Insight into population structure of *Mycobacterium tuberculosis* isolates in the multiethnic province of Alborz, Iran

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ABSTRACT

Background and Objectives: Genetic diversity of *Mycobacterium tuberculosis* clinical isolates from tuberculosis patients in the multiethnic province of Alborz, Iran was assessed.

Materials and Methods: A total of 17 isolates in the period of 2012-2013 were collected and subjected to a Multiple-locus variable-number tandem repeat (VNTR) analysis (MLVA) consisted of 6 variable numbers of tandem repeats (VNTRs) including ETR-A, ETR-B, ETR-C, ETR-D, ETR-E, ETR-F, 5 Mycobacterial Interspersed Repetitive Units including MIRU10, MIRU16, MIRU26, MIRU39, MIRU40, and 1 Queen University of Belfast locus, QUB11.

Results: This classified all isolates into 17 distinct MIRU-VNTR types, a reflection of a highly heterogenic population. Within the 12 used VNTR loci, ten proved highly or moderately discriminant according to the calculated HGD1 scores. No cluster of isolates was identified in the study panel, giving a clustering rate of 0%, several events of SVL (N=5) and DVL (N=4) and TVL (N=3) were detected.

Conclusion: The greater heterogeneity observed here by MLVA-VNTR analysis is most likely due to limited background data in the study region rather than a genuine more heterogeneous population compared to other provinces of the country.

Keywords: *Mycobacterium tuberculosis*; Population; Genotype; Genetic heterogeneity; Polymorphism; Epidemiologic; Locus

INTRODUCTION

Tuberculosis (TB), known to mankind since antiquity, has yet remained a serious threat to the public health in low- and middle-income countries. Fig-

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such as IS6110-RFLP, large sequence polymorphism (LSP), spoligotyping, Single Nucleotide Polymorphism (SNP), mycobacterial interspersed repetitive unit-variable number tandem repeat (MIRU-VNTR) typing and more recently Whole-genome sequencing. According to these findings, MTC strains are highly clonal bacteria representing relatively static genomic content where up to 99.9% nucleotide sequence identity is typically a normal observation (2). Until now seven globally important lineages of human-adapted MTC strains have been defined including lineages scattered globally (e.g. lineages 2 and 4) and other lineages (5 and 6) that tend to operate locally. This is believed to be an indication of the difference in transmission and drug resistance properties of strains (3).

The molecular epidemiology of MTC in human has recently been the subject of numerous studies by Iranian workers (4-7) though, need for further epidemiological analysis of actively circulating MTC genotypes in Iran seems not fulfilled as yet.

The Alborz province, located in the north-west of the country’s capital, Tehran is the latest parliamentary-approved province of Iran formed by division of Tehran province in 2010. Surrounded by Tehran, Qazvin, Mazandaran and Markazi provinces, as many as 2.5 million people lived here in 2018. A typically multiethnic region with its population comprised of almost all the country’s ethnicities and foreign nationals dominated by Afghan and Iraqi settlers. Alborz is represented by a relatively higher TB burden at the national level according to the official medical records. The genetic diversity of MTC population in this province has remained largely unknown so, to generate baseline data, in the present work, we used Multiple-locus variable-number tandem repeat (VNTR) analysis (MLVA).

MATERIALS AND METHODS

Ethics statement. All the study participants were briefed with the research objectives and its procedures with written informed consent obtained from all the subjects. Ethical approval required for this retrospective genotyping work was issued by the relevant authorities at the Ethics Committee of the Alborz University of Medical Sciences.

Collection of samples and isolation. Twenty sputum specimens were collected from randomly selected 20 pulmonary TB-suspected outpatients. These were visiting patients to the university hospitals of Alborz from December 2012 to October 2013. Demographic data of patients were collected along with their consent through questionnaires filled out at the clinic when specimens were taken.

Treating of the samples by Petroff’s method. The collected specimens were transferred to Razi institute in Harāṣar, Karaj where they were subjected to Petroff’s protocol for digestion/decontamination. For bacterial culture, processed (digested and decontaminated) specimens were used to inoculate slopes of plain (glycerinated) and Lowenstein–Jensen medium (LJ) supplemented with sodium pyruvate and glycerol. Incubation of cultured Falcon tubes at 37°C continued for 12 weeks with weekly close inspection in search for traces of bacterial growth. Slopes bearing visual properties of MTC bacterial mass were tested by Acid-fast staining microscopy examination (AFB).

DNA extraction and molecular amplification. Bacterial colonies from MTC-suspected cultures were scraped from the slopes and re-suspended in 200 µL of TB lysis® Solution (SinaClon BioScience, Iran) in microfuge tubes equipped with O-rings. Microtubes were heat-treated at 96°C for 30 min in a boiling water bath. The inactivated bacterial suspension was centrifuged (9,750 g; 10 min) and the DNA holding supernatant was retrieved into two new tubes including one that was stored at -20°C and the second that was used for PCR experiments. The primers were synthesized by Macrogen, (Seoul, South Korea). The PCR mixtures were prepared using 2×Taq Master Mix (Ampliquor, Denmark).

PCR-16S rRNA, PCR-IS6110. Two 543 and 245 bp long fragments of the 16S rRNA gene and IS6110 marker from isolates were assessed according to the Huard (8) and McHugh (9) PCR methods, respectively.

RD-Typing. Four individual PCRs targeting specific portions of RD1, RD4, RD9, and RD12 were performed as previously described by Warren et al. to differentiate members of Mycobacterium tuberculosis complex (10).

MLVA-VNTR typing and data analysis. A ML-VA-VNTR typing method based on the 12 broadly-used loci namely, ETR-A, ETR-B, ETR-C, ETR-D,
ETR-E, ETR-F, MIRU10, MIRU16, MIRU26, MIRU39, MIRU40, and QUB11b, was conducted following the PCR protocol described elsewhere. The number of tandem repeats at the selected loci was determined through visual comparison of the size of the amplification products in Electrophoresis gels against the known size of bands from both the DNA step ladder and those from already-identified bands of *M. tuberculosis* H37Rv genome. Loci with failure PCR amplification or undefined results were re-tested for three times when sizing of the given locus was eventually possible or it was scored a zero.

The allele copy numbers represented in digital Binary-format where each digit signified the number of copies at a particular locus. These were added manually to a Microsoft Excel spreadsheet and used for drawing dendrogram and the minimum spanning tree (MST) to enable classification of the isolates and phylogenetic analysis. In order to determine the individual discriminatory power of the 12 tandem repeat loci, the Nei’s diversity index (D) was calculated as $D = 1 - \Sigma\text{ (allele frequency)}^2$ and the loci were grouped into highly (HGD>0.6), moderately (HGD>0.3) or poorly discriminative (HGD<0.3). Similarly, the Hunter–Gaston discriminatory index (HGD) was used to determine the combinational discriminatory power of the 12 loci on the whole as a genotyping system (11). The HGD represents the average prob- ability that the typing system will assign a different type to two unrelated strains that have been randomly sampled in the microbial population of a given taxon. The calculations were conducted online by using the calculator tool available at the website http://insilico.ehu.es/mini_tools/discriminatory_power/ (12).

For clustering analysis, the assessment of genetic relationships and comparison of strain genotypes, the BioNumerics 7.6 software (Applied Maths, Sint-Martens-Latem, Belgium) was consulted where similarity matrices applying the unweighted pair-grouping method with arithmetic averages algorithm (UPGMA) was used. A minimum spanning tree (MST) was constructed to represent an undirected network of MIRU-VNTR genotypes that are linked together where nearest neighbors are displayed by the smallest possible linkages.

To calculate the recent transmission rate, the N-1 method was employed using the CR=(N-C)/N equilibrium, where N is the total number of cases in the sample, c stands for the number of clusters, and N represents the total number of clustered cases (13). Isolates representing identical MIRU-VNTR profiles were treated as members of the same cluster.

**RESULTS**

The study panel included 20 patients with Iranian nationality. The man to woman ratio was roughly 2:1 (12 males versus 5 females). In bacterial culture, preferential growth was observed for the LJ slopes in all the 20 isolates. All of the 20 isolates, produced the expected 543 bp long PCR target fragment of the 16S rRNA gene proving their identity as *Mycobacterium* species. Besides, 17 isolates produced the expected 245 bp fragment of IS6110 locus and were therefore identified as *M. tuberculosis* complex. This was re-confirmed by RD-typing test.

The allele numbers at each MIRU-VNTR locus ranged between 2 (for loci ETR-B and ETR-D) and 5 (for locus MIRU16). The allelic diversity (h) was in general high with the exception of locus ETR-D that showed 3 repeat units in 99% of isolates, yielding h=0.10 indicated that locus ETR-D was highly conserved.

Successful amplification of 12 loci among the 17 isolates, gave 17 distinct non-clustered MIRU-VNTR profiles.

Comparative analysis of the identified MIRU-VNTR patterns helped to assess relative evolutionary rates of the individual loci. This was analyzed through calculating frequency of these loci in SLVs (Single-Locus Variables), DLVs (Double-locus Variable), or TLVs (Treble-Locus Variable) within the isolates. Out of the 17 isolates, 17 clusters were identified among which 9 were SLV and 13 were DLV and 17 were TLV (Table 1).

Using the minimum spanning tree algorithm, the MIRU-VNTR-based genetic relationships among the isolates in the study panel along with those from Khorasan Razavi (N=93) and Markazi (N=53) provinces were visualized. This was conducted in a way that clonal complexes were defined arbitrarily as groups of isolates gathering according to similarity level of their MIRU-VNTR profiles with a maximal tolerance of three locus difference (Fig. 1).

**DISCUSSION**

The current work seems to be is the first study...
Table 1. Epidemiological and MLVA genotyping details of 17 isolates in the present study.

| Isolate order | Isolate ID | Province | Sex | Nationality | ETR A | ETR B | ETR C | ETR D | ETR E | ETR F | MIRU 10 | MIRU 16 | Min r26 | Min r39 | Min r40 | QUB11 |
|---------------|------------|----------|-----|-------------|-------|-------|-------|-------|-------|-------|---------|---------|--------|--------|--------|--------|
| 1             | ALZ 0043   | Alborz   | F   | Iranian     | 2     | 1     | 4     | 2     | 3     | 2.2   | 2       | 3.2     | 6       | 2       | 2       | 2       |
| 2             | ALZ 0038   | Alborz   | F   | Iranian     | 2     | 1     | 4     | 3     | 3     | 2.2   | 2       | 3.2     | 6       | 2       | 2       | 2       |
| 3             | ALZ 0045   | Alborz   | M   | Iranian     | 2     | 1     | 4     | 3     | 3     | 2.2   | 2       | 3.2     | 6       | 2       | 2       | 2       |
| 4             | ALZ 0040   | Alborz   | M   | Iranian     | 2     | 2     | 4     | 3     | 3     | 2.3   | 3       | 2.2     | 6       | 1       | 1       | 3       |
| 5             | ALZ 0033   | Alborz   | M   | Iranian     | 3     | 1     | 3     | 3     | 3     | 2.2   | 2       | 3.2     | 6       | 1       | 2       | 2       |
| 6             | ALZ 0042   | Alborz   | M   | Iranian     | 3     | 1     | 4     | 3     | 3     | 2.1   | 3       | 3.2     | 6       | 2       | 2       | 2       |
| 7             | ALZ 0036   | Alborz   | F   | Iranian     | 3     | 1     | 4     | 3     | 3     | 2.2   | 2       | 3.2     | 5       | 2       | 2       | 2       |
| 8             | ALZ 0034   | Alborz   | M   | Iranian     | 3     | 1     | 4     | 3     | 3     | 2.2   | 2       | 3.2     | 6       | 2       | 2       | 2       |
| 9             | ALZ 0046   | Alborz   | M   | Iranian     | 3     | 1     | 4     | 3     | 3     | 2.2   | 2       | 4.2     | 6       | 2       | 2       | 2       |
| 10            | ALZ 0041   | Alborz   | F   | Iranian     | 3     | 1     | 4     | 3     | 4     | 2.2   | 2       | 2.2     | 5       | 2       | 2       | 2       |
| 11            | ALZ 0031   | Alborz   | F   | Iranian     | 3     | 1     | 4     | 3     | 4     | 2.2   | 2       | 3.2     | 5       | 1       | 3       | 2       |
| 12            | ALZ 0039   | Alborz   | M   | Iranian     | 3     | 1     | 5     | 3     | 3     | 1.2   | 2       | 3.2     | 5       | 2       | 2       | 2       |
| 13            | ALZ 0050   | Alborz   | M   | Iranian     | 3     | 1     | 5     | 3     | 4     | 2.2   | 2       | 2.2     | 5       | 2       | 2       | 2       |
| 14            | ALZ 0049   | Alborz   | M   | Iranian     | 3     | 2     | 3     | 3     | 5     | 2.3   | 5       | 4.2     | 7       | 3       | 2       | 2       |
| 15            | ALZ 0035   | Alborz   | M   | Iranian     | 3     | 2     | 4     | 3     | 3     | 2.3   | 3       | 1.2     | 5       | 2       | 3       | 3       |
| 16            | ALZ 0032   | Alborz   | M   | Iranian     | 4     | 2     | 2     | 3     | 5     | 2.3   | 6       | 5.2     | 7       | 3       | 2       | 1       |
| 17            | ALZ 0047   | Alborz   | M   | Iranian     | 4     | 2     | 3     | 3     | 5     | 2.3   | 5       | 4.2     | 7       | 2       | 2       | 2       |

No of alleles 3 2 4 2 3 4 4 5 3 3 3 3

Nei’s diversity index 0.51 0.33 0.53 0.11 0.59 0.56 0.47 0.69 0.63 0.46 0.30 0.30

Hunter-Guston (HG) diversity index

| SLV (N=9) | 1.2, 2&3, 2&8, 3&9, 5&7, 6&7, 7&8, 8&9, 10&13 |
| DLV (N=13) | 1&3, 1&8, 6&8, 6&10, 6&12, 2&7, 2&9, 3&8, 5&8, 7&9, 7&10, 7&12, 14&17 |
| TLV (N=17) | 1&7, 1&9, 2&5, 2&6, 2&10, 3&7, 5&9, 5&11, 6&9, 7&11, 7&13, 8&10, 8&12, 9&10, 10&11, 11&13, 12&13 |

on population genetics of MTC in Alborz province using MIRU-VNTR strategy. Detection of 17 distinct genotypes among the 17 isolates in the study panel displayed a high proportion of non-clustered and orphan isolates. A trivial explanation for this observation could be that there is a lack of previous data on genetic structure of MTC in the region. In fact, as shown by the published literature, in populations where extensive genetic studies have been already conducted, observing a lower number of orphan isolates is largely frequent (14, 15). When the MLVA-VNTR results from a further 93 isolates collected in Khorasan Razavi Province as well as 53 isolates collected in Markazi province combined with the results of present work, a number of clustered isolates including those from Alborz were observed. This is supporting evidence on the fact that higher frequency of orphan genotypes in this study is a consequence of less-analyzed genetics of MTC in the region.

Using the “n-l” method on the study panel to calculate Recent Transmission Rate (RTR) was in support of no association between TB cases and recent transmission, a likely indication of association with latent tuberculosis reactivation. This observation seems to be not unusual in Iran as the overall clustering rate reported by Iranian workers ranged 0.0%-75% (6, 16-18). A variety of elements including spatial and temporal dimensions of study, size of examined isolates, ethnicity coverage and socioeconomic status of enrolled patients have been noted as contributors (6).

No sign of multiple alleles was found with any of the loci or isolates under examination in the present study though previous works have displayed the existence of this phenomenon among local populations of MTC in the Iranian environment with a frequency ranging from 1.6% to 26.6% (5, 6, 19, 20). In-patient independent microevolution of discrete MLVA-VN-
TR loci and co-infection with multiple-strains are believed to be the two most relevant explanations for this observation which were not observed in the present work (6).

To visualize the evolutionary relationships of examined isolates along with those from previous studies (Khorasan Razavi province, N=93 and Markazi province, N=53) that used the same MLVA-VNTR genotyping system, a minimum spanning tree and a dendrogram were constructed (Figs. 1 and 2). A number of clonal structures were identified with Alborz isolates distributed across them showing a large genetic heterogeneity, forming several distinct complexes. Furthermore, as displayed by the circular dendrogram drawn using UPGMA method, most of the Alborz isolates scattered through the genotypes which again reiterate on heterogeneity of studied MTC population in the region.

While MIRU-VNTR still remains a broadly used protocol for genotyping at global scale, with the advent of the enabling technology of next-generation sequencing, the whole-genome sequencing (WGS) is now available to TB reference laboratories though mostly in the developed world. The currently high costs and complexity of data analysis in connection with WGS has limited its broad acceptance. Despite delivering greater specificity, the quantitative prospect comparisons in favor of WGS are yet to be completed (21). In reality, as the running costs gradually decline, WGS will very likely replace oth-
er genotyping methods in the near future. If open-source software with capability to connect WGS and MIRU-VNTR typing (so-called MIRU-profiler programs) become available then the prediction of MIRU-VNTR profiles from WGS of *M. tuberculosis* isolates will be very convenient (22). In conclusion, the present work provides baseline data on population genetic of *M. tuberculosis* strains that are operating in Alborz. a step forward for further molecular epidemiology works at local as well as national level.

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