Clonal relationship of synchronous head and neck cancer and esophageal cancer assessed by single nucleotide polymorphism-based loss of heterozygosity analysis

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Abstract

Background: The prognoses of head and neck squamous cell carcinoma (HNSCC) and esophageal squamous cell carcinoma (ESCC) are poor, especially when both tumors occur at the same time. We examined the clonal relatedness of HNSCCs with synchronous ESCCs to confirm whether the second tumors were metastasis or separate second primary malignancies (SPMs) using loss of heterozygosity (LOH) analysis.

Methods: Twenty-one pairs of formalin-fixed paraffin-embedded tissue from HNSCC patients with synchronous esophageal cancer were analyzed by single nucleotide polymorphism (SNP) array using the Illumina HumanCytoSNP FFPE-12 BeadChip (San Diego, CA), which contains approximately 300,000 probes. LOH was identified using Nexus Copy Number software (El Segundo, CA).

Results: Comparing the LOH pattern between HNSCC and paired ESCC, we found that 20 out of 21 paired tissues had a high number of discordant LOHs (LOH identified solely in the primary HNSCC but not in synchronous ESCC at the same genomic location) and a low number of concordant LOHs (LOH at the same genomic location in both HNSCC and ESCC). Only one case fell into the undetermined category. Therefore, these 20 ESCCs were classified as SPMs or second field tumors (SFTs). Moreover, the HNSCC patients with molecularly confirmed esophageal SPM had significantly poorer survival than the other patients.

Conclusions: We propose the use of a genome-wide SNP array as a tool to differentiate metastatic tumors from SPM/SFT. The SNP array offers genome-wide LOH information that earlier microsatellite analysis studies lack. The ability to accurately identify SPM should contribute to a better treatment plan and follow-up care of these patients.

Keywords: Head and neck squamous cell carcinoma, Esophageal carcinoma, Synchronous, Second primary malignancy, SNP array, Loss of heterozygosity, Formalin-fixed-paraffin-embedded tissues

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Background

Patients with head and neck squamous cell carcinoma (HNSCC) have an increased risk of developing second primary malignancies (SPMs) [1]. SPMs can be diagnosed as either synchronous (diagnosis within 6 months after an index tumor) or metachronous (diagnosis more than 6 months after the index tumor) [2]. The common locations of SPMs in head and neck cancer patients are the esophagus, lung or head and neck area [1]. Esophageal cancer is a serious malignancy due to its aggressive behavior. Although esophageal cancer is relatively uncommon in the United States, the incidence is higher in Asia [3]. The worldwide incidence of HNSCC combined with synchronous esophageal cancer varies from 2.7 to 12.5% [4–6]. In Thailand, the incidence of synchronous esophageal squamous cell carcinoma (ESCC) in HNSCC patients between 2009 and 2011 was 12.4% [7].

The molecular mechanism of SPM is still not fully understood. Field cancerization, proposed in 1953 [8], is one of the concepts that explains the development of SPMs. This concept suggests that the mucosa of the upper aerodigestive tract are repeatedly exposed to carcinogens, such as smoking and alcohol, leading to multiple areas of genetic aberrations called “fields”. These accumulated genetic alterations could eventually develop to multiple progressive cancers in the same or independent fields [9].

Currently, SPM is commonly diagnosed based on the clinical criteria proposed by Warren and Gates in 1932 [10]. It is crucial to exclude the metastasis before diagnosis of SPM due to the differences in prognoses and outcomes of disease, which lead to suitable plans of treatment [11]. In clinical practice, metastasis is diagnosed by clinicopathological examination. However, distinguishing a metastatic tumor from an independent primary cancer can be unclear in some situations [12]. Thus, molecular approaches are required to accurately make these distinctions. By comparing molecular patterns between the index tumor and the second tumor, Braakhuis et al. proposed the molecular classification of SPMs after index HNSCC as follows: 1) if the second tumor shares the same molecular alterations with the index tumor, it is defined as recurrence or metastasis; 2) if the index tumor and the second tumor share only some genetic alterations, the second tumor is classified as a second field tumor (SFT); and 3) if the genetic profiles of two tumors are unrelated, the second tumor is considered a SPM [2, 9]. The molecular patterns can also be used to evaluate the clonal relatedness of multiple tumors in the same patient, whether the tumors arise from a common clone or independent clonal origins [13, 14].

Previous studies used multiple microsatellite markers to screen for loss of heterozygosity (LOH) regions to assess the clonal relationship for differentiating between the SPM and metastasis in several cancers [15–18].

Advanced molecular techniques have been used in the studies of clonality, for example, single nucleotide polymorphism (SNP) array and next-generation sequencing [12, 19]. Although next-generation sequencing, in particular whole-genome sequencing, is increasingly used in the molecular study of clonal evolution [20], this technique is still not feasible in many labs due to the high cost and complexity of bioinformatic analysis. SNP array has been frequently used as a clinical diagnostic tool in hematological cancers [21], and its clinical use in solid tumors has gradually become more common [22]. SNP array is still practical and useful for genome-wide analysis of LOH at a high resolution [23].

In this study, we performed LOH analysis using a genome-wide SNP array in patients who developed both primary HNSCC and synchronous ESCC. To evaluate the clonal relatedness of two tumors for differentiating between SPM and metastasis, we compared the LOH patterns between primary HNSCC and synchronous esophageal cancer. Moreover, the survival rate of patients using their LOH ratio was analyzed.

Methods

Patients

The medical records of Songklanagarind Hospital, Prince of Songkla University, were searched for all patients diagnosed with synchronous head and neck cancer and esophageal cancer between January 2002 and December 2012. Inclusion criteria were as follows: 1) the head and neck cancers and esophageal cancers had to be squamous cell carcinoma; 2) both tumors had to be anatomically separated by normal mucosa based on clinical and pathological findings; the esophageal cancer had to be located at middle or lower esophagus to ensure a distance from the head and neck cancer of at least 10 cm; and 3) both tumors had formalin-fixed paraffin-embedded (FFPE) tissues available for subsequent analysis. Patient recruitment and sample collection were performed with protocols approved by the Institutional Review Board.

SNP array analysis

FFPE tissues were sectioned. Archived hematoxylin and eosin-stained tissue slides were evaluated by a pathologist for the area with at least 70% tumor cells for manual macrodissection using a needle tip or scalpel. A minimum amount of 200 ng of DNA extracted from each FFPE tissue was quantified by the Qubit® 2.0 Fluorometer (Thermo-Fisher Scientific, Waltham, MA) and qualified using the Infinium FFPE QC Kit before being processed with the Infinium HD FFPE DNA Restoration Kit (Illumina, San Diego, CA), all according to the manufacturers’ protocols.

SNP array was performed with the HumanCytoSNP FFPE-12 v2.1 DNA Analysis BeadChip (Illumina, San Diego, CA), according to the manufacturer’s instructions.
This array contains approximately 299,140 SNP markers spanning the entire genome with an average probe spacing of 72 kb. The data were analyzed with GenomeStudio Data Analysis Software v. 2011.1 (Illumina, San Diego, CA) and Nexus Copy Number v9.0 (BioDiscovery, Inc., El Segundo, CA) using the reference human genome (hg19/GRCh37).

Analysis of LOH pattern
LOH results were obtained from the SNP array using Nexus Copy Number software’s default settings (SNP-FASST2 Segmentation Algorithm, a minimum of 3 probes per segment and a maximum contiguous probe spacing of 1000 kb). The minimum LOH length was set at 500 kb. Using the GenomicRanges packages in R, two LOHs at the same genomic region of each pair of HNSCC and ESCC were compared and defined as identical when more than 60% of the total length of both LOHs overlapped. Concordant LOH was then defined as the presence of identical LOHs at the same genomic region of both tumors. In contrast, when a unique LOH was identified only in HNSCC or ESCC, this case was defined as discordant LOH. The percentage of concordant (% concordant) or discordant (% discordant) LOH was calculated as the number of concordant or discordant LOH divided by the total number of LOH in HNSCC. The ratio between % discordant and % concordant LOH was subsequently calculated. Of note, the % concordant was initially added with 1 to avoid division by zero, and to create the 0–100 score ranges.

Based on the ratio results, the second tumors were classified according to Braakhuis et al.’s proposed models [2] into four subgroups: tumors with a ratio less than 0.5 were likely to be metastatic; tumors with a ratio between 0.5 and 2 were considered undetermined; tumors with a ratio more than 2 and less than 100 could be either SFT or SPM; tumors with a ratio of 100 were considered SPM (Fig. 1).

Statistical analysis
Statistical analyses were performed using PASW Statistics version 18.0 for Windows (SPSS Inc.) and GraphPad Prism version 6.0 for Windows (GraphPad Software). Survival analysis was performed and visualized using Kaplan-Meier curves. P value was generated by log-rank test.

Results
Patient characteristics
A total of 21 patients diagnosed with HNSCC and synchronous ESCC were included in this study. The major risk factors (i.e., smoking and alcohol) of either HNSCC or ESCC were analyzed. Patient characteristics are summarized in Table 1. The majority of patients (66.7%) were male who were active or former consumers of smoking and alcohol. Nearly half (47.6%) of the HNSCCs were located in hypopharynx. All ESCCs (100%) were located in middle and lower esophagus. The majority of HNSCCs (71.5%) were advanced stage (stage III and IV),

Fig. 1 Diagram showing the classification criteria for LOH result interpretation. SNP, single nucleotide polymorphism; LOH, loss of heterozygosity; HNSCC, head and neck squamous cell carcinoma; SFT, second field tumor; SPM, second primary malignancy
whereas the majority of ESCCs (76.2%) were early stage (stage I and II). The difference in tumor stages implies a chronology of tumor development in which HNSCC might initially develop before ESCC, although both tumors were clinically detected at the same time.

LOH analysis of paired HNSCCs and ESCCs
To determine whether the tumors were two independent primary tumors or metastases, the similarity in genetic profiles were analyzed. Using SNP array, the profiles of LOHs of HNSCC and synchronous ESCC of the same patient were compared. The results represented are depicted in Fig. 2. From the LOH diagram, the majority of LOH patterns of each pair of tumors were distinguishable.

To rule out a LOH pattern that may represent tumor progression, we defined discordant LOH as a LOH identified solely in the primary HNSCC but not in the synchronous ESCC at the same genomic location. The percentages of concordant and discordant LOH were analyzed and the ratios of discordant to concordant LOH were calculated as described in Materials and Methods (Table 2). Twenty cases (95%) had a ratio of more than 2, which classified these cases as SPM or SFT. No patient was classified as metastasis (ratio < 0.5). Only one case (case 2) showed an undetermined result (ratio = 0.5–2). Moreover, 20% of the HNSCC patients with synchronous middle esophageal tumor (3 out of 15) and 50% of the patients with synchronous lower esophageal tumor (3 out of 6) showed the maximum ratio of 100, which was interpreted as molecularly confirmed SPMs.

We further analyzed our LOH data using the 25 microsatellite markers and classification method proposed by Geurts et al. [16]. Our method could determine the molecular diagnosis in cases that were not confidently defined by Geurts’ method. There was only one case with undetermined results from both methods (Table 2).

Survival analysis
To compare the survival between HNSCC patients with molecularly confirmed esophageal SPM (ratio = 100) and the other patients (ratio < 100), Kaplan–Meier analyses were performed using time from diagnosis of the first primary HNSCC to death. The results showed that the HNSCC patients with molecularly confirmed esophageal SPM had significantly poorer survival than the other patients (log-rank test \( P = 0.0466 \)) (Fig. 3). Furthermore, after excluding the case with molecularly undetermined clonality (ratio = 0.5–2), the Kaplan–Meier plot also showed the same pattern but without statistical significance (log-rank test \( P = 0.0738 \)) (Additional file 1: Figure S1).

Discussion
SPM is a leading long-term cause of mortality in patients with HNSCC and is associated with poor prognosis [24, 25]. Field cancerization is one of the major concepts that explains the mechanism of SPM development, in which multifocal tumors could originate from the same or independently genetically altered fields [2, 8]. The index

| Table 1 Patient characteristics |
|---------------------------------|
| No. (%)                         |
| Number                          | 21 |
| Age                             |    |
| mean (years)                    | 58.7 |
| < 65                            | 15 (71.4) |
| ≥ 65                            | 6 (28.6) |
| Gender                          |    |
| Male                            | 20 (95.2) |
| Female                          | 1 (4.8) |
| Smoking                         |    |
| Never                           | 1 (4.8) |
| Former\(^a\)                    | 5 (23.8) |
| Active                          | 9 (42.9) |
| Unknown                         | 6 (28.6) |
| Alcohol consumption             |    |
| Never                           | 0 (0) |
| Former\(^a\)                    | 3 (14.3) |
| Active                          | 12 (57.1) |
| Unknown                         | 6 (28.6) |
| Site of HNSCC                   |    |
| Oropharynx                      | 4 (19.0) |
| Hypopharynx                     | 10 (47.6) |
| Larynx                          | 7 (33.3) |
| Staging of HNSCC                |    |
| I                               | 4 (19.0) |
| II                              | 2 (9.5) |
| III                             | 6 (28.6) |
| IV                              | 9 (42.9) |
| Site of ESCC                    |    |
| upper                           | 0 (0) |
| middle                          | 15 (71.4) |
| lower                           | 6 (28.6) |
| Staging of ESCC                 |    |
| I                               | 1 (4.8) |
| II                              | 15 (71.4) |
| III                             | 5 (23.8) |
| IV                              | 0 (0) |

HNSCC, head and neck squamous cell carcinoma; ESCC, esophageal squamous cell carcinoma; \(^a\)Quit smoking or alcohol consumption at least 1 year before this study began; \(^b\)American Joint Committee on Cancer (AJCC) stage (7th ed)

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tumor and metastasis are developed from the same clone. Within the field of premalignant cells, two tumors separately develop, leading to SFTs. In a case in which two tumors develop from independent fields, the second tumor is defined as SPM [2]. Therefore, the index tumor and metastasis share a strong clonal relationship. While the first tumor and SFT share some similarities in clonal patterns, the first tumor and SPM should have very few or no clonal relatedness [14].

LOH is a common genetic event in cancer development, and LOH analysis is one of the molecular techniques for studying the clonal relationship between two tumors. Many studies performed microsatellite analysis to detect LOH [15–18]. However, PCR-based LOH analysis has a limitation in these techniques use a small number of markers. SNP array is an established technology commonly used in clinical molecular cytogenetic diagnostic labs around the world for both cancers and genetic disorders [21, 23]. SNP array offers genome-wide analysis of LOH at a higher resolution, and therefore we hypothesized that it can also be used for SPM identification with a higher accuracy. Therefore, we conducted analyses using a SNP array with approximately 300,000 probes. Since there is no consensus method for analyzing the LOH profile for determining clonality, we developed classification criteria for interpretation of the LOH results using a ratio of the percentage of discordant and concordant LOH (Fig. 1). A high ratio indicates a high discordant profile, which implies a low clonal relatedness between two tumors.

In clinical practice, it is difficult to differentiate between local recurrence and SFT and also between SFT and SPM [9]. In this study, all of the selected patients were clinically diagnosed as HNSCC with synchronous esophageal SPM. The LOH analysis showed that 20 cases (95%) had the ratio of more than 2, which classified these cases as SPM or SFT. Therefore, molecular diagnosis using LOH analysis was in line with our strict clinical diagnosis, and thus confirmed the validity of using SNP array as a diagnostic tool. Moreover, our method using genome-wide SNP markers could produce more informative results than the method that used fewer numbers of markers [16]. Of note, five cases showed unclear results by Geurts’ method [16]. Using our method, these five cases could be defined as SPM/SFT. Thus, this emphasizes the benefit of using high-resolution technology such as SNP array for LOH analysis.

In a subgroup of patients, the LOH ratio was 100 (i.e. the discordant LOH was 100% and no concordant LOH was detected.) We hypothesize that the two tumors were independently developed from separate fields and had no clonal relatedness. Thus, this group of patients suffered from HNSCC with molecularly confirmed esophageal SPMs.

The synchronous esophageal cancers defined as SFT/SPM (ratio > 2 and < 100) were found in the middle and lower esophagus. In general, the distances between the head and neck area to the middle and lower esophagus regions are approximately 10 and 15 cm, respectively [26]. Our results suggest that the diameter of genetically altered field in these patients could be at least 10 cm. These findings support a previous study showing that fields with genetically altered cells can be as large as up to 7 cm in diameter [14].

A previous study showed that the survival of HNSCC patients with clinically diagnosed esophageal SPM was significantly reduced compared with HNSCC patients without SPM [27]. In this study, the survival analysis showed that HNSCC patients with molecularly confirmed esophageal SPM (ratio = 100) had significantly poorer survival than the other patients (ratio < 100) (log-
| No. | Sex | Age-ranges | HNSCC Site | ESCC Site | Stagea | Stagea | This study | %Discordant LOH | %Concordant LOH | Ratio | Conclusion | Geurts et al., 2005 [16] | Discordant LOHb | Concordant LOH | All discordant LOH | Conclusion |
|-----|-----|------------|------------|-----------|--------|--------|------------|----------------|----------------|--------|------------|-----------------------|----------------|----------------|-------------------|------------|
| 1   | M   | 56–60      | oropharynx | T3        | N0     | M0     | middle     | T2             | N0             | M0     | 95.45      | 4.55                  | 17.2           | 0              | 1                 | 5          | SPM        |
| 2   | M   | 61–65      | oropharynx | T1        | N0     | M0     | middle     | T2             | N1             | M0     | 56.25      | 43.75                 | 1.3            | Undetermined | 0                 | 0          | 0          |
| 3   | M   | 61–65      | oropharynx | T2        | N1     | M0     | middle     | T2             | N1             | M0     | 100        | 0                     | 100            | SPM           | 2                 | 0          | 2          | SPM        |
| 4   | M   | 36–40      | larynx     | T3        | N3     | M0     | middle     | T1             | N0             | M0     | 95.45      | 4.55                  | 17.2           | SFT/SPM       | 5                 | 0          | 5          | SPM        |
| 5   | M   | 51–55      | larynx     | T1        | N0     | M0     | middle     | T2             | N1             | M0     | 92.5       | 7.5                   | 10.9           | SFT/SPM       | 4                 | 2          | 12         | SPM        |
| 6   | M   | 61–65      | larynx     | T1        | N0     | M0     | middle     | T2             | N0             | M0     | 83.33      | 16.67                 | 4.7            | SFT/SPM       | 1                 | 2          | 4          | Metas?      |
| 7   | M   | 61–65      | larynx     | T3        | N2     | M0     | middle     | T2             | N1             | M0     | 100        | 0                     | 100            | SPM           | 3                 | 0          | 10         | SPM        |
| 8   | M   | 76–80      | larynx     | T3        | N0     | M0     | middle     | T2             | N0             | M0     | 93.75      | 6.25                  | 12.9           | SFT/SPM       | 4                 | 1          | 7          | SPM        |
| 9   | M   | 41–45      | hypopharynx| T3        | N2     | M0     | middle     | T3             | N1             | M0     | 75         | 25                    | 2.9            | SFT/SPM       | 3                 | 2          | 6          | SPM        |
| 10  | M   | 41–45      | hypopharynx| T4        | N2     | M0     | middle     | T3             | N1             | M0     | 92.59      | 7.41                  | 11             | SFT/SPM       | 8                 | 0          | 12         | SPM        |
| 11  | M   | 51–55      | hypopharynx| T2        | N2     | M0     | middle     | T2             | N1             | M0     | 100        | 0                     | 100            | SPM           | 0                 | 0          | 0          | ?          |
| 12  | M   | 56–60      | hypopharynx| T2        | N0     | M0     | middle     | T2             | N0             | M0     | 87.1       | 12.9                  | 6.3            | SFT/SPM       | 8                 | 0          | 10         | SPM        |
| 13  | M   | 61–65      | hypopharynx| T3        | N0     | M0     | middle     | T3             | N2             | M0     | 81.82      | 18.18                 | 4.3            | SFT/SPM       | 4                 | 1          | 12         | SPM        |
| 14  | F   | 66–70      | hypopharynx| T3        | N2     | M0     | middle     | T2             | N0             | M0     | 98.25      | 1.75                  | 35.7           | SFT/SPM       | 5                 | 1          | 9          | SPM        |
| 15  | M   | 81–85      | hypopharynx| T1        | N3     | M0     | middle     | T2             | N1             | M0     | 93.75      | 6.25                  | 12.9           | SFT/SPM       | 0                 | 0          | 1          | ?          |
| 16  | M   | 71–75      | oropharynx | T3        | N1     | M0     | lower      | T2             | N0             | M0     | 94.87      | 5.13                  | 15.5           | SFT/SPM       | 4                 | 0          | 4          | SPM        |
| 17  | M   | 46–50      | larynx     | T1        | N0     | M0     | lower      | T2             | N1             | M0     | 100        | 0                     | 100            | SPM           | 7                 | 0          | 8          | SPM        |
| No. | Sex | Age-ranges | HNSCC Site | Stage | ESCC Site | Stage | This study | Geurts et al., 2005 [16] | Conclusion |
|-----|-----|------------|------------|-------|-----------|-------|------------|------------------------|------------|
|     |     |            |            |       |           |       |            | Discordant LOH | Concordant LOH | All discordant LOH |
| 18  | M   | 71–75      | larynx     | T2 N1 M0 | lower     | T2 N0 M0 | 87.5       | 12.5       | 6.5 SFT/SPM       | 9           | 2           | 11 SPM       |
| 19  | M   | 41–45      | hypopharynx| T4 N2 M0 | lower     | T3 N2 M0 | 100        | 0          | 100 SPM           | 0           | 0           | 2 SPM?       |
| 20  | M   | 46–50      | hypopharynx| T3 N2 M0 | lower     | T2 N0 M0 | 95         | 5          | 15.8 SFT/SPM      | 4           | 0           | 7 SPM        |
| 21  | M   | 51–55      | hypopharynx| T2 N0 M0 | lower     | T2 N2 M0 | 100        | 0          | 100 SPM           | 1           | 0           | 3 SPM?       |

LOH, loss of heterozygosity; HNSCC, head and neck squamous cell carcinoma; ESCC, esophageal squamous cell carcinoma; SFT, second field tumor; SPM, second primary malignancy; ?, inconclusive; SPM?, probably second primary malignancy; Metas?, probably metastasis; M, male; F, female; *American Joint Committee on Cancer (AJCC) stage (7th ed); †Discordant LOH not explained by tumor progression.
rank test $P = 0.0466$). This result suggested that tumor clonality may contribute to the survival of cancer patients. A recent review on the evolutionary process on field cancerization by Curtius et al. [28] showed that clonal diversity is involved in the prognoses of many cancers. Thus, primary tumors originating from multiple different fields might be associated with poor prognosis.

To the best of our knowledge, this study is the first SNP array-based genome-wide LOH analysis and clinical correlations to differentiate the cause of HNSCC with synchronous ESCC. Our findings may indicate another important benefit to the clinical application of a SNP array test in solid tumors, which is to help guide clinical decision with regard to patients with synchronous tumors. Further study in larger HNSCC patient cohorts who have clinically-defined metastasis, local recurrent or second tumor with closer anatomical location to the first primary tumor may help clarify the usefulness of SNP array and LOH scoring system for the diagnosis of SPM in clinical practice.

**Conclusions**

There is a 95% concordance between SNP array-based LOH analysis and our strict clinical diagnostic criteria for esophageal SPM, which confirmed the validity of using SNP array as a diagnostic tool. The SNP array offers genome-wide LOH information that earlier microsatellite analysis studies lack. The data generated by SNP array is also more compact and less computational intensive to analyze compared with whole exome or whole genome sequencing data, thus making it an ideal choice for clinical laboratories. The ability to accurately distinguish SPM from metastatic tumor should contribute to a better treatment plan and follow-up care of HNSCC and esophageal cancer patients.

Supplementary information

**Supplementary information** accompanies this paper at https://doi.org/10.1186/s12885-019-6394-6.

**Additional file 1: Figure S1.** Kaplan-Meier curves showing the survival time (in days) between the HNSCC patients with molecularly confirmed esophageal SPM (ratio = 100) and the patients defined as SFT/SPM (ratio > 2), but < 100, excluding a case with undetermined result.

**Abbreviations**

ESCC: Esophageal squamous cell carcinoma; FFPE: Formalin-fixed-paraffin-embedded; HNSCC: Head and neck squamous cell carcinoma; LOH: Loss of heterozygosity; SFTs: Second field tumors; SNP: Single nucleotide polymorphism; SPMs: Second primary malignancies

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**Disclosures**

No potential conflicts of interest were disclosed.

**Authors’ contributions**

SS provided the conception, clinical specimen and clinical data, funding for this study, and wrote the manuscript. SB helped develop the methodology, perform statistical analysis, and write the manuscript. TP performed SNP array experiments. CN did the SNP array data analysis. PS, AN, and TD provided the clinical specimens and reviewed clinical data. NJ supervised the whole study in addition to designing the experiment, writing the manuscript and providing funding. All authors read and approved the final manuscript.

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**Availability of data and materials**

The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

**Ethics approval and consent to participate**

Ethics protocol was approved by Human Research Ethics Committees of the Faculty of Medicine, Prince of Songkla University (EC. SS-2017-101-1-3) and the Faculty of Medicine Ramathibodi Hospital, Mahidol University (ID 09-55-18; MURA2012/383). Since this is a retrospective study using leftover clinical specimens, consent to participate was waived by both committees.

**Consent for publication**

Not Applicable.

**Competing interests**

The authors declare that they have no competing interests.

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1. Vaamonde P, Martin C, Labella T. Second primary malignancies in patients with cancer of the head and neck. Otolaryngol Head Neck Surg. 2003;129(1):65–70.

2. Biahauski BJ, Tabor MP, Leemans CR, van der Waal I, Snow GB, Brahekonoff RH. Second primary tumors and field cancerization in oral and oropharyngeal cancer: molecular techniques provide new insights and definitions. Head Neck. 2002;24(2):198–206.

3. Zhang Y. Epidemiology of esophageal cancer. World J Gastroenterol. 2013;19(34):6508–600.

4. Scherübl H, Von Lampe B, Faiss S, Däubler P, Bohlmann P, Plath T, et al. Screening for oesophageal neoplasia in patients with head and neck cancer. Br J Cancer. 2002;86(2):239–43.

5. Fukushima K, Noguchi Y, Yoshikawa T, Saito A, Doi C, Makino T, et al. High incidence of synchronous cancer of the oral cavity and the upper gastrointestinal tract. Cancer Lett. 1999;144(2):145–51.

6. Kim DH, Gong EJ, Jung H-Y, Lim H, Ahn JY, Choi KS, et al. Clinical significance of intensive endoscopic screening for synchronous esophageal neoplasm in patients with head and neck squamous cell carcinoma. Scand J Gastroenterol. 2014;49(12):1486–92.

7. Laohawiriyakamol S, Sunpaweravong S, Leelamanit V, Pruegsanusak K, Sinkijcharoenchai W. Evaluating synchronous esophageal cancer in head and neck cancer patients using Lugol dye chromoendoscopy. J Med Assoc Thail. 2014;97(11):1164–70.

8. Slaughter DP, Southwick HW, Smejkal W. “Field cancerization” in oral stratified squamous epithelium. Cancer. 1953;6:963–8.

9. Biahauski BJ, Brahekonoff RH, Leemans CR. Second field tumors: a new opportunity for cancer prevention? Oncologist. 2005;10(7):493–500.

10. Warren S, Gates O. Multiple primary malignant tumors: a survey of the literature and a statistical study. Am J Cancer. 1932;16:1358–414.

11. Park JW, Lee S-w. Clinical outcomes of synchronous head and neck and esophageal cancer. Radiat Oncol J. 2015;33(3):172.

12. Begg CB, Ostrovnyaia I, Geyer FC, Papanastasiou AD, Ng CK, Sakr RA, et al. Contralateral breast cancers: independent cancers or metastases? Int J Cancer. 2018;143(2):347–56.

13. Mauguen A, Seshan VE, Ostrovnyaia I, Begg CB. Estimating the probability of clonal relatedness of pairs of tumors in cancer patients. Biometrics. 2018; 74(1):321–30.

14. Tabor MP, Brahekonoff RH, Ruitjer-Schipper HJ, van der Wal JE, Snow GB, Leemans CR, et al. Multiple head and neck tumors frequently originate from a single preneoplastic lesion. Am J Pathol. 2002;161(3):1051–60.

15. Califano J, Leong PL, Koch WM, Eisenberger CF, Sidransky D, Westra WH. Second esophageal tumors in patients with head and neck squamous cell carcinoma: an assessment of clonal relationships. Clin Cancer Res. 1999;5(7):1862–7.

16. Geurts TW, Nederlof PM, van den Brekel MW, van’t Veer LJ, de Jong D, Hart AA, et al. Pulmonary squamous cell carcinoma following head and neck squamous cell carcinoma: metastasis or second primary? Clin Cancer Res. 2005;11(18):6608–14.

17. Orolow I, Tommasi DV, Boom B, Ostrovnyaia I, Cotognola J, Mujumdar U, et al. Evaluation of the clonal origin of multiple primary melanomas using molecular profiling. J Invest Dermatol. 2009;129(8):1972–82.

18. Tang M, Pires Y, Schultz M, Duarte I, Gallegos M, Wistuba II. Microsatellite analysis of synchronous and metachronous tumors: a tool for double primary tumor and metastasis assessment. Diagn Mol Pathol. 2003;12(3):151–9.

19. Meyn JL, Cottu PH, Decraene C, Sterm MH, Couturier J, Lebigot I, et al. A genomic and transcriptomic approach for a differential diagnosis between primary and secondary ovarian carcinomas in patients with a previous history of breast cancer. BMC Cancer. 2010;10:222.

20. Jacoby MA, Duncavage EJ, Walter MJ. Implications of tumor clonal heterogeneity in the era of next-generation sequencing. Trends Cancer. 2015;1(4):231–41.

21. Sato-Otsubo A, Sanada M, Ogawa S. Single-nucleotide polymorphism array karyotyping in clinical practice: where, when, and how? Semin Oncol. 2012; 39(1):13–28.

22. Roth JJ, Santi M, Rorke-Adams LB, Harding BN, Busse TM, Tooke LS, et al. Diagnostic application of high resolution single nucleotide polymorphism array analysis for children with brain tumors. Cancer Genet. 2014;207(4):111–23.

23. Mao X, Young BD, Lu Y-J. The application of single nucleotide polymorphism microarrays in cancer research. Curr Genomics. 2007; 8(4):219–28.

24. Di Martino E, Selhous B, Hausmann R, Minkenberg R, Lohmann M, Esthofen MW. Survival in second primary malignancies of patients with head and neck cancer. J Laryngol Otol. 2002;116(10):831–8.

25. Baxi SS, Pinheiro LC, Patil SM, Pfister DG, Oeffinger KC, Elkin EB. Causes of death in long-term survivors of head and neck cancer. Cancer. 2014; 120(10):1507–13.

26. Floh MH. Netter’s Gastroenterology. 2nd ed. Elsevier; 2010.

27. Chen M-C, Huang W-C, Chan CH, Chen P-T, Lee K-D. Impact of second primary esophageal or lung cancer on survival of patients with head and neck cancer. Oral Oncol. 2010;46(4):249–54.

28. Curtius K, Wright NA, Graham TA. An evolutionary perspective on field cancerization. Nat Rev Cancer. 2018;18(11):59–32.

29. Sunpaweravong S, Sunpaweravong P, Dechaphunkul T, Pongrujikorn T, Bunbanjerdsuk S, Jinawath N. Clinico-molecular study of synchronous head and neck squamous cell carcinoma (HNSSC) and esophageal squamous cell carcinoma (ESCC). J Clin Oncol. 2015;33(suppl)abstr e17080.

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