Alpha₁-antitrypsin levels and phenotypes and hepatitis B serology in liver cancer

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Summary Serum levels of alpha₁-antitrypsin (α₁AT) were measured by radial immunodiffusion and phenotypes were determined by electrophoresing in acrylamide gel in 39 patients with hepatocellular carcinoma (HCC) positive for serum hepatitis B surface antigen (HBsAg), 41 patients with HCC negative for serum HBsAg, and 160 age- and sex-matched hospital controls. There was no difference between the control series and either of the two HCC groups with respect to α₁AT phenotype pattern; also, there was no evidence of association between HCC and either the M₁ allele or any of the α₁AT deficiency phenotypes. However, HCC cases negative for HBsAg had significantly higher serum α₁AT values (mean 665 ± 26 mg/100 ml⁻¹) than HCC cases positive for HBsAg (mean 571 ± 23 mg/100 ml⁻¹), who in turn, had significantly higher α₁AT values than hospital controls (mean 434 ± 13 mg/100 ml⁻¹). These results indicate that (i) in Greece, as in other high HCC incidence countries, genetically determined α₁AT deficiency is not aetiologically important; (ii) the increase of serum α₁AT is an important correlate of HCC with possible aetiologic significance and diagnostic potential and (iii) HBsAg-positive HCC and HBsAg-negative HCC are manifest differently as well as being aetiologically distinct.

Alpha₁-antitrypsin (α₁AT) is one of nine distinct plasma proteins which have been characterized as protease inhibitors (Harpel, 1983); it is a glycoprotein synthesized in the liver and it is responsible for about 90 per cent of the trypsin inhibitory capacity of human serum (Sharp, 1976; Kew et al., 1978; Morse, 1978; Chio & Oon, 1979). α₁AT is under genetic control, and more than 30 codominant alleles at a single chromosomal locus have been identified (Chapuis-Cellier, 1975; Cox, 1978; Morse, 1978). Some of these alleles (particularly the alleles Z, I, S, and perhaps P and other less common ones) have been associated with serum α₁AT deficiency of varying degree (Chapuis-Cellier, 1975; Morse, 1978; Chapuis-Cellier & Arnaud, 1979) and with the presence in the liver of periodic acid-Schiff (PAS)-positive, diastase-resistant globules (Sharp, 1976; Morse, 1978; Palmer et al., 1980). There appears to be an intriguing link between α₁AT and hepatocellular carcinoma (HCC) but the evidence is not clear cut (Sharp, 1976; Kew et al., 1978; Morse, 1978; Kelly et al., 1979). We have studied the association between α₁AT and HCC in a large group of HCC cases and matched controls, using adequate laboratory procedures “blindly” (with respect to disease status), in a European population with an unusually high incidence of HCC (Trichopoulos et al., 1982).

Patients and methods

Two hundred and forty patients were studied; they were Caucasians, of Greek nationality and residence, hospitalized in one of eight large hospitals of Athens during a 15-month period in 1976 and 1977. Among them 80 had HCC (confirmed histologically in 47 cases and by diagnostic alpha-fetoprotein (αFP) values in the remaining 33 cases); for each of them two control patients, matched for age and sex, were selected from the same hospitals with diagnoses other than neoplasm or liver disease. Among HCC patients (“cases”) 69 (86%) were males and the average age was 63 years; among comparison patients (“controls”) the corresponding figures were 138 males (86%) and 62 years. All patients were interviewed, and blood samples were taken from each.

Hepatitis B serologic markers were determined in all of the sera by radioimmunoassay; samples of 39 cases (49%) and 12 controls (8%) were found to be positive for hepatitis B surface antigen (HBsAg) and were considered to have chronic infection. The two HCC groups (HBsAg positive and HBsAg negative) were of similar age and sex (mean age, 63 years in both series; proportion of males 85% and 88% respectively). Therefore, age and sex adjustments were not necessary for comparisons between controls and either or both of the two HCC groups (Trichopoulos et al., 1978).

Serum levels and phenotypes of α₁AT were determined by radial immunodiffusion and

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electrofocusing in acrylamide gel, respectively (Vesterberg, 1973; Chapuis-Cellier, 1975). All determinations were performed blindly in the Department of Clinical Biochemistry of the Hospital “Edouard Herriot” in Lyon, France.

Results

Table I shows the distributions of HCC cases and controls by HBsAg status (positive or negative) and α,AT phenotype. The four distributions are very similar and the observed differences can easily be explained by chance. Thus, comparison of all HCC cases with all controls, with respect to the four most common α,AT phenotypes (and a fifth group combining all the “other” less common ones) gives a χ² with 4 degrees of freedom of 2.56, corresponding to P > 0.5. Furthermore, there is no evidence in the present data that phenotypes associated with α,AT deficiency are over-represented among HCC cases; on the contrary, two heterozygotes for the Z allele were included in the control series whereas none was found among the cases. Lastly, among the cases 6% were homozygous and 14% heterozygous for the M₂ allele; among controls the corresponding figures were almost identical (7% and 11%, respectively) providing no support to the hypothesis that this allele is associated with a substantial increase of HCC risk.

Table II shows mean values of serum α,AT (in mg/100 ml⁻¹) in HCC cases and controls by α,AT phenotype and, for HCC cases, by HBsAg status. The mean value of serum α,AT is higher in HCC cases than in hospital controls (by ~40%) and the difference is highly significant (P < 10⁻⁹). Although HCC cases of both subgroups have elevated α,AT values the elevation is more marked among HBsAg-negative cases than among HBsAg-positive cases; the corresponding mean values are ~50% and 30% higher than the mean value among controls and they differ significantly from each other (P < 0.01) as well as from the mean value of controls (P < 10⁻⁹ and P < 10⁻⁶ respectively). It is of interest that the pattern HCC(HBsAg−) > HCC(HBsAg+) > Controls is evident not only in the total but also within every single α,AT phenotype (with only one marginal exception, in M₁M₃). There was no significant (or even similar) difference in the average level of serum α,AT between HBsAg positive and negative individuals in the control group; the mean values were 489 and 430 mg/100 ml⁻¹ respectively, and the P value for a t-test of their difference is 0.30. Recent reports (Trichopoulos et al., 1980; Lam et al., 1982) indicated that tobacco smoking may cause HBsAg-negative HCC. Since tobacco smokers in general have elevated concentrations of serum α,AT, we have explored whether the very high values of α,AT in HCC (and particularly in HBsAg-negative HCC) could reflect a specific association between α,AT and tobacco-related HBsAg-negative HCC. There is some evidence in the present data that this may be so, but it is far from conclusive. Among patients with HBsAg-negative HCC, markedly elevated concentrations of serum α,AT (>600 mg 100 ml⁻¹) were found in 21/31 current smokers (68%) but only in 5/10 non-smokers (50%); however, the

| AAT phenotype | HCC cases | Controls |
|---------------|-----------|----------|
|               | HBsAg(+)  | HBsAg(-) | Total (%) | HBsAg(+)  | HBsAg(-) | Total (%) |
| M₁M₁          | 20        | 18       | 38 (47)  | 5         | 80       | 85 (53)  |
| M₁M₂          | 1         | 5        | 6 (8)    | 1         | 10       | 11 (7)   |
| M₁M₃          | 13        | 11       | 24 (30)  | 5         | 29       | 34 (21)  |
| M₂M₂          | 3         | 2        | 5 (6)    | 1         | 11       | 11 (7)   |
| M₂M₃          | 1         | 3        | 4 (5)    | 1         | 5        | 6 (4)    |
| M₃M₃          | (0)       |          |          | 5         | 5        | 5 (3)    |
| M₁I           | 1         |          |          | 2         | 2        |          |
| M₁S           | 1         |          |          | 1         | 1        |          |
| M₁F           | 1         |          |          | 1         | 1        |          |
| M₁N₂          | 1         |          | (4)      | 1         | 1        | (5)      |
| M₂P           | 1         |          |          | 1         | 1        |          |
| M₂S           | 1         |          |          | 1         | 1        |          |
| M₃P           | 1         |          |          | 1         | 1        |          |
| M₃S           | 1         |          |          | 1         | 1        |          |
| M₂Z           | 2         |          |          | 2         | 2        |          |
| Total         | 39        | 41       | 80 (100) | 12        | 148      | 160 (100)|
difference is neither statistically significant nor dose-dependent (Table III).

We have also investigated whether, among HCC patients, there is a positive correlation between the serum concentrations of αFP (after log-transformation) and α1AT. There is no such evidence in the present data; among HBsAg-positive HCC patients the correlation coefficient is +0.04, whereas among HBsAg-negative HCC patients the corresponding value is +0.05.

Discussion

The association between α1AT and HCC has been studied from several points of view. Individuals who are homozygous for the Z allele (and perhaps other alleles associated with α1AT deficiency) have an increased risk for HCC, and many of them have the characteristic globular bodies in their livers. However, it is not clear whether individuals who are heterozygous for these alleles are at increased risk for HCC; many studies have noted modest associations but several others have not, even though heterozygotes have consistently the same characteristic globules in their livers (studies reviewed by Sharp, 1976; Kew et al., 1978; Morse, 1978; Kelly et al., 1979; Sizaret et al., 1981; Spech & Liehr, 1982). Kew et al. (1978) have summarized the evidence by stating that α1AT deficiency could be an occasional cause of the tumour in most parts of the world where HCC occurs sporadically, but it is not a numerically important cause in Africa and the Far East where HCC is common. Our findings support this conclusion and permit its generalization to other populations where HCC is common, besides those of Africa and Asia. On the other hand, we did not confirm the associations of HCC with the F or the M2 alleles, which were reported by Theodoropoulos et al. (1976) and Sizaret et al. (1981), respectively. Since the present study is larger than the other two we are inclined to

### Table II

Concentration of serum α1AT in HCC cases (positive and negative for HBsAg) and in controls, by phenotype. Mean values (and standard errors) in mg 100 ml⁻¹

| α1AT phenotype | HCC cases | Controls |
|----------------|-----------|----------|
|                | HBsAg(+)  | HBsAg(−) | Total | Total |
|                | n         | x        | n     | x     | n     | x   |
| M1M1           | 20        | 601      | 18    | 675   | 38    | 636  | 85  |
| M1M2           | 1         | 378      | 5     | 619   | 6     | 579  | 11  |
| M1M3           | 13        | 579      | 11    | 645   | 24    | 609  | 34  |
| M2M2           | 3         | 479      | 2     | 888   | 5     | 643  | 11  |
| M2M3           | 1         | 440      | 3     | 661   | 4     | 605  | 6   |
| M3M3           | 1         | 466      | 1     | 466   | 2     | 578  | 5   |
| other†         | 1         | 466      |       |       |       | 300  |
| other‡         | 2         | 578      | 2     | 578   |       | 312  |
| Total          | 39        | 571      | 41    | 665   | 80    | 619  | 160 |
| (standard error)| (23)     | (26)     | (18)  | (13)  |

†Phenotypes not reported to be associated with α1AT deficiency (M1F, M1N2, M2P, M1P).

‡Phenotypes reported to be associated with α1AT deficiency (M1I, M1S, M2S, M3S, M3Z).

### Table III

Distribution of 41 HCC cases negative for HBsAg by reported smoking habits before the onset of the disease and serum level of α1AT (above or below 600 mg 100 ml⁻¹).

| Serum α1AT | Non (and Ex) smokers | Smokers (cigarettes day⁻¹) | Totalb |
|------------|-----------------------|---------------------------|--------|
|            | 1–10                  | 11–20                     | 21–30  | 31+   | Total |
| 600+       | 5                     | 1                         | 11     | 4     | 5     | 26    |
| <599       | 5*                    | 2                         | 3      | 2     | 3     | 15    |
| Total      | 10                    | 3                         | 14     | 6     | 8     | 41    |

*One of these patients may have been a regular light smoker (repeated interview).

b²for trend (1 d.f.) = 0.5; P > 0.25.
believe that the earlier findings were, perhaps, fortuitous.

Elevated values of serum α1-AT in HCC cases have been noted by Kew et al., (1978), Chio & Oon (1979), and Matsuzaki et al. (1981). Our findings confirm the results of the earlier investigations and indicate that the elevation is sufficiently marked to be aetiologically intriguing and diagnostically useful. Furthermore, we have found that serum α1-AT values are, on the average, higher in HBsAg-negative cases of HCC than in HBsAg-positive cases of this tumour—an observation not reported previously. Although the difference is not sufficiently large to be of clinical importance it suggests that the aetiologic heterogeneity of HCC is reflected not only in the epidemiologic parameters but also in the laboratory findings of the aetiologic subgroups.

The molecular, biochemical and histological aspects of the association between α1-AT and HCC have been reviewed elsewhere (Sharp, 1976; Morse, 1978; Palmer et al., 1980; Spech & Liehr, 1982). However, the differential increase of α1-AT in HBsAg-positive and HBsAg-negative HCC, if real, calls for explanation. Several possibilities exist. Tobacco smoking has been found to increase the serum levels of α1-AT by ~20% (Lellouch et al., 1979) and this proportional excess may also concern the tobacco related HCC cases. It should be noted, also, that cirrhosis is associated with reduced levels of serum α1-AT (Chio & Oon, 1979; Matsuzaki et al., 1981), and cirrhosis (in Greece, at least) is more frequently associated with HBsAg-positive than with HBsAg-negative HCC (Trichopoulos et al., 1982).

Palmer et al. (1980), using ultrastructural, histochemical and immunocytochemical methods have demonstrated the occasional parallel emergence of αFP and α1-AT as tumour tissue markers in malignant hepatoma (HCC). It is, therefore, surprising that a significant positive association was not evident between the serum values of αFP and α1-AT in the present study or in the earlier study of Chio & Oon (1979). Although chance is a likely explanation for the absence of a significant association, it is also possible that the epigenetic emergence of α1-AT as tumour tissue marker is not frequently accompanied by an increase in the serum levels of α1-AT (Palmer et al., 1980).

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