Antioxidant and Antidiabetic Activities of Mempening (Lithocarpus bancanus) Leaves

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ABSTRACT

Background: Lithocarpus bancanus or commonly called as mempening in Talang Mamak Tribe, Indonesia is a plant that is used as a traditional medicine. Objective: This study aim to evaluated antioxidant and antidiabetic activities of L. bancanus leaves extract. Material and Methods: The methanol extract was obtained by maceration of the leaves. The n-hexane, dichloromethane and ethyl acetate fractions were prepared by successive partition process of the methanol extract. Antioxidant activities were evaluated by various antioxidant assays, including DPPH (1,1-diphenyl-2-picrylhydrazyl), FRAP (ferric reducing antioxidant capacity), and ABTS (2,2’-azinobis 3-ethylbenzothiazoline-6-sulfonic acid) method. Total phenolics were estimated based on the Folin–Ciocalteu method, while, aluminum chloride methods were employed to estimate total flavonoids. Antidiabetic activities was determined by inhibiting the activity of α-glucosidase method. Results: antioxidant activity assay against DPPH radical as well as the total phenolic and flavonoid content of L. bancanus leaves showed that the methanol extract possessed IC50 value of 39.469 ± 0.273 µg/mL with total phenol and flavonoid were 11.426 ± 0.432 mg GAE/g and 15.423 ± 0.213 mg QE/g respectively. The FRAP, CUPRAC and ABTS values of methanol extract were 3494.302 ± 0.456, 26665.501 ± 5.940 and 2857977 ± 0.715 µM TE/g dry weight sample respectively. Antidiabetic activity of methanol extract with IC50 value of 30.565 ± 0.331 µg/mL. Conclusion: It could be concluded that leaves of L. bancanus have antioxidant and antidiabetic properties.

Key words: Antioxidant; Antidiabetic; Lithocarpus bancanus; Talang Mamak Tribe.

INTRODUCTION

Diabetes mellitus is the common serious metabolic disorder due to disturbance of carbohydrate, lipid and protein metabolism. It is characterized by hyperglycaemia resulting from insulin resistance or decreased production of insulin by the β-cells of the pancreas. Approximately 90% of all cases of diabetes in developed and developing countries are type-2 diabetes.1,2 Hyperglycaemia is found to increase approximately 90% of all cases of diabetes in developed and developing countries are type-2 diabetes.1,2 Approximately 90% of all cases of diabetes in developed and developing countries are type-2 diabetes.1,2 Hyperglycaemia is found to increase the production of free radicals that are associated with long-term damage, dysfunction, and failure of various organs, especially eyes, kidneys, nerves, hearts, and blood vessels. Several other factors such as hyperlipidaemia and enhanced oxidative stress play a major role in diabetes. The development of diabetes and progression of complications are usually associated with oxidative stress which is as a result of overexpression of reactive oxygen species (ROS) or free radicals. Free radicals are generated during antioxidation of glucose in diabetes mellitus resulting in oxidative stress. ROS is involved in the process of signal transduction in the pancreatic β-cells and has the potential to regulate glucose-stimulated insulin secretion. However, insulin secretion can reduce when excessive ROS synthesis is produced by elevated glucose or fatty acid oxidation.3,4

Traditional medicine is gaining so much interest recently due to their multiple modes of actions with minimal adverse effects in humans. Medicinal plant are rich source of secondary metabolites used in various therapies, including diabetes mellitus. Thus, considering the high Indonesia biodiversity, it is essential to explore potential plant species, including L. bancanus (mempening). This species belongs to Fagaceae family found in the Talang Mamak tribe in Kelayang District, Indragiri Hulu Regency, Riau Province. It is usually used by the peoples as a medicine to treat pain and inflammation. Some species of this genus have been previously reported to contain various secondary metabolites, including terpenoids, steroids and flavonoids as the major components. Likewise, bioactivities from the genus have been evaluated including antioxidants, antidiabetic, anticancer, antimicrobial and other activities.5-8 In regard to explore antidiabetic agent from Talang Mamak medicinal plants, we reported the antioxidant and antidiabetic activity of L. bancanus leaves extract and fractions.

MATERIAL AND METHOD

Chemical reagents

DPPH (1,1-diphenyl-2–picrylhydrazyl), gallic acid, quercetin, ascorbic acid, Trolox®, TPTZ (2, 4, 6-tripyridyl-s-triazine), neocuprine (Nc), α-glucosidase enzyme and p-nitrophenyl-α-D-glucopyranoside (p-NPG) substrate from Sigma-Aldrich Chemical Co (Singapore), Folin-Ciocalteu,
Na₂CO₃, NaNO₂, AlCl₃, NaOH, CuCl₂, 2H₂O 10 mM, neocuproine (Nc), K₂S₂O₈, organic solvents p.a. (n-hexane, dichloromethane, ethyl acetate and methanol from Merck (Germany).

Collection of plant material

Samples were collected from Kelayang (Bukit Tiga Puluh National Park (TNBT) of Indragiri Hulu Regency, Riau Province and identification of sample plants was carried out in the Department of Biology, Faculty of Mathematics and Natural Sciences, University of Riau. Samples were dried and finely ground and stored at 4°C until analysis.

Extraction

Dried L. bancanus leaves (100 g) were ground into powder and then macerated for 48 hour followed by ultrasound for 1 hour and the macerates were collected and concentrated with a rotary evaporator at 50°C. Methanol extract were fractionated with dichloromethane and ethyl acetate respectively. Each fraction were evaporated to get extracts.

DPPH radical scavenging activity assay

Antioxidant activity assay was carried out by using DPPH method (1,1-diphenyl-2-picrylhydrazyl) by the standard method, with a slight modification. Samples with a finally concentration of 1000 µg/mL were diluted by two fold dilution method (1000 - 31.25 µg/mL) in 96 well clear polystyrene microplate. A total of 50 µL of sample was added with 80 µL of DPPH 100 µg/mL then incubated for 30 minutes in a dark place. Absorbance were measured by microplate reader (Berthold, Germany) at 520 nm. The same method were conducted for ascorbic acid and quercetin as positive control.

The % Inhibition value is calculated by the following formula:

\[
\% \text{ Inhibition} = \frac{(A_{s} - A_{t})}{A_{s}} \times 100
\]

Where \( A_s \) represents the absorbance of the DPPH radical solution without sample while \( A_t \) represents the absorbance of the sample with DPPH radical solution. A graph of inhibition percentages (1%) versus concentrations of the sample was plotted to provide value of \( IC_{50} \).

Determination of total phenolic content (TPC)

Determination of the total phenolic sample was carried out by using the Folin-Ciocalteu method. Gallic acid was used as a standard. A total of 100 µL of sample, gallic acid and blank were each mixed with 50 µL of the Folin-Ciocalteu reagent 0.25 N in 96-well microplate. After 5 minutes, 100 µL Na₂CO₃ 7.5% (w/v) was added. The mixture was incubated for 30 minutes in a dark place at room temperature before absorbance was measured at a wavelength of 765 nm by microplate reader. The total phenolic content is expressed as milligrams of equivalent gallic acid per gram dry matter of sample (mgGAE/g) through the calibration curve gallic acid. Linearity range of calibration curve was 10 - 50 µg/mL. (\( y = 0.016x + 0.0081, r = 0.992 \)).

Determination of total flavonoids content (TFC)

Determination of the total flavonoid content of extracts was carried out by using the colorimetric method of aluminum chloride with quercetin as a standard. A total of 50 µL samples, quercetin and blanks were each mixed with 10 µL NaNO₂ 5% (w/v), 10 µL AlCl₃ 10% (w/v) in 96 well microplates. After 5 minutes 100 µL of 1 M NaOH was added. The mixture was added with 30 mL of distilled water and the mixture was incubated in a dark place at room temperature for 30 minutes. Absorbance of the mixture was measured at a wavelength of 510 nm by microplate reader. The total content of flavonoids is expressed as milligrams of equivalent quercetin per gram dry matter of sample (mgQE/g). Linearity range of calibration curve was 10 - 50 µg/mL. (\( y = 0.0162x + 0.0755, r = 0.999 \)).

Ferric reducing antioxidant power assay (FRAP)

Antioxidant activity was measured with FRAP according to the method with Trolox as standard. FRAP reagents was made from 0.2 M acetae buffer solution (pH 3.6), TPTZ solution (2, 4, 6-tripydyl-s-triazine) 10 mM in 40 mM HCl and 20 mM FeCl₃·6H₂O solution were prepared and then the solution was mixed with ratio 10: 1: 1. Some 100 µL of sample was added to 96-well clear polystyrene microplates which contained 100 µL of FRAP reagent. The mixture was incubated for 30 minutes in a dark place at room temperature. The absorbance of sample was measured at wavelength of 595 nm by microplate reader and calculated as micromol of Trolox equivalent per gram of dry weight (mg TE/g dry weight) and using the Trolox as standard curve. Linearity range of calibration curve was 2 - 10 µM/mL (\( y = 0.0641x + 0.0644, r = 0.991 \)).

Cupric reducing antioxidant capacity assay (CUPRAC)

Antioxidant activity was measured using the CUPRAC analysis. Some 50 µL of CuCl₂·2H₂O 10 mM, 50 µL Neocuproine (Nc) 7.5 mM and 50 µL ammonium acetate buffer were added to 96 well clear polystyrene microplates which contained 100 µL of samples, standard and blank. Then, distilled water was added up to 300 µL. The mixture was incubated for 30 minutes in a dark place at room temperature and the absorbance was read at 450 nm by microplate reader and calculated as micromol of Trolox equivalent per gram of dry weight (mg TE/g dry weight) and using Trolox as standard curve. Linearity range of calibration curve was 20 - 100 µM/mL (\( y = 0.0073x + 0.1274, r = 0.999 \)).

2,2'-azobis 3-ethylbenzothiazoline-6-sulfonic acid assay (ABTS)

Antioxidant activity was carried out with the ABTS method. ABTS reagents was prepared by dissolving 0.077 g ABTS powder in 10 ml of distilled water. 10 ml of ABTS solution was reacted with 10 ml of K₂S₂O₈ (5 mM) and was saved in a dark place at room temperature for 16 hours to produce ABTS radical cation. The solution was diluted with distilled water to obtain an absorbance of 1.00 at a wavelength of 734 nm. Some 100 µL of ABTS solution was added to 96 well clear polystyrene microplates which contained 200 µL of sample, standard, and blank. The mixture was incubated for 30 minutes in a dark place at room temperature. The absorbance of the sample was measured at a wavelength 734 nm by microplate reader (Berthold, Germany). and the results were calculated as micromol of Trolox equivalent per gram of dry weight (mg TE/g dry weight) using the Trolox as standard curve. Linearity range of calibration curve was 2 - 10 µM/mL (\( y = 0.0738x + 0.0473, r = 0.999 \)).

Antidiabetic activity assay

The antidiabetic test used a method of inhibiting the activity of α-glucosidase enzyme with p-NPG as a substrate by the standard method, with a slight modification. The sample was diluted by the two fold dilution method of concentration 1000 - 31.25 µg/mL. A total of 10 µL of DMSO (B₀) and 10 µL of sample (S₁) were added with 50 µL of pH 7 phosphate buffer, 25 µL of p-NPG 20 mM and 10 µL of DMSO (B₁) and 10 µL of sample (S₀) were added with 50 µL of phosphate buffer pH 7.25 µL p-NPG 20 mM and 25 µL α-glucosidase 0.2 U/mL were mixed in 96-well microplate and incubated for 30 minutes at 37°C. The reaction was stopped by adding 100 µL of 0.1 M Na₂CO₃ then absorbance was measured by microplate reader at a wavelength of 405 nm.

The % Inhibition value is calculated by the following formula:

\[
\% \text{ Inhibition} = \frac{(B₁ - B₀) - (S₁ - S₀)}{(B₁ - B₀)} \times 100
\]
Where \( B_s \) represents the absorbance without sample and enzyme, \( B_r \) represents the absorbance without sample and contain enzyme, \( S_r \) represents the absorbance contain sample and without enzyme while \( S_s \) represents the absorbance contain sample and enzyme. A graph of inhibition percentages (\%) against concentrations of the sample was plotted to provide value of IC\(_{50}\).

**Statistical analysis**

All assays were carried out in triplicate and their results were expressed as mean ± standard deviation. Data analysed by one - way ANOVA by using IBM SPSS statistics 20 (Version 20.0, IBM. Corp., U.S.A). The significance of difference was calculated by using Duncan’s multiple range test, while Pearson correlation test was conducted to determine the correlation among variable. A \( P < 0.05 \) were considered statistically significant levels. All measurements were carried triplicate.

**RESULT AND DISCUSSION**

**DPPH radical scavenging activity**

DPPH radical scavenging activity from \( L. \) bancanus leaves exhibited various activity (Table 1). Methanol extract and ethyl acetate fraction showed high antioxidant activity with IC\(_{50}\) value of 39.469 ± 0.273 \( \mu g/mL \) and 52.546 ± 0.557 respectively with no significantly different (\( P<0.05 \)) with quercetin. The \( n \)-hexane fraction showed no activity with IC\(_{50}\) values greater than 1000 \( \mu g/mL \) while the dichloromethane fraction exhibited moderate antioxidant activity with IC\(_{50}\) values of 334.464 ± 0.361 \( \mu g/mL \). In this result showed that the solvent with high polarity exhibited high activity, and this might be due to the presence of flavonoids and phenolics.

**Total phenolic (TPC) and flavonoid content (TFC)**

The determination of TPC is based on the reduction of the phosphomolybdate-tungstate complex with its active center is Mo (VI) by phenolic compounds forming a blue product. Total phenolic content of extract and fraction of \( L. \) bancanus leave were differed significantly (\( P<0.05 \)) (Table 2). Methanol extract and ethyl acetate fraction showed high TPC compared to \( n \)-hexane and dichloromethane fractions with value of 11.426 ± 0.106 and 6.525 ± 0.188 mg GAE/g dry sample, respectively. In order to determined TFC, the samples fractions with value of 11.426 ± 0.106 and 6.525 ± 0.188 mg GAE/g dry -hexane and dichloromethane fraction showed high TPC compared to

| Sample                | TPC (mg GAE/g dry sample) | TFC (mg QE/g dry sample) |
|-----------------------|---------------------------|--------------------------|
| \( n \)-Hexane fraction | 0.472 ± 0.241\(^a\)        | 0.463 ± 0.110\(^a\)      |
| Dichloromethane fraction | 0.788 ± 0.125\(^a\)       | 0.925 ± 0.216\(^a\)      |
| Ethyl acetate fraction | 6.525 ± 0.188\(^a\)       | 9.144 ± 0.138\(^a\)      |
| Methanol extract      | 11.426 ±0.106\(^a\)       | 15.422 ± 0.306\(^a\)     |

Note: Data expressed as mean ± standard deviation (\( n = 3 \)). Same letters in each column mean no significant difference (\( P < 0.05 \)).

**Ferric reducing antioxidant power (FRAP)**

The FRAP method was used to measure the ability of antioxidants by reducing ferric in acidic conditions. Complex ferric-tripryridyltriazine (Fe \(^{+3}\)-TPTZ) is reduced to form Fe\(^{+2}\) (Fe\(^{+2}\)-TPTZ) with maximum absorbance at 595 nm. The results showed significantly different among the tested sampel (\( P<0.05 \)), however, the methanol extract and quercetin exhibited no significantly different (Table 3).

**Cupric reducing antioxidant capacity (CUPRAC)**

In this assay, Cu (II) was reduced to Cu (I) by antioxidants. Neocuproin (Nc) chromophore reagent reacts with CuCl\(_2\) to form complex Cu (I) –Nc at pH 7 at a wavelength of 450 nm, from bright blue to yellow-orange. The results showed that extract and fractions differed significantly (\( P<0.05 \)) (Table 3). The methanol extract and ethyl acetate fraction possessed activity with values of 15146.556 ± 5.940 and 15146.556 ± 3.107 \( \mu M \) TE/dry weight, respectively, and these results exhibited high activity compared to \( n \)-hexane and dichloromethane fractions.

Flavonoids are diphenyl propanoids consisting of two of rings connected by chains with three of carbon atoms. The plants extracts contained phenolic and flavonoid compounds which showed effective antioxidant properties and could lower cellular oxidative stress. In this study, we reported correlation between the phenolic and flavonoid content with DPPH radical scavenging with coefficient correlation (\( r \)) = 0.996 and 0.994, respectively (Table 3) and it is in an agreement with Jacobo-Velazquez and coworkers.

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**2,2’-azobisisobutyryl 2,2’-azonobis 3-ethylbenzothiazoline-6-sulfonic acid (ABTS)**

The ABTS \((2,2’-azobisisobutyryl 2,2’-azonobis 3-ethylbenzothiazoline-6-sulfonic acid)\) assay is based on the ability of antioxidants to capture the cation radical of ABTS. In this assay, the radical cation of ABTS is produced from ABTS oxidation by potassium persulfate (K\(_2\)S\(_2\)O\(_8\)) which produces a greenish blue color. Color loss will occur when antioxidant compounds donate H atoms to the ABTS cation radical. The antioxidant activity of a sample in reducing ABTS cation radical compared to Trolox, and showed that methanol extract and ethyl acetate fraction exhibited high TFC with value of 15.422 ± 0.306 and 9.144 ± 0.138 mg QE/dry weight sample, respectively (\( p<0.05 \)).

**Table 1. Antioxidant activity of \( L. \) bancanus leaves against DPPH radical.**

| Sample                | IC\(_{50}\) (\( \mu g/mL \)) |
|-----------------------|------------------------------|
| \( n \)-Hexane fraction | 1151.808 ± 17.458\(^a\)       |
| Dichloromethane fraction | 334.464 ± 0.361\(^a\)       |
| Ethyl acetate fraction | 52.546 ± 0.557\(^a\)        |
| Methanol extract      | 39.469 ± 0.273\(^a\)        |
| Quercetin             | 40.063 ± 1.604\(^a\)        |
| Ascorbic Acid         | 11.043 ± 0.154\(^a\)        |

Note: Data expressed as mean ± standard deviation (\( n = 3 \)). Same letters in each column mean no significant difference (\( P < 0.05 \)).

**Table 2. Total phenolic and flavonoid content of \( L. \) bancanus leaves.**

| Sample                | Total phenolic | Total flavonoid |
|-----------------------|----------------|----------------|
|                      | (mg GAE/g dry sample) | (mg QE/g dry sample) |
| \( n \)-Hexane fraction | 0.472 ± 0.241\(^a\) | 0.463 ± 0.110\(^a\) |
| Dichloromethane fraction | 0.788 ± 0.125\(^a\) | 0.925 ± 0.216\(^a\) |
| Ethyl acetate fraction | 6.525 ± 0.188\(^a\) | 9.144 ± 0.138\(^a\) |
| Methanol extract      | 11.426 ±0.106\(^a\) | 15.422 ± 0.306\(^a\) |

Note: Data expressed as mean ± standard deviation (\( n = 3 \)). Same letters in each column mean no significant difference (\( P<0.05 \)). GAE: Gallat acid equivalents, QE: Quercetin equivalents.
they showed significantly different ($P<0.05$) (Table 3). The value of ABTS from methanol extract and ethyl acetate fraction were 2857.977 ± 0.715 and 1402.082 ± 0.371 μM TE/dry weight sample, respectively, and they exhibited highest ABTS values compared to $n$-hexane and dichloromethane.

**Correlation analyses between phenolic and flavonoid contents with antioxidant and Inhibitor α-glucosidase activities**

Correlation analyses (Table 4) between phenolic and flavonoid content with antioxidant (DPPH, FRAP, CUPRAC, ABTS) and antidiabetic (inhibitor α-glucosidase) activities were performed. Extract and fraction of *L. bancanus* leaves exhibited significant ($P<0.01$) linear correlations between TPC and TFC, TPC and IC$_{50}$ DPPH, TPC and FRAP, TPC and CUPRAC, TPC and ABTS, and TPC and IC$_{50}$ α-glucosidase inhibitor. By comparing the correlation coefficient ($r$) between TPC and CUPRAC exhibited highest $r$ value ($r=1$), followed by TPC and FRAP ($r=0.999$), TPC and ABTS ($r=0.997$), TPC and IC$_{50}$ ($r=0.996$), and TPC and IC$_{50}$ α-glucosidase inhibitor ($r=0.944$) and TPC and IC$_{50}$ DPPH ($r=0.944$). There were linear correlation ($P<0.01$) between TFC and CUPRAC, TFC and ABTS, and TFC and FRAP, TFC and IC$_{50}$ α-glucosidase and TFC and IC$_{50}$ DPPH with $r=0.999, 0.994, 0.993, 0.955$ and 0.739, respectively.

**Antidiabetic activity assay**

The α-glucosidase enzyme is the enzyme which responsible for breaking down disaccharides and complex carbohydrates into glucose. Inhibition of this enzyme can delay the absorption of glucose in the digestive tract, and to prevent an increasing in blood glucose concentration after eating. 29 α-glucosidase inhibition activity is one of method to determined antidiabetic activity. The mechanism of this assay was observed by interfering with the carbohydrate hydrolysis process, inhibits the absorption of glucose and other monosaccharides. Inhibition of this enzyme can effectively to reduce the digestion of complex carbohydrates and their absorption, so as to reduce the increase in postprandial glucose levels in diabetics. 30 The antidiabetic activity results showed significantly different ($P<0.05$), where methanol extract showed high activity followed by ethyl acetate fraction with IC$_{50}$ 30.565 ± 0.331 μg/mL, 44.901 ± 0.128 μg/mL, respectively (Table 5). There are significant correlation between total phenolics and flavanoids and the activity ($P<0.01$) with coefficient correlation ($r=0.944$ and 0.955 (Table 4).

### Table 3. Antioxidant activities (FRAP, CUPRAC and ABTS) of *L. bancanus* leave.

| Sample                  | FRAP (μM TE/g dry sample) | CUPRAC (μM TE/g dry sample) | ABTS (μM TE/g dry sample) |
|-------------------------|---------------------------|-----------------------------|---------------------------|
| $n$-Hexane fraction     | 126.508 ± 0.996           | 380.082 ± 0.544            | 77.517 ± 0.776            |
| Dichloromethane fraction| 190.050 ± 0.581           | 536.926 ± 0.579            | 131.634 ± 0.741           |
| Ethyl acetate fraction  | 1695.339 ± 0.372          | 15145.56 ± 3.107           | 1402.082 ± 0.371          |
| Methanol extract        | 3494.320 ± 0.456          | 26665.501 ± 5.940          | 2857.977 ± 0.715          |
| Quercetin               | 3492.846 ± 2.930          | 25848.774 ± 5.940          | 2742.498 ± 0.589          |
| Ascorbic Acid           | 35220.782 ± 0.674         | 28571.197 ± 5.941          | 2911.909 ± 0.889          |

Note: Data expressed as mean ± standard deviation ($n=3$). Same letters in each column mean no significant difference ($P<0.05$), TE: Trolox equivalents.

### Table 4. Correlation between phenolic and flavonoid contents with antioxidant and antidiabetic (inhibitor α-glucosidase) activities of *L. bancanus* leaves.

| TPC  | TFC  | IC$_{50}$ DPPH | FRAP  | CUPRAC  | ABTS  | IC$_{50}$ α-glucosidase |
|------|------|---------------|-------|---------|-------|-------------------------|
| -    | 0.999*| 0.730*        | 0.996*| 1.000*  | 0.997*| 0.944*                  |
| TFC  | -    | 0.730*        | 0.993*| 0.999*  | 0.994*| 0.955*                  |
| IC$_{50}$ DPPH | -     | -0.697        | 0.722*| 0.701*  | 0.844*| 0.816*                  |
| FRAP | -    | 0.995*        | 1.000*| 0.916*  | 0.947*| 0.920*                  |
| CUPRAC | -     | -0.996        | -     | -       | -     | -                       |
| ABTS | -    | -             | -     | -       | -     | -                       |
| IC$_{50}$ α-glucosidase | -  | -             | -     | -       | -     | -                       |

**Correlation is significant at the 0.01 level (2-tailed).**

*Correlation is significant at the 0.05 level (2-tailed).**

### Table 5. Antidiabetic (Inhibitor α-glucosidase) activity of *L. bancanus* leaves.

| Sample                  | IC$_{50}$ (μg/mL) |
|-------------------------|-------------------|
| $n$-Hexane fraction     | 116.607 ± 1.379a   |
| Dichloromethane fraction| 102.189 ± 1.631b   |
| Ethyl acetate fraction  | 44.901 ± 0.128c    |
| Methanol extract        | 30.565 ± 0.331d    |
| Acarbose                | 18.173 ± 0.122e    |

Note: Data expressed as mean ± standard deviation ($n=3$). Same letters in each column mean no significant difference ($P<0.05$).
CONCLUSION

The leaves extract and its n-hexane, dichloromethane and ethyl acetate fractions of L. bancanus showed high antioxidant and antidiabetic activities, especially ethyl acetate fraction and methanol extracts. It could be concluded that leaves of L. bancanus has antioxidant and antidiabetic properties.

ACKNOWLEDGEMENTS

Thanks to Ministry of Research, Technology and Higher Education of the Republic of Indonesia for supporting this research through postgraduate grant research, Contract Number: 339/UN.19.5.1.3/PP/2018.

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GRAPHICAL ABSTRACT

Lithocarpus bancanus

Extraction

Antioxidant Activities

Phenolic & Flavonoid Contents
PRAF, CUPRAC & ABTS

DPPH radical

Antidiabetic activity

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Cite this article: Almurdani M, Zamri A, Nugroho TT, Karim J, Eryanti Y, Hendra R, et al. Antioxidant and Antidiabetic Activities of Mempening (Lithocarpus bancanus) Leaves. Pharmacog J. 2020;12(2):328-34.