Periventricular heterotopia and white matter abnormalities in a girl with mosaic ring chromosome 6

Satsuki Nishigaki1, Takashi Hamazaki1, Mika Saito1, Toshiyuki Yamamoto2, Toshiyuki Seto1 and Haruo Shintaku1

Abstract

Ring chromosome 6 is a rare chromosome abnormality that arises typically de novo. The phenotypes can be highly variable, ranging from almost normal to severe malformations and neurological defects. We report a case of a 3-year-old girl with mosaic ring chromosome 6 who presented with being small for gestational age and intellectual disability, and whose brain MRI later revealed periventricular heterotopia and white matter abnormalities. Mosaicism was identified in peripheral blood cells examined by standard G-bands, mos 46,XX,r(6)(p25q27)[67]/45,XX,-6[25]/46,XX,dic r(6q)[(p25q27;p25q27)][6]/47,XX,r(6q) × 2[2]. Using array-comparative genomic hybridization, we identified terminal deletion of 6q27 (1.5 Mb) and no deletion on 6p. To our knowledge, this is the first report of periventricular heterotopia and white matter abnormalities manifested in a patient with ring chromosome 6. These central nervous system malformations are further discussed in relation to molecular genetics.

Keywords: Ring chromosome 6, Brain MRI, Periventricular heterotopia, White matter abnormality, Array-comparative genomic hybridization, Small for gestational age

Background

Ring chromosome 6 is a rare chromosome abnormality that arises typically de novo [1]. The phenotypes can vary from almost normal to severe malformations and mental defects [2]. The most common clinical features include intellectual disability, microcephaly, prenatal growth failure, retarded bone age, short neck, and typical facial anomalies [3]. As for brain abnormalities, hydrocephalus is the most important prenatal feature [4]. In addition to cortical atrophy and ventriculomegaly, absence of olfactory tract, agenesis or hypoplasia of the corpus callosum, aqueduct stenosis, and neuronal migration defects are also observed in previous reports [5].

Here, we report a case of mosaic ring chromosome 6 in a 3-year-old girl who presented with failure to thrive, microcephaly, and developmental delay and whose brain magnetic resonance imaging (MRI) later revealed periventricular heterotopia and white matter abnormalities. Neither of these central nervous system anomalies has been previously reported in association with ring chromosome 6.

Case presentation

Clinical description

The patient, a female, is the second child of healthy unrelated parents. Family history, including her 4-year-old sister, was unremarkable. This patient was born by means of cesarean delivery because of intrauterine growth retardation at the 39th week of gestation. Birth weight was 1,785 g (−2SD), length was 43 cm (−2SD) and head circumference was 29 cm (−2SD). Apgar scores were 8 and 9 at 1 and 5 minutes, respectively. She was able to sit at 5 months, walk at 14 months. Although at age 3 years she couldn’t speak in two-word sentences, at age 3 years and 11 months she was able to speak three-word sentences with postpositions. She had a history of repeated febrile seizures.

At the age of 3 years and 11 months, her weight was 9,300 g (−3SD), length 89.7 cm (−2.4SD) and head circumference 41.3 cm (−4.4SD). Short stature prompted endocrinological investigations and karyotyping. Exaggerated growth hormone responses to arginine, levodopa and
clonidine stimulation were within normal range, whereas karyotype analysis revealed mosaic ring chromosome 6 (Fig. 1). She presented minor dysmorphic features including full lips and epicanthal folds. Ophthalmological and otological including aural examination were normal, and ultrasound cardiac examination revealed no anomalies. Kyoto Scale of Psychological Development, which is a standardized developmental test for Japanese children, was performed at 3 years 10 months, revealing postural-motor skills of 3 years 1 month (subquotient 79), cognitive-adaptive skills of 2 years 6 months (subquotient 64), language-social skills of 2 years 9 months (subquotient 70), and overall developmental age of 2 years 8 months (developmental quotient 68).

Cerebral MRI revealed periventricular heterotopia on the bilateral posterior horn of the lateral ventricle and white matter abnormalities in the bilateral parietal and occipital lobe. Neither hydrocephalus nor enlarged lateral ventricles were detected (Fig. 2). Electroencephalogram (EEG) during induced sleep showed poor normal sleep spindles and slightly irregular background activity in the bilateral occipital head regions. The possibility of tuberous sclerosis was also considered with respect to the observed periventricular nodules. Besides the past episodes of febrile seizure, she never had an epileptic attack, and anticonvulsant agent was never prescribed, ruling out the possibility of tuberous sclerosis. This study was approved by institutional review board (Osaka City University).

Molecular and cytogenomic characterization
This study was conducted following approval by the ethics committee at our institution. After obtaining the written informed consent from the patient’s family, blood sample was drawn from the patient. Chromosome analysis was performed on peripheral blood lymphocyte cultures. Standard G-bands analysis revealed mos 46,XX, r(6)(p25q27)[67]/45,XX,-6[25]/46,XX,dic r(6;6)(p25q27; p25q27)[6]/47,XX,r(6)(p25q27) × 2[2]. The microarray-based comparative genomic hybridization (array-CGH) analysis was performed as described previously [6] and revealed a genomic copy number loss at 6q27 (Fig. 3).

Discussion
In general, ring chromosomes are formed by fusion of the distal ends in both arms and often lose genomic material of the terminal ends. The rings may result in clinical features mimicking terminal deletion syndromes. On the other hand, a patient carrying a ring chromosome 6 without terminal deletions has been reported with short stature and minor dysmorphisms but with normal psychomotor development [7]. This indicates that carrying a ring chromosome itself may lead to these clinical features. These prenatal and postnatal growth failures, also seen in the present case, are common features of any autosome ring chromosome. It has been suggested that this is due to the mitotic instability of rings, preventing normal cellular proliferations [8, 9].
Fig. 3 The result of array-CGH. Vertical and horizontal axes indicate the genomic location and signal log_2 ratio, respectively. Dots indicate the location and the signal log_2 ratio of the microarray probes. a Chromosomal view of chromosome 6 shows a genomic copy number loss at q27. b Gene view expands the aberration region. The blue rectangle indicates and the blue bars indicate the aberration region and the locations of the genes. Two genes discussed in the text are marked by red oblongs. The mean log_2 ratio of the aberration region is −0.780567, indicating mosaic deletion.

Fig. 2 a-f: MRI imaging of the brain of this patient. a and b Axial T1W and T2W image, showing left parietal lobe. c Coronal T2W section showing white matter abnormalities in the bilateral parietal lobe. d Coronal T2W section showing white matter abnormalities in the bilateral occipital lobe. e and f Axial and Coronal FLAIR image, showing periventricular heterotopia on the bilateral posterior horn of the lateral ventricle.
Terminal deletion on 6q has been linked to certain brain abnormalities as well as neurological signs and symptoms. Based on a series of the array-CGH analysis, Eash et al. have suggested that the terminal region of chromosome 6q is a critical area for intellectual disability, hydrocephalus, abnormal corpus callosum, seizures, and hypotonia [10]. In the present case where the terminal of 6q27 was deleted, none of those clinical features was present except for intellectual disability.

A few studies have investigated the effects of terminal deletion on 6p. In an adult case report, chromosome 6p subtelomeric deletions were thought to be a possible underlying cause for periventricular white matter abnormalities [11]. Another study found that the most consistent clinical features in the patients with 6p25 deletion included developmental delay, intellectual disability, language impairment, hearing loss, ophthalmologic, cardiac, and craniofacial abnormalities [12]. In the present case where 6p25 was intact, the patient presented with developmental delay, intellectual disability, and language impairment but had neither hearing loss nor ophthalmologic/cardiac abnormalities.

C6orf70 is a gene that maps on 6q27 and is one of the deleted genes in this patient (Fig. 3). Haploinsufficiency of C6orf70 is surmised to play a major role in the pathogenesis of periventricular heterotopia in this patient. Haploinsufficiency or mutation of C6orf70, has recently been suggested as the main cause of the brain malformation complex including periventricular heterotopia [13]. C6orf70 is expressed in developing human brain and is involved in neuronal migration. However, in the previous reports of ring chromosome 6 with 6q27 deletions including C6orf70, periventricular heterotopia has not been pointed out [5, 14].

Regarding the white matter abnormality found in this patient, contiguous gene deletions on 6q27 may play a role. Contiguous gene deletion of PHF10 and DLL1 could be an underlying mechanism of the white matter abnormality observed in this patient (Fig. 3). Conti et al. experimentally demonstrated that down-regulation of PHF10 and DLL1 expression in utero delayed neuronal migration in a rodent model [13]. Additionally, RNA-SET2, which is also mapped within the deleted region in the present case (Fig. 3), was reported to have an association with familial cystic leukoencephalopathy [15]. RNA-SET2 encodes a putative lysosomal hydrase and the loss of function in zebrafish caused accumulation of amyloid precursor protein in the brain [16].

Conclusions
This is the first report of periventricular heterotopia and white matter abnormalities manifested in a patient with ring chromosome 6. Further accumulation of cases with similar deletion in this region of chromosome 6 will identify the genes responsible for this neurological phenotype.

Consent
Written informed consent was obtained from the patients of the patient for publication of this Case report and any accompanying images. A copy of the written consent is available for review by the Editor-in-Chief of this journal.

Abbreviations
MRI: Magnetic Resonance Image; array-CGH: Microarray-based comparative genomic hybridization; EEG: Electroencephalogram.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
MS and TS referred the patient for this study and coordinated the clinical analysis of the patient. SN, TH and TS were responsible for the patient’s clinical genetic examination. MS and HS contributed to the clinical description. TY performed the array CGH and analyzed the data. All authors co-wrote the manuscript. TH and TY revised the manuscript critically for important intellectual content. All authors read and approved the final manuscript.

Author details
1. Department of Pediatrics, Osaka City University Graduate School of Medicine, 1-4-3 Asahi-machi, Abeno-ku, Osaka 545-8858, Japan. 2. Institute for Integrated Medical Sciences, Tokyo Women’s Medical University, 8-1 Kawada-cho, Shinjuku-ward, Tokyo 162-8666, Japan.

Received: 16 April 2015 Accepted: 12 July 2015
Published online: 26 July 2015
13. Conti V, Carabalona A, Pallesi-Pocachard E, Parini E, Levenger RI, Buhler E, et al. Periventricular heterotopia in 6q terminal deletion syndrome: role of the C6orf70 gene. Brain. 2013;136:3378–94.

14. Ciocca L, Surace C, Digilio MC, Roberti MC, Sirleto P, Lombardo A, et al. Array-CGH characterization and genotype-phenotype analysis in a patient with a ring chromosome 6. BMC Med Genomics. 2013;6:3.

15. Henneke M, Diekmann S, Ohlenbusch A, Kaiser J, Engelbrecht V, Kohlschüttner A, et al. RNASET2-deficient cystic leukoencephalopathy resembles congenital cytomegalovirus brain infection. Nat Genet. 2009;41:773–5.

16. Haud N, Kara F, Diekmann S, Henneke M, Willer JR, Hillwig MS, et al. RNASET2 mutant zebrafish model familial cystic leukoencephalopathy and reveal a role for RNase T2 in degrading ribosomal RNA. Proc Natl Acad Sci U S A. 2011;108:1099–103.