Transcutaneous PCO₂ Measurement at Low Temperature for Reliable and Continuous Free Flap Monitoring: Experimental and Clinical Study

Yoshiro Abe, MD
Ichiro Hashimoto, MD, PhD
Keiichi Goishi, MD, PhD
Keisuke Kashiwagi, MD
Masahiro Yamano, MD
Hideki Nakanishi, MD, PhD

Background: Measurement of transcutaneous oxygen pressure (TcPO₂) and transcutaneous carbon dioxide pressure (TcPCO₂) has been used for free flap monitoring. Because these values are obtained with sensor probes heated to 44°C, there is potential for low-temperature burns on skin flaps. We measured TcPO₂ and TcPCO₂ at 37°C in both animals and humans to determine the feasibility and safety of the procedure as a postoperative flap monitoring method.

Methods: Twelve epigastric island flaps were elevated in rabbits, and TcPO₂ and TcPCO₂ were measured at 37°C before and after ligation of the pedicles. In addition, TcPO₂ and TcPCO₂ at 37°C were measured in healthy men. Subsequently, the method was applied to postoperative monitoring of free flaps in 49 clinical cases.

Results: TcPO₂ and TcPCO₂ values were significantly affected by the experimental flap elevation. A rapid increase in TcPCO₂ was observed with both arterial and venous occlusion. In the healthy men, TcPO₂ and TcPCO₂ were measurable at all skin surface sites. In the clinical cases of free flap transfer, TcPO₂ values remained very low for at least 72 hours. TcPCO₂ values ranged from 40 to 70 mm Hg for 72 hours in more than 80% of cases. In 2 cases, TcPCO₂ values increased to more than 90 mm Hg, and exploration surgery was performed. These compromised flaps were saved by reanastomosis of the veins.

Conclusions: Continuous monitoring of TcPCO₂ at 37°C can provide objective information and alert doctors and nurses to the need for checking the free flap. (PRS GO 2013;1;e16; doi:10.1097/GOX.0b013e3182936cd0; Published online 13 May 2013.)

The success rate for free tissue transfer is now more than 95%¹ ² because of improved procedures and increased knowledge of vascular supply patterns of the flaps. Initial success rates are approaching 100%. Nevertheless, a transferred flap can be lost, and microsurgeons expend considerable effort to salvage the failing flap or at least manage the flap conservatively.³ The importance of postoperative flap monitoring for early detection of circulatory disturbance is therefore understood. Although

Disclosure: The authors have no financial interest to declare in relation to the content of this article. This study was supported by departmental resources. The Article Processing Charge was paid for by department resources.
many monitoring methods have been reported, the ideal monitoring method for detecting flap compromise has not been determined.

Flap monitoring methods include transcutaneous oxygen pressure (TcPO$_2$) measurement$^{5,6}$ and transcutaneous carbon dioxide pressure (TcPCO$_2$) measurement. The instrument for measuring TcPO$_2$ and TcPCO$_2$ was originally developed to measure partial pressure of arterial oxygen (PaO$_2$) and partial pressure of arterial carbon dioxide (PaCO$_2$) with a skin probe heated to 44°C. However, a low-temperature skin burn can result when TcPO$_2$ and TcPCO$_2$ were first measured at 37°C in rabbit epigastric island flaps that were subjected to arterial or venous occlusion. TcPO$_2$ and TcPCO$_2$ were then measured at 37°C at 8 different surface skin sites in healthy human adults. Subsequently, a clinical study was conducted in which TcPO$_2$ and TcPCO$_2$ were measured at 37°C in 49 free flaps, two of which proved to be compromised.

The aim of this 3-part study was to investigate the feasibility of monitoring TcPO$_2$ and TcPCO$_2$ in skin flaps with a probe heated to 37°C. TcPO$_2$ and TcPCO$_2$ were first measured at 37°C in rabbit epigastric island flaps that were subjected to arterial or venous occlusion. TcPO$_2$ and TcPCO$_2$ were then measured at 37°C at 8 different surface skin sites in healthy human adults. Subsequently, a clinical study was conducted in which TcPO$_2$ and TcPCO$_2$ were measured at 37°C in 49 free flaps, two of which proved to be compromised.

**MATERIALS AND METHODS**

Measurements in the experimental and clinical studies were performed with a TCM4 monitor (Radiometer Medical Aps, Copenhagen, Denmark). TcPO$_2$ and TcPCO$_2$ are measured simultaneously with the device’s sensor; the probe, which allows measurement of TcPO$_2$ and TcPCO$_2$ at the same site and the same time, contains a Clark-type electrode for measurement of TcPO$_2$ and a pH-sensitive glass electrode for measurement of TcPCO$_2$. The probe, which was fixed to the skin surface with adhesive tape, was thermostatically adjusted to 37°C. Room temperature was controlled between 18°C and 24°C during measurements in both the animal and clinical studies. Experimental procedures were approved by the institutional animal care and use committee of The University of Tokushima and were carried out in accordance with committee guidelines. The human subjects were fully informed of the reason for and the importance of TcPO$_2$ and TcPCO$_2$ measurement.

**Part 1: Animal Experiment For Testing The Effects Of Flap Elevation And Pedicle Occlusion By Low-Temperature Measurement Of TcPo$_2$ And TcPCO$_2$**

Twelve male Japanese white rabbits weighing 3.0–3.5 kg were used for the study. All were maintained under standard housing conditions and allowed water and standard dry rabbit feed ad libitum. The rabbits were anesthetized with intravenous pentobarbital (Somnopentyl) (64.8 mg/mL, Kyoritsu Seiyaku, Tokyo, Japan), and an epigastric island flap (17 cm × 5 cm) was elevated in each rabbit on the basis of the superficial inferior epigastric artery and vein, according to a well-described technique.$^{8–10}$ After the flap was sewn back into its original place, the probe was placed on the skin 4 cm away from the distal edge of the flap. The animals were divided into 2 groups for arterial or venous occlusion produced by ligation of the superficial inferior epigastric artery or vein. TcPO$_2$ and TcPCO$_2$ were measured immediately before and 20 minutes after flap elevation (before ligation of the vascular pedicle) and at 5, 10, 20, 30, 60, 90, and 120 minutes after ligation of the vascular pedicle.

**Part 2: Study Of Low-Temperature Tcpo$_2$ And Tcpco$_2$ Measurement At Different Body Sites In Healthy Humans**

We measured TcPO$_2$ and TcPCO$_2$ at 8 superficial sites in 10 healthy male volunteers (age range, 24–36 years; mean age, 30.4 years): the forehead, cheek, back, abdomen, volar aspect of the forearm, lateral aspect of the thigh, posterior aspect of the leg, and anterior aspect of the tibia. We did this to investigate whether measurement of these values with a probe adjusted thermostatically to 37°C was possible at various donor sites.

**Part 3: Low-Temperature Monitoring Of Tcpo$_2$ And Tcpco$_2$ In Clinical Cases Of Free Flap Transfer**

This part of the study comprised 49 patients [32 men and 17 women ranging in age from 19 to 84 years (mean, 54.8 years)] undergoing free flap transfer at Tokushima University Hospital between January 2002 and July 2011. All 49 patients were of normal nutritional status before surgery. The free flaps comprised 23 anterolateral thigh flaps, 10 latissimus dorsi musculocutaneous flaps, 5 fibular osteocutaneous flaps, 4 medial planter flaps, 4 thoracodorsal artery perforator flaps, 1 radial forearm flap, 1 deep inferior epigastric perforator flap, and 1 transverse rectus abdominis musculocutaneous flap. TcPO$_2$ and TcPCO$_2$ were measured continuously for at least the first 72 hours after the transfer. That is, values were recorded immediately upon the patient’s postsurgical transfer to the intensive care unit and then every hour on postoperative day 1, every 2 hours on postoperative day 2, every 4 hours on postoperative day 3, and thereafter.

**STATISTICAL ANALYSIS**

Measured values are shown as median (range). In the animal flap experiment, differences between values before and after flap elevation and differences between values before and after ligation of vascular pedicles were analyzed by Wilcoxon signed-rank
test for nonparametric data. In the clinical free flap study, values recorded at 24, 48, and 72 hours after the transfer were compared to values recorded immediately after the transfer, and differences were analyzed by Wilcoxon signed-rank test for nonparametric data.

RESULTS

Part 1: Animal Experiment for Testing the Effects of Flap Elevation and Pedicle Occlusion by Low-temperature Measurement of TcPO$_2$ and TcPCO$_2$

In the rabbit epigastric flaps subjected to arterial occlusion ($n = 6$), TcPO$_2$ before flap elevation [44.5 (30–68) mm Hg] decreased significantly to 3.5 (2–8) mm Hg after elevation ($P = 0.031$). After arterial ligation, TcPO$_2$ decreased significantly to 2 (1–3) mm Hg at 5 minutes ($P = 0.031$), and it remained below 2.0 mm Hg to the end of the study period. In this group, TcPCO$_2$ before flap elevation [57 (33–76) mm Hg] increased significantly to 85.5 (67–105) mm Hg after elevation ($P = 0.031$). After arterial ligation, TcPCO$_2$ increased significantly to 98 (71–120) mm Hg at 5 minutes ($P = 0.031$), and it increased gradually to as much as 145.5 (128–168) mm Hg by 90 minutes (Fig. 1A).

In the flaps subjected to venous occlusion ($n = 6$), TcPO$_2$ before flap elevation [65 (26–71) mm Hg] decreased significantly to 13 (2–27) mm Hg after elevation ($P = 0.031$). After venous ligation, TcPO$_2$ decreased significantly to 7 (2–10) mm Hg at 5 minutes ($P = 0.031$), and it remained below 7.0 mm Hg to the end of the study period. In this group, the TcPCO$_2$ value before flap elevation [56 (55–66) mm Hg] increased significantly to 75.5 (67–86) mm Hg after elevation ($P = 0.031$). After venous ligation, TcPCO$_2$ increased significantly to 85 (76–96) mm Hg at 5 minutes ($P = 0.031$), and it continued to increase to as much as 126 (119–158) mm Hg by 90 minutes (Fig. 1B).

Part 2: Study of Low-temperature TcPO$_2$ and TcPCO$_2$ Measurement at Different Body Sites in Healthy Adults

Among the 10 healthy subjects in whom different monitoring sites were investigated, TcPO$_2$ was lower on the face than at other sites. TcPCO$_2$ was higher on the face (up to 70 mm Hg) than elsewhere (values elsewhere never exceeded 60 mm Hg) (Fig. 2).

Part 3: Low-temperature Monitoring of TcPO$_2$ and TcPCO$_2$ in Clinical Cases of Free Flap Transfer

None of the 49 patients who underwent free flap transfer died or suffered a critical setback owing to the original defect during the intraoperative period. There were no complications associated with use of the probe, and presence of the probe elicited no complaints from the patients.

Median TcPO$_2$ was 4 (1–47 mm Hg) immediately after flap transfer (Fig. 3A). TcPO$_2$ values immediately after transfer and at 24, 48, and 72 hours after transfer were below 10 mm Hg in 71.4%, 87.8%, 86.7%, and 93.9% of patients, respectively. Median TcPCO$_2$ was 56 (40–85) mm Hg immediately after flap transfer (Fig. 3B). TcPCO$_2$ values immediately after transfer and at 24, 48, and 72 hours after transfer ranged from 40 to 70 mm Hg in 81.7%, 85.4%, 93.3%, and 84.8% of patients, respectively. TcPO$_2$ and TcPCO$_2$ values at 24, 48, and 72 hours after transfer decreased compared with the values immediately after transfer.

The lowest and highest TcPO$_2$ and TcPCO$_2$ values obtained during the first 72 hours after transfer in each case are shown in Figures 4A and B, respectively. The lowest TcPO$_2$ values ranged from 0 to 20 mm Hg and were less than 10 mm Hg in 96% (47/49) of flaps. The lowest TcPCO$_2$ values ranged from 23 to 64 mm Hg. The highest TcPO$_2$ values ranged from 2 to 47 mm Hg, and the highest TcPCO$_2$ values ranged from 45 to 112 mm Hg. Two cases in which the TcPCO$_2$ values increased to 92 and 112 mm Hg required exploration.

A sudden increase in TcPCO$_2$ was noted postoperatively in 2 patients, one at 6 hours and the other at 60 hours after flap transfer (Table 1 and Figs. 5–7). Immediate exploration was performed, and venous thrombi were removed in both cases. TcPCO$_2$ decreased immediately after venous reanastomosis. The transferred flaps were rescued and fully survived. All flaps that survived the initial operation showed TcPCO$_2$ values below 88 mm Hg. We experienced only 2 cases of flap compromise; however, both positive predictive value and negative predictive value were 100% when TcPCO$_2$ exceeding 90 mm Hg was used to determine that revision surgery was necessary (Fig. 4B). We could not confirm significant changes in TcPO$_2$ values after the reanastomoses because the values remained below 10 mm Hg. Accordingly, the decrease in TcPO$_2$ values could not be used to detect the circulatory disturbance.

DISCUSSION

Free tissue transfer depends on the patency of pedicle vessels for survival. The sooner the circulatory disturbance in the flap is detected, the sooner the exploration and rescue can be performed. Thus, numerous techniques have been investigated and used in search of the ideal free flap monitoring method. Two basic types of measurement methods exist. First, blood flow through the vascular pedicle
can be monitored by means of an implantable Doppler probe or by color duplex sonography. Second, blood flow in the peripheral circulation can be assessed by observation of flap skin color, by capillary refill, by monitoring with a handheld Doppler probe, by near-infrared spectroscopy, by use of a laser Doppler flowmeter, or by measuring the temperature of flaps, tissue oxygen tension, and TcPO$_2$ and TcPCO$_2$ with a probe heated to 44°C.

The TcPO$_2$ and TcPCO$_2$ probe detects oxygen and carbon dioxide coming from capillaries in the dermis through the epidermis and corneum. The oxygen is consumed by the epidermal cells and by the TcPO$_2$ probe on the corneum. The carbon dioxide is produced by the epidermal cells and is not consumed by the probe. This explains why the TcPO$_2$ is lower than the PaO$_2$ and the TcPCO$_2$ is higher than the PaCO$_2$ when TcPO$_2$ and TcPCO$_2$ are measured by unheated probes. When a heated probe is used, the skin temperature reaches approximately 44°C, making the TcPO$_2$ value closer to the PaO$_2$ value. The TcPCO$_2$ value measured at 44°C becomes higher than that measured at 37°C. Although a heated electrode is not required to measure TcPCO$_2$, the measurement is usually carried out under heated conditions for faster reactivity of TcPCO$_2$ upon the change in PaCO$_2$. A low-temperature burn may result from heating the skin to 44°C for more than 3 minutes.

Fig. 1. TcPO$_2$ and TcPCO$_2$ values in rabbit epigastric flaps. Values are expressed as median and range. "Before" refers to before flap elevation, "after" refers to 20 min after flap elevation (before ligation of vascular pedicle), and "time" refers to elapsed time after ligation of vascular pedicle. A, Graph showing TcPO$_2$ and TcPCO$_2$ values before and after rabbit epigastric flap elevation and arterial occlusion. B, Graph showing TcPO$_2$ and TcPCO$_2$ values before and after rabbit epigastric flap elevation and venous occlusion. ††P < 0.01 vs values before flap elevation, by Wilcoxon signed-rank test. **P < 0.01 vs values before ligation of the artery or vein, by Wilcoxon signed-rank test.
hours during continuous monitoring. Therefore, the monitoring site must be altered every 3–4 hours to avoid thermal injury, and the electrode must be recalibrated before placement. Frequent reattachment often causes inaccurate monitoring due to air bubbles trapped in the electrode, improper placement, or a damaged sensor membrane.

Our experimental study revealed that flap elevation and ligation of the vascular pedicle affected the TcPO$_2$ and TcPCO$_2$ values. This suggests that the measurement of both values can perhaps detect any circulatory disturbance caused by arterial or venous ischemia. However, the TcPO$_2$ values remain very low after flap elevation. Trends in the changing TcPO$_2$ and TcPCO$_2$ values were quite similar between arterial occlusion and venous occlusion. Thus, it is difficult to differentiate between the 2 types of occlusion through TcPO$_2$ and TcPCO$_2$ measurements.

In our study of different monitoring sites, TcPO$_2$ and TcPCO$_2$ values obtained at all measurement sites at $37^\circ$C were lower than previously reported TcPO$_2$ and TcPCO$_2$ values obtained at $44^\circ$C or $45^\circ$C. With measurement at $37^\circ$C, TcPO$_2$ and TcPCO$_2$ values on the face differed from those on other body surfaces. These differences are also found with measurement at $44^\circ$C. Thus, the TcPO$_2$ and TcPCO$_2$ values of skin flaps obtained from various donor sites can be measured at $37^\circ$C.

TcPO$_2$ values on clinical free flaps are characteristically low in comparison to TcPCO$_2$ values. TcPO$_2$ values on the skin are reported to vary not only in relation to the actual perfusion situation, density of the capillary network, and temperature but also in relation to regional factors, that is, thickness of the stratum corneum and density of the subdermal sebaceous glands. Flap elevation decreased TcPO$_2$ values in our animal study. The low TcPO$_2$ values...

---

**Fig. 2.** Boxplot showing TcPO$_2$ and TcPCO$_2$ values (median, upper and lower quartiles, and range) at 8 different sites in healthy adults. Values were obtained with probes thermally controlled at $37^\circ$C.

**Fig. 3.** Clinical postoperative TcPO$_2$ and TcPCO$_2$ values obtained with probes thermally controlled at $37^\circ$C. Circles represent individual patients. A, Graph of TcPO$_2$ values obtained immediately after surgery (0 h) and at 24-h intervals thereafter. Bars indicate the median values at each time point. B, Graph of TcPCO$_2$ values obtained immediately after surgery (0 h) and at 24-h intervals thereafter. Bars indicate the median values at each time point. *P < 0.05 vs values at 0 h (ie, immediately after surgery), by Wilcoxon signed-rank test. **P < 0.01 vs values at 0 h (ie, immediately after surgery), by Wilcoxon signed-rank test.
may reflect postoperative edema of the flap skin or the vasospasm that occurs during elevation of the flap.7,8,25

TcPCO₂ values are measurable at 37°C even after clinical free flap elevation. TcPCO₂ is not readily influenced by the cutaneous factors described above.20,23 We suppose that TcPCO₂ is influenced by the severe circulatory disturbance such as venous or arterial ischemia rather than the cutaneous factors.7 Reasons for the decrease in TcPO₂ and TcPCO₂ we observed every 24 hours after the transfer are not clear. The decrease in TcPCO₂ may reflect a settling down of the circulatory condition, whereas the decrease in TcPO₂ may mean continuing edema of the flap skin.

When critical circulatory failure occurs in the free flap, we can expect to detect the failure by the decrease in TcPO₂ and increase in TcPCO₂. The lowest TcPCO₂ values measured in the 2 patients undergoing removal of a thrombus in the vascular pedicle and those measured in patients who did not require exploration were very low and not distinguishable. The highest TcPCO₂ values measured in the 47 patients who did not require exploration were less than 90 mm Hg. The highest values measured in the 2 patients who underwent reoperation were more than 90 mm Hg, and these values decreased to less than 80 mm Hg after removal of the thrombi. This means a line can be drawn at 90 mm Hg of TcPCO₂ to indicate exploratory surgery. This value (90 mm Hg) is the same as that reported in a previous study performed at 44°C.7 TcPCO₂ of more than 90 mm Hg is the critical limit for circulatory failure of a free flap.

The TcPCO₂ monitoring method does not involve direct observation of blood flow to determine the viability of skin flaps. A TcPCO₂ of 90 mm Hg might not be definitive for a diagnosis of flap failure; TcPCO₂ values in some of our flaps that survived were very close to this cutoff value in the present study.

### Table 1. Details of the 2 Cases Requiring Exploration

| Patient         | Place of Defect (Cause) | Flap Type                           | Highest TcPCO₂ (mm Hg) |
|-----------------|-------------------------|-------------------------------------|------------------------|
| 51-year-old man | Foot (injury)           | Fibular o-c flap combined with soleus muscle | 112                    |
| 71-year-old woman | Check (SCC)            | ALT flap combined with VLM           | 92                     |

ALT, anterolateral thigh; o-c, osteocutaneous; SCC, squamous cell carcinoma; VLM, vastus lateralis muscle.
cal assessment that includes the pinprick test and observation of flap color remains the gold standard for evaluation of circulatory failure. We believe that final judgment for exploration must include clinical assessment. However, changes in the skin color of the flap can seem vague and assessment is subjective, especially for an untrained doctor or nurse, and such assessment does not equal continuous nor quantitative monitoring. Monitoring of TcPCO$_2$ at 37°C can be performed continuously without burning the skin. When an alarm is set for a TcPCO$_2$ value of 90 mm Hg, even an untrained doctor or nurse can be alerted to check for circulatory failure because the critical TcPCO$_2$ value is clear, objective, and easily understood.

**CONCLUSIONS**

In our study, TcPO$_2$ and TcPCO$_2$ values measured at 37°C were affected by flap elevation and arterial and venous ischemia. However, the TcPO$_2$ value was not particularly useful for detecting circulatory failure of free flaps because this value remained very low after free flap transfer. On the basis of our study findings, we suggest a TcPCO$_2$ value of 90 mm Hg to be the point at which free flap failure is indicated. The measurement procedure is noninvasive, safe, continuous, quantitative, easy to understand, and commercially available. We believe that this method is useful as an initial alert to the possibility of flap compromise and the need for further checking and that it can reduce the risk of flap failure by early detection of circulatory disturbance in the transferred flap.

**REFERENCES**

1. Hidalgo DA, Disa JJ, Cordeiro PG, et al. A review of 716 consecutive free flaps for oncologic surgical defects: refinement in donor-site selection and technique. *Plast Reconstr Surg*. 1998;102:722–732; discussion 733.
2. Khouri RK, Cooley BC, Kunselman AR, et al. A prospective study of microvascular free-flap surgery and outcome. *Plast Reconstr Surg*. 1998;102:711–721.
3. Weinzweig N, Gonzalez M. Free tissue failure is not an all-or-none phenomenon. *Plast Reconstr Surg*. 1995;96:648–660.
4. Furnas H, Rosen JM. Monitoring in microvascular surgery. *Ann Plast Surg*. 1991;26:255–272.
5. Achauer BM, Black KS, Litke DK. Transcutaneous PO$_2$ in flaps: a new method of survival prediction. *Plast Reconstr Surg*. 1980;65:738–745.
6. Svedman P, Jacobsson S, Ponnert L, et al. Transcutaneous oxygen tension in flaps. *Chir Plast.* 1982;6:201–207.

7. Hashimoto I, Nakanishi H, Takiwaki H, et al. Flap monitoring by transcutaneous PO\(_2\) and PCO\(_2\): importance of transcutaneous PCO\(_2\) in determining follow-up treatment for compromised free flaps. *J Reconsr Microsurg.* 2007;23:269–274.

8. Raskin DJ, Nathan R, Erk Y, et al. Critical comparison of transcutaneous PO\(_2\) and tissue pH as indices of perfusion. *Microsurgery* 1983;4:29–33.

9. Gould JS, Sully L, O'Brien BM, et al. The effects of combined cooling and perfusion on experimental free-flap survival in rabbits. *Plast Reconstr Surg.* 1985;76:104–109.

10. May JW Jr, Chait LA, O'Brien BM, et al. The no-reflow phenomenon in experimental free flaps. *Plast Reconstr Surg.* 1978;61:256–267.

11. Swartz WM, Jones NF, Cherup L, et al. Direct monitoring of microvascular anastomoses with the 20-MHz ultrasonic Doppler probe: an experimental and clinical study. *Plast Reconstr Surg.* 1988;81:149–161.

12. Harrison DH, Girling M, Mott G. Experience in monitoring the circulation in free-flap transfers. *Plast Reconstr Surg.* 1981;68:543–555.

13. Scheufler O, Andresen R. Tissue oxygenation and perfusion in inferior pedicle reduction mammoplasty by near-infrared reflection spectroscopy and color-coded duplex sonography. *Plast Reconstr Surg.* 2003;111:1131–1146.

14. Liss AG, Liss P. Use of a modified oxygen microelectrode and laser-Doppler flowmetry to monitor changes in oxygen tension and microcirculation in a flap. *Plast Reconstr Surg.* 2000;105:2072–2078.

15. Heller L, Levin LS, Klitzman B. Laser Doppler flowmeter monitoring of free-tissue transfers: blood flow in normal and complicated cases. *Plast Reconstr Surg.* 2001;107:1739–1745.

16. Cohn KH, May JW Jr. Thermal-energy dissipation: a laboratory study to assess patency in blood vessels. *Plast Reconstr Surg.* 1982;70:475–480.

17. Mahoney JL, Lista FR. Variations in flap blood flow and tissue PO\(_2\): a new technique for monitoring flap viability. *Ann Plast Surg.* 1988;20:43–47.

18. Hirigoyen MB, Blackwell KE, Zhang WX, et al. Continuous tissue oxygen tension measurement as a monitor of free-flap viability. *Plast Reconstr Surg.* 1997;99:763–773.

19. Takiwaki H, Arase S, Nakanishi H, et al. Transcutaneous PO\(_2\) and PCO\(_2\) measurements in various skin lesions. *J Dermatol.* 1991;18:311–313.

20. Rochat MC, Payne JT, Pope ER, et al. Evaluation of skin viability in dogs, using transcutaneous carbon dioxide and sensor current monitoring. *Am J Vet Res.* 1993;54:476–480.

21. Orenstein A, Mazkereth R, Tsur H. Mapping of the human body skin with the transcutaneous oxygen pressure method. *Ann Plast Surg.* 1988;20:419–425.

22. Takiwaki H, Nakanishi H, Shono Y, et al. The influence of cutaneous factors on the transcutaneous pO\(_2\) and pCO\(_2\) at various body sites. *Br J Dermatol.* 1991;125:243–247.

23. Takiwaki H. Transcutaneous PO\(_2\) and PCO\(_2\) measurement in dermatology. *Acta Derm Venereol Suppl (Stockh).* 1994;185:21–25.

24. Wolff KD, Kolberg A, Mansmann U. Cutaneous hemoglobin oxygenation of different free flap donor sites. *Plast Reconstr Surg.* 1998;102:1537–1543.

25. Caselli A, Latini V, Lapenna A, et al. Transcutaneous oxygen tension monitoring after successful revascularization in diabetic patients with ischaemic foot ulcers. *Diabet Med.* 2005;22:460–465.