Cavidine protects against asthma in neonatal asthmatic mice model by attenuating PI3Ks/NF-κB signaling pathway

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Abstract

Purpose: To evaluate the protective effect of cavidine against asthma in neonatal mice.

Methods: Neonatal mice were treated with cavidine at doses of 5 and 10 mg/kg, po, 2 h prior to asthma induction with ovalbumin (OVA) on the 1st and 14th days of the treatment protocol. The anti-asthma activity of cavidine was evaluated by determining the number of inflammatory cells and cytokine levels in broncho-alveolar lavage fluid (BALF) and OVA-specific IgE and TGF-β1 in the serum of OVA-sensitized mice. The levels of NF-κB and PI3K protein expression were determined in the lung tissues of OVA-sensitized mice.

Results: Cavidine attenuated the number of inflammatory cells and cytokines in BALF of OVA-sensitized mice. The levels of OVA-specific IgE and TGF-β1 decreased significantly in cavidine-treated groups, when compared to asthmatic group of mice, while NF-κB was significantly downregulated (p < 0.01). The altered expression of PI3K signaling protein was attenuated in the lung tissues of cavidine-treated mice sensitized with OVA.

Conclusion: These results reveal that the anti-asthma effect of cavidine in OVA-induced asthmatic neonatal mice occurs via reduction of inflammation and immune responsive cells linked to PI3Ks/NF-κB signaling pathway in lung tissues. These findings suggest that cavidine may be clinically suitable for the management of asthma.

Keywords: Cavidine, Asthma, Inflammation, Immune cell, Neonatal, PI3K signaling

INTRODUCTION

Asthma is an inflammatory lung disorder characterized by infiltration of neutrophils, lymphocytes and macrophages in the lung tissues [1]. The pathogenesis of asthma involves increased secretion of mucus, hyper-responsiveness and remodeling of the airways [2]. Cytokines are responsible for remodeling and inflammation of airways which is activated by T lymphocytes [3]. Cytokines trigger the release of immunoglobulin E, leading to the release of inflammatory mediators by mast cells [4]. Thus, the determination of cytokine levels is an important step for assessing asthma. The enzyme PI3K controls cell signaling, proliferation and differentiation of cells [5]. Moreover, Akt protein is activated by PI3K which acts on mammalian targets of rapamycin and glycogen synthase kinase 3b [6]. In T cells, activation of
PI3K results in progression, survival and differentiation of cells [7]. Several natural compounds have shown promising potential in the management of asthma and other chronic disorders. Cavidine is an alkaloid isolated from Corydalis impatiens [8]. Corydalis impatiens is a Chinese herb traditionally used for the management of cholecystitis, hepatitis and skin injuries [9]. Studies have revealed that cavidine has promising effects against gastric ulcer, colitis and inflammation [10,11]. These effects of cavidine depend on its antioxidant and anti-inflammatory properties [12]. Cavidine shows anti-inflammatory activity by attenuating the NF-κB signaling pathway and pro inflammatory cytokines [11]. The present study was aimed at investigating the protective effect of cavidine against asthma in OVA-sensitized mice model.

**EXPERIMENTAL**

**Animals**

BALB/c mice were purchased from Dashuo Laboratory Animal Reproduction Center, China. They were maintained under standard conditions (12-h day/12-h night light cycle, 60 – 80 % humidity and temperature of 26 ±1°C as per the guidelines of the Association for the Assessment and Accreditation of Laboratory Animal Care International (AAALAC) [13]. The pups were monitored carefully from the day of litter (day 1 (P1). All the protocols used were approved by Institutional Animal Care and Use Committee of Medical College of Yan’an University, China (no. IACUC/YU/2017/03).

**Induction of asthma**

Mice at 12 days of age (P12) were used for the induction of asthma and were divided into five different groups: normal group, asthmatic group, cavidine (5 and 10 mg/kg) groups, and STD group. The mice were sensitized under standard conditions (12-h day/12-h night light cycle, 60 – 80 % humidity and temperature of 26 ±1°C as per the guidelines of the Association for the Assessment and Accreditation of Laboratory Animal Care International (AAALAC) [13]). The pups were monitored carefully from the day of litter (day 1 (P1). All the protocols used were approved by Institutional Animal Care and Use Committee of Medical College of Yan’an University, China (no. IACUC/YU/2017/03).

**Assessment of serum and broncho-alveolar lavage fluid (BALF)**

Tracheotomy was performed at the end of treatment protocol with cervical dislocation, and BALF was collected from the isolated lung. A hemocytometer was used to determine the quantity of inflammatory cells in the BALF, while Wright’s staining was used to estimate differential cell count. The serum levels of OVA-specific IgE and TGF-β 1 in BALF were assayed with ELISA kits as per the manufacturer’s instructions.

**Assessment of inflammatory cytokines in BALF**

ELISA kits were used for the assessment of BALF contents of cytokines such as IL-17, IL-13, IL-6, IL-4 and IFN-y as per the instruction of the kit manufacturer.

**Histopathological studies**

Lung tissues were isolated from each animal and centrifuged at 2000 rpm for 10 min. ELISA kits were used to determine the levels of p-IκBα, IκBα, p-NF-κB p65 and NF-κB p65 in the supernatant solution as per the manufacturer’s instruction.

**Western blot assay**

Lysis buffer containing protease inhibitor was incubated with isolated CD4+ T cells, and protein assay kit was used to determine the concentration of protein in the lysate. The isolated protein was subjected to SDS-polyacrylamide gel electrophoresis, and the fractions were transferred to nitrocellulose membranes and incubated at 4°C overnight with primary antibodies for Notch-1, PTEN mTORc1, p-Akt, Akt and β-actin. Thereafter, peroxidase-conjugated secondary antibodies were incubated with isolated proteins for 1 h at room temperature, and the densities of the bands were determined using chemiluminescence.

**Statistical analysis**

Data are expressed as mean ± SEM (n = 10). Statistical analysis was performed using one-way ANOVA. Post-hoc comparison of means was
carried out by Dunnett’s post hoc test for multiple comparisons. The level of statistical significance was set at $p < 0.05$.

**RESULTS**

**Cavidine attenuated the altered levels of inflammatory cells**

Figure 1 shows the effect of cavidine on cellular inflammation in OVA-sensitized mice. There was an increase in inflammatory cells such as neutrophils, macrophages, lymphocytes and eosinophils in the BALF of the asthmatic group, when compared with normal group of mice. However, treatment with cavidine decreased the number of neutrophils ($1.82 \times 10^5$), lymphocytes ($3.91 \times 10^5$), eosinophils ($1.84 \times 10^5$) and macrophages ($7.48 \times 10^5$) in BALF, relative to untreated asthmatic group of mice.

**Figure 1:** Effect of cavidine on inflammatory cells in OVA-sensitized mice. Data are mean ± SEM (n = 10); $**p < 0.01$ vs normal group; *$p < 0.05$, **$p < 0.01$ vs. asthmatic group

**Cavidine attenuated altered levels of OVA-specific IgE and TGF-β1**

The effect of cavidine was observed on the OVA-specific IgE and TGF-β1 in OVA-sensitized mice. There were increases in the number of OVA-specific IgE up to 157 ng/ml, and TGF-β1 up to 310 pg/ml in the asthmatic group. However, cavidine significantly ($p < 0.01$) decreased OVA-specific IgE (83 ng/mL) and TGF-β1 (127 pg/mL), when compared with the asthmatic group (Figure 2).

**Figure 2:** Cavidine attenuated OVA-specific IgE and TGF-β1 in OVA-sensitized mice. Data are mean ± SEM (n = 10); $##p < 0.01$ vs normal group; *$p < 0.05$, **$p < 0.01$ vs. asthmatic group

**Cavidine attenuated inflammatory scores in OVA-sensitized mice**

Lung architecture was assessed histopathological using H&E staining. As depicted in Figure 4, the peribronchial and perivascular tissues of the OVA-challenged group contained inflammatory cells. However, the cavidine group showed reduction in the inflammatory cells in lung tissues. Moreover, inflammatory score was higher in the lung tissues of the asthmatic group than in normal group. However, treatment with cavidine and Dex significantly ($p < 0.01$) reduced the inflammatory score in in the lung tissues, relative to that in asthmatic mice.

**Cavidine attenuated NF-κB activity in OVA-sensitized mice**

Figure 5 shows that the levels of p-NF-κB p65, NF-κB p65 and p-IκBα increased significantly in tissue homogenates of the asthmatic group, relative to the normal group. However, levels of p-IκBα, p-NF-κB p65 and NF-κB p65 proteins were significantly and dose-dependently

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downregulated in the cavidine groups, when compared to the asthmatic group ($p < 0.01$).

Figure 4: Effect of cavidine on the histopathology of lung of OVA-sensitized mice. I: Histopathology of lung (A: normal; B: asthmatic; C: cavidine 5 mg/kg; D: cavidine 10 mg/kg; E: STD); II: Inflammatory score. Data are mean ± SEM ($n = 10$); **$p < 0.01$ vs normal group; *$p < 0.05$, **$p < 0.01$ vs asthmatic group

Cavidine attenuated the expressions of PTEN mTORc1, Akt and p-Akt proteins

The effect of cavidine on the expressions of PTEN, mTORc1, Akt and p-Akt proteins in tissue homogenates of OVA-sensitized mice is shown in Figure 6. There was enhancement in the relative expression of mTORc1, p-Akt and Akt proteins, and reduction in the expression of PTEN protein in the tissue homogenate of the asthmatic group, relative to the normal group. However, treatment with cavidine ameliorated the altered expressions of PTEN, mTORc1, Akt and p-Akt proteins in tissue homogenates of OVA-sensitized mice.

Figure 6: Effect of cavidine on the expressions of PTEN, mTORc1, Akt and p-Akt proteins in tissue homogenate of OVA-sensitized mice. Data are mean ± SEM ($n=10$); **$p < 0.01$ vs. normal group; *$p < 0.05$, **$p < 0.01$ vs. asthmatic group

DISCUSSION

The prevalence of asthma has increased in the recent years. Anti-inflammatory therapies such as steroids are often used in the management of asthma but many limitations are associated with it. Thus, there is a need for alternative medicine for the management of asthma. The present investigation evaluated the anti-asthmatic effect of cavidine in neonatal mice through determination of the number of inflammatory cells and cytokine levels in BALF, and OVA-specific IgE, and TGF-β in the serum of OVA-sensitized mice. Moreover, histopathology, NF-κB level, and expression of PI3K signaling protein were determined in the lung tissues of the OVA-sensitized mice.

The development of asthma is marked by infiltration of inflammatory cells in lung tissue [14]. Thus, drugs used for the management of asthma function by reducing the number of inflammatory cells [15]. The results of the present investigation showed that cavidine treatment attenuated the number of inflammatory cells in
OVA-sensitized mice. Cytokines also play an important role in the pathogenesis of asthma [16]. This study has shown that treatment with cavidine significantly ameliorated the cytokine levels in BALF of OVA-sensitized mice.

The proliferation and initiation of CD4+ T cells occur due to PI3K signaling which is regulated by several proteins such as PTEN, mTORc1, p-Akt and Akt [17]. The proliferation of CD4+ T cells contributes to the development of asthma [18]. Cavidine attenuated the expressions of PTEN, mTORc1, p-Akt and Akt protein in the lung tissues of OVA-sensitized mice. The NF-κB signaling pathway contributes to the pathogenesis of asthma by regulating immunity and inflammation. Studies have shown that the expression of NF-κB is increased in the lung tissues of asthmatic patients [19]. The data obtained in the investigation reveals that cavidine significantly reduced the expression of NF-κB in OVA-sensitized mice, relative to untreated asthmatic mice.

CONCLUSION

These results reveal that the anti-asthma activity of cavidine in OVA-induced asthma in neonatal mice occurs through attenuation of lung tissue inflammation and immune responsive cells. Thus, cavidine may be clinically effective in the management of asthma.

DECLARATIONS

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Conflict of interest

No conflict of interest is associated with this work.

Contribution of authors

We declare that this work was done by the authors named in this article and all liabilities pertaining to claims relating to the content of this article will be borne by the authors. Qin Hao performed the experimental work and literature survey. Juan Shen had done histopathology and statistical analysis and Lin Zhao supervised and wrote the manuscript.

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