Role of the polypeptide N-acetylgalactosaminyltransferase 3 in ovarian cancer progression: possible implications in abnormal mucin O-glycosylation

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Keywords: GALNT3, epithelial ovarian cancer; DNA hypomethylation, progression-free survival, microarrays, mucin O-glycosylation

Received: December 3, 2013  Accepted: January 25, 2014  Published: January 27, 2014

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ABSTRACT:

Previously, we have identified the polypeptide N-acetylgalactosaminyltransferase 3 (GALNT3) gene as notably hypomethylated in low-malignant potential (LMP) and high-grade (HG) serous epithelial ovarian tumors, compared to normal ovarian tissues. Here we show that GALNT3 is strongly overexpressed in HG serous EOC tumors as compared to normal ovarian tissue. Moreover, the GALNT3 expression significantly correlated with shorter progression-free survival (PFS) intervals in epithelial ovarian cancer (EOC) patients with advanced disease.

Knockdown of the GALNT3 expression in EOC cells led to sharp decrease of cell proliferation and induced S-phase cell cycle arrest. Additionally, GALNT3 suppression significantly inhibited EOC cell migration and invasion. Gene expression profiling and consecutive network and pathway analyses confirmed these findings, as numerous genes and pathways known previously to be implicated in ovarian tumorigenesis, including EOC tumor invasion and metastasis, were found to be downregulated upon GALNT3 suppression, while some tumor suppressor genes were induced. Moreover, GALNT3 downregulation was associated with reduced MUC1 protein expression in EOC cells, probably related to destabilization of the MUC1 protein due to lack of GALNT3 glycosylation activity. GALNT3 knockdown was also accompanied with increase of the cell adhesion molecules β-catenin and E-cadherin, which are normally suppressed by MUC1 in cancer, thus supporting the role of the GALNT3-MUC1 axis in EOC invasion.

Taken together, our data are indicative for a strong oncogenic potential of the GALNT3 gene in advanced EOC and identify this transferase as a novel EOC biomarker and putative EOC therapeutic target. Our findings also suggest that GALNT3 overexpression might contribute to EOC progression through aberrant mucin O-glycosylation.

INTRODUCTION

Epithelial ovarian cancer (EOC) accounts for 4% of all cancers in women and is the leading cause of death from gynecologic malignancies [1]. Despite treatment improvements, long-term survival rates for patients with advanced disease remain disappointing [2]. The molecular basis of EOC initiation and progression is still poorly understood. To establish novel therapeutic and diagnostic strategies against this deadly disease, it is essential to understand its molecular pathology.

Disruption of normal gene regulation is important for carcinogenesis resulting in loss, or gain of genetic function. Recently, the importance of epigenetic
perturbation of gene regulation in cancer [3], including EOC [4], has begun to be more fully appreciated. The most studied epigenetic alteration is DNA methylation, the addition of a methyl moiety to the cytosine-5 position within the context of a Cpg dinucleotide, mediated by DNA methyltransferases [3]. DNA methylation patterns are reset early in the embryogenesis and reestablished early during development. After that, they are thought to be relatively stable. In cancer, the physiological regulation of DNA methylation is disrupted leading to drastic changes of the distribution pattern of 5-methylcytosine. The heavy methylation found in the bulk of chromatin is reduced, while the normally unmethylated Cpg islands located in the promoter and first exon of genes become hypermethylated. Promoter hypermethylation often leads to inactivation of different tumor-suppressing genes and is associated with many important pathways involved in cancer, such as DNA repair, cell cycle regulation, apoptosis, carcinogen metabolism, hormonal response, and cell adherence [5]. Aberrant DNA methylation is also involved in the development of resistance to chemotherapy (CT) [6]. The role of DNA hypomethylation in carcinogenesis is less studied. Recent studies have demonstrated that global decrease in the level of DNA methylation is related to hypomethylation of repeated sequences, increase in genetic instability, as well as re-activation of proto-oncogenes and pro-metastasis genes (reviewed in [7]).

Similar to other malignancies, aberrant DNA methylation, including global hypomethylation of heterochromatin and local Cpg island methylation, occurs in EOC and contributes to ovarian tumorigenesis and mechanisms of chemoresistance [4]. Applying a more global array-based technology, several studies have demonstrated that DNA methylation changes in ovarian cancer are cumulative with disease progression and CT resistance [8-10]. Using a similar approach (methylated DNA immunoprecipitation coupled to Cpg island tiling arrays) we have recently shown that DNA hypermethylation occurs in less invasive/early stages of ovarian tumorigenesis, while advanced disease was associated with DNA hypomethylation of a number of oncogenes, implicated in cancer progression, invasion/metastasis and probably chemoresistance [11]. The polypeptide N-acetylgalactosaminyltransferase 3 gene (GalNAc-T3 gene, also known as GALNT3) was among the genes identified to be notably hypomethylated in low-malignant potential (LMP) and high grade (HG) serous EOC tumors [11]. The GALNT3 gene is a member of the GalNAc-transferases (GALNAC-Ts) gene family; the genes of this family conduct the transfer of N-acetyl galactosamine (GalNAc) to the hydroxyl group of a serine or threonine residue in the first step of O-linked oligosaccharide biosynthesis [12]. So far, 20 members of the GALNAC-Ts gene family have been identified and most of them encode an active polypeptide GALNT functioning in the primary step of the O-glycosylation of different proteins, including mucins [13]. Aberrant mucin-type O-glycosylation represents one of the most abundant posttranslational cancer-associated changes, comprising diverse biologic and pathologic consequences influencing growth and survival of cancer cells and their ability for invasion and metastasis [14]. The membrane-associated mucin-1 (MUC1), as one highly glycosylated protein, is overexpressed in more than 90% of high-grade EOC tumors including metastatic lesions, as MUC1 expression correlated with EOC progression [15].

This prompted us to further investigate if GALNT3 displays elevated expression levels in serous EOC tumors with different malignant potential, and whether this gene is functionally implicated in EOC tumorigenesis, including disease progression and response to treatment. Here we present experimental data, which strongly suggest that GALNT3 is overexpressed HG serous EOC tumors compared to LMP tumors and normal tissues, which probably correlates with its hypomethylated status. We also provide evidence that the GALNT3 gene is involved in EOC cell proliferation and migration/invasion, due to its possible role in MUC1 protein stabilization.

RESULTS

Overexpression of GALNT3 in HG serous EOC tumors: correlation with progression-free survival

Using an epigenomics approach, we have previously identified the GALNT3 gene as hypomethylated in LMP and HG EOC tumors, when compared to normal tissues [11]. Here, we further validated the GALNT3 methylation in independent set of EOC tumors (including LMP and HG tumors) using alternative approach (BSP sequencing). Our BSP analyses confirmed the GALNT3 hypomethylation status in both LMP (including borderline and G1) and HG (G3) serous EOC tumors (see Supplemental Figure 1). We consecutively evaluated GALNT3 protein expression by IHC in serous EOC tumors and ovarian normal tissue samples, using tissue microarrays (TMAs). Our TMAs included triplicate cores of 117 serous EOC tumors, including 13 LMP tumors and 104 HG ovarian tumors. Thirteen normal ovarian tissue samples were also included as controls. Table 1 shows the major clinical characteristics of these patients for whom extensive follow-up clinical data (up to 5-years) were available. The age ranged from 41 to 83 years (median: 66 years). High-grade tumors were all grade 3 (100%) including stage III (69%) and stage IV (31%) tumors. The majority of patients (93%) received a combination of platinum and paclitaxel. The median baseline CA125 was around 800. Forty percent of the patients had a progression or a recurrence within the first 6 months of follow-up; for 39% of the patients the

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progression-free survival (PFS) interval was in the range of 7 to 24 months, and 21% of the patients displayed PFS values higher than 25 months (Table 1).

As seen from Figure 1, GALNT3 displayed significantly higher expression only in HG serous EOC tumors, when compared to normal tissues \((p = 0.0018)\) and LMP tumors \((p = 0.035)\). We further constructed Kaplan–Meier survival curves based on the GALNT3 expression analyses in the cohort of 103 HG serous EOC patients. Our analyses revealed that GALNT3 expression displayed significant inverse association with PFS of serous ovarian adenocarcinoma patients with advanced disease; i.e., women with lower GALNT3 expression had a better survival without progression than those with higher GALNT3 expression \((p=0.034\); see Fig. 1C).

**Phenotype analysis of GALNT3 suppression in EOC cells: possible implications in EOC cell proliferation, cell cycle control, migration and invasion**

Next, we decided to verify if shRNA-mediated GALNT3 gene knockdown could produce any cancer-related phenotypic changes in EOC cells. We tested several EOC cell lines for endogenous GALNT3 protein expression by Western blot analysis (see Figure 8A). Among these, the A2780s cell line displayed strong GALNT3 expression and was further used to generate stably transfected shRNA-GALNT3 clones. Clone selection for further analyses was based on sqRT-PCR and Western blot validation of the GALNT3 mRNA/protein expression in selected clones, compared to empty vector-transfected clones. Among the analyzed clones, the shRNA-GALNT3 knockdown clones sh-G1 and sh-G2 displayed a significant decrease of GALNT3 expression levels compared to the mock-transfected control (Supplemental Figure 2) and were selected for further analyses.

We investigated the impact of the GALNT3 gene suppression on A2780s cell proliferation, cell cycle control, migration, and sensitivity to cisplatin and paclitaxel (drugs, conventionally used for first-line EOC CT). The GALNT3 gene knockdown led to sharp decrease of the number of viable adherent cells (represented by cell index), compared to control cells (Figure 2A). This observation was further supported by the colony formation assay showing that the numbers of clones formed by cells with stably reduced GALNT3 expression were significantly lower compared to control cells (Figure 2B and 2C). Taken together, our observations strongly indicate an influence of GALNT3 on EOC cell proliferation and further on their propensity to form colonies. Moreover, when compared with the control clone, the shRNA-GALNT3 clone sh-G1 exhibited a significant accumulation of cells in the S phase at 6 and 9 hours after removing hydroxyurea (Figure 3). These data indicate that GALNT3 depletion induces S cell cycle arrest which could explain the drastic reduction in the proliferation rates of these cells observed earlier.

Additionally, the GALNT3 suppression significantly inhibited both migration and invasion of A2780s cells. As shown in Figure 4A and 4B, the numbers of A2780

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**Figure 1: Analysis of GALNT3 expression in serous EOC tumors by IHC: correlation with PFS.**

A. Representative IHC images of GALNT3 protein expression in normal ovarian tissues, low-malignant potential (LMP) tumors and high-grade (HG) tumors. B. Box-plot presentation of GALNT3 protein expression levels in normal ovarian tissues, LMP tumors and HG tumors. C. Kaplan-Meier curves for PFS according to the level of GALNT3 IHC intensity in tumor samples of 103 serous EOC patients with advanced disease.

**Figure 2: ShRNA-mediated knockdown of the GALNT3 expression in A2780s cells:**

A. Effect of shRNA-GALNT3 knockdown clones 1 and 2 (sh-G1 and sh-G2) on cell proliferation, compared to the control clone (Ctrl); B. Representative images of colony formation assays following GALNT3 knockdown; C. Quantitative determinations (graph-bars) of data obtained: results are expressed as numbers of colonies formed in the GALNT3 knockdown clones sh-G1- and sh-G2 compared to the ctrl clone. Differences were determined using the Student's \(t\)-test. Error bars denote mean ± SEM; *indicates statistical significance \((p < 0.05)\).
cells that passed through the filter using the sh-G1 and sh-G2 clones were remarkably less than that in the negative control (ctrl) clone, which is indicative for a role for GALNT3 in the regulation of invasion and migration in EOC.

Finally, GALNT3 suppression had no significant impact on A2780s cisplatin and paclitaxel sensitivity (see Supplemental Figure 3).

Molecular mechanisms of GALNT3 action in EOC cells

To better understand the molecular mechanisms of GALNT3 action in A2780s cells, we employed the Agilent Whole Human Genome microarrays, containing 44,000 genes to identify global gene expression changes upon GALNT3 suppression in A2780s cells. We compared the gene expression of the previously selected shRNA-GALNT3 clones (sh-G1 and sh-G2) against the corresponding control clone (ctrl). All microarray experiments were performed in duplicates, as two hybridizations were carried out for the GALNT3 suppressing cell clone against the corresponding control, with extended follow-up, including PFS values, were available for 103 patients.

| Variable           | Range | n/total | %   |
|--------------------|-------|---------|-----|
| Age (years)        | ≥65   | 64/130  | 49  |
|                    | <65   | 66/130  | 51  |
| Median age         |       | 66      |     |
| Tissue/tumor type  | Normal| 13/130  | 10  |
|                    | LMP   | 13/130  | 10  |
|                    | High-grade | 104/130  | 80  |
| Grade              | 3     | 104/104 | 100 |
| Stage              | III   | 72/104  | 69  |
|                    | IV    | 32/104  | 31  |
| Chemotherapy       | platinum+taxol | 97/104  | 93  |
| Other              |       | 13/104  | 7   |
| CA125              | ≥800  | 47/104  | 45  |
|                    | <800  | 53/104  | 55  |
| PFS (months) *     | 0-6   | 41/103  | 40  |
|                    | 7-24  | 40/103  | 39  |
|                    | > 25  | 22/103  | 21  |


*Extended follow-up, including PFS values, were available for 103 patients.

**Figure 3: Effect of shRNA-mediated GALNT3 on cell cycle control in A2780s cells.** Cell-cycle profile was examined by flow cytometry and percentages of cells in G0/G1, S, and G2/M phase in the shRNA-GALNT3 clone 1 (sh-G1) were compared to the mock-transfected control (ctrl) clone. Propidium iodide staining shows an increased fraction of cells in the S-phase and corresponding decrease of cells in both G1- and G2/M-phase at 6 and 9 h after removing hydroxyurea in the shRNA-GALNT3 clone G1 (sh-G1), when compared with the control clone (ctrl).

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using a fluorescent dye reversal (dye-swap) technique. For each comparison, a subset of differentially expressed genes was selected displaying at least 1.5-fold difference in both duplicate microarray experiments. Using these selection criteria, we found 98 genes were upregulated and 375 were downregulated in A2780s cells upon \textit{GALNT3} knockdown. Table 2 shows a list of selected functionally related groups of genes that were differentially expressed (≥ 1.5-fold) in A2780s cells following \textit{GALNT3} suppression. As seen from Table 2, genes with previously shown implication in mechanisms of cell adhesion, cell development, cell growth & proliferation, transport, immune & inflammatory response, metabolism, regulation of transcription and signal transduction, were

### Table 2: Selected differentially expressed gene groups in A2780s cells upon GALNT3 knockdown.

| A. Upregulated genes | B. Downregulated genes |
|----------------------|------------------------|
| **cell adhesion**    | **cell adhesion**       |
| ITGA1, PCDH17, KITLG, NCAM21, AMIG02, PCDH20, PCDH10, PCDH7, HAPLN1, NELL2, MASS1, CCR1 | KERA, THBS1, PALM, KRT8, FLRT3, LAMBI, CXADR, HAPLN3, OMG, PCDH9, CDH2, BA1, plexin C1, NRXN1, NID2, EPDR1, PODXL, COL18A1, COL9A1, COL6A2, SSPN, LUM, TES, KERATIN, FLNC |
| **metabolism**       | **metabolism**          |
| ARSJ, PLAG2A4, TIPARP, DNAJC12, MEOX1, SMA5, TLL1, BCAT1, HS6ST2, LOXL3, PDZRN3, TRIM7, RNF128 | L-arginine, SORL1, MGC:57495, GLT25D2, TRIM34, TRIM43, TRIM6, TRIM49, HSYRA2001105, TRIM48, SMYD3, SOST, MMP10, MMP3, ADPN, NFIB, RBMS1, ERVK6, NAP1L3, UCHL1, UBE1L, GLDC, SUMF1, MMP1, GLUL, PGDS, FLJ10986, PPP1R1A, GFTP2, CHST6, ME3, ST3GAL1, B3GALT3, PRSS12, PSMB9, LRP, CA3, ACYH, ACAS2L, GALNT14, FLJ31568, BASP1, FIGN, TRIM51, NUDT10, OSR2, SCG |
| **regulation of transcription** | **regulation of transcription** |
| SNX10, CHN1, CENTD1, MGST2, NPY1R, SUCNR1, PIK3CA, PPP1B2P2, RGS2, AKA12, RAGE, PKRG1, DNM3, BMP7, DIAPH2, TAOK3, DUSP6, ARRDC3, PRICKLE1, ARRDC3, ARHGP24 | SOX2, MSX1, COL19A1, POSTN, CART1, ZNF426, ZSCAN4, KIAA0222, FOXA1, TAp12A, NFE2L3, ISGF3G, ET7V, NR0B1, AR, IFI16, GATA6, GATA5, NRF2, NFE2, TBX1, MYEF2, LIN28, LHX9, SP100, LHX9, ZNF217, SP140, ARNTL, ZFHX1B, DNAPTP6 |
| **signal transduction** | **signal transduction** |
| SNX10, CHN1, CENTD1, MGST2, NPY1R, SUCNR1, PIK3CA, PPP1B2P2, RGS2, AKA12, RAGE, PKRG1, DNM3, BMP7, DIAPH2, TAOK3, DUSP6, ARRDC3, PRICKLE1, ARRDC3, ARHGP24 | DKK3, DPYSL2, GUCY1B3, S100A11, PLCE1, SPP1, BMP2, BMP4, MAP3K15, PFTK1, MYLK, MARK1, EPB3, PDGFRB, PDGFRRA, PRKCB1, PIM1, PTPRE, PLEKHK1, IRS1, CXXCR4, GPR84, GCG, BA13, GFG18, FGF, CD53, TAC3, NXP2, KIAA1493, ARHGEF4, RHOB, BCR4, RAB3C, BDNF, RGS5, RGS18, SFRP1, MAMDC2, CKB, HRAALS, LOC130576, CKB, GBP4, ANXA3, RAFTLIN, NETO2, MGC52057, ASAM, HHIP, ARPP21, ASAM, mGluR5 |
predominantly or exclusively suppressed, while GALNT3 knockdown was related with the induction of some signal transduction-related genes. Supplemental Table 2 shows the complete list of the differentially expressed genes (≥ 1.5-fold) following GALNT3 knockdown in A2780s cells.

Pathway and network analyses, generated through Figure 5: Functional analysis for a dataset of downregulated genes (≥ 1.5-fold) following GALNT3 suppression in A2780s cells. Top functions that meet a Bonferroni-Holm multiple testing correction p-value of 0.05 are displayed.

Figure 4: Effect of GALNT3 knockdown on A2780s cell migration and invasion. A. Migration was assessed using Boyden-chamber assay. Cells from the shRNA-GALNT3 clones 1 and 2 (sh-G1 and sh-G2) and the control (ctrl) clone were seeded into the upper chambers in 0.1% FBS containing medium at a density of $2.5 \times 10^4$ per well, and 600 μl of 1% FBS containing medium was placed in the lower chamber as a chemoattractant. After 24 h at 37°C in 5% CO2, the cells were fixed with cold methanol and stained with blue trypan solution. Migrated cells on the underside of the filter were photographed and counted by phase contrast microscopy. C. Cell invasion was assayed in a similar way, as the upper chambers were coated with Matrigel. Here, NIH3T3 conditioned medium was added in the lower chamber as a chemoattractant (see Materials and Methods for details). All experiments were performed in triplicate. For each experiment, cell number was calculated as the total count from 10 random fields per filter (at magnification ×40). The bar graphs in panels B and D represent quantitative determinations of data obtained by selecting 10 random fields per filter under phase contrast microscopy and results are expressed as % change of the sh-G1 and sh-G2 clones over the ctrl clone. Differences between shRNA-GALNT3-transfected and vehicle-transfected A2780s cells were determined by a Student’s t-test; error bars denote mean ± SEM; *indicates statistical significance ($p < 0.05$).

Figure 5: Functional analysis for a dataset of downregulated genes (≥ 1.5-fold) following GALNT3 suppression in A2780s cells. Top functions that meet a Bonferroni-Holm multiple testing correction p-value of 0.05 are displayed.
the use of the IPA software confirmed the major functionally related gene groups, found to be differentially expressed in the GALNT3 knockdown clones sh-G1 and sh-G2. Upon using a stringent scoring method (the Bonferroni-Holm multiple testing correction p-value), we have found a number of pathways to be significantly downregulated following GALNT3 suppression (including pathways functionally related to cellular movement, cellular development, cellular growth and proliferation, cellular assembly and organization, cellular function and maintenance, DNA replication, recombination, and repair, cell morphology, cell-to-cell signaling and interaction, cell cycle and gene expression; see Figure 5). However, no significant functional pathways were found to be upregulated upon GALNT3 knockdown when using the same scoring method.

Common networks obtained upon merging the top-scoring networks recognized some important gene nodes and genes that are specifically up- or downregulated upon GALNT3 suppression in A2780s cells (Figure 6). Thus, genes and associated interaction partners that were upregulated upon GALNT3 knockdown (displayed on Figure 6A) comprised members of the ubiquitin C (UBC) and tumor protein p53 (TP53) interaction networks, including genes, predominantly implicated in signal transduction (ANTXR2, ANXA10, ARHGEF15, BMP7, DUSP6, KITLG, PIK3CA, PRKG1, RGS2) and metabolism (ARID5B, BCA1, DPYD, ESRRG, HMGN5, MGST2, PLA2G4A, RNF128, ADAMTS1, UBC). Major gene nodes that were downregulated upon GALNT3 knockdown in A2780s cells are presented in Figure 6B; these were mostly involved in metabolism (MMP1, MMP3, MMP10, SPP1, HTRA1, FOXA1), signal transduction (BMP2, BMP4, CYR61, TGFβ2, BDNF, POSTN, IRS1, IGFBP2) and cell growth & proliferation (CTGF, COL18A1, SFRP1).

**Validation of microarray findings with quantitative PCR (qPCR)**

To validate microarray results, we arbitrarily selected 14 differentially expressed genes and quantified their expression by qPCR in A2780s cells following shRNA-GALNT3 knockdown compared to control (vehicle transfected) A2780s cells. Figure 7 summarizes the gene expression measurements of all validated genes.

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**Figure 6: Network analysis of dynamic gene expression in A2780s cells based on the 1.5-fold common gene expression list obtained following shRNA-mediated GALNT3 knockdown.** A. Upregulated networks; B. Downregulated networks. The three top-scoring networks (upregulated genes) and the five top-scoring networks (downregulated genes) were merged and are displayed graphically as nodes (genes/gene products) and edges (the biological relationships between the nodes). Intensity of the node color indicates the degree of up- (red) or downregulation (green). Nodes are displayed using various shapes that represent the functional class of the gene product (square, cytokine, vertical oval, transmembrane receptor, rectangle, nuclear receptor, diamond, enzyme, rhomboid, transporter, hexagon, translation factor, horizontal oval, transcription factor, circle, other). Edges are displayed with various labels that describe the nature of relationship between the nodes: _ binding only, → acts on. The length of an edge reflects the evidence supporting that node-to-node relationship, in that edges supported by article from literature are shorter. Dotted edges represent indirect interaction.
We found that both methods (microarray analysis and qPCR) detected similar patterns for the up- and down-regulated genes selected for validation.

**Effect of GALNT3 knockdown on MUC1 O-glycosylation and stabilization**

To further elucidate the GALNT3 potential oncogenic potential in EOC, we examined the effect of GALNT3 suppression on the MUC1 mRNA and protein expression in A2780s cells. We initially analyzed the endogenous MUC1 protein expression levels in different EOC cells and found that GALNT3 and MUC1 proteins were co-expressed in the cell lines A2780s and SKOV3 (Figure 8A). Using a semi-quantitative RT-PCR, we found no differences in the MUC1 mRNA expression levels between the shRNA-GALNT3 knockdown A2780s clones (sh-G1 and sh-G2) and the corresponding control (ctrl) A2780s clone (Figure 8B). However, MUC1 displayed considerably lower protein expression levels in the sh-G1 and sh-G2 clones, compared to the ctrl clone (Figure 8C/input). Moreover, VVA lectin pull-down assay for glycosylated proteins was able to detect a MUC1 protein-specific 200 kDa band only in the ctrl clone, but not in the sh-G1 and sh-G2 clones (Figure 8C/pull-down). Similarly, VVA lectin blot following VVA lectin pull-down assay detected GalNAc-conjugated proteins in the range of 165 – 247 kDa exclusively in the ctrl clone (Figure 8D). Taken together, our findings imply that GALNT3 gene may contribute to ovarian carcinogenesis through O-glycosylation and stabilization of the MUC1 oncoprotein and possibly, other mucins. Moreover, because downregulation of MUC1 in cancer cells can inhibit cell migration by promoting the expression of the cell adhesion molecules E-cadherin and β-catenin [16], we also investigated the impact of GALNT3 knockdown on E-cadherin and β-catenin protein expression. As seen from Figure 8E, GALNT3 suppression remarkably augmented the proteins of both these cell adhesion molecules, suggesting that the involvement of the GALNT3-MUC1 pathway in EOC invasion could include the destabilization of the E-cadherin/β-catenin complex formation.

**DISCUSSION**

The mechanisms for the tumorigenesis, progression and biological aggressiveness of EOCs have not been yet fully clarified. We have recently identified GALNT3 gene as a novel potentially hypomethylated target in epithelial ovarian cancer (EOC) by our MeDIP-array experiments and consecutively determined that a 315 nt GALNT3 putative gene control region (located 497 nt downstream of the transcription initiation site, and adjacent to some important control motifs of the GALNT3 promoter domain [17]) was significantly hypomethylated in LMP and HG.
serous EOC tumors, compared to normal tissues [11]. In the present study we demonstrate that GALNT3 is overexpressed exclusively in HG serous ovarian tumors as compared to LMP tumors and normal ovarian tissues; furthermore, we found that GALNT3 expression was significantly associated with PFS in the studied cohort of 103 HG serous EOC patients \( (p = 0.034; \text{see Fig. 1C}) \). Additionally, our functional analyses are strongly indicative for evident oncogenic capacity of GALNT3 in serous EOC, including its potential role in EOC cell proliferation, cell cycle control and cell migration/invasion (see Figs. 2-4). However, our findings suggest that epigenetic mechanisms associated with aberrant DNA hypomethylation might not play important role in controlling the GALNT3 expression in EOC, since we found no correlation between GALNT3 hypomethylation status and its expression in LMP tumors.

To better elucidate the molecular mechanisms and biological pathways implicated in GALNT3-mediated action in EOC cells, we used a complementary gene expression profiling using the DNA microarray technology to monitor cellular changes in gene expression and discover the molecular targets upon GALNT3 suppression. To our knowledge, the present work represents the first effort to define global changes in gene expression upon modulation of GALNT3 gene expression in epithelial cancer cells. The gene expression data and consecutive IPA network and pathway analyses were quite confirmatory of the data obtained by the GALNT3 functional assays. Indeed, microarray data sustained GALNT3 correlation with EOC cell proliferation (including cell cycle control), migration and invasion, since GALNT3 knockdown resulted in reduced expression of genes associated with cell proliferation, cell movement/invasion, and cell cycle control (see Table 2 and Figure 5).

IPA network analysis was indicative for some important gene nodes linked to GALNT3 suppression in EOC cells, as most of these substantiate and/or complement the functional data obtained. Thus, GALNT3 knockdown resulted in upregulation of different members of the p53 and UBC interaction networks, both associated with tumor suppressor activities (Figure 6A). Indeed, p53 is one of the most studied and well characterized tumor suppressor gene in the cancer research field [18], while the UBC interaction network and its members were shown to decrease in anchorage-independent cell growth and increase apoptosis, suggesting UBC may act as a negative regulator of skin carcinogenesis [19]. Interestingly, the p53/UBC interacting partner AKA12 has been characterized as a tumor suppressor gene in many tumor types, acting as an inhibitor of the PKC-Raf/MEK/ERK pro-metastatic pathways [20].

In parallel, upon GALNT3 knockdown, we have observed predominant and strong downregulation of major gene nodes known to be implicated in EOC tumorigenesis (SPP1, CYR61, IRS1, IGFBP2) [21-24], including EOC dissemination/metastasis (MMP1, MMP3, MMP10, BMP2, BMP4, TGFb2, BDNF, POSTN) [25-33]; (see Figure 6B). Additionally, GALNT3 suppression was associated with downregulation of numerous genes from the NfkB pathway (Figure 6B), representing one of the major oncogenic pathways implicated in EOC progression [34] Thus, our findings are strongly indicative for the oncogenic functionality of GALNT3 in EOC carcinogenesis. These results were not unexpected since recent studies suggested that GALNT3 overexpression is predominantly associated with more aggressive tumor behavior and poor outcome in various types of cancer [17, 35-39]. However, GALNT3 expression is also shown to be a negative indicator for disease prognosis in non-small cell lung cancer and colorectal carcinoma [40, 41], including colon carcinoma cells selected for hepatic metastasis [42]. Moreover, dysfunctional GALNT3 gene causes the rare autosomal recessive metabolic disorder familial tumoral calcinosis [43]. The above data suggest that GALNT3 can display distinct oncological roles in different cancer types.

The various genes encoding GALNAC-Ts are differentially expressed in malignant tissue compared to normal tissue and were consistently shown to play distinct roles in cancerogenesis and tumor metastasis. Thus, GALNT6 and GALNT13 display elevated levels in different cancer types, including breast and gastric carcinomas (GALNT6) and lung cancer and neuroblastoma (GALNT13), as both genes are found to be associated with tumor invasion and metastasis [44-47]. Similarly, GALNT14 knockdown significantly suppresses the cell migration and altered cellular morphology of EOC cells [48], and GALNT1 knockdown inhibits proliferation and tumor growth of bladder cancer cells [49], while GALNT10 expression positively correlates with histological type and degree of differentiation of gastric cancer [50]. In contrast, GALNT2 exerts anti-proliferative and anti-metastatic activity in gastric, brain and hepatocellular carcinomas [51-53], while GALNT9 is positively associated with a better clinical outcome in neuroblastoma patients [54], and GALNT12 expression represents a negative marker especially of metastatic gastric and colorectal cancer [55]. Also, GALNT14 expression correlates with pro-apoptotic drug’s sensitivity in pancreatic carcinoma, non–small-cell lung carcinoma and melanoma cell lines [56]. Interestingly, elevated GALNT7 expression contributes to growth and invasiveness of cervical cancer cells [57], while miR-30b/30d targeted GALNT7 suppression promotes the metastatic behavior of melanoma cells [58], which is indicative for the contrasting mode of action of this transferase in different tumor types. Similarly, GALNT6 expression in pancreatic cancer is associated with better overall survival [59]. The above data suggest that the different isoforms of the human GALNAC-Ts family could have distinct implication in tumorigenesis that could be specific for each cancer type.

The altered expression of different members of...
the GALNAC-Ts gene family in different malignancies makes them potential biomarkers [60]. Because aberrant glycoproteins as a result of these enzymes may be involved in promoting tumor invasion and metastasis, these enzymes could also be used as therapeutic targets. Indeed, GALNAC-Ts are responsible for the O-glycosylation of mucins, as this post-translational modification is critical for their function to protect and control the local environment of the cell surface [61], whereby aberrant glycosylation of mucins can contribute to cell differentiation, adhesion, invasion, and metastasis [62]. MUC1, as one highly glycosylated protein, is aberrantly overexpressed in the majority of epithelial cancers [63], including EOC [15]. MUC1 overexpression in EOC contributes to EOC cell–cell adhesion, proliferation, migration and metastasis and MUC1 has been identified as a promising target for EOC therapy [64]. It has been demonstrated that GALNT3 expression is associated with the differentiation or biological behavior of human adenocarcinomas through the initial glycosylation of mucin-type O-linked glycoproteins [65]. Since MUC1 glycosylation could also be regulated by GALNT3 [66], we decided to verify if GALNT3 suppression could affect MUC1 expression in EOC cells. We were able to show that knockdown of GALNT3 protein induced the reduction of MUC1 protein in EOC cells although the transcriptional level of MUC1 was unchanged (Figure 8B and C). Moreover, by using VVA lectin pull-down assay and consecutive VVA lectin blotting analysis, we found significantly diminished protein expression of glycosylated proteins in EOC cells upon GALNT3 suppression (Figure 8D), suggesting that GALNT3 may influence the post-translational modification and stabilization of MUC1 and other glycosylated proteins in EOC cells. Our data are also indicative for the possible implication of the GALNT3-MUC1 axis in EOC cell invasion via the regulation of expression of the cell adhesion proteins E-cadherin and β-catenin. Interestingly, GALNT6 has shown rather similar effects on MUC1 expression/glycosylation in breast cancer cells [44]; this is not quite unexpected, given that human GALNT3 and GALNT6 transferases display very high similarity in DNA and amino acid sequence throughout the coding region, as well as and similar kinetic properties that are distinct from all other human GALNAC-Ts [67].

In conclusion, we have shown that the GALNT3 transferase is significantly overexpressed in HG serous EOC tumors compared to LMP tumors and normal ovarian tissues, as the GALNT3 expression correlated with poor prognosis of EOC patients with advanced disease. Consecutive functional analyses of GALNT3 in EOC cells pointed towards its association with EOC cell proliferation (including cell cycle control), migration and invasion. Gene expression profiling and associated network and pathway analyses confirmed these findings, as numerous genes and pathways known previously to be implicated in ovarian tumorigenesis, including EOC tumor invasion and metastasis, were found to be suppressed upon GALNT3 knockdown, while some tumor suppressor genes were found to be induced. Moreover, GALNT3 downregulation was associated with reduced MUC1 protein expression in EOC cells, related probably with destabilization of the MUC1 protein due to lack of GALNT3 glycosylation activity, which results in consecutive accumulation of cell adhesion molecules such as E-cadherin and β-catenin. Hence, GALNT3 suppression could inhibit cell proliferation and invasion/migration by destabilizing MUC1 and possibly other mucin-type (O-linked) glycoproteins and thus inducing the expression of cell adhesion proteins, although further in-depth analysis will be required to elucidate the precise mechanism of the GALNT3-MUC1 pathway in EOC cells. Our data specify some of the putative mechanisms of abnormal glycosylation in ovarian carcinogenesis and the identification of the GALNT3 enzyme as novel EOC biomarker and/or possible molecular target for EOC therapy. Further studies are needed to more completely elucidate the functional implications of GALNT3 and possibly, of other members of the GALNAC-Ts gene family in ovarian tumorigenesis.

METHODS

Patients and tissue specimens

Snap frozen and formalin-fixed paraffin-embedded (FFPE) tissues of 117 EOC tumors were obtained at the Hotel-Dieu de Quebec Hospital, Quebec, Canada. These included 13 borderline, or LMP tumors and 104 HG adenocarcinomas. None of the patients received CT before surgery (see Table 1 for detailed clinicopathological characteristics). All tumors were histologically classified according to the criteria defined by the World Health Organization [68]. The CT treatment was completed for all patients and the response to treatment was known. Disease progression was evaluated following the guidelines of the Gynecology Cancer Intergroup [68]. Progression free survival (PFS) was defined as the time from surgery to the first observation of disease progression, recurrence or death. Thirteen normal ovarian samples were derived from women subjected to hysterectomy with oophorectomy due to non-ovarian pathologies. The study was approved by the Clinical Research Ethics Committee of the Hotel-Dieu de Quebec Hospital and all patients signed an informed consent for voluntary participation.

Cell cultures

The EOC cell lines OVCAR3 and SKOV3 were purchased from American Tissue Type Collection (Manassas, VA); OV-90, OV2008, TOV-112 and TOV-
21 cell lines were a kind gift from Dr. Anne-Marie Mes-Masson (Montreal University), while A2780s and A2780cp cell lines were a kind gift from Dr. Benjamin Tsang (Ottawa University). The cell lines were passed in different culture media supplemented with 10% fetal bovine serum, as described previously [69].

**Bisulfite sequencing PCR (BSP) analysis**

BSP analysis was performed, as previously described [11]. Briefly, genomic DNAs from normal ovarian tissues, borderline (BL, also known LMP), grade 1 (G1) and grade 3 (G3) EOC tumor specimens were isolated using the Qiagen DNeasy Blood and Tissue Kit. Bisulfite modification of genomic DNAs was done using the Methyl Detector kit (Active Motif, Carlsbad, CA). For BSP, a 315-bp fragment was amplified using primer pairs specific for bisulfite-modified sequences but not harboring CpGs, located at nt + 304 (GATTTTGAGGGGGG GT) to nt + 619 (ATCAAAAAAT AACCCA ACTCT) downstream of the GALNT3 transcription start (ATG) codon. BSP primer selection was performed using the Methyl Primer Express Software v1.0 (Applied Biosystems). PCR was done for 30 cycles (94°C, 45 s; 60°C, 45 s; 72°C, 45 s). PCR products were sent for dideoxy-sequencing analysis at the Genomics Analysis Platform at Laval University (http://www.bioinfo.ulaval.ca/seq/en/).

**Tissue microarrays (TMAs) construction and immunohistochemistry (IHC)**

TMAs were constructed, as previously described [69-71]. Briefly, one representative block of each ovarian tumor and normal ovarian tissue was selected for the preparation of the tissue arrays. Three 0.6 mm cores of tumor were taken from each tumor block and placed, 0.4 mm apart, on a recipient paraffin block using a commercial tissue arrayer (Beecher Instruments, Sun Prairie, WI). The cores were randomly placed on one of two recipient paraffin blocks, 0.4 mm apart, on a recipient paraffin block using a commercial tissue arrayer (Beecher Instruments, Sun Prairie, WI). The cores were heated in an autoclave for 12 min to retrieve antigenicity before blocking with endogenous peroxidase. Following treatment with 3% H2O2 for 10 min to quench the endogenous peroxidase activity, sections were incubated with the GALNT3 mouse monoclonal antibody (Abgent, Inc.) at dilution 1:50. Sections were then incubated with a biotinylated secondary antibody (Dako, Carpinteria, CA) and then exposed to a streptavidin complex (Dako, Carpinteria, CA). Complete reaction was revealed by 3-3’ diaminobenzidine and slides were counterstained with hematoxylin. GALNT3 protein expression was assessed by semi-quantitative scoring of the intensity of staining and recorded as absent (0), weak (1+), moderate (2+) or strong (3+). The relationship between GALNT3 expression in serous ovarian carcinomas and normal ovarian tissues was evaluated by the Wilcoxon two-sample test. A significant association was considered when p-value was below 0.05. A Kaplan-Meier curve and the log-rank test were performed based on PFS values to test the effect of the intensity of GALNT3 (3, 2 versus 0, 1) on disease progression.

**Short Hairpin RNA (shRNA) – mediated GALNT3 knockdown in A2780s cells**

A shRNA, targeting the GALNT3 sequence 5’-TACTGCTGAAGGAAAATCAT-3’, was designed using the siRNA Ambion Target Finder software (http://www.ambion.com/techlib/misc/siRNA_finder.html), and subcloned into the pSilencer 4.1 puro vector (Ambion). A2780s cells were stably transfected with the shRNA-GALNT3 plasmid using the ExGen 500 transfection reagent (Fermentas Canada Inc., Burlington ON), according to the manufacturer’s instructions. Cells were consecutively grown for 2 weeks in selection medium containing 5 μg/ml puromycin (Wisent, Canada) to isolate stable clones. Cells were also mock-transfected with the pSilencer 4.1 puro vector, and the stably-transfected clones were isolated as controls. Stable clones with inhibited GALNT3 expression were evaluated and validated by semi-quantitative RT-PCR and Western blot.

**Cell proliferation assays using impedance measurement with the xCELLigence system**

Cell proliferation (cell index) was checked by the xCELLigence Real-Time Cell Analyzer (RTCA) instrument, as previously described [69, 71]. Cells were seeded in triplicate at 2 × 10^4 cells/well in the E-Plate 16, a specialized 16-well plate used with the RTCA instrument. Each of the 16 wells on the E-Plate 16 contains an integral sensor electrode array so that cells inside each well can be monitored and assayed. Cell growth was monitored for 24 hours.

**Colony formation assay**

Colony formation assay was performed as previously described [69, 71]. A2780s cells were seeded at 500 cells per 60 mm culture dish. After 14 days, the dishes were washed twice in PBS, fixed with cold methanol, stained with Coomassie Blue (Sigma-Aldrich) for 5 min, washed with water and air dried. The number of colonies was determined by imaging with a Multimaging™ Cabinet.
(Alpha Innotech Corporation) and using AlphaEase Fc software.

Cell migration and invasion assays

Cell migration assays were performed in a modified Boyden-chamber assay using a Transwell chamber insert (6.5 mm diameter) separated by a polycarbonate filter of 5 μm pores (Costar, Cambridge, MA), as previously described [69, 71]. Briefly, shRNA-GALNT3 clones (sh-G1 and sh-G2) and the control (ctrl) were seeded into the upper chambers in 0.1% FBS containing medium at a density of 1.5 × 10⁴ per well, and 600 μl of 1% FBS containing medium was placed in the lower chamber as a chemoattractant. After 5 h at 37°C in 5% CO₂, the cells were fixed with cold methanol (15 min) and stained with trypan blue solution (5 mins). Cells on the upper surface of the filter were removed with cotton buds. Migrated cells on the underside of the filter were photographed and counted by phase contrast microscopy, by selecting 10 random fields per filter (at magnification× 40). The experiments were performed in triplicate. Cell invasion was assayed in a similar way, as the 8-μm pore polycarbonate filters were coated with 40 μl of Matrigel™ at concentration of 0.5 mg/ml (BD Biosciences, Franklin Lakes, NJ). Here, 600 μl of NIH3T3 conditioned medium was added in the lower chamber as a chemoattractant. Differences between shRNA-GALNT3 clones (sh-G1 and sh-G2) and the control (ctrl) were determined by a Student’s t-test, where p < 0.05 was considered significant.

Flow cytometry

Flow cytometry analysis was performed, as previously described [69]. Briefly, 7.5 × 10⁴ A2780s cells were treated with 20 mM hydroxyurea (Sigma) for synchronization at the G1/S boundary. After 16 h of incubation, cells were washed once with PBS, and resuspended in 1 ml of complete media. Then, cells were harvested by trypsinization at 0, 3, 6, and 9 h, washed three times with PBS, and fixed with ice-cold 95% ethanol overnight. Cells were washed with PBS (3x) and incubated with propidium iodide (50 μg/ml) (Sigma) in the dark at room temperature for 30 min. Flow cytometric analysis was performed on a Beckman Coulter EPICS XL-MCL analyzer. The cell cycle phase distribution was calculated from the resultant DNA using the cell QuesPro software.

MTT (cytotoxicity) assay

The MTT cell proliferation assay (Sigma) was used to measure the cell growth inhibition effects of cisplatin and paclitaxel in A2780s cell clones suppressing GALNT3, as previously described [69]. Briefly, cell suspensions (at 2 × 10⁴ cells/ml) were transferred to 96-well plates in triplicates and incubated for 3 days with different cisplatin and paclitaxel concentrations (ranging between 1 nM and 100 μM). Then, 20 μl of 3-[4, 5-dimethylthiazol-2-yl]-2, 5-diphenyl-tetrazolium bromide (MTT, 5 mg/ml) was added to each well 4 h before the end of the incubation. After centrifugation and removing the supernatant, 200 μl of dimethyl sulfoxide (DMSO) were added to resolve the crystals and the optical density was measured by microplate reader at 595 nm.

Gene expression profiling and data analysis

Gene expression analysis was carried out as previously described [69-71]. Briefly, total RNA was extracted from the shRNA-GALNT3 knockdown clones sh-G1 and sh-G2, and the corresponding control (mock-transfected) A2780s clone (ctrl). The quality of the RNA samples was examined by capillary electrophoresis using the Agilent 2100 Bioanalyzer (Agilent). Fluorescently labeled cRNA targets were generated from 0.5 μg of total RNA from each corresponding A2780s cell clone, using the Fluorescent Linear Amplification Kit (Agilent) and 10 mM Cyamine 3- or 5-labeled CTP (PerkinElmer), and following user’s manual. Cyanine labeled cRNA from the clone suppressing GALNT3 (sh-G1 and sh-G2) was mixed with the same amount of reverse-color cyanine-labeled cRNA from the corresponding control (ctrl) clone and hybridized on the Agilent Whole Human Genome microarrays, containing 44,000 genes. Array hybridization, washing, scanning, data extraction and analyses were performed as previously described [69]. Network analysis of the microarray data was completed using the Ingenuity Pathway Analysis (IPA) software (see http://www.Ingenuity.com). The microarray data have been deposited to the GEO database (http://www.ncbi.nlm.nih.gov/geo/) with accession number GSE52602.

Semi-quantitative RT-PCR (sqRT-PCR)

Analysis of GALNT3 gene expression in stably transfected GALNT3 knockdown clone (sh-G1) and the corresponding mock-transfected A2780s clone (ctrl) was performed by sqRT-PCR as previously described [69-71]. The GUSB gene was used as an internal standard. Comparative signal intensity was evaluated using the ImageJ software (http://rsb.info.nih.gov/ij/). Primers were designed for these loci with the sequences freely available from the Entrez Nucleotide database and the Primer3 algorithm for primer design (http://www-genome.wi.mit.edu/cgi-bin/primer/primer3_www.cgi) (see Supplemental Table 1 for sqRT-PCR primer description).
Quantitative PCR (qPCR)

Quantitative PCR was performed as previously described [71]. Briefly, total RNA was extracted by RNeasy Plus Mini Kit (QIAGEN) and cDNA was obtained by qScript™ cDNA SuperMix (Quanta BioSciences, Inc.). Primers were designed for these loci with the sequences freely available from the Entrez Nucleotide database and the Primer3 algorithm for primer design (http://www-genome.wi.mit.edu/cgi-bin/primer/primer3_ www.cgi). The primers used for qPCR validation are listed in Supplemental Table 1. PerfeCTa® SYBR® Green FastMix® (Quanta BioSciences, Inc.) was used according to manufacturer’s instructions. PCR reactions were performed on Rotor-Gene RG-3000 Real Time PCR System (Qiagen), with 18S ribosomal RNA used as endogenous control. PCR volume was 20 µl (36-well plate), and conditions were as follow: initial cycle 50°C, 2 min, 95°C, 15 min; 45 cycles at 95°C, 20 s, 60°C, 20 s and 72°C, 20 s; final cycle 72°C, 30 s. Data were analyzed by the Rotor-Gene software using the comparative ΔΔCt method. The relative copy number was calculated based on the target gene/18S RNA ratio.

Western blotting

Western blot analysis was performed as previously described [69-71]. Briefly, protein lysates were prepared by resuspending cell pellets in Laemmlli sample buffer containing 5% β-mercaptoethanol. Protein lysates were separated by 6 to 12% Tris-glycine gel electrophoresis and transferred onto a polyvinylidene difluoride membrane using a semidry apparatus (Bio-Rad). The membrane was blocked with 5% nonfat dry milk, washed with PBS, and membranes were incubated for an additional 2 h at 4°C with rotation. Lectin/glycoprotein complexes were collected by brief centrifugation (1400 rpm, 5 min), and washed three times with lysis buffer, followed by one wash with phosphate-buffered saline (PBS). Glycoproteins were released from the complexes by boiling in 30-50 µl SDS–PAGE sample buffers (5 min). The glycoproteins were resolved by SDS–PAGE, then immunoblotted to detect MUC1 or GalNAc-conjugated proteins.

VVA lectin pull-down assay for O-glycosylated (GalNAc-conjugated) proteins

Lectin extracted from Vicia villosa (VVA lectin) displays high affinity for GalNAc [72] and has been previously used for pull-down assays to detect GalNAc-conjugated proteins [73]. Similarly, we have performed VVA lectin pull-down assay as follows: 600 µg of cell lysate protein was incubated for 3 h at 4°C with 4 µg of biotinylated lectin VVA (EY Laboratories). Twenty µl of streptavidin-agarose (Sigma) was then added, and samples were incubated for an additional 2 h at 4°C with rotation. Lectin/glycoprotein complexes were collected by brief centrifugation (1400 rpm, 5 min), and washed three times with lysis buffer, followed by one wash with phosphate-buffered saline (PBS). Glycoproteins were released from the complexes by boiling in 30-50 µl SDS–PAGE sample buffers (5 min). The glycoproteins were resolved by SDS–PAGE, then immunoblotted to detect MUC1 or GalNAc-conjugated proteins.

ACKNOWLEDGEMENTS

This study was sustained by a grant to D.B. from the Cancer Research Society of Canada. Clinical specimens were provided by the Banque de tissus et de données of the Réseau de recherche sur le cancer of the Fonds de recherche du Québec - Santé which is affiliated with the Canadian Tumor Repository Network.

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