Edinburgh Research Explorer

Induction of Cell Stress in Neurons from Transgenic Mice Expressing Yellow Fluorescent Protein: Implications for Neurodegeneration Research

Citation for published version:
Comley, LH, Wishart, TM, Baxter, B, Murray, L, Nimmo, A, Thomson, D, Parson, SH & Gillingwater, TH 2011, ‘Induction of Cell Stress in Neurons from Transgenic Mice Expressing Yellow Fluorescent Protein: Implications for Neurodegeneration Research’, PLoS ONE, vol. 6, no. 3, e17639, pp. -.
https://doi.org/10.1371/journal.pone.0017639

Digital Object Identifier (DOI):
10.1371/journal.pone.0017639

Link:
Link to publication record in Edinburgh Research Explorer

Document Version:
Publisher's PDF, also known as Version of record

Published In:
PLoS ONE

General rights
Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.
Induction of Cell Stress in Neurons from Transgenic Mice Expressing Yellow Fluorescent Protein: Implications for Neurodegeneration Research

Laura H. Comley1,2,*, Thomas M. Wishart1,2,*, Becki Baxter1,2, Lyndsay M. Murray1,2, Ailish Nimmo1,2, Derek Thomson1,2, Simon H. Parson1,2†, Thomas H. Gillingwater1,2‡

1 Euan MacDonald Centre for Motor Neurone Disease Research, University of Edinburgh, Edinburgh, Lothian, United Kingdom, 2 Centre for Integrative Physiology, University of Edinburgh, Edinburgh, Lothian, United Kingdom

Abstract

Background: Mice expressing fluorescent proteins in neurons are one of the most powerful tools in modern neuroscience research and are increasingly being used for in vivo studies of neurodegeneration. However, these mice are often used under the assumption that the fluorescent proteins present are biologically inert.

Methodology/Principal Findings: Here, we show that thy1-driven expression of yellow fluorescent protein (YFP) in neurons triggers multiple cell stress responses at both the mRNA and protein levels in vivo. The presence of YFP in neurons also subtly altered neuronal morphology and modified the time-course of dying-back neurodegeneration in experimental axonopathy, but not in Wallerian degeneration triggered by nerve injury.

Conclusions/Significance: We conclude that fluorescent protein expressed in thy1-YFP mice is not biologically inert, modifies molecular and cellular characteristics of neurons in vivo, and has diverse and unpredictable effects on neurodegeneration pathways.

Introduction

The development of transgenic mice endogenously expressing spectral variants of fluorescent proteins such as GFP, YFP and CFP (collectively termed XFPs) in neurons, under the control of the neuronal-specific thy1 gene, has revolutionised modern neuroscience research, facilitating experimental breakthroughs and revolutionising our understanding of the form and function of the nervous system in health and disease [1–8]. Early studies on thy1-XFP mice reported that expression of fluorescent proteins in neurons had no discernable effect on neuronal morphology or any detectable level of toxicity [1], in agreement with earlier reports suggesting that XFPs are biologically inert [9]. Thus, there is an underlying assumption that the presence of fluorescent protein has no effect on the cellular or molecular composition of neurons in vivo. However, neurons in these mice contain large amounts of a foreign fluorescent protein. Such fluorescent proteins are known to be capable of altering the cellular and molecular characteristics of living cells in other experimental model systems in vivo and in vitro (e.g. the cardiovascular system), triggering stress responses and pathological changes [10–11]. Given the ever-increasing use of thy1-XFP mice for not only cellular, but also molecular, experiments in the healthy and pathological nervous system, a thorough investigation of the consequences of XFP expression in neurons of these mice is required.

Results and Discussion

We initially tested whether expression of yellow fluorescent protein modifies neuronal cell stress pathways in thy1-YFP line 16 (YFP-16) mice in vivo. YFP-16 mice from our breeding colony (founders originally purchased from Jackson Laboratories) appeared identical to those previously reported in the literature, expressing YFP in the vast majority of neurons in the peripheral nervous system [1]. We used cell stress pathway-specific RT4 profiler PCR arrays to quantify and compare expression of known cell stress-related genes in the spinal cord of thy1-YFP line 16 (YFP-16) mice on a C57Bl/6 background compared to wild-type C57Bl/6 controls from the same breeding colony. Surprisingly, half of all the cell stress genes (41 out of 84) on the PCR array were up-regulated more than 1.5-fold in spinal cord from YFP-16 mice (Figure 1A; Table 1). Of these, several had expression levels increased more than 2-fold, including genes...
involved in cell stress activation, inflammation and apoptosis (Table 1). None of the genes on the array were found to be significantly down-regulated >1.5 fold in YFP-16 mouse spinal cord (Figure 1A).

To confirm that the RNA changes observed in YFP-16 mouse spinal cord resulted in corresponding changes at the protein level, expression levels for 2 proteins selected from the PCR array (caspase 1 and CCL3) were validated in YFP-16 mouse spinal cord using quantitative fluorescent western blot (Figure 1B,C). Expression levels of another cell stress protein not included on the array - stress inducible protein 1 (STI1) - remained unchanged in YFP-16 tissue, showing that not all cell stress proteins responded to the presence of YFP (Figure 1B,C). Increased expression of caspase 1 correlated directly with the amount of YFP present, as levels in YFP-16 mouse spinal cord were 70-fold higher than levels in YFP-H mice where YFP is only expressed in a minority of neurons (Figure 1D,E) [1]. Importantly, immunohistochemistry for caspase 1 expression in YFP-H mouse spinal cord confirmed that increased cell stress responses were restricted to neuronal cells expressing YFP (Figure 1F-G). Together, these data demonstrate that expression of YFP in mouse neurons in vivo activates a robust, but selective, cell stress response at both the RNA and protein level.

We next sought to establish whether activation of cell stress pathways in YFP-expressing neurons had any adverse effects on neuronal morphology. We examined lower motor neurons innervating the levator auris longus muscle in YFP-H mice, allowing us to directly compare the morphological changes of YFP-expressing neurons with neighbouring non-expressing neurons in the same muscles from individual animals (Figure 2A). As previously reported [1], all neuromuscular junctions in 2–4 month old YFP-H mice were fully innervated by a single motor axon collateral (Figure 2A). However, higher-resolution analysis revealed a significant increase in abnormal accumulations of neurofilaments in distal axons and motor nerve terminals when YFP was present. This morphology is similar to cytoskeletal changes considered to represent early pathological changes in neurons [3,12] (Figure 2B-E). Neurofilament accumulations greater than a score of 1 on our 0–5 accumulation ranking scale were very rarely observed in distal axons and motor nerve terminals innervating the entire levator auris longus muscle in wild-type mice (data not shown). This reduces the likelihood that the neurofilament accumulations we observed in YFP-expressing neurons reflected an underlying biological diversity in neurofilament distribution between motor neuron pools that may or may not be selectively labelled in YFP-H mice. Retrospective examination of banked tissue from a now defunct YFP-16 mouse colony with very high YFP expression levels (~7 times greater than those found in our standard YFP-16 colony) revealed more severe morphological abnormalities, suggesting a dose-dependent effect (Figure 3).

It has previously been suggested that modulation of cell stress pathways can alter the response of neurons to neurodegenerative stimuli [13–14]. Therefore, we next addressed whether the presence of YFP in neurons also affected neurodegeneration pathways in vivo. YFP protein persists and retains its fluorescent properties in degenerating neurons, and has previously been used to visualise the breakdown of axons and synapses in a range of different models of neurodegeneration [3–5,15]. The rapid instigation (within 20 hours of nerve lesion), and morphological correlates, of Wallerian degeneration following nerve lesion were not altered in YFP-expressing motor neurons (Figure 2F-G). However, in wasted mice, where loss of the translation elongation factor eEF1A2 causes a severe distal axonopathy [15], the majority of YFP-expressing neurons remained intact whilst neighbouring non-YFP-expressing neurons were in the final stages of degeneration, even at late-symptomatic ages (Figure 2H-L). Therefore, the severity of dying-back pathology, which is mechanistically distinct from Wallerian degeneration [15], was significantly modified in YFP-expressing motor neurons from wasted mice.

In conclusion, our finding that expression of YFP in neurons instigates cell stress pathways means that fluorescent protein expressed in neurons from thy1-YFP mice is not biologically inert. This finding provides important evidence, supported by data from other in vivo and in vitro experimental systems [10–11,16–17], that fluorescent proteins expressed in cells/tissues can modify their molecular composition and cellular activities. Care must now be taken with regard to experimental design and data interpretation, particularly when investigating molecular mechanisms of neurodegeneration in mice endogenously expressing fluorescent protein in neurons. Future studies must take into account the modified molecular characteristics and altered phenotypic responses observed here and perform necessary controls to ensure that any responses they are reporting are a direct result of experimental intervention rather than simply a consequence of the expression of fluorescent protein.

**Materials and Methods**

**Ethics statement**

All animal experiments were approved by a University of Edinburgh internal ethics committee and were performed under license by the UK Home Office (project license number 60/3891).

**Mice**

Breeding colonies of thy1-YFP-H mice [1] and thy1-YFP-16 mice, both on a C57BL/6J background, were originally purchased as breeding pairs from Jackson Laboratories and established and maintained in animal care facilities (standard SPF conditions) at the University of Edinburgh. Wild-type C57Bl/6J mice (not expressing YFP protein) from the same breeding colony...
were used for controls throughout. Mice were sacrificed by inhalation of isoflurane (2% in 1:1 N₂O/O₂). All procedures were carried out under licensed authority from the UK Home Office. The YFP status of mice was ascertained by examining ear punches for YFP-positive neurons. To determine the effects of YFP expression on dying-back pathology, YFP-H mice were crossed with *wasted* mice [15]. *Wasted*/YFP-H mice were genotyped using standard PCR techniques, as described previously [15], and were taken for analysis at post-natal day 24 (late-symptomatic). The banked tissue from a previous, now defunct, colony of strongly-expressing *thy1-YFP-16* mice was obtained from mice originally supplied to the University of Leeds by Jackson Laboratories.

**Cell stress array**

Mouse cell stress focused pathway arrays were performed in a 96-well plate format (PAM-003A), compatible with an ABI 7000 real-time PCR machine, were used to assay cell stress gene expression (3 comparisons for YFP-16 and 3 for wild-type controls, each comparison composed of pooled whole spinal cord tissue from 3 individual 2 month old mice). RNA was extracted as previously described [14]. Samples were added to the reaction plate and signal amplification by PCR was carried out using Sybr-Green ‘1 step qRT-PCR kit’ (Invitrogen). Analysis was carried out using the Analysis Suite spreadsheet provided by Tebubio SuperArrays. Gene functions listed in Table 1 were obtained from the SuperArray product specification sheets.

**Quantitative fluorescent (Li-COR) western blots**

Total protein was isolated from spinal cords and quantitative western blots were performed as described previously [18–20]. Primary antibodies were used as per manufacturers instructions (ST11—BD Biosciences; Caspase 1 and CCL3—Abcam; GFP—Millipore). Odyssey secondary antibodies were used in accordance with manufacturers instructions (Goat anti mouse IRDye 680 and Goat anti rabbit IRDye 680). Blots were imaged using an Odyssey Infrared Imaging System (Li-COR Biosciences). Quantification was performed on single channels with the analysis software provided. Bands were identified according to their molecular weight and the arbitrary fluorescence intensity was calculated by Odyssey software.

**Spinal cord immunohistochemistry**

Entire vertebral columns were removed and immediately immered in 4% paraformaldehyde solution for 1 hour. Spinal cords were removed from the vertebral column and fixed for a further 30 mins before embedding in 2% low-melting point agarose. Embedded spinal cords were mounted on a vibratome and transverse sections cut at a thickness of ~200 µm for immunohistochemical staining. Sections were blocked in 4% bovine serum albumin (BSA) and 1.5% TritonX in 0.1 M PBS for 30 minutes before incubation in primary antibodies directed against caspase 1 (1:500 dilution; Abcam) overnight. After washing for 2 hours in 0.1 M PBS, sections were incubated for 2 hours in a 1:30 dilution of swine anti-rabbit TRITC-conjugated secondary antibody (Dako). Sections were co-stained with the nuclear dye TOPRO-3 (Molecular Probes), whole-mounted in Mowiol® (Calbiochem) on glass slides and cover-slipped for subsequent imaging.

**Surgery**

Mice were anaesthetised by inhalation of halothane (2% in 1:1 N₂O/O₂) before exposing the intercostal nerves innervating the transversus abdominis muscle. Nerves were cut before suturing the skin and allowing the mouse to recover for 20 hours. Post-operative mice were maintained in standard animal house conditions.

---

**Table 1.** Mouse SuperArray data showing greater than 1.5 fold cell stress RNA expression changes in the spinal cord of YFP-16 mice compared with wild-type controls (array cell refers to the location on the 3D bar chart in Fig. 1A).

| Array cell* | Symbol | Fold change | Role in cell stress |
|-------------|--------|-------------|---------------------|
| A00         | Ccl4   | 3.26        | Inflammation        |
| C03         | Cyp7a1 | 3.14        | Oxidative or metabolic stress |
| A08         | Ccl3   | 2.94        | Inflammation        |
| B04         | Cs2f   | 2.77        | Inflammation        |
| A05         | Casp1  | 2.30        | Apoptosis signalling |
| F06         | Nfkbia | 2.26        | Apoptosis signalling |
| B11         | Cyp2c29| 2.23        | Oxidative or metabolic stress |
| F01         | Lta    | 2.18        | Inflammation        |
| A06         | Casp8  | 2.11        | Apoptosis signalling |
| B02         | Chek2  | 2.07        | DNA damage and repair |
| A01         | Anxa5  | 1.95        | Apoptosis signalling |
| A07         | Ccl21b | 1.94        | Inflammation        |
| C01         | Cyp4a10| 1.93        | Oxidative or metabolic stress |
| G05         | Tnfsf10| 1.92        | Apoptosis signalling |
| D05         | Gpx2   | 1.90        | Oxidative or metabolic stress |
| A03         | Bax    | 1.89        | Apoptosis signalling |
| B05         | Cxcl10 | 1.84        | Inflammation        |
| D09         | Hmox1  | 1.84        | Oxidative or metabolic stress |
| B10         | Cyp2b9 | 1.84        | Oxidative or metabolic stress |
| F07         | Nos2   | 1.79        | Inflammation        |
| D08         | Gstm3  | 1.79        | Oxidative or metabolic stress |
| B06         | Cyp1a1 | 1.74        | Oxidative or metabolic stress |
| F02         | Mdm2   | 1.72        | Growth arrest and senescence |
| C07         | Egr1   | 1.72        | Proliferation and carcinogenesis |
| C04         | Ddit3  | 1.68        | Growth arrest and senescence |
| F12         | Rad50  | 1.67        | DNA damage and repair |
| A02         | Atm    | 1.66        | DNA damage and repair |
| E11         | Il1b   | 1.65        | Inflammation        |
| C02         | Cyp4a14| 1.64        | Oxidative or metabolic stress |
| H01         | Gusb   | 1.63        | Oxidative or metabolic stress |
| B07         | Cyp1b1 | 1.63        | Oxidative or metabolic stress |
| G08         | Ugt1a2 | 1.63        | DNA damage and repair |
| C12         | Fmo1   | 1.62        | Oxidative or metabolic stress |
| A04         | Bcl2l1 | 1.62        | Apoptosis signalling |
| E02         | Hspa4  | 1.60        | Heat Shock          |
| A11         | Ccnd1  | 1.59        | Proliferation and carcinogenesis |
| B03         | Cryab  | 1.56        | Oxidative or metabolic stress |
| B09         | Cyp2b10| 1.56        | Oxidative or metabolic stress |
| B12         | Cyp3a11| 1.56        | Oxidative or metabolic stress |
| F11         | Rad23a | 1.52        | DNA damage and repair |
| G04         | Tnfrsf1a| 1.52      | Apoptosis signalling |

doi:10.1371/journal.pone.0017639.t001
Immunohistochemical analysis of axons and neuromuscular junctions

The levator auris longus (LAL) [21–22] and/or transversus abdominis (TVA) [12] muscles and their nerve supply were dissected and fixed in 0.1 M PBS containing 4% paraformaldehyde (Electron Microscopy Science) for 10 minutes. YFP tissue was additionally double-labelled by exposure to α-bungarotoxin conjugated to Alexa Fluor® 647 (Alexa 647-α-BTX; 5 mg/ml, Invitrogen) for 30 minutes to label post-synaptic acetylcholine receptors. Muscles were incubated in 4% bovine serum albumin (BSA) and 1.5% TritonX in 0.1 M PBS for 30 minutes before incubation in primary antibodies directed against 145 kDa neurofilament proteins (1:300 dilution; Millipore) overnight to label axons and nerve terminals. After washing for 2 hours in 0.1 M PBS, muscles were incubated for 4 hours in a 1:30 dilution of swine anti-rabbit TRITC-conjugated secondary antibody (Dako). Muscles were whole-mounted in Mowoil® (Calbiochem).

Microscopy

Fluorescently labelled axon and nerve/muscle preparations were viewed using a laser scanning confocal microscope (63× objective; 1.4NA; Zeiss LSM 710). Confocal Z-series were merged using Zeiss software and images assembled for analysis using Adobe Photoshop. All images were taken using sequential laser capture and with the excitation/emission spectra set to ensure no bleed-through between channels. A minimum of 50 neuromuscular junctions, selected at random, were assessed in each muscle preparation. Quantitative analysis was performed on micrographs with the YFP channel initially excluded, so that the operator did not know the YFP status of the tissue being analysed. For analysis of neurofilament accumulation in normal tissue, endplates were qualitatively assigned a score between 0 and 5, with 0 representing healthy neuromuscular junctions with no evidence of neurofilament accumulation and 5 representing nerve terminals with large abnormal accumulations of neurofilament. The YFP channel was revealed subsequent to quantification to

Figure 2. YFP expression in motor neurons subtly disrupts normal neuronal morphology and alters responses to dying-back pathology but not Wallerian degeneration. A–D – Representative confocal micrographs of NMJs in the levator auris longus muscle from a YFP-H mouse labelled for neurofilaments (NF: red) and postsynaptic motor endplates (BTX: blue). Note that all motor endplates were innervated, but only a small proportion of motor axons were YFP-positive in these mice (A). Panels B–D show high power micrographs of NMJs identified in panel A. Note abnormal accumulations of NF only in the motor nerve terminals of YFP-positive NMJs (B = NF accumulation score of 0, C = 3, D = 5). E – Bar chart (mean ± s.e.m.) showing quantification of NF accumulation in motor nerve terminals from YFP-H mice (0 = no accumulation; 5 = large abnormal accumulation), revealing a significant increase in NF accumulation in YFP-positive terminals (N = 5 mice, n = 9 muscles; *** P < 0.001 Mann-Whitney test). F–G – Representative confocal micrographs of intramuscular axons supplying the transversus abdominis muscle from YFP-H mice (also labelled for NFs; red), before (F) and 20 hours after (G) intercostal nerve lesion. The presence of YFP did not alter the rate or morphological appearance of Wallerian degeneration after nerve injury, with NF fragmentation occurring in YFP-positive and –negative axons in all nerves examined (N = 6 mice, n = 6 nerves). H–K – Representative confocal micrographs of two NMJs in the levator auris longus muscle from a late-symptomatic (P24) wasted/YFP-H mouse labelled to reveal NFs (red) and postsynaptic motor endplates (BTX: blue). Note how the motor axon with YFP (bottom NMJ) remained intact whereas the motor axon without YFP (top) was undergoing retraction, characteristic of a dying-back pathology. L – Bar chart showing quantification of dying-back pathology at the NMJ in late-symptomatic (P24) wasted/YFP-H mice, revealing a significant attenuation of dying-back pathology in motor nerve terminals where YFP was present (i.e. a retention of endplates fully occupied by overlying NFs and a reduction in the numbers of partially occupied endplates; N = 4 mice, n = 7 muscles; *** P < 0.001 Mann-Whitney test). Scale bars = 80 μm (A), 40 μm (B–D), 30 μm (F–G), 50 μm (H–K).

doi:10.1371/journal.pone.0017639.g002
Figure 3. Increased incidence of morphological abnormalities at the NMJ in tissue banked from a strongly-expressing YFP-16 mouse colony. A - Bar chart (mean ± s.e.m.) showing relative expression levels of YFP in the sciatic nerve of YFP-16 mice from the current Edinburgh colony (low expression), compared to YFP levels in tissue banked from a now defunct high-expressing YFP-16 colony previously held at the University of Leeds, quantified using fluorescent western blot (*** P<0.01 unpaired two-tailed t test). B–C - Representative micrographs showing NMJ morphology in low-expressing YFP-16 mice (B) compared to high-expressing YFP mice (C). Motor endplates are labelled and shown in red. Note the presence of large YFP accumulations in both axons (white arrow) and motor nerve terminals (blue) from the high-expressing tissue. Such morphological abnormalities were present in all the preparations examined from high-expressing YFP mice.

doi:10.1371/journal.pone.0017639.g003

References

1. Feng G, Mellor RH, Bernstein M, Keller-Peck C, Nguyen QT, et al. (2000) Imaging neuronal subsets in transgenic mice expressing multiple spectral variants of GFP. Neuron 28: 49–51.
2. Trachtenberg JT, Chen BE, Knott GW, Feng G, Sanes JR, et al. (2002) Long-term in vivo imaging of experience-dependent synaptic plasticity in adult cortex. Nature 420: 788–794.
3. Mi W, Beirovski B, Gillingwater TH, Adalbert R, Wagner D, et al. (2005) The slow Wallerian degeneration gene, Wdls, inhibits axonal spheroïd pathology in gracile axonal dystrophy mice. Brain 128: 405–416.
4. Beirovski B, Adalbert R, Wagner D, Grumme DS, Addicks K, et al. (2005) The progressive nature of Wallerian degeneration in wild-type and slow Wallerian degeneration (Wdls) nerves. BMC Neurosci 6: 6.
5. Kerschensteiner M, Schwab ME, Lichtman JW, Misgeld T (2005) In vivo imaging of axonal degeneration and regeneration in the injured spinal cord. Nat Med 11: 572–577.
6. Sugino K, Hempel CM, Miller MN, Hattox AM, Shapiro P, et al. (2006) Molecular taxonomy of major neuronal classes in the adult mouse forebrain. Nat Neurosci 9: 99–107.
7. Lever J, Weissman TA, Kang H, Draft RW, Lu J, et al. (2007) Transgenic strategies for combinatorial expression of fluorescent proteins in the nervous system. Nature 450: 56–62.
8. Keck T, Mroo-Flogel TD, Vaz Alfonso M, Eysel UT, Bonhoeffer T, et al. (2008) Massive restructuring of neuronal circuits during functional reorganization of adult visual cortex. Nat Neurosci 11: 1162–1167.
9. Hein R, Prasher DC, Tsien RY (1994) Wavelength mutations and posttranslational autoxidation of green fluorescent protein. Proc Natl Acad Sci USA 91: 12501–12504.
10. Liu HS, Jan MS, Chou CK, Chen PH, Kc NJ (1999) Is green fluorescent protein toxic to the living cells? Biochem Biophys Res Commun 260: 712–717.
11. Huang WY, Aramburu J, Douglas PS, Imamo S (2000) Transgenic expression of green fluorescence protein can cause dilated cardiomyopathy. Nat Med 6: 482–483.
12. Murray LM, Comley LH, Thomson D, Parkinson N, Talbot K, et al. (2008) Selective vulnerability of motor neurons and dissociation of pre- and post-synaptic pathology at the neuromuscular junction in mouse models of spinal muscular atrophy. Hum Mol Genet 17: 949–962.
13. Xu C, Bally-Cuif B, Reed JC (2005) Endoplasmic reticulum stress: cell life and death decisions. J Clin Invest 115: 2636–2644.
14. Wishart TM, Pemberton HN, James SR, McCabe CJ, Gillingwater TH (2008) Modified cell cycle status in a mouse model of altered neuronal vulnerability (slow Wallerian degeneration; Wdls). Genome Biol 9: R101.
15. Murray LM, Thomson D, Coradin A, Wishart TM, Gillingwater TH (2008) Loss of translation elongation factor (eEF1A2) expression in vivo differentiates between Wallerian degeneration and dying-back neuronal pathology. J Anat 213: 633–645.
16. Detroit ER, Bower WJ, Halterman MW, Giuliani RE, Bennice L, et al. (2002) Reporter gene transfer induces apoptosis in primary cortical neurons. Mol Therap 5: 723–730.
17. Stevens JC, Chia R, Hendriks WT, Bros-Facer V, van Minnen J, et al. (2010) Modification of superoxide dismutase 1 (SOD1) properties by a GFP-tag-implications for research into amyotrophic lateral sclerosis (ALS). PLoS One 5: e9541.
18. Wishart TM, Paterson JM, Short DM, Meredith S, Robertson KA, et al. (2007) Differential proteomics analysis of synaptic proteins identifies potential cellular targets and protein mediators of synaptic neuroprotection conferred by the slow Wallerian degeneration (Wdls) gene. Mol Cell Proteomics 6: 1318–1330.
19. Wishart TM, Huang JP, Murray LM, Lamont IIJ, Mutsaers EA, et al. (2010) SMN deficiency disrupts brain development in a mouse model of severe spinal muscular atrophy. Hum Mol Genet 19: 4216–4228.

Statistical analysis

All data were collected into Microsoft Excel spreadsheets and analysed using GraphPad Prism software. All bar charts shown are mean ± s.e.m. Statistical significance was considered to be p<0.05 for all analyses. Individual statistical tests used are detailed in figure legends.

Acknowledgments

The authors would like to thank Professor Mike Cousin and members of the Gillingwater and Parson laboratories for helpful advice with this study and comments on the manuscript.

Author Contributions

Conceived and designed the experiments: LHC TMW SHP THG. Performed the experiments: LHC TMW BB LMM AN DT SHP THG. Analyzed the data: LHC TMW BB LMM AN SHP THG. Wrote the paper: LHC TMW SHP THG.
20. Wright AK, Wishart TM, Ingham CA, Gillingwater TH (2010) Synaptic protection in the brain of WldS mice occurs independently of age but is sensitive to gene-dose. PLoS One 5: e15108.

21. Murray LM, Gillingwater TH, Parson SH (2010) Using mouse cranial muscles to investigate neuromuscular pathology in vivo. Neuromuscul Disord 20: 740–743.

22. Murray LM, Lee S, Baumer D, Parson SH, Talbot K, et al. (2010) Pre-symptomatic development of lower motor neuron connectivity in a mouse model of severe spinal muscular atrophy. Hum Mol Genet 19: 420–433.