A new species of Jesogammarus from the Iki Island, Japan (Crustacea, Amphipoda, Anisogammaridae)

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Abstract

A new species of anisogammarid amphipod, Jesogammarus (Jesogammarus) ikiensis sp. n., is described from freshwaters in the Iki Island, Nagasaki Prefecture, Japan, based on results of morphological and molecular analyses. The new species is distinguished from all members of the genus by the combination of small number of setae on dorsal margins of pleonites 1–3, short and small number of setae on posterior margins of peduncular articles of antennae, mandibular article 1 without setae, well developed posterior lobes of accessory lobes of coxal gills on gnathopod 2 and pereopods 3–5, and pectinate setae on palmar margin of female gnathopod 2. A key to all the species of Jesogammarus is provided.

Keywords

Jesogammarus, Anisogammaridae, Amphipoda, Iki Island, Japan, new species, taxonomy

Introduction

The amphipod genus Jesogammarus Bousfield, 1979 has been recorded from fresh and brackish waters of the Japanese archipelago, the Korea peninsula, and the Chinese continent (Bousfield 1979; Morino 1984, 1985, 1986, 1993; Lee and Seo 1990, 1992;
Tomikawa and Morino 2003; Tomikawa et al. 2003, Hou and Li 2004, 2005). To date, 17 species in two subgenera, Jesogammarus Bousfield, 1979 and Annanogammarus Bousfield, 1979, have been recognized.

In 2010, Mr. Y. Tohyama of Hiroshima University provided a few specimens of freshwater amphipod collected from the Iki Island, Nagasaki Prefecture, Japan. They proved to belong to a previously unknown species of Jesogammarus. The Iki Island is located between Kyushu and the Tsushima Island, and 14 km from east to west and 17 km from north to south (Fig. 1). During field surveys of freshwater amphipods in the Iki Island, made in 2010–2015, a significant number of specimens of this species have been accumulated. Close examination of the external morphology and molecular analyses based on mitochondrial DNA sequences revealed that the Iki species is distinct from its congeners, and it is described as a new species.

Materials and methods

Samples

Specimens of Jesogammarus ikiensis sp. n. were collected from four localities in Iki Island, Nagasaki Prefecture, Japan (Fig. 1) by scooping with a fine-mesh hand-net, and preserved in 99% ethanol at the sites. For comparison, DNA sequences data were obtained for specimens of all the Japanese species of Jesogammarus, J. (A.) annandalei (Tattersall, 1922), J. (A.) fluvialis Morino, 1985, J. (J.) fujinoi Tomikawa & Morino, 2003, J. (J.) hinumensis Morino, 1993, J. (J.) hokurikuensis Morino, 1985, J. (J.) jezoensis (Schellenberg, 1937), J. (J.) mikadoi Tomikawa, Morino & Mawatari, 2003, J. (A.) naritai Morino, 1985, J. (J.) paucisetulosus Morino, 1984, J. (J.) shonaiensis Tomikawa & Morino, 2003, J. (J.) spinopalpus Morino, 1985, and J. (A.) suwaensis Morino, 1986. Details of these specimens are shown in Table 1. No sequence data are available for J. (A.) debilis Hou & Li, 2005, J. (J.) fontanus Hou & Li, 2004, J. (J.) hebeiensis Hou & Li, 2004 J. (J.) ilhoii Lee & Seo, 1990, or J. (A.) koreaensis Lee & Seo, 1992.

Morphological observation

All appendages of the examined specimens of Jesogammarus ikiensis sp. n. were dissected in 99% ethanol and mounted in gum-chloral medium on glass slides under a stereomicroscope (Olympus SZX7). Specimens were examined using a light microscope (Nikon Eclipse Ni) and illustrated with the aid of a camera lucida. The body length from the tip of the rostrum to the base of the telson was measured along the dorsal curvature to the nearest 0.1 mm. The nomenclature of the setal patterns on the mandibular palp follows Stock (1974). The specimens are deposited in the Tsukuba Collection Center of the National Museum of Nature and Science, Tokyo (NSMT).
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Figure 1. Sampling localities for Jesogammarus (Jesogammarus) ikiensis sp. n. A Map of Japan and adjacent area showing Iki Island, Nagasaki Prefecture, Japan B the collecting localities of Iki Island: 1, Katsumoto; 2, Ashibe; 3, Ishida; 4, Gonoura.
Table 1. Species, sampling localities, and numbers of specimens used for molecular phylogenetic study.

| Species                      | Voucher | Locality                                      | DDBJ Acc. No.       | Reference  |
|------------------------------|---------|-----------------------------------------------|---------------------|------------|
| Eogammarus kygi              | G1      | Naibetsu River, Eniwa, Hokkaido, Japan        | LC052229 LC052250   | this study |
| Eogammarus posjeticus        | G3      | Akkeshi, Hokkaido, Japan                      | LC052230 LC052251   | this study |
| Jesogammarus annandalei      | G1162   | Lake Biwa, Shiga Prefecture, Japan            | LC052231 LC052252   | this study |
| Jesogammarus fluvialis       | G83     | Samegai, Shiga Prefecture, Japan              | LC052232 LC052253   | this study |
| Jesogammarus fujinoi         | G17     | Gobanmiki, Yamagata, Yamagata Prefecture, Japan | LC052233 LC052254   | this study |
| Jesogammarus hinumenensis    | G52     | Lake Hinuma, Ibaraki Prefecture, Japan        | LC052234 LC052255   | this study |
| Jesogammarus hokurikuensis   | G383    | Takinami, Fukui, Fukui Prefecture, Japan      | LC052235 LC052256   | this study |
| Jesogammarus jesoensis       | G164    | Sapporo, Hokkaido, Japan                      | LC052236 LC052257   | this study |
| Jesogammarus mikadoi         | G13     | Rokugo, Akita Prefecture, Japan               | LC052237 LC052258   | this study |
| Jesogammarus naritai         | G1167   | Lake Biwa, Shiga Prefecture, Japan            | LC052238 LC052259   | this study |
| Jesogammarus paucistulosus   | G1037   | Mito, Ibaraki Prefecture, Japan               | LC052239 LC052260   | this study |
| Jesogammarus ibonaiensis     | G192    | Sakata, Yamagata Prefecture, Japan            | LC052240 LC052261   | this study |
| Jesogammarus ikiensis sp. n. | G515    | Katsumoto, Iki, Nagasagi Prefecture, Japan    | LC052241 LC052262   | this study |
| Jesogammarus ikiensis sp. n. | G665    | Ishida, Iki, Nagasaki Prefecture, Japan       | LC052242 LC052263   | this study |
| Jesogammarus ikiensis sp. n. | G695    | Ishida, Iki, Nagasaki Prefecture, Japan       | LC052243 LC052264   | this study |
| Jesogammarus ikiensis sp. n. | G885    | Ishida, Iki, Nagasaki Prefecture, Japan       | LC052244 LC052265   | this study |
| Jesogammarus ikiensis sp. n. | G886    | Ishida, Iki, Nagasaki Prefecture, Japan       | LC052245 LC052266   | this study |
| Jesogammarus spinopalpus     | G32     | Onjuku, Chiba Prefecture, Japan               | LC052246 LC052267   | this study |
| Jesogammarus suwaensis       | G88     | Lake Suwa, Nagano Prefecture, Japan           | LC052247 LC052268   | this study |
| Jesogammarus suwaensis       | G89     | Lake Suwa, Nagano Prefecture, Japan           | LC052248 LC052269   | this study |
| Spaskogammarus spaskii       | G35     | Akkeshi, Hokkaido, Japan                      | LC052249 LC052270   | this study |

DNA extraction, PCR amplification, and DNA sequencing

Total genomic DNA was extracted from pereopod musculature of each sequenced amphipod (Table 1), by means of the DNeasy blood and tissue kit (Qiagen, Hilden, Germany); the final volume of the DNA solution following extraction was 200 µl. Part of the mitochondrial cytochrome c oxidase subunit I (COI) and 16S ribosomal RNA (rRNA) genes were amplified by polymerase chain reaction (PCR) using the following primer pair: Am-COI-H [CG(AG)GC(CGT)TA(CT)TT(CT)AC(CT)TC(ATC)G(CAC)ACTAT] and Am-COI-T [CGTCG(AGT)GG(CT)AT(ACG)CC(ACGT)
CT(AGT)A(AG)(ATC)CCTA] (Tomikawa et al. 2007); 16STf [GGTAA(T)A(CT) C(T)TA(G)ACC(T)TGCTAAG] (Macdonald et al. 2005) and 16Sbr [CCGGTTT- GAACTCAGATCATGT] (Palumbi et al. 1991). PCR reactions containing 0.5 µl template solution, 2 mM MgCl₂, 2.5 mM dNTP, 10 pmol of each primer, and 5U/µl Taq polymerase (TaKaRa Ex Taq®) in 1X buffer provided by the manufacturer were performed in 10-µl volumes in an PC-320 thermal cycler (ASTEC). Amplification conditions were as follows: an initial denaturation for 7 min at 94 °C; 35 cycles of denaturation for 45 s at 94 °C, annealing for 1 min at 42–50 °C depending on samples, and extension for 1 min at 72 °C; and final extension for 7 min at 72 °C. Amplification products were purified by the silica method (Boom et al. 1990). All sequencing reactions were performed according to the manufacturer’s instructions using the BigDye Terminater v3.1 Cycle Sequencing Reaction Kit (Applied Biosystems, Foster City, CA). Cycle sequencing conditions were 25 cycles of 10 s at 96 °C, 5 s at 50 °C, and 4 min at 60 °C. Sequencing reaction products were purified by ethanol precipitation. Labeled fragments were analyzed using an ABI 3130x Genetic Analyzer (Applied Biosystem). Sequences were obtained from both strands of the gene segments for verification using the same primers. The nucleotide sequences have been submitted to the DNA Databank of Japan (DDBJ) nucleotide-sequence database (linked to the EMBL and GenBank databases) (Table 1).

Molecular phylogenetic analyses

The nucleotide sequences were aligned using the multiple alignment algorithm in Clustal W (Thompson et al. 1994) with default setting (i.e., gap opening penalty = 15, gap extension penalty = 6.66, transition weight = 0.5). Phylogenetic relationships were reconstructed by the Neighbor-Joining method (NJ; Saitou and Nei 1987), the equally weighted maximum parsimony method (MP), and the maximum likelihood method (ML) with MEGA6 software (Tamura et al. 2013). There was no indel in COI sequences of the ingroup taxa. On the other hand, eight indels were found in 16S sequences of the ingroup taxa, which were treated as missing data in all analyses. In the NJ analysis, the Kimura 2-parameter (K2P) model (Kimura 1980) of nucleotide substitution was used to estimate genetic distances. In the MP analysis, a tree was obtained using the Close-Neighbor-Interchange algorithm, in which the initial trees were obtained with the random addition of sequences (10 replicates). The ML analysis used the T92 + G + I model for COI and HKY + G for 16S and COI + 16S; this was selected as the best-fit model using the Bayesian information criterion (BIC) in MEGA6. To estimate statistical support for branching patterns, 1,000 bootstrap replications each (Felsenstein 1985) were performed for the NJ, MP, and ML analyses. As outgroup taxa, three anisogammarid species, *Eogammarus kygi* (Derzhavin, 1923), *E. posjeticus* (Tzvetkova, 1967), and *Spasskogammarus spaskii* (Bulycheva, 1952), were used (Table 1).
Figure 2. A maximum-likelihood tree from a 333 bp sequence of COI gene B maximum-likelihood tree from a 416 bp sequence of 16S rRNA gene. Numbers near the branches are ML/NJ/MP bootstrap values. Bootstraps are shown when ≥ 70%. Vouchers are shown after species names as in Table 1.
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**Remarks**

Monophyly of the subgenera Jesogammarus and Annanogammarus were supported in COI, 16S, and COI + 16S trees (Figs 2, 3). Jesogammarus *ikiensis* sp. n. from Iki Island was included in the clade of the subgenus Jesogammarus. However, phylogenetic position of *J. ikiensis* was not clearly resolved in the phylogenetic trees based on the COI and 16S rRNA genes due to low bootstrap values. Jesogammarus *ikiensis* differs from the all Japanese congeners by large genetic distances (18.6–25.8% for COI and 12.7–18.7% for 16S) (Table 2), which were larger than intraspecific distances among many species of Jesogammarus. In addition, *J. ikiensis* was morphologically distinguished from its congeners. Thus, it can be concluded that *J. ikiensis* from Iki Island as a distinct new species and is described below.

**Systematics**

**Jesogammarus (Jesogammarus) ikiensis** sp. n.  
http://zoobank.org/75FDE441-CD57-41C4-B154-1E3D29ECAC3E  
New Japanese name: Iki-yokoebi  
Figures 3–9

**Material examined.** Holotype: NSMT-Cr 24107, male (13.1 mm, 8 slides), river at Ishida (33°45′1.7″N, 129°44′33.7″E), Iki, Nagasaki Prefecture, Japan, collected by K. Tomikawa and S. Tashiro on 9 March 2012. Paratypes: NSMT-Cr 24108, ovigerous female (10.4 mm, 6 slides), NSMT-Cr 24109, 1 male and 1 ovigerous female in ethanol vial, data same as for holotype; NSMT-Cr 24110, 2 males and 2 ovigerous females in ethanol vial, river at Ishida (33°45′1.7″N, 129°44′33.7″E), Iki, Nagasaki Prefecture, Japan, collected by K. Tomikawa and S. Tashiro on 2 April 2015; NSMT-Cr 24111, male (12.0 mm, 6 slides), NSMT-Cr 24112, ovigerous female (9.4 mm, 5 slides), river at Katsumoto (33°49′30.1″N, 129°42′51.5″E), Iki, Nagasaki Prefecture, Japan, collected by K. Tomikawa and S. Tashiro on 8 March 2012; NSMT-Cr 24113, male (11.9 mm, 5 slides), NSMT-Cr 24114, ovigerous female (10.0 mm, 5 slides), river at Ashibe (33°47′3.1″N, 129°45′3.8″E), Iki, Nagasaki Prefecture, Japan, collected by K. Tomikawa and S. Tashiro on 9 March 2012; NSMT-Cr 24115, male (9.2 mm, 5 slides), NSMT-Cr 24116, female with offsprings (7.4 mm, 5 slides), irrigation ditch at Gonoura (33°43′26″N, 129°41′52″E), Iki, Nagasaki Prefecture, Japan, collected by K. Tomikawa and S. Tashiro on 9 March 2012.

**Description of male (holotype, NSMT-Cr 24107).** Head (Fig. 4) with short rostrum; ventral margin of lateral cephalic lobe weakly concave; antennal sinus rounded; eyes reniform, major axis 0.4 × height of head. Dorsal surfaces of pereonites smooth (Fig. 4). Dorsal margins of pleonites 1–3 (Fig. 9C–E) with three, two, and two setae, respectively. Posterior margin of epimeral plate 1 rounded with seta, anteroventral cor-
Table 2. Uncorrected pairwise differences (%: p-distance) of partial COI (upper right) and 16S rRNA (lower left) gene sequences between species.

| Species  | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 |
|----------|---|---|---|---|---|---|---|---|---|---|----|----|----|----|----|----|
| Eogammarus kygi (N = 1) | 19.5 | 24.3 | 23.7 | 30.6 | 27.3 | 27.0 | 27.9 | 30.0 | 24.3 | 28.5 | 29.1 | 29.4 | 27.6 | 24.3 | 25.2 |
| Eogammarus possjeticus (N = 1) | 16.3 | 24.9 | 24.0 | 27.0 | 27.0 | 26.4 | 25.5 | 27.6 | 23.7 | 28.5 | 27.6 | **28.5** | 28.2 | 23.4 | 24.3 |
| Jesogammarus annandalei (N = 1) | 20.8 | 22.3 | 3.0 | 24.0 | 20.7 | 23.1 | 22.5 | 25.8 | 3.3 | 20.7 | 24.3 | **21.6** | 20.7 | 3.0–3.3 | 22.5 |
| Jesogammarus fluviialis (N = 1) | 20.4 | 22.0 | 2.0 | 24.0 | 19.5 | 22.5 | 21.9 | 25.2 | 2.1 | 20.7 | 22.8 | **21.6** | 20.4 | 1.8–2.1 | 22.8 |
| Jesogammarus fujinoi (N = 1) | 25.1 | 24.0 | 20.6 | 20.7 | 19.8 | 15.3 | 10.5 | 18.9 | 22.5 | 20.4 | 9.3 | **24.6** | 21.3 | 22.8–23.1 | 27.9 |
| Jesogammarus hinunensis (N = 1) | 22.3 | 23.9 | 18.0 | 17.6 | 15.1 | 20.7 | 17.7 | 21.9 | 19.5 | 18.6 | 18.6 | **19.5** | 18.6 | 19.2–19.5 | 25.8 |
| Jesogammarus hokurikuensis (N = 1) | 24.3 | 24.3 | 20.3 | 20.2 | 11.2 | 17.1 | 21.3 | 22.5 | 19.5 | 12.6 | **23.7** | 19.8 | 22.2–22.5 | 26.1 |
| Jesogammarus jesoensis (N = 1) | 20.4 | 22.0 | 2.0 | 24.0 | 19.5 | 22.5 | 21.9 | 25.2 | 2.1 | 20.7 | 22.8 | **21.6** | 20.4 | 1.8–2.1 | 22.8 |
| Jesogammarus mikadoi (N = 1) | 25.1 | 24.0 | 20.6 | 20.7 | 19.8 | 15.3 | 10.5 | 18.9 | 22.5 | 20.4 | 9.3 | **24.6** | 21.3 | 22.8–23.1 | 27.9 |
| Jesogammarus naritai (N = 1) | 20.8 | 21.6 | 1.9 | 1.2 | 19.8 | 17.5 | 19.9 | 19.6 | 20.0 | 20.1 | 22.2 | **20.7** | 20.1 | 0.3–0.6 | 23.4 |
| Jesogammarus paucisetulosus (N = 1) | 24.2 | 25.2 | 18.7 | 18.8 | 16.3 | 15.6 | 16.3 | 16.4 | 18.7 | 18.3 | 19.8 | **21.0** | 21.0 | 20.1–20.7 | 25.2 |
| Jesogammarus shonaiensis (N = 1) | 25.5 | 25.1 | 21.9 | 21.4 | 7.3 | 15.5 | 10.3 | 7.3 | 15.1 | 20.8 | 17.4 | **22.8** | 20.4 | 21.9–22.2 | 27.9 |
| Jesogammarus ikiensis sp. n. (N = 5) | 23.4–23.5 | 24.0–24.2 | 18.2–18.3 | 18.3–18.4 | 18.0–18.2 | 12.7–12.8 | 17.8–17.9 | 18.2–18.3 | 18.6–17.8 | 16.8–17.0 | 17.4 | **18.6** | **20.4–20.7** | 26.1 |
| Jesogammarus spinopalpus (N = 1) | 23.0 | 24.6 | 17.0 | 17.0 | 17.1 | 15.9 | 16.6 | 16.7 | 16.8 | 16.6 | 16.4 | 17.1 | **14.8–15.0** | 19.5–19.8 | 24.6 |
| Jesogammarus suwaisensis (N = 2) | 20.8–21.0 | 21.6–21.8 | 1.9–2.0 | 1.2–1.3 | 20.0–20.2 | 17.5–17.6 | 20.0–20.2 | 19.5–19.6 | 20.0–20.2 | 0.3–0.4 | 18.3–18.4 | **20.8–21.0** | 17.6–17.9 | 16.4–16.6 | 22.8–23.1 |
| Spasskogammarus spasskii (N = 1) | 22.2 | 22.8 | 20.6 | 20.8 | 22.6 | 22.0 | 22.2 | 22.3 | 23.0 | 20.8 | 21.1 | 22.4 | **21.5–21.6** | 21.0 | **20.6–20.7** |
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**Figure 3.** Maximum-likelihood tree from a 749 bp sequence of COI + 16S rRNA genes. Numbers near the branches are ML/NJ/MP bootstrap values. Bootstraps are shown when ≥ 70%. Vouchers are shown after species names as in Table 1.

**Figure 4.** Jesogammarus (*Jesogammarus*) *ikiensis* sp. n., holotype, male, 13.1 mm, NSMT-Cr 24107, Ishida, Iki, Nagasaki Prefecture, Japan. *Habitus*, lateral view.
ner with many setae (Fig. 9I); posterior margin of plate 2 with one seta, posteroventral corner square, anteroventral corner with three setae, ventral submargin with four robust setae (Fig. 9J); posterior margin of plate 3 with two setae, posteroventral corner square, anteroventral to ventral margin with six setae (Fig. 9K). Urosomites 1–3 (Fig. 9F–H) with seven, four, and two robust setae associated with slender setae.

Antenna 1 (Fig. 5A): length 0.7 × body length; peduncular articles 1–3 in length ratio of 1.0 : 0.9 : 0.5; posterodistal corner of peduncular article 1 with one robust seta, posterior margin of peduncular article 2 with one cluster and three pairs of setae, posterior margin of peduncular article 3 with one cluster and one pair of setae; accessory flagellum seven-articulate; primary flagellum 29-articulate, each article with one aesthetasc.

Antenna 2 (Fig. 5B): length 0.7 × antenna 1; posterior margin of peduncular article 4 with three clusters of setae, posterior margin of peduncular article 5 with three clusters of setae and one single seta; flagellum 18-articulate, calceoli present (Fig. 5C).

Mouthparts. Upper lip (= labrum) (Fig. 5D) with rounded distal margin, bearing fine setae. Lower lip (= labium) (Fig. 5E) with broad outer lobes, inner lobes indistinct. Mandibles (Fig. 5F–H) with left and right incisors six- and four-dentate, respectively, left lacinia mobilis five-dentate, right one bifid, bearing many teeth; molar process triturative, with plumose setae; accessory setal rows of left and right mandibles each with seven blade-like setae; left palp three-articulate with length ratio of 1.0 : 3.8 : 3.8, palp article 1 bare, article 2 with 28 setae, article 3 with two clusters and one pair of A-setae, one pair of B-setae, and many C-, D-, and E-setae, article 3 of right palp with three clusters of A-seta and one B-seta. Maxilla 1 (Fig. 5I) with inner and outer plates and palp; medial margin and apical submargin of inner plate with 31 plumose setae; outer plate subrectangular, with 11 serrate teeth apically (Fig. 5J); right palp two-articulate, much longer than outer plate, article 1 lacking marginal setae, article 2 with seven robust and six slender setae on its apical margin, outer margin with three setae, left palp lacking setae on outer margin of article 2. Maxilla 2 (Fig. 5K) with oblique inner row of 23 plumose setae on inner plate; outer plate slightly longer than inner plate. Maxilliped (Fig. 6A) with inner and outer plates and palp; inner plate (Fig. 6C) with six robust setae along apical and inner margins; outer plate (Fig. 6B) with plumose setae on apical margin and robust setae on inner margin; palp four-articulate, article 2 with inner marginal and submarginal rows of setae, article 3 with facial setae, article 4 slightly curved inward, with slender nail.

Gnathopod 1 (= pereopod 1) (Fig. 6D): coxa (= article 1) with six setae on ventral margin; anterior and posterior margins of basis (= article 2) with long setae; carpus (= article 5) length 1.4 × width, anterior margin with seta; propodus (= article 6) length 1.2 × length of carpus and 1.3 × width of propodus, anterior margin with one pair and two clusters of setae, palmar margin (Fig. 6E) oblique, weakly convex, with 16 pegspines (= robust setae); dactylus (= article 7) (Fig. 6E) as long as palmar margin, with posterior accessory blade longer than nail, blade basally elevated.

Gnathopod 2 (= pereopod 2) (Fig. 6F): coxa with seven marginal and one submarginal setae on ventral part, posteroproximal part with two setae; anterior and posterior
margins of basis with long setae; carpus length 1.7 × width, anterior margin with cluster of setae and single seta; propodus almost as long as carpus and 1.5 × width of propodus, anterior margin with two clusters of setae, palmar margin (Fig. 6G) oblique, weakly convex, with 12 peg-spines (= robust setae) and one serrate seta; dactylus (Fig. 6G) as long as palmar margin, with posterior accessory blade longer than nail.

_Pereopod 3_ (Fig. 7A, B): coxa with seven marginal setae on ventral part, posterio-proximal part with two setae; anterior and posterior margins of basis with long setae, anterio-distal corner of basis with robust seta.

_Pereopod 4_ (Fig. 7C, D): coxa expanded with posterior concavity, bearing one seta on anterodistal corner and five setae on posterodistal margin; anterior and posterior margins of basis with long setae, anterodistal corner with robust seta.

_Pereopod 5_ (Fig. 7F, G): coxa bilobed, anterior lobe with apical seta, ventral margin of posterior lobe with three setae; posterior margin of basis weakly expanded, with ten setae; anterior and posterior margins of merus to propodus with robust and slender setae.

_Pereopod 6_ (Fig. 8A, B): coxa bilobed, anterior lobe with apical seta and anterio-proximal setae, ventral margin of posterior lobe with three setae; posterior margin of basis weakly expanded, with 18 setae; anterior and posterior margins of merus to propodus with robust and slender setae.

_Pereopod 7_ (Fig. 7D, E): ventral margin of coxa weakly concave, bearing slender setae on anterior part and three setae on posterodistal margin; posterior margin of basis weakly expanded, with 20 setae; anterior and posterior margins of merus to propodus with robust and slender setae.

_Coxal gills_ on gnathopod 2 and pereopods 3–5 (Figs 6H, 7A, C, E) with two accessory lobes, gills on pereopods 6 and 7 (Fig. 8C, F) each with one accessory lobe.

_Pleopods 1–3_ (Fig. 8G) each with paired retinacula (Fig. 8H) on inner margin of peduncle, and bifid plumose setae (= clothes-pin setae) (Fig. 8I) on inner basal margin of inner ramus.

_Uropods_. Uropod 1 (Fig. 8J): peduncle with robust seta on basofacial part, inner and outer margins each with three robust setae, inner proximal part with three short setae; inner ramus length 0.8 × peduncle, inner margin with three robust setae and outer margin with robust seta and minute seta; outer ramus length 0.9 × inner ramus, inner and outer margins each with two and three robust setae. Uropod 2 (Fig. 8K): peduncle with three robust setae on inner and outer margins, respectively; inner ramus length 0.9 × peduncle, its inner and outer margins with two robust setae, respectively; outer ramus length 0.8 × inner ramus, its outer margin with robust seta. Uropod 3 (Fig. 9A): peduncle length 0.3 × outer ramus; inner ramus length 0.25 × outer ramus (both proximal and terminal articles), with two robust setae on inner margin; outer ramus two-articulate, inner margin of proximal article with five plumose setae, and several robust setae and simple setae, outer margin with robust setae and simple setae, terminal article length 0.2 × proximal article, with short setae apically.

_Telson_ (Fig. 9B) length 1.1 × width, cleft for 59% of length in V-shape; each lobe with one lateral and one apical robust seta.
Figure 5. *Jesogammarus (Jesogammarus) ikiensis* sp. n., holotype, male, 13.1 mm, NSMT-Cr 24107, Ishida, Iki, Nagasaki Prefecture, Japan. **A** peduncular articles 1–3, accessory flagellum, and flagellar articles 1–4 of antenna 1, medial view (posterior-marginal setae on peduncular articles 2 and 3 indicated by arrowheads) **B** peduncular articles 1–5 and flagellar articles 1–3 of antenna 2, medial view (posterior-marginal setae on peduncular articles 4 and 5 indicated by arrowheads) **C** calceolus of antenna 2, medial view **D** upper lip, anterior view **E** lower lip, ventral view **F** left mandible except palp, medial view **G** incisor and lacinia mobilis of right mandible, lateral view **H** palp of right mandible, medial view **I** maxilla 1, dorsal view **J** outer plate of maxilla 1, dorsal view **K** Maxilla 2, dorsal view.
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**Figure 6.** *Jesogammarus* (*Jesogammarus*) *ikiensis* sp. n., holotype, male, 13.1 mm, NSMT-Cr 24107, Ishida, Iki, Nagasaki Prefecture, Japan. A maxilliped, dorsal view B outer plate of maxilliped, dorsal view C inner plate of maxilliped, dorsal view D gnathopod 1, medial view E palmar margin of propodus and dactylus of gnathopod 1, medial view F gnathopod 2, medial view G palmar margin of propodus and dactylus of gnathopod 2, medial view H coxal gill of gnathopod 2, medial view.

**Description of ovigerous female (paratype, NSMT-Cr 24108).** Antenna 1 (Fig. 9L): length 0.7 × body length; peduncular articles 1–3 in length ratio of 1.0 : 0.8 : 0.5; accessory flagellum seven-articulate; primary flagellum 36-articulate.

Antenna 2 (Fig. 9M): length 0.5 × antenna 1; flagellum 12-articulate, calceoli absent.
Figure 7. *Jesogammarus (Jesogammarus) ikiensis* sp. n., holotype, male, 13.1 mm, NSMT-Cr 24107, Ishida, Iki, Nagasaki Prefecture, Japan. **A** coxa–merus and coxal gill of pereopod 3, lateral view **B** carpus–dactylus of pereopod 3, lateral view **C** coxa–merus and coxal gill of pereopod 4, lateral view **D** carpus–dactylus of pereopod 4, lateral view **E** coxal gill of pereopod 5, lateral view **F** coxa–merus of pereopod 5, lateral view **G** carpus–dactylus of pereopod 5, lateral view.

**Gnathopod 1** (Fig. 10A): carpus length 1.7 × width, with cluster of setae and single seta on anterior margin; propodus almost as long as carpus and 1.5 × width of propodus, bearing two clusters and one pair of setae on anterior margin; palmar margin (Fig. 10B) with seven robust setae and two pectinate setae.

**Gnathopod 2** (Fig. 10C): carpus length 2.2 × width, with one cluster, one pair, and one single seta on anterior margin; propodus length 0.9 and 2.0 × carpus and width of propodus, respectively, bearing one cluster and one pair of setae on anterior margin; palmar margin (Fig. 10D) with two robust and 10 pectinate setae.

Posterior margin of bases of pereopods 5–7 more expanded than in male (Fig. 10F–H).

**Brood plates** (= oostegites) (Fig. 10E): broad, with numerous marginal setae.

**Uropod 3** (Fig. 10I): peduncle length 0.3 × outer ramus; inner ramus length 0.3 × outer ramus (both proximal and terminal articles), with robust seta on inner margin; inner margin of proximal article of outer ramus with plumose seta, terminal article length 0.2 × proximal article.

Egg number: 175.

**Variations.** The number of setae and/or setal bundles on posterior margin of peduncular articles of antennae is variable: antenna 1, two or three on article 1, three or four on article 2, one or two on article 3; antenna 2, two to four on article 4, three to
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Figure 8. Jesogammarus (Jesogammarus) ikiensis sp. n., holotype, male, 13.1 mm, NSMT-Cr 24107, Ishida, Iki, Nagasaki Prefecture, Japan. A coxa–merus of pereopod 6, lateral view B carpus–dactylus of pereopod 6, lateral view C coxal gill of pereopod 6, lateral view D coxa–merus of pereopod 6, lateral view E carpus–dactylus of pereopod 6, lateral view F coxal gill of pereopod 7, lateral view G pleopod 1, medial view, distal parts of rami omitted H retinacula on peduncle of pleopod 1, medial view I bifid plumose seta (clothes-pin seta) on inner basal margin of inner ramus of pleopod 1, medial view J uropod 1, dorsal view K uropod 2, dorsal view.

five on article 5. Most specimens have a pair of setae on dorsal margins of pleonites 1–3 but several specimens have three setae. The length ratio of inner ramus of uropod 3 to outer ramus ranged from 0.2 to 0.3 in both sexes. The number of plumose setae on inner margin of outer ramus of uropod 3 varied from two to eight in males and one to three in females. Ovigerous females have 58 to 175 eggs.

Remarks. Jesogammarus ikiensis sp. n. is assigned to the subgenus Jesogammarus in having well developed posterior accessory lobe of coxal gills on gnathopod 2 and pereopods 3–5, and pectinate setae on palmar margin of female gnathopod 2. The new species is distinguished from J. fontanus Hou & Li, 2004, J. hebeiensis Hou & Li, 2004, J. hinumensis Morino, 1993, and J. spinopalpus Morino, 1985 by absence (vs. presence) of setae on article 1 of mandibular palp. Jesogammarus ikiensis is distinguished from J. mikadoi Tomikawa, Morino & Mawatari, 2003 by absence (vs. presence) of setae on dorsal margin of pereonites 5–7 and two or three (vs. more than seven) setae on dorsal margins of pleonites 1–3. Jesogammarus ikiensis is distinguished from J. paucisetulosus Morino, 1984 by medium eye, major axis of eyes 0.4 × height of
Figure 9. Jesogammarus (Jesogammarus) ikiensis sp. n., Ishida, Iki, Nagasaki Prefecture, Japan. Holotype, male, 13.1 mm, NSMT-Cr 24107 (A–K) and paratype, female, 10.4 mm, NSMT-Cr 24108 (L and M). A uropod 3, dorsal view B telson, dorsal view C–E pleonites 1–3, respectively, dorsal views F–H urosomites 1–3, respectively, dorsal views I–K epimeral plates 1–3, respectively, lateral views L peduncular articles 1–3, accessory flagellum, and flagellar articles 1–5 of antenna 1, medial view M peduncular articles 1–5 and flagellar articles 1–3 of antenna 2, medial view.

head (vs. small, less than 0.3), posterodistal corner of peduncular article 1 of antenna 1 with a robust (vs. slender) seta, posterior margin of peduncular article 2 of antenna 1 with three or four (vs. more than five) setae and/or setal bundles, and posterior-marginal setae on peduncular article 4 of antenna 2 shorter (vs. longer) than width of
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**Figure 10.** *Jesogammarus (Jesogammarus) ikiensis* sp. n., paratype, female, 10.4 mm, NSMT-Cr 24108, Ishida, Iki, Nagasaki Prefecture, Japan. A. ischium–dactylus of gnathopod 1, lateral view B. posterodistal part of palmar margin of propodus and part of dactylus of gnathopod 1, medial view C. ischium–dactylus of gnathopod 2, lateral view D. posterodistal part of palmar margin of propodus and part of dactylus of gnathopod 2, medial view E. brood plate of gnathopod 2, lateral view F. coxa–ischium of pereopod 5, lateral view G. coxa–ischium of pereopod 6, lateral view H. coxa–ischium of pereopod 7, lateral view I. uropod 3, ventral view.
Figure 11. *Jesogammarus (Jesogammarus) ikiensis* sp. n., not preserved. **A** precopula pair (male: upper, female: lower) **B** male, approx. 13 mm. Photographed by Ryu Uchiyama.
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Jesogammarus ikiensis differs from the *J. jesoensis* complex including *J. fujinoi* Tomikawa & Morino, 2003, *J. hokurikuensis* Morino, 1985, *J. jesoensis* (Schellenberg, 1937), *J. shonaiensis* Tomikawa & Morino, 2003, by two or three (vs. more than seven) setae on dorsal margins of pleonites 1–3 and three or four (vs. two) setae and/or setal bundles on posterior margin of peduncular article 2 of antenna 1. *Jesogammarus ikiensis* differs from *J. ilhoii* Lee & Seo, 1992 by absence (vs. presence) of pectinate setae on palmar margin of propodus of male gnathopod 2 and two or three (vs. more than ten) setae on dorsal margins of pleonites 1–3.

**Etymology.** The specific name is from the Latinized Japanese *ikiensis* (of Iki), referring to the type locality of the new species.

**Distribution.** Known only from Iki Island.

**Habitat.** River and irrigation ditch.

### Key to species of Jesogamma

Since species of the *J. jesoensis* complex including *J. fujinoi*, *J. hokurikuensis*, *J. jesoensis*, *J. shonaiensis* are difficult to distinguish from each other due to high variability of morphological characters (Kusano and Ito 2003, Tomikawa unpublished data), only the *J. jesoensis* complex is included in the key. In addition, *J. naritai* Morino, 1985 is not morphologically distinguishable from *J. suwaensis* Morino, 1986 (Tomikawa et al. 2007), and the latter is treated as the same as the former in the key.

1. Accessory lobes of coxal gills on gnathopod 2 and pereopods 3–5 well developed, both anterior and posterior lobes subequal in length or posterior lobe longer than anterior one; palmar margin of propodus of female gnathopod 2 with pectinate setae....................................................... 2 (subgenus *Jesogammarus*)
   - Accessory lobes of coxal gills on gnathopod 2 and pereopods 3–5 weakly developed, anterior and posterior lobes unequal in length, often posterior lobe rudimentary; palmar margin of propodus of female gnathopod 2 without pectinate setae....................................................... 10 (subgenus *Annanogammarus*)

2. Article 1 of mandibular palp with setae.........................................................3
   - Article 1 of mandibular palp without setae..................................................6

3. Dorsal margin of pleonites 1–3 each with 1–2 setae; eye large; article 1 of mandibular palp with 1 robust seta; female pereopods densely setose..........
   - Dorsal margin of pleonites 1–3 each with more than 4 setae; eye small to medium; article 1 of mandibular palp with 2 or 3 robust setae; female pereopods not densely setose .................................................................4

4. Peduncular article 1 of antenna 1 with robust seta on posterodistal corner....
   - Peduncular article 1 of antenna 1 with slender seta on posterodistal corner...5

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*J. (J.) hinunemis Morino, 1993*
*J. (J.) spinopalpus Morino, 1985*
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5 Inner ramus of uropod 3 length 1/4 × outer ramus; inner margin of outer ramus of uropod 3 with 4–6 plumose setae.................*J. (J.) fontanus* Hou & Li, 2004
  – Inner ramus of uropod 3 length 1/3 × outer ramus; inner margin of outer ramus of uropod 3 with about 10 plumose setae.................................................................*J. (J.) hebeiensis* Hou & Li, 2004

6 Dorsal margin of pereonites 1–3 each with 2 long setae.................................................*J. (J.) mikadoi* Tomikawa et al., 2003
  – Dorsal margin of pereonites 1–3 without setae ..........................................................7

7 Posterior margin of peduncular article 2 of antenna 1 with fewer than five setae and/or setal bundles; posteromarginal setae on peduncular article 4 of antenna 2 shorter than width of article 4 in male; posterodistal corner of peduncular article 2 of antenna 1 with robust seta (occasionally lacking).........8
  – Posterior margin of peduncular article 2 of antenna 1 with more than 5 setae and/or setal bundles; posteromarginal setae on peduncular article 4 of antenna 2 longer than width of article 4 in both sexes; posterodistal corner of peduncular article 2 of antenna 1 without robust seta....................................................8
  – Posterior margin of peduncular article 2 of antenna 1 with 2 long setae; posterior margin of peduncular article 4 and 5 with more than 5 long-setal bundles .........................12

8 Dorsal margins of pleonites 1–3 each with 2 or 3 setae; posterior margin of peduncular article 2 of antenna 1 with 3 or 4 setae and/or setal bundles .......................*J. (J.) ikiensis* sp. n.
  – Dorsal margins of pleonites 1–3 each with more than 7 setae; posterior margin of peduncular article 2 of antenna 1 with 2 setae and/or setal bundles .. 9

9 Palmar margin of propodus of male gnathopod 2 without pectinate setae.....
  – Palmar margin of propodus of male gnathopod 2 with pectinate setae..............*J. (J.) jesoensis* complex

10 Dorsal margin of pleonite 3 with robust setae; posterior margin of peduncular article 4 and 5 with more than 5 long-setal bundles .................................................*J. (A.) naritai* Morino, 1985
  – Dorsal margin of pleonite 3 without robust setae; posterior margin of peduncular article 4 and 5 with less than 3 short-setal bundles .........................11

11 Posterodistal corner of bases of pereopods 5–7 with long setae ....................11
  – Posterodistal corner of bases of pereopods 5–7 without short setae .............12

12 Dorsal margins of pleonites 1–3 each with 2–4 setae ..................................................*J. (A.) fluvialis* Morino, 1985
  – Dorsal margins of pleonites 1–3 each with more than 10 setae ..................13

13 Posterodistal corner of peduncular article 1 of antenna 1 with robust seta; palmar margin of propodus of female gnathopod 2 with simple setae only ...
  – Posterodistal corner of peduncular article 1 of antenna 1 without robust seta; palmar margin of propodus of female gnathopod 2 with weakly pectinate setae..........................................................*J. (A.) debilis* Hou & Li, 2005
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