Pearls

Leishmania enriettii (Muniz & Medina, 1948): A highly diverse parasite is here to stay

Larissa F. Paranaiba1,*, Lucélia J. Pinheiro2,*, Ana C. Torrecilhas3, Diego H. Macedo4, Armando Menezes-Neto4, Wagner L. Tafuri2, Rodrigo P. Soares4,*

1 Departamento de Parasitologia, Universidade Federal Minas Gerais (UFMG), Belo Horizonte, Minas Gerais, Brazil, 2 Departamento de Patologia, Universidade Federal Minas Gerais (UFMG), Belo Horizonte, Minas Gerais, Brazil, 3 Universidade Federal de São Paulo (UNIFESP), Diadema, São Paulo, Brazil, 4 Centro de Pesquisas Renê Rachou/Fundaçao Oswaldo Cruz (Fiocruz), Belo Horizonte, Minas Gerais, Brazil

* These authors contributed equally to this work.
* larissafparanaiba@gmail.com (LFP); rsoares@cpqrr.fiocruz.br (RPS)

Leishmaniasis are a spectrum of diseases caused by protozoans from the genus Leishmania (Kinetoplastida: Trypanosomatidae) and are divided into two main clinical forms: cutaneous leishmaniasis (CL) and visceral leishmaniasis (VL). Transmission occurs after the bite of sandfly vectors (Diptera: Phlebotominae) when females take a blood meal from the vertebrate host 

In the New World, several species of Leishmania (~20) cause disease to man, the symptoms and epidemiology of which vary depending on species. However, there are species that are nonpathogenic to humans, such as L. enriettii. In 1946, Medina observed ear lesions in 2 farm-reared guinea pigs (Cavia porcellus [Rodentia: Caviida]) from the neighboring state of São Paulo. After lesion analysis, Leishmania was confirmed as the pathogen. The complete L. enriettii description was published by Muniz and Medina in 1948 at the Federal University of Paraná, Brazil [2]. Although this species has been used as a model for cutaneous leishmaniasis (CL), many aspects of its biology remain unknown. In the past 6 years, an increased interest has emerged after the finding of a similar isolate in the red kangaroo (Macropus rufus) in Australia [3]. This article aims to summarize some of the most important publications on this unique pathogen. It demonstrates a high phenotypic plasticity, being able to infect different vertebrate hosts and vectors. It also discusses recent human and veterinary infections due to other L. enriettii complex members.

L. enriettii: Vertebrate hosts

After L. enriettii discovery in C. porcellus in the 1940s [4], the authors failed to infect monkeys, dogs, and wild guinea pigs (C. aperea). They succeeded in infecting only 1 hamster out of 8 animals, and its lesion was poorly infected [2]. This is very likely to occur with wild reservoirs of Leishmania, such as opossums and armadillos, which, in nature, harbor low parasite densities without visible infection. At that time, no molecular approaches were available, opening the possibility of detecting L. enriettii in wild reservoirs other than C. aperea (Fig 1).

In spite of that, L. enriettii epidemiological studies involving hosts in Brazil are scarce, and new information on this parasite did not emerge until almost 50 years later. Two infected guinea pigs from the city of Capão Bonito (São Paulo state) and 3 from Campina Grande do Sul (Paraná state) were found. Species status was confirmed by isoenzyme analysis, with the description of a zymodeme polymorphism in the strains from Paraná [5]. Those data suggest that L. enriettii seems located in the Southern parts of Brazil, but its presence in other states...
should be prospected. A recent interesting study on *Leishmania* detection in road-killed wild animals in São Paulo found 1 *C. apera* (1/4) infected with *Leishmania*. Although this study did not type the species, it provided some evidence that sylvatic *C. apera* could harbor *Leishmania* [6]. Although the above-mentioned rodents have been considered the most common reservoirs for *L. enriettii* in Brazil, the finding of a putative similar isolate in captive red kangaroos (*M. rufus*), northern wallaroos (*Macropus robustus woodwardii*), black wallaroos (*M. bernardus*), and agile wallabies (*M. agilis*) was a landmark in the leishmaniasis epidemiology in Australia. The complete identification at species level only came in 2011 [3], classifying this isolate as a member of the *L. enriettii* complex [7].

**L. enriettii: Possible invertebrate vectors**

Only 20 years after the description of *L. enriettii* (1967), evidence appeared regarding its invertebrate host [8]. A survey of phlebotomine fauna in Parana state identified *Lutzomyia monticola* and *Lutzomyia correalima* in the vicinities where the guinea pigs were infected. Experimental sandfly infections on those guinea pigs were performed where 60% (6/10) of *L. monticola* exhibited a very high infection in their midguts. For this reason, *L. enriettii* was considered a suprapylarian species and included in the subgenus *Leishmania*. The authors recovered those parasites from sandflies and inoculated naive guinea pigs. However, no development of infection was observed, and transmission through *L. monticola* bite has yet to be determined.

Since no *L. monticola* laboratory colony is available, transmission experiments are difficult to perform. For this reason, it is a challenge to ascertain its current status as a *L. enriettii* vector. This vector is widely spread from southern to northern Brazilian states. Although *L. monticola* has no human medical importance, it is very anthropophilic and is often captured together with vectors of CL and VL. More importantly, its distribution overlaps with domestic *C. porcellus* and wild *C. apera*, reinforcing further studies with this vector.

In spite of having many sandfly species in Australia, the first evidence that midges from the subgenus *Forcipomyia* (*Lasiohelea*) (Diptera: Ceratopogonidae) could be an alternative vector of *Leishmania* has emerged. Although this vector fell into most of the Killick-Kendrick criteria, transmission still needs to be demonstrated after its colonization [3]. It is important to mention that this work triggered a recent increase in studying *L. enriettii* by many groups. Recently, 2 other ceratopogonids (*Culicoides nubeculosus* and *Culicoides sonorensis*) and

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**Fig 1. Cavia porcellus infected with Leishmania enriettii in the nose.** (A) Four to 7 weeks of infection and (B) healed lesion after 8–10 weeks of infection.

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Lutzomyia longipalpis were tested for *L. enriettii* infection using 2 strains (Brazilian and Australian) [9]. Only *C. sonorensis* was able to sustain infection, whereas *L. longipalpis* (permissive vector) developed moderate infections. Those data remarkably demonstrated *L. enriettii*’s ability to sustain and develop infection in different invertebrate hosts. Also, those studies stimulate further epidemiological investigations to identify other potential/alternative non–sandfly vectors.

The *L. enriettii* complex members in human and veterinary infections

A detailed and historical review on *Leishmania* and sandflies was recently reported. However, a universal consensus regarding *Leishmania* classification is yet to be achieved [1], especially for the “*L. enrietti* complex.” In addition to *L. enriettii*, it may include *Leishmania martiniquensis* [10], “*Leishmania siamensis*” [11], and the Australian isolate [12]. In spite of the molecular techniques in establishing phylogenetic relationships among *Leishmania* species, a few studies have included *L. enriettii* complex members. The *L. martiniquensis* (strain MAR1) was isolated in Martinique Island, French Antilles, in 1995, causing diffuse CL in an HIV-infected patient [13]. A similar human isolate of *L. martiniquensis* causing VL was also reported from Thailand [14,15]. Finally, “*L. siamensis*” was first reported in Thailand as causing VL [16] and disseminated CL/VL in a patient with HIV. The isolate from this patient was named Trang strain and had 100% identity with *L. enriettii* after molecular analysis [11]. More recently, some isolates of *L. enriettii* complex members, probably “*L. siamensis*” and *L. martiniquensis*, were isolated from CL lesions in Ghana [15]. Detection of this species was also reported from a low number of cases in horses from Florida (2) [17] and Central Europe (6) [18] and in 1 cow from Switzerland [19]. However, as mentioned by Kwakye-Nuako et al. [15], although it appeared in the literature several times, the species “*L. siamensis*” was not formally described, and its name should be used in quotation marks.

The above-mentioned data suggest that members of *L. enriettii* complex are able to infect a wide range of different hosts, from rodents to humans, and to cause many clinical manifestations. In Table 1, we summarized some of the main isolates and reference strains of the *L. enriettii* complex members. Therefore, an international consortium for whole genome sequencing could be useful to ascertain the taxonomic status of this complex using those strains. This is of importance since the real status of “*L. siamensis*” and *L. martiniquensis* are still the subject of debate.

Table 1. Available isolates and strains of the *Leishmania enriettii* complex.

| Species                   | World Health Organization code | Reference       |
|---------------------------|--------------------------------|-----------------|
| *L. enriettii*            | MCAV/BR/1985/L88                | [20–23]         |
|                           | MCAV/BR/95/CUR3                 | [22]            |
|                           | MCAV/BR/1985/COBAI_A_SP        | [23]            |
|                           | MCAV/BR/45/LV90                | [9]             |
|                           | AM-2004                         | [12]            |
| “*L. siamensis*”          | Trang                           | [11]            |
|                           | MHOM/GH/2012/GH5; LV757         | [15]            |
|                           | MHOM/GH/2012/GH10; LV758        | [15]            |
|                           | MHOM/GH/2012/GH11; LV759        | [15]            |
| *L. martiniquensis*       | MHOM/MQ/92/MAR 1                | [10, 13, 24]    |
|                           | MHOM/MQ/92/MAR 2                | [10]            |
|                           | MHOM/TH/2012/LSCM1              | [14]            |

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Concluding remarks

Since its discovery in the 1940s, *L. enriettii* studies have been occurring in pulses. In the 1950s and 1960s, most of the studies were focused on its biology, transmission, and epidemiology. Later on, in the 1970s, some immunopathological and immunological studies appeared. In the 1990s, it use as a model for molecular biology was successfully employed. More recently, the findings of other putative *L. enriettii* complex members such as *L. martiniquensis* and "*L. siamensis*" have generated great interest in understanding the taxonomical relationships among those different isolates. Whole genome sequencing would be a very important tool to investigate such relationships and help to establish their real species status. The species of this complex exhibit a high phenotypic plasticity in being able to infect a wide range of vertebrate hosts, including humans and other vectors. They may also cause different symptoms, ranging from CL to VL. However, many aspects of their epidemiology are still unknown in the geographic areas where those parasites were isolated. Although there is strong evidence of some suspected vectors, another important gap yet to be demonstrated is vectorial transmission by either ceratopogonids or *L. monticula*.

In this context, almost 70 years after *L. enriettii* discovery, many questions and uncertainties about its biology, epidemiology, classification, and immunology remain unanswered. The finding of members of this complex on different continents and its veterinary and human medical importance, especially in patients with HIV, reinforces the need for more studies. Several groups in the world are now investigating those parasites, and an increase in the published papers in the past years has demonstrated that this parasite is here to stay.

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