**Cutaneous amyloid is a biomarker in early ATTRv neuropathy and progresses across disease stages**

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**Abstract**

**Objective:** To determine the sensitivity and specificity of cutaneous amyloid deposition in relation to patient-reported measures in the earliest disease stage of hereditary ATTR amyloidosis (ATTRv). **Methods:** In a cross-sectional study, we analyzed 88 individuals with TTR mutations, 47 of whom were in the earliest disease stage and without clinically evident neuropathy, 12 healthy controls, and 13 disease controls with diabetes. All participants’ neuropathy symptoms and signs were assessed using validated patient and clinician-reported measures and 3-mm skin punch biopsies were immunostained using protein gene product 9.5 and Congo Red. **Results:** Amyloid can be detected in the earliest disease stages in up to 86% of patients with ATTRv amyloidosis. Amyloid was not detected in healthy individuals or individuals with diabetic peripheral neuropathy supporting a sensitivity of 86% and a specificity of 100%. The cutaneous deposition of amyloid correlates with neuropathy sensory symptoms, measured with the Neuropathy Total Symptom Score-6 ($R = 0.46, p < 0.01$); pain measured with the Brief Pain Symptom Inventory ($R = 0.44, p < 0.05$); autonomic symptoms, measured with the Boston Autonomic Symptom Questionnaire ($R = 0.38, p < 0.05$); and quality of life measured with the Norfolk Diabetic Neuropathy Quality of Life Questionnaire ($R = 0.44, p < 0.05$). Individuals with amyloid deposition were more likely to have sensory symptoms, pain, autonomic impairment, and reduced quality of life than ATTRv patients without amyloid deposition. **Interpretation:** These findings have implications for understanding the earliest manifestations of the clinical phenotype of ATTRv-associated neuropathy, for the pathophysiological construct of disease staging, and for timing the introduction of disease-modifying therapy.

**Introduction**

Hereditary ATTR amyloidosis (ATTRv) is a progressive, debilitating, multisystem fatal disease caused by the extracellular deposition of misfolded, mutant transthyretin in tissues throughout the body.¹ ² The clinical manifestations of ATTRv are heterogeneous and vary by mutation, geographical region, and other genetic and environmental factors. This heterogeneity, coupled with low penetrance of the trait, anticipation, and the late onset of the clinical manifestations of the disease in some patients, has led to poor recognition of the disease, particularly in nonendemic areas, and highlights the need for sensitive, specific, and readily available diagnostic tests.

In the presence of a family history of ATTRv or a suggestive clinical phenotype, a pathogenic mutation can be detected by TTR gene sequencing. Pathological confirmation of the disease is obtained by histological confirmation of amyloid deposition. Skin biopsy assessment of intraepidermal nerve fiber density has become the de facto gold standard for the diagnosis of small fiber neuropathy, yet few reports of skin biopsy assessments of cutaneous amyloid exist. Initial studies failed to support the use of skin biopsy to document amyloid deposition.³ ⁴
however, more recent studies have observed amyloid deposition in ATTRv patients although rarely in the absence of the clinically evident disease.

We studied skin biopsy detection of cutaneous amyloid deposition in a large series of ATTRv patients from different geographic regions with different TTR mutations. The patient perspective is of increasing importance in healthcare delivery and clinical trial design. Thus, with patient-centricity in mind, we focus our results on patient input using validated patient-reported outcome (PRO) and quality of life (QOL) questionnaires. We show that amyloid deposition occurs early in the clinical course, even before the appearance of clinically evident disease. In doing so, we determine the optimum number of sections and biopsy sites to maximize diagnostic sensitivity. We also report the relationship between amyloid deposition on skin biopsy and patient- and clinician-reported measures with a focus on the earliest manifestations of ATTRv neuropathy. Finally, we determine the relationship between the clinical neuropathy features and their structural correlates in the skin and introduce an index of amyloid deposition that could function as a therapeutic biomarker and measure of target engagement. These findings provide the basis for a diagnostic and therapeutic approach to individuals with ATTRv with neuropathy across disease stages and mutations.

Materials and Methods

One hundred and thirteen participants (88 individuals with TTR pathogenic variants, 12 healthy controls, and 13 disease controls with diabetes) from the Instituto Nacional de Ciencias Medicas y Nutricion Salvador Zubiran, Mexico City, Mexico; the Raúl Carrea Institute for Neurological Research, FLENI, Ciudad Autónoma de Buenos Aires, Argentina; and the Beth Israel Deaconess Medical Center, Boston, USA. Participants were screened for other potential causes of neuropathy, including diabetes, neurotoxins, thyroid dysfunction, and vitamin B12 deficiency. Subjects were excluded if laboratory abnormalities confirmed another potential cause of neuropathy. Control subjects were healthy individuals of similar age with no known medical illness. Disease controls had diabetic neuropathy with a range of neuropathy severity. The study protocol was approved by local ethical and research committees and all participants gave their written informed consent.

Patient-reported scales

All participants completed symptom scores including the Neuropathy Total Symptom Score (NTSS-6), the Brief Pain Symptom Inventory Short Form (BPI-SF), the Boston Autonomic Symptom Questionnaire, and the Norfolk Diabetic Neuropathy Quality of Life Questionnaire (Norfolk-DN QoL).

The NTSS-6 evaluates the frequency and intensity of individual neuropathy painful and nonpainful sensory symptoms, for example, numbness and/or insensitivity, prickling, and/or tingling sensation.

The BPI-SF assesses the severity of pain and the impact of pain on daily functioning.

QoL was measured with the Norfolk-DN QoL, a 47-item questionnaire that groups items according to small fiber, large fiber, and autonomic nerve function, symptoms, and activities of daily living (ADL).

Clinician-reported scales

All participants had complete history and physical examinations to confirm study eligibility. Neuropathy-specific examinations were quantified by the Neuropathy Impairment Score in the lower limbs (NIS-LL) and the Utah Early Neuropathy Score (UENS). The NIS-LL is a neuropathy score weighted toward motor testing with a range of 0 (no neuropathy) to 88 (severe sensory and motor neuropathy in the lower legs). The Utah Early Neuropathy Score (UENS) is a small fiber sensory examination with a score of 0 (no neuropathy) to 24 (severe bilateral sensory neuropathies). Functional status was assessed by Coutinho staging.

Laboratory studies

Laboratory studies included blood glucose levels, thyroid function tests, vitamin B12 levels, and serum protein electrophoresis with immunofixation to exclude other causes of peripheral neuropathy.

Genetic analysis of TTR mutations

Genomic DNA was extracted from the peripheral venous blood using direct sequencing or saliva testing, using commercial genetic laboratories (Ambry Genetics).

Skin biopsies

All patients underwent 3-mm punch skin biopsies at the lateral distal leg and distal thigh using standard techniques. All specimens were analyzed at Beth Israel Deaconess Medical Center. Specimens were fixed and stained with protein gene product 9.5 (1:1000, rabbit anti PGP 9.5, Chemicon International Inc.). Skin biopsies were fixed and stained in Zamboni fixative, using protein gene product 9.5 (PGP9.5) as previously reported and Congo Red (using a modified Highman’s Congo Red.
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protocol)\(^5,19\) using 50-μm-thick tissue sections. Briefly, sections were stained in Congo Red solution for 10 min, rinsed, differentiated in alkaline alcohol, counterstained with Gill’s hematoxylin, dehydrated, dipped in xylene, and mounted. A total of four tissue sections per biopsy were used to quantify the intraepidermal nerve fiber density (IENFD) using immunofluorescent microscopy.\(^17\) A total of six tissue sections per biopsy were stained with Congo Red. Biopsies were viewed at high power (40x) through all tissue sections under polarized light to detect amyloid. Biopsy sections were viewed under polarized light and were imaged in entirety using a 6.2 megapixel Pixelink camera at 20x resolution to encompass the entire biopsy. The number of amyloid deposits was measured in each image, with a maximum of two amyloid deposits per field of view (to prevent overcounting of large amorphous sheets of amyloid—thus each field of view had 0–2 amyloid deposits). The total number of deposits was then divided by the total length of tissue studied in millimeters multiplied by 100 (for ease of data reporting), with the results expressed as the Amyloid Deposition Index (ADI—the number of amyloid deposits per linear millimeter of tissue measured). The number of tissue sections reviewed in order to detect amyloid was recorded for each subject. The pathology reviewer (C.H.G.) was blinded to the clinical data.

Intraepidermal nerve fiber density (IENFD)

IENFD counting was performed by a physician blinded to diagnosis, and the results were expressed as a linear density (number of fibers per millimeter) as previously described.\(^17\)

Data availability

Anonymized data not published within this article will be made available by request from any qualified investigator.

Statistical analysis

Statistical data analysis was performed using SPSS v17.0 (IBM, Chicago, IL). Results are reported in aggregate, but also by mutation type, Coutinho stage, NIS-LL stage, and amyloid deposition. The amyloid deposition index groups were defined as <15, 15–150, and >150 based on the normal distribution of amyloid index data into three groups. Data are reported as mean, median, range, or standard deviation as noted. Between-group comparisons were made by one-way analysis of variance (ANOVA) with Tamhane T2 post hoc test. Correlations between tests of nerve fiber structure, nerve fiber function, amyloid deposition, and symptom scores were measured using Pearson correlation coefficient (R) if data were normally distributed, with corrections for multiple comparisons.

Results

Demographic information

One hundred and thirteen participants enrolled in the study (88 ATTRv, 12 controls, and 13 individuals with type 2 diabetes). The distribution of TTR mutations is listed in Table 1 and includes 43 participants with Val30-Met mutations and 30 with Ser50Arg mutations. Other mutations, reported as a group, include 2 F64L mutations, 6 G47A mutations, 5 S52P mutations, 1 Y136H, mutation, and 1 V142I mutation. Most participants were in the early disease stage (Coutinho stage 0: n = 47, Coutinho stage 1: n = 32, Coutinho stage 2: n = 8, and Coutinho stage 3: n = 1) The controls were healthy and individuals with diabetes who were otherwise healthy with normal renal function, well-controlled blood pressure, and lipids. Complete demographic information on all participants is provided in Table 1.

Amyloid detection by skin biopsy

Amyloid was detected in skin biopsies of 76 of 88 (86%) individuals with ATTRv, and none of the healthy control participants or individuals with diabetes when using two skin biopsies from different sites with six tissue sections per biopsy (86% sensitivity, 100% specificity). Skin biopsies positive for amyloid deposition are shown in Figure 1A–D. In Figure 1E and 1F the number of skin biopsies obtained and the number of tissue sections required for the detection of amyloid is illustrated. At the distal leg biopsy site, there is a 37% chance that amyloid is detected within a single tissue section, which increases to 74% with six tissue sections (Fig. 1E). By adding a second biopsy with an additional six tissue sections, the positive detection rate of amyloid increases to 86%. Additional results are provided in Table 1.

In patients with no clinical signs of neuropathy (i.e., NIS-LL score of 0), a single tissue section at the distal leg had a 28% amyloid detection rate (Fig. 1F). With six tissue sections from a single biopsy the amyloid detection rate was 52%, with the addition of a second skin biopsy, also with six tissue sections measured, the detection rate increased to 77% (Fig. 1F).

Similar results were obtained for Coutinho grade 0 (i.e., patients graded by their physicians as asymptomatic), a single tissue section at the distal leg had a 48% chance of detecting amyloid (Fig. 1E). Using six tissue sections from a single biopsy, amyloid was detected
Table 1. Demographic information and participant characteristics.

| TTR Mutation | V30M | V30M | V30M | V30M | SS0R | SS0R | SS0R | Other | Other | Other | Healthy control | Disease control |
|--------------|------|------|------|------|------|------|------|-------|-------|-------|----------------|-----------------|
| Coutinho     |      |      |      |      | All  | Stage 0 | Stage I | Stage II | Stage III | All | Stage 0 | Stage I | Total | Stage 0 | Stage I | N/A | N/A |
| Stage Number | 43 | 16 | 18 | 30 | 19 | 11 | 15 | 12 | 3 | 12 | 13 |
| Age Mean     | 40.8 ± 3.14 | 31.9 ± 9.1 | 44.2 ± 12 | 47.7 ± 11.8 | 70 | 38.6 ± 13 | 32.5 ± 9.3 | 50.4 ± 10.8 | 38.8 ± 11.2 | 36.1 ± 8.6 | 49.6 ± 13.5 | 50.1 ± 13.3 | 63.4 ± 9.3 |
| Range SD     | (19–73) | (19–46) | (25–73) | (33–72) | (21–61) | (21–59) | (25–61) | (26–66) | (33–66) | (30–72) | (52–79) |
| Sex Number  | 25 F | 10 F | 12 F | 19 M | 11 M | 7 M | 5 M | 6 M | 6 M | 6 M | 6 F | 4 F |
| Number of amyloid-positive biopsies (%) | 37 (86%) | 11 (68.8%) | 17 (94.4%) | 8 (100%) | 1 (100%) | 25 (83.3%) | 16 (84.2%) | 9 (81.8%) | 14 (93.3%) | 11 (91.6%) | 3 (100%) | 0 |
| Number of NIS-LL mean ± SD (range) | 18.9 ± 23.3 | 16.1 ± 13.3 | 56.9 ± 6.9 | 66.5 | 3.9 ± 5.3 | 2.3 ± 4.1 | 7.1 ± 5.8 | 3.4 ± 4.4 | 2.5 ± 3.7 | 7.5 ± 5.1 | 0 | 7.1 ± 5.1 |
| (0–73) | (0–48) | (51–73) | (0–18) | (0–18) | (0–14) | (0–7) | (0–7) | (0–7) | (0–7) | (2–14) | (2–14) |
| UENS mean ± SD (range) | 14.6 ± 15.8 | 16 ± 10.9 | 37.5 ± 4 | 40 | 5.4 ± 8.1 | 2.6 ± 4.9 | 10.2 ± 10.3 | 3.5 ± 6.0 | 1.5 ± 2.2 | 11.3 ± 10.4 | 0 |
| (0–42) | (0–40) | (30–42) | (0–32) | (0–32) | (0–32) | (0–32) | (0–32) | (0–32) | (0–32) | (3–23) | (3–23) |

NIS-LL, neuropathy impairment score in the lower limb; TTR: transthyretin; UENS: Utah early neuropathy scale.
68% of the time, with a second skin biopsy the detection rate increased to 88%.

Thirteen individuals had no symptoms on any of the questionnaires and had a normal physical examination and were Coutinho stage 0. Amyloid was detected in 8/13 of these individuals (62%).

Patient-reported measures—painful and nonpainful sensory symptoms, autonomic symptoms, and QOL

Symptoms worsened significantly with ATTRv amyloidosis disease clinical progression measured by the NIS-LL.
(NTSS-6 (R = 0.66, p < 0.01)), the BPI-SF (R = 0.61, p < 0.01), the Boston Autonomic Questionnaire (R = 0.58, p < 0.05), and QOL (as measured by the Norfolk-DN QoL (R = 0.64, p < 0.01)) worsened significantly with ATTRv amyloidosis disease clinical progression measured by the NIS-LL.

Similarly, when ATTRv participants were divided into those with no clinical evidence of neuropathy (NIS-LL of 0, N = 41), those with mild neuropathy (NIS-LL scores of 1–10, N = 22) and those with more severe neuropathy (NIS-LL scores >10, N = 25) significant worsening of symptom scores was observed across these divisions. No differences were evident across mutations although the study was not designed or powered to detect such differences.

**Painful and nonpainful sensory symptoms and amyloid deposition**

The NTSS-6 sensory total symptom score increased with a worsening NIS-LL score. All scores were worse in participants with amyloid deposition detected on skin biopsy even in patients without clinically evident neuropathy (NIS-LL = 0) (see Fig. 2 and Table 2). Of the NIS-LL = 0 patients, 48% of participants with amyloid deposition reported sensory symptoms on the NTSS-6. In contrast, only 11% of patients without amyloid deposition reported sensory symptoms on this questionnaire.

All individual symptoms (e.g., prickling and tingling, numbness) were more prominent and more severe and occurred more frequently with disease progression based on the NIS-LL score. All questionnaire sensory symptoms were present in those patients who had amyloid detected on skin biopsy, even in the NIS-LL = 0 group. All items were worse if amyloid deposition was present (see Fig. 2). Numbness was the most prevalent symptom overall, however, in the NIS-LL = 0 patients with amyloid deposition, prickling and tingling were most prevalent. Only deep pain was present in patients without amyloid on skin biopsy.

**Figure 2.** Neuropathy Total Symptom Score-6. The Neuropathy Total Symptom Score-6 (NTSS-6) evaluates the frequency and intensity of individual neuropathy painful and nonpainful sensory symptoms. Participants are grouped by NIS-LL = 0, NIS-LL 1–20, and NIS-LL >20. (A) Symptoms increase with the progression of the peripheral neuropathy as assessed by the Neuropathy Impairment Score of the Lower Limbs (NIS-LL). In all three groups, symptoms are worse in those participants with amyloid detected in the skin biopsy, even in those participants without clinically evident neuropathy (NIS-LL = 0). (B and C) In all participants with amyloid on skin biopsy (N = 76), all sensory symptoms are present and more severe in those participants with amyloid detected on skin biopsy. Numbness is the most prevalent symptom in participants with amyloid detected on skin biopsy. (D and E) All symptoms were present in the NIS-LL = 0 group with detected amyloid (N = 32). In this group, prickling and tingling were the most prevalent symptoms. In the NIS-LL = 0 without amyloid detection on skin biopsy, only occasional deep pain was present (N = 9).
Pain, pain interference, and amyloid deposition

The BPI-SF total score increased with the progression of the peripheral neuropathy as assessed by the Neuropathy Impairment Score of the Lower Limbs (NIS-LL). In all three groups, symptoms are worse in those patients with amyloid detected in the skin biopsy, even in those patients without a clinically evident neuropathy (NIS-LL = 0) (see Fig. 3A). Of the NIS-LL = 0 patients, 38% of participants with amyloid deposition reported pain on the BPI-SF. In contrast, only 11% of patients without amyloid deposition reported pain.

The individual items of the BPI-SF, the “worst,” “least,” “average,” and “pain right now scores” are shown in Figure 3B. All individual pain item scores increased with disease progression and were worse in those participants with amyloid deposition at every disease stage. The “do you have pain right now” item showed the greatest difference between those with and those without amyloid deposition. The percent of patients reporting interference and interference scores increased with disease progression and was significantly higher in patients with evidence of amyloid deposition (see Fig. 3C).

Autonomic symptoms

The orthostatic and vasomotor summary score of the Boston Autonomic Questionnaire increased with neuropathy progression and was significantly worse in those patients with amyloid deposition in all disease stages including those without clinically evident neuropathy (NIS-LL = 0) (see Fig. 3A). Of the NIS-LL = 0 patients, items in the small fiber domain (including, pricking/tingling or electric shocks) were present in 56% of individuals with amyloid deposition, but only 22% of those without amyloid deposition.

Clinician-reported scales—structured, quantified neurological examination

Across all mutation types, the severity of neuropathy, measured by a structured, quantified examination (UENS and NIS-LL) increased across the Coutinho stage (Table 1). The NIS-LL ranged from 0 to 73 and the UENS ranged from 0 to 42. The severity of neuropathy, as defined by NIS-LL and UENS scores, is reported in Table 1. A strong correlation was seen between the NIS-LL scores and the UENS scores. There was 98% concordance between NIS-LL score of 0 and UENS scores of 0.

Skin biopsy analysis

Intraepidermal and dermal nerve fiber density on skin biopsy

The data for IENFD grouped by NIS-LL score are reported in Table 2. IENFD declined significantly at the distal leg with an increase in NIS-LL scores (p < 0.0001, all comparisons). Similar results were observed when grouped by Coutinho staging and UENS scores. ATTRv participants had lower IENFD compared with control participants. Individuals with NIS-LL score = 0 (and those reported as Coutinho Stage 0) had significantly lower nerve fiber densities than control subjects across all mutation types (Table 1). Nearly identical results were seen when classifying by UENS scores of 0 (N = 40), UENS of 1–15 (N = 25), and NIS-LL scores of >15 (N = 23). None of the patients without amyloid deposition met the criteria for a small fiber neuropathy by skin biopsy, whereas, 44% of individuals with amyloid deposition on skin biopsy fulfilled the criteria for a small fiber neuropathy.

Amyloid deposition index

The deposition of amyloid increased with worsening neuropathy assessed by the NIS-LL (see Table 2). Similar results were seen when grouped by Coutinho stage and UENS: the deposition index at Coutinho stage 0 was 50 ± 108, Coutinho stage 1 was 184 ± 257, and at Coutinho stage 2–3 was 277 ± 244, and for the UENS, similar results were noted.
Study participants were divided into those with low levels of amyloid detected (ADI of <15, N = 30), those with moderate levels of amyloid (ADI of 15–150, N = 31) and those with high levels of amyloid detected (ADI >150, N = 27). Results are reported in Table 3. In all cases, the groups with greater levels of detected amyloid had more severe neuropathy, that is, worse sensory symptom scores, worse pain, worse autonomic symptoms, worse QOL, more abnormalities on the quantified neurological examination, more abnormalities to electrophysiological function, and lower nerve fiber densities.

### Relationships among symptoms, signs, and cutaneous neuropathology

The ADI correlated with the symptoms scores: the NTSS-6 (R = 0.46, p < 0.01); the BPI-SF (R = 0.44, p < 0.05), autonomic symptoms, measured with the Boston Autonomic Symptom Questionnaire (R = 0.38, p < 0.05); and QOL measured with the Norfolk Diabetic Neuropathy QOL Questionnaire (R = 0.44, p < 0.01). Similar correlations were observed with the examination scores (UENS [R = 0.53, p < 0.01] and the NIS-LL [R = 0.48, p < 0.01]) and the pathological findings (intraepidermal nerve fiber density at the distal leg [R = 0.52, p < 0.01]).

### Discussion

We report a clinical case series of ATTRv amyloidosis with skin biopsy testing for detection of cutaneous amyloid. In this study, which focuses on patients in the earliest disease stages, the major findings are as follows: First, amyloid is detected frequently using Congo Red staining of skin biopsies in patients with ATTRv amyloidosis; second, amyloid can be detected in the earliest stages of the disease, before symptom onset, and before clinically evident changes in the...
neurological examination; third, an association exists between the deposition of amyloid and the severity of neuropathy assessed using validated patient- and clinician-reported evaluation instruments; and fourth, symptoms, signs, and neuropathological changes to nerve fiber structure worsen with increasing amyloid deposition. These findings have implications for our understanding of the earliest manifestations of the clinical phenotype, for the pathophysiological construct of disease staging, and for timing the introduction of disease-modifying therapy.

Even in genetically confirmed cases, tissue confirmation of amyloid is required for definitive diagnosis of Cutaneous amyloid in ATTRv amyloidosis neuropathy

Figure 3. Brief Pain Symptom Inventory and Norfolk Quality of Life Questionnaire. The Brief Pain Symptom Inventory assesses the severity of pain and the impact of pain on daily functioning. Norfolk Diabetic Neuropathy Quality of Life Questionnaire (Norfolk-DN QoL) is a 47-item questionnaire that groups items according to small fiber, large fiber, and autonomic nerve function, symptoms, and activities of daily living. Participants are grouped by the Neuropathy Impairment Score of the Lower Limbs (NIS-LL) = 0, NIS-LL 1-20, and NIS-LL >20. (A) Pain total score increases with progression of the peripheral neuropathy as assessed by the NIS-LL. In all three groups, symptoms are worse in those participants with amyloid detected in the skin biopsy, even in those participants without clinically evident neuropathy (NIS-LL = 0). (B) All components of the pain total score are worse in those participants with amyloid detected on skin biopsy. (C) All interference items are worse in those participants with amyloid detected on skin biopsy. (D) The Norfolk QOL Total Score increased with disease progression based on the NIS-LL score. All scores were worse in participants with amyloid deposition detected on skin biopsy, even in the participants without clinically evident neuropathy (NIS-LL = 0). (E and F) The percentage of participants endorsing symptoms in QOL domains (physical functioning/large fiber, activities of daily life, symptoms, small fiber neuropathy domain, and autonomic neuropathy domain) was greater in participants with amyloid deposition, even in participants with NIS-LL = 0.
amylodosis, for prognostication and life planning, and prior to initiation of costly disease-modifying therapeutic interventions. A convenient, sensitive, and specific method of tissue confirmation would be a valuable tool in the diagnosis of patients with amyloidosis. At present, several different biopsy sites are considered, including some that require invasive procedures, such as kidney, heart, gastrointestinal tract, salivary gland, and peripheral nerve, but skin biopsy is still not typically included among these. The sensitivity of these investigations varies widely. Sural nerve biopsy, which is performed in some centers, has a sensitivity of up 83%, but may be complicated by pain, sensory loss, infection, and delayed wound healing. Salivary gland biopsies are often performed, particularly in Portugal. This technique has a reported diagnostic sensitivity ranging from 75% to 91%. Fat pad aspirate from abdominal tissue is widely used in the United States and may be the simplest approach to acquire tissue but has low sensitivity even for established disease. Several technical differences may underlie the increased sensitivity of skin biopsy over abdominal fat pad analysis in the detection of amyloid. First, the fat pad needle aspiration provides a smaller sample of tissue than a 3 mm skin punch biopsy; second, the tissue morphology is not maintained using a fat pad aspirate, and the physical integrity of amyloid deposits may be disrupted and difficult to identify; and third, the sensitivity is increased by the availability of additional sections and skin biopsy sites. Assessment of amyloid deposition in the abdominal fat pad obtained by skin biopsy may increase the sensitivity.

Figure 4. The Boston Autonomic Questionnaire. The Boston Autonomic Questionnaire assesses autonomic symptoms across multiple domains. Participants are grouped by the Neuropathy Impairment Score of the Lower Limbs (NIS-LL) = 0, NIS-LL 1–20, and NIS-LL >20. (A) The Boston Autonomic Questionnaire Total Score increased with disease progression based on the NIS-LL score. All scores were worse in participants with amyloid deposition detected on skin biopsy, even in the participants without clinically evident neuropathy (NIS-LL = 0). (B and C) Autonomic symptoms were more severe in participants with amyloid deposition, even in those without clinically evident neuropathy (NIS-LL = 0). (D and E) Individual items of the orthostatic and vasomotor domain were more severe in participants with amyloid deposition, even in those without clinically evident neuropathy (NIS-LL = 0).
Table 3. Participant characteristics subdivided by ADI.

| Examination scores | ADI 0–15 (N = 30) | ADI >15–150 (N = 31) | ADI >150 (N = 27) | ANOVA |
|---------------------|-------------------|-----------------------|-------------------|-------|
| NIS-LL              | 4.3 ± 11.28       | 5.98 ± 11.23          | 24.81 ± 23.7*     | <0.01 |
| UENS                | 3.5 ± 9.37        | 6.48 ± 8.98*          | 18.85 ± 14.83*    | <0.01 |
| Coutinho (median [range]) | 0 [0–1] | 1[0–2] | 1 [0–3] |

Sensory symptoms and pain

| NTS56               | 2.14 ± 3.76       | 4.11 ± 5.58*          | 8.74 ± 6.19*†     | <0.01 |
| BPI - Pain          | 1.48 ± 2.40       | 2.63 ± 2.53           | 4.4 ± 2.1*†       | <0.01 |
| BPI - Interference  | 0.99 ± 2.36       | 2.22 ± 2.91*          | 3.40 ± 3.19*†     | <0.01 |
| Norfolk Total       | 6.62 ± 11.05      | 17.84 ± 21.49*        | 37.81 ± 31.39*†   | <0.01 |
| Norfolk Small fiber | 0.54 ± 1.03       | 1.32 ± 1.89*          | 2.35 ± 1.98*      | <0.01 |
| Norfolk Pain        | 0.12 ± 0.33       | 0.71 ± 1.16*          | 1.62 ± 2.08*†     | <0.01 |
| Norfolk ADL         | 3.08 ± 5.86       | 8.61 ± 11.55*         | 21.73 ± 19.75*†   | <0.01 |
| Norfolk functioning | 0.92 ± 1.67       | 3.48 ± 5.08*          | 4.92 ± 5.8*       | <0.05 |
| Norfolk interference| 1.08 ± 2.19       | 1.74 ± 2.78           | 4.54 ± 3.81*†     | <0.01 |

Autonomic symptoms

| OHQ Score           | 7.23 ± 10.84      | 16.65 ± 22*           | 27 ± 28.55*       | <0.01 |
| BAQ OI              | 1.96 ± 4.20       | 9.40 ± 13.73*         | 11.45 ± 16.76*    | <0.01 |
| BAQ GI              | 9.0 ± 10.27       | 11.34 ± 15.42         | 19.33 ± 18.96*†   | <0.01 |
| BAQ GU              | 7.11 ± 6.78       | 8.13 ± 9.63           | 13.19 ± 12.11*    | <0.05 |
| BAQ sweat           | 3.22 ± 6.14       | 5.30 ± 6.98           | 6.7 ± 9.96*       | <0.05 |
| BAQ total           | 18.88 ± 17.67     | 33.84 ± 35.27*        | 40.77 ± 34.54*    | <0.01 |

Nerve fiber density

| IENFD DL            | 12.79 ± 6.78      | 12.21 ± 8.24          | 2.9 ± 5.78*†      | <0.001 |
| IENFD DT            | 19.69 ± 8.1       | 20.51 ± 10.52 ± 9.1   | 7.47 ± 7.59*†     | <0.001 |

Abbreviations: ADI, Amyloid Deposition Index; ADL, activities of daily living; ANOVA, analysis of variance; BAQ, Boston autonomic questionnaire; BPI, Brief Pain Symptom; DL, distal leg; DT, distal thigh; GI, gastrointestinal; GU, genitourinary; gastrointestinal; GU: genitourinary IENFD, intraepidermal nerve fiber density; NTS56, Neuropathy Impairment Score in the lower limbs; NTSS6, neuropathy total symptom score-6 item; OHQ, orthostatic hypotension questionnaire; OI, orthostatic intolerance; UENS, Utah Early Neuropathy Score.

* p < 0.05 vs ADI 0–15 group using Tamhane T2 post hoc test.
† p < 0.05 vs ADI 15–150 group using Tamhane T2 post hoc test.

Prior studies have reported that cutaneous amyloid can be detected in the skin of an individual with ATTRv mutations. Our approach increases the sensitivity and extends these findings to individuals without clinical evidence of peripheral neuropathy on examination. In the present study, the likelihood of amyloid detection increases from 37% with a single section to 74% with six sections from a single biopsy; and from 74% to 86% with an additional biopsy with six tissue sections. Most noteworthy is the detection of amyloid in 88% of asymptomatic patients (Coutinho Grade 0) and in 77% of patients without clinically evident neuropathy on examination. The majority of patients included in prior diagnostic discrimination studies had clinically evident disease. The increased sensitivity of amyloid detection in this study may be due to several reasons. First, a greater number of tissue sections studied resulted in a direct increase in the sensitivity of testing. Second, 50-μm-thick tissue sections provide greater tissue sampling than thin paraffin-embedded specimens. Third, the modified Highman’s Congo Red protocol appears to provide greater differentiation of amyloid deposits from peripheral tissue than standard Congo Red staining. Finally, the types of TTR mutation studies may influence test results. Our results can be compared with two recent publications that used Congo red staining but with differing results: Ebenezer et al. examined three 50-μm-thick sections from two biopsy sites, with lower sensitivity in TTR-negative cases with and without neuropathy; their results were similar to that shown in Figure 1 but with fewer samples tested, whereas Leonardi et al. examined 7-μm-thick sections and had lower sensitivity for patients with and without neuropathy (the estimated volume of tissue analyzed appears <1/6 of that used in the present study).

Most outcomes in this manuscript are patient-centric, that is, address the patient experience and quality of life using validated PRO and QOL questionnaires. This approach is of growing importance in the delivery of high-quality healthcare. In the present study, the patients endorsed symptoms on questionnaires despite physician’s
determination of Coutinho stage 0. These findings highlight the discordance that may occur when mild symptoms are present but not reported to or recognized by treating physicians. In providing symptom and QOL relationships with objective pathological findings, we reinforce the importance of subjective patient report in the patient with ATTRv amyloid neuropathy. Consistent with the notion that small nerve fibers are affected early and prominently in the course of the ATTRv neuropathy, the present data highlight the relationship between small fiber sensory symptoms (both painful and nonpainful), autonomic symptoms, and amyloid deposition. The strength and consistency of this relationship lend credence to the importance of these subjective manifestations even in the earliest disease stages. The NTSS-6, a sensory symptom questionnaire with both painful and nonpainful items, reveals the negative symptom, numbness is the most prevalent symptom across all disease stages. In contrast, in those individuals with amyloid deposition present in the skin, but without clinically evident disease, that is, the earliest stages of the disease, numbness is less prevalent while positive symptoms, such as pricking, tingling and lancinating pain are more prevalent, suggesting “irritability” or hypersensitivity of intact nociceptors. The Norfolk QOL small fiber questionnaire that includes items such as pricking/tingling or electric shocks corroborates these reported symptoms in the NIS-LL = 0 cohort.

While pain and sensory symptoms are recognized features of ATTRv neuropathy, although arguably underemphasized—they were not among the “potential red flags” suggesting ATTRv neuropathy in a diagnostic guidance
down—the present data emphasize not only the progressive severity of pain and other sensory symptoms across disease stages and their association with cutaneous amyloid deposition, but also the extent to which pain interferes with mood, work, sleep, and other activities of daily life throughout the disease.

As expected, autonomic symptoms were present across disease stages, in particular, symptoms of cardiovascular, gastrointestinal, and sexual autonomic impairment, which were the most prominent autonomic symptoms, and progressed in parallel with disease progression. Of note, symptoms of orthostatic intolerance and cutaneous vaso-motor changes were prominent in patients without clinical evidence of neuropathy and significantly more severe in those individuals with cutaneous amyloid deposition. Throughout the disease stages, all autonomic symptoms were more severe in individuals with cutaneous amyloid deposition providing additional support for the measurement of cutaneous amyloid deposition as a biomarker for disease severity.

Our results show that amyloid deposition can be detected in the earliest disease stages—in a substantial number of patients before significant symptoms and/or clinical signs are evident. In the major, recent large-scale clinical trials, clinically evident peripheral neuropathy has been an inclusion criterion. While neuropathy criteria currently frame the use of disease-modifying treatments, our data suggest that objective pathological evidence of the presence of disease is detectable at an even earlier disease stage, that is, at a stage that would not have met entry criteria in these trials, and raise the possibility of interventions earlier in the course of the disease.

Several lines of evidence support an approach to early treatment. First, although surgical gene therapy with liver transplantation improves prognosis, peripheral neuropathy and organ impairment do not usually reverse, and may, progress. Second, despite unequivocal group benefits, the response to disease-modifying therapies with TTR tetramer stabilizers and gene silencing therapy is not universal and some patients continue to progress despite therapy. Earlier intervention, consistent with strategies advocated in other neurodegenerative diseases, for example, mild cognitive impairment and asymptomatic AD, and premotor Parkinson’s disease may improve these numbers.

This manuscript introduces the notion that cutaneous amyloid deposition is a biomarker for the underlying disease pathology and pathophysiology. The detection of amyloid deposition in the earliest disease stages, even prior to clinically detectable manifestations, supports the utility of cutaneous amyloid deposition as a diagnostic biomarker that could enhance patient care, assist with life planning, and guide therapeutic interventions. Further, the increase in amyloid deposition with disease severity and the correlation with PRO measures, structured examinations, and neuropathology suggests that this biomarker has pathophysiological relevance and lends additional support to the potential utility of this measure as a predictive and prognostic biomarker. Importantly, these findings are consistent with a sural nerve biopsy study that showed earlier loss of small compared with large nerve fibers. In this study, amyloid deposition correlated negatively with the time to onset of symptoms in asymptomatic patients. Longitudinal studies are required to confirm these findings in the skin.

This study has limitations and several questions remain. This is a cross-sectional study. It is not known whether amyloid deposition continues throughout the disease stages, whether deposition is continuous or has stepwise progression, and whether cutaneous amyloid will continue to correlate with the clinical manifestations of the disease. Serum neurofilament was not measured and the relationship between cutaneous amyloid deposition and serum neurofilament levels could not be determined. We were unable to volumetrically quantify the
amount of amyloid within a given skin biopsy section. Despite these limitations, the present data provide a framework for the exploration of these topics in future studies.

**Authors’ Contribution**

RF, AGD, FB, and CG contributed to the conception and design of the study; RF, AGD, FB, MC, SR, JG, JYK, NW, LO, and CG contributed to the acquisition and analysis of data; RF, AGD, FB, and CG contributed to drafting a significant portion of the manuscript or figures.

**Conflict of Interest**

RF received consultant fees and RF and AGD received a grant from Pfizer, which manufactures a drug to treat ATTRv cardiomyopathy. RF received a grant from Akcea, which manufactures a drug to treat ATTRv neuropathy. AGD received a grant from Alnylam, which manufactures a drug to treat ATTRv neuropathy. RF and CG have stock options in CND Life Sciences which is a skin biopsy company. The remaining authors have nothing to report.

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