**ARTICLE**

**SFI1 promotes centriole duplication by recruiting USP9X to stabilize the microcephaly protein STIL**

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In mammals, centrioles participate in brain development, and human mutations affecting centriole duplication cause microcephaly. Here, we identify a role for the mammalian homologue of yeast SFI1, involved in the duplication of the yeast spindle pole body, as a critical regulator of centriole duplication in mammalian cells. Mammalian SFI1 interacts with USP9X, a deubiquitylase associated with human syndromic mental retardation. SFI1 localizes USP9X to the centrosome during S phase to deubiquitylate STIL, a critical regulator of centriole duplication. USP9X-mediated deubiquitylation protects STIL from degradation. Consistent with a role for USP9X in stabilizing STIL, cells from patients with USP9X loss-of-function mutations have reduced STIL levels. Together, these results demonstrate that SFI1 is a centrosomal protein that localizes USP9X to the centrosome to stabilize STIL and promote centriole duplication. We propose that the USP9X protection of STIL to facilitate centriole duplication underlies roles of both proteins in human neurodevelopment.

**Introduction**

In mammalian cells, centrosomes are microtubule organizing centers that participate in cellular process such as ciliogenesis and cellular division (Nigg and Stearns, 2011; Nigg and Holland, 2018). Centrosomes are composed of centrioles embedded within protein-rich matrices. To ensure bipolar spindle formation, centrioles are duplicated exactly once during S phase by forming procentrioles at the base of the existing centrioles (Hinchcliffe et al., 1999; Lacey et al., 1999; Meraldi et al., 1999; Haase et al., 2001). Disruptions in centriole duplication can result in the loss of apical attachment and premature differentiation in the developing brain (Jayaraman et al., 2016; Johnson et al., 2018). Cancer cells can possess supernumerary centrioles, which are associated with abnormal mitoses and DNA damage (Ganem et al., 2009; Godinho et al., 2014; Levine and Holland, 2018; Nigg and Holland, 2018).

Many of the genes mutated in primary microcephaly (MCPH), a neurodevelopmental disorder characterized by a small head and brain, encode centrosomal proteins involved in promoting centriole duplication (Bond et al., 2005; Zhong et al., 2005; Kumar et al., 2009; Guernsey et al., 2010; Nicholas et al., 2010; Yu et al., 2010; Sir et al., 2011; Lin et al., 2013; Kodani et al., 2015). Of these MCPH-associated proteins, PLK4, STIL and SAS6 cooperatively initiate the formation of procentrioles, an early step in centriole duplication (Leidel et al., 2005; Ohta et al., 2014; Moyer et al., 2015). Subsequently, other MCPH-associated proteins (i.e., CDK5RAP2, CEP152, WDR62, CEP63, ASPM, and CPAP) are recruited to the centrosome in a step-wise manner to elongate newly formed procentrioles (Kodani et al., 2015; Jayaraman et al., 2016; Johnson et al., 2018). Thus, MCPH mutations alter centrosome organization and attenuate centriole duplication.

To precisely regulate centriole duplication, ubiquitylation and proteasome-mediated degradation control the abundance of procentriole initiating factors (Cunha-Ferreira et al., 2009; Holland et al., 2010; Puklowski et al., 2011; Arquint et al., 2018). For example, the stabilities of PLK4 and STIL are limited by the E3 ubiquitin ligase SCFF-box (Cunha-Ferreira et al., 2009; Guderian et al., 2010; Holland et al., 2010; Arquint et al., 2018). Conversely, deubiquitylation can protect centrosomal proteins such as CP110 from degradation to promote centriole duplication (Li et al., 2013). Whether proteins such as STIL are also protected from ubiquitin-mediated degradation has not been clear.

SFI1 is an evolutionarily conserved protein first discovered in yeast, where it functions to promote spindle pole body duplication (Kilmartin, 2003; Li et al., 2006). The human homologue of SFI1 localizes to the centrosome and binds the distal centriole component Centrin 2 (Kilmartin, 2003; Martinez-Sanz et al., 2006). Whether human SFI1 functions in centrosome biogenesis...
has been unclear. We found that human SFI1 promotes centriole duplication by stabilizing the procentriole initiating factor STIL. SFI1 limits the K48-linked ubiquitylation and degradation of STIL. In investigating how SFI1 restricts STIL ubiquitylation, we found that, during S phase, SFI1 binds and localizes USP9X, a deubiquitylating enzyme (DUB), to the centrosome. At the centrosome, USP9X binds and deubiquitylates STIL. USP9X is mutated in female-restricted X-linked syndromal mental retardation 99 (MRX99F; Reijnders et al., 2016). Consistent with a role for USP9X in stabilizing STIL, cells from MRX99F-affected individuals have reduced levels of STIL. Thus, SFI1 recruits USP9X to the centrosome to deubiquitylate and stabilize STIL and promote centriole duplication.

Results

SFI1 accumulates at the centrosome during S phase

In Saccharomyces cerevisiae, SFI1 is a key regulator of duplication of the spindle pole body, the functional equivalent of the centrosome (Rüthnick and Schiebel, 2016). Tetrahymena SFI1-related proteins localize around basal bodies, bolstering evidence for an evolutionarily ancient connection between SFI1 and centrosomes (Stemm-Wolf et al., 2013). To assess whether the human homologue of SFI1 is associated with centrosomes, we generated an antibody to human SFI1 and costained HeLa cells for SFI1 and Centrin 2, a centriolar structural component (Fig. 1 A). Human SFI1 was absent from mitotic spindle poles but localized around the interphase centrosome, peaking during S phase (Fig. 1 A). We confirmed the specificity of our antibody by immunoblot using two nonoverlapping siRNAs directed to human SFI1 (Fig. 1 B). To further test the centrosomal enrichment of SFI1, we isolated centrosomes from HeLa cells and found that SFI1 cofractionated with the centrosomal component γ-tubulin (Fig. 1 C). SFI1 protein levels did not decrease in mitosis, suggesting that its cell cycle–dependent localization to the centrosome is not secondary to differential stabilization (Fig. 1 D).

Using GFP-tagged SFI1, we assessed the localization in S phase after 6 and 12 h of transfection. GFP-SFI1 localized to interphase centrosomes (Fig. 1 E). Interestingly, cells exhibited supernumerary Centrin foci that colocalized with the mature centriolar protein CP110 (Chen et al., 2002) 12 h after GFP-SFI1 transfection, suggesting that increased SFI1 expression may promote centriole duplication (Fig. 1, F and G; and Fig. S1 A).

As SFI1 localizes to a cloud around the centrosome during S phase, we examined the localization of SFI1 relative to centrosomal satellites, particles that exist at the periphery of the centrosome (Kubo et al., 1999; Dammermann and Merdes, 2002; Kodani et al., 2015). SFI1 partially colocalized with the satellite component CEP131 in both HeLa and U2OS cells (Fig. S1 B). PCM1 is a prominent component of centriolar satellites and is essential for centriolar satellite integrity (Dammermann and Merdes, 2002; Lopes et al., 2011). To confirm that SFI1 localizes to satellites, we examined the localization of SFI1 in PCM1 knockout and siRNA-depleted cells (Fig. S1, C and D). In the absence of centriolar satellites, SFI1 accumulated at the centrosome, similar to other satellite proteins (Stowe et al., 2012; Staples et al., 2014; Wang et al., 2016).

As centriole duplication requires centriolar and pericentriolar components, we examined the organization of both in SFI1-depleted cells. HeLa cells depleted of SFI1 displayed increased centrosomal localization of CEP135, an MCPH-associated protein required for centriole assembly and stability (Fig. 1 H; Bayless et al., 2012; Lin et al., 2013). Interestingly, the distal centriolar protein CP110 formed an amorphous cloud near the centrioles in SFI1-depleted cells (Fig. 1 I). Immunoblot analysis revealed that SFI1 depletion led to decreased levels of CP100, but not CEP135 (Fig. 1 J), further indicating that SFI1 participates in the organization of the pericentrosomal matrix. As CP110 depletion increases pericentrosomal localization of CEP135 (Schmidt et al., 2009; Al-Hakim et al., 2012), it is possible that the destabilization of CP110 in SFI1-depleted cells underlies the accumulation of CEP135 at the centrosome.

SFI1 is required for centriole duplication

To determine whether SFI1 is required for centriole duplication, we examined whether depletion of SFI1 abrogates centriole duplication. Similar to previous reports, siRNA-mediated depletion of SFI1 reduced the centriole number (Fig. 2, A and B; Balestra et al., 2013). We examined centrosomes in S phase control and SFI1 siRNA-treated cells by serial section electron microscopy. Control cells formed procentrioles associated with the preexisting centrioles, while SFI1-depleted cells failed to duplicate their centrioles (Figs. 2 C and S2 A). Thus, human SFI1, like its yeast homologue, is required for centriole duplication. Because of the severity of the centriole-duplication defects in SFI1-depleted cells, we examined the localization of the early initiating factors for centriole duplication, SAS6 and STIL (Strnad et al., 2007; Vulprecht et al., 2012).

Interestingly, SFI1-depleted cells failed to recruit SAS6 and STIL to the S phase centrosome (Fig. 2, D and E; and quantified in Fig. S2 B). Immunoblot analysis revealed that depletion of SFI1 did not significantly affect SAS6 levels but dramatically reduced STIL levels, suggesting that the failure of centriole duplication in SFI1-depleted cells could be due to the destabilization of STIL (Fig. 2 F). To gain insight into how SFI1 stabilizes STIL, we mapped the domain of STIL stabilized by SFI1. DLL-1 cells expressing full-length STIL, the N-terminal domain, MD1 (aa 186–441), MD2 (aa 442–714), MD3 (aa 715–988), or the C-terminal domain (Moyer et al., 2015) were transfected with scrambled control (SC) or SFI1 siRNA. Immunoblot analysis revealed that SFI1 depletion disrupted the stability of full-length STIL and the MD3 domain of STIL (Fig. 2 G), suggesting that SFI1 stabilizes STIL through aa 715–988.

As SFI1 localizes and stabilizes centrosomal CP10 (Fig. 1, I and J), and CP110 is implicated in centriole duplication (Chen et al., 2002; Yadav et al., 2016), we examined whether restoring CP110 stability could rescue centriole duplication in SFI1-depleted cells. To restore CP110 stability, we depleted NEURL4, a centrosomal ubiquitin ligase that targets CP110 for degradation (Li et al., 2012). Immunoblot analysis revealed that depletion of NEURL4 restored CP110 levels in SFI1 knockdown cells (Fig. S2 C). Immunostaining and quantification of S phase cells revealed that restoring CP110 levels was insufficient to restore centriole duplication (Fig. S2, D and E), indicating that SFI1 promotes centriole duplication via a mechanism independent of CP110.

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SFI1 interacts with and recruits USP9X to the centrosome

As a proximity interactor database for centrosomal proteins detected STIL interactors (Gupta, 2018). As ubiquitin-modifying enzymes transiently interact with their respective substrates, we loosened the stringency for the BioID data to include low confidence STIL proximity interacting proteins and identified several DUBs, including USP9X and USP14, as potential interactors of STIL. We used immunoprecipitation of endogenous STIL to assess whether it interacts with two of these DUBs. Reciprocal immunoprecipitation demonstrated that STIL co-immunoprecipitated with both USP9X and USP14 (Fig. 3, A and B). We confirmed that these interactions were specific, as related DUBs failed to coimmunoprecipitate with STIL (Fig. 3 A). The directness of these interactions remains to be determined.

USP9X has been proposed to control the levels of several centrosomal proteins, such as CEP131 and PCM1, to promote centriole duplication (Li et al., 2017; Wang et al., 2017). To begin to assess whether USP9X controls STIL function, we tested whether it, similar to STIL, participated in centriole duplication. Depletion of USP9X (but not USP14, USP7, USP15, or UCHL1) attenuated centriole duplication, phenocopying depletion of STIL (Figs. 3 C and S3 A). Depletion of USP9X, but not USP14 or the other DUBs, decreased the centrosomal localization and protein levels of STIL (Fig. 3, D and E; and Fig. S3, B and C).

To further assess whether USP9X affects STIL levels, we expressed HA-USP9X or enzymatically inactive HA-USP9X C1566A and analyzed STIL levels 6 h after transfection. Elevated expression of HA-USP9X increased STIL levels while HA-USP9X C1566A did not (Fig. 3, F and G; and Fig. S3 D). As USP9X is a deubiquitylase, we examined whether the stabilization of STIL in HA-USP9X–expressing cells is associated with decreased STIL ubiquitylation. Expression of HA-USP9X reduced K48-linked ubiquitylated STIL (Fig. 3 H and Fig. S3 E). Together, these findings demonstrate that USP9X promotes STIL levels.

As SFI1 and USP9X interact and depletion of each phenocopies the other, we hypothesized that SFI1 localizes USP9X to the centrosome.
centrosomes during interphase. To confirm that USP9X localizes to centrosomes (Li et al., 2017; Wang et al., 2017), we costained HeLa cells with Centrin to mark centrioles and USP9X. Like SFI1, USP9X localized to the centrosome during S phase (Fig. 3 I). siRNA-mediated depletion of USP9X confirmed the specificity of the immunofluorescence (Fig. S3 F). We did not detect USP9X at the centrosome during other stages of the cell cycle, as previously reported (Li et al., 2017). USP9X cofractionated with
γ-tubulin, confirming its localization to the centrosome (Fig. S3 G). USP9X levels were only modestly higher during the G1 and S phases, suggesting that its centrosomal localization is not due to selective degradation at other phases (Fig. S3 H). Together, these results demonstrate that USP9X, like its interactor SFI1, is critical for STIL accumulation at the S phase centrosome.

Depletion of SFI1 also blocked the centrosomal accumulation of USP9X (Figs. 3 J and S3 I). SFI1 was not required to stabilize USP9X (Fig. S3 J), indicating that SFI1 is required specifically for USP9X localization to the centrosome. Depletion of USP9X did not affect either the centrosomal localization or stability of SFI1, demonstrating that SFI1 is required for USP9X centrosomal localization, but not vice versa (Fig. S3, K and L). Thus, SFI1 is essential for localizing USP9X to the centrosome where it stabilizes STIL to promote centriole duplication.

USP9X directly deubiquitylates STIL

As USP9X is a DUB that controls STIL levels, we investigated whether pharmacological inhibition of USP9X decreases STIL stability. We treated HeLa cells with WP1130 (Degrasyn), a

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partially selective inhibitor of USP9X (Peterson et al., 2015). WP1130 sharply decreased STIL levels by one hour and reduced STIL to undetectable levels by two hours (Fig. 4 A). As STIL is required for centriole duplication, we examined whether WP1130 also affected centriole number. Immunofluorescence staining for STIL and Centrin revealed that WP1130 reduced STIL localization at centrosomes, and reduced centriole numbers (Fig. 4, B and C; Fig. S4 A; and quantified in Fig. S4 B). To determine whether WP1130 disrupted the stability of other centrosomal components, we examined the localization and stability of SFI1, CP110 and CEP135. SFI1 and CP110 localization and levels were reduced after 2 h treatment with WP1130 (Fig. S4, C–I). Interestingly, CEP135 formed cytoplasmic aggregates away from the centrosome in WP1130-treated cells but did not affect CEP135 stability (Fig. S4, E, H, and I). These findings indicate that WP1130 disrupts centrosomal organization.

BioID mapping suggested that USP9X is in proximity to STIL (Gupta, 2018). Reciprocal immunoprecipitations confirmed that USP9X and STIL interact (Fig. 4 D). To define which domain of STIL binds USP9X, we immunoprecipitated GFP- and Myc-tagged STIL and the indicated STIL fragments were immunoprecipitated using antibody to GFP. STIL or its fragments were detected by immunoblotting for Myc, respectively. Asterisk denotes non-specific band. Coinmuno precipitating endogenous USP9X was detected by immunoblotting for USP9X.

Figure 4. USP9X deubiquitylates STIL. (A) Total cell lysates from HeLa cells treated with DMSO or 5 µM WP1130 for 1 or 2 h were immunoblotted for USP9X and STIL. Actin served as a loading control. (B) S phase HeLa cells treated with DMSO or WP1130 for 1 or 2 h were stained for Centrin (green) and STIL (red). Scale bars represent 5 µm for all images and 1 µm for inset images. (C) Quantification of the centrosomal fluorescence intensity of STIL in DMSO- and WP1130-treated HeLa cells expressed as the ratio ± SD to the fluorescence intensities of control cells. For all quantifications, 10 cells were analyzed per experiment (n = 3). *P < 0.005 (t test). (D) Reciprocal coimmunoprecipitation (IP) of endogenous USP9X and STIL from HeLa total cell lysates. Efficient precipitation and coprecipitation were detected using antibodies for USP9X and STIL. c-Myc served as a negative control. (E) Control (parental) DLD-1 or stable DLD-1 cells expressing full-length GFP-Myc-tagged STIL or the indicated STIL fragments were immunoprecipitated using antibody to GFP. STIL or its fragments were detected by immunoblotting for Myc, respectively. Asterisk denotes non-specific band. Coimmunoprecipitating endogenous USP9X was detected by im-

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and CEPI35. SFI1 and CP110 localization and levels were reduced after 2 h treatment with WP1130 (Fig. S4, C–I). Interestingly, CEPI35 formed cytoplasmic aggregates away from the centrosome in WP1130-treated cells but did not affect CEPI35 stability (Fig. S4, E, H, and I). These findings indicate that WP1130 disrupts centrosomal organization.
full-length STIL and portions thereof, and used communo-
precipitation to determine which domains are required to in-
teract with USP9X. Interestingly, the MD3 fragment of STIL,
the same domain stabilized by SFI1, interacted with USP9X
(Fig. 4 E). To further determine whether STIL and USP9X in-
teract, we incubated recombinant USP9X or USP7, as a control,
with lysates from DLD-1 cells expressing GFP- and Myc-tagged
full-length SFI1 or the MD3 fragment thereof. Full-length STIL
and the MD3 fragment of STIL were precipitated with GST-
USP9X (catalytic domain aa 1,531–1,972), but not GST-USP7
(Fig. 4 F and Fig. S4 O and P), suggesting that USP9X interacts
with STIL through its MD3 domain.

As we had done to determine which domains are required for
SFI1 to stabilize STIL, we assayed which domain of STIL was
stabilized by USP9X by depleting USP9X in cells expressing full-
length STIL or fragments of STIL. Similar to full-length STIL, the
MD3 fragment, but not other fragments, were destabilized in the
absence of USP9X (Fig. 4 G and Fig. S4 Q). These results are
consistent with the idea that SFI1 and USP9X function together
to stabilize STIL through aa 715–988.

To test whether the MD3 fragment affects STIL stability, we
cloned constructs expressing full-length Myc-tagged STIL and
STIL lacking the MD3 domain (STILΔMD3). We cotransfected
the STIL constructs with GFP to normalize for transfection ef-
niciency and assessed protein levels. Interestingly, STILΔMD3
was significantly less stable than full-length STIL (Fig. S4 R). In
addition to being less stable, STILΔMD3 did not bind USP9X
(Fig. 4 H). Thus, USP9X binds to and stabilizes STIL through its
MD3 domain.

STIL is ubiquitylated and degraded during mitosis by the
anaphase-promoting complex/cyclosome (APC/C) via its KEN
box, a domain defined by the peptide sequence K-E-N-X-X-N
(Arquint and Nigg, 2014). USP9X removes K48 polyubiquitin
chains to protect proteins from proteasome-mediated degrada-
tion (Thrower et al., 2006; Marx et al., 2010). Because USP9X
stabilizes STIL MD3, we hypothesized that STIL is K48 poly-
ubiquitylated during interphase independent of APC/C. By
immunoprecipitating STIL and immunoblotting for K48 poly-
ubiquitin, we found that both STIL and STIL lacking its KEN
box are K48 polyubiquitylated (Figs. 4 I and S4 S), revealing
that STIL ubiquitylation can occur independently of the KEN
box. As the MD3 domain of STIL interacted with and is stabi-
lized by USP9X, we investigated whether MD3 was K48 poly-
ubiquitylated. The MD3 domain was K48 polyubiquitylated
(Fig. 4 I), raising the possibility that USP9X may stabilize STIL
by interacting with and deubiquitylating its MD3 domain.
STILΔMD3 was K48 ubiquitylated, suggesting that APC/
C-mediated ubiquitylation of STIL does not depend upon the
MD3 domain (Fig. S4 T).

To test whether USP9X deubiquitylates the STIL MD3 do-
main, we incubated immunoprecipitated full-length STIL or the
MD3 domain of STIL with recombinant GST, GST-USP7, or GST
fused to the catalytic domain of USP9X (USP9XCD). The catalytic
domain of USP9X was specifically able to deubiquitylate both
full-length STIL and its MD3 domain (Figs. 4 J and S4 U), sug-
prising that USP9X stabilizes STIL by deubiquitylating its MD3
domain, and STIL stabilization is critical for centriole duplication.

Neurodevelopmental USP9X mutations destabilize STIL
A recent study identified loss-of-function mutations in USP9X in
women with MRXS99F (Reijnders et al., 2016). MRXS99F pa-
tients exhibit developmental delays, craniofacial abnormalities,
and congenital brain malformations (Reijnders et al., 2016). To
test whether the loss-of-function mutations associated with
MRXS99F affect USP9X localization, we examined the localiza-
tion of USP9X in MRXS99F patient fibroblasts. Interestingly,
there was a decrease in the centrosomal accumulation of USP9X
in all patient cell lines in comparison to an unaffected control
(Fig. 5 A and quantified in Fig. S5 A). In agreement with our
finding that USP9X is not required to localize SFI1 to the cen-
trosome, SFI1 localization was unaffected in MRXS99F fibro-
blasts (Fig. 5 B and quantified in Fig. S5 B). Consistent with our
finding that USP9X stabilizes STIL, MRXS99F-associated USP9X
mutations reduced centrosomal STIL (Fig. 5 C and quantified in
Fig. S5 C). Examination of centrosome organization in MRXS99F
fibroblasts revealed a decrease in CPI110 and an increase in
CEP135, consistent with the effects of decreased SFI1 or USP9X
function (Fig. S5, D–G).

Given the reduction in centrosomal STIL in MRXS99F fi-
broblasts, we assessed centriole numbers in patient-derived fi-
broblasts. All four MRXS99F fibroblast lines tested displayed
decreased numbers of centrioles (Fig. 5 D), raising the possibility
that MRXS99F-associated intellectual disabilities and brain
malformations arise from defects in centriole duplication during
neurogenesis.

Reduced USP9X expression confers selective growth advan-
tage to various tumors (Schwickart et al., 2010; Pérez-Mancera
et al., 2012; Peng et al., 2013; Kushwaha et al., 2015; Yang et al.,
2016; Zhang et al., 2016; Toloczko et al., 2017; Khan et al., 2018;
Li et al., 2018; Pal et al., 2018). Therefore, we investigated the
stability of additional USP9X targets in MRXS99F fibroblasts.
Immunoblot analysis revealed no consistent alteration of
CEP131, PCMI, MGL1, and ITCH levels in MRXS99F and control
fibroblasts (Fig. S5, H–I; Mouchantaf et al., 2006; Schwickart
et al., 2010; Li et al., 2017; Wang et al., 2017). These results
suggest that USP9X has cell type–specific targets or that there
are different consequences of germline inherited decreased
centrosomal localization of USP9X (MRXS99F) and acquired
dramatically reduced levels (cancer).

As USP9X also controls the stability of STIL and STIL promotes
centriole duplication, we examined whether the decreased
number of centrioles in MRXS99F fibroblasts was associated
with the destabilization of STIL. Whereas three of four
MRXS99F fibroblast lines exhibited modestly reduced levels of
USP9X (Fig. S5 M), all four displayed reduced STIL levels (Fig. 5
E and quantified in Fig. S5 N). In agreement with SFI1 functioning
upstream of USP9X, protein levels of SFI1 were unaltered (Fig. 5
E and quantified in Fig. S5 O). Similar to SFI1 depletion, CEP135
was unaltered in MRXS99F (Fig. S5, H and P–Q). We propose that
MRXS99F may be caused, at least in part, by decreased STIL
stability and concomitant defects in centriole duplication.

Thus, we have identified mammalian SFI1 as a centrosomal
protein that controls the localization of the deubiquitylase
USP9X to stabilize STIL, thereby positively regulating centriole
duplication (Fig. 5 F). Our finding that SFI1 regulates centriole
duplication, similar to the role of the S. cerevisiae SFI1 in spindle pole body duplication, provides evidence that the function of SFI1 in centrosome biogenesis is evolutionarily conserved. As loss-of-function mutations in STIL and USP9X cause human neurodevelopmental disorders (Kumar et al., 2009; Homan et al., 2014; Reijnders et al., 2016), and as USP9X deubiquitylates and stabilizes STIL, we propose that the USP9X-mediated control of STIL levels and centriole duplication is critical for human brain development.

**Discussion**

We found that mammalian SFI1 positively regulates centriole duplication, somewhat akin to the role of yeast SFI1 in promoting spindle pole body duplication. SFI1 localizes to the centrosome during S phase and recruits USP9X, a DUB. USP9X deubiquitylates and stabilizes STIL, a crucial factor for centriole duplication. Consistent with the role of USP9X in STIL stabilization, patient cells with loss-of-function mutations in USP9X have decreased STIL protein and attenuated centriole duplication. These data indicate that SFI1 recruits USP9X to stabilize STIL and that this module is critical for human brain development.

A previous study reported that overexpressed SFI1 colocalizes with Centrin at the distal centriole (Kilmartin, 2003). In contrast, our results using immunofluorescence staining of endogenous SFI1 and GFP-tagged SFI1 indicate that it accumulates as a cloud around the centrosome during S phase. As STIL also begins to associate with the centrosome during S phase, it is possible...
that the timing of SFI1 accumulation at the centrosome helps determine when STIL localizes to centrosomes and when centrioles duplicate.

We found that SFI1 stabilizes centrosomal STIL and supports centriole duplication. SFI1 controls STIL stability by interacting with and localizing the deubiquitylase USP9X to the centrosome. Similar to SFI1, USP9X stabilizes centrosomal STIL, suggesting that a principle way in which SFI1 controls STIL stability is through recruitment of USP9X to the centrosome. As USP9X interacts with and deubiquitylates the MD3 domain (aa 715–988) of STIL and removal of the MD3 domain destabilized STIL, USP9X controls STIL levels via the MD3 antidegron of STIL.

MD3 may cooperate with other domains of STIL to determine its temporal dynamics (Patwardhan et al., 2018). In particular, the MD3 domain is N terminal of the KEN box, itself ubiquitylated by APC/C to degrade STIL during mitosis (Arquint and Nigg, 2014). As SFI1 and USP9X stabilize STIL during interphase, ubiquitylation of STIL through the MD3 domain and KEN box may represent distinct mechanisms of controlling its levels during different parts of the cell cycle. The MD3 domain is C terminal of the recognition site through which β-TrCP binds STIL (Arquint et al., 2018). It will be interesting to determine whether the MD3 domain is ubiquitylated by β-TrCP and when during the cell cycle this might occur.

We found that depletion or pharmacological inhibition of USP9X led to the destabilization of STIL. Unlike knockdown of USP9X, WIPI130 disrupted the stability of additional centrosomal proteins such as SFI1 and CP110. This discrepancy in centrosomal protein stability may result from the ability of WIPI130 to inhibit USP5, USP14, and UCH37 in addition to USP9X. Recent studies have demonstrated that cancer cells are sensitized to WIPI130 in combination with chemotherapeutics (Liu et al., 2015; Fu et al., 2017; Ma et al., 2018). As various cancers have disrupted centrosome numbers, it would be interesting to determine whether the efficacy of WIPI130 treatment is due to its ability to alter centrosome biogenesis and cell proliferation.

Fibroblasts from MRXS99F patients with mutations in USP9X have reduced levels of STIL and defective centriole duplication. While the loss-of-function mutations do not dramatically reduce USP9X levels, they do compromise centrosomal localization, raising the possibility that the mislocalization prevents deubiquitylation and stabilization of STIL. As mutations in STIL also cause congenital brain malformations (Bond et al., 2005; Zhong et al., 2005; Kumar et al., 2009; Guernsey et al., 2010; Nicholas et al., 2010; Yu et al., 2010; Sir et al., 2011; Lin et al., 2013), the reduction in STIL in MRXS99F may underlie the etiology of brain malformations in MRXS99F. Consistent with this hypothesis, deletion of USP9X in the mouse brain causes premature differentiation of cortical progenitors, a phenotype associated with centrosome biogenesis defects (Jayaraman et al., 2016; Premarathne et al., 2017). We propose that the brain malformations in MRXS99F individuals result from centriole duplication defects due to the inability of USP9X to protect STIL from degradation.

Although USP9X has been shown to be required for the stabilization of PCM1, CEP131, ITCH, and MCL1 (Mouchantaf et al., 2006; Schwickart et al., 2010; Li et al., 2017; Wang et al., 2017), we did not detect decreased stability of these proteins in MRXS99F fibroblasts. One possibility is that MRXS99F-associated mutations may compromise deubiquitylation of only a subset of USP9X clients.

In summary, we have found that human SFI1 localizes to the centrosome during S phase and recruits USP9X, where they function to stabilize the procentriole factor STIL, culminating in centriole duplication. These findings provide mechanistic insights into how the timing of centriole duplication is regulated and how human mutations that compromise brain development disrupt centrosome biogenesis.

Materials and methods

Cell culture

HeLa and 293T/17 cells (University of California, San Francisco, tissue culture facility) were cultured in Advanced DMEM (Life Technologies) supplemented with 2% FBS (Life Technologies) and Glutamax-1 (Life Technologies). Antibiotic-antimycotic (Life Technologies) was added to media for siRNA screening studies. Flp-In TReX-DLD-1 cells (a gift from A. Holland and T. Moyer, Johns Hopkins University, Baltimore, MD) engineered to express GFP-Myc–tagged fragments of STIL under tetracycline-dependent control were cultured in DMEM (Life Technologies) supplemented with 10% FBS (Life Technologies) and Glutamax-1 (Life Technologies). HeLa cells were synchronized using a double thymidine block; in brief, cells were cultured in the presence of 2 mM thymidine (Sigma) for 16 h, washed twice with PBS, and incubated with medium devoid of thymidine for 8 h, followed by another 16 h block in thymidine. Cells were then collected at indicated time points following release into fresh culture medium. To induce STIL protein expression, Flp-In TReX-DLD-1 cells were treated with 10 µM tetracycline or doxycyclin (Sigma) for 24–48 h. To inhibit USP9X enzymatic activity, HeLa cells were incubated with 5 µM WIPI130 (Selleckchem) for 1–2 h. Female control and USP9X patient–derived fibroblasts (Table S3) were kindly provided by Drs. Nordgren and Kleefstra (Reijnders et al., 2016; Karolinska Institutet, Stockholm, Sweden, and Radboud University, Nijmegen, Netherlands). Fibroblast lines were grown in AmnioMax C-100 (Life Technologies).

Transfections and siRNA treatment

HeLa cells were transfected with siRNA (Life Technologies; Table S2) using Oligofectamine (Life Technologies) according to manufacturer’s recommendations and analyzed 36–48 h later. HeLa cells were transfected with plasmid using Lipofectamine3000 (Life Technologies) according to the manufacturer’s recommendations and analyzed 6 h later. The pcMV-HA-USP9X and pcMV-HA-USP9X (Cl666A) plasmid was purchased from the Medical Research Council at Dundee University. To test STIL fragment stability, Flp-In TReX-DLD-1 cells were induced with tetracycline for 24 h, before transfection with SC, SFI1, or USP9X siRNA, incubated in media containing tetracycline, and analyzed 48 h later.

Molecular biology

To generate pcMV-EGFP-SFI1, we PCR amplified fragments of SFI1 corresponding to UniProt A8K8P3-1 (RefSeq SFI1 isofor a...
from IMAGE clones 40124134 and 5177555 using Clontech HF polymerase and assembled the fragments using sequential InFusion reactions. pCMV-HA-USP9X and pCMV-HA-USP9X C1566A (DU 10171 and 10685) were purchased from the University of Dundee.

Centrosome isolation
Centrosomes were isolated from asynchronously growing HeLa cells as previously described (Bobinnec et al., 1998; Kodani et al., 2015). In brief, HeLa cells were treated with 2 µM nocodazole previously described (Kodani et al., 2013). In brief, cells were incubated on ice in chilled Dulbecco’s PBS Ca2+ and Mg2+ free (DPBS; Life Technologies), harvested with a cell scraper and lysed on ice in lysis buffer (1% Nonidet-P40, 50 mM Tris, pH 7.5, and 150 mM NaCl in DPBS) supplemented with protease and phosphatase inhibitors (Calbiochem). For each immunoprecipitation, 1 mg total lysate was incubated with 2 µg antibody for 2 h and then incubated with magnetic beads to precipitate Myc-GFP tagged proteins, 1 mg total lysate was incubated with 30 µl GFP-Trap magnetic beads (Chromotek) for 2 h. Immunocomplexes were washed three times in lysis buffer and subsequently boiled in 2× Laemmli reducing buffer (Bio-Rad) supplemented with β-mercaptoethanol (Bio-Rad). To test direct protein interactions, 500 µg total cell lysate from DLD1 cells was incubated with 1 µg GST-USP7 or GST-USP9X catalytic domain (aa 1,531–1,972; Abcam), or GST-USP9X catalytic domain (aa 1,531–1,972) for 1 h and then incubated with glutathione agarose beads (GE Healthcare Life Sciences) for an additional hour. To immunoprecipitate Mhc-GFP–tagged proteins, 1 mg total lysate was incubated with 30 µl GFP-Trap magnetic beads (Chromotek) for 2 h. Immunocomplexes were washed three times in lysis buffer and subsequently boiled in 2× Laemmli reducing buffer (Bio-Rad) supplemented with β-mercaptoethanol (Bio-Rad). To test direct protein interactions, 500 µg total cell lysate from DLD1 cells was incubated with 1 µg GST-USP7 or GST-USP9X catalytic domain (aa 1,531–1,972) for 1 h and then incubated with glutathione agarose beads (GE Healthcare Life Sciences) for an additional hour. Complexes were washed with lysis buffer and reduced in 2× sample buffer. Samples were run on 4–15% gradient TGX precast gels (Bio-Rad) and transferred onto BA85 supported Protran (GE Healthcare Life Sciences) using the Criterion plate electrode blotter (Bio-Rad). Blots were subjected to immunoblot analysis using ECL Lightening Plus (Perkin-Elmer) or Supersignal West Dura (Thermo Fisher Scientific). Samples were quantified and processed using Fiji or Photoshop (Adobe).

Electron microscopy
SC and SFI1-depleted HeLa cells were grown on coverslips (ACLR) and treated with thymidine overnight and released for 2 h to enrich for S phase cells. Cells were fixed in 2.5% glutaraldehyde and 0.03% picric acid diluted in 0.1 M cacodylate buffer, pH 7.4. Samples were postfixed in 1% OsO4 and 1.5% potassium ferrocyanide and contrasted using 1% uranyl acetate. Subsequently, cells were embedded in epon, ultrathin sectioned, stained with lead citrate, and imaged on a Tecnai G2 Spirit BioTWIN electron microscope.

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