Study of bacterial flora associated with mobile phones of healthcare workers and non-healthcare workers

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Received: March 2017, Accepted: May 2017

ABSTRACT

Background and Objectives: Despite improvements in modern diagnosis and therapies, hospital acquired infections remain a leading problem of global health systems. Healthcare workers mobile phones is a reservoir for potential pathogens. Despite the high possibility of being contaminated, mobile phones are rarely clean and are often touched during or after examination of patients and handling of specimens without proper hand washing. The main objective of the present study was to isolate, identify different types of bacteria and their antibiotic sensitivity from mobile phones of healthcare workers and non-healthcare workers.

Materials and Methods: Samples were collected aseptically by rolling over the exposed surfaces of the mobile phones inoculated on the agar plates and incubated aerobically. After incubation, plates were examined for growth. Bacteria were identified and antibiotic sensitivity was tested as per standard microbiological procedures.

Results: In this study a total of 175 samples were examined, out of which 125 samples were from healthcare workers (HCWs), 50 samples were from non-healthcare workers (non-HCWs). Among the mobile phones of HCW’s from ICUs, Acinetobacter baumannii (36.84%) was the predominant organism isolated followed by methicillin resistant Staphylococcus aureus (MRSA) (21.05%). Predominant organism isolated from HCW’s in operation theater was MRSA (46.66%). Out of 50 worker’s non-HCWs mobile phones samples cultured, 23 (46.00%) samples yielded growth of six different types of bacteria.

Conclusion: Our study reveals that there is definite colonization of bacteria on mobile phones of the HCWs. It is not only capable of transferring message but also disease-producing microbes. In order to reduce incidence of nosocomial infections, there should be implementation of hand washing practices and regulations around the use of mobile telephones in hospital settings.

Keywords: Healthcare-associated infections, Mobile phones, Staphylococcus species, Acinetobacter baumannii

INTRODUCTION

Modern technology has contributed significantly to the field of Medicine, by developing newer techniques in diagnosis, patient-care and treatment, which has significantly increased survival of diseased individual. Modern technology, which is growing at
a rapid phase, has also contributed in developing technologies for individual use. This technology includes personal computers, pagers, mobile hand-held devices (wireless tablets, etc.) and mobile phones (1-3). In 1983, in order to improve the communication system, the global system for mobile telecommunication was established in Europe. In India, the first use of mobile phone was in 1995 and today more than 287 million mobile phone users exist, which account for 85% of all the telecommunication users (4, 5). In many countries, mobile phones outnumber landline telephones. Most adult and many children now own mobile phones (6).

Mobile phones increase the speed of communication and contact within healthcare institutions, making healthcare delivery more efficient. Mobile phones dispense laboratory and imaging results, patient data, and photographic images, which are being used by physicians during bedside rounds, in order to engage clinicians, residents, and students. HCWs access pharmaceutical knowledge and literature by mobile phone, which facilitates learning and clinical performance (7, 8). Due to these benefits of mobile phones and computers, their hazard to human health is often overlooked (6). Potential risks of using mobile phones can lead to noise, distractions, loss of concentration, data safety, and disturbance of patient privacy and transfer of micro-organisms possibly leading to nosocomial infections (9). In 2000, World Health Organization (WHO) also described the electromagnetic radiation emitted from phones and base stations as a threat to lives, as it damages the DNA producing sperm cells (10).

The human skin is constantly in contact with micro-organisms and becomes readily colonized by certain microbial species. The adult human is covered with approximately 2m² of skin, with surface area supporting about 10¹² bacterial cells/person (11). During a phone call, the mobile phone comes into close contact with contaminated human body areas with hands to hands, and hands to other areas like mouth, nose and ears (12), which may result in colonization of potential pathogens present on the human skin, on the mobile phones. In 1997, Aronson et al. first suggested the infection potential of telephones (13).

Despite improvements in modern diagnosis and therapies, healthcare-associated infections (HAIs) remain a leading problem of global health systems. In developing countries, approximately 25% of patients are found to acquire HAIs (14). Hands of HCWs play an important role in the transmission of HAIs. Gowns, gloves, bedside stethoscopes, neckties, bed rails, sheets, telephones, horizontal surfaces, door knobs, thermometers, nurse’s clothings and personal bags are contaminated by pathogenic bacteria. During daily rounds, hands of HCWs are contaminated with pathogenic bacteria present on these inanimate objects and these bacteria will be transmitted to the patients. HCWs mobile phones provide a reservoir for these potential pathogens. Despite the high possibility of being contaminated, mobile phones are rarely clean and are often touched during or after examination of patients and handling of specimens without proper hand washing. These mobile phones become exogenous sources of infection, for not only the patients but also potential health hazard for workers as well as for family members (15). Further, sharing of cell phones among HCWs and non-HCWs may directly facilitate the spread of potential pathogenic bacteria to the community (16).

The range of micro-organisms, which are present can vary from one person to another, and HCWs may have different hand flora from ordinary members of the public. Bacterial isolates from cell phones of HCWs may vary in numbers and antibiotic sensitivity compared to cell phones of non-medical personnel. Cell phones of HCWs represent a hospital community and non-HCWs represent an environment of community. Hence, the aim of this study was to examine different types of bacteria present in these two categories of personnel’s.

MATERIALS AND METHODS

Sample collection. Samples were collected aseptically with sterile swabs moistened with sterile normal saline and by rolling over the exposed surfaces of the mobile phones. Maximum care was taken to ensure that all the buttons of the keypad, screen, mouthpiece, earpiece, sides and back of the mobiles were properly swabbed since these areas are the most frequent spots, in contact with the fingers.

Sample inoculation. After collection, the samples were immediately transported to the laboratory and inoculated on 5% sheep blood agar and Mac-Conkey’s agar and plates were incubated aerobically at 37°C for 24 hours. After incubation, plates were ex-
amined for growth and colonial morphology of the isolates. Gram-positive and Gram-negative bacteria were identified as per standard microbiological procedures.

**Antibiotic susceptibility.** Antibiotic sensitivity was tested using the Kirby-Bauer disc diffusion method on Mueller-Hinton agar according to CLSI antibiotic disc susceptibility testing guidelines (17).

The antimicrobial agents tested for Gram-positive cocci were linezolid (30µg), erythromycin (15µg), clindamycin (2µg), ciprofloxacin (5µg), cotrimoxazole (1.25/23.75µg), cefoxitin (30µg) and tetracycline (30µg).

The antimicrobial agents tested for Gram-negative bacilli were pipericillin-tazobactam (100/10 µg), ceftriaxone (30 µg), cefepime (30µg), imipenem (10 µg), cotrimoxazole (1.25/23.75µg), amikacin (30µg), ciprofloxacin (5µg) and ampicillin (10µg).

**RESULTS**

In this study, a total of 175 samples were examined, out of which 125 samples were from HCWs and 50 samples were from non-HCWs. From 125 HCW’s mobile phones, 203 bacteria were isolated. Out of which, 90 (43.68%) were staphylococci as the predominant pathogen, followed by 43 (21.18%) A. baumannii.

Among HCWs, samples were collected from Doctors, nursing staff, medical students and technicians working in various departments like laboratory, ICUs, Operation Theater and general wards. Fifty mobile samples collected from non-HCWs who were not in contact with patients or not visited hospitals during last one month.

Out of 125 HCWs samples, majority of the samples processed were 29 (23.20%) and 21 (16.80%), from Department of Microbiology and Cardio Thoracic Vascular Surgery operation theater (CTVS), respectively. Among healthcare professionals maximum number of samples processed were 54 (43.20%) and 25 (20%), from staff nurses and doctors, respectively. Area-wise and profession-wise distribution of samples is shown in Table 1.

From 125 HCW’s mobile phones, 203 bacteria were isolated. Out of which, 90 (43.68%) were Staphylococcus species, [i.e., MSSA 34 (16.64%), MRSA 31 (15.27%), MRCoNS 12 (5.91%), S. citreus 04 (1.97%)] as the predominant pathogen, followed by 43 (21.18%) A. baumannii. Different types of bacteria grown from HCWs mobile phone are presented in Table 2.

Among the mobile phones of HCW’s from ICUs, A. baumannii (36.84%) was the predominant organism isolated, followed by MRSA (21.05%). Predominant organism isolated from HCW’s in Operation Theater was MRSA (46.66%).

Among 86 (100%) samples positive for staphylococci, excluding S. citreus, 34 (39.53%) were predominantly MSSA. Most of the samples positive

| Area                                   | Doctors | Nurses | Technicians | Students | Area wise distribution of samples |
|-----------------------------------------|---------|--------|-------------|----------|-----------------------------------|
| Department of Microbiology              | 01      | 00     | 12          | 16       | 29                                |
| Pulmonology ward                        | 01      | 07     | 00          | 02       | 10                                |
| Dialysis                                | 00      | 08     | 07          | 00       | 15                                |
| Respiratory Intensive care unit         | 00      | 03     | 01          | 00       | 04                                |
| Medicinal Intensive care unit           | 00      | 06     | 00          | 00       | 06                                |
| Emergency medicinal department          | 05      | 11     | 00          | 00       | 16                                |
| Operation theater complex               | 11      | 01     | 00          | 00       | 12                                |
| General ward                            | 03      | 06     | 00          | 00       | 09                                |
| Cardio thoracic vascular surgery        | 04      | 09     | 04          | 04       | 21                                |
| operation theater                       |         |        |             |          |                                   |
| Infection control Nurses                | 00      | 03     | 00          | 00       | 03                                |
| Profession wise distribution of samples | 25      | 54     | 24          | 22       | 125                               |

Table 1. Distribution of HCW’s mobile phones samples according to profession and area.
Table 2. Number and type of bacteria isolated from HCW’s mobile phones

| Isolated organism                                      | Number of isolated organism (n=203) | Percentage (%) |
|--------------------------------------------------------|-------------------------------------|----------------|
| Methicillin susceptible *Staphylococcus aureus*        | 34                                  | 16.74          |
| Methicillin susceptible coagulase negative *Staphylococci* | 09                                  | 4.43           |
| Methicillin resistant *Staphylococcus aureus*          | 31                                  | 15.27          |
| Methicillin resistant coagulase negative *Staphylococci* | 12                                  | 5.91           |
| *Staphylococcus citreus*                               | 04                                  | 1.97           |
| Diptheroides                                           | 01                                  | 0.49           |
| Gram positive spore bearer                             | 16                                  | 7.88           |
| *Pseudomonas aeruginosa*                               | 39                                  | 19.21          |
| *Acinetobacter baumannii*                              | 43                                  | 21.18          |
| *Klebsiella pneumoniae*                                | 10                                  | 4.92           |
| *Citrobacter spp.*                                     | 02                                  | 0.98           |
| *Escherichia coli*                                     | 02                                  | 0.98           |
| **Total**                                              | **203**                             | **100**        |

Table 3. Distribution of bacteria isolated from HCW’s mobile phones based on the location.

|                         | MSSA | MSCoNS | MRSA | MRCoNS | Diptheroides | Staphylococcus citreus | *Pseudomonas aeruginosa* | *Acinetobacter baumannii* | *Klebsiella pneumoniae* | *Citrobacter spp.* | *Escherichia coli* | Number of total isolated organism (n=203) |
|-------------------------|------|--------|------|--------|--------------|------------------------|-------------------------|--------------------------|------------------------|---------------------|-------------------|------------------------------------------------|
| Dept of Microbiology    | 01   | 01     | 01   | 01     | 01           | 01                     | 01                      | 01                       | 01                     | 01                  | 01                | 50                                           |
| Wards (pulmonalogy,     | 00   | 00     | 00   | 00     | 00           | 00                     | 00                      | 00                       | 00                     | 00                  | 00                | 71                                           |
| general medicin,         |      |        |      |        |              |                        |                         |                          |                         |                     |                   |                                               |
| emergency wards)        |      |        |      |        |              |                        |                         |                          |                         |                     |                   |                                               |
| Dialysis Unit           | 01   | 01     | 01   | 01     | 01           | 01                     | 01                      | 01                       | 01                     | 01                  | 01                | 23                                           |
| Operation theater       | 00   | 00     | 00   | 00     | 00           | 00                     | 00                      | 00                       | 00                     | 00                  | 00                | 15                                           |
| Infection control staff | 00   | 00     | 00   | 00     | 00           | 00                     | 00                      | 00                       | 00                     | 00                  | 00                | 06                                           |
| Nurses                  | 00   | 00     | 00   | 00     | 00           | 00                     | 00                      | 00                       | 00                     | 00                  | 00                | 38                                           |
| ICUS (CTVS, MICU,       | 00   | 00     | 00   | 00     | 00           | 00                     | 00                      | 00                       | 00                     | 00                  | 00                | 203                                          |
| RICU)                   |      |        |      |        |              |                        |                         |                          |                         |                     |                   |                                               |
| **Total**               | 02   | 02     | 02   | 02     | 02           | 02                     | 02                      | 02                       | 02                     | 02                  | 02                | **203**                                      |

MSSA: Methicillin susceptible *Staphylococcus aureus*; MSCoNS: Methicillin susceptible coagulase negative *Staphylococci*; MRSA: Methicillin resistant *Staphylococcus aureus*; MRCoNS: Methicillin resistant coagulase negative staphylococci; GPB: Gram positive spore bearer

ICUS: Intensive care units; CTVS: Cardio thoracic vascular surgery operation theatre; MICU: Medicinal Intensive care unit; RICU: Respiratory Intensive care unit
samples were cultured, out of which, 23 (46.00%) samples yielded growth of six different types of bacteria. Out of which, Gram-positive spore bearer 16 (57.14%) was the predominant organism followed by *Acinetobacter baumannii* (04) (14.28%). Distribution of bacteria from non-HCWs is shown in Table 5.

Antibiotic susceptibility pattern of staphylococci isolated from HCW’s mobile phones is shown in Table 6. *S. aureus* and coagulase negative staphylococcus (CoNS) were 100% susceptible to linezolid.

Among Gram-negative bacteria, 97.67% of *A. baumannii* was susceptible to amikacin, followed by ciprofloxacin (90.69%). 97.43% of *P. aeruginosa* was susceptible to imipenem, followed by amikacin (94.87%). Antibiotic susceptibility pattern of Gram-negative organisms isolated from HCW’s is shown in Table 7.

Antibiotic susceptibility pattern of staphylococci isolated from non-HCWs showed 100% sensitivity to all antibiotics tested and all the four *A. baumannii* were susceptible to piperacillin-tazobactam, ceftriaxone, cefepime, imipenem, amikacin, ciprofloxacin, and only one isolate was susceptible to ampicillin. All

Table 4. Distribution of bacteria isolated from HCW’s mobile phones according to the profession

| Isolated micro-organisms | Doctors (n=25) | Nurses (n=54) | Technicians (n=24) | Student (n=22) |
|--------------------------|---------------|---------------|--------------------|---------------|
| MSSA (34)                | 06            | 13            | 09                 | 06            |
| MSCoNS (09)              | 01            | 03            | 03                 | 02            |
| MRSA (31)                | 09            | 12            | 05                 | 05            |
| MRCoNS (12)              | 05            | 03            | 01                 | 03            |
| *Staphylococcus citreus* (04) | 01            | 02            | 01                 | 00            |
| GPB (16)                 | 04            | 06            | 03                 | 03            |
| Diptheroides (01)        | 00            | 00            | 00                 | 01            |
| *K. pneumoniae* (10)     | 02            | 06            | 02                 | 00            |
| *P. aeruginosa* (39)     | 06            | 21            | 08                 | 04            |
| *A. baumannii* (43)      | 07            | 22            | 09                 | 05            |
| *Citrobacter spp.* (02)  | 00            | 00            | 02                 | 00            |
| *E. coli* (02)           | 00            | 01            | 00                 | 01            |
| **Total**                | **41**        | **89**        | **43**             | **30**        |

MSSA: Methicillin susceptible *Staphylococcus aureus*; MSCoNS: Methicillin susceptible coagulase negative *Staphylococcus*; MRSA: Methicillin resistant *Staphylococcus aureus*; MRCoNS: Methicillin resistant coagulase negative staphylococcus; GPB: Gram positive spore bearer

Table 5. Number and type of bacterial agent isolated from mobile phones of Non-HCWs

| Source type               | Number of samples collected | Number of culture positive samples | Isolated organisms | Number of isolated organisms |
|---------------------------|-----------------------------|-----------------------------------|--------------------|-----------------------------|
| Non Health care workers   | 50                          | 23                                | **Klebsiella pneumoniae** | 03                          |
|                           |                             |                                   | **Acinetobacter baumannii** | 04                          |
|                           |                             |                                   | **Citrobacter spp.** | 02                          |
| **Total**                 | **28**                      |                                   |                    |                      |

MSSA: Methicillin susceptible *Staphylococcus aureus*; MSCoNS: Methicillin susceptible coagulase negative *Staphylococcus*; GPB: Gram positive spore bearer
Table 6. Antibiotic susceptibility pattern of Gram-positive organisms isolated from HCWs

| Antibiotics | S. aureus (n=65) | CoNS (n=21) |
|-------------|-----------------|-------------|
|             | S   | R   | S   | R   | S   | R   | S   | R   |
| LZ          | 65  | 00  | 21  | 00  |     |     |     |     |
| E           | 28  | 37  | 12  | 09  |     |     |     |     |
| CD          | 48  | 17  | 15  | 06  |     |     |     |     |
| CIP         | 52  | 13  | 17  | 04  |     |     |     |     |
| COT         | 45  | 20  | 16  | 05  |     |     |     |     |
| CX          | 34  | 31  | 12  | 09  |     |     |     |     |
| TE          | 56  | 09  | 16  | 05  |     |     |     |     |

LZ: linezolid; E: erythromycin; CD: clindamycin; CIP: ciprofloxacin; COT: cotrimoxazole; CX: cefoxitin; TE: tetracycline; CoNS: Coagulase negative staphylococcus

Table 7. Antibiotic susceptibility pattern of Gram-negative organisms isolated from HCW’s

| Antibiotics | A. baumannii (n=43) | P. aeruginosa (n=39) | K. pneumoniae (n=10) | E. coli (n=02) | Citrobacter (n=02) |
|-------------|---------------------|----------------------|----------------------|----------------|---------------------|
|             | S   | R   | S   | R   | S   | R   | S   | R   | S   | R   |
| PIT         | 26  | 17  | 29  | 10  | 07  | 03  | 01  | 01  | 01  | 01  |
| CTR         | 16  | 27  | 15  | 24  | 08  | 02  | 01  | 01  | 02  | 00  |
| CPM         | 18  | 25  | 11  | 28  | 09  | 01  | 01  | 01  | 02  | 00  |
| IPM         | 31  | 12  | 38  | 01  | 10  | 00  | 02  | 00  | 02  | 00  |
| COT         | 29  | 14  | 28  | 11  | 08  | 02  | 01  | 01  | 02  | 00  |
| AK          | 42  | 01  | 37  | 02  | 10  | 00  | 02  | 00  | 02  | 00  |
| CIP         | 39  | 04  | 32  | 07  | 09  | 01  | 01  | 01  | 02  | 00  |
| A           | 09  | 34  |     |     | 01  | 09  | 00  | 02  | 00  | 02  |

PIT: pipericillin-tazobactam; CTR: ceftriaxone; CPM: cefepime; IPM: imipenem; COT: cotrimoxazole; AK: amikacin; CIP: ciprofloxacin; A: ampicillin

the three *Klebsiella pneumoniae* were susceptible to ceftriaxone, imipenem, amikacin, ciprofloxacin and two isolate were susceptible to pipericillin-tazobactam, cefepime, cotrimoxazole. *Citrobacter spp.* was susceptible to all the antibiotics tested.

**DISCUSSION**

The hospital environment plays a very important role in the transmission of micro-organisms causing HAIs. Micro-organisms can be transferred from person to person or from inanimate objects like stethoscopes, bronchoscopes, pens, computer keyboards, mobile phones and fixed telephones to hand and vice versa. In the present study, one such inanimate object mobile phone was studied for microbial colonization.

The mobile phones have become multi-purpose non-medical devices used in the healthcare facility and in the community. It has increasingly become an important means of communication in the community and in the healthcare facility for collecting epidemiological data and monitoring chronic diseases. Mobile phones are used without restriction in healthcare facilities, including specific, susceptible areas like the operation room and ICUs, regardless of their unknown microbial load (18). In a study, it was discovered that cell phones usually are dirtier than either a toilet seat or the bottom of shoe (4). The constant handling of mobile phones by different users exposes to an array of micro-organisms and thus makes a good carrier for microbes. This is especially so with skin, due to the moisture and optimum temperature of human body especially our palms along with heat generated by mobile phones favors the colonization and multiplication of micro-organisms, so these devices can harbour various potential pathogens and serves as an exogenous source of nosocomial infection among hospitalized patients (6).

In present study, 92.80% of HCWs mobile phones and 57.50% of non-HCWs mobile phones showed microbial growth. Carriage rates of bacterial isolates on cell phones reported by various authors are shown in the Table 8.

Rate of contamination of mobile phones of HCWs...
in present study coincides with studies performed by Marwa et al. (21), Jaya Lakshmi et al. (20), Neha Sharma et al. (22) and Ulger et al. (19). Rate of contamination of mobile phones of non-HCWs reported by Misgana et al. (18) was consistent with the present study. In contrast to the present study, Neha Sharma et al. (22) has reported 80% of contamination of mobile phones among non-HCWs. Arora et al. (15) has reported (41%), lower bacterial contamination of mobile phones. The difference in the contamination rate may be due to the variation of the study participants in adherence to infection prevention, the pattern of mobile phone use, mobile phone keeping habits and personal behavior (18). So these finding definitely indicate that HCWs mobile phones were heavily contaminated compared to non-HCWs mobile phones. The reasons for getting a larger number of isolates from HCWs mobile phones may be a consequence of HCWs having direct contact with patients. Non-compliance of hospital standards for infection prevention may also contribute to the finding of high bacterial contamination.

Bacteria known to cause HAIs have varied by clinical settings and have included MRSA, A. baumannii, and Pseudomonas species (23, 24). Out of 203 bacteria isolated in this study, Staphylococcal species (44.33%) were predominant bacteria, grown from HCWs mobile phones. Similar pattern have been observed and reported by Lawani et al. (25). Staphylococcal species especially S. epidermidis normally found on skin flora, this might be the reason for their high rate of growth from the mobile phones in the present study. S. aureus can cause various illnesses, from minor skin infections to much more serious diseases, which include pneumonia, bacteremia, septicemia etc. MRSA is of particular importance in the medical community, as it has evolved resistance to β-lactam antibiotics (26). Even in the present study, predominant organism isolated from HCW’s in Operation Theater was MRSA (46.66%) and in dialysis unit MRSA (21.73%), followed by MSSA (21.73%).

The second common bacteria isolated from HCWs mobile phones was A. baumannii. It is a Gram-negative cocco-bacilli, which are characterized by their truncated rod shape. The organism is ubiquitous, which can be found in the normal skin flora, as well as in soil and bodies of water, amongst others. Multi-ple drug-resistant strains of A. baumannii (MDR) has been arisen, which combined with its ability to persist in hospital environments for extended periods of time, has led to its emergence as a potentially dangerous nosocomial pathogen (27). A. baumannii (36.84%) was the predominant organism isolated from mobile phones of HCW’s working in ICUs and from nursing (24.71%) professionals in this study.

One of the alarming signs is that multi-drug resistant organisms like MRSA and A. baumannii are isolated from HCWs in critical areas like ICUs, Operation Theater, dialysis units and from doctors and nursing professionals. This could be the reason for high rate of isolation of A. baumannii from ventilator-associated pneumonia (VAP) patients in Respiratory Intensive Care Unit (RICUs) and Staphylococcus species from post operative wards in our tertiary care hospital.

In the present study, staphylococci isolated were 100% susceptible to linezolid and 80% were susceptible to ciprofloxacin. Similar findings were also noted by Dardi (28).

A. baumanii isolates in our study were susceptible to amikacin (97.67%) and ciprofloxacin (90.69%). P. aeruginosa was susceptible to imipenem (97.43%) and amikacin (94.87%). K. pneumoniae were susceptible to imipenem (100%), amikacin (100%) and ciprofloxacin (90%). In study performed by Dardi (29), Gram-negative bacilli isolated from mobile phones were 100% susceptible to amikacin, netilmicin, meropenem, cefazidime, ticarcillin, pipercillin and cefepime.

Out of 50 non-HCWs mobile phone samples exam-

Table 8. Comparison of carriage rate between studies

| Study                        | Percentage of organisms isolated |
|------------------------------|---------------------------------|
|                              | Health care workers | Non-health care workers |
| Present study                | 92.80%              | 57.50 %                  |
| Misgana et al. (19)          | 86.37%              | 56.06 %                  |
| Ulger et al. (20)            | 94.05%              | -                        |
| Jaya Lakshmi et al. (21)     | 91.60%              | -                        |
| Marwa et al. (22)            | 92.50%              | -                        |
| Neha Sharma et al. (23)      | 94%                 | 80 %                     |
ined in this study, 23 (46%) yielded growth. Of these grown isolates, Gram-positive spore bearers (GPSB) were the predominant organisms (n=16, 69.56%). GPSB are non-pathogenic to human beings, may be present on cell phones as contaminants. By excluding growth of GPB, 7 (17.5%) samples yielded the growth of human pathogens. In contrast to the present study, Misgana et al. reported 56.06% (37/66) of growth on cell phones as contaminants. By excluding GPSB are non-pathogenic to human beings, may be the predominant organisms (n=16, 69.56%).

Several studies also revealed that HCWs do not consider mobile phones to be contaminated items and rarely disinfect their phones (2). Hand washing is the most effective method for the prevention of bacterial transmission. Although there are strict rules on hand hygiene in hospitals, it is not possible to provide de-contamination, disinfection or sterilization of each device used personally. Even though the presence of some items can be restricted in the hospital setting, it is not possible to limit the use of mobile phones by HCWs due to their indispensable benefits. The Centers for Disease Control and Prevention (CDC)’s guidelines for environmental infection control in healthcare facilities recommends periodic disinfection after cleaning instruments and surfaces that often come into contact with the hands, such as computer keyboards and mouse, as defined by the infection control committee (29).

CONCLUSION

Our study reveals that there is definite colonization of bacteria on the mobile phones, which are very close to the hand of HCWs. Mobile phones are not only capable of transferring messages but also are disease-producing microbes. They may act as a suitable substrate from which the disease may arise, spread and cause havoc, in the form of nosocomial disease. Our study also reveals that colonization of bacteria on the mobile phones of Non-HCW’s is less, compared to HCW’s. These contaminated phones can play a potential role in the spread of drug-resistant bacteria into the community. There should be regulations around the use of mobile telephones in hospital settings due to their potential to contribute to nosocomial infections. Mobile phones of HCWs could be a friend or foe, depending on how it is used during working hours in the hospital.

REFERENCES

1. Soto RG, Chu LF, Goldman JM, Rampil JJ, Ruskin KJ. Communication in critical care environments: mobile telephones improve patient care. Anesth Analg 2006; 102: 535-541.
2. Ramesh J, Carter AO, Campbell MH, Gibbons N, Powlett C, Moseley H, et al. Use of mobile phones by medical staff at Queen Elizabeth Hospital, Barbados: evidence for both benefit and harm. J Hosp Infect 2008; 70: 160-165.
3. Manning ML, Davis J, Sparnon E, Ballard RM. iPads, droids, and bugs: Infection prevention for mobile hand-held devices at the point of care. Am J Infect Control 2013; 41: 1073-1076.
4. Harish RT, Kairavi JD, Lopa PT, Saklainhaider SM, Tanuja BJ. Role of mobile phone in spreading hospital acquired infection: A Study in different group of health care workers. Natl J Integr Res Med 2011; 2: 61-66.
5. Chinjal AP, Mitesh N.K, Sanjay J.M. Bacteriological profile of cell phones of healthcare workers at tertiary care hospital. JEMDS 2012; 1: 198-202.
6. Jaya Madhuri R, Saraswathi M, Mahitha G, Bhargavi M, Deepika S, Vijaya Lakshmi G. Bacterial contamination of mobile phones and computers in microbiological laboratories. European J Biotechnol Biosci 2015; 3: 51-55.
7. Visvanathan A, Gibb AP, Brady RR. Increasing clinical presence of mobile communication technology: avoiding the pitfalls. Telemed J E Health 2011; 17: 656-661.
8. Vilella A, Bayas JM, Diaz MT, Guinovart C, Diez C, Simó D, et al. The role of mobile phones in improving vaccination rates in travelers. Prev Med 2004; 38: 503-509.
9. Akinyemi KO, Atapu AD, Adetona OO, Coker AO. The potential role of mobile phones in the spread of bacterial infections. J Infect Dev Ctries 2009; 3: 628-632.
10. Ilusanya O, Adesanya O, Adesemowo A, Amushan N. Personal hygiene and microbial contamination of mobile phones of food vendors in Ago-Iwoye Town, Ogun State, Nigeria. Pak J Nutr 2012; 11: 276-278.
11. Philip AM. The normal microbial flora. N Engl J Med 1982; 307: 83-93.
12. Elkholy M, Ewees I. Mobile (cellular) phone contamination with nosocomial pathogens in Intensive Care Units. Med J Cairo Univ 2010; 2: 1-5.
13. Aronson SH. The Lancet on the telephone 1876-1975. Med Hist 1977; 21: 69-87.
14. Kapil A (Ed). Ananthanarayanan and Paniker Text Book of Microbiology (2013). Healthcare- associated infections. 9th edition. Universities Press, pp. 644-649.
15. Arora U, Devi P, Chadha A, Malhotra S. Cell phones a modern stay house for bacterial pathogens. J K Science 2009; 11: 127-129.
16. Chawla K, Mukhopadyay C, Gurung B, Bhate P, Bairy I. Bacterial “Cell” Phones: Do cell phones carry potential pathogens? OJHAS 2009; 8(1): 8. http://www.ojhas.org/issue29/2009-1-8.htm

17. Performance standards for antimicrobial susceptibility testing, “Twenty third information Supplement. 2013. M100-S23. p.66.

18. Girma MM, Ketema A, Gemeda A. Bacterial contamination of mobile phones of healthcare workers at Jimma University Specialized Hospital, Jimma, South West Ethiopia. Int J Infect Control 2014; 11(1): 1-8.

19. Ulger F, Esen S, Dilek A. Are we aware how contaminated our mobile phones with nosocomial pathogens? Ann Clin Microbiol Antimicrob 2009; 8:7.

20. Jayalakshmi J, Appalaraju B, Usha. Cell phones as reservoirs of nosocomial pathogens. J Assoc Phy India 2008; 56: 388-389.

21. Marwa AE, Nadia ME. Mobile phones are silent threat. Int J Curr Microbiol App Sci 2015; 4: 199-205.

22. Neha S, Aruna S, Parihar RS, Khatri PK, Arvind C, Archana B. Prevalence and antibiotic pattern of microbes isolated from mobile phones of health care workers and non-health care workers. Int Curr Microbiol App Sci 2014; 3: 43-60.

23. Khivsara A, Sushma TV, Dhanashree B. Typing of Staphylococcus aureus from mobile phones and clinical samples. Current Science 2006; 90: 910-912.

24. Goldblatt JG, Krief I, Klonsky T, Haller D, Milloul V, Sisahsmith DM, et al. Use of cellular telephones and transmission of pathogens by medical staff in New York and Israel. Infect Control Hosp Epidemiol 2007; 28: 500-503.

25. Lawani EU, Oxford IO. Mobile phones of Healthcare workers: Friend or Foe. GJRA 2015; 4(3).

26. Holmes JW, Williams MD. Methicillin-resistant Staphylococcus aureus screening and eradication in the surgical intensive care unit: Is it worth it? Am J Surg 2010; 200:827-831.

27. Peleg AY, Seifert H, Paterson DL. Acinetobacter baumanii: emergence of a successful pathogen. Clin Microbiol Rev 2008; 21:538-582.

28. Dardi CKB, Jaisreee SP. Study of micro-organisms and their antimicrobial susceptibility pattern on mobile phones of health care workers from a tertiary care rural hospital. Adv Biomed Pharma 2015; 2: 267-273.

29. Boyce JM, Pittet D. Guideline for hand hygiene in health-care settings. MMWR Recomm Rep 2002; 51 (RR-16):1-45.