Evaluation of 15 Functional Candidate Genes for Association with Chronic Otitis Media with Effusion and/or Recurrent Otitis Media (COME/ROM)

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Abstract

DNA sequence variants in genes involved in the innate immune response and secondary response to infection may confer susceptibility to chronic otitis media with effusion and/or recurrent otitis media (COME/ROM). We evaluated single nucleotide polymorphisms (SNPs) in 15 functional candidate genes. A total of 99 SNPs were successfully genotyped on the Sequenom platform in 142 families (618 subjects) from the Minnesota COME/ROM Family Study. Data were analyzed for association with COME/ROM using the Generalized Disequilibrium Test (GDT). Sex and age at exam were adjusted as covariates, relatedness was accounted for, and genotype differences from all phenotypically discordant relative pairs were utilized to measure the evidence of association between COME/ROM and each SNP. SNP rs2735733 in the region of the mucin 5, subtypes A/C gene (MUC5AC) exhibited nominal evidence for association with COME/ROM (P = 0.002). Two additional SNPs from this region had P values < 0.05. Other variants exhibiting associations with COME/ROM at P < 0.05 included the SCN1B SNP rs8100085 (P = 0.013), SFTP5 SNP rs1051246 (P = 0.039) and TLR4 SNP rs2770146 (P = 0.038). However, none of these associations replicated to an independent sample of COME/ROM families. The candidate gene variants examined do not appear to make a major contribution to COME/ROM susceptibility, despite a priori evidence from functional or animal model studies for a role in COME/ROM pathology.

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Introduction

Incidence and prevalence of otitis media (OM) has declined since 2000 when infant immunization with pneumococcal conjugate vaccine was adopted in the United States [1,2,3]. However, OM is still a very common childhood disease and remains a major cause of morbidity and hearing loss in young children [4,5]. Children with an affected sibling are at higher risk, and OM clusters in families across generations [6,7]. Twin studies in the United States, Norway, England and Wales have reported a high degree of heritability for recurrent and chronic OM [8,9,10], defined as more than 3 episodes of acute OM in a year, or middle ear fluid persisting for 3–4 months [11,12].

Our previous research on genetic contributions to chronic otitis media with effusion and/or recurrent otitis media (COME/ROM) include a complex segregation analysis of 173 families suggesting single gene effects [13], a genome-wide linkage scan in 121 families providing support for loci on chromosomes 10q and 19q [14], and an investigation of the human ortholog (FBXO11) of the deaf mouse mutant jff gene, a single gene model of chronic/recurrent OM in our family-based study [15].

We selected 15 candidate genes involved in the innate immune response or the ability to clear infectious agents implicated in OM, and evaluated common variation across the coding regions of these genes for association with COME/ROM.

Methods

Ethics statement

This study was conducted under Institutional Review Board approval at the University of Minnesota, Wake Forest University,
and University of Pittsburgh, and adhered to the tenets of the Declaration of Helsinki. Written consent was provided by all participants or, in the case of children, by their parent or legal guardian.

Subjects

Subjects who had tympanostomy tube surgery for COME/ROM (proband) and their families were recruited for the study, which has been described previously [14,15,16]. All probands were treated with tympanostomy tubes. Participating first and second degree relatives and parents of probands were classified as affected or unaffected based on 1) treatment with tympanostomy tubes, or 2) four data sources. Participants or parents completed a history form documenting history and risk factors for COME/ROM. The study otolaryngologist performed an ear examination to determine presence of OM sequelae without knowledge of the subject’s prior OM history, an audiologist conducted tympanometric testing at three frequencies (226, 630 or 710, and 1400) to detect abnormal middle ear mechanics, and medical records were abstracted to document tympanic membrane and middle ear findings consistent with COME/ROM, and duration of OME. Hearing was screened at 20 dB for speech frequencies. Phenotype information comes from four sources: self or parent reported OM history, ear examination, tympanometric findings, and medical record abstract (see Material S1). Affected status of probands’ relatives was based on having positive findings from at least two of these four sources (Table S1). A total of 142 families with at least two individuals with the diagnosis of COME/ROM were used in the candidate gene analyses.

We carried out a replication analysis in an independent study of otitis media [17] that consisted of 1,583 genotyped individuals from 441 Caucasian families. In order to assure a history of significant ear disease, two or more full siblings who both or all had undergone tympanostomy tube insertion were enrolled. The need for tympanostomy tube insertion established that the subject’s history of middle ear disease was truly significant, resulting in the need for a surgical procedure. A subject was only considered “affected” if he/she had undergone tympanostomy tube insertion at least once for recurrent/persistent OM, while a subject was considered “unaffected” if he/she had never had tympanostomy tubes and had no known history of recurrent/persistent OM. The remaining subjects were considered as having “unknown” disease status. Otoscopic examinations and tympanograms were conducted at entry if requested by the parent for patient information not study data since the condition of the ears at entry did not determine eligibility and the tubes may have been inserted many years prior to study entry.

SNP selection and genotyping

DNA was isolated from blood using the Gentra PureGene method (Qiagen, Valencia CA). Candidate genes were selected on the basis of a role in innate host defense and acute inflammatory response or chronic response and effusion, and prior evidence in the literature for involvement in OM. We selected the largest known isoform, added 5 kb upstream and downstream of the coding region, and used HapMap CEU (Centre d’Etude Polymorphisme Humain) data (HapMap Data Rel.4/phaseII Jul06, on NCBI B35 assembly, dbSNP b125) to identify SNPs with minor allele frequency (MAF)≥0.05. For mucin 5, subtypes A/C (MUC5AC), we used the combined largest mRNA sequence (AF015521; AF298317; AF043909), which encompasses MUC5AC NC_000011.8. We used the pairwise tagging option of Tagger [18] (t2>0.8) to select the minimal tagSNP set for each gene. Genotyping was conducted on the Sequenom platform using the iplex assay [19]. A total of 99 SNPs were successfully genotyped in >95% of samples (see Table 1 for gene summary). Concordance rates were calculated on the basis of 21 replicates included for genotyping on the Sequenom platform at the same time that all samples were assayed. Fifty-four SNPs were 100% concordant; 19 SNPs had one discordant call; 25 SNPs had two discordant calls; and 1 SNP (rs785898 near CD14) had three discordant calls. The concordance rates for nominally associated SNPs were 100% (rs1051246, rs7396030, rs2075859, rs8100085) or 95% (rs2735733, rs2710146; equivalent to one discordant call out of 21 or 20 replicate pairs respectively).

Candidate genes selected on the basis of their function in innate host defense and acute inflammatory response included Toll-like receptor 4 (TLR4) [20], Toll-like receptor 2 (TLR2) [21,22], Beta-defensin 1 (DEFB1) [23], Surfactant protein A1 (SFTPA1) [24,25,26], Surfactant protein D (SFTPD) [27,28], Interleukin-8 (IL8) [29,30,31], Interleukin-1 β (IL1B) [31], Interleukin 10 (IL10) [20], and Lactotransferrin (LTF) [32]. Genes selected for their role in chronic response and effusion were mucin 2 (MUC2) [33], mucin 5, subtypes A/C (MUC5AC) [34], Mannose-binding lectin precursor (MBL2) [35], CD14 antigen precursor (CD14) [36], Sodium channel, voltage-gated, type I-B (SCN1B) [37], and Ecotrophic viral integration site 1 (EVI1) [38].

Statistical analyses

Deviations from Hardy Weinberg equilibrium (HWE) in unaffected founder individuals were determined using the exact test [39]. Four SNPs with small HWE P values were excluded from the analysis and the remaining 99 SNPs have P>0.001. Data were checked for Mendelian errors using PedCheck [40] and pedstats [39] and poorly performing SNPs or a minimal dataset required to resolve errors were removed from analyses. Two monozygotic (MZ) twin pairs were detected and incorporated in the association analysis.

Data were analyzed for association with COME/ROM using the Generalized Disequilibrium Test (GDT) [41]. The GDT utilizes the genotype differences of all phenotypically discordant relative pairs in assessing association within families. This test has been shown to be consistently more powerful than PDT [42], FBAT [43] and several other family-based association tests for a common disease [41]. It allows modeling of covariates and IBD allele sharing, and information from extended pedigrees is efficiently used without breaking extended pedigrees into multiple nuclear families, as implemented in other methods. Compared to other association tests that also examine between-family association, the GDT method has the advantage of protecting from population stratification between families, and being less affected by genetic heterogeneity from multiple susceptibility genes. In GDT analyses, we modeled sex and age at exam (considered a proxy for generation/practical clinical practice) as covariates. Identity-by-descent (IBD) statistics were estimated using the Merlin package [44], and were incorporated in the GDT statistics.

Results

Population characteristics

The genotyped population included 618 individuals from 142 families (Table 2). One hundred thirty-two families were non-Hispanic and of European ancestry. The 10 non-European families consisted of seven families that described themselves as non-Hispanic and of mixed race, one mixed race Hispanic family, one Asian family, and one Native American family. As family-based tests of association using the GDT are robust to population stratification between families [41], all families were retained for analyses.
Associations with COME/ROM

A total of 99 SNPs encompassing 267 kb were successfully genotyped in subjects from the Minnesota COME/ROM Family Study. Age at exam was the factor most significantly associated with COME/ROM. The odds ratio of association between one year increment and COME/ROM was 0.94, with $P = 1.2 \times 10^{-16}$. This strong association suggests the importance of adjusting for the age effect in the candidate region association analysis. Although the sex effect is not statistically significant after adjusting for age, it was retained as a covariate in the candidate region association analyses.

SNP rs2735733 in the $MUC5AC$ gene region exhibited nominal evidence ($P = 0.002$) for association with COME/ROM. At this SNP, the T allele (minor allele frequency (MAF) 0.461) is more frequent in unaffected individuals than in affected individuals, with the allele frequency difference being 0.106. The odds ratio (OR) of the minor allele was estimated to be 0.646, assuming a population prevalence of 0.1. Although mucin 5B ($MUC5B$) was not initially selected for study as an *a priori* candidate, the genomic region genotyped, including the associated region of $MUC5AC$, overlaps with $MUC5B$. In the region of chromosome 11 encompassing $MUC5AC$ ($MUC5B$) and $MUC2$, there were 21 SNPs in total genotyped. Two other SNPs in this region had $P$ values < 0.05 ($P = 0.049$ at rs7396030 and $P = 0.041$ at rs2075859). The largest effect of a single SNP is OR = 1.57 at rs7396030. SNP rs7396030 is in weak LD with rs2075859 and rs2735733, with $r^2 = 0.036$ and 0.013 respectively ($D^2 = 0.293$ and 0.216 respectively); rs2075859 and rs2735733 are in LD, with $r^2 = 0.678$ (and $D^2 = 0.987$). Only founders were used in the LD calculation, conducted using PLINK [45]. Haplotype-based GDT analysis (as implemented in GDT [41]) identified association (although not significant) between haplotype CC at rs2075859 and rs2735733 and COME/ROM, with $P$ value 0.0097, which is consistent with the single SNP association results. Further haplotype analysis based on three SNPs does not produce any significant association (smallest $P = 0.43$).

We identified several other nominal SNP-COME/ROM associations, including the $SCN1B$ rs8100085 ($P = 0.015$), $SFTPD$ rs1051246 ($P = 0.039$), and $TLR4$ rs2770146 ($P = 0.038$; Table 3).

### Table 1. Selected candidate genes and number of SNPs genotyped.

| Candidate gene          | Gene symbol | Chromosome | Gene size* (kb) | Region genotyped (kb) | Total no. SNPs in region† | No. SNPs successfully genotyped | SNPs captured at $r^2$ > 0.8 | Mean $r^2$ |
|------------------------|-------------|------------|-----------------|------------------------|---------------------------|-------------------------------|----------------------------|------------|
| β-defensin 1           | DEFB1       | 8          | 7.4             | 13.0                   | 69                        | 10                            | 51                         | 74%        | 0.982     |
| CD14 antigen precursor | CD14        | 5          | 1.9             | 9.8                    | 9                         | 4                             | 9                          | 100%       | 0.963     |
| Ecotropic viral integration site 1 | EVI1 | 3          | 61.5            | 57.3                   | 73                        | 12                            | 73                         | 100%       | 0.978     |
| Interleukin 1-β        | IL1B        | 2          | 7.0             | 9.1                    | 10                        | 3                             | 8                          | 80%        | 0.952     |
| Interleukin 8          | IL8         | 4          | 3.2             | n/a                    | 4                         | 1                             | 4                          | 100%       | 0.983     |
| Interleukin 10         | IL10        | 1          | 4.9             | 7.8                    | 21                        | 5                             | 9                          | 43%        | 0.977     |
| Lactotransferrin       | LTF         | 3          | 28.9            | 36.3                   | 33                        | 7                             | 29                         | 88%        | 0.966     |
| Mannose-binding lectin precursor | MBL2 | 10         | 6.3             | 13.5                   | 46                        | 10                            | 46                         | 100%       | 0.965     |
| Mucin 2                | MUC2        | 11         | 29.5            | 35.3                   | 26                        | 13                            | 22                         | 85%        | 0.990     |
| Mucin 5, subtypes A/C  | MUC5AC      | 11         | 75.8            | 27.1                   | 18                        | 8                             | 13                         | 72%        | 0.968     |
| Sodium channel, voltage-gated, type I-β | SCN1B | 19         | 9.8             | n/a                    | 1                         | 1                             | 1                          | 100%       | 1.000     |
| Surfactant Protein A1  | SFTPA1      | 10         | 3.3             | n/a                    | 1                         | 1                             | 1                          | 100%       | 1.000     |
| Surfactant Protein D   | SFTPD       | 10         | 11.4            | 17.0                   | 40                        | 8                             | 12                         | 30%        | 1.000     |
| Toll-like receptor 2   | TLR2        | 4          | 21.8            | 23.1                   | 12                        | 8                             | 10                         | 83%        | 0.983     |
| Toll-like receptor 4   | TLR4        | 9          | 13.2            | 17.3                   | 30                        | 8                             | 21                         | 70%        | 0.951     |
| Total                  |             |            | 285.9           | 266.6                  | 393                       | 99                            | 309                        | 77%        | 0.916     |

* Largest isofrom.
† CEU population, minor allele frequency (MAF)>0.05, largest isofrom of gene plus 5 kb upstream and downstream, based on HapMap data rel #24 phase II/Nov 08, on NCBI B36 assembly, dbSNP B126.

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### Table 2. Participant characteristics.

| Trait                        | Value (affected, unaffected) |
|------------------------------|------------------------------|
| Number of families           | 142                          |
| Number of subjects           | 618                          |
| Female                       | 52.8%                        |
| Age, mean ± SD               | 27.1±16.6 (28.2±16.5, 25.4±16.5) |
| Affected with COME/ROM       | 61.5%                        |
| Caucasian                    | 95.3%                        |
| Non-Hispanic                 | 99.0%                        |
| Smokers in the home, mean ± SD | 0.74±0.85                    |
| Attending day care centers   | 52.4% (50.8%, 55.0%)         |
| Prior breastfeeding           | 57.7% (53.5%, 63.4%)         |
| Allergies                    | 30.7% (32.5%, 26.7%)         |

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We also carried out association analysis in families consisting of European American only (eight families were excluded) and observed the same associations. All six associated SNPs listed in Table 3 retain their significance with P values 0.027, 0.027, 0.045, 0.033, 0.002, and 0.022 respectively. An additional SNP (rs2672812) in the MUC5A/MUC5B region also had a P-value <0.05 (P = 0.042).

We carried out a replication analysis of the six SNPs in Table 3 in an independent study [17]. The association results in the replication study are shown in Table 4. The allele frequencies in two studies are comparable. Only one SNP in the MUC2 region, rs7396030, reached a P-value 0.033, 0.002, and 0.022 respectively. An additional SNP (rs2672812) in the MUC5A/MUC5B region was similar to that observed in the University of Minnesota sample, the LD structure across the MUC5AC/MUC5B genomic region, encompassing both MUC2 and MUC5AC/MUC5B.

Table 3. GDT association results (P<0.05) for COME/ROM.

| Chr | SNP    | Position | Allele | Frequency | P     | d     | OR    | Genes     |
|-----|--------|----------|--------|-----------|-------|-------|-------|-----------|
| 9   | rs2770146 | 117552892 | G      | 0.307     | 0.038 | −0.062 | 0.736 | TLR4      |
| 10  | rs1051246 | 81687798  | C      | 0.118     | 0.039 | −0.058 | 0.499 | SFTPD*    |
| 11  | rs7396030 | 1073364   | T      | 0.196     | 0.049 | 0.078  | 1.565 | MUC2      |
| 11  | rs2075859 | 1207064   | T      | 0.374     | 0.041 | −0.067 | 0.744 | MUC5AC,MUC5B |
| 11  | rs2735733 | 1218216   | T      | 0.461     | 0.002 | −0.106 | 0.646 | MUC5AC,MUC5B |
| 19  | rs8100085 | 40214959  | A      | 0.313     | 0.013 | −0.090 | 0.636 | SCN1B     |

Note: Analyses are adjusted for sex and age at exam. * is the allele frequency difference between affected and unaffected individuals. The odds ratio (OR) of association is converted from d and MAF, assuming a population prevalence of 0.1.

This SNP was selected due to its proximity to COME/ROM candidate gene SFTPD, but is located within the mannosol-binding protein-A pseudogene (MBL1P1) gene.

Weisberg PB, Rogers MM, Ingram RH, et al. (2010) Surfactant protein D (sftpd) knockout animals do not clear pathogens in otitis media, scn1b, muc2 and muc5 were among the genes upregulated [37], although polymorphisms of SCN1B do not appear to have been investigated in COME/ROM previously. Surfactant protein D (sftpd) knockout animals do not clear influenza A viral infections [53]. SFTP polymorphisms in humans have been associated with surfactant protein D assembly, function, and concentration [54], as well as severe respiratory

Table 4. GDT association results for SNPs in Table 3 genotyped in the Caucasian subjects from the replication sample.

| Chr | SNP    | Position | Allele | Frequency | P     | d     | OR    | Genes     |
|-----|--------|----------|--------|-----------|-------|-------|-------|-----------|
| 9   | rs2770146 | 117552892 | G      | 0.324     | 0.84  | 0.006 | −0.200 | TLR4      |
| 10  | rs1051246 | 81687798  | C      | 0.129     | 0.65  | −0.012 | −0.454 | SFTPD     |
| 11  | rs7396030 | 1073364   | C*     | 0.802     | 0.022 | 0.057 | 2.295 | MUC2      |
| 11  | rs2075859 | 1207064   | T      | 0.358     | 0.22  | −0.042 | −1.238 | MUC5AC,MUC5B |
| 11  | rs2735733 | 1218216   | T      | 0.447     | 0.11  | −0.052 | −1.617 | MUC5AC,MUC5B |
| 19  | rs8100085 | 40214959  | A      | 0.379     | 0.47  | 0.012 | 0.715 | SCN1B     |

Note: Analyses are adjusted for sex.

*Opposite to allele reported in Table 3.

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syncytial virus infections [27], a known precipitating factor for OM [55]. In a mouse model of spontaneous OM, an exon 3 missense mutation in Tlr4 resulted in lipid A/lipopolysaccharide (LPS) endotoxin insensitivity and an inability to clear Gram-negative bacteria such as Haemophilus influenzae [56], a bacterium frequently found in COME [57]. Similar defects in LPS response due to TLR4 mutations have been shown to exist in humans [58].

This study has several strengths and limitations. The primary strength is the family-based study design, permitting an analytical approach robust to population stratification. Our newly developed GDT approach also allowed us to take into account covariates age and sex. The study included a small sample of non-European families, consequently we are unable to draw any conclusions about whether the observed associations are specific to European American populations. Genotyping inaccuracy could potentially have led to false negative results however, as noted in the Methods, the concordance rates for the SNPs shown in Table 3 exceeded 95%. The candidate gene approach is necessarily limited by the need for evidence of a role in OM pathology, thus we are expanding our search for COME/ROM susceptibility alleles to a genome-wide association study.

Epidemiological studies suggest that six episodes of AOM by age six is not uncommon. From among individuals with medical record abstracts (65% of participants), only 8 subjects (7 probands and 1 sibling) had 6 episodes of OM by age 6, but did not have 3 episodes OM in any 12 month period. All 8 had tubes placement surgery and were also positive for a history of COME/ROM by either ear exam or tympanometry measures, so would have been classified as affected even without meeting more stringent ROM criteria. Although both studies used similar phenotype criteria to determine affected status based on reported history of COME/ROM, medical record review, ear examination and tympanograms, for the primary study, the sibling of the proband did not need to have undergone tympanostomy tube insertion. In contrast, in the replication sample, both siblings were required to have tympanostomy tube insertion. Failure to replicate nominal associations may be a consequence of diagnostic differences between samples, although it is probable that initial associations represent false positive results. Although we cannot rule out the possibility of type 2 error due to lack of power in the initial sample, we can conclude that there are no strong effects on COME/ROM risk by common variants genotyped in these genes.

Since we define COME as middle ear fluid that lasts 3 or more months, and ROM as 3 OM episodes in a year, or 6 episodes by age 6, there is considerable overlap in our sample between COME and ROM. It is not unreasonable to hypothesize that inadequate production of mucins may predispose to ROM whereas excessive production of mucins may predispose to COME, or that different loci contribute to one or the other. Often patients with middle ear fluid are prone to acute OM, so COME may predispose to ROM. In the UMN sample there were 143 participants with a history of both COME and ROM. Disentangling this relationship in future analyses would require a larger sample.

Our results suggest that common variants in the selected candidate genes do not appear to make a major contribution to COME/ROM susceptibility, despite strong a priori hypotheses for a role of these genes in disease pathology, suggesting agnostic genome-wide analyses of COME/ROM populations may be a more productive approach to identify susceptibility alleles.

Supporting Information

Table S1 Criteria for Classifying Family Members as Affected with COME/ROM for the University of Minnesota Study. (DOCX)

Material S1 Criteria for Classifying Family Members as Affected with COME/ROM for the University of Minnesota Study. (DOCX)

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Author Contributions

Conceived and designed the experiments: MMS FS SSR KAD. Performed the experiments: MMS REF. Analyzed the data: W-MC XH DEW MM. Contributed reagents/materials/analysis tools: JCM XH MCL EMM KAD. Wrote the paper: MMS W-MC DEW SSR KAD.

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