Influence of age and sex on the blood biochemical constituent values of broiler breeders during the egg-laying stage

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Received: 25 June 2021 / Accepted: 29 October 2021 / Published online: 11 November 2021
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Abstract
This study aimed to evaluate the influence of age and sex on the blood biochemical constituents of broiler breeders during the egg production stage. The analysis was performed in an industrial broiler breeder farm, and blood samples were collected from males and females at five different ages. Biochemical elements analysed in each serum sample were total proteins, albumin, globulins, uric acid, total cholesterol and triglycerides, alanine aminotransferase (ALT) and aspartate aminotransferase (AST), gamma-glutamyltransferase (GGT), creatine kinase (CK), alkaline phosphatase (PAL), calcium and phosphorus, beside the glycaemic status. At most ages, females had higher values of total proteins, albumin, globulins, triglycerides, cholesterol, glucose, calcium, phosphorus, Ca/P (calcium and phosphorus) ratio and gamma-glutamyltransferase (GGT). The values of uric acid, aspartate aminotransferase (AST), creatine kinase (CK) and alkaline phosphatase (PAL) were higher in males. The lowest protein values were found at 28 and 60 weeks old. The mean albumin values were significantly higher at 44 and 52 weeks old in males and females. A trend of increasing globulin values with increasing age up to 52 weeks old was observed. Although calcium and phosphorus did not vary according to age, the Ca/P ratio was lowest at 36 weeks old. Comparing the means of both sexes, the AST and GGT values were significantly higher in 60-week-old birds. The highest serum levels of CK occurred at 28 and 52 weeks old. The physiological levels of serum biomarkers presented in this work are important for evaluating productivity performance, welfare and disease indication in breeding flocks.

Keywords Protein · Metabolites · Minerals · Liver enzymes · Skeletal muscle enzymes · Blood

Introduction
The Brazilian poultry industry has been outstanding in recent years due to genetic advances in obtaining broilers with excellent weight gain, high meat yield and low feed conversion efficiency (Schmidt and Silva 2018).

Chicken productivity characteristics, such as high performance for weight gain, are transmitted by their male and female progenitors. Thus, the rearing of progenitors should be conducted to prioritize their reproductive characteristics. For females, these characteristics should aim to maximize fertile egg production with low mortality, and for males, these characteristics should guarantee good fertility and result in satisfactory hatch rates and progeny quality (Saxena and Kolluri 2018).

The performance of breeders is closely related to their environment, reflecting their individual interactions with exogenous factors such as environment, food, management and social relationships. Knowledge of the best physiological
responses to these complex interactions is important in obtaining the best performance, as this is an indicator of animal welfare.

Veterinary clinical pathology has stood out as an excellent tool for disease diagnosis, especially those derived from infectious and metabolic origins. Several studies in the poultry area have investigated some blood biochemical components to identify lines with better productive performance. The use of alanine aminotransferase (AST) as a biochemical marker for the disease known as “fatty liver syndrome” and the abdominal fat correlation with very-low-density lipoprotein (VLDL) serum levels are examples of applications of clinical pathology for the selection of animals with better productive performance (Hermier 1997; So et al. 2009).

Although useful, blood biochemical components as disease markers in poultry are still insufficiently explored. Specifically, the study of characteristics that indicate high-performance birds of different ages for both sexes during the production period has not been explored in the literature. This study aimed to determine the physiological variations in certain blood biochemical constituents in male and female broiler breeders at the egg-laying stage, which is between 28 and 60 weeks old, and the influence of sex.

Material and methods

This study was performed in healthy broiler breeders of Cobb lineage at an industrial production plant in Uberlândia, Minas Gerais, Brazil. The birds were raised in negative pressure flock at approximately 78.8 °F, and they remained healthy during all production periods. The flocks’ egg production followed the standards recommended by the line age. During the growing period, these birds were vaccinated for major bird diseases (Marek’s Disease, New Castle Disease, Infectious bronchitis, avian metapneumovirus, avian bobra, coccidiosis, infectious anaemia, encephalomyelitis and Gumboro’s disease). Bird health was measured by nor- mal behaviour, standard egg production, fertility rate and egg hatchability. In addition, health monitoring was carried out every 12 weeks for the diseases mentioned above and Mycoplasma gallisepticum, M. synoviae and Salmonella spp. using serologic analysis or bacteriologic analysis. In the entire period of the experiment, the birds were healthy.

During the egg-laying stage, females consumed layer feed number 1 from 25–48 weeks of age and layer feed 2 after 49 weeks of age (Supplementary table). Males consumed growth feed until 27 weeks of age and rooster feed from 28 weeks of age until the end of production (Supplementary table).

Thirty Cobb birds were randomly selected, with 15 females and 15 males in each of the following age groups: 28, 36, 44, 52 and 60 weeks old. This resulted in a total of 150 birds. Six hours after feed intake, the birds were weighed, and then, 3 mL of blood was collected from each bird through ulnar vein puncture using disposable needles and syringes. As this manuscript is about the standardization of physiological variations in certain blood biochemical constituents in male and female broiler breeders at the egg-laying stage, a control group was not necessary. All the collection process was approved by the Ethics Committee on the Use of Animals at the Federal University of Uberlândia.

Blood samples were transferred to sterile tubes without anticoagulant (dry tubes) immediately after collection. On the same day, the samples were sent to the Clinical Laboratory of the Veterinary Hospital of the Federal University of Uberlândia in an isothermal box and centrifuged at 720 ×g for 10 min. The sera were separated and stored in microtubes at refrigeration temperature (35.6 to 46.4 °F) for 24 h until the biochemical analysis procedure.

Biochemical elements analysed in each serum sample were: total proteins by Biuret’s method (Gornall et al. 1949), albumin by the bromocresol green method (Doumas et al. 1997), globulin calculated as the difference between the total protein value and albumin, uric acid by the Trinder enzyme method (Kageyama 1971), total cholesterol and triglycerides by the Trinder enzyme method (Sharma et al. 1987), alanine aminotransferase (ALT) and aspartate aminotransferase (AST) by the UV-IFCC kinetic method (Bergmeyer et al. 1978), gamma-glutamyl transferase (GGT) by the modified Szasz method (Szasz 1969), creatine kinase (CK) by the UV kinetic method (Schumann et al. 2002), alkaline phosphatase (PAL) by the modified Bowers and McComb method (Bowers and McComb 1966), calcium by the cresolphthalein complex-one method (Connerty and Briggs 1966) and phosphorus by the modified Daly and Ertingshausen method (Daly and Ertingshausen 1972).

A previously calibrated Labmax Plenno® automatic analyzer (Calibra H) (Labtest, Brazil) was used and calibrated with universal control serum (Qualitrol 1), and commercial Labtest Diagnostica® kits were also used. The glycaemic status of each bird in the five age groups was determined shortly after the blood sample collection using the commercial One Touch® Ultra™ glucometer.

The experimental model adopted was a completely randomized design in a factorial scheme composed of two sexes, five ages and 15 replicates, totalling 150 samples. The Kolmogorov-Smirnov test with 5% significance was applied for the normality test, and the data showed normality. A t-test was used to compare differences between the sexes, and ANOVA was used to compare ages. Where there were significant differences, treatment means were separated by Tukey’s test at 5% (P < 0.05). GraphPad Prism 5.1 software was used.
Results

In this study, males and females had a weight gain compatible with their lineage (Cobb-Vantress 2018), with males having significantly higher weight gain than females at all ages (Table 1). The total protein, albumin, globulin, triglyceride and cholesterol serum levels were higher in females than in males at most evaluated ages (Table 1). Regarding age, the lowest protein values in females and males were found at the beginning (28 weeks old) and the end (60 weeks old) of the egg-laying stage. The mean albumin values of males and females were significantly higher at 44 and 52 weeks old. For both females and males, a trend of increasing globulin values with increasing age up to 52 weeks old was observed. Males had higher uric acid levels than females, and the value of uric acid in males and females increased with age. Regarding the age groups, the mean serum triglyceride levels in females and males were similar at all studied ages (Table 1). Females had higher glucose values than males at all studied ages, but there was no difference among ages (Table 1).

The serum concentrations of calcium and phosphorus and the Ca/P ratio were significantly higher in females than in males (Table 2). Serum calcium and phosphorus concentrations in females and males did not vary according to age. Among the age groups, the Ca/P ratio was lowest at 36 weeks old (Table 2).

For ALT, there were no significant differences between males and females in any of the different age groups (Table 2). Higher values of AST and CK were observed in males than in females at 28 and 36 weeks old. Comparing the means of both sexes, enzyme values were significantly higher in 60-week-old birds (Table 2). CK had the highest

### Table 1

Mean values and standard deviations of weight, protein blood concentrations, metabolites and nutrients in broiler breeder males and females during the egg-laying stage

| Parameter          | Sex     | Age (weeks) |
|--------------------|---------|-------------|
|                    | 28      | 36          | 44          | 52          | 60          |
| Weight (g)         |         |             |             |             |             |
| F                  | 3426.67 ± 132.98 | 3728.33 ± 97.88 | 3931.45 ± 127.08 | 4001.33 ± 115.16 | 4003.21 ± 160.57 |
| M                  | 3960.33 ± 115.21 | 4383.00 ± 9.12  | 4626.28 ± 115.12 | 4644.67 ± 100.06 | 4808.33 ± 166.33 |
| F and M            | 3693.50 ± 297.66 | 4055.67 ± 345.40 | 4278.87 ± 372.90 | 4323.00 ± 343.91 | 4405.33 ± 440.24 |
| Total proteins (g/dL−1) |         |             |             |             |             |
| F                  | 5.01 ± 0.69  | 5.38 ± 0.67  | 5.66 ± 0.46  | 6.01 ± 0.53  | 5.08 ± 0.34  |
| M                  | 3.61 ± 0.31  | 5.01 ± 0.68  | 4.35 ± 0.36  | 5.08 ± 0.51  | 4.57 ± 0.67  |
| F and M            | 4.31 ± 0.89  | 5.19 ± 0.69  | 5.08 ± 0.76  | 5.55 ± 0.70  | 4.83 ± 0.57  |
| Albumin (g/dL−1)   |         |             |             |             |             |
| F                  | 2.09 ± 0.25  | 1.93 ± 0.26  | 2.37 ± 0.19  | 2.23 ± 0.18  | 1.90 ± 0.13  |
| M                  | 1.37 ± 0.12  | 1.50 ± 0.13  | 1.73 ± 0.12  | 1.77 ± 0.16  | 1.54 ± 0.23  |
| F and M            | 1.74 ± 0.41  | 1.71 ± 0.32  | 2.08 ± 0.37  | 2.00 ± 0.28  | 1.73 ± 0.26  |
| Globulins (g/dL−1) |         |             |             |             |             |
| A/G ratio F        | 2.92 ± 0.56 | 2.80 ± 0.16 | 3.30 ± 0.41 | 3.79 ± 0.48 | 3.17 ± 0.26 |
| M                  | 2.34 ± 0.52 | 3.34 ± 0.47 | 2.63 ± 0.31 | 3.30 ± 0.56 | 3.03 ± 0.50 |
| F and M            | 2.63 ± 0.57 | 3.07 ± 0.45 | 3.00 ± 0.47 | 3.55 ± 0.57 | 3.10 ± 0.39 |
| F                  | 0.73 ± 0.13 | 0.69 ± 0.11 | 0.73 ± 0.11 | 0.60 ± 0.09 | 0.60 ± 0.04 |
| M                  | 0.58 ± 0.19 | 0.46 ± 0.07 | 0.66 ± 0.08 | 0.55 ± 0.10 | 0.51 ± 0.07 |
| F and M            | 0.66 ± 0.13 | 0.57 ± 0.17 | 0.70 ± 0.11 | 0.57 ± 0.10 | 0.56 ± 0.07 |
| Uric acid (mg/dL−1) |         |             |             |             |             |
| F                  | 5.65 ± 0.96 | 5.22 ± 0.91 | 7.26 ± 1.41 | 6.44 ± 1.32 | 7.58 ± 1.85 |
| M                  | 7.94 ± 3.31 | 7.51 ± 2.89 | 11.07 ± 3.28 | 14.13 ± 3.36 | 11.79 ± 2.84 |
| F and M            | 6.79 ± 2.66 | 6.36 ± 2.34 | 8.69 ± 2.75 | 10.29 ± 4.64 | 9.61 ± 3.12 |
| Triglycerides (mg/dL−1) |        |             |             |             |             |
| F                  | 1120.07 ± 205.97 | 1589.57 ± 246.10 | 1556.40 ± 131.70 | 1538.60 ± 66.66 | 1380.33 ± 215.36 |
| M                  | 77.07 ± 18.45 | 102.86 ± 48.84 | 94.88 ± 24.83 | 92.33 ± 17.45 | 97.25 ± 21.62 |
| F and M            | 597.57 ± 549.53 | 846.21 ± 776.43 | 1048.04 ± 747.14 | 815.47 ± 737.05 | 810.07 ± 668.80 |
| Cholesterol (mg/dL−1) |       |             |             |             |             |
| F                  | 155.13 ± 56.12 | 170.29 ± 36.97 | 187.60 ± 40.12 | 182.47 ± 37.03 | 145.67 ± 35.66 |
| M                  | 94.07 ± 12.74 | 123.00 ± 17.07 | 110.00 ± 13.01 | 119.00 ± 21.56 | 113.64 ± 23.85 |
| F and M            | 124.60 ± 50.63 | 146.64 ± 36.35 | 158.50 ± 48.88 | 150.73 ± 43.91 | 130.21 ± 33.86 |
| Glucose (mg/dL−1)  |        |             |             |             |             |
| F                  | 250.40 ± 16.33 | 248.60 ± 28.16 | 256.53 ± 19.21 | 238.80 ± 16.11 | 248.67 ± 19.08 |
| M                  | 220.80 ± 19.00 | 200.13 ± 16.08 | 209.27 ± 22.22 | 214.93 ± 18.78 | 195.47 ± 16.72 |
| F and M            | 235.60 ± 23.02 | 224.37 ± 33.40 | 232.90 ± 31.53 | 226.87 ± 21.05 | 222.07 ± 32.29 |

Means followed by different capital letters in the same column or lowercase in the same row for the same parameter evaluated differ by the Tukey test (p < 0.05). F, females; M, males
serum levels at 36 and 52 weeks old in females and males (Table 2). There were higher serum concentrations of GGT in 52- and 60-week-old females at the end of the reproductive period, and the average increase for both the sexes in these age groups (Table 2). When comparing the serum levels of PAL in this study, the males had higher levels than females at most of the studied ages, except for 28-week-old birds, where males and females had the same levels at the beginning of the egg-laying stage. Evaluating the means obtained from females and males, the highest value of PAL was found at 28 weeks old (Table 2).

Discussion

Biochemical markers in the blood of high-performance broiler breeders during the egg-laying stage can be an essential tool for future interpretation of these biological markers for performance or even disease in these animals.

In this study, males and females had a weight gain compatible with their lineage (Cobb-Vantress 2018), with males having significantly higher weight gain than females at all ages (Table 1). This was an expected result because the birds on this farm were carefully raised following standardized management and health methods.

Protein serum levels in females were higher than those in males at most of the evaluated ages, except for 36 weeks, in which the values were similar between sexes. The female’s high protein demand for egg production (Penz and Jensen 1991; Capitelli and Crosta 2013) can explain this result. The peak of egg production at 36 weeks can explain the similarity of serum protein values between males and females at this time. During peak production, females need a large amount of protein to form eggs (National Research Council 1994), which can reduce the amount of serum protein. Considering the total protein mean of females and males, the lowest values were found at the beginning (28 weeks old) and the

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**Table 2** Mean values and standard deviations for mineral and enzyme serum concentrations in broiler breeder males and females during the egg-laying stage

| Parameter | Sex | AGE (weeks) | 28 | 36 | 44 | 52 | 60 |
|-----------|-----|-------------|----|----|----|----|----|
| Calcium (mg/dL⁻¹) | F | 31.20A ± 7.91 | 27.94A ± 4.03 | 33.12A ± 2.98 | 31.96A ± 3.68 | 26.24A ± 3.87 |
| | M | 12.53B ± 1.43 | 10.70B ± 1.31 | 12.60B ± 0.85 | 14.40B ± 0.73 | 13.26B ± 5.95 |
| | F and M | 21.87a ± 11.02 | 19.32a ± 9.22 | 25.42 a ± 10.57 | 23.19 a ± 9.29 | 19.97 a ± 8.19 |
| Phosphorus (mg/dL⁻¹) | F | 9.46A ± 3.18 | 9.50B ± 1.75 | 9.87A ± 1.68 | 10.47A ± 1.81 | 8.01A ± 1.64 |
| | M | 4.88B ± 0.76 | 5.91B ± 0.67 | 4.16B ± 0.48 | 4.95B ± 0.52 | 4.48B ± 2.26 |
| | F and M | 7.17a ± 3.25 | 7.71a ± 2.23 | 7.28a ± 3.13 | 7.71a ± 3.10 | 6.31a ± 2.62 |
| Ca/P ratio | F | 3.39A ± 0.58 | 2.96A ± 0.25 | 3.44A ± 0.60 | 3.09A ± 0.34 | 3.33A ± 0.61 |
| | M | 2.61B ± 0.41 | 1.82B ± 0.24 | 3.07A ± 0.45 | 2.94A ± 0.29 | 2.70B ± 0.53 |
| | F and M | 3.00a ± 0.63 | 2.39b ± 0.62 | 3.30a ± 0.53 | 3.01a ± 0.32 | 3.19a ± 0.48 |
| ALT (UI/L⁻¹) | F | 10.46A ± 4.45 | 10.73A ± 2.41 | 11.23A ± 3.65 | 9.80A ± 4.11 | 11.27A ± 2.02 |
| | M | 9.93A ± 1.16 | 9.36A ± 1.01 | 8.78A ± 2.73 | 10.60A ± 3.54 | 13.00A ± 3.14 |
| | F and M | 12.47A ± 11.85 | 9.96A ± 1.75 | 10.38A ± 3.41 | 10.20a ± 3.79 | 12.07a ± 2.70 |
| AST (UI/L⁻¹) | F | 103.60B ± 32.07 | 203.64B ± 22.50 | 137.93A ± 34.43 | 206.67A ± 70.20 | 269.00A ± 27.96 |
| | M | 145.80a ± 35.50 | 278.64A ± 19.65 | 106.78A ± 31.53 | 85.07B ± 35.82 | 274.14A ± 23.63 |
| | F and M | 124.70c ± 39.57 | 241.14b ± 43.05 | 126.25c ± 33.24 | 145.87c ± 82.60 | 271.66 a ± 25.08 |
| CK (UI/L⁻¹) | F | 3800.35B ± 1822.05 | 5312.25B ± 1650.80 | 5410.30A ± 1841.11 | 7388.99A ± 4738.20 | 4252.53A ± 1132.60 |
| | M | 6652.80A ± 3308.14 | 8758.88A ± 2299.59 | 6666.39A ± 3106.51 | 10087.43A ± 4285.64 | 6284.34A ± 3517.50 |
| | F and M | 5177.39b ± 2934.76 | 7035.56ab ± 2581.52 | 6152.54b ± 2496.47 | 8738.21a ± 4646.29 | 5233.40b ± 2707.83 |
| GGT (UI/L⁻¹) | F | 19.67A ± 3.31 | 15.71A ± 3.29 | 30.33A ± 1.59 | 43.60A ± 25.23 | 62.60A ± 37.22 |
| | M | 20.40A ± 3.72 | 17.07A ± 9.36 | 34.22A ± 9.16 | 24.07B ± 3.13 | 30.64B ± 8.98 |
| | F and M | 20.03cd ± 3.48 | 16.39d ± 6.68 | 31.79bc ± 5.59 | 33.83bc ± 20.27 | 47.17a ± 31.13 |
| PAL (UI/L⁻¹) | F | 1017.27A ± 602.74 | 212.93B ± 78.32 | 261.67B ± 81.86 | 181.73B ± 38.69 | 642.14B ± 338.11 |
| | M | 1040.87A ± 671.57 | 937.29A ± 84.92 | 1198.11A ± 1144.14 | 551.13A ± 518.79 | 989.57A ± 484.73 |
| | F and M | 1029.07a ± 627.10 | 575.11bc ± 376.16 | 612.83bc ± 174.03 | 366.43c ± 407.37 | 815.86abc ± 432.20 |

Means followed by different capital letters in the same column or lowercase in the same row for the same parameter evaluated differ by the Tukey test (p < 0.05). F, females; M, males.
end (60 weeks old) of the egg-laying stage. This event probably occurred because of lower egg production in these two stages.

Albumin accounts for 40–50% of the total plasma protein in birds, with regular levels ranging from 0.8 to 2.0 g/dL (Schmidt et al. 2007). Serum values presented by females were higher than those obtained by males. The high protein level that the females demand during egg production and the lower percentage of protein and amino acid levels in the male feed (Supplementary table) also reflected the lower values found for male serum albumin.

At most of the evaluated ages, the serum globulin content was higher in females than in males. Globulins consist of alpha globulins, beta globulins and gammaglobulins. This group of globulins includes acute phase proteins (α-globulins and β-globulins) and immunoglobulins (γ-globulins) (Melillo 2013). Alpha globulins are involved in acute response processes during trauma, inflammation or infection situations, whereas beta globulins still play an unknown clinical role (Melillo 2013). During the egg-laying stage, oestrogen-induced hyperproteinaemia occurs to complete egg formation because most of the yolk proteins are globulins and cause a marked increase in globulin fractions (Campbell and Dein 1984; Capitelli and Crosta 2013). Although albumin and globulin increased in laying females compared to males, the A/G ratio was higher in females. This indicates that the female needs more albumin than male to egg production.

When evaluating females and males, a trend of increasing globulin values with increasing age up to 52 weeks old was observed. Studies in cattle also identified increased globulin serum levels with increasing age (Liberg 1977). Hasegawa et al. (2002) studied the serum protein biochemical profile in broiler breeders and found a value of 2.7 g/dL for globulins in birds at 63 weeks old. This value is near the value found by us.

Males had higher uric acid levels than females (Table 1). Several authors have seen that roosters need lower protein levels than females to improve reproductive performance (Hocking 1990; Silveira et al. 2014), with an intake need of approximately 12% (Hocking 1990). Thus, the feed intake in this study had higher protein content (14%) than in other studies, and this may have resulted in higher nitrogen metabolite production in males. Another possible explanation for the lower level of uric acid in females may be related to the need for an increase in the amount of protein for egg formation, which results in less excretion of this metabolite. In this study, the uric acid was increased according to age in these birds, and the values observed in 52- and 60-week-old birds were higher than those seen in 28- and 36-week-old birds (Table 1). Increased excretion of uric acid in males and females in the final reproductive stage indicates that the same protein intake was associated with lower production.

Serum triglyceride and cholesterol levels were significantly higher in females than in males (Table 1). During egg laying, oestrogen increases hepatic lipid production, mainly triglycerides, to establish a reserve of energy for the embryo (Walzern 1996). A significant increase in cholesterol plasma concentrations occurs in females during the egg-laying stage due to vitellogenesis for egg yolk formation (Harr 2002). Glycaemia in avian species is 150 to 300% higher than that in mammals, considering the same body mass (Braun and Sweazee 2008). Females had higher values of glucose than males at all studied ages (Table 2), probably due to the energy requirement for egg production.

Serum calcium and phosphorus concentrations found in females were significantly higher than those in males (Table 2). These blood levels are dietary-dependent (Vinu et al. 1991). For females, calcium plays an essential role in eggshell formation; it is induced by oestrogen (Harr 2002; Dunbar et al. 2005) and transported by linked proteins, such as vitellogenin and albumin (Capitelli and Crosta 2013). Phosphorus metabolism is closely related to that of calcium, especially regarding absorption at serum levels. Higher calcium concentrations in females require proportionally higher phosphorus levels to maintain homeostasis of these two electrolytes (Proszkowic-Wege larz and Angel 2013). According to age, in the birds from this study, serum phosphorus levels did not vary between females and males. As a result, in this study, the Ca/P ratio was higher in females than males. This was mainly due to calcium serum levels, which were proportionally higher in females (Table 2). Among the age groups, the values were mostly the same, mainly due to the low variation of levels in females.

During the highest egg production stage, one of the most crucial liver roles is yolk production, but this did not result in increased ALT levels in females compared to males at different ages. Although ALT elevations are associated with hepatic or muscular lesions in most birds, even those with severe hepatic lesions do not show significant variations in serum concentration of this enzyme (Hochleithner 1994). Thus, ALT in birds may not be an important bioindicator for birds with liver problems.

AST is not considered a specific enzyme of the hepatic and muscular tissues; however, changes in this enzyme in the blood are primarily associated with disorders in these tissues. In this study, the AST values were higher in males than in females at 28 and 36 weeks of age, possibly due to more significant physical activity during the competition for females. Comparing the means of both sexes, enzyme values were significantly higher in 60-week-old birds (Table 2) and may indicate hepatic or muscular tissue alterations due to age.

Creatine kinase (CK) is a specific enzyme for skeletal muscle, and its increase in birds is mainly associated with muscle changes (Capitelli and Crosta 2013). Comparing the
results between sexes, at 28 and 36 weeks old, the serum levels of CK were higher in males than in females (Table 2). Previous studies have shown that in addition to its utility as a marker of muscle damage, CK can also be used to quantify the energy required for the reproductive strategy of each sex, aiding the study of these strategies (Ramírez et al. 2010). There are increases in CK and AST in the reproductive period, which is associated with adrenaline release, increased exercise and stress (Stout et al. 2010). The elevations in the levels of AST and CK observed in the birds of this study are consistent with the studies of Stout et al. (2010). Males, especially in the early stages of the reproductive period, experience very intense physical activity while competing with each other to form the family nucleus within the breeding stock.

Comparing the female and male means, the highest serum levels of CK occurred at 28 and 52 weeks of age (Table 2). There was management with the birds at these ages. Males underwent movement, containment and individual evaluation to select them according to their body shape to keep the capable ones in the flock. For this process, females are also involved since they are in the same environment as males. This process involves intense muscle activity, which is reflected in the increase in CK blood values.

There was a higher serum GGT concentration in 52- and 60-week-old females, which affected the average between the sexes in these age groups (Table 2). The high serum activity of GGT occurs due to increased production and release of GGT caused by hepatobiliary changes (Meyer 1995). Interestingly, simultaneous increases in GGT and AST levels at these same ages corroborate the suspicion of liver origin changes, mainly due to fatty infiltration, which can cause hepatic injury.

In this study, the males presented higher PAL levels than females at most of the evaluated ages, except at 28 weeks old. This age is the beginning of the egg-laying stage, so males and females had the same levels of PAL. PAL is related to bone metabolism, mainly with osteoblastic activity (Rajman et al. 2006). Compared with females, males had higher serum PAL values. This could possibly be due to a more significant amount of bone isoenzymes because they presented a greater amount of bone tissue due to their more developed skeleton. In another study, a higher level of PAL in males was also found when compared to females at 73 weeks old (Rath et al. 1999). When comparing the means obtained from females and males according to age, the highest value was obtained at 28 weeks old; the higher levels were mainly presented by females. This event probably occurs due to oestrogenic action, which stimulates osteoblasts to deposit calcium in the bone marrow, constituting a calcium mobilization reserve source for eggshell formation (Farmer et al. 1983). At the end of the reproductive period, an elevation of PAL levels was observed in 60-week-old birds in both females and males. There was a strong correlation with increased levels presented by females. This is probably due to the decrease in calcium absorption that occurs in older birds as oestrogen levels decrease, and, with this, there is greater bone resorption.

In breeder healthy, male and female broiler, there was a physiological difference in total proteins, albumin, globulins, A/G ratio, uric acid, triglycerides, cholesterol, glucose, calcium, phosphorus, Ca/P ratio, ALT, CK, GGT and PAL between the sexes within the same age group during the egg-laying stage. During the production period, the weight of the males is controlled as well as females. But certainly, females have different metabolic demands due to egg production, which justifies the difference in the serum levels of the evaluated elements.

Conclusion

In healthy male and female broiler breeders during the egg-laying stage, there was a physiological difference in total proteins, albumin, globulins, A/G ratio, uric acid, triglycerides, cholesterol, glucose, calcium, phosphorus, Ca/P ratio, ALT, CK, GGT and PAL between the sexes within the same age group. The serum biochemical parameter ALT did not significantly differ between the sexes within the same age group or among the different ages.

This study presents an important reference for the biochemical parameters of blood proteins, lipids, enzymes, calcium and phosphorus in broiler breeders. These data will be useful for other studies that aim to improve the use of biochemical markers to assess the physiological state of birds.

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s11250-021-02981-z.

Author contribution Rezende and Mundim planned the research. Rezende and Braga collected sample. Mundim performed laboratory analysis. Guimarães performed the statistical analysis and interpreted the data. Rezende, Mundim and Fonseca interpreted the date. Rezende and Fonseca wrote the manuscript. Fonseca, Mundim and Braga carried out the revision. All authors have read and agreed to the published version of the manuscript.

Funding Part of this study was financed by the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior - Brasil (CAPES) - Finance Code 001, FAPEMIG (Fundação de Amparo à Pesquisa do Estado de Minas Gerais) and CNPq (Conselho Nacional de Desenvolvimento Científico e Tecnológico).

Data availability The datasets generated during and/or analysed during the current study are available from the corresponding author on reasonable request.

Code availability Not applicable.
Declarations

Committee on ethics and biosafety All procedures performed in this study were in accordance with Law No. 11.794, of October 8, 2008, certificated by Protocol CEAU/UFU 004/16, approved on March 15, 2016, by the Ethics Committee on the Use of Animals at the Federal University of Uberlândia.

Informed consent Not applicable.

Conflict of interest The authors declare no competing interests.

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