Phylogeny of European Bat Lyssavirus 1 in Eptesicus isabellinus Bats, Spain

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To better understand the epidemiology of European bat lyssavirus 1 (EBLV-1) in Europe, we phylogenetically characterized *Lyssavirus* from *Eptesicus isabellinus* bats in Spain. An independent cluster of EBLV-1 possibly resulted from geographic isolation and association with a different reservoir from other European strains. EBLV-1 phylogeny is complex and probably associated with host evolutionary history.

The genus *Lyssavirus* comprises 3 species that can infect bats in Europe: *European bat lyssavirus 1* (EBLV-1), *European bat lyssavirus 2*, and *West-Caucasian bat virus* (1,2). Most lyssavirus-infected bats have been found in north-central Europe (Germany, the Netherlands, Denmark, Poland, and France); of these, >95% were serotine bats (*Eptesicus serotinus*) infected by EBLV-1 (3–5). EBLV-1 in other bat species has rarely been described (3,6). EBLV-1–infected bats become increasingly scarce from north to south in Europe, and no cases in northern Spain or Italy have been reported. The same trend has been consistently found within Germany (3) except for an artifact that arose from varied surveillance intensity among different countries. However, several infected serotine bats in southern Spain have been reported (7). These bats have been assigned to the species *E. isabellinus*, which has closely related populations on the African side of the Gibraltar Strait (8). This species is strongly divergent from *E. serotinus* bats (>16% of cytochrome *b* gene) in the northern Iberian Peninsula (9). In Spain, the distribution of EBLV-1 cases in bats apparently coincides with the distribution of *E. isabellinus*; 10 cases of human exposure after contact with infected bats have been reported; each was associated with *E. isabellinus* bats.

Two subtypes have been proposed for EBLV-1: EBLV-1a, which extends from the Netherlands to Russia in a west–east axis, and EBLV-1b, which includes strains that extend south through France and the Netherlands and the only 2 published strains from Iberia (1). We phylogenetically characterized EBLV-1 strains associated with *E. isabellinus* bats, a reservoir in the Iberian Peninsula that differs from *E. serotinus* bats.

The Study

We sequenced 12 bat brains positive for *Lyssavirus* antigen detected by immunofluorescence and reverse transcription–PCR (RT-PCR) as described (10). All viruses were identified as EBLV-1. For phylogenetic analyses, the 400-bp 5′ variable extreme of the nucleoprotein gene of these EBLV-1 strains was amplified by specific EBLV-1 nested RT-PCR and sequenced by using the following primers: SEQVAR1F 5′-ACGCTTAAACACCAGATCAAAG3′, SEQVAR2F 5′-AAAAATGTAACACYYCTACA3′, EBLVSEQVAR1R 5′-CAGTCACAAGATCTGTTCCAT3′, and EBLVSEQVAR2R 5′-TAGTTCAGGATTCTGTC3′.

All rabies-positive serotine bats came from southern Spain (Huelva, Seville, Murcia, and Badajoz) and were molecularly identified as *E. isabellinus* (8). An alignment was performed using ClustalX (www.clustal.org) to combine the obtained sequences and other available EBLV-1 sequences from GenBank, including a Duvenhage virus used as the outgroup (online Appendix Table, www.cdc.gov/EID/content/17/3/520-appT.htm). Before conducting further analyses, we used jModelTest (http://darwin.uvigo.es/software/jmodeltest.html) to select the best fitting substitution model for our sequences according to the corrected Akaike information criterion. Maximum-likelihood phylogenies were reconstructed by using PHYML (http://atgc.lirmm.fr/phyml) software and by using a generalized time-reversible model and the γ parameter estimated in the analyses. Maximum-parsimony analyses were conducted by using PAUP* 4.0b10 (http://paup.csit.fsu.edu/) weighting transitions 15× according to the transitions/transversion ratio estimated in the jModelTest analyses. Confidence in the topologies for the maximum-likelihood and the maximum-parsimony analyses was established with 1,000 bootstrap replicates. A Bayesian phylogenetic inference was obtained by using MrBayes version 3.1 (http://mrbayes.csit.fsu.edu/) with random starting trees without constraints. Two simultaneous runs of 107 generations were conducted, each with 4 Markov chains, and the trees were sampled...
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every 100 generations. Net p-distances between groups were calculated by using MEGA4 (www.megasoftware.net/) (Figure 1).

The genetic structure and relationships between haplotypes were examined within the main lineages through a parsimony-based network built with a median-joining algorithm implemented in the Network 4.5.1 program (11). To evaluate and compare genetic variability and polymorphism among lineages, we estimated the number of haplotypes, mutations, and segregating sites as well as haplotype diversity and nucleotide diversity by using DNAsp version 4.5 (12) for the major clades (Table). Finally, to investigate population dynamics across lineages, the Fu Fs and Tajima D statistics were calculated (Table).

**Table. Genetic diversity statistics for EBLV-1**

| Population | n  | S   | Eta | Hap | Hd  | VarHd | Pi   | ThetaNuc | k        | Tajima D | Fu Fs   |
|------------|----|-----|-----|-----|-----|-------|------|----------|----------|----------|---------|
| EBLV-1a    | 52 | 45  | 48  | 26  | 0.836| 0.00267| 0.00664| 0.02686  | 2.6546   | –2.5693  | –21.676 |
| EBLV-1b    | 25 | 35  | 35  | 18  | 0.970| 0.00038| 0.02202| 0-02317  | 8.8067   | –0.1885  | –4.555  |
| EBLV-1Spain| 13 | 9   | 9   | 7   | 0.795| 0.01191| 0.00538| 0.00725  | 2.1538   | –1.0138  | –2.067  |

*EBLV, European bat lyssavirus; n, no. sequences; S, no. segregating sites; Eta, no. mutations; Hap, no. haplotypes; Hd, haplotype diversity; VarHd, haplotype variance; Pi, nucleotide diversity; ThetaNuc, estimated population mutation rate per site; k, average no. nucleotide differences; and neutrality tests (Tajima D and Fu Fs).*

Figure 1. European bat lyssavirus 1 (EBLV-1) phylogenetic reconstruction based on the first 400 bp of the nucleoprotein gene. The tree was obtained by Bayesian inference run for 10⁷ generations; trees were sampled every 100 generations. The first 25% of trees were excluded from the analysis as burn-in. Black numbers indicate posterior probabilities. Bootstrap supports after 1,000 replicates for each node are also shown for maximum-parsimony (green numbers) and maximum-likelihood (blue numbers) analyses. Net p-distance values (as percentages) between groups are indicated by arrows. A parsimony-based network is presented for each major lineage; sizes of yellow circles are proportional to the number of individuals sharing a given haplotype, and reconstructed haplotypes (median vectors) are shown in red. DUVV, Duvenhage virus.
These 2 statistics are considered to be the most powerful tests for detecting expansion events (13).

Conclusions
All phylogenetic analyses, regardless of the reconstruction criterion used, formed a monophyletic cluster of the EBLV-1 strains from Spain (only the Bayesian inference reconstruction is shown). The Bayesian inference, maximum-likelihood, and maximum-parsimony analyses identified the cluster from Spain and EBLV-1a and EBLV-1b as being monophyletic (Figure 1), although only maximum-likelihood and maximum-parsimony analyses suggested a closer relationship between EBLV-1b and the cluster from Spain. The genetic differentiation of the EBLV-1 strains from the Iberian Peninsula matches their association with another bat species (Figure 2), which suggests that the host bat’s evolutionary history plays a major role in EBLV-1 molecular epidemiology, as has been proposed for rabies virus in bats in North America (14).

The low genetic diversity and the $F_s$ and Tajima $D$ statistics (Table) all suggest rapid population expansion of EBLV-1a, which is consistent with the star-like structure of the network for this lineage (Figure 1). Conversely, haplotype and nucleotide diversity descriptors (Table) have the highest values for EBLV-1b and a complex network structure with differentiated subnetworks. All these elements indicate that this lineage has a complex evolutionary history. The lineage from Spain also has low diversity and a star-shaped network, but neutral evolution cannot be rejected on the basis of the $F_s$ and $D$ statistics. Net distances are similar within and between lineages, except for EBLV-1a, which is slightly more differentiated (Figure 1). Consequently, the suggested EBLV-1 expansion from Spain into Europe (15) is not supported by our results, which record the highest variability and most complex phylogenetic structure for France and the Netherlands (Figure 1). This complex structure suggests either a longer evolutionary history in these areas or a recent contact of distinct bat lineages in this zone.

The results of this study show that the strains from Spain do not belong to subtype 1b because of their association with a different reservoir (E. isabellinus bats). Moreover, what is currently considered to be EBLV-1b seems to include at least 4 lineages that are more genetically diverse and have a complex history. EBLV-1a, however, has low genetic diversity despite its extensive geographic distribution, suggesting a relatively recent and successful expansion of this lineage. These results call into question the current classification of EBLV-1 into 2 single subtypes. To provide a better understanding of EBLV-1 molecular epidemiology in Europe, additional studies that consider different genes should be conducted and the current classification should be revised accordingly.

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