Could lymphatic mapping and sentinel node biopsy provide oncological providence for local resectional techniques for colon cancer? A review of the literature
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Abstract

Background: Endoscopic resectional techniques for colon cancer are undermined by their inability to determine lymph node status. This limits their application to only those lesions at the most minimal risk of lymphatic dissemination whereas their technical capacity could allow intraluminal or even transluminal address of larger lesions. Sentinel node biopsy may theoretically address this breach although the variability of its reported results for this disease is worrisome.

Methods: Medline, EMBASE and Cochrane databases were interrogated back to 1999 to identify all publications concerning lymphatic mapping for colon cancer with reference cross-checking for completeness. All reports were examined from the perspective of in vivo technique accuracy selectively in early stage disease (i.e. lesions potentially within the technical capacity of endoscopic resection).

Results: Fifty-two studies detailing the experiences of 3390 patients were identified. Considerable variation in patient characteristics as well as in surgical and histological quality assurances were however evident among the studies identified. In addition, considerable contamination of the studies by inclusion of rectal cancer without subgroup separation was frequent. Indeed such is the heterogeneity of the publications to date, formal meta-analysis to pool patient cohorts in order to definitively ascertain technique accuracy in those with T1 and/or T2 cancer is not possible. Although lymphatic mapping in early stage neoplasia alone has rarely been specifically studied, those studies that included examination of false negative rates identified high T3/4 patient proportions and larger tumor size as being important confounders. Under selected circumstances however the technique seems to perform sufficiently reliably to allow it prompt consideration of its use to tailor operative extent.

Conclusion: The specific question of whether sentinel node biopsy can augment the oncological propriety for endoscopic resective techniques (including Natural Orifice Transluminal Endoscopic Surgery [NOTES]) cannot be definitively answered at present. Study heterogeneity may account for the variability evident in the results from different centers. Enhanced capacity (perhaps to the level necessary to consider selective avoidance of en bloc mesenteric resection) by its confinement to only early stage disease is plausible although not proven. Specific study of the technique in early stage tumors is clearly essential before proffering this approach.
Background

Advances in technological capability have made feasible the local resection of small colonic tumors by intraluminal and even transluminal endoscopy [1-4]. Although primarily now proposed for supposed benign lesions, in concept, selected germinal cancers could also be resected by these means. However, the insensitivity of preoperative radiological imaging for the detection of nodal metastases (approximately 70% of tumor-containing nodes are less than 5 mm in size [5-9]) and the inability of biopsy analysis to truly reflect the metastatic potential of the primary means that localized resection of the primary for even the earliest cancers risks either the understaging of systemic disease or the rendering of the effort redundant if formal resection becomes indicated by the full pathology of the resected specimen[10]. A reliable means of definitively establishing lymph node status peroperatively, other than en bloc mesenteric resection, would greatly increase the oncological providence of these techniques and could expand their application.

Sentinel node biopsy would seem on first principles well suited to address this breach as it fulfills a similar role in tumors of the breast and skin. This technique has also been recently proposed to accompany endoscopic dissection of early gastric cancers in order to enhance functional outcome by minimizing the extent of surgical resection [11-13]. Adjoining such a 'diagnostic laparoscopy' to an endoscopic resective technique could be justified in selected patients if the outcome of the node biopsy would permit localized excision as the definitive intervention in place of radical operation. Conversely, if the node is revealed as positive for metastases, the surgeon can confidently advocate radical operation in cases when the tumor appears confined. Synchronous laparoscopy has indeed already been advocated for the endoscopic resection of certain difficult or large polyps[14]. Furthermore, it seems likely that increasing experience with transluminal peritoneal access and intervention (i.e. Natural Orifice Transluminal Endoscopic Surgery [NOTES]) could mean that selective lymph node biopsy without abdominal wall ingress will be practicable in the near future[15]. However, lymphatic mapping in intestinal cancers is still considered controversial because of reports of varying accuracy and concerns regarding reliability and reproducibility.

To date however no comprehensive study or review has been performed from the perspective of using lymphatic mapping to facilitate minimally resective techniques for early stage colon tumors. Analyses to date have instead focused primarily on the capacity of the technique to predict recurrence risk through the upstaging of conventionally node negative disease after standard operation has been performed [16-24]. The main focus has therefore been on Stage II rather than Stage I cancers with the sentinel node biopsy and analysis being performed in addition to rather than in place of formal lymphadenectomy[25]. The aim of this review is therefore to formally interrogate the evidence base in its entirety to determine whether lymphatic mapping can, on any basis, be rationally proposed to augment the oncological propriety of localized endoscopic resection specifically for the small, early stage colon cancers that lie within its scope.

Note: Rectal cancers lie outside the premise of this review as the anatomical arrangement of the mesorectum (specifically its bulk, retroperitoneal position and lack of serosal layer) precludes against intraoperative nodal biopsy for rectal cancer. Furthermore, violation of the mesorectum may also compromise any subsequent attempt at formal oncological resection (and hence negatively impact upon patient outcome) should this be indicated by the pathology of the resected specimen.

Methods

Search methods for identification of relevant studies

The following strategy was used to identify relevant publications regarding experience with lymphatic mapping in human patients with colon cancer (see Table 1). Firstly, PubMed software was used to search the Medline database between 1st January 1999 (the year of the earliest series published on the technique in colon cancer) and the 30th July 2008. The Cochrane library (including the Cochrane Central Register of Controlled Trials) and EMBASE databases were also directly searched in a similar fashion. The following expanded Medical Subject Headings were used-‘sentinel node’, ‘lymphatic mapping’, ‘colon cancer’, ‘colon tumo(u)rs’, ‘colorectal cancer/tumo(u)rs’, ‘large intestine’ and ‘gastrointestinal’ (although the intent of the study was confined to colon cancer the later three search terms were included for completeness as some series report mixed cohorts). The reference list of all full publications so identified along with that of consensus papers, review articles, editorials and relevant book chapters were cross-checked for additional relevant publications. Data contained in meeting abstracts were not studied as these were judged unlikely to present sufficient detail for extraction required by our study protocol. Only English language publications were included (language bias however seems unlikely since the literature search in Pubmed and

| Inclusion criteria |
|--------------------|
| English language publication |
| Full publication between 1st January 1999 and July 30th 2008. |
| Human patients with colon cancer |
| In vivo mapping and node identification |
Embde did not reveal any substantial non-english reports. Finally only those studies that used ed method-
ology alone for both the injection and the identification of salient sentinel nodes were analyzed (ex vivo mapping
in only be reliably performed if a specimen with its com-
plete mesentery has already been resected intact while ex
vivo identification can predispose to false negative glands
as redistribution of the dye along the lymphatic chain can
occur in the time between injection and node harvest). Intraoperative marking of the sentinel node rather than
actual excisional biopsy allowed inclusion however as
clearly the intent and purpose is the same.

Assessment of methodological quality of the studies
identified
Each of the studies identified was analyzed for suitability
for inclusion according to the criteria laid down by QUA-
DRAS – an evidenced base tool for the assessment of the
quality of diagnostic assessment studies[26,27]. By this
means it was determined whether the publication was of
sufficient quality to allow inclusion in this review.

Data extraction from included studies
All data extraction was performed by two authors (RAC
and JL) with cross-checking to ensure validation. When
any disparity or disagreements arose the investigators met
with the third author JM as final adjudicator to resolve the
debate. The fields for data capture were pre-specified
before analysis of the studies identified. Data pertaining
to patient demographics, technique methodology
(including both surgeon experience as well as precise tech-
nical details) and sentinel node efficacy by binary classifi-
cation (i.e. detection, accuracy, sensitivity and false
negative rates as well as negative predictive values) for
patients with colon cancer undergoing lymphatic map-
ing were prepared on Microsoft Excel datasheets. Data
from papers that explicitly declared themselves further
sub-analyses of a previous study were included with the
prior publication. Studies from the same authors but
which did not declare themselves to contain overlapping
cohorts were analyzed separately although are flagged in
the subsequent tables to indicate that this is possibility.

Upstaging rate refers to the number of times the sentinel node
was also recorded to allow for subsequent analysis and
consideration.

Sentinel Node Performance Parameters
Where possible, 2 × 2 contingency tables were built com-
prising true positive (both sentinel and non-sentinel
nodes involved), true negative (both sentinel and non-
sentinel nodes clear), false negative (sentinel node clear
but non-sentinel nodes involved). The term 'false positive'
(and hence calculation of 'specificity') is not appropriate
in studies regarding sentinel node in cancer because the
presence of isolated micrometastases in the sentinel node
confers node positivity on the patient. Nor, given the
experimental nature of sentinel node biopsy and the bio-
logical uncertainty of the significance of micrometastases,
is this term appropriate for use when micrometastases
alone are present in the sentinel node. Instead the term
upstaging is used to better reflect the standing of sentinel
nodes that are immunohistochemically positive when
other non-sentinel nodes are clear of disease.

The following definitions were therefore used to ascribe
the performance rates of sentinel node biopsy

Detection rate – refers to the number of times a sentinel
node was actually identifiable = (Number of successful
attempts to retrieve a sentinel node/Number of attempts
*100%).

Accuracy rate refers to the ability of the sentinel node to
reflect the overall status of the lymph basin (whether posi-
tive or negative) = ([(Number of correct predictions of the
nodal status by sentinel node biopsy/Number of patients
undergoing sentinel node biopsy)*100%].

Sensitivity refers to the number of times the sentinel node
reflects the fact that disease is present in the non-sentinel
nodes = (Number of patients with tumor-involved senti-
nel nodes/Number of patients with any lymph node con-
taining tumor)*100%.

The false negative rate reflects the proportion of patients
in whom no cancer was identified in the sentinel node but
who had nodal deposits found in their non-sentinel
nodes compared to the total number of those who had
mumour containing metastases in non-sentinel nodes =
(Number of false negative patients/Number of true posi-
tive cases + number of false-negative cases) * 100%.

Upstaging rate refers to the number of cases in which
sophisticated analysis of the sentinel node reveals tumor
deposits that otherwise would not have been detected =
(Number of patients revealing micrometastases or iso-
lated tumor cells in the sentinel node/Number of patients
undergoing sentinel node biopsy)*100%.
classified as N0 after routine histopathological examination)*100%.

Results

Results of literature search

There were no randomized controlled trails identified by our search methodology. Sixty-three clinical studies regarding lymphatic mapping and sentinel node biopsy for colon cancer in human patients were published in the English language during our review period of interest. Nine of these studies however actually utilized a primarily ex vivo lymph node identification technique (only the dye injection was performed in vivo and the surgeon made no attempt intraoperatively to identify any mapped nodes). These studies [28-37] (along with ten additional studies that utilized a primary ex vivo technique ab initio [38-46]) were therefore excluded from further analysis (see Figure 1 and Table 2). A further two studies were explicitly declared updates or further analyses of already reported patient cohorts.

This left fifty-two studies warranting consideration for inclusion in this study [21,47-97] (see Table 3). Although ten of these studies supplemented their in vivo work with additional ex vivo examination on a occasional basis (particularly in cases of failed mappings or rectal cancers), these were included in the review as their majority of cases underwent their mapping and biopsy entirely in vivo prior to specimen resection. In addition, aside from any contribution to multicenter trials, seven centers provided 27 studies to the literature base. Eight publications have come from the John Wayne Cancer Institute alone and seven have come from the McLaren Regional Center, Michigan State University alone (with a further four publications emerging from these two centers jointly). Two centers (Wake Forest, North Carolina and MD Anderson, Texas) have published two studies each while three centers (Mount Sinai, Miami Beach; Charité-University Medicine Berlin, Germany and the University Medical Center Groningen, Netherlands) have published two studies each. However, although some degree of overlap between patient cohorts is likely in successive publications that exact proportion of patients presented in duplicate was

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**Figure 1**

Flow chart showing the selection and exclusion of publications for this review.
not explicitly stated and so all studies were included albeit with flagging.

**Included studies**

In total fifty two diagnostic studies (listed in Table 3) detailing 3390 patient episodes were included in this analysis. Only two studies (comprising a total of 41 patients)\[52,91\] however set out specifically to evaluate lymphatic mapping selectively for early stage disease. This obviously is not a sufficient cohort to draw robust conclusions about the performance of the technique in this cohort. The clinical and methodological diversity (compounded by variations in data presentation) of the remaining forty-eight studies however was such that formal meta-analysis to allow statistical analysis supplement the power was neither possible nor prudent (QUADROS data not shown). Therefore this systematic review of the literature on this subject must take the form of a narrative synthesis of the evidence base in its entirety[98].

Sentinel node detection rates and false negative scores are listed alongside each publication in Table 3 and displayed graphically in Figure 2. As can be seen considerable variation in both detection rates (58–100%) and, especially, in false negative rates (0–75%) is evident. To be of any clinical utility the technique would seem to require these rates to be consistently better than 90% and 10% respectively (i.e. the considerable heterogeneity regarding variable patient cohorts and tumor characteristics as well as disparate surgeon experience and protocol differences in both the technical performance and pathological analysis) may also underlie these discordant results and will now therefore be critically analyzed in sequence. Tables 4, 5 and 6 detail the salient findings of each study by each of these criteria.

**Patient Demographics (Table 4)**

Ten studies gave no indication of mean (or median) patient age while eleven gave no breakdown of the patient population by gender. The age breakdown of the other studies show no especially striking data (mean age 69 years) although three studies have an unexplained clear preponderance (>70%) of males as have two of females among their cohorts. This suggests that their populations may be somewhat atypical. Only four studies present data regarding the BMIs of their patient population – a potentially important discrepancy that may induce error in both detection and false negative rates as intra-abdominal obesity may obscure discolored nodes in the mesentery. Finally, only 25 (50%) studies examined colon cancer in isolation. The vast majority (26) of the other studies also included rectal cancer. These studies, despite usually declaring the proportions of each tumor studied, most often did not present result sub-analysis. While the mean number of colon cancers studied in each publication is 68, 30 studies included less than 50 of such patients while 14 comprised less than 30 patients with colon cancer.

**Tumor profiles (Table 5)**

Despite the stated aim of most studies being the evaluation of the utility of lymphatic mapping for staging node negative tumors, 19 studies made no attempt to exclude patients with distant metastases or indeed evident
Table 3: Publications included for this analysis along with their concluding results regarding the rates of detection, accuracy, sensitivity, false negative and upstaging as well as the false negative rates associated with lymphatic mapping and sentinel node biopsy for colon cancer.

| Lead Author        | Year | Journal                  | Detection Rate | Accuracy Rate | Sensitivity Rate | False Negative Rate | Negative Predictive Value | Upstaging Rate |
|--------------------|------|--------------------------|----------------|---------------|------------------|----------------------|--------------------------|---------------|
| Saha (a)[47]       | 2000 | Ann Surg Oncol           | 99             | 96            | 91               | 9                    | Ns                       | 18            |
| Wiese (a)[48]      | 2000 | Arch Pathol Lab Med      | 99             | 96            | 91               | 9                    | ns                       | ns            |
| Waters (b)[49]     | 2000 | Am Surg                  | 91             | 100           | 100              | 0                    | 100                      | 6             |
| Bilchik (c)[50]    | 2001 | J Clin Oncol             | 100            | 100           | 100              | 0                    | 100                      | 50            |
| Paramo (d)[51]     | 2001 | Am J Surg                | 71             | 100           | 100              | 0                    | 100                      | 11            |
| Wood (a)+c[52]     | 2001 | Ann Surg Oncol           | 96             | 95            | 88               | 12                   | ns                       | 25            |
| Wood (c)[53]       | 2001 | Surg Endosc              | 100            | 100           | 100              | 0                    | 100                      | 9             |
| Saha (a)+c[54]     | 2001 | Ann Surg Oncol           | 98             | 96            | 90               | 10                   | 95                       | 14            |
| Esse [55]          | 2001 | Dis Colon Rectum         | 58             | 94            | 67               | 33                   | 94                       | 7             |
| Bendavid [56]      | 2002 | J Surg Oncol             | 94             | 94            | 95               | 5                    | 91                       | 42            |
| Paramo (e)[57]     | 2002 | Ann Surg Oncol           | 82             | 98            | 93               | 7                    | 97                       | 11            |
| Wood (c)[58]       | 2002 | J Gastroint Surg         | 97             | 95            | 92               | 8                    | ns                       | 24            |
| Bilchik (c)[59]    | 2002 | Eur J Cancer             | 97             | 95            | 91               | 5                    | ns                       | 24            |
| Kitagawa [60]      | 2002 | Dis Colon Rectum         | 91             | 92            | 92               | 18                   | 88                       | ns            |
| Feg [61]           | 2002 | Am J Surg                | 98             | 79            | 38               | 62                   | 76                       | 8             |
| Broderick-Ville [62]| 2002 | Cancer J                | 92             | 79            | 50               | 50                   | 73                       | 0             |
| Tsioulas (c)[63]   | 2002 | Am Surg                  | 100            | 93            | 67               | 33                   | ns                       | 15            |
| Nastro [64]        | 2002 | Tumori                   | 75             | 100           | 100              | 0                    | 100                      | ns            |
| Bilchik (c)[65]    | 2003 | Cancer Control           | 100            | 93            | 91               | 9                    | ns                       | 14            |
| Cox [66]           | 2003 | Curr Surg                | 100            | 100           | 100              | 0                    | 100                      | 29            |
| Bilchik (c)[67]    | 2003 | J Clin Oncol             | 96             | 96            | 92               | 8                    | ns                       | 29            |
| Turner (c)[68]     | 2003 | Archives Path            | 82             | 92            | 87               | 13                   | ns                       | 29            |
| Trocha (a)+c[69]   | 2003 | J Gastroint Surg         | 98             | 95            | 84               | 16                   | 93                       | 21            |
| Vehil [70]         | 2003 | World J Surg             | 87             | 78            | 50               | 22                   | 71                       | 11            |
| Levine (b)[71]     | 2003 | J Gastroint Surg         | 92             | ns            | 50               | ns                   | 86                       | ns            |
| Saha (a)[72]       | 2004 | Dis Colon Rectum         | 99             | 96            | 86               | 16                   | ns                       | 13            |
| Dan (a)[73]        | 2004 | Arch Surg                | 99             | 97            | 80               | 20                   | 92                       | 3             |
| Brea [74]          | 2004 | Eur J Surg Oncol         | 94             | 97            | 80               | 16                   | 92                       | 9             |
| Bertoglio [75]     | 2004 | J Surg Oncol             | 95             | 92            | 78               | 22                   | 88                       | ns            |
| Read [76]          | 2004 | Dis Colon Rectum         | 79             | 97            | 25               | 75                   | ns                       | 3             |
| Patten (e)[76]     | 2004 | Cancer                   | 98             | 89            | 83               | 17                   | 76                       | 20            |
| Bertagnolli [77]   | 2004 | Ann Surg                 | 92             | 80            | 46               | 54                   | 75                       | 0             |
| Saha (a)[78]       | 2004 | Ann Surg Oncol           | 100            | 95            | 84               | 16                   | 93                       | 5             |
| Saha (a)[79]       | 2004 | Semin Oncol              | 100            | 96            | 92               | 8                    | ns                       | 13            |
| Bembeneke (f)[80]  | 2005 | World J Surg             | 85             | ns            | 92               | 8                    | 95                       | 26            |
| Codignola [81]     | 2005 | J Clin Oncol             | 100            | 96            | 92               | 8                    | ns                       | 13            |
| Dahl [82]          | 2005 | Eur J Surg Oncol         | 100            | 92            | 83               | 17                   | 91                       | ns            |
| Bilchik (a)+c[83]  | 2006 | Arch Surg                | 100            | 95            | 88               | 12                   | ns                       | 23            |
| Tuche [84]         | 2006 | Eur J Surg Oncol         | 97             | 94            | 91               | 9                    | ns                       | ns            |
| Saha (a)[85]       | 2006 | Am J Surg                | 98             | 96            | 90               | 10                   | 93                       | ns            |
| Kelder (g)[86]     | 2006 | Scand J Gastroenterol    | 97             | 93            | 86               | 14                   | 89                       | 33            |
| Thomas (b)[87]     | 2006 | Am Surg                  | 93             | 20            | 46               | 54                   | 73                       | 5             |
| Cavarelli (b)[88]  | 2006 | Am Surg                  | 95             | 95            | 86               | 14                   | ns                       | 8             |
| Kelder (g)[89]     | 2007 | Int J Col Dis            | 97             | 96            | 89               | 11                   | 93                       | ns            |
| Bianchi [90]       | 2007 | Surg End                 | 100            | 95            | 83               | 17                   | 94                       | 9             |
| Murawa [91]        | 2007 | Acta Chir Belg           | 93             | 84            | 83               | 17                   | ns                       | 8             |
| Bembeneke (f)[92]  | 2007 | Ann Surg                | 85             | 86            | 54               | 46                   | 80                       | 21            |
| Sandruci [93]      | 2007 | J Surg Oncol             | 100            | 91            | 92               | 9                    | ns                       | 11            |
| Tiffet [94]        | 2007 | Dis Colon Rectum         | 92             | 81            | 80               | 20                   | 74                       | ns            |
| Lim (e)[95]        | 2008 | Ann Surg Oncol           | 99             | 83            | 59               | 41                   | 78                       | ns            |
| Kusano [96]        | 2008 | Digestive Surgery        | 88.5           | 82.6          | 33               | 67                   | 81                       | ns            |
| Quadros [97]       | 2008 | J Surg Oncol             | 91             | 80            | 67               | 33                   | 67                       | 25            |

The definitions for each are contained in the text. All rates are %; ns denotes data neither stated within nor readily derived from the results presented within the publication. Note: several centers have published more than one series without declaring overlap of their patient cohorts – such publications are flagged by the center of origin by the inclusion of letter subscript as follows: (a) McLaren Regional Center, Michigan, USA; (b) Wake Forest, North Carolina, USA; (c) John Wayne Cancer Institute, California, USA; (d) Mount Sinai, Miami Beach, USA; (e) MD Anderson, Texas; (f) Charité-University Medicine Berlin, Berlin, Germany; (g) University Medical Center Groningen, The Netherlands. Table 4, 5 and 6 also follow a similar format.
mesenteric deposits let alone grossly involved lymph nodes. Indeed some actually specifically included such patients. Of the remainder, seven excluded only patients with distant metastases (making no further consideration of local tumour invasion or mesenteric deposits) while nine required the patient had only ‘clinically localized’ or ‘resectable’ disease to allow their inclusion. Of the twelve papers that exclude gross nodal and distant metastases, five include T4 disease while five do not profile their tumors by T-stage. Overall, at least 21 studies include T4 tumors within their cohorts. Furthermore, 25 of the studies that present such data possess high T3 and T4 to T1 and T2 ratios. Only five studies specifically consider tumor length or diameter as a factor that may affect performance parameters. Interestingly, four of these studies conclude that false negative rates are considerably more likely in patients with larger tumors while the fifth only examined this by taking 4 cm as a cut-off point for analysis. Only one study considered how tumor size may relate to the quantity of dye needed to adequately map it and found a significant positive correlation. Although the mean number of resected nodes in each study is adequate (mean overall is in 15.3), 21 studies include patients who have had considerably less nodes than this resected in their ‘definitive’ operation while 25 do not state either the mean or range of the non-sentinel nodal harvest. This raises concern over the quality control mechanisms in place to ensure the standard of the oncological operation performed in these studies.

**Technical methodology (Table 6)**

Only five studies specifically sought to ensure surgeon experience in the technique prior to commencing their study (although an additional nine had already demonstrated their competence in a prior publication). This is likely particularly pertinent in the ten multicentre studies, only four of whom specifically sought to ensure minimum practical experience among their participants (although a further three studies came from units that had already published experiences and so this likely evinces expertise at least among some of the contributors). Injection methodology overall was relatively similar. 45 protocols utilized an intraoperative subserosal injection of colorimetric mapping agent while three employed a submucosal injection. Twenty eight studies utilized isosulfan blue 1% in isolation while ten used Patent Blue alone and one used Vital Blue. Eight studies used a radioisotope as a mapping agent (alone in one study and in conjunction with blue dye in the others) and the majority injected this agent submucosally preoperatively by additional endoscopy. Two studies also incorporated fluorescein while one used indocyanine green. Six studies specifically included laparoscopic operations with three employing this approach exclusively. All commenting authors agreed however that the technique is easily performed regardless of operative approach and adds minimally to overall operative time[63]. The mean number of sentinel nodes found was consistently approximately two although five studies included within their range numbers in excess of double this. There was though considerable variation among how identified nodes were histologically analyzed. Four studies examined only a single section of the sentinel node while nine used neither immunohistochemistry nor RT-PCR to look for micrometastases or isolated tumor cells (one of these did however subsequently publish an additional, detailed report on their results with such techniques[99]).

Despite the fact that the variation in accuracy and sensitivity rates is frequently decried, only fifteen publications specifically included analysis of their false negative rates (see Table 7 for a tabulated summary of their conclusions). Twelve of these found that increasing tumor stage was inversely related to non-sentinel node tumors and indeed in five studies the detection rate and diagnostic accuracy was 100% among their T1 and T2 cohorts. One other study found that the presence of lymphovascular invasion was significantly associated with false negative rates but that lymph node invasion did not reach significance as a predictor (no data was shown however). The remaining study analyzing its results by tumor stage found no significance difference with either tumor stage or an arbitrarily decided lesion diameter.

**Critical analysis of studies with low performance results**

The nine studies with detection rates < 90% and the thirty-two studies with false negative rates > 10% were then scrutinized from the perspectives gleaned from these analyses. Interestingly, five of the nine studies (>50%) with low detection rates also had false negative rates greater than 20% (actually 22%, 33%, 46% and 75%). Of the 43 studies with detection rates > 90%, only eleven (c. 25%) also had false negative rates greater than 20%.

Of those with detection rates < 90%, two were multicentre trials. Neither these nor six of the seven single centre studies stated they validated surgeon expertise prior to commencing patient enrollment. Furthermore each of these studies was composed of less than 60 patients. Five studies included a proportion of rectal cancers approximating 15% of the population within their study cohorts. Four studies had marked T3/4 to T1/2 preponderance (in the order of 93.3, 78.22 and 83.15 respectively). Only three studies of those with detection rates > 90% included such high proportions of locally advanced disease but one of these specifically excluded clinically apparent lymphadenopathy while the other did not contain any T4 cases. Furthermore one other study included patients with liver metastases and even obvious mesenteric deposits and direct nodal invasion by the primary while every
patient in another study was conventionally node positive. Finally the patients of one report had a mean BMI above 25 kg/m² (although this was not analyzed specifically further). None of the other studies presented any data in this latter regard.

Table 4: Cohort characteristics with regard to baseline demographics of the patients studied.

| Lead Author | % Males | Mean or median age | BMI (kg/m²) | Non-colonic cancers included | % with colon cancer | No with colon cancer |
|-------------|---------|-------------------|-------------|-----------------------------|---------------------|---------------------|
| Saha        | 46      | 71                | ns          | Rectum                      | 86                  | 74                  |
| Wiese       | 44      | 71                | ns          | Rectum                      | 70                  | 70                  |
| Waters      | ns      | ns                | ns          | None                        | 100                 | 22                  |
| Bilchik     | 43      | 70                | ns          | Rectum                      | 83                  | 33                  |
| Paramo      | 46      | 72                | ns          | None                        | 100                 | 35                  |
| Wood        | 40      | 68                | ns          | Rectum                      | 81                  | 61                  |
| Wood        | 36      | 64                | ns          | None                        | 100                 | 11                  |
| Saha        | ns      | ns                | ns          | Rectum                      | ns                  | ns                  |
| Esser       | 55      | 69                | ns          | Rectum                      | 84                  | 26                  |
| Bendavid    | ns      | ns                | ns          | None                        | 100                 | 20                  |
| Paramo      | 51      | 70                | ns          | None                        | 100                 | 55                  |
| Wood        | 49      | 68                | ns          | Rectum                      | 78                  | 78                  |
| Bilchik     | 49      | 68                | ns          | Rectum                      | 100                 | 100                 |
| Kitagawa    | 71      | 61                | ns          | Rectum                      | 21                  | 12                  |
| Feig        | 73      | 68                | ns          | None                        | 100                 | 48                  |
| Broderick-Villa | 50 | 63                | ns          | Rectum                      | 90                  | 40                  |
| Tsioulias   | ns      | ns                | ns          | None                        | 100                 | 14                  |
| Nastro      | ns      | ns                | ns          | None                        | 100                 | 8                   |
| Bilchik     | 12      | 64                | ns          | None                        | 100                 | 30                  |
| Cox         | 35      | Ns                | ns          | None                        | 100                 | 17                  |
| Bilchik     | 48      | 70                | ns          | Rectum                      | 85                  | 102                 |
| Turner      | 53      | 76                | ns          | Rectum                      | 86                  | 44                  |
| Trocha      | 56      | 71                | ns          | Rectum                      | 88                  | 44                  |
| Veihl       | 74      | 75                | 25.3        | None                        | 100                 | 31                  |
| Levine      | 55      | 67                | ns          | Stomach                     | 74                  | 37                  |
| Saha        | 48      | 71                | ns          | Rectum                      | 83                  | 336                 |
| Dan         | 49      | 72                | ns          | Rectum                      | 88                  | 106                 |
| Braatz      | ns      | ns                | ns          | None                        | 100                 | 35                  |
| Bertoglio   | 54      | ns                | ns          | Rectum                      | 77                  | 20                  |
| Read        | ns      | ns                | ns          | None                        | 100                 | 41                  |
| Patten      | ns      | 64.2              | ns          | None                        | 100                 | 50                  |
| Bertagnolli | 65      | 65                | ns          | None                        | 100                 | 72                  |
| Saha        | 47      | 71                | ns          | Rectum                      | 91                  | 52                  |
| Saha        | 48      | 71.3              | ns          | Rectum                      | 80                  | 209                 |
| Bembenek    | 49      | 45 to 83          | ns          | None                        | 100                 | 55                  |
| Codignigola | 36      | 70.8              | ns          | Rectum                      | 93                  | 52                  |
| Dahl        | 57      | 70                | ns          | None                        | 100                 | 30                  |
| Bilchik     | 47      | 74                | ns          | Rectum                      | 73                  | 97                  |
| Tuch        | ns      | 75.5              | ns          | None                        | 100                 | 30                  |
| Saha        | 48      | 74                | ns          | Rectum                      | 82                  | 408                 |
| Kelder      | 53      | 69                | ns          | None                        | 100                 | 30                  |
| Thomas      | 50      | 67                | c. 26.3     | None                        | 100                 | 69                  |
| Covarelli   | 60      | 70                | ns          | None                        | 100                 | 20                  |
| Kelder      | ns      | ns                | ns          | None                        | 100                 | 69                  |
| Bianchi     | 58      | 61                | ns          | None                        | 100                 | 22                  |
| Murawa      | ns      | 61                | ns          | Rectum                      | 48                  | 13                  |
| Bembenek    | 59      | 67                | ns          | None                        | 100                 | 315                 |
| Sandruci    | ns      | 73                | ns          | Rectum                      | 86                  | 30                  |
| Tiffet      | 50      | 73                | 25          | Rectum                      | 75                  | 49                  |
| Lim         | 48      | 67                | ns          | None                        | 100                 | 120                 |
| Kusano      | 70      | 70                | ns          | None                        | 100                 | 26                  |
| Quadros     | 36.5    | 56                | ns          | Rectum                      | 42                  | 22                  |

The order and format is the same as in Table 3.
Table 5: Patient selection criteria along with the resulting tumor profiles in each of the studies and the mean total lymph node harvests.

| Lead Author       | Selection criteria employed | Tumor Size (cm) | T Stage (% of total) | T12:34 ratio | Conventionally Node Negative (%) | No of lymph nodes resected (range) |
|-------------------|-----------------------------|----------------|---------------------|--------------|----------------------------------|----------------------------------|
| Saha              | None                        | ns             | Data not given      | ns           | 37                               | 16 (ns)                          |
| Wiese             | None                        | ns             | 0 14 22 53 11       | 36:63        | 41                               | 16 (ns)                          |
| Waters            | None                        | ns             | Data not given      | ns           | 27                               | 12 (ns)                          |
| Bilchik           | Early stage primary only    | ns             | 0 26 24 50          | 50:50        | 35                               | 15 (2–28)                        |
| Paramo            | No distant metastatic disease | ns             | 0 12 7 81 0        | 7:93         | 29                               | 10 (ns)                          |
| Wood              | Clinically localized primary | ns             | 0 19 29 44 8       | 48:52        | 47                               | 15 (2–28)                        |
| Wood              | Small early stage cancers only | 1.4          | 27 54 9 9 0       | 91:9         | 9                                | 13 (2–20)                        |
| Saha              | None                        | ns             | Data not given      | ns           | 40                               | 20 (ns)                          |
| Esser             | No nodal or distant metastases | ns             | Data not given      | ns           | 19.4                             | 15 (12–16)                       |
| Bendavid          | None                        | ns             | Data not given      | ns           | 65                               | ns                               |
| Paramo            | No distant metastatic disease | ns             | Data not given      | ns           | 27                               | 12 (ns)                          |
| Wood              | Clinically localized        | ns             | Data not given      | ns           | 26                               | 15 (2–28)                        |
| Bilchik           | Clinically localized        | ns             | 0 25 23 46 6       | 48:52        | 43                               | 15 (3–28)                        |
| Kitagawa          | Only if curative surgery    | ns             | Data not given      | ns           | 29:71                            | 43                               | 24 (ns)                          |
| Feig              | None                        | ns             | 7 6 23 58 6        | 36:64        | 33                               | 13 (4–46)                        |
| Broderick-Villa   | No known distant metastases | ?              | 6 8 20 62 4        | 34:66        | 43                               | 8 (1–17)                         |
| Tsioulias         | Clinically localized only   | ns             | Data not given      | ns           | 21                               | 14 (2–21)                        |
| N Castro          | None                        | ns             | Data not given      | ns           | ns                               | ns                               |
| Bilchik           | Early stage primary only    | ns             | 20 46 14 20 0      | 80:20        | 21                               | 14 (2–21)                        |
| Cox               | None                        | ns             | 0 6 36 58 0        | 42:58        | 41                               | 18 (4–33)                        |
| Bilchik           | Early stage primary only    | 3.6            | 14 12 17 53 5      | 42:58        | 36                               | 14 (ns)                          |
| Turner            | None                        | ns             | 0 12 10 75 4       | 22:78        | 52                               | 11 (1–42)                        |
| Trocha            | No distant metastases       | ns             | 26 12 18 42 2      | 56:44        | 38                               | 16 (ns)                          |
| Veihl             | None                        | 4.2            | 0 6 9 71 12        | 15:83        | 48                               | 21 (5–40)                        |
| Levine            | No gross nodal disease      | ns             | Data not given      | ns           | ns                               | ns                               |
| Saha              | Tumor resectable & no metastases | ns             | Data not given      | ns           | 42                               | ns                               |
Table 5: Patient selection criteria along with the resulting tumor profiles in each of the studies and the mean total lymph node harvests. (Continued)

| Study  | Criteria                                                                 | Tumor Profiles | LNH Mean | LNH SD  | LNH Min | LNH Max  | LNH Range |
|--------|---------------------------------------------------------------------------|----------------|----------|---------|---------|-----------|-----------|
| Dan    | None                                                                      | ns             | 17       | 15      | 13      | 53        | 3         | 43        | ns        |
| Braat  | No distant metastases, gross invasion or nodal disease                   | ns             | 0        | 6       | 20      | 51        | 23        | 26:74     | 34        | 9 (1–23)  |
| Bertoglio | Stage I and II & no enlarged nodes only                                 | ns             | Data not given | ns     | 65      | 13        | 6–18      |           |           |
| Read   | Surgery with curative intent only                                        | ns             | Data not given | ns     | 29      | 14        | 7–45      |           |           |
| Patten | No nodal or distant metastases                                          | ns             | Data not given | 37:63  | 39      | 14        | (ns)                  |           |           |
| Bertagnolli | Stage I, II and III only                                                | ns             | 0        | 29      | 16      | 46        | 9         | 35:65     | 67        | 17 (ns)   |
| Saha   | None                                                                      | ns             | 19       | 14      | 11      | 53        | 4         | 43:57     | 33        | 12 (ns)   |
| Saha   | None                                                                      | ns             | 11       | 10      | 15      | 51        | 5         | 34:66     | 41        | 14 (ns)   |
| Bembenek | Conventionally node negative patients only                              | ns             | Data not given | ns     | 100     | 26        | (10–59)    |           |           |
| Codignola | No liver metastases                                                     | ns             | 0        | 2       | 21      | 63        | 14        | 23:77     | 36        | 21 (6–47) |
| Dahl   | No nodal or distant metastases                                           | ns             | Data not given | 11:88  | 40      | 17        | (4–35)                 |           |           |
| Bilchik | Potentially curable cancer with no distant metastases only              | 3.5 (0.2–10.5) | 0        | 17      | 15      | 65        | 3         | 32:68     | 29        | 15 (ns)   |
| Tuech  | No nodal or distant disease                                              | ns             | 0        | 6       | 9       | 85        | 0         | 10:90     | 36        | 20 (12–32) |
| Saha   | None                                                                      | ns             | 15       | 11      | 16      | 52        | 5         | 42:58     | 50        | 15 (ns)   |
| Kelder | No gross nodal or distant metastases                                     | ns             | 0        | 0       | 23      | 73        | 4         | 23:77     | 21        | 14 (ns)   |
| Thomas | None                                                                      | ns             | Data not given | ?      | 38      | ns        |           |           |           |
| Covarelli | None                                                                    | ns             | Data not given | ns     | 35      | ns        |           |           |           |
| Kelder | No gross nodal or distant metastases                                     | ns             | 0        | 1       | 20      | 70        | 9         | 21:79     | 41        | 11 (ns)   |
| Bianchi | No T4 or metastatic disease                                              | ns             | 36       | 4.5     | 9       | 45        | 5         | 50:50     | 73        | 22 (8–38) |
| Murawa | No gross nodal or distant metastases                                     | ns             | 0        | 15      | 20      | 63        | 2         | 37:63     | 41        | 20 (3–96) |
| Bembenek | None                                                                    | ns             | Data not given | ns     | 69      | 20        | (4–79)                |           |           |
| Sandrucci | Stage I or II only                                                      | ns             | Data not given | 100:0  | 69      | 9        | (ns)                  |           |           |
| Tiffet | Excluded if primary unresectable.                                        | ns             | Data not given | 22:78  | 41      | 18        | (4–37)                 |           |           |
| Lim    | No gross nodal or distant metastases                                     | ns             | 0        | 4       | 26      | 66        | 4         | 30:70     | 16        | 13 (ns)   |
| Kusano | None                                                                      | ns             | 0        | 15      | 73      | 12        | 12        | 88:12     | 77        | 13.5 (ns) |
| Quadros | Potential curable cancer, no distant metastases                           | 8.3            | 0        | 0       | 14      | 54        | 31        | 14:86     | 46        | 19 (ns)   |

The order and format is the same as in Table 3.
Table 6: Surgeon and technical factors associated with nodal mapping, identification and pathological analysis methodology.

| Lead Author | Surgeon Experience sought | Multicentre | Laparoscopic resection (%) | Mapping agent used | Supplementary ex vivo identification | Mean/median No. of sentinel nodes (range) | Sentinel node analysis |
|-------------|---------------------------|-------------|---------------------------|--------------------|--------------------------------------|-------------------------------------------|-----------------------|
| Saha        | No                        | No          | ns                        | Isosulfan blue 1%  | No                                   | 1.6 (1 to 4)                             | Yes                   |
| Wiese       | No                        | No          | 0                         | Isosulfan blue 1%  | No                                   | 1.9 (1 to 4)                             | Yes                   |
| Waters      | No                        | No          | ns                        | Isosulfan blue 1%  | No                                   | ns                                        | Yes                   |
| Bilchik     | Yes                       | Yes         | ns                        | Isosulfan blue 1%  | No                                   | 2 (1 to 3)                               | Yes                   |
| Paramo      | No                        | No          | 0                         | Isosulfan blue 1%  | No                                   | 1.4 (1 to 4)                             | Yes                   |
| Wood        | No                        | No          | 12                        | Isosulfan blue 1%  | Yes (10%)                            | 2 (1 to 4)                               | Yes                   |
| Wood        | No                        | No          | 100                       | Isosulfan blue 1%  | No                                   | 2 (1 to 3)                               | Yes                   |
| Saha        | No                        | Yes         | ns                        | Isosulfan blue 1%  | No                                   | 1.7 (1 to 4)                             | Yes                   |
| Esser       | No                        | No          | 0                         | Isosulfan blue 1%  | No                                   | 1.7 (0 to 5)                             | No                    |
| Bendavid    | No                        | No          | ns                        | Isosulfan blue 1%  | No                                   | 3.9 (0 to 5)                             | Yes                   |
| Wiese       | No                        | No          | ns                        | Isosulfan blue 1%  | Yes (15%)                            | 2 (1 to 4)                               | Yes                   |
| Wood        | No                        | No          | ns                        | Isosulfan blue 1%  | Yes (11%)                            | 2 (1 to 4)                               | Yes                   |
| Bilchik     | No                        | No          | 16                        | Isosulfan blue 1%  | Yes (1%)                             | 2 (1 to 4)                               | Yes                   |
| Kitagawa    | No                        | No          | 0                         | Technetium         | No                                   | 3.5 (0 to 8)                             | No                    |
| Feig        | No                        | Yes         | 0                         | Isosulfan blue 1%  | No                                   | 2.6 (0 to 7)                             | Yes                   |
| Broderick-Villa | Yes                     | No         | 100                       | Isosulfan blue 1%  | Yes (23%)                            | 1.5 (0 to 5)                             | Yes                   |
| Tsoulias    | No                        | No          | 100                       | Isosulfan blue 1%  | No                                   | 1.7 (1 to 3)                             | Yes                   |
| Nastro      | No                        | No          | 0                         | Technetium and blue dye | No                                  | ns                                        | Yes                   |
| Bilchik     | Yes                       | No          | 23                        | Isosulfan blue 1%  | No                                   | 2 (1 to 3)                               | Yes                   |
| Cox         | No                        | No          | 0                         | Isosulfan blue 1%  | Yes (58%)                            | 6 (2 to 11)                              | Yes                   |
| Bilchik     | Yes                       | No          | ns                        | Isosulfan blue 1%  | No                                   | 1.75 (ns)                               | Yes                   |
| Turner      | Yes                       | No          | ns                        | Isosulfan blue 1%  | No                                   | 3 (ns)                                   | Yes                   |
| Trocha      | No                        | No          | 0                         | Technetium and blue dye | No                                  | 2.5 (ns)                                 | Yes                   |
| Veilh       | No                        | Yes         | 0                         | Isosulfan blue 1%  | No                                   | 2 (1 to 8)                               | Yes                   |
| Levine      | No                        | No          | 0                         | Isosulfan blue 1%  | No                                   | 1.9 (1 to 6)                             | No                    |
| Saha        | Yes                       | No          | ns                        | Isosulfan blue 1%  | No                                   | 2.1 (ns)                                 | Yes                   |
| Dan         | No                        | No          | 0                         | Isosulfan and fluorescein | No                                  | 2.5 (ns)                                 | Yes                   |
| Braat       | No                        | No          | ns                        | Patent Blue        | Yes (57%)                            | 1.7 (1 to 4)                             | Yes                   |
| Bertoglio   | No                        | No          | 0                         | Vical Blue         | No                                   | 2.9 (1 to 3)                             | Yes                   |
| Read        | No                        | No          | 0                         | Isosulfan blue 1%  | No                                   | 2 (ns)                                   | No                    |
| Patten      | No                        | No          | 0                         | Technetium and blue dye | No                                  | 3.5 (0 to 11)                             | Yes                   |
| Bertagnolli | No                        | Yes         | 0                         | Isosulfan blue 1%  | No                                   | 2.1 (ns)                                 | Yes                   |
| Saha        | No                        | No          | 0                         | Technetium and Isosulfan blue | No                                  | 3 (1 to 4)                               | Yes                   |
| Saha        | Yes                       | Yes         | ns                        | Isosulfan, Technetium and fluorescein | No                                  | 2 (1 to 4)                               | Yes                   |
| Bembenek    | No                        | No          | 0                         | Patent Blue        | No                                   | 2 (ns)                                   | Yes                   |
| Codognola   | No                        | No          | 0                         | Patent Blue        | No                                   | 2.02 (ns)                                | Yes                   |
| Dahl        | No                        | No          | 0                         | Patent Blue        | Yes (6%)                              | 2.2 (0 to 6)                             | Yes                   |
| Bilchik     | Yes                       | Yes         | 15                        | Isosulfan blue 1%  | Yes (4%)                              | 3 (ns)                                   | Yes                   |
| Tuech       | Yes                       | No          | ns                        | Patent Blue        | Yes                                   | 1.5 (ns)                                 | Yes                   |
| Saha        | Yes                       | Yes         | ns                        | Isosulfan blue 1%  | No                                   | 2.2 (ns)                                 | Yes                   |
| Kelder      | No                        | No          | 0                         | Patent Blue        | No                                   | 2.7 (1 to 4)                             | Yes                   |
| Thomas      | No                        | No          | 0                         | Isosulfan blue 1%  | No                                   | 2.1 (1 to 5)                             | Yes                   |
| Covarelli   | No                        | No          | 0                         | Technetium and blue dye | No                                  | 1.3 (ns)                                 | Yes                   |
| Kelder      | Yes                       | Yes         | 0                         | Patent Blue        | No                                   | 2.3 (ns)                                 | Yes                   |
| Bianchi     | No                        | No          | 100                       | Patent Blue        | Yes (5%)                              | 2 (ns)                                   | Yes                   |
| Murawa      | No                        | No          | 0                         | Patent Blue        | No                                   | 1.6 (0 to 4)                             | No                    |
| Bembenek    | No                        | Yes         | 7                         | Patent Blue        | No                                   | ns                                       | Yes                   |
| Sandrucci   | No                        | No          | ns                        | Patent blue and technetium | No                                  | 2.2 (ns)                                 | Yes                   |
| Tiffet      | No                        | No          | 0                         | Patent blue and technetium | No                                  | 2.6 (ns)                                 | Yes                   |
| Lim         | No                        | No          | ns                        | Technetium and Isosulfan blue | Yes                                   | 4 (ns)                                   | Yes                   |
| Kusano      | No                        | No          | 62%                       | Indocyanine green  | No                                   | 2.6 (0 to 5)                             | No                    |
| Quadros     | No                        | No          | 0                         | Technetium and patent blue | Yes                                   | 3.5 (ns)                                 | Yes                   |

The order and format is the same as in Table 3.
Of the sixteen studies with false negative rates above 20%, twelve presented no critical analysis of their false negative rates. Nor did any of these studies place any emphasis on surgeon experience in their stated inclusion criteria. Four studies were performed on a multicentre basis but none explicitly ensured surgeon expertise prior to commencement (in one such study the mean number of operations per surgeon was less than three) and nine studies included non-colonic tumors in between 10 and 26% of the cohort size. Furthermore one study specifically commented that tumors adherent to the retroperitoneum were included while five had T3/4 tumors accounting for approximately 80% of their patient cohorts. Three had a mean tumor size of greater than 4.2 cm with one concluding that its false negatives case were associated with significantly bigger tumor sizes (7.2 cm in the false negative cases versus 5.2 cm mean size overall). One additional study also found a strong trend in favor of an association between false negative rate and larger tumor sizes (4.5 cm v. 3.5 cm, p = 0.09). Two studies had a mean number of resected lymph nodes of eight and nine respectively while at least twelve studies included some patients with less than ten nodes in their resected specimens (eight including some with five or less nodes examined). Two others presented no mean data on this subject and three presented no range. Four did not serially section the sentinel node while six did not employ immunohistochemistry or RT-PCR.

With respect to the fifteen publications with false negative rates between 10 and 20%, only three studies included specific analysis of their rates. Of the fifteen, two were multicentric trials and both sought to ensure surgical expertise and had false negative rates each of 11 and 12%. Nine studies included rectal cancers in their cohorts (in between 9% and 74% of their cohorts). Four studies had no explicit exclusion criteria while six sought only to exclude distant disease deposits. No study provided any data on patient BMI. Only one study specifically excluded T4 disease. Eight had a significant (>60%) T3/4 preponderance (being >70%). The mean number of resected lymph nodes was greater than ten in fourteen. Four studies included patients with less than this number while the one presented no data regarding the range. Two studies did not employ serial sectioning of the identified sentinel nodes and two did not utilize immunohistochemical or RT-PCR methods of examination.

**Discussion**

Sufficient lymph basin resection is an oncologic sine qua non qua of operation with curative intent for solid organ malignancies as nodal status remains the primary portent of prognosis and adjuvant therapy prescription. For colon cancer, staging propriety by convention demands that this equate to en bloc resection of the entire mesenteric lymphatic delta. While the manner of performance of standard resectional operation for colon cancer by laparoscopy or laparotomy means that the extent of access is already determined (and so supplementing bowel resection with full mesenteric resection is readily facilitated) this is not for case for endoscopic resectional techniques. Therefore these techniques are currently limited to the address of benign lesions or neoplastic disease with minimum likelihood of lymphatic dissemination. A minimally invasive means of reliably confirming the lymph node status could greatly enhance the oncological propriety of these approaches and extend their indications towards their actual technical capacity. While it is clear that sentinel node biopsy is not indicated to minimize the extent of therapeutic lymphadenectomy in colon cancer (i.e. the resection of nodes containing tumor deposits), it could theoretically have a role in helping select those with truly early stage disease for endoscopic resection.

Furthermore, although it is often considered that adjoining mesenteric lymph node dissection to the intestinal resection impacts little on the patient undergoing conventional oncological operation for colon cancer [100-103], this assumption does not necessarily stand up to close scrutiny. The high-tie of the arterial inflow arcade necessary to adequately perform mesenteric nodal dissection however segues wider longitudinal intestinal margins than would otherwise be necessary for local clearance purposes alone. The negative impact of wide dissection and resection may include post-operative bowel dysfunction, nerve plexus praxia or adjacent organ injury [104-106]. Therefore, tacit knowledge, as well as some prior clinical investigation [107-109], would suggest that minimizing the extent of mesenteric dissection could pay dividends for the patient even if standard surgical access is employed. These considerations have however been lost amid the concerns regarding the efficacy of lymphatic mapping in this disease and the current emphasis on ensuring adequate staging by resection of considerable ‘minimum’ numbers of lymph nodes [110-112].

The focus of sentinel node biopsy in colon cancer has therefore been on upstaging conventionally node negative patients after conventional radical operation[113]. This means that false-negative sentinel nodes do not impose any negative effect on the patient but also has conferred a priori license to investigators to modify the technique and broaden the patient cohort. To propose that it could be used a means of predicting a negative basin and obviating mesenteric dissection places great emphasis on ensuring that the technique’s rationale in colon cancer is biologically sound and that methodological discrepancies are eliminated. However the literature base to date contains multiple un-standardized variables and so assessment of the true utility of lymphatic mapping when applied selectively to early stage disease is considerably confounded.
Figure 2
Spread of (a) Detection rates and (b) False Negative Rates among the studies of in vivo sentinel node mapping in colon cancer included in this review. In the figures, studies arising from either of the two centers with the most publications (ie The John Wayne Cancer Institute, California and the McLaren Reginal Cancer Centre, Michigan) on the topic are indicated separately to minimize any visual bias resulting from their inclusion. The remaining studies are divided into multi-center studies and all others.
The high accuracy rates in some studies do at least suggest that its concept is biologically tenable (indeed of those centers that have published multiple series, four conclude strongly in favor of the reliability of the technique) and that methodological variability may underlie the discordant results (as they do for the same technique in malignancies at different sites). However, this heterogeneity also confounds any attempt at meta-analysis to selectively extract data regarding the reliability of sentinel node biopsy selectively in early stage cancer. A similar conclusion was also reached by another group performing a detailed statistical systematic review of the literature from another perspective[114]. Although their review contains significant differences in study inclusion and exclusion criteria to ours, it is interesting that they nonetheless concluded that the technique had a 9% median risk of a false negative result in comparison to an 8.4% median risk for the technique in breast cancer. However, the general tendency to pool results in the reporting of studies frustrated their attempt to relate T-stage to sensitivity.

The first evident confounder to obtaining clarity regarding the accuracy of the technique is the number of publications emanating from single centers. Although it seems likely that these seven centers (who together have contributed over half of the published evidence base) have overlapped their patient cohorts at least to some extent in successive publications, the exact proportions is rarely explicitly declared. Equally however it cannot be assumed that these studies entirely overlapped their experiences and so to ensure fairness and transparency every study meeting the inclusion exclusion is included in this study with those coming from the same center being flagged in the Tables. The next main obstacle within each publication complicating deliberation of the technique’s applicability for early stage colon cancer is the marked contamination of rectal cancers throughout the evidence base. The consequences of doing so without presentation of complete subgroup analyses gives an artificial impression of reduced feasibility and accuracy rates overall because lymphatic mapping is clearly more arduous and less reliable in this site[115]. Furthermore, specific account of tumor size and mural penetration in colon cancer is too often lacking for any firm conclusions to be made regarding the efficacy of selectively mapping early stage disease. The main inclusion criterion to date is primarily the requirement for operation for symptomatic colon cancer despite the fact that the disease tends to present late (often at stage when curative surgery is not possible and adjuvant treatment is already indicated regardless of the nodal status). An elongated diameter of the cancer may also mean that different segments of the tumor map to different nodes as watershed areas are crossed. Furthermore, direct spread through the bowel wall may compress and alter the original lymphatic flow and so cause the mapping agent to deviate from the

| Authors      | Year | Comment                                                                                                                                 |
|--------------|------|-----------------------------------------------------------------------------------------------------------------------------------------|
| Bendavid     | 2002 | The one false negative case occurred in a patient with liver metastases. Also ‘evidently metastatic nodes’ did not receive colourant       |
| Paramo       | 2002 | No specific analysis presented                                                                                                         |
| Wood         | 2002 | All five false negatives occurred in T3 or T4 tumors (in one case the only positive non-sentinel node was involved by direct extension). Three occurred in 1st 30 cases |
| Bilchik      | 2002 | All five false negatives occurred in T3 or T4 tumors. Three occurred in the first fifty cases.                                          |
| Kitagawa     | 2002 | Four false negative cases were advanced T3 and/or had massive lymph node metastases                                                    |
| Feig         | 2002 | Also ‘several patients’ (of ten) classified as false negative had ‘palpable lymph nodes’                                              |
| Broderick-Villa | 2002 | Learning curve strongly associated with false negative rate (67% in first half, 32% in second half). No significant association with T-stage, LN involvement or tumor diameter > or < 4 cm |
| Veihl        | 2003 | Amount of dye relative to tumor size was an important predictor of identification of node. False negative more common in cases with larger nodes (4.5 cm v 3.4 cm, p = 0.09) |
| Bilchik      | 2006 | Of the six false negatives, four were attributable to tumor obliteration of the lymphatic channels                                        |
| Saha         | 2006 | 95% of patients with skip metastases were T3 or T4                                                                                     |
| Thomas       | 2006 | Two patients with liver metastases along with two others with gross mesenteric disease had false positive sentinel nodes. No relationship between BMI and disease |
| Kelder       | 2006 | In one of the two false negatives, the non-SLNs were involved by extra-nodal tumor invasion                                              |
| Bembenek     | 2007 | Significant association with learning curve/center experience, BMI (cut-off level being 22 patients and a BMI of 25 respectively) & LVI. No significant association between detection and T stage, age, sex, vascular invasion, no of nodes, total no of nodes. |
| Sauranucci   | 2007 | 'Skip metastases' were all correlated with 'T2 lesions with massive lymphatic involvement'                                           |
| Tiffet       | 2007 | Three of 12 false negatives were in patients with direct tumor involvement of adjacent non-sentinel epicolic nodes while four were in N2 patients. False negative rate markedly lower in the subgroup with T1 and T2 tumors only. and in those with BMI < 30 kg/m² |
original first order draining nodes. Extensive nodal involvement, including total destruction of the nodes by metastases, may also compromise the identification of true sentinel nodes[22,116]. Such concerns have been confirmed in the analyses presented here as well as in ex vivo experiences of mapping colon cancer and indeed in other tumors [117-119].

Surgeon expertise and experience has already been determined a major feature for lymphatic mapping in breast cancer[120] and seems likely to also be one for colonic mapping (a point made clear in one study attempting ex vivo biopsy that found many of their failed mapping were associated with intraluminal dye injection)[39]. The relationship between low detection rate and higher false negative rate is a likely related phenomenon while lack of technical proficiency may in addition compound any technical difficulties associated with adverse patient factors (such as BMI or previous laparotomy) to further undermine the performance of the technique. A specific learning curve can also certainly be expected if the surgeon looks for the nodes intraoperatively. However the lack of specific inclusion criteria for this seems to suggest that many of the studies published to date are in fact reflections of expertise acquisition rather than reports of technique reliability. The timing of the vital blue dye injection prior to resection may also be a crucial issue. A prolonged latency in identifying the node (depending on the intraoperative situation) may allow overflowing of the dye and potentially complete decoloration of the first stained node. Also the discrepancy in standardizing nodal analysis between the reports is concerning as standard assessment based on hematoxylin and eosin staining of one level of a paraffin-embedded block reportedly can miss as many as 33% of metastases[121]. While micrometastatic disease may of itself be of considerable clinical importance [115], these cells in sentinel nodes may also confer significance by their prediction of the presence of non-sentinel nodal metastases. Finally the numbers within the nodal harvest rates of many studies raises serious concern over quality assurance utilized given that this aspect is of utmost importance when a novel technique is compared to established practice.

Certain groups have however clearly managed to overcome confounding factors of the technique and consistently obtain negative predictive values of similar quality to those that currently justify conservative resection fields in other specialties. It is perhaps no surprise that these investigators tend to fastidiously analyse their false negative results as did the pioneers of the technique in breast cancer and melanoma. On the other hand, other authorities have not hesitated to declare the technique in colon cancer either invalid or of dubious clinical value[40,122] on the basis of their own experience and perception of the evidence base. Most of the latter group did not however present data detailing their efforts to examine the potential biological reasons behind their results. As much therefore as it is clear that certain proponents in both single centre and multicentre trials can perform sentinel node biopsy with high accuracy for colonic malignancy, it is equally clear that others cannot. The fact that advanced endoscopic resection is at present only wrought in superspecialized centers may mean however that only such selected, expert departments need attempt to investigate, validate and standardize the performance of sentinel node biopsy to accompany it. Also, the need for high detection and accuracy rates implicit if the results were going to impinge on patient care could drive efforts to develop methods to improve the yield of lymphatic mapping by use of additional or alternative dyes (perhaps fluorescent markers[73,123] or radioisotopes[69,78]) as have been used for other cancers [124-126].

The general tendency of a relationship between advancing stage an increased likelihood of metastases being present in non-sentinel nodes also supports the basic contention that lymphatic mapping in colon cancer may be particularly efficacious in germinal cancers – a perspective made particularly compelling by the inherent suitability of early stage disease for truly minimally invasive resective techniques. The lesions that could be resected by endoscopic means are by definition smaller and therefore likely confined to a single lymphatic delta. Furthermore T3 or T4 disease identifiable by staging is not feasibly resected endoscopically and so the tumor stages with the highest frequency of being node-positive are excluded. The fact that 20–40% of patients with T3/4 lesions but without demonstrable lymph node metastases actually die of their disease also supports exclusion of these patients from non-radical operation. De facto confinement of the patient cohort to T1 and T2 stage disease (perhaps by including adjunctive staging measures such as endoscopic ultrasound[127]) may therefore obviate some of the main confounders of lymphatic mapping and potentially allow offering of localized tumor resection as definitive surgery. However the lack of specific study or discriminating subgroup analysis by T-stage means that this aspect of lymphatic mapping in colon cancer has yet to be definitively explored and so this remains speculative.

If sentinel node biopsy is ever to be used as means to alleviate mesenteric resection in cases that are truly node negative, consideration must be given to the cases where the sentinel node is positive or indeed falsely negative. In the former case, subsequent radical lymphadenectomy should still be performed. Ideally therefore the sentinel node analysis should be performed intraoperatively to allow direct progress to the definitive excisional surgery (whether of the primary alone or of the traditional
extended operation). Considerable precedent exists for such analysis in breast cancer [128-131] and it is even possible that additional innovative technology may allow for in situ virtual biopsy of the salient nodes [132,133]. In the situation that the sentinel node is falsely negative, suspicious features present in the resected specimen may encourage revisional surgery in some cases. Although re-intervention after a previous laparotomy may be difficult, the risk of added morbidity because of a prior minimally invasive ‘cherry-picking’ of sentinel nodes is likely to be less. However any true consideration of the risk involved needs balancing with an accurate measure of the potential benefit of avoiding mesenteric resection. The consequence of missed positive lymph nodes is mainly one of potential understaging and hence depriving the patient of systemic chemotherapy. However it should also be realized the added benefit of chemotherapy over standard surgery for T1 and T2 lesions is low and perhaps that the clinical significance of minimal nodal disease in these patients is lower than that of more advanced T-stage (as is the case for early gastric cancer [134]). The actual clinical risk of detriment due to mesenteric lymphadenopathy per se is also low [85]. It is worth however here emphasizing again that these deliberations only apply to colon cancer and not its counterpart in the rectum. The anatomic characteristics of the mesorectum (namely its bulk, extraperitoneal situation and lack of a serosal layer) mitigate against selective sentinel node biopsy in isolation for rectal cancer however as does the fact that transgression of its planes may complicate any subsequent total mesorectal resection in cases of nodal positivity and so impact upon the potential for curative resection. Finally the consequences of missed nodal disease in the pelvis and, therefore the occurrence of pelvic recurrence, may be catastrophic for the patient.

Conclusion
Sentinel node mapping could never substitute for a properly performed oncologic colorectal resection when this is indicated. The concept however that lymphatic mapping may have sufficient capability to provide the oncological propriety for the curative surgery for early stage cancers without en bloc mesenteric resection seems biologically plausible but cannot yet be definitively judged given the lack of clarity and consistency in the literature to date. Specific study of the technique in those early stage tumors likely to be selected for endoscopic resection is clearly therefore essential before this approach can be considered in clinical practice.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
RAC conceived of the study, participated in the study design and performance (performed the initial literature review and analysis) and composed the manuscript. JL contributed to the study design and performance and guided the manuscript composition. JM conceived of the study and participated in its design and coordination. All authors read and approved the final manuscript.

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