Cellular Heterogeneity in Cultured Human Chondrocytes Identified by Antibodies Specific for α2(XI) Collagen Chains

B. Swoboda,* R. Holmdahl,† H. Stöß,‡ and K. von der Mark*

*Max-Planck-Society, Clinical Research Unit for Rheumatology at the University of Erlangen-Nürnberg, D-8520 Erlangen, Federal Republic of Germany; † University of Uppsala, Department of Medical and Physiological Chemistry, S-75123 Uppsala, Sweden; and ‡ Department of Pathology, University of Erlangen-Nürnberg, D-8520 Erlangen, Federal Republic of Germany

Abstract. Collagen type XI is a component of hyaline cartilage consisting of α1(XI), α2(XI), and α3(XI) chains; with 5–10% of the total collagen content, it is a minor but significant component next to type II collagen, but its function and precise localization in cartilaginous tissues is still unclear. Owing to the homology of the α3(XI) and α1(II) collagen chains, attempts to prepare specific antibodies to native type XI collagen have been unsuccessful in the past.

In this study, we report on the preparation and use for immunohistochemistry of a polyclonal antibody specific for α2(XI) denatured collagen chains. The antibody was prepared by immunization with the isolated α2(XI) chain and reacts neither with native type XI collagen nor type I, II, V, or IX by ELISA or immunoblotting, nor with α1(XI) or α3(XI), but with α2(XI) chains. Using this antibody, it was possible to specifically localize α2(XI) in cartilage by pretreating tissue sections with 6 M urea. In double immunofluorescence staining experiments, the distribution of α2(XI) as indicative for type XI collagen in fetal bovine and human cartilage was compared with that of type II collagen, using a monoclonal antibody to α1(II). Type XI collagen was found throughout the matrix of hyaline cartilage. However, owing to cross-reactivity of the monoclonal anti-α1(II) with α3(XI), both antibodies produced the same staining pattern. Cellular heterogeneity was, however, detected in monolayer cultures of human chondrocytes. Numerous cells stained intracellularly for α2(XI) but not for α1(II)/α3(XI) and vice versa, besides the majority of the cells that stained with both antibodies. These findings suggest that (a) chondrocytes may not always simultaneously synthesize α1(II) and α2(XI) collagen chains in cell culture; and (b) that a subpopulation of chondrocytes may exist in vitro that synthesize type XI collagen molecules containing α2(XI) but not α3(XI) subunits.

The assembly of the highly hydrated, space-filling extracellular matrix of hyaline cartilage involves the interaction of specific proteoglycans and the collagen types II, IX, and XI that are unique for this tissue (for review, see Mayne and von der Mark, 1983; van der Rest and Mayne, 1987; Eyre and Wu, 1987), with the exception of vitreous humor (Swann and Scotsman, 1980) and certain transient embryonic epithelia (Linsenmayer et al., 1977; von der Mark et al., 1977; Thorogood et al., 1986). Type VI collagen recently described in cartilage (Ayad et al., 1984; Keene et al., 1988) also exists in skin and other connective tissues (Timpl and Engel, 1987; Keene et al., 1988), and type X only occurs in hypertrophic cartilage (Schmid and Linsenmayer, 1985; Capasso et al., 1984; Gibson and Flint, 1985). The size, degree of cross-linking, and the orientation of collagen fibres determines to a large extent the biomechanical properties of hyaline cartilage. Thus, the role of the individual collagen types in the assembly of collagens fibrils and the three-dimensional collagenous meshwork is the subject of intense investigation.

Using electron microscopical technique, Vaughan and coauthors (1987) were able to show that type IX collagen molecules, which are covalently cross-linked to type II collagen (Eyre et al., 1987), decorate the periphery of type II collagen fibrils in a D-periodic pattern. Type VI collagen forms a fine fibrous meshwork interwoven with the type II collagen fibrils (Keene et al., 1988). Type XI collagen apparently does not assemble into separate supramolecular structures in cartilage, but is integrated into type II collagen fibrils; it may control the fibril diameter (Mendler et al., 1989).

Type XI collagen is thought to be a 1:1:1 heterotrimer consisting of α(XI), α2(XI), and α3(XI) chains (Burgeson and Hollister, 1979; Morris and Bachinger, 1987). Parent type XI collagen molecules consisting of either [α1(XI), α2(XI), α3(XI)] or [α3(XI)] have never been observed. Type XI collagen has the character of a "hybrid" collagen: α1(XI) and α2(XI) are similar to α1(V) and α2(V) in terms of solubility and migration on SDS-PAGE (Burgeson and Hollister, 1979; Reese and Mayne, 1981; von der Mark et al., 1982); α3(XI) is probably coded by a common type II collagen gene and differs

© The Rockefeller University Press, 0021-9525/89/09/1363/7 $2.00
The Journal of Cell Biology, Volume 109, September 1989 1363-1369 1363
from α(II) by posttranslational modifications (Furuto and Miller, 1983; Eyre and Wu, 1987), possibly also by amino acid substitutions that give rise to a different CNBr-peptide pattern (von der Mark et al., 1982).

Collagen types IX and XI always coextract together with type II collagen; e.g., from human articular cartilage (Burgeson and Hollister, 1979), chick sternal cartilage (Reese and Mayne, 1981; von der Mark et al., 1982), and from bovine and porcine cartilage (Eyre and Wu, 1987). In chondrocyte cultures modulated by bromodesoxoxyuridine or grown to senescence, the transition from type II synthesis to type I collagen is accompanied by a change from type XI to type V collagen (Mayne et al., 1984), suggesting coordinate regulation of types II and XI. In this study, however, we provide evidence by immunofluorescence double staining that in cultured chondrocytes the α2 subunit of type XI collagen may not always be simultaneously synthesized with α1(II). Using polyclonal antibodies specific for α2(XI) and monoclonal antibodies recognizing α1(II) and α3(XI) chains (Holm Dahl et al., 1986), we show that in monolayer cultures of human chondrocytes numerous cells stain for α2(XI) but not for α1(II)/α2(XI) collagen chains and vice versa, although a majority of cells stain with both antibodies. The rabbit antibody specific for denatured α2(XI) collagen chains was obtained by immunization with purified α2(XI) and can also be used to identify type XI collagen in cartilage after denaturing tissue sections with urea.

Materials and Methods

Collagen Preparation

Epiphysial cartilage from long bones of 3–6 mo calf fetuses was dissected free of adhering connective tissue, cut in pieces, weighed, and homogenized at 4°C with an Ultraturrax (Janke and Kunkel, FRG) in 0.15 M NaCl, 0.05 M Tris-HCl, pH 7.5, containing protease inhibitors (1 mM EDTA, 1 mM N-ethylmaleimide and 1 mM phenyl methane sulfonylfluoride) and 50 μg/ml Nε-thylmaleimide and 1 mM phenyl methane sulfonylfluoride) and 50 μg/ml. The extract was centrifuged for 30 min at 50,000 g, and the residual pellet was subsequently extracted with 4 M guanidinium hydrochloride 0.05 M Tris-HCl, pH 7.4, then with 0.5 M acetic acid, and finally with 5 mg pepsin/g wet weight cartilage in 0.5 M HCl at 4°C for 24 h as described previously (von der Mark et al., 1982). The pepsin extract was neutralized, and the collagens were separated into 3 fractions by subsequent precipitation with 0.7, 1.2, and 2 M NaCl in 0.5 M acetic acid as described elsewhere (Shimokomaki et al., 1980). The 1.2 M NaCl fraction containing type XI collagen was dissolved in 0.5 M acetic acid and reprecipitated with 0.86 and 1.2 M NaCl.

Collagens type II and IX from bovine fetal cartilage were obtained in the same preparation and purified according to von der Mark et al. (1982). Collagen types I and V were extracted and purified from human placenta as described by Benz et al. (1978).

Isolation of α2(XI)

α1, α2, and α3(XI) collagen chains were separated by chromatography on carboxymethyl cellulose at 42°C in 0.04 M sodium acetate, pH 4.8, containing 6 M urea as described previously (von der Mark et al., 1982). The purity of the α-chains was checked by SDS-gel electrophoresis in 7% acrylamide minigels using the buffer system of Laemmli (1970).

Antibodies

Rabbits were immunized by subcutaneous injection with 1 mg purified α2(XI) in 1 ml 1 M NaCl/0.05 M Tris-HCl, pH 7.5, emulsified in 1 ml complete Freund's adjuvant. Blood was collected from the ear vein after 2 mo following two booster injections, 1 mg α2(XI), in incomplete Freund's adjuvant 4 and 6 wk after the first immunization. The antibody titer was checked by ELISA (see below). The antiserum was absorbed with collagen types I, II, V, and IX collagen followed by affinity chromatography on type II collagen as described previously (von der Mark et al., 1976). Rabbit antibodies against human type II collagen were obtained by immunization with native type II collagen extracted from human fetal epiphysial cartilage. Specific antibodies were isolated after cross absorption with types I, V, and IX collagen followed by affinity chromatography on type II collagen as described previously (von der Mark et al., 1976).

The monoclonal antibody CII D3 is a Ig G2a, kappa isotype antibody cloned after immunization of a DBA/I mouse with native chick type II collagen (Holm Dahl et al., 1986). The antibody has been shown to react with native type II collagen from chick, mouse, rat, bovine, and human cartilages, but not with type I collagen (Holm Dahl et al., 1986).

ELISA

Antibody tiers were determined by ELISA using collagen-coated microtitre plates (coating solutions: 50 μg collagen/ml; Engvall and Perlmann, 1971). The second antibody was peroxidase-conjugated goat anti-rabbit-Ig (Bio-Rad Laboratories, Richmond, CA) diluted 1:5,000. Color substrate was 3',5'-tetramethylbenzidine.

Immunoblotting

Collagen type I, II, V, and IX were Western blotted onto nitrocellulose filters (Bio-Rad Laboratories) after electrophoresis in 7% polyacrylamide-SDS gels (Towbin et al., 1979). After blocking with low fat milk protein, blotted proteins were immunolabeled with rabbit anti α2(XI) or monoclonal anti type II collagen (CII D3) both diluted 1:100 with PBS, using peroxidase-labeled goat anti-rabbit-Ig (Bio-Rad Laboratories) as second antibodies, or rabbit anti-mouse-Ig (Bio-Rad Laboratories), and chloronaphthol as color substrate.

Cell Culture

Human chondrocytes were prepared from femoral head cartilage of an autopsied 22-wk-old fetus. The cartilage was dissected free of adhering connective tissue, and digested with trypsin (2 mg/ml; Serva, Heidelberg, FRG) in PBS for 45 min at 37°C followed by clostridium collagenase (2 mg/ml; CIL II; Worthington, Freehold, NJ) in Ham's F12 medium containing 10% FCS at 37°C. When the cartilage was digested (usually after 8–12 h), chondrocytes were filtered through 2 layers of a 20 mesh nylon net and washed by centrifugation. Cells were plated at 2 × 10⁶ cells/60 mm tissue culture dish and grown in F12 medium containing 10% FCS, Penicillin/Streptomycin (100 U/ml), glutamine (200 μM), and sodium ascorbate (50 μg/ml) (Dessau et al., 1978).

Collagen Biosynthesis

First passage chondrocytes monolayer cultures were labeled for 8 h with 10 μCi/ml of [1-2,3]-[3H]proline (30 Ci/mmol; Amersham Buchler GmbH, Braunschweig, FRG) in DME without serum, containing sodium ascorbate (50 μg/ml), β-aminopropionitrile (50 μl/ml), and glutamine (180 μg/ml). The newly synthesized collagens secreted into the medium and retained in the cell layer were digested with pepsin at 4°C for 12 h and analyzed by SDS-gel electrophoresis and fluorography as described (Herrmann et al., 1980).

Immunofluorescence

8-μm-thick sections were cut from fetal bovine and human cartilage and mounted on polylysine-coated glass slides. To reveal α2(XI)-staining, sections were treated with testicular hyaluronidase (2 mg/ml; Worthington, Freehold, NJ) in Ham's F12 medium containing 10% FCS at 37°C. Sections were treated with testicular hyaluronidase (2 mg/ml) for 20 min at 20°C and then with 6 M urea, 0.05 M sodium acetate, pH 4.8, for 20 min in a humidified atmosphere at 50°C. Sections were washed in PBS and stained with affinity-purified rabbit anti α2(XI) (10 μg/ml) followed by fluorescein-conjugated goat anti-rabbit-Ig. For double staining with monoclonal antibody CII D3, fluorescein-conjugated goat anti-mouse-Ig (1:40 dilution; Jackson Laboratory, Bar Harbor, ME) and rhodamin-conjugated goat anti-rabbit-Ig (1:40 dilution; Bio-Rad Laboratories). Sections were mounted in 10% mowiol 4-88, (Hoechst AG, FRG), 25% glycerol, 0.1 M Tris-HCl, pH 8.5, and viewed under a Diaplan microscope (E. Leitz, Wetzlar, FRG) equipped for epifluorescence.

Chondrocyte monolayer cultures were fixed with 70% ethanol for 10 min and stained with antibodies as described previously (von der Mark and von der Mark, 1977).

The Journal of Cell Biology, Volume 109, 1989 1364
Results

Preparation of Antibodies Specific for Type XI Collagen

Owing to the high degree of homology of α3(XI) with α1(II), previous attempts in our laboratory have failed to obtain antibodies against native type XI collagen without cross reaction with type II collagen. For this reason, α1(XI) and α2(XI) chains from fetal bovine cartilage were isolated and purified by carboxymethylcellulose chromatography as described previously for chicken cartilage (von der Mark et al., 1982) (Fig. 1). The homogeneity of the α chains was checked by SDS-PAGE (Fig. 2). Rabbits were immunized with purified α2(XI) chains; the antiserum was absorbed with collagen types I, II, IX, and V collagen and purified by affinity chromatography on α2(XI) coupled to Sepharose. The specificity of the antibody for α2(XI) and the absence of cross-reactivity with native collagen types I, II, and IX was determined by ELISA (Fig. 3). Interestingly, the purified α2(XI) antibodies failed to react with native type XI collagen (Fig. 3).

They were also specific for α2(XI) in immunoblotting and stained neither α1(XI) nor α3(XI), nor α1(II), nor human α2(V) chains (Fig. 4 B). The monoclonal antibody CII D3 prepared against chick type II collagen reacted with bovine (not shown) and human type II collagen in the immunoblot, as well as with bovine α3(XI), but not with α1(XI) or α2(XI) (Fig. 4 A).
Localization of Type XI Collagen in Fetal Cartilage

In cryostat sections of bovine fetal cartilage, the antibody to \( \alpha_2(\text{XI}) \) did not reveal any immunofluorescence reaction even after hyaluronidase treatment. However, after denaturing the tissue collagen with 6 M urea, pH 4.8, for 20 min at 50°C, the entire cartilaginous area was labeled (Fig. 5, b and d). Perichondral tissue and endochondral bone were unlabeled. Double immunofluorescence staining with the monoclonal antibody to type II collagen revealed extensive colocalization of both collagen types in hypertrophic (Fig. 5, a and b) as well as in resting (Fig. 5, c and d) fetal bovine cartilage. Similar results were obtained after staining of fetal human cartilage (not shown).

Identification of Intracellular Type XI Collagen in Chondrocyte Cultures

Chondrocytes from human fetal femoral heads were grown in monolayer cultures for 1 wk and labeled metabolically with \([\text{H}]\)proline to assess type XI collagen synthesis. Both secreted and cell-bound collagens were digested with pepsin at 4°C, and analyzed by SDS-PAGE (Fig. 6). Besides a majority of \( \alpha_1(\text{II}) \) and some \( \alpha_2(\text{II}) \) chains, \( \alpha_1(\text{XI}) \) and \( \alpha_2(\text{XI}) \) as well as fragments of type IX collagen were identified. The band of \( \alpha_3(\text{XI}) \) is probably covered by \( \alpha_1(\text{II}) \).

When subcultures were fixed and stained with antibodies to human type II collagen, only \( \sim 50\% \) of the cells showed intracellular reactions (Fig. 7 b), indicating the start of cellu-
Discussion

In this study, we report on the preparation of an antibody specific for α2(XI) collagen that can be used for the immunolocalization of type XI collagen in hyaline bovine and human fetal cartilages. While previous attempts to obtain antibodies specific for type XI collagen by immunization with native type XI collagen molecules have failed as a result of cross-reactivity with types II and V collagen, immunization with purified α2(XI) chains led to the production of antibodies specific for this polypeptide.

The availability of an antibody recognizing only denatured α2(XI) chains but not native type XI collagen might be useful for the detection of collagen degradation in cartilage. Recently, Dodge and Poole (1989) have reported on the immunohistochemical detection and quantitative analysis of type II collagen chains and fragments in rheumatoid cartilage or bovine cartilage explants treated with Interleukin 1.

The anti α2(XI) antibody can, however, also be used for the identification of intact type XI collagen in tissue sections. Similar to immunofluorescence localizations of type V collagen using monoclonal antibodies that required acid or pepsin pretreatment of tissue sections (von der Mark and Ocalan, 1982; Linsenmayer et al., 1983), reaction with type XI collagen in tissue sections could be achieved after treatment of the section with urea. In double staining experiments with a monoclonal antibody against type II collagen, we found the same distribution for type XI collagen and type II collagen in the extracellular matrix of fetal bovine and human cartilages. In accordance with this finding, Mendler et al. (1989) have shown by immunogold-labeling using anti α2(XI) that type XI collagen is incorporated into type II collagen fibrils; also in their study the antigenic epitopes of type XI collagen were exposed only after partial disruption of the fibrils.

Similarly, by immunofluorescent investigations (von der Mark and Ocalan, 1982; Linsenmayer et al., 1983) and fibril reconstitution experiments (Adachi and Hayashi, 1985), it has become evident that type V collagen is integrated into type I collagen fibrils (Birk et al., 1988) and may be involved in the control of fibril growth. Thus, type XI collagen is frequently considered as the type V collagen-analogue of hyaline cartilage (Burgeson and Hollister, 1979; Reese and Mayne, 1981; Eyre and Wu, 1987).

Several lines of evidence indicate coordinate synthesis of type II and type XI collagen in hyaline cartilage: (a) αI(II) and α3(XI) have a common genetic origin; (b) type II and type XI collagen are consistently found together in cartilage; (c) a concurrent change from type II and XI collagen synthesis to type I and V collagen synthesis follows the modulation of chondrocytes by BUdR or growth to senescence (Mayne et al., 1984). Here we present data, however, which indicate that, at the level of an individual chondrocyte, the subunits of type II and XI collagen are not necessarily simultaneously synthesized. Using antibodies specific for α2(XI) and αI(II), we have identified cells staining intracellularly for α2(XI) but not for αI(II) collagen, and vice versa. The fact that in the same culture α2(XI)-positive and αI(II)/α3(XI)-negative, as well as αI(II)/α3(XI)-positive and α2(XI)-negative cells were observed excluded the possibility of antigen masking, cross-reactivity of antibodies, or lack of species cross-reactivity. Antigen masking in individual cells is also unlikely in view of the observation that even chondrocytes that have developed a pericellular matrix in cell culture still take up antibody after alcohol fixation and reveal intracellular fluorescence reactions (Dessau et al., 1981).
These observations suggest that either different populations of chondrocytes emerge in monolayer culture synthesizing either α1(II) or α2(XI) (or, as many cells, both simultaneously), or that the synthesis of the subunits of type II and type XI does not occur synchronously in all cells. The latter hypothesis is supported by the observation that in some cells staining with both antibodies, the reaction for α1(II)/α3(XI) seems to be more intense in the Golgi region, while the α2(XI) antibody stained preferentially the peripheral cytoplasmic domains, probably the region of the RER. However, this differential staining pattern could also be a result of the inability of the anti α2(XI) antibody to react with the native type XI collagen molecules in the Golgi region, and a higher affinity of the monoclonal antibody to native, densely packed type II collagen molecules in the Golgi region as compared to nascent α1(II) and α3(XI) chains in the RER. Studies at the electronmicroscopical level would be required to answer this question.

Since fibril assembly occurs extracellularly, this finding is not in conflict with the idea that after secretion of complete type II and XI collagen molecules seem habitually to be incorporated into the same fibrils (Mendler et al., 1989).

The observation that some cells stained for α2(XI) but not with the monoclonal antibody specific for α1(II)/α3(XI), suggests that even the synthesis of the subunits of type XI collagen may not occur simultaneously in some cells. The question remains open, whether in such cells the α2(XI) chains assemble into triplehelical molecules together with α1(XI) but not α3(XI), or whether the α2(XI) chains are degraded intracellularly. The possibility that molecules are synthesized and secreted containing only α1(XI) and α2(XI) but not α3(XI) subunits is not unlikely in view of the fact that also type V collagen molecules have been identified containing only α2(V) but not α1 or α3(V) subunits (Haralson et al., 1980).

We wish to thank Ms. Klara Lauerer and Barbara Reiß for expert technical assistance, Thorsten Kirsch for valuable advice in FPLC techniques and Mrs. M. Rauber for typing the manuscript. We are also grateful to Dr. S. Goodman for critical reading of the manuscript.
This work was financially supported by the German Secretary for Research and Technology (BMFT, VM 8619/2).

Received for publication 2 March 1989 and in revised form 27 April 1989.

References

Adachi, E., and T. Hayashi. 1985. In vitro formation of fine fibrils with a D-periodic banding pattern from type V collagen. Collagen Relat. Res. 5:225-232.

Ayad, S., H. Evans, J. B. Weiss, L. Holt. 1984. Type VI collagen but not type V collagen is present in cartilage. Collagen Relat. Res. 4:165-168.

Bentz, H., H. P. Bächinger, R. Gianvilli, and K. Kühl. 1978. Physical Evidence for the assembly of A and B chains of human placental collagen in a single triple helix. Eur. J. Biochem. 92:563-567.

Birk, D. E., J. M. Fitch, J. M. Babiarz, and T. F. Linsenmayer. 1988. Collagen and cartilage contains mixed fibrils of collagen types I and V. Proceedings of the National Academy of Sciences USA. 85:7952-7956.

Birk, D. E., and J.-J. Wu. 1987. Type XI collagen is a heterotrimer with the composition (1α, 2α, 3α) retaining non-triple-helical domains. J. Biol. Chem. 262:11345-11350.

Bruneau et al. 1984. Collagen Type XI in Chondrocyte Cultures. Biochemistry. 20:5443-5448.

Capasso, O., G. Tajana, and R. Cancenedda. 1984. Location of 64K collagen types in cartilage: identification of several new collagen chains. Biochem. Biophys. Res. Commun. 87:1124-1131.

Capasso, O., G. Tajana, and R. Cancenedda. 1984. Location of 64K collagen types in cartilage: identification of several new collagen chains. Biochem. Biophys. Res. Commun. 87:1124-1131.

Capasso, O., G. Tajana, and R. Cancenedda. 1984. Location of 64K collagen types in cartilage: identification of several new collagen chains. Biochem. Biophys. Res. Commun. 87:1124-1131.