T-cell Subset Regulation in Atopy

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Abstract Presentation of processed allergen by antigen-presenting cells to T-helper (Th) lymphocytes, which is influenced costimulatory signals, cytokines, chemokines, and regulatory T cells (Tregs), determines the development of different types of T-cell immunity. The discovery of Tregs revolutionized the primary concepts of immune regulation interpreted within the framework of a binary Th1/Th2 paradigm. Tregs play a central role in the maintenance of peripheral homeostasis, the establishment of controlled immune responses, and the inhibition of allergen-specific effector cells. Recently, some other T-cell subsets appeared, including Th17 and Th9 cells, which control local tissue inflammation through upregulation of proinflammatory cytokines and chemokines. This review aims to discuss our understanding of the T-cell subset reciprocal interaction in atopy.

Keywords Atopy · T cells · Tolerance

Introduction

The activity of T lymphocytes is largely dependent on specialized introduction of antigenic materials by highly specialized antigen-presenting cells [1]. Thus, dendritic cells (DCs) are essential for the differentiation of naïve T cells into T-helper (Th)1, Th2, Th9, Th17, and Th22 effector T-cell subsets [2] via release of cytokines and other cofactors (Fig. 1). The differentiation of naïve T cells into Th2 cells is induced in the presence of interleukin (IL)-4. Allergen-specific Th2 cells acting through the effects of released IL-4 and IL-13 induce immunoglobulin class switching to IgE and clonal expansion of naive and IgE-positive memory B cells. Cross-linking of IgE bound to FceRI (high-affinity receptor for IgE) on the effector mast cells and basophils results in the release of vasoactive amines (including histamine); lipid mediators (eg, prostaglandin D, platelet-activating factor, leukotriene [LT]C4, LTD4, and LTE4); chemokines (CXC-chemokine ligand 8 [CXCL8], CXCL10, CC-chemokine ligand 2 [CCL2], CCL4, and CCL5); and cytokines such as IL-4, IL-5, and IL-13, leading to the development of type 1 immediate hypersensitivity reaction [3]. Then, after 6 to 12 h, should a late-phase allergic reaction occur, it appears due to the migration of allergen-specific T cells, which are reactivated and clonally expand under the influence of chemokines and other cytokines at the site of allergen exposure. The major effector cells are eosinophils; however, Th1 cells, mast cells, and basophils also contribute at this phase [3]. The cellular chronic late-phase response, which is mostly driven by allergen-specific T cells activated by continuous allergen exposure, is the driving force for the persistent inflammation and/or tissue remodeling responsible for the chronic symptoms of allergic disease [4].

Effector T-cell Subsets

Naive T lymphocytes are primed by DCs that mature and migrate from peripheral tissues to the T-cell areas of
secondary lymphoid organs, where they produce regulatory cytokines [5]. Two distinct subsets of DCs have been described in humans. Myeloid DCs (mDCs) [6] express Toll-like receptor (TLR2) through TLR6 and TLR8 and produce IL-12 in response to the bacterial and viral stimuli. Plasmacytoid DCs (pDCs) express TLR7 and TLR9 and release large quantities of type 1 interferons during the outcome of antiviral immune responses [5, 7–9]. pDCs directly suppress the ability of mDCs to generate effector T cells [10]. pDCs are capable of stimulating the development of regulatory T cells (Tregs), probably via the inducible costimulatory molecule (ICOS) ligand–dependent pathway [8, 10]. It has been shown that the depletion of pDCs in the lungs results in lack of tolerance to inhaled antigens [10, 11]. The other two DC populations that are present at inflammatory sites of the skin in atopic dermatitis are the classical Langerhans cells and the inflammatory dendritic epidermal cells [12]. It has been recognized that inflammatory dendritic epidermal cells activate the Th1-cell subset. Classical Langerhans cells induce the Th2-cell subset [13]. Furthermore, the expression of IgE FcεRI on the surface of DCs and Langerhans cells is not specific to atopic eczema but rather determined by the type of inflammatory status in various inflammatory skin diseases. [14].

It has been demonstrated that stem cell factor, the ligand for c-kit, which is a receptor tyrosine kinase type III, affects the synthesis of IL-6 and Jagged-2, the ligand of Notch. Engagement of Notch on the T cells with Jagged on DCs and T-cell receptors with major histocompatibility complex-II–coupled peptide induces priming of Th cells [15]. Thus, Notch affects the Th-cell differentiation by promoting the Th2 and Th17 development but shows no effect on Th1 response [16]. Apoptosis-resistant DCs effectively generate antigen-specific Th2 cells in vitro and in vivo and induce IgE responses in vivo independent of the sensitization status of the host [17].

Th1 and Th2 Cells

The subsets of CD4+ Th lymphocytes are categorized on the basis of their distinct cellular functions and cytokine secretion capacities [18]. Originally, two subsets—Th1 and Th2 lymphocytes—were described, which accounted for a binary Th1/Th2 paradigm. Since this time, new Th subsets have been recognized as important players in immune regulation. They include well-characterized Tregs as well as newly described proinflammatory Th17 or Th9 cell lineages. In the near future, it is likely that new Th subsets will be described and characterized. The imbalance between...
various T-cell subsets accounts for different immune pathologies, including atopy.

Peripheral T-cell clones differentiate into these subsets using self-reinforcing transcriptional circuitries that involve major transcriptional regulators: T-box expressed in T cells (Tbet) in Th1 cells, trans-acting T-cell–specific transcription factor (GATA-3) in Th2 cells, forkhead box P3 (FoxP3) in Tregs, and retinoic-related orphan receptor (ROR)γt/RORα in Th17 cells. It has been recognized that the counter-regulation between the T-cell effector subsets is crucial in immune regulation and in maintaining the balance between different types of the immune response [19–22]. Thus, the activation of Tbet inhibits both Th2 cell–mediated eosinophil recruitment and Th17 cell–mediated neutrophil recruitment into the airways [23]. An association between a specific Tbet haplotype and allergic asthma in children has been suggested [24].

Th2 cells predominantly mediate IgE responses and allergic inflammation and are also involved in immunity to parasites [3, 25]. There are likely to be several mechanisms of Th2 profile dominance in atopic disease. It has been postulated that Th1 cells are prone to activation and apoptosis, as their high IFN-γ–producing fraction and CXCR3+ T cells in particular show increased apoptosis in atopic individuals [26]. Th1 cells are involved in cell-mediated defense against intracellular microorganisms and in promotion of memory IgG responses. Their phenotype is dominated by IL-2, IFN-γ, and tumor necrosis factor-β cytokine profiles. Th1 cells differentiate after stimulation with IL-12, IL-18, and IL-17. It must be emphasized that Th1 cells also engage in the effector mechanisms of allergic disease. It has been recognized that they induce apoptosis of keratinocytes in atopic dermatitis. In asthma, these cells induce apoptosis of epithelium and bronchial smooth muscle cells [27, 28].

The phenotype of Th2 cells is characterized by secretion of proinflammatory cytokines IL-4, IL-5, and IL-13, which are clustered on chromosome 5q in close proximity with granulocyte-macrophage colony-stimulating factor. These cytokines are associated with pathogenesis of IgE and eosinophilia. Th2 polarizing factors include monocytic chemotactic protein 1 and OX40 ligand [29]. OX40 ligation upregulates IL-4 production, which in turn promotes Th2 polarization [29]. Thymic stromal lymphopoietin (TSLP) is a novel growth factor produced by epithelial cells that promotes the proliferation and differentiation of committed B-cell progenitors. It can substitute for the activity of IL-7, the B-cell growth and maturation factor. TSLP plays an essential role in allergic inflammation at the epithelial cell and DC interface [30] and activates human mDCs to induce inflammatory Th2 responses [31]. TSLP levels are increased in asthma [32]. TSLP-induced DCs mature and migrate into the draining lymph nodes and trigger the adaptive phase of allergic immune response. TSLP induces OX40L expression in DCs, which induce the differentiation of allergen-specific naive CD4+ T cells to inflammatory Th2 cells [33].

Recently, IL-25 (IL-17E), a member of the IL-17 family of immunoregulatory cytokines, was shown to regulate Th2-type immunity. Blocking of IL-25 in an experimental model of allergic asthma prevented airway hyperresponsiveness and suppressed IL-5 and IL-13 synthesis, as well as eosinophil infiltration, goblet cell hyperplasia, and serum IgE secretion [34].

Regulatory T Cells

The discovery of Tregs contributed tremendously to the understanding of the mechanisms of peripheral tolerance and induction of Th1 or Th2 immunity [35]. Tregs are characterized by their IL-10 and transforming growth factor (TGF)-β secretion capacities [36]. Tregs suppress allergen-induced specific T-cell activation. They also suppress effector cells of allergic inflammation such as mast cells, basophils, and eosinophils [37, 38] as well IgE production [38–40]. Notably, Treg-derived cytokines IL-10 and TGF-β induce synthesis of noninflammatory IgG4 and IgA isotypes, respectively [37]. Several subsets of Tregs showing distinct phenotypes and regulatory effects have been recognized. They include the naturally occurring, thymus-selected CD4+CD25+FoxP3+ Tregs and the inducible type 1, IL-10–secreting Tregs [41]. Furthermore, regulatory subsets of CD8+ T cells, γδ T cells, DCs, IL-10–producing B cells, natural killer cells, and resident tissue cells, which may promote the generation of CD4+ Tregs, have been described [42].

The mechanisms by which Tregs induce peripheral tolerance to allergens may go beyond the effects of cytokines produced by these cells. They may directly interact with DCs and compete with naïve T cells in a physical manner by creating aggregates around DCs and in this way inhibit their maturation [43]. In addition, Tregs downregulate the expression of CD80/CD86 on DCs [43]. Naturally occurring Tregs express high cytotoxic T lymphocyte antigen-4 (CTLA-4). CTLA-4 is a CD28 family member that binds CD80/CD86. However, it does so with a higher affinity than CD28. In contrast to stimulatory effects of CD28, CTLA-4 inhibits T-cell activation [44–46].

Regarding other IL-10–producing cells (eg, monocytes), compared with healthy controls, their percentage has been shown to be significantly increased in atopic patients [47]. They preferentially differentiate into SOCS3-expressing, alternatively activated macrophages, which induce Th2 immune response [47]. The physiologic relevance of this phenomenon is unclear. IL-10–treated DCs poten-
press the airway inflammation and hypersensitivity as well as Th2 cytokine production. These effects are exerted through endogenous production of IL-10 [48]. At the molecular level, IL-10 inhibits CD28 and ICOS costimulations of T cells via Src homology 2 domain-containing protein tyrosine phosphatase (SHP)-1 [49]. IL-10 receptor–associated tyrosine kinase (Tyk)2 acts as a constitutive reservoir for SHP-1 in resting T cells. Tyrosine phosphorylates SHP-1 upon IL-10 binding. SHP-1 rapidly binds to CD28 and ICOS costimulatory receptors and dephosphorylates them within minutes. Binding of phosphatidylidylinositol-3-kinase to either costimulatory receptor is inhibited, and downstream signaling is consequently blocked. In an experimental model, spleen cells from SHP-1–deficient mice showed increased proliferation with CD28 and ICOS stimulation in comparison with wild-type mice, in which cells were not suppressed by IL-10. It has been postulated that the inhibition of the CD28 or ICOS costimulatory pathways by SHP-1 may provide a novel approach for direct T-cell suppression [49]. Src homology 2 domain-containing inositol 5-phosphatase 1 has been shown consistently to inhibit allergic reactions as a negative regulator of cytokine and immune receptor signaling. On the contrary, its deficiency leads to spontaneous development of allergic-like inflammation in the murine lung [50].

TGF-β is another key player in immune tolerance. It exerts its activity in the intestinal mucosa and can induce immune tolerance to dietary antigens [51]. BALB/c mice treated orally with ovalbumin (OVA) and TGF-β showed augmented reduction of OVA-specific IgE and IgG1 antibodies, T-cell reactivity, and immediate-type skin reactions when compared with mice treated orally with OVA alone [51]. Thus, local administration of TGF-β may become a strategy to prevent and treat allergic diseases.

Tregs can directly inhibit the effector cells of allergic inflammation [52]. Tregs directly inhibit the FcgR1-dependent mast cell degranulation through cell–cell contact involving OX40–OX40 ligand interactions [53]. The depletion or inactivation of Tregs causes enhancement of the anaphylactic response [2, 53]. IL-10 regulates mast cell maintenance and proliferation in peripheral tissues [54] and reduces proinflammatory cytokine release from mast cells [55]. IL-10 also inhibits eosinophil activities [56].

IL-10 is a potent suppressor of allergen-specific IgE, whereas it induces IgG4 production [37]. IL-10 decreases ε transcript expression, thus downregulating IgE production, while enhancing γ4 transcript expression and increasing IgG4 production [57]. Supporting evidence has been provided by studies on Treg–B-cell interaction through glucocorticoid-induced TNFR-related protein (GITR)/GITR ligand [58]. The induction of IgG4 and suppression of IgE in healthy individuals shows the direct influence of Tregs on B cells [39].

During allergen-specific immunotherapy, the development of peripheral T-cell tolerance is characterized mainly by the generation of allergen-specific Tregs, which suppress proliferative and cytokine responses against the major allergens [37, 38, 59–67]. Moreover, local FoxP3CD25+ T cells in the nasal mucosa and their increased numbers after immunotherapy were demonstrated [68]. Treg-derived IL-10 not only generates tolerance in T cells but also regulates specific isotype formation and skewsthe specific response from an IgE to an IgG4-dominated phenotype [38]. IL-10 is a potent suppressor of total and allergen-specific IgE, while it simultaneously increases IgG4 production. During allergen-specific immunotherapy, increases in allergen-specific IgG4 and IgG1, with 10- to 100-fold increases in their serum levels, were observed [69, 70].

Th17 Cells

Th17 cells have been shown to induce host protection against extracellular pathogens and tissue inflammation [20, 71, 72]. Differentiation of Th17 (IL-17A– and IL-17 F–producing) cells is induced by IL-6, IL-21, IL-23, and TGF-β [2]. Th17 cells have been implicated in the pathogenesis of autoimmune diseases [73]. They are characterized by a strong IL-17–producing capacity. IL-17 family cytokines include IL-17A through F, which control inflammatory responses in the tissue by triggering secretion of proinflammatory cytokines and chemokines [71, 74]. Tregs can suppress Th17 cells and autoimmunity. Activation of Th17 cells can start tissue inflammation [75]. IL-17 has been shown to coordinate granulocyte influx in allergic airway inflammation models [76, 77]. A member of the IL-17 family, IL-17 F (IL-25), is designated to the Th2-type group. It promotes Th2-type response; induces eosinophilia; increases serum IgE and IgG1; and upregulates tissue expression of IL-4, IL-5, and IL-13 [78–80].

Th9 and Th22 Cells

Th9 represents a novel, distinct population of effector T-helper cells involved in tissue inflammation [2]. It shows no suppressive functions. This population is characterized by IL-9 and IL-10 secretion. These cells differentiate from naïve cells after IL-4 and TGF-β stimulation. It has been shown that IL-9 together with TGF-β can contribute to Th17 cell differentiation, and Th17 cells themselves can produce IL-9 [81]. Th9 cells have been shown to induce tissue inflammation in autoimmune diseases, however, the role of this T-cell subset in allergic responses remains to be elucidated. Another novel T-cell subset has been demonstrated in T cells that independently express IL-22 with low expression levels of IL-17 and play a role in atopic dermatitis. IL-22 can be protective in animal models of
Conclusions

Many factors contribute to development of the Th2 profile in atopic individuals. Th2-derived IL-4 and IL-13 exert an important role in class switching in B lymphocytes that results in IgE production. Recent evidence on T-cell subset reciprocals regulation and counterbalance between Th1 and Th2 cells to Th17 and Tregs has greatly influenced our understanding of immunoregulation and peripheral tolerance. Different types of Treg control several facets of allergic inflammation. They regulate IgE versus IgG4 and drive the allergen-specific antibodies toward the noninflammatory and nonanaphylactic phenotype. They also directly or indirectly inhibit effector cells and consequently restrict allergic inflammation. The understanding of these insights into the mechanisms of immunoregulation leads to novel therapeutic approaches.

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Disclosure

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