Vancomycin heteroresistance in coagulase negative *Staphylococcus* bloodstream infections from patients of intensive care units in Mansoura University Hospitals, Egypt

Ghada El-Saeed Mashaly and Rasha Hassan El-Mahdy*

**Abstract**

**Background:** Vancomycin heteroresistance in coagulase negative *Staphylococci* (CoNS) is a recent health concern especially in serious infections like bloodstream infections as it may lead to failure of therapy. Little information is available about the prevalence vancomycin heteroresistance in CoNS causing bloodstream infections in intensive care units (ICUs) patients of Mansoura University Hospitals (MUHs).

**Methods:** This prospective study enrolled 743 blood samples collected from ICUs patients presented with clinical manifestations of bloodstream infections over the period extending from January 2014 to March 2016. Samples were processed, coagulase negative *Staphylococci* were identified by routine microbiological methods and the absence of coagulase activity. Species were identified by API Staph 32. Oxacillin resistant CoNS were identified by cefoxitin disc diffusion method. Susceptibility testing of isolated CoNS to vancomycin was carried out using vancomycin agar dilution method. *Mec A* gene detection by PCR was done for oxacillin resistant isolates. Screening for vancomycin heteroresistance was done on brain heart infusion (BHI) agar containing 4 μg/mL vancomycin. Confirmation of vancomycin heteroresistance was carried out by population analysis profile (PAP).

**Results:** A total of 58 isolates were identified as CoNS from patients of clinically suspected bloodstream infections. The identified species were 33 (56.9%) *Staphylococcus epidermidis*, 12 (20.7%) *Staphylococcus capitis*, 7 (12.1%) *Staphylococcus haemolyticus*, and 3 isolates (5.2%) *Staphylococcus lugdunensis*. Three isolates were unidentified by API Staph 32. Forty-four (75.9%) isolates were oxacillin resistant. *Mec A* gene was detected in all oxacillin resistant isolates. Screening for vancomycin heteroresistance was done on brain heart infusion (BHI) agar containing 4 μg/mL vancomycin. Confirmation of vancomycin heteroresistance was carried out by population analysis profile.

**Conclusions:** Vancomycin heteroresistant CoNS causing bloodstream infections is growing unrecognized health hazard in ICUs patients. These isolates have susceptible vancomycin MICs. Screening methods are recommended and should be considered to improve clinical outcome in these high risk patients.

**Keywords:** Heteroresistance, Vancomycin, CoNS, ICU
glycopeptides leads to diminished glycopeptides susceptibility among CoNS [4]. Later, another type of resistance named heteroresistance has been also described in clinical CoNS isolates in which vancomycin-intermediate subpopulation of cells exist in susceptible microbial population [5, 6].

Although, there is decrease susceptibility to glycopeptides in CoNS [6, 7], little studies were conducted on heteroresistance to glycopeptides in CoNS in comparison to S. aureus [8–10]. The objective of the present study was to determine the prevalence of heteroresistance vancomycin coagulase negative Staphylococci among patients with BSIs in ICUs of Mansoura university hospitals.

Patients and methods
This study was conducted on patients who had BSI in the ICUs of Mansoura university hospitals from January 2014 to March 2016. Nine ICUs were included in this study; five medical and four surgical ICUs. Blood samples were collected from patients admitted to these ICUs and presented with symptoms with suspected bloodstream infections according to CDC criteria [11]. Bloodstream infections caused by CoNS are considered only if at least two blood cultures positive for CoNS were collected within 5 days [12]. Staphylococci were identified by standard microbiological methods [13]. CONS was confirmed by negative coagulase (coa) gene [14]. Identification of Species was done by API Staph 32 (bioMérieux) according to the manufacture instructions.

The data including: sex, age, clinical diagnosis and systemic antimicrobial therapy were retrieved from the medical records.

Oxacillin susceptibility testing
Resistance to oxacillin was evaluated by cefoxitin (30 µg) (Mast Diagnostics, Merseyside-UK) disc diffusion method according to the Clinical and Laboratory Standards Institute (CLSI) criteria [15]. Oxacillin resistant CoNS isolates were tested for the presence of mecA gene. The PCR for mecA gene was done as previously described [16].

Vancomycin susceptibility testing
Agar dilution method was done to determine the vancomycin susceptibility according to CLSI guidelines [15]. Standard vancomycin susceptible S. aureus ATCC 29213 was included as quality control.

Isolated strains were evaluated for vancomycin heteroresistance using the brain heart infusion (BHI) screen agar method as formerly described [17]. Briefly, bacterial suspension of 0.5 McFarland turbidity was prepared from an overnight culture on blood agar plate. Four 10-µL drops of the suspension were dropped onto a BHI agar plate (Mast Diagnostics, Merseyside-UK) containing 4 µg/mL vancomycin (Sigma Chemical Co., St. Louis, Mo.). Plates were incubated at 35 °C ± 2 for 48 h; the number of colonies in each drop was counted. A strain was considered to be heteroresistant to vancomycin if ≥ 1 droplet had ≥ 2 colonies.

Population analysis profile (PAP) for vancomycin heteroresistance
The PAP method was used to confirm vancomycin heteroresistance in subpopulations. All isolates that grown on BHI screen agar plate were included in PAP experiments [18]. Inoculums of (100 µL) 0.5 McFarland turbidity bacterial suspension and serial tenfold dilutions were cultured on BHI agar plates containing vancomycin at the concentrations 2, 4, 6, 8, 10, 12, 14 and 16 µg/mL at 35 °C. After 48 h, the number of colonies on plate in each concentration of drug was counted. Each strain was tested twice.

Heteroresistant strain was defined as any CoNS strain that has a susceptible vancomycin MIC by agar dilution method, can grow on BHI with 4 µg/mL vancomycin and has by PAP susceptibility testing a MIC greater than or equal to 8 µg/mL in the two experiments [19].

Results
Total of 743 of clinically suspected bloodstream infections were involved in this study. Fifty-eight isolates of CoNS were detected by absence of coA gene. S. epidermidis was the most common isolated species. Distribution of CoNS was summarized in Fig. 1. Resistance rates of the isolated CoNS to oxacillin were reported to be 76.6% (36/47). All isolates were categorized as vancomycin susceptible using agar dilution method according to the CLSI guidelines [15]. The MICs of vancomycin range from 0.25 to 2 and 0.25 to 1 µg/mL in oxacillin resistant and oxacillin sensitive isolates respectively. Vancomycin MICs by agar dilution method among different CoNS species are shown in Table 1.

![Fig. 1 Distribution of CoNS species](image)
Nine of these CoNS isolates can grow on the BHI agar containing 4 μg/mL of vancomycin. vancomycin MICs ranged from (1–2 μg/mL) by agar dilution method. By PAP these nine isolates were in the vancomycin intermediate susceptibility zone ranging from 10 to 16 μg/mL. All these heteroresistant isolates were oxacillin resistant. Majority of heteroresistant strains were oxacillin resistant. Majority of heteroresistant isolates were oxacillin resistant. Majority of heteroresistant strains were S. epidermidis (66.7%). The PAPs of these nine strains are shown in Fig. 2.

Most of heteroresistant isolates were isolated from patients with previous vancomycin exposure (6/9). The characteristics of the vancomycin heteroresistant CoNS isolates and Characteristics of patients are shown in Table 2.

**Discussion**

Coagulase-negative *Staphylococci* were considered as contaminants of bacterial cultures. However, this group especially *S. epidermidis* has emerged as an important pathogen and a major cause of serious infections in ICU patients [1, 20]. In this study, *S. epidermidis* was the most common isolated member of CoNS followed by *S. capitis*. This was in agreement with prior reports conducted in adult ICUs [5, 20].

In the current study, resistance of CoNS to oxacillin was 76.6%. Our study supports previous reports that show increased resistance of CoNS to oxacillin in which oxacillin resistance among CoNS reached 82.4% [21]. Owing to increased resistance to methicillin among CoNS, vancomycin is frequently considered as the first choice in antimicrobial therapy [22]. In contrast to oxacillin, all CoNS isolates in the present study were sensitive to vancomycin. This was in agreement with other studies conducted on different samples demonstrated that all isolates of CoNS were sensitive to vancomycin [23–25]. Vancomycin resistance is still infrequent in CoNS. However, heterogeneous resistance was reported among CoNS and was associated with failure of vancomycin therapy. These heterogeneous microcolonies may be a precursor of vancomycin resistance [5, 6]. These heterogeneous resistant strains have a susceptible vancomycin MIC, but they can grow in presence of 4 μg/mL vancomycin which is greater than their MIC [19].

Despite the presence of many methods for diagnosis of vancomycin heteroresistance in *Staphylococci* as modified E test, PAP, PAP-AUC and disc diffusion method [26–28], still PAP method is the most reliable method.
and considered as the gold standard for the detection of heteroresistant *Staphylococci* [29]. In the present study, nine isolates (15.5%) showed vancomycin heteroresistance profile identified by growth on BHI containing 4 µg/mL vancomycin and confirmed by PAP. The prevalence of heteroresistance was similar to previous reports that range from (7.4 to 18.3%) [28, 21]. All heteroresistant isolates were oxacillin resistant and most of them were associated previous vancomycin therapy. In spite of susceptibility of all isolates to vancomycin, there was an increase in vancomycin MIC range of oxacillin resistant isolates (0.25–2 µg/mL) towards the intermediate cutoff. In prior studies, increase in vancomycin MIC and rise of heteroresistant *Staphylococci* among BSI was attributed to previous use of β lactams or glycopeptides, patients admission to ICU and high rate of oxacillin resistance [28, 4]. Many previous reports show the possible relation of reduced susceptibility to glycopeptides and methicillin resistant CoNS isolates [30]. More studies extending longer durations are needed to investigate if there is a gradual increase in vancomycin MICs in oxacillin resistant CoNS like vancomycin MICs creep in methicillin resistant *S. aureus*.

**Conclusion**

This study alerts about the emergence of oxacillin-resistant CoNS showed heteroresistance to vancomycin. These heteroresistance strains were susceptible to vancomycin and cannot be detected by conventional standard methods. This implies the need of screening test for heteroresistance to avoid therapeutic failure. So, we should be aware of the potential reduction in vancomycin susceptibility of this pathogen, and this should be considered in determination of both empirical and rational therapy.

**Abbreviations**

CoNS: coagulase negative *Staphylococci*; BSIs: bloodstream infections; ICUs: intensive care units; BHI: brain heart infusion; PAP: population analysis profile.

**Authors’ contributions**

GM and RE designed the study, carried out the microbiological tests, analyzed and interpreted data, contributed in writing the manuscript. All authors read and approved the final manuscript.

**Acknowledgements**

Not applicable.

**Competing interests**

The authors declare that they have no competing interests.

**Availability of data and materials**

Please contact author for data requests.

**Consent for publication**

Not applicable.

**Ethics approval and consent to participate**

The protocol was approved by the institutional research board at faculty of medicine, Mansoura University. consent to participate were taken from all participants.

**Funding**

None.

**Publisher’s Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Received: 8 July 2017 Accepted: 13 September 2017 Published online: 19 September 2017

**References**

1. Stoll BJ, Hansen N, Fanaroff AA, Wright LL, Carlo WA, Ehrenkranz RA, Lemons JA, Donovan EF, Stark AR, Tyson JE, et al. Late-onset sepsis in very low birth weight neonates: the experience of the NICHD Neonatal Research Network. Pediatrics. 2002;110(2 Pt 1):285–91.

2. Venkatesh MP, Placencia F, Weisman LE. Coagulase-negative staphylococcal infections in the neonate and child: an update. Semin Pediatr Infect Dis. 2006;17(3):120–7.

3. Schwalbe RS, Stapleton JT, Gilligan PH. Emergence of vancomycin-resistant *Staphylococcus* among BSI was attributed to previous use of β-lactams or glycopeptides, patients admission to ICU and high rate of oxacillin resistance [28, 4]. Many previous reports show the possible relation of reduced susceptibility to glycopeptides and methicillin-resistant CoNS isolates [30]. More studies extending longer durations are needed to investigate if there is a gradual increase in vancomycin MICs in oxacillin-resistant CoNS like vancomycin MICs creep in methicillin-resistant *S. aureus*.

**Table 2** Characteristics of Vancomycin heteroresistant CoNS isolates and patients features

| Strain No | *Staphylococcus* species | OX | VA MIC | PAP MIC | Age (yrs) | Sex | ICU | Underlying disease | VA exposure |
|-----------|-------------------------|----|--------|---------|-----------|-----|-----|-------------------|------------|
| 14        | *S. epidermidis*         | R  | 2      | 16      | 68        | M   | MICU| Hypertensive encephalopathy | Yes        |
| 40        | *S. epidermidis*         | R  | 2      | 12      | 59        | M   | MICU| Liver cirrhosis       | NO         |
| 52        | *S. epidermidis*         | R  | 1      | 14      | 62        | F   | MICU| Pneumonia            | Yes        |
| 31        | *S. epidermidis*         | R  | 2      | 12      | 66        | M   | SICU| Cancer colon         | No         |
| 41        | *S. epidermidis*         | R  | 1      | 10      | 38        | F   | SICU| Motor car accident   | Yes        |
| 21        | *S. epidermidis*         | R  | 2      | 10      | 25        | M   | SICU| Motor car accident   | Yes        |
| 51        | *S. capitis*             | R  | 2      | 14      | 56        | M   | SICU| Liver recipient      | Yes        |
| 4         | *S. capitis*             | R  | 2      | 14      | 67        | F   | MICU| Cerebral stroke      | Yes        |
| 18        | *S. haemolyticus*        | R  | 4      | 12      | 69        | M   | MICU| Diabetic ketoacidosis| No         |

*R* resistant, *Va* vancomycin, *SICU* surgical ICU, *MICU* medical ICU, *M* male, *F* female
susceptibility to vancomycin in Staphylococcus epidermidis. Infect Control Hosp Epidemiol. 1999;20(3):167–70.
6. Sieradzki K, Roberts RB, Serur D, Hargrave J, Tomasz A. Heterogeneously vancomycin-resistant Staphylococcus epidermidis strain causing recurrent peritonitis in a dialysis patient during vancomycin therapy. J Clin Microbiol. 1999;37(1):39–44.
7. Nunes AP, Teixeira LM, Ionio NL, Bastos CC, de Sousa Fonseca L, Souto-Padron T, dos Santos KR. Heterogeneous resistance to vancomycin in Staphylococcus epidermidis, Staphylococcus haemolyticus and Staphylococcus warren clinical strains: characterisation of glycopeptide susceptibility profiles and cell wall thickening. Int J Antimicrob Agents. 2006;27(4):307–15.
8. Yusof A, Engelhardt A, Karlsson A, Bylund L, Vldh P, Mills K, Wootton M, Walsh TR. Evaluation of a new Etest vancomycin-teicoplanin strip for detection of glycopeptide-intermediate Staphylococcus aureus (GISA), in particular, heterogeneous GISA. J Clin Microbiol. 2008;46(9):3042–7.
9. Sun W, Chen H, Liu Y, Zhao C, Nichols WW, Chen M, Zhang J, Ma Y, Wang H. Prevalence and characterization of heterogeneous vancomycin-intermediate Staphylococcus aureus isolates from 14 cities in China. Antimicrob Agents Chemother. 2009;53(9):3642–9.
10. van Hal SJ, Wehrhahn WC, Barbagianakos T, Mercer J, Chen D, Paterson DL, Gospell IB. Performance of various testing methodologies for detection of heteroresistant vancomycin-intermediate Staphylococcus aureus in bloodstream isolates. J Clin Microbiol. 2011;49(4):1489–94.
11. Horan TC, Andrus M, Dudek MA. CDC/NHSN surveillance definition of health care-associated infection and criteria for specific types of infections in the acute care setting. Am J Infect Control. 2008;36(5):309–32.
12. Beekman SE, Diekema DJ, Doern GV. Determining the clinical significance of coagulase-negative Staphylococci isolated from blood cultures. Infect Control Hosp Epidemiol. 2005;26(6):559–66.
13. Koneman E, Allen S, Janda W, Schreckenberger P, Winn Jr W. The Gram positive cocci. I. Staphylococci and related organisms. Color atlas and textbook of diagnostic microbiology. 5th ed. Lippincott/The Williams & Wilkins Co, Philadelphia, PA. 1997:539–576.
14. Tiwari HK, Sapkota D, Sen MR. Evaluation of different tests for detection of Staphylococcus aureus using coagulase (coa) gene PCR as the gold standard. Nepal Med Coll J (NMCC). 2008;10(2):29–31.
15. Wayne P. Clinical and laboratory standards institute; performance standards for antimicrobial susceptibility testing: twenty-fourth informational supplement, M100-S24. Clin Lab Stand Inst (CLSI). 2014;34(1):1–219.
16. Murakami K, Minamide W, Wada K, Nakamura E, Teraoka H, Watanabe S. Identification of methicillin-resistant strains of Staphylococcus by polymerase chain reaction. J Clin Microbiol. 1991;29(10):2240–4.
17. Satola SW, Farley MM, Anderson KE, Patel JB. Comparison of detection methods for heteroresistant vancomycin-intermediate Staphylococcus aureus, with the population analysis profile method as the reference method. J Clin Microbiol. 2011;49(1):177–83.
18. Kim MN, Pai CH, Woo JH, Ryu JS, Hiramatsu K. Vancomycin-intermediate Staphylococcus aureus in Korea. J Clin Microbiol. 2000;38(10):3879–81.
19. Hiramatsu K, Arita K, Hanaki H, Kawasaki S, Hosoda Y, Hori S, Fukuchi Y, Kobayashi I. Dissemination in Japanese hospitals of strains of Staphylococcus aureus heterogeneously resistant to vancomycin. Lancet. 1997;350(9092):1670–3.
20. Natoli S, Fontana C, Favaro M, Bergami A, Testore GP, Minelli S, Bossa MC, Casapulla M, Broglio G, Beltrame A, et al. Characterization of coagulase-negative staphylococcal isolates from blood with reduced susceptibility to glycopeptides and therapeutic options. BMC Infect Dis. 2009;9:83.
21. Nunes AP, Schuenck RP, Bastos CCR, Magnanini MMF, Long JB, Ionio NLP. Santos KRRd: heterogeneous resistance to vancomycin and teicoplanin among Staphylococcus spp. isolated from bacteremia. Braz J Infect Dis. 2007;11(3):345–50.
22. Al-Tayyar IA, Al-Zoubi MS, Hussein E, Khudairat S, Sarosiek K. Prevalence and antimicrobial susceptibility pattern of coagulase-negative staphylococci (CoNS) isolated from clinical specimens in northern of Jordan. Iran J Microbiol. 2015;7(6):294–301.
23. Soltani R, Khalili H, Abdollahi A, Rasoolinejad M, Dashi-Khavdaki S. Nosocomial Gram-positive antimicrobial susceptibility pattern at a referral teaching hospital in Tehran, Iran. Future Microbiol. 2012;7(7):903–10.
24. Tayebi Z, Seyediavadi SS, Goudarzi M, Rahimi MK, Boromandi S, Bostanabad SZ, Mirzai A, Mahdyoum M. Frequency and antibiotic resistance pattern in gram positive uropathogens isolated from hospitalized patients with urinary tract infection in Tehran, Iran. J Genes Microbiol Immun 2014;2014:1–9.
25. Park KH, Kim ES, Kim HS, Park SJ, Bang KW, Park HJ, Park SY, Moon SM, Chong YP, Kim SH, et al. Comparison of the clinical features, bacterial genotypes and outcomes of patients with bacteremia due to heterogeneous vancomycin-intermediate Staphylococcus aureus and vancomycin-susceptible S. aureus. J Antimicrob Chemother. 2012;67(8):1843–9.
26. EL-Halfawy OM, Valvano MA. Antimicrobial heteroresistance: an emerging field in need of clarity. Clin Microbiol Rev. 2015;28(1):191–207.
27. Wong SS, Ho PL, Woo PC, Yuen KY. Bacteremia caused by staphylococci and antimicrobial susceptibility pattern of coagulase-negative staphylococci isolated in blood cultures from patients with hematological malignancies during three decades. Eur J Clin Microbiol Infect Dis. 2011;30(11):1349–54.