Genetics of diaphragmatic hernia

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INTRODUCTION

Congenital diaphragmatic hernia (CDH), accounting for roughly 8% of all major congenital malformations, is a severe physical deformity associated with high morbidity and mortality and occurs in less than 5 cases in 10,000 births [1, 2]. Due to improved treatment options survival has greatly improved up to 88% [3] but long-term morbidity commonly remains an important issue [4]. CDH is caused by a discontinuity in the diaphragm allowing abdominal organs to penetrate into the thoracic cavity, to interfere with heart and lung development thus causing a series of severe pathophysiologic events: pulmonary hypoplasia, pulmonary hypertension (PH) following increased pulmonary vascular resistance (PVR) and cardiac impairment are hallmarks of CDH [5]. Notably, most cases occur sporadically, however, genetic causes have long been discussed to explain a proportion of cases. These range from aneuploidy to complex chromosomal aberrations and specific mutations often causing a complex phenotype exhibiting multiple malformations along with CDH. This review summarises the genetic variations which have been observed in syndromic and isolated cases of congenital diaphragmatic hernia.

GENETIC VARIATIONS IN CONGENITAL DIAPHRAGMATIC HERNIA

The significance of copy number variations (CNVs)

Several studies have examined the association between the CNVs and certain diseases and provide evidence for both causative and predisposing relations [10–12]. To what extent these genomic imbalances could potentially contribute to CDH development has been unclear but recent scientific evidence sheds light on this relatively new genetic approach, revealing a subset of as much as 10% of CDH cases as attributable to CNVs [13]. Notably, most CNVs that have so far been identified to play a role in the development of CDH are deletions, which in the context of haploinsufficiency are considered deleterious in the process of diaphragm development. However, several CDH-related chromosomal loci are affected by copy number gains. To date, CNVs putatively responsible for CDH could be found in numerous loci. Zhu et al. compared 196 CDH cases to 987 healthy controls and identified six CNVs significantly associated with CDH (Suppl. Table 1) [14]. Of these, a gain in copy number of parts of HLX1 found in five patients with CDH is easiest to associate to the developing diaphragm since Hlx could be determined to the murine embryonic septum transversum and diaphragm [15, 16]. Also, Hx-signal was colocalized with Myod in myoblasts of the upper somites [15] suggesting its contribution to myogenesis. These findings make HLX1 an interesting candidate gene in CDH.

Disruption of essential signalling pathways can lead to CDH

Disp1 is a protein similar to the hedgehog-receptor Ptc acting in the Sonic Hedgehog (Shh) signalling pathway [17]. In mouse

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studies, Disp1 is specifically expressed in the PPFs of the developing premature diaphragm [18]. Yet, Shh-expressing cells were shown to neither contribute to the muscular, nor to the nonmuscular part of the developing murine diaphragm in fate-mapping studies, and Ptc1 was not found in the PPFs [18]. Therefore, the Disp1 function remains unknown in diaphragm development. However, murine Shh-signalling contributes to phrenic nerve outgrowth [18], which in neural tube cells is executed by the transcription factors GlI2 and GlI3 and a promoter element of NR2F2 (COUP-TFII) responsive to Shh-signalling was identified in a murine cell line [20]. A similar activation of NR2F2 during human diaphragm development has yet to be uncovered. But since Shh-signalling is crucial for embryonic lung [21] and phrenic nerve development and mutations of GLI2 and GLI3 are described in human cases of CDH [22, 23], Shh-signalling could still influence the developing human diaphragm. Furthermore, Kantarci et al. discovered two distinct deletions on chromosome 1 encompassing both HLX and DIP1 associated with CDH that consistently remain of uncertain significance [18]. In contrast, there is evidence for functional Wnt/β-catenin-signalling in the developing diaphragm upstream of which acts WT1 to promote β-catenin expression and cell proliferation especially in the posterior diaphragm [24]. In the embryo, canonical Wnt-signalling is an essential driver for development and requires multiple cellular proteins like WNT5A and FZD2 [25]. Consistently, there is evidence suggesting disrupted Wnt-signalling could contribute to CDH in a male infant harbouring a deletion in 17q12.2 encompassing FZD2 [26]. In line with the importance of Wnt/β-catenin-signalling, Scott et al. reported on a male infant exhibiting a deletion on chromosome 11p13 covering WT1 and PAX6 [27]. Notably, PAX6 is required in myogenesis and its contribution to the CDH phenotype could not be excluded.

Low copy repeats at 8p23 are responsible for CNVs leading to CDH

A mutational hotspot for structural chromosomal aberrations (e.g., translocations, inversions) and microaberrations (e.g., point mutations, base deletions, indels) is the short arm of chromosome 8. Yu et al. and Keitges et al. determined CNVs at 8p23.1 in CDH patients with additional ventricular septal defect (VSD), atrial septal defect (ASD) or incomplete atrioventricular canal defect, or both ASD and VSD [28, 29]. In one case, only two genes, namely GATA4 and NEIL2, were affected by the deletion pinpointing on another CDH candidate region located at 8p23. NEIL2 is involved in base excision repair and is therefore not considered a good CDH candidate. At that time, loss of function mutations of GATA4 were already associated to cardiac anomalies [30, 31] and today we know that GATA4 is important for both embryonic cardiac and lung development [32, 33]. Furthermore, the examination of diaphragm development in mice in which Gata4 was specifically deleted in the PPFs identified muscle connective tissue fibroblasts as a source of CDH in Gata4-deficient organisms [34]. Developing CDH can therefore be associated with GATA4 deficiency in the nonmuscular mesenchyme. Additionally, Wat et al. reported on a male infant with a heterozygous de novo del(8p23.1) mutation encompassing GATA4, exhibiting multiple cardiac anomalies along with anterior, right-sided CDH [35] similar to those observed in Gata4+/−/− mice [33]. Notably, its monozygotic twin brother presented with cardiac defects only, omitting CDH despite the presence of a similar mutation also involving GATA4 [35]. These findings from animal studies suggest a striking value of GATA4 transcriptional activity in the developing diaphragm. Notably, both deletions encompassed SOX7, another transcription factor that acts upstream of GATA4 and might multiply the disrupting effects of GATA4 deficiency either if SOX7 itself is deleted or affected by missense mutations [36].

NR2F2 (COUP-TFII) – Key element in CDH?

NR2F2 (COUP-TFII), mentioned previously as a potential target of Shh-signalling in neural cells, was also found to be expressed in the murine nonmuscular mesenchyme of the PPFs and is frequently affected by deletions in patients with CDH [37-39]. Consistently, Nr2f2-knockout mice develop diaphragmatic hernia [40]. Interestingly, multiple pathways unite upon this essential transcription factor (Supplemental Figure 1), which was found to colocalize with WT1, likewise expressed in PPF connective tissue [37] and to interact with GATA4 and FOGL2 [41]. We are unable to provide possible mechanisms of action for all of the CNVs described in impaired diaphragm development. However, a comprehensive list of additional CNVs associated with CDH is presented in Supplementary Table 1.

POINT MUTATIONS ASSOCIATED WITH CDH SNVS

The pinpointer for candidate genes encompassed by CNVs

Besides complex CNVs, also single nucleotide variations (SNVs) are associated with cases of CDH. Some of these mutation sites correspond to regions where CNVs have already been detected thus highlighting significant genes within the deleted or duplicated loci, e.g., NR2F2 [42] or GATA4 [43] (Suppl. Table 2). Similar to GATA4, GATA6 is another zinc-finger transcription factor involved particularly in heart and lung organogenesis [44, 45] and interestingly, CDH cases exhibiting GATA6 variants were also reported [46, 47]. Notably, GATA6 expression is restricted to the mesenchymal part of the PPFs [48]. The activity levels of GATA4 and GATA6 are both modulated by the zinc-finger transcription factor FOGL2 (also known as ZFPM2) thus activating or repressing transcriptional activity. Hence, mutations affecting FOGL2 could also interfere with the diaphragm, heart, and lung development. At E13.5, FOGL2 and GATA4 were found to be expressed in nuclei in cells of the PPF. Conversely at E16.5, GATA4 and FOGL2 were also determined in the muscularized diaphragm though only FOGL2 switched localization to cytosolic [37]. To our knowledge, Ackerman et al. were the first to discover FOGL2 variants in association with CDH [49]. In their experiments, FOGL2+/− mice displayed pulmonary dysplasia and the absence of an accessory right lobe along with an intact but muscular posterior and peripheral diaphragm [49]. Furthermore, FOGL2−/− mice exhibited severely downregulated HGF expression in the PPFs. How FOGL2 might influence the developing diaphragm is not yet fully understood. However, the authors concluded, that impaired migration of muscle cell progenitors due to defective HGF/c-MET signalling led to the non-muscularized diaphragm and hypothesised HGF expression might be directly regulated by FOGL2. Also, FOGL2 is thought to act as a corepressor of NR2F2 [50]. Consistently, SNVs in MET were found in cases of CDH [23, 51] supporting the concept of defective migration as a cause for high diaphragmatic vulnerability.

THE ROLE OF VITAMIN A HOMOEOSTASIS AND SIGNALLING PATHWAYS

There is scientific consensus about the importance of vitamin A and its derivatives in embryonic lung development [52, 53]. Tissue-specific retinaldehyde dehydrogenases (RALDHs) generate retinoic acids (RAs) in two different isoforms, namely all-trans retinoic acid and 9-cis retinoic acid both binding to different intracellular receptors (RAR or RXR), which then form heterodimers and translocate to the nucleus, where they act as transcription factors by binding to specific retinoic acid response elements (RAREs, RXRES). During alveologenesis, RALDH2 (ALDH1A2) and ALDH1H1 specifically expressed in lung tissue are of significant importance for generating RAs [54], which are thought to perpetuate mesodermal proliferation, to induce FGF10 and to affect shh-signalling during the period of initial budding and
branching morphogenesis [52, 55, 56]. Also, retinoic acids act upstream of homeobox transcription factors like HOXB4 to promote lung differentiation [57]. Interestingly, there are mutated variants of HOXB4 associated with CDH [23]. Supplemental Figure 1 provides an overview on vitamin A’s metabolism and target genes in lung development. The observation, that vitamin A deficiency during rat dams’ pregnancy did not only impede lung development but did also cause diaphragm defects in the pups led to the hypothesis that vitamin A signalling would be involved in its morphogenesis [58, 59]. Indeed, Aldh1a2, retinol-binding protein Rbp, and Rarβ were found to be strongly expressed in the PPs of rat pups [60–62] and generated Rar-knockout strains exhibiting diaphragmatic defects additional to pulmonary hypoplasia further highlight the importance of vitamin A in diaphragm development [61]. Conversely, Clugston et al. evaluated the presence of Rara, Rary and Rxor but did not determine Rarβ transcripts in the developing murine diaphragm [63]. Subsequently performed studies on human newborns revealed a reduction of retinol and RBP in the chord blood by up to 50% in CDH cases [64]. In consistence with the emerging concept of vitamin A-dependent diaphragm development, it was shown NR2F2 (COPU-TFI) transcription factors could act as repressors of and in turn be regulated by retinoic acid signalling [65, 66] and by this, a possible mechanism in CDH pathogenesis was revealed. Supporting the concept of essential vitamin A signalling in diaphragm development, both point mutations in and CNVs of genes involved in vitamin A metabolism and signalling were described affecting several levels of retinoid signalling. Starting at the top of the signalling cascade, SNVs in STRA6 (encoding a membrane receptor involved in the uptake of vitamin A) and a CNV covering STRA6 potentially compromising vitamin A uptake in affected cells were discovered [67, 68]. Interestingly, Aldh1a2 was found to be affected by duplications on chromosome 9 in CDH cases [69, 70].

Furthermore, one of those duplications also covered the retinoid acid receptors RARA and RXRA, the terminal endpoints of retinoid signalling [69]. Given the numerous mutations in members of the retinoid signalling pathway discovered in CDH patients (Supplementary Table 1 and 2), we provide evidence for its significant contribution to diaphragm development and CDH pathogenesis.

**DISCUSSION**

We comprehensively summarise point mutations and copy number variations described in the literature associated with CDH. Both have contributed equally to the identification of CDH candidate genes and are nowadays indispensable in its deep diagnosis. However, declaring detected genes explicitly causative would be a bold statement with regards to the complexity of genetics and organogenesis during embryonic development. Because deleted or duplicated chromosomal loci often carry many different genes, a disease-causing genotype-phenotype association is a difficult venture. Also, the deletion or duplication of promoter regions or regulatory units could possibly escape our notice. Moreover, the fact that in some cases unaffacted relatives harbour the same genetic anomaly further underlines the complexity of CDH pathogenesis. Nevertheless, our summary of genetic findings associated with CDH offers the possibility for clinicians and geneticist to classify rare mutations in affected infants as potentially relevant, and subsequently to screen parents for the respective genetic variant and offer genetic counselling. By doing so, future pregnancies rated high-risk upon the presence of certain mutations could be closely monitored in order to improve the outcome of the newborn. The development of diaphragmatic hernias can be considered the endpoint of a single (rare), dual, or even multiple hit event in multiple tissues or the interaction between cells of different ancestry. We consider mutational events in the pathogenesis of CDH as being of predisposing character as it remains difficult to make single specific events solely responsible for the development of diaphragmatic defects. Mutated genes can be assigned to multiple signalling pathways (e.g., retinoid, hedgehog, wnt), play a role in various areas of cellular homeostasis or are crucial during embryonic development. The majority resembles transcription factors of which some had been exhaustively investigated and their presence in the premature murine diaphragm or pleuroperitoneal folds have been determined. However, we think more transcription factors and proteins could be found in the premature diaphragm and further extend the spectrum of CDH candidate genes and possible pathogenic backgrounds. There seems to be more independent factors explaining why some individuals with certain genetic anomalies do develop CDH and others do not. Likewise, genetics alone could hardly explain the local restriction of CDH defects. Furthermore, there is evidence for essential vitamin A signalling in the developing primitive lungs and diaphragm. Disruption of the retinoid signalling pathway would therefore be able to constitute a basis for an in vivo applicable dual hit hypothesis, suggesting that lung hypoplasia could in part occur independently of a diaphragmatic defect or even precede it. In the literature, possible pathways by which vitamin A could impede diaphragm development are poorly discussed. Hence, its contribution to the pathogenicity of CDH remains largely unknown. By demonstrating RA-inducible NR2F2 activity, at least one possible mechanism of RA-regulated CDH development has been revealed. Further research needs to be done to verify if there are interactions between retinoic acid and other transcription factors known to be crucial for diaphragm development.

**DATA AVAILABILITY**

All data generated or analysed during this study are included in this published article (and its supplementary information files).

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**ADDITIONAL INFORMATION**
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