Abstract

Seaweeds are marine organisms capable of producing diverse kinds of chemical compounds with promising pharmacological use. The study evaluated the proximate and elemental composition and the potential antioxidant (using CUPRAC, ABTS\(^+\), and DPPH assays) and antibacterial activities (using microtiter plate dilution assay) of Ulva prolifera O. F. Müller. The seaweed has a total phenolic content of 829 ± 2.00 mg GAE/g. Antioxidant efficiency of U. prolifera exerted high ability of reducing copper ions, potent ABTS\(^+\) and DPPH scavenging activities in a concentration-dependent manner with IC\(_{50}\) value of 24.7 mg GAE/mL, 43.52 mg GAE/mL, and 54.1 mg GAE/mL, respectively, more effective than ascorbic acid. In vitro antibacterial activity assay showed that U. prolifera exhibited inhibitory activity against bacterial pathogens such as Methicillin-resistant Staphylococcus aureus (MIC = 125 mg/mL), S. epidermidis (MIC = 125 mg/mL), and Pseudomonas fluorescens (MIC = 500 mg/mL). This investigation is the first documented report regarding antibacterial activity of U. prolifera against P. fluorescens. In addition, results showed elemental composition to be in decreasing order of Ca > K > Mg > Na > Mn > Fe > Zn > Cu > Pb > Cd > Cr. Proximate composition of U. prolifera showed high carbohydrate and protein content with a percentage composition of 36.20 ± 0.27% and 23.72 ± 0.31% (dry weight), respectively. This study is the first report in the Philippines that shows the potential of U. prolifera as an excellent candidate organism as source of chemical compounds with relevant application to the pharmacological industry.

Keywords: antioxidant activity, nutritional composition, phenolic content, polyphenols, seaweed

Introduction

The demand of human society for a healthy lifestyle and increased longevity has led people (consumers) to consume functional foods with plentiful sources of chemical compounds and other nutraceuticals. Of these well-known natural foods, seaweeds are considered as one of the most valuable natural resources for macronutrients (carbohydrates, proteins and lipids), micronutrients (vitamins and minerals), and other significant chemical substances for animal and human health (Corsetto et al., 2020; Cotas et al., 2020; Arguelles, Monsalud, & Sapin, 2019; Arguelles, Laurena, Monsalud, & Martinez-Goss, 2018; Sivaramakrishnan et al., 2017). Recent studies shows that several unique compounds from seaweeds, such as acrylic acid, brominated phenols, heterocyclic carbons, polysaccharides, terpenes, peptides, chlorophyllides and sterols, were isolated and being developed as new active ingredients for pharmaceuticals (Cotas et al., 2020; Yuan et al., 2018; Yaich et al., 2011). These substances have diverse biological and chemical activities like antimicrobial, anticancer, anti-diabetic, anti-inflammatory, antioxidant and anti-hypertensive properties that can be used to discover novel drugs for pharmaceutical application (Kordjazi, Etemadian, Shabanpour, & Pourashouri, 2019; Sivaramakrishnan et al., 2017).

Macroalgae are also regarded as popular food ingredients and are widely consumed in several countries. However, the nutritional profiles of these organisms have not been fully elucidated since only a few studies on the nutritional composition of seaweeds have been conducted (Arguelles et al., 2019). Edible seaweeds abound insoluble proteins, dietary fibers or
carbohydrates, minerals, polyunsaturated fatty acids, and vitamins (Wells et al., 2017; Senapati et al., 2016). Several studies reported that the protein concentration of dried seaweed biomass differs from 30-40% (Rhodophyta) to around 5-10% (Phaeophyta) both of which have low lipid content, 1-5% of the dry weight algal biomass. Also, these organisms are an excellent alternative source of minerals (36% of its total dry biomass) and high fiber content (3-50% of total dry biomass) (Senapati et al., 2016; Wells et al., 2017; Matanjun, Mohamed, Mustapha, & Muhammad, 2008). The biochemical compositions of seaweeds vary in different species. These variations are caused by several factors such as seasonal changes, environmental factors (salinity, minerals, water temperature, nutrients, and light), and geographical differences (Haq et al., 2019; Wells et al., 2017; YaiCh et al., 2011). The genus Ulva (Chlorophyta) has high carbohydrates, proteins, dietary fiber, vitamins, and minerals, making this group of seaweeds a significant natural food source for commercial production. These macroalgae are also being tapped as potential value-added food because of reported health benefits such as therapeutic bioactive peptides and prebiotics (dietary fiber) source as well as other functional properties such as anti-diabetic, antimicrobial, and antioxidant activities (Manandhar et al., 2019; Arguelles et al., 2018; Wells et al., 2017; Yuan et al., 2018).

Seaweeds are renewable natural resources that grow abundantly in different coastal areas (islands and reefs) of the Philippines. U. prolifera O. F. Müller is a green seaweed that is classified to the order Ulvales and family Ulvaceae. It is a bloom-forming algae usually associated with the occurrence of the green tide phenomenon in some coastal areas of the Philippines. Previously, this seaweed was reported to have bioactive compounds such as polyphenols, peptides, chlorophyll, and carotenoids. These compounds exhibit important chemical properties useful for nutraceutical, pharmacological industry, and functional food for humans (Corsetto et al., 2020; Cotas et al., 2020; Haq et al., 2019; Arguelles et al., 2018; Zhao, Yang & Liu, 2016). U. prolifera is edible and is considered an economically viable food option in Japan and China. The algae is nutritious and regarded as low-calorie food rich in essential amino acids, fatty acids, vitamins, dietary fiber, and proteins. It also contains chlorophyll b and different minerals, such as calcium, magnesium, and iron. Polysaccharides are the major component of U. prolifera. The use of these carbohydrate polymers expands from food and pharmaceutical industries (Haq et al., 2019; Zhao, Yang & Liu, 2016). This investigation is the first documented report in the Philippines showing the bioactive properties of a green tide-causing macroalga, U. prolifera, with potential use as natural antioxidants, control of bacterial diseases and as an alternative source of minerals for nutraceutical application. The study was conducted to evaluate the proximate and elemental composition of the dried algal biomass. In addition, the total polyphenolic content (TPC), antibacterial and antioxidant (using copper reduction antioxidant capacity (CUPRAC), 2,2′-Azino-bis (3-ethylbenzothiazoline-6-sulfonic acid) (ABTS+), and 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging assays) activities of U. prolifera were analyzed. Furthermore, correlation analysis between phenolic concentration and antioxidant property of the seaweed extract was established.

Material and Methods

Chemicals

The chemicals used were of analytical grade and obtained from Sigma–Aldrich (Steinheim, Germany), Sigma–Aldrich (Singapore City, Singapore) or Sigma–Aldrich (St. Louis, MO, USA). Meanwhile, the solvents used were of HPLC grade and analytical grade and obtained from Sigma–Aldrich (Steinheim, Germany, St. Louis, MO, USA and Singapore City, Singapore) or Merck (Darmstadt, Germany).

Seaweed Sampling and Preparation

The green macroalga, U. prolifera, was gathered on January 18th, 2020 during low tide conditions alongside the coast of Pagkalitan (Lat. 13° 38’ 4.2” N; Long. 121° 2’ 39” E), Batangas, Philippines. The collected sample was initially cleaned with seawater to remove sand debris and other loosely attached organisms. The seaweed was immediately transferred to the laboratory using sterilized polythene bags kept in a cooler with ice. The macroalga was washed several times with distilled water and scrubbed thoroughly using brush bristles to remove the seaweed’s epiphytes, sand particles, and necrotic parts. U. prolifera was air-dried under direct sunlight for six days and pulverized (using a dry grinder) before extraction (Arguelles et al., 2019). The pulverized samples were sieved to get the uniform size of seaweed particles, then placed in an air-tight glass container and kept in a freezer (set at -20°C) until the needed chemical assays were conducted. The seaweed was identified using morphological characteristics and identification keys of Trono (1997) and Algae Base (web site: www.algaebase.org) (Guiry & Guiry, 2020).

Preparation of Seaweed Extract

The powdered U. prolifera (1 g) was extracted with 30 mL acidified methanol (1 HCl:80 CH_3OH:10
H₂O) in an ultrasonic bath for 30 min and stirred for 1 h. The extraction mixture was centrifuged for 20 min at a speed of 12,000 rpm at 20 °C. U. prolifera extract was concentrated under reduced pressure set at 40 °C using a rotary evaporator. The concentrated extract was kept at 4 °C to maintain its activity in other biological/chemical assays needed in the study (Arguelles, 2021).

**Proximate Composition Analysis**

U. prolifera biomass was dried for 24 h at 60 °C and then grind into a powder prior to proximate composition analysis using standard methods. The ash, protein (N × 4.38), crude fiber, and fat contents were determined using protocols set by the AOAC (2011). The total carbohydrate content was obtained via the difference method following the equation below:

\[
\%\text{Carbohydrates} = 100 - (\%\text{Moisture Content} + \%\text{Protein} + \%\text{Fat} + \%\text{Ash})
\]

**Elemental Composition Analysis**

The powdered biomass of U. prolifera was subjected to elemental composition analysis using the standard protocols (AOAC, 2011). The analysis was done to detect and quantify magnesium (Mg), cadmium (Cd), potassium (K), chromium (Cr), calcium (Ca), zinc (Zn), manganese (Mn), iron (Fe), sodium (Na), copper (Cu), and lead (Pb) using an Atomic Absorption Spectrophotometer Perkin Elmer AAnalyst 400.

**Determination of Total Phenolic Content (TPC)**

The TPC of U. prolifera extract was assessed following the Folin-Ciocalteu procedure of Nuñez Selles et al. (2002). Briefly, 0.5 mL of U. prolifera extract was thoroughly mixed in equal volume with 10% sodium carbonate solution and Folin-Ciocalteau’s reagent for 1 min. The mixture was then kept at ambient temperature for about 5 min. The reaction mixture’s final volume was increased to 5 mL using sterile water. The reaction mixture’s optical density reading (OD) was noted at 720 nm using an Ultraviolet-Visible spectrophotometer. The TPC is presented as a microgram of gallic acid equivalent (GAE) per gram of the algal sample (standard curve equation: \( y = 0.0682x - 0.0214, r^2 = 0.997 \)).

**Antioxidant Activities**

Polyphenolic compounds from seaweeds are known to have diverse antioxidant and biological activities. The antioxidant properties of U. prolifera extract were determined using radical scavenging assays for DPPH, ABTS⁺, and CUPRAC assays. Three methods of antioxidant assays were used in the investigation to elucidate the different mechanisms responsible for the antioxidant activities of U. prolifera extract.

**ABTS⁺ (2,2’-Azino-bis (3-ethylbenzothiazoline-6-sulfonic acid) Scavenging Assay**

The potential of U. prolifera extract to scavenge ABTS⁺ free radical was determined using a method based on the decolorization of ABTS⁺ (Re et al., 1999). In this assay, radical cations (ABTS⁺) are generated by the reaction of sodium persulfate (bluish) and ABTS⁺. These radical cations are reactive in several antioxidants (including polyphenols) and convert the ABTS⁺ to its neutral (colorless) form. An aliquot (40 \( \mu \)L) of different prepared concentrations of U. prolifera extract (12.5–62.5 \( \mu \)g GAE/mL) and 40 \( \mu \)L of 90% methanol (for the control) were homogenized with 3 mL of ABTS⁺ radical mixture (with initial OD reading of 0.72 ± 0.05 at 734 nm). The solutions were thoroughly mixed and kept at ambient temperature for 5 min. The absorbance reading of each sample was noted at 734 nm using water to zero the spectrophotometer (Re et al., 1999). The ABTS⁺ inhibition (%) was estimated using the following formula:

\[
\text{ABTS Inhibition(\%)} = \left( \frac{A_{\text{control}} - A_{\text{sample}}}{A_{\text{control}}} \right) \times 100
\]

The percentage of ABTS⁺ inhibition (%) was marked against separately prepared concentrations of U. prolifera extract. The prepared concentration of the seaweed extract that exhibited 50% ABTS radical scavenging activity was considered as the \( IC_{50} \). The standard antioxidant used in this assay is ascorbic acid (\( IC_{50} = 147.9 \mu \)g/mL).

**2,2-diphenyl-1-picrylhydrazyl (DPPH) Radical Scavenging Assay**

The scavenging activity of U. prolifera extract against DPPH free radical was analyzed following the protocol done by Ribeiro, Barbosa, Queiroz, Knodler, and Schieber (2008). Briefly, 100 \( \mu \)L of the U. prolifera extract (10.0 – 50.0 \( \mu \)g GAE/mL) was added into 5.0 mL of 0.1 mM DPPH methanolic solution. It was then homogenized using a vortex mixer and set aside for 20 min at ambient temperature. The optical density (OD) readings of the U. prolifera extract and the controls were noted at 517 nm using a UV-VIS spectrophotometer. The percent DPPH radical scavenging inhibition (%) was estimated using the following formula:

\[
\text{DPPH Inhibition(\%)} = \left( \frac{A_{\text{control}} - A_{\text{sample}}}{A_{\text{control}}} \right) \times 100
\]
where: \( A_{\text{control}} \) = optical density of the control (DPPH solution without algal extract) and \( A_{\text{sample}} \) = optical density of the algal sample (DPPH solution plus algal extract). Gallic acid was used as the control in this assay. The effective concentrations (IC \( 50 \)) of the \( U. \) prolifera extract, as well as the control needed to neutralize 50\% of DPPH free radicals, were analyzed using the standard curve obtained from scavenging capacities and various concentrations of algal extract and is expressed in \( \mu \)g/mL (Arguelles et al., 2019).

**Copper Reduction Antioxidant Capacity (CUPRAC) Assay**

The antioxidant activity of \( U. \) prolifera extract (5.0 – 25.0 \( \mu \)g GAE/mL) to reduce cupric ions (\( \text{Cu}^{2+} \)) was done following the method proposed by Alpinar et al. (2009). In this antioxidant assay, 1 mL each of 1 M ammonium acetate buffer (pH 7), 0.0075 M neocuproine, and 0.01 M \( \text{CuCl}_2 \) solutions were mixed in prepared sterile test tubes containing 0.5 mL of \( U. \) prolifera extract (5.0, 10.0, 15.0, 20.0 and 25.0 \( \mu \)g GAE/mL) and ascorbic acid as the standard antioxidant (Arguelles, 2021). The total volume for each reaction mixtures prepared was adjusted to 4.1 mL using sterile distilled water and kept for 30 min at ambient temperature. The absorbance readings (OD) of \( U. \) prolifera extract and ascorbic acid concentrations were recorded at 450 nm (Arguelles, 2021).

**Antibacterial Activity**

The National Institute of Molecular Biology and Biotechnology (BIOTECH), University of the Philippines Los Baños (UPLB) provided the test organisms used in this investigation. A total of three Gram-negative bacteria (\( \text{Aeromonas hydrophila} \) BIOTECH 10089, \( \text{Escherichia coli} \) BIOTECH 1825, and \( \text{Pseudomonas fluorescens} \) BIOTECH 1123) and three Gram-positive bacteria (\( \text{Staphylococcus epidermidis} \) BIOTECH 10098, \( \text{Methicillin-Resistant Staphylococcus aureus} \) BIOTECH 10378 and \( \text{Bacillus cereus} \) BIOTECH 1509) were tested against \( U. \) prolifera extract using the microtiter plate dilution assay. The reference bacterial pathogens were pre-cultivated using Luria Bertani (LB) Broth medium and kept at 37 \(^\circ\)C with shaking for 24 h. To maintain the cultures, the purity and viability for each bacterial pathogens are regularly monitored via morphological characterization and biochemical tests every week (Arguelles, 2021).

A two-fold serial dilution technique was used to establish the minimum bactericidal concentration (MBC) and minimum inhibitory concentration (MIC) of \( U. \) prolifera extract (Arguelles, 2021). Briefly, 100 \( \mu \)L of test bacterial pathogens (with a corresponding cell density of 1 x 10\(^6\) cells/mL) were combined with 100 \( \mu \)L of \( U. \) prolifera extract set in different dilutions (1000 \( \mu \)g/mL-7.8125 \( \mu \)g/mL). The antibacterial assay was done in triplicate using a 96-well microtiter plate and kept for 12 h in an incubator set at 35\(^\circ\)C. After incubation, MICs of \( U. \) prolifera extract, as opposed to the tested bacterial pathogens, were noted. Streptomycin was used as the positive control against the tested pathogenic bacteria. Minimum bactericidal concentration (MBC) was determined following the protocol done by Arguelles (2021) with few modifications. Loopful of samples in microtiter plate wells that exhibited no bacterial growth from the MIC assay were inoculated onto tryptic soy agar plates and were kept at 35\(^\circ\)C for 24 h. After incubation, the petri plates were evaluated for bacterial colony growth for each dilution subculturing. The absence of colony growth would mean that \( U. \) prolifera extract was bactericidal at that specific dilution (Arguelles, 2021).

**Statistical Analyses**

The experimental data are presented as means ± standard deviations (mean ± SD) of three assay replicates. The statistical tests for the linear correlation coefficient necessary for the correlation analysis were analyzed using MS Office Excel 2007.

**Results and Discussion**

**Proximate Composition Analysis**

Algae are valuable sources of micronutrients such as vitamins and minerals and macronutrients such as carbohydrates, protein, lipids, and fiber, directly relevant to human health (Arguelles & Martinez-Goss, 2021; Arguelles et al., 2018; Osuna-Ruiz et al., 2019). These organisms are considered good alternative nutrient sources of protein, lipids and carbohydrates that can enrich the traditional plant-based sources for human consumption (Ismail, 2017). Seaweeds are good alternative sources of proteins and lipids that can be used in animal feed formulation and the development of highly nutritious foods that can replace current crop protein sources (such as bean) consumed by humans. These organisms can enhance the nutritional quality of human foods and animal feeds while at the same time reducing the cost of production. In addition, seaweed-based food and feeds also improve humans’ immunity and overall health using bioactive polysaccharides and other important chemical constituents that possess important biological activities such as antibacterial, anti-inflammatory, and antiviral activities. Seaweeds are also considered alternative sources of important microelements because of the high concentration of minerals in the algal biomass. These micronutrients can be utilized by humans and
other animals as food additives for food and industrial applications (Arguelles & Martinez-Goss, 2021; Alwaleed, 2019; D’Armas, Jaramillo, D’Armas, Echavarria, & Valverde, 2019). The proximate composition of *U. prolifera* is presented in Table 1. The seaweed contains a high carbohydrate, protein, and lipid concentration with a percent composition of 36.20 ± 0.27% and 23.72 ± 0.31%, 12.50 ± 0.11%, respectively. Such results prove that *U. prolifera* is a good source of carbohydrate, protein, and lipids that can be utilized as a functional food for humans. Carbohydrate content of *U. prolifera* is less than those obtained from other seaweed species (dry biomass) such as *Sargassum polycystum* (90.01%), *Digenea simplex* (42.40%) and *Cystoseira myrica* (78.7%) (Alwaleed, 2019). On the other hand, the crude fat content of *U. prolifera* is higher than those obtained by Raja et al. (2020) from *Gracilaria corticota* and *Halimeda opuntia* with a percent composition of 10.09 ± 0.06% and 7.26 ± 0.048%, respectively. Differences in lipid and carbohydrate contents among seaweeds can be due to factors like the effects of climate and geography on the development and growth of seaweeds (Ismail, 2017; Matanjun et al., 2008). The crude protein content of *U. prolifera* is greater than those obtained by Osuna-Ruiz et al. (2019) from tropical seaweeds of Sinaloa, Mexico which includes *U. prolifera* (7.85 ± 0.90%), *Codium isabelae* (8.90 ± 0.06%), *Rhizoclonium riparium* (7.85 ± 0.90%), and *Spyridia lamentosa* (10.61 ± 0.34%). Variations in the protein content among different groups and species of seaweeds are caused by differences in the protein levels present on the algal tissue that is highly dependent on the concentration of nutrients (nitrogen) in seawater (D’Armas et al., 2019).

Table 1. Proximate composition of *U. prolifera* dry biomass.

| Proximate composition | Percent composition (%) |
|-----------------------|-------------------------|
| Moisture Content      | 4.66 ± 0.20             |
| Ash Content           | 15.89 ± 0.04            |
| Crude Protein         | 23.72 ± 0.31            |
| Crude Fat             | 12.50 ± 0.11            |
| Crude Fiber           | 6.03 ± 0.15             |
| Carbohydrate          | 36.20 ± 0.27            |

Table 1 showed other dominant biochemical components like ash (15.89 ± 0.04%) and crude fiber (6.03 ± 0.15%). The ash content shows the concentration of inorganic substances in seaweeds. This parameter is influenced by the existing minerals and salt attached to the macroalgae. *U. prolifera* are rich in minerals which are beneficial for feed formulation in animals. On the other hand, crude fiber represents the concentration of indigestible components of the macroalgae. In this study, low concentration of crude fiber was observed in *U. prolifera* and is comparable to that obtained by D’Armas et al. (2019) from other species of seaweeds such as *Kappaphycus alvarezii* (3.18 ± 0.11%), *Padina pavonica* (4.94 ± 0.16%), *Ulva lactuca* (0.96 ± 0.03), *Centrocercus clavulatum* (2.02 ± 0.11%), and *Acanthophora spicifera* (2.42 ± 0.24%).

**Elemental Composition Analysis**

Essential minerals for human consumption include sulfur, potassium, phosphorus, sodium chloride (salt), calcium, iron, manganese, and magnesium. Also, trace elements such as copper, cobalt, iodine, zinc, molybdenum, and selenium are essential but should be consumed by humans in regulation and not in excessive amounts (Arguelles, 2021; D’Armas et al., 2019; Matanjun et al., 2008). The result of the elemental composition analysis of *U. prolifera* is shown in Table 2. The elemental distribution in *U. prolifera* was observed to be in decreasing order of Ca > K > Mg > Na > Mn > Fe > Zn > Cu > Pb > Cd > Cr. High concentration of calcium (Ca) (20.18 ± 32.91 ppm) followed by magnesium (Mg) (14.98 ± 365 ppm), sodium (Na) (9.863 ± 0.17 ppm), and manganese (Mn) (533.47 ± 2.77 ppm) were observed in the seaweed biomass. The high concentration of these elements in *U. prolifera* biomass is comparable to that observed by Akköz, Arslan, Unver, Özcan, and Yılmaz (2011) in *Enteromorpha intestinalis* (a green seaweed) for calcium (Ca) (22.818 ± 42.38 ppm), magnesium (mg) (15.764 ± 132.36 ppm), sodium (Na) (8.690.27 ± 69.38 ppm), and manganese (Mn) (691.33 ± 11.32 ppm). This study demonstrates that *U. prolifera* can be used as an alternative source of important microelements (for human use) because of the high concentration of essential minerals present in the algal biomass. In addition, low concentration of other trace elements such as iron (Fe) (251.91 ± 0.25 ppm), copper (Cu) (16.57 ± 0.18 ppm), zinc (Zn) (18.15 ± 0.45 ppm), chromium (Cr) (1.74 ± 0.13 ppm), lead (Pb) (1.37 ± 0.25 ppm), and cadmium (Cd) (1.01 ± 0.14 ppm) was observed from the algal biomass. Such observation can be used to indicate the low occurrence of heavy metal pollution in the sampling area. This concentration level in *U. prolifera* biomass is favorable for human consumption since these elements are only required in minimal concentrations (D’Armas et al., 2019; Alfonso et al., 2018).
Normally, the elemental composition of green macroalgae has high levels of Mg, K, Na, and Ca, with reported concentrations greater than those obtained from other groups of seaweed (Alfonso et al., 2018; Akköz et al. 2011). Also, previous study observed that green seaweeds such as U. lactuca are also rich in Fe with an estimated concentration of 41 mg/100 g or 410 ppm (Yaich et al., 2011). These observations are in accordance with what was observed in the current study regarding the elemental composition of U. prolifera. Variations in the concentration of elemental composition from within species and strains of the same seaweed group usually occur depending on the environment where the seaweed is exposed. Seaweeds are capable of bio-accumulating elements from their surrounding environment. Thus, concentration of elements in the seaweed tissue may be controlled by the quality and concentration of the elements present in seawater (Alfonso et al., 2018).

### Total Phenolic Content (TPC)

The TPC of U. prolifera extract was analyzed using the Folin-Ciocalteu reagent and reported as gallic acid equivalents (standard calibration curve used has an equation of y = 0.0682x - 0.0214, r² = 0.997). The TPC of the seaweed extracts is 829 ± 2.00 μg GAE/g. The concentration of polyphenolic compounds observed in this study is greater than that obtained by Akköz et al. (2011) from ethanol extracts of E. intestinalis and Cladophora glomerata with 32 ± 0.003 μg GAE/g and 47 ± 0.001 μg GAE/g, respectively. Also, TPC of U. prolifera is comparable to those observed by Nunes, Ferraz, Valente, Barreto, and Pinheiro de Carvalho (2017) from other seaweed species such as U. lactuca, Chondrus crispus, Asparagopsis taxiformis, Nemalion elminthoides, and Grateloupia lanceola obtained from Madeira archipelago with TPC content of 556.1 μg GAE/g, 362.8 μg GAE/g, 576.3 μg GAE/g, 655.2 μg GAE/g, and 257.9 μg GAE/g, respectively. The type of solvent influences the concentration of polyphenolic compounds that can be extracted from a sample. Generally, polar organic solvents, such as acidified methanol, exhibit greater capacity to extract polyphenolic compounds in seaweed samples than water (Arguelles, 2021; Nunes et al., 2017). Also, other factors such as differences in the geographic location of the sampling site, growth stage (of the seaweed), seasonal variation and genetic diversity can cause variation in the concentration of polyphenolic compounds present in seaweed sample (Mekinie, 2019; Arguelles et al., 2019; Nunes et al., 2017).

### Antioxidant Activities

The diversity of polyphenolic compounds in seaweeds and other structural features plays a crucial role in the reaction mechanism by which these macroalgae can exhibit antioxidant properties. In this study, the antioxidant activities of U. prolifera extract from Pagkalitan, Batangas were analyzed using three methods: DPPH and ABTS⁺ radical scavenging assays as well as CUPRAC assay. These antioxidant assays were used in this study to understand better the mechanisms involved in U. prolifera antioxidant activities. Previous study has also documented the antioxidant activities of several strains of Philippine seaweeds (Arguelles, 2021; Arguelles et al., 2019); however, this study documented the first report on the antioxidant activities of U. prolifera found in the coastal areas of Pagkalitan, Batangas in the Philippines.

### ABTS⁺ (2,2’-Azino-bis (3-ethylbenzo-thiazoline-6-sulfonic acid) Scavenging Activity

ABTS⁺ antioxidant assay is an excellent method in determining the antioxidant property of chain-breaking antioxidants (lipid peroxyl radical scavenger) and natural antioxidants capable of hydrogen donation (aqueous phase radical scavenger) (Liu et al., 2018; Sivaramakrishnan et al., 2017). The ABTS⁺ scavenging activity and IC₅₀ value of U. prolifera extract is shown in Table 3. ABTS⁺ scavenging activity against U. prolifera extract demonstrated that the maximum antioxidant activity was 56.41 ± 1.32% at phenolic concentration of 62.50 mg GAE/mL. Further, the result of the antioxidant assay shows that the scavenging activities of U. prolifera extract increased with the concentration of polyphenolic compounds in the algal extract (12.5 - 62.50 μg GAE/mL). The computed IC₅₀ value of the algal extract is 54.10 μg/mL, which is more potent than that obtained from standard antioxidant, ascorbic acid (IC₅₀, 147.9 μg/mL). Lower IC₅₀ concentrations are indicative of high antioxidant activity. In collation with the IC₅₀ of other macroalgae

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**Table 2. Concentrations of macro and micro-elements (ppm) of U. prolifera O.F. Müller**

| Seaweed   | Ca     | Mg     | Na   | Mn   | K     | Fe   | Zn   | Cu   | Pb   | Cr   | Cd   |
|-----------|--------|--------|------|------|-------|------|------|------|------|------|------|
| U. prolifera | 22.818.994±.42.38 | 15.764±.132.36 | 9.863±.017 | 533.47±.2.77 | 13.989±.1.188 | 251.91±.0.25 | 18.15±.0.45 | 16.57±.0.18 | 1.37±.0.25 | 1.74±.0.13 | 1.01±.0.14 |

* All values are reported as mean ± standard deviation (n = 3)
Table 3. ABTS* radical scavenging activity and IC$_{50}$ value of phenolics from U. prolifera and ascorbic acid.

| Phenolic concentration (μg/mL) | ABTS Inhibition (%) | Concentration (μg/mL) | ABTS Inhibition (%) |
|-------------------------------|---------------------|-----------------------|---------------------|
| U. prolifera                  | Ascorbic Acid       |                       |                     |
| 12.5  | 14.70 ± 0.10         | 37.5                  | 11.70 ± 0.54        |
| 25.0  | 25.22 ± 0.06         | 75.0                  | 24.56 ± 0.62        |
| 37.5  | 35.81 ± 0.10         | 112.5                 | 36.70 ± 0.51        |
| 50.0  | 46.90 ± 0.92         | 150.0                 | 50.87 ± 0.82        |
| 62.5  | 56.41 ± 1.32         | 187.5                 | 64.53 ± 1.64        |
| IC$_{50}$*                     | 54.10 μg/mL         | IC$_{50}$*             | 147.9 μg/mL         |

*IC$_{50}$ is the effective concentration that inhibits the activity of ABTS (2,2-Azino-bis (3-ethylbenzothiazoline-6-sulfonic acid) diammonium salt) cation radical by 50%. Computed by interpolation.

Table 4. DPPH free radical scavenging activity and IC$_{50}$ value of phenolics from U. prolifera and ascorbic acid.

| Phenolic concentration (μg GAE/mL) | DPPH Inhibition (%) | Concentration (μg/mL) | DPPH Inhibition (%) |
|-----------------------------------|---------------------|-----------------------|---------------------|
| U. prolifera                      | Ascorbic Acid       |                       |                     |
| 10.0                             | 9.22 ± 0.00         | 20.0                  | 16.64 ± 0.00        |
| 20.0                             | 19.29 ± 0.06        | 40.0                  | 34.11 ± 0.61        |
| 30.0                             | 34.88 ± 0.11        | 60.0                  | 50.34 ± 0.00        |
| 40.0                             | 46.11 ± 0.23        | 80.0                  | 67.32 ± 0.61        |
| 50.0                             | 57.18 ± 0.23        | 100.0                 | 83.58 ± 0.67        |
| IC$_{50}$                        | 43.52 μg/mL         | IC$_{50}$*             | 59.6 μg/mL          |

*IC$_{50}$ is the effective concentration that inhibits DPPH free radical by 50%. Computed by interpolation.

such as Halimeda tuna (IC$_{50}$ 16.1 μg/mL) and Acetabularia acetabulum (IC$_{50}$ 6.3 μg/mL), U. prolifera had a comparatively potent antioxidant activity (Sivaramakrishnan et al., 2017). The results suggested that U. prolifera extract may contain phenolic compounds that serve as antioxidants capable of inhibiting oxidation through free radical scavenging activity.

2,2-diphenyl-1-picrylhydrazyl (DPPH) Radical Scavenging Activity

Polyphenols are capable of scavenging DPPH by electron donation or hydrogen atom provision and transforming it to a colorless product (2, 2-diphenyl-1-hydratine), causing a decrease in optical density of the sample extract at 517 nm (Liu et al., 2018; Sivaramakrishnan et al., 2017). As the result obtained in ABTS* antioxidant activity assay, the scavenging activity against DPPH free radical also increased when the concentration of U. prolifera extract (10-50 μg/mL) increases (Table 4). The IC$_{50}$ value of the seaweed extract (43.52 μg/mL) exhibited greater antioxidant activity than that obtained from the control antioxidant, ascorbic acid, with IC$_{50}$ of 59.6 μg/mL. In collation with IC$_{50}$ of other seaweeds obtained from Nain Village, Manado, North Sulawesi (Indonesia) such as G. salicornia (12.81 μg/mL), Halymenia durvillei (14.17 μg/mL), Halimeda macroloba (18.54 μg/mL), Turbinaria decurrens (10.01 μg/mL), and Sargassum olygocystum (15.38 μg/mL), U. prolifera had a comparatively less potent DPPH antioxidant activity (Sanger, Rarung, Kaseger, Assa & Agustin, 2019). Seaweeds’ antioxidant activity may be associated with polyphenols such as phloroglucinol, vanillic acids, quinin acids, phenylethanol and dieckol in the U. prolifera extract. These active substances are known to have scavenging free radical ability (Grina et al., 2020; Mekiniae et al., 2019). Also, other biotic and abiotic factors such as strains of the alga and seasonal variation have a significant effect on the synthesis of phenolic compounds and thus affecting the antioxidant properties of the seaweed (Mekiniae et al., 2019; Yuan et al., 2018; Liu et al., 2018).

Copper Reduction Antioxidant Capacity (CUPRAC)

CUPRAC assay was used in this study to assess the capacity of U. prolifera extract to inhibit oxidation via metal chelation mechanism. In this assay, cupric ions are reduced to become cuprous by an electron released by the action of an antioxidant. The cuprous ion creates a stable complex compound with neocuproine, which exhibits maximum absorbance at 450 nm. The extract of U. prolifera exhibited a dose-dependent copper ion reduction ability (Table 5). The maximum absorbance of 0.506 was observed at 25μ GAE/mL concentration of the algal extract.

Table 5. Copper reduction antioxidant capacity (CUPRAC) and IC$_{50}$ value of phenolics from U. prolifera and ascorbic acid.

| Phenolic concentration (μg GAE/mL) | CUPRAC value (Absorbance at 450 nm) | Concentration (μg/mL) | CUPRAC value (Absorbance at 450 nm) |
|-----------------------------------|------------------------------------|-----------------------|------------------------------------|
| U. prolifera                      | Ascorbic Acid                       |                       |                     |
| 5.00                              | 0.15 ± 0.04                        | 10.00                 | 0.11 ± 0.02                        |
| 10.00                             | 0.23 ± 0.11                        | 20.00                 | 0.23 ± 0.07                        |
| 15.00                             | 0.33 ± 0.01                        | 30.00                 | 0.33 ± 0.04                        |
| 20.00                             | 0.42 ± 0.00                        | 40.00                 | 0.43 ± 0.12                        |
| 25.00                             | 0.51 ± 0.01                        | 50.00                 | 0.54 ± 0.11                        |
| IC$_{50}$*                        | 24.70 μg/mL                        | IC$_{50}$*             | 46.30 μg/mL                       |

*IC$_{50}$ is the effective concentration that gives CUPRAC value of 0.5 absorbance reading at 450 nm. Computed by interpolation.
DPPH, ABTS$^+$ and CUPRAC assays with correlation coefficients (R) of 0.9979, 0.9997, and 0.9995, respectively. This finding would mean that the strong antioxidant property of the seaweed extract is caused by polyphenols present in *U. prolifera*. This correlation is similar to those observed from previous studies from other seaweeds such as *Fucus spiralis*, *S. vulgare*, *Cystoseira stricta*, and *Chaetomorpha crasssa* on the relationship of phenolic concentration and their antioxidant activities (Grina et al., 2020; Arguelles et al., 2019; Gazali et al., 2019). The significant correlation between antioxidant activity and phenolic concentration suggests that *U. prolifera* extracts can cause oxidation inhibition via metal chelation and free radical scavenging mechanisms. Isolation, identification, and mass production of these active compounds are needed to maximize the potential use of these natural resources for pharmacological studies (Arguelles et al., 2019).

**Antibacterial Activity**

Seaweeds have diverse bioactive compounds in response to stress caused by environmental pressure. These compounds, such as polyphenols, terpenes, carotenoids, and alkaloids, possess biological activities that are significant in discovering novel antibiotics needed in the pharmacological industry (Manandhar et al., 2019; Sameeh, Mohamed, & Elazzazy, 2016). The potential of a green-tide causing species of seaweed, *U. prolifera*, for antimicrobial activity against six pathogenic bacteria was done in this study. The concentrated crude extract of *U. prolifera* was subjected to *in vitro* antibacterial (microtiter plate dilution) assay against six medically important bacterial pathogens. *U. prolifera* exhibited an extended spectrum of inhibitory activity against *S.epidermidis* and Methicillin-Resistant *S. aureus* (MRSA), each with MIC and MBC values of 125 and 250 µg/mL, respectively (Table 6). This result is more potent than that observed for streptomycin (standard antibiotic drug) used in the study against MRSA and *S. epidermidis* where MIC and MBC values of 250 and 500 µg/mL, respectively, were noted. The antibacterial activity of *U. prolifera* against MRSA is comparable to that observed by Sameeh et al. (2016), where ethanol extracts of *P. boergesenii*, *S. cinereum*, and *Cystoseira myrica* with MIC value of 25, 50 and 50 µg/mL, respectively (El-Sheekh, Mousa, & Farghl, 2020). Also, *U. prolifera* was able to inhibit *Pseudomonas fluorescens* showing MIC and MBC values of 500 µg/mL and 1000 µg/mL, respectively. A

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**Correlation Analysis**

The correlation analysis and its corresponding coefficients among different phenolic concentrations and antioxidant properties of *U. prolifera* are presented in Figures 1-2. The algal extract’s phenolic content strongly correlated with antioxidant activities using assay is similar to those obtained from the ABTS$^+$ and DPPH assay in which 62.5 µg GAE/mL and 50.0 µg GAE/mL concentrations exhibited the greatest antioxidant activity. The IC$_{50}$ value of *U. prolifera* extract is 24.70 µg/mL which is more potent than that obtained from ascorbic acid with IC$_{50}$ 46.30 µg/mL. This result is relatively comparable to that observed by Gazali, Zamani, and Nurjanah (2019) for *Chaetomorpha crasssa* (IC$_{50}$ value 167.39 µmol trolox/g) from Lhok Bubon, Aceh, Indonesia. The current study proves that *U. prolifera* extract may contain polyphenols that have excellent copper reducing ability as a reaction mechanism of the antioxidant property. These phenolic compounds in *U. prolifera* extract resulted in a potent antioxidant capacity better than ascorbic acid (standard antioxidant) that can be used as an alternative natural antioxidant beneficial for the pharmacological industry.

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**Figure 1.** Simple regression correlation between phenolic content and antioxidant activity via ABTS$^+$ (+) and DPPH (-) radical scavenging assay of *U. prolifera*.

**Figure 2.** Simple regression correlation between phenolic content and antioxidant activity via copper reduction antioxidant capacity (CUPRAC) assay of *U. prolifera*.
The Philippine coast is known to have several species of seaweeds with diverse biological properties. However, only a few studies are available showing the potential of these marine resources as a source of novel antibiotics that can be used against medically important pathogens. This investigation is the first report in the Philippines regarding antibacterial activities of *U. prolifera* against *S. aureus*, *Methicillin-resistant S. aureus*, *Staphylococcus epidermidis*, *Bacillus cereus*, *Aeromonas hydrophila*, and *Escherichia coli*. The result of this investigation is comparable to earlier studies done by Arguelles et al. (2019) and Jesumani, Du, Pei, Aslam, and Huang (2020). In these investigations, Gram-negative bacteria were less vulnerable to algal extract than to Gram-positive bacteria. This observation is caused by the difference in cell wall composition and structure of Gram-negative bacteria. Gram-negative bacteria contain a thick murine layer and an outer membrane that serve as an added protection and a barrier that inhibits substances such as antibiotics (Kordjazi et al., 2019; Manandhar et al., 2019; Mekinie et al., 2019).

The Philippine coast is known to have several species of seaweeds with diverse biological properties. However, only a few studies are available showing the potential of these marine resources as a source of novel antibiotics that can be used against medically important pathogens. This investigation is the first report in the Philippines regarding antibacterial activities of *U. prolifera* against *S. aureus*, *Methicillin-resistant S. aureus*, and *P. fluorescens*. In addition, this study is the first documented report regarding the antibacterial activity of *U. prolifera* against *P. fluorescens*. These findings suggest that bioactive substances such as polyphenols are present in the seaweed extract, which causes bacterial growth inhibition. Further studies should be done on the purification and identification of these chemical substances to understand the reaction mechanism that causes this biological activity.

**Conclusion**

This study presented for the first time in the Philippines the nutritional profile and functional properties of *U. prolifera*. The seaweed showed potent antibacterial activities against bacterial pathogens and potent antioxidant activity that can be exploited for pharmacological use. *U. prolifera* can produce chemical compounds in high concentration such as carbohydrates, proteins, lipids, and other essential microelements with direct relevance for nutraceutical industries. The potent biological activities and high concentration of important chemical constituents proved the suitability of this seaweed for large-scale production. Using other solvents (such as water, methanol, ethanol, ethyl acetate, and the like) as an extractant is recommended to elucidate further the differences in the biological activities of the extracts. Additional experimental assays are required to know the identity and action mechanism of the bioactive phytoconstituents present in *U. prolifera* extract and assess the biological activities *in vivo*.

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