Interactions between tall oatgrass invasion and soil nitrogen cycling

Eve-Lyn S. Hinckley1,2 · Hannah R. Miller1,2 · Ann Lezberg3 · Brian Anacker3

Received: 5 July 2021 / Accepted: 20 May 2022 / Published online: 7 June 2022 © The Author(s) 2022

Abstract

Increases in nitrogen (N) inputs to the biosphere can exacerbate the introduction and spread of invasive non-native plant species. Often, with elevated soil N levels, invasive plants establish and further enrich soil N pools, changing overall ecosystem function. This study examined the relationship between soil N cycling and an increasingly prevalent, invasive plant species, tall oatgrass (Arrhenatherum elatius subsp. elatius), in foothills ecosystems between the Colorado Rocky Mountains and the Denver-Boulder Metropolitan area—similar to many Western US grasslands and woodlands. It focused on investigating differences in soil N transformations, inorganic N pools, and vegetation characteristics across invaded and uninvaded plots at three sites in two seasons (summer and autumn). There was a statistically significant effect of invasion on rates of net N mineralization, but it was dependent on site and season (p = 0.046). Site had a statistically significant effect on soil moisture and aboveground biomass C:N (p < 0.04). The interactions of invasion x site were statistically significant for ammonium pools (p < 0.03). These findings suggest that A. elatius invasion can be associated with accelerated N cycling, but that the nature of the relationship differs by location and season in the foothills. More broadly, this study contributes to determining how the N cycle is shifting in grassland ecosystems subject to increasing pressures from anthropogenic change.

Keywords Biogeochemistry · Invasion ecology · Nitrification · Nitrogen mineralization · Plant-soil interactions · Restoration · Species diversity · Tallgrass prairie

Introduction

Worldwide, increases in atmospheric N deposition to the biosphere have exacerbated the introduction and spread of invasive non-native plant species and lowered plant diversity (Ehrenfeld 2003; Davidson et al. 2011; Field et al. 2014). Prior research demonstrates that high soil N environments resulting from elevated N inputs can give invasive plants a competitive advantage (Dukes and Mooney 1999; Clark and Tilman 2008). In addition, invasive plants can affect soil N cycling via several mechanisms, including N fixation (e.g., Asner et al. 2008; Kurokawa et al. 2010), production of N-rich tissues (e.g., Kurokawa et al. 2010; Lee et al. 2017), and biomass growth (e.g., Aguilera et al. 2010). Often, a positive feedback ensues: the invasive continues to enrich the soil inorganic N pool, perpetuating conditions that favor its success or the success of other non-natives (Corbin and D’Antonio 2003; Liao et al. 2008; Shaben and Myers 2010), or changing plant species composition and accelerating N cycling (Bobink et al. 2010). This interaction between elevated N inputs and plant species invasion can create notable changes in ecosystem function that make restoration activities challenging for land managers.

For the last several decades, the foothills and prairie ecosystems on the eastern slope of the Colorado Rocky Mountains have experienced the interacting effects of elevated atmospheric N deposition and changes to plant species composition due to invasive species introductions. Crawford and colleagues (2020) report that atmospheric N deposition as nitrate—once nearly double that of the less populated western slope (Baron et al. 2000)—is declining, likely due to air pollution regulation. However, ammonium-N inputs are
increasing, which may be sourced from agricultural areas in Eastern Colorado. At the same time, the invasive perennial grass, tall oatgrass (Arrhenatherum elatius subsp. elatius), has replaced large areas once dominated by native prairie species (EnvironPlan Partners 2018). Originally introduced in the mid-nineteenth century by ranchers, spread of A. elatius was likely controlled by grazing. However, with cessation of grazing around the 1960s, as well as fire suppression and possibly elevated atmospheric N deposition levels, it spread rapidly, and its expansion continues today, making it the predominant plant species in invaded areas (EnvironPlan Partners 2018). During the last decade, land managers have been evaluating several approaches to contain A. elatius, including prescribed fire, pesticide applications, and grazing.

Previous research on the relationship between A. elatius invasion and the soil N cycle have focused on several factors related to structural and soil chemical effects of the invasive. For example, Berendese et al. (1992) found that areas receiving N fertilizer inputs within hayfields of the Netherlands favored persistence of A. elatius compared with unfertilized areas that were dominated by native red fescue (Festuca rubra). In addition, they found that clipping A. elatius caused more allocation of N to its aboveground tissues, which led to increased ecosystem losses of N, compared with uninvaded areas. Other studies determined that A. elatius increases rates of decomposition above even that of other aggressive invasive plant species (Holub et al. 2012).

The goal of our study was to determine if A. elatius is associated with an altered soil N cycle in sites where it is a serious threat to ecosystem health and integrity. Specifically, we addressed the question: what is the nature of the relationship between A. elatius and the soil N cycle in foothills and grassland ecosystems? Given that invasive plants often speed up N cycling through high quality litter and rapid growth, we hypothesized that A. elatius would be associated with higher soil inorganic N (ammonium and nitrate). In addition to evaluating differences in net N cycling rates, we also measured other ecosystem factors that might affect N cycling rates (soil moisture) or change in response to invasion (aboveground biomass and aboveground biomass C:N). Our study is a first step to inform future efforts that address the interactions among A. elatius, soil N cycling, and management practices. More broadly, it can provide a foundation for conducting process-based study of the relationships among atmospheric N deposition, soil N cycling, and ecosystem functioning from the plains to the alpine.

**Methods**

**Study area**

We conducted this research at three grass-dominated study sites in the foothills of the Colorado Front Range, all managed by Boulder Open Space and Mountain Parks (OSMP) in Boulder, Colorado, US (Fig. S1, Table 1). We chose three sites to represent some of the different vegetation structural types in which A. elatius grows across the foothills area. We refer to the sites with labels that represent the growth form of the tallest layer of vegetation at each site. These sites include: “shrubland”, located in Coyote Canyon, a steep ravine with north-facing (invaded) and south-facing (largely uninvaded) slopes supporting a mix of scattered shrubs and grass-dominated herbaceous vegetation (Fig. S1a); “grassland”, located south of the National Center for Atmospheric Research (NCAR) mesa where mixed grass prairie has been invaded by broad swaths of dense A. elatius, the highest percent cover observed in this study (Fig. S1b); and “savanna”, located on Shanahan Ridge in an open woodland area with dense patches (~5–10 m) of A. elatius invasion interspersed with other grasses and trees (Fig. S1c).

Average monthly air temperature in the Denver-Boulder Metropolitan area ranges from approximately 0 °C in winter to 21 °C in summer and mean annual precipitation to the foothills of the Colorado Front Range is ~790–890 mm (Wetherbee et al. 2019).

**Study design**

At each of three study sites, we established three 2 m × 2 m plots within A. elatius-invaded and uninvaded areas, respectively (n = 18 across sites and treatments). The patchiness of A. elatius invasion precluded random plot location. Thus, we chose plot locations based on

| Site       | Slope (%) | Soil type¹ | Bulk density (g cm⁻³) | Soil pH² | Soil C:N² | Soil N (%)² |
|------------|-----------|------------|-----------------------|---------|----------|-------------|
| Grassland  | 15        | Colluvial land, gravelly sandy loam | 0.89 | 6.69±0.17 | 13.5±1.1 | 0.40±0.01 |
| Shrubland  | 28        | Colluvial land, gravelly sandy loam | 1.02 | 6.25±0.60 | 17.0±1.4 | 0.24±0.04 |
| Savanna    | 1         | Nederland very cobbly sandy loam    | 0.83 | 6.17±0.21 | 15.4±2.7 | 0.34±0.04 |

¹Soil type determined from NRCS survey area (version 17, 5 June 2020)

²n = 3 per site
the following criteria: invaded plots were placed within patches of 40–80% A. elatius cover, and uninvaded plots were located at least 2 m from the nearest A. elatius individual, with 0% A. elatius cover within the plot. At the grassland and savanna sites, the plots were placed in uninvaded and invaded areas along a common slope and aspect. At the shrubland site, plots were located along two transects parallel to the slope.

**Field sampling**

Each site was sampled two times during the study period (Sept 2019–Oct 2020) for soil N processing rates, inorganic N pools, and gravimetric soil moisture. We targeted two key phenological seasons to make these measurements: Summer, Jun–Jul 2020, to capture peak biomass, and autumn, Sept–Oct 2020, to measure plant senescence and seasonal transition. For each sampling period, paired soil cores (3 cm diameter × 10 cm depth) were collected at each plot, roots were removed (to exclude plant N uptake), and the cores were placed in plastic bags, one for immediate analysis of soil inorganic N species, and the other returned to its borehole for incubation in the field, following the in-field buried bag technique described by Hart et al. (1994). Both cores were extracted for inorganic N concentrations and then the rates of net N cycling processes were calculated by differing the results of the final, incubated core and the initial core (see “Data Analysis”). Following incubation (~30 days), soil cores were transported to the laboratory for analysis in the same manner as the ones subjected to immediate analysis (see description below). Once during the study period, we collected soil cores for soil C:N, soil pH, and bulk density (Sept 2019, and Sept and Oct 2020, respectively).

In addition to soil measurements, we sampled vegetation within each plot for aboveground biomass, C:N ratios, and A. elatius abundance (as percent aerial cover). We collected samples from 0.5 m × 0.5 m quadrats (one per plot) in Sept 2019. Standing stems plus leaves and litter (“thatch”) cover were clipped to the ground surface within the quadrat; it is important to note that these collections occurred when A. elatius had senesced at the end of summer/early autumn. In the laboratory, aboveground biomass samples were dried at 60 °C for 48 h, then weighed to determine dried biomass per unit area. After cutting and mixing each sample, a homogenous subsample of aboveground biomass tissue was pulverized with a mortar and pestle and analyzed for total C and N, as described below. We estimated plant species cover within a 1 m² area of each plot using visual cover and then identified the presence of all remaining species within the remaining 4 m² plot. Dominant plant species were any species that had >20% areal cover.

**Laboratory analysis**

Immediately upon return from the field, soil cores were hand-picked to remove roots and rocks. Subsamples of field-moist soil were extracted in 2 M potassium chloride (KCl), shaken for 2 h, and then filtered using Whatman 1 filters, modified from Binkley et al. (1986) and Hart et al. (1994). Extractants were analyzed for ammonium and nitrate concentrations with a Lachat QuikChem 8500 Flow Injection Autoanalyzer, detection limit of 0.005 mg NH₄⁺ –N L⁻¹ and 0.004 mg NO₃⁻ –N L⁻¹, respectively. In addition, a sub-sample of each soil sample was analyzed for gravimetric soil moisture (dried at 105 °C for 48 h) and subsamples of oven-dried (60 °C for 48 h) and ground soil and vegetation tissue were analyzed for total C and N by combustion on a Thermo Finnigan Flash EA 1112. All analyses were completed in the Arikaree Environmental Laboratory at University of Colorado, Boulder.

**Data analysis**

We calculated the rate of net N mineralization by differencing the mass of inorganic N (ammonium) in final and initial soil extracts and dividing by the incubation period to get the mass of N produced per gram of dry soil per day for each of two seasons. We also calculated the rate of nitrification but it was highly correlated with mineralization (r² = 0.94, p < 0.001), and, therefore, not analyzed further. Net N mineralization rate is reported on an areal basis using bulk density to scale. We fit three linear models, one per response variable—net N mineralization rates, soil ammonium pools, and soil moisture—to examine the relationship between A. elatius invasion and soil properties. Models share the same predictors (site, invasion, season, and their interactions) as fixed effects and have 36 observations. We fit two additional models for aboveground vegetation (aboveground biomass and C:N ratios). Finally, we created a structural equation model (SEM) to relate A. elatius cover to soil and vegetation variables simultaneously. Due to the limited nature of our sample size, we rely on the linear models for the interpretation of our results in the main text and provide the SEM results in the Supplementary Material.

We used R (version 1.2.5033) for all analyses, including the following packages: car, dplyr, forcats, ggplot2, MASS, semTools, semPlot, tidyr, and vegan. The complete dataset (10 variables, 36 observations) is included in the Supplementary Material (Table S1).

**Results**

The characteristics of the three sites (i.e., locations and soils) are summarized in Table 1. The linear models demonstrated that invasion by A. elatius was associated with...
higher rates of net N mineralization, but this result was dependent on site as well as season (site × invasion × season, \( p = 0.046 \); Fig. 1a; and 1b Table 2). Rates were highest in summer, in invaded plots, and at the grassland and shrubland sites; the savanna did not match our initial expectations and had lower net N mineralization in the invaded than the uninvaded plots. Soil ammonium pools were not well predicted (\( r^2 = 0.06 \)), but invasion was associated with lower ammonium pools in some contexts (site × invasion \( p = 0.028 \); Fig. 1c and 1d; Table 2). Soil nitrate pools were not significant relative to predictors (\( P = 0.094 \)), and, thus, are not discussed further in detail.

Soil moisture was not significantly related to invasion, but site and seasonal differences were obvious (site \( p < 0.001 \); season \( p = 0.03 \); Fig. 1e and 1f; Table 2). Similarly, aboveground biomass was not significantly related to invasion (\( p = 0.06 \)). This result was likely due to high variability among our plots, yet there was a pattern of higher average aboveground biomass in invaded plots at all three sites (Fig. 2a). Aboveground biomass C:N was higher in invaded plots at the grassland and shrubland sites, but not at the savanna site (site \( p = 0.03 \); site × invasion \( p = 0.015 \); Fig. 2b).

For the five linear models, we tested if each one met the assumptions of normality of residuals and homogeneity of variance. The assumptions were met for three

![Fig. 1 Comparisons of soil response variables across sites and seasons. Summer patterns are in panels a, c, and e, autumn patterns are in panels b, d, and f. n = 3 per treatment per site and bars are ± 1SE](image)

Table 2 Statistical results for each model

| Response variable      | Invasion | Site  | Season | Site × invasion | Site × season | Invasion × season | Site × invasion × season | \( r^2 \) |
|------------------------|----------|-------|--------|-----------------|---------------|-------------------|--------------------------|--------|
| Net N mineralization   | 0.02*    | 0.15  | 0.002**| 0.01*           | 0.16          | 0.35              | 0.046*                   | 0.50   |
| Ammonium               | 0.43     | 0.38  | 0.65   | 0.028*          | 0.50          | 0.43              | 0.91                     | 0.06   |
| Soil moisture          | 0.83     | <0.001**| 0.03* | 0.42           | 0.29          | 0.42              | 0.90                     | 0.40   |
| Aboveground biomass    | 0.06     | 0.43  | –      | 0.60            | –             | –                 | –                        | 0.28   |
| Aboveground biomass C:N| 0.48     | 0.03* | –      | 0.015*          | –             | –                 | –                        | 0.49   |

*\( p < 0.05 \); **\( p < 0.01 \)
of the five models. For the two offending models (soil ammonium pools and net N mineralization rates), model assumptions could be met via transformation of the data (i.e., square root or log transformation). In the case of ammonium pools, data transformation made no difference to the assessment of data significance. In the case of net N mineralization rates, data transformation changed the p value for the three-way interaction from 0.046 to 0.067. Upon inspection, the square root transformation that we used appeared to decrease the leverage of a single outlying data point indicating a very high net N transformation rate; this change allowed model assumptions to be met but nudged the p value above alpha 0.05. Nevertheless, for convenience, we share the results from using conventional linear models and untransformed data.

The SEM was difficult to fit (i.e., failed to converge) due to our small sample size and the large among-site differences. After removing the savanna site, we were able to get a model to converge; it showed that A. elatius cover was a direct, significant predictor of vegetation variables (p < 0.001), which was, in turn, a significant predictor of soil variables (p = 0.034). More information is presented as a supplement to our study, but not discussed further here.

**Discussion**

This study addressed the question of whether A. elatius, an invasive grass in the Colorado Front Range, is associated with altered cycling of N, a nutrient that is often limiting to plant growth in terrestrial ecosystems. Elevated atmospheric N deposition from anthropogenic sources in the Denver-Boulder Metropolitan area contribute additional N inputs to ecosystems of the Colorado Front Range (Wetherbee et al. 2019; Crawford et al. 2020). We hypothesized that with greater abundance of A. elatius, soil N cycling would speed up—a common effect of invasive plant species, due to higher quality litter and rapid growth. While we know that A. elatius is not an N-fixing species, process-based studies of the plant-soil system that examine the degree to which the invasive is associated with altered soil N cycling (following Lee et al. 2017) have been largely unstudied. Our research was designed to explore this unknown.

We found that there was a significant trend toward higher rates of net N mineralization associated with invasion in two of the three sites that we studied (Fig. 1a and 1b); the grassland site, an open grassland area, and the shrubland site, a steep grassy ravine with regular shrub occurrence. We believe that the same pattern was not observed at the savanna site because plots were located across more variable soil and vegetation environments, including some under the canopy and overlying the litter of coniferous trees, which often have lower rates of net N cycling processes (Aber et al. 1991; Compton et al. 1998). In addition, there was a greater presence of other invasive species in both A. elatius-invaded and uninvaded plots (e.g., Bromus japonicus), which may influence soil N cycling and/or plant community dynamics at the savanna site. The net N rates that we observed are consistent with patterns in other ecosystems along the Colorado Front Range: at > ~ 30 kg N ha⁻¹ in two of the three invaded areas and > ~ 9.6 kg N ha⁻¹ in all uninvaded areas. Across the summer months alone, net N mineralization rates exceed atmospheric N deposition of ~ 4 kg N ha⁻¹ yr⁻¹ (e.g., Chen et al. 2020; Crawford et al. 2020). Contrary to our initial expectations, our findings highlight variable results across the three sites, regarding the association between A. elatius and an accelerated soil N cycle.

Although soil moisture levels—a first-order control on microbial activity—were significantly lower in summer than during the autumn sampling period (Fig. 1e and 1f; Table 2), microbes were active in the summer and released inorganic N in excess of their metabolic demands (i.e., > 0 mg N m⁻² day⁻¹), with more soil inorganic N released in invaded than uninvaded areas. Measurement of comparatively higher rates of net N mineralization in
summer is consistent with patterns observed by McCulley et al. (2009) in short-, mixed, and tallgrass prairie ecosystems, as well as Owen et al. (2003) in a Taiwanese grassland ecosystem. However, it is contrary to Liu et al. (2010) who observed the lowest net N cycling rates during the growing season in temperate grasslands of Inner Mongolia.

In contrast, the lower rates of net N mineralization that we observed during autumn sampling indicate potential consequences for changes to the ecosystem N balance (Fig. 1b). During autumn, net N mineralization rates were at least three-fold lower than during the summer but remained positive (indicating release of inorganic N into the soil) in invaded areas (Fig. 1a and 1b; Table 2). However, in uninvaded areas, net N mineralization rates at the shrubland and grassland sites were lower and more likely to be negative—indicating net N immobilization—in autumn (Fig. 1b). A tendency toward low net N mineralization rates is consistent with previous studies in tallgrass prairie ecosystems (Risser and Parton 1982; McCulley et al. 2009). Our results suggest that the soil N cycle in uninvaded areas may be more closed when plants senescence in the autumn; that is, any inorganic N released by microbes is more likely to be immobilized in their tissues and remain within the soil. Yet microbes in A. elatius-invaded areas may still release inorganic N in excess of their metabolic demand. Inorganic N may be subject to other ecosystem fates (e.g., leaching) or remain in the soil, leaving it enriched for the start of the following growing season.

It is important to consider the potential fates of inorganic N released into the soil. We observed an effect of site X invasion on soil ammonium pools (p = 0.06), which provides some important insights about variation in soil N dynamics across Colorado Front Range grasslands. Overall, soil ammonium pools were within the range of those reported for comparable prairie ecosystems (McCulley et al. 2009). Interestingly, of the three sites, the savanna had the highest soil ammonium pools in both summer and autumn (Fig. 1c and 1d), with invaded plots associated with higher pools than uninvaded plots. Yet, the savanna also had lower net N mineralization rates than the other sites, particularly in invaded areas (Fig. 1a and b). In contrast, the grassland, the site with greatest percent cover of A. elatius in invaded areas and comparatively higher net N mineralization rates, had lower inorganic N pools overall (Fig. 1). There, invaded areas had more ammonium during the summer than uninvaded areas, while the opposite pattern occurred in the autumn.

These contrasting examples suggest that the dynamics of both the plant and microbial communities likely differ by site (Table 2). At the savanna site, it appears that any inorganic N produced is stored longer in the soil than at the other two sites. At the grassland site, net N mineralization rates are higher and inorganic N produced may be assimilated by the more densely populated A. elatius (e.g., during autumn tillering), leading to less inorganic N stored in the soil. In the case of both sites, stored inorganic N could be subject to other ecosystem fates with the addition of snowmelt or rainfall. Past research in California, U.S. grasslands highlighted the seasonal pattern of high nitrification rates with the transition from dry to wet seasons (see Parker and Schimel 2011; Xiang et al. 2008). Similarly, Yahdjian et al. (2010) reported pulses of higher net nitrification following significant rainfall events in Patagonian grasslands. Stored inorganic N may be subject to additional ecosystem fates that we did not measure, such as leaching or denitrification to the atmosphere. Of the three sites, it appears that the savanna has the greatest potential for significant episodic N transformations or stimulation of N loss with the addition of water.

An important question that follows from our observations at the grassland and shrubland sites is: Why is A. elatius associated with accelerated soil N cycling rates at some sites but not all? Although our study was not designed to test causality, we did measure several factors that could help to explain higher N cycling rates in invaded areas. We were interested in quantifying aboveground biomass production (stems and leaves), plus thatch, in invaded compared with uninvaded areas (e.g., Stanley et al. 2011). We were also interested in exploring whether A. elatius had higher aboveground biomass N content than that of grass tissues present in uninvaded areas, similar to invasive species in other grassland ecosystems (e.g., Rossiter-Rachor et al. 2009; James 2008). As demonstrated primarily through studies of N saturation in forested ecosystems (Aber et al. 1998), higher leaf and litter N content can result in accelerated N cycling processes. In addition, more thatch cover can alter soil moisture content by minimizing evaporative losses and holding more water locally (Liang et al. 2017), a particularly important control on tallgrass prairie species (Craine et al. 2010). Thatch cover can also limit rates of primary productivity in native tallgrass species by reducing photosynthetically active radiation to growing shoots and preventing cooling of emergent leaves (Knapp and Seastedt 1986).

Prior evidence that A. elatius produces more thatch than native prairie communities prompted an aboveground biomass harvest in September 2019. We measured approximately two-fold higher aboveground biomass at the grassland and shrubland sites in invaded compared with uninvaded areas. At the savanna, the difference between invaded and uninvaded areas was less dramatic, but the site exhibited the same pattern as the others, with invaded areas ~ 200 g m⁻² greater in aboveground biomass than uninvaded areas (Fig. 2a). Stanley et al. (2011) also observed higher thatch production by A. elatius and other exotic species compared with native plant species in the Pacific Northwest (US). Interestingly, patterns in soil moisture during autumn did not differ substantially
between invaded and uninvaded plots (Table 2, Fig. 1f). However, our snapshots of soil moisture during soil sampling are insufficient to determine its relationship with A. elatius; this question remains outstanding and requires further investigation.

We also considered the C:N ratios of aboveground biomass tissues (leaves, stems, and thatch), as an indication of whether A. elatius dominated vegetation is N-enriched relative to native prairie dominated vegetation. We measured a significant effect of site x invasion on aboveground biomass C:N ($p = 0.015$), however, only at the savanna did we find slightly lower C:N ratios in invaded compared with uninvaded areas. In all other cases, we observed the opposite pattern (Fig. 2b). Our results are challenging to interpret since our harvests in invaded areas were not 100% A. elatius; there were some non-native species present in uninvaded plots, and we sampled aboveground biomass after tissues had senesced. To better understand differences in aboveground biomass C:N ratios, it would be useful to compare tissue chemistry in live stems vs standing dead, and to analyze thatch separately.

In conclusion, we observed different relationships between A. elatius invasion and soil N cycling across sites in the Colorado foothills region. Similar complexity has been noted in studies evaluating the relationships between Cheatgrass (Bromus tectorum) invasion and nutrient cycling (Concilio et al. 2015). However, the differences observed in measured variables between invaded and uninvaded areas and among sites lead us to some important conclusions and logical next steps. First, the inconsistent directional difference between net N mineralization rates in invaded and uninvaded areas across sites suggests that while one might observe high-density A. elatius growth across the foothills of the Colorado Front Range, the relationship between soil N cycling and the invasive may be influenced by other factors that differ across locations. It is possible, for example, that rather than causing changes to soil N cycling, A. elatius exploits areas that have higher net N mineralization rates. A future study to evaluate causality of invasion and variation in soil N cycling rates would be useful. Second, understanding whether restoration objectives are feasible following widespread A. elatius invasion will be important. Many native grasses like Big Bluestem (Andropogon gerardii) are adapted to low-N soil environments (Averett et al. 2004). Thus, a logical next step is to determine whether native grass species of interest, such as A. gerardii, can persist in soils previously colonized by A. elatius. Such research will be important to inform how best to maintain or restore native plant community composition in tallgrass prairie ecosystems.

**Acknowledgements** The authors thank the University of Colorado-Boulder, Fall 2019 Environmental Studies Field Methods in Ecosystem Science students and M. Huber for assistance with sampling. J. Larson for assistance with plant community composition measurements, B. Bednarek and L.P. Hartwell for laboratory analyses, and the University of Colorado-Boulder Research and Innovation Office Faculty Fellows Program. We also thank three anonymous reviewers who helped to improve the quality of the manuscript.

**Author contribution statement** ESH, BA, and AL conceived of the ideas and designed methodology. HRM and ESH collected the data. BA, ESH, and HRM analyzed the data; ESH led the writing of the manuscript. All authors contributed critically to the drafts and gave final approval for publication.

**Funding** This research was funded by City of Boulder Open Space and Mountain Parks (Grant # 19–01-0047).

**Availability of data and material** Data are provided as a supplement to this paper.

**Code availability** The code used in this study is available from the corresponding author on reasonable request.

**Declarations**

**Conflict of interest** The authors declare no conflicts of interest.

**Ethics approval** Not required for this study.

**Consent to participate** Not required for this study.

**Consent for publication** Not required for this study.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article’s Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

**References**

Aber JD, Melillo JM, Nadelhoffer KJ, Pastor J, Boone RD (1991) Factors controlling nitrogen cycling and nitrogen saturation in northern temperate forest ecosystems. Ecol Appl 1(3):303–315

Aber JD, McDowell W, Nadelhoffer K, Magill A, Berntson G, Kamakea M, McNulty S, Currie W, Rustad L, Fernandez I (1998) Nitrogen saturation in temperate forest ecosystems: hypotheses revisited. Bioscience 48(11):921–934

Aguilera AG, Alpert P, Dukes JS, Harrington R (2010) Impacts of the invasive plant Fallopia japonica (Houtt) on plant communities and ecosystem processes. Biol Invasions 12(5):1243–1252
Asner GP, Hughes RF, Vitousek PM, Knapp DE, Kennedy-Bowdoin T, Boardman J, Martin RE, Eastwood M, Green RO (2008) Invasive plants transform the three-dimensional structure of rain forests. Proc Natl Acad Sci 105(11):4519–4523

Averett JM, Klips RA, Nave LE, Frey SD, Curtis PS (2004) Effects of soil carbon amendment on nitrogen availability and plant growth in an experimental tallgrass prairie restoration. Restor Ecol 12(4):568–574

Baron JS, Rueh HM, Wolfe AM, Nydick KR, Allsott EJ, Minear JT, Moraska B (2000) Ecosystem responses to nitrogen deposition in the Colorado Front Range. Ecosystems 3(4):352–368

Berendse F, Elberse WT, Geerts RHME (1992) Competition and nitrogen loss from plants in grassland ecosystems. Ecology 73(1):46–53

Binkley D, Aber J, Pastor J, Nadelhoffker K (1986) Nitrogen availability in some Wisconsin forests: comparisons of resin bags and on-site incubations. Biol Fertil Soils 2(2):77–82

Bobbinck R, Hicks K, Galloway J, Spranger T, Alkemade R, Ashmore M, Bustamante M, Cinderby S, Davidson E, Dentener F, Emmett B (2010) Global assessment of nitrogen deposition effects on terrestrial plant diversity: a synthesis. Ecol Appl 20(1):30–59

Chen Y, Wieder WR, Hermes AL, Hinckley E-LS (2020) The role of physical properties in controlling soil nitrogen cycling across a tundra-forest ecotone of the Colorado Rocky Mountains, USA. CATENA 186:104369

Clark CM, Tilman D (2008) Loss of plant species after chronic low-level nitrogen deposition to prairie grasslands. Nature 451:712–715

Compton JE, Boone RD, Motzkin G, Foster DR (1998) Soil carbon and nitrogen in a pine-oak sand plain in central Massachusetts: role of vegetation and land-use history. Oecologia 116(4):536–542

Concilio AL, Prevey JS, Omasta P, O’Connor J, Nippert JB, Seastedt TR (2015) Response of a mixed grass prairie to an extreme precipitation event. Ecosphere 6(10):1–12

Corbin JD, D’Antonio CM (2003) Too much of a good thing: Restoration, immobilization, and nitrification in an experimental tallgrass prairie restoration. Restor Ecol 11(2):119–125

Dukes JS, Mooney HA (1999) Does global change increase the success of biological invaders? Trends Ecol Evol 14(13):135–139

Ehrenfeld JG (2003) Effects of exotic plant invasions on soil nutrient cycling processes. Ecosystems 6(6):503–523

EnviroPlan Partners (2018) Tall oatgrass ecological study. City of Boulder open space and mountain parks report. 201 pp. (Online: https://www-static.boulder.col orado.gov/docs/tall-oatgrass-full-1-2018030911337.pdf)

Field CD, Duse NB, Payne RJ, Britton AJ, Emmett BA, Helliwell RC et al (2014) The role of nitrogen deposition in widespread plant community change across semi-natural habitats. Ecosystems 17(5):864–877

Hart SC, Stark JM, Davidson EA, Firestone MK (1994) Nitrogen mineralization, immobilization, and nitrification. Methods Soil Anal Part 2 Microbiol Biochem Prop 5:985–1018

Holub P, Tůma I, Záhora J, Fiala K (2012) Different nutrient use strategies of expansive grasses Calamagrostis epigejos and Arrhenatherum elatius. Biologia 67(4):673–680

James JF (2008) Leaf nitrogen productivity as a mechanism driving the success of invasive annual grasses under low and high nitrogen supply. J Arid Environ 72(10):1775–1784

Knapp AK, Seastedt TR (1986) Detritus accumulation limits productivity of tallgrass prairie. Bioscience 36(10):662–668

Kurokawa H, Pelzzer DA, Wardle DA (2010) Plant traits, leaf palatability and litter decomposability for co-occurring woody species differing in invasion status and nitrogen fixation ability. Funct Ecol 24(3):513–523

Lee MR, Bernhardt ES, van Bodegom PM, Cornelissen JHC, Kattge J, Laughlin DC, Wright JP (2017) Invasive species’ leaf traits and dissimilarity from natives shape their impact on nitrogen cycling: a meta-analysis. New Phytol 213(1):128–139

Liang X, Su D, Wang Z, Qiao X (2017) Effects of turfgrass thatch on water infiltration, surface runoff, and evaporation. J Water Resour Prot 9(7):799–810

Liao C, Peng R, Luo Y, Zhou X, Wu X, Fang C, Li B (2008) Altered ecosystem carbon and nitrogen cycles by plant invasion: a meta-analysis. New Phytol 177(3):706–714

Liu XR, Dong YS, Ren QJ, Li SG (2010) Drivers of soil net nitrogen mineralization in the temperate grasslands in Inner Mongolia, China. Nutr Cycl Agroecosyst 87(1):59–69

McCulley RL, Burke IC, Lauenroth WK (2009) Conservation of nitrogen increases with precipitation across a major grassland gradient in the Central Great Plains of North America. Oecologia 159(3):571–581

Owen JS, Wang MK, Sun HL, King HB, Wang CH, Chuang CF (2003) Comparison of soil nitrogen mineralization and nitrification in a mixed grassland and forested ecosystem in central Taiwan. Plant Soil 251(1):167–174

Parker SS, Schimel JP (2011) Soil nitrogen availability and transformations differ between the summer and the growing season in a California grassland. Appl Soil Ecol 48(2):185–192

Risser PG, Parton WJ (1982) Ecosystem analysis of the tallgrass prairie: nitrogen cycle. Ecology 63(5):1342–1351

Rossiter-Rachor NA, Setterfield SA, Douglas MM, Hutley LB, Cook GD, Schmidt S (2009) Invasive Andropogon gayanus (gamba grass) is an ecosystem transformer of nitrogen relations in Australian savanna. Ecol Appl 19(6):1546–1560

Shaben J, Myers JH (2010) Relationships between Scotch broom (Cytisus scoparius), soil nutrients, and plant diversity in the Garry oak savanna ecosystem. Plant Ecol 207:81–91

Stanley AG, Kaye TN, Dunwiddie PW (2011) Multiple treatment combinations and seed addition increase abundance and diversity of native plants in Pacific Northwest prairies. Ecol Restor 29(1–2):35–44

Wetherbee GA, Benedict KB, Murphy SF, Elliott EM (2019) Inorganic nitrogen wet deposition gradients in the Denver-Boulder metropolitan area and Colorado Front Range—Preliminary implications for Rocky Mountain National Park and interpolated deposition maps. Sci Total Environ 691:1027–1042

Xiang SR, Doyle A, Holden PA, Schimel JP (2008) Drying and rewetting effects on C and N mineralization and microbial activity in surface and subsurface California grassland soils. Soil Biol Biochem 40(9):2281–2289

Yahdjian L, Sala OE (2010) Size of precipitation pulses controls nitrogen transformation and losses in an arid Patagonian ecosystem. Ecosystems 13(4):575–585