Expression of HO1 and PER2 can predict the incidence of delirium in trauma patients with concomitant brain injury

Matti Steimer¹,2, Sandra Kaiser¹,2, Felix Ulbrich¹,2, Johannes Kalbhenn¹,2, Hartmut Bürkle¹,2 & Nils Schallner*¹,2

Intensive care unit (ICU)-acquired delirium is associated with adverse outcome in trauma patients with concomitant traumatic brain injury (TBI), but diagnosis remains challenging. Quantifying circadian disruption by analyzing expression of the circadian gene period circadian regulator 2 (PER2) and heme oxygenase 1 (HO1), which determines heme turnover, may prove to be potential diagnostic tools. Expression of PER2 and HO1 was quantified using qPCR from blood samples 1 day and 7 days after trauma. Association analysis was performed comparing mRNA expression levels with parameters of trauma (ISS—injury severity score), delirium, acute kidney injury (AKI) and length of ICU stay. 48 polytraumatized patients were included (equal distribution of TBI versus non-TBI) corrected for ISS, age and gender using a matched pairs approach. Expression levels of PER2 and HO1 were independent of age (PER2: P = 0.935; HO1: P = 0.988), while expression levels were significantly correlated with trauma severity (PER2: P = 0.009; HO1: P < 0.001) and longer ICU length of stay (PER2: P = 0.018; HO1: P < 0.001). High expression levels increased the odds of delirium occurrence (PER2: OR = 4.32 [1.14–13.87]; HO1: OR = 4.50 [1.23–14.42]). Patients with TBI showed a trend towards elevated PER2 (OR = 3.00 [0.84–9.33], P = 0.125), but not towards delirium occurrence (P = 0.556). TBI patients were less likely to develop AKI compared to non-TBI (P = 0.022). Expression levels of PER2 and HO1 correlate with the incidence of delirium in an age-independent manner and may potentially improve diagnostic algorithms when used as delirium biomarkers.

Trial registration: German Clinical Trials Register (Trial-ID DRKS00008981; Universal Trial Number U1111-1172-6077; Jan. 18, 2018).

Trauma patients in the ICU often exhibit impaired sleep–wake-cycles¹, impaired cognition and impaired organ function, which might become manifest clinically as delirium and acute kidney injury (AKI). Disruption in circadian rhythm is now believed to play a major role in development of delirium² and contribute to the pathogenesis of kidney disease³,⁴. Both delirium and AKI are independently associated with adverse patient outcome and mortality³,⁵. Their diagnosis however remains challenging⁶. A better physiological and biochemical understanding of delirium and AKI development is key to finding objective laboratory parameters to predict, diagnose and prognosticate outcome. Particularly in the study of delirium, multiple potential biomarkers in both serum and cerebrospinal fluid have been proposed⁷–¹², however none have been shown to provide added clinical value¹¹. Despite the proposed role of disrupted circadian rhythmicity in development of delirium the molecular machinery behind it has not been investigated in search of a biomarker yet.

The core clock genes that regulate autonomous circadian rhythm include the Period genes (Per 1–3)¹³. These are associated with susceptibility to organ dysfunction, including the brain and cardiovascular system¹⁴–¹⁷. Eckle et al. for instance identified transcriptional and post-translational regulation of PER2 as well as the stabilization of the resulting protein as an integral step in the adaption of cardiomyocytes to ischemia. Consequently, PER2−/− mice had larger infarct sizes than their wild-type controls. With regard to delirium, increasing PER2 expression has been shown to be effective against midazolam-induced delirium in mice¹⁸.

¹Department of Anesthesiology & Critical Care Medicine, Medical Center - Faculty of Medicine, University of Freiburg, Hugstetter Str. 55, 79106 Freiburg, Germany. ²Faculty of Medicine, University of Freiburg, Freiburg, Germany. *email: nils.schallner@uniklinik-freiburg.de
The cytoprotective enzyme heme oxygenase 1 (HO1) serves as the rate-limiting step in heme metabolism and is strongly upregulated following various cellular stressors\(^1\). HO1 catalyzes the degradation of heme into iron, biliverdin and CO. Biliverdin is then further reduced to bilirubin by oxidation of NAD(P)H to NAH(P)\(^+\). HO1 expression may serve as a potential predictive blood biomarker since it is closely linked to circadian regulation as it: (a) catabolizes the heme moiety contained in the circadian regulatory proteins NPAS-2 and RevErb-\(\alpha\), and (b) produces carbon monoxide (CO) as a bioactive product. Endogenous CO in turn modulates the transcriptional activity of NPAS-2, RevErb-\(\alpha\) and CLOCK that control expression of PER2\(^2\). Moreover, antioxidant qualities of HO1 (and CO) are well documented. Especially in the case of AKI HO1 induction has been identified as a rapid cytoprotective response in multiple animal models of AKI\(^21\text{-}26\).

Here, we hypothesized that PER2 and HO1 expression could serve as potential serum biomarkers to diagnose and predict delirium and/or AKI. We posited that alterations in PER2 and HO1 mRNA expression would indicate circadian disruption. Being at high risk to develop both delirium and AKI\(^27\text{-}28\), we tested this hypothesis in polytraumatized patients. As circadian rhythm is controlled by the hypothalamus, a trauma patient collective afforded us the opportunity to simultaneously investigate the effect of traumatic brain injury (TBI) on circadian disruption and clinical outcome. We studied a sub-group of patients with TBI and measured PER2 and HO1 mRNA expression and asked if expression correlated with subjects developing delirium and/or AKI.

**Results**

**Patient characteristics and recruitment.** A total of 72 patients were screened of which three patients were excluded due to pre-existing kidney disease. Of the remaining 69, a total of 48 polytrauma patients were included in the study (Fig. 1b). Using a matched pairs approach, two 24-patient groups with TBI and non-TBI patients were formed. These groups did not significantly vary regarding age at trauma, gender, trauma ISS, pre-existing cognitive or cardiovascular disease, diabetes mellitus, time spent in the ICU and maximum leucocyte counts during the first week after trauma (Table 1).

**HO1 expression correlates with bilirubin concentration.** To verify that expression levels mirror actual enzymatic activity, we tested HO1 expression and bilirubin concentration for correlation, as bilirubin is the final product of heme degradation. Indeed, HO1 expression levels on the first day correlated with concurrent bilirubin concentration (\(r = 0.315, P = 0.029, r^2 = 0.378, P < 0.001\), Fig. 2a) on day one and after one week.
The correlation of systemic HO1 expression after one week and bilirubin concentration at the same time missed significance ($r = 0.253, P = 0.149, r^2 = 0.100, P < 0.067$; data not shown).

Table 1. Patient characteristics. Presented in total and within matched groups. $P$-values of between-group comparisons of patients with traumatic brain injury (TBI) and without (non-TBI) calculated by Mann Whitney or Fisher’s exact test. All $P > 0.05$. SD standard deviation, IQR inter-quartile ratio, ICU intensive care unit.

|                         | Total | TBI  | Non-TBI | $P$  |
|-------------------------|-------|------|---------|------|
| n                       | 48    | 24   | 24      |      |
| Age [years]             |       |      |         |      |
| Mean                    | 53.54 | 55.13| 51.96   | 0.422|
| SD                      | 16.73 | 17.70| 15.92   |      |
| <40                     | 8     | 4    | 4       |      |
| 40–75                   | 36    | 18   | 18      |      |
| >75                     | 4     | 2    | 2       |      |
| Gender                  |       |      |         |      |
| Male                    | 42    | 21   | 21      |      |
| Female                  | 6     | 3    | 3       |      |
| Injury severity score   |       |      |         |      |
| Mean                    | 32.52 | 32.71| 32.33   | 0.947|
| SD                      | 12.60 | 13   | 12.46   |      |
| <25                     | 16    | 8    | 8       |      |
| 25–36                   | 20    | 10   | 10      |      |
| >36                     | 12    | 6    | 6       |      |
| Pre-existing neurocognitive disorder | 4 | 3 | 1 | 0.609 |
| Pre-existing cardiovascular disease | 19 | 10 | 9 | >0.999 |
| Diabetes mellitus       | 6     | 4    | 2       | 0.666|
| ICU length of stay [days] |       |      |         |      |
| Mean                    | 11.63 | 12.13| 11.13   | 0.770|
| SD                      | 11.25 | 11.82| 10.62   |      |
| Max. leucocyte count first week [1000/µl] |       |      |         |      |
| Median                  | 14.54 | 13.78| 15.20   | 0.617|
| IQR                     | [11.24 to 17.25] | [11.13 to 15.96] | [11.36 to 18.64] |      |

Figure 2. Correlation of systemic HO1 expression with bilirubin plasma concentration. Linear regression curves of systemic HO1 expression on admission and (a) bilirubin concentration on admission (Spearman: $r = 0.315, P = 0.029$, Linear Regression: $r^2 = 0.376, P < 0.0001$) and (b) bilirubin expression after 1 week (Spearman: $r = 0.424, P = 0.011$, Linear Regression: $r^2 = 0.377, P < 0.0001$). HO1 heme oxygenase 1.

($r = 0.424, P = 0.011, r^2 = 0.377, P < 0.001$, Fig. 2b). The correlation of HO1 expression after one week and bilirubin concentration at the same time missed significance ($r = 0.253, P = 0.149, r^2 = 0.100, P < 0.067$, data not shown).

Biomarkers correlate with trauma severity and duration of ICU treatment. To review our hypotheses (Fig. 1a), the relationships between input variable and mediator as well as between mediator and output data were tested. Indeed, a higher ISS correlated with significantly higher mRNA expression levels of PER2 ($r = 0.375, P = 0.009, r^2 = 0.166, P = 0.004$, Fig. 3a) and HO1 ($r = 0.603, P < 0.001, r^2 = 0.346, P < 0.001$, Fig. 3b), and among each other, ($r = 0.424, P = 0.003, r^2 = 0.144, P = 0.008$, data not shown). Similarly, higher
PER2 (r = 0.340, P = 0.018, r² = 0.067, P = 0.008, Fig. 3c) and HO1 mRNA expression levels (r = 0.604, P < 0.001, r² = 0.483, P < 0.001, Fig. 3d) correlated with significantly longer ICU length of stay. Bilirubin concentration on day one showed no significant correlation with the ISS (r = 0.182, P = 0.216, data not shown). Higher bilirubin concentration on day one did, however, correlate significantly with longer ICU length of stay (r = 0.420, P = 0.003; r² = 0.349, P < 0.001, Fig. 3e).

Biomarkers correlate with occurrence of delirium. A significant correlation was discovered between the expression of both PER2 and HO1 on day one after trauma and the maximum nuDesc (PER2: r = 0.319, P = 0.027, Fig. 4a; HO1: r = 0.409, P = 0.004, Fig. 4b). Correlation of delirium and PER2 expression remained significant 1 week post trauma (r = 0.366, P = 0.024, r² = 0.083, P = 0.048, Fig. 4c). In accordance with age as a major risk factor for the development of delirium²⁹, maximum nuDesc in the first week after trauma correlated significantly with a patient's age at the time of trauma (r = 0.370, P = 0.010, r² = 0.105, P = 0.024, Fig. 4d). No correlation could be found between a patient's age at the time of trauma and PER2 and HO-1 expression (PER2: r = 0.012, P = 0.935, r² = 0.005, P = 0.634; HO1: r = 0.002, P = 0.988, r² = 0.029, P = 0.247, Fig. 4e), suggesting an age-independent correlation between these gene expression patterns and delirium. Trauma severity measured by ISS did not correlate with maximum nuDesc after trauma (r = 0.070, P = 0.638, r² = 0.001, P = 0.825, data not shown). Mean values of PER2 and HO1 mRNA expression on day one after trauma (P = 0.013) and after one week (P = 0.016 and P = 0.004 respectively) were significantly elevated in delirious patients (Table 2). When dividing patients into categories of high and low mRNA expression in relation to ISS scoring, delirium (nuDesc > 1) was significantly more frequent in patients within the group of high PER2 (P = 0.032, Fig. 4f) and HO1 (P = 0.019,
Figure 4. Correlation of PER2 and HO1 expression with delirium during hospitalization. (a–c) Linear regression curves between mRNA expression (PER2/HO1) as well as patients’ age at admission and maximal nuDesc. (a,b) Correlation of PER2 (Spearman: r = 0.319, P = 0.027, Linear Regression: r² = 0.057, P = 0.104) and HO1 expression (r = 0.409, P = 0.004, r² = 0.075, P = 0.080) on first day of trauma with their maximal nuDesc during hospitalization. (c) Correlation of PER2 after 1 week and with their maximal nuDesc during hospitalization (r = 0.366, P = 0.024, r² = 0.083, P = 0.048). (d) Correlation of age at admission with the maximal nuDesc during the first week of hospitalization (r = 0.370, P = 0.010, r² = 0.105, P = 0.024). (e) (Absent) Correlation between patients’ age at admission and PER2 (Spearman: P = 0.935, Linear Regression: P = 0.634) and HO1 expression (P = 0.988, P = 0.247) on the first day of trauma. (f) Contingency diagrams analyzed via Fisher’s exact test showing distribution of delirium/non-delirium between patient groups of high and low mRNA expression levels of PER2 (Fisher’s exact test: P = 0.032, Koopman asymptotic score: RR = 2.92, 95% CI 1.15–4.64; Baptista-Pike: OR = 4.32, 95% CI 1.14–13.87) and HO1 (P = 0.019, RR = 2.40, 95% CI 1.18–5.07; OR = 4.50, 95% CI 1.23–14.42). Categories of high and low expression levels of PER2 and HO1 were formed from linear regression of ISS and mRNA expression levels on first day of trauma (cf. Fig. 3a,b). (g) Higher individual dispersion of nuDesc during hospital stay in patients of the category “delirium” than “non-delirium” (Mann–Whitney-U test: P < 0.0001, cf. Methods/clinical assessment). nuDesc nursing delirium screening scale, 95% CI 95% confidence interval, OR odds ratio, PER2 period circadian regulator 2, HO1 heme oxygenase 1, ISS injury severity score.
Table 2. Median HO1 and PER2 mRNA expression between groups of TBI/non-TBI and delirium/non-delirium over time. Sample acquisition on first day and after 1 week. *p*-values of between-group comparisons of PER2 and HO1 mRNA expression (median and IQR) calculated via Mann Whitney test between groups of patients with and without delirium as well as between TBI and non-TBI groups. PER2 period circadian regulator 2, HO1 heme oxygenase 1, TBI traumatic brain injury, IQR inter-quartile ratio.

|          | TBI                  | Non-TBI              | Delirium          | Non-delirium          |          |
|----------|----------------------|----------------------|-------------------|-----------------------|----------|
| n        | 24                   | 24                   | 19                | 29                    |          |
| PER2 expression |                     |                      |                   |                       |          |
| Day 1    | 3.419 [2.38 to 4.085]| 2.319 [1.647 to 3.828]| 0.125 3.490 [2.830 to 5.927]| 2.325 [1.635 to 3.533]| 0.013    |
| Day 8    | 2.702 [1.808 to 3.829]| 2.235 [1.033 to 3.382]| 0.322 3.483 [2.388 to 5.477]| 3.583 [2.847 to 4.294]| 0.191    |
| HO1 expression |                     |                      |                   |                       |          |
| Day 1    | 5.130 [3.721 to 6.273]| 5.634 [3.302 to 10.46]| 0.646 6.130 [5.130 to 10.66]| 4.371 [2.998 to 6.242]| 0.004    |
| Day 8    | 3.666 [2.808 to 5.147]| 4.053 [3.016 to 5.227]| 0.504 4.895 [2.942 to 5.793]| 3.583 [2.847 to 4.294]| 0.191    |

Fig. 4f) expression. Relative risk to develop delirium was 2.92 (95% CI 1.15–4.64) for patients with high PER2 and 2.40 (95% CI 1.18–5.07) for patients high HO1 expression. Correspondingly, the odds ratios of developing delirium with high expression levels were 4.32 (PER2, 95% CI 1.14–13.87) and 4.5 (HO1, 95% CI 1.23–14.42), respectively. Patients classified as delirious according to their maximum nuDesc showed significantly higher SDs of nuDesc during their hospital stay (*p* < 0.0001, Fig. 4g).

Bilirubin concentration neither correlated with maximum nuDesc (*r* = 0.065, *p* = 0.663, *r*² = 0.011, *p* = 0.486, data not shown) nor were patients with high bilirubin levels more likely to develop delirium compared to those with low levels (*p* = 0.770, data not shown).

**TBI influences PER2 expression but not HO1 expression or occurrence of delirium.** As shown in Table 2, no significant differences in mRNA expressions were apparent for either PER2 or HO1 between groups of TBI and non-TBI patients. However, there was a tendency towards elevated values of PER2 on day one after trauma in TBI patients with a relative risk of 2.00 (95% CI 0.94–4.52) and an OR of 3.00 (95% CI 0.84–9.33, *p* = 0.125). A similar non-significant tendency was apparent when comparing the distribution of high and low PER2 mRNA expression on day one after trauma comparing TBI and non-TBI (*p* = 0.135, Fig. 5a). Distribution of high and low HO1 mRNA expression and bilirubin concentration showed no difference between TBI and non-TBI patients (HO1: *p* = 0.770; bilirubin: *p* = 0.561). Similarly, analysis of delirium occurrence in TBI and non-TBI patients revealed no difference in delirium when comparing TBI with non-TBI groups (*p* = 0.556).

**TBI patients show smaller likelihood of developing AKI.** As a second clinical readout, the occurrence of AKI was investigated in relation to the biomarkers defined above and in relation to the presence or absence of TBI. Compared with non-TBI trauma patients, TBI patients showed a significantly lower risk of developing AKI (*p* = 0.022, Fig. 5b). However, higher prevalence of AKI did not significantly coincide with either high PER2 (*p* > 0.999, Fig. 5c) or HO1 mRNA expression (*p* = 0.219). Patients with high bilirubin concentrations on day one non-significantly tended towards developing AKI (*p* = 0.073, data not shown).

**Six-month mortality does not differ between TBI versus non-TBI and between biomarker risk groups.** Of the 48 trauma patients included in the study, four patients died within 6 months after ICU admission. Descriptive statistical analysis (retrospectively and limited by an underpowered sample size) identified both TBI (RR = 0.33, 95% CI 0.049–2.16; OR = 0.30, 95% CI 0.023–2.23) and high PER2 mRNA expression (RR = 0.56, 95% CI 0.082–3.53; OR = 0.53, 95% CI 0.039–3.85) to be associated with less 6-month mortality, while high HO1 mRNA expression (RR = 1.40, 95% CI 0.26–7.43; OR = 1.44, 95% CI 0.21–9.80), high bilirubin concentration (RR = 1.31, 95% CI 1.04–1.31; OR = Infinity, 95% CI 1.00–Infinity), delirium (RR = 4.58, 95% CI: 0.70–30.69; OR = 5.25, 95% CI 0.71–70.17) and AKI (RR = 7.00, 95% CI 1.32–33.01; OR = 10.00, 95% CI 1.20–71.00) carried an elevated risk of death (Fig. 5d). However, each of these associations remained non-significant (TBI: *p* = 0.609, high PER2: *p* > 0.999, high HO1: *p* > 0.999, high bilirubin: *p* = 0.103, delirium: *p* = 0.286, AKI: *p* = 0.071).

**Discussion**

In this study, we measured systemic PER2 and HO1 mRNA expression on day one after trauma. We conclude that both HO1 and PER2 may serve as novel blood biomarkers for trauma severity, length of ICU-stay and, most interestingly, the risk of developing trauma-associated delirium.

There is an unmet need for objective laboratory parameters to improve diagnosis and treatment algorithms for delirium. Many have been tested but have so far proven incapable of having diagnostic value in a clinical setting[1]. Delirious patients commonly demonstrate compromised sleep–wake-cycles, and molecular circadian disruption is considered to play a major role in delirium pathogenesis[2]. Nonetheless, circadian gene expression has not been investigated as potential biomarkers for delirium. Our data suggests that PER2 and HO1 mRNA expressions may prospectively assess delirium and trauma severity. Further we measured concentrations of bilirubin, the final product of heme degradation by HO1. By demonstrating a correlation between HO1 expression and bilirubin concentration we can confirm the notion that mRNA expression mirrors actual enzymatic activity.
Expression of PER2 was not only significantly elevated on day one, but remained elevated one week post trauma. These findings suggest not only predictive value, as is the case with HO1, but also provided diagnostic value for occurrence and severity of trauma-associated delirium. Although the patient’s age at trauma is a known major risk factor for developing delirium it could not explain the predictive aspects of PER2 and HO1, suggesting age-independence and hence potentially additive diagnostic value. Trauma severity did not contribute to identifying patients at risk for trauma-associated delirium.

Although circadian disruption is thought to contribute to kidney disease, no predictive or diagnostic capacity of PER2 or HO1 expression could be found for AKI. However, this may be due to the limited amount of AKI cases in our patient cohort. Work by others strongly suggests that HO1 plays a major role in stress response to acute kidney injury. Patients with more GT-repeats in the HO1-gene promoter region have been shown to be less likely to develop AKI. Therefore, further studies should investigate the predictive and diagnostic capacity of HO1 and PER2 for those with AKI.

Similarly, 6-month mortality could not be attributed to high HO1 and PER2 expression as well as TBI. With a total of only four deceased patients, our cohort of n = 48 was underpowered for this endpoint and similar to AKI, larger cohort studies might prove to yield more conclusive results. On a descriptive statistical level limited by our patient cohort, our data suggest that TBI and high PER2 expression may be favorable for survival, while high HO1 expression, high bilirubin, delirium and AKI carried a greater risk of death. The last two of which are already considered to contribute to increased mortality.

Intracranial bleeding has been shown to cause disruption in circadian gene expression in mice. It was investigated whether TBI has an effect on the likelihood of developing delirium and AKI. TBI did not show any significant impact on occurrence of trauma-associated delirium or systemic mRNA expression levels of PER2 and HO1. Thus, our findings cannot support our hypothesis of TBI on top of polytrauma causing more severe molecular circadian disruption and thereby contributing to the pathogenesis of delirium and AKI. It is possible, however, that our definition of TBI as a radiographical diagnosis might have been too broad. Our categorization may not have allowed for sufficient effect sizes and significant differences between TBI and non-TBI groups. In addition, systemic changes in mRNA expression were measured in peripheral blood leucocytes, which might not be representative of the regional changes in PER2 and HO1 expression in brain tissue following TBI.
patients tended to have elevated PER2 expression on day one after trauma when compared with non-TBI patients. Further studies evaluating regional PER2 and HO1 expression following TBI, e.g. in the cerebrospinal fluid might help fill this gap.

Interestingly, PER2 expression in peripheral blood cells as a surrogate for disturbance of circadian rhythmicity has recently been studied in both polytrauma and general ICU patient collectives\(^{31,32}\). In both cases, low PER2 expression was associated with systemic inflammation and thereby may mirror progression towards multi-organ dysfunction syndrome\(^{33}\). Strikingly, in our patient cohort the likelihood of AKI was significantly reduced in TBI patients compared with the non-TBI control group while non-significantly tending towards elevated PER2 expression. Taking into account work by others concerning nephro-protective qualities of PER2\(^{34}\), along with our findings, it is tempting to speculate that TBI induces PER2 expression, which on the one hand indicates central circadian disruption that contributes to the development of delirium while on the other hand exerting anti-inflammatory organ protection peripherally by preserving kidney function. Fittingly, on a descriptive statistical level and limited to our patient cohort, TBI and PER2 induction showed a trend towards a beneficial impact on survival.

Several limitations of this study have to be noted in relation to the findings discussed above. First, our patient sample size was limited and restricts generalizability of our findings. Second, the ICU as a "polluted" environment in terms of external circadian entrainment factors poses challenges for making conclusions about true endogenous circadian signaling. Third, we performed static analysis of circadian gene expression at two time points that might not reflect true circadian oscillation of these genes.

**Conclusions**

In summary, systemic mRNA expression of PER2 and HO1 is induced after trauma that correlates with trauma severity. Furthermore, the extent of expression correlates with the incidence of delirium in an age-independent manner. TBI in our cohort did not contribute to delirium development. However, it was associated with reduced incidence of AKI, suggesting a link between TBI and secondary organ injury. Utilizing expression of PER2 and HO1 as biomarkers could potentially improve diagnosis algorithms for delirium by adding an objective laboratory parameter.

**Methods**

**Study.** This observational biomarker study focused on TBI in the setting of polytrauma and regulation of PER2 and HO1 expression in leucocytes along with ICU-relevant clinical outcome data (Fig. 1a). In this single-center cohort study, patients were studied under a protocol that was approved by the Institutional Ethics Committee of the University of Freiburg (Protocol No. 293/15). All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki Declaration. Data reporting is done in accordance with the STROBE guidelines. Informed consent from the patient, legal guardian or by proxy was provided. The trial was registered with the German Clinical Trials Register (Trial-ID DRKS00008981; Universal Trial Number U1111-1172-6077; first registration Jul 31, 2015; amendment Jan. 18, 2018).

**Patients and procedures.** Between 13.07.2018 and 06.08.2019, patients who were admitted to the ICU in the Department of Anesthesiology & Critical Care Medicine, University of Freiburg, were screened.

**Inclusion criteria:**

1. Adult patients with polytrauma (ISS > 16) ± TBI.
2. Admission to ICU within 24 h after trauma.
3. Blood sample acquisition possible within 24 h after admission.
4. Possibility to receive informed consent from the patient, legal guardian or by proxy.

**Exclusion criteria:**

1. Pre-existing kidney disease or pre-existing impaired kidney function.
2. Pre-existing diagnosed dementia.
3. Admission later than 24 h after trauma.
4. Death of patient within 24 h after admission.
5. Traumatic or non-traumatic brain injury within 14 days prior to admission.

During the recruitment process, two patient groups (TBI versus non-TBI) were created, limiting differences in gender, age and trauma severity (ISS) using a matched pairs approach. For this purpose, three sub-groups for both age and trauma severity were defined: (1) patients younger than 40 years, (2) patients between 40 and 75 years and (3) patients older than 75 years. These were further grouped by trauma severity into those with an ISS of 24, an ISS of 25 and 36 and an ISS of > 36. Patient groups did not differ in any of the sub-group categorizations (cf. Table 1). TBI was defined as a radiographically diagnosed brain trauma in a routine trauma cranial computer tomogram performed upon hospital admission. Data for clinical assessment was obtained from the digital patient file. Patient mortality was determined from either the digital patient file or via contact with the patients and/or their relatives six months after admission.
**Clinical assessment.** Delirium was assessed by trained ICU staff every 8 h using the nursing delirium screening scale (nuDesc)\(^\text{35}\). In patients with deep sedation for therapeutic reasons, delirium screening was resumed as soon as sedation was terminated. Maximum nuDesc scores during the hospital stay were used to correlate with biomarker expression. To perform between-group comparisons patients were classified into groups of “delirium” and “non-delirium” according to their maximum nuDesc. However, the maximum nuDesc is a static evaluation of delirious symptoms and may not be specific to delirium alone when applied at a single point of time. Therefore, the classification was validated based on a dynamic criterion of delirium that is mentioned in the DSM-5 (fifth edition of the Diagnostic and Statistical Manual of Mental Disorders): fluctuation of delirious symptoms. Symptom fluctuation was assessed through SD (standard deviation) of all daily nuDesc values during patient’s hospital stay. This SD was then defined as the independent variable in a between-group comparison of “delirium” and “non-delirium” via Mann–Whitney-U test. Acute kidney injury was assessed using the first KDIGO (kidney disease improving global outcomes) criterion (0.3 mg/dl serum creatinine increase within 48 h (Stage 1)). The injury severity score (ISS) was obtained from the emergency ward admission sheet that is routinely assessed in polytrauma patients by the admitting staff.

**Biomarkers.** Blood was drawn from arterial or venous catheters during the morning hours on the first day (within 24 h after admission) and one week after trauma (Tempus Blood RNA Tube, AB#4342792). RNA isolation from leucocytes via spin-column purification (Tempus Spin RNA Isolation Kit, AB#1710145) was done as recommended by the manufacturer. Equal amounts of RNA were reversely transcribed into cDNA via reverse transcriptase PCR (iScript CDNA Synthesis Kit, BioRad#1708890; P eqStar 96 Universal Gradient, PeqLab), followed by semi-quantification via real-time PCR (StepOnePlus Real Time PCR-System, A&V Applied Biosystems) with nucleic acid stain (PowerUp SYBR Green Master Mix, AB#1708020).

Primer Sequences were:

| Primer       | Forward Sequence                  | Reverse Sequence                  |
|--------------|-----------------------------------|-----------------------------------|
| PER2 forward | TCCCTCGGCTTGAAACGGC                |                                   |
| PER2 reverse | GAAAGAGCTTCTCGAGCCTCA              |                                   |
| HO1 forward  | GTGATAGAAAGGAGGCAAGACG            |                                   |
| HO1 reverse  | GAATCTTGACATTGTTGCTGG             |                                   |
| Rpl13a forward | CCGACGCTGCGAGGTAT                |                                   |
| Rpl13a reverse | CACCATCGCTTTTCCCTGTC              |                                   |

Gene expression analysis was performed relatively using ΔΔCt-method. Rpl13a served as reference gene as it provides stable expression profiles in various tissues and cell types including peripheral blood and across various experimental conditions\(^\text{36–39}\). Lacking pre-trauma sampling, cDNA samples of five healthy individuals without a history of trauma or known preconditions were recruited in order to obtain normal baseline expression patterns. These pooled technical baseline values were than used to calculated relative changes in mRNA expression in trauma patients.

Furthermore, maximum plasma concentrations of bilirubin (as the final product of heme degradation) from blood gas analyses were obtained from the digital patient file matching the corresponding time of blood withdrawal for gene expression analysis. Plasma bilirubin quantification was done by direct spectrophotometry using a blood gas analyzer (ABL 800 Flex, Radiometer).

**Statistics.** Data analysis was done using Prism 8 software (GraphPad Software Inc.). Based on previous data\(^\text{40}\), an a priori power analysis (contingency tables PER2/HO1 high/low vs. delirium/non-delirium; effect size Cohen’s w 0.6; α = 0.05; power 95%, df 2) indicated a total sample size of n = 43 to be sufficient to characterize biomarker expression levels. Patients for whom data on outcomes were missing were excluded from the study. When performing measurements of central tendency in ordinal-scaled data (PER2 expression, HO1 expression, bilirubin concentration, maximum leucocyte count of first week) the median value is presented along with the corresponding IQR (inter-quartile ratio). In instances of cardinal-scaled data (age, ISS (injury severity score), time on ICU) the mean value is presented with SD. Between-group differences in metrical data (age, ISS, time on ICU, PER2 expression, HO1 expression, bilirubin concentration, maximum leucocyte count of first week) were analyzed via Mann–Whitney-U test. Between-group comparisons in categorical data were performed using Fisher’s exact test (yes/no: pre-existing neurological disease, pre-existing cardiovascular disease, diabetes mellitus, TBI, delirium, AKI, 6-month mortality; high/low: PER2 expression, HO1 expression, bilirubin concentration). Tests for correlation were performed between metrical data sets (ISS, PER2 expression, HO1 expression, bilirubin concentration, time on ICU, age, maximum nuDesc) using Spearman correlation (r, P) as well as linear regression (r², P). Categories of high and low expression levels of PER2 and HO1 were formed from linear regression of ISS and mRNA expression levels of PER2 or HO1 as well as maximum bilirubin concentration on day one after trauma. A linear regression formula of the format Y = a × x + b with the corresponding curve was calculated. Then, data points with mRNA expression/bilirubin concentration located above the regression curve, mathematically corresponding to values higher than expected when using the regression formula for the individual data point, were defined to be of high expression/concentration. Data points with mRNA expression/bilirubin concentration located below the regression curve, mathematically corresponding to values lower than expected using the regression formula, were defined to be of low expression/concentration. Descriptive statistics of relative risk (RR) and odds ratio (OR) were assessed via Koopman asymptotic score (RR) and Baptista-Pike (OR). RR and OR are presented along with their 95% confidence interval (95% CI). With all statistical analyses, a P-value...
of less than 0.05 was considered to be significant. A P-value of less than 0.15 was considered to be indicative of a possible association, and is referred to as "non-significant tendency".

**Data availability**

The data that support the findings of this study are available from the corresponding author upon reasonable request.

Received: 4 December 2020; Accepted: 16 July 2021

Published online: 28 July 2021

**References**

1. Telias, I. & Wilcox, M. E. Sleep and circadian rhythm in critical illness. *Crit. Care* **23**, 82 (2019).
2. Fitzgerald, J. M. et al. Delirium: A disturbance of circadian integrity? *Med. Hypotheses* **81**, 568–576 (2013).
3. Nagasawa, Y., Hasuike, Y., Kuragano, T. & Ishihara, M. Circadian rhythm and CKD: Is melatonin a key player or bi-payer? *Intern. Med.* **58**, 1531–1532 (2019).
4. Olave, O. A., Masten, S. H., Mohandas, R. & Gumz, M. L. Circadian clock genes in diabetic kidney disease (DKD). *Carr. Diabetes Rep.* **19**, 42 (2019).
5. Singbartl, K. & Kellum, J. A. AKI in the ICU: Definition, epidemiology, risk stratification, and outcomes. *Kidney Int.* **81**, 819–825 (2012).
6. Ely, E. W. et al. Delirium as a predictor of mortality in mechanically ventilated patients in the intensive care unit. *JAMA* **291**, 1753–1762 (2004).
7. Inouye, S. K. Nurses’ recognition of delirium and its symptoms: Comparison of nurse and researcher ratings. *Arch. Intern. Med.* **161**, 2467 (2001).
8. Rowley-Conwy, G. Barriers to delirium assessment in the intensive care unit: A literature review. *Intens. Crit. Care Nurs.* **44**, 99–104 (2018).
9. Hall, R. J. et al. CSF biomarkers in delirium: A systematic review: CSF biomarkers in delirium. *Int. J. Geriatr. Psychiatry* **33**, 1479–1500 (2018).
10. Khan, B. A., Zawahiri, M., Campbell, N. L. & Boustani, M. A. Biomarkers for delirium—A review. *J. Am. Geriatr. Soc.* **59**, S256–S261 (2011).
11. Toft, K. et al. Serum biomarkers of delirium in the elderly: A narrative review. *Ann. Intens. Care* **9**, 76 (2019).
12. Kotfis, K., Bott-Olejnik, M., Szylińska, A. & Rotter, I. Could neutrophil-to-lymphocyte ratio (NLR) serve as a potential marker for delirium prediction in patients with acute ischemic stroke? A prospective observational study. *J. Clin. Med.* **8**, 1075 (2019).
13. Colwell, C. S. Linking neural activity and molecular oscillations in the SCN. *Nat. Rev. Neurosci.* **12**, 533–569 (2011).
14. Wiebking, N., Maronde, E. & Rami, A. Increased neuronal injury in clock gene Per-1 deficient-mice after cerebral ischemia. *Carr. Neurovasc. Res.* **10**, 112–125 (2013).
15. Ecke, T. et al. Adora2b-elicited Per2 stabilization promotes a HIF-dependent metabolic switch crucial for myocardial adaptation to ischemia. *Nat. Med.* **18**, 774–782 (2012).
16. Lefa, M., Campbell, K. S., Feng, H.-Z., Jin, J.-P. & Esser, K. A. Development of dilated cardiomyopathy in Bmal1-deficient mice. *Am. J. Physiol. Heart Circ. Physiol.* **303**, H475–H485 (2012).
17. Viswambharan, H. et al. Mutation of the circadian clock gene Per2 alters vascular endothelial function. *Circulation* **115**, 2188–2195 (2007).
18. Gile, J., Scott, B. & Eckle, T. The period 2 enhancer nobiletin as novel therapy in murine models of circadian disruption resembling delirium. *Crit. Care Med.* **46**, e600–608 (2018).
19. Choi, A. M. & Alam, J. Heme oxygenase-1: Function, regulation, and implication of a novel stress-inducible protein in oxidant-induced lung injury. *Am. J. Respir. Cell Mol. Biol.* **15**, 9–19 (1996).
20. Schallner, N. et al. Carbon monoxide preserves circadian rhythm to reduce the severity of subarachnoid hemorrhage in mice. *Stroke* **48**, 2565–2573 (2017).
21. Blydt-Hansen, T. D. Gene transfer-induced local heme oxygenase-1 overexpression protects rat kidney transplants from ischemia/reperfusion injury. *J. Am. Soc. Nephrol.* **14**, 745–754 (2003).
22. Shiraishi, H. et al. Heme oxygenase-1 gene ablation or expression modulates cisplatin-induced renal tubular apoptosis. *Am. J. Physiol.-Ren. Physiol.* **278**, F726–F736 (2000).
23. Agarwal, A., Balla, J., Alam, J., Croatt, A. J. & Nath, K. A. Induction of heme oxygenase in toxic renal injury: A protective role in cisplatin nephrotoxicity in the rat. *Kidney Int.* **48**, 1298–1307 (1995).
24. Shima, H. et al. Protective effect of heme oxygenase induction in ischemic acute renal failure. *Crit. Care Med.* **28**, 809–817 (2000).
25. Tracz, M. J. et al. Renal hemodynamic, inflammatory, and apoptotic responses to lipopolysaccharide in HO-1−/− mice. *Am. J. Pathol.* **170**, 1820–1830 (2007).
26. Bolsethy, S., Zarjou, A. & Agarwal, A. Heme oxygenase 1 as a therapeutic target in acute kidney injury. *Am. J. Kidney Dis.* **69**, 531–545 (2017).
27. For the Traumabase Group. Prevalence and risk factors for acute kidney injury among trauma patients: A multicenter cohort study. *Crit. Care* **22**, 344 (2018).
28. Von Rueden, K. T. et al. Delirium in trauma patients: Prevalence and predictors. *Crit. Care Nurse* **37**, 40–48 (2017).
29. Zaal, I. J., Destin, J. W., Peelen, L. M. & Slooter, A. J. C. A systematic review of risk factors for delirium in the ICU. *Crit. Care Med.* **43**, 40–47 (2015).
30. Leaf, D. E. et al. Does melatonin alter gene expression in hippocampal tissue in aged mice? *J. Neurosci. Res.* **30**, 684–691 (2001).
31. Godin, P. J. & Buchman, T. G. Uncoupling of biological oscillators: A complementary hypothesis concerning the pathogenesis of multiple organ dysfunction syndrome. *Crit. Care Med.* **24**, 1107–1116 (1996).
32. Correa-Costa, M. et al. Carbon monoxide protects the kidney through the central circadian clock and CD39. *Proc. Natl. Acad. Sci.* **115**, E2302–E2310 (2018).
33. Luetz, A. et al. Different assessment tools for intensive care unit delirium: Which score to use? *Crit. Care Med.* **38**, 409–418 (2010).
34. Ho, K. H. & Patrizi, A. Assessment of common housekeeping genes as reference for gene expression studies using RT-qPCR in mouse choroid plexus. *Sci. Rep.* **11**, 3278 (2021).
35. Xiao, J. et al. Identification of reference genes in blood before and after entering the plateau for SYBR green RT-qPCR studies. *Peerj* **5**, e3726 (2017).
38. Bian, Z. et al. RPL13A as a reference gene for normalizing mRNA transcription of ovarian cancer cells with paclitaxel and 10-hydroxycamptothecin treatments. Mol. Med. Rep. 11, 3188–3194 (2015).
39. Curtis, K. M. et al. EF1alpha and RPL13a represent normalization genes suitable for RT-qPCR analysis of bone marrow derived mesenchymal stem cells. BMC Mol. Biol. 11, 61 (2010).
40. Kaiser, S. et al. Neuroprotection after hemorrhagic stroke depends on cerebral heme oxygenase-1. Antioxidants 8, 496 (2019).

Author contributions
Study concept and design: N.S. and H.B. Acquisition of data: M.S., S.K., F.U. and J.K. Statistical analysis: M.S. Analysis and interpretation of data: M.S. and N.S. Drafting of the manuscript: M.S., H.B. and N.S. Critical revision of the manuscript for intellectual content: all authors. Study supervision: N.S. All authors read and approved the final manuscript.

Funding
Open Access funding enabled and organized by Projekt DEAL. German Research Foundation (DFG) Grant to NS: SCHA 1838/4-1. NS is funded by the Berta-Ottenstein-Programme for Advanced Clinician Scientists, Faculty of Medicine, University of Freiburg. The article processing charge is funded by the University of Freiburg in the funding program “Open Access Publishing”.

Competing interests
The authors declare no competing interests.

Additional information
Correspondence and requests for materials should be addressed to N.S.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher’s note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article’s Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

© The Author(s) 2021