Intermuscular Adipose Tissue: A Brief Review of Etiology, Association With Physical Function and Weight Loss in Older Adults

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Adipose tissue redistributes during aging resulting in increased intermuscular adipose tissue (IMAT), intramuscular, and intramyocellular lipid while subcutaneous fat decreases. IMAT has been associated with lower muscle strength, power, and quality, chronic inflammation, impaired glucose tolerance, and elevated total cholesterol in older adults. This review focused on trials investigating the role of age, physical activity and diet on IMAT. The studies agreed that IMAT increases with age and seems to be responsive to physical activity, particularly the combination of aerobic and resistance exercise. However, some reported this could occur with or without weight loss, and some reported that high IMAT at baseline may blunt the muscle quality adaptive response to physical training. Larger and longer trials are needed to differentiate the independent or synergistic effects of resistance and/or aerobic training, and obesity and weight loss combined with resistance, aerobic or combination of aerobic and resistance training on IMAT. (Ann Geriatr Med Res 2019;23:3-8)

Key Words: Adipose tissue, Skeletal muscle, Weight loss, Aging

INTRODUCTION

The loss of skeletal muscle mass and function with aging (i.e., sarcopenia) is a well-known biological phenomenon. These losses are accompanied by shifts in adipose tissue and accumulation of fat in other non-adipose depots. The main anatomical depots for adipocytes are subcutaneous fat (SQF), visceral adipose tissue (VAT), intermuscular adipose tissue (IMAT), intramuscular (IMC), intramyocellular lipid (IMCL), and bone marrow. This review begins with evidence regarding age-related redistribution of adipose tissue, cellular origins, and physiological consequences of IMAT, and ends with a review of research performed during the past decade that explored the impact of physical activity, weight loss, and obesity on IMAT in older adults.

REDISTRIBUTION OF ADIPOSE TISSUE WITH AGING

Advancing age results in a redistribution of fat, with IMAT, IMC (fat within muscle but between fibers), and IMCL tending to increase, whereas SQF decreases. The IMAT is located between muscle groups and clearly separated from SQF by well-defined fascia. In contrast, IMC triglycerides accumulate within the muscle cells and are believed to primarily account for IMCL. IMAT can be evaluated and quantitated by magnetic resonance imaging (MRI) or computerized tomography (CT), whereas quantification of IMC and IMCL requires proton magnetic resonance spectroscopy (1H-MRS) or lipid histochemistry of muscle biopsy.

CELLULAR ORIGINS OF ADIPOSE TISSUE IN MUSCLE

The cellular origins of adipose accumulation within muscle fibers (intramuscular IMC/IMCL) may arise directly via the accumulation of lipid within myofibers, or intramyocellular lipid. As with IMAT, the accumulation of IMCL has been associated with insulin insensitivity, inflammation, and skeletal muscle functional deficits. The intramuscular lipid pool is both a dynamic fat-storage depot that can expand during periods of elevated lipid availability and a fatty acid source that can be utilized during periods of increased energy expenditure in active individuals. Although numerous studies have investigated the lifestyle determinants of IMCL content, the results are far from consistent, and studies attempting to unravel the mechanisms behind IMCL metabolism are in their infancy.

It has also been suggested that stem cells of the skeletal muscle may be among the drivers of adipocyte accumulation in ectopic regions. Satellite cells are a well-described stem cell population in skeletal muscle. Another stem cell type has also been described more recently, known as fibro/adipogenic progenitors, or mesenchymal interstitial cells. These cells, unlike skeletal muscle satellite cells...
that are resistant to adipogenic differentiation, readily differentiate into adipocytes under conditions of muscle injury or glucocorticoid treatment,⁸,¹² both of which may occur more frequently in middle and older age. Another regulator of adipogenesis is Wnt10b, which has been reported to suppress IMC and increase insulin sensitivity while inhibiting adipogenic differentiation in aged muscle-derived stem cells.¹³,¹⁴ Skeletal muscle also has leptin receptors, and altered leptin signaling can increase both intra- and intermuscular adipose accumulation.¹⁵ Paradoxically, caloric restriction that leads to decreasing leptin levels, or even leptin deficiency, results in increased bone marrow adiposity,⁶,¹⁷ while decreasing lipid stores and lipid droplet size in skeletal muscle.¹⁸

**PHYSIOLOGIC CONSEQUENCES AND RELATIONSHIP TO PHYSICAL ACTIVITY AND WEIGHT LOSS**

Inter- and intramuscular fat mass have been associated with lower muscle strength, power, and quality;

chronic inflammation;³⁰,³¹ impaired glucose tolerance;²³,³² and elevated total cholesterol³⁶,³⁸ in older adults. Despite consistent evidence implicating ectopic adipose tissue in aging-related loss of muscle function, the morphologic and/or molecular mechanisms are yet to be elucidated.

Both intra- and intermuscular fatty infiltration in skeletal muscle decrease sensitivity to insulin, which is required for normal protein synthesis.⁵ This mechanism may explain why fatty infiltration in and around skeletal muscle is detrimental to muscle mass and strength. In addition, the accumulation of lipid in skeletal muscle with aging or disuse is not identical across different muscle groups and fiber types. Type 1 or slow-twitch fibers accumulate more IMCL lipid with age than do fast-twitch fibers,¹³,¹⁵ and type II or fast-twitch muscle fibers are known to be preferentially lost in the progression of sarcopenia.

A number of trials have investigated the roles of age, physical activity, and diet on IMAT, and a summary of these trials is presented in Table 1. There have been fewer trials investigating the role of intermuscular fat and intermuscular myocellular lipids, and these have primarily focused on obese individuals and younger age groups, men, or athletes. As stated earlier, this area of research is nascent compared to that on IMAT, and thus this short review will focus on studies on the effect of physical activity and/or weight loss or obesity on IMAT in older adults. Studies from the past decade are summarized in Table 1.

**SUMMARY OF THE REVIEWED STUDIES**

To summarize these studies, Goodpaster et al.³⁶ reported that 12 months of physical activity prevented age-related increase in IMAT. Santanasto et al.³⁷ also reported a decrease in IMAT with physical activity combined with weight loss in obese sedentary older adults. Manini et al.³⁸ using a 6-month dietary restriction and physical activity intervention in obese sedentary older women, reported reductions in SAT and IMAT within the calf, but not the thigh, and that these changes were positively associated with faster walking speed. Nicklas et al.³⁹ used 5-month caloric restriction and resistance training (RT+CR) intervention in older obese and overweight men and women. The authors found that post-intervention, body and thigh composition measurements were all lower with RT+CR, except that IMAT did not demonstrate a decrease. However, they found that individuals with lower % baseline fat and IMAT showed greater improvement in the 400-m walk, knee strength, and power. The researchers concluded that the individuals with higher baseline adiposity experienced less overall improvement. Marcus et al.⁴⁰ also reported no effect of a 12-week, 3 times weekly combined resistance and aerobic plus balance intervention in older adults with a risk of falling. The authors reported that muscle quality only improved in those participants with low IMAT at baseline, and concluded that high IMAT blunts the muscle quality adaptive response to physical training. In contrast, Santanasto et al.⁴¹ conducted a pilot randomized controlled trial of physical activity (combined aerobic and resistance training) and weight loss in moderately obese older adults and reported that the decreased IMAT and VAT in response to the intervention was significantly associated with improved Short Physical Performance Battery (SPPB) independent of change in total fat mass. Other authors reported that IMAT was greater among older African than Caucasian men despite lower adiposity, and that IMAT was associated with type 2 diabetes regardless of race.⁴² Using the Health, Aging, and Body Composition study data, Delmonico et al.⁴³ reported that IMAT increased with age in both men and women but was independent of weight loss, weight gain, or weight stability. More recently, Englund et al.⁴⁴ reported that 6 months of a physical activity program that included walking, lower extremity resistance exercise, balance, and flexibility in older adults with limited mobility who had vitamin D deficiency resulted in improvements in body composition, SQF, IMAT, and strength. Addition of nutritional supplements resulted in further declines in IMAT.

Other interesting findings include those of Tuttle et al.⁴⁵ who reported that in obese older adults with diabetes, the gastrocnemius muscle had the highest IMAT, and that this was inversely related to walking and physical performance testing. Finally, Marcus et al.⁴⁶ conducted a 3 nonconsecutive days/week 12-week eccentric resistance training intervention in a small sample of adults (n=88) with a wide age range (30–67 years). The authors reported that eccentric resistance training decreased IMAT in the thigh in this sample of older adults who had a wide age range, and also had a variety of metabolic and mobility deficits, making interpretation of these results challenging.
| Research topic, participants | Study design, methods, outcome measures | Results | Conclusion | Investigators |
|-----------------------------|----------------------------------------|---------|------------|---------------|
| Physical activity | RCT (PA or ED) 12 month. IMAT from mid-thigh CT. | Muscle strength decreased in ED, but maintained in PA. No significant increase in IMAT in PA. SQF no difference between groups. | PA can prevent loss of strength and IMAT accumulation. | Goodpaster et al.36) |
| Race and obesity | BMI, DXA, SQF, and pQCT of calf muscle. | Afro-Caribbeans had greater IMAT and lower SQF at all levels of adiposity. Difference in IMAT, SQF independent of age, height, calf skeletal muscle, and total adipose tissue. | IMAT greater among African than Caucasian men despite lower adiposity. IMAT associated with T2D in both groups. | Miljkovic et al.42) |
| Aging and weight gain | Health ABC cohort study. Thigh CT scan, CSA muscle, strength, muscle quality. | Weight gain did not prevent loss of muscle strength. IMAT increased with age in men and women. IMAT increased with weight gain, loss, or weight stability. | Aging associated with decreased strength and quality regardless of body weight changes. IMAT changes independent of weight changes. | Delmonico et al.43) |
| Age and eccentric resistance exercise | Two aims. Observe IMAT change with age and 3 nonconsecutive days/wk for 12-week eccentric resistance training (age 55 and over). Thigh MRI. | Increasing IMAT with age. 11% decrease in thigh IMAT and 7% increase in thigh lean tissue in response to eccentric training. | Eccentric resistance training decreased thigh IMAT in a range of adults with metabolic and mobility deficits. | Marcus et al.40) |
| Exercise and muscle location | IMAT right calf by MRI, 6 min. walk test and PPT. | Gastrocnemius muscle highest IMAT and volume. No group differences. Calf IMAT inversely related to 6 min walk and PPT. | Calf IMAT muscle specific and associated with poorer physical performance. | Tuttle et al.45) |
| Weight loss and physical function | RCT comparing PA plus weight loss (PA+WL) or PA plus successful aging education (PA+SA) program. DXA, CT Biodex, SPPB. | 6 months, PA+WL lost greater thigh fat and muscle area; PA+WL lost 12.4% strength; PA+SA lost 1.0%. Muscle fat infiltration decreased significantly in PA+WL. Thigh fat area decreased 6-fold compared to lean area in PA+WL. Change in SPPB inversely correlated with change in fat, but not with change in lean or strength. | Weight loss resulted in additional functional improvements over exercise alone, primarily due to loss of body fat. | Santanasto et al.37) |
| 70 older adults with fall history | Resistance, endurance, and balance exercise 3 nonconsecutive days/wk for 12 weeks. MVC, thigh MRI to determine cross-sectional area of lean tissue and IMAT. MQ=force per unit area of lean tissue. Changes in MQ, lean and IMAT. | No significant changes in lean or IMAT in any group with training. MQ increased only in baseline low IMAT group. Middle and high IMAT groups did not demonstrate a significant change in MQ following training. | High thigh IMAT blunted the adaptive MQ response to training. | Yoshida et al.30) |
| Dietary restriction and exercise (DR+E) in obese older adults | RCT 6 months DR+E or ED. Thigh and calf muscle SQF, and IMAT by MRI. Physical function by long-distance corridor walk and knee extension strength. | DR+E significantly reduced body mass. Thigh and calf muscle volumes responded similarly between groups. Knee extension strength not changed by DR+E, but trend increased walking speed in the DR+E group. DR+E reductions in SQF and IMAT within calf, but not the thigh, associated with faster walking speed. | DR+E preserved lower extremity muscle size and function and reduced regional lower extremity adipose tissue. Reductions in calf SQF and IMAT associated with positive adaptations in physical function. | Manini et al.38) |
### Table 1. Continued

| Research topic, participants | Study design, methods, outcome measures | Results | Conclusion | Investigators |
|-----------------------------|----------------------------------------|---------|------------|---------------|
| CR for weight loss and RT on muscle and physical function | RCT of 5-month progressive, 3 d/wk, moderate-intensity RT with weight loss (RT+CR) or RT without weight loss (RT). Biodex maximal knee strength; muscle power. DXA and CT muscle quality, overall physical function, and total body and thigh composition. | Fat mass, % fat, and all thigh fat volumes decreased in both groups. Only RT+CR group lost lean mass. Post-intervention body and thigh composition were all lower with RT+CR except IMAT. Lower % baseline fat and IMAT had greater improvement in the 400-m walk, knee strength and power. | RT improved body composition (including reduced IMAT) and muscle strength and physical function. Higher baseline adiposity had less improvement. | Nicklas et al.39) |
| PA and weight loss. 36 overweight to moderately obese older adults. Mean age 70.6±6.1 years | 12-month pilot RCT (PA+WL) or PA plus SA education. PA was treadmill walking supplemented with lower extremity resistance and balance training. WL based on Diabetes Prevention Project with 7% weight loss by cutting fat calories. CT and DXA. VAT and thigh IMAT. SPPB | Decreased IMAT and VAT significantly associated with improved SPPB independent of change in total fat mass. PA+WL improved SPPB, whereas PA+SA did not. No intergroup differences. | Decreases in IMAT and VAT important mechanisms underlying improved function following intentional weight loss plus physical activity. | Santanasto et al.41) |
| Nutritional supplementation and physical activity. 149 mobility limited and vitamin D deficient older adults. 46.3% women Mean age 78.5±5.4 years. | Six-month trial. All participated in a PA program of walking, lower-extremity strength exercises, balance, and flexibility. Randomized to daily nutritional supplement (150 kcal, 20 g whey protein, 800 IU vitamin D, 119 mL beverage) or placebo (30 kcal, non-nutritive, 119 mL). DXA CT thigh composition and muscle strength, power, and quality. | Both groups demonstrated improvements in muscle strength, body composition, and thigh composition. Nutritional supplementation led to further losses of IMAT and increased normal muscle density. | Six months of physical activity resulted in improvements in body composition, SQF, IMAT and strength. Addition of nutritional supplement showed further declines in IMAT and improved muscle density. | Englund et al.44) |

IMAT, intermuscular adipose tissue; RCT, randomized controlled trial; PA, physical activity; ED, education; CT, computed tomography; SQF, subcutaneous fat; BMI, body mass index; DXA, dual energy X-ray absorptiometry; pQCT, peripheral quantitative computed tomography; T2D, type 2 diabetes; ABC, aging and body composition; CSA, cross sectional area; MRI, magnetic resonance spectroscopy; PPT, physical performance test; WL, weight loss; SA, successful aging; SPPB, short physical performance battery; MVC, maximal voluntary contraction; MQ, muscle quality; DR, dietary restriction; E, exercise; RT, resistance training; CR, caloric restriction; VAT, visceral adipose tissue.
CONCLUSION

Studies to date consistently show that IMAT increases with age and appears to be responsive to physical activity, particularly the combination of aerobic and resistance exercise. However, there is less agreement regarding whether this response may occur with or without weight loss. Many of the studies cited in this brief review had small study samples. The two larger trials in obese and non-obese older adults suggest an effectiveness of physical activity on IMAT, although additional trials are needed to differentiate the independent or synergistic effects of resistance or aerobic training alone or in combination on IMAT, as well as the effects of obesity and weight loss combined with resistance, aerobic, or a combination of aerobic and resistance training. Ongoing clinical trials may provide a greater understanding of the relationships between aging, physical activity, weight loss, physical function, and IMAT.

CONFLICTS OF INTEREST DISCLOSURES

The researcher claims no conflicts of interest.

REFERENCES

1. Kirkland JL, Tchkonia T, Pirskkhala T, Han J, Karagiannides I. Adipogenesis and aging: does aging make fat go MAD? Exp Gerontol 2002;37:757-67.
2. Coen PM, Goodpaster BH. Role of intramyocellular lipids in human health. Trends Endocrinol Metab 2012;23:391-8.
3. Kindler JM, Lewis RD, Hamrick MW. Skeletal muscle and pediatric bone development. Curr Opin Endocrinol Diabetes Obes 2015;22:467-74.
4. Komolka K, Albrecht E, Wimmers K, Michal JJ, Maak S. Molecular heterogeneities of adipose depots - potential effects on adipose-muscle cross-talk in humans, mice and farm animals. J Genomics 2014;2:31-44.
5. Rivas DA, McDonald DJ, Rice NP, Haran PH, Dolnikowski GG, Fielding RA. Diminished anabolic signaling response to insulin induced by intramuscular lipid accumulation is associated with inflammation in aging but not obesity. Am J Physiol Regul Integr Comp Physiol 2016;310:R561-9.
6. Gepner Y, Shelef I, Schwarzfuchs D, Cohen N, Bril N, Rein M, et al. Intramyocellular triacylglycerol accumulation across weight loss strategies; Sub-study of the CENTRAL trial. PLoS One 2017;12:e0188431.
7. Vettor R, Milan G, Franzin C, Sanna M, De Coppi P, Rizzuto R, et al. The origin of intermuscular adipose tissue and its pathophysiological implications. Am J Physiol Endocrinol Metab 2009;297:E987-98.
8. Dong Y, Silva KA, Dong Y, Zhang L. Glucocorticoids increase adipocytes in muscle by affecting IL-4 regulated FAP activity. FASEB J 2014;28:4123-32.
9. Farup J, Madaro L, Puri PL, Mikkelsen UR. Interactions between muscle stem cells, mesenchymal-derived cells and immune cells in muscle homeostasis, regeneration and disease. Cell Death Dis 2015;6:e1830.
10. Penton CM, Thomas-Ahner JM, Johnson EK, McAllister C, Montanaro F. Muscle side population cells from dystrophic or injured muscle adopt a fibro-adipogenic fate. PLoS One 2013:e85455.
11. Uezumi A, Fukada S, Yamamoto N, Takeda S, Tsuichida K. Mesenchymal progenitors distinct from satellite cells contribute to ectopic fat cell formation in skeletal muscle. Nat Cell Biol 2010;12:143-52.
12. Agley CC, Rowleson AM, Velloso CP, Luzanus NR, Harridge SD. Human skeletal muscle fibroblasts, but not myogenic cells, readily undergo adipogenic differentiation. J Cell Sci 2013;126:5610-25.
13. Abiola M, Favier M, Christodoulou-Vafeiadou E, Pichard AL, Martelly I, Guillet-Deniau I. Activation of Wnt/beta-catenin signaling increases insulin sensitivity through a reciprocal regulation of Wnt10b and SREBP-1c in skeletal muscle cells. PLoS One 2009;4:e8509.
14. Taylor-Jones JM, McGeehe RE, Rando TA, Lecka-Czernik B, Lipschitz DA, Peterson CA. Activation of an adipogenic program in adult myoblasts with age. Mech Ageing Dev 2002;123:649-61.
15. Aurumleit F, Bowser M, Upadhyay S, Shi XM, Fulzele S, Johnson MH, et al. Absence of functional leptin receptor isoforms in the POUND (Lepr(db/lb)) mouse is associated with muscle atrophy and altered myoblast proliferation and differentiation. PLoS One 2013;8:e72330.
16. Devlin MJ. Why does starvation make bones fat? Am J Hum Biol 2011;23:577-85.
17. Devlin MJ, Cloutier AM, Thomas NA, Panus DA, Lotimun S, Pinz I, et al. Caloric restriction leads to high marrow adiposity and low bone mass in growing mice. J Bone Miner Res 2010;25:2078-88.
18. Shaw CS, Clark J, Wagenmakers AJ. The effect of exercise and nutrition on intramuscular fat metabolism and insulin sensitivity. Annu Rev Nutr 2010;30:13-34.
19. Beavers KM, Ambrosius WT, Nicklas BJ, Rejeski WJ. Independent and combined effects of physical activity and weight loss on inflammatory biomarkers in overweight and obese older adults. J Am Geriatr Soc 2013;61:1089-94.
20. Gallagher D, Kuznia P, Heshka S, Albu J, Heymsfield SB, Goodpaster B, et al. Adipose tissue in muscle: a novel depot similar in size to visceral adipose tissue. Am J Clin Nutr 2005;81:903-10.
21. Goodpaster BH, Carlson CL, Visser M, Kelley DE, Scherzinger A, Harris TB, et al. Attenuation of skeletal muscle and strength in the elderly: The Health ABC Study. J Appl Physiol (1985) 2001;90:2157-65.
22. Goodpaster BH, Thaete FL, Kelley DE. Thigh adipose tissue distribution is associated with insulin resistance in obesity and in type 2 diabetes mellitus. Am J Clin Nutr 2000;71:885-92.
23. Koster A, Stenholm S, Alley DE, Kim LJ, Simonsick EM, Kanaya AM, et al. Body fat distribution and inflammation among obese older adults with and without metabolic syndrome. Obesity (Silver Spring) 2010;18:2354-61.
24. Pou KM, Massaro JM, Hoffmann U, Vasan RS, Maurovich-Horvat P, Larson MG, et al. Visceral and subcutaneous adipose tissue volumes are cross-sectionally related to markers of inflammation and oxidative stress: the Framingham Heart Study. Circulation 2007;116:1234-41.
25. Sepe A, Tchkonia T, Thomou T, Zamboni M, Kirkland JL. Aging and regional differences in fat cell progenitors - a mini-review. Gerontology 2011;57:66-75.
26. Snijder MB, Visser M, Dekker JM, Goodpaster BH, Harris TB, Kritchevsky SB, et al. Low subcutaneous thigh fat is a risk factor for unfavourable glucose and lipid levels, independently of high abdominal fat. The Health ABC Study. Diabetologia 2005;48:301-8.
27. Visser M, Kritchevsky SB, Goodpaster BH, Newman AB, Nevitt M, Stamm E, et al. Leg muscle mass and composition in relation to lower extremity performance in men and women aged 70 to 79: the health, aging and body composition study. J Am Geriatr Soc 2002;50:897-904.
28. Yoshida Y, Marcus RL, Lastayo PC. Intramuscular adipose tissue and central activation in older adults. Muscle Nerve 2012;46:813-6.
29. Zoico E, Di Francesco V, Mazzali G, Vettor R, Fantin F, Bissoli L, et al. Adipocytokines, fat distribution, and insulin resistance in elderly men and women. J Gerontol A Biol Sci Med Sci 2004;59:M935-9.
30. Beasley LE, Koster A, Newman AB, Javaid MK, Ferrucci L, Kritchevsky SB, et al. Inflammation and race and gender differences in computed tomography-measured adipose depots. Obesity (Silver Spring) 2009;17:1062-9.
31. Cartier A, Côté M, Lemieux I, Pérusse L, Tremblay A, Bouchard C, et al. Age-related differences in inflammatory markers in men: contribution of visceral adiposity. Metabolism 2009;58:1452-8.
32. Dubé MC, Lemieux S, Piché ME, Corneau L, Bergeron J, Riou ME, et al. The contribution of visceral adiposity and mid-thigh fat-rich muscle to the metabolic profile in postmenopausal women. Obesity (Silver Spring) 2011;19:953-9.
33. Durheim MT, Strentz CA, Bateman LA, Mahe SK, Kraus WE. Relationships between exercise-induced reductions in thigh intermuscular adipose tissue, changes in lipoprotein particle size, and visceral adiposity. Am J Physiol Endocrinol Metab 2008;295:E407-12.
34. Choi SJ, Files DC, Zhang T, Wang ZM, Messi ML, Gregory H, et al. Intramyocellular lipid and impaired myofiber contraction in normal weight and obese older adults. Gerontol A Biol Sci Med Sci 2016;71:557-64.
35. Gnaiger E, Boushel R, Søndergaard H, Munch-Andersen T, Damsgaard R, Hagen C, et al. Mitochondrial coupling and capacity of oxidative phosphorylation in skeletal muscle of Inuit and Cauca-
sians in the arctic winter. Scand J Med Sci Sports 2015;25 Suppl 4:126-34.
36. Goodpaster BH, Chomentowski P, Ward BK, Rossi A, Glynn NW, Delmonico MJ, et al. Effects of physical activity on strength and skeletal muscle fat infiltration in older adults: a randomized controlled trial. J Appl Physiol (1985) 2008;105:1498-503.
37. Santanasto AJ, Glynn NW, Newman MA, Taylor CA, Brooks MM, Goodpaster BH, et al. Impact of weight loss on physical function with changes in strength, muscle mass, and muscle fat infiltration in overweight to moderately obese older adults: a randomized clinical trial. J Obes 2011;2011:516576.
38. Minnick TM, Buford TW, Lott DJ, Vandenbome K, Daniels MJ, Knaggs JD, et al. Effect of dietary restriction and exercise on lower extremity tissue compartments in obese, older women: a pilot study. J Gerontol A Biol Sci Med Sci 2014;69:101-8.
39. Nicklas BJ, Chmelo E, Delbono O, Carr JJ, Lyles MF, Marsh AP. Effects of resistance training with and without caloric restriction on physical function and mobility in overweight and obese older adults: a randomized controlled trial. Am J Clin Nutr 2015;101:991-9.
40. Marcus RL, Addison O, Kidde JP, Dibble LE, Lastayo PC. Skeletal muscle fat infiltration: impact of age, inactivity, and exercise. J Nutr Health Aging 2010;14:362-6.
41. Santanasto AJ, Newman AB, Strotmeyer ES, Boudreau RM, Goodpaster BH, Glynn NW. Effects of changes in regional body composition on physical function in older adults: a pilot randomized controlled trial. J Nutr Health Aging 2015;19:913-21.
42. Miljkovic I, Yerges LM, Li H, Gordon CL, Goodpaster BH, Kuller LH, et al. Association of the CPT1B gene with skeletal muscle fat infiltration in Afro-Caribbean men. Obesity (Silver Spring) 2009;17:1396-401.
43. Delmonico MJ, Harris TB, Visser M, Park SW, Conroy MB, Velasquez-Meyer P, et al. Longitudinal study of muscle strength, quality, and adipose tissue infiltration. Am J Clin Nutr 2009;90:1579-85.
44. Englund DA, Kirn DR, Koochek A, Zhu H, Travison TG, Reid KF, et al. Nutritional supplementation with physical activity improves muscle composition in mobility-limited older adults, the VIVE2 study: a randomized, double-blind, placebo-controlled Trial. J Gerontol A Biol Sci Med Sci 2017;72:95-101.
45. Tuttle LJ, Sinacore DR, Mueller MJ. Intramuscular adipose tissue is muscle specific and associated with poor functional performance. J Aging Res 2012;2012:172957.