Original Article

Design, Synthesis and Biological Evaluation of New 1, 4-Dihydropyridine (DHP) Derivatives as Selective Cyclooxygenase-2 Inhibitors

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Abstract

As a continuous research for discovery of new COX-2 inhibitors, chemical synthesis, in vitro biological activity and molecular docking study of a new group of 1, 4-dihydropyridine (DHP) derivatives were presented. Novel synthesized compounds possessing a COX-2 SO\textsubscript{2}Me pharmacophore at the \textit{para} position of C-4 phenyl ring, different hydrophobic groups (R\textsubscript{1}) at C-2 position and alkoxy carbonyl groups (COOR\textsubscript{2}) at C-3 position of 1, 4-dihydropyridine, displayed selective inhibitory activity against COX-2 isozyme. Among them, compound 5e was identified as the most potent and selective COX-2 inhibitor with IC\textsubscript{50} value of 0.30 µM and COX-2 selectivity index of 92. Molecular docking study was performed to determine probable binding models of compound 5e. The study showed that the p-SO\textsubscript{2}Me-phenyl fragment of 5e inserted inside secondary COX-2 binding site (Arg\textsuperscript{513}, Phe\textsuperscript{518}, Gly\textsuperscript{519}, and His\textsuperscript{90}). The structure-activity relationships acquired reveal that compound 5e with methyl and ethoxycarbonyl as R\textsubscript{1} and COOR\textsubscript{2} substitutions has the necessary geometry to provide selective inhibition of the COX-2 isozyme and it can be a good basis for the development of new hits.

Keywords: Synthesis; 1, 4-Dihydropyridine (DHP) Derivatives; COX-2 Inhibitors; Molecular modeling.

Introduction

Cyclooxygenase (COX) also known as prostaglandin synthase (PGH) is an apotent mediator of inflammation. Non-steroidal anti-inflammatory drugs (NSAIDs) bind to cyclooxygenase, thereby inhibiting the production of prostaglandins. However, inhibition of COXs may lead to undesirable side effects. Nowadays, it is well established that there are at least two COX isozymes, COX-1 and COX-2 (1). The constitutive COX-1 isozyme is produced in a variety of tissues and appears to be important to the maintenance of physiological functions such as gastric protection and vascular homeostasis (2, 3). As COX-2 is usually specific to inflamed tissue, there is much less gastric irritation associated with COX-2 inhibition. This has led intense efforts in searching for potent and selective COX-2 inhibitors which could provide anti-inflammatory drugs with fewer risks. Several classes of compounds having selective COX-2 inhibitory activity have been reported in...
and synthetically important class of compounds in the field of drugs and pharmaceuticals and have attracted attention of synthetic chemists due to their pharmacological properties (8, 9). The Hantzsch reaction is a well-known method for synthesizing dihydropyridines (10). Hantzsch reaction is a kind of multi component reactions (MCRs) which have gained wide applicability in the field of synthetic organic chemistry as they increase the efficiency of the reaction and decrease the number of laboratory operations along with quantities of solvent and chemicals (11, 12).

In this study novel 1, 4-dihydropyridine derivatives were prepared according to Hantzsch reaction and evaluated for in vitro COX-1/COX-2 isozyme inhibition. We also performed docking studies to determine the orientation of the synthesized compounds in the COX-2 active site which led to the better understanding of the structure-activity relationship in designed COX-2 inhibitors.

**Experimental**

**General**

All chemicals and solvents used in this study were purchased from Merck AG and Aldrich Chemical. Melting points were determined using a Thomas-Hoover capillary apparatus. Infrared spectra were acquired using a Perkin Elmer Model 550 SE spectrometer. A Bruker AM-300 NMR spectrometer was used to acquire $^1$H NMR spectra with TMS as internal standard. Coupling constant ($J$) values are estimated in hertz (Hz) and spin multiples are given as s (singlet), d (double), t (triplet), q (quartet), m (multiplet), and br (broad). Low-resolution mass spectra were acquired with an MAT CH5/DF (Finnigan) mass spectrometer that was coupled on line to a Data General DS 50 data system. Electron-impact ionization was performed at an ionizing energy of 70 eV with a source temperature of 250°C. Elemental microanalyses, determined for C and H, were within ±0.4% of theoretical values. All chemicals and solvents used in this study were purchased from Merck AG and Aldrich Chemical. Melting points were determined with a Thomas-Hoover capillary apparatus. Infrared spectra were acquired using a Perkin Elmer Model 1420
was poured into 80 mL ice water the precipitate was filtered off and washed with water. The crude products were purified by recrystallization from ethanol to give final products.

Methyl-1, 4, 5, 6, 7, 8-hexahydro-2, 7, 7-trimethyl-4-(4-(methylsulfonyl)phenyl)-5-oxoquinoline-3-carboxylate (5a)

Yield, 78%; mp: 244-245 °C; IR (KBr disk) \( \nu \) (cm\(^{-1}\)): 1150, 1300 (SO\(_2\)); 1400-1600 (aromatic); 1689 (C=O); 3356 (NH); \( \delta \) (CDCl\(_3\), 500 MHz): 0.91 (s, 3H, CH\(_3\)), 1.12 (s, 3H, CH\(_3\)), 2.16-2.20 (m, 2H, dihydroquinoline H\(_4\)), 2.26-2.29 (m, 2H, dihydroquinoline H\(_5\), \( J=15.8 \) Hz), 2.45 (s, 3H, CH\(_3\)), 3.07 (s, 3H, SO\(_2\)Me), 3.64 (s, 3H, CO\(_2\)CH\(_3\)), 5.17 (s, 1H, dihydroquinoline H\(_5\)), 5.84 (s, 1H, NH), 7.54 (d, 2H, methanesulfonyl phenyl H\(_2\) & H\(_6\)\'), 7.81 (d, 2H, methanesulfonyl phenyl H\(_3\) & H\(_5\)\'), 8.2 Hz); LC-MS (ESI) \( m/z \): 404.3 (M+1, 100); Anal. Calcd. for C\(_{21}\)H\(_{25}\)NO\(_5\)S: C, 62.51; H, 6.25; N, 3.47. Found: C, 62.81; H, 6.45; N, 3.59.

Methyl-2-amino-1, 4, 5, 6, 7, 8-hexahydro-2, 7-dimethyl-4-(4-(methylsulfonyl)phenyl)-5-oxoquinolone-3-carboxylate (5b)

Yield, 64%; mp: 177-179°C; IR (KBr disk) \( \nu \) (cm\(^{-1}\)): 1150, 1300 (SO\(_2\)); 1400-1600 (aromatic); 1697 (C = O); 3350 (NH); \( \delta \) (CDCl\(_3\), 500 MHz): 0.99 (s, 3H, CH\(_3\)), 1.12 (s, 3H, CH\(_3\)), 2.20 (d, 1H, dihydroquinoline H\(_5\)), \( J=16.3 \) Hz), 2.29 (d, 1H, dihydroquinoline H\(_5\), \( J=16.3 \) Hz), 2.48 (s, 2H, dihydroquinoline H\(_6\)), 3.07 (s, 3H, SO\(_2\)Me), 3.62 (s, 3H, CO\(_2\)CH\(_3\)), 4.81 (s, 1H, NH), 6.27 (br s, 2H, NH\(_2\)), 7.49 (d, 2H, methanesulfonyl phenyl H\(_2\) & H\(_6\)\'), \( J=8.2 \) Hz), 7.86 (d, 2H, methanesulfonyl phenyl H\(_3\) & H\(_5\)\'), \( J=8.2 \) Hz); LC-MS (ESI) \( m/z \): 405.1 (M+1,
Methyl-2-ethyl-1, 4, 5, 6, 7, 8-hexahydro-7,7-dimethyl-4-(4-(methylsulfonyl)phenyl)-5-oxoquinoline-3-carboxylate (5c)

Yield, 87%; mp: 165-166°C; IR (KBr disk) ν (cm⁻¹): 1150, 1300 (SO₂); 1400-1600 (aromatic); 1657 (C=O); 3352 (NH); 1HNNMR (CDCl₃, 500 MHz): δ 0.89 (s, 3H, CH₃); 1.12 (t, 3H, CH₃), 1.71 (t, 3H, CH₃), 1.36 (t, 3H, CH₃), 1.29 (d, 2H, methanesulfonyl phenyl H, J=1.21 Hz), 3.04 (s, 3H, SO₂Me), 3.64 (s, 3H, CO₂H), 5.19 (s, 1H, dihydroquinoline H₃), 5.80 (br s, 1H, NH), 7.51 (d, 2H, methanesulfonyl phenyl H, J=8.1 Hz), 7.81 (d, 2H, methanesulfonyl phenyl H, J=8.2 Hz); LC-MS (ESI)m/z: 432.2 (M+1, 100); Anal. Calcd. for C₂₄H₂₃NO₅S: C, 64.21; H, 6.95; N, 3.12.

Ethyl-1, 4, 5, 6, 7, 8-hexahydro-7,7-dimethyl-4-(4-(methylsulfonyl)phenyl)-5-oxo-2-propylquinoline-3-carboxylate (5f)

Yield, 54%; mp: 163-164°C; IR (KBr disk) ν (cm⁻¹): 1150, 1300 (SO₂); 1400-1600 (aromatic); 1658(C = O); 3305 (NH); 1HNNMR (CDCl₃, 500 MHz): δ 0.88 (s, 3H, CH₃), 1.05 (t, 3H, CH₃), 1.12 (s, 3H, CH₃), 1.27 (t, 3H, CH₃), 1.71 (m, 4H, 2CH₂), 2.16-2.20 (d, 1H, dihydroquinoline H₃, J=1.62 Hz); 2.26-2.29 (m, 2H, dihydroquinoline H₆ & H₇), 2.39-2.42 (d, 1H, dihydroquinoline H₅, J=15.0 Hz), 2.45 (d, 1H, dihydroquinoline H₅, J=16.6 Hz), 3.04 (s, 3H, SO₂Me), 3.63 (s, 3H, CO₂H), 4.27 (m, 1H, CH), 5.19 (s, 1H, dihydroquinoline H₆), 6.07 (s, 1H, NH), 7.50 (d, 2H, methanesulfonyl phenyl H, J=8.3 Hz), 7.80 (d, 2H, methanesulfonyl phenyl H, J=8.3 Hz); LC-MS (ESI)m/z: 446.2 (M+1, 100); Anal. Calcd. for C₂₄H₂₅NO₅S: C, 64.69; H, 7.01; N, 3.14. Found: C, 63.89; H, 6.95; N, 3.32.

Ethyl-1, 4, 5, 6, 7, 8-hexahydro-7,7-dimethyl-4-(4-(methylsulfonyl)phenyl)-5-oxo-2-phenylquinoline-3-carboxylate (5g)

Yield, 87%; mp: 187.9-189.5°C; IR (KBr disk) ν (cm⁻¹): 1150, 1300 (SO₂); 1400-1600 (aromatic); 1687(C = O); 3344 (NH); 1HNNMR (CDCl₃, 500 MHz): δ 0.89 (s, 3H, CH₃), 0.97 (s, 3H, CH₃), 1.14 (s, 3H, CH₃), 2.19-2.32 (m, 3H, dihydroquinoline H₅ & H₆), 2.45-2.48 (d, 1H, dihydroquinoline H₅, J=15.6 Hz), 3.03 (s, 3H, SO₂Me), 3.86 (m, 2H, CH₂), 5.28 (s, 1H, dihydroquinoline H₆), 6.07 (s, 1H, NH), 7.36 (m, 2H, benzyl H & H₂), 7.45 (m, 3H, benzyl H, H₆ & H₇), 7.68 (d, 2H, methanesulfonyl phenyl H₆ & H₇, J=7.6 Hz), 7.85 (d, 2H, methanesulfonyl phenyl H₆ & H₇, J=7.5 Hz); LC-MS (ESI)m/z: 480.2 (M+1, 100); Anal. Calcd. for C₂₅H₂₅NO₅S: C, 67.62; H, 6.69; N, 2.92. Found: C, 63.96; H, 6.25; N, 3.12.

Ethyl-1, 4, 5, 6, 7, 8-hexahydro-7,7-dimethyl-4-(4-(methylsulfonyl)phenyl)-5-oxo-2-phenylquinoline-3-carboxylate (5g)

Yield, 87%; mp: 187.9-189.5°C; IR (KBr disk) ν (cm⁻¹): 1150, 1300 (SO₂); 1400-1600 (aromatic); 1687(C = O); 3344 (NH); 1HNNMR (CDCl₃, 500 MHz): δ 0.89 (s, 3H, CH₃), 0.97 (s, 3H, CH₃), 1.14 (s, 3H, CH₃), 2.19-2.32 (m, 3H, dihydroquinoline H₅ & H₆), 2.45-2.48 (d, 1H, dihydroquinoline H₅, J=15.6 Hz), 3.03 (s, 3H, SO₂Me), 3.86 (m, 2H, CH₂), 5.28 (s, 1H, dihydroquinoline H₆), 6.07 (s, 1H, NH), 7.36 (m, 2H, benzyl H & H₂), 7.45 (m, 3H, benzyl H, H₆ & H₇), 7.68 (d, 2H, methanesulfonyl phenyl H₆ & H₇, J=7.6 Hz), 7.85 (d, 2H, methanesulfonyl phenyl H₆ & H₇, J=7.5 Hz); LC-MS (ESI)m/z: 480.2 (M+1, 100); Anal. Calcd. for C₂₅H₂₅NO₅S: C, 67.62; H, 6.69; N, 2.92. Found: C, 63.96; H, 6.25; N, 3.12.
t-Butyl-1, 4, 5, 6, 7, 8-hexahydro-2,7,7-trimethyl-4-(4-methanesulfonyl-phenyl)-5-oxoquinoline-3-carboxylate (5h)

Yield, 95%; mp: 163-164°C; IR (KBr disk) (cm⁻¹): 1150, 1300 (SO₂); 1400-1660 (aromatic); 1694 (C = O); 3300-3500 (NH); 1'H NMR (CDCl₃, 500 MHz): 8 0.95 (s, 3H, CH₃), 1.11 (s, 3H, CH₃), 1.37 (s, 9H, CH₃), 2.14 (d, 1H, dihydroquinoline Hₜ, J=16.3 Hz), 2.26 (d, 1H, dihydroquinoline Hₜ, J=15.9 Hz), 2.36-2.39 (d, 2H, dihydroquinoline Hₜ), 2.41 (s, 3H, CH₃), 3.03 (s, 3H, SO₂Me), 5.10 (s, 1H, dihydroquinoline Hₜ), 5.91 (br s, 1H, NH), 7.54 (d, 2H, methanesulfonyl phenyl Hₜ & Hₜ', J=8.2 Hz), 7.81 (d, 2H, methanesulfonyl phenyl Hₜ & Hₜ', J=8.2 Hz); LC-MS (ESI)m/z: 446.2 (M⁺+1, 100); Anal. Calcd. for C₂₉H₂₉NO₅S: C, 67.62; H, 6.09; N, 2.92. Found: C, 67.32; H, 7.01; N, 3.14. Found: C, 64.89; H, 7.21; N, 3.32.

Benzyl-1, 4, 5, 6, 7, 8-hexahydro-2, 7, 7-trimethyl-4-(4-methanesulfonylphenyl)quinoline-3-carboxylate (5i)

Yield, 87%; mp: 136-138.9 °C; IR (KBr disk) (cm⁻¹): 1150, 1300 (SO₂); 1400-1660 (aromatic); 1694 (C = O); 3557 (NH); 1'H NMR (CDCl₃, 500 MHz): 8 0.86 (s, 3H, CH₃), 1.04 (s, 3H, CH₃), 2.01-2.07 (d, 1H, dihydroquinoline Hₜ, J=16.3 Hz), 2.17-2.20 (d, 1H, dihydroquinoline Hₜ, J=16.4 Hz), 2.20-2.36 (q, 2H, dihydroquinoline Hₜ), 2.38 (s, 3H, CH₃), 2.96 (s, 3H, SO₂Me), 4.98 (s, 2H, CH₂), 5.10 (s, 1H, dihydroquinoline Hₜ), 7.11-7.12 (m, 2H, benzyl Hₜ & Hₜ'), 7.26 (m, 3H, benzyl Hₜ, Hₜ & Hₜ'), 7.41 (d, 2H, methanesulfonyl phenyl Hₜ & Hₜ'), J=8.2 Hz), 7.67 (d, 2H, methanesulfonyl phenyl Hₜ & Hₜ', J=8.3 Hz); LC-MS (ESI)m/z: 480.2 (M⁺+1, 100); Anal. Calcd. for C₂₉H₂₉NO₅S: C, 67.62; H, 6.09; N, 2.92. Found: C, 67.32; H, 5.84; N, 3.02.

Molecular Modeling

The active compound was selected for docking studies which performed using Autodock software Version 4.0. The ligand molecule was constructed using the Chem Draw and was energy minimized for 1000 iterations reaching a convergence of 0.01 kcal/mol Å. The coordinates of the X-ray crystal structure of COX-2 enzyme was obtained from the RCSB Protein Data Bank (3NT1) and the protein structure was prepared for docking. First of all, co-crystallized ligand and all water molecules were removed from crystal protein. Polar hydrogens were added and non polar hydrogens were merged, finally Kallman unitedatom charge and atom type parameter was added to 3NT1. Grid map dimensions (20×20×20) were set surrounding active site. Lamarckian genetic search algorithm was employed and docking run was set to 50. The aim of docking is to search for suitable binding configuration between the ligands and the rigid protein. These docked structures were very similar to the minimized structures provided initially. The quality of the docked structures was determined by measuring the intermolecular energy of the ligand-enzyme assembly (13).

Result and Discussion

A group of 1,4-dihydropyridine derivatives possessing a MeSO₂ at the para-position of the C-4 phenyl ring, alkyl groups (R₃) at the C-2 position and alkoxy carbonyl groups (COOR₇) at the C-3 position were prepared and evaluated for their ability to inhibit COX-1 and COX-2 using chemiluminescent kit (Cayman chemical, MI, USA) according to our previously reported method (14). Potent and selective COX-2 inhibitor, celecoxib was used as a reference compound in the COX activity assay. All experiments were carried out at least three times and the data of inhibitory effects were summarized in Table 1.

As shown in Table 1, all compounds except 5i and 5g (IC₅₀ > 100 µM) displayed moderate to good inhibitory activities against COX-2 and were more potent inhibitor of COX-2 (IC₅₀ = 0.3-1.38 µM range) than COX-1 (IC₅₀ = 22.9-46.1 M range) with COX-2 selectivity indexes (SI) in the range of 18.2-92.0. However, in all cases, the measured activities were lower than that of celecoxib. Our results indicated that different hydrophobic substituents at C-2 and C-3 position of 1, 4-dihydropyridine core affected the activity of the target molecules. In compounds series possessing methoxy carbonyl as COOR₇ group (5a-d), replacement of methyl (5a, IC₅₀ = 0.48 µM) at C-2 position with other alkyl groups such
as ethyl (5c, IC\textsubscript{50} = 0.59 µM) and isopropyl (5d, IC\textsubscript{50} = 0.62 µM) slightly decreased the COX-2 inhibitory activity. Compound 5b showed approximately similar potency (IC\textsubscript{50} = 0.44 µM) to compound 5a. This may be due to isosteric replacement of methyl group with NH\textsubscript{2} group in compound 5b. It is found that replacement of methoxycarbonyl with ethoxycarbonyl as R\textsubscript{2} group in compound 5a resulted in compound 5e with improved COX-2 inhibitory effect (IC\textsubscript{50} = 0.30 µM). Introduction of larger groups such as propyl and phenyl at C-2 position of compound 5e led to compounds 5f and 5g with significant loss of activities. The experimental results showed that t-butoxycarbonyl as COOR\textsubscript{2} group is well tolerated and the corresponding compound, 5h exhibited IC\textsubscript{50} value of 0.40 M. In addition, modification of ethoxycarbonyl group to benzyloxy carbonyl group in compound 5e led to compound 5i with no activity (IC\textsubscript{50}>100 µM). This may be due to large size of substitution and resulting steric hindrance.

Molecular docking studies helped to understand the various interactions between the most active ligand (5e) and enzyme active sites in details. According to docking studies results (Figure 2), it is clear that p-SO\textsubscript{2}Me-phenyl moiety of compound 5e inserts deep inside the COX-2 active site pocket and forming hydrogen bond with Arg\textsuperscript{513} (distance = 4.8 Å) and His\textsuperscript{90} (distance = 3.1 Å). In addition, the N-H of the 1,4-dihydropyridine scaffold interacts with C=O of Val\textsuperscript{349} (distance = 4.0 Å). Moreover, the carbonyl group of central ring and ethoxycarbonyl bind to Arg\textsuperscript{520} (distance = 2.8 Å) and Gly\textsuperscript{526} (distance = 3.9 Å) through hydrogen bonds, respectively. Molecular docking studies associated with experimental results showed that compound 5e possesses the pharmacophoric requisites for COX-2 inhibition.

### Conclusion

In conclusion, new 1,4-dihydropyridine derivatives were synthesized and evaluated for COX-1/COX-2 inhibition. Among them, compound 5e exhibited good COX-2 inhibitory activity and selectivity (IC\textsubscript{50}=0.30 µM and COX-2 selectivity index=92). Experimental results in conjunction with molecular docking studies indicated that compound 5e with methyl and ethoxycarbonyl groups as R\textsubscript{1} and COOR\textsubscript{2} substitutions could interact appropriately...
with COX-2 active site. Therefore, this compound provides a promising lead for further development.

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