Role of Nanodiamonds in Drug Delivery and Stem Cell Therapy

Shakeel Ahmed Ansari 1*, Rukhsana Satar 2, Mohammad Alam Jafri 1, Mahmood Rasool 1, Waseem Ahmad 1, Syed Kashif Zaidi 1

1Center of Excellence in Genomic Medicine Research, King Abdulaziz University, Jeddah-21589, Kingdom of Saudi Arabia
2Department of Biochemistry, Ibn Sina National College for Medical Sciences, Jeddah-21418, Kingdom of Saudi Arabia

*Corresponding author: Shakeel Ahmed Ansari, Center of Excellence in Genomic Medicine Research, King Abdulaziz University, Jeddah-21589, Kingdom of Saudi Arabia. Tel: +96-026401000 (Ext: 25478), Fax: +96-6126952521, E-mail: saansari@kau.edu.sa

Received: August 19, 2015; Revised: February 16, 2016; Accepted: March 05, 2016

Context: The use of nanotechnology in medicine and more specifically drug delivery is set to spread rapidly. Currently many substances are under investigation for drug delivery and more specifically for cancer therapy.

Evidence Acquisition: Nanodiamonds (NDs) have contributed significantly in the development of highly efficient and successful drug delivery systems, and in stem cell therapy. Drug delivery through NDs is an intricate and complex process that deserves special attention to unravel underlying molecular mechanisms in order to overcome certain bottlenecks associated with it. It has already been established that NDs based drug delivery systems have excellent biocompatibility, non-toxicity, photostability and facile surface functionalization properties.

Results: There is mounting evidence that suggests that such conjugated delivery systems well retain the properties of nanoparticles like small size, large surface area to volume ratio that provide greater biocatalytic activity to the attached drug in terms of selectivity, loading and stability.

Conclusions: NDs based drug delivery systems may form the basis for the development of effective novel drug delivery vehicles with salient features that may facilitate their utility in fluorescence imaging, target specificity and sustained-release.

Keywords: Biomedical applications; Drug delivery; Hydrogels; Nanodiamonds; Surface functionalization

1. Context
   Nanoparticles mediated drug delivery has attracted the attention of researchers for safely transferring the drug of choice to the site of interest in a biological system apart from ensuring their biocompatibility (1,2). These nanoparticles prevent high systemic loads of drugs at specific targeted sites that usually results in manifestation of unwanted toxicity. The process of nanoparticle based drug delivery occurs either by delivering drugs into the cytoplasm or through extracellular domains of transmembrane signaling molecules (3). Since human digestive tract contains greater acid/enzyme content, it degrades protein and peptide drugs before they are absorbed into the bloodstream. Therefore, acid labile drugs need adequate protection against unfavorable acidic environment of the gut. Apart from this, several charged compounds and macromolecules possess weak partitioning properties which make them poorly absorbed through biological membranes. Hence the need of nanoparticle based drug delivery systems arises which can inhibit acid and peptidase mediated degradation thereby allowing sufficient time for the drug to be absorbed via gastrointestinal tract epithelium (4). Additionally, nanoparticle based delivery vehicles may be exploited to release pharmacologically relevant drug concentration at specific target sites such as malignant tumors.

2. Evidence Acquisition

2.1. Benefits and Limitations of Existing Nanoparticle Based Drug Delivery Platforms
   During past couple of decades, several studies have significantly contributed for the development of novel nanoparticle based platforms with different compositions and biological properties for plethora of drug and gene delivery applications (5-7). Quantum dots, chitosan and polylactic/glycolic acid based nanoparticles...
have also been suggested for in vitro RNAi delivery (8). The brain cancer is one of the most difficult to manage malignancy mainly because of the inability of therapeutic agents to pass through the blood brain barrier (BBB) and reach brain tumor, therefore, loperamidine and doxorubicin were successfully bound to nanomaterials to cross the intact BBB in order to release them in desired therapeutic concentrations to the brain (9). Moreover, peptide-based nanotubes were also exploited to target vascular endothelial growth factor receptor and cell adhesion molecules like integrins, cadherins and selectins, as a new therapeutic approach to control disease progression (10).

Inspite of observed promising benefits of nanoparticle based drug delivery platforms in addressing numerous key issues associated with current clinical practice for the treatment of difficult to treat diseases, there are certain well-known challenges in translating this exciting technology for the clinical application. The potential toxicity of nanoparticles remains one of the major concerns in the field of nanoparticle based drug delivery. Therefore, an in-depth knowledge about the nature and mechanism of nanoparticle-induced toxicity is required to facilitate their successful clinical translation. For example, their toxicity on central nervous system and their interactions with the cells and tissues are poorly understood (11). Furthermore, nanoparticles usually have short circulation half-life due to faster elimination through opsonization by phagocytic cells inside the human body. Hence, the nanoparticle surfaces need to be modified in a way to avoid phagocytosis by circulating macrophages.

Recent years have witnessed significant increase in the application of carbon based nanoparticles in the field of biomedicine and electronics (12) which has led to the emergence of NDs as novel constituents of carbon based nanoparticles. They have excellent biocompatibility, photostability and facile surface functionalization properties which could be utilized to promote longer circulation half-lives, to improve drug pharmacokinetics and reduce side effects of therapeutically active substances. These outstanding features of NDs might also help in devising drug-delivery vehicles with the ability to overcome the action of drug efflux pumps, thus minimizing cancer chemo-resistance and maximizing the therapeutic efficacy of the anticancer drugs (13-14).

2.2. Nanodiamonds (NDs)

NDs are carbon-based nanomaterials that provide large surface area. They can be functionalized with different ligand molecules, which can be used to conjugate various compounds or drugs (15). Same as bulk diamond, the basic crystal structure of an ND consists of a nanocrystal having tetrahedral bonded carbon atoms in the form of a three-dimensional cubic lattice which imparts the properties of diamond and an onion shaped carbon shell containing a coat of functional groups on the surface (16,17). However, ND is often described as a crystalline diamond core surrounded by an onion like amorphous graphite shell (18). The sp²/sp³ bonds in ND are quite flexible endowing it with the ability to assume two geometrical forms for example, the stretched face of diamond can behave as a graphene plane whereas the puckered graphene may become a diamond surface. This unique characteristic feature of ND particles promises a greater degree of template flexibility particularly around the curved surface where electrons are unstable (19,20).

NDs can be easily doped with nitrogen and possess nitrogen-vacant (NV) defect centers in their crystal lattice which make them suitable as photoluminescent probes for several in vivo and in vitro applications (15). The presence of the NV centers - a nitrogen atom next to a vacancy in NDs, leads to useful fluorescence properties. They emit bright fluorescence at 550-800 nm from NV centers produced by high-energy ion beam irradiation and subsequent thermal annealing. The excellent emission property, together with negligible cytotoxicity and easiness of surface functionalization, makes NDs a promising fluorescent probe for single-particle tracking in heterogeneous environments as well as using them as biomarkers and in biolabelling studies (21). Research activities concerning the bioimaging and drug delivery applications of NDs have also been discussed in detail by previous researchers (22). The focus was on the prospective of using NDs in bioimaging, their interaction with biological objects and possibility of modifying their surface by attaching biological/medical molecules for drug delivery applications (23).

Several studies suggested that NDs can be easily functionalized by covalent and non covalent method, and their biocompatibility is superior over other carbon containing nanomaterials like single walled and multi walled nanotubes, and carbon blacks (24). They have been classified on the basis of their particle size and on the method of synthesis (Figure 1).

2.3. Surface Functionalization of NDs

The surface of NDs can be modified by many functional groups for imparting extra stability (Figure 2).
These surface modifications can be achieved through covalent and/or non-covalent bonds with different methods, though each method has its own advantages and disadvantages. Covalent modifications of NDs have resulted in stable drug complexes. However, the used processes are typically complex (26, 27), suffering from difficulty in toxic solvents removal introduced during preparation, and inconsistent slow release of the drug. On the contrary, drugs can be attached to NDs easily by non-covalent methods, however with lower stability. Loading of drug by this method can be modulated simply by controlling con-
centration of involved inorganic molecules such as NaOH and NaCl. This method also imparts slow-release of drug into the cells, thereby reducing the possibility of undesired side effects of drug. Therefore, at present, non-covalent method is preferably chosen to build ND-based drug delivery systems. Arnault has also discussed the control of the surface chemistry of NDs for improving their colloidal behavior (28). NDs possess high surface free energy, which allows to form clusters of about 10-100 nm even when they are dispersed in a solution. Additionally, numerous functional molecules and variety of drugs have been attached successfully on their surface by non-covalent interactions (29). NDs can be embedded in a silica shell terminated by a hydrophilic, azide- or alkyne-substituted PEG polymer. It was observed that in contrast to unmodified NDs, this nanoarchitecture brings high colloidal stability in buffers and biological media. Furthermore, such NDs allow the possibility of modifying particles by biomolecules in high yields using click chemistry (30, 31).

Hydrophilic moieties containing chlorine, ketonic and carboxylic group have been introduced earlier on the surface of NDs by a thermal induction and plasma treatment method (32-34). Moreover, covalent attachment was also obtained onto NDs by using alkyl chains, fluorine and silicon (35-37). Surface modified ND films were used to adsorb small molecules like allyl alcohol, sulfonic acids, thiols, and complex structures like DNA and enzymes in preparing sensors and for the delivery of therapeutic agents (38-42). The predominant functional moieties present on the surface of NDs include carboxylic (COOH) and ketonic (C=O) groups apart from having alcoholic and ether groups (43-46). Moreover, amine group was introduced on the surface of NDs by reacting chlorinated diamond with gaseous ammonia at elevated temperature, using sub-micron particles by several investigators (47,48). In another novel approach, ultra-bright fluorescent NDs were developed and their thermal and kinetic optimum of nitrogen-vacancy (NV) center formation was identified (49). The presence of NV centers imparted excellent biocompatibility, bright fluorescence and high magnetic sensitivity at ambient conditions. These fluorescent NDs have great potential in designing nanosensors (50). The core/shell structures of NDs surrounded by porous silica shells have demonstrated a remarkable increase in drug loading efficiency. Additionally, such NDs decorated with carbon dots have shown excellent potential as bioimaging probes (51). Attachment of various drugs to NDs has been listed in Table 1.

A large number of covalent and non-covalent methods have been used in past to modify NDs. Surface modification of NDs by N,O-carboxymethyl chitosan and benzoquinone to assist the binding of drug on these modified NDs via non-covalent methods have been reported elsewhere. The results were illustrative of lower cytotoxicity and absence of photo-bleaching. Furthermore, performing in vivo studies, biological imaging, drug delivery and other theranostic applications have been envisaged (52-55). Contrarily, Liu et al. (56) reported the attachment of paclitaxel to surface modified NDs prepared by covalent method. It was observed that application of 0.1-50 μg.mL⁻¹ modified ND based paclitaxel significantly decreases the cell viability of A549 human lung carcinoma cells by inducing mitotic arrest and apoptosis, consequently blocking the tumor growth.

3. Results

3.1. In vitro and In vivo Toxicity Studies of NDs

Negligible cytotoxicity has been reported for NDs with little changes in gene expression under physiological conditions (75). No significant changes were observed in the expression of TNFα and Bcl-x genes after incubation with the acid purified NDs when compared with controls. Their therapeutic potential was confirmed by observing ND-induced morphological changes in HT-29 human colorectal adenocarcinoma cells and effect on cell viability on murine macrophages (75). Chow and co-workers noticed that adsorption of doxorubicin was increased significantly on NDs after addition of NaCl (10 mg.mL⁻¹) to the
reaction system. Moreover, NDs bound doxorubicin exhibited less toxicity on RAW264.7 mouse macrophage cell line and HT-29 human colon cancer cells as compared to doxorubicin alone. The possible explanation might be that NDs provided a protective effect to doxorubicin by favoring the slow release of drug and preventing the normal cells from its side effects. They also used this drug complex for the treatment of liver and breast cancer, and observed significant killing of drug resistant cells at relatively lower dosages, resulting in the decreased toxicity of the drug. Doxorubicin was released from this complex steadily and exhibited increased retention in blood circulation and tumor causing enhanced inhibition of tumor growth as compared to usual doxorubicin treatment in tumor models (76). These studies provide a promising foundation to find ND-based drug development and their potential clinical applications.

It was shown that optimum binding of doxorubicin on NDs occurs only at high pH and availability of at least 10% of the ND surface area (77). A comparative study reported that NDs were significantly non-toxic to macrophages and neuroblastoma cells, preventing the formation of reactive oxygen species as compared to carbon black, multi-walled carbon nanotubes and single-walled carbon nanotubes (78, 79). In another study, A549 lung cancer cells and 3T3-L1 embryonic fibroblasts were incubated with NDs for 10 days, and no adverse effect on cell or gene expressions was observed in cancer cell lines during their progression and adipogetic differentiation (68). Vaijayanthimala et al. (12) observed that NDs can be readily taken up by 3T3-L1 pre-adipocytes, HeLa cancer cells and 489-2 osteoprogenitors cells in large amount suggesting their greater biocompatibility. Ease in the adsorption of doxorubicin (anticancer drug), purvalanol A (drug for liver cancer), hydroxytamoxifen (drug for breast cancer), dexamethasone (anti-inflammatory drug), insulin (drug for diabetes) and TAT (a cell penetrating peptide) onto NDs have been reported, preventing premature release and improving therapeutic efficacy (76, 80-83).

The toxicity of fluorescent NDs (FND) in long term in vivo imaging was evaluated in C. elegans (84). FNDs were given to C. elegans either by feeding with colloidal FND solution or microinjecting FND suspension into the gonads. The toxic potential of FND was ascertained by assessing its effect on longevity and reproductive potential. The result obtained in study demonstrated that FNDs were devoid of any toxicity and did not cause evident stress to the worms. FNDs were found to be highly efficient in continuous imaging of entire alimentary canal and mapping of several biological processes including development of the living organism from fertilized egg for several days.

### 3.2. Application of NDs in Drug Delivery System

A new drug delivery system consisting of ND-embedded in contact lens (for drug sequestration and

---

Table 1. Ligands used to modify the surface of NDs

| Ligands                          | Applications                                                                 | References |
|---------------------------------|-----------------------------------------------------------------------------|------------|
| Carboxylic acid                 | Adsorption of mycotoxins, aflatoxin B1 and ochratoxin A                      | (57,58)    |
| Benzoyl peroxide and dicarboxylic acid | Binding anticancer drugs                                                     | (59)       |
| Fluorine                        | Development of ND based bioconjugates for labeling, drug delivery and other applications | (60-63)    |
| Lysine                          | Development of non-viral vectors for transferring genetic materials across cellular membranes | (64,65)    |
| Alkyl lithium, ethylenediamine, glycine ethyl ester generated alkyl | Biolabelling and specific binding of drugs for efficient drug delivery | (47)       |
| Glutaraldehyde                  | Enzyme immobilization                                                       | (66)       |
| Thiolated                       | Therapeutic and drug delivery applications                                   | (67)       |
| CO, OH, and NH-groups           | Binding protein drug with excellent drug release properties                  | (68-70)    |
| N,O-carboxymethyl chitosan      | Production of highly dispersed suspensions of detonated NDs in polar media directly from dry powders | (71)       |
| Hydrocarbon and Perfluorobutyl radical | Chemical applications                                                        | (72,73)    |
| Hydrogenation and hydroxylation | Analytical applications                                                       | (58)       |
| Hydrogen                        | Integration of epoxy polymer composites for corrosion resistant coatings, as well as biomedical applications by attaching complex molecules such as proteins and DNA | (74,59)    |
| Chlorine, Glycine               |                                                                            | (47)       |
sustained release) was tested using timolol maleate (drug used for treating ocular disease, glaucoma) in the presence of lysozyme in trabecular meshwork cells (85). This study established that the ND-embedded lenses can be utilized for drug sequestration, controlled release and enzyme activation purposes. In another study, NDs were used to deliver an anti-cancer drug 10-hydroxy camptothecin (HCPT) in the presence of NaOH (pH 8.2) to enhance the adsorption of HCPT on NDs. It was determined that NDs-HCPT complex showed slow and steady release of the drug under slightly acidic condition and was highly effective on tumour cells as compared to free HCPT (86). Similar studies were also carried out by adsorbing cisplatin onto NDs. Contrarily, the drug in this case was released at high rate in the presence of PBS buffer (pH 6) from the ND complex (87). It should be noted that subcutaneously injected NDs for three months in a mouse model did not elicit any inflammatory response (88). Xing et al. (89) observed improved expression of DNA repair proteins, p53 and MOGG-1 when embryonic stem cells were challenged with NDs. Bakowicz and Mitura (90) also reported the absence of adverse immune response in rats following 10 days of intraperitoneal injection of NDs. These studies suggested excellent biocompatibility of NDs as they did not evoke any inflammatory immune response at injection site when administered through various routes in animal models.

Several molecules like insulin, cytochrome c, lysozyme, apoobelin were immobilized on NDs by non-covalent method to improve their functionality (91-94). In another study, L-polylysine and polyarginine were used to produce their respective ND conjugates containing greater number of primary amino groups (12, 95). Polyaniline, polyethyleneimine and poly(lactic acid) were also used to functionalize NDs by amide bond formation (96-99). Moreover, Sulfur, thiols and sulfonic acids were also used for modifying NDs in preparing plasmonic structures and for conjugating very complex (bio) molecules efficiently (100-102). An excellent review has appeared recently dealing with all the possible transformations of NDs in great detail (103).

Carbon nanoparticles possess small size, penetrate through the cells easily and can be surface functionalized. These properties enabled their application in designing drug delivery vehicles for attaching small molecule drugs and peptides (104-107). However, carbon nanotubes and graphene are the two forms of carbon based nanoparticles that showed cytotoxicity, oxidative stress and apoptosis (108-110). Thus, the need arises to have safer drug delivery systems that employ surface modified nanoparticles. The NDs are such versatile tools for building drug delivery systems owing to their chemical inertness, optical transparency, robust hardness high specific area, and excellent biocompatibility. Promising data, apart from low toxicity and high cellular uptake, have been reported for use of NDs in complete cell culture media (111). NDs, as a drug carrier, can be used as a thin film for efficient interaction of drug, and in the form of spontaneous clusters known as ND hydrogel that favors attachment of drug. It should be noted that ND based films can be introduced immediately after surgical removal of a tumor to target residual cancerous cells to prevent the tumor from recurring. Thus, they have gained extensive attraction in developing highly efficient drug delivery systems for treating superficial tumors such as skin, breast, head and neck cancers, and superficial skin inflammations where the drug is delivered transdermally to tumors or inflammation sites thereby reducing the toxicity on normal tissues. Table 2 shows the list of studies where NDs were used to develop drug delivery systems.

The simplest way to develop ND films was achieved by self-assembly process in which ND hydrogels were

| Drugs | Applications | References |
|-------|--------------|------------|
| Doxorubicin | Treatment of colorectal carcinoma (75, 76, 80, 86, 24, 124) |
| 10-hydroxy camptothecin (HCPT) | Intracellular transportation of chemotherapeutic drug (86) |
| Paclitaxel | Drug delivery and cancer therapy for lung carcinoma (56) |
| Insulin | For delivering insulin and other therapeutic treatments (81) |
| Purvalanol A | Treatment of liver carcinoma (80) |
| 4-hydroxytamoxifen | Treatment of breast carcinoma (80) |
| Si RNA | In vivo imaging (84) |
| polymyxin B | Development of drug delivery platforms (24) |
| Epirubicin | Overcoming chemoresistance in hepatic cancer stem cells (128) |

| Ansari SA. et al. | Iran J Biotech. 2016;14(3):e1320 | 135 |
fixed on poly-lysine coated glass substrate and the thickness of ND-poly-lysine film was obtained through layer-by-layer technique (112-115). The films thus formed could assemble several biological molecules. The cellular gene expression studies and MTT assays revealed excellent biocompatibility and safety of the ND films. The self-assembled doxorubicin on ND films showed anti-inflammatory effects as well as slow and controlled release when tested against RAW264.7 cells apart from demonstrating significant reduction in toxic side effects on the normal tissues. ND bound doxorubicin sandwiched in a Parylene C polymer microfilm was found to be stable; slowly released even after a month due to the potent sequestration ability of the doxorubicin-ND complex and the release modulating nature of the thin Parylene layer (116). Moreover, Xi et al. (117) utilized NDs to enhance delivery of DOX in a preclinical glioma model using a convection-enhanced delivery method and demonstrated remarkably enhanced efficacy of doxorubicin. It was found that NDs markedly enhanced DOX uptake and retention in glioma cells. Thus, ND-DOX delivered by this method was significantly more efficient in killing tumor cells than uncomplexed DOX.

3.3. Applications of NDs Based Hydrogel
The research on NDs based hydrogels has mainly concentrated on their evaluation as a vehicle for targeted transportation, controlled release, declined drug concentrations and reduced side effects. This material also provides better penetration of the drug complex to gain access to the interior of the cells. Some of the improvements using NDs based hydrogels for drug delivery are as follows. Zhang et al. (107) reported heterofunctionalization of a 2-8 nm ND for a drug delivery system. This multifunctional system has simultaneous capabilities of targeting, imaging and enhancing therapy. ND based hydrogels were conjugated with the anticancer cell proliferation-inhibiting and apoptosis-stimulating paclitaxel-DNA via fluorescently labeled oligonucleotide strands, and with anti-EGFR monoclonal antibodies. The feasibility of these conjugates was studied on human breast cancer cell lines (MDA-MB-231 and MCF7). Fluorescently labeled oligonucleotide linkers enabled the intracellular observation and quantification of resultant ND conjugates and anti-EGFR monoclonal antibodies provided the internalization and delivery of anticancer agents into EGFR-overexpressing cells. The results showed enhanced cellular internalization and therapeutic activity of covalently attached chemotherapeutic and targeting moieties on ND surface. Liu et al. (118) also successfully covalently linked 3-5 nm sized NDs with paclitaxel for drug delivery and cancer therapy. The results showed that ND-paclitaxel significantly reduced the cell viability and induced mitotic arrest, apoptosis and anti-tumorigenes in A549 (human lung carcinoma) cells, thereby inducing cell death in A549 cells. Furthermore, ND-paclitaxel markedly blocked the tumor growth and formation of lung cancer cells in xenograft SCID mice.

3.4. NDs in Stem Cells
Stem cells have increasingly become the focus of attention in translational research as well as in the development of human cellular therapy for difficult to treat diseases. There is an urgent need for the development of novel and improved methods to tract the fate of a specific stem cell under in vivo conditions. A significant amount of data is available to support the fact that nanoparticles including NDs can be utilized for tracking stem cells in vivo without affecting their normal biology. It has been demonstrated that the presence of fluorescently labeled superparamagnetic iron oxide particles (~0.9 μm) and carboxylated NDs (~0.25 μm) did not adversely affect normal morphology, osteogenic and adipogenic differentiation potential, CD marker expression, cytokine secretion and other biochemical parameters of adipose-derived mesenchymal stem cells (119). In addition, several studies have clearly established excellent biocompatibility of NDs in a variety of cell lines without noticeable cytotoxicity (88). However, it has also been reported that NDs slightly stimulate expression of DNA repair proteins such as p53 and MOGG-1 in embryonic stem cells suggesting DNA damage. Furthermore, it was observed that oxidized NDs have higher potential to cause DNA damage compared to raw NDs, possibly due to specific surface chemistry. The lung stem cells are being extensively considered for the treatment of lung diseases particularly as regenerative therapy for repairing damaged or lost lung tissues in patients. Therefore, accurate tracking of stem cells under in vivo conditions constitutes an important step in the process of understanding how stem cells differentiate and regenerate themselves. Numerous optical methods and photo-probes have been employed for this purpose but most of these methods are limited due to undesired photo-bleaching, toxicity and interference from background tissue. Wu et al. (120) have demonstrated that FNDs can be utilized to track and identify engrafted
lung stem/progenitor cells by recognizing CD45–CD54+CD157+ surface markers employing fluorescence-activated cell sorting or fluorescence lifetime imaging microscopy techniques in order to evaluate their regenerative capabilities. It was also observed that FND labelling did not interfere with normal stem cell properties including self-renewal and differentiation into functional pneumocytes. The NDs have also been used for target-based delivery of drugs in the stem cells. For instance, epirubicin, an anthracycline anticancer drug attached to NDs was delivered in hepatic cancer stem cells to overcome chemoresistance. It was observed that epirubicin bound to NDs created a rapidly synthesized and stable NDs-epirubicin complex that promoted the endocytic uptake and enhanced tumor cell retention. These attributes mediated effective killing of both cancer stem cells and non-cancer stem cells in vitro and in vivo (121).

4. Conclusions

Owing to excellent biocompatibility, bright photoluminescence based on color centers and outstanding photostability, NDs could provide much needed foundation to develop application-specific drug delivery vehicles to treat various cancers. This can be achieved by preparing NDs in a systematic manner by combining disciplines such as nanomanufacturing, drug discovery and molecular biology/medicine which will serve as collaborative roles in downstream realization of clinical ND. Moreover, the studies showed that the surface of NDs can be modified by several functional groups via covalent and non-covalent methods for imparting extra stability to them. Hence, in near future, NDs based drug delivery systems can provide hope for effective treatment of aggressive diseases and treating resistant diseases in large. The progress in the development of a number of methods of ND functionalization with different kinds of drugs will definitely open possibilities to apply constructed systems not only of drugs but also gene and protein delivery. Moreover, continuous research on NDs is needed to seek further development of including magnetism (FRET and magnetic resonance) and nonlinear optical properties (optical trapping) in NDs so that they can be successfully employed for bioimaging, labeling and drug delivery systems.

5. Future Perspective

There has been a considerable progress in the applications of ND in the field of biomedical sciences including bio-imaging and targeted drug delivery in the past few years. However, some bottlenecks still exist that have to be overcome. For medical (clinical) applications, size of NDs is a limiting factor. Aggregation of NDs is a serious problem, especially for sizes smaller than 50 nm. Therefore, an effective method is needed to prevent cluster aggregation. Majority of the current studies have mainly used cellular models or microorganisms for their experiments. The interaction of NDs with animal organs and the fate of ND/drug clearance in the animal body have not been systematically studied. If ND-based drug delivery is to be converted into successful clinical application, well designed comparative studies using important drug delivery methods are also warranted. Research on NDs continues to seek further development of novel methods such as addition of magnetism and nonlinear optical properties such as optical trapping, FRET and magnetic resonance in NDs. These properties will aid in successful validation of bioimaging, labeling and drug delivery tracing using NDs for future biomedical applications.

Acknowledgements

The authors are thankful to Dr. Peter Natesan Pushparajan for making the first figure of the manuscript.

Authors’ Contributions

Study concept and design: Shakeel Ahmed Ansari and Rukhsana Satar. Acquisition of data: Rukhsana Satar and Mohammad Alam Jafri. Drafting of the manuscript: Rukhsana Satar, Syed Kashif Zaidi and Mahmood Rasool. Critical revision of the manuscript for important intellectual content: Waseem Ahmad and Mohammad Alam Jafri.

References

1. Rasool M, Naseer MI, Arif MM, Manan A, Ullah I; Bin AN, Ok KM, Husain QM, Kamal MA, Bibi F. Recent developments in nanomedicines for management of various health issues via metabolism and physico-chemical properties. Curr Drug Metab. 2015a;16:389-396. DOI: 10.2174/1389200215666141125121212

2. Rasool M, Malik A, Manan A, Ansari SA, Asif M, Qazi MH, Kamal MA, Pushparaj PN. Scope and applications of nanomedicines for the management of multiple sclerosis. Curr Drug Metab. 2015b;16:602-608. DOI: 10.2174/138920021666150807105457

3. Rasool M, Malik A, Manan A, Ansari SA, Naseer MI, Qazi MH, Asif M, Gan SH, Kamal MA. Nanoparticle-based therapy in genomics. Curr Drug Metab. 2015c;16:354-361. DOI: 10.2174/138920021666141208152121

4. Ansari SA, Satar R, Panda DS, Zaidi SK, Chibber S, Khan MJ,
Khan TA, Jafari MA, Alqahtani MH. Surface engineering of multifunctional nanocomposites for biomedical applications: a brief update. Iran J Biotechnol. 2014;12:e11124. DOI: 10.5812/ijb.11124

5. Singh R, Lillard Jr JW. Nanoparticle-based targeted drug delivery. Exp Mol Pathol. 2009;86:215-223. DOI: 10.1016/j.yexmp.2008.12.004

6. Fonseca-Santos B, Gremiao MP, Chorilli M. Nanotechnology-based drug delivery systems for the treatment of Alzheimer’s disease. Int J Nanomed. 2015;10:4981-5003. DOI: http://dx.doi.org/10.2174/IJN.87148

7. Rigon RB, Oyafuso MH, Fujimura AT, Gonçalez ML, do Prado AH, Gremiao MPD, Chorilli M. Nanotechnology-based drug delivery systems for melanoma antimutural therapy: A review. Biomed Res Int. 2015;Article ID 841817. DOI: http://dx.doi.org/10.1155/2015/841817

8. Chen AA, Derfus AM, Khetani SR, Bhatia SN. Quantum dots based drug delivery. J Occupat Med Toxicol. 2007;2:16-22. DOI: 10.1186/1745-6673-2-16

9. Suri SS, Fenniri H, Singh B. Nanotechnology-based drug delivery. J Med All Sci. 2012;31-40.

10. Burgess NC, Sharp TH, Thomas F, Wood CW, Thomson AR, Zaccai NR, Brady RL, Serpell LC, Woolfson DN. Modular design of self-assembling peptide-based nanotubes. J Am Chem Soc. 2015;26:10554-10562. DOI: 10.1021/jacs.5b03973

11. Feng X, Chen A, Zhang Y, Wang J, Shao J, Wei L. Central nervous system toxicity of metallic nanoparticles. Int J Nanomed. 2009;10:4321-4340. DOI: http://dx.doi.org/10.2147/IJN.S78308

12. Vajayanthimala V, Chang HC. Functionalized fluorescent nanodiamonds for biomedical applications. Nanomedicine 2009;4:47-55. DOI: 10.2217/17435889.4.1.47

13. Osaka E, Ho D. Nanodiamond and its application to drug delivery. J Med All Sci. 2012;3:21-40.

14. Igor I, Andrey AS, Torsten R, Steffen S, Sang-Yun, L, Denis A. Molecular-sized fluorescent nanodiamonds. Nat Nanotechnol. 2014;9:54-58. DOI: 10.1038/nnano.2013.255

15. Lam R, Ho D. Nanodiamonds: Applications in Biology and nanoscale medicine. Diam Rel Mater. 2009;13:89-95.

16. Iakoubovskii K, Baidikova MV, Studentt J, Wouters BH, Stesmans A, Kober J, Baidikova M. Structure and defects of detonation synthesis nanodiamond. J Nan Technol. 2015;10:953-954. DOI: http://doi.org/10.1246/cl.1998.953

17. Iakoubovskii K, Matsushita K, Furuya K. High-resolution electron microscopy of detonation nanodiamond. Nanotechnol. 2008;19:16-21. DOI: http://dx.doi.org/10.1088/0957-4484/19/15/155705

18. Shengfu J, Tianlai J, Kang X, Shuben L. Optical properties of nanodiamond layers. Appl Surf Sci. 1998;133:231-346.

19. Krueger A, Lang D. Functionality is key: Recent progress in the surface modification of nanodiamond. Adv Func Mat. 2012;22:890-906. DOI: 10.1002/adfm.201102670

20. Jarre G, Heyer S, Memmel E, Meinhart D, Krueger A. Synthesis of nanodiamond derivatives carrying amino functions and quantification by a modified Kaiser test. Beilstein J Org Chem. 2014;10:2729-2734. DOI: 10.3762/bjoc.10.288

21. Kazi S. A review article on nanodiamonds discussing their properties and applications. Int J Pharm Sci Inv. 2014;3:40-45.

22. Perevedentseva E, Lin YC, Jani M, Cheng CL. Biomedical applications of nanodiamonds in imaging and therapy. Nanomed. 2013;8:2041-2060. DOI: 10.2217/nmn.13.183

23. Reina G, Orlanducci S, Cairone C, Tamburri E, Lenti S, Cianchetta I, Rossi M, Terranova ML. Rhodamine Nanodiamond as a system model for drug carrier. J Nanosci Nanotechnol. 2015;11:1022-1029. DOI: http://dx.doi.org/10.1166/jn.2015.9736

24. Mochalin VN, Amanda P, Xue-Mei L, Gogotsi Y. Adsorption of drugs on nanodiamond: toward development of a drug delivery platform. Mol Pharm. 2013;10:3728-3735. DOI: 10.1021/mp400213z

25. Shenderova OA, Gruen DM. Ultrananocrystalline diamond: Synthesis, properties and applications. Elsevier. 2012;P:584.

26. Basem M, Fhalyl K, Li S, Jafarian A, Khashab NM. Applications of nanodiamonds in drug delivery and catalysis. J Nanotechnol. 2014;1:332-343. DOI: http://dx.doi.org/10.1166/jn.2014.9141

27. Lai L, Barnard AS. Anisotropic adsorption and distribution of immobilized carbonyl on nanodiamond. Nanoscal. 2014;6:14185-14189. DOI: 10.1039/C4NR05363J

28. Arment JC. Novel Aspects of Diamond, Topics in Applied Physics. 2015;P:85-122.

29. Chang BM, Hsin-Hung L, Long-Jyun K, Wen-Der L. Highly fluorescent nanodiamonds protein-functionalized for cell labeling and targeting. Adv Func Mater. 2013;23:5737-5745. DOI: 10.1002/adfm.201301075

30. Rehor I, Slegerva J, Kucka J, Proks V. Fluorescent nanodiamonds embedded in biocompatible translucent shells. Small 2014;10:1029-1034. DOI: 10.1002/smll.201400033

31. Passeri D, Rinaldi F, Ingallina C, Carafa M, Rossi M, Terranova ML, Marianecci J. Biomedical applications of nanodiamonds: an overview. J Nanosci Nanotechnol. 2015;15:972-988. DOI: http://dx.doi.org/10.1166/jn.2015.9734

32. Ohtani B, Kim YH, Yano T, Hashimoto K, Fujishima A, Uosaki K, Kohei H. Surface functionalization of doped CVD diamond via covalent bond. an XPS study on the formation of surface-bound quaternary pyridinium salt. Chem Lett. 1998;15:953-954. DOI: http://dx.doi.org/10.1246/cl.1998.953

33. Ghodbane S, Haensen T, Cossinier Y, Szunertis S, Steinmuller-Nethl D, Boukhherroub R, Ahmed SIU, Schaefer JA. Investigation of the surfaces of nanocrystalline diamond films oxidized by different processes. Langmuir 2010;26:18798-18805. DOI: 10.1021/la1032652

34. Kulisch H, Kulisch W, Popov C, Merz R, Merz B, Reithmaier JP. Plasma immobilized carboxyl on nanodiamond. Adv Func Mater. 2010;20:1777-1782. DOI: 10.1002/adfm.201002631

35. Nicholas BM, Butler JE, Russell JN, Hamers RJ. Photochemical functionalization of hydrogen terminated diamond surfaces: a structural and mechanistic study. J Phys Chem B. 2005;109:20938-20947. DOI: 10.1021/jp0545389

36. Ray MA, Tyler T, Hook B, Martin A, Cunningham G, Shenderova JL. Cool plasma functionalization of nano-crys-
1. Zou Q, Wang MZ, Li YG. Analysis of the nanodiamond particle fabrication by detonation. Nanoscale 2007;16:2087-2089. DOI: 10.1016/j.nanos.2007.07.016

2. Lud SQ, Neppfl G, Richter G, Bruno P, Gruen DM, Jordan R, Feulner P, Stutzmann M, Garrido JA. Controlling surface functionality through generation of thiol groups in a self-assembled monolayer. Langmuir 2010;26:15895-15900. DOI: 10.1021/la102225r

3. Garguilo JM, Davis BA, Buddie M, Kock FAM, Nemanich RJ. Fibrinogen adsorption onto microwave plasma chemical vapor deposited diamond films. Diam Relat Mater. 2004;13:595-599. DOI: 10.1016/j.diamond.2003.10.008

4. Loh KP, Lim CW, Gao X, Qi D, Wee ATS. High resolution electron energy loss and X-ray near edge absorption spectroscopic studies of diamond film functionalized with allyl alcohol. Diam Relat Mater. 2006;15:711-715. DOI: 10.1016/j.diam.2006.01.001

5. Xu Z, Jecidigunta S, Kumar A. A study of polyaniline deposited nanocrystalline diamond films for glucose detection. J Nanosci Nanotechnol 2007;7:2092-2095. DOI: http://dx.doi.org/10.1166/jnn.2007.775

6. Bijnens N, Vermeeren V, Daenen M, Grieten L, Haenen K, Mochalin V, Osswald S, Gogotsi G. Contribution of functional groups to the Raman spectrum of nanodiamond powders. J Phys Chem C 2009;113:3208-3211. DOI: 10.1021/jp902057q

7. Huang H, Chen M, Bruno P, Lam R, Robinson E, Gruen D, Ho D. Nanodiamond-mediated delivery of water-insoluble therapeutic agents to the cytosolic compartment of mammalian cells. J Phys Chem B 2009;113:2966-2971.

8. Mochalin V, Osswald S, Gogotsi G. Contribution of functional groups to the Raman spectrum of nanodiamond powders. Chem Mater. 2009;21:273-279. DOI: 10.1021/cm802057q

9. Comet M, Pichot V, Siegert B, Britz F, Spitzer D. Detonation nanodiamonds for doping Kevlar. Nanoscale 2010;2:4286-4292. DOI: http://dx.doi.org/10.1039/B601325B

10. Ansari SA, et al. Covalent linkage of functionalized detonation nanodiamond-paclitaxel for drug delivery and cancer therapy. Nanotechnology 2010;21:315106. DOI: http://dx.doi.org/10.1088/0957-4484/21/31/315106

11. Tsubota T, Tani S, Ida S, Nagata M, Matsumoto Y. Chemical modification of diamond surface with various carboxylic acids by radical reaction in liquid phase. Diam Relat Mater. 2004;13:1093-1097. DOI: 10.1016/j.diamond.2003.10.065

12. Gibson NM, Luo TJ, Brenner DW, Shenderova O. Immobilization of mycotoxins on modified nanodiamond substrates. Biointerphases 2011;6:210-217. DOI: http://dx.doi.org/10.1116/1.3672489

13. Ida S, Tsubota T, Tani S, Nagata MY, Matsumoto C. Chemical modification of the diamond surface using benzoyl peroxide and dicarboxylic acids. Langmuir 2003;19:9693-9698. DOI: 10.1021/la03433k

14. Krueger A, Liang Y, Jarre G, Stegk J, Schmiedt M. Surface functionalization of detonation diamond suitable for biomedical applications. J Mater Chem. 2006;16:2322-2328. DOI: 10.1039/B601325B

15. Krueger A, Ozawa M, Jarre G, Liang Y, Stegk J, Lu L. De-agglomeration and functionalisation of detonation diamond. Physica Status Solidi A 2010;207:959-971. DOI: 10.1007/s00763-010-9526-0

16. Yu SJ, Kang MW, Chang HC. Bright fluorescent nanodiamonds: no photobleaching and low cytotoxicity. J Am Chem Soc. 2005;127:17604-17605. DOI: 10.1021/ja056708i

17. Fu CC, Lee HY, Chen K. Characterization and application of single fluorescent nanodiamonds as cellular biomarkers. Proc Nat Acad Sci. 2007;104:727-732. DOI: 10.1073/pnas.0605490104

18. Wang HD, Yang QQ, Niu CH. Functionalization of nanodiamond particles with N,O-carboxymethyl chitosan. Diam Relat Mater. 2010;19:441-444. DOI: 10.1016/j.diamond.2010.01.032

19. Purtov KV, Petunin AI, Burov AE. Nanodiamonds as carriers for address delivery of biologically active substances. Nanoscal Res Lett. 2010;5:631-636. DOI: 10.1007/s11671-010-9526-0

20. Liu KK, Zheng WW, Wang CC. Covalent linkage of nanodiamond-paclitaxel for drug delivery and cancer therapy. Nanotechnology 2010;21:315106. DOI: http://dx.doi.org/10.1088/0957-4484/21/31/315106

21. Tsubota T, Tani S, Ida S, Nagata M, Matsumoto Y. Chemical modification of diamond surface with various carboxylic acids by radical reaction in liquid phase. Diam Relat Mater. 2004;13:1093-1097. DOI: 10.1016/j.diamond.2003.10.065

22. Gibson NM, Luo TJ, Brenner DW, Shenderova O. Immobilization of mycotoxins on modified nanodiamond substrates. Biointerphases 2011;6:210-217. DOI: http://dx.doi.org/10.1116/1.3672489

23. Ida S, Tsubota T, Tani S, Nagata MY, Matsumoto C. Chemical modification of the diamond surface using benzoyl peroxide and dicarboxylic acids. Langmuir 2003;19:9693-9698. DOI: 10.1021/la03433k

24. Krueger A, Liang Y, Jarre G, Stegk J. Surface functionalization of detonation diamond suitable for biomedical applications. J Mater Chem. 2006;16:2322-2328. DOI: 10.1039/B601325B

25. Krueger A, Ozawa M, Jarre G, Liang Y, Stegk J, Lu L. De-agglomeration and functionalisation of detonation diamond. Physica Status Solidi A 2010;207:959-971. DOI: 10.1007/s00763-010-9526-0

26. Krueger A, Boedeker T. De-agglomeration and functionalization of detonation nanodiamond with long alkyl chains. Diam Relat Mater. 2008;17:1367-1370. DOI: 10.1016/j.diam.2008.01.033

27. Krueger A, Stegk J, Liang Y, Jarre G. Biotinylated nanodiamond: simple and efficient functionalization of detonation diamond. J Mater Chem. 2006;16:2322-2328. DOI: 10.1039/B601325B

28. Kaur R, Chitanda JM, Michel D. Lysine-functionalized nanodiamonds: synthesis, physicochemical characterization, and nucleic acid binding studies. Int J Nanomed. 2012;7:3851-3866. DOI: http://dx.doi.org/10.2147/IJN.S2877

29. Huang L, Chang H. Adsorption and immobilization of cytochrome c on nanodiamonds. Langmuir. 2004;20:5879-5884. DOI: 10.1021/la045736

30. Ansari SA, et al. Functionalized nanodiamonds as an effective and novel matrix for immobilizing β-galactosidase. Food Bioprod Proc. 2015;95:298-303. DOI: 10.1016/j.fbp.2014.10.014

Iliran J Biotech. 2016;14(3):e1320
67. Hsu MH, Chuang H, Cheng FY, Huang YP, Han CC, Chen JCY. Directly thiolated modification onto the surface of detonation nanodiamonds. ACS Appl Mater Interf. 2014;6:7198-7203. DOI: 10.1021/am500324z

68. Burleson T, Yusuf N, Stanishevski A. Surface modification of nanodiamonds for biomedical application and analysis by infrared spectroscopy. J Achiev Mater Manuf Eng. 2009;37:258-263.

69. Zhu YW, Shen XQ, Wang BC, Xu XY, Feng ZJ. Chemical mechanical modification of nanodiamond in aqueous system. Int J Pharm. Sci. Inv. 2015;3:50-55.

70. Puzyr AP, Burov AE, Bondar VS. Modification and comparative study of commercial nanodiamonds. Full Nanotub Carbon Nanot. 2015;23:93-97. DOI: 10.1080/1536383X.2013.794338

71. Wang GD, Yang Q, Catherine HN. Functionalization of nanodiamond particles with N,O-carboxymethyl chitosan. Diam Relat Mat. 2010;19:441-444. DOI: 10.1016/j.diamond.2010.01.032

72. Vozyakovskii P, Ginzburg BM, Rashidov D. Structure, mechanical, and tribological characteristics of polyurethane modified with nanodiamonds. Polym Sci. 2010;52:1044-1050.

73. Vozyakovskii P, Kalinin AV, Agibalova LV. Modification of detonation synthesized nanodiamonds by a hydrocarbon radical as a method of producing highly dispersed water suspensions of diamond. J Sup Mat. 2011;33:244-249.

74. Ando T, Nishitani-Gamo M, Rawles RE, Yamamoto K, Kamo M, Sato Y. Chemical modification of diamond surfaces using a chlorinated surface as an intermediate state. Diam Relat Mater. 1996;5:1136-1142. DOI: 10.1016/0925-9636(96)00529-8

75. Huang H, Pierstorff E, Osawa E. Active nanodiamond hydrogels for chemotherapeutic delivery. Nanolett. 2007;7:3305-3314. DOI: 10.1021/nl071521o

76. Chow EK, Zhang XQ, Chen M. Nanodiamond therapeutic delivery agents mediate enhanced chemoresistant tumor treatment. Sci Transl Med. 2011;3:73ra21. DOI: 10.1126/scitranslmed.3001713

77. Ma XW, Zhao XL, Liang XJ. Nanodiamond delivery circumvents tumor resistance to doxorubicin. Acta Pharmacol Sin. 2011;32:543-544. DOI: 10.1038/aps.2011.58

78. Schrand A, Dai L, Schlager J. Differential biocompatibility of carbon nanotubes and nanodiamonds. Diam Relat Mater. 2007;16:2118-2123. DOI: 10.1016/j.diamond.2007.07.020

79. Liu KK, Wang CC, Cheng CL. Endocytic carboxylated nanodiamond for the labeling and tracking of cell division and differentiation in cancer and stem cells. Biomaterials 2009;30:4249-4259. DOI: 10.1016/j.biomaterials.2009.04.056

80. Chen M, Pierstorff ED, Lam R. Nanodiamond-mediated delivery of water-insoluble therapeutics. ACS Nano. 2009;3:2016-2022. DOI: 10.1021/nn900480m

81. Shimkunas RA, Robinson E, Lam R, Lu S, Xu X, Zhang XQ, Huang H, Osawa E, Ho D. Nanodiamond-insulin complexes as pH-dependent protein delivery vehicles. Biomaterials 2009;30:5720-5728. DOI: 10.1016/j.biomaterials.2009.07.004

82. Adnan A, Lam R, Chen HN. Atomic simulation and measurement of pH dependent cancer therapeutic interactions with nanodiamond carrier. Mol Pharm. 2011;8:368-374. DOI: 10.1021/mp1002398

83. Li X, Shao J, Qin Y, Shao C, Zheng T, Ye L. TAT-conjugated nanodiamond for the enhanced delivery of doxorubicin. J Mater Chem. 2011;8:7966-7973. DOI: 10.1039/C1JM10653H

84. Mohan N, Chen CS, Hsieh HH, Wu YC, Chang HC. In vivo imaging and toxicity assessments of fluorescent nanodiamonds in Caenorhabditis elegans. Nano Lett. 2010;10:3692-3699. DOI: 10.1021/nl101909

85. Kim HJ, Zhang K, Moore L, Ho D. Diamond nanogel-embedded contact lenses mediate lysozyme-dependent therapeutic release. ACS Nano. 2014;8:2998-3005. DOI: 10.1021/nn5002968

86. Li J, Zhu Y, Li W, Zhang Y, Peng Y, Huang Q. Nanodiamonds as intracellular transporters of chemotherapeutic drug. Biomaterials 2010;31:8410-8418. DOI: 10.1016/j.biomaterials.2010.07.058

87. Guan B, Zou F, Zhi FH. Nanodiamond as the pH-responsive vehicle for an anticancer drug. Small 2010;6:1514-1519. DOI: 10.1002/sml.200902305

88. Puzyr AP, Baron AV, Purtof KV. Nanodiamonds with novel properties: a biological study. Diam Relat Mater. 2007;16:2124-2128. DOI: 10.1016/j.diamond.2007.07.025

89. Xing Y, Xiong W, Zhu L, Osawa E, Hussin S, Dai L. DNA damage in embryonic stem cells caused by nanodiamonds. ACS Nano. 2011;5:2376-2384. DOI: 10.1021/nn200279k

90. Bakowicz K, Mitura S. Biocompatibility of NCD. J Wid Band Mat. 2009;2:269-272. DOI: 10.1106/152451102024429

91. Huang TS, Tzeng Y, Liu YK, Chen YC, Walker KR, Guntupalli R, Liu C. Immobilization of antibodies and bacterial binding on nanodiamond and carbon nanotubes for biosensor applications. Diam Relat Mater. 2004;13:1098-1102. DOI: 10.1016/j.diamond.2003.11.047

92. Chao J, Perevedentseva E, Chung PH. Nanometer-sized diamond particle as a probe for biolabeling. Biophys J. 2007;93:2199-2208. DOI: 10.1529/biophysj.107.108134

93. Nguyen TTB, Chang HC, Wu VWK. Adsorption and hydrolytic activity of lysozyme on diamond nanocrystallites. Diam Relat Mater. 2007;16:872-876. DOI: 10.1016/j.diamond.2007.01.030

94. Perevedentseva E, Cai PJ, Chiu YC, Cheng CL. Characterizing protein activities on the lysozyme and nanodiamond complex prepared for bio applications. Langmuir 2011;27:1085-1091. DOI: 10.1021/la103155c

95. Chang IP, Hwang KC, Chiang CS. Preparation of fluorescent magnetic nanodiamonds and cellular imaging. J Am Chem Soc. 2008;130:15476-15481. DOI: 10.1021/ja804253y

96. Zhang XQ, Chen M, Lam R, Xu X, Osawa E, Ho D. Polymer-functionalized nanodiamond platforms as vehicles for gene delivery. ACS Nano. 2009;3:2609-2616. DOI: 10.1021/nn900865g

97. Gomez H, Ram MK, Alvi F, Stefanakos E, Kumar A. Graphene-conducting polymer nanocomposite as novel electrode for supercapacitors. J Phys Chem C. 2010;114:18797-18804. DOI: 10.1021/jp106379e

98. Kovalenko I, Bucknall DG, Yushin G. Detonation nanodiamond and onion-like-carbon-embedded polyaniline for supercapacitors. Adv Funct Mater. 2010;20:3979-3986. DOI: 10.1002/adfm.201000906

99. Zhang Q, Mochalin VN, Neitzel I, Knoke IY, Han J, Klug CA, Zhou GJ, Lelkes PI, Gogotsi Y. Fluorescent PLLA-nanodiamond composites for bone tissue engineering. Biomaterials 2011;32:87-94. DOI: 10.1016/j.biomaterials.2010.08.090

100. Nakamura T, Ohana T, Hagiwara Y, Tsubota T. Kinetics of nuclear microplasma atmospheric ion generation correlated with discharge current. Phys Chem. 2009;11:730-734.

101. Schietinger S, Barth M, Aichele T, Benson O. Plasmom
enhanced single photon emission from a nano assembled metal diamond hybrid structure at room temperature. *Nanolett*. 2009;9:1694-1698. DOI: 10.1021/nl900384c

102. Shenderova O, Koscheev A, Zaripov N, Petrov I, Skryabin Y, Detkov P, Turner S, Van Tendeloo G. Surface chemistry and properties of ozone purified denotation nanodiamonds. *J Phys Chem*. 2011;115:9827-9837. DOI: 10.1021/jp1102466

103. Zhong YL, Loh KP. The chemistry of -CH bond activation on diamond. *Chem Asian J*. 2010;5:1532-1540. DOI: 10.1002/asia.201000027

104. Jia NQ, Lian Q, Shen HB. Intracellular delivery of quantum dots tagged antisense oligodeoxynucleotides by functionalized multiwalled carbon nanotubes. *Nano Lett*. 2007;7:2976-2980. DOI: 10.1021/nl071114c

105. Zhang XK, Meng LJ, Lu XH. Targeted delivery and controlled release of doxorubicin to cancer cells using modified single wall carbon nanotubes. *Biomaterials* 2009;30:6041-6047. DOI: 10.1016/j.biomaterials.2009.07.025

106. Huang P, Xu C, Lin J. Folic acid-conjugated graphene oxide loaded with photosensitizers for targeting photodynamic therapy. *Theranostics*. 2011;1:240-250. DOI: 10.7150/thno/v01p0240

107. Zhang LM, Lu ZX, Zhao QH. Enhanced chemotherapeutic efficacy by sequential delivery of siRNA and anticancer drugs using PEI-grafted graphene oxide. *Small* 2011;7:460-464. DOI: 10.1002/smll.201001522

108. Manna SK, Sarkar S, Barr J. Single-walled carbon-nanotube induces oxidative stress and activates nuclear transcription factor-kB in human keratinocytes. *Nano Lett*. 2005;5:1676-1684. DOI: 10.1021/nl0507966

109. Zhang YB, Ali SF, Dervishi E. Cytotoxicity effects of graphene and single wall carbon nanotubes in neural phaeochromocytoma-derived PC12 cells. *ACS Nano*. 2010;4:3181-3186. DOI: 10.1021/nn1007176

110. Wang JY, Sun PP, Bao YM. Cytotoxicity of single-walled carbon nanotubes on PC12 cells. *Toxicol In Vitro*. 2011;25:242-250. DOI: 10.1016/j.tiv.2010.11.010

111. Magrez A, Kasas S, Salicio V. Cellular toxicity of carbon-based nanomaterials. *Nano Lett*. 2006;6:1121-1125. DOI: 10.1021/nl060162e

112. Hartl A, Schmich E, Garrido JA. Protein-modified nano-crystalline diamond thin films for biosensor applications. *Nat Mater*. 2004;3:736-742. doi:10.1038/nmat1204

113. Liu Y, Khabashesku VN, Halas NJ. Fluorinated nanodiamond as a wet chemistry precursor for diamond coatings covalently bonded to glass surface. *J Am Chem Soc*. 2015;127:3712-3713. DOI: 10.1021/ja042389m

114. Huang HJ, Pierstorff E, Osawa E. Protein-mediated assembly of nanodiamond hydrogels into a biocompatible and biofunctional multilayer nanofilm. *ACS Nano*. 2008;2:203-212. DOI: 10.1021/nn700867

115. Huang HJ, Dai LM, Wang DH. Large-scale self-assembly of dispersed nanodiamonds. *J Mater Chem*. 2008;18:1347-1352. DOI: 10.1039/B716676A

116. Lam R, Chen M, Pierstorff E. Nanodiamond-embedded microfilm devices for localized chemotherapeutic elution. *ACS Nano*. 2008;2:2095-2102. DOI: 10.1021/nn800465x

117. Xi G, Robinson E, Mania-Farnell B, Vanin EF, Shim KW. Convection-enhanced delivery of nanodiamond drug delivery platforms for intracranial tumor treatment. *Nanomedicine*. 2014;10:381-391. DOI: 10.1016/j.nano.2013.07.013

118. Liu K, Cheng C, Chang C. Biocompatible and detectable carboxylated nanodiamond on human cell. *Nanotechnology*. 2007;18:325102. DOI: http://dx.doi.org/10.1088/0957-4484/18/32/325102

119. Blaber SP, Hill CJ, Webster RA, Say JM, Brown LJ. Effect of labeling with iron oxide particles or nanodiamonds on the functionality of adipose-derived mesenchymal stem cells. *Plos One*. 2013;8:e52997. DOI: 10.1371/journal.pone.0052997

120. Wu TJ, Tseng YK, Chang WW, Cheng CA, Kuo Y, Chien CH, Chang HC, Yu J. Tracking the engraftment and regenerative capabilities of transplanted lung stem cells using fluorescent nanodiamonds. *Nat Nanotechnol*. 2013;8:682-689. DOI: 10.1038/nnano.2013.147

121. Wang X, Low XC, Hou W, Abdullah LN, Toh TB, Rashid MMA, Ho D, Chow EKH. Epirubicin-adsorbed nanodiamonds kill chemoresistant hepatic cancer stem cells. *ACS Nano*. 2014;8:12151-12166. DOI: 10.1021/nn503491e