Killing of *Staphylococcus aureus* and *Salmonella enteritidis* and neutralization of lipopolysaccharide by 17-residue bovine lactoferricins: improved activity of Trp/Ala-containing molecules

Ya Hao1,2, Na Yang1,2, Xiumin Wang1,2, Da Teng1,2, Ruoyu Mao1,2, Xiao Wang1,2, Zhanzhan Li1,2 & Jianhua Wang1,2

Bovine lactoferricin (LfcinB) has potent antibacterial, antifungal and antiparasitic activities but is also hemolytic. Our objective was to identify LfcinB17-31 derivatives with reduced hemolysis and improved antimicrobial activity via substituting Cys3, Arg4, Gln7, Met10, and Gly14 with more hydrophobic residues. Two peptides, Lfcin4 and Lfcin5, showed higher activity against *Staphylococcus aureus* and *Salmonella enteritidis* and lower hemolytic activity than the parent peptide LfcinB17-31. These peptides permeabilized the outer and inner membranes of *S. enteritidis*; however, Lfcin5 did not permeabilize the inner membrane of *S. aureus*. Gel retardation and circular dichroism spectra showed that Lfcin4 and Lfcin5 bound to bacterial genomic DNA. Lfcin4 inhibited DNA, RNA and protein synthesis. Both peptides induced the peeling of membranes and the lysis of *S. enteritidis*. At doses of 10 and 15 mg/kg, Lfcin4 and Lfcin5 reduced the bacterial counts in infected thigh muscles by 0.03–0.10 and 0.05–0.63 log10 CFU/g of tissue, respectively, within 10 h. Lfcin4 and Lfcin5 enhanced the survival rate of endotoxemic mice; reduced serum IL-6, IL-1β and TNF-α levels; and protected mice from lipopolysaccharide-induced lung injury. These data suggest that Lfcin4 and Lfcin5 may be antimicrobial and anti-endotoxin peptides that could serve as the basis for the development of dual-function agents.

*Staphylococcus aureus* and *Salmonella enteritidis* are leading causes of infection at numerous sites in the body (including skin, soft tissue, blood, and lungs) in both humans and animals1–3. The use of antibiotics is critical for the treatment of these infectious bacterial diseases. However, multidrug-resistant isolates of *S. aureus* and *S. enteritidis* have been present for many years4. Thus, it is of critical importance to identify alternatives to traditional antibiotics. Additionally, the endotoxin lipopolysaccharide (LPS) found on cell membranes of Gram-negative bacteria, a primary trigger of sepsis, can stimulate host cells to produce a large amount of proinflammatory cytokines that play a pivotal role in the pathogenesis of multiple organ failure5,6. Currently, the neutralization of LPS is one of the therapeutic strategies being investigated for sepsis; however, no therapeutic agents thus far have been efficacious enough to protect the host from tissue damage and organ failure induced by LPS. Thus, there is a pressing need for new effective agents that can neutralize LPS.

The bovine lactoferricin (LfcinB) peptide is generated by the gastric pepsin cleavage of the N-terminal region of the iron-binding protein lactoferrin. LfcinB consists of 25 amino acids, corresponding to residues 17–41 in lactoferrin, and is a bioactive peptide that possesses antibacterial, antifungal, antiviral, antiparasitic, antitumor,
and immunological activities\textsuperscript{8–17}. A shorter version of LfcinB containing amino acids 17 to 31 (LfcinB17–31) was found to have slightly less antimicrobial activity against \textit{S. aureus} and \textit{Escherichia coli} than LfcinB1–41\textsuperscript{18}.

A previous study showed that the core hexapeptide (RRWQR) of LfcinB can cause membrane disturbances in model membranes and penetrate HeLa cells\textsuperscript{19}. Additionally, the Trp residues at positions 6 and 8 of LfcinB17–31 are crucial for its high antibacterial activity\textsuperscript{20}. The largest loss of antimicrobial activity is found when either of these Trp residues is replaced by Ala. A single Ala substitution at Cys3, Gln7 or Gly14 results in an approximately 1-fold increase in the antibacterial activity of LfcinB17–31 against \textit{E. coli}, and other Ala substitutions display reduced antibacterial activity against \textit{S. aureus}\textsuperscript{20}. Substitutions of Cys3, Gln7 and Ala14 with Trp have revealed that the antimicrobial activity increases with the amount of Trp residues, especially against \textit{S. aureus}\textsuperscript{21}. The aromatic Trp residues are hypothesized to function as an anchor in membrane proteins and act as a needle that pulls the $\alpha$-helix across phospholipid membranes\textsuperscript{22}. Additionally, the Arg residues were reported to be critical for antimicrobial activity against \textit{Mycobacterium avium}\textsuperscript{23}. All of the Arg substitutions (at Lys2, Lys11 and Lys12) were slightly more active than the parent peptide. Conversely, the substitution of Lys by Arg4, Arg5 and Arg9 resulted in significantly less antimicrobial activity than the corresponding original peptide\textsuperscript{23}.

Some antimicrobial peptides (AMPs) exhibit remarkable specificity for particular bacterial species. LfcinB can damage the outer membrane of bacteria by interaction with LPS or teichoic acid, and it is hypothesized that the main function of LfcinB is to increase the permeability of the cytoplasmic membrane of both Gram-positive and Gram-negative bacteria, a mechanism closely resembling that of the class L antibiotic peptides\textsuperscript{24,25}. LfcinB inhibits DNA synthesis in \textit{E. coli} UC 6782 and interferes with DNA, RNA and protein synthesis in \textit{Bacillus subtilis}\textsuperscript{25}. It was determined that the mode of action of LfcinB in \textit{B. subtilis} may differ from that in \textit{E. coli} and that LfcinB may induce an SOS-like response in \textit{E. coli}\textsuperscript{25}. Ho et al. identified the intracellular targets of LfcinB using \textit{E. coli} K12 proteome microarrays and found that LfcinB binds to two response regulators, BasR and CreB, members of the two-component system family. They further showed that LfcinB inhibited the growth of bacteria by directly influencing the phosphorylation of a two-component system protein\textsuperscript{25}. However, the exact mode of action of LfcinB or its derivatives against \textit{S. aureus} and \textit{S. enteritidis} has not yet been elucidated.

In the present study, on the basis of previous reports wherein some residues of Phe1, Lys2, Trp6, Trp8, Arg9, Lys11, Lys12, and Ala15 are found to be essential for antibacterial activity\textsuperscript{18–22,27}, we performed a systematic analysis of residues in the 3rd, 4th, 7th, 10th, and 14th positions of LfcinB17–31 by substituting the hydrophobic amino acids (Ala, Phe and Trp) at each position. LfcinB17–31 and its derivatives were synthesized and their antimicrobial activities against \textit{Gram-positive} and \textit{Gram-negative} bacteria were evaluated. One possible mechanism of action of LfcinB derivatives against \textit{S. aureus} and \textit{S. enteritidis} was further elucidated. Additionally, the \textit{in vivo} antibacterial activities of LfcinB derivatives were examined in a mouse thigh model of \textit{S. aureus} infection, as well as their endotoxin-neutralizing activities in \textit{C57BL/6} mice challenged with LPS.

\section*{Results}

\textbf{Physicochemical parameters, structures and electrostatic potentials on the surface of LfcinB17–31 and its derivatives.} The amino acid sequences and key physicochemical parameters of the LfcinB17–31 derivatives are summarized in Table 1. The physicochemical properties of Lfcin2 were almost equal to those of the parent peptide, Lfcin1. The net charge of peptides Lfcin3-Lfcin6 was lower than that of Lfcin1. The Lfcin3, Lfcin4 and Lfcin5 peptides had the highest $\alpha$-helical content at 64.7% and Lfcin6 had the lowest $\alpha$-helix content at 35.2%. Lfcin2 contained a lower helical content (52.9%) compared to Lfcin1. The hydrophobicity of Lfcin3-Lfcin6 increased from $-1.207$ to $-0.376$, indicating that they are more hydrophobic than Lfcin1 and Lfcin2; this is likely due to the addition of a methyl group from the Gly/Cys/Gln to Ala residue substitution\textsuperscript{28}. The instability index of Lfcin2-Lfcin6 ranged from 28.82 to 46.67. The Boman index, which is a measure of peptide instability, also increased from 32.67 to 46.47.

The two-dimensional (2D) and three-dimensional (3D) structure modeling of Lfcin3, Lfcin4 and Lfcin5 revealed similarities to their parent peptide, which adopts a typical $\alpha$-helix (Supplementary Figs S1A and S1B). However, Lfcin4 and Lfcin5 appeared to have a more irregular coil in the C-terminus (35.3%), whereas Lfcin6 lost...
as shown in Table 2, Lfcin4 and Lfcin5 showed higher activity against E. coli, S. enteritidis and S. aureus than against Lfcin1. The MIC values for Lfcin4 and Lfcin5 against S. enteritidis CVCC3377, S. aureus ATCC25923 and S. aureus ATCC43300 were in the range of 2 to 64 μg/ml. Lfcin6 exhibited the lowest antimicrobial activity against the test strains. Similar to Lfcin1, higher MICs (>128 μg/ml) for Lfcin2 to Lfcin6 against S. pullorum CVCC1789 and CVCC1802, and P. aeruginosa CVCC10419 and CICC21630 were observed. Moreover, Lfcin4 and Lfcin5 displayed very low hemolytic activity against mouse erythrocytes (Fig. 1A). Even at a high concentration (512 μg/ml), the hemolytic activity of Lfcin4 and Lfcin5 was 4.87% and 2.2%, respectively, which is lower than that of the parent peptide, Lfcin1 (approximately 7.5% hemolysis at 256 μg/ml).

### Circular dichroism (CD) spectra of Lfcin4 and Lfcin5.

Anionic surfactants such as sodium dodecyl sulfate (SDS) and fluorinated alcohols such as trifluoroethanol (TFE) are often used to mimic the cell membrane environment. To analyze the structural features of Lfcin4 and Lfcin5, the CD spectra of peptides were measured in ddH₂O, 5–40 mM SDS and 50% TFE buffer, respectively. The secondary structures of Lfcin4 and Lfcin5 in ddH₂O were characterized predominantly by antiparallel and random coils with a characteristic positive maximum at 230 nm and a negative minimum at 200 nm (Fig. 1B and C). The CD spectra of Lfcin4 and Lfcin5 showed an increase in α-helical content in SDS and TFE buffer (>11.8% α-helix) (Supplementary Tables S2 and S3) relative to the parent peptide (<10% α-helix). This result indicates that both SDS and TFE buffer are favorable for the formation of an α-helical structure.

### Lfcin4 and Lfcin5 affect the cell surface hydrophobicity of bacterial cells.

As shown in Fig. 2A and B, the cell surface hydrophobicity of S. aureus ATCC25923 and S. enteritidis CVCC3377 treated with peptides was positively correlated with the concentration of Lfcin4 and Lfcin5. For S. aureus ATCC25923, treatment with Lfcin4 resulted in an increased cell surface hydrophobicity from 5.27% to 18.87% (Fig. 2A), whereas Lfcin5 caused a relatively small change in cell surface hydrophobicity (from 5.27% to 8.95%). The cell surface hydrophobicity of S. enteritidis CVCC3377 increased from 9.43% to 16.45% and from 9.43% to 16.74% following treatment with Lfcin4 and Lfcin5, respectively, which is markedly higher than the untreated controls. Lfcin5 exhibited higher cell surface hydrophobicity than Lfcin4 (Fig. 2B). Changes in cell surface hydrophobicity are one of the noticeable effects of peptide interactions with bacterial cells, which may be correlated with structural alterations of bacterial cell surfaces.

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Table 2. MIC values of LfcinB17-31 and its derivatives against bacteria. aChina Veterinary Culture Collection Center (CVCC). bChina Center of Industrial Culture Collection (CICC). cAmerican Type Culture Collection (ATCC). dNational Center for Medical Culture Collection (CMCC). The data are representative of three independent experiments.

| Strains                        | Lfcin1 | Lfcin2 | Lfcin3 | Lfcin4 | Lfcin5 | Lfcin6 |
|-------------------------------|--------|--------|--------|--------|--------|--------|
|                               | μM     | μM     | μM     | μM     | μM     | μM     |
| Gram-negative bacteria        |        |        |        |        |        |        |
| Escherichia coli CVCC195*     | 16     | 32     | 15     | 32     | 31     | 64     |
| E. coli CVCC1515*             | 16     | 32     | >59    | >128   | 31     | 64     |
| E. coli CVCC1530 (serotype O157:H7)* | 32    | 64    | 59     | 128    | 31     | 64     |
| Salmonella typhimurium ATCC14028F | 16        | 32    | 29     | 64     | 61     | 128    |
| S. enteritidis CVCC3377*      | 8      | 16     | 15     | 32     | 1.9    | 4      |
| S. pullorum CVCC1789*         | >64    | >128   | >59    | >128   | >61    | >128   |
| S. pullorum CVCC1802*         | >64    | >128   | >59    | >128   | >61    | >128   |
| S. cholerasuis CVCC503*        | 64     | 128    | 59     | 128    | 61     | 128    |
| Pseudomonas aeruginosa CICC10419* | >64 | >128  | >59    | >128   | >61    | >128   |
| P. aeruginosa CICC21630*       | >64    | >128   | >59    | >128   | >61    | >128   |
| Gram-positive bacteria        |        |        |        |        |        |        |
| Staphylococcus aureus ATCC43300* | >64 | >128  | >59    | >128   | >61    | >128   |
| S. aureus ATCC25923F          | 1      | 2      | 0.92   | 2      | 0.95   | 2      |
| Streptococcus suis CVCC606*   | >64    | >128   | >59    | >128   | 61     | 128    |
| Enterococcus faecium CMCC1.2136F | 8      | 16     | 15     | 32     | 15     | 32     |
| Bacillus subtilis ATCC6633F    | 32     | 64     | 29     | 64     | 15     | 32     |
Scanning electron microscopy (SEM) observations. Scanning electron microscopy (SEM) was used to directly observe the effects of 1× MIC Lfcin4 or Lfcin5 treatment on the cell morphology and integrity of *S. aureus* ATCC25923 and *S. enteritidis* CVCC3377. As shown in Fig. 2C, untreated *S. aureus* cells displayed an intact cell morphology. After treatment with 1× MIC Lfcin4 or Lfcin5 for 2 h, the *S. aureus* cells appeared smaller, suggesting that the cells shrunk or released intracellular components. In untreated control groups, a normal intact cell morphology was observed in *S. aureus*. For *S. enteritidis* CVCC3377, treatment with Lfcin4 or Lfcin5 induced membrane peeling in approximately 30% of cells and lysis in approximately 50% of cells.

Lfcin4 and Lfcin5 permeabilize model and bacterial membranes. Permeabilization of model membranes. To investigate whether Lfcin4 or Lfcin5 causes membrane disruption of bacterial cells, calcein was used as a marker for membrane damage. The release of calcein from liposomes was fluorometrically monitored following exposure to Lfcin4 or Lfcin5 peptides. As shown in Fig. 3A, both Lfcin4 and Lfcin5 induced the leakage of 2-oleoyl-1-palmitoyl-sn-glycero-3-phosphocholine/2-oleoyl-1-palmitoyl-sn-glycero-3-glycerol (POPC/POPG) vesicles. In *E. coli*, Lfcin4 treatment resulted in 45.5%, 90.6% and 98.4% dye leakage from vesicles at 5 min when used at 1×, 2× and 4× MIC, respectively. Treatment with Lfcin5 caused a similar result, with 30.1–70.3% leakage within 5 min (data not shown). These results indicate that both Lfcin4 and Lfcin5 have potent ability to permeabilize the lipid bilayer in a time- and concentration-dependent manner and that Lfcin4 is more effective at permeabilizing bilayers than Lfcin5.

Permeabilization of the outer membrane in *S. aureus* and *S. enteritidis*. The hydrophobic fluorophoric probe N-phenyl-1-naphthylamine (NPN) is normally prevented from entering cells because it cannot penetrate the outer membrane. Once the outer membrane is damaged, NPN is partitioned into the interior of the outer membrane, thereby exhibiting strong fluorescence in this hydrophobic environment. As shown in Fig. 3B and C, both peptides instantly permeabilized the outer membrane of *S. aureus ATCC25923* and *S. enteritidis CVCC3377* within 1 min, and higher concentrations of Lfcin4 and Lfcin5 resulted in a stronger NPN signal, indicating that the peptides made the outer membrane more permeable. These results suggest that both Lfcin4 and Lfcin5 induce a time- and concentration-dependent NPN fluorescence increase in intact bacterial cells and that the permeabilization capacity of Lfcin4 is higher than that of Lfcin5.
Permeabilization of the inner membrane in *S. aureus* and *S. enteritidis*. Propidium iodide (PI) is commonly used as a cell death marker because it is excluded by intact plasma membranes and can only enter cells that have lost their membrane integrity. The fluorescence of cells stained with PI was used as an indicator to evaluate cell membrane integrity and was measured by flow cytometry. In the absence of peptides, the percentage of *S. aureus* ATCC25923 and *S. enteritidis* CVCC3377 cells stained with PI was 0.04% and 0.76%, respectively, indicating the presence of intact healthy cell membranes. The percentage of PI-positive *S. aureus* ATCC25923 cells treated with 1×MIC Lfcin4 for 5 min, 30 min and 120 min was 4.30%, 44.20% and 50.10%, respectively (Fig. 3D); for *S. enteritidis* CVCC3377, Lfcin4 treatment resulted in 1.17%, 2.94% and 18.90% positive PI staining (Fig. 3F). A dose-dependent increase in PI fluorescence indicated that Lfcin4 damaged the cell membranes of *S. aureus* ATCC25923 and *S. enteritidis* CVCC3377. After treatment with 1×MIC Lfcin5, the percentage of PI-positive *S. enteritidis* CVCC3377 cells was 1.99% (5 min), 3.12% (30 min) and 21.30% (120 min); however, Lfcin5 did not permeabilize the inner membrane of *S. aureus* ATCC25923 (Fig. 3E and G), in agreement with the SEM images.

Lfcin4 and Lfcin5 bind to bacterial genomic DNA. An electrophoretic mobility shift assay (EMSA) was used to evaluate the binding affinity of each peptide to bacterial genomic DNA. Lfcin4 and Lfcin5 interacted with the genomic DNA (Fig. 4A, Supplementary Fig. S2). Lfcin4 impaired the migration of the genomic DNA from *E. coli* O157 and *S. aureus* ATCC25923 at mass ratios over 1.0 (peptide:DNA) and from *S. enteritidis* CVCC3377 at a mass ratio greater than 2.5, demonstrating the intrinsic DNA-binding ability of Lfcin4. For Lfcin5, no DNA bands were observed on the gel for *E. coli* or *S. aureus* at mass ratios of 10; however, a fraction of the genomic DNA of *S. enteritidis* migrated into the gel. This finding suggests that DNA-binding activity of Lfcin4 is higher than that of Lfcin5. The DNA-binding properties of peptides were further supported by the following CD assay.

As shown in Fig. 4B, C and D, the DNA CD spectra were dramatically changed when peptides bound to DNA from *E. coli*, *S. aureus* and *S. enteritidis*, which contained a negative band at approximately 250 nm and a positive band at approximately 275 nm. There was a greater decrease in the CD amplitude but no obvious shift, suggesting that Lfcin4 and Lfcin5 interact with genomic DNA and alter the DNA conformation.

Lfcin4 inhibits the synthesis of DNA, RNA and protein precursors. Despite the focus on bacterial membranes as targets for AMPs, several AMPs have additional intracellular targets. The incorporation of radioactive precursors into DNA, RNA, peptidoglycans, and proteins was used to evaluate the effects of Lfcin4 and
Lfcin5 on macromolecular synthesis in *S. aureus* ATCC25923 cells. Lfcin4 inhibited the synthesis of DNA, RNA and proteins compared to untreated controls. An inhibition of 30–50% was observed in *S. aureus* (Fig. 5B,C and D); increased label incorporation into peptidoglycans was also observed (Fig. 5A). Conversely, treatment with Lfcin5 appeared to promote all macromolecular biosynthetic pathways examined in *S. aureus* ATCC25923 cells (Fig. 5) but requires further study.

**Efficacy of Lfcin4 and Lfcin5 in vivo.** *Murine thigh infection model.* The neutropenic mouse thigh infection model has been extensively used to evaluate the *in vivo* antimicrobial activity of AMPs. The viable bacterial counts from infected thighs after treatment with peptides are shown in Fig. 6. Both Lfcin4 and Lfcin5 were
Effective at reducing the bacterial load in treated mice. At the start of therapy, in treated mice, 8.97 (the Lfcin4 group) and 9.07 (the Lfcin5 group) log10 CFU/g of \( S. \) aureus were detected in the thigh. Five hours after treatment, the bacterial counts in mice treated with Lfcin4 and Lfcin5 were all reduced by \(-0.01\) to \(-0.10\) and \(-0.01\) to \(-0.57\) log10 CFU/g, respectively, compared to untreated mice (Fig. 6). Ten hours after treatment, an increase of \(0.19\) to \(0.21\) log10 CFU/g was detected in untreated control mice. Treatment with 10 or 15 mg/kg Lfcin4 resulted in a reduced bacterial burden by \(0.03\) and \(0.10\) log10 CFU/g, respectively. This reduction was less than that observed following vancomycin treatment (\(-0.06\) log10 CFU/g) (Fig. 6A). At 5 h, treatment with 15 mg/kg Lfcin5 reduced the bacterial count by \(0.57\) log10 CFU/g. Treatment with 10 mg/kg vancomycin resulted in a decrease of \(0.1\) log10 CFU/g (Fig. 6B). At 10 h post-treatment with 15 mg/kg Lfcin5, approximately 76.6% of \( S. \) aureus cells had been killed. This indicates that treatment with 15 mg/kg Lfcin5 is more effective at reducing the bacterial burden than treatment with 10 mg/kg vancomycin (\(-1.2\)% and Lfcin4 (20.5%) (Fig. 6B).

Lfcin4 and Lfcin5 protect mice from lethal challenge with LPS. To evaluate the therapeutic activity of peptides in the endotoxemia model, mice were injected with colistin (15 mg/kg) or Lfcin4 or Lfcin5 (10, 15 and 20 mg/kg)
at 0.5 h, 8 h and 24 h after challenge with LPS. Mice in the control group were injected with PBS instead of LPS and survived the experimental period (Fig. 7A). The mice injected with LPS began to die 12 h after inoculation; all mice had succumbed within 24 h. Treatment with 10, 15 or 20 mg/kg Lfcin4 improved the survival rates to 75%, 100% and 100%, respectively. Mice treated with 10, 15 or 20 mg/kg Lfcin5 responded in a dose-dependent fashion, with survival rates of 50%, 62.5% and 75%, respectively. The survival rate of mice treated with 15 mg/kg

Figure 5. Effects of Lfcin4 and Lfcin5 on macromolecular synthesis in *S. aureus* ATCC25923. Incorporation of $^3$H-glucosamine hydrochloride (peptidoglycan) (A), $^3$H-leucine (protein) (B), $^3$H-thymidine (DNA) (C), and $^3$H-uridine (RNA) (D) was determined in cells treated with 1 × MIC Lfcin4 and Lfcin5. Van: vancomycin (2 × MIC); Ery: erythromycin (2 × MIC); Cip: ciprofloxacin (8 × MIC); Rif: rifampicin (4 × MIC). Antibiotics were used as controls. Results are presented as the mean ± SEM (n = 3).

Figure 6. Efficacy of Lfcin4 and Lfcin5 against *S. aureus* ATCC25923 in the neutropenic murine thigh infection model. (A) Lfcin4 treatment. (B) Lfcin5 treatment. Control: mice treated with saline only; Van (10 mg/kg): mice treated with a single intravenous (tail) dose of vancomycin (10 mg/kg); Lfcin4/Lfcin5 (10 mg/kg) and Lfcin4/Lfcin5 (15 mg/kg); mice treated with a single intravenous (tail) dose of 10 or 15 mg/kg Lfcin4 or Lfcin5, respectively. Results are presented as the mean ± SEM. Differences between groups were determined by one-way ANOVA followed by SPSS analysis (n = 4 per group). *p < 0.05 compared to the control group.
Figure 7. Effects of Lfcin4 and Lfcin5 on LPS-induced responses in vivo. (A) Groups of eight C57BL/6 mice were intraperitoneally injected with LPS from E. coli O111:B4 (30 mg/kg of body weight). Thirty minutes after LPS injection, Lfcin4, Lfcin5, colistin, or saline was administered intraperitoneally. Survival was recorded every 12 h and followed for up to 7 d. (B,C) C57BL/6 mice were intraperitoneally injected with 10 mg/kg LPS followed by intraperitoneal administration of 15 mg/kg Lfcin4, Lfcin5 or colistin 30 min later. Mice treated with buffer only served as a control. Cytokines were measured in blood from mice sacrificed at 2 h, 8 h or 20 h after LPS injection. The results are presented as the mean ± SEM. Differences between groups were determined by one-way ANOVA followed by SPSS analysis (n = 3 per group). #p < 0.05 compared to the control group and *p < 0.05 compared to the LPS-treated group (B). Light microscopy images (10× magnification, scale bar: 100 μm) of lung tissue from representative mice sacrificed at 8 h, 1 d, 4 d, and 7 d after LPS injection (C).
both Lfcin4 and Lfcin5 protect mice from lethal LPS challenge in vivo.

To determine whether the protective activities of Lfcin4 and Lfcin5 are associated with inflammatory cytokines, the serum levels of proinflammatory cytokines interleukin-6 (IL-6), interleukin-1β (IL-1β) and tumor necrosis factor-α (TNF-α) were determined in endotoxemic mice at different time points using an enzyme-linked immunosorbent assay (ELISA). As shown in Fig. 7B, the concentrations of serum IL-6, IL-1β and TNF-α were 779.5–115.8, 100.1–6.3 and 158.9–67.8 pg/ml, respectively, for endotoxemic mice treated with Lfcin4 and were 761.6–240.2, 155.2–47.7 and 173.8–78.3 pg/ml, respectively, for mice treated with Lfcin5. The levels of IL-1β and TNF-α were significantly lower than those from the corresponding LPS control groups (717.2–294.9, 258.0–193.2 and 330.3–120.5 pg/ml, respectively) and the colistin groups. These data suggest that treatment with either Lfcin4 or Lfcin5 inhibits the production of proinflammatory cytokines.

To determine whether Lfcin4 or Lfcin5 protects mice from LPS-induced lung injury, the lungs of mice were examined histologically at 8 h–7 d after treatment. As shown in Fig. 7C, no pathological change was found in the lungs of mice not injected with LPS, whereas LPS-challenged mice treated with saline developed acute lung injury, characterized by alveolar septum distention, inflammatory cells, an infiltration of red blood cells, and a certain degree of lung tissue distortion. In contrast, after treatment with Lfcin4 or Lfcin5, there was apparently less lung damage at 8 h–4 d and no obvious pathological differences in the lung tissues were observed at 7 d. The efficacy of both Lfcin4 and Lfcin5 was higher than that of colistin. These data demonstrate that, similar to colistin, both Lfcin4 and Lfcin5 protect mice from lethal LPS challenge in vivo.

**Discussion**

Lfcin is a multifunctional peptide that targets antibiotic-resistant bacterial strains, fungi, viruses, parasites, and tumors, as well as directly binds to the bacterial endotoxin LPS and is thereby already being used in different preclinical and clinical trials. In this work, we designed five 17-residue LfcinB17-31 derivatives (Trp-rich and Ala-rich) and first studied their antibacterial activities; Lfcin4 and Lfcin5 were found to be the most active peptides among those tested against both S. aureus and S. enteritidis, displaying more than 2- to 8-fold increases in antibacterial activity over the parent peptide, Lfcin1 (LfcinB17-31) (Table 2).

The α-helical conformation is necessary for the activity of some AMPs such as magainin and cecropin. In this study, there was also a positive correlation between the α-helical content and antibacterial activity of LfcinB17-31 derivatives. Both Lfcin4 and Lfcin5 had the highest α-helical content at 64.7% and the most potent antimicrobial activities (Tables 1 and 2), which may be related to Ala at positions 3, 7 and 14 since Ala is favorable for α-helix formation. Conversely, Lfcin6, with the lowest α-helical content, was inactive, wherein Arg and Met modifications in positions 4 and 10 by Trp resulted in a decrease in α-helix content from 60% to 35.2% (Tables 1 and 2). Furthermore, the CD spectra verified that both Lfcin4 and Lfcin5 contain an amphipathic α-helical structure (Fig. 1B and C, Supplementary Table S2). Therefore, it appears that the α-helical structure of the LfcinB17-31 derivatives is the most important structural parameter affecting antibacterial activity, while the antimicrobial activity of these peptides was less dependent on their positive charge.

Rekdal et al. reported that an increase in hydrophobicity leads to an increase in antibacterial activity. In our study, the enhanced activity of certain peptides may be due to the increased hydrophobicity by the replacement of Cys, Arg, Gln, Met, and Gly residues at positions 3, 4, 7, 10, and 14 in Lfcin1 by an aliphatic Ala or an aromatic Trp or Phe with larger hydrophobic side chains, which contribute largely to the depth of penetration into the hydrophobic core of a bilayer and the strength of interaction with negatively charged membranes, respectively. Lfcin4 and Lfcin5 contain the largest number of hydrophobic residues. These results are consistent with previous conclusions that the increase in activity of Lfcin peptides (LFB A3, LFB Q7 and LFB G14) may be due to increased hydrophobicity that comes with the replacement of Cys, Gln or Gly residues by Ala. Additionally, Trp is essential for the pore-forming properties of the peptide. LfcinB uses Trp solely as a hydrophobic anchoring force to help thread the peptide through the membrane by its hydrophobic character. However, it is also evident that there is an upper limit to the number of Trp residues that is tolerated, as peptide Lfcin6 (containing four Trp residues at positions 3, 6, 8, and 10) in this study exhibited much less antimicrobial activity than Lfcin4 (containing three Trp residues at positions 4, 6 and 8) and Lfcin5 (containing three Trp residues at positions 6, 8 and 10). This finding is in agreement with a previous study that reported that the LFB derivatives containing three additional Trp residues had lower antimicrobial activity than the LFB peptide with two Trp residues. Cation-π interactions and hydrogen bonds occur mainly between aromatic residues (e.g., Trp and Phe) and residues with positively charged side chains (e.g., Arg and Lys), which may allow the peptides to penetrate deeper into the membrane. All of these factors combined (i.e., α-helical content, hydrophobicity and intermolecular interactions) may have contributed to a more efficient activity of Lfcin4 and Lfcin5 against bacteria than their parent peptide.

The LfcinB peptide exerts a minor permeabilizing effect on the cytoplasmic membrane of susceptible bacteria; however, it does not lyse the bacteria. In our study, both Lfcin4 and Lfcin5 induced a concentration-dependent leakage of entrapped calcein, with the former having more potent interactions with negatively charged phospholipids than the latter. Additionally, the effects of Lfcin4 and Lfcin5 on Gram-negative bacteria differ from those on Gram-positive bacteria. Compared to Lfcin5, Lfcin4 demonstrates relatively stronger outer membrane permeabilization capacity against S. aureus and S. enteritidis. Moreover, Lfcin4 permeabilized the inner membrane of S. aureus and induced the leakage of cytoplasmic content, although Lfcin5 did not cause any significant damage to the integrity of the inner membrane; only shrinkage of cells was observed (Figs 2C and 3E). It is very interesting that the effect of Lfcin5 on S. enteritidis was more rapid and more profound than that of Lfcin4 (Fig. 3F and G), as observed by SEM (Fig. 2C). These results suggest a possible intracellular target for Lfcin4 and Lfcin5.
The interactions between the peptides and bacterial genomic DNA examined in our study imply that both Lfcin4 and Lfcin5 bind to genomic DNA from *S. aureus*, *S. enteritidis* and *E. coli* (Fig. 4). This result is contrary to the previous finding that LfcinB did not bind to pGEM-3Z/GAL plasmid DNA25. Moreover, we found that Lfcin4 inhibited DNA, RNA and protein synthesis in *S. aureus* whereas Lfcin5 did not (Fig. 5), which is similar to LfcinB17-41 and other Trp-rich peptides derived from human lysozymes that have been shown to translocate across lipid bilayers and inhibit macromolecular synthesis25,39. Therefore, it appears that Lfcin4 and Lfcin5 use multiple mechanisms to kill bacteria.

Conversely, in this study, Lfcin4 and Lfcin5 demonstrated excellent in vivo activity against *S. aureus* in mice (Fig. 6), as does human Lfcin1-1140. A dose-dependent effect was observed, where increasing doses of Lfcin4 and Lfcin5 from 10 to 15 mg/kg of body weight resulted in a greater reduction of viable bacteria in infected thighs of 0.85 log10 CFU/g of tissue compared to control mice (Fig. 6). We also showed that both Lfcin4 and Lfcin5 increased the survival of endotoxemic mice and protected the lungs from acute injury induced by LPS and that they were more effective than colistin (Fig. 7A and C). Moreover, these peptides significantly inhibited the release of cytokines (IL-6, IL-1β and TNF-α) in mice challenged with LPS (Fig. 7B). Together, these results suggest that both Lfcin4 and Lfcin5 are potential endotoxemia therapeutics.

In conclusion, we designed a series of LfcinB17-31 derivatives based on net charge, α-helical content and hydrophobicity. Both Lfcin4 and Lfcin5 showed stronger antimicrobial activity than the parent peptide and had very low hemolysis. Lfcin4 and Lfcin5 disrupt bacterial cell membranes and interact with bacterial DNA. DNA, RNA and protein synthesis were inhibited in *S. aureus* by Lfcin4 but not by Lfcin5. Both Lfcin4 and Lfcin5 protected mice from *S. aureus* infection and from LPS-induced lethality. Our findings strongly suggest that Lfcin4 and Lfcin5 (Trp-rich and Ala-rich) could serve as promising antimicrobial and anti-endotoxin agents for further clinical applications.

Materials and Methods

Peptide design. A series of LfcinB17-33 derivatives was designed based on several structural parameters, including net charge, α-helical content and hydrophobicity, by replacing the Cys residue at position 3 with Ala, Arg4 with Phe or Trp, Gla7 with Ala, Met10 with Trp, and Gly14 with Ala (Table 1). The physicochemical properties of these peptides were determined using the ProtParam tool on the ExPaSy server (http://web.expasy.org/protparam/) and the Antimicrobial Peptide Calculator and Predictor tool from the APD database (http://aps.unmc.edu/AP/prediction/protein_prediction_main.php). The α-helical index was calculated by NPSA (http://npsa-pbil.ibcp.fr/cgi-bin/npsa_automat.pl?page=npsa_hnn.html). The structures of the peptides were predicted by Emboss explorer (http://emboss.bioinformatics.nl/) and the I-TASSER sever (http://zhanglab.ccmb.med.umich.edu/I-tasser). Electrostatic surface potential was evaluated by PyMOL. All peptides were over 95% pure and were synthesized by SBS Genetech Co., Ltd. (Beijing, China) and ChinaPeptides Co., Ltd. (Shanghai, China), respectively.

Antibacterial and hemolytic activity. The MIC values of LfcinB17-33 derivatives against bacteria were determined by the microtiter broth dilution method41. Briefly, 10 μl of serially diluted aliquots of peptides were added to each well of 96-well microtiter plates, followed by the addition of 90 μl exponential phase bacteria (105 CFU/ml) and incubated for 16–18 h at 37 °C. The MIC value was determined as the lowest peptide concentration at which the peptide completely inhibited the growth of bacteria.

The hemolytic activities of Lfcin4 and Lfcin5 were evaluated based on the amount of released hemoglobin from erythrocyte suspensions of healthy mouse blood according to a previous method42. Blood cells were centrifuged at 2000 rpm for 5 min at 4 °C and washed three times with a saline solution (0.9% NaCl). Erythrocytes were resuspended in 0.9% NaCl, and 100 μl aliquots were mixed with 100 μl of peptides at different concentrations ranging from 2 to 1024 μg/ml. The mixtures were then added to 96-well plates. After incubation at 37 °C for 1 h, cells were centrifuged at 5000 rpm for 5 min, and the absorbance of the supernatant was measured at 540 nm on a microtiter plate reader. Controls for 0% and 100% hemolysis release were determined with 0.9% NaCl (A0%c) and 0.1% Triton X-100 (A100c), respectively. Hemolytic percentages are expressed as follows: hemolysis (%) = [1 – (Asample – A0%c)/(A100c – A0%c)] × 100%, where Asample is the absorbance value of each sample. All experiments were performed at least three times.

CD analysis. The secondary structures of Lfcin4 and Lfcin5 were investigated in ddH2O, a 50% TFE buffer and an SDS solution by CD spectroscopy. The CD spectra of Lfcin4 and Lfcin5 were determined on an MOS-450 spectropolarimeter (Bio-Logic, Grenoble, France) as previously described42. Peptides were dissolved in ddH2O, 5–40 mM SDS or a 50% TFE buffer. Samples were loaded into a quartz cuvette (1.0 mm path length), and spectra were recorded from 180 to 260 nm at 25 °C with a step resolution of 2.0 nm, a scanning speed of 100 nm/min and an integration time of 2 s.

Cell surface hydrophobicity assay. The effects of AMPs on the cell surface hydrophobicity of *S. aureus* ATCC25923 and *S. enteritidis* CVCC3377 were measured using the hexadecane partitioning method43. Cells were grown to stationary phase in Mueller-Hinton broth at 37 °C, harvested by centrifugation at 5000 rpm for 4 min, washed twice with sterile normal saline, and suspended in 0.1 M KNO3 (pH 6.2) to an OD600nm of 0.4. Lfcin4 or Lfcin5 was added to a final concentration of 1 ×, 2 × or 4 × MIC and incubated for 10 min at 37 °C. Sterile normal saline was used as a negative control, and the optical density at 600 nm was measured as OD600. Subsequently, a 1.2-ml cell suspension was mixed with 0.2 ml of hexadecane by vortexing for 1 min, and then, the mixture was incubated for 15 min. The OD600 nm value of the aqueous phase was determined as OD600. The hydrophobicity percentages were calculated by the following equation: hydrophobicity (%) = (1 – OD600/OD600) × 100%. All experiments were repeated three times.
SEM observations. Mid-log phase S. aureus ATCC25923 and S. enteritidis CVCC3377 suspensions (1 × 10^6 CFU/ml) were incubated with 1 × MIC Lfcin4 or Lfcin5 for 2 h at 37 °C. The samples were centrifuged at 4000 rpm for 5 min, washed three times with 0.1 M PBS (pH 7.2) and fixed with 2.5% glutaraldehyde at 4 °C for 2 h. Subsequently, the cells were washed three times with 0.1 M PBS and post-fixed with 1% osmium tetroxide (OsO4) for 2 h. The samples were dehydrated through a graded ethanol series (50–70–85–95–100%), CO2-dried, followed by platinum coating and then observed under a QUANTA200 SEM (FEI, Philips, Netherlands).

Membrane permeability assay. Effects of Lfcin4 and Lfcin5 on model membranes. Large unilamellar vesicles (LUVs) composed of POPC/POPG (1:3) were used as a model anionic membrane.41 LUVs were dissolved in chloroform, dried under nitrogen gas and vacuum-dried for 2 h. The lipid film was rehydrated in 20 mM HEPES (150 mM NaCl, 1 mM EDTA, pH 7.4). To make calcein-encapsulated unilamellar liposomes, the lipid film was suspended in 5 mM sodium HEPES (containing 100 mM calcein, pH 7.5). The liposome suspensions were freeze-thawed in liquid nitrogen five times and extruded ten times through two stacked polycarbonate filters (100 nm pore size) in an Avanti Mini-Extruder. Untrapped calcein was removed using a Sephadex G-50 column (Amersham Pharmacia Biotech, Little Chalfont, UK). Peptides were then added to the LUVs. After a 10-min incubation at room temperature, the release of calcein form liposomes was monitored for 10 min on a Tecan Infinite M200 PRO plate reader (TECAN, Austria) with an excitation wavelength of 485 nm and an emission wavelength of 535 nm. The total calcein fluorescence (Ft) was determined by the addition of 0.1% Triton X-100. Dye leakage was calculated according to the following equation: leakage rate (%) = ((Ft - Fs)/(Ft - F0)), where F0 indicates the fluorescence of each sample at T = 0.

Effects of Lfcin4 and Lfcin5 on model membranes. The effects of Lfcin4 and Lfcin5 on the outer membrane of S. aureus ATCC25923 and S. enteritidis CVCC3377 were determined by measuring the uptake of NPN.44,45 Mid-log phase cells were washed with sterile normal saline three times and suspended in 5 mM HEPES buffer (pH 7.2, containing 5 mM glucose) to an OD at 600 nm of 0.5. Subsequently, NPN (10 mM) and peptide (1 ×, 2 × or 4 × MIC) solutions were added to 96-well plates. The relative fluorescence intensity was measured using an Infinite M200 PRO plate reader. The excitation and emission wavelengths were set at 328 and 438 nm, respectively. Treatment with PBS served as a negative control, and treatment with ampicillin or colistin was used as a positive control.

Effects of Lfcin4 and Lfcin5 on the inner membrane. The permeabilization of the inner membrane was performed according to a previous report by Li et al. with slight modifications.39 S. aureus ATCC25923 and S. enteritidis CVCC3377 cells in mid-log phase were washed with 10 mM PBS buffer three times, resuspended in the same buffer (1 × 10^5 CFU/ml), and incubated with or without 1 × MIC peptides at 37 °C for 5, 30 and 120 min. Bacterial cells were then washed twice with PBS, fixed with 50 μg/ml PI and analyzed using a BD FACS Calibur Flow Cytometer (Franklin Lakes, New Jersey, USA). Data were analyzed using CellQuest Pro software (BD, USA).

Effects of Lfcin4 and Lfcin5 on bacterial genomic DNA. Genomic DNA was extracted from S. aureus ATCC25923, E. coli O157 and S. enteritidis CVCC3377 using a TIANamp Bacteria DNA Kit (TIANGEN Biotech Co., Ltd., Beijing). The gel retardation experiment was performed by mixing the bacterial DNA with different concentrations of Lfcin4 or Lfcin5 in 20 μl binding buffer (10 mM Tris-HCl, pH 8.0, 5% glycerol, 1 mM dithiothreitol, 1 mM EDTA, 200 mM KCl, and 50 μg/ml bovine serum albumin).41 The ratios of peptide to DNA were 0, 0.5, 1, 2.5, 5.0, and 10.0 (w/w). The mixtures were incubated for 10 min at 37 °C and then subjected to electrophoresis on a 0.7% agarose gel.

To examine whether peptide binding causes secondary structure changes in bacterial DNA, CD measurements were carried out on an MOS-450 spectropolarimeter using a quartz cuvette with a 1.0 mm path length. The ratios of peptide to bacterial DNA were 0, 5 and 10. The spectra were recorded from 220 to 320 nm at 25 °C. The CD data represent the average of 10 scans with a 20 s bandwidth.

Effects of Lfcin4 and Lfcin5 on macromolecular synthesis. The effects of each peptide on the incorporation of L-[methyl-3H] thymidine, [5-3H] uridine, D-[6-3H(N)] glucosamine hydrochloride, and L-[3,4,5-3H] leucine into DNA, RNA, peptidoglycans, and proteins were investigated in S. aureus ATCC25923. Mid-log phase S. aureus cells (10^6 CFU/ml) were incubated with 1 × MIC peptide or antibiotics at 37 °C for 15 min. Radioactive precursors of [H] thymidine, [H] uridine, [H] leucine, and [3H]-glucosamine (40 μCi/ml) were added to cultures to monitor macromolecular synthesis and incubated for 20 min at 37 °C. Cold 25% trichloroacetic acid (TCA) was added to the mixture and placed on ice for 30 min. After centrifugation, pellets were washed twice with 25% TCA, dried and counted with scintillation fluid on a MicroBeta 1450 scintillation counter (Perkin Elmer)49.

In vivo experiments. Animal care and all experimental protocols were approved by the Laboratory Animal Ethical Committee and its Inspection of the Feed Research Institute of Chinese Academy of Agricultural Sciences (AEC-CAAS-20090609) and were performed in accordance with ARRIVE guidelines.67.

Thigh infection model. The thigh infection protocol was performed as previously described.68 Female BALB/c mice (6 weeks old, 22 ± 2 g) were rendered neutropenic (neutrophils, 100/mm^3) by intraperitoneal injection with 150 and 100 mg cyclophosphamide per kg of body weight for 4 d and 1 d before infection, respectively. Mice were inoculated with 100 μl of S. aureus ATCC25923 cells (1.0 × 10^7 CFU/ml) and given a single dose (0.2 ml) of Lfcin4 or Lfcin5 (10 or 15 mg/kg of body weight) or vancomycin (10 mg/kg of body weight) by tail vein injection at 2 h post-infection. Mice were humanely euthanized by cervical dislocation at 5 and 10 h after treatment. Thighs were removed and homogenized in sterile PBS for a CFU assay for S. aureus ATCC25923.
LPS-induced endotoxemia model. Male C57BL/6 mice (8 weeks old, 22 ± 2 g) were intraperitoneally injected with 30 mg/kg of LPS (0.2 ml) from E. coli O111:B4 cells as previously described15. Mice were intraperitoneally injected with Lfcin4 or Lfcin5 (10, 15 or 20 mg/kg of body weight in 0.2 ml) or colistin (15 mg/kg of body weight in 0.2 ml) at 0.5, 8 and 24 h after inoculation with LPS, respectively. Mice injected with LPS or saline only served as positive or mock-treated controls, respectively. Survival was recorded every 12 h and followed for up to 7 d.

Serum was separated from mice at 2, 8 and 20 h following the administration of LPS (10 mg/kg). The levels of IL-6, IL-1β and TNF-α in endotoxemic mice were detected at Jiaxuan Biotech. Co., Ltd. (Beijing, China) using an ELISA kit according to the manufacturer’s protocol.

Thirty minutes after intraperitoneal injection of LPS (10 mg/kg of body weight in 0.2 ml), mice were intraperitoneally injected with Lfcin4, Lfcin5 or colistin (15 mg/kg of body weight in 0.2 ml). Mice were sacrificed at 8 h, 1 d, 4 d, and 7 d after LPS injection. The lungs were removed, washed with PBS and fixed in 4% paraformaldehyde at 4 °C for 24 h. After they were washed with PBS and dehydrated with a graded series of ethanol (75–95%), the tissues were infiltrated with xylene and embedded in paraffin wax. Sections were cut, stained with hematoxylin and eosin and examined under a light microscope. Mice injected with LPS or only PBS served as positive or mock-treated controls, respectively.

**Statistical analysis.** Statistical analysis was performed using software SPSS21.0 (SPSS, USA). Data are expressed as the mean ± standard error of the mean (SEM). One-way repeated analysis of variance (ANOVA) (Institute Inc., Cary, NC, USA), Duncan method and the Mann-Whitney rank test were used to determine the statistical significance. A p value of <0.05 was considered statistically significant.

**References**

1. Diekema, D. J. et al. Survey of infections due to *Staphylococcus* species: frequency of occurrence and antimicrobial susceptibility of isolates collected in the United States, Canada, Latin America, Europe, and the Western Pacific region for the SENTRY Antimicrobial Surveillance Program, 1997-1999. *Clin. Infect. Dis.* 32 Suppl 2, S114–S132 (2001).

2. Wang, Z., Cao, B., Liu, Y. M., Gu, L. & Wang, C. Investigation of the prevalence of patients co-colonized or infected with methicillin-resistant *Staphylococcus aureus* and vancomycin-resistant enterococci in China: a hospital-based study. *Chin. Med. J. (Engl.)* 122, 1283–1288 (2009).

3. Maiti, S. et al. Effective control of *Salmonella* infections by employing combinations of recombinant antimicrobial human β-defensins hBD-1 and hBD-2. *Antimicrob. Agents Chemother.* 58, 6898–6903 (2014).

4. Lu, Q. et al. Pathogen and antimicrobial resistance profiles of culture-proven neonatal sepsis in Southwest China, 1990-2014. *J. Paediatr. Child Health* 52, 939–943 (2016).

5. Miller, M. F. & Loch-Caruso, R. Comparison of LPS-stimulated release of cytokines in punch versus transwell tissue culture systems of human gestational membranes. *Reprod Biol. Endocrinol.* 8, 121 (2010).

6. Kalle, M. et al. Host defense peptides of thrombin modulate inflammation and coagulation in endotoxin-mediated shock and *Pseudomonas aeruginosa* sepsis. *PloS One* 7, e51313 (2012).

7. Russell, S., Hayes, M. A., Simko, E. & Lumsden, J. S. Plasma proteomic analysis of the acute phase response of rainbow trout (*Oncorhyncus mykiss*) to intraperitoneal inflammation and LPS injection. *Dev. Comp. Immunol.* 30, 393–406 (2006).

8. Tomita, M. et al. Potent antibacterial peptides generated by pepsin digestion of bovine lactoferrin. *J. Dairy Sci.* 74, 4137–4142 (1991).

9. Bellamy, W., Takase, M., Wakahayashi, H., Kawase, K. & Tomita, M. Antibacterial spectrum of lactoferrin B, a potent bactericidal peptide derived from the N-terminal region of bovine lactoferrin. *J. Appl. Bacteriol.* 73, 472–479 (1992).

10. Yamauchi, K., Tomita, M., Giehl, T. J. & Ellison, R. T. 3rd. Antibacterial activity of lactoferrin and a pepsin-derived lactoferrin peptide fragment. *Infect. Immun.* 61, 719–728 (1993).

11. Gifford, J. L., Hunter, H. N. & Vogel, H. J. Lactoferricin: a lactoferrin-derived peptide with antimicrobial, antiviral, antitumor and immunological properties. *Cell Mol. Life Sci.* 62, 2588–2598 (2005).

12. Jenssen, H. & Hancock, R. E. W. Antimicrobial properties of lactoferrin. *Biochemie* 91, 19–29 (2009).

13. Oo, T. Z., Cole, N., Garthwaite, L., Willcox, M. D. & Zhu, H. Evaluation of synergistic activity of bovine lactoferricin with antibiotics in corneal infection. *J. Antimicrob. Chemother.* 65, 1243–1251 (2010).

14. Liu, Y., Han, E., Xie, Y. & Wang, Y. Comparative antimicrobial activity and mechanism of action of bovine lactoferricin-derived synthetic peptides. *Biometals* 24, 1069–1078 (2011).

15. Silva, T. et al. Structural diversity and mode of action on lipid membranes of three lactoferrin candidacidal peptides. *Biochim. Biophys. Acta* 1828, 1329–1339 (2013).

16. Puñkun, A. et al. A heterodimer comprised of two bovine lactoferrin antimicrobial peptides exhibits powerful bactericidal activity against *Burkholderia pseudomallei*. *World J. Microbiol. Biotechnol.* 29, 1217–1224 (2013).

17. Wang, S. et al. The effect of Lfcin-B on non-small cell lung cancer H460 cells is mediated by inhibiting VEGF expression and inducing apoptosis. *Arch. Pharm. Res.* 38, 261–271 (2015).

18. Rekdal, Ø., Andersen, J., Vorland, L. H. & Svendsen, J. S. Construction and synthesis of lactoferricin derivatives with enhanced antibacterial activity. *J. Peptide Sci.* 5, 32–45 (1999).

19. Schibili, D. J., Hwang, P. M. & Vogel, H. J. The structure of the antimicrobial active center of lactoferricin B bound to sodium dodecyl sulfate micelles. *FEBS Lett.* 446, 213–217 (1999).

20. Strom, M. B., Rekdal, Ø. & Svendsen, J. S. Antibacterial activity of 15-residue lactoferricin derivatives. *J. Peptide Res.* 56, 265–274 (2000).

21. Strom, M. B. et al. Important structural features of 15-residue lactoferricin derivatives and methods for improvement of antimicrobial activity. *Biochem. Cell Biol.* 80, 65–74 (2002).

22. Silva, T. et al. Killing of *Mycobacterium avium* by lactoferricin peptides: improved activity of arginine- and D-amino-acid-containing molecules. *Antimicrob. Agents Chemother.* 58, 3461–3467 (2014).

23. Vorland, L. H., Ulvatne, H., Rekdal, Ø. & Svendsen, J. S. Initial binding sites of antimicrobial peptides in *Staphylococcus aureus* and *Escherichia coli*. *Scand. J. Infect. Dis.* 31, 467–473 (1999).

24. Vorland, L. H. et al. Interference of the antimicrobial peptide lactoferricin B with the action of various antibiotics against *Escherichia coli* and *Staphylococcus aureus*. *Scand. J. Infect. Dis.* 31, 173–177 (1999).

25. Ulvatne, H., Samuelsén, Ø., Haukland, H. H., Krämer, M. & Vorland, L. H. Lactoferricin B inhibits bacterial macromolecular synthesis in *Escherichia coli* and *Bacillus subtilis*. *FEBS Microbiol. Lett.* 237, 377–384 (2004).

26. Ho, Y. H., Sung, T. C. & Chen, C. S. Lactoferricin B inhibits the phosphorylation of the two-component system response regulators BasA and CreB. *Mol. Cell. Proteomics* 11, M111.014720 (2012).

27. Haug, B. E. & Svendsen, J. S. The role of tryptophan in the antibacterial activity of a 15-residue bovine lactoferricin peptide. *J. Pept. Sci.* 7, 190–196 (2001).
28. Kovacs, J. M., Mant, C. T. & Hodges, R. S. Determination of intrinsic hydrophobicity/hydrophobicity of amino acid side chains in peptides in the absence of nearest-neighbor or conformational effects. Biochim. Biophys. Acta 1668, 175–189 (2005).
29. Brouwer, C. P. & Welling, M. V. Various routes of administration of 99mTc-labeled synthetic lactoferrin antimicrobial peptide hLF and its antimicrobial activity. J. Nucl. Med. 49, 533–539 (2008).
30. Teng D. et al. A dual mechanism involved in membrane and nucleic acid disruption of AvBD103b, a new avian defensin from the king penguin, against Salmonella enteritidis. Cytoskeleton 69, 144–155 (2012).
31. Cho, J. & Lee, D. G. The characteristic region of arenicin-1 involved with a bacterial membrane targeting mechanism. Biochem. Biophys. Res. Commun. 405, 422–427 (2011).
32. Pavia, K. E., Spinella, S. A. & Elmore, D. E. Novel histone-derived antimicrobial peptides use different antimicrobial mechanisms. Biochem. Biophys. Res. Commun. 418, 133–138 (2012).
33. Han, F. E. et al. Comparing bacterial membrane interactions and antimicrobial activity of porcine lactoferrin-derived peptides. J. Dairy Sci. 96, 3471–3478 (2013).
34. Li, L., Shi, Y., Su, G. & Le, G. Selectivity for and destruction of Salmonella typhimurium via a membrane damage mechanism of a cell-penetrating peptide pPG20 analogue. Int. J. Antimicrob. Agents 40, 337–343 (2012).
35. Xiong, Y. Q., Bayer, A. S. & Yeaman, M. R. Inhibition of intracellular macromolecular synthesis in Staphylococcus aureus by thrombin-induced platelet microbicidal proteins. J. Infect. Dis. 185, 348–356 (2002).
36. Kilkenny, C., Browne, W. J., Cuthill, I. C., Emerson, M. & Altman, D. G. Improving bioscience research reporting: the ARRIVE guidelines for reporting animal research. PLOS Biol. 8, e1000412 (2010).
37. Zhang, Y. et al. In vitro and in vivo characterization of a new recombinant antimicrobial peptide, MP1102, against methicillin-resistant Staphylococcus aureus. Appl. Microbiol. Biotechnol. 99, 6255–6266 (2015).
38. Hu, L., Sun, C., Wang, S., Su, F. & Zhang, S. Lipopolysaccharide neutralization by a novel peptide derived from phosvitin. Int. J. Biochem. Cell Biol. 45, 2622–2631 (2013).

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Author Contributions
X.M.W., D.T. and J.H.W. conceived and designed the research. Y.H., R.Y.M., N.Y., and X.W. developed the CD, FACS and SEM methods, and interpreted the results. Y.H. performed the hemolysis experiment. N.Y. provided help and support with MIC and mice experiments. Z.Z.L. analyzed the peptide structures. Y.H., X.M.W. and J.H.W. wrote the manuscript. All authors reviewed the manuscript.

Additional Information
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