Population-Based Estimate of Prostate Cancer Risk for Carriers of the HOXB13 Missense Mutation G84E

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Abstract

The HOXB13 missense mutation G84E (rs138213197) is associated with increased risk of prostate cancer, but the current estimate of increased risk has a wide confidence interval (width of 95% confidence interval (CI) >200-fold) so the point estimate of 20-fold increased risk could be misleading. Population-based family studies can be more informative for estimating risks for rare variants, therefore, we screened for mutations in an Australian population-based series of early-onset prostate cancer cases (proband). We found that 19 of 1,384 (1.4%) probands carried the missense mutation, and of these, six (32%) had a family history of prostate cancer. We tested the 22 relatives of carriers diagnosed from 1998 to 2008 for whom we had a DNA sample, and found seven more carriers and one obligate carrier. The age-specific incidence for carriers was estimated to be, on average, 16.4 (95% CI 2.5–107.2) times that for the population over the time frame when the relatives were at risk prior to baseline. We then estimated the age and birth year-specific cumulative risk of prostate cancer (penetrance) for carriers. For example, the penetrance for an unaffected male carrier born in 1950 was 19% (95% CI 5–46%) at age 60 years, 44% (95% CI 18–74%) at age 70 years and 60% (95% CI 30–85%) at age 80 years. Our study has provided a population-based estimate of the average risk of prostate cancer for HOXB13 missense mutation G84E carriers that can be used to guide clinical practice and research. This study has also shown that the majority of hereditary prostate cancers due to the HOXB13 missense mutation are ‘sporadic’ in the sense that unselected cases with the missense mutation do not typically report having a family history of prostate cancer.

Introduction

The HOXB13 missense mutation G84E (rs138213197) has been reported to be associated with increased risk of prostate cancer [1]. The original study identified 72 case carriers among 5,083 tested, but just one control carrier among 1,401 tested. Therefore, their point estimate of a 20-fold increased risk was highly imprecise (width of 95% confidence interval (CI) >200-fold) and could be misleading. A more recent paper reported that the mutation was about 8 times more prevalent in cases with a family history of prostate cancer compared with controls; their ratio is also very imprecise as only two controls were carriers [2]. Moreover, one cannot interpret this comparison as a legitimate estimate of increased risk; see [3]. Another recent paper reported a conventional case-control study of 1,525 cases and 1,757 controls that found just two control carriers, and the width of the 95% confidence interval for the odds ratio estimate of 5.8 was 20-fold [4]. That is, estimating risk for rare variants using case-control studies is problematic [5]. Only a newly published study from Sweden had a reasonable number of carrier controls (N = 24 and 37 for the two study populations reported) to estimate risk [6].

For rare variants, a population-based family study can be more informative [5]. To provide accurate and unbiased estimates, it is important that the residual disease correlation within families is taken into account. Failure to adequately adjust for this can lead to upwardly biased penetrance estimates for rare, moderate risk genotypes [7]. The mixed model is one approach that can allow for the simultaneous effect of the HOXB13 missense mutation G84E and a polygenic component. We have, therefore, used the family cancer histories of a population-based series of early-onset prostate cancer cases (proband), unselected for family history, to estimate from a mixed model the age-specific cumulative risk of prostate cancer (penetrance) for carriers.

Results and Discussion

Of the 1,505 case probands identified, we successfully genotyped 1,384 (92%). The genotyped and non-genotyped case probands had similar average age at diagnosis (52.4 versus 52.1 years, respectively). We found that 19 (1.4%) case probands carried the HOXB13 G84E missense mutation. Table 1 shows their pedigree structures at baseline and the clinical details sufficient to estimate penetrance. Six case probands (32%) had a
family history of prostate cancer; three had one, and three had two, affected relatives. An additional 22 relatives were genotyped, of whom seven were carriers. Of the nine affected relatives, three were carriers, one was an obligate carrier and the other five could not be genotyped. There were four unaffected carrier relatives (aged 42, 50, 73 and 81 years).

The age-specific incidence for carriers was estimated to be, on average, 16.4 (95% CI 2.5–107.2) times that for the Australian population. There was no evidence that the hazard ratio decreased with age (P = 0.7) or year of birth (P = 0.3), but we had little power to address this issue. The estimated penetrance for carriers, by assumption, varied by year of birth; for example, the estimated penetrance for a male born in 1950 was 19% (95% CI 5–46%) at age 60 years, 44% (95% CI 18–74%) at age 70 years and 60% (95% CI 30–85%) at age 80 years (see Figure 1 and Table 2).

The major strength of our study is that all case probands were sampled from a population-based cancer registry, irrespective of their family history of prostate cancer. Another strength is that we have allowed for residual phenotype correlation within families. Consequently, our estimates of penetrance are unbiased [7].

Our study was focussed on early-onset disease, and sampling restricted to probands diagnosed before the age of 60 years. Therefore, if there are factors that modify risk for carriers, our estimates will be relevant to carriers enriched for those factors. It would be of interest to conduct the same study using probands diagnosed at a later age to see if penetrance is lower. It will also be of interest to see what happens to these families in the future, especially as the proband’s brothers move into higher risk age groups. A limitation of this study is the imprecision of the risk estimates (i.e. wide confidence intervals), a consequence of the small numbers of carrier probands and affected relatives, and the limited proportion of relatives with genotype information. Our estimates of age-specific incidence for carriers is consistent with previous studies, [1,2,4,6]. Given we found no evidence that the hazard ratio differed by calendar year, with the caveat of limited power to do so, the varying penetrances depending on year of birth are a consequence of the changing incidence rates over calendar time, most likely due to changes in screening. Up to 50% of our cases would have been unlikely to have come to clinical attention in the absence of PSA testing [8]. Larger family studies of cases with a wider range of age at diagnosis are needed to obtain more precise penetrance estimates. It is also an open question as to whether this penetrance estimate applies to other mutations in the HOXB13 gene.

Our study has provided an estimate of the cumulative risk of prostate cancer for HOXB13 missense mutation G84E carriers that can be used to guide clinical practice and research. This study has also shown that, similar to high-risk mutations in other cancer susceptibility genes, the majority of hereditary prostate cancers due to the HOXB13 missense mutation are ‘sporadic’ in the sense that unselected cases with the missense mutation do not typically report having a family history of prostate cancer [9].

### Table 1. Clinical details of the 19 HOXB13 missense mutation G84E carriers (rs138213197), their male relatives and their mothers at baseline, sufficient to calculate the penetrance estimates. *

| Family | Age at onset | Gleason's score | Stage | Affected | Male relatives (relationship/observation time/carrier status) |
|--------|--------------|----------------|-------|----------|------------------------------------------------------------|
|        |              |                |       |          | Proband's diagnosis details | Male relatives | Carrier status of mother |
|        |              |                |       |          | Age at | Gleason's score | Stage | Unaffected | Affected | Status of mother |
| 1      | 54           | 7              | IIA   | None     | MU/69/7, S/31/7, S/29/7 | ?              |
| 2      | 53           | 6              | I     | None     | F/80/7 | ?              |
| 3      | 52           | 7              | IIA   | None     | S/18/7 | ?              |
| 4      | 53           | 5              | IV    | None     | B/50/7, B/43/7, B/42/7, B/35/7, F/78/7 | +              |
| 5      | 58           | 7              | IIA   | None     | F/72/7, S/33/7, S/28/7 | ?              |
| 6      | 52           | 7              | IIA   | None     | B/50/7, F/76/7 | ?              |
| 7      | 56           | 7              | IIA   | None     | B/54/7, S/30/7, S/28/7, PU/79/7 | ?              |
| 8      | 53           | 9              | III   | None     | S/25/7, S/21/7 | ?              |
| 9      | 50           | 7              | IIA   | None     | F/81/7, S/24/7, MU/80/7, MGF/85/7, PU/78/7, PU/73/7 | –              |
| 10     | 52           | 7              | III   | None     | F/76/7, S/1/7 | ?              |
| 11     | 54           | 6              | IIA   | None     | B/49/7, B/43/7, S/30/7 | ?              |
| 12     | 53           | 6              | III   | MU/57/7, | F/46/7, S/28/7, HB/42/7, HB/34/7, MGF/65/7 | +              |
| 13     | 55           | 7              | IIA   | F/68/7, PU/73/7 | MU/80/7, PFG/90/7 | –              |
| 14     | 47           | 7              | III   | F/63/7, | MU/70/7, MU/67/7, MGF/83/7, PFG/83/7, S/14/7 | –              |
| 15     | 54           | 9              | IV    | None     | B/50/7, F/64/7, S/18/7 | ?              |
| 16     | 52           | 7              | IIB   | F/63/7, MU/56/7 | MU/64/7, MU/36/7 | ?              |
| 17     | 55           | 6              | IIB   | MU/61/7, MU/71/7 | B/66/7, B/57/7, S/32/7, S/29/7, S/28/7, F/67/7, MU/67/7, MU/80/7, MGF/87/7 | –              |
| 18     | 55           | ?              | ?     | None     | B/47/7, F/78/7, PU/74/7, S/17/7, S/12/7 | ?              |
| 19     | 44           | 7              | IIA   | F/70/7 | B/53/7, B/51/7, B/48/7, MU/80/7, MGF/87/7 | –              |

* B, brother; F, father; HB, half-brother; MU, maternal uncle; MGF, maternal grandfather; PU, paternal uncle; PFG, paternal grandfather; S, son.

Observation time began at date of birth and ended at the earliest of the following: date of diagnosis of prostate cancer, date at death, or 31 Dec 2011. Carrier status: + carrier, (+) obligate carrier, – non-carrier, ? unknown carrier status.

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Materials and Methods

Ethics statement

The study protocol was approved by the Human Research Ethics Committee of the Cancer Council Victoria. All participants provided written consent to participate in the study.

Participants

Between 1998 and 2008, all men with histopathologically confirmed carcinoma of the prostate were identified through the population-complete Victorian Cancer Registry. Recruitment included all men younger than 55 years at diagnosis but the upper age limit varied over time according to available numbers, with quotas being filled by randomly sampling additional cases aged from 55 to 59 years. Overall, 68% of eligible men approached participated in the study.

Cases were mailed questionnaires in which they were asked to identify, and provide a detailed cancer history for, their first- and second-degree relatives. Strenuous efforts were made to verify reported prostate cancer diagnoses by use of multiple sources, including cancer reported by relatives, pathology reports, medical records, and death certificates. All probands and selected relatives were asked to provide a blood sample for DNA analysis.

Genotyping

All probands who donated a blood sample were genotyped for the HOXB13 missense mutation G84E (rs138213197). For the identified carriers, all relatives from whom a blood sample had been obtained were also genotyped for this missense mutation. Genotyping was performed with the TaqMan assay (Applied Biosystems) using a 384 well format on a LightCycler480, and data were interpreted using LightCycler480 1.5.0 software (Roche Diagnostics, Castle Hill, Australia). The DNA plates contained 31 (1.7%) blind duplicate DNA samples and 6 negative control (blank) wells per 384 well plate that returned high quality genotyping measures (consistent genotyping calls and no evidence of contamination). All identified carriers were confirmed by Sanger sequencing using BigDye v.3.1 chemistry (Applied

Table 2. Penetrance (age-specific cumulative risk) and 95% confidence intervals of prostate cancer for carriers of the HOXB13 missense mutation G84E (rs138213197) by selected years of birth.

| Birth year | 1920 | 1930 | 1940 | 1950 |
|------------|------|------|------|------|
| Age        | Penetrance | 95% CI | Penetrance | 95% CI | Penetrance | 95% CI | Penetrance | 95% CI |
| 50         | 0.4  | (0.1,2.3) | 0.4  | (0.1,2.3) | 0.4  | (0.1,2.3) | 1.5  | (0.2,7.2) |
| 55         | 1.5  | (0.2,7.3) | 1.6  | (0.2,7.4) | 3.2  | (0.5,13.2) | 7.0  | (1.5,23.5) |
| 60         | 4.6  | (0.8,17.2) | 5.0  | (1.0,18.4) | 11.6 | (2.7,33.0) | 19.2 | (5.3,45.9) |
| 65         | 10.3 | (2.3,30.5) | 16.2 | (4.2,41.2) | 23.2 | (6.8,51.4) | 32.1 | (11.0,62.2) |
| 70         | 19.0 | (5.2,45.6) | 28.9 | (9.5,58.6) | 37.7 | (14.1,68.1) | 44.2 | (18.0,74.1) |
| 75         | 34.2 | (12.2,64.5) | 40.0 | (15.4,70.3) | 48.6 | (21.0,77.5) | 53.1 | (24.4,80.9) |
| 80         | 45.7 | (19.0,75.3) | 49.9 | (22.0,78.6) | 56.4 | (27.0,83.1) | 60.0 | (30.1,85.4) |

Figure 1. Penetrance (age-specific cumulative risk) (solid line), and 95% confidence limits intervals (dashed lines), of prostate cancer for carriers of the HOXB13 missense mutation G84E (rs138213197) by selected years of birth.

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Biosystems) and chromatograms visualized using chromas (URL: http://www.technelysium.com.au/chromas.html).

**Statistical methods**

Prostate cancer incidence \( \lambda_i(t) \) for individual \( i \) at age \( t \) was assumed to depend on the underlying genotype through a model of the form \( \lambda_i(t) = \lambda_0(t) \exp(G_i(t) + P_i) \) where \( \lambda_0(t) \) is the age- and birth cohort-specific population incidence at age \( t \), \( G_i(t) \) is the log relative risk corresponding to the \( \text{HOXB13} \) missense mutation \( G84E \) carrier status of individual \( i \) at age \( t \), and \( P_i \) is the polygenic component that is assumed to be normally distributed with mean 0 and variance \( \sigma_P^2 \). The polygenic component is approximated by the hypergeometric polygenic model [10]. \( \sigma_P \) was fixed to equal 2.01 based on a previous segregation analysis [11]. Five-year age-specific population incidences for 1982–2008 in Australia were obtained from the AIHW [12] and smoothed using locally weighted regression techniques [13].

Given probands were sampled independently of family cancer history, for both the penetrance and the HR analyses, we adjusted for ascertainment by conditioning the likelihood for each pedigree on the proband’s \( \text{HOXB13} \) missense mutation \( G84E \) carrier status, prostate cancer status, and age at diagnosis. We included all first- and second-degree relatives who were ascertained, irrespective of affected status.

Pedigree analyses were performed with the program MENDEL [14]; all other calculations were performed in Stata 11.

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**Author Contributions**

Conceived and designed the experiments: GG JH MS. Performed the experiments: MS. Analyzed the data: RM JH. Contributed reagents/materials/analysis tools: GS LB JD MJ. Wrote software used for analyses: RM JD LB MJ.

**References**

1. Ewing CM, Ray AM, Lange EM, Zuhlke KA, Robbins CM, et al. (2012) Germline mutations in \( \text{HOXB13} \) and prostate-cancer risk. N Engl J Med 366: 141–149.
2. Breyer JP, Avritt TG, McReynolds KM, Dupont WD, Smith JR (2012) Confirmation of the \( \text{HOXB13} \) \( G84E \) Germline Mutation in Familial Prostate Cancer. Cancer Epidemiol Biomarkers Prev.
3. Byrnes GB, Southey MC, Hopper JL (2008) Are the so-called low penetrance breast cancer genes, \( \text{ATM, BRIP1, PALB2 and CHEK2} \), high risk for women with strong family histories? Breast Cancer Res 10: 208.
4. Alkhari MR, Trachtenberg J, Lee J, Tam S, Bristow R, et al. (2012) Association Between Germline \( \text{HOXB13} \) \( G84E \) Mutation and Risk of Prostate Cancer. J Natl Cancer Inst.
5. Hopper JL, Bishop DT, Easton DF (2005) Population-based family studies in genetic epidemiology. Lancet 366: 1397–1406.
6. Karlsson R, Aly M, Clements M, Zheng L, Adolfsen J, et al. (2012) A Population-based Assessment of Germline \( \text{HOXB13} \) \( G84E \) Mutation and Prostate Cancer Risk. Eur Urol.
7. Gong G, Hannon N, Whittemore AS (2010) Estimating gene penetrance from family data. Genet Epidemiol 34: 373–381.
8. Erzioni R, Cha R, Feuer EJ, Davidow O (1998) Asymptomatic incidence and duration of prostate cancer. Am J Epidemiol 148: 775–785.
9. Cui J, Hopper JL (2000) Why are the majority of hereditary cases of early-onset breast cancer sporadic? A simulation study. Cancer Epidemiol Biomarkers Prev 9: 805–812.
10. Lange K (1997) An approximate model of polygenic inheritance. Genetics 147: 1423–1430.
11. MacInnis RJ, Antoniou AC, Eeles RA, Severi G, Guy M, et al. (2010) Prostate cancer segregation analyses using 4390 families from UK and Australian population-based studies. Genet Epidemiol 34: 42–50.
12. AIHW (Australian Institute of Health and Welfare) (2011) ACIM (Australian Cancer Incidence and Mortality) Books. Canberra: AIHW.
13. Royston P (1991) Lowess smoothing: Stata Tech Bull 3: 7–9.
14. Lange K, Weeks D, Boechke M (1988) Programs for Pedigree Analysis: MENDEL, FISHER, and eGENE. Genet Epidemiol 5: 471–472.