Reciprocal and dynamic polarization of planar cell polarity core components and myosin

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Abstract The Ciona notochord displays planar cell polarity (PCP), with anterior localization of Prickle (Pk) and Strabismus (Stbm). We report that a myosin is polarized anteriorly in these cells and strongly colocalizes with Stbm. Disruption of the actin/myosin machinery with cytochalasin or blebbistatin disrupts polarization of Pk and Stbm, but not of myosin complexes, suggesting a PCP-independent aspect of myosin localization. Wash out of cytochalasin restored Pk polarization, but not if done in the presence of blebbistatin, suggesting an active role for myosin in core PCP protein localization. On the other hand, in the pk mutant line, aimless, myosin polarization is disrupted in approximately one third of the cells, indicating a reciprocal action of core PCP signaling on myosin localization. Our results indicate a complex relationship between the actomyosin cytoskeleton and core PCP components in which myosin is not simply a downstream target of PCP signaling, but also required for PCP protein localization.

DOI: 10.7554/eLife.05361.001

Introduction Despite the importance of the planar cell polarity (PCP) pathway in ensuring the proper orientation of cells in a range of embryonic and adult tissues, the molecular mechanisms of this pathway are not fully understood. The PCP pathway is ancient in metazoans, and its requirement for proper development and physiology has been well described in insects (e.g., Drosophila) and chordates. The function of the PCP pathway can be seen in the coordinated polarization of subcellular components such as cilia and multi-celled structures such as bristles in epithelial sheets and organs like the Drosophila wing, vertebrate inner ear, and kidney, as well as in dynamic processes, such as cell migration, convergence and extension of mesoderm, neural tube closure, and axonal guidance (Ybot-Gonzalez et al., 2007; Wallingford, 2012; Tissir and Goffinet, 2013; Papakrivopoulou et al., 2014). One of the complicating issues of PCP signaling in Drosophila, where it has been most extensively addressed, is that it appears to consist of several semi-independent pathways that may act either in parallel or in sequence (Gray et al., 2011; Lawrence and Casal, 2013). The best characterized of these pathways is the core PCP pathway, which acts locally to coordinate polarity between neighboring cells. The key components of the core pathway include the transmembrane proteins Strabismus/Van Gogh and Frizzled, and the cytoplasmic proteins Prickle and Dishevelled. Some components of this pathway are shared with the wnt/β-catenin signaling pathway, although unlike the Wnt/β-catenin pathway the core PCP pathway is thought to act primarily by directing cytoskeletal organization, rather than by regulating transcription.

While the core PCP pathway coordinates local polarity in groups of cells, there appears to be a requirement for an additional signaling pathway(s) that brings this local coordination into register.
with the axes of the organ or embryo. The precise nature of this so-called ‘global’ signaling pathway has been elusive, although several candidate pathways are supported experimentally. One model for the Drosophila global polarizer stresses the importance of mechanical forces generated during morphogenesis leading to the alignment of cells (Eaton and Jülicher, 2011). Two signaling pathways have also been proposed to act as global polarizers—the Wg/Wnt4 pathway (Wu et al., 2013), and the Dachsous (Ds)/Fat/Four-jointed (Fj) pathway (Casal et al., 2006; Thomas and Strutt, 2012; Matis et al., 2014; Olofsson et al., 2014). The Ds/Fat/Fj module has been most extensively characterized and appears to signal via formation of heterodimers with the extracellular domains of Ds and Fat (which are atypical cadherins), putatively in a gradient across the tissue (Ambegaonkar et al., 2012; Bosveld et al., 2012; Brittle et al., 2012). Most recently, it was reported that the Ds/Fat/Fj module affects polarity through microtubule orientation, which in turn directs core PCP polarization (Matis et al., 2014). Fat and Ds orthologs also play roles in vertebrate development, but their precise roles in regulating aspects of PCP remain to be clarified (Mao et al., 2011; Saburi et al., 2012). One final complicating factor in assessing the role of the Fat/Ds/Fj pathway in planar polarity is the apparent overlap with the growth-stimulating Hippo pathway (Lawrence and Casal, 2013).

The notochord of the ascidian Ciona provides a particularly tractable model for the study of the PCP pathway in tissue morphogenesis (Kourakis et al., 2014). Ascidians are invertebrate chordates, and as members of the chordate subphylum Tunicata they belong to the group of animals that are the closest extant relatives of the vertebrates (Delsuc et al., 2006). While the presence of the notochord—a stiff axial rod of mesodermal cells lying under the nerve cord—is a uniting feature of the Chordata, tunicate notochords are much simpler than those of vertebrates, and in Ciona the fully formed notochord consists of only 40 cells arranged in a stack one-cell wide (Jiang et al., 2005; Kourakis et al., 2014). We have described two discrete developmental phases in notochord morphogenesis that shows polarized cell behavior. Initially, the notochord precursor cells undergo a mediolaterally oriented intercalation behavior, which forms the notochord column along the AP axis. The role of the core PCP pathway in the convergent extension of the Ciona notochord is seen in the pk
mutant aimless (aim) of Ciona savignyi, which has defects in notochord cell intercalation behavior, morphology, and extracellular matrix deposition (Jiang et al., 2005; Veeman et al., 2008).

Following the completion of intercalation, the individual notochord cells initially assume a flattened-disk shape (Jiang and Smith, 2007). Over the next approximately 3–4 hr (corresponding to stages 21–24 [Hotta et al., 2007]), the cells undergo a dramatic elongation in the A/P axis, which results in the continued elongation of the entire tail. During the process of elongation, a second polarity becomes evident in the A/P axis (at a right angle to the mediolateral polarity seen earlier at intercalation). The first sign of A/P polarity can be seen shortly after the completion of intercalation (stage 22) in the localization of Pk protein to the anterior poles of the cells (Kourakis et al., 2014). As elongation continues the core PCP protein Strabismus (Stbm) also localizes to the anterior pole, while nuclei move to the posterior poles of the cells where they take on a flattened shape (Figure 1) (Jiang et al., 2005; Kourakis et al., 2014). The exception to this pattern is the posterior-most notochord cell whose nucleus is usually found in reverse orientation relative to the other cells (Jiang et al., 2005; Kourakis et al., 2014). In aim embryos, the nuclei of intercalated cells are still polarized, but they are randomly oriented to either the anterior or posterior pole of each cell (Kourakis et al., 2014). Following elongation, the notochord enters a new phase in its development (from stage 24 onward). A matrix is secreted into extracellular pockets that form between A and P faces of the cells (Dong et al., 2009; Denker and Jiang, 2012; Deng et al., 2013). As this process continues, the pockets of matrix between the cells expand and then fuse to make a single, uninterrupted lumen along the length of the notochord.

In this manuscript, we examine the relationship between core PCP signaling and the actin/myosin network. We report here that the notochord cells have anteriorly polarized myosin machinery. While existing models depict the polarization of myosin as downstream of PCP signaling, we instead present evidence for a more complex series of interactions in which the core PCP components and the myosin machinery act in a reciprocal fashion to promote cell polarization.

Results

Myosin is polarized in notochord cells

The Ciona intestinalis genome is predicted to contain six members of the non-muscle myosin II family (Chiba et al., 2003). One of these, myosin10/11/14/9, is reported to be expressed exclusively in the notochord (Satou et al., 2001) and is found in a two-gene operon with a myosin regulatory light chain (MRLC) (Satou et al., 2008). This MRLC is one of five found in the C. intestinalis genome and has previously been used to mark myosin assemblies in notochord cells (Dong et al., 2011). By convention, Ciona genes are named according to their vertebrate orthologs (Stolfi et al., 2014). This particular Ciona MRLC shares orthology to vertebrate MRLCs 9 and 12 (phylogeny not shown) and will be referred to here as MRLC9/12.

Figure 1. Ciona intestinalis late-tailbud embryo (stage 23) expressing an electroporated Histone 2A/Red Fluorescent Protein (H2A-RFP) in the notochord. Insets show two cells to illustrate the polarization of the nuclei to the posterior of the cells. Scale bar is 50 μm.
DOI: 10.7554/eLife.05361.003
To visualize myosin assemblies in developing notochord cells, we constructed fusion proteins of MRLC9/12 and Myosin10/11/14/9 with Venus fluorescent protein and myc antigen, respectively. The resulting cDNAs were fused to the Brachyury cis-regulatory region to drive notochord expression (Corbo et al., 1997), and then introduced to embryos by electroporation. Low doses of plasmid were used to encourage highly mosaic expression. This allows the visualization of the exogenous proteins in cells isolated from the expression of neighboring cells, which would otherwise make the determination of anterior vs posterior protein localization difficult (Jiang et al., 2005; Kourakis et al., 2014). At the earliest developmental stage analyzed, stage 19—when the notochord cells are mid-intercalation, both Myosin10/11/14/9-myc and MRLC9/12-Venus were observed throughout the cells, with the highest localization on the perinotochordal surfaces where cells contact the basement membrane that surrounds the developing notochord (white arrows, stage 19; Figure 2A; see also Veeman et al., 2008). At the immediate completion of intercalation (stage 21, Figure 2B), the localization of both fusion proteins was still the strongest at the basal/lateral edges. However, less than an hour later, at stage 22, both proteins were found localized to the anterior pole of each notochord cell (arrowhead, Figure 2C), as well as continuing to be found at the basal/lateral edges. Quantification of MRLC9/12-Venus fluorescence signal along the anterior/posterior axis of notochord cells confirms a distinct transition in MRLC9/12-Venus localization between stages 21 and 22 (insets, Figure 2B,C). During notochord cell elongation, and at its completion (stage 24), the anterior localization of both proteins persists (Figure 2D). The onset of anterior/posterior localization of myosin assemblies closely matches the localization of Pk-myc, with A/P localization of Pk-myc starting at stage 22 (Kourakis et al., 2014). However, the Pk-myc and MRLC9/12 proteins do not appear to completely colocalize spatially. At stage 22, Pk-myc, Myosin10/11/14/9-myc, and MRLC9/12-Venus are each found to be enriched on the anterior side of the cells. However, as elongation progresses, Pk-myc localization becomes restricted to a small region within the center of this domain (Jiang et al., 2005; Kourakis et al., 2014), while Myosin10/11/14/9-myc and MRLC9/12-Venus remain more dispersed (Figure 2).

Colocalization of phospho-MRLC with core PCP components
To examine the localization of active endogenous myosin complexes in the notochord, embryos were stained with an antibody to mono-phosphorylated (ser19) MRLC (pMRLC) (Ikebe and Hartshorne, 1985). Specific staining was observed along the cortices of the cells (white arrowheads, Figure 3A). Because the endogenous protein is not mosaically expressed, it is not possible to definitively resolve the expression to either the anterior or posterior face of the cells. Co-staining for electroporated Pk-myc and pMRLC shows relatively little overlap between the two proteins, with Pk-myc concentrated to a smaller domain at the center of the cell, as described above, while pMRLC distributed along the face of the cell. In contrast, a much higher degree of colocalization was observed for pMRLC and Stbm-Venus (Figure 3B, yellow arrowheads). As was reported previously (Jiang et al., 2005), Stbm is distributed evenly along the anterior face of the cell, rather than being concentrated centrally like Pk.

Core PCP protein localization is lost upon disruption of the actin/myosin cytoskeleton
To examine possible relationships between polarization of the myosin machinery, core PCP (Pk-myc and Stbm-Venus) and nuclear polarity, embryos were treated with chemical inhibitors that disrupt the actin cytoskeleton (cytochalasin B and latrunculin) or myosin II motor activity (blebbistatin). Treatments were performed both to embryos with fully polarized notochord cells (stage 23), as well as to younger embryos (stage 21) before polarity is evident.

At stage 23, cytochalasin B caused notochord nuclei to detach from posterior membranes and drift anteriorly toward the middle of the cell (Figure 4B,E; and Video 1). In these same cells, we observed a loss of polarization of both Pk-myc (Figure 4A,B) and Stbm-Venus (Figure 4D,E). Embryos were scored for cytochalasin B treatment at both mid-stage 23 and late-stage 23, and only 6% (n = 35) and 5% (n = 20) of cells showed Pk-myc polarization, respectively. Even in cells scored as having polarized Pk-myc, the tight localization seen at the center of the anterior membrane was lost, replaced by labeling throughout the anterior membrane. By contrast, cells from embryos treated with DMSO at mid-stage 23 and late-stage 23 had 81% (n = 59) and 100% (n = 5) of cells with Pk-myc properly polarized, respectively. Similar results were seen for Stbm-Venus localization. In embryos treated with cytochalasin B at stage 23, only 6% (n = 50) had detectable Stbm-Venus localization vs 75% (n = 52) of
control DMSO-treated cells. Latrunculin B, which inhibits actin polymerization by a different mechanism than cytochalasin B (Morton et al., 2000), also caused a loss of nuclear, Stbm-Venus, and Pk-myc polarization (Figure 4C,F). Only 5% and 4% of latrunculin B treated embryos showed localized Stbm-Venus and Pk-myc, respectively, vs 92% of controls (n = 57, 25, and 25). The stage 23 treatments demonstrated that cytochalasin B could disrupt the notochord cell polarity once it was established. To examine the effect of cytochalasin B on the establishment of polarity, embryos were treated at stage 21 and scored at stage 23. In these embryos, no polarization of Pk-myc was evident in any of the cells scored (n = 5) vs 92% of cells from vehicle (DMSO)-treated embryos (n = 12) having normal anterior polarization of Pk-myc.
Blebbistatin, which blocks myosin motor function, was also able to cause nuclear and Pk-myc depolarization (Figure 4G), but with varying effectiveness. In one set of experiments, we observed Pk-myc polarization in only 2 of 18 cells scored, while in second set, we observed Pk-myc polarization in 31 of 43 cells. The DMSO-treated controls showed 34/41 and 12/13 cells with proper Pk-myc polarity, respectively. Blebbistatin-treated cells scored as having Pk-myc polarization often lacked the restricted domain of labeling at the center of the membrane seen in DMSO-treated controls, instead showing a more even distribution throughout the anterior membrane.

As expected, treatment of the embryos with either cytochalasin B or latrunculin B profoundly disrupted the actin network in notochord cells. Staining of control embryos with phalloidin shows strong cortical actin in the notochord cells (Figure 4H). Cytochalasin B treatment resulted in fragmentation of the actin network (Figure 4I), with apparent loss of the equatorial contractile ring (indicated by arrows in 4H) (Sehring et al., 2014), while latrunculin B resulted in a complete loss of phalloidin staining (Figure 4J). The embryos shown in Figure 4H–J were also stained for the protein atypical protein kinase C (aPKC), which is localized to the cell cortex of notochord cells, but unlike Pk and Stbm is found at both the anterior and posterior poles of the cells (Denker et al., 2013) (Figure 4H′, arrowhead). Surprisingly, a stronger disruption in the localization of aPKC was seen with cytochalasin B treatment than with latrunculin B (Figure 4H′,J′). We speculate that aPKC may associate with the fragmented actin filaments seen with cytochalasin treatment, resulting in it spreading across the face of the cells, while in the complete absence of filamentous actin (latrunculin B) the aPKC is not displaced and is able to remain concentrated at the center of the cell.

**Myosin remains localized in the presence of cytoskeletal inhibitors**

While we observed that cytochalasin, latrunculin, and blebbistatin treatments all lead to loss of core PCP protein and nuclei localization, the same is not true for myosin localization. Using MRLC9/12-Venus as a marker, we observed that treatment of embryos with cytochalasin B or latrunculin B from
Figure 4. Loss of planar cell polarity protein localization following actin/myosin disruptions. (A–G) Single notochord cells from embryos expressing H2A-RFP (nuclear RFP), and either a Pk-myc or Stbm-Venus fusion protein (as indicated). Arrowheads indicate anteriorly localized reporter protein. Embryos were treated with either DMSO (vehicle), cytochalasin B, latrunculin B, or blebbistatin (also as indicated). (H, I, and J) Cortical actin staining of notochord cells with phalloidin in embryos treated with vehicle only (DMSO), cytochalasin B, or latrunculin B, Figure 4. continued on next page
stage 21–23 did not disrupt MRLC9/12-Venus polarization (Figure 5A–D), although the cells failed to extend to the same extent as the DMSO-treated controls, and the nuclei were displaced to the center of the cells. Additionally, in those cells treated with cytochalasin B, but not latrunculin B, MRLC9/12-Venus staining was punctate, albeit still anteriorly localized.

In older embryos, cytochalasin B appeared to be more disruptive. While treatment of embryos at early stage 23 to the start of stage 24 only resulted in a modest disruption of MRLC9/12-Venus polarization (Figure 5D), with 80% of treated cells retaining polarity vs 90% of control cells, treatment of embryos with cytochalasin B at late-stage 23 had a stronger effect, with 57% of cells scored (n = 44) having polarized MRLC9/12-Venus (vs 100% of cells in DMSO-treated embryos (n = 13); Figure 5D).

The transition to cytochalasin B dependence from early to late-stage 23 may be tied to dramatic changes in cell architecture as the notochord begins the process of lumen formation, first visible at stage 24, a period during which the orderly A/P arrangement and polarity of cells changes dramatically (Dong et al., 2009).

The dependence of the anterior polarization of MRLC9/12-Venus on myosin activity was also examined using blebbistatin. Because blebbistatin inhibits myosin II Mg-ATPase activity without disassembling actin/myosin complexes (Straight et al., 2003), we would expect to observe a disruption in polarization of MRLC9/12-Venus only if the polarization required myosin motor activity. In embryos treated with 10 μM blebbistatin from stage 21 through stage 23, a dramatic block in cell elongation was observed, although there was no apparent disruption of MRLC9/12-Venus polarization indicating that MRLC9/12-Venus polarization does not require myosin activity (Figure 5E,F).

In summary, the core PCP proteins and myosins respond very differently to cytoskeletal inhibitors. The initiation and maintenance of Stbm-Venus and Pk-myc localization depends on an intact actin cytoskeleton, and addition of the myosin inhibitor blebbistatin leads to loss of Pk-myc and Stbm-Venus polarization. These same treatments also disrupted nuclei, which normally polarize posteriorly in a PCP-dependent manner (Jiang et al., 2005), indicating an extensive disruption to the normal polarity of notochord cells. In contrast, the anterior polarization of MRLC9/12-Venus is largely resistant to these treatments, indicating that the polarization of the myosin machinery in Ciona notochord cells can occur independently of the localization of the core PCP proteins Pk and Stbm.

**Dynamic repolarization upon removal of cytochalasin B**

Despite the loss of A/P polarization of nuclei, Pk-myc, and Stbm-Venus in cytochalasin B-treated cells, we observed that polarity could be restored when the cytochalasin B was washed out. Within approximately 60 min of placing the embryos in cytochalasin B-free seawater, nuclei were observed to move back to the posterior membranes and resume their flattened shapes (Figure 6A; Video 2). Likewise, Pk-myc was observed to relocalize to anterior membranes upon wash out of cytochalasin B (Figure 6B). In assessing Pk-myc repolarization, a sample of embryos was first collected at the end of the cytochalasin B treatment to assure the efficacy of the treatment. In cells from this first group of embryos, anterior localization of Pk-myc was observed in only one of 33 cells scored. 1 hr after cytochalasin B was washed out, 13 out of 18 cells scored had re-established anterior
localization of Pk-myc (Figure 6B). However, when blebbistatin was added during the recovery from cytochalasin B treatment, the ability of the cells to properly repolarize was greatly reduced (Figure 6D). Only 14 of 45 cells (31%) scored from 19 embryos showed proper repolarization of Pk-myc in the presence of blebbistatin vs 20 of 24 cells (83%) from 15 embryos treated with vehicle (DMSO). In contrast to cytochalasin B-treated embryos, embryos treated with latrunculin B were unable to recover either nuclear or Pk-myc polarization in the notochord, even after extensive washing.

Cytochalasin B causes extensive disruption of notochord cells. The cells undergo a dramatic shape change, the polarization of Pk-myc, Stbm-Venus, and nuclei is lost, and the localization of aPKC is disrupted. Despite this, polarization of MRLC9/12-Venus persists, and much of the actin network remains cortical, although fragmented (Figure 4I). Inhibition of myosin motor activity with blebbistatin gives similar effects on Pk-myc and nuclei. Thus, in blebbistatin-treated cells or cytochalasin B-treated cells, while the actin and myosin machinery are inactivated, they appear to remain polarized and cortical. Removal of embryos from cytochalasin B allows the recovery of Pk-myc and nuclei, but not in the presence of blebbistatin. Together, these results support an active role for the actin/myosin machinery in repolarization of core PCP proteins and nuclei. Consistent with this, latrunculin treatment, while not disrupting MRLC9/12-Venus localization, caused a much more extensive disruption of the cortical actin network than did cytochalasin B (Figure 4J). These cells do not recover nuclear or Pk-myc polarization when removed from latrunculin, presumably because of the more extensive loss of the actin network. In summary, despite the loss of Stbm and Pk localization in cytochalasin-treated cells a polarity persists that can restore proper core PCP and nuclei polarity. This persistent polarity appears to be the actin/myosin machinery.

Neither establishment, maintenance nor recovery of nuclear, Pk or MRLC9/12 polarity requires the microtubule network

Previous reports in Drosophila have shown an important role for the microtubule network in PCP function (Shimada et al., 2006; Harumoto et al., 2010; Olofsson et al., 2014). The microtubule
network in the Ciona notochord can be readily visualized with a GFP-labeled version of the microtubule-binding protein Ensconsin (Figure 7A; [Roure et al., 2007]), and addition of 10 μm nocodazole results in a complete disruption of the network (Figure 7B). To test for a role of microtubules in the establishment of polarity, embryos were treated with nocodazole at stage 21. At stage 21, notochord intercalation is complete, but no signs of A/P polarity are evident. The resulting embryos observed at stage 23 showed relatively normal notochord development with no perturbations in either nuclear position (Figure 7C,D) or in Pk-myc localization (Figure 7E). Of 26 nocodazole-treated cells examined, 21 (81%) had normal Pk-myc localization vs 23 of 25 cells (92%) for a DMSO-treated control. The addition of vinblastine, another microtubule inhibitor, at stage 21 is consistent with our nocodazole results; after vinblastine treatment at stage 21, 210 of 211 cells (99.5%) showed proper nuclear polarity, compared to 227 of 228 (99.6%) for the DMSO control (data not shown). Moreover, the maintenance of Pk-myc polarization was also not disrupted by the...
addition of nocodazole at stage 23. In embryos treated with 10 μm nocodazole for 1 hr starting at stage 23, we observed that 42 of 45 cells (93%) scored had anterior-localized Pk-myc (vs 32 of 36 cells (89%) from DMSO-treated embryos).

Disruption of microtubules by the addition of nocodazole also had little effect on the anterior localization of MRLC9/12 in notochord cells. To test for a role in the establishment of polarity, nocodazole was added for 1 hr at late-stage 20, when notochord cell intercalation is nearly complete. In those cells scored, 90 of 102 had properly polarized MRLC9/12 (88.2%), while all DMSO-only treated notochord cells scored (111/111, 100%) showed anterior MRLC9/12 (Figure 7F,G). When treated later, at stages 22–23 (well after intercalation is complete) polarization of MRLC9/12-Venus was nearly unaffected in nocodazole-treated embryos; 166 of 167 (99%) of notochord cells counted showed anterior MRLC. Control animals showed localization in 100% of cells observed (61/61). Finally, addition of nocodazole did not disrupt recovery of polarity following cytochalasin B treatment. We observed that nuclei were able to properly repolarize (65/66, 98.5%) following cytochalasin wash out in the presence of 10 μm nocodazole. (Figure 7H).

Our results do not support a role for microtubules in the polarization of nuclei, Pk, or MRLC9/12. When nocodazole is added at a developmental stage immediately preceding that in which polarization of Pk-myc and MRLC9/12 is first evident, very little disruption of the polarity of these molecules is observed later. It is unknown what mechanism leads to the polarization of these molecules between stages 21 and 22, and an earlier, but yet unknown, polarity cue may be present. We have observed that the addition of nocodazole earlier interferes with intercalation, and that the onset of A/P polarization and intercalation appear to be linked, leaving open the possibility of an earlier role of microtubules in A/P polarity. Nevertheless, the role of microtubules in the localization of core PCP proteins in the Ciona notochord differs significantly from the localization of other PCP proteins.
of these proteins in other models for PCP (Shimada et al., 2006; Harumoto et al., 2010; Olofsson et al., 2014).

**Complex localization of Myosin10/11/14/9 and MRLC9/12 in the absence of PCP signaling**

The functional link between myosin polarization and the core PCP pathway was further investigated by examining Myosin10/11/14/9-myc and MRLC9/12-Venus localization in the *C. savignyi* aim line, which carries a null mutation in the *pk* gene (Jiang et al., 2005). Despite the absence of Pk activity, a few notochord cells in aim embryos are still able to intercalate. These notochord cells, which are usually but not exclusively in the posterior notochord, individually display a polarity, as is evident by the localization of the nuclei to either the anterior or posterior poles of intercalated cells. However, the polarity is not coordinated between cells, and they show no bias with respect to the A/P axis (Kourakis et al., 2014).

We assayed the localization of Myosin10/11/14/9-myc or MRLC9/12-Venus in 34 cells from 23 aim embryos, 25 having posterior nuclei and 9 having anterior nuclei (Figure 8A). We observed two distinct phenotypes for myosin localization. For those cells with posterior nuclei (i.e., the same as observed in wild-type embryos), we observed that both Myosin10/11/14/9-myc and MRLC9/12-Venus were polarized anteriorly (Figure 8B,D) as normal. By contrast, for those cells with anterior nuclei, we were unable to detect either posterior or anterior enrichment of myosin (with one exception), Figure 8A,C,E.

Thus, despite disruption of core PCP protein localization using cytoskeletal inhibitors, or by loss of Pk activity as is the case with the aim mutant, myosin can show a wild-type polarization pattern. Nevertheless, we do observe a requirement for Pk activity in properly polarizing Myosin10/11/14/9-myc and MRLC9/12-Venus in a subset of notochord cells in aim embryos.

![Figure 8](https://example.com/figure8.png)

**Figure 8.** Myosin10/11/14/9 and MRLC9/12 localization in homozygous aim embryos. (A) Distribution of nuclear and Myosin10/11/14/9 (green) or MRLC9/12 (orange) localization phenotypes (ant., anterior; post., posterior; unloc., unlocalized) in aimless embryos. Each dot represents a single scored cell. (B and C) MRLC9/12-Venus (green) localization in cells with posterior and anterior nuclei (red), respectively. (D and E) Myosin10/11/14/9-myc (green) localization in cells with posterior and anterior nuclei (red), respectively. Anterior is to the right in all panels. DOI: 10.7554/eLife.05361.012
Discussion

The results presented here demonstrate a dynamic interaction between PCP components and the actin/myosin cytoskeleton in establishing and maintaining notochord cell AP polarity. A linear pathway from the activation of the core PCP components Fz/Dsh to polarization of the actin cytoskeleton via Daam1 and RhoA has been well described, as reviewed by Wallingford (2012). PCP-directed polarization of actin is also seen in the *Drosophila* wing, where the actin-rich prehair is targeted to the distal side, opposite where pk and stbm are enriched (Wong and Adler, 1993). Our results in the *Ciona* notochord indicate a more complex interaction in which these components are mutually reinforcing rather than being in a simple linear pathway. Additionally, and in contrast to results in other experimental systems (Shimada et al., 2006; Harumoto et al., 2010; Vladar et al., 2012; Matis et al., 2014), we find no evidence of a role for the microtubule network in the localization of either PCP components Pk and Stbm, or MRLC9/12.

We report here for the first time that myosin machinery (Myosin10/11/14/9 and MRLC9/12) is anteriorly polarized in *C. intestinalis* notochord cells and shows extensive colocalization with the core PCP protein Stbm, suggesting a functional interaction between them. This is further supported by the fact that the disruption of the actin/myosin cytoskeleton with cytochalasin, latrunculin, or blebbistatin results in the loss of polarization of Pk and Stbm. Conversely, these same treatments leave the polarization of MRLC9/12 largely intact. Thus, Myosin10/11/14/9 in the notochord appears to localize in the absence of core PCP protein localization, similar to the situation for the atypical myosin Dachs in *Drosophila*, which is also polarized independently of the core PCP pathway (Brittle et al., 2012).

It should be noted, however, that no direct ortholog of Dachs is found in chordate genomes (Mao et al., 2006). Moreover, the persistence of MRLC9/12 localization when Pk and Stbm are delocalized (e.g., cytochalasin treatment), and the requirement for myosin activity during re-polarization after cytochalasin treatment, suggest that myosin polarization in Ciona notochord cells could act upstream of core PCP localization. However, it is important to note that in the developing notochord, Pk, Myosin10/11/14/9, and MRLC9/12 all appear to simultaneously localize in the A/P axis between stages 21 and 22 (Figure 2 and [Kourakis et al., 2014]). Thus, the timing of events would argue against a simple model in which myosin localization precedes and directs Pk polarization. Moreover, it has also been observed that polarization of Stbm lags behind that of Pk (and thus also of myosin) (Jiang et al., 2005), indicating that the core PCP machinery is still assembling at the time we first see polarized myosin.

The mechanism and identity of the polarizing cue that leads to the simultaneous polarization of Pk, Myosin10/11/14/9, and MRLC9/12 remains mysterious. The observations presented here in a Pk null mutant line (aim) provide clues to the nature of this signal. Myosin polarization was disrupted in approximately one third of the notochord cells scored in aim embryos. While the pk allele in aim contains an ∼200 base-pair deletion that spans one intron and an adjacent exon, creating a premature stop codon and a likely null allele (Jiang et al., 2005), and the Ciona genome contains only a single pk gene, one possible explanation for the variability between cells is that residual Pk-independent PCP activity remains in a subset of the notochord cells in aim embryos. However, by other measures of cell polarity, such as extension and protrusive activity along the mediolateral axis during intercalation, and the membrane localization of Dishevelled, we see no evidence for residual PCP activity in aim embryos, and moreover, all notochord cells appear to be uniformly disrupted (Jiang et al., 2005; Veeman et al., 2008; Kourakis et al., 2014). Thus, we surmise that many cells in the notochord are able to properly polarize myosin without input from the core PCP pathway. The notochord cells in aim embryos show a highly consistent pattern in which those having posterior localized nuclei (i.e., the wild-type pattern) have anterior polarized myosin, while those with inverted nuclei polarity (anterior) lacked detectable myosin polarity, rather than having inverted myosin polarity. We speculate that those cells in aim embryos showing wild-type nuclear polarity are correctly aligned with a global polarizing cue, and can polarize their myosin complex, while those cells showing anterior nuclei are not in alignment, and myosin does not polarize properly. The role of the core PCP pathway, which is lacking in the aim cells, is thus to act locally to bring the cells into proper alignment. Despite this observation, disruption of Pk and Stbm localization with cytochalasin C, latrunculin, or blebbistatin has relatively little effect on MRLC9/12 localization. One potentially important difference between these two examples is that the aim mutants are null for pk from fertilization, while the cytoskeletal inhibitors are added post-intercalation. Thus, while we see no evidence of A/P polarization until approximately
1 hr after the completion of intercalation (e.g., Figure 2), earlier events during intercalation may be required for the establishment of A/P polarity. The nature of these events and the mechanism that switches notochord polarity from mediolateral to A/P remain unknown.

**Materials and methods**

**Animal culture**

Adult *C. intestinalis* were collected from Santa Barbara Harbor or purchased from M-REP (Carlsbad, CA, USA) and kept at a facility supplied with raw seawater at the University of California, Santa Barbara. Gametes were isolated, mixed, and dechorionated as previously described. Anatomical landmarks were used to stage animal development (*Hotta et al., 2007*).

**Transgenesis**

Plasmid constructs were electroporated into one-cell stage embryos (*Christiaen et al., 2009*). The electroporation procedure typically results in mosaic expression, allowing us to score expressing cells in a background of non-expressing neighbors. Nuclei were visualized either using an existing *C. intestinalis* stable transgenic line expressing GFP under control of the brachyury promoter (*Joly et al., 2007*), or by electroporation with Bra > GFP::H2A::RFP (*Kourakis et al., 2014*).

**Cytoskeletal inhibitors**

Cytochalasin B (C2743, Sigma, St. Louis, MO) was provided at 10 mg/ml in DMSO and diluted in seawater to 10 μg/ml. Blebbistatin (B0560, Sigma) was prepared as 10 mM in DMSO and diluted in seawater to a final concentration of 10 μM. Nocodazole (M1404, Sigma) was diluted to 10 mM in DMSO and used at a final concentration of 10 μM in seawater. Latrunculin B was prepared as a 10 mg/ml stock solution and diluted to a working concentration of 2 μg/ml or 10 μg/ml in sea water; both concentrations yielded similar phenotypes. Vinblastine (V1377, Sigma) was added to a final concentration of (6 μg/ml). For each of these drugs, DMSO alone in seawater served as a negative control. An additional positive control was performed for working concentrations of nocodazole; for any trial using the microtubule inhibitor, we also confirmed that exposure to it resulted in cleavage arrest in embryos.

**Plasmid constructs**

A *C. intestinalis* MRLC cDNA (KH2012:KH.C11.143; [Satou et al., 2005]) was PCR amplified (forward primer: ATGTCGAGCCGACGAACTAAAA; reverse primer: AATGTCATCTTTTTCTTTAGCGCCAT) and cloned into pDONR221. This was recombined with the Brachyury promoter (forward primer: TAACGACGATTGTTCCGTCA; reverse primer: TATAGGTTTGTAACTCGCACTGAGC) donor construct into pSP72-R3-ccdBCmR-R5-Rfa-Venus (*Roure et al., 2007*) to generate *Bra>MRLC-Venus*.

The *C. intestinalis* Stbm cDNA (KH2012:kh.C4.173 [Satou et al., 2005]) generated previously (*Jiang et al., 2005*) was cloned into pDONR221, and recombined with the Brachyury promoter donor construct (above) into pSP72-R3-ccdBCmR-R5-Venus-Rfa (*Roure et al., 2007*) to generate *Bra>Venus-Stbm*.

The Bra promoter sequence was cloned into pSP72BSSPE-Swal-Rfa and recombined with an entry vector encoding amino acids 18–283 of human ensconsin (*pEntr>Ensconsin-3xGFP*) (*Roure et al., 2007*) to generate *Bra>Ensconsin-3xGFP*.

A gBlock fragment (IDT, Coralville, IA) containing approximately 1 kb of the C-terminal portion of Myosin 10/11/14/9 (Kh. C11.456; [Satou et al., 2005]) cDNA was generated to include a myc tag (GCATCAATGCAGAAGCTGATCTCAGAGGAGGACCTG) inserted immediately 5’ of the stop codon. This fragment was cloned into the full-ORF clone of Myosin 10/11/14/9 (cima8817) at the SacII and Nde1 sites. This results in a full-length Myosin 10/11/14/9 cDNA with a C-terminal myc tag, including 5’ and 3’ UTRs. This donor construct was recombined with a *Bra* promoter containing destination vector to generate *Bra>Myosin 10/11/14/9-myc*.

**Immunohistochemistry**

Embryos were fixed in 2% paraformaldehyde in seawater for 1 hr then washed 4 times in PBST, 10 min each. Embryos were incubated in PBST + 5% goat serum for 1 hr at room temperature, or overnight at 4°C, with the primary antibody diluted 1:1000 (anti-GFP, anti-myc, anti-RFP; Invitrogen, Grand Island, NY), 1:650 (anti-HA; Millipore, Billerica, MA), 1:300 (anti-PKCζ; Santa Cruz Biotechnology,
Dallas, TX), or 1:250 (anti-phospho-Myosin Light Chain 2 [Ser19]; Cell Signaling Technology, Danvers, MA). Animals were washed 5× for 10 min in PBST and then placed in an appropriate secondary antibody, and incubated as described for the primary antibody. Secondary antibodies used were antimouse or anti-rabbit Alexa Fluor-labeled antibodies (Invitrogen) with a range of excitation/emission spectra, depending on the experiment. Samples were washed 4 to 10× in PBST following secondary incubation. For microscopy, labeled embryos were immobilized onto cover slips coated with 0.08% Poly-L-lysine. Fixed embryos were cleared in 80% glycerol or in an isopropyl alcohol series followed by 2:1 benzyl alcohol:benzoyl benzoate (BABB).

**Injection of aimless embryos**

Embryos were collected from natural spawning of aimless mutants, dechorionated, and injected as described previously (Kourakis et al., 2014). An injection solution containing 0.5 μg of Myosin10/11/14/9-myc or MRLC9/12-Venus, 0.25 μg of Bra > H2A::RFP, and 0.5 mg/ml Fast Green (Sigma) was injected into one cell of a two or four-cell embryo. Embryos were fixed and immunostained (as described above) when they reached stage 23.

**Acknowledgements**

This work was supported by award GM088997 from the National Institutes of Health.

**Additional information**

**Funding**

| Funder                      | Grant reference | Author                                      |
|-----------------------------|-----------------|---------------------------------------------|
| National Institutes of Health | GM088997        | Erin Newman-Smith, Matthew J Kourakis, Wendy Reeves, Michael Veeman, William C Smith |

The funder had no role in study design, data collection and interpretation, or the decision to submit the work for publication.

**Author contributions**

EN-S, MJK, WR, MV, WCS, Conception and design, Acquisition of data, Analysis and interpretation of data, Drafting or revising the article

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