Preclinical Evaluation of the Thrombogenicity and Endothelialization of Bare Metal and Surface-Coated Neurovascular Stents

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ABSTRACT

BACKGROUND AND PURPOSE: Stent-assisted coiling is routinely used for the endovascular treatment of complex or wide-neck intracranial aneurysms. However, in-stent thrombosis, thromboembolic events, and ischemic complications remain a major concern associated with stent implants. Therefore, a novel low-profile neurovascular stent with a bare metal surface was investigated with regard to thrombogenicity and endothelialization and compared with the same stent coated with albumin or heparin.

MATERIALS AND METHODS: The bare metal and heparin- or albumin-coated stents were loaded in heparin-coated tubing, which was then filled with heparinized human blood (n = 5) and circulated at 150 mL/min and 37°C for 60 minutes. Before and after circulation, measurement of various inflammation and coagulation markers and scanning electron microscopy were performed. Endothelialization of the stents was investigated in an in vitro model including human umbilical vascular endothelial cells.

RESULTS: Our results showed that platelet loss and platelet activation and activation of the coagulation cascade, which are induced by the bare metal stents, were significantly reduced by heparin or albumin coating. Adverse effects on erythrocytes, leukocytes, and the complement cascade were not induced by the bare metal or coated stents. Moreover, the bare metal and albumin-coated stents showed good endothelialization properties.

CONCLUSIONS: Albumin and heparin coatings clearly improve the thrombogenicity of the stents and thus may represent fundamental progress in the treatment of intracranial aneurysms. Moreover, preclinical evaluation of neurovascular stents under physiologic conditions supports and accelerates the development of more biocompatible neurovascular stents.

ABBREVIATIONS: HUVECs = human umbilical vascular endothelial cells; SEM = scanning electron microscopy; TAT = thrombin-antithrombin III

Endovascular treatment of complex or wide-neck intracranial aneurysms has been proved feasible by using the stent-assisted coiling technique, which was primarily described in 1997.1 In this procedure, self-expandable, nitinol-made stents, braided or laser cut, are delivered into the parent vessel by using microcatheters with an inner lumen between 0.0165 and 0.027 inches to retain coils within the aneurysm.1,2 Thus, the implanted stent prevents coil protrusion within the parent vessel, reduces coil compaction and hence recanalization, and provides a scaffold for endothelialization.2-5

However, the endoluminal presence of the implant can lead to activation of platelets and leukocytes following adverse effects like thromboembolic complications, hemorrhage, or proinflammatory reactions. Platelet activation is associated with the release of granules containing a plethora of molecules mainly promoting platelet activation and blood coagulation. Therefore, antiplatelet therapy is required, which can prevent intra-aneurysmal thrombosis and thus aneurysm occlusion but also increases the risk of hemorrhagic complications.6 Moreover and despite antiplatelet therapy, ischemic complications remain a major concern associated with stent implants. A retrospective review of stent-assisted coiling procedures in 624 patients between 2000 and 2011 revealed intra- and postprocedural in-stent thrombosis and thromboembolic events in 9.0% of the patients, whereas low or no response to antiplatelet agents and early interruption of therapy are possible causes for ischemic events.2

A retrospective study of 36 patients treated with stent-assisted...
coiling after aneurysm rupture showed that in 17%, stent-related thromboembolic complications occurred. Intra- and postproce-
dural clopidogrel administration was related to an incidence of
thrombotic events of 20% and 29%, respectively. Even pretreat-
ment with clopidogrel could not prevent thrombotic events,
which were shown in 6% of cases.

Therefore, preventing or strongly decreasing activation of
platelets and the coagulation cascade, while enhancing or at least
not impairing endothelialization, is a primary objective in the
treatment of endovascular diseases. Stent design, influence on
fluid dynamics, and particularly surface modifications are current
options to decrease activation of platelets and the coagulation
cascade, and they also address hemorrhagic stroke, because the
necessity for antiplatelet therapy can be reduced. Previous studies
show that surface coatings with the antithrombotic molecules
heparin and albumin, which is the most abundant protein in
blood, increase the biocompatibility of different biomaterials.

In our study, an in vitro model was established, which allows
investigation of neurovascular stents with regard to thrombo-
genicity and coagulation activation.

Small-diameter tubing and low flow rates simulating intracra-
nial vessel conditions were used to investigate novel low-profile
neurovascular stents with bare metal surfaces or 2 coatings, in-
cluding albumin and heparin, with regard to thrombogenicity.
Furthermore, a predisposition toward endothelialization was in-
vestedigated in an in vitro rotation model including human umbil-
ical vascular endothelial cells (HUVECs).

MATERIALS AND METHODS

For this study, the neurovascular stent Acclino (n = 24; Acand-
is, Pforzheim, Germany) was used. Acclino is a low-profile,
laser-cut stent with an electropolished surface and strut di-
mensions ranging between ~55 μm (wall thickness) and ~35
μm (strut width).

The stent is deliverable within the 1.9F microcatheter Neuro-
Slider 17 (Acandis) with a lumen of 0.0165 inches (~0.42 mm).
The small catheter lumen and the size of the stent (3.5 and 4.5 mm
in expanded configurations) allow the application in distal intra-
cranial vessels with diameters between 2 and 4 mm, whereas the
closed-cell structure allows retrieval and repositioning up to 90%
of its deployment.

For the current study, specific Acclino stent samples with a
wall thickness of ~75 μm were manufactured to reproduce a
worst-case scenario with regard to blood flow interruption caused
by the stent structure.

Blood Sampling

Blood-sampling procedures were approved by the ethics commit-
tee of the University of Tuebingen, Tuebingen, Germany. Blood
was collected from healthy volunteers (n = 5), who gave signed
informed consent, and was anticoagulated with 1.5 IU/mL Hep-
arin-Natrium (Rathiopharm, Ulm, Germany). Exclusion criteria
for the blood donors were the following: smokers; drug intake in
the last 2 weeks before blood sampling, especially hemostasis-
afflicting agents like acetylsalicylic acid; oral contraceptives; non-
steroidal antiphlogistics; and others, to guarantee optimal hemo-
static function.

FIG 1. Closed-loop in vitro circulation model for neurovascular stent
testing by using fresh human whole blood. Schematic overview (A)
and photograph (B) of the in vitro circulation model consisting of the
following: 1) heparin-coated tubing (inner diameter = 3.2 mm,
length = 75 cm), 2) connection tubing, 3) peristaltic pump (flow rate =
150 mL/min), 4) test device. C, Representative macroscopic images
of uncoated and albumin- and heparin-coated stents after circulation
for 60 minutes at 37°C (∼20 magnification).

Neurovascular Stent Coating

For albumin coating, 8 neurovascular stents were first incubated
for 2 hours at room temperature in 1% human serum albumin
(HSA; CSL Behring, Marburg, Germany), followed by a washing
step in saline and a 30-minute incubation in a suspension con-
taining 0.5-mg of 1-ethyl-3-(3-dimethylaminopropyl)carbo-
imid hydrochloride (Thermo Scientific, Karlsruhe, Germany)
for albumin cross-linking. Afterward, the stents were incubated
again in 1% HSA for 2 hours at room temperature followed by
another washing step in saline. After being coated, the stents were
air-dried and stored at 4°C until analysis in the in vitro thrombo-
genicity model. The heparin coating of the stents (n = 8) for the
thrombogenicity and endothelialization experiments was per-
formed according to standard methods. To guarantee sterile
conditions in the endothelialization model, we sterilized the
stents by using ethylene oxide.

In Vitro Thrombogenicity Model

This novel model was established to investigate the effect of neu-
rovascular stents on various cascade reactions of the human he-
mostatic system (coagulation cascade, cell alteration, comple-
ment system, and inflammation) during blood circulation (Fig
1A, B).

For each blood donor (n = 5), 4 polyvinyl chloride tubes (in-
ner diameter = 3.2 mm, length = 75 cm; Tygon; Saint-Gobain
Performance Plastics, Chamy, France) were coated with heparin
by Enson (Enson, Cape Coral, Florida) and left empty or loaded
with either 1 bare metal, 1 albumin-, or 1 heparin-coated stent.
Subsequently, each tubing was filled with a total of 6-mL heparin-
ized human blood, closed by silicone connection tubing, and cir-
culated at 150 mL/min in a water bath at 37°C for 60 minutes.
For each donor, 6 mL of heparinized blood served for the measure-
ment of baseline values before circulation. Before and after circu-
lation, the blood was directly transferred to the corresponding
terminating media and was further processed for enzyme-linked
immunosorbent assays and whole blood analysis.
The endothelialization experiments \( (n = 11005) \) were performed as previously described.12 Briefly, 2-mL bioreactor chambers were loaded with either a sterilized bare metal, an albumin-, or a heparin-coated stent. Isolation of HUVECs was adapted from a previously described method for the isolation of endothelial cells from residual saphenous vein biopsies.12 For each sample, \( 2.5 \times 10^6 \) cells were resuspended in 2.5-mL VascuLife EnGS medium (CellSystems, Troisdorf, Germany) containing a VascuLife EnGS LifeFactors Kit, 50-\( \mu \)g/mL gentamicin, and 0.05-\( \mu \)g/mL amphotericin B (PAA Laboratories, Co¨lbe, Germany).

Following the addition of the cell suspension, the bioreactor chambers were assembled in the rotation device and rotated at 37°C and 5% CO2. After 72 hours of incubation, the stents were washed in phosphate buffered saline and subsequently prepared for scanning electron microscopy analysis.

### Statistics

Data are depicted as means with standard error of the mean. Data were analyzed by using repeated-measures ANOVA with the Bonferroni multiple-comparison test to analyze differences among groups. All analyses were performed by using the statistical software package GraphPad Prism (Version 5; GraphPad Software, San Diego, California). Statistical significance was defined as \( P < .05 \).

### RESULTS

#### Hemocompatibility Testing of Neurovascular Stents

Contact of whole blood with artificial surfaces like stents, catheters, implants, and so forth is associated with various alterations of the hemostatic system. To evaluate the hemocompatibility of the bare metal \( (n = 5) \), albumin- \( (n = 5) \), or heparin-coated \( (n = 5) \) stents for neurovascular application, we developed an in vitro model to guarantee a defined and continuous blood flow mimicking blood circulation in the brain (Fig 1A, -B). Directly after

Cambridge, United Kingdom) and polymorphonuclear neutrophil elastase (Demeditec Diagnostics, Kiel-Wellsee, Germany) levels. As a marker for coagulation, we determined thrombin-antithrombin III (TAT) complexes (Siemens Healthcare, Marburg, Germany). Moreover, we analyzed SC5b-9 (TEComedical, B¨unde, Germany) to detect activation of the complement system.

#### Whole Blood Count Analysis

The number of erythrocytes, leukocytes, and platelets and hematocrit and hemoglobin values were measured by a cell counter system (ABX Micros 60; Axon Lab AG, Baden, Switzerland) in baseline samples and after circulation in all groups.

#### Scanning Electron Microscopy

Scanning electron microscopy (SEM) was performed to investigate the thrombogenicity and the endothelialization rate of the different stent surfaces after contact with human whole blood and HUVECs, respectively. After each experiment, the stents were rinsed in phosphate buffered saline and fixed in glutaraldehyde solution (2% in phosphate buffered saline, pH 7.4) overnight at 4°C. After another washing step with phosphate buffered saline, the stents were dehydrated by using ascending concentrations of ethanol (Emsure; Merck, Darmstadt, Germany) and critical point dried. After sputtering with gold palladium, we performed SEM analysis by using the EVO LS 10 scanning electron microscope (Carl Zeiss, Jena, Germany).

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**FIG 2.** Neurovascular stents do not change hematocrit levels, erythrocyte number, or hemoglobin concentration. Heparin-coated tubing was loaded with bare metal, albumin-, or a heparin-coated neurovascular stents and filled with 6 mL of fresh heparinized human blood. Tubing without a stent served as a negative control. Hematocrit level (A), erythrocyte numbers (B), and hemoglobin concentrations (C) were analyzed before and after 60 minutes of circulation at 150 mL/min. Data are given as means and standard error of the mean \( (n = 5) \) and were analyzed by using repeated-measures ANOVA with the Bonferroni multiple-comparison test. One asterisk indicates \( P < .05 \); two asterisks, \( P < .01 \); three asterisks, \( P < .001 \).

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**Endothelialization Model**

The endothelialization experiments \( (n = 3) \) were performed as previously described.12 Briefly, 2-mL bioreactor chambers were loaded with either a sterilized bare metal, an albumin-, or a heparin-coated stent. Isolation of HUVECs was adapted from a previously described method for the isolation of endothelial cells from residual saphenous vein biopsies.12 For each sample, \( 2.5 \times 10^6 \) cells were resuspended in 2.5-mL VascuLife EnGS medium (CellSystems, Troisdorf, Germany) containing a VascuLife EnGS LifeFactors Kit, 50-\( \mu \)g/mL gentamicin, and 0.05-\( \mu \)g/mL amphotericin B (PAA Laboratories, Co¨lbe, Germany).

Following the addition of the cell suspension, the bioreactor chambers were assembled in the rotation device and rotated at 37°C and 5% CO2. After 72 hours of incubation, the stents were washed in phosphate buffered saline and subsequently prepared for scanning electron microscopy analysis.

**Measurement of Hemostatic Markers**

The analysis of markers indicating activation of platelets and leukocytes and the coagulation cascade and complement system was performed in plasma by enzyme-linked immunosorbent assays, according to the manufacturer’s instructions. To investigate released, plasmatic markers indicating platelet and leukocyte activation, we analyzed PF-4 (Abcam, (Emsure; Merck, Darmstadt, Germany) and critical point dried. After sputtering with gold palladium, we performed SEM analysis by using the EVO LS 10 scanning electron microscope (Carl Zeiss, Jena, Germany).
circulation, macroscopic images, which are exemplarily shown in Fig 1C from 1 blood donor, were obtained of each stent.

Whole Blood Count Analysis. Before and after circulation in the in vitro model, the whole blood was analyzed in all groups. Circulation induced a significant decrease of hematocrit (P < .05, Fig 2A), red blood cell counts (P < .01, Fig 2B), and hemoglobin levels (P < .001, Fig 2C) in the control group containing no stent. No significant changes were observed in the bare metal and albumin- and heparin-coated stent groups compared with the control group.

Platelet and Leukocyte Activation. The concentration of PF-4, a small chemokine released from platelet α granules, is significantly increased during circulation in both control groups containing no stent and a bare metal stent, respectively (P < .05, Fig 3A). In the groups containing albumin- or heparin-coated stents, the release of PF-4 was significantly reduced (P < .05). Platelet activation was also estimated by the analysis of platelet counts. Compared with a platelet count of 217,200 ± 9,300 μL before circulation, a significant loss of platelets in the control group containing no stent (P < .001) was observed, which was further increased in the bare metal stent group (P < .001, Fig 3B). Again, the decrease in platelet numbers was less severe in the coated stent groups (P < .01).

Despite platelet activation, leukocyte activation based on granule release and leukocyte counts was investigated. The plasma concentration of polymorphonuclear neutrophil-elastase significantly increased because of circulation in the in vitro model (P < .01, Fig 3C). In the stent groups, no significant differences compared with the control group were detected. Similar results were obtained for the leukocyte counts (Fig 3D), in which no significant differences were detected among the groups after circulation.

Effects on the Coagulation Cascade and the Complement System. Activation of the coagulation cascade was estimated by the formation of the TAT complex indicating thrombin generation before and after circulation in all groups (Fig 4A). Compared with a TAT concentration of 41.1 ± 10.6 μg/L in the group containing no stent, a significant increase in TAT concentration was found in the bare metal stent group (518.3 ± 98.9 μg/L, P < .001). Stents coated with albumin or heparin induced a significantly lower activation of the coagulation cascade (P < .001), reaching values comparable with the group containing no stent.

The formation of Sc5b-9, the final complex of the complement...
system, significantly increased on circulation ($P < .001$, Fig 4B). No significant differences in Sc5b-9 generation were observed between the control and the stent-containing groups.

**Blood Cell Adhesion on Stents.** After circulation, each stent was analyzed by using scanning electron microscopy, whereby a clear difference in thrombogenicity was observed. SEM images of each stent were exemplarily shown from 1 donor at magnifications of $\times 500$, $\times 1000$, and $\times 2500$ (Fig 5). The bare metal stents showed a profound adhesion of blood cells, particularly platelets, and the formation of an attenuated fibrin network for all blood donors. In contrast, no or only a few platelets were detected on the surface of the albumin- and heparin-coated stents.

Overall, the albumin- and heparin-coated stents exhibited very little thrombogenicity compared with the bare metal stents, which also did not induce strong thrombogenic reactions (Table).

**Endothelialization Properties of Neurovascular Stents**

Next to hemocompatibility, the rate of endothelialization is another factor in characterizing the performance of a stent. Adhesion of HUVECs on the surface of the bare metal ($n = 3$), albumin- ($n = 3$), and heparin-coated ($n = 3$) stents was thus investigated after 72 hours of dynamic incubation in a specific model. SEM images showed that the bare metal stents and the albumin-coated stents displayed good endothelialization performance. Most interesting, the adhesion of HUVECs on the surface of the heparin-coated stents was decreased compared with the bare metal and albumin-coated stents (Fig 6 and Table).

**DISCUSSION**

The findings of our study indicate that surface coating of neurovascular stents with albumin or heparin significantly reduces platelet loss, platelet activation, and activation of the coagulation cascade. Other blood cell counts, activation of leukocytes, and the
complement system were not altered by the bare metal or coated stents.

To investigate the hemostatic performance of a low-profile neurovascular stent and different surface coatings, we established a dynamic in vitro model with focus on cerebral-like flow conditions and lumen size. A flow rate of 150 mL/min \(^{-1}\) was set according to blood flow in the middle cerebral artery, resulting from measurement in 92 patients. A tubing diameter of 3.2 mm was chosen, which is an appropriate size for distal intracranial vessels like the middle cerebral artery.

In our study, platelet counts and SEM analysis showed that a high number of platelets adhere to the surface of the bare metal stents. Platelet adhesion and aggregation were seen in particular at junctions between stent struts, thus indicating the role of flow dynamic phenomena like fluid stagnation or turbulence.

Previous studies investigating biomaterials showed that heparin coating is most frequently used to increase the hemocompatibility and decrease the thrombogenicity of various biomaterials. Furthermore, albumin coating of arterial prostheses decreased activation of the coagulation cascade and platelet and leukocyte adhesion.

In our study, both coatings showed excellent performance in preventing platelet adhesion, resulting in platelet counts similar to those in the control group without stents. Platelet activation investigated by measuring platelet granule release was also decreased by both coatings, indicating good antithrombotic surface properties.

The amount of TAT complex was also extremely reduced by the coatings; this finding shows the important role of surface material in the (intrinsic) coagulation cascade.

With regard to endothelialization, various studies have investigated the effect of heparin- or albumin-coated biomaterials in different in vitro and in vivo models.

In our in vitro study, endothelial cell adhesion was shown on the surface of the bare metal and albumin-coated stents, whereas the heparin coating seems not to promote cell adhesion. Compared with the in vivo situation, our endothelialization experiments were performed in an isolated system containing endothelial cells and cell culture media with 2% fetal calf serum. Under physiologic conditions, the adsorption of blood plasma proteins, such as albumin and fibrinogen, occurs immediately after implantation of the biomaterial. Hence, the low amount of plasma proteins in our cell culture experiments might have influenced the endothelialization performance of the different stents; thus, further in vivo investigations are warranted.

**CONCLUSIONS**

Although the application of low-profile stents has greatly expanded the possibilities in treating intracranial aneurysms, improving the performance of stents with regard to biocompatibility is still necessary and can be achieved by implant surface modifications. In this in vitro study, albumin and heparin coatings reduced the thrombogenicity of the stents in comparison with the bare metal stent. Therefore, a surface that decreases platelet and coagulation activation can represent crucial progress in the treatment of intracranial aneurysms.

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