Rational design and latest advances of codelivery systems for cancer therapy

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ABSTRACT

Current treatments have limited effectiveness in treating tumors. The combination of multiple drugs or treatment strategies is widely studied to improve therapeutic effect and reduce adverse effects of cancer therapy. The codelivery system is the key to realize combined therapies. It is necessary to design and construct different codelivery systems in accordance with the variable structures and properties of cargoes and vectors. This review presented the typical design considerations about codelivery vectors for cancer therapy and described the current state of codelivery systems from two aspects: different types of vectors and collaborative treatment strategies. The commonly used loading methods of cargoes into the vectors, including physical and chemical processes, are discussed in detail. Finally, we outline the challenges and perspectives about the improvement of codelivery systems.

1. Introduction

Cancer is one of the deadliest threats to global human health [1]. Cancer cells are very crafty and possess a variety of defenses against treatments. As a result, cancer is difficult to be cured, which leads to a high fatality rate. Research into the prevention and treatment of cancer is in full swing. The commonly used clinical method of cancer treatments include surgery, chemotherapy, and radiotherapy. Chemotherapy, as a systemic treatment, has a therapeutic effect on some tumors that tend to spread throughout body or have metastasized to advanced tumors [2]. Yet chemotherapy still has some hurdles to overcome. (i) Chemotherapy would cause systemic toxicity, which is often accompanied by obvious side effects such as hair loss, diarrhea, and liver/kidney damage. (ii) Most chemotherapeutic drugs are poorly soluble in aqueous solution and unstable during circulation. (iii) The tumor targeting and cellular uptake of chemotherapeutic drugs need to be improved. (iv) Multidrug resistance (MDR) is an issue that needs to be seriously considered after long-term administration [3]. Clinically, multiple chemotherapeutic drugs are administered in sequence to improve the efficacy of chemotherapy and delay MDR. The collaborative treatment strategy of paclitaxel (PTX) and cisplatin (DDP) is a first-line treatment for lung cancer. Docetaxel (TXT) combined with adriamycin (AMD) in the treatment of breast cancer is an effective and safe neoadjuvant chemotherapy. Various drug delivery systems are also proposed to extend the half-life of drugs, increase drug targeting, improve drug bioavailability, and reduce adverse reactions to chemotherapy drugs [4]. Over the past decade, plenty of effective drug delivery vectors, such as liposomes [5], polymersomes [6,7], nanoparticles [8–10], micelles [11–13], exosomes [14,15], hydrogels [16,17], and drug-polymer conjugates [18,19], have been developed. To further improve the delivery efficiency, delivery vector functionalization is the focus at present [20].

The combination of different therapeutic strategies exhibits ideal therapeutic effects on cancer, meanwhile higher requirements are put forward for the rational design of delivery systems. Vectors that deliver only one type of drug are no longer sufficient for cancer therapy. To realize combination therapy similar to clinical treatments, vectors delivering two or more kinds of reagents, which termed as codelivery vectors, were proposed. For instance, doxorubicin (DOX) and PTX are delivered simultaneously by nanoparticles or micelles for cancer therapy [21,22]. The combination of vincristine sulfate and verapamil hydrochloride by poly(ε-caprolactone-glycolide acid) nanoparticles increases the sensitivity of tumor cells to chemotherapy drugs [23]. Codelivery vectors can combine gene therapy, phototherapy, or immunotherapy by loading different reagents such as nucleic acid, photosensitive molecules, and quantum dots [24,25]. For example, Zhu et al. [26] combined camptothecin (CPT) and upconversion nanoparticles (NaYF4:Yb/Er) into a
hybrid vesicle for synergistic chemotherapy and photodynamic therapy (PDT).

To build effective codelivery systems, the interaction of each component, the balance of underlying conflicting factors, and the synergistic treatment strategies should be considered overall. Besides, in accordance with the typical cancer drug-delivery process of an intravenously administered nanomedicine, which including circulation, accumulation, penetration, internalization, and drug release, it is important to choose the type of vectors and the approach of drug loading in a proper way [27]. All these factors come together to guarantee high overall therapeutic efficiency. Similar to the barrel effect, any weak factor will reduce the performance of the codelivery systems. Although some codelivery systems have been reviewed, these reviews mainly focus on the properties and applications of nanocarriers [28–30]. Few reviews on the structure of codelivery vectors and the loading methods of cargoes have been reported to date. In this review, the design strategies and considerations of codelivery systems are discussed in detail. Then we highlight the latest advances of codelivery systems for cancer therapy. Typical examples about synergistic therapy are also illustrated.

2. Design considerations

The design of codelivery systems requires an overall consideration of the interaction between vectors and cargoes, as well as the synergistic action between the loaded reagents. Plenty of vectors, such as micelles, organic/inorganic nanoparticles, and microgels, have been successfully used for codelivery systems [31–33]. The different structures of drugs and vectors bring a variety of strategies to achieve the purpose of codelivering different drugs. Though all roads lead to Rome, the optimal design route needs to be explored, and some general rules need to be summarized. Although nanocarriers have some advantages owing to their nanoscale, such as the enhanced permeability and retention effect, further modification of the vectors is necessary to enhance the diagnostic and therapeutic effects. For example, targeting molecules are conjugated to the surface of the vectors to enhance the targeting of the vectors to the lesion site [34]; the vectors can be loaded with imaging agents to realize the integration of diagnosis and treatment [35]. As the functionalization of the vectors has been summarized in several excellent reviews in recent years [28,29,36], they will only be briefly described here to cover the subject. In this section, we will mainly focus on the loading methods of drug molecules and collaborative strategies of codelivery systems.

2.1. Loading methods

Loading methods can be categorized into physical and chemical processes. The commonly used physical interactions include electrostatic interaction, hydrophobic interaction, and simple encapsulation. Physical process operations are generally straightforward. Simple stirring for a certain period may accomplish the packaging process. However, owing to the unstable encapsulation, premature leakage may occur during the circulation. Chemical processes usually involve multiple chemical reactions and the necessary refinements. Stimuli-responsive bonds or structures should be introduced between the drug molecule and backbone to achieve the controlled release of intact drug molecules. Fig. 1 illustrates the typical loading methods of physical and chemical processes.

2.1.1. Physical processes

The top three commonly used physical processes are hydrophobic interaction, electrostatic interaction, and simple physical encapsulation. The hydrophobic action is a phenomenon that hydrophobic molecules or segments aggregate to avoid water. Monolayer liposomes and micelles have hydrophobic internal cavities that can hold hydrophobic drugs [37, 38]. The bilayer liposomes can be loaded with hydrophobic drugs between the bilayer, and the inner cavity can be loaded with water-soluble drugs. The hydrophilic shell of the vectors increases the biocompatibility and circulation time of the delivery system [39]. Most chemotherapeutic drugs are poorly water-soluble. The first formulation, liposomal DOX,
was used clinically in 1995. Since then, many liposomes and lipid-based products have been placed on the market or undergo clinical trials [40]. Liposomes can be designed flexibly [41,42]. As shown in Fig. 2A, Lee et al. [43] prepared a polymer-caged nanobin which comprises a doxorubicin-encapsulated liposomal core protected by a pH-responsive cisplatin prodrug-loaded polymer shell.

In 1984, polymer micelles were first proposed by Bader and co-workers as drug vectors [44]. Nowadays, different kinds of micelles have been developed as effective vectors with good biodegradability, effective encapsulation, and long circulation in the blood [45–47]. Fig. 2B shows a typical micelle that codeliver DOX and PTX by hydrophobic interaction [48]. The solubility and bioavailability of hydrophobic drugs are improved by this delivery system. Dendrimers possess well-defined, highly branched, and spherical three-dimensional structures [49]. Their hydrophobic interior cavities and large number of reactive end groups enable dendrimers to be a candidate for drug/gene delivery vectors [50–53]. As shown in Fig. 2C, the inner cavity or periphery of the dendrimer can be a suitable host for the guest molecules. Taratula et al. [54] synthesized multifunctional theranostic platforms based on phthalocyanine-loaded dendrimer for image-guided drug delivery and PDT because hydrophobic interactions are non-specific. Unwanted aggregation of hydrophobic molecules may occur during the preparation process, which will reduce the drug loading efficiency and increase the heterogeneity of the formulation [55].

Electrostatic interaction is an equilibrium achieved through the mutual attraction of positive and negative charges. Some drug molecules are electrically charged under physiological conditions. For example, DOX is positively charged under physiological conditions owing to the ionization of carboxyl groups [56]. Methotrexate (MTX) is negatively charged in solution because of the ionization of carboxyl groups [57]. Most protein-based reagents are negatively charged in a neutral environment [58]. Some materials also exhibit electrical properties in the physiological environment and can be applied to prepare carriers. Chitosan is electropositive due to the large number of amino groups, and hyaluronic acid is electronegative due to the carboxyl groups. They are widely used in the preparation of biomedical materials [59,60]. Therefore, electrostatic interaction is an alternative intermolecular force that can be used to bind up drug molecules. For instance, as shown in Fig. 3A, the negatively charged new indocyanine green (ICG) (IR820) molecules were trapped on the surface of the cationic micelle [24]. Polyamidoamine dendrimers with positive charges were used to increase the solubility of ketoprofen which is a non-steroidal anti-inflammatory drug [61]. For immunotherapy, the use of electrostatic interaction to deliver antigens is an effective method [58]. In fact, the most widely used area of electrostatic interaction is the preparation of non-viral gene delivery systems [62,63]. The nucleic acids present electronegative due to the dissociation of hydrogen ions of phosphate groups. So nucleic acids can be condensed by positively charged polymer chains or adsorbed on the surface of positively charged nanoparticles. As shown in Fig. 3B, a series of cationic gene carriers were prepared with high transmission efficiency [62–66].

Drug molecules can also be simply encapsulated in the cavity of the carriers, just like putting the goods into the cardboard box, sealing the box and sending it to the receiving address. Mesoporous silica nanoparticles (MSNs), carbon nanotubes, and other nanoparticles with the suitable pore structures can be used to transport molecules in this way [67,68]. The key to this method is to select suitable lids to plug the cavity. It is necessary to consider a suitable occlusion strategy to prevent the leakage of drugs during the circulation in vivo. As shown in Fig. 4A, core-shell-shellac up-conversion (UC) nanoparticles were designed for simultaneous photo-responsive drug release, PDT, and cell imaging [69]. The nanoparticle consists of an UC core, a silica intermediate shell filled with a photosensitizer, and a cyclodextrin (β-CD)–gated mesoporous silica outmost shell. Similarly, Liu et al. [70] designed hollow silica mesoporous nanoparticles that facilitate drug delivery by cascading pH stimulation in the tumor microenvironment. The hollow silicon mesoporous nanoparticles are loaded with DOX inside and blocked with PEG-modified β-CD outside as lids. Tactfully, Zeng et al. [71] designed a drug-gated mesoporous antitumor nanoplatfrom based on pH-sensitive dynamic covalent bonds as shown in Fig. 4B. The doors are blocked by the drug itself through a pH-sensitive dynamic benzoate-imine covalent bond. This strategy cleverly bypasses the use of auxiliary capping agents.

Other physical interactions, such as the hydrogen-bonding [72,73], coordination interaction [74], and π–π stacking [75], are also applied to the preparation of codelivery systems. Because these interactions are directional, special requirements for molecular structure are proposed. For instance, as shown in Fig. 5A, macromolecular assembly with 5-fluorouracil (5-FU) or cyanuric acid (CA) requires specific hydrogen-bonding recognition between 5-FU/CA and polymer-displayed melamine [76]. The coordination interaction is molecular forces between electron donors and acceptors, including z-donor/organometallics or Lewis bases/acidcs [77]. The bond energy of coordination bonds is generally weaker than covalent bonds but stronger than hydrophobic interactions [78]. Lv et al. [79] prepared a series of micelles with high drug loading capacity by

Fig. 2. (A) The liposomes with DOX-encapsulated core and pH-responsive cisplatin prodrug-loaded polymer shell [43]. Copyright 2016 Elsevier B.V. (B) DOX and PTX were simultaneously encapsulated into the TAT/FOL modified micelles (TAT: TAT peptide; FOL: folate) [48]. Copyright 2015, Elsevier B.V. (C) Schematic diagram of drug loading in dendrimer cavities [53]. Copyright 2013, Elsevier Ltd. DOX, doxorubicin; PTX, paclitaxel.
Fig. 3. (A) The negatively charged new indocyanine green (IR820) molecules are attracted to the cationic micelle carrier for phototherapy and imaging [24]. Copyright 2019, American Chemical Society. (B) Typical cationic gene carriers for cancer therapy. (For interpretation of the references to colour in this figure legend, the reader is referred to the Web version of this article.)

Fig. 4. (A) Schematic illustration of a core-shell-shell structured nanoparticle which consists of an up-conversion (UC) core, a silica intermediate shell filled with a photosensitizer, and β-cyclodextrin (β-CD)-gated mesoporous silica outmost shell [69]. Copyright 2016, American Chemical Society. (B) Schematic illustration of the DOX-self-gated mesoporous silica nanomaterials with pH-responsive drug release property, and dynamically PEGylated and DOX self-gated mesoporous silica nanomaterials with site-specific drug release and cell uptake at weak acidic tumor tissue/cells [71]. Copyright 2017, WILEY-VCH.
introducing coordination interactions between electron acceptor-containing polymers and electron donor-containing chemotherapy drugs (Fig. 5B). \( \pi \)-\( \pi \) stacking is a special spatial arrangement of aromatic compounds, which refers to a weak interaction that often occurs between aromatic rings. Many chemotherapeutic drugs possess aromatic rings inside the molecules and can be loaded on/in the carrier by \( \pi \)-\( \pi \) stacking interaction [80,81]. As shown in Fig. 5C, Wei et al. [75] conjugated DOX to the amphiphilic polymer chains, and HCPT was loaded along with DOX via \( \pi \)-\( \pi \) stacking interaction.

2.1.2. Chemical processes

Chemical processes are performed by the reactions of complementary groups. The functional groups similar to the hydroxyl group (-OH), carboxyl group (-COOH), and amino group (-NH\(_2\)) could be used to graft drug molecules onto the vectors. It should be noted that, to achieve the controlled release of the carried reagents, cleavable bonds or structures...
need to be introduced between the drug molecule and the vector. Many indicators in tumor cells are different from normal cells, such as higher reactive oxygen species (ROS) concentration, higher glutathione (GSH) concentration, and relative lower pH [82,83]. The thioethal bond which response to ROS and the disulfide bond which response to GSH have already introduced into some factors for controlled release [84,85]. As shown in Fig. 6, Bai et al. [86] grafted CPT to β-CD with ROS-responsive linker and GSH-responsive linker. The drug release can be controlled by the break of these cleavable bonds. Similarly, Kong et al. [87] combined CPT and NIR fluorophores via disulfide bonds to monitor drug release in vivo. This strategy can use the tumor microenvironment to achieve GSH-responsive release of CPT. High drug loading capacity and precise structures of vectors can be obtained by chemical processes. But, generally, chemical processes are more complex than physical processes. Tedious experimental procedures and purification procedures are required. There is a trade-off between the method and performance.

2.2. Collaborative strategies

Physical and chemical processes have both strengths and weaknesses. It is necessary to select the appropriate mode or cooperative strategy based on the structure and property of both cargoes and carriers. The collaborative strategies could be a combination of two or more chemotherapeutic drugs, nucleic acids, proteins, and functional molecules. The synergistic action between the loaded reagents needs to be considered in advance. And no matter which combination, there must be no conflict between each component. Proper sequential administration of different drugs may lead to a synergistic effect. For example, the combined therapy of gemcitabine (GEM) and docetaxel (DOC) is order-dependent. The GEM administration, a large number of tumor cells enter the interphase. The break of these cleavable bonds. Similarly, Kong et al. [87] combined linker and GSH-responsive linker. The drug release can be controlled by the disintegration of the vectors and realize the following controlled release of the loaded drug molecules [89].

Some components of carriers have special functions. Superparamagnetic nanoparticles, such as Fe3O4 nanoparticles, can be used as contrast agents for magnetic resonance imaging (MRI) or as vectors for the targeted drug delivery [90,91]. Gold nanorods can be used as photothermal agents owing to the photothermal conversion ability. And gold nanoparticles (Au NPs) with high X-ray absorption coefficients are explored as contrast agents for computed tomography (CT) imaging [92]. Using these materials as the matrix, multifunctional vectors, which can be applied for imaging, photothermal therapy (PTT), and PDT, are available. For instance, the combination of gold nanocages with DOX and photosensitizer ICG exerted the simultaneous chemotherapeutic and photothermal/photodynamic treatment [93]. In general, simply loading multiple cargoes into a carrier is not the goal. The purpose is to amplify the therapeutic effect by collaborating among the components. Besides, the convenience and cost of carrier preparation are also factors to be considered. Table 1 summarized some typical examples of codelivery systems for cancer therapy.

3. Advances of codelivery systems

Many conventional vectors can be used to build codelivery systems by rational design. The synergistic treatment of two or more drugs could be realized by codelivery systems. This section introduces codelivery systems from two perspectives: different types of vectors and collaborative treatment strategies.

Table 1

| Vectors         | Drug combination                                  | Physical/chemical processes                     | Therapy model                           | References  |
|-----------------|---------------------------------------------------|-------------------------------------------------|-----------------------------------------|-------------|
| Inorganic       | DOX & Mir-31 (drug & nucleic acid)                 | Physical encapsulation, electrostatic interaction | Hela tumor-bearing BALB/c nude mice     | [94]        |
| nanoparticles   |                                                   |                                                  | Hepatoma-bearing BALB/c nude mice       |             |
| Inorganic       | Gold nanorod & SF & p53 (photothermal/imaging agent & drug & nucleic acid) | Physical encapsulation, electrostatic interaction | A549 tumor-bearing BALB/c nude mice     | [96]        |
| nanoparticles   | DOX & TRAIL (drug & cytokine)                     | π-π stacking interaction, chemical bonding       |                                         |             |
| Micelles        | CPT & TPP (drug & drug)                           | Chemical bonding                                 | 4T1 tumor-bearing BALB/c nude mice      | [97]        |
| Micelles        | CPT & IR820 (drug & phototherapy drug)            | Chemical bonding, electrostatic interaction      | 4T1 tumor-bearing BALB/c nude mice      | [24]        |
| Micelles        | PTX & GFP siRNA (drug & nucleic acid)              | Hydrophobic interaction, electrostatic interaction | MDA-MB-435-GFP cells                   | [98]        |
| Liposomes       | Ir & Cb (drug & drug)                             | Chemical bonding                                 | MCF-7 tumor-bearing nude mice           | [99]        |
| Hydrogels       | DOX & PTX (drug & drug)                           | Hydrophobic interaction                          | 4T1 tumor-bearing BALB/c nude mice      | [100]       |
| Micelles        | DOX & PTX (drug & drug)                           | Electrostatic interaction, chemical bonding      | B16F10 tumor-bearing mice               | [101]       |
| Organic         | Au NPs & MTX (photothermal/imaging agent & drug)  | Physical encapsulation, chemical bonding         | KB cells                                | [102]       |
| nanoparticles   | DOX & PTX (drug & drug)                           | Electrostatic interaction, hydrophobic interaction | A549 tumor-bearing BALB/c nude mice     | [33]        |
| Organic         | SN38&LND (drug & inhibitor)                       | Physical encapsulation                           | 4T1 tumor-bearing BALB/c nude mice      | [103]       |
| nanoparticles   | NY ESO-1 epitope & CpG (antigen & nucleic acid)    | Electrostatic interaction                        | HLA-A2 transgenic C57BL/6 mice         | [56]        |
| Organic         | DOX & Mir-21 (drug & nucleic acid)                 | Physical encapsulation, electrostatic interaction | LN229-luc tumor-bearing BALB/c nude mice| [104]       |
| nanoparticles   | AMD3100 & DNA (antagonist & nucleic acid)          | Chemical bonding, electrostatic interaction      | CXCR4 & U2OS cells                      | [105]       |
| Organic         | Phycocyanin & polypyrrole (photosensitizer & photothermal agent) | Physical encapsulation                          | MDA-MB-231 & HEK-293 cells             | [106]       |

Au NPs, gold nanoparticles; DOX, doxorubicin; PTX, paclitaxel; CPT, camptothecin; TPP, triphenylphosphonium bromide; LND, lonidamine.
3.1. Different types of vectors

3.1.1. Inorganic nanoparticles

The development of nanotechnology has promoted the production of various inorganic nanoparticles as drug vectors, such as silica nanoparticles [107], iron-based nanoparticles [108], Au NPs [92] and fullerenes [109]. Each nanoparticle has its unique characteristics, which makes the vector versatile.

MSNs are used in a variety of nanotechnology applications owing to its high surface area and pore volume [110,111]. The pore structure is orderly, the pore size distribution is single and adjustable, and the mesoporous shape is diverse. A large number of hydroxyl groups on the surface of MSNs can be chemically modified, which provides convenience for subsequent encapsulation of drugs or functionalization. As shown in Fig. 7, a codelivery system based on MSN was reported, where DOX molecules were loaded into the pore of MSNs by physical encapsulation, and microRNAs were conjugated onto the surface via disulfide bonds [94]. MSNs have good biocompatibility but no other special functions. The single function limits the development of MSNs as multifunctional vectors.

Compared with MSNs and organic nanoparticles, metal-based nanoparticles exhibit many special properties due to the metal bonding, small size effect, and surface effect. Iron oxide nanoparticles with a diameter of 50–100 nm have been already developed as contrast agents for MRI [112]. The ultra-small ultra-paramagnetic iron oxide particles with a diameter of about 10 nm can be used to achieve thermotherapy and active targeted treatment of cancer [113]. For example, as shown in Fig. 8, PTX and curcumin (Cur) were codelivered by pluronic-coated Fe3O4 nanoparticles [114]. Au NPs are widely used as drug and gene delivery vectors due to the good biocompatibility and versatility [115]. After modification, Au NPs can also codeliver a variety of reagents. Shiao et al. [116] designed a kind of aptamer-functionalized Au NPs to codeliver DOX and the photosensitizer 5,10,15,20-tetrakis(1-methylpyridinium-4-yl)porphyrin (TMPyP4) for improving the drug effectiveness. It should be noted that with the decrease of the particle size, some metal-based nanoparticles may interfere with the defense mechanism of antioxidants and thus exhibit higher cytotoxicity [117]. Using inorganic nanoparticles as codelivery carriers often require modification of the nanoparticles. Two or more interactions, whether physical or chemical, may be exploited to achieve the codelivery of reagents.

3.1.2. Micelles

Amphiphilic copolymers are capable of forming micelles by self-assembly in aqueous solutions. Hydrophilic segments, such as PEG, can extend the circulation time in vivo. And hydrophobic segments, such as poly (L-lysine), can provide space for hydrophobic drugs [33]. Micelles have been considered to be one of the potential drug delivery systems [118,119]. Many polymer micelles have been evaluated in vivo and clinical trials. Hydrophobic drugs usually loaded into the micelles via hydrophobic interactions. For instance, Lv et al. [33] designed a series of micelles to codeliver DOX and PTX, where DOX molecules were encapsulated by electrostatic interaction and PTX molecules were encapsulated hydrophobic interaction. Duong and Yung [48] packaged DOX and PTX into one micelle by hydrophobic interaction for synergistic codelivery. The loading capacity of drugs in conventional micelles is usually low, and therefore it is the pursuit to further improve the loading capacity of micelle-based delivery systems [120].

Some kinds of micelles are composed of polyprodrugs. One highlight of the polyprodrugs is the use of the hydrophobicity of the chemotherapeutic drugs to fabricate carriers. The loading capacity and the stability of the drugs would be increased [85,86,121]. For instance, the hydroxyl group on adjacent mitoxantrone (MTO) molecules can be linked by ROS-responsive cleavage linkers to form polyMTO [84]. The polyprodrugs similar to a cluster bomb, which will improve drug accumulation at the tumor site and the reduce side effects along with chemotherapy. As shown in Fig. 9, CPT and triphenylphosphonium bromide (TPP) molecules can be grafted onto the dextran backbone to achieve dual drug delivery. Huang et al. [99] connected hydrophobic anticancer drug chlorambucil (Cb) and hydrophilic anticancer drug irinotecan (Ir) through a hydrolyzable ester bond to make an amphiphilic prodrg, which then formed micelles. This nanoparticle is composed entirely of anticancer drugs. Two kinds of anticancer drugs were delivered together to achieve synergistic treatment and overcome the MDR of tumors. Cong et al. [122] designed a dual sensitive dual drug backboned shattering polymer (DDBSP) which composed of a PP2A inhibitor demethylcantharidin and cisplatin. DDBSP self-assembled micelle can be triggered intracellularly to break down in a chain-shattering manner to release the dual drugs payload (Fig. 10). Using the hydrophobic cavity inside the micelle that structured by polyprodrugs to carry drugs is an alternative way to design codelivery systems [123].

3.1.3. Liposomes

Liposomes are phospholipid vesicles formed by lipid bilayer membranes. Vesicles have isolated hydrophilic and lipophilic phase spaces. Hydrophilic drugs can be encapsulated in the inner aqueous phase, whereas hydrophobic drugs can be encapsulated in the lipid layer [124,125]. A number of liposome delivery systems are on the market such as conventional doxorubicin liposomes (e.g., Evacet, Myocet), long-cycle
doxorubicin liposomes (e.g., Doxil, caelyx), cytarabine liposomes (e.g., Depocyt), and paclitaxel liposomes (e.g., Taxo). Fig. 11 illustrates the structure of a typical liposome drug delivery system [126]. Liu et al. [100] designed a multilayer liposome vesicle that can improve the loading efficiency and sustained release of DOX and PTX, maximizing the combined therapeutic effect and minimizing the systemic toxicity. Liposomes can also be loaded with dyes or imaging agents. Sheng et al. [127] used nanoliposomes to carry perfluorooctyl bromide and ICG for enhanced multimodal imaging-guided phototherapy. This strategy combines CT contrast imaging, PDT, and PTT. Although many liposome drug systems have been proposed, the low drug loading capacity and poor stability hinder the large-scale application of liposomes.

3.1.4. Hydrogels

Owing to their adjustable chemical and physical properties, hydrogels have been vigorously developed as biomaterials [128–130]. Drugs or reagents can be encapsulated directly in hydrogels. The engineered hydrogels are increasingly considered as promising tools for transporting chemotherapeutic drugs and immunotherapeutics, with decreased systemic toxicities. As shown in Fig. 12, Zhao et al. [101] synthesized an injectable hydrogel with hydrophobic microdomains by glycol chitosan and benzaldehyde capped poly(ethylene glycol)-b-poly(propylene glycol)-b-poly(ethylene glycol) (PEO-PPO-PEO). DOX and PTX are physically encapsulated inside this hydrogel. The tumor microenvironment-responsive crosslinkers gives hydrogels the ability to release drugs in a controlled manner [131]. Zhang et al. [132] prepared various polyplexes with pentablock copolymer micelle (PB) and Pluronic F127 (PL) which can condense plasmid DNA (pDNA) and encapsulate PTX. Then a synthetic barrier gel based on poly(ethylene glycol) diacylate was developed to enable the released vectors to instantly and continuously transf ect cultured cells.

Hydrogels with micron or nanometer size, which termed as microgels and nanogels, also have important applications as vectors [133]. In addition to having the usual properties of hydrogels, microgels and nanogels are more suitable as carriers owing to their small size effect. As shown in Fig. 13, Lu et al. [102] synthesized a kind of polyacrylamide hybrid nanogels for targeted cancer chemotherapy via codelivery of Au NPs and MTX. Au NPs can be easily incorporated into/onto polyacrylamide (PAm) nanogels, whereas MTX was chemically conjugated to and physically adsorbed on Au-PAm hybrid nanogels. Delivery systems derive from microgels and nanogels can combine chemotherapy and gene therapy. Costa et al. [134] developed a new p53-encoded pDNA microgel with porosity, biocompatibility, and photodegradability, which is suitable for the dual release of pDNA and anticancer drug DOX through ultraviolet light irradiation. Microgels and nanogels exhibit outstanding design ability by combining the advantages of the nanocarrier and hydrogel.
3.1.5. Other organic nanoparticles

In addition to the vectors mentioned previously, many types of organic nanoparticles, such as dendrimers, hyperbranched polymers, polymer mesoporous microspheres, and some complexes, are also developed as delivery systems [135–138]. Dendrimers and hyperbranched polymers have hydrophobic cavities that are suitable for loading cargoes. Plenty of functional groups on the surface provide convenience for the subsequent modification. For example, Qian et al. [104] prepared star-branched amphiphilic copolymers for codelivery of DOX and miRNA (miR-21i) (Fig. 14). DOX was loaded into the hydrophobic core of the branched copolymer through hydrophobic interaction, and the outer layer was provided positive charges by polydimethylaminoethyl methacrylate (PDMAEMA) for binding the miR-21i. Dendrimers are branched macromolecules with many arms protruding from the central core [135]. Although the dendrimer is clear in structure and adjustable in size, its complex preparation process limits its wide application. The construction of codelivery systems using polymer mesoporous microspheres is similar to that of MSNs. Cargoes can be physically encapsulated in pores [139].

Complexes at the nanoscale are usually formed by entanglement of polymer chains. Many polymer gene vectors are formed by the complexation of cationic polymer chains with nucleic acid [140]. The polyelectrolyte complexes protect nucleic acids from degradation and promote transport across cell membranes. To achieve the joint transmission of genes and other reagents, several physical or chemical processes would be applied together in addition to electrostatic interaction.
As shown in Fig. 15, Oupický et al. [105] used AMD3100, a cyclam derivative as CXC chemokine receptor 4 (CXCR4) inhibitor, and disulfide-containing bisacrylamide to form cationic polymer chains. Because there are four nitrogen atoms in the center of the ring structure, the polycations, named RPA, can provide positive charges to condense with DNA. This strategy used drugs to structure polycations by a chemical process, and pDNA was loaded by a physical process. Chen et al. [141] synthesized the amphiphilic graft polymer polyethyleneimine-ferrocene (PEI-Fc), which can form micelles in aqueous solution through the hydrophobic side groups of ferrocenes. The oil/water (O/W) method was used to load DOX into micelles. DNA molecule were complexed by PEI through electrostatic interactions. Oh et al. [106] fabricated polypyrrole (PPy) nanoparticles by using bovine serum albumin-phycocyanin (Pc) complex, where PPy and Pc endow PTT and PDT to the nanoparticles respectively.

In addition to the conventional vectors mentioned above, some new...
kinds of vectors have been proposed [142, 143]. For example, Gu et al. [144] reported Trojan horse-like injectable engineered adipocytes that can serve as a drug-delivery depot for sustained drug release with suppressed primary tumor growth and postsurgical tumor recurrence. As shown in Fig. 16, the adipocytes were engineered with the encapsulation of rumenic acid, as an anticancer fatty acid, and a ROS-responsive DOX prodrug for chemotherapy and simultaneously inducing an immunogenic tumor phenotype. This section is discussed based on different types of vectors. One drug can be delivered by different types of vectors. To improve the codelivery efficiency of multiple drugs, it is important to select and design the vector reasonably.

3.2. Collaborative treatment strategies

In many cases, the purpose of constructing the codelivery systems is to combine different treatment strategies. In addition to chemotherapy, other therapeutic strategies, such as gene therapy, PDT, PTT, and immunotherapy, are also effective in tumor treatment. This requires the vectors can deliver multiple functional reagents, such as antibodies [145], nucleic acids [146], photosensitizer [147]. To observe the interactions between drugs and diseases, it is necessary to bring therapy and diagnosis together. Simultaneously delivering of imaging agents, including optically active small molecules, metals and metal oxides, ultrasonic contrast agents, and radionuclides, is required [148].

3.2.1. Combination of multiple chemotherapy strategies

Since the 1960s, combined chemotherapy with two or more chemotherapeutic drugs has been shown to be an effective strategy for cancer therapy [149]. Goldman et al. [150] demonstrated that two-in-one nanomedicine can terminate the origin of adaptive resistance by ensuring the delivery of two drugs with limited deterministic space to target cells, thereby inducing greater antitumor efficacy. In contrast, free drugs or two nanoparticles (each carry one drug) were less effective at binding because of the random distribution of cells. These findings suggested that two-in-one nanomedicines may become an important strategy for targeting adaptive resistance.

The combination of multiple chemotherapy strategies would reduce the side effects caused by chemotherapy drugs, delay the appearance of MDR, and improve the treatment effect [21]. The strategy for the combined administration of chemotherapeutic drugs depends on the antitumor mechanism of each drug. For example, DOX and PTX codelivery strategies have developed rapidly in recent years. DOX is one of the most effective anthracycline antitumor drugs. It can interfere with DNA by insertion and then induce apoptosis in cancer cells. PTX is a representative antimicrotubule agent [151]. PTX specifically acts on the G2 and M phases of the cell cycle, making microtubules unable to form spindles and spindle filaments during mitosis, and preventing tumor cells from dividing and reproducing. Chemotherapy enhancement was observed by administering PTX and DOX in a reasonable order.

Yang et al. [103] synthesized a kind of ROS-sensitive PEGylated bilirubin nanoparticles (BRNPs) to encapsulate dimer-7-ethyl-10-hydroxycamptothecin (d-SN38) and dimer-lonidamine (d-LND). As shown in Fig. 17, dimerization of the drugs significantly increases the drug loading capacity and the encapsulation efficiency of nanoparticles. SN38, which is an active metabolite of CPT in vivo, can act on DNA topoisoasemerase to kill cancer cells [152–154], whereas hexokinase inhibitor LND exhibits antitumor activity by affecting energy metabolism [155]. This collaborative treatment strategy strongly avoided the non-specific drug action. Similarly, Du et al. [156] fabricated a dual drug-paired polydrug nanoparticle (DOX, 25 wt%; cisplatin (CDDP), 12.5 wt%). This work provides an innovative strategy for reversing MDR and combating DOX-resistant breast and CDDP-resistant ovarian cancers. As the mechanism of antitumor action of various drugs becomes clear, more collaborative therapies are proposed, which accelerate the design and preparation process of novel codelivery systems.

3.2.2. Combination of chemotherapy and gene therapy

Gene therapy refers to the introduction of foreign normal genes into target cells to correct or compensate the defective and abnormal genes [157–159]. Therapeutic genes, including pDNA, miRNA, and siRNA, play important roles in the regulation of cell development, differentiation, metabolism, and apoptosis. To realize the codelivery of chemotherapeutic drugs and genes, the commonly used method is to encapsulate chemotherapeutic drugs in the carrier and adsorb genes on the surface [160–163]. Zhu et al. [98] reported on cationic micelles based on
well-defined PDMAEMA-PCL-PDMAEMA triblock copolymers for the combinatorial delivery of siRNA and PTX. The zeta potential of the prepared cationic micelles is from $+29.3$ to $+35.5$ mV, which is suitable to complex with siRNA. PTX molecules are loaded in the hydrophobic cavity of the micelles. Rattle-structured rough nanocapsules (Au@HSN-PGEA, AHPs) composed of in situ formed Au NR cores and polycationic mesoporous silica shells were constructed for trimodal complementary cancer therapy [95]. As shown in Fig. 18, the outer shell of the nanorattles was functionalized with a superior polycation, CD-PGEA (two-armed ethanolamine-functionalized poly(glycidyl methacrylate) with one $\beta$-CD core) to carry antioncogene p53 for gene therapy. The interior space around the Au NR cores was reserved for sorafenib (SF, a hydrophobic antiproliferative and antiangiogenic drug) loading. A well-designed combination of chemotherapy and gene therapy can produce synergies to improve the effectiveness of cancer treatment and push back MDR. As more and more therapeutic genes are proposed, the combination of gene therapy with other therapeutic strategies is being widely studied.

3.2.3. Combination of chemotherapy and immunotherapy

In recent years, immunotherapy has great prospects for inhibiting metastatic cancer [164]. There were previous reports that the combination of chemotherapy and immunotherapy may result in enhanced anticancer effects and can be used to promote an immunogenic tumor phenotype [165–168]. As shown in Fig. 19, Wang et al. [169] engineered a therapeutic hydrogel that, when formed in situ, allows the local release of GEM and an anti-PD-L1 blocking antibody (aPDL1) with distinct release kinetics. This strategy promotes immune-mediated tumor regression in the tumor-bearing mice, with the prevention of tumor recurrence after primary resection. Similarly, using this therapeutic hydrogel, codelivery of aPDL1 and Zebularine (Zeb), an epigenetic modulator which can enhance the antitumor immune response by inducing the expression of tumor-associated antigens, was achieved [170]. The combination of chemotherapy and immunotherapy also increased the therapeutic effect of chemotherapy-insensitive tumors. For example, docetaxel (DTX), a chemotherapeutic agent, and cholesterol-modified Toll-like receptor 9 (TLR9) agonist CpG (cho-CpG) oligonucleotide are co-loaded in synthetic high-density lipoprotein nanoparticles (sHDL) nanodiscs for chemoinmunotherapy of colon adenocarcinoma. This collaborative treatment strategy can be used to improve survival outcomes significantly compared with chemotherapy alone.

3.2.4. Combination of chemotherapy and PTT/PDT

PTT and PDT, are the two most important phototherapy strategies. Copper sulfide NPs [171], gold NPs [172], graphene oxide [173], and carbon nanotubes [174] are commonly used for PTT. Chlorine e6 (Ce6) [175], ICG [176], and rose bengal [177] are widely used for PDT. Under the irradiation of a particular wavelength laser, photosensitizers can convert light energy into thermal energy ($>42^\circ$C) or ROS. Tumor cells will be destroyed in this abnormal environment. The combination of phototherapy and chemotherapy has been shown to be feasible and effective. As shown in Fig. 20, Mao et al. [178] developed cyclic cRGDfK peptide and Ce6 conjugated silk fibroin-based nanoparticles for 5-FU delivery and PDT. Chemotherapy and PDT are well combined. Liu et al. [179] constructed an azobenzene (AZO)-containing conjugated polymer-based nanocarrier for codelivery of CPT and Ce6. Under 670 nm laser irradiation, Ce6 generated ROS for PDT and induced tumor hypoxia. The hypoxia-responsive drug release occurred subsequently for chemotherapy. Yu et al. [93] developed a chemotherapeutic agent and photosensitizer corelease system which was constructed by filling the interior of gold nanocages (AuNCs) with DOX, ICG and 1-tetradeccanol, and modifying the surface with biotinylated poly (ethylene glycol) via Au-S bonds. Under the irradiation of 808 nm laser, AuNCs will heat up rapidly and release DOX and ICG simultaneously. Some ROS-responsive linkers can be introduced into the carrier to achieve controlled release of cargoes [180].

Chemotherapy can also be used in conjunction with other therapies...
through codelivery vectors, such as synergistic chemotherapy and protein therapy [181,182]. In addition to chemotherapy, combinations of different treatment strategies, such as PDT/PTT combined therapy, have been shown to be effective [106,183]. More radically, Chang et al. [184] constructed a multifunctional cascade bioreactor based on hollow mesoporous Cu2MoS4 loaded with glucose oxidase for synergetic cancer therapy by chemo-dynamic therapy/starvation therapy/phototherapy/immunotherapy. As can be seen from the aforementioned example, collaborative treatment strategies achieved by codelivery systems are promising treatments for cancer.

4. Conclusions and perspectives

To achieve ideal therapeutic effect of tumor treatment, the combination of several drugs or strategies had been proposed. The codelivery system is the key to realize combined therapies. The different structures of drugs and vectors bring a variety of directions to the design of codelivery systems. The interaction between cargoes and carriers, as well as the synergistic action between loaded reagents should be fully considered. It is necessary to select appropriate modes or cooperative strategies in accordance with the structure and property of cargoes and carriers. Encapsulating of reagents into vectors by physical processes is usually easy to operate. However, the stability of such kind of delivery systems needs to be explored in detail. The delivery systems constructed by chemical processes are relatively stable. Concomitantly, chemical processes usually involve multiple chemical reactions, where the way of drug release has to be precisely designed. Stimuli-responsive chemical bonds or structures should be introduced between drug molecules and backbones to achieve the controlled release of drugs. This review presented the typical design considerations about codelivery vectors for cancer therapy and described their current states from two aspects: different types of vectors and collaborative treatment strategies. As shown in Fig. 21, the first aspect focuses on the design and construct of vectors, whereas the latter focuses on the combined application of different treatment strategies.

Synergies between different drugs or strategies have been have exhibited excellent therapeutic effects via codelivery systems. The integration of diagnosis and treatment could be realized by carefully selecting constitute substrates. In the meantime, many obstacles still need to overcome. In the design process of the vectors, the advantages...
and disadvantages of the matrix should be emphasized. For example, although metal-based nanoparticles have appended functions, they are usually modified to serve as vectors, and their biocompatibility needs to be improved. Much attention should be paid to the potential inter-restriction of each part of codelivery systems, including matrices and cargoes. A delicate equilibrium between each part requires precise regulation. Besides, the degraded components of vectors should have no negative influences on systems. For the future development of codelivery systems, more definite pharmacological actions and new strategies provide infinite possibilities for the design and preparation of codelivery systems.

Based on laboratory research, multiple vectors and strategies are available to prepare codelivery with ideal performances. However, owing to the physiological and pathological differences between humans and rodents, even the difference of social attributes, the treatment plan that relatively effective in rodents may not necessarily efficacy in a human trial. For clinical application, administration methods and many stringent biosafety assessments of vectors need to be assessed. In addition, controlling of production cost and simplifying of preparation processes are also important factors from laboratory to clinic. Therefore, the design of codelivery systems for cancer therapy should be considered holistically. Accompanied by the introduction of novel technologies, such as the application of molecular machines in the field of drug delivery systems, more codelivery systems with promising performances and therapeutic effects will be proposed.

Author contributions

Q.Y.M. and H.L.C. contributed to writing of the manuscript and also in the preparation of the original draft. H.H. and F.J.X. contributed to writing, reviewing, and editing of the manuscript. F.J.X. supervised.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this article.
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