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Misidentification of \textit{Burkholderia pseudomallei}, China

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We report a case of melioidosis in China and offer a comparison of 5 commercial detection systems for \textit{Burkholderia pseudomallei}. The organism was misidentified by the VITEK 2 Compact, Phoenix, VITEK mass spectrometry, and API 20NE systems but was eventually identified by the Bruker Biotype system and 16S rRNA sequencing.

\textit{Burkholderia pseudomallei} is the cause of melioidosis, a serious disease endemic to Southeast Asia and northern Australia (1). Because of the increase in international travel, the disease is now occurring in areas to which \textit{B. pseudomallei} is not endemic. In these previously unaffected areas, laboratory staff might be unfamiliar with the organism or use identification systems that are not suitable for its detection, potentially leading to misidentification (2). We report the misidentification of \textit{B. pseudomallei} by various commercial detection systems.

On May 15, 2019, a man 33 years of age in Guangxi Province, China, sought treatment for leg pain at a local hospital in Guangxi Province. Physicians diagnosed his condition as gout and prescribed oral febuxostat. However, the pain progressively worsened, and the patient began to have difficulty walking. On June 10 he was admitted to Guangzhou First People’s Hospital. Laboratory analysis of serum samples taken at admission showed moderate systemic inflammation with elevated levels of procalcitonin (0.296 ng/mL; reference value <0.05 ng/mL), C-reactive protein (61.7 mg/L; reference value <0.05 ng/mL), and neutrophils (9.42×10^9/L; reference value 1.1–3.2×10^9/L). His temperature fluctuated between 38.5°C and 39.8°C, peaking in the evening. Magnetic resonance imaging results suggested osteomyelitis. We conducted surgical debridement and collected pus from the lesion for microbiological analysis. We used the matrix-assisted laser desorption/ionization time-of-flight (MALDI-TOF) mass spectrometry VITEK 2 Compact system (bioMérieux,
https://www.biomerieux.com) to identify the isolate as *Aeromonas sobria* with 93% probability. According to the VITEK 2 Compact system, the isolate was sensitive to amikacin, meropenem, imipenem, ceftazidime, ciprofloxacin, trimethoprim/sulfamethoxazole, and piperacillin/tazobactam but resistant to cefepime and aztreonam. We made a preliminary diagnosis of *Aeromonas* infection and treated the patient with piperacillin/tazobactam (500 mg, 4×/d) and levofloxacin (500 mg/d). However, we doubted the accuracy of this identification because *Aeromonas sobria* rarely causes extraintestinal disease (3). To examine this suspicion, we collected blood samples and incubated them in the Bact/ALERT 3D automated microbial detection system (bioMérieux). We cultured the samples on sheep blood and chocolate agar, revealing gram-negative rod-shaped bacteria (Figure; Appendix Figure 1, https://wwwnc.cdc.gov/EID/article/27/3/19-1769-App1.pdf). We then tested the samples with a variety of commercial detection systems. The VITEK 2 Compact system again identified the blood sample as *Aeromonas sobria* with 90% probability. However, the Bruker MALDI-TOF Biotyper system (Bruker Daltonics, https://www.bruker.com) identified the isolate as *B. pseudomallei* with an identification score of 2.18 (a score of >2.0 is considered an accurate identification). BD Phoenix M50 (Becton Dickinson, http://www.bd.com) identified it as *Alcaligenes faecalis* with 98% probability; VITEK MS (bioMérieux) identified it as *B. thailandensis* with an identification score of 2.23; API 20NE (bioMérieux) identified it as *Pseudomonas fluorescens* with 75.8% probability (Table).

To confirm the identity of the organism, we extracted DNA from blood cultures using a bacterial genomic DNA isolation kit (Sangon Biotech Co., Ltd, https://www.sangon.com). The 16S rRNA gene was amplified and sequenced by Sangon Biotech Co., Ltd. The isolate showed 100% identity and 100% coverage with a sequence of *B. pseudomallei* collected in India in 2019 (GenBank accession no. CP040552.1). On June 25, we diagnosed melioidosis in the patient. The patient recovered and was discharged after 14 days of the original piperacillin/tazobactam and levofloxacin treatment regimen. The global recommendations from the US Public Health Emergency Medical Countermeasures Enterprise suggest that physicians treat melioidosis with intravenous ceftazidime or meropenem, according to the severity of the disease; alternatively, physicians can prescribe oral trimethoprim/sulfamethoxazole or amoxicillin/clavulanic acid (4).

We conducted multilocus sequence typing as described previously (5). This isolate belongs to sequence type (ST) 550, corresponding with isolates previously documented in Vietnam in 2005 (6). The patient in this study had never been to Vietnam, but Guangxi Province borders that country. We constructed a phylogenetic tree with 1,000 bootstrap replicates using the unweighted pair group method with arithmetic averages in MEGA X software.

### Table. Identification of *Burkholderia pseudomallei* by various detection systems, China, 2019

| Detection method | Identification result (Characteristics) |
|------------------|----------------------------------------|
| Vitek 2 Compact  | *Aeromonas sobria* (90% probability)   |
| Phoenix          | *Alcaligenes faecalis* (98% probability)|
| Bruker Biotyper MS | *Burkholderia pseudomallei* (2.18 score)* |
| Vitek MS         | *Burkholderia thailandensis* (2.23 score)* |
| API 20NE         | *Pseudomonas fluorescens* (75.8% probability) |
| 16S rRNA         | *Burkholderia pseudomallei* (GenBank accession no.CP040552.1) |

*An identification score >2.0 indicates an accurate identification.
The accuracy of the identifications made by VITEK 2 (63%–81%), Phoenix (0%–28%), and API 20NE (37%–99%) systems varied substantially (7,8). Zakharova et al. found that commercially available biochemical identification systems commonly misidentified B. pseudomallei as Chromobacterium violaceum or B. cepacia complex (9). We found that although the isolate in this study was misidentified by multiple systems, most systems accurately identified the genus. MALDI-TOF mass spectrometry is a rapid, accurate, and highly reproducible technique for bacterial identification. Several studies have explored the potential of MALDI-TOF mass spectrometry for the identification of B. pseudomallei. We prefer the Bruker Biotyper system, which is more accurate because the VITEK databases lack reference spectra for B. pseudomallei (10). In conclusion, scientists must be aware of the potential misidentification of B. pseudomallei by automated identification systems, especially those in regions to which B. pseudomallei is not endemic.

About the Author

Mr. Wu is a member of the Department of Laboratory Medicine of Guangzhou First People’s Hospital, Guangzhou. His primary research interest is bacterial infections.

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Autochthonous Case of Pulmonary Histoplasmosis, Switzerland

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