Chemotherapy Response Assay Test and Prognosis for Breast Cancer Patients Who Have Undergone Anthracycline- and Taxane-Based Chemotherapy

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INTRODUCTION

Anthracycline and taxane have been very essential drugs in the treatment of breast cancer patients in neoadjuvant and adjuvant settings. In particular, anthracycline has been very widely used in spite of its major limitation, cardiotoxicity [1-3]. Some studies have reported that an anthracycline-based regimen had more favorable results in HER2 overexpression tumors. Recently, it has been demonstrated that this result is mediated by topoisomerase IIα (TOP2A). Anthracycline binds to TOP2A and stabilizes DNA double strand breaks, resulting in cell cycle arrest and apoptosis. The Top2A gene is located close to the HER2 gene on chromosome 17, and TOP2A gene co-amplification has been found in about 40% of HER2-positive type breast cancers [4,5]. Nevertheless, anthracycline is still a fascinating chemotherapeutic regimen for HER2-positive type breast cancer patients. Anthracycline-based chemotherapy has been used as a good neoadjuvant and adjuvant treatment choice for both HER2-positive type and triple negative breast cancers (TNBC). It has been shown to produce higher pathological complete response (pCR) rates for TNBC in a neoadjuvant setting and better survival rates in an adjuvant setting [6-10].

Several studies have been conducted on the usefulness of the adenosine triphosphate-based chemotherapy response assay (ATP-CRA) in breast cancer [11-13]. This assay measures intracellular ATP, the basic energy source for living cells, which rapidly disappears when cells lose viability. The ATP-CRA has an advantage of a high success rate in primary culture and require only a small number of cells [12-16]. Thus, it is widely used as a chemosensitivity assay modality for the treatment of various
cancers in clinical settings. However, only a few studies have been performed concerning the relationship between chemotherapy response assay test results and breast cancer patient prognosis [17]. We performed this study to evaluate the ATP-CRA results and prognoses for breast cancer patients who had undergone anthracycline and taxane based chemotherapy.

METHODS

From August 2004 to December 2009, we performed ATP-CRA for 257 patients with breast cancer at the Department of Surgery, Ewha Womans University Mok-dong Hospital, Seoul, Republic of Korea. Among these patients, we enrolled patients who had undergone curative surgery and had received doxorubicin plus taxane or doxorubicin plus cyclophosphamide followed by taxane chemotherapy. A total of 102 patients were included in this study. The mean follow-up duration was 29.8 ± 15.6 months.

For ATP-CRA, two specimens were taken per patient by true-cut biopsy or core needle biopsy. We stored these tissues in Hank’s Balanced Salt Solution (HBSS; Gibco BRL, Rockville, USA) composed of 100 IU/mL of penicillin (Sigma, St. Louis, USA), 100 mg/mL of streptomycin (Sigma), 100 mg/mL of gentamicin (Gibco BRL), 2.5 mg/mL of amphotericin B (Gibco BRL), and 5% fetal bovine serum (Gibco BRL). The stored tumor tissues were sent to a commercial laboratory center for determination of their sensitivities to each chemotherapeutic regimen. The most commonly used chemotherapeutic regimens and their concentrations were as follows: 5-FU (10 µg/mL), paclitaxel (8.5 µg/mL), docetaxel (3.7 µg/mL), doxorubicin (1.5 µg/mL), methotrexate (0.37 µg/mL), and cyclophosphamide (4-OH-cyclophosphamide, 1.2 µg/mL). If sufficient cancer cells were available to determine their sensitivity, these cells were tested with three treated drug concentrations (TDC; 0.2×, 1×, 5×). If not, their ATP–CRA results were determined using only one TDC (1×). Doxorubicin and taxane sensitivities were determined using a cell death ratio cut-off of 40% and 24%, respectively, on the basis of TDC (1×). Receiver operating characteristic (ROC) curves were used to determine these cut-off values.

The doxorubicin sensitive group was compared to the resistant group with respect to their clinical characteristics (age, operation methods, chemotherapy type, and chemotherapeutic regimen), pathologic results, disease-free survival (DFS), and overall survival (OS). In respect to estrogen receptor (ER) status, we defined ER-positive when more than 10% of cancer cells were stained with more than 1+ of intensity. In the case of HER2-positive, it was defined as c-erb-B2+++ or when the ratio of HER2 loci/chromosome 17 centromeres was more than 2.2. Univariate and multivariate analysis were performed to identify whether ATP-CRA results could be an independent prognostic factor for DFS.

All data were analyzed using SPSS version 17.0 (SPSS Inc., Chicago, USA). Statistical significance was analyzed using the Student's t-test and Pearson’s chi-square test. DFS and OS were calculated using the Kaplan-Meier method and log-rank test. Cox’s proportional hazards regression model was used for univariate and multivariate analyses, and p-values < 0.05 were considered statistically significant.

RESULTS

Patient characteristics

During the study period, a total number of 257 ATP-CRAs were performed in our hospital and among these, 102 were included in this study. We divided these patients into two groups according to doxorubicin sensitivity. There were 19 patients in the sensitive group with a mean age of 49.0 ± 9.3 years (range, 29 to 69 years) and 83 patients in the resistant group with a mean age of 46.8 ± 8.8 years (range, 30 to 69 years). No significant differences were noted with regard to patient age, operation type, or chemotherapeutic regimen between the two groups. However, there were more patients who received neoadjuvant chemotherapy (p = 0.021), and more patients who were sensitive to taxane in the doxorubicin sensitive group (p = 0.001, Table 1).

Pathologic results according to doxorubicin sensitivity

The pathologic results of all patients according to doxorubicin sensitivity have been presented in Table 2. No significant differ-

Table 1. Patients’ demographics according to doxorubicin sensitivity

| Variables               | Sensitive group (n=19) | Resistant group (n=83) | p-value |
|-------------------------|------------------------|------------------------|---------|
| Age (yr)*               | 49.0±9.3               | 46.8±8.8               | 0.365   |
| Operation               |                         |                        | 0.430   |
| BCS                     | 5 (26.3)               | 32 (38.6)              |         |
| MRM                     | 14 (73.7)              | 51 (61.4)              |         |
| Chemotherapy type       |                         |                        | 0.021   |
| Neoadjuvant             | 9 (47.4)               | 17 (20.5)              |         |
| Adjuvant                | 10 (52.6)              | 66 (79.5)              |         |
| Chemotherapy regimen    |                         |                        | 0.097   |
| AC followed by T        | 13 (68.4)              | 71 (85.5)              |         |
| AT                      | 6 (31.6)               | 12 (14.5)              |         |
| Taxane sensitivity      |                         |                        | 0.001   |
| Sensitive               | 13 (68.4)              | 21 (25.3)              |         |
| Resistant               | 6 (31.6)               | 62 (74.7)              |         |

Values are presented as mean±SD or number (%). BCS=breast conserving surgery; MRM=modified radical mastectomy; AC=doxorubicin+cyclophosphamide; T=taxane; AT=doxorubicin+taxane. *Mean±SD.
ences were observed in histologic type, HER2-positive status, histologic grade, nuclear grade, tumor size, and nodal status between the two groups. However, HER2-positive type cancer, TNBC, and ER-negative cancers were more frequently observed in the sensitive group ($p < 0.05$). All patients had metastases of axillary lymph nodes (LNs) or lymphovascular invasion.

Table 2. Pathologic results according to doxorubicin sensitivity

| Variables                          | Sensitive group $n=19$ | Resistant group $n=83$ | $p$-value |
|------------------------------------|------------------------|------------------------|-----------|
| Histology                          |                        |                        | 0.548     |
| IDC                               | 19 (100.0)             | 78 (94.0)              |           |
| ILC                               | 0                      | 4 (4.8)                |           |
| Others                            | 0                      | 1 (1.2)                |           |
| Molecular subtype                  |                        |                        | 0.001     |
| Luminal A                         | 2 (10.5)               | 51 (61.5)              |           |
| Luminal B                         | 5 (26.3)               | 10 (12.0)              |           |
| HER2 type                         | 3 (15.8)               | 8 (9.6)                |           |
| TNBC                              | 9 (47.4)               | 14 (16.9)              |           |
| Estrogen receptor                  |                        |                        | 0.006     |
| Positive                          | 7 (36.8)               | 61 (73.5)              |           |
| Negative                          | 12 (63.2)              | 22 (26.5)              |           |
| HER2 status                       |                        |                        | 0.082     |
| Positive                          | 8 (42.1)               | 18 (21.7)              |           |
| Negative                          | 11 (57.9)              | 65 (78.3)              |           |
| Histologic grade                  |                        |                        | 0.077     |
| 1                                 | 0                      | 17 (20.5)              |           |
| 2                                 | 9 (47.4)               | 35 (42.2)              |           |
| 3                                 | 9 (47.4)               | 28 (33.1)              |           |
| Unknown                           | 1 (5.2)                | 5 (6.0)                |           |
| Nuclear grade                     |                        |                        | 0.216     |
| 1                                 | 0                      | 10 (12.0)              |           |
| 2                                 | 8 (42.1)               | 37 (44.6)              |           |
| 3                                 | 10 (52.7)              | 32 (38.6)              |           |
| Unknown                           | 1 (5.2)                | 4 (4.8)                |           |
| Tumor size (cm)                   |                        |                        | 0.479     |
| $\leq 2$                          | 7 (36.8)               | 31 (37.3)              |           |
| $>2, \leq 5$                      | 9 (47.4)               | 46 (55.4)              |           |
| $>5$                              | 3 (15.8)               | 6 (7.2)                |           |
| No. of metastatic LNs             |                        |                        | 0.326     |
| 0                                 | 2 (10.5)               | 8 (9.6)                |           |
| $\leq 3$                          | 11 (57.9)              | 40 (48.2)              |           |
| 4-9                               | 2 (10.5)               | 25 (30.2)              |           |
| $\geq 10$                         | 4 (21.1)               | 10 (12.0)              |           |

Values are presented as number (%). IDC = invasive ductal carcinoma; ILC = invasive lobular carcinoma; TNBC = triple negative cancer; LNs = lymph nodes.

Figure 1. Disease-free survival according to doxorubicin sensitivity.

Figure 2. Overall survival according to doxorubicin sensitivity.

Table 3. Univariate and multivariate analysis for DFS in all patients

| Variables                              | Univariate analysis | Multivariate analysis |
|----------------------------------------|---------------------|-----------------------|
|                                       | CI                  | $p$-value             | CI              | $p$-value |
| ER status                              | 0.147-1.244         | 0.109                 | 0.207-2.208     | 0.517 |
| HER2 status                            | 0.468-4.295         | 0.538                 | 0.191-2.413     | 0.549 |
| Histologic grade                       | 0.880-4.915         | 0.096                 | 0.484-5.192     | 0.446 |
| Nuclear grade                          | 0.860-6.351         | 0.096                 | 0.297-4.482     | 0.837 |
| Tumor size                             | 0.694-3.276         | 0.300                 | 0.335-1.884     | 0.602 |
| No. of metastatic LNs                  | 0.997-3.301         | 0.051                 | 1.203-4.771     | 0.013 |
| Doxorubicin sensitivity                | 1.286-13.827        | 0.034                 | 1.541-17.029    | 0.008 |
| Taxane sensitivity                     | 0.721-6.045         | 0.175                 |                   |       |

DFS = disease free survival; CI = confidence interval; ER = estrogen receptor; LNs = lymph nodes.
**Patient prognosis according to doxorubicin sensitivity**

In our study, a total of 14 cancer recurrences were observed, 7 of which were in the sensitive group. Among these, 3 patients died of breast cancer. We compared DFS and OS according to doxorubicin sensitivity. In the sensitive group, DFS and OS were significantly worse than those in the resistant group (Figures 1, 2).

Univariate analysis for DFS indicated that only doxorubicin sensitivity was significantly associated with high risk of recurrence ($p = 0.006$; confidence interval [CI], 1.550-12.994). With respect to taxane sensitivity, no significant effect was observed on DFS ($p = 0.175$; CI, 0.721-6.045). We performed a multivariate analysis for significant factors in univariate analysis and established breast cancer prognostic factors, and according to the results, a higher nodal stage ($p = 0.013$; CI, 1.203-4.771) and doxorubicin sensitivity ($p = 0.008$; CI, 1.541-17.029) were associated with poor DFS (Table 3).

**DISCUSSION**

Anthracycline and taxane have been the most commonly used chemotherapeutic agents for breast cancer patients. In neoadjuvant treatment, it had been reported that anthracycline-based chemotherapy produced more superior treatment results in TNBC/basal like breast cancer (BLBC) compared to non-TNBC/BLBC. Furthermore, these results were improved by adding taxane to the anthracycline [8-10]. With respect to adjuvant settings, anthracycline-based chemotherapy has been effective for patients with HER2-positive type and positive TOP2A and also for TNBC patients when compared with CMF chemotherapy [18,19]. In HER2 over-expression tumors, the effect of anthracycline has been shown to be mediated by TOP2A. Anthracycline binds to TOP2A and stabilizes DNA double strand breaks, which results in cell cycle arrest and apoptosis, and it has been known that the TOP2A gene is located close to the HER2 gene on chromosome 17. TOP2A gene co-amplification has been found in about 40% of HER2-positive type breast cancers [4,5].

Among the various chemosensitive assay tests, ATP-CRA showed a relatively high diagnostic accuracy for predicting response to drugs, and this result was similar with or superior to those of other tests. The ATP-CRA measured intracellular ATP, which is a basic energy source for living cells, and which rapidly disappears when cells lose viability. This has an advantage of a high success rate in primary culture, in addition to requiring only a small number of cells. Therefore ATP-CRA has been widely used as a chemosensitivity assay modality for the treatment of various cancers [11-16]. Among studies on the correlation between ATP-CRA results and patient prognosis, Konecny et al. [20] reported that patients with chemosensitivity had a favorable prognosis (progression-free survival [PFS], OS) compared to patients with chemoresistance in primary FIGO III ovarian cancer. In breast cancer, Ahn et al. [17] reported that the doxorubicin sensitive group had a lower early recurrence rate.

It has been previously reported that in neoadjuvant chemotherapy, TNBC/BLBC, and HER2-positive type are more sensitive to anthracycline and taxane chemotherapy. Despite this result, these cancer subtypes have worse prognoses among residual lesions after chemotherapy [7-10]. Furthermore, it has been suggested that ER-negative cancers showed a better response to primary chemotherapy than ER-positive cancers [21,22]. Actually, in our study, there were relatively more HER2-positive type, TNBC, ER-negative, and taxane-sensitive breast cancer patients in the sensitive group compared to those in the resistant group (Table 2). Thus, we think that doxorubicin sensitivity can reflect tumor biology, and more tumor types with a worse prognosis may be found in the doxorubicin sensitive group. For this reason, prognoses of patients in the sensitive group might be poorer than those of the resistance group. We determined doxorubicin and taxane sensitivity using cell death ratio cut-offs of 40% and 24%, respectively.

Ahn et al. [17] suggested that tumor recurrence within first two years after operation was associated with chemotherapy resistance and therefore, we determined cut-off values for doxorubicin and taxane using ROC curves based on their study. However, these cut-off values have been a little bit different in every study, and a consensus value has yet to be established [12]. Thus, we believe that additional studies addressing this issue should be performed, as well as studies utilizing a larger number of patients.

We performed univariate and multivariate analyses to determine whether the doxorubicin sensitivity could be an independent prognostic factor. Univariate analysis for DFS indicated that only doxorubicin sensitivity was associated with poor DFS ($p = 0.006$; CI, 1.550-12.994) whereas in the multivariate analysis, doxorubicin sensitivity ($p = 0.008$; CI, 1.541-17.029) and higher nodal stage ($p = 0.013$; CI, 1.203-4.771) were associated with poor DFS.

In the present study, we could not find any significant relationship between well known prognostic factors of breast cancer and prognosis except LN status. This may have been due to a small number of patients ($n = 102$) and relatively short follow-up period (29.8 ± 15.6 months). With respect to OS, we could not get a meaningful result because of the limited number of cancer-related deaths in the univariate and multivariate analysis.

There are some limitations in our study. This study was performed according to doxorubicin sensitivity, although all patients were treated with combined or sequential chemotherapy (doxorubicin and taxane or doxorubicin and cyclophospha-
Can This Assay Predict the Prognosis of Breast Cancer Patients?

The main purpose of a chemosensitive test is to choose the most effective chemotherapeutic agent or exclude non-effective agents for cancer patients. Additionally, if prognostic information for those patients is made available in this study, it may present a further benefit of this test. According to our study, the doxorubicin sensitive group was related with poor DFS and OS. Additionally, most of the patients in the sensitive group were HER2-positive types and TNBC. Considering these facts, we could predict the prognosis of patients who received anthracycline and taxane based chemotherapy with doxorubicin sensitivity on the basis of ATP-CRA results. However, as mentioned, higher numbers of patients and longer study periods are needed, as well as further studies on the precise cut-off values for drug sensitivity, and the most effective way to show the results of combined or sequential chemotherapy.

**CONFLICT OF INTEREST**

The authors declare that they have no competing interests.

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