Distribution of arbuscular mycorrhizal fungi (AMF) in Terceira and São Miguel Islands (Azores)

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Abstract

Background

The data, presented here, come from samples collected during three research projects which aimed to assess the impact of land-use type on Arbuscular Mycorrhizal Fungi (AMF) diversity and community composition in pastures of Terceira Island (Azores, Macaronesia, Portugal) and also in the native forest of two Azorean Islands (Terceira and São Miguel; Azores, Macaronesia, Portugal). Both projects contributed to improving the knowledge of AMF community structure at both local and regional scales.
New information

Little is known on the AMF communities from Azores islands and this study reports the first survey in two Azorean Islands (Terceira and São Miguel). A total of 18,733 glomeromycotan spores were classified at the species level from 244 field soil samples collected in three different habitat types – native forests (dominated by Juniperus brevifolia and Picconia azorica), semi-natural and intensively-managed pastures. Thirty-seven distinct spore morphotypes, representing ten glomeromycotan families, were detected. Species of the family Acaulosporaceae dominated the samples, with 13 species (38% of the taxa), followed by Glomeraceae (6 spp.), Diversisporaceae (4 spp.), Archaeosporaceae (3 spp.), Claroideoglomeraceae (3 spp.), Gigasporaceae (3 spp.), Ambisporaceae and Paraglomeraceae, both with the same number of AMF species (2 spp.), Sacculosporaceae (1 sp.) and Entrophospora (family insertae sedis). Members of the family Acaulosporaceae occurred almost exclusively in the native forests especially associated with the Picconia azorica rhizosphere, while members of Gigasporaceae family showed a high tendency to occupy the semi-natural pastures and the native forests of Picconia azorica. Members of Glomeraceae family were broadly distributed by all types of habitat which confirm the high ecological plasticity of this AMF family to occupy the more diverse habitats.

Keywords

Arbuscular mycorrhizal fungi (AMF), native forest, Juniperus brevifolia, Picconia azorica, semi-natural and intensive pastures

Introduction

Arbuscular mycorrhizal fungi (AMF) are one of the most important groups of below-ground biota (Jeffries et al. 2003; Barea et al. 2005). These obligate symbionts live in association with approximately 80% of vascular plants and have essential ecological roles, namely, they facilitate plant growth through enhancing uptake of several macro- and micro-nutrients of low mobility (e.g. P, Zn, Cu) in soil (Brundrett and Tedersoo 2018). Arbuscular mycorrhizas can also provide other ecological functions such as influencing the microbial and chemical environment of the mycorrhizosphere, stabilising soil aggregates (Rillig and Mummeny 2006) and conferring plant tolerance to several abiotic (Göhre and Paszkowski 2006; Li et al. 2013; Chitarra et al. 2016) and biotic (Vos et al. 2012; van der Heijden et al. 2015) stresses.

AMF are, therefore, beneficial for plant performance, playing a crucial role for the sustainability of natural and agricultural ecosystems (Barea et al. 2011) and important ecosystem services (Chen et al. 2018). However, despite their ecological role, little is known about how their community structure varies in relation to habitat type in the Azores archipelago.
The Azores archipelago has an extended area of grasslands (Martins 1993), including natural grasslands, semi-natural pastures and intensive pastures (Cardoso et al. 2009). It also has the unique native forest, Laurisilva, which has more endemic plants and animals than any other habitat in the region. In the last 500 years, as a consequence of human activity, much of this native forest has been replaced by man-made habitats and has been subjected to fragmentation (Borges et al. 2005). Thus, immediate action to restore and expand native forest is required to avoid the ongoing loss of endemic species (Terzopoulou et al. 2015). AMF play an important role in habitat restoration, by improving plant nutrition and performance under environmental stress by facilitating plant adaptation in both nursery and field conditions (Babu and Reddy 2011). Therefore, understanding the AMF diversity in the native forest will help to define strategies for management and restoration of such endangered forests. An important step in restoration strategies is the re-establishment of adapted native plant species (Ferrol et al. 2004). A good understanding of mycorrhizal associations in undisturbed localities could then be used to provide information about AMF inoculum production for use in the rehabilitation of degraded ecosystems.

In this contribution, we list the species of Arbuscular Mycorrhizal Fungi (AMF) found in ecological studies, comparing anthropogenically disturbed pastures and forests of Terceira Island (Azores, Macaronesia, Portugal) and also in the native forests of São Miguel Island (Azores, Macaronesia, Portugal).

General description

Purpose: In this contribution, we list the AMF species found in pastures from different land-use types of Terceira Island to investigate the effect of disturbance on AMF community structure. Native forests from Terceira and São Miguel Island were also sampled to observe patterns of AMF species composition and distribution, in order to provide baseline information for later use in establishing strategies for conservation of Picconia azorica and Juniperus brevifolia, in particular and native Azorean forests, in general.

Project description

Study area description: All data used in this study came from surveys about AMF diversity and composition in different ecosystems (pasturelands and native forests) conducted in two Islands of the Azorean archipelago, Terceira and São Miguel (Melo et al. 2014, Melo et al. 2017, Melo et al. 2018) (Fig. 1). The sampling areas were cattle-grazed upland pastures of two different types from Terceira and four fragments of native forests from each Island (Table 1) (Fig. 2). The two pasture types include semi-natural pastures with low grazing intensity and frequency (managed for more than 50 years, with low stocking density, grazed only in summer and with a relatively high diversity of grasses and forbs) and intensively-managed pastures with high grazing intensity and frequency (managed for more than 30 years, with high stocking density, grazing during all year and characterised also by a depauperate vascular flora of five or fewer dominant species) (Melo et al. 2014). The semi-natural pastures, Pico Galhardo (TER_SP_PG) and Terra
Brava (TER_SP_TB) (Fig. 1) are included in Terceira Natural Park and are dominated by the perennial grasses *Holcus lanatus* and *Agrostis castellana*, have a high floristic diversity (Dias 1996, Borges 1997), often including other grasses such as *Anthoxanthum odoratum, Lolium multiflorum, Holcus rigidus* and *Poa trivialis* and non-forage species including *Lotus uliginosus, Rumex acetosella* ssp. *angiocarpus, Potentilla anglica, Hydrocotyle vulgaris, Plantago lanceolata* and *Lobelia urens* (For more details see Melo et al. 2014). The intensively-managed pastures, Agualva 1 (TER_IP_R1) and Agualva 2 (TER_IP_R2) (Fig. 1) resulted from the conversion of undisturbed native forest to wood production of non-native trees and then to permanent pastures. They are now surrounded by an exotic eucalyptus plantation. The vegetation is dominated by *Holcus lanatus* and *Lolium perenne*, but also has high populations of *Trifolium repens*, *P. lanceolata*, *Cyperus esculentus*, *Mentha suaveolens*, *Cerastium fontanum* and *Rumex conglomeratus* (Dias 1996, Borges 1997).

| Site           | Longitude | Latitude |
|----------------|-----------|----------|
| TER_SP-PG      | -27.2232  | 38.7296  |
| TER_SP_TB      | -27.2112  | 38.7341  |
| TER_IP_R1      | -27.1922  | 38.8296  |
| TER_IP_R2      | -27.1950  | 38.7568  |
| TER_NF_PG      | -27.2272  | 38.7341  |
| TER_NF_LA      | -27.3310  | 38.7496  |
| TER_NF_TB      | -27.1979  | 38.7323  |
| TER_NF_SE      | -27.3677  | 38.7618  |
| SMG_NF_LO      | -25.4649  | 37.7856  |
| SMG_NF_TR      | -25.1854  | 37.7940  |
| SMG_NF_LG      | -25.1433  | 37.7862  |
| SMG_NF_RQ      | -25.3020  | 37.7407  |

In Terceira Island, the native forests included two fragments from Natural Park – Pico Galhardo (TER_NF_PG) and Lagoinha (TER_NF_LA) (Fig. 1) (Melo et al. 2017), both dominated by the Azorean cedar *Juniperus brevifolia*, a rare conifer species that is endemic to Azores, which dominates at high-elevation (> 650 m), with subdominant endemic woody perennials, including *Laurus azorica* (Lauraceae), *Ilex perado azorica* (Aquifoliaceae), *Erica azorica* (Ericaceae), *Vaccinium ciliatrum* (Ericaceae) and *Frangula azorica* (Rhamnaceae) (Elias et al. 2016). Nevertheless, in Lagoinha, invasive woody species, including *Cryptomeria japonica, Pittosporum undulatum* (Pittosporaceae), *Eucalyptus globules* and *Acacia melanoxylon* (Fabaceae), have begun to establish. The remaining two native fragments from Terceira Island include two populations of *Picconia*.
azorica – Terra Brava (TER_NF_TB) and Serreta (TER_NF_SE) (Fig. 1). Terra Brava is located in the very wet Laurisilva at 650 m altitude (Fig. 1) being dominated by endemic woody plants, predominantly L. azorica, I. azorica, Frangula azorica, V. cylindraceum, E. azorica, Myrsine africana and, occasionally, by J. brevifolia and P. azorica. Serreta (NFSE) is located at low altitude (95 m) (Fig. 1) and is characterised by a low diversity of plants, dominated by Morella faya and P. azorica and, occasionally, by L. azorica. These forests are located in the most thermophilic areas of Azores and are almost extinct (Dias 1996). The highest canopy is dominated by a dense cover of P. undulatum and, rarely, by L. azorica. This forest is mixed with other invasive woody species, including Metrosideros excelsa, E. globules, A. melanoxylon, Sphaeropteris cooperi, Fuchsia magellanica and Rubus inermis. The herbaceous stratum is dominated by Dryopteris azorica, Hedera helix var. azorica, Smilax aspera and Gomphocarpus fruticosus (Dias 1996).

Figure 1. Location of sampling sites in Terceira and São Miguel Islands (Azores).

Figure 2. Habitat types in Terceira and São Miguel Islands, Azores: a. native forest of Juniperus brevifolia; b. native forest of Picconia azorica; c. semi-natural pastures; d. intensively-managed pastures.
In São Miguel Island, the four native fragments are two populations of *J. brevifolia* up to 700 m altitude (Lombadas and Tronqueira) and two populations of *P. azorica* in the lowlands around 95 m altitude in Lombo Gordo and Ribeira Quente (Fig. 1). Lombadas (SMG_NF_LO) is included in the Natural Reserve of Lagoa do Fogo in São Miguel (Fig. 1). Although the surrounding vegetation is dominated by the introduced species *C. japonica*, *Clethra arborea*, *A. melanoxylon* and *E. globulus*, this forest still retains several endemic elements, including *J. brevifolia*, *V. cylindraceum* (Ericaceae), *L. azorica*, *Euphorbia stygiana* (Euphorbiaceae), *F. azorica*, *E. azorica*, *I. azorica* and *Culcita macrocarpa* (Culcitaceae) (Silva 2001). Tronqueira (SMG_NF_TR) is located in hyperhumid native forest (Fig. 1), a type of forest that has been largely replaced by other land uses (Moreira et al. 2012), resulting in an abundance of exotic plants, such as *C. japonica* and *C. arborea*. The tree layer is composed of the endemic woody plants *J. brevifolia*, *L. azorica*, *L. africana* (Myrsinaceae) and *E. azorica*, while the shrub layer is mostly formed by *V. cylindraceum* and *Viburnum treleasei* (Adoxaceae). Lombo Gordo (SMG_NF_LG) is covered by a coastal scrubland (Fig. 1) where *P. azorica* dominates in certain areas, but is mixed with other native and invasive woody species including *M. faya*, *E. azorica*, *P. undulatum*, *Arundo donax*, *Hedychium gardnerianum* and *Phormium tenax* (Martins et al. 2011). Ribeira Quente (SMG_NF_RQ) is also a coastal scrubland (Fig. 1) dominated by the endemic plants *L. azorica* and *P. azorica* but also associated with other native and invasive woody species, such as *P. undulatum*, *M. faya* and *A. melanoxylon*.

**Design description:** Arbuscular mycorrhizal fungi (AMF) diversity and composition were investigated at three habitat types: native forests of *J. brevifolia* and *P. azorica*, semi-natural pastures and intensively-managed pastures. Each habitat type was represented by two sites. At each site from semi-natural (Pico Galhardo (10.39 ha) – TER_SP_PG; Terra Brava (8.81 ha) – TER_SP_TB) and intensively (Agualva 1 (5.03 ha) – TER_IP_R1; Agualva 2 (3.06 ha) – TER_IP_R2) managed pastures, ten soil samples were collected in August 2007 (i.e. a total of 40 soil samples) (Project CD_Melo_PhD). In natural forests of *J. brevifolia* from Terceira (Pico Galhardo (13.97 ha) – TER_NF_PG; Lagoinha (3.05 ha) – TER_NF_LA) and from São Miguel (Lombadas (37.42 ha) – SMG_NF_LO; Tronqueira (51.70 ha) – SMG_NF_TR), 21 soil samples were collected from seven marked *J. brevifolia* plants in each site at three different sampling times (September 2012; May 2013; September 2013) in both islands, resulting in a total of 84 soil samples (2 islands × 2 sites/island × 7 samples/site × 3 sampling dates) (CD_Melo_Postdoc; FCT - PTDC /AGR-ALI/122152/2010). In natural forests of *P. azorica* from Terceira (Terra Brava (9.72 ha) - TER_NF_TB; Serreta (7.67 ha) – TER_NF_SE) and from São Miguel (Lombo Gordo (41.69 ha –SMG_ NF_LG; Ribeira Quente (6.32 ha) – SMG_NF_RQ), 30 soil samples were collected from ten marked *P. azorica* plants in each site, during the same sampling times for *J. brevifolia*, in both islands resulting in 120 soil samples (2 islands × 2 sites/island × 10 samples/site × 3 sampling dates) (CD_Melo_Postdoc; FCT - PTDC /AGR-ALI/122152/2010).
Sampling methods

Sampling description: In semi-natural and intensively-managed pastures, the soil samples with associated roots were randomly collected with a shovel, from the rooting zone of the dominant plant species, *H. lanatus*, to a depth of 0 - 20 cm. In native fragments of *P. azorica*, the distance between samples taken on each site was a minimum of 25 m and maximum of 40 m and the distance between sample sites was about 20 km in Terceira and 15 km in São Miguel. Each soil sample was geo-referenced and consisted of four subsamples collected from different points (approximately N, S, E and W) around the rooting zone of each *P. azorica* plant with a shovel to a depth of 0 - 20 cm or 0 - 30 cm, depending on the soil conditions and the depth of rhizosphere system. The litter layer was removed during sampling and replaced afterwards. Subsequent samples were taken from the same marked plants following the cardinal points. In the case of native fragments of *J. brevifolia*, the distance between samples taken on each site was between 25 m and 40 m and the distance between sample sites was about 5 km in Terceira and 24 km in São Miguel. The sample collection followed the same procedure as for *P. azorica* (Melo et al. 2017). For all habitat types, each soil sample consisted of approximately 2 kg of rhizosphere soil. In the lab, the soil samples were air-dried, sieved through a 2 mm mesh and stored at 4ºC before analysis.

Quality control: Frequently, spores directly extracted from the soil are low in number and contaminated by other organisms, which makes their identification difficult. Consequently, it is necessary to establish trap cultures to promote sporulation and provide specimens for detailed examination. Open pot-trap cultures (Gilmore 1968) were established from each soil sample collected at semi-natural and intensive pastures with one-week-old *Zea mays* seedlings (Melo et al. 2014). Soil samples collected from the native forests were used to establish two of such cultures, one with one-week-old *Z. mays* seedlings and another one with micropropagated *J. brevifolia* and *P. azorica* seedlings (Melo et al. 2017, Melo et al. 2018). Establishment of single or multi-spore cultures of the different AM fungal morphotypes with *Plantago lanceolata* as host plant was attempted in pots of river sand. Spores with a healthy appearance (oily contents; without evidence of contamination by non-AMF) of each AM fungal morphotype were used as inoculum by placing them on a seedling root system under a dissecting microscope, immediately before transplanting into the pot (Melo et al. 2017, Melo et al. 2018). Specimens were given a voucher number, linked to their culture attempt number. Individual microscope slides were numbered serially so that photographic images could be traced back to their specimen of origin and details were recorded in a database to allow complete tracking of culturing history and linkage of related voucher specimens. The new cultures were placed in a climate-controlled plant growth chamber. When needed, cultures were watered with deionised water. Individuals of morphologically characterised spore types, extracted from field soil, trap cultures or single spore cultures, were used for DNA analysis. Molecular characterisation, including DNA extraction, PCR, cloning, RFLP, sequencing and phylogenetic analyses, is published in Melo et al. (Melo et al. 2017, Melo et al. 2018).
**Step description:** Glomeromycotan spores were extracted from 50 g of air-dried soil from each sample (field soil, trap cultures and single or multi-spore cultures) by wet sieving and sucrose centrifugation (Walker 1992) and stored at 4°C in autoclaved water, pending examination. Different spore types were initially separated in water under a stereomicroscope. Representatives of each morphotype were identified through a compound microscope in a 4:1 mixture of polyvinyl alcohol lacto-glycerol (PVLG) and Melzer's reagent, photographed and stored as semi-permanent slide preparations. Counts were made for the total number of spores of each morphotype under a dissecting microscope after classification into either known species or types that could not be placed in a current species, based on colour, size, surface ornamentation, hyphal attachment, reaction to Melzer's reagent and wall structure. Identification of spores was carried out by use of primary literature and experience from more than 40 years of taxonomic study of the *Glomeromycota* by C. Walker (e.g., Walker and Trappe 1981, Walker et al. 1984, Koske and Walker 1985, Walker et al. 1986, Walker and Diederichs 1989, Walker 1992, Walker and Vestberg 1998, Schüßler et al. 2011, Krüger et al. 2012, Redecker et al. 2013, Walker et al. 2018, Wijayawardene et al. 2018, Schüßler and Walker 2019) and joint authorship with A. Schüßler of the website amf-phylogeny.com, which lists all accepted species in the phylum. The illustrated manual of Blaszkowski (2012) was also used. Comparisons were made, where possible, with the type and the authenticated specimens and with other literature such as original species descriptions, websites (e.g. http://invam.wvu.edu) and examination of other well-documented specimens, when available.

**Geographic coverage**

**Description:** Terceira and São Miguel Islands, the Azores, Macaronesia, Portugal.

**Coordinates:** 37.423 and 38.959 Latitude; -27.532 and -24.917 Longitude.

**Taxonomic coverage**

**Taxa included:**

| Rank      | Scientific Name | Common Name                      |
|-----------|-----------------|----------------------------------|
| phylum    | *Glomeromycota* | Arbuscular mycorrhizal fungi     |

**Collection data**

**Collection name:** AMF data base

**Collection identifier:** Catarina Melo

**Parent collection identifier:** Christopher Walker

**Specimen preservation method:** PVLG-Melzer slide
Usage rights

Use license: Creative Commons Public Domain Waiver (CC-Zero)

Data resources

Data package title: Distribution of Arbuscular Mycorrhizal Fungi in Terceira and S. Miguel (Azores, Portugal)

Resource link: http://ipt.gbif.pt/ipt/resource?r=arbuscular_mycorrhizal_fungi_terceira_azores

Alternative identifiers: https://www.gbif.org/dataset/c72a7a97-9de0-4854-aa80-90df6389ff12

Number of data sets: 1

Data set name: arbuscular_mycorrhizal_fungi_terceira_azores

Download URL: http://ipt.gbif.pt/ipt/resource?r=arbuscular_mycorrhizal_fungi_terceira_azores

Data format: Darwin Core Archive

Data format version: 1.4

Description: The following data table includes all the records for which a taxonomic determination of the species was possible. The dataset submitted to GBIF (Melo et al. 2019) is structured as a sample event dataset, with two tables: event (as core) and occurrences. The data in this sampling event resource have been published as a Darwin Core Archive (DwCA), which is a standardised format for sharing biodiversity data as a set of one or more data tables. The core data table contains 226 records (eventID). One extension data table also exists with 665 occurrences. An extension record supplies extra information about a core record. The number of records in each extension data table is illustrated in the IPT link. This IPT link archives the data and thus serves as the data repository. The data and resource metadata are available for downloading in the downloads section.

| Column label     | Column description                                           |
|------------------|--------------------------------------------------------------|
| Table of Events  | Table with sampling events data                             |
| eventID          | Identifier of the events, unique for the dataset            |
| samplingProtocol | The sampling protocol used to capture the species           |
| eventRemarks     | Remarks of the plant species from where the specimens were extracted |
| sampleSizeValue  | The numeric amount of time spent in each sampling            |
| sampleSizeUnit   | The unit of the sample size value                            |
| Field                  | Description                                                                 |
|-----------------------|-----------------------------------------------------------------------------|
| eventDate             | Date or date range when the record was collected                           |
| Year                  | Year of the event                                                          |
| Month                 | Month of the event                                                         |
| country               | Country of the sampling site                                               |
| locality              | Name of the locality                                                       |
| stateProvince         | Name of the region of the sampling site                                     |
| island                | Name of the island                                                         |
| locationID            | Identifier of the location                                                 |
| habitat               | The surveyed habitat                                                       |
| DecimalLatitude       | Approximate centre point decimal latitude of the field site in GPS coordinates |
| DecimalLongitude      | Approximate centre point decimal longitude of the field site in GPS coordinates |
| coordinateUncertaintyInMetres | Uncertainty of the coordinates                                            |
| coordinatePrecision   | Precision of the coordinates                                               |
| georeferenceSources    | Method used to obtain coordinates                                          |
| eventRemarks          | The list of Projects supporting the sampling event                         |
| Table of Occurrences  | Table with species density data (beginning of new table)                  |
| CatalogNumber         | Unique identification code for species density data                        |
| eventID               | Identifier of the events, unique for the dataset                           |
| occurrenceID          | Identifier of the record, coded as a global unique identifier              |
| licence               | Reference to the licence under which the record is published               |
| institutionID         | The identity of the institution publishing the data                         |
| institutionCode       | The code of the institution publishing the data                             |
| basisOfRecord         | The nature of the data record                                              |
| kingdom               | Kingdom name                                                               |
| phylum                | Phylum name                                                                |
| class                 | Class name                                                                 |
| order                 | Order name                                                                 |
| family                | Family name                                                                |
| genus                 | Genus name                                                                 |
| specificEpithet       | Specific epithet                                                           |
| scientificNameAuthorship | The authorship information for the scientificName formatted according to the conventions of the applicable nomenclaturalCode |
A total of 53,208 glomeromycotan spores, representing 97 distinct morphotypes, were classified from 244 field soil samples. However, only 37, including 18,733 spores, could be classified morphologically at the species level, eight of which were also characterised by molecular methods (Tables 2, 3). The families with most AMF species were Acaulosporaceae (13 spp.), followed by Glomeraceae (6 spp.), Diversisporaceae (4 spp.) and Archaeosporaceae, Claroideoglomeraceae and Gigasporaceae, all with 3 spp. Spores from the Acaulosporaceae were found in almost all samples (85%), followed by those from Gigasporaceae (48%), Glomeraceae (39%), Archaeosporaceae (18%) and Claroideoglomeraceae (13%). Of the 37 AMF identified, 18 AMF occurred only in Terceira (49%), most of which were from Acaulosporaceae. 17 AMF were found in both Islands (46%) and just two from the Paraglomeraceae and Sacculousporaceae, respectively only occurred in São Miguel (5%).

Table 2.
Species occurrence per habitat type in Terceira Island at four sampling dates (Su – August 2007; Au1 – September 2012; Sp – May 2013; Au2 – September 2013): semi-natural pastures (SPPG; SPTB); intensively-managed pastures (IPR1; IPR2) and native forests of J. brevifolia (NFPG; NFLA) and P. azorica (NFTB; NFSE). (*) Indicates de AMF species, characterised by molecular methods.

| Terceira | AMF | SPPG | SPTB | IPR1 | IPR2 | NFPG | NFLA | NFTB | NFSE |
|----------|-----|------|------|------|------|------|------|------|------|
|          |     | Su   | Su   | Su   | Su   | Au1  | Au2  | Au1  | Au2  |
| Acaulospora brasiliensis * | -    | -    | -    | -    | -    | +    | +    | +    | +    |
| Acaulospora cavernata * | -    | -    | -    | -    | -    | -    | -    | +    | +    |
| Acaulospora delicata | +    | -    | -    | -    | -    | -    | -    | +    | -    |
| Acaulospora elegans | -    | -    | +    | -    | -    | -    | -    | +    | -    |
| Acaulospora excavata | -    | +    | -    | -    | -    | -    | -    | -    | -    |
| AMF                     | SPPG | SPTB | IPR1 | IPR2 | NFPG | NFLA | NFTB | NFSE |
|------------------------|------|------|------|------|------|------|------|------|
| Su                     | Su   | Su   | Su   | Su   | Au   | Au   | Sp   | Sp   |
| **Acaulospora koskei** | *    | +    | -    | +    | +    | -    | -    | -    |
| **Acaulospora lacunosa** | -    | -    | -    | -    | +    | +    | +    | +    |
| **Acaulospora laevis** | *    | +    | +    | +    | -    | -    | -    | -    |
| **Acaulospora mellea** | *    | -    | -    | -    | -    | -    | +    | +    |
| **Acaulospora paulinae** | +    | +    | +    | +    | -    | -    | -    | -    |
| **Acaulospora spinosa** | -    | -    | -    | -    | +    | +    | +    | +    |
| **Acaulospora thomii** | +    | -    | -    | -    | -    | -    | -    | -    |
| **Acaulospora tuberculata** | -    | +    | -    | -    | -    | -    | -    | -    |
| **Ambispora appendicula** | -    | -    | -    | +    | -    | -    | -    | +    |
| **Ambispora fennica** | -    | -    | -    | -    | -    | -    | -    | +    |
| **Archaeospora myriocarpa** | +    | +    | +    | +    | -    | -    | -    | -    |
| **Archaeospora schenckii** | +    | -    | -    | -    | -    | -    | -    | -    |
| **Archaeospora trappei** | -    | -    | -    | -    | +    | -    | -    | -    |
| **Cetraspora pellucida** | +    | +    | +    | +    | -    | -    | -    | -    |
| **Claroideoglomus claroideum** | -    | -    | -    | -    | -    | -    | -    | +    |
| **Claroideoglomus etunicatum** | -    | -    | +    | -    | -    | -    | -    | -    |
| **Claroideoglomus lamellosum** | -    | -    | +    | -    | -    | -    | -    | -    |
| **Corymbiglomus globiferum** | -    | -    | +    | -    | -    | -    | -    | -    |
| **Diversispora celata** | -    | +    | -    | -    | -    | -    | -    | -    |
| **Diversispora epigaea** | -    | -    | -    | -    | -    | -    | -    | -    |
| **Diversispora spurca** | -    | -    | -    | +    | -    | -    | -    | -    |
| **Entrospora infrequens** | -    | -    | +    | -    | -    | -    | -    | -    |
| **Funneliformis mosseae** | -    | -    | -    | -    | -    | -    | -    | -    |
| **Gigaspora margarita** | -    | -    | -    | -    | -    | -    | -    | +    |
| **Paraglomus brasiliatum** | -    | -    | +    | -    | -    | -    | -    | -    |
| **Rhizophagus clarus** | +    | +    | +    | +    | -    | -    | -    | +    |
| **Sclerocystis rubiformis** | +    | +    | +    | +    | +    | -    | -    | +    |
| **Sclerocystis sinuosa** | -    | -    | -    | -    | +    | -    | -    | -    |
### Terceira

| AMF                        | SPPG | SPTB | IPR1 | IPR2 | NFPG | NFLA | NFTB | NFSE | Su | Su | Su | Au | Sp | Au1 | Sp | Au2 | Sp | Au1 | Sp | Au2 | Sp | Au2 |
|----------------------------|------|------|------|------|------|------|------|------|----|----|----|----|----|-----|----|-----|----|-----|----|-----|----|-----|
| *Scuttelospora calospora*  | +    | +    | +    | -    | -    | -    | -    | -    | +  | +  | +  | +  | +  | +   | +  | +   | +  | +   | +  | +   |
| *Septoglomus constrictus*  | -    | -    | -    | -    | -    | -    | -    | -    | -  | -  | -  | +  | -  | -   | +  | +   | -  | +   | -  | +   |

### São Miguel

| AMF                        | NFLO | NFTR | NFLG | NFRQ |
|----------------------------|------|------|------|------|
| Au1 | Sp | Au2 | Au1 | Sp | Au2 | Au1 | Sp | Au2 | Au1 | Sp | Au2 | Au1 | Sp | Au2 |
| *Acaulospora brasiliensis*  | +    | +    | +    | +    | +    | +    | -    | -    | -   | -   | -   | -   | -   | -   |
| *Acaulospora cavernata*     | -    | -    | -    | -    | -    | -    | -    | -    | -   | -   | -   | -   | -   | -   |
| *Acaulospora koskea*        | +    | -    | +    | +    | +    | +    | -    | -    | -   | -   | -   | -   | -   | -   |
| *Acaulospora laevis*        | +    | -    | -    | -    | -    | -    | -    | -    | -   | -   | -   | -   | -   | -   |
| *Acaulospora mellea*        | -    | -    | -    | -    | -    | -    | +    | +    | +   | +   | +   | +   | +   | +   |
| *Ambispora appendicula*     | -    | +    | +    | -    | +    | +    | -    | -    | -   | -   | -   | -   | -   | -   |
| *Ambispora fennica*         | -    | -    | -    | -    | -    | -    | +    | -    | -   | -   | -   | -   | -   | -   |
| *Archaeospora trappei*      | +    | -    | -    | +    | -    | -    | +    | +    | +   | +   | +   | +   | +   | +   |
| *Claroideoglomus claroideum*| -    | -    | -    | +    | -    | -    | +    | +    | +   | +   | +   | +   | +   | +   |
| *Diversispora celata*       | -    | +    | -    | -    | -    | -    | -    | -    | -   | -   | -   | -   | -   | -   |
| *Diversispora epigaea*      | -    | +    | +    | -    | -    | -    | -    | -    | -   | -   | -   | -   | -   | -   |
| *Gigaspora margarita*       | -    | -    | -    | -    | -    | -    | +    | +    | +   | +   | +   | +   | +   | +   |
| *Paraglomus albidum*        | -    | -    | -    | -    | -    | -    | -    | -    | -   | -   | -   | -   | -   | -   |
| *Rhizophagus clarus*        | -    | +    | +    | +    | +    | -    | -    | -    | -   | -   | -   | -   | -   | -   |
| *Sacculospora baltica*      | -    | -    | -    | +    | +    | -    | -    | -    | -   | -   | -   | -   | -   | -   |
| *Sclerocystis rubiformis*    | +    | -    | -    | +    | -    | -    | -    | -    | -   | -   | -   | -   | -   | -   |
| *Sclerocystis sinuosa*      | -    | +    | -    | -    | -    | -    | -    | -    | -   | -   | -   | -   | -   | -   |
| *Scuttelospora calospora*   | -    | -    | +    | +    | +    | -    | -    | -    | -   | -   | -   | -   | -   | -   |
| *Septoglomus constrictus*   | -    | +    | +    | -    | -    | -    | +    | +    | +   | +   | +   | +   | +   | +   |

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**Table 3.**

Species occurrence per habitat type in São Miguel Island at three sampling dates (Au1 – September 2012; Sp – May 2013; Au2 – September 2013): native forests of *J. brevifolia* (NFLO; NFTR) and *P. azorica* (NFLG; NFRQ). (*) Indicates de AMF species, characterised by molecular methods.
Members of the family Acaulosporaceae occurred almost exclusively in the native forests, especially associated with the P. azorica rhizosphere (Tables 2, 3). The most frequently found members of this family were Ac. brasiliensis (48%), followed by Ac. lacunosa (26%) and Ac. mellea (17%), all exclusively detected in the native forests (Tables 2, 3) (Fig. 3). Five members of this family were only found in pastures, especially in semi-natural areas, including Ac. excavata, Ac. paulinae, Ac. thomii, Ac. tuberculata and Ac. myriocarpa (Table 2), which may indicate a tendency of some members of this family to occupy less-disturbed habitats (Turrini and Giovannetti 2011, Velázquez et al. 2016) (Fig. 3). Moreover, some AMF species were of restricted distribution. Ac. spinosa was detected only in the rhizosphere of P. azorica of Serreta (TER_NT_SE); Ac. thomii occurred only in the semi-natural pasture of Pico Galhardo (TER_SP_PG); and Ac. excavata and Ac. tuberculata were both found only in semi-natural pastures of Terra Brava (TER_SP_TB) (Table 2) (Fig. 3). Within the Glomeraceae, the AMF species with greater occurrence were Rhizophagus.
clarus (20%), Sclerocystis rubiformis (12%) and Septoglomus constrictus (10%) (Tables 2, 3) (Fig. 3). The members of this family were distributed throughout all habitats, confirming the great ecological plasticity of Glomeraceae members to colonise a broad range of habitats (Öpik et al. 2006) (Tables 2, 3). Gigasporaceae was the second most frequent family, being represented by Scutellospora calospora (25%) and by Gi. margarita (24%) (Tables 2, 3) (Fig. 3). Members of this family also occurred in less disturbed habitats, particularly in the native forests of P. azorica (e.g. Gi. margarita) (Tables 2, 3) (Fig. 3), although some members of this family were also detected in semi-natural pastures (Table 2). The most frequent members of the family Archaeosporaceae was Ar. myriocarpa (9%) detected only in pasture systems (Table 2) and Ar. trappei (8%) only found in native forests (Tables 2, 3). Within the Claroideoglomeraceae, the most frequent species was Cl. claroideum (8%), detected only in the native forests (Tables 2, 3) and Cl. etunicatum (5%) only found in intensively-managed pastures (Table 2) (Fig. 3).

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Author contributions

Conceptualisation, CDM; Investigation, CDM; Formal Analysis, CDM and PAVB; Writing—Original Draft Preparation, CDM; Writing-Review & Editing, CDM, CW, HF and PAVB; Funding Acquisition, AM; and CW and CDM performed AMF species identification. All authors interpreted the results and contributed to the final manuscript and CDM led the writing of the manuscript. All authors gave final approval for publication.
References

- Babu AG, Reddy SM (2011) Diversity of arbuscular mycorrhizal fungi associated with plants growing in fly ash pond and their potential role in ecological restoration. Current Microbiology 63 (3): 273-280. https://doi.org/10.1007/s00284-011-9974-5
- Barea JM, Pozo MJ, Azcón R, Azcón-Aguilar C (2005) Microbial co-operation in the rhizosphere. Journal of Experimental Botany 56 (417): 1761-1778. https://doi.org/10.1093/jxb/eri197
- Barea JM, Palenzuela J, Cornejo P, Sánchez-Castro I, Navarro-Fernández C, Lopéz-García A, Estrada B, Azcón R, Ferrol N, Azcón-Aguilar C (2011) Ecological and functional roles of mycorrhizas in semi-arid ecosystems of Southeast Spain. Journal of Arid Environments 75 (12): 1292-1301. https://doi.org/10.1016/j.jaridenv.2011.06.001
- Blaszkowski J (2012) Glomeromycota. W. Szafer Institute of Botany Polish Academy of Sciences, Krakow, 304 pp.
- Borges PAV (1997) Pasture arthropod community structure in Azorean islands of different geological age. Ph.D. Dissertation, Imperial College, University of London, London.
- Borges PAV, Aguiar C, Amaral J, Amorim IR, André G, Arraiol A, Baz A, Dinis F, Enghoff H, Gaspar C, Ilharco F, Mahnert V, Melo C, Pereira F, Quattau JA, Ribeiro SP, Ribes J, Serrano ARM, Sousa AB, Strassen RZ, Vieira L, Vieira V, Vitorino A, Wunderlich J (2005) Ranking protected areas in the Azores using standardised sampling of soil epigean arthropods. Biodiversity and Conservation 14 (9): 2029-2060. https://doi.org/10.1007/s10531-004-4283-y
- Brundrett MC, Tedersoo L (2018) Evolutionary history of mycorrhizal symbioses and global host plant diversity. New Phytologist 220 (4): 1108-1115. https://doi.org/10.1111/nph.14976
- Cardoso P, Aranda S, Lobo J, Dinis F, Gaspar C, Borges PAV (2009) A spatial scale assessment of habitat effects on arthropod communities of an oceanic island. Acta Oecologica 35 (5): 590-597. https://doi.org/10.1016/j.actao.2009.05.005
- Chen M, Arato M, Borghi L, Nouri E, Reinhardt D (2018) Beneficial services of arbuscular mycorrhizal fungi – from ecology to application. Frontiers in Plant Science 9 https://doi.org/10.3389/fpls.2018.01270
- Chitarra W, Pagliarani C, Maserti B, Lumini E, Siciliano I, Cascone P, Schubert A, Gambino G, Balestrini R, Guerrieri E (2016) Insights on the Impact of arbuscular mycorrhizal symbioses on tomato tolerance to water stress. Plant Physiology https://doi.org/10.1104/pp.16.00307
- Dias E (1996) Vegetação Natural dos Açores. Ecologia e sintaxonomia das florestas naturais. Ph.D. Dissertation, University of Azores, Ponta Delgada.
- Elias R, Gil A, Silva L, Fernández-Palacios JM, Azevedo EB, Reis F (2016) Natural zonal vegetation of the Azores Islands: characterization and potential distribution. Phytocoenologia 46 (2): 107-123. https://doi.org/10.1127/phyto/2016/0132
- Ferrol N, Calvente R, Cano C, Barea J, Azcón-Aguilar C (2004) Analysing arbuscular mycorrhizal fungal diversity in shrub-associated resource islands from a desertification-threatened semiarid Mediterranean ecosystem. Applied Soil Ecology 25 (2): 123-133. https://doi.org/10.1016/j.apsoil.2003.08.006
• Gilmore AE (1968) Phycomycetous mycorrhizal organisms collected by open-pot culture methods. Hilgardia 39 (4): 87-105. https://doi.org/10.3733/hilg.v39n04p087
• Göhre V, Paszkowski U (2006) Contribution of the arbuscular mycorrhizal symbiosis to heavy metal phytoremediation. Planta 223 (6): 1115-1122. https://doi.org/10.1007/s00425-006-0225-0
• Jeffries P, Gianinazzi S, Perotto S, Turnau K, Barea J (2003) The contribution of arbuscular mycorrhizal fungi in sustainable maintenance of plant health and soil fertility. Biology and Fertility of Soils 37: 1-16. https://doi.org/10.1007/s00374-002-0546-5
• Koske RE, Walker C (1985) Species of Gigaspora (Endogonaceae) with Roughened Outer Walls. Mycologia 77 (5): 702-720. https://doi.org/10.1080/00275514.1985.12025159
• Krüger M, Krüger C, Walker C, Stockinger H, Schüssler A (2012) Phylogenetic reference data for systematics and phylotaxonomy of arbuscular mycorrhizal fungi from phylum to species level. The New Phytologist 193 (4): 970-84. https://doi.org/10.1111/j.1469-8137.2011.03962.x
• Li X, Gai J, Cai X, Li X, Christie P, Zhang F, Zhang J (2013) Molecular diversity of arbuscular mycorrhizal fungi associated with two co-occurring perennial plant species on a Tibetan altitudinal gradient. Mycorrhiza 24 (2): 95-107. https://doi.org/10.1007/s00572-013-0518-7
• Martins AM (1993) The Azores-Westernmost Europe: where evolution can be caught red-handed. Boletim do Museu Municipal do Funchal 2: 181-198. URL: http://dspace.cm-funchal.pt:8080/jspui/handle/100/1063
• Martins J, Moreira O, Silva L, Moura M (2011) Vegetative propagation of the endangered Azorean tree, Picconia azorica. Life and Marine Sciences 28: 39-46.
• Melo CD, Walker C, Rodríguez-Echeverría S, Borges PAV, Freitas H (2014) Species composition of arbuscular mycorrhizal fungi differ in semi-natural and intensively managed pastures in an isolated oceanic island (Terceira, Azores). Symbiosis 64 (2): 73-85. https://doi.org/10.1007/s13199-014-0303-1
• Melo CD, Luna S, Krüger C, Walker C, Mendonça D, Fonseca HA, Jaizme-Vega M, Machado AC (2017) Arbuscular mycorrhizal fungal community composition associated with Juniperus brevifolia in native Azorean forest. Acta Oecologica 79: 48-61. https://doi.org/10.1016/j.actao.2016.12.006
• Melo CD, Luna S, Krüger C, Walker C, Mendonça D, Fonseca HAC, Jaizme-Vega M, Machado AC (2018) Communities of arbuscular mycorrhizal fungi under Picconia azorica in native forests of Azores. Symbiosis 74 (1): 43-54. https://doi.org/10.1007/s13199-017-0487-2
• Melo CD, Walker C, Freitas H, Machado AC, Borges PAV (2019) Distribution of arbuscular mycorrhizal fungi in Terceira and São Miguel (Azores, Portugal). 1.8. GBIF. Release date: 2019-12-20. URL: https://doi.org/10.15468/fypcjn
• Moreira OCB, Martins J, Silva L, Moura M (2012) Seed germination and seedling growth of the endangered azorean cherry Prunus azorica. HortScience 47 (9): 1222-1227. https://doi.org/10.21273/hortsci.47.9.1222
• Öpik M, Moora M, Liira J, Zobel M (2006) Composition of root-colonizing arbuscular mycorrhizal fungal communities in different ecosystems around the globe. Journal of Ecology 94 (4): 778-790. https://doi.org/10.1111/j.1365-2745.2006.01136.x
• Redecker D, Schüßler A, Stockinger H, Stürmer S, Morton J, Walker C (2013) An evidence-based consensus for the classification of arbuscular mycorrhizal fungi
(Glomeromycota). Mycorrhiza 23 (7): 515-531. https://doi.org/10.1007/s00572-013-0486-y

- Rillig M, Mummey D (2006) Mycorrhizas and soil structure. New Phytologist 171 (1): 41-53. https://doi.org/10.1111/j.1469-8137.2006.01750.x

- Schüßler A, Krüger M, Walker C (2011) Revealing natural relationships among arbuscular mycorrhizal fungi: Culture line BEG47 represents Diversispora epigaea, not Glomus versiforme. PLOS One 6 (8): e23333. https://doi.org/10.1371/journal.pone.0023333

- Schüßler A, Walker C (2019) Archaeospora ecuadoriana sp. nov. from a mountainous biodiversity hotspot area in Ecuador, and transfer of Palaeospora spainiae to Archaeospora, as A. spainiae comb. nov. Mycorrhiza 29 (5): 435-443. https://doi.org/10.1007/s00572-019-00913-2

- Silva L (2001) Plantas invasoras no Arquipélago dos Açores: caracterização geral e estudo de um caso, Clethra arborea Aiton (Clethraceae). Ph.D. Dissertation. University of Azores, Ponta Delgada.

- Terzopoulou S, Rigal F, Whittaker RJ, Borges PAV, Triantis KA (2015) Drivers of extinction: the case of Azorean beetles. Biology Letters 11 (6). https://doi.org/10.1098/rsbl.2015.0273

- Turrini A, Giovannetti M (2011) Arbuscular mycorrhizal fungi in national parks, nature reserves and protected areas worldwide: a strategic perspective for their in situ conservation. Mycorrhiza 22 (2): 81-97. https://doi.org/10.1007/s00572-011-0419-6

- van der Heijden MA, Martin F, Selosse M, Sanders I (2015) Mycorrhizal ecology and evolution: the past, the present, and the future. New Phytologist 205 (4): 1406-1423. https://doi.org/10.1111/nph.13288

- Velázquez MS, Stürmer SL, Bruzone C, Fontenla S, Barrera M, Cabello M (2016) Occurrence of arbuscular mycorrhizal fungi in high altitude sites of the Patagonian Altoandina region in Nahuel Huapi National Park (Argentina). Acta Botanica Brasilia 30 (4): 521-531. https://doi.org/10.1590/0102-33062016abb0223

- Vos CM, Tesfahun AN, Panis B, De Waele D, Elsen A (2012) Arbuscular mycorrhizal fungi induce systemic resistance in tomato against the sedentary nematode Meloidogyne incognita and the migratory nematode Pratylenchus penetrans. Applied Soil Ecology 61: 1-6. https://doi.org/10.1016/j.apsoil.2012.04.007

- Walker C, Trappe JM (1981) Acaulospora spinosa sp. nov. with a key to the species of Acaulospora. Mycotaxon 12 (2): 515-521.

- Walker C, Reed LE, Sanders FE (1984) Acaulospora nicolsonii, a new endogonaceous species from Great Britain. Transactions of the British Mycological Society 83 (2): 360-364. https://doi.org/10.1016/s0007-1536(84)80165-5

- Walker C, Pfeiffer CM, Bloss HE (1986) Acaulospora delicata sp. nov. – an endomycorrhizal fungus from Arizona. Mycotaxon 25: 621-628.

- Walker C, Diedrichs C (1989) Scutellospora scutata sp. nov., a newly described endomycorrhizal fungus from Brazil. Mycotaxon 35: 357-361.

- Walker C (1992) Systematics and taxonomy of the arbuscular endomycorrhizal fungi (Glomales)- a possible way forward. Agronomie 12: 887-897. https://doi.org/10.1051/agro:19921026

- Walker C, Vestberg M (1998) Synonymy amongst the arbuscular mycorrhizal fungi: Glomus claroideum, G. multisubstenum and G. fistulosum. Annals of Botany 82 (5): 601-624. https://doi.org/10.1006/anbo.1998.0714
• Walker C, Harper CJ, Brundrett MC, Krings M (2018) Chapter 20 - Looking for Arbuscular Mycorrhizal Fungi in the Fossil Record: An Illustrated Guide. In: Krings M, Harper C, Cúneo N, Rothwell G (Eds) Transformative Paleobotany. Academic Press, 481–517 pp. [ISBN 9780128130124]. https://doi.org/10.1016/B978-0-12-813012-4.00020-6

• Wijayawardene NN, Pawlowska J, Letcher PM, Kirk PM, Humber RA, Schüssler A, Wrzosek M, Muszewska A, Okrasińska A, Istel Ł, Gęsiorska A, Mungai P, Lateef AA, Rajeshkumar KC, Singh RV, Radek R, Walther G, Wagner L, Walker C, Wijesundara DSA, Papizadeh M, Dolatabadi S, Shenoy BD, Tokarev YS, Lumyong S, Hyde KD (2018) Notes for genera: basal clades of Fungi (including Aphelidiomycota, Basidiobolomycota, Blastocladiomycota, Calcarisporiellomycota, Caulochytriomycota, Chytridiomycota, Entomophthoromycota, Glomeromycota, Kickxellomycota, Monoblepharomycota, Mortierellomycota, Mucoromycota, Neocallimastigomycota, Olpidiomycota, Rozellomycota and Zoopagomycota). Fungal Diversity 92 (1): 43-129. https://doi.org/10.1007/s13225-018-0409-5