ASPP2 expression predicts the prognosis of patients with hepatocellular carcinoma after transcatheter arterial chemoembolization

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Abstract. Transcatheter arterial chemoembolization (TACE) induces ischemia-hypoxia and local chemotherapy-induced cytotoxicity which destroys cancerous cells. However, some patients do not respond to TACE. The causes for such a lack of response remain unclear. Recent studies have revealed that self-regulation of apoptosis-stimulating p53 protein 2 (ASPP2) may play an important role in promoting cell survival under hypoxic conditions as well as chemotherapy resistance via autophagy in various types of cancer. We measured the expression of ASPP2, autophagy-related proteins and apoptotic proteins by western blot assays. Multivariate logistic regression analysis was used to identify the independent risk factor. The present study found that ASPP2 expression was negatively correlated with that of BECN-1 (Beclin-1) in hepatocellular carcinoma (HCC) tissues. The expression of ASPP2 was lower while that of Beclin-1 was higher in patients who underwent recurrence of HCC following TACE, than in those who do not undergo such a relapse. ASPP2 expression was also lower in cancerous tissues subjected to TACE, compared with that of directly resected cancerous tissue. The expression of LC3-II was also higher in patients with post-operative recurrence of HCC than in those without relapse. In vitro experiments showed that administration of an autophagy inhibitor, together with hypoxia activation and 5-FU treatment, promoted apoptosis in HepG2 liver cancer cells and primary HCC cells. Multivariate logistic regression analysis revealed that ASPP2 expression in cancer tissue following TACE is an independent risk factor for HCC recurrence as well as overall survival. Higher levels of ASPP2 expression were notably associated with higher objective responses evaluated via mRECIST. Thus, patients with resectable HCC showing high levels of ASPP2 expression may benefit from neoadjuvant TACE prior to resection.

Our study provided a novel biomarker for HCC prognosis following TACE, based on cell survival mechanisms related to autophagy.

Introduction

Transcatheter arterial chemoembolization (TACE) is a selective therapeutic strategy which is optimal for hepatocellular carcinoma (HCC) patients who are not indicated for direct surgical resection, such as those with a single cancerous nodule (1). The value of preoperative adjuvant TACE as an optional therapeutic strategy to improve prognoses for advanced HCC patients with resectable HCC is a subject of controversy (2,3). Some studies have shown that preoperative adjuvant TACE does not improve the condition of patients with resectable HCC, and that these patients do not benefit from preoperative TACE followed by surgical resection (4,5). The findings demonstrated that TACE is not beneficial in advanced HCC patients, and that preoperative TACE leads to worsen liver function, which is associated with higher mortality. However, another study demonstrated that patients showing an objective response to preoperative TACE may have a better prognosis following surgical resection, than non-responders (6). A study from Chinese Taiwan evidenced the association of preoperative TACE with improved long-term outcomes for patients, despite not having an effect on disease-free survival (DFS) or overall survival (OS) (7). Majno et al (8) reported that 62% of HCC patients, who were initially deemed unresectable, experienced a downstaging of their tumors due to necrosis induced by TACE, followed by a significant improvement in DFS after liver resection. For some patients with unresectable tumors, TACE administration further improved their suitability for resection. Therefore, objective response and the presence of HCC downstaging following preoperative TACE, may be considered as a predictor of improved prognosis following neoadjuvant TACE and liver resection.

TACE may cause changes in various markers, such as stemness markers, markers of hypoxia and tumor stromal markers (9,10). Furthermore, Xu et al (11) reported that administration of TACE prior to surgery may activate hypoxia-inducible factor 1α (HIF-1α), which is responsible for a poor prognosis of HCC. Increased expression of these molecular markers may biologically promote aggressive HCC. Autophagy may protect cancerous cells from cell death.
due to adverse conditions, such as hypoxia, starvation, and chemotherapy-induced cellular apoptosis (12,15). Therefore, evaluation of autophagy in cancer tissue following TACE may be valuable for assessing the resistance to hypoxia and chemotherapy and to predict therapeutic objective response as well.

Apoptosis-stimulating p53 protein 2 (ASPP2) may stimulate the apoptosis of cancer cells with, or without, exerting a pro-apoptotic function via p53 signaling. It is defined as a tumor suppressor which promotes apoptosis and inhibits cell growth (16,17). Furthermore, in HCC, XIAP expression is reduced by ASPP2, which facilitates sensitivity to chemotherapeutics in a p53-free manner (18). A recent study reported that, in response to oxidative stress, Beclin-1-mediated autophagy is pivotal to the survival of pediatric leukemia cells (19). In addition, administration of exemestane was found to result in increased Beclin-1 and LC3 in cancer cells, where stromal levels of Beclin-1 are associated with enhanced proliferation of cancer cells and a lower sensitivity to neoadjuvant endocrine treatment (20,21). Therefore, under stress conditions, autophagy in cancer tissues may contribute to cell survival and cell proliferation.

The present study investigated the expression of ASPP2 and the autophagy marker, Beclin-1, in carcinoid and para-carcinoid tissues of HCC patients following TACE. Results of in vitro experiments indicated that inhibition of autophagy promoted cell apoptosis in HepG2 liver cancer cells under adverse conditions. We provide a novel biomarker for predicting the objective response and clinical prognosis following TACE based on cell survival via autophagy under hypoxia conditions and local chemotherapy.

Materials and methods

Patient enrollment. This observational study was approved by the Medical Ethics Committee of the People's Hospital of Danyang (Danyang, Jiangsu). Written consent for collection of the tissue samples was acquired from all patients prior to surgery. HCC samples were collected directly from resected tissues of 165, and 437 patients subjected to TACE in combination with liver resection, including unresectable patients who needed tumor staging therapy via preoperative TACE, and resectable patients who received preoperative neoadjuvant TACE. Samples were collected from primary HCC patients between January 2015 and February 2018 at the Department of Radiology, People's Hospital of Danyang. In the present study, patient enrollment was based on the following criteria: i) diagnoses were confirmed via histological testing; ii) patients did not exhibit distant metastasis; iii) patients had no severe complications or other malignancies. Staging of tumors was based on criteria stipulated by the Barcelona Clinic Liver Cancer (BCLC) staging system (22). Overall survival (OS) was defined as duration from the time of liver resection to that of death or the last follow-up. Progression-free survival (PFS) was defined as duration from the date of resection without any tumor progression, including deterioration of liver function, recurrence or metastasis. Baseline characteristics and survival data, based on follow-ups of all enrolled patients, were collected for statistical analyses. Resected tissues were prepared for studies aimed at the determination of protein expression and isolation of primary HCC cells.

Western blot analysis. Resected tissues were homogenized in RIPA lysis buffer (Beyotime) to obtain total protein, the concentration of which was measured using a BCA kit (Thermo Fisher Scientific, Inc.). An amount of 10 µg of protein in each sample were adjusted to the same volume and separated via 10% SDS-PAGE. Proteins in the gel were electrically transferred onto a polyvinylidene difluoride (PVDF) membrane (Millipore). Unoccupied sites on the membrane were blocked using 5% milk for 1 h, following which the PVDF membrane was trimmed into pieces according to the relative molecular weights of the target proteins. Trimmed membranes were immersed in solutions containing primary antibodies (all provided by Abcam) for Beclin-1 (dilution 1:1,000, cat. no. ab210498), ASPP2 (dilution 1:2,000, cat. no. ab181377), LC3-I (dilution 1:800, cat. no. ab66278), LC3-II (dilution 1:1,000, cat. no. ab51520), cleaved caspase 3 (dilution 1:1,500, cat. no. ab32042), cleaved caspase 8 (dilution 1:2,000, cat. no. ab119809) and GAPDH (dilution 1:2,000, cat. no. ab181602) for 1 h at room temperature, followed by measurement of the protein signals via the Odyssey System (Lico Biosciences, Germany) and band intensities using the BioRad Image Lab Software 3.0 (Bio-Rad Laboratories).

Cultivation of primary HCC cells and a liver cancer cell line. Liver cancer (HepG2) cells were cultivated in Dulbecco's modified Eagle's medium (DMEM) (Gibco; Thermo Fisher Scientific, Inc.) with 10% fetal bovine serum, FBS at 37°C under 5% CO_2.

Tumor as well as normal specimens were collected from the primary tumor and tumor-surrounding parenchyma in pre-cooled DMEM-F12 (10% FBS) of 5 HCC patients (Table I). Next, the cells were dissociated from the tissues in pre-heated 0.1% collagenase IV, followed by filtration and resuspension of the cell pellet in cool medium. After 3 rounds of centrifugation at 81 x g for 12 min, the hepatocytes were separated as a pellet. The pellet containing liver cells was resuspended in ice-cold DMEM-F12 with 20% FBS, following which triple centrifugation was performed at 81 x g for 12 min at 4°C in order to separate the purified hepatocyte population (pellet) from the non-parenchymal cells (supernatant). The pellet contained partially purified hepatocytes. To further purify the hepatocytes, the pellet was placed in a dish without any collagen I. The supernatant was carefully removed 4 h later, and tumor cells and non-tumor cells were inoculated on a 24-well plate for 48 h. At 16 h before co-culture, the original medium was refreshed, and cells were rinsed twice in PBS. Cell culture was continued in medium containing 1% FBS. Hydroxychloroquine (HCQ) (10 µM) was administrated to inhibit the autophagy process in in vivo experiment to explore the role of autophagy in the cellular protection in primary HCC cells or in HepG2 cells.

MTT assay. MTT assay was used to measure the cell viability as previously reported (14). Cells were seeded in 96-well plates (2~3 x 10^3 cells/well), and 5-FU (20 µg/ml) or HCQ (10 µM)
was added for 2-4 days. Hypoxic stimulation was given at an oxygen concentration of 5% (5% CO₂ and 90% N₂). The cells were cultured with 100 μl sterile MTT dye (Sigma-Aldrich; Merck KGaA) for 4 h, followed by the removal of medium and supplementation with 150 μl of DMSO (dimethyl sulfoxide). After shaking, the absorbance was measured.

**Immunohistochemical staining.** The carcinoma tissues from patients were immediately fixed by 5% formalin and then embedded in paraffin blocks. Before immunohistochemical staining, antigen was retrieved according to thermal remediation method and the slides were incubated with the 3% goat serum for 30 min. The paraffin block was cut into 5-μm sections and then incubated with primary antibody for rat anti-human ASPP2 (BD Pharmingen) in 5% rabbit serum overnight at 4°C followed by washing and incubation with ABC (avidin-linked biotin complex) rabbit anti-rat Ig. The signals were detected using DAB (3,3’-diaminobenzidine) substrate and hematoxylin was used for counterstaining. The slides were dehydrated, coverslipped, and observed under a Zeiss microscope (Zeiss, Germany). A negative control in the absence of primary antibodies and incubated with isotype-matched immunoglobulins was also used for immunostaining. All captured images were analyzed by ImageJ software (National Institutes of Health, Bethesda, MD, USA) to quantify the expression level according the ratio of fluorescent intensity of primary antibody-stained section to negative control. The mean of expression level was used to divide the subjects into two groups.

**Patient follow-up.** Follow-up of patients ended in February 2018. During the follow-up, patients were required to attend an examination every 2 months to determine α-fetoprotein (AFP) levels in the serum, submit to imaging examinations including abdominal ultrasonography and undergo CT or MRI scan every 6 months. In addition, an 18F-FDG PET-CT scan every 6 months was also suggested to clarify distant metastasis status.

Metastasis was evaluated via a combination of 18F-FDG positron emission tomography and CT (PET-CT) every 6 months, following hepatectomy. Recurrence of HCC and liver complications were the major causes of death.

**Statistical analysis.** Data analysis was conducted using SPSS software 20.0 (IBM, Corp.). Relative expression of Beclin-1 and ASPP2 are expressed as mean ± standard deviation, and compared via the pairwise Student’s t-test. Correlation between the Cezanne and staining scores of Beclin-1 and ASPP2 were detected using linear correlation analysis. Independent risk factors affecting 1-year survival of patients were identified via multiple logistics regression analysis, while survival was assessed via Kaplan-Meier curves. P<0.05 was considered to indicate a statistically significant difference.

**Results**

**Patient characteristics.** In total, 165 patients who underwent direct surgical resection and 437 patients who underwent liver resection following TACE were retrospectively enrolled in the study. Among patients with direct resection, all patients were candidates for resection. Among the 437 patients who received TACE before resection, 205 patients were candidates for direct resection and 232 were unresectable at initiation. Baseline characteristics of the unresectable and resectable patients are presented (Tables II and III). Baseline characteristics of the unresectable HCC patients were compared before surgical resection, and no significant differences were found between the baseline characteristics of the high ASPP2 expression and low ASPP2 expression groups. The tested baseline characteristics included age, sex, platelet count (PLT), serum levels of blood urea nitrogen (BUN), tumor size, number of cancerous nodules, histological grading, Child-Pugh classification, microvessel invasion (MVI), portal vein tumor thrombus and ascites. However, AFP levels were significantly higher in unresectable HCC patients with a higher ASPP2 expression. Among the 437 patients who underwent surgical resection following TACE, 267 patients were considered as responders, defined as exhibiting a complete response (CR), or a partial response (PR) according to modified RECIST criteria for HCC (23). The remaining 170 patients exhibited stable disease (SD) or progressive disease (PD), defined according to modified RECIST criteria. In resectable and unresectable patients, a higher ASPP2 expression was associated with a positive therapeutic response to TACE according to modified RECIST criteria (Tables II and III).

**Expression of ASPP2 and Beclin-1 in the HCC tissues of patients with or without TACE.** In order to investigate ASPP2 expression in carcinoma tissues following TACE, carcinoma and para-carcinoma tissues were collected from patients who underwent surgical resection directly or following TACE. Next, ASPP2 expression in carcinoma and para-carcinoma tissues was assessed by western blot
assay. ASPP2 expression in carcinoma tissues of patients who received surgical resection either directly or following TACE was significantly lower, compared with that in the para-carcinoma tissues (Fig. 1A and B). The ratio of ASPP2 in para-carcinoma to carcinoma tissues (Para-CA/CA) was significantly higher in patients receiving surgical resections following TACE, than that in those receiving surgical resections directly (Fig. 1C). Beclin-1 expression in carcinoma tissues and para-carcinoma tissues of HCC patients who received surgical resection directly or following TACE was assessed by western blot assay. Beclin-1 expression in carcinoma tissues was observably increased, compared with that of para-carcinoma tissues in HCC patients following TACE. However, there was no observable difference in Beclin-1 expression between carcinoma and para-carcinoma tissues from HCC patients who did not undergo TACE before surgical resection (Fig. 1D and E). In addition, Beclin-1 expression in the histological samples of patients who underwent TACE, was negatively correlated with ASPP2 expression (Fig. 1F). These data indirectly demonstrated that a reduction in ASPP-2 expression, which occurred in the carcinoma tissues of patients following TACE, was associated with autophagy which was dependent on Beclin-1.

**Culture in hypoxia and chemotherapy-induced cell apoptosis promote autophagy in HepG2 liver cancer cells and primary HCC cells from HCC patients.** Expression of LC3-II and LC3-I in carcinoma and para-carcinoma tissues of samples from patients who underwent surgical resection following TACE was assessed by western blot assay. Results indicated that LC3-II expression was increased (Fig. 2A) and the LC3-II to LC3-I ratio was higher in carcinoma tissues compared with those in para-carcinoma tissues (Fig. 2B), suggesting that autophagy flux in cancer cells was increased following TACE. Examination of primary HCC cells isolated from a patient who underwent palliative surgical resection following TACE failure indicated that administration of 5-FU at a low concentration (20 µg/ml) and in vitro hypoxia stimulation led to an increase in LC3-II expression and a higher ratio of LC3-II/LC3-I (Fig. 2C). Application of the autophagy inhibitor, hydroxychloroquine

| Variable                        | High ASPP2 expression (n=106) | Low ASPP2 expression (n=126) | P-value |
|---------------------------------|-------------------------------|------------------------------|---------|
| Mean age, years                 | 54.61±10.54                   | 56.85±12.67                  | NS      |
| Gender, male n (%)              | 77 (72.64)                    | 80 (63.49)                   | NS      |
| PLT (10^3/µl)                   | 187.3±68.91                   | 179.1±76.16                  | NS      |
| BUN (mg/dl)                     | 18.7±8.18                     | 17.6±10.15                   | NS      |
| Child-Pugh classification, n (%)|                               |                              | NS      |
| A                               | 28 (26.40)                    | 45 (35.71)                   |         |
| B                               | 66 (62.26)                    | 60 (47.61)                   |         |
| C                               | 12 (11.34)                    | 21 (16.68)                   |         |
| Hepatitis B, n (%)              | 80 (75.47)                    | 88 (69.84)                   |         |
|AFP (ng/ml)                      | 2,376.98±317.12               | 2,087.33±286.89              | 0.036*  |
| Tumor size, n (%)               |                               |                              | NS      |
| ≤5 cm                           | 59 (55.66)                    | 54 (42.86)                   |         |
| >5 cm                           | 47 (44.34)                    | 72 (57.14)                   |         |
| BCLC stage, n (%)               |                               |                              | NS      |
| 0-B                             | 63 (59.33)                    | 57 (45.24)                   |         |
| C-D                             | 43 (40.67)                    | 65 (54.76)                   |         |
| Histological grading, n (%)     |                               |                              | NS      |
| Well                            | 25 (23.54)                    | 36 (28.57)                   |         |
| Moderate                        | 48 (45.67)                    | 51 (40.48)                   |         |
| Poor                            | 33 (30.79)                    | 38 (30.16)                   |         |
| MVI %                           | 48 (45.67)                    | 66 (52.38)                   | NS      |
| Portal vein thrombi, n (%)      | 28 (26.41)                    | 47 (37.30)                   | NS      |
| Ascites n, %                    | 44 (41.51)                    | 50 (39.68)                   | NS      |
| Objective therapeutic response, n (%)|                             |                              | 0.017*  |
| CR and PR                       | 62 (58.49)                    | 26 (20.63)                   |         |
| SD and PD                       | 44 (41.51)                    | 100 (79.37)                  |         |

Chi-square test or t-test were used for statistical analysis. *P<0.05 indicates statistical significance. HCC, hepatocellular carcinoma; ASPP2, apoptosis-stimulating p53 protein 2; PLT, platelet level; BUN, blood urea nitrogen; AFP, α-fetoprotein; BCLC, Barcelona Clinic Liver Cancer; MVI, microvessel invasion; CR, complete response; PR, partial response; SD, stable disease; PD, progressive disease; NS, not significant.
HCQ (10 µM), promoted apoptosis of primary HCC cells under hypoxic cultivation and 5-FU treatment, as evidenced by the increased expression of cleaved caspase-3 and cleaved caspase-8 (Fig. 2D) and inhibition of proliferation, as measured via MTT assays (Fig. 2E). Furthermore, we assessed the expression of LC3-II as well as the ratio of LC3-II to LC3-I in the HepG2 cell line, under 5-FU treatment and hypoxia cultivation. The results indicated a significant increase in the expression of LC3-II and a higher ratio of LC3-II to LC3-I which was markedly similar to that observed in the primary HCC cells (Fig. 2F). The application of the autophagy inhibitor did not result in the change of biological behavior, including apoptosis, proliferation, and chemotherapy resistance (data not shown). The above findings illustrated that autophagy initiated by hypoxia or a chemotherapy drug may promote the survival of primary HCC cells and protect them from adverse conditions.

Expression of ASPP2 predicts the prognosis of neoadjuvant TACE and liver resection. In order to verify the prognostic significance of ASPP2 expression in predicting the survival of HCC patients following TACE and surgical resection, we constructed a cohort in the 437 patients treated with TACE following surgical resection. We collected and analyzed data related to survival and objective response. Immunohistochemical (IHC) assays and histopathological examinations were used to obtain a semi-quantitative evaluation of ASPP2 expression in tissues. These patients were divided into a low expression group and a high expression group according to a cut-off value determined by median expression levels. As shown in Table IV, patients with a higher ASPP2 expression had a low 1-year mortality rate following combination therapy with surgical resection (high expression vs. low expression, 65.3 vs. 26.7%; \( \chi^2 = 12.91; P=0.003 \)). Multiple logistics regression revealed that low ASPP2 expression levels, MVI and histopathological grading of carcinoma differentiation were an independent risk factor for TACE failure. PVTT (OR=2.17; 95% CI 1.34-3.61; \( P=0.020 \)), Child-Pugh classification (OR=2.76; 95% CI 1.35-4.12; \( P=0.031 \)) and ASPP2 (OR=2.12; 95% CI 1.68-3.75; \( P=0.004 \)) were independent risk factors for TACE failure.

| Variable                        | High ASPP2 expression (n=93) | Low ASPP2 expression (n=112) | P-value |
|--------------------------------|-------------------------------|-----------------------------|---------|
| Mean age, years                | 54.23±13.64                  | 53.85±10.61                 | NS      |
| Gender, male n (%)             | 59 (63.45)                   | 77 (68.91)                  | NS      |
| PLT (10^3/µl)                  | 194.34±46.81                 | 197.14±51.56                | NS      |
| BUN (mg/dl)                    | 15.61±7.18                   | 16.71±9.62                  | NS      |
| Child-Pugh classification, n (%)|                               |                             | NS      |
| A                              | 35 (37.45)                   | 47 (42.34)                  |         |
| B                              | 52 (56.12)                   | 50 (44.51)                  |         |
| C                              | 6 (5.43)                     | 15 (13.15)                  |         |
| Hepatitis B, n (%)             | 76 (82.13)                   | 74.78                       |         |
| AFP (ng/ml)                    | 1,965.98±567.12              | 1,876.33±789.10             | NS      |
| Tumor size, n (%)              |                               |                             | NS      |
| ≤5 cm                          | 32 (34.12)                   | 30 (26.61)                  |         |
| >5 cm                          | 61 (65.88)                   | 82 (73.21)                  |         |
| BCLC stage, n (%)              |                               |                             | NS      |
| 0-B                            | 81 (87.12)                   | 93 (83.17)                  |         |
| C-D                            | 12 (12.88)                   | 19 (16.83)                  |         |
| Histological grading, n (%)    |                               |                             | NS      |
| Well                           | 51 (55.54)                   | 54 (48.13)                  |         |
| Moderate                       | 24 (25.67)                   | 36 (32.47)                  |         |
| Poor                           | 18 (18.89)                   | 32 (20.40)                  |         |
| MVI, n (%)                     | 39 (41.87)                   | 51 (45.38)                  | NS      |
| Portal vein thrombi, n (%)     | 0 (0.00)                     | 0 (0.00)                    | NS      |
| Ascites, n                     | 12.67                        | 16.83                       |         |
| Objective therapeutic response, n (%)|                       |                             | 0.024a |
| CR and PR                      | 49 (52.69)                   | 21 (18.75)                  |         |
| SD and PD                      | 44 (47.31)                   | 91 (81.25)                  |         |

Chi-square test or t-test were used for statistical analysis. *P<0.05, indicates statistical significance. HCC, hepatocellular carcinoma; ASPP2, apoptosis-stimulating p53 protein 2; PLT, platelet level; BUN, blood urea nitrogen; AFP, α-fetoprotein; BCLC, Barcelona Clinic Liver Cancer; MVI, microvessel invasion; CR, complete response; PR, partial response; SD, stable disease; PD, progressive disease; NS, not significant.
predicting factors for the 1-year death risk, after adjustment and exclusion of influence from other confounding factors, including tumor size (OR=1.54; 95% CI 0.87-1.98; P=0.078), age (OR=1.23; 95% CI 0.65-1.78; P=0.21), BCLC stage (OR=1.98; 95% CI 0.91-2.80; P=0.067), MVI (OR=1.58; 95% CI 0.96-2.77; P=0.058) and AFP levels (OR=1.21; 95% CI 0.76-1.88; P=0.31) (Table IV). Survival analysis revealed that, among the 232 initially unresectable patients, overall survival and progression-free survival were improved in patients exhibiting a higher ASPP2 expression (dotted line) compared to those with a low expression group (solid line) (Fig. 3A and B). Intriguingly, among patients who underwent direct resection, TACE only improved the overall survival and progression-free survival of those resectable patients exhibiting high ASPP2 expression (Fig. 3C and D). These data revealed that ASPP2 expression may be a predictor of the prognosis for TACE.
therapy followed by liver resection, regardless of the anatomic condition at initiation.

**Discussion**

Transcatheter arterial chemoembolization (TACE) is considered as a first-line therapeutic strategy for intermediate stage hepatocellular carcinoma (HCC) in the EASL-EORTC clinical practice guideline (24). For HCC patients considered unresectable at initiation, TACE is performed to downstage HCC prior to surgical resection (1,2). Whether TACE is indicated for resectable HCC patient, as well as whether such patients would benefit from TACE remains a controversial topic. A meta-analysis based on HCC patients with macrovascular invasion reported that administration of TACE was associated with a lower 1-year and 3-year survival rate compared to resection (25). These data demonstrated that refractoriness of TACE may have an adverse influence on

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**Figure 2.** Autophagy-induced anti-apoptotic effect in primary HCC cells in vitro following hypoxia cultivation or administration of chemotherapy. (A) Expression of LC3-II and (B) ratio of LC3-II to LC3-I in para-carcinoma and carcinoma tissues of HCC patients following TACE. n=5, \( ^*P<0.01 \). (C) 5-FU treatment or hypoxia stimulation activated autophagy in the HCC primary cells of patients without an objective therapeutic response following TACE. n=5, \( ^*P<0.01 \). (D) Inhibitor of autophagy, HCQ (hydroxychloroquine), promoted the expression of cleaved caspase-3 and cleaved caspase-8 in the HCC primary cells of patients without an objective therapeutic response following TACE measured by MTT assay. n=5, \( ^*P<0.05 \). (E) Inhibitor of autophagy, HCQ (hydroxychloroquine), repressed proliferation of HCC primary cells of patients without objective therapeutic response following TACE measured by MTT assay. n=5, \( ^*P<0.05 \). All experiments were repeated at least thrice. Student’s t-test was used for statistical analysis. P<0.05 indicates statistical difference. TACE, transcatheter arterial chemoembolization; HCC, hepatocellular carcinoma.
prognosis and that TACE should be administrated only to those patients conforming to selective criteria. For recurrent but resectable HCC, resection with preoperative TACE may not improve prognosis compared to single resection. It has been confirmed that TACE may improve the prognosis for unresectable patients compared to supportive therapy, and may lead to the downstaging of HCC in some patients who are considered unresectable at initiation (8). In those patients for whom HCC was not down-staged following TACE, disease-free survival was significantly worse than that of patients with a remarkable response to TACE (8).

Furthermore, in spite of the known efficacy and a level of safety associated with it, administration of TACE in intermediate HCC patients is beset with several concerns and limitations (1,26,27). The most important of these is an incomplete and weak response to TACE due to insufficient arterial supply or large tumor size (>5 cm) (26). The second issue is related to a hypoxic microenvironment, which inevitably results from arterial embolization, that promotes angiogenic factors such as the vascular endothelial growth factor (VEGF) (26,27). The surge in serum VEGF levels following TACE was found to be correlated with tumor size, vascular invasion and patient survival. Tolerance of hypo-perfusion of chemotherapy drugs in a hypoxia environment in HCC patients with larger tumors may increase their malignant biological behavior and induce therapeutic resistance to TACE. It has been reported that the expression of hypoxia-induced factor 1 (HIF-1) and VEGF was increased following TACE (28,29). Increased expression of some stemness markers was also found in HCC tissues following TACE. These compensatory changes in biological behavior may reduce sensitivity to TACE and aggravate resistance to regional chemotherapy. In addition, TACE may eventually result in the deterioration of liver function, damage to the hepatic artery, an increased post-embolization liver failure risk and, most importantly, a poor quality of life, leading to a poor prognosis in those patients who do not respond to TACE and do not exhibit tumor downstaging (30). As shown in many previous investigations, the clinical benefits are correlated with the presence of objective response and are evident only in patients with Child-Pugh B, European Cooperative Oncology Group (ECOG) status 1, who did not present with severe liver cirrhosis (31). Therefore, whether patients benefit from liver resection following preoperative TACE may be dependent on fundamental liver function and inhibition of compensatory mechanisms underlying resistance to chemotherapy and ischemia.

In solid tumors, autophagy functions as a survival mechanism against various stressors including metabolic stress, starvation, hypoxia, chemotherapy, and radiotherapy (12-15). In physiological situations, autophagy plays an important role in organelle turnover, protein degradation, cellular differentiation and aging. Under stress, autophagy protects cells by eliminating damaged organelles and proteins via autophagosomes. Under mild and physiological hypoxia (0.19-3% O2), autophagic response is dependent on HIF-1, which has an anti-apoptotic effect on cancer cells (32,33). Autophagy may promote cell survival. In HCC, under nutrition derivation Beclin1-induced autophagy, chemotherapy resistance in pancreatic cancer cells was increased as well (34). Moreover, Beclin1-induced autophagy mediated cell survival in leukemia cells (35). Previous research indicated that activation of autophagy was correlated with cell survival as well as resistance to adverse conditions in cancer cells (36). In addition, according to a previous study, Beclin-1, considered as a molecular autophagy marker, may be downregulated by apoptosis-stimulating p53 protein 2 (ASPP2), which was considered as a transcriptional cofactor of p53 and can act as a tumor suppressor. Therefore, down-regulation of ASPP may protect cancer cells from autophagy instead of conventional p53-related signaling (16,17). Chen et al (37) found that mechanically, ASPP2 binds to BECN1, leading to a decrease in binding of PIK3C3 and the UV radiation resistance-associated gene (UVRAG), and an increase in the binding of Rubicon in the PIK3C3 complex. The present study demonstrated that down-regulation of ASPP2 enhanced pro-survival, chemoresistant properties of HCC cells via autophagy, both in vitro and in vivo. In HCC, administration of TACE led to local ischemia of carcinoma tissues and the regional release of chemotherapy. Therefore, we speculated that autophagy activation in carcinoma may promote resistance to chemotherapy and hypoxia environments following TACE, leading to an incomplete

| Variable                          | P-value | OR   | 95% CI      |
|-----------------------------------|---------|------|-------------|
| PVTT (no vs. yes)                 | 0.020*  | 2.17 | 1.34-3.61   |
| Expression of ASPP2 (high vs. low) | 0.004*  | 2.12 | 1.68-3.75   |
| Child-Pugh classification (classification A=1, classification B=2, classification C=3) | 0.031*  | 2.76 | 1.68-3.75   |
| Tumor size (≤5 cm vs. >5 cm)      | 0.078   | 1.54 | 0.87-1.98   |
| AFP levels (≤1,500 ng/ml vs. >1,500 ng/ml) | 0.31    | 1.21 | 0.76-1.88   |
| Age (≤50 years vs. >50 years)     | 0.21    | 1.23 | 0.65-1.78   |
| BCLC stage (0-B vs. C-D)          | 0.067   | 1.98 | 0.91-2.80   |
| MVI (no vs. yes)                  | 0.058   | 1.58 | 0.96-2.77   |

Multiple logistics regression analysis was used for statistical analysis and *P<0.05, indicates statistical significance. OR, odds ratio; CI, confidence interval; PVTT, portal vein tumor thrombus; ASPP2, apoptosis-stimulating p53 protein 2; AFP, α-fetoprotein; BCLC, Barcelona Clinic Liver Cancer; MVI, microvessel invasion.
response to TACE. We found that inhibition of autophagy in primary HCC cells of a patient exhibiting TACE failure in vitro, promoted apoptosis and attenuated chemotherapy resistance. Our results demonstrated that activation of autophagy in HCC cells under stress, protected cancer cells from apoptosis. It was also observed that decreased ASPP2 expression in carcinoma tissues of HCC samples in patients following TACE, was associated with increased Beclin-1 expression.
These findings derived from basic research suggest that ASPP2 expression may act as a predictor of prognosis following TACE. We further measured ASPP2 expression in HCC tissues from HCC patients following TACE, where a high ASPP2 expression was associated with a higher objective therapeutic response rate, indicating a better prognosis for unresectable HCC patients. Furthermore, administration of neoadjuvant TACE improved survival only in resectable patients with a high ASPP2 expression. Clinical observation data combined with the above stated findings from basic experiments demonstrated that patients with a high ASPP2 expression following TACE show a higher response rate than those with a low expression, which may be associated with increased sensitivity to TACE after inhibition of autophagy via downstream tumor-suppressor p53 or ASPP2/Beclin1 signaling pathways.

In conclusion, our study provides a novel biomarker for predicting prognosis and proposes a potential approach for choosing suitable patients who may benefit from TACE combined with resection.

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Availability of data and materials
The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors’ contributions
JM and ZT conceived the study and designed the experiments. XP and FM contributed to the data collection; performed the data analysis and interpreted the results. JM wrote the manuscript; JM and ZT contributed to the critical revision of the article. All authors read and approved the final manuscript and agree to be accountable for all aspects of the research in ensuring that the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Ethics approval and consent to participate
This observational study was approved by the Medical Ethics Committee of The People’s Hospital of Danyang (Danyang, Jiangsu). Written consent for collection of tissue samples was acquired from all patients prior to surgery.

Patient consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

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