Enzyme-Free Glucose Biosensors Based on MoS₂ Nanocomposites

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Abstract
High-performance glucose biosensors are highly desired for healthcare. To meet these demands, glucose biosensors, particularly enzyme-free glucose biosensors, have received much attention. Two-dimensional materials, e.g., graphene, with high surface area, excellent electrical properties, and good biocompatibility, have been the main focus of biosensor research in the last decade. This review presents the recent progress made in enzyme-free glucose biosensors based on MoS₂ nanocomposites. Two different techniques for glucose detections are introduced, with an emphasis on electrochemical glucose biosensors. Challenges and future perspectives of MoS₂ nanocomposite glucose biosensors are also discussed.

Keywords: Glucose biosensors, Two-dimensional materials, Enzyme-free, Electrochemical

Introduction
Glucose concentration in the human blood is an important health indicator. For example, healthy people typically have a blood glucose level around 3.9–6.1 mM (1 mM ≈ 18 mg/dL), and glucose concentrations outside of this range may indicate kidney dysfunction, diabetes, etc. [1]. Driven by the ever-increasing demand for healthcare, many efforts have been devoted to enzymatic glucose biosensors based on glucose oxidase (GOx) since the report of enzyme electrode by Updike and Hicks in 1967 [2]. Despite the simplicity, efficiency, high sensitivity, and selectivity of enzymatic glucose biosensors, two major challenges, i.e., high cost and instability, still cannot be satisfactorily addressed. This is because enzymatic glucose sensors use enzymes, such as GOx, to detect glucose indirectly, which often involves expensive catalysts and complicated device construction, and thus, relatively high cost [3, 4]. In addition, enzymes like GOx are vulnerable to temperature, humidity, pH, and nonphysiological chemicals due to their intrinsic thermal and chemical instability [5, 6]. The immobilization of GOx on an electrode surface often adds another layer of difficulty in fabricating enzymatic glucose sensors with good stability and reproducibility [7, 8].

Thanks to the development of the Internet of Things, sensors of low cost and high reliability have attracted increasingly more attention. In the pursuit of glucose sensors that meet these demands, enzyme-free electrochemical biosensors have gained popularity due to a number of advantages, including simplicity, high sensitivity, and stability [9–12]. Enzyme-free electrochemical biosensors directly detect glucose via electrocatalytic oxidation, which avoids the use of costly enzyme as well as improves stability in ambient conditions. Such biosensors are expected to open new opportunities for incorporation with portable devices and real-time glucose detection [13]. The key to realize practical enzyme-free glucose sensors is inexpensive, reliable, biocompatible, and abundant catalysts. To this goal, nanocomposites, e.g., composites of two-dimensional materials and nanoparticles, have been widely adopted as biosensor electrode materials [14]. In particular, bioelectronics based on two-dimensional (2D) materials becomes an exciting new interdisciplinary field, owing to many unique physical and chemical properties of 2D materials, including large specific surface area, excellent conductivity, and facile synthesis. For instance, the large specific surface area of 2D materials enables easy surface functionalization via hybridization. The high conductivity allows efficient charge transfer and collection in 2D materials. Among various 2D materials, graphene and its derivatives

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are undoubtedly the most studied material in biosensors [15]. Another type of 2D materials that has been widely explored in electronics and optoelectronics also show merits of being bioelectronics materials. Transition metal dichalcogenides (TMDs), especially molybdenum disulfide (MoS₂), possess similar advantages of large specific surface area, chemical inertness, and surface functionality. Intercalation of foreign ions or molecules into TMD nanosheets can be readily achieved given their unique atomic structure [16]. An important property that makes MoS₂ stand well out from other 2D materials is its high catalytic activity arises from exposed edges [13]. However, the disadvantage of MoS₂ is also quite obvious. Compared with graphene, 2D MoS₂ sheets have much lower electrical conductivity. Re-stacking of MoS₂ nanosheets further limits the charge transfer as well as active reaction sites.

Marginal efforts have been made in enzyme-free glucose sensors based on 2D MoS₂ until the very recent years. The poor performance of MoS₂-based electrochemical devices has been well coped with by using a number of methods that successfully solved the low intrinsic conductivity. A few MoS₂-based electrochemical glucose biosensors have been reported with performance exceeding its graphene counterparts [17]. Apart from electrochemical biosensors, low-cost non-electrochemical methods have also been recently studied for 2D MoS₂ by taking advantage of the progress of MoS₂ made in electronics and optoelectronics [18]. In this review, we summarize the recent advances in MoS₂-based glucose biosensors. Particular attention is given to MoS₂-based electrochemical glucose biosensors, which are described in the “Electrochemical Glucose Biosensor-Based MoS₂ Nanocomposites” section. In the “High Sensitivity Glucose Detection Using MoS₂ Field-Effect Transistors” section, MoS₂ field-effect transistors for glucose detection of glucose are briefly introduced. Finally, the conclusion and future perspectives of MoS₂ nanocomposite glucose biosensors are presented.

**Electrochemical Glucose Biosensor-Based MoS₂ Nanocomposites**

For a few decades, metals or alloys have been the main catalyst options for the direct electrocatalytic oxidation of glucose [19]. In the last decade, two-dimensional materials with large surface area as well as unique chemical and physical properties open new opportunities for many fields including electrochemical sensing, energy storage, and electronics [20]. In terms of electrochemical biosensing, nanocomposites of different 2D materials and catalysts show clear advantages over traditional catalysts. The synergistic coupling between these materials, namely synergistic effects, can lead to distinct enhancement in catalytic activity [21]. A good number of such nanocomposites, particularly those based on graphene or graphene derivatives, have been developed and applied to enzyme-free glucose sensors. Layered MoS₂ is expected to possess similar advantages as it shares the material properties of graphene. Particularly, layer MoS₂ nanosheets possess a large number of edges, which, similar to functionalized graphene sheets, act as active sites for catalytic reactions [22, 23].

Indeed, Huang et al. synthesized MoS₂ nanostructures by a hydrothermal method [24]. A glassy carbon electrode modified with the MoS₂ nanostructures and chitosan/Au nanoparticle composites showed distinct overpotential reduction for bisphenol A oxidation. The nanocomposite sensor showed an efficient electrocatalytic oxidation of bisphenol A as evidenced by the significantly increased current in the cyclic voltammograms. A good linear detection range from 0.05 to 100 μM is obtained for bisphenol A sensing. Also, a very detection limit of 5 nM is estimated. This work clearly demonstrated the excellent electrocatalytic activity and synergistic effects of Au/MoS₂ nanocomposites. Similarly, MoS₂-based nanocomposites have been used for enzyme-free glucose detection. MoS₂ flowers with a large surface area were synthesized by a hydrothermal method using cetyltrimethylammonium bromide (CTAB) as a surfactant [25]. The morphology of the microflowers can be controlled by the pH of the reaction solution, concentration of CTAB surfactant, and annealing temperature. The MoS₂ microflowers obtained at an annealing temperature of 500 °C showed good crystalline quality and hence improved charge transfer. Interestingly, electrochemical enzyme-free glucose sensing tests showed that the MoS₂ microflower electrode without any functionalization can offer a high sensitivity of 570.71 μA mM⁻¹ cm⁻². Additionally, the sensor shows a wide linear detection range of up to 30 mM.

The synergistic effects of MoS₂ catalysts doped or hybridized by foreign metals, such as Cu, Ni, Co, and Fe, also hold true for enhanced electrochemical catalysis of glucose. Huang et al. managed to combine the advantages of both the strong electrocatalytic activity of copper for glucose oxidation and the large surface area and active edge sites of MoS₂ nanosheets [26]. The Cu nanoparticles decorated MoS₂ nanosheets showed electrocatalytic activity towards glucose oxidation. A high sensitivity of 1055 μA mM⁻² cm⁻² and a linear detection range of up to 4 mM have been reported for the Cu/MoS₂ nanocomposite glucose sensor. The sensitivity nearly doubled the value measured from the MoS₂ microflower electrode. The sensor also demonstrated good selectivity in detecting glucose against uric acid, ascorbic acid, and dopamine. The interference current caused by these chemicals is only about 2.1–5.2% of that from glucose, and such a low interference current level can be considered negligible at physiological concentration.
Another attractive candidate is nickel (Ni), which has been extensively studied for Ni/graphene hybrids. Similar to Cu, Ni is also an earth-abundant metal. The redox couple of Ni$^{3+}$/Ni$^{2+}$ offers an impressive catalytic activity in alkaline media. Therefore, Huang et al. used MoS$_2$ nanosheet as catalyst support to immobilize Ni nanoparticles [27]. MoS$_2$ nanosheet was synthesized from MoS$_2$ powder in ethanol/water mixed-solvent via liquid exfoliation. Ni nanoparticles were reduced on MoS$_2$ nanosheet by heating a MoS$_2$ nanosheet–ethylene glycol solution at 60 °C for 1 h after an addition of NiCl$_2$·6H$_2$O precursor and N$_2$H$_4$·H$_2$O and NaOH solutions. A glucose sensor electrode was prepared by depositing Ni-MoS$_2$ hybrid on a glassy carbon electrode. Cyclic voltammogram of the Ni/MoS$_2$ hybrid-modified electrode clearly revealed glucose oxidation with a higher current than a Ni-modified reference electrode. The improved electrocatalytic activity was attributed to more active sites on MoS$_2$ nanosheets as well as reduced Ni nanoparticle aggregation on a 2D material support. The amperometric results confirmed a good linear detection range up to 4 mM, a high sensitivity of 1824 μA mM$^{-1}$ cm$^{-2}$, and a low detection limit of 0.31 μM at a signal/noise ratio of 3 (S/N = 3). Compared with the Cu/MoS$_2$ nanocomposite glucose sensor, there is further improvement in sensitivity by using Ni/MoS$_2$ nanocomposites. The impact of the interfering species, including dopamine, ascorbic acid, and uric acid, on glucose sensing was also found to be marginal. More importantly, the sensor showed good reproducibility and high stability. A negligible 3.4% reduction in the response of the sensor was measured after ambient storage for 4 weeks. In addition, Anderson et al. reported a highly sensitive non-enzymatic glucose biosensor by incorporating colloidal silver nanoparticles with MoS$_2$ [28]. The introduction of Ag nanoparticles was to address the intrinsic poor conductivity of MoS$_2$. An excellent sensitivity of 9044.6 μA mM$^{-1}$ cm$^{-2}$ and a low detection limit of 0.03 μM were reported. However, the linear detection range is only up to 1 mM.

The electrocatalytic activities of MoS$_2$ can be further improved by hybridizing with graphene. The low intrinsic conductivity of MoS$_2$ undermines its high catalytic activity. The charge transfers among MoS$_2$ nanomaterials are slow in electrochemical reactions or general electronic applications. On the other hand, graphene has superior electrical conductivity and can serve as an immediate solution to slow electron transport in MoS$_2$ nanomaterials [29]. Jeong et al. fabricated three-dimensional (3D) MoS$_2$/graphene aerogel nanocomposites by a one-pot hydrothermal method [13]. Although enhanced electrocatalytic activities have been observed by using the 3D porous structure compared with the 2D reference sample, the use of glucose oxidase complicated the fabrication and faced the same issues of enzymatic sensors. Geng et al. synthesized Ni-doped MoS$_2$ nanoparticles decorated on reduced graphene oxide (Ni-MoS$_2$/rGO) by a facile and scalable method [30]. As shown in Fig. 1a, graphene oxide synthesized by the Hummers and Offeman method was mixed with CH$_3$COOH and deionized water. Ni-Mo precursor solution was prepared by adding (NH$_4$)$_2$Mo$_3$S$_4$ and Ni(CH$_3$COO)$_2$4H$_2$O with different molar ratios into the graphene oxide solution. The Ni-MoS$_2$/rGO suspension was obtained after centrifugation and drying at 80 °C. The collected Ni-MoS$_2$/rGO suspension was then calcined for 4 h at 600 °C in N$_2$ atmosphere. The obtained Ni-MoS$_2$/rGO nanocomposites were used for non-enzymatic glucose sensing. Figure 1b shows the amperometric response of a sensor electrode modified by Ni-MoS$_2$/rGO nanocomposites to the successive addition of glucose solution. A clear increase in current was observed after each addition of glucose. Additionally, the inset in Fig. 1b shows the sensor was capable of responding to a glucose concentration as low as 5 μM. The exacted current signal as a function of glucose concentration is plotted in Fig. 1c, which clearly shows a broad linear detection range of the sensor, 0.005–8.2 mM, well covering the typical human blood glucose level. The calculated sensitivity and detection limit is 256.6 μA mM$^{-1}$ cm$^{-2}$ and 2.7 μM (S/N = 3), respectively. Although the sensitivity and detection limit of the sensor is distinctly lower than the earlier ones, the linear detection range is well improved. The work further reported improved electron transport rates and electrocatalytic activity with a diffusion coefficient of 1.83 × 10$^{-3}$ cm$^2$s$^{-1}$ and catalytic rate constants of 6.26 × 10$^5$ cm$^3$mol$^{-1}$s$^{-1}$ by using the Ni-MoS$_2$/rGO composites. As shown in Fig. 1d, when the sensor was stored under dry conditions, its current response to 1 mM glucose remains nearly unchanged for 15 days, showing good stability. The influence of common interference chemicals, NaCl, dopamine, uric acid, ascorbic acid, and V$_{pp}$, was also examined. The results are shown in Fig. 1e, and the impact of these chemicals on signal current is marginal. The current change caused by 0.1 mM of NaCl, dopamine, uric acid, ascorbic acid, and V$_{pp}$, a typical concentration of the interference chemicals in normal serum, is only 0.76%, 2.77%, 6.03%, 0%, 2.51%, and 0.63% of the current response to 2.5 mM glucose. Finally, the work demonstrated a good match between the measured concentration by the Ni-MoS$_2$/rGO sensor and the reported hospital values, showing a great potential of the sensors for practical applications [30].

An alternative way to improve the charge transfer in MoS$_2$ nanocomposites is to hybridize with another highly conductive and bio-compatible carbon material, carbon nanotubes (CNTs). Meanwhile, this way can well limit the re-stacking of MoS$_2$ nanomaterials, thereby providing more active reaction sites. CNTs have also
been widely synthesized in 3D structures and applied in energy storage, energy harvesting, sensing, etc. [31–33]. Li et al. prepared a 3D nanocomposites of MoS2 nanosheets hybridized with cobalt oxide nanoparticles and CNTs [34]. The cobalt oxide nanoparticles were used to enhance electrocatalytic activities and the CNTs to improve conductivity. One-pot hydrothermal method used to synthesize the Co-MoS2/CNT nanocomposites is briefly shown in Fig. 2a. A mixture of CNTs, 0.1 mmol Co(CH3COO)2 4H2O, 1.35 mmol Na2MoO4, and 7.5 mmol L-cysteine was transferred into a Teflon-lined stainless autoclave and kept at 180 °C for 24 h. The product was then cooled, centrifuged, and rinsed with deionized water and absolute ethanol. The cleaned Co-MoS2/CNT nanocomposites were finally dried in a vacuum oven at 60 °C for 24 h. The scanning electron microscopy (SEM) and transmission electron microscopy (TEM) images of the Co-MoS2/CNT nanocomposites are shown in Fig. 2b, c. Typical 3D bundled CNTs with diameters around 20 nm was observed. The TEM image clearly shows the hollow CNTs attached to MoS2 nanosheets. Such a structure serves as a highly conductive matrix to support MoS2 nanosheets and immobilize Co nanoparticles. Such the densely packed Co-MoS2/CNT nanocomposites not only provide a good amount of catalytic active edges but also allow efficient charge transfer during reactions. More importantly, the densely packed Co-MoS2/CNT structure and the fairly large interlayer spacing of 0.65 nm for MoS2 (larger than that of 0.34 nm for CNTs) effectively suppress re-stacking of MoS2/CNT composite. Typical cyclic voltammetry and amperometric measurements were carried out to examine the performance of the enzyme-free sensor made of Co-MoS2/CNT nanocomposites. The amperometric responses of Co-MoS2/CNT sensing electrode measured (at 0.65 V vs. Ag/AgCl) with successive addition of glucose are shown in Fig. 2d. A distinct step increase in response current to glucose addition was observed. A good linear detection range was achieved up to 5.2 mM, as shown in Figs. 2e. The calculated sensitivity is 131.69 μA mM⁻¹ cm⁻². Despite the relatively low sensitivity, an extremely low detection limit of 80 nM obtained (S/N = 3) from Fig. 2f.

Similar to the synergistic effects displayed by metal-2D material hybrids, bimetallic alloys and nanostructures have also showed improved catalytic performance and showed good potential for many applications, including sensing [35], energy harvesting [36, 37], etc. Li et al. recently synthesized Au-Pd bimetallic nanoparticles for non-enzymatic hydrogen peroxide and glucose sensing [5]. The fabrication of the Au-Pd/MoS2 sensor electrode is illustrated in Fig. 3a. The MoS2 nanosheets were prepared by liquid exfoliation. Au-Pd bimetallic nanoparticles were synthesized by chemical reduction. The prepared Au-Pd/MoS2 nanocomposites were then deposited on a glassy carbon electrode for chemical sensing. As shown in Fig. 3b, good current steps were observed with a successive addition of glucose. The
linear detection range measured as 0.5–20 mM is well beyond normal human blood glucose level (Fig. 3c). Instead of using conventional bimetallic nanoparticles which often made from expensive metals, Ma et al. designed a gold nanoparticle-polypyrrole (PPY) co-decorated MoS₂ nanocomposite [38]. The metal/conductive polymer hybrids are also expected to improve the surface area and conductivity of a sensor electrode. Moreover, the use of conductive polymers can further reduce the cost of electrochemical sensors. The fabricated MoS₂-PPY-Au/glassy carbon electrode showed an incredible low detection of 0.08 nM, nearly interference-free selectivity, and long stability over 3 weeks. However, the sensor sensitivity is only 37.35 μA·μM⁻¹·cm⁻² and the detection range is rather limited (0.1–80 nM).

In addition to metals, metal oxides of high catalytic activities have also been attempted for improved electrochemical catalysis. Apart from its high electrocatalytic activities, the low cost of metal oxides is another advantage that cannot be overstressed for low-cost electrochemical sensors. Among various metal oxides, Cu₂O nanomaterials with different morphologies are promising for catalysis in various applications. Fang et al. have been studied MoS₂ decorated with Cu₂O nanoparticles for non-enzyme glucose sensing [39]. The amperometric measurements of the Cu₂O/MoS₂ hybrid-modified electrode show a good linear range from 0.01 to 4 mM. The extracted detection limit is about 1 μM. The sensitivity was calculated as high as 3108.87 μA·μM⁻¹·cm⁻², which is higher than most MoS₂-based non-enzyme glucose sensors. The results also indicate a good potential of metal oxides for low-cost non-enzyme glucose sensors. The comparison of the enzyme-free glucose biosensors based on MoS₂ nanocomposites is presented in Table 1.

**High-Sensitivity Glucose Detection Using MoS₂ Field-Effect Transistors**

MoS₂ field-effect transistors (FETs) possess a number of advantages, such as high switching current ratio, low leakage current, small subthreshold swing, and high mobility [41, 42]. Thanks to its excellent electronic properties and mechanical robustness, MoS₂ transistors show good promise for low energy, low cost, and wearable electronics [43, 44]. Biosensors, photodetectors, gas...
sensors, and their flexible counterparts based on MoS$_2$ transistors have recently been reported [45]. The advantages of MoS$_2$ transistors make these sensors highly sensitive, low in power consumption, portable, etc. MoS$_2$ FETs have been reported as various sensors for humidity, H$_2$O$_2$, NO, NO$_2$, NH$_3$, DNA, etc. [46–49]. Shan et al. reported the first MoS$_2$-based field-effect transistor for glucose detection [40]. As shown in Fig. 4a, a back-gate MoS$_2$ FET was fabricated on a SiO$_2$/Si substrate. Source and drain electrodes were patterned by photolithography and e-beam lithography. Au/Ni (70 nm/10 nm) contacts were deposited by evaporation. It should be noted that the MoS$_2$ channel material of about 2 $\mu$m $\times$ 3 $\mu$m was mechanically exfoliated and transferred to the pre-patterned electrodes, as displayed in Fig. 4b. The fabricated transistor was placed in a sample cell and tested.

The measured $I_{ds}$–$V_{ds}$ curves with different concentrations of glucose solutions clearly showed increases of source-drain current with increase of glucose concentration. It should be noted that GO$_x$ enzyme was added to the glucose concentration. Therefore, the sensing is not

| Sensing materials | Applied potential (V vs. Ag/AgCl) | Linear range (mM) | Detection limit | Sensitivity ($\mu$A mM$^{-1}$ cm$^{-2}$) | Response time (s) | Reference |
|-------------------|----------------------------------|-------------------|-----------------|--------------------------------------|-----------------|-----------|
| Cu-MoS$_2$ hybrid | 0.65                             | 0–4               | --              | 1055                                 | --              | [26]      |
| Ni-MoS$_2$ hybrid | –                                | 0–4               | 0.31 $\mu$m     | 1824                                 | < 2             | [27]      |
| AuNPs/MoS$_2$    | 0.93                             | 0.00005–0.1       | 5 nM            | --                                   | 2               | [24]      |
| MoS$_2$ microflowers | 0.35                          | 0–30             | --              | 570.71                               | --              | [25]      |
| MoS$_2$-Au/Pt    | 0.6                              | 0.01–19.07        | 0.39 mM         | 142.68                               | --              | [35]      |
| Au-Pd/MoS$_2$    | 0.3                              | 0.5–20           | 0.40 mM         | 184.9                                | --              | [5]       |
| Ni-MoS$_2$ /rGO   | 0.55                             | 0.005–8.2         | 2.7 $\mu$m      | 256.6                                | < 2             | [30]      |
| AgNPs/MoS$_2$    | 0.31                             | 0.0001–1         | 0.03 $\mu$m     | 9044.6                                | --              | [28]      |
| Cu$_2$O/MoS$_2$  | 0.4                              | 0.01–4.0         | 1.0 $\mu$m      | 3108.71                               | < 2             | [39]      |
| Bilayer MoS$_2$  | –                                | 30–300           | 300 nM          | 260.75                                | < 1             | [40]      |
| MoS$_2$-PPY-AuNPs | 0.45                         | 0.1–80 nM        | 0.08 nM         | 37.35                                 | < 2             | [38]      |
| Co@MoS$_2$/CNTs  | 0.65                             | --               | 80 nM           | 131.69                                | 4.5             | [34]      |
completely enzyme-free. The enhanced channel current upon increasing glucose concentration was attributed to the enzymatic glucose oxidation. The electrons produced from the reaction were transferred to the n-type MoS$_2$ channel and hence increased its conductivity. To illustrate the sensor response to glucose, the real-time $I_{ds}$ measurement was carried out with successive addition of different concentrations of glucose, as shown in Fig. 4c. The measurements started with a pure PBS solution and a higher concentration solution with 1 mM more glucose placed the previous one every minute. The source-drain current response to different concentrations of glucose is plotted in Fig. 4d. Clearly, the MoS$_2$ FET sensor shows a very large linear range for glucose detection, up to 30 mM. The tests were repeated for a low concentration of glucose solutions to probe the detection limit and sensitivity of the MoS$_2$ FET sensor. As shown in the inset of Fig. 4d, the sensor can clearly detect the presence of glucose with a concentration as low as 300 nM. The sensitivity of the MoS$_2$ FET glucose biosensor is calculated to be 260.75 mA mM$^{-1}$. In addition to high sensitivity and low detection limits, the device also showed high stability for up to 45 days. However, the current device has to involve the addition of GO$_x$ enzyme in the glucose solution under test, which makes it less practical for portable applications.

**Conclusion and Perspectives**

This mini-review presents the recent efforts made towards developing enzyme-free biosensor-based MoS$_2$ nanocomposites. These publications all presented facile and low-cost means to high-performance glucose sensors, in terms of sensitivity, linear detection range, and detection limit. These studies undoubtedly open new opportunities towards low-cost and sensitive glucose sensors. The advancements are largely depending on the recent progress made in the synthesis of novel nanocomposites of 2D materials, metallic nanomaterials, and catalytic oxide nanoparticles. It can be expected that more efforts would be invested in this direction, and the experience accumulated is highly beneficial to future studies on related materials for sensing applications.

However, at the same time, one should realize that many efforts are yet required for clinical or any other practical
applications. The stability and reproducibility of these devices are yet to be improved. Either limited storage time or in dry conditions were so far used. Secondly, the chemical synthesis methods are facile and low cost, but whether the methods are scalable remains unclear. New techniques, such as inkjet printing, may be used for repeatable large-scale fabrication of sensors. Although MoS_2-based electrochemical sensors show competitive performance compared with the carbon material-based counterparts, the advantages, e.g., catalytic edge sites of MoS_2, are not substantial. There is clearly a lot of room to really take advantage of the unique properties of MoS_2 for further improvements in non-enzymatic glucose sensing. Furthermore, the development of flexible glucose biosensor-based MoS_2 nanocomposites is important for flexible sensing in healthcare and should be more competitive in the market, which will surely become a research hotspot in the future.

Finally, MoS_2 FET-based sensors show excellent performance in glucose sensing. Given the recent development of MoS_2 FETs, this direction seems very promising in developing low-cost glucose sensors and other types of chemical sensors. It should be reemphasized that the current work reported on MoS_2 FET glucose sensors was only functional to GO_x-doped glucose solution. Future work needs to find alternatives to avoid the use of GO_x for more practical deployment of MoS_2 FET glucose sensors.

Abbreviations

2D: Two-dimensional; 3D: Three-dimensional; CNT: Carbon nanotubes; CTAB: Cetyltrimethylammonium bromide; FETs: Field effect transistors; GO_x: Glucose oxidase; Initials: Full name; MoS_2: Molybdenum disulfide; Ni: Nickel; SEM: Scanning electron microscopy; TEM: Transmission electron microscopy; TMDs: Transition metal dichalcogenides

Authors’ Contributions

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Availability of Data and Materials

Not applicable.

Competing Interests

The authors declare that they have no competing interests.

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References

1. Song J, He W, Shen H, Zhou Z, Li M, Su P et al (2019) Self-assembly of a magnetic DNA hydrogel as a new biomaterial for enzyme encapsulation with enhanced activity and stability. Chem Commun 55(17):2440–2452
2. Uppdeick S, Hinks G (1967) The enzyme electrode. Nature 214(5092):986–988
3. Lu L, Li H, Qu F, Zhang X, Shen G, Yu R (2011) In situ synthesis of palladium nanoparticle–graphene nanohybrids and their application in nonenzymatic glucose biosensors. Biosensors and Bioelectronics 26(8):3500–3504
4. Zhang Y, Xiao X, Sun Y, Sh Y, Dai H, Ni P et al (2013) Electrochemical deposition of nickel nanoparticles on reduced graphene oxide film for nonenzymatic glucose sensing. Electroanalysis 25(4):959–966
5. Li X, Du X (2017) Molybdenum disulfide nanosheets supported Au-Pd binmetallic nanoflowers for non-enzymatic electrochemical sensing of hydrogen peroxide and glucose. Sensors Actuators B Chem 259:536–543
6. Yang J, Zhang W, Gunasekaran S (2010) An amperometric non-enzymatic glucose sensor by electrodepositing copper nanocubes onto vertically well-aligned multi-wall carbon nanotube arrays. Biosensors Bioelectronics 26(1):279–284
7. Salimi A, Sharifi E, Nourbakhsh A, Soltanian S (2007) Immobilization of glucose oxidase on electrodeposited nickel oxide nanofibers: direct electron transfer and electrocatalytic activity. Biosensors Bioelectronics 22(12):3146–3153
8. Kang X, Mai Z, Zou X, Cai P, Mo J (2007) A novel glucose biosensor based on immobilization of glucose oxidase in chitosan on a glassy carbon electrode modified with gold-platinum alloy nanorods/multiwall carbon nanotubes. Anal Biochem 368(1):71–77
9. Li H, Zhang L, Mao Y, Wen C, Zhao P (2019) A simple electrochemical route to access amorphous Co-Ni hydroxide for non-enzymatic glucose sensing. Nanoscale Res Lett 14(1):1–12
10. Mu Y, Jia D, He Y, Miao Y, Wu H (2011) Nano nickel oxide modified non-enzymatic glucose sensors with enhanced sensitivity through an electrochemical process strategy at high potential. Biosens Bioelectron 26(6):2948–2952
11. Zhu H, Li L, Zhou W, Shao Z, Chen X (2016) Advances in non-enzymatic glucose sensors based on metal oxides. J Mater Chem B 8(46):7333–7349
12. Wang G, Lu X, Zhai T, Ling Y, Wang H, Tong Y et al (2012) Free-standing nickel oxide nanoflake arrays: synthesis and application for highly sensitive non-enzymatic glucose sensors. Nanoscale 4(10):3123–3127
13. Jeong J, Yang M, Kim DS, Lee TJ, Choi BG (2017) High performance electrochemical glucose sensor based on three-dimensional MoS_x/gaphene aerogel. J Colloid Interface Sci 506:379–385
14. Li C, Adamcik J, Mezenzega R (2012) Biodegradable nanocomposites of amyloid fibrils and graphene with shape-memory and enzyme-sensing properties. Nat Nanotechnol 7(7):421
15. Gao N, Fang X (2015) Synthesis and development of graphene; “inorganic semiconductor nanocomposites”. Chem Rev 115(16):8294–8343
16. Alarawi A, Rahaliningam V, Fu H, Varadhan P, Yang R, He J (2019) Enhanced photoelectrochemical hydrogen production efficiency of MoS_2-Si heterojunction. Optics Express 27(8):A352–A363
17. Parlik Q, Incel A, Uzun L, Turner APF, Tiwari A (2017) Structuring Au nanoparticles on two-dimensional MoS_2 nanosheets for electrochemical glucose biosensors. Biosens Bioelectron 89:545–550
18. Zheng L, Han S, Liu H, Yu P, Fang X (2016) Hierarchical MoS_2 nanosheet@TiO_2 nanotube array composites with enhanced photocatalytic and photocurrent performances. Small 12(11):1527–1536
19. Wang Q, Cui X, Chen J, Zheng X, Liu C, Xue T et al (2012) Well-dispersed palladium nanoparticles on graphene as a non-enzymatic glucose sensor. RSC Adv 2(15):6245–6249
20. Han D, Zhang J, Weng Z, Kong D, Tao Y, Ding F et al (2019) Two-dimensional materials for lithium/sodium-ion capacitors. Mater Today 11:30–45
21. Liang Y, Li Y, Wang H, Zhou J, Wang J, Regier T et al (2011) CoO_4 nanocrystals on graphite as a synergistic catalyst for oxygen reduction reaction. Nat Mater 10(10):780–786
22. Jaramillo TF, Jorgensen KP, Bonde J, Nielsen JH, Horch S, Chorkendorff I (2007) Identification of active edge sites for electrochemical H_2 evolution from MoS_2 nanocatalysts. Science 315(5834):100–102
23. Karunadasa HI, Montalvo E, Sun Y, Majda M, Long JR, Chang CJ (2012) A molecular MoS_2 edge site mimic for catalytic hydrogen generation. Science 335(6069):698–702
24. Huang K, Liu Y, Liu Y, Wang L (2014) Molybdenum disulfide nanoflower-alloys Compounds 672:600
25. Zhai Y, Li J, Chu X, Xu M, Jin F, Li X et al (2016) MoS_2 microflowers based on graphene nanohybrids and their application in nonenzymatic glucose biosensing. Biosensors and Bioelectronics 89:545
26. Jaramillo TF, Jorgensen KP, Bonde J, Nielsen JH, Horch S, Chorkendorff I (2007) Identification of active edge sites for electrochemical H_2 evolution from MoS_2 nanocatalysts. Science 315(5834):100–102
27. Karunadasa HI, Montalvo E, Sun Y, Majda M, Long JR, Chang CJ (2012) A molecular MoS_2 edge site mimic for catalytic hydrogen generation. Science 335(6069):698–702
28. Huang K, Liu Y, Liu Y, Wang L (2014) Molybdenum disulfide nanoflower-alloys Compounds 672:600
29. Jaramillo TF, Jorgensen KP, Bonde J, Nielsen JH, Horch S, Chorkendorff I (2007) Identification of active edge sites for electrochemical H_2 evolution from MoS_2 nanocatalysts. Science 315(5834):100–102
30. Karunadasa HI, Montalvo E, Sun Y, Majda M, Long JR, Chang CJ (2012) A molecular MoS_2 edge site mimic for catalytic hydrogen generation. Science 335(6069):698–702
31. Huang K, Dong Z, Li Y, Li J, Tang W, Yang H et al (2013) MoS_2 nanosheet functionalized with Cu nanoparticles and its application for glucose detection. Mater Res Bull 48(11):4544–4547
27. Huang J, He Y, Jin J, Li Y, Dong Z, Li R (2014) A novel glucose sensor based on MoS₂ nanosheet functionalized with Ni nanoparticles. Electrochimica Acta 136:41–46
28. Anderson K, Poulsen B, Dudgeon J, Li S, Ma X (2017) A highly sensitive nonenzymatic glucose biosensor based on the regulatory effect of glucose on electrochemical behaviors of colloidal silver nanoparticles on MoS₂. Sensors 17(8):1807
29. Xu H, Han X, Dai X, Liu W, Wu J, Zhu J et al (2018) High detectivity and transparent few-layer MoS₂/glassy-graphene heterostructure photodetectors. Adv Mater 30(13):1706661
30. Geng D, Bo X, Guo L (2017) Ni-doped molybdenum disulfide nanoparticles anchored on reduced graphene oxide as novel electroactive material for a non-enzymatic glucose sensor. Sensors Actuators B Chem 244:131–141
31. Li M, Carter R, Douglas A, Oakes L, Pint CL (2017) Sulfur vapor-infiltrated 3D carbon nanotube foam for binder-free high areal capacity lithium–sulfur battery composite cathodes. ACS Nano 11(5):4877–4884
32. Gong Y, Wang L, Xiong H, Shao M, Xu L, Xie A et al (2019) 3D self-supported Ni nanoparticle@MoS₂ nanotubes anchored on NiMoN pillars for the hydrogen evolution reaction with high activity and anti-oxidation ability. J Mater Chem A 7(22):13671–13678
33. Pan P, Liu L, Guo Q, Wang J, Yang J, Guan X et al (2017) Three-dimensional N-doped carbon nanotube@carbon foam hybrid: an effective carrier of enzymes for glucose biosensors. RSC Advances 7(43):26574–26582
34. Li X, Ren K, Zhang M, Sang W, Sun D, Hu T et al (2019) Cobalt functionalized MoS₂/carbon nanotubes scaffold for enzyme-free glucose detection with extremely low detection limit. Sensors Actuators B Chem 293:122–128
35. Zhou J, Zhao Y, Bao J, Huo D, Fa H, Shen X et al (2017) One-step electrodeposition of Au-Pt bimetallic nanoparticles on MoS₂ nanoflowers for hydrogen peroxide enzyme-free electrochemical sensor. Electrochimica Acta 250:152–158
36. Liu X, Chen K, Ma S, Hao Z, Liang S, Zhou L et al (2019) Synthesis of Au/ CDSe Janus nanoparticles with efficient charge transfer for improving photocatalytic hydrogen generation. Nanoscale Res Lett 14(1):349
37. Lee H, Lim J, Lee C, Back S, An K, Shin JW et al (2018) Boosting hot electron flux and catalytic activity at metal–oxide interfaces of PtCo bimetallic nanoparticles. Nat Commun 9(1):1–8
38. Ma K, Sinha A, Dang X, Zhao H (2019) Electrochemical preparation of gold nanoparticles-polylypyrrole co-decorated 2D MoS₂ nanocomposite sensor for sensitive detection of glucose. J Electrochem Soc 166(2):B147
39. Fang L, Wang F, Chen Z, Qiu Y, Zhai T, Hu M et al (2017) Flower-like MoS₂ decorated with Cu₂O nanoparticles for non-enzymatic amperometric sensing of glucose. Talanta 167:593–599
40. Shan J, Li J, Chu X, Xu M, Jin F, Wang X et al (2018) High sensitivity glucose detection at extremely low concentrations using a MoS₂-based field-effect transistor. RSC Adv 8(15):7942–7948
41. Desai SB, Madhvapathy SR, Sachid AB, Llinas JP, Wang Q, Ahn GH et al (2016) MoS₂ transistors with 1-nanometer gate lengths. Science 354(6308):99–102
42. Kim S, Konar A, Hwang W, Lee JH, Lee J, Yang J et al (2012) High-mobility and low-power thin-film transistors based on multilayer MoS₂ crystals. Nat Commun 3(1):1–7
43. Zhu J, Xu H, Zou G, Zhang W, Chai R, Choi J et al (2019) MoS₂–OH bilayer-mediated growth of inch-sized monolayer MoS₂ on arbitrary substrates. J Am Chem Soc 141(13):5392–5401
44. Pu J, Yomogida Y, Liu K, Li L, Iwasa Y, Takenobu T (2012) Highly flexible MoS₂ thin-film transistors with ion gel dielectrics. Nano Lett 12(8):4013–4017
45. Sahatiya P, Badhulika S (2017) Strain-modulation-assisted enhanced broadband photodetector based on large-area, flexible, few-layered Gr/ MoS₂ on cellulose paper. Nanotechnology 28(45):455204
46. Farimani AB, Min K, Aluru NR (2017) Strain-modulation-assisted enhanced broadband photodetector based on large-area, flexible, few-layered Gr/ MoS₂ on cellulose paper. Nanotechnology 28(45):455204
47. Liu B, Chen L, Liu G, Abbas AN, Fathi M, Zhou C (2014) High-performance chemical sensing using Schottky-contacted chemical vapor deposition grown monolayer MoS₂ transistors. ACS Nano 8(5):5304–5314
48. Li H, Yin Z, He Q, Li H, Huang X, Lu G et al (2012) Fabrication of single-and multilayer MoS₂ film-based field-effect transistors for sensing NO at room temperature. Small 8(1):63–67
49. Zhao J, Li N, Yu H, Wei Z, Liao M, Chen P et al (2017) Highly sensitive MoS₂ humidity sensors array for noncontact sensation. Adv Mater 29(34):1702076

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