Effect of Substitution at Amine Functionality of 2,6-Diaminopyridine-Coupled Rhodamine on Metal-Ion Interaction and Self-Assembly

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ABSTRACT: 2,6-Diaminopyridine-coupled rhodamines 1 and 2 have been synthesized, and the effect of substitution on amine functionality toward metal-ion interactions and self-assembly is thoroughly investigated. Both the compounds effectively recognize different metal ions of biological significance fluorimetrically and colorimetrically with a high degree of selectivity and sensitivities. While compound 1 is sensitive to Fe\(^{3+}\) ions, compound 2 is responsive to both Fe\(^{3+}\) and Al\(^{3+}\) ions in aqueous CH\(_3\)CN (4/1, v/v; 10 mM tris HCl buffer, pH 6.8). The sensing mechanism involves the metal-ion chelation-induced spirolactam ring opening of the rhodamine scaffold that results in both color and fluorescence changes, while the extent of interactions with the metal ions is truly governed by the chemical structure of the compounds. Both 1 and 2 are proficient in detecting Fe\(^{3+}\) and Al\(^{3+}\) ions in human lung cancer cells (A549). As new findings, unlike 1, compound 2 formed a faint pink gel in the toluene-hexane mixture solvent (1:1, v/v), and the gel state of 2 selectively recognizes Ag\(^{+}\) ions by exhibiting a phase change from gel to purple sol. Experimental findings establish the role of the formamide moiety in forming the self-assembly.

INTRODUCTION

Since last few decades, the development of convenient and directly responsive techniques for recognition of ionic analytes has been desirable in the area of supramolecular chemistry because of their environmental and physiological relevance.\(^1\) In this endeavor, fluorogenic probes with suitable binding sites are preferable because of high selectivity and sensitivity of fluorescence particularly in the solution state.\(^1\) In relation to this, colorimetric sensors are much convenient than fluorogenic sensors as the recognition event can be read out visually without using any costly instrument.\(^2\) Therefore, the construction of both fluorogenic as well as chromogenic probes is of special attention.\(^3,4\) In developing these probes, the use of rhodamine fluorochromes is notable for their excellent photophysical properties that include long absorption and emission wavelengths and high quantum yields.\(^4\) For rhodamine compounds, the sensing is attributed to their metal ion-induced switching from the spiroring system (colorless) to the ring-opened form (pink colored, highly fluorescent).\(^4,5\) In exploring metal-ion sensing, the use of rhodamine is stimulating although the event is somewhat solvent and pH-dependent.

In addition to these attributes, the stimuli-responsive self-aggregation behavior of the molecular architecture is often of considerable importance in reporting naked eye detection of ionic analytes.\(^7\) Compounds of this class require the presence of suitable functional groups with hydrophobic and hydrophilic balance in the backbone which encourages the formation of three-dimensional-crosslinked network in solution involving weak noncovalent interactions. Encapsulation of the solvent in such networks produces a highly viscous semisolid phase called the gel. Exploitation of this supramolecular gel in sensing involves the interaction of externally added ionic analytes with the gelator molecules that causes perturbation to the intermolecular forces occurring among the gelator molecules. As a consequence, the self-aggregated superstructure is destroyed, and the semisolid phase is converted into solution. Apart from the sensing study, supramolecular gels are incredibly important in the field of material to medicine.\(^8\) However, designing of such gelator molecules is complicated and receives increasing attention in material chemistry research.\(^9\) The literature survey reveals that, although huge numbers of rhodamine-based fluororeceptors are reported,\(^10\) only few are explored in gel chemistry.\(^10\) Rajasekar and Das has introduced a series of rhodamine-N-glycosylamine conjugates that form gels in various organic solvents.\(^10\) However, the use of rhodamine-based supramolecular gelators in sensing of the metal ion is completely unknown in the literature.
Herein, we report the synthesis of designed rhodamine-coupled 2,6-diaminopyridine 1 and its derivative 2 toward metal ion sensing and self-assembly under different conditions. While in the semiaqueous medium compound 1 fluorimetrically as well as colorimetrically distinguished Fe$^{3+}$ ions from other different metal ions examined, compound 2 exhibited strong interaction for Fe$^{3+}$ and Al$^{3+}$ ions under identical conditions. Both the compounds were eligible for in vitro detection of the said metal ions in human lung cancer cells (A549). Experimental results signified the role of the formamide moiety in stabilizing the adduct formation with metal ions. Incorporation of formamide functionality resulted in significant increase in the association constant values for compound 2 with Fe$^{3+}$ and Al$^{3+}$ ions.

Incorporation of the formamide moiety on 1 not only influenced the interaction strength toward metal ions but also allowed compound 2 to self-assemble into a fibrous structure involving intermolecular hydrogen bonding. Thus, compound 2 was capable of forming a faint pink gel in a nonpolar solvent viz toluene-hexane (1:1, v/v). The metal ion responsiveness of 2 was significantly altered in the gel state compared to the solution state. The gel was transformed into sol with a color change from faint pink to deep violet selectively in the presence of an Ag$^+$ ion. To our opinion, this attribute is a solution state. The gel was transformed into sol with a color change of the solution from non emissive spectrum of 1 at 570 nm selectively in the presence of both Fe$^{3+}$ and Al$^{3+}$ ions.

**RESULTS AND DISCUSSION**

**Synthesis.** Scheme 1 describes the synthesis of compounds 1 and 2. The reaction of 2,6-diaminopyridine with rhodamine acid chloride 3 yielded the compound 1, which on reaction with ammonium formate in dry acetonitrile afforded rhodamine-labeled formamide 2. All the compounds were adequately characterized by different spectroscopic methods. Good quality crystals of the compound 2 were obtained from the CH$_3$OH solvent, and the structure was solved by single crystal X-ray analysis (Figure 1b).

**Metal Ion Binding in Solution.** The metal-ion responsive behaviors of 1 and 2 toward different metal ions were studied in CH$_3$CN/H$_2$O (4:1, v/v, 10 mM tris HCl buffer, pH = 6.8). The binding interactions were followed using UV–vis, fluorescence, and nuclear magnetic resonance (NMR) spectroscopic tools. Before examining the interactions, the effect of pH on both the compounds 1 and 2 was studied. As can be seen from Figures S1 and S2, the solutions of 1 and 2 are colorless from pH 4 to 12. Below pH 4, solutions become pink in color because of spirolactam ring-opening. The UV–vis spectra of 1 as recorded at different pHs reveal strong absorption at 560 nm in the pH range 2–3 because of the xanthenoid form of rhodamine (Figure S1b). Compound 2 exhibited a similar result (Figure S2b). Thus, the metal-ion interaction study of 1 and 2 was performed at pH 6.8 where the spirolactam ring is retained.

The change in emission of 1 was observed upon gradual addition of metal ions to the solution of 1 in aq. CH$_3$CN (CH$_3$CN:H$_2$O = 4:1, v/v, 10 mM tris HCl buffer, pH = 6.8). Compound 1 exhibited a weak emission at 570 nm ($\lambda_{\text{exc}} = 480$ nm) which remained unperturbed upon interaction with all the metal ions examined except Fe$^{3+}$ ions (Figure S3). Figure 2a, in this context, shows the change in the fluorescence ratio $[(I - I_0)/I_0]$ of 1 at 588 nm in the presence of 20 equiv amounts of metal ions ($c = 3.0 \times 10^{-5}$ M; $\lambda_{\text{exc}} = 480$ nm) at 588 nm upon the addition of 20 equiv amounts of metal ions ($c = 3.0 \times 10^{-5}$ M); (b) change in emission of 1 upon addition of 20 equiv amounts of Fe$^{3+}$ ions in CH$_3$CN/water (4/1, v/v; 10 mM tris HCl buffer, pH 6.8) (Inset: emission intensity of 1 as a function of Fe$^{3+}$ ion concentration and related color change of the solution upon exposure to UV light).

As we move from compound 1 to 2, a different sensitivity for metal ions was observed in fluorescence. In this case, upon interaction with the metal ions, the nonemissive spectrum of 2 was disturbed significantly and furnished strong band at 585 nm selectively in the presence of both Fe$^{3+}$ and Al$^{3+}$ ions (Figure S4).

![Figure 1](image1.png)

**Figure 1.** (a) Structures of compounds 1 and 2 and (b) single crystal X-ray structure of compound 2. Hydrogen atoms are removed for clarity.

![Figure 2](image2.png)

**Figure 2.** (a) Change in the fluorescence ratio of 1 ($c = 3.80 \times 10^{-5}$ M, $\lambda_{\text{exc}} = 480$ nm) at 588 nm upon the addition of 20 equiv amounts of metal ions ($c = 3.0 \times 10^{-5}$ M); (b) change in emission of 1 upon addition of 20 equiv amounts of Fe$^{3+}$ ions in CH$_3$CN/water (4/1, v/v; 10 mM tris HCl buffer, pH 6.8) (Inset: emission intensity of 1 as a function of Fe$^{3+}$ ion concentration and related color change of the solution upon exposure to UV light).
obvious changes in the emission spectra (Figures 3a and S5). In reference to 1, additional inclination of 2 for Al$^{3+}$ is believed to be due to its N-formyl oxygen donor center. However, in comparison to 1, the emission of 2 at 585 nm was perturbed in almost equal extent and reached the plateau on gradual addition of only 10 equiv amounts of Fe$^{3+}$ and Al$^{3+}$ ions. Again, the nonfluorescent solutions became reddish brown in the presence of both Fe$^{3+}$ and Al$^{3+}$ ions under UV exposure (Figure 3b,c).

Figure 3. (a) Change in the fluorescence ratio of 2 ($c = 3.80 \times 10^{-5}$ M, $\lambda_{ex} = 480$ nm) at 585 nm upon addition of 10 equiv. amounts of metal ions ($c = 3.0 \times 10^{-3}$ M, $\lambda_{ex} = 480$ nm). Change in emission of 2 ($c = 3.80 \times 10^{-5}$ M) with 10 equiv. amounts of (b) Fe$^{3+}$ and (c) Al$^{3+}$ ($c = 3.0 \times 10^{-3}$ M) ions in CH$_3$CN–H$_2$O (4:1, v/v, Tris–HCl buffer, pH = 6.8) containing 1% DMF. [Inset: Emission intensity of 2 as a function of metal ion concentration and color change of the solutions during titration upon exposure to UV light].

Figure 4. (A) Suggested mode of interaction of 1 with metal ions in solution. (B) Partial $^1$H NMR (400 MHz, CDCl$_3$) of (a) compound 1 ($c = 1.97 \times 10^{-2}$ M) and (b) 1 with the equiv. amount of Fe$^{3+}$. (C) Suggested mode of interaction of 2 with metal ions in solution. (D) Partial $^1$H NMR of (a) 2 ($c = 5.49 \times 10^{-3}$ M), (b) 2 with Al$^{3+}$ (1:1) and (c) 2 with Fe$^{3+}$ (1:1) in DMSO–$d_6$. 

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The change in fluorescence response of 1 at 588 nm in the presence of other metal ions was also recorded, and data are cited in Figure S6, which corroborates its high selectivity and sensitivity toward Fe³⁺ ions. This experimental finding distinguishes the Fe³⁺ ion from the other metal ions studied. On the other hand, competitive experiments revealed mutual interference of Fe³⁺ and Al³⁺ ions for 2 in the detection process. The noninterfering nature of other metal ions as shown in Figure S7 indicates that compound 2 is able to sense both Fe³⁺ and Al³⁺ ions fluorimetrically over the other metal ions considered in the study with a high degree of selectivity and sensitivity. The distinction of these two metal ions was possible by F⁻ ions as well as by their concentration variation as discussed later.

In the interaction, 1:1 stoichiometry of the 1-Fe³⁺ complex was evaluated from the Benesi-Hildebrand plot and the binding constant (K_b) for the formation of the 1-Fe³⁺ complex was estimated to be 3.06 x 10⁵ M⁻¹ (Figure S8). The detection limit of I for Fe³⁺ was calculated to be 4.48 x 10⁻⁶ M (Figure S9). Similarly, the linear nature of the plots of change in emission intensity against the reciprocal of concentration of metal ions suggests 1:1 stoichiometric complexes of 2 with Fe³⁺ and Al³⁺ ions with association constants of 7.85 x 10⁵ M⁻¹ and 2.17 x 10⁴ M⁻¹, respectively (Figure S10). Detection limits of 2 for Fe³⁺ were determined to be 2.79 x 10⁻⁶ M which is almost comparable to that of Al³⁺ (2.43 x 10⁻⁶ M) (Figure S11). Comparison of the data indicates that a structural change from amine 1 to formamide 2 not only increase the metal-binding capability but also allows the lower limit of detection of the aforementioned metal ions.

To our opinion, the selective response of 1 toward the Fe³⁺ ion is ascribed to the strong participation of the binding center with the metal ion as shown in Figure 4A. In Fourier transform infrared (FTIR), the amide carbonyl stretching at 1698 cm⁻¹ for the spiroring moiety was changed into a lower wave number 1632 cm⁻¹ in the presence of Fe³⁺ ions and thereby suggested the opening of the spiroring (Figure S12). Also, in 1H NMR, the rhodamine ring protons of types Hα moved to the downfield direction with broadening in the presence of the equiv amount of Fe³⁺ ions (Figure 4B). Because of the interaction, the proton of the free amine (a-type) underwent a downfield chemical shift of ∼1 ppm and became broad (paramagnetic effect of Fe³⁺). This suggested the involvement of the pyridine nitrogen in metal coordination.

Similarly, strong metal ion chelation into the binding cleft of 2 according to the mode suggested in Figure 4C is responsible for the generation of the xanthene moiety. As a result of this, in FTIR, the stretching frequencies of both N-formyl and amide carbonyls shifted to the lower regions by 3–8 and 23–24 cm⁻¹, respectively, and became broad (Figure S13). Comparison of 1H-NMR spectra of 2 in the presence and absence of the equiv. amount of Fe³⁺ and Al³⁺ ions also supports this proposition. Almost similar changes were observed in the 1H-NMR. In both cases, strong metal–ligand interactions resulted in significant broadening of the formamide –NHs. Moreover, a downfield chemical shift by 0.54 ppm of the rhodamine ring protons (Hα,β) along with 0.28 ppm downfield movement of the methylene protons strappingly corroborated the spiroring opening of 2 in the presence of Fe³⁺ and Al³⁺ ions, respectively (Figure 4D).

The ground-state interactions were understood by recording the changes in the UV–vis spectrum of 1 and 2 with the same metal ions in CH₃CN–H₂O. In UV–vis study, compound 1 initially showed strong absorption at 276 and 315 nm. The intensities of these peaks were dramatically enhanced along with the manifestation of a new absorption at 559 nm in the presence of Fe³⁺ (Figure 5a). The growth of the absorption peak at 559 nm is the consequence of spiroring opening in 1 that corresponds to the generation of the delocalized xanthene moiety in 1A. In the event, the colorless solution of 1 became pink in color. It is further mentionable that although the colorless solution of 1 became faint pink in color in the presence of Al³⁺ ions (ε = 1.91 x 10⁴ M⁻¹ cm⁻¹) (Figure 5b), the response of the compound for this ion was considerably less than Fe³⁺ (ε = 3.34 x 10⁴ M⁻¹ cm⁻¹). This was not observed with other metal ions during the titration (Figure S14).

On the other hand, in case of 2, the strong absorptions at 274 and 308 nm were irregularly perturbed, and no characteristics changes were observed for all metal ions except Fe³⁺ and Al³⁺ ions (Figure S15). In the presence of Fe³⁺ (ε = 5.51 x 10⁵ M⁻¹ cm⁻¹) and Al³⁺ (ε = 4.51 x 10⁵ M⁻¹ cm⁻¹), new absorption at 558 nm was again progressively intensified with a change in color of the solution from colorless to pink (Figure S5c,d). This corroborated that Fe³⁺ and Al³⁺-induced spiroring opening in 2. Compound 2 remained silent in the titrations with other metal ions (Figure S16).

The reversible nature of metal complexation was tested by titrating the ensembles of 1 and 2 with Fe³⁺/Al³⁺ ions with different anions separately. Initially, the fluorescence and
absorption of the Fe³⁺ complex of 1 were studied with various anions in CH₃CN/water. During fluorescence titration of the ensemble of 1-Fe³⁺ with the anions such as F⁻, OAc⁻, and H₂PO₄⁻, the peak at 588 nm was progressively decreased and became labeled off (Figure S17). Under similar conditions, other anions brought moderate change in emission of the ensemble. The effectiveness of the metal ion (characterized by the fading of the peak at 588 nm) varied as F⁻ ≈ OAc⁻ ≈ H₂PO₄⁻ > Cl⁻ ≈ Br⁻ ≈ I⁻ ≈ ClO₄⁻. To our opinion, this order appears from strong interaction of more basic anions with the metal ion.⁵a A similar trend was noted in UV−vis titration. The absorption band at 559 nm vanished upon successive addition of F⁻, OAc⁻, and H₂PO₄⁻ ions and resulted in colorless solution (Figure S17). These findings pointed out the regeneration of the spiroring in the rhodamine part through the removal of the coordinated metal ion.

Similar experiments were carried out with the ensembles of 2 with Fe³⁺ and Al³⁺ ions. In fluorescence and UV−vis, during titration with the AcO⁻ ion, the peak at 585 and 558 nm, respectively, became labeled off (Figures S18−S19). Interestingly, while F⁻ brought a marked change in the optical properties of the 2-Fe³⁺ ensemble, under identical conditions, moderate decrease in intensities of the peaks at 585, 558 nm for the 2-Al³⁺ ensemble in emission and absorption spectra, respectively, was observed (Figure S20). Thus, the F⁻ ion is able to discriminate the Fe³⁺ and Al³⁺ complexes of 2 by exhibiting strong affinity for Fe³⁺ which in turn encouraged the demetallation process from the probe binding site. Rest of the anions undertaken showed a similar trend as that of 1 for both the ensembles (Figures S18−S19).

For compound 2, as we were not able to fully distinguish between Al³⁺ and Fe³⁺ ions by F⁻, we checked the complexation characteristics with variation of their concentrations. Importantly, successful distinction was achieved at low concentrations of Al³⁺ and Fe³⁺ ions. At concentration of 1 × 10⁻⁴ M of metal ions, compound 2 distinguished Al³⁺ from Fe³⁺ by showing different UV−vis spectra as well as color of the solution of 2. Figure 6 explains this.

**Metal-Ion Sensing in the Gel State.** In contrast to 1, compound 2 was capable of forming self-assembled superstructures in the solvent of particular polarity. While the presence of most of the protic/polar solvents led to either precipitation or poor solubility, in toluene-hexane (1:1, v/v), it exhibited instant gelation (mgc = 12 mg/mL). Under identical conditions, the nongelation behavior of 1 signified the key role of the formamide functionality in 2 that augments the self-aggregation by amplifying a number of intermolecular hydrogen bond contacts. Table S1 reports the gelation profiles of 1 and 2 in various solvents.

The gel of 2 obtained from toluene−hexane (1:1, v/v) exhibited thermoreversible gel−sol phase transformation, and the gel melting temperature (Tg) increased linearly with the gelator’s concentration (Figure S21). To understand the
hydrogen bond-mediated aggregation, FTIR spectra of 2 in amorphous and gel states were compared. Owing to hydrogen bonding, the stretching frequency of N-formyl carbonyl was shifted to the lower region by 3 cm\(^{-1}\) in the gel state (Figure S22). Such hydrogen bonding features of the formamide motif are widely explored.\(^{12}\) In the present case, the engagement of formamide in the formation of the hydrogen-bonded dimer in the solid state is confirmed from packing of the crystal (Figure 7a, Table S2). We believe that the rhodamine moiety with hydrophobic surfaces stabilizes the hydrogen-bonded network in the less polar solvent through hydrophobic–hydrophobic interaction. However, weak forces enable the gelator 2 to produce non-twisted rod-like fibers and fiber-bundles in the aggregated state, as evident from scanning electron microscopy (SEM) images (Figure 7b,c).

After characterizing the gel, it was explored in metal-ion sensing. In this purpose, same metal ions as used earlier were undertaken and added as the solid on the top of the gel at room temperature. In presence of 2 equiv amounts of all metal ions, the gel state remained unaffected except Ag\(^+\) which resulted in violet-colored sol formation within 2 h (Figure 8A).

![Figure 8](image)

Figure 8. (A) Representation of the phase change of the toluene–hexane (1:1, v/v) gel of 2 (12 mg/mL) in the presence of 2 equiv amounts of (a) Pb\(^{2+}\), (b) Ag\(^+\), (c) Cu\(^{2+}\), (d) Ni\(^{2+}\), (e) Zn\(^{2+}\), (f) Cd\(^{2+}\), (g) Fe\(^{3+}\), (h) Fe\(^{3+}\), (i) Al\(^{3+}\), (j) Co\(^{2+}\), and (k) Hg\(^{2+}\) ions after 2 h; normalized (B) UV–vis, and (C) emission spectra of 2 in sol and gel states.

The inability of other metal ions in bringing such gel-to-sol transition of 2 indicated the selectivity for Ag\(^+\) ions. However, addition of tetrabutylammonium chloride to the Ag\(^+\) ion containing sol did not cause reappearance of the gel. This is presumably due to poor solubility of the 2-Ag\(^+\) complex in the medium which prevents the demetallation reaction.

The Ag\(^+\)-induced disruption of the gel was probably associated with the strong metal coordination involving the amide ion (obtained from spiroring opening) and pyridyl nitrogen atoms\(^{13}\) that disturbed the intermolecular association of the molecules. A strong band at 575 nm in UV–vis of the broken gel established the spiroring opening (Figure 8B). In fluorescence, because of such spiroring opening, a strong emission at 657 nm was observed (Figure 8C). In FTIR, the stretching frequency of the amide carbonyl was shifted to a lower region by 24 cm\(^{-1}\) (Figure S23).

Cell Imaging. The utility of compounds 1 and 2 to detect Fe\(^{3+}\) and Al\(^{3+}\) ions in live cells was investigated through fluorescence. When A549 cells were treated with only 40 \(\mu\)M of either compound 1 or compound 2 alone, they do not show any fluorescence under the microscope (Figures 9A and S24A). When cells were incubated with 3 mM Fe\(^{3+}\) or Al\(^{3+}\) along with the compound 1 or 2, cells showed fluorescence. It appears from Figure 9E that compound 1 shows intense fluorescence with Fe\(^{3+}\) ions. This fluorescence was reduced to a significant extent when 3 mM F\(^-\) ion was added (Figure 9G). The cells incubated individually with 3 mM Fe\(^{3+}\) and Al\(^{3+}\) along with compound 2 also exhibited fluorescence as shown in Figure S24.

Similarly, compounds 1 and 2 with Al\(^{3+}\) showed intense fluorescence as shown in Figures 9I and S24G, respectively. Notably, this fluorescence was reduced to the significant extent when the 3 mM F\(^-\) ion was added further. For example, when 3 mM F\(^-\) was added to compounds 1 and 2 with Al\(^{3+}\), the fluorescence was decreased as shown in Figures 9K and S24I. No fluorescence in Figure 9C represents that the cells treated with acetonitrile only with the dose used for preparation of solutions of compounds used in these experiments. However, there is no effect of acetonitrile on cell viability within the dose used in our experiment. We used each compound up to 40 \(\mu\)M, and cell viability was observed to be about 82 and 72% for compounds 1 and 2, respectively, at the dose (Figure S25).

**CONCLUSIONS**

In conclusion, 2,6-diaminopyridine-coupled rhodamine B derivatives 1 and 2 have been synthesized and the effect of substitution on amine functionality toward metal-ion interactions and self-assembly are thoroughly investigated. The compounds effectively recognized different metal ions of their...
biological significance in organic as well as in a semiaqueous system by exhibiting both color and fluorescence changes with a high degree of selectivity and sensitivity. Experimental findings reveal that the compound 1 which contains the amine group can be coupled to formyl functionality producing a new structure 2 to modulate the metal-ion selectivity under different conditions. While compound 1 is sensitive to Fe³⁺, compound 2 is responsive to Fe³⁺ as well as Al³⁺ ions in the aqueous CH₃CN medium with a higher association constant. The distinction of the Fe³⁺ ion form Al³⁺ has been possible by F − as a chelating ion as well as by concentration variation of the metal ions. Compounds are cell permeable and capable to detect the mentioned metal ions in human lung cancer cells (a549).

Moreover, unlike 1, compound 2 forms a faint pink gel from the toluene-hexane (1:1, v/v) mixture solvent. Experimental observations establish the role of the formamide moiety in 2 that augments the self-aggregation by amplifying the number of intermolecular hydrogen bond contacts compared to 1. The gel is metal ion responsive and selectively recognizes Ag⁺ ions by exhibiting violet-colored sol formation. Thus, the selectivity and sensitivity can easily be altered by the incorporation of different functionalities on the –NH₂ group of 1. There are reasonable numbers of silver responsive gelators found in the literature (Table S3). Importantly, the gel phase sensing of metal ions by rhodamine derivatives is completely unknown, and therefore, compound 2 draws attention, in this regard.

### EXPERIMENTAL SECTION

#### Materials and Methods.

2,6-Diaminopyrididine, rhodamine B, POCl₃, and ammonium formate were procured from Spectrochem. Metal perchlorates in the study were obtained from Sigma-Aldrich and were handled with care. All solvents as used in the synthesis were properly dried and distilled before use. Solvents for NMR experiments were procured from Aldrich. Thin layer chromatography was done on Merck precoated silica gel 60–F₂₅₄ plates. ¹H and ¹³C NMR spectra were recorded on a Bruker 400 MHz instrument. High-resolution mass data were obtained from an XEVO GS-2 QToF Waters mass spectrometer using the electron spray ionization technique. FTIR measurements of the compounds were on a PerkinElmer L120-00A spectrometer (ν max in cm⁻¹) using KBr pellets. Fluorescence and UV–vis are studied using a PerkinElmer spectrophotometer (LS55) and Shimadzu UV-2450 spectrophotometer, respectively. SEM images were obtained on the EVO LS-10 ZEISS instrument.

#### Synthesis. Compound 1.³ᵃ Rhodamine B (0.5 g, 1.12 mmol) was taken in 1,2-dichloroethane (20 mL) and stirred for 20 min. Then, POCl₃ (0.21 mL, 2.24 mmol) was added to this stirred solution. The reaction mixture was heated under reflux for 2 h. The mixture was cooled. and excess POCl₃ was evaporated under reduced pressure to have compound 3 as a solid mass, which was used directly in the next step without purification. Compound 3 was dissolved in dry CH₂Cl₂ (30 mL), and to this, a solution of 2,6-diaminopyridine (0.49 g, 4.48 mmol) containing Et₃N (0.25 mL, 1.77 mmol) was added. The mixture was stirred for 10 h at room temperature, and the completion of reaction was verified by thin layer chromatography. Finally, the solvent was removed, and water was added. The mixture was extracted with ethyl acetate. The organic layer was collected and dried over anhydrous sodium sulphate. Evaporation of ethyl acetate gave a crude mass which was next purified by column chromatography using 40% ethyl acetate in petroleum ether to give a pure compound 1 (0.72 g) in 58% yield. Mp 208 °C, ¹H NMR (CDCl₃, 400 MHz): δ 7.98 (d, 1H, J = 8 Hz), 7.78 (d, 1H, J = 8 Hz), 7.50–7.47 (m, 2H), 7.29–7.26 (m, 1H), 7.13 (d, 1H, J = 8 Hz), 6.40–6.37 (m, 4H), 6.13 (dd, 2H, J₁, J₂ = 6 Hz, J₂, J₃ = 4 Hz), 6.01 (d, 1H, J = 8 Hz), 4.06 (s, 2H), 3.32–3.27 (m, 8H), 1.13 (t, 12H, J = 8 Hz); ¹³C NMR (CDCl₃, 100 MHz): δ 167.9, 156.0, 153.9, 153.4, 149.1, 148.3, 138.5, 133.3, 131.1, 128.1, 124.4, 123.0, 109.1, 107.0, 104.9, 103.2, 97.2, 66.1, 44.2, 12.6 (one carbon in the aromatic region is unresolved); FTIR (KBr) ν cm⁻¹: 3498, 2967, 2853, 1699, 1634, 1444, 1221; Mass (EI): 534.0 (M + 1)⁺.

**Compound 2.** To a stirred solution of compound 1 (500 mg, 0.94 mmol) in dry acetonitrile (10 mL), anhydrous ammonium formate (120 mg, 1.90 mmol) was added, and the resulting mixture was heated to reflux at 95 °C (bath temperature) for 10 h. After removal of acetonitrile under reduced pressure, a residue was diluted with ethyl acetate (2 × 20 mL). The organic layer was separated, dried over anhydrous Na₂SO₄, and then reduced pressure, a residue was diluted with ethyl acetate (40 mL). The mixture was stirred for 10 h at room temperature, and the resulting mixture was heated to reflux at 95 °C (bath temperature) for 10 h. After removal of acetonitrile under reduced pressure, a residue was diluted with ethyl acetate (2 × 20 mL). The organic layer was separated, dried over anhydrous Na₂SO₄, and then concentrated under reduced pressure to afford the compound 2 (269 mg, 51%) as a white solid, mp 232 °C. ¹H NMR (DMSO-d₆, 400 MHz): δ 10.21 (d, 1H, J = 8 Hz), 8.50 (d, 1H, J = 8 Hz), 8.09 (d, 1H, J = 8 Hz), 7.91 (d, 1H, J = 8 Hz), 7.64 (t, 1H, J = 8 Hz), 7.60–7.51 (m, 2H), 7.00 (d, 1H, J = 8 Hz), 6.40–6.37 (m, 3H), 6.34 (s, 2H), 6.23–6.21 (m, 2H), 3.33–3.19 (m, 8H), 1.05 (t, 12H, J = 8 Hz); ¹³C NMR (DMSO-d₆, 100 MHz): δ 168.1, 163.0, 150.4, 154.3, 152.1, 149.8, 149.3, 148.3, 140.1, 134.6, 128.9, 128.7, 127.5, 124.3, 123.5, 111.0, 108.0, 107.9, 106.7, 97.5, 65.7, 44.1, 12.7; FTIR (KBr) ν cm⁻¹: 3279, 2972, 2929, 1701, 1670, 1617; HRMS (TOF MS) (ES⁺): calcd 534.2864 (M + CO + H)⁺; found, 534.2852 (M + CO + H)⁺.

**General Procedures for Fluorescence and UV–Vis Titrations.** Stock solutions of the compounds were prepared in the desired solvent in a concentration of 3.80 × 10⁻³ M. Stock solutions of metal ions were in the same solvent in a concentration of 3.0 × 10⁻³ M. Working solution of each compound (2 mL) was placed in the cuvette. Stock solutions of different metal ions were added in different amounts. Upon adding metal ions, the change in emission of the compound was recorded. The same stock solutions were used in the similar way in UV–vis titrations.

**Binding Constant Determination.**¹¹ The binding constant (Kₐ) was determined by the Benesi–Hildebrand plot using the expression: \(I_0/\Delta I = \left[\varepsilon_M / (\varepsilon_M - \varepsilon_C)\right] (K_aC_{M} + 1)\), where \(\varepsilon_M\) and \(\varepsilon_C\) are molar extinction coefficients of the compound and the complex, respectively, at a selected wavelength; \(I_0\) represents the emission intensity of free compounds at that specific wavelength, and \(C_{M}\) is the concentration of the metal ions. The linear fit of \(I_0/\Delta I\) with the inverse of the metal ion concentration indicated 1:1 stoichiometry of the complex. The ratio of the intercept to slope was used for binding constant (Kₐ) determination.

**Calculation of the Detection Limit.**²⁶ The detection limit was determined using the fluorescence titration data. The emissions of 1 and 2 at 588 and 585 nm, respectively, was measured six times to achieve the standard deviation of blank measurement. The fluorescence intensities at 588 and 585 nm for 1 and 2, respectively, were plotted against concentrations of metal ions to determine the slope. Then, detection limits were calculated using the equation: Detection limit = 3σ/k, where σ
and $k$ refer the standard deviation of blank measurement and the slope, respectively.

**Gelation Test.** The necessary amount of compound 2 was taken in a solvent (1 mL) of choice and slightly warmed to make a homogeneous solution. Gelation which was checked via the usual inversion of the vial method occurred when the temperature of the undisturbed solution reached to room temperature. For SEM imaging, the sample of the gel was dried under vacuum, and then, it was coated with a thin layer of gold metal.

**Method of Determination of Gel–Sol Transition Temperature.** The gel transition temperature ($T_g$) at which gel is melted and started to flow was measured by the dropping ball method. In this method, a small glass ball was placed on the top of the gel taken in a test tube. The tube was then slowly heated in a thermostated oil bath. The temperature at which the ball fell and reached the bottom of the test tube was assigned as $T_g$ of the gel.

**Cell Culture.** A549 cell was purchased from the National Centre for Cell Sciences, Pune, India. Cells were cultured in the Dulbecco’s modified Eagle’s medium supplemented with 10% fetal bovine serum at 37 °C under humidified conditions containing 5% CO₂.

**Cellular Uptake.** About 50,000 cells were grown on a cover slip in each well of a 24-well plate. After overnight incubation, media were discarded and washed with 1X PBS. Then, cells were incubated with appropriated concentration of the compounds 1 and 2 with or without Al³⁺, Fe³⁺, and $F^-$. We took 40 μM of compounds 1 and 2 with or without 3 mM of Al³⁺, Fe³⁺, and $F^-$. Then, those cover slips were placed on a clean grease-free glass slide upside down. Cell images were captured under 40× magnification of AxioscopeA1 (Carl Zeiss) fluorescence microscope.

**Cell Viability by Trypan Blue Method.** About $10^5$ A549 cells were plated in each 35 mm plates. Cells were then treated with different concentrations of compounds 1 and 2 (10 μM, 20 μM and 40 μM). After 24 h, treatment cells were incubated with 0.2% of Trypan blue. Then, cells were counted with a hemocytometer chamber under a light microscope.

## ASSOCIATED CONTENT

### Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/10.1021/acsomega.0c01384.

Emission and absorption spectra, change of fluorescence, Benesi-Hildebrand plot, binding curves, detection limit, NMR and mass data, pictorial representations, cell viabilities, copies of $^1$H, $^{13}$C NMR, FTIR and HRMS spectra (PDF)

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### Notes
The authors declare no competing financial interest.

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