Platelet-derived growth factors induced lymphangiogenesis: evidence, unanswered questions and upcoming challenges

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Abstract
Crosstalk between angiogenesis and lymphangiogenesis in embryonic development continues during postnatal life and has specific mechanisms involving factors that initiate activation of the intracellular cascade for their specific receptors. Platelet-derived growth factors (PDGFs) and their corresponding receptors (PDGFRs) are known as important regulators of blood vessel development in both normal and pathologic angiogenesis. Despite some recent papers which reported a potential role of the PDGF/PDGFR axis in lymphatic spread of tumor cells, a few papers have suggested the potential role of PDGFs in tumor lymphangiogenesis development. The present paper summarizes the potential lymphangiogenic role of the PDGF/PDGFR axis, underlying upcoming challenges in the field.

Key words: lymphangiogenesis, platelet-derived growth factor receptor, platelet-derived growth factor, tumor microenvironment.

PDGF/PDGFR system: structure and signaling pathways
The platelet-derived growth factor (PDGF) family was identified in platelets and in serum as fibroblast, smooth muscle cell and glia culture cell mitogen factors [1]. Four members of this family have been evidenced, noted as A, B, C and D, the latter two types being recently discovered. The active form of the PDGF presents as a homodimer or a heterodimer; in the PDGF family four homodimers (PDGF-AA, PDGF-BB, PDGF-CC and PDGF-DD) and only one heterodimeric form (PDGF-AB) are recognized. The PDGF dimers are assembled within the cell [2]. Configurations of certain types have the assembly of isotypes A and B accomplished without a specific orientation [3, 4]. This is the reason behind a great deal of structural similarities between the A and B type of PDGF receptors [5]. Certain cells may therefore produce PDGF-AA or PDGF-BB, if any of the genes that encode PDGF synthesis are translated in a singular fashion [5]. In case of simultaneous transcription of both the genes, the result will be a mixture that will include PDGF-AA, PDGF-BB and PDGF-AB [4]. In addition to this, noticing the primary sequence of the polypeptide chain, it is seems that PDGF-C and -D are not strongly related to PDGF-A and -B, which explains the impossibility to form heterodimers with the two isotypes [5]. The possibility to form heterodimers between types C and D remains at the level of a hypothesis [1]. PDGF-C and PDGF-D possess an...
N-terminal domain named the CUB domain with a function unknown for the time being. It appears that during the growth factor secretion through proteolytic processing, the CUB domains are not required.

There are two binding receptors for PDGFs, named PDGFRα and PDGFRβ. PDGFR-dependent receptor dimerization is at first needed for signal auto-phosphorylation and transmission [1]. PDGFRβ presents high affinity for PDGF-B and PDGF-D [6]. PDGFRα has high specificity for PDGF-A [7] and PDGF-C [8] but also for PDGF-B (the receptor affinity being greater for PDGF-A than for PDGF-B). The gene expression for the two receptor types might be in an individual manner or together in a single cell [1]. The PDGF signaling receptor structure unity may be PDGFRαα, PDGFRββ or PDGFRαβ. This highly depends on the gene expression for the PDGF receptor pattern, as well as on the viability of different family dimers. The studies carried on PDGF receptors have been based especially on the β form. Through the intracellular residues of phosphotyrosine, receptor dimers come in contact with a certain number of SH2 domains containing molecules that ensure the bond with a number of signaling pathways [1, 9]. In the cytoplasmic matrix the protein categories which bind PDGFRβ, namely PLC-γ, PI3K [10], Ras, GAP SHP2 phosphatase and the family of Src kinases, are included [1, 9, 11]. These pathways seem to be in connection with genes that activate immediately [9]. The phospho-tyrosine residues within PDGFRβ molecules are implicated in binding and activating a certain signaling pathway. The differences that appear in receptor binding are found in the case of the Ras GAP pathway, which binds only PDGFRβ, but not PDGFRα, and in the case of the Crk adaptor protein family, which binds only PDGFRα [1]. Receptors have different roles during development [12], an aspect that is mostly associated with pattern expression. It is unknown whether the role played by the β type (in the case of its absence) could be totally compensated by the presence of the α form.

The PDGF stimulates cells in a specific manner, through binding to the cell surface receptors, and that determine DNA synthesis and cell proliferation [7, 13]. The PDGFR rather than the factor itself may possess activities or sites needed for proliferation response initiation in target cells. There is evidence that if the cells are preincubated with PDGF, the activity of PDGFR is much reduced. The number of binding sites decreases to 10% after the cell is pre-incubated with 200 ng/ml of PDGF. Dimer BB of the PDGF family is expressed by endothelial cells (ECs) and is implicated in signal induction through binding only to PDGFRβ expressed by perivascular cells and ECs [14]. However, numerous experimental studies have shown that PDGF is expressed by ECs within a temporal-spatial pattern. The PDGF-B endothelial expression is at first largely spread, but afterwards it becomes restricted to ECs from the sprouting tips [14]. It seems that the tip cell maintains a tight contact with pericytes. This contact is a key element that facilitates PDGF-B release.

Pericyte recruitment is one of the essential levels which concerns angiogenesis induced by PDGF family members, and a phenomenon that completely depends on the PDGF-B/PDGFRβ system signaling pathway [15]. This affinity might be an explanation for almost complete loss of pericytes in the nervous system (and also in other organs) in the absence of the B dimer or PDGFRβ. Also in angiogenesis, the function of PDGF-B is expressed much better in the arterial endothelium than in the venous one [14].

Implication of PDGFs and PDGFRs in pathological lymphangiogenesis and lymph node metastasis

The PDGF-BB is a direct lymphangiogenic factor that stimulates lymphatic metastasis [16–19]. The PDGFRα as well as PDGFRβ expressed in an isolated manner in lymphatic endothelial cells (LECs) and all three PDGF prototypes (AA, BB and CC) are capable of inducing lymphangiogenesis [20]. In addition, the growth factors from the PDGF family, as well as the hepatocyte growth factor, have a stimulating effect on the vascular endothelial growth factor family members (with proangiogenic and lymphangiogenic effects) [21–24]. It seems that within the vascular endothelial growth factor (VEGF) family, dimer A has in itself a stimulating effect on PDGF-B expression in ECs [25, 26], while FGF-2 stimulates the perivascular expression of PDGFRβ.

Co-stimulation of PDGFs through both VEGF and fibroblast growth factor (FGF) actions determines the recruitment of perivascular cells in vitro as well as determining the formation of a functional vasculature in vivo. In order to accomplish these two effects of PDGF, the preservation of the PDGF peri-endothelial gradient is of a great importance. The two effects of PDGF are suppressed by neutralizing antibody action as well as by PDGF of exogenous origin [14]. In vitro, PDGF-BB stimulates MAP-kinase activity as well as the motility of isolated LECs [18]. In vivo, this factor induces lymph vessel growth. PDGF-BB expression in murine fibrosarcoma induces tumor lymphangiogenesis, leading to lymph node metastasis [18]. Tumor cells have the capacity to induce blood vessel and lymphatic vessel formation for the tumor itself.

Besides PDGF family members, of great importance in this matter is the intervention of the vascular endothelial growth factor (VEGF-C and VEGF-D),
which interacts with a lymphatic endothelial receptor VEGFR-3. As well as in the case of PDGF-B, both VEGF-C and VEGF-D are overexpressed during lymphatic metastasis in several cancer types including pancreatic and gastric cancer [27, 28].

In recent studies performed in order to evidence the new mechanisms regarding pathological lymphangiogenesis, it has been observed that PDGF-BB expression in primary transplanted murine sarcomas leads to lymph node metastasis in the mouse model [29]. Of all PDGF family members, it seems that only PDGF-BB is capable of activating both α and β receptors.

In order to experimentally demonstrate the direct implication of PDGF-BB in tumor lymphangiogenesis and lymph node metastasis, a murine lymphangiogenic model has been used. By using a laboratory lymphangiogenic corneal mouse model together with confocal microscopy, it has been demonstrated that PDGF-BB is capable of inducing lymphangiogenesis in the mouse cornea. In normal conditions, the corneal tissue does not possess blood or lymphatic vessels, but after the implantation of the growth factor, in about 2 weeks the corneal vasculature was well developed. The inclusion of homodimer B determines the development of new lymphatic vessels but with a diameter above the average. New blood vessels started from pre-existing vessels located in the limbus [29]. It was demonstrated that PDGF-AA as well as PDGF-AB presents similarities from this point of view with PDGF-BB, although with some differences. It seems that unlike PDGF-BB, PDGF-AA determined the formation of a lesser number of lymphatic vessels, while the other member of the family induced the formation of a similar number as PDGF-BB. The topic of discussion would be, in this case, the importance of the β receptor considering that both PDGF-AB and PDGF-BB (but not PDGF-AA), play the ligand role in this case. Therefore, PDGF-BB is the factor with a higher lymphangiogenic potential compared to the rest of the family members. The phenomenon of PDGF-BB-induced lymphatic vessel formation is based on a remodeling process.

The direct role of PDGF-BB in lymphangiogenesis sustains the fact that its action does not depend on the VEGF-C/D/VEGFR-3 pathway. Nor does it depend on the levels of angiopoietin 2, which are not affected by cell stimulation with PDGF-BB cells that express PDGFR [29, 30].

The pathway through PDGFs acts in order to induce lymphangiogenesis depending on various intracellular signaling components, common for PDGF-BB and PDGF-AA but also for VEGF-C [4, 20].

The Prox-1 gene is responsible for the lack of venous vessel conversion to a lymphatic vessel; therefore embryos will present a lack of lymphatic vessel structure [31-33]. There is no connection between Prox-1 gene expression and PDGF-BB level, so overexpression of the gene does not induce high levels of PDGF-BB. Therefore, the manner through which PDGF-BB acts for lymphatic vessel development and differentiation uses other pathways than those activated by Prox-1. Despite all these aspects, some authors support a different hypothesis regarding the relationship between Prox-1 and PDGF-BB. Initially, the Prox-1 gene induces lymphatic vessel development, starting from the venous ECs. Some papers assume the existence of independent control of the Prox-1 gene, which regulates lymphatic vessel growth, a development that probably is led by PDGF system activity, later. All tumor types were able to induce lymphatic vessel development, but only PDGF-BB positive tumors have intratumor lymphatic vessels, which infiltrate deeper in the tumor tissue and may facilitate lymphatic metastasis through various mechanisms [19]. Therefore, the main functional implications of PDGF-BB as a direct endogenous pro-lymphangiogenic factor but also as a lymphatic metastasis promoter are on one hand the direct stimuli of tumor cell growth and on the other hand inducers of angiogenesis and lymphangiogenesis.

Recent studies have suggested the role of PDGF-A in tumor invasion, lymph node metastasis and poor prognosis in different types of malignancies. Donnem et al. [34] found for the first time in 2010 a significant association between PDGF-A expression and lymphatic spread in non-small cell lung carcinoma, together with co-expression of PDGF-B and VEGFR-3. This first direct evidence about PDGF-A involvement in lymphangiogenesis and lymphatic metastasis was strongly supported by recently published papers about PDGF-A and PDGFRα involvement as predictors of renal tumors. The PDGFRα is constitutively expressed in proximal and distal convoluted tubules and collecting ducts of normal renal parenchyma, being also positive in clear cell renal cell carcinoma and papillary renal cell carcinoma [35]. Also, limited data are available about the role of the PDGF-A/PDGFRα axis in nephroblastomas [36]. Together with PDGF-BB induction of lymphangiogenesis in the tumor microenvironment, PDGF-AA is also involved in tumor metastasis induction and progression, being an important factor in the recruitment of metastatic cells and their commitment through a favorite metastatic site. Metastatic preference of breast and prostate cancer cells for bone is explained by PDGF-AA secretion from osteoclasts and osteoblasts followed by recruitment of highly metastatic PDGFRα positive variants of tumor cells [37-40].

Scattered and controversial data about the role of PDGF-AB in tumor lymphatic vessel development are available at this moment. It seems that
PDGF-AB has a potential role in tumor lymphangiogenesis by acting through cancer-associated fibroblast and tumor-associated macrophage recruitment followed by activation of both PDGFRα and PDGFRβ through an inflammatory mediated mechanism [41, 42]. Certified or potential roles of PDGF family members and their known (for PDGF BB/PDGFRβ) and still hypothetical (for other PDGF family members) mechanisms of action are summarized in Figure 1.

**The role of PDGF-B and PDGFRβ in human gastric carcinoma lymphangiogenesis**

Functional implications of the PDGF/PDGFR axis in pathological lymphangiogenesis concern human gastric carcinoma. The PDGF-B and PDGFRβ RNA expression was found significantly higher in the case of patients who presented lymphatic metastasis as well as in the case of patients with diffuse human gastric carcinoma (mainly intestinal type gastric carcinoma) [43]. The PDGF plays a direct role not only in tumor angiogenesis but also in lymphangiogenesis. The PDGF stimulates tumor growth by tumor cell growth induction and is implicated in the stimulation of angiogenesis, and pericyte recruitment. Apart from this, it successfully influences the transendothelial transport of some chemotherapeutic agents in a paracrine manner [43]. After the quantitative mRNA analysis for PDGF-B and PDGFRβ by means of real-time PCR, it can be concluded that patients with positive lymph node metastasis had significantly higher levels of PDGF-B/PDGFRβ than those who presented negative lymph nodes. In the case of diffuse human gastric carcinoma, it had a higher probability of lymphatic metastasis, this being a basic characteristic that differentiates it from other types of gastric tumors and tumors that affect the digestive system in general. Therefore, patients with such a type of gastric tumor had much higher levels of PDGFs. Hence, it is possible to state that there is an important association between tumor severity and PDGF levels.

**PDGF-A, tumor lymphangiogenesis and esophageal carcinoma**

In squamous cell carcinoma of the esophagus, the exact role played by regulating growth factors still remains unknown. A significant increase of lymphatic vessel density in the case of VEGF-A and VEGF-C was observed [44, 45]. In addition to this, high expression of mRNA for VEGF-A, VEGF-C and PDGF-A is observed. Both the density of lymphatic vessels and that of blood vessels were significantly higher.

The PDGFs induce the growth of the tumor through direct stimulation of cell growth from different types of tumors. Experimental studies on angiogenesis and lymphangiogenesis were based especially on evidencing the functional implications of the vascular endothelial growth factor. In contrast, as per the recent studies, it is evident that PDGFs are a class of potential angiogenic factors. Particularly, the angiogenic activity of PDGFs is stimulated by the presence of other proangiogenic factors [14].

**Breast cancer, PDGFs and PDGFRs**

Known and extensively studied as promoters of bone marrow metastases in breast cancer [46–49], PDGFs and corresponding PDGFRs are considered, currently, as potential targets for antitumor and anti-metastatic drugs rather than antiangiogenic or anti-lymphangiogenic targets. Indirect evidence about the role of PDGFs/PDGFRs in breast cancer lymphangiogenesis and breast cancer cell spreading was found in the literature. The PDGF-D is highly expressed in human breast cancer and facilitates tumor growth and lymph node metastasis, making it a potential target in breast cancer. At the same time, PDGF-D increases drug delivery and hence improves the efficacy of chemotherapy through vessel normalization [50, 51]. Some studies have reported a correlation between nodal metastases, peritumoral and intratumoral lymphatic microvessel density in breast cancer, but direct evidence about the PDGF/PDGFR axis in lymphangiogenic onset and support in breast malignancies has not been provided. Recently, Schito et al. reported that human LECs start to proliferate and migrate when they are co cultured with breast cancer cells expressing HIF-1α and PDGF-B and demonstrated colocalization of HIF-1α and PDGF-B, which were correlated with lymphatic vessel area and histological grade by immunohistochemical analysis of paraffin-embedded human breast cancer specimens [52]. These data are supplemented by Semenza (2013), who described the cancer-stromal cell interactions mediated by hypoxia-inducible factors able to promote angiogenesis, lymphangiogenesis, and metastasis by PDGFR activation together with other stromal factors [53]. Limited data support the PDGF-B/PDGFRβ axis as a tumor promoter from in situ carcinoma to the invasive stage of breast cancer, but none have concerned its involvement in the early steps of lymphangiogenesis [54]. Our preliminary immunohistochemical studies showed high PDGF-B, PDGFRα and PDGFRβ expression during progression from in situ carcinoma to invasive, metastatic breast cancer in both tumor cells and surrounding lymphatic vessels (Figure 2).

No data about PDGF/PDGFR involvement in the lymphangiogenic process from molecular types of breast cancer are available now.
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**Figure 1.** PDGFs' cellular sources, their isoforms and known (for PDGF-B, continuous arrow) or potential (for other forms, marked with dotted arrow) mechanism of action for inducing lymphangiogenesis in tumor conditions with effects on tumor cells and metastasis development and progression

**Implications of PDGFs in lymphatic vessel remodeling**

The basis of lymphatic vessel remodeling remains incompletely elucidated. In the context of the role of PDGF in vessel development, studies have revealed that PDGF signals determine the branching of the vasculature directly from the coelomic vessels [55]. Considering the experiments that have been performed on culture cells, it has...
been demonstrated that endothelial migration is PDGFRα dependent signaling [56]. It seems that PDGF family members are not long-term chemotactic factors for ECs, but are mandatory for upstream regulation in order to activate or induce the secretion of different signaling proteins [57]. Recently, it was demonstrated that PDGFRβ induced signaling in the vasculature may determine different types of transcriptional responses. Different members of the PDGF family are linked to a series of transcriptional targets under different circumstances. Based on these remarks, the specific cellular responses that occur after PDGF action can be explained.

Normal versus pathological

Despite the fact that the same signaling molecule is implicated, the expression, function and the PDGF mechanism of action largely vary in normal compared to pathological conditions. This differential expression concerns the receptors associated with this family as well. Although lymphangiogenesis (normal and pathological) as well as the behavior of the PDGF are aspects about which very little is known or understood, a parallel can be made between the normal and the tumor tissue regarding the PDGF action.

There are two main theories regarding lymphatic vessel development: the first stating that the conversion of the venous EC into a lymphatic one by Prox-1 induction [58] and the second starting from lymphangioblasts.

The main factor involved in normal angiogenesis and lymphangiogenesis induction is the vascular endothelial growth factor (VEGF) [59, 60], PDGF being just a factor that participates in this process. The VEGF induces migration, proliferation and cell growth followed by the future development of blood or lymphatic vessels [61].

In normal conditions, PDGF acts in a predominant paracrine manner in order to encourage angiogenesis and lymphangiogenesis.

In the case of all tumors, the lymphatic vessel profile is changed. This fact has been demonstrated in numerous studies based on experimental models. Most of these studies target the functional implications of VEGF and VEGFR in pathological angiogenesis (mostly) and lymphangiogenesis [62], the implications of PDGF and PDGFR being recently discovered.

Within tumors, high levels of PDGF (especially homodimer B), as well as the overexpression of PDGFRβ, are detected mainly in the peripheral lymphatic vessels. The newly formed pathological lymphatic vessels, unlike the physiological

Figure 2. PDGFRα expression in primary breast cancer cells (A), peritumoral lymphatic vessels (B) and corresponding nodal metastasis (C)
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One possible answer could be the development of different stromal components from various normal tissues with a high variability in the cell types able to secrete PDGFs. Also the autocrine and paracrine mechanism of action could be specific for each normal and tumor microenvironment. But all these hypotheses need to be validated by future studies.

The PDGF members are expressed in various cell types in physiological conditions. It seems that the strongest expression for PDGF-A was identified in the heart, the skeletal muscle and in the pancreas, while the BB dimer presents the highest expression in the heart and placenta, and moderate levels in other organs.

Within the PDGF family, only form A and form B are capable of forming the one heterodimer that has been evidenced so far: PDGF-AB. Even PDGF-C presents high expression in the heart, the kidney, the adrenal gland and the pancreas, but lower levels have been evidenced in the liver and the ovary. The highest expression of D form appears in the pancreas, ovaries, and heart, while their expression in the brain, lungs, and skeletal muscles has not been documented [4]. Also, the expression of PDGF-C is absent in the spleen and the colon [14]. The differential expression has been identified not only in normal cells but also in tumor cell lines [69]. The literature reveals that different PDGF members are expressed within the same organ, but they may appear with a different structure. The structural variety is in this case associated with a functional one.

Is it possible for structural differences to appear within the same PDGF form? Supposing that this might be possible, the basis of this phenomenon may be represented by a mutation that appears in the gene encoding the synthesis of the PDGF form.

Not only the growth factor itself presents differential expression but also the growth factor associated receptor. Thus, PDGFRα is expressed by tumor cells in glioblastomas and PDGFRβ is expressed by the ECs in the newly formed blood vessels [69]. In the transplanted tumor models, the number and the density of perivascular cells are much higher, because of cancerous cells over-expressing PDGF-B. It seems that perivascular cells are capable of responding to ectopic sources of PDGF-B through the migration phenomenon from the ECs. Regarding this aspect, PDGF-B is not
only a mitogen factor for perivascular cell precursors but at the same time it guides the migration during tumor angiogenesis.

In lymphangiogenesis, intra- and peritumoural lymphatic vessels are key elements that facilitate tumor cell spreading in regional lymph nodes. This fact has been evidenced in numerous common types of cancer such as breast cancer, colorectal carcinoma and malignant melanoma. Amongst the PDGF family members, PDGF-B expression has been evidenced in breast cancer. It seems that dimer B of PDGFs is a chemotactic factor for primary LECs. The effect of PDGF in perivascular cells and in tumor stroma points out the possibility that PDGF acts along with other angiogenic molecules in order to sustain both angiogenesis and lymphangiogenesis.

Through stopping PDGFR activity, the decrease of cell proliferation may be determined, and the increase of apoptosis rate may be induced because the high levels of PDGF and PDGFR are often associated with a bad prognosis and a low response to chemotherapy. Due to the fact that tumor cells and tumor associated ECs express PDGFRs, the receptor stopping pathway is based on the use of a PDGFR-tyrosine kinase, in order to block the PDGFR phosphorylating domain.

**Upcoming challenges!**

Besides this lymphangiogenic growth factor with direct action, the other factors intervening in lymphangiogenesis are fibroblastic growth factor 2, angiopoietin 2, neuropilin 2 and Prox-1. Is it possible for PDGF action to resemble that of VEGF during embryogenesis in the context of lymphatic vessel formation? Is it possible to derive two types of polypeptides with similar action? Despite all this, it seems that ECs do not express the β receptor in physiological conditions. Is it possible, under some specific circumstances, for the VEGF to replace the action of the PDGF, considering the fact that the latter has a higher lymphangiogenic action compared to that of PDGF? Is it possible to neglect the PDGF action in embryonic lymphangiogenesis opposed to that of VEGF? It may be so.

In the end, almost all experimental studies on the functional implications of the PDGF are mostly based on pathological/tumor lymphangiogenesis, its pro-lymphangiogenic action, in normal conditions but mentioned along with that of VEGF. Despite all this, the action of the VEGF in pathological conditions should not be neglected. The capacity of tumor cells to secrete VEGF-C in order to induce lymphatic vessel formation is well known, the next step being represented by the dissemination towards the adjacent lymph nodes. Is it possible to delay PDGF interventions in normal lymphangiogenesis compared to that of VEGF? Phenotypic modification of lymphatic vessels through the intervention of microenvironmental factors is well known. Changing phenotype, in normal and pathological conditions, is in need of the interference of other isoforms of the PDGF family, apart from those known. If the PDGF family has members with antibody-like domains, is it possible for PDGF members to possess a similar capacity of isotype switching? Antibody isotype switching confers on them the capacity to interact with a large range of antigens. The isotype switching phenomenon is closely related to the gene that encodes the antibody chain and confers their great capacity to form about 10 types in order to interact with the antigens. Is it possible that because of the great phenotype variety those lymphatic vessels can present, there is a great variety of growth factors? (not only regarding the PDGF family).

Under the circumstances in which this phenomenon could be possible, in pathological conditions, it is very difficult to find characteristic inhibitors of great specificity for each type of growth factor generated.

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**Conflict of interest**

The authors declare no conflict of interest.

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