Volume Exclusion and H-Bonding Dominate the Thermodynamics and Solvation of Trimethylamine-N-oxide in Aqueous Urea

Jörg Rösgen* and Ruby Jackson-Atogi

Pennsylvania State University College of Medicine, Hershey, Pennsylvania 17033, United States

ABSTRACT: Trimethylamine-N-oxide (TMAO) and urea represent the extremes among the naturally occurring organic osmolytes in terms of their ability to stabilize/destabilize proteins. Their mixtures are found in nature and have generated interest in terms of both their physiological role and their potential use as additives in various applications (crystallography, drug formulation, etc.). Here we report experimental density and activity coefficient data for aqueous mixtures of TMAO with urea. From these data we derive the thermodynamics and solvation properties of the osmolytes, using Kirkwood−Buff theory. Strong hydrogen-bonding at the TMAO oxygen, combined with volume exclusion, accounts for the thermodynamics and solvation of TMAO in aqueous urea. As a result, TMAO behaves in a manner that is surprisingly similar to that of hard-spheres. There are two mandatory solvation sites. In plain water, these sites are occupied with water molecules, which are seamlessly replaced by urea, in proportion to its volume fraction. We discuss how this result gives an explanation both for the exceptionally strong exclusion of TMAO from peptide groups and for the experimentally observed synergy between urea and TMAO.

INTRODUCTION

Virtually all organisms use organic osmolytes to counter biochemical stress.1 The denaturant urea is among those osmolytes and can occur as a stressor, e.g., in the kidney,2 but it is also used against osmotic stress.1 In either case, urea is found in mixtures with at least one other osmolyte, typically a methylamine, such as trimethylamine-N-oxide (TMAO).1 This compound has generated considerable research interest, not only because it is the strongest known protein stabilizer among the natural osmolytes3 and a crystallizing agent;4 it also has been found to correct medicinally significant issues, such as prion aggregation5 and cellular folding defects.6

It has remained somewhat elusive what the mechanism of TMAO’s action on protein is, and how it counters the denaturing effect of urea. The range of opinions is broad and includes suggestions that TMAO stabilizes proteins by classical preferential exclusion7−14 or by altering the structure of water15−17 and that TMAO counteracts urea by directly interacting with urea,18,19 by urea-independent preferential exclusion from the protein,7,20−22 or by reverting changes of the structure of water caused by urea.15 Since many of these ideas have to do with the bulk solution, it is expedient to focus on ternary mixtures of TMAO with urea in water.

Here we report thermodynamic experiments that are combined with rigorous statistical mechanics23 to derive structural properties of the solution.24 This so-called Kirkwood−Buff approach provides a link between thermodynamics and preferential interactions on the one side and the structure of the solution on the other.11,12,25−27 The basic idea of this approach is that deviations from random distribution of molecules around each other produce characteristic changes in the partial molar volumes and chemical activities of the solution components,23 and that conversely knowledge of these thermodynamic properties allows us to derive information about the proximity of molecules in solution.24 The so-called Kirkwood−Buff integrals (integrated pair correlation functions) are a measure of the excess or deficit of one type of molecule around another.23

Using this Kirkwood−Buff approach, we find that, in aqueous urea, TMAO behaves as if it were a hard-sphere gas and the effective size of the hard-sphere depends on its occupancy with either water or urea molecules at two mandatory solvation sites. This result gives a rationale for the strong exclusion of TMAO from peptide groups9,28 which is substantially enhanced by the water/urea spacers intercalated between TMAO and the peptide group. It also explains the experimentally observed (though small)20 synergy between urea and TMAO, which is caused by the increased exclusion of TMAO from the peptide group, as small spacers (water) are replaced by larger ones (urea).

RESULTS

The primary goal here is to derive the solvation behavior of aqueous mixtures of urea and TMAO, which requires partial molar volumes and chemical activities of the solution components (see eqs 16 and 17).

Volumes. The partial molar volumes are shown in Figure 1, where the horizontal lines represent the limiting values in plain
water. The partial molar volumes of both urea (top panel) and TMAO (bottom panel) depend very little on the concentration of the respective other osmolyte within the range of concentrations used in the vapor pressure experiments (3–4 M).

The partial molar volume of water is virtually unchanged (not shown). Our data on urea compare favorably with previous data reported by Lee and Chalikian30 (points in the top panel of Figure 1). The partial molar volume of TMAO comes very close to the data reported by Di Michele et al.30 (points in the bottom panel of Figure 1). However, there is a slight offset. This could be due to deviations between the assumed and actual concentrations of TMAO. Di Michele et al. used a very hygroscopic31 preparation of anhydrous TMAO without further treatment, and their data become identical with ours if we assume that their TMAO contained 3–4% water.

![Figure 1](image)

**Figure 1.** Partial molar volumes of urea and TMAO in their ternary mixtures with water. Straight horizontal lines represent limiting partial molar volumes in pure water. Top panel: urea partial molar volume (thick lines) in 0 m, 2 m, 4 m, and 6 m TMAO (bottom to top). Propagated fitting errors are given by thin lines, and points represent previous data.29 Bottom panel: Same for TMAO in 0–15 m urea (3 m steps from bottom to top). Points are literature data.30 The parameters from a fit of the density data to eq 7 are given in Table 1.

**Table 1. Density Fitting Results to Eq 7 in mL/mol**

| Parameter | Value (mL/mol) |
|-----------|----------------|
| \( \bar{v}_{1U} \) | 44.2 |
| \( \bar{v}_{1T} \) | 73.4 |
| \( \bar{v}_{2U}/(10^{-3}/m) \) | 276 |
| \( \bar{v}_{2T}/(1/m) \) | -1.1 |
| \( \bar{v}_{3U}/(10^{-3}/m^2) \) | -48 |
| \( \bar{v}_{3T}/(1/m^2) \) | 0.11 |
| \( \bar{v}_{4U}/(10^{-3}/m^3) \) | 7 |
| \( \bar{v}_{4T}/(10^{-3}/m^3) \) | 7 |
| \( \bar{v}_{5U}/(10^{-3}/m^4) \) | -0.5 |
| \( \bar{v}_{5T}/(10^{-3}/m^4) \) | 0 |
| \( \bar{v}_{6U}/(10^{-3}/m^5) \) | -41 |
| \( \bar{v}_{6T}/(10^{-3}/m^5) \) | -8 |
| \( \bar{v}_{7U}/(10^{-3}/m^6) \) | 77 |
| \( \bar{v}_{7T}/(10^{-3}/m^6) \) | -8 |
| \( \bar{v}_{8U}/(10^{-3}/m^7) \) | 4 |
| \( \bar{v}_{8T}/(10^{-3}/m^7) \) | 0.5 |

**Figure 2.** Osmotic coefficients and activity coefficients in aqueous mixtures of urea with TMAO. (Top panel) Osmotic coefficients. Lines are the fit: blue, addition of urea to water (solid) or 1 m TMAO (dashed); red, addition of TMAO to water (solid) or 1 m urea (dashed); magenta, urea/TMAO = 2/1. Data: thin open circles, urea in water (this work); thick open circles, urea in water;29 open squares, urea in 1 m TMAO; filled circles, TMAO in water; filled squares, TMAO in 1 m urea; triangles, urea/TMAO = 2/1. Error bars show the standard deviation of three to nine measurements. (Bottom panel) Molar activity coefficients as a function of the TMAO concentration: red lines, \( \gamma_c \) of TMAO; blue lines, \( \gamma_c \) of urea. The successively shorter lines that are offset relative to unity correspond to 0–3 M urea, spaced by 0.5 M increments.

**Osmotic Coefficient and Activity Coefficients.** The osmotic coefficient data are shown in Figure 2 along with the fit (eq 12). Our urea data (thin open circles) compare well with previous data (thick open circles).32 The osmotic coefficient of TMAO (filled circles) is symptomatic for molecules that have repulsion between each other.33 We decided to sample the osmotic coefficient of a 2:1 mixture of urea:TMAO (triangles), because this is a ratio found in living organisms.34,35 At this ratio, the osmotic coefficient remains close to unity; i.e., the water behaves close to ideally in the molality scale.

The molar activity coefficients can be described as previously by33

\[
\gamma_i(c_{1U}, c_T) = \gamma_0 \frac{2c}{2c - c_i} \left(1 + \frac{4c(2 - c_i)}{8c_i(1 - c_i)}\right)
\]

(1)

where \( i \) stands for either urea (U) or TMAO (T). The parameters \( c_i \) and \( c_i \) are the inverse effective volumes occupied by a single osmolyte and a pair of osmolyte molecules, and \( c_i \) is an interaction parameter. The parameter \( \gamma_0 \) accounts for the effect of the second osmolyte on the infinite dilution activity coefficient of the first one. All parameters depend on the other osmolyte’s concentration, e.g.,

\[
c_{1,U} = c_{1,U,0} + c_{1,U,1}T + c_{1,U,2}T^2/2
\]

(2)

The parameters are given in Table 2.

The resulting molar activity coefficients (Figure 2, bottom panel) depend primarily on the molarity of TMAO, and to a much lesser degree on the molarity of urea. For example, the change in \( \gamma_c \) (blue lines) as a function of TMAO (going from left to right) is several fold larger than as a function of urea.
going from the uppermost curve to the lowest). For this reason, $\gamma_c$ is plotted as a function of the TMAO concentration. Both activity coefficients increase upon addition of the respective other osmolyte, suggesting mild mutual exclusion.33 This effect is, however, small compared to the strong increase of the activity coefficient of TMAO with its own concentration (see bottom panel of Figure 2).

Solvation. From the combined volumetric and osmometric data we derived the Kirkwood–Buff integrals (KBIs) of all components in the ternary mixture, using eq 16. Remember that the KBIs are a measure of the excess or deficit of one type of molecule around another type. The results are shown in Figure 3. The first observation is that again, as in the case of the activity coefficients (Figure 2), all concentration dependencies are almost exclusively due to TMAO. That is, the change along the TMAO axis is generally larger than the urea-dependent offset within each group of curves. Only the self-solvation KBIs of the osmolytes $\mathcal{G}_{UU}$ and $\mathcal{G}_{TT}$ depend slightly on the concentration of urea. Thus the cluster of lines corresponding to increasing urea concentration becomes visible in these two cases. The urea concentration increases from the highest line (0 m) to the lowest one (3.6 m) in both cases. The overall level of the TMAO self-solvation, $\mathcal{G}_{TT}$, is at least 3 times more negative than expected for an osmolyte of its size. The magnitude of $\mathcal{G}_{TT}$ is in the order of that of disaccharides.37 But as expected,37 the hydration terms $\mathcal{G}_{WT}$, $\mathcal{G}_{WU}$ and $\mathcal{G}_{WW}$ are small in comparison.

Hard-Sphere-like Behavior of TMAO. Figure 4 demonstrates that TMAO closely follows a solvation pattern expected for a hard-sphere gas, i.e., a system in which the one and only interaction between molecules is hard-core repulsion (molecules cannot overlap each other). At low concentrations, such spheres occupy the space randomly. As the concentration increases, their proximity is determined by the question of how well they can be packed. The lower panel shows $\mathcal{G}_{TT}$ at 0 M urea (continuous red line) and in the limit of 0 M TMAO (dashed red line). The latter is shown as a function of the urea molarity, and the dash-dotted line is calculated according to eq 5. The deviations between the best prediction and the data are smaller than the uncertainty in the data ($\sim 10$ mL/mol).

Table 2. Parameters for the Second-Order Polynomials for Each Parameter in Eq 1, for Either Urea (U) or TMAO (T): Given Are the Zeroth-, First-, and Second-Order Coefficients for a Second-Order Polynomial of the Type Given by Eq 2

| Parameter | Zeroth | First | Second |
|-----------|--------|-------|--------|
| $\mathcal{G}_{UU}$ | 1.453 M | −0.040 | 0 |
| $\mathcal{G}_{WT}$ | 2.066 M | −0.078 | 0 |
| $\mathcal{G}_{WU}$ | 8.311 M | 0.022 | 0.030/M |
| $\mathcal{G}_{TT}$ | 1 | 0.074 | 0 |
| $\mathcal{G}_{UU}$ | 19.84 M | −6.1 | 1.32/M |
| $\mathcal{G}_{WT}$ | 11.02 M | 0 | 0 |
| $\mathcal{G}_{WU}$ | 21.14 M | −3.94 | 0 |
| $\mathcal{G}_{TU}$ | 1 | 0.11 | 0 |
a hydrated hard-sphere that has the size of a doubly hydrated TMAO molecule (eq 24). The same behavior is found with the water self-hydration, $G_{WW}$ (Figure 4, top panel). There is no straightforward way to calculate the other, urea-related KBIs from the hard-sphere model.

**DISCUSSION**

Much has been written about the interactions among TMAO, urea, and water in solution, and the main focus has been on the water interactions. We start by first considering the hydration of TMAO, and then examine the other (previously more neglected) binary interactions in solution, and close with a discussion of water structure.

**Strong, Two-fold Hydration of TMAO.** Our finding that aqueous TMAO behaves similarly to a hard-sphere of the size of TMAO dihydrate strongly suggests that TMAO has two mandatory hydration waters. This is supported by various other lines of research. Anhydrous TMAO cannot be produced by evaporating the two hydration waters; rather, TMAO has to be sublimed at high temperature.\(^{31}\) *Ab initio* calculations also show tightly coordinated water.\(^{39}\) Dielectric relaxation finds TMAO as a dihydrate in solution, and in the same research, the very large enthalpy of H-bond formation was pointed out.\(^{37}\) Based on NMR it was found that all TMAO in water is present as large ensembles found that both water oxygen and urea nitrogen are found around the TMAO oxygen at hydrogen-bonded distances, with one hydrogen atom between.\(^{19,43}\) Thus, both urea and water snap into hydrogen-bonded positions as they approach the TMAO oxygen.

**The Two Mandatory Ligands of TMAO Can Be Water Molecules, Urea Molecules, or Both.** It has been found both by Raman spectroscopy\(^ {30}\) and neutron/X-ray scattering\(^ {43}\) that TMAO does not self-aggregate. We can add now the significant finding to which degree TMAO does not aggregate; viz., its self-solvation $G_{TT}$ is indistinguishable from that of a hard-sphere gas—there is no significant attraction. According to the effective size of the hard-sphere, the TMAO must be doubly hydrated, as discussed above. Upon addition of urea, these waters are progressively replaced by urea molecules, and this should lead to an increase of the effective hard-sphere size. Indeed, $G_{TT}$ changes upon urea addition toward more negative values, which correspond to larger sphere diameters (Figures 3 and 4), as discussed in the following around eq 5.

**Water and Urea Are Essentially Equivalent Ligands for TMAO.** Is there actual (weak affinity) binding of urea to TMAO? Meersman et al. argued that urea binds to TMAO with an affinity that is comparable to the interaction of urea with peptide groups.\(^ {35}\) However, they also pointed out that their analysis was assuming a binding model, rather than the proper exchange model.\(^ {43}\) Therefore we reanalyze their data, as shown in Figure 5 as continuous lines. We focus on the first two reported binding events, because a potential third “urea site” is barely populated.\(^ {43}\) The reaction model is

$$
\begin{align*}
\text{T-2W+2U} & \Rightarrow \text{T-W-U+W+U} \\
\text{T-W-U+W} & \Rightarrow \text{T-2U+2W}
\end{align*}
$$

where $T$, $U$, and $W$ represent TMAO, urea, and water, respectively, and $K_1$ and $K_2$ are the exchange constants for the two reactions. The populations for each species were calculated and fit to the data numerically. The resulting “affinities” are quite weak, so that the midpoint of the “binding” of the first urea is around 7.5 M urea. In comparison, the midpoint of the binding of urea to a peptide NH is around 4 M.\(^ {44}\) In the face of such weak affinities the question arises, how do the exchange curves in Figure 5 compare to random occupancy with water and urea? If we take the volume fraction as an approximate model for the likelihood that a site is randomly occupied by a species,\(^ {44}\) we first need to consider that direct TMAO–TMAO interaction at the oxygen site does not occur (see previous paragraph). So, we normalize the volume fractions of water and urea to refer to the non-TMAO volume $(1 - \phi_T)$, and obtain for the populations of the three species

$$
\begin{align*}
\phi_W^2 & = \phi_T^2 (1 - \phi_T) \\
2\phi_W\phi_U & = \phi_T^2 (1 - \phi_T) \\
\phi_U^2 & = \phi_T^2 (1 - \phi_T)
\end{align*}
$$

The resulting curves are shown as dashed lines in Figure 5. The population of the dihydrate, $P_{WW}$, closely follows the exchange curve, and thus double occupancy with water is practically random. For the other two species, there is a marginal deviation from random distribution, such that the single urea species is populated slightly less than random, and the double urea species slightly more. This view of nearly random interaction is also supported by recent molecular dynamics simulations, where TMAO was found to be equally H-bonded to water and urea at about 35% by volume of each urea and water,\(^ {22}\) and a rough equivalence of both was noted.\(^ {40}\) The idea of random interaction of water and urea with TMAO is also consistent with the spectroscopic finding that there is no specific urea–TMAO binding.\(^ {38}\) A computational finding that both urea and water interact strongly with TMAO should then be taken to mean that though both are strongly interacting with TMAO...
measurements Meersman et al. derived the radial distribution functions of TMAO (2.66 Å). The resulting curve is shown in red in Figure 6.

The Urea–TMAO Interaction $G_{UT}$ Is Dominated by Steric Exclusion Effects. From neutron and X-ray diffraction measurements Meersman et al. derived the radial distribution function between a urea nitrogen and the TMAO oxygen as shown in Figure 6 (blue line). Although there is a clear first contribution (red) reveals more clearly the second solvation peak around 5.5 Å, which may come from both the second urea nitrogen and water-separated urea–TMAO pairs. Beyond this peak the radial distribution function levels off. Significantly, the blue scattering curve integrates within error to the same value as the red curve, viz., the measured $G_{UT} \approx 90 \text{ mL/mol at } \sim 2 \text{ M TMAO.}$ This result highlights the dominance of steric exclusion, where the H-bonding related deviations of the actual $g(r)$ from eq 6 merely have an alignment effect that averages out to zero.

Hydration of Urea and TMAO Is Unchanged upon Mixing. Previous findings support our observation that the hydration of the osmolytes, $G_{TW}$ and $G_{UW}$ hardly depends on osmolyte concentration (except for the nonspecific hard-sphere-like behavior of $G_{TW}$ as a function of $c_{TMAO}$). Near-infrared spectra do not show any evidence that the hydration of either urea or TMAO changes upon mixing. Also NMR, combined with molecular dynamics, showed that the hydration shell structure of TMAO is maintained over a wide concentration range.

Water Structure. There are two schools of thought on the origin of the behavior of osmolytes, viz. either direct interaction of water/osmolyte with target molecules or indirect effects through alterations of the structure of water. Typical uses of the term “water structure” refer to just one small aspect of the properties of water. Thus it is not surprising that one can reach diametrically opposite conclusions regarding the effect of osmolytes merely by choosing how the term “water structure” is used. It has been suggested that the term “water structure” may be even undefinable in view of such issues. It is therefore expedient to refer to the actual observations (e.g., H-bond angles) rather than to use a word with multiple disparate uses.

What is the appropriate definition of “water structure” when we want to understand the thermodynamics of the solution? Perhaps the most general definition is given by the KBIs, which provide a rigorous link between the positions of molecules in solution and thermodynamics. The KBIs are directly linked with preferential interactions and thermodynamics. The KBIs are directly linked with preferential interactions and thermodynamics.

The KBIs are directly linked with preferential interactions and thermodynamics.

![Figure 6. Radial distribution function of urea nitrogen around TMAO oxygen: blue line, data for about 2 M each of urea and TMAO; red line, effect of direct steric exclusion between urea and TMAO (eq 6).](image)
seems to be governed by mere packing, as judged by its hard-sphere like behavior.

**CONCLUSIONS**

We have used a combination of our current experimental volume and chemical activity data with Kirkwood–Buff theory to elucidate the behavior of TMAO in aqueous urea. A second look on scattering data provided a strong confirmation and some additional insight. The picture that emerges shows a dominant role of hydrogen bonding along with simple steric exclusion. TMAO is a roughly spherical molecule and behaves as such to a degree that its self-solvation and hydration resemble the properties of a hard-sphere gas. However, it has two mandatory solvation sites that can be occupied by water or urea, substantially enlarging the effective hard-sphere diameter. The hydrogen bonding at these sites is sufficiently strong to prevent both water and urea from approaching the oxygen with anything else other than their hydrogen atoms.

What implications does this have for osmolyte–protein interactions? It is well known that TMAO is strongly excluded from peptide groups. Such preferential exclusion (also termed “osmophobicity”) favors the native state because the denatured state exposes more surface, leading to more exclusion. Aqueous TMAO appears to interact with the peptide groups only through its hydration waters.38,40 Thus, the H-bonds donated to the TMAO from peptide groups must be significantly weaker than those donated from water and urea. The mechanism of stabilization by TMAO does not change as urea is added. Only the spacing between TMAO and peptide groups is increased as the TMAO solvation waters are replaced by urea. That is, TMAO should be more excluded from the peptide group through an intercalated water.

What is then expected upon addition of urea is that the situation does not fundamentally change. The only difference is that now TMAO has the option to interact with the peptide group through either an intercalated urea, or an intercalated water. The mechanism of stabilization by TMAO does not change as urea is added. Only the spacing between TMAO and the peptide groups is increased as the TMAO solvation waters are replaced by urea. Thus, the H-bonds donated to the TMAO from peptide groups must be significantly weaker than those donated from water and urea.

**MATERIALS AND METHODS**

**Densimetry.** TMAO dihydrate from Sigma and ultrapure urea from USB were dried at 60 °C for at least 20 h. The masses used to prepare the solutions were measured using a Mettler Toledo AT20 analytical balance equipped with an antistatic device. The densities of the solutions were measured in an Anton Paar DMA 5000M density meter, using automatic viscosity correction.

**Vapor Pressure Osmometry.** The TMAO dihydrate used for the osmotic coefficient measurements was freshly synthesized and recrystallized. The water vapor pressure was measured in a Wescor Vapro 5520 osmometer, with modifications described previously. The measured osmolality values were divided by the total molality of urea and TMAO to obtain the osmotic coefficient, which is a measure of the deviation of the water chemical activity from ideal behavior.

**Volume Data Evaluation.** The density of the urea TMAO mixtures was measured to obtain the partial molar volumes. Using Wolfram Mathematica, the data were fit to the relation

\[
\rho = \frac{1 + m_U M_U + m_T M_T}{\rho_0 + \sum_{i+k>0} m_i m_T \bar{v}_{i+k,1}/1!}/k!
\]

where the numerator is the mass of the solution per kg of water and the denominator the mass per kg of water (note that the molecular weights \( M_i \) have to be given in kg/mol). Among the parameters, \( \rho_0 \) is the density of plain water, \( \bar{v}_{i,0} \) and \( \bar{v}_{i,1} \) are the limiting partial molar volume in plain water (of urea and TMAO, respectively), and the \( \bar{v}_{i,k} \) terms in general are mixed derivatives of the volume with respect to the molalities of urea and TMAO, \( m_U \) and \( m_T \). The partial molar volumes of urea and TMAO are then

\[
\bar{v}_U = \sum_{i,k \geq 0} m_i m_T \bar{v}_{i+1,k,1}/1!/(k!)
\]

\[
\bar{v}_T = \sum_{i,k \geq 0} m_i m_T \bar{v}_{i+k,1}/1!/(k!)
\]

and the partial molar volume of water

\[
\bar{v}_W = \frac{1 - c_U \bar{v}_U - c_T \bar{v}_T}{c_W}
\]

directly follows, knowing the molar concentrations of urea and TMAO (\( c_U \) and \( c_T \)) and the molecular weights of all species (18.015, 60.06, and 75.11 g/mol), and considering

\[
c_W = \frac{\rho - c_U M_U - c_T M_T}{M_W}
\]

**Vapor Pressure Data Evaluation.** The vapor pressure data were used to derive the chemical activities of urea and TMAO through the method of Schönert. The osmotic coefficient data were fitted to

\[
\varphi = 1 + \frac{1}{m_U + m_T} \sum_{i,k=0,i+k>1} g_{i,k}(i + k - 1) m_i m_T^{k-1}
\]

and it was found that terms up to second order are sufficient. There are additional isopiestic data available on the dependence of \( \varphi \) on urea concentration in the absence of TMAO. Including these data requires terms up to seventh order in urea concentration. Equations for the chemical activities of water, urea, and TMAO follow directly:

\[
\ln a_W = -\frac{\varphi m_U + m_T}{m_W}
\]

\[
\ln a_U = \ln m_U + \sum_{i>1} g_{i,0} i m_U^{i-1} + \sum_{i,k>0} g_{i,k} i m_U^{i-1} m_T^{k-1}
\]

\[
\ln a_T = \ln m_T + \sum_{i>1} g_{0,i} i m_U^{i-1} + \sum_{i,k>0} g_{i,k} i m_U^{i-1} m_T^{k-1}
\]

**Kirkwood–Buff Integrals.** Solvation properties can be calculated from the partial molar volumes and chemical activities as follows. The excess or deficit of molecules of type \( i \) and \( j \) around each other (relative to the bulk) is given by integrated pair correlation functions, the so-called KBIs,

\[
G_{ij} = \frac{1}{c_i c_j} \frac{\langle A_{ij} \rangle}{|A|} - \frac{\delta_{ij}}{c_i}
\]
where the elements of matrix A are given by

$$A_{ij} = \frac{1}{c_W} \left( \frac{\partial \ln a_i}{\partial N_j} \right) + \frac{\bar{v}_i}{k_{RT}}$$

(17)

Here, $\kappa$ is the compression coefficient, $\delta_i$ the Kronecker delta, $T$ the absolute temperature, $R$ the gas constant, $l/l'$ the determinant of $A$, and $l/l'$ denotes a cofactor of the matrix. After eq 17 is substituted into eq 16, $k$ has little impact on $G_{WT}$. This is because $k$ is then not in an isolated position in the denominator any more, and is negligibly small for our purposes, so we just set it to its value in plain water. In order to link the derivatives in this equation back to the vapor pressure results, they are rewritten as either

$$\left( \frac{\partial \ln a_i}{\partial N_k} \right) = m_W \left( \frac{\partial \ln a_i}{\partial N_k} \right)$$

(18)

for all cases except when $i = k = W$, or

$$\left( \frac{\partial \ln a_W}{\partial N_W} \right) = m_W - \frac{\partial \ln a_W}{\partial N_W} - \frac{\partial \ln a_W}{m_W} \left( \frac{\partial \ln a_W}{\partial N_W} \right)$$

(19)

by use of the Gibbs--Duhem relation.

**Kirkwood–Buff Integrals for Hard-Spheres.** As shown in the Results section, TMAO behaves like a hard-sphere gas in urea solution. Calculating the KBIs for hard-spheres is conveniently done through the Carnahan–Starling equation. Though this is relation for a single-component hard-sphere gas, it is possible to infer the implicit properties of the water as follows. In a two-component solution (here, water and TMAO), the KBIs are

$$G_{WT} = k_{RT} - \frac{\bar{v}_T + \bar{v}_W}{a_{TT}}$$

(20)

and

$$G_{WW} = k_{RT} - \frac{\bar{v}_W + \bar{v}_W}{a_{WW}}$$

(21)

for the self-solvation of TMAO and self-hydration of water, and

$$G_{WT} = k_{RT} - \frac{\bar{v}_W}{a_{WT}} = k_{RT} - \frac{\bar{v}_W}{a_{WW}}$$

(22)

for the hydration of TMAO, where $a_i = (\partial \ln a_i/\partial \ln c_i)$ and $c_i = (\partial \ln a_i/\partial c_i)$. The molar activity coefficient of component $i$ is $\gamma_i$. From eq 22 we see that $a_{NW} = a_{TT}/\gamma_i$ holds. Taking into account $\gamma_W = (a_{WW} - 1)/c_W$ allows use of eqs 20 to 22 to obtain $G_{NW}$ and $G_{WT}$ from $G_{TT}$, if $a_{WT}$ and the partial molar volumes are known:

$$G_{TT} = k_{RT} - \frac{\bar{v}_T + (\bar{v}_T - 1)/c_T}{a_{TT}}$$

(23)

$$G_{WT} = k_{RT} - \frac{\bar{v}_T - \bar{v}_T}{\gamma_W}$$

(24)

$$G_{WW} = \frac{k_{RT}(\bar{v}_W - \bar{v}_T) - \bar{v}_W \bar{v}_W + \bar{v}_T \bar{v}_T - \bar{v}_T^2}{\gamma_W}$$

(25)

Multiplying the Carnahan–Starling equation for a hard-sphere gas by the concentration $c$ and performing a derivative with respect to $c$, gives $a_{SS}$ as follows:

$$a_{SS} = 1 + 2q + 2q - \bar{v}_W q \frac{\partial \bar{v}_W}{\partial c_S}$$

(26)

where $q = c_S/\bar{v}_W$ is the volume fraction of the hard-sphere, $c_S$ its molarity, and $\bar{v}_W$ its partial molar volume. Equation 26 can then be used in place of $a_{SS}$ to calculate the various KBIs (eqs 23–25). When the size of the hard-spheres is different from $\bar{v}_T$, we change $\bar{v}_T$ in eq 23 accordingly. That is, the partial molar volume of TMAO mono- or dihydrate is larger than that of TMAO itself. However, when it comes to the partial molar volume in the context of $G_{WT}$, there is no such thing as a mono- or dihydrate. This is because $G_{WT}$ considers the spacing between TMAO and the surrounding water molecules independently of whether or not they are mandatory hydration waters. Accordingly, we do not change $\bar{v}_T$ in eq 24 when the size of the hard-spheres is assumed to be different from $\bar{v}_T$.

Note that in the limit of $c_S \to 0$, we recover the classically expected result for the excluded volume (equaling the second virial coefficient), $G_{WT} - G_{TT} = 4\bar{v}_T$.

**AUTHOR INFORMATION**

Corresponding Author

Jorg.Rosgen@psu.edu

Notes

The authors declare no competing financial interest.

**ACKNOWLEDGMENTS**

Support by NIH to J.R. through R01 GM049760 is gratefully acknowledged.

**REFERENCES**

(1) Yancey, P.; Clark, M.; Hand, S.; Bowls, R.; Somero, G. Science 1982, 217, 1214–1222.

(2) Nakashishi, T.; Balaban, R. S.; Burg, M. B. Am. J. Physiol. 1988, 255, C181–91.

(3) Auton, M.; Bolen, D. W. Proc. Natl. Acad. Sci. U.S.A. 2005, 102, 15065–15068.

(4) Doolittle, R. Biophys. Chem. 2003, 100, 307–313.

(5) Tatzelt, J.; Prusiner, S.; Welch, W. EMBO J. 1996, 15, 6363–6373.

(6) Brown, C.; Hong-Brown, L. Q.; Welch, W. J. Bioenerg. Biomembr. 1997, 29, 491–502.

(7) Lin, T. Y.; Timasheff, S. N. Biochemistry 1994, 33, 12695–12701.

(8) Courtenay, E.; Capp, M.; Anderson, C.; Record, M. J. Biochemistry 2000, 39, 4455–4471.

(9) Bolen, D.; Baskakov, I. J. Mol. Biol. 2001, 310, 955–963.

(10) Gallagher, K. R.; Sharp, K. A. J. Am. Chem. Soc. 2003, 125, 9853–9860.

(11) Abu, M.; Smith, P. E. J. Phys. Chem. B 2004, 108, 7382–7388.

(12) Shintu, S. Proc. Natl. Acad. Sci. U.S.A. 2004, 101, 1195–1199.

(13) Athawale, M. V.; Dorick, J. S.; Gardner, S. Biophys. J. 2005, 89, 858–866.

(14) Jiao, Y.; Smith, P. J. Chem. Phys. 2011, 135, 014502.

(15) Zou, Q.; Bennion, B.; Daggett, V.; Murphie, K. J. Am. Chem. Soc. 2002, 124, 1192–1202.

(16) Bennion, B.; Daggett, V. Proc. Natl. Acad. Sci. U.S.A. 2004, 101, 6433–6438.

(17) Reuz, Y.; Bakker, H. J. Phys. Chem. B 2009, 113, 4038–4044.

(18) Paul, S.; Patey, G. J. Am. Chem. Soc. 2007, 129, 4476–4482.

(19) Meier, B.; Bowron, D.; Soper, A.; Koch, M. Biophys. J. 2009, 97, 2559–2566.

(20) Kello, C.; Barrick, D. Protein Sci. 2003, 12, 1522–1529.

(21) Rostrom, L. M.; Bolen, D. W. Protein Science 2007, 16, 293–298.

(22) Kubokwa, H.; Hu, C.; Pettitt, B. J. Am. Chem. Soc. 2011, 133, 1849–1858.

(23) Kirkwood, J. G.; Buff, F. P. J. Chem. Phys. 1951, 19, 774–777.

(24) Ben-Naim, A. J. Chem. Phys. 1977, 67, 4884–4890.

(25) Ben-Naim, A. Cell Biophys. 1988, 12, 255–269.

(26) Chitra, R.; Smith, P. J. Phys. Chem. B 2002, 106, 1491–1500.

(27) Rössig, J.; Pettitt, B. M.; Bolen, D. W. Biophys. J. 2005, 89, 2988–2997.

(28) Auton, M.; Bolen, D.; Rössig, J. Proteins 2008, 73, 802–813.

(29) Lee, S.; Chalikian, T. J. Phys. Chem. B 2009, 113, 2443–2450.

(30) Di Michele, A.; Freda, M.; Onori, G.; Paolantoni, M.; Santucci, A.; Sassi, P. J. Phys. Chem. B 2006, 110, 21077–21085.
(31) Meisenheimer, J. Liebigs Ann. Chem. 1913, 297, 273−300.
(32) Scatchard, G.; Hamer, W. J.; Wood, S. E. J. Am. Chem. Soc. 1938, 60, 3061−3070.
(33) Rösgen, J.; Pettitt, B.; Bolen, D. Biochemistry 2004, 43, 14472−14484.
(34) Yancey, P.; Somero, G. Biochem. J. 1979, 183, 317−323.
(35) Pang, P.; Griffith, R.; Atz, J. Am. Zool. 1977, 17, 365−377.
(36) Rösgen, J.; Pettitt, B. M.; Bolen, D. W. Protein Sci. 2007, 16, 733−743.
(37) Shikata, T.; Itatani, S. J. Solution Chem. 2002, 31, 823−844.
(38) Hovagimyan, K. G.; Gerig, J. T. J. Phys. Chem. B 2005, 109, 24142−24151.
(39) Noto, R.; Martorana, V.; Emanuele, A.; Fornili, S. J. Chem. Soc., Faraday Trans. 1995, 91, 3803−3808.
(40) Hu, C.; Lynch, G.; Kokubo, H.; Pettitt, B. Proteins 2010, 78, 695−704.
(41) Munroe, K.; Magers, D.; Hammer, N. J. Phys. Chem. B 2011, 115, 7699−7707.
(42) Petersen, C.; Bakulin, A.; Pavel'ev, V.; Pshenichnikov, M.; Bakker, H. J. Chem. Phys. 2010, 133, 164514.
(43) Meersman, F.; Bowron, D.; Soper, A.; Koch, M. Phys. Chem. Chem. Phys. 2011, 13, 13765−13771.
(44) Lim, W.; Rösgen, J.; Englander, S. Proc. Natl. Acad. Sci. U.S.A. 2009, 106, 2595−2600.
(45) Wallqvist, A.; Covell, D. G.; Thirumalai, D. J. Am. Chem. Soc. 1998, 120, 427−428.
(46) Sharp, R.; Madan, B.; Manas, E.; Vanderkooi, J. J. Chem. Phys. 2001, 114, 1791−1796.
(47) Zhang, W.; Capp, M.; Bond, J.; Anderson, C.; Record, M. J. Phys. Chem. B 2003, 107, 5175−532.
(48) Rafflenbeul, L.; Rösgen, J. Methods Cell Biol. 2008, 84, 679−735.
(49) Rösgen, J.; Rafflenbeul, L.; Bolen, D. W. Methods Enzymol. 2011, 492C, 61−125.
(50) Matteoli, E.; Lepori, L. J. Chem. Phys. 1984, 80, 2856−2863.
(51) Lide, D. CRC Handbook of Chemistry and Physics; CRC Press: Boca Raton, FL, 2004.
(52) Carnahan, N. F.; Starling, K. E. J. Chem. Phys. 1969, 51, 635−636.
(53) Hansen, J.-P.; McDonald, I. R. Theory of Simple Liquids, 2nd ed.; Academic Press: London, 1990.