Androgen receptor expression is required to ensure development of adult Leydig cells and to prevent development of steroidogenic cells with adrenal characteristics in the mouse testis

Peter J. O’Shaughnessy1*, Rod T. Mitchell2, Ana Monteiro1, Laura O’Hara2,3, Lyndsey Cruickshanks2, Hedi Claahsen-van der Grinten4, Pamela Brown2, Margaret Abel5 and Lee B. Smith2,6

Abstract

Background: The interstitium of the mouse testis contains Leydig cells and a small number of steroidogenic cells with adrenal characteristics which may be derived from the fetal adrenal during development or may be a normal subset of the developing fetal Leydig cells. Currently it is not known what regulates development and/or proliferation of this sub-population of steroidogenic cells in the mouse testis. Androgen receptors (AR) are essential for normal testicular function and in this study we have examined the role of the AR in regulating interstitial cell development.

Results: Using a mouse model which lacks gonadotropins and AR (hpg ARKO), stimulation of luteinising hormone receptors in vivo with human chorionic gonadotropin (hCG) caused a marked increase in adrenal cell transcripts/protein in a group of testicular interstitial cells. hCG also induced testicular transcripts associated with basic steroidogenic function in these mice but had no effect on adult Leydig cell-specific transcript levels. In hpg mice with functional AR, treatment with hCG induced Leydig cell-specific function and had no effect on adrenal transcript levels. Examination of mice with cell-specific AR deletion and knockdown of AR in a mouse Leydig cell line suggests that AR in the Leydig cells are likely to regulate these effects.

Conclusions: This study shows that in the mouse the androgen receptor is required both to prevent development of testicular cells with adrenal characteristics and to ensure development of an adult Leydig cell phenotype.

Keywords: Testis, Androgen receptor, Leydig, Adrenal, Development, Stereology

Background

The two major steroidogenic cell types in the male are the adrenocortical cell and the testicular Leydig cell. Both cell types share a basic steroidogenic cell phenotype typified by expression of proteins such as CYP11A1, STAR and HSD3B with added expression of cell-specific enzymes such as CYP11B1 (required for corticosterone production in adrenocortical cells) or HSD17B3 (required for testosterone production in adult Leydig cells). Embryonic stem cells, mesenchymal cells or fibroblasts can be induced towards a steroidogenic lineage [1–3] and the transformed cells will follow an adrenocortical or Leydig cell lineage or take on the phenotype of the more basic steroidogenic cell [1, 4, 5]. Leydig cells and adrenocortical cells are closely linked, therefore, and there is evidence that the two cell types originate from a single cell population [6]. While many of the factors involved in steroid cell differentiation are known [7], it is not clear what controls the final phenotype that the cells...
will adopt, although the local environment is likely to make a major contribution.

During fetal development the adrenals and testes form from a single adreno-gonadal primordium [6] which functionally divides in the mouse at about embryonic day (e) 11.5 [8]. Following adreno-gonadal separation the testes have been shown to contain a number of cells with adrenal characteristics and there is evidence that these may be derived from a cohort of adrenal cells that migrates to the developing gonad and remains there after adrenal/gonadal separation [9]. Shortly after the split of the adreno-gonadal primordium the fetal population of Leydig cells develops in the testicular interstitium and so it is also possible that some or all of the adrenal-like cells may be a normal sub-population of the testicular interstitium [9].

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Using a number of different models of androgen receptor (AR) ablation, previous studies have shown that AR are required for normal Leydig cell development in the adult [19–23]. In testicular feminised (Tfm) mice, which lack AR due to a mutation in the gene, and in ARKO mice adult Leydig cell number is reduced by up to 50% [20, 22] and the profile of steroidogenic enzyme expression is markedly altered [20]. Differences in enzyme expression may be complex as LH levels are increased in ARKO and Tfm mice [22, 24] but the results suggest that both LH and the AR may interact to ensure that there is normal proliferation and differentiation of testicular steroidogenic cells and that these cells adopt a specific Leydig cell phenotype.

To examine the role of LH and androgen in regulating development of interstitial steroidogenic cells (both Leydig cells and cells with adrenal characteristics) we have used the hypogonadal (hpg) mouse crossed with animals lacking ARs. Three models have been used; the hpg mouse which lacks circulating gonadotrophins [25] and is responsive to both LH and androgens, the hpg.SCARKO mouse which lacks AR specifically on the Sertoli cells and the hpg.ARKO mouse which lacks AR ubiquitously. The testicular phenotype of these animals has been described previously [26] and they differ from SCARKO, ARKO and Tfm models in that they lack gonadotrophins. This means that the Leydig cells in all animals will be largely inactive and under-developed but they will also be highly sensitive to the effects of exogenous hormone stimulation [27–29].

Results

hCG-induced Leydig cell hyperplasia in the hpg mouse is dependent on androgen receptors

Treatment with human chorionic gonadotropin (hCG; homologous protein to LH that acts on the LH-receptor) increased testicular volume (Table 1) and caused an 8 to10-fold increase in total Leydig cell number (Fig. 1) in both hpg and hpg.SCARKO mice compared to untreated animals. In hpg.ARKO mice, treatment with hCG had no effect on testicular volume and increased Leydig cell number by 3-fold, indicating a requirement for androgen action to achieve the full hyperplastic effect of hCG (Fig. 1). All data shown as mean ± sem in Fig. 1 (and subsequent figures) are also shown as primary raw data in Additional file 1.

Expression of transcript/proteins common to most steroidogenic cells is unaffected by the absence of androgen receptors

Expression of Hsd3b1 and Por transcripts was relatively high in untreated hpg, hpg.SCARKO and hpg.ARKO mice and showed little response to hormone treatment (Fig. 2a). This was also reflected in the clear immunohistochemical staining for HSD3B (Fig. 2b) seen in all groups with or without hCG treatment. The main effect of hCG was to increase the number of cells expressing HSD3B, consistent with the total increase in Leydig cell numbers induced by hCG, although there was some variation in staining intensity. In contrast to Hsd3b1, Cyp11a1 and Star transcript levels were very low in untreated mice but were clearly stimulated by hCG in all three groups (Fig. 2a).

Similarly, CYP11A1 was largely undetectable by immunohistochemistry in untreated mice but was clearly stimulated by hCG in all three groups (Fig. 2a).

Table 1 Effect of hCG on testicular volume

| genotype         | Testicular volume (mm³) | plus hCG |
|------------------|------------------------|---------|
| hpg              | 1.80 ± 0.16            | 6.12 ± 0.73* |
| hpg.SCARKO       | 1.70 ± 0.14            | 4.38 ± 0.12* |
| hpg.ARKO         | 0.83 ± 0.08            | 1.30 ± 0.06 |

*Significantly (P < 0.05) different to control by ANOVA and t-test
animals from any group but showed marked interstitial expression in all groups following hCG (Fig. 2c).

**hCG induces adult Leydig cell-specific transcripts only in the presence of androgen receptors**

Transcript levels of Cyp17a1, Lhr, Hsd17b3, Ins13, Sult1e1 and Hsd3b6, which are all Leydig cell specific [13, 30–32], were significantly increased by treatment with hCG in hpg and hpg.SCARKO mice. In contrast, there was little or no effect of hCG on transcript levels in hpg.ARKO mice (Fig. 3a). Immunohistochemical staining for CYP17A1 showed that the protein was detectable in untreated hpg, and hpg.ARKO mice (Fig. 3b) despite low transcript levels. Following hCG treatment there was a clear increase in the number of cells expressing CYP17A1 in the hpg mice. In hpg.ARKO mice there was also an increase in interstitial cells expressing CYP17A1 following hCG treatment, consistent with the increase in Leydig cell number, but a large number of unstained cells with a Leydig cell morphology were also present in the interstitium (Fig. 3b, red arrows). Interestingly, in the untreated hpg.ARKO mice there was clear immunostaining of the spermatogonia (Fig. 3b, black arrows). This staining was not seen in hpg mice (Fig. 3b) or in hpg.SCARKO mice (not shown) and was largely undetectable in the hpg.ARKO after hCG-treatment.

**hCG induces adrenal transcripts/protein in a number of interstitial cells in the absence of androgen receptors**

Expression of transcripts associated with adrenocortical steroidogenic cells (Mc2r, Cyp11b1, Cyp21a1, Akr1b7, Mrap and Hao2) showed marked stimulation in response to hCG in hpg.ARKO mice (Fig. 4a). In hpg and hpg.SCARKO mice, in contrast, hCG either had no effect on these transcripts or caused only a small increase in expression (Fig. 4a). Consistent with transcript data, CYP11B1 was undetectable by immunohistochemistry in control or hCG-treated hpg or hpg.SCARKO mice (not shown). In hpg.ARKO mice occasional interstitial cells stained for CYP11B1 but most were negative. Treatment with hCG, however, led to more intense staining although the numbers of steroidogenic cells showing expression remained low (Fig. 4b).

**AR expression in the Leydig cells inhibits development/differentiation of cells with adrenal characteristics in the testis**

Data above show that preventing differentiation of cells with adrenal characteristics in the testis depends on AR expression. To determine the cell types in which AR expression is essential for this effect we examined testicular adrenal transcript levels in different cell-specific models of AR deletion. Data in Fig. 5a shows that the Sertoli cell AR are unlikely to be the major factor responsible for the effects seen as there was no change in Mc2r in the SCARKO mouse, although there was an increase in Cyp11b1 transcript levels in the same animals. In contrast, there were more marked changes in both Mc2r and Cyp11b1 in mouse models lacking AR in the PTM cells (Fig. 5a). It is likely, however, that PTM.ARKO and PTM.SCARKO mice also lack AR in at least some of the Leydig cells as PTM cells are reported to act as Leydig cell precursors [23, 33]. To test this we examined Mc2r and Cyp11b1 transcript levels in Leydig cell-ARKO (LCARKO) mice (which are on a different background to the other mouse models [23, 34, 35]) (Fig. 5a). Results show that both Mc2r and Cyp11b1 are significantly increased in testes specifically lacking AR from Leydig cells, suggesting that the site of action of AR necessary for Leydig cell specification (as opposed to adrenal-like specification) is primarily the Leydig cells themselves.

To examine, directly, the effect of AR depletion in mouse Leydig cells we used RNAi to knockdown AR in a mouse Leydig tumor cell (MLTC-1) line. Exposure of MLTC-1 cells to AR miRNA significantly reduced Ar transcript levels with or without hCG treatment (Fig. 5b) (mean 67.5% knockdown). A reduction of Ar in these cells was associated with a significant reduction in transcript levels of the LC-specific androgen target gene Ins13 [36]. Suppression of AR also resulted in a significant increase in the adrenal enzyme Cyp11b1 (Fig. 5b).

**Discussion**

During fetal development the gonads and adrenals initially form adjacent to each other from the same
Fig. 2 hCG-induced expression of transcript/proteins common to most steroidogenic cells is unaffected by the absence of androgen receptors. Adult *hpg*, *hpg.SCARKO* and *hpg.ARKO* mice were treated daily with 4IU of HCG (or vehicle) for 1 week. A expression of *Hsd3b1*, *Por*, *Cyp11a1* and *Star* was measured by qPCR and is expressed relative to Leydig cell number in each group. The presence of an asterisk (*) indicates that the effect of hCG was significant (P < 0.05) for that mouse group relative to the respective control. B and C Immunohistochemical expression of HSD3B and CYP11A1 in testes from *hpg*, *hpg.SCARKO* and *hpg.ARKO* mice with or without hCG treatment. The bar represent 50 μm. Primary raw data are shown in Additional file 1.
primordium and there is evidence that the same SF1-positive cells give rise to adrenocortical cells and fetal Leydig cells [6, 37]. Subsequently, the adrenals and gonads separate, at about embryonic day 11 in the mouse, and some cells from the adrenal may become assimilated into the developing gonad [9] despite normal inhibition of the process by WNT4 [8]. Under this scenario, these cells would give rise to the adrenal-like cells seen in the adult testis. Cells with adrenal characteristics may also arise as a small subgroup of the developing Leydig cells [10, 11] and it is possible that there may be more than one such population of cells in the testicular interstitium. Whatever their origin, these adrenal-like cells in the mouse testis remain quiescent postnataally and it is only in humans that they have been shown to proliferate and to become steroidogenically active if stimulated by high ACTH [38]. Results described here now suggest the quiescent nature of these cells in mice is dependent upon testicular ARs which act to inhibit inappropriate, LH-dependent, postnatal development of cells with adrenal characteristics. Results also show that, in the absence of ARs, trophic stimulation of steroidogenic cells in the testicular interstitium leads to development of a basic steroidogenic cell type lacking enzymes specific for the adult Leydig cells.

Data from the hpg mouse, and other models of gonadotrophin or LH deficiency, have shown previously that LH is required for the establishment of a normal population of active Leydig cells. In the absence of LH or the LH-receptor Leydig cell number is markedly reduced [39–41] and Leydig cell activity is minimal [40, 41] while treatment with LH/hCG increases both cell number and activity [27, 42, 43]. Earlier studies have suggested that the adult hpg testis contains adult-type Leydig cells [44] and that the main effect of LH treatment is likely to be proliferation and maturation of these cells rather than induced stem cell differentiation [44]. This is consistent with other studies which have shown
increased [3H]thymidine labelling of Leydig cells in the hpg mouse following LH treatment [27]. In addition, it is clear from the pattern of HSD3B and CYP11A1 staining after hCG treatment, in the current study, that there is an overall increase in Leydig cell numbers throughout the interstitium. This contrasts with the marked increase in peritubular Leydig cells that occurs during Leydig cell stem cell differentiation following ethanedimethane sulphonate (EDS) treatment [15, 45, 46].

All steroidogenic cells express a base-line level of the proteins CYP11A1 and STAR which are required to form C21 steroids from cholesterol and POR is expressed if there is further metabolism by P450 enzymes in the endoplasmic reticulum. HSD3B is also generally expressed in steroidogenic cells although it is lacking in some cases (e.g. most human fetal adrenal cells [47, 48]). The different steroidogenic cell types are distinguished largely through the additional enzymes

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Fig. 4 hCG-treatment induces adrenal transcripts/protein in the absence of androgen receptors. Adult hpg, hpg SCARKO and hpg ARKO mice were treated daily with 4 IU of hCG (or vehicle) for 1 week. a Expression of Mc2r, Cyp11b1, Akr1b7, Cyp21a1, Hao2 and Mrap was measured by qPCR and is expressed relative to Leydig cell number in each group. The presence of an asterisk (*) indicates that the effect of hCG was significant (P < 0.05) for that mouse group relative to control. b Immunohistochemical expression of CYP11B1 in testes from hpg ARKO mice with or without treatment with hCG. Primary raw data are shown in Additional file 1.
Fig. 5 (See legend on next page.)
and hormone receptors that they express. In this study, the common steroidogenic transcripts were either expressed constitutively in the interstitial steroidogenic cells of the hpg or were induced by LH/hCG but, in either case, there was no influence from the presence or absence of ARs. This would suggest that LH/hCG acts to induce generic steroidogenic potential in the interstitial steroidogenic cells of the testis and that additional stimulation of the ARs is essential for the induction of a Leydig cell phenotype, consistent with previous data from androgen-resistant Tfm and LCARKO mice [20, 23]. Both gonadal and adrenal steroidogenic cells are dependent on SF-1 which regulates downstream expression of genes required for steroidogenesis [49]. It has been shown that forced expression of SF-1 in stem cells will transform those cells into steroidogenic cells [4]. The phenotype that the transformed cells adopt is variable and depends on a combination of stem cell type, SF-1 dosage and other transcriptional factors [2–4] but, in vivo, other factors are likely to determine the tissue-specific cell type. The results reported here now suggest that the AR is a required factor for generation of the adult Leydig cell phenotype in vivo. An alternative explanation would be that the hpg.ARKO mouse contains only fetal-type Leydig cells and LH/hCG induces proliferation and stimulates activity in these cells [50]. This would certainly be consistent with the failure of adult Leydig cell markers to increase after hCG. Recent studies have shown, however, that while there is a relative increase in fetal Leydig cells in the adult ARKO mouse, adult Leydig cells still outnumber fetal Leydig cells by a factor of about 4 to 1 [19]. Thus, lack of ARs does not appear to prevent at least initial differentiation and development of the adult Leydig cell population, consistent with previous results [23].

It appears likely that the cells showing induced adrenal transcript expression in the hpg.ARKO mice after hCG injection are the cells with adrenal characteristics cells described earlier [9]. This would be consistent with the relatively small number of cells in the hpg.ARKO testes that show an increase in adrenal enzymes in response to hCG treatment. Overall, the failure of hCG to induce adrenal transcript expression in hpg mice indicates that ARs normally inhibit trophic hormone stimulation of cells with adrenal characteristics in the testis. Interestingly, and consistent with this data, recent studies have shown that androgens act to inhibit recruitment and differentiation of progenitor cells into steroidogenic cells in the mouse adrenal [51]. Relevant data has also emerged from the study of Gata4 and Gata6 double knockout mice. In these animals there is loss of Leydig cells [52, 53] but survival and, perhaps, proliferation of cells with adrenal characteristics in the testis [52]. This indicates that there may be some synergism between the ARs and GATA factors in these interstitial cells, as seen in other tissues [54].

Androgen receptors are expressed in the Sertoli cells, the PTMC, vascular smooth muscle, endothelial cells and the Leydig cells in the normal post-natal mouse testis [26, 55] and a similar pattern of expression is seen in hpg mice [26]. Since the effects of hormone stimulation are similar in hpg and hpg.SCARKO mice, the results from this study show that androgen must mediate its effects on Leydig/adrenal cell development either through the cells themselves or through the PTMC, but not through the Sertoli cells. Previous studies using the Leydig cell specific LCARKO mouse have shown that it recapitulates many of the effects on Leydig cell transcript expression seen in the ARKO mouse [23] and we show here that there is increased adrenal transcript expression in LCARKO mice, and also when AR is specifically knocked down in vitro in a Leydig cell line. This suggests that the main effects of androgen on Leydig cell development are mediated through ARs in the Leydig cells. As the origin of testicular cells with adrenal characteristics is uncertain, however, it is not clear whether ARs expressed in these cells or the Leydig cells regulates their development in the testis. Nevertheless, data from AR knockout in MLTC-1 cells suggests that there may be a requirement for continued AR-mediated suppression of plasticity, since a reduction of AR in MLTC-1 cells is able to switch ‘mature’ Leydig cells towards an adrenal cell fate.

Recent work has suggested that Hsd3b1 in the mouse testis is only expressed in fetal Leydig cells [56] which would suggest that the adult hpg mouse testis contains a
significant number of fetal Leydig cells. This is consistent with the “independent” nature of these cells which do not require LH stimulation during development [57, 58] or input from the Sertoli cells [59]. Expression of Hsd3b1 was also relatively unaffected by the presence or absence of ARs indicating that ARs are not important for fetal Leydig cell development, as previously suggested [19].

These studies underline the importance that the ARs play in regulating development and function of cells in the testicular interstitium. This data, and previous studies [44], show that the interstitial tissue of the adult hpg mouse testis contains adult Leydig cells and cells with adrenal characteristics and that hCG/LH treatment stimulates Leydig cell proliferation/activity while having little effect on the other cells. The hpg.ARKO mouse also contains adult Leydig cells and cells with adrenal characteristics but appears to have an additional population of “basal” steroidogenic cells. In these animals, hCG treatment causes proliferation of the adult Leydig cells but has little effect on adult Leydig cell-specific transcripts (Fig. 3). hCG-treatment also causes the basal steroidogenic cells to undergo proliferation and stimulation (Figs. 2 and 3) and leads to activation of adrenal-like cells (Fig. 4).

Conclusions
These studies have dissected the roles that LH (hCG) and the AR play in the development of interstitial cells in the mouse testis. As shown before, LH/hCG stimulates proliferation and maturation of Leydig cells but we show here that this process requires the presence of AR in the Leydig cells. These AR are also required to prevent activation of cells with adrenal characteristics under LH/hCG stimulation and act, therefore, to induce the formation of normal interstitial cell populations in the adult mouse testis.

Methods
Animals and treatments
ARKO, SCARKO, hpg.ARKO and hpg.SCARKO mice were generated at the University of Oxford as published previously [16] from mice obtained originally from Prof G Verhoeven and Dr. K De Gendt (Catholic University of Leuven, Belgium) and from mouse colonies bred at the University of Oxford [16]. SCARKO and ARKO mice were produced by crossing female mice carrying an Ar allele with a floxed exon 2 (Ar<sup>fl</sup>) with male mice expressing Cre under the regulation of the Amh or the ubiquitous promoter Pkg-1 [60–62]. LC-ARKO mice were generated at the University of Edinburgh by mating Ar<sup>fl</sup> female mice (obtained as above) with male mice expressing Cre under the regulation of the Fabp4 promoter (provided by Kerry McInnes, University of Edinburgh) [23] while PTM-ARKO were generated as previously described [34] by mating Ar<sup>fl</sup> female mice with male mice expressing Cre under the regulation of the Myh11 promoter (originally from Michael Kotlikoff (Cornell University, USA)) [35]. Mice were killed, as necessary, by exposure to a rising concentration of CO₂.

Adult mice (10 weeks) were injected sub-cutaneously with 4 IU recombinant hCG (Serono Ltd. Middlesex, UK) in 0.2 ml PBS (phosphate buffered saline, pH 7.4, Sigma Aldrich, Dorset, UK) once daily for 7 days. hCG acts as an LH analog and stimulates Leydig cell function through the LH-receptor. Mice were killed, as above, on day 8 (24 h after the last injection) and testes snap frozen in liquid N₂ or fixed in Bouin’s overnight.

Steroeology
For stereoeological analysis, testes were embedded in Technovit 7100 resin, cut into sections (20 μm), and stained with Harris’ hematoxylin. The total testis volume was estimated using the Cavalieri principle [63]. The optical dissector technique [64] was used to count the number of Leydig cells in each testis. Leydig cells were recognised by their position, round nucleus and relatively abundant cytoplasm [39, 65]. The numerical density of each cell type was estimated using an Olympus BX50 microscope fitted with a motorized stage (Prior Scientific Instruments, Cambridge, UK) and Stereologer software (Systems Planning Analysis, Alexandria, VA, USA). Stereological data for untreated hpg, hpg.SCARKO and hpg.ARKO animals in this study includes data from a previous study [62] plus additional animals. Testicular volume data (Table 1) also includes data reported previously [26, 62] with the inclusion of additional samples.

Immunohistochemistry
For immunohistochemistry, testes were embedded in paraffin and sections (5 μm) were mounted on glass slides, dewaxed, and rehydrated. Sections were incubated with anti-CYP11A1 (rabbit polyclonal, donated by Dr. AH Payne, Stanford University), anti-HSD3B2 (rabbit polyclonal, donated by Prof Ian Mason, University of Edinburgh), anti-CYP11B1 (mouse polyclonal, donated by Prof Celso E. Gomez-Sanchez, University of Mississippi) after heat-induced antigen retrieval. Bound primary antibody was detected using a peroxidase-conjugated secondary antibody, followed by fluorescyl-tyramide amplification step with visualization using 3,3-diaminobenzidine tetrahydrochloride (Dako UK Ltd., Cambridgeshire, UK). To reduce “mouse-on-mouse” effects when using an anti-mouse second antibody blocking reagent (M.O.M. Vector Laboratories Ltd., Peterborough,
(100 nM) and template in a total volume of 10 μl. Reactions contained 5 μM L-Glutamine, 0.5 mM Sodium pyruvate, 1.2 g/L Sodium bicarbonate and 15 mM HEPES), 10% fetal bovine serum and 1% penicillin/streptomycin. Cells were transfected with either an AR miRNA lentivirus consisting of a CMV promoter, an mKate2 fluorescent reporter and 2 chained miRNAs that target the androgen receptor (“Lc-cppt-CMV-mKate2-ARmiR2&4”), a ‘scrambled’ lentivirus (“Lv-cppt-CMV-mKate2-negmiRx2”) or a control with no virus. Viruses were produced by Dr. Pamela Brown at the SURF University of Edinburgh Biomolecular Core Facility. Transfection was performed at an MOI of 5 with medium supplemented with a final concentration of 6 μg/mL Polybrene (Sigma). Cells were then sorted by Fluorescence Activated Cell Sorting for mKate2 expression to increase the purity of the transfection cell population before being selected for blastocystin resistance. To treat cells with hCG, cells growing in complete medium were allowed to reach confluence, then replaced with serum-free medium and incubated for four hours. The serum-free medium was then removed and replaced with either serum-free medium supplemented with 1.5 IU hCG final concentration for “+hCG” groups or serum-free medium only for “-hCG” groups. The cells were then incubated for 16 h before being collected and processed for RNA extraction. Each condition was repeated four times with two or three flasks each time used as technical replicates.

Measurement of mRNA levels
For data shown in Figs. 1, 2, 3 and 4, testis RNA was extracted using TRIzol (Life Technologies, Paisley, UK) and the RNA was reverse transcribed using random hexamer-primed cDNA was prepared using the VILO cDNA synthesis kit (Invitrogen, ThermoFisher Scientific, Paisley, UK) according to manufacturer’s instructions. The primers used for qPCR were designed using the online Roche Universal Probe Library (UPL) software (Roche Diagnostics Ltd., Burgess Hill, UK) and they are listed in Additional file 2. The qPCRs were carried out using an ABI Prism 7900 Sequence Detection System (ThermoFisher Scientific). Transcript levels per testis were normalised to luciferase using the ΔΔCt method [70] as above. Transcript levels in MLTC1 cells were normalised to an internal housekeeping gene assay for Actb (Roche).

Statistical analysis
Data were analysed using t-tests, analysis of variance (ANOVA) (with differences between individual groups identified by Fisher’s post-hoc test or by t-test) or the Kruskal-Wallis test as appropriate. Where necessary, data were transformed using the Box-Cox method [71] to avoid heterogeneity of variance. Statistical analysis was carried out using Minitab v15 (Minitab Ltd., Coventry, UK).

Additional files

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Availability of data and materials
All data generated or analysed during this study are included in this published article and its additional files.
The authors declare that they have no competing interests.

Not applicable.

Consent for publication of Edinburgh (Animal Welfare Ethical Review Body).

Ethics approval

PJOS, MHA and LBS conceived and designed the study. AM, LOH, LC, HCG, EH16 4TJ, UK. 3Centre for Discovery Brain Sciences, University of Edinburgh, 'The Queen

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