Cytogenetic description of the Amazonian brown brocket Mazama nemorivaga (Artiodactyla, Cervidae)

Bruno Ferreto Fiorillo¹, Javier Adolfo Sarria-Perea¹², Vanessa Veltrini Abril¹², José Maurício Barbanti Duarte¹

¹ Núcleo de Pesquisa e Conservação de Cervídeos (NUPECCE), Departamento de Zootecnia, Faculdade de Ciências Agrárias e Veterinárias, UNESP, Campus de Jaboticabal, 14884-900, Jaboticabal, SP, Brazil
² Programa de Pós-Graduação em Genética e Melhoramento Animal, Faculdade de Ciências Agrárias e Veterinárias, UNESP, Campus de Jaboticabal, 14884-900, Jaboticabal, SP, Brazil

Corresponding author: Vanessa Veltrini Abril (vanessa.abril@gmail.com)

Academic editor: E. Gornung | Received 14 November 2012 | Accepted 9 January 2013 | Published 18 March 2013

Citation: Fiorillo FB, Sarria-Perea JA, Abril VV, Barbanti Duarte JM (2013) Cytogenetic description of the Amazonian brown brocket Mazama nemorivaga (Artiodactyla, Cervidae). Comparative Cytogenetics 7(1): 25–31. doi: 10.3897/CompCytogen.v7i1.4314

Abstract

The Amazonian brown brocket Mazama nemorivaga (Cuvier, 1817) is a small to medium-sized deer from the Amazon rainforest and ecotones. The first karyotype described was 2n=67 to 69 + 2-7 B and FN=69-72, in which all chromosomes were acrocentric and the X chromosome was the only submetacentric chromosome. However, important aspects of the species chromosome evolution were not resolved because of the lack of information on chromosome banding. The G-banding pattern of Mazama nemorivaga karyotype showed the presence of an XX/XY₁ Y₂ sex chromosome system as a product of an X-autosome tandem fusion, which results in a basic 2n=68, FN=70 in females and 2n=69, FN=70 in males. The fact that this karyotype only differs from that of Capreolus capreolus pygargus (Pallas, 1771; 2n=70, FN=72+B) by X-autosome tandem fusion may corroborate the basal condition of Mazama nemorivaga and its proximity to the ancestral karyotype of the American Odocoileini. A derived karyotype 2n=67, XY₁ Y₂, FN=70 + 3B from the Brazilian state of Mato Grosso (the western Amazon) may be evidence of differentiation between western and eastern populations.

Keywords

chromosome banding, centric fusion, B chromosomes, multiple sex chromosome system
Introduction

The Amazonian brown brocket *Mazama nemorivaga* (Cuvier, 1817) is a small to medium-sized deer that possesses a large muzzle, small ears, protruding eyes, small and spike-like antlers in males and a uniform dark brown coat. This species occurs mainly in the Amazon rainforest and ecotones of Brazil, French Guiana, Surinam, Guyana, Venezuela, Colombia, Ecuador, Peru, and most likely in Bolivia. Although classified as a species of Least Concern by the Red List of the International Union for Conservation of Nature (IUCN) because of its wide distribution, the species would be seriously affected by the effects of deforestation (Rossi et al. 2010). This species belongs to the Rangiferini tribe, which was recently included within the subfamily Capreolinae (González et al. 2010). It was once classified as a subspecies of the gray brocket *M. gouazoubira superciliaris* (Cabrera, 1961), but recent molecular analyses demonstrated that *M. nemorivaga* is a clearly differentiated species, located basally in the exclusively South American clade of Odocoileinae that also groups together *Blastocerus* (Illiger, 1815), *Ozotoxerus* (Linnaeus, 1758), *Hippocamelus* (Molina, 1782), and *Pudu* (Molina, 1782), and which includes *M. gouazoubira* (Fisher, 1814) (Duarte et al. 2008). Cytogenetics also support this finding, since the karyotype of *M. nemorivaga* presents as 2n=67-69 + 2-7 B and FN=69-72, with a submetacentric X, a very different karyotype from that of *M. gouazoubira*: 2n=70+0-3B and FN=70 with an acrocentric X (Rossi et al. 2010). However, the lack of studies on chromosome banding patterns in this species makes it difficult to resolve important aspects of its chromosome evolution. The present study provides much of this information through the analysis of G, C and NOR banding patterns. It also brought new insights for the understanding of the complex chromosome evolution of South American deer.

Material and methods

Seven wild-caught specimens of *M. nemorivaga* were analyzed: one male and four females from the city of Santarém (Pará State, Brazil), one male from the city of Imperatriz (Maranhão State, Brazil), which lies within the eastern Amazon, and one male from the city of Juína (Mato Grosso State, Brazil), which lies within the western Amazon (Figure 1). The males from Imperatriz and Juína and one female are currently being kept in the captive breeding facilities of the “Deer Research and Conservation Center” (NUPECCE – Núcleo de Pesquisa e Conservação de Cervídeos, at São Paulo State University’s, Jaboticabal campus); the other four animals died before the analysis and the cell cultures were prepared from frozen skin samples. The sampling was not homogeneous throughout the *M. nemorivaga* distribution area due to the difficulty in capturing free-living animals and the small number of captive specimens in Brazil.

Metaphase chromosomes were obtained from lymphocyte (Moorhead et al. 1960) and fibroblasts (Verma and Babu 1995) cultures using peripheral blood and skin
fragments, respectively. The chromosomes were studied using G-banding (Seabright 1971), C-banding (Sumner 1972), and Ag-NOR staining (Howell and Black 1980). Approximately 40 metaphases from each specimen were analyzed in order to determine the diploid number (2n) and the fundamental number (FN). The chromosomes were classified following Abril and Duarte (2008) as metacentric, submetacentric or acrocentric according to their arm ratio, and were then organized into groups according to their relative lengths (RL): A (biarmed chromosomes with RL > 2.5%), C (biarmed chromosomes with RL < 2.5%), D (acrocentric chromosomes with RL < 3.0%), E (acrocentric chromosomes with RL > 3.0%) and B (microchromosomes or extranumerary chromosomes with RL > 1.0%). B chromosomes were not considered in the calculation of the diploid or fundamental numbers because there was intraindividual variation (Abril and Duarte 2008).

Figure 1. Origin of analyzed animals of the species *M. nemorivaga*, where A = animals from Imperatriz, MA (T265), B = animals from Santarém, PA (T261, T262, T263, T264, T266), C = animals from Juina, MT (T275, T295). Modified version of the map by Colinvaux et al. 2000.
Results and discussion

The karyotypes of six animals from the eastern Amazon were found to be standard karyotypes with \(2n=68/69 + 2-7B \ XX/ \ XY_1 \ Y_2 \ FN=70\), composed of 66 acrocentric autosomes from the E group, with the X chromosome being a medium-sized submetacentric chromosome and the males presenting one small metacentric \(Y_1\) chromosome and one small acrocentric \(Y_2\) chromosome. The male from the western Amazon had a variant karyotype with \(2n=67 + 7B \ XY_1 \ Y_2 \ FN=70\), composed of one pair of large submetacentrics from the A group and 62 acrocentric autosomes from the E group; the
sex chromosomes were the same as in the males from the eastern Amazon. The large submetacentric pair in this specimen showed homology with the 4q and 32p of the standard karyotype. The X chromosome showed an interstitial C-band in the middle of the q arm, and its distal half was homologous to the \( Y_1 \) chromosome, which confirm X-autosome tandem fusion. All chromosomes except the \( Y_1 \) chromosome presented extended pericentromeric C-bands. The NOR regions were seen in autosomes as satellites that were distal to the telomeres of the q arms of the two larger acrocentrics (first and second pairs). Additionally, a number of very small supernumerary or B chromosomes, varying from two to seven chromosomes, were present in all of the animals.

The results showed a basic karyotype of \( 2n=68/69 + 2-6B \) FN=70, with a sex chromosome system of the XX/XY, \( Y_2 \) type for \( M. nemorivaga \). This kind of sex chromosome system has been widely described for several deer species of the subfamily Muntiacini (Fontana and Rubini 1990), but it is rare in the subfamily Capreoleinae; it has been reported only in the red brocket \( M. americana \) from Brazil (Sarria-Perea 2004, Abril et al. 2010). This rearrangement might has been achieved by both species independently, since they belong to two phylogenetically distant clades (Duarte et al. 2008, Gonzalez et al. 2010).

When comparing the karyotype of \( M. nemorivaga \) to the other species of the subfamily Capreoleinae, its standard karyotype seems to be more similar to that of the Eurasian roe deer species \( Capreolus capreolus \) (Linnaeus, 1758; with \( 2n=70, \) FN=72) and \( C. c. pygargus \) (with \( 2n=70+10B, \) FN=72) than to that of other phylogenetically closer South American species such as \( Hippocamelus bisulcus \) (Molina, 1782) (Vila et al. 2010) and \( Pudu puda \) (Molina, 1782), which both possess a \( 2n=70, \) FN=74 karyotype (Fontana and Rubini 1990). The karyotype of \( M. nemorivaga \) differs from the roe deer karyotypes only by X-autosome tandem fusion and this particular sex chromosome system can be considered as an autapomorphic feature, as the product of an independent process of chromosome evolution. This proximity to the karyotype of the roe deer agrees with molecular phylogenetics that arranges \( M. nemorivaga \) as a basal species onto the South American Rangiferini clade (Duarte et al. 2008, Gonzalez et al. 2010). The proximity may also suggest that the ancestral karyotype of the New World Rangiferini is likely to be \( 2n=70, \) FN=72 rather than the \( 2n=70, \) FN=70 karyotype that has previously been reported in the literature (Fontana and Rubini 1990).

The single animal from the western Amazon analyzed here had a different karyotype, which was composed of 64 autosomes, with one pair from the A group. According to its G-banding patterns, this new submetacentric pair was the product of the Robertsonian translocation between the chromosomes 4 and 32 from standard karyotype. The fixation of this kind of rearrangement in some local population increases a probability of reproductive isolation of this population. Such a process has indeed been observed in a sympatric brocket deer species \( M. americana \) (Abril et al. 2010).

These results seem to indicate that \( M. nemorivaga \) has a karyotype similar to the ancient one, but it followed an independent and complex chromosomal evolutionary pathway. The existence of polymorphic karyotypes likely indicate some degree of population differentiation and this can be found in other neotropical brocket deer species (Abril and
Duarte 2008; Abril et al. 2010). Despite the small number of analyzed animals in this study, these are the first results concerning the karyotypic status of the *M. nemorivaga* in Brazil, and they are fundamental for a new review of chromosomal evolution in Cervidae.

**Acknowledgments**

We would like to thank Paulo Antonio Tosta for his aid in laboratory methodologies and FAPESP for financial support. We are grateful to team of NUPECCE (Núcleo de Pesquisa e Conservação de Cervídeos) for their support in this work.

**References**

Abril VV, Duarte JMB (2008) Chromosome polymorphism in the Brazilian dwarf brocket deer, *Mazama nana* (Mammalia, Cervidae). Genetics and Molecular Biology 31:53–57. doi: 10.1590/S1415-47572008000100011

Abril VV, Carnelossi EAG, González S, Duarte JMB (2010) Elucidating the Evolution of the Red Brocket Deer *Mazama americana* Complex (Artiodactyla; Cervidae). Cytogenetic and Genome Research 128(1–3): 177–87. doi: 10.1159/000298819

Colinvaux PA, De Oliveira PE, Bush MB (2000) Amazonian and neotropical plant communities on glacial time-scales: The failure of the aridity and refuge hypotheses. Quaternary Science Reviews 19: 141–169. doi: 10.1016/S0277-3791(99)00059-1

Cuvier GF (1817) Dictionnaire des Sciences Naturelles Zoologie. Mammifères. FG Levraut, Paris 7: 1–534

Duarte JMB, Gonzalez S, Maldonado JE (2008) The surprising evolutionary history of South American deer. Molecular Phylogenetics and Evolution 49: 17–22. doi: 10.1016/j.ympev.2008.07.009

Fontana F, Rubini M (1990) Chromosomal evolution in Cervidae. Biosystems 24: 157–174. doi: 10.1016/0303-2647(90)90008-O

Gonzalez S, Duarte JMB, Maldonado JE (2010) Molecular Phylogenetics and Evolution. In: Duarte JMB, Gonzalez S (Eds) Neotropical cervidology: biology and medicine of Latin American Deer. FUNEP–IUCN, 12–17.

Howell WW, Black DA (1980) Controlled silver staining of nucleous organizer regions with a protective colloidal developer: a 1 step method. Experientia 36: 1014–1015. doi: 10.1007/BF01953855

Jiménez JE (2010) The southern pudu (*Pudu puda*). In: Duarte JMB, González, S (Eds) Neotropical cervidology: biology and medicine of Latin American deer. FUNEP – IUCN, 140–150.

Moorhead PS, Nowell PC, Mellman WJ, Battips DM, Hungerford DA (1960) Chromosome preparations of leukocytes cultured from human peripheral blood. Experimental Cell Research 20: 613–616.

Rossi RV, Bodmer R, Duarte JMB, Trovati RG (2010) Amazonian Brown brocket deer *Mazama nemorivaga* (Cuvier 1817). In: Duarte JMB, Gonzalez S (Eds) Neotropical cervidology: Biology and Medicine of Latin American Deer. FUNEP–IUCN, 202–210.
Sarria-Perea JA (2004) Comparação entre alguns citótipos da espécie *Mazama americana*: Quão grande é a diferença entre eles? Master Dissertation, Jaboticabal, Brasil: Faculdade de Ciências Agrárias e Veterinárias, 51 pp.
Seabright M (1971) A rapid banding technique for human chromosomes. Lancet 11: 971–972. doi: 10.1016/S0140-6736(71)90287-X
Sumner AT (1972) A simple technique of demonstrating centromeric heterochromatin. Experimental Cell Research 75: 304–306. doi: 10.1016/0014-4827(72)90558-7
Verma RS, Babu A (1995) Human chromosomes: principles and techniques. 2.ed. New York, McGraw Hill, 419 pp.
Vila AR, Galvez CES, Aldridge D, Ramilo E, González PC (2010) South Andean huemul *Hippocamelus bisulcusi* (Molina 1782). In: Duarte JMB, Gonzalez S (Eds) Neotropical cervidology: Biology and Medicine of Latin American Deer. FUNEP – IUCN, 89–100.