Abstract: Accumulating evidence shows that oxidative stress plays an essential role in the pathogenesis and progression of many diseases. The imbalance between the production of reactive oxygen species (ROS) and the antioxidant systems has been extensively studied in pulmonary, neurodegenerative cardiovascular disorders; however, its contribution is still debated in gastrointestinal disorders. Evidence suggests that oxidative stress affects gastrointestinal motility in obesity, and post-infectious disorders by favoring the smooth muscle phenotypic switch toward a synthetic phenotype. The aim of this review is to gain insight into the role played by oxidative stress in gastrointestinal pathologies (GIT), and the involvement of ROS in the signaling underlying the muscular alterations of the gastrointestinal tract (GIT). In addition, potential therapeutic strategies based on the use of antioxidants for the treatment of inflammatory gastrointestinal diseases are reviewed and discussed. Although substantial progress has been made in identifying new techniques capable of assessing the presence of oxidative stress in humans, the biochemical-molecular mechanisms underlying GIT mucosal disorders are not yet well defined. Therefore, further studies are needed to clarify the mechanisms through which oxidative stress-related signaling can contribute to the alteration of the GIT mucosa in order to devise effective preventive and curative therapeutic strategies.

Keywords: oxidative stress; gastrointestinal diseases; gastrointestinal muscle inflammation; antioxidants

1. Introduction

Oxidative stress in living organisms results from the imbalance between the production of reactive oxygen species (ROS) and the ability to neutralize them. The disparity between excessive reactive molecules and weak endogenous defense leads to damage to cell structures and molecules such as lipids, proteins, and DNA, ultimately contributing to the pathogenesis of a wide range of diseases. ROS, when available in appropriate low amounts, act as signal transduction molecules driving cell activities and also provide cell protection [1]. On the other hand, if generated in excess, as in inflammation, ROS can trigger the production of additional highly reactive species [2]. Crucially is the oxidative modification of key enzymes or regulatory sites, whose redox modification triggers cell signaling alteration and programmed cell death. Oxidative stress and inflammation are closely linked. Oxidative stress can cause inflammation and this, in turn, induces oxidative stress generating a vicious circle [3,4] that results in cell damage, which promotes a pro-inflammatory environment [5].

Literature data confirm the key role of oxidative stress in etiology of numerous and different diseases (Figure 1), including metabolic syndrome [6], atherosclerosis [7], cardiovascular disease [8,9], cancer [10,11], neurodegenerative disorders [12,13], diabetes [14], infertility [15], renal diseases [16], and gastrointestinal and hepatic diseases [17].
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Figure 1. Scheme of oxidative stress-induced diseases in humans.

Being involved in the absorption of nutrients and in the immune response, the gastrointestinal tract (GIT) plays a key role also in the production of ROS. Several evidences highlight how the pathogenesis of various GIT diseases, including colorectal and gastric cancers [18–20], inflammatory bowel disease (IBD) [21,22], and peptic ulcers [23], is due, at least in part, to oxidative stress.

The GIT tissue is structured into four layers: the mucosa (epithelium, lamina propria, and muscular mucosae), the submucosa, the muscularis propria (inner circular muscle layer, intermuscular space, and outer longitudinal muscle layer), and the serosa.
The intestinal epithelia are exposed continuously to a wide variety of potentially harmful substances and act as a selective barrier between the tissues and luminal environment of the GIT. There are several stressors, which induce the generation of free radicals and result in oxidative stress and GIT inflammatory responses involving the epithelium and immune/inflammatory cells [24]. Although there is enough information on the role played by oxidative stress in the damage of intestinal mucosa, little is known about the involvement of the surrounding muscle layers.

Knowledge of the biochemical mechanisms underlying the alterations induced by oxidative stress at the GIT level, as well as of the physiological responses of the different GIT layers to such stress, is mandatory to better understand either pathogenesis of GIT diseases or to develop new and more effective therapeutic strategies.

This review summarizes the current understanding of the role of oxidative stress in GIT pathophysiology, also discussing the specific molecular mechanisms involved, focusing particular attention on the implication of the muscular layers of the GIT.

2. Oxidative Stress

Oxidative stress occurs when, in tissues and organs, the formation of highly reactive molecules e.g., ROS, reactive nitrogen species (RNS), and reactive sulfur species (RSS), overcome the endogenous antioxidant defense system capacities, leading to cellular damage and dysfunctions that result in a wide range of diseases. The reactive species are constantly generated within cells at low concentrations as a result of normal metabolic processes. They can also result from the exposure to external factors like radiation (X-rays and UV), ozone, air pollutants, cigarette smoke, bacteria, viruses, drugs, etc. [25], or as the outcome of an acute or chronic cellular stress. The reactive species can be free radicals and non-radical oxidants. The free radicals are unstable because of unpaired electrons presence in their outer electron orbit. Since free radicals are highly unstable and reactive, tend to neutralize themselves by reacting with other molecules causing their oxidation [26]. Therefore, by reacting with important biological molecules, including DNA, lipids and proteins, they can cause damage on various levels [27]. Proteins, being among the main components of the cells, represent major targets for free radicals [28]. Free radicals can induce some protein modifications, i.e., unfolding or alteration of protein structure, most of which, fortunately, are essentially harmless events [29]. While the reversible oxidative changes are involved in the regulation of protein activity, irreversible protein changes can lead to their inactivation with consequent lasting harmful cellular effects [29].

The intracellular sources of chemical reactive species are mainly mitochondria, endoplasmic reticulum, lysosomes, peroxisomes, cytosol, and plasma membrane [30] (Figure 2). ROS derive from the chemical reduction of molecular oxygen and, among the main ones, we find: the free radicals, such as superoxide anion radical (O$_2$$^\cdot$−), hydroxyl radical (•OH), as well as non-radical oxidant, such as hydrogen peroxide (H$_2$O$_2$) and hypochlorous acid (HClO) [31]. Among the RNS, the major players are peroxynitrite radical (ONOO$^-$), ozone, and nitric oxide (•NO) [32]. The new identified RSS include thiol radical (RS), and RSS both formed by the reaction between ROS and thiols. Similarly, RSS include radical species, such as (RSR$^*$), glutathionyl radical (GSSG$^*$), and non-radicals ones, such as reactive sulfane species (RSR), reactive sulfur substances (SO$_2$, SO$_3$), etc. [33,34]. In particular, RSS are able to trigger both oxidation and reduction reactions with particular tropism for sulfur-containing molecules, such as peptides and proteins [33,34].
The most important sites of ROS production are the enzymes of the mitochondrial electron transport respiratory chain. Other enzymes catalyze chemical reactions contributing to the ROS formation, among them the homologs of nicotinamide adenine dinucleotide phosphate (NADPH) oxidase, phospholipase A2 (PLA2), uncoupled nitric oxide synthase (NOS) as well as cyclooxygenases (COX), xanthine oxidase (XO), lipoxygenases (LOXs), glucose oxidase, and myeloperoxidase (MPO) [24,35,36]. NADPH oxidase, an enzyme present in the plasma membrane, was initially discovered in the phagosomes of macrophages, neutrophils, and monocytes. There are six homologs of NADPH oxidase, NOX1, NOX3-5, and dual oxidase (DUOX) 1 and 2, with several intracellular localizations [37]. Literature data showed that NOX1 and DUOX2 have significant roles in Helicobacter pylori-induced gastric inflammation, inflammatory bowel disease (IBD), and tumors [24,38]. XO is present in the cytoplasm and also on the outer surface of the plasma membrane; it is mainly expressed in the liver and small intestinal mucosa within the gastrointestinal tract [39]. LOXs are non-heme iron enzymes that can generate ROS catalyzing oxidation of arachidonic acid (AA). MPO is a heme-enzyme localized in lysosomes of neutrophils, macrophages, and monocytes. Several data demonstrate that MPO activity is increased in inflamed mucosa in ulcerative colitis and also in H. pylori-infected subjects [24], playing a role in the development of H. pylori-induced atrophic gastritis. The chronic oxidative stress related to...
ulcerative colitis and *H. pylori*-infection could also lead to cancer, often associated with these diseases [40,41]. NOS is a heme-containing monooxygenase that generates NO. There are three different isoforms of NOS: neuronal NOS (nNOS), endothelial NOS (eNOS), and endotoxin or cytotoxin-inducible NOS (iNOS) [42]. In GIT, NOS expression and activity are very important because the generation of NO maintains normal functions of mucosa and plays a cytoprotective role. Indeed, NO regulates blood flow, epithelial secretion, and barrier function of gastric mucosal [43] and represents one of the main enteric neurotransmitters mediating GI muscle relaxation [43,44]. However, NO can also have deleterious effects, and iNOS expression was found increased in chronic ulcerative colitis and peptic ulcer patients [24]. COX enzyme releases AA from the membrane phospholipids and catalyzes AA conversion to prostanoids. COX has two isoforms, COX-1 and COX-2, both of which are expressed in normal human gastric mucosa. COX-1 is constitutively expressed, while COX-2 is induced by inflammation and tumorigenesis [45]. COX-2 has also been reported to have cytoprotective functions in human colon and gastric cancer cells where it was induced during high osmotic stress [46]. Therefore, reactive oxygen species, including oxygen free radicals, are generated by the activity of several types of oxidases. Initially, O\textsubscript{2} is reduced by the addition of electrons, thereby producing O\textsubscript{2}•− that can react with other endogenous molecules to generate secondary oxidizing molecules, such as ONOO•. Thereafter, the reduction of O\textsubscript{2}•− leads to the by-product H\textsubscript{2}O\textsubscript{2} that is characterized by a long life span and relative stability. The latter is enzymatically converted into water and O\textsubscript{2}, or possibly into different metabolites, thus extinguishing the radical cascade [6].

Both O\textsubscript{2}•− and H\textsubscript{2}O\textsubscript{2} are also important signaling molecules, particularly in vascular smooth muscle cells where they can trigger specific biochemical pathways that regulate the defense mechanisms following exposure to oxidative stress. At the center of these pathways are for example mitogen-activated protein kinases, and tyrosine kinases, and transcription factors [47]. Particularly transcription factors, such as activator protein-1 (AP-1), NF-κB, and/or NF-E2-related factor (NRF2) have been reported to also participate in redox-modulated cell signaling [48,49].

### 3. Antioxidants

If the body’s antioxidant defense system fails to neutralize the excess free radicals, the imbalance between oxidants and the defense system can lead to pathological conditions, including cancer [10,11], cardiovascular disease [7,8], neurodegenerative disorders [12,13], atherosclerosis [7], and others. Halliwell and Gutteridge defined antioxidants as “any substance that delays, prevents or removes oxidative damage to a target molecule” [50–52]. All living organisms are endowed with endogenous antioxidant defenses capable of contrasting and removing reactive chemical species. However, these defenses are insufficient to totally remove reactive species and completely prevent oxidative damage to cells, tissues, and organs [4]. The endogenous antioxidants can act at various levels: blocking the formation of radicals, neutralizing them by oxidizing themselves, or delaying the oxidation reactions of other molecules. Moreover, some antioxidants, acting as metal chelators, transform metal pro-oxidants into more stable chemical forms.

The antioxidants were be classified by Gutteridge and Halliwell into three categories: primary, secondary, and tertiary antioxidants, on the bases of their mechanism of action [51]. Primary antioxidants inhibit oxidant formation; secondary antioxidants function as scavengers of ROS, and tertiary antioxidants repair the oxidized molecules. Currently, antioxidants are substantially classified as enzymatic or non-enzymatic (Figure 3).
3.1. Enzymatic Antioxidants

Among the enzymatic antioxidants that contribute to the defense against the reactive species, we find catalase (CAT), superoxide dismutase (SOD), glutathione peroxidase (GPX), and glutathione reductase (GSR). Enzymatic antioxidants have both primary and secondary defense functions and represent an endogenous antioxidant system. Glutathione peroxidase, SOD, and catalase are the primary defense that prevents the formation or neutralize reactive species [53]. In particular, SOD and catalase provide major antioxidant defenses against ROS.

SOD catalyzes the dismutation of $O_2^−$ into $O_2$ and $H_2O_2$. In humans beings are present three isoforms of SOD [54]: cytosolic copper and zinc-containing enzyme (Cu-Zn-SOD), present in the mitochondrial inter-membranous space; manganese-requiring mitochondrial enzyme (Mn-SOD), present in the mitochondrial matrix; and extracellular Cu-Zn containing SOD (EC-SOD) [55]. $H_2O_2$ not scavenged by GPX located at the level of the mitochondrial matrix crosses the mitochondrial membrane towards the cytosol, where it can be scavenged either by cytosolic Cu-Zn-SOD or CAT [56]. Increased levels of all three SOD isoforms are present in intestinal tissues from IBD patients, particularly in the epithelium [57], and in patients with ulcer healing [58]. Increased expression of Mn-SOD is associated with colorectal cancer, and it was also found increased in normal mucosa of gastric adenocarcinoma as well as squamous cell oesophageal carcinoma tissues [59]. Moreover, a gastrointestinal mucosal injury could be prevented by the presence of SOD [60].
CAT, present mainly in peroxisomes, dismutates $\text{H}_2\text{O}_2$ to $\text{H}_2\text{O}$ and $\text{O}_2$ [61]. In humans, CAT has been found virtually in all organs although it is produced largely in liver, kidney, and erythrocytes. Lower catalase activity was observed in colorectal cancer [62], gastric adenocarcinoma, in $H.\text{pylori}$-infected stomach [62], and in Crohn’s disease [63].

GPX converts glutathione (GSH) into its oxidized form (GSSG), reduces $\text{H}_2\text{O}_2$ to $\text{H}_2\text{O}$, and lipid hydroperoxides (ROOH) to the corresponding stable alcohols. The GPX reaction is paired to glutathione reductase (GSR), which maintains reduced glutathione (GSH) levels. GSR, GPX, and glutathione S-transferases (GST), form the glutathione system that in the GIT mucosa acts as an antioxidative barrier. This enzyme, generating GSH, is important for the protection of cell membranes, red blood cells, and hemoglobin to oxidative stress [64]. GPX is found in the mitochondria, cytoplasm, and extracellular space [65], and protects cells from the harmful consequences of peroxide decomposition. In humans, there are eight isotypes of GPX. While GPX1 is ubiquitous, GPX2 is specific for the gastrointestinal tract and protects the gut against the absorption of dietary hydroperoxides [66]. Moreover, GPX2 defends the gastrointestinal tract against ROS derived from gut inflammation associated with commensal bacteria [67]. Importantly, glucose-6-phosphate dehydrogenase, while not directly neutralizing the radicals, can be considered an antioxidant enzyme. This oxidoreductase maintains the level of NADPH, thus helping to keep glutathione in its reduced state (GSH) [68] and creating a reducing environment [53].

Thioredoxin reductase (TrxR) together with thioredoxin (Trx) forms the thioredoxin system. There are three TrxR isoforms: TrxR1 found in the cytoplasm, TrxR2 in mitochondria, and TrxR3 present only in specialized tissues. TrxR, by transferring reducing equivalents from NADPH to thioredoxin, keeps it in its reduced form [69]. It has been shown that a compensatory upregulation of TrxR mRNA in gastrointestinal cancer was induced by oxidative stress provoked by bile acids [70].

3.2. Non-Enzymatic Antioxidants

Among the endogenous non-enzymatic antioxidants, there are glutathione and Trx. Glutathione is ubiquitously expressed mostly in its reduced form, GSH. Glutathione is a strong antioxidant, certainly one of the most important among those that the body can produce. Relevant is its action against both free radicals and molecules such as hydrogen peroxide, nitrates, nitrates, benzoates, and others. An important element for its functioning is NADPH. In fact, this molecule, a derivative of vitamin PP (nicotinic acid), functions as a redox cofactor of the enzyme GSR, which reduced glutathione (GSH) from oxidized glutathione (or GSSG) through electrons transferred from NADPH to GSSG [53].

Trx contains two free sulfhydryl groups of two cysteine residues. It is involved in the biosynthesis of deoxynucleotides, since it reduces the oxidized ribonucleotide reductase by yielding their hydrogens to the two oxidized sulfhydryl groups of the ribonucleotide reductase. Trx is present in the cytoplasm, membranes, and mitochondria but also in the extracellular space [71]. It showed a cytoprotective action in various inflammatory conditions, and was found to regulate the activity of redox-sensitive transcription factors, which are part of the antioxidant defence system.

Ubiquinone, also known as CoQ, is a lipophilic molecule existing in three different redox states: fully oxidized, partially reduced (ubisemiquinone), and fully reduced (ubiquinol) [72]. It is found in the plasma membrane and in several intracellular membrane including mitochondrial ones where it plays a key role in energy production and ROS generation. In its fully reduced form, CoQ is a potential antioxidant. Experimental studies have shown a protective role of ubiquinone against protein carbonylation and oxidative damage to DNA [73,74]. Furthermore, it is also been shown that ubiquinone can prevent peroxidative damage to membrane phospholipids [75] and regenerate other powerful antioxidants, such as $\alpha$-tocopherol and ascorbate, by recycling them back to their reduced active forms, thus increasing resources cellular antioxidants [76]. These properties make ubiquinone suitable as a food supplement to improve cellular bioenergetics and to counteract some age-related diseases [76].
Activator protein-1 (AP-1), nuclear factor kappa B (NF-κB), and nuclear factor (erythroid-derived 2)-like 2 related factor (NRF2) are three transcription factors that have been reported to be involved in redox-modulated signaling pathways. Indeed, oxidative stress up-regulates NF-κB activity, and AP-1 and NRF2 activation depends on the environmental and/or intracellular redox state. Under normal conditions, NRF2 is found blocked in the cytosol by its inhibitor, KEAP1. Oxidative modification of KEAP1 and NRF2 phosphorylation result in the release of NRF2 from KEAP1 [77] and its translocation into the nucleus, where it binds with antioxidant response elements involved in activation of gene expression, thereby protecting cells from free radical damage. Therefore, NRF2, through its interaction with antioxidant response element (ARE), is able to modulate the expression of defensive genes coding detoxifying enzymes and antioxidant proteins [78].

3.3. Exogenous Antioxidants

Besides the endogenous enzymatic and non-enzymatic antioxidant defenses, other antioxidants are also utilized by the body, which must necessarily be introduced through diet and for this reason are defined exogenous. In addition to endogenous antioxidants, exogenous ones act through different mechanisms and in different cellular compartments. They are mainly free radical scavengers: they neutralize free radicals, repair oxidized membranes, and decrease reactive oxygen species production [79]. Among the exogenous antioxidants, we find: vitamins (A, C, E, and K), enzyme cofactors (Q10), nitrogen compounds (uric acid), minerals (zinc, Zn and selenium, Se), and polyphenols (flavonoids and phenolic acid) [80]. Metals such as manganese, zinc, copper, iron, and selenium up-regulate the catalytic activity of antioxidant enzymes [81]. It has been indicated that an inadequate dietary intake of these trace minerals may compromise the effectiveness of antioxidant defense mechanisms [82].

Exogenous antioxidants have generated a growing interest in preventing or reducing oxidative stress. In fact, many epidemiological researches have highlighted how the use of foods containing antioxidants and scavengers has a potential protective effect against the disorders caused by oxidative stress [83–87]. By increasing the body’s natural antioxidant defenses, or by supplementing with dietary antioxidants, various chronic diseases can be prevented or their progression can be slowed down. Natural antioxidants such as flavonoids, tannins, and polyphenols act by donating electrons to intermediate radicals and play a role in the inhibition of lipid peroxidation. For example, vitamin E, particularly its active form α-tocopherol, protects cells from lipid peroxidation and helps in the prevention of chronic diseases associated with oxidative stress [88,89].

The antioxidant phytochemicals contained in vegetables and fruits are considered a benefits to the health. Indeed, several studies demonstrated that they have antioxidant abilities both in vitro and in vivo [90,91]. Moreover, literature data highlighted that antioxidant phytochemicals can also have anti-inflammatory action [92]. In fact, natural compounds such as curcumin, resveratrol, and anthocyanins could reduce inflammation via inhibition of prostaglandin production, NF-κB activity, and specific oxidative enzymes, as well as by increasing anti-inflammatory cytokine (e.g., IL-10) or decreasing pro-inflammatory cytokine (i.e., IL-1β) production [93,94].

4. Available Methods to Assess Oxidative Stress in Clinical and Research Approaches

Currently, one of the most interesting challenges in studying oxidative stress is identifying biomarkers that can be used in clinical diagnostics. According to the World Health Organization, a biomarker is “any substance, structure or process that can influence or predict the incidence of outcomes or diseases and be measured in the body or its products” [95].

Although oxidative stress markers can often be measured easily and scientific evidence suggests that oxidative stress can influence the onset and evolution of numerous diseases, they are not always considered clinically relevant biomarkers. In fact, a biomarker is clinically useful when it is specific for certain pathology (diagnostic marker), or has value
in predicting the evolution of the disease or is related to the degree of disease (prognostic marker). Furthermore, to be clinically useful, a biomarker must also be reasonably stable, present in easily accessible tissue and cost-effective to evaluate. Venous blood and urine are commonly used in clinical practice to detect oxidative stress, but in particular pathological conditions measurements of oxidative stress can also be conducted in cerebrospinal fluid [96], and other tissues [97,98].

In 2015, Frijhoff and colleagues examined the biomarkers used to assess oxidative stress, focusing on those most suitable for clinical and diagnostic use [99]. Possible markers of oxidative stress include the amount production of ROS produced, some downstream effects induced by ROS, and antioxidant defenses. Direct quantification of ROS, due to their short half-life, is quite a daunting task. Possible methods of measurement in biological systems include electron spin resonance, fluorescence magnetic resonance, and mass spectrometry techniques [100,101], but their use was limited to cell cultures and other in vitro applications. Flow cytometry is the most widely used method in clinical practice and research. In recent years, many fluorescent probes have been developed for the detection of reactive species, with a different degree of specificity and sensitivity [102]. The most commonly used fluorescent probes in clinical diagnostics for the detection of reactive species in cells by flow cytometry are shown in Table 1.

### Table 1. Fluorescent probes used for the measurements of reactive oxygen and nitrogen species by flow cytometry.

| Probe (Localization)          | ROS/RNS                  | Limitations                                                                 |
|-----------------------------|--------------------------|----------------------------------------------------------------------------|
| DCFH-DA (intracellular)     | •OH, ONOO⁻, •NO₂, H₂O₂  | DCF radicals production, MDR substrates or inducers, Antioxidants          |
| DAF-2 DA/DAF-FMDA (intracellular) | •NO                 | MDR substrates or inducers, Esterase inhibitors                             |
| DHR123 (intracellular)      | H₂O₂                    | MDR substrates or inducers, DHR radicals production                       |
| HE (intracellular)          | O₂⁻                    | Intercalating agents                                                      |
| C11-BODIPY₅₈₁/₅₉₁ (membrane) | •OH, •ROO              | Antioxidants                                                              |

C11-BODIPY₅₈₁/₅₉₁: 4,4-difluoro-5-(4-phenyl-1,3-butadienyl)-4-bora-3a,4a-diaza-s-indacene-3-undecanoic acid; DAF-2 DA: 4,5-diaminofluorescein diacetate; DAF-FMDA: 4-amino-5-methylamino-2′,7′-difluorofluorescein diacetate; DCFH-DA: dihydrochlorofluorescein diacetate; DHR123: dihydrorhodamine 123; H₂O₂: hydrogen peroxide; HE: hydroethidine; MDR: multidrug resistance; •NO: nitrogen monoxide; NO₂⁻: nitrogen dioxide; O₂⁻: superoxide radical; •HO: hydroxyl radical; ONOO⁻: peroxynitrite; •ROO: peroxyl radicals.

In addition to direct measurement of free radicals, a different approach is to measure reaction products of biological molecules with oxidizing species that may indicate systemic or tissue-specific oxidative stress. Indeed, as previously mentioned, molecules such as lipids, DNA, and proteins can be modified by the interaction with ROS, producing stable products that can be easily quantified [27,28]. Table 2 summarizes the main oxidation products of biological molecules used as markers of oxidative stress. Among the lipid oxidation products, useful as markers of oxidative stress, and involved in a variety of chronic diseases, trans-4-hydroxy-2-nonenal (4-HNE) and malondialdehyde (MDA) are the most studied. Several methods are available for the detection of both MDA and 4-HNE but the most reliable are the immunohistochemical and ELISA methods, in particular for HNE [103]. F2-isoprostanes (F2-IsoPs) are other markers used for the evaluation of oxidative stress in vivo. F2-IsoPs are formed in lipid membranes as a reaction between polyunsaturated fatty acids and ROS, and are therefore released in free form by the action of phospholipases. The measurement of F2-IsoP in biological fluids, as well as in the condensation of the breath, can provide an estimate of the systemic oxidative stress, while the measurement of esterified F2-IsoP in specific tissues can quantify a circumscribed oxidative stress. The most reliable methods for their quantification, the gas/liquid chromatography coupled with the mass spectroscopy techniques (HPLC/GC-MS), are laborious and require specialized and expensive instrumentation [104], while commercial immunoassays are often less reliable [105].
Table 2. Markers obtained from ROS-induced modifications.

| Markers                        | Methods                          | Limitations                                      |
|-------------------------------|----------------------------------|-------------------------------------------------|
| Lipid oxidation               |                                 |                                                 |
| HNE                           | HPLC, GC-MS, Immunoassay         | Sugars, aminoacids, Bilirubin and albumin, antibody specificity |
| MDA                           | Spectrophotometric/fluorimetric (TBARS), HPLC, Immunoassay | Antibody specificity                            |
| F2-IsoPs                      | Gas/liquid chromatography, Immunoassay | Antibody specificity                            |
| DNA oxidation                 | ELISA assays, HPLC-ECD, HPLC/GC-MS, Western blot, immunohistochemistry | Antibody specificity                            |
| 8oxodG                        | HPLC/GC-MS, Western blot, immunohistochemistry | Antibody specificity                            |
| 5-chlorouracil                |                                 |                                                 |
| Protein oxidation             |                                 |                                                 |
| ALEs, AGes                    | HPLC, Western blot, immunohistochemistry, ELISA | Structural variety of products, Antibody specificity |
| Carboxyls                     | Spectrophotometric, HPLC, ELISA, Western blot | Possible nitration of tyrosine residues in the sample, Antibody specificity |
| 3-NO-Tyr                      | HPLC/GC-MS, ELISA, Flow cytometry | Antibody specificity                            |
| AOPP                          | MS, colorimetric assays          |                                                 |
| oxLDL                         | ELISA, Flow cytometry            | Antibody specificity                            |
| IMA                           | ABC test, ELISA                  | Antibody specificity                            |

8oxodG: 7,8-dihydroxy-8-oxo-2′-deoxyguanosine; ABC test: binding capacity of albumin for cobalt; AGEs: advanced glycation end products; ALEs: advanced lipoxigenation end products; AOPP: advanced oxidation protein products; F2-IsoPs: F2-isoprostanes; GC: gas chromatography; HNE: 4-hydroxy-2-nonenal; HPLC: high-performance liquid chromatography; IMA: ischemia-modified albumin; MS: mass spectroscopy; MDA: malondialdehyde; oxLDL: malondialdehyde; TBARS: thiobarbituric acid reactive substances.

The nucleic acids DNA and RNA also represent a target of oxidative stress, particularly in their guanine bases. Oxidized nucleosides are excreted in the urine and their quantification can be interpreted as the cumulative total body oxidative stress. They are therefore able to provide information on systemic oxidative stress. Several commercial ELISAs are available to measure DNA damage with 7,8-dihydro-8-bone-2′-deoxyguanosine (8oxodG) and RNA damage with 7,8-dihydro-8-bone-2′-guanosine (8oxoGuo). The clinical use of chromatography coupled with mass spectrometry to detect oxidized nucleosides can be excessively expensive. Nucleic acid oxidation products have also been shown to predict the development of certain diseases [106].

Protein carbonyl groups originate from the oxidative cleavage of proteins by various mechanisms. They are usually detected after derivatization with 2,4-dinitrophenylhydrazine (DNP). The resulting carbonyl-2,4-dinitrophenylhydrazine adduct [107] can be detected spectrophotometrically, by ELISA, or by immunohistochemical, cytochemical, and western blot techniques using specific anti-DNP antibodies [108,109]. For clinical use, ELISA (commercially available) and HPLC tests are the only applicable methods. The functional groups of proteins can react with different molecules oxidized by ROS, such as polyunsaturated fatty acids and carbohydrates, generating respectively advanced peroxidation (ALE) and advanced glycation (AGE) end products [110,111]. This physiological process is particularly accentuated in conditions of hyperglycemia, hyperlipidemia, and oxidative stress. Protein adducts can be identified by mass spectrometry-based techniques, but their use is still limited in routine clinical analysis [112]. Furthermore, specific antibodies or spectrofluorimetric measurements based on the fluorescent properties of adducts are available [113,114]. Low-density oxidized lipoproteins (oxLDL), present in peripheral blood, are biomarkers of oxidative stress in cardiovascular disease, atherosclerosis, diabetes and obesity [115,116]. OxLDLs are measured in plasma or isolated LDLs by immunological methods using specific antibodies.

The redox state of cells, tissues or the whole organism can also be assessed by measuring the change in antioxidant defense systems in response to increased oxidative stress. The main players in this context include cysteine protein residues, the pool of antioxidants, ROS-generating enzymes, and transcription factors involved in their regulation (Table 3) [117].
Table 3. Markers of antioxidant defenses.

| Markers                         | Methods                  | Limitations            |
|---------------------------------|--------------------------|------------------------|
| Cysteine modifications          |                          |                        |
| S-glutathioylation              | MS, ELISA, WB            | Specialized instrumentation |
| GSH/GSSG, SH                    | Spectrophotometric, Flow cytometry |                        |
| ROS-regulated transcription factors |                        |                        |
| Nrf2, NF-kB                    | RT-PCR, WB, immunohistochemistry | Antibody specificity |
| NOX, MPO, XO, NOS               | RT-PCR, WB, ELISA, Immunological techniques | Antibody specificity |
| Antioxidant enzymes             | ELISA, WB, PCR, RT-PCR, Immunological techniques | Antibody specificity |
| SOD, CAT, GPX, GR              |                          |                        |

CAT: catalase; GPX: glutathione peroxidase; GR: glutathione reductase; GSH: glutathione; MPO: Myeloperoxidase; MS: mass spectroscopy; NOX: nitric oxide synthases; NOS: NADPH oxidase; PCR: reverse-transcription polymerase chain reaction; RT-PCR: reverse-transcription polymerase chain reaction; SOD: superoxide dismutase; WB: Western blot; XO: xanthine oxidase.

The cysteine residues exposed on the cell surface are particularly sensitive to oxidation by ROS. Once oxidized, they can be reduced again by the reaction with GSH and/or by specific enzymatic activities (e.g., by thioredoxins, glutaredoxins and isomerase of the disulfide protein) [118,119]. The measurement of GSH, GSSG and their ratio in the blood was considered an index of the systemic redox status [120]. Different methods have been used to determine GSH in biological samples (spectrophotometry, HPLC, capillary electrophoresis, nuclear magnetic resonance and mass spectrometry) [120]. As mentioned above, Nrf-2 regulates the cellular response to oxidative stress by promoting the transcriptional activation of genes containing antioxidant response elements (ARE) in their promoter regions. Among these, the genes encoding antioxidant and detoxifying enzymes, such as glutathione S-transferase, glutathione synthetase, heme oxygenase 1, and NAPD-oxidoreductase.

Some enzymes involved in ROS production, such as NOS, NOX, and MPO can be found in the peripheral blood and can therefore be used as markers of oxidative stress. However, expensive equipment would be required to detect MPO, also due to its low concentration in the blood, and this severely limits its use in clinical practice. Finally, antioxidant enzymes, such as CAT and SOD, can also be used as markers of oxidative stress. Conventional methods for evaluating enzymes are: gene expression by reverse transcription polymerase chain reaction (RT-PCR), direct protein quantification by western blot or other immunological techniques (e.g., immunocytochemistry and immunohistochemistry), and evaluation of the enzymatic activity [121]. Each of the methods illustrated so far for the quantification of oxidative stress has intrinsic limitations. These limits could be overcome by simultaneously using multiple evaluation criteria for oxidative stress. A redox state index was therefore proposed, the OXY-SCORE, [122–124], a global index of oxidative stress, which results from the integrated evaluation of a series of pro-oxidant and anti-oxidant biomarkers.

In conclusion, beyond the specific tests and biomarkers of oxidative stress already used in clinical diagnostics, the assays currently used in experimental research could be validated, in the near future, in the clinical setting. For this to happen, the reduction of economic costs, the standardization of methodological procedures, and the overcoming of the high variability of the results are necessary, also determined in part by intra-individual differences in risk factors, pathologies, and lifestyles. High costs and poor reproducibility of the results are the two elements that currently constitute the greatest obstacles to their large-scale use in clinical diagnostics.
5. Oxidative Stress in Gastrointestinal Disease

The gastrointestinal tract is a key source of ROS. Many cell types within the mucosa of the GIT produce ROS as part of normal physiology, yet the gut mucosa is also a target of various oxidants that can lead to pathological conditions. Redox signaling regulates the physiological function of gastrointestinal epithelium mainly through NADPH oxidases (NOXs), and commensal bacteria also contribute to intestine epithelial homeostasis through NOX1- and DUOX2-derived ROS. Commensal gut microbiota molecules, such as N-butyrate, are essential for controlling mitochondrial oxidative stress and inflammatory responses, pathogen growth, and adherence, as well as in improving metabolism and energy expenditure during exercise [125]. Despite the protective barrier provided by the epithelial layer, ingested materials and pathogens can cause inflammation by activating the epithelium, polymorphonuclear neutrophils, and macrophages to produce inflammatory cytokines and other mediators that contribute further to oxidative stress. Moreover, an excess of ROS could also induce discontinuation of the GI tract barrier, thereby increasing intestinal permeability and contributing to the inflammation observed in a variety of gastrointestinal diseases. Among the main sources of ROS and RNS, NADPH oxidase exerts an important role in the gastrointestinal tract. This group of enzymes includes numerous membrane-bound multimeric NOX isoforms and DUOX complexes, which are present in different tracts of the gut [126]. In particular, while DUOX complexes are found in all the tracts of the intestine, NOX1 is present only in the ileum, cecum, and colon epithelium. NOX2 is expressed mainly by professional phagocytes, while NOX4 is present in the epithelium, fibroblasts, and smooth muscle cells, and its expression was induced by stimuli such as TGF-β and hypoxia [37,127–129]. Other sources of intestinal ROS are mitochondria, particularly electron transport chain and NO synthases (NOS) [126]. It has been demonstrated that mitochondria have a prominent role in the modulation of gut functions as intestinal barrier protection, mucosal immune response [130,131], and maintenance of an eubiotic intestinal microbiota. A crosstalk between the gut microbiota and mitochondria during exercise is known in the literature. In particular, endurance exercise induces systemic mitochondrial biogenesis, prevents mitochondrial DNA depletion and mutations, and increases mitochondrial oxidative and antioxidant capacity. However, overtraining and chronic stress promote gut inflammation in athletes, which results in a plethora of stressors that favour the lipopolysaccharide translocation and the proliferation of pathobionts [125]. Moreover, oxidative stress exerts an important role in dysbiosis through the variation of microbial diversity in the gut. Intestinal inflammation, and consequent leukocyte infiltration, trigger oxidative stress by a generation of ROS and RNS promoting the loss of anaerobic bacteria [132,133].

The loss of redox homeostasis is implicated in the pathogenesis of several gastrointestinal disorders, such as Barrett’s esophagus, peptic ulcer, celiac disease, inflammatory bowel disease, and several adenocarcinomas (Figure 4) [134].

The Barrett’s esophagus is characterized by an increased production of $O_2^-$ anion, and consequent lipid peroxidation, paralleled by the inactivation of SOD [135]. The increase of $O_2^-$ in Barrett’s esophagus was triggered by one of the NOX isoforms. i.e., NOX5 [136], whose overexpression results mediated by the calcium-dependent activation of Rho kinase ROCK2 [137]. Moreover, it has also been observed that NOX5-S, a variant lacking calcium-binding domains, triggers the acid-induced generation of H$_2$O$_2$ and DNA damage in Barrett’s cells, thus contributing to the progression from Barrett’s esophagus to adenocarcinoma [138].
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As previously stated, NOX1 and DUOX2 have significant roles in Helicobacter pylori-induced gastric inflammation, which plays a cardinal role in peptic ulcer disease and gastric cancer. In particular, peptic ulcer is characterized by overproduction of $\text{O}_2^-$ and $\text{H}_2\text{O}_2$ derived mainly from leukocyte and neutrophils infiltrate [139,140], and from NOXs activity [141]. The oncogenesis of gastric carcinoma was found associated with the up-regulation of both NOX-1 and spermine oxidase. The up-regulation of spermine oxidase activity induced by $H.\ \text{pylori}$ in gastric epithelial cells increased $\text{H}_2\text{O}_2$ as a byproduct during the conversion of polyamine spermine into spermidine with consequent oxidative DNA damage [142].

Oxidative stress was also found to mediate most of the cytotoxic effects induced by gluten peptides in intestinal epithelial cells in celiac disease. In addition, ROS also enhanced the inflammatory cascade via NF-κB, and increased transglutaminase levels [134].

Progression of inflammatory bowel disease appeared to be determined by a quite complex mechanism of balance between pro-inflammatory redox-sensitive pathways, such as NLRP3 inflammasome and NF-κB, and the adaptive up-regulation of the antioxidant enzymes Mn-SOD and glutathione peroxidase 2 (GPX2) [134].

The overproduction of ROS due to mitochondrial dysfunction plays an important role also in the pathogenesis of inflammatory bowel disease (IBD) [126]. IBD, including Crohn’s disease and ulcerative colitis, were characterized by chronic inflammation of the GI tract. In ulcerative colitis, only the colon mucosal layer is affected, whereas in Crohn’s disease, inflammation may occur in all layers of the GI tract wall. Although the exact genesis of IBD is not fully understood, the association of ROS with IBD appeared evident from the observation that increased ROS and decreased antioxidant levels represented the major pathogenetic mechanisms in IBD [63,143]. Although the two forms of IBD share similar
characteristics, \( \text{H}_2\text{O}_2 \) and \( \text{HOCl} \) showed an important role in the pathophysiology of ulcerative colitis, whilst \( \text{HO}^- \) and \( \text{O}_2^- \) are found to be responsible for Crohn’s disease \[144\]. Further, in ulcerative colitis, a loss of mucosal antioxidant defence contributes to inflammation and disease progression. Murine studies showed that the severity of ulcerative colitis is related to SOD \[145\] and that antioxidants significantly reduce inflammatory responses \[146\]. Similarly, in inflamed mucosa of Crohn’s disease patients, the increase in XO, Mn-SOD activity, iNOS, and tumor necrosis factor-\( \alpha \) (TNF-\( \alpha \)) resulted associated with decreased antioxidant levels \[57,147\]. A deficiency in antioxidant molecules could lead to increased levels of lipid peroxides or ROS, which could act locally or be secreted into the circulation to produce different systemic effects in the patient \[148\].

6. Role of Oxidative Stress in Gastrointestinal Muscular Alterations

There are numerous data on the effects of oxidative stress in skeletal muscle. For example, it has been observed that in the skeletal muscle cells, the production of ROS increased with age and was associated with a loss of function, but could also significantly increase in the case of chronic inflammation \[149\]. As far as smooth muscle was concerned, most of the data are related to lung and vascular diseases. In fact, many experimental data showed that vascular smooth muscle cells are involved, together with endothelial cells, in the development of atherosclerosis, a pathology characterized by persistent inflammation and oxidative stress \[150\].

In any case, the analysis of the experimental data highlights the role played by mitochondria in oxidative stress.

Mitochondria are involved in many cellular functions, including the production of adenosine triphosphate (ATP), redox homeostasis, ROS and NADPH generation, calcium metabolism, and apoptosis \[151\]. Moreover, mitochondria can also detect warning signs and induce inflammation by activating and controlling the innate immune system \[152\]. Given the importance of mitochondria, alterations in their functions can have a profound effect on immunology and cell biology.

For instance, abnormalities in mitochondrial function have been described in human airway smooth muscle (ASM) cells from asthmatic patients \[153\], and in bronchial epithelial cells from ex-smokers with chronic obstructive pulmonary disease (COPD) \[154\]. In both pathologies, there are excessive mitochondrial ROS production, damaged mitochondrial structures with depletion of cristae, increased branching, elongation, and swelling of the mitochondria. Moreover, ASM cells from severe asthmatic patients present also a lack in the NRF2 antioxidant system \[153\].

Furthermore, it has been observed that oxidative damage can cause lesions of endothelial cells and deleterious vasodilatory effects, which could induce functional alterations in the smooth muscle cells of the vessel wall \[155,156\].

As regards the pathologies of the gastrointestinal tract, while the mucosal alterations associated with oxidative stress have been extensively investigated, the knowledge on oxidative stress-mediated muscle alterations has been only recently expanding. Most of the literature data come from studies on animal models, and the role of oxidative stress in the pathogenesis of muscular gastrointestinal diseases has not yet been extensively studied in humans.

Past studies performed on the murine model showed that molecules involved in oxygen-free radical production or in protection against oxygen radicals differed among the different gastrointestinal tracts and suggested that the large intestine was better provided with protective enzymes and non-enzymatic factors against oxidative stress than the small intestine \[39\]. Accordingly, it was observed that the large intestine was the most sensitive gastrointestinal tract in which oxidative stress-induced an alteration of intestinal motility. In fact, treatment of tissue segments from the large intestine with hydrogen peroxide compromised the contractile response into muscarinic agonists \[39\].

Later, Gonzalez and co-workers studied the in vitro modulation of rat colonic circular muscle contractions by dextran sodium sulfate (DSS)-induced inflammation coming to the
same conclusions. They observed that H$_2$O$_2$ altered the excitation-contraction coupling ending process suppressing the spontaneous phasic contractions and reducing responses by acetylcholine (ACh) stimuli [157]. The suppression of contractile capacity induced by oxidative stress was also confirmed by a study conducted on normal and inflamed canine colon, which demonstrated as H$_2$O$_2$-induced oxidative stress activated NF-kB in colonic circular smooth muscle cells, resulting in suppression of their contractility [158].

Similarly, it has been reported that in the intestines of rats, oxidative stress associated with aging reduced the tone of the internal anal sphincter (IAS) via RhoA/ROCK down-regulation. In particular, the decrease of RhoA/ROCK expression, both at the transcriptional and translational levels, was reverted by the activity of SOD, thus demonstrating its link with oxidative stress [159]. However, these data disagree with a previous observation made on rat gastric muscle in which oxidative stress induced an activation of Rho kinase II with a consequent increase in Ach-induced contraction [160]. Nevertheless, in a later study, Singh and co-authors described a bimodal effect of oxidative stress in IAS basal tone. Mild oxidative stress led to an increase in IAS tone associated, at least in part, with neuronal nitric oxide synthase (nNOS) inhibition; on the other hand, higher levels of oxidative stress caused a decrease in IAS tone. Both these effects were associated with changes in RhoA/ROCK [159]. The bimodal effect of oxidative stress was being previously hypothesized.

H$_2$O$_2$ has been shown to have not only harmful effects, since it results in an important signaling molecule that stimulates cell growth/proliferation and DNA synthesis in different types of cells [161,162]. More recently, Song et al. showed in feline ileal smooth muscle cells as short-term oxidative stress induced by H$_2$O$_2$ activated the signal transduction of mitogenic pathways, which are thought to represent a protective response against oxidant injury [163].

Only recently, the effects of oxidative stress on smooth muscle have been analyzed in human gastrointestinal muscle tissues. Scirocco et al. provided the first direct evidence of a muscular oxidative imbalance in the human gastric muscle that impaired antral smooth muscle relaxation both in vivo and in vitro [164]. This observation was in accordance with previous data showing the association between metabolic disorders and oxidative stress [165], and its effects on vascular and cardiac smooth muscle relaxation in the obesity condition [166]. In gastric muscle, the oxidative imbalance mainly affects the cAMP-signaling pathway and the expression of eNOS, two key components of the vasoactive intestinal polypeptide (VIP)-induced relaxation. The key role of oxidative stress in the impairment of muscle activity was confirmed by the ability of apocynin, an NADPH-inhibitor, to restore relaxation as well as antioxidant cell capacity and eNOS expression [164].

Interestingly, oxidative-stress mediated muscle impairment was also observed in vitro when the colonic muscle was exposed to supernatants obtained from a culture of mucosal biopsies of patients with irritable bowel syndrome (IBS). Colonic human smooth muscle strips and cells showed a decreased basal tone, a significant cell shortening, and a reduced Ach-induced contraction after exposure to supernatants [167]. The oxidative-related muscle damage induced by mucosal supernatants occurred likely through the generation of superoxide rather than hydrogen peroxide damage since it was reverted by apocynin but not by catalase [167].

Studies on Crohn’s disease have shown an increase in ROS levels and a decrease in antioxidant defenses resulting in a state of oxidative stress at the level of stenosis tracts [168]. Intestinal fibrosis with stricture formation represents a severe complication in Crohn’s disease. It has recently been reported that fibrosis could be triggered by NOX4-dependent ROS production that, activating myofibroblasts cells to produce collagen, lead to an increased thickness of the bowel wall [148,169].

The role of free radicals, lipid peroxides, and antioxidant activities in the occurrence of muscular phenotypic switch remains to be clarified. An oxidative imbalance-driven phenotypic switch of human colonic smooth muscle cells (HSMC) has been demonstrated in response to bacterial lipopolysaccharide (LPS) exposure via its interaction with toll-like
receptor 4 (TLR4), constitutively expressed by colonic SMC [170]. LPS-induced contractile dysfunction in SMC, consisting of a time- and dose-dependent decrease in cell length and contraction, was associated with ROS production, GSH content depletion, hyperpolarization of mitochondrial membrane, and rearrangement of actin microfilament cytoskeleton. Most of these effects were partially prevented by the NADPH oxidase inhibitor apocynin or the NF-κB inhibitor MG132, supporting the important pathogenic role of oxidative stress (Figure 5) [171]. Thus, the exposure to bacterial endotoxin directly and persistently impaired gastrointestinal smooth muscle activity, strongly indicating that HSMC could actively participate in the dysmotility observed during the infective burst.

**Figure 5.** Effects of oxidative stress on gastrointestinal smooth muscle cells. Oxidative stress can cause both cell structure-function alterations and inflammation with the promotion of pro-inflammatory environments. As concern cell phenotype, ROS alter the production of cytoskeletal proteins, like smooth muscle myosin heavy chain (SMMHC) and smoothelin (SM) that leads to the impairment of contraction and cell length. The final consequence of these alterations is the cellular switch from contractile to the synthetic phenotype. In the presence of oxidative stress, molecular signaling results are also altered. In particular, signaling that leads to the amplification of damage (NF-κB-signaling) are activated, complexes involved in the correct maintenance of contractile phenotype (RhoA/ROCK-signaling) are down regulated, and eNOS expression and cAMP-signaling pathways resulted are altered. ROS have a dual effect on cells: they induce apoptosis but also favour the cell cycle progression as compensatory mechanism. An increase of ROS, or conversely a decrease of antioxidants, induce cells to an oxidative state that manifests itself also with hyperpolarization of mitochondrial membrane. The last effect of oxidative stress on cells is the alterations of protein production, in particular extracellular proteins, like collagen, and pro-inflammatory cytokines IL-6 result increased, while anti-inflammatory cytokines IL-10 result decreased. The presence of these effects were partially prevented by the NADPH oxidase inhibitor apocynin or the NF-κB inhibitor MG132, supporting the important pathogenic role of oxidative stress (Figure 5) [171]. Thus, the exposure to bacterial endotoxin directly and persistently impaired gastrointestinal smooth muscle activity, strongly indicating that HSMC could actively participate in the dysmotility observed during the infective burst.

In addition, prolonged exposure to LPS also triggers a redox imbalance that leads to profound modifications of the contractile microfilament network, with a decrease of the contractile differentiation markers smooth muscle myosin heavy chain and smoothelin, and the induction of cell proliferation, thus inducing a persistent cellular phenotype switch from a contractile to a synthetic phenotype [172]. These effects were potently counteracted by antioxidant drugs alpha-tocopherol and N-acetylcysteine (NAC), which were able to reverse the cytopathic effects of LPS and to restore normal muscle cell function.
Very interestingly, oxidative damage has also been reported on vascular smooth muscle cells (VSMCs), major cell types in the vessel wall, which has different functions. As colonic SMC, VSMCs present two different phenotypes: contractile and synthetic [173]. When subject to inflammation and oxidative stress, VSMCs switch from the contractile to the synthetic phenotype. It was hypothesized that the phenotypic switch may play an important role in several cardiovascular diseases [174]. Indeed, the phenotypic switch, while causing the loss of the contractile ability of the VSMC, gives it the ability to recruit inflammatory cells, thus inducing a remodeling of the vessel wall at the basis of the vascular damage [173]. The crosstalk between oxidative stress and inflammation therefore represents a key element in endothelial dysfunction and consequent vascular damage. Endothelial dysfunction is known to be characterized by an impaired ability to regulate vascular tone and altered anti-inflammatory and anticoagulant properties [155,174]. For this reason, endothelial dysfunction has been accepted as primum movens in the development of hypertension [175] and atherosclerosis [156,176].

By contrast, the exact molecular mechanisms underlying the intestinal muscle alterations induced by oxidative stress remain to be clarified. This would be very important in order to provide new insights in the pathophysiology of persistent gut dysmotility disorders that occur, for instance, in post-infective conditions (i.e., post-infective IBS) [177] or during remission of IBD.

Furthermore, shedding light on the role of oxidative stress in gut dysmotility disorders is essential for the development of new therapeutic approaches, also with a possible reappraisal of antioxidants molecules in the clinical management of these disorders.

7. Therapeutic Strategies Based on the Use of Antioxidants

Currently, the treatments of gastrointestinal inflammatory diseases, such as CD and UC, include a combination of immunosuppressive agents and anti-inflammatory, although sometimes adverse effects may occur [178,179]. The treatment can also include probiotics and prebiotics in order to normalize the microbiota favoring the species with a greater antioxidant capacity [180]. Importantly, modifications in diet and lifestyle are always recommended. Pharmacological treatments aim at blocking TNF-α or NF-κB-mediated inflammation. Given the close connection between inflammation and oxidative stress, the use of antioxidant supplements could modulate endogenous mechanisms of ROS production, through inhibition of oxidizing enzymes, or enhancing the antioxidant system activity (Table 4). As mentioned above, the role of oxidative stress, either as the main cause or secondary effect of the inflammation, was been demonstrated in inflammatory bowel disease, gastritis, and peptic ulcer disease and to a lesser extent, in celiac disease, inflammatory bowel syndrome, and esophagus cancer. Hence, modulation of oxidative stress represents an evidence-based rational choice to improve gastrointestinal disorders.

Among the antioxidants, polyphenols are widely used in the treatment of gastrointestinal diseases. In particular, they have antioxidant, anti-inflammatory, and immunomodulatory action. Polyphenols are able to inhibit the cytokines production, such as IL-8, IL-1β, and TNF-α, to promote the activities of intracellular antioxidants, including SOD and GPX, and to directly scavenge free radicals. In addition, polyphenols protect the intestinal mucosal by reducing intestinal permeability via tight junction stabilization. They also enhance the healthy microbiota in the gut [181]. Polyphenols include flavonoids and phenolic acids (Figure 3). Among the polyphenols, resveratrol is used in the treatment of H. Pylori-related disease [182] and to improve IBD [183–185], thanks to its anti-inflammatory and antioxidant properties. Moreover, this compound has also been reported to inhibit the proliferation of gastric cancer cells [186].

Studies in animal models have also shown that antioxidants such as curcumin, boswellic acids, planar aromatics, and rosmarinic acid were able to activate the NRF2 pathway [187–191]. The antioxidant supplements may also decrease ROS levels by affecting specific enzymes. For example, allopurinol inhibited the action of xanthine oxidase that generates O2•−. Curcumin hindered inflammation by decreasing COX activities, IL-
16, and the AKT/mTOR pathway. Several clinical studies have reported the benefits of curcumin in IBD patients without serious side-effects [192,193]. Flavonoids prevent ROS production by inhibiting XO [194], COX, LOX, GST, and NADH oxidase [195]. Moreover, several flavonoids are also able to chelate free Fe²⁺ and Cu²⁺ that could increase ROS generation [196].

Table 4. Therapeutic antioxidant strategies.

| Compounds          | Effects                                                                 | GI Disease                  | Ref.          |
|--------------------|------------------------------------------------------------------------|----------------------------|--------------|
| Melatonin          | Anti-inflammatory and anti-oxidant action, Lipid peroxidation reduction | PI-IBS                     | [185]        |
|                    | MAPK and NF-κB Activation, iNOS expression induction, Stimulation of nitrite production in intestinal tissue, Inhibition of cytokines production, Promotion of intracellular antioxidants activity | IBD                        | [181]        |
| Polyphenols        | Scavenging direct of free radicals, Protection of intestinal mucosal, Enhancing of eubiosis | IBD                        | [181]        |

SPECIFIC POLYPHENOLS

| Compounds          | Effects                                                                 | GI Disease                  | Ref.          |
|--------------------|------------------------------------------------------------------------|----------------------------|--------------|
| Resveratrol        | Anti-inflammatory and anti-oxidant activity, Inhibition of gastric cancer cells proliferation | IBD                        | [182,183,186]|
| Allopurinol        | Inhibition of XO activity, Inhibition of COX activity                  | ID                         | [192,193]    |
| Curcumin           | Inhibition of IL-1β, Inhibition of AKT/mTOR pathway                    | IBD                        | [192,193]    |
| Flavonoids         | Scavenging direct of free radicals, Protection of intestinal mucosal, Chelation of free Fe²⁺ and Cu²⁺ | IBD                        | [184,194–196]|

PI-IBS: post-infective Irritable Bowel Syndrome; IBD: Inflammatory Bowel Disease; ID: Inflammatory Bowel Disease; NOX: NADPH oxidase; XO: xanthine oxidase; COX: cyclooxygenases; LOXs: lipoxygenases; GST: glutathione transferase; iNOS: inducible nitric oxide synthase.

Recently, new technologies and nanomaterials have been developed that can improve the targeted administration of antioxidant drugs in gastrointestinal inflammation [197,198]. In particular, nanomaterials can be designed in such a way as to have antioxidant properties themselves, thus acting as nano-antioxidants, or as carriers and/or containers of natural compounds with antioxidant activity, mainly polyphenols [199]. Many literature data have reported the use of exosomes, lipid, and magnetic polymers; natural and synthetic polymers; ultrasonic microtubules; and nanoemulsions for the targeted release of antioxidants, in particular curcumin [200–202], resveratrol, and vitamin E [203] in the treatment of gastrointestinal inflammation, mostly IBD [204] and gastric ulceration [205]. It is very important to note that, although the use of nanostructures is associated with better bioavailability or enhancing pharmacokinetics, at high concentrations, they could be very toxic, releasing the drug out of control, and increasing oxidative stress through the generation of ROS. Therefore, they need further studies before any preclinical and clinical applications [199].

8. Conclusions

Generally, our body counterbalances the production of reactive species with the endogenous antioxidant defense system or by antioxidants introduced with the diet. When this balance is not maintained, oxidative stress occurs. Oxidative stress contributes to the evolution of numerous and diverse pathologies also those of the gastrointestinal tract. As shred of evidence shows that the use of antioxidants can improve the evolution of many diseases, the development of antioxidant therapies represents a promising avenue also for the gastrointestinal pathologies treatment. Therefore, the knowledge of the peculiar
oxidative pathway involved in each disease could allow both the identification of disease markers and the development of preventive and curative therapeutic strategies.

**Author Contributions:** R.V. conceived the idea, drafted the manuscript, and drew the figures relative to oxidative stress; L.P. contributed to the design, drafting of the manuscript, and drew the figure relative to gastrointestinal diseases. M.C. drew the figure relative to SMC. C.S. contributed to conceiving the idea and revising the manuscript; P.M. contributed to conceiving the idea, drafting the manuscript, and concurring to the final version of the review. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was partially funded by the Peretti Foundation (3603) to P.M., and Sapienza 2018-000324_19_RS_SEVERI to C.S.

**Acknowledgments:** Editorial assistance was provided by Veronica Scascafratte, and Chiara Serrao.

**Conflicts of Interest:** The authors declare no conflict of interest.

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