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Environmental controls on daytime net community calcification on a Red Sea reef flat

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Abstract Coral growth and carbonate accumulation form the foundation of the coral reef ecosystem. Changes in environmental conditions due to coastal development, climate change, and ocean acidification may pose a threat to net carbonate production in the near future. Controlled laboratory studies demonstrate that calcification by corals and coralline algae is sensitive to changes in aragonite saturation state ($\Omega_a$), as well as temperature, light, and nutrition. Studies also show that the dissolution rate of carbonate substrates is impacted by changes in carbonate chemistry. The sensitivity of coral reefs to these parameters must be confirmed and quantified in the natural environment in order to predict how coral reefs will respond to local and global changes, particularly ocean acidification. We estimated the daytime hourly net community metabolic rates, both net community calcification (NCC) and net community productivity (NCP), at Sheltered Reef, an offshore platform reef in the central Red Sea. Average NCC was $8 \pm 3$ mmol m$^{-2}$ h$^{-1}$ in December 2010 and $11 \pm 1$ mmol m$^{-2}$ h$^{-1}$ in May 2011, and NCP was $21 \pm 7$ mmol m$^{-2}$ h$^{-1}$ in December 2010 and $44 \pm 4$ mmol m$^{-2}$ h$^{-1}$ in May 2011. We also monitored a suite of physical and chemical properties to help relate the rates at Sheltered Reef to published rates from other sites. While previous research shows that short-term field studies investigating the NCC–$\Omega_a$ relationship have differing results due to confounding factors, it is important to continue estimating NCC in different places, seasons, and years, in order to monitor changes in NCC versus $\Omega$ in space and time, and to ultimately resolve a broader understanding of this relationship.

Keywords Coral · Calcification · CaCO$_3$ · Acidification · Aragonite saturation

Introduction

Biologically mediated calcification is an essential process on coral reefs. Corals build the foundation of the reef, enabling the reef to keep pace with changes in local sea level (Stoddart 1969; Grigg 1982; Kleypas and Langdon 2006; Kleypas et al. 2006). Coralline algae, foraminifera and sand serve as the infill and cement that fortify the reef foundation (Adey 1998; Kleypas and Langdon 2006; Kleypas et al. 2006). Coral colonies of diverse morphologies create the complex habitat that supports the extremely high biodiversity of reef ecosystems (Kleypas and Langdon 2006; Kleypas et al. 2006). Net community calcification (NCC) is the production of biogenic carbonate minerals by calcifying organisms minus the loss of that material by dissolution.
The growth of calcifying reef organisms is presently threatened by environmental changes due to climate change and ocean acidification (Kleypas et al. 2006; Pandolfi et al. 2011), in addition to the local stresses imposed by coastal development, harmful fishing practices, invasive species, and disease.

Ocean acidification (OA) is the process by which anthropogenic CO₂ dissolves in the surface ocean and depresses the pH and carbonate saturation state (Ω) of seawater. Results from controlled experiments and field studies indicate that a decline in aragonite saturation state (Ωₐ) reduces coral and algal calcification rates and increases dissolution rates of carbonate sediments and reef matrix (Marubini and Atkinson 1999; Langdon et al. 2000; Leclercq et al. 2000; Marubini et al. 2001; Langdon and Atkinson 2005; Jokiel et al. 2008; Kuffner et al. 2008; Martin and Gattuso 2009). The combined effect of reduced calcification rates and increased dissolution rates is reduced NCC. The negative impact on calcification rates is presumably because the biologically mediated precipitation of a given mineral is more energetically costly when the mineral saturation state and pH of the seawater are depressed. Corals have been shown to mediate calcification by regulating the saturation state and pH of the internal fluid from which the carbonate skeleton precipitates (Venn et al. 2011, 2013; McCulloch et al. 2012). Experiments have established that calcification in hermatypic scleractinian corals is enhanced by light (Wainwright 1963; Chalker and Taylor 1975; Barnes 1982; Gattuso et al. 1999; Marubini et al. 2001; Allemand et al. 2004; Muscatine et al. 2005) and photosynthesis (Goreau and Goreau 1959; Gattuso et al. 1999; Allemand et al. 2004). This may confound the NCC–Ωₐ relationship observed in the field. Recent experiments have suggested that nutrient uptake and heterotrophic feeding also impact calcification rates (Marubini and Davies 1996; Houbreque et al. 2003) and may in fact decrease the sensitivity of calcification to Ωₐ (Langdon and Atkinson 2005; Silverman et al. 2007a, b; Cohen and Holcomb 2009). Although the mechanistic links between light, nutrition and calcification are not yet fully understood, it is clear that the impacts of multiple environmental factors on both calcification and dissolution must be considered when investigating the impact of OA on NCC in an observation-based field study.

While mesocosm studies have demonstrated that NCC is sensitive to changes in Ωₐ, confirming this dependence in the natural environment is challenging. The large differences in the correlations observed at different sites has highlighted the need for a deeper understanding of environmental controls on metabolic rates at various timescales before extrapolating correlations globally or over several decades (Pandolfi et al. 2011; Shamberger et al. 2011; Andersson and Gledhill 2013). Each empirical NCC–Ωₐ correlation is influenced by several factors including the relative rates of NCC and net community production (NCP) (Andersson and Gledhill 2013) and contemporaneously variable temperature, light, and nutrition (or feeding) levels. The relationship determined for short-term local studies can also be affected by the fluctuating residence time of the water. For example, water that is exposed to a certain NCC or NCP for a long time will exhibit lower Ω than water exposed to that same NCC or NCP for a shorter period of time (Shaw et al. 2012; Zhang et al. 2012; Falter et al. 2013; McMahon et al. 2013). In order to gain a predictive understanding of how coral reefs worldwide will respond to OA, we must investigate the relationships between metabolic rates and environmental conditions in diverse regions, reef settings (e.g., fringing reef or outer shelf reef), and reef zones (e.g., fore-reef, reef flat, lagoon) (Kleypas and Langdon 2006; Atkinson and Cuet 2008).

In this study we estimated NCC and NCP rates on Sheltered Reef, a platform reef on the mid-shelf of the Red Sea near Jeddah, Saudi Arabia. The goal of this study was to examine the relationship between NCC and NCP and the physiochemical environment on an hourly timescale and to compare the results with those from previously published studies in the Red Sea and other regions. A study of NCC in the Red Sea is of particular interest in comparison with similar studies elsewhere, because the Red Sea has relatively high temperature (mean global sea temperature is 27.6 °C, mean for this study was 29 ± 0.9 °C) and Ωₐ (mean global Ωₐ is 3.8, mean for this study was 4.6 ± 0.1) (Kleypas et al. 1999, Silverman et al. 2007b).

### Methods

#### Study site

Our study reef is a small (275 × 125 m²) platform reef within the Quita Dukais offshore reef platform in the eastern Red Sea, north of Jeddah (21°59′N, 38°51′E, Fig. 1). We named it Sheltered Reef (SR) because it is on the leeward side of a larger reef. Samples were collected on 7 and 8 December 2010, and 21–23 May 2011. The temperature around Quita Dukais ranges from 25 °C in February to 31 °C in August and is about 28 °C in both December and May (Fig. 2). Light levels are at a minimum in December and reach a maximum in May (Fig. 2). The water is oligotrophic (Table 1).

SR has rich coral cover on the steep walls and rim. The reef flat is about 1 m deep (Fig. 3) with maximum tidal range of approximately 30 cm and hosts a community consisting of 41 % rhodoliths (free-living coralline algal crusts; Foster 2001; Donnan and Moore 2003; Fig. 4b), 28 % algal turf (non-calcifying algae), 15 % crustose...
coralline algae, 8% sand, and 5% live coral (Fig. 4a). The corals present are mainly *Stylophora* spp., *Porites* spp., and *Platygyra* spp.

SR is a particularly interesting site because the community is largely composed of coralline algae, in both rhodolith and encrusting forms. This group of calcifying organisms fulfills many important functional roles in coral reef systems. A rhodolith bed is a complex three-dimensional matrix that provides habitat for numerous associated invertebrates and macroalgae (Foster 2001; Donnan and Moore 2003). Crustose coralline red algae serve as a settling cue for juvenile coral recruits (Morse et al. 1994; Birrell et al. 2008). Crustose coralline algae also cement and consolidate the reef foundation, supporting the construction of diverse habitats, guarding against erosion, and in some cases serving as the principal driver of carbonate accumulation on reefs (Bjork et al. 1995). Coralline algae may be more susceptible to ocean acidification than coral with aragonite skeletons because they are made of high-magnesium calcite (HMC), and biogenic HMC is typically a more soluble carbonate mineral than aragonite (Bischoff et al. 1987; Morse et al. 2006; Anthony et al. 2008; Jokiel et al. 2008; Kuffner et al. 2008; Martin and Gattuso 2009). This study is one of few that have examined calcification in natural communities dominated by coralline algae (Chisholm 2000).

**Determination of metabolic rates**

We estimated daytime NCC and NCP rates at SR using an Eulerian flow respirometry method (Odum 1956; Langdon et al. 2010), in which we compared the alkalinity and dissolved inorganic carbon (DIC) concentration of the upstream open ocean end-member (unlabeled white points, Fig. 1) and the reef flat end-member (points A, B, and C, Fig. 1). We used the changes in salinity-normalized alkalinity and salinity-normalized DIC together with estimates of current speeds and reef geometry to calculate

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1 Normalization to constant salinity (Normalized Alkalinity = Alkalinity × 40/Salinity) removes the effects of evaporation and precipitation on alkalinity and DIC. This should not make a large difference in most reef settings, but we chose to do this because we were confident in our salinity measurements (accuracy = 0.001 PSU, resolution 0.0002 PSU), so there was little danger of confounding the results with faulty salinity measurements.
Table 1  Maximum and minimum levels in incoming short wave radiation (PAR, μE m$$^{-2}$$ s$$^{-1}$$) in December and May. Nitrate and phosphate concentrations (μM), temperature (°C), aragonite saturation state ($\Omega_a$), partial pressure of CO$_2$ (pCO$_2$, μatm)$^a$ and pH$^a$ were similar on the reef flat in December and May. Values listed as mean ± standard deviation, except net community calcification (NCC) and net community productivity (NCP) values (mmol m$^{-2}$ h$^{-1}$) listed with standard error

|       | n  | Temperature °C | $\Omega_a$ | pCO$_2$ μatm | pH$^a$ | PAR$^b$ μE m$^{-2}$ s$^{-1}$ | [NO$_3$] μM | [PO$_4$] μM | NCC mmol m$^{-2}$ h$^{-1}$ | NCP mmol m$^{-2}$ h$^{-1}$ |
|-------|----|----------------|------------|---------------|-------|-----------------|-------------|-------------|-----------------|-----------------|
| December | 12 | 29.6 ± 0.3     | 4.6 ± 0.3  | 340 ± 50      | 8.10 ± 0.05 | 1350 ± 130      | 0.2 ± 0.1   | 0.04 ± 0.03   | 8 ± 3           | 21 ± 7          |
| May    | 42 | 28.4 ± 0.6     | 4.5 ± 0.3  | 340 ± 40      | 8.10 ± 0.03 | 1550 ± 660      | 0.2 ± 0.2   | 0.1 ± 0.03    | 11 ± 1          | 44 ± 4          |
| Average of December and May | 2 | 29.0 ± 0.9     | 4.6 ± 0.1  | 340 ± 2       | 8.10 ± 0.00 | 1440 ± 130      | 0.2 ± 0.0   | 0.07 ± 0.04   | 9 ± 2           | 33 ± 12         |

$^a$ Calculated from alkalinity and DIC using the CO2SYS program (Pierrot and Wallace 2006), applying the total pH scale (mol kg$^{-1}$ sea water), the carbonate species dissociation constants of Mehrbach et al. (1973) as refit by Dickson and Millero (1987), and the aragonite solubility constant of Mucci (1983)

$^b$ Average of short wave radiation flux was integrated over 1100–1300 h on the three sampling days. The flux of PAR is estimated as 43 % of total incoming radiative energy flux (Baker and Frouin 1987) which is divided by 0.21 (Onsetcomp.com) to convert W m$^{-2}$ to μE m$^{-2}$ s$^{-1}$

NCC and NCP rates, respectively. By budgeting these two parameters, we are able to estimate the net metabolic rates.

The two reactions of interest are calcification and photosynthesis. Calcification results in the loss of two equivalents of alkalinity and one mole of DIC for each mole of CaCO$_3$ produced:

$$\text{Ca}^{2+}_{(aq)} + \text{CO}_3^{2-}_{(aq)} \leftrightarrow \text{CaCO}_3(s)$$

(Reaction1)

In contrast, photosynthesis results in a decrease in DIC by one mole and a negligible change in alkalinity [ΔALK:ΔDIC = 28:550, for photosynthesis on coral reefs (Atkinson and Smith 1983; Atkinson and Falter 2003)] for each mole of organic carbon produced:

$$550 \text{CO}_2 + 30 \text{NH}_4^+ + \text{HPO}_4^{2-} \leftrightarrow 550 \text{H}_{2}\text{O} + 30 \text{P} + 500 \text{O}_2 + 28 \text{H}^+$$

(Reaction2)

where PQ is photosynthetic quotient or the moles of O$_2$ produced for each mole of CO$_2$ assimilated.

Therefore, the budget for alkalinity describes NCC:

$$\text{NCC} = 0.5 \rho_w h \frac{\Delta A_T}{\Delta t} - 0.5 \rho_w u h \frac{\Delta A_T}{L}$$

(1)

where NCC is the instantaneous net calcification rate (mmol m$^{-2}$ h$^{-1}$). NCC is the sum of the rate of change of the total alkalinity ($\Delta A_T$) inventory along the transect plus the advective flux of $A_T$ into and out of the transect, assuming the diffusive flux is small (Falter et al. 2008). Again, we have assumed that the diffusive flux is small (Falter et al. 2008). The terms in Eq. 1 are analogous to those in Eq. 1, except the change in DIC is corrected by 0.5ΔA$_T$ to account for the change in DIC that derives from the calcification process (Reaction 1). $F_{ASGE}$ is the flux of carbon due to the air–sea gas exchange of CO$_2$. However, because CO$_2$ equilibrates slowly with the atmosphere this term is negligible (<0.35 mmol m$^{-2}$ h$^{-1}$) (Wanninkhof 1992; Frankignoulle et al. 1996; Sweeney et al. 2007) and is not included in the calculation.

The uncertainty for NCC and NCP is calculated using the differential method (the relative error is the square root of the sum of the squares of the relative errors) to propagate the uncertainty of all input parameters, $\Delta A_T$, $\Delta DIC$, $u$, $h$, L, addressed individually below. This uncertainty for both NCC and NCP is about 24 % [2 mmol m$^{-2}$ h$^{-1}$ for NCC and 12 mmol m$^{-2}$ h$^{-1}$ for NCP (Table 1)]. The values estimated by this method represent an average over an appreciable area due to lateral mixing of reef waters (Kinsey 1985).

Sample collection and analysis

Water samples for determination of DIC, alkalinity, salinity, and nutrients were collected from the reef–water interface using a hand-held Niskin sampler. We measured DIC and alkalinity using a Marianda VINDTA-3C analysis system, in Dr. Daniel McCorkle’s lab at Woods Hole Oceanographic Institution (WHOI). Alkalinity was
determined by nonlinear curve-fitting to data obtained by open-cell titrations, and DIC concentrations were determined by coulometric analysis. Both measurements were standardized using certified reference materials obtained from Dr. A. Dickson at the Scripps Institution of Oceanography. The analytical precision for alkalinity based on replicate samples was 1.2 μmol kg\(^{-1}\) \((n = 90\) pairs\), and for DIC was 3.4 μmol kg\(^{-1}\) \((n = 90\) pairs\). Salinity was measured using a Guildline salinometer at WHOI (accuracy = 0.001 PSU, resolution 0.0002 PSU; D. Wellwood, pers. comm., March 2013).

We sampled the open ocean end-member from the waters surrounding the reef (unlabeled white points in Fig. 1c) in the morning and afternoon of each day. We evaluated spatial variability in the open ocean end-member by collecting water from 1-m depth at three different locations within the open ocean region on each morning of three consecutive days in December. The variability between these locations for alkalinity and DIC was less than the analytical precision of the measurement (Electronic Supplementary Material, ESM, Table S1). Therefore, although we always aimed to sample upstream of the reef each morning and afternoon, we assumed the open ocean end-member to be uniform in space.

We sampled on the reef flat at three points along the long axis of the reef (points A, B, C in Fig. 1c) twice per day for 2 d in December, and four to five times per day for 3 d in May. To calculate NCC and NCP, we compared the reef flat values to the open ocean values that corresponded most closely to the sampling time of each reef flat end-member. This yielded two to five NCC and NCP estimates at each point each day. Samples were collected between 0900 and 1600 h, during peak sunlight hours.

**Input parameters**

The calculations of NCC and NCP require input of the water velocity \((u)\), water depth \((h)\), and the distance \((L)\) over which the water traveled across the reef flat. A 2 MHz Nortek Aquadopp Profiler, located at point B (Fig. 1c), sampled the current profile for 4 min at hourly intervals with 2.5 cm vertical resolution, and 1 s temporal resolution. This sampling program was designed for a separate year-long study, and the data were generously provided to us by our colleagues, Drs. Steve Lentz and James Churchill. We understand that this sampling resolution was not ideal for our study and may have introduced aliasing problems. When determining cross-reef transport, we based the calculation on the linear interpolation between sampling bursts. The depth-averaged Stokes drift (wave transport) was negligible in May (\(~0.1\) cm s\(^{-1}\)) and only 0.3–1 cm s\(^{-1}\) in December (S. Lentz, pers. comm. June 2013); therefore, Stokes drift was not included in calculations of cross-reef transport, and is a source of additional uncertainty for the four points in December. Because the water was shallow (\(~1\) m) and the benthic topography was rough, we assumed that the water column was well mixed, and we used the vertically integrated water velocity (analytical error 0.1 cm s\(^{-1}\)).

The changes in depth were measured using a Seagaue Wave and Tide Recorder (SBE 26 plus). The Seagaue was located at the northern end of the reef, so the raw data were corrected by 21 ± 5 cm to account for the depth of the Aquadopp relative to the Seagaue. We used the water depth and water velocity at the Aquadopp because the product of depth and velocity is equal everywhere on the reef flat when flow is non-divergent and non-convergent.

The distance, \(L\), was estimated as the length from the sampling point to the reef edge, following the direction of flow at the sampling time. The reef edge was defined by a GPS track generated by swimming the perimeter of the reef with a hand-held GPS unit (white loop in Fig. 1, black loops in Fig. 5). The distance was calculated from each sampling point to the edge of the reef following the direction of flow described by the velocity measurements. The error in distance was estimated as the difference between subsequent estimates. We estimated a minimum error of 5 m to account for uncertainties in our knowledge of the exact positions along the reef edge and positions at points A, B, and C. An additional source of uncertainty arises from the fact that a given parcel of water may follow a meandering path and may experience changes in speed as it moves across the reef. However, because we did not follow a parcel of water, we were compelled to make the simplest assumption that the water followed a linear path across the reef (Fig. 5). Using the assumption of linear flow yields a minimum estimate of \(L\), a maximum estimate of metabolic rate.

**Environmental parameters**

\(\Omega\), temperature, light, and nutrient (ammonium, nitrate, nitrite) concentrations all influence NCC and NCP (Kleypas and Langdon 2006; Kleypas et al. 2006). In addition, nitrogen fixation and consumption of dissolved and particulate organic matter can also be important sources of nutrition for reef communities (Kinsey 1985; Erez 1990; Ribes et al. 2003). Inorganic nutrient concentrations were measured at Oregon State University using a continuous segmented flow system consisting of a Technicon AutoAnalyzer II (SEAL Analytical) and an Alpkem RFA 300 Rapid Flow Analyzer (Alpkem), as described in Aprill and Rappe (2011). The precisions for nitrite and nitrate + nitrite were 0.02 and 0.15 μM, respectively.

\(\Omega\) was calculated from the measured alkalinity, DIC, temperature, and salinity. Although the reef in this study is
Table 2  Instantaneous net community calcification (NCC) and net community productivity (NCP) at Sheltered Reef in December 2010 and May 2011 (mmol m\(^{-2}\) h\(^{-1}\))

| Date       | Time  | Color Code | Point | NCC se | NCP se | Ω   | Temp °C | PAR (μE m\(^{-2}\) s\(^{-1}\)) |
|------------|-------|------------|-------|--------|--------|------|---------|-----------------------------|
| 7-Dec-10   | 11:50 | Yellow     | A     | 13     | 6      | 23   | 11      | 5.1 29.9                    |
|            |       |            | B     | 3      | 2      | 12   | 12      | 4.5 29.7                    |
|            |       |            | C     | 2      | 5      | 10   | 5       | 4.3 29.5                    |
|            | 12:37 | Blue       |       |        |        |      |         | 1193                        |
|            | 13:25 |           |       |        |        |      |         |                             |
| 8-Dec-10   | 11:45 | Yellow     | A     | 4      | 1      | 12   | 5       | 4.7 28.8                    |
|            |       |            | B     | 19     | 3      | 53   | 15      | 4.7 29.5                    |
|            |       |            | C     | 4      | 1      | 17   | 3       | 4.2 29.4                    |
|            | 12:27 | Blue       |       |        |        |      |         | 1359                        |
|            | 13:10 |           |       |        |        |      |         |                             |
| 21-May-11  | 10:20 | Red        | A     | 5.1    | 0.1    | 16.2 | 0.2     | 4.4 27.9                    |
|            |       |            | B     | 19     | 3      | 49   | 8       | 4.6 28.2                    |
|            |       |            | C     | 13     | 4      | 47   | 10      | 4.4 27.8                    |
|            | 10:50 | Yellow     |       |        |        |      |         | 1652                        |
|            | 11:20 | Green      | A     | 13     | 4      | 52   | 21      | 4.4 28.1                    |
|            |       |            | B     | 9      | 2      | 36   | 9       | 4.7 28.5                    |
|            |       |            | C     | 23     | 11     | 64   | 27      | 4.8 28.6                    |
|            | 12:50 | Blue       |       |        |        |      |         | 1960                        |
|            | 13:20 |           |       |        |        |      |         |                             |
| 22-May-11  | 10:20 | Red        | A     | 7      | 1      | 24   | 17      | 4.1 27.6                    |
|            |       |            | B     | 8      | 1      | 14   | 8       | 4.1 27.6                    |
|            |       |            | C     | 7      | 2      | 23   | 17      | 4.1 27.6                    |
|            | 10:50 | Yellow     |       |        |        |      |         | 482                         |
|            | 11:20 | Green      | A     | 6.6    | 0.4    | 45   | 16      | 4.3 27.8                    |
|            |       |            | B     | 6      | 1      | 32   | 14      | 4.2 27.8                    |
|            |       |            | C     | 5      | 1      | 39   | 7       | 4.2 27.8                    |
|            | 12:25 | Blue       |       |        |        |      |         | 565                         |
|            | 13:05 |           |       |        |        |      |         |                             |
|            | 13:45 | Purple     | A     | 13     | 2      | 70   | 2       | 4.4 28.0                    |
|            |       |            | B     | 17     | 1      | 72   | 3       | 4.5 28.2                    |
|            |       |            | C     | 16     | 1      | 67   | 10      | 4.4 28.2                    |
|            | 14:50 |           |       |        |        |      |         | 983                         |
| 23-May-11  | 10:00 | Red        | A     | 13     | 2      | 48   | 4       | 4.6 28.6                    |
|            |       |            | B     | 17     | 3      | 60   | 15      | 4.7 28.9                    |
|            |       |            | C     | 21.1   | 0.2    | 22   | 5       | 4.2 28.5                    |
|            | 10:35 | Yellow     |       |        |        |      |         | 1775                        |
|            | 11:10 | Green      | A     | 3      | 3      | 13   | 9       | 4.6 28.7                    |
|            |       |            | B     | 12     | 4      | 44   | 16      | 4.9 29.0                    |
|            |       |            | C     | 16     | 4      | 63   | 5       | 4.7 29.0                    |
|            | 11:52 | Blue       |       |        |        |      |         | 1930                        |
|            | 12:35 |           |       |        |        |      |         |                             |
|            | 13:10 | Purple     | A     | 6      | 2      | 22   | 9       | 4.4 28.5                    |
|            |       |            | B     | -2.1   | 0.1    | 1    | 2       | 4.7 28.8                    |
|            |       |            | C     | 0.89   | 0.04   | 10   | 3       | 5.0 29.6                    |
|            | 13:45 |           |       |        |        |      |         | 1802                        |
|            | 14:15 | Blue       | A     | 10     | 2      | 46   | 10      | 4.4 28.6                    |
|            |       |            | B     | 12     | 3      | 43   | 9       | 4.5 28.8                    |
|            |       |            | C     | 16     | 7      | 68   | 31      | 4.9 29.3                    |
|            | 14:45 |           |       |        |        |      |         | 1528                        |

'se' is standard analytical error. Colors correspond to the approximate times of sampling transects in Fig. 5. Actual transect times are indicated in gray. Sequential transects were averaged when adding the advection and time-dependent terms, resulting in a new time assignment as indicated in black and illustrated by each data set straddling the color bars for each transect. Aragonite saturation state (Ω), temperature (Temp °C) and photosynthetically available radiation (PAR, μE m\(^{-2}\) s\(^{-1}\)) are also listed.

The advection terms for NCC and NCP at point A at 1325 h on 7 December are more than four standard deviations above the means (60 and 177 mmol m\(^{-2}\) h\(^{-1}\), respectively). The advection data are non-normally distributed with these outliers included, but distributions are normal when these outliers are excluded. The lowest NCC advection value (−5 mmol m\(^{-2}\) h\(^{-1}\)) also occurs at the same time, but at point C. By conditional logic it is unlikely for this to randomly occur on the exact same date and time as the anomalously highest value. Also, it is unlikely that such a high rate of net dissolution would occur in the very middle of the day when all of the other data from all other times exhibit positive net calcification. For these reasons, we suspect something was amiss with the sample at point C of the second transect on 7 December, and excluded this value as well. The text describes results excluding the two outliers for advection terms for NCC and the single outlier for NCP.
primarily composed of coralline algae that produce HMC, we calculated $\Omega$ with respect to aragonite because studies have found that biogenic Mg calcites exhibit a wide range of solubilities even for Mg calcites of similar Mg content (Morse et al. 2006). Therefore, we used $\Omega_a$, which is most relevant to the previously published literature. The calculation was carried out using the CO2SYS program (Pierrot and Wallace 2006), applying the carbonate species dissociation constants of Mehrbach et al. (1973) as refit by Dickson and Millero (1987), and the aragonite solubility constant of Mucci (1983). The values of $\Omega_a$ reported here can be multiplied by a factor of 0.8–1.0 to approximate $\Omega_{HMC}$ (Morse et al. 2006).

Temperature was measured using several tools, including a YSI sonde; a conductivity, temperature, depth (CTD) logger; and numerous Hobo temperature loggers. Incoming short wave radiation (SWR) was measured in W m$^{-2}$ at a meteorological tower located 43 km away on the campus of King Abdullah University of Science and Technology (22°17.823′N, 39°05.567′E).

Results

Measurements

The average photosynthetically active radiation (PAR) from 1100 to 1300 h was 1350 ± 130 W m$^{-2}$ in December and 1530 ± 660 W m$^{-2}$ in May (mean ± SD) (Table 1). The measured reef flat temperatures were approximately 29 °C on the sampling days in both seasons (Table 1), and the salinity was approximately 39 PSU. The average reef flat $\Omega_a$ was approximately 4.6 in both seasons (Table 1).

Average nitrate and phosphate concentrations were 0.2 ± 0.0 and 0.07 ± 0.04 µM (mean ± SD), respectively ($n = 2$ seasons, see Table 1 for seasonal averages). These values are typical of coral reef systems worldwide (nitrate typical range: 0.05–0.5 µM, phosphate typical range: 0.05–0.3 µM; Atkinson and Falter 2003).

The input parameters for calculating NCC and NCP were water depth, current speed, and length of reef substrate traversed. Average water depth was just under 1 m (Fig. 3). The water speed ranged from 0 to 6 cm s$^{-1}$ and was highly variable (Fig. 3). The lengths of reef over which the water passed ranged from 20 to 200 m (Fig. 5).

NCC and NCP

Daytime hourly NCC and NCP were estimated at three points on the reef flat (Fig. 1c, points A, B, C) during each season. The daytime NCC rates ranged from 2 to 19 mmol m$^{-2}$ h$^{-1}$ in December and from −2 to 23 mmol m$^{-2}$ h$^{-1}$ in May (Table 2). The daytime NCP rates ranged from 10 to 53 mmol m$^{-2}$ h$^{-1}$ in December and from 1 to 72 mmol m$^{-2}$ h$^{-1}$ in May (Table 2). The average daytime NCC for the reef was $8 \pm 3$ mmol m$^{-2}$ h$^{-1}$ in December and $11 \pm 1$ mmol m$^{-2}$ h$^{-1}$ in May (Table 1). Integrating over 12-h days and assuming nighttime NCC around zero (Yates and Halley 2003; Shamberger et al. 2011; Albright et al. 2013), this equates to NCC of $91$ mmol m$^{-2}$ d$^{-1}$ in December and 129 mmol m$^{-2}$ d$^{-1}$ in May. These values are within the range of previously published studies (Table 3; ESM Table S2), and at the extremes of the range ($110 \pm 19$ mmol m$^{-2}$ d$^{-1}$) of the long-term “standard performance” for coral/algal flats between 23°S and 21°N (Kinsey 1983).

The average daytime NCP for the reef was $21 \pm 7$ mmol m$^{-2}$ h$^{-1}$ in December and $44 \pm 4$ mmol m$^{-2}$ h$^{-1}$ (mean ± SE) in May (Table 1). These are within the range observed in previously published studies (Table 3; ESM Table S2).

Regressions

NCC was strongly correlated with NCP ($r^2 = 0.71$, $p < 0.0001$; Table 4; Fig. 6). The correlation between NCC and $\Omega_a$ was also significant ($p = 0.05$; Fig. 7; Table 4). However, the variance explained was low due to the large scatter ($r^2 = 0.1$; Table 4). Both NCC and NCP showed a weak but significant correlation with nitrate concentration (Fig. 8). Regressions of both NCC and NCP against light and temperature were weak and insignificant (Table 4).

Discussion

Seasonal differences

Both NCC and NCP were higher in May (11 ± 1 and 44 ± 4 mmol m$^{-2}$ h$^{-1}$, respectively) than in December (8 ± 3 and 21 ± 7 mmol m$^{-2}$ h$^{-1}$, respectively). While temperature, nutrient concentrations, and $\Omega_a$ showed similar values in both December and May (Table 1), seasonally averaged light showed a larger difference between the seasons (Table 1; Fig. 2) and is a plausible driver of the difference in NCC and NCP between the sampling seasons. It is plausible that temperature is also a driver of metabolic variability on seasonal timescales, but we were not able to test this because the maximum variability in temperature did not coincide with our sampling seasons (Fig. 2). Long-term studies with higher-resolution (i.e., weekly or monthly) sampling to capture the full intra- and inter-seasonal variability would be required to quantify the relationships between metabolic rates and light, temperature, and nutrient concentrations on a seasonal timescale.
Comparison with previously published data

The NCC and NCP values obtained for SR are within the range of values obtained for coral reefs worldwide (Table 3; ESM Table S2). The range in average daytime metabolic rates reported over the past three decades appears to be quite large (0–18 mmol m$^{-2}$ h$^{-1}$ for NCC, and −2 to 100 mmol m$^{-2}$ h$^{-1}$ for NCP). This large range may be due to differences in $\Omega_a$ or community structure (including calcifying organism, sediments, carbonate framework, and the organisms involved in dissolution) and indeed, when comparing metabolic rates at different reefs, care must be taken to consider the community composition, environmental conditions, and methods used (Kinsey 1985; Atkinson and Cuet 2008).

NCC-$\Omega_a$ relationship and confounding factors

The relationship between NCC and $\Omega_a$ is of keen interest because the x-intercept of the relationship represents the threshold $\Omega_a$ at which the community transitions from positive to negative net production. A coral reef cannot...
Persist indefinitely if it is not able to sustain positive net production of calcium carbonate in the long term. Predicting long-term impacts of ocean acidification by extrapolating from short-term studies is problematic for three main reasons. First, the timescale of observation (hourly, daily) is typically very short relative to the timescale for which predictions are needed (annual to decadal). Thus, one must recognize the implicit assumption in basing long-term predictions on short-term observations that the relationship between NCC and $\Omega_a$ is the same on both short and long timescales (Langdon et al. 2000). Second, the range of $\Omega_a$ values encountered in a short-term study is likely to be very small and not extend to the values that are predicted for the end of the century (Cyronak et al. 2013; Eyre et al. 2014). The fact that the x-intercept from mesocosm studies is significantly lower, $1.2 \pm 0.2$ (Table 5). The broad range in $\Omega_a$ at which NCC = 0 might be explained by differences in dissolution rates between sites, and between field sites and mesocosms. The greater the long-term (annual) average dissolution rate, the greater the long-term average $\Omega_a$ has to be for a positive rate of NCC to be achieved over long timescales (years–decades) (Cyronak et al. 2013; Eyre et al. 2014). The fact that the x-intercept from the mesocosm studies is lower than that from field studies might reflect lower dissolution rates in mesocosms because they do not replicate the environments and biota where dissolution is likely to be most active, i.e., in the sediments and the reef framework. If this interpretation is correct, it would follow that the dissolution rate is much higher at the reef in Japan studied by Ohde and van Woesik (1999) and much lower at Lady Eliot Island Reef studied by Shaw et al. (2012).

It is also interesting to compare the slopes of the NCC versus $\Omega_a$ relationships. It is useful to apply a normalization before attempting such a comparison because the absolute rate varies considerably due to differences in abundance and community composition of the calcifiers found at each study site, and due to the differences in carbonate framework and sediment environments and biota responsible for dissolution. Borrowing from the literature on laboratory experiments with corals, a commonly used normalization is to express the rates as a percentage of the rate at some reference saturation state, such as the pre-industrial value 4.6 (Langdon and Atkinson 2005; Kleypas and Langdon 2006; Chan and Connolly 2013). In a meta-analysis of 30 laboratory studies, Chan and Connolly (2013) found that these factors positively influence NCC, then plotting NCC against $\Omega_a$ will lead to the false conclusion that too much of the change in NCC is explained by $\Omega_a$ (i.e., the slope of the NCC–$\Omega_a$ relationship will be overestimated; Venti et al. 2014). In contrast, if the co-varying factor has a negative effect on NCC the plot of NCC against $\Omega_a$ will underestimate the true slope of the NCC–$\Omega_a$ relationship. Increasing the duration of field studies, improving the precision of the NCC measurements, and measuring a suite of environmental parameters (temperature, light, NCP, community composition, etc.) will be necessary to address these issues.

NCC–$\Omega_a$ relationship compared to previously published studies

In this study, $\Omega_a$ where NCC = 0 (intercept) was $3.1 \pm 2.7$ (SE) (Table 5). This parameter has been found to vary widely between studies from a low of $1.2 \pm 1.6$ (Shaw et al. 2012) to a high of $4.9 \pm 0.3$ (Ohde and van Woesik 1999). The mean of this parameter for all field studies is $2.9 \pm 0.5$ (SE) (Table 5). It is interesting that the x-intercept from mesocosm studies is significantly lower, $1.2 \pm 0.2$ (Table 5).
the average response of corals was a $15 \pm 8 \%$ decrease in calcification per unit change in $\Omega_a$. In this study, NCC was found to be considerably more sensitive to $\Omega_a$ with a $68 \pm 33 \%$ change in calcification per unit change in $\Omega_a$ (Table 5). The sensitivity to $\Omega_a$ varies from a low of $30 \pm 11 \%$ to a high of $85 \pm 8 \%$ per unit change in $\Omega_a$ in field studies (Table 5). The fact that coral reef communities in nature seem to be more sensitive to a change in $\Omega_a$ than individual corals in a laboratory setting (where carbonate sand or framework are absent) is consistent with the finding...
that dissolution may be more sensitive than calcification to declining $\Omega_a$ (Cyronak et al. 2013; Eyre et al. 2014). It is worth noting that mesocosms seem to capture an intermediate response with sensitivities in the range of 27–34 % per unit change in $\Omega_a$.

There are several possible reasons that coral reef communities in nature might exhibit a higher sensitivity to a change in $\Omega_a$ than in laboratory and mesocosm studies. One obvious reason is that laboratory studies are generally performed under optimal or near-optimal light and temperature conditions. It is quite possible that corals in the field experiencing sub-optimal environmental conditions will be less able to cope with the additional stress of reduced pH and will therefore show a greater sensitivity to a reduction in $\Omega_a$. A second reason could be that food scarcity and interactions with other species (competition, predator–prey, disease) could reduce a coral’s energy reserves and hence its ability to devote the extra energy needed to elevate pH from a lower baseline at the site of calcification. Thirdly, the differences in sensitivity of NCC to $\Omega_a$ may be reflecting the limited ability of laboratory and mesocosm experiments to replicate the environments and biota involved in dissolution, and thus the limited ability of those experiments to capture the full sensitivity of dissolution to $\Omega_a$ that is exhibited in the field (Cyronak et al. 2013; Eyre et al. 2014). The fact that laboratory and mesocosm studies may be underestimating both the sensitivity to a reduction in $\Omega_a$ and the critical $\Omega_a$ below which reef framework starts to dissolve is cause for concern as this means that the threat of OA may be more pressing than we previously thought.

**Reconciling the two explanations**

While it is reasonable that both the x-intercept and sensitivity of the NCC–$\Omega_a$ relationship are higher in nature than in the laboratory and in mesocosms for the reasons given above, it is also likely that some of the difference may be explained by the fact that in field studies it is easy to confound the effects of $\Omega_a$ on NCC with other factors that control NCC. It is likely that the truth lies somewhere between the two explanations offered above. The x-intercept and slope in natural coral reef communities may be greater than those found in laboratory and mesocosm studies because of poorly replicated community structure lacking sufficient representation of sand and carbonate framework involved in dissolution. On the other hand, some of the extremely high x-intercepts and steep slopes
from field studies may be artifacts produced by the confounding effects of NCP and light on NCC (Falter et al. 2012). Resolving a clearer understanding of the NCC–$\Omega_a$ relationship requires additional estimates of NCC along with thorough characterization of the full suite of environmental conditions: NCP, $\Omega_a$, temperature, PAR, percent

from Table 3 Average daytime net community calcification (NCC) and net community productivity (NCP) rates (mmol m$^{-2}$ h$^{-1}$) and environmental data, including percent calcifier cover (%CC), aragonite saturation state ($\Omega_a$), and photosynthetically active radiation (PAR, $\mu$E m$^{-2}$ s$^{-1}$) for several field and mesocosm studies

| Reference                          | %CC | Season/treatment          | $\Omega_a$ | T on Reef (°C) | Daily Integrated PAR (E m$^{-2}$ d$^{-1}$) | Daytime NCC mmol m$^{-2}$ h$^{-1}$ | Daytime NCP mmol m$^{-2}$ h$^{-1}$ |
|------------------------------------|-----|---------------------------|------------|---------------|------------------------------------------|-----------------------------------|-----------------------------------|
| This study                         | 61  | December 2010             | 4.62       | 30            | 30                                       | 8                                 | 21                                |
| Falter et al. (2012)               | 61  | May 2011                  | 4.50       | 28            | 37                                       | 11                                | 44                                |
| Falter et al. (2012)               | 70  | Winter 2009               | 4.17       | 24.7          | 20.3                                     | 18                                | N/A                               |
| Falter et al. (2012)               | 70  | Summer 2008               | 3.53       | 23.7          | 40.9                                     | 16                                | N/A                               |
| Shaw et al. (2012)                 | 40  | Feb 2010                  | 4.09       | 28.0          | N/A                                      | 9                                 | N/A                               |
| Shaw et al. (2012)                 | 40  | April 2010                | 4.40       | 26.6          | N/A                                      | 10                                | N/A                               |
| Shaw et al. (2012)                 | 40  | July 2010                 | 3.99       | 23.2          | N/A                                      | 11                                | N/A                               |
| Shamberger et al. (2011)           | 25  | Feb 2009                  | 2.71       | 23            | 28.4                                     | 15.7                              | 3.3                               |
| Shamberger et al. (2011)           | 25  | June 2008, August 2009    | 2.87       | 26.3          | 39.0                                     | 9.9                               | 13.41                             |
| Bates et al. (2010)                | 21  | Jan–April 2003            | 3.08       | 21            | N/A                                      | 5.3                               | N/A                               |
| Bates et al. (2010)                | 21  | July–August 2003          | 3.35       | 29            | N/A                                      | 4.8                               | N/A                               |
| Bates et al. (2010)                | 21  | September–December 2002   | 3.26       | 24            | N/A                                      | 2.4                               | N/A                               |
| Silverman et al. (2007a, b)        | 10  | Winter 2000–2002          | 3.98       | 21.8          | 17.7                                     | 2.3                               | 14                                |
| Silverman et al. (2007a, b)        | 10  | Summer 2000–2002          | 3.97       | 24.2          | 20.6                                     | 2.5                               | 17                                |
| Yates and Halley (2006)            | 10  | Feb 2000                  | 3.00       | 26.18         | 40.4                                     | 0.48                              | 3.52                              |
| Yates and Halley (2006)            | 22  | Feb 2000                  | 3.05       | 25.43         | 49.4                                     | 1.73                              | 7.79                              |
| Watanabe et al. (2006)             | 16  | April 2000                | 3.78       | 28.3          | N/A                                      | 6.0                               | —0.23                             |
| Watanabe et al. (2006)             | 16  | September 2000            | 3.83       | 29.4          | 32.4                                     | 6.1                               | N/A                               |
| Ohde and van Woesik (1999)         | 36  | October/September 1993–1995 | 5.38      | 30.38        | N/A                                      | 7.07                              | 9.98                              |
| Ohde and van Woesik (1999)         | 36  | June/July 1994            | 5.80       | 31.30         | N/A                                      | 12                                | 16                                |
| Gattuso et al. (1997)              | 15  | July 16–17 1992           | 3.61       | 27.3          | 35.6                                     | 3.25                              | 15                                |
| Gattuso et al. (1996)              | 29  | July, August 1992, Austral winter | 5.38 | 27.1 | 43.2 | 13.5 | 70 |
| Gattuso et al. (1996)              | 40  | December 1993, Austral summer | 5.04 | 27.2 | 43.2 | 17.5 | 100 |
| Andersson et al. (2009)            | 25  | Control June 2006         | 3.33       | 27.5          | N/A                                      | 6.033                             | N/A                               |
| Andersson et al. (2009)            | 25  | Treatment June 2006       | 1.85       | 27.43         | N/A                                      | 1.6                               | N/A                               |
| Langdon and Atkinson (2005)        | 100 | Winter                    | 3.01       | 23.8          | 15.4                                     | 15.4                              | 23                                |
| Langdon and Atkinson (2005)        | 100 | Winter                    | 2.27       | 23.8          | 17.9                                     | 12                                | 32                                |
| Langdon and Atkinson (2005)        | 100 | Winter                    | 1.65       | 24.4          | 18.4                                     | 3                                 | 35                                |
| Langdon and Atkinson (2005)        | 100 | Summer                    | 3.00       | 27.3          | 26.3                                     | 16                                | 45                                |
| Langdon and Atkinson (2005)        | 100 | Summer                    | 1.80       | 27.3          | 24.6                                     | 9                                 | 55                                |
| Langdon et al. (2000)              | 40  | Two long experiments 1995–1998 | 1.60      | 26.5 | N/A | 0.4 | N/A |
| Langdon et al. (2000)              | 40  | Last glacial maximum pCO$_2$ | 3.10       | 26.5 | N/A | 3 | N/A |
| Langdon et al. (2000)              | 40  | Present day pCO$_2$       | 5.20       | 26.5          | N/A                                      | 10                                | N/A                               |
| Langdon et al. (2000)              | 40  | 2 × present day pCO$_2$   | 3.35       | 26.5          | N/A                                      | 4.3                               | N/A                               |
| Langdon et al. (2000)              | 40  | 2 × present day pCO$_2$   | 2.05       | 26.5          | N/A                                      | 2.7                               | N/A                               |

See ESM Table S2 for details on each data source
calcifier cover, and nutrient levels. With broader geographic and temporal coverage, we will then be able to average out the short-term confounding factors and capture a general relationship, with predictive power to understand the impact of long-term OA on NCC.

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### Table 4

|                  | p   | r²  | Slope | Slope error | p   | r²  | Slope | Slope error |
|------------------|-----|-----|-------|-------------|-----|-----|-------|-------------|
| **Temperature**  |     |     |       |             |     |     |       |             |
| NCC              | 0.7 | 0.003 | -0.5 | 1           | 0.1 | 0.06 | 0.003 | 0.002       |
| NCP              | 0.1 | 0.06 | -8    | 5           | 0.7 | 0.004 | 0.003 | 0.007       |
| **Ωₐ**          |     |     |       |             |     |     |       |             |
| NCC              | 0.05 | 0.10 | 7     | 4           | 0.04 | 0.11 | -29    | 13          |
| NCP              | 0.5 | 0.01 | 8     | 13          | 0.02 | 0.13 | -115   | 48          |

The regressions are weak (low r²) and all are insignificant (p > 0.05) except for NCC versus Ωₐ and NCC and NCP versus [NO₃⁻²⁻]

### Table 5

|                  | Slope | y-intercept | x-intercept | NCC at Ωₐ = 4.6 | % ΔNCCₐ/ΔΩₐ |
|------------------|-------|-------------|-------------|-----------------|------------|
| **Field studies**|       |             |             |                 |            |
| This study       | 7.37  | -23         | 3.1 ± 2.7   | 10.9            | 68 ± 33    |
| Falter et al. (2012) | 20.7  | -57.6       | 2.8 ± 1.3   | 37.6            | 55 ± 16    |
| Shaw et al. (2012) | 3.4   | -4.2        | 1.2 ± 1.6   | 11.4            | 30 ± 11    |
| Shamberger et al. (2011) | 10.2  | -17.3       | 1.7 ± 0.8   | 29.6            | 35 ± 8     |
| Silverman et al. (2007b) | 4.32  | -14.8       | 3.4 ± 0.5   | 5.1             | 85 ± 8     |
| Ohde and van Woekik (1999) | 8.5   | -40.9       | 4.9 ± 0.3   | -1.8            | -          |
| Average          |       |             | 2.9 ± 0.5   |                |            |
| **Mesocosm studies** |       |             |             |                 |            |
| Andersson et al. (2009) | 3.4   | -5.3        | 1.1 ± 0.4   | 10.3            | 29 ± 4     |
| Langdon and Atkinson (2005) | 7.8   | -7.3        | 0.9 ± 0.6   | 28.7            | 27 ± 6     |
| Kleypos and Langdon (2006) | 34.1  | -58.7       | 1.7 ± 0.3   | 98.2            | 34 ± 3     |
| LeClercq et al. (2002) | 1.20E-02 | 9.80E-02 | -8.2 | 0.2 | 7.8 |
| Average          |       |             | 1.2 ± 0.1   |                |            |

The x-intercept is the aragonite saturation state (Ωₐ) at which net communacation calcification equals zero (NCC = 0). The percent change (Δ) in NCC per unit change in Ωₐ is also listed, where the NCC rates are expressed as a percentage of the rate at the pre-industrial Ωₐ value of 4.6

a Normalized pre-industrial sensitivity not calculated because 4.6 is outside the range of saturation states observed on this reef

b Only uses the pre-nutrient addition data
c Excluded the x-intercept for LeClercq et al. (2002) because a saturation state below zero is not realistic
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