Association of anti-oxidative capacity of HDL with subclinical atherosclerosis in subjects with and without non-alcoholic fatty liver disease

Sara Karami¹, Hossein Poustchi², Negar Sarmadi¹, Amir Reza Radmard³, Fatemeh Ali Yari¹, Abbas Pakdel¹,⁴*† and Parisa Shabani⁵*†

Abstract

Background: Non-alcoholic fatty liver disease (NAFLD) patients are at a substantial risk for developing cardiovascular disease (CVD). High-density lipoprotein (HDL) is well known to have protective effects against the development of atherosclerotic CVD. One of the major antiatherogenic effects of HDL is its anti-oxidative function.

Objectives: This study investigated the association of anti-oxidative capacity of HDL with subclinical atherosclerosis in NAFLD and non-NAFLD subjects.

Methods: A total of 143 subjects including 51 NAFLD and 92 control subjects were included in this case–control study. HDL oxidative index (HOI) was determined spectrophotometrically using a cell-free method in the presence of a fluorescent substrate dichlorofluorescein diacetate (DCFDA). Paraoxonase 1 (PON1) activity, superoxide dismutase (SOD) activity, and malondialdehyde (MDA) plasma levels were assessed in both groups.

Results: The NAFLD patients with impaired HDL anti-oxidative function (HOI ≥ 1) had higher MDA levels, aspartate amino transferase (AST), liver stiffness (LS), and carotid intima-media thickness (cIMT) values compared to the controls. HDL oxidative index (HOI) was positively correlated with MDA levels and cIMT and negatively correlated with SOD activity.

Conclusions: Higher circulating levels of MDA were associated with the impaired anti-oxidative function of HDL in NAFLD. The impaired anti-oxidative capacity of HDL might be related to NAFLD severity and subclinical atherosclerosis in NAFLD patients.

Keywords: HDL oxidative index, Non-alcoholic fatty liver disease, Cardiovascular disease, Carotid intima-media thickness

Introduction

Non-Alcoholic Fatty Liver Disease (NAFLD) is the most leading cause of chronic liver disease which affects an estimated 20–30% of the general population [1]. NAFLD is strongly associated with other metabolic conditions such as obesity, type 2 diabetes mellitus, and cardiovascular diseases [1]. Despite liver-related morbidity and mortality, cardiovascular disease (CVD) is the most...
important cause of mortality in the NAFLD population [2].

NAFLD patients exhibit atherogenic dyslipidemia with low levels of high-density lipoprotein cholesterol (HDL-C) [3]. Several studies demonstrated an association of higher plasma high-density lipoprotein (HDL) cholesterol levels with a lower risk of atherosclerotic cardiovascular disease (CVD) morbidity and mortality. However, recent studies suggest that HDL particle functionality might be a better predictor of CDV risk than HDL cholesterol mass levels [4]. HDL particle has numerous antiatherogenic functions. One of the important antiatherogenic properties of HDL is its anti-oxidative function, the ability to suppress LDL oxidation which in turn decreases the atherogenicity of LDL particles [5].

NAFLD presence was shown to be associated with the size and functional heterogeneity of HDL particles [6, 7]. NAFLD patients had an abnormal distribution of HDL subpopulations and the lipid composition of HDL particles [7]. Proteomics analysis showed altered protein composition of HDL particles in the NAFLD patients [8]. HDL size differed by fatty liver index and NAFLD patients had lower levels of larger HDL particles [6]. Association of HDL ApoA1 with liver fat content was only found in the HDL lacked apoC3 [9]. This heterogeneity results in the alteration of different HDL functionalities such as anti-oxidative function. Several studies addressed the anti-oxidative function of HDL in CVD and other disorders with high cardiovascular risk [10–13]. Moreover, previous studies showed impaired HDL cholesterol efflux capacity in NAFLD [14, 15]. However, no study evaluated the anti-oxidative function of HDL in NAFLD patients.

Oxidative stress which occurs because of the imbalance between antioxidants and reactive oxygen species (ROS) is involved in the initiation and promotion of NAFLD [16, 17]. One of the mechanisms which cause HDL dysfunction is a modification with lipid peroxidation products such as malondialdehyde (MDA) [18, 19]. It has been shown that MDA impaired HDL’s athero-protective functions. MDA generates lysine adducts on apoA-1 and blocks the cholesterol efflux function of HDL [20]. Incubation of macrophages with MDA modified HDL also led to an increased ability to generate ROS [21]. The antioxidant enzymes including superoxide dismutase and PON1 diminish ROS and lipid peroxidation. Paraoxonase 1 (PON1) associates with HDL and catalyzes the hydrolysis of lipid peroxides [22].

Increasing data have demonstrated that NAFLD patients have abnormal circulating markers of oxidative stress, such as increased malondialdehyde (MDA), superoxide dismutase activity, and PON1 activity [23, 24]. Having a disturbed anti-oxidative system along with a high risk of CVD in these patients suggests that impaired antioxidant activities of HDL, as an important player in CVD, may be the potential link between these phenomena. In this study, we aimed to investigate differences in HDL antioxidative capacity between NAFLD patients and controls. Moreover, we sought to compare carotid intima-media thickness as well as other relevant biomarkers between NAFLD patients and controls with preserved and reduced HDL antioxidative capacity.

Material and methods

Study participants

In the current study, 92 Controls and 51 NAFLD patients were selected from Golestan Cohort Study [25]. The study was a case–control study and approved by the medical ethics committee of the Semnan University of Medical Sciences (Ethics Code: IR.SEMUMS.REC.1397.331) and all participants gave written informed consent. All participants were male, aged 50–81 years old. The diagnosis of NAFLD was based on abdominal ultrasonography. Subjects were excluded if they met one of the following criteria: a history of alcohol consumption (> 30 g/d), diabetes, viral hepatitis, autoimmune liver disease, hemochromatosis, Wilson’s disease. None of the patients were taking medication that has been reported to induce hepatitis, statins and antioxidants.

Ultrasonography and elastography

Ultrasound assessment was performed using an Accuvix XQ ultrasound unit (Medison, South Korea) equipped with a 3–7 MHz curved array and a 5–12 MHz linear array transducer for the evaluation of liver, abdominal fat, and carotid arteries as previously described [25]. Visceral Adipose Tissue thickness (VAT), carotid intima-media thickness (cIMT), and liver stiffness (LS) were measured as described before [25].

Anthropometric and laboratory evaluations

Anthropometric parameters were measured under standardized protocols. Fasting blood samples were obtained from the participants following overnight fasting. Serum and plasma were isolated and frozen in aliquots at − 80 °C for further analysis. Biochemical parameters including Fasting blood glucose (FBG), serum total cholesterol (TC), triglycerides (TG), high-density lipoprotein cholesterol (HDL-C), low-density lipoprotein cholesterol (LDL-C), and levels of alanine amino transferase (ALT), aspartate amino transferase (AST), gamma glutamyl transferase (GGT) were measured by automated enzymatic methods using commercial kits (Pars Azmoon, Iran).

Paraoxonase 1 (PON1) enzymatic activity was measured according to a method described before. Paraoxon
Measurement of high-density oxidative index (HOI)
HOI measures the ability of apoB-depleted plasma to inhibit LDL oxidation in the presence of dichlorofluorescein diacetate (DCFDA) (Sigma-Aldrich, Germany). A cell-free assay was performed as previously described with some modifications [5]. Before analysis, apolipoprotein (apo) B-depleted plasma was prepared as previously described [5]. LDL was diluted in PBS to a final cholesterol concentration of 100 μg/mL and oxidized by CuSO4 (100 μmol/L) for 6 h at 37 °C. Oxidized LDL with a concentration of 1.4 μg/mL, DCFDA with a concentration of 0.725 μg/mL and apo B-depleted plasma from the participants with PBS to a final volume of 175 μL were added and incubated at 37 °C for 60 min. Fluorescence intensity was measured with an excitation wavelength of 485 nm and emission wavelength of 530 nm using the synergy h1 hybrid multi-mode microplate reader (BioTek, USA). The HDL oxidative index (HOI) was calculated as the ratio of fluorescence intensity in the presence of apo-B-depleted plasma samples divided by the fluorescence intensity in the absence of apo-B-depleted plasma. An HOI < 1 was considered as anti-oxidative HDL function and an HOI ≥ 1 was considered as pro-oxidative HDL.

Statistical analysis
The Kolmogorov-Smirnov test was used to test for the normal distribution of data. Categorical data are presented as percentages, continuous data as the mean ± SD or median (IQR) as appropriate. Comparison between two groups was performed using the independent-sample t-test or Mann-Whitney U test for continuous data and the chi-squared test for categorical data. Differences among the four subgroups as classified based on the presence of NAFLD and HOI were determined using ANOVA followed by Tukey’s post hoc tests. Pearson correlation was applied to determine the correlations between HOI and the other parameters in the whole study population. Non-normally distributed data were log-transformed before ANOVA and Pearson correlation tests. Significant differences were defined by P < 0.05 (*) or (#), P < 0.01 (**) or (##), and P < 0.0001 (****) or (####). All of the statistical analyses were performed using the IBM SPSS Statistic 27 and GraphPad prism 9.

Results
Basic clinical and laboratory characteristics of the study groups are presented in Table 1. BMI, waist circumference (WC), waist-to-hip ratio, fasting blood glucose (FBG), TG, visceral fat, systolic blood pressure, diastolic blood pressure, AST, ALT, GGT, and LS were significantly higher in the NAFLD group compared with the control group. NAFLD patients had lower HDL-C than the control subjects. There was not any significant difference in age, total cholesterol (TC), LDL-C, and cIMT between the two groups.

There was not any significant difference in the mean of HOI between NAFLD and control groups (P = 0.405). We found that 61 (66.30%) controls and 30 (58.82%) patients had a preserved anti-oxidative function with an HOI < 1, and 31 (33.69%) controls and 21 (41.17%) patients presented pro-oxidative HDL serum measurements with an HOI ≥ 1 (P = 0.468). We stratified the subjects into two groups, HOI < 1 and HOI ≥ 1. The subjects with HOI ≥ 1 had higher levels of MDA (P = 0.042), lower PON1 activity (P = 0.027), lower SOD activity (P = 0.010), and higher cIMT values (P = 0.022) (Additional file 1: Figure S1).

We further stratified the study population into 4 groups according to combined strata of status and HOI above and below 1 and compared MDA, antioxidant enzymes, NAFLD markers, and cIMT (Additional file 1: Table S1). The variables that were significantly different between HOI < 1 and HOI ≥ 1 subgroups of control or NAFLD groups were presented in Fig. 1. Interestingly, NAFLD patients with HOI ≥ 1 had higher MDA (P = 0.043), LS (P = 0.008), AST (P = 0.028) and cIMT (P = 0.030) than the NAFLD patients with HOI < 1.

Correlation analysis demonstrated that HDL anti-oxidant capacity (HOI) correlated positively with plasma MDA levels (r = 0.298, P = 0.001) and cIMT (r = 0.197, P = 0.025) in the whole population. There was a significant negative correlation between HOI and the SOD activity (r = −0.242, P = 0.004) in the whole population (Table 2).

Discussion
Cardiovascular diseases are the main cause of death in NAFLD patients [27]. The impaired anti-oxidative capacity of HDL turned out to be a good predictor of cardiovascular disorders [5]. Our findings showed that impaired anti-oxidative capacity of plasma HDL was associated with higher levels of MDA, hepatic fibrosis markers, and cIMT in NAFLD.
A previous study on the anti-oxidative properties of HDL in patients with coronary syndrome found higher HOI in patients with acute coronary syndrome or stable coronary artery disease compared with controls [11]. Another study reported total HDL antioxidant capacity in systemic lupus erythematosus patients who have an increased risk of cardiovascular diseases were significantly reduced compared to the controls [28]. But there was no significant difference in anti-oxidative properties of HDL in MI patients with or without ST elevation and non-MI participants [10]. We also did not find a significant difference in HOI values between the NAFLD and the control groups. The frequency of HOI $\geq 1$ was higher in NAFLD compared to the controls, albeit not significant. Consistently, it has been reported that impaired anti-oxidative capacity of plasma HDL was more frequent in women with polycystic ovary syndrome (PCOS) compared with the control group [13]. These conflicting results might be due to the different study populations and methods used in the studies.

In the current study, we found a higher level of MDA in the NAFLD group compared to the control group. In parallel, previous studies reported increased levels of MDA in the NAFLD patients compared to controls [23, 24, 29]. Additionally, a study showed increased levels of MDA in HDL subfractions isolated from the plasma of acute coronary syndrome patients compared to control subjects and it was along with a pro-inflammatory effect of HDL [30]. Consistently, we found higher levels of plasma MDA in the NAFLD patients with pro-oxidative HDL (HOI $\geq 1$) compared to the patients with HOI < 1. Our findings also showed a significant positive correlation between HOI values and MDA levels. Moreover, our results showed higher levels of PON1 paraoxonase activities in the NAFLD patients than in the controls. Conversely, other studies showed lower

| Parameter                  | Control (n = 92) | NAFLD (n = 51) | P value |
|----------------------------|-----------------|---------------|---------|
| Age (years)                | 59.0 (55.0–66.0)| 56.0 (54.0–61.0)| 0.273   |
| BMI (kg/m²)                | 24.8 ± 3.6      | 29.3 ± 3.4     | <0.001  |
| WC (cm)                    | 93.2 ± 11.1     | 103.6 ± 9.6    | <0.001  |
| WHR                        | 0.97 (0.92–1.00)| 0.98 (0.94–1.03)| 0.002   |
| Weight (kg)                | 68.7 (61.0–79.5)| 80.7 (73.5–89.0)| <0.001  |
| TC (mg/dL)                 | 205.5 ± 39.4    | 207.3 ± 37.0   | 0.779    |
| LDL-C (mg/dL)              | 121.6 ± 33.7    | 124.6 ± 28.7   | 0.594    |
| HDL-C (mg/dL)              | 57.4 ± 12.5     | 51.9 ± 11.1    | 0.007    |
| TG (mg/dL)                 | 112.0 (82.0–158.0)| 148.5 (112.5–186.0)| 0.016   |
| Systolic blood pressure (mmHg)| 131.5 ± 21.3   | 144.9 ± 23.7   | 0.001    |
| Diastolic blood pressure (mmHg)| 79.9 ± 11.0   | 86.8 ± 12.4    | 0.001    |
| FGF (mg/dL)                | 92.71 ± 9.00    | 99.57 ± 11.2   | <0.001   |
| Visceral fat (%)           | 46.5 ± 20.7     | 70.2 ± 18.0    | <0.001   |
| AST (U/L)                  | 18.5 (15.0–22.0)| 22.0 (19.5–27.0)| <0.001   |
| ALT (U/L)                  | 15.5 (11.0–22.0)| 31.5 (23.0–41.0)| <0.001   |
| GGT (U/L)                  | 22.37 (17.40–29.10)| 28.65 (23.53–38.50)| <0.001   |
| LS (kPa)                   | 3.8 (3.3–4.3)   | 5.0 (4.2–6.6)  | <0.001   |
| cIMT (mm)                  | 0.8 (0.75–0.87) | 0.81 (0.74–0.91)| 0.863    |
| HOI                        | 0.92 ± 0.19     | 0.95 ± 0.24    | 0.405    |
| HOI ≥ 1 frequency (%)      | 33.69           | 41.17          | 0.468    |
| SOD (U/mg protein)         | 10.7 ± 1.5      | 8.6 ± 2.4      | <0.001   |
| MDA (µM)                   | 33.2 (25.0–42.3)| 34.0 (30.4–47.7)| 0.771    |
| PON1 (U/L)                 | 24.4 (17.7–31.1)| 42.2 (27.7–52.2)| <0.001   |
| Hypertension (%)           | 27.90           | 47.36          | 0.021    |

Comparisons between groups were performed using Independent Student’s t test or Mann–Whitney U test as appropriate. Continuous data are expressed as mean ± standard deviation (SD) or median and (interquartile range), categorical data as percentages.

BMI: body mass index; WC: waist circumference; WHR: waist to hip ratio; TC: total cholesterol; LDL-C: low density lipoprotein cholesterol; HDL-C: high density lipoprotein cholesterol; TG: triglycerides; AST: aspartate amino transferase; ALT: alanine amino transferase; GGT: gamma glutamyl transferase; LS: liver stiffness; cIMT: carotid intima-media thickness; MDA: malondialdehyde; PON1: paraoxonase 1; SOD: superoxide dismutase; HOI: HDL oxidative index.

P < 0.05 was considered statistically significant.
PON1 serum concentration in the NAFLD group compared to the control group [31, 32]. However, in consistent with our findings, a large population study that used fatty liver index for the diagnosis of NAFLD showed that in men PON-1 activity was significantly higher in the NAFLD population compared to the non-NAFLD population [33]. PON1 has been shown to be critical for the anti-oxidative function of HDL particles. A previous study demonstrated a lower PON-1 activity in patients with an HOI ≥ 1 than the subjects with an HOI < 1 [4]. Although we found lower levels of PON1 paraoxonase activity in all subjects with HOI ≥ 1 compared to the subjects with HOI < 1, we didn’t see any significant difference in PON1 between NAFLD patients with HOI ≥ 1 and the patients with HOI < 1. Considering SOD activity, we observed decreased activity of SOD in NAFLD patients. Similarly, other studies reported lower activity of SOD in NAFLD patients [23, 34, 35]. Our findings showed a lower SOD activity in all subjects with HOI ≥ 1 compared to the subjects...
cIMT is considered a surrogate marker of cardiovas-
comprehensive antioxidant function of HDL and cIMT [26].

young adults did not find an association between the
between HOI and cIMT. However, a previous study on
< 1. We also found a significant positive correlation
jects with HDL
high
levels of pro-oxidative and other detrimental factors
which cause HDL dysfunctional. However, owing to the
multiple anti-oxidative and anti-inflammatory properties
of HDL, the possibility that an impaired anti-oxidative
capacity of HDL may affect disease severity in NAFLD
patients cannot be ruled out.

Finally, we observed higher levels of cIMT in the sub-
jects with HDL ≥ 1 compared to the subjects with HOI
< 1. Moreover, we found that NAFLD patients with the
impaired antioxidant function of HDL (HOI ≥ 1) had
higher AST and LS. Previous studies in NAFLD patients
demonstrated the association of circulating oxidative
stress biomarkers with disease severity [17]. In the light
of the important role of oxidative stress in the progres-
sion of NAFLD, one can speculate that NAFLD patients
with higher liver enzymes and liver fibrosis have higher
levels of pro-oxidative and other detrimental factors
which cause HDL dysfunctional. However, owing to the
multiple anti-oxidative and anti-inflammatory properties
of HDL, the possibility that an impaired anti-oxidative
capacity of HDL may affect disease severity in NAFLD
patients cannot be ruled out.

Conclusions

In summary, our results showed that an impaired anti-
oxidative capacity of HDL in NAFLD patients was asso-
ciated with higher NAFLD markers. This association
suggests either this disease state might attenuate the anti-
oxidative effect of HDL or HDL anti-oxidative function-
ality might contribute to NAFLD pathogenesis.

Abbreviations

NAFLD: Non-alcoholic fatty liver disease; CVD: Cardiovascular disease; HDL: High-density lipoprotein; DCFDA: Dichlorofluorescein diacetate; PON1: Paraoxonase 1; HOI: HDL oxidative index; SOD: Superoxide dismutase (SOD); MDA: Malondialdehyde (MDA); cIMT: Carotid intima-media thickness; LDL-C: Low-density lipoprotein cholesterol; ALT: Alanine amino transferase; AST: Aspartate amino transferase; GGT: Gamma glutamyl transferase; LDL: Low-density lipoprotein cholesterol; WC: Waist circumference; BMI: Body mass index; WHR: Waist to hip ratio; GGT: Gamma glutamyl transferase; ALT: Alanine amino transferase; AST: Aspartate amino transferase; WC: Waist circumference; BMI: Body mass index; WHR: Waist to hip ratio; GGT: Gamma glutamyl transferase; ALT: Alanine amino transferase; AST: Aspartate amino transferase; WC: Waist circumference; BMI: Body mass index; WHR: Waist to hip ratio; GGT: Gamma glutamyl transferase; ALT: Alanine amino transferase; AST: Aspartate amino transferase; WC: Waist circumference; BMI: Body mass index; WHR: Waist to hip ratio; GGT: Gamma glutamyl transferase; ALT: Alanine amino transferase; AS

| Parameter      | Correlation | P value |
|----------------|-------------|---------|
| MDA            | 0.298       | 0.001   |
| PON1           | −0.117      | 0.201   |
| SOD            | −0.242      | 0.004   |
| Age            | 0.056       | 0.503   |
| cIMT           | 0.188       | 0.031   |
| Visceral fat   | 0.01        | 0.909   |
| WC             | 0.011       | 0.900   |
| BMI            | 0.042       | 0.622   |
| WHR            | 0.012       | 0.891   |
| AST            | 0.059       | 0.488   |
| ALT            | 0.067       | 0.432   |
| GGT            | 0.075       | 0.374   |
| LS             | 0.062       | 0.474   |
| FBG            | 0.077       | 0.358   |
| HDL-C          | −0.044      | 0.603   |
| LDL-C          | 0.032       | 0.705   |
| TG             | −0.009      | 0.919   |

Table 2 Correlation coefficients between HOI and metabolic and anthropometric parameters

with HOI < 1 but SOD was not significantly different
in the NAFLD patients with HOI ≥ 1 compared to the
patients with HOI < 1.

There are some limitations to this study. The study
was cross-sectional in nature, which does not provide
information about possible causal relationships among
NAFLD markers, HDL anti-oxidative capacity, and oxy-
dative stress markers. So, further prospective studies
are required to specifically address the potential role of HDL
anti-oxidative function in the development and progres-
sion of NAFLD. We used apolipoprotein B–depleted
plasma samples instead of isolated HDL to measure
the anti-oxidative capacity of HDL. Polyethylene glycol
precipitation is considered as a reproducible and rapid
method to extract HDL from plasma samples [5, 11],
but there is a possibility that anti-oxidative effects are
influenced by other proteins rather than HDL associated
proteins. However, a previous study showed comparable
HOI values determined with apo B depleted sample and
ultracentrifugation isolated HDL.

Supplementary Information

The online version contains supplementary material available at https://doi.org/10.1186/s13098-021-00741-5.

Additional file 1: Figure S1. Comparison of cIMT, plasma levels of MDA,
PON1 and SOD in the subjects with preserved anti-oxidative HDL capacity
(HOI < 1) and impaired anti-oxidative HDL capacity (HOI ≥ 1). The differ-
ences between two groups were analyzed by independent Student’s t
-test and were presented as mean ± SD or median (IQR). MDA malondi-
alone at 1). The differ-
ences between two groups were analyzed by independent Student’s t
test and were presented as mean ± SD or median (IQR). MDA malondi-
aldehyde; PON1: Paraoxonase 1; SOD: Superoxide dismutase (SOD);
MDA: Malondialdehyde (MDA); cIMT: Carotid intima-media thickness; LDL-C:
Low-density lipoprotein cholesterol; ALT: Alanine amino transferase; AST:
Aspartate amino transferase; GGT: Gamma glutamyl transferase.

Please note that the supplementary information contains additional data and analyses related to the study, including a figure comparing carotid intima-media thickness (cIMT), plasma levels of malondialdehyde (MDA), paraoxonase 1 (PON1), and superoxide dismutase (SOD) between patients with preserved and impaired anti-oxidative HDL capacity. This information is crucial for understanding the implications of the findings and for further research in the field.
aspartate amino transferase, LS liver stiffness; cIMT carotid intima-media thickness; MDA malondialdehyde.

Acknowledgements
The research presented in this article is part of the dissertation of Sara Karami to receive a master's degree in clinical biochemistry. The authors wish to thank the study participants, doctors, and co-researchers of Digestive Diseases Research Institute of Tehran University of Medical Sciences.

Authors' contributions
PS designed the study, analyzed the data and wrote the manuscript. AP and HP supervised the study and edited the manuscript. ARR did ultrasonography and fibroscan. SK, FA, and NS performed the experiments and collected the human subjects: All authors read and approved the final manuscript.

Availability of data and materials
The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Declarations
Ethics approval and consent to participate
All participants gave written informed consent. This study was approved by the corresponding ethics committee of the Semnan University of Medical Sciences. The funding body played no role in the design of the study and collection, analysis, and interpretation of data and in writing the manuscript.

Consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

Author details
1Department of Biochemistry, School of Medicine, Semnan University of Medical Sciences, Semnan, Iran. 2Liver and Pancreatobiliary Diseases Research Center, Digestive Diseases Research Institute, Tehran University of Medical Sciences, Tehran, Iran. 3Department of Radiology, Sharari Hospital, Tehran University of Medical Sciences, Tehran, Iran. 4Nervous System Stem Cells Research Center, Semnan University of Medical Sciences, Semnan, Iran. 5Department of Integrative Medical Sciences, Northeast Ohio Medical University, Rootstown, OH, USA.

Received: 11 August 2021 Accepted: 16 October 2021
Published online: 26 October 2021

References
1. Vernon G, Baranova A, Younossi Z. Systematic review: the epidemiology and natural history of non-alcoholic fatty liver disease and non-alcoholic steatohepatitis in adults. Aliment Pharmacol Ther. 2011;34(3):274–85.
2. McCulloch A, Previs SF, Dasarath J, Lee K, Osme A, Kim C, et al. HDL flux is higher in patients with nonalcoholic fatty liver disease. Am J Physiol Endocrinol Metab. 2019;317(5):E852–62.
3. Wang K, Shan S, Zheng H, Zhao X, Chen C, Liu C. Non-HDL-cholesterol to HDL-cholesterol ratio is a better predictor of new-onset non-alcoholic fatty liver disease than non-HDL-cholesterol: a cohort study. Lipids Health Dis. 2018;17(1):1–8.
4. Schrütka L, Distelmaier K, Hohsinnser P, Sulzgruber P, Lang IM, Maurer G, et al. Impaired high-density lipoprotein anti-oxidative function is associated with outcome in patients with chronic heart failure. J Am Heart Assoc. 2016;5(12):e004169.
5. Schrütka L, Distelmaier K, Hohsinnser P, Sulzgruber P, Lang IM, Maurer G, et al. Impaired high-density lipoprotein anti-oxidative function is associated with outcome in patients with chronic heart failure. J Am Heart Assoc. 2016;5(12):e004169.
6. Amor AJ, Pinyol M, Solà E, Catalan M, Coñán M, Herrera Z, et al. Relationship between noninvasive scores of nonalcoholic fatty liver disease and nuclear magnetic resonance lipoprotein abnormalities: a focus on atherogenic dyslipidemia. J Clin Lipidol. 2017;11(2):551-61.e7.
7. Orozco Morales JA, Medina Urrutia AX, Torres Tamayo M, Jorge Galazera E, Reyes Barrera J, Dies Suarez P, et al. Effects of fatty liver on the size and composition of high-density lipoprotein cholesterol subpopulations in adolescents with type 2 diabetes mellitus. Pediatr Diabetes. 2020;21(7):1140–9.
8. Rao PK, Merath K, Drigalenko E, JadHAV AYL, Komorowski RA, Goldblatt MI, et al. Proteomic characterization of high-density lipoprotein particles in patients with non-alcoholic fatty liver disease. Clin Proteomics. 2018;15:10.
9. Morze J, Koch M, Aroner SA, Budhoff M, McClelland RL, Mukamal KJ, et al. Associations of HDL subspecies defined by ApoC3 with non-alcoholic fatty liver disease: the multi-ethnic study of atherosclerosis. J Clin Med. 2020. https://doi.org/10.3390/jcm9113522.
10. Annema W, Willermens HM, de Boer JF, Dikkers A, van der Giet M, Nieuwland W, et al. HDL function is impaired in acute myocardial infarction independent of plasma HDL cholesterol levels. J Clin Lipidol. 2016;10(6):1319–28.
11. Patel PJ, Khera AV, Fjiri J, Wileys NK, Roder DJ. The anti-oxidative capacity of high-density lipoprotein is reduced in acute coronary syndrome but not in stable coronary artery disease. J Am Coll Cardiol. 2011;58(20):2068–73.
12. Morgantini C, Natali A, Boldrini B, Imaizumi S, Navab M, Fogelman AM, et al. Anti-inflammatory and antioxidant properties of HDLs are impaired in type 2 diabetes. Diabetologia. 2017;60(10):2617–23.
13. Zhang J, Zhang Y, Liu H, Bai H, Wang Y, Jiang C, et al. Antioxidant properties of high-density lipoproteins are impaired in women with polycystic ovary syndrome. Fertil Steril. 2015;103(5):1346–54.
14. Fadaei R, Pouochtchi H, Meshkani R, Moradi N, Golmohammadi T, Merat S. Impaired HDL cholesterol efflux capacity in patients with non-alcoholic fatty liver disease is associated with subclinical atherosclerosis. Sci Rep. 2018;8(1):11691.
15. van den Berg EH, Gruppen EG, Ebtehaj S, Bakker S, Tietge UJF, Bullaart RPF. Cholesterol efflux capacity is impaired in subjects with an elevated Fatty Liver Index, a proxy of non-alcoholic fatty liver disease. Atherosclerosis. 2018;277:21–7.
16. Wang B, Yang RN, Zhu YR, Xing JC, Lou XW, He YJ, et al. Involvement of xanthine oxidase and paraoxonase 1 in the process of oxidative stress in nonalcoholic fatty liver disease. Mol Med Rep. 2017;15(1):387–95.
17. Chen Z, Tian R, She Z, Cai J, Li H. Role of oxidative stress in the pathogenesis of nonalcoholic fatty liver disease. Free Radical Biol Med. 2020;152:116–41.
18. Ito T, Ito T, Suzuki C, Yahata T, Ikeda K, Hamaoka K. The application of a modified d-ROMs test for measurement of oxidative stress and oxidized high-density lipoprotein. Int J Mol Sci. 2017;18(2):454.
19. Monserrat-Mesquida M, Quetglas-Llabrés M, Abbate M, Montserrat M, Mascaro CM, Cañeses M, et al. Oxidative stress and pro-inflammatory status in patients with non-alcoholic fatty liver disease. Antioxidants. 2020. https://doi.org/10.3390/antioxidants9090795.
20. Shao B, Pennathur A, Pagani J, Oda MN, Wittzum JL, Oram JF, et al. Modifying apolipoprotein A-I by malondialdehyde, but not by an array of other reactive carbonyls, blocks cholesterol efflux by the ABCA1 pathway. J Biol Chem. 2010;285(24):18473–84.
21. Schill RL, Knaack DA, Powers HR, Chen Y, Yang M, Schill DJ, et al. Modification of HDL by reactive aldehydes alters select cardioprotective properties of HDL in macrophages. FEBS J. 2020;287(6):695–707.
22. Sehitogullari A, Aslan M, Sayır F, Kahraman A, Demir H. Serum paraoxonase-1 enzyme activities and oxidative stress levels in patients with esophageal squamous cell carcinoma. Redox Rep. 2014;19(5):199–205.
23. Aya A, Azarmehr N, Mansourian M, Doustismitagh AAH. Inactivation of the superoxide dismutase by malondialdehyde in the nonalcoholic fatty liver disease: a combined molecular docking approach to clinical studies. Arch Physiol Biochem. 2019. https://doi.org/10.1080/13813455.2019.1659827.
24. Świderska M, Maciejczyk M, Zalewska A, Pogorzelska J, Flisiak R, Chabowski A. Oxidative stress biomarkers in the serum and plasma of patients with non-alcoholic fatty liver disease (NAFLD) Can plasma AGE be a marker of NAFLD? Oxidative stress biomarkers in NAFLD patients. Free Radic Res. 2019;53(8):841–50.

25. Merat S, Pouschti H, Hemming K, Jafari E, Radmard AR, Nateghi A, et al. PolyPill for prevention of cardiovascular disease in an urban iranian population with special focus on nonalcoholic steatohepatitis: a pragmatic randomized controlled trial within a cohort (polyiran—liver)—study protocol. Arch Iran Med. 2015;18(8):515–23.

26. Breton CV, Yin F, Wang X, Avol E, Gilliland FD, Araujo JA. HDL anti-oxidant function associates with LDL level in young adults. Atherosclerosis. 2014;232(1):165–70.

27. Lee S, Park G-M, Lee J-Y, Lee BU, Park JH, Kim BG, et al. Association between non-alcoholic fatty liver disease and subclinical coronary atherosclerosis: an observational cohort study. J Hepatol. 2018;68(5):1018–24.

28. Gaál K, Tarr T, Lőrincz H, Borbás V, Seres I, Harangi M, et al. High-density lipoprotein antioxidant capacity, subpopulation distribution and paraoxonase-1 activity in patients with systemic lupus erythematosus. Lipids Health Dis. 2016;15(1):1–8.

29. Ito F, Ito T. High-density lipoprotein (HDL) triglyceride and oxidized HDL: new lipid biomarkers of lipoprotein-related atherosclerotic cardiovascular disease. Antioxidants. 2020;9(5):362.

30. Carnuta MG, Stancu CS, Toma L, Sanda GM, Niculescu LS, Deleanu M, et al. Dysfunctional high-density lipoproteins have distinct composition, diminished anti-inflammatory potential and discriminate acute coronary syndrome from stable coronary artery disease patients. Sci Rep. 2017;7(1):1–13.

31. Milaciu MV, Vesa ŞC, Bočzan IC, Ciumăeanu L, Sâmpelean D, Negrean V, et al. Paraoxonase-1 serum concentration and PON1 gene polymorphisms: relationship with non-alcoholic fatty liver disease. J Clin Med. 2019;8(12):2200.

32. Sammy W, Hassanian MA. Paraoxonase-1 activity, malondialdehyde and glutathione peroxidase in non-alcoholic fatty liver disease and the effect of atorvastatin. Arab J Gastroenterol. 2011;12(2):80–5.

33. van den Berg EH, Gruppen EG, James RW, Bakker SJL, Dullaart RPF. Serum paraoxonase 1 activity is paradoxically maintained in nonalcoholic fatty liver disease despite low HDL cholesterol. J Lipid Res. 2019;60(1):168–75.

34. Koruk M, Taysi S, Savas MC, Yilmaz O, Akcay F, Karakok M. Oxidative stress and enzymatic antioxidant status in patients with nonalcoholic steatohepatitis. Ann Clin Lab Sci. 2004;34(1):57–62.

35. Videla LA, Rodrigo R, Orellana M, Fernandez V, Tapia G, Quiñones L, et al. Oxidative stress-related parameters in the liver of non-alcoholic fatty liver disease patients. Clin Sci. 2004;106(3):261–8.

36. Willett P, Tschirner L, Allara L, Reuber K, Seekircher L, Gao L, et al. Carotid intima-media thickness progression as surrogate marker for cardiovascular risk: meta-analysis of 119 clinical trials involving 1,00,667 patients. Circulation. 2020;142(7):621–42.

Publisher's Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.