Acknowledgments

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About the Author

Dr. Donnelly is a professor of applied statistics at the University of Oxford and a professor of statistical epidemiology at Imperial College London. As a statistician and epidemiologist, her research interest is the spread and control of infectious diseases, with a particular focus on outbreaks.

Limited Scope of Shorter Drug Regimen for MDR TB Caused by High Resistance to Fluoroquinolone

Pravin K. Singh, Amita Jain

Author affiliation: King George Medical University, Lucknow, India

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Resistance to second-line tuberculosis drugs for patients with multidrug-resistant tuberculosis has emerged globally and is a potential risk factor for unfavorable outcomes of shorter duration drug regimens. We assessed the proportion of patients eligible for a shorter drug regimen in Uttar Pradesh, India, which had the highest rate of multidrug-resistant tuberculosis in India.

India has the largest burden of multidrug-resistant (MDR) tuberculosis (TB) worldwide (1). The success rate for MDR TB treatment is low (47%), largely caused by death, suboptimal adherence of patients to long treatment courses, and frequent drug-related adverse events (2).

In 2016, the World Health Organization recommended a shorter drug regimen (9–12 months) for patients with MDR TB or rifampin-resistant TB who had not received second-line drugs (SLDs) and in whom resistance to fluoroquinolones and injectable SLDs is considered highly unlikely (3). A shorter regimen is a promising step toward high treatment success rates. Recently, this regimen was instituted in Uttar Pradesh, which has ≈20% of the total
MDR TB and rifampin-resistant TB burden in India (2). We assessed the proportion of rifampin-resistant TB patients in Uttar Pradesh who would be eligible for a shorter regimen under a programmatic setting.

Under the Revised National Tuberculosis Control Program for India, all TB patients are tested for drug susceptibility for at least rifampin by the GeneXpert MTB/RIF assay (http://www.cepheid.com), which is available in every district laboratory. Two samples per patient are usually collected, and if rifampin-resistant TB is confirmed, the second sample is transported to the state laboratory for susceptibility testing of SLDs by line-probe assay (LPA) (LPA-SLD Genotype MTBDRsl; Bruker, https://www.hain-lifescience.de). Because it might take ≈1 week to obtain an LPA-SLD result, a shorter treatment regimen is initiated without delay for all patients with rifampin-resistant TB, except for patients who have received SLDs, those with extrapulmonary TB, and those who are pregnant (4). If resistance to SLD is detected, patients are given an appropriate, longer treatment regimen.

Over a 2-month period (July–August 2018), sputum samples from 708 patients with rifampin-resistant TB (1 sample/patient) were collected before treatment with any SLDs and submitted to our reference laboratory for LPA-SLD. Smears from each sputum sample were examined for acid-fast bacilli (AFB). AFB-positive samples were subjected directly to LPA, AFB-negative samples were cultured in Mycobacterium growth indicator tube liquid medium, and LPA was performed indirectly for culture isolates, if recovered. LPA testing on smear-positive sputum samples with inconclusive results (Mycobacterium tuberculosis not detected/indeterminate resistance/invalid) was repeated by using the indirect method. Resistance patterns for fluoroquinolones and injectable SLD (as determined by LPA-SLD) were analyzed.

Of 708 samples, drug susceptibilities were determined for 541 (498 by direct LPA and 43 by indirect LPA). For the remaining 167 samples (AFB negative, 112; AFB scanty, 52), results were inconclusive because of no growth or contamination (Figure). Plausible reasons for lower rate of culture recovery include testing of only AFB-negative/scanty samples, expected low culture yield if patients have a history of tuberculosis treatment or if a sample was of suboptimal quality (5), and a harsh decontamination process used for the sample. This last reason is unlikely because the culture contamination rate was not lower than standard range.

In our study, a high proportion (21.4%) of patients were smear negative, although they were given a diagnosis of rifampin-resistant TB by GeneXpert. Low analytical sensitivity for smear microscopy (compared with that for GeneXpert) could be a potential reason. This observation highlights the need of submitting 2 quality samples for LPA.

Of 541 conclusive LPA-SLD results, the proportion of strains resistant to only fluoroquinolone was 50.1%, to injectable SLD 1.5%, and to fluoroquinolone and injectable SLD 8.3%. Fluoroquinolone resistance (with or without injectable SLD resistance) was high, indicating that 58.4% of patients were ineligible for a shorter regimen. This estimate raises concern because, in an earlier trial of a shorter regimen, fluoroquinolone resistance was a key risk factor for a bacteriologically unfavorable outcome (6).

Our estimate might not be a true representation for fluoroquinolone resistance burden because we included only
rifampin-resistant TB patients in our study, not patients with rifampin-susceptible but fluoroquinolone-resistant TB. Nevertheless, our estimate might be extrapolated for a programmatic setting in Uttar Pradesh because the number of patients in the study was ≈7.0% of MDR TB and rifampin-resistant TB patients reported in Uttar Pradesh during 2017 (1).

In India, a high incidence of fluoroquinolone-resistant TB has been reported (7,8). It is believed that widespread and indiscriminate use of fluoroquinolones was a key contributor. However, primary resistance caused by transmission of fluoroquinolone-resistant strains might be a potential reason for such a high rate of fluoroquinolone resistance. This hypothesis is well supported by increasing evidence for countries with high burdens of TB where MDR TB is increasing in patients who have not received TB treatment (9,10). In our study, we could not stratify our data into new and previously treated patients because of the observational design and limited access to patient information.

Considering high levels of fluoroquinolone resistance and possible intolerance/resistance to certain drugs in a regimen, only ≈40.0% of rifampin-resistant TB patients would be eligible for a shorter drug regimen in our setting. Thus, a shorter regimen should not be an obvious choice for empiric treatment of MDR TB or rifampin-resistant TB patients in this region.

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About the Authors
Dr. Singh is a microbiologist at the Intermediate Reference Laboratory, Department of Microbiology, King George Medical University, Lucknow, Uttar Pradesh, India. His research interests are diagnosis and epidemiology of infectious diseases.

Dr. Jain is a professor and head of the Department of Microbiology, King George Medical University, Lucknow, Uttar Pradesh, India. Her research interests are tuberculosis, virology, and clinical microbiology.

References
1. World Health Organization. Global tuberculosis report 2018 [cited 2019 Jun 24]. https://www.who.int/tb/publications/global_report
2. India TB. Report 2018—Revised National Tuberculosis Control Program Annual Status Report 2018: Central TB Division, Directorate General of Health Services, Government of India; 2018 [cited 2019 Jun 24]. https://tbcindia.gov.in
3. World Health Organization. Treatment guidelines for drug-resistant tuberculosis, 2016 update [cited 2019 Jun 24]. https://apps.who.int/medicinedocs/en/m/abstract/Js23097en
4. Programmatic Management of Drug Resistance, 2017. Guidelines on programmatic management of drug resistant tuberculosis in India 2017: Central TB Division, Directorate General of Health Services, Government of India; 2018 [cited 2019 Jun 24]. https://tbcindia.gov.in/index1.php?lang=1&level=2&sublinkid=4780&id=3306
5. Shi J, Dong W, Ma Y, Liang Q, Shang Y, Wang F, et al. GeneXpert MTB/RIF outperforms Mycobacterial culture in detecting Mycobacterium tuberculosis from salivary sputum. BioMed Res Int. 2018;2018:1514381. https://doi.org/10.1155/2018/1514381
6. Aung KJ, Van Deun A, Declercq E, Sarker MR, Das PK, Hossain MA, et al. Successful ‘9-month Bangladesh regimen’ for multidrug-resistant tuberculosis among over 500 consecutive patients. Int J Tuberc Lung Dis. 2014;18:1180–7. https://doi.org/10.5588/ijtld.14.0100
7. Agrawal D, Udwadia ZF, Rodriguez C, Mehta A. Increasing incidence of fluoroquinolone-resistant Mycobacterium tuberculosis in Mumbai, India. Int J Tuberc Lung Dis. 2009;13:79–83.
8. Jain A, Dixit P, Prasad R. Pre-XDR and XDR in MDR and ofloxacin and kanamycin resistance in non-MDR Mycobacterium tuberculosis isolates. Tuberculosis (Edinb). 2012;92:404–6. https://doi.org/10.1016/j.tube.2012.05.010
9. Zhao M, Li X, Xu P, Shen X, Gui X, Wang L, et al. Transmission of MDR and XDR tuberculosis in Shanghai, China. PLoS One. 2009;4:e4370. https://doi.org/10.1371/journal.pone.0004370
10. Shah NS, Auld SC, Brust JC, Mathema B, Ismail N, Moodley P, et al. Transmission of extensively drug-resistant tuberculosis in South Africa. N Engl J Med. 2017;376:243–53. https://doi.org/10.1056/NEJMoA1604544

Address for correspondence: Amita Jain, Department of Microbiology, King George Medical University, Lucknow, Uttar Pradesh, India; email: amita602002@yahoo.com