Original research

Influence of standard corneal cross-linking in keratoconus patients on macular profile

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Abstract

Purpose: To determine the effect of corneal cross-linking (CXL) on retinal structure and function.
Methods: The current study was conducted on 42 eyes of 21 patients with keratoconus (KCN) who were candidates for CXL due to disease progression. The Optovue optical coherence tomography (OCT) (Optovue Inc., Fremont, USA) from macula and multifocal electroretinography (mERG) were performed on all patients prior to surgery and at 1- and 6-month follow-up. Structural and functional parameters of macula including retinal thickness in OCT, and amplitude and latency of electroretinogram were compared between eyes that underwent surgery and control fellow eyes during the study period.
Results: A statistically significant increase in central foveal, foveal, parafoveal, and perifoveal thickness was observed at 1-month follow-up. The changes were non-significant at 6 months. Although a statistically significant reduction in amplitude and increase in latency in both rings 2 and 3 were observed at 1 month in mERG, only amplitude changes in ring 2 remained significant at 6 months.
Conclusion: Transient anatomical and functional alterations following CXL were observed in the current study.

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Keywords: Corneal cross-linking; Keratoconus; Ultraviolet; Optical coherence tomography; Phototoxicity; Multifocal electroretinography

Introduction

Keratoconus (KCN) is a bilateral, non-inflammatory, and progressive ectasia, and its characteristic findings are corneal thinning and protrusion.1 Clinical findings of the disease include progressive myopia associated with irregular astigmatism as well as characteristic finding in slit-lamp examinations.2 Its prevalence is 1 per 2000 persons, and it is the most common cause of corneal transplantation in developed countries.3,4 Therapeutic interventions for such patients range from hard contact lenses to intra-stromal corneal rings and corneal transplantation. In the meantime, the corneal cross-linking (CXL) to prevent the progression of the disease has become more popular in the last decade.

Cross-linking is a bond that links one polymer chain to another and can be a covalent or ionic bond in which the physical features of the polymer is changed.4 This procedure is extensively used in biology engineering to increase material strength and in dentistry for the hardening of filling materials.5,6 Riboflavin, also called vitamin B2, is a photosensitizer, health-essential, colored micronutrient for humans and animals. In the CXL procedure, a combination of riboflavin and ultraviolet-A (UV-A) is used to strengthen the biomechanical characteristics of the cornea. Riboflavin solely has no cross-linking property, but following absorption of UV-A energy, its electrons jump to higher level orbits, and its circular structure splits, which leads to oxygen free-radical and superoxide anions formation.7,8 These hyper-active molecules can cause cross-linking in corneal collagen, proteoglycan, and/or nucleic
acids through forming covalent bonds. For the first time in 1970, Siegel et al. described the cross-linking reaction in which the lysyl oxidase catalyzed the production of aldehyde bonds in collagen and elastin. The CXL was conducted for the first time in 2003 on human eyes; henceforward several clinical trials were conducted to evaluate its effectiveness on the prevention of KCN progression. Recent studies showed that the CXL with riboflavin drops can biochemically increase the strength and stability of cornea. The procedure can increase the intra- or inter-fibrillar cross-links following photo-sensitized oxidation resulted by exposing riboflavin as a photomediator to UV-A. In the presence of riboflavin, almost 95% of irradiated UV-A are absorbed in 300 µm of the anterior corneal stroma, and to increase its penetration, the corneal epithelium is usually removed before applying the riboflavin. Hence, according to most studies, after removing the epithelium, at least 400 µm corneal thickness is required to prevent corneal endothelial damage.

Although CXL is generally considered a safe procedure, possible side effects have been reported including delay in corneal epithelium improvement, as well as bacterial, herpetic, and Acanthamoeba keratitis, corneal edema, reduced corneal sensitivity, affecting cornea limbal stem cells by inhibiting their growth and development, stromal sterilized infiltration, corneal haziness, corneal thinning and toxicity, reduced permeability and drug penetration, postsurgical pain, persistent corneal edema, and endothelial cells damage. As most of UV-A is absorbed by the cornea and lens, possible side effects on deeper layers such as the retina have not been extensively investigated.

Although most ultraviolet-B (UV-B) radiation striking the human eye is absorbed either by the cornea or by the high level of ascorbic acid in aqueous humor, a certain proportion is able to reach the lens epithelium where it can cause damage. UV-A light is able to reach more deeply into the lens where it can react with various chromophores to generate hydroperoxide, superoxide anion, and singlet oxygen.

There is evidence that UV and short wave light may have hazardous effect on the retina, so it is necessary to investigate possible changes in the retina following CXL. Evolving evidence shows that cumulative long-term exposure to UV radiation may be harmful to the retina and possibly leads to age-related macular degeneration (AMD), irrespective of age. Although much research was conducted to discover this relationship, the cellular and molecular mechanisms of retinal phototoxicity are still unknown.

The current study aimed at evaluating the possible anatomical and functional changes in retinal and macular regions after CXL, using quantitative metrics measured by Optovue optical coherence tomography (OCT) and multifocal electroretinography (mERG).

Methods

All patients with KCN who were candidate for CXL (CCL-Vario Crosslinking; Peschke Meditrade GmbH, Zurich, Switzerland) and visited in the cornea clinic at ophthalmology center in Tabriz, Iran, during March 2015 to October 2017 were included.

All patients underwent full ophthalmology examinations, uncorrected visual acuity (UCVA), best spectacle-corrected visual acuity (BSCVA), measuring refraction and evaluation of corneal topography using a Pentacam (Oculus, Germany). Thirty patients with KCN and without corneal scar who were the candidates for cross-linking due to disease progression (0.5 diopter increase in maximum keratometry or corneal steep meridian keratometry, increased topographic astigmatism over 0.5 diopter, or at least 20 µm corneal thickness decrease during 6 months) were enrolled in the study. Lack of willingness to cooperate with the study, comorbidity of retinochoroidal disease, history of any intraocular surgery, history of laser surgery, the thinnest point of the corneal thickness <400 µm, crystalline lens opacity, severe dry eye, autoimmune systemic diseases, and need for cross-linking in the opposite eye within the 6-month follow-up period were the exclusion criteria.

The protocol of cross-linking used in the current study was the standard protocol, called the Dresden Protocol, as described elsewhere. Briefly, after local anesthesia with ocular topical anesthetics (Tetracaine hydrochloride ophthalmic solution 0.5%), central 8 mm epithelial zone was removed. After application of 0.1% riboflavin-5'-phosphate and 20% dextran solution on the epithelium free surface every 3–5 min for 30 min, UV-A irradiation (370 nm, 3 mW/cm²) during the 30 min alongside the application of the aforementioned solution for every 5 min was performed. At the end of surgery, topical antibiotics were administered and bandage soft contact lens were placed.

Optovue OCT (Optovue Inc., Fremont, USA) and mERG (Diagnosys LLC, Littleton, MA, USA) of macula were performed for all patients before CXL and repeated in 1 and 6 months after surgery. The macular parameters including macular thickness in central fovea (center point of fovea), fovea (the central 1 mm of macula), parafovea (the area located between fovea and perifovea with a radius of 3 mm), and perifovea (outermost area outside the parafovea with a radius of 5 mm) regions were separately measured in full, inner, and outer retinal maps (according to the analysis and calculations of the Optovue OCT). The segmentation was performed automatically by the device software. The mERG amplitude and latency changes were extracted up to 20° for 5 pericentral rings around the central fovea, by the mERG, and compared to fellow control eyes of the same patients. All images were provided under the constant condition using the same software. The electroretinography was performed monocularly, after maximally dilation of pupils with 1% tropicamide eye drops (after 30 min) and 15 min of adaptation in a bright room based on the International Society for Clinical Electrophysiology of Vision (ISCEV) guidelines. To reduce patients' discomfort, the anesthetic eye drop (Anestocaine; Sina Darou, Tehran, Iran) was used before tests.

In all stages, the patients' BSCVA was measured using the Snellen chart and exchanged to the logarithm of the minimum of angle resolution (logMAR) for statistical analyses.
Finally, the collected data were analyzed with IBM SPSS software (version 13; SPSS, Chicago, IL) using the paired samples $t$ test, independent $t$ test, and the Wilcoxon and the Mann-Whitney U tests. Data obtained from macular thickness had normal distribution in all full, inner, and outer retinal maps whereas based on mERG findings, the amplitude and latency in both groups had non-normal distribution. Hence, a non-parametric test was used.

The aims and objectives of the study as well as the follow-up procedure were explained to the patients, and they were enrolled into the study by interest, and informed consents were obtained. This study was approved by the review board ethics committee of the training hospital and Tabriz University of Medical Sciences, Tabriz, Iran.

**Results**

A total of 30 patients who met the inclusion criteria were enrolled into the study and underwent corneal standard cross-linking based on the evidence of KCN progression. The opposite eyes of the patients were followed up as the control group. Out of the enrolled patients, 5 subjects withdrew from the study due to lack of interest, and 4 patients were excluded due to the need for cross-linking to the opposite eye. Finally, 42 eyes from 21 patients with KCN were studied. 12 (57.1%) patients were male, and 9 (42.9%) were female. The mean age of the patients was 20.9 ± 8.7 years, ranging from 10 to 32. Nine patients (42.9%) and 12 (57.1%) underwent CXL on their right and left eyes based on clinical indication, respectively. The means of UCVA and BSCVA for the cross-linked eyes were 0.27 ± 0.08 and 0.72 ± 0.16, respectively, and in the opposite eyes, they were 0.30 ± 0.10 and 0.81 ± 0.17, respectively. Visual changes in the cross-linked eyes in 1- and 6-month follow-ups were statistically insignificant ($P = 0.073$ and 0.094, respectively).

The mean thickness of macula in central fovea, fovea, parafovea, and perifovea before surgery in the eyes that underwent cross-linking were 213.14 ± 18.56 (ranged 182–265), 251.28 ± 20.12 (218–291), 325.38 ± 11.72 (307–352), and 295.61 ± 11.38 (280–318) micro-meters (μm), respectively; the means in the control eyes were 214.2 ± 25.59 (183–281), 252.55 ± 21.93 (217–293), 324.9 ± 17.46 (302–353), and 297.1 ± 16.97 (276–327) μm, respectively.

In addition, the mean inner retinal thickness in central fovea, fovea, parafovea, and perifovea regions before surgery in the eyes that underwent cross-linking were 29 ± 18.24 (6–71), 65.9 ± 11.66 (51–91), 126.95 ± 9.21 (111–142), and 111.35 ± 5.68 (103–125) μm, respectively. The means in the control eyes were 32.5 ± 22.19 (13–86), 68.95 ± 11.48 (50–91), 127.15 ± 15.11 (88–146), and 110.4 ± 9.88 (85–125) μm, respectively.

The mean outer retinal thickness in the central fovea, fovea, parafovea, and perifovea regions before surgery in the eyes that underwent cross-linking were 184.1 ± 10.30 (164–207), 180.5 ± 23.46 (117–206), 199.15 ± 8.79 (174–210), and 184.15 ± 7.80 (173–196) μm, respectively; the means in the control eyes were 183.75 ± 15.28 (158–207), 185.5 ± 11.29 (167–202), 198.1 ± 18.45 (179–234), and 186.9 ± 17.63 (168–217) μm, respectively (Table 1).

### Table 1

Macular thickness in central fovea, fovea, parafovea, and perifovea separated in full/inner/outer retina before corneal cross-linking (CXL), 1- and 6-month follow-up in both eyes.

|                         | CXL Before | CXL 1 m | CXL 6 m | Control Before | Control 1 m | Control 6 m | P value |
|-------------------------|------------|---------|---------|----------------|-------------|-------------|---------|
| **Full retinal map**    |            |         |         |                |             |             |         |
| Foveal thickness        | 251.28 (20.12) | 254.28 (24.47) | 250.52 (20.29) | 252.55 (21.93) | 253.80 (19.38) | 254.20 (20.45) | 0.052   |
| Central foveal thickness| 213.14 (18.56) | 214.76 (21.05) | 212.85 (18.61) | 214.20 (25.59) | 219.60 (31.75) | 213.60 (25.94) | 0.251   |
| Parafoveal thickness    | 325.38 (11.72) | 325.52 (16.91) | 318.76 (23.66) | 324.90 (17.46) | 327.30 (12.60) | 327.65 (13.08) | 0.934   |
| Perifoveal thickness    | 295.61 (11.38) | 297.76 (11.43) | 295.95 (12.00) | 297.10 (16.97) | 297.70 (16.29) | 297.55 (15.97) | 0.163   |
| **Inner retinal map**   |            |         |         |                |             |             |         |
| Foveal thickness        | 65.90 (11.66) | 70.75 (12.34) | 65.45 (12.00) | 68.95 (11.48) | 70.35 (11.31) | 69.60 (10.77) | 0.000   |
| Central foveal thickness| 29.00 (18.24) | 35.60 (15.68) | 29.60 (16.69) | 32.50 (22.19) | 32.60 (20.58) | 32.80 (20.36) | 0.019   |
| Parafoveal thickness    | 126.95 (9.21) | 135.35 (7.52) | 129.15 (8.39) | 127.15 (15.11) | 128.20 (16.13) | 127.10 (15.17) | 0.000   |
| Perifoveal thickness    | 111.35 (5.68) | 117.05 (4.41) | 112.30 (6.03) | 110.40 (9.88) | 110.85 (10.85) | 110.45 (10.27) | 0.000   |
| **Outer retina map**    |            |         |         |                |             |             |         |
| Foveal thickness        | 180.50 (23.46) | 185.70 (13.29) | 185.30 (9.52) | 185.50 (11.29) | 186.65 (10.51) | 186.15 (9.85) | 0.250   |
| Central foveal thickness| 184.10 (10.30) | 180.05 (17.38) | 182.80 (11.78) | 183.75 (15.28) | 179.65 (17.64) | 180.80 (17.00) | 0.300   |
| Parafoveal thickness    | 199.15 (8.79) | 194.65 (14.95) | 195.35 (9.79) | 198.10 (18.45) | 198.30 (15.06) | 197.60 (16.21) | 0.173   |
| Perifoveal thickness    | 184.15 (7.80) | 183.45 (11.45) | 182.80 (7.40) | 186.90 (17.63) | 186.40 (15.16) | 186.41 (16.58) | 0.712   |

Central fovea (center point of fovea), fovea (the central 1 mm of macula), parafovea (the area located between fovea and parafovea with a radius of 3 mm), and perifovea (the outermost area outside the parafovea with a radius of 5 mm).

CXL: Corneal cross-linking, Thickness unit: μm.

Numbers are Mean ± SD.
In the analysis of macular thickness changes, the results of the paired samples $t$ test showed no significant changes in central fovea, fovea, parafovea, and perifovea regions of full and outer retinal maps in the intervention group before and after cross-linking. Nevertheless, in 1-month follow-up, the changes in each of the 4 regions showed significant changes in the inner retinal map assessments. The macular thickness in inner retinal map for central fovea, fovea, parafovea, and perifovea regions increased from $29 \pm 18.24, 65.9 \pm 11.66, 126.95 \pm 9.21, 111.35 \pm 5.68$ to $35.6 \pm 15.68$ ($P = 0.019$), $70.75 \pm 12.34$ ($P < 0.000$), $135.35 \pm 7.52$ ($P < 0.000$), and $117.05 \pm 4.41$ mm ($P < 0.000$), respectively. The increasing trend of macular thickness was not observed in 6-month follow-up examinations, and the changes in macular thickness in all evaluated regions were statistically insignificant. Also, macular thickness the control eyes in 1- and 6-month follow-ups showed no clinically significant changes compared to the baseline values (Table 1).

Data obtained from mERG were assessed as mean amplitude and latency in hexagonal regions of 5 concentric rings (Fig. 1). The mERG findings showed that the mean amplitude in 5 concentric rings around central fovea in the eyes that underwent cross-linking were $22.67 \pm 19.79, 45.50 \pm 18.59, 26.22 \pm 13.65, 17.87 \pm 8.42$, and $15.08 \pm 7.99$ nV/deg$^2$, respectively, and $22.31 \pm 19.50, 44.76 \pm 21.83, 28.14 \pm 12.44, 18.58 \pm 9.51$, and $16.66 \pm 7.51$ nV/deg$^2$ in the control eyes, respectively. Also, mean latency in 5 concentric rings around central fovea in the CXL eyes were $38.12 \pm 7.63, 34.52 \pm 4.14, 34.22 \pm 1.99, 36.76 \pm 5.02$, and $35.39 \pm 0.95$ millisecond (ms), respectively, while it was $37.32 \pm 11.81, 34.49 \pm 3.06, 35.02 \pm 4.18, 35.58 \pm 2.64$, and $36.42 \pm 2.19$ ms in the control eyes, respectively.

Baseline values were similar statistically before surgery between the groups (Table 2).

In functional evaluations of macula by the mERG, the amplitude and latency measures of the rings 2 and 3 showed statistically significant changes at 1-month follow-up, compared with the pre-surgical measures. Changes in other rings were statistically insignificant in the 1-month follow-up assessments. The mean amplitude in the ring 2 reduced from $45.50 \pm 18.59$ in pre-surgical evaluations to $25.25 \pm 21.52$ nV/deg$^2$ ($P = 0.011$) and in the ring 3 from $26.22 \pm 13.65$ to $14.20 \pm 10.93$ nV/deg$^2$ ($P = 0.033$) in the 1-month follow-up. In addition, the mean latency increased significantly in the ring 2 from $34.52 \pm 4.14$ in pre-surgical assessments to $38.67 \pm 8.36$ ms ($P = 0.047$) and in the ring 3 from $34.22 \pm 1.99$ to $37.62 \pm 6.04$ ms ($P = 0.008$) in the 1-month follow-up.

Also, the amplitude changes in the ring 2 were statistically significant in the 6-month follow-up, while this parameter showed no significant changes in other regions. The mean amplitude in the ring 2 reduced from $45.50 \pm 18.59$ in
Discussion

CXL is a therapeutic option for progressive KCN.14 As with other new treatments, its safety is of particular importance. Previous studies have emphasized that in combination with riboflavin and in the presence of a minimal corneal thickness of 400 μm, insignificant amounts of UV-A radiance reach inner ocular tissues, including the lens and retina, and that there is no evidence of UV-A induced damage in these tissues during CXL procedure.35 Recent in vitro studies have revealed that corneal endothelial damage due to UV exposure is a possible risk.36 Furthermore, some in vivo reports have raised concerns for corneal endothelial damage after standard CXL procedure.15,37 Therefore, it is possible that other intraocular complications occur after CXL.

The adverse effects of UV radiation on the eye have been suspected and recently have been studied in the field of epidemiology. Some studies evaluated the phototoxic effects of UV-A radiation on intraocular structures such as crystalline lens. Several in vivo and in vitro studies also considered UV light as a risk factor of lens damage and cataract. Different hypotheses also justified such toxic effects by the induced oxidative stress.38 Azzam et al. reported that UV-A can imitate the senile cataract morphology and make changes in lens optics by damaging the lens cell membrane.39 Another study by Hedge et al. indicated UV light as one of the most important risk factors for the induction of cataract. They also added that this effect is probably formed though the creation of oxygen free-radicals in aqueous humor and crystalline lens.40 However, a recent study on the intraocular complications of CXL evaluated the changes of crystalline lens density following the CXL procedure in patients with KCN and reported no significant changes in the density of crystalline lens following the procedure compared with the control group, based on the Pentacam Scheimpflug imaging system evaluations.41

Based on the extensive literature review conducted, this is, to the best of our knowledge, one of the few studies investigating CXL adverse event on macular profile among patients with progressive KCN. The study findings showed that the CXL had some transient changes that may be due to toxicity on macula, and alterations mostly involved inner areas of the retina. However, retinal changes were eliminated over time, and only small retinal functional changes were observed in 6-month follow-up evaluations.

Light toxicity frequently affects outer retina and retinal pigment epithelium (RPE) early after exposure. Chromophores are theorized to mediate the light-induced damage to the retina. Chromophores usually exist in the retina and RPE, namely the photoreceptors, flavoproteins, heme proteins, melanosomes, and lipofuscin. Therefore, RPE and outer retina are expected to be influenced more than inner retina.42 However, in our study, the micro-anatomical changes were more profound in inner layers. On the other hand, it should be noted that presence of astigmatism influenced the OCT measurements, and corneal characteristics were changed following CXL that had been taken approximately 3 months or more to be stabilized.43 So, corneal characteristics alterations might explain the micro-anatomical changes of inner retina in the OCT. It should be noted that the lack of examination of corneal characteristic changes, as one of the important contributing factors of OCT analysis, was one of our study disadvantages.

pre-surgical assessments to 32.95 ± 19.38 ms (P = 0.028) in the 6-month follow-up. The latency changes were insignificant in all under study regions in the 6-month follow-up (Table 2).

In the 1- and 6-month follow-ups, no significant changes were observed in the control eyes (Table 2).

### Table 2

Amplitude and latency of multifocal electroretinography (mERG) in 1–5 rings before corneal cross-linking (CXL), 1- and 6-month follow-up in both eyes.

| Ring1 | Before | 1 m | 6 m | Control | Before | 1 m | 6 m |
|-------|--------|-----|-----|---------|--------|-----|-----|
| Latency | 38.12 (7.63) | 38.14 (7.36) | 37.36 (7.03) | 37.32 (11.81) | 34.55 (9.17) | 37.70 (11.28) | 1.000 | 0.157 | 0.055 | 0.910 |
| Amplitude | 22.67 (19.79) | 21.05 (20.01) | 22.31 (19.50) | 23.96 (15.83) | 25.29 (16.70) | 0.747 | 0.889 | 0.423 | 0.074 |

| Ring2 | Before | 1 m | 6 m | Control | Before | 1 m | 6 m |
|-------|--------|-----|-----|---------|--------|-----|-----|
| Latency | 34.52 (4.14) | 38.67 (8.36) | 35.90 (4.88) | 34.49 (3.06) | 34.06 (3.20) | 33.87 (2.98) | 0.047 | 0.309 | 0.604 | 0.204 |
| Amplitude | 45.50 (18.59) | 25.25 (21.52) | 32.95 (19.38) | 44.76 (21.83) | 45.80 (21.02) | 45.70 (25.42) | 0.011 | 0.028 | 0.959 | 0.950 |

| Ring3 | Before | 1 m | 6 m | Control | Before | 1 m | 6 m |
|-------|--------|-----|-----|---------|--------|-----|-----|
| Latency | 34.22 (1.99) | 37.62 (6.04) | 35.47 (2.76) | 35.02 (4.18) | 34.55 (4.41) | 35.57 (4.19) | 0.008 | 0.009 | 0.183 | 0.339 |
| Amplitude | 26.22 (13.65) | 14.20 (10.93) | 17.92 (12.22) | 28.14 (12.44) | 25.95 (12.93) | 26.28 (14.53) | 0.033 | 0.115 | 0.120 | 0.210 |

| Ring4 | Before | 1 m | 6 m | Control | Before | 1 m | 6 m |
|-------|--------|-----|-----|---------|--------|-----|-----|
| Latency | 36.76 (5.02) | 39.04 (7.34) | 36.68 (5.98) | 35.58 (2.64) | 35.31 (1.95) | 35.51 (2.27) | 0.054 | 0.575 | 0.871 | 0.837 |
| Amplitude | 18.75 (8.42) | 10.97 (8.11) | 13.03 (7.63) | 18.58 (9.51) | 18.83 (10.42) | 18.53 (10.15) | 0.107 | 0.233 | 0.938 | 0.950 |

| Ring5 | Before | 1 m | 6 m | Control | Before | 1 m | 6 m |
|-------|--------|-----|-----|---------|--------|-----|-----|
| Latency | 35.39 (0.95) | 35.35 (3.57) | 35.93 (2.51) | 36.42 (2.19) | 36.50 (2.35) | 36.23 (1.79) | 0.652 | 0.917 | 0.910 | 0.200 |
| Amplitude | 15.08 (7.99) | 10.07 (8.06) | 9.91 (7.89) | 16.66 (7.51) | 16.32 (8.37) | 16.18 (8.35) | 0.126 | 0.393 | 0.255 | 0.203 |

CXL: Corneal cross-linking, mERG: Multifocal electroretinography, Amplitude unit: nV/deg², Latency unit: ms. Numbers are Mean ± SD.
The small sample size was one of the limitations of the current study. On the other hand, light toxicity starts early after exposure, so early changes after CXL may remain undetectable. Further studies on larger populations and more frequent evaluations in shorter intervals after the CXL are recommended. Future studies may use adaptive optics to show ultrastructural changes. Also, the complementary tests for more anatomical evaluations of the macula such as ganglion cell complex thickness, and functional evaluations such as microperimetry, color vision test (CVT), macular autofluorescence pigment density (MAFPD), and photostress recovery time (PSRT) may show the retinal changes after CXL.

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