Antibacterial and *In vitro* Antioxidant Effect of Dodonaea Viscosa Leaves

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Authors' contributions

This work was carried out in collaboration between both authors. Both authors read and approved the final manuscript.

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ABSTRACT

Free extremists are highly responsive substances linked to the pathophysiology of various infections, such as threatening development and disruption. As a result, there is a need to investigate compounds having anti-free radical properties or cell-supporting properties. The assessment's main goal is to look into the *in vitro* malignant growth avoidance expert development of Dodonaea visciosa's hydro alcholic focus point on various *in vitro* models. The D.viscosa hydroalcoholic concentrate was coordinated and exposed to a targeted phytochemical evaluation. DPPH rummaging, decreasing power, and nitric oxide enthusiast gazing investigation were used to examine D.viscosa's *in vitro* cell support activity. Furthermore, the antibacterial progress of plant extract was evaluated on various microorganisms using agar plate dispersing and agar very much spread techniques. Using measures with IC50 values of 68.42, 36.88, and 100 g/ml independently, hydroalcoholic concentrate of D.viscosa exhibited productive restriction of free moderates in DPPH rummaging, lowering power, and nitric oxide gazing. The whole antibacterial activity against the bacterium was genuinely checked out at various concentrate points. D.viscosa hydroalcoholic concentrate is a probable source of commonplace cell fortifications and serves as an effective free outrageous scrounger.

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1. INTRODUCTION

There is just one unpaired electron in a free extremist's body. In order to form each of the more stable species, they either add or subtract electrons from various particles that are attempting to recombine with their electrons. Ro (Receptive Oxygen Species) are oxygen subcontractors [1] that are continuously delivered in your system by different external components [2] and various metabolic exercises, such as the high-sway breathing. Cell growth and phagocytosis (oxygen burst) are predicted to benefit from a free oxygen species component, with intracellular hailing moving forward as a result. Free progressives produced by sunshine, UV light, ionising radiation, programmed reactions, and the metabolic cycle, on the other hand, have a broad range of over-the-top effects. A proper relationship of guard device in the body takes out or kills receptive oxygen species carried in the living thing. As soon as these free fans or responsive oxygen species approach, they create tissue damage, bio particles, and further, illness conditions, especially degenerative issues like ageing, diabetes, arthritis, carcinogenesis, and cardiovascular pollutions [3-6].

There are fundamental molecules that play a vital part in the oxidative processes induced by free radicals, therefore providing humans with resistance [7]. Cell fortresses are being used for a longer period of time to agree on responsive oxygen species as a consequence of this significant breaking point. A large portion of the harmful development expectation specialists is created misleadingly within a day's time. Several projected cell strongholds, such as butylated hydroxyl anisole (BHA), butylated hydroxyl toluene (BHT), Tertiary butylated hydroxyl quinone (TBHQ), and Gallic horrendous esters, are financially available. When taken in vivo, such intended threatening development neutralisation specialists have been shown to have probable consequences and some amount of sickness-causing nature [8-12]. From here on out, their usage will be limited. Specialized plant-based anti-malignant growth compounds are used to prevent free radicals from causing illness in the present structure. As a result, there has been a significant increase in interest in surveying supporting plants for the existence of brand name cell strongholds.

Pharmacological properties of plant-initiated combinations like as Flavonoids, Terpenes and Alkaloids have lately received much interest due of their antioxidant, antibacterial and anti-singing properties. [13,14]. Dodonaeaviscosa Leaves (Sapindaceous) is an evergreen languishing brier that is most often found in the Western Ghats and Tamilnadu. The Muthuvan gatherings of Kerala used the leaves for the cure of brain desolations and spinal tortures, according to folklore. To alleviate swellings and spinal tortures, a percolating water decoction of leaves is used, and steam inner breath is used to minimise cold. D. viscosa is also used in normal therapeutic practise to relieve stomach pain, stacks, and ulcers. D. viscosa has been shown to have antibacterial, neighbourhood quieting, and smooth muscle relaxing properties in previous studies [15,16]. Similarly, the current evaluation was designed to evaluate the cell support potential of D. viscosa Hydro alcoholic concentrate on several invitro models.

2. MATERIALS AND METHODS

2.1 Plant Material

The leaves of D. viscosa were gathered in Trichy, Tamil Nadu, India, in August 2019. The botanist examined and verified the plant material. The plant components were dried in the dark, then chopped into small pieces, beaten with a mechanical processor, sifted through a 40-cross segment sifter, and in a sealed container for future use.

2.2 Extraction of Plant Material

Hydro alcohol was used to remove the powdered leaves of D. viscosa from the leaves at room temperature. The dissolvable was collected and separated after complete extraction. Under lower temperatures of 50-55°C, the dissolvable was locked in. Desiccators were used to store the concentrated hydro alcohol disengages for later use.

2.3 Qualitative Phytochemical Analysis

The existence of several phytoconstituents in the severe hydro alcoholic concentrate of D. viscosa leaves was tested by conventional phytochemical shows. Alkaloids (Dragendorff reagent, Mayer's reagent, Hager's reagent, and Wagner's reagent), flavonoids (Shinoda-Paw test), steroids
(Liberman Burchard test and Salkowski’s reaction), terpenes (Vanillin sulfuric damaging reagent), and carbohydrates (Fehling’s test and Molisch test) were all investigated.

2.4 In vitro Antioxidant Activity

2.4.1 In vitro antioxidant activity by DPPH assay

The approach given by Braca et al., 2001 was used to examine the scanning development for DPPH free devotees. An aliquot of 3 ml of ethanol containing 0.004 percent DPPH strategy and 0.1 ml of plant eliminate was combined with varied concentrations of plant eliminate. After thoroughly mixing the concoction, it was allowed to sit at room temperature for 30 minutes before being discarded. It was necessary to decolorize DPPH in order to measure its absorbance at 517 nm. 0.1 ml of an individual vehicle was used as a control at the plant elimination/ascorbic disaster site. Not exactly \([A0 - A1] / A0\times100\), where A0 was the control absorbance and A1 was the plant remove/ascorbic destructive absorbance, the rate counteract.

2.5 Lessening Power Assay

Soluble potassium ferricyanide (2.5 ml) was mixed with several plant mixtures in various solvents (2.5 ml). For 20 minutes, this mix was held at 50°C in a water shower. After cooling and centrifuging at 3000 rpm for 10 minutes, 2.5 ml of 10% trichloro acidic disaster was added. Refined water and a delayed ferric chloride game plan (0.5 ml) were added to the blueprint's top layer (2.5 ml). At 700 nm, the absorbance was measured. With the exception of testing, control was set up in the same manner. Supplement E was used as a control at various fixations. The response blend's expanded absorbance indicates an increase in declining power.

2.6 NITRIC Oxide Radical Scavenging Assay

Sodium nitroprusside (5 mm) in standard phosphate cushion saline (0.025 M, pH 7.4) was used to centralise various concentrations of concentrates, and chambers were heated to 29°C for three hours. Control tests were conducted without the test drugs but with the same level of help. It took 1 ml of Griess reagent to knock out the hatching models after three hours. After diazotization with sulphanilamide and subsequent coupling with naphthyl ethylenediamine hydrochloride, the absorbance of the covering produced during this process was measured at 550 nm using a spectrophotometer. In an uncommonly common test, ascorbic damage produced a similar structure [8].

2.7 Bacterial Cultures

The Microbial Type Culture Collection in Chandigarh, India, provided the bacterial social hierarchies. Microorganisms such as Staphylococcus aureus (MTCC 3160), Bacillus subtilis, Escherichia coli (MTCC40), Klebsiella pneumonia (MTCC3384), P. aeruginosa (MTCC741), and Proteus mirabilis (MTCC741) Staphylococcus aureus (MTCC3384), Pseudomonas aeruginosa (MTCC741) (MTCC425) are included in this list. Each bacterial social order was maintained alive in supplement agar and stored at 4 degrees Celsius.

2.8 Arrangement of Inoculum

Sterilized peptone water from the sub-refined creature was used to transfer many villages (5 ml). To guarantee homogeneity, the suspensions were stirred for 15 seconds, although this did not result in turbidity matching the 0.5 McFarland standard (OD = 0.12-0.15 at k = 530 nm, corresponding with 1-5 x 106 CFU/ml). Agar plate dispersing framework and agar well dissipating approach were used to execute the antibacterial measure. Mueller Hinton agar (MHA) was used as a plate-based medium for evaluating microscopic live animals. Using a cleaned q-tip, the bacterial inoculum was dispersed pretty evenly throughout the MHA plates.

For the agar plate scattering technique, sterile channel paper circles (6mm in diameter) were submerged in different mixtures of the test substance, allowed to dry, and then placed on top of the prepared agar plate. With the use of a fitting drill, an in general was built up in the plates for the agar well dissipating procedure (0.6cm). The well was filled with 100 litres of the test substance. At 37°C, the plates were brooded till further notice. Controls were preserved for each bacterial strain, where pure solvents were used instead of the concentrate. As a terrible control, clean purified water was used. By calculating the zone distance across, the result was obtained. The evaluation was greatly improved, and the average attributes were incorporated.
3. RESULTS

It was found that D. viscosa had a breaking point of 66.42 g/ml when hydrogen was activated in solid form. Table 1 shows the findings of the experiment. With an IC50 of 32.88 g/ml, D. viscosa induced solid nitric oxide glancing through advancement. The results were shown in Table 2. The declining force of D. viscosa increases with an increase in the focus in this study. At a concentration of 100 g/ml, the D. viscosa displayed the greatest over the top progress. The antibacterial enhancement of the plant removes was evaluated using an agar plate spread approach and an agar well dissipating framework in this study. The samples collected at various concentrations showed promising antibacterial activity against the microorganism used in the study. Tables 4 and 5 summarised the results.

Table 1. *In vitro* DPPH extremist rummaging impact of *D. viscosa*

| Concentration (µg/ml) | Vitamin C | *D. viscosa* |
|----------------------|-----------|--------------|
| 10                   | 9.7±0.98  | 8.76±0.65    |
| 20                   | 18.9±0.92 | 18.76±0.80   |
| 40                   | 33.4±0.85 | 32.86±0.69   |
| 80                   | 67.7±0.92 | 65.54±1.17   |
| 100                  | 84.2±1.02 | 78.54±1.29   |
| **IC50 (µg/ml)**     | 61.47     | 68.42        |

Table 2. *In vitro* nitric oxide rummaging impact of *D. viscosa*

| Concentration (µg/ml) | Vitamin C | *D. viscosa* |
|----------------------|-----------|--------------|
| 10                   | 40.96±0.58| 32.56±0.42   |
| 20                   | 51.46±0.76| 44.05±0.62   |
| 40                   | 63.12±0.87| 60.76±0.78   |
| 80                   | 82.14±0.92| 78.76±0.87   |
| 100                  | 95.56±0.95| 86.21±1.21   |
| **IC50 (µg/ml)**     | 12.54     | 36.88        |

Table 3. *In vitro* lessening power capacity of *D. viscosa* extract

| Concentration (µg/ml) | Vitamin E | *D. viscosa* |
|----------------------|-----------|--------------|
| 10                   | 0.40±0.02 | 0.24±0.01    |
| 20                   | 0.50±0.05 | 0.35±0.04    |
| 40                   | 0.68±0.07 | 0.72±0.08    |
| 80                   | 1.02±0.08 | 1.0±0.07     |
| 100                  | 1.78±0.07 | 1.72±0.1     |

Table 4. Antibacterial movement of *D. viscosa* extract by plate dispersion technique

| Microorganism       | 20    | 40    | 80    | 100   |
|---------------------|-------|-------|-------|-------|
| S. aureus           | 24    | 26    | 29    | 33    |
| B. subtilis         | 16    | 19    | 20    | 29    |
| E. coli             | 20    | 23    | 27    | 35    |
| K. pneumonia        | 15    | 19    | 22    | 27    |
| P. aeruginosa       | 12    | 14    | 19    | 25    |
| P. mirabilis        | 11    | 12    | 17    | 24    |
Table 5. Antibacterial action of *D. viscosa* remove by well dissemination strategy

| Microorganism       | *D. viscosa* extract concentration (µg/ml) |
|---------------------|-------------------------------------------|
|                     | 20 | 40 | 80 | 100 |
| S. aureus           | 22 | 25 | 29 | 34 |
| B. subtilis         | 16 | 19 | 25 | 29 |
| E. coli             | 18 | 23 | 25 | 33 |
| K. pneumonia        | 13 | 16 | 18 | 23 |
| P. aeruginosa       | 14 | 19 | 21 | 25 |
| P. mirabilis        | 11 | 13 | 16 | 21 |

4. DISCUSSION

Some masochistic occurrences, including as tissue obliteration, malignant growths, aggravating, and neurodegenerative illnesses [2] are caused by free fans. When cells and organs are able to regulate oxidative strain, the dangerous growth balancing expert capacity of tolerating plants and their phytoconstituents, which stimulates free preposterous covering potential, is legitimate [3,4].

An excellent DPPH enthusiast is often employed to examine cell support and to declare important hazardous development anticipated expert potential. Phosphorylation of DPPH results in proton free revolutionary with clear support area when exposed to proton free inhibitors over the top. The cell support chemicals' absorbance drops at 517nm as a result of the DPPH reaction. The DPPH impediment measure is based on the potential of steady DPPH decolorization moderated by cell fortress activity. The difference in responding DPPH extremist to 1, 1-diphenyl-2-picryl hydrazine within illness counteraction experts is the reaction.

The DPPH fanatic examines the capability of cell fortifications from a broad viewpoint, taking into account their ability to produce hydrogen. Furthermore, the accommodating plants' phone support cutoff has a favourable association with the social event of the phenolic chemical contained in the concentrates. By giving the hydrogen atom, *D. viscosa* shown efficient blocking of DPPH free enthusiast in the force evaluation. It then demonstrates the interaction between plant concentrates and moderate age, which stimulate mild appearance.

As the Fe3+ ferricyanide complex is broken down, electrons are transferred to the ferrous (Fe2+) particles and a blue green concealment is formed at 700nm. This concealment shows the decreasing far reaches of the dangerous development balance experts and thus shapes the blue green concealment in the response mix. Research on the role of *D. viscosa* in ferric particle reduction is ongoing. Free crazy chain ending by giving hydrogen molecules, which additionally responds to peroxide proclaims and diminishes the peroxidation collaboration, results in the lowering limit of plant boundaries as a consequence of the nature of lowers and their properties. [5].

Macrophages, endothelial cells, and the cerebrum all rely on nitric oxide (NO) to maintain regular cycles. In a number of masochist conditions, the levels of NO are increased. A five-carbon metabolic oxidative pathway is used to convert arginine to citrulline and NO in normal tissues, regulating the age of NO in these tissues. [7].

Organs are affected by the NO impacts that are supplied to them and their essential and significant limitations are affected. As a measure of NO-seeking potential, free ridiculous scroungers reduce the absorbance of 550 nm. The difference in NO absorbance is measured to determine the cell support practicality of plants dispensed with by NO rummaging. When nitric oxide reacts with oxygen or superoxide, it produces NO2, N2O4, N3O4, NO3, and NO2, which are all staggeringly open. The organs are put under oxidative stress and underhandedness by these response revolutionaries [8].

Because of its cell support capability, the *D. viscosa* dispense with exhibited convincing nitric oxide seeking in the continuing study. The occurrence of flavonoids such as quercetin, groove in, and myricetin is substantially a direct result of *D. viscosa*’s illness evasion expert growth. Gram positive microorganisms are weaker against plant kills than Gram-negative living beings, according to mounting research [17,18]. The present scenario is a direct result of Gram-positive microbes having a single cell mass and Gram-negative germs having a perplexing cell divider [19]. Previous studies
have shown that using a bio autography framework, D.viscosa may increase its antibacterial properties [20].

5. CONCLUSION

The continuous evaluation ensures that the D.viscosa plant's cell support limit is eliminated in various inviro models. Furthermore, phytochemical and disconnection examinations are often used to identify phytoconstituents that are at danger for cell support advancement. Furthermore, the evaluation reveals the efficacy of D.viscosa kill against pathogenic animals that cause certain human contaminations.

CONSENT

It is not applicable.

ETHICAL APPROVAL

It is not applicable.

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COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Fresquet F, Pourageaud F, Leblais V, Brandes RP, Savineau JP, et al. Role of reactive oxygen species and gp91phox in endothelial dysfunction of pulmonary arteries induced by chronic hypoxia. Brit J Pharmacol. 2006;148:714-723.
2. Kikuzaki H, Nakatani N. Antioxidant effects of some ginger constituents. Journal of Food Science. 1993;58:1407-1410.
3. Hallwell B, Gutteridge JMC. Free radicals in biology and medicine. 3rd edn. Oxford University Press, Oxford, UK. 1999;543.
4. Finkel J, Holbrook NJ. Oxidants, oxidative stress and the biology of aging. Nature. 2000;408:239-247.
5. Das M, Mukherjee SB, Shaha C. Hydrogen peroxide induces apoptosis like death in Leishmania donovani promastigotes. J Cell Sci. 2001;114:2461-2469.
6. Naskar K, Guha bakshi DN. Vegetarian pattern of the sundarbans n mangrove swamps of the sundarbans. An ecological perspective. Naya proksah: Calcutta. India. 1995;27.
7. Villoglu YS, Mazza G, Gao L, Oomah BD. Antioxidant activity and total phenolics in selected fruits, vegetables, and grain products. J Agri food chem. 1998;46:4113-4117.
8. Barlow SM. Toxicology aspects of antioxidants used as food additives. In Hudson BJF (Ed), Food antioxidants. Elsevier, USA. 1990;253-307.
9. Branen AL. Toxicology and Biochemistry of butylated hydroxyanisole and butylated hydroxyl toluene. Journal of American Oil Chemical Society. 1975;52:59-63.
10. Chan HWS. Autioxidation of unsaturated lipids. Academic press, London. 1987;296.
11. Namiki M. Antioxidants/antimutagens in food. Critical Reviews in Food science and Nutrition. 1990;29:273-300.
12. Prkorny J. Natural Antioxidants for food use. Trends in food science Technology. 1991;9:223-227.
13. Sahu SC, Dhal NK, Mohanty RC. “Potential Medicinal plants used by the Tribal of Deogarh district, Orissa, India” studies on Ethano. Medicine. 2010;4:53-61.
14. Newman DJ, Cragg GM. Natural products as source of new drugs over the last 25 years. Journal of Natural Products. 2007;70:461-477.
15. Khalil NM, Sperotto JS, Manfron MP. Antinflammatory activity and acute toxicity of Dodonaeaviscosa. Fitoterapia. 2006;77(6):478-480.
16. Rojas A, Cruz S, Ponce-Monter H, Mata R. Smooth muscle relaxing compounds from Dodonaeaviscosa. Planta Medica. 1996;62(2):154–159.
17. Lin J, Opoku AR, Geheeb-Keller M Hutching AD, Terblanche SE, Jagar AK, van Staden J. Preliminary screening of some traditional zulu medicinal plants for anti-inflammatory and antimicrobial activities. Journal of Ethnopharmacology. 1999;68:267-274.
18. Parekh J, Chanda S. In vitro antimicrobial activities of extract of Launnaea procumbent Rob. (Labiateae), Vitis vinifera
(Vitaceae) and *Cyperus rotundus*. 20. Khurram M, Khan MA, Hameed A, (Cyperaceae). African Journal of Biomedical Research. 2006;9:89-93. Antibacterial activities of *Dodonaea viscosa* using contact bioautography technique. Molecules. 2009;14(3): 1332-41.

19. Yao J, Moellering R. Antibacterial agents. manual of clinical microbiology. ASM Wahington DC. 1995;1281-129.