Advances in microbial remediation for heavy metal treatment: a mini review

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Abstract
In recent years, microbiological treatment to remediate contamination by heavy metals has aroused public attention as such pollution has seriously threatens ecosystems and human health and impedes sustainable development. However, the aspect of actual industrial wastewater and solid waste remediation by microorganisms is not explored sufficiently. And what we focus on is technical field of microbial remediation. Therefore, in this review, we discuss and summarize heavy metal treatment via microbiological approaches in different media, including wastewater, solid waste from industrial factories and polluted sites. We also clarify the technical applicability from the perspective of biosorption, bioleaching, biominerization, etc. In particular, the exploration of the combination of microbiological approaches with chemical methods or phytoextraction are scrutinized in this review relative to real waste heavy metal remediation. Furthermore, we highlight the importance of hyperaccumulator endophytes.

Keywords: Heavy metals, Biological treatment application, Hyperaccumulator endophytes

1 Introduction
Heavy metals are usually released by different anthropogenic sources such as nonferrous metallurgical industry, mining, mineral processing, electroplating, leather tanning, chemical industry, etc. [1–3]. The contamination by heavy metals has received global attention as water contamination has caused serious negative effects on human health through bioaccumulation in the food chain. Large volumes of wastewater originated from the metallurgical processes, and the effluent produced by the nonferrous metallurgical industry are around 730 million tons per year in China [4, 5]. Therefore, wastewater from nonferrous metallurgical industry is the major source of heavy metals. Approximately 10 million tons of electroplating sludge are produced annually in China [6], which is the secondary solid waste arising out of the precipitation of rinse water and spent electrolytes/etchants in the electroplating treatment industries [7, 8]. Tannery sludge contains diverse undesirable substances such as organics, inorganics, metallic elements [9, 10]. It is currently a huge challenge to remediate tannery sludge consists of high concentration chromium [11].

Recently, a series of technologies, such as chemical precipitation, electrochemical treatment, reverse osmosis, ion-exchange, physical adsorption and membrane filtration have been developed to remove heavy metal ions from liquid waste, which focus on reducing metal bioavailability by transforming the heavy metal to the a less toxic state [3, 12]. Several treatment schemes have been proposed for solid waste, such as landfilling [13], chemical leaching [14], stabilization/solidification [15], extraction [16], etc. However, most of technologies appear to be expensive, inefficient, or the treatment process lacks selectivity, and are especially ineffective at low metal concentrations. Bioremediation is a promising technology that uses living green plants or microorganisms to remove pollutants from soils, surface water and groundwater. Microorganisms can also detoxify metals by bioreduction, biosorption, bioleaching, biomineralization, etc. Biosorption/bioaccumulation is the most widely
biotechnique to remove metal ions form effluent, among which pollutants could be adsorbed onto the walls of microbial cell by (1) precipitation, (2) chelation, (3) complexation, or (4) ionic interactions [17, 18]. Recently, bioleaching is developed to recycle different metals in the treated leachate or residues via leaching them from solid wastes [19, 20]. In addition, biomineralization usually refers to a process of precipitation of mineral materials, by which microbes transform aqueous metal ions into amorphous or crystalline precipitates [21]. Hitherto, biological technologies based on microorganisms for heavy metal waste treatment has received considerable and growing attention over the years [22, 23], because of outstanding advantages, including high efficiency, low cost, and environment friendly. The metabolic diversity and activity of microorganisms has allowed their commercial exploitation in the field of waste cleanup (Fig. 1). Previous studies focused on the isolation and application of microbes with heavy metal resistance in synthetic heavy metal solutions. Nevertheless, nowadays researchers prefer pilot or large-scale waste treatment in actual (i.e., real) case, or using pollutants from industrial plants and contaminated sites. As a result, we have summarized the heavy metal treatment by microbial technology in both wastewater and discharged solid pollutants from industrial plants and polluted sites. Likewise, we need to focus on the biotechniques applicability from the perspective of technical field of microbial remediation. Subsequently, the exploitation of hyperaccumulator endophytes as an efficient bioremediation strategy is discussed.

2 Biological treatment of heavy metal in different types of waste

2.1 Electroplating effluent and sludge

The electroplating industry produces a variety of metallic coatings, ranging from technological to decorative applications, discharges large amounts of wastewater contaminated with heavy metals, which is of environmental concern due to their high toxicity [24, 25]. However, most of traditional techniques for electroplating effluent treatment have drawbacks such as low removal efficiency and production of large amounts of sludge, which has been identified as hazardous waste because of the high-concentration of potentially toxic metals [26, 27]. Therefore, a combination of biological and conventional approach is a proper strategy to decontaminate metal pollutant for the remediation of electroplating wastewater and sludge.

Several studies have been conducted on the treatment of metals in electroplating effluent using different biological methods, as summarized in Table 1. Ye et al. [29] reported metal removal from aqueous solutions (including Cr, Cu, Ni and Zn) by the mixture of Candida lipolytica and dewatered sewage sludge. The biosorption of Cr increased the live C. lipolytica extracellular secreta

![Fig. 1](image_url)
| Mixture or microorganism | Source | Capacity/Efficiency | Mechanism | Reference |
|--------------------------|--------|---------------------|-----------|-----------|
| Leptospirillum ferrophilum CS13, Acidithiobacillus caldus S2, Sulfobacillus acidiphilus S5 | Electroplating sludge | Removal rate of various heavy metals was over 99% | Bioleaching | [28] |
| Water treatment sludge | Electroplating wastewater | Cu^{2+} and Cr^{6+} adsorption capacities are about 1.7 and 3.5 mg g^{-1} | Biosorption | [21] |
| Mixture of Candida lipolytica and sewage sludge | Electroplating wastewater | 6.66 mg g^{-1} | Bioreduction and biosorption | [25] |
| Sulfate-reducing bacteria (43.3% of Desulfotignum) enriched sludge | Electroplating effluent | 100% within 3 d | Biological precipitation | [26] |
| Acidithiobacillus ferrooxidans | Dewatered metal plating sludge | Metal solubilization: 97% of Zn, 96% of Cu, 93% of Ni, 84% of Pb, 67% of Cd and 34% of Cr in 20 days | Bioleaching | [27] |
| Acidithiobacillus ferrooxidans | Electroplating sludge | Cr: 80.9% and Ni: 65.8% (with acid leaching) + 6.0% Cr and 11.7% Ni (additional extraction by bioleaching) | Combined biodegradation and acid leaching | [29] |
| Anthrobacter sp. X34 | Simulated groundwater | 0 | Biomineralization | [20] |
| Bacillus sp. Y9–2 | | 73% precipitated in 5 days | Biomineralization | [31] |
| Rahnella sp. Y9602 | | 95% precipitated in 5 days | Biomineralization | [32] |
| Chlorella vulgaris and calcined eggshells | Acid mine drainage | 99.7% of Fe, 99.5% of Cu, 99.9% of Zn, 99.8% of Mn, 100% of As, and 100% of Cd in 6 days | Biosorption | [30] |
| Sulfate-reducing bacteria | Underground mine drainage | 90.5% of Cd, 89.3% of Zn | Biomineralization | [25] |
| Acidithiobacillus ferrooxidans | Synthetic acid mine drainage | 53.62% of Fe | Biomineralization | [26] |
| Bacillus subtilis ITSUKMW1, Acinetobacter junii VITSUKMW2, and Escherichia coli VITSUKMW3 | Synthetic Cr^{6+} solution | 99% reduction of Cr^{6+} (100 mg L^{-1}) in 64 h | Bioreduction | [33] |
| Bacillus cerus Pseudomonas putida | Tannery effluent | 51.9% of Cr (VI) | Bioreduction | [35] |
| Cladosporium perangustum, Penicillium commune, Paecilomyces lilacinus, Fusarium equiseti | Tannery wastewater | 100% of Cr (VI), 99.92% of total Cr, 95.91% total Pb, 100% of Pb (II) | Biosorption | [36] |
| Acidithiobacillus ferrooxidans LX5, Acidithiobacillus thiooxidans T56 | Bioleachate derived from tannery sludge | Chromium-iron agent was bioleached from bioleachate | Bioleaching and recycling | [37] |
| Kitasatospora sp | Tannery wastewater | 100% of Cr (VI) 98% of Cr (III) precipitation | Biosorption and chemical precipitation | [38] |
| Stenotrophomonas sp. WW501, Proteus mirabilis, Microbacterium paraoxydans, Bacterium FX2, Bacillus tequilensis | Tannery sludge | 90% of Cr (VI) within 65 h (single culture of WW501) | Bioreduction | [39] |
| Mixture of Leptochloa fusca and Pantoaea stewartii AS11, Microbacterium arborescens HU33, Enterobacter sp. U38 | Tannery effluent | 135 ± 4.2, 19 ± 1.3, 8550 ± 42, 450 ± 9.5, 174 ± 3.1, 15 ± 0.72, 11 ± 0.21, 5 ± 0.29, 7 ± 0.65 (mg kg^{-1} Root) | Bioaccumulation and phytoextraction | [40] |
| Pantoaea sp. strain TYRI15, Microbacterium arborescens TY5104, Bacillus endophyticus PIRI25, Bacillus pumilus PIRI30, Bacillus sp | Textile effluent | 97% of Cr, 89% of Fe, 88% of Ni, 72% of Cd | Bioaccumulation and phytoextraction | [41] |
slime layers and decreased the intracellular synthesis of metal binding proteins or peptides. However, only aluminum-based water treatment sludge was assessed for removal of high concentrations of Cu and Cr (VI) from an electroplating wastewater along with other heavy metals [24]. Two continuous-flow pilot-scale systems were used to demonstrate that sulfate-reducing bacteria-based metal process could not only remove sulfate and nickel with low cost and less sludge, but also facilitate the subsequent removal of total phosphorus and nitrogen [28]. The main genera in the sulfate-reducing bacteria community were Desulfovibrio (relative abundance of 43.3%). In summary, the microbes enriched from activated sludge is a better choice for treatment of actual plating wastewater containing various wastes.

Electroplating sludge is a potential secondary resource. Earlier, to compare the microbial and chemical leaching technique to recover heavy metals in dewatered electroplating sludge, Bayat and Sari [42] studied the bioleaching of sludge involving Acidithiobacillus ferrooxidans in a completely mixed batch reactor. Their results illustrated that Zn, Cu, Ni and Pb had good solubilization efficiency (84–97%) during the bioleaching process, but Cd and Cr solubilization was relatively low (67% and 34%, respectively). Recently, combined microbial and acid leaching was followed for sludge treatment because acid leaching could reduce the toxicity of heavy metals in electroplating sludge [43]. The solubilized Cr content (80.9%) was improved compared to the former research (Table 1). The bio-treatment of electroplating sludge was also investigated for bioleaching of metals by mixed microbes (Leptospirillum ferrophilum CS13, Acidithiobacillus caldus S2, Sulfobacillus acidophilus S5) at semi-pilot scale [44]. A 300 L aerated packed reactor was designed to effectively leach heavy metals from electroplating sludge within a few hours using the mixed microbial stock solution.

### 2.2 Pollution of mining waste

The booming economy in China has led to scale expansion of mining in the past decades. Nonferrous mining can impact water and soil quality according to the chemical composition of the ores and waste dump materials [30]. Mining wastewater is a kind of mine impacted water composed of mine drainage, open cast mining and waste rock yard drainage, most of which contains high concentrations of sulfate and heavy metals. Mine impacted waters can be acidic (acid mine drainage, AMD), or with pH values ranging from 6 to 9 defined as neutral mine drainage (NMD) [31]. Moreover, mine impacted waters could be treated by abiotic and biotic means which can be classified as active and passive treatments [32].

Many research reports are available on the heavy metal pollution from mine areas worldwide, as summarized in Table 1. AMD is a major environmental problem that is greatly contaminating water bodies in and around abandoned mine area. A pilot scale AMD treatment plant was built at the Yongdong mine located in South Korea. The effluent contained excess Fe, Cu, Zn, Mn, As, and Cd. A hybrid system containing calcined eggshells and microalgae was used to remove heavy metals from AMD in a 40-L bioreactor [45]. The results showed that the simultaneous removal of Fe, Cu, Zn, Mn, As, and Cd from the AMD effluent was 99 to 100% in 6 days. A semi-passive experiment mimicking the belowground situ conditions was carried out at Silver King mine site in Canada, where leaching of Zn and Cd occurs. Metals were removed from underground mine drainage (it was NMD) fed into the 200 L bioreactors, Zn and Cd removal extent was 20.9% and 39% in winter, respectively. However, the Zn and Cd removal efficiency increased to 89.3% and 90.5% in summer, respectively [34]. Given that the influent average concentration of Zn and Cd was 640.4 μg/L and 10.5 μg/L respectively, this biological approach could only be applied in the low metal concentration for groundwater treatment. In order to enhance lime neutralization with AMD by reducing the production of ferric hydroxide and waste gypsum, a microbial enhanced-lime neutralization passive treatment technology was developed for AMD on a semi-pilot scale [33]. This system promoted As (III) in solution adsorption by minerals and transformation of soluble Fe and SO$_4^{2−}$ into secondary iron minerals by biomineralization with A. ferrooxidans.

There are more than 114 Sb mines in China, accounting for approximately 90% of global Sb production [46]. Thus, remediation of Sb-rich mine water is generally regarded as a priority issue for local government. Sun et al. [47], designed an onsite field-scale bioreactor system (consisted of five treatment tanks) to passively treat Sb mine contaminated water in Southwest China. The polluted water came from an active upstream Sb mine that produces 8–25 m$^3$/d of mine water with up to 7 mg/L soluble Sb (III). With the aerobic tanks inoculated with indigenous mine water microorganisms, the bioreactor removed more than 90% of total soluble Sb and 80% of soluble Sb (III). Three indigenous bacteria were isolated from chromite mining sites in India. The development of sequential processes (isolated → acclimatized → consortia) improving Cr (VI) resistant isolates, proved to be a feasible microbial reduction approach for the specific chromite mine [48].

### 2.3 Leather tanning effluent and sludge

Both leather production and consumption are developing fast in China. In order to convert raw skin/hides into leather, approximately 80% of chromium-containing tanning agents have been utilized in the leather-based
industries [49, 50]. The enforceable maximum contaminant level of total chromium in drinking water and public water systems is 50–100 μg/L [51], whereas chromium in industrial discharge ranges from 0.1 to 400 mg/L. [52] Consequently, treatment of tanning effluent for contamination alleviation becomes urgent and necessary. Chromium is a transition metal with wide industrial application, existing in oxidation states of trivalent to hexavalent. The trivalent and hexavalent forms are the most stable among the major oxidation states of chromium [53]. Compared with Cr (III), Cr (VI) is considered to be more toxic due to its high solubility and mobility. In addition, Cr (VI) is both genotoxic and carcinogenic, identified as one of the chemicals causing a threat to human health [35]. Several studies have shown that Cr (III) compounds at high concentrations cause oxidatively-generated DNA damage [36]. Hence, any form of chromate removal/recycling is an alternative option to the conventional reduction of chromium, especially the treatment of whole real tannery waste, as summarized in Table 1.

For every 20% of leather produced from raw material, more than 72% of the chromium from tanning agents is converted into solid and liquid waste [38]. Thus, microbial reduction of Cr (VI) from tannery solid and liquid waste has been studied by many researchers. Nutrients required for microbial metabolism, such as carbon/energy and nitrogen sources are scanty in real tannery effluent. Supplementation with such nutrients was proved to increase the efficiency of bioremediation [37]. Tannery effluent decolorization, dechlorination, and Cr⁶⁺ remediation were obtained with 0.8% glucose and 0.2% ammonium chloride (w/w) in 3:1 diluted wastewater within 3 d of Bacillus cereus incubation. Simultaneously, effluent bioremediation was attempted with an immobilized coculture of B. cereus and Pseudomonas putida, which enhanced the remediation of Cr⁶⁺ (51.9%) compared to the single culture removal (41.7%). Apart from bacteria, filamentous fungi are an attractive option for tanning waste treatment. High-strength tannery wastewater treatment by nylon mesh immobilized fungal inocula in a stirred tank bioreactor demonstrated good reduction in different contamination parameters within 120 h, e.g. COD (82.52%), color (86.19%), Cr (VI) (100%), Total Cr (99.92%), Pb (II)(100%), Total Pb (95.91%) and NO₃⁻ (9.94%) [39].

Some researches focused on developing effective resource utilization of leather waste. For example, a combined chemical–biological system was designed for Cr remediation and reclamion. Lime and cement dust were used for chemical precipitation of Cr (III) that was recycled in the leather tanning, while the actinomycete Kitasatospora sp. was used in a vertical glass column filled with porous media for Cr (VI) adsorption [54]. Ma et al. [55] reported chromium recovery of bioleachate derived from tannery sludge by microbial leaching and its reuse, and a chromium-iron tanning agent was developed to reduce the costs of Cr and Fe separation. Chromium-iron tanned leather showed equivalent to around one year’s natural aging of heat aging properties after hot air aging tests. Resource utilization of leather processing wastes is probably a potential way in the future. Due to microbial consumption of organic matter, the amount of the tannery sludge was decreased by 27% with culturing mixed bacteria (five Cr resistant strains) anaerobically for 14 d [56]. Chromate (VI) reductase was found to be localized inside the extracellular membrane or adsorbed to its surface. This is probably a mechanism for the removal of Cr (VI) via Stenotrophomonas sp.

3 Exploitation of hyperaccumulating endophytic microbes as an efficient heavy metal bioremediation strategy

3.1 Endophytes isolation and characterization

Heavy metal hyperaccumulators, particularly those inhabiting in contaminated areas, are plant species capable of absorbing much larger amounts of metal compounds than general plants. Endophytic microbes inhabit internal tissues of hyperaccumulators and form a range of different relationships with the host plant, including symbiotic, mutualistic, and trophobiotic without causing disease. In such plant-bacteria combination, plants support the microbial community, and in return, microbes improve plant growth and pollutant detoxification. Hence, these hyperaccumulator endophytes also exhibit higher heavy metal tolerance and accumulating abilities compared to other microorganisms [57–59]. Endophytes from heavily polluted sites with radionuclides and other toxic heavy metals may be potentially rich bioresources in heavy metal decontamination. We can exploit the endophytic microorganisms as an effective bioremediation strategy. Earlier, a bacterium strain LRE07 (Serratia sp.) was isolated from the root of the cadmium hyperaccumulator Solanum nigrum L., which could bind cadmium and zinc efficiently in its growing microbial cells both in single-ion and multi-ions system [58]. However, S. nigrum L. inoculated with an endophyte Pseudomonas sp. Lk9 isolated from its host plant, increased phytoextraction rates of all metals from multi-metal contaminated soils (17.4%, 48.6% and 104.6% for Cd, Zn and Cu, respectively) [59]. A total of 14 bacterial endophytes were isolated from Alnus firma root and assayed for tolerance to lead, among which isolate MN3–4 was identified as Bacillus sp. [60]. Moreover, Bacillus MN3–4 could produce siderophores and indoleacetic acid to transform the Pb form in soils or increase Pb accumulation by plants.
3.2 Endophytes with plant growth promoting properties
Plant growth promoting endophytes (PGPE), identified as precious bioresources in bioaugmentation with phytoremediation, promote plant growth and heavy metal uptake via various mechanisms. Many isolates from hyperaccumulators have been proved for their plant growth promoting (PGP) features as well as their resistance to different heavy metals [61–63]. Four Cd-resistant endophytic strains assisted their host plants to cope with toxicity stress responses. They produced indole-3-acetic acid (IAA), 1-aminocyclopropane-1-carboxylic deaminase, phosphate solubilizing activity and siderophores, which were identified as plant growth promoting mechanisms [40]. Multiple heavy metal-resistant (Cd, Zn, Pb and Cu) PGPE from root nodules of *Robinia pseudoacacia* in a mining area were selected to improve phytoremediation efficiency. *Mesorhizobium loti* HZ76 and *Agrobacterium radiobacter* HZ6 were symbiotic with the highest potential for heavy metal resistance and PGP properties [41]. Besides bacteria, the stress-mitigating effects of endophytic fungi have been investigated. *Penicillium funiculosum* LHL06, isolated from soybean roots, could help the host plants to modulate physio-biochemical, molecular, and proteomic responses to multiple heavy metals (Ni, Cu, Pb, Cr, and Al) toxicity [64]. Fungus LHL06 can upregulate gibberellins, IAA production and downregulate heavy metal ATPase genes in its host plants compared to non-fungi-inoculated plants.

3.3 Application of endophytic microbes in real contaminants
A plant-bacterial system with constructed wetlands (CWs) was developed for the efficient remediation of tannery effluent, among which three different endophytic bacteria were used for bioaugmentation [65]. The results showed that the combined use of plants and endophytes lead to enhanced performance of heavy metal (Cr, Fe, Mn, Ni, Pb, Ba, Cd, and Co) removal from wastewater compared to the plants only, as summarized in Table 1. The combined utilization of the salt-tolerant plant *Leptochloa fusc a* and the chromium-resistant endophytes *Prosopis juliflora* (bacteria from another plant) for tannery effluent treatment was the highlight of this study. Another pilot-scale vertical flow constructed wetlands setup was likewise implemented to treat dye-rich real textile effluent for one year. The endophyte-assisted CWs promoted a substantial removal of heavy metals (Cr 97%, Fe 89%, Ni 88%, Pb 72%), simultaneously, decreased chemical oxygen demand (81%), biochemical oxygen demand (72%), color (74%), nitrogen (84%) and phosphorus (79%) [66]. The above-mentioned study could likely be applied in future field-scale and time-effective bioremediation of real industrial effluent consisted of organic and inorganic contaminants. Plants inoculated with different consortia were used to carry out an endophyte-assisted phytoremediation experiment in a metal contaminated mine soil [67, 68]. Inoculation of endophytes improved the plants’ physiological status by increasing the chlorophylls and carotenoids content. Moreover, positive influence of plant growth and endophyte inoculation on soil characteristics were reflected in PGP features, such as higher values of acid phosphatase activity, microbial community diversity, etc. The phytoremediation of vanadium-contaminated soil was elevated via affecting the rhizosphere and endosphere microecosystem by endophyte inoculant.

4 Biological technique patterns applicable principles
As shown in Table 2, we analyzed and organized the techniques applicability, heavy metal form, and concentration range involved in this study. Biosorption can be applied using live or dead organism, which include bacteria, fungi, algae, actinomycete. Microorganism combine with different bioresource as biosorbers adsorb and accumulate heavy metals (soluble form) from wastewater or acid mine drainage under low to moderate concentration. Microbial leaching is suitable for extracting metals (insoluble form) from secondary resources. Bioleaching not only decrease the amount of sludge to be disposed of but also prevent release of metals into the environment. *Acidithiobacillus ferrooxidans* and *Acidithiobacillus thiooxidans* are widely employed in the bioreduction process [42, 43, 55]. But various microbes like *Pseudomonas fluorescens* [69], *Leptospirillum ferriphilum* [70], and *Aspergillus niger* [71] have been tested for metals bioleaching. Biominalization, ubiquitous in nature, is a process of mineral crystal formation induced/controlled by microbes. Sulfate-reducing bacteria, *A. ferrooxidans* and urease-positive fungi play a potential role in the synthesis of novel biominerals (schwertmannite, calcite, otavite) that can remove toxic metal As or Cd [33, 34, 72]. Microorganisms can also detoxify metals by bioreduction (e.g., Cr (VI) to Cr (III), SeO₄²⁻ to Se), the valence of metal ions is transformed during the process. In cleaning up heavy metal-contaminated water or soils (soluble form), phytoextraction is typically employed together with bioaccumulation by endophytes. The approach is suitable for large, low contamination areas in situ.

5 Conclusions
We have compared the advantages and disadvantages of biological approaches in Table 2. Biological or other technologies for purifying or recycling real wastes containing heavy metals have their own strength and weakness. Thus, the future trend is to develop treatment of
| Remediation pattern | Applicability | Metal valence and initial concentration range (mg/L) | Reactor | Advantages | Disadvantages | Reference |
|---------------------|---------------|-----------------------------------------------------|---------|------------|--------------|-----------|
| Biosorption         | Electroplating or tannery wastewater, acid mine drainage, ex-situ, low to moderate concentration | Total Cr: 11.4–47.76  
Cr²⁺: 8.27–39.65  
Cu²⁺: 0.86–22.78  
Co³⁺: 0.032  
Total Pb: 1.14  
Pb²⁺: 0.26–0.96  
Hg²⁺: 0.65  
Zn²⁺: 0.89–28.1  
Ni²⁺: 0.18–30.92  
Fe³⁺: 140.83–225.80  
Mn²⁺: 9.23–11.95  
As³⁺: 0.16–0.67  
Cd²⁺: 0.15–0.36 | Continuous flow column, a stirred tank, the photobioreactor | The rapid adsorption of most metal ions within 30 min, high removal rate | Concentration range below 50 mg/L except Fe³⁺ | [24, 29, 39, 45] |
| Combination of biosorption and chemical precipitation | Tannery effluent, ex-situ, moderate to high concentration | Total Cr: 2250  
Cr²⁺: 625 (biosorption)  
Cr³⁺: 1625–2410 (chemical precipitation)  
Cr (VI): 212–279,400  
Cr (III): 30  
Zn: 29.1–700  
Cu: 500–691  
Ni: 600–8850  
Pb: 600–850  
Cd: 20–341  
Fe: 2338  
Al: 43.1–800 | Column filled with sand (sand can absorb 70% Cr²⁺) | High metal concentration, lime could enhance the pH to meet the standard discharge requirements | Produce large amount of sludge and Cr³⁺ could be oxidized easily. | [54] |
| Bioleaching         | Sludge, bioleachate derived from sludge, ex-situ, moderate to high concentration | Insoluble metal form, concentration range (mg/kg)  
Total Cr: 212–279,400  
Cr (VI): 30  
Cr (III): 2126  
Zn: 29.1–700  
Cu: 500–691  
Ni: 600–8850  
Pb: 600–850  
Cd: 20–341  
Fe: 2338  
Al: 43.1–800 | A stirred aeration packed reactor, mixed batch reactor | A cleaner process and higher removal efficiency than chemical leaching | A longer period of operation (slow kinetics) compared to chemical leaching | [42, 44, 55] |
| Bioleaching incorporating acid leaching | Electroplating sludge, ex-situ, moderate to high concentration | Concentration range (mg/kg)  
Total Cr: 155000  
Ni: 8500 | Bioelectrical reactor | Shorter period than bioleaching only: 9 h (acid leaching) + 60 h (bioleaching) | Additional power is supplied to maintain the bioelectrical reactor | [43] |
| Biomineralization   | Groundwater or acid mine drainage, sulfate rich metal-laden wastewater | U⁶⁺(UO₂)²⁻: 47.6  
Total Ni: 441  
Total Cr: 5.66  
Soluble Sb: 7  
Cd²⁺: 0.001  
Zn²⁺: 0.64–7.24  
As³⁺: 1 | The plug-flow ditch system, two continuous flow pilot-scale bioreactor | Produce less toxic neutralized residues, less sludge, low cost, low efficiency and concentration | [23, 28, 33, 34, 47] |
| Bioreduction        | Effluent or sludge containing Cr⁶⁺ | Cr⁶⁺: 5–300 | Bench-scale bioreactor | Simple and effective strategy, no secondary pollution | Cr³⁺ needs to be treated | [37, 48, 56] |
| Combination of phytoextraction and bioaccumulation | Effluent or contaminated soil, in-situ, suitable for large, low contamination areas | Total Cr: 9.7–247  
Cr⁶⁺: 0.78  
Cr³⁺: 246  
Cu²⁺: 1.12 | Pilot-scale vertical flow constructed wetlands | High public acceptance, low cost, easy to implement | Large area, limited to shallow contamination, time-consuming | [65–68] |
Table 2: Mechanisms, advantages, disadvantages, reactor, and applicability of the biological techniques (Continued)

| Remediation pattern | Applicability | Metal valence and initial concentration range (mg/L) | Reactor | Advantages | Disadvantages | Reference |
|---------------------|---------------|-----------------------------------------------------|---------|------------|---------------|-----------|
|                     |               | Fe$^{3+}$: 14.3–17.38 |         |            |               |           |
|                     |               | Mn$^{2+}$: 4.33 |         |            |               |           |
|                     |               | Ni$^{2+}$: 1.22–7.6 |         |            |               |           |
|                     |               | Pb$^{2+}$: 0.24 |         |            |               |           |
|                     |               | Ba$^{2+}$: 1.15 |         |            |               |           |
|                     |               | Cd$^{2+}$: 0.88–2.66 mg/L or 3.84 mg/kg soil |         |            |               |           |
|                     |               | Co$^{2+}$: 1.06 |         |            |               |           |
|                     |               | V$^{4+}$: 66–31.9 mg/kg soil |         |            |               |           |

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heavy metals by the combination of multiple approaches to exploit the optimum experimental protocol which benefit from the vantage and avoid the shortcomings. In addition, a very limited number of field applications have been performed for heavy metal bioremediation. Consequently, research for future technology enhancement should be aimed at evaluating the feasibility of industrial waste treatment at large scale and application of on-site treatment systems. Then try to resolve problems of field engineering application.

Abbreviations
C. lipolytica: Candida lipolytica; AMD: Acid mine drainage; NMD: Neutral mine drainage; A. ferrooxidans: Acidithiobacillus ferrooxidans; B. cereus: Bacillus cereus; PGPE: Plant growth promoting endophytes; PGP: Plant growth promoting; IAA: Indole-3-acetic acid; CW: Constructed wetlands

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Luo S conceived and designed this manuscript and made substantial contributions to the revision of this work. Wang S interpreted data from references and wrote the original manuscript. Liu T wrote and revised the contributions to the revision of this work. Wang S, Luo S, Li J, Wang D interpreted data from references and wrote the original manuscript. Liu T wrote and revised the contributions to the revision of this work. Wang S, Luo S, Li J, Wang D, contributed to the revision of this work. Authors read and approved the final edition.

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Availability of data and materials
The data of Table 1 in this study is available in the references, which has been mentioned in the article.

Competing interests
No conflict of interest exits with the submission of this manuscript, and the been mentioned in the article.

References
1. Nouha K, Kumar RS, Tyagi RD. Heavy metals removal from wastewater using extracellular polymeric substances produced by Cloacabacterium normanense in wastewater sludge supplemented with crude glycerol and study of extracellular polymeric substances extraction by different methods. Bioresour Technol. 2016;212:209–9.
2. Liu SH, Zeng GM, Niu QY, Liu Y, Zhou L, Jiang LH, Tan P, Zhang C, Cheng M. Bioremediation mechanisms of combined pollution of PAHs and heavy metals by bacteria and fungi: a mini review. Bioresour Technol. 2017;224:25–33.
3. Ayangbenro AS, Babalola OO. A new strategy for heavy metal polluted environments: a review of microbial biosorbents. Int J Environ Res Public Health. 2017;14(1):94.
4. Cai R, Gao C, Zhang J, Huang X. Resource and utilization of metallurgical wastewater. J Salt Chem Ind. 2013;42:1–7.
5. Wu P, Jiang LY, He Z, Song Y. Treatment of metallurgical industry wastewater for organic contaminant removal in China: status, challenges, and perspectives. Environ Sci Wat Res Technol. 2017;36(6):1015–31.
6. Lu H, Yu S. Spatio-temporal variational characteristics analysis of heavy metal pollution in water of the typical northern rivers, China. J Hydrol. 2018;559:787–93.
7. Li P, Peng C, Li F, Song S, Juan A. Copper and nickel recovery from electroplating sludge by the process of acid-leaching and electro-depositing. Int J Environ Res. 2011;5(3):797–804.
8. Peng G, Tian G. Using electrode electrolytes to enhance electrokinetic removal of heavy metals from electroplating sludge. Chem Eng J. 2010;165(2):388–94.
9. Tang P, Zhao Y, Xia F. Thermal behaviors and heavy metal vaporization of phosphatized tannery sludge in incineration process. J Environ Sci. 2008;20(9):1146–52.
10. Goswami L, Mukhopadhyay R, Bhattacharya SS, Das P, Goswami R. Detoxification of chromium-rich tannery industry sludge by Eudrillus eugeniae: insight on compost quality fortification and microbial enrichment. Bioreasour Technol. 2018;266472–81.
11. Yang YL, Ma HR, Chen XP, Zhu C, Li X. Effect of incineration temperature on chromium speciation in real chromium-rich tannery sludge under air atmosphere. Environ Res. 2020. https://doi.org/10.1016/j.envres.2019.109159.
12. Azeem A, Azari A, Rezaalizami M, Ansarpour M. Removal of heavy metals from industrial wastewaters: a review. ChemBioEng Rev. 2017;4(1):37–59.
13. Pant D, Joshi D, Upreti MK, Kotnala RK. Chemical and biological extraction of metals present in E waste: a hybrid technology. Waste Manag. 2012;32(5):979–90.
14. Podstawczyk D, Witek-Krowiak A, Dawiec A, Bhattacharya SS, Chakraborty A. Biosorption of copper (II) ions by fox meal: empirical modeling and process optimization by response surface methodology (RSM) and artificial neural network (ANN) simulation. Ecol Eng. 2015;82:264–79.
15. Lu D, Xu X, Hu H, Han X. Characterization of pb2+ biosorption by psychrotrophic strain Pseudomonas sp. I3 isolated from permafrost soil of Mohe wetland in Northeast China. J Environ Manag. 2017;196:15–8.
16. Pant D, Joshi D, Upreti MK, Kotnala RK. Chemical and biological extraction of metals present in E waste: a hybrid technology. Waste Manag. 2012;32(5):979–90.
17. Scanzazzato T, Panossian Z, Tenorio J, Perez-Herranz V, Espinosa D. A review of cleaner production in electroplating industries using electrodialysis. J Clean Prod. 2017;168:1590–602.
18. Zhou C, Wang Q, Lv M, Chen L. Microorganism remediation strategies towards heavy metals. Chem Eng J. 2019;360:1553–63.
19. Aaslestad MJ, Martinez RJ, Sobecky PA, Webb SM, Talieft M. Uranium immobilization in heavy metal sludges using engineered mineral earth as a result of bacterial phosphatase activity: insights from bacterial isolates from a contaminated subsurface. Environ Sci Technol. 2007;41(16):5701–7.
20. Ghorapade A, Ahammed MM. Water treatment sludge for removal of heavy metals from electroplating wastewater. Environ Eng Res. 2017;23(2):92–8.
21. Orescanin V, Durgo K, Mikelic IL, Halkijevic I, Kuspilic M. Toxicity assessment of untreated/treated electroplating sludge using human and plant bioassay. J Environ Sci Health A. 2018;53:1295–30.
22. Dhami NK, Quirin ME, Mukherjee A. Carbonate biominalization and heavy metal remediation by calcium-fusion isolated from fungal koji. Caves Ecol. 2017;103:106–17.
23. Yin K, Wang Q, Lv M, Chen L. Microorganism remediation strategies towards heavy metals. Chem Eng J. 2019;360:1553–63.
24. Beazley MJ, Martinez RJ, Sobecky PA, Webb SM, Talieft M. Uranium immobilization in heavy metal sludges using engineered mineral earth as a result of bacterial phosphatase activity: insights from bacterial isolates from a contaminated subsurface. Environ Sci Technol. 2007;41(16):5701–7.
25. Ghorapade A, Ahammed MM. Water treatment sludge for removal of heavy metals from electroplating wastewater. Environ Eng Res. 2017;23(2):92–8.
26. Orescanin V, Durgo K, Mikelic IL, Halkijevic I, Kuspilic M. Toxicity assessment of untreated/treated electroplating sludge using human and plant bioassay. J Environ Sci Health A. 2018;53:1295–30.
27. Scanzazzato T, Panossian Z, Tenorio J, Perez-Herranz V, Espinosa D. A review of cleaner production in electroplating industries using electrodialysis. J Clean Prod. 2017;168:1590–602.
28. Zhou C, Gu S, Yu H, Zhang T, Cheng H, Sun Q, Xiao R. Environmental risk assessment of pyrometallurgical residues derived from electroplating and pickling sludges. J Clean Prod. 2018;177:699–707.
29. Hu K, Xu X, Chen Y. An assessment of sulfate reducing bacteria on treating wastewater from electroplating plant. J Hazard Mater. 2020. https://doi.org/10.1016/j.jhazmat.2020.122376.
30. Ye J, Yin H, Mai B, Peng H, Qin H, He B, Zhang N. Biosorption of chromium from aqueous solution and electroplating wastewater using mixture of Candida lipolytica and dewatered seawater sludge. Bioreasour Technol. 2010;101(11):3893–902.
31. Dhal B, Thatoi HN, Das NN, Pandey BD. Environmental quality of the Boula-Nuasahi chormite mine area in India. Mine Water Environ. 2011;30:191–6.
32. Nordstrom DK, Blowes DW, Paceck CJ, Hydrogeochemistry and microbiology of mine drainage: an update. Appl Geochem. 2015;57:3–16.
32. Johnson DB, Hallberg KB. Acid mine drainage remediation options: a review. Sci Total Environ. 2005;338(1):3-14.
33. Song Y, Wang H, Yang J, Zhou L. Evaluation and optimization of a new microbial enhancement plug-flow ditch system for the pretreatment of acid mine drainage: semi-pilot test. RSC Adv. 2018;8(31):10399-46.
34. Nielsen G, Hatam I, Abuan KA, Janin A, Coudert L, Biais JF, Mercier G, Baldwin SA. Semi-passive in-situ pilot scale biosorator successfully removed sulfate and metals from mine impacted water under subarcatic climatic conditions. Water Res. 2018;140:268-79.
35. Elahi A, Acog I, Bhalari DA, Rehman A. Successive use of microorganisms to remove chromium from wastewater. Appl Microbiol Biotechnol. 2020;104(3):795-802.
36. EFSA Panel on Contaminants in the Food Chain (CONTAM). Scientific opinion on the risks to public health related to the presence of chromium in food and drinking water. EFSA J. 2014;12(3):3959.
37. Tripathi M, Garg SK. Dechlorination of chloroorganics, decolorization, and simultaneous bioremediation of Cr(6+) from real tannery effluent employing indigenous Bacillus cereus isolate. Environ Sci Pollut Res. 2014;21(7):5227–41.
38. Hülfer S, Taeger T. Sustainable leather manufacturing—a topic with growing importance. J Am Leather Chem Assoc. 2004;99(10):423–8.
39. Sharma S, Malavaria P. Isolation, characterisation, and application of heavy metal-resistant and plant growth-promoting endophytic bacteria from root nodules of Robinia pseudoacacia in a Pb/Zn mining area. Microbiol Res. 2018;217:51–9.
40. Bayat B, Sari C. Comparative evaluation of microbial and chemical leaching processes for heavy metal removal from dewatered metal plating sludge. J Hazard Mater. 2010;174(1–3):763–9.
41. Wu P, Zhang LI, Lin C, Xie Y, Yong X, Wu X, Zhou J, Ji H, Wei P. Extracting heavy metals from electroplating sludge by acid and bioelectrochemical leaching using Acidithiobacillus ferrooxidans. Hydrometallurgy. 2020;210:105225.
42. Zhou W, Zhang L, Peng Ji, Ge Y, Tian Z, Sun J, Cheng H, Zhou H. Cleaner recycling of chromium from tannery wastewater using combined chemical–biological treatment. Process Saf Environ Prot. 2016;104:1–10.
43. Ma H, Zhou J, Hua L, Cheng F, Zhou L, Qiao X. Chromium recovery from tannery sludge by biodegrading and its reuse in tanning process. J Clean Prod. 2017;142:2752–60.
44. Liu H, Wang Y, Zhang H, Huang G, Yang Q, Wang Y. Synchronous detoxification and reduction treatment of tannery sludge using Cr(VI) resistant bacterial strains. Sci Total Environ. 2019;687:34–40.
45. Xiao L, Luo S, Zeng G, Wei W, Lan Y, Chen L, Guo H, Cao Z, Yang L, Chen J, Xi Q. Biosorption of cadmium by endophytic fungus (EF) Microsporum sp. LSE10 isolated from cadmium hyperaccumulator Solanum nigrum L. Bioresour Technol. 2010;101:1668–74.
46. Luo S, Wan Y, Xiao X, Guo H, Chen L, Xi Q, Zeng G, Liu C, Chen J. Isolation and characterization of endophytic bacterium LR07 from cadmium hyperaccumulator Solanum nigrum L. and its potential for remediation. Appl Microbiol Biotechnol. 2011;89:1637–44.
47. Chen L, Luo S, Li X, Wan Y, Chen J, Liu C. Interaction of Cd-hyperaccumulator Solanum L. and functional endophyte Pseudomonas sp. Lk9 on soil heavy metals uptake. Soil Biochem. 2014;68:300–8.
48. Shin MN, Shim J, You Y, Myung H, Bang SK, Cho M, Kamala-Kannan S, Oh BT. Characterization of lead resistant endophytic Bacillus sp. MN3-4 and its potential for promoting lead accumulation in metal hyperaccumulator Alnus firma. J Hazard Mater. 2012;198:114–20.
49. Liao S, Chen L, Chen JL, Xiao X, Xu T, Wang Y, Rao C, Liu C, Liu Y, Lai C, Zeng GM. Analysis and characterization of cultivable heavy metal-resistant bacterial isolates from Cd-hyperaccumulator Solanum nigrum L. and their potential use for phytoremediation. Chemosphere. 2011;85:1130–8.
50. Plociniczak T, Chodor M, Pacwa-Plociniczak M, Piotrowska-Segert Z. Metal-tolerant endophytic bacterial associated with Silene vulgaris support the Cd and Zn phytoextraction in non-host plants. Chemosphere. 2019;219:520–60.
51. Wang Q, Ma L, Zhou Q, Chen B, Zhang X, Wu Y, Pan F, Huang L, Yang X, Feng Y. Inoculation of plant growth promoting bacteria from hyperaccumulator facilitated non-host root development and providing promising agents for elevated phytoremediation efficiency. Chemosphere. 2019;223:769–76.
52. Bilal S, Shahzad R, Khan AL, Harrazi A, Kim CK, Lee UJ. Phytotribiotics enabled endophytic Penicillium funiculosum LH06 protects Glycine max L. from synergistic toxicity of heavy metals by hormonal and stress-responsive proteins modulation. J Hazard Mater. 2019;379:120824.
53. Ashraf S, Afzal M, Naveed M, Shahid M, Ahmad ZZ. Endophytic bacteria enhance remediation of tannery effluent in constructed wetlands vegetated with Leptochloa fusca. Int J Phytoremediat. 2018;20(2):121–8.
54. Hussain Z, Arslan M, Malik MH, Mohsin M, Iqbal S, Afzal M. Treatment of the textile industry effluent in a pilot-scale vertical flow constructed wetland system augmented with bacterial endophytes. Sci Total Environ. 2018;645:966–73.
55. Burges A, Epedel L, Blanco F, Becerril JM, Garbiu G. Ecosystem services and plant physiological status during endophyte-assisted phytoremediation of metal contaminated soil. Sci Total Environ. 2017;584:329–38.
56. Wang L, Lin H, Dong Y, Li B, He Y. Effects of endophytes inoculation on rhizosphere and endosphere microecology of Indian mustard (Brassica juncea) grown in vanadium contaminated soil and its enhancement on phytoremediation. Chemosphere. 2020;240:124891.
57. Potyris A, Lens PNL, van de Vossenberg J, Rene ER, Grybos M, Guibaud G, Kierzak J, van Hullebusch ED. Comparison of Cu, Zn and Fe bioaccumulation from Cu-metallurgical slags in the presence of Pseudomonas fluorescens and Acidithiobacillus thiooxidans. Appl Geochem. 2016;68:39e62.
58. Funari V, Makinen J, Salminen J, Braga R, Dinelli E, Revizor H. Metal removal from municipal solid waste incineration fly ash: a comparison between chemical leaching and bioleaching. Waste Manag. 2017;60:397–406.
59. Shah SS, Palmeri MG, Regina S, Sponchiado P, Bevilacqua D. Environmentally sustainable and cost-effective bioleaching of aluminum from low-grade bauxite ore using marine-derived Aspergillus niger. Hydrometallurgy. 2020;205:105368.
60. Li Q, Laslo JC, Geoffrey MG. Bionmineralization of metal carbonyles by Neurospora crassa. Environ Sci Technol. 2014;48:14099–106.

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