Afoxolaner against fleas: immediate efficacy and resultant mortality after short exposure on dogs

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Abstract – The speed of efficacy of afoxolaner (NexGard®) against Ctenocephalides felis fleas was evaluated in two studies. Study A assessed the efficacy against existing fleas whereas study B assessed the efficacy against new infesting fleas. In study A, 12 dogs were allocated to the untreated group and 20 dogs to the treated group. All dogs were infested by 100 fleas each at Day −1, treated at Day 0 and flea combed at 2 h or at 6 h post treatment. In study B, 6 dogs were allocated to the untreated group and 10 to the treated group. They were infested with 100 fleas each on Days 2, 7, 14, 21 and 28. Fleas were removed and counted at 6 h post-infestation. Immediate and persistent efficacies were evaluated by counting fleas on the dogs. To evaluate induced mortality after exposure on dogs, fleas collected alive were placed in an insectarium for 24 h and assessed for viability. The immediate efficacy on dogs was significant at 6 h with 100%. The induced death of the fleas collected live from dogs 2 h after exposure was 99.7%. Concerning new infesting fleas, the observed efficacy at 6 h and the induced mortality were significantly different (p < 0.05) from the control at all time-points. At 6 h, the prophylactic efficacy was > 97% at Day 2 and Day 8 and > 90% at Day 14. The induced mortality after 6 h of exposure on dogs varied between 73.3% and 100% for the whole study.

Key words: Ctenocephalides felis, Dogs, Afoxolaner, Short exposure, Immediate killing, Induced mortality.

Introduction

Fleas represent the most prevalent parasites in domestic carnivores [2, 17, 18]. The cat flea, Ctenocephalides felis, is the main flea species infesting both dogs and cats [8, 14]. In addition to causing discomfort to pets and their owners, cat fleas are associated with several diseases. Indeed C. felis is primarily responsible for flea bite allergy dermatitis (FAD) in dogs and cats [5, 7, 19, 20]. The cat flea is also the primary intermediate host of the tapeworm Dipylidium caninum, the common intestinal cestode of dogs and cats [9, 20]. C. felis can also transmit flea-borne spotted fever (Rickettsia felis), and some Bartonella species, such as B. henselae, the agent of Cat Scratch Disease [1, 13, 16]. Fleas are also vectors of some filarioids such as Acanthocheilonema reconditum infesting dogs in many parts of the world [4].

Although the use of insecticides such as fipronil, imidacloprid, selamectin and spinosad have revolutionised flea control

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in recent years, treatment and prevention of cat flea infestations remain a major concern for pet owners and veterinarians [2, 24]. The research continues for new flea control compounds that are effective, fast acting, long lasting, and easy to administer. Licensed products must reach standards of efficacy determined by pharmaceutical regulatory authorities worldwide (European Medicine Agency (EMA) in Europe, Environmental Protection Agency (EPA) or Food and Drug Administration (FDA) in USA). To meet EMA guidelines, the efficacy of a drug against fleas must reach at least 95%, 48 h post infestation for a given time-point in several controlled standard studies conducted [10].

Over the past decade speed of kill on fleas has been recognized as an increasingly important criterion of efficacy [2] prompting the evaluation of flea efficacy after shorter exposures than the ones required by regulatory agencies worldwide. The speed of kill is also an important criterion to estimate the risk reduction for transmission of vector-borne diseases, especially in regard to tick-borne pathogens [2, 17, 18].

Afoxolaner is a new insecticide/acaricide molecule from the isoaxalone family that acts on the insect γ-aminobutyric acid receptor (GABA) and glutamate receptors, inhibiting GABA & glutamate-regulated uptake of chloride ions, resulting in excess neuronal stimulation and death of the arthropod [12, 27]. Afoxolaner is the active ingredient of NexGard®©, a recently approved oral ectoparasiticide product for dogs. NexGard®© administered to dogs has been demonstrated to provide an efficacy higher than 95% within 12 h against adult fleas (Ctenocephalides felis and C. canis) for at least 3 weeks, and within 24 h for at least 5 weeks [15]. The two studies presented here were intended to evaluate the killing activity of afoxolaner against adult C. felis fleas after short exposure times of 2–6 h on dogs, either on existing fleas (immediate or curative efficacy) or on new infesting fleas (prophylactic efficacy).

**Materials and methods**

The two studies were parallel group design, randomised, single centre, controlled efficacy studies. Study A assessed the efficacy of afoxolaner against existing fleas, whereas study B assessed efficacy against new infesting fleas. In total, 48 dogs were included in the two studies, 32 in study A and 16 in study B. All 32 study A dogs and 16 study B dogs were infested with 100 (±5) unfed adult C. felis on Day −7, which were removed and counted on Day 0 for allocation purposes.

In study A, 32 mongrel dogs were randomly allocated to the untreated control group (12 dogs) or to the NexGard®© orally treated group (20 dogs). All dogs were infested by 100 fleas each at Day −1 and were treated at Day 0. Dogs were flea combed at 2 h (6 control and 10 treated dogs) or at 6 h (6 control and 10 treated dogs) post treatment.

In study B, 16 dogs were allocated to the negative untreated control group (n = 6) or to the NexGard®© treated group (n = 10).

The design of the two studies was in accordance with the World Association for the Advancement of Veterinary Parasitology (W.A.A.V.P.) guidelines for evaluating the efficacy of parasiticides for the treatment, prevention and control of flea and tick infestation on dogs and cats” [6]. The studies were authorized by both Merial and ClinVet ethics committees, and were conducted in respect of Good Clinical Practices as described in International Cooperation on Harmonisation of Technical Requirements for Registration of Veterinary Medicinal Products (VICH) guideline GL9 [10].

**Animals**

The dogs were acclimated at the study facility for at least seven days before Day 0; they were clinically healthy as determined by a veterinarian on Day −7; they were older than 6 months, not clinically pregnant, and they had not been treated with a topical or systemic acaricide/insecticide within 12 weeks preceding Day 0. Study animals were kept individually in cages and no contact between dogs was possible. The animals were fed once a day. Food and water were provided in stainless steel bowls and the water was replenished at least twice daily.

At Day 0, dogs in the NexGard®© treated group were dosed once orally with the appropriate soft chew formulations containing afoxolaner, following approved label recommendations. The dogs were weighed on Day −1. As all treated dogs weighed between 10 and 25 kg, they received a 3 g chew containing 68 mg of afoxolaner. The dogs were observed hourly (+15 min) for 4 h after the last animal had been treated for possible adverse events.

**Flea infestations**

A laboratory-bred European strain (ClinVet, Bloemfontein, South Africa) of Ctenocephalides felis was used for all infestations. For the evaluation of efficacy, each dog was infested with 100 (±5) unfed adult C. felis once on Day −1 in study A and on Days 2, 7, 14, 21 and 28 in study B.

**Flea counts and removal**

Flea removal and counts on dogs were performed on Day 0 (study A) at 2 h (for 6 control and 10 treated dogs) and at 6 h (for 6 control and 10 treated dogs) in study A. In study B, flea removal and counts were performed on Days 2, 7, 14, 21 and 28, 6 h after each infestation. For combing, a fine-toothed flea comb was used to recover fleas present in the animal’s fur. The method of combing was by several strokes of the comb in each area of the animal, each time moving in the same direction following the pattern of the hair coat. Movement, from one part of the animal’s fur to the next was via strokes overlapping each other, so that no area of fur was missed. After completion of the combing procedure for all body areas, the whole procedure was repeated once more so that all areas were combed twice. The fleas were categorized in three categories as: live (i.e. normal behaving); moribund (i.e. dying fleas); and dead fleas.

**Flea assessments in insectarium**

At all time-points in both studies, live collected fleas were placed in vials at the time of collection, identified with at least
the dog’s identification, study number and date, and maintained in an insectarium for 24 h. They were held in an environmental chamber at ~75% relative humidity and 24–28 °C (±2 °C). These fleas were evaluated for viability on Day 1 (study A), and Days 3, 8, 15, 22 and 29 (study B).

**Effectiveness criteria**

The primary criterion was the number of live fleas counted on the dog at the time of removal. It allowed the assessment of the killing efficacy observed on dogs, either immediate/curative or persistent/prophylactic.

A secondary criterion was assessment of the viability of live fleas kept in an insectarium for 24 h after collection off dogs. This secondary criterion measures the afoxolaner-induced flea mortality after they have been removed off of dogs, and prevented any additional drug exposure. It allowed the assessment of induced mortality after a very short exposure to afoxolaner, i.e. the mortality rate after a given exposure time on treated dogs compared to control dogs, based on the 100 fleas deposited on each dog at each infestation.

**Statistical analysis**

The observed efficacy on dogs against fleas counted after a short exposure (30 min, 2 h or 6 h) was calculated according to the following formula:

\[
\text{Efficacy (\%)} = 100 \times \frac{(m_c - m_t)}{m_c},
\]

where

- \(m_c\) = geometric or arithmetic mean of live fleas collected from the dogs in the negative control group at each collection time-point;
- \(m_t\) = geometric or arithmetic mean of live fleas collected from the dogs in the administration group at each collection time-point.

The induced death rate was calculated for the administration groups according to the following formula:

\[
\text{Efficacy (\%)} = 100 \times \frac{(m_c - m_t)}{m_c},
\]

where

- \(m_c\) = geometric or arithmetic mean of live fleas in insectarium from the negative control group;
- \(m_t\) = geometric or arithmetic mean of live fleas in insectarium from the treated group.

The number of live fleas observed per dog in the insectarium is related to the original 100 fleas put on each dog. The remaining live fleas after a certain period of time are placed in the insectarium and then assessed at 24 h. The comparison between treated and untreated dogs is linked to the 100 fleas put on all animals in both groups. Therefore, the induced death is a cumulative assessment of the killing efficacy (immediate and delayed) after a certain exposure time.

The untransformed flea count data were compared using a non-parametric Mann-Whitney U test. SAS® Version 9.3 was used for all the statistical analyses. The level of significance of the formal tests was set at 5%, all tests were two sided.

**Results**

In study A, no adverse events were noted.

In study B, one control dog presented a footpad laceration. No adverse event was related to the administration of afoxolaner.

Study A assessed the immediate curative efficacy on existing fleas (Table 1). The curative efficacy against fleas on dogs was significant after just 6 h with 100% effect. The induced flea mortality observed after 24 h in the insectarium was 99.7% for the fleas collected 2 h after exposure, which was significant versus the control (\(p = 0.00138\)).

Study B assessed the prophylactic efficacy after only a short exposure time on dogs against new infesting fleas during the month after treatment.

The persistent efficacy observed on Days 7, 14, 21, 28 after 6 h of exposure on treated dogs varied between 29% and 96.7% based on arithmetic means with the highest percentage of efficacy obtained on Day 7 and the lowest at Day 28 (Table 2). The live flea counts differed significantly (\(p < 0.05\)) between control and treated groups at all assessment time-points.

The mortality observed in the insectarium from fleas collected live after 6 h of exposure on dogs varied between 73.3% and 100% based on geometric means with the highest percentage efficacy obtained on Days 7 and 14. The flea survival differed significantly (\(p < 0.05\)) from the control on all assessments.

**Discussion**

Afoxolaner is a new systemic insecticide and acaricide. It was therefore important to assess its speed of action on fleas. These studies demonstrated that afoxolaner completely killed existing fleas on dogs in 6 h and induced a significant mortality after just 2 h of exposure. It induced the mortality of 97.6 to 100% of new infesting fleas after 6 h of exposure on dogs until Day 21. This property can be related to both the quick absorption and plasma peak [14], but also to the volume of blood ingested by the fleas. Afoxolaner is detected in plasma 20–30 min after oral administration and reaches its peak in 2–4 h. The frequency of flea bites is also a factor explaining the quick intoxication of fleas, which will ingest the insecticide present in the blood [25]. New infesting fleas may bite as soon as 20 min after infesting their host and almost all fleas feed in the first 2 h. The number of flea bites per day is unknown. Fleas can stop their meal when they are disturbed and it is possible that they may bite more often but without ingesting a sufficient volume. The curative efficacy and its speed may then be related to both the number of bites, the volume ingested by fleas, and by the number of fleas that will bite in a certain time on the host. The 5.5% curative efficacy observed at 2 h may be due to a low number of fleas having bitten, but when looking at the induced mortality of 99.7% of these fleas, it is more
The live fleas removed from dogs were put in the insectarium for 24 h. Induced mortality was evaluated as the survival of fleas measured in the insectarium after 24 h.

Day of treatment (Day 0). Time (in hours) after treatment (Day 0).

### Table 1. Immediate efficacy and induced mortality of afoxolaner against fleas after 2 h or 6 h of exposure on dogs.

| Day –1 flea infestation | Control group (n = 6) | Treated group (n = 10) | p-value | Control group (n = 6) | Treated group (n = 10) | p-value |
|-------------------------|-----------------------|------------------------|---------|-----------------------|------------------------|---------|
| Day 0 + 2 h             | Control mean          | Arithmetic mean        | Geometric mean (Eff%) | 0.0005* |
|                         | 75.5 (57–89)          | 74.5                   | 70.5 (5.5)         | >0.05   | 69.5 (47–86)           | 0.6     | 0.2 (99.7)          | 0.00138* |
| Day 0 + 6 h             | Control mean          | Arithmetic mean        | Geometric mean (Eff%) | 0.0005* |
|                         | 67.8 (40–88)          | 0                      | 0 (100)            | 0.0138* | 62.7 (28–86)           | /       | /                   | / |

* The live fleas removed from dogs were put in the insectarium after 24 h. Induced mortality was evaluated as the survival of fleas measured in the insectarium after 24 h.

### Table 2. Observed prophylactic efficacy of afoxolaner and induced mortality of fleas after 6 h of exposure on dogs.

| Day of treatment | Control group (n = 6) | Treated group (n = 10) | p-value |
|------------------|-----------------------|------------------------|---------|
| Day 14           | Arithmetic mean        | Arithmetic mean        | Geometric mean (Eff%) |
|                  | 78.8 (51–93)          | 75.7                   | 71.5 (3.2)         | 0.0007* |
| Day 21           | Arithmetic mean        | Arithmetic mean        | Geometric mean (Eff%) |
|                  | 76.1 (50–90)          | 72.3                   | 67.7 (2.8)         | 0.0006* |
| Day 28           | Arithmetic mean        | Arithmetic mean        | Geometric mean (Eff%) |
|                  | 79.7 (56–93)          | 77.5                   | 71.5 (2.8)         | 0.0006* |

1. Computed as the anti-logarithm of the arithmetic mean of the log-counts and then subtracting 1.
2. (Two-sided) Probability value associated with the comparison of the population means of the two treatment groups.
* Significant difference.

probably related to the volume of drug ingested. After 6 h of exposure to afoxolaner, all existing fleas were dead, which would mean that fleas having ingested more drug, died quicker.

Until now, the majority of anti-flea treatments were based on topical spot-on applications [2, 24], with the active molecules diffusing onto the skin and acting by contact with the arthropods (e.g. dinotefuran, fipronil, imidacloprid) or being absorbed to act systemically (i.e. selamectin). Recently, oral formulations appeared on the market, based on either nitenpyram [21], spinosad [3, 23, 26], afoxolaner [27] or fluralaner [22]. They provide a fast curative efficacy on existing flea infestation. Nitenpyram kills existing fleas on dogs between 15 min and 24 h (3–8 h in some papers) but it has no persistent activity until 24 h (3–8 h in some papers) but it has no persistent activity [21, 26]. Spinosad oral administration provides a curative efficacy, starting to kill fleas within 30 min and killing >95% in about 4 h, combined with up to 4 weeks preventive efficacy on new flea infestations when counted at 48 h after weekly flea infestations [23, 26].

With afoxolaner, the prophylactic efficacy and the induced mortality were significant during the full month after 6 h of exposure, with an induced mortality over 96.7% through Day 21. The decrease in observed efficacy at 6 h on dogs the third and the fourth week may be related to a lower concentration circulating in the blood. However, the dose ingested was still enough to induce the death of the fleas during the month as shown by the mortality rate in the insectarium and by the on dog efficacy studies published at 12 h and 24 h counts [12].

The new oral insecticide-acaricide afoxolaner is able to induce the death of fleas after a short exposure time on treated dogs.

## Conflict of interest

This clinical study was funded by Merial S.A.S., 29 avenue Tony Garnier, 69007 Lyon of which Frédéric Beugnet and Lénaïg Halos are employees. ClinV et, of which Josephus Fourie and Christa deVos are employees, is an independent, South African, Contract Research Organisation contracted to conduct the study. All authors voluntarily publish this article and have no personal interest in these studies other than publishing the scientific findings that they have been involved in via planning, initiating, monitoring and conducting the investigations and analysing the results. This document is provided for scientific purposes only. Any reference to a brand or trademark herein is for informational purposes only and is not intended for a commercial purpose or to dilute the rights of the respective owner(s) of the brand(s) or trademark(s). NexGard® is a registered trademark of Merial. All other marks are the property of their respective owners.

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