Structure and Function of Rat Liver Polysome Populations. II. Characterization of Polyadenylate-containing mRNA Associated with Subpopulations of Membrane-bound Particles

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ABSTRACT Poly(A)+RNA fractions prepared from free and loosely and tightly membrane-bound polysome populations (poly(A)+RNA_free, poly(A)+RNA_loose, and poly(A)+RNA_tight) were used to drive cDNA in homologous and heterologous hybridization reactions. A large fraction by mass of sequences was shared among the three poly(A)+RNA populations, but shared sequences exhibited distinct frequency distributions within the different populations. 13–15 in vitro translation products of poly(A)+RNA_free and poly(A)+RNA_loose detected by gel electrophoresis were shared. Most of these were produced in different relative quantities by the two RNA populations. Five or six higher mol wt polypeptides were produced by poly(A)+RNA_loose that were not detected as products of either poly(A)+RNA_free or poly(A)+RNA_tight. We suggest that loosely bound polysomes may not be artifactually derived as reflected in their quantitatively distinct poly(A)+RNA population.

Two tightly membrane-bound RNP fractions were prepared from rat liver on the basis of their release from or retention on purified rough microsomes or a crude membrane fraction after in vitro disaggregation of polysomes with high-salt and puromycin. Homologous and heterologous hybridizations involving their poly(A)+RNA fractions revealed that a large portion by mass of sequences was shared but that these sequences exhibited distinct frequency distributions in the two fractions. The RNA fractions produced an identical set of in vitro translation products but individual polypeptides were produced in different relative quantities. This indicates that the two RNP fractions do not arise by any random artifactual process and suggests that they may represent functionally distinct populations.

Numerous investigations have revealed the existence of membrane-bound ribosomes or polyribosomes that apparently exhibit distinct interactions with rough microsomal or rough endoplasmic reticulum (RER) fractions (See references in 24, 42). Rosbash and Penman (38, 39) coined the terms “loose” and “tight” to describe ribosomes that are either susceptible or resistant to release from a HeLa cell membrane fraction by treatment with EDTA, puromycin, or ribonuclease. They interpreted their findings to suggest that two distinct classes of membrane-bound polyribosomes exist in HeLa cells. Subsequent work by Mechler and Vassalli (25, 26) with myeloma cells did not support this interpretation. Their findings suggested that “loose” and “tight” ribosomes are derived from different ends of the same polyribosome structure, the former being only indirectly attached to the membrane by means of the mRNA molecule due to a shortage of 60s subunit binding sites on RER of cultured cells. Such “dangling” ribosomes are present in relatively small quantities in rough membrane fractions from liver (8). Instead, a significant fraction of liver membrane-bound polyribosomes are released by incubation of membrane fractions in buffers containing high monovalent cation concentrations (37). “Loosely bound” polyribosomes of this type have been identified in other systems (17, 45). In many cases these high-salt-releasable polyribosomes have been shown to exhibit similar characteristics (37) or synthesize the same polypeptides as free polyribosomes (15, 29, 45). This has led to the conclusion by some investigators (37, 45) that they represent artifactually adsorbed free polysomes. However,
much of the available data concerning this question is difficult to interpret since either (a) the integrity of mRNA was not shown to be preserved during polyribosome isolation or (b) the extent of free polysome contamination of rough membrane fractions was not determined. The latter is particularly significant since contaminating free polysomes will be included in the high-salt releasable fraction. In addition, to these two polyribosome classes, two classes of membrane-bound mRNA or mRNP ribonucleoprotein have been defined in numerous systems on the basis of membrane release or retention after various treatments that cause polyribosome breakdown and release of most of the membrane-bound ribosomes (2, 3, 10, 14, 18, 20-23, 25-27, 31). In rat liver divergent results have been obtained. The membrane-associated fraction has been found by different investigators to account for 20-90% of the total, and there is controversy over whether the results indicate the existence of a direct interaction between mRNA and the membrane (10, 14, 18). In this case the available data are extremely limited since the two mRNA fractions have not been isolated in degraded form and characterized.

We have addressed the question concerning the existence of functionally distinct subpopulations of membrane-bound polyribosomes and mRNP by using hybridization and in vitro translation technologies to characterize and compare their poly(A)* mRNA populations. Our data indicate that, although the populations exhibit considerable sequence overlap, distinct quantitative differences exist among them. This suggests that the populations may be functionally distinct.

**Materials and Methods**

Isolation of free and total membrane-bound polysomes, synthesis and kinetic fractionation of cDNA, and hybridization reactions were conducted exactly as described previously (28).

**Tissue Fractionation and Isolation of Loosely and Tightly Membrane-Bound Polysome Fractions**

Adult male rats of the Holtzman strain were used. Rats were given chow and water ad libitum and were not starved before sacrifice. Rats were maintained on a 12 h light/dark cycle and were kept in the dark between 8 pm and 8 am. Sacrifice occurred between 2 and 4 pm. Loosely and tightly membrane-bound polysomes were prepared by the procedure of Ramsey and Steele (36, 37) with some modifications to preserve integrity of mRNA, increase recovery of polysomes from the livers of fed rats, and decrease the levels of cross-contamination.

**Isolation of Loosely Membrane-Bound Polysomes:** Rats were decapitated and the livers perfused via the inferior vena cava with ice-cold 0.25 M sucrose containing 5 mM MgCl$_2$ and 100 μg/ml sodium heparin. Perfused livers were excised and homogenized in 3 vol of a solution containing 0.25 M sucrose, 50 mM HEPES, pH 7.6, 5 mM MgCl$_2$, 75 mM KCl, 1.3 mM GSH (reduced glutathione), and 250 μg/ml heparin. The homogenate was centrifuged in a Beckman SW 27 rotor (Beckman Instruments, Inc., Fullerton, CA) for 2 min at 6,900 rpm and 12 min at 131,000 g$_{av}$. The supernate was discarded and the pellet homogenized with three to four strokes in a loose-fitting Potter-Elvejem homogenizer in 3 vol/g liver of the initial homogenization buffer containing 17% rat liver high-speed supernatant fluid (36). The homogenate was centrifuged as described above and the supernate, containing residual free polysomes, discarded.

Loosely membrane-bound polysomes were extracted from the pellet by homogenizing with three to four strokes as above in 2 vol of the initial homogenization buffer containing 17% rat liver high-speed supernatant fluid and 0.25 M KCl rather than 0.075 M. The homogenate was incubated for 20 min on ice and centrifuged as described above. The supernate containing loosely membrane-bound polysomes was adjusted to 1 mg/ml heparin, layered over sucrose cushions prepared as described previously (28), and polysomes pelletized by centrifuging at 303,500 g$_{av}$ in a Beckman 60 Ti rotor (Beckman Instruments, Inc.) for 20 h.

**Isolation of Tightly Membrane-Bound Polysomes:** A 131,000 g$_{av}$ pellet was prepared as described above, except that the initial homogenization buffer contained 0.25 M KCl rather than 0.075 M and 500 μg/ml sodium heparin rather than 250 μg/ml. The pellet was washed free of residual contaminating free and loosely membrane-bound polysomes by homogenizing in 3 vol/g liver of the initial homogenization buffer containing 15% rat liver high-speed supernatant fluid, and recentrifuging. The supernate was discarded and the 131,000 g$_{av}$ pellet homogenized in 50% rat liver high-speed supernatant fluid containing 1% Triton X-100, 20 mM MgCl$_2$, 250 mM KCl, 3 mM GSH and centrifuged at 1,470 g$_{av}$ in a Sorvall SS34 rotor (Du Pont Instruments-Sorvall Biomedical Div., Du Pont Co., Newtown, CT) for 5 min to pellet nuclei. The supernate was adjusted to 50 mM MgCl$_2$, 2 mg/ml heparin, 1.3% sodium deoxycholate and centrifuged at 15,000 g$_{av}$ for 5 min to pellet insoluble material. Polysomes were pelletized from the supernate as described above for loosely membrane-bound polysomes.

**Isolation of High-Salt, Puromycin-Mediated Membrane-Released and Membrane-Associated RNP Fractions:** Polyribosome breakdown and release of most of the membrane-bound ribosomes (2, 3, 10, 14, 18, 20-23, 25-27, 31). In rat liver the results are due to the extensive manipulation and incubation periods involving the ribonuclease-rich 131,000 g$_{av}$ pellet. In method B, the RNP fractions were prepared from high-speed washed, purified, rough microsomes obtained from a postmitochondrial supernatant. Poly(A)$^+$RNA obtained by this procedure was intact (see Results), but was derived from a subcellular fraction containing only 5-10% of the total tightly membrane-bound polysomes.

**Method A:** A 131,000 g$_{av}$ pellet was prepared and washed once as described for tightly membrane-bound polysomes. The resulting pellet was homogenized gently in 3 vol/g liver of the initial homogenization buffer containing 2 mM puromycin diHCl, 15% rat liver high-speed supernatant fluid, 0.1 mM EDTA, and 1.5 mM, rather than 5 mM MgCl$_2$. The homogenate was incubated at 4°C for 1 h and recentrifuged as above. The supernate containing the membrane-released tightly bound RNP fraction was decanted and stored on ice. The pellet was gently homogenized in the same buffer, incubated at 4°C for 15 min, and recentrifuged. The supernate was decanted and added to the membrane-released tightly bound RNP fraction. The pellet was washed once more in the same buffer lacking puromycin and the supernate discarded. Nuclei were removed from the pellet as described above for tightly membrane-bound polysomes using 1% Triton X-100 treatment, except that the buffer contained 1 mg/ml yeast tRNA as an additional ribonuclease inhibitor. Membrane-associated tightly bound RNP was liberated from RER by sodium deoxycholate treatment of the 1,470 g$_{av}$ supernatant fluid as described above. Both tightly membrane-bound RNP fractions were pelleted through 1.8 M sucrose cushions prepared in the same buffer as for the isolation of polysomes.

**Method B:** Postmitochondrial rough microsomes were prepared essentially as described previously (10) except that all solutions contained 0.5 mg/ml sodium heparin and 3 mM GSH. The microsomes that banded at the 1.35 M-2.0 M sucrose interface were collected and diluted with 2 vol of a solution containing 10% rat liver high-speed supernatant fluid, 0.75 M KCl, 5 mM MgCl$_2$, 50 mM HEPES, pH 7.6, 500 μg/ml sodium heparin, and 3 mM GSH. 50 ml of the rough microsome solution were layered over 10 ml of 1 M sucrose in the same buffer in a 1 x 4½ inch polycarbonate bottles. Microsomes were washed free of contaminating free and loosely membrane-bound polysomes by centrifuging in a Beckman 45 Ti rotor (Beckman Instruments, Inc.) at 30,000 rpm for 35 min. The supernate was decanted and the walls of the bottles carefully swabbed. The pellet was gently homogenized in 20 ml of a solution containing 25% rat liver high-speed supernatant fluid, 2 mM puromycin diHCl, 0.66 M KCl, 2 mM MgCl$_2$, 500 μg/ml sodium heparin, 50 mM HEPES, pH 7.6, and 3 mM GSH. After incubation for 1 h at 4°C, this mixture was layered over 30 ml of 0.5 M sucrose prepared in the same buffer lacking puromycin and pelleted as above. The supernatant containing the membrane-released tightly bound RNP fraction was decanted into a fresh centrifuge bottle and spun in a Beckman 45 Ti rotor (Beckman Instruments, Inc.) at 40,000 rpm for 2½ h to pellet RNP. The high-salt, puromycin-stripped microsomal pellet containing the membrane-associated tightly bound RNP fraction was extracted directly in guanidinium thiocyanate as described below.

**Isolation of Poly(A)$^+$RNA Fractions**

Polyosomal pellets were extracted by the SDS-phenol-chloroform procedure of Palmiter (32) as described previously (28). RNP and microsomal pellets were extracted by the guanidinium thiocyanate-CsCl procedure of Chirgwin et al. (12). Poly(A)$^+$RNA fractions were isolated by oligo(dT) cellulose chromatography using the procedure of Bantle et al. (4) as described previously (28).
In Vitro Translation and Analysis of Products

Poly(A)*RNA was translated in a nuclease-treated rabbit reticulocyte lysate system as described by Gonzales and Kasper (15). Poly(A)*RNA was translated at a concentration of 5–15 μg/ml. [3H]methionine (600–1400 Ci/mmol) was included at a concentration of 50 μCi/ml. Under these conditions incorporation of radioactivity into TCA-insoluble products varied from 10–50-fold over background with the poly(A)*RNA fractions. Translation was allowed to proceed for 45 min at 30°C and the reaction quenched by boiling on ice. 2 μl aliquots of the reaction mixture were spotted onto Whatman No.1 filter paper (Whatman Inc., Paper Div., Clifton, NJ). Filters were washed by boiling for 10 min in 10% TCA and rinsed twice each in 5% TCA, methanol, and ether. Filters were dried and acid-precipitable radioactivity determined by scintillation spectrophotometry in 10 ml of OCS scintillator cocktail (Amersham Corp., Arlington Heights, IL). The translation products were analyzed by loading equal quantities of acid-precipitable counts onto 10% polyacrylamide, 1.5-mm slab gels using the system of Laemmli (19). The gels were fixed and treated with Enhance (New England Nuclear, Boston, MA) as described by the manufacturer. Dried gels were fluorographed using Kodak XAR-5 film and fluorographs were subjected to microdensitometry.

Sizing of Poly(A)*RNA and cDNA Populations

Poly(A)*RNA populations were sized by hybridization of excess [3H]poly(U) to formamide-sucrose gradient fractions as described previously (28). Number-average sizes were estimated by using the formula: X cpm/μ, where cpm is proportional to the number of molecules of length l.

RESULTS

Isolation and Characterization of Loosely and Tightly Membrane-Bound Polysomal Poly(A)*RNA Populations

Loosely and tightly membrane-bound polysomes were isolated and their respective poly(A)*RNA fractions obtained as described in Materials and Methods. Polysomes prepared by pelleting through sucrose cushions, as described, have previously been shown to be free of contaminating nuclear or cytoplasmic RNP particles. Cross-contamination levels were estimated to be <1% by the procedure used previously (28). Yields were 0.61 mg/g liver of loosely membrane-bound and 2.19 mg/g liver of tightly membrane-bound polysomal RNA (average of two preparations). Loosely membrane-bound polysomes thus comprise ~22% of the total membrane-bound polysome population. Polysome profiles have demonstrated the presence of large polyribosome structures with monomers comprising a relatively small proportion of each fraction (37). Yields of poly(A)*RNA were somewhat variable and ranged from 0.5–1.0% of the total polysomal RNA populations. Yield variability was due to aggregation rather than degradation of RNA during preparation. This aggregation is the result of the extremely high g-force necessary for maximum recovery of polysomes from subcellular fractions prepared from the livers of fed rats. All poly(A)*RNA preparations were sized on formamide sucrose gradients, as described in Materials and Methods. Number-average lengths of poly(A)*RNAloose and poly(A)*RNAtight were ~1,725 and 1,400 nucleotides, respectively.1

1 Abbreviations used in this paper: cDNAfree, cDNAbound, cDNAloose, cDNAtight, cDNAmean, complementary DNA fractions transcribed from poly(A)*RNA derived from membrane-associated RNP, total membrane-bound polysomes, free polysomes, loosely membrane-bound polysomes, tightly membrane-bound polysomes, and membrane-released RNP, respectively; FOA, 5-fluoro-orotic acid; GSH, reduced glutathione; Kmean, rate constant for the most abundant component of the heterologous hybridization involving poly(A)*RNAloose and cDNAloose; Ktight, rate constant for the most abundant component of the heterologous hybridization involving poly(A)*RNAtight and cDNAtight; Ploose, fraction of hybridizable cDNAloose comprising the most abundant component of the heterologous hybridization involving poly(A)*RNAloose and cDNAloose; Ptight, fraction of hybridizable cDNAtight comprising the most abundant component of the heterologous hybridization involving poly(A)*RNAloose and cDNAloose; poly(A)*RNAloose, poly(A)*RNAmean, poly(A)*RNAfree, poly(A)*RNAmean, poly(A)*RNAfree, poly(A)*RNAmean, poly(A)*RNAfree, poly(A)*RNAmean, poly(A)*RNAfree.
Comparing the kinetics of the two analogous and heterologous reactions in Fig. 1A indicates that poly(A)\textsuperscript{+}RNA\textsubscript{too} contains all of the sequences that are present in poly(A)\textsuperscript{+}RNAtight. Poly(A)\textsuperscript{+}RNA\textsubscript{too} was also hybridized to kinetically fractionated cDNA\textsubscript{too} and cDNA\textsubscript{tight} to poly(A)\textsuperscript{+}RNAtight. This figure indicates that poly(A)\textsuperscript{+}RNA\textsubscript{too} is comprised of rare RNA species. However, it should be realized that the difference of 9% observed here may not be significant, and is not strong evidence that the two populations are qualitatively unique. The kinetics of the reactions suggests that poly(A)\textsuperscript{+}RNA\textsubscript{too} is relatively enriched in sequences that are rare in poly(A)\textsuperscript{+}RNAtight. The hybridization curves in Fig. 1b indicate that poly(A)\textsuperscript{+}RNA\textsubscript{too} may contain some sequences that are either lacking or present at very low levels in poly(A)\textsuperscript{+}RNAtight.

The hybridization reactions with kinetically-enriched cDNA\textsubscript{bound} was kinetically fractionated into abundant and less-abundant fractions as described (28). Abundant cDNA\textsubscript{bound} comprised 39% and less-abundant cDNA\textsubscript{bound} 61% of the total. Abundant cDNA\textsubscript{bound} (A) and less-abundant cDNA\textsubscript{bound} (B) were hybridized to an excess of poly(A)\textsuperscript{+}RNA\textsubscript{too} (O) and the extent of reaction determined as described (28). RNA concentrations ranged from ~4 to 400 µg/ml. Smooth curves were drawn without the aid of a computer.

### Table 1: Complexity and Frequency Distribution of Loosely Membrane-bound and Tightly Membrane-bound Polysomal Poly(A)\textsuperscript{+}RNA Populations

| Polysome class | Abundance class | Percent cDNA hybridized* | Rate Constant | Nucleotide complexity | No. of mRNA species| Copies/cell** |
|----------------|-----------------|--------------------------|---------------|-----------------------|------------------|--------------|
| Loose I        | 18.4            | 317                      | 2420          | 2.05 x 10\textsuperscript{3} | 1                | 39,000       |
| II             | 13.0            | 40.3                     | 136           | 1.38 x 10\textsuperscript{2} | 8                | 5800         |
| III            | 29.7            | 9.4                      | 5.53          | 5.36 x 10\textsuperscript{2} | 311              | 149          |
| IV             | 38.8            | 0.014                    | 0.866         | 5.71 x 10\textsuperscript{2} | 33,100           | 1            |
| Tight I        | 13.1            | 266                      | 1450          | 1.38 x 10\textsuperscript{2} | 1                | 205,000      |
| II             | 35.4            | 28.3                     | 218           | 2.45 x 10\textsuperscript{2} | 1                | 11,600       |
| III            | 35.1            | 1.64                     | 5.52          | 6.04 x 10\textsuperscript{2} | 431              | 470          |
| IV             | 16.3            | 0.020                    | 0.052         | 3.86 x 10\textsuperscript{2} | 27,600           | 7            |

* Normalized to a terminal hybridization value of 100%. Observed terminal values were 87.2 and 89.0% for loosely bound and tightly bound poly(A)\textsuperscript{+}RNA, respectively.

† Liter per mole-s. Values have not been adjusted to those that would be observed under standard salt conditions. Those values may be obtained by consulting the appropriate tables.

‡ Rate constant expected for an RNA abundance class reaction in isolation.

§ Rate constant expected for an RNA abundance class reaction in isolation.

¶ Number of unique mRNA species 1725 (loosely bound) or 1400 (tightly bound) nucleotides in length.
tions indicate that on the average poly(A)$+\text{RNA}_{\text{loose}}$ sequences are present in poly(A)$+\text{RNA}_{\text{tight}}$ at only slightly reduced concentrations.

Since only sequences that are abundant in both populations will react at low rot values in the heterologous hybridizations, the first components of these hybridizations can be analyzed to reveal the extent to which abundant sequences are shared (13). Computer-best fits (28) revealed that the first component of the heterologous reaction involving cDNA$\text{tight}$ exhibits a rate constant ($K_{\text{tight}}$) of $36.4\text{ liter/mole-s}$ and contains $27.6\%$ of the reacting cDNA$\text{tight}$ ($P_{\text{tight}}$). The values for the reciprocal heterologous reaction involving cDNA$\text{loose}$ were: $K_{\text{loose}} = 77.5$ and $P_{\text{loose}} = 15.7$. If these values represent only abundant shared sequences, then $K_{\text{loose}}/P_{\text{loose}}$ should be equal to $K_{\text{tight}}/P_{\text{tight}}$ since the complexity of the reacting component is the same regardless of which population it is a part. These values are 281 and 232, respectively, which are reasonably close. Using an intermediate value of $256\text{ l/mole-s}$, approximately four abundant species are shared. It can be estimated that these abundant shared sequences are present at $\approx 2000$ copies/cell in poly(A)$+\text{RNA}_{\text{loose}}$ and $9000$ copies/cell in poly(A)$+\text{RNA}_{\text{tight}}$. Thus, these sequences are represented in kinetic component II of poly(A)$+\text{RNA}_{\text{loose}}$ and poly(A)$+\text{RNA}_{\text{tight}}$. It should be noted that, since there is about four times more poly(A)$+\text{RNA}_{\text{tight}}$ than poly(A)$+\text{RNA}_{\text{loose}}$ in the cell, these abundant species are present at about an equal relative concentration in the two RNA populations.

Are Loosely Membrane-bound Polysomes Artificially Adsorbed Free Polysomes?

Nonspecific adsorption of free polysomes to rough microsomes or RER, by means of the large ribosomal subunits as a result of cell disruption in a low-to-medium monovalent cation concentration buffer, would be expected to be a random event with respect to mRNA species. Therefore, if this accounts for the observation of loosely-membrane bound polysomes, poly(A)$^+\text{RNA}_{\text{loose}}$ should have characteristics identical or very similar to poly(A)$^+\text{RNA}_{\text{free}}$. A comparison of the homologous hybridization data of poly(A)$^+\text{RNA}_{\text{loose}}$ and poly(A)$^+\text{RNA}_{\text{tight}}$ revealed a major difference. About $31\%$ of poly(A)$^+\text{RNA}_{\text{loose}}$ is comprised of about nine sequences that are absent or greatly reduced in poly(A)$^+\text{RNA}_{\text{tight}}$. Sequences of comparable abundance are completely absent from poly(A)$^+\text{RNA}_{\text{tight}}$.

To elucidate further distinctions between the two RNA populations, heterologous reactions were conducted. Hybridizations of poly(A)$^+\text{RNA}_{\text{loose}}$ to kinetically-fractionated abundant and less abundant cDNA$\text{free}$ (28) are shown in Fig. 3 A and B. Fig. 3 A indicates that abundant poly(A)$^+\text{RNA}_{\text{free}}$ sequences are present on the average at about a 10-fold reduced concentration in poly(A)$^+\text{RNA}_{\text{loose}}$. However, the most abundant species appear to be present at roughly equal frequency in the two populations. Fig. 3 B indicates that the least abundant poly(A)$^+\text{RNA}_{\text{free}}$ sequences are present on the average at about a two fold reduced concentration in poly(A)$^+\text{RNA}_{\text{loose}}$. Although clear plateaus were not achieved, the behavior of the curves at high rot values suggests that some sequences in poly(A)$^+\text{RNA}_{\text{free}}$ are either absent or present at greatly reduced concentration in poly(A)$^+\text{RNA}_{\text{loose}}$. The heterologous reaction of cDNA$\text{loose}$ to poly(A)$^+\text{RNA}_{\text{free}}$ shown in Fig. 4 indicates that poly(A)$^+\text{RNA}_{\text{loose}}$ sequences are present at an overall reduced concentration in poly(A)$^+\text{RNA}_{\text{free}}$. The heterologous curve platter at $\approx 13\%$ lower than the homologous curve which indicates that $13/87$ or $15\%$ of poly(A)$^+\text{RNA}_{\text{loose}}$ sequences by mass may be either absent or present at greatly reduced concentrations in poly(A)$^+\text{RNA}_{\text{free}}$.

In an attempt to determine whether there exist poly(A)$^+\text{RNA}_{\text{loose}}$ sequences that are completely absent from poly(A)$^+\text{RNA}_{\text{free}}$, we performed the following experiment. A $10^3$ mass-excess of poly(A)$^+\text{RNA}_{\text{free}}$ was hybridized to cDNA$\text{loose}$ to a rot value of $320\text{ mol-s/l}$ at which the heterologous cDNA$\text{loose}$-poly(A)$^+\text{RNA}_{\text{free}}$ hybridization had reached saturation. The hybridized and unhybridized molecules were separated by hydroxyapatite chromatography and the two cDNA fractions isolated as described previously (28). The cDNA that did not hybridize should be enriched in those sequences that are absent or greatly reduced in poly(A)$^+\text{RNA}_{\text{free}}$.

FIGURE 3 Homologous and heterologous hybridization reactions with kinetically-fractionated cDNA$\text{loose}$. cDNA$\text{loose}$ was kinetically-fractionated into abundant and less-abundant fractions as described (28). Abundant cDNA$\text{loose}$ and less-abundant cDNA$\text{free}$ comprised 32% and 68% of the total. Abundant cDNA$\text{loose}$ (A) and less-abundant cDNA$\text{loose}$ (B) were hybridized to an excess of poly(A)$^+\text{RNA}_{\text{free}}$ (○) or poly(A)$^+\text{RNA}_{\text{loose}}$ (◊) and the extent of reaction assayed as described (28). RNA concentrations ranged from $4 \times 400\mu\text{g/ml}$. Smooth curves were drawn without the aid of a computer.

FIGURE 4 Heterologous hybridization reaction of cDNA$\text{loose}$ to poly(A)$^+\text{RNA}_{\text{free}}$. cDNA$\text{free}$ was hybridized to an excess of poly(A)$^+\text{RNA}_{\text{free}}$ and the extent of reaction assayed as described (28). The solid curve is the corresponding homologous reaction shown in Fig. 1 B. The broken curve representing the heterologous reaction was drawn with the aid of a computer (28). RNA concentrations ranged from 8 to $1700\mu\text{g/ml}$.
do not overlap. The kinetics of the hybridization curves in Fig. 5 B indicate that some of the sequences contained in this 20% that are present in poly(A)"RNAfree are reduced in relative concentration by about 100-fold.

Isolation and Characterization of High Salt, Puromycin-mediated Membrane-released and Membrane-associated Tightly-bound Poly(A) "RNA Fractions

Previous experiments in our laboratory (10) and others (14, 18) have demonstrated that a fraction of liver poly(A)"RNA remains associated with rough microsomes when various treatments are applied in vitro or in vivo to cause breakdown of polysome structures and release of most of the ribosomes from the surface of the membrane. These experiments have been interpreted by some (10, 14) but not others (18) to indicate the existence of a direct interaction between mRNP and the membrane. If such an interaction exists for some membrane-bound mRNA molecules but not others, a functional distinction might exist between the two populations that is reflected in their complexity and frequency distribution, as well as in their degree of uniqueness with respect to each other. The finding that the two populations are distinct in some manner would eliminate the possibility that the two fractions are observed as the result of a random incomplete release of ribosomes and mRNP from the membrane.

Earlier experiments demonstrated that after removal of >90% of the ribosomes from rough microsomes by treatment with 0.5 M KCl in the presence of 1 mM puromycin, ~40% of 3-h pulse-labeled mRNA and 40% of steady state poly(A)"RNA remained associated with microsomes. However, under the conditions used mRNA was degraded. Since we wished to characterize the translational activity of the membrane-associated RNA(poly(A)"RNAfree) and the released RNA(poly(A)"RNAfree), we developed the fractionation scheme referred to as method B in Materials and Methods. Poly(A)"RNA free and poly(A)"RNA ml isolated using this method exhibited number-average lengths of ~1,460 and 1,500 nucleotides, respectively, on formamide-sucrose gradients (Fig. 6) and are translationally active in the rabbit reticulocyte lysate system (see below). Nine independent analytical experiments were conducted to determine the distribution of mRNA be-

between the membrane-associated and membrane-released RNP fractions using this fractionation procedure (Fig. 7). 52.3 ± 17.3% of 3-h pulse-labeled mRNA and 62.1 ± 4.0% of steady state poly(A)"mRNA remained associated with membranes after removal of 85.4 ± 2.5% of large ribosomal subunits.

To obtain poly(A)"RNAfree and poly(A)"RNA ml that are representative of the total poly(A)"RNAhigh population for use in hybridization experiments, we used fractionation method A described in Materials and Methods. This was desirable since recent evidence suggests that postmitochondrial membrane-bound polysomes may be functionally distinct from rapidly sedimenting RER bound polysomes (16, 35, 43). Using method A, a lower percentage of mRNA was recovered in the membrane-associated fraction. This is probably the result of partial degradation of mRNA which occurred using this procedure.
Since mRNP that is released from the membrane due to nucleolytic degradation will contaminate the released fraction, any differences observed between the two RNA fractions must be considered minimal. Using method A, ~31% of 3-h pulse-labeled mRNA and 23% of poly(A)RNA were recovered in the membrane-associated fraction when 85% of total ribosomes were removed.

Poly(A)RNArel and poly(A)RNAassoc were isolated using method A and cDNA prepared as described in Materials and Methods. The homologous hybridization curves are presented in Fig. 8A. Note that the slight differences in the sizes of the driver and tracer populations for the two reactions can only account for a ~10% difference in hybridization kinetics, which would not be discernible and would be in the opposite direction from the difference in kinetics that is apparent. The bulk of poly(A)RNAassoc hybridizes considerably faster than does poly(A)RNArel, although a highly complex component is discernible in poly(A)RNArel that is absent from poly(A)RNArel. An analysis of the homologous hybridizations is given in Table II. The best fits to the data were obtained by assuming four discrete components for poly(A)RNAassoc and three discrete components for poly(A)RNArel. 54% of poly(A)RNArel is comprised of 42 abundant RNA species including one highly abundant species. 11% of the mass of poly(A)RNAassoc consists of a component containing ~19,000 RNA species. In contrast, poly(A)RNArel has components which are neither as abundant nor as rare as those in poly(A)RNAassoc. The combined complexity of the two populations of 29,500 RNA species is very close to that of poly(A)RNAlight of 28,000 species. Although this suggests that they may be nonoverlapping populations, the heterologous curves in Fig. 8B and C indicate that this is not so. The heterologous cDNAassoc-poly(A)RNArel curve (Fig. 8C) plateaus at the same level as the corresponding homologous curve, which indicates that poly(A)RNArel contains all of the sequences of poly(A)RNAassoc. The heterologous cDNAassoc-poly(A)RNAassoc curve (Fig. 8B) plateaus at ~10% below the level of the homologous curve, which indicates that 10% by mass of poly(A)RNArel sequences may be either absent or present at greatly reduced concentrations in poly(A)RNAassoc.

If all of these sequences were derived from the rare abundance class of poly(A)RNArel, they would constitute ~2,000 RNA species. Again, however, differences in plateau levels of this magnitude may not be significant. Therefore, we cannot conclude that the populations are qualitatively unique.

Comparison of In Vitro Translation Products

To obtain further evidence that distinct poly(A)+ mRNA populations are associated with the various subpopulations of membrane-bound polysomes, we analyzed their in vitro translation characteristics in a rabbit reticulocyte lysate system. The translational efficiencies of poly(A)+RANfree, poly(A)+RANlight, poly(A)+RNArel, and poly(A)+RNAassoc are presented in Fig. 9. The translational efficiencies of poly(A)+RANfree and poly(A)+RNAassoc are comparable and are about threefold greater than that of poly(A)+RNArel. As would be expected, the translational efficiency of poly(A)+RNAassoc is intermediate between that of poly(A)+RNArel and poly(A)+RNAassoc. The latter fraction exhibits the lowest efficiency, about one-half that of poly(A)+RNAassoc and one-third that of poly(A)+RNArel.

Translation products were analyzed by one-dimensional gel electrophoresis. A fluorograph of a gel containing the [35S]-methionine-labeled polypeptides produced by all five poly(A)RNA populations is shown in Fig. 10. The gel patterns can be divided into two groups on the basis of qualitative differences. poly(A)+RANfree (Fig. 10, lane c) and poly(A)+RANlight (Fig. 10, lane b) produced a qualitatively

### Table II

**Complexity and Frequency Distribution of High-salt, Puromycin-mediated Membrane-associated and Membrane-released Tightly Bound Polysomal Poly(A)+RNA Populations**

| Polysome class | Abundance class | Percent cDNA hybridized* | Rate Constant | Nucleotide complexity | No. of mRNA species | Copies/cell** |
|----------------|-----------------|--------------------------|---------------|----------------------|---------------------|---------------|
| Membrane-associated | I               | 8.80                     | 1,220         | 13,900               | 3.74 x 10^3     | 1             | 600,000 |
|                  | II              | 46.4                     | 98.3          | 212                  | 2.45 x 10^4     | 41            | 9,200  |
|                  | III             | 33.4                     | 2.59          | 7.75                 | 6.70 x 10^5     | 1,130         | 336     |
|                  | IV              | 11.4                     | 0.053         | 0.467                | 1.11 x 10^7     | 18,800        | 20      |
| Membrane-released | I               | 10.2                     | 307           | 3,010                | 1.55 x 10^3     | 3             | 89,100 |
|                  | II              | 41.0                     | 30.5          | 74.2                 | 6.30 x 10^4     | 124           | 2,190  |
|                  | III             | 47.8                     | 0.478         | 0.981                | 4.77 x 10^6     | 9,890         | 29      |

See Table I footnotes.

* Calculated as described in Table I footnote †. The mass-average sizes of the driver and tracer populations for the membrane-associated RNA reaction were 604 and 590 nucleotides and for the released RNA fraction 562 and 513 nucleotides.

** Calculated as described in Table I footnote ‡. The membrane-associated RNA fraction represents ~62% of the total tightly membrane-bound polysomal poly(A)+RNA or 0.62 x 0.156 = 0.124 pg RNA/cell. The membrane-released RNA fraction makes up the remaining 38% or 0.38 x 0.156 = 0.044 pg/cell.
similar polypeptide pattern distinct from that of the three tightly membrane-bound poly(A)+RNA fractions (Fig. 10, lanes d-f). Within the two groups there are clear quantitative differences in the abundance of specific polypeptides as well as some qualitative differences.

Densitometric tracings of lanes b and c (Fig. 10) representing the translation products of poly(A)⁺RNAfree and poly(A)⁺RNAtight are compared in Fig. 11a. The two low mol wt polypeptides labeled 1 and 3 in the poly(A)⁺RNAfree tracing have not been detected as products of poly(A)⁺RNAtight. The abundant polypeptides labeled 2, 16, and 17 synthesized by poly(A)⁺RNAfree are barely detectable as products of poly(A)⁺RNAtight. Approximately six to eight polypeptides >70,000 daltons have been detected as products of poly(A)⁺RNAfree and these were distinct from the high molecular weight polypeptides synthesized by the tightly membrane-

![Figure 9](image)

**Figure 9**: Translational efficiencies of poly(A)⁺RNA populations. Poly(A)⁺RNAfree (C), poly(A)⁺RNAtight (O), poly(A)⁺RNAmembrane ( ), poly(A)⁺RNAreleased ( ), and poly(A)⁺RNAassociate ( ) were translated at 5-15 µg/ml final concentration in the rabbit reticulocyte lysate system as described in Materials and Methods. 2-µl aliquots of the reaction mixtures were spotted onto filter paper, boiled in 10% TCA, washed in 5% TCA, ethanol, and ether, dried, and radioactivity determined in 10 ml OCS scintillator cocktail.

**Figure 10**: Comparison of in vitro translation products of poly(A)⁺RNA fractions. The five poly(A)⁺RNA fractions in Fig. 9 were translated in the rabbit reticulocyte lysate system as described in Materials and Methods. RNA concentrations were within the linear range for each fraction. A volume of lysate corresponding to 1.70 × 10⁵ dpm of acid insoluble radioactivity from each RNA fraction was subjected to SDS PAGE by the method of Laemmli (19) as described in Materials and Methods. The gel was fixed, treated with Enhance, dried, and fluorographed. Kodak XAR-5 film was exposed for 20 h at -70°C. (Lane a) M₄ standards, phosphorylase B, 92,500; albumin, 69,000; ovalbumin, 46,000; carbonic anhydrase, 30,000. (Lane b) poly(A)⁺RNAfree translation products; (lane c) poly(A)⁺RNAmembrane translation products; (lane d) poly(A)⁺RNAreleased translation products; (lane e) poly(A)⁺RNAassociate translation products; (lane f) poly(A)⁺RNAassociate translation products.

![Figure 11](image)

**Figure 11**: Comparison of in vitro translation products of poly(A)⁺RNA fractions. Densitometric tracings of the translation products shown in lanes b, c, e, and f of Fig. 10 were obtained using a Joyce-Loebel microdensitometer. (a) A comparison of the translation products of poly(A)⁺RNAfree and poly(A)⁺RNAtight shown in Fig. 10, lanes b and c. (b) A comparison of the translation products of poly(A)⁺RNAassociate and poly(A)⁺RNAreleased shown in Fig. 10, lanes e and f.
bound poly(A)\textsuperscript{+}RNA fractions. Polypeptides 4 and 8–17 are all synthesized in greater quantities by poly(A)\textsuperscript{+}RNA\textsubscript{assoc}, whereas polypeptides 5 and 6 are synthesized in greater quantities by poly(A)\textsuperscript{+}RNA\textsubscript{free}.

Densitometric tracings of the poly(A)\textsuperscript{+}RNA\textsubscript{assoc} and poly(A)\textsuperscript{+}RNA\textsubscript{rel} translation products are shown in Fig. 11 B. These were qualitatively very similar, but the same quantitative differences were consistently observed. The polypeptide-labeled 18 is 2.5-fold enriched in poly(A)\textsuperscript{+}RNA\textsubscript{rel}, polypeptides 19–21 are produced in similar quantities by the two fractions, and polypeptides 22–24 and 17 are enriched in the products of poly(A)\textsuperscript{+}RNA\textsubscript{assoc}. Polypeptide 17 has been identified as proalbumin by immunoprecipitation.

DISCUSSION

Our results demonstrate that poly(A)\textsuperscript{+}RNA fractions derived from free polysomes, loosely and tightly membrane-bound polysomes, and membrane-released and membrane-associated RNP have unique characteristics. From this we can eliminate the possibilities that (a) loosely membrane-bound polysomes are the result of random artifactual adsorption of free polysomes to membrane fractions and (b) membrane-associated mRNP is the result of random entrapment of released mRNA or random incomplete release from membranes. Our results do not eliminate artifactual origins that are nonrandom in nature. It is logical to assume, however, that any nonrandom processes that might occur during cell fractionation would themselves be indicative of structural differences among RNP or polysomes.

Recently, Adesnik and Maschio (3) characterized poly(A)\textsuperscript{+}RNA fractions derived from rat liver rough microsomal loosely and tightly bound polysome populations by analysis of their hybridization characteristics. From their data they concluded that loosely bound polysomes are artifactualy adsorbed free polysomes. Any or all of the following may account for the different results obtained by these investigators and ourselves: (a) Adesnik and Maschio (3) isolated polysomes from a rough microsomal fraction representing only a small (7) and perhaps functionally distinct (16, 35) portion of the total RER. In this study we isolated polysomes from a crude membrane fraction containing at least 95% of the total RER. (b) Adesnik and Maschio (3) isolated poly(A)\textsuperscript{+}RNA populations that were partially degraded while our poly(A)\textsuperscript{+}RNA populations were essentially intact. (c) In no case did Adesnik and Maschio demonstrate that their free and loosely membrane-bound polysomal poly(A)\textsuperscript{+}RNA populations were either qualitatively or quantitatively identical, since hybridizations were only carried out to relatively low rot values and plateaus were not observed. In most of our experiments clear plateaus were observed, which allowed us to conclude that the two fractions are at least quantitatively unique. (d) We have observed that rough microsomal fractions isolated by procedures similar to those used by Adesnik and Maschio (3) may be extensively contaminated with trapped free polysomes, i.e., polysomes that can be extracted by low salt washes (Mueckler and Pitot, unpublished data, see also reference 24). Since these contaminating free polysomes were not removed from microsomal fractions before extraction of high-salt releasable polysomes, they would be included in the latter fraction. Thus it is reasonable that Adesnik and Maschio (3) observed similar hybridization characteristics between these fractions, as a significant portion of their loosely bound polysome fraction may actually have consisted of trapped free polysomes. We have determined that low-salt-extractable polysomes comprise <1% of our loosely membrane-bound polysome fraction, which comprises 22% of total membrane-bound polysomes. Adesnik and Maschio (3) found that their loosely membrane-bound polysome fraction comprised 33% of total microsomal-bound polysomes. Thus, about one-third of their loosely membrane-bound polysomes may, in fact, have been trapped free polysomes.

Adesnik and Maschio (3) also conducted hybridization experiments with microsomal-associated and nonassociated RNA after in vivo disaggregation of polysome structures with ethionine. Their data suggested that this treatment resulted in a random release of mRNA from microsomal membranes. They concluded that the retention of mRNA on microsomal membranes observed by us (10) after in vitro treatment with 0.5 M KCl-puromycin is not functionally significant. However, it is clear that the mRNA fractions examined in each case are not equivalent. After disaggregation of polysomes in vivo, they did not wash microsomal membranes with high-salt buffer to fractionate mRNA into membrane-associated and membrane-released components. Since it had been reported earlier (18) that ribosomes and mRNA are not released from microsomal membranes after ethionine treatment in vivo unless these are washed with high-salt buffer, the significance of the RNA fractions examined by Adesnik and Maschio (3) is questionable. The release they may have been due to random nucleolytic degradation of mRNA in vitro after mRNP had been denuded of ribosomes in vivo.

The in vitro translation experiments and homologous and heterologous hybridizations involving poly(A)\textsuperscript{+}RNA\textsubscript{assoc} and poly(A)\textsuperscript{+}RNA\textsubscript{rel} indicate that (a) these two RNA fractions contain a qualitatively identical set of abundant mRNAs that are differentially distributed between them, (b) poly(A)\textsuperscript{+}RNA\textsubscript{assoc} is enriched in species that are very abundant and very rare and (c) poly(A)\textsuperscript{+}RNA\textsubscript{rel} may contain some species that are absent from poly(A)\textsuperscript{+}RNA\textsubscript{assoc}, but the former contains all of the species present in the latter. Although these results suggest the possibility of a functional distinction between these RNA fractions, they do not suggest what this distinction is. To shed light on this question we are currently conducting hybridization experiments with several specific cloned cDNA probes to determine the relative distribution of specific mRNAs between these RNA populations.

Shields (41) used in vitro translation to characterize membrane-associated and membrane-released mRNA populations of dog pancreas. He found a much smaller percentage of mRNA in the membrane-associated fraction (3–15%) and the in vitro translation products of the two fractions were quantitatively and qualitatively very similar. These data are not necessarily in conflict with the results presented here, since there is no reason to suspect that the processes examined need be identical in liver and pancreas. Pancreatic acinar cells are highly specialized for the synthesis and secretion of a relatively small number of polypeptides. Hepatocytes, however, are functionally much more diverse. Thus, it is reasonable that these two cell types might exhibit unique characteristics.

The results of pollysome disaggregation experiments have usually (10, 14), but not always (18), been interpreted to indicate the existence of a direct association between mRNA or mRNP and rough microsomal membranes, but there is no direct structural evidence to support this. Therefore, we refer to these fractions as “membrane-released” and “membrane-associated” in a purely operational sense. In particular, it cannot be ruled out that the membrane-bound ribosomes that
are resistant to release by high-salt and puromycin treatment are involved in the binding of mRNA. If so, this interaction probably doesn’t involve nascent chains since the two RNA fractions encode for an identical set of abundant polypeptides, and puromycin is present at a large molar excess during fractionation. The nature of membrane interaction involving the resistant ribosomes is obscure (1). In the case of ethionine disaggregation of liver polysomes in vivo, ribosomes are apparently not involved in the binding of mRNA to microsomal membranes (14). Under conditions where the integrity of mRNA was preserved, Endo and Natori (14) also found a much larger proportion of mRNA associated with microsomal membranes than was reported by Adesnik and Maschio (3), and mRNA coding for serum albumin was found to be preferentially retained. These results again suggest that the randomization of mRNA sequences observed by Adesnik and Maschio (3) was the result of release caused by random nucleolytic degradation. If an interaction exists between mRNA and rough membranes in vivo as well as in vitro, it is not likely to be involved in the selection of messages for translation on membrane-bound polysomes, as experiments with myeloma cells in vivo (25) have demonstrated that this process is dependent on translation, and probably involves nascent chains (6).

Loosely membrane-bound polysomes have been shown to be active in the synthesis of cytochrome b₅ (15), ribosomal structural proteins (29), and histones (45). All of these are also synthesized on free but not on tightly membrane-bound polysomes at significant levels. Unfortunately, cross-contamination levels were only determined in the case of the ribosomal structural proteins (29). More work is needed to determine whether the results presented here are indicative of a functionally distinct subpopulation of membrane bound polysomes or simply of a nonrandom adsorption of free polysomes to membranes during cell fractionation. The necessity of working with polysome preparations that exhibit minimal cross-contamination between low and high-salt extractable fractions cannot be over-emphasized.

The finding that classes of proteins that must transverse the nuclear envelope are synthesized on loosely bound polysomes suggests a possible function for the loose interaction. RER is known to be intimately associated with the nuclear envelope. It is possible that the loose interaction functions to establish a concentration gradient of polysomes synthesizing nuclear proteins in the vicinity of the nucleus. In this regard Lønn (23) has demonstrated that Balbiani ring 75S RNA, which is associated with RER in a high-salt, puromycin-resistant linkage (22), is concentrated in the cytoplasm proximal to the nucleus. This message apparently codes for secretory polypeptides.

Similarly, in the case of integral membrane proteins such as cytochrome b₅ whose synthesis occurs on loosely bound polysomes and whose integration is not cotranslational, the loose interaction may function to allow the proper topographical and spatial orientation of newly completed polypeptides for post-translational integration. The loose interaction may conceivably involve any of the component structures of polyribosomes. Specificity for the interaction could reside within the mRNA molecule itself or the polypeptide chain. In analogy with the signal hypothesis (6), the loose interaction could be initiated by transient ionic interactions between the growing nascent chain and specific membrane proteins exposed on the cytoplasmic face of the membrane. This would allow the large ribosomal subunits to bind to the receptor sites on the membrane surface (1), establishing the high-salt sensitive binding that is observed in vitro. This, in turn, would allow disposal of newly completed polypeptides at the surface of the membrane, facilitating subsequent integration.

It should be mentioned that, while the free monovalent cation concentration of our initial homogenization buffer was close to physiological, nonphysiological concentrations are required to release loosely membrane-bound polysomes. This is consistent with the idea that this interaction exists in vivo as well as in vitro.

In conclusion, we have provided considerable evidence that the poly(A)⁺RNA fractions associated with free and loosely membrane bound polysomes, and membrane-associated and membrane released mRNA, are discrete in the sense that they are not identical. However, the data indicates that there are substantial qualitative similarities between these populations, much of which cannot be attributed to cross-contamination. Further experiments are required to determine whether the differences observed are of functional significance.

We thank Dr. Frank Gonzalez for his help with the in vitro translation system.

This work was supported in part by National Institutes of Health grants CA-07175, CA-22484, and CA-09135.

Received for publication 28 September 1981, and in revised form 2 March 1982.

REFERENCES

1. Adelman, M. J., D. S. Sabatini, and G. Blobel. 1973. Ribosome-membrane interaction. Nondestructive disassembly of rat liver rough microsomes into ribosomal and membrane components. J. Cell Biol. 56:206-219.
2. Adelman, M. J., M. Lande, T. Martin, and D. S. Sabatini. 1976. Retention of mRNA on the endoplasmic reticulum membranes after in vivo disassembly of polysomes by an inhibitor of initiation. J. Cell Biol. 71:307-312.
3. Adesnik, M., and F. Maschio. 1981. Segregation of specific classes of messenger RNA into free and membrane-bound polysomes. Eur. J. Biochem. 114:221-228.
4. Banerje, J., A. H. Maxwell, and W. E. Hahn. 1976. Specificity of oligo(dT) cellulose chromatography in the isolation of polyadenylated RNA. Anal. Biochem. 72:413-427.
5. Bishop, J. O., J. G. Morton, M. Rees, and M. Richardson. 1974. Three abundance classes in HeLa cell messenger RNA. Nature (Lond.). 250:199-204.
6. Blobel, G., and B. Dobberstein. 1975. Transfer of proteins across membranes. I. Presence of proteinolytically processed and unprocessed nascent immunoglobulin light chains on membrane-bound ribosomes of mouse myeloma. J. Cell Biol. 67:835-851.
7. Blobel, G., and V. R. Potter. 1967. Studies on free and membrane-bound ribosomes in rat liver. I. Distribution as related to total cellular RNA. J. Mol. Biol. 26:279-292.
8. Blobel, G., and V. R. Potter. 1967. Studies on free and membrane-bound ribosomes in rat liver. II. Interaction of ribosomes and membranes. J. Mol. Biol. 26:293-310.
9. Capetanakis, Y. G., and A. Alonso. 1980. Comparison of polyosomal and nuclear poly(A) containing RNA populations from normal rat liver and Novikoff hepatoma. Nucleic Acids Res. 8:3193-3214.
10. Cardelli, J. A., and H. C. Pirot. 1976. Direct association of messenger RNA labeled in the presence of fluorouracil with membranes of the endoplasmic reticulum in rat liver. J. Cell Biol. 70:47-58.
11. Chamberlin, M. E., G. A. Galau, R. J. Britten, and E. H. Davidson. 1978. Studies on eukaryotic and reassociating kinases. V. Effects of disparity in tracer and driver fragment lengths. Nucleic Acids Res. 5:2073-2094.
12. Chirgwin, J. M., A. E. Przybyla, R. J. MacDonald, and W. J. Rutter. 1979. Isolation of biologically active ribonucleic acid from sources enriched in ribonuclease. Biochemistry. 18:5294-5299.
13. Clouston, P. M., P. J. Mason, and J. O. Bishop. 1981. Comparison of poly(A)⁺mRNA prepared from membranes and free polysomes of mouse liver. Proc. Natl. Acad. Sci. U. S. A. 78:3697-3701.
14. Endo, Y., and Y. Natori. 1978. Direct association of messenger RNA-containing ribosome-electron spin labled polysomes, lipid and membrane fractions of endoplasmic reticulum in ethanol-treated rat liver. Biochim. Biophys. Acta. 562:281-291.
15. Gonzalez, F. J., and C. B. Kasper. 1980. Phenobarbital induction of P-450 oxidoreductase messenger ribonucleic acid. Biochemistry. 19:1790-1796.
16. Gonzalez, F. J., and C. B. Kasper. 1981. Subcellular fractionation of rat liver. Methods Enzymol. 69:307-313.
17. Harrison, T. M, G. G. Brownlee, and C. Milstein. 1974. Studies on polysome-membrane interactions in mouse myeloma cells. Eur. J. Biochem. 47:413-420.
18. Kropp, J. D., and D. S. Sabatini. 1977. Release of poly(A)⁺ messenger RNA from rat liver rough microsomes upon disassembly of bound polysomes. J. Cell Biol. 74:414-427.
19. Lauemni, V. K. 1970. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. Nature (Lond.). 227:680-683.
20. Lake, M., and T. Hultin. 1980. Characterization of a poly(A)⁺RNA-containing structural component directly associated with cytoplasmic membranes in dormant artemia cysts. Biochim. Biophys. Acta. 606:286-295.
21. Lande, M. A., M. Adesnik, M. Sumida, Y. Tashiro, and D. S. Sabatini. 1975. Direct...
association of messenger RNA with microsomal membranes in human diploid fibroblasts. 

J. Cell Biol. 165:513-528.

22. Löven, V. 1977. Direct association of Balbiani ring 75S RNA with membranes of the endoplasmic reticulum. Nature (Lond.) 270:630-631.

23. Löven, V. 1978. Delayed flow-through cytoplasm of newly synthesized Balbiani ring 75S RNA. Cell. 13:727-733.

24. McIntosh, P. R., and K. O'Toole. 1976. The interaction of ribosomes and membranes in animal cells. Biochim. Biophys. Acta 457:171-212.

25. Mechler, B. 1981. Membrane-bound ribosomes of myeloma cells VI. Initiation of immunoglobulin mRNA translation occurs on free ribosomes. J. Cell Biol. 88:42-50.

26. Mechler, B., and P. Vassani. 1975. Membrane-bound ribosomes of myeloma cells. III. The role of the messenger RNA and the nascent polypeptide chain in the binding of ribosomes to membranes. J. Cell Biol. 67:25-37.

27. Milcarek, C., and S. Penman. 1974. Membrane-bound polyribosomes in HeLa cells: association of polyadenylic acid with membranes. J. Mol. Biol. 89:327-338.

28. Mueckler, M., and H. C. Piat. 1981. Structure and function of rat liver polysome populations. I. Complexity, frequency distribution, and degree of uniqueness of free and membrane-bound polysomal polyadenylate-containing RNA populations. J. Cell Biol. 90:495-506.

29. Nabeshima, Y., K. Tsurgui, and K. Ogata. 1975. Preferential biosynthesis of ribosomal structural proteins by free and loosely bound polysomes from regenerating rat liver. Biochim. Biophys. Acta 416:30-42.

30. Ordahl, C. P., T. R. Johnson, and A. I. Caplan. 1976. Sheared DNA fragment sizing: comparison of techniques. Nucl. Acids Res. 3:2985-2999.

31. Ouelfate, A. J., M. B. Silverberg, and T. A. Matl. 1981. Association of polyadenylic acid-deficient messenger ribonucleic acid with membranes in mouse kidney. Biochemistry. 20:3561-3567.

32. Palmiter, R. D. 1974. Magnesium precipitation of ribonucleoprotein complexes. Expedition techniques for the isolation of undegraded polysomes and messenger ribonucleic acid. Biochemistry. 13:3606-3615.

33. Pearson, W. R., E. H. Davidson, and R. J. Britten. 1977. A program for least squares analysis of reassociation and hybridization data. Nucl. Acids Res. 4:1727-1735.

34. Peavy, D. E., J. M. Taylor, and L. S. Jeffersoon. 1978. Correlation of albumin production rates and albumin mRNA levels in livers of normal, diabetic, and insulin-treated diabetic rats. Proc. Natl. Acad. Sci. U. S. A. 75:5879-5883.

35. Pickett, C. B., N. R. Rosenberg, and R. L. Jeter. 1977. The physical association between rat liver mitochondria and rough endoplasmic reticulum. Exp. Cell Res. 132:225-234.

36. Ramsey, J. C., and W. J. Steele. 1976. A procedure for the quantitative recovery of homogeneous populations of undegraded free and bound polysomes from rat liver. Biochemistry. 15:1704-1712.

37. Ramsey, J. C., and W. J. Steele. 1976. Differences in size, structure, and function of free and membrane-bound polyribosomes of rat liver. Evidence for a single class of membrane-bound polyribosomes of rat liver. Biochem. J. 160:1-8.

38. Rosbash, M., and S. Penman. 1971. Membrane-associated protein synthesis of mammalian cells. I. The two classes of membrane-associated ribosomes. J. Mol. Biol. 59:227-241.

39. Rosbash, M., and S. Penman. 1971. Membrane-associated protein synthesis of mammalian cells. II. Isopycnic separation of membrane-bound polyribosomes. J. Mol. Biol. 59:243-253.

40. Savage, M. J., J. M. Sals-Trepat, and J. Bonner. 1978. Measurement of the complexity and diversity of polyadenylic acid containing messenger RNA from rat liver. Biochemistry. 17:462-467.

41. Shields, D. 1979. Association of messenger ribonucleic acid with mammalian microsomal membranes: characterization by analysis of cell-free translation products. Biochemistry. 18:2622-2627.

42. Shore, G. C., and J. R. Tata. 1977. Functions for polyribosome-membrane interactions in protein synthesis. Biochin. Biophys. Acta. 519:173-193.

43. Shore, G. C., and J. R. Tata. 1977. Two fractions of rough endoplasmic reticulum from rat liver. I. Cytoplasmic messenger RNA's which code for albumin and mitochondrial proteins are distributed differently between the two fractions. J. Cell. Biol. 72:726-743.

44. Wettner, J. G., and N. Davidson. 1968. Kinetics of renaturation of DNA. J. Mol. Biol. 31:349-370.

45. Zauderer, M., P. Liberti, and C. Baglioni. 1973. Distribution of histone messenger RNA among free and membrane-associated polyribosomes of a mouse myeloma cell line. J. Mol. Biol. 79:577-586.