Structure Prediction and Binding Site Analysis of Hepatotoxic Microcystin-LR Degrading MlrC-Like Protein from *Burkholderia* sp. using Computational Approaches

Dhananjaya P. Singh, Ratna Prabha, Vivek Keshri and P.C. Abhilash

ICAR-National Bureau of Agriculturally Important Microorganisms, Indian Council of Agricultural Research, Kushmaur, Maunath Bhanjan-275103, UP, India

Institute of Environmental and Sustainable Development, Banaras Hindu University, Varanasi- 221005, UP, India

**Abstract:** Microcystin-LR (MCYST-LR) is a well characterized hepatotoxic heptapeptide produced by various species of cyanobacteria including *Microcystis aeruginosa*. *Burkholderia* is a genus of bacteria with cyanobacterial toxins degrading property. This study predicts the structure of microcystin degrading MlrC-like protein from *Burkholderia* sp. strain CCGE1002 that has microcystin degradation capability. Binding interaction of MlrC-like protein with MCYST-LR was studied. Three-dimensional model of MlrC-like protein was generated using composite modeling based I-TASSER server. The model was further assessed through different computational approaches. The generated model was found comparable to experimental structures. MCYST-LR was used for docking with predicted model to investigate ligand-protein interaction. The study provides the structural insight into the binding mode of MlrC-like protein of *Burkholderia* sp. with MCYST-LR and could be further helpful in designing modeling inhibitors for MCYST-LR.

**Keywords:** Microcystin, MCYST-LR, Structure Prediction, Docking, *Burkholderia* sp.

**Introduction**

Cyanobacteria are the most prominent photosynthetic prokaryotes in freshwater (lakes, ponds, rivers and estuaries) and marine ecosystem. Under favourable environmental conditions, many species of cyanobacteria produce blooms (Briand *et al.*, 2003) that are capable of producing highly toxic secondary metabolites (Pearson *et al.*, 2010). Microcystins (MCYSTs) are structurally diverse collection of heptapeptide hepatotoxins which are specifically secreted by different cyanobacterial species e.g. *Microcystis*, *Nostoc*, *Anabaena* and *Planktothrix* (Carmichael, 1994). Almost 90 variants of these cyclic hepatotoxins possess specifically β-amino acid residue, 3-amino-9-methoxy-2,6,8-trimethyl-10-phenyl-deca-4,6-dienoic acid (Adda) and other amino acids at diverse functional positions which differ in view of different activities like methylation, epimerization, hydroxylation, peptide sequence and toxicity (Sivonen and Jones, 1999; Zurawell *et al.*, 2005). Synthesis and secretion of microcystins by different cyanobacterial species into the water mainly causes toxicity of water bodies and due to this, the problem has gained worldwide attention (Carmichael *et al.*, 1985). Many variants of MCYSTs including the most potent MCYST-LR (leucine-arginine) are prominent hepatotoxins and chronic usage even in little quantity in drinking water leads to tumor growth in the human liver (Carmichael, 1994; Falconer, 1991). Owing to these facts, these toxins are grouped as “probable carcinogenic to humans (group 2B)” via the International Agency for Research on Cancer (Grosse, 2006). Microcystins possess stable chemical structure in water and thus different water treatment approaches including filtration, coagulation and flocculation are found inefficient in decreasing the concentration of these toxins in the fresh water reservoirs (Manage *et al.*, 2009). Thus, the risk associated with their toxicity remains very high. Presence of toxic cyanobacterial blooms in numerous natural freshwater bodies and possible use of such water in human consumption initiated...
work on biodegradation of cyanotoxins (Lam et al., 1995). Diversity of microbial species with ability of microcystin degradation and nodularin with tentative recognizance of novel degradation intermediates were reported (Manage et al., 2009). It was shown that MCYST can undergo biodegradation by certain aquatic bacteria like strains of the genus Sphingomonas (Saitou et al., 2003; Valeria et al., 2006) owing to the existence of an enzymatic degradation mechanism. Certain other bacteria like Pseudobacter toxinivorans, Sphingosinicella microcystinivorans in Japan and Burkholderia (Lemes et al., 2008) were also reported as biodegraders of these cyanotoxins. The biodegradation mechanism is reported to involve a gene cluster mlrA, mlrB, mlrC and mlrD (Bourne et al., 2001; Saitou et al., 2003; Imanishi et al., 2005; Manage et al., 2009) via which particular variants, e.g., [D-Leu]MC-LR are recognized as biodegraded and/or biotransformed through aquatic microbes (Matthiensen et al., 2000). *Burkholderia* sp. (strain CGE1002) is an agriculturally important bacteria contributing to nitrogen fixation (Ormeño-Orrillo et al., 2012). This bacterium possesses MCYST degrading MlrC-like protein. Several 3D structure modeling based studies has been conducted to yield suitable and wide spectrum application for the investigation of mechanisms of degradation (Suresh et al., 2008), pollutant interactions (Srivastava et al., 2011; Librando and Pappalardo, 2012) and biotransformation of MCYST-LR (Jones et al., 1994; Lam et al., 1995). However, the three-dimensional configuration of MlrC-like protein or its interaction with MCYST-LR has not been worked out, although it could lead to valuable information related to the molecular structure, role and efficient binding with the protein. Theoretical approaches have been applied for structure prediction (Suresh et al., 2015). We report 3D structure of MlrC-like protein implementing a combined in silico approach like threading and ab-initio for identification via composite modeling. The interaction and identification of possible binding sites of MlrC-like protein of Burkholderia sp. with MCYST-LR was prediction through the docking perspective and probable interactions with binding sites were also recognized.

**Materials and Methods**

Sequence retrieval and 3D structure prediction

Amino acid sequence of MCYST degrading MlrC-like protein (GenBank Accession no: ADG18989) of *Burkholderia* sp. strain CCGE1002 was taken from NCBI (http://www.ncbi.nlm.nih.gov). Primary properties of this protein sequence i.e., total number of composed amino acid residue, molecular weight, theoretical pl, entire amino acid content, number of residues with negative and positive charges and atomic composition were computed using Expasy’s ProtParam server (Wilkins et al., 1999).

An adequate template was required for comparative modeling of MlrC-like protein. For this purpose, protein sequence of MlrC-like protein was aligned against Protein Data Bank (PDB) (Berman et al., 2000) through BLAST (Altschul et al., 1990). Default parameters were used during alignment (algorithm: Blastp; expected threshold: 10; matrix: BLOSUM62; word size: 3; filter low complexity regions). No single template was identified to provide full length query coverage.

For 3D structure prediction of full length protein, I-TASSER server (Zhang, 2008) was used. Score for template modeling i.e., TM-score was numerated via following formula for assessment of the identity between target and template protein structures in terms of topology (Zhang, 2008):

\[
\text{TM Score} = \frac{1}{L} \sum_{i=1}^{L} \frac{1}{d_i^3 / d_o^3 - 1}
\]

Here, \(L\) represents length of the protein under investigation; \(d_i\) is distance of the \(i\)th pair of the residues among two structures; the scale \(d_o = \sqrt{L - 15 - 1.8}\) is used for the normalization of the TM-score in a manner so that the extent of the average TM-score for particular protein pairs is not influenced by the protein size.

**Model Validation**

Estimation of model quality is a crucial step in computational protein modeling as the accuracy of a model is responsible for its appropriateness for diverse biological and biochemical experimental purposes (Srivastava et al., 2012). Confidence score was calculated for the structure predicted from I-TASSER server for quality estimation (Zhang, 2008). Confidence score is calculated on the basis of threading template alignments significance and convergence parameters employed in the structure assembly refinement simulations of I-TASSER (Zhang, 2008). In addition to this, modeled structure was analyzed for energy minimization followed by complete assessment of model quality with Qualitative Model Energy Analysis (QMEAN) server (Benkert et al., 2009). The quality of model was checked with the help of Errat (Bowie et al., 1991), VERIFY3D (Eisenberg et al., 1997) and Ramchandran plot (constructed using RAMPAGE server (Lovell et al., 2002)).

**Docking Studies**

Docking studies were carried out through AutoDock4.2 with AutoDockTools (ADT) to explore the residues of the MlrC-like protein involved in interaction with MCYST-LR toxin. As the binding site position of MCYST-LR was not known, thus blind docking
Physicochemical Properties

Physico-chemical properties of target protein i.e., MlrC-like protein of *Burkholderia* sp. strain CCGE1002 as computed using Exspasy’s ProtParam server (Wilkins et al., 1999) showed the molecular weight 54390.1, number of amino acids 511 and theoretical pI 5.86 (Table 1).

Structure Prediction and Validation

BlastP search against PDB disclosed the unavailability of template with >50% sequence identity. As the modeling confidence deteriorates below 50% sequence identity and below 25% identity, confidence in accuracy is poor (Kopp and Schwede, 2004) therefore, we used I-TASSER server which implemented composite modeling approach for structure determination of MlrC-like protein of *Burkholderia* Target protein structure was modeled by means of restraints from PDB template 3iuuA. The whole structure prediction process included identification of template, structure reassembly, atomic model production and selecting the best model (Zhang, 2009). The PDB template 3iuuA composed of two domains (i) domain of unknown function (DUF1485) and (ii) MlrC C-terminus domain. Domain search revealed the presence of both of these domains in MlrC-like protein of *Burkholderia* (Table 2).

The calculation of a structural alignment between two protein structures is crucial step in protein modeling. Rather than sequence alignment methods, structure alignment approaches focus specifically on improving the structural resemblance of input proteins (Gupta et al., 2012). C-alpha atom root-mean-square deviation (RMSD) was is calculated via the superposition among the model of MlrC-like protein and crystal structure of threading template putative metallopeptidase from *Mesorhizobium* (3iuuA) was 0.49 Å (Fig. 1).

The PDB models which were most close (in terms of structure) to the modeled protein along with TM-score for their structural alignment are listed in Table 3. TM-score >0.5 represents a model with accurate topology while TM-score <0.17 signalizes random similarity. TM-score of our MlrC-like protein model was 0.93±0.06 which reflects enhanced structural similarity of the target sequence with the templates. TM-score is a sensitive scale for evaluation of the quality of protein structure templates (Zhang and Skolnick, 2004) and the MlrC-like protein fits well with the quality parameters. I-TASSER provided C-score i.e., confidence score for evaluation of the quality of predicted models. C-score lies in a range of -5 to +2, where higher C-score value represents a model with high confidence and vice-versa (Silao et al., 2007) and the models with C-score > -1.5 possess correct fold (Roy et al., 2010). The calculated C-score for predicted 3D structure of MlrC-like protein was 1.53 which favored the model.

The distorted geometries of protein models can be repaired with energy minimization procedure by movement of atoms for releasing internal constraints. Energy minimization was performed using Chimera (Pettersen et al., 2004) with default parameters (steepest descent minimization algorithms, maximum number of minimization steps (i.e., 100), step size 0.02Å and update interval 10). In the minimization procedure, we could not find the improvement in model quality as 10 residues from most favored regions moved in additional allowed regions while there was no any change in residue information of other generously allowed and disallowed regions of Ramachandran plot. Thus, the model prior to minimization was used for further analysis. The Ramachandran plot drawn through the RAMPAGE (Lovell et al., 2002) validated the model with 84.7% residues in favored region, 9.0% residues in allowed area and 6.3% residues in outlier area (Fig. 2). ERRAT is specifically used algorithm for verification of protein structure and assessment of model building and refinement. This algorithm is found on the investigation of statistics of non-bonded interactions among diverse atom types and helps in taking decisions with consistency. After the errat, the overall quality factor was 61.034. Results obtained from VERIFY3D revealed that 92.17% residues possess an average 3D-1D score higher than 0.2 which reflects a finely built model as most of the residues are suitable in their folded conformation.

Moreover, for approximation of the quality of predicted structure comprehensively, we employed QMEAN server (Benkert et al., 2009) that employs a composite scoring function QMEAN Z-score (Benkert et al., 2008; 2009) as an approximate of the total quality of a model by connecting it to experimentally solved reference structures of like sizes (model size +/- 10%) present in PDB (Berman et al., 2000). The calculated QMEAN Z-score for the modeled structure was -0.5 which reflects the model is close to the experimental data.
score of our model was -1.06 that represents reasonable homology model with correct folds.

Docking studies were carried out for the validated model.

Fig. 1. Three dimensional structure superposition of Cα atoms of template 3iuuA (blue) and MlrC like protein model (orange)

Fig. 2. Ramachandran plot of structure of MlrC-like protein showing residues in favoured, allowed and outlier region

Fig. 3. Chemical structure of oligopeptidic toxin MCYST-LR. The two amino acid residue ARG and LEU is shown in magenta and green color respectively
Fig. 4. The hydrogen bond interaction of ligand MCYST-LR (orange) with modeled MlrC like protein residues. The ligand interacts with THR399 and ASP400 residue of the modeled protein with hydrogen bond (green dotted line). The black, blue and red color represents carbon, nitrogen and oxygen atoms.

Table 1. Physico-chemical properties of target protein sequence

| Properties                      | Description                                      |
|---------------------------------|--------------------------------------------------|
| Formula                         | $C_{2406}H_{3816}N_{700}O_{702}S_{19}$           |
| Number of amino acid            | 511                                              |
| Molecular Weight                | 54390.1                                          |
| Theoretical pl                   | 5.86                                             |
| Amino Acid Composition          |                                                   |
| A                               | 84                                               |
| C                               | 3                                                |
| D                               | 32                                               |
| E                               | 28                                               |
| F                               | 17                                               |
| G                               | 48                                               |
| H                               | 15                                               |
| I                               | 23                                               |
| K                               | 10                                               |
| L                               | 43                                               |
| M                               | 16                                               |
| N                               | 10                                               |
| P                               | 29                                               |
| Q                               | 12                                               |
| R                               | 40                                               |
| S                               | 22                                               |
| T                               | 18                                               |
| V                               | 46                                               |
| W                               | 7                                                |
| Y                               | 8                                                |
| No. of negatively charged residue | 60                                           |
| No. of positively charged residue | 50                                           |
| Atomic Composition             |                                                   |
| Atom                            | Number                                           |
| C                               | 2406                                             |
| H                               | 3816                                             |
| N                               | 700                                              |
| O                               | 702                                              |
| S                               | 19                                               |
| Total                           | 7643                                             |
| GRAVY                           | 0.059                                            |
Table 2. Comparison of the domains of template (3iuuA) and target protein (ADG18989.1). Domains were identified with the help of Pfam (Punta et al., 2012)

| Accession number | Domain       | Description                      | Start position | End position |
|------------------|--------------|----------------------------------|----------------|--------------|
| PDB: 3iuuA       | DUF1485      | Protein of unknown function (DUF1485) | 7              | 289          |
| PDB: 3iuuA       | MlrC_C       | MlrC C-terminus                  | 297            | 472          |
| GenBank: ADG18989.1 | DUF1485 | Protein of unknown function (DUF1485) | 1              | 293          |
| GenBank: ADG18989.1 | MlrC_C | MlrC C-terminus                  | 303            | 482          |

Table 3. PDB structures structurally closest to the predicted model along with TM-score of the structural alignment

| Rank | PDB ID | TM-Score |
|------|--------|----------|
| 1    | 3iuuA  | 0.933    |
| 2    | 3k70E  | 0.417    |
| 3    | 3iuuA  | 0.411    |
| 4    | 3effA  | 0.409    |
| 5    | 4e2xA  | 0.396    |
| 6    | 3u44A  | 0.394    |
| 7    | 3bulA  | 0.389    |
| 8    | 2b3yB  | 0.387    |
| 9    | 2y0eA  | 0.387    |
| 10   | 3vH4A  | 0.386    |

Table 4. Various energies of the best conformation of MCYST-LR docked into the modeled MlrC-like protein calculated by Autodock4 software

| Binding energy | Ligand efficiency | Inter-mol energy | Electro-static energy | Torsional energy | Total Internal |
|----------------|-------------------|------------------|-----------------------|------------------|----------------|
| -3.0           | -0.04             | -8.08            | -0.59                 | 5.07             | -3.18          |

Table 5. Receptor-ligand residues involved in hydrogen bond formation

| Residues of receptor (MlrC-like protein) | Residues of ligand (MCYST-LR) |
|-----------------------------------------|-------------------------------|
| A:HIS381:HE2                            | :O9                           |
| A:ARG430:O                              | :H19, :H21                    |
| A:GLU435:OE1                            | :H73, :H74                    |

Docking Studies

The chemical structure of oligopeptidic toxin MCYST-LR (PDB: 1LCM) is shown in Fig. 3. Docking analysis was performed by Autodock4. While molecular docking, sampling and scoring, both are equally important components. All the potential binding modes of particular molecule are sampled in accordance to the other molecule. Conformational adjustments may be contemplated while the sampling procedure (Huang and Zou, 2014). After scoring of docking results, it was identified that MCYST-LR showed greater binding affinity with MlrC C-terminus domain of MlrC-like protein of Burkholderia.

The complex with best docking score is envisaged in Fig. 4 and different energies determined by AutoDock4 for the best pose is listed in Table 4. It was observed that residues Leu353, His381, Leu382, Ser383, His384, Arg386, Arg388, Asn427, Agr430, Ala431, Ala432, Gly433 and Glu435 of MlrC-like protein were in close contact with MCYST-LR (within 4.5 Å) among which three residues HIS381, ARG430 and GLU435 forms the critical hydrogen bonds with MCYST-LR toxin (Fig. 4 and Table 5).

Conclusion

Protein structure prediction approaches are not expected to substitute protein structure determination experimentally but can contribute significantly for overcoming the gap among number of sequences and structures available in respective databases. We predicted 3D structure of MCYST-LR degrading protein of Burkholderia sp. strain CCCE1002. Structure prediction was done by threading based modeling since no close homolog was available for most of the targets. Predicted structure was further analyzed by different computational approaches for stability, efficacy, binding site prediction and docking with MCYST-LR. The protein structure assessment results provided the evidence that the predicted model is stable and accurate. Results suggested that the MlrC-like protein of Burkholderia sp. strain CCCE1002 can be used as significantly potential target for modeling future inhibitors for biological
degradation and/or biotransformation of MCYST-LR from freshwater bodies.

Acknowledgement

Authors are thankful to National Agricultural Innovation Project (NAIP), Indian Council of Agricultural Research, India for providing financial support in the form of National Agricultural Bioinformatics Grid (NABG) project.

Funding Information

This work was supported by the project "Establishment of National Agricultural Bioinformatics Grid (NABG)" under National Agricultural Innovation Project (NAIP) ICAR (India)

Author’s Contributions

Dhananjaya P. Singh: Designed the experiment, write the MS, thoroughly revised and modified the MS.

Ratna Prabha: Performed the work, write the MS.

Vivek Keshri: Performed the work.

PC Abhilash: Thoroughly revised and modified the MS.

Ethics

None.

Conflict of Interest

Authors declared no conflict of interest.

References

Altschul, S.F., W. Gish, W. Miller, E.W. Myers and D.J. Lipman, 1990. Basic local alignment search tool. J. Mol. Biol., 215: 403-410. DOI: 10.1016/S0022-2836(05)80360-2

Benkert, P., M. Kuenzli and T. Schwede, 2009. QMEAN Server for Protein Model Quality Estimation. Nucleic Acids Res., 37: W510-514. DOI: 10.1093/nar/gkp322

Benkert, P., S.C. Tosatto and D. Schomburg, 2008. QMEAN: A comprehensive scoring function for model quality assessment. Proteins, 71: 261-277. DOI: 10.1002/prot.21715

Berman, H.M., J. Westbrook, Z. Feng, G. Gilliland and T.N. Bhat et al., 2000. The protein data bank. Nucleic Acids Res., 28: 235-242. DOI: 10.1093/nar/28.1.235

Bourne, D.G., P. Riddles, G.J. Jones, W. Smith and R.L. Blakeley, 2001. Characterisation of a gene cluster involved in bacterial degradation of the cyanobacterial toxin microcystin LR. Environ. Toxicol., 16: 523-534. DOI: 10.1002/tox.10013

Bowie, J.U., R. Lüthy and D. Eisenberg, 1991. A method to identify protein sequences that fold into a known three-dimensional structure. Science, 253:164-70. DOI: 10.1126/science.1853201

Briand, J.F., S. Jacquet, C. Bernard and J.F. Humbert, 2003. Health hazards for terrestrial vertebrates from toxic cyanobacteria in surface water ecosystems. Vet. Res., 34: 361-377. DOI: 10.1051/vetres:2003019

Carmichael, W.W., 1994. The toxins of cyanobacteria. Sci. Am., 270: 64-72. DOI: 10.1038/scientificamerican0194-78

Carmichael, W.W., C.L.A. Jones, N.A. Mahmood, W.C. Theiss and P. Krogh, 1985. Algal toxins and water-based diseases. Crit. Rev. Environ. Cont., 15: 275-313. DOI: 10.1080/10643388509381734

Eisenberg, D., R. Lüthy and J.U. Bowie, 1997. VERIFY3D: Assessment of protein models with three-dimensional profiles. Meth. Enzymol., 277: 396-404. DOI: 10.1016/S0076-6879(97)77022-8

Falconer, I.R., 1991. Tumor promotion and liver injury caused by oral consumption of cyanobacteria. Environ. Toxicol. Water Qual., 6: 177-184. DOI: 10.1002/tox.253060207

Grosse, Y., R. Baan, K. Straif, B. Secretan and F. El Ghissassi et al., 2006. Carcinogenicity of nitrate, nitrite and cyanobacterial peptide toxins. Lancet. Oncol., 7: 628-629. DOI: 10.1016/S1470-2045(06)70789-6

Gupta, S.K., M. Srivastava, B.A. Akhoon, S.K. Gupta and N. Grabe, 2012. In silico accelerated identification of structurally conserved CD8+ and CD4+ T-cell epitopes in high-risk HPV types. Infect. Genet. Evol., 12: 1513-1518. DOI: 10.1016/j.meegid.2012.02.022

Hetényi, C. and D. van der Spoel, 2002. Efficient docking of peptides to proteins without prior knowledge of the binding site. Protein Sci., 11: 1729-1737. DOI: 10.1110/ps.0202302

Hetényi, C. and D. van der Spoel, 2006. Blind docking of drug-sized compounds to proteins with up to a thousand residues. FEBS Lett., 580: 1447-1450. DOI: 10.1016/j.febslet.2006.01.074

Hetényi, C. and D. van der Spoel, 2011. Toward prediction of functional protein pockets using blind docking and pocket search algorithms. Protein Sci., 20: 880-893. DOI: 10.1002/pro.618

Huang, S. and X. Zou, 2014. A knowledge-based scoring function for protein-RNA interactions derived from a statistical mechanics-based iterative method. Nucleic Acids Res., 42: e55-e55. DOI: 10.1093/nar/gku077

Imanishi, S., H. Kato, M. Mizuno, K. Tsuji and K. Harada, 2005. Bacterial degradation of microcystins and nodularin. Chem. Res. Toxicol., 18: 591-598. DOI: 10.1021/tr049677g
Jones, G.J., D.G. Bourne, R.L. Blakeley and H. Doelle, 1994. Degradation of the cyanobacterial hepatotoxin microcystin by aquatic bacteria. Nat. Toxins., 2: 228-235. DOI: 10.1002/nt.262002012

Kopp, J. and T. Schwede, 2004. Automated protein structure homology modeling: A progress report. Pharmacogenomics, 5: 405-416. DOI: 10.1517/14622416.5.4.405

Lam, A.K., P.M. Fedorak and E.E. Prepas, 1995. Degradation of the cyanobacterial hepatotoxin microcystin-LR, as determined by HPLC and protein phosphatase bioassay. Environ. Sci. Technol., 29: 242-246. DOI: 10.1021/es00001a030

Lemes, G.A., R. Kersanach, S. Lda Pinto, O.A. Dellagostin and J.S. Yunes et al., 2008. Biodegradation of microcystins by aquatic Burkholderia sp. from a South Brazilian coastal lagoon. Ecotoxicol. Environ. Saf., 69: 358-365. DOI: 10.1016/j.ecoenv.2007.03.013

Librando, V. and M. Pappalardo, 2012. Engineered enzyme interactions with poly cyclic aromatic hydrocarbons: A theoretical approach. J. Mol. Graph. Model., 36: 30-35. DOI: 10.1016/j.jmgm.2012.02.005

Lovell, S.C., I.W. Davis, W.B. Arendall III, P.I.W. de Bakker and J.M. Word et al., 2002. Structure validation by Cα geometry: ϕ,ψ and Cβ deviation. Proteins: Structure Funct. Genet., 50: 437-450. DOI: 10.1002/prot.10286

Manage, P.M., C. Edwards, B.K. Singh and L.A. Lawton, 2009. Isolation and identification of novel microcystin-degrading bacteria. Applied Environ. Microbiol., 75: 6924-6928. DOI: 10.1128/AEM.01928-09

Matthiesen, A., J.S. Metcalf, A.H.F. Ferreira, J.S. Yunes and G.A. Codd, 2000. Mycotoxins and phycotoxins in perspective at the turn of the millennium. Proceedings of the 10th International IUPAC Symposium on Mycotoxins and Phycotoxins, (SMP’00), Guaruja, Brazil, pp: 527-536.

Ormeño-Orrillo, E., M.A. Rogel, L.M. Chueire, J.M. Tiedje and E. Martinez-Romero et al., 2012. Genome sequences of Burkholderia sp. strains CCGE1002 and H160, Isolated from legume nodules in Mexico and Brazil. J. Bacteriol., 194: 6927-6927. DOI: 10.1128/ JB.01756-12

Pearson, L., T. Mihali, M. Moffitt, R. Kellmann and B. Neilan, 2010. On the chemistry, toxicology and genetics of the cyanobacterial toxins, microcystin, nodularin, saxitoxin and cylindrospermopsin. Mar. Drugs., 8: 1650-1680. DOI: 10.3390/md8051650

Pettersen, E.F., T.D. Goddard, C.C. Huang, G.S. Couch and D.M. Greenblatt et al., 2004. UCSF Chimera-a visualization system for exploratory research and analysis. J. Comput. Chem., 25: 1605-1612. DOI: 10.1002/jcc.20084

Punta, M., P.C. Coggill, R.Y. Eberhardt, J. Mistry and J. Tate et al., 2012. The Pfam protein families database. Nucleic Acids Res., 40: D290-301. DOI: 10.1093/nar/gkr1065

Roy, A., A. Kucukural and Y. Zhang, 2010. I-TASSER: A unified platform for automated protein structure and function prediction. Nat. Protoc., 5: 725-738. DOI: 10.1038/nprot.2010.5

Saitou, T., N. Sugiuara, T. Itayama, Y. Inamoti and M. Matsumura, 2003. Degradation characteristics of microcystins by isolated bacteria from Lake Kasumigaura. J. Water SRT-Aqua., 52: 13-18. http://aqua.iwaponline.com/content/52/1/13

Sitao, W., J. Skolnick and Y. Zhang, 2007. Ab initio modeling of small proteins by iterative TASSER simulations. BMC Biol., 5: 17-17. DOI: 10.1186/1741-7007-5-17

Sivonen, K. and G. Jones, 1999. Cyanobacterial Toxic. In: Toxic Cyanobacteria in Water: A Guide to Public Health Significance, Monitoring and Management, Chorus, I. and J. Bartram (Eds.), E&FN Spon, London, pp: 41-111.

Srivastava, M., P.C. Abhilash and N. Singh, 2011. Remediation of lindane using engineered nanoparticles. J. Biomed. Nanotechnol., 2011: 172-174. DOI: 10.1166/jbn.2011.1255

Srivastava, M., S.K. Gupta, P.C. Abhilash and N. Singh, 2012. Structure prediction and binding sites analysis of curcin protein of Jatropha curcas using computational approaches. J. Mol. Model., 7: 2971-2979. DOI: 10.1007/s00894-011-1320-0

Suresh, P.S., A. Kumar, R. Kumar and V.P. Singh, 2008. An insilico approach to bioremediation: Laccase as a case study. J. Mol. Graph. Model., 26: 845-849. DOI: 10.1016/j.jmgm.2007.05.005

Suresh, V., L. Liu, D. Adjeroh and X. Zhou, 2015. RPI-Pred: Predicting ncRNA-protein interaction using sequence and structural information. Nucleic Acids Res., 43: 1370-1379. DOI: 10.1093/nar/gkv020

Valeria, A.M., E.J. Ricardo, P. Stephan and W.D. Alberto, 2006. Degradation of microcystin-RR by Burkholderia sp. strains CCGE1002 and H160, Isolated from legume nodules in Mexico and Brazil. J. Bacteriol., 194: 6927-6927. DOI: 10.1128/JB.01756-12

Pearson, L., T. Mihali, M. Moffitt, R. Kellmann and B. Neilan, 2010. On the chemistry, toxicology and genetics of the cyanobacterial toxins, microcystin, nodularin, saxitoxin and cylindrospermopsin. Mar. Drugs., 8: 1650-1680. DOI: 10.3390/md8051650
Zhang, Y., 2009. I-TASSER: Fully automated protein structure prediction in CASP8. Proteins, 77: 100-113. DOI: 10.1002/prot.22588

Zhang, Y., 2008. I-TASSER server for protein 3D structure prediction. BMC Bioinform., 9: 40-40. DOI: 10.1186/1471-2105-9-40

Zhang, Y. and J. Skolnick, 2004. Scoring function for automated assessment of protein structure template quality. Proteins, 57: 702-710. DOI: 10.1002/prot.20264

Zurawell, R.W., H. Chen, J.M. Burke, E.E. Prepas, 2005. Hepatotoxic cyanobacteria: A review of the biological importance of microcystins in freshwater environments. J. Toxicol. Environ. Health, 8: 1-37. DOI: 10.1080/10937400590889412