The associations between TMAO-related metabolites and blood lipids and the potential impact of rosuvastatin therapy

Xiaowei Xiong1†, Jian Zhou2†, Qiang Fu1, Xiaowei Xu1, Shaobin Wei1, Shenghua Yang1 and Buxing Chen3*

Abstract

Background: Trimethylamine N-oxide (TMAO)-related metabolites are associated with the pathogenesis of atherosclerotic cardiovascular disease (ASCVD) and are known to disrupt lipid metabolism. The aims of this study were to evaluate the associations between TMAO-related metabolites and blood lipids and determine how lowering the lipid profile via rosuvastatin therapy influences TMAO-related metabolites.

Methods: A total of 112 patients with suspected ASCVD were enrolled in this study. The levels of plasma TMAO-related metabolites, including TMAO, choline, carnitine, betaine, and γ-butyrobetaine (GBB), were analyzed by stable isotope dilution liquid chromatography-tandem mass spectrometry (LC/MS/MS) before and after rosuvastatin therapy in all patients. Statistical methods were used to detect the associations between TMAO-related metabolites and blood lipids and determine how rosuvastatin therapy alters the levels of these metabolites.

Results: A significant positive correlation was found between TMAO and triglycerides (TG) ($r = 0.303$, $p < 0.05$). Furthermore, significant negative correlations were found between TMAO and high-density lipoprotein cholesterol (HDL-c) and between betaine and low-density lipoprotein cholesterol (LDL-c) ($r = -0.405$ and $-0.308$, respectively, both $p < 0.01$). Compared to baseline, significantly lower TMAO levels and higher carnitine, betaine and GBB levels were observed after rosuvastatin therapy, while the lipids decreased significantly ($p < 0.05$). The significant correlation between TMAO and TG or between betaine and LDL-c disappeared after rosuvastatin therapy ($r = 0.050$ and $-0.172$, respectively, both $p > 0.05$). However, a significantly positive association between carnitine and TC and a negative association between carnitine and LDL-c or between betaine and TG were found after adjustment for sex, age, body mass index (BMI) and lipids ($p < 0.05$).

Conclusions: This study suggests that TMAO-related metabolites are significantly associated with blood lipids, although some of them are changed postrosuvastatin therapy. Lower TMAO and higher TMAO precursors were observed after rosuvastatin therapy compared to baseline. This study indicates that elevated TMAO precursors after rosuvastatin therapy and their potential impact on ASCVD should be considered in the clinic.

Keywords: Trimethylamine N-oxide, Atherosclerotic cardiovascular disease, Rosuvastatin, Blood lipids

Introduction

Trimethylamine N-oxide (TMAO) is an oxidative product of trimethylamine (TMA), which is generated from choline, choline-containing substances, L-carnitine, γ-butyrobetaine (GBB) and betaine by the gut microbiota [1–3]. TMAO and its related precursors mentioned above are collectively termed TMAO-related...
metabolites. Many studies have found that TMAO-related metabolites are associated with the formation and development of atherosclerotic cardiovascular disease (ASCVD) [1–4], and elevated TMAO-related metabolite levels are associated with a heavy atherosclerotic burden, poor prognosis of ASCVD and a high risk of major adverse cardiovascular events (MACEs) [5–8]. Atherosclerosis is a type of chronic metabolic disease that is characterized by chronic inflammation and lipid metabolism disorders [9]. The associations between TMAO-related metabolites and inflammation have been studied and demonstrated in cultured cells, mouse models, and even clinical trials [10–13]. Moreover, some studies also found that the compositions of the gut microbiota are associated with variations in blood lipids [14, 15], and TMAO-related metabolites, such as TMAO, carnitine and GBB, could disrupt cholesterol metabolism in animal models [2, 4, 16]. Although a previous study found that plasma betaine was negatively associated with lipids, such as low-density lipoprotein cholesterol (LDL-c) and triglycerides (TG) [17], there is still little information about the relationships between TMAO-related metabolites and blood lipids. In addition, as an effective lipid-lowering drug, rosuvastatin was found to influence gut microbiota composition and the function of genes related to the metabolism of TMAO-related metabolites [18, 19]. However, information about its effect on TMAO-related metabolites is still ambiguous. Therefore, the aims of this study were to investigate the associations between TMAO-related metabolites and blood lipids and examine the potential impact of rosuvastatin therapy on these metabolites and associations.

Methods

Study population

This is a prospective study based on a registered clinical trial at http://clinicaltrials.gov/ (No.02305862). Patients with suspected ASCVD who were willing to receive coronary angiography and carotid magnetic resonance imaging (MRI) were consecutively recruited from Beijing Tiantan Hospital between January 2013 and December 2016. The demographic characteristics, personal history and diagnosis were based on personal statements, clinical records, current drug usages, and auxiliary examinations in the hospital. The exclusion criteria were age < 18 years, stent implantation, uncontrolled hypertension, acute myocardial infarction, malignant tumor, heart failure, active infections, severe liver or kidney dysfunction, stroke, diabetes mellitus (DM), familial hyperlipidemia, and antibiotic usage. A total of 135 patients were recruited in this study, including 35 patients who received irregular statin therapy for less than 1 month and had a clearance period of at least 2 weeks. A total of 121 patients received rosuvastatin therapy because of hyperlipidemia, ASCVD, and/or other proper purposes. All patients were initially administered 20 mg rosuvastatin per night, and 4 of these patients were adjusted to 10 mg because of impaired liver function during the follow-up time. No patients suspended usage during the follow-up. All patients were given a recommended light diet before sampling and after discharge and monitored half-monthly at the outpatient department or by telephone interviews. Finally, 112 patients with complete follow-up data after 3 months of follow-up constituted the study cohort. The study flowchart is displayed in Fig. 1, and the baseline characteristics of the cohort are presented in

![Flow chart](Fig. 1)
Supplementary Table S1. This study abided by the principles of the Declaration of Helsinki and was approved by the Ethics Committee of Beijing Tiantan Hospital (Approval No: KY2014-020-02). All patients in this study signed informed consent forms. The data can be obtained from the corresponding author after the permission of the Ethics Committee of Beijing Tiantan Hospital.

Statistical analyses
Categorical variables were compared by the chi square test and expressed as frequencies (n) and percentages (%). Normality of continuous variables was evaluated using a one-sample Shapiro–Wilk test. Student’s t test was performed to compare the differences between groups, or the Mann–Whitney test was used when variables were not normally distributed. A paired test was used to compare the differences in lipids and TMAO-related metabolites before and after rosuvastatin therapy. All data are expressed as the mean ± standard deviation (SD). Pearson analyses were performed to evaluate the correlations between TMAO-related metabolites and lipids, and Spearman analyses were performed when variables were not normally distributed. Partial correlation analyses were performed to explore the correlations between TMAO-related metabolites and lipids by adjusting for sex, age, body mass index (BMI) and estimated glomerular filtration rate (eGFR). Multivariate analyses were performed using a linear regression model to identify the associations between TMAO-related metabolites and lipids and investigate the effect of rosuvastatin therapy on TMAO-related metabolites. The variables included in the multivariate analyses were sex, age, BMI, eGFR, TG, TC, LDL-c, HDL-c, ApoA1 and ApoB. The statistical analyses were performed using SPSS version 17.0 (SPSS Inc., Chicago, IL). A P value < 0.05 was considered statistically significant.

Results
Baseline characteristics
In this study, female patients with suspected ASCVD were older (64.57 ± 6.94 vs. 60.34 ± 9.77 years, P < 0.05) and had lower rates of smoking (12.8% vs. 75.4%, P < 0.05) and drinking (2.1% vs. 41.5%, P < 0.05) than male patients (Supplementary Table S2), and these results are consistent with epidemiological characteristics. All patients were divided into low-TMAO and high-TMAO groups according to the median TMAO level (3.92 μM), and their baseline characteristics are shown in Table 1. Compared with patients in the low-TMAO level group, patients in the high-TMAO level group had older age, higher BMI, lower eGFR and HDL-c levels, and higher TG, choline and carnitine levels (P < 0.05).

Laboratory testing
Fasting venous blood samples were collected using EDTA-containing pyrogen-free tubes after admission and immediately centrifuged at 4 °C and 3000 rpm for 10 min to obtain plasma samples. Then, the plasma samples were stored at −80 °C until analyses. The TMAO, choline, carnitine, betaine and GBB levels were measured by stable isotope dilution liquid chromatography-tandem mass spectrometry (LC/MS/MS) according to previous methods [20]. Briefly, plasma samples (20 μl) were mixed with 80 μl of 5 μM internal standards of TMAO-trimethyl-d9 (d9-TMAO), choline-trimethyl-d9 (d9-choline), carnitine-trimethyl-d9 (d9-carnitine), betaine-trimethyl-d9-methylene-d2 (d11-betaine) and GBB-trimethyl-d9 (d9-GBB) in methanol. They were detected in MRM MS mode: d9-TMAO at m/z 85.0 → 65.9 amu, d9-choline at m/z 113.2 → 68.9 amu, d9-carnitine at m/z 162.0 → 102.8 amu, d11-betaine at m/z 118.0 → 59.0 amu and d9- GBB at m/z 146.1 → 87 amu. Different concentrations of TMAO, carnitine, choline, betaine and GBB standards and a fixed amount of internal standards were added to 4% bovine serum albumin to generate the calibration curves for the quantification of blood TMAO-related metabolites. The standard curve coefficient of determination (R2) reached 0.999, and the accuracy across a span of different concentrations all reached 95%. After protein precipitation, the supernatant was recovered after centrifugation at 20000 g at 4 °C for 10 minutes. Then, 70 μl of the supernatant was absorbed into vials for loading, and the injection volume by the autosampler was 3 μl. The detections were performed on an API 5500Q-TRAP mass spectrometer (AB SCIEX, Massachusetts, USA). Analytes were monitored in positive MRM MS mode by using the following characteristic precursor-production ion transitions: TMAO at m/z 75.7 → 59.0 amu, choline at m/z 104.0 → 59.8 amu, carnitine at m/z 162.0 → 85.0 amu, betaine at m/z 118 → 57.7 amu and GBB at m/z 155.1 → 87.0 amu. All stable iso-labeled internal standards used in this study were purchased from Cambridge Isotope Laboratories, Inc. (Andover, MA, USA). Lipid parameters, including TG, total cholesterol (TC), high-density lipoprotein cholesterol (HDL-c), LDL-c, apolipoprotein A1 (ApoA1) and apolipoprotein B (ApoB), were measured by a biochemistry autoanalyzer (Beckman Coulter Dxc 800, California, USA) in the clinical laboratory center of Beijing Tiantan Hospital. Hyperlipidemia was defined as TC ≥ 6.22 mmol/L, LDL-c ≥ 4.14 mmol/L, TG ≥ 2.26 mmol/L or HDL-C < 1.04 mmol/L, which was based on the criteria recommended by the U.S. Adult Treatment Panel III.
Table 1 Baseline characteristics of the study cohort

| Variables                  | TMAO ≤ 3.92 μM (n = 56) | TMAO > 3.92 μM (n = 56) | P       |
|---------------------------|--------------------------|--------------------------|---------|
| Male, n (%)               | 36 (64.3)                | 29 (51.8)                | 0.180   |
| Age, years                | 60.18 ± 8.51             | 64.05 ± 8.96             | 0.021   |
| BMI, kg/m²                | 25.53 ± 3.15             | 26.91 ± 3.73             | 0.036   |
| Smoke, n (%)              | 28 (50.0)                | 27 (48.2)                | 0.850   |
| Alcohol, n (%)            | 16 (28.6)                | 12 (21.4)                | 0.383   |
| Hypertension, n (%)       | 43 (76.8)                | 42 (75.0)                | 0.825   |
| Hyperlipidemia, n (%)     | 27 (48.2)                | 31 (55.4)                | 0.449   |
| CHD, n (%)                | 45 (84.4)                | 49 (87.5)                | 0.303   |
| eGFR, mL/min/1.73 m²      | 101.01 ± 7.77            | 96.48 ± 7.69             | 0.002   |
| ApoB, mmol/L              | 1.69 ± 0.68              | 2.19 ± 1.15              | 0.006   |
| TC, mmol/L                | 4.27 ± 0.95              | 4.32 ± 1.03              | 0.795   |
| HDL-c, mmol/L             | 1.32 ± 0.42              | 1.09 ± 0.32              | 0.002   |
| LDL-c, mmol/L             | 2.64 ± 0.75              | 2.57 ± 0.74              | 0.589   |
| ApoA1, mmol/L             | 1.27 ± 0.19              | 1.26 ± 0.18              | 0.753   |
| ApoB, mmol/L              | 1.11 ± 0.33              | 1.09 ± 0.37              | 0.835   |
| TMAO, μM                  | 2.83 ± 1.34              | 8.43 ± 4.85              | <0.001  |
| Choline, μM               | 12.50 ± 2.81             | 14.01 ± 3.09             | 0.008   |
| Carnitine, μM             | 75.21 ± 15.69            | 82.58 ± 15.44            | 0.014   |
| Betaine, μM               | 39.26 ± 10.90            | 38.06 ± 10.09            | 0.548   |
| GBB, μM                   | 0.09 ± 0.02              | 0.10 ± 0.02              | 0.059   |

Data are expressed as the mean ± standard deviation or n (%)

TMAO Trimethylamine N-oxide, BMI Body mass index, eGFR Estimated glomerular filtration rate, CHD Coronary heart disease, TG Triglycerides, TC Total cholesterol, HDL-c High-density lipoprotein cholesterol, LDL-c Low-density lipoprotein cholesterol, ApoA1 Apolipoprotein A1, ApoB Apolipoprotein B, GBB γ-butyrobetaine

Associations between TMAO-related metabolites and lipids before rosuvastatin therapy

First, we found that TMAO was positively correlated with patient age and BMI and negatively correlated with eGFR (r = 0.265, 0.247 and −0.275, respectively, all P < 0.05), and betaine was negatively correlated with BMI (r = −0.285, P < 0.05) (Supplementary Table S3 and Fig. S1). Second, the associations among the TMAO-related metabolites were significant except for the association between betaine and GBB (Supplementary Table S4), and these associations still existed after partial correlation analyses adjusting for sex, age, BMI and eGFR (Supplementary Table S5). Third, compared with patients without hyperlipidemia (Supplementary Table S6), patients with hyperlipidemia had significantly higher TMAO levels (6.93 ± 5.42 μM vs. 4.23 ± 2.73 μM, P < 0.05), and the differences in other TMAO-related metabolites between the two patient groups were not significant (P > 0.05). Further analyses found that patients with hyper-TG or hypo-HDL-c had higher TMAO levels than patients without hyper-TG or hypo-HDL-c (7.94 ± 5.12 μM vs. 4.79 ± 4.00 μM or 6.97 ± 5.37 μM vs. 4.99 ± 3.95 μM, both P < 0.05), respectively. Patients with hyper-LDL-c had lower betaine levels than patients without hyper-LDL-c (33.17 ± 7.73 μM vs. 39.44 ± 10.61 μM, P < 0.05).

In univariate analyses (Supplementary Table S7), TMAO was positively correlated with TG and negatively correlated with HDL-c (r = 0.303 and −0.405, respectively, both P < 0.05), and betaine was negatively correlated with LDL-c (r = −0.308, P < 0.05). The correlations mentioned above still existed after partial correlation analyses adjusting for sex, age, BMI and eGFR (Table 2) or multivariate linear regression analyses adjusting for sex, age, BMI, eGFR and lipids TC, TG, LDL-c, HDL-c, ApoA1 and ApoB (Supplementary Tables S8 and S9, P < 0.05), and betaine was also significantly associated with TC after multivariate linear regression analyses (P < 0.05).

Table 2 Partial correlation analyses for the associations between TMAO-related metabolites and lipids before rosuvastatin therapy (n = 112)

| Variables                  | TMAO | Choline | Carnitine | Betaine | GBB |
|---------------------------|------|---------|-----------|---------|-----|
|                           | r    | P       | r         | P       | r   |
| TG                        | 0.294| 0.002   | −0.020    | 0.839   | 0.016| 0.008| 0.936| 0.135| 0.161|
| TC                        | 0.156| 0.106   | 0.093     | 0.338   | 0.073| 0.451| 0.093| 0.334| 0.075| 0.439|
| HDL-c                     | −0.316| 0.001  | −0.098    | 0.313   | −0.009| 0.923| −0.105| 0.275| 0.003| 0.977|
| LDL-c                     | 0.040| 0.680   | −0.037    | 0.702   | −0.090| 0.351| −0.269| 0.005| 0.043| 0.654|
| ApoA1                     | −0.081| 0.404  | −0.075    | 0.441   | 0.070| 0.468| −0.086| 0.373| −0.035| 0.715|
| ApoB                      | 0.021| 0.828   | 0.096     | 0.320   | 0.042| 0.668| −0.028| 0.772| 0.076| 0.429|

TMAO Trimethylamine N-oxide, TG Triglycerides, TC Total cholesterol, HDL-c High-density lipoprotein cholesterol, LDL-c Low-density lipoprotein cholesterol, ApoA1 Apolipoprotein A1, ApoB Apolipoprotein B, GBB γ-butyrobetaine. Partial correlation was performed by controlling for sex, age, body mass index and estimated glomerular filtration rate.

Changes in TMAO-related metabolites after rosuvastatin therapy

Rosuvastatin therapy significantly decreased TG, TC, LDL-c and ApoB levels but increased HDL-c and ApoA1 levels (Supplementary Table S10, P < 0.05). Moreover,
lower TMAO levels and higher carnitine, betaine and GBB levels were observed compared to baseline (3.82 ± 2.72 μM vs. 5.63 ± 4.52 μM, 83.23 ± 12.80 μM vs. 78.89 ± 15.93 μM, 4.67 ± 12.62 μM vs. 3.86 ± 10.47 μM and 0.11 ± 0.03 μM vs. 0.10 ± 0.02 μM, respectively, all \( P < 0.05 \)). The changes in TMAO-related metabolites were still significant after adjustment for sex, age, BMI and eGFR by multivariate analyses (Table 3, \( P < 0.05 \)). Compared with patients in the low-TMAO group (Supplementary Table S11), patients in the high-TMAO group had a greater reduction in TMAO levels after rosuvastatin therapy (3.85 ± 3.02 μM vs. 1.63 ± 1.50 μM, \( P < 0.05 \)).

In addition, (Supplementary Table S12), the correlation between TMAO and TG or between betaine and LDL-c disappeared after rosuvastatin therapy in univariate analysis (\( r = 0.050 \) and \( -0.175 \), respectively, both \( P > 0.05 \)), but correlations between TMAO and ApoB and between carnitine and LDL-c or between betaine and TG was still significant (Supplementary Tables S13 and S14, \( P < 0.05 \)), and a significantly positive association was also found between carnitine and TC (Supplementary Table S13, \( P < 0.05 \)).

**Discussion**

In this study, we systematically investigated the associations between TMAO-related metabolites and blood lipids and assessed the potential impact of rosuvastatin therapy. We found that TMAO and betaine were significantly associated with TG, HDL-c, TC and/or

### Table 3

Multivariate linear regression analyses for the changes in TMAO-related metabolites after rosuvastatin therapy (\( n = 112 \))

| Variables | TMAO a | Choline b | Carnitine b | Betaine b | GBB b |
|-----------|--------|-----------|-------------|-----------|-------|
| B         | P      | B         | P           | B         | P     |
| Sex       | −0.198 | 0.618     | −0.939      | 0.080     | −2.422 | 0.292 |
| Age       | 0.027  | 0.380     | 0.065       | 0.111     | −0.122 | 0.488 |
| BMI       | −0.010 | 0.854     | 0.007       | 0.923     | −0.070 | 0.825 |
| eGFR      | −0.037 | 0.276     | 0.059       | 0.188     | −0.055 | 0.775 |
| TMAO a    | 0.386  | <0.001    | 0.355       | <0.001    | 0.371  | <0.001 |
| Choline a | 0.371  | <0.001    |             |           |       |       |
| Carnitine a | 0.756 | <0.001    |             |           |       |       |
| Betaine a | 0.756  | <0.001    |             |           |       |       |
| GBB a     | 0.756  | <0.001    |             |           |       |       |

*a indicates before rosuvastatin therapy

*b indicates after rosuvastatin therapy

TMAO Trimethylamine N-oxide, BMI Body mass index, eGFR Estimated glomerular filtration rate, GBB γ-butyrobetaine. Variables including sex, age, BMI and eGFR were entered into the multivariate linear regression model.

### Table 4

Partial correlation analyses for the associations between TMAO-related metabolites and lipids after rosuvastatin therapy (\( n = 112 \))

| Variables | TMAO | Choline | Carnitine | Betaine | GBB |
|-----------|------|---------|-----------|---------|-----|
| \( r \)   | \( P \) | \( r \) | \( P \) | \( r \) | \( P \) |
| TG        | 0.076 | 0.434   | 0.051     | 0.598   | 0.003 | 0.978 |
| TC        | −0.158 | 0.101   | 0.020     | 0.833   | −0.104 | 0.281 |
| HDL-c     | −0.172 | 0.074   | 0.046     | 0.633   | 0.052 | 0.591 |
| LDL-c     | −0.171 | 0.075   | −0.071    | 0.460   | −0.298 | 0.002 |
| ApoA1     | −0.059 | 0.539   | 0.035     | 0.716   | −0.026 | 0.785 |
| ApoB      | −0.122 | 0.205   | −0.075    | 0.440   | −0.255 | 0.007 |

TG Triglycerides, TC Total cholesterol, HDL-c High-density lipoprotein cholesterol, LDL-c Low-density lipoprotein cholesterol, ApoA1 Apolipoprotein A1, ApoB Apolipoprotein B, GBB γ-butyrobetaine. Partial correlation was performed by controlling for sex, age, body mass index and estimated glomerular filtration rate.
 LDL-c levels in univariate and/or multivariate analyses. Compared to baseline, lower TMAO levels and higher carnitine, betaine and GBB levels were observed after rosuvastatin therapy, while the lipids decreased significantly. Meanwhile, the associations between TMAO and TG and HDL-c or the association between betaine and LDL-c disappeared postrosuvastatin therapy. The associations between carnitine and TC and LDL-c and between betaine and TG were significant in multivariate linear regression analyses after rosuvastatin therapy, although there were no associations before rosuvastatin therapy.

Disorders of the gut microbiota and its function are associated with a series of chronic metabolic diseases, such as hyperlipidemia, ASCVD, DM and obesity [1, 4, 14, 21–25]. As important gut microbiota metabolites, TMAO-related metabolites have also been studied in recent years because of their proatherogenic effects. First, the levels of TMAO-related metabolites were influenced by diet habits, gut microbiota compositions and hepatic flavin-dependent monooxygenases and varied significantly in different individuals [26, 27]. Second, TMAO-related metabolite levels were also influenced by the physical conditions of the patients. Studies have indicated that TMAO is mainly excreted by the kidney, and impaired kidney function or reduced eGFR could decrease TMAO elimination [28, 29]. In addition, TMAO levels can increase with advancing age [30]. In this study, we found a similar phenomenon, as mentioned above. TMAO levels were positively correlated with patient age and BMI and negatively correlated with patient eGFR. In addition, we also found that betaine was negatively associated with BMI, and this result was also consistent with previous studies that found a negative correlation between betaine and BMI [17, 31]. In this study, we also assessed the associations among TMAO-related metabolites and found that TMAO was positively associated with choline and GBB, and most TMAO-related precursors were positively correlated with each other, except betaine and GBB. All the data mentioned above indicate that although choline, carnitine and GBB are called precursors of TMAO [32], there is no one-to-one relationship among them because of various influencing factors [27, 33].

In addition to the associations between TMAO-related metabolites and ASCVD, studies also found that changes in gut microbiota compositions were associated with variations in blood lipids [14, 15], and TMAO-related metabolites are involved in lipid metabolism [1, 2, 4]. Researchers found that TMAO could reduce the expression of the bile acid synthetic enzyme cytochrome P450 7A1 (CYP7A1), which is the key enzyme in cholesterol metabolism [2]. Mice administered a diet containing choline, carnitine or TMAO exhibited decreased reverse cholesterol transport [1, 2, 4]. A TMAO-containing diet could also reduce the expression of intestinal cholesterol transporters, which could excrete cholesterol through enterocytes [2]. All these data indicate that TMAO-related metabolites may affect cholesterol-containing lipid levels, and there may be some associations between TMAO-related metabolites and blood lipids. In this study, we found that patients with hyperlipidemia had significantly higher TMAO levels than patients without hyperlipidemia, but the differences in other TMAO-related metabolites between the two patient groups were not significant. Further analyses according to the types of hyperlipidemia indicated that patients with hyper-TG or hypo-HDL-c tended to have higher TMAO levels than patients without hyper-TG or hypo-HDL-c. Patients with hyper-LDL-c tended to have lower betaine levels than patients without hyper-LDL-c. The results of univariate correlation analyses also supported that TMAO had a positive correlation with TG and a negative correlation with HDL-c, and betaine had a negative correlation with LDL-c. Because of the significant correlations between TMAO-related metabolites and the personal characteristics or lipids mentioned above, we further analyzed the correlations between TMAO-related metabolites and lipids by partial correlation analyses and multivariate analyses and found that the associations mentioned above were still significant before rosuvastatin therapy. In addition, a positive association between betaine and TC was also found in multivariate analyses. We failed to find a negative association between betaine and TG, as previously reported [17], and the results of this study support an atherogenic role of TAMO [1, 8, 11]. However, the conflicting associations between betaine and LDL-c and TC make the negative association between betaine and ASCVD controversial [17, 31]. All these results indicate that TMAO-related metabolites are significantly associated with blood lipids, but these associations do not exist between every marker of TMAO-related metabolites and lipids. Although many studies have suggested significant associations between TMAO-related metabolites and ASCVD [1, 2, 4, 34, 35], their exact mechanisms are still unclear. Together with the laboratory findings about the influence of TMAO-related metabolites on inflammation and cholesterol metabolism [2, 4, 10, 11, 16], the findings of the associations between TMAO-related metabolites and lipids in our study may help to explain part of the associations between TMAO-related metabolites and ASCVD.

Previous studies found that many drugs could affect the composition of the gut microbiota [19, 36–38], but there is scarce information about their effect on the metabolites of the gut microbiota. Rosuvastatin is an excellent lipid-lowering drug that is used for ASCVD.
treatment and prevention and is commonly prescribed in developed and developing countries. Recent studies found that rosuvastatin could profoundly affect the gut microbiota composition [19, 38] and even the levels of TMAO-related metabolites [19]. The results of a small cohort study found that patients in the rosuvastatin therapy group had higher betaine and GBB levels than those in the placebo group, and the TMAO and carnitine levels in the rosuvastatin therapy group also tended to decrease and increase separately [19]. In this study, we found that blood lipids were significantly decreased after rosuvastatin therapy, and significantly lower TMAO levels and higher carnitine, betaine and GBB levels were also observed compared to baseline. These changes in TMAO-related metabolites still existed after multivariate analyses. The results of our study further support the findings of a previous study [19]. Although there was a lack of placebo controls in this study, the changed TMAO-related metabolites after rosuvastatin therapy were considered to be associated with the potential impact of rosuvastatin therapy according to the self-control study. Whether it is necessary and how to treat these elevated precursors is an ambiguous question. Therefore, more experimental studies are needed to clarify the potential mechanisms involved and identify potential therapeutic targets for use in clinical therapy.

Although previous studies found an influence of rosuvastatin therapy on gut microbiota and its metabolites [19, 38, 39], it is still unclear whether rosuvastatin could influence the bioenzymes associated with the metabolism of TMAO precursors directly. Therefore, it is difficult to explain the contradictory phenomenon of the decreased TMAO and its increased precursors after rosuvastatin therapy. In addition, although TMAO and lipids had a decreasing trend at the same time, it is also unclear whether there were some correlations between the reductions in TMAO and lipids. As previous studies reported that the compositions of gut microbiota were associated with the variations of lipids and could be influenced by statins [15, 19, 38, 39], the altered compositions of gut microbiota may be responsible for part of the lipid changes after rosuvastatin therapy. To some extent, the reduction in TMAO may be correlated with part of the reduction in lipids according to the altered gut microbiota after rosuvastatin therapy.

In addition, following the changes in TMAO-related metabolites and lipids after rosuvastatin therapy, the associations between them were also altered. The correlation between TMAO and TG or between betaine and LDL-c disappeared after rosuvastatin therapy, but significantly negative associations between carnitine and LDL-c and between betaine and TG emerged in univariate and/or multivariate analyses. Although we found a significant positive association between betaine and TC in multivariate analyses before rosuvastatin therapy, it disappeared after rosuvastatin therapy, and a positive association between carnitine and TC was found in multivariate analyses. Although studies have reported that carnitine is atherogenic [2, 7], the conflicting associations between carnitine and LDL-c and TC make the positive association between carnitine and ASCVD controversial.

Comparisons with other studies and what the current study adds to the existing knowledge

As previous studies found that TMAO-related metabolites could influence cholesterol metabolism [2, 4, 16] and that the gut microbiota was associated with variations in lipids [14, 15], this study found that TMAO-related metabolites were also significantly associated with blood lipids, and these associations were changed but still existed after rosuvastatin therapy. In addition, similar to the results of a previous study [19], this study further demonstrated the potential impact of rosuvastatin therapy on TMAO-related metabolites.

Study strengths and limitations

To the best of our knowledge, this is the first study to systematically explore the associations between TMAO-related metabolites and lipids and the impact of rosuvastatin therapy. There are also some limitations. First, enrolled patients with carotid plaque, suspected ASCVD and relatively old age inevitably create a certain amount of selection bias. Second, although we used a self-control method to evaluate the changes in TMAO-related metabolites before and after rosuvastatin therapy, it is still a weakness because of the lack of a placebo control. Third, although we tried to enlarge the sample size and our study represents the largest cohort to date evaluated for the associations between TMAO-related metabolites and blood lipids, it is still limited by the relatively small sample size. Fourth, although all the patients were given a
recommended diet, instructed to avoid change and were even monitored weekly during the study, it was still difficult to avoid the potential influence of diet habits and a lack of reported specific diets. Finally, the usage of multiple drugs may inevitably lead to a potential impact on the gut microbiota and TMAO-related metabolites.

Conclusions
This study suggests that TMAO-related metabolites are significantly associated with blood lipids, although some of them were changed after rosuvastatin therapy. Lower TMAO levels and higher betaine, carnitine and GBB levels were observed after rosuvastatin therapy compared to baseline. This study adds to our knowledge about the role of TMAO-related metabolites in ASCVD and indicates that although rosuvastatin therapy could decrease blood lipids and TMAO significantly, the elevated TMAO precursors postrosuvastatin therapy and their potential impact on ASCVD should be considered in the clinic.

Abbreviations
ASCVD: Atherosclerotic cardiovascular disease; DM: Diabetes mellitus; TMAO: Trimethylamine N-oxide; GBB: γ-Butyrobetaine; MACE: Major adverse cardiovascular events; TG: Triglycerides; TC: Total cholesterol; HDL-c: High-density lipoprotein cholesterol; LDL-c: Low-density lipoprotein cholesterol; ApoA1: Apolipoprotein A1; ApoB: Apolipoprotein B; BMI: Body mass index; eGFR: Estimated glomerular filtration rate; CYP7A1: Cytochrome P450 7A1.

Supplementary Information
The online version contains supplementary material available at https://doi.org/10.1186/s12944-022-01673-3.

Additional file 1: Figure S1. The associations between the TMAO-related metabolites and personal characteristics (n = 112). Table S1. Baseline characteristics of patients without prior statin therapy (Group 1) and patients who received irregular statin therapy for less than one month (Group 2). Table S2. Baseline characteristics of males and females. Table S3. Correlations between the TMAO-related metabolites and personal characteristics (n = 112). Table S4. Correlations among the TMAO-related metabolites before statin therapy. Table S5. Partial correlation analyses for the associations among the TMAO-related metabolites before statin therapy. Table S6. Comparisons of the TMAO-related metabolites between different types of hyperlipidemia. Table S7. Correlations between the TMAO-related metabolites and blood lipids before rosuvastatin therapy (n = 112). Table S8. Multivariate linear regression analyses for the associations between betaine and blood lipids before rosuvastatin therapy. Table S9. Multivariate linear regression analyses for the associations between carnitine and blood lipids before rosuvastatin therapy. Table S10. The changes of TMAO-related metabolites after rosuvastatin therapy (n = 112). Table S11. The changes of TMAO-related metabolites between low-TMAO and high-TMAO groups after rosuvastatin therapy. Table S12. Correlations between the TMAO-related metabolites and blood lipids after rosuvastatin therapy (n = 112). Table S13. Multivariate linear regression analyses for the associations between carnitine and blood lipids after rosuvastatin therapy. Table S14. Multivariate linear regression analyses for the associations between betaine and blood lipids after rosuvastatin therapy.

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Not applicable.

Authors’ contributions
Xiaowei Xiong and Jian Zhou took part in the study design, data analyses and writing the manuscript. Qiang Fu, Xiaowei Xu, Shaoxin Wei and Shenghua Yang recruited the patients, isolated the blood samples, and assayed the levels of TMAO-related metabolites. Buxing Chen initiated and designed the study. All authors read and approved the final version of this manuscript.

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Availability of data and materials
The data can be obtained from the corresponding author following the permission of the Ethics Committee of Beijing Tiantan Hospital.

Declarations
Ethics approval and consent to participate
This study was registered at http://clinicaltrials.gov/ (NCT02305862), abided by the principles of the Declaration of Helsinki, and was approved by the Ethics Committee of Beijing Tiantan Hospital (Approval No. KY2014-D20-02). All patients in this study signed informed consent forms.

Consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

Author details
1 Department of Cardiology and Macrovascular Disease, Beijing Tiantan Hospital, Capital Medical University, Beijing, China. 2 Department of Cardiovascular Medicine, Shanghai East Hospital, School of Medicine, Tongji University, Shanghai, China. 3 Department of Cardiology, Beijing University of Chinese Medicine Third Affiliated Hospital, No. 51, Xiaoguan Street, Andingmenwai, Chaoyang District, Beijing 100029, China.

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References
1. Wang Z, Klipfell E, Bennett BJ, Koeth R, Levison BS, Dugar B, et al. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. Nature. 2011;472:57–63.
2. Koeth RA, Wang Z, Levison BS, Buffa JA, Org E, Sheehy BT, et al. Intestinal microbiota metabolism of L-carnitine, a nutrient in red meat, promotes atherosclerosis. Nat Med. 2013;19:576–85.
3. Jonsson AL, Backhed F. Role of gut microbiota in atherosclerosis. Nat Rev Cardiol. 2017;14:79–87.
4. Koeth RA, Levison BS, Culley MK, Buffa JA, Wang Z, Gregory JC, et al. γ-Butyrobetaine is a proatherogenic intermediate in gut microbial metabolism of L-carnitine to TMAO. Cell Metab. 2014;20:799–812.
5. Tang WH, Wang Z, Fan Y, Levison B, Hazen SE, Donahue LM, et al. Prognostic value of elevated levels of intestinal microbe-generated metabolite trimethylamine-N-oxide in patients with heart failure: refining the gut hypothesis. J Am Coll Cardiol. 2014;64:1908–14.
6. Tang WH, Wang Z, Shrestha K, Borowski AG, Wu Y, Troughton RW, et al. Intestinal microbiota-dependent phosphatidylcholine metabolites, diastolic dysfunction, and adverse clinical outcomes in chronic systolic heart failure. J Card Fail. 2015;21:91–6.
7. Skagen K, Troseid M, Ueland T, Holm S, Abbas A, Gregersen I, et al. The carnitine-butyrobetaine-trimethylamine-N-oxide pathway and its association with cardiovascular mortality in patients with coronary atherosclerosis. Atherosclerosis. 2016;247:64–9.
8. Senthong V, Li XS, Hudec T, Coughlin J, Wu Y, Levison B, et al. Plasma trimethylamine N-oxide, a gut microbe-generated phosphatidylcholine metabolite, is associated with atherosclerotic burden. J Am Coll Cardiol. 2016;67:2620–8.
9. Xiong X, Liu H, Hua L, Zhao H, Wang D, Li Y. The association of HDL-apoCIII with coronary heart disease and the effect of statin treatment on it. Lipids Health Dis. 2015;14:127.

10. Chen ML, Zhu XH, Ran L, Lang HD, Yi L, Mi MT. Trimethylamine-N-oxide induces vascular inflammation by activating the NLRP3 Inflammasome through the SIRT3-SOD2-mtROS signaling pathway. J Am Heart Assoc. 2017;6:e006347.

11. Cheng X, Qiu X, Liu Y, Yuan C, Yang X. Trimethylamine-N-oxide promotes tissue factor expression and activity in vascular endothelial cells: a new link between trimethylamine-N-oxide and atherosclerotic thrombosis. Thromb Res. 2019;177:110–6.

12. Chou RH, Chen CY, Chen IC, Huang HL, Lu YW, Kuo CS, et al. Trimethylamine-N-oxide, circulating endothelial progenitor cells, and endothelium in patients with stable angina. Sci Rep. 2019;9:4249.

13. Rohrmann S, Linseisen J, AllenSpach M, von Eckardstein A, Müller D. Plasma concentrations of trimethylamine-N-oxide are directly associated with dairy food consumption and low-grade inflammation in a German adult population. J Nutr. 2016;146:283–9.

14. Fu J, Bondor MJ, Cenit WC, Tsigelchaia EA, Maatman A, Dekens JA, et al. The gut microbiome contributes to a substantial portion of the variation in blood lipids. Circ Res. 2015;117:817–24.

15. Le Chatelier E, Nielsen T, Qin J, Pritsch E, Hildebrand F, Falony G, et al. Richness of human gut microbiome correlates with metabolic traits. Nature. 2013;500:541–6.

16. Ding L, Chang M, Guo Y, Zhang L, Xue C, Yanagita T, et al. Trimethylamine-N-oxide (TMAO)-induced atherosclerosis is associated with bile acid metabolism. Lipids Health Dis. 2018;17:286.

17. Lever M, George PM, Atkinson W, Molyneux SL, Elmslie JL, Slow S, et al. Plasma lipids and betaine are related in an acute coronary syndrome cohort. PloS One. 2011;6:e21666.

18. Zhermakova A, Kuriokshikov A, Bondor MJ, Tsigelchaia E, Schirmer M, Vatanen T, et al. Population-based metagenomics analysis reveals markers for gut microbiome composition and diversity. Science. 2016;352:565–9.

19. Kummen M, Solberg OG, Storm-Larsen C, Holm K, Ragnarsson A, Trøseid M, et al. Rosuvastatin alters the genetic composition of the human gut microbiome. Sci Rep. 2020;10:5397.

20. Trøseid M, Ueland T, Hov JR, Svardal A, Gregersen I, Dahl CP, et al. Statin therapy is associated with lower prevalence of gut microbiota dysbiosis. Nature. 2020;581:310–5.

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10. Wang Z, Levison BS, Hazen JE, Donahue L, Li XM, Hazen SL. Measurement of trimethylamine-N-oxide by stable isotope dilution liquid chromatography tandem mass spectrometry. Anal Biochem. 2014;453:35–40.

11. Guasch-Ferré M, Hu FB, Ruiz-Canela M, Bulló M, Toledo E, Wang DD, et al. Plasma metabolites from choline pathway and risk of cardiovascular disease in the PREDIMED (prevention with Mediterranean diet) study. J Am Heart Assoc. 2017;6:e006524.

12. Menon V, Kumar A, Patel DR, John JS, Wolksi KE, McErlean E, et al. Impact of baseline glycemic control on residual cardiovascular risk in patients with diabetes mellitus and high-risk vascular disease treated with statin therapy. J Am Heart Assoc. 2020;9:e014328.

13. Kühn T, Rohrmann S, Sookthai D, Johnson T, Ketzke V, Kaaks R, et al. Intra-individual variation of plasma trimethylamine-N-oxide (TMAO), betaine and choline over 1 year. Clin Chem Lab Med. 2017;55:261–8.

14. Geng J, Yang C, Wang B, Zhang X, Hu T, Gu Y, et al. Trimethylamine-N-oxide promotes atherosclerosis via CD36-dependent MAPK/JNK pathway. Biomed Pharmacother. 2018;97:941–7.

15. Mohammadi A, Najar AG, Yaghoubi MM, Jahani Y, Vahabzadeh Z. Trimethylamine-N-oxide treatment induces changes in the ATP-binding cassette transporter A1 and scavenger receptor A1 in murine macrophage J774A.1 cells. Inflammation. 2016;39:393–404.

16. Wu H, Esteve E, Tremaroli V, Khan MT, Caucas R, Mannérás-Holm L, et al. Metformin alters the gut microbiome of individuals with treatment-naïve type 2 diabetes, contributing to the therapeutic effects of the drug. Nat Med. 2017;23:850–8.

17. Freedberg DE, Toussaint NC, Chen SP, Ratner AI, Whittier S, Wang TC, et al. Proton pump inhibitors Alter specific taxa in the human gastrointestinal microbiome: a crossover trial. Gastroenterology. 2015;149:883–885.e889.

18. Liu Y, Song X, Zhou H, Zhou X, Xia Y, Dong X, et al. Gut microbiome associates with lipid-lowering effect of Rosuvastatin in vivo. Front Microbiol. 2018;9:530.

19. Vieira-Silva S, Falony G, Belda E, Nielsen T, Aron-Wisnewsky J, Chakaroun R, et al. Statin therapy is associated with lower prevalence of gut microbiota dysbiosis. Nature. 2020;581:310–5.