Vitamin D deficiency in girls from South Brazil: a cross-sectional study on prevalence and association with vitamin D receptor gene variants

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Abstract

Background: Vitamin D deficiency has been associated with a multitude of disorders including diabetes, defective insulin secretion as well as rickets and poor bone health. Vitamin D is also a concern during childhood and adolescence and has been reported in girls from South Brazil. We determined the prevalence of vitamin D deficiency in girls from South Brazil and investigated whether the genotypic distribution of the BsmI, ApaI and TaqI polymorphisms of the VDR gene and their haplotypes were associated with vitamin D levels.

Methods: Cross-sectional study including 234 apparently healthy girls aged 7 to 18 years. Height and weight were measured for calculation of body mass index (BMI) percentiles for age. Plasma levels of 25-hydroxyvitamin D [25(OH)D] were assessed. Participants were genotyped for ApaI (rs7975232), TaqI (rs731236), and BsmI (rs1544410) SNPs.

Results: The median and interquartile range (25-75%) of BMI percentile was 62.0 (33.3 – 84.9). The frequency of overweight/obesity was 24.9%. Circulating levels of 25(OH)D (≥ 30 ng/mL) were adequate in 9.4%; insufficient in 54.3% (20–29 ng/mL); and deficient in 36.3% (< 20 ng/mL). Genotype frequencies were GG = 47.0%, GA = 41.5%, and AA = 11.5% for BsmI; GG = 16.7%, GT = 52.6%, and TT = 30.8% for ApaI; TT = 46.2%, TC = 44.9% and CC = 9.0% for TaqI. Genotypes with no gene variance (ancestral wild genotype) of BsmI (GG vs. GA + AA, two-tailed Student’s t-test p < 0.001), ApaI (GG vs. GT + TT, two-tailed Student’s t-test p = 0.031) and TaqI (TT vs. TC + CC, two-tailed Student’s t-test p = 0.005) SNPs and the GGT haplotype (two-tailed Student’s t-test p = 0.036) were significantly associated with lower 25(OH)D levels.

Conclusions: 25-hydroxyvitamin D deficiency and insufficiency were highly prevalent in this sample. The BsmI, ApaI and TaqI wild variants of the VDR gene, as well as the GGT haplotype, were associated with lower vitamin D levels, suggesting that VDR gene polymorphisms could be linked to higher susceptibility to vitamin D deficiency in a sub-population of children and adolescents.

Keywords: 25-hydroxyvitamin D, VDR gene polymorphisms, Pediatric female population

Background

Vitamin D is mainly recognized for its effects on bone metabolism and for being an important determinant of growth and body development during childhood and adolescence. However, a high prevalence of vitamin D deficiency has been observed in several pediatric and adolescent populations [1-3] in association with overweight/obesity, blood pressure and glucose metabolism [4-6]. While these studies report no gender-related differences in vitamin D levels, recent evidence shows that girls have significantly lower vitamin D levels than boys [7]. Our group has also observed a high prevalence of vitamin D deficiency in girls with precocious pubarche and controls with normal pubertal development in South Brazil [8].

The vitamin D receptor (VDR) gene regulates up to 200 genes [9], and mediates most effects of vitamin D on gene expression via formation of a heterodimer with...
the retinoid x receptor, which binds to promoter regions of many target genes [9]. Several polymorphisms have been described for the VDR gene (ID: 7421), located on chromosome 12 (12q13.11), consisting of 11 exons and spanning 63495 bp. BsmI, ApaI (both located in intron 8) and TaqI (located in exon 9) are the most studied variants. VDR gene polymorphisms have been linked with specific health outcomes, including low bone density in postmenopausal women [10], type 2 diabetes or metabolic syndrome [11,12] and low 25-hydroxyvitamin D (25(OH)D) concentration [13,14]. Given the high prevalence of vitamin D deficiency in children and adolescents described earlier, it may be speculated that VDR gene polymorphisms could be linked to higher susceptibility to develop vitamin D deficiency.

Therefore, the aims of the present study were to assess the genotypic distribution of the BsmI, ApaI and TaqI polymorphisms of the VDR gene in a population of girls from South Brazil and to determine whether these gene variants and their haplotypes are associated with 25(OH)D levels.

Methods

Subjects

This cross-sectional study was carried out between April 2008 and January 2011 and included 234 apparently healthy girls aged 7 to 18 years who had parental consent to participate in the study. Two hundred and thirteen girls recruited at four public schools from the four main regions of the city of Curitiba (North, South, East, and West), in the state of Paraná, Brazil (latitude −25°), and 21 girls enrolled at a vaccination facility or University adolescent clinic in the city of Porto Alegre (latitude −30°), state of Rio Grande do Sul, were included in the study. None of the girls took calcium or vitamin D supplements. Two girls used birth control pills and 19 made occasional use of bronchodilators or nasal corticosteroids for asthma or rhinitis. Most of these girls were also included in the control group of a previous study [8].

Approval for this study was obtained from the Institutional Review Boards and the local Ethics Committees of Hospital de Clínicas de Porto Alegre and Universidade Federal do Paraná. Written informed consent was obtained from all participants or their caretakers.

Study protocol

All subjects underwent physical examination. On that occasion, information on thelarche and menarche age was collected through interview. Anthropometric measurements included height and weight for subsequent calculation of body mass index (BMI). Height was obtained by using a stadiometer fixed to the wall and weight was obtained by using a digital balance, with resolution of 100 grams. The subjects were evaluated barefoot and wearing light clothing. Individual height and BMI values were converted into percentiles according to age based on 2000 Centers for Disease Control and Prevention charts [15]. For that, the software EpiInfo/AnthropometricData (version 3.5.1) was used.

25(OH)D, the main vitamin D circulating metabolite, was assessed in blood samples drawn between 8:00 and 10:00 AM from an antecubital vein, after an overnight fast. Blood samples were also collected for genomic DNA extraction.

Girls without thelarche at the time of enrollment were defined as prepubertal. Subjects were classified as normal weight (BMI ≤ 85 percentile), overweight (85 percentile ≤ BMI ≤ percentile 95) or obese (BMI > 95 percentile). Serum 25(OH)D status was classified as sufficient (≥ 30 ng/mL), insufficient (20–29 ng/mL) or deficient (< 20 ng/mL).

Assays

Serum 25(OH)D (sensitivity = 1.5 ng/mL) was measured with radioimmunoassay (DiaSorin, Stillwater, USA) with intra and inter-assay coefficients of variation of <12.0% and <15.0% respectively.

Genotype analysis

Genomic DNA was extracted from peripheral blood leukocytes. Molecular genotyping for the Apal (rs7975232) and TaqI (rs731236) SNPs (change of the G → T and T → C, respectively) was performed by polymerase chain reaction (PCR) followed by restriction fragment length polymorphism (RFLP) analysis [16]. Forward 5'-GTTCCAGCAGAAATGGGACACA-3' and reverse 5'-AGCTTCTGGATCATCTTGGCATAG-3' primer sequences yielded a 740 bp PCR product. Protocol conditions consisted of denaturation at 95°C for 2 min followed by 35 cycles (95°C, 30sec; 59.2°C, 30sec; 72°C, 80sec) and final extension at 72°C for 5 min. PCR products were digested overnight by the restriction enzymes Apal or TaqI (New England Biolabs, USA) at 37°C or 65°C, respectively. Apal digestion revealed genotypes TT (740 bp), TG (740, 559, and 181 bp) or GG (559 and 181 pb), while TaqI digestion denoted genotypes TT (740 bp), TC (740, 635 and 105 pb) or CC (635 and 105 pb) at 2% agarose gel electrophoresis.

BsmI (rs1544410) SNP (change of the G → A) genotyping was performed through real-time PCR (7500 Fast Applied Biosystems, California, USA) with allelic discrimination assays (Taqman MGB Probes®) according to the manufacturer’s instructions (Applied Biosystems, California, USA).

We calculated Lewontin’s D’ (|D’|) $r^2$ between each pair of genetic markers for estimating the linkage disequilibrium. Haplotypes were constructed from the combination of the three VDR polymorphisms (BsmI, Apal
and TaqI), and their frequencies were inferred using the PHASE 2.1.1 program. The first letter of each haplotype refers to BsmI polymorphism, the second to the ApaI polymorphism, and the third to TaqI polymorphism.

**Statistical analysis**

The sample size estimation was based on the study by Bhanushali et al. [13], in which a different frequency of the SNP TaqI was found in relation to vitamin D status in adults. Therefore, considering the same difference, an alpha of 5% and a beta of 90%, the sample size was estimated as 228 girls, 94 with genotype TT and 134 with genotypes TC + CC.

Data were described as mean ± standard deviation (SD) (Gaussian variables) or median and interquartile range (25-75%) (non-Gaussian variables). Comparisons between means were analyzed by the unpaired two-tailed Student’s t-test or one-way analysis of variance (ANOVA) followed by Tukey's post hoc test. Log10 transformation was used to normalize the distribution of non-Gaussian variables and mean values were back-transformed for presentation. Data were adjusted by season and/or age at the time of blood collection by linear regression.

Categorical variables and the agreement of genotype frequencies with Hardy-Weinberg equilibrium for each SNP were analyzed using the Pearson chi-square test [χ²]. Odds ratios (OR) and 95% confidence intervals (95%CI) were obtained using χ² risk estimate. Multiple linear regression analysis models were set up to determine the independent effects of season, age, and BMI percentile for each of the polymorphisms (BsmI, ApaI and TaqI) and the Ht1 haplotype, having plasma 25(OH)D D as the dependent variable. Data were considered as statistically significant at p < 0.05. The Statistical Package for the Social Sciences 16 (SPSS, Chicago, IL) was used in the analyses.

**Results**

All the 234 healthy girls who were enrolled following parental consent completed the interview, physical examination and blood collection. They were studied during spring/summer [35 (15%)] or fall/winter [199 (85%)]. Mean chronological age was 13.0 ± 1.9 years. The median BMI percentile was 61.9 (33.3 - 84.9). The frequency of overweight was 15.7%, and of obesity 9.2%. Only 4.0% of the participants were prepubertal, and 34.2% had not had menarche.

Table 1 shows clinical features and 25(OH)D levels in girls according to tertiles of age. Height and BMI percentiles were adequate for all age categories. 25(OH)D levels did not differ among age categories. Mean serum 25(OH)D was 21.3 ± 6.8 ng/mL. Stratification by 25(OH)D status revealed sufficient circulating levels (≥ 30 ng/mL) in only 9.4% of the overall group; 54.3% were insufficient (20–29 ng/mL) and 36.3% were deficient (< 20 ng/mL). No associations were found between season at the time of the blood collection and 25(OH)D levels (spring/summer = 21.4 ± 6.7 ng/mL vs. fall/winter = 21.3 ± 6.9 ng/mL; p = 0.955). No significant differences were found between season at the time of the blood collection and 25(OH)D status (χ² = 0.249; p = 0.883).

Concerning VDR polymorphisms, all three SNPs (BsmI, ApaI and TaqI) were in Hardy-Weinberg equilibrium. Genotype frequencies were GG = 47.0% (n = 110), GA = 41.5% (n = 97), and AA = 11.5% (n = 27) for BsmI SNP; GG = 16.7% (n = 39), GT = 52.6% (n = 123), and TT = 30.8% (n = 72) for ApaI SNP; TT = 46.2% (n = 108), TC = 44.9% (n = 105), and CC = 9.0% (n = 21) for TaqI SNP. Twenty-one randomly selected samples (around 10%) were genotyped twice, and the reproducibility of genotyping was 100%.

The BsmI (G → A) polymorphism was in partial linkage disequilibrium with the Apal (G → T) polymorphism (|D|² = 0.964; r² = 0.330), and in almost complete linkage disequilibrium with the TaqI (T → C) polymorphism (|D|² = 0.919; r² = 0.807). The Apal (G → T) polymorphism was also in partial linkage disequilibrium with the TaqI (T → C) polymorphism (|D|² = 0.970; r² = 0.319). Seven haplotypes were inferred in the sample (Ht1: GGT, Ht2: GGC, Ht3: GTT, Ht4: GTC, Ht5: AGT, Ht6: ATT and Ht7: ATC). Haplotype frequencies were Ht1: 0.419; Ht2: 0.004; Ht3: 0.242; Ht4: 0.013; Ht5: 0.006; Ht6: 0.019 and Ht7: 0.297.

No differences were found in age, height and BMI percentiles, age at menarche or at thelarche between genotypes for all the three polymorphisms. In contrast, presence of the wild-type genotypes BsmI, ApaI and TaqI SNPs of the VDR gene was significantly associated with lower 25(OH)D levels (BsmI p < 0.001; ApaI p = 0.031; TaqI p = 0.005), even after adjustment for season at the time of blood collection and age (Figure 1). Compared to the wild genotype, 25(OH)D deficiency (< 20 ng/mL) was less frequent with BsmI GA + AA (p = 0.014) and TaqI TC + CC (p = 0.034) genotypes (Figure 2). The OR for 25(OH)D deficiency among girls with the wild genotype was 1.96 (95%CI: 1.14–3.37) and 1.78 (95%CI: 1.04–3.06), for BsmI and TaqI SNPs, respectively. There was no difference in frequency of 25(OH)D deficiency between Apal polymorphisms and the wild genotype (Figure 2) (95%CI: 0.27–1.08; p = 0.078).

In addition, taking into consideration the haplotype frequencies and the results for individual polymorphism analyses, Ht1 was considered as the risk haplotype and Ht7 as the protective haplotype for circulating 25(OH)D levels. Patients with two copies of Ht1 (Ht1/ Ht1) presented lower 25(OH)D levels as compared to
patients having one or no copies of Ht1 (Ht1/0 + 0/0) (19.16 ± 7.08 vs. 21.70 ± 6.66 ng/mL, p = 0.036) and patients having one copy of Ht1 and one copy of Ht7 (Ht1/Ht7) or two copies of Ht7 (Ht7/Ht7) (22.68 ± 6.34 ng/mL, p = 0.008), even after adjustment for season at the time of the blood collection and age.

Multiple linear regression analysis assessing the effects of season, age, BMI percentile, and polymorphic genotypes on plasma 25(OH)D are shown in Table 2. BsmI, ApaI and TaqI polymorphisms, Ht1 haplotype (GGT) and age were significant independent predictors of plasma 25(OH)D, explaining 7.4%, 4.3%, 5.6% and 4.4% of variance in plasma 25(OH)D concentrations, respectively.

Discussion
In the present study, a high prevalence of vitamin D deficiency or insufficiency was found in 7 to 18-year old girls from South Brazil. In addition, VDR wild-type genotypes BsmI, ApaI and TaqI were associated with lower 25(OH)D levels both individually and as the GGT haplotype, even after adjustment for age and season at the time of blood collection, and were independent positive predictors of 25(OH)D levels. To the best of our knowledge, this is the first report relating VDR gene polymorphisms and haplotypes to 25(OH)D levels in healthy children and adolescent girls from the general population.

Previous studies have reported low levels of 25(OH)D in different pediatric and adolescent populations [2,3]. In our sample, only 9.4% of the participants had adequate circulating concentrations of 25(OH)D, which is far below the 39.0% reported for a 16–20 year old population from a rural town in the state of São Paulo, Brazil (latitude −23°) [17]. Since the latitude and solar radiation are similar in the cities included in those previous studies and in the cities studied by us, diverse dietary habits and the age of participants may explain the discrepancies in the frequency of sufficient 25(OH)D status. In this sense, one limitation of the present study is the lack of control for some confounders, such as dietary vitamin D intake and duration of sun exposure. However, no associations were found between 25(OH)D levels and season at the time of blood collection. One potential explanation for the lack of seasonal difference is that below a latitude of approximately 35°, UVB radiation is sufficient for vitamin D synthesis all year round [18].

Concerning young populations, several studies have shown that vitamin D levels are lower in the presence of overweight/obesity, and that low levels of 25(OH)D could influence the risk of developing metabolic disorders

Table 1 Anthropometric and clinical features of girls according to tertiles of age

| Variable                        | <12.04 (77) | 12.04-14.05 (80) | >14.05 (77) | p   |
|---------------------------------|------------|-----------------|------------|-----|
| Age (years)                     | 10.9 ± 1.1 | 13.0 ± 0.6      | 15.2 ± 1.1 | <0.001 |
| *Height percentile              | 66.2 (39.9, 82.1) | 55.65 (31.45, 74.40) | 41.87 (19.75, 77.13) | 0.066 |
| *BMI percentile                 | 62.9 (31.8, 86.4) | 59.9 (35.1, 85.1) | 61.8 (31.7, 82.1) | 0.967 |
| Age at thelarche (years)        | 9.9 ± 1.0  | 10.3 ± 1.1      | 10.5 ± 1.2  | 0.013 |
| Age at menarche (years)         | 10.9 ± 1.2 | 11.6 ± 1.0      | 12.1 ± 1.3   | 0.002 |
| 25(OH)D (ng/mL)                 | 20.77 ± 6.97 | 22.10 ± 6.01    | 21.09 ± 7.33 | 0.492 |

Values are expressed as mean ± SD or median and interquartile range (25-75%). p value by one-way analysis of variance (ANOVA).
* Log10 transformation.
Equal letters mean statistical difference between groups.

Figure 1 Vitamin D levels according to BsmI (A), ApaI (B), and TaqI (C) SNPs in VDR gene grouped by presence of polymorphic allele.

Values are expressed as means (central circle) and 95% CI (lower and upper limit). Two-tailed Student’s t-test, *p < 0.05 BsmI GG (n = 110) and GA + AA (n = 124); ApaI GG (n = 39) and GT + TT (n = 195); TaqI TT (n = 108) and TC + CC (n = 126).
and cardiovascular disease in pediatric and adolescent populations [4-6]. Obesity-associated vitamin D insufficiency is likely caused by deposition of skin and dietary vitamin D3 in body fat compartments, resulting in decreased bioavailability [19].

In addition, studies have shown that obese girls with vitamin D deficiency presented lower insulin sensitivity [4], and adolescents with reduced levels of vitamin D had increased risk of metabolic syndrome [5]. Moreover, a recent study with children aged 8 to 18 years, in which 47% were obese, reported a negative association between BMI, percentage of body fat, subcutaneous and visceral adipose tissue and 25(OH)D levels in white and black subjects [3]. Despite the age similarity, the frequency of obesity was very low in our sample, and therefore we were unable to assess these associations.

Only a few studies are available in the literature relating VDR polymorphisms and 25(OH)D levels, especially in young and healthy populations. In a study with adolescent girls, the BsmI polymorphism was marginally, but not significantly, associated with 25(OH)D levels [20]. Regarding adult populations, the results are conflicting. In India, one study reported that TaqI polymorphism was associated with higher 25(OH)D levels in healthy adults aged 25 to 60 years [13], while another study analyzing BsmI and TaqI SNPs did not confirm this association [21]. In addition, other SNPs in different regions of the VDR gene have been associated with levels of 25(OH)D in adolescent girls [20] and other populations [14].

In contrast, studies focusing on specific conditions, such as the overweight Insulin Resistance Atherosclerosis family study, in which only the BsmI SNP [22] was analyzed, or the Metabolites in Multiple Sclerosis study, covering Apal and TaqI SNPs [23], did not find an association between VDR polymorphism and 25(OH)D levels. However, these studies were carried out with disease-affected adult populations, whereas our results refer to younger, healthy subjects. In studies with older overweight subjects, BsmI, Apal, and TaqI SNPs of VDR gene were not associated with 25(OH)D levels. Nevertheless, the lack of association may be explained, at least

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**Table 2 Multiple linear regression analysis for plasma 25(OH)D (ng/mL)**

| 25(OH)D vs. independent variables | Coefficient (B) ± SE | p      | r²   |
|-----------------------------------|---------------------|--------|------|
| **Model 1**                       |                     |        |      |
| Season                            | −0.082 ± 1.249      | 0.948  | 0.074|
| Age (years)                       | 0.469 ± 0.225       | 0.038  |      |
| *BMI percentile                    | −0.004 ± 0.015      | 0.804  |      |
| BsmI GG vs. GA + AA               | 3.114 ± 0.881       | <0.001 |      |
| **Model 2**                       |                     |        |      |
| Season                            | −0.149 ± 1.270      | 0.907  | 0.043|
| Age (years)                       | 0.518 ± 0.228       | 0.024  |      |
| *BMI percentile                    | −0.005 ± 0.016      | 0.739  |      |
| Apal GG vs. GT + TT               | 2.575 ± 1.189       | 0.031  |      |
| **Model 3**                       |                     |        |      |
| Season                            | −0.131 ± 1.261      | 0.917  | 0.056|
| Age (years)                       | 0.497 ± 0.227       | 0.030  |      |
| *BMI percentile                    | −0.005 ± 0.015      | 0.732  |      |
| TaqI TT vs. TC + CC               | 2.505 ± 0.890       | 0.005  |      |
| **Model 4**                       |                     |        |      |
| Season                            | −0.115 ± 1.268      | 0.928  | 0.044|
| Age (years)                       | 0.510 ± 0.228       | 0.027  |      |
| *BMI percentile                    | −1.098 ± 1.382      | 0.428  |      |
| Ht1 haplotype vs. Ht1/0 + 0/0     | 2.632 ± 1.218       | 0.032  |      |

* Log₁₀ transformation.
in part, by the fact that most of the participants in that study were using vitamin D supplements [24,25].

VDR gene polymorphisms have also been studied in some other conditions. BsmI, ApaI, and TaqI polymorphic genotypes were associated with suppressed cytokine response in pulmonary tuberculosis [26] and appear to be related with higher glucose levels [12] and diabetes [27,28]. Apal gene variant has been linked to higher bone metabolism and bone mineral density [10]. We also have recently described that the wild genotype of the Apal polymorphism is more frequent in girls with precocious pubarche than in controls with normal pubertal development [8].

The BsmI, Apal and TaqI polymorphism are located at the 3' untranslated region (3' UTR) of the VDR gene. The BsmI and Apal SNPs are located in intron 8, and TaqI is a silent SNP in exon 9. Although 3' UTR has been recognized as a region involved in the modulation of gene expression, especially through the regulation of mRNA stability and efficiency of protein translation [29], the functional role of the BsmI, Apal, and TaqI VDR gene polymorphisms has not been established. Moreover, the most common haplotype for the VDR gene in the present study (Ht1) is the same found in Caucasians and Asians, followed by Ht7 and Ht3, as found for Caucasians [30]. A strong linkage disequilibrium between these three SNPs has been shown in the present and previous studies in different populations [31].

In the present study, girls with the wild genotype for BsmI and TaqI SNPs or presenting the Ht1 haplotype were found to be at higher risk for 25(OH)D deficiency. In turn, Ht7 was regarded as the protective haplotype for circulating 25(OH)D levels. Therefore, it is possible to hypothesize that the VDR genotype influences the susceptibility to vitamin D deficiency in children and adolescents. Thus, the present results open new possibilities for further studies in this field, especially on the genotype-related mechanisms of calcium and vitamin D metabolism and prevention of 25(OH)D deficiency in young subjects.

Conclusions

In conclusion, data from this study confirm the high prevalence of vitamin D deficiency and insufficiency in children and adolescent populations. The present findings also suggest that the BsmI, Apal and TaqI wild variants of the VDR gene and the GGT haplotype are associated with lower 25(OH)D levels. Further studies with populations of different ethnic origins are needed to confirm the clinical relevance of the present results.

Abbreviations

25(OH)D: 25-hydroxyvitamin D; 95%CI: 95% confidence interval; ANOVA: Analysis of variance; BMI: Body mass index; OR: Odds ratio; RFLP: Restriction fragment length polymorphism; SD: Standard deviation; VDR: Vitamin D receptor.

Competing interests

The authors declare that they have no competing interests.

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Authors’ contributions

LPGM, FS and MCSB contributed to acquisition of data, analysis and interpretation of data and manuscript review. BRS and PMS contributed to conception and study design, acquisition, analysis and interpretation of data, drafting manuscript and final review. All authors approved the final version of the manuscript.

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References

1. Harinarayan CV, Ramalakshmi T, Prasad UV, Sudhakar D: Vitamin D status in Andhra Pradesh: a population based study. Indian J Med Res 2006, 127(3):211–218.
2. Kumar J, Muntrer P, Kaskell FJ, Halipern SM, Melamed ML: Prevalence and associations of 25-hydroxyvitamin D deficiency in US children: NHANES 2001–2004, Pediatrics 2009, 124(3):e362–e370.
3. Rajakumar K, de las Heras J, Chen TC, Lee S, Holick MF, Aniansan SA: Vitamin D status, adiposity, and lipids in black American and Caucasian children. J Clin Endocrinol Metab 2010, 96(5):1560–1567.
4. Ashraf A, Alvarez J, Saenz K, Gower B, McCormick K, Frankfurt F: Threshold for effects of vitamin D deficiency on glucose metabolism in obese female African-American adolescents. J Clin Endocrinol Metab 2009, 94(9):3200–3206.
5. Reis JP, von Muhlen D, Miller ER 3rd, Michos ED, Appel LJ: Vitamin D status and cardiometabolic risk factors in the United States adolescent population, Pediatrics 2009, 124(3):e371–e379.
6. Johnson MD, Nader NS, Weaver AL, Singh R, Kumar S: Relationships between 25-hydroxyvitamin D levels and plasma glucose and lipid levels in pediatric outpatients. J Pediatr 2010, 156(3):444–449.
7. Al-Musharaf S, Al-Othman A, Al-Daghrī NM, Krishnaswamy S, Yosuf DS, Alkhadry KM, Al-Saleh Y, Al-Atta S, Akkol MS, Moharan Q, et al: Vitamin D deficiency and calcium intake in reference to increased body mass index in children and adolescents. Eur J Pediatr 2012, Feb 7, [Epub ahead of print].
8. Santos BR, Mascarenhas LP, Satler F, Bogsuzewski MC, Spritzer PM: Vitamin D receptor gene polymorphisms and sex steroid secretion in girls with precocious pubarche in Southern Brazil: a pilot study. J Endocrinol Invest 2011, Oct 3. [Epub ahead of print].
9. Dilworth FJ, Chambon P: Nuclear receptors coordinate the activities of chromatin remodelling complexes and coactivators to facilitate initiation of transcription. Oncogene 2001, 20(4):3047–3054.
10. Dundar U, Solak M, Kavuncu V, Ozdemir M, Cakir T, Yildiz H, Evcik D: Evidence of association of vitamin D receptor Apa I gene polymorphism with bone mineral density in postmenopausal women with osteoporosis. Clin Rheumatol 2009, 28(10):1187–1191.
11. Oh JY, Barrett-Conner E: Association between vitamin D receptor polymorphism and type 2 diabetes or metabolic syndrome in community-dwelling older adults: the Rancho Bernardo Study. Metabolism 2002, 51(3):356–359.
12. Ortepp JR, Mettikul J, Albrecht M, von Koff H, Hanrath P, Hoffmann R. The vitamin D receptor gene variant and physical activity predicts fasting glucose levels in healthy young men. *Diabetes Med* 2003, 20(6):451–454.

13. BhanuVishi AA, Lajpal N, Kulkarni SS, Chavan SS, Bagadi SS, Das BR. Frequency of foki and taqI polymorphism of vitamin D receptor gene in Indian population and its association with 25-hydroxyvitamin D levels. *Indian J Hum Genet* 2009, 15(3):108–113.

14. Smolders J, Damoiseaux J, Menheere P, Tervaert JW, Hupperts R. Fok-I vitamin D receptor gene polymorphism (rs10735810) and vitamin D metabolism in multiple sclerosis. *J Neuroimmunol* 2009, 207(1–2):117–121.

15. Kuczewska RJ, Ogden CL, Grummer-Strawn LM, Flegal KM, Guo SS, Wei R, Mei Z, Curtin LR, Roche AF, Johnson CL. CDC growth charts: United States. *Adv Data* 2000, 314:1–27.

16. Wilten D, Furtado L, Kohek MB, Spritzer PM. CAPN10 UC SNP-43, UC SNP-19 and UC SNP-63 polymorphisms and metabolic syndrome in polycystic ovary syndrome. *Gynecol Endocrinol* 2007, 23(3):173–178.

17. Peters BS, dos Santos LC, Fisberg M, Wood RJ, Martini LA. Prevalence of vitamin D insufficiency in Brazilian adolescents. *Ann Nutr Metab* 2009, 54(1):15–21.

18. Webb AR, Kline L, Holick MF. Influence of season and latitude on the cutaneous synthesis of vitamin D3: exposure to winter sunlight in Boston and Edmonton will not promote vitamin D3 synthesis in human skin. *J Clin Endocrinol Metab* 1988, 67(2):373–378.

19. Wortsman J, Matsuzaka LV, Chen TC, Hu C, Holick MF. Decreased bioavailability of vitamin D in obesity. *Am J Clin Nutr* 2000, 72(3):690–693.

20. d’Alesio A, Garabedian S, Sabatier JP, Guaydier-Souqueges G, Marcelli C, Lemacon A, Walrant-Debray O, Jehan F. Two single-nucleotide polymorphisms in the human vitamin D receptor promoter change protein-DNA complex formation and are associated with height and vitamin D status in adolescent girls. *Hum Mol Genet* 2005, 14(23):3539–3548.

21. Vupputuri MR, Goswami R, Gupta N, Ray D, Tandon N, Kumar N. Prevalence and functional significance of 25-hydroxyvitamin D deficiency and vitamin D receptor gene polymorphisms in Asian Indians. *Am J Clin Nutr* 2006, 83(6):1411–1419.

22. Engelmann CD, Fingerlin TE, Langefeld CD, Hicks PJ, Rich SS, Wagenknecht LE, Bowden DW, Norris JM. Genetic and environmental determinants of 25-hydroxyvitamin D and 1,25-dihydroxyvitamin D levels in Hispanic and African Americans. *J Clin Endocrinol Metab* 2008, 93(9):3381–3388.

23. Smolders J, Damoiseaux J, Menheere P, Tervaert JW, Hupperts R. Association study on two vitamin D receptor gene polymorphisms and vitamin D metabolites in multiple sclerosis. In Contemporary challenges in autoimmunity. Edited by Shoerden Y, Gershwin M. Oxford: Blackwell Publishing, 2009:515–520.

24. Hübner EA, Jurutka PW, Egan JH, Hu C, LeRoy EC, Martinez ME, Thompson PA, Jacobs ET. Association between polymorphic variation in VDR and RXRA gene on cytokine response in pulmonary tuberculosis. *J Immunol* 2010, 184(1):438–441.

25. Bu FX, Arnas L, Lappe J, Zhou Y, Gao G, Wang HW, Recker R, Zhao L. Comprehensive association analysis of nine candidate genes with serum 25-hydroxy vitamin D levels among healthy Caucasian subjects. *Hum Genet* 2010, 128(5):549–556.

26. Selvaraj P, Vidyarani M, Alagarasu K, Prabhu Anand S, Narayanan PR. Regulatory role of promoter and 3’ UTR variants of vitamin D receptor gene on cytokine response in pulmonary tuberculosis. *J Clin Immunol* 2008, 28(6):306–313.

27. Panierakis C, Goulleimlos G, Mamoulakis D, Petrali E, Papavasiliou E, Galanakis E. Vitamin D receptor gene polymorphisms and susceptibility to type 1 diabetes in Crete, Greece. *Clin Immunol* 2009, 133(2):278–281.

28. Nosratabadi R, Arababadi MK, Salehabad VA, Shamsizadeh A, Mahmoodi M, Sayadi AR, Kennedy D. Polymorphisms within exon 9 but not intron 8 of the vitamin D receptor are associated with the nephropathic complication of type-2 diabetes. *Int J Immunogenet* 2010, 37(6):493–497.

29. Ogunkolade BW, Boucher BJ, Prahl JM, Bustin SA, Burin JM, Noonan K, North BV, Mannan N, McDermott MF, DeLuca HF, et al. Vitamin D receptor (VDR) mRNA and VDR protein levels in relation to vitamin D status, insulin secretory capacity, and VDR genotype in Bangladeshi Asians. *Diabetes* 2002, 51(7):2294–2300.

30. Thakkarista A, De Stree C, Attia J. Haplotype analysis of VDR gene polymorphisms: a meta-analysis. *Osteoporos Int* 2004, 15(9):729–734.

31. Utterlinden AG, Fang Y, Van Meurs JB, Pols HA, Van Leeuwen JP. Genetics and biology of vitamin D receptor polymorphisms. *Gene* 2004, 338(2):143–156.