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Article  (Published Version)

Jeng, Jing-Yi, Carlton, Adam J, Goodyear, Richard J, Chinowsky, Colbie, Ceriani, Federico, Johnson, Stuart L, Sung, Tsung-Chang, Dayn, Yelena, Richardson, Guy P, Bowl, Michael R, Brown, Steve D M, Manor, Uri and Marcotti, Walter (2022) AAV-mediated rescue of Eps8 expression in vivo restores hair-cell function in a mouse model of recessive deafness. Molecular Therapy - Methods and Clinical Development, 26. pp. 355-370. ISSN 2329-0501

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AAV-mediated rescue of Eps8 expression in vivo restores hair-cell function in a mouse model of recessive deafness

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The transduction of acoustic information by hair cells depends upon mechanosensitive stereociliary bundles that project from their apical surface. Mutations or absence of the stereociliary protein Eps8 cause deafness in humans and mice, respectively. Eps8 knockout mice (Eps8−/−) have hair cells with immature stereocilia and fail to become sensory receptors. Here, we show that exogenous delivery of Eps8 using Anc80L65 in P1–P2 Eps8−/− mice in vivo rescued the hair bundle structure of apical-coil hair cells. Rescued hair bundles correctly localize Eps8, WHIRLIN, MYO15, and BAIAP2L2, and generate normal mechanoelectrical transducer currents. Inner hair cells with normal-looking stereocilia re-expressed adult-like basolateral ion channels (BK and KCNQ4) and have normal exocytosis. The number of hair cells undergoing full recovery was not sufficient to rescue hearing in Eps8−/− mice. Adeno-associated virus (AAV)-transduction of P3 apical-coil and P1–P2 basal-coil hair cells does not rescue hair cells, nor does Anc80L65-Eps8 delivery in adult Eps8−/− mice. We propose that AAV-induced gene-base therapy is an efficient strategy to recover the complex hair-cell defects in Eps8−/− mice. However, this therapeutic approach may need to be performed in utero since, at postnatal ages, Eps8−/− hair cells appear to have matured or accumulated damage beyond the point of repair.

INTRODUCTION

Hearing loss (HL) is a very common condition that can arise as a congenital birth defect or can develop postnatally. According to a recent report from the World Health Organization (WHO), by 2050 it is projected that nearly 2.5 billion people globally will suffer some degree of HL, with genetic predisposition being a key factor. Indeed, in developed countries, pre-lingual HL has an incidence of ∼1 in 500, and in ∼80% of cases there is a genetic cause.1 To date, mutations in 124 genes have been identified as causing monogenic non-syndromic HL in humans, several of which are expressed in the stereociliary bundles of hair cells.2,3

Cells use actin-based plasma membrane protrusions to detect cues from their environment, which are then used to regulate several physiological processes such as motility, adhesion, and even mechanosensation.1–6 In the cochlea, specialized microvilli-like structures emerge from the apical surface of the sensory inner hair cells (IHCs) and outer hair cells (OHCs). These membrane protrusions, called stereocilia, are key for transducing acoustic information into the hair-cell receptor potentials, which are then used to drive the action potential activity in the auditory fibers.7 This mechanoelectrical transduction is operated by mechanically gated ion channels positioned near the tips of hair-cell stereocilia.8 Stereocilia are composed of a compact array of parallel and uniformly polarized actin filaments,1,9,12 and the length of each stereocilium is scaled precisely to give rows of stereocilia (hair bundle) that form a staircase-like architecture. The hair-cell stereociliary bundles are composed of three rows of stereocilia, which are cross-linked by several types of extracellular links.11,12 The architecture of the hair bundles is so precise that the height and width of stereocilia within each row is similar, not only within a single hair bundle but also between the bundles of adjacent hair cells. In altricial rodents, this sophisticated control over the growth of stereocilia occurs during late embryonic and early postnatal stages,13–15 and is tightly regulated by several actin-binding proteins and unconventional myosin motors.5,12 One of the key molecules regulating the length of the actin filaments in the stereocilia of hair cells is epidermal growth factor receptor substrate 8 (EPS8).16–18

Received 30 April 2022; accepted 15 July 2022; https://doi.org/10.1016/j.omtm.2022.07.012.
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Here, we investigated the ability of adeno-associated virus (AAV) vector Anc80L65 to rescue the complex morphological and functional defects present in hair cells from Eps8−/− mice. This AAV vector, containing the coding sequence of wild-type Eps8 with an N-terminal FLAG tag (referred to hereafter as Anc80L65-Eps8), was injected through the round window membrane into the cochlea of P1–P2 mice. We found that transduced hair cells expressing exogenous Eps8 showed a variable degree of hair bundle morphological recovery. Transduced hair cells with fully rescued hair bundles had normal mechanoelectrical transduction and were able to progress in their maturation. We propose that AAV delivery of exogenous Eps8 in vivo is an efficient method to recover the complex hair-cell dysfunctions in Eps8−/− mice, but it may need to be performed in utero to recover hearing function, as the work presented here suggests the existence of a critical window for functional hair-cell recovery.

RESULTS

Transduction of cochlear hair cells using AAVs in vivo

The transduction efficiency of the synthetic adeno-associated viral vector Anc80L65 was determined from the number of GFP-positive hair cells (injected Anc80L65-GFP, in B and Eps8-positive hair cells (injected Anc80L65-Eps8, as in C and D) and in each cochlear preparation normalized by the total hair cells identified with phalloidin. In (E) and (F), the number of mice/cochlea (one mouse = one cochlea) is indicated next to the data. (G) ABR thresholds for frequency-specific pure tone stimulation from 3 to 42 kHz recorded from non-injected (black) and injected (magenta: surgery and RWM injection of Anc80L65-GFP) C57BL/6N wild-type mice at 19–26 days. The number of mice tested is shown. The dashed line represents the upper threshold limit of our system, 95 dB. Single animal recordings are plotted as faded lines. (H) Average ABR thresholds for click stimuli recorded from the same mice described in (G). Number of mice tested is shown above the average data, and single data points are plotted as small open symbols. (I) DPOAE thresholds measured from a subset of mice used in (E). The frequency range tested was between 6 and 24 kHz. Dashed line: upper threshold limit of our system, 80 dB. Single-animal recordings are shown as faded lines. Data are mean ± SD.
auditory brainstem responses (ABRs) from injected and non-injected C57BL/6N wild-type mice between P19 and P26 (Figures 1G–1I). ABRs, which measure the summed sound-induced activity of the auditory pathway, showed thresholds for tone and click stimuli that were indistinguishable between injected and non-injected mice (Figure 1G, p = 0.8852, two-way ANOVA; Figure 1H, p = 0.3574, t-test). Distortion product otoacoustic emissions (DPOAEs), which are a readout of active non-linear cochlear amplification due to OHC activity, were also comparable between the two experimental conditions (Figure 1I, p = 0.0527, two-way ANOVA). Because transduction efficiency of Anc80L65 was higher in apical-coil IHCs, most of the following experiments were performed in this region.

Anc80L65-EPS8 promotes stereocilia growth in Eps8 knockout mice

EPS8 has been shown to localize at the tips of stereocilia of both IHCs and OHCs and is essential for their normal elongation. In adult wild-type mice, both IHCs and OHCs are composed of three rows of stereocilia (tall, intermediate, and short), which can be easily identified using scanning electron microscopy (Figure 2A; see also Fettiplace and Kim). In Eps8−/− adult mice, the hair bundles of both IHCs and OHCs are comparable with those present during the early stages of immature development, displaying several rows of very short stereocilia (Figure 2B). Therefore, we investigated whether the injection of Anc80L65 containing the coding sequence of wild-type Eps8 with an N-terminal FLAG tag (referred to hereafter as Anc80L65-Eps8) in the RWM in P1–P2 pups was able to rescue the normal growth of stereocilia in Eps8−/− mice. We found that about 3 weeks after AAV delivery in Eps8−/− mice (i.e., at P22–P26), the hair bundles of both IHCs and OHCs showed largely improved structure (Figure 2C). However, the degree of recovery was quite variable, with some hair bundles from both IHCs (Figures 2C and S1) and OHCs (Figures 2C and S2) being indistinguishable from those present in wild-type mice (Figure 2A), and others showing an intermediate or a very immature phenotype (Figures 2C, S1, and S2). When mice were injected with Anc80L65-Eps8 at P3, we observed almost no recovery (Figure 2D).

We then established whether the rescued hair bundles from Eps8−/− mice transduced with Anc80L65-Eps8 at P1–P2 showed a normal distribution of EPS8 using immunofluorescence labeling with antibodies against EPS8 and, for the transduced hair cells, the FLAG tag fused to the N terminus of EPS8 (see section “materials and methods”). Similar to wild-type mice (Figures 3A and 3B; see also Zampini et al. and Furness et al.), hair cells from Eps8−/− mice transduced with Anc80L65-Eps8 were able to localize EPS8 at the tips of their stereocilia, while Eps8−/− mice, both IHCs and OHCs have extra rows of short stereocilia (B, see insets), which is in agreement with previous findings. In (C), arrows indicate hair bundles from Eps8−/− mice that have regained the normal morphology. Arrowheads indicate hair bundles that show a variable degree of recovery. (D) Images from a P22 Eps8−/− mouse that was transduced with Anc80L65-Eps8 in vivo at P3. Almost every IHC retained an immature hair bundle morphology (arrowheads), with some stereocilia having elongated (arrows).
transducing shortest two rows of stereocilia is less well understood, (Figures 4F and 4G). Although the growth and maintenance of the Eps8 with the different experimental conditions: highlighted in the orthogonal projections images of the stereocilia (D). (F) Height of the tallest row of stereocilia, which is that expressing EPS8 in adult mice, in IHCs from with Anc80L65-EPS8 at P2 (C–E). Hair cells were immunostained for EPS8 and, for the Anc80L65-EPS8 transduced mice, also for FLAG tag. Note the characteristic local-
and E, middle panels) as also shown in the merge images (C and E, right panels). The elongation of the stereocilia in EPS8+/– mice that underwent transduction is better highlighted in the orthogonal projections images of the stereocilia (D). (F) Height of the tallest row of stereocilia, which is that expressing EPS8 in adult mice, in IHCs from the different experimental conditions: EPS8+/+ (four cochleae), EPS8+/– (four cochleae), and following the injection of Anc80L65-EPS8 (EPS8+/– + Anc80L65) (seven cochleae). Number of mice/cochlea tested (one mouse = one cochlea) is shown above the average data and single data points are plotted as small open symbols. ANOVA Tukey‘s post test: p = 0.0003 and p < 0.0001 for ANOVA; Figure 3F). The height of the bundles of wild-type and MYO15.33,34 We found that only the rescued hair bundles from P25–P29 EPS8−/− mice were able to localize BAIAP2L2 in their shorter rows of stereocilia (Figures 4F and 4I), which has been shown to be dependent not only on EPS8 but also on the motor protein MYO15.25

taller stereocilia (Figures 3C, 3D, and 3E). Transduced hair cells also showed a stereocilia tip localization of the FLAG tag, which overlapped with the EPS8 staining (Figures 3C, 3D, and 3E). The length of the tallest row of stereocilia in the IHCs was quantified, as these were easier to measure than those of OHCs, by labeling the actin core with phalloidin (see section "materials and methods"). We found that the stereocilia height in IHCs from non-rescued EPS8−/− mice was significantly smaller than that measured in cells from wild-type and EPS8+/− mice with rescued hair bundles (p < 0.0001, one-way ANOVA; Figure 3F). The height of the bundles of wild-type and EPS8−/− mice with fully rescued hair bundles were not significantly different (p = 0.8863).

EPS8, MYO15, and WHIRLIN are all hair bundle proteins that localize at the tip of the taller rows of stereocilia of P25–P29 wild-type mice (Figures 4A–4C), and are involved in their selective lengthening and widening relative to the other shorter rows by promoting F-actin polymerization at their tips.16,31,32 In apical-coil IHCs, we found that both MYO15 and WHIRLIN were absent in P25–P29 EPS8−/− mice at the tip of the tallest stereocilia (Figures 4D and 4E), but co-localized with EPS8 at the tips of the taller stereocilia in P25–P29 EPS8−/− mice transduced with Anc80L65-Eps8 at P1 (Figures 4F and 4G). Although the growth and maintenance of the transducing shortest two rows of stereocilia is less well understood, they require EPS8 during early stages of development17,33 and several other proteins, including the I-BAR protein BAIAP2L2,25 EPS8L2,18 and MYO15.33,34 We found that only the rescued hair bundles from P25–P29 EPS8−/− mice were able to localize BAIAP2L2 in their shorter rows of stereocilia (Figures 4F and 4I), which has been shown to be dependent not only on EPS8 but also on the motor protein MYO15.25

Cellular differentiation, which is defined by Atoh1 expression during embryonic stages, changes along the length of the cochlea with basal hair cells being about 2–3 days ahead compared with apical cells.35 Because of this differential development along the cochlea, at P0–P1 basal-coil, but not apical-coil, hair cells exhibit a functional mechanoelectrical transducer current and larger basolateral membrane currents.16–38 We found that, in contrast to apical-coil IHCs, the bundles of hair cells located toward the basal end of the cochlea of EPS8−/− mice transduced with Anc80L65-Eps8 did not elongate, and both EPS8 and WHIRLIN showed a more diffuse distribution along the length of several rows of stereocilia (Figures 4J and 4K).

Despite the morphological recovery of the hair bundles of at least some of the hair cells from EPS8−/− mice transduced with Anc80L65-Eps8 at P1–P2, we did not observe any functional recovery of hearing measured using either ABR or DPOAE (Figure S3).
Anc80L65-Eps8 rescues the mechanotransducer currents in Eps8−/− mice

Hair cells from Eps8−/− mice have been shown to have a wide range of morphological and physiological dysfunctions. Therefore, we investigated whether the absence of hearing in Eps8−/− mice injected with Anc80L65-Eps8 at P1–P2 was due to the limited ability to restore key functional characteristics of developing hair cells separate from hair bundle structure. To test this possibility, we focused the following work on the IHCs since they show a higher viral-transduction rate than OHCs (Figure 1), and their taller stereocilia makes it easier to recognize their morphology using our upright microscope with differential interference contrast (DIC) optics and a water immersion objective (see section “materials and methods”). In the Eps8−/− mice transduced with Eps8, the following experiments were performed from IHCs that exhibited three well-defined rows of stereocilia with homogeneous height, which are about 21% of those viewed under the field of view of our microscopes.

The functional rescue of the hair bundles from Eps8−/− mice transduced with Anc80L65-Eps8 at P1–P2 was investigated by recording the mechanoelectrical transducer (MET) current. MET currents were recorded from P9 apical-coil IHCs by displacing their stereociliary bundles using a 50-Hz sinusoidal force stimulus from a piezo-driven fluid jet. Moving the bundles toward the taller stereocilia (i.e., in the excitatory direction) elicited a large inward MET current in IHCs from wild-type mice (Figure 5A),...
Eps8–/– mice (Figure 5B), and Eps8–/– mice transduced with the viral vector (Figure 5C). By stepping the membrane potential from –124 mV to more depolarized values in 20-mV increments, the transducer current decreased in size at first and then reversed near 0 mV in all experimental conditions (Figure 5D), consistent with the non-selective permeability of MET channels to cations. At –124 mV, the maximal MET current in IHCs from wild-type mice (–944 ± 260 pA, n = 6 at –124 mV) was significantly smaller than that measured in Eps8–/– mice (–1,807 ± 525 pA, n = 7, p = 0.0026, one-way ANOVA, Tukey’s post test; Figure 5E). The larger MET current in Eps8–/– mice correlates with their larger number of stereociliary rows, and most likely MET channels, as previously shown.17 Hair cells from AAV-transduced Eps8–/– mice showed an MET current that was no longer significantly higher than wild-type mice (–1,259 ± 273 pA, n = 6, p = 0.3546, Tukey’s post test; Figure 5E). The maximal MET current at +96 mV (Figure 5F) follows a similar trend to –124 mV. Eps8–/– mice transduced with Anc80L65-Eps8 showed a similar current size at +96 mV (+921 ± 234 pA, n = 6) to wild-type (+818 ± 212 pA, n = 6, p = 0.8339) but was significantly reduced compared with that measured in cells from Eps8–/– mice (+1,424 ± 416 pA, n = 7, p = 0.0076).

One important feature of the MET channel is its resting open probability (P_o), which allows the flow of an MET current through open transducer channels in the absence of mechanical stimulation. The resting MET current can be measured as the difference between the holding current and the current present during inhibitory bundle deflection, which completely closes the MET channel (Figures 5A–5C). At both negative and positive membrane potentials, the resting MET current was evident in IHCs from wild-type mice (Figures 5A, 5G, and 5H), but absent in cells from Eps8–/– mice (Figures 5B, 5G, and 5H), in agreement with a previous study.17 The larger resting MET current at positive membrane potentials is due to an increased P_o of the MET channel resulting from a reduced driving force for Ca^{2+} influx.17,41 Eps8–/– mice transduced with Anc80L65-Eps8 showed a rescue of the resting MET current, which was not significantly different from that measured in IHCs of wild-type mice (p = 0.5610, at –124 mV, p = 0.2150; at +96 mV, one-way ANOVA, Tukey’s post test; Figures 5C, 5G, and 5H).

The above data indicate that injection of Anc80L65-Eps8 into the cochlea of early-postnatal Eps8–/– pups (P1–P2) is able to restore the biophysical characteristics of the MET current in IHCs that have normal hair bundle morphology.
IHCs from Eps8−/− mice transduced with Anc80L65-Eps8 mature into normal sensory receptors

The basolateral membrane properties of IHCs from Eps8−/− mice have been shown to retain features of early stages of development, since they fail to express adult-like ion channels during maturation.17 We investigated, therefore, whether P19–P28 IHCs from Eps8−/− mice transduced with the Anc80L65-Eps8 viral vector were able to mature into functional sensory receptors (Figures 6A–6K).

characteristic K+ current present in adult IHCs is IK,f which is a rapid-activating, large-conductance Ca2+-activated K+ current carried by BK channels.42–44 Another mature-type K+ current in IHCs is that carried by KCNQ4 channels (IK,n).37,45 Both IK,f and IK,n were present in wild-type IHCs (Figures 6A and 6D), and both currents were absent in Eps8−/− mice (Figures 6B, 6E, and 6G) as previously shown.17 IHCs from Eps8−/− mice transduced with Anc80L65-Eps8, which showed normal-looking hair bundles, showed both currents (Figures 6C and 6K).

Figure 6. Rescue of basolateral membrane properties in adult IHCs from Eps8−/− mice injected with Anc80L65-Eps8

(A–C) Current responses from IHCs of wild-type (Eps8+/+, P19: A), knockout (Eps8−/−, P28: B), and Eps8+/− injected with Anc80L65-Eps8 (P22, C) mice. Current recordings were elicited by using depolarizing voltage steps (10-mV increments) from the holding potential of −84 mV to the various test potentials shown by some of the traces. The characteristic fast-activating BK current (IK,f) present in adult IHCs (A), which is better appreciated in the expanded timescale (see insets), was absent in Eps8−/− mice (B) but rescued in cells transduced with Anc80L65-Eps8 (C). (D–F) Current responses from the same mouse models described in (A)–(C), but elicited using hyperpolarizing and depolarizing voltage steps (10-mV increments) from the holding potential of −84 mV to better highlight the inward currents. Adult IHCs from both Eps8+/+ (D) and Eps8−/− mice injected with Anc80L65-Eps8 (F) express the characteristic KCNQ4-mediated delayed rectifier IK,n in IHCs from Eps8−/− mice (E) retained the inward rectifier IK1, which is normally expressed only during pre-hearing stages.36 (G) Maximum intensity projections of confocal z stacks taken from the apical cochlear region of Eps8−/− (top) and Eps8+/− rescued (bottom) mice using antibodies against BK channels carrying IK,f (red) and the hair-cell marker Myo7a (white). (H) Steady-state current-voltage curves obtained from IHCs of Eps8+/+ (P19–P28), Eps8−/− mice (P20–P28) and Eps8+/− injected (P22) mice. (I) Size of the fast-activating BK outward K+ current IK,f, which was measured at −25 mV and at 1 ms from the onset of the voltage step.37 (J) Size of the KCNQ4-mediated IK,n, which was measured as the difference between the peak and steady state of the deactivating inward current at −124 mV.36 (K) Size of the inward rectifier IK1 measured as the steady-state inward current at −124 mV.36 In (I)–(K), single cell value recordings (open symbols) are plotted behind the average data. Number of IHCs investigated is shown above the average datapoints. One-way ANOVA: p < 0.0001 (I–K) is indicated by the star symbol. For post test, see section “results”. Data are plotted as means ± SD.
6F). BK channels were also normally localized near the neck region of the IHCs (Figure 6G) as previously shown.\(^\text{46}\) Over the entire voltage range investigated (−124 and +46 mV), the total K\(^+\) current was significantly smaller in non-transduced Eps8\(^{−/−}\) mice compared to wild-type mice (Figure 6H). When \(I_{K,f}\) and \(I_{K,n}\) were investigated in isolation, their size was not significantly different in IHCs from wild-type and Eps8\(^{−/−}\) mice transduced with Anc80L65-Eps8 (\(p = 0.3487\), one-way ANOVA; Figure 6H). When \(I_{K,n}\) was estimated by measuring increases in cell membrane capacitance (\(\Delta C_m\)) following depolarizing voltage steps that activate the Ca\(^{2+}\) current but not the resting membrane potentials were similar between the different mice investigated (\(p = 0.3815\) and \(p = 0.9357\), respectively, two-way ANOVA; Figure 6H). Exocytosis was not control (D) and transduced (F) cells, the post-synaptic ribbons of CtbP2 and GluR2 puncta in IHCs from Eps8\(^{+/+}\) (black) and Eps8\(^{−/−}\) (blue) transduced with Anc80L65-Eps8 (magenta) mice. Please note that, for Eps8\(^{−/−}\) mice, we could not quantify the GluR2 labeling because it shows a very diffuse profile. (G) Number of CtbP2 and GluA2 puncta in IHCs from Eps8\(^{+/+}\), wild-type and viral transduced Eps8\(^{−/−}\) mice; \(I_{K1}\), wild-type and viral transduced Eps8\(^{−/−}\) mice was comparable.

We then investigated exocytosis from the ribbon synapses of IHCs transduced with Anc80L65-Eps8 (Figures 7A–7C). Exocytosis was estimated by measuring increases in cell membrane capacitance (\(\Delta C_m\)) following depolarizing voltage steps that activate the Ca\(^{2+}\) current (see section “materials and methods”). A previous study has shown that, in Eps8\(^{−/−}\) mice, IHCs retain a high-order Ca\(^{2+}\) dependence of glutamate-containing vesicle release,\(^\text{17}\) which is normally only present during pre-hearing stages of development in apical-coil IHCs.\(^\text{37,40}\) We found that IHCs from Eps8\(^{−/−}\) mice elicit comparable graded-receptor potentials in response to depolarizing current injections, which is the normal behavior for adult cells.\(^\text{42}\) However, the resting membrane potentials were similar between the different mice investigated (\(p = 0.3487\), one-way ANOVA; Figure 6H), indicating that the contribution of the K\(^+\) currents active at rest (\(I_{K,n}\)) at −71 and −11 mV, showing that Eps8\(^{−/−}\) mice transduced with Anc80L65-Eps8 show a normal linear Ca\(^{2+}\) dependence of exocytosis. Data were approximated using a power function \(\Delta C_m = k I_{Ca}^n\), where \(k\) is a scaling coefficient and the exponent \(n\) is the power. (D–F) Maximum intensity projections of confocal z stacks of apical-coil IHCs from >P20 Eps8\(^{+/+}\) (D), Eps8\(^{−/−}\) (E), and Eps8\(^{−/−}\) transduced with Anc80L65-Eps8 at P1 (F) mice. IHCs were labeled with antibodies against CtbP2 (ribbon synaptic marker: red) and GluR2 (post-synaptic marker: green). Insets show a magnified view of the co-localization between the CtbP2 and GluR2. Note that, in the Eps8\(^{−/−}\) IHCs (E), but not control (D) and transduced (F) cells, the post-synaptic marker GluR2 shows a diffuse distribution (see arrows in the magnified view of E). Scale bars in the insets represent 2 \(\mu\)m. (G) Number of CtbP2 and GluA2 puncta in IHCs from Eps8\(^{+/+}\) (black) and Eps8\(^{−/−}\) (blue) transduced with Anc80L65-Eps8 (magenta) mice. Please note that, for Eps8\(^{−/−}\) mice, we could not quantify the GluR2 labeling because it shows a very diffuse profile. (H) Number of co-localized CtbP2 and GluR2 puncta in Eps8\(^{+/+}\) (black) and Eps8\(^{−/−}\) transduced with Anc80L65-Eps8 (magenta) mice. Data in (G) and (H) are plotted as mean values and individual counts are shown as open symbols. Numbers above or below the data represent the IHCs (and mice) used for each genotype. Average values are mean ± SD.
transduced with Anc80L65-Eps8 at P1 develop a normal linear Ca\textsuperscript{2+}-dependent exocytosis, as indicated by the nearly unitary exponent resulting from the fit with the power function (0.94 ± 0.23, n = 5; Figure 7C). This power value was not significantly different to that previously measured in Eps8\textsuperscript{+/−} mice (1.17 ± 0.26, n = 5, p = 0.8872), but significantly different from Eps8\textsuperscript{−/−} mice (3.35 ± 1.31, n = 5, p = 0.0010, Tukey’s post test, one-way ANOVA).\textsuperscript{17}

To investigate the synapses between the IHCs and the spiral ganglion afferent terminals, which are key to proper sound encoding, we used antibodies to label the presynaptic ribbon protein RIBEYE (CTBP2) and the post-synaptic AMPA-type glutamate receptor GluR2.\textsuperscript{49,50} Compared with wild-type Eps8\textsuperscript{+/+} hair cells, we found that both CTBP2 and GluR2 puncta were present at the pre- and post-synaptic sites, respectively, in IHCs from both the Eps8\textsuperscript{+/+} and Eps8\textsuperscript{−/−} mice transduced with Anc80L65-Eps8 (Figures 7D–7F). The number of CTBP2 puncta in IHCs was not significantly different between Eps8\textsuperscript{+/+}, Eps8\textsuperscript{−/−}, and Eps8\textsuperscript{−/−} mice transduced with Anc80L65-Eps8 (p = 0.3839, one-way ANOVA; Figure 7G). Post-synaptic GluR2 labeling was organized in its characteristic puncta-like distribution in Eps8\textsuperscript{+/+} mice (Figure 7D), but we found that Eps8\textsuperscript{−/−} mice showed a more diffuse organization of GluR2, which prevented the reliable quantification of discrete puncta (Figure 7E, arrows). Anc80L65-Eps8 restored the normal puncta-like organization of GluR2 in the Eps8\textsuperscript{−/−} mice (Figure 7F). A similar diffuse distribution of GluR2 was recently described in Tmc1;Tmc2 double-knockout mice, in which the MET current is absent,\textsuperscript{31} similar to the Eps8\textsuperscript{−/−} mice (Figure 7E; see also Zampini et al.).\textsuperscript{17} The number of GluR2 puncta was not significantly different between the non-transduced Eps8\textsuperscript{+/+} mice and the Eps8\textsuperscript{−/−} mice transduced with Anc80L65-Eps8 (p = 0.1367, t test; Figure 7G) and showed statistically similar co-localization with CTBP2 puncta-like organization (p = 0.5877, t test; Figure 7H).

Together, these findings show that virally transduced Eps8\textsuperscript{−/−} IHCs can produce normal-looking hair bundles and can fully mature into functional sensory receptors.

**OHC stereocilia from Eps8\textsuperscript{−/−} mice transduced with AAV do not attach to the tectorial membrane**

OHC stereocilia are attached to the tectorial membrane and react against this structure in response to sound stimuli. This connection can be confirmed by the presence of imprints of the tallest stereocilia in the tectorial membrane or a protein that connects the stereocilia to the tectorial membrane, such as stereocilin.\textsuperscript{32} Labeling of whole-mount cochlear preparations from P22 wild-type (C57BL/6N) mice with antibodies to stereocilin revealed puncta primarily localized at the tips of the tallest row of stereocilia, with some staining also associated with the stereocilia of the shorter rows (Figure S5A). In contrast, in Eps8\textsuperscript{−/−} mice, in which the stereocilia are shorter, stereocilin is no longer present to the tips of the taller stereocilia (Figure S5B). In Eps8\textsuperscript{−/−} mice in which the cochlea were injected with Anc80L65-Eps8, and OHC stereocilia showed near-normal growth, stereocilin nevertheless failed to localize to the tips of the tallest row of stereocilia (Figure S5C). Consistent with these observations, examination of the underside of the tectorial membrane (TM) in preparations stained with wheat-germ agglutinin revealed that imprints were only present in TMs derived from wild-type cochlea, and not from those of Eps8\textsuperscript{−/−} or Anc80L65-Eps8 transduced Eps8\textsuperscript{−/−} mice (Figures S5D–S5F). These findings show that the even few OHCs that were showing normal-looking hair bundles were unable to interact with the TM, thus explaining the lack of DPOAEs in these mice (Figure S3).

**Injection of Anc80L65-Eps8 into the cochlea of adult mice fails to rescue Eps8 localization at the tip of stereocilia**

We tested the ability of the AAVs to transduce cochlear hair cells of older mice. Despite the successful transduction of Anc80L65-Eps8 in adult Eps8\textsuperscript{−/−} mice (injected at P19–P20), transduced IHCs failed to localize Eps8 to the tips of their taller stereocilia (Figure 8A). Instead, Eps8 appears to be present along the entire length of these stereocilia, which predominantly remained short. A few hair cells still possessed extra rows of short stereocilia normally associated with immature hair bundles. A few stereocilia within bundles of transduced hair cells did elongate; however, these still showed a diffuse labeling of Eps8 (Figure 8A, arrows). Even though the bundle structure of the IHCs from Eps8\textsuperscript{−/−} mice transduced with Anc80L65-Eps8 was similar to that of IHCs from Eps8\textsuperscript{−/−} mice, they were able to localize MYO15 and WHIRL1 to their stereocilia (Figures 8C–8E). However, similar to the distribution of Eps8 (Figure 8A), or that of WHIRL1 in baso-coil IHCs (Figures 4J and 4K), these proteins showed a more diffuse distribution along the length of several rows of stereocilia (Figures 8C–8E, arrows).

**DISCUSSION**

The correct development of cochlear hair cells into fully mature sensory receptors is a tightly regulated process requiring the interaction of several proteins, including the actin-regulatory protein Eps8.\textsuperscript{16,17} The absence of Eps8 has been shown to halt the normal development of hair cells, especially that of the IHCs, holding them at pre-hearing immature stages, preventing them from processing acoustic stimuli.\textsuperscript{17} We demonstrated that exogenous delivery of Eps8 using the AAV vector Anc80L65 in Eps8\textsuperscript{−/−} mice in vivo can restore the functional characteristics of transduced apical-coil hair cells in adult mice. Exogenous Eps8 was able to rescue not only the normal staircase structure of the stereociliary bundle and have normal mechanoelectrical transduction but also the expression of adult-like basolateral ion channels and the distribution of post-synaptic glutamate receptors. Despite the full recovery of some hair cells, we observed very little to no recovery of hearing function. This may be due to the failure of transduced OHC bundles to attach to the TM. We also found that transduced hair cells from P3 apical coil or P1–P2 basal coil of the cochlea maintained the immature hair bundle structure characteristic of Eps8\textsuperscript{−/−} mice, and did not localize Eps8 to stereocilia tips, similar to when exogenous delivery of Eps8 was performed in P19–P20 Eps8\textsuperscript{−/−} mice with functionally mature IHCs. We propose that exogenous gene augmentation using AAV vectors in vivo is a suitable strategy to rescue complex morphological and functional defects present in
hairstones is a mouse model of recessive deafness. However, by postnatal ages, most of the hair cells of Eps8$^{−/−}$ mice may have matured or accumulated damage beyond the point for certain morphological or physiological features to be repaired. Therefore, we propose that, for genes that regulate early hair-cell development (i.e., embryonic onset of function), gene-based therapeutic approaches may need to be performed in utero in order to recover hearing function.

**In vivo delivery of AAV-Eps8 in early-postnatal Eps8-deficient mice restores the function of hair cells**

The precise staircase structure of the stereociliary bundles of cochlear hair cells develops primarily during pre-hearing stages (prior to ~P12 in mice) through a tightly controlled process of elongation and thickening, as well as an elimination of redundant stereocilia.\(^5\),\(^3\) This precise regulation of the stereociliary actin cytoskeleton is defined by several proteins, most of which are already expressed in the emerging stereocilia during embryonic stages of development,\(^2\),\(^3\),\(^3\),\(^3\) and their absence leads to largely undeveloped hair bundles in both IHCs and OHCs, and consequent hearing loss. Eps8 is one of these key proteins since it is expressed from late embryonic stages of development in mice and it is required for the initial elongation of the immature stereociliary bundles of hair cells, as well as the reabsorption of the supernumerary stereocilia.\(^1\),\(^6\),\(^7\) Interestingly, MYO15, WHIRLIN, and BAIAP2L2 did not localize to stereocilia tips in Eps8$^{−/−}$ mice, suggesting EPS8 is critical for the proper transport and/or retention of these proteins at stereocilia tips. In Eps8$^{−/−}$ mice, the immature hair bundle structure causes a wide range of cellular dysfunctions, including a very small or absent resting MET current (Figure 5), possibly due to a reduced tension at the tip links connecting adjacent stereocilia that normally keep the MET channel partially open at rest.\(^7\) This resting depolarizing MET current normally drives the resting firing activity in the auditory fibers of the adult cochlea. However, the MET apparatus is fully functional before hearing onset in mice,\(^8\),\(^9\),\(^4\),\(^5\) and the resting MET current has been shown to be required for the normal maturation of the IHC biophysical properties\(^10\) and their afferent synapses.\(^5\) We also observed this in Eps8$^{−/−}$ mice (Figures 6 and 7). As such, EPS8 regulates the physiological and morphological maturation of IHCs and in its absence IHCs remain arrested at very early stages of development. All these morphological and physiological defects lead to profound deafness in Eps8$^{−/−}$ mice.

When exogenous EPS8 was delivered to Eps8$^{−/−}$ pups (P1–P2) via Anc80L65 in vivo, we found that some transduced apical-coil hair cells were able to develop normal bundles, including the reabsorption of the extra numeral stereociliary rows (Figure 2). These normal-looking hair bundles generated an MET current in pre-hearing hair cells that was indistinguishable from that of wild-type mice, and showed a normal distribution pattern of EPS8, MYO15, and WHIRLIN, including the presence of BAIAP2L2 at the shorter rows (Figures 3 and 4). The recovery of the MET current meant that some transduced IHCs from Eps8$^{−/−}$ mice were able to progress with their normal development and acquire the biophysical basolateral membrane characteristics and morphological features of fully mature sensory receptors (Figures 6 and 7).

In vivo delivery of AAV-Eps8 in early-postnatal Eps8-deficient mice restores the function of hair cells

To proveباحتة، إزالة الألوان من المكتبة، وتعويضها بـ EPS8 at P20. Adult cochleae were immunostained for EPS8 (red) and stereocilia were labeled with phalloidin (green). Note that transduced IHCs still show several rows of stereocilia, most of which remained short. Moreover, EPS8 is no longer localized at the tip of the taller stereocilia. (B–E) Confocal images using the same experimental conditions described in (A), but using antibodies against MYO15 (B and C) and WHIRLIN (D and E), in addition to EPS8. Note that the diffuse staining of all the proteins, similar to the labeling seen in basal-coil IHCs from Eps8$^{−/−}$ mice transduced with Anc80L65-EPS8 at P1–P2. Scale bar in the inset of (E), which also applies for (C), represents 1 μm.

Figure 8. Hair bundles in IHCs transduced with Anc80L65-Eps8 in the adult cochlea

(A) Confocal images of the hair bundles from IHCs of 3-month-old Eps8$^{−/−}$ mice transduced with Anc80L65-EPS8 at P20. Adult cochleae were immunostained for EPS8 (red) and stereocilia were labeled with phalloidin (green). Note that transduced IHCs still show several rows of stereocilia, most of which remained short. Moreover, EPS8 is no longer localized at the tip of the taller stereocilia. (B–E) Confocal images using the same experimental conditions described in (A), but using antibodies against MYO15 (B and C) and WHIRLIN (D and E), in addition to EPS8. Note that the diffuse staining of all the proteins, similar to the labeling seen in basal-coil IHCs from Eps8$^{−/−}$ mice transduced with Anc80L65-EPS8 at P1–P2. Scale bar in the inset of (E), which also applies for (C), represents 1 μm.
mice, with EPS8, MYO15, and WHIRLIN labeling the entire length of the stereocilia (Figure 8). Together, these results indicate a role for EPS8 in the proper targeting of these tip-complex proteins to stereocilia tips, which is perhaps counterintuitive given MYO15 is the motor protein responsible for transporting these proteins to stereocilia tips.

**AAV gene delivery as a therapy to restore the function of early-onset genes in the mouse cochlea**

More than 100 genes have now been linked to hereditary hearing impairment (https://hereditaryhearingloss.org/), most of which are expressed in the sensory hair cells of the mammalian cochlea. During the last decade or so, several AAV vectors have been used to transduce exogenous genes in vivo into dysfunctional hair cells of different mouse models of hereditary HL. Although the success of this approach was limited, at least initially, recent development in the viral capsids used for gene augmentation in vivo (Anc80L65, AAV-S, AAV9-PHP.B) has resulted in more efficient hair-cell transduction and functional recovery. In agreement with these previous reports, we found that Anc80L65 leads to a high transduction rate, at least in the apical coil of the cochlea, which was the region used for the ex vivo functional recovery experiments. It is therefore unlikely that a low infection efficiency of Anc80L65 is responsible for the very little or no functional recovery of low-frequency hearing in Eps8−/− mice injected with the exogenous EPS8. It is also worth noting that our surgery procedure did not cause any damage to the cochlea since wild-type mice subjected to the same procedure as Eps8−/− mice showed normal auditory functions at adult ages.

One possible explanation for the absence of hearing recovery is that, at the time we performed the AAV gene delivery (primarily P1), most of the hair cells are at a point in development such that they have either matured or accumulated damage beyond the point of repair using therapeutic gene delivery. This is because EPS8 exerts a key role in bundle growth from embryonic stages of development. In fact, the transduction of Anc80L65-Eps8 in hair cells from the basal-coil region of the cochlea of Eps8−/− mice, which are known to be approximately 2–3 days ahead in terms of development compared with apical cells, failed to rescue their immature-like hair bundles. Similar findings were also seen when attempting to correct mature hair cells from P19–P20 Eps8−/− mice. Evidence of a critical period for therapeutic gene delivery in the cochlea seems to be consistent with the published literature. Most of the successful functional recovery of mouse models for hereditary monogenic hearing impairment has involved genes that primarily contribute to the development and/or function of the postnatal mouse cochlea (e.g., Tmc1, Vglut3, Otof). Gene-based therapy targeting proteins that are directly involved in the initial growth of the stereociliary bundles of cochlear hair cells has mainly focused on harmonin, whirlin, and clarin-1, which lead to different forms of Usher syndromes when mutated (Usher 1C, Usher 2, and Usher 3A, respectively). Harmonin is present at the tip of the stereociliary bundle from about P0 onward, and AAV gene augmentation at P1 with Anc80L65-harmonin injected in the cochlea of a mouse model for Usher 1C led to a reasonable hearing rescue in the low-frequency (<16 Hz) but not high-frequency regions. Clarin-1 and whirlin are expressed in hair bundles just before birth, and AAV gene delivery leads to a very limited or no recovery of hearing in corresponding mouse models for Usher 3A (clarin-1, ABR threshold ~90 dB only at about 10 kHz) and Usher 2 (whirlin: ABR threshold >80 dB only at 8 kHz or deaf). We speculate that, for genes involved in the early stages of morphological and functional development of cochlear hair cells, there is a critical window for successful therapeutic gene delivery for treating hearing loss, and, in some cases (e.g., EPS8 and some Usher proteins), in utero delivery of genes via AAVs may be required as previously performed for other genes. Alternatively, it is possible that co-expression of other proteins or co-administration of small molecules that can expand this critical window will increase success in restoring hearing function.

**MATERIALS AND METHODS**

**Ethics statement**

The animal work was licensed by the UK Home Office under the Animals (Scientific Procedures) Act 1986 (PPL_PCC05E93) and was approved by the University of Sheffield Ethical Review Committee (180626_Mar). Eps8 knockout mice (Eps8−/−) on a C57BL/6N background were bred as either Eps8−/− × Eps8−/− or Eps8−/− × Eps8+/−. Both males and females where used for this study. For ex vivo experiments, mice were killed by cervical dislocation followed by decapitation. For in vivo ABRs and DPOAEs, mice were anesthetized using intraperitoneal injection of ketamine (100 mg/kg body weight, Fort Dodge Animal Health, Fort Dodge, USA) and xylazine (10 mg/kg, Rompun 2%, Bayer HealthCare, NY, USA). At the end of the in vivo recordings, mice were either culled by cervical dislocation or recovered from anesthesia with intraperitoneal injection of atipamezole (1 mg/kg). For in vivo gene delivery, mice were anesthetized with isoflurane (2.5%) under oxygenation (0.8%). Mice under recovery from anesthesia were returned to their cage, placed on a thermal mat, and monitored over the following 2–5 h.

**AAV production**

The original Eps8 cDNA was a gift from Giorgio Scita and Andrea Disanza. The EGFP-mouse Eps8-beta-globin-polyA coding cDNA was synthesized using PCR and was cloned into the pAAV vector (with cytomegalovirus [CMV] promoter) using the SacI and XbaI restriction enzyme sites. The FLAG-tag-Eps8 coding cDNA was synthesized using PCR and was cloned into the pAAV vector (with CMV promoter) using the EcoRI and HindIII restriction enzyme sites. All constructs have been verified by multiple restriction enzyme digestions and sequencing. AAV-Anc80L65.CMV.FLAG-mEps8, bGH (Anc80L65-Eps8) and AAV-Anc80L65.CMV.PI.eGFP.WPRE. bGH (Anc80L65-GFP) were produced by the Penn Vector Core at the University of Pennsylvania (USA). The pAAV-Anc80L65 plasmid was purchased from Addgene (92307).

**AAV gene delivery in vivo for both pups and adult mice**

The surgical protocol used for AAV injection into the cochlea of P1–P3 and adult mice was performed under anesthesia (see section “ethics statement”) and body temperature was maintained using a
heating mat. The right ear was accessed via an incision just below the pinna as previously described. When the RWM was identified, it was gently punctured with a borosilicate pipette. This was followed by the injection of the AAV into the cochlea (pressure controlled by mouth) of 1 µL of Anc80L65-GFP (2.58 × 10^{13} vg/mL) or Anc80L65-Eps8 (8.90 × 10^{12} vg/mL), which was the maximal titer achievable from the supplied AAV. Following the injection, the pipette was retracted from the RWM and the wound was closed with veterinarian glue.

**Tissue preparation for ex vivo recordings**

Patch clamp recordings were performed from hair cells within the 9–12 kHz region of the cochlear apical coil. Cochleae were dissected out in extracellular solution composed of (in mM) 135 NaCl, 5.8 KCl, 1.3 CaCl₂, 0.9 MgCl₂, 0.7 NaH₂PO₄, 5.6 D-glucose, and 10 HEPES-NaOH. Sodium pyruvate (2 mM), amino acids and vitamins were added from concentrates (Thermo Fisher Scientific, UK). The pH was adjusted to 7.48 with 1 M NaOH, and the final osmolality of the solution was measured (∼308 mmol kg⁻¹). The dissected apical coil was then transferred to a microscope chamber and immobilized via a nylon mesh attached to a stainless-steel ring. The chamber (volume 2 mL) was perfused from a peristaltic pump and mounted on the stage of an upright microscope (Olympus BX51, Japan) with Nomarski DIC optics. For electrophysiology, the hair bundle structure of the pinna was assessed using a 60× water immersion objective, and an additional 2× magnification prior to the eyepiece (15×). The microscope chamber was continuously perfused with extracellular solution by a peristaltic pump (Cole-Palmer, UK).

**Whole-cell electrophysiology**

Patch-clamp recordings were performed at room temperature (20°C ± 2°C) using an Optopatch amplifier (Cairn Research, UK) as previously described.²⁵ Patch pipettes were pulled from soda glass capillaries and had a typical resistance in extracellular solution of 2–3 MΩ. The patch pipette intracellular solution contained (in mM) 131 KCl, 3 MgCl₂, 1 EGTA-KOH, 5 Na₃ATP, 5 HEPES-KOH, and 10 Na-phosphocreatine (pH was adjusted with 1 M KOH to 7.28; 294 mmol kg⁻¹). Data acquisition was controlled by pClamp software using Digidata 1440A (Molecular Devices, USA). To reduce the electrode capacitance, patch electrodes were coated with surf wax (Mr. Zog’s SexWax, USA). Recordings were low-pass filtered at 2.5 kHz (eight-pole Bessel), sampled at 5 kHz, and stored on a computer for offline analysis (Clampfit, Molecular Devices; Origin 2020; OriginLab, USA). For voltage-clamp experiments, membrane potentials were corrected offline for the residual series resistance Rₑ after compensation (usually 80%) and the liquid junction potential (LJP) of −4 mV, which was measured between electrode and bath solutions. Holding currents were plotted as zero current to allow a better comparison between recordings. Voltage-clamp protocols are referred to a holding potential of −84 mV or −64 mV depending on the protocol used.

For MET current recordings, the bundles of hair cells were displaced using a fluid jet from a pipette driven by a 25-mm diameter piezoelectric disc. The pipette was pulled from borosilicate glass to a final overall length of 5.3–5.5 cm. The fluid jet pipette tip had a diameter of 8–10 µm and was positioned near the hair bundles to elicit a maximal MET current. Mechanical stimuli were applied as 50-Hz sinusoids (filtered at 1 kHz, eight-pole Bessel). Prior to the positioning of the fluid jet by the hair bundles, any steady-state pressure was removed by monitoring the movement of debris in front of the pipette. The use of the fluid jet allows for the efficient displacement of the hair bundles in both the excitatory and inhibitory directions, which is essential to perform reliable measurements of the resting open probability of the MET channels.

Real-time changes in membrane capacitance (ΔCₘ) were measured as previously described.⁷⁷ Briefly, ΔCₘ was measured using a 4-kHz sine wave (13 mV RMS) applied to hair cells from a holding potential of −81 mV and was interrupted for the duration of the voltage step. The capacitance signal from the Optopatch was amplified (×50), filtered at 250 Hz, sampled at 5 kHz, and measured by averaging the Cₘ traces after the voltage step (around 200 ms) and subtracting from pre-pulse baseline. ΔCₘ was recorded in the presence of K⁺ channel blockers TEA (30 mM), 4-AP (15 mM), and linopirdine (80 µM) in the extracellular solution.

**ABRs**

Following the onset of anesthesia (see section “ethics statement”) and the loss of the retraction reflex with a toe pinch, mice were placed in a soundproof chamber (MAC-3 acoustic chamber, IAC Acoustic, UK). Male and female mice were placed on a heated mat (37°C) with the animal’s pinna being positioned at a distance of 10 cm from the loudspeaker. Two subdermal electrodes were placed under the skin behind the pinna of each ear (reference and ground electrode), and one electrode half-way between the two pinna on the vertex of the cranium (active electrode) as previously described.⁸⁰ Sound stimuli were delivered to the mouse ear by a loudspeaker (MFI-S, Multi Field Speaker, Tucker-Davis Technologies, USA), which was calibrated with a low-noise microphone probe system (ER10B+, Etymotic, USA). Experiments were performed using customized software⁹¹ driving an RZ6 auditory processor (Tucker-Davis Technologies). ABR thresholds, which were delivered as white-noise clicks and pure-tone stimuli of frequencies at 6, 12, 18, 24, 30, 36, and 42 kHz, were defined as the lowest sound level where any recognizable feature of the waveform was visible. Stimulus sound pressure levels were up to 95 dB SPL, were presented in steps of 5-dB SPL (averaged over 256 repetitions). Tone bursts were 5 ms in duration with a 1-ms on/off ramp time presented at a rate of 42.6/s.

**DPOAEs**

DPOAEs were used to assess OHC function in vivo by the synchronous presentation of two stimulus tones (primaries f₁ and f₂). DPOAEs were recorded at 2f₁−f₂ in response to primary tones f₁ and f₂, where f₂/f₁ = 1.2. The f₂ level (L₂) was set from 20 to 80 dB (maximum level set for our system) in 10-dB increments, and the f₁ level (L₁) was set equal to L₂. Frequency pairs of tones between f₂ = 6.5 kHz and f₁ = 26.3 kHz were presented directly into the left ear.
canal of mice by means of a coupler, which was connected to two calibrated loudspeakers using 3-cm plastic tubes (MF1-S, Multi Field Speaker, Tucker-Davis Technologies, USA).

Recordings were performed in a soundproof chamber (MAC-3 Acoustic Chamber, IAC Acoustic, UK) and the emission signals were recorded by a low-noise microphone (ER10B+, Etymotic Research, USA) connected to the coupler mentioned above. Experiments were performed using BioSigRZ software driving an RZ6 auditory processor (Tucker-Davis Technologies). The DPOAE thresholds were defined by the minimal sound level where the DPOAEs were above the standard deviation (SD) of the noise. The determined DPOAE thresholds were plotted against the geometric mean frequency of f1 and f2. Stimulus sound pressure levels were up to 80 dB SPL, presented in steps of 10 dB. The response signal was averaged over 500 repetitions.

**SEM**

For SEM, the dissected mouse cochleae were initially fixed by a very gentle intralabyrinthine perfusion using a 100-µL pipette tip through the round window. The fixative contained 2.5% glutaraldehyde in 0.1 M sodium cacodylate buffer plus 2 mM CaCl2 (pH 7.4). Following perfusion, the cochleae were immersed in the above fixative for 2 h at room temperature. After the fixation, the organ of Corti was exposed by removing the bone from the apical coil, and the cochlea, then incubated in solutions of saturated aqueous thiocarbohydrazide (20 min) alternating with 1% osmium tetroxide in cacodylate buffer (20 min) twice (the OTOTO technique). The cochleae were then dehydrated through an ethanol series and critical point dried using CO2 as the transitional fluid (Leica EM CPD300) and mounted on specimen stubs using conductive silver paint (Agar Scientific, Stansted, UK). The apical coil of the organ of Corti was examined at 10 kV using a Tescan Vega3 LMU scanning electron microscope.

**Immunofluorescence microscopy**

The majority of this work was performed by fixing the inner ear with 4% paraformaldehyde in phosphate-buffered saline (PBS, pH 7.4) for 20 min at room temperature. Cochleae were washed three times in PBS for 10 min and fine dissected. Samples were incubated in PBS supplemented with 5% normal goat or horse serum and 0.5% Triton X-100 for 1 h at room temperature. The samples were immunolabelled with primary antibodies overnight at 37°C, washed three times with PBS, and incubated with the secondary antibodies for 1 h at 37°C. Antibodies were prepared in 1% serum and 0.5% Triton X-100 in PBS. F-actin was stained with Texas Red-X phalloidin (1:400, Thermo Fisher, T7471) or Alexa Fluor 488 phalloidin (1:400, Thermo Fisher, A12379) in the secondary antibody solution. Primary antibodies were mouse-IgG1 anti-Eps8 (1:1,000, BD Biosciences, 610143), rabbit-IgG anti-BAILAP2L2 (1:500, Atlas Antibodies, HPA003043), mouse-IgG1 anti-BK channel (1:200, NeuroMab, 75–408), rabbit-IgG anti-MYO7a (1:500, Proteus Biosciences, 25–6790), anti-MYO15 and anti-WHIRLIN (1:400 and 1:200, respectively, gift from Dr. Thomas Friedman, NIH, USA), mouse anti-CtBP2 (1:200, Biosciences, #612044), mouse anti-GluR2 (1:200, Millipore, MAB397), and rabbit-anti-FLAG (1:200, Cell Signaling, #14793). Secondary antibodies were species appropriate Alexa Fluor secondary antibodies. Samples were mounted in VECTASHIELD (H-1000). The images from the apical cochlear region (8–12 kHz) were captured with Nikon A1 confocal microscope equipped with Nikon CFI Plan Apo 60 x oil objective or a Zeiss LSM 880 Airyscan equipped with Plan-Apochromat 63 x Oil DIC M27 objective for super-resolution images of hair bundles. Both microscopes are part of the Wolfson Light Microscope Facility at the University of Sheffield. Image stacks were processed with Fiji Imagej software (https://imagej.net/Fiji).

For the immunostaining to investigate OHC insertion into the TM, the inner ear was fixed with 4% paraformaldehyde in PBS (pH 7.4) for 1 h at room temperature. Cochleae were then decalciﬁed in 0.5 M EDTA for 3 days at 4°C, washed in PBS, and dissected as previously described. These were sub-dissected by iterative trimming with a razor blade, and the tectorial membrane (TM) segments carefully peeled off from the tissue pieces with a pair of ﬁne forceps. These TM pieces were transferred into dishes containing PBS with 10% horse serum and 0.1% Triton X-100 (PBS/HS/TX) to which FITC-conjugated wheat-germ agglutinin (FL-1021, Vector Laboratories) was added at a dilution of 1:1,000. These were incubated overnight at 4°C with gentle agitation, washed in PBS containing 0.1% TX (PBS/TX), and mounted on glass slides in Vectashield with the underside of the TM facing upward. Pieces of organ of Corti with the TMs removed were transferred to PBS/HS/TX for 1 h, followed by PBS/HS/TX containing rabbit anti-stereocilin (1:150, gift from Dr. Saaid Safieddine, Institute Pasteur, France) and 1:1,000 Phalloidin-Atto 488 (49049, Sigma-Aldrich). The tissue was then incubated overnight at 4°C with gentle agitation, washed three times in PBS/TX and stained for 3 h in Alexa 555 donkey anti-rabbit (A31572, Invitrogen), washed three times in PBS/TX, and mounted in Vectashield (Vector Laboratories). Super-resolution scans of these preparations were obtained on a Zeiss LSM 880 with an Airyscan detector, using a 63×1.4 NA oil-immersion objective. Z-projections were created using Imaris Viewer 9.8.0.

**Measurements of stereocilia dimensions**

Image analysis and any further processing for visualization was performed using Fiji. Airyscan processed z stacks were resliced along the y or x axis using the Reslice feature. The tallest stereocilia of each bundle was identified and measured in the phalloidin channel using the Segmented Line tool of ImageJ. Bundles were considered EPS8 positive if EPS8 puncta were visible at the tips of stereocilia within the bundle. Non-transduced hair cells within the same field were used to measure EPS8-negative stereocilia height. For these measurements, IHCs were selected based on the presence of a tip-localized EPS8 signal, regardless of the amount of improved bundle height apparent prior to quantification. Only the tallest stereocilia of each IHC was measured.

**Statistical analysis**

Statistical comparisons of means were made by Student’s two-tailed t test or, for multiple comparisons, analysis of variance (one-way or
two-way ANOVA followed by a suitable post test); p < 0.05 was selected as the criterion for statistical significance. Only mean values with a similar variance between groups were compared. Average values are quoted in text and figures as mean ± SD.

Data availability statement
The data that support the findings of this study are available from the corresponding authors upon reasonable request.

SUPPLEMENTAL INFORMATION
Supplemental information can be found online at https://doi.org/10.1016/j.omtm.2022.07.012.

ACKNOWLEDGMENTS
The authors thank Saaid Saffieddine (Institut Pasteur, France) and Omar Akl (University of California San Francisco, USA) for helping us establishing the surgical procedure required to deliver the AAV into the cochlea, Michelle Bird (University of Sheffield) for her assistance with the mouse husbandry, and Catherine Gennery and Laila Moushatq-Kheradmandi for their genotyping work. We also thank Saaid Saffieddine for the anti-stereocillin antibody; Thomas Friedman, Inna Belyantseva, and Jonathan Bird for the anti-MY015 and anti-Whirlin antibodies; and Giorgio Scita and Andrea Disanza (Istituto FIRC di Oncologia Molecolare) for the Eps8 cDNA. The Eps8−/− mice were obtained from Pier Paolo Di Fiore (University of Milano).

This work was supported by the BBSRC (BB/S006257/1) to M.R.B. and W.M.; BBSRC (BB/T016337/1) to R.J.G. and G.P.R.; and WaItt Foundation, the Grohne Foundation, Core Grant application NCI CCGS (CA014195), NIH (R21 DC018237), NSF NeuroNex Award (2014862), and the Chan-Zuckerberg Initiative Imaging Scientist Award to U.M. The gene therapy suite used for AAV injections at Sheffield has been funded by the Dudley and Geoffrey Cox Charitable Trust fund to W.M. For the purpose of open access, the author has applied a CC BY public copyright license to any author accepted manuscript version arising from this submission.

AUTHOR CONTRIBUTIONS
All authors helped with the collection and analysis of the data. W.M. and U.M. conceived and coordinated the study. J.-Y.J. and W.M. wrote the paper. All authors approved the final version of the manuscript. All authors agree to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. All persons designated as authors qualify for authorship, and all those who qualify for authorship are listed.

DECLARATION OF INTERESTS
The authors declare no competing interests.

REFERENCES
1. Shearer, A.E., Hildebrand, M.S., and Smith, R.J.H. (2017). Hereditary hearing loss and deafness overview. In GeneReviews® [Internet] (University of Washington, USA), https://www.ncbi.nlm.nih.gov/books/NBK1434/.
2. Delmaghani, S., and El-Amraoui, A. (2020). Inner ear gene therapies take off: current promises and future challenges. J. Clin. Med. 9, 2309.
3. Van Camp, G., and Smith, R.J.H. (2017). The hereditary hearing loss homepage. https://hereditaryhearingloss.org.
4. Chhabra, E.S., and Higgs, H.N. (2007). The many faces of actin: matching assembly factors with cellular structures. Nat. Cell Biol. 9, 1110–1112.
5. Barr-Gillespie, P.G. (2015). Assembly of hair bundles, an amazing problem for cell biology. Mol. Biol. Cell 26, 2727–2732.
6. Houdusse, A., and Titus, M.A. (2021). The many roles of myosins in filopodia, microvilli and stereocilia. Curr. Biol. 31, R586–R602.
7. Fettplace, R., and Kim, K.X. (2014). The physiology of mechanoelectrical transduction channels in hearing. Physiol. Rev. 94, 951–986.
8. Fort, M., Fettplace, R., Nam, J.-H., and Ricci, A.J. (2009). Localization of inner hair cell mechanotransducer channels using high-speed calcium imaging. Nat. Neurosci. 12, 553–558.
9. Tilney, L.G., Tilney, M.S., and DeRosier, D.J. (1992). Actin filaments, stereocilia, and hair cells: how cells count and measure. Annu. Rev. Cell Biol. 8, 257–274.
10. Bartles, J.R. (2000). Parallel actin bundles and their multiple actin-bundling proteins. Curr. Opin. Cell Biol. 12, 72–78.
11. Goodyear, R.J., Marcotti, W., Kros, C.J., and Richardson, G.P. (2005). Development and properties of stereociliary link types in hair cells of the mouse cochlea. J. Comp. Neurol. 485, 75–85.
12. Vélez-Ortega, A.C., and Frolenkov, G.I. (2019). Building and repairing the stereocilia cytoskeleton in mammalian auditory hair cells. Hear. Res. 376, 47–57.
13. Roth, B., and Bruns, V. (1992). Postnatal development of the rat organ of Corti. II. Hair cell receptors and their supporting elements. Anat. Embryol. (Berl) 185, 571–581.
14. Kaltenbach, J.A., Falzarano, P.R., and Simpson, T.H. (1994). Postnatal development of the hamster cochlea. J. Growth and differentiation of stereocilia bundles. J. Comp. Neurol. 350, 187–198.
15. Zine, A., and Romand, R. (1996). Development of the auditory receptors of the rat: a SEM study. Brain Res. 721, 49–58.
16. Manor, U., Disanza, A., Grati, M., Andrade, L., Lin, H., Di Fiore, P.P., Scita, G., and Kachar, B. (2011). Regulation of stereocilia length by myosin XVa and whirin depends on the actin-regulatory protein Eps8. Curr. Biol. 21, 167–172.
17. Zampini, V., Rüttiger, L., Johnson, S.L., Franz, C., Furness, D.N., Waldhaus, J., Xiong, H., Hackney, C.M., Holley, M.C., Officer, N., et al. (2011). Eps8 regulates hair bundle length and functional maturation of mammalian auditory hair cells. PLoS Biol. 9, e1001048.
18. Furness, D.N., Johnson, S.L., Manor, U., Rüttiger, L., Tocchetti, A., Officer, N., Olt, J., Goodyear, R.J., Vijayakumar, S., Dai, Y., et al. (2013). Progressive hearing loss and gradual deterioration of sensory hair bundles in the ears of mice lacking the actin-binding protein EphSl2. Proc. Natl. Acad. Sci. USA 110, 13898–13903.
19. Croce, A., Cassata, G., Disanza, A., Gagliani, M.C., Tacchetti, C., Malabarba, M.G., Carlier, M.-F., Scita, G., Baumeister, R., and Di Fiore, P.P. (2004). A novel actin barred-end-capping activity in EPS-8 regulates apical morphogenesis in intestinal villi and stereocilia. Curr. Biol. 14, 986–998.
20. Menna, E., Disanza, A., Cagnoli, C., Schenk, U., Gelosoimo, G., Fritelli, E., Hertzog, M., Officer, N., Savallachis, C., Krenchkamp, H.-J., et al. (2009). Eps8 regulates aminotriphosphatidyl in hippocampal neurons in response to brain-derived neurotrophic factor (BDNF). PLoS Biol. 7, e1000138.
21. Belyantsev, I.A., Boger, E.T., and Friedman, T.B. (2003). Myosin XVa localizes to the tips of inner ear sensory cell stereocilia and is essential for staircase formation of the hair bundle. Proc. Natl. Acad. Sci. USA 100, 13958–13963.
22. Mbure, P., Mustapha, M., Varella, A., Weil, D., El-Amraoui, A., Holme, R.H., Rump, A., Hardisty, R.E., Blanchard, S., Coumara, R.S., et al. (2003). Defects in whirin, a PDZ domain molecule involved in stereocilia elongation, cause deafness in the whirler mouse and families with DFNB31. Nat. Genet. 34, 421–428.
23. Tadeney, A.L.D., Akturk, A., Devanney, N., Mathur, P.D., Clark, A.M., Yang, J., and Tarchini, B. (2019). GPSM2-GNAI specifies the tallest stereocilia and defines hair bundle row identity. Curr. Biol. 29, 921–934.e4.
24. Tarchini, B., Tadener, A.L.D., Devanney, N., and Cayouette, M. (2016). A link between planar polarity and staircase-like bundle architecture in hair cells. Development 143, 3926–3932.

25. Carlton, A.J., Halford, J., Underhill, A., Jung, J.Y., Avenarius, M.R., Gilbert, M.L., Ceriani, F., Ebisne, K., Brown, S.D.M., Bowl, M.R., et al. (2021). Loss of Baspa22 destabilizes the transducing stereocilia of cochlear hair cells and leads to deafness. J. Physiol. 599, 1173–1198.

26. Behlouli, A., Bonnet, C., Abdi, S., Bouaita, A., Lelli, A., Hardelin, J.-P., Schietroma, C., Rous, Y., Louha, M., Cheknane, A., et al. (2014). EPS8, encoding an actin-binding protein of cochlear hair cell stereocilia, is a new causal gene for autosomal recessive profound deafness. Orphanet J. Rare Dis. 9, 55.

27. Yu, S., Chen, W.X., Zhang, Y.F., Ni, Y., Lu, P., Wang, B., Wang, Y., Wu, B., Ni, Q., Wang, H., and Xu, Z.M. (2021). Apparent homozygosity for a novel splicing variant in EPS8 causes congenital profound hearing loss. Eur. J. Med. Genet. 64, 104362.

28. Landegger, L.D., Pan, B., Askew, C., Wassmer, S.L., Gluck, S.D., Galvin, A., Taylor, R., Forge, A., Stankovic, K.M., Holt, J.R., and Vandenbergh, L.H. (2017). A synthetic AAV vector enables safe and efficient gene transfer to the mammalian inner ear. Nat. Biotechnol. 35, 280–284.

29. Kujawa, S.G., and Liberman, M.C. (2009). Adding insult to injury: cochlear nerve degeneration after ‘temporary’ noise-induced hearing loss. J. Neurosci. 29, 14077–14085.

30. Shera, C.A., and Guinan, J.J., Jr. (1999). Evoked otocoustic emissions arise by two fundamentally different mechanisms: a taxonomy for mammalian OAEs. J. Acoust. Soc. Am. 105, 782–798.

31. Belyantseva, I.A., Boger, E.T., Naz, S., Frolenkov, G.L., Sellers, J.R., Ahmed, Z.M., Griffith, A.J., and Friedman, T.B. (2005). Myosin-XVα is required for tip localization of whorl and differential elongation of hair-cell stereocilia. Nat. Cell Biol. 7, 148–156.

32. Delprat, B., Michel, V., Goodyear, R.J., and Richardson, G. (2016). A link between planar polarity and staircase-like bundle architecture in hair cells. J. Physiol. 599, 1515–1531.

33. Mano, U., and Kachar, B. (2008). Dynamic length regulation of sensory stereocilia. Semin. Cell Dev. Biol. 19, 502–510.

34. Johnson, S.L., Kennedy, H.J., Holley, M.C., and Marcotti, W. (2012). The resting transducer current drives spontaneous activity in prehearing mammalian cochlear inner hair cells. J. Neurosci. 32, 10479–10483.

35. Jeng, J.Y., Haraszti, C., Carlton, A.J., Corns, L.F., Marchetta, P., Johnson, S.L., Goodyear, R.I., Legan, K.P., Rüttiger, L., Richardson, G.P., and Marcotti, W. (2021). MET currents and otocoustic emissions from mice with a detached tectorial membrane indicate the extracellular matrix regulates Ca2+ near stereocilia. J. Physiol. 599, 2015–2036.

36. Géléoc, G.G.S., and El-Amraoui, A. (2003). Noise-induced cochlear neuropathy is selective for fibers with low spontaneous rates. J. Neurophysiol. 110, 577–586.

37. Liberman, L.D., Wang, H., and Liberman, M.C. (2011). Opposing gradients of ribbon size and AMPA receptor expression underlie sensitivity differences among cochlear-nerve/hair-cell synapses. J. Neurosci. 31, 801–808.

38. Lee, J., Kawai, K., Holt, J.R., and Géléoc, G.S. (2021). Sensory transduction is required for normal development and maturation of cochlear inner hair cell synapses. Elife 10, e69433.

39. Goodyear, R.J., and Richardson, G.P. (2018). Structure, function, and development of the tectorial membrane: an extracellular matrix essential for hearing. Curr. Top. Dev. Biol. 110, 217–244.

40. Manor, U., and Kachar, B. (2008). Dynamic length regulation of sensory stereocilia. Semin. Cell Dev. Biol. 19, 502–510.

41. Johnson, S.L., Kennedy, H.J., Holley, M.C., and Marcotti, W. (2012). The resting transducer current drives spontaneous activity in prehearing mammalian cochlear inner hair cells. J. Neurosci. 32, 10479–10483.

42. Jeng, J.Y., Haraszti, C., Carlton, A.J., Corns, L.F., Marchetta, P., Johnson, S.L., Goodyear, R.I., Legan, K.P., Rüttiger, L., Richardson, G.P., and Marcotti, W. (2021). MET currents and otocoustic emissions from mice with a detached tectorial membrane indicate the extracellular matrix regulates Ca2+ near stereocilia. J. Physiol. 599, 2015–2036.

43. Géléoc, G.G.S., and El-Amraoui, A. (2020). Disease mechanisms and gene therapy for neurofibromatosis type II. Trends Genet. 36, 757–768.

44. Liberman, L.D., Wang, H., and Liberman, M.C. (2011). Opposing gradients of ribbon size and AMPA receptor expression underlie sensitivity differences among cochlear-nerve/hair-cell synapses. J. Neurosci. 31, 801–808.

45. Lee, J., Kawai, K., Holt, J.R., and Géléoc, G.S. (2021). Sensory transduction is required for normal development and maturation of cochlear inner hair cell synapses. Elife 10, e69433.

46. Goodyear, R.J., and Richardson, G.P. (2018). Structure, function, and development of the tectorial membrane: an extracellular matrix essential for hearing. Curr. Top. Dev. Biol. 110, 217–244.

47. Manor, U., and Kachar, B. (2008). Dynamic length regulation of sensory stereocilia. Semin. Cell Dev. Biol. 19, 502–510.

48. Johnson, S.L., Kennedy, H.J., Holley, M.C., and Marcotti, W. (2012). The resting transducer current drives spontaneous activity in prehearing mammalian cochlear inner hair cells. J. Neurosci. 32, 10479–10483.

49. Jeng, J.Y., Haraszti, C., Carlton, A.J., Corns, L.F., Marchetta, P., Johnson, S.L., Goodyear, R.I., Legan, K.P., Rüttiger, L., Richardson, G.P., and Marcotti, W. (2021). MET currents and otocoustic emissions from mice with a detached tectorial membrane indicate the extracellular matrix regulates Ca2+ near stereocilia. J. Physiol. 599, 2015–2036.

50. Géléoc, G.G.S., and El-Amraoui, A. (2020). Disease mechanisms and gene therapy for neurofibromatosis type II. Trends Genet. 36, 757–768.

51. Liberman, L.D., Wang, H., and Liberman, M.C. (2011). Opposing gradients of ribbon size and AMPA receptor expression underlie sensitivity differences among cochlear-nerve/hair-cell synapses. J. Neurosci. 31, 801–808.

52. Lee, J., Kawai, K., Holt, J.R., and Géléoc, G.S. (2021). Sensory transduction is required for normal development and maturation of cochlear inner hair cell synapses. Elife 10, e69433.

53. Goodyear, R.J., and Richardson, G.P. (2018). Structure, function, and development of the tectorial membrane: an extracellular matrix essential for hearing. Curr. Top. Dev. Biol. 110, 217–244.

54. Manor, U., and Kachar, B. (2008). Dynamic length regulation of sensory stereocilia. Semin. Cell Dev. Biol. 19, 502–510.

55. Lee, J., Kawai, K., Holt, J.R., and Géléoc, G.S. (2021). Sensory transduction is required for normal development and maturation of cochlear inner hair cell synapses. Elife 10, e69433.
63. Akil, O., Dyka, F., Calvet, C., Emptor, A., Lah lou, G., Nouaille, S., Bouet de Monvel, J., Hardelin, J.P., Hauswirth, W.W., Avan, P., et al. (2019). Dual AAV-mediated gene therapy restores hearing in a DFNB9 mouse model. Proc. Natl. Acad. Sci. USA 116, 4496–4501.

64. Al-Moyed, H., Cepeda, A.P., Jung, S., Moser, T., Kügler, S., and Reisinger, E. (2019). A dual-AAV approach restores fast exocytosis and partially rescues auditory function in deaf otoferlin knock-out mice. EMBO Mol. Med. 11, e9396.

65. El-Amraoui, A., and Petit, C. (2005). Usher I syndrome: unravelling the mechanisms that underlie the cohesion of the growing hair bundle in inner ear sensory cells. J. Cell Sci. 118, 4593–4603.

66. Mathur, P., and Yang, J. (2015). Usher syndrome: hearing loss, retinal degeneration and associated abnormalities. Biochim. Biophys. Acta 1852, 406–420.

67. Boëda, B., El-Amraoui, A., Bahloul, A., Goodyear, R., Daviet, L., Blanchard, S., Dulon, D., Papal, S., Patni, P., Cortese, M., Vincent, P.F., Tertrais, M., Emptoz, A., Zallocchi, M., Meehan, D.T., Delimont, D., Askew, C., Garige, S., Gratton, M.A., Ingham, N.J., Pearson, S., and Steel, K.P. (2011). Using the auditory brainstem response (ABR) to determine sensitivity of hearing in mutant mice. Curr. Protoc. Mouse Biol. 24, 23, 2551–2791.

68. Pan, B., Askew, C., Galvin, A., Heman-Ackah, S., Asai, Y., Indzhlykulian, A.A., Jodelka, F.M., Hastings, M.L., Lentz, J.J., Vandenberghe, L.H., et al. (2017). Gene therapy restores auditory and vestibular function in a mouse model of Usher syndrome type 1c. Nat. Biotechnol. 35, 264–272.

69. Zallocchi, M., Meehan, D.T., Delmont, D., Askew, C., Garige, S., Gratton, M.A., Rothermund-Franklin, C.A., and Cosgrove, D. (2009). Localization and expression of clarin-1, the Clrn1 gene product, in auditory hair cells and photoreceptors. Hear. Res. 255, 109–120.

70. Dulon, D., Papal, S., Patni, P., Cortese, M., Vincent, P.F., Tertrais, M., Emptor, A., Tlili, A., Bouleu, Y., Michel, V., et al. (2018). Clarin-1 gene transfer rescues auditory synaptopathy in model of Usher syndrome. J. Clin. Invest. 128, 3382–3401.

71. Isgirg, K., Shteamer, J.W., Belvantsvea, L.A., Drummond, M.C., Fitzgerald, T.S., Vijiayakumar, S., Jones, S.M., Griffith, A.J., Friedman, T.B., Cunningham, L.L., and Chien, W.W. (2017). Gene therapy restores balance and auditory functions in a mouse model of usher syndrome. Mol. Ther. 25, 780–791.

72. Chien, W.W., Isgirg, K., Roy, S., Belvantsvea, L.A., Drummond, M.C., May, I.A., Fitzgerald, T.S., Friedman, T.B., and Cunningham, L.L. (2016). Gene therapy restores hair cell stereocilia morphology in inner ears of deaf whirler mice. Mol. Ther. 24, 17–25.

73. Bedrosian, J.C., Gratton, M.A., Brigande, J.V., Tang, W., Landau, J., and Bennett, J. (2006). In vivo delivery of recombinant viruses to the fetal murine cochlea: transduction characteristics and long-term effects on auditory function. Mol. Ther. 14, 328–335.

74. Hu, C.J., Lu, Y.C., Tsai, Y.H., Cheng, H.Y., Takeda, H., Huang, C.Y., Xiao, R., Hsu, C.J., Tsai, J.W., Vandenberghe, L.H., et al. (2020). Efficient in utero gene transfer to the mammalian inner ears by the synthetic adeno-associated viral vector Anc80L65. Mol. Ther. Methods Clin. Dev. 18, 493–500.

75. Duan, M., Vernial, F., Spencer, N., and Mezzina, M. (2004). Treatment of peripheral sensorineural hearing loss: gene therapy. Gene Ther. 11, SS1–SS6.

76. Müller, M., von Hörnstein, K., Hoss, S., and Smolders, J.W.T. (2005). A physiological place-frequency map of the cochlea in the CBA/J mouse. Hear. Res. 282, 63–73.

77. Zallocchi, M., Jody, A., Belvantsvea, L.A., Goodyear, R., Daviet, L., Blanchard, S., Chien, W.W., Isgrg, K., Shteamer, J.W., Belyantseva, I.A., Drummond, M.C., Fitzgerald, T.S., Friedman, T.B., Cunningham, L.L., and Chien, W.W. (2017). Gene therapy restores auditory and vestibular function in a mouse model of Usher syndrome type 1c. Nat. Biotechnol. 35, 264–272.

78. Chien, W.W. (2017). Gene therapy restores balance and auditory functions in a mouse model of Usher syndrome type 1c. Nat. Biotechnol. 35, 264–272.

79. Johnson, S.L., Olt, J., Cho, S., von Gersdorff, H., and Marcotti, W. (2017). The coupling between Ca2+ channels and the exocytotic Ca2+ sensor at hair cell ribbon synapses varies tonotopically along the mature cochlea. J. Neurosci. 37, 2471–2484.

80. Ingham, N.J., Pearson, S., and Steel, K.P. (2011). Using the auditory brainstem response (ABR) to determine sensitivity of hearing in mutant mice. Curr. Protoc. Mouse Biol. 1, 279–287.

81. Furness, D.N., and Hackney, C.M. (1986). High-resolution scanning-electron microscopy of stereocilia using the osmium-thiocarbohydrazide coating technique. Hear. Res. 21, 243–249.

82. Schindelin, J., Arganda-Carreras, I., Frise, E., Kaynig, V., Longair, M., Pietzsch, T., Preibisch, S., Rueden, C., Saalfeld, S., Schmid, B., et al. (2012). Fiji: an open-source platform for biological-image analysis. Nat. Methods 9, 676–682.

83. Legan, P.K., Goodyear, R.J., Morín, M., Mencia, A., Pollard, H., Olavarrieta, L., Korchagina, J., Modamio-Hoybjor, S., Mayo, F., Moreno, F., et al. (2014). Three Usher syndrome gene products: localized, expressed, and associated abnormalities. Biochim. Biophys. Acta 1852, 406–420.
Supplemental information

AAV-mediated rescue of \textit{Eps8} expression \textit{in vivo}
restores hair-cell function in a mouse
model of recessive deafness

Jing-Yi Jeng, Adam J. Carlton, Richard J. Goodyear, Colbie Chinowsky, Federico Ceriani, Stuart L. Johnson, Tsung-Chang Sung, Yelena Dayn, Guy P. Richardson, Michael R. Bowl, Steve D.M. Brown, Uri Manor, and Walter Marcotti
Figure S1. Hair bundle morphology in IHCs from $Eps8^{−/−}$ mice transduced with exogenous Eps8.

(A) Scanning electron micrographs (SEM) showing examples of the stereociliary bundles of apical-coil IHCs in 5 adult $Eps8^{−/−}$ mice that were transduced with Anc80L65-Eps8 in vivo at P1-P2. (B) SEM images of the IHC hair bundles at a higher magnification from one the images in Figure 2C (second from the bottom). Arrowheads: indicate examples of hair bundles that show very little or no recovery. Arrows: highlight examples of hair bundles that are considered to have recovered their normal staircase structure with 3 rows of stereocilia.
Figure S2

Figure S2. Hair bundle morphology in OHCS from $Eps8^{-/-}$ mice transduced with exogenous $Eps8$.

(A,B) SEM showing examples of the stereociliary bundles of apical-coil IHCs in 2 adult $Eps8^{-/-}$ mice that were transduced with Anc80L65-Eps8 in vivo at P1-P2. Arrowheads: indicate hair bundles that show very little recovery. Arrow: highlight a hair bundle that was considered to have recovered its normal staircase structure with 3 rows of stereocilia.
Figure S3: Anc80L65-Eps8 do not rescue hearing in Eps8<sup>−/−</sup> mice.

(A) ABR thresholds for frequency-specific pure tone stimulation from 3 kHz to 42 kHz recorded from Eps8<sup>−/−</sup> mice injected at P1-P2 with Anc80L65-Eps8 (P21-P35). The number of mice tested is shown. The dashed line represents the upper threshold limit of our system, 95 dB. Single animal recordings are plotted as faded lines. (C) DPOAE thresholds measured from a subset or all the mice also used in panel (A). The frequency range tested was between 6 kHz and 24 kHz. Dashed line: upper threshold limit of the system, 80 dB. Single animal recordings are shown as faded lines. Data are plotted as means ± SD.
Figure S4. Rescue of IHC voltage responses in Eps8<sup>−/−</sup> mice injected with Anc80L65-Eps8

(A-C) IHC voltage responses elicited by applying a series of depolarizing current injections (10 pA or 100 pA increments), from their respective membrane potentials, from Eps8<sup>+/+</sup> (P19: A), Eps8<sup>−/−</sup> (P20: B) and Eps8<sup>−/−</sup> injected with Anc80L65-Eps8 (P22, C) mice. Note that while Eps8<sup>+/+</sup> and rescued Eps8<sup>−/−</sup> mice exhibited normal graded and sustained responses to current injections, Eps8<sup>−/−</sup> retained the characteristic spiking behaviour that is normally only present during in pre-hearing stages. (D) Average IHC resting membrane potential (V<sub>m</sub>) was not significantly different between the different mouse models (P = 0.3487, one-way ANOVA). Single-cell V<sub>m</sub> values (open symbols) are plotted behind the average data. Number of IHCs investigated is shown above the average data points. Data are plotted as means ± SD.
Figure S5. OHC stereocilia from rescued $Eps8^{-/-}$ mice fail to attach to the tectorial membrane

(A-C) OHCs from whole-mount preparations of adult wild-type (A), $Eps8^{-/-}$ (B) and $Eps8^{-/-}$ rescued (C) cochleae double-labelled with an anti-stereocilin antibody (magenta) and phalloidin (green). Images are generated from 0.6 micron Z-projections from super-resolution confocal scans encompassing the upper regions of the stereocilia. Strongest stereocilin label is seen at the tips of the tallest row of stereocilia in the wild-type (A, arrows), whilst in the shortened hair bundles of the $Eps8^{-/-}$ stereocilin labelling is only present between the stereocilia (B, arrows) and not at the tips of the stereocilia that are expected to adhere to tectorial membrane (TM). In the $Eps8^{-/-}$ mice rescued with Anc80L65-Eps8, although hair bundles are observed with the normal staircase pattern of stereocilia rows, stereocilin is absent from the tips of the tallest row and is only observed between the stereocilia (C, arrows). (D-F) Airyscan confocal Z-projections (1.4 microns in depth) from mid-coil regions of the TM stained with FITC-conjugated wheat-germ agglutinin. Each projection is from the underside of the TM, in the region where the imprints from OHCs of row 1 would be present. Clear imprints are readily visible in the wild-type (arrows) but none are observed in the $Eps8^{-/-}$ (E) or $Eps8^{-/-}$ Anc80L65-Eps8 rescued (F) cochlea.
Figure S6

Figure S6. Stereocilia from Eps8<sup>-/-</sup> mice do not express the characteristic bundle proteins
(A-D) Confocal images of the hair bundles from apical-coil (A-C) and basal-coil (D) IHCs of >P20 Eps8<sup>-/-</sup> mice immunostained for EPS8 (red) together with MYO15 (A), WHIRLIN (B,D) and BAIAPL2 (C) (blue). Stereocilia are labelled with phalloidin (white). None of the above proteins is expressed at the stereocilia.