Association of HDL-Related Loci with Age-Related Macular Degeneration and Plasma Lutein and Zeaxanthin: the Alienor Study

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Abstract

Background: Several genes implicated in high-density lipoprotein (HDL) metabolism have been reported to be associated with age-related macular degeneration (AMD). Furthermore, HDL transport the two carotenoids, lutein and zeaxanthin, which are highly suspected to play a key-role in the protection against AMD. The objective is to confirm the associations of HDL-related loci with AMD and to assess their associations with plasma lutein and zeaxanthin concentrations.

Methods: Alienor study is a prospective population-based study on nutrition and age-related eye diseases performed in 963 elderly residents of Bordeaux, France. AMD was graded according to the international classification, from monochromatic colour retinal photographs. Plasma lutein and zeaxanthin were determined by normal-phase high-performance liquid chromatography. The following polymorphisms were studied: rs493258 and rs10468017 (LIPC), rs3764261 (CETP), rs12678919 (LPL) and rs1883025 (ABCA1).

Results: After multivariate adjustment, the TT genotype of the LIPC rs493258 variant was significantly associated with a reduced risk for early and late AMD (OR=0.64, 95%CI: 0.41-0.99; p=0.049 and OR=0.26, 95%CI: 0.08-0.85; p=0.03, respectively), and with higher plasma zeaxanthin concentrations (p=0.03), while plasma lipids were not significantly different according to this SNP. Besides, the LPL variant was associated with early AMD (OR=0.67, 95%CI: 0.45-1.00; p=0.05) and both with plasma lipids and plasma lutein (p=0.047). Associations of LIPC rs10468017, CETP and ABCA1 polymorphisms with AMD did not reach statistical significance.

Conclusion: These findings suggest that LIPC and LPL genes could both modify the risk for AMD and the metabolism of lutein and zeaxanthin.

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Introduction

Age-related macular degeneration (AMD) is a degenerative disease of the central part of the retina (macula), responsible for half of the cases of blindness in industrialized countries [1]. This disease affects 2.5 million subjects in Europe [2] and 1.75 million in the USA [3]. It comprises two late forms both associated with severe visual impairment (neovascular and atrophic AMD), generally preceded by early, asymptomatic, retinal abnormalities (drusen, pigmentary abnormalities). It is a multifactorial disorder involving genetic and environmental factors [4]. Since 2005, major associations with genes in the complement pathway (complement factors H, B, C2 and C3) and the ARMS2 gene have been identified [4]. Besides, the influence of environmental factors (in particular smoking and diet) has been highlighted by epidemiological studies [5–8]. In particular, lutein and zeaxanthin, two carotenoids of dietary origin, are suspected to play a key-role in the prevention against AMD [9].

Indeed, lutein and zeaxanthin are highly concentrated in the macula (30 to 10 000 times higher than in others tissues), in which they constitute the macular pigment (MP). MP may help protect against AMD because of its physical property of blue light filtration and its local antioxidant activity [9,10]. Epidemiological studies have reported a decreased risk for AMD in subjects with high plasma concentration or dietary intake of lutein and zeaxanthin [11–14]. The metabolism of these lipophilic molecules is closely interlinked with cholesterol metabolism [9]. Indeed, they are mainly carried by HDL [15]. In addition, several lipid transporters, such as SR-BI, CD36 or NCP1/L1, are implicated in the transport of carotenoids [16]. Genetic variations in the corresponding genes of these transporters have been reported to be associated with alterations of plasma carotenoid status [16–18]. Alterations in cholesterol metabolism might therefore influence MP accumulation, with consequences on the risk for AMD.

Associations of AMD with several genes implicated in HDL metabolism have been reported recently. In 2010, two genome-wide association studies [19,20] (GWAS) reported associations of AMD with the hepatic lipase (LIPC) gene, and suggested further associations with the cholesteryl ester transfer protein (CETP) gene, the lipoprotein lipase (LPL) gene and the ATP-binding cassette transporter A1 (ABCA1) gene. These findings have been partly replicated in a few studies (Table 1) [21–31].

Regarding LIPC, two SNPs have been studied in relation to AMD: rs493258, which was initially identified by the GWAS studies on AMD [19,32], and rs10468017, which was initially identified by a GWAS study on HDL concentrations [33]. Both SNPs, located on chromosome 15 (15q21.3), are in linkage disequilibrium (r²=0.42), and most studies on AMD tested only the rs10468017 SNP (Table 1). With regard to rs493258, in Caucasians, all four available studies have shown a significantly decreased risk for AMD in subjects bearing the minor (T) allele. With regard to rs10468017, in Caucasians, the risk for AMD was significantly reduced in subjects bearing the minor (T) allele in 6 studies, while 4 other studies showed no statistically significant associations. Two studies in Han Chinese showed no associations of AMD with rs493258 or rs10468017 [29,31].

As for ABCA1, most studies, including one study performed in Han Chinese subjects, showed a decreased risk for AMD in subjects bearing the minor (T) allele, although one study did not find any associations [28]. The CETP minor allele (A) was also generally associated with increased risk for AMD, although the association failed to reach statistical significance in 4 studies. Finally, results concerning LPL were more inconsistent: one of the original GWAS studies found a significantly increased risk for AMD [19], while the other GWAS study found a non significant decreased risk [20], consistently with three more recent case-control studies (including one in Han Chinese subjects).

These genetic associations suggest that lipid metabolism, and in particular HDL-cholesterol, is implicated in the pathophysiology of AMD. However, the relationship between AMD and HDL cholesterol may not be straightforward. Indeed, epidemiological studies evaluating associations between plasma lipids (including HDL-cholesterol) and AMD have been inconsistent, with conflicting results [34–39]. A recent study also reported that the associations of AMD with LIPC and with HDL-cholesterol seem to be independent [26], thereby implying that the relationship between HDL-related genotypes and AMD may be mediated by other mechanisms. Finally, the direction of associations of AMD with HDL-related loci is not consistent: while for LIPC, the HDL-raising allele is associated with lower risk for AMD, while for ABCA1 and CETP, they confer higher risk for AMD (Table 1). Thus, as noted by Neale et al [20], the association of LIPC and other HDL-related loci with AMD may not represent a causal effect of HDL cholesterol, but could rather indicate a shared underlying biological mechanism involving the cholesterol pathway.

As explained above, the metabolism of lutein and zeaxanthin, being strongly associated both with AMD and HDL cholesterol, is a potential candidate for such common mechanism between AMD and HDL related genetic loci. However, very few data are available on the potential relationship of LIPC and other HDL-related genes with plasma lutein and zeaxanthin. Only one study of 129 subjects showed some associations of plasma carotenoids and vitamin E with LIPC and CETP [18].

Here, we assessed the associations of several genetic loci implicated in HDL metabolism with AMD and with plasma lutein and zeaxanthin concentrations in the framework of a population-based study. The main originality of the present study resides firstly, in its population-based design, which overcomes major issues in selection bias inherent to case-control studies. Secondly, we provide results for the associations of HDL-related loci with early AMD, the pre-clinical stage of AMD, while most previous studies have included only late AMD cases. Finally, we provide some data on the associations of these loci with plasma lutein and zeaxanthin, which might, at least partially, explain the association of AMD with these genetic loci.
Subjects and Methods

Study aims
The Alienor (Antioxydants, Lipides Essentiels, Nutrition et maladies OculaiRes) Study is a population-based study aiming at assessing the associations of age-related eye diseases (AMD, glaucoma, cataract, dry eye syndrome) with nutritional factors (in particular antioxidants, macular pigment and fatty acids), determined from estimation of dietary intakes and plasma measurements [40]. It also takes into account other major determinants of eye diseases, including gene polymorphisms, environmental and vascular factors.

This research followed the tenets of the Declaration of Helsinki. Participants gave written consent for the participation in the study. The design of the Alienor study has been approved by the Ethical Committee of Bordeaux (Comité de Protection des Personnes Sud-Ouest et Outre-Mer III) in May 2006.

Table 1. Summary of studies about AMD and genes implicated in cholesterol metabolism.

| Study (First author, Year) | Design   | Ethnicity | Subjects (n, AMD definition) | Associations with AMD (OR (95% CI)) for minor allele |
|----------------------------|----------|-----------|-----------------------------|---------------------------------------------------|
|                            |          |           |                             | LIPC rs10468017  | LIPC rs493258  | ABCA1 rs1883025 | CETP rs3764261 | LPL rs12678919 |
| Chen, 2010 [19]            | GWA      | Caucasian | Large drusen, late GA, late NV AMD, n=2157. Control, n=1150. | 0.87 (0.82-0.91)  | 0.83 (0.76-0.89) | 1.19 (1.21-1.27) | 1.26 (1.11-1.43) |
| Neale, 2010 [20]           | GWA      | Caucasian | Late NV AMD, n=979. Control, n=1709. | 0.82 (0.77-0.88)  | 0.86 (0.82-0.91) | 0.77 (0.66-0.90) | 1.12 (1.04-1.20) | 0.85 (0.71-1.02) |
| Reynolds, 2010 [26]        | Case-control | Caucasian | Late AMD (NV and GA), n=318. Controls, n=140. | CT: 0.6 (0.4-0.9) T: 0.4 (0.2-0.9) |
| Seddon, 2010 [25]          | Case-control, AREDS | Caucasian | Late AMD (NV and GA), n=545. Controls, n=275. | CT: 0.9 (0.7-1.2) TT: 0.5 (0.2-0.9) |
| Fauser, 2011 [27]          | Case-control | Caucasian | AMD (at least 10 small, hard drusen and pigmented changes or at least one intermediate size drusen), n=1201. Control, n=562. | 0.93 (0.78-1.09)  | 0.69 (0.52-0.86) | 1.14 (0.97-1.33) | 0.98 (0.77-1.26) |
| Peter, 2011 [24]           | Multicenter, Case-control | Caucasian | Women, Late AMD, n=146. Control, n=1269. | CT: 0.4 (0.2-0.9) TT: 0.6 (0.2-2.0) CT: 0.8 (0.4-1.5) TT: 0.2 (0.1-0.7) | CA: 1.1 (0.6-2.1) AA: 1.5 (0.6-3.7) | 0.6 (0.3-1.5) |
| Sobrin, 2011 [23]          | Multicenter, Case-control | Caucasian | Late AMD (NV and GA), n=3958. | 1.07 (0.94-1.23) |
| Yu, 2011 [21]              | Case-control, AREDS | Caucasian | Late AMD (NV and GA), n=1082. Control, n=221. | 0.68 (0.53-0.87)  | 0.63 (0.50-0.81) | 1.22 (0.97-1.55) |
| Yu, 2011 [22]              | Meta-analysis of case-control studies | Caucasian | Late AMD, n=2594. Control, n=4134. | 0.84; p<.0001  | 1.15, p<.0001 |
| Cipriani, 2012 [30]        | GWA      | Caucasian | Late NV AMD or late GA, n=893. Control, n=2199. | 0.91(0.80-1.03)  | 0.89 (0.79-0.99) | 1.07 (0.95-1.19) |
| Tian, 2012 [31]            | Multicenter, Case-control | Han | Late AMD, n=535. Control, n=469. | OR not shown p=0.53 | OR not shown p=0.94 |
| Yu, 2012 [28]              | Longitudinal study, AREDS | Caucasian | Incidence of late NV AMD (large drusen➔NV AMD). | TT: 0.64 (0.38-1.08)  | 1.02 (0.85-1.23) | 1.27 (1.08-1.49) |
| Zhang, 2013 [29]           | Case-control | Han | Late NV AMD, n=157. Control, n=204. | 0.75 (0.51-1.10)  | 0.99 (0.70-1.39) | 0.65 (0.46-0.92) | 1.16 (0.78-1.70) | 0.86 (0.55-1.34) |

Abbreviations: AREDS: Age-Related Eye Disease Study; GA: Geographic Atrophy; GWA: Genome-Wide Association; NV: Neovascular

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Subjects of the Alienor Study were recruited from an ongoing population-based study on the vascular risk factors for dementia, the Three-City (3C) Study [41]. The 3C Study included 9294 subjects aged 65 years or more from three French Cities (Bordeaux, Dijon and Montpellier), among whom 2104 were recruited in Bordeaux. They were initially recruited in 1999-2001 and followed-up about every two years since. The Alienor Study consists of eye examinations, which are offered to all participants of the 3C cohort in Bordeaux after the third follow-up (2006-2008) (Figure 1). Among the 1450 participants re-examined between October 2006 and May 2008, 963 (66.4%) participated in the Alienor Study’s baseline eye examination.

Eye examination
The eye examination took place in the Department of Ophthalmology of the University Hospital of Bordeaux at Alienor baseline (2006-2008). It included a recording of ophthalmological history, measures of visual acuity, refraction, two 45° non mydriatic colour retinal photographs (one centered on the macula, the other centered on the optic disc), measures of intraocular pressure and central corneal thickness and break-up time test.

Retinal photographs were performed using a non mydriatic retinograph (TRC NW6S, Topcon, Japan) and were interpreted in duplicate by two specially trained technicians. Inconsistencies between the two interpretations were adjudicated by a senior grader. Finally, all cases of late AMD were reviewed and confirmed by retina specialists (JFK, MND and MBR).

None of the persons implicated in interpretation of retinal photographs had access to plasma measurements, genetic determinations or information on risk factors.

Classification of AMD
Retinal photographs were interpreted according to the international classification [42] and to a modification of the grading scheme used in the Multi-Ethnic Study of Atherosclerosis for drusen size, location and area [43]. Late AMD was defined by the presence of neovascular AMD or geographic atrophy within the grid (3000 microns from the Foveola). Neovascular AMD included serous or hemorrhagic detachment of the retinal pigment epithelium (RPE) or sensory retina, sub-retinal or sub-RPE hemorrhages and fibrous scar tissue. Geographic atrophy was defined as a discrete area of retinal depigmentation, 175 microns in diameter or larger, characterized by a sharp border and the presence of visible choroidal vessels. Five cases of late AMD had no gradable photographs and were classified by using ophthalmological history of AMD and AMD therapy (in particular antiangiogenic agents and photodynamic therapy), and confirmed by their treating ophthalmologist.

Early AMD was defined by the presence of soft distinct drusen and/or soft indistinct drusen and/or reticular drusen and/or pigmentary abnormalities. Soft distinct and indistinct drusen were larger than 125 microns in diameter and with uniform density and sharp edges or decreasing density from the center outwards and fuzzy edges, respectively. Pigmentary abnormalities were defined as areas of hyperpigmentation and/or hypopigmentation (without visibility of choroidal vessels).
Genotyping

Genotyping was performed on DNA extracted from leukocytes at baseline (1999-2001) and kept frozen at -80°C. Centralized facilities for genotyping are provided by the Lille Genopôle. Genotyping of single nucleotide polymorphisms (SNPs) rs1061170 (Complement Factor H, CFH, Y402H) were determined using TaqMan assays (Applied Biosystems, Inc., [ABI], Foster City, CA), as described by the suppliers. Furthermore, a genome wide scan was performed at the Lille Genopôle [44]. For the present study, the following genotypes were extracted from the GWAS data: rs10490924 (ARMS2/HTRA1), rs493258 and rs10468017 (LIPC), rs3764261 (CETP), rs12678919 (LPL) and rs1883025 (ABCA1). Genotypes data for rs10468017 (LIPC), rs3764261 (CETP) and rs12678919 (LPL) were available. For the others SNPs namely rs10490924 (ARMS2/HTRA1), rs493258 (LIPC), rs9621532 and rs1883025 (ABCA1), genotypes were imputed using Markov Chain based haplotype (MACH v1.0.16a) [45,46] software and the 1000 Genomes Project data [47] ($r^2 = 0.96$; $r^2 = 0.94$; $r^2 = 0.99$ and $r^2 = 0.96$, for rs10490924, rs493258, rs9621532 and rs1883025, respectively).

Plasma measurements

Plasma measurements were determined from fasting blood samples collected at the 3C baseline visit (1999-2001) into heparinized evacuated tubes and centrifuged at 1000 g for 10 min. Plasma lipids (HDL and LDL-cholesterol, triglycerides) were measured at the Biochemistry Laboratory of the University Hospital of Dijon from baseline fasting blood samples.

Plasma lutein and zeaxanthin measurements were performed at DSM Nutritional Products (Kaiseraugst, Switzerland). Their concentrations were determined by normal-phase HPLC, using dedicated analytical methods [48]. Plasma samples were analysed for zeaxanthin (sum of all-E and Z-isomers) and lutein (sum of all-E and Z-isomers). The xanthophylls were extracted from plasma (100 mL) with a 20% mixture of n-hexane and chloroform (1100 mL) after dilution with water (100 mL) and proteins precipitation with ethanol (200 mL). After centrifugation, an aliquot (800 mL) of the clear supernatant fluid was dried under nitrogen at room temperature. The dried residue was quantitatively redissolved in the mobile phase (200 mL n-hexane and acetone; 19%, by vol). The resulting solution was injected (100 mL) into a normal-phase HPLC system equipped with an autosampler (15°C), a column oven (40°C), an HPLC pump, and an ultraviolet-visible detector. Data were analysed with a data acquisition system (Atlas: Thermo Labsystems). The separation was done on a polar column (Lichrosorb, SI60, 5 mm, 250 * 4 mm; Stagroma, Reinach, Switzerland) with a mixture of n-hexane and acetone (19%, by vol) at a flow rate of 1 mL/min. Xanthophylls were detected at a wavelength of 452 nm. To assess the daily and long-term laboratory performance of the HPLC plasma analytics, dedicated control plasma was used. None of the people involved in plasma carotenoid determination had any access to ocular clinical findings or genetic data at any time of the study.

Covariates

Socio-demographic and lifestyle data were collected through face-to-face standardized interview in 1999-2001. They included age, gender and smoking (never smoker, <20 pack-years (PY) and ≥20 PY, where number of PY = packs (20 cigarettes) smoked per day X years of smoking).

Statistical analyses

For comparison between subjects included and not included in analyses, Student test was used for quantitative variables and Chi² test for qualitative variables.

Associations of AMD with all genetic polymorphisms were estimated using logistic Generalized Estimating Equations (GEE) models [49], which allow taking into account the data from both eyes and their intra-individual correlations. We used information for eyes instead of subjects in order to increase statistical power for AMD analyses. For each genetic polymorphism, odds-ratios adjusted for age and gender were estimated using early AMD, and late AMD as the dependent variable, and the SNP genotype variable, age, gender as the independent variables. Odds-ratios adjusted for potential confounders were obtained by adding these confounders as independent variables to the models. Potential confounders retained in the final multivariate models were factors strongly associated with AMD in our cohort (smoking [50], CFH [50] and ARMS2 [51] polymorphisms). Among 963 subjects, 76 (7.9%) had ungradable photographs in both eyes, 53 (5.5%) refused the blood sampling and 8 (0.8%) had missing data for photographs and blood sampling. Among the 826 remaining subjects, available data for the different SNPs were somewhat variable, leaving between 715 and 737 subjects for each analysis.

Associations of SNPs genotype with plasma lipids (triglycerides, HDL and LDL-cholesterol) and lutein and zeaxanthin were performed using analysis of variance adjusted for age, gender and lipid lowering medication. Correlations of plasma lutein and zeaxanthin with plasma lipids were performed using Pearson correlations.

Associations between plasma lutein and zeaxanthin with LIPC and LPL SNPs were evaluated with linear regression models. Plasma lutein and plasma zeaxanthin were used as the dependent variable and the LIPC and LPL genotypes, age, gender, HDL-cholesterol, LDL-cholesterol, triglycerides and lipid lowering medication as the independent variables. Among the 963 participants, 106 (11.2%) participants had missing data for plasma lutein and zeaxanthin: 61 (6.3 %) refused the blood sampling and 47 (4.9%) had unavailable data for plasma lutein and zeaxanthin. Among the 855 remaining subjects, the LIPC rs493258 genotype was available in 761 subjects, the LIPC rs10468017 in 748 and the LPL rs12678919 genotype in 738 subjects.

For AMD variables, eyes with no AMD were the reference. For SNPs genotype, the most frequent polymorphism was used as a reference. Quantitative variables (age, plasma lutein and zeaxanthin, plasma HDL-cholesterol, LDL-cholesterol and triglycerides) were handled as continuous variables in all analyses.
All statistical analyses were performed using SAS version 9.2 (SAS Institute Inc, Cary, NC; procedure GENMOD for the GEE analysis).

**Results**

Table 2 showed baseline characteristics of Alienor participants. Mean of age was 80.2 years (± 4.4), 62.0 % of the sample were women and 32.8 % declared lipid lowering medication use. 64.6 % had never smoked, 18.2 % had smoked less than 20 pack-years and 17.2 % more than 20 pack-years. Mean of BMI was 26.4 kg/m^2 (± 3.9). Means of plasma lutein and zeaxanthin were 166.6 (± 87.5) and 40.8 (± 30.2) µg/L.

As shown in Table 3, after adjustment for age and gender, the TT genotype of LIPC rs493258 SNP was associated with a decreased risk for early and late AMD (model 1). These associations remained significant after further adjustment for smoking, CFH and ARMS2 genotypes and lipid lowering medication (model 2). The CT genotype of LIPC rs10468017 SNP was significantly associated with a decreased risk for early AMD in model 1. However, statistical significance was not maintained after full adjustment. The associations of the CT genotype with late AMD and of the TT genotype with early and late AMD were in the same direction, but far from significant.

We found an inverse association between the AG genotype of LPL rs12678919 SNP and early AMD in model 1, but these associations were of borderline significance (p=0.05) after further adjustment for smoking, CFH, ARMS2 and lipid lowering medication. No associations were found between ABCA1 and CETP polymorphisms and early or late AMD.

As shown in Table 4, plasma lutein and zeaxanthin significantly correlated with plasma cholesterol, and in particular HDL-cholesterol, while they correlated negatively with plasma triglycerides.

As shown in Table 5, associations of LIPC rs493258 TT genotype with plasma zeaxanthin and LPL rs1267819 with plasma lutein were maintained after further adjustment of plasma lipids and lipid lowering medication.

**Table 2. Baseline characteristics of Alienor sample.**

| Characteristic | Alienor sample |
|---------------|----------------|
| Age (yrs) n   | 963            |
| Mean (sd)     | 80.2 (4.4)     |
| Gender (women) n | 597 (62.0) |
| Smoking n (%) | 951            |
| Never         | 614 (64.6)     |
| < 20 pack-years | 173 (18.2) |
| ≥ 20 pack-years | 164 (17.2) |
| Lipid lowering medication (yes) n | 963 |
| n (%)         | 316 (32.8)     |
| Body Mass Index (kg/m^2) n | 955 |
| Mean (sd)     | 26.4 (3.9)     |
| AMD characteristics n (%) |
| n (%)         | 879            |
| No AMD        | 580 (66.0)     |
| Early AMD     | 250 (28.4)     |
| Late AMD      | 49 (5.6)       |
| Plasma characteristics, mean (sd) |
| Lutein (µg/L), n=855 | 166.6 (87.5) |
| Zeaxanthin (µg/L), n=855 | 40.8 (30.2) |
| Lutein+zeaxanthin (µg/L) n=855 | 207.3 (108.6) |
| HDL-cholesterol (mmol/L), n=901 | 1.59±0.39 |
| LDL-cholesterol (mmol/L), n=901 | 3.63 (0.85) |
| Total cholesterol (mmol/L), n=901 | 5.78±0.98 |
| Triglycerides (mmol/L), n=901 | 1.23±0.60 |
| Genetic polymorphism, n (%) |
| LIPC rs49325 | N=806          |
| CC            | 237 (29.4)     |
| CT            | 389 (48.3)     |
| TT            | 180 (22.3)     |
| LIPC rs10468017 | N=792         |
| CC            | 415 (52.4)     |
| CT            | 315 (39.8)     |
| TT            | 62 (7.8)       |
| APOE          | N=889          |
| At least 1 Allele E2 | 120 (13.5) |
| At least 1 Allele E4 | 150 (17.9) |
| ABCA1 rs1883025 | N=806        |
| CC            | 422 (52.4)     |
| CT            | 320 (39.7)     |
| TT            | 64 (7.9)       |
| LPL rs12678919 | N=782         |
| AA            | 581 (74.3)     |
| AG            | 186 (23.8)     |
| GG            | 15 (1.9)       |
| CETP rs3784261 | N=788         |
| CC            | 415 (52.7)     |
| AC            | 310 (39.3)     |
| AA            | 63 (8.0)       |
| CFH           | N=878          |
| CC            | 399 (45.4)     |
| CT            | 378 (43.1)     |
| TT            | 101 (11.5)     |
Discussion

This study confirms the association of AMD with LIPC, and suggests that an association of LIPC with the metabolism of lutein and zeaxanthin could, at least partly, mediate it. With regard to LIPC rs493258, our study was consistent with five previous studies in Caucasians, showing a decreased risk for late AMD in subjects bearing the T allele. Consistently with one previous study [24], association with LIPC rs493258 was significant for subjects with the TT genotype but not for those with the CT genotype, suggesting a recessive mode of effect. Moreover, we also identified a significantly reduced risk for early AMD, also on a recessive mode.

Although our results for LIPC rs10468017 did not reach statistical significance, they were consistent with other studies, showing a decreased risk for AMD in subjects bearing the T allele [21,22,24-26]. Three other studies in Caucasians did not show significant associations of LIPC rs10468017 with AMD [23,27,28]. Although the rs10468017 and rs493258 are linkage disequilibrium (LD) (r²=0.42), LIPC functional variant(s), tagged by rs10468017 and rs493258 markers have not been identified yet, and therefore it is possible that rs493258 may be in LD with the functional variant, whereas rs10468017 SNP, which is further in the promoter region, may not be. Regarding the effect of LIPC variants on HDL-cholesterol level, tedious investigations have highlighted functional variants such as rs1800588 (-514C>T) and variant rs493258 is in LD with the functional variant, whereas rs10468017 SNP, which is yet, and therefore it is possible that rs493258 may be in LD

In our study, we found a significant association between plasma zeaxanthin concentration and LIPC rs493258 variant, but not with LIPC rs10468017 SNP, after adjustment for potential confounders, including plasma lipids. Lutein and zeaxanthin are fat-soluble micronutrients carried by plasma lipoprotein [52], mainly by HDL [15]. As expected, in the present study, plasma lutein and zeaxanthin correlated with plasma cholesterol, and in particular HDL-cholesterol. It is well established that lutein and zeaxanthin absorption is not passive and some genes like SCARB1, ABCG5, BCMO1 and CD36 are implicated in their transport across cellular membranes [16–18]. In a French study, the LIPC rs1800588 variant was associated with plasma concentrations of some carotenoids (γ-tocopherol, α-carotene and β-carotene) but not with xanthophylls [18]. By contrast, we report an association of LIPC rs493258 variant with plasma zeaxanthin, but not with plasma lutein or other carotenoids (data not shown). Finally, a recent study showed a variation of macular pigment density (formed by lutein and zeaxanthin in the retina) according to another

### Table 2 (continued).

| LIPC         | N=806 | 1031 | 281 | 66 |
|--------------|-------|------|-----|----|
| rs493258     |       |      |     |    |
| CC (n=413)   | 1.0 (ref) | 1.0 (ref) | 1.0 (ref) | 1.0 (ref) |
| CT (n=687)   | 0.83 | 0.82 | 0.88 | 0.85 |
| (95 % CI)    | (0.58-1.18) | (0.42-1.62) | (0.61-1.28) | (0.41-1.77) |
| p-value      | 0.30 | 0.57 | 0.51 | 0.67 |
| TT (n=324)   | 0.60 | 0.28 | 0.64 | 0.25 |
| (95 % CI)    | (0.39-0.92) | (0.09-0.83) | (0.41-0.99) | (0.08-0.84) |
| p-value      | 0.02 | 0.02 | 0.046 | 0.03 |

### Table 3. Associations between AMD and genetic polymorphisms of lipids metabolism genes.

#### Model 1: OR adjusted for age and gender

| LIPC         | No AMD | Early AMD | Late AMD |
|--------------|--------|-----------|----------|
| rs493258     |        |           |          |
| CC (n=413)   | 1.0 (ref) | 1.0 (ref) | 1.0 (ref) |
| CT (n=687)   | 0.70 | 0.64 | 0.72 | 0.58 |
| (95 % CI)    | (0.50-0.98) | (0.32-1.25) | (0.51-1.02) | (0.28-1.16) |
| p-value      | 0.04 | 0.19 | 0.07 | 0.12 |
| TT (n=324)   | 0.76 | 0.40 | 0.73 | 0.21 |
| (95 % CI)    | (0.44-1.31) | (0.09-1.79) | (0.41-1.29) | (0.03-1.56) |
| p-value      | 0.33 | 0.23 | 0.27 | 0.13 |

#### Model 2: OR adjusted for age, gender, smoking, CFH and ARMS2 polymorphism, lipid lowering medication

| LIPC         | No AMD | Early AMD | Late AMD |
|--------------|--------|-----------|----------|
| rs493258     |        |           |          |
| CC (n=413)   | 1.0 (ref) | 1.0 (ref) | 1.0 (ref) |
| CT (n=687)   | 0.63 | 0.81 | 0.61 | 0.93 |
| (95 % CI)    | (0.42-0.93) | (0.37-1.80) | (0.45-1.00) | (0.41-2.13) |
| p-value      | 0.02 | 0.61 | 0.05 | 0.87 |
| TT (n=324)   | 1.43 | 2.44 | 1.41 | 3.06 |
| (95 % CI)    | (0.48-4.23) | (0.59-10.09) | (0.47-4.18) | (0.75-12.53) |
| p-value      | 0.52 | 0.22 | 0.54 | 0.12 |

### ABCA1

| rs1883025 | 1062 | 292 | 70 | 1031 | 281 | 66 |
|-----------|------|-----|----|------|-----|----|
| CC (n=755) | 1.0 (ref) | 1.0 (ref) | 1.0 (ref) | 1.0 (ref) | 1.0 (ref) | 1.0 (ref) |
| CT (n=560) | 0.97 | 0.80 | 1.01 | 0.62 |
| (95 % CI) | (0.70-1.34) | (0.41-1.55) | (0.73-1.42) | (0.30-1.28) |
| p-value | 0.84 | 0.51 | 0.93 | 0.19 |
| TT (n=109) | 1.04 | 0.48 | 1.18 | 0.41 |
| (95 % CI) | (0.60-1.81) | (0.10-2.30) | (0.65-2.12) | (0.09-1.85) |

Alienor Study Bordeaux, France (2006-2008).

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impeded us to detect these associations, the minor alleles this last study: we showed an increased risk for late AMD for rs493258. [19,20] reported significant associations with AMD. Results being less frequent for GG. On the contrary, we found a borderline decreased risk for early AMD in the GG genotype, which did not reach statistical significance. The relatively small sample size may have significance. The relatively small sample size may have from the present study were in the same direction (decreased risk for rs1883025 T carriers and increased risk for CETP rs3764261 A carriers), but did not reach statistical significance. The estimated odds-ratios were in the expected range. Therefore, we cannot exclude that some of the observed associations were due to chance finding, although our findings are generally consistent with previous studies in this field.

In conclusion, the present population-based study confirms the association of LIPC polymorphism with AMD and suggests a potential association between LIPC and plasma zeaxanthin concentrations. We also detected associations of LPL with AMD and plasma lutein. These findings suggest that LIPC and LPL may be implicated both in AMD pathogenesis and lutein and zeaxanthin transport. Further study of the LIPC locus may contribute to our understanding and points another pathway in the pathogenesis of AMD. These findings open up new opportunities in the research for prevention and treatment of AMD. Further studies, in particular large prospective population-based studies, will be needed to confirm these findings.

### Table 3 (continued).

|                        | Model 1: OR adjusted for age and gender | Model 2: OR adjusted for age, gender, smoking, CFH and ARMS2 polymorphism, lipid lowering medication |
|------------------------|----------------------------------------|---------------------------------------------------------------------------------------------------|
|                        | No AMD Early AMD Late AMD No AMD Early AMD Late AMD |
| p-value                |                                       |                                                                                                  |
| CETP                   |                                       |                                                                                                  |
| rs3764261 (n)          |                                       |                                                                                                  |
| CC (n=730)             | 1.0 (ref)                              | 1.0 (ref)                                                          1.0 (ref)                                                       1.0 (ref)                                                   |
| AC (n=558), OR         | 1.7 (95 % CI) (0.85-1.61)              | 1.04 (95 % CI) (0.53-2.03)                                                                      1.24 (p-value 0.20) (0.63-2.66)                                |
| AA (n=102), OR         | 0.59 (95 % CI) (0.29-1.20)             | 1.38 (95 % CI) (0.42-4.54)                                                                      0.54 (p-value 0.20) (0.44-5.22)                                |
|                         |                                       |                                                                                                  |
|                         |                                       |                                                                                                  |

Alienor Study (Bordeaux, France, 2006-2008) P value for generalized estimating equation model. Abbreviations: AMD: age-macular degeneration; doi: 10.1371/journal.pone.0079848.b003

LIPC polymorphism (rs6078) [53]. Different variants of the hepatic lipase gene could therefore interfere in different ways with the metabolism of specific carotenoids. Regarding the LPL gene, one GWAS [20] and three case-control studies [24,27,29] found a non-significant decreased risk for AMD in G allele carriers of the rs1267819 SNP. By contrast, one study [19] found a significant increased risk for AMD in G carriers. Our results for late AMD are consistent with this last study: we showed an increased risk for late AMD for the GG genotype, which did not reach statistical significance, probably because of small sample size for GG. On the contrary, we found a borderline decreased risk for early AMD in AG carriers.

With regard to CETP and ABCA1 genes, two initial GWAS [19,20] reported significant associations with AMD. Results from the present study were in the same direction (decreased risk for ABCA1 rs1883025 T carriers and increased risk for CETP rs3764261 A carriers), but did not reach statistical significance. The relatively small sample size may have impeded us to detect these associations, the minor alleles being less frequent for CETP and ABCA1 than for LIPC rs493258. Alienor participants seem to have higher levels of plasma lutein and zeaxanthin than AREDS2 participants [54]: 207.3 (±108.6) versus 179.0 95 % CI (171.0-188.0) µg/L, possibly resulting from differences in dietary habits.

Strengths of our study include the population-based design, with photographic assessment of AMD in all subjects. Previous studies of associations of AMD with HDL-related loci were all case-control, hospital-based studies, where selection bias is a major issue, in particular for controls. Moreover, in other studies, definition of AMD cases was not always clear and control choice maybe questionable. In the present study, we used the international classification of AMD, based on retinal photographs, and distinguished early from late AMD.

A potential limitation to our results is the questionable representativeness of the sample. First, about two thirds of the participants in The 3C Study accepted the eye examination. However, subjects included in the Alienor study were not different from those who did not participate for most parameters of interest in our study [40]. Moreover, the prevalence of AMD in our study was similar to that observed in the same age group in other studies performed in Europe [2,55] and other industrialized countries [3] and the distribution of genetic polymorphisms was similar to that observed in other studies in Caucasians [21–27]. We also observed expected associations of AMD with major risk factors (smoking, CFH and ARMS2) in this sample [50,51].

The relatively small number of late AMD cases (n=70 eyes) limited statistical power for analysis of subcategories (atrophic/neovascular). In addition, associations of LIPC rs10468017, CETP rs3764261 and ABCA1 rs1883025 with early and late AMD did not reach statistical significance in our sample, although the estimated odds-ratios were in the expected range. Finally, a potential limitation of our study is the high number of comparisons performed. Therefore, we cannot exclude that some of the observed associations were due to chance finding, although our findings are generally consistent with previous studies in this field.

In conclusion, the present population-based study confirms the association of LIPC polymorphism with AMD and suggests a potential association between LIPC and plasma zeaxanthin concentrations. We also detected associations of LPL with AMD and plasma lutein. These findings suggest that LIPC and LPL may be implicated both in AMD pathogenesis and lutein and zeaxanthin transport. Further study of the LIPC locus may contribute to our understanding and points another pathway in the pathogenesis of AMD. These findings open up new opportunities in the research for prevention and treatment of AMD. Further studies, in particular large prospective population-based studies, will be needed to confirm these findings.
Figure 2. Associations between plasma lutein, zeaxanthin and genetic polymorphisms of lipids metabolism. Alienor Study, Bordeaux, France. P Anova adjusted for age and gender and lipid lowering medication.
doi: 10.1371/journal.pone.0079848.g002
Figure 3. Associations between plasma lipids and genetic polymorphisms of lipids metabolism. Alienor Study Bordeaux, France. P Anova adjusted for age, gender and lipid lowering medication. 
doi: 10.1371/journal.pone.0079848.g003
Table 4. Correlations of plasma lutein and zeaxanthin with plasma lipids.

|                  | Zeaxanthin | Lutein + zeaxanthin | HDL-cholesterol | LDL-cholesterol | Total cholesterol | Triglycerides |
|------------------|------------|---------------------|-----------------|-----------------|-------------------|---------------|
| Lutein           | 0.61 (<0.0001) | 0.98 (<0.0001)     | 0.30 (<0.0001)  | 0.16 (<0.0001)  | 0.21 (<0.0001)    | -0.16 (<0.0001)|
| Zeaxanthin       | 0.77 (<0.0001) | 0.20 (<0.0001)     | 0.18 (<0.0001)  | 0.21 (<0.0001)  | 0.12 (0.0007)    |               |
| Lutein + zeaxanthin | 0.30 (<0.0001) | 0.18 (<0.0001)     | 0.23 (<0.0001)  |                | -0.16 (<0.0001)  |               |

* Pearson Coefficient correlation (P value)

Table 5. Associations between plasma lutein and zeaxanthin with *LIPC* and *LPL* polymorphisms.

| LIPC rs493258, n=759 | Plasma Lutein (µg/l) | Plasma Zeaxanthin (µg/l) |
|----------------------|-----------------------|--------------------------|
|                      | Mean (SD) | Difference | CI 95% | P | Mean (SD) | Difference | CI 95% | P |
| CC reference, n=227  | 166.9 (92.5) | - | - | - | 38.9 (24.5) | - | - | - |
| CT (CC), n=364       | 161.2 (84.1) | -7.82 | -21.61 - 5.96 | 0.27 | 38.5 (22.9) | -0.82 | -5.77 - 4.13 | 0.76 |
| TT (CC), n=168       | 173.2 (89.4) | 3.82 | -12.76 - 20.39 | 0.65 | 46.3 (48.2) | 6.78 | 0.83 - 12.74 | 0.03 |
| LIPC rs10468017, n=746 |          |          |          |          |          |          |          |          |
| CC reference, n=391  | 165.1 (89.6) | - | - | - | 39.6 (24.8) | - | - | - |
| CT (CC), n=297       | 165.2 (85.0) | -1.53 | -13.96 - 10.91 | 0.81 | 41.5 (37.7) | 1.38 | -3.18 - 5.93 | 0.55 |
| TT (CC), n=58        | 178.7 (88.2) | 13.09 | -9.65 - 35.83 | 0.26 | 46.2 (33.0) | 6.87 | -1.46 - 15.19 | 0.11 |
| LPL rs1267819, n=736 |          |          |          |          |          |          |          |          |
| CC reference, n=547  | 164.8 (87.0) | - | - | - | 39.5 (24.8) | - | - | - |
| AG (AA), n=177       | 163.9 (82.8) | -1.49 | -15.41 - 12.43 | 0.83 | 43.2 (45.3) | 3.13 | -1.98 - 8.25 | 0.23 |
| GG (AA), n=12        | 226.8 (100.6) | 47.76 | 0.48 - 95.03 | 0.047 | 46.1 (18.3) | 4.64 | -12.72 - 22.01 | 0.90 |

* Adjusted for age, gender, HDL-cholesterol, LDL-cholesterol, plasma triglycerides and lipid lowering medication using multivariate linear regression.

Adjusted linear regressions. Alienor Study (Bordeaux, France, 2006-2008).
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Author Contributions

Conceived and designed the experiments: JFK MND MBR FM JCL PA JFD PBG CD. Performed the experiments: JFK MND MBR FM JCL PA JFD PBG CD. Contributed reagents/materials/analysis tools: JFK MND MBR FM JCL PA JFD PBG CD. Wrote the manuscript: BM CM.

References

1. Resnikoff S, Pascolini D, Etya'ale D, Kocur I, Pararajasegaram R et al. (2004) Global data on visual impairment in the year 2002. Bull World Health Organ 82: 844-851. PubMed: 15640920.

2. Augood CA, Vingerling JR, de Jong PT, Chakravarty U, Seland J et al. (2006) Prevalence of age-related maculopathy in older Europeans: the European Eye Study (EUREYE). Arch Ophthalmol 124: 529-535. doi: 10.1001/archopht.124.5.529. PubMed: 1669879.

3. Friedman DS, O'Colmain BJ, Muñoz B, Tomary SC, McCarty C et al. (2004) Prevalence of age-related macular degeneration in the United States. Arch Ophthalmol 122: 564-572. doi: 10.1001/archopht.122.4.564. PubMed: 15078975.

4. Lim LS, Mitchell P, Seddon JM, Holz FG, Wong TY (2012) Age-related macular degeneration. Lancet 379: 1728-1738. doi:10.1016/S0140-6736(12)60282-7. PubMed: 22539989.

5. Chong EW, Kreis AJ, Wong TY, Simpson JA, Guymer RH (2008) Dietary omega-3 fatty acid and fish intake in the primary prevention of age-related macular degeneration: a systematic review and meta-analysis. Arch Ophthalmol 126: 826-833. doi: 10.1001/archopht.126.6.826. PubMed: 18541848.

6. Thornton J, Edwards R, Mitchell P, Harrison RA, Buchan I et al. (2005) Smoking and age-related macular degeneration: a review of association. Eye 19: 935-944. doi:10.1038/sj.eye.6701978. PubMed: 16151432.

7. Chan D (1998) Cigarette smoking and age-related macular degeneration. Optom Vis Sci 75: 476-484. doi: 10.1097/00006324-199807000-00015. PubMed: 9703035.

8. van Leeuwen R, Boekhoorn S, Vingerling JR, Witteman JC, Klaver CC et al. (2005) Dietary intake of antioxidants and risk of age-related macular degeneration. JAMA 294: 3101-3107. doi:10.1001/jama.294.24.3101. PubMed: 16380590.

9. Kijlstra A, Tian Y, Keltie ER, Berendschot TT (2012) Lutein: more than just a filter for blue light. Prog Retin Eye Res 31: 303-315. doi: 10.1016/j.preteyres.2012.03.002. PubMed: 22465791.

10. Whitehead AJ, Mares JA, Danis RP (2006) Macular pigment: a review of current knowledge. Arch Ophthalmol 124: 1038-1045. doi: 10.1001/archopht.124.7.1038. PubMed: 16832030.

11. Delcourt C, Carriere I, Delage M, Barbanger-Gateau P, Schalch W (2006) Plasma lutein and zeaxanthin and other carotenoids as modifiable risk factors for age-related maculopathy and cataract: the POLA Study. Invest Ophthalmol Vis Sci 47: 2329-2335. doi: 10.1167/iovs.05-1235. PubMed: 16723441.

12. (1993) Antioxidant status and neovascular age-related macular degeneration. Eye Disease Case-control study group. Arch Ophthalmol 111: 104-109.

13. Gale CR, Hall NF, Phillips DJ, Martyn CN (2003) Lutein and zeaxanthin status and risk of age-related macular degeneration. Invest Ophthalmol Vis Sci 44: 2461-2465. doi:10.1167/iovs.02-0929. PubMed: 12766044.

14. Ma L, Dou HL, Wu YQ, Huang YM, Huang YB et al. (2012) Lutein and zeaxanthin intake and the risk of age-related macular degeneration: a systematic review and meta-analysis. Br J Nutr 107: 350-359. doi: 10.1017/S0007114510004260. PubMed: 21899805.

15. Clevidence BA, Bieri JG (1993) Association of carotenoids with human plasma lipoproteins. Methods Enzymol 214: 33-46. doi: 10.1016/0076-6879(93)10511-J. PubMed: 8468147.

16. Borel P (2012) Genetic variations involved in interindividual variability in carotenoid status. Mol Nutr Food Res 56: 228-240. doi:10.1002/mnu.201100322. PubMed: 21657063.

17. Borel P, de Edelenyi FS, Vincent-Baudry S, Malezet-Desmoulins C, Margotat A et al. (2011) Genetic variants in BCMO1 and CD36 are associated with plasma lutein concentrations and macular pigment optical density in humans. Ann Med 43: 47-59. doi: 10.1007/s11809-011-00359. PubMed: 21091228.
(2009) Common variants at 30 loci contribute to polygenic dyslipidemia. Nat Genet 41: 56-65. doi:10.1038/ng.291. PubMed: 19060906.

Kathiresan S, Willer CJ, Peloso GM, Demissie S, Musunuru K et al. (2009) Common variants at 30 loci contribute to polygenic dyslipidemia. Nat Genet 41: 56-65. doi:10.1038/ng.291. PubMed: 19060906.

van Leeuwen R, Tomany SC, Wang JJ, Klein R, Mitchell P et al. (2004) Is medication use associated with the incidence of early age-related maculopathy? Pooled findings from 3 continents. Ophthalmology 111: 1169-1175. doi:10.1016/j.ophtha.2003.10.024. PubMed: 15177967.

Delcourt C, Michel F, Colvez A, Lacroux A, Delage M et al. (2001) Associations of cardiovascular disease and its risk factors with age-related macular degeneration: the POLA study. Ophthalmol Epidmiol 8: 237-249. doi:10.1076/opep.8.4.237.1613. PubMed: 11471092.

Wachter A, Sun Y, Dasch B, Krause K, Pauleikhoff D et al. (2004) [Munster age- and retina study (MARS). Association between risk factors for arteriosclerosis and age-related macular degeneration]. Ophthalmologe 101: 50-53. doi:10.1007/s00347-003-0868-1. PubMed: 14872268.

Nowak M, Swietochowska E, Marek B, Szapska B, Wielgoszynski T et al. (2005) Changes in lipid metabolism in women with age-related macular degeneration. Clin Exp Med 4: 183-187. doi:10.1007/s10238-004-0054-z. PubMed: 15750765.

Abalain JH, Carre JL, Leglise D, Robinet A, Leglise D et al. (2002) Is age-related macular degeneration associated with serum lipoprotein and lipoparticle levels? Clin Chim Acta 326: 97-104. doi:10.1016/S0009-8881(02)00288-7. PubMed: 12417100.

Tan JS, Mitchell P, Smith W, Wang JJ (2007) Cardiovascular risk factors and the long-term incidence of age-related macular degeneration: the Blue Mountains Eye Study. Ophthalmology 114: 1143-1150. doi:10.1016/j.ophtha.2006.09.033. PubMed: 17275090.

Delcourt C, Korobelink JF, Barberger-Gateau P, Deleyer MN, Rougier MB et al. (2010) Nutrition and age-related eye diseases: the Allenor (Antioxydants, Lipides Essentiels, Nutrition et maladies OcAuRais) Study. J Nutr Health Aging 14: 854-861. doi:10.1007/s12603-010-0131-9. PubMed: 21125205.

Group CS (2003) Vascular factors and risk of dementia: design of the Three-City Study and baseline characteristics of the study population. Neuroepidemiology 22: 316-325.

Bird AC, Bressler NM, Bressler SB, Chisholm IH, Coscas G et al. (1995) An international classification and grading system for age-related maculopathy and age-related macular degeneration. The International ARM Epidemiological Study Group. Surv Ophthalmol 39: 367-374. doi:10.1016/0925-698X(95)00001-0. PubMed: 7604380.

Klein, Klein BE, Knudtson MD, Wong TY, Cotch MF et al. (2006) Prevalence of age-related macular degeneration in 4 racial/ethnic groups in the multi-ethnic study of atherosclerosis. Ophthalmology 113: 373-380. doi:10.1016/j.ophtha.2005.12.013. PubMed: 16513457.

Lambert JC, Heath S, Even G, Campion D, Sleepers K et al. (2009) Genome-wide association study identifies variants at CLU and CR1 associated with Alzheimer's disease. Nature 41: 1094-1099. doi:10.1038/ng.2439. PubMed: 19060906.

Li Y, Willer CJ, Ding J, Scheet P, Abecasis GR (2010) MaCH: using sequence and genotype data to estimate haplotypes and unobserved genotypes. Genet Epidemiol 34: 816-834. doi:10.1002/gepi.20533. PubMed: 20156334.

Li Y, Willer C, Sanna S, Abecasis G (2009) Genotype imputation. Annu Rev Genomics Hum Genet 10: 387-406. doi:10.1146/annurev.genom.9.081307.164242. PubMed: 19715440.

1000 Genomes Project Consortium, Abecasis GR, Altshuler D, Auton A, Brooks LD, Durbin RM, Gibbs RA, Hurles ME, McVean GA (2010) A map of human genome variation from population-scale sequencing. Nature 467: 1061-1073. doi:10.1038/nature09534. PubMed: 20981092.

Hartmann D, Thürmann PA, Spitzer V, Schalich W, Mannner B et al. (2004) Plasma kinetics of zeaxanthin and 3-dehydro-lutein after multiple oral doses of synthetic zeaxanthin. Am J Clin Nutr 79: 410-417. PubMed: 14985215.

Zeger SL, Liang KY, Albert PS (1988) Models for longitudinal data: a generalized estimating equation approach. Biometrics 44: 1049-1060. doi:10.2307/2531734. PubMed: 3233245.

Delcourt C, Delyfer MN, Rougier MB, Amouyel P, Colín J et al. (2011) Associations of complement factor H and smoking with early age-related macular degeneration: the ALIENOR study. Invest Ophthalmol Vis Sci 52: 5955-5962. doi:10.1167/iovs.10-6235. PubMed: 21642625.

Delcourt C, Delyfer MN, Rougier MB, Lambert JC, Amouyel P et al. (2012) ARMS2 A69S Polymorphism and the Risk for Age-Related Maculopathy: The ALIENOR Study. Arch Ophthalmol 130: 1734-1740. PubMed: 22893087.

Tysander V, Choubert G, Grolier P, Borel P (2002) Carotenoids, mostly the xanthophylls, exchange between plasma lipoproteins. Int J Vitam Nutr Res 72: 300-308. doi:10.1024/0300-9831.72.5.300. PubMed: 12463105.

Meyers KJ, Johnson EJ, Bernstein PS, Iyengar SK, Engelman CD et al. (2013) Genetic determinants of macular pigments in women of the carotenoids in age-related eye disease study. Invest Ophthalmol Vis Sci 54: 2333-2345. doi:10.1167/iovs.12-10867. PubMed: 23404124.

Lutein + Zeaxanthin and Omega-3 Fatty Acids for Age-Related Maculopathy: The ALIENOR Study. Arch Ophthalmol 130: 1734-1740. PubMed: 22893087.

Tysander V, Choubert G, Grolier P, Borel P (2002) Carotenoids, mostly the xanthophylls, exchange between plasma lipoproteins. Int J Vitam Nutr Res 72: 300-308. doi:10.1024/0300-9831.72.5.300. PubMed: 12463105.

International ARM Epidemiological Study Group. Surv Ophthalmol 39: 367-374. doi:10.1002/0930-6257(05)00001-0. PubMed: 7604380.