Effect of Cyanophycin Metabolism in Recombinant Sinorhizobium (Ensifer) meliloti 1021 on the Symbiosis with Alfalfa (Medicago sativa)

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Abstract

The aim of this study was to investigate the effect of the synthesis and degradation of the nitrogen-rich polymer cyanophycin in rhizobia on symbiotic nitrogen fixation and crop yield in legumes. For this, cyanophycin synthetase from Anabaena sp. PCC7120 was expressed in the bacteroids of the symbiont S. meliloti 1021 either alone or together with an intracellular cyanophycinase from the same bacterium, either in the wild type or in a polyhydroxybutyrate-negative (PHB) mutant, and the effect on the growth of alfalfa host plants was studied. All strains induced the formation of nitrogen-fixing nodules in the host, but clear differences in various parameters were noticeable. Alfalfa plants infected with the wild type expressing only cyanophycin synthetase showed significantly lower shoot nitrogen contents and higher nitrogen fixation rates than plants inoculated with wild type, but the wild type phenotype was exceeded in S. meliloti 1021 expressing cyanophycinase in addition to cyanophycin synthetase. Growth of plants infected with the PHB mutant expressing only cyanophycin synthetase was severely impaired in comparison to growth of plants infected with the PHB mutant expressing both or neither of the two proteins. Transmission electron micrographs of sections of nodules induced by wild type S. meliloti 1021 producing cyanophycin synthetase and cyanophycinase showed that the rough endoplasmic reticulum and Golgi membranes were extended further in comparison to the nodules induced by the wild type, the PHB mutant, or all other recombinant S. meliloti strains, indicating higher metabolic activities in these nodules.

Keywords: Cyanophycin; Nitrogen fixation; Rhizobia; Symbiosis; Crop yield; Alfalfa

Abbreviations

ATP: Adenosine Triphosphate; PCR: Polymerase Chain Reaction

Introduction

Cyanophycin (multi-L-arginyl-poly-L-aspartic acid, cyanophycin granule polypeptide or CGP) was discovered over one hundred years ago during light microscopic examinations of cyanobacteria [1]. The polymer consists of equimolar amounts of arginine and aspartic acid arranged as a polyspartate backbone with arginine moieties linked to the β-carboxyl group of each aspartate via a peptide bond [2]. CGP is synthesized in the cytoplasm of most cyanobacteria [3-5] and also of the β-carboxyl group of each aspartate via a peptide bond [2]. CGP is usually insoluble in water at physiological pH, but is soluble under acidic (pH<2) or alkaline (pH>9) conditions [8]. In addition, the polymer consists of equimolar amounts of arginine and aspartic acid arranged as a polyspartate backbone with arginine moieties linked to the β-carboxyl group of each aspartate via a peptide bond [2]. CGP is synthesized in the cytoplasm of most cyanobacteria [3-5] and also of several non-photosynthetic bacteria [6,7] and deposited in the cells as water-insoluble membrane-less granules.

CGP is usually soluble in water at physiological pH, but is soluble under acidic (pH<2) or alkaline (pH>9) conditions [8]. In addition, a water-soluble form of CGP was observed in cells of a recombinant Escherichia coli strain [7]. Intracellular synthesis of CGP is catalyzed by cyanophycin synthetase, which is encoded by cphA [7]. Degradation of CGP is catalyzed by two different types of enzymes acting consecutively: (i) an exopeptidase also referred to as cyanophycinase (CphB), produces β-Asp-Arg dipeptides [9], and (ii) a peptidase that hydrolyzes these dipeptides to the amino acid monomers [10]. A detailed overview of the distribution of CGP metabolism genes and thorough in silico analyses of the respective genes and of the gene products has been published [11]. The genome of Anabaena sp. PCC7120 harbors two cph clusters, each consisting of cphB and cphA. However, it was shown that the cyanophycin synthetase of the first cluster (CphA1) contributed more to the synthesis and accumulation of CGP than the second cyanophycin synthetase (CphA2) [12].

The cyanophycin synthetase genes (cphA) of many cyanobacteria [13-15] as well as those of the chemotrophic bacteria Acinetobacter sp. strain DSM 587 [6] and Desulfotobacterium hafniense DCB-2 [7] were cloned and expressed in E. coli. Heterologous expression of cphA was also demonstrated in recombinant strains ofRalstonia eutropha, Corynebacterium glutamicum, Pseudomonas putida and Bacillus megaterium [16]. CGP has also been produced in a few eukaryotes [17-21].

The Gram-negative soil bacterium Sinorhizobium meliloti fixes atmospheric nitrogen in symbiotic association with its host plants, including Medicago truncatula [barrel medic] and Medicago sativa (alfalfa) [22]. A successful symbiosis involves a series of specific interactions between the legume host and the rhizobial symbiote [22]. Symbiosis is initiated by the plant root by the release of flavonoids, which attract S. meliloti cells from the surrounding environment and induce the formation of nodulation (Nod) factors by the bacteria. The Nod factors then induce root hair curling and root nodule development. S. meliloti cells invade the developing root nodules via infection threads, extended invaginations of the root hair cell membrane [23]. From these, bacteria are then released into the cells of the developing nodule [24]. Once released from the infection threads, the bacterial cells differentiate into bacteroids that are able to convert atmospheric nitrogen into ammonium, which is then delivered to the plant host [25,26].

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nitrogen to ammonia [25]. The host plant supplies bacteroids with carbon compounds, including dicarboxylic and amino acids [26], which are used by the bacteroids to support the reduction of nitrogen to ammonia in amounts sufficient for plant growth [24].

The cyanophycin synthetase and cyanophycinase genes (cphA1 and cphB1) from *Anabaena* sp. PCC7120 were heterologously expressed in wild type *S. meliloti* 1021 and in a phbC-negative mutant of this strain. CGP was accumulated up to 44% (wt/wt) of cell dry weight in the recombinant cells harbouring only *cphA1* alone without *cphB1*. In contrast, no CGP accumulation was observed in cells harbouring both *cphA1* and *cphB1*. The aspartate-arginine dipeptides released from CGP degradation were shown to be used by the cells to fuel PHB biosynthesis [27].

In rhizobia inhabiting indeterminate nodules, such as *S. meliloti*, *M. ciceri*, and *R. leguminosarum* bv. *vicieae*, PHB accumulates in free-living cells, but generally not in bacteroids [28]. The reason for this is unknown, but GC analysis of bacteroids of *M. ciceri* CC1192 has shown that small amounts of PHB are accumulated [29]. PHB granules were also observed in mature bacteroids of *R. leguminosarum* bv. *vicieae* carrying mutations in the broad-specificity amino acids transporters *aat* and *bra* [26]. Blocking the amino acid cycling pathway between the plant and the bacteroid prevents the synthesis of aspartate from oxaloacetate, which increases carbon flow from dicarboxylate to pyruvate, which in turn is channelled into PHB synthesis [26].

This study was aimed at investigating the effect of CGP biosynthesis alone or its accumulation and degradation in the nodules induced by recombinant cells of strains *S. meliloti* 1021 (pBBR1MC5-3::cphA1*<sub>1021</sub>*), *S. meliloti* 1021 phbC<sup>Km</sup> (pBBR1MC5-3::cphA1<sub>1021</sub>*), *S. meliloti* 1021 (pBBR1MC5-3::cphB1<sub>1021</sub>*::cphA1<sub>1021</sub>*), and *S. meliloti* 1021 phbC<sup>Km</sup> (pBBR1MC5-3::cphB1<sub>1021</sub>*::cphA1<sub>1021</sub>*) on the symbiosis with alfalfa plants.

**Materials and Methods**

**Bacterial strains and cultivation conditions**

The bacterial strains and plasmids used in this study are listed in Table 1. Cells of *S. meliloti* were grown at 30°C in Tryptone Yeast (TY) medium [30]. Antibiotics were used at the following concentrations: kanamycin (300 μg/ml), streptomycin (100 μg/ml), and tetracycline (12.5 mg/ml). Aspartic acid and arginine were added as supplements at concentrations of 0.2% (wt/vol). For induction 0.1 mM isopropyl-β-D-thiogalactopyranoside (IPTG) was added.

**Nodulation experiments**

Seeds of alfalfa (*Medicago sativa*) var. "Eugenia" (Reviereberatung Wolmersdorf, Germany) were surface-sterilized by soaking in 70% ethanol for 1 min, followed by 10 min soaking in 1% (vol/vol) freshly prepared solution of sodium hypochlorite and rinsed several times with sterile H<sub>2</sub>O [31]. The treated seeds were germinated on 1% water agar in sealed Petri plates at room temperature in the dark for 48 h.

Germinated seeds were transferred to plant growth tubes (50 ml polypropylene falcon centrifuge tube, Greiner Bio-One GmbH, Germany) containing 25 ml sterile nitrogen-free B+D plant nutrient solution solidified using 1% agar [32] covered with 0.5-1 cm of sterile, washed vermiculite saturated with B+D plant nutrient solution at one seedling per tube. The outer surface of plant growth tubes were covered with aluminium foil surrounding the medium area to provide dark conditions for the plant root. Four days after germination, seedlings were inoculated with the *S. meliloti* strains. The cells were grown for 48 h in TY medium [supplemented with the appropriate antibiotics, 0.1 mM IPTG, 0.2%, wt/vol arginine, and 0.2%, wt/vol aspartic acid, harvested by centrifugation (15 min, 2,800 x g), washed twice with saline solution (0.9%, wt/vol NaCl), and diluted to an optical density at 600 nm of 0.2. Each germinated seedling was inoculated with 0.5 ml of the bacterial suspension, and each of the control seedlings was mock inoculated with 0.5 ml of saline solution (0.9%, wt/vol NaCl). After inoculation, the tubes were wrapped with semi-transparent parafilm (Rio Tinto Alcan, Canada) until the growing plants reached the top of the tube; it was removed afterwards. Each treatment included 48 replicates, where each replicate consisted of one tube containing one plant. The plants were placed in randomized blocks in a growth chamber (16 h at 25°C [day], 8 h at 20°C [night], light intensity 18,000 lux and 60% relative humidity). The inoculated plants were grown for 55 days and watered with nitrogen-free B+D plant nutrient solution as necessary.

**Nitrogenase activity in nodulated roots** [30], shoot length, root length, number of leaves, number of nodules, nodule fresh weight, shoot dry weight and %N (wt/wt) of shoot dry weight [33] were determined after the plants had been harvested. Bacteria were isolated from the nodules at 35 and 55 days. The nodules were surface-sterilized by soaking for five to ten seconds in 95% ethanol, followed by two to four minutes soaking in freshly-prepared 3% (vol/vol) solution of sodium hypochlorite, rinsed several times with sterile H<sub>2</sub>O, then crushed in a few drops of TY medium. The resulting suspension was streaked onto TY agar plates supplemented with the appropriate antibiotics.

**Nitrogenase activity assay**

Nitrogen fixation activity was determined by the acetylene reduction method as follows: Seven millilitres of acetylene gas were injected into stoppered 70-ml bottles containing alfalfa root segments with nodules from 35- or 55-day old plants inoculated with *S. meliloti* stains. The bottles were incubated for 1-3 h before analysis at room temperature. A 100 μl gas sample from each tube was analyzed for the presence of acetylene and ethylene using an Agilent 6850 gas chromatograph (Waldbrohn, Germany) equipped with a BP21 capillary column [0.25 mm diameter, 250 nm film thickness, 50 m length, SGE Darmstadt, Germany] and a flame ionization detector. The flow rate of the hydrogen carrier gas was set at 30 ml/min. The injection temperature was 100°C, and the column temperature was 35°C. Under these conditions, ethylene typically eluted after 4.405 min, while acetylene eluted after 4.530 min.

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**Table 1:** Bacterial strains and plasmids used in this study. Abbreviations: Km′, Kanamycin resistance; Sm′, Streptomycin resistance; Tc′, Tetracycline resistance.

| Strains or plasmids | Relevant characteristics | References or sources |
|---------------------|--------------------------|-----------------------|
| *S. meliloti* Rm1021 Rm4775 | SU47 atv21, Sm′ S. meliloti 1021::phbC<sup>Km</sup> | [47] [27] |
| **Plasmids** | | |
| pBBR1MC5-3::cphA1<sub>1021</sub> | T<sub>c</sub>, mob′, tra, lacPOZ′, cphA1 in addition to carboxy-terminal region of cphB1 from *Anabaena* sp. PCC7120 colinear to lacPOZ′ | [27] |
| pBBR1MC5-3::cphB1<sub>1021</sub>*::cphA1<sub>1021</sub> | pBBR1MC5-3 harbouring cphB1 and cphA1 from *Anabaena* sp. strain PCC7120 | |
DNA manipulation

The following oligonucleotides were used as primers to amplify cphA1-or the cphB1-cphA1-gene cluster by colony PCR from bacterial colonies recovered from surface-sterilized alfalfa nodules induced by the recombinant S. meliloti strains after 35 or 55 days post-inoculation: for amplifying cphA1+ primer P1 (5'-GCTATGGGGATCCCT- CATCTGTACCAAAGG-3') and primer P2 (5'-GTTTTTACCTAGT-GG CCTTCCA-3') were used; for amplifying the cph1 gene cluster (cphB1+cphA1), primer P3 (5'-TTTGGGGACACGGACATA C-3') and primer P2 were used. The oligonucleotides were purchased from Thermo Scientific (Germany). For the amplification reaction Taq polymerase from Invitrogen [San Diego, CA] was used according to the instructions of the manufacturer.

Transmission electron microscopy

Nodules taken 35 and 55 days after inoculation with S. meliloti strains were fixed with 2.5% glutaraldehyde in 0.1 M phosphate-buffered saline (PBS; pH 7.3) according to Sörensen [34] for 1 h. After washing three times with PBS (30 min each), nodules were post-fixed in 1% osmium tetroxide in 0.1 M PBS (pH 7.3) for 1.5 h and washed once with PBS (pH 7.3) for 10 min. The water was then removed by using a graded water/ethanol series (30, 50, 70, 90 and 96% ethanol followed by two changes of absolute ethanol) followed by propylene oxide (30 min each). To obtain thin sections, the dehydrated samples were embedded in Spurr's resin with propylene oxide [35] at the following mixture ratios (1:3 for 4 h, 1:1 for 4 h, 3:1 for 4 h, respectively). The samples were soaked twice in Spurr's resin without propylene oxide for 4-8 h for the first soak and over night for the second soak. The final embedding and polymerization was done in fresh Spurr's resin without propylene oxide overnight at 70°C. After polymerization, longitudinal ultrathin sections with a thickness of 70-80 nm were cut with an ultramicrotome (Leica Mikroskopie und Systeme, Wetzlar, Germany) using a diamond knife (Diatome AG, Biel, Switzerland). The sections were then placed on 200-mesh copper grids that were covered with a thin film of pioloform. Sections were then stained for 30 min with saturated uranyl acetate in 50% ethanol. After brief washing with double-distilled water, the sections were stained for five minutes with lead citrate according to Reynolds [36]. After brief washing with double-distilled water, imaging was performed with an 80 kV electron microscope 'EM Philips 410' (Eindhoven, Netherlands).

Statistical analysis

All data for nitrogenase activity in nodulated roots as well as for shoot length, root length, number of leaves, number of nodules, nodule fresh weight, shoot dry weight, or %N (wt/wt) of shoot dry weight for plants 55 days after inoculation are given as means with standard errors. The statistical significance P<0.05 for root length and number of nodules, P<0.01 for nitrogenase activity, shoot length, number of leaves, nodule fresh weight, shoot dry weight, and %N (wt/wt) of shoot dry weight of the results was assessed using Duncan's t test.

Results

Alfalfa seedlings were inoculated with wild type S. meliloti 1021 (Rm1021), the phbC-negative mutant S. meliloti 1021 phbCQKm (Rm4775), or with recombinant strains harbouring plasmids pBBR1MCS-3::cphA1 and pBBR1MCS-3::cphB1. The inoculated plants were grown for 55 days in a growth chamber under controlled conditions as described in the Materials and Methods section. All strains induced elongate, cylindrical, pink, nitrogen-fixing nodules similar to the nodules induced by the wild type strain.

The results of time course analysis of plant growth, nodule development, and nitrogen fixation of alfalfa plants inoculated with the various S. meliloti strains are shown in Figures 1 and 2. Alfalfa plants inoculated with the phbC-negative mutant Rm4775, which is unable to synthesize PHB, demonstrated an insignificantly [P>0.01] higher acetylene reduction activity 55 days after inoculation in comparison to the acetylene reduction activity levels of plants inoculated with the wild type (Figure 2). However, they showed fewer nodules, significantly [P<0.01] reduced shoot dry mass, and lower shoot nitrogen content (Figure 1 and Table 2). The plants inoculated with the recombinant phbC-negative mutant harbouring cphA1 showed significantly (P<0.01) lower shoot dry mass, shoot nitrogen content, acetylene reduction activity, and nodule fresh weight as well as a significantly (P<0.05) lower number of nodules (Table 2), and also much poorer growth (Figure 3) in comparison to alfalfa plants inoculated with the wild type strain. Furthermore, these plants showed the lowest growth of all inoculated plants in the experiment (Figures 1 and 2) (Table 2).

In contrast, plants inoculated with the recombinant phbC-negative mutant harbouring cphB1 in addition to cphA1 performed very differently. Higher acetylene reduction activity and nodule fresh weights were obtained with plants inoculated with this strain in comparison to plants inoculated with either the wild type strain or the phbC-negative mutant or the recombinant phbC-negative mutant harbouring only cphA1 (Figures 1 and 2). Moreover, the shoot dry weights of these plants were slightly higher than of those infected with the phbC-negative mutant and significantly (P<0.01) higher than of those infected with the recombinant phbC-negative mutant harbouring only cphA1 (Table 2).

The root systems of the plants 55 days after inoculation with the wild type strain showed a significantly higher acetylene reduction activity (32 %) than plants inoculated with the recombinant strain Rm1021 harbouring both cphB1 and cphA1. In contrast, the latter plants exhibited enhanced growth as indicated by (albeit insignificantly, P>0.01) higher nodule fresh weight, shoot dry weight, and shoot nitrogen content. The number of nodules was also (insignificantly, P>0.05) higher (Table 2). Furthermore, the same plants showed an insignificantly (P>0.01) higher shoot dry weight, but a significantly (P<0.01) higher shoot nitrogen content in comparison to the plants inoculated with the recombinant strain of Rm1021 harbouring only cphA1 (Table 2).

On the other hand, the plants inoculated with the recombinant Rm1021 strain harbouring only cphA1 without cphB1 revealed approximately the same shoot dry weights, insignificantly higher number of nodules (P>0.05), higher nodule fresh weight (P>0.01), lower shoot nitrogen content (P>0.01), and significantly (P<0.01) higher acetylene reduction activity in comparison to plants infected with the wild type strain.

Plasmid test assay

A 3,105-bp fragment corresponding to the cphA1 fragment could be amplified by PCR from bacterial colonies recovered from surface-sterilized alfalfa nodules induced by the two recombinant strains Rm1021 (pBBR1MCS-3::cphA1) and Rm4775 (pBBR1MCS-3::cphA1). In addition, a 4,134-bp fragment corresponding to the cphB1-cphA1 fragment could be amplified by PCR from the bacterial colonies recovered from surface-sterilized alfalfa nodules induced by the two recombinant strains Rm1021 (pBBR1MCS-3::cphB1) and Rm4775 (pBBR1MCS-3::cphA1). The presence of these plasmids in at least some bacterial cells during symbiosis 35 or 55 days after inoculation.
Discussion

In this study, we addressed the question whether or not the capability of CGP biosynthesis alone or of a combination of biosynthesis and degradation of CGP improves the capacity of recombinant PHB-positive or PHB-negative *S. meliloti* 1021 strains to fix atmospheric nitrogen, and whether or not the inoculated alfalfa host plants exhibited enhanced growth.

One striking result was that inactivation of *phbC* in the genome of Rm1021 negatively affected symbiosis. The plants, which were inoculated with the *phbC*-negative mutant strain (Rm4775), showed significantly lower shoot dry mass and shoot nitrogen content in comparison to plants inoculated with the wild type strain. This negative effect was also observed by Aneja et al. [38], who reported that alfalfa plants inoculated with a *phbC*-negative mutant of *S. meliloti* showed a delayed appearance of nodules, a reduced total number of nodules, and also a reduced shoot dry mass in comparison to plants inoculated with the wild type strain.

Bacteroids occupying indeterminate nodules, such as *S. meliloti*, *M. ciceri* and *R. leguminosarum* bv. *viciae*, do not accumulate PHB during symbiosis, but possess large numbers of PHB granules during the initial stages of invasion and cell division within the infection thread [23]. These PHB granules disappear during differentiation, and mature were observed: (i) The sizes of the electron-dense bodies within the bacteroids of the recombinant strains Rm1021 and Rm4775 harbouring *cphA1* alone were greater and the bodies exhibited irregular shapes, presumably due to the accumulation of CGP (Figure 4). Similar irregular shapes of CGP granules were observed in the cytoplasm of the bacterial cells of the strains when cultivated in YMB medium [27] and also in the cytoplasm of the cells of *Acinetobacter baylyi* strain ADP1 when cultivated in MSM medium [37]; (ii) Higher levels of rough endoplasmatic reticulum (RER) and Golgi membranes were observed in plant cells containing bacteroids of the recombinant strain Rm1021 (pBBR1MCS-3::*cphB1*::*cphA1*::ΩKm) in comparison to plant cells containing bacteroids of the wild type strain or bacteroids of the other *S. meliloti* strains (Figure 4).

Electron micrographs of sections of alfalfa nodules induced by the PHB- *S. meliloti* strains showed that PHB inclusions were present in the cytoplasm of the bacterial cells in the infection thread and also just after release of the bacterial cells into the host plant cells, but absent from the mature bacteroids.

Compact electron-dense bodies were observed within the cytoplasm of mature bacteroids in all nodules sections of all treatments as well as in the wild type bacteroids (Figure 4). Only two differences
bacteroids are notably devoid of visible PHB granules [39]. The reason for this is not known; however, it has been shown that the activities of β-ketothiolase, acetocacetyl-CoA reductase, PHB depolymerase, and β-hydroxybutyrate dehydrogenases are similar in free-living cells and bacteroids of *M. ciceri* CC1192. Because gas chromatographic analysis also revealed only small amounts of PHB in bacteroids, PHB synthesis is presumably accompanied by an equivalent rate of degradation [29]. Although PHB mutants are capable of nodulation and of establishing effective symbiosis [40,41], it is possible that intracellular PHB may fuel cell division and growth during root infection and invasion [28].

The major exchange of nutrients between the symbiotic partners is the transfer of reduced carbon from the plant to support the nitrogenase activity of the bacteroids and the transfer of fixed nitrogen as ammonium ions from the bacteroids [25] in the opposite direction. The transferred ammonium is assimilated as glutamine in the plant cytoplasm by glutamine synthetase, and is then transferred to form the amino acids, most likely through transamination that generates the amino group, and next, the amino group is transferred to one of the carbon skeletons of the amino acids. ATP, which is required for the reduction of nitrogen to ammonia, is derived from the reduced carbon provided by the plant [24,43]. CGP biosynthesis was also identified as an energy-consuming process [21]. As a result, CGP biosynthesis may compete with nitrogen fixation for ATP produced by the metabolism of C4-dicarboxylates as sole energy source via the TCA cycle.

CGP biosynthesis in bacteroids probably has complex effects on nitrogen fixation due to the consumption of ATP and of amino acids. The consequences are lower plant growth, lower plant yields, and lower plant nitrogen content. The legume-rhizobial symbiosis responds to inefficient nitrogen fixation in complex ways, such as increased nodule mass as well as increased nodule number. An increase in nodule mass similar to what may be observed in inefficient symbioses is observed in the symbiosis induced by the recombinant strain Rm1021 (pBBR1MCS-3::cphA17120), although this symbiosis also exhibited a very high nitrogen fixation activity. However, this higher nitrogenase activity is not accompanied by higher rates of nitrogen assimilation by the plant, and the nitrogen contents of these plants are significantly reduced at 55 days. CGP is a nitrogen-rich polymer, and at least a portion of the fixed nitrogen may become locked up in CGP instead of being available to the plant. Similarly, production of CGP in transgenic tobacco and potato plants decreased the growth rate by the depletion of the amino acid pools of the plant [25]. A reduced ability to produce essential proteins might occur due to depletion of the amino acids, most likely of arginine and/or aspartate, caused by CGP production and storage [17]. Similarly, cell densities of transgenic *Saccharomyces cerevisiae* expressing *cphA* reached lower values than those of the negative controls without *cphA*, indicating that synthesis and accumulation of CGP resulted in slower growth [20].

Generally, simultaneous biosynthesis and degradation of CGP in bacteroids slightly improved the growth of the plants inoculated with the respective strains, although this increase could not be shown to be statistically significant. The slightly enhanced growth of the plants infected with these strains may be explained by additional

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### Table 2: Growth parameters of alfalfa plants 55 days after inoculation with different *S. meliloti* strains

| S.No | Shoot length (cm) | Root length (cm) | Leaves/plant | Nodules/plant | Nodule fresh wt. (mg/plant) | Shoot dry wt. (mg/plant) | %N of shoot dry wt. | Nitrogenase activity |
|------|------------------|-----------------|--------------|---------------|-----------------------------|--------------------------|---------------------|------------------|
| 1    | 55.68 ± 4.96     | 16.53 ± 0.62    | 92.43 ± 10.21 | 30.30 ± 2.06  | 71.26 ± 6.44                | 289.35 ± 8.05            | 8.10 ± 0.21         | 1675 ± 164        |
| 2    | 58.82 ± 4.21     | 16.38 ± 0.68    | 96.70 ± 6.9  | 32.77 ± 1.60  | 94.47 ± 7.76                | 289.57 ± 2.73            | 4.90 ± 0.31         | 3093 ± 10         |
| 3    | 59.33 ± 4.90     | 14.77 ± 0.57    | 93.07 ± 8.43 | 31.40 ± 2.08  | 85.09 ± 1.10                | 317.68 ± 23.69           | 8.70 ± 0.23         | 1144 ± 70         |
| 4    | 47.25 ± 4.02     | 16.55 ± 0.82    | 69.5 ± 6.80  | 28.70 ± 2.46  | 66.45 ± 4.33                | 218.49 ± 24.67           | 4.77 ± 0.26         | 1777 ± 77         |
| 5    | 24.13 ± 3.05     | 17.77 ± 0.85    | 41.7 ± 5.99  | 22.53 ± 1.90  | 39.85 ± 1.59                | 84.78 ± 1.37             | 4.10 ± 0.23         | 300 ± 50          |
| 6    | 51.33 ± 4.82     | 16.93 ± 0.59    | 76.77 ± 9.09 | 28.57 ± 2.26  | 82.99 ± 12.54               | 235.62 ± 17.60           | 4.83 ± 0.20         | 1854 ± 223        |

*Note: Identical (a, b, c) or (A, B, C) superscripts within the same column denote values not significantly different at two significance levels: *P*<0.05 for a and b; *P*<0.01 for A and B. The values of all parameters are mean values ± standard errors as obtained from the analyses of 30 plants inoculated with the respective strains. Plants were grown in a growth chamber under the conditions described in the Materials and Methods section.*
TEM photographs of thin sections of alfalfa nodules at 35 days post-inoculation with various strains of Sinorhizobium meliloti 1021 on the Symbiosis with Alfalfa (Medicago sativa). J Microb Biochem Technol 8: 375-381. doi: 10.4172/1948-5948.1000312

The large expansion of rough endoplasmic reticulum (RER) and Golgi bodies observed in the nodules induced by the wild type strain occurred in all nodules induced by the recombinant strains harbouring only cphA1 and are likely to be CGP granules. Further examination should be done to differentiate between the various electron-dense bodies within the bacteroids.

The compact electron-dense bodies that were observed within the cytoplasm of mature bacteroids of all Sinorhizobium meliloti strains probably consisted of polyphosphate [44]. Larger electron-dense bodies than those seen in the nodules induced by the wild type strain occurred in all nodules induced by the recombinant strains to which an artificial CGP metabolism had been introduced were achieved at significantly reduced levels of nitrogen fixation. This observation may also suggest a more efficient transfer of nitrogen to the plant via CGP degradation products in these symbioses, and should be investigated more closely.

The existence of the latter enzyme, which is responsible for cleavage of the dipeptides into the amino acid monomers, was confirmed by in vitro experiments [27]. At least in case of the PHB cphB1 strain, similar or even enhanced growth parameters compared to the wild type were achieved at significantly reduced levels of nitrogen fixation. This observation may also suggest a more efficient transfer of nitrogen to the plant via CGP degradation products in these symbioses, and should be investigated more closely.

Although the increases in growth observed in plants infected with strains to which an artificial CGP metabolism had been introduced were not statistically significant, the differences were consistently observed, and the system has not yet been optimized in any way. An increase of some growth parameters by about 10% is therefore remarkable. This study will pave the way to further enhance the growth and to increase the yield of legume crops.

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