Influence of foliar application of algae extract and nitrogen fertilization on yield and quality of sugar beet grown in reclaimed sandy soil

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Abstract
The current examination was done at North- Eltahrer, Alnubaria Albuhaira Governorate, during 2018/19 and 2019/20 seasons. The field analyze was led to consider the impact of foliar application with alga extricate (Spirulina platensis) and Nitrogen fertilization on creation and nature of sugar beet (Beta vulgaris var. MK4200) was planted in a sandy soil under sprinkler water system framework. A split – split plot design with three replicates was utilized in the two seasons. Nitrogen focus was masterminded in the primary plots, while Algae were allotted in the sub plot. the Results demonstrated that foliar utilization of alga extract using1.0 g/l or 2.0 g/l produced significantly higher values of photosynthetic pigments), vegetative growth traits of sugar beet plants (root diameter, root and foliage fresh weight), (root and top dry weight/plant) and yield and yield components (root and top weight (g/plant and kg/fed), Sugar yield and quality traits. Higher estimations of development ascribe, and yield and its segments were gotten with adding 120 N kg/fed. The interaction between alga concentrates and Nitrogen fertilization altogether influenced complete photosynthetic pigments, all development characters, yield and yield components. The blend of 2.0 g/L alga extracts alongside 120 N kg/fed. can be prescribed to get the most extreme root yield/fed. Sugar yield was essentially and emphatically corresponded with Sucrose% followed by juice purity% and Brix% as well. Negative relationship was identified between juice purity%, and brix%.

Keywords: Spirulina platensis; Nitrogen fertilizer; Sandy soil; Sugar beet.

INTRODUCTION
Sugar beet is one of the most basic economic crops. The proceeded with creation of this significant ware requires more harvest the board methods. Moreover, the expansive leaved yield builds soil richness, particularly in recently developed soils which, portrayed with low of natural issue and water holding limit just as high supplement misfortunes by filtering; which prompts show lack manifestations of large scale and miniature supplements (Shafeek et al., 2013). Green growth separate being natural and bio-degradable in nature is considered as a significant wellspring of sustenance for practical horticulture particularly in the recently recovered soil. Synthetic investigation of green growth extricates and have uncovered the presence of a wide assortment of plant development controllers, for example, auxins and cytokinins in differing sums (Zhang and Ervin, 2004). In this way, green growth
separate invigorate root foundation, root prolongation and advance vegetative development of plants. Foliar use of green growth extract has been accounted for to actuate numerous beneficial outcomes, where showering plants had prompted improve crop yield and quality, increment supplement take-up, protection from ice and stress conditions. Green growth was early considered as a significant gathering of microorganisms fit for fixing air nitrogen, just as it causes critical expansion in root development, fresh and dry weight of roots, complete biomass, yield segment, photosynthetic pigments and development advancing hormones (Ghalab and Salem, 2001). Because of the useful movement, there is an expansion of photosynthetic contraption through raising the substance of all out sugars, starch, amino acids and protein (Raupp and Oltmanns, 2006) and (Yassen et al., 2007). *Spirulina platensis* is a rich wellspring of potassium and contains impressive measures of Ca, Cu, Fe, Mg, Mn, P and Zn, consequently it expands take-up and gathering of these components in plants. This thus clarifies the critical increment of vegetative development and yield and its parts just as substance of nitrogen, phosphorus and protein in leaves additionally leaf chlorophyll content for most harvests particularly those become under semiarid and desert conditions (Abd El-Mawgoud et al., 2010) and (Marrez et al., 2014).

Nitrogen is the main agronomic variable known to influence sugar beet yield and quality and it ought to be figured out how to deliver high root weight with high Sucrose focus and virtue levels with insignificant top development (Seadh et al., 2007). A satisfactory flexibly of nitrogen is fundamental for ideal yield and the financial yield is firmly identified with the sugar amassing measure. (Clinton et al. 2000) revealed that nitrogen is the main manure component for sugar beet development and yield, and they added that the option of N up to 100 kg N/prevailing fashion. Significantly improved length, diameter and weight of sugar beet roots in any case the best Sucrose yield was acquired with 120 kg/ha and ideal N treatment rates for sugar beet rely upon the locale. Expanding N level up to 100 kg N/fed. Discouraged essentially Sucrose and virtue rates (El-Harriri and Gobarah2001) and (Badr et.al., 2017). Expanding N levels from 60 to 120 kg N/prevailing fashion. Influenced fundamentally root length, width and new weight/plant, root sugar yields. Something else, Sucrose and virtue rates were not influenced by the used N levels (Ismail 2002). (Ramadan et al. 2003) demonstrated that Sucrose and immaculateness rates just as recoverable sugar content altogether diminished with expanding N application from 0 to150 kg N/prevailing fashion. in sandy soil. Then again, contamination credits [Na, K and α-amino N (meq/100 g)] and Sucrose misfortune to molasses altogether expanded with raising N levels up to 150 kg N/fed. (Azzazy 2004) announced that root length, root and top yields and TSS were essentially expanded as the applied N level was expanded from 60 up to 100 kg N/fed. while, Sucrose and virtue rates were essentially diminished. (Ismail and Abo EL-ghait 2005) showed that expanding N levels from 65 to 112 kg N/ha-1 improved root yield and its segments, while, yield and quality (all out solvent solids, Sucrose and immaculateness rate) were diminished. (Montemurro and Maiorana 2007) revealed
that adding 250 kg N ha⁻¹ created the most elevated estimations of length, distance across and new weight of roots just as root, top and sugar yields ha⁻¹. Additionally, (Stevens et al., 2007) demonstrated that surplus N lessens the extent of the sugar which can be solidified and amount of N application as the viable factor to upgrade sugar yield and quality. From ecological scene the inordinate nitrogen treatment may bring about a critical reduction in NUE and likely contamination of climate which lessen the monetary advantage of horticulture creation and human wellbeing (Zhenxie et al., 2008). Expanding N application decreased nitrogen use productivity (NUE) in sugar stick (Koochekzadeh et al., 2009). Principal part (PC) examination that was utilized to clarify most of the complete variety (Abo Elenen, et al., 2019). Gomaa et al. (2017) reported that nitrogen fertilizer application at a rate of 100.5 kg N/ fed. accompanied with 48 kg K2O/ fed. was found to be the most favorable for improving the yield and quality of sugar beet grown in a sandy soil.

The goal of this analysis to contemplate the impact of foliar application with alga extract (Spirulina platensis) and Nitrogen fertilization on creation and nature of sugar beet (Beta vulgaris var. MK4200).

MATERIALS AND METHODS

Site Description

Two field trials were directed at The newly reclaimed lands served by Nubaria Agricultural Research Station, is located at the East side of Cairo-Alexandria desert road, at Al-Naser Central village, North Tahrir, El-Beheira Governorate, Egypt, Throughout 2018/19 and 2019/20 sugar beet growing seasons to study the effect different rates of Nitrogen fertilization and foliar spray of Spirulina platensis filtrate on yield components and technological characters of Sugar Beet (Beta vulgaris var. MK4200), was planted in a sandy loam soil under surface irrigation regime.

Former to any soil preparation of the experimental site, a multiple soil sample was gathered from the soil depth of (0-30 cm), air-dried, sieved by 2 mm sieve and analyzed. The physical and chemical properties of soil as illustrated in Table (1), were determined according to Klute, 1986 and Page et al., 1982.

Plant Material

Seeds of the tested sugar beet varieties were commercial and obtained from the under certification pool of sugar beet varieties hold by Sugar Crops Research Institute (SCRI). Beta vulgaris subsp. vulgaris was polygerm variety, MK4200

The Algae, Spirulina platensis and preparation for the study

S. platensis strain was isolated from Al-Khadra Lake, Wadi Al-Natroon, El-Baheira governorate, Egypt. Isolation, purification, Morphological identification and production to massive scale were performed at Algal Biotechnology Unit, National Research Center. Homogenate was prepared by a suspension of dry S. platensis in deionized water (in a ratio 1:10) and mixing (Thermomix TM5; USA) at 37 °C for 40 min. (500 rpm). The obtained solution was centrifuged for 20 min. (4600 rpm) (Heraeus Megafuge 40, rotor TX-750, Thermo Scientific, and Waltham, MA, USA). Supernatant was separated and treated as an algal filtrate (AF)—100%. Three samples were taken for determination of Macrominerals and trace elements by Algal Biotechnology Unit, National Research Center.

Samples were irradiated, together with standard reference materials; two standard reference materials were used: the European
Inter-Institutes Committee (CII) “Alfa-Alfa” and the Bureau Communautaire de Référence “Lucerna Flour”. Major components of the used alga filtrate are appeared in Table 2.

Table 1. Some physical and chemical properties of the experimental soil site during 2018/2019 and 2019/2020 sugar beet growing seasons.

| Soil parameters | Seasons          |
|-----------------|------------------|
| Particles size distribution % | 2018/2019 | 2019/2020 |
| Sand            | 58.41            | 56.10     |
| Silt            | 11.00            | 12.00     |
| Clay            | 30.59            | 31.91     |
| Textural class  | Sandy loam       | Sandy loam|
| Chemical properties |          |          |
| PH (1:1) (soil: water suspension) | 7.91     | 8.10     |
| EC (1:1) (soil: water extract), dS/m | 3.71     | 3.79     |
| Soluble cations (1:2) (meq/l) |          |          |
| K⁺              | 0.46             | 6.59      |
| Ca²⁺            | 0.61             | 8.30      |
| Mg²⁺            | 3.49             | 4.11      |
| Na⁺             | 12.11            | 12.49     |
| Soluble anions (1:2) (meq/l) |          |          |
| HCO₃⁻           | 5.98             | 6.22      |
| Cl⁻             | 7.91             | 8.41      |
| SO₄²⁻           | 4.41             | 4.11      |
| Calcium carbonate, % | 12.27 | 12.91 |
| Total Nitrogen, % | 0.77     | 0.87      |
| Available P (mg/ kg) | 4.52   | 4.61      |
| Organic matter, % | 1.51     | 1.60      |

Experimental design

The accomplished work included forty eight experimental units as a factorial arranged split plot design with Nitrogen fertilizer rates (Without, 60, 90 and 120 kg/fed.) in main plots and Algae, *Spirulina platensis* was sprayed as foliar application at four filtrate concentrations (Without, 0.5, 1.0 and 2.0 g/L/400 liter water/fed) were allocated in the subplots, Mineral fertilization consists of nitrogen as in form of ammonium nitrate (NH4NO3 33.5 %), with Three replicates were used. subplots consisted of six rows (50 cm spacing) by 7.0 m in length (3 m ×7.0 m = 21.0 m²) i.e. 1/200 Fed. Sugar beet plants were cultivated on 14th Oct. in both investigated seasons. The experimental setup was repeated for the successive trial in 2018/19 and 2019/20.

All recommended agriculture practices for sugar beet cultivation other than experimental treatments needed for growing as sowing, irrigation, hoeing and pest control were taken whenever it were required and accomplished according to the approvals of Ministry of Agriculture, Egypt.

Table 2. Macrominerals and trace elements contents of the The Algae, *Spirulina platensis*. *Elements determined by Algal Biotechnology Unit, National Research Center Values are the mean (RSD in brackets) of five determinations.
### Elements Sample1 Sample2 Sample3 Mean *a*Refs

| Elements | Sample1 | Sample2 | Sample3 | Mean  | *a*Refs |
|----------|---------|---------|---------|-------|---------|
| N        | 11.2    | 12.3    | 11.8    | 11.8  | 9.3–24.42 |
| P        | 1.65    | 1.54    | 1.6     | 1.6   | 1.3–9.21.3 |
| K        | 0.900 (0.141) | 0.88     | 1.0     | 0.9   | 0.233–1.9361.4 |
| Mg       | 0.067 (0.021) | 0.22     | 0.14    | 0.1   | 0.120–0.5451.4 |
| Na       | 0.35 (0.6) | 0.01     | 0.18    | 0.2   | 0.027–5.431.4 |
| Ca       | 0.432 (0.005) | 0.33     | 0.381   | 0.4   | 0.042–2.610 **+** |
| Fe       | 1176 (25) | 1936    | 1553    | 1555.0 | 220–20161.4 |
| Zn       | 18.5 (0.4) | 21       | 19.8    | 19.8  | 18.4–1851.4 |
| Mn       | 54.5 (0.4) | 68       | 61.3    | 61.3  | 13–2051.4 |
| Cu       | 16.2 (1.1) | 18       | 17.3    | 17.2  | 2.6–4801.3 |

*a* References: 1. (Ortega-Calvo et al., 1993); 2. (Dillon et al., 1995); 3. (Mannino and Benelli, 1980); 4. (Johnson and Shubert, 1986).

### Data Recorded:

The outer two ridges were considered as a belt, while plants of the 2nd ridge was used for growth analysis. The other ridges, 3rd, 4th and 5th were kept for determination of yield, yield components and technological quality.

For determining growth analysis, a random sample of five plants was taken from each plot of every replication after 6 months (at harvest) from sowing during both seasons to determine the following parameters:

#### A- Growth characters:

1. Root length (cm).
2. Root diameter (cm).
3. Root fresh weight (g/plant).
4. Leaf fresh weight (g/plant).

#### B- Yield components:

1- Root yield (Ton/Fed.)
2- Top yield (Ton/Fed.)
3- Sugar yield (Ton/Fed.)

#### C- Quality traits

Saccharometer according to Le-Docte (1927). Total Soluble Solids (TSS %) was determined by using digital referactometer. Juice purity percentage (%) was determined according to the method of Carruthers and Oldfield (1961). Potassium (K) and Sodium (Na) were measured in the root dry weight at harvest time, by using the Flame photometer. α-amino nitrogen was also calculated by double beam filter photometry using the blue number method (Sheikh_Aleslami 1997).

**Sucrose loss to molasses (SLM)**

\[
\text{SLM} = 0.343*(K+Na) + 0.0393* \alpha-\text{ amino N} + 0.31
\]

Extractable sugar % was calculated using the following equation according to (Cooke & Scott 1993):

\[
\text{Quality} = \text{QZ} = (ZB \times 100)/ (\alpha-\text{ amino N})
\]

**Extractable white sugar % (ZB %)**

\[
\text{ZB} = (\text{Pol} - 0.29) - 0.343 \times (K + Na) - \alpha-\text{ amino N} (0.0939).
\]

Where: K, Na and α-amino N were determined as meq/100 g beet.

### Statistical analysis:

The obtained data of the two seasons and their combined analysis were computed according to Snedecor and Cochran (1980). Combined analysis between two seasons were calculated according to Mc Intosh, M. S. (1983). The treatment means were compared by using the least significant difference (L.S.D) test (Waller and Duncan, 1969) at 5% level of significance.

### Results and Discussion

#### A-Growth characters:

A-1-Effect of nitrogen fertilizer
Information introduced in Table (3) showed that increasing application Nitrogen fertilization rates from 60 to 120 kg N/fed. was related with critical expansions in completely contemplated characters’ root length and diameter (cm) and fresh weight (root and leaf). The most noteworthy estimations of previously mentioned characters were gotten with the expansion of 120 kg N/fed. compared with other treatments. These outcomes affirm that the function of nitrogen in division just as building natural metabolites which thusly trans situated to be put away in sugar beet roots. It could be inferred that high rates of nitrogen application empower creation of new vegetative organ which uncovered an articulated augmentation in leaf yield. Comparable outcomes were acquired by (Montemurro and Maiorana 2007), (Badr 2006) who expressed that expanding Nitrogen fertilization 100 kg N/fed. of delivered the most elevated development qualities. Badr et al. (2017) development ascribes essentially expanded by expanding Nitrogen fertilization levels from 40 to 80 and 120 kg N/fed.

**Table 3.** The effect of foliar application with alga extract and nitrogen fertilizer on growth characters of sugar beet (combined analysis of the two seasons 2018/2019 and 2019/2020).

| Treatment     | Root length (cm) | Root diameter (cm) | Root fresh W.(gm) | Leaf fresh W.(gm) |
|---------------|------------------|--------------------|-------------------|-------------------|
| 60 Nkg/fed   | 28.02            | 29.26              | 765.58            | 439.92            |
| 90Nkg/fed    | 32.76            | 32.87              | 848.67            | 499.75            |
| 120Nkg/fed   | 36.98            | 37.11              | 973.42            | 605.33            |
| **LSD 0.05** | **0.35**         | **0.42**           | **13.65**         | **7.67**          |
| Control      | 30.64            | 31.70              | 820.84            | 480.89            |
| 0.5 g/L      | 32.32            | 32.64              | 851.44            | 503.78            |
| 1.0 g/L      | 33.32            | 33.39              | 875.11            | 526.44            |
| 2.0 g/L      | 34.06            | 34.58              | 902.78            | 548.89            |
| **LSD 0.05** | **0.26**         | **0.12**           | **6.90**          | **2.08**          |

**A-2-Effect of foliar application with alga extract**

Information in Table (3) showed that a critical impact of alga extract on root diameter, length root and root, leaf fresh weight/plant. Splashing beet plants with (2 g/L) Spirulina extract brought about higher estimations of root thickness just as new weight of leaves and roots when contrasted with different levels. The obvious impact of alga concentrate might be credited with its impact in expanding cell layer penetrability and advancing plant productivity in the retention of supplements, for example, nitrogen, which has an immediate connection with leaf chlorophyll fixation. In addition, green growth concentrate may assume a part through its substance of cytokinins in postponing the maturing of leaves by decreasing the debasement of chlorophyll. Likewise, alga extricate as a bio-controller influencing the harmony among photosynthesis and breath measures in plants (Raupp & Oltmanns, 2006, Yassen et al., 2007 and Enan et.al 2016).

**A-3-The Interaction**

The interaction between foliar application with alga extract and Nitrogen fertilization significantly affected growth characters. The results in Table (4) showed that were responded to the increase of foliar applied
Spirulina extract level, but was highly magnitude by adding 120 kg/N. Therefore, the highest value of growth characters recorded by 120 kg/N and 2 gm/L alga extract finding may be due to algae extract contains cytokines which induce the physiological activities and increase total chlorophyll in plants which, reflects on the activity of photosynthesis and the synthesized materials which will positively reflects on the growth characteristics (Ghalab & Salem, 2001) and (Enan et al.,2016).

**Table 4.** The interaction effect of foliar application with alga extract and Nitrogen fertilization on growth characters of sugar beet (combined analysis of the two seasons 2018/2019 and 2019/2020).

| Treatments  | Root length (cm) | Root diameter (cm) | Root fresh w.(gm) | Top fresh w.(gm) |
|-------------|------------------|--------------------|-------------------|------------------|
| **60 N kg/fed** |                  |                    |                   |                  |
| Control     | 25.83            | 28.03              | 725.67            | 410.33           |
| 0.5 g/L     | 27.73            | 28.87              | 759.67            | 438.00           |
| 1.0 g/L     | 28.80            | 29.57              | 782.33            | 450.00           |
| 2.0 g/L     | 29.70            | 30.57              | 794.67            | 461.33           |
| Control     | 30.50            | 31.57              | 814.38            | 476.44           |
| **90 N kg/fed** |                  |                    |                   |                  |
| Control     | 32.57            | 32.47              | 838.00            | 487.00           |
| 0.5 g/L     | 33.57            | 33.03              | 859.33            | 505.00           |
| 1.0 g/L     | 34.40            | 34.40              | 883.00            | 530.67           |
| 2.0 g/L     | 35.60            | 35.50              | 922.67            | 556.00           |
| Control     | 36.67            | 36.60              | 956.67            | 586.34           |
| **120 N kg/fed** |                  |                    |                   |                  |
| Control     | 37.60            | 37.57              | 983.77            | 624.49           |
| 0.5 g/L     | 38.07            | 38.77              | 1030.82           | 654.67           |
| 1.0 g/L     | 38.07            | 38.77              | 1030.82           | 654.67           |
| 2.0 g/L     | 38.07            | 38.77              | 1030.82           | 654.67           |
| **L.S.D**   | **0.45**         | **0.22**           | **11.95**         | **3.60**         |

**B-Yield and yield components:**

**B-1- Effect of Nitrogen fertilizer**

All yield segments altogether expanded because of expanding Nitrogen fertilization levels from 60 to 90 and 120 kg N/fed. in Table (5). Preparing sugar beet plants with120 kg N/fed. of delivered the most noteworthy estimations of all contemplated yield components.

Use of 90 kg N/fed. brought about the best discoveries after the most elevated level of Nitrogen fertilization with critical contrasts correlation with different levels. While, the most reduced ones were acquired because of plant did got Nitrogen fertilization (60 kg N/fed.). Such impact of nitrogen on these attributes might be gotten back to its function in working up metabolites and initiation of compounds that partner with aggregation of starches, which made an interpretation of from leaves to creating roots just as expanding division and lengthening of cells, subsequently expanding root size. The current outcomes are in accordance with those got by (Salami et al.,2013),( Awad et al., 2013a and b),( Amin et al.,2013) ,(Hamad et al.,2015) and (Badr et al.,2017). The most elevated estimations of root and sugar (21.01and 3.15 ton/fed.) were delivered from preparing beet plants with 120 kg N/fed. The least estimations of root yield/fed. were acquired from treatment (60 kg N/fed). The increment in root yield because of utilization of nitrogen preparation can be clarified through the way that nitrogen has a crucial function in working up metabolites, initiating compounds and upgraded root
length, diameter just as root new weight lastly root yield.

**B-2--Effect of foliar application with alga extract**

Data in Table(5) indicate a significant effect on root, top and sugar yields/fad due to the fertilization with Spirulina extract levels Feeding sugar beet plants with 1.0 or 2.0 g/l alga extract/fad without significant difference between them gave higher values of these traits compared with check treatment and those that received 0.5 g/l alga extract.

Application of 2.0 g/l alga extract led to an increase in root, top and sugar yields/fad amounted to 20.6, 7.15 and 3.02 ton/fed, compared to the untreated treatment with alga extract. The increase in root, top and sugar yields/fad may be due to the increase in plant growth parameters (Tables 3 and 4), which in turn is reflected on root, top yields/fed depending on the theory of the Source and downstream.

**Table 5.** The effect of foliar application with alga extract and Nitrogen fertilization on yield and yield components of sugar beet (combined analysis of the two seasons 2018/2019 and 2019/2020).

| Treatments         | Root yield (ton/fed) | Top yield (ton/fed) | Sugar yield (ton/fed) |
|--------------------|----------------------|---------------------|-----------------------|
| 60 N/kg/fed        | 15.74                | 4.33                | 2.15                  |
| 90N/kg/fed         | 16.21                | 6.58                | 2.44                  |
| 120N/kg/fed        | 21.01                | 7.07                | 3.15                  |
| **L.S.D**           | **0.66**             | **0.11**            | **0.06**              |
| Control            | 13.92                | 3.22                | 2.05                  |
| 0.5 g/L            | 15.11                | 4.47                | 2.21                  |
| 1.0 g/L            | 18.44                | 5.15                | 2.63                  |
| 2.0 g/L            | 20.6                 | 7.15                | 3.02                  |
| **L.S.D**           | **0.74**             | **0.21**            | **0.2**               |

**B-3-The Interaction**

The interaction between foliar application with alga extract and Nitrogen fertilization significantly affected all yield characters. The results in Table (6) showed that were responded to the increase of foliar applied Spirulina extract level, but was highly magnitude by adding 120 kg/N. Therefore, the highest value of yield characters.

Except for Sugar yield recorded higher values by spraying beets with 1.0 or 2.0 g/l Spirulina extract, and adding 60 kg N/fed.

It qualified to make reference to that sugar beet quality is convoluted cycle affected by a few elements (Tawfik et al. 2010). The specialized nature of sugar beet is basic for prudent sugar fabricating. Specifically, it relies upon the synthetic organization of the beet. The substance creation of sugar beet is the main boundary influencing its preparing. Sugar manufacturing plants require beet with high groupings of Sucrose and low centralizations of melassigenic substances to boost the measure of extractable sugar (Kenter and Hoffmann, 2005).

**C- Quality traits:**
Table 6. The interaction effect of foliar application with alga extract and Nitrogen fertilization on yield components of sugar beet (combined analysis of the two seasons 2018/2019 and 2019/2020).

| Treatments        | Root yield (ton/fed) | Top yield (ton/fed) | Sugar yield (ton/fed) |
|-------------------|----------------------|---------------------|-----------------------|
| 60 N kg/fed       |                      |                     |                       |
| Control           | 10.23                | 3.11                | 1.21                  |
| 0.5 g/L           | 11.27                | 3.51                | 1.39                  |
| 1.0 g/L           | 12.2                 | 3.95                | 1.94                  |
| 2.0 g/L           | 16.31                | 4.98                | 2.25                  |
| 90 N kg/fed       |                      |                     |                       |
| Control           | 17.39                | 5.12                | 2.47                  |
| 0.5 g/L           | 17.41                | 5.32                | 2.49                  |
| 1.0 g/L           | 18.29                | 5.91                | 2.56                  |
| 2.0 g/L           | 18.53                | 6.70                | 2.84                  |
| 120 N kg/fed      |                      |                     |                       |
| Control           | 19.03                | 6.92                | 2.91                  |
| 0.5 g/L           | 20.51                | 7.53                | 3.02                  |
| 1.0 g/L           | 22.63                | 8.71                | 3.41                  |
| 2.0 g/L           | 24.23                | 9.01                | 3.59                  |
| L.S.D             | 1.6                  | 0.68                | 0.25                  |

It is notable that the main components which influence the nature of sugar beet are the level of sugar, immaculateness and TSS of root. Besides, different substances that disable white sugar recuperation. These are called root pollutants. Dissolvable substances, for example, potassium, sodium, alpha-amino N and different nitrogenous mixes, which can't be wiped out before the Sugar is solidified, increment the measure of Sucrose lost to molasses.

C-1-Effect of Nitrogen fertilization

Data in Fig (1&2) indicated that expanding Nitrogen fertilization up to 60 N/fed. Fundamentally expanded Sucrose rate, TSS, α-amino N, K rate, Na rate, extractable sugar rate and sugar lost molasses rate. Be that as it may, purity% was not influenced by nitrogen rates expanding nitrogen manure prompted diminishing juice virtue rate. These outcomes are in concurrence with those got by Ramadan et al. (2003), Leilah et al. (2005) and Gomaa et al. (2017) in sandy soil.

Higher rates of Nitrogen fertilization (120 N/fed.) had altogether impact on α-amino-N, K and Na content These outcomes might be because of the explanation that high pace of N expanded solvent non-sugar in root juice (pollutants) and they meddle with sugar extraction which reflected by raising the level of sugar misfortunes to molasses and therefore decreasing sugar recuperation.
C-2- Effect of foliar application with alga extract

significantly influenced by the applied alga extract concentrations. Meantime, the fertilization with 1.0 g/l and/or 2.0 g/l alga extract led to higher values of extractable sugar percentage, Sucrose % and purity %. Meanwhile TSS %, alpha-amino-N, K%, Na% and sugar lost molasses % were significantly influenced by the applied alga extract concentrations. However increases alga extract concentrations decrease TSS %, alpha-amino-N, K%, Na% and sugar lost molasses %. These finding are in agreement with that mentioned by (Enan et al., 2016).
Figure 4. The effect of foliar application of alga extract on Sugar beet quality traits.

C-3- The Interaction
The interaction between foliar application with alga extract and Nitrogen fertilization significantly effect on Sucrose's %, TSS% and QZ%. The results in Table (7) showed that the highest values of Sucrose's% and QZ % by spraying beets with 1.0 or 2.0 g/l Spirulina extract, and adding 60 kg N/fed. (18.70%, 19.20% and 77.37 %, 81.18%). Except for TSS% give lowest value by spraying beet with 0.5 g/l Spirulina extract and adding 60 kg N/fed (18.00%). Meanwhile alpha-amino-N, K%, Na%, extractable sugar % and Sugar lost molasses (SLM %) not significant effect by foliar application with alga extract and Nitrogen fertilization.

Table 7. The interaction effect of foliar application with alga extract and Nitrogen fertilization on quality traits of sugar beet (combined analysis of the two seasons 2018/2019 and 2019/2020).

| Treatments       | Sucrose % | TSS%  | QZ %  |
|------------------|-----------|-------|-------|
| 60 N kg/fed      | Control   | 17.20 | 18.90 | 79.65 |
|                  | 0.5 g/L   | 17.83 | 18.00 | 77.73 |
|                  | 1.0 g/L   | 18.70 | 18.87 | 77.37 |
|                  | 2.0 g/L   | 19.20 | 19.47 | 81.18 |
| 90 N kg/fed      | Control   | 17.43 | 18.50 | 75.92 |
|                  | 0.5 g/L   | 17.87 | 18.70 | 77.73 |
|                  | 1.0 g/L   | 18.10 | 19.27 | 80.30 |
|                  | 2.0 g/L   | 18.47 | 19.90 | 81.40 |
| 120 N kg/fed     | Control   | 17.60 | 20.53 | 75.92 |
|                  | 0.5 g/L   | 17.67 | 21.57 | 75.94 |
|                  | 1.0 g/L   | 18.13 | 22.17 | 78.19 |
|                  | 2.0 g/L   | 18.47 | 22.90 | 78.32 |
| L.S.D            |           | **0.46** | **0.25** | **1.70** |
**Principal components analysis**

Data in Figure (5) show Sugar yield was significantly and positively correlated with Sucrose% followed by juice purity%, then and brix%. Negative correlation was detected between juice purity%, and brix %. These results are agreement with those stated by (Gadallah and Mehareb 2020) and (Mehareb and El-Mansoub 2020), who mentioned that purity% was negatively and significantly correlated with brix %. On the other hand, results in Figure (5) showed that top yield was significantly and positively correlated with Root weight followed by Top yield and Top weight.

![Principal components biplot (99.91%)](image)

**Figure 5.** show highly significant and positive correlation between root yield and root length followed by root yield and root diameter then root yield and root weight.

**Conclusion**

Spirulina extract when applied as a foliar application, can contribute to the supply of plant nutrients and substitute nutrients. On the results of this study, it can be recommended to spray sugar beet with 1.0 or 2.0 g/L of Spirulina extract along with 120 N k/fed for maximum yields of root but sugar yield, Sucrose % and purity adding 90 N k/fed to spray with 1.0 or 2.0 g/L of Spirulina extract under sandy soil conditions.

**References**

Abd El-Mawgoud, A.M.R., Tantawy, A.S., El-Nemr, M.A. and Sassine, Y.N. (2010) ‘Growth and yield responses of strawberry plants to chitosan application’ Europ. *J. Scientific Res* 39 (1), pp. 161-168.

Abo Elenen. Fouz, F. M., Helmy, Samar. A. M., Mehareb, Eid. M. and Bassiony. Noran. A. M. (2019) ‘Genetic diversity and principal component analysis for agronomic and technological characterization of sweet sorghum germplasm under Egyptian conditions’
Amin, Gehan A., Elham A. Badr and M.H.M. Afifi (2013) ‘Root yield and quality of sugar beet in response to biofertilizer and foliar application with micronutrients’ World Applied Sciences Journal 27 (11), pp. 1385-1389.

Awad, N. M.M.; Sahar, F.Tawfik and Sahr, M.I. Moustafa (2013a) ‘Influence of foliar spray of some micronutrients and nitrogen fertilizer on productivity of sugar beet under newly reclaimed soils’ J. Agric. Res. Kafr El-Sheikh Univ. 39 (2), pp. 181-194.

Azzazy, N.B. (2004) ‘Effect of nitrogen and boron fertilization on yield and quality of sugar beet under new reclaimed soil condition’ Egypt J. Appl. Sci. 19 (8), pp. 1099-1113.

Badr, Elham A. Gehan. Sh. Bakhoun, Gehan A. Amin and Howida, H. Khedr (2017) ‘Effect of unconventional fertilizers on Root quality and yield components of sugar beet (Beta vulgaris L.) plants’, Bull. NRC 41 (Bi.2), pp. 260-272.

Badr, E. A. (2006) ‘Effect of organic, biofertilization and plant density on yield and quality sugar beet’, Ph.D .Thesis Fac., Agric. (SabaBesh) Alex . Univ., Egypt .

Carruthers, A. and J.F.T. Oldfield (1961) ‘Methods for the assessment of beet quality’ Int. Sug. J. 63, pp. 7 2-74.

Clinton C. Shock, Majid Seddigh, Lamont D. Saunders, Timothy D. Stieber, and John G. Mille (2000) ‘Sugar beet nitrogen uptake and performance following heavily fertilized Onion’ Agron. J. 92, pp. 10-15.

Cooke, D.A. and Scott, R.K. (1993) ‘The Sugar Beet Crop, Science Practice’ Pablished by Chapman and Hall, London.

Dillon J., Phue A. and Dubacq J. (1995) ‘Nutritional value of the algae Spirul ina’ World. Rev. Nutr. Diet. Karger 77, pp. 32-46.

El-Harriri, D.M. and Mirvat E. Gobarah (2001) ‘Response of growth, yield and quality of sugar beet to nitrogen and potassium fertilizers under newly reclaimed sandy soil’ J. Agric. Sci. Mansoura Univ. 26 (10) pp. 5895 - 5907.

Enan, S.A.A.M., A. M. El-Saady and A. B. El-Sayed (2016) ‘Impact of Foliar Feeding With Alga Extract and Boron on Yield and Quality of Sugar Beet Grown in Sandy Soil’ Egypt. J. Agron. Vol. 38, (2), pp. 319-336.

Gadallah A.F.I. and E.M. Mehareb (2020) ‘Yield and quality of some sugarcane varieties as affected by irrigation number’ SVU-International Journal of Agricultural Science 2 (2) pp. 144-165.

Ghalab, A.M. and Salem, S.A. (2001) ‘Effect of biofertilizer treatments on growth, chemical composition and productivity of wheat grown under different levels of NPK fertilization’ Annal Agric. Sci. 46, pp. 485-509.

Gomaa, M. A., G. Abdel-Nasser, M. F. ‘Maareg and M. M. El-Kholi (2017) Sugar Beet Response to Nitrogen and Potassium Fertilization Treatments in Sandy Soil’ J. Adv. Agric. Res. (Fac. Agric. Saba Basha) 22 (2), pp. 272-287.

Hamad, A. M.; H. M. Sarhan and S. S. Zalat (2015) ‘Effect of nitrogen; potassium fertilizer and plant distribution patterns on yield and quality of sugar beet (Beta vulgaris L.)’ J. Plant Production, Mansoura Univ. 6 (4), pp. 517 – 527.

Ismail, A.M.A (2002) ‘Evaluation of some sugar beet varieties under different nitrogen levels in El-Fayium’ Egypt, J. Appl. Sci. 17(2), pp. 75-85.
Ismail, A.M.A. and R. A. Abo EL-ghait (2005) ‘Effect of nitrogen sources and levels on yield and quality of sugar beet’, Egypt J. Agric. Res. 83 (1), pp. 229-235.

Jackson, M. L. (1973) ‘Soil Chemical Analysis’, Prentice-Hall, Inc. Analytical Chemists, Official Methods of Analysis’, 11th Ed., Washington, D. C.

Johnson PE, Shubert LE (1986). Accumulation of mercury and other elements by Spirulina (Cyanophyceae). Nutr. Rep. Int. 34, pp. 1063-1070.

Kenter C. and Hoffmann C. M. (2005) ‘Seasonal patterns of Sucrose concentration in relation to other quality parameters of sugar beet (Beta vulgaris L.)’ Journal of the science of food and Agriculture 86 (1), pp. 62-70.

Klute A. (1986) ‘Methods of Soil Analysis, Part I, Physical and Mineralogical Methods 2nd edition’ American Society of Agronomy, Madison, Wisconsin, USA.

Koochekzadeh, A., G. Fathi, M.H. Gharineh, S.A. Siadat, S. Jafari and Kh. Alami-Saeid, (2009) ‘Impacts of rate and split application of N fertilizer on sugarcane quality’, Int. J. Agric. Res. 4, pp. 116-123.

Le-Docte, A. (1927) ‘Commercial determination of sugar beet root using the Sachs Le-Docta process’, Int. Sugar J., (29), pp. 488-492.

Leilah, A.A.; M.A. Badawi; E. M. Said; M.H. Ghonema and M.A. E. Abdou. (2005) ‘Effect of planting dates, plant population and nitrogen fertilization on sugar beet productivity under the newly reclaimed sandy soils in Egypt’ Scientific Journal of King Faisal University (Basic and Applied Sciences) 6(1), pp. 95-110.

Leilah, A.A.; S.E; El-Kalla, A.T. El-Kassaby; M.A. Badawi and Mahasen M. Fahmi (2007) ‘Yield and quality of sugar beet in response to levels and times of nitrogen application and foliar spraying of Urea’ Scientific Journal of King Faisal University (Basic and Applied Sciences) 8 (1), pp. 87-100.

Mannino S, Beneli TG (1980) ‘Constituenti minerali di biomasse di Spirulina maxima. In Materassi R (ed.)’, Prospettive della coltura di Spirulina in Italia. CNR, Rome, pp. 131-135.

Marrez, D. A., Naguib, M. M., Sultan, Y. Y., Daw, Z.Y. and Higazy, A.M. (2014) ‘Evaluation of chemical composition for Spirulina platensis in different culture media’ Res. J. Pharmaceutical, Biol. and Chem. Sci. 5 (4), pp. 1161-1171.

McIntosh, M.S. (1983) ‘Analysis of combined experiments’ Agron. J. 75, pp. 153–155.

Mehareb, E. M and A El-Mansoub, M. M. (2020) ‘Genetic parameters and Principal components biplot for agronomical, insect and Pathological traits in some sugarcane genotypes’ SVU-International Journal of Agricultural Science 2 (2), pp. 77-93.

Montemurro, F. and M Maiorana: (2007) ‘Nitrogen utilization, yield, quality and soil properties in a sugar beet crop amended with municipal solid waste compost’ Compost-Science-and-Utilization. 15(2), pp. 84-92.

Ortega, C. J. J.; Mazuelos, C.; Hermosin, B. and Saiz-Jimenez, C. (1993) ‘Chemical composition of Spirulina and eukaryotic algae food products marketed in Spain’ J. Appl. Phycol. 5, pp. 425-435.

Page AI, Miler RH and Keeny DR. (1982) ‘Methods of Soil Analysis part II. Chemical and Microbiological Methods’.2nd ed. Amer. Soc. Agron., Madison, Wisconsin, USA.

Ramadan, B.S.H.; H.R. Hassan and Fatma A. Abdo (2003) ‘Effect of minerals and biofertilizers on photosynthetic pigments, root quality, yield components and anatomical structure of sugar beet (Beta vulgaris L.) Plants grown under reclaimed soils’ J. Agric. Sci. Mansoura Univ. 25 (12), pp.7389-7398.

Raupp, J. and Oltmanns, M. (2006) ‘Farmyard manure, plant based organic fertilizers, inorganic fertilizer-which sustains soil organic matter best’, Aspects of Applied Biology 79, pp. 273-276.
Raupp, J. and Oltmanns, M. (2006) ‘Farmyard manure, plant based organic fertilizers, inorganic fertilizer-which sustains soil organic matter best’ Aspects of Applied Biology 79, pp. 273-276.

Salami, M. and S. Saadat (2013) ‘Study of potassium and nitrogen fertilizer levels on the yield of sugar beet in jolge cultivar’, J. Novel Appl. Sci. 2(4), pp. 94–100.

Seadh; S.E. Farouk S. and M.I. EL-Abady,(2007): Response of sugar beet to potassium sulfate under nitrogen fertilizer levels in newly reclaimed soils conditions’ African Crop Science Conference Proceedings (8), pp. 147-153.

Shafeek, M.R., Helmy, Y.I., Omer, Nadia M. and Rizk, Fatma A. (2013) ‘Effect of foliar fertilizer with nutritional compound and humic acid on growth and yield of broad bean plants under sandy soil conditions’ J. Appl. Sci. Res. 9 (6), pp. 3674-3680.

Sheikh_Aleslami, R., (1997) ‘Laboratorial Methods and their Application to Control’ Food and Sugar Industries, Process Mersa Publications, Tehran, Iran.

Snedecor, G.V. and W. G. Cochran (1990) ‘Statistical Methods 8th Ed’. The Iowa State Univ. Presss. Ames. 158-160.

Stevens W.B., A.D. Blaylock, J.M. Krall, B.G. Hopkins and J. W. Ellsworth (2007) ‘Sugar beet yield and nitrogen use efficiency with preplant broadcast, banded or point-injected nitrogen application’ Agron. J. (99), pp. 1252-1259.

Tawfik,M.M. Mirvat, I. Gobarah and Magda, H. Mohamed (2010) ‘Management practice for increasing potassium fertilizer efficiency of sugar beet in north Delta Egypt’ International Journal of Academic Res. 2(3), pp. 220-225.

Waller, R.A., and D.B. Duncan (1969) ‘A Bayes rule for the symmetric multiple comparisons problem’, J. Amer. Statist. Ass. 64, pp. 1484–1503.

Wettstein, D. (1957) ‘Chlorophyll, Letal und der submikro svopische formmech Sallplastiden’, Exptl. Cell Ser. (12), pp. 427-433.

Yassen, A.A., Badran, N.M. and Zaghloul, S.M. (2007) ‘Role of some organic residues as tools for reducing metals hazard in plant’, World J. Agric. Sci. 3(2), pp. 204-209.

Yassen, A.A., Badran, N.M. and Zaghloul, S.M. (2007) ‘Role of some organic residues as tools for reducing metals hazard in plant’ World J. Agric. Sci. 3(2), pp. 204-209.

Zhang, X. and Ervin, H. (2004) ‘Seaweed extract and humic acid contain cytokinins ’ Crop. Sci. 44 (5), pp. 1509.

Zhenxie Yi, Pu Wang, Hongbin Tao, Hongfang Zhang and Lixia Shen (2008) ‘Effects of types and application rates of nitrogen fertilizer on the development and nitrogen utilization of summer maize,’ Frontier of Agric. 3(1), pp. 44-49.