Atypical haemolytic uraemic syndrome associated with a CD46 mutation triggered by Shigella flexneri

Vicky Brocklebank1, Edwin K.S. Wong2, Rick Fielding1, Timothy H.J. Goodship1,2 and David Kavanagh1,2

1Renal Services Centre, Freeman Hospital, Newcastle upon Tyne, UK and 2The Institute of Genetic Medicine, Newcastle University, Newcastle upon Tyne, UK

Correspondence and offprint requests to: David Kavanagh; E-mail: david.kavanagh@ncl.ac.uk

Abstract
We present a case of haemolytic uraemic syndrome (HUS) triggered by Shigella flexneri. Of the Shigella species, only S. dysenteriae type 1 is said to produce Shiga toxin and consequently cause HUS. Investigation of the complement system in this patient revealed a CD46 mutation. In individuals with mutations in complement genes incomplete penetrance of atypical HUS (aHUS) is seen, suggesting that a trigger, such as infection, is required for disease to manifest. In an era of complement modulatory therapy for aHUS it is important to be alert to unusual presentations of diarrhoeal-associated disease.

Keywords: CD46; complement; haemolytic uraemic syndrome; shiga toxin; Shigella

Introduction
Haemolytic uraemic syndrome (HUS) is characterized by the clinical triad of microangiopathic haemolytic anaemia, thrombocytopenia and acute kidney injury (AKI). The typical form is more common and is caused by enteric infection with Shiga-toxin (Stx)-producing bacteria [1]. The most frequent pathogens responsible are Stx-producing Escherichia coli, particularly serotype O157:H7 (more recently O104:H4 [2]) and Shigella dysenteriae type 1 [1].

The non-shiga-toxin-associated form of HUS or atypical HUS (aHUS) is, in the majority of individuals, associated with inherited and/or acquired defects in the complement system resulting in complement over-activation [3]. Increased understanding of the pathogenesis of disease has led to the successful introduction of the complement inhibitor eculizumab in the treatment of aHUS [4].

Although there are anecdotal reports of eculizumab use in Stx-associated HUS [5], the lack of evidence from a controlled trial leaves clinicians uncertain as to its efficacy and therefore it is not routinely used in Stx-HUS.

We report a case of HUS associated with Shigella flexneri diarrhoea where subsequent investigation revealed a CD46 mutation.

Case report
A 38-year-old woman was referred by her primary care physician following return from a visit to Holland where she had experienced an episode of AKI. She had attended the emergency department with a 2-day history of watery diarrhea and vomiting. She was found to have AKI with a serum creatinine of 663 μmol/L. Admission haemoglobin was 10.9 g/dL, and platelet count was 113 × 10⁹/L with schistocytes on blood film. The serum lactate dehydrogenase (LDH) was raised at 1346 u/L and coagulation profile was normal.

Her past medical history consisted of severe asthma with long-term steroid use. Her renal function was normal with a creatinine of 90 μmol/L 2 months previously. Following volume resuscitation her renal function did not improve and the microangiopathic haemolytic anaemia worsened. She did not have neurological or any other extra-renal manifestations. Treatment with high-dose intravenous steroids and therapeutic plasma exchange was initiated, and she received twice-daily exchanges for a total of 10 days.

Shigella flexneri was isolated in both blood and stool cultures. Intravenous piperacillin-tazobactam was administered and a putative diagnosis of Stx-HUS was made. Blood samples taken prior to the commencement of therapeutic plasma exchange demonstrated that ADAMTS13 activity was 78% of normal and C3 [1.25 g/L (0.68–1.38)] and C4 [0.24 g/L (0.18–0.60)] levels were normal. Polymerase chain reaction (PCR) analyses for Stx1, Stx2 and STEC were negative. Serum creatinine peaked at 744 μmol/L and she did not require renal replacement therapy. When she was discharged 21 days after admission her serum creatinine was 140 μmol/L.

The patient was reviewed 17 days following discharge with a serum creatinine of 85 μmol/L, a urine protein:creatinine ratio of 32 mg/mmol, and a normal platelet count.
and LDH. Although there was no family history of renal disease, because *S. flexneri* does not usually cause Stx-HUS and the PCR for Stx had been negative, we investigated for aHUS.

Convalescent concentrations of complement components were C3 1.09 g/L (0.68–1.80), C4 0.17 g/L (0.18–0.60), factor H 0.73 g/L (0.35–0.59) and factor I 63 mg/L (38–58). Mutation screening of *CFH*, *CFI*, *CD46*, *C3* and *CFB* revealed a heterozygous *CD46* splice site mutation (IVS2+2T>G). This change has previously been identified in a patient with aHUS and demonstrated to result in abnormal splicing [6, 7]. FACS analysis of granulocytes from the patient demonstrated that cell surface expression of CD46 was reduced by 50% (Figure 1). In the following year she has had no further episodes of HUS, her serum creatinine is 88 μmol/L and urine protein:creatinine ratio is 6 mg/mmol.

**Discussion**

This patient presented with diarrhoea and had clinical findings consistent with a renal thrombotic microangiopathy (not biopsy proven). A presumptive diagnosis of Stx-HUS secondary to *S. flexneri* infection was made and her renal function fully recovered. However, it is unexpected for *S. flexneri* infection to cause Stx-HUS. Of the *Shigella* species, only *S. dysenteriae* type 1 is said to produce the Shiga toxin responsible for HUS [8]. Although *S. dysenteriae* type 1 is the predominant cause of *Shigella*-associated HUS, a few cases of *S. flexneri* HUS have been reported [9]. The pathogenesis of non-*S. dysenteriae*-associated HUS is unclear. Recent studies have failed to detect shiga toxin sequences in non-*S. dysenteriae* type 1 [10] although historical reports suggest infrequent Stx-producing strains [11]. In this case the Stx-PCR was negative and we therefore re- evaluated the diagnosis and in particular investigated for atypical HUS.

Studies of familial aHUS have shown that not all individuals carrying a mutation in a complement gene will develop aHUS. It is now established that other factors, including a trigger, are often needed for aHUS to manifest [3]. In this patient we identified a functionally significant mutation (IVS2+2T>G) in the gene encoding the transmembrane complement regulator CD46. We presume that the *S. flexneri* diarrhoeal illness triggered aHUS in this patient. A preceding non-*E. coli* O157 diarrhoeal illness has been reported in 28% of patients with aHUS [12]. *Escherichia coli* O157 infections have also triggered the first presentation of aHUS in a patient with a CD46 mutation [13]. Another report described two patients with Stx-HUS resulting in end-stage renal disease (ESRD) who developed recurrent HUS following renal transplantation and were subsequently found to have complement gene mutations (*CFH*; p.V412M, *MCP* IVS2+2 T>G) [14].

The prognosis in individuals with aHUS associated with only a *CD46* mutation is good (0–6% risk of ESRD or death within 1 year of the first episode [12]). In keeping with this, our patient recovered normal renal function. In those individuals with a CD46 mutation, who do progress to ESRD, the outcome following transplantation is better than patients with a mutation in *CFH*, *CFI*, *C3* and *CFB* [15]. This is because the renal allograft will usually correct the recipient’s membrane-bound CD46 defect, whereas there is a high rate of recurrent disease post transplant in those individuals with serum complement protein defects.

This case highlights the importance of screening for underlying complement abnormalities in unusual presentations of diarrhoea-associated HUS. Making a diagnosis of aHUS in this patient is important because of the implications for her future management and for her relatives. She remains at risk (70–90% [12]) of recurrent episodes triggered by infection and so she has been advised to present whenever she is unwell so that appropriate investigations can be performed. In the event of a recurrence, therapeutic

---

**Fig. 1.** Flow cytometry analysis of CD46 expression on lymphocytes in a normal subject (a) and the patient (b) measured on the same day. FACS analysis was performed on the day of venepuncture. Mean fluorescence intensity for the patient was 1651 and 3658 for the control subject, consistent with haploinsufficiency of CD46.
plasma exchange would not be expected to be beneficial because CD46 is cell surface bound and not a circulating protein [12, 15]. However, she would be a candidate for treatment with the complement inhibitor eculizumab, which is a monoclonal antibody directed against C5 [16]. In addition, her relatives have been counselled to present for assessment should they become unwell, although they have currently declined genetic screening.

Screening for underlying complement mutations is essential if an aHUS patient has progressed to ESRD and requires renal transplantation. This allows an individualized approach to the use of eculizumab to prevent the morbidity and mortality associated with recurrent aHUS in the graft [17].

In conclusion, HUS presenting in the context of a diarrhoeal infection should not be presumed to be Stx-associated. Non-Stx diarrhoea commonly triggers aHUS in individuals with underlying complement defects. Such patients may benefit from complement modulatory therapy.

Conflict of interest statement. D.K. and T.H.J.G. have received honoraria for consultancy work from Alexion Pharmaceuticals. The results presented in this paper have not been published previously in whole or part, except in abstract format.

References

1. Tarr PI, Gordon CA, Chandler WL. Shiga-toxin-producing *Escherichia coli* and haemolytic uremic syndrome. *Lancet* 2005; 365: 1073–1086
2. Wu CJ, Hsueh PR, Ko WC. A new health threat in Europe: Shiga toxin-producing *Escherichia coli* O104:H4 infections. *J Microbiol Immunol Infect* 2011; 44: 390–393
3. Kavanagh D, Goodship TH, Richards A. Atypical hemolytic uremic syndrome. *Semin Nephrol* 2013; 33: 508–530
4. Legendre CM, Licht C, Muus P et al. Terminal complement inhibitor eculizumab in atypical hemolytic-uremic syndrome. *N Engl J Med* 2013; 368: 2169–2181
5. Lapeyraque AL, Malina M, Fremeaux-Bacchi V et al. Eculizumab in severe Shiga-toxin-associated HUS. *N Engl J Med* 2011 364: 2561–2563
6. Fremeaux-Bacchi V, Moulton EA, Kavanagh D et al. Genetic and functional analyses of membrane cofactor protein (CD46) mutations in atypical hemolytic uremic syndrome. *J Am Soc Nephrol* 2006; 17: 2017–2025
7. Richards A, Kathryn Liszewski M, Kavanagh D et al. Implications of the initial mutations in membrane cofactor protein (MCP; CD46) leading to atypical hemolytic uremic syndrome. *Mol Immunol* 2007; 44: 111–122
8. O’Loughlin EV, Robins-Browne RM. Effect of Shiga toxin and Shiga-like toxins on eukaryotic cells. *Microbes Infect* 2001; 3: 493–507
9. Khan WA, Griffiths JK, Bennish ML. Gastrointestinal and extra-intestinal manifestations of childhood shigellosis in a region where all four species of *Shigella* are endemic. *PLoS One* 2013; 8: e64097
10. Strauch E, Lurz R, Beutin L. Characterization of a Shiga toxin-encoding temperate bacteriophage of *Shigella sonnei*. *Infect Immun* 2001; 69: 7588–7595
11. Prado D, Cleary TG, Pickering LK et al. The relation between production of cytotoxin and clinical features in shigellosis. *J Infect Dis* 1986; 154: 149–155
12. Loirat C, Fremeaux-Bacchi V. Atypical hemolytic uremic syndrome. *Orphanet J Rare Dis* 2011; 6: 60
13. Sellier-Leclerc AL, Fremeaux-Bacchi V, Dragon-Durey MA et al. Differential impact of complement mutations on clinical characteristics in atypical hemolytic uremic syndrome. *J Am Soc Nephrol* 2007; 18: 2392–2400
14. Alberti M, Valoti E, Piras R et al. Two patients with history of STEC-HUS, posttransplant recurrence and complement gene mutations. *Am J Transplant* 2013; 13: 2201–2206
15. Noris M, Caprioli J, Bresin E et al. Relative role of genetic complement abnormalities in sporadic and familial aHUS and their impact on clinical phenotype. *Clin J Am Soc Nephrol* 2010; 5: 1844–1859
16. Wong EK, Goodship TH, Kavanagh D. Complement therapy in atypical haemolytic uraemic syndrome (aHUS). *Mol Immunol* 2013; 56: 199–212
17. Kavanagh D, Richards A, Goodship T et al. Transplantation in atypical hemolytic uremic syndrome. *Semin Thromb Hemost* 2010; 36: 653–659

Received for publication: 13.3.14; Accepted in revised form: 13.3.14