Lethal Encephalopathy in an Infant with Hypophosphatasia despite Enzyme Replacement Therapy

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Established Facts

- Morbidity and mortality in hypophosphatasia (HPP) are associated with residual alkaline phosphatase activity and skeletal manifestations.
- Pyridoxine-responsive seizures due to impaired cerebral passage are commonly discussed as most severe CNS manifestations in HPP.
- Effects of enzyme replacement therapy on neurologic symptoms are uncertain.

Novel Insights

- Progression of hypophosphatasia (HPP)-associated encephalopathy can occur even with early onset enzyme replacement therapy.
- HPP-associated encephalopathy due to neuronal ALP deficiency may occur despite pyridoxine responsiveness of neonatal seizures.
- Monitoring of neurological pathologies including encephalopathy has to be implemented for patients with HPP (at risk).

Keywords

Alkaline phosphatase · Asfotase alfa · Bone · Encephalopathy · Enzyme replacement therapy · Hypophosphatasia · Pyridoxine · Rare diseases · Vitamin B6

Abstract

Hypophosphatasia (HPP) is an inborn error of metabolism caused by loss-of-function mutations in the biomineralization-associated alkaline phosphatase gene, encoding tissue-nonspecific alkaline phosphatase (TNSALP). Symptoms include skeletal hypomineralization and extra-skeletal mani-
festations such as pyridoxine (B6)-responsive seizures due to impaired cerebral B6 passage. Since the introduction of enzyme replacement therapy (ERT), skeletal manifestations and B6-responsive seizures were reported to improve significantly. Nevertheless, there is an increasing evidence of B6-independent neurological manifestation of HPP including HPP-associated encephalopathy. Here, we present for the first time the brain alterations of an infant with neonatal HPP who died of neurological complications at the age of 5 months despite early initiation of ERT. CSF analysis showed normal concentrations of biogenic amines reflecting sufficient intracellular B6 availability. Postmortem histopathology revealed severe, localized affection of the cerebral cortex including cortical lesions in layers 2 and 3 in direct proximity to TNSALP-expressing neurons and hippocampal sclerosis. Our findings confirm that TNSALP deficiency may lead to a severe encephalopathy. We hypothesize that HPP-associated encephalopathy resistant to currently available ERT may develop in addition and probably independently of typical B6-responsive seizures in some patients. Prospective, controlled studies with close neurological follow-up including brain imaging are needed to identify patients at risk for severe neurological symptoms despite ERT.

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Introduction

Hypophosphatasia (HPP) is a rare metabolic disorder caused by inactivating mutations in the biomineralization-associated alkaline phosphatase (ALPL), a gene encoding tissue-nonspecific alkaline phosphatase (TNSALP) [1]. The survival and prognosis for patients with life-threatening neonatal HPP have significantly improved since the introduction of enzyme replacement therapy (ERT) with asfotase alpha [2]. Moreover, struc-

![Fig. 1. Comparison of MR images at the age of 4 days (left) and 2.5 weeks (right) before death. Coronal T2-TSE images show a significant brain atrophy with enlarged ventricles and widening of the subarachnoid space at the age of 4.5 months, 2.5 weeks before death (b) compared with the postnatal image (a). Axial diffusion-weighted images reveal bilateral diffusion restriction at the subependymal border of the head of the caudate nucleus (d, arrows) (with correspondingly decreased ADC values [not shown]), adjacent to the third ventricle (f, arrows) and within the hippocampi (f, h, arrows) which were not present at birth (c, e, g). Axial T1-weighted gradient-echo images show regularly progressing myelination of the internal capsule and the splenium of the corpus callosum corresponding to the age of the patient (j) compared with the postnatal MRI (i).](image)
tural brain anomalies and a poorly understood form of encephalopathy have been reported in single patients with neonatal HPP as well as in animal models [3–5].

Here, we present the case of a child with neonatal pyridoxine (PN)-responsive seizures, compound heterozygous genotype, and lethal outcome at the age of 5 months despite start of therapy with recombinant, bone-targeted ERT during the first week of life. We report for the first time histopathological changes of the brain in human HPP in context with a detailed analysis of the clinical course during neurological deterioration. Our findings are in line with a severe encephalopathy, presumably on the basis of cerebral TNSALP deficiency despite early ERT.

**Case Report/Case Presentation**

**Clinical Course**

The male index patient was delivered at term after an uneventful pregnancy and was the first child of nonconsanguineous, Caucasian parents. During the first 48 h, the neonate showed hyperexcitability and had 2 tonic-clonic seizures, which ceased after a single intramuscular dose of 50 mg PN hydrochloride (PN-HCl). EEG and cranial MRI on the second day of life showed no pathologies (Fig. 1). Laboratory investigation revealed undetectable levels of serum alkaline phosphatase (ALP). Genetic analysis confirmed the diagnosis of HPP by detection of compound heterozygosity for *ALPL* (c.667C>T, c.984_986delCTT). Both mutations have been found as homozygous alleles in neonatal HPP [6, 7]. ERT (asfotase alpha, Strensiq®, 1 mg/kg subcutaneously 6 days/week) was initiated on day 6. Mild skeletal hypomineralization of the skull (Fig. 2) was observed on plain radiographs, but the axial and appendicular skeleton appeared normal. The child was discharged after 3 weeks with mild muscular hypotonia for which he received physiotherapy and was seen for regular neurological follow-up. Adhesion to treatment was excellent documented by a rise of serum ALP to supraphysiologic levels.

At 12 weeks of age, the patient developed dyskinesia of the face and upper extremities, which ceased after intramuscular application of 12.5 mg PN-HCl followed by low-dose oral PN-HCl supplementation (5 mg/day). He showed reduced alertness and muscular hypotonia for which he received physiotherapy and was seen for regular neurological follow-up. Adhesion to treatment was excellent documented by a rise of serum ALP to supraphysiologic levels.

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Hypophosphatasia-Associated Encephalopathy

Analysis of biogenic amines gave normal results (Table 1). Vitamin B6 vitamers were markedly elevated in plasma and CSF. Analyses of neurotransmitters and PN derivates in CSF/plasma showed no pathologic findings. PL, pyridoxal 5'-phosphate; MMA, methylmalonic acid; 5-HIAA, 5-hydroxyindoleacetic acid; HVA, homovanillic acid; DOPAC, 3,4-dihydroxyphenylacetic acid; MHPG, 3-methoxy-4-hydroxyphenylglycol; L-DOPA, L-dihydroxyphenylalanine; 3-OMD, 3-O-methyldopa; 5OH Trp, l-5-hydroxytryptophan; 5MTHF, l-5-methyltetrahydrofolate; PN-HCl, PN hydrochloride.

**Table 1.**: Analysis of neurotransmitters and PN derivates in CSF/plasma

| Age at sample acquisition: 149 d | Specific therapy, mg/kgBW/day | asfotase alfa 2 mg/kg | PN-HCl 30 mg/kg |
|---|---|---|---|
| B6 derivates | CSF | PLP | 1,540 | nmol/L | <60 |
| | | PL | 2,310 | nmol/L | 70–83 |
| | | PM | 24 | nmol/L | <4 |
| | | PN | 1,430 | nmol/L | <1 |
| | | PA | 172 | nmol/L | <2 |
| | Serum | MMA | 759 | nmol/L | 73–271 |
| | | PLP | 5,010 | nmol/L | 10–289 |
| | | PL | 5,830 | nmol/L | 0.4–85 |
| | | PM | 28 | nmol/L | <1 |
| | | PN | 19,600 | nmol/L | <1 |
| | | PA | 5,900 | nmol/L | 4–584 |
| Biogenic amines | CSF | 5-HIAA | 261 | nmol/L | 150–800 |
| | | HVA | 671 | nmol/L | 310–1,100 |
| | | HVA/5-HIAA | 2.6 | – | – |
| | | DOPAC | 31 | nmol/L | – |
| | | MHPG | 77 | nmol/L | 98–188 |
| | | L-DOPA | 8.2 | nmol/L | <25 |
| | | 3-OMD | 25.3 | nmol/L | <300 |
| | | 5 OH Trp | 11.5 | nmol/L | <10 |
| | | 5MTHF | 260 | nmol/L | 64–182 |
| | | Neopterin | 24 | nmol/L | 15–35 |
| | | Biopterin | 18.2 | nmol/L | 20–70 |

PN-HCl was increased to 40 mg/kg/day intravenously and due to lack of clinical improvement switched to oral pyridoxal 5'-phosphate (PLP) 40 mg/kg/day after 1 week. In addition, magnesium gluconicum (700–2,100 mg/day) and calcium folinate therapy (3 mg/kg/day) were started to optimize residual ALP activity and intracellular metabolism.

Cerebrospinal fluid CSF and plasma samples were taken at age 5 months, while the patient was on PN-HCl 30 mg/kg/day. Vitamin B6 vitamers were markedly elevated in plasma and CSF. Analysis of biogenic amines gave normal results (Table 1).

Vitamin B6-responsive seizures as observed in our patient have been described as a main neurological manifestation of neonatal HPP with a detrimental outcome and are probably caused by reduced PL passage over the blood-brain barrier as well as impaired cellular uptake into neurons [1, 11]. In a subset of patients with severe HPP, neurologic symptoms may represent the main resistance. Following consensus between parents and medical staff, the patient was moved to palliative care and died at the age of 5.5 months 9 days after extubation due to respiratory failure.

Postmortem Analysis of the Brain

The brain was edematous and had a fresh weight of 666 g on macroscopic examination. The cerebral white matter appeared shrunken and grayish. The cerebral cortex, cerebellum, and brainstem showed no macroscopic abnormalities.

A mild to moderate spongiosis was seen on histopathological examination in the superficial cortical layers II and III but was less pronounced in layer III (Fig. 3a). Shrunken red neurons and occasional apoptotic features were present in layer II, and anti-HLA-DR staining revealed an increase in microglial cells (Fig. 3b). These changes were most pronounced in the frontal lobe, including the cingulate gyrus, and the temporal and parietal cortex. The mammillary bodies displayed a bilateral severe spongiosis and shrunken neurons accompanied by microglia activation (Fig. 3c, d) without bleeding or endothelial proliferations. Similar lesions were observed in a patchy pattern along the walls of the third and lateral ventricles, especially close to the head of the caudate nucleus (Fig. 3e, f). The hippocampus showed a severe loss of neurons in the pyramidal layer within the sectors CA1 and CA4, and the subiculum, accompanied by a severe increase in microglial cells (Fig. 3i–k).

A prominent reactive astrogliosis could be observed in the cerebral white matter (Fig. 3g), whereas the myelination was appropriate for the patient’s age (Fig. 3h). No lesions were detectable in the basal ganglia, thalamus, cerebellum, or brain stem.

Postmortem Analysis of Bone

Specimens from rib, femur, spine, and calvaria were taken and analyzed. Characteristic changes of HPP were found in the zone of enchondral ossification: the layer of columnar cartilage was lengthened with few blood vessels and ballooning of chondrocytes. A broad layer of disorganized and uncalcified osteoid with islands of growth cartilage was present at the border to the growth plate. Osteoblasts were rarely present.

Discussion

TNSALP is a membrane-bound ectophosphatase encoded by ALPL which is found in a vast range of skeletal and extra-skeletal tissues including the central nervous system (CNS) [8–10]. The diversity of tissue-specific substrates, such as pyrophosphates in mineralized tissues and pyridoxal (PL) in the CNS, explains the broad spectrum of symptoms in patients with TNSALP deficiency.

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manifestation of disease (Table 2) and evidence of a severe HPP-associated, vitamin B6-independent encephalopathy has emerged [12–14].

In ALPL knockout mice (Akp2−/−), the effects on neural tube development were described as exceeding B6-associated symptoms and accounting for compromised autonomic nervous system functions [3]. This accounts for the synaptic dysfunction and myelin abnormalities found in both spinal cord and cerebral cortex of Akp−/− mice [15]. Recently, specific metabolic changes involving mainly inhibitory neurotransmitters (GABA, adenosine) and myelin synthesis (NAA, NAG) could be proven in Akp2−/− mice by MRI SPECT [16]. In vitro, TNSALP was found to play an essential role in principal functions such as cell proliferation and differentiation as well as in axonal development in hippocampal neurons in murine-cultured multipotent neural stem cells [17, 18]. Based on histopathological findings in Akp2−/− mice, a primary defect in neural tube development during embryonic life was speculated as the underlying mechanism [3]. This points to a neurologic phenotype beyond B6-dependent seizures with a possible onset during fetal development and a complex pathophysiology, including impaired inhibitory pathways as well as involvement of the limbic system.

Human HPP-associated encephalopathy has so far only been documented by MRI: a severe fatal multicystic encephalomalacia with band-like cortical necrosis has been reported in a child refractory to PN treatment [4]. A patient was found to have MRI alterations in the dorsal-lateral medulla oblongata at the age of 39 days and in the further course of disease a volume loss in the white matter and basal ganglia and a cystic degeneration of the white matter [19]. In contrast to these severe changes, reversible

Fig. 3. Postmortem histopathological examination of the brain. Histopathological examination of the brain reveals spongiosis (a) and microglial activation in layer 2 (b), less pronounced in layer 3, of the cerebral cortex. The mammillary bodies show severe spongiosis and shrunken neurons (c) and prominent microglial activation (d). A lesion subventricular close to the head of the caudate nucleus reveals spongiosis (e), shrunken neurons and apoptosis (e inset) as well as microglial activation (f). The cerebral white matter shows a prominent gliosis (g), whereas no defects are found in the myelination (h). The hippocampus is severely damaged with spongiosis and nerve cell loss in sectors CA1 and CA4 of the pyramidal layer (arrows i, j) and the subiculum (arrow heads i, j), which is confirmed by a concomitant microglial activation (k). (a, c, e, i hematoxylin-eosin stain, b, d, f, k HLA DR immunohistochemistry, g, GFAP immunohistochemistry, Kluver-Barrera myelin stain; original magnification a–f, h: ×4, g: ×40, h: ×10, i–j: ×4).
Table 2. Characteristics and outcome of published HPP cases with primarily neurologic symptoms [4, 5, 11, 19, 20, 26–32]

| Author          | Year | ALPL mutation | Proximity to Ca-binding site | Patient ID (Fig. 4) | Initial ALP, IU/L | Initial neurologic symptoms | Manifestation day | Pyridoxine response | Asfotase treatment | Outcome |
|-----------------|------|---------------|-----------------------------|---------------------|------------------|---------------------------|------------------|--------------------|--------------------|---------|
| Béthenod [26]   | 1967 | NA            | NA                          |                     | Decreased        | Agitation, generalized seizure d5 | 1                | n                  | n                  | 72 d    |
| Baumgartner-Sigl et al. [11] | 2007 | c.677T>C, c.1112C>T | y (Met226) | 1 | 29 | Multifocal myoclonic jerks/tonic seizures | 7 | y | n | 9 mo |
| Balasubramaniam [27] | 2010 | c.891C>A, c.1471G>A | y (Tyr297) | 2 | 14 | Right-sided clonic seizure | 5 | t | n | 5 wk |
| Balasubramaniam [27] | 2010 | c.318G>C | n | <5 | 1 | 0 | 1 | n | 7 d |
| Demirbilek et al. [5] | 2012 | Homozygous c.659G>C | y (Gly220) | 3 | 7 | Generalized tonic-clonic seizures | 2 | y | n | 8 mo |
| Whyte [28] | 2012 | c.920C>T, c.1171C>T | n | 6 | Seizures | NA | y | y | Alive |
| Belachew [29] | 2013 | c.875_881delCAGGGGAinsT, c.1559delT | y (Gly293/Asp294) | 4 | <5 | Facial grimacing, flexion of upper/extension of lower extremities | 1 | y | y | Alive |
| Hofmann et al. [4] | 2013 | c.667C>T, c.1323C>A | y (Arg223) | 5 | 1 | Apnea | 1 | t | n | 10 wk |
| Leung [30] | 2013 | NA | NA | NA | Seizures | 1 | NA | n | 36 d |
| Leung [30] | 2013 | NA | NA | NA | Seizures | 36 | NA | n | 253 d |
| Leung [30] | 2013 | NA | NA | NA | Seizures | 1 | NA | n | 281 d |
| Leung [30] | 2013 | NA | NA | NA | Seizures | 1 | NA | n | 125 d |
| de Roo et al. [19] | 2014 | c.668G>A, c.43_46delACTA | y (Arg223) | 6 | <1 | Generalized seizures | 3 | t | n | 77 d |
| Gülzel Nur [31] | 2016 | Homozygous c.799_804delCACTTC | y (His267-Phe268) | 7 | 1 | Seizures | 1 | t | n | ~6 mo* |
| Fukazawa et al. [20] | 2018 | Homozygous 1559delT | n | 27 | Brief tonic convulsions | 5 | y | y | Alive |
| Oyachi [32] | 2018 | c.1559delT | n | 11 | Seizures | 2 | y | y | Alive* |

Outcome is stated as reflected in each publication or as personally corresponded by the author. Neurologic and skeletal symptoms are summarized and condensed from the original publications. Affected amino acid residues in proximity to the Ca-binding site are graphically depicted in Fig. 4. ALP, alkaline phosphatase; y, yes; n, no; t, temporary; NA, not available; HPP, hypophosphatasia; ALPL, biomineralization-associated alkaline phosphatase. * According to personal correspondence with the author. † Time of death.
bilateral lesions in the dorsal brain stem including the medulla oblongata have been described [20]. MRI in our patient was unremarkable at day 4, but showed a severe atrophy and signal hyperintensities periventricular and in the hippocampi at age 4.5 months. These periventricular changes could be confirmed upon postmortem histological analysis and showed similarities to Wernicke encephalopathy (thiamine deficiency). In our patient, white matter was gliotic without signs of myelin defects. Interestingly, we observed cortical lesions in layers 2 and 3 in direct proximity to TNSALP-expressing neurons in layer 4 as reported in human neocortex [21]. The changes in our patient were reminiscent of mitochondrial DNA depletion syndrome 4A (Alpers type). Hippocampal lesions are described in many inborn errors of metabolism closely related to the manifestation of epilepsy. A similar pattern of cortical lesions may be observed after hypoxic-ischemic brain damage, yet we can exclude hypoxic episodes in our patient until his neurological deterioration at age 4.5 months, where MRI already demonstrated severe brain atrophy. Asfotase alfa was applied from first week of life at the maximum recommended dosage (6 mg/kgBW) and further increased above recommendations (12 mg/kgBW) at first signs of EEG alterations.

In our patient, administration of high-dose PN-HCl or PLP as the active form of vitamin B6 was clinically ineffective to arrest the progression of severe encephalopathy. Normal concentrations of biogenic amines in CSF reflect sufficient intracellular PLP availability as PLP is an important cofactor in the synthesis of these neurotransmitters. Incomplete rescue of the neurologic phenotype by vitamin B6 administration has also been observed in 2 Akp2−/− mouse strains, although to different extents: while 2/3 of animals responded to treatment in the first mouse model, only temporary amelioration was observed in the second [3, 22].

So far, HPP-associated encephalopathy has only been described in severe neonatal forms of the disease with residual ALP enzyme activity below 10% (Table 2). With such low residual activity, formation of PL is severely impaired, as illustrated by increased PLP concentrations in the plasma and CSF of patients with HPP before supplementation of B6. It has therefore been discussed if additional vitamin B6 supplementation may have a deleterious effect by increasing extracellular PLP concentrations to a toxic level [30] [23]. HPP-associated encephalopathy has been observed irrespective of vitamin B6 supplementation [4, 19] and at least 1 patient with higher residual ALP activity responded to increased doses of vitamin B6 treatment [20]. Still, excessive concentrations of PLP in the extracellular space cannot be excluded as a contributing factor in HPP-associated encephalopathy. Interestingly, our patient exhibited only mild features of mineralization deficits at birth despite severe ALP deficiency. While demineralization signs in radiographs occurred in the clinical course, the discrepancy between moderate skeletal affection and severe neurologic manifestation suggests specific pathomechanisms in these 2 organ systems with a certain degree of independent affection. Thus, a mild skeletal phenotype at birth does not rule out lethal HPP. In our case, the mild initial phenotype with continuous deterioration and missing data on neurologic complications under treatment posed/contributed to/an extremely difficult decision basis for caretakers and parents.
Kinetic characterization of ALPL mutations revealed that some mutations were efficient in hydrolyzing inorganic pyrophosphate while inefficient in hydrolyzing PLP, which could explain why some patients with neonatal HPP manifest neurologic disease and others do not [24]. Interestingly, most patients with primarily neurologic manifestation carry mutations in proximity to the Ca-binding site of TNSALP, distant from the active site pocket (Fig. 4). An eventual genotype-phenotype association caused by substrate or tissue specificity remains to be determined.

Outcome data from patients with perinatal HPP and ERT are sparse. Our patient received ERT from a very early stage of disease. Like in other metabolic disorders, ERT in HPP is not expected to pass the blood-brain barrier and rescue a CNS disease. Nonbone targeted or intrathecal ERT may be needed to treat a CNS disease associated with neonatal HPP. While the postnatal impact of cerebral TNSALP deficiency might be positively addressed/influenced by such an option, developmental deficiencies, as observed in animal studies, might be irreversible. At the present state of therapeutic options, the aspects of clinical neurological development including HPP-associated encephalopathy have to be implemented in information provided to parents as well as current monitoring guidelines for children with severe HPP [25].

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Statement of Ethics

This study protocol was reviewed, and the need for approval was waived by the Ethical Committee of the Medical University of Vienna, as published research complies with the guidelines for human studies in accordance with the World Medical Association Declaration of Helsinki. Written informed consent to perform autopsy and for publication of their clinical details and/or clinical images was obtained from the parents of the patient.

Conflicts of Interest Statement

The authors have no conflicts of interest to declare.

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Author Contributions

Conception and design of the work were conceived by A.R., D.A.E., C.H., M.S., B.P., and G.H. Acquisition of data was performed by A.R., J.P., D.A.E., K.S., M.F., S.L., M.S., and G.H. Data were analyzed by A.R., C.H., J.P., D.A.E., M.F., S.L., M.S., and G.H. Interpretation of data for the work was performed by A.R., C.H., J.P., D.A.E., K.S., M.F., S.L., M.S., B.P., and G.H. The work for important intellectual content was drafted/revised by A.R., C.H., B.P., and G.H.

Data Availability Statement

All datasets generated during and/or analyzed during the current study are not publicly available but are available from the corresponding author on reasonable request.
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