INTRODUCTION

Phytochemicals are the bioactive plant compounds present in functional foods like fruits, vegetables, grains etc. These bioactive compounds not just promote good health but play a major role in prevention of chronic diseases [1]. These bioactive plant compounds which are also known as secondary plant metabolites, possess various benefits including antibacterial properties against pathogenic and spoilage microbes [2]. The increasing antibiotic resistance, limited spectrum and multiple side effects with synthetic agents at an alarming rate, prevails the natural antimicrobials. Unlike the synthetic antibiotics, natural antimicrobials don’t target a particular reaction rather act on various groups of biomolecules in a pathogen thus there is no occurrence of resistance which makes them more alluring candidate [3]. However these bioactive compounds or secondary metabolites are limited in their efficacy because of poor absorption with limited solubility in lipophilic phases and instability at different physiological pH values [4]. The antibacterial efficacy of these secondary metabolites in plants depends on the chemical structure of active components and their concentration. Various phytochemicals present in plants with antibacterial effect includes saponin, flavonoids, thioureas, glucosinolates, phenolics, and organic acids. Among them, Chlorogenic acid (CGA), a polyphenolic derivative of caffeic acid and quinic acid holds as a promising physiologically active substance present in foods such as coffee beans, tea and different fruits like apples, grapes, plums etc. [5]. CGA has been reported to have antimicrobial activity against gram negative as well as gram positive bacteria [6]. The antimicrobial activity of polyphenolic derivatives has been imputed by their structure, their pH and sodium chloride concentration which results in physiological changes in pathogens and consequent cell death [7]. It also possesses other activities such as chemopreventive, antimutagenic, antioxidative, hypoglycemic, antiviral and anti-inflammatory. Its properties are attributed to the phenolic hydroxyl groups and it is characterized by relatively low toxicity and side effects. Till date there have been several literatures on antimicrobial efficiency of CGA against various strains having mechanism of disrupting the cell membranes of microbes [6] and inhibiting the bacterial fatty acid synthesis [8]. It has also been studied against various resistant strains as a synergistic agent along with standard antibiotics [9, 4]. Despite its safety and effectiveness the use of CGA is limited by its low bioavailability [10]. Low bioavailability is attributed to extensive metabolism, thus despite of having a good hydrophilicity (40 mg/ml), it has low bioavailability (17-24%) and permeability (Biopharmaceutical classification system class III) which in turn affects its biological activities [11]. Moreover, CGA undergoes oxidation during storage or processing [12]. Complexation with phospholipid has not only shown to increase bioavailability by limiting the metabolism and increase in lipophilic solubility, but will also form stable phytovesicular form of CGA because of complexation [13]. Also CGA possesses polar hydroxyl groups in its structure which favours binding with phospholipid to form a phytovesicular complex [14].

Present work is thus aimed to develop and optimize phytovesicular complexes of CGA to improve its bioavailability, permeability and stability which should in turn enhance its antibacterial efficiency against these gram positive and gram negative bacteria. This enhanced activity can act as a source of an antibiotic alternative for drug development against array of infections. Four different bacterial strains Staphylococcus aureus, Bacillus subtilis, Klebsiella pneumonia and Escherichia coli were selected which causes various infections like soft tissue infections, joint infections, Urinary tract infections, nosocomial infections etc. Antibacterial efficiency against them was evaluated using agar diffusion assay, measuring minimum inhibitory concentration (MIC), time kill assay. Further the role of pH and temperature on antibacterial was also evaluated and compared against plain CGA.

MATERIALS AND METHODS

Materials
CGA was purchased from Chemsworth, Surat-394230, India. Dimethyl sulfoxide (DMSO), Dichloromethane, N-hexane were
obtained from Merck Pvt. Ltd. India. Various bacterial growth mediums like Mueller-Hinton agar (MHA), Mueller-Hinton Broth(MHB) were purchased from Himedia Lab. Pvt. Ltd. Phospholipon 90H was obtained as a gratis sample from Lipoid, Germany. All the other chemicals were of analytical grade.

**Bacterial stains**

Test organisms used were obtained from National collection of industrial microorganisms (NCIM), Pune. Two gram positive strains [Staphylococcus aureus (NCIM 2079) and Bacillus subtilis (NCIM 2063)] and two gram negative strains [Klebsiella pneumonia (NCIM 2036) and Escherichia coli (NCIM 2345)] were selected. The microorganisms were maintained and stored at 4 °C.

**Culture conditions**

Organisms obtained were sub cultured in nutrient broth and nutrient agar. A loopful of organisms were transferred from the cultures obtained, in a freshly prepared nutrient broth suspension in an aseptic environment. Similarly a loopful of culture was streaked on an agar slant and incubated at 37 °C for 24 h. The sub culturing was done every two months.

**Standard antibiotic**

Gentamycin (GEN) was used as a standard antibiotic to compare the antibacterial activity of CGA. Gentamycin (Gentabal) of Labonite pharmaceuticals India Ltd. was obtained from a local pharmacy shop.

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### Table 1: Coded levels and actual values for optimization of CGA phytovesicle using central composite design

| Type of variables | Variables | Levels for optimization |
|-------------------|-----------|-------------------------|
| Independent       | X1 (Drug: Lipid ratio) | -1, 0, +1 |
|                   | X2 (Reaction time)     | 1h, 2h, 3h |
|                   | X3 (Reaction temperature) | 40 °C, 50 °C, 60 °C |
| Dependent         | Y1 (Partition Coefficient) | Maximum |
|                   | Y2 (Entrapment efficiency) | Maximum |

**Analytical method of analyzing CGA**

Estimation of CGA content in the complex was carried out by UV spectrophotometer (Jasco (V-630), Japan.) Also Calibration of CGA was performed in methanol, ultrapure water and phosphate buffer saline pH 6.8 (PBS). A standard stock solution of 1000 ppm was prepared, and from this, a 10 ppm solution was prepared to determine the λmax and calibration curves. Various validation parameters like linearity, intraday precision study, interday accuracy study, robustness, ruggedness were determined in accordance with the ICH guidelines [16].

**Entrapment efficiency and partition coefficient**

Phytovesicles equivalent to 10 mg of drug were accurately weighed in 100 ml beaker to which 50 ml of 6.8 pH phosphate buffer was added. The beaker was then subjected to stirring on a magnetic stirrer for 4 h and then allowed to stand for 1h. Clear supernatant was decanted while the sludge was subjected to centrifugation at 5000 rpm for 15 min. The supernatant obtained after centrifugation was filtered with Whatman filter paper (0.45 µm) and was measured for UV absorbance at 328.8 nm. Entrapment efficiency was calculated by following formula Eq. 2.0 [17]

\[ \text{Entrapment efficiency} = \frac{\text{Amount of drug in complex} - \text{Amount of free drug obtained} + \text{Amount of drug taken}}{\text{Total amount of drug taken}} \times 100 \]  

Solubility of CGA and CGA phytovesicles in both oil and water phase was determined by calculating partition coefficient using shake flask method with water and n-octanol. 10 mg equivalent of phytovesicles were taken in a conical flask to which 10 ml of water and n-octanol was added and then placed upon rotary shaker for 24 h. The mixture was then transferred to a separating funnel and was allowed to stand for 30 min. The amount of drug in both phases was determined by UV spectrophotometer in triplicate. Same procedure was carried out for the plain drug. Concentration in both the phases was calculated using calibration curve equation and partition coefficient was calculated by using equation 3.0 [13]

\[ \text{Partition coefficient} = \frac{C_o}{C_w} \]  

Where Co-concentration in oil phase, Cw-concentration in water phase.

**Characterization of phytovesicles**

**Fourier transform infra-red (FT-IR) spectroscopy**

The IR spectra of the samples (CGA, Phospholipon 90H, phytovesicles) were recorded using a Fourier transform infrared spectrophotometer (Shimadzu-IRAFFINITY-1, Japan) in the region of 4000 cm\(^{-1}\) to 400 cm\(^{-1}\). Pellets were prepared by triturating sample with potassium bromide using glass pestle and mortar in the ratio 1:100. The mixture was then introduced into the sample holder and scanned. The interaction between CGA and Phospholipon 90H was studied with the spectra obtained [18].

**Differential scanning calorimetry (DSC)**

Thermograms of CGA, Phospholipon 90H, Phytovesicles and their physical mixture were recorded using a differential scanning calorimeter (DSC 60 PLUS, Shimadzu, Japan). The thermal behaviour was studied by heating each individual sample in aluminium seal sample pan under nitrogen gas flow at the heating rate of 5 °C/min from 0 to 300 °C [19].

**Particle size analysis and zeta potential**

The particle size, polydispersity index and zeta potential of phytovesicles were determined by a Zetasizer Nano ZS90 (Malvern Instruments Ltd., Massachusetts, USA). For measurement phytovesicles (10 mg) were dispersed in 10 ml deionized water and measured [18].

Formulation of CGA phytovesicles by central composite design

Central composite design was used to study the influence of different process and formulation variables such as drug-phospholipid ratio (X1), reaction time (X2) and reaction temperature(X3) on the quality attributes of phytovesicular complex i.e. partition coefficient and entrapment efficiency (table 1). The effect of these three independent variables at three different levels was evaluated by carrying out trials for 20 different combinations. The data obtained was statically analyzed and validated by Design expert® software (Version 11.0. Stat-Ease Inc., Minneapolis, MN). The design constructed gave the following polynomial Eq. 1.0.

\[ Y = b_0 + b_1X_1 + b_2X_2 + b_3X_3 + b_11X_1^2 + b_22X_2^2 + b_33X_3^2 + b_{12}X_1X_2 + b_{13}X_1X_3 + b_{23}X_2X_3 \]  

Where Y is the response for all the 20 combinations, b, is the intercept while b1 to b33 are coefficients to the variables X1, X2, X3 (main factors). The interaction terms (X1X2, X2X3, and X1X3) showed concurrent changes in response as the factors change. While the linearity was measured by polynomial terms (X1, X2, X3). Briefly, CGA and Phospholipon 90H were refluxed with 20 ml of dichloromethane at different molar ratios, reaction times and temperatures. The volume was then reduced to 3-5 ml by continuously stirring and to it 10 ml of n-hexane was added to precipitate the complex. n-hexane was allowed to evaporate at room temperature for 24-48 h. Percentage yield was calculated and the dried complex was further stored in a glass bottle at room temperature [15].
**Scanning electron microscopy (SEM)**

SEM imaging of CGA phytovesicles, Phospholipon 90H and CGA was performed by a scanning electron microscope (JEDL, JSM—6390LV, Tokyo, Japan). The sample was mounted on specimen stubs using carbon tape and was coated over with gold using ion sputtering device (JEOL 1600). The measurements were performed at 20kV using different magnifications [19].

**Anti-microbial activity**

**Agar diffusion assay**

Antibacterial activity of CGA and CGA phytovesicles against Staphylococcus aureus, Bacillus subtilis, Klebsiella pneumonia and Escherichia coli was determined by agar diffusion assay. All the bacterial isolates were grown in nutrient broth and standardized in saline solution to a turbidity of 0.5 McFarland standards. Using sterile cotton swabs the bacterial inoculum (50 µl) were then spread onto MHA plates by lawn culture. Wells were created onto the MHA plates using sterilized cork borer (6 mm). Test solutions (100 µl) prepared by dissolving the CGA and CGA phytovesicles in DMSO at a concentration of 2, 4, 8, 16, 32 mg/ml were then carefully loaded into the wells. Gentamycin (GEN) was used as a standard while the lowest concentration showing no visible growth. All measurements were performed in triplicates [20].

**Minimum inhibitory concentration (MIC)**

Minimum inhibitory concentration of CGA and CGA phytovesicles for Staphylococcus aureus, Bacillus subtilis, Klebsiella pneumonia and Escherichia coli was determined by broth dilution method. Stock solutions of CGA and CGA phytovesicles were prepared at 1000 µg/ml. 11 sterile test tubes were labelled and arranged. Tubes 2-11 were loaded with 500 µl of MHB. While to tubes 1 and 2, 1000 µl of CGA stock solution was added. Two fold serial dilution was then performed by pipetting out 500 µl of the mixture from test tube 2 to 3 and consecutive transferring mixture till tube 9. 500 µl of mixture from tube 9 was discarded. Tube 10 and tube 11 served as growth control (with no antibiotic) and broth control respectively. All test tubes were inoculated except the 11th test tube with 500 µl of bacterial inoculum (0.5 McFarland standard). The tubes were then further incubated at 37 °C for 24 h and observed for growth and inhibition zones around the wells which were then further compared with antibiotic. All measurements were performed in triplicates [20].

**Effect of pH on antibacterial activity**

A change in the antibacterial activity with changes in pH was determined. Test solutions of CGA and CGA phytovesicles, at the concentration equivalent to MIC, were prepared in pH 4 sodium acetate buffer, pH 7 and pH 10 phosphate buffer. To the sterile test tube 1000 µl of test sample, 1000 µl of MHB and 500 µl of bacterial inoculum (for all the 4 strains Staphylococcus aureus, Bacillus subtilis, Klebsiella pneumonia and Escherichia coli) was added and further incubated in rotary shaker at 37 °C for 3h. 10 fold dilutions were made before plating the sample from each test tube onto a MHA plate. After incubation of plates at 37 °C for 24h the CFUs were counted. All measurements were performed in triplicates [20].

**Table 2: Optimization of CGA phytovesicular complex using central composite design**

| Batch number | X₁ drug: lipid ratio | X₂ reaction time | X₃ reaction temperature | Y₁ partition coefficient | Y₂ entrapment efficiency |
|--------------|----------------------|------------------|-------------------------|--------------------------|--------------------------|
| 1            | 3                    | 2                | 33.1821                 | 0.653±0.07               | 73.72±1.34               |
| 2            | 3                    | 4                | 33.1821                 | 0.662±0.01               | 74.47±0.87               |
| 3            | 3                    | 3                | 33.1821                 | 0.835±0.19               | 66.13±1.12               |
| 4            | 2                    | 3                | 33.1821                 | 0.778±0.24               | 68.5±1.34                |
| 5            | 2                    | 2                | 33.1821                 | 0.954±0.21               | 69.02±0.99               |
| 6            | 2                    | 2                | 33.1821                 | 0.841±0.28               | 75.26±1.13               |
| 7            | 3                    | 1                | 33.1821                 | 0.977±0.36               | 96.89±1.24               |
| 8            | 1                    | 3                | 33.1821                 | 0.354±0.26               | 84.61±1.09               |
| 9            | 2                    | 2                | 33.1821                 | 0.798±0.32               | 83.44±0.91               |
| 10           | 0.318207             | 2                | 33.1821                 | 0.502±0.11               | 71.6±0.67                |
| 11           | 2                    | 2                | 66.1879                 | 0.435±0.39               | 75.4±0.35                |
| 12           | 2                    | 2                | 50                      | 1.036±0.20               | 81.4±0.78                |
| 13           | 3                    | 3                | 50                      | 1.546±0.29               | 81.89±1.4                |
| 14           | 2                    | 3                | 50                      | 1.693±0.17               | 64.6±0.56                |
| 15           | 2                    | 2                | 50                      | 0.719±0.04               | 78.45±1.64               |
| 16           | 2                    | 0.318207         | 50                      | 0.922±0.43               | 72.27±0.85               |
| 17           | 2                    | 2                | 50                      | 0.687±0.23               | 80.5±1.76                |
| 18           | 2                    | 2                | 50                      | 0.735±0.22               | 77.25±0.93               |
| 19           | 2                    | 2                | 50                      | 0.592±0.44               | 82.47±1.45               |
| 20           | 1                    | 1                | 60                      | 0.492±0.27               | 69.6±0.27                |

Values were expressed as mean±SD (n=3).
RESULTS AND DISCUSSION

Optimization of CGA phytovesicles by central composite design

CGA phytovesicles were optimized by determining the effect of three different variables-drug-phospholipid ratio (X₁), reaction time (X₂) and reaction temperature (X₃) which greatly influenced partition coefficient (Y₁) and entrapment efficiency (Y₂) of CGA phytovesicles. A higher value of entrapment efficiency and partition coefficient depicted better complex formation between phospholipid and CGA with increased lipophilic solubility respectively. The results are summed up in Table 2. The values measured for entrapment efficiency ranged from 64.6-96.89% while the values of partition coefficient ranged from 0.354 to 1.036. Batch [F7] showed highest entrapment of 96.89% and partition coefficient value of 0.977 as compared to 0.036 of plain CGA and thus it was further selected as the optimized batch. The regression equations in terms of coded factors (Eq. 4 and 5) obtained for-partition coefficient gave correlation coefficient of 0.7338 exhibiting a good fit to quadratic model while for entrapment efficiency it gave correlation coefficient of 0.6703 with good fit of 2FI model.

\[
Y_1 = 0.8279 + 0.0802 X_1 - 0.0060 X_2 - 0.0215 X_3 - 0.1092 X_1 X_2 - 0.0335 X_1 X_3 - 0.0733 X_2 X_3 - 0.0943 X_1^2 + 0.0163 X_2^2 - 0.0938 X_3^2 \quad \text{... Eq. 4.0} \]

\[
Y_2 = 76.34 + 2.76 X_1 - 1.01 X_2 + 1.52 X_3 - 0.0738 X_1 X_2 + 8.40 X_1 X_3 - 0.7538 X_2 X_3 \quad \text{...... Eq. 5.0} \]

\(r^2=0.7595; p=0.0480\)

Statistically significant coefficients with p value of 0.0480 for Y₁ and 0.0120 for Y₂ were obtained for both the responses. Regression analysis showed that drug-phospholipid ratio (X₁) had a direct relationship and reaction time (X₂) with reaction temperature (X₃) had an inverse relationship on partition coefficient (Y₁). Indicating that partition coefficient values increased with increasing drug phospholipid ratio and decreased with increasing reaction time and temperature. This is because increase in drug phospholipid ratio causes higher quantity of phospholipid to remain in an uncomplexed form forming additional layers encircling CGA phytovesicles rendering good lipophilic character to it [21]. While with increasing reaction time and temperature the phytovesicles formed were sticky and less free flowing owing to the phase transitioning of phospholipid at higher temperature resulting into less solubility.

On the other hand for entrapment efficiency (Y₂), drug-phospholipid ratio (X₁) and reaction temperature(X₃) had direct relationship while reaction time(X₂) having an inverse relationship. It was found that highest reaction time of 3.6 h gave entrapment of 83.44% while at 1h it gave entrapment of 96.86%. This may be due dissociation in hydrogen bonding between CGA phenolic group and choline group of Phospholipon 90H with longer reaction times. Thus it shows the appropriate time required for CGA to complex with that of phospholipid while the increase in entrapment efficiency with increasing drug-phospholipid ratio and reaction temperature can be attributed to increased phospholipid content at required transition temperature which favors better entrapment of CGA complex [22].

Fig. 1: Contour plots of CGA phytovesicles (a) Partition coefficient (b) Entrapment efficiency

Fig. 2: 3D surface plots of CGA phytovesicles (a) Partition coefficient (b) Entrapment efficiency
Similar relationship was seen in contour plots and 3D response curves (fig. 1 and 2 respectively). Depending on these observations optimal values of the variables were drawn i.e. 1:3, 1h, 60 °C respectively. Batch F7 was thus selected as the optimized batch showing highest entrapment with highest partition coefficient where the drug CGA and phospholipid were in the molar ratio of 1:3 thus having better prospects to complex sufficiently with optimum reaction time of 1h with 60 °C of reaction temperature.

The models obtained for both the dependent variables were further validated by preparing an additional batch as per the optimal values of independent variables-1:3, 1h, 60 °C. The predicted values for partition coefficient and entrapment efficiency of CGA phytovesicles from quadratic and 2FI model (respectively) and the actual values from the prepared batch were compared. The partition coefficient and entrapment efficiency of the prepared batch as per optimized conditions were found to be 0.9554 and 93.65±0.25% respectively as compared to 0.9265 and 90.85±0.02 % of the predicted values given by model showing its practicability. Bias was calculated using the Eq. 6.0 was found (-3.1%) for partition coefficient and (-3.08%) for entrapment efficiency demonstrating the robustness of the generated models [23].

Validation of analytical method

Validation of the UV method was performed as per the guidelines approved by ICH. A linear relationship was found between concentrations (2 -22µg/ml) for all the three; methanol, ultrapure water and pH 6.8 PBS with correlation coefficient (r ²) of 0.9989, 0.9947, 0.9995 respectively. The λmax was obtained at 328.2 nm for methanol and 6.8 pH PBS whereas 323.4 nm for ultrapure water. Accuracy of the analytical method was determined using recovery assay methods where the sample solutions were spiked with analyte at three different concentrations (18, 20, 22µg/ml) for all the three; methanol, ultrapure water and pH 6.8 PBS whereas 323.4 nm for ultrapure water. Accuracy of the analytical method was determined using recovery assay methods where the sample solutions were spiked with analyte at three different concentrations (18, 20, 22µg/ml) for all the three; methanol, ultrapure water and pH 6.8 PBS whereas 323.4 nm for ultrapure water. Relative standard deviations of intra and inter day measurements in all the solvents was less than 2%. The limit of quantification and limit of detection values obtained were 0.06 µg/ml and 0.02 µg/ml for methanol and ultrapure water; 0.10 µg/ml and 0.03 µg/ml for pH 6.8 PBS. The results obtained were in agreement with the literature reported by Navarra et al. [24] and Shikov et al. [25].

Entrapment efficiency and partition coefficient

Entrapment efficiency of optimized CGA phytovesicles was 96.89±1.24 % [14] while the partition coefficient of CGA phytovesicle was found to be 0.977±0.36 as compared to 0.036±0.43 of CGA [26]. This increase in partition coefficient is attributed to the complexation between CGA and phospholipid. The CGA phytovesicle showed better solubility (30 times more) in oil phase (4.830 mg/ml) with solubility in water phase of 4.939 mg/ml as compared to plain CGA which showed 0.16 mg/ml of solubility in oil phase and 4.414 mg/ml solubility in water phase. Thus the phytovesicle formed was more amphiphilic in nature. Leptihn et al. [27] reported that pka, LogP and hydrophobicity play a major role for a molecule to exert antimicrobial activity. CGA in phytovesicular form showed higher logP values with optimum hydrophobicity thus having propensity for improved antimicrobial activity [27].

Characterization of optimized CGA phytovesicles

FTIR spectroscopy

Formation of phytovesicle was confirmed by comparing the spectra of the CGA and CGA phytovesicle. The characteristic peaks of CGA were identified as, phenolic and carboxylic O -H stretching at 3647.39 cm⁻¹, 3672.47 cm⁻¹, 3900.07 cm⁻¹; C=O ketonic stretching at 1714.72 cm⁻¹; aromatic C=C stretching at 1456.26 cm⁻¹ and 1539.20 cm⁻¹; aromatic ring stretching at 908.47 cm⁻¹, 817.82 cm⁻¹ (fig. 3). The spectra of Phospholipon 90H gave characteristic C-H stretching band of long chain fatty acid at 2916.37 cm⁻¹ and 2848.86 cm⁻¹[28]. Also, a C=O band at 1467.83 cm⁻¹; a -a (so3)³ band at 923.90 cm⁻¹ and a P -O-C stretch at 968.27 cm⁻¹[29]. The spectra of the physical mixture showed all the characteristic bands as those present in CGA and Phospholipon 90H. While in the spectrum of CGA phytovesicle was different from CGA and Phospholipon 90H with the absence of 908.47 cm⁻¹, 817.82 cm⁻¹, 92.39 cm⁻¹, 968.27 cm⁻¹ peaks. The peak at 3672.47 cm⁻¹ shifted left to 3687.90 cm⁻¹. Furthermore, the absence of aromatic stretch peaks has been previously attributed to successful formation of phytovesicular complex [30] indicating the hydrogen bonding between phenolic group of CGA with choline group of Phospholipon 90H[14].

Fig. 3: FTIR spectra of (a) CGA (b) Phospholipon 90H (c) Physical mixture (d) CGA phytovesicle
Differential scanning calorimetry (DSC)

Fig. 4 shows the thermograms of CGA, Phospholipon 90H, physical mixture and CGA Phytovesicles. CGA showed two sharp endothermic peaks at with onset at 195.01 °C and 202.56 °C which is in consistent with the literature reports [31]. Thermogram of Phospholipon 90H showed a sharp endothermic peak at onset of 101.43 °C and additional endothermic peaks at onset of 57.20 °C, and 177.69 °C. The physical mixture of CGA and Phospholipon 90H retained characteristics peaks of both CGA and Phospholipon 90H, however a new characteristic peak was seen at onset of 70.45 °C which can be attributed to the probable depression in melting endotherm of Phospholipon 90H, complex or eutectic formation. CGA phytovesicle exhibited an endotherm with an onset at 61.52 °C and also an endotherm close to characteristic physical mixture peak at 70.45 °C which clearly indicates the Phytovesicular complex formation. Additionally, disappearance of other characteristic endotherms of CGA and Phospholipon 90H, corroborates the formation of phytovesicular complex.

Particle size analysis

Particle size, zeta potential and SEM: The mean particle diameter of optimized phytovesicular complex was found to be 808.6±1.8 nm and the polydispersity index obtained as 0.78±0.06 indicating a narrow range of particle size distribution [32]. The Zeta potential was found to be (-) 18.6±6.3 mV (fig. 5) indicating the stability of the formulation. The micrographs of CGA phytovesicles show distinct structures unlike that of the characteristic structures of CGA and Phospholipon 90H (fig. 6).
Anti-microbial activity

Agar diffusion assay

Table 3: Agar diffusion assay-zone of inhibitions of CGA, CGA phytovesicle along with standard antibiotic Gentamycin against all the four bacterial strains

| Bacterial strain                  | Zone of inhibition in (mm) | CGA       | CGA Phytovesicle | DMSO | GEN       |
|-----------------------------------|-----------------------------|-----------|------------------|------|-----------|
| Staphylococcus aureus             |                             | 20±1.57   | 25±1.47          | -    | 33±1.15   |
| Klebsiella pneumonia              |                             | 15±2.5    | 22±3.02          | -    | 29±3.78   |
| Escherichia coli                  |                             | 17±1.6    | 23±1.52          | -    | 30±1.27   |
| Bacillus subtilis                 |                             | 16±2.64   | 17±2.08          | -    | 31±2.0    |

(-) No zone of inhibition, Values were expressed as mean±SD (n=3), CGA, Chlorogenic acid; DMSO, Dimethyl sulfoxide; GEN, Gentamycin

Agar diffusion assay is a standard susceptibility testing for antimicrobials where the antimicrobial agent diffuses into the agar plates and inhibits the growth of the test organism forming growth inhibition zones. Balouiri et al. [33] referred to it as a technique to screen or quantify the antimicrobial effect. CGA in phytovesicular form will give a larger zone of inhibition than plain CGA because of its higher penetrability across cell membrane with higher partition coefficient value [33]. The results obtained from agar diffusion assay are illustrated in table 3. CGA was found to be highly active against all the 4 strains. The zone of inhibition for CGA phytovesicle ranged between 17-25 mm as compared to CGA showing zone of 15-20 mm at 32 mg/ml. With increasing concentrations of both the samples, evenly increasing zones of inhibition were found with maximum activity at 32 mg/ml and a zone of 25 mm (fig. 7). While the zones obtained with standard antibiotic Gentamycin at equivalent concentration were in the range 30-33 mm [34].

Minimum inhibitory concentration

MIC values for CGA and CGA phytovesicles are given in table 4 which showed that CGA inhibited all the 4 strains. The MICs for CGA for all the 4 strains were in the range 500-550 µg/ml. While CGA phytovesicles showed the MIC obtained was two times less i.e. 250-300 µg/ml for Staphylococcus aureus, Bacillus subtilis, Klebsiella pneumonia and 200-250µg/ml for Escherichia coli. The reason for lower MIC values can be the improved physicochemical properties of CGA phytovesicles as compared to that of plain CGA [35]. A relatively higher lipophilic solubility of CGA phytovesicle is responsible for higher microbial membrane permeability ultimately leading to its disruption and cell death. This shows the efficiency of CGA in phytovesicular form with higher antibacterial activity.

Table 4: Minimum inhibitory concentrations of CGA and CGA phytovesicle against all the four bacterial strains

| Bacterial strain          | CGA      | CGA phytovesicle |
|---------------------------|----------|------------------|
| Gram positive             |          |                  |
| Staphylococcus aureus     | 500-550 µg/ml | 250-300 µg/ml |
| Bacillus subtilis         | 500-550 µg/ml | 250-300 µg/ml |
| Gram negative             |          |                  |
| Klebsiella pneumonia      | 500-550 µg/ml | 250-300 µg/ml |
| Escherichia coli          | 450-500 µg/ml | 200-250 µg/ml |

Values are expressed as per the subsequent dilution concentration (n=3)
Time kill assay

The effect of time on bactericidal activity of CGA and CGA phytovesicles in all the 4 strains was determined and shown in fig. 8. There was sharp decrease in the number of viable cells obtained with *Escherichia coli* and *Bacillus subtilis* after 6h of incubation and a linear decrease in the number of viable cells was found with *Staphylococcus aureus* and *Klebsiella pneumonia* after 12h. This higher resistance with the latter two strains can be explained with the basis of different cell wall composition [36]. A relatively higher lethal effect was observed with CGA phytovesicle after 6h of incubation as compared to moderate lethal effect after 12h with CGA. This strong bactericidal effect at MIC with a log reduction (1.5 times) in the range of 0.90-2.04 in CFUs at 24h for different strains as compared to 1.38-2.17 of plain CGA can be attributed to the efficiency of phytovesicular form of CGA in crossing the lipid bilayer of the cell and exerting its antimicrobial properties. Moreno *et al.* [37] reported that the antimicrobial effect of phenolic derivatives is related to inactivation of enzymes in the cell, which are further dependent on rate at which alterations in the cellular membrane and cellular penetration takes place. CGA in phytovesicular form crosses the lipid bilayer of the cell membrane more easily as compared to plain CGA. This loss in integrity of cell membrane causes cell death [37]. Thus a more lethal effect was obtained in phytovesicular form than plain CGA which increased the cell membrane permeability [38].

Effect of pH on antibacterial activity

Almajano *et al.* [39] outlined that antimicrobial effect of phenolic derivatives is governed by pH. Thus the effect of pH on antimicrobial activity on CGA and CGA phytovesicles was assessed and shown in fig. 9. It was found that at pH 10, no bactericidal effect was exerted, while at pH 7 a very weak bactericidal effect was exerted. On the other hand very lethal bactericidal effect was seen at pH 4. The reason for this was described by Narita *et al.* [40] that phenolic derivatives are more stable under acidic pH conditions. At higher pH (≥7) CGA undergoes irreversible structural changes [41]. It has been reported by Wen *et al.* [42] that antimicrobial activity of weak acids such as CGA depends on the ionization state and proportion of undissociated molecules of caffeic acid and quinic acid. Hence pH appeared to be a crucial factor in the bactericidal activity of CGA and parallel effect was seen on the antibacterial activity of CGA phytovesicles with less CFUs at lower pH as compared to plain CGA.
Effect of temperature on antibacterial activity

An effect of temperature on the bactericidal effect of CGA and CGA phytovesicles was studied which showed that a moderate bactericidal effect was observed at 37 °C and a marked bactericidal effect was obtained at 50 °C for all the 4 strains especially on Escherichia coli (fig. 10). While CGA phytovesicles showed further reduction in the CFUs with increased temperature. This was explained by Kabir et al. [20] and Lou et al. [43] who reported that the role of temperature in antimicrobial action was due to increased permeability of CGA through the bacterial cell walls. They also highlighted that the mechanism for temperature mediated membrane permeability may be because of cell wall disruption at specific temperature leading to cell death [20]. Hence the role of temperature in bactericidal effect of CGA and CGA phytovesicles was determined.

CONCLUSION

In present study, phytovesicular delivery of CGA provided a greater antibacterial activity over plain CGA. Increased antibacterial activity of CGA in phytovesicular form can be attributed to its improved lipophilic solubility (partition coefficient), which in turn should facilitate an increase in its permeability across bacterial cell wall bilayer. Thereby, reducing the MIC and kill time nearly to half of its plain drug form. The antibacterial activity was largely dependent on both pH and temperature. Phytovesicles exerted a marked antimicrobial action on both Gram negative [Klebsiella pneumonia and Escherichia coli] and Gram positive [Staphylococcus aureus and Bacillus subtilis] microbes. In order of sensitivity to CGA phytovesicles, the microorganisms are Eschericha coli> Bacillus subtilis> Staphylococcus aureus> Klebsiella pneumonia. All these strains are associated with various types of infections including Urinary tract infections, wound infections, meningitis, nosocomial infections etc. Demonstration of enhanced antimicrobial activity against these test bacteria indicate that there is a possibility of CGA phytovesicle as an alternative source of potential antibacterial agent.

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AUTHORS CONTRIBUTIONS

Hemangi Trivedi: Conceptualization, Methodology, Software, Validation, Formal analysis, Investigation, Data Curation, Writing-Original Draft, Writing-Review and Editing, Project administration.

Prashant K. Puranik: Conceptualization, Methodology, Validation, Resources, Writing-Review and Editing, Visualization, Supervision.

CONFLICT OF INTERESTS

The authors report no conflicts of interest. The authors alone are responsible for the content and writing of this article.

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