Monocyte and Blood Interleukin-12 Levels in Patients with Obstructive Jaundice

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Patients with obstructive jaundice have an increased incidence of peri-operative complications and immune dysfunction. This study was to investigate interleukin (IL)-12 (a cellular immunity stimulant), levels in jaundiced patients. 23 jaundiced patients and 17 controls were studied. There was significantly increased monocyte IL-12 production in jaundice, as measured by an ELISA, compared with that in controls (p<0.01 by Mann-Whitney U test). A similar increase is seen in both benign and malignant jaundice. There was no difference between plasma IL-12 levels in the two groups. It is concluded that in jaundice, monocytes have a significantly increased capacity to release IL-12. This suggests that IL-12 may play a role in the immune dysfunction in jaundiced patients.

KEY WORDS: Interleukin-12 obstructive jaundice monocytes

INTRODUCTION

Patients with obstructive jaundice are associated with an increased incidence of perioperative complications, such as renal failure, sepsis, haemorrhagic disorders and endotoxaemia1-3. For some years it has been known that these patients have depressed Reticulo-Endothelial system (RES) phagocytic functions5-8. It has been reported recently that the secretory functions from monocyte and neutrophils, for example, superoxide production, cytokine secretions, and lysosomal enzyme secretions are increased, and this may be related in some way to a patient's clinical outcome9-12. In other studies, it has been shown that patients with jaundice demonstrate lymphocyte activation as indicated by increased IL-2 receptor levels13. Interleukin-12, otherwise known as natural killer stimulatory factor (NKSF), is a recently defined cytokine which has major effects on both lymphocytes and NK cells. Both proliferation and function of these two cell types are stimulated by IL-1214-19. The monocyte/macrophage is a key cell type in immune regulation. It produces an array of cytokines including IL-1, IL-6, IL-8, TNF alpha. This cell is also a key contributor to IL-12 in vivo20. A markedly raised production of IL-1, IL-6, and TNF from monocytes has been shown to be related to patients' poor prognosis in sepsis, trauma, and burn injuries21-23.

In order to determine the monocyte cytokine production and the possible relationship to cellular immunity, we have studied the monocyte production and blood level of interleukin-12.

PATIENTS, METHODS, AND RESULTS

23 patients with biliary obstructions (median age 71 years, 14 male, 9 female) were studied. Of these patients 14 had malignant diseases (cholangiocarcinoma or pancreatic cancer) and the others gall stones or benign biliary strictures. 17 controls were studied (10 had herniae or gall stones without jaundice, 7 colon cancer without liver metastasis, median age 64 years).

Peripheral blood was taken before any clinical intervention. The blood was processed at 4°C. Plasma was obtained by centrifugation at 2500rpm for 25 minutes. Monocytes were separated by a standard Ficoll-Hypaque separation and purified by adherence techniques. The monocyte population was assessed by...
a non-specific esterase stain (monocyte population was over 90% after adherence). Cells were stimulated in vitro with lipopolysaccharide (10μg/ml) for 24 hours and cell free-supernatant were stored at -80°C for later assay.

IL-12 was measured using a specific ELISA with monoclonal 20C2 as capture antibody and R7926 as detecting antibody (Kindly supplied by Genetic Institute, Cambridge, USA). Secondary antibody was horse-reddish peroxidase conjugated IgG (Biorad) and the colour was developed using orthophenylene-diamine (OPD) and absorbance measured at 492 nm on a Tiertek Multiscanner. Recombinant human interleukin-12 was used as standard.

There was a significantly increased IL-12 production in jaundice, 205.6±2.8 pg/ml (mean±SEM), compared with 182.2±4.2 pg/ml in controls (p < 0.01 by Mann-Whitney U test, figure 1A). When jaundiced patients were separated into groups with either benign or malignant disease, a similar increase is seen in both jaundiced groups.

There was no difference between plasma IL-12 levels in the jaundiced and control groups (170.0±4.5 pg/ml and 171.0±8.0 pg/ml respectively, p > 0.05, figure 1B). The differences between benign and malignant in control or that in jaundiced are also not significant. Monocyte IL-12 level in jaundiced patients had no correlation with the plasma bilirubin, bile salt, or aspartate aminotransferase.

**DISCUSSION**

This paper describes a previously unknown phenomenon, patients with biliary obstruction have a raised monocyte interleukin-12 production.

IL-12 is a recently identified cytokine which is produced by lymphocyte and monocyte/macrophage cells, It has major effects on lymphocyte and natural killer (NK) cells (also known as NK cell stimulatory factor, NKSF). This cytokine therefore has an important role in natural and cellular immunity. It has been reported recently that IL-12 may be in the future have some promise as an agent in the treatment of certain infections, such as leishmaniasis.

Obstructive jaundice is associated with increased incidence of perioperative complications. It is also established that these patients frequently suffer from endotoxaemia. Both local and systemic endotoxemia may therefore be responsible for the immune malfunction in jaundiced patients.

It is not clear by which pathway monocytes increase their production in these patients. It has been reported that IL-12 shares structural homology with IL-6, a proinflammatory cytokines. IL-6 production from 'jaundiced' monocyte is reported to be increased and this increase may be due to endotoxaemia which occurs in jaundiced patients. Increased IL-12 production may therefore share a similar pathway, this deserves further studies.

In summary, peripheral blood monocyte from obstructive jaundiced patients showed a greatly increased IL-12 production in both benign and malignant aetiologies and this may contribute to the immune malfunction in these patients.

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**REFERENCES**

1. Pain JA, Cahill CJ, Bailey M.E. (1985). Perioperative complications in obstructive jaundice: therapeutic considerations. *Br J Surg.* 72: 942–945.
2. Wait RB, Kahng K.V. (1989). Renal failure complicating obstructive jaundice. *Am J Surg.* 157: 256–263.
3. Allison M.E.M, Prentice CRM, Kennedy A.C. and Blumgart L.H. (1977). Renal function and other factors in obstructive jaundice. *Br J Surg.* 66: 392–397.

4. Dixon J.M, Armstrong C.P, Duffy R.A. and Davies G.C. (1984). Upper gastrointestinal bleeding. A significant complication after surgery for relief of obstructive jaundice. *Ann Surg.* 199: 271–275.

5. Halpeax B.N, Biozzi G, Nicol T. and Bilbby D.L.J. (1957). Effect of experimental biliary obstruction on the phagocytic activity of the reticulo-endothelial system. *Nature.* 200: 503–504.

6. Drivas G, James O, Wardle N. (1976). Study of reticulo-endothelial phagocytic capacity in patients with cholestasis. *Br Med J,* 1: 1568–1569.

7. Clements W.D, Halliday M.I, McCaigue M.D, Barclay R.G. and Rowlands B.J. (1993). Effects of extrapancreatic obstructive jaundice on Kupffer cell clearance capacity. *Arch Surg.* 128: 200–205.

8. Pain J.A. (1987). Reticuloendothelial function in obstructive jaundice. *Br J Surg.* 74: 1091–1094.

9. Jiang W.G and Puntis M.C.A. (1993). Monocyte and blood hexosaminidase in patients with obstructive jaundice. *Hepatol.* 7: 15–23.

10. Semeraro N, Montemurro P, Chetta G, Altonare D.F, Giordano D. and Colucci M. (1989). Increased procoagulant activity of peripheral blood monocyte in human and experimental obstructive jaundice. *Gastroenterol.* 96: 892–899.

11. Levy R, Schlaeffel F, Keynan A, Nagauer O, Yaari A. and Sikuler. (1993). Increased neutrophil function induced by bile duct ligation in a rat model. *Hepatol.* 17: 908–914.

12. Bemelman M.H.A, Buurman W.A, Gouma D.J. (1993). The presence of TNF and soluble TNF receptors in obstructive jaundice. *Gastroenterol.* 104: suppl, A665.

13. Wagner F, Assemi C, Lersch C, Hart R. and Classen M. (1990). Soluble interleukin-2 receptor and soluble CD8 in liver cirrhosis and obstructive jaundice. *Clin Exp Immunol.* 82: 344–349.

14. Kobayashi M, Fitz L, Ryan M, Hewick RM, Clark S.C, Chan S, Loudon S, Sherman F, Perussia B, Trinchieri G. (1989): Identification and purification of natural killer cell stimulatory factor (NKSF), a cytokine with multiple biologic effects on human lymphocytes. *J Exp Med* 170: 827–845.

15. Wolf S.F, Temple P.A, Kobayashi M, Young D, Diég M, Lowe L, Dzialo R, Fitz L, Ferenz C, Hewick R.M. et al. (1991): Cloning of cDNA for natural killer cell stimulatory factor, a heterodimeric cytokine with multiple biologic effects on T and natural killer cells. *J Immunol* 146: 3074–3081.

16. Gately M.K, Wolitzky A.G, Quinn P.M, Chizzonite R. (1992) Regulation of human cytotropic lymphocyte responses by interleukin-12. *Cell Immunol* 143: 127–142.

17. Manetti R, Paronchi P, Giudici M.G, Pecchini M.P, Maggi E, Trinchieri G. Romagnani S. (1993): Natural killer cell stimulatory factor (interleukin 12 [IL-12]) induces T helper type 1 (TH1)-specific immune responses and inhibits the development of IL-4-producing Th cells. *J Exp Med* 177: 1199–1204.

18. Perussia B, Chan S.H, D’Andrea A, Tsuji K, Santoli D, Pospisil M, Young D, Wolf S.F, Trinchieri G. (1992): Natural killer (NK) cell stimulatory factor or IL-12 has differential effects on the proliferation of TCR-alpha beta+ TCR-gamma delta+ T lymphocytes, and NK cells. *J Immunol* 149: 3495–3502.

19. Robertson MJ, Soiffer RJ, Wolf SF, Manley TJ, Donahue C, Young D, Herrman S.H, Ritz J. (1992): Response of human natural killer (NK) cells to NK cell stimulatory factor (NKSF): cytotropic activity and proliferation of NK cells are differentially regulated by NKSF. *J Exp Med* 175: 779–788.

20. D’Andrea A, Rengaraju M, Valiante N.M, Chehimi J, Kubin M, Aste M, Chan S.H, Kobayashi M, Young D, Nickbarg E. et al. (1992): Production of natural killer cell stimulatory factor (interleukin 12) by peripheral blood mononuclear cells. *J Exp Med* 176: 1387–1398.

21. Pain J.A. and Bailey M.E. (1986). Experimental and clinical study of lactulose in obstructive jaundice. *Br J Surg.* 73: 775–778.