Cell proliferation and invasion ability of human choriocarcinoma cells lessened due to inhibition of Sox2 expression by microRNA-145

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Abstract. To date, the mechanism underlying the development of human choriocarcinomas has not been elucidated. It is hypothesized that the Sox2 protein plays a pivotal role in the proliferation and invasion capacity of tumor cells. A microRNA (miR-145) was cloned and used to study the expression of Sox2 and its regulatory effect on the proliferation and invasion capacity of the human choriocarcinoma cell line JAR. In the present study, Sox2 mRNA and protein expression decreased in JAR and JEG-3 cells following transfection with the miR-145 expression virus. Cell proliferation assays indicated that miR-145 expression affected cell cycle regulation and suppressed the proliferation of choriocarcinoma cells in vitro. In addition, xenograft experiments confirmed the suppression of tumor growth in vivo due to cell cycle arrest. Therefore, endogenous mature miR-145 expression may have an important role in the pathogenesis of human choriocarcinomas via interference with the Sox2 target gene by epigenetic modification. This information is of potential significance for the identification of therapeutic targets in human choriocarcinoma.

Introduction

Human choriocarcinomas, which are a rare form of cancer that develops in the uterus from tissue that would normally become the placenta, are a trophoblastic gestational disease. These carcinomas have been studied largely to investigate conditions associated with pregnancy, such as preeclampsia (1). However, the mechanism underlying choriocarcinoma development remains to be elucidated.

microRNAs (miRNAs) are small RNA molecules (21-23 nt) that act as negative regulators of gene expression, either by blocking mRNA translation into protein or by RNA interference (2,3). Previous studies have revealed that dysregulation of specific miRNAs is associated with certain types of cancer and are considered to act either as oncogenes or tumor suppressors, depending on the target gene (2,4-10). Examples include, miR-15a, which has been associated with chronic lymphocytic leukemia (2,5,6), and also miR-21 and miR-17, which are upregulated, while miR-143 and miR-145 are down-regulated in colorectal cancer (11,12). The presence of the let-7 family of miRNAs is a prognostic factor in lung cancer (5,6) and miR-182 inhibits the proliferation and invasion ability of the human lung adenocarcinoma cells via its effect on human cortical actin-associated protein (13). To date, investigation of the decreased expression of miR-199b in human choriocarcinoma by Chao et al. indicated that epigenetic mechanisms play an important role in increasing the expression levels of protein phosphatase 2A inhibitor and contribute to the pathogenesis of human choriocarcinoma (14). However, no studies have linked miR-145 expression with the proliferation and invasion capacity of human choriocarcinoma cells (2).

The transcription activator Sox2 was originally studied in the context of sexual determination during the development of *Drosophila* embryos and thus, its name is an acronym for ‘sex determination Y-box2’ (15-17). Numerous studies have indicated a primary role for Sox2 factor in the maintenance of embryonic stem cell pluripotency, and in later stages of development, in the repression of trophectoderm and epiblast genes. In addition, Sox2 appears to have a significant role in the differentiation of the nervous system (16). Extensive studies have indicated that Oct4, Sox2 and Nanog are required for self-renewal and pluripotency of embryonic stem cells (17,18). Investigation of the expression and methylation profiles of Sox2 in placentas and gestational trophoblastic disease by Li et al. indicated that epigenetic mechanisms play an important role in the transcriptional regulation of Sox2 and contribute to the pathogenesis of gestational trophoblastic disease (19). By contrast, Xu et al. reported that endogenous miR-145 represses the 3' untranslated regions (3'UTRs) of Oct4, Sox2 and Klf4, and that increased miR-145 expression inhibits human embryonic stem cell self-renewal, represses expression of pluripotency genes and induces lineage-restricted differentiation.

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tion (18). In addition, Sox2 was closely associated with certain tumors, its inappropriate activation being involved in the development processes of human tumors, including the abnormal methylation modification of the promoter region of the Sox2 gene. Nakatsuigawa et al (20) analyzed the functions of Sox2 in cancer stem-like cells/cancer-initiating cells derived from human lung adenocarcinoma. Nakatsuigawa et al revealed that the Sox2 protein was detected in >80% of cancer stem-like cells/cancer-initiating cells in primary lung carcinoma tissues. However, Sox2 mRNA knockdown of the human lung cancer stem-like cells/cancer-initiating cells by gene-specific siRNA eliminated tumorigenicity in vitro and in vivo. These observations indicate that Sox2 has a role in the maintenance of stemness and tumorigenicity of human lung carcinoma and is a potential target for treatment.

In view of this evidence, in the current study, the miR-145 expression vector was transfected into the human choriocarcinoma cell lines JAR and JEG-3 to determine its specific role in Sox2 regulation and inhibition of cell proliferation, and invasion. These results are of potential importance for the identification of therapeutic targets in human choriocarcinoma.

Materials and methods

Cell lines and animals

The human choriocarcinoma cell lines JAR and JEG-3 were grown in DMEM (HyClone, Logan, UT, USA) supplemented with 10% fetal bovine serum (PAA Laboratories Inc., Queensland, Australia), penicillin (100 U/ml), streptomycin (100 U/ml) and 2 mM L-glutamine (all were purchased from Hyclone). The JAR cells were incubated at 37°C in a humidified atmosphere of air containing 5% CO2. All experiments with BALB/c nude mice of 6-7 weeks of age and 20-22 g of weight were carried out at the Laboratory Animal Center of Tongji University with Institutional Animal Care and Use Committee approval in accordance with institutional guidelines.

Recombinant lentivirus generation and vector construction.

The method for creating the recombinant lentivirus package was as previously described (18). An RNAi pLL3.7 (LentiLox 3.7) lentiviral system was used to create lentiviral vectors (Clontech, Beijing, China). For vector pLL3.7-mir145 (pre-miRNA of miR-145 expression element), oligonucleotide pairs for pre-miRNA of miR-145 and linker sequences with Hpal and Xhol sites were chemically synthesized. The sequences of the oligonucleotides were: top strand, 5'-CGg tta acc ACC TTT TCC TCA CGG TCC AGT TTT CCC AGG AAT CCC TTA GAT GCT AAG ATG GGA CCT GGA AAT ACT GTT CCT CTT CCT GTC ATG GTT etc gag CG-3'; and bottom strand, 5'-Gcc tgg aga ACC ACC ACC TCA AGA ACA GTA TTT CCA GGA ATC CCC ATC ATC TCA GCT TCT AAG GGA TCC CTG GGA AAA CTG GAC CGT GAG GAC AAC GTG gtt aac CG-3' (sequences corresponding to miR-145 seed sequences are capitalized and bold, and restriction enzyme sites are lower case and bold) (18). To build the expression plasmid, the pairs of oligos were annealed and inserted into the multiple cloning sites between the Hpal and Xhol sites in the pLL3.7 vector. The negative control plasmid pLL3.7-mir145-Mut was similarly constructed, with the exception that 23 nucleotides in sequences corresponding to miR-145 were mutated (GTC CAG TTT GCC CAG AAG TCC CT to Gaa Ct Gaa TTA gCA cGA AgC aCT, mutations shown in lower-case). The pLL3.7-mir145 or pLL3.7-mir145-Mut was recombined in the package cell line 293T to create lentiviruses. Recombinant viruses were propagated in 293T cells, purified and titered by standard methods, as previously described by our laboratory (21). The corresponding viruses were designated Ldv-mir145 and Ldv-mir145-Mut. Co-transfection of human iPSC cells used 4x107 PFU/ml Ldv-mir145 or Ldv-mir145-Mut lentivirus according to the manufacturer's instructions. The cells were seeded in a six-well plate in DMEM (Sigma-Aldrich, St. Louis, MO, USA) supplemented with 10% fetal bovine serum, 100 U/ml penicillin and 100 μg/ml streptomycin at 37°C in a humidified atmosphere of air containing 5% CO2, until cells were 80% confluent.

RNA extraction and analysis by quantitative real-time PCR (qRT-PCR).

Total RNA from each cell was isolated using TRIzol reagent (Invitrogen, Carlsbad, CA, USA) according to the manufacturer's instructions. The RNA samples were treated with DNase I (Sigma-Aldrich), quantified and reverse-transcribed into cDNA using the ReverTra Ace-α First Strand cDNA Synthesis kit (Toyobo, Osaka, Japan). qRT-PCR was conducted using a RealPlex4 real-time PCR detection system from Eppendorf Co., Ltd., (Hamburg, Germany), with SYBR Green RealTime PCR Master mix (Toyobo) used as the detection dye. qRT-PCR amplification was performed over 40 cycles with denaturation at 95°C for 15 sec and annealing at 58°C for 45 sec. Target cDNA was quantified using the relative quantification method. A comparative threshold cycle (Ct) was used to determine gene expression relative to a control (calibrator) and steady-state mRNA levels were reported as an n-fold difference relative to the calibrator. For each sample, the marker genes Ct values were normalized using the formula \( \Delta Ct = Ct_{\text{genes}} - Ct_{\text{iPS RNA}} \). To determine relative expression levels, the following formula was used: \( \Delta \Delta Ct = \Delta Ct_{\text{all groups}} - \Delta Ct_{\text{blank control group}} \). The values used to plot relative expressions of markers were calculated using the expression 2\(^{-\Delta \Delta Ct} \). The mRNA levels were calibrated based on levels of 18S rRNA. The cDNA of each gene was amplified using primers as previously described (19).

Methyl-thiazolyl-tetrazolium (MTT) assay for cell proliferation.

Each group of JAR and JEG-3 cell lines was seeded at 2x10^4 per well in 96-well plates and cultured in DMEM supplemented with 10% FBS at 37°C with 5% CO2, until cells were 85% confluent. The MTT (Sigma Chemicals, St. Louis, MO, USA) reagent (5 mg/ml) was added to the maintenance cell medium at various time points and incubated at 37°C for an additional 4 h. The reaction was terminated with 150 μl dimethylsulfoxide (DMSO, Sigma Chemicals) per well and the cells were lysed for 15 min, and the plates were agitated every 5 min. Absorbance values were determined using the enzyme linked immunosorbent assay (ELISA) reader (Model 680; Bio-Rad, Hercules, CA, USA) at 490 nm.

Flow cytometric (FCM) analysis of cell cycle by propidium iodide (PI) staining.

Each group of JAR and JEG-3 cell lines was seeded at 3x10^5 per well in 6-well plates and cultured until 85% confluent. Each group of cells was washed with
PBS three times, then collected by centrifuging (Allegra X-22R; Beckman Coulter, Miami, FL, USA) at 1000 x g for 5 min. The cell pellets were resuspended in 1 ml PBS, fixed in 70% ice-cold ethanol and kept in a freezer for >48 h. Prior to FCM analysis, the fixed cells were centrifuged, washed twice with PBS and resuspended in PI staining solution (Sigma Chemicals) containing 50 µl/ml PI and 250 µg/ml RNase A (Sigma Chemicals). The cell suspension, which was kept in the dark, was incubated for 30 min at 4°C and analyzed by FACS (FCM-500, Beckman Coulter). A total of 20,000 events were acquired for analysis using CellQuest software.

Luciferase report assay. All steps of the luciferase reporter assay were as previously described (18,22,23). NIH-3T3 cells were seeded at 3x10^4 per well in 48-well plates and co-transfected with 400 ng pLL3.7-miR145, pLL3.7 or pLL3.7-miR145-Mut, 20 ng pGL3cm-Sox2-3'UTR-WT or pGL3cm-Sox2-3'UTR-Mut, and pRL-TK (Promega, Madison, WI, USA) using Lipofectamine 2000 reagent according to the manufacturer's instructions. Luciferase activity was measured 48 h after transfection using the dual-luciferase reporter assay system (Promega).

**RNA extraction and northern blot analysis.** Northern blotting was performed as previously described (13,24). For all groups, 20 µg good quality total RNA was analyzed on a 7.5 M urea 12% PAA denaturing gel and transferred to a Hybond N+ nylon membrane (Amersham, Freiburg, Germany). Membranes were crosslinked using UV light for 30 sec at 1,200 mJ/cm². Hybridization was performed with the miR-145 antisense starfire probe, 5'-AGG GAT TCC TGG GAA AAC TGG AC-3' (IDT, Coralville, IA, USA), to detect the 22-nt miR-199a fragments according to the manufacturer's instructions. After washing, membranes were exposed for 20-40 h to Kodak XAR-5 films (Sigma-Aldrich). As a positive control, all membranes were hybridized with a human U6 snRNA probe, 5'-GCA GGG GCC ATG CTA ATC TTC TCT GTA TCG-3'. Exposure times for the U6 control probe varied between 15 and 30 min.

**Western blot analysis.** Total protein extracts from each group of cells were resolved by 12% SDS-PAGE and transferred on PVDF (Millipore, Billerica, MA, USA) membranes. After blocking, the PVDF membranes were washed 4 times for 15 min with TBST at room temperature and incubated with the primary antibody [rabbit anti-human Sox2 polyclonal antibody (1:200; Chemicon, Temecula, CA, USA)]. After extensive washing, membranes were incubated with secondary peroxidase-linked goat anti-rabbit IgG (1:1000; Santa Cruz Biotechnology, Inc., Santa Cruz, CA, USA) for 1 h. After washing 4 times for 15 min with TBST at room temperature, the immunoreactivity was visualized by enhanced chemiluminescence (ECL kit; Pierce Biotechnology, Inc., Rockford, IL, USA) and the membranes were exposed to Kodak XAR-5 films.

**Soft agar colony formation assay.** The method used was as previously described (25). Soft agar assays were constructed in 6-well plates. The base layer of each well consisted of 2 ml with final concentrations of 1X medium (DMEM+10% FBS) and 0.6% low melting point agarose. The plates were chilled at 4°C until solid. Subsequently, a 1.0-ml agar growth layer, consisting of 1x10⁴ cells suspended in 1X media and 0.3% low melting point agarose, was poured onto the base layer. The plates were again chilled at 4°C until the growth layer congealed. Additional 1X media without agarose (1.0 ml) was added to the top of the growth layer on day 0 and again on day 15 of growth. The cells were allowed to grow at 37°C for 1 month and total colonies counted. The assays were repeated a total of 3 times. Results were statistically analyzed by paired t-test using the PRISM Graphpad program (Graphpad Software, La Jolla, CA, USA).

**Transwell migration assay.** All steps were as previously described (26). The cells (2x10⁵) were resuspended in 200 µl serum-free medium and seeded on the top chamber of the 6.5 mm polycarbonate transwell filters (8.0 µm pores; Corning Inc., Corning, NY, USA). The full medium (600 µl) containing 10% FBS was added to the bottom chamber. The cells were allowed to migrate for 24 h at 37°C in a humidified incubator with 5% CO₂. The cells attached to the lower surface of the membrane were fixed in 4% paraformaldehyde at room temperature for 30 min and stained with 4,6-diamidino-2-phenylindole (DAPI; C1002; Beyotime Institute of Biotechnology, Jiangsu, China), and the number of cells on the lower surface of the filters was counted under the microscope. A total of 5 fields were counted for each transwell filter.

**In vivo xenograft experiments.** Logarithmically growing ovarian cancer-initiating cells (~1x10⁶) were inoculated into BALB/c nude/mude mice. Each experimental group consisted of four mice. After 4 weeks of observation, the mice were sacrificed and tumors were obtained (27). The tumor weight was measured and tumor volume was calculated according to the formula: tumor volume (mm³) = length (mm) x width (mm) x height (mm).

**Statistical analysis.** Each experiment was performed at least three times and data were expressed as the mean±SE. The differences were evaluated using Student’s t-tests. P<0.05 was considered to indicate a statistically significant result.

**Results**

**miR-145 binding with the 3'-UTR sites in Sox2.** Using an online research tool, the miRBase Target database (http://www.mirbase.org) (28,29), the precursor miRNA (pre-miRNA) sequences, mature miRNA sequences, chromosomal locations and length of miR-145 and the target gene Sox2 were analyzed. Seven putative miRNA target sites were identified in the 3'-UTR of Sox2 mRNA, depending on species. This study focused on human miR-145, which targets the human Sox2 3'-UTR, although conservation in this sequence indicates the possibility of binding to varying degrees, across species (Fig. 1). Plasmid DNA encoding each Sox2 mRNA 3'-UTR site [wild-type (wt) Sox2, empty plasmid and mutant Sox2] was co-transfected with the miR-145 expression lentivirus (wt miR-145, empty lentivirus and mutant miR-145 lentivirus) into the mouse embryonic fibroblast cell line NIH-3T3, to examine regulation of Sox2 gene expression by mature miR-145. The
miR-145 specifically influences expression of Sox2 protein in human choriocarcinoma cell lines. Northern blot analysis demonstrated that the hybridized signal of mutant miR-145 in the JAR and JEG-3 choriocarcinoma cell lines was weaker than in cells transfected with wt miR-145. qRT-PCR and western blot analyses were used to determine the effect of exogenous and endogenous miR-145 expression on Sox2 expression. qRT-PCR analyses revealed decreased Sox2 mRNA expression in wt miR-145 lentivirus-transfected JAR and JEG-3 cells than in untransfected and mutant miR-145-transfected cells. The relative mRNA expression after normalization to 18S ribosomal RNA (rRNA), which served as an internal control, is shown in Fig. 1. Notably, western blotting revealed that Sox2 levels in untransfected cells (JAR or JEG-3 cell lines) and mutant miR-145 transfected cells (JAR or JEG-3 cell lines) were 0.667±0.026 or 0.876±0.036, and 0.669±0.020 or 0.879±0.028 relative to those of GAPDH, respectively (Fig. 1). These values were significantly higher than those for the wt miR-145 transfected group (JAR: 0.429±0.019; JEG-3: 0.547±0.040 relative to GAPDH), which indicated that exogenous miR-145 downregulates Sox2 expression. Therefore, miR-145 expression may influence endogenous Sox2 expression.

Figure 1. miR-145 and Sox2 expression in different groups. (A) The human Sox2 microRNA (miRNA) 3′-untranslated region (3′-UTR) contains miR-145 binding sites. The mature miR-145 sequences of multiple species were analyzed and contrasted using bioinformatics tools. The typical secondary structure of precursor miRNAs (pre-miRNAs) was compared with miR-145. Pre-miRNA contains stem-loop and hairpin structures and the common binding site is located in an unstable region with a multi-branching loop-like RNA structure. Mature miRNAs are bound in the 3′-UTR of the target gene. Complementarity between miR-145 and the putative human Sox2 3′-UTR site target (1-20 bp downstream) showed that the conserved bases of the putative miR-145 target sequence are present in the human Sox2 3′-UTR. (B) The expression of miR-145 and its interference with the target gene Sox2 were assessed by luciferase assays. Wild-type (wt) reporter or mutated control luciferase plasmids were transfected into NIH-3T3 cells with miR-145 or mutant miR-145 expression viruses. Luciferase activity within the Sox2 3′-UTR sites was inhibited by miR-145 (*P<0.01 vs. pLL3.7; #P>0.05 vs. pLL3.7; n=3). (C) Northern blot hybridized signals of miR-145 in human choriocarcinoma cells. Northern blot hybridized signals of miR-145 in (Ca) JAR cells and (Cb) JEG-3 cells. Northern blot analysis showed a weaker hybridized signal in mutant miR-145-transfected cells than in wt miR-145-transfected cells. The human U6 probe was used as a loading control. (D) Sox2 mRNA expression assay in different human choriocarcinoma cells by quantitative real-time PCR (qRT-PCR). qRT-PCR indicated lower expression of Sox2 mRNA in the (Da) wt miR-145-transfected JAR cells and (Db) wt miR-145-transfected JEG-3 cells than in the corresponding untransfected or mutant miR-145-transfected cells. Relative mRNA expression is shown after normalization to 18S rRNA, which served as an internal control. (E) Western blot showing the expression of Sox2 in various groups. Western blots showing the expression of Sox2 in (Ea) JAR cells and (Eb) JEG-3 cells. Sox2 levels were significantly higher in the untransfected or mutant miR-145-transfected cells than in the miR-145-transfected cells. Data indicate that exogenous miR-145 downregulates Sox2 expression (*P<0.01 vs. untransfected; #P>0.05 vs. untransfected; n=3).
exogenous miR-145 expression inhibited the growth of human choriocarcinoma JAR and JEG-3 cell lines in vitro. In addition, migration and invasion ability were shown to be reduced in JAR and JEG-3 cells with stably repressed Sox2 mediated by miR-145 transfection using transwell migration analysis and soft agar colony formation assays, respectively (Fig. 2). Transwell migration invasion assays showed that the number of invading wt miR-145-transfected JAR cells was significantly lower than the numbers of invading untransfected and mutant miR-145-transfected JAR cells (invading cell numbers: miR-145 transfected group, 13±2; untransfected cells, 24±2; mutant miR-145 transfected cells, 26±2). The results of transwell migration invasion assays in JEG-3 cells were similar to those of JAR cells. These results indicate that the repression of Sox2 expression by miR-145 significantly attenuates the invasion and migration ability of human choriocarcinoma cells. Soft agar colony formation assays consistently indicated that miR-145-transfected cells formed substantially fewer colonies compared with controls or mutant miR-145-transfected cells when plated at low density (Fig. 2). In addition, miR-145 transfected, mutant miR-145 transfected and untransfected JAR or JEG-3 cells were stained with PI, and analyzed by flow cytometry to detect changes in cell cycle progression. As shown in Fig. 3, the majority of the wt miR-145-transfected JAR cells were arrested in the G0/G1 phase of the cell cycle and the percentage of cells in the S phase were markedly decreased. However, no significant differences were observed in the cell cycle distribution of the mutant miR-145-transfected and untransfected JAR cells. The results suggest that miR-145-mediated repression of Sox2 expression significantly attenuates the invasion and migration capacity of human choriocarcinoma cells.
suggested that wt miR-145 expression affected cell cycle regulation in human choriocarcinoma cells in vitro.

Expression of wt miR-145 in JAR cells inhibited subcutaneous tumor growth in nude mice. The effect of miR-145 expression on tumor growth was investigated in vivo by subcutaneous inoculation of the miR-145 lentivirus-transfected JAR cells and mutant miR-145 lentivirus-transfected JAR cells into two groups of nude mice. All the mice in the mutant miR-145 group developed tumors ~37 days after injection, whereas tumors were detected in only one in four mice from the miR-145 transfected group at this time. Although both groups developed tumors, the tumors formed by wt miR-145-transfected cells grew more slowly than those in the mutant miR-145-transfected group (Fig. 4). In addition, when the mice were sacrificed 62 days after injection, tumor weights in the mutant miR-145 transfected group were significantly heavier than those in the wt miR-145 lentivirus-transfected
group. Furthermore, miR-145 expression in the JAR cell line was associated with a significant decrease in tumor volume (Fig. 4). These results suggested that miR-145 expression in the human choriocarcinoma cell line suppressed \textit{in vivo} tumor growth.

**Discussion**

Increasing evidence has shown that miRNA plays an important role in the proliferation and invasion ability of numerous types of cancer cells. However, the miRNAs that regulate human choriocarcinoma cell growth and invasion have not yet been reported. In the present study, it was observed that miR-145 interfered with Sox2 expression via putative sites located in the 3'-UTR region. Therefore, it was hypothesized that miR-145 suppresses the human choriocarcinoma cell line JAR by downregulation of Sox2 expression. Putative miRNA target sites in the 3'-UTR of Sox2 mRNA were used to construct a wt miR-145 expression lentivirus, which was then transfected into the JAR human choriocarcinoma cell line. Luciferase activity assays indicated that the activity of the Sox2 3'-UTR site was significantly inhibited by wt miR-145, while that of the mutated Sox2 3'-UTR site was unchanged, which suggested that miR-145 targeted Sox2. In addition, qRT-PCR and western blot analysis demonstrated that Sox2 protein expression was reduced in wt miR-145 lentivirus-transfected JAR cells compared with the levels detected in mutant miR-145 lentivirus-transfected or untransfected cells. FCM analysis revealed that the majority of wt miR-145-transfected cells were arrested in the G0/G1 phase of the cell cycle with reduced percentages in the S and G2/M phases, which suggested that miR-145 expression affected the cell cycle regulation of choriocarcinoma cells \textit{in vitro}. Similarly, exogenous miR-145 expression was shown to inhibit the growth of the JAR cell line \textit{in vitro} using MTT assays. Soft agar colony formation assay and transwell migration invasion assays showed that the number of invading wt miR-145 transfected cells was significantly lower than the numbers of invading untransfected and mutant miR-145 transfected cells. Finally, xenograft experiments indicated that miR-145 expressed in the JAR cell line also suppressed tumor growth \textit{in vivo}.

This is in contrast to a previous report which found that the expression of high levels of Sox2 was associated with malignancy in human lung cancer stem-like cells/cancer-initiating cells (20). In studies of human choriocarcinoma, it was observed that expressed levels of Sox2 in human gestational trophoblastic neoplasia cells were higher than in the normal trophoblast cells (19). Thus, we considered that there was an association between Sox2 expression and malignancy in human choriocarcinoma. By contrast, certain studies have shown that Sox2 plays potentially important roles in the pathogenesis of human choriocarcinomas.

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