A prognostic nomogram based on LASSO Cox regression in patients with alpha-fetoprotein-negative hepatocellular carcinoma following non-surgical therapy

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Abstract

Background: Alpha-fetoprotein-negative hepatocellular carcinoma (AFP-NHCC) (< 8.78 ng/mL) have special clinicopathologic characteristics and prognosis. The aim of this study was to apply a new method to establish and validate a new model for predicting the prognosis of patients with AFP-NHCC.

Methods: A total of 410 AFP-negative patients with clinical diagnosed with HCC following non-surgical therapy as a primary cohort; 148 patients with AFP-NHCC following non-surgical therapy as an independent validation cohort. In primary cohort, independent factors for overall survival (OS) by LASSO Cox regression were all contained into the nomogram1; by Forward Stepwise Cox regression were all contained into the nomogram2. Nomograms performance and discriminative power were assessed with concordance index (C-index) values, area under curve (AUC), Calibration curve and decision curve analyses (DCA). The results were validated in the validation cohort.

Results: The C-index of nomogram1 was 0.708 (95%CI: 0.673–0.743), which was superior to nomogram2 (0.706) and traditional modes (0.606–0.629). The AUC of nomogram1 was 0.736 (95%CI: 0.690–0.778). In the validation cohort, the nomogram1 still gave good discrimination (C-index: 0.752, 95%CI: 0.691–0.813; AUC: 0.784, 95%CI: 0.709–0.847). The calibration curve for probability of OS showed good homogeneity between prediction by nomogram1 and actual observation. DCA demonstrated that nomogram1 was clinically useful. Moreover, patients were divided into three distinct risk groups for OS by the nomogram1: low-risk group, middle-risk group and high-risk group, respectively.

Conclusions: Novel nomogram based on LASSO Cox regression presents more accurate and useful prognostic prediction for patients with AFP-NHCC following non-surgical therapy. This model could help patients with AFP-NHCC following non-surgical therapy facilitate a personalized prognostic evaluation.

Keywords: Alpha-fetoprotein-negative hepatocellular carcinoma, Nomogram, Prognosis, LASSO cox regression, Non-surgical therapy
Background
Hepatocellular carcinoma (HCC) is the sixth-most commonly diagnosed cancer and the fourth leading cause of tumor-related death worldwide in 2018 [1]. Accumulated evidence demonstrates that inefficient diagnosis of HCC is still a major cause of high mortality, especially in patients harboring early or small HCC [2]. Since the identification of alpha-fetoprotein (AFP) in 1970s, it has been the only serologic marker that is widely used for the HCC diagnosis [3]. For decades, HCC screening relied primarily on ultrasound imaging and AFP. Due to technical limitations, ultrasound images are often unrecognizable for HCC nodules less than 1 cm [4]. AFP is the most important and traditional serological diagnostic indicator for HCC, but about 30–40% of overall HCC patients have normal AFP levels (<20 ng/mL). This is referred to as AFP-negative hepatocellular carcinoma (AFP-NHCC) [5]. AFP-NHCC is an important type of liver cancer that currently causes many HCC patients to lose early diagnosis and treatment, especially in patients harboring early or small HCC [2]. Since the identification of alpha-fetoprotein (AFP) in 1970s, it has been the only serologic marker that is widely used for the HCC diagnosis [3]. For decades, HCC screening relied primarily on ultrasound imaging and AFP. Due to technical limitations, ultrasound images are often unrecognizable for HCC nodules less than 1 cm [4]. AFP is the most important and traditional serological diagnostic indicator for HCC, but about 30–40% of overall HCC patients have normal AFP levels (<20 ng/mL). This is referred to as AFP-negative hepatocellular carcinoma (AFP-NHCC) [5]. AFP-NHCC is an important type of liver cancer that currently causes many HCC patients to lose early diagnosis and treatment, especially in HCC with tumors less than 3 cm [6]. Although imaging technology has greatly improved the level of HCC detection, ultrasound images often fail to recognize small HCC nodules or distinguish malignant nodules from benign ones [5,7], and the diagnosis rate for patients with AFP-NHCC is only 10.4% [8]. Studies have shown that patients with AFP-NHCC often have special clinicopathologic characteristics and prognosis, they have higher tumor differentiation, earlier TNM staging, smaller tumor size, and higher survival rates [9]. Therefore, it is essential to identify the independent prognostic risk factors of such patients, and construct prognostic prediction models.

Many staging systems have been employed to predict how HCC patients will respond over time, such as the BCLC, ALBI, Child-Pugh and TNM staging system. However, studies have shown that the traditional staging system is flawed to varying degrees. The BCLC staging system was reported to have the greatest potential in predicting patients with AFP-NHCC [10]. However, evidence has shown that the classification of the BCLC score is limited to the advanced stages of HCC [11]. Child-Pugh does not consider tumor-related factors, which are important for the prognosis of HCC patients [12], as the prognosis of HCC patients, tumor-related factors are crucial. ALBI model incorporates few factors, clinical indicators are easily available and easy to apply, but there are no tumor-related indicators to evaluate [13], as a model to evaluate the prognosis of HCC needs to be validated in a multicenter large sample. The TNM staging system is easy to use and is considered to be the best staging system for solid tumors, but its effect on the staging and prognosis of HCC is debatable because it only considers tumor characteristics but not liver function, which usually plays an important role in the prognosis of HCC patients [14]. Furthermore, these systems are not specifically designed to predict outcomes of patients with AFP-NHCC. Recently, nomograms for prediction of survival and recurrence of patients with AFP-NHCC after radical resection have been developed in two previous studies [15,16], respectively. However, patients with AFP-NHCC with transarterial chemoembolization (TACE) and radiofrequency ablation (RFA) were not included in both studies. At the same time, there are many ways to establish independent risk factors of prognostic prediction model.

LASSO Cox regression is a method for variable selection and shrinkage in Cox proportional hazards model, proposed by Tibshirani et al. in 1997 [17]. LASSO Cox regression analysis constructs a penalty function to obtain a more refined model. And a number of studies have shown that it plays an important role in cancer research: Li et al. applied LASSO Cox regression to the establishment of a prognostic model for lung adenocarcinoma [18]; Xiong et al. establishment of an outcome model for bladder cancer [19]; Jiang et al. establishment of a prognostic model gastric cancer [20]; Wu et al. establishment of a prognostic model pancreatic cancer [21]. Meanwhile, Liu et al. applied LASSO Cox regression to the establishment of a prognostic model for hepatocellular carcinoma [22,23]. However, the above studies all used LASSO Cox screening genes, and there were few reports on the screening of clinical indicators. Therefore, the aim of our study was applying a new method to establish and validate a new model that combines clinical pathological factors, biochemical indicators, for predicting the prognosis of patients with AFP-NHCC. In addition, a comparison between the constructed nomograms and traditional staging systems was conducted to determine whether the nomograms provided more accurate prediction in prognosis.

Methods
Patient selection
We performed a retrospective cohort study on 558 AFP-negative (<8.78 ng/mL) HCC patients following nonsurgical therapy using data from the Beijing Ditan Hospital between January 2008 and December 2016. The HCC diagnosis data included biopsy, radiology. First, we selected patients based on hepatic angiography, pathology in combination with ultrasonography, computed tomography (CT), and magnetic resonance imaging (MRI). Next, we only included patients with complete clinical data. Our exclusion criteria included: (1) other viral infections such as human immunodeficiency virus (HIV); (2) metastatic liver cancer; (3) pregnant women; (4) incomplete data; (5) following surgical therapy; and (6) patients with liver transplantation and survival time<15 days. The primary cohort of our study included 410 clinically diagnosed patients with
AFP-NHCC, retrospectively studied via an information system between January 2008 and December 2014. The patients were followed for 5 years (death follow-up stopped) and first hospitalization records were kept. One hundred forty eight patients with AFP-NHCC between January 2015 and December 2016 as an independent validation cohort. The patients were followed for 3 years (death follow-up stopped) and first hospitalization records were kept. It is recommended that all HCC patients undergo regular follow-up visits according to clinical guidelines after completion of hospital admission, usually every 3 months for the first 2 years and once a year for the next 3 to 5 years. Patients who did not come to our hospital on time for review were given treatment information and living conditions by telephone follow-up (telephone follow-up by our clinicians), the last follow-up occurred in December 2019. The outcome of our study was overall survival (OS), defined as the time from the diagnosis of HCC to the last follow-up or death. The study was approved by the ethics committee of Beijing Ditan Hospital, Capital Medical University and was conducted in accordance with the standards of the Declaration of Helsinki.

Laboratory measurements
Patients are routinely examined at the first visit. Data provided include: sex, age, tumor multiplicity, tumor size, ascites, cirrhosis, etiology, portal vein tumor thrombus (PVTT), neutrophil to lymphocyte ratio (NLR), hemoglobin (HGB), platelet (PLT), creatinine (CR), alanine aminotransferase (ALT), aspartate aminotransferase (AST), total bilirubin (TBIL), albumin (ALB), gamma-glutamyl transpeptidase (y-GGT), prothrombin time activity (PTA), lactate dehydrogenase (LDH), carbohydrate antigen 199 (CA199), C reactive protein (CRP), carcino-embryonic antigen (CEA), AFP. When laboratory values at the time of HCC diagnosis were higher than clinical normal, it was classified as elevated. The tumor was staged using the BCLC staging system and liver function was scored using Child-Pugh.

Statistical analysis
We convert continuous variables into classification variables, which makes the model more objective and simpler. The cut-off value of the classification variable was the normal value of clinical laboratory examination. The cut-off value of numeric values as follows: HGB was 120 g/L, PLT was 100×10^9/L, CR was 111 μmol/L, ALT was 50 U/L, AST was 40 U/L, TBIL was 18.8 μmol/L, ALB was 40 g/L, LDH was 250 U/L, GGT was 60 U/L, PTA was 70%, CEA was 5 ng/ml, CA199 was 37 u/ml, CRP was 5 mg/L. At the same time, it is consistent with the cut-off value of previous literatures published by our team [24]. The cut-off value of NLR was 5 determined according to relevant literature [25, 26]. In this study, we used the primary cohort to plot nomogram1 based on LASSO Cox regression, and used the primary cohort to plot nomogram2 based on Forward Stepwise Cox regression. Based on the established nomogram1 and nomogram2, the C-index and calibration curves were derived based on Cox regression analysis, NRI and IDI scores were performed, and decision curves were plotted. Nomogram1 was found to be superior to nomogram2 in the primary and validation cohorts. The total score for each patient was calculated based on nomogram 1, and three groups of patients with high, medium, and low prognostic risk (based on the total score) were divided according to interquartile values. The Kaplan-Meier curve was applied in MedCalc software, and the risk group was compared using the three-point factor, log-rank test, and the two-tailed P value < 0.05 was statistically significant.

Statistical analysis was done using SPSS 24.0 (IBM, Chicago, IL, USA), R software (version 3.6.3; http://www.Rproject.org) and MedCalc19.2.0. Categorical variables were classified based on clinical findings. SPSS 24.0 was used to perform the Forward Stepwise Cox regression. R version 3.6.3 for “foreign” package, “survival” package, and “rms” package, were used to plot nomograms and calibration plots, and calculate C-index; “glmnet” package was used to perform the LASSO Cox regression; “nricens” package for NRI calculation; “stdca” package for decision curve; “time ROC” package for time-dependent ROC curve. MedCalc19.2.0 for low-risk group, middle-risk group and high-risk group Kaplan-Meier curve plotting.

Results
Basic characteristics
In the primary cohort, 410 patients with AFP-NHCC met the inclusion and exclusion criteria and were included in this study. A total of 148 patients with AFP-NHCC were included in the validation cohort. The clinical characteristics of patients in the two independent cohorts are shown in Table 1. Most of the study population was male (79.8% vs 83.8%; p = 0.286), and the patients were older than 50 years (77.8% vs 83.1%; p = 0.173) in the primary and validation cohort. In addition, (78.5% vs 81.1%; p = 0.876) of the patients presented positive HBsAg. Most patients remained at BCLC stage 0-B (78.5% vs 80.4%; p = 0.632) and Child-Pugh A-B (87.1% vs 92.6%; p = 0.168), and (24.1% vs 18.2%; p = 0.212) had a tumor size ≥5 cm (Table 1). In the primary cohort, 224 patients died from cancer within 5 years.

Biomarker selection
All available clinical indicators, including clinicopathological features and biomarkers (Table 1), were subjected to LASSO Cox regression, with a significant correlation between sex, age, tumor size, tumor number, cirrhosis, PVTT,
Table 1 Basal clinicopathologic characteristics in training and validation cohort

| Characters            | Training set  | Validation set | P  |
|-----------------------|---------------|----------------|----|
|                       | n = 410(%)    | n = 148(%)     |    |
| Sex                   |               |                |    |
| Male                  | 327 (79.8)    | 124 (83.8)     | 0.286 |
| Female                | 83 (20.2)     | 24 (16.2)      |    |
| Age                   |               |                |    |
| <50                   | 91 (22.2)     | 25 (16.9)      | 0.173 |
| ≥50                   | 319 (77.8)    | 123 (83.1)     |    |
| Tumor size(cm)        |               |                |    |
| <3                    | 230 (56.1)    | 84 (56.8)      | 0.212 |
| 3–5                   | 81 (19.8)     | 37 (25.0)      |    |
| ≥5                    | 99 (24.1)     | 27 (18.2)      |    |
| Tumor multiplicity    |               |                |    |
| Single                | 266 (64.9)    | 77 (52.0)      | 0.0006 |
| Multiple              | 144 (35.1)    | 71 (48.0)      |    |
| Cirrhosis             |               |                |    |
| Yes                   | 377 (92.0)    | 135 (91.2)     | 0.780 |
| No                    | 33 (8.0)      | 13 (8.8)       |    |
| PVTT                  |               |                |    |
| Yes                   | 145 (35.4)    | 40 (27.0)      | 0.065 |
| No                    | 265 (64.6)    | 108 (73.0)     |    |
| Ascites               |               |                |    |
| Yes                   | 162 (39.5)    | 53 (35.8)      | 0.428 |
| No                    | 248 (60.5)    | 95 (64.2)      |    |
| HBsAg                 |               |                |    |
| Positive              | 330 (78.5)    | 120 (81.1)     | 0.876 |
| Negative              | 80 (21.5)     | 28 (18.9)      |    |
| NLR                   |               |                |    |
| <5                    | 344 (83.9)    | 132 (89.2)     | 0.119 |
| ≥5                    | 66 (16.1)     | 16 (10.8)      |    |
| HGB(g/L)              |               |                |    |
| <120                  | 159 (38.8)    | 58 (39.2)      | 0.930 |
| ≥120                  | 251 (61.2)    | 90 (60.8)      |    |
| PLT(*10^9/L)          |               |                |    |
| <100                  | 219 (53.4)    | 80 (54.1)      | 0.894 |
| ≥100                  | 191 (46.6)    | 68 (45.9)      |    |
| CR (μmol/L)           |               |                |    |
| <111                  | 391 (95.4)    | 141 (95.3)     | 0.962 |
| ≥111                  | 19 (4.6)      | 7 (4.7)        |    |
| ALT(U/L)              |               |                |    |
| <50                   | 347 (84.6)    | 130 (87.8)     | 0.343 |
| ≥50                   | 63 (15.4)     | 18 (12.2)      |    |
| AST(U/L)              |               |                |    |
| <40                   | 268 (65.4)    | 113 (76.4)     | 0.014 |
| ≥40                   | 142 (34.6)    | 35 (23.6)      |    |
| Characters | Training set n = 410(%) | Validation set n = 148(%) | P    |
|------------|------------------------|-------------------------|------|
| TBIL (μmol/L) |                         |                         |      |
| <18.8      | 218 (53.2)             | 87 (58.8)               | 0.240|
| ≥18.8      | 192 (46.8)             | 61 (41.2)               |      |
| ALB (g/L)  |                         |                         |      |
| <40        | 275 (67.1)             | 103 (69.6)              | 0.574|
| ≥40        | 135 (32.9)             | 45 (30.4)               |      |
| LDH (U/L)  |                         |                         |      |
| <250       | 365 (89.0)             | 131 (88.5)              | 0.865|
| ≥250       | 45 (11.0)              | 17 (11.5)               |      |
| GGT (U/L)  |                         |                         |      |
| <60        | 273 (66.1)             | 99 (66.9)               | 0.861|
| ≥60        | 139 (33.9)             | 49 (33.1)               |      |
| PTA (%)    |                         |                         |      |
| <70        | 142 (34.6)             | 42 (28.4)               | 0.165|
| ≥70        | 268 (65.4)             | 106 (71.6)              |      |
| CEA (ng/ml) |                         |                         |      |
| <5         | 345 (84.1)             | 113 (76.4)              | 0.034|
| ≥5         | 65 (15.9)              | 35 (23.6)               |      |
| CA199 (u/ml)|                         |                         |      |
| <37        | 364 (89.3)             | 133 (90.5)              | 0.664|
| ≥37        | 44 (11.7)              | 14 (9.5)                |      |
| CRP (mg/L) |                         |                         |      |
| <5         | 266 (64.9)             | 99 (71.2)               | 0.180|
| ≥5         | 144 (35.1)             | 40 (28.8)               |      |
| Child-Pugh |                         |                         |      |
| A          | 230 (56.1)             | 92 (62.2)               | 0.168|
| B          | 127 (31.0)             | 45 (30.4)               |      |
| C          | 53 (12.9)              | 11 (7.4)                |      |
| BCLC stage |                         |                         |      |
| 0-B        | 322 (78.5)             | 119 (80.4)              | 0.632|
| C-D        | 88 (21.5)              | 29 (19.6)               |      |
| ALBI grade |                         |                         |      |
| I          | 149 (36.3)             | 76 (51.4)               | 0.004|
| II         | 207 (50.5)             | 59 (39.9)               |      |
| III        | 56 (13.7)              | 13 (8.8)                |      |
| TNM        |                         |                         |      |
| I          | 159 (38.8)             | 53 (35.8)               | 0.005|
| II         | 66 (16.1)              | 45 (28.4)               |      |
| III        | 131 (32.0)             | 43 (29.1)               |      |
| IV         | 54 (13.2)              | 10 (6.8)                |      |
ascites, HBV, HGB, CR, AST, ALB, LDH, γ-GGT, CA199, CRP and OS at minimum values (Fig. 1a). Further disciplinary regression was performed to take 1-s.e. criteria PVTT, ascites, HGB, γ-GGT, CRP as independent risk factors for prognosis in patients with AFP-NHCC (Fig. 1b). Inclusion of all clinical indicators in Cox univariate analysis, there was a significant correlation with OS in sex, tumor size, tumor multiplicity, cirrhosis, PVTT, ascites, HBsAg, NLR, HGB, PLT, CR, AST, TBIL, ALB, LDH, γ-GGT, PTA, CA199, and CRP. Cox multivariate analysis was then performed to identify the factors that were distinguished in the Cox univariate analysis. The results showed that sex, PVTT, CR, γ-GGT, and CRP were independent risk factors for prognosis in patients with AFP-NHCC (Table 2).

Development the prediction model
Nomogram1 and nomogram2 were constructed to predict 3 and 5-year OS based on prognostic factors determined by both instruments (Fig. 2). Nomogram1 and nomogram2 used consistency index (C-index), AUC and time-dependent ROC curves in the primary cohort, respectively, and the calibration curves were plotted.
The C-index of nomogram1 in the primary cohort was 0.708 (95% CI: 0.673–0.743); the AUC (ROC curve) was 0.736 (95%CI: 0.690–0.778), with sensitivity (62.05%), specificity (76.34%), PPV (76.0%) and NPV (62.6%). The C-index of nomogram2 was 0.706 (95%CI: 0.673–0.739); the AUC was 0.714 (95%CI: 0.667–0.757), with sensitivity (74.55%), specificity (61.83%), PPV (70.2%) and NPV (66.9%). Continuity cut point 0.05NRI (−0.037), subtype

| Characteristic            | Univariate analysis |          | Multivariate analysis |          |
|---------------------------|---------------------|----------|-----------------------|----------|
|                           | HR (95% CI)         | P        | HR (95% CI)           | p        |
| Sex                       | Male/Female         | 1.577 (1.163–2.138) | 0.003 | 1.655 (1.191–2.300) | 0.003 |
| Tumor size(cm)            | <3/3–5/5≥5          | 1.274 (1.093–1.484) | 0.002 |          |          |
| Tumor multiplicity        | Single/Multiple     | 1.514 (1.160–1.976) | 0.002 |          |          |
| Cirrhosis                 | Yes/N0              | 2.265 (1.201–4.271) | 0.012 |          |          |
| PVTT                      | Yes/N0              | 2.052 (1.497–2.811) | <0.001 | 1.547 (1.162–2.059) | 0.003 |
| Ascites                   | Yes/N0              | 2.314 (1.779–3.010) | <0.001 | 1.304 (0.918–1.852) | 0.138 |
| HBsAg                     | Positive/Negative    | 0.622 (0.459–0.844) | 0.002 |          |          |
| NLR                       | <5, ≥5              | 1.723 (1.246–2.384) | 0.001 |          |          |
| HGB(g/L)                  | <120, ≥120          | 0.447 (0.343–0.581) | <0.001 | 0.785 (0.568–1.084) | 0.142 |
| PLT(*10^9/L)              | <100,≥100           | 0.690 (0.503–0.948) | 0.022 |          |          |
| CR (μmol/L)               | <111, ≥111          | 3.486 (2.116–5.745) | <0.001 | 2.200 (1.254–3.858) | 0.006 |
| AST(U/L)                  | <40, ≥40            | 1.908 (1.463–2.489) | <0.001 |          |          |
| TBLI (μmol/L)             | <18.8, ≥18.8        | 1.602 (1.232–2.084) | <0.001 |          |          |
| ALB(g/L)                  | <40, ≥40            | 0.468 (0.342–0.639) | <0.001 |          |          |
| LDH(U/L)                  | <250, ≥250          | 1.993 (1.389–2.861) | <0.001 |          |          |
| GGT (U/L)                 | <60, ≥60            | 2.147 (1.647–2.799) | <0.001 | 1.495 (1.087–2.056) | 0.013 |
| PTA(%)                    | <70, ≥70            | 0.574 (0.440–0.748) | <0.001 |          |          |
| CA1999(u/ml)              | <37, ≥37            | 2.479 (1.738–3.534) | <0.001 |          |          |
| CRP (mg/L)                | <5, ≥5              | 2.545 (1.955–3.313) | <0.001 | 1.823 (1.341–2.478) | <0.001 |

Table 2: Univariate and multivariate cox hazards analysis of the training cohort
cut point 0.5NRI (−0.037), IDI: 0.006 (p = 0.29). We get similar results in the validation cohort. All suggest that nomogram1 is better than nomogram2.

**Performance of the nomogram**

In the primary cohort, the C-index (0.708) and AUC (0.736) of nomogram1 outperformed the nomogram2 and the other models (Table 3). Time-dependent ROC curves suggest that nomogram 1 and nomogram 2 are similar, but significantly better than traditional modes such as Child-Pugh, BCLC, ALBI and TNM staging systems (Fig. 3b). Calibration plots for 3-year OS probabilities show the best agreement between nomogram1 predictions and actual observations (Fig. 4b, d).

The decision curve analysis of nomogram 1 and nomogram 2 is also meaningful (Fig. 5). In the primary cohort the cue differences were small, but in the validation cohort nomogram1 and nomogram2 were better at predicting OS than either the all-patient death scenario or the no-patient death scenario if the patient threshold probability was >

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**Table 2**

| Points | 0 | 10 | 20 | 30 | 40 | 50 | 60 | 70 | 80 | 90 | 100 |
|--------|---|----|----|----|----|----|----|----|----|----|-----|
| PVTT   |   |    |    |    |    |    |    |    |    |    |     |
| Ascites| no|    |    |    |    |    |    |    |    |    |     |
| GGT    | less than 50|    |    |    |    |    |    |    |    |    |     |
| HGB    | more than 120|    |    |    |    |    |    |    |    |    |     |
| CRP    | less than 5    |    |    |    |    |    |    |    |    |    |     |
| Total Points | 0 | 30 | 90 | 150| 210| 270| 330| 390| 450|    |     |

**Fig. 2**

- **a** Nomogram1 including PVTT, Ascites, γ-GGT, HGB and CRP, for three- and five-years overall survival (OS) in patients with AFP-negative HCC.
- **b** Nomogram2 including Sex, PVTT, CR, γ-GGT and CRP, for three- and five-years overall survival (OS) in patients with AFP-negative HCC. The nomogram1 and nomogram2 are valued to obtain the probability of three- and five-years survival by adding up the points identified on the points scale for each variable.
20%. Moreover, the net benefit is comparable; in this range, the predicted OS of nomogram 1 is more advantageous than nomogram 2.

**Application of the Nomogram model for risk stratification**

Based on the nomogram1 we developed in this study, we subdivided the patients into low-risk, middle-risk, and high-risk groups, and the patients with AFP-NHCC showed good prognostic classification in both the primary cohort and the validation cohort. In the primary cohort, there were 122 cases in the low-risk group, 175 cases in the middle-risk group, and 113 cases in the high-risk group. Intergroup OS was (52.111 ± 1.386) months, (40.960 ± 1.622) months, and (26.219 ± 2.138) months ($p < 0.001$) (Fig. 6a). In the validation cohort, there were 50 cases in the low-risk group, 70 cases in the middle-risk group, and 113 cases in the high-risk group. Intergroup OS was (35.121 ± 0.777) months, (25.983 ± 1.670) months, and (18.721 ± 2.685) months ($p < 0.001$) (Fig. 6b).

**Discussion**

It is important to have an accurate prognosis of the tumor after relevant treatment. In this study, we identified 3 years and 5 years of independent risk factors for OS in patients with AFP-NHCC in the primary cohort, and established novel, effective, and verified nomogram1 to predict individuals at 3 years and 5 years OS. PVTT, ascites, HGB, γ-GGT and CRP have been included in the nomogram1. In addition, nomogram1 shows a higher discriminatory power, and the C-index of the two independent cohorts are 0.708 and 0.752, respectively. In the subsequent study, the calibration curve and decision curve analysis were used to assess the predicted precision and clinical utility of the nomogram1. Our model showed a better net benefit and higher consistency between the nomogram prediction and the actual observation.

Previous studies have demonstrated the prominent role of tumor burden and grade, liver function, degree of hepatic dysfunction, and performance status in the prognosis of HCC. For many years, the traditional Child-Pugh rating system is the most widely used method for assessing liver function and predicting therapeutic efficacy [27]. Our study also found that the C-index (0.629) and AUC (0.672) of Child-Pugh in the primary cohort were significantly higher than ALBI, TNM, and BCLC. We also obtained the same results in the validation cohort. However, patients with AFP-NHCC have special clinicopathologic characteristics and prognosis, they have higher tumor differentiation, earlier TNM staging, smaller tumor size, and higher survival rates [9]. Most models are built without taking into account the specificity of liver biology function in patients with AFP-NHCC and do not fully reflect an accurate prognosis. The above studies indicate that our model is more accurate for the prognosis of patients with AFP-NHCC. We found C-index (0.708), AUC (0.736) and time-dependent ROC curves of our model was better than the other models, such as Child-Pugh, BCLC, ALBI, TNM. The calibration curve for probability of OS showed good homogeneity between prediction by our model and actual observation. DCA demonstrated that our model was clinically useful. We get similar results in the validation cohort.

A variety of nomograms have been developed to predict the prognosis of certain cancers and have been shown to be more accurate than traditional staging systems. However, most of them are limited to Forward Stepwise Cox regression risk factor screening, which is not conducive to small sample size, multi-indicator model screening [28]. LASSO Cox regression analysis constructs a penalty function to obtain a more refined model. It compresses the regression coefficients (the sum of the absolute values of the mandatory coefficients is less than a fixed value) and sets some regression coefficients to 0. LASSO Cox regression analysis can not only solve the problem of over-fitting, but also extract useful features effectively [17]. Meanwhile, LASSO Cox regression is more applicable in decisions with more clinical indicators. In this study, we find that the model variables were screened by LASSO Cox regression has better accuracy and resolution than the model variables were screened by Forward Stepwise Cox regression.

### Table 3 C-index and AUC of prognostic staging systems for Training and Validation cohort

| Models  | Training cohort | Validation cohort |
|---------|----------------|-------------------|
|         | C-index 95% CI | AUC 95% CI        | C-index 95% CI | AUC 95% CI        |
| Nomogram1 | 0.708 0.673–0.743 | 0.736 0.690–0.778 | 0.752 0.691–0.813 | 0.784 0.709–0.847 |
| Nomogram2 | 0.706 0.673–0.739 | 0.714 0.667–0.757 | 0.714 0.647–0.781 | 0.677 0.595–0.751 |
| ALBI     | 0.616 0.583–0.649 | 0.659 0.611–0.705 | 0.664 0.593–0.735 | 0.691 0.610–0.764 |
| TNM      | 0.606 0.573–0.639 | 0.656 0.607–0.702 | 0.639 0.578–0.720 | 0.669 0.587–0.744 |
| BCLC     | 0.609 0.582–0.636 | 0.628 0.579–0.674 | 0.668 0.547–0.669 | 0.631 0.547–0.708 |
| CHILD    | 0.629 0.598–0.660 | 0.672 0.625–0.718 | 0.677 0.612–0.742 | 0.722 0.643–0.793 |
Number of studies have shown that it plays an important role in cancer research: especially in lung adenocarcinoma [18]; bladder cancer [19]; gastric cancer [20]; and pancreatic cancer [21]. Meanwhile, Liu et al. applied LASSO Cox regression to the establishment of a prognostic model for hepatocellular carcinoma [22, 23]. Although, the above studies all used LASSO Cox screening genes, and there were few reports on the screening of clinical indicators. In the primary cohort, we find the C-index (0.708) and AUC (0.736) of nomogram1 is superior to the C-index (0.706) and AUC (0.714) of nomogram2. We got better results in the validation cohort.

Our final nomogram included five independent risk factors, liver function parameters: ascites (C-index: 0.606) and γ-GGT (C-index: 0.596); inflammatory indicator: CRP (C-index: 0.596); and a tumor-related index: HGB (C-index: 0.604) and PVTT (C-index: 0.582). The importance of various risk factors can be shown by the Decision Tree (Fig. 7). CRP is an exquisitely sensitive marker of inflammation and tissue damage [29]. CRP is
synthesized in the liver and is secreted into the plasma as a pentamer, belonging to the family of pentraxins, together with serum amyloid protein [30]. Based on recent studies, the serum CRP levels are correlated with the poor prognosis in HCC: Chun et al. found that CRP can predict overall survival and recurrence rates after hepatectomy in patients with HCC patients [31]; Na et al. found that CRP was independently associated with OS in non-surgical HCC patients [32]; and She et al. found that CRP is a biomarker of AFP-negative HBV-related hepatocellular carcinoma [5]. While the molecular mechanism underlying tumor-related CRP elevation in HCC or other cancers remains unknown, several possible mechanisms have been proposed. For instance, cancer growth and tumor-host cell interaction could increase CRP levels [33]. Additionally, CRP levels might reflect an inflammatory response activated as a secondary process in reaction to tumor necrosis or other local tissue damage. Moreover, cancer cells produce cytokines via autocrine pathways, such as IL-6 and IL-8, which in turn induce CRP production [34]. The significance of inflammatory signaling through the STAT3 pathway has been emphasized by numerous studies of HCC and other malignancies. PVTT is the most common form of macrovascular invasion of HCC. Multiple case series have suggested that the PVTT is a common phenomenon with a prevalence rate ranging from 10% to over 60% [35–38]. The ascites, jaundice, hepatic encephalopathy, and liver failure may induce by the formation of PVTT [39]. The presence of PVTT in patients with HCC has been consistently demonstrated by different series to be associated with poor prognoses, with a hazard ratio of death close to 2 [38, 40]. Clinically, PVTT is associated with large tumor size, increased tumor number, higher tumor grade, worse Child-Pugh class and higher serum alpha-fetoprotein (AFP). In this study, PVTT was found to be a strong prognostic indicator in patients with AFP-NHCC. It is to be verified with a large multi-center sample. As a crucial enzyme in glutathione metabolism, γ-GGT was continually elevated in metabolic-induced hepatic injury [41]. Salvatore et al. demonstrated that the level of serum γ-GGT elevated with the process of liver carcinogenesis and promoted tumor progression in an HCC animal model of male.

Fig. 4 Calibration curve of the nomogram1 and nomogram2 in the primary and validation cohort, with the x-axes are actual survival estimated by the nomogram, the y-axes are observed survival calculated by the Kaplan-Meier method. a Five-year survival OS in the primary cohort-nomogram1. b Three-year OS in the validation cohort-nomogram1. c Five-year survival OS in the primary cohort-nomogram2. d Three-year OS in the validation cohort-nomogram2.
Wistar rats [42]. Serum levels of γ-GGT could also help with the selection of further treatment and clinical outcomes for patients with HCC [43]. Carr et al. found that patients with significantly high GGT values were prone to poor overall survival in cases of low AFP HCC [44]. The results of this study and previous studies indicate that the increase of γ-GGT is closely related to the prognosis of patients with AFP-NHCC [15, 45]. Anemia is common in cancer patients. HGB levels have been shown to have an impact on survival both before and during anti-cancer therapy [46]. The pre-treatment anemia in HCC patients was found to be 7.0%, which is less than the 12.8% of cancer-related anemia [47]. The causes of anemia in HCC patients include nutritional deficiencies, hemolysis, blood loss, and tumor cell infiltration of the bone marrow [48]. In addition, chronic liver injury can also lead to anemia in HCC patients [49]. It has also been shown that downregulation of iron-regulated genes, including heparin, ceruloplasmin, transferrin, and transferrin receptors, disrupts the systemic
iron homeostasis, leading to anemia in patients with HCC [50]. It has been demonstrated that HGB is an independent prognostic factor in patients with HCC, which is similar to the results we obtained [47]. Ascites is the hallmark of portal hypertension [51]. Besides, ascites may also be associated with large tumor burden and vascular invasion of HCC, suggesting that the cause of ascites could be attributed to both tumoral and cirrhotic factors [52, 53]. Hence, ascites is not only an indicator of deterioration of liver functional reserve but could also be a signal of tumor progression. Studies have shown that ascites reduces long-term survival in patients with liver cancer.

Fig. 6 Kaplan-Meier survival curves of nomogram1. a In the primary cohort. b In the validation cohort.
cancer [54]. It is similar to the results of our current study. Tumor size is one of the most important parameters of tumor burden. However, evaluation of tumor number failed to predict the prognosis of HCC patients in the present study, although other studies have demonstrated its prominent role in prognostic prediction [55]. Meanwhile, tumor markers have not been included in this study. After research and repeated comparison, our nomogram exhibited superior discrimination ability for the prediction of prognostic in patients with AFP-NHCC. Hence, our novel nomogram could be used to guide routine follow-up for patients. PVTT, ascites, HGB, γ-GGT and CRP should be given great importance in patients with AFP-NHCC. In addition, patients given a high score by the nomogram should undergo more high-end imaging examinations, such as MRI or CT exams, and the interval time of follow up should be reduced, even if the last test results have no causes for concern.

Although our nomogram performed well, several limitations need to be addressed. Firstly, patients with AFP-NHCC were limited and all came from the same hospital, and not validated internally, which may create bias in evaluating the predictive value of these markers; in the validation cohort the follow-up time was shorter, and close monitoring and five-year follow-up data are still required for patients in the validation cohort. Secondly, because the present study was a retrospective study for predicting anticipated future performance, our results need to be confirmed by prospective cohort studies. Thirdly, all patients we included are following non-surgical therapy, whether our nomogram can applicable the patients who following radical resection remains uncertain. Hence, future prospective studies require multiple centers, a larger scale, and more detailed information to validate these results.

Conclusions
We applied new methods to develop and validate nomogram to predict the OS in patients with AFP-NHCC following non-surgical therapy. Nomogram1 presented in this study is statistically easier than previous model screening methods and more accurate than ALBI, TNM, Child-Pugh, BCLC, and provides a useful tool for prognosis. The combination of ascites, γ-GGT, CRP, HGB, and PVTT as economic, simple, effective, and promising biomarkers, possessed a high diagnostic efficiency in the progression of patients with AFP-NHCC, especially in patients following non-surgical therapy.
Abbreviations
HCC: Hepatocellular carcinoma; AFP-NHC: Alpha-fetoprotein-negative hepatocellular carcinoma; OS: Overall survival; HR: Hazard ratio; CI: 95% confidence interval; AFP: Alpha fetoprotein; PVT: Portal vein tumor thrombus; ALB: Albumin; ALP: Alkaline phosphatase; ALT: Alanine aminotransferase; AST: Aspartate aminotransferase; TBL: Total bilirubin; LDH: Lactate dehydrogenase; γ-GGT: γ-Glutamyl transpeptidase; NLR: Neutrophil to lymphocyte ratio; HGB: Hemoglobin; PLT: Absolute platelet count; PTA: Prothrombin activity; RFA: Radiofrequency ablation; TACE: Transcatheter arterial chemoembolization; TNM: The Tumor, node, metastasis; CRP: C-reactive protein; CEAI: Carcino-embryonic antigen; CA199: Carbohydrate antigen 199; BCLC: Barcelona Clinic Liver Cancer

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Authors’ contributions
ZYY and XLL designed the study; DZJ collected and analyzed the data and wrote the manuscript; XHW, FNY, PW and HWY provided patients data; YYJ was responsible for the interpretation of data and revision. All authors participated in interpretation of the findings. ZYY revised the manuscript, and all authors have read and approved the final version of the manuscript.

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Availability of data and materials
The datasets used or analyzed during the current study are available from the corresponding author on reasonable request. In order to protect study participant privacy, our data cannot be shared openly.

Ethics approval and consent to participate
This study is in accordance with the Declaration of Helsinki and has been approved by the ethical committee of Beijing Ditan Hospital, Capital Medical University. The original data used in the study were managed by Beijing Ditan Hospital, Capital Medical University. The study is retrospective and only clinical information of patients will be collected without interfering with patients’ treatment plans or posing physiological risks to patients. The investigators will make every effort to protect the information provided by the patients from disclosure of personal privacy. Informed consent is not required for retrospective studies for the following main reasons: (1) we only collect clinical laboratory information from patients, and the risks are manageable; (2) some patients have lost contact or died and are unable to track their signed informed consent forms.

Consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

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References
1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2018;68(6):394–424.
2. Tang A, Hallouch O, Chernyak V, Kamaya A, Srinl CB. Epidemiology of hepatocellular carcinoma: target population for surveillance and diagnosis. Abdomin Radiol (New York). 2018;43(1):13–25.
3. Johnson PJ. Role of alpha-fetoprotein in the diagnosis and management of hepatocellular carcinoma. J Gastroenterol Hepatol. 1999;14(Suppl)S32–6.
4. Monfardini L, Ors F, Caserta R, Salleri C, Deila Vgna P, Bonomo G, et al. Ultrasound and cone beam CT fusion for liver ablation: technical note. Int J Hyperthermia. 2018;35(1):500–4.
5. She S, Xiang Y, Yang M, Ding X, Liu X, Ma L, et al. Creative protein is a biomarker of AFP-negative HBV-related hepatocellular carcinoma. Int J Oncol. 2015;47(2):543–54.
6. Wang M, Devarajan K,Singal AG, Marrero JA, Dai J, Feng Z, et al. The Doylestown algorithm: a test to improve the performance of AFP in the detection of hepatocellular carcinoma. Cancer Prev Res (Phila). 2016;9(2):172–9.
7. Tsuchiya N, Savada Y, Endo I, Saito K, Uemura Y, Nakatsuma T. Biomarkers for the early diagnosis of hepatocellular carcinoma. World J Gastroenterol. 2015;21(37):10573–83.
8. Chen ZZ, Huang L, Wu YH, Zhai WJ, Zhu PP, Gao YF. LncSox4 promotes the self-renewal of liver tumour-initiating cells through Stat3-mediated Sox4 expression. Nat Commun. 2016;7:12598.
9. Bai DS, Zhang C, Chen P, Jin SJ, Jiang GQ. The prognostic correlation of AFP level at diagnosis with pathological grade, progression, and survival of patients with hepatocellular carcinoma. Sci Rep. 2017;7(1):12870.
10. Zhang XF, Qi X, Meng B, Liu C, Yu L, Wang B, et al. Prognosis evaluation in alpha-fetoprotein negative hepatocellular carcinoma after hepatectomy: comparison of five staging systems. Eur J Surg Oncol. 2010;36(8):718–24.
11. Gómez-Rodríguez R, Romero-Gutiérrez M, Antuza-Vanasa T, González-Frutos C, Giampi D, la Cruz Pérez G, et al. The value of the Barcelona clinic liver cancer and alpha-fetoprotein in the prognosis of hepatocellular carcinoma. Revista Española de Enfermedades Digestivas. 2012;104(6):298–304.
12. Na SK, Yim SY, Suh SJ, Jung YK, Kim JH, Seo SY, et al. ALBI versus Child-Pugh grading systems for liver function in patients with hepatocellular carcinoma. J Surg Oncol. 2018;117(5):912–21.
13. Hu SY, Hisu CY, Liu PH, Hsia CY, Su CW, Huang YH, et al. Albumin-bilirubin (ALBI) grade-based nomogram to predict tumor recurrence in patients with hepatocellular carcinoma. Eur J Surg Oncol. 2019;45(5):776–81.
14. Zhang Y, Chen SW, Liu LL, Yang X, Cai SH, Yun JP. A model combining TNM stage and tumor size shows utility in predicting recurrence among patients with hepatocellular carcinoma after resection. Cancer Manag Res. 2018;10:3707–15.
15. Gan W, Huang JL, Zhang MX, Fu YP, Yi Y, Jing CY, et al. New nomogram predicts the recurrence of hepatocellular carcinoma in patients with negative preoperative serum AFP subjected to curative resection. J Surg Oncol. 2018;117(7):1540–7.
16. Wang X, Mao M, He Z, Zhang L, Li H, Lin J, et al. Development and validation of a prognostic Nomogram in AFP-negative hepatocellular carcinoma. Int J Biol Sci. 2019;15(22):1–8.
17. Tibshirani R. The lasso method for variable selection in the cox model. Stat Med. 1996;15(4):385–95.
18. Li Y, Ge D, Gu J, Xu F, Zhu Q, Lu C. A large cohort study identifying a novel prognosis prediction model for lung adenocarcinoma through machine learning strategies. BMC Cancer. 2019;19(1):886.
19. Xiong Y, Yuan L, Xiong J, Xu H, Luo Y, Wang G, et al. An outcome model for human bladder cancer: a comprehensive study based on weighted gene co-expression network analysis. J Cell Mol Med. 2020;24(3):2342–55.
20. Yang J, Zhang Q, Hu Y, Li T, Yu J, Zhao L, et al. ImmunoScore signature: a prognostic and predictive tool in gastric cancer. Ann Surg. 2018;267(3):504–13.
21. Wu M, Li X, Zhang T, Liu Z, Zhao Y. Identification of a nine-gene signature and establishment of a prognostic Nomogram predicting overall survival of pancreatic cancer. Front Oncol. 2019;9:996.
22. Liu GM, Xie WX, Zhang CY, Xu JW. Identification of a four-gene metabolic signature predicting overall survival for hepatocellular carcinoma. J Cell Physiol. 2020;235(2):1624–36.
23. Liu GM, Zeng HD, Zhang CY, Xu JW. Identification of a six-gene signature predicting overall survival for hepatocellular carcinoma. Cancer Cell Int. 2019;19:138.
24. Yu L, Liu X, Wang X, Dang Z, Jiang Y, Wang X, et al. Impact of gender as a prognostic factor in HBV-related hepatocellular carcinoma: the survival strength of female patients in BCLC stage 0-8. J Cancer. 2019;10(18):4237–44.
25. Chan AW, Chan SL, Wong GL, Wong WW, Chong CC, Lai PB, et al. Prognostic nutritional index (PNI) predicts tumor recurrence of very early/early stage
hepatocellular carcinoma after surgical resection. Ann Surg Oncol. 2015; 22(13):4138–48.

26. Halazun KJ, Najjar M, Abdelmessih RM, Samstein B, Giesemer AD, Guarner JV, et al. Recurrence after liver transplantation for hepatocellular carcinoma: a new MORAL to the story. Ann Surg. 2017;265(3):557–64.

27. Bruix J, Sherman M. Management of hepatocellular carcinoma: an update. Hepatology (Baltimore, Md). 2011;53(3):1002–20.

28. Iasonos A, Schrag D, Raj GV, Panageas KS. How to build and interpret a nomogram for cancer prognosis. J Clin Oncol. 2008;26(8):1364–70.

29. Chen JM, Kwon HJ, Sohn J, Kim SG, Park JY, Bae HJ, et al. Prognostic factors after early recurrence in patients who underwent curative resection for hepatocellular carcinoma. J Surg Oncol. 2011;103(2):148–51.

30. Na GH, Kim DG, Han JH, Kim EY, Lee SH, Hong TH, et al. Inflammatory markers as selection criteria of hepatocellular carcinoma in living-donor liver transplantation. World J Gastroenterol. 2014;20(2):659–601.

31. Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. Nature. 2008;454(7203):436–44.

32. Bruix J, Sherman M. Management of hepatocellular carcinoma: an update. Hepatology (Baltimore, Md). 2011;53(3):1002–20.

33. Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. Nature. 2008;454(7203):436–44.

34. Heikkilä K, Ebrahim S, Lawlor DA. A systematic review of the association between circulating concentrations of C reactive protein and cancer. J Epidemiol Community Health. 2007;61(9):824–33.

35. Quirk M, Kim YH, Saab S, Lee EW. Management of hepatocellular carcinoma with portal vein thrombosis. World J Gastroenterol. 2015;21(12):3462–71.

36. Kim JH, Lee JM, Ryu KS, Park YG, Hur SY, et al. The prognostic impact of duration of anemia during chemotherapy in advanced epithelial ovarian cancer. Oncologist. 2011;16(8):1154–61.

37. Qiu MZ, Xu RH, Ruan DY, Li ZH, Luo HY, Teng KY, et al. Incidence of anemia, leukocytosis, and thrombocytosis in patients with solid tumors in China. Tumour Biol. 2010;31(6):633–41.

38. Aapro M, Østerborg A, Gascón P, Ludwig H, Beguin Y. Prevalence and management of cancer-related anaemia, iron deficiency and the specific role of iron. Ann Oncol. 2012;23(8):1954–62.

39. Minagawa M, Makuuchi M. Treatment of hepatocellular carcinoma. Drug Discoveries Therapeutics. 2014;8(3):134–8.

40. Arcone R, Gualandi G, Ciliberto G. Identification of sequences responsible for acute-phase induction of human C-reactive protein. Nucleic Acids Res. 1988;16(8):3195–207.

41. Chan SL, Mo FK, Johnson PJ, Liem GS, Chan TC, Poon MC, et al. Prospective validation of the Chinese University prognostic index and comparison with other staging systems for hepatocellular carcinoma in an Asian population. J Gastroenterol Hepatol. 2011;26(2):340–6.

42. Bruix J, Sherman M. Management of hepatocellular carcinoma: an update. Hepatology (Baltimore, Md). 2011;53(3):1002–20.

43. Wang Z, Song P, Xia J, Inagaki Y, Tang W, Kokudo N. Can gamma-glutamyl transferase levels, insulin resistance and liver fibrosis in patients with chronic liver diseases. PLoS One. 2012;7(12)e51165.

44. Hsu CY, Lee YH, Huang YH, Hsia CY, Su CW, Lin HC, et al. Ascites in patients with hepatocellular carcinoma: prevalence, associated factors, prognostic impact, and staging strategy. Hepatol Int. 2013;7(1):188–98.

45. Hsu CY, Lee YH, Huang YH, Hsia CY, Su CW, Lin HC, et al. Ascites in patients with hepatocellular carcinoma: prevalence, associated factors, prognostic impact, and staging strategy. Hepatol Int. 2013;7(1):188–98.

46. Spacek LA, Solga SF. Comment on ‘When to perform hepatic resection for intermediate-stage hepatocellular carcinoma’. Hepatology (Baltimore, Md). 2016;63(3):1050.

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