Assessment of Salt Stress to *Arabidopsis* Based on the Detection of Hydrogen Peroxide Released by Leaves Using an Electrochemical Sensor

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Abstract: Salt stress will have a serious inhibitory effect on various metabolic processes of plant cells, this will lead to the excessive accumulation of reactive oxygen species (ROS). Hydrogen peroxide (H$_2$O$_2$) is a type of ROS that can severely damage plant cells in large amounts. Existing methods for assessing the content of H$_2$O$_2$ released from leaves under salt stress will cause irreversible damage to plant leaves and are unable to detect H$_2$O$_2$ production in real time. In this study, on the strength of a series of physiological indicators to verify the occurrence of salt stress, an electrochemical sensor for the detection of H$_2$O$_2$ released from leaves under salt stress was constructed. The sensor was prepared by using multi-walled carbon nanotube-titanium carbide–palladium (MWCNT-Ti$_3$C$_2$Tx-Pd) nanocomposite as substrate material and showed a linear response to H$_2$O$_2$ detection in the range 0.05–18 mM with a detection limit of 3.83 µM. Moreover, we measured the determination of H$_2$O$_2$ released from *Arabidopsis* leaves at different times of salt stress by the sensor, which was consistent with conventional method. This study demonstrates that electrochemical sensing is a desirable technology for the dynamic determination of H$_2$O$_2$ released by leaves and the assessment of salt stress to plants.

Keywords: salt stress; hydrogen peroxide; electrochemical sensor; nanomaterial; plant leaves

1. Introduction

At present, about one-fifth of the irrigated soil in the world is affected by salinization [1]. Under the condition of soil salinization, salt stress has caused certain economic losses to agricultural production [2–4]. Salt stress caused by saline alkali land has a significant impact on normal plant life activities, and high salt can induce osmotic stress, ion stress, and oxidative stress in plants [5–9]. In a high-salinity environment, osmotic stress occurs first in plants, then ion stress occurs later, which will destroy ion homeostasis in plants, thus affecting plant growth. Osmotic stress and ion stress can induce oxidative stress in plants and will lead to the excess production of ROS, which can cause cell damage and death. ROS are an important signaling molecule that play an important role in biotic and abiotic stress in plants. It can sense and integrate environmental signals quickly in cells in order to improve the tolerance of plants to the environment [10]. Among ROS, H$_2$O$_2$ is one of the most stable forms which participates in electron transfer during photosynthesis in and out of the body [11–13]. H$_2$O$_2$ also acts as a signal molecule and plays an important role in the signal transduction of plants because of its long-life span and ability to cross cell membranes [14–16].
The traditional determination method of H$_2$O$_2$ released by plants is to dye plant tissues or roots with diaminobenzidine tetrahydrochloride (DAB) which can combine with H$_2$O$_2$ to form insoluble brown precipitation [17]. However, this method can neither achieve real-time monitoring of H$_2$O$_2$ release from leaves nor detect the specific amount of H$_2$O$_2$, and it can also cause irreversible damage to plant leaves. The method of chemiluminescence has been used to quantify the amount of H$_2$O$_2$ in plant tissues [18], but it is not convenient to use because it requires luminescent detection equipment to monitor the reaction. Scopoletin and 7-hydroxy-6-methoxy-2H-1-benzopyran-2-one are fluorescent substrates for peroxides and have also been used to determine H$_2$O$_2$ concentrations [19]. However, it is influenced by many factors and its sensitivity is relatively low. The H$_2$O$_2$ content is measured with spectrophotometry using phenol red as an H$_2$O$_2$ indicator dye. However, this method requires the fragmentation of plant tissues, which causes irreversible damage to the plants [20]. At present, non-destructive probe techniques are becoming more and more popular in the detection of H$_2$O$_2$ released by plants, such as Amplex Red, europium-tetracycline complex, and other probes [21]. However, these probes have certain toxic effects on plants. Nowadays, analytical methods based on electrochemical sensors have attracted widespread concern in the analysis of plants owing to its sensitivity, in situ detection ability, and so on [22].

In the process of the electrochemical sensor preparation, the selection of electrode substrate material is the key factor affecting the conductivity of the sensor. With the deepening of nanomaterials research, the application of enzyme-free H$_2$O$_2$ electrochemical sensors provides a new idea for researchers [23]. MWCNTs stand out among many nanomaterials with enzyme-like electrocatalytic activity because of their significant structural diversity and stable physical and chemical properties [24]. In addition, the electrochemical properties of MWCNT nanocomposite are often better than each component due to the synergistic effect [25,26]. As a new two-dimensional material, Ti$_3$C$_2$Tx shows excellent electrochemical ability because of its good conductivity and catalytic activity [27–29]. At the same time, Ti$_3$C$_2$Tx shows excellent catalytic ability for H$_2$O$_2$. At present, there have been a lot of research on electrochemical sensors constructed with Ti$_3$C$_2$Tx to further detect H$_2$O$_2$, and it has shown good detection performance [30,31]. Among several metal nanomaterials, Pd nanoparticles exhibit excellent electrocatalytic performance for hydrogen peroxide, and Pd further exhibits enhanced electron transfer and reduced overpotential properties [32–34]. Furthermore, the abundance of Pd makes it an inexpensive alternative used in a variety of fields compared to other novel metals [35].

In this work, an electrochemical sensor based on MWCNT-Ti$_3$C$_2$Tx-Pd nanocomposite was constructed to detect H$_2$O$_2$ released from *Arabidopsis* leaves and aim to assess salt stress (Scheme 1). The electrochemical information of H$_2$O$_2$ produced by *Arabidopsis* leaves under salt stress was compared with the results obtained by conventional staining. Compared with current methods, samples of leaves would not to be lysed or fixed, which maintain the living and original forms of biological samples. At same time, the sensor can detect the release of H$_2$O$_2$ in different time periods after salt stress to measure the real-time dynamic detection of H$_2$O$_2$ release from leaves. This electrochemical method provides a certain basis for the detection of harmful substances produced by plants under other stress conditions.
Scheme 1. Schematic illustration of structure and working principle of the electrochemical sensor.

2. Results and Discussions

2.1. Characterization of the MWCNT-Ti3C2Tx-Pd Nanocomposite

To improve the performance of the electrochemical sensor, MWCNT-Ti3C2Tx-Pd nanocomposite was synthesized to enhance the signal of the electrode. In order to explore the synthesis of nanocomposite, the shape and composition of the material were characterized with a transmission electron microscope (TEM). Figure 1A showed that the MWCNTs were fibrous and their size could reach the micron level. As can be seen from Figure 1B, the Ti3C2Tx presented multilayer flake, which effectively expanded the electroactive area and provided more attachment sites for Pd nanoparticles. From Figure 1C, it can be seen that MWCNTs and Ti3C2Tx can be combined together through π-π interactions. The image showed that the MWCNT-Ti3C2Tx nanocomposite was successfully synthesized. At present, studies have shown that palladium nanoparticles have a significant catalytic effect on H2O2 [34]. Therefore, we reduced PdCl2 to Pd nanoparticles on the surface of MWCNT-Ti3C2Tx based on the redox method. It can be seen from Figure 1D that Pd was well adsorbed on the Ti3C2Tx. In addition, the energy dispersive X-ray spectroscopy measurement result of the MWCNT-Ti3C2Tx-Pd nanocomposite was shown in Figure 1E, which confirmed the existence of Pd and Ti elements in the nanocomposite. These results demonstrated the successful synthesis of the MWCNT-Ti3C2Tx-Pd nanocomposite.

2.2. Electrochemical Characterizations of the Modified Electrodes

CV is a method that is commonly used to explore electrochemical properties of molecules adsorbed on electrode surfaces. Figure 2A showed the conductivity of Ti3C2Tx, MWCNT, MWCNT-Ti3C2Tx, and MWCNT-Ti3C2Tx-Pd in the 10 mM K3Fe(CN)6 solution with 0.1 M KCl. The response signal of the MWCNT-Ti3C2Tx-Pd nanocomposite modified electrode indicated that its electron transfer speed was fast. As shown in Figure 2B, the current response of Ti3C2Tx nanocomposite modified electrode was relatively low. This result also showed that Ti3C2Tx had a poor electron transfer ability when it existed alone. The performance of MWCNTs and MWCNT-Ti3C2Tx nanocomposite modified electrodes were better than that of Ti3C2Tx modified electrodes. Moreover, the closed area of the CV curve of MWCNT-Ti3C2Tx-Pd nanocomposite modified electrode was large, which further showed that the active surface area of the electrode was expanded, and the sensitivity of the sensor was greatly improved. Figure 2B showed that the reduction potential of H2O2 was approximately 0 V, and the MWCNT-Ti3C2Tx-Pd nanocomposite modified electrode exhibited an excellent detection effect on H2O2 solution. This result indicated that Pd nanoparticles had an efficient catalytic effect on H2O2 [36].
Figure 1. Characterization of the nanocomposite. TEM images of MWCNTs (A), Ti$_3$C$_2$Tx (B), MWCNT-Ti$_3$C$_2$Tx (C), MWCNT-Ti$_3$C$_2$Tx-Pd (D,E). The energy dispersive X-ray spectroscopy of MWCNT-Ti$_3$C$_2$Tx-Pd nanocomposite.

This electrode-modified nanomaterial MWCNT-Ti$_3$C$_2$Tx-Pd was further explored for the kinetic properties of electrochemical sensors by using the CV method. This procedure was performed by measuring the response signals at different scan rates using MWCNT-Ti$_3$C$_2$Tx-Pd modified electrodes in 5 mM H$_2$O$_2$. As can be seen from Figure 2C, the response signal of this sensor gradually increased with the increase of the scan rate within the 20 mV/s–190 mV/s scan rate. According to the above exploration, we fit and analyze the reduction peak currents ($I_p$) and the square root of the scan rate ($\sqrt{v}$) at different scan rates. It was found that the current value was directly proportional to the square root of the scanning rate, and its linear equation was $I_p = -11.28 \sqrt{v} - 62.28$ ($R^2 = 0.998$). This result further showed that the electrochemical behavior of H$_2$O$_2$ on MWCNT-Ti$_3$C$_2$Tx-Pd nanocomposite modified electrodes was a diffusion-controlled process.
In order to enhance the conductivity of the electrochemical sensor, the dispersion effect of the nanocomposite is one of the most important factors to be considered. Therefore, MWCNTs were dispersed with water, PBS, PVP (2 mg/mL), 5% PDDA, and 98% DMF as dispersants, respectively. As can be seen from Figure 3A, MWCNTs dispersed by water and PBS had a poor dispersion effect compared with the other three dispersants. In order to further explore the dispersion properties of PVP, PDDA, and DMF, the dispersion effects of these three dispersants were evaluated with CV. Figure 3B showed that MWCNTs dispersed by 5% PDDA had the best conductive effect. Therefore, 5% PDDA was used as dispersant for subsequent experiments. The ratio between MWCNTs and Ti3C2Tx are also an important factor affecting the performance of the electrochemical sensor. According to previous experiments, when the concentration of MWCNTs exceed 4 mg/mL, it was not more stable on the electrode surface. The optimization of material proportion was also explored in 5 mM H2O2 solution using the CV method. It can be seen from Figure 3C that when the ratio of MWCNTs to Ti3C2Tx reached 4:1 (4 mg MWCNT and 1 mg Ti3C2Tx dissolved in 1 mL system), the sensor showed a better catalytic effect on H2O2 solution. Therefore, this proportion was used for follow-up research.

Figure 2. (A) CV curves in the presence of 10 mM K3Fe(CN)6 in 0.1 M KCl solution for Ti3C2Tx (a), MWCNT (b), MWCNT–Ti3C2Tx (c) and MWCNT–Ti3C2Tx–Pd (d). (B) CV curves of Ti3C2Tx (a), MWCNT (b), MWCNT–Ti3C2Tx (c) and MWCNT–Ti3C2Tx–Pd (d) in PBS containing 5 mM H2O2. (C) CV curves of the MWCNT–Ti3C2Tx–Pd in PBS containing 5 mM H2O2 at different scan rates (20, 30, 40, 50, 60, 70, 80, 90, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190 mV/s) and plot of peak current vs. square root (v1/2) of scan rates (D).

2.3. Optimization of Experimental Conditions

In order to enhance the conductivity of the electrochemical sensor, the dispersion effect of the nanocomposite is one of the most important factors to be considered. Therefore, MWCNTs were dispersed with water, PBS, PVP (2 mg/mL), 5% PDDA, and 98% DMF as dispersants, respectively. As can be seen from Figure 3A, MWCNTs dispersed by water and PBS had a poor dispersion effect compared with the other three dispersants. In order to further explore the dispersion properties of PVP, PDDA, and DMF, the dispersion effects of these three dispersants were evaluated with CV. Figure 3B showed that MWCNTs dispersed by 5% PDDA had the best conductive effect. Therefore, 5% PDDA was used as dispersant for subsequent experiments. The ratio between MWCNTs and Ti3C2Tx are also an important factor affecting the performance of the electrochemical sensor. According to previous experiments, when the concentration of MWCNTs exceed 4 mg/mL, it was not more stable on the electrode surface. The optimization of material proportion was also explored in 5 mM H2O2 solution using the CV method. It can be seen from Figure 3C that when the ratio of MWCNTs to Ti3C2Tx reached 4:1 (4 mg MWCNT and 1 mg Ti3C2Tx dissolved in 1 mL system), the sensor showed a better catalytic effect on H2O2 solution. Therefore, this proportion was used for follow-up research.
was measured when the polarization voltage was 0 V. At the same time, in order to make the response signal of the sensor also increased gradually, and this result further showed that the electrochemical sensor had a good performance.

2.4. Amperometric Measurements of H₂O₂

The electrocatalytic behavior of MWCNT-Ti₃C₂Tx-Pd nanocomposite for H₂O₂ reduction was studied using the CV method. Figure 4 showed that the MWCNT-Ti₃C₂Tx-Pd nanocomposite modified electrode was placed in H₂O₂ solution with concentrations of 0, 1 mM, 5 mM, and 10 mM, respectively, to electrochemical detection using the CV method. As can be seen from Figure 4, with the increase of the H₂O₂ concentration, the current response signal of the sensor also increased gradually, and this result further showed that the potential of H₂O₂ reduction is the 0 V. Figure 4 showed that H₂O₂ can be easily electrochemically reduced on MWCNT-Ti₃C₂Tx-Pd nanocomposite in a wide concentration range, which also showed that the H₂O₂ electrochemical sensor had a good performance.

Figure 3. (A) Effect of dispersed carbon nanotubes with different dispersants (a: water; b: PBS; c: 2 mg/mL PVP; d: 5% PDDA; e: 98% DMF) and their electrochemical properties in the 10 mM K₃Fe(CN)₆ solution containing 0.1 M KCl (B). (C) CV curves of MWCNTs and Ti₃C₂Tx modified electrode in PBS containing 5 mM H₂O₂.

Figure 4. CV curves at different H₂O₂ concentrations.

In order to further detect the performance of a MWCNT-Ti₃C₂Tx-Pd nanocomposite modified electrode to H₂O₂, the response ability of the electrochemical sensor to H₂O₂ was measured when the polarization voltage was 0 V. At the same time, in order to make H₂O₂ disperse evenly in the system quickly, the solution was stirred with a magnetic stirrer. The response of the electrochemical sensor to different concentrations of H₂O₂ was further evaluated by chronoamperometry. It was obvious from Figure 5A that the current response signal also increased gradually with the increase of the added H₂O₂ in the same time period.
It was obvious that the MWCNT-Ti$_3$C$_2$Tx-Pd nanocomposite modified electrode reacted rapidly to the change of H$_2$O$_2$ concentration, and the electrochemical sensor also responded well to the small concentration of H$_2$O$_2$. As shown in Figure 5B, the linear fitting between the concentration of the detected H$_2$O$_2$ solution and the corresponding current value was calculated. The linear equation of the modified electrode was $y = -20.76 \times x - 20.85$ from 0.05 mM to 18 mM, and the linear correlation coefficient $R^2 = 0.993$. According to the linear regression equation, the sensitivity was calculated to be 293.85 µA mM$^{-1}$ cm$^{-2}$, and the minimum detection limit was calculated to be 3.83 µM (LOD = 3SD/slope) based on the linear regression curve at a relatively low concentration range. Meanwhile, it could be clearly seen from Table 1 that the electrochemical sensor constructed in this study had excellent sensitivity compared with the H$_2$O$_2$ sensor reported in previous studies. These results further showed that MWCNT-Ti$_3$C$_2$Tx-Pd nanocomposite was an excellent material for the detection of H$_2$O$_2$, which also greatly improved the performance of the electrochemical sensor.

![Figure 5. (A) The current response of the MWCNT-Ti$_3$C$_2$Tx-Pd nanocomposite modified electrode was measured when different concentrations of H$_2$O$_2$ were continuously added to the saturated PBS. (B) Calibration plot of cathodic peak currents vs. different H$_2$O$_2$ concentrations and the error bar is expressed as the mean of the standard error ($n = 3$).](image)

**Table 1.** Performance comparison of different types of H$_2$O$_2$ electrochemical sensors.

| Material                  | Sensitivity (µA mM$^{-1}$ cm$^{-2}$) | Detection Limit (µM) | Linear Range (mM) | Reference  |
|---------------------------|--------------------------------------|----------------------|-------------------|------------|
| PANI-HRP-SWCNTs           | 200                                  | 900                  | 2.5–50            | [37]       |
| C$_3$O$_2$-LSC-2          | 47.83                                | 1.5                  | 0.005–11.4        | [38]       |
| MWCNTs-PtNPs             | 142.8                                | 10                   | 0.01–0.1          | [39]       |
| SWCNTs-Au-AgNPs          | 13.1                                 | 26.8                 | 0.3–1.8           | [40]       |
| MWCNT-Ti$_3$C$_2$Tx-Pd   | 293.85                               | 3.83                 | 0.05–18           | this work  |

2.5. Stability and Reproducibility

In the actual monitoring process, the electrode will be affected by the external environment. Therefore, the stability and reproducibility of the nanocomposite modified electrode becomes very critical for the performance of the electrochemical sensor. The stability experiment of the material was carried out by placing the MWCNT-Ti$_3$C$_2$Tx-Pd modified electrode in a 5 mM H$_2$O$_2$ solution and using the CV method to explore it. The modified electrodes were measured every two days and stored in a 4 °C environment for the next measurement after each measurement. As can be seen from Figure 6A, the electrodes still reacted well to H$_2$O$_2$ on the sixth day. This result showed that the electrochemical sensor exhibited good stability, which laid a foundation for the long-term determination of H$_2$O$_2$ released from leaves.
Proline is an osmotic regulator. When plants are subjected to salt stress, they become yellow and wilted and the growth conditions were worse than those in the control group. Traditional DAB staining showed that the leaves of the salt treatment group was darker than the control group. This result demonstrated that plants under salt stress will produce a large amount of H$_2$O$_2$, which will cause serious damage to plants.

Five separate electrodes (a, b, c, d, e) in PBS containing 5 mM H$_2$O$_2$ were used to determine the detection limit of H$_2$O$_2$ in the control group. The level of H$_2$O$_2$ was lower than that of the control group after salt treatment (Figure 7F,G). In order to compare the changes of ion content more intuitively, we calculated the ratio of Na$^+$ and K$^+$ and found that the Na$^+$/K$^+$ value (salt stress group) was higher than that of the control group after salt treatment (Figure 7H). Proline is an osmotic regulator. When plants are subjected to salt stress, they will release a large amount of proline to regulate osmotic balance. As we expected, it was found that the proline level of the salt stress group was significantly higher than that of the control group (Figure 7I). The above physiological data indicated that plants were indeed damaged by salt stress.

2.6. Physiological Characteristics of Salt Stress to Arabidopsis Leaves

Wild-type Arabidopsis plants with the same growth vigor were selected and divided into a control group and salt stress group. After the salt stress group was treated with 100 mM NaCl for seven days, the physiological characteristics of Arabidopsis were determined to verify the salt stress. Compared with the control group, the fresh weight of the shoots in the salt stress group was significantly reduced (Figure 7A). Under salt stress conditions, the contents of chlorophyll a, chlorophyll b, and total chlorophyll in the control group were significantly higher than those in the salt stress group (Figure 7B–D). Meanwhile, the level of MDA in the salt stress group was significantly higher than that in the control group (Figure 7E).

Since salt stress usually leads to ionic stress, we determined the Na$^+$ and K$^+$ content of leaves. Under the condition of salt stress, the Na$^+$ content of the salt stress group was significantly higher than that of the control group, while the K$^+$ content was significantly lower than that of the control group (Figure 7F,G). In order to compare the changes of ion content more intuitively, we calculated the ratio of Na$^+$ and K$^+$ and found that the Na$^+$/K$^+$ value (salt stress group) was higher than that of the control group after salt treatment (Figure 7H). Proline is an osmotic regulator. When plants are subjected to salt stress, they will release a large amount of proline to regulate osmotic balance. As we expected, it was found that the proline level of the salt stress group was significantly higher than that of the control group (Figure 7I). The above physiological data indicated that plants were indeed damaged by salt stress.

2.7. Detection of H$_2$O$_2$ Released from Arabidopsis Leaves

In order to evaluate the amount of H$_2$O$_2$ released by Arabidopsis under salt stress, traditional DAB staining and electrochemical sensing technology were used to perform the qualitative and quantitative analysis of H$_2$O$_2$ released from leaves. It can be seen from the phenotypic map of markers in Figure 8A that the leaves of plants under NaCl stress turned yellow and wilted and the growth conditions were worse than those in the control group. Traditional DAB staining showed that the leaves of the salt treatment group was darker than the control group. This result demonstrated that plants under salt stress will produce a large amount of H$_2$O$_2$, which will cause serious damage to plants.
Figure 7. (A) Fresh weight of aboveground part of each plant. (B) Chlorophyll a content per gram of leaves. (C) Chlorophyll b content per gram of leaves. (D) Total Chlorophyll content per gram of leaves. (E) MDA content per gram of leaves. (F) Na⁺ content per gram of leaves. (G) K⁺ content per gram of leaves. (H) Ratio of Na⁺ and K⁺ per gram of leaves. (I) The content of proline per gram of leaves. The sample size of each group is 3. *p < 0.05, **p < 0.01, ***p < 0.001 and ****p < 0.0001 as determined by the student t test.

Figure 8. (A) Phenotypic map and DAB staining of leaves before and after salt treatment. (B) Results of measuring H₂O₂ released from leaves based on electrochemical method (R₀ represents the ratio of current to leaves mass. n = 3). *p < 0.05 and **p < 0.01 as determined by the student t test. ns: not significant.

In order to perform the quantitative analysis of H₂O₂ released from Arabidopsis leaves, a prepared electrochemical sensor was used. Since the quality of each leaf was different, R₀ (ratio of current to leaves mass) was used to analyze the results. Figure 8B showed that...
the H$_2$O$_2$ released by leaves also gradually increased with the increase of time, and that the H$_2$O$_2$ release of the salt treatment group was higher than the control group. The result was consistent with that of DAB staining. At the same time, we can calculate the amount of H$_2$O$_2$ released from each leaf based on the measured current value. Therefore, the dynamic quantitative monitoring of H$_2$O$_2$ released by leaves under salt stress was achieved with an electrochemical method. This provides a new method for the evaluation of some markers produced by plants under salt stress.

3. Materials and Methods

3.1. Reagents and Apparatus

K$_3$Fe(CN)$_6$, polyvinylpyrrolidone (PVP) and phthalic diglycol diacrylate (PDDA) were purchased from Sigma (Saint Louis, MO, USA). H$_2$O$_2$ was purchased from Shanghai Wokai Biotechnology Co., Ltd. (Shanghai, China). The MWCNTs, Ti$_3$C$_2$T$_x$, and PdCl$_2$ were purchased from Nanjing Xianfeng Nano Technology Co., Ltd. (Nanjing, China). Potassium chloride, ethanol, sodium chloride (NaCl), sodium borohydride (NaBH$_4$), hydrochloric acid (HCl), N,N-dimethylformamide (DMF), L-proline, trioxohydrindene monohydrate, glacial acetic acid, phosphoric acid, trichloroacetic acid (TCA), and thiobarbituric acid (TBA) were obtained from Sinopharm Chemical Reagent Co., Ltd. (Shanghai, China). 5-Sulfosalicylic acid was obtained from the Tianjin Fuchen Chemical Reagents Factory (Tianjin, China). Acetone was obtained from Yantai Yuandong Fine Chemicals Co. Ltd. (Yantai, China). Toluene was purchased from the Laiyang Economic and Technological Development Zone fine chemical plant. Phosphate buffer solution (PBS) was purchased from Biological Industries. DAB was sourced from Biotopped.

The morphology characterization of nanomaterials was analyzed with an HT-7800 transmission electron microscope (Hitachi Limited, Tokyo, Japan). A CHI 400C electrochemical workstation (Chen Hua Company, Shanghai, China) with a standard three-electrode configuration was used to perform electrochemical properties. Platinum wire, a Ag/AgCl electrode, and a glassy carbon electrode (GCE, diameter 3 mm) were used as the counter electrode, reference electrode, and working electrode, respectively. The absorbance value was measured with a UV-visible spectrophotometer (Beijing General Analysis Instrument Co., LTD., Beijing, China). Ion content was determined using a flame photometer (Cole Parmer Instrument Co., made in the UK, St. Neots, UK).

3.2. Preparation of MWCNT-Ti$_3$C$_2$T$_x$-Pd Nanocomposite

The dispersing effect of different kinds of dispersants is evaluated by using MWCNTs. MWCNTs were dissolved with different dispersants including ultrapure water, PBS (0.1 M, pH 7.4), PVP (2 mg/mL), 98% DMF and 5% PDDA. After that, the MWCNTs were subjected to ultrasound for 2 h and stirred for 4 h at room temperature. The dispersed MWCNTs were washed by solution (ethanol: ultrapure water = 1:1) and centrifuged at 15,300 r/min for 10 min. This process was repeated 3–5 times, and finally ultrapure water was added to obtain MWCNT uniform dispersion.

The synthesis of MWCNT-Ti$_3$C$_2$T$_x$-Pd nanocomposite was to add MWCNTs and Ti$_3$C$_2$T$_x$ to 1mL of the optimized dispersant, and then add 20 µL 0.5 mol/L PdCl$_2$ solution (dissolved in 10% HCl) and 200 µL 2.5 mg/mL NaBH$_4$ (dissolved in ultrapure water). Finally, the composites were subjected to ultrasonic (2 h), stirring (4 h), and centrifugal washing (3–5 times). After the PDDA was washed off, the composite was resuspended with ultrapure water to obtain a uniform and stable MWCNT-Ti$_3$C$_2$T$_x$-Pd solution.

3.3. Fabrication of the Modified Electrode

Before modifying the electrode, the undecorated GCE was polished with alumina powder (0.3 µm to 0.05 µm) to gain a smooth surface, and then a mixed solution of ultrapure water and ethanol was used to wash the GCE to remove the physically adsorbed substances. After that, the GCE was dried with nitrogen. Subsequently, MWCNT-Ti$_3$C$_2$T$_x$-
Pd nanocomposite dispersion (10 µL) was dropped to the conductive area of the electrode, and then the electrode was dried at room temperature.

3.4. Electrochemical Measurements

Cyclic voltammetry (CV) was used to characterize the electrochemical performance of the electrochemical sensor. PBS was used as the electrolyte for the determination of H₂O₂. Different concentrations of H₂O₂ solution were obtained by diluting 30% H₂O₂ solution in PBS. The determination of H₂O₂ solution was carried out under the condition of stirring. Differential pulse voltammetry was used to investigate the release of H₂O₂ from leaves under salt stress conditions. All electrochemical measurements were made at room temperature.

3.5. Physiological Characteristics of Wild-Type Arabidopsis before and after Salt Treatment

Wild-type Arabidopsis grew in an artificial climate incubator in the Plant Physiology Laboratory, Shandong Normal University. After 27 days of normal growth for the Arabidopsis, the samples were treated with 100 mM NaCl for one week for subsequent physiological analysis of leaves under salt stress and electrochemical determination of the H₂O₂.

3.5.1. Determination of the Fresh Weight

Wild-type Arabidopsis with the same growth was selected and divided into the control group and salt stress group. The salt stress group was treated with 100 mM NaCl for 7 days, and the fresh weight of the aboveground part of the control group and salt stress group was weighed.

3.5.2. Determination of Chlorophyll Content

0.2 g fresh leaves were weighed and cut into filaments, then put into a mixture of ethanol (95%): acetone (80%) = 1:1. After 48 h of extraction in darkness, we measured the absorbance values at wavelengths of 663 nm and 645 nm, and calculated the chlorophyll a, chlorophyll b, and total chlorophyll according to its formula.

3.5.3. Determination of Malondialdehyde (MDA) Content

Seedlings (0.2 g) were mixed with 2.5 mL of 0.1% TCA and ground to homogenate, then the extract was mixed with 2.5 mL of 0.5% TBA. The mixture was soaked in boiling water for 10 min and quickly cooled on ice. The supernatant was taken and its absorbance values at wavelengths 532 nm and 600 nm were measured [41].

3.5.4. Determination of Na⁺, K⁺ Content and Calculation of Na⁺ to K⁺ Ratio

0.3 g leaves were weighed and bathed in boiling water for 3 h. Then Na⁺ and K⁺ were measured in a constant volume of 25 mL and calculated with a flame spectrophotometer.

3.5.5. Determination of Proline Content

0.2 g leaves were weighed and ground by adding sulfosalicylic acid, then the extract was bathed in boiling water for 10 min and centrifuged. The supernatant was removed, mixed with glacial acetic acid and acidic ninhydrin solution, and bathed in boiling water for 1 h. After cooling, toluene solution was added, and the upper toluene solution was taken to measure its absorbance value at 520 nm.

3.6. Preparation and Analysis of Leaves Model

In the DAB staining part, the pH of the DAB dye solution (1 mg/mL) was first adjusted to 5.8 with NaOH. Secondly, the leaves were then added to the DAB solution and left to stand in the dark at 28 °C for 8 h. After standing, the DAB solution was discarded, 10 mL of 80% ethanol was added, and boiled in water for 5 min. Finally, the waste liquid was discarded, 10 mL of absolute ethanol was added, and water was added and boiled until
the leaves turned white. The samples were photographed for observation and stored in anhydrous ethanol.

During electrochemical detection, the leaves were picked from the plant, weighed, and then put into 5 mL PBS for electrochemical determination. The experiment adopted the method of differential pulse voltammetry to measure the H$_2$O$_2$ released from leaves at the potential of $-0.3$ V to 0.5 V. Meanwhile, the release of H$_2$O$_2$ from the leaves every two hours in 0–8 h was measured with an electrochemical sensor.

4. Conclusions

In this work, we prepared an electrochemical sensor based on MWCNT-Ti$_3$C$_2$T$_x$-Pd that showed a good performance for the determination of H$_2$O$_2$. Based on the successful construction of the salt stress model by measuring a series of physiological indicators, we used a self-made electrochemical sensor to indirectly evaluate salt stress suffered by *Arabidopsis* thaliana by detecting the H$_2$O$_2$ released by the leaves. The research results showed that the content of H$_2$O$_2$ released by *Arabidopsis* was improved under the condition of salt stress and accumulated more with longer time. The results obtained with an electrochemical sensor were consistent with those obtained by traditional DAB staining, which further showed that the sensor had a good new performance in detecting H$_2$O$_2$. More importantly, the method based on electrochemical sensor overcomes the limitations of DAB staining and measures the quantitative determination of H$_2$O$_2$ under the minimum damage to plants. Therefore, this work provides a new method for the dynamic monitoring of hazard markers produced by plants under abiotic stress conditions.

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References

1. Kopittke, P.M.; Menzies, N.W.; Wang, P.; McKenna, B.A.; Lombi, E. Soil and the intensification of agriculture for global food security. *Environ. Int.* 2019, 132, 105078. [CrossRef] [PubMed]
2. Rengasamy, P. World salinization with emphasis on Australia. *J. Exp. Bot.* 2006, 57, 1017–1023. [CrossRef] [PubMed]
3. Ruan, C.-J.; da Silva, J.A.T.; Mopper, S.; Qin, P.; Lutts, S. Halophyte improvement for a salinized world. *J. Exp. Bot.* 2015, 66, 2399–2415. [CrossRef]
4. Morton, M.J.L.; Awlia, M.; Al Tamimi, N.; Saade, S.; Pailles, Y.; Negrão, S.; Tester, M. Salt stress under the scalpel–dissecting the genetics of salt tolerance. *Plant J.* 2019, 97, 148–163. [CrossRef] [PubMed]
5. Farooq, M.; Gogoi, N.; Hussain, M.; Barthakur, S.; Paul, S.; Bharadwaj, N.; Migdadi, H.M.; Alghamdi, S.S.; Siddique, K.H.M. Effects, tolerance mechanisms and management of salt stress in grain legumes. *Plant Physiol. Biochem.* 2017, 118, 199–217. [CrossRef]
6. Zeng, Y.; Li, L.; Yang, R.; Yi, X.; Zhang, B. Corrigendum: Contribution and distribution of inorganic ions and organic compounds to the osmotic adjustment in Halostachys caspica response to salt stress. *Sci. Rep.* 2015, 5, 15867. [CrossRef] [PubMed]
7. Ran, X.; Wang, X.; Gao, X.; Liang, H.; Liu, B.; Huang, X. Effects of salt stress on the photosynthetic physiology and mineral ion absorption and distribution in white willow (Salix alba L.). *PLoS ONE* 2021, 16, e0260086. [CrossRef]
8. Hameed, A.; Ahmed, M.; Hussain, T.; Aziz, I.; Ahmad, N.; Gul, B.; Nielsen, B. Effects of Salinity Stress on Chloroplast Structure and Function. *Cells* 2021, 10, 2023. [CrossRef]
9. Liang, W.; Ma, X.; Yan, P.; Liu, L. Plant salt-tolerance mechanism: A review. *Biochem. Biophys. Res. Commun.* 2018, 495, 286–291. [CrossRef] [PubMed]
10. Mittler, R.; Zandalinas, S.I.; Fichman, Y.; Van Breusegem, F. Reactive oxygen species signalling in plant stress responses. *Nat. Rev. Mol. Cell Biol.* 2022, 23, 666–679. [CrossRef] [PubMed]
11. Vicente, C.S.L.; Ikuyo, Y.; Mota, M.; Hasegawa, K. Fine wood nematode-associated bacteria contribute to oxidative stress resistance of *Bursaphelenchus xylophilus*. *BMC Microbiol.* 2013, 13, 1–8. [CrossRef] [PubMed]
12. Smirnoff, N.; Arnaud, D. Hydrogen peroxide metabolism and functions in plants. *New Phytol.* 2018, 221, 1197–1214. [CrossRef] [PubMed]

13. Quan, L.-J.; Zhang, B.; Shi, W.-W.; Li, H.-Y. Hydrogen Peroxide in Plants: A Versatile Molecule of the Reactive Oxygen Species Network. *J. Integr. Plant Biol.* 2008, 50, 2–18. [CrossRef]

14. Cuypers, A.; Hendrix, S.; Dos Reis, R.A.; De Smet, S.; Deckers, J.; Gielen, H.; Jozeefczak, M.; Loix, C.; Vercampt, H.; Vangronsveld, J.; et al. Hydrogen Peroxide, Signaling in Disguise during Metal Phytotoxicity. *Front. Plant Sci.* 2016, 7, 470. [CrossRef] [PubMed]

15. Černý, M.; Habáňová, H.; Berka, M.; Lučková, M.; Brzobohatý, B. Hydrogen Peroxide: Its Role in Plant Biology and Crossstalk with Signalling Networks. *Int. J. Mol. Sci.* 2018, 19, 2812. [CrossRef]

16. Singh, S.; Husain, T.; Kushwaha, B.K.; Suhel, M.; Fatima, A.; Mishra, V.; Singh, S.K.; Bhatt, J.A.; Rai, M.; Prasad, S.M.; et al. Regulation of ascorbate-glutathione cycle by exogenous nitric oxide and hydrogen peroxide in soybean roots under arsenate stress. *J. Hazard. Mater.* 2020, 409, 123686. [CrossRef] [PubMed]

17. Jambunathan, N. Determination and Detection of Reactive Oxygen Species (ROS), Lipid Peroxidation, and Electrolyte Leakage in Plants. In *Methods in Molecular Biology*; Springer Science and Business Media LLC: Berlin/Heidelberg, Germany, 2010; Volume 639, pp. 291–297.

18. Warm, E.; Lattes, G.G. Quantification of hydrogen peroxide in plant extracts by the chemiluminescence reaction with luminol. *Phytochemistry* 1982, 21, 827–831. [CrossRef]

19. Corbett, J.T. The scopoletin assay for hydrogen peroxide A review and a better method. *J. Biochem. Biophys. Methods* 1989, 18, 297–307. [CrossRef]

20. Miyata, K.-I.; Miyashita, M.; Nose, R.; Otake, Y.; Miyagawa, H. Development of a Colorimetric Assay for Determining the Amount of H2O2 Generated in Tobacco Cells in Response to Elicitors and Its Application to Study of the Structure-Activity Relationship of Flagellin-Derived Peptides. *Biosci. Biotechnol. Biochem.* 2006, 70, 2138–2144. [CrossRef] [PubMed]

21. Šnyrychová, L.; Bertok, T.; Filip, J.; Jerigova, M.; Mahmoud, K.A.; Tkac, J. Highly stable Ti3C2Tx (MXene)/Pt nanoparticles-modified glassy carbon electrode for H2O2 and small molecules sensing applications. *Biosens. Bioelectron.* 2018, 99, 108–114. [CrossRef]

22. Dhara, K.; Mahapatra, D.R. Recent advances in electrochemical nonenzymatic hydrogen peroxide sensors based on nanomaterials: A review. *J. Mater. Sci.* 2019, 54, 12319–12357. [CrossRef]

23. Wang, Z.; Dai, Z. Carbon nanomaterial-based electrochemical biosensors: An overview. *Nanoscale* 2015, 7, 6420–6431. [CrossRef]

24. Wang, J.; Chen, X.-J.; Liao, K.-M.; Wang, G.-H.; Han, M. Pd nanoparticle-modified electrodes for nonenzymatic hydrogen peroxide determination and detection. *Analyst* 2018, 143, 99–108. [CrossRef]

25. Tang, N.; Zheng, J.; Sheng, Q.; Zhang, H.; Liu, R. A novel H2O2 sensor based on the enzymatically induced deposition of polyaniline at a horseradish peroxidase/aligned single-wall carbon nanotubes modified Au electrode. *Analyst* 2010, 136, 781–786. [CrossRef] [PubMed]
38. Zhang, Y.; Huang, Y.; Feng, C.; Zhang, Y.; Wu, H. Electrochemical assay of hydrogen peroxide based on hybrids of Co3O4/biomass-derived carbon. Ionics 2019, 25, 6051–6059. [CrossRef]

39. Chou, T.-C.; Wu, K.-Y.; Hsu, F.-X.; Lee, C.-K. Pt-MWCNT modified carbon electrode strip for rapid and quantitative detection of H2O2 in food. J. Food Drug Anal. 2018, 26, 662–669. [CrossRef] [PubMed]

40. Ko, E.; Tran, V.-K.; Geng, Y.; Chung, W.S.; Park, C.H.; Kim, M.K.; Jin, G.H.; Seong, G.H. Continuous electrochemical detection of hydrogen peroxide by Au-Ag bimetallic nanoparticles in microfluidic devices. J. Electroanal. Chem. 2017, 792, 72–78. [CrossRef]

41. Liu, T.; Guo, S.; Lian, Z.; Chen, F.; Yang, Y.; Chen, T.; Ling, X.; Liu, A.; Wang, R.; Zhang, B. A P4-ATPase Gene GbPATP of Cotton Confers Chilling Tolerance in Plants. Plant Cell Physiol. 2014, 56, 549–557. [CrossRef] [PubMed]