not significantly correlated with a specific TS genotype.

These novel data warrant further investigation in larger prospective series and may help to patient's selection if finally validated.

Abbreviations
NSCLC: Non-small cell lung cancer; TS: Thymidylate synthase; EGFR: Epidermal Growth Factor Receptor; VNTR: Variable number of tandem repeats; TSER: Thymidylate Synthase gene promoter enhancer region; SNP: Single nucleotide polymorphism; ORR: Overall response rate; PFS: Progression-free survival; OS: Overall survival; CT: Computerized tomography; RECIST: Response Evaluation Criteria in Solid Tumors; CTCAE: Common Terminology Criteria for Adverse Events; PCR: Polymerase chain reaction; RFLP: Restriction fragment length polymorphism; A/FS: Active/former smokers.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
EA participated in the design of the study, contributed to the patients’ identification and medical history charts revision and performed samples’ processing and statistical analyses as well as manuscript drafting. EC participated in the design of the study, contributed to the patients’ identification and medical history charts revision and performed samples’ processing and statistical analyses as well as manuscript drafting. IL carried out samples’ processing and laboratory analysis. JS participated in the design of the study and contributed to the laboratory analysis. VC contributed to samples’ processing, laboratory analysis and paper drafting. MS contributed to results’ interpretation and manuscript’s drafting. MM contributed to drafting the manuscript and revising it, contributing for important intellectual content. PM aided to conceive the study, and participated in its design and helped to draft the manuscript. LZ contributed to drafting the manuscript and revising it critically for important intellectual content. APG participated in the design of the study and contributed to the laboratory analysis. CR contributed to drafting the manuscript and revising it critically for important intellectual content. IGB conceived the study and its design, contributed to patients’ selection, drafted the manuscript and gave final approval. All authors read and approved the final version of the manuscript.

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