End-tidal carbon dioxide variation after a 100- and a 500-ml fluid challenge to assess fluid responsiveness

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Abstract

Background: EtCO₂ variation has been advocated replacing cardiac output measurements to evaluate fluid responsiveness (FR) during sepsis. The ability of EtCO₂ variation after a fluid challenge to detect FR in the context of general anaesthesia has not been investigated. Forty patients were prospectively studied. They underwent general anaesthesia for major surgeries. CO was measured by transoesophageal Doppler, and EtCO₂ was recorded as well as other haemodynamic parameters [heart rate (HR), mean arterial pressure (MAP), pulse pressure (PP)] at baseline, after 100-ml fluid load over 1 min, and at the end of the 500-ml fluid load. We measured the variation of EtCO₂ at 100 (ΔEtCO₂100) and 500 ml (ΔEtCO₂500), and ROC curves were generated. A threshold for ΔEtCO₂ to predict FR was determined with receiver operating curves (ROC) analysis. The primary end point was the ability of EtCO₂ variation after a 500-ml fluid load to diagnose FR.

Results: Fifteen patients (38 %) were fluid responders. ROC analysis showed that for a threshold of 5.8 % (ΔEtCO₂500), sensitivity was 0.6 IC 95 % [0.33; 0.86] and specificity was 1.0 IC 95 % [1.0; 1.0]. An absolute increase of more than 2 mmHg of EtCO₂ is specific to diagnose fluid responsiveness (spe = 96 % [88–100 %], sens = 60 % [33–88 %], AUC = 0.80 [0.96–0.65]). HR, MAP, and PP variations and ΔEtCO₂100 did not bring information to predict or diagnose FR. During fluid challenge, the correlation between CI variation and EtCO₂ variation was r = 0.566, p < 0.001.

Conclusions: During surgery, when alveolar ventilation and CO₂ production are constant, ΔEtCO₂500 is fairly reliable to assess FR. When the variation of EtCO₂ is >5.8 %, all patients were responders, but no conclusion could be done when this variation was <5.8 %. ΔEtCO₂100 failed to predict FR.

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based on heart and lung interactions are limited by their dependence on relatively large tidal volumes, arrhythmias, or the need for an arterial line [7]. Passive leg raising (PLR) is not convenient during surgery. Mini-fluid challenges have been described as a reliable method to predict VR if cardiac output is monitored [8]. Another approach is to perform a fluid challenge and assess a posteriori its effectiveness. Even though some of published studies show that PP, HR, or MAP variations do not give valuable information about VR after volume expansion; a majority of anaesthesiologists still use these parameters to assess VR. This can be explained by the fact that cardiac output monitoring is thought to be too invasive [9]. The end-tidal carbon dioxide (EtCO₂) correlates with cardiac output in experimental settings [10]. EtCO₂ variation has been recently used to assess cardiac output variation during PLR and allows one to predict fluid responsiveness of septic patients in intensive care units [11, 12].

Measuring EtCO₂ might be a reliable method to assess the effect of a fluid challenge in the operating room when cardiac output monitoring is not available. But this strategy has not been prospectively evaluated during general anaesthesia with a population including non-septic patients. The correlation between EtCO₂ and CI has been shown to be stronger in a context of circulatory insufficiency than in a normal haemodynamic state [10]. The primary end point was the ability of EtCO₂ variation after a 500-ml fluid load to diagnose fluid responsiveness. Secondary end points were to test whether changes in EtCO₂ during volume expansion correlate with changes in cardiac output and the ability of the mini-fluid challenge (100 ml) to predict fluid responsiveness.

**Methods**

Our institution review board (Comité de Protection des Personnes Lyon Est III ref: 2013-027 B, ANSM Number ID RCB : 2013-A00729-36) approved the study protocol and waived signed informed consent. Inform consent was obtained and notified in the medical record.

**Participants**

The study was conducted over a 2.5-month period in 2013 in a secondary care university hospital in Lyon. Participants were those admitted for emergency surgery with a high risk of haemodynamic disorders.

The decision to administer a fluid bolus for volume expansion was left to the discretion of the attending anaesthesiologist.

Inclusion criteria were patient undergoing general anaesthesia with mechanical ventilation and monitored with an oesophageal Doppler. The attending anaesthesiologist decision to administer a fluid bolus for volume expansion was a prerequisite for the inclusion of the patient. The need of oesophageal Doppler monitoring was defined in our setting as patients undergoing high-risk surgeries or high-risk patients. High-risk surgeries were hip fractures, peritonitis, and abdominal haemostatic surgery. High-risk patients were patients with heart failure or significant arteriopathy. Exclusion criteria were patient refusal age <18 years, pregnancy, fluid overload before anaesthesia, patients undergoing laparoscopic surgery, and patients with no available Doppler signals or contraindications to this monitoring technique. All patients meeting the inclusion criteria were included in the study except if they refused to participate if Doppler signal was lacking, or if principal investigators were not available.

**Experimental design**

Fluid volume expansion was performed, during the surgery (after the induction of anaesthesia and placement of different monitoring devices). All the data were prospectively recorded, and fluid responsiveness was determined after the fluid bolus and calculated afterthought. The experimental design is detailed in Fig. 1. The manuscript is written to fit with the STARD statement, to allow readers to assess internal and external validity [13].

**Fluid used and administration**

Hydroxyethyl starch (Fresenius Kabi, Germany) was used. A total of 500 ml divided into one bolus of 100 ml and remaining 400 ml were administered. The first fluid bolus of 100 ml was administered at a rate of 100 ml in 1 min (verified using a stopwatch) using a 50-ml syringe. After 1 min, the remaining 400 ml was administered over 11 min. During the 13 min of fluid administration, there was no modification of the ventilatory settings. Also, no bolus or change in the rate of administration of vasoconstrictor and anaesthetic agents was performed during this period.

**Cardiac output monitoring**

Patients were routinely monitored with a transoesophageal Doppler probe (HemoSonic™ 100, ARROW International®, 61 mm length, <7-mm section, 5 MHz transducer) which enables recording of continuous descending aortic blood velocity, aortic diameter, aortic ejection volume, aortic blood flow, and other valuable parameters such as acceleration of the flow (Acc) and systolic flow time corrected by heart rate (Ftc). Acc has been described to reflect inotropic change [14], and Ftc has been incorrectly described as a measure of left ventricular preload [15]. An algorithm enables cardiac output computation. The same observer (MJL) performed placement of the probes for all patients.
MJL had a good experience with oesophageal monitoring with more than 2-year practice and more than 200 examinations performed. Aortic diameter was visualised and sampled by the transoesophageal probe. The Doppler probe was inserted through the mouth to the place where the maximal aortic blood flow velocity signal could be captured. The quality of the signal was assessed by the visualisation of the two walls of the aorta, and a consistent aortic diameter [16]. Doppler-derived indices were averaged on ten systolic ejections. Cardiac index was calculated as CO divided by body surface. Measurements were recorded before fluid challenges to assessing variability and the least significant change (LSC).

**EtCO2 monitoring**
The Infinity® EtCO2 Microstream SmartPod device was used to monitor EtCO2, a side stream device based on infrared absorption of a specific wavelength. This technology provides measures of EtCO2. The instantaneous values were recorded at 0, 1, and 13 min.

**Other data and monitoring**
Perioperative monitoring included continuous electrocardiogram, pulse oximetry, and non-invasive blood pressure every 5 min. Systolic, diastolic, and mean arterial pressures were recorded. Pulse pressure (PP), defined by systolic minus diastolic pressures, was calculated. Airway pressure, peak pressure, plateau pressure, respiratory rate, and tidal volume were monitored as well as partial inspiratory pressures of O2 and CO2. The end-tidal anaesthetic agent concentration of desflurane or sevoflurane was monitored to assess deepness of anaesthesia.

**Reference standard**
Patients with more than a 15 % increase in cardiac index with a 500-ml fluid load were defined as fluid responders (R), the others being non-responders (NR) This definition is consistent with many diagnosis accuracy studies of fluid responsiveness. The observer was blind from the reference standard as fluid responsiveness was determined after the end of the fluid challenge and was calculated a posteriori.

**Statistical analysis**
We calculated with the method of Obuchowsky et al. [17] that 39 patients were needed in order to detect an area under the ROC curve of 0.75 with a power of 0.9 and an alpha risk of 0.05. The ratio between responder and non-responder in the studied population was 0.77. Normal distribution was tested by the d’Agostino–Pearson test. Pairwise comparisons of values were made with the paired Student’s t test or Wilcoxon test. The two-tailed Student t test or Mann–Whitney U test was performed for comparisons between responders and non-responders. In cases of relevancy, data were expressed as variations from baseline computed as the difference between final and baseline value divided by the baseline value and expressed as ΔCI and ΔEtCO2 for CI and EtCO2 variation. Correlations were tested by the Spearman method. The relationships between variables underwent linear regression analysis method. Before volume expansion, multiple measurements were recorded during steady haemodynamic and respiratory conditions defined as no need of vasoconstrictor, fluid challenge, and also no respiratory setting modification and no spontaneous breathing detected on the respirator. To assess the reproducibility of the reference standard and the evaluated test, we performed ten successive measures during a

![Fig. 1 Experimental design. Data were collected at 0, 1, and 13 min. Mean arterial pressure (MAP), systolic arterial pressure (SAP), diastolic arterial pressure (DAP), corrected systolic flow time (Ftc), and end-tidal carbon dioxide (EtCO2)](image-url)
stable haemodynamic period. The coefficient of variation was calculated as the standard deviation divided by the mean. The precision was twice the coefficient of variation, and the LSC was computed as 1.96 times the square root time the coefficient of error [18]. The LSC was the minimum change that can be considered as a real change. Data were expressed as mean ± standard deviation (SD) or as median [interquartile range: IQR] when appropriate. ROC curves were built, and AUC was expressed as 95% confidence interval. Confidence interval was built with the “bootstrap” technique with 2000 repetitions and the same ratio between case and control.

ROC curves were then compared by the Delong test to a 0.5 built ROC curve [19]. Then, ROC curves were used to define three classes of response: negative, inconclusive, and positive. These classes were defined by the author to implement a 10% diagnosis tolerance in the analysis as it is proposed in a grey zone approach. An EtCO2 variation with a value lower than the 90% sensitivity threshold was defined as negative. An EtCO2 variation greater than the 90% specificity threshold was defined as positive. Remaining EtCO2 variations were defined as inconclusive. The proportion of the study population within these limits was calculated. Statistical analysis was performed with R Packages, referenced below [20]. Significant results were defined by a p value <0.05.

Results
All patients who met inclusion criteria were screened (Fig. 2). Patient characteristics are given in Table 1. The patients were scheduled for orthopaedic (65%) and abdominal surgery (35%). All patients underwent general anaesthesia with mechanical ventilation without spontaneous breathing at the time of the study. Twelve patients had surgery in a context of sepsis and nine had a previous history of cardiac failure (Table 1). The precision of EtCO2 was 2.2 ± 1.3%, and the LSC was 3.2 ± 0.2%. We did not deplore any adverse events as a consequence of the Doppler monitoring or EtCO2 measurements. We did not have any missing values of EtCO2 or CO at the three different times of the study.

Responders and non-responders
Fifteen patients (38%) were considered to be fluid responders after a 500-ml bolus. CI increased in all patients by 7.8 [3.1; 20.0] %, in R group by 32 [20; 42] %, and in NR groups, by 3.7 [0; 7.2] %. Distribution of EtCO2 variations in responders and non-responders is described in Fig. 3.
Description of the Variation of CI and EtCO₂, pulse pressure, heart rate, Ftc, ventilatory change and end-tidal anaesthetic agent concentration during volume expansion

Baseline haemodynamic and respiratory parameters were not significantly different between fluids responders and non-responders (Table 2). No significant differences were found for Vt and minute ventilation between before and after fluid expansion and between both groups at each moment (Table 2). The end of the trial, after the 500-ml volume expansion, showed a significant increase in CI and EtCO₂ in the responders group and the Ftc increased in both groups (Table 2). The end-tidal anaesthetic agent concentration of desflurane or sevoflurane variation between before and after the fluid challenge was 2.6 [−1.8; 11.4] % in the R group and 0 [0; 11.1] % in the NR group with no significant difference (p = 0.9).

Correlation between EtCO₂ and CI

The Spearman correlation test between CI and EtCO₂ was not significant (r = 0.178, p = 0.272). However, there was a significant correlation (r = 0.566, p < 0.001) between CI variation (ΔCI) and EtCO₂ variation (ΔEtCO₂). The slope of the linear regression was 0.172.

Ability of ΔEtCO₂ to predict fluid responsiveness after a mini-fluid challenge (100 ml)

The correlation between EtCO₂ variation and CI variation after a 100-ml fluid challenge was significant: r = 0.39, p = 0.013. The change in EtCO₂ after a 100-ml infusion provided an AUC-ROC = 0.74 [0.60; 0.89]; threshold = 3.0 %; sens = 33 [13–60] %, spe = 100 [100–100] %.

Ability of ΔEtCO₂ to diagnose fluid responsiveness after a 500-ml volume expansion

The area under the receiver operating characteristic curve (ROC-AUC), sensitivity, specificity, positive predictive value, and negative predictive value likelihood ratios are given in Table 3. ΔEtCO₂ after a 500-ml expansion (ΔEtCO₂500) was able to diagnose fluid responsiveness with a threshold of 5.8 %. In a pragmatic approach, an absolute increase of more than 2 mmHg of EtCO₂ can diagnose fluid responsiveness (spe = 96 [88–100] %, sens = 60 [33–88] %, AUC = 0.80 [0.96–0.65]). The AUC-ROC for heart rate variation (ΔHR₅₀₀), MAP

Table 1 Characteristics of the population studied

| Characteristics                      | N = 40 |
|--------------------------------------|--------|
| Age (years)                          | 75 [57–92] |
| Male sex                             | 17 (43) |
| Weight (kg)                          | 69 [51–86] |
| Height (cm)                          | 166 [156–177] |
| BMI (kg m⁻²)                         | 25 [20–30] |
| Surgery                              |        |
| Orthopaedic surgery                  | 26 (65) |
| General surgery                      | 14 (35) |
| Sepsis                               | 12 (30) |
| Antecedent                           |        |
| ASA score                            | 2.0 [1.6–2.8] |
| Atrial fibrillation                  | 5 (13) |
| Cardiac failure                      | 9 (23) |
| COPD                                 | 4 (10) |
| Treatment prior surgery              |        |
| Beta blockers                        | 12 (30) |
| Other antiarrhythmic agents          | 5 (13) |
| Ventilation                          |        |
| Tidal volume (ml kg⁻¹)               | 6.6 [4.9–8.4] |
| PEEP (cmH₂O)                         | 5 [4–7] |
| FiO₂ (%)                             | 58 [50–70] |
| EtCO₂ (mmHg)                         | 31 [28–34] |
| SpO₂ (%)                             | 99 [97–100] |
| Anaesthesia protocol                 |        |
| Sufentanil                           | 39 (98) |
| Propofol TIVA                        | 5 (13) |
| Halogenated                          | 35 (88) |
| Curare                                | 16 (40) |
| Regional anaesthesia                 | 14 (35) |
| Vasoconstrictors                     |        |
| Vasoconstrictor use                  | 11 (28) |
| Phenylephrine (µg kg⁻¹ min⁻¹)        | 0.2 [0.0–0.3] |

Data are expressed as median and (25th–75th) or as number and proportion of patients (n = 40). Body mass index (BMI), chronic obstructive pulmonary disease (COPD), positive end-expiratory pressure (PEEP), fraction of inspired oxygen (FiO₂), end-tidal carbon dioxide (EtCO₂), arterial oxygen partial pressure (PaO₂), arterial carbon dioxide partial pressure (PaCO₂), pulse oxygen saturation (SpO₂), total intravenous anaesthesia (TIVA), vasoconstrictor (VC).
variation (ΔMAP500), and PP variation (ΔPP500) was not significantly different from 0.5. ROC curve of ΔHR500, ΔMAP500 and ΔEtCO2500 is shown in Fig. 4.

Clinical ranking of the response with a 10 % diagnosis tolerance

After a 500-ml fluid load, EtCO2 variation within the population can be classified as follows: 10 patients had a negative test, 10 patients had a positive test, and 20 patients had an inconclusive test.

Discussion

This study shows that in a context where alveolar ventilation and CO2 production are constant, the change in EtCO2 provides information to assess the CI changes during a 500-ml volume expansion. EtCO2 variation after a mini-fluid challenge is probably not usable to predict the response to a fluid bolus of 500 ml.

Three factors can explain a change in EtCO2: pulmonary blood flow (usually equal to cardiac output), CO2 metabolic production, and alveolar ventilation [21]. Naturally, if two of these factors are constant, the change in

Table 2 Haemodynamic data before and after a 500-ml fluid load in responders and non-responders

| Variable | T0        | T500 ml    | p value |
|----------|-----------|------------|---------|
| HR (min⁻¹) |           |            |         |
| R        | 69 [65; 75] | 73 [66; 76] | 0.27    |
| NR       | 69 [61; 77] | 70 [63; 74] | 0.33    |
| MAP (mmHg) |           |            |         |
| R        | 59 [51; 66] | 67 [55; 83] | 0.12    |
| NR       | 63 [58; 67] | 64 [56; 72] | 0.47    |
| SAP (mmHg) |           |            |         |
| R        | 91 [84; 99] | 111 [92; 115] | 0.08    |
| NR       | 95 [88; 107] | 95 [90; 112] | 0.40    |
| DAP (mmHg) |           |            |         |
| R        | 50 [41; 54] | 55 [44; 66] | 0.07    |
| NR       | 50 [47; 56] | 53 [45; 60] | 0.60    |
| PP (mmHg) |           |            |         |
| R        | 44 [39; 51] | 49 [41; 61] | 0.16    |
| NR       | 46 [37; 51] | 44 [40; 60] | 0.42    |
| FtC (s/√s) |           |            |         |
| R        | 312 [296; 329] | 331 [325; 371] | <0.01 |
| NR       | 305 [289; 322] | 334 [303; 369] | <0.01 |
| EtCO2 (mmHg) |           |            |         |
| R        | 31 [30; 32] | 33 [31; 33] | 0.02    |
| NR       | 32 [29; 34] | 31 [28; 32] | 0.04    |
| MV (L/min) |           |            |         |
| R        | 5.1 [4.4; 6.9] | 5.2 [4.4; 6.9] | 0.52    |
| NR       | 6.4 [5.8; 7.2] | 6.4 [5.7; 7.3] | 0.81    |

Table 3 Main characteristics of ROC curves built for hemodynamic variables of interest

| Variable | AUC        | Best thresholds (%) | Compared with AUC = 0.5 | Specificity | Sensitivity | PPV | NPV | LR+ | LR− | Youden index |
|----------|------------|---------------------|------------------------|-------------|-------------|-----|-----|-----|-----|--------------|
| varEtCO2500 | 0.82 [0.67; 0.97] | 5.89 | 0.006 | 1 [1.0; 1.0] | 0.6 [0.33; 0.87] | 1.0 | 0.81 | Infinite | 0.4 | 0.6 |
| varEtCO2100 | 0.74 [0.60; 0.89] | 3.0 | 0.034 | 1 [1.0; 1.0] | 0.33 [0.13; 0.6] | 1.0 | 0.68 | Infinite | 0.67 | 0.33 |
| varMAP500 | 0.62 [0.44; 0.83] | 0.31 |           |             |             |     |     |     |     |              |
| varPP500 | 0.62 [0.43; 0.82] | 0.31 |           |             |             |     |     |     |     |              |
| varHR500 | 0.65 [0.44; 0.82] | 0.29 |           |             |             |     |     |     |     |              |
| FtC | 0.49 [0.37; 0.67] | 0.09 |           |             |             |     |     |     |     |              |

Mean arterial pressure (MAP), pulse pressure (PP), heart rate (HR), corrected systolic flow time (Ftc), and end-tidal carbon dioxide (EtCO2). Data are expressed as variation from baseline (i.e., ΔEtCO2 for EtCO2 variation) after different volumes of infused fluid 100 and 500 ml. Area under the curve (AUC), positive predictive value (PPV), negative predictive value (NPV), positive likelihood ratio (LR+), negative likelihood ratio (LR−)
the third factor can explain the variation of EtCO2. Two mechanisms can also explain EtCO2 variation. First, a fluid challenge increases venous return and pulmonary blood flow; thus, a greater amount of CO2 delivery to the lungs and removal by alveolar ventilation should be observed. Second, a fluid challenge recruits collapsed pulmonary blood capillaries and, therefore, reduces West zones 1 and 2 and increases zone 3 [22]. Consequently, the ventilation/perfusion ratio decreases and dead space is reduced enabling more CO2 to be extracted by alveolar ventilation.

Our results are consistent with previously published studies. EtCO2 has been described to be able to track CI changes in experimental [10] and clinical settings [23]. Additionally, changes in EtCO2 have been used in many clinical situations as cardiac resuscitation for monitoring blood flow generated by precordial compression [24] or assessing the prognosis of cardiac arrest [25, 26]. Two recent studies focusing on patients with septic shock using PLR and volume expansion have shown a significant correlation between changes in EtCO2 and CI. Some anaesthesiologists already assess fluid responsiveness by measuring EtCO2 variation during a fluid challenge, though this strategy has not been specifically evaluated [27]. As previously described, the correlation between the absolute values of EtCO2 and CI was not significant. This is explained by the fact that except for very low CI, EtCO2 is mainly influenced by many other factors than CI. Of course, during the short meantime of a volume expansion, these factors can be considered as unchanged.

The threshold value of the EtCO2 variation after 500-ml volume infusion was 5.8 %, or an increase of 2 mmHg, which is low, but EtCO2 is a very stable variable if no modification of ventilation or cell metabolism occurs, so even a small change can be significant. This idea is reinforced by the fact that the least significant change (LSC) is smaller than the best threshold for a 500-ml load. Moreover, this is about the same threshold as the previously published one [11, 12]. Again, the threshold is near the LSC, and it seems to us that it would be hard to use in the clinical settings for a 100-ml load.

In previous studies, the correlation between changes in EtCO2 and CI was stronger during PLR than during volume expansion [11, 12]. Three main explanations can be given.

First, a passive leg raising is shorter than a fluid challenge. This increases the risk of a confounding factor such as ventilation or metabolism variation to be present.

Second, the study protocol was applied during anaesthesia for a surgery and not in a context of circulatory insufficiency. The link between EtCO2 and CI is predicted to be weaker when there is no circulatory insufficiency [10].

Third, the preload increase induced by a mini-fluid challenge is inferior compared with a passive leg raising. Our results concerning the mini-fluid challenge are consistent with a study performed recently in a population of septic patients [28]. From our point of view, as long as the fluid challenge is great enough and the infusion time is short, changes in EtCO2 could be used as a surrogate for CO changes.

As expected, HR, PP, and MAP variations were unable to predict fluid responsiveness. In a pragmatic approach, with a clinical 10 % tolerance of sensitivity and specificity, as in the grey zone model, we can only conclude for half of the patients. Of course, these results are poorly discriminant. Nevertheless, the discrimination is higher than with parameters mostly used to assess fluid responsiveness as MAP and HR.

We acknowledge that there are some limitations. The variation of EtCO2 could be explained by variation in VCO2 due to variation in the depth of anaesthesia, but we did not find a significant difference between both groups in the end-tidal anaesthetic agent concentration. We did not use a specific device to monitor the depth of anaesthesia. Nevertheless, end-tidal anaesthetic agent concentration remains the reference to assess depth of anaesthesia [29].

The weak correlation between EtCO2 variation, and CI variation after a mini-fluid challenge was largely explained by the imprecision of both measurements: in fact, at that same time the mean variation of CI and EtCO2 in the responder group is under the least significant change in the EtCO2 and CI.

Our patients benefited from protective mechanical ventilation with positive end-expiratory pressure and low tidal volumes for its positive effect [30]. Consequently, we did not compare our data to pulse pressure variation (PPV) or stroke volume variation (SVV) because we set the tidal volume under 8 ml/kg. These settings make SVV or PPV unable to predict fluid responsiveness [6, 31].

The AUC-ROC after a 500-ml fluid load has a lower confidence interval which is below 0.75. Accordingly, our study has not sufficient power to exclude the fact that the AUC in the population is below this limit. Therefore, we cannot exclude the fact that EtCO2 variation after a 500-ml fluid load could be of limited clinical interest according to Ray et al. [32].

To conclude in a setting with a constant alveolar ventilation and CO2 production, if no data on cardiac output or pulse pressure variation are available, EtCO2 is the only parameter that was discriminant to assess fluid responsiveness. When the variation of EtCO2 is >5.8 %,
all patients were responders, but no conclusion could be done when this variation was <5.8 %. A 100-ml mini-fluid challenge was not discriminant and cannot be used to predict fluid responsiveness regarding the least significant change in the EtCO₂. Conversely to widespread belief, MAP, PP, and HR variations were not accurate in predicting fluid responsiveness. The strategy of fluid expansion based on EtCO₂ variation could be a part of a haemodynamic optimisation protocol regarding its high specificity and its tremendous advantage upon classical cardiac output monitoring to be non-invasive, at no additional cost, and available in all operating rooms.

Authors’ contribution
M.J.L conceived of the study and the design of the study, collected data, performed statistical analysis, and wrote the manuscript. VP, FB, JLF, ML and PBH participated in the study’s design and helped to draft the manuscript. JSD participated in the study’s design and coordination and helped to draft the manuscript. All authors read and approved the final manuscript.

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Competing interests
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