We are IntechOpen, the world’s leading publisher of Open Access books
Built by scientists, for scientists

5,000
Open access books available

125,000
International authors and editors

140M
Downloads

Our authors are among the

154
Countries delivered to

TOP 1%
most cited scientists

12.2%
Contributors from top 500 universities

WEB OF SCIENCE™
Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com
Plasma Exchange in Severe Attacks Associated with Neuromyelitis Optica Spectrum Disorder

Bonnan Mickael and Cabre Philippe
Service de Neurologie, Centre Hospitalier Universitaire Zobda Quitman, Fort-de-France,
French West Indies

1. Introduction
Neuromyelitis optica (NMO) is an inflammatory disorder restricted to the spinal cord and optic nerves. Contrary to multiple sclerosis (MS), relapses of NMO are often strikingly severe and most NMO patients present stepwise neurological impairments. NMO treatments are aimed to prevent the relapses with the administration of various promising immunosuppressive drugs. However, relapse treatment is still a tricky problem. Since the largely used steroid treatment usually fails to control severe attacks, specific add-on treatments have to be considered in order to limit the stepwise increase of residual impairment. Given that a strong humoral response characterizes NMO physiology, one might assume plasma exchange (PLEX) to be particularly well adapted in severe NMO relapses.

This chapter will analyze the relevant data of PLEX in the setting of NMO spectrum disorder. We will first outline the physiological grounds leading to the rationale of the PLEX treatment and the technical aspects of the procedure. Then we will assess the clinical results obtained in each type of attacks. Finally we will try to build an original concept linking the clinical results and the timing of PLEX onset with the dynamic of the inflammation inside the lesions.

2. Physiopathology of NMO

The physiopathology of demyelinating disorders is complex and may grossly be divided in two distinct parts: a cellular response implying lymphocytes, macrophages and granulocytes in NMO; and a humoral response involving many circulating components including antibodies, complement and cytokines which will be extravasated into the inflammatory sites where they will participate to the inflammatory cascade of events leading to the local lesion. We will briefly review the main physiopathological aspects of NMO and focus on the humoral response, which is especially suitable to be eliminated by PLEX upstream to the lesion.

2.1 Physiopathology of NMO lesions

2.1.1 Pathology

A characteristic pathological pattern has been described in NMO (Lucchinetti et al., 2002). Lesions are infiltrated by neutrophils and eosinophils and wall capillaries are hyalinized.
vasculocentric pattern of activated complement and immunoglobulin of IgG and IgM types is observed that mirrors the normal expression of AQP4. AQP4 expression is definitely reduced in normal appearing white matter and lost throughout the lesions. These modifications are the hallmark of NMO and could occur alone or associated with a wide range of lesions from mild demyelination to large necrosis. This pattern of lesion was classified in the pattern II of the Lassmann’s classification of the inflammatory lesions (Lucchinetti et al., 2000, 2002).

Contrary to MS, T cells are rare in NMO lesions and probably had no major effect on the formation of the lesions (Saadoun et al., 2011). However T cells are probably involved upstream in physiopathological cascade in the earlier phases of the disease where a complex interplay leads to antigen sensitization and possibly in the initial opening of the blood-brain barrier (BBB) (Pohl et al., 2011).

2.1.2 Specific antibodies and their epitopes

The NMO-IgG antibody is an IgG1 directed against a surface epitope of the protein aquaporin-4 (AQP4) (Lennon et al., 2005). This antibody is detected with tissue-based immunofluorescence assays with a sensitivity and specificity for clinically defined NMO of more than respectively 60% and 90%. NMO-IgM theoretically coexists with NMO-IgG in about 10% of cases but significance is poorly known (Jarius et al., 2010a). Clinically diagnosed NMO patients share common clinical and evolutional characteristics according to the NMO-IgG status. Beyond the surrogate marker value of NMO-IgG, this marker is now used as a major diagnostic criterion (Wingerchuk et al., 2006) and delineates the NMO spectrum disorders that gather in a same entity both typical NMO and unusual or truncated clinical forms (Wingerchuk et al., 2007).

Astrocytes closely interact with endothelial cells to maintain the CNS BBB. These cells express AQP4 in the apical domain of the membrane feet expansions situated close to the surrounded blood vessels. They are generally found as single tetramers, closely arranged in orthogonal arrays. This transmembrane protein is critically involved in the homeostasis of the water in the brain and interfaces with blood vessels, especially in the clearance of free water. Loss of perivascular AQP4 in the basal state results in cellular swelling, ostensibly due to a failure to eliminate water generated from cellular metabolism (Amiry-Moghaddam et al., 2003). Thus in NMO, since the interaction of NMO-IgG and AQP4 leads to a functional knock-out phenotype of AQP4, edema develops as a result of functional impairment of AQP4 although BBB is expected to be still intact, which may explain the paradoxical lack of gadolinium enhancement in most NMO lesions. Apart from water homeostasis, the removal of AQP4 from astrocytes membrane is associated with an impaired homeostasis of glutamate via the loss of function of EAAT2, a major glutamate transporter associated with AQP4 in a macromolecular complex (Hinson et al., 2008). The disruption of glutamate homeostasis initiates an excitotoxic mechanism damaging oligodendrocytes and ultimately leading to demyelination (Marignier et al., 2010).

Virtually all the CNS astrocytes express AQP4, however some regions are enriched in AQP4. Those regions are the spinal cord gray matter, the posterior optic nerve, the floor of the fourth ventricle and the circum ventricular organs especially the area postrema, explaining the restriction of the sites of lesion characterizing NMO (Pittock et al., 2006a). Interestingly
circumventricular organs are also the only sites of the CNS expressing fenestrated capillaries favoring local passive diffusion of circulating antibodies.

2.1.3 NMO-IgG and complement as key factors

Clinical activity may correlate with the underlying NMO-IgG titres. NMO-IgG detection is a strong predictor of recurrence after an initial spinal or optic attack (Jarius et al., 2008, 2010b; Weinschenker, et al 2006). In few patients, NMO-IgG was high during flares and became negative during the stabilized disease following treatment, and in contrary, an initially seronegative patient became positive during a further attack (Weinstock-Guttman et al., 2008). In the seminal work of Takahashi et al. (2007), NMO-IgG levels were positively correlated with both clinical severity (i.e. blindness) and radiological severity. Moreover a strong positive correlation was obtained between the NMO-IgG titres at the nadir of exacerbations and the spinal cord lesion length on MRI (Takahashi et al., 2007). In contrast, low NMO-IgG titres were observed during remission induced by immunosuppressive maintenance therapy (Jarius et al., 2008).

In vitro, the binding of NMO-IgG to the extracellular domain of AQP4 reversibly down-regulates its plasma expression. In the presence of active complement, this binding leads to strong complement activation and rapid cell destruction. NMO serum IgM is not AQP4-specific and abundant IgM deposits in the NMO lesions may have passively diffused after the BBB disruption by the seminal focal complement activation initiated by NMO-IgG (Hinson et al., 2007).

In an animal model of EAE with passive transfer of NMO-IgG, the transfer exacerbated EAE signs and the typical pathological characteristics were reproduced in treated rats (Kinoshita et al., 2009; Bradl et al., 2009). Direct injection of NMO-IgG in rat brains could reproduce the pathology but only when complement is coinjected (Saadoun et al., 2010).

The NMO-IgG ability to lesion AQP4-transfected cells in the presence of complement was assessed with serum drawn from patients with mild and severe attacks. The percentage of cells lesioned by complement was strongly higher in presence of sera from patients with severe attacks, although lesion induced by sera from patients with mild attacks did not differ from negative controls or MS patients (Hinson et al., 2009). Thus the severity of the disease may be partly determined by intrinsic NMO-IgG characteristics to activate the complement.

2.2 Proof of concept of PLEX in NMO

As we already described, NMO lesions are associated with a strong IgG, IgM and complement deposition, typical of the pattern II in Lassmann’s classification. The NMO-IgG is involved in a complement-dependant toxicity against the astrocytes. All of these components -IgG, IgM, and complement- are targeted by plasma exchanges. By means of 5 exchanges, all the exchanged molecules will drop to less than 20% of their initial level. By this way, antibodies and complement, which are the core of the pattern II lesions, are excluded from the circulating pool and cannot migrate anymore to the lesions.

Although PLEX has long been used in various demyelinating disorders (Keegan et al., 2002), there is some clue that the pattern is a key determinant of PLEX efficiency. In a retrospective
study, Keegan et al. (2005) reported that all the patients suffering from demyelinating disorders and improved by PLEX had a biopsy proven pattern II lesion. None of the patients with any other kind of lesion improved. However all these patients were MS without NMO-IgG and none were NMO (Keegan et al., 2005; Kale et al., 2009).

All the aforementioned findings stress that circulating NMO-IgG and complements are the two main actors of the NMO pathogeny and why clearing them from blood with PLEX should be appropriate for special benefits.

3. Plasma exchange procedure

3.1 Principles and goals

Basically, the goal of PLEX (or plasmapheresis) is to remove a given volume of patient’s plasma containing harmful targeted substances and to reinfuse an artificial plasma substitute in its place - the plasma exchange (Brecher, 2002).

3.2 Technique

PLEX are carried on in a nephrology or a resuscitation ward. Two high flow rate accesses are mandatory: an input line from patient to device ('artery') and a return line from device to patient ('vein'). In continuous filtration, two needles are placed in both arms or groins in order to draw out the blood of the body through an extracorporeal line connected to one needle, then blood is processed and reinfused continuously through the other needle. In case of discontinuous filtration, the separation and remixing are done in small batches through a single venous access in the groin where in and out cycles may alternate. Anticoagulation (citrate or heparin) is added to the blood pre-plasma filter to prevent from clotting. The removed blood is processed (apheresis procedure) in a cell separator that continuously separates plasma from cellular components (consisting of red and white blood cells and platelets) either by a centrifugation ring with permanent in and out flow, or by filtration through a porous membrane. Small molecules like cytokines as well as large molecules, such as albumin and immunoglobulin, are easily extruded from the blood compartment with a reported sieving coefficient >0.95 at a blood flow rate of 100 mL/min. The cleared cellular components are then combined with the replacement fluid (donor plasma or artificial albumin mixed with a saline solution) and returned to the patient through the needle in the other arm. A PLEX session is usually performed in 2 to 6 hours, depending on patient’s height, weight, viscosity of the blood and technical parameters.

3.3 Kinetics of the target exchanged components.

All the targeted components are distributed in the interstitium (extra-vascular compartment) by variable part. Large molecular weight compounds equilibrate slowly between the vascular space and the interstitium. Calculations of the rate of removal are simplified to first order kinetics. The relation curve of the achieved concentration of a plasma component \([C]\) after a unique exchange of a given plasma volume \(V\) is an exponential inverse: 
\[
\frac{[C]}{[C_0]} = e^{-\frac{V}{\text{Estimated plasma volume (liters)}}}.
\]

The whole plasma volume can be approximated in an adult with the following formula:

\[
\text{Estimated plasma volume (liters)} = 0.07 \times \text{weight (kg)} \times (1 - \text{hematocrit}).
\]
The larger volume of plasma exchanged during each session clears a larger amount targeted circulating component. An exchange of one body plasma volume leads to the immediate clearance of 50% of the circulating component. A 1.3 body mass volume exchange that removes about 72% of C is generally agreed. Beyond, the volume to process increases massively for too little gain. However, according to the distribution of C in the interstitium, the achievement of the clearance of C will necessitate the use of multiple PLEX sessions separated by the time necessary for the equilibration of C concentration between interstitium and vascular spaces. The number and frequency of sessions should be evaluated according to the biological characteristics of the components to remove (synthesis level, vascular distribution, diffusion ability). An empirically driven number of 4 to 6 sessions is usually scheduled. The durability of the immunomodulatory effect after PLEX is difficult to assess and will depend on the turnover rate of the targeted humoral components. Concomitant intensive immunosuppressive therapy (i.e. steroids, mitoxantrone, mycophenolate mofetil, rituximab) will be required to sustain the obtained depletive effect.

3.4 Risks and side effects
PLEX are contraindicated in case of ongoing infectious disease, precarious hemodynamics and active hemorrhage (heparin). Immediate side effects are related to the extracorporeal line: hemodynamic instability, vaso-vagal syndrome, numbness or tingling, venous puncture hazards with excessive local bleeding, septicemia or allergy. Since blood coagulation factors are all depleted by PLEX, hemostasis is affected in variable ways: first, a hypocoagulation state is immediately achieved by the global depletion of all the coagulation factors for half a day; at day 2, short life pro-coagulant factors are regained but antithrombin-III synthesis is delayed leading to a hypercoagulable state until day 3. Preventive anticoagulation with heparin is always required since the high risk of thrombosis. Persistently low fibrinogen levels have been described with the concomitant use of high dosage steroid infusion. In summary PLEX are generally well tolerated and now commonly and safely used.

4. PLEX in severe attacks
Various regimens of high doses of intravenous methylprednisolone are used in first line of treatment ranging from 3 g infused in 3 days, to 10 g in 5 to 10 days, depending on authors. There is no evidence in favor of one regimen or another and efficacy assessment has never been addressed. Moreover, even if steroids reduce the inflammatory cellular response by triggering apoptosis of lymphocytes, they are clearly not sufficient because poor outcomes are still a common issue even when steroid treatment is given immediately after onset. We wish to develop here the evidence for the effectiveness of PLEX that we have been largely using as an add-on therapy for more than 10 years.

Of note, steroids were always used to treat relapse. When used the same day as PLEX procedure, steroids were infused at the end of each PLEX session. However methylprednisolone pharmacokinetics is characterized by a short half-life and PLEX demonstrated to have no effect on steroids biodisposibility (Assegba et al., 1988; Stigelman et al, 1984).
4.1 Spinal attacks

PLEX proved to be efficient in central demyelinating diseases in a randomized sham-controlled study (Weinshenker et al., 1999, 2001). Keegan et al. (2002) reviewed the clinical data from 59 patients who received PLEX for inflammatory demyelinating diseases, including 10 NMO and 6 acute transverse myelitis (ATM) cases. A moderate or marked improvement was obtained in half of NMO and ATM patient groups. The late final outcome at one year was more or less obtained during the first month after treatment in both groups, without regard to success or failure of treatment (Keegan et al., 2002; Brunot et al., 2011). A small number of case reports and few small studies were reported with variable issues. Judging improvement is even more complex due to the subjective classification of improvement in mild/moderate/marked instead of a quantified clinical exam (Brunot et al., 2011; Keegan et al., 2002; Munemoto et al., 2011; Llufriu et al., 2009). Moreover the natural history of single spinal relapse in NMO has never been addressed, so any improvement bias after PLEX is inappreciable in the absence of a control group. Finally, most authors used PLEX as a rescue treatment given late after the onset. For example PLEX was delayed from onset by a mean of 33±30 days in Brunot et al. (2011) and a median of 30 days [6 to 90 days] in Llufriu et al. (2009).

Although a synergistic effect of steroids and PLEX was long expected due to their complementary action, none of these studies compared PLEX-treated attacks with conventional steroid treatment given alone with add-on PLEX-treated attacks.

We previously refined these results in a study of outcome after severe spinal attacks associated with NMO spectrum disorders (Bonnan et al., 2009a). We included 96 spinal attacks from 43 patients, divided in two groups: 1) a steroid-only group designed from historical patients treated with steroids alone; 2) an active group treated both with PLEX and steroids. Steroid infusion was started immediately after patient admission. PLEX decision was raised at the same time and started as soon as possible during the two days later. As a major difference with other groups, PLEX was never initiated as a delayed rescue treatment after a standard steroid treatment failure. Since PLEX therapy is mainly expected to minimize residual impairment, we used the \( \Delta EDSS \) (calculated as the difference between residual and basal EDSS) as the main outcome.

If we except 5 PLEX delayed due to difficult medevac reasons, PLEX were initiated by a mean of 5.4 ± 3.1 days after attack onset with a median of 4 sessions.

There was no significant difference between the PLEX-treated and steroid-only groups for basal and acute EDSS (3.9 ± 2.9 vs 4.2 ± 2.9, and 7.9 ± 1.0 vs 8.0 ± 1.4; p=NS), however residual EDSS (5.1 ± 2.4 vs 6.8 ± 1.9, p<0.01) and mean \( \Delta EDSS \) (1.2 ± 1.6 vs 2.6 ± 2.4, p<0.01) were significantly lower in the PLEX-treated group than in the steroid-only group.

Basal EDSS dramatically influenced therapy outcome as shown in Table 1. During the first attack, although acute EDSS were similar in both groups (7.6 ± 1.2 vs 7.1 ± 1.5, p=NS), \( \Delta EDSS \) and residual EDSS were dramatically reduced in the PLEX-treated group (2.1 ± 1.9 vs 5.8 ± 2.0, p<0.01) given that acute EDSS was similar in this sub-group. In the two other sub-groups of basal impairment (EDSS 1.0 to 5.5 and EDSS ≥6.0), residual EDSS and \( \Delta EDSS \) tended to be lower in PLEX-treated attacks but no statistical signification could be obtained due to the small size of these groups.

www.intechopen.com
Table 1. Disability measured as EDSS during spinal attacks stratified with basal impairment. St: steroid-only treated group; St+PLEX: steroid and PLEX-treated group. Values are given as mean±SD (from Bonnan et al., 2009a).

| EDSS         | St (n=17) | St+PLEX (n=7) | p  | St (n=26) | St+PLEX (n=13) | p  | St (n=24) | St+PLEX (n=9) | p  |
|--------------|-----------|---------------|----|-----------|---------------|----|-----------|---------------|----|
| Basal        | 0.00      | 0.00          |    | 0.99      | 3.9±0.8       | 0.59| 7.4±1.0   | 7.1±0.8       | 0.52|
| Acute        | 7.1±1.5   | 7.6±1.2       | 0.52| 7.6±1.3   | 7.6±1.1       | 0.67| 8.9±0.9   | 8.6±0.6       | 0.24|
| Residual     | 5.9±1.9   | 2.1±1.9       | <01| 5.8±1.6   | 5.1±1.1       | 0.21| 8.5±1.1   | 7.6±1.0       | 0.05|
| ΔEDSS        | 5.9±1.9   | 2.1±1.9       | <01| 2.0±1.5   | 1.2±1.6       | 0.10| 1.1±0.8   | 0.5±0.8       | 0.11|

The classical Lazarus effect, defined as a very short-term dramatic improvement (Weinshenker et al., 2000), was rather unusual in our group but our study was not designed to analyse short-term improvement. The patients who experienced this effect have all received a very early treatment (less than 2 days). In Magana et al. (2011), patients who exhibited functional improvement did so within a median of 4 days (third PLEX), although a minority (6%) exhibited a delayed response (more than 2 months).

Minor side effects occurred in 24% of PLEX treated attacks and resulted in PLEX interruption once (84-year-old patient with pulmonary embolism).

In summary, PLEX-treated patients achieved a significantly better outcome after a spinal attack, especially if PLEX was given during the first attack. The exact effect of PLEX in previously impaired patients should be validated in a larger multicentric cohort. As PLEX proved to be a promising treatment in spinal attacks, it would now be unethical to design a study with a sham-treated control group.

Predictors of good outcome were studied in a large group of PLEX including 26 NMO patients (Bonnan et al., 2009a). The only good outcome predictor was normal or brisk reflexes in acute phase (Magana et al., 2011). Surprisingly, a short PLEX delay was associated with a good outcome in a first study (Keegan et al., 2002) but had no effect in a second study, although one should remind that median PLEX delay (23 days) was delayed in this later compared to our group. The same PLEX response rate was obtained irrespective of NMO-IgG serostatus in our cohort and in the Mayo Clinic cohort (Magana et al., 2011).

As a practical consequence, faced with a patient suffering from a severe relapse, the knowledge of NMO-IgG status should not be required to start PLEX as soon as possible, since PLEX was found efficient in NMO-IgG negative patients.

4.2 Optic attacks

Visual impairment in NMO is very severe. We previously showed that an immediate unilateral blindness occurred in a third of patients after the first optic neuritis (ON), and generally two attacks are sufficient to cause a definitive loss of vision (Merle et al., 2007). Few PLEX were undertaken after ON and a quick dramatic recovery is usual as we also observed (Bonnan et al., 2009b; unpublished results). Depending authors, PLEX were used immediately (Bonnan et al., 2009b) or as a delayed add-on therapy (Schilling et al., 2006;
Watanabe et al., 2007a; Trebst et al., 2009; Yoshida et al., 2010). After pooling severe ON patients (acute visual acuity < 1/10°) from available studies (Schilling et al., 2006; Ruprecht et al., 2004; Trebst et al., 2009) with ours (Bonnan et al., 2009b and unpublished results), data were gathered for 39 eyes. PLEX were given in median of 19 days in patients who recovered a visual acuity more than 1/10° (considered here as a treatment success) but 41 days in treatment failure. A clear effect of PLEX delay was observed since success rate was 8/8 (100%) during the first 11 days, than 4/7 (57%) from days 12 to 22, and 7/13 (53%) from days 23 to 73. Moreover, even when patients recovered, averaged residual VA tended to be lower in delayed PLEX patients (Figure 1).

Interestingly, the spontaneous recovery (> 1/10°) after severe ON treated by steroids alone was about 40% in our cohort (from Merle et al., 2007), which is very close to the recovery obtained in the two last groups of late PLEX. In conclusion, strong clues support that PLEX change the outcome of severe ON only when they are given early, however broader studies are still lacking to confirm this hypothesis.

![Fig. 1. Metanalysis of residual Visual Acuity as a function of PLEX delay (days) in severe ON attacks (acute VA<1/10°). See text.](www.intechopen.com)

### 4.3 Brain attacks

Apart from optico-spinal attacks, severe brain attacks are described in NMO, especially involving hypothalamus and medulla. Those lesions are usually severe and associated with blindness, central endocrine disorders or quadriplegia with respiratory failure. Brain lesions are common but are mostly asymptomatic (Pittock et al., 2006b). However symptomatic lesions involving supratentorial white matter are exceptional and extensive (Pittock et al., 2006b). Even if a favourable outcome after PLEX has been reported in a few severe cases (Viegas et al., 2009; Watanabe et al., 2009ab), comparative data are still lacking.

Posterior reversible encephalopathy syndrome (PRES) is an encephalopathy with consciousness and visual disturbances with rapidly reversible changes on MRI consistent with vasogenic edema. PRES are triggered by blood pressure instability or fluid stresses...
due to various causes. It seems to occur more often than coincidental in NMO patients: 5 out of 70 consecutive NMO-IgG patients evaluated at Mayo Clinic (Magana et al., 2009); 2 out of 5 in Hadassah Medical School, Israel (Eichel et al., 2008). Authors proposed that the auto-immune mediated disruption of the AQP4 water channel function may predisposes to PRES at comparable levels of acute illness (Magana et al., 2009). PLEX was involved as a trigger in one case with a good final outcome. In few cases, PLEX were implemented as curative treatment with an overall good outcome (Eichel et al., 2008; Magana et al., 2009).

5. Timing of PLEX: evolving to a key concept

Besides knowing PLEX are effective and safe, the central question remains: is PLEX necessary as-soon-as and as-often-as possible? Prospective, randomised, multi-centre clinical trials would be required to definitively answer the question. For most authors, to date PLEX are considered as an add-on rescue treatment after steroid failure. The European recommendation from EFNS is to start with an early steroid course no matter the severity (Sellner et al., 2010). Early escalation with PLEX is only recommended after a failure of a second course of steroids, that is to say that PLEX initiation may be postponed for more than a week. As we demonstrated before, PLEX efficiency is depends on the timing of initiation, ranging from immediate dramatic improvement (the Lazarus effect) to no effect according to whether they are given early or very late. We propose to regress to the dynamic of the inflammatory NMO lesions to explain why PLEX efficiency is strongly dependant of the timing of their onset.

5.1 Evidence for reversible dysfunction preceding irreversible tissue loss.

As we described above, the lesion is the consequence of a cascade of reversible events, susceptible to an external action. One could abruptly divide this cascade in two main points: a direct toxic action upon astrocytes and a bystander effect on oligodendrocytes and axons. Astrocyte dysfunction is initiated by the binding of NMO-IgG to the extracellular domain of AQP4 on the foot of the astrocyte membrane. In vitro, this binding reversibly down-regulates AQP4 plasma expression. The presence of fresh complement leads to a strong complement activation and a rapid cell destruction. IgG titres strongly correlates with the cytotoxic effect (Kalluri et al., 2010). By the other way, the removal of AQP4 from astrocytes membrane, due to internalization or cell death, impairs the clearance of free glutamate due to the dysfunction of the transporter EAAT2. The glutamate progressively accumulates and initiates an excito-toxic mechanism upon oligodendrocytes, ultimately leading to demyelination (Marignier et al., 2010).

The time sequence of these events was studied in lesions induced by direct mouse brain injection with NMO-IgG and complement (Saadoun et al., 2010). Loss of AQP4 and GFAP, and myelin breakdown were evident 7h following the injection. The inflammatory cells infiltration became evident later. Within 12h, axonal injury became prominent. By day 7, axonal loss and dying neurons were evident. Finally, one could suppose that a very early intervention targeting astrocytes dysfunction may prevent the progression to the bystander effect.
5.2 Evidence supporting an early treatment

The influence of treatment delay upon outcome has been addressed in a single study of first ON receiving steroid treatments (Nakamura et al., 2010). The outcomes were both visual acuity and the width of the retinal fibers layer evaluated with optic tomography (OCT). Patients were divided into two groups: one group with a good visual outcome, including a high residual visual acuity and high RFL; and a second group with a poor visual outcome in terms of low acuity and low RFL. Very interestingly, the two groups were similar in all the parameters except one: patients with a good outcome received steroids with a lower mean delay after ON onset, by a mean of 1.8±1.1 days compared to 7.8±3.8 in patients with a poor outcome. This study is the first proof that a delayed infusion of steroids is associated with a poorer outcome. A similar effect of treatment delay, although unknown, should be expected in spinal attacks. Even if no proof is yet available after a PLEX treatment, these clues could be gathered that early PLEX would improve the prognosis (see above).

In spinal attacks treated with PLEX, early initiation of treatment was one out of the predictors of good outcome (Keegan et al., 2002). In a larger study encompassing attacks from various demyelinating disorders, success rates were stratified by delay: improvement occurred in 83% when given before day 15, but fell to 43 after 2 months (Llufriu et al., 2009). Moreover the dramatically very short-term improvement, called Lazarus effect (Weinshenker et al., 2000), is sometimes observed after severe attacks receiving a very early treatment with PLEX and steroids. However, this earliness responsibility on the Lazarus effect remains elusive since no study is available on this rather unusual effect.

5.3 Lesion stages and PLEX action: ‘time is cord and eyes’

In the light of the available data, we postulate a link between the staging of NMO lesion and the PLEX effect upon clinical and radiological outcome (Figure 2).

**Stage 1 (first hours):** acute attack provokes for hours an astrocyte dysfunction (by NMO-IgG binding on AQP4 leading to internalization) mainly expressed by an edema. This purely edematous lesion could be immediately reversible by the clearance of NMO-IgG preventing the loss of astrocytes and the excitotoxic cascade. Clinical and radiological recovery after PLEX is dramatic and explains the Lazarus effect. **Stage 2 (days):** the loss of EAAT2 induces an excitotoxic effect of glutamate on oligodendrocytes leading progressively to demyelination and axonal loss. Astrocytes loss initiates a self-sustained excitotoxic process henceforth independent from NMO-IgG persistence. Even if the extraction of NMO-IgG and complement by PLEX ends the astrocytes aggression. A variable amount of them has been already lost and excitotoxic effects upon oligodendrocytes are evident. Variable amount of tissue is lost as visible on MRI and recovery is incomplete. **Stage 3 (weeks):** astrocytes, oligodendrocytes and axonal loss is prominent, engulfed in large areas of necrosis. PLEX is almost useless. Neural tissue remains cavitated or atrophic on MRI and no recovery will be expected.

We propose to reconsider PLEX as a major part of the treatment of severe NMO attacks and suggest that PLEX could be given systematically in severe relapses of NMO, extended transverse myelitis or bilateral severe ON resistant to steroids. Moreover, when given they should be started as soon as possible with steroids.
6. PLEX as preventive treatment

Since NMO-IgG positivity is both predictive of attacks and severity, achieving a low concentration of plasmatic antibodies remains a goal to achieve. Besides immunosuppressive drugs, PLEX have been used to achieve a sustained depletion of NMO-IgG and complement. Favourable cases have been reported but studies are lacking (Miyamoto et al., 2009). Miyamoto et al. (2009) proposed to use PLEX as preventive treatment as an add-on therapy after immunosuppressive drugs failure.

7. Preventive treatments and future avenues

The natural history of NMO leads all the patients to a deep impairment in a stepwise fashion without progressive phase. In our study, 5 years after the onset, 70% of patients suffered from a unilateral loss of vision and almost half of them from a bilateral loss of vision (Merle et al., 2007). After 8 years, half of the patients had suffered from a severe myelitis and become chair-bound (Cabrera-Gomez et al., 2009). The mortality rate was very high before immunosuppressive drugs but dropped since they are largely used (Cabre et al., 2009). The exact role of recurring PLEX along the remaining attacks to tune the outcome to a low impairment has not yet been addressed but remains most probable considering this striking epidemiological change of mortality in French West Indies.
7.1 Immunosuppressive maintenance

Contrary to MS, cumulative evidence accumulates that interferon beta had no effect on activity rate and worsens some patients, especially in lupus-associated NMO (Uzawa et al., 2010). When IFN is compared to immunosuppressive drugs, a dramatic reduction of annualized relapse rate (ARR) is obtained (Papeix et al., 2007). Rituximab and mycophenolate mofetil were well tolerated and dramatically reduced ARR (Jacob et al., 2008; Jacob et al., 2009). A favorable action of mitoxantrone was reported in few cases (Weinstock-Guttman et al., 2006). In a group of 32 patients treated with mitoxantrone in our centre, a dramatic drop in ARR was obtained from a mean rate of 1.8 to 0.3 and sustained over 5 years (Cabre, personal communication). The choice in one these three drugs should mostly be driven by safety concerns since no comparative study or recommendation is readily available. Low dose steroids were reported to be effective in a few patients, however these data needs further studies with more patients. Various others treatments (cyclophosphamide, azathioprine, venous immuno-globulins) have been used in isolated cases where no general conclusion could be drawn upon ARR action. However cyclophosphamide is commonly used to treat lupus in overlapping cases of NMO (Polgar et al., 2011).

7.2 New strategies for the future

Since the lesion severity mostly depends on the initial and definitive depth of the loss of AQP4 and astrocytes, future treatments strategies may be directed upon AQP4 preservation. Small molecules or monoclonal antibodies could be used to prevent NMO-IgG binding to AQP4 and to block the physiopathological cascade upstream (Verkman et al., 2011; Yu et al., 2011). Another strategy may deplete pathogenic antibodies by apheresis using dedicated immunoadsorption systems as previously described in myasthenia gravis (Zisimopoulou et al., 2008) and in various extra neurological disorders. However the value of this technique is less clear in disorders like MS (De Andres et al., 2000; Moldenhauer et al., 2005) where pathology is broader than a specific antibody. No experience is yet available in the NMO setting. Lymphocytapheresis was successfully described in isolated cases of resistant attacks (Aguilera et al., 1984, Nozaki et al., 2006). A complementary approach may target the complement system with newly developed anti-complement recombinant antibodies at various levels, with preliminary promising results (Saadoun et al., 2010). Such future treatments may be aimed at preventing or curing the attacks. During attacks, neuroprotective treatments could be used to prevent the oligodendrocytes loss induced by the excitotoxic action of glutamate.

Animal models gave clues to dynamic mechanisms evolving over time and appear suitable to address the effect of those various early therapeutic interventions directed to halt or prevent ongoing lesions (Saadoun et al., 2010).

8. Conclusion

PLEX, in synergy with steroids, could be a major treatment of relapses, aimed at preventing cumulative disability. PLEX is a safe and efficient add-on therapy in NMO. Since PLEX proved to be effective regardless of NMO-IgG status, NMO-IgG status should not be required to initiate PLEX. These preliminary results suggest that PLEX may modify the
Plasma Exchange in Severe Attacks Associated with Neuromyelitis Optica Spectrum Disorder

short prognostic of NMO relapses. Immunosuppressive drugs are necessary to prevent further relapses but no recommendation is yet available.

Animal models have confirmed that mechanisms leading to lesion evolve over hours and days. Those models should be able to confirm that early therapeutic intervention directed to halt the ongoing lesions should be even more dramatic in an early narrow therapeutic window.

The next steps should be to concentrate upon large multicentric therapeutic trials in order to validate the therapeutic procedure. However we are aware that good trials against placebo could be difficult to accept since this is an extremely devastating disease. The take-away messages are: undertaking PLEX in severe relapses and the importance of starting treatment as soon as possible.

9. References

Aguilera AJ, Carlow TJ, Smith KJ & Simon TL. (1985). Lympho-cytaplasmapheresis in Devic's syndrome. Transfusion, Vol. 25, No. 1, pp. 54-6.

Amiry-Moghaddam M, Otsuka T, Hurn PD, Traystman RJ, Haug FM, Froehner SC, & al. (2003). An alpha-syntrophin-dependent pool of AQP4 in astroglial end-feet confers bidirectional water flow between blood and brain. Proc Natl Acad Sci USA, Vol.100, pp. 2106-11.

Assogba U, Baumelou A, Pecquinot MA, Raymond F, Durande JP, Lenoir G et al. (1988) Removal of prednisone and prednisolone during plasma exchange. Ann Med Interne, Vol. 139, pp. 38-9.

Bonnan M, Valentino R, Olindo S, Mehdaoui H, Smadja D & Cabre P. (2009). Plasma exchange in severe spinal attacks associated with neuromyelitis optica Spectrum disorder. Mult Scler, Vol.15, No. 4, pp. 487-92.

Bonnan M, Brasme H, Diaby MM, Vlaicu M, Le Guern V & Zuber M. (2009). Severe bouts of neuromyelitis optica: dramatic improvement after plasma exchanges. Rev Neuro, Vol.165, No. 5, pp. 479-81.

Bradl M, Misu T, Takahashi T, Watanabe M, Mader S, Reindl M, & al. (2009). Neuromyelitis optica: pathogenicity of patient immunoglobulin in vivo. Ann Neurol, Vol.66, No.5, pp. 630-43.

Brecher ME. Plasma exchange: why we do what we do. (2002). J Clin Apher, Vol.17, No.4, pp. 207-11.

Brunot S, Vukusic S, Fromont A, Couvreur G, Mousson C, Giroud M, Confavreux C & Moreau T. (2011). Plasma exchanges in severe and acute inflammatory demyelinating attacks of the central nervous system. Presse Med, Vol.40, No.5, pp. e271-8.

Cabre P, Gonzalez-Quevedo A, Lannuzel A, Bonnan M, Merle H, Olindo S, & al. (2009). Descriptive epidemiology of neuromyelitis optica in the Caribbean basin. Rev Neurol, Vol.165, pp. 676-83.

Cabrera-Gomez JA, Bonnan M, Gonzalez-Quevedo A, Saiz Hinajeros A, Marignier R, Graus F, & al. (2009). Neuromyelitis optica positive antibodies confer a worse course in...
relapsing-neuromyelitis optica in Cuba and French West Indies. *Mult Scler*, Vol.15, pp. 828-33.

de Andrès C, Anaya F & Gimenez-Rolden S. (2000). Plasma immunoadsorption treatment of malignant multiple sclerosis with severe and prolonged relapses. *Rev Neurol*. 2000, Vol.30, No.7, pp. 601-5.

Eichel R, Meiner Z, Abramsky O & Gotkine M. (2008). Acute disseminating encephalomyelitis in neuromyelitis optica: closing the floodgates. *Arch Neurol*, Vol.65, No.2, pp. 267-71.

Hinson SR, Pittock SJ, Lucchinetti CF, Roemer SF, Fryer JP, Kryzer TJ & Lennon VA. (2007). Pathogenic potential of IgG binding to water channel extracellular domain in neuromyelitis optica. *Neurology*, Vol.69, No.24, pp. 2221-31.

Hinson SR, Roemer SF, Lucchinetti CF, Fryer JP, Kryzer TJ, Chamberlain JL, Howe CL, Pittock SJ & Lennon VA. (2008). Aquaporin-4-binding autoantibodies in patients with neuromyelitis optica impair glutamate transport by down-regulating EAAT2. *J Exp Med*, Vol.205, No.11, pp. 2473-81.

Hinson SR, McKeon A, Fryer JP, Apiwattanakul M, Lennon VA & Pittock SJ. (2009). Prediction of neuromyelitis optica attack severity by quantitation of complement-mediated injury to aquaporin-4-expressing cells. *Arch Neurol*, Vol.66, pp. 1164-7.

Jacob A, Mattiello M, Weinshenker BG, Wingerchuk DM, Lucchinetti C, Shuster E, Carter J, Keegan BM, Kantarci OH & Pittock SJ. (2009). Treatment of neuromyelitis optica with mycophenolate mofetil: retrospective analysis of 24 patients. *Arch Neurol*, Vol.66, No.9, pp. 1128-33.

Jacob A, Weinshenker BG, Violich I, McLinskey N, Krupp L, Fox RJ, Wingerchuk DM, Boggild M, Constantinescu CS, Miller A, Dean Angela T, Mattiello M & Cree BA. (2008). Treatment of neuromyelitis optica with rituximab: retrospective analysis of 25 patients. *Arch Neurol*, Vol.65, No.11, pp. 1443-8.

Jarius S, Aboul-Enein F, Waters P, Kuenz B, Hauser A, Berger T, Lang W, Reindl M, Vincent A & Kristoferitsch W. (2008). Antibody to aquaporin-4 in the long-term course of neuromyelitis optica. *Brain*, Vol.131, No.11, pp. 3072-80.

Jarius S, Franciotta D, Bergamaschi R, Wildemann B & Wandinger KP. (2010). Immunoglobulin M antibodies to aquaporin-4 in neuromyelitis optica and related disorders. *Clin Chem Lab Med*, Vol.48, No.5, pp. 659-63.

Jarius S, Frederikson J, Waters P, Paul F, Akman-Demir G, Marignier R, Franciotta D, Ruprecht K, Kuenz B, Rommer P, Kristoferitsch W, Wildemann B & Vincent A. (2010). Frequency and prognostic impact of antibodies to aquaporin-4 in patients with optic neuritis. *J Neurol Sci*, Vol.298, No. 1-2, pp. 158-62.

Kale N, Pittock SJ, Lennon VA, Thomsen K, Roemer S, McKeon A & Lucchinetti CF. (2009). Humoral pattern II multiple sclerosis pathology not associated with neuromyelitis Optica IgG. *Arch Neurol*, Vol.66, No.10, pp. 1298-9.

Kalluri SR, Illes Z, Srivastava R, Cree B, Menge T, Bennett JL, Berthele & Hemmer B. (2010). Quantification and functional characterization of antibodies to native aquaporin 4 in neuromyelitis optica. *Arch Neurol*, Vol.67, No.10, pp. 1201-8.
Keegan M, Pineda AA, McClelland RL, Darby CH, Rodriguez M & Weinshenker BG. (2002). Plasma exchange for severe attacks of CNS demyelination: predictors of response. *Neurology*, Vol.58, No.1, pp. 143-6.

Keegan M, Konig F, McClelland R, Bruck W, Morales Y, Bitsch A, Panitch H, Lassmann H, Weinshenker B, Rodriguez M, Parisi J & Lucchinetti CF. (2005). Relation between humoral pathological changes in multiple sclerosis and response to therapeutic plasma exchange. *Lancet*, Vol.366, No.9485, pp. 579-82.

Kinoshita M, Nakatsuji Y, Kimura T, Moriya M, Takata K, Okuno T, Kumanogoh A, Kajiyama K, Yoshikawa H & Sakoda S. (2009). Neuromyelitis optica: Passive transfer to rats by human immunoglobulin. *Biochem Biophys Res Commun*, Vol.386, No.4, pp. 623-7.

Lennon VA, Kryzer TJ, Pittoc S, Verkman AS & Hinson SR. (2005). IgG marker of optic-spinal multiple sclerosis binds to the aquaporin-4 water channel. *J Exp Med*, Vol.202, No.4, pp. 473-7.

Llufriu S, Castillo J, Blanco Y, Ramio-Torrenta L, Rio J, Valles M, Lozano M & al. (2009). Plasma exchange for acute attacks of CNS demyelination: Predictors of improvement at 6 months. *Neurology*, Vol.73, pp. 949-53.

Lucchinetti CF, Mandler RN, McGavern D, Bruck W, Gleich G, Rabesohoff RM, Trebst C, Weinshenker B, Wingerchuk D, Parisi JE & Lassmann H. (2002). A role for humoral mechanisms in the pathogenesis of Devic’s neuromyelitis optica. *Brain*, Vol.125, No.7, pp. 1450-61.

Lucchinetti C, Brück W, Parisi J, Scheithauer B, Rodriguez M & Lassmann H. (2000). Heterogeneity of multiple sclerosis lesions: implications for the pathogenesis of demyelination. *Ann Neurol*, Vol.47, No.6, pp. 707-17.

Magana SM, Matiello M, Pittoc S, McKeon A, Lennon VA, Rabinstein AA, Shuster E, Kantarci OH, Lucchinetti CF & Weinshenker BG. (2009). Posterior reversible encephalopathy syndrome in neuromyelitis optica spectrum disorders. *Neurology*, Vol.72, No.8, pp. 712-7.

Magana SM, Keegan BM, Weinshenker BG, Erickson BJ, Pittoc S, Lennon VA & al. (2011). Beneficial plasma exchange response in central nervous system inflammatory demyelination. *Arch Neurol*, Vol.68, pp. 870-8.

Marignier R, Nicolle A, Watrin C, Touret M, Cavagna S, Varrin-Doyer M, Cavillon G, Rogemond V, Confavreux C, Honnorat J & Giraudon P. (2010). Oligodendrocytes are damaged by neuromyelitis optica immunoglobulin G via astrocyte injury. *Brain*, Vol.133, No.9, pp. 2578-91.

Merle H, Olindo S, Bonnan M, Donnio A, Richer R, Smadja D & Cabre P. (2007). Natural history of the visual impairment of relapsing neuromyelitis optica. *Ophthalmology*, Vol.114, No.4, pp. 810-7.

Miyamoto, K. & Kusunoki, S. (2009). Intermittent plasmapheresis prevents recurrence in neuromyelitis optica. *Ther Apher Dial*, Vol.13, pp. 505-8.

Moldenhauer A, Haas J, Wascher C, Derfuss T, Hoffmann KT, Kiesewetter H & Salama A. (2005). Immunoadsorption patients with multiple sclerosis: an open-label pilot study. *Eur J Clin Invest*, Vol.35, No.8, pp. 523-30.
Munemoto M, Otaki Y, Kasama S, Nanami M, Tokuyama M, Yahiro M, Hasuike Y, Kuragano T, Yoshikawa H, Nonoguchi H & Nakanishi T. (2011). Therapeutic efficacy of double filtration plasmapheresis in patients with anti-aquaporin-4 antibody-positive multiple sclerosis. J Clin Neurosci, Vol.18, No.4, pp. 478-80.

Nakamura M, Nakazawa T, Doi H, Hariya T, Omodaka K, Misu T, Takahashi T, Fujihara K & Nishida K. (2010). Early high-dose intravenous methylprednisolone is effective in preserving retinal nerve fiber layer thickness in patients with neuromyelitis optica. Graefes Arch Clin Exp Ophthalmol, Vol.248, No.12, pp. 1777-85.

Nozaki I, Hamaguchi T, Komai K & Yamada M. (2006). Fulminant Devic disease successfully treated by lymphocytapheresis. J Neurol Neurosurg Psychiatry, Vol.77, No.9, pp. 1094-5.

Papeix C, Vidal JS, de Seze J, Pierrot-Deseilligny C, Tourbah A, Stankoff B, Lebrun C, Moreau T, Vermersch P, Fontaine B, Lyon-Caen O & Gout O. (2007). Immunosuppressive therapy is more effective than interferon in neuromyelitis optica. Mult Scler, Vol.13, No.2, pp. 256-9.

Pittock SJ, Weinshenker BG, Lucchinetti CF, Wingerchuk DM, Corboy JR & Lennon VA. (2006). Neuromyelitis optica brain lesions localized at sites of high aquaporin 4 expression. Arch Neurol, Vol.63, No.7, pp. 964-8.

Pittock SJ, Lennon VA, Krecke K, Wingerchuk DM, Lucchinetti CF & Weinshenker BG. (2006). Brain abnormalities in neuromyelitis optica. Arch Neurol, Vol.63, No.3, pp. 390-6.

Pohl M, Fischer MT, Mader S, Schanda K, Kitic M, Sharma R, Wimmer I, Misu T, Fujihara K, Reindl M, Lassmann H & Bradl M. (2011). Pathogenic T cell responses against aquaporin 4. Acta Neuropathol, Vol.122, No.1, pp. 21-34.

Polgar A, Rosa C, Muller V, Matolcsy J, Poor G & Kiss EV. (2011). Devic’s syndrome and SLE: challenges in diagnosis and therapeutic possibilities based on two overlapping cases. Autoimmun Rev, Vol.10, No.3, pp. 171-4.

Ruprecht K, Klinker E, Dintelmann T, Rieckmann P & Gold R. (2004). Plasma exchange for severe optic neuritis: treatment of 10 patients. Neurology, Vol.63, No.6, pp. 1081-3.

Saadoun S., Waters P., Bell BA., Vincent A., Verkman AS. & Papadopoulos MC. (2010). Intra-cerebral injection of neuromyelitis optica immunoglobulin G and human complement produces neuromyelitis optica lesions in mice. Brain, Vol.133, pp. 349–61.

Saadoun S, Waters P, Macdonald C, Bridges LR, Bell BA, Vincent A & al. (2011). T cell deficiency does not reduce lesions in mice produced by intracerebral injection of NMO-IgG and complement. J Neuroimmunol, Vol.235, pp. 27-32.

Schilling S, Linker RA, Konig FB, Koziolok M, Bahr M, Muller GA & al. (2006). Plasma exchange therapy for steroid-unresponsive multiple sclerosis relapses: clinical experience with 16 patients. Nervenarzt, Vol.77, pp. 430-8.

Sellner J, Boggild M, Clanet M, Hintzen RQ, Illes Z, Montalban X, Du Pasquier RA, Polman CH, Sorensen PS & Hemmer B. (2010). EFNS guidelines on diagnosis and management of neuromyelitis optica. Eur J Neurol, Vol.17, No.8, pp. 1019-32.

Stigelman WH Jr, Henry DH, Talbert RL, (1984) Townsend RJ. Removal of prednisone and prednisolone by plasma exchange. Clin Pharm, Vol.3, pp. 402-7.
Takahashi T, Fujihara K, Nakashima I, Misu T, Miyazawa I, Nakamura M, Watanabe S, Shiga Y, Kanaoka C, Fujimori J, Sato S & Itoyama Y. (2007). Anti-aquaporin-4 antibody is involved in the pathogenesis of NMO: a study on antibody titre. *Brain*, Vol.130, No.5, pp. 1235-43.

Trebst C, Reising A, Kielstein JT, Hafer C & Stangel M. (2009). Plasma exchange therapy in steroid-unresponsive relapses in patients with multiple sclerosis. *Blood Purif*, Vol.28, No.2, pp. 108-15.

Uzawa A, Mori M, Hayakawa S, Masuda S & Kuwabara S. (2010). Different responses to interferon beta-1b treatment in patients with neuromyelitis optica and multiple sclerosis. *Eur J Neurol*, Vol.17, No.5, pp. 672-6.

Verkman AS, Ratelade J, Rossi A, Zhang H & Tradtrantip L. (2011). Aquaporin-4: orthogonal array assembly, CNS functions, and role in neuromyelitis optica. *Acta Pharmacol Sin*, Vol.32, pp. 702-10.

Viegas S, Weir A, Esiri M, Kuker W, Waters P, Leite MI & al. (2009). Symptomatic, radiological and pathological involvement of the hypothalamus in neuromyelitis optica. *J Neurol Neurosurg Psychiatry*, Vol.80, pp. 679-82.

Watanabe S, Nakashima I, Miyazawa I, Misu T, Shiga Y, Nakagawa Y, Fujihara K & Itoyama Y. (2007). Successful treatment of a hypothalamic lesion in neuromyelitis optica by plasma exchange. *J Neurol*, Vol.254, No.5, pp. 670-1.

Watanabe S, Nakashima I, Miyazawa I, Shiga Y, Fujihara K & Itoyama Y. (2007). Therapeutic efficacy of plasma exchange in NMO-IgG-positive patients with neuromyelitis optica. *Mult Scler*, Vol.13, No.1, pp. 128-32.

Weinshenker BG, O'Brien PC, Pettersson TM, Noseworthy JH, Lucchinetti CF, Dodick DW, Pineda AA, Stevens LN & Rodriguez M. (1999). A randomized trial of plasma exchange in acute central nervous system inflammatory demyelinating disease. *Ann Neurol*, Vol.46, No.6, pp. 878-86.

Weinshenker BG. (2001). Plasma exchange for severe attacks of inflammatory demyelinating diseases of the central nervous system. *J Clin Apher*, Vol.16, No.1, pp. 39-42.

Weinshenker BG, Wingerchuk DM, Vukusic S, Linbo L, Pittcock SJ, Luccinetti CF & Lennon VA. (2006). Neuromyelitis optica IgG predicts relapse after longitudinally extensive transverse myelitis. *Ann Neurol*, Vol.59, No.3, pp. 566-9.

Weinshenker BG. (2000). Plasma exchange for acute attacks of demyelinating disease: detecting a Lazarus effect. *Ther Apher*, Vol.4, No.3, pp. 187-9.

Weinstock-Guttman B, Ramathan M, Lincoff N, Napoli SQ, Sharma J, Feichter J & Bakshi R. (2006). Study of mitoxantrone for the treatment of recurrent neuromyelitis optica (Devic disease). *Arch Neurol*, Vol.63, No.7, pp. 957-63.

Weinstock-Guttman B, Miller C, Yeh E, Stosic M, Umbauer M, Batra N, Munschauer F, Zivadinov R & Ramathan M. (2008). Neuromyelitis optica immunoglobulins as a marker of disease activity and response to therapy in patients with neuromyelitis optica. *Mult Scler*, Vol.14, No.8, pp. 1061-7.

Wingerchuk DM, Lennon VA, Luccinetti CF, Pittcock SJ & Weinshenker BG. (2007). The spectrum of neuromyelitis optica. *Lancet Neurol*, Vol.6, No.9, pp. 805-15.

Wingerchuk DM, Lennon VA, Pittcock SJ, Luccinetti CF & Weinshenker BG. (2006). Revised diagnostic criteria for neuromyelitis optica. *Neurology*, Vol.66, No.10, pp. 1485-9.
Yoshida H, Ando A, Sho K, Akioka M, Kawai E, Arai E, Nishimura T, Shinde A, Masaki H, Takahashi K, Takagi M & Tanaka K. (2010). Anti-aquaporin-4 antibody-positive optic neuritis treated with double-filtration plasmapheresis. J Ocul Pharmacol Ther, Vol.26, No.4, pp. 381-5.

Yu X, Green M, Gilden D, Lam C, Bautista K & Bennett JL. (2011). Identification of peptide targets in neuromyelitis optica. J Neuroimmunol, Vol.236, No.1-2, pp. 65-71.

Zisimopoulou P., Lagoumintzis G., Kostelidou K., Bitzopoulou K., Kordas G., Trakas N. & al. (2008) Towards antigen-specific apheresis of pathogenic autoantibodies as a further step in the treatment of myasthenia gravis by plasmapheresis. J Neuroimmunol, Vol. 201-2, pp. 95–103.
Immunology is the branch of biomedical sciences to study the immune system physiology both in healthy and diseased states. Some aspects of autoimmunity draw our attention to the fact that it is not always associated with pathology. For instance, autoimmune reactions are highly useful in clearing off the excess, unwanted or aged tissues from the body. Also, generation of autoimmunity occurs after the exposure to the non-self antigen that is structurally similar to the self, aided by the stimulatory molecules like the cytokines. Thus, a narrow margin differentiates immunity from auto-immunity as already discussed. Hence, finding answers for how the physiologic immunity turns to pathologic autoimmunity always remains a question of intense interest. However, this margin could be cut down only if the physiology of the immune system is better understood. The individual chapters included in this book will cover all the possible aspects of immunology and pathologies associated with it. The authors have taken strenuous effort in elaborating the concepts that are lucid and will be of reader's interest.

How to reference
In order to correctly reference this scholarly work, feel free to copy and paste the following:

Bonnan Mickael and Cabre Philippe (2012). Plasma Exchange in Severe Attacks Associated with Neuromyelitis Optica Spectrum Disorder, Recent Advances in Immunology to Target Cancer, Inflammation and Infections, Dr. Jagat Kanwar (Ed.), ISBN: 978-953-51-0592-3, InTech, Available from: http://www.intechopen.com/books/recent-advances-in-immunology-to-target-cancer-inflammation-and-infections/plasma-exchange-in-severe-attacks-associated-with-neuromyelitis-optica-spectrum-disorder
