Selective and non-selective loss of immunoregulatory molecules (HLA-A,B,C antigens and LFA-3) in transitional cell carcinoma

A.M.E. Nouri, M.E.F. Smith, D. Crosby & R.T.D. Oliver

Medical Oncology, The London Hospital, Whitechapel, London E1 2BB, UK; and 1Director's Laboratory, Imperial Cancer Research Fund, Lincoln's Inn Fields, London WC2, UK.

Summary: The expression of the major histocompatibility complex (MHC) class I and II antigens and lymphocyte function-associated antigen-3 (LFA-3) was investigated using immunohistochemical staining of bladder tissue sections from 18 patients with transitional cell carcinoma (TCC) and two normal bladder specimens. The expressions of HLA-A,B,C antigens varied greatly between different tumours. Complete loss was observed in one of 18 cases. Moderate to strong expression of HLA-A,B,C antigens was observed in 10 of 18 cases with the remaining seven cases showing either weak expression or expression on only a proportion of the tumour cells. Selective loss of HLA-Bw6 was seen in one of 18 cases. In many cases heterogenous and often focal expression of HLA-D products was seen. In one case tumour cells not expressing HLA-DR antigens were adjacent to strongly HLA-DR expressing non-neoplastic bladder epithelium, indicating a lack of inducible HLA-DR in the tumour cells. LFA-3 was undetectable in two of 18 cases with the remaining 16 cases showing moderate to strong expression of the molecule. These findings indicate that a substantial proportion of bladder tumours have one or more of a wide range of different alterations in the expressions of immunoregulatory molecules that could contribute to escape from immune surveillance.

There has been considerable debate as to whether immunosurveillance is of relevance in providing protection against development of malignancy (for review see Oliver, 1985). The recent evidence that use T cell growth factor, interleukin-2 can produce long term remission in patients with renal cell carcinoma and malignant melanoma (Rosenberg et al., 1987), has led to a rebirth of interest in this debate, particularly with the demonstration that alterations in tumour major histocompatibility complex (MHC) expression may be important in determining which patients respond to treatment (Cohen et al., 1987).

The recognition of neo-antigens by T cells occurs in the context of MHC molecules which are polymorphic cell surface glycoproteins expressed in two forms: class I (HLA-A,B,C) and class II (HLA-D). These molecules act as restriction elements for presentation of foreign antigens to mostly CD8 positive T cytotoxic (CTL) and CD4 positive T helper cells respectively (Zinkernagel & Doherty, 1979). Individual HLA allele products, even those mapping to the same locus, vary in their ability to present different antigen epitopes (Gotch et al., 1987; McMichael et al., 1986).

Class I antigens are composed of a heavy chain and a light chain, β2 microglobulin, whose association with the heavy chain is required for surface expression of class I antigens (Arce Gomez et al., 1978). An additional factor contributing to efficiency of CTL activity is the interaction between CD2 molecules on T cells and lymphocyte function-associated antigen-3 (LFA-3) on target cells in a non-antigen-specific manner (Krensamy et al., 1984).

Evidence that loss of MHC class I antigen on tumours led to increased malignancy first came from studies of murine tumours (for review see Festenstein & Schmidt, 1981). Later studies showed that transfaction of the missing MHC class I gene into these tumour cells led to a reversal of tumourigenicity or of metastatic potential (Hui et al., 1985; Wallich et al., 1985) and more importantly, after immunisation with such altered tumour, exposed mice were resistant to the original class I deficient tumour (Hui et al., 1985).

Increasingly it is being recognised that loss of class I antigen expression may be a frequent event in a human malignancy. Class I antigen loss has been demonstrated in a substantial proportion of colon adenocarcinomas (Smith et al., 1988; Rees et al., 1988; Momburg et al., 1986), breast carcinoma (Fleming et al., 1981) and Burkitt lymphomas (Masucci et al., 1987) providing a possible mechanism for escape from immune surveillance.

There has long been interest in the possibility that bladder cancer may be a tumour where immune surveillance is important because of a strong association of prognosis with degree of lymphocyte infiltration, occurrence of spontaneous regression and the presence of anti-tumour cytotoxic circulating lymphocytes (for review see Oliver, 1985).

In an attempt to assess whether alterations in tumour expression of immunoregulatory molecules does occur in this group of tumours, tumour sections from a series of patients with bladder cancer have been screened with monoclonal antibodies (Mab) to MHC class I and II antigens and to LFA-3 molecule.

Materials and methods

Clinical material

Post-mortem bladder tissue which was microscopically and histologically normal was sampled from two individuals within 24 h of death. Operative specimens were also collected from TCC patients admitted for surgery to the London Hospital. There were five females with an age range of 50–76 years (mean age of 66 years) and 13 males with an age range of 48–85 years (mean age of 68 years). All but three cases (5, 8 and 10, Table I) had surgery at least once before this tissue sampling. In each case the specimen was divided into two portions; one was sent for histological examination and the other was stored in liquid nitrogen until use. Histological examination of the cases indicated that they were all TCC with nine of 18 showing no invasion and nine showing varying degrees of invasion.

Monoclonal antibodies and HLA determinants

The Mabs used as primary reagents in the form of tissue culture supernatants, are listed as follows together with their specificities: BBM.1 detects β2m (Brodskey et al., 1979); HCA10 detects HLA-A,B,C free heavy chain (i.e. not associated with β2m) with a preference for HLA-B products (Stam et al., 1986); PA 2.6 and W6/32 detect all β2m-associated HLA-A,B,C antigens (Brodskey et al., 1979); GAP-A detects HLA-A3 (Berger et al., 1982); 116.5.28 detects HLA-Bw3 (K. Gelishorpe, unpublished data); TS2/9 detects LFA-3 (Krensamy et al., 1983); SPLV3 detects HLA-DQ (Spits et al., 1984); TU39 detects all HLA-D antigens (Pawelec et al., 1982);
Table 1 Expression of HLA-A,B,C and LFA-3 molecules on bladder tumours

| Patient | Cases | HLA-A,B,C Heavy chain | β2-M (BBM1.1) | A2 + B17 (MA2.1) | A2 + Av69 (BB7.2) | A3 (GAP43) | Bw4 (116.5.28) | B26 (126.39) | LFA-3 (TS2/9) |
|---------|-------|-----------------------|---------------|-----------------|------------------|-----------|----------------|-------------|--------------|
| 1       | N     | 4s                    | 4s            | -               | -                | -         | 4s             | 4s          | -            |
| 2       | N     | 4s                    | 4s            | -               | -                | -         | 4s             | 4s          | -            |
| 3       | N     | 4s                    | 4s            | -               | -                | -         | 4s             | 4s          | -            |
| 4       | N     | 4s                    | 4s            | 4s              | 4s               | 4m        | 4s             | 4s          | 4s           |
| 5       | I     | 4s                    | 4s            | 4m              | 4m               | 4w        | -              | 3m          | 4s           |
| 6       | N     | 4s                    | 4s            | 4s              | 4m               | 4w        | 4m             | 4w          | 4s           |
| 7       | I     | 4s                    | 0             | 4s              | 4m               | 3w        | -              | 4w          | 4s           |
| 8       | I     | 4s                    | 4w            | 4s              | 4s               | 4w        | n.d.           | 4s          | 4m           |
| 9       | I     | 4s                    | 4w            | 4s              | 4w               | 4w        | 4s             | 4s          | 4s           |
| 10      | N     | 4m                    | 0             | 4s              | 4m               | 4w        | -              | 4w          | 4m           |
| 11      | N     | 4w                    | 0             | 4w              | 0                | 0         | 4w             | 4w          | 4m           |
| 12      | N     | 4w                    | 4w            | 4w              | 4w               | 4w        | 0              | 4w          | 4m           |
| 13      | I     | 3m                    | 0             | 4m              | -                | -         | 4w             | 0           | 4m           |
| 14      | N     | 2s                    | 1s            | 2s              | -                | 2w        | 1s             | 4m          | 4m           |
| 15      | I     | 2s                    | 1m            | 3m              | 3s               | n.d.      | -              | 4m          | 4m           |
| 16      | N     | 1w                    | 0             | 1w              | -                | -         | 0              | 0           | 4m           |
| 17      | I     | 1w                    | 0             | 1w              | -                | n.d.      | -              | 0           | 4m           |
| 18      | I     | 0                     | 0             | 0               | 0                | 0         | 0              | 0           | 0            |

Assessment: no stromal or tumour cell staining = -; stromal staining present but tumour negative = 0, stromal and tumour cell staining present = grades 1, 2, 3 and 4. Grade 1 < 10%, grade 2 ≥ 10% but < 50%, grade 3 ≥ 50% but < 95% and grade 4 ≥ 95% of tumour cells stain positively. The strength of antigen expression was graded as strong = s, moderate = m and weak = w. n.d. = for not done, I = invasive tumour and N = non-invasive tumour.

L243 detects HLA-DR (Lampson & Levy, 1980); B7/21 detects HLA- DP (Watson et al., 1983) and anti-CD3, -CD4 and -CD8 Mabs (Ortho Pharmaceutical) detect total T, T helper and T cytotoxic lymphocytes subsets respectively. The immunohistochemical specificity of the Mabs directed against HLA-A,B,C allele products was demonstrated by their reactivity on tumour stroma in a series of 12 breast carcinomas, which has been HLA-typed (Dr J.G. Bodmer, unpublished data).

Immunohistochemistry

Frozen sections were cut using a cryostat at a thickness of 7 μm, placed on microscope slides and kept at −80°C until used. The sections were stained as described by Smith et al. (1989). The area of tumour sections varied from 9 to 35 mm². Selected cases with adequate material had repeat testing to establish acceptable reproducibility.

Assessment

Immunohistochemical staining on epithelium and stroma was assessed in normal bladder (where available) and neoplastic bladder tissues. HLA-A,B,C type was deduced from the reaction of the Mabs directed against polymorphic HLA-A,B,C determinants with tumour stroma. Expression of antigens on tumour cells were graded semi-quantitatively by comparison to the degree of expression on the stromal cells (see Table 1).

Results

HLA-A,B,C antigens

Staining of tissue sections from two normal bladders showed strong positive staining in 100% of epithelial cells with Mab against class I antigens (W6/32 and PA2.6) and LFA-3 (TS2/9) and no staining for class II antigens defined by monoclonal antibody L243.

The tumour cells of one of 18 cases showed a complete loss of all HLA-A,B,C molecules (Table 1, case 18, Figure 1). Using Mabs against class I monomorphic determinants, W6/32 and PA2.6, comparable strength of antigen expression between all tumour cells and tumour stromal cells was seen in ten of 18 cases. Of the remaining seven cases, three showed moderate to strong staining on a proportion of tumour cells and four showed weak or absent staining on all cells.

Although the expression of HLA-A,B,C free heavy chain (unassociated with β2-microglobulin) as detected by Mab HC-10 usually mirrored the expression of mature (β2-microglobulin associated) heavy chain as detected by Mab PA2.6, in cases 7, 10 and 13 free heavy chain was undetectable despite moderate to strong expression of mature heavy chain (Table 1).

Of greater interest was the observation that there were patients whose tumour showed selective loss of some class I antigens. Selective loss was defined as complete absence of a specific HLA-A or C product from tumour cells in the presence of moderate to strong expression of monomorphic HLA antigens on tumour stroma. Because of uniformly weak or absent expression of all HLA-A and B antigens detected by monomorphic determinants, five of 18 cases could not be scored for selective loss. One of the remaining 13 tumours showed unequivocal selective loss confirmed by retesting on three separate occasions. This was one of the 12 tumours whose stroma was positive for Bw6. None of seven A-2 positive none of four A-3 positive and none of three Bw4 positive tumours showed selective loss (Table 1, Figure 2a,b). Partial degrees of selective HLA-A,B,C allele product loss are difficult to identify with certainty, though cases showing strong staining with monomorphic HLA-A,B,C Mabs but weak staining with some polymorphic Mabs (such as case 7) which demonstrate weak expression of Bw6 but strong expression of A-2 may be examples of this phenomenon. Using
this lower definition of loss, only four of 18 could be said to have totally normal expression of HLA-A,B,C with the available Mabs against monomorphic and polymorphic determinants.

The pattern of antigen expression on tumour cells using Mabs specific for class II antigens was heterogeneous and often focal. Although nine of nine were positive for the ‘core’ HLA-D antigen recognised by Mab TU39. Seven of nine were positive for DR, five of nine positive for DQ and three of nine positive for DP. HLA-DP and DQ expression mirrored HLA-DR and in no case was a tumour observed to be HLA-DR negative yet HLA-DP or DQ positive. There was not any direct correlation between the expression of HLA-D antigens and that of degree of T cell infiltration (see below) into the tumour area. In one case (11) strong expression of HLA-DR antigen was observed on normal epithelium adjacent to HLA-DR negative neoplastic epithelium (Figure 3).

LFA-3 was expressed with moderate to strong intensity in 16 of 18 cases. There were two cases of complete LFA-3 loss (cases 15, 18, Table I, Figure 4) and in both cases the HLA-A,B,C antigens were either lost or down-regulated.

In a subset of the cases, the nature of tumour infiltrating lymphocytes was investigated using Mabs specific for different T cell subtypes. T cells (CD4>> CD8) positive were found to be present in tumour stroma (11 of 12 cases) the intra-epithelially (eight of 12).

Discussion

Although post-mortem material is not ideal for such studies because of risk of autolysis and problems of induction of class II antigen by terminal bladder infection, the results do confirm the observations from larger studies (Gardiner et al., 1985) that class I but not class II is expressed on normal urothelium.

Against this background the results of these tumour studies have demonstrated that HLA-A,B,C antigen expression is demonstrated in over a third of TCCs. Also, there was a definite case (13) of selective loss of HLA-Bw6 antigen in the presence of normal expression of other HLA-A,B,C products, two cases of complete loss of LFA-3 molecules (both of which also had either complete or partial loss of all class I antigens) and, finally, a single example where HLA-D antigen expression was observed on normal epithelium adjacent to negative neoplastic epithelium. All of these abnormalities in the expression of a immunoregulatory molecules could be capable of conferring a selective growth advantage to tumour cells by enabling escape from T cell immune attack directed against tumour specific antigens.

The selective loss of HLA-A,B,C allele products from neoplastic cells has previously been reported in other tumour types. These include Burkitt lymphoma (Masucci et al., 1987) and colorectal adenocarcinoma (Smith et al., 1989; Rees et al., 1988; Momburg et al., 1989). Such a selective loss of class I antigens could confer resistance to immune attacks against tumour given the varying ability of different HLA-A,B,C allele products to present antigen epitopes to cytotoxic T cells. Thus, the loss of a single allele product could be functionally equivalent, in terms of escape from T cell attack, to the loss of all HLA-A,B,C molecules. Selective losses of HLA-A,B,C allele products could result from a single genetic mutation, whereas loss of all HLA-A,B,C antigens (e.g. case 18) would usually require more than a single mutation.

The incidence of colorectal adenocarcinomas with normal HLA-A,B,C antigen expression (11 of 30 cases) (Smith et al., 1989) was similar to that reported in this study of bladder tumours. However, in contrast to the situation demonstrable in the bladder, most HLA-A,B,C loss in colorectal carcinomas was of individual HLA-A,B,C allele products and not a generalised loss of all HLA-A,B,C antigens as in bladder where there was an unexpectedly high frequency of loss of free heavy chain. This deserves further investigation.

The absence of detectable HLA-A,B,C free heavy chain in tumours expressing mature HLA-A,B,C antigen was seen in
three cases (7, 10 and 13). The basis of this interesting observation may be a slow synthesis of HLA-A,B,C antigen by tumour cells.

Class II antigen expression as detected by TU39 Mab was often focal and of variable intensity on tumour cells of all nine case studies. Locus product specific Mabs for DR, DP and DQ also showed similar areas of positivity on the tumour cells. Similar results regarding the expression of class II antigens on neoplastic bladder epithelium were also reported by Gardiner et al. (1985).

The significance of class II antigen expression on bladder tumour cells is not clear, but it is conceivable that the detection of tumour antigen by infiltrating T cells results in the production of cytokines which in turn stimulate induction of IFN-α, and other products. The focal nature of T cell infiltrate in both tumour stroma and intra-epithelial areas is agreement with this hypothesis. Supportive evidence comes also from the observation that it has been possible to use interleukin-2 (IL-2) to expand activated T cells from six of 18 of these tumour biopsies using IL-2 after culture of tumour cell suspension (A. Nouri, in preparation).

The complete loss of LFA-3 molecules from tumour cells occurred in 11 of 18 cases. This may give tumour additional advantage to escape from immune surveillance given the importance of LFA-3 binding to CD2 molecules on CD8 cytotoxic cells to initiate cytotoxicity. The fact that LFA-3 loss is not, however, confined to bladder neoplasia, having been also demonstrated in colorectal adenocarcinoma (Smith et al., 1989) and Burkitt lymphoma (Gregory et al., 1987) would also support this observation. It is interesting to note that the two cases (15 and 18) showing LFA-3 losses also showed complete or major down-regulation of HLA-A,B,C antigens.

Given the focal nature of these antigen losses and the small size of tumour sample tested, it is obvious that the frequency of loss reported in this study is a minimum incidence. Attempts were made to correlate the presence or absence of invasion (according to UICC system) with class I antigen expression on tumour cells. No significant association was demonstrated in the small numbers tested but it is of interest that the tumour not expressing any HLA-A,B,C antigen or LFA-3 (case 13) showed a complete pattern of immune recognition. More extensive testing is required to clarify whether loss of these immuno-regulatory molecules does lead to selective growth advantage to tumour.

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