SILAM for quantitative proteomics of liver Akt1/PKBα after burn injury

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Received October 6, 2011; Accepted November 17, 2011

DOI: 10.3892/ijmm.2011.861

Abstract. Akt1/protein kinase Bα (Akt1/PKBα) is a downstream mediator of the insulin signaling system. In this study we explored mechanism(s) for its role in burn injury. Akt1/ PKBα in liver extracts from mice with burn injury fed with \(^{(2}H_{2})\)-L-Leu was immunoprecipitated and isolated with SDS-PAGE. Two tryptic peptides, one in the kinase loop and a control peptide just outside of the loop were sequenced via nano-LC interfaced with quadruple time-of-flight tandem mass spectrometry (Q-TOF tandem MS). Their relative isotopologue abundances were determined by stable isotope labeling by amino acids in mammalian (SILAM). Relative quantifications based on paired heavy/light peptides were obtained in 3 steps. The first step included homogenization of mixtures of equal amounts of tissue from burned and sham-treated animals (i.e., isotope dilution) and acquisition of uncorrected data based on parent monoisotopic MS ion ratios. The second step included determination of isotopic enrichment of the kinase from burned mice on Day 7 and the third step enrichment correction of partially labeled heavy and light monoisotopic ion ratios for relative quantification of bioactivity (loop peptide) and expression level (control peptide). Protein synthesis and enrichment after injury were found to be dependent on tissue and turnover of individual proteins. Three heavy and light monoisotopic ion ratios for albumin peptides from burned mice indicated ~55% enrichment and ~16.7-fold downregulation. In contrast, serum amyloid P had ~66% enrichment and was significantly upregulated. Akt1/PKBα had ~56% enrichment and kinase level in response to the burn injury was upregulated compared with the control peptide.

However, kinase bioactivity, represented by the Cys\(^{206}\) peptide, was significantly reduced. Overall, we demonstrated that i) quantitative proteomics can be performed without completely labeled mice; ii) measurement of enrichment of acyl-tRNAs is unnecessary and iii) Cys\(^{206}\) plays an important role in kinase activity after burn injury.

Introduction

Stable isotope labeling by amino acids in cell culture (SILAC) provides relative quantification of in vitro protein synthesis and functional proteomics under conditions that mimic disease states (1-5). Typically, two cell populations are cultured for six doublings times; control cell medium contains the natural amino acid (e.g., \(^{15}\)C-Lys and/or \(^{15}\)C-Arg, 99% natural abundance), and the second medium contains the same levels of amino acids with heavy isotopes (e.g., \(^{13}\)C-Lys and/or \(^{13}\)C-Arg, 98% abundance) and disease related stimuli. The two cell populations are mixed with equal amounts of total protein and digested peptides are accurately measured by mass spectrometry with a mass difference of 6 Da for singly charged parent ions. Six doubling times allows isotopic enrichment of the precursor acyl-tRNA pool to reach ~98% in cancer cell lines. Labeling above 95% is generally required for comparative and quantitative proteomics by MS. Thus, the relative abundance of any paired peptide's monoisotopic MS ion chromatogram with SILAC can be used to measure protein synthesis in comparison with the controls under in vitro conditions. The SILAC approach has been used with C. elegans fed with \(>98\%\) \(^{15}\)N-labeled E. coli (6), skeletal muscle from chickens fed with a synthetic diet containing 50% of \(^{(2}H_{2})\)-Val (7,8), partially labeled rat diet with >99% ape \(^{15}\)N algal cells for 44 days (9) and FI-F4 offspring of mice fed with a Lys-free diet containing 1% of L-\(^{13}\)C-Lys (10). Complete labeling has been reported to be achieved by the F2 generation. Metabolic labeling with \(^{15}\)N can be performed efficiently and economically, however, data interpretation can be challenging since the monoisotopic peak can shift, due to the distribution of positional isotopomers as a function of labeling time (11,12). Global labeling with \(^{15}\)N has been used as a tool for characterization of enrichment under partially labeling conditions (13-15). Stable isotope-labeled amino acids, such as L-\(^{15}\)C-Lys and L-\(^{13}\)C-Arg, provide ideal residues at C-terminal labeling positions for trypsin digestion. Protein synthesis depends on 2 factors: acyl-tRNA levels and protein turnover rate. These factors are tissue, cell type, time

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Abbreviations: Akt1/PKBα, Akt1/protein kinase Bα; CID, collision induced dissociation; DDA, data-dependent acquisition; FSR, fractional synthesis rate; MS/MS, mass spectrometry/mass spectrometry; PTM, post-translational modification; Q-TOF tandem MS, quadruple time-of-flight tandem mass spectrometry; SILAM, stable isotope labeling by amino acids in mammalian; TBSA, total burn surface area

Key words: Akt1/PKBα, SILAM, MS/MS, burn injury
and treatment dependent. To minimize individual variability, full incorporation of L-\(^{13}\)C\(_6\)-Lys can be achieved in mice; however, it is very costly.

Analysis of the hypermetabolic/inflammatory response under acute phase conditions is very challenging for several reasons: i) significant changes in protein expression are associated with high levels of reactive oxygen species (ROS) and post-translational protein modifications (PTM); i.e. not only protein levels but also biological activities have to be quantified; ii) individual protein enrichments and incorporated isotope distributions may vary with the partially isotope enriched precursor t-RNA pool (\(\geq50\%\)) and corresponding protein turnover rates during the acute phase response.

The metabolic alterations produced by stresses such as burn trauma are associated with a hypermetabolic/inflammatory state, that includes: increased protein catabolism (with resulting muscle wasting) and insulin resistance. Muscle wasting can lead to muscle weakness that can result in hypoventilation, prolongation of dependence on mechanical ventilation, prolonged rehabilitation and even death (16-19). Insulin resistance is a well established state in critically ill patients and plays a major role in metabolic derangements. Binding of insulin to its receptor (IR) activates the IR tyrosine kinase, which then phosphorylates IR substrates (IRSs). Phosphorylations of IRS1 and IRS2 transfer the signal from IR to phosphatidylinositol-3-kinase (PI3-kinase) (19,20). PTMs of the insulin signaling system are considered to be major disease-dependent events that regulate glucose transport via GLUT-4 translocation and downstream protein synthesis (21-27).

Akt1/PKB\(\alpha\) is a critical downstream mediator of the IR/IRS/PI3-kinase pathway of the insulin signaling system (28-32). Akt1/PKB\(\alpha\) consists of three structural features: the N-terminal pleckstrin homology (PH) domain, a large central kinase domain and a short C-terminal hydrophobic motif. High specific binding of the PH domain with membrane lipid products of PI3-kinase recruits Akt1/PKB\(\alpha\) to the plasma membrane where phosphorylations of Thr\(^{308}\) (kinase domain) and Ser\(^{373}\) (hydrophobic motif) occur. Phosphorylation of Thr\(^{308}\) partially stimulates kinase activity; however, additional phosphorylation of Ser\(^{473}\) is required for full activity. Akt1/PKB\(\alpha\) activation is associated with a disordered to ordered transition of a specific \(\alpha\)C helix via an allosteric mechanism. A salt bridge between the side-chain of Lys\(^{297}\) and the phosphate group of pThr\(^{308}\) in this \(\alpha\)C helix contributes to an ordered activation segment from DFG\(^{292}\) to APE\(^{319}\) (33-36). Reversible dephosphorylations of Thr\(^{308}\) and Ser\(^{373}\) by protein phosphatase 2A (PP2A) and PH domain leucine-rich repeat protein phosphatase (PHLPP\(\alpha\)) also occur in the Akt1/PKB\(\alpha\) activation/deactivation cycle (37-40).

In addition to the role of reversible phosphorylation/dephosphorylation in the regulation of Akt1/PKB\(\alpha\) activity, this kinase is also reversibly inactivated by S-nitrosylation under conditions that result in a persistently increased production of NO (28,41-43); such as after burn injury as previously reported (44). Thiol titration and NMR data have indicated that a disulfide bond (Cys\(^{297}\)-Cys\(^{311}\)) exists in the kinase PH domain (45). A second disulfide bond in the kinase activation loop (Cys\(^{297}\)-Cys\(^{311}\)) has been reported to be associated with dephosphorylation under oxidative stress \textit{in vitro} (46). In addition, it has been shown that Cys\(^{272}\) of Akt1/PKB\(\alpha\) is mutated to a Ser residue, the kinase becomes resistant to NO donor-induced S-nitrosylation and inactivation; suggesting that this residue is a major S-nitrosylation acceptor site (43). \textit{In vivo} S-nitrosylations of the insulin receptor (\(\beta\)) and Akt1/ PKB\(\alpha\) result to reductions in their kinase activities (42). These data suggest that the redox status of Akt1/PKB\(\alpha\), regulated by NO, is a second factor in post-translational modifications that modulate kinase activity (via dynamic conformational changes) and thus GLUT-4 trafficking and protein synthesis.

Recent technical developments have made it feasible to study the molecular mechanisms of these important processes with quantitative proteomics. These techniques include: i) sensitive and site-specific procedures for the detection of S-nitrosylation based upon nano-LC interfaced with tandem MS/MS methods (47,48); ii) the Biotin-Switch method for thiol state discrimination between free, disulfide bonded and S-nitrosylated cysteine residues (49-54); problems associated with quantitative analysis by this technique have been discussed previously (48); iii) stable isotope labeling by amino acids in mammals (SILAM) with partially \(^{13}\)N-labeled rats and completely L-\(^{13}\)C\(_6\)-Lys-labeled mice, and iv) highly specific anti-Akt1/PKB\(\alpha\) monoclonal antibodies that can be used to immunoprecipitate the protein and yield clear SDS-PAGE bands with Coomassie Brilliant Blue R-250 staining.

Burn injury impairs IRS1 mediated signaling and attenuated IR-IRS-PI3K-Akt/PKB activation has been the focus of research studies at our Institute (22,24,26,28,41,43,44,47,50,57). Significantly reduced phosphorylations of Ser\(^{373}\) and Thr\(^{308}\), as well as Akt/PKB kinase activity were observed under burn injury [55% total burn surface area (TBSA), Day 3] and insulin simulation (41). Cys\(^{296}\) is found to be capable of capturing free radicals, such as NO, produced under burn injury (50). However, quantification of Cys\(^{296}\) PTM in the kinase loop remains a challenging issue.

To address these considerations and explore the feasibility of the basic SILAC approach in animals under acute phase and partial isotope labeling conditions, a mouse model with full-thickness 40% total burn surface area (TBSA) burn was used to proteomically characterize liver Akt1/PKB\(\alpha\) from the perspectives of both protein expression and biological activity. (Isopropyl-\(^{2}\)H\(_4\))-L-Leu was selected for labeling based on 4 considerations: i) L-Leu is an essential amino acid with high abundance in proteins. ii) compared with other amino acids, (isopropyl-\(^{2}\)H\(_4\))-L-Leu is relatively inexpensive; iii) none of the 7 deuterium atoms of (isopropyl-\(^{2}\)H\(_4\))-L-Leu are attached at the \(\alpha\)-carbon which may be exchangeable with hydrogen atoms; thus in contrast to studies with (\(^{13}\)C\(_6\))-L-Leu, MS offset can be eliminated as a confounding variable; iv) the hydrophobicity of L-Leu reduces false negative discovery during C18 reversed phase trapping and desalting.

**Materials and methods**

*Chemicals.* Acetonitrile (ACN, LC-MS Chromosolv), formic acid (FA), glacial acetic acid, LC-MS grade water, dithiothreitol (DTT), (Glu\(_4\))-fibrinopeptide B, N,N-dimethylformamide (DMF), were obtained from Sigma Chemical Co. (St. Louis, MO). SDS-PAGE ready gels (12% Tris-HCl, #161-1102), Laemmli sample buffer (#161-0737) and Coomassie Brilliant
Blue R-250 (#161-0436) were obtained from Bio-Rad. The trypsin profile IGD kit (#PP0100) was obtained from Sigma. Anti-Akt1/PKBα monoclonal antibody (cat #05-796, Lot #26860) and inactive Akt1/PKBα (cat #14-279) were purchased from Upstate (Charlottesville, VA). Immobilized monomeric avidin beads were a product of Pierce Chemical Co., Rockford, IL (cat #20228). Iodoacetyl-LC-biotin (cat #21333) were purchased from Pierce.

**Blood sampling.** The mice were anesthetized with ether inhalation, and blood was drawn from the retro-orbital plexus. The blood samples were kept on ice and after centrifugation, serum samples were frozen in liquid nitrogen and stored at -80°C.

**Burn injury.** Male CD-1 mice (Charles River, Wilmington, MA) weighing 25-30 g were used throughout this study. The Institutional Animal Care Committee approved the study protocol. The animal care facility is accredited by the Association for Assessment and Accreditation of Laboratory Animal Care. A standardized thermal injury consisting of full-thickness scald involving 40% of the body surface area was performed on the mice (n=3) using previously described methods (55). Briefly, after anesthesia with intraperitoneal ketamine (60 mg/kg) and xylazine (1.3 mg/kg), the animals were shaven, and placed in a mold on top of a boiling water bath for 10 sec, exposing the back to a 40% body surface area full-thickness burn injury. Each mouse was given an intraperitoneal injection of saline (50 ml/kg) to replace intravascular volume known to be lost after thermal injury. Sham-treated (n=3, control) mice were anesthetized, confined to the mold without burning, dipped in water at room temperature, given saline resuscitation, and otherwise treated in the same fashion as the injured animals.

The routine protocol for pain management was 72 h of scheduled analgesia followed by p.r.n. analgesia based on overt signs of distress. This procedure was viewed as adequate pain management for this potentially painful procedure. The analgesic was buprenorphine (0.05 mg/kg s.c.) and/or a topical agent (EMLA cream) until discomfort disappeared. The buprenorphine was given 10-15 min prior to burn.

**Diets.** The diet with depletion of L-Leu was obtained as L-amino acids defined diet for rats and mice. (Dyets, Inc., Bethlehem, PA; Dyet #510133, meets the 1995 NRC nutrient requirements). For producing the light diet, natural L-Leu was added to the diet powder (1% by weight) and producing the heavy diet, the same amount of (isopropyl-2H)-L-Leu was added to the diet powder. For both diets, a minimal amount of water was added to make biscuits of adequate size. The biscuits were dried at room temperature for 1 week as suggested by Dyets in order to maintain the original nutrients.

**Metabolic labeling with (isopropyl-2H)2-L-Leu and natural L-Leu diets.** Burned and sham-treated mice were caged separately and maintained in a temperature controlled facility with a 12-h light/dark cycle. Burned mice were fed with the diet containing (isopropyl-2H)2-L-Leu and sham-treated mice fed with the diet containing natural L-Leu. Five grams of light and heavy diets were provided daily to each mouse for 7 days. The animals had ad libitum access to water.

**Tissue homogenization.** Equal amounts of tissue from burned and sham-treated animals were mixed and homogenized in cell lysis buffer (Cell Signaling Technology, cat #9803, 5 ml/g tissue) containing phenylmethanesulfonyl fluoride (BioChemika, 2 mM, freshly prepared 100X stock in ethanol just prior to use) on ice with 30 strokes. The homogenates were sonicated for 2 min at 4°C, followed by 3 freeze-thaw cycles in liquid nitrogen and 4°C. The homogenates (5 ml) were extracted with chloroform (1 ml) with vortexing for 15 sec and centrifuged at 14,000 x g for 2 min. Supernatants were collected and re-centrifuged at 14,000 x g for 2 min in order to remove traces of chloroform. The final supernatants were diluted with PBS to a final volume of 7 ml, and then passed through a membrane filter (0.22 µm, Millipore, cat # SLFG025NS). All tissue processing was performed at 4°C.

**Immunoprecipitation.** Protein G bead slurry (50%, 100 µl) was transferred into an Eppendorf vial (1.5 ml), and washed 2 times with PBS at 4°C by gently rotating for 1 min. The packed beads (50 µl) were collated by centrifuging at 3000 x g (5000 rpm) for 20 sec. PBS (50 µl) was added to the beads to produce a final immunoprecipitation volume of 100 µl. Anti-Akt1 monoclonal antibody (5 µg, Upstate cat #05-796, clone AW24) was added and the mixture was gently stirred at 4°C for 1 h. Without washing the beads, processed homogenate was added and immunoprecipitation was performed for 2 h with gently rotating at 4°C.

**Biotinylation.** Laemmli sample buffer (50 µl, 2X, pH adjusted to 8.0, Bio-Rad, cat #161-0737) was added to the washed immunocomplex beads (packed 50 µl, washed with 1 ml PBS 3 times for 5 min/cycle). This was followed by addition of freshly prepared iodoacetyl-LC-biotin solution (10 µl, stock solution; 2 mg in 1 ml of DMF). Cysteine acylation was performed at room temperature for 15 min with stirring. The reaction was quenched by addition of 2-mercaptoethanol (5 µl). The beads were then heated at 95°C for 5 min and kept at room temperature for 30 min prior to loading on SDS-PAGE gels.

**SDS-PAGE isolation and in-gel digestion.** SDS sample solution (15 µl) was added to ready gel wells (Bio-Rad, 12% gel, cat #161-1102). SDS-PAGE was performed with Tris/Glycine/SDS running buffer (Bio-Rad, cat #161-0732) at 200 V for 45 min. The gels were removed and washed with deionized water for 5 min on a rocking platform and excess water was discarded. Proteins were stained with Coomassie Brilliant Blue R-250 for at least 1 h with gently shaking. The gels were washed three times with destaining solution (acetic acid:methanol:DH2O = 10:40:50) for 2 h. Protein bands were excised as 1x1 mm pieces, and placed into siliconized polypropylene Eppendorf tube (1.5 ml). In-gel digestion was performed with a Sigma Trypsin Profile IGD kit (cat #P0100) according to the manufacturer’s protocol.

**Avidin purification.** Immobilized monomeric avidin beads (30 µl, 50% aqueous slurry, Pierce, cat #20228) were placed in siliconized polypropylene Eppendorf tubes (0.6 ml), and washed with PBS. The digested peptides (70 µl) were added to the packed avidin beads (15 µl) and the mixture was placed on a rocking platform for 30 min to capture the biotinylated peptides. The supernatant was collected, dried via Speed-Vac,
Table I. Protein enrichment levels on Day 7 after third degree burn of 40% total burn surface area.

| Sequence, (charge state observed) | Enrichments %, (SD) |
|-----------------------------------|---------------------|
| Albumin                           | 264 SQTFPNADFAEITK²⁵⁷, (2) 53 (3.2) |
|                                   | 55⁹ HKPKATAEQKL⁶⁶⁹, (2) 57 (3.8) |
|                                   | 43⁹ APQVSTPTLVÆAR²⁵², (2) 54 (4.1) |
| Amyloid P component               | 6⁶ SQSIFSVSK⁷⁵, (2) 66 (4.9) |
|                                   | 8⁸ GVEYLGYGQ⁹⁹, (2) 65 (4.6) |
|                                   | 14⁷ APPSIVLQEQDNYGGFQR¹⁶⁶, (3) 69 (5.2) |
| Liver Akt1/PKBα                    | ⁹ EGWLHKR⁰⁰, (2) 59 (4.0) |
|                                   | 1⁸⁴ EVIVAKDEVAHTLTENR²⁰⁰, (3) 56 (2.7) |
|                                   | ²⁹⁰ TDFGLB*KEGIFGATMK³⁰⁷, (3) 55 (3.3) |

B*, 2-mercaptoethanol derivatized cysteine; L, partially labeled L-Leu residues.

and resuspended in mobile phase A (as described below, 15 µl) for control peptide <sup>29⁰</sup>FYGAEIYSALDYLHSEK<sup>2⁸⁸</sup> analysis. The beads were then washed with PBS (200 µl x 3), followed by ACN/water (10/90 = v/v, 200 µl x 3). The biotinylated peptides were recovered by addition of formic acid (30 µl, 70%) and gently rocking for 5 min at room temperature. This recovery step was repeated three times and the supernatants were combined, dried via Speed-Vac, and resuspended in mobile phase A (15 µl) for biotinylated loop peptide <sup>2⁹⁰</sup>TDFGLCK<sup>²⁸⁷</sup> analysis.

**LC-MS/MS analysis.** All experiments were performed using a Waters CapLC-Q-TOF<sup>®</sup> system (Waters Corporation, Milford, MA). An analytical column (75 mm ID x 150 mm, C18 PepMap300, 5 mm, LC Packings) was used to connect the stream select module of the CapLC with the voltage supply adapter for ESI. Peptide mixtures were loaded onto the precolumn (C18 resin) at a flow rate of 15 µl/min. Dead volume from the CapLC injector to the precolumn was measured to be ~1.5 µl. After washing with mobile phase C (auxiliary pump, 0.1% formic acid in water/ACN, 2% ACN) for 2 min, the trapped peptides were back washed from the precolumn onto the analytical column using the 10-position stream switching valve. Freshly prepared mobile phases A and C were sonicated under vacuum for about 25 min and mobile phase B was treated in this way for 5 min. The mobile phases were degassed every week, and the CapLC pumps were wet primed for 20 cycles. A linear gradient was used to elute the peptide mixture from mobile phase A (0.1% FA in water/ACN, 2% ACN) to mobile phase B (0.1% FA in ACN). The linear gradient was segmented as follows: isocratic elution with 2% B for 3 min, increasing B from 10% to 70% (3-40 min) and isocratic 70% B for 5 min. Mobile phase B was then reduced to 2% over 2 min. The injector syringe (25 µl) was washed with degassed mobile phase A and the injection volume was set as full loop mode (10 µl). The gradient flow rate was set at 1.5 µl/min before the 16/1 Nanotee splitter and the pressure drop from the analytical column was about 800 psi. The pressure drop (or the flow splitting ratio) was adjusted and maintained with 20 µm ID capillary tubing at the waste outlet position of the Nanotee splitter. The gradient flow rate was ~95 nl/min. The electrospray voltage was set to ~3000 V to obtain an even ESI plume at the beginning of the gradient (high water content). As a routine sensitivity check, the PicoTip Emitter position and other parameters were adjusted to achieve ~45 counts/sec for the capillary tubing background peak (m/z 429). Sample cone and extraction cone voltages were set at 45 and 3 V, respectively. The instrument was operated in a positive ion mode with the electrospray source maintained at 90°C. The instrument was calibrated with synthetic human (Glu<sup>-</sup>)-fibrinopeptide B (100 fmol/µl in ACN/water = 10:90, 0.1% formic acid, v/v) at an infusion rate of 1 µl/min in TOF MS/MS mode. The peptide was selected at m/z = 785.8 and focused into the collision cell containing argon gas at ~3x10<sup>-5</sup> Torr; the collision energy was set at 35 V. Instrument resolution for the (Glu<sup>-</sup>)-fibrinopeptide B parent ion, m/z = 785.84, was found to be 5,250 FWHM. All data were acquired and processed using MassLynx 4.1 software. Data-dependent acquisition (DDA) was set from m/z 630 to 660 for the biotinylated (doubly charged m/z 639.83) and control peptides (triply charged m/z 647.99). Scan time was ~1.9 sec and the inter-scan delay was 0.1 sec. MS to MS/MS switch criteria were dependent on reporter ion intensity (3 counts/sec) and detection window (2.3 Da, charge status). The instrument was switched from MS/MS back to MS after 5 sec without intensity restriction.

**Evaluation of the biotinylated Cys<sup>²⁹⁶</sup> site.** In contrast to the highly abundant proteins in serum, biotinylated Cys<sup>²⁹⁶</sup> is expressed at very low levels after burn injury and site-specific pinpointing requires careful evaluation of low S/N spectra. Confirmations of the biotinylated Cys<sup>²⁹⁶</sup> residue were performed by the following three step procedure: i) for parent ion discoveries a narrow MS survey scan was used in order to increase MS sensitivity. Under these conditions, only a few false positive ions were observed and these were eliminated manually. ii) The positively discovered parent ions were analyzed with PepSeq of MassLynx V4.1 software. iii) For the loop peptide, with MS/MS scores <35 and S/N ~3, manual interpretations of candidate parent ions were performed with the following procedure: continuum MS/MS spectra were obtained and the upper 80% was centroided in order to obtain the sequence data associated with biotinylated y ions with expected mass shifts.
Cysteine residue monoisotopic mass $C_3H_5NOS = 103.01$ Da was replaced with the iodoacetyl-LC-biotin derivatized adduct monoisotopic mass $C_{21}H_{35}N_5O_4S_2 = 485.21$ Da.

**Results and Discussion**

The essential issue for any isotope dilution method is to precisely characterize the heavy isotope-labeled internal standard in terms of atomic and chemical purities. To determine the relative quantification of liver Akt1/PKBα after burn injury, two typical serum acute phase proteins, negative regulated albumin and positive regulated amyloid P component, as well as liver Akt1/PKBα were characterized in a preliminary study with tissue and blood samples from 3 mice with burn injury (40% TBSA). To eliminate possible mathematical and biological complexities associated with multiple isoto-
pomer population distributions for individual tryptic peptides produced with partial labeling conditions, peptides with only one instance of (isopropyl-\(^2\text{H}_7\))-L-Leu were selected for measurement of relative labeling efficiencies. A tryptic peptide may be positively charged at its N-terminal \(\alpha\)-amine group and the side-chain amine group of a Lys residue or the guanido group of an Arg residue; thus, a doubly charged parent ion, \((M+2\text{H}^+\rangle\), may be observed under ESI mass spectrometry. In addition, triply charged tryptic parent ions, \((M+3\text{H}^+\rangle\), may be obtained for peptides containing proline or histidine residues or with larger size peptides. Peptides with one instance of (isopropyl-\(^2\text{H}_7\))-L-Leu incorporation yield paired light and heavy parent ions with monoisotopic mass ion differences of 3.5 Da (doubly charged) or 2.3 Da (triply charged) under ESI conditions. Charge state dependent DDA allows these multiply charged peptides to be focused in the CID chamber under charged argon cleavages. The singly charged and predominantly light and heavy MS/MS y ion series from the light and heavy parent ions are produced with mass difference of 7 Da. This allows unambiguous relative quantification of parent ion enrichments from possible false positive discoveries. The enrichments of three proteins are shown in Table I.

The negative acute phase protein albumin and the positive acute phase protein amyloid P component had enrichments of ~55 and ~66\%, respectively (Table I). The enrichment level (56\%) of liver Akt1/PKB\(\alpha\) was found to be very similar to that of albumin. These enrichment values represent ultimate (isopropyl-\(^2\text{H}_7\))-L-Leu incorporation in each individual protein which can be used as the isotopic correction factor for the light and heavy parent ion MS ratio obtained by mixing exactly the same weight of labeled and non-labeled liver tissues (Fig. 1).

The enrichments of albumin (Table I), implies that approximately equal amounts of the ion at m/z 723.97 come from burned and sham animal sera. Thus, the ion intensity of 2680 under the m/z 720.37 peak needs to be subtracted (enrichment correction) by about the same ion intensity at m/z 723.97, i.e., the corrected m/z 720.37 is 2529. The corrected monoisotopic ion ratio, m/z 720.37: m/z 723.97 = 16.7 based on the enrichment listed in Table I, indicates that albumin was downregulated by 16.7 fold after burn injury.
band from serum (2 µl) from sham-treated animals. In this case, relative quantification failed due to lack of a control peptide; suggesting amyloid P component is upregulated significantly after the burn injury.

Akt1/PKBα was immunoprecipitated from the mixture of (isopropyl-²H₇)-L-leucine-labeled liver from burned mice an equal amount tissue from sham-treated animals and processed further by the methods described above. The relative expression of Akt1/PKBα from livers of burned and sham-treated mice (Fig. 4). The triply charged monoisotopic parent ion at m/z 647.99 was detected for the control peptide SFYGAIVSALDYLHSEK₂⁶⁶, calculated (M+3H⁺) = 647.99. Two heavy monoisotopic parent ions at m/z 650.34 and 652.36 indicate one and two instances of (isopropyl-²H₇)-L-Leu incorporation into the control peptide. Akt1/PKBα protein expression, estimated from both heavy ions, was found to be upregulated after correction for both enrichment and background.

Figure 3. Relative quantification of the upregulation of amyloid P component. Amyloid P component light and heavy peptides (⁶⁵SQSLFSYSVK prol) were identified at m/z 573.27 (sham) and m/z 576.79 (burn) with doubly charged MS difference of 3.52 Da. A monoisotopic MS ion intensity ratio of m/z 573.27: m/z 576.79 = 0.5 demonstrated that there is little contribution of the m/z 573.27 from sham mice after mixing equal volumes of serums (2 µl). This suggests that amyloid P component is significantly upregulated (undetectable in sham-treated mice).

Figure 4. Changes in liver Akt1/PKBα expression after burn injury. A triply charged monoisotopic parent ion at m/z 647.99 was found for the control peptide SFYGAIVSALDYLHSEK₂⁶⁶, calculated (M+3H⁺) = 647.99. Two heavy monoisotopic parent ions at m/z 650.34 and 652.36 indicate one and two instances of (isopropyl-²H₇)-L-Leu incorporation into the control peptide. Akt1/PKBα protein expression, estimated from both heavy ions, was found to be upregulated after correction for both enrichment and background.
poration into the control peptide. Triply charged MS differences were found to be 2.3 Da (calculated MS difference for one heavy Leu incorporation: 7/3=2.3 Da) and 4.3 Da (calculated MS difference for two heavy Leu incorporation: 14/3=4.6 Da), respectively. MS/MS y ion series (from y3 to y12 as shown in Fig. 5) of the m/z 647.98 confirms the control peptide for studying protein expression. The intensities of the m/z 650.34 and 652.36 ions represent the heavy Akt1/PKBα populations from burn injured mice, whereas the m/z 647.99 ion is derived from the sham-treated animals. Estimated total ion intensity was ~9.5 after burn injury and ~6.5 after sham treatment. Due to the low S/N of ~2 for the heavy ions and the enrichment of 56% in Table I, Akt1/PKBα protein expression, was observed to be upregulated in response to burn injury the mice. However, the heavy ion for the biotinylated peptide, 290ITDFGLCK297, which was used as a marker for kinase activity, was significantly reduced (almost to background) after injury (Fig. 6).

The biotinylated light peptides 290ITDFGLCK297 of Akt1/PKBα from sham-treated mice was detected at m/z 639.88 and was doubly charged as indicated by the natural abundance carbon isotope peak at m/z 640.33. Three biotinylated y2, y4 and y6 ions at m/z 632.44, 802.51 and 1064.59 confirmed the loop peptide sequence with biotin modification (Fig. 7). The corresponding y2 ion of iodoacetyl-LC-biotin derivatized, 296CK, was confirmed at m/z 632.44 (expected m/z 632.33 equal to 485.21 + 145.10 + 2.02), DDA with low S/N occurred in vivo studies was performed with a continuum mode to enhance the parent ion discovery. On the other hand, centroided spectra are found to be necessary to obtain accurately the diagnostic modifications in the y ions.

The biotinylated heavy peptide 290ITDFGLCK297 in Akt1/PKBα from burn injured mice (n=3) was identified at m/z 643.38 with its natural carbon isotopic peak at m/z 643.88. An MS difference of 3.5 Da between the paired light and heavy parent ions clearly indicated that one instance of (isopropyl-2H7)-L-Leu was incorporation into the doubly charged parent ion at m/z 643.38 with doubly charged state (n=3). MS ion intensity at m/z 643.38 indicated that loop peptide containing Cys296 was almost undetectable after burn injury. These observations were evaluated with MS/MS sequencing; peptide charge status and MS shift of the isotope-labeled paired peptides in vivo.

The incorporation efficiencies of isotopically labeled peptides in Table I demonstrates that amyloid P component was 10% higher than albumin; suggesting a faster turnover rate during upregulation. Serum albumin and liver Akt1/PKBα appear to have similar enrichments ~55%. Classically, individual protein fractional synthesis rate (FRS) is calculated by the relative isotope enrichment ratio of the labeled protein vs. precursor acyl-tRNA over the labeling time period for a given isotope tracer (56,57). FSR is a very important parameter for assessing effects of clinical interventions by comparisons between patients and healthy controls. In general, information about protein synthesis obtained with FRS and partially labeled SILAM are very similar and either can be used to optimize therapeutic strategies.

Our previous studies with thermally injured rats have demonstrated that there is no apparent alteration in binding of insulin to its receptors in liver, skeletal muscle or adipose tissue (22). Thus, acute and chronic insulin resistances induced by surgical trauma, burn injury, hemorrhage and sepsis are primarily post-receptor effects in the insulin-like growth factor/PI3-kinase/Akt pathway. Phosphorylations of specific Ser and Thr residues in the C-terminus of insulin receptor substrate-1 (IRS-1) induces its degradation via the ubiquitin-proteasome pathway (unpublished data); which may be an early biological effect after receptor binding. Impaired Akt1/PKBα kinase activity after injury may be a later downstream event. Deficiency of Akt1/PKBα causes decreased somatic cell and body size (58), while knockout of Akt2/PKBα leads to insulin resistance (59). Akt1/PKBα is involved in cellular survival
pathways, by inhibiting apoptotic processes and stimulating protein synthesis pathways. It is also a key signaling protein in cellular pathways of skeletal muscle differentiation (60,61).

Currently, assays of Akt1/PKBα activity in vitro and in vivo are performed with antibodies specific for the phosphorylated Ser273 and Thr308 residues which are critical for stabilizing the global and loop active conformations of the kinase. Difficulties with using anti-phospho-Ser, but not anti-phospho-Tyr antibodies, have occurred in our phosphoproteomic research (unpublished data). NO production is elevated after burn injury (n=3).
injury and in patients with type 2 diabetic (61,63) and it has been shown that the Cys297-Cys311 disulfide bond in the kinase loop may be formed in association with dephosphorylation under oxidative stress in vitro (46). Reversible S-nitrosoylation at Cys296 in the kinase loop is another PTM which complements reversible phosphorylation at Thr308 in the regulation of kinase activity (37). It has been shown that Akt1/PKBα undergrades transient phosphorylation/dephosphorylation which regulates the kinase active conformation cycle (37); kinase disulfide bond formation, Cys307-Cys311, and dephosphorylation at pThr308 are induced simultaneously by H2O2 oxidative stress in vitro (46); and high levels of nitric oxide production occurs in both burn injured rats (62) and diabetic patients (63). The free thiol group of Cys296 undergoes loop conformational changes by capture of nitric oxide, or chemical modifications with other reactive oxygen species produced under the burn injury; thus blocking substrate recognition. Intact loop peptide with a trace amount of free cysteine in the peptide population, TTDGFLCK307, after burn injury was developed as an unambiguous index for bioactivity. However, this peptide is not related to kinase protein expression in responses to the burn, since varying degrees of different thiol modifications in the loop may occur at the same time. In contrast, the control peptide, FYGAEIVSALDYLHSEK328, located just outside of the kinase loop, was a useful index of protein level.

Kinase bioactivity measured with the tandem MS was comparable with previously reported data measured with immune complex kinase assay and anti-pThr308 as well as anti-Ser37 monoclonal antibodies (41), whereas protein levels were slightly increased in responses to injury. MS/MS sequence analysis and (isopropyl-2H5)-L-Leu incorporation in the paired peptides indicated that after thermal injury kinase activity is significantly reduced, despite increased protein expression. Overall, our findings indicate that neither complete labeling of nor measurement of acyl-tRNA enrichment are necessary or critical for quantitative proteomics with SILAM. Cys296 thiol state is considered as one of the important factors for the kinase activity. The limitations of partially labeled SILAM for clinical studies are first that specific protein enrichment must be measured in tissues labeled with heavy isotopes under stress conditions; unfortunately, many proteins of clinical interest occur at low abundance and thus there is a high rate of false negative discovery. Second, sequence confirmation of Cys307-Cys311 and dephosphorylation at pThr308 are induced simultaneously by H2O2 oxidative stress in vitro (46); and high levels of nitric oxide production occurs in both burn injured rats (62) and diabetic patients (63). The free thiol group of Cys296 undergoes loop conformational changes by capture of nitric oxide, or chemical modifications with other reactive oxygen species produced under the burn injury; thus blocking substrate recognition. Intact loop peptide with a trace amount of free cysteine in the peptide population, TTDGFLCK307, after burn injury was developed as an unambiguous index for bioactivity. However, this peptide is not related to kinase protein expression in responses to the burn, since varying degrees of different thiol modifications in the loop may occur at the same time. In contrast, the control peptide, FYGAEIVSALDYLHSEK328, located just outside of the kinase loop, was a useful index of protein level.

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Acknowledgements

This study was supported in part by grants from the National Institutes of Health (NIGMS P50 GM21000) and Shriners Hospitals for Children. We would like to thank Dr E.A. Carter and Ms. F. Lin for their assistance with these studies.

References

1. Beynon RJ and Pratt JM: Metabolic labeling of proteins for proteomics. Mol Cell Proteomics 4: 857-872, 2005.
2. Harsha HC, Molina H and Pandey A: Quantitative proteomics using isotope labeling with amino acids in cell culture. Nat Protoc 3: 505-516, 2008.
3. Beynon RJ: The dynamic of the proteome: strategies for measuring protein turnover on a proteome-wide scale. Brief Funct Genomic Proteomic 3: 382-390, 2005.
4. Moresco JJ, Dong MQ and Yates JR: Quantitative mass spectrometry as a tool for nutritional proteomics. Am J Clin Nutr 89: 1095S-1104S, 2008.
5. Nair KS, Jaleel A, Asmann YW, Short KR and Raghavakaimal S: Proteomic Research: potential opportunities for clinical and physiological investigators. Am J Physiol Endocrinol Metab 286: E863-E874, 2004.
6. Krügsveld J, Ketting RE, Mahmoudi T, Johansen J, Artal-Sanz M, Verruiter CP, Plasterk RH and Heck AJ: Metabolic labeling of C. elegans and D. melanogaster for quantitative proteomics. Nat Biotechnol 21: 927-931, 2003.
7. Doherty MK, Whitehead C, McCormack H, Gaskell SJ and Beynon RJ: Proteome dynamics in complex organisms: using stable isotopes to monitor individual protein turnover rates. Proteomics 5: 522-533, 2005.
8. Hayter JR, Doherty MK, Whitehead C, McCormack H, Gaskell SJ and Beynon RJ: The subunit structure and dynamics of the 20s proteosome in chicken skeletal muscle. Mol Cell Proteomics 4: 1767-1781, 2005.
9. Wu CC, MacCoss MJ, Howell KE, Matthews DE and Yates JR III: Metabolic labeling of mammalian organisms with stable isotopes for quantitative proteomic analysis. Anal Chem 76: 4951-4959, 2004.
10. Kruger M, Moser M, Usar S, Thiessen E, Lubner CA, Former F, Schmutz S, Zanivan S, Fassler R and Mann M: SILAC mouse for quantitative proteomics uncovers kindling-3 as an essential factor for red blood cell function. Cell 134: 353-364, 2008.
11. Gouw JW, Tops BB, Mortensen P, Heck AJ and Krügsveld J: Optimizing identification and quantification of 13C-labeled proteins in comparative proteomics. Anal Chem 80: 7796-7803, 2008.
12. Huttlin EL, Hegeman AD, Harms AC and Sussman MR: Comparison of full versus partial metabolic labeling for quantitative proteomics analysis in Arabidopsis thaliana. Mol Cell Proteomics 6: 860-881, 2007.
13. McClatchy DB, Dong, MQ, Wu CC, Venable JD and Yates JR III: 13C Metabolic labeling of mammalian tissue with slow protein turnover. J Proteome Res 6: 2005-2010, 2007.
14. Liao L, McClatchy DB, Park SK, Xu T, Lu B and Yates III J: Quantitative analysis of brain nuclear phosphoproteins identifies developmentally regulated phosphorylation events. J Proteome Res 7: 4743-4755, 2008.
15. Bach A and Bonaldi T: Quantitative proteomics as a new piece of the systems biology puzzle. J Proteomics 71: 357-367, 2008.
16. Biolo G, Fleming KY, Maggi SP, Nguyen TT, Herndon DN and Wolfe RR: Inverse regulation of protein turnover and amino acid transport in skeletal muscle of hypercatabolic patients. J Clin Endocrinol Metab 87: 3378-3384, 2002.
17. Bodine SC, Stitt TN, Gonzalez M, Kline WO, Stover GL, Bauerlein R, Zlotchenko E, Scrimgeour A, Lawrence JC, Glass DJ and Yancopoulos GD: Akt/mTOR pathway is a crucial regulator of skeletal muscle hypertrophy and can prevent muscle atrophy in vivo. Nat Cell Biol 3: 1014-1019, 2001.
18. Bruning JC, Winnay J, Cheatham B and Kahn CR: Differential signaling in insulin receptor substrate 1 (IRS-1) and IRS-2 in IRS-1-deficient cells. Mol Cell Biol 17: 1513-1521, 1997.
19. Carvalho E, Rondinone C and Smith U: Insulin resistance in fat cells from obese Zucker rats - evidence for an impaired activation and translocation of protein kinase B and glucose transporter 4. Mol Cell Biochem 206: 7-16, 2000.
20. Araki E, Lipes MA, Patti ME, Bruning JC, Haag B, Johnson RS and Kahn CR: Alternative pathway of insulin signaling in mice with targeted disruption of the IRS-1 gene. Nature 372: 186-190, 1994.
21. Khoury W, Klausner JM, Benabraham R and Sodol O: Glucose control by insulin for critically ill surgical patients. J Trauma 57: 1132-1138, 2004.
22. Carter EA, Burks D, Fischman AJ, White M and Tompkins RG: Insulin resistance in thermally-injured rats is associated with post-receptor alterations in skeletal muscle, liver and adipose tissue. Int J Mol Med 14: 653-658, 2004.
23. Groeneveld AB, Beishuaizen A and Visser FC: Insulin: a wonder drug in the critically ill? Critical Care 6: 102-105, 2002.
24. Iketu T, Okamoto T, Yonezawa K, Tompkins RG and Martyn IA: Analysis of thermal injury-induced insulin resistance in rodents. J Biol Chem 272: 25289-25295, 1997.
25. White MF: Insulin signaling in health and disease. Science 302: 1710-1711, 2003.

26. Zhang Q, Carter EA, Ma BY, White M, Fischman AJ and Tompkins RG: Molecular mechanism(s) of burn-induced insulin resistance in murine skeletal muscle: role of IRS phosphorylation. Life Sci 77: 3068-3077, 2005.

27. Ishiki M and Klip A: Minireview: recent developments in the regulation of glucose transporter-4 traffic: new signals, locations, and partners. Endocrinology 146: 5071-5078, 2005.

28. Kaneki M, Shimizu N, Yamada D and Chang K: Nitrosative stress and pathogenesis of insulin resistance. Antioxid Redox Signal 9: 1-11, 2007.

29. Song G, Quyang G and Bao S: The activation of Akt/PKB signaling pathway and cell survival. J Cell Mol Med 9: 59-71, 2005.

30. Neels JG and Olefsky JM: A new way to burn fat. Science 312: 1756-1758, 2006.

31. Tian R: Another role for celebrity, Akt and insulin resistance. Circ Res 96: 139-140, 2006.

32. Lawlor MA and Alessi DR: PKB/Akt: a key mediator of cell proliferation, survival and insulin responses? J Cell Sci 114: 2903-2910, 2001.

33. Yang J, Cron P, Thompson V, Good VM, Hess D, Hemmings BA and Barford D: Molecular mechanism for the regulation of protein kinase B/Akt by hydrophobic motif phosphorylation. Mol Cell 9: 1213-1226, 2002.

34. Yang J, Cron P, Good VM, Thompson V, Hemmings BA and Barford D: Crystal structure of an activated Akt/protein kinase B ternary complex with GSK3- peptide and AMP-PNP. Nat Structure Biol 9: 940-944, 2002.

35. Huang X, Begley M, Morgenstern KA, Gu Y, Rose P, Zhao H and Zhu X: Crystal structure of an inactive Akt2 kinase domain. Structure 11: 21-30, 2003.

36. Kumar CC and Madison V: Akt crystal structure and Akt-specific inhibitors. Oncogene 24: 7493-7501, 2005.

37. Fayard E, Tintignac LA, Baudry A and Hemmings BA: Protein kinase B/Akt signaling: AKTion on multiple fronts. Trends Biochem Sci 29: 233-242, 2004.

38. Brazil DP, Park J and Hemmings BA: PKB binding proteins: getting in on the Akt. Cell 111: 293-303, 2002.

39. Huang BX and Kim HY: Interdomain conformational changes in Akt activation revealed by chemical cross-linking and tandem mass spectrometry. Mol Cellular Proteomics 5: 1045-1053, 2006.

40. Sugita H, Kaneki M, Sugita M, Yasukawa T, Yatsuhasha S and Martyn JA: Burn injury impairs insulin-stimulated Akt/PKB activation in skeletal muscle. Am J Physiol Endocrinol Metab 288: E585-E591, 2004.

41. Carvalho-Filho MA, Ueno M, Hirabara SM, Seabra AB, Carvalheira JB, de Oliveira MG, Velloso LA, Curi R and Saad MJ: S-nitrosation of the insulin receptor, insulin receptor substrate 1, and protein kinase B/Akt: a novel mechanism of insulin resistance. Diabetes 54: 959-967, 2005.

42. Yasukawa T, Tokunaga, E, Ota H, Sugita H, Martyn JA and Kaneki M: S-Nitrosylation-dependent inactivation of Akt/protein kinase B in insulin resistance. J Biol Chem 280: 7511-7518, 2005.

43. Carter EA, Derojas-Walker T, Tamir S, Tannenbaum SR, Yu YM and Tompkins RG: Nitric oxide production is intensely and persistently increased in tissue by thermal injury. Biochem J 340: 201-204, 1994.

44. Auguin D, Barthe P, Auge-Senegas MT, Stern MH, Noguchi M and Reumondand C: Solution structure and backbone dynamics of the Pleckstrin homology domain of the human protein kinase B (PKB/Akt). Interaction with inositol phosphates. J Biomed NMR 28: 137-155, 2004.

45. Murata H, Ibarra Y, Nakamura H, Yodoi J, Sumikawa K and Kondo T: Glutaredoxin exerts an antiapoptotic effect by regulating the redox state of Akt. J Biol Chem 278: 50226-50233, 2003.

46. Lu XM, Lu M, Fischman AJ and Tompkins RG: A new approach for sequencing human IRS1 phosphorytosome-containing peptides using CapLC-Q-TOF micro mass spectrometry. J Mass Spectrom 40: 599-607, 2005.

47. Lu XM, Lu M, Tompkins RG and Fischman AJ: Site-specific detection of S-nitrosylated PKB/Akt1 from rat soleus muscle using CapLC-Q-TOF micro mass spectrometry. J Mass Spectrom 40: 1140-1148, 2005.

48. Jaffrey SR, Erdjument-Bromage H, Ferris CD, Tempst P and Snyder SH: Protein S-nitrosylation: a physiological signal for neuronal nitric oxide. Nat Cell Biol 3: 193-197, 2001.

49. Kondo T: Glutaredoxin exerts an antiapoptotic effect by regulating the redox state of Akt. J Biol Chem 278: 50226-50233, 2003.

50. Greco TM, Hodara R, Parastatis I, Heijnen HFG, Dennehy MK, Liebler DC and Ischiropoulos H: Identification of S-nitrosylation motifs by site-specific mapping of the S-nitrosocysteine proteome in human vascular smooth muscle cells. Proc Natl Acad Sci USA 103: 7420-7425, 2006.

51. Kuncewicz T, Sheta EA, Goldkniapf IL and Kone BC: Proteomic analysis of S-nitrosylated proteins in mesangial cell. Mol Cellular Proteomics 2: 156-163, 2003.

52. Martinez-Ruiz A and Lamas S: Detection and proteomic identification of S-nitrosylated proteins in endothelial cells. Arch Biochem Biophys 423: 192-199, 2004.

53. Carter EA, Tompkins RG, Babich JW, Correia J, Bailey EM and Fischman AJ: Thermal injury in rats alters glucose utilization by skin, wound, and small intestine, but not by skeletal muscle. Metabolism 45: 1161-1167, 1996.

54. Vogt JA, Hunzinger C, Schroer K, Holzer K, Bauer A, Schrattenholz A, Cahill MA, Schillo S, Schwall G, Stegmann W and Albuszies G: Determination of fractional synthesis rates of mouse hepatic proteins via metabolic 15N-labeling, MALDI-TOF MS and analysis of relative isotopologue abundances using average masses. Anal Chem 77: 2034-2042, 2005.

55. Zhao Y, Lee WN, Lim S, Go VL, Xiao J, Cao R, Zhang H, Recker RR and Xiao GG: Quantitative proteomics: measuring protein synthesis using 15N amino acid labeling in pancreatic cancer cells. Anal Chem 81: 764-771, 2009.

56. Cho H, Thorvaldsen JL, Chu Q, Feng F and Birnbaum MJ: Akt1/ PKB alpha is required for normal growth but dispensable for maintenance of glucose homeostasis in mice. J Biol Chem 276: 38349-38352, 2001.

57. Chi H, Mu J, Kim JK, Thorvaldsen JL, Chu Q, Crenshaw EB, Kaestner KH, Bartolome M, Shilman GI and Birnbaum MJ: Insulin resistance and a diabetes mellitus-like syndrome in mice lacking the protein kinase Akt2 (PKB beta). Science 292: 1728-1731, 2001.

58. Wilson EM and Rotwein P: Selective control of skeletal muscle differentiation by Akt1. J Biol Chem 282: 5106-5110, 2007.

59. Rotwein P and Wilson EM: Distinct actions of Akt1 and Akt2 in skeletal muscle differentiation. J Cell Physiol 219: 503-511, 2005.

60. Gan HT and Chen JD: Roles of nitric oxide and prosta glandins in pathogenesis of delayed colonic transit after burn injury in rats. Am J Physiol Regul Integr Comp Physiol 288: R1316-R1324, 2005.

61. Torres SH, Sanzis JB, de L. Briceno M, Hernandez N and Finol HJ: Inflammation and nitric oxide production in skeletal muscle of type 2 diabetic patients. J Endocrinol 181: 419-427, 2004.