COVID-19

Lung Ultrasound and Respiratory Pathophysiology in Mechanically Ventilated COVID-19 Patients—an Observational Trial

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Abstract

The relationship between respiratory system mechanics, lung ultrasound (LUS) abnormalities, and mortality in mechanically ventilated patients with COVID-19-associated respiratory failure is unknown. We assessed the pattern of respiratory mechanics and LUS, their changes over time, and the differences between survivors and non-survivors. We additionally analyzed the relationship between LUS findings and the severity of gas exchange impairment and interleukin 6 (IL-6). This was a two-center retrospective, observational trial carried out in the intensive care units of the hospitals of Bolzano and Merano, Italy, from March 15 to April 20, 2020. We enrolled 41 consecutive patients. Seven patients (17%, 95% CI 8.5–31.3%) died. Mean compliance of the respiratory system on ICU admission was 41.6 (± 18.8) ml/mbar (42.5 (± 19.6) for survivors, 38.0 (± 16.3) for deceased, \( p = 0.605 \)). Non-survivors had a significantly lower compliance over time, decreasing from day 14 after symptom onset, compared with survivors (\( p = 0.008 \)). Mean LUS score on admission was 11.2 (± 3.7) and survivors had lower LUS scores on admission than non-survivors (10.5 (± 3.6), 13.9 (2.8), respectively, \( t \) test, \( p = 0.029 \)). LUS score correlated with IL-6 concentrations (\( r = 0.52, p = 0.001 \)) and arterial pCO\(_2\) (\( r = 0.30, p = 0.033 \)) and was inversely correlated with oxygenation (\( r = -0.34, p = 0.001 \)). No correlation was found between LUS and respiratory system compliance (\( r = -0.02, p = 0.299 \)). Non-survivors from COVID-19-associated respiratory failure had a significant decrease in compliance after day 14 of symptom onset. Compliance did not correlate with the degree of abnormalities found in LUS, but LUS score correlated with oxygenation, pCO\(_2\), and IL-6.

Keywords COVID-19 · ARDS · Mechanical ventilation · Lung ultrasound · Respiratory mechanics

Background

A new form of viral pneumonia caused by the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) was identified in Wuhan, China, in December 2019 [1–3]. Due to its transmission dynamics, the virus spread quickly across the globe within a very short time and on March 11, 2020, the WHO declared the outbreak a pandemic [4]. Studies from

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China reported a high incidence of critical illness (23–32%) and acute respiratory distress syndrome (ARDS) (17–29%) among hospitalized patients [1, 5–7].

The pathophysiology of coronavirus disease 2019 (COVID-19)-associated respiratory failure is still the subject of debate and ongoing research. Even though most COVID-19 patients needing mechanical ventilation fulfil the Berlin-criteria [8], severe respiratory failure due to COVID-19 pneumonia is distinct from classical ARDS and (at least at the early stage of the disease) characterized by a dissociation between relatively well-preserved lung mechanics and the severity of hypoxemia.Gattinoni et al. suggested that two COVID-19 phenotypes can be distinguished: type L, characterized by a maintained lung compliance, near normal lung weight, and low lung recruitability, and type H, characterized by a poor lung compliance, an increase in lung weight, and high lung recruiting capacity [9]. Type L can progress to type H as the disease progresses and depending on the phenotype, different treatment strategies might be adapted [9]. Yet, further evidence is needed to prove this hypothesis. To date, little is known about the changes of respiratory system compliance over time in mechanically ventilated COVID-19 patients and the relationship between respiratory system mechanics and mortality.

Lung ultrasonography (LUS) allows rapid and reliable diagnosis of interstitial–alveolar syndrome, lung consolidation or pleural effusion, and is superior to chest radiography [10]. When evaluating patients with pneumonia and/or ARDS, LUS gives results similar to chest CT [11]. During the COVID-19 pandemic, point of care LUS has been widely adopted and characteristic findings of COVID-19 pneumonia have been described [12–15]. However, little is known about the relationship between LUS findings and the severity of gas exchange impairment and between LUS and lung mechanics in COVID-19 pneumonia. Moreover, it is unknown whether LUS abnormalities are linked to the degree of systemic inflammation (e.g., assessed by interleukin 6 (IL-6) plasma concentration) or to mortality.

Our study aimed at assessing the pattern of respiratory mechanics and LUS in mechanically ventilated patients with COVID-19-associated respiratory failure, their changes over time, and the differences between survivors and non-survivors. We additionally analyzed the relationship between LUS findings and the severity of gas exchange impairment and IL-6.

Methods

In this observational trial, the medical records of consecutive patients admitted to the intensive care units (ICUs) of the hospitals of Bolzano and Merano, South Tyrol, Italy, from March 15 to April 20, 2020, due to COVID-19-associated respiratory failure were retrospectively analyzed. The study was approved by the Ethics Committee of South Tyrol (protocol number 46-2020). Study design and reporting were performed according to the Strengthening the Reporting of Observational Studies in Epidemiology (STROBE) statement [16].

Clinical Management

All patients were treated at the discretion of the attending physicians. COVID-19-associated respiratory failure was managed according to the current guidelines [17], including lung protective ventilation, prone positioning, use of neuromuscular blocking agents (if indicated), and a conservative fluid balance. Laboratory tests were performed according to the clinical indications and protocols in use in each institution.

Data Collection

Ventilatory parameters, LUS data, and laboratory values were routinely recorded in the ICU patient data managing system. The following data were extracted and stored anonymously: sex, age, mode of ventilation, PEEP, driving pressure, respiratory system compliance, $\text{PaO}_2$, $\text{FiO}_2$, $\text{PaCO}_2$, IL-6 plasma concentration (Elecsys® IL-6, Roche Diagnostics, Espoo, Finland), LUS score, ICU length of stay (ICU-LOS), and outcome (alive or deceased). Respiratory system compliance was calculated in patients under controlled mechanical ventilation as tidal volume (TV) / (plateau pressure – PEEP).

Lung Ultrasound

The scan was performed with the patient in supine position. A convex probe with abdominal preset was selected, without compound and with general frequency setting. Focus was set at the pleural line, depth at 6–8 cm, and gain low (i.e., to see pleural sliding). The two ultrasound scanners used (MyLab30 Esaote spa, Genova, Italy, and X-porte Fujifilm-Sonosite Inc., Amsterdam, The Netherlands) were dedicated exclusively to COVID-19 patients. The probe and hardware were cleaned after each use according to the local protocols. In order to obtain the LUS score, the surface of each hemithorax was divided into four lung fields [18]. For each lung field, a numerical score was given: 0 for A lines, 1 for well separated B lines, 2 for coalescent B lines, and 3 for the presence of parenchymal consolidations (aerial bronchogram or atelectasis). We used the sum of the score assigned to each lung field on both sites (i.e., maximal score of 24 points) for analysis. The LUS examination was done routinely and not driven by a clinical indication (e.g., a deterioration of gas exchange).
Statistical Analysis

We presented data as percentages for counted and as mean and standard deviation (SD) for numerical data. IL-6 values were log transformed to achieve normality prior to analysis. Group comparisons for baseline values or mean values over all time points were carried out by means of the Chi-square test or the t-test. Linear mixed models (LMM) were fitted to the data when utilizing all time points using the patient as a random effect with a random intercept and the fixed effects status, hospital, time (grouped in quintiles) and the covariate LUS. Additionally, correlation coefficients between continuous variables have been assessed by means of the partial correlation coefficient controlling for patient and hospital. For analysis of changes over time, the day of symptom onset rather than the timepoint of ICU admission was considered as baseline (i.e. day 1). All p values are two-sided and values below 0.05 have been considered significant. SPSS (IBM version 26.0) was used for statistical analysis.

Results

Patient Characteristics

We included 41 consecutive patients. All patients were intubated and mechanically ventilated on ICU admission. Table 1 summarizes patients’ demographics, comorbidities, respiratory parameters, and laboratory values on admission, divided between survivors and non-survivors.

Table 1  Baseline demographic characteristics, comorbidities, respiratory parameters, and laboratory values, divided between survivors and non-survivors. We revealed these data on admission

|                                | Survivors n = 34 (82%) | Non-survivors n = 7 (17%) | Overall n = 41 | p value |
|--------------------------------|------------------------|---------------------------|----------------|---------|
| Age—mean (SD)                  | 59.4 (11.8)            | 73 (5.1)                  | 61.7 (12.1)    | 0.005*  |
| Sex—no. (%)                    |                        |                           |                |         |
| Male                           | 26 (81%)               | 6 (19%)                   | 32 (78%)       | 0.591+  |
| Female                         | 8 (29%)                | 1 (11%)                   | 9 (22%)        |         |
| BMI—mean (SD)                  | 27.1 (4.3)             | 25.3 (5)                  | 26.7 (4.4)     | 0.335*  |
| Enrolment site                 |                        |                           |                |         |
| Bolzano—no. (%)                | 15 (79%)               | 4 (21%)                   | 19 (46%)       | 0.529*  |
| Merano—no. (%)                 | 19 (86%)               | 3 (14%)                   | 22 (54%)       |         |
| Comorbidities—no. (%)          |                        |                           |                |         |
| Arterial hypertension          | 13 (32%)               |                           |                |         |
| Coronary artery disease        | 1 (2%)                 |                           |                |         |
| Atrial fibrillation            | 2 (5%)                 |                           |                |         |
| Diabetes                       | 5 (12%)                |                           |                |         |
| COPD                           | 4 (10%)                |                           |                |         |
| Chronic kidney disease         | 5 (12%)                |                           |                |         |
| History of smoking             | 5 (12%)                |                           |                |         |
| Obesity                        | 5 (12%)                |                           |                |         |
| History of malignancy          | 7 (17%)                |                           |                |         |
| None                           | 11 (27%)               |                           |                |         |
| Time (days) from symptom onset to ICU admission—mean (SD) | 9.8 (3.7) | 12.8 (9.8) | 10.2 (5) | 0.167* |
| Length of ICU stay—days, mean (SD) | 17.8 (7.7) | 14.9 (10.5) | 17.3 (8.2) | 0.399* |
| Number of prone positionings during ICU stay—mean (SD) | 1.9 (1.8) | 2.0 (1.4) | 1.9 (1.7) | 0.891* |
| PaO2/FiO2 on day 1 of ICU admission—mean (SD) | 183.1 (71) | 185.3 (61.3) | 183.4 (68.7) | 0.940* |
| PaO2 on day 1 of ICU admission—mean (SD) | 90.4 (24.2) | 77.2 (22.7) | 88.4 (24.2) | 0.221* |
| PaCO2 on day 1 of ICU admission—mean (SD) | 46.9 (15.5) | 41.1 (11) | 46 (14.9) | 0.394* |
| Respiratory system compliance on day 1—ml/mbar, mean (SD) | 42.5 (19.6) | 38.0 (16.3) | 41.6 (18.8) | 0.605* |
| PEEP on day 1 of ICU admission—mbar, mean (SD) | 13.3 (2.2) | 13.7 (1.9) | 13.3 (2.2) | 0.625* |
| IL-6 plasma concentration on admission—pg/ml, mean (SD) | 530.8 (1545) | 552.9 (763.7) | 534.3 (1433.2) | 0.304* |

*p test
+ Chi-square test
Respiratory System Compliance

Mean respiratory system compliance on day 1 of ICU admission was 42.5 (± 19.6) ml/mbar for survivors and 38.0 (± 16.3) ml/mbar for non-survivors. We found no significant difference between survivors and non-survivors on admission (t test, p = 0.605) or as mean over all time points (t test, p = 0.066, Fig. 1a). However, when comparing compliance over time, we revealed that non-survivors had a significantly lower compliance from day 14 after symptoms onset compared with surviving patients (p = 0.008; Fig. 1b).

Lung Ultrasonography

Of the 41 patients included in this study, 34 underwent regular LUS examinations with a median of 5 LUS exams per patient during the ICU stay (i.e., an exam about every 72 h). Mean LUS score on admission was 11.2 (± 3.7) and was significantly different between survivors and non-survivors (10.5 (± 3.6), 13.9 (2.8), respectively, t test, p = 0.029). Non-survivors had higher but not significantly different mean LUS scores over all time points than survivors (12.9 ± 4.6, 9.9 ± 3.4, respectively, t test, p = 0.061, Fig. 2a). In contrast to the respiratory system compliance, there was no time-dependent effect when comparing these two groups (LMM status effect, p = 0.141, Fig. 2b).

We found a significant inverse correlation between the LUS score and the PaO₂/FiO₂ ratio (r = −0.34, p = 0.001, Fig. 3a). Furthermore, we found a significant correlation between the LUS score and PaCO₂ (r = 0.30, p = 0.033, Fig. 3b) and a significant correlation between LUS score and IL-6 plasma levels (r = 0.52, p = 0.001, Fig. 3c). We found no correlation when comparing LUS and compliance (r = −0.02, p = 0.299).

Discussion

In our study, we describe a series of 41 consecutive, mechanically ventilated patients with COVID-19-associated...
respiratory failure. Overall mortality rate was 17% (95% CI 8.5–31.3%) with non-survivors being significantly older than survivors. We found that mechanically ventilated ICU patients not surviving COVID-19 had a lower compliance after 2 weeks from symptom onset and a higher LUS score compared with survivors. Moreover, we showed that LUS score is a potential indicator of respiratory function and inflammatory status, as it correlates with gas exchange and IL-6 plasma concentration.

COVID-19-associated respiratory failure differs from classical ARDS. Though the Berlin ARDS criteria [8] are often met at presentation, a number of characteristics are atypical, such as a near normal lung volume with normal respiratory mechanics and compliance [9]. Gattinoni et al. named this initial presentation phenotype L. Yet, some patients undergo transition from phenotype L to phenotype H, the latter resembling classical ARDS with reduced lung volume, increased lung weight, and decreased compliance [9]. However, more than two phenotypes, type L and H might reflect two stages of the disease; COVID-19 pneumonia can over time develop classical ARDS, like any pneumonia caused by other pathogens [19]. This is in line with our findings, showing that some patients continue to have normal lung mechanics while others develop a reduced compliance approximately 2 weeks after symptom onset. This time point corresponds to about day 4 of ICU admission as mean time from symptom onset to ICU admission in our cohort is 10 days. Expectedly, patients with a decrease in lung compliance over time, i.e., patients developing classical ARDS, have a higher mortality rate than patients with preserved lung mechanics, i.e., patients with COVID-19 pneumonia without classical ARDS. Our findings on temporal changes in lung mechanics and their relationship to disease severity are in agreement with changes in CT findings over time found by Li et al. [20]. They described that pulmonary lesions with extensive ground-glass opacities and consolidations generally peaked around 14 days after the onset of the disease.
Chest CT scan is the gold standard imaging modality for pneumonia and ARDS [21]. CT scan is not universally available during a pandemic of a highly infectious disease with a large number of patients requiring hospital and intensive care unit admission. LUS, on the other hand, is a nearly universally available, a highly reproducible bedside technique with a short learning curve, and a reliable tool to identify interstitial syndrome, lung consolidation, lung collapse, and pleural effusion [10, 22–26]. Recently, its utility for diagnosis and follow-up of patients with COVID-19 pneumonia has been shown [12, 27, 28]. We found a significant correlation between the degree of abnormality in LUS (expressed as LUS score) and abnormalities in pulmonary gas exchange: The higher the LUS score, the lower the PaO₂/FiO₂ ratio and the higher the PaCO₂. Thus, the LUS findings of a severe interstitial syndrome and/or of consolidations are predictive of a ventilation/perfusion mismatch with shunt and dead space ventilation, leading to hypoxemia and hypercapnia. This correlation between LUS and morphologic and functional lung alterations was reflected on survival: LUS score on admission was significantly higher in non-survivors compared with survivors (and marginally significantly higher over time). On the other hand, we did not find a correlation between LUS score and respiratory system compliance. This is in contrast to chest CT scan, where the size of the so called “baby lung” in an ARDS patients correlates with compliance [29]. There are several potential limitations concerning the evaluation of lung mechanics with ultrasound that could explain our findings, e.g., the limited penetration of ultrasound waves into the lung parenchyma and the related difficulty to assess lung volume.

We found a correlation between LUS score and the plasma concentration of IL-6. IL-6 is a cytokine with a wide variety of context-dependent effects on immunity and inflammation that seems to be an important factor in the COVID-19 pathogenesis [30, 31]. Increased IL-6 plasma levels correlated with morbidity and mortality in COVID-19 patients [32, 33]. Furthermore, this cytokine might be directly involved in the pathogenesis of acute lung injury, presumably via recruitment of inflammatory cells in the lung [34–36]. It is a hallmark cytokine released during macrophage activation syndrome and cytokine storm, which are both potential key processes in COVID-19 disease exacerbation [37]. In contrast to other studies, we found no difference in IL-6 plasma concentration between survivors and non-survivors on admission. We observed, however, a correlation between LUS score and plasma IL-6 concentration during the time course of the disease, underlining the role of this cytokine in the pathogenesis and the severity of COVID-19 pneumonia. LUS score might therefore reflect the degree of inflammatory damage in the lung parenchyma. Yet, this issue remains object of further investigations.

**Limitations**

The retrospective design of the study and the limited number of patients are the main limitations of our study. LUS is an operator dependent technique, but during the emergency related to the pandemic, it was not possible to assess the inter-operator agreement. However, only experienced clinicians performed LUS using already tested international protocols.
Conclusion

In this series of mechanically ventilated COVID-19 patients, respiratory system compliance differed between survivors and non-survivors with the latter having a significant decrease in compliance after day 14 of symptom onset. Compliance did not correlate with the degree of abnormalities found in LUS, but LUS score correlated with survival, oxygenation, ventilation, and IL-6. LUS gives a dynamic bed-side view of the respiratory function and pathophysiology in patients with COVID-19-associated respiratory failure.

Authors’ Contribution MR, IR, MF, and SR had full access to all the data in the study and take responsibility for the integrity and the accuracy of the data analysis. MR, IR, GS, and SR conceived the study. MR, IR, EF, AZ, and SR collected the data for the study. AZ, EF, PZ, MG, and MB participated in analysis and interpretation of data, helped to draft the manuscript, and critically revised the manuscript for important intellectual content. MF was in charge of the statistical analysis. All authors read and approved the final manuscript.

Compliance with Ethical Standards

The study was approved by the Ethics Committee of South Tyrol (protocol number 46–2020). Study design and reporting were performed according to the Strengthening the Reporting of Observational Studies in Epidemiology (STROBE) statement [16].

Conflict of Interest

The authors declare that they have no conflict of interest.

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