Vitamin D and early rheumatoid arthritis

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Abstract

Background: Previous studies have linked rheumatoid arthritis (RA) risk and disease activity with vitamin D-deficiency (low serum 25-hydroxyvitamin D (25OHD)), but a causal role for vitamin D in RA is still unclear, with conflicting results from many previous studies, partly due to heterogeneity in study design and patient populations. In this study we aimed to (1) analyse serum 25OHD in early inflammatory arthritis, (2) compare 25OHD with disease activity and fatigue in early RA and (3) determine whether low 25OHD is associated with progression to RA.

Methods: An analysis of 790 patients recruited to the Birmingham Early Inflammatory Arthritis Cohort and followed longitudinally to determine clinical outcomes. The following were recorded at baseline: demographic data, duration of symptoms, duration of early morning stiffness (EMS), tender and swollen joint counts, Visual Analogue Scale (VAS) pain/fatigue/EMS, PHQ-9, HAQ and FACIT-Fatigue scores, DAS28-ESR, DAS28-CRP, CRP, ESR, anti-CCP antibody status, rheumatoid factor status, and serum 25OHD (ng/ml). Diagnosis was recorded at 0 and 12 months onwards as either RA, Undifferentiated Inflammatory Arthritis (UIA; synovitis not meeting other classification/diagnostic criteria), Clinically Suspect Arthralgia (CSA; arthralgia of an inflammatory type without synovitis), or Other.

Results: Baseline demographic data were similar between all groups, with median symptom duration of 16.8–34.0 days. Baseline 25OHD was not significantly different between groups [median, interquartile range (IQR): RA 46.7, 30.0–73.3; UIA 51.4, 30.0–72.3; CSA 47.7, 30.3–73.0; Other 39.9, 28.6–62.2]. In RA (n = 335), there were no significant differences between 25OHD and measures of disease activity or fatigue. No association between 25OHD and progression from UIA or CSA to RA was observed.

Conclusions: There was no clear association between serum 25OHD and baseline diagnosis, RA disease activity, or progression from UIA or CSA to RA. Future studies of other vitamin D metabolites may better define the complex role of vitamin D in RA.

Keywords: Vitamin D, Inflammation, Rheumatoid arthritis, Psoriatic arthritis, Undifferentiated inflammatory arthritis

Background

Rheumatoid arthritis (RA) is a chronic inflammatory disease characterised by synovitis affecting the small/medium joints and extra-articular manifestations [1]. Early diagnosis and treatment maximise chances of inducing remission thereby preventing permanent disabling joint damage [2]. Currently there are few clinical or biochemical markers to predict who will respond to RA treatment, and approaches to predicting which patients with undifferentiated inflammatory arthritis (UIA) or clinically suspect arthralgia (CSA) will progress to RA are imperfect [3, 4]. However, recent studies have suggested that vitamin D status may be a potential contributor to inflammatory disorders such as RA [5]. In vitro, vitamin D metabolites modulate inflammation through changes in T helper and regulatory T cell function [5, 6]. In vivo, vitamin D metabolites including the
main circulating metabolite 25-hydroxyvitamin D (25OHD), and the active form of vitamin D 1,25-dihydroxyvitamin D (1,25(OH)₂D) have been linked to RA disease progression [7–10], and vitamin D supplementation may have beneficial effects for patients with established RA [11, 12]. What is less clear is the role of vitamin D status in the onset and early progression of RA. In 2015, Cooles et al. reported no relationship between 25OHD and CRP, ESR, symptom duration, tender joint count (TJC), swollen joint count (SJC), or patients’ global health visual analogue scale (GH-VAS) in a small cohort of 73 RA patients [13], and similarly Neilen et al. found no evidence of low vitamin D in blood donor samples from patients in the pre-RA phase [14]. Conversely, other studies have reported an association of serum 25OHD or 1,25(OH)₂D status with some markers of disease activity in early RA [15, 16].

The aim of the current study was to use a large prospective cohort of early inflammatory arthritis patients to: (i) compare total serum 25OHD levels between early inflammatory arthritis patients with different baseline diagnoses; (ii) explore the relationship between 25OHD levels and measures of RA disease activity and of fatigue, and; (iii) determine whether low serum 25OHD is linked to progression from CSA or UIA to RA.

Methods
We enrolled patients with inflammatory arthritis or CSA newly seen in rheumatology clinics into the Birmingham Early Inflammatory Arthritis cohort (BEACON) between 2013 and 2019 from rheumatology outpatient departments at Sandwell and West Birmingham NHS Trust and University Hospitals Birmingham NHS Trust. Demographic information including age, sex, BMI, smoking status and ethnicity were recorded. In addition, clinical data including duration of symptoms, early morning stiffness (EMS), Tender Joint Count (TJC) and Swollen Joint Count (SJC), DAS28-ESR, DAS28-CRP, visual analogue scale (VAS) pain, fatigue and ESR, PHQ9, HAQ and FACIT-Fatigue scores were collected, alongside CRP, ESR, anti-CCP antibody and RF levels, at baseline (0 months (M)). Classification criteria for RA [17] were applied at 0 M and at 12 M or beyond. In addition, we looked at two distinct groups of patients representing different time points in the transition to RA. Patients with synovitis not meeting criteria for RA or other rheumatological diseases (i.e. UIA) and inflammatory-type arthralgia without synovitis (CSA). Serum 25OHD concentrations were analysed using NHS laboratory assays and reported as ng/mL.

Box-and-whisker diagrams depicting the median and interquartile range (IQR) were used to describe non-parametric variables, and percentages for categorical variables. Significance between groups was determined by non-parametric testing with either Pearson’s correlation coefficient, Kruskal-Wallis test and Chi-squared tests as appropriate, using SPSS v25. Ethical approval was granted by the NRES Committee West Midlands - The Black Country (REC 12/WM/0258).

Results
Overall 790 patients were included in the study. Baseline age, sex, smoking status, BMI and symptom duration are shown in Table 1. There were significant differences noted between groups for age and symptom duration (p < 0.001 for both). The median duration of symptoms at time of clinic visit was 26 days, however there was significant variation (IQR 13.3–54.4). Data on ethnicity were also recorded: 60.7% of patients considered themselves as White British, 20.4% Asian or Asian British and 7.8% Black or Black British.

We initially compared 25OHD levels between clinical groups [RA, UIA, CSA and other diagnoses (e.g. psoriatic arthritis, gout)] at initial clinical presentation to a Rheumatologist and prior to the commencement of disease modifying anti-rheumatic drug (DMARD) therapy for the patient’s arthritis where clinically indicated. There were no significant differences between groups (Fig. 1). However, there was a wide distribution of 25OHD values across all groups [median IQR (ng/mL): RA = 46.7 (30.0–73.0), UIA = 51.5 (30.0–72.7), CSA = 47.7 (30.3–73.0), Other = 39.9 (28.6–62.2)]. Of note, a significantly higher number of RA patient samples were taken from March to September (Suppl Table 1), a time of year when most adults will be able to synthesise vitamin D from sunlight [18]. In the RA group, those tested between March and September had on average a higher baseline vitamin D, although this was not statistically significant (Suppl Fig. 1, p = 0.165).

Subsequently, we analysed whether serum 25OHD levels correlated with clinical disease activity measures and other disease related variables in patients diagnosed with RA at baseline. 25OHD levels were recorded for 335 of these patients; there were no clear associations between serum 25OHD levels and baseline EMS duration, TJC, SJC, DAS28-ESR, DAS28-CRP, VAS pain, VAS fatigue, VAS EMS, PHQ9, HAQ, FACIT-Fatigue, CRP or ESR (Table 2).

Finally, we took CSA and UIA patients who had been enrolled in the study for at least 12M, and compared baseline serum 25OHD levels in those who progressed to other diagnoses at 12 M with those whose diagnosis remained unchanged. For the CSA group (n = 150), follow-up data were not available for 36 patients (24%), and a further 9 patients did not have vitamin D levels measured at baseline, leaving 105 patients eligible for inclusion in the subgroup analysis. Of these, 60 remained CSA, 28 progressed to RA, 11 progressed to UIA and 6
Table 1 Baseline patient characteristics

|                                | All  | RA   | UIA  | CSA  | Other |
|--------------------------------|------|------|------|------|-------|
| No. patients                   | 790  | 368  | 139  | 169  | 114   |
| Age*, years mean (SD)          | 52.4 (15.5) | 55.5 (15.2) | 51.4 (15.4) | 46.3 (13.7) | 52.3 (16.5) |
| Missing (no.)                  | n = 0 | n = 0 | n = 0 | n = 0 | n = 0 |
| Sex, % female                  | 64.3  | 68.8  | 59.0  | 72.8  | 43.9  |
| Missing (no.)                  | n = 0 | n = 0 | n = 0 | n = 0 | n = 0 |
| Ethnicity, %                   |       |       |       |       |       |
| White British                  | 60.7  | 58.2  | 59.7  | 60.4  | 71.1  |
| White Other                    | 4.7   | 4.6   | 3.6   | 6.6   | 3.5   |
| Asian or Asian British         | 20.4  | 20.4  | 22.3  | 21.9  | 15.8  |
| Black or Black British         | 7.8   | 9.0   | 10.8  | 4.8   | 5.3   |
| Other                          | 2.7   | 2.8   | 2.8   | 3.6   | 0.9   |
| Missing or not stated (no.)    | n = 27| n = 18| n = 1 | n = 4 | n = 4 |
| Smoking status, %              |       |       |       |       |       |
| Current                        | 19.9  | 20.5  | 21.7  | 21.3  | 18.4  |
| Previous                       | 31.5  | 29.9  | 34.5  | 28.4  | 37.8  |
| Never                          | 47.2  | 49.2  | 49.3  | 49.7  | 43.2  |
| Missing (no.)                  | n = 8 | n = 2 | n = 2 | n = 1 | n = 3 |
| BMI, kg/m² mean (SD)           | 28.7 (6.4) | 28.7 (6.5) | 28.0 (5.6) | 28.9 (7.1) | 29.5 (6.1) |
| Missing (no.)                  | n = 41| n = 24| n = 5 | n = 8 | n = 4 |
| Symptom duration*, days median (IQR) | 26.0 (13.3–54.4) | 24.1 (13.0–52.2) | 24.1 (13.0–57.0) | 34.0 (18.0–70.1) | 25.7 (10.5–55.7) |
| Missing (no.)                  | n = 12| n = 7 | n = 1 | n = 2 | n = 2 |

*C current, CSA clinically suspect arthralgia, N never, no. number of cases, P previous, RA rheumatoid arthritis, SD standard deviation, UIA unclassified inflammatory arthritis

* denotes statistically significant difference between groups (p < 0.001, Kruskall Wallis test)

progressed to other forms of inflammatory arthritis. Baseline vitamin D levels were not significantly different between groups (p = 0.851, Fig. 2). In the UIA group (n = 120), follow-up data were not available for 35 patients (28.9%), and vitamin D data were missing for a further 4 patients, leaving 81 patients eligible for inclusion in this subgroup analysis. Of these 81 patients, 58 patients remained UIA, 19 progressed to RA and 4 progressed to other forms of chronic inflammatory arthritis. Vitamin D levels were not significantly different between those who remained UIA, and those who progressed to RA or other diagnoses (p = 0.908, Fig. 3). Importantly, in the CSA and UIA groups, 25OHD levels at baseline were not significantly different in those without follow up data compared with those for whom follow up data were available (Suppl. Figure 2, p = 0.863).

Some CSA and UIA patients were commenced on DMARDs (methotrexate, hydroxychloroquine or sulfasalazine, alone or in combination) during follow-up if the treating clinician felt that was appropriate. Progression of either CSA or UIA to RA may be influenced by DMARD therapy commenced during the clinical follow up period as this may reduce rate of progression to RA in those at risk. Of the CSA patients who remained as having CSA, only one (baseline vitamin D level 68 ng/mL) had been commenced on a DMARD. Of the CSA patients who progressed to RA, eight (baseline vitamin D level 66.5 ng/mL (48.0–79.5)) had been commenced on a DMARD before RA development. In the CSA population, DMARD use during follow-up is thus unlikely to be a factor limiting transition to RA. Of the UIA patients who remained as having UIA, 27 had been commenced on a DMARD (baseline vitamin D level 46.9 ng/mL (28.3–67.0)) while of those who progressed to RA, 14 had been commenced on a DMARD before RA developed (baseline vitamin D level 50.1 ng/mL (32.0–60.0)). It is possible that some of the UIA patients on DMARDs who remained UIA during follow-up may have actually developed RA had they not commenced a DMARD. However, of those UIA patients started on a DMARD, there was no difference in the baseline vitamin D level between those who progressed to RA and those who did not.

**Discussion**

This study exploring the relationship between vitamin D levels and clinical characteristics at presentation and final clinical diagnoses after follow up is, to our knowledge, the first of its kind to be conducted in an early
arthritis cohort. Furthermore, we assessed more disease related variables than previous studies, including markers relating to fatigue (VAS fatigue and FACIT-Fatigue scores). Nielen et al. have previously reported that vitamin D deficiency does not increase risk of progression to RA in patients at risk on the basis of positive serology and musculoskeletal symptoms in the absence of synovitis [14]. Similarly, in our cohort, despite there being a range of different 25OHD values in all groups, we did not find any relationship between 25OHD and clinical variables in early RA and the development of arthritis/RA in those at risk with CSA/UIA. This latter finding may be worth further exploration in a larger adequately powered cohort.

The main strengths of this study were the sample size, which is larger than previous early arthritis cohorts in which vitamin D has been analysed, and therefore provides novel insight into vitamin D at this critical time point in the trajectory of inflammatory joint disease. We compared serum 25OHD levels with a wide range of clinical and biochemical markers of disease, including both subjective and objective measures, giving a comprehensive insight into their relationship with serum 25OHD. Previous studies have found conflicting results as to whether there is a relationship between vitamin D levels and markers of disease activity [19]. Data presented in the current study strongly suggest that low serum 25OHD is not linked to any marker of disease activity in early RA.

Furthermore, the inclusion of patients with UIA provides a novel insight into this facet of arthritis.

There are important limitations to our data. Firstly, although the study had large numbers of patients, there was significant loss to follow up in all groups at 12 months. This reduced power to detect significant differences in 25OHD levels between those UIA and CSA patients who progressed to RA and those who did not progress to RA. Secondly, a significantly higher number of RA patient samples were taken from March to September; a time of year when most adults will be able to synthesise vitamin D from the sunlight [18]. In the RA group those tested between March and September had on average a higher baseline vitamin D, although this was not statistically significant. Nevertheless, this could have impacted on our ability to detect any relationship between low 25OHD and markers of RA disease activity. In addition, we were unable to reliably determine which patients were receiving vitamin D supplements, and when they may have begun these in relation to the onset of their joint symptoms, owing to the frequent (and inconsistently recorded) use of over-the-counter (OTC) supplements. GPs are encouraged to recommend OTC supplements to patients with vitamin D insufficiency (30-50 nmol/L serum 25OHD) rather than issuing a prescription for vitamin D [18]. Accordingly, GP records could not be used as a means of determining which patients were taking vitamin D supplements at baseline. Many patients do not tell doctors they take vitamin supplements as they do not necessarily consider them medications, and furthermore, patients are usually unaware of their preparation’s formulation (vitamin D2 vs D3) or strength making it impossible to adjust for vitamin D supplementation use in our analysis.

This study focused on total serum 25OHD concentrations as this is the analysis used in most routine assay services. However, there are multiple vitamin D metabolites which affect both innate and adaptive immune processes at the molecular level, and these have been shown to play a role in the aetiopathogenesis of RA. Studies have shown the active form of vitamin D, 1,25(OH)2D, promotes tolerogenic DC cell function [20, 21], maintains the balance of Th1/Th2 cells [22], suppresses the proinflammatory TH17 phenotype [23–25], and promotes T-regulatory (Treg) cell function [26, 27], whilst facilitating migration of T cells to the site of inflammation and phenotype stabilisation [28–30]. Similarly, the vitamin D receptor (VDR), which plays an important role in facilitating PBMC responses to 1,25(OH)2D in immune homeostasis [31], is altered in RA [32] and VDR polymorphisms are linked to the development of arthritis [33]. Thus, it is likely that simple measurement of serum 25OHD provides only a limited perspective on the potential impact of vitamin D on diseases such as
RA. Further studies are needed to determine if other forms of vitamin D are more effective determinants of its anti-inflammatory action.

The compartment in which vitamin D is measured may also be important. Serum can be obtained through simple phlebotomy, but the levels and behaviour of vitamin D and its metabolites at the site of inflammation, the synovial joint, may differ relative to peripheral blood. Our group has reported differences in vitamin D and its metabolites between serum and synovial fluid in patients with established RA [34]. Although we did not measure cytokine expression, evidence from in vitro and murine studies suggests the differences inflammatory cytokine milieu in the synovial tissue compartment will modulate the effects of vitamin D further [35]. Thus, future studies should focus on analysis of multiple vitamin D metabolites and their relationship with inflammatory factors within specific microenvironment sites of RA disease. Further research is thus needed to better elucidate the role of vitamin D and its metabolites not only in the peripheral blood, but at the site of inflammation, the synovial joint. Understanding how vitamin D changes with, and as a result of, the inflammatory milieu in the synovial joint will provide new insights, and potential novel treatment approaches. In recent studies of T cells from RA patients we observed much stronger responses to 1,25(OH)2D in T cells from the peripheral blood compared to paired inflamed joint synovial fluid from the same patient [36]. This suggests that once RA disease is established, there is insensitivity to 1,25(OH)2D in immune cells, with this effect severely limiting the anti-inflammatory benefits of 1,25(OH)2D. Thus, it is possible that, even in early RA disease, much higher levels of vitamin D than previously thought are required to induce beneficial effects on synovial inflammation.

**Conclusion**

This study did not show any clear associations between serum 25OHD and baseline diagnosis, in patients with a range of different inflammatory arthritides, disease activity in patients with RA, or progression from UIA or CSA to RA. Our data thus provide further clarity on an issue where previous studies have yielded conflicting data; that is, whether RA disease activity is linked to serum vitamin D levels. Future research should focus on validating these findings including those related to vitamin D at baseline and subsequent disease trajectories, in another cohort powered to detect statistical significance. Furthermore, it will be important to assess different vitamin D

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**Table 2** Correlations between serum 25OHD concentrations at baseline and clinical characteristics of RA patients at baseline

| No. patients | Correlation co-efficient | P value |
|--------------|--------------------------|---------|
| Duration EMS | 335                      | 0.074   | 0.191 |
| ESR          | 335                      | −0.045  | 0.418 |
| CRP          | 321                      | −0.015  | 0.785 |
| TJC68        | 333                      | 0.075   | 0.170 |
| SJC66        | 333                      | 0.029   | 0.601 |
| TJ28         | 333                      | 0.077   | 0.163 |
| SJC28        | 333                      | 0.017   | 0.758 |
| VAS Patient global | 327                  | −0.008  | 0.157 |
| DAS28-ESR    | 315                      | 0.037   | 0.513 |
| DAS28-CRP    | 324                      | 0.047   | 0.402 |
| VAS pain     | 319                      | 0.012   | 0.831 |
| VAS EMS      | 115                      | −0.103  | 0.272 |
| VAS fatigue  | 319                      | −0.021  | 0.713 |
| FACIT fatigue| 287                      | 0.005   | 0.926 |
| HAQ          | 286                      | −0.014  | 0.808 |
| PHQ9         | 297                      | −0.006  | 0.913 |

CRP C-reactive protein, DAS disease activity score, EMS early morning stiffness, ESR erythrocyte sedimentation rate, FACIT functional assessment of chronic illness therapy, HAQ health assessment questionnaire, PHQ patient health questionnaire, SJC swollen joint count, TJC tender joint count, VAS visual analogue scale

**Fig. 2** Concentrations of serum 25OHD for all CSA patients at baseline according to final diagnosis after 12 months. Boxes represent median and interquartile range (IQR), whiskers represent the range. Values for each group are as follow: (i) Remained CSA, n = 60, median 47.1, IQR 29.6–68.8; (ii) Progressed to UIA, n = 11, median 42.0, IQR 28.5–88.5; (iii) Progressed to RA, n = 28, median 51.8, IQR 25.9–76.0; (iv) Progressed to Other; n = 6, median 54.0, IQR 40.8–81.5. There were no statistically significant differences between groups (p = 0.851)
metabolites, their carrier proteins, and enzymes involved in vitamin D metabolism that might affect the local action of vitamin D in the joint, to allow a more comprehensive understanding of the role of vitamin D in RA pathogenesis.

Supplementary information
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Additional file 1: Suppl. Table 1. Number of patients in each diagnostic group who had their serum 25OHD measured between March and September.

Fig. 3 Concentrations of serum 25OHD for all UIA patients at baseline according to final diagnosis after 12 months. Boxes represent median and interquartile range (IQR), whiskers represent the range, and beyond this any extreme outlier values are also shown. Values for each group are as follows: (i) Remained UIA, n = 58, median 52.0, IQR 29.5–71.0; (ii) Progressed to RA (n = 19, median 48.8, IQR 32.0–61.0); (iii) Progressed to Other; n = 4, median 52.5, IQR 24.7–82.3. There were no statistically significant differences between groups (p = 0.908).

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Authors’ contributions
SRH compiled datasets, analysed the data and wrote the manuscript. GJ compiled datasets and contributed to data analysis. DL designed the study and edited the manuscript. IS collected patient samples and developed the patient datasets. AF collected patient samples, developed patient datasets and designed the study. MH designed the study and wrote the manuscript. KR designed the study, organised patient sample collection, developed the patient datasets and wrote the manuscript. The author(s) read and approved the final manuscript.

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Availability of data and materials
All data generated or analyzed during this study are included in this published article [and its supplementary information files] or are available from the corresponding author on reasonable request.

Ethics approval and consent to participate
Ethical approval for this study was granted by the NRES Committee West Midlands - The Black Country (REC 12/WM/0258). All participants gave written informed consent to participate.

Consent for publication
Not applicable.

Competing interests
The authors have no competing interests to declare.

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20. Anderson AE, Smolen JS. Diagnosis and Management of Rheumatoid Arthritis. A review. J Am Med Assoc. 2018;320(3):1360–72.
21. Symmons DP, Jones MA, Scott DL, Prior P. Longterm mortality outcome in patients with rheumatoid arthritis: early presenters continue to do well. J Rheumatol. 1998;25(6):1072–7.
22. Silvagni E, Di Battista M, Bonifacio AF, Zucchi D, Governato G, Sciri CA. One year in review 2019: novelties in the treatment of rheumatoid arthritis. Clin Exp Rheumatol. 2019;37(4):S19–34.
23. Boets DM, Raza K, van der Helm-van Mil AHM. Which patients presenting with arthralgia eventually develop rheumatoid arthritis? The current state of the art. RMD Open. 2017;3(2):e000479.
24. Jeffery LE, Raza K, Hewison M. Vitamin D in rheumatoid arthritis—towards clinical application. Nat Rev Rheum. 2016;12(4):201–10.
25. Colin EM, Asmawidjaja PS, van Hamburg JP, Mus AM, van Driel M, Hazer JM, et al. 1,25-dihydroxyvitamin D3 modulates Th17 polarization and interleukin-22 expression by memory T cells from patients with early rheumatoid arthritis. Arthritis Rheum. 2010;62(1):132–42.
26. Lin J, Liu J, Davies ML, Chen W. Serum vitamin D level and rheumatoid arthritis disease activity: review and meta-analysis. PLoS One. 2016;11(1): e0146531.
27. Oelzer P, Muller A, Deschner F, Muller M, Abendroth K, Hein G, et al. Relationship between disease activity and serum levels of vitamin D metabolites and PTH in rheumatoid arthritis. Calcif Tissue Int. 1998;62(3):193–8.
28. Merlino LA, Curtis J, Mikuls TR, Cerhan JR, Criswell LA, Saag KG. Vitamin D intake is inversely associated with rheumatoid arthritis: results from the Iowa women’s health study. Arthritis Rheum. 2004;50(1):72–82.
29. Rossini M, Maddali Bongi S, La Montagna G, Miniscoli G, Malavolta N, Benini L, et al. Vitamin D deficiency in rheumatoid arthritis: prevalence, determinants and associations with disease activity and disability. Arthritis Res Ther. 2010;12(6):R216.
30. Soubrier M, Lambert C, Combe B, Gaudin P, Thomas T, Sibilla J, et al. A randomised, double-blind, placebo-controlled study assessing the efficacy of high doses of vitamin D on functional disability in patients with rheumatoid arthritis. Clin Exp Rheumatol. 2018;36(6):1056–60.
31. Chandrashekara S, Patted A. Role of vitamin D supplementation in improving disease activity in rheumatoid arthritis: an exploratory study. Int J Rheum Dis. 2017;20(7):825–31.
32. Coiles FA, Pratt AG, Lendrem DW, Ng WF, Aspray TJ, Isaacs JD. Retrospective analysis of the role of serum vitamin D in early rheumatic disease. Rheumatology. 2015;54(2):1374–5.
33. Nielen MM, van Schaardenburg D, Lems WF, van de Stadt RJ, de Koning L, et al. The D-1 vitamin metabolite 1,25(OH)2D in serum is associated with disease activity and anti-Citrullinated protein antibodies in active and treatment naive, early rheumatoid arthritis patients. Scand J Immunol. 2018;88(3):e12704.
34. Taylor WJ, Marcheson A, Arreghini M, Sokoll K, Helliwell PS. A comparison of the performance characteristics of classification criteria for the diagnosis of psoriatic arthritis. Seminars Arthritis Rheumatism. 2004;34:575–84.
35. NICE Clinical knowledge summaries (CKS). Vitamin D Deficiency in Adults – treatment and prevention. 2018.
36. Harrison SR, Li D, Jeffery LE, Raza K, Hewison M, Vitamin D, Autoimmune disease and rheumatoid arthritis. Calcif Tissue Int. 2020;106:558–75.
37. Anderson AE, Swan DJ, Wong OY, Buck M, Etherington O, Harry RA, et al. Tolerogenic dendritic cells generated with dexamethasone and vitamin D3 regulate rheumatoid arthritis CD4+ T cells partly via transforming growth factor-beta1. Clin Exp Immunol. 2017;187:113–23.
38. Bscheider M, Butcher EC. Vitamin D immunoregulation through dendritic cells. Immunology. 2016;148:227–36.
39. Zhang Z, Chen F, Li J, Luo F, Hou T, Xu J, et al. 1,25(OH)2D3 suppresses proinflammatory responses by inhibiting Th1 cell differentiation and cytokine production through the JAK/STAT pathway. Ann Transl Res. 2018;6(9):1751–6.
40. Nistala K, Adams S, Cambrok H, Ursu S, Olovito B, de Jager W, et al. Th17 plasticity in human autoimmune arthritis is driven by the inflammatory environment. Proc Natl Acad Sci USA. 2010;107(33):14751–6.
41. van Hamburg JP, Asmawidjaja PS, Davelaar N, Mus AM, Colin EM, Hazes JM, et al. Th17 cells, but not Th1 cells, from patients with early rheumatoid arthritis are potent inducers of matrix metalloproteinases and proinflammatory cytokines upon synovial fibroblast interaction, including autocrine interleukin-17A production. Arthritis Rheumatism. 2011;63(1):73–83.
42. Zhou L, Wang J, Li J, Li T, Chen Y, June RR, et al. 1,25-Dihydroxyvitamin D3 ameliorates collagen-induced arthritis via suppression of Th17 cells through miR-124 mediated inhibition of IL-6 signaling. Front Immunol. 2019;10:178.
43. Jeffery LE, Queoshi OS, Gardner D, Hou TZ, Biggs Z, Sokic B, et al. Vitamin D Antagonises the suppressive effect of inflammatory cytokines on CTLA-4 expression and regulatory function. PLoS One. 2015;10(7):e0131539.
44. Jeffery LE, Burke F, Mura M, Zheng Y, Queoshi OS, Hewison M, et al. 1,25-Dihydroxyvitamin D3 and IL-2 combine to inhibit T cell production of inflammatory cytokines and promote development of regulatory T cells expressing CTLA-4 and FoxP3. J Immunol. 2009;183(9):5485–67.
45. Baek P, Korf H, Overbergh L, Versyft A, Thorez L, Van Lommel L, et al. The vitamin D analog, TSK27, promotes a human CD4+ CD25 high CD127 low regulatory T cell profile and induces a migratory signature specific for homing to sites of inflammation. J Immunol. 2011;186:132–42.
46. Cantorna MT, Snyder L, Lin Y-D, Yang L, Vitamin D and 1,25(OH)2D regulation of T cells. Nutrients. 2015;7:3011–21.
47. Moradi B, Schnitzer P, Hagnemann S, Rosshart N, Gottherram B, Kretzer JP, et al. CD4+CD25+high CD127 low− regulatory T cells are enriched in rheumatoid arthritis and osteoarthritides–joints–analysis of frequency and phenotype in synovial membrane, synovial fluid and peripheral blood. Arthritis Res Ther. 2014;16:R97-R.
48. Bhalia AK, Amento EP, Clemens TL, Holick MF, Krane SM. Specific high-affinity receptors for 1,25-dihydroxyvitamin D3 in human peripheral blood mononuclear cells: presence in monocytes and induction in T lymphocytes following activation. J Endocrinol Metabol. 1983;57(6):1308–10.
49. He XJ, Ding Y, Xiang W, Dang XQ. Roles of 1,25(OH)2D3 and vitamin D receptor in the pathogenesis of rheumatoid arthritis and systemic lupus erythematosus by regulating the activation of CD4+ T cells and the PKCε/ERK signaling pathway. Cell Physiol Biochem. 2016;40(3):743–56.
50. Mukhtar M, Sheikh N, Susgana SK, Batooll A, Fatima N, Mehmood R, et al. Vitamin D receptor gene polymorphism: an important predictor of arthritis development. Biomed Res Int. 2019;2019:8326468. https://doi.org/10.1155/2019/8326468.
51. Li D, Jeffery LE, Jenkinson C, Harrison SR, Chun RF, Adams JS, et al. Serum and synovial fluid vitamin D metabolites and rheumatoid arthritis. J Steroid Biochem Mol Biol. 2019;187:1–8.
52. Albtobeli E, Angeletti PM, Piccolo D, De Angelis R. Synovial fluid and serum concentrations of inflammatory markers in rheumatoid arthritis, psoriatic arthritis and osteoarthritis: a systematic review. Curr Rheumatol Rev. 2017;13(1):70–9.
53. Jeffery LE, Henley P, Marium N, Filer A, Sansom DM, Hewison M, et al. Decreased sensitivity to 1,25-dihydroxyvitamin D3 in T cells from the rheumatoid joint. J Autoimmun. 2018;88:50–60.

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