Comparison of two kinetics models for estimating N mineralization affected by different quality of organic matter in Typic Hapludults

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Abstract: N availability in the soil is determined by the rate of mineralization. N mineralization can be estimated using several models. Aim of research is to compare of two kinetics models for estimating N mineralization affected by different inputs of organic matter and soil fertility in Typic Hapludults. Research used completely randomized design with two factors. The first factor was land use: (1). Soil from land cultivated by cassava crops for less than 10 years, and (2). Soil from land cultivated by cassava crops for more than 30 years. The second factor was type of organic matter: (1). Groundnut biomass, (2). Maize biomass, (3). Groundnut-maize biomass, with ratio of 1:1, (4). Groundnut – maize biomass, with ratio of 2:1, (5). Groundnut – maize biomass, with ratio of 1:2, and (6). Without organic matter. The result shows that based on the value of R², first-order kinetics equation is more suitable to describe N mineralization than double-pool kinetics equation because R-square value higher than double-pool kinetics and is positively correlated with N mineralization parameters (N0, k and N0,k).

Keywords: kinetic equations, nitrogen mineralization, organic matter quality

Introduction

Ultisol has low fertility due to intensive leaching (Van Noordwijk et al., 1996). Ultisol has acid soil reaction, low of cation exchange capacity (CEC) and organic matter (Shaw et al, 2010). The addition of organic matter into soil through harvest remaining, compost, and cover crops prunings can improve total reserve of soil organic matter and soil fertility.

Agricultural practices will reduce total N in soil. Crops need N in large amount compared to other nutrients. However, N availability is low due to its mobility in soil is very high. N availability in soil is determined by the rate of mineralization where it is affected by various factors such as soil pH, moisture, temperature and quality of organic matter (Handayanto et al., 1997; Griffin and Honeycutt, 2000; Samuel et al., 2002; Cookson et al., 2002; Kyveryga et al., 2004; Fritschi et al. 2005; Agehara and Warncke, 2005).

There are several approaches to evaluate the kinetic of N mineralization as an indicator of N availability in soil. Two models widely known for estimating N mineralization are first-order kinetics equation and double exponential model involving two parts describing fast and slow organic pools (Heumann and Bottcher, 2004; Cartes et al., 2009).

Recently, logarithmic, parabolic and hyperbolic models are also developed. However, the exponential model is more often used. Mineralization of organic matter process involves microorganism. Microorganism activity is determined by the dynamic of enzyme which is often described as exponential. Thus the mineralization process then described using exponential model (Li et al., 2003).

Several kinetic models are often used to estimate the kinetic of N mineralization, then a model is selected based on the highest correlation coefficient and the lowest standard error (Shariatmadari et al., 2006). Aim of research was to compare of two kinetics models for estimating N mineralization affected by different inputs of organic matter and soil fertility in Typic Hapludults.
Materials and Methods

Soil preparation and experimental treatment

Sample of soil was taken from a depth of 0-20 cm (topsoil), belong to Typic Hapludults order (ICALRD, 2008). The result of soil analysis is presented in Table 1. Research used completely randomized design with two factors. The first factor was land use: (1). Soil from land cultivated by cassava crops for less than 10 years, and (2). Soil from land cultivated by cassava crops for more than 30 years. The second factor was type of organic matter: (1). Groundnut biomass, (2). Maize biomass, (3). Groundnut-maize biomass, with a ratio of 1:1, (4). Groundnut -maize biomass, with a ratio of 2:1, (5). Groundnut – maize biomass, with a ratio of 1:2, and (6). Without organic matter.

Table 1. Chemical properties of soil samples

| Soil analysis      | Soil from land cultivated by cassava crops for less than 10 years | Soil from land cultivated by cassava crops for more than 30 years |
|-------------------|---------------------------------------------------------------|---------------------------------------------------------------|
| pH (H₂O)          | 5.1                                                           | 4.6                                                           |
| N-total (%)       | 0.073                                                         | 0.037                                                         |
| C–organic (%)     | 2.06                                                          | 0.70                                                          |
| P-Bray 1 (ppm P₂O₅) | 15.9                                                         | 6.8                                                           |
| CEC (cmol (+)/kg) | 6.54                                                         | 4.12                                                          |
| K (cmol (+)/kg)   | 0.14                                                          | 0.05                                                          |
| Ca (cmol (+)/kg)  | 1.68                                                          | 0.50                                                          |
| Mg (cmol (+)/kg)  | 0.33                                                          | 0.15                                                          |
| Al exchange (cmol (+)/kg) | 1.40                                                   | 2.50                                                          |
| Al saturation (%) | 33                                                            | 54                                                            |

Dosage of all selected biomasses applied were equivalent to 5 t/ha. Samples of soil sieved through a 2 mm sieve. 10 g of soil was weighed and kept into a plastic bottle then each treatment of organic matter was given. The moisture content of soil samples were maintained same as in field capacity.

These bottles were stored in incubators with various temperatures of 20°C, 25°C and 30°C. After 0, 2, 4, 8, 6, 10 and 12 weeks, content of N (NH₄⁺ and NO₃⁻) was observed using 1 M KCl extract with Kjeldahl method . The relationship between kinetics parameters of N mineralization and nutrient uptake were obtained by treated cassava in greenhouse same as incubators. Crops were cultivated in pots contained 15 kg of soil in field capacity. Crops were harvested at the age of four months and analyzed for the N uptake and soil analysis involving microbial biomass C and N, water-soluble C and N, Particulate Organic Carbon (POC) and Particulate Organic Nitrogen (PON) (Soon et al., 2007; Okore et al., 2007).

Observation of N mineralization kinetic

Kinetics was observed by two models, i.e. irst-order kinetics equation and double-pool kinetics equation. First-order kinetics equation was estimated by determining N mineralization potential (N₀, in mg/kg), mineralization rate constant (k in day⁻¹), the apparent energy activation (Ea in J mol⁻¹), using a simple reaction-type model developed by Sugihara et al. (1987):

\[ N = N₀ \left[ 1 - \exp \left( -kt \right) \right] \quad (1) \]

N is the amount of inorganic N (mg N kg⁻¹) on day t, N₀ is the N mineralization potential (mg/kg), k is mineralization rate constant (day⁻¹).

Double-pool kinetics equation was estimated by Ando et al. (1992) and Heumann and Bottcher (2004):

\[ N = Na(1 - \exp \left( -kt \right)) + Nr(1 - \exp \left( -kt \right)) \quad (2) \]

N is the amount of inorganic N (mg N/kg) on day t, Na is fast pool mineralization (mg/kg), ka is fast pool mineralization rate constant (day⁻¹), Nr is slow pool mineralization (mg/kg), kr is slow pool mineralization rate constant (day⁻¹).

Statistical analysis

First-order kinetics and double-pool kinetics equations were fitted with the curve fit procedure using SigmaPlot 12 program (SYSTAT software inc, 2011).
Results and Discussion

First-order and double-pool kinetics equations to estimate N mineralization

N mineralization estimation using first-order kinetics equation shows that organic matter from groundnut and maize biomasses affected mineralization rate constant (k). The k value of soil from land cultivated by cassava crops for less than 10 years has same pattern as soil from land cultivated by cassava crops for more than 30 years. Organic matter from mix of groundnut : maize (2: 1) has the highest k value. It is followed by groundnut biomass, groundnut : maize (1: 1), groundnut : maize (1: 2), maize and without organic matter (Table 2).

These results indicates that the mix of organic matter from groundnut biomass with low C:N ratio and maize biomass with high C:N ratio can increase mineralization rate compared to only maize biomass. It is due to C:N ratio decrease. Sholihah et al. (2012) reported that mineralization rate has correlation to C:N ratio (0.582*). Organic matter with low C:N ratio has higher mineralization rate compared to organic matter with high C:N ratio (Abera et al., 2012). A similar pattern is found for N potential observation.

Mineralization rate constant (k) has positive correlation with temperature. The k value increases around 32% by temperature rise from 20°C to 30°C for the less than 10 years cassava crops cultivation and about 27% for the more than 30 years cassava crops cultivation. Guntinas et al. (2012) reported that k value is strongly influenced by temperature and sharply increase for temperature between 25°C and 35°C. It is due to the increase of organic matter decomposition rate (Nordmeyer and Richter, 1985) resulted by changes in biochemical composition of organic matter and transport processes e.g. diffusion process as well as increasing the number of microorganisms (MacDonald et al., 1995). The highest k value for the mix organic matter of groundnut : maize (2:1) from the less than 10 years cassava crops cultivation is 0.098 and from the more than 30 years cassava crops cultivation is 0.0083. The low k value can be assumed come from the low activity of soil microorganisms (Nira and Nishimune, 1993) and it shows poor N mineralization (Nishio et al., 1994).

N mineralization measured by double-pool kinetics equation estimates mineralization rate of fast and slow pools. This study indicates that mineralization rate constant (ka and kr) and N mineralization (Na and Nr) are affected by temperature and organic matter. Increasing of temperature will be followed by the increase of k and N value. The ka value of the less than 10 years cassava crops cultivation is 0.028 per day at temperature of 20°C and 0.031 per day at temperature of 30°C, while kr value respectively are 0.026 and 0.029 per day. The ka value of the more than 30 years cassava crops cultivation is 0.023 per day at temperature of 20°C, 0.025 and 0.029 per day at temperature of 25°C and 30°C, while kr value respectively are 0.021, 0.023 and 0.026 per day at temperature of 20°C, 25°C and 30°C (Table 2). Temperature is one of factors affecting N mineralization rate (Sierra, 2002). Study of Stanford and Smith (1972) showed that range of k values is 0.035 up to 0.095 per week, with the average of 0.054 per week. In addition, Sierra (2002) finds that the range of k value in tropical soil is 0.007 up to 0.014 per day.

In this study, N mineralization is also determined by type of organic matter. N mineralization of groundnut biomass is greater than maize biomass. The mix of organic matter from groundnut and maize biomass increase N mineralization compared to only maize biomass. Mineralization of the fast pool (Na) was higher than slow pool (Nr). The range of Na value of the less than 10 years cassava crops cultivation is 943 - 1253 mg/kg, while the range of Nr value is 560 -1016 mg/kg. At temperature of 30°C, application of groundnut : maize biomass (1:1) has the highest N mineralization (3.570 mg/kg), 284% higher than no biomass application. The range of Na value of the more than g 30 years cassava crops cultivation-soil sample is 579-1011 mg/kg, while the range of Nr value is 549-792 mg/kg. Application of groundnut biomass has the highest N mineralization (2318 mg.kg⁻¹), 147% higher than no biomass application. Plant biomass affects N mineralization. Groundnut biomass or the mix of groundnut and maize biomasses increase N mineralization. Groundnut is a legume crop having a low ratio of C: N that is easy to be mineralized.

Based on double-pool kinetics equation, N mineralization of fast pool is higher than the slow pool, same as mineralization rate constant (k) of fast pool that was greater than slow pool. This indicates that mineralization of easily decomposed-organic matter can happen quickly compared to some resistant organic which can be mineralized linearly with time. The results are consistent with study of Li et al. (2003) which the value of Na and ka is greater than the value of Nr and kr. Result of this study shows that the average of R-square value in first-order kinetics equation is higher than double-pool kinetics equation. It implies N mineralization estimation using first-order kinetics equation is better than double-pool kinetics equation (Table 2).
Comparison of two kinetics models for estimating N mineralization

Table 2. Value of k and potential N mineralization on first-order kinetics equation and double pool kinetics equation in Typic Hapludult, Lampung

| Treatments                                      | First-order kinetics equation | Double-pool kinetics equation |
|------------------------------------------------|-------------------------------|-------------------------------|
|                                                 | k   | N0   | R²  | Na   | ka   | Nr   | kr   | R²  |
| The less than 10 years cassava crops cultivation-soil sample |     |      |     |      |      |      |      |     |
| Groundnut (G)                                   | 0.0090 | 743.0 | 0.996 | 1719 | 0.027 | 1078 | 0.023 | 0.980 |
| Maize (M)                                       | 0.0074 | 989.8 | 0.989 | 1222 | 0.030 | 959  | 0.028 | 0.955 |
| G : M (1:1)                                     | 0.0090 | 1000.9 | 0.995 | 1969 | 0.027 | 1601 | 0.025 | 0.982 |
| G : M (2:1)                                     | 0.0098 | 1150.5 | 0.994 | 1569 | 0.034 | 1301 | 0.032 | 0.956 |
| G : M (1:2)                                     | 0.0078 | 1056.2 | 0.989 | 519  | 0.026 | 737  | 0.025 | 0.897 |
| Without OM                                      | 0.0063 | 641.1 | 0.966 | 519  | 0.027 | 417  | 0.025 | 0.964 |
| Means                                           | 0.0082 | 930.3 | 0.988 | 1253 | 0.029 | 1016 | 0.026 | 0.957 |
| The more than 30 years cassava crops cultivation-soil sample |     |      |     |      |      |      |      |     |
| Groundnut (G)                                   | 0.0080 | 587.5 | 0.996 | 1374 | 0.030 | 944  | 0.027 | 0.980 |
| Maize (M)                                       | 0.0076 | 650.7 | 0.997 | 1192 | 0.031 | 796  | 0.027 | 0.982 |
| G : M (1:1)                                     | 0.0083 | 1057.8 | 0.998 | 1251 | 0.033 | 1037 | 0.031 | 0.987 |
| G : M (2:1)                                     | 0.0072 | 994.1 | 0.982 | 996  | 0.035 | 898  | 0.033 | 0.939 |
| Without OM                                      | 0.0058 | 544.7 | 0.981 | 485  | 0.027 | 451  | 0.027 | 0.925 |
| Means                                           | 0.0073 | 776.8 | 0.991 | 1011 | 0.031 | 792  | 0.029 | 0.965 |

Relationship between kinetics parameters of N mineralization with N uptake

The parameters of N mineralization kinetics are positively correlated with plant dry weight and N uptake (Table 3). It implies that N mineralization kinetics can be used to estimate N uptake by plants. Organic matter addition into soil will increase microorganisms activity in N and C mineralization processes. The increase of N0, k and N0.k will increase N mineralization kinetics. Moreover, it can increase the availability of N in the soil resulting N uptake by plants increase (Geisseler et al., 2009). De-Zhi et al. (2006) reported that N mineralization has positive correlation with N uptake in paddy in which without N source-fertilizer, approximately 98% of N absorbed by paddy is taken from N mineralization. In double-pool kinetics equation, N mineralization kinetics parameter.e.g, fast (Na) and slow (Nr) pools mineralization has positive correlation with plant dry weight and N uptake (Table 3). In this study, fast and slow pools N mineralization is contributed to cassava plant growth.

Table 3. Correlation between N kinetics parameters in firstorder and double-pool kinetics equations with plant dry weight and N uptake

| Observations                                      | First-order kinetics equation | Double-pool kinetics equation |
|---------------------------------------------------|-------------------------------|-------------------------------|
|                                                   | N0   | k     | N0.k | Na   | ka   | Nr   | kr   |
| The less than 10 years cassava crops cultivation-soil sample |      |       |      |      |      |      |      |
| Plant dry weight                                  | 0.510* | 0.534* | 0.521* | 0.785** | 0.579* | 0.756** | 0.501 |
| N uptake                                          | 0.549* | 0.516* | 0.516* | 0.693** | 0.454  | 0.656** | 0.537* |
| The more than 30 years cassava crops cultivation-soil sample |      |       |      |      |      |      |      |
| Plant dry weight                                  | 0.560* | 0.674** | 0.543* | 0.648** | 0.171  | 0.564* | 0.675** |
| N uptake                                          | 0.552* | 0.832** | 0.616** | 0.646** | 0.205  | 0.457  | 0.796** |

Based on the result of correlation between kinetics parameters of N mineralization with plant dry weight and N uptake, first-order kinetics equation has better correlation than double-pool kinetics equation. The first –order kinetics equation has the value of N0, k and N0.k which is positively correlated with plant dry weight and N uptake, while the double pool kinetics equation has the value of Na, Nr and Kr which is correlated with plant dry weight and N uptake (Table 3).
Comparison between first order and double-pool kinetics equations

R-square value is generally used for estimating equation. R-square value closer to 1.00 indicates good equation. Table 2 shows that the average value of R square at various temperatures in the first-order kinetics equation is higher than the double-pool kinetics equation.

First order kinetics equation for explaining the dynamics of soil N mineralization was first introduced by Stanford and Smith (1972), which was developed through laboratory procedures with experimental incubation for 30 weeks under aerobic condition with controlled humidity and temperature. In first-order kinetics equation, N mineralization potential is defined as an easily-mineralized pool N in soil (Gill et al., 2011). In recent years, incubation procedure to estimate N mineralization in a controlled laboratory developed by Stanford and Smith (1972) has been followed by many researchers (Li et al., 2003; Cordovil et al., 2007; Dessureault-Rompre et al., 2010; Guntinas et al., 2012).

In 1980, another equation introduced by Molina et al. (1980) and Richter et al. (1980) is a model with two components or double exponential equation. This equation is developed based on two components of organic N fraction, namely active and resistant fractions. Active fraction is rapidly undergoing a process of mineralization (fast pool), while the resistant fraction is relatively difficult in mineralization (slow pool). This equation assumes that N mineralization process always involves microbial and enzyme activities which is growing exponentially so that the estimation of N mineralization should be based on the exponential equation (Li et al., 2003).

Research of Ando et al. (1992) suggested that first order equation can be used to estimate N mineralization in soil with anaerobic (wet condition), otherwise in aerobic condition, double exponential equation is more suitable to estimate N mineralization. Dou et al. (1996) suggested that N mineralization depends on incubation time. For 30 weeks-incubation, double exponential equation is a good model to explain N mineralization, while for 15 weeks-incubation either first-order kinetics or double exponential equation can be used to estimate N mineralization.

Benbi and Richter (2002) reported that for incubation during 82 days, double exponential equation could not be used to estimate N mineralization, because organic N pool consists of various components which can be characterized by its half-life. If the incubation time is too short compared to the half-life of organic N fraction, N mineralization may look as linear. Related to incubation time, Gill et al. (2011) suggested that double-pool kinetics equation is better to estimate N mineralization if the incubation period is between one to two years, and even for long-term experiment which need more than two years-incubation time. Benbi and Richter (2002) suggested that for much longer incubation time until mineralization rate is approaching a constant value.

Quality of organic matter also affects equation model. Organic matter with wide C:N ratio is more suitable using double-pool kinetics equation for describing N mineralization. In organic matter with wide C:N there are two organic N pools, i.e. fast pool consisting organic matter with low C: N ratio and slow pool consisting high C: N ratio (Benbi and Richter, 2002). Azeez and Van Averbeke (2010) found that N release rapidly on days 0-30, constant on days 45-55 then decrease on days 70-90 when analyzing N mineralization from three manures respectively chicken (C:N ratio = 9.6), cow (C:N ratio = 10.3) and goat (C:N ratio =11.3).

Some researchers have suggested that soil samples preparation also affects N mineralization. In undisturbed soil, first-order kinetics equation is suitable to describe N mineralization, while double pool kinetics equation is suitable to describe disturbed soil. The high release of N in disturbed soil due to micro aggregate destruction during soil samples handling (Benbi and Richter, 2002).

Based on explanation above, there are several factors that cause N mineralization can be estimated with an equation, namely: (1) incubation time, (2) type of organic matter or organic matter quality, (3) soil condition (dry, wet, disturbed and not disturbed). In this study, the incubation time was only 12 weeks and categorized as short-term. This caused the first-order kinetics equation to have R square value higher than double-pool kinetics equation.

**Conclusion**

In this study, N mineralization is depend on soil fertility, temperature and organic matter quality. N mineralization is higher at fertile soil, high temperature (30°C) and low of C:N ratio. First-order kinetics equation is more suitable to describe N mineralization than double-pool kinetics equation. The first-order kinetics equation has higher R-square value (correlation coefficient) than double-pool kinetics equation and positively correlated with N mineralization parameters (N0, k and N0.k). Several factors that
cause N mineralization can be estimated with an equation, namely: (1) incubation time, (2) type of organic matter or organic matter quality, and (3) soil condition (dry, wet, disturbed and not disturbed).

References

Abera, G., Wolde-mesqet, E. and Bakken, I.R. 2012. Carbon and nitrogen mineralization dynamics in different soils of the tropics amended with legume residues and contrasting soil moisture contents. *Biogeochemistry of Soils* 48: 51–66.

Agehara, S. and Warncke, D.D. 2005. Soil moisture and temperature effect on nitrogen release from organic nitrogen sources. *Soil Science Society of America Journal* 69: 1844–1855.

Ando, H., Aragones, R.C. and Wada, G. 1992. Mineralization pattern of soil organic N of several soils in the tropics. *Soil Science and Plant Nutrition* 38:2: 227–234.

Azeez, J.O. and Van Averbeke, W. 2010. Nitrogen mineralization potential of three animal manures applied on a sandy clay loam soil. *Bioresource Technology* 101: 5645–5651.

Benbi, D.K. and Richter, J. 2000. A critical review of some approaches to modelling nitrogen mineralization. *Biogeochemistry of Soils* 35: 168–183.

Cartes, P., Jara, A., Demanet, R. and de la Luz Mora, M. 2009. Urease activity and nitrogen mineralization kinetics as affected by temperature and urea input rate in southern Chilean Andisol. *Journal of Soil Science and Plant Nutrition* 9 (1): 69–82.

Cookson, W.R., Cornforth, I.S. and J.S. Rowarth, J.S. 2000. Winter soil temperature (2–15 °C) effect on nitrogen transformations in clover green manure amendend and unamendend soils: a laboratory and field study. *Soil Biology and Biochemistry* 34: 1401–1415.

Cordovil, C.M.D.S., Cabral, F. and Coutinho, J. 2007. Potential mineralization of nitrogen from organic wastes to ryegrass and wheat crops. *Bioresource Technology* 98: 3265–3268.

Dessureault-Rompré, J., Burton, D.L., Sharifi, M., Cooper, J., Grant, C.A. and Drury, C.F. 2010. Relationships among mineralizable soil nitrogen, soil properties, and climatic indices. *Soil Science Society of America Journal* 74: 1218-1227.

De-Zhi, Y., Wang, D., Sun, R. and Lin, J 2006. N Mineralization as affected by long-term N and its relationship with crop N uptake. *Pedosphere* 16 (1): 125-130.

Dou, Z., Toth, J.D., Jabro, J.D., Fox, R.H. and Fritton, D.D. 1996. Soil nitrogen mineralization during laboratory incubation: dynamics and model fitting. *Soil Biology and Biochemistry* 28: 625–632.

Fritsch, F.B., Roberts, B.A., Rains, D.W., Travis, R.L. and Hutmacher, R.B. 2005. Recovery of residual fertilizer-N and cotton residues-N by acacia and pima cotton. *Soil Science Society of America Journal* 69: 718-728.

Geisseler, D., Horwath, W.R. and Doane, T.A. 2009. Significance of organic nitrogen uptake from plant residues by soil microorganisms as affected by carbon and nitrogen availability. *Soil Biology and Biochemistry* 41: 1281–1288.

Gill, M.V., Carballo, M.T. and Calvo, L.F. 2011. Modelling N mineralization from bovine manure and sewage sludge composts. *Bioresource Technology* 102: 863–871.

Griffin, T.S. and Honeycutt, C.W. 2000. Using growing degree days to predict nitrogen availability from livestock manures. *Soil Science Society of America Journal* 64 : 1876-1882.

Guntinas, M.E., Leirós, M.C., Trasar-Cepeda, C. and Gil-Sotres, F. 2012. Effects of moisture and temperature on net soil nitrogen mineralization: A laboratory study. *European Journal of Soil Biology* 48 : 73-80.

Handayanto, E., Giller, K.E. and Cadisch, G. 1997. Regulating N release from plant residues by mixing residues of different quality. *Soil Biology and Biochemistry* 29 : 1417-1426.

Heumann, S. and Bottcher, J. 2004. Temperature function of the rate coefficients of net N mineralization in sandy arable soils Part I. Derivation from laboratory incubation. *Journal of Plant Nutrition and Soil Science* 167 : 381-389.

ICALRD (Indonesian Center for Agricultural Soil Resources Research and Development). 2008. Identification and soil suitability mapping for soybean at Sumatera. p. 146 (In Indonesian).

Kvyerыва, P.M., Blackmer, A.M., Ellsworth, J.W. and Isla, R. 2004. Soil pH effect on nitrification of fall applied anhydrous ammonia. *Soil Science Society of America Journal* 68 : 545-551.

Li, H., Han, Y. and Cai, Z. 2003. Nitrogen mineralization in paddy soils of the Taihu Region of China under anaerobic conditions: dynamics and model fitting. *Geoderma* 88 (5): 161-175.

MacDonald, N.W., Zak, D.R. and Pregitzer, K.S. 1995. Temperature effects on kinetics of microbial respiration and net nitrogen and sulfur mineralization. *Soil Science Society of America Journal* 59: 233-240.

Molina, J.A.E., Clapp, C.E. and Larson, W.E. 1980. Potentially mineralizable nitrogen in soil: the simple exponential model does not apply to the first 12 weeks of incubation. *Soil Science Society of America Journal* 44 :442-443.

Nira, R. and Nishimune. A. 1993. Studies on nitrogen mineralization properties of Tokachi soils by kinetic analysis. *Soil Science and Plant Nutrition* 39 (2) : 321-329.

Nishio, T., Sekiya, H., Toriyama, K. and Kogano, K. 1994. Changes in gross rates of nitrogen transformations in soil caused by conversion of paddy fields to upland fields. *Soil Science and Plant Nutrition* 40 (2) : 301-309.

Nordmeyer, H. and Richter, J. 1985. Incubation experiments on nitrogen mineralization in loess and sandy soils. *Plant and Soil* 83 : 433–445.

Okore, I. K., Tijani-Eniola, H., Agboola, A.A. and Aiyelari, E.A. 2007. Impact of soil clearing methods and cropping systems on labile soil C and...
Comparison of two kinetics models for estimating N mineralization

N pools in the humid zone Forest of Nigeria. 
*Agriculture, Ecosystems and Environment* 120 : 250–258.
Richter, J., Nuske, A., Boehmer, U. and Wehrmann, J. 1980. Simulation of nitrogen mineralization and transport in Loess-Parabrownearthes: plot experiments. *Plant and Soil* 54, 329–337.
Samuel, A., Sylvie, Manuel, R.G.V. and Robert, O. 2002. Impact of residue quality and location in soil on the C and N mineralisation of residues from cropping systems from Cerrados, Brasil. *World Congress Soil Science*, 14-21 August 2002. Thailand. Paper No.800. p. 800-1 – 800-10.
Shariatmadari, H., Shirvani, M. and Jafari, A. 2006. Phosphorous release kinetics and availability in calcareous soils of selected arid and semiarid toposquences. *Geoderma* 132 : 261-272.
Shaw, J.N., Hajek, B.F. and Beck, J.M. 2010. Highly weathered mineralogy of select soils from Southeastern U.S. Coastal Plain and Piedmont landscapes. *Geoderma* 154 : 447-456.
Sholihah, A., Prijono, S., Utami, S.R. and Handayanto, E. 2012. N mineralization from residues of crops grown with varying supply of 15N concentrations. *Journal of Agricultural Science* 4 (8) : 117-123.
Sierra, J. 2002. Nitrogen mineralization and nitrification in a tropical soil : effects of fluctuating temperature conditions. *Soil Biology and Biochemistry* 34 : 1219–1226.
Soon, Y.K., Arshad, M.A. Haq, A. and Lupwayi, N. 2007. The influence of 12 years of tillage and crop rotation on total and labile organic carbon in a sandy loam soil. *Soil and Tillage Research* 95 : 34-46.
Stanford, G. and Smith, S.J. 1972. Nitrogen mineralization potentials of soils. *Soil Science Society of America Proceeding* 36 : 465-472.
Sugihara, S., Konno, T. and Ishii, K. 1987. Kinetics of nitrogen mineralization of organic nitrogen in soil. Bull. Natl. Inst. Agro-Environ. Sci 1: 127-166 (in Japanese with English summary).
Van Noordwijk, M., Hairiah, K., Guritno, B., Sugito, Y. and Ismunandar, S. 1996. Biological management of soil fertility for sustainable agriculture on acid upland soils in Lampung (Sumatra). *Agrivita* 19:131-136.
Comparison of two kinetics models for estimating N mineralization

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