Selection and structural characterization of anti-TREM2 scFvs that reduce levels of shed ectodomain

Graphical abstract

Highlights

- scFvs were generated against the immunoglobulin-like domain of the receptor TREM2

- Crystal structures revealed scFv binding to epitopes outside the TREM2 CDRs

- Oligomeric scFv species reduced levels of shed TREM2 ectodomain in a HEK293 model

- The scFvs form renewable structural and functional biology tools for TREM2 research

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In brief

TREM2 variants are associated with an increased risk of dementia. Szykowska et al. generated scFv antibody fragments to TREM2 immunoglobulin-like domain and solved crystal structures of scFv-TREM2 complexes to reveal their binding modes. Oligomeric scFv species showed enhanced functional activity reducing levels of shed TREM2 ectodomain in a HEK293 model.
Selection and structural characterization of anti-TREM2 scFvs that reduce levels of shed ectodomain

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SUMMARY

Mutations in TREM2, a receptor expressed by microglia in the brain, are associated with an increased risk of neurodegeneration, including Alzheimer’s disease. Numerous studies support a role for TREM2 in sensing damaging stimuli and triggering signaling cascades necessary for neuroprotection. Despite its significant role, ligands and regulators of TREM2 activation, and the mechanisms governing TREM2-dependent responses and its cleavage from the membrane, remain poorly characterized. Here, we present phage display generated antibody single-chain variable fragments (scFvs) to human TREM2 immunoglobulin-like domain. Co-crystal structures revealed the binding of two scFvs to an epitope on the TREM2 domain distal to the putative ligand-binding site. Enhanced functional activity was observed for oligomeric scFv species, which inhibited the production of soluble TREM2 in a HEK293 cell model. We hope that detailed characterization of their epitopes and properties will facilitate the use of these renewable binders as structural and functional biology tools for TREM2 research.

INTRODUCTION

Late-onset Alzheimer’s disease (LOAD) is the most common type of dementia, characterized by accumulation of extracellular amyloid-β (Aβ) aggregates and intracellular neurofibrillary tangles of hyper-phosphorylated tau, with a long prodromal phase followed by cognitive decline. Despite the urgency and necessity to develop therapeutics, there are currently no approved drugs which cure or slow the underlying progression of Alzheimer’s disease (AD) and patients are still reliant on symptomatic treatments discovered in the mid-last century (Long and Holtzman, 2019).

After some late-stage clinical trials targeting Aβ failed to meet their desired endpoints and several large genome-wide association studies (GWAS) linked genes coding for components of the immune response to AD, neuro-inflammation has become an area of intense research for therapeutics (Block et al., 2007; Lambert et al., 2013). Among the genes identified by GWAS is TREM2 (Triggering Receptor Expressed on Myeloid cells), which encodes a single transmembrane receptor expressed in myeloid-derived cells, including microglia in the central nervous system (CNS) (Guerreiro et al., 2013). Homozygous loss-of-function mutations in TREM2, or the associated adaptor protein DAP12, were previously identified to cause aggressive early-onset dementia in Nasu-Hakola disease (Paloneva et al., 2003). Since then, several point mutations in the extracellular domain of TREM2 have been linked to neurodegenerative disorders, highlighting the importance of TREM2 functions in brain health and homeostasis (Konishi and Kiyama, 2018; Ulrich and Holtzman, 2017). The most notable amino acid substitution, R47H, leads to
4-fold increased risk of developing LOAD (Jonsson et al., 2013; Guerreiro et al., 2013). A stronger genetic effect is observed only in carriers of apolipoprotein E (APOE) ε4, a potential TREM2 ligand, which has been implicated in TREM2 pathology (Kraemmann et al., 2017; Shi and Holtzman, 2018; Parhizkar et al., 2019).

TREM2 has numerous putative functions, including regulation of lipid and cholesterol metabolism, phagocytosis of myelin and Aβ, and generation of a microglial barrier around Aβ plaques (Jay et al., 2017; Nugent et al., 2020; Ulland and Colonna, 2018; Ulolland et al., 2017; Yeh et al., 2016; Yuan et al., 2016). TREM2 has been reported to bind ligands as diverse as Aβ, lipids, myelin, and lipoproteins (Wang et al., 2015; Yeh et al., 2016; Zhao et al., 2018). Recent findings have indicated that TREM2-dependent signaling is essential for the transcriptional definition of disease-associated microglia, a phenotype that is believed to be neuroprotective as it upregulates genes involved in phagocytosis (e.g., Keren-Shaul et al., 2017).

TREM2 contains an immunoglobulin-like (Ig-like) domain followed by a flexible stalk region, a transmembrane domain, and a short cytoplasmic tail. The stalk region can be cleaved by ADAM10/17 proteases to generate a soluble TREM2 fragment (sTREM2), while the C-terminal intramembranous domain is further cleaved by gamma-secretase (Wunderlich et al., 2013). sTREM2 can be detected in cerebrospinal fluid (Wang et al., 2020) and is increased in patients with neuronal injury or CNS inflammatory diseases (Piccio et al., 2008; Rauchmann et al., 2019). In addition, sTREM2 was found to be increased in patients at early symptomatic stages of AD and correlated well with levels of phosphorylated tau in patients with tau pathology (Rauchmann et al., 2019; Suárez-Calvet et al., 2016). The importance of TREM2 cleavage in AD pathology is highlighted by the H157Y polymorphism at the protease cleavage site, which leads to excessive shedding of sTREM2 and increased risk of AD (Schlepckow et al., 2017; Thornton et al., 2017). It is unclear whether the increased risk is due to the resulting increased generation of additional sTREM2, which might be a biologically active molecule (Zhong et al., 2019), alterations in intracellular signaling or functional properties of the remaining C-terminal fragments.

There is minimal published structural data for TREM2 ectodomain motifs responsible for ligand engagement or regulation of TREM2 functions (Kober et al., 2016; Sudom et al., 2018). While...
the Ig-like domain R47H mutant has been proposed to be defec-
tive in ligand binding, its crystal structure was solved only
recently (Song et al., 2017; Sudom et al., 2018). The authors
concluded that the arginine substitution in CDR1 causes exten-
sive remodeling in the neighboring CDR2 loop of TREM2 result-
ing in local structural disorder and the loss of electron density.
The same loop has been identified to interact with putative li-
ghands in wild-type TREM2 crystals soaked with phosphatidyl-
serine (Sudom et al., 2018). However, structures incorporating
other ligands or the molecular mechanisms of TREM2 signaling
remain to be fully elucidated.

Because of the potential therapeutic impact of targeting
TREM2, we decided to generate single-chain variable antibody
fragments (scFvs) against the human TREM2 ectodomain with
which to study TREM2 structure and function. Agonist anti-
bodies have already been reported in recently published work
(Ellwanger et al., 2021; Fassler et al., 2021; Ibach et al., 2021;
Price et al., 2020; Schlepckow et al., 2020; Wang et al., 2020).
The group of Schlepckow have shown that a full-length antibody
specific to mouse TREM2 can decrease shedding and activate
TREM2 signaling in vitro, and also lead to a significant reduction
in amyloid plaques in 6-month-old amyloid-beta precursor pro-
cin knockin mice (Schlepckow et al., 2020). Antibodies might
also be useful biochemical tools for studying the function of
sTREM2 (Zhong et al., 2019).

There are currently no published structures of antibodies in
complex with TREM2. Here, we present our work on the genera-
tion and characterization of four scFvs selected against the Ig-
like domain (herein Ig domain) of human TREM2. The co-crystal
structures of two of these antibody fragments, scFv-2 and scFv-
4, have been solved in complex with TREM2, revealing interac-
tions with epitopes distal to the putative ligand-binding site.
Two antibody fragments, scFv-3 and scFv-4, reduced the shed-
ding of sTREM2 from HEK293 cells.

RESULTS
Phage display selection of anti-TREM2 scFvs
To facilitate the development of renewable antibody binders
against TREM2, we performed phage display selections using
the SciLifeLib synthetic library of human scFvs similarly con-
structed and designed as reported previously (Preger et al.,
2020; Säll et al., 2016). A TREM2 antigen comprising the Ig
domain of human TREM2 (His19-Asp131) was expressed re-
combinantly from Sf9 insect cells and biotinylated on a C-termi-
nal Avi tag for immobilization on streptavidin-coated magnetic
beads. Following four rounds of phage selections and initial
binding and specificity analyses (Figure S1), four scFvs (scFv-
1, 2, 3, and 4) were selected for further characterization (Figure 1).
All four scFv fragments were relatively poorly expressed when
produced in E. coli and, therefore, scFvs were prepared by
secretion from Sf9 insect cells, which allowed for higher yields,
increased solubility and a reduction in endotoxin contamination
(Figures 2A and 2B). The scFvs showed a tendency to dimerize
as observed by size-exclusion chromatography (SEC), as well
as delayed elution from interaction with the Superdex media as well as a tendency for dimerization (D-dimer, M-monomer).

Figure 2. Optimization of scFv production in a baculoviral expression system
(A) Expression yields of scFvs produced in E. coli TOP10 cells or Sf9 insect cells.
(B) Coomassie blue-stained SDS-PAGE gel showing the final purity of scFvs after purification from Sf9 cells (image generated from different purification ex-
periments).
(C) Size-exclusion chromatography of each scFv antibody fragment purified on a Superdex 200 16/60 column. Delayed elutions indicated scFv interactions with
the Superdex media as well as a tendency for dimerization (D-dimer, M-monomer).
The binding of the scFvs, expressed from Sf9 cells, to TREM2, was characterized by surface plasmon resonance (SPR) to establish their relative binding affinities (Figure S2; apparent $K_D$ values [Preger et al., 2020] are reported to account for potential avidity effects caused by scFv oligomerization). For these experiments, the human TREM2 antigen was cloned into a mammalian expression vector and produced as a longer biotinylated construct consisting of the Ig domain and stalk region of TREM2 (His19-Ser174) that was then immobilized onto a streptavidin-coated sensor chip. Initial measurements on mixed oligomer samples revealed that all antibody fragments showed strong binding to TREM2 except for scFv-1, which proved to be a weaker binder with an apparent equilibrium dissociation constant ($K_D$) $> 100$ nM (Figure S1). The binding kinetics of scFv-3 displayed moderate affinity with a fast dissociation rate ($K_D$ $\approx 5$ nM; Figure S2B). By contrast, scFv-2 and scFv-4 were characterized by high-affinity binding, $K_D$ values of $\sim 1$ nM (Figures S2A and S2C), although the accuracy of scFv-4 binding kinetics determination was limited by some non-specific interactions with the SPR sensor chip as well as the possible avidity effects due to scFv oligomerization (Figure 2C).

Structure determination of TREM2-scFv antibody complexes

We hypothesized that scFv antibodies binding at different sites on the TREM2 ectodomain might differentially affect receptor signaling, internalization, or cleavage and, therefore, that an appreciation of their binding sites is important. Hence, we decided to attempt to determine the crystal structures of TREM2 bound to scFv-2, scFv-3, and scFv-4 antibodies whose binding kinetics suggested strong interactions. Crystallization screens were established using the TREM2 Ig domain (His19-Asp131), which has been structurally characterized previously (Kober et al., 2016; Sudom et al., 2018), as well as using the TREM2 Ig domain and stalk (His19-Ser174), which is relevant to shedding by ADAM proteases at H157, but susceptible to degradation in vitro due to the inherent flexibility of the C-terminal stalk region (Sudom et al., 2018). Both TREM2 proteins were produced in Expi293F/C212 human cells in the presence of kifunesin and deglycosylated using Endo H treatment.

Diffracting crystals were first obtained for the complexes of the smaller TREM2 Ig domain with both scFv-2 and scFv-4, enabling their structure determination at 3.36 and 3.07 Å resolution.

### Table 1. Data collection and refinement statistics (molecular replacement)

|                      | 6Y6C (scFv-4) | 6YMQ (scFv-4) | 6YYE (scFv-2) |
|----------------------|---------------|---------------|---------------|
| **Data collection**  |               |               |               |
| Space group          | P 4 1 2 2     | P 2 1 2 1 2   | 1 2 2 2       |
| Cell dimensions      |               |               |               |
| $a$, $b$, $c$        | 111.66, 111.66, 232.20 | 167.51, 180.77, 125.51 | 113.94, 126.20, 225.01 |
| $\alpha$, $\beta$, $\gamma$ | 90, 90, 90  | 90, 90, 90  | 90, 90, 90 |
| Resolution ($\AA$)   | 100.63–2.26 (2.341–2.26) | 75.99–3.07 (3.18–3.07) | 110.07–3.36 (3.36–3.57) |
| $R_{merge}$          | 0.2129 (2.041) | 0.2811 (1.86) | 0.266 (1.55) |
| $I/\langle I \rangle$ | 9.96 (1.25)   | 4.2 (1.0)     | 7.4 (1.7)     |
| Completeness (%)     | 99.80 (99.34) | 99.83 (99.90) | 92 (68.9)     |
| Redundancy           | 21.4 (14.3)   | 6.4 (6.6)     | 13.4 (12.7)   |
| Wilson B factor      | 39.44         | 69.70         | 68.7          |
| **Refinement**       |               |               |               |
| Resolution ($\AA$)   | 2.26          | 3.07          | 3.36          |
| No. of reflections   | 69,383 (6,776) | 71,714 (7,075) | 16,272 (70)   |
| $R_{work}/R_{free}$  | 0.191/0.216   | 0.2730/0.2871 | 0.298/0.328   |
| No. of atoms         |               |               |               |
| Protein              | 5368          | 15,625        | 5038          |
| Ligand/ion           | 28            | 92            | 14            |
| Water                | 375           | 13            | –             |
| **B factors**        |               |               |               |
| Protein              | 42.31         | 88.15         | 91.39         |
| Ligand/ion           | 67.13         | 90.45         | 91.5          |
| Water                | 47.65         | 58.05         | –             |
| **RMSD**             |               |               |               |
| Bond lengths ($\AA$) | 0.002         | 0.001         | 0.003         |
| Bond angles ($)      | 0.51          | 0.40          | 0.56          |
| Solvent (%)          | 73.85         | 69            | 77.18         |
| Average B factor ($\AA^2$) | 42.0   | 88.14        | 91.0          |
| Anisotropy           | 0.484         | 0.641         | 0.041         |

RMSD, root-mean-square deviation.
respectively (see Table 1 for data collection and refinement statistics). Both crystal lattices were characterized by a high solvent content (Figure S3) and, despite screening over 180 crystals, the resolution of the scFv-2 complex could not be improved. Nevertheless, this level of resolution was sufficient to define the scFv-2/TREM2 interface. A higher-resolution structure of the scFv-4 complex was, however, subsequently obtained using the longer TREM2 construct comprising both the Ig domain and stalk region and refined to 2.26 Å resolution. Generally, the scFv-2/TREM2 complex contained fewer crystal contacts than scFv-4, leading to a more spacious lattice and possibly lower resolution (Figure S3). Despite similar efforts, no crystals were obtained for TREM2 in complex with scFv-3. This could reflect its faster dissociation kinetics or perhaps its greater monomer/dimer heterogeneity at high concentrations.

**ScFv-2 and scFv-4 bind to partially overlapping epitopes on TREM2 distal to the putative ligand-binding site**

Here, we focus our subsequent work on the higher-resolution structure of the scFv-4 complex with the TREM2 Ig domain and stalk (Figure 3A) and the scFv-2 complex with the TREM2 Ig domain alone (Figure 3B). In both structures, the globular Ig domain of TREM2 displayed a β sandwich fold that consisted of nine β strands, including strands A-G common to all Ig-like domains and a C'-C'' insertion typical of the V-set Ig domain. The overall structure does not differ significantly to the previously solved structures of wild-type TREM2 (PDB: 5ELI and 5UD7), except for some variability in the flexible loops (Figure S4A). The putative ligand-binding site on TREM2 is proposed to be located at one end of the β sandwich, which presents the three complementarity-determining regions (CDRs1-3) and includes

![Figure 3](https://example.com/figure3.png)
the AD-risk allele sites Arg47, as part of CDR1, and Arg62 in the C' strand (Kober et al., 2016; Song et al., 2017; Sudom et al., 2018). Of note, both scFv-2 and scFv-4 were found to bind to the opposite end of TREM2 (Figures 3A–3C). Here, the β sandwich of TREM2 splayed between β strands C and F to provide an extended surface for the scFvs to bind that included parts of β strands A, F, and G, and loop C-C' (Figures 3A–3C). However, the superposition of the two TREM2-scFv complexes revealed that the binding epitopes on TREM2 were only partially similar (Figure 3C). For instance, scFv-4 formed interactions with the C-C' loop in TREM2 that were largely absent in the scFv-2 complex. Overall, the structures suggest that the scFvs should find broad application with the binding epitopes being located favorably for scFv interaction with both the ligand-bound and unbound states, as well as mutant states.

**Interactions of scFv-4 with the TREM2 ectodomain**

Overall, the variable heavy (VH) and variable light (VL) domains of scFv-4 displayed canonical immunoglobulin folds tethered by an engineered glycine-serine-rich linker that was disordered in the refined structure. The binding of scFv-4 to TREM2 was centered on the CDR3 loop of the scFv-4 VH domain (herein CDR-H3), flanked by interactions from CDRs-H1 and H2 and by the VL domain CDR-L2 (Figures 4A and 4B).

The resolution of the scFv-4 complex structure enabled a detailed analysis of the side chain interactions in the binding interface (Figures 4A and 4B). A striking feature of the CDR sequence selections in scFv-4 was the enrichment of tyrosine residues (Figure 1). Indeed, CDR-H3 contained a sequence of six consecutive tyrosines, interrupted only by Gly104 (numbering corresponding to PDB entry sequences), which was likely selected to avoid the steric clashes that would result from any other amino acid at this position. Tyrosine residues 33, 59, 102, 103, 106, 107, and 187 in scFv-4 contributed to extensive hydrophobic packing interactions with the non-polar TREM2 residues Val23, Pro102, Leu107, Leu125, Val128, and Ala130 (Figures 4A and 4B). Additional side chain- and water-mediated hydrogen bond interactions were provided by scFv-4 loops CDR-H3 and CDR-L2 (Figure 4B). The most extensive hydrogen bond interactions occurred between the CDR-L2 with the C-C' loop of TREM2. Here, the scFv-4 residues Tyr187, Ser191, and Ser194 formed hydrogen bonds with TREM2 residues Lys47, Gly58, and Glu56, respectively (Figure 4B). Tyr102 from CDR-H2 and H3 also inserted between TREM2 β strands C and F to form a further hydrogen bond with TREM2 Gln53 (Figure 4B). No significant differences were observed in scFv-4 binding to the TREM2 Ig domain in the equivalent structure using the shorter TREM2 construct His19-Asp131 (Figure S4B).

While the primary scFv-4 interaction was mediated by the TREM2 Ig domain as described above, we identified crystal contacts between the TREM2 stalk and CDR-H1 from a neighboring scFv-4 subunit. This additional packing allowed the stalk to be traced in the electron density maps up to Asp137 in the C terminus and several hydrogen bond interactions were observed to support the stalk interaction (Figure S5).

**Interactions of scFv-2 with the TREM2 ectodomain**

The binding of scFv-2 to TREM2 was mediated by the three CDR loops of the VH domain (CDRs-H1, H2, and H3), as well as by CDR-L3 from the light domain (Figures 5A and 5B). Overall, these loops were enriched in tyrosine and serine residues that contributed to a mix of hydrophobic and hydrogen bond interactions. Of note, TREM2 Glu127 (βG strand) was within hydrogen bonding distance of three scFv-2 side chains, including Ser54, Ser56, and Ser59, from CDR-H2. CDR-H2 Ser59 was additionally within hydrogen bonding distance of TREM2 Gln25 (βA). The sole glycine in CDR-L3 (Gly231) also contributed a hydrogen bond from its carbonyl group to the side chain of TREM2 His103.

Together the scFv-2 and scFv-4 complex structures show the potential value of these antibody fragments as crystallization chaperones. More importantly, however, they define the epitopes of these scFvs, allowing for the interpretation of functional properties in the context of the binding site on the surface of TREM2, which has been poorly described for other anti-TREM2 antibodies.

**ScFv-3 and scFv-4 reduce shedding of sTREM2 from HEK293 cells overexpressing TREM2 and DAP12**

The H157Y polymorphism, which leads to accelerated cleavage of TREM2 and increases AD risk, suggests that regulation of shedding of sTREM2 might be important in human biology (Jiang et al., 2016; Thornton et al., 2017). Based on the potential interactions of scFv-4 with the stalk domain, as indicated by the crystal contacts, and the potential for TREM2 internalization induced by scFv dimers, we decided to determine the effect of the scFv antibodies on sTREM2 shedding. We devised a sandwich ELISA, capturing
sTREM2 from cell culture supernatants, and confirmed that scFvs were not masking the sTREM2 epitopes required by the ELISA capture and detection antibodies (Figure S6). Recently another metalloprotease, meprin β, was shown to cleave TREM2, with the main cleavage site between Arg136 and Asp137 (Berner et al., 2020). We confirmed that both the Ig domain as well as a region of the stalk C-terminal to Ala138 were required for detection in our ELISA protocol. Thus, the detected sTREM2 product is unlikely to be resulting from meprin β cleavage (Figure S6). Batimastat, a broad-spectrum chemical inhibitor of matrix metalloproteinases, including ADAM proteases, was used as a positive control to inhibit shedding. HEK293 cells stably transfected with Trem2, either mock-transfected or transfected with Trem2/DAP12, were treated with each of the scFvs for 5 h to reach the appropriate threshold for sTREM2 detection in the supernatant. ScFv-3 and scFv-4 (10 μg/mL) significantly reduced shedding of sTREM2 by 22% and 38%, respectively (Figure 6A).

Oligomerization of scFvs has been observed and can be exploited to enhance the properties of selected molecules (Kortt et al., 2001). We decided to investigate the effect on shedding of the apparent monomer and dimer species present in our scFv-3 and scFv-4 samples. After these species were acutely separated by SEC and processed rapidly to minimize re-equilibration between low- and high-molecular-weight (MW) species, it was found that the monomer species for both scFv-3 or scFv-4 had minimal or no activity (Figure 6B), whereas the dimers decreased production of sTREM2 by more than 50% (Figure 6B). Concentration-response experiments for the scFv-3 and scFv-4 dimers showed that maximum effect plateaued between 50% and 70% with calculated half-maximal effective concentration values of 2.5 and 3 nM, respectively (Figures 6C and 6D). Of note, scFv-2, which showed a high binding affinity for TREM2, caused no changes in sTREM2 release despite having a similar binding epitope to scFv-4 (Figure 6A). This antibody fragment purified mostly as a monomer, further highlighting the importance of scFv oligomerization for reduction of shedding. None of the scFvs impacted cell viability, as measured by counting cell nuclei using Hoechst staining and fluorescence-based viability (data not shown).

Finally, to demonstrate selective binding to TREM2, we conjugated the dimeric fractions of scFv-3 and scFv-4 to Alexa Fluor 647 and repeated the HEK293 treatment protocol with subsequent imaging of the fluorescent scFvs (Figure 7). Labeling of the cells was dependent upon the heterologous expression of TREM2/DAP12. Most of the Alexa signal was present intracellularly and was resistant to acidic wash, consistent with TREM2-mediated internalization under the conditions tested. ScFv-4 appeared to show superior specificity compared with scFv-3 based on the staining of non-transfected HEK293 cells (Figure 7).

DISCUSSION

In this paper, we have presented our work on the structural characterization of scFvs raised against human TREM2 by phage display. We screened a library of scFv antibody fragments against fully folded and glycosylated TREM2 Ig domain, which resulted in the generation of three successful candidates. Despite challenging purification procedures, we were successful in producing recombinant antibody fragments and demonstrated them to be suitable for X-ray crystallography. We have solved the structures of two scFvs bound to TREM2 and provide the first crystallographic structures of antibody interactions with TREM2. These are also the first antibody fragments reported to unambiguously bind to the TREM2 globular Ig domain. In addition, we have shown that higher-MW oligomeric species of scFvs are able to display enhanced functional activity, suggesting parallels with the activity of bivalent mAbs and diabodies.

Since TREM2 is a potential therapeutic target for AD, the pharmaceutical industry has initiated several projects aimed at developing TREM2 antibody therapeutics, with at least two candidates having commenced clinical trials. Data have been reported for two, AL002 and 4D9, which allow for an interesting comparison with scFv-4 (Schlepcow et al., 2020; Wang et al., 2020). Other functionally active antibodies have either not been widely published (WO2016023019A2) or the epitope has not been defined and, therefore, are not included in the discussion below (Cheng et al., 2018). X-ray crystallographic data, which accurately define the antibody/TREM2 interfaces, are not publicly available for any of these reagents. AL002 is an agonist antibody with a poorly defined epitope most likely in the stalk region between residues 112 and 174, while 4D9 has an epitope defined by peptide mapping to residues Asp137-Gly145 (Schlepcow et al., 2020; Wang et al., 2020). As such, they target TREM2 epitopes N-terminal of the z-secretase cleavage site at H157, whereas our X-ray crystallography data show that scFv-4 binds the Ig domain of TREM2. The binding sites do show some cross-species variation, explaining the observation that 4D9 is specific.
C...that reduction of sTREM2 production by bivalent antibodies or possibly that they might affect cleavage of TREM2 by virtue of sterically hindering access by ADAMs. Consistent with that hypothesis, both full-length antibodies (4D9 and AL002) have been shown to reduce shedding of sTREM2. This interesting property has been confirmed for the first time in human with AL002 (Wang et al., 2020). This function also appears to be dependent upon the use of a bivalent antibody, as the 4D9 Fab fails to inhibit shedding of sTREM2 by cells. These observations are in agreement with our data, where higher-MW oligomers of scFv-3 and scFv-4 inhibit up to 70% of sTREM2 production.

At present, the mechanism of action for the inhibition of shedding by all of these antibody reagents is unclear. It might occur either by inhibition of cleavage of TREM2 at the cell surface or by promoting internalization of full-length TREM2, thereby removing substrate availability. Schlepckow et al. propose that 4D9 exerts some of its effects by stabilizing TREM2 at the plasma membrane, explaining the potentiation of signaling in response to liposomes by 4D9. However, it is also possible that reduction of sTREM2 production by bivalent antibodies or oligomeric scFvs is due to crosslinking of receptor and internalization, thereby reducing cell surface substrate for shedding. It is noteworthy that a bivalent antibody is necessary for 4D9 functional effects and, to date, we have not observed any functional responses to monomeric scFv-4, only observing activity in preparations containing appreciable dimeric species. Furthermore, Alexa Fluor-labeled scFv-4 and scFv-3 label HEK293 cells in a TREM2 expression-dependent manner, leading to intracellular immunofluorescence. Interestingly, the scFv-2 antibody fragment has the lowest propensity to form oligomeric species and did not show any inhibitory activity in the sTREM2 shedding assay. Our data are therefore consistent with the effect of scFv-3 and scFv-4 upon shedding of sTREM2 being due to promoting the internalization of TREM2.

Both AL002 and 4D9 show agonist properties in vitro. AL002 elicits TREM2-mediated secondary intracellular signaling, demonstrated using an NFAT reporter cell line and phosphorylation of a protein of equal MW to DAP12 in bone-derived macrophages (Wang et al., 2020). Similarly, 4D9 shows agonism, but only when a full-length antibody was used to challenge cells; Syk phosphorylation did not occur following exposure of cells to a 4D9 Fab fragment. The requirement for a bivalent antibody for the function of AL002 has not been reported, but these observations with 4D9 led the authors to hypothesize that the functional effects of 4D9 are dependent upon a bivalent antibody and crosslinking of TREM2 molecules (Schlepckow et al., 2020). Taken together, the literature and our own data highlight the importance of multivalency for the functional activities of anti-TREM2 antibodies. On account of the need to prepare bivalent mAb and Fab antibodies, a full investigation of the agonist...
and antagonist properties of the scFvs obtained in our studies is the subject of a separate piece of work.

While we have presented evidence that scFvs capable of dimerization reduce sTREM2 shedding from the HEK293 cells, the functional impact of monomeric scFvs is unclear due to the dynamic equilibrium between monomeric and dimeric forms. However, we have observed a decrease in activity upon storage of a purified fraction of dimers and also negligible activity of freshly prepared and rapidly utilized monomers, suggesting that the monomer represents an inactive conformation (data not shown). Similarly to the mAbs discussed above, multivalent scFvs will have the potential to crosslink TREM2 ectodomains into higher-order structures, thereby creating significant steric hindrance to proteolytic complexes, altering their characteristics as substrates, or triggering endocytosis.

By using phage display to raise recombinant scFvs we have assured that the source of the molecules is renewable and can be easily shared. During our studies, we have encountered difficulties caused by the use of commercially available antibodies with unknown TREM2 epitopes, especially when used in ELISA, pull-down studies or immunocytochemistry. The presented scFvs bind away from the proposed ligand recognition site, qualifying them for development into additional tools for the study of TREM2-ligand interactions and receptor processing.

Our data represent the first crystallographic characterization of binding interactions between functional antibody fragments and TREM2 and we hope that these additional tools will help elucidate the complex pathophysiology of TREM2 in AD.

**STAR METHODS**

Detailed methods are provided in the online version of this paper and include the following:

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SUPPLEMENTAL INFORMATION

Supplemental information can be found online at https://doi.org/10.1016/j.str.2021.06.010.

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AUTHOR CONTRIBUTIONS

Conceptualization, A.S., J.B.D., N.A.B.-B., and A.N.B.; provision of resources, Y.C., J.Y., D.Q., S.M.M., S.G., P.J.A., and J.W.; investigation, A.S., J.Y., D.Q., C.P., and E.W.; supervision, Y.C., T.B.S., S.G., H.P., P.J.A., E.DiD., E.M., J.B.D., E.M., T.B.S., and E.DiD. were supported by a DPhil studentship sponsored by Eisai. J.B.D., E.M., T.B.S., and E.DiD. were supported by a grant from Alzheimer’s Research UK (ARUK-2015DD-OX).

DECLARATION OF INTERESTS

Y.C., S.J.N., P.J.A., and J.W. are employees of Eisai Ltd., and Eisai Inc.

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# STAR METHODS

## KEY RESOURCES TABLE

| REAGENT or RESOURCE | SOURCE | IDENTIFIER |
|---------------------|--------|------------|
| **Antibodies**      |        |            |
| Rabbit monoclonal anti-TREM2 antibody | Abcam | Cat#ab209814 |
| Biotinylated goat polyclonal anti-TREM2 antibody | R&D systems | Cat#BAF1828; RRID: AB_2208688 |
| **Bacterial and Virus Strains** |        |            |
| MACH1 T1            | ThermoFisher | Cat#C862003 |
| DH10Bac             | ThermoFisher | Cat#10361012 |
| **Chemicals, Peptides, and Recombinant Proteins** |        |            |
| pTT5-SP-TREM2-6xHis- (19-131) | This paper | N/A |
| pTT5-SP-TREM2-6xHis-avi- (19-174) | This paper | N/A |
| pHTBV1.1-SecNH-Bio:TREM2-10xN-His-C-avi (19-174) | This paper | N/A |
| pFB-Sec-Bio5:TREM2 N6xHis-C-avi (19-131) | This paper | N/A |
| pFB-Sec-Bio5:TREM2 N6xHis-C-avi (19-138) | This paper | N/A |
| pFB-Sec-NH:TREM2 N-6xHis (19-174) | This paper | N/A |
| TREM2 standard peptide | SinoBiological | Cat#11084-H08H |
| TMB ELISA substrate | ThermoFisher | Cat# 34028 |
| HRP conjugated streptavidin | ThermoFisher | Cat#N100 |
| Alexa Fluor 647 NHS ester | ThermoFisher | Cat#A37573 |
| **Deposited Data** |        |            |
| scFv-4 with TREM2 (19-174) | This paper | PDB: 6Y6C |
| scFv-4 with TREM2 (19-131) | This paper | PDB: 6YMQ |
| scFv-2 with TREM2 (19-131) | This paper | PDB: 6YYE |
| SP140 PHD-Bromodomain complex with scFv (data phasing) | Fairhead et al., 2019 | PDB: 6GSR |
| TREM2 Ig-domain (data phasing) | Sudom et al. (2018) | PDB: 5UD7 |
| **Experimental Models: Cell Lines** |        |            |
| HEK293 | ATCC | Cat#CRL1573 |
| HEK293 transfected with TREM2-DAP12 | Thornton et al. (2017) | N/A |
| Expi293F (for protein expression) | ThermoFisher | Cat#A14527 |
| Sf9 (for protein expression) | ThermoFisher | Cat#11496015 |
| **Oligonucleotides** |        |            |
| List of oligonucleotides used in this study | This study | Table S1 |
| **Recombinant DNA** |        |            |
| pH-sec (Signal peptide origin) | Aricescu et al. (2006) | Addgene plasmid # 99845 |
| pHTBV1.1-SecNH-Bio | Strain-Damerell et al. (2014) | N/A |
| pFB-Sec-Bio5 | SGC | N/A |
| pFB-Sec-NH | SGC | Addgene plasmid # 39189 |
| pTT5 | Zhang et al. (2013) | Addgene plasmid # 52326 |
| **Software and Algorithms** |        |            |
| Phenix version 1.17.1 | Emsley et al. (2010); Liebschner et al. (2019) | https://www.phenix-online.org/ |
| ChimeraX version: 0.93 | Goddard et al. (2018) | https://www.rbvi.ucsf.edu/chimerax/ |
| Buster 2.10.3 | Bricogne et al. (2016) | https://www.globalphasing.com/buster/ |
| PyMOL 2.3.4 | The PyMOL Molecular Graphics System, Version 2.3 Schrödinger, LLC | https://pymol.org/2/#download |
| GraphPad Prism version 8.42 for Windows | GraphPad Software, La Jolla California USA | https://www.graphpad.com |
RESOURCE AVAILABILITY

Lead contact
Further information and requests for reagents and resources may be directed to and will be fulfilled by the Lead Contact Alex N. Bullock, (alex.bullock@cmd.ox.ac.uk)

Materials availability
Plasmids generated in this study will be deposited to Addgene and made available on request.

Data and code availability
The coordinates and structure factors for the crystal structures reported in this article have been deposited in the PDB with accession codes 6YYE (TREM2-scFv-2), 6Y6C (TREM2-scFv-4) and 6YMQ (TREM2-scFv-4).

EXPERIMENTAL MODEL AND SUBJECT DETAILS

Cell lines
All plasmids were generated in MACH1 or DH5α (Thermofisher Scientific C862003) cells grown in LB media at 37 °C and bacmid DNA for insect cell expression and baculovirus generation were generated in DH10Bac (Thermofisher Scientific 10361012). For phage display, the enriched phages were amplified overnight by infection of E. coli XL1-blue (Stratagene, La Jolla, CA, USA).

Baculoviral protein expression was performed in Sf9 cells (Thermofisher Scientific 11496015) grown at 27 °C in SF900 II-SFM media. Mammalian protein expression was performed in Expi293F™ cells (female, Thermofisher Scientific A14527) in Expi293™ expression medium (Thermofisher Scientific) at 37 °C, 8% CO2.

WT HEK293 (female; ATCC CRL1573) and HEK293 cells stably expressing WT hTREM2 and hDAP12 were cultured in DMEM (GIBCO) with 10% heat-inactivated foetal bovine serum (FBS) (Gibco) at 37 °C, 5% CO2. The cell culture medium for the HEK293 Trem2-DAP12 cells was supplemented with 1 mg/mL puromycin hydrochloride (Cayman Chemical).

METHOD DETAILS

Cloning of TREM2 and scFvs

Cloning of TREM2 for ELISA validation
DNA encoding human TREM2 (amino acids 19-131; gi|9507203; MGC) used for antigen generation and TREM2 (a.a. 19-138) for ELISA validation were cloned into the baculovirus secretion plasmid pFB-Sec-Bio5, which provides a C-terminal Avi-tag sequence for biotinylation. The stalk-containing TREM2 (a.a. 19-174) used for ELISA validation was cloned into baculovirus secretion plasmid pFB-Sec-NH which provides a N-terminal hexahistidine tag. See Table S1 for oligonucleotides and Key Resources Table for vectors and cell lines.

Cloning of TREM2 for SPR
A TREM2 construct containing the native signal peptide (a.a. 1-174) was cloned into the pTT5 vector, followed by an Avi tag and TEV-cleavable hexahistidine tag (GLNDIFEAQKIEWHEGSENLYFQSHHHHHH) at its C-terminus. The SP sequence was cloned from pHL-sec vector into pTT5 DNA inserts (Aricescu et al., 2006; Zhang et al., 2013).

Cloning of TREM2 for X-ray crystallography
For structural studies, human TREM2 (a.a. 19–131) was cloned into the mammalian expression vector pTT5, following an N-terminal signal peptide (MGILPSGPMPALLSLVSLLSVMGCVAETG) with hexahistidine tag preceeded by additional linker (a.a. GTK) at its C-terminus.

The longer construct TREM2 (a.a. 19-174) containing the stalk was inserted by ligation independent cloning (LIC) (Strain-Damerell et al., 2014) into the vector pHTBV1.1-SecNH-Bio, which provides an N-terminal Gp64 signal peptide and TEV-cleavable hexahistidine tag as well as a C-terminal Avi tag.

Cloning of scFvs for baculovirus expression
ScFv antibody fragments identified by phage display were cloned for baculovirus expression with scaffold specific primers into the vector pFB-Sec-NH.

Expression and purification of TREM2 and scFvs in baculovirus expression system

Bacmid generation and transfection
TREM2 viruses for baculovirus expression were generated by transfection with bacmid DNA:JetPrime (Polyplus) complexes. Sf9 cells at a density of 0.2x10^6 in adherent culture were transfected for 4 hours (h) at 27 °C followed by medium exchange to SF900 II-SFM containing 0.1% penicillin and streptomycin (Thermofisher Scientific) with 2% FBS (Thermofisher Scientific). P1 viruses were harvested after 7 days at 27 °C and 120 µL was used to infect 3 mL of Sf9 cells for P1 virus amplification (72 h and 450 rpm shaking and throw 6 mm). Following another round of viral amplification, 0.75 L of Sf9 cells in SF900 II-SFM media were infected with virus P2 at 7 mL/L at density 2x10^6 cells/mL in glass flasks (27 °C, 100 rpm for 72 h, throw 25 mm).
**Purification of TREM2 for antigen generation and ELISA validation**

The clarified supernatants were loaded onto 5 mL Ni-NTA (Qiagen)/per 1 L at 10 mL/min. Resin was washed with 20 column volumes (CV) of 50 mM HEPES pH 7.4 and 300 mM NaCl, 5% glycerol, 5 mM imidazole followed by 10 CV of 10 mM imidazole and 600 mM NaCl containing buffer. Proteins were eluted with buffer containing 500 mM imidazole. Proteins were purified further by size exclusion chromatography (Superdex 200, 16/60) and in vitro biotinylated using recombinant BirA enzyme (enzyme to substrate ratio of 1:50, w/w). The reaction was performed in a buffer containing 10 mM Tris-HCl, 50 mM NaCl and 10 mM MgOAc, pH 8.0 for 14 h, at 4°C in the presence of 0.1 mM biotin and 10 mM ATP. Subsequently, the buffer was exchanged by dialysis to 20 mM HEPES, 300 mM NaCl, 5% glycerol and 0.5 mM TCEP, pH 7.5.

TREM2 (His19-Ala138) and (His19-Ser174) for ELISA validation were purified in the same manner but were stored at -80°C after size exclusion.

**Expression and purification of scFvs**

Viruses for scFvs were generated as above. Immobilised metal affinity chromatography (IMAC) was performed as for TREM2, except for a change to pH 6.8 which decreased precipitation. Eluted proteins were cleaved with TEV protease overnight (O/N) at 4°C followed by loading onto a 1 mL Protein A FF (GE healthcare) column at 3 mL/min. Resin was washed with 20 CV of base buffer (BB: 25 mM HEPES pH 7.5, 150 mM NaCl) followed by a wash with 10 CV of BB containing 0.05% Triton-X. Proteins were eluted with 0.1 M acetic acid pH 3 at 0.5 mL/min and immediately buffer exchanged on PD10 columns (scFv-1: 120 mM Citrate pH 7 with 50 mM L-arginine, 50 mM L-glutamic acid, scFv-2: 100 mM HEPES pH 7.2, 25 mM NaCl, 100 mM L-arginine, 100 mM L-glutamic acid, scFv-3: 100 mM Tricine pH 8, scFv-4: 100 mM Tricine pH 8) and frozen until further use. Later, scFv-3 and scFv-4 oligomers were separated by double SEC on Superdex 200 16/60 in 20 mM Tris-HCl pH 7.5, 300 mM NaCl and 5% glycerol. Separated oligomers were pooled and buffer exchanged to PBS using vivaspin 2 10 MWCO concentrators (GE healthcare). Proteins were clarified by centrifugation at 30,000 x g and immediately frozen in liquid nitrogen and used for the assay within a week.

**Expression and purification of TREM2 in mammalian cells**

**Transient transfection of TREM2 plasmids**

For crystallisation studies of TREM2 (a.a. 1-131), Exp293™ cells (Thermofisher Scientific A14527) in Exp293™ expression medium (Thermofisher Scientific) at 2.5 × 10⁶ were transfected with a mixture of 1 mg of pTT5:TREM2 (19-131) DNA to 6 mg L-PEI 25 K (Poly-science) per 1 L at 10 mL/min. The reaction was performed in a buffer containing 10 mM Tris-HCl, 50 mM NaCl and 10 mM MgOAc, pH 8.0 for 14 h, at 4°C in the presence of 0.1 mM biotin and 10 mM ATP. Subsequently, the buffer was exchanged by dialysis to 20 mM HEPES, 300 mM NaCl, 5% glycerol and 0.5 mM TCEP, pH 7.5.

TREM2 (His19-Ala138) and (His19-Ser174) for ELISA validation were purified in the same manner but were stored at -80°C after size exclusion.

**Antibody selection and validation**

Antibody generation through phage display selection was performed as described previously, using the in vitro biotinylated TREM2A (19-131) from Sf9 cells as antigen and a human synthetic single-chain fragment variable (scFv) library (Preger et al., 2020). Briefly, four rounds of selections were performed using streptavidin beads (Dynabeads M-280 streptavidin, Invitrogen) to immobilise the biotinylated TREM2 antigen. In the first two rounds, phages were incubated with immobilized antigen after a pre-selection on naked beads. In round 3 and 4, phage and antigen were incubated in solution before capturing on beads. The antigen-phage incubation time was decreased from 3 h in the first round to 1.5 h in rounds 2, 3, and 4. Furthermore, the selection pressure was augmented by increasing the number of washing steps and decreasing the amount of antigen added (200, 100, 50 and 10 pmol, respectively) between the different rounds. The enriched phages were recovered using trypsin digestion and amplified overnight by infection of E. coli XL1-blue. Amplification in round 1 was done using agar plates and in rounds 2, 3 and 4 in solution. Amplified phages were precipitated using PEG/NaCl and used for the next round of selection. Phagemid DNA from rounds 3 and 4 were purified and the scFv genes were transferred to an expression vector as described previously (Preger et al., 2020).
Following selections, cloning and transformation, a total of 188 colonies were picked from rounds 3 and 4 and analysed further. ELISA revealed 8 colonies as potential binders, of which 5 were identified as unique clones after DNA sequencing. Based on these results (Figure S1, HTRF data not shown), four scFv candidates, scFv-1, scFv-2, scFv-3, scFv-4, were selected to be produced in large-scale and tested in additional binding kinetics and co-crystallisation trials.

**ScFv binding kinetics**

ScFv-1 was determined to be a weaker binder of TREM2 (Figure S1) and thus was not analysed in detail. Approximate binding kinetics of Protein A purified ScFv-2, scFv-3 and scFv-4 were determined by SPR performed on a Biacore 8K instrument. Biotinylated TREM2 (19-174) produced in Expi293F™ human cells was immobilised on a streptavidin-coated chip supplied in the Biotin CAPture kit (28-9202-34, GE Healthcare) at 0.5 μg/mL concentration. ScFvs were diluted in 10 mM HEPES, 150 mM NaCl, 0.05% P-20 and injected at a flow rate of 30 μL/min for 60 seconds followed by dissociation for 120 seconds. The binding response was calculated after subtracting signal coming from a blank flow cell and the buffer. Langmuirian 1:1 model was used to calculate the binding kinetics. For scFv-3 and scFv-4 avidity contributed to obtained values. Additionally, scFv-4 showed some non-specific interaction with the reference sensor, and so calculated values are used as approximation only.

**Crystallisation**

**scFv-4 PDB: 6YMQ**

After Protein A purification, scFv-4 in 100 mM Tricine, pH 8 and TREM2 (a.a. 1-131) were mixed at molar ratio 1:1:1 and buffer exchanged into 20 mM Tricine pH 8, 200 mM NaCl, 200 mM L-arginine, 200 mM L-glutamic acid. The complex was concentrated to 14 mg/mL and crystals were grown by sitting drop vapour diffusion in solution containing 39% MPD, 0.2 M ammonium acetate, 0.1 M citrate pH 5.5 at 20°C. Crystals were mounted directly from the drop and vitrified in liquid nitrogen.

**scFv-4 PDB: 6Y6C**

Antibody scFv-4 was buffer exchanged into 25 mM HEPES, 300 mM NaCl and 5% glycerol and mixed with TREM2 (a.a. 1-174) at molar ratio 1:1:1. The complex was concentrated to 12.5 mg/mL and crystal plates were set up after 32,000 x g spin for 20 min. Protein crystals appeared after 1 day using a reservoir solution containing 0.2 M potassium chloride, 35% pentaerythritol propoxylate 5/4, 0.1 M HEPES pH 7.5 at 20°C. Crystals were cryo-protected after addition of reservoir solution containing 20% ethylene glycol and vitrified in liquid nitrogen.

**scFv-2 PDB: 6YYE**

Protein A purified antibody scFv-2 in 100 mM citrate buffer and 250 mM L-arginine and 50 mM L-glutamic acid was mixed with TREM2 (a.a 19-131) at a molar ratio 1.3:1 and buffer exchanged on PD10 column to 60 mM Tricine 8, 200 mM NaCl containing 200 mM L-arginine, 200 mM L-glutamic acid. The complex was concentrated to 16 mg/mL and precipitation was removed by centrifugation. Crystals were grown in 15 mM nickel chloride, 0.1 M Tris pH 8.5, 2.2 M ammonium acetate at 20°C. Crystals were cryo-protected with reservoir containing 20% ethylene glycol before vitrification.

**Data collection and structure determination**

For scFv-4 structures (6YMQ, 6Y6C), data were collected on beamline I24 at the Diamond Light Source using the X-ray wavelength of 0.9688 Å and processed using Xia2 package and DIALS (G. Winter 2010; Winter et al., 2018).

For the lower resolution scFv-2 dataset (6YYE), data were collected on beamline I03 at the Diamond Light Source using the X-ray wavelength 0.9763 Å. The initial model was built using PhenixRefine and coot modelling using Xia and DIALS dataset. Later the model was used to refine against AutoProc processed dataset with ellipsoidal truncation by Staraniso (Evans, 2006; Kabsch et al., 2010; Vonrhein et al., 2011, 2018). The model was finished by refinement in Buster v.2.10.3 (Bricogne et al., 2016) and final rounds of Phenix refine (Liebschner et al., 2019).

The first dataset of scFv-2 (6YYE) was phased using the structure of another scFv (PDB: 6G8R) generated from the same library and solved previously (Fairhead et al., 2019). TREM2 was phased using PDB 5UD7 (Sudom et al., 2018). The resulting scFv-2 co-structure (PDB 6YYE) was used for phasing of PDB 6YMQ. Phenix molecular replacement was used to phase the data and coot to build a model (Emsley et al., 2010; Liebschner et al., 2019). Models were improved by iterative cycles of Phenix Refine and coot rebuilding.

| Ramachandran statistics       | 6Y6C (scFv-4) | 6YMQ (scFv-4) | 6YYE (scFv-2) |
|-------------------------------|--------------|--------------|--------------|
| Ramachandran favoured (%)     | 97.21        | 92.59        | 93.73        |
| Ramachandran allowed (%)      | 2.79         | 7.41         | 5.97         |
| Ramachandran outliers (%)     | 0.0          | 0.0          | 0.3          |
| Rotamer outliers (%)          | 0.35         | 0.92         | 2.35         |
| Clash score                   | 2.56         | 4.76         | 6.01         |
Molecular graphics and analyses were performed with UCSF ChimeraX, developed by the Resource for Biocomputing, Visualization, and Informatics at the University of California, San Francisco, with support from National Institutes of Health R01-GM129325 and the Office of Cyber Infrastructure and Computational Biology, National Institute of Allergy and Infectious Diseases (Goddard et al., 2018). Crystal lattice images were generated using PyMOL (Schrödinger, LLC.). See Key Resources Table for all software.

Mammalian cell culture for cell biology

The HEK293 TREM2-DAP12 cell line was a kind gift from Professor Peter St-George Hyslop and generated as described previously (Thornton et al., 2017). HEK293 wild type cell line was purchased from ATCC (CRL1573). WT HEK293 and HEK293 cells stably expressing WT hTREM2 and hDAP12 were cultured in T-175 flasks (Greiner Bio-one) in DMEM (GIBCO) with 10% heat-inactivated foetal bovine serum (FBS) (Gibco) at 37°C, 5% CO₂. The cell culture medium for the HEK293 TREM2-DAP12 cells was supplemented with 1 mg/mL puromycin hydrochloride (Cayman Chemical).

sTREM2 ELISA

HEK293 transfected with TREM2-DAP12 were plated in 96-well plates (Corning Biocoat, 354640) at 30x10⁵ cells per well, grown for 24 h, washed with culture medium and treated with antibodies for 5 h. After antibody incubation, the medium was collected and transferred into a new plate, centrifuged at 1,000 x g followed by supernatant transfer into a fresh plate that was stored at -80°C.

TREM2 ELISA was performed by coating high bind 96-well plates (Sigma, M4436-040EA) with 8 μg/mL TREM2 antibody (Abcam, ab209814) O/N at 4°C followed by 1 h blocking using PBS with 1% BSA (Sigma). 100 μL of the defrosted cell supernatants were added at the appropriate dilution in DMEM + 1% BSA alongside a standard curve with TREM2 peptide (SinoBiological, 11084-H08H) and incubated for 2 h, at RT. 25 μL of the biotinylated detection antibody (R&D, BAF1828) at 1.5 μg/mL was added for an hour followed by the addition of HRP conjugated streptavidin diluted at 1:30,000 (ThermoFisher Scientific, N100). TMB ELISA substrate (ThermoFisher Scientific 34028) was added for 20 min followed by 2 M sulphuric acid (Immuno Chemistry technologies). Results were obtained by measuring optical density (OD) at 450 nm using a plate reader (SpectraMax M2). OD measurements in the linear range (0.8-1.9) were chosen for analysis and TREM2 concentration was interpolated from the TREM2 standard curve. To ensure that scFv binding to TREM2 did not interfere with the assay antibodies, the standard peptide (a.a 1-174, SinoBiological 11084-H08H) was mixed with each one of the scFvs in DMEM +1% BSA and results were compared to the standard curve only. Also, as another control, to assess TREM2 epitope recognition by the ELISA antibodies, TREM2 proteins (His19-Ala138 and His19-Ser174) were diluted to 2441 pg/mL in DMEM +1% BSA using five-point serial dilutions and analysed using the protocol described above. Additionally, the medium conditioned in cells for 5 h was spiked with scFvs and ELISA was performed in comparison to non-spiked sample (data not shown).

Alexa fluor 647 cell staining and imaging

Purified scFv-3 and scFv-4 oligomers and control non-TREM2 scFvs were incubated with Protein A and Alexa Fluor 647 NHS ester (Thermofisher Scientific A37573) at 1:10 molar ratio for 20 h, at 4°C and excess dye was removed by NAP5 columns (GE healthcare). HEK293 and HEK293 transfected with TREM2-DAP12 were prepared as for the ELISA and treated with scFv-4 for 5 h. Cells were fixed in 4% paraformaldehyde in PBS (Santa Cruz Biotechnology) for 15 min and kept at 4°C until imaging. Nuclei were stained with Hoechst 33342 for 1 h in the dark and cells were imaged using an OperaPhenix confocal microscope with 40x magnification, 405/456 nm and 640/706 nm, excitation/emission respectively.

QUANTIFICATION AND STATISTICAL ANALYSIS

All statistical analyses were carried out using three independent biological replicates and two or three technical repeats unless specifically stated otherwise. Error bars are standard deviation (SD) and EC₅₀ was interpolated using non-linear regression (inhibitor vs. response- four parameters) in GraphPad Prism Software 8.4.2. Each statistical test was performed on raw data normalised to cell number or signal before normalisation to the percentage of control was performed. One-way ANOVA with Dunnett’s test comparison to control untreated cells was utilised to calculate significance. p values are indicated in the respective figure legends with symbols (*P <.05 and **P <.01; ****P <.0001).