The role of new carbapenem combinations in the treatment of multidrug-resistant Gram-negative infections

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Multi-drug resistant (MDR) Gram-negative bacteria represent a growing threat, with an increasing prevalence of carbapenem-resistant Enterobacterales (CRE) infections, for which treatment options are limited. New treatment combinations composed of a β-lactam antibiotic plus a potent β-lactamase inhibitor (BLI) with anti-carbapenemase activity have been developed, including two carbapenem/BLI combinations that are commercially available—meropenem/vaborbactam (Vabomere® in the US, Vaborem® in Europe; Melinta Therapeutics) and imipenem/cilastatin/relebactam (Recarbrio®, Merck Sharp & Dohme), plus one other (meropenem/nacubactam) in early clinical development. This review provides a summary of the preclinical evidence supporting the use of carbapenem/BLI combinations and presents the clinical evidence across a range of MDR Gram-negative infections, with a focus on the use of meropenem/vaborbactam. All three BLIs have shown in vivo activity against Klebsiella pneumoniae carbapenemase and other class A carbapenemases. In 2019, meropenem/vaborbactam was listed in the WHO’s list of essential medicines, because of its activity against priority 1 antibiotic-resistant pathogens. Meropenem/vaborbactam has considerable in vitro and in vivo activity against CRE, and in vitro evidence showing a low potential for resistance at clinically relevant doses. In randomized trials, meropenem/vaborbactam was non-inferior to piperacillin/tazobactam in patients with complicated urinary tract infection and more effective than the best-available treatment in patients with serious CRE infections. Meropenem/vaborbactam is well tolerated and, based on clinical experience, demonstrated lower toxicity compared with the combination regimens that have previously been the standard of care. In conclusion, carbapenem/BLI combinations represent an important therapeutic strategy in patients with MDR Gram-negative infections.

Introduction

Major public health organizations around the world, including the WHO, US CDC, and the ECDC, recognize the growing threat of MDR Gram-negative bacteria. All of the priority 1 (critical) pathogens identified by the WHO in their 2017 report on antibiotic research priorities are Gram-negative MDR organisms (Acinetobacter baumannii, Pseudomonas aeruginosa, and Enterobacterales), and all but two of the nine priority 2 and 3 pathogens are Gram-negative MDR.

Carbapenems act by inducing the lysis of bacterial cells. These agents do not easily cross the outer membrane of Gram-negative organisms, and evidence that carbapenem monotherapy may no longer be an appropriate empirical choice for many patients with severe Gram-negative infections. Of particular concern is the increasing prevalence of carbapenem-resistant Enterobacterales (CRE) infections, for which treatment options are limited (see the related article by Bassetti and Garau in this Supplement). This has led to the development of new agents that combine a β-lactam antibiotic with a potent β-lactamase inhibitor (BLI), including three carbapenem/BLI combinations: meropenem/vaborbactam (Vabomere® in the US (Melinta); Vaborem® in Europe (Menarini)), imipenem/cilastatin/relebactam (Recarbrio®, Merck Sharp & Dohme) and meropenem/nacubactam (in early clinical development; Meiji Seika Pharma and Roche).

This article reviews the rationale for this type of combination in the treatment of patients with MDR Gram-negative infections, and describes the combinations currently available or in late-stage development, with a focus on the preclinical and clinical data for meropenem/vaborbactam.

Rationale for carbapenem/β-lactam combinations

Carbapenems act by inducing the lysis of bacterial cells. These agents do not easily cross the outer membrane of Gram-negative organisms, but rely on transport by porins. Once inside the periplasmic space, the carbapenems acylate the penicillin-binding proteins (PBPs) responsible for peptidoglycan...
formation in the cell wall. Inhibition of PBPs weakens the wall, leading to cell lysis and death.⁷

Many antibiotics have the same mechanism of action, but are broken down by β-lactamase enzymes in the periplasmic space before they can cause appreciable cell lysis. β-Lactamases fall into several categories based on their structure and hydrolytic activity (Table 1).⁸ Class A, C and D β-lactamases of the Ambler classification utilize active-site serine residues, whereas class B β-lactamases utilize zinc ions and are called metallo-β-lactamases (MBLs). The ability of the new β-lactamase inhibitor compounds to act on different enzymes is variable and is minimal against type B β-lactamases (MBLs). However, they are active against several clinically relevant class A and D enzymes [OXA-23, OXA-48, Klebsiella pneumoniae carbapenemase (KPC)].⁹ Currently, KPC-2 is the most widespread β-lactamase responsible for carbapenem resistance.⁹

The recently developed combination antibiotics include both a carbapenem and a BLI with inhibitory activity against a range of β-lactamase enzymes, including several carbapenemases.¹⁰⁻¹² However, these new agents have limited or no activity against MBLs, meaning that they will be less effective in regions where MBLs are prevalent.

**Carbapenem/β-lactamase inhibitor combinations**

Currently, there are two carbapenem/BLI combinations commercially available: meropenem/vaborbactam and imipenem/cilastatin/relebactam. Meropenem/vaborbactam was approved by the US FDA in 2017 for the treatment of complicated urinary tract infection (cUTI) in adults,¹³ and by the EMA in 2018 for the treatment of cUTI (including pyelonephritis), complicated intra-abdominal infection (cIAI) or hospital-acquired pneumonia [HAP; including ventilator-assisted pneumonia (VAP)] in adults, as well as bacteraemia that occurs in association (or suspected association) with any of these infections.¹⁴ In the EU, meropenem/vaborbactam is also indicated for the treatment of infections due to aerobic Gram-negative organisms in adults with limited treatment options.¹⁴ Imipenem/cilastatin/relebactam is also indicated for the treatment of infections due to aerobic Gram-negative organisms in adults with limited treatment options.¹⁴ Impinem/cilastatin/relebactam was approved by the FDA in adults with limited or no alternative treatment options for the treatment of cUTI or cIAI in 2019,¹⁵ and for treatment of HAP and VAP in 2020,¹⁶ and by the EMA for the treatment of infections due to aerobic Gram-negative organisms in adults with limited treatment options in 2020.¹⁷ A third combination, meropenem/nacubactam, is being developed by NacuGen Therapeutics and is in early clinical development.¹⁸

The BLIs in these three combination agents do not have a β-lactam structure. Relebactam and nacubactam are diazabicyclooctane molecules, both structurally related to avibactam (approved for use in combination with ceftazidime),¹²,¹⁹ whereas vaborbactam is a cyclic boronic acid molecule.¹⁰,¹² All three BLIs have a similar spectrum of activity against β-lactamase enzymes (Table 2), including KPC,¹²,¹⁹ and have shown in vivo activity against KPC and other class A carbapenemases.²¹⁻²⁴

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### Table 1. Nomenclature of clinically important enzymes⁸

| Molecular (Ambler) class | Functional group or subgroup | Common name⁹ | Clinically relevant enzyme(s) or enzyme family(ies) | Characteristic substrate profileb | Characteristic inhibitor profilec |
|--------------------------|-----------------------------|-------------|-----------------------------------------------|---------------------------------|---------------------------------|
| A 2a                     | Penicillinase               | PC1/blaZ    | Narrow-spectrum PENs                          | CLA, TZB                       |
| A 2b                     | Penicillinase               | TEM-1, SHV-1| Narrow-spectrum PENs, early CEPHs             | CLA, TZB                       |
| A 2be                    | ESBL                        | TEM-10, SHV-2, CTX-M-15 | Narrow-spectrum PENs, early CEPHs, ES-CEPHs, monobactams | CLA, TZB, AVI |
| A 2br                    | IRT                         | TEM-30 (IRT-2) | PENs, early CEPHs                             | TZB, AVI                       |
| A 2e                     | ESBL cephalosporinase       | CepA        | ES-CEPHs                                      | CLA but not ATM                |
| A 2f                     | Carbapenemase               | KPC         | All FDA-approved β-lactams                    | AVI, REL, VAB                  |
| A 2f                     | Carbapenemase               | SME         | PENs, early CEPHs, carbapenems, monobactams; not ES-CEPHS | CLA, AVI, VAB |
| B1, B3                   | MBL, carbapenemase          | IMP, NDM, VIM, SPM | All PENs, CEPHs, carbapenems; not monobactams | EDTA; no clinically approved inhibitor |
| B2                       | MBL, carbapenemase          | L1, CphA    | Carbapenems preferred                        | EDTA; no clinically approved inhibitor |
| C 1                      | Cephalosporinase            | AmpC        | CEPHs                                         | ATM, AVI, VAB                  |
| D 2d                     | Oxacillinase                | OXA-1       | PENs, especially oxacillin/ cloxacillin       | Variable                       |
| D 2df                    | Carbapenemase               | OXA-23, OXA-48, OXA-181, OXA-232 | PENs, especially oxacillin/ cloxacillin, carbapenems | AVI (OXA-48) |

⁹ESBL, extended-spectrum β-lactamase; IRT, inhibitor-resistant TEM; MBL, metallo-β-lactamase.

CEPH, cephalosporin; ES-CEPHs, expanded-spectrum cephalosporins; PEN, penicillin.

⁹ATM, aztreonam; AVI, avibactam; CLA, clavulanic acid; REL, relebactam; TZB, tazobactam; VAB, vaborbactam.
Class D enzymes

Citrobacter, Enterobacter cloacae spp. (including K. pneumoniae, Escherichia coli, Enterobacter AmpC, CTX-M and SHV enzyme variants. In a comparative in vitro analysis, meropenem/vaborbactam showed more potent reduces the risk of renal toxicity. Peptidase I inhibitor, which prolongs the half-life of imipenem and peptidase I when administered alone. Cilastatin is a renal dehydro- with cilastatin, because it is rapidly inactivated by renal dehydro-

Across all the isolates tested, the MIC90 of meropenem/vaborbactam was 1 mg/L, which was four times more potent than ceftazidime/avibactam and >64 times more potent than meropenem alone. In an in vitro hollow-fibre model, meropenem/vaborbactam at concentrations equivalent to those achieved by administration of the approved human dosage showed significant activity against a range of KPC-producing CRE strains, including K. pneumoniae, E. cloacae and E. coli isolates (Figure 1). Importantly, the isolates used in this study included strains with a range of KPC enzymes and with porin mutations that could confer meropenem resistance. Meropenem/vaborbactam was active against all strains except KP1092 and KP1254, which harboured loss-of-function mutations in the genes for OmpK36 porins. Data from that study and another indicate that strains with non-functional or poorly functional OmpK35 or OmpK36 porins are the least susceptible to meropenem/vaborbactam, as are those with an increase in blaKPC gene copy number. These are the same mechanisms that confer resistance to ceftazidime/avibactam, but meropenem/vaborbactam has shown potent activity against isolates resistant to ceftazidime/avibactam. In fact, the in vitro data suggest that, at concentrations simulating exposure after human dosing, meropenem/vaborbactam retains activity against CRE strains that harbour these resistance mechanisms, so there is a low risk of resistance development with clinical use of meropenem/vaborbactam. The increasing rates of reported ceftazidime/avibactam resistance may make meropenem/vaborbactam the preferred agent for KPC infections, especially considering the possible lower risk of selection for cross-resistance. However, meropenem/vaborbactam resistance was found in 8% (5/62) of the KPC-producing strains isolated from patients with bloodstream infections in an Italian series. Molecular characterization revealed that resistance was due to porin mutations and was associated with reduced susceptibility to both ceftazidime/avibactam and carbapenems. The in vivo activity of meropenem/vaborbactam was demonstrated in a range of murine models of infection, including thigh infections, lung infections or pyelonephritis caused by KPC-producing CRE, and a neutropenic thigh infection caused by P. aeruginosa and A. baumannii. In all of these models, meropenem/vaborbactam demonstrated significant bacterial killing, providing support for clinical investigation.

**Pharmacokinetic studies**

Studies in healthy volunteers demonstrated that meropenem and vaborbactam have similar pharmacokinetic properties. Both compounds are widely distributed throughout the body and are rapidly eliminated, predominantly through renal excretion.

The clinically tested doses of meropenem and vaborbactam were chosen based on pharmacokinetic and pharmacodynamic studies in animal and in vitro models of infection. The studies in in vitro models of infection allowed for detection of resistance, which is not usually possible in animal models of infection, and thus identified conditions and pharmacodynamic parameters associated with resistance prevention. The resulting dosage regimen that accomplished this objective was meropenem 2 g plus vaborbactam 2 g administered as a 3 h infusion every 8 h. This dose of meropenem/vaborbactam maximizes the percentage of the dosing interval during which free-drug levels exceed the MIC for the target organisms, based on the time-dependent killing profile of meropenem. This may minimize the likelihood of

### Table 2. Inhibitory activity of vaborbactam, relebactam and nacubactam against various β-lactamase enzymes

| β-Lactamase | Vaborbactam | Relebactam | Nacubactam |
|-------------|-------------|------------|------------|
| Class A enzymes | + | + | + |
| TEM | - | - | - |
| SHV | + | + | NA |
| CTX-M | + | + | + |
| KPC | + | + | + |
| Class B enzymes | - | - | - |
| MBL | NA | NA | - |
| IMP | - | - | - |
| Class C enzymes | + | + | + |
| AmpC | - | - | - |
| CMY-2 | NA | NA | + |
| Class D enzymes | + | + | + |
| OXA | - | - | - |

- No inhibitory activity; +, inhibitory activity; NA, not available.

Limited data available.

The carbapenems in the combinations are similarly effective in the management of serious infections, but meropenem shows greater activity than imipenem against Gram-negative bacilli, whereas imipenem is generally more active than meropenem against Gram-positive cocci. Imipenem is always administered with cilastatin, because it is rapidly inactivated by renal dehydro-peptidase I when administered alone. Cilastatin is a renal dehydro-peptidase I inhibitor, which prolongs the half-life of imipenem and reduces the risk of renal toxicity.

In 2019, meropenem/vaborbactam was added to WHO’s list of essential medicines because of its activity against priority 1 antibiotic-resistant pathogens. The rest of this article focuses on the research with this agent.

**Meropenem/vaborbactam**

**Preclinical studies**

In vitro data show that meropenem/vaborbactam is highly active against a range of KPC-positive CRE at concentrations of ≤4 mg/L (the susceptibility breakpoint defined by the US FDA) or ≤8 mg/L (the breakpoint defined by EUCAST). These include K. pneumoniae, Escherichia coli, Enterobacter spp. (including Enterobacter cloacae), Klebsiella oxytoca, Serratia marcescens and Citrobacter spp. The overall MIC required to inhibit 90% of isolates (MIC90) for meropenem/vaborbactam was ≤1 mg/L for all isolates, including isolates harbouring the KPC-2 and KPC-3, AmpC, CTX-M and SHV enzyme variants. In a comparative analysis, meropenem/vaborbactam showed more potent in vitro antimicrobial activity than meropenem alone, ceftazidime/avibactam, tigecycline, ceftazidime alone, minocycline, gentamicin or polymyxin B against clinical isolates of KPC-positive Enterobacteriaceae from a global collection. Across all the isolates tested, the MIC90 of meropenem/vaborbactam was 1 mg/L, which was four times more potent than ceftazidime/avibactam and >64 times more potent than meropenem alone.
resistance developing in strains that are not susceptible to meropenem alone.

The closely matched pharmacokinetic profiles of meropenem and vaborbactam in humans (including the shared route of excretion) result in comparable changes in pharmacokinetics in patients with renal impairment and similar recommended changes in dose in these patients.\(^{41}\) Dose adjustment is required in patients with an estimated glomerular filtration rate of <50 mL/min/1.73 m\(^2\) or creatinine clearance (\(C_L_{Cr}\)) ≤ 39 mL/min.\(^{13,44}\) The high rate of clearance with continuous renal replacement therapy necessitates prolonged infusion dosing, in addition to dosage adjustments. The usual dose of meropenem/vaborbactam is 2 g/2 g every 8 h, but patients with a \(C_L_{Cr}\) of 20–39 mL/min should receive 1 g/1 g every 8 h, those with a \(C_L_{Cr}\) of 10–19 mL/min should receive 1 g/1 g every 12 h, and those with a \(C_L_{Cr}\) of <10 mL/min should receive 0.5 g/0.5 g every 12 h.\(^{14}\) According to simulations, these doses will achieve the target pharmacokinetic/pharmacodynamic drug exposures in >90% of patients.\(^{44}\)

The combination shows good penetration of lower respiratory tract tissues after intravenous (IV) administration, with epithelial lining fluid concentrations of 65% for meropenem and 79% for vaborbactam (based on free drug).\(^{43}\) Given the efficacy of meropenem/vaborbactam in patients with serious CRE infections in the TANGO II trial, including those with HAP/VAP (described in the next section),\(^{45}\) the ability of meropenem/vaborbactam to enter the bronchial epithelial lining fluid is likely to translate into a clinical benefit for patients with HAP/VAP, and to reduce the risk of resistance development in this population.

Based on the well-established pharmacokinetic profile of meropenem and the pharmacokinetic profile of vaborbactam defined during Phase I development, the clinical development of meropenem/vaborbactam was able to proceed without the need for Phase II (dose-finding) studies.

**Clinical studies**

Because Phase II studies were not required for meropenem/vaborbactam, the efficacy and safety of the combination was able to be investigated in two sequential Phase III studies: TANGO I in patients with cUTI and acute pyelonephritis (AP)\(^{46}\) and TANGO II in patients with serious CRE infections.\(^{45}\)

TANGO I was the first study to investigate the efficacy of the proposed meropenem/vaborbactam dosage in a population of patients with complicated infections. This multicentre, randomized, double-blind, non-inferiority study compared meropenem/vaborbactam with piperacillin/tazobactam in 550 patients with cUTI or AP, with Enterobacterales as the most common causative pathogens.\(^{46}\)

Patients in both groups were treated with IV agents for 1–15 days (mean 8 days), but the overall mean antibiotic treatment duration was 10 days when oral step-down therapy was included. Two different primary endpoints were defined for the two key regulatory bodies. For the US FDA, the primary endpoint was overall success at the end of IV treatment, defined as a composite of clinical cure (complete resolution or significant improvement in symptoms) and microbial eradication (<10\(^4\) cfu in urine). For the EMA, the primary endpoint was microbial eradication (<10\(^3\) cfu in urine) at the test-of-cure visit (7 days after the end of treatment).\(^{46}\)

The non-inferiority of meropenem/vaborbactam was demonstrated for both primary endpoints. The rate of overall success (US FDA criterion) was 98.4% in the group receiving meropenem/vaborbactam and 94.0% in the group receiving piperacillin/tazobactam (difference 4.5%). Because the lower limit of the 95% CI was greater than the prespecified non-inferiority margin of −15%, meropenem/vaborbactam was shown to be significantly non-inferior to piperacillin/tazobactam (\(P<0.001\)). Additionally,
because the lower limit of the 95% CI was also greater than 0%, meropenem/vaborbactam was also shown to be superior to piperacillin/tazobactam (P = 0.01). Secondary endpoints (overall success at test of cure, clinical cure at the end of IV treatment and microbial eradication at test of cure) all showed a similar pattern, with comparable or slightly higher rates of each outcome in the meropenem/vaborbactam group compared with piperacillin/tazobactam. The incidence and type of adverse events were similar in the two groups. This pathogen-specific, randomized, controlled trial compared meropenem/vaborbactam as a single agent with best-available therapy (BAT), usually administered as a combination of multiple antibiotics, in 77 patients with CRE infections including bacteraemia (36%), cUTI or AP (45.3%), cIAI (9.3%) and HAP or VAP (9.3%). The most common pathogen in the intent-to-treat population was KPC-producing K. pneumoniae with a high-level of meropenem resistance (MIC50 of 64 mg/L in both groups).

Patients were randomized 2:1 to open-label treatment with meropenem/vaborbactam or BAT selected by the investigator; this could include any monotherapy or combination of polymyxins, carbapenems, aminoglycosides or tigecycline, or monotherapy with ceftazidime/avibactam. An interim analysis showed that the risk/benefit profile favoured meropenem/vaborbactam, so the independent Data Safety Monitoring Board recommended stopping randomization to the BAT group, leading to early discontinuation of study recruitment.

Among patients with a confirmed CRE infection, clinical cure rates were significantly higher in the group receiving meropenem/vaborbactam compared with BAT (65.6% versus 33.3% at the end of treatment (P = 0.03) and 59.4% versus 26.7% at test of cure (P = 0.02)). The 28 day mortality rate was numerically lower with meropenem/vaborbactam (15.6%) than with BAT (33.3%; P = 0.20) (Figure 2); only one of the five deaths in the meropenem/vaborbactam group was related to sepsis compared with four of the five deaths in the BAT group. The difference in all-cause mortality at Day 28 across all indications was driven by mortality differences in subjects with HAP/VAP or bacteraemia, the sickest subjects enrolled in the study.

Moreover, meropenem/vaborbactam was associated with fewer adverse events (Figure 2), including severe, serious, and drug-related adverse events, compared with BAT. The difference in renal events was notable, with 24.0% of patients in the BAT group developing renal-related adverse events compared with 4.0% in the meropenem/vaborbactam group; the incidence of acute renal failure was 12.0% with BAT versus 2.0% with meropenem/vaborbactam.

Although TANGO II was a randomized study, patients with prior antibiotic failure, who are expected to have a lower response to salvage therapy, were all enrolled in the meropenem/vaborbactam arm.
vaborbactam group. Therefore, a post hoc analysis was conducted in patients who had not received prior antibiotics (i.e. the population receiving meropenem/vaborbactam or BAT as first-line therapy for CRE). This analysis showed more marked differences in clinical cure and microbiological cure rates between the meropenem/vaborbactam and the BAT group than was seen in the overall population. These data confirm that, in critically ill patients, prompt initiation of the antibiotic therapy is linked to better outcomes. First-line use of meropenem/vaborbactam was associated with a clinical cure rate of 82.6% at end of treatment and 69.6% at test of cure compared with 33.3% and 26.7% in the first-line BAT group, representing an improvement of 49.3% and 42.9% at these time-points, respectively. Because the incidence of renal adverse events was higher in the BAT group, the risk/benefit profile clearly favoured meropenem/vaborbactam.

**Real-world clinical use**

Importantly, three real-world observational studies conducted in the USA have confirmed the effectiveness and safety of meropenem/vaborbactam during real-world clinical use. The first was a single-centre prospective observational study conducted in 20 consecutive patients who received meropenem/vaborbactam between December 2017 and April 2019. The other two were multicentre, retrospective studies: one in 40 patients with Gram-negative infections between October 2017 and June 2019, and the other in 131 patients with CRE infections treated with either meropenem/vaborbactam ($n = 26$) or ceftazidime/avibactam ($n = 105$) between February 2015 and October 2018.

In the single-centre study, patients had bacteraemia ($n = 8$), pneumonia ($n = 6$; VAP in $5/6$), tracheobronchitis ($n = 2$; ventilator-associated in $1/2$), skin and soft tissue infections ($n = 2$), pyelonephritis ($n = 1$) and peritonitis with intra-abdominal abscess ($n = 1$). This was a vulnerable group of patients: 14 (70%) were in the ICU at the onset of the infection, and seven (35%) required renal replacement therapy with intermittent or continuous haemodialysis. In addition, the median Acute Physiology and Chronic Health Evaluation (APACHE II) score was 20 and median Charlson comorbidity index (CCI) was 4. The most common pathogen was *K. pneumoniae*, and 19 of the 20 isolates (95%) were resistant to ertapenem.

Patients in the multicentre studies were similarly vulnerable. In the study of patients with Gram-negative infections, median APACHE II score was 17, median CCI was 6, 70% were in the ICU, and 90% had at least one risk factor for developing an MDR infection. In that study, the most common infection was pneumonia ($n = 13$), followed by UTI ($n = 8$), intra-abdominal infections ($n = 5$) and skin and soft tissue infections ($n = 5$); 11 patients had bacteraemia—primary in 2 and secondary in 9. Overall, 45 pathogens were isolated, most commonly *K. pneumoniae* (47%), *E. cloacae* (20%) and *E. coli* (13%). In the study of patients with CRE infections, 57% of patients were in ICU, 41% had bacteraemia and median APACHE II scores were 26 and 27 in the meropenem/vaborbactam and ceftazidime/avibactam groups, respectively. In that study, the most commonly identified CRE organisms were *Klebsiella* spp. (69%) and other Enterobacteriales (21%).

All three studies defined clinical success as the composite of 30 day survival, absence of recurrence at 30 days after starting treatment, and resolution of signs and symptoms of infection while receiving treatment. This endpoint was achieved by 20 patients (65%) in the single-centre study, by 34/40 patients (85%) in the study of patients with Gram-negative infections, and by 18/26 patients (69%) with meropenem/vaborbactam and 65/105 patients (62%) with ceftazidime/avibactam in the study of patients with CRE infections. In patients with Gram-negative infections, better clinical success was seen in patients with a community-acquired infection (86%) than a nosocomial infection (50%). Severe adverse events were reported in one patient in the single centre study (eosinophilia) and one in the study of patients with Gram-negative infections (Stevens-Johnson syndrome). In the study of patients with CRE infections, the incidence of nephrotoxicity was 14% with meropenem/vaborbactam and 29% with ceftazidime/avibactam.

The management of Gram-negative infections is often complicated by the presence of multiple morbidities and need of venous access devices, catheters or ventilators in fragile subjects. Real-world case reports indicate that meropenem/vaborbactam can be effective even in these vulnerable individuals. *Jorgensen et al.* described the successful use of meropenem/vaborbactam to treat bacteraemia caused by carbapenem-resistant *S. marcescens* and carbapenem-resistant *Enterobacter aerogenes* in an asplenic patient with HIV infection and renal failure requiring dialysis. In another case, Athans and colleagues used meropenem/vaborbactam to treat a subphrenic abscess and persistent bacteraemia caused by KPC-producing carbapenem-resistant *K. pneumoniae* in a liver transplant recipient who developed hepatic artery thrombosis and graft failure. The infection had not responded to courses of ceftazidime/avibactam or polymyxin B, initially administered as monotherapy, then with added gentamicin and tigecycline. This combination was discontinued because of renal toxicity. Once meropenem/vaborbactam was started, the patient’s renal function improved and their infection cleared, allowing the patient to undergo a successful second transplantation. The authors of both these reports highlighted the advantage of being able to use a single agent rather than combination therapy to treat CRE infections in clinically complex patients, thereby reducing the risk of toxicity.

**Conclusions**

Given the increasing threat posed by carbapenem-resistant pathogens and the limited treatment options for patients with CRE infections, the development of these new combinations of carbapenems and BLIs represents an important therapeutic advance. Meropenem/vaborbactam demonstrates considerable *in vitro* and *in vivo* activity against these pathogens, with a low potential for resistance at clinically relevant doses. The available clinical data show its efficacy and tolerability in patients with complicated CRE infections, with a low potential for toxicity compared with the combination regimens that have been the standard of care until now. In addition, observational studies and case reports have confirmed the efficacy of meropenem/vaborbactam in a real-world setting, including in very clinically complex patients with immunosuppression, renal dysfunction, or extensively drug-resistant organisms. Therefore, it is not surprising that this agent is now listed by the WHO as an essential medicine. With time, further data will emerge.
about how to optimally use meropenem/vaborbactam and other carbapenem-BLI combinations in the treatment of CRE.

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References
1 Centers for Disease Control and Prevention. Antibiotic resistance threats in
the United States, 2019. https://www.cdc.gov/drugresistance/pdf/threats-re
port/2019-ar-threats-report-508.pdf.
2 European Food Safety Authority, European Centre for Disease Prevention
and Control. The European Union Summary Report on Antimicrobial
Resistance in zoonotic and indicator bacteria from humans, animals and
food in 2017/2018. EFSA Journal 2020; 18: 6007.
3 World Health Organization. Global priority list of antibiotic-resistant bac-
teria to guide research, discovery, and development of new antibiotics.
https://www.who.int/medicines/publications/WHO-PPL-Short_Summary_
25Feb-ET_NM_WHO.pdf.
4 Hawkey PM, Warren RE, Livermore DM et al. Treatment of infections caused
by multidrug-resistant Gram-negative bacteria: report of the British Society
for Antimicrobial Chemotherapy/Healthcare Infection Society/British Infection
Association Joint Working Party. J Antimicrob Chemother 2018; 73: ii2–78.
5 Norgaard SM, Jensen CS, Aalestrup J et al. Choice of therapeutic interven-
tions and outcomes for the treatment of infections caused by multidrug-
resistant Gram-negative pathogens: a systematic review. Antimicrob Resist
Infect Control 2019; 8: 170.
6 Schmid A, Wolfensberger A, Nemeth J et al. Monotherapy versus combina-
tion therapy for multidrug-resistant Gram-negative infections: systematic
review and meta-analysis. Sci Rep 2019; 9: 15290.
7 Papp-Wallace KM, Endimiani A, Taracila MA et al. Carbapenems: past,
present, and future. Antimicrob Agents Chemother 2011; 55: 4943–60.
8 Bush K, Bradford PA. Epidemiology of β-lactamase-producing pathogens.
Clin Microbial Rev 2020; 33: e00047–19.
9 Munoz-Price LS, Pairoli L, Bonomo RA et al. Clinical epidemiology of the glo-
bal expansion of Klebsiella pneumoniae carbapenemases. Lancet Infect Dis
2013; 13: 785–96.
10 Lomovskaya O, Sun D, Rubio-Aparicio D et al. Vaborbactam: spectrum of
β-lactamase inhibition and impact of resistance mechanisms on activity in
Enterobacteriaceae. Antimicrob Agents Chemother 2017; 61: e01443-17.
11 Barnes MD, Taracila MA, Good CE et al. Nacubactam enhances merope-
nom activity against carbapenem-resistant Klebsiella pneumoniae producing
KPC. Antimicrob Agents Chemother 2019; 63: e00432-19.
12 Zhanel GG, Lawrence CK, Adam H et al. Imipenem-relebactam and merope-
nem-vaborbactam: two novel carbapenem-β-lactamase inhibitor combina-
Drugs 2018; 78: 65–98.
13 US Food and Drug Administration. Vaborber (meropenem and vabor-
bactam) for injection, for intravenous use [prescribing information]. https://
www.accessdata.fda.gov/drugsatfda_docs/label/2018/209776s001lbl.pdf.
14 European Medicines Agency. Vaborber 1 g/1 g powder for concentrate for
solution for infusion [summary of product characteristics]. https://www.
ema.europa.eu/en/documents/product-information/vaborber-epar-product-
information_en.pdf.
15 US Food and Drug Administration. Recarbrio (imipenem, cilastatin, and
relebactam) for injection, for intravenous use [prescribing information]. https://
www.accessdata.fda.gov/drugsatfda_docs/label/2019/212819s
000lbl.pdf.
16 US Food and Drug Administration. FDA approves antibiotic to treat
hospital-acquired bacterial pneumonia and ventilator-associated bacterial
pneumonia. https://www.fda.gov/news-events/press-announcements/fda-
approves-antibiotic-treat-hospital-acquired-bacterial-pneumonia-and-venti-
lator-associated.
17 European Medicines Agency. Recarbrio 500 mg/500 mg/250 mg powder
for solution for infusion [summary of product characteristics]. https://www.
ema.europa.eu/en/documents/product-information/recarbrio-epar-product-
information_en.pdf.
18 World Health Organization. 2019 Antibacterial agents in clinical develop-
ment: an analysis of the antibacterial clinical development pipeline. https://
apps.who.int/iris/bitstream/handle/10665/330420/9789240000193-eng.pdf.
19 Mushtaq S, Vickers A, Woodford N et al. Activity of nacubactam (RG6080/
OP0595) combinations against MBL-producing Enterobacteriaceae. J Antimicrob
Chemother 2019; 74: 953–60.
20 Tehrani K, Martin NL. β-lactam/β-lactamase inhibitor combinations: an
update. Medchecomm 2018; 9: 1439–56.
21 Monogue ML, Giovagnoli S, Bissantz C et al. In vivo efficacy of meropenem
with a novel non-β-lactam-β-lactamase inhibitor, nacubactam, against
gram-negative organisms exhibiting various resistance mechanisms in a
murine complicated urinary tract infection model. Antimicrob Agents
Chemother 2018; 62: e02596-17.
22 Griffith DC, Sabet M, Tarazi Z et al. Pharmacokinetics/pharmacodynamics
of vaborbactam, a novel β-lactamase inhibitor, in combination with merope-
nem. Antimicrob Agents Chemother 2019; 63: e01659-18.
23 Reyes S, Abdelraouf K, Nicolau DP. In vivo activity of human-simulated
regimens of imipenem alone and in combination with relebactam against
Pseudomonas aeruginosa in the murine thigh infection model. J Antimicrob
Chemother 2020; 75: 2197–205.
24 Asempa TE, Motos A, Abdelraouf K et al. Efficacy of human-simulated
epithelial lining fluid exposure of meropenem-nacubactam combination
against class A serine β-lactamase-producing Enterobacteriaceae in the neu-
traphilic murine lung infection model. Antimicrob Agents Chemother 2019;
63: e02382-18.
25 Zhanel GG, Simor AE, Vercaigne L et al. Imipenem and meropenem: com-
parison of in vitro activity, pharmacokinetics, clinical trials and adverse
effects. Can J Infect Dis 1996; 9: 215–28.
26 Tune BM, Fravert D, Hsu CY. Thienamycin nephrotoxicity. Mitochondrial in-
jury and oxidative effects of imipenem in the rabbit kidney. Biochem
Pharmacol 1989; 38: 3779–83.
27 World Health Organization. Model list of essential medicines, 21st list.
https://apps.who.int/iris/bitstream/handle/10665/325771/WHO-MVP-EMP-
IAU-2019-06-Eng.pdf?ua=1.
28 Hackel MA, Lomovskaya O, Dudley MN et al. In vitro activity of meropenem-
vaborbactam against clinical isolates of KPC-positive Enterobacteriaceae. Antimicrob Agents Chemother 2018; 62: e01904–17.
29 Castanheira M, Rhomberg PR, Flamm RK et al. Effect of the β-lactamase
inhibitor vaborbactam combined with meropenem against serine
 carbapenemase-producing Enterobacteriaceae. Antimicrob Agents
Chemother 2016; 60: 5454–8.
30 Castanheira M, Doyle TB, Kanto V et al. Meropenem-vaborbactam activ-
ity against carbapenem-resistant enterobacteriales isolates collected in U.S.
hospitals during 2016 to 2018. Antimicrob Agents Chemother 2020; 64: e01951-19.
31 Sabet M, Tarazi Z, Rubio-Aparicio D et al. Activity of simulated human
dosage regimens of meropenem and vaborbactam against
carbapenem-resistant Enterobacteriaceae in an in vitro hollow-fiber model. *Antimicrob Agents Chemother* 2018; 62: e01969-17.

32 Sabet M, Tarazi Z, Nolan T et al. Activity of meropenem-vaborbactam in mouse models of infection due to KPC-producing carbapenem-resistant *Enterobacteriaceae*. *Antimicrob Agents Chemother* 2017; 62: e01446-17.

33 Sun D, Rubio-Aparicio D, Nelson K et al. Meropenem-vaborbactam resistance selection, resistance prevention, and molecular mechanisms in mutants of KPC-producing *Klebsiella pneumoniae*. *Antimicrob Agents Chemother* 2017; 61: e01694-17.

34 Wilson WR, Klime EG, Jones CE et al. Effects of KPC variant and porin genotype on the in vitro activity of meropenem-vaborbactam against carbapenem-resistant *Enterobacteriaceae*. *Antimicrob Agents Chemother* 2019; 63: e02048-18.

35 Tsivkovski R, Lomovskaya O. Potency of vaborbactam is less affected than that of avibactam in strains producing KPC-2 mutations that confer resistance to ceftazidime-avibactam. *Antimicrob Agents Chemother* 2020; 64: e01936-19.

36 Pogue JM, Bonomo RA, Kaye KS. Ceftazidime/avibactam, meropenem/vaborbactam, or both? Clinical and formulary considerations. *Clin Infect Dis* 2019; 68: S19–24.

37 Galiani P, Lombardo D, Bussini L et al. Epidemiology of meropenem/vaborbactam resistance in kpc-producing *Klebsiella pneumoniae* causing bloodstream infections in northern Italy. 2018. *Antibiotics (Basel)* 2021; 10: 536.

38 Weiss WJ, Pulse ME, Nguyen P et al. Activity of meropenem-vaborbactam against carbapenem-resistant *Enterobacteriaceae* in a murine model of pyelonephritis. *Antimicrob Agents Chemother* 2018; 62: e01439-17.

39 Sabet M, Tarazi Z, Griffith DC. Activity of meropenem-vaborbactam against *Pseudomonas aeruginosa* and *Acinetobacter baumannii* in a neutropenic mouse thigh infection model. *Antimicrob Agents Chemother* 2019; 63: e01665-18.

40 Griffith DC, Loutit JS, Morgan EE et al. Phase 1 study of the safety, tolerability, and pharmacokinetics of the β-lactamase inhibitor vaborbactam (RPX7009) in healthy adult subjects. *Antimicrob Agents Chemther* 2016; 60: 6326–32.

41 Rubino CM, Bhavnani SM, Loutit JS et al. Single-dose pharmacokinetics and safety of meropenem-vaborbactam in subjects with chronic renal impairment. *Antimicrob Agents Chemther* 2018; 62: e02103-17.

42 Rubino CM, Bhavnani SM, Loutit JS et al. Phase 1 study of the safety, tolerability, and pharmacokinetics of vaborbactam and meropenem alone and in combination following single and multiple doses in healthy adult subjects. *Antimicrob Agents Chemther* 2018; 62: e02228-17.

43 Wenzler E, Gottfried MH, Loutit JS et al. Meropenem-RPX7009 concentrations in plasma, epithelial lining fluid, and alveolar macrophages of healthy adult subjects. *Antimicrob Agents Chemther* 2015; 59: 7232–9.

44 Bhavnani SM, Trang M, Griffith DC et al. Vaborbactam pharmacokinetic-pharmacodynamic (PK-PD) target attainment analyses as support for dose selection in patients with normal renal function and varying degrees of renal impairment [abstract 1851]. *Open Forum Infect Dis* 2017; 4: S530–1.

45 Wundrnick RG, Giamarellos-Bourboulis EJ, Rahav G et al. Effect and safety of meropenem-vaborbactam versus best-available therapy in patients with carbapenem-resistant *Enterobacteriaceae* infections: the TANGO II randomized clinical trial. *Infect Dis Ther* 2018; 7: 439–55.

46 Kaye KS, Bhowmick T, Metallidis S et al. Effect of meropenem-vaborbactam vs piperacillin-tazobactam on clinical cure or improvement and microbial eradication in complicated urinary tract infection: the TANGO I randomized clinical trial. *JAMA* 2018; 319: 788–99.

47 Jorgensen SCJ, McDonald P, Mynatt RP et al. Averting the post-antibiotic era: successful use of meropenem/vaborbactam for carbapenem-resistant *Serratia marcescens* and *Enterobacter aerogenes* bacteraemia in a haemodialysis patient. *J Antimicrob Chemother* 2018; 73: 3529–31.

48 Bassetti M, Giacobbe DR, Patel N et al. Efficacy and safety of meropenem-vaborbactam versus best available therapy for the treatment of carbapenem-resistant *Enterobacteriaceae* infections in patients without prior antimicrobial failure: a post hoc analysis. *Adv Ther* 2019; 36: 1771–7.

49 Shields RK, McCreary EK, Marini RV et al. Early experience with meropenem-vaborbactam for treatment of carbapenem-resistant *Enterobacteriaceae* infections. *Clin Infect Dis* 2019; 71: 667–71.

50 Alosaimy S, Jorgensen SCJ, Lagnf AM et al. Real-world multicenter analysis of clinical outcomes and safety of meropenem-vaborbactam in patients treated for serious Gram-negative bacterial infections. *Open Forum Infect Dis* 2020; 7: ofaa051.

51 Ackley R, Rushdy D, Meredith J et al. Meropenem-vaborbactam versus ceftazidime-avibactam for treatment of carbapenem-resistant *Enterobacteriaceae* infections. *Antimicrob Agents Chemther* 2020; 64: e02313-19.

52 Athans V, Neuner EA, Hassouna H et al. Meropenem-vaborbactam as salvage therapy for ceftazidime-avibactam-resistant *Klebsiella pneumoniae* bacteremia and abscess in a liver transplant recipient. *Antimicrob Agents Chemther* 2019; 63: e01551-18.