Evaluation of the lymphoproliferation of mononuclear cells in cutaneous leishmaniasis patients treated with Libidibia ferrea

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Abstract
American integumentary leishmaniasis (ATL) is a neglected disease that mostly affects vulnerable populations. Its broad spectrum of clinical manifestations is related to the type of immune response produced by the host and the species of Leishmania involved. In recent years, the use of medicinal plants has become a therapeutic alternative in the treatment of infectious parasitic diseases. This research aimed to evaluate the lymphoproliferative responses of peripheral blood mononuclear cells (PBMCs) of patients with cutaneous leishmaniasis (CL) before and after treatment, and healthy individuals. The lymphoproliferative response was evaluated in cell culture using stimuli of the dichloromethane fraction (DCM) obtained from Libidibia ferrea, Glucantime® and phytohemagglutinin - PHA using a BrdU Cell Proliferation after 72 h of incubation. In cultures treated with the DCM fraction, intense induction of lymphoproliferation was observed (p<0.0001), as was also observed in response to the PHA mitogen, and there was a significant difference when compared with the conventional treatment (p<0.0135). In the post-treatment and healthy groups, although the compound induced lymphoproliferation, there was no statistical difference. These results suggest that the organic compound played an important inducing role in lymphoproliferation, which highlights the importance of continuity involving new studies in order to evaluate its immunomodulatory activity.

Keywords: Cell proliferation, leishmaniasis, natural products, therapeutic alternative.

Avaliação da linfoproliferação de células mononucleares de pacientes com leishmaniose cutânea frente a Libidibia ferrea

Resumo
A leishmaniose tegumentar americana (LTA) é uma doença negligenciada que acomete majoritariamente populações vulneráveis. Seu amplo espectro de manifestações clínicas é relacionado ao tipo de resposta imunológica produzida pelo hospedeiro e a espécie de Leishmania envolvida. Nos últimos anos, o uso de plantas medicinais vem sendo uma alternativa terapêutica no tratamento de doenças infectoparasitárias. Esta pesquisa teve como objetivo avaliar as respostas linfoproliferativas de células mononucleares do sangue periférico (PBMCs) de pacientes com leishmaniose cutânea (LC) antes e após tratamento e de indivíduos saudios. A resposta linfoproliferativa foi avaliada em cultura de células frente a estímulos da fração diclorometânica (DCM) obtida de Libidibia ferrea, Glucantime® e phytohemagglutinin - PHA utilizando BrdU Cell Proliferation após 72h de incubação. Nas culturas tratadas com a fração DCM foi observada em todos os grupos intensa indução da linfoproliferação (p<0.0001), apresentando diferença significativa ao comparar com o tratamento convencional no grupo de pacientes pós-tratamento (p<0.0135). Estes resultados sugerem que o composto orgânico desempenhou importante ação indutora na linfoproliferação ressaltando a importância da continuidade com novos estudos a fim de avaliar sua atividade imunomodulatória.

Palavras-chave: Proliferação celular, leishmaniose, produtos naturais, alternativa terapêutica.
Introduction

American tegumentary leishmaniasis (ATL) is classified by the World Health Organization (WHO) as a neglected tropical disease (NTD). In Brazil, it is considered a public health problem, whose case notification data from the Department of Informatics of the National Health System (SINAN, 2019) indicated 16,135 confirmed cases of ATL in 2019. The northern region had the highest incidence, with 7,123 cases of ATL in 2019, with 1,302 cases reported in the state of Amazonas. ATL is a disease caused by intracellular protozoa of the genus Leishmania (Kinetoplastida, Trypanosomatidae), with Leishmania (Leishmania) amazonensis, Leishmania (Vianna) guyanensis and Leishmania (Vianna) braziliensis being the main species that cause ATL in Brazil (Brasil, 2017).

In the Amazon, ATL manifests itself in the cutaneous, diffuse cutaneous and mucosal forms, with cutaneous leishmaniasis (CL) being the most prevalent in the state of Amazonas (Brasil, 2017). CL is characterized by a diversity of clinical forms, the lesions of which can be localized or disseminated. The variability of clinical presentations depend on the species of Leishmania involved in the infection and the development of the patient’s immune response (Brasil, 2017).

Regarding the mechanisms of immune regulation, there is a dichotomy between Th1 and Th2, since they induce important cytokine profiles that are capable of defining the clinical course of infection (Espirit al., 2014). In this context, the main human defense against leishmaniasis is its ability to adjust the cell-mediated immune response, which is demonstrated by lymphoproliferation and the production of high levels of IFN-γ and TNF-α, making it capable of controlling or even eliminating the parasite. Thus, lymphoproliferation is an important evaluation parameter, as it demonstrates the ability of a given cell or group of cells to proliferate in situations that may be adverse (Teixeira, Teixeira, Andrade, Barral-Netto, & Barral, 2006; Bacellar et al., 2002).

For more than 50 years, the drugs recommended by the Brazilian Ministry of Health for treating leishmaniasis have been composed of pentavalent antimonials (Sb⁵⁺) (Carvalho et al., 2019). Despite their effectiveness, Sb⁵⁺ presents a number of problems, such as parenteral administration, long duration of treatment, high toxicity and high cost. Thus, many patients who contract leishmaniasis abandon treatment, which is a factor that favors the appearance of resistant strains (Pradhan, Schwartz, Patil, Grabe, & Goldust, 2021).

Due to the high toxicity of the drugs that are currently used, new therapeutic alternatives have been studied, and Libidibia ferrea is one of the candidates. It is a plant that belongs to the Fabaceae family, and is popularly known in Brazil as “pau-ferro” or “jucá”. L. ferrea has a multiplicity of therapeutic uses and its different plant parts are used in the treatment of various diseases. Therefore, given its medical importance, the Brazilian Ministry of Health has included this species in the National List of Important Medicinal Plants for the Health System and indicates that it has analgesic, antipyretic, anti-inflammatory and/or antispasmodic (Brasil, 2006), and healing and antioxidant properties (Carvalho, Sampaio, Araujo, Pinto, & Rocha, 2016; Falcão et al., 2019).

Antileishmanial activity was observed by Comandolli-Wyrepkowski et al. (2017), who confirmed that L. ferrea presented efficiency in the treatment of CL in in vitro and in vivo experiments. In this sense, considering the therapeutic relevance of L. ferrea, this study aimed to evaluate the action of a fraction obtained from the methanolic extract of L. ferrea on the lymphoproliferation of PBMC of people exposed to Leishmania sp. and compare it to those not exposed, for the purpose of providing subsidies for new Leishmaniasis treatment strategies.

Materials and Methods

Plant material and fractionation

Samples of the fruit of L. ferrea were collected in Manaus, Amazonas, Brazil (3° 4’ 34” S 60° 0’ 11” W). These were deposited in the Herbarium of the National Institute for Amazonian Research – INPA and identified under the number 246150. The collected fruits were sorted and, after removing the seeds, were dried, pulverized and extracted with the solvents hexane and methanol (Comandolli-Wyrepkowski et al., 2017). The dichloromethane fraction (DCM) of the L. ferrea extract was obtained by liquid-liquid partition. An aliquot of 20 g of methanic extract solubilized in 200 mL of H₂O:MeOH (9: 1) was used and extracted three times with each of the solvents used in the partition (hexane and dichloromethane). After this process, the fraction was concentrated in a rotary evaporator (YAMATO®).

Ethical aspects

This work was approved by the Ethics Committee for Research with Human Beings (CEP) of the Federal University of Amazonas (UFAM) - CAAE/UFAM: 29406319.2.0000.5020. All the volunteers who agreed to participate in the study signed the informed consent form.

Sample population

The biological samples used in this study were obtained from 30 people from the municipality of Rio Preto da Eva (2° 41’ 55” S, 59° 42’ 3’W), Amazonas state, Brazil, who sought treatment at a basic healthcare unit (UBS), in the period from March to June 2020. The participants in this study were divided into three groups: before treatment, after treatment and negative control, according to Table 1.

All the primoinfected patients underwent a physical examination and the diagnosis for CL was based on the detection of Leishmania sp. amastigotes, from scarification of the inner edge of the lesion and smears stained with rapid the Panotic Kit (Laborclin™). Samples of the edge of the lesion of each patient were inoculated in NNN (Neal, Novy and Nicolle) culture media for parasitological confirmation of promastigote forms of the parasite.

Blood sampling and PBMC extraction

Around 8 mL of heparinized blood was collected from all study participants. The blood samples were transferred to
centrifuge tubes, diluted in complete RPMI (Roswell Park Memorial Institute) medium at a 1:1 ratio and had 8 mL Histopaque™ 1077 carefully added. The tubes were centrifuged at 2,200 rpm for 30 minutes. The peripheral mononuclear cells (PBMCs) were collected and transferred to centrifuge tubes, washed twice with RPMI medium, and then centrifuged at 1,800 rpm for 10 minutes at 4 °C. The PBMCs were resuspended in 1 mL of complete RPMI medium and counted in the Neubauer chamber with the aid of Trypan blue. The concentration of PBMCs was adjusted to 2 x 10⁶ cells/mL in complete RPMI medium.

**Table 1.** Participants in the study from Rio Preto da Eva, Amazonas, Brazil organized by group and the inclusion criteria used in the research.

| Group | Inclusion criteria |
|-------|--------------------|
| Before treatment (primoinfection) | Untreated patients with first *Leishmania* infection |
| After treatment | Patients post Glucantime® treatment for cutaneous leishmaniasis with healed lesions. Blood collection of this group was performed after full healing of the lesion and clinical evaluation to ensure that the lesion was no longer active, at least 30 days and up to 12 months after treatment. |
| Negative control | Healthy individuals, without clinical manifestations or lesions caused by *Leishmania* spp. The healthy individuals were from the metropolitan area of Manaus, and epidemiological questionnaire was applied to ensure that these individuals had not been in an endemic area in recent months. |

The number of participants in each group was n = 10.

**In vitro cell stimulation assays**

The PBMCs were transferred to culture microplates of flat-bottomed 96-well plates; each well containing 100 µL/PBMC [2x10⁵]. The tests were individually carried out with the PBMC samples from each participant. The wells of the microplates, with 100 µL/well fr each test substance (final volume of 200 µg/well), were organized in triplicate as follows: wells containing cells stimulated with phytohemagglutinin-PHA [10 µg/mL], considered the positive control for lymphoproliferative response; wells without stimulation (with only RPMI medium) were considered the negative control; wells with cells treated with Glucantime® [100 µg/mL]; and wells with cells treated with the DCM fraction [120 µg/mL].

Prior to this, both substances were dialyzed in RPMI medium, and the DCM fraction was first dissolved in DMSO-dimethylsulfoxide (CH₃)₂SO (1%). The PBMCs were incubated for 72 h at 37 °C (5% CO₂; 95% humidity) and, after 18 hours of incubation, BrdU (5-bromo-2'-deoxyuridine) [10 µM/mL] was added. An ELISA (enzyme-linked immunosorbent assay) colorimetric test was performed according to the manufacturer’s instructions (BrdU Cell Proliferation ELISA Kit, Biotrak, Amersham, UK).

Lymphoproliferation was evaluated based on the quantification of BrdU incorporation, by means of the average increase in optical density through spectrophotometry (absorbance of 450 nm). Estimates of cell proliferation were determined as a relative value between the arithmetic mean of positive and negative control triplicates (white, well with RPMI medium), in relation to wells with stimulated cells.

**Statistical analysis**

The statistical analysis was carried out using GraphPad Prism® software v.6.0 for Windows, for which the significance (p<0.05) was calculated using parametric tests. A one-way ANOVA statistical test was used for comparisons among groups of study subjects and the Tukey test was used for comparisons among the drug treatments.

**Results and Discussion**

During the treatment, epidemiological investigations and clinical evaluations were performed on the 20 patients diagnosed with CL. Of these, ten were positive direct examination primoinfected patients, and ten were patients who had achieved clinical cure, which was evaluated according to wound healing, after treatment with Glucantime®. There was a predominance of CL in males (85%), and 76.47% were in the age group between 19 and 56 years. The higher prevalence of males, who are young and are involved in forest extractivism are part of the epidemiological profile of ATL in the Brazilian Amazon (Figueira et al., 2017; Hernández & Núñes, 2019).

Regarding the CL lesions, 91.30% were ulcerated lesions, with a mean size of 2.0 cm, 62.86% of which were located in the lower limbs and 13.04% in the upper limbs and trunk (Figure 1). Figure 2A, B and C shows the results of the lymphoproliferation response in each group, considering the positive control (PHA mitogen) and the negative control (no stimulation).

![Figure 1](image_url) - Description: Figure 1 shows the distribution of CL lesions among the study population.

![Figure 2A](image_url) - Description: Figure 2A shows the results of the lymphoproliferative response in the primoinfected group.

![Figure 2B](image_url) - Description: Figure 2B shows the results of the lymphoproliferative response in the group treated with Glucantime®.

![Figure 2C](image_url) - Description: Figure 2C shows the results of the lymphoproliferative response in the group treated with DCM fraction.

Of the 30 subjects involved in this study, mononuclear cells from 21 participants (7 samples/group) were analyzed in vitro. The lymphoproliferative response was expressed in high rates (≥60%) (Figure 2B) and negative control (p=0.0624) (Figure 2C) groups with the stimulus by the DCM fraction, conventional treatment with Glucantime® and PHA mitogen.

The lymphoproliferation of PBMC was strongly positive (100%) and significant in response to PHA mitogen (Figure 2) and was verified in all groups of participants. PHA mitogen is commonly used as a lymphoproliferation enhancer and has been used in other studies (Reis et al., 2009; Mohammadi, Duthie, Reed, Javadi, & Khamesipour, 2021).
confirmed against *L. (L.) amazonensis* and *L. (V.) guyanensis* and did not present cytotoxic effects in murine macrophages (Comandolli-Wyrepkowski *et al*., 2017). To date, the mechanism of action of the pharmacological components of *L. ferrea in vivo* is not yet known. However, promising effects of *L. ferrea* for the treatment of CL were observed by Comandolli-Wyrepkowski *et al.* (2017), when submitting Golden hamsters (*Mesocricetus auratus*) that were infected with *L. (L.) amazonensis* and *L. (V.) guyanensis* to topical treatment with a hydrogel containing the DCM fraction of *L. ferrea*. Control of the growth of the CL lesions in animals and low parasitic load in culture analyses were noted after treatment.

The phytochemical prospecting of the extract of *L. ferrea* showed the presence of phenolic compounds and flavonoids in greater evidence (Comandolli-Wyrepkowski *et al*., 2017). Phenolic compounds are recognized as being involved in anti-inflammatory and antioxidant activity, and act directly on specialized cells of the immune system (Hakime-Silva *et al*., 2013). Vila-Nova *et al.* (2012) demonstrated leishmanicidal activity of the phenolic compound coumarin (scoporone) isolated from *Platymiscium floribundum*, and of the flavonoids (rutin and quercetin) that were isolated from *Dimorphandra gardneriana*. These are both plants that are native to the Caatinga biome of Brazil, and in this case were against *Leishmania infantum*, which are the protozoa that cause visceral leishmaniasis. These compounds were able to inhibit the enzyme acetyl-cholinesterase (AChE) and, consequently, disrupt the biosynthesis of phosphatidylycholine, an essential constituent of the surface membranes of the parasite.

Among some of the important phenolic compounds found in *L. ferrea* is gallic acid. Associating these phenolic compounds and amphotericin B, Alves (2019) demonstrated the activation of macrophages and a reduction in amastigotes through macrophage, indicating therapeutic action and activation of the immune response, and leading to the healing of lesions in *Mus musculus* lineage BALB/c. Some studies with herbal compounds, such as *Campsiandra laurifolia* (Chagas, Henrique Müller, Soares & Garcez, 2010), *Portulaca hirsutissima* and *Portulaca werdermannii* (Costa *et al*., 2007), *Nelumbo nucifera* (Khan *et al*., 2016), did not demonstrate direct action on the parasitic agent, but rather, as potential anti-inflammatory treatments with high immunomodulatory capacity in *Leishmania* sp. infections.

Considering the PBMCs from all the groups showed a significant lymphoproliferative response (≥77%; *p*<0.0001) to the dichloromethane fraction (DCM) (Figure 2). In particular, the group of post-treatment patients, in particular, presented almost 100% proliferated cells induced by DCM. Thus, the DCM fraction induced more cell proliferation than Glucantime®, which was different to that observed for the primoinfected and negative control groups, since the values between the substances were equivalent.

This result of expressive lymphoproliferation may be related to the therapeutic biological potential of the fraction of *L. ferrea* obtained, whose antileishmanial activity was

![Figure 1](image1.png) **Figure 1.** Clinical aspects of lesions in male patients primo-infected with cutaneous leishmaniasis (CL) treated in Rio Preto da Eva, Amazonas, Brazil, during March to May 2020. IM: Registration code.

![Figure 2](image2.png) **Figure 2.** PBMC lymphoproliferation of participants evaluated in Rio Preto da Eva, Amazonas state, Brazil, during March and May 2020. A - primo-infected patients with cutaneous leishmaniasis; B - post-treatment cutaneous leishmaniasis patients; C - healthy people without cutaneous leishmaniasis. DCM: dichloromethane fraction; PHA: phytohemagglutinin. Vertical lines represent standard error. *Horizontal lines represent statistical significance between treatments (p<0.005).
stirulated with total antigen of \textit{L. (V.) braziliensis}. The authors pointed out that in an \textit{in vivo} study, T cell stimulation is expected to decrease as treatment progresses to cure, since the reduction in parasitic load can cause the number of responsive T cells to decrease. It is worth remembering that in both studies, blood collection of post-treatment patients was performed at least 30 days after the end of Glucantime\textsuperscript{®} treatment.

The toxicity of meglumine antimoniate goes far beyond the adverse effects on the vital organs of CL patients while being treated. Lopez (2015) observed that antimoniate interfered in the process of cell proliferation, in addition to contributing cytotoxic, genotoxic and mutagenic effects in human leukocytes exposed to different concentrations of Glucantime\textsuperscript{®}. Given this, the combination therapy of potential drugs and antimoniate have numerous advantages, such as reduced toxicity, ease in the treatment regimen and reduced development of resistance to this drug.

Lymphoproliferation is considered an important factor in the control of infection or even in the elimination of Leishmania sp. (Brasil, 2017). Therefore, the discovery of phytopharmaceuticals with immunomodulatory capabilities are crucial for the development of alternative treatments or for combination with recommended drugs, in order to provide a potential synergism and therapeutic success in treating leishmaniasis. This study contributes to a more diligent look at the importance of the immunomodulatory effect of \textit{L. ferrea} that may favor the healing of lesions in patients with CL. However, for the best elucidation of the regulation of inflammation induced by the DCM fraction, it is necessary to perform cytokine dosage to understand these immune mechanisms.

Conclusion

These findings suggest that the organic fraction DCM induced stimulatory action on activation and lymphoproliferation when incubated with the PBMC of patients before and after treatment for CL, which may, at least in part, be due to an immunomodulatory response. This immunomodulatory response needs to be highlighted and better investigated in future trials in order to strengthen the data presented here in order to demonstrate which molecular mechanisms are involved and the stimulatory effect of the immune response caused by the dichloromethane fraction of \textit{L. ferrea}, a phytopharmaceutical candidate for the alternative treatment of cutaneous leishmaniasis.

Acknowledgments

The authors would like to thank the Amazonas State Research Support Foundation - FAPEAM (POSGRAD) for its financial support, and the Edible Fungi Cultivation Laboratory for its collaboration with the spectrophotometry experiments.

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