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Paternity in wild ring-tailed lemurs (Lemur catta): implications for male mating strategies

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In group-living species with male dominance hierarchies where receptive periods of females do not overlap, high male reproductive skew would be predicted. However, the existence of female multiple mating and alternative male mating strategies can call into question single-male monopolization of paternity in groups. Ring-tailed lemurs (Lemur catta) are seasonally breeding primates that live in multi-male, multi-female groups. Although established groups show male dominance hierarchies, male dominance relationships can break down during mating periods. In addition, females are the dominant sex and mate with multiple males during estrus, including group residents, and extra-group males—posing the question of whether there is high or low male paternity skew in groups. In this study, we analyzed paternity in a population of wild L. catta from the Bezà Mahafaly Special Reserve in southwestern Madagascar. Paternity was determined with 80–95% confidence for 39 offspring born to nine different groups. We calculated male reproductive skew indices for six groups, and our results showed a range of values corresponding to both high and low reproductive skew. Between 21% and 33% of offspring (3 of 14 or three of nine, counting paternity assignments at the 80% or 95% confidence levels, respectively) were sired by extra-troop males. Males siring offspring within the same group during the same year appear to be unrelated. Our study provides evidence of varying male reproductive skew in different L. catta groups. A single male may monopolize paternity across one or more years, while in other groups, >1 male can sire offspring within the same group, even within a single year. Extra-group mating is a viable strategy that can result in extra-group paternity for L. catta males.

KEYWORDS
Bezà Mahafaly, extra-group mating, Madagascar, reproductive skew, sexual selection

†This paper is dedicated to the memory of our co-author Robert W. (Bob) Sussman (1941–2016), a pioneer of lemur research and a highly respected scholar of primatology and biological anthropology. If not for Bob’s hard work and determination in helping to establish the Bezà Mahafaly Special Reserve, neither this study, nor countless others based on research at Bezà Mahafaly, would have been possible.

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1 | INTRODUCTION

Reproductive skew refers to the distribution of reproductive success among same-sex individuals in a population (Clutton-Brock, 1998). Among males, several factors can determine the extent of reproductive skew. For example, high male reproductive skew is expected where female fertile periods do not overlap and where stable male dominance hierarchies determine mating priority (Altmann, 1962). Conversely, male reproductive skew is expected to be lower where there are more male competitors (Kutsukake & Nunn, 2006), greater numbers of females (Kappeler & Port, 2008), or where overlap in female receptive periods disrupts the ability of single males to monopolize mating opportunities (Ostner, Nunn, & Schülke, 2008).

Apart from its interesting behavioral causes and consequences, reproductive skew is important to document for its influence on microevolutionary dynamics. When reproduction is concentrated to a limited number of males within a population (assuming an equal sex ratio), skewed reproduction ultimately lowers the variance effective size of a population (NeV), thereby increasing the effects of genetic drift; this is particularly relevant to protected and threatened populations. Further, reproductive skew has the potential to unite offspring cohorts at the half-sib level (since infants all share the same father), thereby promoting the opportunity for kin selection to act (Altmann, 1979; Widdig, 2013). Finally, reproductive skew influences the opportunity for sexual selection, and thereby the evolution of sexually dimorphic traits (Shuster & Wade, 2003).

Among primates with multiple male residents per group, paternity can be highly skewed, with one or just a few males siring group offspring (e.g., capuchin monkeys, Cebus capucinus: [Jack & Fedigan, 2006; Muniz et al., 2010]; red-fronted lemurs, Eulemur rufifrons: [Kappeler & Port, 2008; Wimmer & Kappeler, 2002]). Conversely, paternity may show relatively low skew, with reproductive success being distributed among several males (e.g., Macaca assamensis: [Suaka, Wajjwalku, Ostner, & Schülke, 2014]) who may or may not be related. Related males siring infants in the same group would have effects similar to high paternity skew: increased relatedness of offspring cohorts that would favor kin selection (Altmann, 1979; Widdig, 2013). Sires can also come from within the group (residents) or outside of the group (extra-group males).

Occasionally, extra-group male parentage can be considerable, such as in Verreaux's sifaka, Propithecus verreauxi, where non-residents sire between 17% and 65% of group offspring each year (Lawler, 2007; Lawler, Richard, & Riley, 2003). Similarly, in one population of langurs (Semnopithecus entellus), nonresident males sired 21% of the infants in multi-male groups (Launhardt, Borries, Hardt, Epplen, & Winkler, 2001). Rhesus macaques, Macaca mulatta, on Cayo Santiago show extra-group paternity rates as high as >59% in some years (Georgiev et al., 2016). Rates of extra-group paternity are expected to be highest where resident males have greater difficulty monopolizing copulations with females (e.g., groups with a female-biased sex ratio: [Lawler et al., 2003]).

In this study, we investigated male reproductive skew, extra-group male paternity, and evaluated the relatedness of sires in the ring-tailed lemur, Lemur catta, a seasonally breeding primate endemic to Madagascar that lives in multi-male, multi-female groups and is female dominant (Jolly, 1966; Kappeler, 1990; Pereira, Kaufman, Kappeler, & Overdorff, 1990; Sauther, Sussman, & Gould, 1999). This primate species is of unique interest because it is characterized by extremes in reproductive traits (i.e., asynchronous estrus that lasts for <1 day [Pereira, 1991; Sauther, 1991]) that would seem to favor male monopolization potential, but other aspects of this species' behavior might be expected to promote low paternity skew. For example, established groups usually have a single high-ranking male who is dominant over other group males (Sauther, 1991; Sauther & Sussman, 1993) and this alpha male tends to mate first and mate guard for longer periods than other males (but see Gould, 1994; Parga, 2003; Sauther, 1991). As such, paternity skew could be high, especially if a first-mate fertilization advantage operates in this species (Pereira & Weiss, 1991). However, male dominance relationships can be highly unstable during mating periods (Gould, 1994, 1997; Gould & Ziegler, 2007; Jolly, 1966; Koyama, 1988; Parga, 2009), and females (due to their social dominance) exercise a high degree of mate choice (Gould, 1994; Koyama, 1988; Pereira & Weiss, 1991; Sauther, 1991; Taylor, 1986; Taylor & Sussman, 1985), limiting the monopolization ability of high-ranking males. Consequently, females mate with multiple males from within and outside of the group (Gould, 1994; Koyama, 1988; Sauther, 1991; Sussman, 1992), which should be expected to decrease reproductive skew. Clearly, the level of male reproductive skew or extent of extra-group paternity is impossible to predict solely with behavioral measures, necessitating genetic analysis to address these issues.

2 | METHODS

2.1 | Study site and sample collection

Across seven capture years between 1987 and 2006, we collected 243 biological samples (blood or hair) from safely captured, sedated L. catta belonging to groups in and around the gallery forest portion of the Bezà Mahafaly Special Reserve (BMSR), Madagascar. Beginning in 2003, all groups in and surrounding Parcel 1 of BMSR were sampled. The lemurs were studied as part of long-term research on L. catta ecology, health, behavioral endocrinology, and demography (Gould, Ziegler, & Wittwer, 2005; Gould & Ziegler, 2007; Sauther et al., 2006; Sussman & Ratsirarson, 2006). These samples derived from 14 distinct L. catta groups, each with 3–8 males and 2–9 females. Beginning in 2003, yearly records were kept of membership within reserve groups. Records of group membership in BMSR prior to this date were made opportunistically. For further details on methodology, including capture protocol, refer to Sauther et al. (2006) and Parga, Sauther, Cuzzo, Youssouf Jacky, and Lawler (2012). Since 2003, all animal handling was conducted with Institutional Animal Care and Use Committee (IACUC) approval from the University of Colorado and/or the University of North Dakota. This research also adhered to the American Society of Primatologists (ASP) Principles for the Ethical Treatment of Non-Human Primates, and conformed to the legal requirements of the government of Madagascar.
2.2 Genetic analyses

The protocol for DNA extraction and amplification has been previously described (Parga et al., 2012, 2015; Pastorini, Fernando, Forstner, & Melnick, 2005, 2015). The following microsatellites were used: Lc5, Lc6, Lc7, Lc8, Lc9, Lc10 (Pastorini et al., 2005), 69HDZ267, 69HDZ299 (Zaonarivelo et al., 2007), Efr09 (Jekielek & Strobeck, 1999), Efr02 (Wimmer, 2000), L-2 (Merenlender, 1993), Em7 (Pastorini, Fernando, Melnick, & Forstner, 2004), Em12 (Parga et al., 2015), and Pv1 (Lawler, Richard, & Riley, 2001). MICRO-CHECKER, version 2.2.3 (van Oosterhout, Hutchinson, Wills, & Shipley, 2004) was used to evaluate the data for null alleles and scoring errors. One microsatellite (Lc9) showed evidence of scoring errors and null alleles, so was discarded. Approximately, 55% of samples were re-genotyped from separate extractions to verify allelic data, as in Parga et al. (2012).

CERVUS (Kalinowski, Taper, & Marshall, 2007; Marshall, Slate, Kruuk, & Pemberton, 1998) was used to determine maternity and paternity using maximum likelihood methods for individuals born into the study groups who could be identified as natal offspring based on age. Fifty-eight animals from 11 groups met the criterion of being natal offspring at the time of capture. Natal offspring were those that, at the time of capture, were either infants (1 yr), subadults (2 yrs), or young adults (3 yrs). All sampled adult males in the population (N = 141) were included as potential sires. Simulation parameters in CERVUS included 10,000 cycles, 72% of loci typed, and 188 candidate fathers (we assumed that the 141 males we sampled only represented 75% of possible sires). The proportion of loci mistyped was kept at the default value of 0.01. For every infant born, LOD (natural logarithm of the likelihood-odds ratio) scores were calculated for each possible sire, indicating the likelihood that each male sired the offspring in question. The male with the highest LOD score was identified as the sire, excluding males who were too young at the time of infant conception to have sired offspring. Sire-offspring pairs had to share at least one allele at each locus and were allowed no mismatching loci. Following standard convention, parentage assignments were made at two confidence levels: 80% and 95% (Marshall et al., 1998). Extra-group paternity was determined when a male was identified as a sire but did not belong to the infant’s social group at the time the infant was conceived, or by excluding all males residing in the group at the time of the infant’s conception due to allelic mismatches at one or more loci.

As with high paternity skew, instances in which male kin sire same-group infants can increase offspring relatedness (Altmann, 1979). We therefore used ML-Relate (Kalinowski, Wagner, & Taper, 2006) to generate maximum likelihood estimates of pairwise relatedness (r) for males siring offspring within the same group in the same year. This program provides 95% confidence sets for relationships between pairs of individuals. For each set of male sires of same-year and same-group offspring, the most likely putative relationship between the two males (unrelated) was tested against the second most likely alternative (half-sibling) using likelihood ratio tests, with a 0.05 significance level.

Methods to estimate pairwise relatedness and determine kin relationships from molecular data alone are not always accurate (Csillery et al., 2006; Van Horn, Altmann, & Alberts, 2008). Therefore, to test the predictive accuracy of ML-Relate for our population, using CERVUS paternity and maternity assignments reaching 95% confidence, we created known sets of dyads in four different relatedness categories: parent-offspring (N = 5), full-siblings (N = 3), half-siblings (N = 5), and unrelated individuals (N = 5). ML-Relate identified the correct kin relationship (via a significant P-value; data not shown) in four of five cases for parent-offspring pairs (r = 0.5–0.67; mean r = 0.54), one of three cases for full-siblings (r = 0.39–0.52; mean r = 0.48), two of five cases for half-siblings (r = 0.27–0.57; mean r = 0.35), and one of five cases for unrelated individuals (r = 0.0–0.37; mean r = 0.13). Though ML-Relate did not consistently perform well at hypothesis-testing to determine proper kin relationships in our cohort of dyads who were known to be related, the r values generated by ML-Relate were either at or above what would be expected for each relationship category (0.5 for parent-offspring and full sib pairs, 0.25 for half-sibs). Accordingly, whereas higher relatedness values (r ≥ 0.2) generated by ML-Relate in this dataset cannot be consistently used to identify the proper kin category among relatives, low values of relatedness (r < 0.1) can be trusted in this dataset to identify unrelated individuals, as only dyads in our constructed cohort known to be unrelated show such low r values.

2.3 Determining reproductive skew

The software Skew Calculator 2013 (https://www.eeb.ucla.edu/Faculty/Nonacs/Pl.html) was used to calculate Nonacs’ B, a binomial skew index (Nonacs, 2000, 2003) and to test whether the observed male reproductive skew per group was significantly different from a random distribution. The B index was calculated for groups producing more than one offspring for which a sire could be identified at the 80% or 95% confidence level (N = 6) across the period spanning 2000–2005, which were the years across which the most complete paternity data for the greatest number of study groups was available. Groups for which such data were lacking (N = 7) were not used to calculate reproductive skew. This 6 year period was deemed appropriate for the calculation of the skew index because it falls within the time frame of some alpha male tenure durations at this location (e.g., in BMSR, three males were documented as having alpha tenures that lasted for 6 years [Sauther et al., 1999]). B index values closer to one indicate high reproductive skew, values closer to zero indicate low skew, and negative values indicate a more even distribution of paternity than would be expected by chance. B index values were calculated with 95% confidence intervals.

When calculating the B index, the Skew Calculator software is capable of incorporating data on male tenure for sires and non-sires; therefore, we included data on the duration of male membership in each group (number of years each male was a group member) where such data were available, which was from 2003 to 2005 for most groups, with the exception of Green group, for which data on male group membership were available even earlier, beginning in 2001. Though male tenure data were not consistently available across all 6 years of the period of infant production we considered (2000–2005), we deemed it more accurate to include all available data on male tenure rather than exclude it, especially when such data were available.
for groups for at least half (or more, the case of Green group) of the period under consideration.

Nonparametric statistics were run in Statistica 12 (Stat Soft Inc., Tulsa, OK, 2013). In particular, Spearman rank correlation tests were used to determine whether aspects of group composition across the multi-year period of analysis (the total number of group males, the average number of group males per year, the average number of group females per year) showed a significant relationship with the amount of paternity skew, as measured by the B index. Mann–Whitney U tests were used to determine if there was a significant difference in the number of males, number of females, or sex ratio between groups with and without extra-group paternity. All tests were two-tailed, with alpha set at 0.05.

3 | RESULTS

Average heterozygosity and allele number were high for the microsatellite loci used in paternity analyses (Table 1). These loci had a combined exclusionary power of 0.9996 when assigning the first parent (both parents being unknown), and 0.9999 when assigning the second parent. Out of 114 males in the population, 21 males were identified as sires (including a pair of identical male twins who were genetically indiscernible) at the 80% confidence level. If strictly counting only assignments at the 95% confidence level, 17 males were identified as sires (including the identical male twins). Although sires produced between 1 and 3 infants per year, each sire only produced offspring within a single group per year (Figure 1).

3.1 | Paternity assignments and Nonacs’ B index

Male parentage could be assigned at the 80% or 95% significance level for 39 of the 58 offspring who were identified as natal at the time of capture. The 39 offspring came from nine different groups (Figure 1).

### Table 1: Heterozygosity and allele number for loci used in paternity analysis

| Locus     | k | H₀   | Hₑ   |
|-----------|---|------|------|
| Lc5       | 9 | 0.750| 0.778|
| Lc6       | 8 | 0.750| 0.734|
| Lc7       | 10| 0.900| 0.838|
| Lc8       | 7 | 0.733| 0.757|
| Lc10      | 10| 0.807| 0.794|
| 69HDZZ267 | 10| 0.800| 0.816|
| 69HDZZ299 | 7 | 0.700| 0.795|
| Efr02     | 10| 0.741| 0.758|
| Efr09     | 12| 0.800| 0.740|
| L-2       | 12| 0.850| 0.825|
| Em7       | 5 | 0.588| 0.621|
| Em12      | 17| 0.850| 0.864|
| Pv1       | 13| 0.841| 0.869|
| Average   | 10| 0.778| 0.784|

The average number of group males across the period of analysis (Spearman: $r_s = 0.26$, $N = 6$, $P = 0.62$), average number of males per group per year ($r_s = 0.38$, $N = 6$, $P = 0.46$) or average number of females per group per year ($r_s = 0.26$, $N = 6$, $P = 0.62$). Indeed, there was substantial overlap in each of these variables in groups having both high and low reproductive skew (Table 2).

3.2 | Extra-group male parentage

Individual males only sired offspring within a single group per year, and both resident males and extra-group males sired offspring. Among
cases in which paternity assignments were made at the 95% confidence level, in three of nine (33%) cases, sires either did not belong to the social group at the time of conception (triangles), or whether the birth was a case of extra-group paternity (X). Cases of extra-group paternity where a sire could not be identified with at least 80% confidence, but where all residents were excluded as sires due to one or more allelic mismatches (N = 2), are not shown.

Admittedly, male group membership was unknown at the time of conception for many infants, so the actual values of extra-group paternity measured here may be underestimated.

To test whether the number of resident males per group, the number of resident females, or the group sex ratio had a significant effect on extra-group male parentage, a series of Mann–Whitney U tests were conducted on groups in which male and female membership data were available at the time of infant conception (N = 6). Although few groups showed extra-group paternity, we found much overlap in the composition of groups with and without extra-group paternity (Table 3). Neither the number of males (U = 11, N₁ = 3, N₂ = 8, P = 0.92), the number of females (U = 8.0, N₁ = 2, N₂ = 8, P = 1.0) nor the group sex ratio (U = 5.5, N₁ = 2, N₂ = 8, P = 0.53) showed a significant difference between groups with and without extra-troop male paternity (Table 3).

### 3.3 Relatedness among sires

In three instances, two different sires sired offspring in the same group during the same year (Figure 1, Table 4). In each instance, the sires appeared to be unrelated to one another, showing remarkably

| Sire     | 1985 // | 1987 // | 1993 // | 1999 // | 2000 | 2001 | 2002 | 2003 | 2004 | 2005 |
|----------|---------|---------|---------|---------|------|------|------|------|------|------|
| BM8      | Δ       |         |         |         | Δ    |      |      |      |      |      |
| BM36     |         | Δ       |         |         |      |      |      |      |      |      |
| BM45     |         |         | Δ       |         |      |      |      |      |      | Δ    |
| BM90     |         |         |         | Δ       |      |      |      |      |      |      |
| BM163    |         |         |         |         | Δ    |      |      |      |      |      |
| BM222    |         |         |         |         | Δ    |      |      |      |      |      |
| BM113    |         |         |         |         |      |      |      |      |      |      |
| BM115    |         |         |         |         |      |      |      |      |      |      |
| BM117    |         |         |         |         |      |      |      |      |      |      |
| BM153    |         |         |         |         |      |      |      |      |      |      |
| BM173    |         |         |         |         |      |      |      |      |      |      |
| BM189/191 (identical twins) | Δ | Δ | Δ | Δ | Δ | Δ | Δ | Δ | Δ | Δ |
| BM195    |         |         |         |         |      |      |      |      |      |      |
| BM101    |         |         |         |         |      |      |      |      |      |      |
| BM143    |         |         |         |         |      |      |      |      |      |      |
| BM453    |         |         |         |         |      |      |      |      |      |      |
| BM133    |         |         |         |         |      |      |      |      |      |      |
| BM136    |         |         |         |         |      |      |      |      |      |      |
| BM140    |         |         |         |         |      |      |      |      |      |      |
| BM124    |         |         |         |         |      |      |      |      |      |      |

**FIGURE 1** All paternity assignments made at the 80% and 95% confidence levels, indicating the number of offspring produced by each male. Each infant’s group of birth is shown, indicating whether the sire was a resident (circles), whether male group membership was unknown at the time of conception (triangles), or whether the birth was a case of extra-group paternity (X). Cases of extra-group paternity where a sire could not be identified with at least 80% confidence, but where all residents were excluded as sires due to one or more allelic mismatches (N = 2), are not shown.

### TABLE 2 Nonacs’ B index values per group and group parameters

| Group | Total # known males in group during period | Ave # of males per year | Ave # of females per year | # Sires | # Infants | B    | CI     | P   |
|-------|-------------------------------------------|-------------------------|---------------------------|---------|-----------|------|--------|-----|
| Black | 11                                        | 5.3                     | 4.7                       | 4       | 6         | -0.039 | -0.137–0.168 | 0.798 |
| Blue  | 8                                         | 5                       | 8.3                       | 4       | 7         | 0.071  | -0.116–0.496 | 0.156 |
| Green | 13                                        | 5.2                     | 5                         | 3       | 3         | 0.072  | -0.282–0.539 | 0.290 |
| Orange| 10                                        | 5.3                     | 6.3                       | 2       | 4         | 0.378  | 0.046–0.794  | <0.001 |
| Teal  | 14                                        | 7.7                     | 5.3                       | 1       | 5         | 0.734  | 0.227–0.734  | <0.0001 |
| Yellow| 5                                         | 3.7                     | 5.7                       | 4       | 8         | 0.158  | -0.091–0.524 | 0.049 |
low estimates of relatedness \((r = 0.0–0.04)\), although for only one pair did the test to evaluate the unrelated status of the males against the second most likely alternative (half-siblings) reach statistical significance (Table 4). Nevertheless, all three estimates of relatedness between these “same group sires” were markedly lower than the relatedness values calculated for our cohort of male dyads known to be half-siblings \((r = 0.27–0.57); \text{see Methods}\), which makes it unlikely that the “same group sires” shared any considerable degree of relatedness.

### 4 | DISCUSSION

The *L. catta* groups in our population showed variability in male reproductive skew. One group showed considerable skew, with a single male siring >1 offspring in the same group in the same year as well as across multiple years. Another group showed more moderate skew (two sires of four offspring), while all other groups showed lower skew, with as many as four different males siring group offspring across a 6-year period (sometimes with >1 sire in the same group in the same year). No consistent relationship was found between levels of paternity skew and group composition (average number of group males per year, average number of group females per year, or total number of males across the 6-year period of analysis). Paternity skew is of interest because it is expected to favor kin selection via the creation of closely related cohorts of offspring; related males siring same-group offspring would have a similar effect (Altmann, 1979).

However, not only did most groups in our study show low skew, but different males who sired infants within the same group in the same year appeared to be unrelated.

When compared to other primates in which paternity skew has been measured using the B index, our *L. catta* groups showed a broad range and some of the highest skew values calculated among multi-male primate groups. Multi-male primate groups often show low or intermediate skew, with >1 male siring group offspring (e.g., northern muriqui, *Brachyteles hypoxanthus*, \(B = 0.012\) [Strier, Chaves, Mendes, Fagundes, & Di Fiore, 2011]; rhesus macaques, *Macaca mulatta*, \(B = 0.0485–0.1068\) [Dubuc, Muniz, Heistermann, Engelhardt, & Widdig, 2011; Georgiev et al., 2016; Widdig et al., 2004]; Assamese macaques, *M. assamensis*, \(B = 0.087\) [Sukmak et al., 2014]). Half of our groups showed similarly low skew, with values approximating (or lower than) these published values. On the opposite end, our group with the highest skew—a single male siring offspring across a handful of years—exceeded published values of the B index for any primate to date. Even primates known to show high paternity skew (e.g., mountain gorillas (*Gorilla beringei*), \(B = 0.107–0.432\) [Bradley et al., 2005]; white-faced capuchins (*Cebus capucinus*), \(B = 0.083–0.473\) [Muniz et al., 2010]) have B indices that are lower than that calculated for our group with the highest skew. However, we hasten to point out that this high skew is not the norm for our *L. catta* groups, and that the majority of our groups showed moderate to low paternity skew, comparable to those primate species in which reproduction is more equitably shared among males. Additionally, the confidence intervals for each group’s skew index showed much overlap, likely because the B index is sensitive to sample size [Nonacs, 2000, 2003], and our sample size of sires and infants from which each of these skew values were calculated was small.

Although a review of the various reproductive skew models (Nonacs and Hager, 2011) is beyond the scope of this paper, between the two major sets of skew models—transactional, where dominants gain a benefit from the presence of subordinates and therefore “tolerate” their reproductive activity in the group (Keller and Reeve, 1994; Reeve, 2000) and “tug-of-war”/“limited control” (Clutton-Brock, 1998; Johnstone, 2000; Reeve, Emlen, & Keller, 1998), where

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### Table 3 Comparison of groups with and without extra-group paternity

| Group | Conception year | # Males | # Females | Sex ratio | Extra-group paternity? |
|-------|-----------------|---------|-----------|-----------|------------------------|
| Black | 2003            | 3       | 5         | 0.6       | No                     |
| Black | 2005            | 8       | 4         | 2         | No                     |
| Blue  | 2003            | 5       | 8         | 0.63      | No                     |
| Blue  | 2004            | 5       | 9         | 0.56      | No                     |
| Green | 2003            | 7       | 6         | 1.17      | No                     |
| Green | 2004            | 5       | 6         | 0.83      | Yes                    |
| Orange| 2003            | 3       | 7         | 0.43      | No                     |
| Orange| 2004            | 6       | 6         | 1         | Yes                    |
| Red   | 2002            | 4       | Unknown   | Unknown   | Yes                    |
| Yellow| 2003            | 5       | 5         | 1         | No                     |
| Yellow| 2004            | 4       | 6         | 0.67      | No                     |

*Table 3* Tests evaluating whether sires of same-group offspring were significantly more likely to be unrelated than related at the half-sibling level.

| Group | Conception year | Sires                  | r     | p    |
|-------|-----------------|------------------------|-------|------|
| Black | 2001            | BM 115 & BM 173        | 0.04  | 0.07 |
| Blue  | 2003            | BM 136 & BM 140        | 0.020 | 0.066|
| Yellow| 2002            | BM 195 & BM SD         | 0.0   | 0.009|

*r*, estimated relatedness.
dominants are unable to keep subordinates from reproducing (e.g., mountain gorillas: Bradley et al. [2005])—L. catta appear to fall in the latter category. The aggressive competition that occurs among males during mating periods (Gould & Ziegler, 2007; Jolly, 1966; Koyama, 1988; Parga, 2006, 2009; Sauther, 1991) suggests that high-ranking male L. catta are not conceding reproductive units to rivals (Reeve & Keller, 2001; Vehrencamp, 1983), but rather are unable to exclude other males from mating with females and siring group offspring. Indeed, in L. catta groups with established male dominance hierarchies, there is often a high-ranking male who has first access to estrous females (but see Gould, 1994, 1996; Sauther, 1991), but this male is unable to monopolize mating with the estrous female (Koyama, 1988; Sauther, 1991). The tendency for L. catta females to mate multiply results in low male “mating skew” (Port & Kappeler, 2010) which in turn appears to translate into low paternity skew, the pattern evident for most groups in our study.

Extra-group males in this study sired approximately 21–33% of offspring. This level of extra-group paternity is comparable to that found in some other group-living primates (i.e., langurs, Semnopithecus entellus: 21% [Launhardt et al., 2001], rhesus macaques, Macaca mulatta: 25–59% [Georgiev et al., 2016; Widdig et al., 2004], chimpanzees, Pan troglodytes: 0–10.5% [Boesch, Kohou, Nene, & Vigilant, 2006; Newton-Fisher, Emery Thompson, Reynolds, Boesch, & Vigilant, 2010; Vigilant, Hofreiter, Siedel, & Boesch, 2001], Verreaux’s sifaka, Propithecus verreauxi: 17–65% [but see Kappeler & Schaller, 2008; Lawler, 2007; Lawler et al., 2003]). Thus, male visits to other groups during the breeding season (Gould, 1994; Sauther, 1991; Sussman, 1992) function as a viable mating strategy for L. catta males (Sauther & Sussman, 1993).

Extra-group paternity is considered more likely to occur where group males experience difficulty monopolizing copulations with group females, such as where estrus periods overlap (Isvaran & Clutton-Brock, 2007), or in groups having a female-biased sex ratio (i.e., Verreaux’s sifaka, Propithecus verreauxi [Lawler et al., 2003]). However, estrus synchrony is uncommon in L. catta (Pereira, 1991; Sauther, 1991), making this an insufficient explanation for extra-group paternity in our population. Furthermore, neither the number of resident males per group, the number of females per group, nor group sex ratio was significantly different between groups with and without extra-group paternity—though admittedly, our comparison is based upon very few instances (N = 3) of extra-group paternity.

A contributing factor to extra-group paternities is often female mate choice (Soltis, Thomsen, & Takenaka, 2001). Indeed, in species where extra-group paternity is considerable (e.g., rhesus macaques, Macaca mulatta, with an average of 25% extra-group paternity [Widdig et al., 2004] that can be as high as >59% in some years [Georgiev et al., 2016]), female choice has been implicated in the mating success of non-resident males. Similarly, we suggest that our findings of extra-group paternity and low skew within some groups is due to female mate choice for multiple males (Gould, 1994; Koyama, 1988; Parga, 2006; Pereira & Weiss, 1991; Sauther, 1991; Taylor, 1986) in this female dominant species (Jolly, 1966; Kappeler, 1990; Pereira et al., 1990; Sauther et al., 1999) coupled with resident males’ variable mate guarding activity (Parga, 2003, 2010; Sauther, 1991). Some L. catta males do not mate guard or mate guard for only minutes following ejaculation, whereas other males post-copulatory guard for hours; none appears to mate guard throughout the female’s entire estrus period (Parga, 2003, 2010; Sauther, 1991). The result is that females mate with both resident and extra-group males (Gould, 1994; Koyama, 1988; Sauther, 1991; Sussman, 1992), with both types of males siring offspring (this study).

It is worth noting that very few of the males sampled in this study sired offspring (21 of the 141 males). Even if a sire had been identified for each infant for whom no sire was detected (or where paternity assignments did not reach statistical significance), the total number of sires (assuming a different male for every offspring), this would still only represent 28% (40/141) of males sampled. Hence, male reproductive skew at the population level may be considerable in L. catta. Admittedly, males who were not identified as sires may have sired offspring in the time period before or after this study, especially as males of this species frequently disperse between groups (which can include into and out of study areas) every few years (Koyama, Nakamichi, Ichino, & Takahata, 2002; Sussman, 1992). Furthermore, no male in this population sired offspring in >1 group per year, although some males sired >1 infant within a single group per year. Although we lack data on mating season dynamics, this pattern of paternity suggests that males may only be able to target a single group successfully during the mating season for reproductive activities—whether their own or another—but not both. This finding underscores the temporal limitations of male mating effort, and the difficulty for male primates of juggling competing activities while pursuing mating opportunities (Alberts, Altmann, & Wilson, 1996).

In conclusion, paternity skew in our L. catta population varied among groups, with some having high to moderate paternity skew, but most groups showing low paternity skew; non-resident males also occasionally sired offspring. Still to be determined is the link between specific behavioral male mating strategies in this species and paternity success. Unfortunately, we did not consistently have male dominance rank data available for groups in the study, so whether high-ranking males show greater reproductive success than other males is still an open question. Studies in non-wild L. catta populations suggest that both dominant males and novel males (new immigrants) show paternity success (Pereira & Weiss, 1991; White et al., 2007), but more sampling is needed to determine whether such males have superior reproductive success in wild populations. Additionally, how mating dynamics differ in groups having high versus low paternity skew is currently unknown and is a topic for future research.

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REFERENCES

Alberts, S. C., Altmann, J., & Wilson, M. L. (1996). Mate guarding
constrains foraging activity of male baboons. Animal Behaviour, 51,
1269–1277.

Altmann, J. (1979). Age cohorts as paternal sibships. Behavioral Ecology and
Sociobiology, 6, 161–164.

Altmann, S. A. (1962). A field study of the sociobiology of the rhesus
monkey, Macaca mulatta. Annals of the New York Academy of Sciences,
102, 338–435.

Boesch, C., Kohou, G., Nene, H., & Vigilant, L. (2006). Male competition
and paternity in wild chimpanzees of the Tai forest. American Journal of
Physiological Anthropology, 130, 103–115.

Bradley, B. J., Robbins, M. M., Williamson, E. A., Steklis, H. D., Steklis, N. G.,
Eckhardt, N., Boesch, C., & Vigilant, L. (2005). Mountain gorilla tug-of-
war: Silverbacks have limited control over reproduction in multilple
groups. Proceedings of the National Academy of Sciences United States of
America, 102, 9418–9423.

Clutton-Brock, T. H. (1998). Reproductive skew, concessions and limited
control. Trends in Ecology & Evolution, 13, 288–292.

Csillery, K., Johnson, T., Berald, D., Clutton-Brock, T., Coltman, D.,
Hansson, B., Spong, G., & Pemberton, J. M. (2006). Performance of
marker-based relatedness estimators in natural populations of outbred
vertebrates. Genetics, 173, 2091–2101.

Dubuc, C., Muniz, L., Heistermann, M., Engelhardt, A., & Widdig, A. (2011).
Testing the priority-of-access model in a seasonally breeding primate
species. Behavioral Ecology and Sociobiology, 65, 1615–1627.

Georgiev, A. V., Christie, D., Rosenfeld, K. A., Ruiz-Lambides, A. V.,
Maldonado, E., Thompson, M. E., & Maestripieri, D. (2016). Breaking the
succession rule: The costs and benefits of an alpha-status take-over by
an immigrant rhesus macaque on Cayo Santiago. Behaviour, 153,
325–351.

Gould, L. (1994). Patterns of affiliative behavior in adult male ringtailed lemurs
(Lemur catta) at the Beza-Mahafaly Reserve, Madagascar (dissertation). St.
Louis, MO: Washington University. 253 p.

Gould, L. (1996). Male-female affiliative relationships in naturally occurring
ringtailed lemurs (Lemur catta) at the Beza-Mahafaly Reserve,
Madagascar. American Journal of Primatology, 39, 63–78.

Gould, L. (1997). Intermale affiliative behavior in ringtailed lemurs (Lemur
catta) at the Beza-Mahafaly Reserve, Madagascar. Primates, 38, 15–30.

Gould, L., & Ziegler, T. E. (2007). Variation in fecal testosterone levels, inter-
male aggression, dominance rank and age during mating and post-
mating periods in wild adult male ring-tailed lemurs (Lemur catta).
American Journal of Primatology, 69, 1325–1339.

Gould, L., Ziegler, T. E., & Wittwer, D. J. (2005). Effects of reproductive and
social variables on fecal glucocorticoid levels in a sample of adult male
ring-tailed lemurs (Lemur catta) at the Beza Mahafaly Reserve,
Madagascar. American Journal of Primatology, 67, 5–23.

Isvaran, K., & Clutton-Brock, T. (2007). Ecological correlates of extra-group
paternity in mammals. Proceedings of the Royal Society, (London) B, 274,
219–224.

Jack, K. M., & Fedigan, L. M. (2006). Why be alpha male? Dominance and
reproductive success in wild white-faced capuchins (Cebus capucinus).
In A. Estrada, P. A. Garber, M. S. M. Pavelka, & L. Luecke (Eds.), New
perspectives in the study of Mesoamerican primates: Distribution, ecology,
behavior, and conservation (pp. 367–386). New York, NY: Springer.

Jekielek, J., & Strobeck, C. (1999). Characterization of polymorphic brown
lemur (Eulemur fulvus) microsatellite loci and their amplification in the
family Lemuridae. Molecular Ecology, 8, 901–903.

Johnstone, R. A. (2000). Models of reproductive skew: A review and
synthesis. Ethology, 106, 5–26.

Jolly A. (1966). Lemur behavior: A Madagascar field study. Chicago, IL:
University of Chicago Press. 187 p.

Kalinowski, S. T., Wagner, A. P., & Taper, M. L. (2006). ML-RELATE: A
computer program for maximum likelihood estimation of relatedness and
relationship. Molecular Ecology Notes, 6, 576–579.

Kalinowski, S. T., Taper, M. L., & Marshall, T. C. (2007). Revising how
the computer program CERVUS accommodates genotyping error
increases success in paternity assignment. Molecular Ecology, 16,
1099–1006.

Kappeler, P. M. (1990). Female dominance in Lemur catta: More than just
female feeding priority? Folia Primatologica, 55, 92–95.

Kappeler, P. M., & Port, M. (2008). Mutual tolerance or reproductive
competition? Patterns of reproductive skew among male redfronted
lemurs (Eulemur fulvus rufus). Behavioral Ecology and Sociobiology, 62,
1477–1488.

Kappeler, P. M., & Schaffler, L. (2008). The lemur syndrome unresolved:
Extreme male reproductive skew in sifakas (Propithecus verreauxi), a
sexually monomorphic primate with female dominance. Behavioral
Ecology and Sociobiology, 62, 1007–1015.

Keller, L., & Reeve, H. K. (1994). Partitioning of reproduction in animal
societies. Trends in Ecology & Evolution, 9, 98–102.

Koyama, N. (1988). Mating behavior of ring-tailed lemurs (Lemur catta) at
Berenty, Madagascar. Primates, 29, 163–175.

Koyama, N., Nakamichi, M., Ichino, S., & Takahata, Y. (2002). Population and
social dynamics changes in ring-tailed lemur troops at Berenty,
Madagascar between 1989 and 1999. Primates, 43, 291–314.

Kutsukake, N., & Nunn, C. L. (2006). Comparative tests of reproductive
skew in male primates: The roles of demographic factors and incomplete control.
Behavioral Ecology and Sociobiology, 60, 695–706.

Launhardt, K., Borries, C., Hardt, C., Epplen, J. T., & Winkler, P. (2001).
Paternity analysis of alternative male reproductive routes among the
langurs (Semnopithecus entellus) of Ramnagar. Animal Behaviour, 61,
53–64.

Lawler, R. R. (2007). Fitness and extra-group reproduction in male Verreaux’s sifaka: An analysis of reproductive success from
1989–1999. American Journal of Physical Anthropology, 132, 267–277.

Lawler, R. R., Richard, A. F., & Riley, M. A. (2001). Characterization and
screening of microsatellite loci in a wild lemur population (Propithecus
verreauxi verreauxi). American Journal of Primatology, 55, 253–259.
Lawler, R. R., Richard, A. F., & Riley, M. A. (2003). Genetic population structure of the white sifaka (Propithecus verreauxi verreauxi) at Beza Mahafaly Special Reserve, southwest Madagascar (1992–2001). Molecular Ecology, 12, 2307–2317.

Marshall, T. C., Slate, J., Kruuk, L. E. B., & Pemberton, J. M. (1998). Statistical confidence for likelihood-based paternity inference in natural populations. Molecular Ecology, 7, 639–655.

Merenlender, A. M. (1993). The effects of sociality on the demography and genetic structure of Lemur fulvus rufus (polygamous) and Lemur rubriventer (monogamous) and the conservation implications [dissertation]. New York, NY: University of Rochester. 166 p.

Muniz, L., Perry, S., Manson, J. H., Gilkenson, H., Gros-Louis, J., & Vigilant, L. (2010). Male dominance and reproductive success in wild white-faced capuchins (Cebus capucinus) at Lomas Barbudal, Costa Rica. American Journal of Primatology, 72, 1118–1130.

Nei, M. (1978). Estimation of average heterozygosity and genetic distance from a small number of individuals. Genetics, 89, 583–590.

Newton-Fisher, N. E., Emery Thompson, M., Reynolds, V., Boesch, C., & Vigilant, L. (2010). Paternity and social rank in wild chimpanzees (Pan troglodytes) from the Budong Forest, Uganda. American Journal of Physical Anthropology, 142, 417–428.

Nonacs, P. (2000). Measuring and using skew in the study of social behavior and evolution. American Naturalist, 156, 577–589.

Nonacs, P. (2003). Measuring the reliability of skew indices: Is there one best index? Animal Behaviour, 65, 615–627.

Nonacs, P., & Hager, R. (2011). The past, present and future of reproductive skew theory and experiments. Biological Reviews, 86, 271–298.

Ostner, J., Nunn, C. L., & Schülke, O. (2008). Female reproductive synchrony predicts skewed paternity across primates. Behavioral Ecology, 19, 1150–1158.

Parga, J. A. (2003). Copulatory plug displacement evidences sperm competition in Lemur catta. International Journal of Primatology, 24, 889–899.

Parga, J. A. (2006). Male mate choice in Lemur catta. International Journal of Primatology, 27, 107–131.

Parga, J. A. (2009). Dominance rank reversals and rank instability among male Lemur catta: The effects of female behavior and ejaculation. American Journal of Physical Anthropology, 138, 293–305.

Parga, J. A. (2010). Male post-ejaculatory mounting in the ring-tailed lemur (Lemur catta): A behavior solicited by females? Ethology, 116, 832–842.

Parga, J. A., Sauther, M. L., Cuozzo, F. P., Yousouf Jacky, I. A., & Lawler, R. R. (2012). Evaluating ring-tailed lemurs (Lemur catta) from southwestern Madagascar for a genetic population bottleneck. American Journal of Physical Anthropology, 147, 21–29.

Parga, J. A., Sauther, M. L., Cuozzo, F. P., Yousouf Jacky, I. A., Gould, L., Sussman, R. W., Lawler, R. R., & Pastorini, J. (2015). Genetic evidence for male and female dispersal in wild Lemur catta. Folia Primatologica, 86, 66–75.

Pastorini, J., Fernandez, P., Melnick, D. J., & Forstner, M. R. J. (2004). Isolation of 10 microsatellite markers for mongoose lemurs (Eulemur mongoz). Molecular Ecology Notes, 4, 67–69.

Pastorini, J., Fernandez, P., Forstner, M. R. J., & Melnick, D. J. (2005). Characterization of new microsatellite loci for the ring-tailed lemur (Lemur catta). Molecular Ecology Notes, 5, 149–151.

Pastorini, J., Sauther, M. L., Sussman, R. W., Gould, L., Cuozzo, F. P., Fernandez, P., Nievergelt, C. M., & Mundy, N. I. (2015). Comparison of the genetic variation of captive ring-tailed lemurs with a wild population in Madagascar. Zoo Biology, 34, 463–472.

Perea, M. E. (1991). Asynchrony within estrus synchrony among ringtailed lemurs (Primates:Lemuridae). Physiology & Behavior, 49, 47–52.

Perea, M. E., & Weiss, M. L. (1991). Female mate choice, male migration, and the threat of infanticide in ringtailed lemurs. Behavioral Ecology and Sociobiology, 28, 141–152.

Perea, M. E., Kaufman, R., Kappeler, P. M., & Overdorff, D. J. (1990). Female dominance does not characterize all of the Lemuridae. Folia Primatologica, 55, 96–103.

Port, M., & Kappeler, P. M. (2010). The utility of reproductive skew models in the study of male primates, a critical evaluation. Evolutionary Anthropology, 19, 46–56.

Reeve, H. K. (2000). A transactional theory of within-group conflict. American Naturalist, 155, 365–382.

Reeve, H., & Keller, L. (2001). Tests of reproductive-skew models in social insects. Annual Review of Entomology, 46, 347–385.

Reeve, H. K., Emlen, S. T., & Keller, L. (1998). Reproductive sharing in animal societies: Reproductive incentives or incomplete control by dominant breeders? Behavioral Ecology, 9, 267–278.

Sauther, M. L. (1991). Reproductive behavior of free-ranging Lemur catta at Beza Mahafaly Special Reserve, Madagascar. American Journal of Physical Anthropology, 84, 463–477.

Sauther, M. L., & Sussman, R. W. (1993). A new interpretation of the social organization and mating system of the ringtailed lemur (Lemur catta). In P. M. Kappeler, & J. U. Ganzhorn (Eds.), Lemur social systems and their ecological basis (pp. 111–121). New York, NY: Plenum Press.

Sauther, M. L., Fish, K. D., Cuozzo, F. P., Miller, D. S., Hunter-Ishikawa, M., & Culbertson, H. (2003). Patterns of health, disease, and behavior among wild ring-tailed lemurs, Lemur catta: Effects of habitat and sex. In A. Jolly, R. W. Sussman, & N. Koyama (Eds.), Ringtailed lemur biology: Lemur catta in Madagascar (pp. 313–331). New York, NY: Springer Science + Business Media.

Sauther, M. L., Sussman, R. W., & Gould, L. (1999). The socioecology of the ringtailed lemur: Thirty-five years of research. Evolutionary Anthropology, 8, 120–132.

Shuster, S. M., & Wade, M. J. (2003). Mating systems and strategies. Princeton: Princeton University Press. 533 p.

Solits, J., Thomsen, R., & Takenaka, O. (2001). The interaction of male and female reproductive strategies and paternity in wild Japanese macaques, Macaca fuscata. Animal Behaviour, 62, 485–494.

Strier, K. B., Chaves, P. B., Mendes, S. L., Fagundes, V., & Di Fiore, A. (2011). Low paternity skew and the influence of maternal kin in an egalitarian, patrilocal primate. Proceedings of the National Academy of Sciences United States of America, 108, 18915–18919.

Sukmak, M., Wajjwalku, W., Ostner, J., & Schülke, O. (2014). Dominance rank, female reproductive synchrony, and male reproductive skew in wild Assamese macaques. Behavioral Ecology and Sociobiology, 68, 1097–1108.

Sussman, R. W. (1992). Male life history and intergroup mobility among ringtailed lemurs (Lemur catta). International Journal of Primatology, 13, 395–413.

Sussman, R. W., & Ratsirarson, J. (2006). Beza Mahafaly Special Reserve: A research site in Southwestern Madagascar. In A. Jolly, R. W. Sussman, N. Koyama, & H. Rasamimanana (Eds.), Ringtailed lemur biology: Lemur catta in Madagascar (pp 43–51). New York, NY: Springer Science + Business Media.

Taylor, L. L. (1986). Kinship, dominance, and social organization in a semi-free ranging group of ringtailed lemurs (Lemur catta) [dissertation]. St. Louis, MO: Washington University. 218 p.

Taylor, L., & Sussman, R. W. (1985). A preliminary study of kinship and social organization in a semi-free-ranging group of Lemur catta. International Journal of Primatology, 6, 601–614.
Van Horn, R. C., Altmann, J., & Alberts, S. C. (2008). Can’t get there from here: Inferring kinship from pairwise genetic relatedness. *Animal Behaviour*, 75, 1173–1180.

dvan Oosterhout, C., Hutchinson, W. F., Wills, D. P. M., & Shipley, P. (2004). MICRO-CHECKER: Software for identifying and correcting genotyping errors in microsatellite data. *Molecular Ecology Notes*, 4, 535–538.

Vehrencamp, S. L. (1983). A model for the evolution of despotic versus egalitarian societies. *Animal Behaviour*, 31, 667–682.

Vigilant, L., Hofreiter, M., Siedel, H., & Boesch, C. (2001). Paternity and relatedness in wild chimpanzee communities. *Proceedings of the National Academy of Sciences United States of America*, 98, 12890–12895.

White, F. J., Overdorff, D. J., Keith-Lucas, T., Rasmussen, M. A., Kallam, W. E., & Forward, Z. (2007). Female dominance and feeding priority in a prosimian primate: Experimental manipulation of feeding competition. *American Journal of Primatology*, 69, 295–304.

Widdig, A. (2013). The impact of male reproductive skew on kin structure and sociality in multi-male groups. *Evolutionary Anthropology*, 22, 239–250.

Widdig, A., Bercovitch, F. B., Streich, W. J., Sauermann, U., Nurnberg, P., & Krawczak, M. (2004). A longitudinal analysis of reproductive skew in male rhesus macaques. *Proceedings of the Royal Society, (London) B*, 271, 819–826.

Wimmer, B. (2000). Untersuchung der Paarungssysteme und der Populationsstruktur von Lemuren an Coquerels Zwergmaki (Mirza coquereli), dem grauen Mauslemur (Microcebus murinus), dem Rotstirnmaki (Eulemur fulvus rufus) und dem Larvensifaka (Propithecus verreauxi verreauxii) [PhD thesis]. Munich, Germany: Universität München.

Wimmer, B., & Kappeler, P. M. (2002). The effects of sexual selection and life history on the genetic structure of redfronted lemur, *Eulemur fulvus rufus*, groups. *Animal Behaviour*, 64, 557–568.

Zaonarivelo, J. R., Andriantompohavana, R., Shore, G. E., Engberg, S. E., McGuire, S. M., Louis, Jr., E. E., & Brenneman, R. A. (2007). Characterization of 21 microsatellite marker loci in the ring-tailed lemur (*Lemur catta*). *Conservation Genetics*, 8, 1209–1212.