Reviews on Biological Activity, Clinical Trial and Synthesis Progress of Small Molecules for the Treatment of COVID-19

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Abstract
COVID-19 has broken out rapidly in nearly all countries worldwide, and has blossomed into a pandemic. Since the beginning of the spread of COVID-19, many scientists have been cooperating to study a vast array of old drugs and new clinical trial drugs to discover potent drugs with anti-COVID-19 activity, including antiviral drugs, antimalarial drugs, immunosuppressants, Chinese medicines, Mpro inhibitors, JAK inhibitors, etc. The most commonly used drugs are antiviral compounds, antimalarial drugs and JAK inhibitors. In this review, we summarize mainly the antimalarial drugs chloroquine and hydroxychloroquine, the antiviral drugs Favipiravir and Remdesivir, and JAK inhibitor Ruxolitinib, discussing their biological activities, clinical trials and synthesis progress.

Keywords COVID-19 · Remdesivir · Favipiravir · Chloroquine · Hydroxychloroquine · Ruxolitinib

Abbreviations
ACE2 Angiotensin-converting enzyme 2
ARDS Acute respiratory distress syndrome
BCL-2 β-Cell lymphoma 2
CLQ-OH Hydroxychloroquine
CNS Central nervous system

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COVID-19 Corona virus disease 2019
CRP Levels of C-reactive protein
CXCR4 Cxc chemokin receptor 4
CXCL-12 Cxc chemokin ligand 12
DAAs Direct-acting antivirals
EC\textsubscript{50} Concentration for 50\% of maximal effect
EC\textsubscript{90} Concentration for 90\% of maximal effect
EFDA European Food and Drug Administration
FDA Food and Drug Administration
G-CSF Granulocyte colony-stimulating factor
hrs ACE2 Human recombinant soluble angiotensin-converting enzyme 2
IC\textsubscript{50} The half maximal inhibitory concentration
ICU Intensive care unit
IL Interleukins
INF Interferon
JAK Janus kinase
MCP-1 Monocyte chemoattractant protein-1
MERS-CoV Middle East respiratory syndrome coronavirus
MIP-1α Recombinant human chemokine CCL3
PBPK Physiologically-based pharmacokinetic models
RBD Receptor binding domain
RdRp RNA dependent RNA polymerase
SARS-CoV Severe acute respiratory syndrome coronavirus
STAT Signal transducer and activator of transcription
SHLH Secondary hemophagocytic lymphohistiocytosis
T-ALL T cell acute lymphoblastic leukemia
TNF Tumor necrosis factor

1 Introduction

Coronavirus disease 2019 (COVID-19) has broken out rapidly in nearly all countries worldwide, and has blossomed into a pandemic. COVID-19 infections normally manifest with symptoms of high temperature, cough, myalgia, weakness, polypnea and other symptoms [1]. In grievous cases, it can also cause acute respiratory distress syndrome (ARDS), and result in fluids around in the lungs, eliciting infectious shock. Since the beginning of the COVID-19 spread, scientists have investigated cooperatively, examining abundant old drugs and new clinical treatments, such as Chinese medicines [2], vaccine development [3, 4], convalescent plasma [5–7], interferon-based therapies [8], monoclonal antibodies [9], cell-based therapies [10], immunopathology therapies [11] and small molecule drugs, aiming to discover drugs with potent anti-COVID-19 activity. However, the pathway to develop a new drug or vaccine usually takes more than 1 year or even 3–5 years. Monoclonal antibodies, cell-based therapies, interferon-based therapies, and immunopathology therapies are unacceptable for their high cost. Considering cost and time constraints, small molecule drugs, including existing drugs, e.g., those used to treat influenza,
HBV, HCV, HIV, antimalarial and anti-filovirus drugs, have evoked great interest among researchers as they might allow more rapid development [12].

As traditional small molecule drugs used for treating malaria and certain autoimmune diseases, chloroquine (CLQ) and hydroxychloroquine (CLQ-OH) (Fig. 1) were demonstrated recently to exhibit certain activity against COVID-19 both in vitro and in vivo. Preliminary clinical results showed that CLQ has potential for use in the treatment of COVID-19 patients [13]. Compared with CLQ, CLQ-OH is more effective and has better safety properties in vitro [14]. Recently, the Gautret group [15] conducted a clinical trial using CLQ-OH in combination with Azithromycin to treat patients with COVID-19; the results showed that this was effective. But, afterwards, it was reported that use of these two antimalarial drugs may be fatal in some cases [16].

Shah et al. [17] reviewed a total of 61 antiviral drugs to screen efficient drugs against COVID-19 as shown in Tables 1, 2, 3, 4, 5, 6 and Figs. 2, 3, 4, 5, 6, 7. Some biological activities against COVID-19 are also listed based on literature reports [18–37].

Yan and Muller [37] provided a detailed analysis and described the use of the parent nucleoside of remdesivir, GS-441524 (Fig. 8), over remdesivir for the treatment of COVID-19, and appealed to Gilead Science to ditch GS-441524.

Riva et al. [38] identified 100 known drugs that can inhibit COVID-19 replication in mammalian cells, and found 21 compounds exhibiting effective dose response relationships with antiviral activity and confirmed their dose/activity relationships. Then, they found 13 compounds harboring EC50 values < 500 nM in at least one cell line (Table 7); for structures of these compounds see Fig. 9.

In addition to antimalarial and antiviral drugs, Mpro has also attracted the interest of researchers as a drug target for COVID-19 as it can mediate virus duplication and transcription. Jin and co-workers [39] reported the mechanism of Mpro inhibitor N3 via computer-aided drug design, and confirmed the crystal structure of COVID-19 Mpro and N3. Their results showed that N3 had the strongest antiviral COVID-19 effects at a concentration of 10 μM, the inhibition against COVID-19 EC50 value was 16.77 μM. They also discovered the crystal structure of the Mpro-Carmofur complex, confirming that Carmofur covalently links to the Cys145 residue via the carbonyl group, while the fatty acid group married with the hydrophobic S2 subsite of Mpro [40]. Su and co-workers [41] investigated the inhibition of COVID-19 Mpro by natural products derived from Chinese traditional medicines. They found that baicalin and baicalein showed non-covalent,

![Fig. 1 Structure of chloroquine (CLQ) and hydroxychloroquine (CLQ-OH)](image-url)
non-peptidomimetic inhibition of COVID-19 M\textsuperscript{pro}, and had strong antiviral activities both in vitro and in a cell-based system. The in vitro study results and favorable safety data from clinical trials showed that baicalein has great potential to become a candidate for a much needed anti-coronaviral drug. Dai and co-workers [42] designed two M\textsuperscript{pro} inhibitors 11a and 11b, which showed perfect anti-COVID-19 activity. The structure–function relationship showed that the aldehyde group of the two compounds can covalently link to the M\textsuperscript{pro} Cys145

| Entry | Name | Biological activity against COVID-19 | Mechanism |
|-------|------|-------------------------------------|-----------|
| 1     | ABT450 | ND | Inhibits NS3/4A serine protease |
| 2     | Asunaprevir | IC\textsubscript{50}: 53.9 ± 1 μM [18] | Protease inhibitor |
| 3     | Beclabuvir | ND | Inhibits NS5B protein |
| 4     | Boceprevir | IC\textsubscript{50}: 2.7 ± 0.05 μM [19] | NS3/4A protease inhibitor |
| 5     | Dasabuvir | ND | Inhibits the action of NS5B polymerase |
| 6     | Danoprevir | CC\textsubscript{50}: > 50 μM [20] | NS3/4A protease inhibitor |
| 7     | Daclatasvir | EC\textsubscript{50}: 0.8 μM [20] | NS5A inhibitor |
| 8     | Faldaprevir | ND | Protease inhibitor |
| 9     | Elbasvir | ND | NS5A inhibitor |
| 10    | Grazoprevir | K\textsubscript{i}: 172.33 nM [21] | Blocks NS3 |
| 11    | Mericitabine | ND | Inhibitor of RdRp |
| 12    | Radalbuvir | ND | NS5B inhibitor |
| 13    | Simeprevir | EC\textsubscript{50}: 4.08 μM, CC\textsubscript{50}: 19.33 μM [22] | HCV protease inhibitor |
| 14    | Ombitasvir | ND | NS5A inhibitor |
| 15    | Sofosbuvir | EC\textsubscript{50}: 381 ± 34 μM [20] | Inhibits viral RNA synthesis by inhibiting NS5B protein |
| 16    | Ravidasvir | ND | NS5A inhibitor |
| 17    | Telaprevir | IC\textsubscript{50}: 10.7 ± 0.4 μM [18] | NS3/4a protease inhibitor |
| 18    | Velpatsvir | EC\textsubscript{50}: 0.77–2.74 μM [23] | NS5A protein inhibitor |
| 19    | Vedroprevir | ND | Inhibits HCV NS3 |
| 20    | Vaniprevir | ND | NS3/4A protease inhibitor |
| 21    | Uprifosbuvir | EC\textsubscript{50}: > 50 μM [24] | NS5B polymerase inhibitor |

**Table 1** Hepatitis C virus (HCV) antiviral agents against COVID-19

**Table 2** Hepatitis B virus (HBV) antiviral agents against COVID-19

| Entry | Name | Biological activity against COVID-19 | Mechanism |
|-------|------|-------------------------------------|-----------|
| 1     | Famciclovir | ND | Inhibits viral DNA polymerase |
| 2     | Entecavir | EC\textsubscript{50}: > 10 μM, CC\textsubscript{50}: > 50 μM [23] | Inhibits reverse transcription |
| 3     | Telbivudine | ND | Reverse transcriptase inhibitor |
| 4     | Foscarnet | ND | Viral DNA polymerases inhibitor |
| Entry | Name            | Biological activity against COVID-19 | Mechanism                                      |
|-------|-----------------|--------------------------------------|------------------------------------------------|
| 1     | Amprenavir      | EC_{50}: 31.32 μM, CC_{90}: > 81 μM [25] | Protease inhibitor                              |
| 2     | Adefovir        | ND                                   | Reverse transcriptase inhibitor                 |
| 3     | Azidothymidine  | ND                                   | Inhibits reverse transcriptase                  |
| 4     | Darunavir       | K_{d}: 57.30 nM (3CL protease), 6.09 nM (RdRp) and 46.16 nM (papain-like protease) [26] EC_{50} > 100 μM [27] | Inhibits HIV protease enzyme                    |
| 5     | Delavirdine     | ND                                   | Non-nucleoside reverse transcriptase inhibitor  |
| 6     | Didanosine      | ND                                   | Nucleoside reverse transcriptase inhibitor      |
| 7     | Elvitegravir    | ND                                   | Integrase inhibitor                             |
| 8     | Efavirenz       | EC_{50}: > 9.6 μM, CC_{50}: 37.6 ± 10.7 μM [23] | Inhibits non-nucleoside reverse transcriptase enzyme |
| 9     | Ritonavir       | C_{50}: 8.63 μM, CC_{50}: 74.11 μM [25] | HIV Protease inhibitor                          |
| 10    | Indinavir       | EC_{50}: 59.14 μM CC_{50}: > 81 μM [25] | Protease inhibitor                              |
| 11    | Maraviroc       | EC_{50}: 2.7 μM [28]                | C–C chemokine receptor type 5 allosteric modulator |
| 12    | Lopinavir       | EC_{50}: 5.73 μM, CC_{50}: 74.44 μM [25] | Protease inhibitor                              |
| 13    | Raltegravir     | ND                                   | HIV-1 integrase inhibitor                       |
| 14    | Nevirapine      | ND                                   | Non-nucleoside reverse transcriptase inhibitor  |
| 15    | Sequinavir      | EC_{50}: 8.83 μM, CC_{50}: 44.43 μM [25] | Protease inhibitor                              |
| 16    | Stavudine       | ND                                   | Inhibits HIV reverse transcriptase enzyme       |
| 17    | Zalcitabine     | ND                                   | Inhibits nucleoside reverse transcriptase       |
| 18    | Tenofovir       | EC_{50}: 100 μM [29]                | HIV-1 reverse transcriptase inhibitor           |
residue. Zhang and co-workers [43] reported the complex structure of COVID-19 M\(^{\text{pro}}\) and 11r, found that 11r showed excellent inhibitory activity and potent anti-COVID-19 activity. 11r could be used as a lead compound to develop potent inhibitors of COVID-19 M\(^{\text{pro}}\) (Fig. 10).

Vitner et al. [44] examined a Glucosyl Ceramide synthase (GCS) inhibitor GENZ-123346 (analogue of Cerdelga) and GENZ-66761 (structure unknown) for their antiviral effects on COVID-19. Both drugs can inhibit COVID-19 virus, and could be assessed further in preclinical and clinical trials (Fig. 11).

Cytokine storm is a driver of pathology and mortality in viral infections. In COVID-19-infected patients, cytokine storm increases the risk of death and other severe symptoms [45]. Plenty of COVID-19 patients with cytokine storm syndrome encounter a sharp respiratory function obstacle [46]. Secondary hemophagocytic lymphohistiocytosis (sHLH) is a hyper inflammatory syndrome characterized by noteworthy augmentation of cytokines, with multi-organ failure and high mortality rate [47]. COVID-19 patients with cytokine storm syndrome exhibit increased levels of several interleukins: IL-2, IL-6, IL-7, granulocyte colony-stimulating factor (G-CSF), interferon-gamma (IFN-γ) and tumor necrosis factor alpha (TNF-α). The study showed that COVID-19 patients who died have higher plasma levels of ferritin and IL-6 [48]. Four United States Food and Drug Administration (FDA)-approved Janus kinase (JAK) inhibitors have proved useful for the treatment of COVID-19 [49]: Jakafi/Ruxolitinib; Xeljanz/Tofacitinib; Olumiant/Baricitinib; and Rinvoq/Upadacitinib (Fig. 12). Among these four JAK inhibitors, Ruxolitinib can significantly reduce the IL-6 and TNF-α level in spleen [50].

Table 4  Influenza antiviral agents against COVID-19

| Entry | Name                 | Active against | Biological activity against COVID-19 | Mechanism               |
|-------|----------------------|----------------|-------------------------------------|-------------------------|
| 1     | Arbidol (Umifenovir) | Influenza      | EC\(_{50}\): 4.11 \(\mu\)M [29]     | Inhibits membrane fusion |
| 2     | Favipiravir          | Influenza      | EC\(_{50}\): 22.5 \(\mu\)M [30]     | Inhibits viral RNA dependent RdRp |
| 3     | Amantadine           | Influenza A    | IC\(_{50}\) > 100 \(\mu\)M [31]     | The influenza virus A-M2 proton channel agonist |
| 4     | Zanamivir            | Influenza viruses | ND                                 | Neuraminidase inhibitor |
| 5     | Oseltamivir          | Influenza viruses A | EC\(_{50}\): > 100 \(\mu\)M [32] | Inhibits the neuraminidase enzyme |

Table 5  Ebola antiviral agents against COVID-19

| Entry | Name            | Active against | Biological activity against COVID-19 | Mechanism         |
|-------|-----------------|----------------|-------------------------------------|-------------------|
| 1     | Galidesivir     | Ebola          | EC\(_{50}\): > 100 \(\mu\)M [32]     | RNA polymerase inhibitor |
| 2     | Remdesivir      | Ebola virus, Respiratory syncytial virus | EC\(_{50}\): 0.77 \(\mu\)M [33] | Viral RNA polymerase |
| Entry | Name                  | Active against                  | Biological activity against COVID-19 | mechanism                                                   |
|-------|-----------------------|---------------------------------|--------------------------------------|-------------------------------------------------------------|
| 1     | Acyclovir             | Cytomegalovirus infections      | ND                                   | Inhibits viral DNA polymerase                               |
| 2     | Baricitinib           | Rheumatoid arthritis            | IC<sub>50</sub>: 400–800 nM [34]    | Inhibits Janus kinase                                       |
| 3     | Brivudin              | Herpes zoster                   | ND                                   | Locks the action of DNA polymerases                        |
| 4     | Camostate             | Pancreatitis                    | EC<sub>50</sub>: 107 nM [35]         | Serine protease inhibitor                                  |
| 5     | CGP42112A             | Vasodilation and blood pressure reduction | ND                                   | Angiotensin AT2 receptor agonist                           |
| 6     | Dihydroxy propyladenine | Herpes Virus                 | ND                                   | Inhibits viral replication                                 |
| 7     | Ganciclovir           | Cytomegalovirus                 | ND                                   | Inhibits viral DNA polymerases                             |
| 8     | Idoxuridine           | Herpes simplex virus            | ND                                   | Interferes viral DNA replication                           |
| 9     | Marboram/Methisazone | Small pox virus                 | ND                                   | Inhibits mRNA and protein synthesis                        |
| 10    | Nitrazoxanide         | Broad-spectrum antiviral        | EC<sub>50</sub>: 2.21 μM [36]        | Pyruvate:ferred oxinoxidoreductase (PFOR) enzyme            |
| 11    | NSC306711 (Ferristatin II) | Flavivirus               | ND                                   | Degradation of Transferrin receptor-1                      |
Of all the above-mentioned drugs that have potential activity in inhibiting COVID-19, CLQ, CLQ-OH, Favipiravir, Remdesivir and Ruxolitinib attracted our interest due to extensive research on these drugs worldwide.

Fig. 2 Structure of hepatitis C virus (HCV) antiviral agents against COVID-19
Fig. 2 (continued)

Fig. 3  Structure of hepatitis B virus (HBV) antiviral agents against COVID-19
Fig. 4 Structure of human immunodeficiency virus (HIV) antiviral agents against COVID-19
Characteristics of COVID-19

Coronavirus is a positive sense, single-chain RNA virus [51]. Coronaviruses are classified as α-, β-, γ-, and δ-coronavirus. Only α-coronavirus and β-coronavirus can infect humans [52]. γ-Coronavirus and δ-coronavirus can affect humans indirectly through animals [53]. The coronavirus (severe acute respiratory syndrome coronavirus (SARS-CoV)-2) causing COVID-19 is a β-coronavirus and shares about 80% RNA sequence consistency with SARS-CoV [54]. The SARS-CoV-2 genome encodes four main non-structural proteins: helicase, Mpro, RNA-dependent papain-like protease, and RNA polymerase [55], which are absolutely necessary for the survival of SARS-CoV-2. Initial analyses found that the four main SARS-CoV-2 enzymes mentioned above are highly conserved [56], and the spike glycoprotein is indispensable for SARS-CoV-2 invading the host cell [57]. Similar to SARS-CoV, SARS-CoV-2 encodes a large spike protein with two domains (S1 and S2); SARS-CoV-2 virus binds and enters a host cell via this spike protein [58, 59] (Fig. 13).

New research has shown that SARS-CoV-2 engages a receptor binding domain (RBD), binding to angiotensin-converting enzyme 2 (ACE2) to invade its human host cell [60] (Fig. 14).

Nevertheless, non-conserved mutations are highly accumulated in regions S1 and S2 which interact directly with ACE2. Mercurio et al. [61] performed a comparative in silico modeling analysis, and gained new insights into the spike protein of SARS-CoV-2. SARS-CoV-2 spike protein can interact with the ACE2
receptor on human cells at the RBD. This analysis can supply an ideal pipeline to identify characterized antibodies that might target the SARS-CoV-2 spike protein RBD to prevent interacting with human ACE2. Laurini et al. [62] reported an atomistic-based, reliable in silico structure of the viral transmembrane spike

Fig. 7 Structure of other antiviral agents against COVID-19

Fig. 8 Structure of GS-441524 (remdesivir)
### Table 7  Drugs with known activity against COVID-19 in cell lines

| Entry | Drug name | EC50 value (µM) | Cell line |
|-------|-----------|-----------------|-----------|
| 1     | AMG2674a  | 0.023           | Vero E6 cells |
| 2     | AMG2674   | 0.3             | 293T-ACE2 cells |
| 3     | AMG2674   | 0.41            | Hub-7-ACE2 cells |
| 4     | Astemizole| ~ 1.2           | Vero E6 cells |
| 5     | Astemizole| 0.87            | 293T-ACE2 cells |
| 6     | Astemizole| 1.3             | Hub-7-ACE2 cells |
| 7     | Clofazimine| 0.31             | Vero E6 cells |
| 8     | Clofazimine| ND              | 293T-ACE2 cells |
| 9     | Clofazimine| 0.49             | Hub-7-ACE2 cells |
| 10    | Elopiprazole| 1.6            | Vero E6 cells |
| 11    | Elopiprazole| 0.13             | 293 T-ACE2 cells |
| 12    | Elopiprazole| ~ 2.7            | Hub-7-ACE2 cells |
| 13    | Hanfanychin A| ~ 1.2            | Vero E6 cells |
| 14    | Hanfanychin A| 0.56            | 293T-ACE2 cells |
| 15    | Hanfanychin A| 0.64            | Hub-7-ACE2 cells |
| 16    | MLN-3897  | 0.65            | Vero E6 cells |
| 17    | MLN-3897  | 0.41            | 293T-ACE2 cells |
| 18    | KW 8232   | ~ 1.2           | Vero E6 cells |
| 19    | KW 8232a  | ~ 0.091         | 293T-ACE2 cells |
| 20    | KW 8232   | 1               | Hub-7-ACE2 cells |
| 21    | N-tert-Butylisoquine| ~ 1.2 | Vero E6 cells |
| 22    | N-tert-Butylisoquine| 0.29 | 293T-ACE2 cells |
| 23    | N-tert-Butylisoquine| 0.37 | Hub-7-ACE2 cells |
| 24    | SB 616234-A| ~ 1.2            | Vero E6 cells |
| 25    | SB 616,234-A| ~ 0.29           | 293 T-ACE2 cells |
| 26    | SB 616234-A| 0.64            | Hub-7-ACE2 cells |
| 27    | SDZ-62-434| 0.63            | Vero E6 cells |
| 28    | SDZ-62-434| 0.12            | 293T-ACE2 cells |
| 29    | SDZ-62-434| 0.11            | Hub-7-ACE2 cells |
| 30    | SL-11128  | ~ 0.25          | Vero E6 cells |
| 31    | SL-11128  | ND              | 293T-ACE2 cells |
| 32    | SL-11128  | ~ 2.5           | Hub-7-ACE2 cells |
| 33    | YH-1238   | ~ 0.95          | Vero E6 cells |
| 34    | YH-1238   | 1.1             | 293T-ACE2 cells |
| 35    | Apilimoda  | 0.0203          | Vero E6 cells |
| 36    | Apilimoda  | 0.012           | 293T-ACE2 cells |
| 37    | Apilimoda  | 0.088           | Hub-7-ACE2 cells |
| 38    | VBY-825   | 0.3             | Vero E6 cells |
| 39    | VBY-825a  | 0.071           | 293T-ACE2 cells |
| 40    | ONO 5334  | 0.41            | 293T-ACE2 cells |
| 41    | ONO 5334a | 0.042           | Vero E6 cells |
| 42    | ONO 5334a | 0.078           | 293T-ACE2 cells |
glycoprotein (S-protein)/ACE2 complex (Fig. 15), showing that residues D38, K31, E37, K353, Y41 on ACE2 and Q498, T500, R403 on the SARS-CoV-2 S-protein RBD are true hot spots to shaping and determining the stability of the relevant protein–protein interface. These results could be used in the structure-based design and development of neutralizing antibodies, vaccines, and protein–protein inhibitors against COVID-19.

Monteil et al. [63] afforded a molecular explanation for the severe lung failure and death due to COVID-19, and proved that APN01 (human recombinant soluble ACE2, also named hrsACE2) can inhibit SARS-CoV-2 infections, which can be used for treatment of COVID-19 patients.

3 Chloroquine

CLQ is an old antimalarial drug used to treat malaria, amebiasis, rheumatoid, arthritis and lupus erythematosus syndrome. It inhibits the heme polymerase in malarial trophozoites via preventing heme conversion to hemozoin [64]. CLQ has strong antiviral effects on SARS-Cov infection [65] and Ebola in vitro [66], and is also able to inhibit influenza A virus replication in vitro [67]. Moreover, CLQ also has been shown to have some level of anti-HIV [68] and anti-H5N1 [69] activity.

3.1 Biological Activity of CLQ

CLQ can increase the pH value of host cell vacuoles. In lysosomes of the host cell, CLQ can change the catalytic activity of acidic hydrolases, affecting protein degradation, endosomal macromolecule composition, and post-translational modifications in Golgi [70]. In macrophages and antigen-presenting cells, an antirheumatic response is present that interrupts the immunological process [71]. Recent literature reports [72–74] assist our understanding of the three probable mechanisms of CLQ activity (Fig. 16).
Gao et al. [13] found an EC$_{50}$ value of 1.13 μM and a CC$_{50}$ value greater than 100 mM for CLQ anti-COVID-19. Wang et al. [33] evaluated the efficiency of CLQ and six other drugs against SARS-CoV-2 in vitro, obtaining an EC$_{50}$ value for CLQ of 1.13 μM and EC$_{90}$ value of 6.90 μM in Vero E6 cells.

*Fig. 9* Compounds exhibiting good effectiveness in COVID-19 cell lines
Fig. 9 (continued)

Carmofur    EC$_{50}$ = 24.30µM;  
11a    IC$_{50}$ = 0.053±0.005µM;  
11b    IC$_{50}$ = 0.040±0.002µM;  
11r    IC$_{50}$ = 2.39±0.63µM; 
N$_3$    EC$_{50}$ = 16.77µM

Fig. 10  Structure and biological activity of Carmofur, Baicalein, 11a, 11b, 11r and N$_3$
Fig. 11  Structure of GENZ123346

Fig. 12  Structure of four Janus kinase (JAK) inhibitors

Fig. 13  Schematic representation of a coronavirus virion. Image reproduced from Ref. [59] with permission from Elsevier

Fig. 14  Comparison of severe acute respiratory syndrome coronavirus (SARS-CoV)-2 receptor binding domain (RBD)/human angiotensin-converting enzyme 2 (hACE2) complex structures (a) and the constructed SARS-CoV-2 RBD/hACE2 peptide complex (b). Image reproduced from Ref. [60] with permission from bioRxiv
3.2 Clinical Trials of CLQ

Since the outbreak of COVID-19, CLQ has been used extensively to treat COVID-19 patients. Sun et al. [75] reported using CLQ phosphate at a dose of 500 mg to treat COVID-19 with a duration not exceeding 10 days in elderly patients; these authors summarized the main adverse reactions of CLQ in elderly patients: cardiac
arrest, effects on skeletal musculoskeletal system, nerve irritability, medicated psychosis, granulocytopenia, aplastic anemia, thrombocytopenia, irreversible visual impairment, and others. Verscheijden et al. [76] established the best-evidence pediatric CLQ (free base) doses: 35 mg/kg, children 0–1 month, 47 mg/kg; children 1–6 months, 55 mg/kg; children 6 months to 12 years; and adolescents 44 mg/kg. Gao et al. [13] revealed that CLQ used in an intervention group had potent efficacy compared with a control group. Borba et al. [77] performed a parallel phase IIb clinical trial study to evaluate the safety and efficacy of high (0.6 g) and low (0.45 g) dose CLQ. Double-blinded and randomized participants were treated in intervention and control groups; the results suggested that the high dose of CLQ is hazardous to COVID-19 patients. Huang et al. [78] undertook a multicenter prospective observational study. Patients received a CLQ dose of 500 mg once or twice daily; patients treated with non-CLQ therapy were used as historical controls. In total, 197 patients used CLQ treatment, and 176 patients were treated as the historical control group. The duration of fever was clearly shortened. No serious adverse events were found in patients treated with CLQ. Patients treated with half dose CLQ experienced a lower rate of adverse events than with full dose. This study provides evidence for the safety and efficacy of CLQ in treatment of COVID-19 patients. Smit et al. [79] introduced pharmacokinetic and safety properties of CLQ for treatment of COVID-19, revealing that, for the use of CLQ to treatment of COVID-19 infection, early achievement of high ‘target’ concentrations is necessary, but, due to the high loading dose of CLQ and slow elimination, it can cause severe life-threatening toxicity and increased risk of mortality. Sinkeler [80] conducted a retrospective and observational study in a total of 397 patients aged over 18 years, found the CLQ gradually increased the QTc interval during treatment, which may result in ventricular tachycardia in patients. Sinkeler [80] suggested to measure the QTc interval before adjusting the dose of CLQ or withdrawing this potentially beneficial medication. While the pathogenesis of COVID-19 is still unknown, and because it does not show any anti SARS-Cov effect in an in vivo model, CLQ could be useless in treatment of COVID-19 patients and might even be harmful [81]; hence, before the pathogenesis of COVID-19 is known and the effect of CLQ is evaluated in clinical trials, it should be used cautiously.

3.3 Synthesis of CLQ

Surrey and Hammer [82] reported the first synthesis of CLQ (Scheme 1).

Ethyl ethoxalylacetate reacting with m-chloroaniline (I) in the presence of HOAc under mild condition obtained II, then ring-closure at high temperature in a short time yields III and its isomer IIIa; III and IIIa were refluxed in strong base solution to obtain a mixture of IV and IVa, with a final coupling of IV and V at high temperature obtaining CLQ. This synthetic route was unacceptable due to steps (ii) and (iii) producing nearly half of an isomer byproduct and steps (ii) and (iv) proceeding at high temperature.
Jonnson and Buell [83] reported a second generation synthesis of CLQ (Scheme 2).

Methyl acrylate and \( m \)-chloroaniline (I) were refluxed in HOAc to obtain II, then II was protected by TsCl and ring-closure obtained IV. Finally, IV and V were

\[
\begin{align*}
\text{Cl} & \quad \text{NH}_2 \\
\text{O} & \quad \text{O} \\
\text{HOAc} & \quad \text{a} \\
\text{Cl} & \quad \text{NH}_2 \\
\text{Cl} & \quad \text{OH} \\
\text{Cl} & \quad \text{N} \\
\text{H} & \quad \text{N} \\
\text{Cl} & \quad \text{N} \\
\text{N} & \quad \text{Cl} \\
\text{N} & \quad \text{Cl} \\
\text{Cl} & \quad \text{Cl} \\
\text{IVa} & \quad \text{IV} \\
\end{align*}
\]

Reaction conditions: a) ethyl ethoxalylacetate, \( m \)-chloroaniline, HOAc, 40-50 °C, 4 h, then r.t. 15-18 h, yield 78.1%; b) 250 °C, 15-18 min, yield (III) and (IIIa) 86.8%; c-1) 35% NaOH, 2 h, then con HCl; c-2) mineral oil 270 °C 5 min, yield 96.2%

**Scheme 1** First generation synthesis of Chloroquine (CLQ)

\[
\begin{align*}
\text{Cl} & \quad \text{NH}_2 \\
\text{O} & \quad \text{O} \\
\text{HOAc} & \quad \text{a} \\
\text{Cl} & \quad \text{NH}_2 \\
\text{Cl} & \quad \text{OH} \\
\text{Cl} & \quad \text{N} \\
\text{H} & \quad \text{N} \\
\text{Cl} & \quad \text{N} \\
\text{N} & \quad \text{Cl} \\
\text{N} & \quad \text{Cl} \\
\text{Cl} & \quad \text{Cl} \\
\text{IVa} & \quad \text{IV} \\
\end{align*}
\]

Reaction conditions: a) HOAc, reflux, 12 h; b) (1)tosyl chloride, Et3N; (2) 35%NaOH, reflux, 3 h, then HCl, a and b total yield 80%; c) PCl5, AlCl3, reflux; d. phenol, 154 °C, 6 h, yield 68%

**Scheme 2** Compound II should be added an C-Cl at meta position

Jonnson and Buell [83] reported a second generation synthesis of CLQ (Scheme 2).
refluxed in the presence of phenol to obtain CLQ. The total yield of this procedure is about 24.7%, which is higher than the first generation process, but, in step d, the reaction temperature is as high as more than 150 °C, at which temperature byproducts are easily produced, and the solvent phenol is harder to recover.

Margolis et al. [84] reported a third generation synthesis of CLQ (Scheme 3). I and II were refluxed without solvent and ring-closure obtained IV, then chlorination in the presence of POCl₃ obtained V. Finally, V and VI coupling via Suzuki coupling obtained CLQ. The totally yield of this procedure is approximately 58% in mild conditions, which is suitable for large-scale production.

4 Hydroxychloroquine

4.1 Biological Activity of CLQ-OH

CLQ-OH is similar to CLQ, and as an old antimalarial drug, also has potential effects on COVID-19 and lower toxicity than CLQ [85]. Moreover, it showed better anti-SARS-CoV-2 efficiency compared to CLQ in vitro [86]. Yao et al. [14] used physiologically-based pharmacokinetic models (PBPK) models to ensure the most efficacious concentrations of CLQ-OH and its safety profile, and found its EC₅₀ value to be 0.72 μM in vitro. Fantini et al. [74] used an assembly of structure-molecular modeling methods and found that CLQ-OH can prevent binding of the SARS-CoV-2 spike to gangliosides, which is helpful in understanding the mechanism of CLQ-OH as an anti-COVID-19 drug (Figs. 17, 18).

Zhang et al. [87] systematically evaluated the treatment of COVID-19 with CLQ and CLQ-OH for efficacy and safety, and asserted a potential mechanism involving COVID-19-induced injury of multiple organs as well as the pharmacological effects of CLQ-OH on COVID-19 (Fig. 19); the authors concluded that COVID-19 is actually a multisystem disease, with respiratory symptoms as the major clinical manifestation. Therefore, the treatment of COVID-19 with CLQ and CLQ-OH should be evaluated systematically, and patients should be monitored carefully for cardiovascular conditions to prevent lethal adverse events.

![Scheme 3 Third generation synthesis of CLQ](image)

Reaction conditions: a) triethyl orthoformate, reflux 4 h, yield 82%; b) Ph₂O reflux; c) POCl₃, reflux, yield 95%; d) 4 mol% Pd(OAc)₂, 8 mol% DPEphos, 2.5 eq K₃PO₄, N₂, Dioxane, 85 °C, 18 h, yield 74%.
**Fig. 17** Molecular model of CLQ-OH interaction with sialic acids. Image reproduced from Ref. [74] with permission from Elsevier

**Fig. 18** Molecular modeling simulations of CLQ and CLQ-OH binding to ganglioside. Image reproduced from Ref. [74] with permission from Elsevier

**Fig. 19** Potential mechanisms of SARS-COV-2-induced injury of multiple organs and pharmacological effects of CLQ-OH on COVID-19. *Blue arrows* indicate the actions of SARS-COV-2. ACE2 is key for SARS-COV-2 entering cells in human organs. *Red dashed lines* indicate the potential mechanisms of the therapeutic and toxic effects of CLQ-OH on SARS-COV-2 and organs in COVID-19 patients. Image reproduced from Ref. [87] with permission from Elsevier
4.2 Clinical Trials of CLQ-OH

Based on clinical studies [88], CLQ-OH is considered to be safer than CLQ. Gautret et al. [89] conducted a phase III clinical trial treating 80 mildly infected inpatients with CLQ-OH in combination with Azithromycin, demonstrating its therapeutic effect. Furthermore, the cost of this treatment is negligible. The suggested dose of CLQ-OH is 400 mg daily on the 1st day, then 200 mg daily for the next 3 days [90]. A total of 62 COVID-19 patients were assigned stochastically to the CLQ-OH (0.4 g daily, 0.2 g bid) and the control groups. After 5 days of treatment, patients in the CLQ-OH groups were clearly recovering in a shorter time [91]. To evaluate toxicity in patients who received high doses of CLQ-OH, a total of 63 patients were included [92], 58 females and 5 males. The mean dosage of CLQ-OH was 3.9 mg/kg, but 14 patient had doses higher than 5 mg/kg. A total of 36 patients were treated with CLQ-OH for more than 5 days. Only one (1.58%) patient exhibited CLQ-OH toxic retinopathy over a mean of 8 years treatment period. Patients on doses of > 5 mg/kg of CLQ-OH may be put at higher risk for retinal toxicity. Gautret et al. [15] used CLQ-OH, alone (0.6 g daily) or in combination with Azithromycin to treat COVID-19 patients in a small clinical trial. The results showed that Azithromycin combined with CLQ-OH was more efficient than CLQ-OH alone. A recent report [93] also confirmed significant efficacy of CLQ-OH in combination with Azithromycin. On the contrary, Sharma et al. [94] found opposite consequences in CLQ-OH combination with Azithromycin in a small, non-randomized clinical trial. Moreover, the risk of arrhythmia and prolonged QT interval was increased. Singh et al. [95] studied the efficacy of CLQ-OH in a treatment group compared with a control group in COVID-19, and concluded that there was no positive result in the CLQ-OH group, and that the death rate even increased in the CLQ-OH group compared to the control group. Gendelman et al. [96] performed a retrospective study based on a database including COVID-19 patients treated with CLQ-OH from February to March. A total sample of 14,520 were screened for COVID-19 infection, with 1317 being found positive. These findings raise doubts about the safety and efficacy of CLQ-OH against COVID-19 infection. Lauriola et al. [97] performed a retrospective study including 337 consecutive COVID-19 patients; 297 patients received CLQ-OH and azithromycin combination treatment, 17 patients CLQ-OH treatment alone and 63 patients did not recieve either of these two drugs due to contraindications. In this study, 146 patients died, including 102 in the combination treatment, 7 in the CLQ-OH treatment and 35 in the no treatment groups.

4.3 Synthesis of CLQ-OH

Alexander et al. [98] first reported the synthesis of CLQ-OH (Scheme 4).

I and II were refluxed in xylene to get III, then ammoniation and hydrogenation by Raney Nickel in high pressure obtained IV; finally, IV and V refluxed in phenol
CLQ-OH. The overall yield of this procedure is as low as 17.6%, in step (b) intermediate (III) ammoniation and reduction by Raney Nickel requires high pressure, and in step (c) the solvent phenol is hard to recover. Hence, this method is unacceptable for large-scale production.

Ashok et al. [99] reported another synthetic route for the preparation of CLQ-OH (Scheme 5). Compound I was protected by ethylene glycol then reacted with III in refluxing toluene obtained IV, IV via deprotection, ammoniation and hydrogenation with Raney Nickel in high pressure to get VI, VI reacted with VII obtained CLQ-OH; finally, treatment of CLQ-OH with sulfuric acid yields CLQ-OH sulfate. The overall yield of this procedure is about approximately 40% via

Reaction conditions: a) NaCl, xylene, reflux 2 h, yield 30%; b) ammoniacal methanol, Raney Nickel, 1000 pounds pressure, rt 24 h, yield 89%; c) phenol, KI, 125-130 °C, 18 h, yield 66%

Scheme 4 Method for preparation of Hydroxychloroquine (CLQ-OH) by Alexander et al. [98]

CLQ-OH. The overall yield of this procedure is as low as 17.6%, in step (b) intermediate (III) ammoniation and reduction by Raney Nickel requires high pressure, and in step (c) the solvent phenol is hard to recover. Hence, this method is unacceptable for large-scale production.

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Reaction conditions: a) ethylene glycol, PTSA, 80-90 °C, 20-25 h, yield 100%; b) toluene, 125-130 °C, 15-18 h, yield 83%; c) con HCl, water, 30-40 °C, 4-5 h, yield 100%; d) ammonical methanol, Raney Nickel 40-80 °C, 20 kg hydrogen pressure, 4-5 h, yield 80%; e) NaOH, KI, 110-115 °C, 40-50 h, yield 76%; f. H2SO4, MeOH, 5-100 °C, 3-4 h, yield 77%

Scheme 5 Method for preparation of CLQ-OH by Ashok et al. [99]
six steps, but in step (d), the Raney Nickel reduction of intermediate (IV) requires high pressure, which is a potential safety hazard.

In 2010, Min et al. [100] reported a method for preparation of CLQ-OH (Scheme 6). I and II reacted in nitrogen conditions at high pressure obtained CLQ-OH with a yield of 78%; treatment sulfuric acid obtained CLQ-OH sulfate. This procedure led to CLQ-OH sulfate via two steps with an approximate total yield of 77%, but in step (a) the reaction proceeded in high pressure, which has safety implications.

Yu et al. [101] and Frank et al. [102] used continuous-flow method for high-yield preparation of CLQ-OH (Scheme 7), which is not suitable for large-scale production. Iodization of I with hydroiodic acid and reaction with III in the presence of K₂CO₃ yielded IV, then oxime was obtained with hydroxylamine, and hydrogenation using Raney Nickel in high pressure obtained VI; finally, VI and VII reacted at high temperature in base conditions yielded CLQ-OH. This procedure yielded CLQ-OH in mild conditions, but the final step using ethanol as a solvent at high temperature is unacceptable, as ethanol volatilizes easily.

**Scheme 6** Method for preparation of CLQ-OH by Min et al. [100]

**Scheme 7** Preparation of CLQ-OH with a continuous-flow method
5 Favipiravir

Favipiravir (T-705) has been developed by Toyama Chemical Co., Ltd (Tokyo, Japan) as a broad spectrum antiviral drug against RNA viruses [103]. It shows good efficacy in the treatment of influenza infections [104] and pathogenic avian influenza A (H5N1). Favipiravir was also used as a potential drug for Ebola virus [105] and severe influenza [106], with EC₅₀ values of 0.014–0.55 μg/mL to seasonal influenza and oseltamivir-resistant virus [107]. Favipiravir has an antiviral effect on SARS-CoV-2 by inducing a reduction in virus-induced cytopathic effect [108]. Observations show that Favipiravir induced the mutagenic effect responsible for the inhibition of replication; it was shown to act through lethal mutagenesis for several viruses, predominantly by competing with guanosine to cause transition mutations.

5.1 Biological Activity of Favipiravir

Favipiravir is a virus RdRp inhibitor. EC₉₀ values for H5N1 resistance are in the 1.3–7.7 μM range [109]. Research in a P388D1 cell-based system [110] showed that Favipiravir can clearly inhibit the generation of TNF-α. Shirak et al. [111] established an influenza infection animal model to study the activity of Favipiravir, and proved that Favipiravir was significantly effective in alleviating influenza infection in mice. Janowski et al. [112] demonstrated that Favipiravir has EC₅₀ values of 246 ± 76 μM (VA1), > 1000 μM (HAstV4) and 4.73 μM (IAV).

5.2 Clinical Trials of Favipiravir

Survival and virological characteristics were observed [113]. Randomized, multicenter phase II [114] and phase III [115] trials demonstrated clinical efficacy and safety of Favipiravir in Influenza. Lou [116] assessed the antiviral activity of Favipiravir and Baloxavir against COVID-19. Patients were randomized and assigned into three groups: Baloxavir group: dose 80 mg daily orally on Day 1 and Day 4, for patients still positive in virological test, given again on Day 7; Favipiravir group: The first dose 1.600 g or 2.2 g, followed by dose 0.6 g, 3 times daily; Control group: Continue existing antiviral treatment. The results showed that Favipiravir did not have any dramatic efficiency against COVID-19 even at high concentration. Cai et al. [117] conducted an Open-Label Controlled trial to test the efficacy of Favipiravir compared to Lopinavir (LPV)/Ritonavir (RTV) as anti-COVID-19 agents. Patients were randomly allocated to Favipiravir group (dose 1.6 g twice a day for the 1st day; dose 0.6 g twice a day for 2–14 days) and LPV/RTV group (dose 0.4 g/0.1 g twice a day); the results showed that the Favipiravir group patients exhibited higher efficacy than the LPV/RTV group.

5.3 Synthesis of Favipiravir

Takamatsu and Yonezawa [118] disclosed a method for producing Favipiravir and its intermediates in high yields (Scheme 8). Hydroxylation of I at room
temperature in the presence of KOAc gives II. II reacted with III in the presence of ammonium hydroxide to give IV in 83.2% yield. Finally, cyan oxidation of IV by hydrogen peroxide in strong base solution gives Favipiravir in total yield of 74%. This procedure obtained Favipiravir via three steps under mild conditions. The process has the advantages of easy post-processing, low toxicity, and high yield for high purity of Favipiravir, and is suitable for large-scale production.

Hara et al. [119] reported a method for producing Favipiravir via six steps (Scheme 9), with total yield of 53% under mild conditions. Compound I reacted with glyoxal in the presence of NaOH solution at lower temperature to obtain II, bromination, chlorination and dehydration obtained IV, fluoro-substitution in the presence of KF and tetra-n-butylammonium bromide (TBAB) gives V, hydrolyzation of 2-fluoro group of V in sulfuric acid and 3-cyan group of VI in NaOH solution obtained Favipiravir.

Reaction conditions: a) DMF, H2O, KOAc, 25-35 °C, 2 h; b) NH4OH, pH = 9.4, acetone/toluene, dicyclohexylamine, 20-30 °C, 45min, step a and b totally yield 83.2%; c) toluene, H2O, NaOH, 30% H2O2, 20-30 °C, 1 h, yield 89%.

Scheme 8 Synthesis of Favipiravir by Takamatsu and Yonezawa [118]

Scheme 9 Preparation of Favipiravir by Hara et al. [119]
Liu et al. [120] reported a method for preparation of Favipiravir (Scheme 10) via 8 steps with a total yield of 24%. Esterification of I gave II, bromination by NBS obtained III, diazotization of III and hydrolyzation in the presence of ammonium hydroxide gave IV, ester group amidation obtained V, chlorination and dehydration obtained VI, fluoro-substitution chloro group of V in the presence of KF and TBAB got VII, hydrolyzation 3-cyan group of VII in the presence of hydrogen peroxide and 2-fluro group of VIII in weak base solution obtained Favipiravir. This procedure needs long steps and the diazotization reaction in step (c) has potential safety issues.

Li et al. [121] reported a method for preparation of Favipiravir (Scheme 11) via 4 steps with a total yield of less than 10%. Oxidation of I using hydrogen peroxide in the presence of HOAc gives II. Chlorination and dehydration of II obtained III, and then fluoro-substitution in presence of KF and TBAB gives

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**Scheme 10** Preparation of Favipiravir by Liu et al. [120]

**Scheme 11** Preparation of Favipiravir by Li et al. [121]
IV. Hydrolyzation of 3-cyan group of IV in the presence of hydrogen peroxide gave V, followed by hydrolyzation of 2-fluoro group of V in weak base solution, obtained Favipiravir.

6 Remdesivir

Remdesivir (Fig. 20) also named GS-5734, is a broad-spectrum antiviral RdRp inhibitor against MERS, Ebola, SARS and the like. Research showed an EC₅₀ value of 0.77 μM in Vero E6 cells inhibiting COVID-19 [122]. Holshue [123] reported the first case of a US COVID-19 patient. After the COVID-19 spread worldwide in March, phase III clinical trials were launched in the US and other countries extensively. On 2 May, the FDA issued Remdesivir emergency use for COVID-19 treatment.

6.1 History of Remdesivir

The HCV genome encodes two proteins, NS2 and NS5, that are important targets for drug design [124, 125]. The discovery of small molecules inhibiting virus replication has attracted more attention [25]. Abundant direct-acting antiviral (DAA) small molecules have been designed and tested in clinical trials over the last few decades. A number of DAAs have been proven to have anti-HCV activity. Nucleoside inhibitors (NIs) are the most outstanding due to their prominent therapeutic effect [126]. Most NIs used clinically are N-nucleosides, which have a 2′-C-Me in the sugar branch. The first 2′-C-Me branched N-nucleosides prepared in 1960s were used to treat HCV infection; their in vitro activity against HCV was tested in the 2000s [127]. N-Nucleosides firstly metabolize into nucleoside triphosphates in cells, then link with the NS5B polymerase and insert into the viral RNA, inhibiting viral RNA extension and virus replication [128].
In 2012, Cho et al. [129] synthesized a few 2′-C-Me C-nucleosides targeting NS5B, and screened compound (1) as a HCV inhibitor (Fig. 21).

On the basis of compound (1), abundant analogues of compound (1) were synthesized to improve selectivity (Table 8) [130].

Among these analogues of compound (1), it was found that compound (2) has perfect activity in the NS5B enzyme assay. Then compound (2) should be used as a lead nucleoside for further optimization (Table 9).

In 2013, Cho et al. [131] prepared the first HCV inhibitor C-nucleoside GS-6620 (Fig. 22) with high efficiency in phase I clinical trials.

In 2017, based on GS-6620, Siegel et al. [132] discovered and synthesized Remdesivir, and found it had good activity against Ebola virus.
6.2 Biological Activity of Remdesivir

Remdesivir is RdRp inhibitor, suppressing virus genome replication [51]. RdRp controls the replication of virus RNA. Once Remdesivir metabolizes into the corresponding NTP, the latter competes with ATP for incorporation into the nascent RNA strand [133], leading to termination of virus RNA synthesis and prevention of the growth of RNA. Even though the virus can probe and delete C-nucleosides, resulting in tolerance, Remdesivir appears to be able to overcome this problem and maintain efficiency [134]. Yin et al. [135] reported the complex structure of Remdesivir and SARS-CoV-2 RdRp, where Remdesivir was covalently inserted into RdRp. This complex structure supplies a mechanism whereby Remdesivir inhibits SARS-CoV-2 replication, providing a platform for development of new drugs against COVID-19 (Fig. 23).

Remdesivir has EC$_{50}$ values about 0.07 μM for either SARS-CoV or MERS-CoV [136, 137]. Emmiede et al. [138] tested the efficiency of Remdesivir against MERS-CoV virus. The result showed that Remdesivir can clearly inhibit virus replication. Wang et al. [33] first examined the effect of Remdesivir against COVID-19 in Vero E6 cells and found an EC$_{50}$ value 0.77 μM for Remdesivir inhibiting COVID-19. Elfiky et al. [139] targeted a few anti-polymerase drugs targeting RdRp, and found Remdesivir, Sofosbuvir, Galidesivir and Tenofovir as potent drugs against COVID-19. Choy et al. [140] reported the efficiency of Remdesivir and three other drugs against COVID-19 in Vero E6 cells, with EC$_{50}$ values of 23.15 μM (Remdesivir), 26.63 μM (Lopinavir), 2.55 μM (Homorlingtonine) and 0.46 μM (Emetine), respectively. Zhang et al. [141] found that the 1’-cyano group of Remdesivir has dual roles in inhibition of nucleotide addition and proofreading. Pruijssers et al. [142] reported that Remdesivir potently inhibits SARS-CoV-2 replication in human lung cells and primary human airway epithelial cultures (EC$_{50}$=0.01 μM), in Vero E6 cells (EC$_{50}$ = 1.65 μM), respectively. Wu et al. [143] found that Remdesivir and GS-441524 can inhibit cell proliferation and the expression of fibrotic markers (fibronectin, pSmad3, and aSMA) in NRK-49F and HK2 cells. Brandi et al. [144] used rhesus macaque model of COVID-19, and showed that Remdesivir can be used in transient lower respiratory tract disease.
Fig. 23 The complex structure of Remdesivir and SARS-CoV-2 RNA bound to RdRp complex. Image reproduced from Ref. [135] with permission from Wiley-VCH
6.3 Clinical Trials of Remdesivir

The first case of a COVID-19 patient in the US appeared in January 2020 [90]; his condition improved after 8 days’ Remdesivir treatment, and no obvious adverse effect was observed during the treatment. Stephanie et al. [145] then described 12 US COVID-19 mild-to-moderate patients treated with Remdesivir, and all patients recovered at the end of this clinical trial. A randomized, controlled clinical trial was conducted [146] to investigate the safety and efficiency of Remdesivir against COVID-19 at the University of Nebraska Medical Center (UNMC). The clinical trial results showed no evidence that Remdesivir can improve clinical outcomes. A few phase III clinical trials have been conducted to evaluate the efficiency and safety of Remdesivir against mild and moderate COVID-19 patients [147–150]; the results do not support the efficacy and safety of Remdesivir. Grein et al. [151] reported 61 COVID-19 patients receiving compassionate use Remdesivir: 8 patients were not analyzed for various reasons, 22 patients were from the US, 22 patients from the EU or Canadian, 9 patients were Japanese. The results showed that 68% patients got well, 57% patients received mechanical ventilation, 47% patients were in recovery, and 13% patients died. Even though there were two absolutely contrary results in US and Chinese phase III clinical trials, on 2 May, the FDA authorized Remdesivir for compassionate use against COVID-19 for various reasons. Beige et al. [152] undertook a phase III trial using Remdesivir (dose 0.2 g daily in the first day, dose 0.1 g 2–10 days) in adult patients with COVID-19. A total of 1063 patients underwent randomization. The results, based on findings from analysis, showed that Remdesivir can shorten time to recovery. Goldman et al. [153] proceeded with a phase III clinical trial with Remdesivir (dose 0.2 g daily first day, 0.1 g the remaining days) against COVID-19 in 397 patients. Patients were allocated to two groups (10 days treatment and 5 days treatment). At baseline, prolonging treatment time did not improve clinical status.

6.4 Synthesis of Remdesivir

The first generation [154] for the synthesis of Remdesivir was as follows (Scheme 12). Coupling I and II in the presence of n-BuLi, and chlorotrimethylsilane (TMSCl) or 1,2-bis(chlorodimethylsilyl)ethane, NaH, and n-BuLi at ultralow temperature gave III, cyanation of III by TMSCN in the presence of Lewis acid BF₃-Et₂O at ultralow temperature obtained IV, benzyl deprotection using BCl₃ gave V, V was reacted with VIII in the presence of NMI and OP(OMe)₃ to give VI, with Remdesivir finally achieved by chiral HPLC. In the first generation for synthesizing of Remdesivir, Remdesivir was obtained in total yield of less than 2%. The first four steps were conducted at −78 °C and the β-anomer VII was separated by chromatography [155], which hindered this synthetic route from large-scale development.

Second generation [138] Remdesivir was synthesized diastereoselectively via 7 steps in total 4% yield which is still unacceptable but suitable for large-scale production (Scheme 13). Compound I reacted with II in the presence of TMSCl.
and Grignard reagent at low temperature yielded III, then cyanation of III using TMSCN and TMSOTf in the presence of TfOH at ultralow temperature obtained IV, benzyl deprotection using BCl₃ gave V, using 2,2-dimethoxypropane selectively protected two hydroxy of V to obtain VI, and then VI and X were reacted in the presence of DIPEA followed by deprotection to give Remdesivir.

Vieira et al. [156] described a route to synthesize a key Remdesivir intermediate (IV) by using a continuous flow chemistry method, providing improved control over the reaction conditions and increasing the diastereoselectivity (Scheme 14). Coupling of I and II in the presence of TMSCl and Grignard reagent catalyzed by NdCl₃ at low temperature obtained III in yields of 69%, and cyanation of III using TMSCN in the presence of TMSOTf and TfOH at lower temperatures obtained intermediate IV in yields of 78%. This synthetic route obtained the key intermediate IV in total yield 54% in two steps. But this process requires continuous flow conditions which is unacceptable in industrial production.

Xue et al. [157] disclosed an improved methodology for the key C-glycosylation step for synthesis of Remdesivir using i-Pr₂NH as a cost-effective additive in yield 75% (Scheme 15). The reaction was conducted in the presence of i-Pr₂NH, and n-BuLi at ultralow temperature within 2 h using III as a protecting amino group of I. This procedure is unacceptable in large-scale production due to the ultralow temperature conditions.
Reagents and conditions: a) TMSCl, PhMgCl, i-PrMgCl-LiCl, THF, -20 °C, yield 40%; b) TMSCN, TfOH, TMSOTf, CH2Cl2, -78 °C, yield 85%; c) BCl3, CH2Cl2, -20 °C, yield 86%; d) 2,2-dimethoxyp propane, H2SO4, acetone, rt, yield 90%; e) X, MgCl2, (i-Pr)2NEt, MeCN, 50 °C, yield 70%; f) conc HCl, THF, rt, yield 69%. g) OP(OPh)Cl2, Et3N, CH2Cl2, -78 °C, then 4-nitrophenol, Et3N, 0 °C, yield 80%; h) i-Pr2O, yield 39%.

Scheme 13 Second generation synthesis of Remdesivir

Reaction conditions: a-i) TMSCl, PhMgCl, i-PrMgCl, THF, -20 °C, a-ii) NdCl3, n-Bu4NCl, THF, -20 °C, i and ii total yield 69%; b) TfOH, TMSOTf, TMSCN, DCM, -40 °C, yield 78%.

Scheme 14 Large-scale cyanation process using continuous flow chemistry synthesis of Remdesivir key intermediate

Scheme 15 Improved methodology for the synthesis of Remdesivir key intermediate by Xue et al. [157]
Wang et al. [158] reported a gram-scale catalytic asymmetric synthesis of a key step of Remdisivir in high chiral purity and yield (Scheme 16). Compound I was reacted with II in the presence of 2,6-lutidine, and 4 Å MS catalyzed by chiral catalyst A at lower temperature gave III in 89% yield with chiral purity higher than 99%. III was then deprotected by 37% HCl to obtain Remdesivir at a yield of 73%. In this procedure, Remdesivir was synthesized asymmetrically in short steps in high yield under mild conditions at gram-scale, and is thus suitable for large-scale production.

7 Ruxolitinib

7.1 Biological Activities of Ruxolitinib

Ruxolitinib is an FDA-approved targeting JAK1 and JAK2 inhibitor. It prevents the tyrosine phosphorylation of STAT1/3/5, which are downstream of cytokine receptors and drive T-ALL proliferation. Walker et al. [159] proved that Ruxolitinib and venetoclax worked synergistically to treat T-ALL in vitro, but were not effective in vivo. CXCR4-CXCL12 was implicated as the potential pathway that drives T-ALL infiltration into the central nervous system (CNS). By deleting the CXCR4 gene from T-ALL, they found prolonged survival in vivo with decreased neurologic clinical scores. Thus, T-ALL CNS infiltration should be blocked via CXCR4 inhibition. Ruxolitinib may be able to inhibit CXCR4 [160] (Fig. 24).

The 7H-pyrrolo[2,3]pyrimidine core of Ruxolitinib links to the hinge area of Cxc Chemokin Receptor 4 (CXCR4) by hydrogen bond, the nitrile function of Ruxolitinib binds to Ser936 by hydrogen bond, the pyrazole ring linking the side chain with the hinge binding adenine mimic acts as a structural template. Since it is not involved in direct interactions with the enzyme, bioisosteric replacement with a triazole ring is a promising strategy to increase synthetic accessibility.
In in vitro experiments, Ruxolitinib was diluted into 50 μM in DMSO. In in vivo studies, Ruxolitinib dissolved in DMSO was added to 5% DMA in H₂O. To survey the relevance of the JAK/STAT and BCL2 pathways on T-ALL proliferation and cell survival, Jurkat (mature T-ALL) and Loucy (early precursor T-ALL with high BCL2 expression) were assessed following treatment with Ruxolitinib. These cell lines were treated with different doses of Ruxolitinib over 72 h and assessed using a trypan blue exclusion assay and MTT (3-(4, 5-dimethylthiazolyl-2)-2,5-diphenyltetrazolium bromide) proliferation assay. Ruxolitinib decreased the survival and proliferation of both Jurkat and Loucy cell lines after 24, 48, and 72 h of treatment in 2.5 μM. Ruxolitinib failed to treat T-ALL in vivo because of leukemia CNS infiltration. In 2011, the FDA and European Food and Drug Administration (EFDA) approved Ruxolitinib first in class in treatment of myelofibrosis [161]. Tuttle et al. [162] used two mouse models to prove that interferon receptor genes are overexpressed in mice, and that JAK1 inhibitors can clearly restrain cytokine storm. Increased evidence proves that mortality with COVID-19 infections is caused mainly by the overexpression of a immune response to SARS-CoV-2, resulting in a cytokine storm and ARDS [163]. Many cytokines and chemokines involved in the cytokine storm employ JAKs for signal transduction. Cytokine analysis of COVID-19 patients shows that C-reactive protein (CRP) and interleukin (IL)-6 levels are significantly higher in patients who eventually died compared to those who survived [50]. Similar to some other mortal lung infections, the overexpression of immune response to the COVID-19 virus causes a cytokine storm, along with infiltration and activation of diverse immune cells, then generation secondary cell factors and chemotactic factors [164]. In this study, patients admitted to intensive care units (ICUs) showed significantly higher levels of IL-2, IL-10, IL-7, IP-10, MCP-1, MIP-1α, G-CSF, and TNF-α relative to non-ICU patients. All in all, these discoveries support the combination of antiviral treatment and targeted immunosuppression as a therapeutic program in COVID-19 [165].

7.2 Clinical Trials of Ruxolitinib

Several clinical trials have evaluated the safety and efficacy of Ruxolitinib of IL-6 and JAK/STAT signaling. Jung et al. [50] conducted a phase II clinical trial to
evaluate Ruxolitinib treatment of myelofibrosis. Their results proved the efficiency of Ruxolitinib to treat myelofibrosis. Studies on Ruxolitinib treatment of sHLH patients showed encouraging results on overall survival [166]. Cao et al. [167] carried out a phase II clinical trial to treat severe COVID-19 patients using Ruxolitinib. A total of 20 patients were distributed to the Ruxolitinib and standard of care (SoC) group, 21 patients were distributed to placebo group (SoC treatment) randomly. The results showed that the patients in Ruxolitinib and SoC group recovered faster than the placebo group. Most importantly, there were no deaths in the Ruxolitinib and SoC group.

7.3 Synthesis of Ruxolitinib

Rodgers et al. [168] synthesized racemic Ruxolitinib via three steps with a total yield of 48% (Scheme 17). Condensation of I with malonic acid in strong organic base condition obtained III, then III reacted with IV in the presence of 1,8-Diazabicyclo[5.4.0]undec-7-ene (DBU) gave V; finally, V was deprotected by TFA to racemate Ruxolitinib with a total yield of 48%.

Haydl et al. [169] reported Rhodium-catalyzed asymmetric coupling of (I) with pyrazole derivatives (II) giving enantioenriched allylic pyrazoles, which can be used to synthesize the targeted drug Ruxolitinib (Scheme 18). They developed a Rhodium-catalyzed, enantioselective synthesis of (R)-Ruxolitinib in the presence of chiral ligand using cheap material I, II and V in high chiral purity and yield. Above all, there were only three reaction steps.

Deepshikha et al. [170] provided a method for preparation of Ruxolitinib and its phosphate giving a chiral purity of 99.96% but with a total yield as low as 5%, which is unacceptable (Scheme 19). In this synthetic route, I was protected by 2-(trimethylsilyl)ethoxymethyl chloride, then direct Suzuki coupling with III gave compound IV with a yield of 80.8% in two steps. IV reacted with V in base condition, Reagents and conditions: a) THF, t-BuOK, 0 °C, 64 h, yield 89%; b) CH₃CN, DBU, rt, overnight, yield 93%; c) DCM, TFA, rt, 6 h, yield 58%.

Scheme 17 Preparation of racemic Ruxolitinib by Rodgers et al. [168]
Reagents and conditions: a) Cyclohexylallene, 4-bromopyrazole, PPTS (20 mol%), \([\{\text{Rh(cod)}\text{Cl}\}_2]\) (2.0 mol %), L2 (5.0 mol%), toluene, 80 °C, 24 h, yield 95%, 90% ee; b) 9-BBN, THF, rt; then H2O2, NaOH, rt, yield 99%; c) (COCl)2, DMSO, NEt3, -78 °C then rt, yield 97%; d) NH4OH, THF, rt, yield 90%; e) B2pin2, [Pd(dppf)Cl2](5.0 mol%), KOAc, DMSO, 90 °C; f. [PdCl2(PPh3)2] (5.0 mol%), K2CO3, 1,4-dioxane/H2O (2:1), 120 °C, step e and f total yield: 81%.

Scheme 18 Gram-scale synthesis of (R)-Ruxolitinib by Haydl et al. [169]

8 Conclusions

After the spread of COVID-19 worldwide, tens of thousands medical scientists and pharmacologists have made great efforts to search for potent drugs that can inhibit COVID-19, and they successfully found a few drugs like CLQ, CLQ-OH, Favipiravir and Remdesivir that are useful in the treatment of COVID-19 patients. But, in clinical trials to date, CLQ and CLQ-OH may increase mortality due to their
Favipiravir has high efficiency in treating Chinese patients, but it also needed a high dose (600 mg dose, 2–3 times daily); especially, there is a lack of clinical data to prove its efficacy and safety outside of China. Gilead Sciences conducted clinical trials with Remdesivir after the spread of COVID-19 worldwide, and the FDA authorized Remdesivir for compassionate use in treating COVID-19 patients in May 2020. To date, COVID-19 patients who die with cytokine storm syndrome have higher levels of IL-6 in plasma; four JAK inhibitors, Ruxolitinib, Tofacitinib, Baricitinib and Upadacitinib, have proved useful in the treatment of COVID-19 patients. Most importantly, there were no deaths using Ruxolitinib to treat COVID-19 patients in a phase II clinical trial.
As far as we know, there is no specific medicine for treatment of COVID-19 patients, but COVID-19 is still spreading rapidly worldwide. Combinations of antiviral drugs such as Remdesivir or GS-441524 and JAK inhibitor drugs may be a useful therapeutic schedule to reduce mortality before a specific medicine appears.

In this review, we introduced lots of small molecules that exhibit potent activity in inhibiting COVID-19, especially CLQ, CLQ-OH, Favipiravir, Remdesivir and Ruxolitinib, presenting their biological activities, clinical trials and synthesis processes, which may help researchers to systematically understand the processes of these potential drugs.

Scheme 20 Preparation of Ruxolitinib by Zhang et al. [171] synthetic route one

As far as we know, there is no specific medicine for treatment of COVID-19 patients, but COVID-19 is still spreading rapidly worldwide. Combinations of antiviral drugs such as Remdesivir or GS-441524 and JAK inhibitor drugs may be a useful therapeutic schedule to reduce mortality before a specific medicine appears.

In this review, we introduced lots of small molecules that exhibit potent activity in inhibiting COVID-19, especially CLQ, CLQ-OH, Favipiravir, Remdesivir and Ruxolitinib, presenting their biological activities, clinical trials and synthesis processes, which may help researchers to systematically understand the processes of these potential drugs.
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Scheme 21 Preparation of Ruxolitinib by Zhang et al. [171] synthetic route 2
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