Ginsenoside Rd ameliorates muscle wasting by suppressing the signal transducer and activator of transcription 3 pathway

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Abstract

Background  The effects of some drugs, aging, cancers, and other diseases can cause muscle wasting. Currently, there are no effective drugs for treating muscle wasting. In this study, the effects of ginsenoside Rd (GRd) on muscle wasting were studied.

Methods  Tumour necrosis factor-alpha (TNF-α)/interferon-gamma (IFN-γ)-induced myotube atrophy in mouse C2C12 and human skeletal myoblasts (HSkM) was evaluated based on cell thickness. Atrophy-related signalling, reactive oxygen species (ROS) level, mitochondrial membrane potential, and mitochondrial number were assessed. GRd (10 mg/kg body weight) was orally administered to aged mice (23–24 months old) and tumour-bearing (Lewis lung carcinoma [LLC1] or CT26) mice for 5 weeks and 16 days, respectively. Body weight, grip strength, inverted hanging time, and muscle weight were assessed. Histological analysis was also performed to assess the effects of GRd. The evolutionary chemical binding similarity (ECBS) approach, molecular docking, Biacore assay, and signal transducer and activator of transcription (STAT) 3 reporter assay were used to identify targets of GRd.

Results  GRd significantly induced hypertrophy in the C2C12 and HSkM myotubes (average diameter 50.8 ± 2.6% and 49.9% ± 3.7% higher at 100 nM, vs. control, P ≤ 0.001). GRd treatment ameliorated aging- and cancer-induced (LLC1 or CT26) muscle atrophy in mice, which was evidenced by significant increases in grip strength, hanging time, muscle mass, and muscle tissue cross-sectional area (1.3-fold to 4.6-fold, vs. vehicle, P ≤ 0.05; P ≤ 0.01; P ≤ 0.001). STAT3 was found to be a possible target of GRd by the ECBS approach and molecular docking assay. Validation of direct interaction between GRd and STAT3 was confirmed through Biacore analysis. GRd also inhibited STAT3 phosphorylation and STAT3 reporter activity, which led to the inhibition of STAT3 nuclear translocation and the suppression of downstream targets of STAT3, such as atrogin-1, muscle-specific RING finger protein (MuRF-1), and myostatin (MSTN) (29.0 ± 11.2% to 84.3 ± 30.5%, vs. vehicle, P ≤ 0.05; P ≤ 0.01; P ≤ 0.001). Additionally, GRd scavenged ROS (91.7 ± 1.4% reduction at 1 nM, vs. vehicle, P ≤ 0.001), inhibited TNF-α-induced dysregulation of ROS level, and improved mitochondrial integrity (P ≤ 0.05; P ≤ 0.01; P ≤ 0.001).

Conclusions  GRd ameliorates aging- and cancer-induced muscle wasting. Our findings suggest that GRd may be a novel therapeutic agent or adjuvant for reversing muscle wasting.

Keywords  cachexia; ginsenoside Rd; muscle wasting; sarcopenia; signal transducer and activator of transcription 3

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Introduction

Myotoxic drugs, aging, cancers, and other diseases, such as chronic kidney disease and heart failure, can cause muscle wasting. Sarcopenia is a multifactorial syndrome in geriatric patients that is associated with a sedentary lifestyle, oxidative stress, mitochondrial integrity, altered proteostasis, and inflammation. Cachexia is a wasting syndrome caused by underlying illness and associated with anorexia, inflammation, insulin resistance, and increased muscle protein breakdown. Cancer cachexia is the term used to refer to muscle atrophy that occurs in cancer patients. It is characterized by a significant increase in the levels of pro-inflammatory cytokines, such as TNF-α, IFN-γ, and interleukin-6 (IL-6). Approximately 80% of cancer patients are affected by cachexia, which leads to a poor prognosis and accounts for 20%–30% of cancer-associated mortality. Muscle wasting decreases the quality and duration of life; therefore, it is essential to develop novel therapeutic strategies against it. Currently, there are no effective therapeutics that have a protective effect against muscle atrophy.

Balancing myofibrillar protein synthesis and breakdown is essential in maintaining muscle mass. Several signal transduction pathways, including the AKT/mammalian target of rapamycin (mTOR), unfolded protein response, FoxO, Janus kinase (JAK)/signal transducer and activator of transcription (STAT), and ubiquitin-proteasome system (UPS) pathways, are involved in muscle hypertrophy and wasting. Muscle hypertrophy is associated with an increase in muscle mass, and it is the result of an increase in the size of pre-existing skeletal muscle fibres due to up-regulated protein synthesis without an apparent variation in myofibre number. The mTOR pathway is a key driver of protein synthesis; therefore, its inhibition by therapeutic drugs results in muscle atrophy, which is associated with shrinkage of myofibres and involves a shift toward decreased protein synthesis and increased protein degradation. It is reported that protein degradation is driven by the autophagy-lysosome pathway, two ubiquitin E3 ligases (muscle atrophy F-box [atrogin-1] and muscle RING finger [MuRF-1]), and the activation of proteasome degradation. In cancer patients, the IL-6-JAK/STAT3 pathway is highly up-regulated and associated with muscle atrophy. Additionally, JAK/STAT signalling is highly expressed in old mice, and its inhibition leads to the recovery of muscle function. Furthermore, STAT3 phosphorylation leads to the activation of atrogin-1, MuRF-1, and MSTN in cancer patients with cachexia. Mitochondrial quality and function are impaired during oxidative stress and in cancer. This leads to excessive production of reactive oxygen species (ROS) and contributes to muscle wasting. STAT3 and ROS are important in muscle-wasting conditions, which makes them potential targets in the treatment of such conditions.

*Panax ginseng* C. A. Meyer (*Panax ginseng*) has been shown to improve muscle strength and reduce muscle damage in mice. Therefore, we hypothesized that ginseng-derived components might have a protective effect against muscle atrophy. Thus, we screened ginseng components and found that GRd has a protective effect against muscle atrophy (Figure S1). It has been previously shown that GRd inhibits epidermal growth factor receptor (EGFR) signalling and suppresses cancer stem cell—like properties in colorectal cancer (CRC) cells, which result in the reduction of CRC metastasis. Similarly, the anti-tumour activities of GRd in several other cancer types have been reported; however, the effect of GRd on muscle wasting is currently unknown. In this study, we showed that GRd exhibits a muscle anti-atrophic effect in C2C12 cells and HSkMs. Furthermore, GRd efficiently protected against muscle atrophy in mice with cancer cachexia and aging-induced atrophy.

Materials and methods

Reagents/cell cultures

Ginsenoside Rd (purity ≥98%) was purchased from ChemFaces (Wuhan, China), and other ginseng-derived components were obtained from Korean Ginseng Corporation (Daejeon, Korea). Mouse C2C12 myoblast and Lewis lung carcinoma (LLC1) were obtained from ATCC (ATCC® CRL-1772™ and ATCC® CRL-1642™, respectively). CT26 and human skeletal muscle (HSKM) were purchased from Korean Cell Line Bank (80009, KCLB) and Gibco™ (A12555, Gibco™), respectively. Other detailed information is provided in Supplementary Materials and Methods.

Animal study

All animal experiments were approved by the Soonchunhyang University Animal Care and Use Committee and maintained in a pathogen-free environment with 12 h light–dark cycles and fed with regular normal chow. For the sarcopenia experiment, C57BL/6 male mice (3- to 4-month-old mice, n = 8, control adults) or (23- to 24-month-old mice, n = 16, old) were orally administered either vehicle (0.9% NaCl) or GRd (10 mg/kg BW) daily for 36 days (5 weeks). For the cancer cachexia experiment, LLC1 and CT26 cells (2.0 × 10⁶ cells/mouse) were injected subcutaneously (s.c.) in the right hind flank into male C57BL/6 and BALB/c mice (10-week-old, respectively (n = 12). When tumours were palpable on day 6, the mice were blindly and randomly divided into two groups and orally administered either vehicle or GRd (10 mg/kg BW) daily for 16 days. Tumour volume was calculated with the formula 0.52*(length*width²), and the measurement was performed using a digital calliper. The grip strength was measured every 7 and 3 days for sarcopenia and cancer cachexia, respectively, using the grip strength meter.
(Ugo Basile, Italy). The hanging test was performed using Kondziela’s inverted screen test.

**STAT3 reporter assay**

STAT3 reporter assay was described previously with slight modifications. The plasmid used for STAT3 reporter is pRRL.sin-18.ppt. STAT3-GFP.pre (STAT3-GFP, Addgene plasmid #110495).

**Binding target prediction of GRd**

The evolutionary chemical binding similarity (ECBS) method, a machine learning model originally developed to predict chemical binding similarity, was applied to predict the potential targets of GRd. Among the variants of the ECBS model, the target-specific ensemble ECBS (TS-ensECBS) model was used to screen binding targets because of its high accuracy. A collection of TS-ensECBS models generated for a total of 7690 targets enabled comprehensive target-binding predictions for GRd. The predicted target scores were analysed to estimate the potential binding targets of GRd (Table S1).

**Biacore assay**

Biacore assay was performed with a Biacore 2000 (GE Healthcare, USA); 100 mM NHS and 200 mM EDC were mixed in equal volumes and injected into a closed HC1000M chip (XanTec Bioanalytics) at a rate of 30 μL/min.

**Statistical analysis**

All data were performed with two to five independent replicates (as indicated in the figure legends) and the results are presented as mean ± SEM (standard error mean) and analysed by GraphPad Prism software. The experimental groups were compared using one-way or two-way analysis of variance (ANOVA) test followed by Tukey’s multiple comparisons test, as indicated. The main effects of GRd treatment, time, and an interaction were determined with Mixed-effects model (REML). The main effects model was used to assess the trajectory of GRd across treatments, while the interaction model was used to assess the differential effect of time based on treatment received, with a focus on the factors of time, treatment received, and the interaction between time and treatment received. The mixed model takes into account the correlation between repeated measures taken within a subject and makes use of all available data on each subject. P-values < 0.05 were considered statistically significant.

**GRd ameliorates myotube atrophy in HSkMs**

GRd ameliorated atrophy in the mouse C2C12 myotubes; therefore, we evaluated the anti-atrophic effects of GRd in human myotubes. Myotube atrophy in HSkMs was induced with TNF-α, after which the effect of GRd on myotube diameter was investigated. TNF-α induced myotube atrophy, which was evidenced by a smaller myotube diameter (Figure 1A, D). Consistent with the results obtained from the tests performed on mouse C2C12 myotubes, it was found that myotube diameter was much larger following treatment with GRd than with TNF-α alone (Figure 1C, D). To confirm this in an independent method, HSkM was treated with HT29-derived conditioned media. The decreased diameter of HskM myotubes with HT29-conditioned media was rescued by GRd (Figure S1C, D). These data indicate that GRd ameliorates TNF-α or HT29-conditioned media-induced atrophy in human myotubes.

**Results**

**GRd protects C2C12 myotubes against TNF-α/IFN-γ (TI)-induced myotube atrophy**

We sought to identify ginseng-derived components that have an in vitro protective effect against TI-induced muscle wasting. After 4 days of differentiation, C2C12 myotubes were treated with ginseng-derived components together with TI. As expected, myotubes diameter was decreased following the treatment with TI. Interestingly, GRd had the highest anti-atrophic effect among the ginseng components tested in TI-treated C2C12 myotubes, as evidenced by the hypertrophy in GRd-treated C2C12 myotubes compared to that in the other groups (Figure S1A, B). Thus, we focused on GRd and determined its dose–response effect against myotube atrophy. C2C12 myotubes were treated with several concentrations of GRd (10–1000 nM) with or without TI, after which myotube atrophy was determined. The results showed that GRd significantly increased the diameter of C2C12 myotubes in a dose-dependent manner (Figure 1A, B). The decreased diameter of C2C12 myotubes treated with conditioned media obtained from CT26 or LLC1 was consistently increased by GRd (Figure S1C, D), which suggests that GRd can protect against pathological muscle wasting.

**Additional methods**

Additional details regarding the methods and materials are provided in the Supplementary Materials and Methods.
Based on the observed anti-atrophic effect of GRd on myotubes *in vitro*, we investigated whether GRd protects against muscle wasting *in vivo*. The effects of GRd on muscle wasting were assessed in aged mice (23–24 months old) and control adult mice (4 months old). The animals were treated with vehicle or GRd daily for 5 weeks (*Figure 2A*). As expected, the aged mice exhibited lower grip strength than the control adult mice did; however, GRd significantly improved grip strength (*Figure 2B*; main effect of GRd treatment, \( P < 0.001 \), although body weight was not affected (*Figure S2A*). The grip strength of GRd treatment increased over time (*Figure 2B*; main effect of time, \( P = 0.0034 \)). To further corroborate the results of the grip strength test, a hanging test was performed using Kondziela’s inverted screen. Hanging time was shorter among the aged mice than among the control mice but was significantly improved by GRd (*Figure 2C*). Interestingly, the weights of the gastrocnemius (GA), soleus (SOL), and extensor digitorum longus (EDL) muscles were significantly higher in the GRd-treated group than in the control old mice. However, there was no difference in the weight of the tibialis anterior (TA) muscle between the groups (*Figure 2D,E*). Furthermore, liver, spleen, and lung weights were not different between the vehicle- and GRd-treated mice; however, heart weight was significantly lower in the GRd-treated mice than in the control old mice (*Figure 2B*). To evaluate differences in muscle tissue between groups, haematoxylin and eosin (H&E) staining was carried out, after which the cross-sectional area (CSA) of the GA muscle fibre was determined. The GA muscle CSA was much smaller in the aged group than in the control group (*Figure 2F*); however, it was significantly higher in the GRd-treated group than in the vehicle-treated group. The average CSA in the GRd-treated group was 33.83% higher than that in the control mice.
GRd in muscle wasting

vehicle-treated group but similar to that in the control group (Figure 2F). These results indicate that GRd can improve muscle mass and function in aged mice.

**GRd ameliorates Lewis lung carcinoma (LLC1)- and CT26-induced cancer cachexia in vivo**

Cancer-induced cachexia is characterized by loss of skeletal muscle in patients; therefore, we determined whether GRd can protect against muscle loss in a mouse model of LLC1-induced cancer cachexia. LLC1 cells were subcutaneously injected into the flanks of C57BL/6 mice, after which the mice were orally administered GRd daily for 16 days starting from day 6 when the tumours were palpable. Grip strength was determined every 3 days during the treatment period, and hanging ability was evaluated 1 day before the animals were sacrificed (Figure 3A). Decreases in body weight and muscle weight as well as low grip strength and short hanging time were indicative of the successful establishment of LLC1-induced cancer cachexia in tumour-bearing mice (Figures 3B,C and S3A). Tumour sizes were smaller in GRd-treated mice than in control mice at the tested concentrations, which indicated the anti-tumour effect of GRd
Interestingly, GRd noticeably ameliorated body wasting, muscle atrophy, weakness, and loss of strength, which are features of cancer cachexia (Figures 3B–E and S3A). Body weight was 11.28% lower in the vehicle-treated tumour-bearing group (LLC1 + vehicle) than in the healthy control group. However, it was found that GRd (LLC1 + GRd) ameliorated body weight loss (Figure 3B). Moreover, grip strength progressively decreased over time in the vehicle-treated tumour-bearing group but was significantly improved by GRd from day 6 (Figure 3C; main effect of treatment, \( P < 0.001 \); the main effect of time, \( P < 0.001 \); interaction, \( P < 0.001 \)), and hanging test was conducted on day 20. (D, E) Skeletal muscles, including GA, SOL, TA, and EDL, were dissected after 21 days, and the weights were determined. Representative images of skeletal muscles were shown. (F) GA muscles were stained with H&E staining, and representative images were shown. The average cross-sectional area (CSA) of GA muscle fibre was quantified by ImageJ and plotted depending on the frequency as indicated. The data were shown as mean ± SEM. One-way (B, C, E, F) or two-way (C, F) ANOVA followed by Tukey’s multiple comparisons were used to compare between data (ns, not significant; \( ^{*} P \leq 0.05 \), \( ^{**} P \leq 0.01 \), \( ^{***} P \leq 0.001 \), control health vs. LLC1 + vehicle; \( ^{*} P \leq 0.05 \), \( ^{**} P \leq 0.01 \), \( ^{***} P \leq 0.001 \), control health vs. LLC1 + GRd; \( ^{\&} P \leq 0.05 \), \( ^{\&\&} P \leq 0.01 \), \( ^{\&\&\&} P \leq 0.001 \), LLC1 + vehicle vs. LLC1 + GRd).

(Figure S3B). Heart, lung, kidney, liver, and spleen weights were not different between the vehicle- and GRd-treated groups (Figure S3C). H&E staining was performed on GA muscles excised from the treated mice. It was found that, compared to the healthy control mice, the vehicle-treated tumour-bearing mice had smaller muscle fibres with smaller average CSAs (Figure 3F). However, GRd protected myofibres from the decrease in CSA (Figure 3F).

A mouse model of CT26 CRC-induced cachexia was also used in the study to validate the therapeutic efficacy of GRd in different cancer cachexia models. BALB/c mice were subcutaneously injected with CT26 cells and administered GRd or vehicle (Figure 4A). The results were similar to those

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**Figure 3** GRd ameliorates Lewis lung carcinoma (LLC1)-induced cancer cachexia in vivo. (A) Experiment design of LLC1-induced cancer cachexia. C57BL/6 mice were injected subcutaneously (s.c.) with LLC1 cells. Then, after 6 days of injection, mice were orally administrated with GRd (10 mg/kg body weight/day) or vehicle daily for 16 days. Mice without tumour inoculation was used as control health. Control (\( n = 6 \)), LLC1 + vehicle (\( n = 6 \)), and LLC1 + GRd 10 mg/kg (\( n = 6 \)). (B) Tumor-free body weight was measured in the day 21. (C) Muscle performance for grip strength was evaluated every 3 days (the main effect of treatment, \( P < 0.001 \); the main effect of time, \( P < 0.001 \); interaction, \( P < 0.001 \)), and hanging test was conducted on day 20. (D, E) Skeletal muscles, including GA, SOL, TA, and EDL, were dissected after 21 days, and the weights were determined. Representative images of skeletal muscles were shown. (F) GA muscles were stained with H&E staining, and representative images were shown. The average cross-sectional area (CSA) of GA muscle fibre was quantified by ImageJ and plotted depending on the frequency as indicated. The data were shown as mean ± SEM. One-way (B, C, E, F) or two-way (C, F) ANOVA followed by Tukey’s multiple comparisons were used to compare between data (ns, not significant; \( ^{*} P \leq 0.05 \), \( ^{**} P \leq 0.01 \), \( ^{***} P \leq 0.001 \), control health vs. LLC1 + vehicle; \( ^{*} P \leq 0.05 \), \( ^{**} P \leq 0.01 \), \( ^{***} P \leq 0.001 \), control health vs. LLC1 + GRd; \( ^{\&} P \leq 0.05 \), \( ^{\&\&} P \leq 0.01 \), \( ^{\&\&\&} P \leq 0.001 \), LLC1 + vehicle vs. LLC1 + GRd).

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obtained for the other cancer cachexia model. Several features of CT26 cancer-induced cachexia, such as loss of grip strength, weakness, and muscle atrophy, were partially ameliorated by GRd (Figures 4B–F and S3D–F; main effect of GRd treatment, \( P < 0.001 \)). Collectively, these results provide evidence that GRd can prevent cancer-induced muscle wasting in vivo.

**GRd interacts with STAT3**

An evolutionary chemical binding similarity (ECBS) approach, which was previously developed based on a classification similarity-learning framework, was used to identify the molecular targets of GRd.\(^9\) The binding targets of GRd were predicted using a collection of target-specific ensemble models generated for 7690 targets.\(^10\) The predicted binding scores for each target were used to prioritize potential binding target candidates (Table S1). STAT3 was found to be one of the top-ranked candidates. It is of particular interest as it is an important mediator of muscle wasting and is considered a drug target in muscle atrophy treatment. To independently validate the ECBS findings, we conducted a molecular docking study to investigate the interaction between GRd and STAT3. The results suggested that GRd had favourable molecular interactions with STAT3 at its binding sites.

**Figure 4** GRd ameliorates (CT26)-induced cancer cachexia in vivo. (A) Experiment design of CT26-induced cancer cachexia. BALB/c mice were injected subcutaneously with CT26 cells. Then, after 6 days of injection, mice were orally administrated with GRd (10 mg/kg body weight/day) or vehicle daily for 16 days. Mice without tumour inoculation was used as control health. Control (\( n = 6 \)), CT26 + vehicle (\( n = 6 \)), and CT26 + GRd 10 mg/kg (\( n = 6 \)). (B) Tumor free body weight was measured in the day 21. (C) Muscle performance for grip strength was evaluated every 3 days (the main effect of treatment, \( P < 0.001 \); the main effect of time, \( P < 0.001 \); interaction, \( P < 0.001 \)), and hanging test was conducted on day 20. (D, E) Skeletal muscles, including GA, SOL, TA, and EDL were dissected after 21 days, and the weights were determined. Representative images of skeletal muscles were shown. (F) GA muscles were stained with H&E staining, and representative images were shown. The average cross-sectional area (CSA) of GA muscle fibre was quantified by ImageJ and plotted depending on the frequency as indicated. The data were shown as mean ± SEM. One-way (B, C, E, F) or two-way (C, F) ANOVA followed by Tukey’s multiple comparisons were used to compare between data (ns, not significant; \( ^* P \leq 0.05 \), \( ^{**} P \leq 0.01 \), \( ^{***} P \leq 0.001 \), control health vs. CT26 + vehicle. \( ^{\dag} P \leq 0.05 \), \( ^{\ddag} P \leq 0.01 \), \( ^{\ddagger} P \leq 0.001 \), control health vs. CT26 + GRd. \( ^{\&} P \leq 0.05 \), \( ^{\&&} P \leq 0.01 \), \( ^{&&} P \leq 0.001 \), CT26 + vehicle vs. CT26 + GRd). 

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ing energy was found to be $-8.1$ kcal/mol (Figure 5A). In the docking model, the sugar moiety in GRd primarily interacted with negatively charged residues of STAT3, such as Lys591 and Arg609, whereas the steroid backbone had hydrophobic interactions with Phe716, Tyr640, and Lys626 (Figure 5A). Direct binding between GRd and STAT3 was experimentally confirmed through Biacore analysis, in which STAT3 was immobilized on a chip and GRd was injected to determine its affinity for STAT3. The results indicated that GRd was bound to STAT3 with a dissociation constant (Kd) of 36.3 $\mu$M (Figure 5B). A STAT3 reporter assay was also performed to verify the interaction between GRd and STAT3. In the assay, the level of green fluorescent protein (GFP), which is a surrogate marker of STAT3, depends on STAT3 transcriptional activity since the reporter has four STAT3 binding sites.18 C2C12 cells were transduced with a lentiviral STAT3 reporter and then treated with TI in the presence or absence of GRd (Figure 5C). As expected, the GFP level was 1.422-fold higher following treatment with TI than it was following the control treatment; however, it was significantly reduced by GRd (Figure 5C), suggesting that GRd regulates STAT3 activity.

**GRd regulates the STAT3 pathway by suppressing the phosphorylation and nuclear localization of STAT3**

Active STAT3 is phosphorylated during muscle wasting21–24; therefore, we investigated whether GRd changes the level of phosphorylated STAT3 in vitro. Interestingly, TNF-α or TI-induced up-regulation of STAT3 phosphorylation was significantly reduced by GRd (Figure 5D). In a further study, we investigated the lower levels of pSTAT3 in vivo by analysing muscle samples obtained from aged mice and cancer (LLC1 or CT26)-bearing mice. Consistently, the pSTAT3 level was much higher in these mice than in the control mice but was significantly reduced by GRd (Figure 5E,F). STAT3 nuclear translocation was also evaluated since activated STAT3 is localized within nuclei.23,24 C2C12 cells were treated with TI in the absence or presence of GRd, after which STAT3 localization was determined using a confocal microscope. It was found that STAT3 level was highly up-regulated and that STAT3 was localized within the nucleus following treatment with TI; however, these effects were inhibited by GRd.

**Figure 5** GRd inhibits muscle wasting through direct binding with STAT3. (A) Molecular docking between GRd and STAT3. (B) Biacore analysis was used to analyse the binding ability of GRd to STAT3. (C) C2C12 cells were transduced with STAT3 reporter, and then treated for 8 h with TNF-α at 20 ng/mL and IFN-γ at 100 U/mL (TI) in the presence or absence of GRd (100 nM). STAT3 reporter expressions were measured by flow cytometry. (D) C2C12 myotubes were treated for 24 h with TNF-α at 20 ng/mL and IFN-γ at 100 U/mL in the presence or absence of GRd (100 nM) as indicated (n = 4). Then, the protein levels of pSTAT3/TSTAT3 were evaluated by western blot. Representative images were shown and images were measured by ImageJ software. The data were shown as mean ± SEM. (E, F) GA tissue lysates were isolated from aged mice (n = 3), LLC1 (n = 6), or CT26-implanted mice (n = 6). Then, the protein levels of pSTAT3/TSTAT3 were evaluated by western blot. Representative images were shown and images were measured by ImageJ software. The data were shown as mean ± SEM. One-way (C–F) ANOVA followed by Tukey’s multiple comparisons were used to compare between data (*$P \leq 0.05$; **$P \leq 0.01$; ***$P \leq 0.001$).
Our findings were also confirmed on muscle tissues excised from aged mice and cancer-bearing mice (Figure 6B–D). Taken together, the current findings suggest that GRd binds to STAT3, resulting in the suppression of STAT3 phosphorylation and nuclear translocation.

MSTN and muscle-specific E3 ubiquitin ligases, MuRF-1 and atrogin-1, are central mediators of muscle wasting and their expression is induced following STAT3 activation. Thus, we hypothesized that these molecules might be affected by GRd. To test this, C2C12 cells were treated with TNF-α alone or with the TI combination in the absence or presence of GRd. The levels of MuRF-1, atrogin-1, and MSTN were then determined via immunoblotting. As expected, the levels of all three proteins were increased by TNF-α alone or TI but significantly reduced by GRd (Figure 7A). To verify these changes in animal models, muscle-derived protein lysates obtained from aged mice and cancer (LLC1 or CT26)-bearing mice were analyzed. Consistently, GRd significantly inhibited the expression of MuRF-1, atrogin-1, and MSTN in these groups of mice (Figure 7B–D).

**GRd reduces ROS levels and protects mitochondrial integrity**

STAT3-induced ROS accumulation in muscle wasting was observed and linked to compromised mitochondrial integrity. Thus, we hypothesized that GRd may reduce ROS levels by suppressing STAT3 activation and MSTN expression. To test this, we evaluated GRd-induced changes in total ROS level in C2C12 cells by flow cytometry and fluorescence microscopy. As expected, cellular ROS level was increased by TNF-α but significantly reduced by GRd (Figures 8A and S4). Interestingly, GRd had a strong ROS scavenging activity. The results showed that GRd decreased the levels of free radicals by 92.72% even at a concentration of 1 nM (Figure 8B). Thus, we also determined the effect of GRd on mitochondrial ROS levels using the MitoSox™ Red Mitochondrial Superoxide Indicator. Mitochondrial ROS level was also decreased by GRd, which was consistent with the effect of GRd on total ROS level (Figure 8C). Higher levels of ROS are associated with a reduction in mitochondrial function, which is indicated by a decrease in mito-

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**Figure 6** GRd inhibits STAT3 nuclear translocalization. (A) C2C12 myotubes were incubated for 30 min with TI ± GRd (100 nM) as indicated and stained with antibodies against pSTAT3 (green) and counterstained with DAPI (blue) to determine the localization of STAT3 in the nucleus. (B–D) Cryosections of GA muscles from aged mice, LLC1, and CT26-implanted mice were stained with antibodies against pSTAT3 (green) and counterstained with DAPI (blue). Representative images were shown. Arrows indicate nuclear localization of STAT3.
Chondrial membrane potential ($\Delta \Psi_m$). Thus, we assessed $\Delta \Psi_m$ and mitochondrial number using MitoProbe DilC1(5) assay kit and MitoTracker™ Orange, respectively. The results showed that $\Delta \Psi_m$ and mitochondrial number in C2C12 cells decreased following treatment with TNF-$\alpha$; however, this was partially inhibited by GRd (Figure 8D,E). Additionally, mitochondrial DNA (mtDNA) copy number, an indicator of mitochondrial biogenesis level, was decreased by TNF-$\alpha$ but partially increased by GRd (Figure 8F). These results indicate that GRd reduces ROS level and protects mitochondrial integrity via ROS scavenging and suppression of STAT3 signalling, thereby conferring protection against muscle wasting.

Figure 7 GRd suppresses the downstream pathways of STAT3. (A) C2C12 myotubes were treated for 24 h with TNF-$\alpha$ at 20 ng/mL, IFN-$\gamma$ at 100 U/mL or GRd (100 nM) as indicated, and then protein levels of muscle atrophy-related genes were evaluated by western blot. The data were shown as mean ± SEM of three to four independent experiments. (B–D) GA tissue lysates were isolated from aged mice-, LLC1-, and CT26-implanted mice, respectively. Then, the protein levels of muscle atrophy-related genes were evaluated by western blot. Representative images were shown, and images were measured by ImageJ software. The data were shown as mean ± SEM ($n = 6$). One-way (A–D) ANOVA followed by Tukey’s multiple comparisons were used to compare between data (*$P \leq 0.05$; **$P \leq 0.01$; ***$P \leq 0.001$).

Thus, we assessed $\Delta \Psi_m$ and mitochondrial number using MitoProbe DilC1(5) assay kit and MitoTracker™ Orange, respectively. The results showed that $\Delta \Psi_m$ and mitochondrial number in C2C12 cells decreased following treatment with TNF-$\alpha$; however, this was partially inhibited by GRd (Figure 8D,E). Additionally, mitochondrial DNA (mtDNA) copy number, an indicator of mitochondrial biogenesis level, was decreased by TNF-$\alpha$ but partially increased by GRd (Figure 8F). These results indicate that GRd reduces ROS level and protects mitochondrial integrity via ROS scavenging and suppression of STAT3 signalling, thereby conferring protection against muscle wasting.
Muscle atrophy is associated with various conditions such as cancers, aging, and chronic diseases, including diabetes, chronic kidney diseases, and chronic obstructive pulmonary disease. It has major negative effects on the quality of life and contributes to a high rate of mortality. In this study, we showed that GRd has the potential to alleviate muscle wasting in pathophysiological conditions such as cancer and sarcopenia. GRd exerted strong anti-atrophic effects in both

Discussion

Muscle atrophy is associated with various conditions such as cancers, aging, and chronic diseases, including diabetes, chronic kidney diseases, and chronic obstructive pulmonary disease. It has major negative effects on the quality of life and contributes to a high rate of mortality. In this study, we showed that GRd has the potential to alleviate muscle wasting in pathophysiological conditions such as cancer and sarcopenia. GRd exerted strong anti-atrophic effects in both
C2C12 and HSkM myotubes. Furthermore, GRd treatment protects against cancer-induced (LLC1 and CT26) and aging-induced muscle atrophy following oral administration to mice, which was evidenced by increases in grip strength and CSA of muscle fibres. The results showed that, mechanistically, GRd binds to STAT3 and reduces its phosphorylation, thereby suppressing the expression of several atrophy-inducing factors such as MSTN, MuRF-1, and atrogin-1. Therefore, the STAT3 pathway is a major pathway that GRd suppresses to protect against muscle atrophy.

IL-6/JAK/STAT3 signalling plays an important role in muscle wasting by regulating inflammatory responses. 27 It is reported that STAT3 phosphorylation results in an increase in MSTN, atrogin-1, and MuRF-1 expression through the activation of the CCAAT/enhancer-binding protein delta. 25 Moreover, genetic ablation of STAT3 in a mouse model results in partial amelioration of muscle wasting under the induction of diabetes, chronic kidney disease, and cachexia. 25 Similarly, in a previous study, pharmacological inhibition of STAT3 with STAT3 inhibitors, such as AG490 (JAK2 inhibitor) and sunitinib, led to the amelioration of muscle wasting in several muscle wasting models. 28 In the present study, an ECBS approach and molecular docking analysis were used to predict that STAT3 is a target of GRd. Then, we performed Biacore analysis and experimentally confirmed the interaction between STAT3 and GRd. The results showed that, mechanistically, GRd binds to STAT3 and reduces its phosphorylation, thereby suppressing the expression of several atrophy-inducing factors such as MSTN, MuRF-1, and atrogin-1. Therefore, the STAT3 pathway is a major pathway that GRd suppresses to protect against muscle atrophy. Further, the results showed that aging- and cancer-induced muscle atrophy was effectively suppressed by GRd through the suppression of STAT3 phosphorylation and nuclear translocation.

It was also shown that IL-6 binding to its receptor induces STAT3 phosphorylation, which further leads to skeletal muscle proteolysis and muscle wasting. 29 Muscle wasting factors, such as MSTN, ROS, activin, and growth differentiation factor 15, as well as inflammatory cytokines, such as TNF-α, IFN-γ, and IL-6, promote consistent STAT3 activation, catabolism by activating UPS, and the autophagy-lysosome system. 30 Further, the increased circulatory proinflammatory cytokines IL-6, TNF-α, and TGF-β are associated with decreased mitochondrial biogenesis and function, and an imbalance of mitochondrial dynamics. 31 TNF-α and H2O2 are potent inducers of MSTN, which triggers ROS generation. Thus, MSTN and TNF-α form a feed-forward loop that induces muscle wasting through proteasomal-mediated catabolism. 32 In the present study, it was found that GRd scavenges ROS and inhibits STAT3 phosphorylation, which results in a decrease in ROS level and down-regulation of MSTN expression. Further, ROS-induced decreases in ΔΨm, mitochondrial number, and mtDNA copy number were significantly inhibited by GRd, suggesting that GRd is a potent inhibitor of proteasomal catabolism via the STAT3 pathway. Thus, we propose for the first time that GRd binds to STAT3, reducing the muscle wasting activity of STAT3 by binding to it. However, it is also plausible that GRd might exert its activity through other mechanisms, besides STAT3. Recently, Tang et al. demonstrated that GRd treatment enhanced AMP-activated protein kinase (AMPK)/Sirtuin 1 (SIRT1) interaction, leading to the reduction of oxidative stress, mitochondrial dysfunction, and apoptosis in diabetic retinopathy condition. 33 As SIRT1 was shown to prevent muscle dysfunction, 34 it could be possible that the anti-atrophic effects of GRd might be associated with SIRT1 activation, which warrants further study.

In the Orient, particularly in Korea and China, ginseng roots and root extracts have been traditionally used for many centuries as medicine to revitalize the body and mind, prevent aging, and increase vigour. The major active components of *Panax ginseng* are ginsenosides, polysaccharides, volatile oils, glycopeptides, amino acids, and vitamins. There are major ginsenosides (*Rb1*, *Rb2*, *Rc*, *Rd*, *Re*, and *Rg1*) which comprise more than 80% of the total ginsenosides, and minor ginsenosides (*F1*, *F2*, *Rg3*, *Rh1*, *Rh2*, compound *Y*, compound *Mc*, and compound *K*) that are present at a low concentration in ginseng. 35 GRd has been shown to have anti-cancer effects against gastric cancer. 36 Additionally, by counteracting the effects of miR-18a and down-regulating Smad2 expression, GRd suppresses the metastasis of breast cancer. 37 Recently, it was reported that GRd inhibits the metastasis of CRC cells to the liver, lungs, and kidneys in mice. 38 Consistently, it was found in the present study that GRd reduced tumour volumes in the mice with LLC1- and CT26-induced cachexia. Together with the findings that GRd could protect C2C12 and HSkM against myotube atrophy, we rationalize that the mechanisms by which GRd exerts its effect in cancer-induced cachexia might be associated with dual effects, including reducing cancer burden and protecting against muscle atrophy, which warrants further studies. Several ginsenosides (*Rb1* and *Rb2*) show protective effects against muscle wasting via the AKT signalling pathway. 38 However, the effects of GRd on TI-induced cellular atrophy were better than those of other ginseng components in our studies. GRd has already been clinically used to treat acute ischaemic stroke and has been shown to have fewer side effects compared to glucocorticoids, indicating that it can safely improve patient outcomes following ischaemic stroke. 39 Thus, we propose that GRd should be clinically tested for its effect against muscle atrophy.

**Conclusions**

In this study, we found that GRd protects against aging- and cancer-induced skeletal muscle wasting by binding to STAT3, enhancing muscle function, and suppressing the expression of...
of protein degradation factors such as MSTN, atrogin-1, and MuRF-1. Additionally, GRd reduces ROS levels and protects mitochondrial integrity (Figure 5S). Taken together, our findings show that GRd may be a suitable therapeutic agent for suppressing muscle atrophy under various pathophysiological conditions.

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