Merged testing for colorectal cancer syndromes and re-evaluation of genetic variants improve diagnostic yield: Results from a nationwide prospective cohort

Sara Svensson1,2 | Theofanis Zagoras3,4 | Christos Aravidis5 | Marie Stenmark Askmalm1,2 | Erik Björck6,7 | Åke Borg8 | Ekaterina Kuchinskaya6,7 | Mef Nilbert8 | Margareta Nordling3,9 | Anna Rohlin3,4 | Gustav Silander10 | Kristina Lagerstedt-Robinson6,7 | Samuel Gebre-Medhin1,2

Abstract
Approximately 5% of patients with colorectal cancer (CRC) have a Mendelian predisposition for the disease. Identification of the disease-causing genetic variant enables carrier testing and tailored cancer prevention within affected families. To determine the panorama and genetic variation of Mendelian CRC syndromes among referrals at the cancer genetics clinics in Sweden, 850 patients clinically selected for CRC genetic investigation were included in a prospective study that tested for all major hereditary polyposis and nonpolyposis CRC conditions. Genetically defined syndromes were diagnosed in 11% of the patients. Lynch syndrome was predominant (n = 73) followed by familial adenomatous polyposis (n = 12) and MUTYH-associated polyposis (n = 8); the latter of which two patients presented with CRC before polyposis was evident. One patient with a history of adolescent-onset CRC and polyposis had biallelic disease-causing variants diagnostic for constitutional mismatch repair deficiency syndrome. Post-study review of detected variants of unknown clinical significance (n = 129) resulted in the reclassification of variants as likely benign (n = 59) or as diagnostic for Lynch syndrome (n = 2). Our results reveal the panorama of Mendelian CRC syndromes at the cancer genetics clinics in Sweden and show that unified testing for polyposis and nonpolyposis CRC conditions as well as regular reexamination of sequence data improve the diagnostic yield.

Keywords
colorectal cancer, genetic testing, hereditary, polyposis, syndrome, variant classification
INTRODUCTION

Approximately 5% of patients with colorectal cancer (CRC) have a constitutional disease-causing genetic variant (DV) that causes autosomal dominant (AD) or autosomal recessive (AR) predisposition to the disease. Lynch syndrome (LS) is by far the most common known Mendelian CRC condition with a prevalence approximately 1:300, followed by familial adenomatous polyposis (FAP) and MUTYH-associated polyposis (MAP; AR inheritance) with estimated prevalences approximately 1:10 000 to 1:40 000, respectively. There are several less common Mendelian conditions that predispose to CRC, such as juvenile polyposis syndrome (JPS), PTEN hamartoma tumor syndrome (PHTS), Peutz–Jeghers syndrome (PJS), polymerase proofreading-associated polyposis (PPAP), and a handful additional more recently defined conditions. Because DV may have prognostic, therapeutic, and prophylactic implications, screening for DV in selected patients with CRC and/or colorectal polyposis is routine in many health-care systems. In addition, identification of DV permits carrier testing and personalized health care for relatives. Patients with colorectal polyposis with or without CRC have traditionally been genetically screened FAP and MAP. Conversely, patients with non-polyposis CRC have traditionally been genetically screened for LS. To date, the outcomes of simultaneous testing for Mendelian CRC have mostly been evaluated in retrospective cohorts of selected or unselected patients. In this work, we have addressed the panorama and genetic variation of Mendelian CRC syndromes in a prospective national cohort of patients clinically selected for CRC genetic diagnostics.

MATERIALS AND METHODS

2.1 Patients

From 2014 to 2019, 861 patients with suspected Mendelian predisposition to CRC were invited to participate in the Swedish-extended genetic analysis of colorectal neoplasia (SWEN) study, a prospective study with inclusion from all cancer genetics clinics in Sweden, including the university hospitals in Umeå, Uppsala, Stockholm, Linköping, Gothenburg, and Lund (Figure 1). Patients obtained oral and written study information and provided written informed consent. This study was approved by The Regional Ethical Review Board in Lund (application no. 2013/468 and no. 2015/211) and by the Swedish Ethical Review Agency (application no. 2019-02312). The inclusion criteria of the study were as follows: (1) age of majority (≥18 years), (2) CRC clinical genetic investigation decided upon at a cancer genetics clinic in Sweden according to national clinical guidelines (Supplementary Material 1), and (3) written informed consent. After exclusion of 11 patients due to patient withdrawal (n = 5), no sample available (n = 2), incomplete molecular genetic analysis (n = 1), previously genetically screened for DV in MMR genes (n = 2), or inclusion merely on the basis of family history (n = 1), 850 patients were successfully enrolled in the study and cataloged regarding gender, neoplasms, colorectal polyps, and age at diagnosis. Records of unspecified numbers of colorectal polyps stating “polyposis”, or “many”, “numerous” or “massive numbers of” polyps were interpreted as 10 or more polyps and denoted polyposis.

2.2 Molecular genetic analyses

All patients were screened for DV in genes associated with established Mendelian CRC syndromes including LS (i.e., DNA mismatch repair [MMR] protein-encoding genes MLH1, MSH2, MSH6, and PMS2, and the MSH2-adjacent gene EPCAM), FAP (APC), MAP, JPS (BMPR1A and SMAD4), PHTS, and PJS (STK11). A majority of the patients (n = 702) were also screened for DV in the genes responsible for PPAP (POLE gene; POLD1 gene), and GALNT12 (putative gene for familial CRC type X; FCCTX), and the GREM1 gene (upstream duplication only; candidate gene for hereditary-mixed polyposis syndrome; HMPS). All molecular genetic analyses were performed with genomic DNA extracted from venous blood according to standard laboratory procedures. DNA was sequenced using targeted capture MPS (massive parallel sequencing) assay.
either using SureselectXT Custom and sequencing using Illumina HiSeq 2000 (n = 616) and analyzed as previously described, or using AmpliSeq following sequencing using an IonTorrent S5 (n = 234) as previously described and analyzed using IonReporter. All variants identified by capture MPS were confirmed using Sanger sequencing using standard protocols. Samples analyzed with IonTorrent sequencing regarding PMS2 were also subjected to nested PCR followed by Sanger sequencing as described. For detection of deletions/duplications, samples analyzed with IonTorrent were also analyzed using multiplex ligation-dependent probe amplification (MLPA) regarding the genes APC, EPCAM, GALNT12, MLH1, MSH2, MSH6, MUTYH, PMS2, PTEN, SMAD4, and STK11 (MRC-Holland, Amsterdam, The Netherlands) according to the manufacturer’s instructions.

### TABLE 1  Summary of clinical characteristics of all 850 patients by confirmed Mendelian colorectal cancer syndrome

| Mendelian syndrome | Total | Sex | Age at diagnosis (years)** | Sex Female (%) | Mendelian syndrome |
|--------------------|-------|-----|---------------------------|----------------|-------------------|
|                    |       |     |                           |                | LS                |
| All patients       | 850   | 527 (62)* | 49 (16–100) | | 113 | 5 | 5 | 1 |
| Cancer of any type | 806   | 503 (62) | 49 (17–100) | | 73 | 5 | 5 | 1 |
| Colorectal cancer  | 693   | 412 (59) | 50 (17–100) | | 61 | 4 | 5 | 1 |
| Polyposis          | 94    | 52 (55) | 49.5 (16–86) | | 12 | 8 | 1 | |
| ≥2 Cancers         | 163   | 123 (75)** | 50 (17–82) | | 21 | | | |
| ≥2 Colorectal cancers | 32    | 20 (63) | 56 (37–81) | | 8 | | | |
| Mendelian syndrome | 94    | 51 (54) | 45*** (16–81) | | 73 | 12 | 8 | 1 |
| Syndrome not detected | 756  | 476 (63) | 49 (17–100) | | | | | |

*P < 0.00001 for female sex/male sex. **P = 0.001 for ≥2 cancers in females/≥2 cancers in males. ***P < 0.01 for Mendelian syndrome/syndrome not detected. †Age at first cancer or polyposis.

### TABLE 2  Summary of cancers in cohort by confirmed mendelian colorectal cancer syndrome

| Mendelian syndrome | Total | Sex | Age (years)** | Sex Female (%) | Mendelian syndrome |
|--------------------|-------|-----|---------------|----------------|-------------------|
|                    |       |     |               |                | LS | FAP | MAP | CMMRD |
| All cancers        | 1025  | 670 (65) | 52 (16–100) | | 113 | 5 | 5 | 1 |
| Colon              | 594   | 353 (59) | 51 (17–100) | | 62 | 4 | 4 | 1 |
| Rectum             | 138   | 84 (61) | 47 (27–81) | | 10 | 1 | | |
| Uterus             | 74    | 74 (100) | 55 (27–82) | | 19 | | | |
| Breast             | 62    | 61 (98) | 53.5 (33–82) | | 6 | | | |
| Ovary              | 37    | 37 (100) | 49 (26–81) | | 4 | | | |
| Prostate           | 16    | 0 (0) | 66 (54–72) | | 2 | | | |
| Bladder            | 12    | 1 (8) | 52 (29–50) | | 1 | | | |
| Small bowel        | 11    | 6 (54) | 50 (32–75) | | 2 | | | |
| Stomach            | 9     | 2 (22) | 48 (36–70) | | 1 | | | |
| Kidney             | 9     | 5 (56) | 65 (38–72) | | 1 | | | |
| Pancreas           | 8     | 8 (100) | 70.5 (55–78) | | 1 | | | |
| Ureter             | 5     | 1 (20) | 55.5 (52–64) | | 3 | | | |
| Brain              | 3     | 2 (67) | 57 (55–67) | | 1 | | | |
| Thyroid            | 2     | 2 (100) | 58 (46–70) | | 1 | | | |
| Other              | 45    | 34 (76) | 48 (17–76) | | | | | |

Lynch syndrome.
Familial adenomatous polyposis.
MUTYH-associated polyposis.
Constitutional mismatch repair deficiency syndrome.
Data were analyzed using Gene Marker (Soft Genetics). Variant classification was performed according to the America College of Medical Genetics (ACMG)11 and ClinVar.12 Two main categories of variants were reported: DV including variants of class 4 (likely pathogenic variants) and variants of class 5 (pathogenic variants), and variants of unknown significance (VUS; class 3). Likely benign variants (class 2) and benign variants (class 1) were regarded as normal findings and were not reported. DV and VUS were re-evaluated in October 2021, that is, 2–7 years after initial reporting.

2.4 | Statistics

For statistical analysis, two-tailed t-test, chi-square test of goodness of fit, or chi-square test of independence were used (https://www.socscistatistics.com). Values of \( p < 0.05 \) were considered statistically significant.

3 | RESULTS

3.1 | Clinical characteristics of cohort

The cohort contained 527 females and 323 males (Table 1). A total of 1025 cancers were recorded with a median age at diagnosis of 52 years (Table 2). Cancer of any type, CRC, and polyposis occurred without significant differences between the sexes (Table 1). More females than males were recorded with \( \geq 2 \) cancers (Table 1). Females remained supernumerary to males when individuals with \( \geq 2 \) cancers were excluded (404 females vs. 283 males; \( P = 0.00001 \)). Similarly, females remained supernumerary to males when individuals with breast cancer and reproductive organ cancers were removed (381 females vs. 306 males; \( P = 0.0042 \)). Females were also more numerous than males among patients with solitary CRC and no other diagnosis (292 females vs. 226 males; \( P < 0.040 \)).

3.2 | Confirmed Mendelian CRC syndromes

Mendelian CRC syndromes were genetically confirmed in 94 patients (11% of cohort); LS was predominant (\( n = 73 \)), followed by FAP (\( n = 12 \)) and MAP (\( n = 8 \)) (Table 1). One patient with a history of adolescent-onset CRC and polyposis displayed biallelic DV in an MMR gene diagnostic for constitutional mismatch repair deficiency (CMMRD) (Table 1, patient ID C068; Supplementary Material 2). No patients were genetically confirmed with PPAP, JPS, PHTS, or PJS. All patients with LS had cancer and none of them had polyposis (Table 1). Polyposis was recorded in all patients with FAP, MAP, and CMMRD (Table 1). Age at cancer or polyposis was lower (median age 45 years) in patients with Mendelian CRC syndrome compared to patients with no syndrome detected (median 49 years; Table 1). In two patients

| Gene | Total | MLH1 | MSH2 | MSH6 | PMS2 | EPCAM | APC | BMPR1A | MUTYH | POLE | POLD1 | SMAD4 | STK11 | GALNT12 |
|------|-------|------|------|------|------|-------|-----|--------|-------|------|-------|-------|-------|-------|
| All DV | 112 | 20 | 28 | 15 | 11 | 1 | 12 | 24 | 1 | 11 | 1 | 1 | 1 | 1 |
| All VUS | 129 | 11 | 10 | 29 | 20 | 20 | 11 | 13 | 2 | 15 | 6 | 5 | 14 | 5 | 2 |
| VUS reclassified as LB | 59 | 4 | 4 | 5 | 8 | 2 | 9 | 6 | 14 | 5 | 14 | 5 | 2 |
| VUS reclassified as DV | 2 | 1 | 1 | 2 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |

aDisease-causing variant.
bIncluding two biallelic variants.
cPresent in patient with disease-causing variant in EPCAM.

\( ^{c} \)Likely benign variant.
with MAP, CRC was diagnosed >10 years before polyposis was discovered (age 51 vs. 64 years; age 39 vs. 50 years, respectively).

3.3 | Detected genetic variants and variant reclassification

A total of 112 DV and 129 VUS were reported (Table 3). Among the MMR genes, MSH2 displayed the greatest number of DV diagnostic for LS (Table 3). Two DV in PMS2 were confirmed as biallelic compound heterozygote variants diagnostic for CMMRD (Table 3, patient ID C068; Supplementary Material 2). In MUTYH, 16 DV were biallelic and accordingly diagnostic for MAP, whereas 8 DV were monoallelic representing heterozygote carriers for MAP (Table 3). Post-study re-evaluation of all detected variants resulted in the reclassification of 62 variants from VUS to likely benign (n = 59), from VUS to DV (n = 2), or from DV to VUS (n = 1; Table 3; Supplementary Material 2). The most frequent reasons for reclassification from VUS to likely benign were >1/500 allele frequency in any population (n = 21) and variants in POLE or POLD1 located outside the exonuclease domain (n = 17; Supplementary Material 2).

4 | DISCUSSION

In this work, we have investigated a prospective cohort of patients selected for CRC genetic investigation at the cancer genetics clinics in Sweden during 2014–2019. We aimed to determine the occurrence and genetic variation of Mendelian CRC syndromes using a comprehensive gene panel covering both nonpolyposis and polyposis conditions. A greater number of female patients was observed overall and throughout most clinical subcategories in our cohort, indicating a possible referral or pre-referral gender bias. Since the literature is scarce on the topic,13,14 we can only speculate that women in Sweden for reasons yet to be determined are more prone to request or accept or be offered cancer genetic investigation.

Median age of onset for CRC in our cohort was 49 years, which is >20 years lower than for CRC in the general population in Sweden,15 indicating compliance with national referral guidelines for CRC genetic investigation. LS was by far the most frequent Mendelian CRC syndrome detected in our study, followed by FAP and MAP both at approximately 1% levels, that is, mutual ratios for LS, FAP, and MAP repeatedly shown in other studies of selected and unselected patients with CRC and in line with reported prevalences in general populations.2,3,5-7 Given their rarity, the absence of identified patients with PPAP, PJS, PTHS, and PJS is not surprising as the size of our cohort is limited. Also, as symptoms may present early in life, some patients with these conditions might have been diagnosed before adulthood by pediatricians and thus have escaped our study.

We found one patient with CMMRD, a severe early-onset multiorgan cancer predisposition syndrome caused by biallelic DV in MMR genes, predominantly in PMS2.16 Molecular genetic diagnosis of CMMRD can be challenging as it may resemble neurofibromatosis type 1 presenting with café-au-lait macules and brain tumors, or FAP or MAP presenting with adenomatous polyposis.16 Our case with CMMRD had polyposis and CRC and would have escaped detection if genetic testing had been restricted to CRC polyposis conditions. Indeed, the clinical overlap between Mendelian CRC syndromes is now increasingly recognized,8,17 including accumulation of colorectal polyps in some cases of LS18 and attenuated forms of MAP and FAP.19 None of our patients with LS had ≥10 synchronous polyps. However, two of our patients with MAP presented with early-onset CRC more than a decade before polyposis was evident. This is in line with the findings of a study done by Nielsen et al., who showed that >30% of patients with CRC and proven biallelic DV in MUTYH do not have polyps and approximately 20% have <10 adenomas.20 Consequently, as already endorsed by others,21 multigene panel testing including all high-penetrant Mendelian CRC syndromes should be promoted in patients with early-onset CRC and/or polyposis.

DV in MSH2 and MLH1 were responsible for the majority of cases of LS in our study, which is largely in line with previous retrospective studies in Sweden,19,22-24 and in studies from other Western countries.25 We found no DV in the POLE and POLD1 exonuclease (proofreading) domains, but several VUS were detected in other regions of these genes. Possibly, as our understanding of the clinical impact expands, some of the current VUS located outside the proofreading domains may prove clinically relevant. Similarly, after variant re-evaluation, no DV but several VUS remained in the genes associated with the hamartomatous polyposis syndromes.

More VUS than DV were initially identified in our cohort. However, after re-evaluation approximately 45% of VUS were subsequently reclassified as likely benign variants. This shows that the ongoing sharing of data to open variant databases is crucial for improved variant classification. The importance of data sharing is further emphasized by the two cases of VUS in PMS2 and MLH1, respectively, which were reassessed as causative for LS in our work. Although systematic reanalysis of sequence data has been reported to increase diagnostic yield,26-29 clinical practice for reanalysis and reinterpretation remains to be established.30 Nevertheless, as outlined above we conclude that in the absence of multigene panel testing and follow-up of VUS a total of five patients in our cohort could potentially have escaped the molecular genetic diagnosis of a Mendelian CRC syndrome.

During the study period, the routine laboratory diagnostic workup for simplex cases with an LS-associated tumor diagnosed between ages 40–49 years included tumor tissue MMR protein immunohistochemical and mutation testing of BRAF, and subsequent targeted molecular genetic testing of any indicated MMR gene(s). Consequently, an unknown number of patients could have been diagnosed with LS through this alternative route, that is, outside the present cohort study. After variant re-evaluation in an earlier retrospective study with emphasis on LS in Sweden, we observed only one such simplex case among a total of 52 patients with confirmed LS implying few additional cases diagnosed with LS during the period of the current study.
The role of heterozygous DV in MUTYH as risk alleles for CRC remains controversial.\textsuperscript{20,31} We found eight heterozygote carriers for MAP, which corresponds to 0.94% of all patients in our cohort. The MAP carrier frequency in Northern Europe has been approximated to 1%-2%.\textsuperscript{20,33} Since MAP carriers seem not to be accumulated in our cohort, our data do not support heterozygous DV in MUTYH as risk alleles for early-onset CRC.

Introduction of paired sample tumor tissue DNA profiling for diagnostic and treatment decision purposes\textsuperscript{32–34} as well as exome or genome sequencing of noncancer patients now contribute to the identification of cancer predisposition alleles, either as sought or unsolicited findings. These rapidly emerging fields bypass the traditional triage for genetic testing of patients with suspected cancer predisposition based on age at diagnosis and family history. As technology develops and costs for testing drop it could be anticipated that a large proportion of germline cancer susceptibility soon will be identified through these novel diagnostic routes.

5 | CONCLUSION

In summary, we describe the panorama and genetic variation of Mendelian CRC syndromes in a prospective cohort of patients with a suspected predisposition to CRC at the cancer genetics clinics in Sweden. Our data show that unified testing for the different Mendelian CRC syndromes as well as an intermittent reexamination of sequence data improve the diagnostic yield.

ACKNOWLEDGMENTS
We would like to thank the patients who participated in the study. The coworkers at the participating molecular diagnostic laboratories are greatly acknowledged. We also thank the health-care regions in Sweden for support and infrastructure.

CONFLICT OF INTEREST
The authors declare no conflict of interest.

DATA AVAILABILITY STATEMENT
The data that supports the findings of this study are available in the supplementary material of this article.

REFERENCES
1. Stoffel EM, Boland CR. Genetics and genetic testing in hereditary colorectal cancer. Gastroenterology. 2015;149(5):1191-1203 e2. doi: 10.1053/j.gastro.2015.07.021
2. Win AK, Jenkins MA, Dowty JG, et al. Prevalence and penetrance of major genes and polygenes for colorectal cancer. Cancer Epidemiol Biomarkers Prev. 2017;26(3):404-412. doi: 10.1158/1055-9965.EPI-16-0693
3. Ollikoina AP, Peltomaki PT, Aaltonen LA, Rajamaki K. From APC to the genetics of hereditary and familial colon cancer syndromes. Hum Mol Genet. 2021;30(20):R206-R224. doi: 10.1093/hmg/ddab208
4. Valle L, Villar E, Tavtigian SV, Stoffel EM. Genetic predisposition to colorectal cancer: syndromes, genes, classification of genetic variants and implications for precision medicine. J Pathol. 2019;247(5):574-588. doi: 10.1002/path.5229
5. Stoffel EM, Koepp E, Everett J, et al. Germline genetic features of young individuals with colorectal cancer. Gastroenterology. 2018; 154(4):897-905.e1. doi: 10.1053/j.gastro.2017.11.004
6. Chubb D, Broderick P, Frpmton M, et al. Genetic diagnosis of high-penetration susceptibility for colorectal cancer (CRC) is achievable for a high proportion of familial CRC by exome sequencing. J Clin Oncol. 2015;33(5):426-432. doi: 10.1200/JCO.2014.56.5689
7. Yurgelun MB, Kulpke MH, Fuchs CS, et al. Cancer susceptibility gene mutations in individuals with colorectal cancer. J Clin Oncol. 2017; 35(10):1086-1095. doi: 10.1200/JCO.2016.71.0012
8. Rolnin A, Rambech E, Kvist A, et al. Expanding the genotype-phenotype spectrum in hereditary colorectal cancer by gene panel testing. Fam Cancer. 2017;16(2):195-203. doi: 10.1007/s10689-016-9934-0
9. Tesi B, Lagerstedt-Robinson K, Chiag SC, et al. Targeted high-throughput sequencing for genetic diagnostics of hemophagocytic lymphohistiocytosis. Genome Med. 2015;7:130. doi: 10.1186/s13073-015-0244-1
10. Vaughn CP, Robles J, Swensen JJ, et al. Clinical analysis of PMS2: mutation detection and avoidance of pseudogenes. Hum Mutat. 2010;31(5):588-593. doi: 10.1002/humu.21230
11. Richards S, Aziz N, Bale S, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med. 2015;17(5):405-424. doi: 10.1038/gim.2015.30
12. Landrum MJ, Chitipiralla S, Brown GR, et al. ClinVar: improvements in accessing data. Nucleic Acids Res. 2020;48(D1):D835-D844. doi: 10.1093/nar/gkaa972
13. Espenschied C, Pepper J, McFarlane RE. What about the guys? An assessment of gender differences in hereditary colorectal cancer testing. J Clin Oncol. 2017;35(4):537. doi: 10.1200/JCO.2017.35.4_suppl.537
14. Childers KK, Maggard-Gibbons M, Macinko J, Childers CP. National Distribution of cancer genetic testing in the United States: evidence for a gender disparity in hereditary breast and ovarian cancer. JAMA Oncol. 2018;4(6):876-879. doi: 10.1001/jamaoncol.2018.0340
15. National Board of Health and Welfare. Cancer statistics in Sweden; 2018. https://www.socialstyrelsen.se/en/statistics-and-data/
16. Aronson M, Colas C, Shen A, et al. Diagnostic criteria for constitutional mismatch repair deficiency (CMMRD): recommendations from the international consensus working group. J Med Genet. 2022;59(4):318–327. doi: 10.1136/jmedgenet-2020-107627
17. Sutcliffe EG, Bartenbacher Thompson A, Stettner AR, et al. Multi-genie panel testing confirms phenotypic variability in MUTYH-associated polyposis. Fam Cancer. 2019;18(2):203-209. doi: 10.1007/s10689-018-0116-2
18. Kalady MF, Kravochuck SE, Heald B, Burke CA, Church JM. Defining the adenoma burden in Lynch syndrome. Dis Colon Rectum. 2015; 58(4):388-392. doi: 10.1001/jcrc.2015.0333
19. Grover S, Kastrinos F, Steyerberg EW, et al. Prevalence and phenotypes of APC and MUTYH mutations in patients with multiple colorectal adenomas. JAMA. 2012;308(5):485-492. doi: 10.1001/jama.2012.8780
20. Nielsen M, Morreau H, Vasen HF, Hes FJ. MUTYH-associated polyposis (MAP). Crit Rev Oncol Hematol. 2011;79(1):1-16. doi: 10.1016/j.critrevonc.2010.05.011
21. Heald B, Hampel H, Church J, et al. Collaborative Group of the Americas on inherited gastrointestinal cancer position statement on multi-gene panel testing for patients with colorectal cancer and/or polyposis. Fam Cancer. 2020;19(3):223-239. doi: 10.1007/s10689-020-00170-9
22. Lagerstedt Robinson K, Liu T, Vandrovocova J, et al. Lynch syndrome (hereditary nonpolyposis colorectal cancer) diagnostics. *J Natl Cancer Inst.* 2007;99(4):291-299. doi:10.1093/jnci/djk051

23. Lagerstedt-Robinson K, Rohlin A, Aravidis C, et al. Mismatch repair gene mutation spectrum in the Swedish lynch syndrome population. *Oncol Rep.* 2016;36(5):2823-2835. doi:10.3892/or.2016.5060

24. Henriksson I, Henriksson K, Ehrencrona H, Gebre-Medhin S. Hereditary colorectal cancer diagnostics in southern Sweden: retrospective evaluation and future considerations with emphasis on lynch syndrome. *J Community Genet.* 2019;10(2):259-266. doi:10.1007/s12687-018-0385-1

25. Thompson BA, Spurdle AB, Plazzer JP, et al. Application of a 5-tiered scheme for standardized classification of 2,360 unique mismatch repair gene variants in the InSiGHT locus-specific database. *Nat Genet.* 2014;46(2):107-115. doi:10.1038/ng.2854

26. Hiatt SM, Amaral MD, Bowling KM, et al. Systematic reanalysis of genomic data improves quality of variant interpretation. *Clin Genet.* 2018;94(1):174-178. doi:10.1111/cge.13259

27. Sun Y, Xiang J, Liu Y, et al. Increased diagnostic yield by reanalysis of data from a hearing loss gene panel. *BMC Med Genomics.* 2019;12(1):76. doi:10.1186/s12920-019-0531-6

28. Schmitz-Abe K, Li Q, Rosen SM, et al. Unique bioinformatic approach and comprehensive reanalysis improve diagnostic yield of clinical exomes. *Eur J Hum Genet.* 2019;27(9):1398-1405. doi:10.1038/s41431-019-0401-x

29. Liu P, Meng L, Normand EA, et al. Reanalysis of clinical exome sequencing data. *N Engl J Med.* 2019;380(25):2478-2480. doi:10.1056/NEJMct1812033

30. Vears DF, Senecal K, Borry P. Genetic health professionals’ experiences with initiating reanalysis of genomic sequence data. *Fam Cancer.* 2020;19(3):273-280. doi:10.1007/s10689-020-00172-7

31. Patel R, McGinty P, Cuthill V, Hawkins M, Clark SK, Latchford A. Risk of colorectal adenomas and cancer in monoallelic carriers of MUTYH pathogenic variants: a single-Centre experience. *Int J Colorectal Dis.* 2021;36(10):2199-2204. doi:10.1007/s00384-021-03983-x

32. Hampel H, Pearlman R, Beightol M, et al. Assessment of tumor sequencing as a replacement for lynch syndrome screening and current molecular tests for patients with colorectal cancer. *JAMA Oncol.* 2018;4(6):806-813. doi:10.1001/jamaoncol.2018.0104

33. Bryjalsen A, Hansen TVO, Stoltze UK, et al. Nationwide germline whole genome sequencing of 198 consecutive pediatric cancer patients reveals a high incidence of cancer prone syndromes. *PLoS Genet.* 2020;16(12):e1009231. doi:10.1371/journal.pgen.1009231

34. Goudie C, Witkowski L, Cullinan N, et al. Performance of the McGill interactive pediatric OncoGenetic guidelines for identifying cancer predisposition syndromes. *JAMA Oncol.* 2021;7(12):1806-1814. doi:10.1001/jamaoncol.2021.4536

**SUPPORTING INFORMATION**

Additional supporting information may be found in the online version of the article at the publisher's website.

**How to cite this article:** Svensson S, Zagoras T, Aravidis C, et al. Merged testing for colorectal cancer syndromes and reevaluation of genetic variants improve diagnostic yield: Results from a nationwide prospective cohort. *Genes Chromosomes Cancer.* 2022;61(10):585-591. doi:10.1002/gcc.23049