Physical, Chemical and Biological Effects on Soil Bacterial Dynamics in Microscale Models

Sara König*, Hans-Jörg Vogel1, Hauke Harms2,3 and Anja Worrich2*

1 UFZ – Helmholtz Centre for Environmental Research, Department of Soil System Science, Halle (Saale), Germany, 2 UFZ – Helmholtz Centre for Environmental Research, Department of Environmental Microbiology, Leipzig, Germany, 3 German Centre for Integrative Biodiversity Research (iDiv) Halle-Jena-Leipzig, Leipzig, Germany

Soil is populated by highly diverse microbial communities mediating important processes and functions. The distribution of microbes, however, is neither uniform nor random. Instead, it is dictated by physical, chemical, and biological processes and conditions and varying over small spatial and temporal scales. The feedbacks between these processes make the soil-microbe complex a self-organizing system capable of adapting to the continuously changing conditions mainly driven by the highly fluctuating water content. For making meaningful predictions on the spatiotemporal dynamics of soil microbes and their functions, we need to integrate knowledge from physics, chemistry, and biology in our modeling approaches. Here, we review modeling studies with a focus on spatiotemporal dynamics of bacteria and bacterial functions in soil microhabitats. We compare these studies along four dimensions: specific aim, model type (individual-based, population-based), scale, and considered physical, chemical, biological processes and aspects. A special emphasis is laid on modeling approaches considering processes and aspects influencing the spatial distribution of bacteria such as motility, vector-based dispersal and biofilm formation. This includes factors like soil structure, carbon and oxygen gradients, temporal variations in hydration conditions or anthropogenic disturbance events. By assessing the importance of different microscale bacterial processes, this review should contribute to the ongoing discussion on challenges related to the upscaling from the microscopic via the profile to the landscape scale. Recent technical advances to observe bacteria in soils or soil-like environments combined with multidisciplinary collaborations will help to shed light on currently understudied physical, chemical and biological interactions in the soil-microbe complex.

Keywords: soil processes, soil structure, bacterial models, simulation, upscaling

INTRODUCTION

Soils harbor myriads of microorganisms providing ecosystem services such as nutrient cycling, carbon storage, water filtering, structure stabilization, pest control, and root growth promotion (Nannipieri et al., 2003; Pulleman et al., 2012; Schloter et al., 2017). Microbial activity is controlled by the physical, chemical, and biological conditions prevailing on the microscale but also feeds back on the soil conditions making the soil-microbe complex a self-organizing system...
(Young and Crawford, 2004). These interactions are highly dynamic along with continuously changing environmental conditions. Water content, for example, is linked to precipitation patterns and affects the distribution of microbes and their supply with substrates and oxygen. To analyze and predict these interactions, we need to take into account the temporal and spatial dynamics at the microscale (µm–cm). Although emerging visualization techniques and microscale laboratory methods like microfluidics have substantially enhanced our possibilities to study microorganisms in complex spatial settings (Alekket et al., 2018; Baveye et al., 2018; Juyal et al., 2019), simulation models are still the most efficient tool to systematically analyze their behavior over a broad range of conditions.

A widely used approach for simulating bacterial dynamics in a spatially explicit environment is individual-based modeling (IBM) (Kreft et al., 2013; Hellwegner et al., 2016). In IBMs, bacterial cells are considered as single entities enabling the simulation of different and dynamic properties within each cell including their stochastic behavior and evolution (Grimm, 1999). They operate in a discrete space and often consider processes like bacterial motility thereby introducing spatial dynamics even in absence of explicit spatial heterogeneity. A well-known application is species coexistence with non-hierarchical relationships, known as the rock-paper-scissor model (Kerr et al., 2002; Menezes et al., 2019). With simple process-rules in a discrete homogeneous space, these models simulate spatial patterns in highly dynamic coexistence behavior. IBMs can run in complex 2- or 3-D spatial structures mimicking soil systems (Portell et al., 2018) or be combined with pore network models (Ebrahimi and Or, 2015). Whereas soil bacterial communities consist of vast numbers of cells, IBMs are often restricted by computational power, especially if they are combined with complex physical structure models (O’Donnell et al., 2007). To overcome this constraint, population-based modeling (PBM) can be applied. In PBM, all individuals of one population are assumed to be equal in process behavior and properties, and their dynamics are often described by differential equations. The deterministic nature of PBMs allows for simulating a broad range of environmental conditions and the influence of stochasticity therein (Esser et al., 2015). Although PBMs often exclude space, there is an increasing number of microbial PBMs operating on discrete or continuous spatial areas, for example mimicking bacterial activity in the rhizosphere (Zelenev et al., 2000; Dupuy and Silk, 2016).

Here, we aim to identify physical, chemical, and biological aspects influencing the dynamics of bacteria and their functions on the microscale and give an overview on existing modeling approaches integrating such aspects. We discuss neglected but important aspects as well as missing experimental data, and what challenges we need to tackle prospectively to upscale soil bacterial processes from the microscale to scales relevant for land management. We want to point out that there is also huge effort and progress in modeling the dynamics of fungi in soils. However, for fungi the choice of the appropriate scale is much more difficult than for bacteria as mycelia may extend over and exert an influence on several hectares of soil while consisting of hyphae with a thickness of only several micrometers. Therefore, models simulating fungal networks are distinguished in micro-, intermediate and macro-scale models. They focus on processes such as tip growth, substrate uptake and branching of single hyphae (Schnepf et al., 2011), dynamics of hyphae constituting the mycelium and the extension of sparse fungal colonies over millimeters to centimeters (Pajor et al., 2010; Boswell and Davidson, 2012) and fungal biomass distribution, substrate decomposition, and translocation (Falconer et al., 2005, 2012, 2015). Due to the huge differences in fungal and bacterial life-history traits, a comparison of modeling approaches considering the interactions with their physical, chemical, and biological environment is beyond the scope of this mini review.

**MODELING PHYSICAL, CHEMICAL AND BIOLOGICAL ASPECTS OF SOIL BACTERIAL ECOSYSTEMS**

Soil is fairly different from other habitats. Soils contain a vast amount of microbial diversity, which can exceed the diversity in aquatic ecosystems by several orders of magnitude (Or et al., 2007). Overall, the specificity of soil manifests itself in complex spatiotemporal dynamics of microbial communities arising from the interaction with their physical, chemical, and biological environment. This interplay drives soil ecosystem functions and services and has been considered in modeling approaches albeit to different extents (Figure 1 and Table 1).

**Physical Aspects of Soil Bacterial Ecosystems**

Soils expose a high specific surface area (i.e., surface area per unit of mass) with less than 1% being covered by microbes (Chenu et al., 2002; Tecon and Or, 2017). This results in a patchy distribution of microbes and the emergence of spatially isolated bacterial microcolonies residing in poorly connected locations (Nunan et al., 2003; Young et al., 2008). The connectivity is controlled by a tortuous, dynamically changing pore network emerging from the complex arrangement of soil particles. The pore size and geometry determines in how far bacteria have access to their habitat. Narrow pores and pore neck diameters smaller than 0.2 µm prevent cell-to-cell contact and bacterial entry (Chenu et al., 2002). In addition, surface roughness of particles influences bacterial adhesion by shaping local diffusion fields (Mills, 2003; Long and Or, 2007; Wang and Or, 2010). When not fully saturated with water, soils exhibit three types of interfaces: solid-gas, solid-liquid, and liquid-gas. The liquid is retained in corners or adsorbed as thin films on surfaces (Tuller et al., 1999). Water flow and solute transport changes substantially with soil water content from slow seepage through smaller pores when dry to fast preferential flow through macropores at high water content (Or et al., 2007). Therefore, preferential flow paths transport higher loads of soluble carbon making them hotspots of biological activity (Bundt et al., 2001; Dechesne et al., 2014).

Using a 3-D IBM, Resat et al. (2012) modeled growth dynamics of a community consisting of species with different growth and survival strategies in response to heterogeneities...
### TABLE 1 | Overview of different spatially explicit microbial models, thematic focus, implementation details and considered physical, chemical, and biological aspects (IBM, individual-based model; PBM, population-based model).

| Topic                                                                 | Type       | Implemented spatial processes                                                                 | Considered aspects                                                                 | Scale     | Area       | References               |
|----------------------------------------------------------------------|------------|------------------------------------------------------------------------------------------------|-----------------------------------------------------------------------------------|-----------|------------|--------------------------|
| 1 Resource acquisition in face of low nutrient supply and competition | IBM        | Nutrient diffusion, reproductive expansion                                                     | Porous structure                                                                  | Nutrient conditions | Ecological interactions | Allison, 2005 |
|                                                                      |            |                                                                                                |                                                                                   | µm–mm     | grid       |                          |
| 2 Benefit of dispersal networks for bacterial degradation             | PBM        | Bacterial dispersal, nutrient diffusion, reproductive expansion                                | –                                                                                  | Nutrient conditions | Ecological interactions | Banitz et al., 2011 |
|                                                                      |            |                                                                                                |                                                                                   | mm–cm     | grid       |                          |
| 3 Metabolic bacterial networks in heterogeneous pore networks        | IBM        | Bacterial dispersal, nutrient diffusion, chemotaxis, reproductive expansion                   | Porous structure, Hydration conditions                                             | –         | Bacterial traits | Borer et al., 2019 |
|                                                                      |            |                                                                                                |                                                                                   | µm–mm     | 2-D pore   | network                  |
| 4 Biofilm development in porous media                                | PBM        | Bacterial dispersal, nutrient diffusion, reproductive expansion, water flow, biofilm movement, biofilm attachment/detachment biofilm | Porous structure                                                                  | –         | –          | Bozorg et al., 2011 |
|                                                                      |            |                                                                                                |                                                                                   | mm–cm     | grid       |                          |
| 5 Pattern formation through chemotaxis                               | IBM        | Bacterial dispersal, chemotaxis, reproductive expansion                                        | –                                                                                  | Chemical signals | Bacterial traits | Center and Thuiller, 2015 |
|                                                                      |            |                                                                                                |                                                                                   | µm–mm     | grid       |                          |
| 6 Effect of heterogeneity on carbon cycling models and upscaling     | PBM        | Bacterial dispersal, nutrient diffusion                                                        | –                                                                                  | Nutrient conditions | Bacterial distribution | Chakrawal et al., 2019 |
|                                                                      |            |                                                                                                |                                                                                   | mm–cm     | grid       |                          |
| 7 Physical interactions between bacteria and roots                   | PBM        | Bacterial dispersal, exudate diffusion, root expansion, chemotaxis, reproductive expansion     | –                                                                                  | Nutrient conditions | Ecological interactions | Dupuy and Silk, 2016 |
|                                                                      |            |                                                                                                |                                                                                   | µm–cm     | root       |                          |
| 8 Effects of hydration dynamics on composition and spatial organization of communities | IBM        | Bacterial dispersal, nutrient diffusion, chemotaxis, oxygen diffusion, reproductive expansion | Hydration conditions                                                              | –         | –          | Ebrahimii and Or, 2016 |
|                                                                      |            |                                                                                                |                                                                                   | µm–mm     | 3-D pore   | network                  |
| 9 Model toolbox for describing population dynamics                   | IBM        | Bacterial dispersal, chemical diffusion                                                        | –                                                                                  | Chemical signals | Ecological interactions | Gorochowski et al., 2012 |
|                                                                      |            |                                                                                                |                                                                                   | NA        | 3-D pore   | network                  |
| 10 Influence of environmental heterogeneities on colony expansion    | IBM        | Reproductive expansion                                                                         | Surface roughness                                                                 | –         | Bacterial traits | Grauka and Hallatschek, 2019 |
|                                                                      |            |                                                                                                |                                                                                   | µm–mm     | grid       |                          |
| 11 Carbon and nitrogen dynamics                                       | IBM        | Nutrient diffusion, oxygen diffusion, reproductive expansion                                    | –                                                                                  | Nutrient conditions | –                         | Gras et al., 2010 |
|                                                                      |            |                                                                                                |                                                                                   | mm–cm     | grid       |                          |
| 12 Effect of community composition on litter decomposition           | IBM        | Nutrient diffusion, reproductive expansion                                                      | –                                                                                  | Nutrient conditions | Bacterial traits     | Kaiser et al., 2014 |
|                                                                      |            |                                                                                                |                                                                                   | µm–mm     | grid       |                          |
| 13 Key processes of resilience of bacterial degradation              | PBM        | Bacterial dispersal, nutrient diffusion, reproductive expansion                                | –                                                                                  | –         | Bacterial traits | König et al., 2017 |
|                                                                      |            |                                                                                                |                                                                                   | mm–cm     | grid       |                          |

(Continued)
### TABLE 2 | Continued

| Topic | Type | Implemented spatial processes | Considered aspects | Scale | Area | References |
|-------|------|--------------------------------|-------------------|-------|------|-----------|
|       |      |                                |                   |       |      |           |
|       |      |                                | physical          |       |      |           |
| 14    | IBM  | Nutrient diffusion, reproductive expansion | Surface roughness, Hydration conditions | – | – | µm-mm grid | Long and Or, 2005 |
| 15    | PBM  | Bacterial dispersal, reproductive expansion | – | – | Ecological interactions (competition) | NA grid | Lowery and Ursel, 2019 |
| 16    | PBM  | Nutrient diffusion, reproductive expansion | Hydration conditions | – | Bacterial traits | µm-mm | 3-D pore network | Monga et al., 2014 |
| 17    | IBM  | Bacterial dispersal, predator dispersal, nutrient diffusion, reproductive expansion | – | Nutrient conditions | Ecological interactions (trophic structure) | mm–cm grid | Moore et al., 2013 |
| 18    | IBM  | Root expansion, bacterial reproductive expansion | – | pH, Nutrient conditions | bacterial distribution | µm-mm | root | Muci et al., 2012 |
| 19    | IBM  | Nutrient diffusion, reproductive expansion | Hydration conditions | Nutrient conditions | Ecological interactions (competition) | µm-mm | 3-D pore network | Portell et al., 2018 |
| 20    | IBM  | Bacterial dispersal, nutrient diffusion | – | Nutrient conditions | Dormancy | µm-mm | 3-D pore network | Resat et al., 2012 |
| 21    | PBM  | Nutrient diffusion, reproductive expansion | Porous structure | – | Dormancy, bacterial traits (metabolism) | µm–cm | 2-D pore network | Stolpovsky et al., 2012 |
| 22    | PBM  | Nutrient diffusion, reproductive expansion | Porous structure, Hydration conditions | – | Dormancy, bacterial distribution | µm-mm | 3-D pore network | Vogel et al., 2015 |
| 23    | IBM  | Population dispersal, single cell motility, nutrient diffusion | Surface roughness, Hydration conditions | – | – | mm | 2-D pore network | Wang and Or, 2013 |
| 24    | PBM  | Root expansion, bacterial reproductive expansion, moving nutrient source at root tip | Hydration conditions | Nutrient conditions | – | mm–cm | root | Zelenev et al., 2000 |

Note that “implemented spatial processes” differ from “Considered aspects” in the sense that the first refers to explicitly implemented model processes, while the latter refers to aspects whose effects on the bacterial dynamics were tested within the specific study.

in the structure and carbon distribution at the pore and aggregate scale. They showed that, although pore geometry had an influence, the trends of the growth dynamics at both scales were largely similar. This indicates that microscale simulations may be able to represent predicted dynamics occurring at larger scales. While this study assumed a fully saturated porous system, other modeling approaches revealed the importance of aqueous phase fragmentation on microbial processes and spatiotemporal dynamics. Using a roughness network, Long and Or (2007) simulated bacterial growth at different matric potentials and found that high matric potentials, which correspond to higher water contents and larger nutrient...
storage on the surface, led to higher abundances and faster expansion of the population. In subsequent studies, it was also shown that small reductions of the matric potential induced a sharp decrease in cell velocities to almost complete inhibition of colony expansion at \(-5 \text{ kPa}\) (Wang and Or, 2010). The functional consequences of macropore topology, water saturation and bacterial distribution on biodegradation of a soluble substrate were assessed by coupling a lattice Boltzmann model and a compartmental biodegradation model in a 3-D pore scale model. CT images were used to reproduce environmental variations at the millimeter scale and the importance of physical and biological effects was tested in a full factorial design. A sensitivity analysis revealed the importance of physical factors for the biodegradation kinetics. Both, water saturation and pore topology influenced biodegradation by affecting diffusion and thus substrate availability for bacteria (Vogel et al., 2015).

**Chemical Aspects of Soil Microbial Ecosystems**

Under unsaturated conditions the matric potential is the major determinant of the soil water potential. Nevertheless, under specific conditions such as long drought periods, after fertilizer application, in certain locations near the soil surface or in the rhizosphere, the osmotic potential may become an important component of the water potential (Or et al., 2007). Moreover, irrigation increases soil salinity especially in arid regions where evaporation is high (Lambers, 2003). In saline soils, microbial biomass and activity tends to be reduced (Rietz and Haynes, 2003). In addition, processes such as bacterial chemotaxis and motility can be repressed (Steil et al., 2003). Conversely, rapid water infiltration can increase the water potential with effects on cell viability and integrity (Halverson et al., 2000). So far, modeling studies considering soil salinity effects on microbes are virtually absent. This may be due to the fact that the impact of the osmotic potential on convective and diffusive nutrient fluxes is negligible (Or et al., 2007; Tecon and Or, 2017) and empiric information on its effects on microbial processes are scarce. The situation is very similar for the soil redox potential, which is indicative of oxidation-reduction conditions. For a long time, the redox potential has received little attention in agricultural sciences. Soils were considered to be oxic and anoxia only expected in flooded and poorly drained soils. In addition, the measurement of the redox potential is associated with methodological difficulties including the high variability in space and time and the pH dependence (Husson, 2013). Oxygen concentrations vary with soil depth due to diffusion limitations and consumption by plant roots and microbes (Noll et al., 2005). Moreover, recent studies emphasized that anoxic microsites can comprise up to 85% of the pore volume also in unsaturated soils (Keiluweit et al., 2018). Anoxic regions arise in the interior of soil aggregates due to high microbial activity directly consuming the oxygen at the periphery and allowing aerobic and anaerobic microbial communities to coexist (Sexstone et al., 1985; Long and Or, 2005; Schlüter et al., 2018). In addition, oxygen availability determines the phylogenetic and physiological composition of soil bacterial communities as it selects for specialized metabolic types (Pett-Ridge and Firestone, 2005). Under anoxic conditions, the availability of alternative electron acceptors determines the type of anaerobic metabolism. Using an IBM with a 3-D angular pore network, Ebrahimi and Or (2016) exposed differently sized soil aggregates to variable carbon, water, and oxygen contents to estimate biogeochemical fluxes from soil profiles. They found that large aggregates support anaerobic activity under unsaturated conditions. Upscaling the model to the profile scale revealed a narrow region of denitrification activity in the soil profile, while deeper regions were devoid of aerobic and anaerobic activities due to low carbon concentrations (Ebrahimi and Or, 2016). Recently, Borrer et al. (2018) investigated the impact of oxygen and carbon counter-gradients formed at the microscale inside soil aggregates and found that these gradients drive the segregation of aerobic and anaerobic bacterial populations which supported their coexistence (Borrell et al., 2018).

In several studies, soil pH was proposed as the main factor determining the diversity of soil microbial communities (Fierer and Jackson, 2006; Lauber et al., 2009). In an arable soil, bacterial diversity was twice as high at pH 8 as at pH 4. In contrast, effects of pH on fungi were only minor (Rousk et al., 2010). Using molecular and geostatistical methods to analyze the distribution of microbial communities involved in N-cycling at the landscape scale, Bru et al. (2011) found that spatial variation in community abundances could largely be explained by environmental parameters with pH being dominant.
However, similar to redox conditions, pH is often a bulk measure and conveys an imprecise picture of the conditions prevailing at the microscale. Despite its relevance for microorganisms, comparably few modeling studies integrate effects of pH. To study bacterial colonization of the root surface, Muci et al. (2012) used the cellular automata approach combined with an IBM and tested the influence and feedbacks of different environmental conditions including pH (Muci et al., 2012). Here, a lower pH decreases growth and activity of the bacteria, but was not systematically varied to test the direct effect of the pH on bacterial dynamics.

**Biological Aspects of Soil Microbial Ecosystems**

Microorganisms live in multispecies communities and interact with their neighbors via diverse mechanisms. In addition to the structural soil properties discussed above, microbial interactions are an important driver of population structure and dynamics at the microscale. Population dynamics models predict that in populations with strong antagonistic interactions, coexistence is globally unstable and competitive exclusion eventually leads to dominance of one species (Fukami, 2015). In spatially structured systems, such interactions become manifest in the emergence of a spatial pattern allowing for coexistence. Using a modified Lotka–Volterra model in a 2-D environment, Lowery and Ursell (2019) demonstrated that structured environments fostered the coexistence of two microbial species that could not coexist under homogenous conditions (Lowery and Ursell, 2019). Portell et al. (2018) simulated organic matter decomposition of a community consisting of three strains of *Arthrobacter sp.* with different competitive abilities using the previously described pore network approach with soil CT images in combination with an IBM. Here, a spatially heterogeneous distribution of organic matter was important for tripartite coexistence. Effects of social interactions between microbes on organic matter turnover and nitrogen retention were also tested with an IBM developed by Kaiser et al. (2015). Microbes exploiting the catalytic activities of others increased nitrogen retention and organic matter accumulation. Also, Wang and Or (2013) used their previously introduced hybrid modeling framework to test the influence of hydration dynamics on the coexistence of bacterial species and found that fragmented and disconnected aquatic niches emerging under dry conditions provide shelter for less competitive species. With microcosm experiments on sand, Tecon et al. (2018) showed that the contact frequency of bacterial cells increases with decreasing water content supporting these observations with a previously developed IBM. Microbes can also be involved in mutualistic interactions. A recently developed IBM coupled to a metabolic network model for soil habitats can represent trophic interactions among bacterial species, predicting the spatial organization and segregation of bacterial populations in response to oxygen and carbon gradients (Borer et al., 2019). However, spatial structure in microbial communities can also arise through self-organization. Depending on the nature of the interaction, spatial segregation or local mixing may occur (Wang and Or, 2014; Cordero and Datta, 2016). In an IBM considering a cooperating consortium, self-organization was found to be important to enable particulate carbon turnover (Ebrahimi et al., 2019).

The spatial organization and distribution of microbes in soil can also be influenced by interactions with other organisms including plants, earthworms and fungi. The effects of the latter on bacterial dispersal and biodegradation in homogeneous and heterogeneous systems were investigated via 2-D reaction-diffusion modeling. The results indicated that fungi can help bacteria to overcome areas of restricted motility (e.g., under dry conditions) thereby improving biodegradation of pollutants (Banitz et al., 2011). The rhizosphere is a hotspot for microbial activity, and several modeling studies investigated the interaction between bacteria and plant roots (Zelenev et al., 2000; Muci et al., 2012; Dupuy and Silk, 2016). For example, Dupuy and Silk (2016) observed in a PBM the benefits for bacteria attaching to the growing root compared to free-living bacteria. For investigating environmental conditions leading to wavelike patterns of bacteria in the rhizosphere, Zelenev et al. (2000) also developed a PBM describing bacterial population dynamics on a growing root releasing nutrients.

**MODELING SPATIALLY STRUCTURED DISTURBANCES**

Anthropogenic disturbances (i.e., land management, climate change) can considerably affect the soil microbial community composition or activity directly (de Vries et al., 2012; Griffiths and Philippot, 2013) or indirectly due to effects on the physical, chemical, or biological microenvironment (Figure 1). However, investigating effects of spatial disturbance on soil microorganisms is quite challenging, especially on the microscale. Some experimental studies made use of microcosm approaches and observed interesting spatial effects on community structure and function (Altermatt et al., 2011; Veraart et al., 2011; Baho et al., 2012; Worrich et al., 2016), although not on the microscale. Here, microfluidics or micromodels are promising tools to mimic soil pore structure especially when combined with new imaging techniques allowing for small-scale observations (Aleklett et al., 2018; Baveye et al., 2018). Recently, Co et al. (2019) applied microfluidics and observed an increase in antibiotic tolerance in spatially structured compared to intermixed populations (Co et al., 2019). However, the usage of microfluidics is an emerging field and until fully established, the application of simulation models to investigate microscale spatial disturbance effects is a powerful tool. Yet, there are surprisingly few microbial models focusing on this. In previous work, we established a PBM for simulating the response of bacterial population dynamics and activity to spatially heterogeneous disturbances (Banitz et al., 2011; König et al., 2017). We used bacterial degradation as an example for an important function of the soil microbial ecosystem and systematically tested spatial effects of differently distributed, lethal disturbances on the recovery, the long-term resistance (König et al., 2018b), and collapse probability (König et al., 2018a). Although we used a simple spatial structure (i.e., applied no explicit pore structure) and applied an
artificial disturbance mimicking, for instance, antibiotic pulses; we observed a high sensitivity of the functional stability to the spatial disturbance distribution in all studies. In general, a higher fragmentation of the disturbance increases functional stability and can even compensate a larger disturbance area (König et al., 2019). This relationship needs to be evaluated for natural systems with more complex spatial structures using experimental systems and more detailed pore-scale models.

THE PROBLEM OF UPSCALING

Many current research questions related to soil management or the response of terrestrial ecosystems to climate change need to be addressed on larger scales to derive management and mitigation options. For answering those large scale questions, we need to identify if and to what extent processes and interactions on the microscale have to be included at least effectively or conceptually into models operating on the profile or even on the landscape scale. Upscaling information from the highly dynamic soil-microbe complex is an ambitious task hampered by lack of data for parametrization and validation, computational constrains, spatial and temporal oscillation artifacts, or interactions of biological features with physical and chemical factors (e.g., microbial-mineral interactions) (Wang et al., 2014; Wieder et al., 2015). Moreover, already in microscale research different scales are analyzed probably hampering the comparability. Here, the reviewed model scales range from μm to cm scale, depending on the specific research question. For instance, models incorporating metabolic networks using methods like flux-balance analysis need to simulate the μm scale to depict single bacterial cells (Borer et al., 2019), while models focusing on the physical interaction between bacteria and roots need to simulate at larger scales to capture root size (Dupuy and Silk, 2016). We need to carefully consider for which research questions the effort of upscaling microbial processes is necessary to understand the whole systems dynamics. For instance, the integration of biological processes into soil models will likely increase our process understanding of biogeochemical cycles and carbon storage, especially under critical environmental conditions such as droughts (Fry et al., 2018). Spatial and temporal upscaling of models can be proceeded by applying different approaches like simplifying the model, averaging model outputs or model equations, or using effective parameters (Bierkens et al., 2000). Many soil models use simplified microbial dynamics by integrating turnover rates or microbial biomass as implicit bulk variable, for example, for simulation of carbon or nitrogen cycling on different scales (Manzoni and Porporato, 2009; Schimel and Schaeffer, 2012). Simplification of models allows for better understanding emergent model dynamics and feedbacks and reduces data requirements, which is often a problem for soil model parametrization, especially for biological parameters (Wieder et al., 2015; Baveye et al., 2018). A prerequisite for simplification should be that complex effects or processes are sufficiently understood. Then, one can successively reduce its complexity while continuously checking if the model outcome still fits our observations. Recently, it was shown that a soil organic carbon model with explicit microbial dynamics can reproduce long-term and global soil carbon data (Woolf and Lehmann, 2019). For integrating microscale effects into the prediction of greenhouse gas emission Ebrahimi and Or (2018) used a microbial numerical model to parameterize an analytical model with simplified aggregate structure allowing for upscaling to the landscape scale, though the prediction accuracy of average emission rates was only moderate in this case. The authors previously published several studies on microbial modeling on the aggregate or pore scale (e.g., Ebrahimi and Or, 2014, 2015), strengthening the implication that knowledge on the microscale is needed to reasonably scale-up processes of the soil-microbe complex. Extrapolation of microbial dynamics or mathematical formulations from the microscale to the macroscale requires a thorough understanding of the underlying mechanisms (Wang et al., 2014; Chakrawal et al., 2019).

FUTURE DIRECTIONS

Several exciting modeling approaches considering different aspects affecting the spatiotemporal dynamics of soil microbial ecosystems arose in the last two decades. Our review reveals that we are on a good way to integrate physical, chemical and biological aspects for a systemic view on microscale effects in soil microbiology. However, further experimental work is needed to understand processes and effects which are not sufficiently understood such as salinity, pH effects or spatial disturbances. As already discussed, microfluidics are a promising approach for digging deeper into microbial interactions (Aleklett et al., 2018; Baveye et al., 2018) and the increasing resolution of imaging techniques enables us to observe spatial patterns of microbes in their soil microhabitats (Juyal et al., 2019). Also, more modeling work is needed integrating aspects of all disciplines to carefully investigate microbial dynamics and functions on the microscale to gain a systemic understanding of the soil microbial ecosystem. This understanding is necessary to enable soil modelers to include microbial processes in models operating on larger scales, without describing those processes in full detail. Another problem is a possible discrepancy between measured field values and actual local conditions on the microscale. The latter may have an effect on microbial activity which will be under- or overestimated when parametrizing our models with measured field data differing from local conditions. One of the key challenges for achieving the integration of microbial microscale dynamics to profile or landscape models is model validation. The observation of microbial data such as abundance, community composition, or respiration rate gets more and more into focus of long-term field experiments (Geisseler and Scow, 2014; Romero-Oliveras et al., 2017). However, it will take decades to get reliable information on the long-term impact of climate change or agricultural practices on microbial dynamics and functions. Until then, a possibility is to validate our models by comparison of time series of specific microbial-related functions, as it was previously done, for
instance, for soil carbon dynamics (Wieder et al., 2013; Woolf and Lehmann, 2019).

It remains a scientific challenge to model the soil-microbe complex and its related functions and we suggest collaborating closely with researchers from soil physics, chemistry, and biology to incorporate knowledge of different disciplines on all relevant scales.

AUTHOR CONTRIBUTIONS

SK and AW designed the scope of the manuscript, performed the literature research, and prepared the manuscript. H-JV and HH contributed further ideas and substantially revised the manuscript.

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Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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