Interactive Electric Fields Attract KELEA (Kinetic Energy Limiting Electrostatic Attraction) and Can Lead to the Activation of Water

Abstract

Water can acquire a kinetic activity by absorbing an environmental force termed KELEA (kinetic energy limiting electrostatic attraction). This activity leads to increased volatility of the water that can be measured as the progressive weight loss in closed, but not completely sealed containers. A fundamental role of KELEA may be to prevent the fusion and possible annihilation of electrostatically attracted opposite electrical charges. As such, it may become concentrated in situations in which there is convergence of force fields of opposing electrical charges. At least conceptually, this may arise with a bidirectional electrical current, especially if conducted through coiled electrical wires. This paper reports on preliminary studies based on this premise. Although not proving the premise, the reported experiments do indicate a simple method for activating water. Moreover, the described procedure should allow for further exploration of the underlying mechanism of water activation. The procedure involves the use of electrically conductive dual wire in the form of a coil, through which a direct current (DC) is transmitted. Water samples placed close to the coil, but insulated from the heat of the coil, showed increased volatility, which persisted well beyond the period of exposure to the electrical field. The paper further provides confirmation of previously reported data showing that activated water can indirectly lead to the activation of nearby water. The reported observations are of both practical and theoretical importance.

Keywords: ACE; Alternative cellular energy; KELEA; Water; Hydrogen bonding; Electricity; Direct current

Abbreviations: ACE: Alternative Cellular Energy; DC: Direct Current; ICE: Insufficiency of Cellular Energy; Av: Average; KELEA: Kinetic Energy Limiting Electrostatic Attraction; gm: Gram; mg: Milligram; ml: Milliliter; “-” inches

Introduction

Cells require energy to perform their various functions and many illnesses can be attributed to an insufficiency of cellular energy (ICE). It is generally assumed that apart from photosynthesis in plants and in certain bacteria, that cellular energy is obtained solely through food metabolism [1]. This view has been challenged with the description of the third or alternative cellular energy (ACE) pathway [2]. It has been proposed that the ACE pathway is expressed as a dynamic (kinetic) quality of the body’s fluids resulting from the absorption of an external force termed KELEA (kinetic energy limiting electrostatic attraction) [2]. The fundamental role of this force may be to prevent the fusion and possible annihilation of opposite electrical charges when they are being increasing electrostatically attracted to one another [2]. KELEA may also explain the repulsion of like electrical charges. Experimental studies indicate that KELEA can also reduce the strength of intermolecular hydrogen bonding in water [3], enabling the water molecules to better support metabolic and other cellular functions [4]. KELEA activation of water and other fluids can lead to changes in various physical properties. Among these changes are lower surface tension, reduced specific heat and increased volatility [5]. Repeated measurements of the weight of closed but not completely sealed water containing vials, provide a reliable method of comparing the volatility of different water samples [3]. This method was used in the present study.

Electrical current comprises the movements of electrons that create moving electrical and magnetic field changes in the surrounding environment [6-8]. The interaction of oppositely moving electrical fields created by bidirectional current is likely to attract KELEA. In other words, the forced proximity of bidirectional electrical fields from direct current (DC) passage through wires may provide a concentrated source KELEA, which could potentially lead to the activation of nearby fluids. This effect would, presumably, be greater if the conducting electrical wires were in the form of a compact coil. This reasoning, strengthened by preliminary data, led to the following experiment.

Materials and Methods

Water in Glass Vials: Before beginning the experiment, all residual vials of activated water from previous experiments were removed from the room containing the weighing balance and from a non-adjacent room in which the electrical current was to be generated. Thirty-one previously unused 1.25 oz glass vials with screw caps were purchased from The Container Store, Pasadena, CA. The vials were brought into the weighing room, numbered and individually weighed using a Sartorius Balance that reads to 0.1 mg. A gallon of distilled water (Arrowhead) was purchased
from a local store on the day of the experiment. Using a measuring cup, each glass vial was near completely filled with water and capped tightly by hand. The water filled vials were let stand for 5 hours before being individually weighed. The numbered vials were reweighed 12 hours later. Ten vials were selected at random without regard to any of the earlier measurements. Five of the vials were for the study described in this paper (test vials) and five vials were used for a study involving converging and opposing light emissions. The results of the light study are reported elsewhere [9]. The remaining 21 water containing vials provided the control vials for both experiments.

**Presentation of data**

The cumulative change in weight from time zero was measured in mg for all of the available control and test vials throughout the experiment. Particular emphasis was placed on the initial 48 hours, which included the 5 hours of exposure to the electrical current. For ease of presentation and because of the relatively little variability among the control vials, the average value, along with the highest and lowest readings of the control vials at each of the time points are shown in the Results. For the 5 test vials the average and actual cumulative weight change values for each of the 5 vials are shown for each time point.

**Results**

The empty vials with caps weighed an average of 28.02 grams (gm) prior to being filled with water (range 27.88 to 28.24). With added water, the tightly capped vials weighed an average of 63.590 gm, with the measured amounts of added water ranging from 34.56 – 37.09 ml. Upon re-weighing the individual vials 12 hours later; the average weight of the capped vials was essentially unchanged (63.590 gm). Matched comparisons of the weights of each of the 31 individual vials at the two time points showed slight variability of from minus 1.1 mg to plus 0.7 mg. This degree of occasional minor variation was attributed to not quite waiting long enough for the balance readings to fully stabilize. The finding of no appreciable weight change in the closed vials containing untreated water was anticipated since it was consistent with prior observations in other experiments. The decision was made to proceed with the electrical field experiment using 5 randomly selected test vials. Another 5 vials were assigned to a different experiment, the results of which are to be presented elsewhere [9]. Twenty-one vials were left as the control vials for both experiments.

**Table 1** includes the data acquired from the 21 control vials and from the 5 test vials during the first 48-hour period, which includes the initial 5-hour exposure of the test vials to the interactive electrical fields. After 2 hours of exposure to the electrical fields, an appreciable loss of weight had occurred in each of the 5 test vials (average 2.7 mg). There was no weight loss in the control vials, which actually showed a slight increased (negative loss) in their measured weights, averaging –0.5 mg. The weight loss in the test vials progressively increased over the later time points. The very small measurable negative weight losses in the control vials were recorded with most of the control vials up to the 8th hour. At hour 10 and, thereafter, a progressive reduction in the average weight of the control vials, compared to the weight at time zero, was observed. The average levels of weight losses in the control vials at 12, 24 and 48 hours (1.5 mg, 2.8 mg and 4.8 mg respectively) were far less than those in the test vials (20.1, 28.9 and 40.0 mg respectively).

The vials were periodically weighed at least weekly for the initially planned 840 hours (35 days) duration of the experiment. The results showed a continuing loss of weight, much more in the test than in the control vials. This is shown by comparing the weight loss measurements at 84 hours with both the weights recorded at 48 hours and with those recorded at 840 hours. It is also shown by comparing the 840 hour measurements with those obtained 24 hours earlier (816 hours). In is noted in this last
comparison that test vial number 5 lost significantly less weight (2.4 mg), than the remaining 4 test vials (7.9, 9.1, 9.5 and 9.0 mg respectively). The final 24-hour weight loss of test vial number 5 was, in fact, somewhat less than the 24-hour weight loss of the control vials, which averaged 3.1 mg.

The experiment was extended to 62 days (one day short of 9 weeks). The average daily weight losses of the 5 test vials from day 35 to day 62 were 7.9, 9.4, 8.1, 8.7 and 3.2 mg respectively. The first 4 of these values exceeded the average daily weight loss of the control vials over the same 27-day period of 3.1 mg with a range of average daily losses within individual control vials ranging from 2.1mg to 4.9 mg.

Table 1: Activation of Test Water Samples Using Coiled Electric Cord Conducting Direct Current (DC).

| Control vs. Test Vials | Cumulative Weight Loss (Mg) of Vials From the Time in Hours after Starting the 5-Hour Exposure Electric Fields |
|------------------------|---------------------------------------------------------------------------------------------------------------|
|                        | 1     | 2     | 3     | 4     | 5     | 6     | 8     | 10    | 12    | 24    | 36    | 48    | 84\(^{\text{a}}\) | 816   | 840   |
| Control Ave\(^{*}\)    | -0.5\(^{\circ}\) | -0.5  | -0.5  | -0.6  | -0.7  | -0.5  | -0.3  | 1.4   | 1.5   | 2.8   | 3.2   | 4.8   | 7.5   | 120.0 | 123.1 |
| Lowest                 | -0.2  | -1.1  | -1.2  | -1.3  | -1.4  | -1.5  | -0.1  | -0.1  | 0     | -0.3  | 1.3   | 1.3   | 77.1  | 79.6  |
| Highest                | -0.9  | 1.2   | 1.5   | 2.1   | 2.1   | 3.1   | 3.6   | 6.1   | 6.4   | 8.9   | 10.4  | 12.9  | 15.8  | 181.1 | 185.4 |
| Test Average           | 0.8   | 2.7   | 4.2   | 6.7   | 8.2   | 10.1  | 13.6  | 18.1  | 20.1  | 28.9  | 35.8  | 40.0  | 51.4  | 329.9 | 337.5 |
| Test Vial 1            | 1.3   | 2.6   | 4.1   | 6.5   | 7.7   | 9.5   | 12.8  | 17.2  | 18.8  | 25.4  | 28.9  | 31.8  | 41.1  | 315.2 | 323.1 |
| Test Vial 2            | 0.7   | 2.9   | 4.4   | 6.7   | 8.3   | 9.9   | 13.5  | 17.5  | 19.6  | 26.1  | 31.4  | 35    | 50.1  | 353.9 | 363.0 |
| Test Vial 3            | 0.9   | 2.7   | 4.5   | 7.3   | 8.7   | 10.8  | 14.5  | 19.2  | 21.4  | 28.7  | 35.0  | 38.7  | 48.1  | 374.0 | 383.5 |
| Test Vial 4            | 0.8   | 3.3   | 5.0   | 7.6   | 9.8   | 11.7  | 15.9  | 20.4  | 22.3  | 29.3  | 34.5  | 37.9  | 50.8  | 385.9 | 394.9 |
| Test Vial 5            | 0.4   | 1.8   | 3.2   | 5.3   | 6.7   | 8.4   | 11.5  | 16.0  | 18.6  | 34.8  | 49.3  | 56.4  | 66.8  | 220.5 | 222.9 |

\(^{*}\)Average weight of the control vials.

\(#\) The negative values indicate that the measured weights of some of the control vials exceeded the weights recorded at time zero.

\(^{\text{a}}\)Seventeen control vials were available at this time, since 4 had been used in a side study.

**Discussion**

The three basic findings of this paper are:

i) That capped vials of distilled water placed in the vicinity of interactive electric fields generated by DC flowing through a two-wired coiled electrical cord will lose weight in comparison to control vials of the same water not exposed to the electrical fields.

ii) That the weight loss of the test vials, which is viewed as a measure of kinetic activation of the water, continues well beyond the time that the vials were exposed to the interactive electric fields.

iii) That although at a slower rate than for the test vials, the control vials began to show a continuing and persisting reduction in weight beginning several hours after the experiment was begun.

The interpretation of this third finding is that the weight losses in the control vials reflect a low level of indirect water activation from maintaining vials of control and activated water near one another. The placing of all of the vials adjacent to the weighing balance helped in facilitating the weighing process. It is clear that there was no significant weight loss in any of the vials for the 12 hours that preceded the time zero beginning of the test procedures; a finding consistent with many similar prior observations. It is also apparent that shortly after terminating the 5-hour exposure of the test vials to the interactive electrical fields that the control vials began to show discernable weight loss. Although the results are not included in this paper, a parallel energy exposure study was performed on 5 other vials of water. Essentially, the water in these vials was activated by exposure to fluctuating, interactive light sources during the first and the third 12-hours’ periods of the experiment. Except for these two 12-hour periods, these vials along with the 5 test vials from the present experiment were stored within 1-2 feet of the 21 control vials. The delay in onset of weight loss of the control vials is consistent with the transfer of KELEA from activated water to nearby water [4].

The ability of activated water to indirectly activate other water without the need for direct contact formed the basis of Johann Grander’s water activating device (www.grander.com). This approach is also being employed in a Los Angeles bottled...
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Since the human body can be considered as a container of the body’s fluids, this consideration can also extend to the proposed beneficial effects of certain electrical fields on human health. Indeed, devices such as the Beam Ray of Royal Raymond Rife [16] and the Multi-wave oscillator of Georges Lakhovsky [17] may act by establishing water activating electrical fields reactive with the body’s water.

This reasoning is also consistent with the suggested role of the fluctuating electrical activity of the brain, which may act as an antenna for attracting KELEA for transfer to the body’s water [18]. The working hypothesis is that for some individuals, consuming KELEA activated water or being exposed to KELEA generating devices, such as certain types of electrical fields, may enhance the KELEA absorbing activity of the brain. Conversely, certain other types of electrical stimuli may be inhibitory to the brain’s KELEA antenna function. The system described in this paper should help facilitate studies on these intriguing topics.

Conclusion

DC transmitted through two-wired coiled electrical cord can increase the volatility of nearby water, as assessed by measuring weight loss of water in closed, but not completely sealed glass vials. The increased volatility is attributed to a loosening of the intermolecular hydrogen bonding between water molecules due to the absorption of an environmental force termed KELEA (kinetic energy limiting electrostatic attraction). KELEA is presumably attracted to the transmission of DC, possibly as a result of opposite electrical charges moving into closer proximity to each other. This is consistent with KELEA being a basic mechanism to ensure the lack of fusion and possible annihilation of the electrostatically attracted, opposite electrical charges. Once water is activated, it can attract additional KELEA in a manner that can lead to its further activation and to that of nearby water. The use of interactive electrical fields will likely find value in some of the many health, agriculture and industrial applications of KELEA activated water [5,14,15].

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