A new species of *Bembidion* Latrielle 1802 from the Ozarks, with a review of the North American species of subgenus *Trichoplataphus* Netolitzky 1914 (Coleoptera, Carabidae, Bembidiini)

Drew A. Hildebrandt¹, David R. Maddison²†

¹ 710 Laney Road, Clinton, MS 39056 USA ² Department of Zoology, Oregon State University, Corvallis, OR 97331, USA

† urn:lsid:zoobank.org:author:038776CA-F70A-4744-96D6-B9B43FB56BB4  ‡ urn:lsid:zoobank.org:author:075A5E9B-5581-457D-8D2F-0B5834CDE04D

Corresponding author: David R. Maddison (david.maddison@science.oregonstate.edu)

Academic editor: T. Erwin  |  Received  31 July 2011  |  Accepted 25 August 2011  |  Published 16 November 2011

Citation: Hildebrandt DA, Maddison DR (2011) A new species of *Bembidion* Latrielle 1802 from the Ozarks, with a review of the North American species of subgenus *Trichoplataphus* Netolitzky 1914 (Coleoptera, Carabidae, Bembidiini). In: Erwin T (Ed) Proceedings of a symposium honoring the careers of Ross and Joyce Bell and their contributions to scientific work. Burlington, Vermont, 12–15 June 2010. ZooKeys 147: 261–275. doi: 10.3897/zookeys.147.1872

Abstract

A new species of *Bembidion* (*Trichoplataphus* Netolitzky) from the Ozark Plateau of Missouri and Arkansas is described (*Bembidion ozarkense* Maddison and Hildebrandt). It is distinguishable from the closely related species, *B. rolandi* Fall, by characteristics of the male genitalia, and sequences of the genes cytochrome oxidase I and 28S ribosomal DNA. A brief review of the North American species of *Trichoplataphus* is presented, including a key to species.

Keywords

*Bembidion*, Bembidiini, Trechinae, COI, 28S rDNA, taxonomy, systematics

Introduction

While identifying beetles of the genus *Bembidion* Latrielle that the senior author collected in Arkansas and Missouri some years ago, he came across a series of specimens
of subgenus *Trichoplataphus* Netolitzky that keyed to *Bembidion rolandi* Fall using the latest revision of the group (Lindroth 1963). However, the collecting locality was far west of the known range of that species (Bousquet and Larochelle 1993). He contacted the junior author for his opinion on the species identification, which initiated a study that has resulted in the present work.

The purpose of this paper is to describe the new species that was discovered and provide a review of the North American species of the subgenus *Trichoplataphus*. The most recent revision of the subgenus was included in Lindroth’s treatment of the genus for the northern U.S. and Canada (Lindroth 1963), and this paper builds on that work by addition of the new species, and modification of Lindroth’s key.

It is with great pleasure that we dedicate this paper to Dr. Ross Bell. For one of us he has been a very generous, informative and helpful correspondent, for the other a mentor and close colleague. Both of us have benefited immeasurably by our interactions with him. This paper is a small contribution to the fauna of an area that has interested Ross since his youth, when he first collected the endemic *Chlaenius viduus* Horn in the Ozarks.

**Methods**

Approximately 580 specimens of *Bembidion* (*Trichoplataphus*) were examined as part of this review; no effort was made to do a complete survey of specimens in existing collections. Specimens were examined from the following collections; each collection listing begins with the codon used in the text.

**CMNH** Carnegie Museum of Natural History, Pittsburgh, PA, USA (Robert L. Davidson)

**DAHC** Drew A. Hildebrandt collection, Clinton, MS, USA

**OSAC** Oregon State Arthropod Collection, Oregon State University, Corvallis, OR (David R. Maddison)

**TAMU** Texas A&M University, College Station, TX, USA (Edward C. Riley)

**Collecting methods.** Specimens were collected by hand, by splashing or pouring water on gravel bars or by raking the gravel by hand during the day to dislodge the beetles hiding under the gravel; or at night when the beetles were out, actively moving about on the surface. During the day, raking the gravel by hand works best for collecting specimens of this subgenus because of their behavior when disturbed. Unlike members of most other Bembidiini, individuals of *Trichoplataphus* tend to run under the water and cling to stones, and thus they are more difficult to catch (Davidson 1978).

Specimens for morphological studies were killed and preserved in sawdust or woodchips to which ethyl acetate was added. Specimens for DNA sequencing were collected into 95% or 100% ethanol, with best results obtained if the abdomen was slightly separated from the rest of the body to allow better penetration.
Morphological methods. Methods for studying adult structures, and terms used, are given in Maddison (1993).

All measurements were made on dried, pinned or pointed specimens using an Olympus SZ6045 zoom stereo microscope with a calibrated ocular micrometer. Character examinations were made at magnifications ranging from 10x to 63x; all measurements were made at 30x. Standardized body length (SBL) was measured following the protocol of Kavanaugh (1979).

Taxon sampling for DNA studies. We sequenced DNA from 18 specimens of *Bembidion* (*Trichoplataphus*), including all known North American species, as well as *Bembidion mimekara* Toledano and Schmidt from China (Table 1). Preliminary analyses of multiple genes across *Bembidion* (Maddison, unpublished) indicate that the sampled *Trichoplataphus* form a clade, and that, among the species sampled, *B. mimekara* is the sister group to the North American species. DNA vouchers are housed in the David Maddison voucher collection at Oregon State University.

DNA sequencing. Methods for obtaining DNA sequences are described in Maddison (2008). In brief, we obtained ca. 1000 bases of sequence data in the D1 through D3 domains of 28S ribosomal DNA (28S or 28S rDNA) and 650 to 750 bases of cytochrome oxidase I (COI). Fragments for these genes were amplified using the Polymerase Chain Reaction on an Eppendorf Mastercycler Thermal Cycler, using either Eppendorf Hotmaster Taq or TaKaRa Ex Taq and the basic protocols recommended by the manufacturers. Primers and details of the cycling reactions used are given in Maddison (2008). In particular, we used the primer pair LS58F and LS998R and the pair NLF184/21 and LS1041R to amplify and sequence 28S rDNA. For COI, two amplification and sequencing strategies were used: use of primer pairs B1490 and Bcoi2R (see Maddison 2008), or the LCO1490 and HCO2198 primers (Hebert et al. 2003). Amplified products were cleaned, quantified, and sequenced at the University of Arizona's Genomic and Technology Core Facility using either a 3730 or 3730 XL Applied Biosystems automatic sequencer.

Assembly of multiple chromatograms for each gene fragment and initial base calls were made with Phred (Green and Ewing 2002) and Phrap (Green 1999) as orchestrated by Mesquite's Chromaseq package (Maddison and Maddison 2009a; Maddison and Maddison 2009b), with subsequent modifications by Chromaseq and manual inspection. Multiple peaks at a single position in both reads were coded using IUPAC ambiguity codes.

Sequences have been deposited in GenBank with accession numbers JF800039 through JF800074.

Alignment. Alignment for both genes could be unambiguously determined, as there were no insertion or deletion events evident in the COI sequences, and only one in 28S, in which *B. mimekara* has 2 bases not present in the other species.

Molecular phylogenetic analysis. Models of nucleotide evolution were chosen with the aid of ModelTest version 3.7 (Posada 2005). For 28S rDNA, the model chosen by the Akaike Information Criterion (AIC) was a General Time Reversible (GTR) rate matrix with a proportion of sites being invariant (the GTR + I model); for COI it was a GTR rate matrix with site variation following a gamma distribution (the GTR + G model).
Table 1. *Bembidion* (*Trichoplataphus*) specimens sampled for DNA sequences. Four-digit numbers under “#” are D.R. Maddison DNA voucher numbers. The specimen marked with a “*” is the holotype of *B. ozarkense*, n.sp.

| Species          | #       | Locality                                                                 |
|------------------|---------|---------------------------------------------------------------------------|
| *B. mimekara* Toledano & Schmidt | 1366, 2448 | China: Yunnan: Gongshan County, Dulongjiang Township, Kongdan, east bank of Dulong Jiang. 1510 m, 27.87764°N, 098.33618°E |
| *B. planum* (Haldeman) | 1977 | USA: Arkansas: Henderson Creek at route 23, Ozark Mountains |
| *B. planum* (Haldeman) | 1423 | USA: Indiana: Crawford Co., English, Camp Fork Creek, 150m 38.3334°N, 86.4646°W |
| *B. planum* (Haldeman) | 2542 | USA: Kansas: Jefferson Co., Little Slough Ck, W of Kiowa Rd. 39.23355°N, 95.39497°W |
| *B. planum* (Haldeman) | 1974 | USA: Missouri: Washington Co., Big River at route 21, 245m, 37.8121°N, 90.7723°W |
| *B. planum* (Haldeman) | 1975 | USA: North Carolina: Mitchell Co., Penland, North Toe River, 744m, 35.9293°N, 82.1149°W |
| *B. fugax* (LeConte) | 2285 | USA: Pennsylvania: Perry Co., Susquehanna River 5 km N New Buffalo, 40.4909°N, 76.9535°W |
| *B. grandiceps* Hayward | 1689 | USA: Iowa: Webster Co., Des Moines River near Stratford, 275m, 42.3101°N, 93.9371°W |
| *B. grandiceps* Hayward | 1997 | USA: Iowa: Webster Co., Des Moines River near Stratford, 275m, 42.3101°N, 93.9371°W |
| *B. grandiceps* Hayward | 2540 | USA: Kansas: Jefferson Co., Little Slough Ck, W of Kiowa Rd. 39.23355°N, 95.39497°W |
| *B. grandiceps* Hayward | 2543 | USA: Kansas: Jefferson Co., Little Slough Ck, W of Kiowa Rd. 39.23355°N, 95.39497°W |
| *B. ozarkense*, sp. n. | 1976 | USA: Missouri: Carter Co., Current River at Van Buren, 135m, 36.9904°N, 91.0100°W |
| *B. ozarkense*, sp. n. | 1995 | USA: Missouri: Washington Co., Irondale, Big River, 230m, 37.8302°N, 90.6895°W |
| *B. ozarkense*, sp. n. | 2552* | USA: Missouri: Carter Co., Current River at Van Buren, 135m, 36.9904°N, 91.0100°W |
| *B. ozarkense*, sp. n. | 2554 | USA: Missouri: Carter Co., Current River at Van Buren, 135m, 36.9904°N, 91.0100°W |
| *B. rolandi* Fall | 1319 | Canada: Nova Scotia: Economy River at route 2, 45.3868°N, 63.8992°W |
| *B. rolandi* Fall | 1996 | USA: Virginia: Rockbridge Co., Maury River, Glasgow, 215m, 37.6329°N, 79.4431°W |
| *B. rolandi* Fall | 2553 | USA: Pennsylvania: Perry Co., Susquehanna River 5 km N New Buffalo, 40.4909°N, 76.9535°W |

Likelihood analyses of nucleotide data were conducted using Garli version 1.0.699 (Zwickl 2006). For each matrix, 1000 non-parametric bootstrap search replicates were conducted.
Results from molecular analyses

Specimens of each of the species have distinctive COI and 28S sequences (Fig. 1). *B. rolandi* shows consistent differences from similar specimens from the Ozarks, which we are describing as *B. ozarkense*. All specimens of *B. rolandi* differ from all specimens of *B. ozarkense* at 18 bases among the 766 sequenced sites in COI (2.3% divergent), as well as 2 of the 995 sites sampled of 28S rDNA (0.2%). These two species are reciprocally monophyletic in the inferred phylogenetic trees (Fig. 1).

Descriptions and identification of taxa

Subgenus *Trichoplataphus* Netolitzky, 1914

The subgenus *Trichoplataphus* of the genus *Bembidion* (Coleoptera: Carabidae: Trechinae: Bembidiini) contains 19 described species in the Palaearctic Region (Toledano and Schmidt 2010), and four species reported from North America (Bousquet and Larochelle 1993; Lindroth 1963; Lorenz 2005).

Adult specimens of the subgenus are easily separated from most other groups in North America by the irregular scattering of setiferous punctures on the abdominal sternum (Lindroth 1963). The only other species in North America with adults that have extra setiferous punctures on the abdominal sternum is *Bembidion hasti* Sahlberg, in which the setae are arranged in a regular, transverse row on each sternum.

Members of *Trichoplataphus* are found on open, bare gravel bars and banks, sometimes mixed with clay or sand (Fig. 2; Larochelle and Larivière 2003). Although mostly confined to running waters, they have also been collected among gravel and pebbles on banks of islands in Lake Champlain (Davidson 1978). These beetles are macropterous and have been found to fly to light (Larochelle and Larivière 2003).

The North American species of *Trichoplataphus* are:

*Bembidion ozarkense* sp. n.
*Bembidion rolandi* Fall, 1922
*Bembidion grandiceps* Hayward, 1897
*Bembidion fugax* (LeConte, 1848)
*Bembidion planum* (Haldeman, 1843)

Identification of Species Using Morphological Data

Species of this subgenus are very difficult to tell apart using morphological features, as the known differences are very subtle. Future morphological studies may reveal better characters to separate them. In the meantime, the key we present below (a modified version of couplets 77 through 80 in Lindroth 1963) should help in identifying speci-
Figure 1. Maximum-likelihood bootstrap trees for A 28S rDNA and B COI. Each terminal taxon is a single specimen, whose state or province of origin is shown, using standard abbreviations, after the species name; the four digits at the end of the name form the voucher number. Numbers below each branch are the percentage of bootstrap replicates showing that branch; values are not shown within each species. Branch lengths are proportional to number of substitutions per site, as reconstructed by Garli. *Bembidion* (*Trichoplataphus*) *mimekara*, the outgroup, is not shown because of its long branch length.
Mentum with epilobes of normal size for a Bembidion, projected notably anteriad of the mentum tooth (Fig. 3A) .................................................. 77a

From the Ozark Plateau of Missouri and Arkansas; aedeagus with tip relatively blunt, and not sharply bent downward (Fig. 4A and 4B) ........... B. ozarkense

Mentum with epilobes much reduced, projected about as far anteriad as the mentum tooth, and with a prominent concavity on the lateral side of each epilobe (Fig. 3B) ........................................................................................................ 78

Prothorax strongly constricted at base (Lindroth 1963, fig. 144); although constricted, parallel-sided base of prothorax very short. Elytra quite flat at apex ........................................................................................................ B. grandiceps

**Figure 2.** Type locality of Bembidion ozarkense, the Current River at Van Buren (USA: Missouri: Carter County).
– Prothorax less constricted. Elytra slightly sloping at extreme tip. ..........79
79 6th elytral stria hardly weaker than 5th behind shoulder but obliterated to-
wards apex. Frontal furrows prolonged and strongly diverged behind poste-
rior supra-orbital punctures ......................................................... B. fugax
– 6th elytral stria much weaker than 5th, often obliterated throughout. Front-
al furrows ended just behind posterior supra-orbital puncture and little di-
verged .......................................................................................... B. planum

Bembidion ozarkense Maddison & Hildebrandt, sp. n.
urn:lsid:zoobank.org:act:042B6160-C581-4028-BC44-421C1448C831
http://species-id.net/wiki/Bembidion_ozarkense
Figs 3A, 4A, 4B, 5A, 6

Holotype. Male (in OSAC), here designated, labeled “USA: Missouri: Carter Co.,
Current River at Van Buren, 135m, 36.9904°N, 91.0100°W, 24.iv.2005. DRM
05.015. D.R. Maddison” / “David R. Maddison DNA2552 DNA Voucher” [pale
green paper] / “HOLOTYPE Bembidion ozarkense Maddison & Hildebrandt” [red
paper]”. Genitalia in glycerine vial with specimen; extracted DNA stored separately.
GenBank accession numbers for DNA sequences of the holotype are JF800056 (28S)
and JF800065 (COI).

Paratypes. 221 specimens as follows: USA: Missouri: Carter Co., Current River
at Van Buren, 135m, 36.9904°N, 91.0100°W (88 specimens); Carter Co., Current
River at Van Buren, 135m, 36.9924°N, 91.0167°W (3); Carter Co., Current River
at Van Buren, 135m, 36.9911°N, 91.0133°W (3); Maries Co., Maries River near Ar-
gyle, 200m, 38.2700°N, 92.0007°W (10); Reynolds Co., Clark National Forest. Sut-
ton’s Bluff Campground (6); Washington Co., Irondale, Big River, 230m. 37.8302°N,
90.6895°W (16); Washington Co., Big River at route 21, 245m. 37.8121°N,
90.7723°W (4); Washington Co., Big River at route 21, 245m, 37.8132°N, 90.7734°W
(1); Arkansas: Crawford Co., Lee Creek (1); Marion Co., Buffalo Point St.Pk. (42);
Searcy Co., 5 mi. W. Big Flat. Big Creek (47). Male genitalia have been examined from
at least one specimen from each paratype locality.

Paratypes of B. ozarkense have been deposited in the CMNH, DAHC, OSAC and
TAMU, and in the collections of The Natural History Museum, London (BMNH),
the California Academy of Sciences (CAS), the Field Museum of Natural History
(FMNH), the Museum of Comparative Zoology (MCZ), Muséum National d’Histoire
Naturelle in Paris (MNHN), the University of Arizona (UAIC), the University of Al-
berta Strickland Museum (UASM), and the National Museum of Natural History,
Smithsonian Institution (USNM).

Eighty-two specimens examined but not designated as paratypes are: USA: Mis-
souri : Pulaski Co., Devil’s Elbow. Big Piney River. 10km E. Waynesville. 734 ft. 37 50
A new species of Bembidion Latrielle 1802 from the Ozarks...

Figure 3. Menta of Bembidion. (Trichoplataphus). Scale bar is 0.1 mm. Features other than the menta have been digitally faded A B. ozarkense, USA: Missouri: Carter Co., Current River at Van Buren, 135m, 36.9904°N, 91.0100°W B B. grandiceps, USA: Iowa: Webster Co., Des Moines River near Stratford, 275m, 42.3101°N, 93.9371°W, voucher DNA1689.

52.4 N 92 03 42.0 W (2); ARKANSAS: Marion Co., Buffalo Point S.P. (15); Searcy Co., 5.3 mi. W. Big Flat. Big Creek. (65). The Missouri specimens are omitted from the paratype series as there are no males whose genitalia could be examined; the Arkansas specimens are omitted as they are slightly damaged.

Type locality. USA: Missouri: Carter Co., Current River at Van Buren, 135m, 36.9904°N, 91.0100°W. At the type locality (Fig. 2), adults of B. ozarkense were com-
Figure 4. Aedeagus of male *B. ozarkense* and *B. rolandi*. Scale bar is 0.1 mm

**A** *B. ozarkense*, USA: Missouri: Carter Co., Current River at Van Buren, 135m, 36.9904°N, 91.0100°W Voucher DNA1976

**B** *B. ozarkense*, USA: Missouri: Washington Co., Irondale, Big River, 230m, 37.8302°N, 90.6895°W, Voucher DNA1995

**C** *B. rolandi*, USA: Virginia: Rockbridge Co., Maury River, Glasgow, 215m, 37.6329°N, 79.4431°W, Voucher DNA1996

**D** *B. rolandi*, Canada: Nova Scotia: Economy River at route 2, 45.3868°N, 63.8992°W, Voucher DNA1319.
A new species of Bembidion Latrielle 1802 from the Ozarks...
B. ozarkense tends to be darker than B. rolandi, with less-rufous elytra, and with the posterior medial portion of the dorsal surface of the head darker, rarely with the rufous region of many B. rolandi; the second antennomere of B. ozarkense is often infuscated centrally, whereas it is usually entirely pale rufous in B. rolandi. However, while these tendencies are noticeable in larger series, there is enough overlap between species that they are of marginal use when comparing individual specimens.

The only morphological characteristic that we have found to reliably distinguish the two species is in the male aedeagus: the tip of B. rolandi is thin, and abruptly bent downward (Fig. 4C and 4D; n=8), traits not found in B. ozarkense (Fig. 4A and 4B; n=30). DNA sequence data can also be definitively used to identify the species; either COI or 28S rDNA will suffice.

However, the known distribution ranges are distinctly separate, and can be used in the absence of male genitalia or DNA sequences to identify specimens.

**Geographic distribution.** The known specimens of this species are from the Ozark Plateau of Missouri and Arkansas (Fig. 6).
Bembidion rolandi Fall, 1922
http://species-id.net/wiki/Bembidion_rolandi

**Diagnosis.** As discussed above, specimens of this species cannot be separated from *B. ozarkense* using external characteristics, although they can be distinguished by male genitalia; see the diagnosis under *B. ozarkense* for details.

**Geographic distribution.** Currently recorded from **Canada**: NB, NS, ON, QC; **USA**: DC, MA, MD, ME, NJ, NY, OH, PA, VA, VT, WV (Bousquet and Larochelle 1993 and R. Davidson, pers. comm.).

Bembidion grandiceps Hayward, 1897
http://species-id.net/wiki/Bembidion_grandiceps

**Diagnosis.** Most specimens can be separated from *B. fugax* and *B. planum* by a combination of: broad head and pronotum; prothorax markedly constricted at base with the lateral margins just in front of the hind angle parallel, although this parallel region is much shorter than in *B. rolandi* or *B. ozarkense*; elytra quite flat at apex; and broad, markedly delimited frontal furrows that are clearly diverging between eyes and extended beyond the posterior supraorbital puncture.

**Geographic distribution.** We have seen specimens from **USA**: TX, KS, IA. Reports from the literature (Hayward 1897) of the presence of *B. grandiceps* in **USA**: DC, MA, NJ, NY, and PA are likely incorrect, and may be based on *B. fugax*.

Bembidion fugax (LeConte, 1848)
http://species-id.net/wiki/Bembidion_fugax

**Junior synonyms.** Octhedromus planipennis LeConte, 1850; Bembidion champlaini Casey, 1918.

**Diagnosis.** Most specimens can be separated from *B. grandiceps* using the characters listed for that species. Also, specimens of *B. fugax* have a more narrow head and pronotum and the elytral apex slopes slightly.

This species can be separated from *B. planum* by the 6th elytral stria being as, or nearly as, impressed behind the shoulder as the 5th, and the frontal furrows are prolonged and markedly diverging behind the posterior supra-orbital puncture. In addition, *B. fugax* has deeper furrows with steeper sides, more polished especially at the bottom, and more sharply etched at the bottom, with the raised area between the eye and the frontal furrow being more bulbous, and shinier; *B. planum* has broad, shallow furrows, sides gently sloped, more microsculpture on the slopes and especially bottoms, not sharply etched at the bottom, and with the raised area between the eye and the frontal furrow being flatter and not as shiny. Specimens tend to be darker and slightly larger than those of *B. planum* and in most specimens the pronotal microsculpture is weakly impressed and obsolete on the disk.
**Geographic distribution.** Recorded from **USA**: DC, IL, IN, MA, MD, NJ, NY, OH, PA, TN, VA, VT (Bousquet and Larochelle 1993 and R. Davidson, pers. comm.).

*Bembidion planum* (Haldeman, 1843)
http://species-id.net/wiki/Bembidion_planum

**Junior synonyms.** *Bembidium guexii* Chaudoir, 1868: 242; *Bembidion vulsum* Casey, 1918: 55; *Bembidion filicorne* Casey, 1918: 56.

**Diagnosis.** Most specimens of this species can be separated from *B. grandiceps* using the characters listed for that species. Also, specimens of *B. planum* have a narrower head and pronotum and the elytral apex slopes slightly.

Most specimens of this species can be separated from *B. fugax* by the more weakly impressed 6th compared with 5th elytral stria, and the frontal furrows end just behind the posterior supra-orbital puncture and are little divergent. Specimens tend to be paler and slightly smaller than those of *B. fugax*, and in most specimens have strongly-impressed microsculpture over the entire pronotum.

**Geographic distribution.** Recorded from **Canada**: NB, NS, ON, QC; **USA**: AR, CT, DC, IA, IL, IN, KY, MA, MD, ME, MI, MN, MO, MS, NC, NH, NJ, NY, OH, PA, RI, SC, TN, VA, VT, WI, WV (Bousquet and Larochelle 1993 and Maddison, unpublished).

**Acknowledgements**

We thank the curators for the collections from which we borrowed specimens.

We owe special thanks to Robert Davidson (CMNH) for sharing his particular knowledge of the habitats and habits of this group, for informing us of some characters that help separate *B. fugax* and *B. planum*, and for providing a thoughtful review of the manuscript.

We also thank the following for collecting specimens of *Trichoplataphus* for our DNA studies: Kojun Kanda, Taro Eldridge, and Zach Falin (*B. grandiceps* from Kansas); Monica Hughes (*B. planum* from Arkansas); David Kavanaugh (*B. mimekara* from China). Thanks as well to A.E. Arnold for help in collecting at numerous other localities.

This project was enabled by funds generously provided by the University of Arizona, and the Harold E. and Leona M. Rice Endowment Fund at Oregon State University.
A new species of Bembidion Latrielle 1802 from the Ozarks...

References

Bousquet Y, Larochelle A (1993) Catalogue of the Geadephaga (Coleoptera: Trachypachidae, Rhysoidea, Carabidae including Cicindelini) of America north of Mexico. Memoirs of the Entomological Society of Canada 167: 1–397. doi: 10.4039/entm125167fv

Davidson RL (1978) Bembidion rolandi Fall (Coleoptera: Carabidae) in Vermont, with a note on its habitat and behaviour. Cordulia 4: 93–94.

Green P (1999) Phrap. Version 0.990329. [http://phrap.org]

Green P, Ewing B (2002) Phred. Version 0.020425c. [http://phrap.org]

Hayward RT (1897) On the species of Bembidium of America North of Mexico. Transactions of the American Entomological Society 24: 32–143.

Hebert PDN, Cywinska A, Ball SL, DeWaard JR (2003) Biological identifications through DNA barcodes. Proceedings of the Royal Society of London Series B-Biological Sciences 270: 313–321. doi: 10.1098/rspb.2002.2218

Kavanaugh DH (1979) Studies of the Nebriini (Coleoptera: Carabidae), III. New Nearctic Nebria species and subspecies, nomenclatural notes, and lectotype designations. Proceedings of the California Academy of Sciences 42: 87–133.

Larochelle A, Larivière M-C (2003) A natural history of the ground-beetles (Coleoptera: Carabidae) of America north of Mexico. Pensoft, Sofia & Moscow, 583 pp.

Lindroth CH (1963) The ground-beetles (Carabidae, excl. Cicindelinae) of Canada and Alaska. Part 3. Opuscula Entomologica Supplementum XXIV: 201–408.

Lorenz W (2005) Systematic list of extant ground beetles of the world (Insecta Coleoptera “Geadephaga”: Trachypachidae and Carabidae incl. Paussinae, Cicindelinae, Rhysoidea). Second edition. Published by the author, Hörmannstrasse 4, D-82327 Tutzin, 530 pp.

Maddison DR (1993) Systematics of the Holarctic beetle subgenus Bracteon and related Bembidion (Coleoptera: Carabidae). Bulletin of the Museum of Comparative Zoology 153: 143–299.

Maddison DR (2008) Systematics of the North American beetle subgenus Pseudoperyphus (Coleoptera: Carabidae: Bembidion) based upon morphological, chromosomal, and molecular data. Annals of Carnegie Museum 77: 147–193. doi: 10.2992/0097-4463-77.1.147

Maddison DR, Maddison WP (2009a) Chromoseq: a Mesquite module for analyzing sequence chromatograms. Version 0.97. [http://mesquiteproject.org/packages/chromaseq]

Maddison WP, Maddison DR (2009b) Mesquite: a modular system for evolutionary analysis. Version 2.71. [http://mesquiteproject.org]

Posada D (2005) Modeltest: A tool to select the best-fit model of nucleotide substitution. Version 3.7. [http://darwin.uvigo.es]

Toledano L, Schmidt J (2010) Revision of the Bembidion kara Andrewes, 1921 species group and notes on the Palaearctic species of Bembidion subgenus Trichoplatus Netolitzky, 1914 (Coleoptera, Carabidae, Bembidiini). Entomologische Blätter für Biologie und Systematik der Käfer 106: 371–406.

Zwickl DJ (2006) Genetic algorithm approaches for the phylogenetic analysis of large biological sequence datasets under the maximum likelihood criterion. PhD Dissertation, Austin, Texas: The University of Texas at Austin.