Duck Liver–associated Outbreak of Campylobacter Infection among Humans, United Kingdom, 2011

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Campylobacter spp.–related gastroenteritis in diners at a catering college restaurant was associated with consumption of duck liver pâté. Population genetic analysis indicated that isolates from duck samples were typical of isolates from farmed poultry. Campylobacter spp. contamination of duck liver may present a hazard similar to the increasingly recognized contamination of chicken liver.

Although bacteria in the genus Campylobacter commonly cause gastroenteritis, identified outbreaks are relatively rare. In England and Wales, 21 identified campylobacteriosis outbreaks during 1992–1994 (1) and 50 during 1995–1999 (2) accounted for 0.2% and 0.4% of reported outbreaks of gastroenteritis, respectively. Water and milk were the main sources of Campylobacter spp. outbreaks in the United Kingdom and the United States, although becoming less so (2,3). Poultry consumption and restaurant dining are the most common foodborne illness risks, although many foodstuffs are implicated (2,3). Outbreaks associated with chicken liver pâté or parfait have increased: 14 outbreaks were associated with these items in England and Wales during 2007–2009 compared with 11 during the 15 preceding years (4). There were also large outbreaks in Scotland (5,6). The peer-reviewed literature identifies chicken as the type of poultry liver or refers to poultry without specifying type.

Multilocus sequence typing is increasingly used to identify animal origins of human campylobacteriosis (7). The presence of multiple Campylobacter strains (6) in individual outbreaks linked to chicken liver is consistent with documentation that chickens harbor multiple strains (8), that pâté is prepared from multiple livers (5,6), or both. We describe epidemiologic evidence for a duck liver pâté–associated outbreak and compare sequence types (STs) of isolates with animal and food isolate datasets.

The Study

The outbreak involved a group of 3 persons and a group of 29 persons who ate lunch at a catering college restaurant. A probable case-patient was defined as a restaurant diner with diarrhea onset within 7 days after eating at the restaurant on May 12, 2011. Infections were confirmed by laboratory test results.

Environmental health officers inspected the restaurant kitchen and reviewed food preparation processes on May 17. The lunches had been ordered in advance, and officers recorded the food choices made by each diner. Menu choices and occurrence of illness were verified by face-to-face interviews (22 diners), postal interviews (9 diners), and other diners for 1 diner who had died. When food consumption history differed from the diner’s lunch order, which occurred mainly through sharing of food, consumption history was used. Fisher exact test p-values and odds ratios with CIs were calculated for the association of each menu option with illness. All case-patients reported exposure to pâté. Lower CIs were estimated by using the Cornfield method in Stata 11 (StataCorp LP, College Station, TX, USA). Repeat analysis was restricted to patients with laboratory-confirmed illness and those who were not ill.

Symptomatic patients were requested to provide fecal samples. In addition, a sample of duck liver, not from the batch used to prepare the meals in question, was obtained from the supplier on June 13 and tested for contamination of duck liver samples. In addition, a sample of duck liver, not from the batch used to prepare the meals in question, was obtained from the supplier on June 13 and tested for C. jejuni contamination. Lower CIs were estimated by using the Cornfield method in Stata 11 (StataCorp LP, College Station, TX, USA). Repeat analysis was restricted to patients with laboratory-confirmed illness and those who were not ill.

Of the 32 diners, 18 (56%) reported diarrhea: 8 had laboratory-confirmed campylobacteriosis, 6 had samples that were negative for Campylobacter infection, and 4 were not tested (Figure 1). Median duration of illness was 4 days; 1 case-patient died. Five case-patients described severe diarrhea (profuse, explosive, uncontrollable, or watery), 5
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reported fever or shivering, and 2 reported abdominal pain. Consumption of duck liver pâté was strongly associated with illness. No other positive associations were identified (Table). When analysis was restricted to confirmed cases, campylobacteriosis was strongly associated with pâté (lower CI of odds ratio 5.5; p = 0.001).

Through review of cooking processes, we found that ≈1 kg of duck livers was seared and flambéed in batches without ensuring that adequate internal cooking temperatures were achieved. The seared livers were blended with other ingredients and chilled. No other high-risk ingredients or processes were identified. No illness among staff members was recorded on or immediately preceding May 12. A catering student who made and tasted the pâté became ill on May 16. No food samples remained.

_Campylobacter_ isolates were available from 6 of 8 confirmed case-patients and the duck liver. One isolate was positive for _C. coli_ and 5 for _C. jejuni_. The _C. jejuni_ STs were ST356 (3 cases), ST50, and ST607. These STs are genetically diverse (Figure 2), but each clustered with chicken and farmed duck rather than wild waterfowl isolates. The duck liver isolate, ST5097, clustered with wild waterfowl isolates (Figure 2).

**Conclusions**

The attack rate of 86% among persons who ate duck liver pâté was similar to rates for outbreaks associated with chicken liver pâté (5,6). Pâté consumption was strongly associated with illness and laboratory-confirmed infection. Diners who did not eat this dish were unaffected. Pan-frying of chicken livers is effective for killing internal _Campylobacter_ spp. if the internal temperature reaches 70°C and is sustained for at least 2 minutes and if total cooking time is at least 5 minutes (14). The cooking process for the pâté, as reviewed by environmental health officers, was insufficient to kill bacteria inside the livers. This finding corroborates the epidemiologic evidence.

Aseptic testing of 30 chicken livers showed internal infection in 90% (14); testing of 50 chicken and 50 duck livers identified _Campylobacter_ spp. contamination in 20 and 18, respectively (15). The high level of internal and external contamination in chicken liver in these studies and failure of insufficient cooking to destroy the bacteria in the current outbreak suggest that internal contamination of duck liver also occurs. Undercooked duck liver may therefore present a hazard similar to that presented by undercooked chicken liver. Cooking time should be sufficient to destroy bacteria throughout the liver. Deliberate undercooking was identified in 68% of 25 poultry liver–associated campylobacteriosis outbreaks that occurred during 1992–2009.

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**Table.** Association between food consumed and campylobacteriosis among diners at a catering college restaurant, United Kingdom, 2011

| Food item               | Foods diners ate | Foods diners did not eat | Attack rate, % | Odds ratio* | p value† |
|-------------------------|------------------|--------------------------|----------------|-------------|----------|
|                         | Symptomatic      | Asymptomatic             | Symptomatic    | Asymptomatic|          |
| Starters                |                  |                          |                |             |          |
| Duck liver pâté         | 18               | 3                        | 0              | 11          | 86       |
| Vegetable broth         | 2                | 11                       | 16             | 3           | 15       |
| Main courses            |                  |                          |                |             |          |
| Pot roasted breast of lamb | 12             | 9                        | 6              | 5           | 57       |
| Poached plaice in white wine sauce | 5    | 5                        | 13             | 9           | 50       |
| Vegetarian polenta romaine | 1               | 0                        | 17             | 14          | 100      |
| Desserts                |                  |                          |                |             |          |
| Vanilla gateaux chantilly | 12              | 9                        | 6              | 5           | 57       |
| Chocolate pudding soufflé | 5               | 5                        | 13             | 9           | 50       |
| Cheese                  | 1                | 0                        | 17             | 14          | 100      |

*95% Cornfield CIs are in parentheses. Where odds ratio is undefined, lower CI is presented.
†By Fisher exact test.
Outbreaks associated with chicken and duck liver pâté and parfait are being increasingly identified in the United Kingdom and are likely to occur in other countries because the cooking procedures described in the United Kingdom outbreaks are not based on recipes restricted to the United Kingdom. Sporadic cases associated with similar home cooking of poultry liver products are also likely to occur, but such cases will be difficult to identify unless specifically sought.

The diversity of isolates in this outbreak resembles that in an outbreak of campylobacteriosis related to chicken liver pâté (6). As with that outbreak, the diversity in the outbreak in this study could reflect individual livers co-infected with >1 *Campylobacter* strain, >1 infected liver in the food item, or both. This diversity suggests that bacterial invasion of chicken and duck livers is possible for a wide range of fairly distantly related *Campylobacter* spp. strains, including those of *C. jejuni* and *C. coli*. The clustering of *C. jejuni* isolates from this outbreak with STs associated with farmed duck and farmed chicken and the genetic separation from wild duck and wild goose isolates (Figure 2) suggests that the farm environment may favor some *Campylobacter* spp. subtypes sufficiently to overcome natural host associations. An alternative hypothesis is that among a wide range of subtypes infecting ducks, those that are found in other farm animals are more effective at causing human disease. The single *Campylobacter* isolate from a later, non-outbreak-associated batch of duck liver clustered with isolates from wild waterfowl rather than the outbreak isolates or other isolates from farmed ducks. The limited data on *Campylobacter* populations in poultry other than chickens restrict our ability to interpret this discrepancy. Further work to characterize the *Campylobacter* populations of wild and farmed ducks may facilitate more reliable inference.

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**References**

1. Pebody RG, Ryan MJ, Wall PG. Outbreaks of *Campylobacter* infection: rare events for a common pathogen. Commun Dis Rep CDR Rev. 1997;7:R33–7.
etymology

Campylobacter [kam"pɔ-lo-bakˈtər]

From the Greek kampylos (curved) and baktron (rod), a genus of gram-negative curved or spiral rods that is among the most common causes of foodborne diarrheal illness worldwide. Illness caused by Campylobacter spp. was first described by Theodor Escherich in 1886, but they were not successfully isolated from human fecal samples until 1972. For many years, they were classified among the vibrios, but Sebald and Vérón proposed the genus Campylobacter in 1963 for these “slender, curved bacilli” that differ from the classical cholera and halophilic vibrios.

Sources

1. Dekeyser P, Gossuin-Detrain M, Butzler JP, Stermon J. Acute enteritis due to related vibrio: first positive stool cultures. J Infect Dis. 1972;125:390–2. http://dx.doi.org/10.1053/jinfdis.1972.390
2. Dorland’s Illustrated Medical Dictionary. 32nd ed. Philadelphia: Elsevier Saunders; 2012.
3. Karmali MA, Fleming PC. Campylobacter enteritis. Can Med Assoc J. 1979;120:1525–32.
4. Samie A, Obi CL, Barrett LJ, Powell SM, Guerrant RL. Prevalence of Campylobacter species, Helicobacter pylori, and Arcobacter species in stool samples from the Venda region, Limpopo, South Africa: studies using molecular diagnostic methods. J Infect. 2007;54:558–66. http://dx.doi.org/10.1016/j.jinf.2006.11.003
5. Sebald M, Vérón M. Base DNA content and classification of Campylobacter jejuni in poultry giblets [in German]. Zentralbl Veterinarmed B. 1990;37:31–4. http://dx.doi.org/10.1051/jj1990:0012023

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Frost JA, Gillespie IA, O’Brien SJ. Public health implications of Campylobacter outbreaks in England and Wales, 1995–9: epidemiological and microbiological investigations. Epidemiol Infect. 2002;128:111–8. http://dx.doi.org/10.1017/S0950268802006799
3. Friedman CR, Neimann J, Wegener HC, Tauxe RV. Epidemiology of Campylobacter jejuni infection in the United States and other industrialized nations. In: Nachamkin I, Blaser MJ, editors. Campylobacter. Washington (DC): ASM Press; 2000. p. 121–38.
4. Little CL, Gormley FJ, Rawal N, Richardson JF. A recipe for disasters: outbreaks of campylobacteriosis associated with poultry liver pâté in England and Wales. Epidemiol Infect. 2010;138:1691–4. http://dx.doi.org/10.1017/S0950268810001974
5. O’Leary MC, Harding O, Fisher L, Cowden J. A continuous common-source outbreak of campylobacteriosis associated with changes to the preparation of chicken liver pâté. Epidemiol Infect. 2009;137:383–8. http://dx.doi.org/10.1017/S0950268809000013
6. Forbes KJ, Gormley FJ, Dallas JF, Labovitiado O, MacRae M, Owen RJ, et al. Campylobacter immunity and coinfection following a large outbreak in a farming community. J Clin Microbiol. 2009;47:111–6. http://dx.doi.org/10.1128/JCM.01731-08
7. Sheppard SK, Dallas JF, Strachan NJ, Macrae M, McCarthy ND, Wilson DJ, et al. Campylobacter jejuni genotyping to determine the source of human infection. Clin Infect Dis. 2009;48:1072–8. http://dx.doi.org/10.1086/597402
8. Colles FM, McCarthy ND, Sheppard SK, Layton R, Maiden MC. Comparison of Campylobacter populations isolated from a free-range broiler flock before and after slaughter. Int J Food Microbiol. 2010;137:383–8. http://dx.doi.org/10.1017/S0950268808001003
9. Manning G, Dowson CG, Bagnall MC, Ahmed IH, West M, Newell DG. Multilocus sequence typing for comparison of veterinary and human isolates of Campylobacter jejuni. Appl Environ Microbiol. 2003;69:6370–9. http://dx.doi.org/10.1128/AEM.69.11.6370-6379.2003
10. Sheppard SK, Dallas JF, McGregor M, McCarthy ND, Sproston EL, Gormley FJ, et al. Campylobacter genotypes from food animals, environmental sources and clinical disease in Scotland 2005/6. Int J Food Microbiol. 2009;134:96–103. http://dx.doi.org/10.1016/j.ijfoodmicro.2009.02.010
11. Colles FM, Ali JS, Sheppard SK, McCarthy ND, Maiden MCJ. Campylobacter populations in wild and domesticated Mallard ducks (Anas platyrhynchos). Environ Microbiol Rep. 2011;3:574–80. http://dx.doi.org/10.1111/j.1758-2229.2011.00265.x
12. Colles FM, Dingle KE, Cody AJ, Maiden MC. Comparison of Campylobacter populations in wild geese with those in starlings and free-range poultry on the same farm. Appl Environ Microbiol. 2008;74:3583–90. http://dx.doi.org/10.1128/AEM.02491-07
13. Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S. MEGA5: Molecular Evolutionary Genetics Analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. Mol Biol Evol. 2011;28:2731–9. http://dx.doi.org/10.1093/molbev/msr121
14. Whyte R, Hudson JA, Graham C. Campylobacter jejuni in chicken livers and their destruction by pan frying. Lett Appl Microbiol. 2006;43:591–5. http://dx.doi.org/10.1111/j.1472-765X.2006.00202.x
15. Khalafalla FA. Campylobacter jejuni in poultry giblets [in German]. Zentralbl Veterniärmed B. 1990;37:31–4. http://dx.doi.org/10.1111/j.1439-0450.1990.tb01023.x

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