**ABCB1 C3435T Gene Polymorphism Frequency and Correlation with Clinical Parameters in Multiple Sclerosis**

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**Abstract**

**Background:** People with Multiple Sclerosis (MS) show varying responses to the same drugs, suggesting a genetic factor. In addition, certain ABCB1 gene polymorphisms have been associated with resistance to many drugs. In the era of individualized treatment, identifying possible genetic causes of drug nonresponsiveness in MS patients may enable the prediction of nonresponse before treatment. Research is needed to determine relationships between ABCB1 polymorphisms and patients’ clinical parameters and drug response in MS.

**Objective:** This study investigated the presence of ABCB1 C3435T polymorphism among patients with MS and evaluated possible associations between C3435T variants and disease activity and clinical parameters in MS.

**Materials and Methods:** The study included 100 patients aged 18 and over who were definitively diagnosed with MS according to the 2010 McDonald diagnostic criteria and were receiving immunomodulatory therapy, as well as a group of 100 healthy individuals. Clinical and demographic characteristics of the MS group were recorded. All study participants underwent ABCB1 C3435T genotyping. A blood sample was collected from each participant and used for DNA isolation and single-nucleotide polymorphism analysis.

**Results:** There was no statistically significant difference between MS patients and the healthy subjects with regard to ABCB1 C3435T variants. Mean score on the Extended Disability Status Scale was significantly higher in MS patients with the CT variant of the ABCB1 polymorphism compared to those with CC and TT variants, indicating that disability was more severe in MS patients with the CT genotype of the ABCB1 C3435T polymorphism.

**Conclusion:** Considering the role of P-glycoprotein in drug pharmacokinetics, the results of this study suggest a possible benefit of assessing MS patients for ABCB1 gene polymorphisms. The literature includes very little information on this topic. Although the number of patients in this study was limited, the higher level of disability in MS patients heterozygous for the ABCB1 C3435T polymorphism is a novel contribution to the literature.

**Keywords:** ABCB1 gene; C3435T; Disability; Multiple sclerosis; Polymorphism

**Statement of Significance**

Multiple Sclerosis (MS) is a debilitating and heterogeneous disease whose management continues to present major challenges. People with MS exhibit a wide range of clinical presentations as well as varying responses to the same drugs, which both indicate that genetic factors are involved in disease course and treatment response. The ability to predict treatment response in MS patients would prevent unnecessary drug usage, limit adverse side effects, and enable earlier direction of patients to effective treatment methods. The ABCB1 gene, which encodes P-glycoprotein 1, has been associated with resistance to many drugs. While the literature includes studies investigating the role of ABCB1 polymorphisms in other autoimmune diseases, there is little information available regarding its relevance in MS. Therefore, the present study evaluated associations between the ABCB1 C3435T polymorphism and MS patients’ clinical parameters such as disease activity and drug changes. The results showed that the heterozygous CT genotype of ABCB1 was associated with significantly higher disability scores, as well as a trend toward higher annual relapse rate and fewer drug changes. This suggests that ABCB1 may have an important role in MS that warrants further research in order to make progress toward individualized therapy for people with MS.

**Introduction**

Multiple Sclerosis (MS) is an autoimmune inflammatory demyelinating neurodegenerative disease of Central Nervous System (CNS) with relapsing-remitting and progressive forms [1]. MS is more common in young adults and women. Age of onset is between 20 and 40 in approximately 65% of patients. Prevalence studies have shown that the incidence is 2-3 times higher among women than men [2]. The prevalence of MS, which is known to be affected by genetic and environmental factors, varies in different regions. It is considered that this variation is caused by different seasonal characteristics, geographic locations, and ethnic profiles of the population [3]. Although the prevalence and incidence of MS in Turkey have not been definitively determined, it is likely a medium-risk region. A prevalence of 101.4 per 100,000 was reported based on clinical observations [4]. Although its etiology remains unclear, MS is believed to be affected by genetic, environmental, viral, and autoimmune factors [5].

Because inflammation in MS can appear throughout the brain,
spinal cord, and optic nerve, it can manifest with any symptom related to the central nervous system. Common signs and symptoms include weakness in the extremities, sensory symptoms, ataxia, bladder problems, fatigue, visual symptoms such as diplopia and blurred vision, dysarthria, and cognitive symptoms such as memory, concentration, and attention disorders [6].

MS is currently diagnosed using the McDonald criteria. These criteria were first introduced in 2001 and revised in 2005, 2010, and 2017. Diagnosis is made based on clinical symptoms and findings. Supportive laboratory findings are obtained through analysis of Magnetic Resonance Imaging (MRI), Cerebrospinal Fluid (CSF), and Visual Evoked Potentials (VEP) [7-9].

MS has several forms which vary in terms of disease course. The most common form encountered in clinical practice is Relapsing Remitting MS (RRMS). Approximately 80%-90% of patients present with RRMS [10]. Primary Progressive MS (PPMS), which accounts for about 10%-15% of cases, is characterized by progressive worsening of symptoms without acute relapses [11]. In Secondary Progressive MS (SPMS), neurodegeneration is more predominant than the inflammatory process. Seventy-five percent of RRMS patients naturally progress to SPMS [12]. An extremely small proportion of patients exhibit a chronically progressive course with acute relapses but no recovery between, a form called Relapsing Progressive MS (RPMS) [13,14].

There is currently no treatment to cure or prevent MS. Treatment options can be grouped into four subheadings: treatment of acute relapses, symptomatic treatment, disease-modifying therapy, and rehabilitation. Acute relapses are treated using corticosteroids and, less frequently, Adrenocorticotropic Hormone (ACTH). Disease Modifying Therapies (DMTs) including immune-modulatory and immunosuppressant drugs delivered by injection, infusion, and oral route to slow down the progress of MS and to improve patients’ quality of life [1-10]. Most of these drugs were approved for the treatment of RRMS. Only mitoxantrone is also approved for the secondary progressive and relapsing progressive forms and ocrelizumab for PPMS.

Patients treated with Interferon-beta (IFN-β) and natalizumab may develop Neutralizing antibodies (NAbs) that can adversely affect therapeutic response. A study revealed that MS patients treated with IFN-β who had persistent NAb positivity exhibited more frequent disease activity [15].

Treatment is less effective in NAb-positive patients. Other than this factor, there is no other predictive indicator for DMT response. Identifying the reasons underlying nonresponse to DMTs and predicting which patients will benefit from treatment can both improve medical outcomes and reduce expenses. Genetic variations such as ABCB1 polymorphism may serve as possible predictors of treatment failure [15].

Carrier proteins are very large protein families that transport various drugs, xenobiotics, and endogenous compounds across membranes. They have recently attracted attention because of their role in antineoplastic drug resistance and their effects on drug pharmacokinetics [16].

Numerous drugs are substrates of P-glycoprotein 1 (P-gp). P-gp has a major impact on drug distribution due to its role in the blood-brain barrier and blood-CSF barrier, where it prevents the passage of drugs to the brain and CSF and transports substrates back into the bloodstream [16-18].

Inter-individual variations in drug responses are attributed to polymorphisms or rare phenotypes [19]. The carrier protein that best demonstrates the effect of polymorphisms on pharmacokinetics is P-gp, encoded by the ABCB1 gene. Of the 29 Single-nucleotide Polymorphisms (SNPs) identified for ABCB1, C3435T in exon 26 and G2677T in exon 21 alter the substrate specificity of P-gp to eliminate its carrier protein function. The C3435T and G2677T SNPs may show ethnic variation and imbue resistance to drugs that are substrates of P-gp, thus influencing treatment responses and the prevalence of some diseases [20].

Studies have demonstrated a possible correlation between ABCB1 polymorphism and non-responsiveness to certain drugs. There are also numerous studies regarding ABCB1 polymorphism in relation to autoimmune diseases, immunosuppressant drugs used after transplantation, the determination of psychiatric drug levels, and non-responsiveness to drugs [16,21-23].

In light of these data, this study was conducted to determine the frequency of the ABCB1 C3435T polymorphism in 100 MS patients being treated with various drugs with different mechanisms of action, and compare them with those of 100 healthy controls. The objective of the study was to identify potential correlations between C3435T variants and disease activity and clinical parameters in MS patients.

Materials and Methods

This study was jointly planned by the Medical Biology and Neurology Departments of Atatürk University in Erzurum, Turkey. One hundred MS patients aged 18 and over who presented to the MS outpatient clinic of the Erzurum Ataturk University Medical School Neurology Department were included in the study. All patients were diagnosed according to the 2010 McDonald diagnostic criteria and were taking drugs with various mechanisms of action. One hundred healthy, unrelated individuals who had no systemic disease and were not taking any form of medication were included as a control group. Demographic and clinical data pertaining to the patients were noted, including age, gender, and clinical findings such as age at MS onset, disease duration, total number and annual rate of relapses, and Extended Disability Status Scale (EDSS). The patients’ current treatment regimens were not modified for the study.

The study protocol was carried out in accordance with the 1989 revised Declaration of Helsinki and was approved by the Ethics Committee of the Erzurum Regional Training and Research Hospital.

Genomic DNA extraction

Genomic DNA isolation was done using whole blood samples (2cc) collected from each patient into EDTA tubes) using a commercial kit (EZ1 DNA Blood Kit; Qiagen, Germany). DNA was extracted from 200 µL aliquots of whole blood according to the manufacturer’s protocol. DNA concentration was diluted to 10 ng/µL for working solutions and the isolated DNA was stored at -20°C.

ABCB1 polymorphism analysis

The present study focused on one Single-nucleotide Polymorphism (SNP) of the ABCB1 gene: C3435T (rs1045642). Polymerase Chain Reaction (PCR) and melting curve analyses were performed under the same conditions in a 96-well plate using a Light Cycler 480 (Roche Diagnostics, Penzberg, Germany). Genotyping was done with Light SNP typing assay (TIB-MolBiol, Berlin, Germany) by analyzing melting curves with the LightCycler 480 II system (Roche Applied Science, Mannheim, Germany). Samples with a final volume of 20 µL were
prepared by combining 2 μL of purified genomic DNA (~50 ng), 2 μL of Fast Start DNA Master HybProbe (Roche Diagnostics, Mannheim, Germany), 1 μL of Light SNiP Reagent Mix (TIB-MolBiol, Berlin, Germany), 1.6 μL of 25 mM MgCl₂, and 13.4 μL of distilled H₂O. Real-time PCR was performed as follows: denaturation at 95°C for 10 min, followed by 45 cycles of 95°C for 10 s, 60°C for 10 s, and 72°C for 15 s. After the amplification phase, a melting curve analysis was performed at 95°C for 30 s, 40°C for 2 min, 75°C for 0 s, followed by cooling phase at 40°C for 30 s. Collected data were analysed using LightCycler 480 Gene Scanning software version 1.2 (Roche Diagnostics).

**Statistical Analysis**

The study data were analysed using SPSS® version 23.0 (IBM Corp., Armonk, NY, USA) statistical software package. Frequency distribution, mean, and standard deviation values were used for comparisons between the groups. Kolmogorov-Smirnov test was used to assess normality of data distributions. Kruskal-Wallis variance of analysis was used to compare number of drug changes and annual relapse rate between polymorphism groups due to the non-normal distribution of the data. ANOVA was used for comparison of Extended Disability Status Scale (EDSS) scores because the data were normally distributed.

**Results**

The study included 100 MS patients and 100 healthy individuals.

| ABCB1 3435C>T Haplotype | MS* (n) | Control (n) |
|-------------------------|--------|------------|
| CC                      | 23     | 21         |
| CT                      | 46     | 40         |
| TT                      | 31     | 34         |
| Total                   | 100    | 100        |

**Table 1:** Distribution of C3435T genotypes and allele frequencies in MS patients and controls *MS: Multiple sclerosis.*

|                      | TT  (n=31) | CC   (n=23) | CT  (n=46) | Total (n=100) |
|----------------------|------------|------------|------------|---------------|
| Male/Female          | 1.4:3      | 1.2:5      | 1.3:8      | 1.2:6         |
| Age, years (mean ± SD)| 35.13 ± 8.10 | 36.22 ± 11.69 | 36.37 ± 9.30 | 36.41 ± 9.53   |
| Age at onset, years (mean ± SD) | 30.95 ± 8.20 | 31.96 ± 9.17 | 28.78 ± 9.00 | 30.19 ± 8.81   |
| Duration of disease, years (mean ± SD) | 4.18 ± 2.18 | 6.26 ± 5.79 | 7.59 ± 6.54 | 6.23 ± 5.34   |
| Annual relapse rate (mean ± SD) | 0.86 ± 0.01 | 1.02 ± 0.03 | 0.97 ± 0.01 | 0.85 ± 0.02   |
| Number of drug changes (mean ± SD) | 0.46 ± 0.01 | 0.49 ± 0.01 | 0.42 ± 0.01 | 0.38 ± 0.01   |
| EDSS (mean ± SD) | 1.89 ± 0.01 | 1.80 ± 1.31 | 2.48 ± 1.32 | 2.14 ± 1.26   |
| Type of multiple sclerosis | | | | |
| RRMS | 28 | 21 | 38 | 87 (87%) |
| SPMS | 2 | 2 | 8 | 12 (12%) |
| PPMS  | 1 | 0 | 0 | 1 (1%) |

**Table 2:** Demographic and clinical characteristics of multiple sclerosis patients genotyped for ABCB1.

| Drugs used by multiple sclerosis patients as disease-modifying therapy | Total patient number (n=100) | TT (n=31) | CT (n=46) | CC (n=23) |
|---------------------------------------------------------------------|-----------------------------|----------|----------|----------|
| Interferon beta 1a (sc) 22 μg or 44 μg dose 3 times weekly         | 31                          | 14       | 12       | 5        |
| Interferon beta 1a (im) 30 μg once weekly                          | 9                           | 2        | 4        | 3        |
| Interferon beta 1b 20 mg/day or 40 mg 3 times/week                 | 12                          | 2        | 6        | 4        |
| Glatiramer acetate                                                | 11                          | 4        | 5        | 2        |
| Dimethyldifumarate oral 240 mg twice daily                         | 9                           | 2        | 4        | 3        |
| Teriflunomide Oral 7-14 mg/day                                    | 7                           | 2        | 3        | 2        |
| Fingolimod Oral 0.5 mg/day                                        | 14                          | 3        | 9        | 2        |
| Natalizumab (iv) 300 mg once monthly                              | 7                           | 2        | 3        | 2        |

**Table 3:** Drugs used by MS patients as disease-modifying therapy.
levels was found and found that gene polymorphisms affect drug blood levels by determining that drug blood level (digoxin) is higher in patients with TT genotype [26].

Kerb et al. suggested that ABCB1 polymorphism is an important parameter in the development of drug resistance [27]. The C3435T polymorphism is the most common ABCB1 gene polymorphism, resulting from a single base change (C-T) at position 3435 in exon 26. It does not cause an amino acid change [28].

The effects of ABCB1 SNPs are a newly emerging area of research in MS etiology. No statistical differences have been previously reported between MS patients and controls in terms of ABCB1 2677G>T and 3435C>T. SNPs in ABC transporter genes can function as pharmaco genetic markers associated with clinical response to drug therapy in multiple sclerosis [29].

In the present study, the mean age at MS onset was 19 years, and the patients’ mean age was 36.4 years. The male to female ratio was 1:2.6, which is consistent with the literature. Similarly, the most common form of MS in the present study was RRMS, accounting for 87% of the patients. An additional 12% of the patients had SPMS and 1% had PPMS.

Conclusion

Our study constitutes a small population of MS patients in Turkey. The results of this study show that EDSS values were significantly higher among patients with the CT polymorphism when compared with the other two groups. This suggests that disability is more severe in MS patients having the CT genotype of the ABCB1 polymorphism. Presence of CT polymorphism more extensive and large studies are needed to determine whether MS patients are a predictive parameter for progression. This study may contribute to the literature in that sense. Subsequent studies will investigate larger populations with a broader scope.

Author Contributions

Study design/planning: Eda Balkan; Data collection/entry: Nuray Bilge; Data analysis/statistics: Nuray Bilge; Data interpretation: Eda Balkan; Manuscript preparation: Eda Balkan; Literature search/analysis: Eda Balkan; Funds collection: Eda Balkan and Nuray Bilge.

Ethical Standards

The study was conducted in compliance with international, national, and institutional regulations. The Ataturk University Medical Faculty Ethics Committee approved the study. All persons provided informed consent prior to inclusion in the study.

References

1. Fazzitoli MM, Jordy SS, Tilbery CP (2009) Psychiatric disorders in multiple sclerosis patients. Arc Neuro-Psiquiatr 67: 664-667.
2. Compston A, Coles A (2008) Multiple sclerosis. Lancet 372: 1502-1517.
3. Mirza M (2002) Etiology and epidemiology of multiple sclerosis. Erciyes Medical Journal 24: 40-47.
4. Börü UT, Alp R, Sur H (2006) Prevalence of multiple sclerosis door-to-door survey in Maltepe, Istanbul, Turkey. Neuroepidemiology 27: 17-21.
5. Hafler DA, Slavik JM, Anderson DE, O’Connor KC, De Jager P, et al. (2005) Multiple sclerosis. Immunological reviews 204: 208-231.
6. Cavitt B (2009) Clinical findings and symptoms in multiple sclerosis. Türkiye Klinikleri Journal of Neurology Special Topics 2: 9-15.
7. McDonald WI, Compston A, Edan G, Goodkin D, Hartung HP, et al. (2001) Recommended diagnostic criteria for multiple sclerosis: Guidelines from the International Panel on the diagnosis of multiple sclerosis. Ann Neurol 50: 121-127.
8. Polman CH, Reingold SC, Edan G, Filippi M, Hartung HP, et al. (2005) Diagnostic criteria for multiple sclerosis: Revisions to the McDonald criteria. Ann Neurol 58: 840-846.

9. Polman CH, Reingold SC, Banwell B, Clanet M, Cohen JA, et al. (2011) Diagnostic criteria for multiple sclerosis: Revisions to the McDonald criteria. Ann Neurol 69: 292-302.

10. Kantarci OH, Weisshenker BG (2005) Natural history of multiple sclerosis. Neurol Clin 23: 17-38.

11. Koch M, Kingwell E, Rieckmann P, Tremlett H (2009) The natural history of primary progressive multiple sclerosis. Neurology 73: 1996-2002.

12. Runmarker B, Andersen O (1993) Prognostic factors in a multiple sclerosis incidence cohort with twenty-five years of follow-up. Brain 116: 117-134.

13. Vukusic S, Confavreux C (2010) Natural history of multiple sclerosis. Presse Médicale 39: 359-362.

14. Lublin FD, Reingold SC (1996) Defining the clinical course of multiple sclerosis: Results of an international survey. Neurology 46: 907-911.

15. Allan CB (2018) Multiple sclerosis mechanism and immunotherapy. Neuron 97: 742-768.

16. Gül IG, Eryılmaz G, Karamustafalıoğlu OK (2016) P-glycoprotein and its role in treatment resistance. Current Approaches in Psychiatry 8: 19-31.

17. Azzalos A (2007) Drug-drug interactions affected by the transporter protein, P-glycoprotein (ABCB1, MDR1) II. Clinical aspects. Drug Discov Today 12: 838-843.

18. Balayssac D, Audhier N, Cayre A, Coudere F (2005) Does inhibition of P-glycoprotein lead to drug-drug interactions? Toxicol Lett 156: 319-329.

19. Kayalp SQ (2000) Medical pharmacology for rational treatment all 1 and 2 skin. Tibbi Farmakoloji 9: 96-107.

20. Fromm MF (2002) Genetically determined differences in P-glycoprotein function: Implications for disease risk. Toxicology 1: 181-182.

21. Lee YH, Bae SC, Song GG (2016) Association of the ABCB1 C3435T polymorphism with responsiveness to and toxicity of DMARDs in rheumatoid arthritis: A meta-analysis. J Rheumatol 75: 707-715.

22. Paul TM, Vikramraj KA, Christina J, Mariaselvam M, Negi S (2015) Multidrug resistance 1 (MDR1) 3435C>T gene polymorphism influences the clinical phenotype and methotrexate-inducedadverse. Eur J Clin Pharmacol 71: 959-996.

23. Capron A, Mourad M, de Meyer M, De Pauw L, Eddour DC, et al. (2010) CYP3A5 and ABCB1 polymorphisms influence tacrolimus concentrations in peripheral blood mononuclear cells after renal transplantation. Pharmacogenomics 11: 703-714.

24. Mirza M (2002) The etiology and the epistemology of the multiple sclerosis. Erzurum Med J 24: 40-47.

25. Fojo A, Lebo R, Shimizu N, Chin JE, Roninson IB, et al. (1986) Localization of multidrug resistance-associated DNA sequences to human chromosome. Somat Cell Mol Genet 12: 415-20.

26. Hoffmeyer S (2000) Functional polymorphisms of the human multidrug-resistance gene: Multiple sequence variations and correlation of one allele with pglycoprotein expression and activity in vivo. Proc Natl Acad Sci 97: 3473-3478.

27. Kebr R (2001) ABC drug transporters: Hereditary polymorphisms and pharmacological impact in MDR 1, MRP 1 and MRP 2. Pharmacogenomics 2: 51-64.

28. Min DJ, Ellingrod VL (2002) C3438T mutation in exon 26 of the human MDR 1geneand cyclosporine pharmacokinetics in healthy subjects. Ther Drug Monit 24: 400-404.

29. Cotte S, von Ahsen N, Kruse N (2009) ABC-transporter gene-polymorphisms are potential pharmacogenetic markers for mitoxantrone response in multiple sclerosis. Brain 132: 2517-2530.