Influence of Emerging Technologies on the Utilization of Plant Proteins

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Protein from plant sources is claimed alternatives to animal sources in the human diet. Suitable protein sources need high protein digestibility and amino acid bioavailability. In terms of protein functionality and food applications, they also need high-quality attributes, such as solubility, gelling, water- and oil-holding capacities, emulsifying, and foaming. Thermal processing can improve the nutritional quality of plants with some disadvantages, like reducing the assimilation of micronutrients (vitamins and minerals). Emerging technologies—such as ultrasound, high-pressure, ohmic heating, microwave, pulsed electric field, cold plasma, and enzymatic processes—can overcome those disadvantages. Recent studies demonstrate their enormous potential to improve protein techno-functional properties, protein quality, and decrease protein allergenicity. However, the literature lacks a broader evaluation, including protein digestibility, industrial-scale optimization, and exploring applications to these alternative protein sources.

Keywords: plant-based proteins, food processing, eco-friendly technologies, nutritional quality, in vitro protein digestibility, food safety

INTRODUCTION

Proteins are vital macronutrients in human nutrition, supplying the essential amino acids. It performs relevant functional roles in food formulation, processing, storage, and consumption. They also benefit food sensory and quality attributes, depending on their functional properties, such as solubility, gelling, water- and oil-holding capacities, emulsifying, and foaming (1, 2). The main current challenges regarding protein food sources are supply and distribution guarantees. Regarding animal protein production and consumption, there are concerns about adverse impacts on human health, natural resource depletion, climate change, and animal welfare (3). These concerns lead to a constant growing adoption of vegetarian and vegan diets. Food security awareness for the increasing world population (about 10 billion by 2050) drives the demand for sustainable protein sources (4). The most prominent new protein sources are non-conventional plants (including agro-industrial by-products), fungi, algae, and insects (5, 6). Although food-grade insects are excellent protein sources, they are not widely accepted by consumers, mainly due to cultural aspects. Fungi and algae are limited in terms of supply. Thus, plant-based proteins keep drowning the most attention among others.

In general, thermal processing improves plant protein nutritional quality. However, some disadvantages are high time- and energy-consuming procedures, large water expenditure, and losses of desirable compounds in the final product (7). The main chemical changes produced by heating are degrading heat-labile micronutrients, reducing vitamins and minerals assimilation, and
when the Maillard reaction occurs, generating toxic compounds and reducing the essential amino acids bioavailability (8). Elevated processing temperatures may also induce crosslinking, protein-protein interactions, and amino acid racemization (9).

Alternatively, some emerging food processing technologies have been investigated for the best protein employment, such as ultrasound, microwave, supercritical fluids, pulsed electric field, high-pressure, ohmic heating, cold plasma, and enzymatic processes (10, 11). Figure 1 illustrates the use of these techniques for valorizing plant-based proteins (11), which may contribute to environmental preservation by reducing wastewater production, organic solvents utilization, and processing time (12). Under mild temperatures, a balance can be reached between the processing feasibility with reduced environmental impact and the increased nutritional aspects and techno-functionalities of the proteins.

Plants have been studied and used worldwide as protein sources, including legumes, cereals, pseudocereals, and seeds (6). However, plant-based proteins are negatively associated with a diminished nutritional quality due to minor components that impair protein bioavailability to the human body. These antinutritional factors are protease (trypsin) inhibitors, polyphenols (tannins), phytates, fibers, haemagglutinins, and non-starch polysaccharides (8). Thus, a proper plant protein processing selection may affect the digestibility and nutritional value by inactivating or eliminating these compounds, modifying the profile of bioactive peptides, and changing the protein structure (13). Besides, some plant proteins present food allergenicity, including nuts, wheat, and soybean (14). Those emerging technologies are also being investigated to create hypoallergenic products (15) due to changing the protein conformation and the IgE epitopes of the allergens, making them less available to antibody receptors, which declines the protein allergenicity, as well as increase the protein digestibility (16).

Therefore, this critical review aims to discuss the advances and perspectives of the emerging processing methods—non-thermal or performed at mild temperatures/short times—to improve plant protein quality and properties. It is focused on the nutritional aspects, protein digestibility, protein allergenicity, and techno-functional properties.

TECHNO-FUNCTIONAL PROPERTIES OF PLANT PROTEINS

Protein functionality is critical in determining the applicability of plant proteins flours, concentrates, and isolates (17, 18). The biofunctionality is related to protein physiological and nutritional properties (e.g., antioxidant and antibacterial activity) (11). On the other hand, techno-functionality is associated with the impact on the physicochemical characteristics of food products, affecting the texture, appearance, stability, emulsifying, solubility, foaming, gelling, water- and oil-holding capacities, cohesion-adhesion, elasticity, and viscosity (19).

Table 1 presents some protein techno-functional properties and their relationship with food sensory and physicochemical characteristics.

Food macromolecules (e.g., polysaccharides, lipids, and proteins) are inherently functional by their molecular structure and ability to interact and form complexes. Protein molecular structures have important roles in determining food functionality and can be used as targets to alter protein functionality (20). Intrinsic and environmental factors determine the functional properties, stability, and shelf-life of foods.
containing functional proteins. The main intrinsic factors are the protein structure, conformation, amino acid composition, surface functional groups, net and surface electric charge, and hydrophobicity/hydrophilicity. Extrinsic factors are the medium pH, salts and solvents, ionic strength, temperature, and shear stress (21, 22). Protein extraction and processing may change those functional properties. Thus, it is also essential to study the parameters setup impact on a diversity of functional and physicochemical properties of food products.

Most proteins are functional due to their globular component properties, especially solubility, which is attributed to the amphiphilicity of these molecules. Proteins have both inwardly bounded apolar (hydrophobic) amino acids and outwardly exposed polar (hydrophilic) side-chain amino acid residues. This arrangement allows dipole-dipole interactions with solvents by twisting and unfolding the amonic acid side chains, placing the polar groups at the protein’s surface. It leads to networks that can form gels and develop films, hold water, absorb fat, foam, emulsify, and dissolve under various pH conditions (20). Also, the relative amount of α-helices, random coils, and the α-helix/β-sheet ratio in protein secondary structures of soybean and corn meals were positively correlated with protein solubility, while the percentage of β-sheet structures was negatively correlated with this same ability (23).

Some studies assessed the techno-functional properties of plant proteins, such as soybean, chickpea, kidney bean (24), mung bean, pea (22, 25), cowpea, lentils (26), amaranth, quinoa (16), cashew nut (27), sorghum (28), avocado (29), and mustard (30). Few studies investigated these properties in edible oil processing by-products, such as rapeseed meal protein solubility (31, 32). The utilization of plant proteins is limited due to their extremely low solubility at neutral pH, except for the soybean, pea, canola (9), and cowpea (33).

Other studies also evaluated some plant proteins’ foaming capacity and stability, like soybean, pea, chickpea, lupin, and rapeseed (34, 35). These sources have excellent foaming properties, comparable to egg protein, mostly due to high solubility, high surface hydrophobicity, low molecular weight, and net charge (36).

### TABLE 1 | Functional properties and their relationships with physicochemical and sensory properties of proteins (15, 49, 106, 107).

| Functional property | Definition | Physicochemical property and mode of action | Sensory property | Examples of plant proteins | Products and food systems |
|---------------------|------------|------------------------------------------|-----------------|----------------------------|--------------------------|
| Solubility          | Interaction of protein surface hydrophilic groups with water | Hydrophilicity, H-bonding and surface ionization, protein solvation, pH-dependent | Flavor, taste, mouthfeel, turbidity | Soybean, almond and rice proteins | Beverages |
| Foaming             | Formation films to entrap air and foam stabilization | Hydrophilicity, Hydrophobicity, film formation in the air/water interface | Mouthfeel, smoothness | Seeds protein | Desserts, ice cream, cakes, mousses |
| Emulsifying         | Formation and stabilization of emulsions | Hydrophilicity, Hydrophobicity, film formation in oil/water interface | Mouthfeel, flavor, smoothness | Seeds protein | Meat analogs, soups, sauces, desserts, cakes, ice cream, salad dressings |
| Gelling             | Capacity to form gels | Thermal aggregation, water entrapment, and immobilization, protein matrix formation | Mouthfeel, texture, smoothness | Seeds protein | Desserts, meat analogs, and bakery products |
| Oil-holding capacity | Fat entrapment | Hydrophobicity | Flavor, odor, smoothness | Seeds protein | Beverages, sausages, meat analogs, bakery products |
| Water-holding capacity | Water entrapment | Ionic hydration, H-bonding, hydrodynamic shape, and size, water-binding, | Texture, consistency | Soybean and cereal proteins | Meat analogs, cakes, bakery products |
| Viscosity           | Thickening | Hydrophobicity, Hydrophobicity, and size, water-binding, | Taste, consistency, mouthfeel | Soybean | Soups, salad dressings, sauces, desserts |
| Elasticity          | Stretchiness | Hydrophobicity, disulfide crosslinking deformable gels | Texture, crispiness, chewiness | Gluten protein | Meat analogs, extruded and bakery products |
| Cohesion and adhesion | Protein acts as an adhesive material | H-bonding, Ionic-bonding | Chewiness, stickiness | Seeds protein | Meat analogs, pasta, extruded snacks, and bakery products |
Some plant proteins have highlighted **emulsifying properties**, like the bell pepper, which formed stabilized emulsions with small oil droplet sizes (37); peas (34); chickpeas, with high emulsion activity index (EAI) at pH 10 (35); soybean, with a high emulsifying capability and emulsion stabilization against creaming during storage (38); and rapeseed, with higher emulsifying stabilities than soybean products (39).

Furthermore, few studies evaluated the **water- and oil-holding capacities** (WHC and OHC) of plant proteins. Bell peppers are suggested to food products requiring high WHC (37), while the OHC of peanut protein isolates was remarkably higher than commercial soybean protein isolates (40).

Although few studies evaluated the **gelling properties** of plant proteins, there are results about rapeseed products (flours, concentrates, and isolates) reporting poor gelation properties (41). However, soybean protein isolates have been used as gelling agents in several semi-solid food products, mainly for meat analogs (42).

Therefore, in terms of techno-functionality, there is little research reporting the solubility, emulsifying, foaming, water- and oil-holding capacities, and gelling properties for plant proteins. Potentially, plant-based proteins may be used by the food industry in formulations for protein supplements, meat analogs, beverages, snacks, desserts, bakery, whipped creams, soups, sauces, and salad dressings (22). From here, one can consider that exploring plant-based proteins aiming to develop technological alternatives for food formulation is an open field, including evaluating the required processing technologies for extraction and modulating the techno-functionalities.

**EMERGING TECHNOLOGIES FOR PROTEIN VALORIZATION, RECOVERY, AND IMPROVEMENT OF PROTEIN QUALITY**

The employability of plant proteins is related to the availability of their use, in addition to their intrinsic properties. The extraction yield from a food source is related to protein structure (primary to quaternary). Withal, the complexity of the food matrix influences the extraction process; proteins are generally bonded to other macromolecules (e.g., carbohydrates); the presence of salts and different pH values change proteins' charge, ionic strength, conformation, and solubility; previous matrix processing (e.g., defatting or heating) and the presence of water/solvents alters the matrix structure; and the fractionation of the plant sources (43–46). Also, the structural, functional, and sensory properties of extracted proteins are influenced by the conditions under which plant proteins are processed, e.g., temperature, time, pH, and ionic strength (44).

Protein extraction can be categorized into wet and dry methods (11). Subsequently to the extraction step, many technologies can be employed to purify or concentrate the protein of interest, aiming to obtain a food ingredient for different purposes. **Figure 2** presents a diagram with the methods most used for recovering proteins from agri-food materials, which could be applied in a biorefinery concept.

The main consequence of many extraction methods is protein denaturation, which alters the proteins' secondary, tertiary, and

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**FIGURE 2** Flow diagram representing the extraction methods most used for recovering proteins from agri-food. Adapted from Contreras et al. (9).
quaternary structures due to changes in temperature, pH, and organic solvents and salts presence (47). In general, denaturation makes the protein more susceptible to digestive enzymes, improving protein digestibility (21).

Alkaline extraction (pH 8–11), followed by isoelectric precipitation (pH 4–5) of solubilized proteins, is the most usual technique employed for the extraction of plant proteins to make enriched flours (up to 65% of protein), protein concentrates (65–90%), and protein isolates (more than 90%) (48). Several studies used the alkaline technique to produce protein isolates of plants, such as seeds, cereals, and legumes (9). However, the enormous water, energy, and chemicals requirement is a significant drawback of alkaline extraction (19). Besides, high protein purity and yield are not guaranteed, which are affected by the processing conditions (e.g., equipment configuration, extraction time, temperature, pH, ionic strength, net charge, presence of salts, protein content, and protein solubility) (26, 49). Also, extreme extraction conditions (e.g., highly acid or alkaline medium and high temperature) may reduce protein nutritive value by changing the amino acid profile and degrading bioactive compounds (9, 11, 50). Furthermore, acid precipitation and neutralization tend to reduce protein solubility and negatively impact other techno-functional properties, such as gelling and foaming (51). Therefore, the alternative technique using membranes (e.g., ultrafiltration) is a less-energy consuming alternative to concentrate proteins instead of isoelectric precipitation, reaching the manufacture of protein ingredients (21, 26) and resulting in improved protein recovery yield, preserved techno-functional protein properties, and higher purity.

The existing drawbacks of conventional extraction methods (e.g., high temperature, energy consumption, wastewater, and organic solvents utilization) (19) may be overcome by emerging technologies using mild conditions. The most prominent alternatives are ultrasound, high-pressure, microwave, pulsed electric field, ohmic heating, and enzymatic processes. They potentially increase the protein extraction yield while reducing chemicals and water consumption (9–11, 50, 52, 53). However, a key question is whether these novel technologies can extract proteins from agri-food sources efficiently and cost-effectively. Data about the required energy and costs are scarce, and these approaches were mainly performed at a laboratory scale or are still in the early stage of their industrial applications; therefore, the large-scale feasibility still needs further studies.

Although these emerging processes were evaluated about protein valorization and recovery, studies concerning protein quality improvement (e.g., protein digestibility and inactivation of the so-called “antinutritional factors”) are rare for plant protein sources. Few evaluations have been made for the inactivating trypsin inhibitors in plant proteins in soybeans, chickpeas, and beans (54). However, as presented in the following, reasonable indications are that those methods are alternatives for plant-based protein processing.

High-Pressure Processing

High-pressure processing (HPP)—also known as high hydrostatic pressure (HHP) or high isostatic pressure (HIP)—is a non-thermal technology using hydrostatic pressures up to 1,000 MPa into a product in controlled temperature and time conditions (55). HPP affects the structure of the non-covalent bonds, increases the surface hydrophobicity, and causes protein denaturation, aggregation, or gelation (56). The protein secondary and tertiary structures significantly change at pressures higher than 200 MPa due to the consequent denaturation and aggregation of plant proteins with increased pressure, which changes the conformation and coagulation of their native structures because of the disruption of interactive forces, mainly hydrophobic and electrostatic bonds (57). HPP also improves protein functionality and digestibility of cereals and legumes (58), inactivating the antinutritional factors on a laboratory scale (59).

High-pressure homogenization (HPH)—also called dynamic high-pressure (DHP) or ultra-high pressure homogenization (UHPH)—imposes high-pressure conditions by pumping liquid food through a tiny gap in a valve, increasing velocity and causing high shear stresses. Consequently, it causes changes in food rheological properties. HPH utilizes the combined effectiveness of high-frequency vibration, high-velocity impact, quick pressure drop, cavitation, and intense shear stress in a short time, which causes a significant effect on proteins conformation (60). HPH was also recently applied to food products aiming at microbial inactivation and changes in the protein techno-functional properties (55).

Typical HPH pressures are moderate and usually up to 100 MPa (61), while HPP can reach ten times more. In addition to the range of applied pressures, another difference between HPP and HPH is the molecules’ movements during treatment (lower in HPP and increased in HPH), leading to different protein structures after the treatments, bringing many interaction possibilities between polypeptides and protein aggregation. Also, HPP is governed by the ordering principle (33), whereas during HPH, high shear forces perturb protein structures (62).

Table 2 shows examples of high-pressure technology applied to plant proteins. The use of elevated pressure favors extracting protein from plants, decreasing solvent consumption, increasing extraction yields, and shortening the extraction time. High-pressure can modify protein techno-functional properties and reduce protein allergenicity (63, 64). HPP has also been used to extract proteins from some agri-food residues (e.g., wheat bran, grape pomace, and corn stover) (9), reduce food allergenicity, and inactivate some compounds detrimental to protein digestion.

Ultrasound

Ultrasound (US) is a non-thermal technology using high-intensity and low-frequency sound waves, ranging from 20 to 100 kHz (11). The basic principle of ultrasound technology is the cavitation phenomenon, where air bubbles are formed within the liquid phase, their volume increase, and finally explodes (19). US accelerates the mass transfer of compounds, provides high shear forces in the extractive agent (50), and improves solubility due to cellular structure’s high stress and deformation (19). The mass transfer is also facilitated because microchannels may occur when bubbles collapse. The increased temperature, turbulence, and mixing effects by cavitation in the US also increase extraction...
### TABLE 2 | Examples of emerging technologies application on plant proteins.

| Process | Objective of the study | Processing conditions | Results | Protein yield | References |
|---------|------------------------|-----------------------|---------|---------------|------------|
| HPP     | Soybean                | Protein allergenicity  | 350 MPa | Reduced allergenicity by 46.6% | * (64) |
|         | Soybean protein isolate| Reduction of antinutritional factors | 200–700 MPa | Efficient to eliminate the phytates, however, not effective to reduce trypsin inhibitor | * (108) |
|         | Soybean protein isolate| In vitro protein digestibility | 400–600 MPa | Increased IVPD 68% | * (109) |
|         | Soybean protein isolate| Functional properties  | 100–300 MPa | Foaming increased and viscosity decreased | * (110) |
|         | Soybean slurry (by-product) | Protein extraction | 50–125 MPa | Good results of extraction yield at 100 MPa | 82% (65) |
|         | Kidney bean protein isolate | Functional properties | 300–600 MPa | Production of isolate with higher functionality | 23.5% (10) |
|         | Peanut protein isolate | Functional properties | 50–200 MPa | Improved water- and oil-holding capacities, but not improved gelling property | * (40) |
|         | Sweet potato protein | In vitro protein digestibility | 200–600 MPa | Increased IVPD from 53.8 to 59.1% in 30 min | * (111) |
|         | Sweet potato protein | Functional properties | 250–550 MPa | 400 MPa was a good choice for preparing novelty food products with structural modification | * (112) |
|         | Sweet potato protein | Gelation behavior | 400 MPa | Textural properties of gels were improved by sulfur-containing amino acids, especially by cysteine | * (56) |
|         | Macuna bean protein isolate | Protein extraction and color evaluation | 200–600 MPa | Inactivation of polyphenol oxidase and improvement of the color of protein isolate | 8–34% (113) |
|         | Pea protein isolate | Functional properties | 200–600 MPa | Improvement of emulsion and foaming capacities | * (114) |
|         | Fababean | Functional properties | 103–207 MPa | Improvement in solubility and foaming capacity and decreased emulsifying capacity | * (115) |
|         | Lentil protein isolate | Functional properties | 34–103 MPa | Decreased surface hydrophobicity and increased zeta potential | * (116) |
|         | Hazelnut | Functional properties | 25–150 MPa | Improvement of solubility, foaming, emulsifying capacity, and emulsifying stability | * (117) |
|         | Mung bean, chickpea, pea, lentil, and faba bean yogurts | Rheological analyses | 600 MPa | Viscosity and viscoelastic properties of plant protein gels was comparable to commercial dairy yogurts | * (118) |
|         | Potato protein isolate | Gelation properties | 300–500 MPa | High pressures can allow the formation of gels from potato protein isolate as a novel plant-based protein source | * (119) |
| Process | Objective of the study | Processing conditions | Results | Protein yield | References |
|---------|------------------------|-----------------------|---------|---------------|------------|
| Cowpea  | Gelation properties    | 400–600 MPa 5 min     | HHP-induced gels were less hard and adhesive than heat-induced ones | *         | (33)       |
| US      |                        |                       |         |               |            |
| Soybean | Inactivation of trypsin inhibitor | 20 kHz 20 min | Inactivation of trypsin inhibitor by 55% | *         | (120)      |
| Soybean protein isolate | Emulsifying property | 200–600 W 150–450 W 15–30 min | Improved emulsifying capability | *         | (38)       |
| Soybean protein isolate | Gelation properties | 20 kHz 150–450 W | Under 300 W, the gel hardness reached a maximum of 998.9 g, with water binding capacity of 87% | *         | (121)      |
| Soybean okara (by-product) | Protein extraction | 20 kHz 65 W 15 min | US improved the extraction of up to 10% | 70%       | (66)       |
| Soybean milk | In vitro protein digestibility and inactivation of trypsin inhibitor | 25 kHz 400 W 1–16 min | US significantly reduced trypsin inhibitor activity up to 52% and improved the digestibility of proteins in soymilk | *         | (122)      |
| Millet protein concentrate | Functional properties | 20–100 W 18.4–73.9 W/cm² 5–20 min | Improvement of solubility and emulsifying capacity | *         | (71)       |
| Pea protein concentrate | Functional properties | 412.5–712.5 W 336–582 s 2–30 min | Emulsifying properties were greatly improved | *         | (18)       |
| Pea protein isolate | Foaming property | 20 kHz Amplitude of 30–90% 30 min | Foaming ability increased from 145.6 to 200% and foaming stability increased from 58 to 73.3% | *         | (25)       |
| Soybean and rice protein isolates and pea protein | Functional properties | 20 kHz 562.5–712.5 W 120–600 s | Functional properties are improved as the dispersibility of protein materials increases (712.5 W, 600 s) | *         | (123)      |
| Potato protein | In vitro protein digestibility and functional properties | 20–60 kHz 2–30 min 40°C | Solubility and digestibility of potato protein was significantly improved by online ultrasound-assisted pH shifting treatment | *         | (124)      |
| Barley protein isolate | Functional properties | 20 kHz 100% amplitude | Improved protein solubility and colloidal stability especially at alkaline pH | *         | (125)      |
| Sunflower protein isolate | Functional properties | 20–40 kHz 5–30 min | Improved solubility, emulsifying, foaming and oil-holding capacity and decreased water-holding capacity | *         | (126)      |
| Olive kernel | Protein and phenolic compounds extraction | 400 W 24 kHz 100% amplitude | Potential use for protein extraction | 25%       | (67)       |
| Tamarind seed protein isolate | Functional properties | 100–200 W 15–30 min | Solubility, emulsifying, foaming capacity, water-and oil-holding capacity was the highest when both time and intensity of treatment were high | *         | (127)      |

(Continued)
| Process | Objective of the study | Processing conditions | Results | Protein yield | References |
|---------|------------------------|-----------------------|---------|---------------|------------|
| Bell pepper seed protein isolate | Protein extraction and functional properties | 350 W | High oil-holding capacity, low solubility, and low foaming property | 6% | (37) |
| Pea | Functional properties | 68 W/100 mL 20 kHz | Both pH-shifting at pH 12 and power ultrasound treatments were effective in modifying the properties of pea | * | (128) |
| **PEF** | | | | | |
| Grape juice | Impact on the protein structure | 35 kV/cm 4 µs pulses at 1,000 Hz | No evidence that PEF affects the primary structure of proteins and amino acid content | * | (129) |
| Rapeseed stems and leaves | Protein and polyphenols extraction | 0.2–20 kV/cm | Enhanced protein yield | 80% | (130) |
| Alfalfa leaves | Protein extraction | * | Increase of protein extracted by PEF | 57% | (131) |
| Olive kernel | Protein and phenolic compounds extraction | Pulse voltage of 40 kV | Increased the total phenolic content and proteins of the recovered extracts | 25% | (67) |
| Pea, rice, and gluten protein concentrates | Functional properties | 60,000 pulses 1.65 kV/cm | PEF was able to modify protein structure by inducing unfolding, intramolecular rearrangement, and formation of aggregates. These effects were strongly dependent on protein nature and pH | * | (68) |
| Blackberries | Protein and phenolic compounds extraction | 40 kV–10 kA 0.5 Hz 13.3 kV/cm | The maximum anthocyanin yield was found after applying PEF treatment | 38 mg/100 g | (69) |
| **MH** | | | | | |
| Beans | Inactivation of trypsin inhibitor | 2,450 MHz 5–20 min | Effective for inactivation of trypsin inhibitor (97–100%) of different varieties of beans | * | (132) |
| Soybean | Inactivation of trypsin inhibitor | 2,450 MHz 500 W 2–4 min | Trypsin inhibitor was completely inactivated | * | (45) |
| Soybean milk | In vitro protein digestibility and inactivation of trypsin inhibitor | 2.45 GHz 70–100 °C 2–8 min | Increased digestibility by 7% Trypsin inhibitor activity reduced to 1% | * | (54) |
| Soybean milk | In vitro protein digestibility and inactivation of trypsin inhibitor | 2,450 MHz 70–100 °C 2–10 min | Digestibility of soymilk significantly increased up to 93% after 10 min microwave processing at 85 °C | * | (122) |
| Soybean milk | Protein extraction, digestibility and functional properties | 540–810 W 70–90 °C 140–180 rpm | The optimal microwave-assisted extraction conditions were 675 W, 80 °C and 160 rpm | 24% | (133) |
| Rapeseed meal | In vitro protein digestibility | 800 W 2–6 min | Microwave for 2 and 4 min increased IVPD and for 6 min decreased IVPD | * | (100, 134) |
| Peanut peptides | Degree of hydrolysis | 9.5 min 600 W 50 °C | DH reach 26.1% | * | (135) |
| Chickpea | Comparison of process time with conventional methods | 400–600 W 14–56 s | Reduction of cooking times from microwave | * | (76) |

(Continued)
| Process | Objective of the study | Processing conditions | Results | Protein yield | References |
|---------|------------------------|-----------------------|---------|---------------|------------|
| Chickpea | In vitro protein digestibility and inactivation of trypsin inhibitor | 2,450 MHz 15 min | IVPD were improved, and trypsin inhibitor activity was significantly decreased | * | (136) |
| Coffee silverskin protein (by-product) | Protein extraction | 434.7 W 10–20 min | Microwave-assisted extraction have potential to be a rapid and effective tool for protein extraction from coffee silverskin | 43.53% | (46) |
| CAPP | Pea protein isolate | Functional properties | Air DBD 8.8 kVPP 3kHz 10 min | Improvement of protein solubility, water- and oil-holding capacities | * | (88) |
| | Peanut protein isolate | Functional properties | DBD 36 kV 1–4 min | Improvement of emulsion stability, solubility, and water-holding capacity | * | (137) |
| | Wheat flour | Functional properties | Air 20 V 9kHz 120 s | Increase in the dough strength | * | (138) |
| | Soybean protein isolate | Functional properties and allergenicity | DBD 40–60 kV 80–100kHz 1–10min | CAPP induced reactive oxygen species resulting in modifications in the secondary and ternary structures. Functional properties such as emulsifying and foaming properties (60 to 194%) were influenced. The IgE-binding level was decreased by up to 75% (120Hz, 5 min) | * | (83) |
| | Rice flour | Amino acid composition | DBD 60–70 kV 5–10 min | Higher content of amino acids for samples treated with cold plasma (glutamic acid, asparagine, serine, histidine, threonine, tryptophan, isoleucine, phenylalanine, and proline) | * | (139) |
| | Wheat grain and flour | Functional properties | DBD 80 kV 5–30 min | Plasma treatment increased the flour hydration, pasting and viscosity properties of wheat flour | * | (86) |
| EAEP | Soybean | Protein extraction | Protease M® pH 4.5 50–100°C 10–120 min | Good results of protein yield | 59.3% | (140) |
| | Peanut | Protein extraction | Alcalase® 1.5% 60°C pH 9.5, 5h | Good results of protein yield | 71.4% | (141) |
| | Sesame bran | Protein extraction | Viscozyme L® Alcalase® 25–55°C 10–120 min | Good results of protein yield | 88.8% | (50) |
| Process                          | Objective of the study                                      | Processing conditions                                      | Results                                      | Protein yield | References |
|---------------------------------|------------------------------------------------------------|------------------------------------------------------------|----------------------------------------------|---------------|------------|
| Rice bran                       | Protein extraction                                         | Alcalase® 50°C                                            | Good results of protein yield               | 44.8%         | (142)      |
| Oat bran                        | Protein extraction and functional properties               | Amyloglucosidase 55°C pH 11.5, 60 min                     | Good results of protein yield               | 82%           | (143)      |
| Moringa oleifera seed           | Protein extraction                                         | Protex 7L® 45°C 15 min                                    | Good results of protein yield               | 75.4%         | (144)      |
| Rapeseed meal                   | Protein extraction                                         | Viscozyme® Alcalase® 80 min                                | Good results of protein yield               | 82.1%         | (145)      |
| Almond cake                     | Protein extraction and protein digestibility              | FoodPro Alkaline Protease® 50°C pH 9.0 120 rpm 1h        | 64% of protein digestibility               |               | (146)      |
|                                 |                                                            |                                                            |                                              |               |            |
| EH                              |                                                            |                                                            |                                              |               |            |
| Palm kernel cake                | Improve nutrient utilization                               | Mannanase 1–20% 2–12 h                                    | Mannanase improved nutrient release of reducing sugar, total sugar and proteins | *             | (147)      |
| Chickpea protein isolate        | Functional properties                                     | Alcalase® pH 8.0 50°C 210 min                             | Improvement of protein recovery, solubility, and emulsifying properties | *             | (148)      |
| Peanut protein isolate          | Functional properties                                     | Papain 130°C                                              | Enhanced DH and increased protein solubility | *             | (97)       |
| Beans                           | In vitro protein digestibility                            | Proteases 28°C 150 rpm 5 h                                | Enzyme treatment improved the IVPD of the four bean varieties | *             | (149)      |
| Lupin protein isolates          | Functional properties                                     | Alcalase 2.4L®, Papain® 7098L®, and Neutrase 0.8L®       | The enzymatic hydrolysis increased their techno-functional properties (protein solubility, foam activity, and emulsifying capacity) independently of the enzyme preparation | *             | (99)       |
| DSI                             |                                                            |                                                            |                                              |               |            |
| Pea and rice protein isolate    | Functional properties                                     | 107°C pH 9–11                                             | Enhanced solubility, emulsifying, foaming, and gelling for protein treated by DSI | *             | (101)      |
| RW                              |                                                            |                                                            |                                              |               |            |
| Chickpea protein isolate        | Functional properties                                     | 90°C 20 min                                               | RW samples had a better water-holding capacity and emulsifying stability compared to freeze-drying samples | *             | (35)       |
| GI                              |                                                            |                                                            |                                              |               |            |
| Sunflower meal                  | In vitro protein digestibility                            | 10–20 kGy                                                 | Improved the IVPD (85.5%)                   | *             | (100)      |
| Rapeseed                        | Phytic acid concentration                                 | 15–45 kGy                                                 | 100% inactivation of phytic acid            | *             | (100)      |

*Data not found in the respective study.

CAPP, Cold-atmospheric pressure plasma; DBD, Dielectric barrier discharge; DH, Degree of hydrolysis; DSI, Direct steam injection; EAEP, Enzyme-assisted extraction processing; EH, Enzymatic hydrolysis; GI, Gamma irradiation; HPP, High pressure processing; IVPD, In vitro protein digestibility; MH, Microwave heating; PEF, Pulsed electric field; RW, Refractance-window; US, Ultrasound.
efficiency (19). Consequently, the US can modify proteins by affecting H-bonds, increasing protein recovery, and reducing extraction time and protein aggregates (9). Also, improved protein functionality can occur. The cavitation bubbles on the protein surface result in micro-jetting and particle breakdown, improve solvent permeation into the food matrix and change the protein allergen conformation and reactivity (9, 11).

As a novel technology, the US appealed to environmental sustainability. High-intensity ultrasound is a quick and cost-effective technology used to modify globular proteins’ structural and functional properties (25). The US was used to recover valuable proteins from food industry by-products, e.g., soybean okara (65, 66) and olive kernel (67). Table 2 summarizes the US technology application to plant proteins.

**Pulsed Electric Field**

Pulsed electric field (PEF) treatment consists of electric pulses of short duration ($10^{-4}$ to $10^{-2}$ s) and relatively high amplitude (0.1–80 kV/cm). It induces a critical electrical potential across cell membranes, enabling an easier extraction of proteins (9, 11). PEF is a promising non-thermal food processing method that is efficient in cell disintegration (12) and microbial inactivation. PEF also induces changes in the protein hydrophobicity and structure (secondary and tertiary) and dissociates the non-covalent bondings, improving the protein functionality (68). PEF technology may cause lethal damage to cells or induce sub-lethal stress by transient permeabilization of cell membranes and electrophoretic movement of charged species between cellular compartments. Therefore, PEF application can facilitate the selective recovery of valuable compounds without deteriorating the treated matrix, favoring the subsequent separation and purification stages (69). Exposure of viable cells to PEF increases cell membrane permeability due to electroporation. This phenomenon can be used in biorefineries to extract and introduce molecules into the cells. The PEF application on plant cell tissues can positively change the membrane transport properties, facilitating the extraction of targeted molecules (e.g., proteins). Figure 3 schematically represents the impacts of PEF on any viable cells, where the possible outcomes depend on the PEF setup protocols (amplitude, number, shape, and pulses duration) and additional techniques (e.g., electrophoresis) (12).

The PEF advantages are increased mass transfer, improved extraction yield, decreased processing time, reduced compounds degradation (e.g., flavors and proteins), and reduced energy costs (68). The application of PEF is a potential alternative to recover high-added value compounds from food matrices and residues, which reduces waste disposal and extends limited resources. However, these studies were performed at the laboratory scale, and further investigations are needed to address the large-scale feasibility and energy costs.

Recent studies used PEF on the recovery of bioactive compounds from plants (e.g., betalain from red beet, betacarotene from carrot, sucrose from sugar beetroot, inulin from chicory, and phenolics from grapes), agri-food residues, such as pomace, peels, kernels, and oilseed meals (e.g., carotenoids, chlorophylls, sterols, and polyphenols), and marine microalgae without killing the cells (12, 49, 70). However, the literature using PEF to improve protein digestibility and reduce antinutritional factors of plant proteins is scarce, and further research on this topic may be worthy. Table 2 presents applications of PEF to plant proteins.

**Ohmic Heating**

Ohmic heating (OH) is an advanced thermal processing method that applies electrical current to generate heat inside a food material by the well-known Joule effect (71). OH emerged as an alternative method to food thermal pasteurization and sterilization (11) and can promote higher yields and lower processing time than the conventional methods, bringing significant nutrient retention and preservation of the food

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**FIGURE 3** | Schematic representation of the effect on cells exposed to pulsed electric fields. Adapted from Golberg et al. (12).
quality (60). The existence of electrolytic components, such as salt and acids, allows the electric current to pass through food materials, which is the basis of the OH technique, generating heat internally (72). Besides heating, OH causes electroproteination of cell membranes, increases the electrical conductivity and permeability, and positively influences the extraction rates of different biomolecules (11).

It was shown that OH inactivates trypsin inhibitors due to the electrochemical effects, depending on the electric voltage used. Studies using OH (50 Hz, 220 V, 3 min) showed more efficient inactivation of the trypsin inhibitors than the electric stove over the same processing time (59). The heat produced by the OH depends on the food material's electrical field strength and electrical conductivity. Low electrical conductivity heats slower than higher ones if the same electrical field strength is used. Food materials with 0.01–10 S/m of electrical conductivities are considered proper for the OH technique (73). OH was a pretreatment to soybean oil recovery, in which 600 V for 10 min promoted a 73% yield at 90°C (74). However, no studies using OH to assist protein extraction or improve plant proteins' functional properties and digestibility were found. Thus, there are opportunities for new applications for this emerging technology.

**Microwave**

Microwave heating (MH) uses non-ionizing electromagnetic waves from 300 MHz to 300 GHz (75). Microwaves cause heating due to the interaction of the alternating electromagnetic field with the food chemical constituents. MH of food materials occurs due to dipole and ionic mechanisms, and the dielectric properties and penetration depth are the most important characteristics affecting the process (76).

Microwave is mostly applied to foods for short cooking time, with less energy consumption. However, this technique can also influence the extraction of proteins (46) and other nutrients in food matrix (e.g., polyphenols and polysaccharides) (77). Moreover, the advanced MH use may improve the nutritional food quality by combining operating conditions that preserve more nutrients (e.g., vitamins and heat-labile amino acids like lysine, tryptophan, and sulfur amino acids) and sensory aspects (78–80). MH affects the conformational properties of food proteins (secondary structure) and accelerates their denaturation without changing their primary structure (60). The microwave processing disrupts H-bonds, increases the food matrix porosity, and allows dissolved ions migration. These food changes facilitate the extraction and the improvement of the protein functional properties (11). MH is also useful for inactivating the antinutritional factors usually present in plant proteins (59) and improves protein digestibility (8). Table 2 presents MH applications to plant proteins.

Literature shows some studies that use microwave heating to reduce the cooking time of plants, reduce the concentration of antinutritional factors, and enhance protein digestibility (8). However, there is scarce evidence of efficiently using this technology to extract proteins from plant sources and agro-industrial waste.

**Cold Plasma**

Plasma technology (or gas discharge plasma) involves producing and using ionized gas molecules to treat a material for superficial effects, e.g., polymers functionalization and food decontamination. The gas ionization produces reactive species, e.g., ions, electrons, excited atoms, ultraviolet (UV) photons, and free radicals (81–83), that can cause different changes on the food surfaces (solids food) and food bulk. Plasma can be generated at different temperatures and classified into thermal and non-thermal (also called cold) plasma (81, 84). Depending on the pressure condition, plasma can also be classified as low-, high-, or atmospheric-pressure plasma (85).

Cold atmospheric plasma processing (CAPP) is an emerging, sustainable, and environmentally friendly technology that has received significant attention in the food industry for the recent decade (82, 83, 86). CAPP can be generated by corona discharge, atmospheric-pressure plasma jet (APPJ), dielectric barrier discharge (DBD), microwave discharge (85, 87), and radiofrequency (RF) (88). It produces reactive oxygen species, including singlet oxygen and ozone, and exciting molecular nitrogen (81, 89). CAPP has an inactivation effect on microorganisms, such as food pathogens, spores, and viruses (84, 85, 89). Also, it is valid for surface modification (90, 91), inactivation of deteriorating enzymes (82), food packaging modification (85), and reduction of food allergenicity (83). According to processing conditions, CAPP influences food proteins' conformation and functional properties, e.g., plasma source, design, treatment power, pressure, reactive gas type, exposed time, and sample nature (84, 85).

Reactions initiated by reactive oxygen species (ROS) with the synergistic effect of reactive nitrogen species (RNS) are the leading cause of protein structure modification, triggering the cleavage of proteins into peptides (84). Also, CAPP can damage proteins, amino acids, nucleic acids, and lipids (81, 84, 89). The plasma protein denaturation mechanism is associated with reactive species interaction with amino acids and secondary structure by losing α-helix and β-sheet (85). Besides, it was observed that sulfur and aromatic amino acid content decreased after plasma exposure (84, 92).

There are few studies regarding CAPP effects on protein modification and the techno-functional properties of plant proteins and by-products. Table 2 brings some applications of CAPP technology to plant proteins present in the current literature. There is no published report on the extraction and protein quality enhancement of plant proteins in terms of protein digestibility by plasma technology. Also, no studies about the protein recovery of plant proteins from by-products using CAPP were found in the literature.

**Enzymatic Processes**

Enzymes are biocatalysts in many industries (e.g., food, chemical, and pharmaceutical). The enzyme-assisted extraction processing (EAEP) simultaneously extracts oil and protein from plants. EAEP has a low environmental impact due to avoiding organic solvents and can be labeled as an eco-friendly technique that produces valuable products in mild conditions without losing quality (50).
hydrolysis (EH) technique may generate protein hydrolysates, such as di- and tri-peptides, and bioactive peptides from plant proteins, which contribute to the production of bio-functional foods (94, 95). EH is better than conventional chemical hydrolysis (e.g., acid or alkali) due to high product quality, reduced processing time, and mild conditions (17).

The endopeptidases (e.g., Alcalase®) and the exopeptidases (e.g., Flavourzyme®) used in the EAEP process break down some peptide bonds (17, 96). The resulting carboxyl groups and free amino acids increase the isolated proteins’ nutritional value, digestibility, and functional properties (9, 49, 97). EH decreases molecular weight, increases hydrophilicity, and changes protein conformation (17, 97).

Multi-enzyme cocktails may be a strategy to improve hydrolytic reactions (96). The enhanced functionality in protein hydrolysates depends on the extent of hydrolysis and process conditions (e.g., pH, temperature, and enzyme/substrate ratio) (17). The degree of hydrolysis (DH) is the percentage of peptide bonds cleaved per gram of protein over the total number of peptide bonds (98). DH is crucial for obtaining desirable results and avoiding excessive protein hydrolysis that can impair functionalities and sensory attributes (38, 98).

EH can result in hypoallergenic foods due to peptides’ production that does not trigger IgE antibody binding activity, lowering protein allergenicity (99). High-quality protein hydrolysates have applications in food formulations destined for individuals with special diets (e.g., infants, elderly, allergic individuals, or medical nutrition) or those seeking a higher protein intake (e.g., athletes) (17, 49). However, there are some challenges regarding protein hydrolysate applications. The significant drawbacks of protein hydrolysates are their palatability and the bitter taste due to the release of bitter hydrophobic peptides (8). Table 2 shows examples of applications of enzymatic hydrolysis to plant proteins.

Other Emerging Technologies

Other processes commented in the literature allow the extraction and conformation changes, leading to high yield and improved protein quality. Examples are gamma irradiation (100), direct steam injection processing (DSI) (101), and refractance-window (RW) and cast-tape drying (CTD) (35).

DSI may lead to protein denaturation due to product exposure to high temperatures for short periods. DSI changes proteins’ 3D conformation, like disulfide bonds and free S-H, and the protein surface becomes more hydrophilic, enhancing the functional properties (e.g., solubility) without affecting the essential amino acid composition (101).

CTD and RW are emerging techniques used for dehydrating liquid and semi-liquid foods (102, 103). The heat transfer occurs fastly to the product bottom by conduction (104). The processing time is relatively short, leading to minor changes in product nutritive value, conserving, or improving the protein functional properties (35).

Gamma irradiation (GI) may improve protein extraction and cause conformational changes (secondary structure rearrangement), crosslinking, and break covalent bonds. The protein structure and concentration and the presence of oxygen are the main factors affecting this process. However, GI may promote amino acid oxidation (8, 100).

Table 2 shows DSI, RW, and GI applications to plant proteins. There are rare studies on extracting plant proteins and evaluating their protein digestibility and techno-functional properties.

Combined Emerging Technologies

Some studies show the combined effect of some presented emerging technologies (e.g., US, HPP, MH, OH) with the enzymatic processes on enhancing protein recovery, extraction yield, and improving protein functional properties (10, 105). The improvement of protein extraction and functional properties results can be explained due to the synergistic effect of the mentioned technologies related to the increase of mass transfer and the influence of the conformational and protein structure changes, for example. Table 3 shows applications of combined processes on plant proteins. These studies showed combined techniques as more efficient in enhancing protein functional properties. Nevertheless, more research is required to validate their application to improve the nutritional quality of plant proteins.

PERSPECTIVES AND CONCLUDING REMARKS

The emerging technologies for food processing are alternatives to traditional thermal processing to enhance plant proteins’ nutritional quality or techno-functional properties. Protein quality is the main concern when replacing traditional sources, particularly meat by plants. It was observed that the selected processing technologies could interact with the protein structure (from primary to quaternary) and inactivate/eliminate the antinutritional factors, which indicate changes toward higher protein digestibility. This effect was demonstrated for high-pressure, ultrasound, microwave, ohmic heating, gamma irradiation, and enzymatic processes. In particular, trypsin inhibitors were inactivated by ultrasound, microwave, and ohmic heating, while gamma irradiation was suitable to inactivate the phytic acid. Protein allergenicity is also a concern for adding alternative protein sources in a diet, and it was demonstrated that enzymatic processes, cold plasma, and high-pressure were also efficient in reducing the allergenicity of these alternative proteins.
High-pressure processing is the most extensively explored process and presented many good results for techno-functional properties (foaming, emulsifying, and water- and oil-holding capacities), reducing allergenicity and antinutritional factors, and, as a consequence, improving plant protein digestibility and food applications. Those techno-functionality improvements result from changing the protein structure (from secondary to quaternary). Most of the emerging technologies discussed here can promote similar changes to the protein molecules, and, in this way, they are potential methods to impact those properties positively. In some studies, these properties were affected by ultrasound, pulsed electric field, microwave, cold plasma, direct steam injection, refractance-window drying, and enzymatic processes. In addition to the mentioned benefits, both ultrasound and enzymatic processes are suitable for increasing protein extraction yield and bioactive compounds recovery.

The promising results found about these technologies can be explored to nutritionally enrich traditional plant origin sources, such as soybean, peas, beans, and chickpeas. This work claims the broad perspective of these emerging technologies for achieving the adequate protein quality of plant origin sources. However, there are scarce studies evaluating protein digestibility and amino acid composition of plant proteins when using these emerging technologies. There is no published data on ohmic heating and cold plasma impact on plant protein quality enhancement or functional properties. Furthermore, combining some of these technologies has been indicated as a suitable strategy for better results, and many further investigations can be carried out to find optimal conditions.

Finally, the industrial application feasibility needs additional development at scales more extensive than those implemented at the laboratory. An actual industrial processing method must consider economic, environmental, and food security issues, which will supply sufficient proteins to the growing global population. Besides, the technological selection must meet human nutritional and sensory requirements and the consumer’s cultural aspects. A key question is whether plant proteins can be extracted efficiently and cost-effectively and preserve nutritional value for human consumption worldwide.

**AUTHOR CONTRIBUTIONS**

AS conceived and designed the work, performed the data search, figure and tables preparation, and wrote the draft. JL and YM revised the draft. BC conceived and designed the work, and revised the draft. All authors contributed to the article and approved the submitted version.

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