Clearing the undergrowth: detection and quantification of low level impurities using $^{19}$F NMR†

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A new method for the analysis of low level impurities in sparsely fluorinated species allows measurement of clean high dynamic range $^{19}$F spectra, fully decoupled and free of interfering signals from $^{13}$C isotopomers.

The high sensitivity and wide chemical shift range of $^{19}$F NMR make it potentially very attractive for characterising fluorine-containing impurities. In pharmaceutical chemistry, for example, a quarter of current drugs contain one or more fluorines, and regulatory authorities require all impurities above 0.1% of a main active pharmaceutical ingredient to be identified and quantified. Both 1D $^{19}$F NMR and $^{19}$F DOSY have been used for the detection of minor fluorinated impurities. One major technical problem is the difficulty of exciting quantitatively the very wide chemical shift range of $^{19}$F, but solutions now exist for both 1D and DOSY experiments. However, there remains the problem of $^{13}$C isotopomer signals. At around 0.54% of the intensity of $^{13}$C isotopomer signals, these are in the same range as impurity signals of interest and often have similar chemical shifts, and therefore complicate their identification and quantitation. The obvious solution is to use broadband $^{13}$C decoupling to collapse the heteronuclear $J$-couplings. This can work well for $^1$H spectra, albeit at the expense of some sample heating. However, $^{19}$F is quite sensitive to chemical environment and its large secondary isotope shift means that the decoupled ($^{19}$F–$^{13}$C) signals have slightly different chemical shifts from the parent ($^{19}$F–$^{12}$C) signals, so decoupling just halves the number of $^{19}$F–$^{13}$C signals, rather than hiding them all under the parent. Here we show how to acquire clean $^{19}$F spectra without interference from $^{13}$C isotopomers and with no heteronuclear ($^1$H or $^{13}$C) splittings. The new method does not use $^{13}$C decoupling, minimising sample heating, and should greatly facilitate the detection and quantification of low-level impurities by $^{19}$F NMR.

Fig. 1 shows $^{19}$F spectra of a slightly degraded sample of rosuvastatin (1, Scheme 1), used for treating dyslipidaemia, spiked with small amounts of precursors 2 and 3. The proton-decoupled spectrum of Fig. 1a (multiplet structure renders the proton coupled spectrum, shown in Fig. S4 of the ESI, uninformative) is complicated by the presence of both one-bond and long-range $^{13}$C satellites; one of the two satellite signals due to the presence of $^{13}$C at the ortho position with respect to fluorine is almost degenerate with (8 ppb from) the signal of 2.

Acquiring a spectrum with this resolution with full broadband decoupling is uncomfortably close to the limits of many instruments, because of the long high-power irradiation required, but if the one-bond $^{13}$C satellite signals are suppressed (see Section S1 of the ESI), low power irradiation can be used to decouple the remaining longer-range (≥ two-bond) couplings. This gives the spectrum of Fig. 1b, in which a singlet signal is seen for the 2.2% of ortho-$^{13}$C 1. Had full $^{13}$C decoupling been used, the ipso-$^{13}$C signal of 1, midway between the one-bond satellites in Fig. 1a, would have been degenerate with that of impurity 1a (a diastereomer). In the spectrum of Fig. 1c, in contrast, which was obtained with the new method, no resolvable signals at all are seen from $^{13}$C isotopomers, and there is no interference with the signals of the minor components of the sample.

The new method, using the pulse sequence of Fig. 2, is compatible with several different hardware configurations; the results shown here used a single high band radiofrequency (RF) amplifier and a ($^1$H/$^{19}$F),$^{13}$C triple-resonance probe with a double-tuned high band coil. The experiment consists of three parts: a low-pass filter to suppress one-bond $^{13}$C satellite signals; a $J_{CF}$– modulated spin echo; and time-shared acquisition during which the $^{19}$F signal is recorded under $^1$H decoupling. The low-pass $J$ filter, which converts $^{19}$F antiphase signals into unobservable heteronuclear multiple quantum coherences when $A = 1/(2J_{CF})$, suppresses the one-bond $^{13}$C satellite signals. Since a $^{19}$F spin echo is needed to refocus the

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† Electronic supplementary information (ESI) available. See DOI: 10.1039/c6cc08836h
fluorine chemical shift, there is time to use two $^{13}$C $90^\circ$ pulses in a two-stage filter; if a wide range of $^{13}$JC$_{F}$ values is present, further stages can be added.

The modulated spin echo, which is analogous to a heteronuclear 2D $^{1}$H resolved experiment, makes the phases of the remaining $^{13}$C satellite signals depend on the evolution time $t_1$, while the desired signals from the $^{12}$C isotopomers are unaffected. Weighted averaging of experiments with different $t_1$ cancels the modulated signals, leaving a clean spectrum. In practice the most effective way to perform this averaging is by double Fourier transformation and integral projection onto $F_2$ of the $F_1$ range spanned by the lineshape of the parent signal. This suppresses all satellite signals that would be resolvable in the 1D spectrum, while preserving the quantitative character of the spectrum. The final $^{13}$C $90^\circ$ pulse deals with the problem of the phasetwist lineshape of a 2D spectrum by suppressing the sine-modulated dispersive part of the signal. The remaining cosine-modulated signal can then be selected by zeroing the imaginary component after the first Fourier transformation, leading to signals that are doubled in $F_1$ but have 2D absorption mode lineshapes. The choice of increment $1/\text{sw}_1$ in $t_1$ is determined by the range of couplings to be suppressed ($\text{sw}_1 > |J_{CH}|$, and the number of increments $n_i$ by the $T_2$ of the parent signal ($n_i > \text{sw}_1 T_2$). Relaxation losses during $t_1$ lead to a small sensitivity penalty for the new method, about a factor of 2 here (apparent on comparing Fig. 1a and c).

The data acquisition section of the pulse sequence uses time-shared decoupling because the $^{1}$H and $^{19}$F channels share the same coil in the probe used. In normal circumstances, a simple WALTZ$^{27,28}$ or similar decoupling waveform would suffice to decouple $^{1}$H from $^{19}$F, but the very high dynamic range of the sample means that the weak systematic signal modulations such methods induce would here give rise to significant decoupling sidebands (see Fig. S3, ESI$^\dagger$). These are suppressed very effectively here by the use of bivel level adiabatic decoupling.$^{29}$

As well as decoupling $^{1}$H from $^{19}$F during acquisition, it can be helpful to decouple in the earlier parts of the sequence, to suppress any echo modulation caused by strong $^{1}$H–$^{1}$H coupling. This is common in aromatic spin systems (as for example in Fig. S2 of the ESI$^\dagger$). Here the quality of decoupling is less critical, so bivel level decoupling is not needed.
shown in Fig. 3 for the antifungal drug fluconazole, which has an analogue of Fig. 2 to excite the regions around $107$ and $-111$ ppm respectively, revealing the degradation products $4a$, $4b$ and $4c$.

13C isotopomer signals can pose significant challenges in identifying and quantifying impurities down to the 0.1% level. The novel approach introduced here of filtering out, rather than decoupling, these signals offers the possibility of acquiring clean, high dynamic range 19F spectra without interference from species containing 13C. A slightly simpler approach can be used in proton spectra.

This work was supported by AstraZeneca and by the Engineering and Physical Sciences Research Council (grant number EP/ N033949/1). All raw experimental data, and the pulse sequence code, can be downloaded from DOI: 10.15127/1.304823.

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