Solitary fibrous tumor (SFT) is a spindle cell tumor which is mainly composed of fibroblastic component. The first description of SFT was in 1931 in pleura. At that time, it used to be called “hemangiopericytoma;” however, gradually it was revealed that SFT can be seen everywhere in the human body. Histopathologically, many of the fibroblastic tumors can mimic SFT, and immunohistochemistry is necessary for differential diagnosis. Several markers have been used, and recently, STAT6 has been introduced as a diagnostic marker. No research study has been reported from Iran.

OBJECTIVES: In this report, we are trying to introduce our experience about the clinicopathologic findings of 35 cases of SFT during the last 10 years from our center with an emphasis on the diagnostic role of STAT6 in this tumor.

RESULTS AND DISCUSSION: Our experience showed that SFT is more common in middle-aged men with a wide age range, although it is very rare in children. It can be present in every location with a wide size range from less than 1 cm to more than 15 cm. The presence of abnormal staghorn vessels is one of the main diagnostic histopathologic characteristics. STAT6 showed a sensitivity and specificity of 91% and 86% in the diagnosis of SFT. Other markers such as CD99, CD34, and BCL2 have also been considered useful in the differential diagnosis of this tumor, although the diagnostic accuracy seems to be lower than STAT6. High mitosis, presence of necrosis, and atypia are important criteria for predicting malignant and aggressive behavior in SFT. Among 35 cases in our study, 17% showed malignant behavior as recurrent or metastatic tumors.

CONCLUSION: Solitary fibrous tumor is a common soft tissue tumor that can be seen everywhere and needs careful histopathologic and immunohistochemical evaluation for the correct diagnosis and prediction of aggressive or benign behavior.

KEYWORDS: Solitary fibrous tumor, Immunohistochemistry, STAT6

Introduction

Solitary fibrous tumor (SFT) is a spindle cell tumor which is mainly composed of fibroblastic component. The first description of SFT was in 1931 in pleura. At that time, it used to be called “hemangiopericytoma;” however, gradually it was revealed that SFT can be seen everywhere in the human body. Histopathologically, SFT is one of the main diagnostic histopathologic characteristics. STAT6 showed a sensitivity and specificity of 91% and 86% in the diagnosis of SFT.

This tumor has been reported mainly to occur in middle-aged adults (20-70 years of age), and it is equal in women and men. It usually presents as slow growing painless mass with large size (5-10 cm) and well-defined border. Magnetic resonance imaging usually shows heterogeneous signal intensity. Grossly, SFTs are well-defined, firm, and homogeneous with or without capsule. Cut section shows homogeneously white tissue. Histologic sections show moderate cellularity with bland spindle to oval-shaped cells with no specific pattern. No significant atypia is seen. The intervening stroma is mostly collagenous, however, lipomatous (fatforming) differentiation may also occur. Many large branching and staghorn thin-walled hemangiopericytic vessels are also present. This characteristic blood vessels with pericytes were what originally was defined by Stout in 1942. In addition, some tumors show a predominant fat-containing component. The number of mitotic figures is variable. Malignant transformation is defined as infrequent event in which a high mitotic count is noted, (>4/10 high power field [HPF]) with atypia and necrosis. Rare histologic variants were also introduced such as mast cell rich, myxoid, and giant cell rich variant.
Most SFTs behave indolently but sometimes they are aggressive and unpredictable, with bland cytomorphology. Contrary to sufficient local control, estimated relapse rates for malignant SFTs are about 30%.9,10

Histologic findings besides the immunohistochemical staining help to differentiate SFT and other benign spindle cell tumors such as schwannoma, fibromatosis, neurofibroma, and so on.11

Markers such as CD99, CD34, and BCL2 are positive in SFTs and can be helpful, but recently a gene fusion NAB2-STAT6 is proposed as the molecular hallmark of SFT, encoding a chimeric protein that combines the early growth response (EGR)-binding domain of NAB2, a repressor of primary growth response (EGR) transcription factors that regulate dissociation and proliferation, with the transactivation domain of STAT6, a transcription factor that intercedes cytokine signaling.12,13 Molecular diagnostics of the fusion gene and immunohistochemical expression of nuclear STAT6 may be beneficial in diagnosing SFT, particularly in cases that are not clearly classified.14 Subsequent studies have shown that STAT6 c-terminus nuclear immunoreactivity is a viable surrogate marker for the fusion gene and is both a sensitive and particular marker in the histopathologic diagnosis of SFTs.14

In this report, we tried to investigate clinicopathologic characteristics of SFT in our center and evaluating the value of

Figure 1. A solitary fibrous tumor of lung: (A) low power H&E view, (B) high power H&E view, (C) CD34, and (D) STAT6.
STAT6 as a marker for the diagnosis of SFT and differential diagnosis of SFT from its mimics.

Patients and Methods
In this cross-sectional study, we evaluated the positivity of STAT6 in 35 cases of SFT during 10 years (2009–2019) in the affiliated hospitals of Shiraz University of Medical Sciences. Another 35 cases with spindle cell morphology and diagnosis other than SFT were considered as negative controls (7 cases in each category: fibromatosis, schwannoma, neurofibroma, malignant peripheral nerve sheath tumor, dermatofibrosarcoma protuberance)

In this study, we extracted all 70 cases reported as SFT and other spindle cell tumors from the pathology archive of the affiliated hospitals of Shiraz University of Medical Sciences during 10 years. All the hematoxylin–eosin stains were extracted and reviewed by 2 pathologists (B.G. and F.S.) to confirm the diagnosis. The atypia was classified as mild, moderate, and severe according to hypercellularity, nuclear pleomorphism, and hyperchromasia. The severity of atypia was defined whether each item being mild, moderate, and severe (presence of highly atypical bizarre cells was defined as severe).

There were 35 cases of SFT in the study (23 males and 12 females) with a mean age of 50.77 ± 19.65 [range: 1-99]. Other spindle cell tumors composed of 27 males and 8 female patients with the age range of 61.06 ± 16.97.

Clinical findings extracted from the clinical chart of the SFT patients including demography, treatment modalities, and outcome. The atypia was classified as mild, moderate, and severe according to hypercellularity, nuclear pleomorphism, and hyperchromasia. The severity of atypia was defined whether each item being mild, moderate, and severe (presence of highly atypical bizarre cells was defined as severe).

For all of the 35 patients, other immunohistochemical stainings were also done composed of Ki67, CD99, CD34, and BCI2. Then the best paraffin block was isolated and STAT6 performed by immunohistochemistry (IHC) for all of the 70 cases and controls.

We performed IHC on the corresponding formalin-fixed paraffin-embedded tissue block. The largest and well-fixed block of each patient was selected. Tissue slides (4 mm thick) were stained according to routine IHC procedure.

Section of tonsil served as STAT6-positive control in each immunohistochemical run. The IHC slides were evaluated by the pathologist. STAT6 was regarded as positive if nuclear staining is present.

Results
Thirty-five cases of SFT included into the statistical analysis. Twenty-two of SFTs were extrathoracic and 13 of them were intrathoracic.

Furthermore, tumor size was 8.16 ± 5.18 [range: 1-18] cm. Mild, moderate, and severe atypia were observed in 62.86% (22/35), 14.29% (5/35), and 5.71% (2/35), respectively. Also, 17.14% (6/35) of the patients did not show atypia. Necrosis was seen in 14.29% (5/35) of cases. Malignant SFT
was diagnosed if >4 mitoses/10 HPF were present, which was present in 28.57% (10/35) of our SFT cases (Figures 1–3). The mean number of mitosis in 10 HPF was 3.47 ± 4.48 (ranged: 1-25). It should be emphasized that mitosis should be considered as a sign of malignancy accompanied with infiltrative margins, pleomorphism, hypercellularity, and necrosis.14

Also, all of cases, except one case, had underwent surgical resection. Four patients reported a recurrence of SFT; and 2 patients had metastasis and expired. Details of clinicopathologic findings of all 35 SFT patients are summerized in Table 1.

Table 2 shows the value of different IHC markers in SFT, and Table 3 shows the comparison of the value of the diagnosis of SFT versus its mimickers by STAT6 and other markers. As Table 2 shows, more than 80% of the cases of SFT were positive for STAT−6, so this marker has the highest percentage of positivity in the SFT cases. The lowest percentage has been in CD99 which was positive in 60% of the cases. CD34 was the least specific marker which was positive in more than 70% of the non−SFT cases, which is shown in Table 3. Figure 4 shows the summery of diagnostic value of each marker in the diagnosis of SFT. As the figure shows, the best diagnostic value was in STAT−6. Only 3 cases with the diagnoses other than SFT were positive for STAT−6 (all 3 cases were fibromatosis). It is worthy to note that all of our cases showed intense nuclear staining.

Figure 3. Comparing different IHC markers in diagnosing 35 definite SFT patients.

Table 1. Clinicopathologic characteristics of the cases of SFT.

| SEX/AGE | LOCATION     | SIZE | MITOSIS/10 HPF | NECROSIS | ATYPIA | SURGERY | OUTCOME  |
|---------|--------------|------|----------------|----------|--------|---------|----------|
| 1 M/68  | Thigh        | 4    | 1-2            | –        | Mild   | +       | No recurrence |
| 2 M/47  | Pelvis       | 7    | 2              | –        | Mild   | +       | No recurrence |
| 3 F/90  | Lung         | 5    | 2              | –        | Mild   | +       | No recurrence |
| 4 M/50  | Lymph node   | Bx   | 6              | –        | Mild   | +       | Recurrence  |
| 5 M/51  | Retroperitoneum | 15  | 7              | –        | Moderate | + | Metastasis/expired |
| 6 M/59  | Mediastinum  | 2    | 9              | 20%      | Moderate | + | Metastasis/expired |
| 7 M/59  | Pelvis       | 15   | 5              | 10%      | Mild   | +       | No recurrence |
| 8 M/83  | Lung         | 15   | 1              | –        | Mild   | +       | No recurrence |
| 9 M/40  | Axilla       | 2    | 1              | –        | Mild   | +       | No recurrence |
| 10 F/75 | Lung         | 10   | 1-2            | –        | Mild   | +       | No recurrence |
| 11 M/59 | Orbit        | 1.5  | 4              | –        | Mild   | +       | Recurrence  |
| 12 F/53 | Kidney       | Bx   | 2              | –        | Mild   | +       | No recurrence |
| 13 M/56 | Brain        | 3    | 8              | –        | Moderate | + | No recurrence |
| 14 F/45 | Omentum      | 10   | 1              | –        | Mild   | +       | No recurrence |
| 15 F/53 | Retroperitoneum | 8   | 1              | –        | Mild   | +       | No recurrence |

(Continued)
Table 1. (Continued)

| SEX/AGE | LOCATION | SIZE | MITOSIS/10 HPF | NECROSIS | ATYPIA     | SURGERY | OUTCOME  |
|---------|----------|------|---------------|----------|------------|---------|----------|
| 16 F/42 | Kidney   | 11   | 1             | –        | Mild       | +       | No recurrence |
| 17 M/27 | Parotid  | 4    | 2             | –        | Mild       | +       | No recurrence |
| 18 F/1  | Tongue   | 1    | 5             | –        | Mild       | +       | No recurrence |
| 19 F/40 | Lung     | 13   | 6-7           | 10%      | Moderate   | +       | Recurrence |
| 20 M/64 | Axilla   | Bx   | 7-8           | –        | Severe     | +       | Recurrence |
| 21 M/1  | Buttock  | Bx   | 2             | –        | –          | +       | No recurrence |
| 22 F/45 | Bladder  | 9    | 1             | –        | –          | +       | No recurrence |
| 23 M/33 | Pelvis   | 11   | 1             | –        | Mild       | +       | No recurrence |
| 24 M/67 | Lung     | 15   | 25       | 25%      | Severe     | +       | No recurrence |
| 25 M/24 | Lung     | 5    | 1             | –        | Mild       | +       | No recurrence |
| 26 F/37 | Eyelid  | 2    | 2             | –        | Mild       | +       | No recurrence |
| 27 M/64 | Pleura   | 18   | 1             | –        | Mild       | +       | No recurrence |
| 28 M/45 | Pleura   | Bx   | 1             | –        | –          | +       | No recurrence |
| 29 F/40 | Vagina   | 2    | 1             | –        | Mild       | +       | No recurrence |
| 30 M/67 | Lung     | 15   | 2             | –        | –          | +       | No recurrence |
| 31 M/70 | Breast   | Bx   | 1             | –        | –          | –       | No recurrence |
| 32 F/38 | Face     | 3    | 1-2           | –        | Mild       | +       | No recurrence |
| 33 M/45 | Inguinal | 9    | 6             | –        | Mild       | +       | No recurrence |
| 34 M/70 | Mediastinum | 12   | 1           | 10%      | Mild       | +       | No recurrence |
| 35 M/69 | Lung     | 9    | 1             | –        | –          | +       | No recurrence |

Abbreviations: Bx, biopsy; M, Male; F, Female.

Table 2. Comparison of IHC markers with other clinicopathologic findings.

| VARIABLE | TOTAL (N = 35) | INTRA THORACIC (N = 13) | EXTRA THORACIC (N = 22) | P-VALUE |
|----------|----------------|--------------------------|--------------------------|---------|
| Gender   | 23 (65.7%)     | 10 (76.9%)               | 13 (59.1%)               | .463<sup>a</sup> |
|          | 12 (34.3%)     | 3 (23.1%)                | 9 (40.9%)                |         |
| Age (M ± SD [R])<sup>b</sup> | 50.77 ± 19.65 [1-99] | 61.77 ± 44.27            | 44.27 ± 17.86            | .009<sup>c</sup> |
| Atypia   |                |                          |                          |         |
| Negative | 6 (17.1%)      | 3 (23.1%)                | 3 (13.6%)                |         |
| Mild     | 22 (62.9%)     | 7 (53.8%)                | 15 (68.2%)               | .837<sup>a</sup> |
| Moderate | 5 (14.3%)      | 2 (15.4%)                | 3 (13.6%)                |         |
| Severe   | 2 (5.7%)       | 1 (7.7%)                 | 1 (4.5%)                 |         |

(Continued)
Table 2. (Continued)

| VARIABLE | TOTAL (N = 35) | INTRA THORACIC (N = 13) | EXTRA THORACIC (N = 22) | P-VALUE |
|----------|----------------|--------------------------|--------------------------|---------|
| Necrosis (+) | 5 (14.3%) | 4 (30.8%) | 1 (4.5%) | .052a |
| Mitosis (N/10 HPFs) | 3.47 ± 4.48 [1-25] | 4.46 ± 6.72 | 2.89 ± 2.39 | .322a |
| Tumor size (cm) | 8.16 ± 5.18 [1-18] | 10.82 ± 5.09 | 6.53 ± 4.64 | .028a |
| CD34 (+) | 25 (71.4%) | 11 (84.6%) | 14 (63.6%) | .259a |
| BCL2 (+) | 27 (77.1%) | 10 (76.9%) | 17 (77.3%) | 1a |
| CD99 (+) | 21 (60%) | 5 (38.5%) | 16 (72.7%) | .075a |
| Ki67 (+) | 18 (51.4%) | 8 (61.5%) | 10 (45.5%) | .489a |
| STAT6 (+) | 31 (88.6%) | 12 (92.3%) | 19 (86.4%) | 1a |
| Surgery (+) | 34 (97.1%) | 13 (100%) | 21 (95.5%) | 1a |
| Outcome | | | | .648a |
| NR | 29 (82.6%) | 10 (76.9%) | 19 (86.4%) | |
| R | 4 (11.4%) | 3 (23.1%) | 1 (4.5%) | |
| M + E | 2 (5.7%) | | | |

Abbreviations: NR, no recurrence; R, recurrence; M + E, metastasis and expired; HPFs, high power fields.
aChi-square test or Fisher's exact test.
bM ± SD [R]: mean ± standard deviation [range].
Wilcoxon rank-sum test (Kolmogorov-Smirnov test: 0.047).
Independent t-test (Kolmogorov-Smirnov test: 0.09).
Independent t-test (Kolmogorov-Smirnov test: 0.152).
Staining of more than 5% was considered positive.
Positive outcomes: recurrence or metastasis and expired.

Table 3. Comparison of STAT6 positivity in SFT and non-SFT spindle cell mimickers.

| VARIABLE | STAT6 + | CD99 + | CD34 + | BCL2 + |
|----------|---------|--------|--------|--------|
| SFT | 31 (88.6%) | 21 (60%) | 25 (71.4%) | 27 (77.1%) |
| Non-SFT | 3 (8.6%) | 2 (5.7%) | 25 (71.4%) | 5 (14.3%) |
| P value | <.05 | <.05 | >.05 | <.05 |

Figure 4. High-power and Low-power view of malignant solitary fibrous tumor.
Discussion
Clinicopathologic findings of our study showed that SFT is more common in middle-aged men and can be seen in every location in human body. This tumor is very rare in children, and in our cross-sectional study, only 2 cases have been identified in pediatric population. The size range is variable from less than 1 cm to more than 15 cm. Histopathologically, SFT is a fibroblastic tumor with characteristic hemangiopericytic vessels which needs to be differentiated from other mimickers such as neural tumors and other fibrohistiocytic tumors. In this study, we tried to study the frequency of STAT6 as an IHC marker versus other markers in the diagnosis of SFT. The main findings of our study showed that nuclear staining of STAT6 marker had the sensitivity and specificity of 88.6% and 91%, respectively. Diagnostic rate of other markers in the diagnosis of SFT were lower than STAT6 (BCL2: 77.1%; CD34: 71.4%; CD99: 60%). This finding is compatible with previous reports.15 NAB2-STAT6 has been proved to have pivotal role in SFT tumorigenesis,16 and as a IHC marker, has an excellent diagnostic performance for the diagnosis of SFTs, as well as differentiating it from other mimicking pathologies.17-19 The reported sensitivity of STAT6 in the diagnosis of SFT has been from 87% to 100% in the previous reports.20-22

In our study, 17% of the cases with the final diagnosis of SFT showed recurrence and metastasis, composed of 4 recurrent tumors and 2 metastatic cases, which is higher than previous studies (10%). mitosis >4/10 HPFs, 2 showed necrosis, and 4 cases showed moderate-to-severe atypia. Also, in univariate analysis we showed that higher mitosis rate in 10 HPFs (P value < .0001) and more severe atypia (P value: .017) were correlated with worse outcome, that is, recurrence or metastasis. Other previous reports showed that up to one third of SFTs may show malignant features, atypia, and necrosis, which might be associated with malignant tumor behaviors.23-26 In the previous reports, 15% to 28.57% of cases showed malignant features.23 Malignant SFTs should be differentiated from other sarcomas such as malignant peripheral nerve sheath tumors, synovial sarcoma, fibrosarcoma, and so on.27

Conclusion
Solitary fibrous tumor is a tumor with wide-spread location preferences which can accurately be diagnosed by histology and IHC studies, especially the newly introduced STAT6. The overall prognosis is good, but atypia and mistosis are accurate predictor of malignant behavior.

Author Contributions
Bita Geramizadeh: Idea of research, looking at slides, analysis of data, and writing the paper. Fatemeh Safavi: Looking at slides and analysis of data and help to write the paper.

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REFERENCES
1. Doyle LA, Vivero M, Fletcher CD, Mertens F, Hornick JL. Nuclear expression of STAT6 distinguishes solitary fibrous tumor from histologic mimics. Mod Pathol. 2014;27:390-395.
2. Wignall OJ, Moskovic EC, ‘Thway K, Thomas JM. Solitary fibrous tumors of the soft tissues: review of the imaging and clinical features with histopathologic correlation. AJR Am J Roentgenol. 1995;165:W55-W62.
3. Fukunaga M, Nagasuna H, Nikaido T, Harada T, Ushigome S. Extraperitoneal solitary fibrous tumor: a report of seven cases. Mod Pathol. 1997;10:443-450.
4. Stout AP, Murray MR. Hemangiopericytoma: a vascular tumor featuring Zim- merman’s pericytes. Ann Surg. 1942;116:26-33.
5. ‘Thway K, Ng W, Noujaim J, Jonas RL, Fisher C. The current status of solitary fibrous tumor: diagnostic features, variants, and genetics. Int J Surg Pathol. 2016;24:281-292.
6. Lee J-C, Fletcher CD. Malignant fat-forming solitary fibrous tumor (so-called “lipomatous hemangiopericytoma”): clinicopathologic analysis of 14 cases. Am J Surg Pathol. 2011;35:1177-1183.
7. Dong SS, Wang N, Yang CP, et al. Giant cell rich solitary fibrous tumor in the asopher- ynx: a case report and review of literature. Orca Targen Ber. 2020;13:6819-6826.
8. Ronchi A, Cozolino I, Marino FZ, et al. Extra pleural solitary fibrous tumor: a distinct entity from pleural solitary fibrous tumor. An update on clinical, molecu- lar, and diagnostic features. Diagn Histopathol. 2020;26:144-150.
9. Bylicki O, Rouvière D, Cassier P, et al. Assessing the multimodal management of advanced solitary fibrous tumors of the pleura in a routine practice setting. J Thorac Oncol. 2015;10:309-315.
10. Lococo F, Cesario A, Candillo G, et al. Malignant solitary fibrous tumors of the pleura: retrospective review of a multicenter series. J Thorac Oncol. 2013;8:1708-1716.
11. Chen BJ, Maritio-Enriquez A, Fletcher CD, Hornick JL. Loss of retinoblastoma protein expression in spindle cell/pleomorphic lipomas and cytogenetically related tumors: an immunohistochemical study with diagnostic implications. Am J Surg Pathol. 2012;36:1119-1128.
12. Geramizadeh B, Marras M. Role of immunohistochemistry in the diagnosis of solitary fibrous tumor, a review. Iran J Pathol. 2016;11:195-203.
13. Chmielecki J, Crago AM, Rosenberg M, et al. Whole-exome sequencing identifies a recurrent NAB2-STAT6 fusion in solitary fibrous tumors. Nat Genet. 2015;47:131-132.
14. Schweizer L, Koellech C, Sahm F, et al. Meningeal hemangiopericytoma and solitary fibrous tumors carry the NAB2-STAT6 fusion and can be diagnosed by nuclear expression of STAT6 protein. Acta Neuropathol. 2013;125:651-658.
15. Fletcher CD, Bridge JA, Lee JC. WHO Classification of Tumours of Soft Tissue and Bone. 4th ed. Lyon: IARC Press; 2013.
16. Robinson DR, Wu Y-M, Kalyana-Sundaram S, et al. Identification of recurrent NAB2-STAT6 gene fusions in solitary fibrous tumor by integrative sequencing. Nat Genet. 2013;45:180-185.
17. Demicco EG, Park MS, Araujo DM, et al. Solitary fibrous tumor: a clinicopatho- logical study of 110 cases and proposed risk assessment model. Mod Pathol. 2012;25:1298-1306.
18. Yoshida A, Tsuta K, Ohno M, et al. STAT6 immunohistochemistry is helpful in the diagnosis of solitary fibrous tumors. Am J Surg Pathol. 2014;38:552-559.
19. Vogels RJ, Vletter M, Verdeijken-Jonkers YM, et al. Solitary fibrous tumors— clinicopathologic, immunohistochemical and molecular analysis of 28 cases. Diagn Pathol. 2014;9:242.
20. Tai H-C, Chuang I-C, Chen T-C, et al. NAB2–STAT6 fusion types account for clinicopathological variations in solitary fibrous tumors. Mod Pathol. 2015;28:1324-1335.
21. Oualian S, Trautmann M, Orosji E, et al. Differential diagnosis of solitary fibrous tumours: a study of 454 soft tissue tumors indicating the diagnostic value of nuclear STAT6 relocation and ALDH1 expression combined with in situ proximity ligation assay. Int J Oncol. 2015;46:2599-2605.
22. Guner G, Bishop JA, Bezerra SM, et al. The utility of STAT6 and ALDH1 expression in the differential diagnosis of solitary fibrous tumor versus prostate-specific stromal neoplasms. Hum Pathol. 2016;54:184-188.
23. Gold JS, Antonescu CR, Hajdu C, Ferrone CR, Hussain M, et al. Clinicopatho- logic correlates of solitary fibrous tumors. Cancer. 2002;94:1057-1068.
24. Vallat-Decouvelaere AV, Dry SM, Fletcher CD. Atypical and malignant solitary fibrous tumors in extrathoracic locations: evidence of their comparability to intra-thoracic tumors. Am J Surg Pathol. 1998;22:1501-1511.
25. Schirosi L, Lantuejoul S, Cavazza A, et al. Pleureo-pulmonary solitary fibrous tumors: a clinicopathologic, immunohistochemical and molecular study of 88 cases confirming the prognostic value of de Perrot staging system and p53 expression, and evaluating the role of c-kit, BRAF, PDGFRA (alpha/beta), c-met, and EGFR. Am J Surg Pathol. 2008;32:1627-1642.
26. England DM, Hochholzer L, McCarthy MJ. Localized benign and malignant fibrous tumors of the pleura. A clinicopathologic review of 223 cases. Am J Surg Pathol. 1989;13:640-658.
27. Saeed O, Zhang S, Cheng L, Lin J, Alruewai F, et al. STAT6 expression in solitary fibrous tumor and histologic mimics: a single institution experience. Appl Immunohistochim Med Mol Pathol. 2020;28:311-315.