Prevalence of *Staphylococcus aureus* and *Staphylococcus sciuri* Isolated from Apparently Healthy and Hospitalized Patients in Ekiti State, Nigeria

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Authors’ contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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ABSTRACT

The emergence of microbial resistance to antibiotics on daily basis has become a major global challenge. The increasing prevalence of methicillin resistant *Staphylococcus aureus* (MRSA) infections among hospitalized and non-hospitalized patients due to emergence of unique community-associated strains has become a great problem in Nigeria in particular and the entire world in general. In this study, a total of four hundred (400) clinical specimens were collected from hospitalized patients at Ekiti State University Teaching Hospital (EKSUTH), Ado-Ekiti and Federal Teaching Hospital (FETHI), Ido-Ekiti and from apparently healthy individuals from both
communities. Standard bacteriological procedures were employed to isolate and identify these organisms. Antibiotic susceptibility test was carried out using the modified Kirby Bauer disc diffusion method. Polymerase chain reaction (PCR) assay was used to identify S. aureus nuc gene, as well as mecA and aac resistance genes. Seventy seven (19.3%) S. aureus isolates were phenotypically identified; 43 (21.5%) hospital-associated (23 from EKSUTH and 20 from FETHI) and 34 (17.0%) community-associated (27 from Ado-Ekiti and 7 from Ido-Ekiti communities). No S. sciuri was phenotypically isolated in both locations. Fifty five (71.4 %) of the 77 S. aureus isolates were nuc gene positive, while 11 (20%) of the 55 isolates were mecA gene positive, implying that they were MRSA. Of the 11 isolates, 6 (19.4%) were hospital-associated MRSA while 5 (20.8 %) were community-associated MRSA. Phenotypic resistance of S. aureus to cefoxitin in the two locations ranged between 77.8 – 100%, but 80% of cefoxitin-resistant nuc gene positive S. aureus lacked mecA gene. S. aureus isolates exhibited high phenotypic resistance to tetracycline, erythromycin and fusidic acid but susceptible to clindamycin and gentamicin. The prevalence of clinical MRSA infection of 20% in this study is relatively high. However, there was 0% prevalence of S. sciuri colonization. Cefoxitin disc test demonstrated low specificity as a phenotypic marker of methicillin resistance. There is need to institute control measures for MRSA infections and colonization in this environment.

Keywords: Staphylococcus aureus; Staphylococcus sciuri; antibiotic susceptibility.

1. INTRODUCTION

Staphylococcal infections are a major cause of nosocomial infections. The spectrum of infections can range from a minor boil or skin abscess to life threatening infections such as septicaemia or endocarditis. There are many species of staphylococci but most infections are caused by Staphylococcus aureus. A number of studies done on S. aureus both in Nigeria and elsewhere have focused on nasal carriage of the organism and its impact on skin and soft tissue infections [1]. Most nosocomial pathogens associated with multiple antimicrobial resistance mechanism are Staphylococci. For many years, S. aureus was the only species recognized as an important human pathogen whereas coagulase-negative staphylococci (CONS) were viewed mostly as clinically non relevant contaminant or commensals. Only recently, the importance of CONS strains as a major cause of nosocomial infections began to be established [2]. Infections with S. aureus are especially difficult to treat because of evolved resistance to antimicrobial drugs such as penicillin and newer narrow spectrum drugs and β-lactamase resistance penicillin-antimicrobial drug [3]. The widespread incidence of antibiotics resistance across various strains of S. aureus has been attributed to horizontal transfer of genes encoding antibiotics resistance and virulence [4]. Methicillin resistant S. aureus (MRSA) is the bacterium responsible for several difficult-to-treat infections in human. Strains of methicillin resistant S. aureus (MRSA) that are common causative agents of nosocomial disease world-wide often pose therapeutic dilemma to clinicians because of the nature of these strains [5]. It is any strain of S. aureus that has developed through the process of natural selection, resistance to β-lactam antibiotics which include methicillin, dicloxacillin, nafcillin, oxacillin and cephalosporins. Strains unable to resist these antibiotics are classified as methicillin sensitive S. aureus (MSSA). The evolution of this resistance makes MRSA infection very difficult to treat with standard antibiotics and thus making it more dangerous. Resistance in MRSA is related to a chromosomal mecA gene that encodes the production of an abnormal penicillin binding protein called PBP2a. These proteins are membrane bound enzymes which are target for all β-lactam antibiotics. They have decreased affinity for binding β-lactam antibiotics resulting not only to methicillin resistance but also to all β-lactams including penicillin and cephalosporins [6]. Staphylococcus sciuri strains have been found to be an important pathogen responsible for UTI [7], wound infection [8] endocarditis [9], peritonitis septic shock endophthalmitis [10] and pelvic inflammatory diseases [11]. S. sciuri may be found as a colonizing organism in humans with low carrier rates in nasopharynx, skin and urogenitals [12,13]. The mecA gene is native to S. sciuri which has been found to have the same similarity with mecA gene in S. aureus [14]. Inspite of the close sequence similarities between the mecA of S. sciuri and the antibiotic resistance mecA gene of S. aureus, S. sciuri strains have been found to be uniformly susceptible to β-lactam antibiotics. It has been noted that S. sciuri may be an
evolutionary precursor of the methicillin resistance gene mecA of the pathogenic strains of MRSA [13]. Though in Nigeria, S. aureus is one of the major nosocomial infection pathogen [14]. S. sciuri also has become another major pathogen in nosocomial infection [14]. which result in morbidity and mortality. The aim of this study therefore was to compare the prevalence of Staphylococcus aureus among hospitalized patients and apparently healthy persons in the community with that of Staphylococcus sciuri among hospitalized patients and apparently healthy persons in the community.

2. METHODOLOGY

2.1 Study Area

This study was carried out in two hospitals (Federal Teaching Hospital FETHI, Ido Ekiti and Ekiti State Teaching Hospital EKSUTH Ado-Ekiti) and Ado-Ekiti and Ido-Ekiti communities both in Ekiti State, Southwest, Nigeria.

2.2 Study Population

These consist of selected hospitalized patients in FETHI and EKSUTH and apparently healthy individuals from Ido and Ado-Ekiti communities from whom appropriate clinical samples were obtained.

2.3 Inclusion Criteria

The inclusion criteria were hospitalized patients with clinical symptoms and signs of infection, age range 15 to 60 years, and not less than one month of hospitalization, and apparently healthy individuals from the two communities in the same age group.

2.4 Specimen Collection and Transport

Urine, wound, blood and pus samples were obtained from hospitalized patients while nasal samples and urine were obtained from healthy individuals. Samples were collected into appropriately label specimen bottles using aseptic techniques to prevent contamination, and transported immediately to the laboratory for analysis.

2.5 Sample Analysis

2.5.1 Isolation of staphylococci

Wound, pus and nasal specimens were inoculated onto MacConkey agar and Chocolate agar plates and incubated at 37°C overnight to primarily isolate staphylococci species. The staphylococci colonies were then subcultured onto Mannitol salt agar (MSA) and Tryptic Soy Agar (TSA) for isolation of S. aureus and S. sciuri respectively [15]. Urine specimens were first cultured on Cysteine Lactose Electrolyte Deficient (CLED) agar and incubated at 37°C for 24 hrs (Cheesbrough, 2006). Isolated staphylococci were then sub-cultured onto MSA and TSA for isolation of S. aureus and S. sciuri respectively. Pure colonies of suspected S. aureus and S. sciuri were then stored on nutrient agar slopes for further phenotypic tests.

2.5.2 Speciation of isolates

S. aureus was identified based on Gram reactions, catalase positive test, coagulase positive test, fermentation of mannitol while S. sciuri was identified based on catalase positive test, tube coagulase negative, oxidase test positive, urease test positive, esculin hydrolysis, resistance to novobiocin and bacitracin, acid production from galactose and sucrose but not raffinose fermentation [16].

2.6 Molecular Analysis

2.6.1 DNA extraction

The DNA of the isolates was extracted by suspending 4-5 bacteria colonies in 500 µl of Tris Borate EDTA (TBE) buffer in Eppendorf tubes appropriately labeled. The cells were boiled at 100°C for 10 minutes and were cooled rapidly on ice for 30 minutes. 3ul of Proteinase K was added to the lysed cells and the mixture was incubated for 15-20 minutes at 55-60°C. The enzyme was denatured by boiling at 100°C for 10 minutes and was centrifuged at 13, 400 rpm. The supernatant containing the DNA was collected for PCR and was stored at -20°C for further use.

2.6.2 Amplification of nuc, mecA and aac-genes

Polymerase chain reaction (PCR) was used to detect nuc gene that is common to all S. aureus and resistance genes; mecA that encodes methicillin resistance and aac that encodes aminoglycoside resistance.

2.7 PCR Procedural Steps

2.7.1 Gel Electrophoresis

Preparation of agarose gel: At the completion of the amplification, PCR products were resolved
on 1% agarose gel prepared by dissolving 1g of agarose powder in 100 ml of 1X TBE buffer solution inside a clean conical flask. The 1% agarose solution was heated in a microwave oven for 2-3 minutes and was observed for clarity which was an indication of complete dissolution. The mixture was then allowed to cool to about 50 °C after which 0.5 µl of ethidium bromide was then added. It was allowed to cool further and then poured into a tray sealed at both ends with support to form a mould with special combs placed in it to create wells. The comb was carefully removed after the gel had set.

2.8 Analysis and Interpretation of Gel Pattern

The DNA fragment of 276bp and 310bp were separated for nuc and mecA gene respectively on a 1% agarose gel with a 100bp DNA ladder.

2.9 Data Entry and Statistical Analysis

All data were analysed using statistical package for the social sciences (SPSS) version 16.0. Data were presented using frequency tables as appropriate and cross tabulation to study relationships associated between variables. Statistical significance was set at 5% using chi-square.

3. RESULTS

A total of 400 clinical samples were collected from hospitalized patients in two hospitals and apparently healthy individuals in two communities in Ekiti State, Nigeria. Table 1 shows the distribution of organisms isolated from the study areas. Seventy seven (19.25%) S. aureus was phenotypically isolated from the 400 samples collected; 23 (5.75%) from Ekiti State University Teaching Hospital (EKSUTH), Ado- Ekiti, 20 (5%) from Federal Teaching Hospital (FETHI), Ido-Ekiti, 27 (6.75%) from Ado-Ekiti community and 7 (1.75%) from Ido-Ekiti community. In this study, no S. sciuri was isolated phenotypically in all the locations. The prevalence of S. sciuri in these study areas by the standard phenotypic detection method is therefore 0%.

The comparison of Cefoxitin resistance to all other antibiotics of all the isolate obtained in all the location is shown in table 2 only Erythromycin and Gentamicin have no statistical significant.

Table 1. Primers used for the amplification of genes

| Primers   | Sequence 5'-3'          | gene  | Product size(bp) | Annealing temp.(°C) | Reference       |
|-----------|-------------------------|-------|------------------|---------------------|-----------------|
| SANucF    | GCGATGTGATGGTAGACCGTT   | SA    | 276              | 58                  | Brakstad et al. [17] |
| SANucR    | AGCCAGCCCTTGACGAACCTAAAGC | Nuc   |                  |                     |                 |
| MecA F    | GTAGAATGATCTGACGTCCGATAA | MecA  | 310              | 52                  | Geha et al. [18] |
| MecA R    | CCAATTCACATTGTGTTCGGTCTAA |       |                  |                     |                 |
| AAC-3-IV F| AGTTGACCCAGGGCG          | AAC-3-IV | 286        | 55                  |                 |
| AAC-3-IV R| GTGTGCTGCTGGTCACAGC      |       |                  |                     |                 |

Table 2. Distribution of staphylococcus isolates detected by phenotypic method in two locations in Ekiti State, Nigeria

| Staphylococcus isolate | Hospital (%) | Community (%) | Total (%) | 95% CI | P value |
|------------------------|--------------|---------------|-----------|--------|---------|
|                        | FETHI | EKSUTH | Ido-Ekiti | Ado- Ekiti |        |         |
| No of samples          | 100   | 100    | 100       | 100     | 400     |         |
| No of S. aureus        | 20 (20)| 23 (23) | 7 (7)     | 27 (27) | 77 (19.3) | 0.3851  |
| No of S. sciuri        | 0     | 0      | 0         | 0       | 0       |         |
| Total isolate          | 43 (21.5)| 34 (17.0)| 77 (19.3) | 0.3851 | 0.2449  |

FETHI = Federal Teaching Hospital; EKSUTH = Ekiti State University Teaching Hospital, CI = Confidence Interval
Table 3 shows the distribution *S. aureus* isolates with *nuc* and *mecA* genes in the study areas. Aminoglycoside resistance, *aac* gene, was not detected in any isolate. Fifty five (71.43%) of the 77 phenotypically confirmed *S. aureus* isolates were positive for *nuc* gene, while 11 (20%) of these were positive for *mecA* gene. The prevalence of MRSA in these study areas using the gold standard PCR detection method is therefore 20%.

4. DISCUSSION

Emergence of resistance to antibiotics on daily basis has become a serious global health challenge. The increasing prevalence of MRSA infection among non hospitalized patients due to emergence of unique community associated *S. aureus* and hospitalized patients has become a great problem in Nigeria in particular and the globe in general. MRSA can be found in any individual regardless of their age. Isolates from the hospitalized patients were from urine, wound and aspirate while from the community the isolation was from urine and nasal specimen. Fifty-six percent of the *S. aureus* isolates was hospital associated while 44% were community associated. In this study, phenotypic resistance of *S. aureus* to cefoxitin in the two locations used ranges between 77.8% - 100% which when compared to previous work done in Ekiti by Olowe et al. [19] shows 46.5% resistance. The antimicrobial susceptibility of cefoxitin resistance to *S. aureus* in this study shows that *S. aureus* is multi-resistant to tetracycline as described previously by Yanagihara et al. [20]. *S. aureus* isolates in the hospitals were found to show more resistance to erythromycin while fusidic acid shows a high rate of resistance to isolates of *S. aureus* in the community. Gentamycin shows a high rate of sensitivity ranging between 73.9-79.3% in Ado Ekiti, while it shows 100% sensitive in Ido Ekiti. Macrolides and aminoglycosides show a very low resistance in this study as against Taiwo et al. [21]. The high rate of clindamycin and gentamycin susceptibility in all isolates from both the hospital and community in this study does not support previous work done by Gould et al. [22] and Yanagihara et al. [23]. This suggests that the susceptibility of clindamycin and gentamycin varies with location and frequency of use of antibiotics, likewise fusidic acid and erythromycin. The susceptibility of clindamycin to MRSA in this study shows that clindamycin can be used as drugs of choice in the treatment of MRSA in combination with gentamycin. Hospital acquired MRSA still remains a major challenge in the medical health care in Nigeria. Hospital acquired *S. aureus*’ genetic resistance to Cefoxitin of 27% in each location is very high compared to the Community acquired resistance of 5.4-9% and 1.8-5.4% in each location as shown in this study. The distribution of MRSA detection in both the hospital and community in Ekiti shows that genetically hospital associated MRSA is 19.4% and community associated, 20.8%. Compared to previous studies by Olowe et al. [24] of 19.2%, 1.4% by Adesida et al. [25] 1.5% by Shittu et al., (2006) in Southwest Nigeria appears to increase as the year goes by. But compared to studies by Terry, of 22.2% and Taiwo et al., (2004) of 34.7% in other part of the country, the prevalence high rate of MRSA varies according to location and time which supports Okon et al., studies in (2013). 38.8% by Fayomi et al. [24] in Ido Ekiti compared to the 27% prevalent rate in this study shows that the rate has reduced but still on the high side. The MRSA isolated in this studies was found to be multi-resistant to tetracycline and fusidic acid as earlier reported by Vaez et al. [25] Cefoxitin has greatly shown a high rate of resistance of 95% phenotypically in this study, this shows that cefoxitin has develop a high rate of resistance when compare to studies done by Olowe et al. [26].

MSSA (methicillin sensitive *staphylococcus aureus*) also shows multiresistance to tetracycline, erythromycin, and fusidic acid in this study. According to this study, 80% of phenotypically cefoxitin resistant *S. aureus* does not possess the resistance gene mecA (MSSA-mecA negative). This suggests that disc diffusion test for the detection of MRSA gives a high false prevalence rate as described by Syed et al. [27]. PCR amplification still remains the gold standard for the detection of MRSA and MSSA. According to this study, isolation of MRSA in relation to gender, occupation and age group shows no relationship. Phenotypically, MRSA was found to be high in urine 92.9%, wound swab 100% and nasal swab of 87% but shows no statistically significant between the isolation of MRSA and sample site. The high rate of detection of MRSA in the hospitalized patient shows that HA-MRSA still remains a very big challenge.
### Table 3. Comparative antibiotic susceptibility of hospital and community associated Staphylococcus aureus isolates in Ekiti State, Nigeria

| Antibiotics    | S. aureus isolates | P value |
|----------------|--------------------|---------|
|                | Hospital-associated (n=43) | Community-associated (n=34) |
|                | S | I | R | S | I | R |
| Cefoxitin      | 0 | 11 | 32 | 6 | - | - |
| Fusidic acid   | 25 | 13 | 5 | 37 | 4 | 22 |
| Clindamycin    | 5 | 14 | 24 | 10 | 12 | 12 |
| Erythromycin   | 37 | 2 | 4 | 3 | 12 | 20 |
| Gentamicin     | 4 | 1 | 20 | 3 | 28 | 3 |
| Tetracycline   | 22 | 1 | 31 | 3 | 3 | 3 |

* Statistically significant difference. S = sensitive, I = intermediate, R = resistant. Note that for statistical analysis, intermediate isolates were considered resistant.

### Table 4. Distribution of Staphylococcus aureus isolates detected by molecular method in two locations in Ekiti State, Nigeria

| Staphylococcus aureus isolate | Hospital (%) | Community (%) | Total (%) | 95% CI | P value |
|-------------------------------|--------------|---------------|-----------|--------|---------|
|                               | FSH, Ido-Ekiti | EKSUTH Ado-Ekiti | Ido-Ekiti | Ado-Ekiti |         |         |
| No positive for nuc gene      | 16 | 15 | 5 | 19 | 55 |        |         |
| No positive for mecA gene (MRSA isolate) | 3 (18.8) | 3 (20) | 1 (20) | 4 (21.1) | 11 |        |         |
| Total MRSA isolate            | 6 (19.4) | 5 (20.8) | 11 (20) |         | 0.5279 - 1.746 | 1.000 |

*FSH = Federal Teaching Hospital; EKSUTH = Ekiti State University Teaching Hospital, MRSA = methicillin resistant Staphylococcus aureus*
Fig. 1. PCR amplification of Nuc gene for *staphylococcus aureus*
Fig. 2. PCR amplification of mecA gene of some of the Cefoxitin resistant isolates
The route of *S. sciuri* is usually transmitted from animal to human and in this study, the individuals involved had no close contact with animals frequently. The absence of *S. sciuri* in the locations of this study might be due to the low population density as well as the requirement of enriched environment for isolation as previously described by Ivana et al. or no contact with infected animal and animal dairy. The frequently colonization of the hospital environment which makes rate of isolation to be sporadic and most probably transient as described by Ahoyo et al. [28] might also be a contributing factor to why *S. sciuri* was not isolated during the cause of this study. Non isolation of *S. sciuri* does not primarily indicate that the organism does not exist in the area of study and not necessarily means that it does exist. This study suggests that for the detection of interaction between *S. aureus* and *S. sciuri* based on the presence of *mecA* gene will greatly depend on the isolation of both organisms in the same location and individual to clearly give the better understanding and analysis of the nucleotides of the amino acid sequences as done previously by Couto et al. [29-35]. Though, previous study done already shows that there is a relationship in the nucleotide sequence of the *mecA* in MRSA and *S. sciuri*.

There is a limited study on the isolation and prevalence of *S. sciuri* in Ekiti State therefore more studies are needed in the area of isolation and detection of *S. sciuri* in humans in this region, and Also the interaction and relationship between *S. aureus* and *S. sciuri* based on their *mecA*. Until these two organisms are isolated together in this region, the study of their interactions will still be difficult.

5. CONCLUSION

The high prevalent rate of MRSA in this study shows that MRSA continues to be a major challenge in our society and nation and disc diffusion test is not 100% precise in detecting MSSA. PCR remains the gold standard to detect and isolate both MRSA and MSSA. On the isolation and detection of *S. sciuri* in human, more work needs to be done in order to have the complete understanding and knowledge about the organism, then the interaction between *S. aureus* and *S. sciuri* can be fully understand. There is a need to put more control measures in place to eradicate MRSA to reduce one of the major healthcare challenges. Therefore, everyday awareness on proper hygiene and usage of antibiotics should be put in place.
6. RECOMMENDATION

Since MRSA still pose a lot of threat to the medical services, the healthcare unit and the general public should be exposed to more education on proper hygiene and sanitation periodically. Proper handwashing techniques and procedures should henceforth be in place in all healthcare service provider units. Further studies should be encouraged in the aspect of identifying S. sciuri in clinical specimens using molecular methods, and to understand its interaction with S. aureus in human infection or colonization.

CONSENT

As per international standard or university standard, patients’ written consent has been collected and preserved by the author(s).

ETHICAL APPROVAL

The ethical clearance was obtained from Ekiti State University Teaching Hospital (Protocol number: EKSUTHA67/2015/08/004)

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Sheng-Yun L, Fang-Yu C, Ching-Chung C, Keong-Diong L, Yhu-Chering H. Methicillin-resistant staphylococcus aureus nasal colonization among adult patients visiting emergency department in a medical centre in taiwan. PLoS One.2011;6:e18820.
2. Kloos WE, Schleifer KH, Smith RF. Characterization of Staphylococcus sciuri sp. nov. and its subspecies. Int J Syst Bacteriol. 1976;26:22-37.
3. Lowy FD. Antimicrobial resistance: the example of Staphylococcus aureus. J. Clin. Invest. 2003; 111(9):1265–73.
4. Chan CX, Beiko RG, Ragan MA. Lateral transfer of genes and gene fragments in Staphylococcus extends beyond mobile elements”. J Bacteriol. 2011;193(15).
5. Taiwo SS, Onile BA, Akanbi AA, Il. Methicillin-resistant Staphylococcus aureus (MRSA) isolates in Ilorin, Nigeria. African Journal of Clinical and Experimental Microbiology. 2004;5(2):189–197
6. Weems JJ. The many faces of Staphylococcus aureus infections. Postgraduate Med. 2001; 110(4):24-36.
7. Stepanovic SI, Dakic N, Opavski P, Jezek L, Ranin. Influence of the growth medium composition on biofilm formation by Staphylococcus sciuri. Ann. Microbiol. 2003;53:63–74.
8. Shittu AO, Lin J, Kolawole DO. Antimicrobial susceptibility patterns of Staphylococcus aureus and characterization of MRSA in Southwestern Nigeria. Wounds. 2006;18:77-84.
9. Hedin G, Widerstrom M. Endocarditis due to Staphylococcus sciuri. Eur. J. Clin. Microbiol. Infect. Dis. 1998;17:673-675.
10. Couto IH, de Lencastre E, Severina W, Kloos J, Webster I. Santos Sanches, and A. Tomasz. Ubiquitous presence of a mecA homologue in natural isolates of Staphylococcus sciuri. Microb. Drug Resist. 1996;2:377–391.
11. Stepanovic SI, Dakic S, Djukic B, Lozuk M, Svabic-Vlahovic. Surgical wound infection associated with Staphylococcus sciuri. Scand. J. Infect. Dis. 2002;34:685–686.
12. Couto ISW, Wu A, Tomasz H, de Lencastre. Development of methicillin resistance in clinical isolates of Staphylococcus sciuri by transcriptional activation of the mecA homologue native to S. sciuri. J. Bacteriol. 2003;185:645–653.
13. Wu SW, DE Lencastre H, Tomasz A. Recruitment of the mecA gene homologue of Staphylococcus sciuri into a resistance determinant and expression of the resistant phenotype in Staphylococcus aureus. J Bacteriol. 2001;183:2417–24.
14. Ivana D, Donald M, Dragana V, Branislava S, Adebayo S, Petr J, Tomasz H, Srdjan S. Isolation and Molecular Characterization of Staphylococcus sciuri in the Hospital Environment. J Clin. Microbiol. 2005;43: 2782–2785.
15. Shittu AJ, Lin D, Morrison D, Kolawole. Isolation and molecular characterization of multiresistant Staphylococcus sciuri and Staphylococcus haemolyticus associated with skin and soft-tissue infections. J. Med. Microbiol. 2004;53:51–55.
16. Kloos WE, Ballard DN., Webster JA., Hubner RJ, Tomasz A, Couto I, Sloan GL, Dehart HP, Fiedler F, Schubert K, De Lencastre H, Santos Sanches I, Heath HE, Leblanc PA, Ljungh A. Ribotype delineation and description of Staphylococcus sciuri subspecies and their
potential as reservoirs of methicillin resistance and staphyloloytic enzyme genes. Int. J. Syst. Bacteriol. 1997; 47:313-323.

17. Brakstad OG, Aasbakk K, Maeland JA. Detection of *Staphylococcus aureus* by polymerase chain reaction amplification of the nuc gene. J. Clin. Microbiol. 1992;30:1654-1660.

18. Geha DJ, Katayama Y, Hiramatsu K. Cloning and nucleotide sequence determination of the entire mec DNA of premethicillin-resistant *Staphylococcus aureus* N315. Antimicrob Agt.& Chemo. 1994;43:1449-1458.

19. Olowe OA, Eniola KIT, Olowe RA, Olayemi AM. Antimicrobial susceptibility and betalactamase detection of MRSA in Osogbo, S.W. Nigeria. Nat Sci. 2007;5:44-48.

20. Yanagihara K, Araki N, Watanabe S, Kinebuchi T, Kaku M, Maesaki S, Yamaguchi K, Matsumoto, T, Mikamo, H, Takesue Y, Kadota, JI, Fujita, J, Iwatsuki, K, Hino, H, Kaneko, T, Asagoe, K, Ikeda, M, Yasuoka A, Kohno S. Antimicrobial susceptibility and molecular characteristics of 857 methicillin-resistant Staphylococcus aureus isolates from 16 medical centers in Japan (2008-2009): nationwide survey of community-acquired and nosocomial MRSA. Diagnostic Microbiology and Infectious Disease. 2012;72:253-257.

21. Gould, SWJ, Cuschieri, P, Rollason, J, Hilton, AC, Easmon, S, Fielder, MD. The need for continued monitoring of antibiotic resistance patterns in clinical isolates of *Staphylococcus aureus* from London and Malta. 2010;9:1-7.

22. Adesida S, Boelen H, Babajide B, Kehinde A, Snijders S, van Leeuwen W, Coker A, Verburgh H, van Belkum A. Major epidemic clones of *Staphylococcus aureus* in Nigeria. Microbial. Drugs Resistance. 2005;11(2):115-121.

23. Okon K.O, Basset P, Uba A, Lin J, Oyawoye B, Shittu AO, Blanc DS. Co-occurrence of predominant PVL-positive (ST152) and multidrug-resistant (ST241) *Staphylococcus aureus* clones in Nigerian hospitals. J. Clin. Microbiol. 2009;47:3000-3003.8

24. Fayomi OD, Oyediran EIO, Adeyemo AT, Oyekale OT. Prevalence and antibiotics resistance of Methicillin-Resistance *Staphylococcus aureus* among inpatients at a tertiary health facility in Ido-Ekiti Nigeria. Int. J. Lab. Med. 2009;4:2

25. Vaez H, Tabaraei A, Moradi A, Ghaemi EA. Evaluation of methicillin-resistant *Staphylococcus aureus* (MRSA) isolated from patients in Golestan, Province-North of Iran. Afr. J. Microbiol. Res. 2011;5 (4):432-436.

26. Olowe OA, Kukoyi OO Taiwo S.S, Ojurongbe OO, Opaleyee OO, Bolaji OS, Abiodun AA, Makanjuola OB, Ogbolu DO, Alli OT. Phenotypic and molecular characteristics of methicillin-resistant *Staphylococcus aureus* isolates from Ekiti State, Nigeria; 2013.

27. Syed ZB, Safia A, Naheed Z. Antimicrobial susceptibility pattern of *Staphylococcus aureus* on clinical isolates and efficacy of Laboratory test to diagnose MRSA: A multicentre study. J Ayub Med Coll Abbottabad. 2011;23(1):139-142.

28. Ahoyo TA, Yehouenou Pazou E, Baba-Moussa L, Attolou Gbohou A, Boco M, Dramane KL and Aminou T. *Staphylococcus sciuri* outbreak at Tertiary Hospital in Benin. J Med Microb Diagn. 2013;2:126

29. Couto IIS, Sanches R, Sa-Leao H, de Lencastre. Molecular characterization of *Staphylococcus sciuri* strains isolated from humans. J. Clin. Microbiol. 2000;38:1136–1143.

30. Annad KB, Agrawal P, Kumar S, Kapila K. Comparison of cefoxitin disc diffusion test, oxacillin screen agar and PCR for mecA gene for detection of MRSA. Indian J. Med Microbiol. 2009; 27:27-29

31. Clinical and Laboratory Standard Institutes (CLSI). Performance Standard for Antimicrobial Disk Susceptibility Test. Approved Standard. Eleventh Edition. CLSI document M02-A11. 2012;32:1.

32. Clinical and Laboratory Standard Institutes (CLSI). Disc diffusion supplementary table. Performance Standard for Antimicrobial Disk Susceptibility Test. 940 West Valley Road Suite 1400, Wayne, PA19807; 2013.

33. Rushdy AA, Salama MS, Othman AS. Detection of Methicillin/oxacillin resistant S. aureus isolated from some clinical hospitals in Cairo using mecA/nuc genes and antibiotic susceptibility profile. Int J. Agric and Biol. 2007;9(6):800–806

34. Stepanovic SP, Jezek D, Vukovic I, Dakic P, Petras. Isolation of members of the *Staphylococcus sciuri* group from urine and their relationship to urinary tract...
infections. J. Clin. Microbiol. 2003;41: 5262–5264.
35. Valones MAA, Guimarães RL, Brandão LAC, Eleutério de Souza PR, Carvalho AT, Crovela S. Principles and application of polymerase chain reaction in medical diagnostic fields: A Review. Brazil J of microbiol. 2009;40:1-11.

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