Periodontitis and alzheimer’s disease

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Abstract
Background: Alzheimer’s disease (AD), the main cause of dementia in the adult population, is characterized by a progressive loss of cognitive function. It is considered that neuroinflammation plays a fundamental role in its onset and progression. The bacteria present in the disbiotic microbiome generated during the course of periodontitis (PE) are capable of inducing a systemic inflammatory response, exacerbating the production of proinflammatory mediators that have the potential to spread to the systemic circulation.

Material and Methods: A literature review was made using the databases Scielo, PubMed, EBSCO and key words “Alzheimer disease”, “Periodontitis”, “Neurodegeneration”, “Inflammation mediators”, “Elderly”.

Results: Several hypotheses point to similar pathophysiological pathways in the establishment of AD and PE, sharing cellular and molecular proinflammatory characteristics. In periodontitis, locally produced cytokines and pro-inflammatory products spread from the ulcerated periodontal pocket into the systemic circulation, or around the trigeminal nerve terminals, which allows the passage of bacteria or their products to the brain. This fact leads to the formation of plaques of amyloid peptide and intraneuronal neurofibrillar tangles (NFTs) that activate the glial cells producing a significant increase in proinflammatory cytokines in the affected regions that lead to a loss of neuronal synapses and neurodegeneration, contributing to the progression of AD.

Conclusions: This review of the literature contributes to the understanding of the pathological pathways shared by both diseases such as oxidative damage and inflammation. There is not enough evidence to determine an association between these two pathologies, so it is considered necessary to conduct studies to determine if periodontitis is capable of inducing or exacerbating the neuroinflammation that will trigger AD.

Key words: Alzheimer’s disease, periodontitis, neurodegeneration, inflammatory mediators, elderly.
Introduction

Alzheimer's disease (AD) is the most common form of dementia in elderly, constituting the main cause of dependence and disability in this population group. It appears more frequently in people over 65 years of age, though there is an affected group of younger age and is more prevalent in women than in men (1). According to the World Health Organization's report of 2018, there are 50 million people in the world living with dementia, of which two-thirds are affected by AD (2). As life expectancy increases, it is estimated that the incidence of AD increases to 152 million people by 2050, becoming a major health problem (1,3).

Early onset AD is believed to be extremely determined by genetic factors, while late or sporadic onset, which includes 95% of patients, results from the interaction of risk and environmental factors. Risk factors for AD onset include family history, education, high-fat diet, hypertension, diabetes, dyslipidemia, distress, insomnia, sedentary lifestyle, smoking, alcoholism, substance abuse, age, history of head trauma or susceptibility genes such as Amyloid Precursor Protein (APP), Beta-secretase (BACE) and Apolipoprotein E (APOE) (4).

Currently, several experimental models explain the pathology of AD with different onset and progression causes, however, it is considered that neuroinflammation plays a central role in the amyloid cascade hypothesis. The most accepted theory to explain the pathogenesis of AD is the amyloid cascade, where the neuroinflammation is the initial event conducting to the accumulation and aggregation of β-amyloid peptide (Aβ), followed by the formation of neurofibrillar tangles (NFT) formed by a hyperphosphorylated form of the Tau protein. The presence of Aβ induces hyperphosphorylation of Tau proteins and the formation of NFTs and thus, axonal dysfunction that generates higher accumulation of Aβ in synaptic spaces. The Aβ aggregates make up the senile plaques and, together with NFTs, constitute the histopathological features of AD (5-10).

It has been speculated that systemic infections may play an important role in establishing an inflammatory state in the central nervous system. In this regard, PE is one of the most common oral infections in the adult population, it is considered the second most common oral disease in humans and in older age groups a major cause of tooth loss (11,12). It derives from the establishment of a dysbiotic microbiome that is generated by keystone pathogens or pathobionts (13). These bacteria or its virulence factors can induce both direct damage to periodontal tissues or indirect damage by inducing the host’s immune response, which leads to the formation of subgingival pocket and evoking chronic inflammation in periodontal tissues and causes alveolar bone loss (14,15). This dysbiotic microbiome is capable of inducing a low-grade systemic inflammatory response by activating the host’s immune response and exacerbated production of pro-inflammatory mediators such as interleukin (IL)-1α, IL-1β, IL-6, tumor necrosis factor (TNF)-α, prostanoids and matrix metalloproteinases (MMPs) that have the potential to spread to the bloodstream (16). At the local level, the dysbiotic microbiome modulates the destruction of periodontal connective tissue and bone resorption (16). At the systemic level, it results in the host’s exposure to various cytokines and inflammatory mediators (17). The periodontal pathogens associated with periodontitis are rich in endotoxins and Lipopolysaccharide (LPS) that stimulate the activity of immune cells and the production of cytokines. Cytokines, such as IL-1β, IL-6 and TNF-α can spread to brain by bloodstream or by peripheric nerve terminals. Once in the brain, cytokines, bacteria or its virulence factors could, eventually, stimulate glial cells and induce neuroinflammation, which can contribute to the onset or progression of AD (18).

The aim of this article is to determine the pathological pathway through AD and PE can be associated, to allow an approach and higher knowledge about these two diseases so prevalent in elderly.

Material and Methods

A literature review was made using the databases Scielo, PubMed, EBSCO, MEDLINE and DOAJ, for this purpose the keywords “Alzheimer’s disease”, “Periodontitis”, “Neuroimmune mechanism”, “Neurodegeneration”, “Inflammation mediators” and “elderly” were used. The review was carried out in the period between December 2019-April 2020. 25 studies met the specific parameters and were included, among them bibliographic review studies, clinical trials, cases and controls, experimental studies and bibliographic meta-analysis.

Results

- Alzheimer and periodontitis: links

Similar pathological pathways have been reported in the establishment of AD and PE, including cellular and molecular characteristics such as oxidative damage and inflammation (Fig. 1). A model of association suggest that during the course of PE, locally produced pro-inflammatory cytokines diffuse from the ulcerated periodontal pocket into the systemic circulation around the trigeminal nerve terminals. Also, it has been suggested that bacteria or its virulence factors could spread from periodontal tissues to the brain through peripheric nerves, being the trigeminal nerve the main pathway. This molecules could induce neuroinflammation and, under chronic condition of neuroinflammation, mediators activated pro-inflammatory microglia phenotype, known as M1. (19-21).
In a state of health, microglia, the mononuclear phagocytes destined to limit injury in the central nervous system, maintain a neuroprotective function by eliminating Aβ peptides and expressing protective factors such as Insulin Growth Factor (IGF-1), Transforming Growth Factor-β and Nerve Growth Factor (22). Under inflammatory condition, the microglia differentiate toward a modulator phenotype or M2, which is characterized by secreting IL-10 and tumor growth factor (TGF)-β1. When this inflammatory state becomes persistent, the microglia modify its phenotype from the modulatory M2 to the pro-inflammatory M1. The M1 cells increase the production of IL-1β, IL-6 and TNF-α and are able to modify the function of astrocytes. The physiological astrocyte modifies its phenotype to a reactive astrocyte that responds by increasing the secretion of pro-inflammatory cytokines and the production of the amyloid precursor protein (APP) and of β and γ-secretases, generating the production of Aβ. The modification of the astrocyte function generates a metabolic, energetic and oxidative imbalance in the neuron that will respond by increasing the production of Aβ and the hyperphosphorylation of the microtubule-associated protein, Tau. These molecular events generate neuro and synaptotoxicity, constituting the main histopathological markers of AD (Fig. 2).

In general terms, microglia play a neuroprotective role, but in inflammatory conditions a neuroinflammatory role, thus being the central cell at the onset of AD (23). The amyloid cascade hypothesis establishes Aβ as the central axis of neurodegeneration. Indeed, the Aβ peptide once secreted by astrocytes or neurons can undergo enzymatic modifications as non-enzymatic, generating its insolubilization. Insolubilization of Aβ peptides generate their accumulation in the brain and the establishment of senile plaques. Both soluble and insoluble Aβ are detected in the brains of patients affected by AD and, in this context, the presence of insoluble Aβ in the brain is directly correlated with the loss of synapse and cognitive impairment, being considered a marker of AD (24-26).

A neurodegenerative process secondary to the aggregation of Aβ in AD is the formation of NFTs, formed by a hyperphosphorylated form of the Tau protein that is expressed primarily in neurons. Both Aβ peptides and the immune response induced before Aβ peptides generate hyperphosphorylation of Tau proteins and the formation of NFTs and thus, axonal dysfunction that generates higher accumulation of Aβ in the synaptic spaces. In the initial stages of AD, these lesions are found in the entorhinal cortex and in the region of the dentate gyrus (GD) of the hippocampus —regions of the brain
associated with memory processing and storage—and during the progression they extend from the GD to the field of Amón (CA)3, CA2 and CA1 to later affect the different areas of the cerebral cortex. The presence of these protein aggregates and dysfunctional proteins induce an immune-inflammatory response in the affected regions. Thus, the presence of pro-inflammatory molecules secreted by microglia, astrocytes, endothelial cells or neurons in response to these proteins, such as IL-1β and IL-6, TNF-α or INF-γ generate the vicious cycle of neuronal injury (27-29). Thus, NFTs are considered a true progression of AD marker, and insoluble Aβ, a marker for diagnosis.

- Role of the periodontal bacteria
In animal models it has been shown that chronic neuroinflammation induced by inoculation or injection of Porphyromonas gingivalis Lipopolysaccharide (LPS) causes exacerbation in the production of pro-inflammatory cytokines and mediators which leads to hyperphosphorylation of Tau protein and an increase in Aβ levels contributing to neuronal deterioration (30,31). Numerous bacteria have been studied to determine a possible role of infection at the onset of AD. Thus, Treponema denticola and Treponema pallidum have been detected in the trigeminal ganglion and cortex and Porphyromonas gingivalis in the fourth ventricle and cerebrospinal fluid (32,33). These data allow us to speculate that bacteria are able to invade the brain and possibly generate a local inflammatory response and neurotoxicity (Fig. 3) (33).

The mechanism of bacterial invasion to the brain are still unknown, however three models are proposed. The first establish that the bacteria spread into the bloodstream and, once in the cerebral vessels, the inflammatory response associated with cerebral vascular atherosclerosis could induce the breakdown of the blood-brain barrier (BBB). The second determine that oral bacteria would migrate through the peripheral terminations of the trigeminal nerve to the trigeminal ganglion and then, to the brain. The third route establishes that bacteria could migrate by lymphatic circulation. Indeed, the IV ventricle and the oral regional lymph nodes drain to a common lymph node, the mid-deep cervical lymph node (32,34). Although the three theories explain the possible routes of migration of oral bacteria to the brain, it has not yet been proven by which of them these bacteria can enter more easily.

- Periodontitis as a source of systemic inflammation.
One of the distinctive features of AD is the presence of activated glial cells that produce significant levels of inflammation. Suggesting that processes capable of regulating the expression of inflammatory molecules would contribute to their progression once established. Pro-inflammatory molecules derived from the periphery could increase the set of inflammatory molecules in the brain by at least two mechanisms, through systemic circulation or peripheral nerve endings. Pro-inflammatory molecules in the systemic circulation could enter to the CNS in multiple ways. They could enter through
areas of the brain that lack a BBB, but they could also enter in areas with BBB by various mechanisms: 1) diffusion through fenestrated capillaries of the BBB, 2) using specific cytokine transporters, 3) increasing the permeability of BBB or 4) activating brain endothelial cells to produce cytokines that induce signaling molecules such as nitrous oxide or prostanoids (35,36).

Once in the brain, these pro-inflammatory molecules increase the amount of cytokines, but also stimulate glial cells to synthesize additional pro-inflammatory cytokines (37). Several studies have determined that peripheral cytokines have the ability to stimulate afferent fibers of peripheral nerves, leading to increased levels of brain cytokines, or they may also have the ability to enter the brain through channels or compartments associated with peripheral nerves (36,37). The last hypothesis suggests that pro-inflammatory molecules originating in the oral cavity can reach the brain through neurological pathways (37).

Additionally, bacterial products or bacteria themselves can increase the concentration of brain cytokines. Various bacteria have been identified as cofactors in the etiology and pathogenesis of AD (22), including *Chlamydia Pneumoniae*, *Borrelia burgdorferi* and *Treponema denticola*. Similarly, the findings indicate that periodontal bacteria, *Aggregatibacter actinomycetemcomitans*, *Porphyromonas gingivalis*, *Prevotella intermedia*, *Fusobacterium nucleatum*, *Tannerella forsythia*, *Eikenella corrodens* and *T. denticola*, can invade the brain through two pathways: through bloodstream or through peripheral nerves (38,39).

Finally, a bidirectional relationship between both diseases has also been proposed, there being a coherent relationship so that AD can predispose to the development of PE, this is explained because patients with AD have a worse oral hygiene, either because their dexterity manual to carry out daily oral hygiene is diminished or absent or due to the inability of the subject himself to go to the dentist for professional care (40).

**Discussion**

This review of the literature contributes to the understanding of the pathological pathways that both diseases share. There is still not enough evidence to determine an association between the two pathologies, however the available evidence indicates a positive trend towards the association between them. The existence of a causal relationship between them is unknown due to the heterogeneity of the designs used. For this reason, it is considered necessary to carry out studies to determine if PE is capable of inducing or exacerbating the neuroinflammation that will trigger AD.

**References**

1. Knopman DS, De-Kosky ST, Cummings JL, Chui H, Corey-Bloom J, Relkin N, et al. Practice parameter: Diagnosis of dementia (an evidence-based review). Report of the Quality Standards Subcommittee of the American Academy of Neurology. Neurology. 2001;56:1143-53.
2. Calusolaro V, Antognoli R, Okoye C, Monzani F. The use of antipsychotic drugs for treating behavioral symptoms in Alzheimer’s disease. Frontiers in Pharmacology. 2019;10:1-8.
3. Ferri CP, Prince M, Brayne C, Brodaty H, Fratiglioni L, Ganguli M, et al. Global prevalence of dementia: a Delphi consensus study. Lancet. 2005;366:2112-7.
4. Kamer AR, Craig RG, Dasanayake AP, Brys M, Gloods-Sobranska L, De-Leon MJ. Inflammation and Alzheimer’s disease: Possible role of periodontal diseases. Alzheimers Dement. 2008;4:242-50.
5. Bateman RJ, Xiong C, Benzinger TL, Fagan AM, Goate A, Fox NC, Marcus DS, Carins NJ, Xie X, Blazey TM et al. Dominantly Inherited Alzheimer Network. Clinical and biomarker changes in dominantly inherited Alzheimer’s disease. N Engl J Med. 2012;367:795-804.
6. Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer’s disease: progress and problems on the road to therapeutics. Science. 2002;297:353-6.
7. Castello MA, Soriano S. On the origin of Alzheimer’s disease. Trials and tribulations of the amyloid hypothesis. Ageing Res Rev. 2014;13:10-2.
8. Ferreira ST, Clarck JR, Bomfim TR, De-Felice FG. Inflammation defecting signaling and neuronal dysfunction in Alzheimer’s disease. Alzheimers Dement. 2014;10:372-80.
9. Cochran JN, Hall AM, Roberson ED. The dendritic hypothesis for Alzheimer’s disease pathophysiology. Brain Res Bull. 2014;103:18-28.
10. Baranello RJ, Bharani KL, Padmaraju V, Chopra N, Lahiri DK, Greig NH, et al. Amyloid-beta protein clearance and degradation pathways and the role in Alzheimer’s disease. Curr Alzheimer Res. 2015;12:32-46.
11. Ide M, Harris M, Stevens A, Sussams R, Hopkins V, Culliford D, et al. Periodontitis and cognitive decline in Alzheimer’s disease. PLoS One.2016;11:1-9.
12. McGeer PL, McGeer EG. Inflammation, autotoxicity and Alzheimer disease. Neurobiol Aging. 2001;22:799-809.
13. Socransky SS, Haffajee AD. Periodontal microbial ecology. Periodontol 2000. 2005;38:135-87.
14. Javed F, Ahmed HB, Mikami T, Almas K, Romanos GE, Al-Hezaimi K. Cytokine profile in the gingival crevicular fluid of rheumatoid arthritis patients with chronic periodontitis. J Investig Clin Dent. 2014;5:1-8.
15. Ximenez-Fyvie LA, Haffajee AD, Socransky SS. Comparison of the microbiota of supra and subgingival plaque in health and periodontitis. J Clin Periodontol 2000. 2007;8:648-57.
16. McGeer PL, McGeer EG. Inflammation, autotoxicity and Alzheimer disease. Neurobiol Aging. 2001;22:799-809.
17. Riviere GR, Riviere KH, Smith KS. Molecular and immunological evidence of oral Treponema in the human brain and their association with Alzheimer’s disease. Oral Microbiol Immunol. 2002;17:113-46.
18. Kramer AR, Craig RG, Dasanayake AP, Brys MJ. Inflammation and Alzheimer’s disease: Possible role of periodontal disease. Alzheimers Dement. 2008; 4:242-50.
19. Elobeid A, Sylvia L, Marina L. Altered proteins in the aging brain. J Neuropathol Exp Neurol. 2016;75:316-25.
20. Thal DR, Rub U, Orantes M, Braak H. Phases of Aβ deposition in the human brain and its relevance for development of AD. Neurology. 2002;58:1791-800.
21. Braak H, Thal DR. Stages of the pathologic process in Alzheimer’s disease: age categories from 1-100 years. J Neuropathol Exp Neurol. 2011;71:171-81.
22. Weitz T, Town T. Microglia in Alzheimer’s disease: it’s all about context. Int J Alzheimers Dis. 2012;2012:2-13.
23. Schram MT, Euser SM, de Craen AJ, Wittman JC, Frolích M, Hofman A, et al. Systemic markers of inflammation and cognitive decline in old age. J Am Geriatr Soc. 2007;55:708-16.
24. Barron M, Gartlon J, Dawson LA, Atkinson PJ, Pardon MC. A state of delirium. Deciphering the effect of inflammation on tau pathology in Alzheimer’s disease. Exp Gerontol. 2017;94:163-7.
25. Bolos M, Llorens-Martín M, Jurado-Arjona J, Hernández F, Rábano A, Avila J. Direct evidence of internalization of tau by microglia receptor. J Alzheimers Dis. 2015;50:77-87.
26. Fiala M, Cribs DH, Rosenthal M, Bernard G. Phagocytosis of amyloid-beta and inflammation: two faces of innate immunity in Alzheimer’s disease. J Alzheimers Dis. 2007;11:457-63.
27. Liu CC, Zhao N, Fu Y, Wang N. ApoE4 accelerates early seeding of amyloid pathology. Neuro. 2017;96:1024-32.
28. Garcia-Alloza A, Robbins EM, Zhang-Nunes SX, Purcell SM, Betensky RA, Raju S, et al. Characterization of amyloid deposition in the APPps1 mouse model of Alzheimer’s disease. Neurobiol Dis. 2006; 24: 516-24.
29. Grace EA, Busciblio J. Aberrant activation of focal adhesion proteins mediates fibrillar amyloid-beta induced neuronal dystrophy. J Neurosci. 2003;23:493-502.
30. Kitazawa M, Oddo S, Yamasaki TR, Green KN, La-Ferla FM. Lipopolysaccharide-induced inflammation exacerbates tau pathology by a cyclin-dependent kinase 5-mediated pathway in a transgenic model of Alzheimer’s disease. J Neurosci. 2005;25:8843-53.
31. Lee JW, Lee YK, Suk DY, Choi DY, Ban SB, Oh KW. Neuroinflammation induced by lipopolysaccharide causes cognitive impairment through enhancement of beta-amyloid generation. J Neuroinflammation. 2008;5:37-50.
32. Singhrao SK, Poole S, Harding A, Kesavalu L, Crean S. Porphromonas gingivalis periodontal infection and its putative links with Alzheimer’s Disease. Mediators Inflamm. 2015;1:1-10.
33. Dominy SS, Lynch C, Ermini F, Benedykt M, Mareczk A, Konradi A, et al. Porphromonas gingivalis in Alzheimer’s disease brains: evidence of disease causation and treatment with small-molecule inhibitors. Sci Adv. 2019;5:2-21.
34. Nee LE, Lipa CF. Alzheimer’s disease in 22 twin pairs: 13-year follow-up—hormonal, infectious and traumatic factors. Dement Geriatr Cogn Disord. 1999;10: 148-51.
35. McGeer PL, McGeer EG. Inflammation, autotoxicity and Alzheimer disease. Neurobiol Aging. 2001;22:799-809.
36. Danzter R, Konsman JP, Bluthe RM, Kelley KW. Neural and humoral pathways of communication from the immune system to the brain: parallel or convergent? Auton Neurosci. 2000;85:60-5.
37. Miller AJ, Muñoz-Gonzalez A, Rothwell NJ, Hopkins SJ. Local cytokine induction by LPS in the rat aortic arch and its relationship to the febrile response. Am J Physiol. 1997; 272:857-61.
38. D’Aiueto F, Graziani F, Tett F, Gabriele M, Tonetti MS. Periodontitis: from local infection to systemic diseases. Int J Immunopathol Pharmacol. 2005;18:1-11.
39. Ishida N, Ishihara Y, Ishida K, Tada H, Funaki-Kato Y, Hagiwara M, et al. Porphyromonas gingivalis periodontal infection and its putative links with Alzheimer’s disease. Mediators Inflamm. 2015;1:1-10.
40. Leira Y, Domínguez C, Seoane J, Pias J. Is periodontal disease associated with Alzheimer’s disease? A systematic review with metaanalysis. Neuroepidemiology. 2017;48:21-31.

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Conflict of interest

The authors declare that in this study there are not conflicts of interest.

Authors contributions

Sansores-España and Díaz-Zúñiga conceived the review idea; Martínez-Aguilar, Sansores-España and Díaz-Zúñiga literature review and article selection; Cárriho-Avila, Melgar-Rodríguez, Martínez-Aguilar led the manuscript writing. All authors approved the final version of the submitted manuscript.