Microsatellite instability and Epstein-Barr virus combined with PD-L1 could serve as a potential strategy for predicting the prognosis and efficacy of postoperative chemotherapy in gastric cancer

Na Yang¹*, Yanhua Wu¹*, Meishan Jin², Zhifang Jia⁴, Yueqi Wang¹, Donghui Cao¹, Lili Qin¹, Xueying Wang¹,³, Min Zheng¹, Xueyuan Cao⁴ and Jing Jiang¹

¹ Division of Clinical Research, First Hospital of Jilin University, Changchun, Jilin Province, China
² Division of Pathology, First Hospital of Jilin University, Changchun, Jilin Province, China
³ Department of Epidemiology and Biostatistics, School of Public Health, Jilin University, Changchun, Jilin Province, China
⁴ Department of Gastric and Colorectal Surgery, First Hospital of Jilin University, Changchun, Jilin Province, China
* These authors contributed equally to this work.

ABSTRACT

Background. Microsatellite instability (MSI) and Epstein-Barr virus (EBV)-positive molecular subtypes exhibit complex immune responses in gastric cancer (GC), and PD-L1 has emerged as a prognostic biomarker associated with the cancer immune microenvironment. This study aimed to determine the prognostic value of molecular subtypes and whether the addition of PD-L1 would accurately predict the prognosis and guide postoperative chemotherapy for GC patients.

Methods. We performed molecular subtyping of tissue microarray slides from 226 GC patients who were treated with radical gastrectomy. The MSI status and PD-L1 expression were evaluated through immunohistochemistry (IHC) and EBV status through situ hybridization. Multiplex polymerase chain reaction (PCR) was also performed on 50 cases to validate the accuracy of IHC in defining MSI status. Differences in overall survival (OS) were assessed using the Kaplan-Meier method, log-rank test and Cox proportional hazards regression model.

Results. Among the 226 GC patients, 52 (23.2%) patients were classified as the MSI subtype, 11 (4.9%) were EBV+ subtype, and 161 (71.9%) were MSS (Microsatellite stable) /EBV− subtype according to TCGA analysis. Two patients were both positive for MSI and EBV infection. EBV+ cases showed higher PD-L1 positivity than MSI cases and MSS/EBV− cases (81.8% vs. 50.0% vs. 35.4%, P = 0.003). Compared with the non-MSS/EBV− (MSS or EBV+ cases) subgroup, GC patients with MSS/EBV− were associated with the worst outcomes (HR = 1.610, 95% CI [1.046–2.479], P = 0.031). MSS/EBV− GCs alone could benefit from postoperative chemotherapy (HR = 0.452, 95% CI [0.299–0.682], P < 0.001), and PD-L1-positive expression could also predict a better prognosis (HR = 0.612, 95% CI [0.389–0.962], P = 0.033) in this subgroup. Considering both chemotherapy efficacy and PD-L1 expression in the MSS/EBV− subgroup, chemotherapy could improve the prognosis for PD-L1-negative
MSS/EBV− GCs (HR = 0.357, 95% CI [0.217–0.587], \( P < 0.001 \)) but not PD-L1-positive MSS/EBV− GCs.

**Conclusions.** Molecular subtyping combined with PD-L1 expression could serve as a potential strategy to better predict prognosis and guide postoperative chemotherapy of GC patients.

**Subjects** Molecular Biology, Gastroenterology and Hepatology, Oncology, Histology, Medical Genetics  
**Keywords** Microsatellite instability, Epstein-Barr virus, PD-L1, Gastric cancer, Postoperative chemotherapy, Overall survival

**INTRODUCTION**

Gastric cancer (GC) is still an important global health problem ([Lyons et al., 2019](#)) and ranks fifth in terms of incidence but third in terms of mortality, with over 1,000,000 new cases and an estimated 769,000 deaths worldwide in 2020 ([Sung et al., 2021](#)). Notably, more than 40% of the worldwide GC cases and deaths occur in China. Based on the latest data from the National Central Cancer Registry of China (NCCRC) in 2015, GC was the second most prevalent cancer and third leading cause of cancer-related deaths in China ([Wang et al., 2019](#)). At present, GC patients are still mainly treated with complete tumor resection, and postoperative adjuvant radiotherapy and chemotherapy or neoadjuvant chemotherapy can improve the survival time of patients with advanced GC ([Tan, 2019](#)). Surgical resection combined with chemotherapy remains the first-line treatment for advanced GC in China. Even though the GC survival rate has increased due to the improved systemic management over recent decades, the prognosis of GC patients remains poor in China, with a disappointing low 5-year relative survival rate of 35.9% ([Yang et al., 2018](#)). More importantly, a variable response to treatment has been observed based on the one-size-fits-all approach to treatment, showing that GC is a heterogeneous disease ([Digklia & Wagner, 2016](#)). Thus, there is an urgent need for novel biomarkers to precisely predict the prognosis of GCs and to achieve personalized treatment.

The Cancer Genome Atlas (TCGA) has recently proposed four comprehensive molecular subtypes for GC based on gene expression profile studies ([Cancer Genome Atlas Research Network, 2014](#)), which provide new prognostic biomarkers and potential therapeutic implications, particularly the Epstein-Barr virus (EBV)-positive subtype and microsatellite instability (MSI) subtype ([Kawazoe et al., 2017](#)). Recent studies have suggested that patients with different molecular subtypes of GC may have different prognoses, and some subtypes, such as MSI, may affect the efficacy of chemotherapy for GC ([Smyth et al., 2017](#)). However, in this regard, molecular typing alone may not be sufficient, and more markers should be considered for prognostic evaluation and choice of treatment. Previous studies indicate that the EBV and MSI subtypes may show complex reactions in the immune system, which exhibit high levels of tumor infiltrating lymphocytes ([Chang et al., 2018](#)). It indicates that the immune microenvironment of the tumor and tumor infiltration may be different in GC patients with different molecular types. PD-L1 has emerged as the most promising...
biomarker, which is associated with the cancer immune microenvironment (Jin et al., 2017). PD-L1 expression in tumor or tumor infiltrating immune cells are used as an adjunct diagnostic criteria for the application of PD-1/PD-L1 pathway inhibitors (Muro et al., 2016), but the prognostic value of PD-L1 is controversial in GC patients (Liu et al., 2020). In recent years, higher PD-L1 expression has been found to be closely associated with EBV-positive and MSI GCs (Lee et al., 2017; Derks et al., 2016). However, it is unclear whether the value of PD-L1 expression in predicting GC prognosis is contingent upon EBV and MSI statuses (Choi et al., 2020). Overall, given the complex immune interactions that occur within the tumor microenvironment and available evidence, the addition of PD-L1 expression based on stratification by EBV and MSI statuses would provide better prognostic insight and elucidate the therapeutic implications in GC, whereas the available reports regarding this pattern are insufficient.

In this study, we focused on the molecular subtypes stratified by EBV and MSI statuses according to the TCGA study and analyzed their prognostic value. Furthermore, we examined the level of PD-L1 expression and explored its prognostic value, as well as its role in response to postoperative chemotherapy among different molecular subtypes for GC patients.

MATERIALS & METHODS

Study population
A total of 226 cases were obtained from the Department of Gastric and Colorectal Surgery, First Hospital of Jilin University (Changchun, China) from February 2011 to August 2016. All included patients were diagnosed with GC pathologically, each of whom was treated with radical gastrectomy and did not receive any chemotherapy or radiotherapy prior to surgery. Patients with distant metastasis or a positive surgical margin were excluded. We retrieved the information of clinical characteristics from the electronic medical record system, which includes age, gender, tumor size, WHO classification, postoperative chemotherapy, T stage, N stage, histological grade, vascular invasion, and neural invasion. The AJCC Cancer Staging Manual (8th edition) was applied to determine the TNM stage.

Follow-up data collection
Prospective follow-up of GC patients was conducted regularly at three months and six months after gastrectomy, as well as every year afterwards until death or being lost to follow-up. The prognosis and chemotherapy of patients were collected during the follow-up. Overall survival (OS) was calculated from surgery to death (from any cause) or the final date of follow-up (if the patients were alive) or the last date of follow-up (if the patients were lost to follow-up). Cases would not be included in the survival analysis if they were lost to follow-up at the first time of telephone interview or if they died of perioperative complications. Postoperative chemotherapy was defined as at least three cycles of chemotherapy after surgery, which mainly included the following three regimens: XELOX (oxaliplatin and capecitabine), FOLFOX-4 (5-fluorouracil, leucovorin and oxaliplatin) and other (such as oxaliplatin, capecitabine or other combinations).
Immunohistochemistry

Immunohistochemistry (IHC) was carried out formalin-fixed, paraffin-embedded 226 GC tissues to detect the expression of four MMR proteins and PD-L1 with antibodies to MLH1 (ab92312, dilution: 1/200, Abcam, Cambridge, UK), MSH2 (ab92473, dilution: 1/200, Abcam, Cambridge, UK), MSH6 (ab92471, dilution: 1/200, Abcam, Cambridge, UK), PMS2 (ab110638, dilution: 1/200, Abcam, Cambridge, UK), and PD-L1 (E1L3N, dilution: 1/200, Cell Signaling Technology, Cambridge, UK). Among them, tissue microarray slides (MiniCore, Alphelys, France) were used for MMR proteins detection and whole sections were used for PD-L1 detection. First, the slides were baked in a 65 °C oven for 20 min, followed by dewaxing. Antigen retrieval was performed in sodium citrate aqueous solution in 100 °C for one minute and forty seconds (20 min and ethylenediamine tetraacetic acid for PD-L1 without high pressure). After this, the slides were washed 3 times with PBS for 5 min each time. The slides were incubated with endogenous peroxidase blocker for 10 min at room temperature, then washed 3 times with PBS for 5 min each time. Then, the slides were incubated with primary antibody for 60 min (90 min for PD-L1) at room temperature, followed by secondary antibody for 15 min at room temperature and washed 3 times with PBS for 5 min each time. 3, 3′-diaminobenzidine tetrahydrochloride (DAB) H₂O₂ was added for 2 min. Counterstaining was done with hematoxylin.

MSI analysis

Positive expression of each MMR protein in IHC was defined as the presence of nuclear staining of tumor cells, regardless of the proportion or intensity (Smyth et al., 2017). Stromal cells of tumor tissue served as the internal negative control. Two pathologists judged the immunohistochemical staining for the expression of four MMR proteins in a blinded manner. MSS was determined with expressions of all MMR proteins, and MSI was determined with at least one MMR protein showing a complete loss of nuclear reactivity (Smyth et al., 2017). For cases with any MMR protein loss, the experiments were repeated on whole sections, and the results were reanalyzed by two pathologists for final confirmation.

To validate the accuracy of the IHC method, we performed multiplex polymerase chain reaction (PCR) on 50 cases randomly selected from the MSI (8 cases) and MSS (42 cases) groups. Genomic DNA was extracted from tumor tissues and matched to adjacent tissues using the tissue DNA isolation mini kit (Vazyme Biotech Co., Ltd., Beijing, China) according to the manufacturer’s instructions. Multiplex PCR was performed with six mononucleotide repeat markers (BAT-25, BAT-26, NR-21, NR-24, NR-27, and MONO-27) using a multiplex PCR-capillary electrophoresis MSI detection kit (Sinomdgene Co. Ltd., Beijing, China). Amplified PCR products were separated, and their sizes were evaluated by capillary electrophoresis on an Applied Biosystems® 3130 or 3500 Dx genetic analyzer (Los Angeles, CA, USA). GeneMapper® software (Applied Biosystems) was used to analyze allelic sizes. By comparing the allelic position of the microsatellite markers in tumor and matched normal tissues, cases with instability at two or more microsatellite markers were classified as MSI-high (MSI-H), whereas cases with instability at 1 microsatellite marker and those without instability were classified as MSI-low (MSI-L) and microsatellite stable.
(MSS), respectively (Goel et al., 2010). Both MSI-L and MSS tumors were combined as MSS in our analysis, according to previous studies (Chao et al., 2019).

**EBV in situ hybridization**

The evaluation of EBV infections were confirmed via DNA in situ hybridization (ISH-5021, ZSGB-BIO, China) using tissue microarrays (MiniCore, Alphelys, France). Tumors with brown nuclear staining were considered as EBV-positive (Pereira et al., 2018).

**PD-L1 expression analysis**

Cell membrane and cytoplasm staining were performed to detect PD-L1 expression in tumor cells (TCs). The IRS system was used to assess the staining intensity and percentages of TCs (Wu et al., 2017). Staining intensity and expression prevalence were graded using a 4-point scale as follows: 0 (no immunostaining; <5% expression), 1 (weak staining; 5 to 19% expression), 2 (moderate staining; 20 to 49% expression), or 3 (strong staining; ≥ 50% expression), and the percentages of PD-L1-positive TCs were recorded as the following 4 categories: 0 (<5% expression), 1 (5 to 19% expression), 2 (20 to 49% expression) and 3 (≥ 50% expression). In the IRS system, specimens were scored on the basis of the intensity and percentage scores, ranging from 0 to 6. A total score of more than 2 indicated a PD-L1-positive status.

**Statistical analysis**

The data were analyzed by SPSS version 21.0 for Windows (SPSS Inc., Chicago, IL, USA). The difference of PD-L1 positivity between the three molecular subgroups was tested using Pearson $\chi^2$ or Fisher’s exact test when appropriate. Differences in OS by molecular features and other characteristics were assessed using the Kaplan–Meier method, log-rank test and Cox proportional hazards regression model. Hazard ratios (HRs) and their 95% confidence intervals (CIs) for OS were calculated. All tests were two-sided, and $P$ values < 0.05 were considered to be statistically significant.

**Ethics approval**

This research was carried out in accordance with the Declaration of Helsinki and was approved by the Human Ethics Review Committee of the First Hospital of Jilin University (Project identification code: 2017-090).

**Consent to participate**

All participants signed the written informed consents before sample and information collection.

**RESULTS**

**Patients’ characteristics**

Median follow-up was 39.7 months (range 1.0–86.7). The baseline characteristics of the 226 GC patients finally enrolled are shown in Table 1. Postoperative chemotherapy was administered to 101 (44.7%) patients. Of note, the majority of patients were in stage III (70.4%) in the entire cohort. The expressions of representative molecular features in GC
tissue are shown in Figure 1. MSI status was determined in all 226 cases by IHC. Overall, 172 (76.1%) patients exhibited MSS and 54 (23.9%) patients exhibited MSI (Table 1). The positive rates of MLH1, MSH2, MSH6, and PMS2 protein expression were 85.8%, 96.0%, 86.7%, and 85.8%, respectively (Figs. 1A–1H). It has become a consensus that immunohistochemistry can be a useful and reliable method in the detection of MSI status in GC compared with microsatellite marker analysis, which could achieve high concordance (91.1%) (Bae et al., 2015). In the current study, of the total 50 cases, 6 cases showed a discrepancy between IHC and PCR results, including 2 cases of MSI-H and 4 cases of MSS/MSI-L, determined on the basis of PCR-based analysis, showing a similar high concordance (88%, Table S1), which is reliable enough for the determination of the MSI status of GCs. Among all patients assessed by ISH, 13 (5.8%) patients were positive for EBV infection (Table 1 and Fig. 1I), and 93 patients (41.2%) were positive for PD-L1 expression in TCs (Table 1 and Fig. 1K). Examples of PD-L1 graded staining in Fig. S1 and the MSI-H and MSS cases based on PCR analysis are shown in Fig. S2.

**Molecular subgroups classified by MSI and EBV statuses**

In the present study, we defined EBV-positive and MSI subgroups separately according to TCGA classification (Cancer Genome Atlas Research Network, 2014). Patients with MSI status were assigned to the MSI subgroup (n = 52, 23.2%), patients with EBV infection were assigned to the EBV+ subgroup (n = 11, 4.9%), and patients without MSI and EBV infection were assigned to the MSS/EBV− subgroup (n = 161, 71.9%). It is worth noting that 2 patients were both positive for MSI and EBV infection, which were not classified in the subsequent subgroup analysis.

**MSS/EBV− subgroup was associated with poor prognosis in GC**

Log-rank test investigated that MSS/EBV− GCs showed the worst OS compared to MSI and EBV+ GCs (P = 0.016, Fig. 2A). The associations between other clinical characteristics and GC survival are shown in Table S2. Results of multivariate Cox regression also showed that patients with MSS/EBV− appeared to have the worst outcomes (HR = 1.547, 95% CI [0.965–2.481], P = 0.077, Table 2) compared to those with MSI. When MSI and EBV+ cases were combined as a reference group (non-MSS/EBV− subgroup), multivariate analysis revealed that GCs with MSS/EBV− had poorer OS (HR = 1.610, 95% CI [1.046–2.479], P = 0.031, Table S3). Also, PD-L1 expression tended to correlate with improved survival, but multivariate analysis showed marginal significance (HR = 0.684, 95% CI [0.458–1.021], P = 0.063, Table 2).

**MSS/EBV− subgroup could benefit from postoperative chemotherapy**

To determine the effect of postoperative chemotherapy in the MSS/EBV− subgroup, which tended to have the worst clinical outcomes, we analyzed the association of chemotherapy and GC survival stratified for different molecular subgroups. The results showed that postoperative chemotherapy can improve the OS of the MSS/EBV− subgroup (log-rank P < 0.001, Fig. S3C) but showed less benefit when compared with the MSI (log-rank P = 0.327, Fig. S3B) and EBV+ subgroups (log-rank P = 0.355, Fig. S3A). Besides, further multivariate analysis in MSS/EBV− subgroup showed that postoperative chemotherapy
Table 1  Baseline characteristics of enrolled 226 gastric cancer patients.

| Characteristics          | N (%)   |
|--------------------------|---------|
| Gender                   |         |
| Male                     | 172(76.1) |
| Female                   | 54(23.9)  |
| Age(years)               |         |
| <60                      | 92(40.7)  |
| ≥60                      | 134(59.3) |
| Tumor size(cm)           |         |
| <5                       | 67(29.6)  |
| ≥5                       | 159(70.4) |
| WHO classification       |         |
| Tubular adenocarcinoma   | 165(73.0) |
| Mucoid adenocarcinoma    | 28(12.4)  |
| Signet ring cell         | 30(13.3)  |
| Others                   | 3(1.3)   |
| Histological grade       |         |
| Low grade                | 54(23.9)  |
| High grade               | 172(76.1) |
| Vascular invasion        |         |
| Negative                 | 45(19.9)  |
| Positive                 | 181(80.1) |
| Neural invasion          |         |
| Negative                 | 67(29.6)  |
| Positive                 | 159(70.4) |
| CTx                      |         |
| Yes                      | 101(44.7) |
| No                       | 125(55.3) |
| TNM stage                |         |
| I/II                     | 67(29.6)  |
| III                      | 159(70.4) |
| Depth of invasion        |         |
| T1/T2                    | 9(4.0)   |
| T3/T4                    | 217(96.0) |
| Lymph metastasis         |         |
| N0                       | 38(16.8)  |
| N1/N2/N3                 | 188(83.2) |
| MSI status               |         |
| MSI                      | 54(23.9)  |
| MSS                      | 172(76.1) |
| EBV                      |         |
| Positive                 | 13(5.8)   |
| Negative                 | 213(94.2) |
| PD-L1                    |         |
| Positive                 | 93(41.2)  |
| Negative                 | 133(58.8) |

Notes.
MSI, Microsatellite instability; MSS, Microsatellite stable; EBV, Epstein-Barr Virus; PD-L1, Programmed cell death ligand 1; CTx, Chemotherapy.

could improve OS of the MSS/EBV\(^{-}\) GCs (HR = 0.452, 95% CI [0.299–0.682], \(P < 0.001\), Table 3)

**PD-L1 expression could predict the prognosis of GC patients with MSS/EBV\(^{-}\) subtype**

PD-L1 positivity was associated with the EBV\(^{+}\) cases. EBV\(^{+}\) cases manifested more PD-L1\(^{+}\) tumors compared with the MSI and MSS/EBV\(^{-}\) cases (81.8% vs. 50.0% vs. 35.4%, \(P = 0.001\)).

Then, we evaluated the prognostic value of PD-L1 expression in different subgroups to accurately assess the prognosis of GC patients with adverse molecular typing. We found
Figure 1  Molecular features of tumor issue. (A–H) Immunohistochemistry for four MMR proteins in the nucleus of tumor cells: (A) Positive MLH1 expression. (B) The loss of the expression of MLH1. (C) Positive MSH2 expression. (D) The loss of the expression of MSH2. (E) Positive MSH6 expression. (F) The loss of the expression of MSH6. (G) Positive PMS2 expression. (H) The loss of the expression of PMS2. (I) EBV-positive case with strong brown nuclear ISH staining. (J) Negative detection of EBV in tumor cells. (K) PD-L1 expression in tumor cells (both in cell membrane and cytoplasm staining). (L) Negative detection of EBV in tumor cells. Magnification: 200×.

Figure 2  Kaplan–Meier estimates of overall survival (OS) of gastric cancer patients according to (A) three molecular subgroups (B) Chemotherapy in MSS/EBV−/PD-L1+ and MSS/EBV+/PD-L1− subgroup. MSI, Microsatellite instability; MSS, Microsatellite stable; EBV, Epstein-Barr Virus; PD-L1, Programmed cell death ligand 1; CTx, Chemotherapy.

in the MSS/EBV− subgroup, which tended to show the worst clinical outcomes, that PD-L1-positive GCs were found to be related to a better prognosis compared to those who were PD-L1-negative (P < 0.001, Fig. S3F). In contrast, there was no significant survival difference between patients who were PD-L1-positive and those without PD-L1
Table 2  Multivariate analyses of risk factors affecting overall survival (OS) in 224 gastric cancer patients.

| Characteristics          | HR     | 95% CI          | P    |
|--------------------------|--------|-----------------|------|
| Molecular subtypes       | MSI (n = 52)  | 1.00            |      |
|                          | EBV+ (n = 11) | 0.755           | 0.275–2.076 | 0.077 |
|                          | MSS/EBV− (n = 161) | 1.547           | 0.965–2.481 |
| PD-L1                    | Positive | 1.00            |      |
|                          | Negative | 0.684           | 0.458–1.021 | 0.063 |
| TNM stage                | I/II    | 1.00            |      |
|                          | III     | 2.117           | 1.208–3.709 | 0.009 |
| Vascular invasion        | Negative | 1.00            |      |
|                          | Positive | 2.216           | 1.135–4.330 | 0.020 |
| CTx                      | No      | 1.00            |      |
|                          | Yes     | 0.477           | 0.331–0.686 | <0.001 |

Notes. MSI, Microsatellite instability; MSS, Microsatellite stable; EBV, Epstein-Barr virus; PD-L1, Programmed cell death ligand 1; CTx, Chemotherapy; HR, Hazard ratio; CI, Confidence interval. 95% CI and P values were calculated with multivariate Cox regression, adjusting for the variables that P < 0.100 from the univariate analysis, such as PD-L1, WHO classification, histological grade, vascular invasion, neural invasion, postoperative chemotherapy and TNM stage.

Table 3  Multivariate analyses of risk factors affecting overall survival (OS) in MSS/EBV− subgroup (N = 161).

| Characteristics          | HR     | 95% CI          | P    |
|--------------------------|--------|-----------------|------|
| PD-L1                    | Negative | 1.00            |      |
|                          | Positive | 0.612           | 0.389–0.962 | 0.033 |
| Histological grade       | Low grade | 1.00            |      |
|                          | High grade | 1.834           | 1.083–3.107 | 0.024 |
| TNM stage                | I/II    | 1.00            |      |
|                          | III     | 3.342           | 1.798–6.212 | <0.001 |
| CTx                      | No      | 1.00            |      |
|                          | Yes     | 0.452           | 0.299–0.682 | <0.001 |

Notes. MSI, microsatellite instability; MSS, microsatellite stable; EBV, Epstein-Barr Virus; PD-L1, programmed cell death ligand 1; CTx, Chemotherapy; HR, Hazard ratio; CI, Confidence interval. 95% CI and P values were calculated with multivariate Cox regression, adjusting for the variables that P < 0.100 from the univariate analysis, such as WHO classification, histological grade, vascular invasion, neural invasion, postoperative chemotherapy and TNM stage.

expression for the MSI (P = 0.656, Fig. S3E) and EBV+ (P = 0.765, Fig. S3D) subgroups. Moreover, further multivariate analysis showed that PD-L1-positive could serve as an independent predictive factor for better OS in the MSS/EBV− subgroup (HR = 0.612, 95% CI [0.389–0.962], P = 0.033, Table 3).

PD-L1 expression could predict the response to chemotherapy of GC patients with MSS/EBV− subtype
Because both PD-L1 and chemotherapy can predict better OS in the MSS/EBV− subgroup, we explored the association of PD-L1 expression with the efficacy of chemotherapy in the MSS/EBV− subgroup. As shown in Fig. 2B, the log rank test showed that postoperative
Chemotherapy can improve OS of the MSS/EBV⁻/PD-L1⁺ subgroup (log-rank $P = 0.005$), while showed less benefit for the MSS/EBV⁻/PD-L1⁻ subgroup and the result was not statistically significant (log-rank $P = 0.061$). Further multivariate analysis indicated that chemotherapy was associated with an improved prognosis for PD-L1-negative MSS/EBV⁻ GCs (HR = 0.357, 95% CI [0.217–0.587], $P < 0.001$; Table S4), while it was not a significant prognostic factor for PD-L1-positive MSS/EBV⁻ GCs (HR = 0.499, 95% CI [0.223–1.117], $P = 0.090$; Table S4).

DISCUSSION
Recently, more evidence has been emerged about the potential prognostic and therapeutic implications of molecular subtypes, particularly through the stratification according to EBV and MSI status. According to The Cancer Genome Atlas (TCGA) study and recent clinical trials, EBV and MSI GC subgroups exhibited high expression of PD-L1 and could benefit from therapy with PD-1/PD-L1 antibodies. However, very few studies employed combinatorial analysis of MSI, EBV-infection and PD-L1 to evaluate the prognosis and chemotherapy efficacy of gastric cancer. As postoperative chemotherapy is still the first-line treatment for GC patients, so we try to combine conventional chemotherapy with molecular subtyping and PD-L1 expression, which may help to select the patients who would benefit the most from chemotherapy and provide evidence for individualized treatment of patients with GC. In the present study, we observed that GC patients with MSS/EBV⁻ were associated with the worst prognosis compared with those with MSI and EBV⁺. Beyond that, we found that PD-L1-positive expression can predict better OS in the MSS/EBV⁻ subgroup. At the same time, MSS/EBV⁻ GCs can also benefit more significantly from chemotherapy after surgery. Nevertheless, MSS/EBV⁻ GCs with PD-L1⁺ cannot benefit from postoperative chemotherapy.

Among the 226 GC patients in our study, 54 (23.9%) patients were defined as having MSI by IHC, which was similar to that reported in TCGA cohort (22%) (Cancer Genome Atlas Research Network. 2014) and in ACRG Korean cohort (23%) (Angell et al., 2019). The incidence of EBV-associated GC ranged from 4–18% among different regions (Pikutula et al., 2020), our result (4.9%) is identical to the data of a recent Korean study (25/569, 4.4%) (Cho, Kang & Kim, 2019) and Taiwan study (65/1248, 5.2%) (Huang et al., 2019) that used the same ISH method. A review of PD-L1 expression in gastric cancer (Vrána et al., 2018) showed that the percentage of PD-L1⁺ in the TCs ranges from 9% to 49.1% (most of the trials were performed in Asian populations), similar to our results (41.2% in TCs). Previous studies have demonstrated a mutually exclusive relationship between GCs that were EBV-positive and deemed to have MSI (Choi et al., 2020). However, two cases representing both MSI and EBV⁺ were observed in a recent study (2/287, 0.70%) (Pereira et al., 2018) and in our study (2/226, 0.88%). Previous studies have demonstrated that MSI and EBV⁺ subtypes were separately associated with better prognoses (Sohn et al., 2017), and our result is in line with the results from Pereira et al. (2018) and De Rosa et al. (2018) that suggested MSS/EBV⁻ GCs showed the worst OS compared with MSI and EBV⁺ GCs. The potential mechanism underlying better survival of these two GC subtypes
is that intense inflammatory infiltration observed in EBV-positive and MSI GCs may play an important role in antitumor effects. CD8+ cytotoxic T cells were found to be more prevalent in these two subgroups based on the stimulation of many neo-antigens (existing in MSI and EBV+ tumors), which can promote the elimination of TCs (Choi et al., 2020).

As a standard regimen, postoperative chemotherapy has been recommended for patients with stage II or III GC. Nevertheless, its application to all stage II or III GC patients is unnecessary and may even be detrimental to some patients (Jiang et al., 2017). Among these, GCs with MSI may experience a detrimental effect from chemotherapy (especially for 5-FU based treatment), and this finding has been supported by several studies (Kim et al., 2015). Our study yielded a similar result, namely no significant benefit from postoperative chemotherapy (5-FU-based) was observed for MSI and EBV+ GCs. In this regard, cancer immunotherapy might offer a promising new option and has been demonstrated effective in MSI and EBV+ GCs. A recent phase II trial described the high overall response rates in MSI-H and EBV-positive GCs with pembrolizumab treatment in metastatic GCs (ORR: 85.7% for MSI-H and 100.0% for EBV-positive) (Kim et al., 2018). A possible explanation is that both MSI and EBV+ GCs are characterized by elevated mutation rates and high expression of PD-L1 and PD-L2, which can be used as possible targets for immunotherapy (Sohn et al., 2017), especially PD-1/PD-L1 directed therapy (Le et al., 2017). On the other hand, our results indicated that MSS/EBV− GCs, which was associated with a poorer prognosis, showed improved OS with the administration of chemotherapy after surgery.

In our study, we observed that both EBV-positive and MSI GCs showed a relatively higher proportion of PD-L1+ expression than MSS/EBV− GCs. This result is consistent with reports in previous studies (Angell et al., 2019), which can be explained by observed amplification of the PD-L1 gene in EBV-positive GCs by TCGA study (Cancer Genome Atlas Research Network. 2014) and adaptive immune resistance in the tumor microenvironment. Intratumor lymphocytes can release interferon-γ to induce the upregulation of PD-L1 in TCs (Pereira et al., 2018). However, the positive expression rate of PD-L1 in MSS/EBV− TCs varies greatly in different studies though there is few relevant studies, which could be attributed to the various method of PD-L1 expression determination and cut-off value of calling positive status used among different studies. In the study of De Rosa et al. (2018), only 3% PD-L1+ cases were found in MSS/EBV− subgroup and a case was considered positive if membrane immunostaining was present in at least 5% of tumor cells, independently of the intensity. Choi et al. (2020) found 14% PD-L1-positive cases in MSS/EBV− when considering both staining intensity and percentage of stained tumor cells, and any membranous staining was regarded as positive expression. In the present study, a higher proportion of PD-L1+ cases in MSS/EBV− group (35%) are reported when scoring PD-L1 expression on the basis of both the intensity and percentage of PD-L1-positive TCs and considering both cell membrane and cytoplasm staining. We combined molecular subtypes with PD-L1 expression and found that PD-L1+ can predict a better OS in MSS/EBV− subgroup but not in the other two subgroups. In contrast to our results, De Rosa et al. (2018) reported that PD-L1+ cannot predict the prognosis for GC patients with MSS/EBV−; however, the reliability of their result is doubtful due to a very limited number of cases with PD-L1+ (n = 2) in the MSS/EBV− subgroup.
PD-L1 expression in TCs might be attributed to the response to endogenous host immunity (Kim et al., 2016). For the MSS/EBV− subtype with a lower immune infiltration, the expression of PD-L1 might suggest the activation of the immune response in the tumor microenvironment, indicating a better prognosis.

Although MSS/EBV− GCs can benefit from chemotherapy, it is still unclear whether there are new markers, especially PD-L1, that could subgroup them and better guide the selection of chemotherapy or immunotherapy. In the current study, chemotherapy could improve the prognosis for PD-L1-negative MSS/EBV− GCs but not for PD-L1-positive MSS/EBV− GCs. One possible explanation is that the survival benefit of PD-L1+ in this subtype could be attenuated by the immunosuppression caused by chemotherapy toxicity (Choi et al., 2019). Thus, for MSS/EBV− GCs with PD-L1−, with an obvious benefit from chemotherapy, chemotherapy after surgery should be recommend for GCs of this subgroup. Even though MSS/EBV− GCs with PD-L1+ exhibit better OS, we cannot give up evaluating the potential use of immune checkpoint inhibitors targeted at PD-L1 for this distinct subgroup of GCs considering the low efficacy of chemotherapy.

Therefore, we suggest that stratification by MSI and EBV statuses combined with PD-L1 expression should be used as a potential strategy to predict prognosis and identify GC patients who are most likely to respond to chemotherapy after surgery. This predictive strategy has the potential to guide patient subtype-oriented therapeutic decisions and avoid treating a certain proportion of resectable GC patients with unnecessary treatment.

Some limitations in the present study should be addressed. Firstly, only 13 patients were identified as EBV+ due to the low incidence of EBV-associated GC among Chinese people, and this could lead to a low level of statistical efficacy in EBV subgroup analysis. Further study with more patients is needed to verify our results. Secondly, the median follow-up time (39.7 months) was relatively short in our study. Clinical outcomes were not observed in 42% patients, and longer follow-up was needed. With the development of clinical practice, there have been PD-L1 test kit (PD-L1 IHC 22C3 pharmDx) approved by FDA as a companion diagnostic assay for the use of pembrolizumab in GC clinical treatment (Fashoyin-Aje et al., 2019). However, due to our earlier research, we used PD-L1 antibodies for scientific research, and the consistency between them needs further study.

**CONCLUSIONS**

In summary, we observed that MSS/EBV− GCs were associated with the worst OS compared to that of GC patients with MSI and EBV+ subtypes. PD-L1 expression was associated with the MSI and EBV+ subtypes. MSS/EBV− GCs could also gain a survival benefit from postoperative chemotherapy. On the other hand, PD-L1 positivity could predict a better prognosis for MSS/EBV− GCs, whereas MSS/EBV− GCs with PD-L1+ cannot gain survival benefit from chemotherapy. Thus, molecular typing plus PD-L1 expression could improve prognostic prediction for GC patients and help choose more effective treatments for specific subsets of GCs.
ACKNOWLEDGEMENTS
The authors would like to thank Song Ying for her work on follow-up of participants and Changsong Guo for his work on preparation of laboratory samples.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding
This work was supported by the National Natural Science Foundation of China under Grant numbers 81874279, 81703293; National Key R&D Program of China under Grant number 2017YFC0907405; the Science and Technology Department of Jilin Province under Grant numbers 20200201442JC, 20200201326JC. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures
The following grant information was disclosed by the authors:
National Natural Science Foundation of China: 81874279, 81703293.
National Key R&D Program of China: 2017YFC0907405.
Science and Technology Department of Jilin Province: 20200201442JC, 20200201326JC.

Competing Interests
The authors declare there are no competing interests.

Author Contributions
• Na Yang and Yanhua Wu conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.
• Meishan Jin performed the experiments, prepared figures and/or tables, and approved the final draft.
• Zhifang Jia and Donghui Cao conceived and designed the experiments, prepared figures and/or tables, and approved the final draft.
• Yueqi Wang, Lili Qin, Xueying Wang and Min Zheng analyzed the data, prepared figures and/or tables, collected clinical data, and approved the final draft.
• Xueyuan Cao and Jing Jiang conceived and designed the experiments, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.

Human Ethics
The following information was supplied relating to ethical approvals (i.e., approving body and any reference numbers):
This research was approved by the Human Ethics Review Committee of the First Hospital of Jilin University (Project identification code: 2017-090).

Data Availability
The following information was supplied regarding data availability:
The raw measurements are available in the Supplemental File.
Supplemental Information
Supplemental information for this article can be found online at http://dx.doi.org/10.7717/peerj.11481#supplemental-information.

REFERENCES

Angell HK, Lee J, Kim KM, Kim K, Kim ST, Park SH, Kang WK, Sharpe A, Ogden J, Davenport A, Hodgson DR, Barrett JC, Kilgour E. 2019. PD-L1 and immune infiltrates are differentially expressed in distinct subgroups of gastric cancer. *Oncoimmunology* 8:e1544442 DOI 10.1080/2162402x.2018.1544442.

Bae YS, Kim H, Noh SH, Kim H. 2015. Usefulness of Immunohistochemistry for Microsatellite Instability Screening in Gastric Cancer. *Gut Liver* 9:629–635 DOI 10.5009/gnl15133.

Cancer Genome Atlas Research Network 2014. Comprehensive molecular characterization of gastric adenocarcinoma. *Nature* 513:202–209 DOI 10.1038/nature13480.

Chang YH, Heo YJ, Cho J, Song SY, Lee J, Kim KM. 2018. Computational measurement of tumor immune microenvironment in gastric adenocarcinomas. *Scientific Reports* 8:13887 DOI 10.1038/s41598-018-32299-0.

Chao X, Li L, Wu M, Ma S, Tan X, Zhong S, Bi Y, Lang J. 2019. Comparison of screening strategies for Lynch syndrome in patients with newly diagnosed endometrial cancer: a prospective cohort study in China. *Cancer Communications* 39:42 DOI 10.1186/s40880-019-0388-2.

Cho J, Kang SY, Kim KM. 2019. MMR protein immunohistochemistry and microsatellite instability in gastric cancers. *Pathology* 51:110–113 DOI 10.1016/j.pathol.2018.09.057.

Choi E, Chang MS, Byeon SJ, Jin H, Jung KC, Kim H, Lee KL, Kim W, Park JH, Kim KH, Kim JS, Choi IS, Han DS, Ahn HS, Heo SC. 2020. Prognostic perspectives of PD-L1 combined with tumor-infiltrating lymphocytes, Epstein-Barr virus, and microsatellite instability in gastric carcinomas. *Diagnostic Pathology* 15:69 DOI 10.1186/s13000-020-00979-z.

Choi YY, Kim H, Shin SJ, Kim HY, Lee J, Yang HK, Kim WH, KimYW, Kook MC, Park YK, Kim HH, Lee HS, Lee KH, Gu MJ, Choi SH, Hong S, Kim JW, Hyung WJ, Noh SH, Cheong JH. 2019. Microsatellite instability and programmed cell death-ligand 1 expression in stage II/III gastric cancer: post hoc analysis of the CLASSIC randomized controlled study. *Annals of Surgery* 270:309–316 DOI 10.1097/sla.0000000000002803.

De Rosa S, Sahnane N, Tibiletti MG, Magnoli F, Vanoli A, Sessa F, Chiaravalli AM. 2018. EBV+ and MSI gastric cancers harbor high PD-L1/PD-1 expression and high CD8+ intratumoral lymphocytes. *Cancers* 10:102 DOI 10.3390/cancers10040102.

Derks S, Liao X, Chiaravalli AM, Xu X, Camargo MC, Solcia E, Sessa F, Fleitas T, Freeman GJ, Rodig SJ, Rabkin CS, Bass AJ. 2016. Abundant PD-L1 expression in Epstein-Barr Virus-infected gastric cancers. *Oncotarget* 7:32925–32932 DOI 10.18632/oncotarget.9076.
Digklia A, Wagner AD. 2016. Advanced gastric cancer: current treatment landscape and future perspectives. World Journal of Gastroenterology 22:2403–2414 DOI 10.3748/wjg.v22.i8.2403.

Fashoyin-Aje L, Donoghue M, Chen H, He K, Veeraraghavan J, Goldberg KB, Keegan P, McKee AE, Pazdur R. 2019. FDA Approval Summary: pembrolizumab for recurrent locally advanced or metastatic gastric or gastroesophageal junction adenocarcinoma expressing PD-L1. Oncologist 24:103–109 DOI 10.1634/theoncologist.2018-0221.

Goel A, Nagasaka T, Hamelin R, Boland CR. 2010. An optimized pentaplex PCR for detecting DNA mismatch repair-deficient colorectal cancers. PLOS ONE 5:e9393 DOI 10.1371/journal.pone.0009393.

Huang SC, Ng KF, Yeh TS, Cheng CT, Lin JS, Liu YJ, Chuang HC, Chen TC. 2019. Subtraction of Epstein-Barr virus and microsatellite instability genotypes from the Lauren histotypes: combined molecular and histologic subtyping with clinicopathological and prognostic significance validated in a cohort of 1, 248 cases. International Journal of Cancer 145:3218–3230 DOI 10.1002/ijc.32215.

Jiang Y, Li T, Liang X, Hu Y, Huang L, Liao Z, Zhao L, Han Z, Zhu S, Wang M, Xu Y, Qi X, Liu H, Yang Y, Yu J, Liu W, Cai S, Li G. 2017. Association of adjuvant chemotherapy with survival in patients with stage II or III gastric cancer. JAMA Surgery 152:e171087 DOI 10.1001/jamasurg.2017.1087.

Jin S, Xu B, Yu L, Fu Y, Wu H, Fan X, Wei J, Liu B. 2017. The PD-1, PD-L1 expression and CD3+ T cell infiltration in relation to outcome in advanced gastric signet-ring cell carcinoma, representing a potential biomarker for immunotherapy. Oncotarget 8:38850–38862 DOI 10.18632/oncotarget.16407.

Kawazoe A, Kuwata T, Kuboki Y, Shitara K, Nagatsuma AK, Aizawa M, Yoshino T, Doi T, Ohtsu A, Ochiai A. 2017. Clinicopathological features of programmed death ligand 1 expression with tumor-infiltrating lymphocyte, mismatch repair, and Epstein-Barr virus status in a large cohort of gastric cancer patients. Gastric Cancer 20:407–415 DOI 10.1007/s10120-016-0631-3.

Kim SY, Choi YY, An JY, Shin HB, Jo A, Choi H, Seo SH, Bang HJ, Cheong JH, Hyung WJ, Noh SH. 2015. The benefit of microsatellite instability is attenuated by chemotherapy in stage II and stage III gastric cancer: results from a large cohort with subgroup analyses. International Journal of Cancer 137:819–825 DOI 10.1002/ijc.29449.

Kim ST, Cristescu R, Bass AJ, Kim KM, Odegaard JI, Kim K, Liu XQ, Sher X, Jung H, Lee M, Lee S, Park SH, Park JO, Park YS, Lim HY, Lee H, Choi M, Talasaz A, Kang PS, Cheng J, Loboda A, Lee J, Kang WK. 2018. Comprehensive molecular characterization of clinical responses to PD-1 inhibition in metastatic gastric cancer. Nature Medicine 24:1449–1458 DOI 10.1038/s41591-018-0101.

Kim JW, Nam KH, Ahn SH, Park DJ, Kim HH, Kim SH, Chang H, Lee JO, Kim YJ, Lee HS, Kim JH, Bang SM, Lee JS, Lee KW. 2016. Prognostic implications of immunosuppressive protein expression in tumors as well as immune cell infiltration....
within the tumor microenvironment in gastric cancer. *Gastric Cancer* **19**:42–52 DOI 10.1007/s10120-014-0440-5.

Le DT, Durham JN, Smith KN, Wang H, Bartlett BR, Aulakh LK, Lu S, Kemberling H, Wilt C, Luber BS, Wong F, Azad NS, Rucki AA, Laheru D, Donehower R, Zaheer A, Fisher GA, Crocenzi TS, Lee JJ, Greten TF, Duffy AG, Ciombor KK, Eyring AD, Lam BH, Joe A, Kang SP, Holdhoff M, Danilova L, Cope L, Meyer C, Zhou S, Goldberg RM, Armstrong DK, Bever KM, Fader AN, Taube J, Housseau F, Spetzler D, Xiao N, Pardoll DM, Papadopoulos N, Kinzler KW, Eshleman JR, Vogelstein B, Anders RA, Diaz Jr LA. 2017. Mismatch repair deficiency predicts response of solid tumors to PD-1 blockade. *Science* **357**:409–413 DOI 10.1126/science.aan6733.

Lee KS, Kwak Y, Ahn S, Shin E, Oh HK, Kim DW, Kang SB, Choe G, Kim WH, Lee HS. 2017. Prognostic implication of CD274 (PD-L1) protein expression in tumor-infiltrating immune cells for microsatellite unstable and stable colorectal cancer. *Cancer Immunology and Immunotherapy* **66**:927–939 DOI 10.1007/s00262-017-1999-6.

Liu X, Choi MG, Kim K, Kim KM, Kim ST, Park SH, Cristescu R, Peter S, Lee J. 2020. High PD-L1 expression in gastric cancer (GC) patients and correlation with molecular features. *Pathology, Research and Practice* **216**:152881 DOI 10.1016/j.prp.2020.152881.

Lyons K, Le LC, Pham YT, Borron C, Park JY, Tran CTD, Tran TV, Tran HT, Vu KT, Do CD, Pelucchi C, La Vecchia C, Zgibor J, Boiffetta P, Luu HN. 2019. Gastric cancer: epidemiology, biology, and prevention: a mini review. *European Journal of Cancer Prevention* **28**:397–412 DOI 10.1097/cej.0000000000000480.

Muro K, Chung HC, Shankaran V, Geva R, Catenacci D, Gupta S, Eder JP, Golan T, Le DT, Burtness B, McRee AJ, Lin CC, Pathiraja K, Lunceford J, Emancipator K, Juco J, Koshiji M, Bang YJ. 2016. Pembrolizumab for patients with PD-L1-positive advanced gastric cancer (KEYNOTE-012): a multicentre, open-label, phase 1b trial. *The Lancet Oncology* **17**:717–726 DOI 10.1016/s1470-2045(16)00175-3.

Pereira MA, Ramos M, Faraj SF, Dias AR, Yagi OK, Zilberstein B, Cecconello I, Alves VAF, de Mello ES, Ribeiro Jr U. 2018. Clinicopathological and prognostic features of Epstein-Barr virus infection, microsatellite instability, and PD-L1 expression in gastric cancer. *Journal of Surgical Oncology* **117**:829–839 DOI 10.1002/jso.25022.

Pikuła A, Kwietniewska M, Rawicz-Pruszyński K, Cisiel B, Skórzewska M, Gęca K, Franciszkiewicz-Pietrzak K, Kurylicio A, Mielko J, Polkowski WP. 2020. The importance of Epstein-Barr virus infection in the systemic treatment of patients with gastric cancer. *Seminars in Oncology* **47**:127–137 DOI 10.1053/j.seminoncol.2020.04.001.

Smyth EC, Wotherspoon A, Peckitt C, Gonzalez D, Hulkki-Wilson S, Eltahir Z, Fassan M, Rugge M, Valeri N, Okines A, Hewish M, Allum W, Stenning S, Nankivell M, Langley R, Cunningham D. 2017. Mismatch repair deficiency, microsatellite instability, and survival: an exploratory analysis of the medical research council adjuvant gastric infusional chemotherapy (MAGIC) trial. *JAMA Oncology* **3**:1197–1203 DOI 10.1001/jamaoncol.2016.6762.
Sohn BH, Hwang JE, Jang HJ, Lee HS, Oh SC, Shim JJ, Lee KW, Kim EH, Yim SY, Lee SH, Cheong JH, Jeong W, Cho JY, Kim J, Chae J, Lee J, Kang WK, Kim S, Noh SH, Ajani JA, Lee JS. 2017. Clinical significance of four molecular subtypes of gastric cancer identified by the cancer genome atlas project. Clinical Cancer Research 23:4441–4449 DOI 10.1158/1078-0432.Ccr-16-2211.

Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. 2021. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA: A Cancer Journal for Clinicians Epub ahead of print Feb 04 2021 DOI 10.3322/caac.21660.

Tan Z. 2019. Recent advances in the surgical treatment of advanced gastric cancer: a review. Medical Science Monitor 25:3537–3541 DOI 10.12659/msm.916475.

Vrána D, Matzenauer M, Neoral Č, Aujeský R, Vrba R, Melichar B, Rušarová N, Bartoušková M, Jankowski J. 2018. From tumor immunology to immunotherapy in gastric and esophageal cancer. International Journal of Molecular Sciences 20:13 DOI 10.3390/ijms20010013.

Wang SM, Zheng RS, Zhang SW, Zeng HM, Chen R, Sun KX, Gu XY, Wei WW, He J. 2019. Epidemiological characteristics of gastric cancer in China, 2015. Zhonghua Liu Xing Bing Xue Za Zhi 40:1517–1521 DOI 10.3760/cma.j.issn.0254-6450.2019.12.003.

Wu Y, Cao D, Qu L, Cao X, Jia Z, Zhao T, Wang Q, Jiang J. 2017. PD-1 and PD-L1 co-expression predicts favorable prognosis in gastric cancer. Oncotarget 8:64066–64082 DOI 10.18632/oncotarget.19318.

Yang L, Zheng R, Wang N, Yuan Y, Liu S, Li H, Zhang S, Zeng H, Chen W. 2018. Incidence and mortality of stomach cancer in China, 2014. Chinese Journal of Cancer Research 30:291–298 DOI 10.21147/j.issn.1000-9604.2018.03.01.