Glycodelin A is a prognostic marker to predict poor outcome in advanced stage ovarian cancer patients

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Christoph Scholz¹†, Sabine Heublein²†, Miriam Lenhard³, Klaus Friese²,³, Doris Mayr⁴ and Udo Jeschke²* 

Abstract

Background: Glycodelin is a cell surface glycoprotein offering a unique gender specific carbohydrate configuration. Sialylated carbohydrate structures, which are unusual for mammals, characterize Glycodelin isolated from amniotic fluid (Glycodelin A, GdA). Glycodelin in general exerts multiple, partly opposing functions ranging from immunosuppression to cell differentiation. As these markedly influence tumorigenesis, this study aimed to clarify whether expression of different Glycodelin isoforms is related to clinicopathological characteristics and prognosis of ovarian cancer patients. Further the use of Glycodelin as a serum marker in benign and malignant ovarian diseases was evaluated.

Methods: Ovarian cancer specimens (n = 152) were stained for Glycodelin with carbohydrate and peptide specific antibodies. Associations between Glycodelin expression and histological grading, FIGO stage as well as patient’s prognosis were examined. Glycodelin was correlated to expression of gonadotropin receptors and mucin-1, which are discussed as ovarian cancer tissue markers. In addition, Glycodelin serum concentrations were analyzed in patients suffering from benign (n = 73) or malignant (n = 38) ovarian neoplasias.

Results: Glycodelin A was found to be an independent prognostic marker for poor prognosis in advanced ovarian cancer patients. GdA staining correlated with gonadotropin receptor (FSHR and LHCGR) and with hCG expression. Gd expression showed a positive correlation with a tumour-associated epitope of mucin 1 (TA-MUC1). Further, compared to ovarian cancer, serum Gd was increased in patients with benign ovarian tumors.

Conclusion: Glycodelin A might be related to tumor aggressiveness and poor clinical outcome in advanced epithelial ovarian cancer. Glycodelin serum levels found in patients suffering from benign ovarian tumors, might contribute to a more global attenuation during progression of these precursor lesions.

Keywords: Ovarian cancer, Glycodelin, Immunohistochemistry, Prognosis

Background

Epithelial ovarian cancer (EOC) represents the most lethal malignancy of the female genital tract. Nowadays ovarian cancer patients’ prognosis mostly relies on completeness of surgical tumor resection [1,2], clinical staging and histological tumor grading[3]. Thus there is a compelling need to identify and validate tumor specific antigens which are suitable to individualize therapeutic strategies. Interestingly, during EOC evolvement and progression host anti-tumor immune defense seems to be actively blocked by tumor derived mediators. By creating this highly suppressive environment, EOC manages to extensively grow and spread in the peritoneal cavity.

Glycodelin (Gd), a potent immunosuppressive agent of the reproductive tract, is supposed to contribute to this immune tolerant phenotype. Gd is a glycoprotein whose immune-regulatory actions have been highlighted within different biological processes [4-6] and which is abundantly found in the female reproductive tract [7-9]. Structure wise it is part of the lipocalin superfamily and exerts its potent immune-regulatory activity via its unique, heavily sialysiated glycosylation pattern. Apart

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from its physiologic role as an immunomodulatory agent during implantation of the fetal semiallotransplantant it is also expressed by malignant tissues and contributes to the tumor-micromilieu [10,11]. Nevertheless, the physiological importance of Gd-expression in malignant diseases remains unknown. Gd is one of very few proteins that show a gender specific glycosylation pattern. Glycodelin, isolated from amniotic fluid (glycodelin A, GdA) is composed of two identical subunits closely connected by non-covalent bonds and a carbohydrate content of 17.5% [12]. A similar glycoprotein, Glycodelin S (GdS), was found in seminal plasma, but with a different glycosylation compared to GdA. While GdA is heavily sialylated, GdS is characterized by fucose-rich carbohydrate structures [13].

In the current study Gd was detected by antibodies raised against peptide sequences, which are not gender specific or specific for GdA or GdS, and a GdA specific monoclonal antibody [14,15]. In this work we aimed to clarify whether Gd expression in EOC is of prognostic significance. Further Glycodelin was correlated to expression of gonadotropin receptors and Mucin-1, which are discussed as ovarian cancer tissue markers. Finally we tested whether Glycodelin might be a potentially useful serum biomarker to detect ovarian cancer.

Materials and methods
Tissue acquisition
All tissue samples (n = 152) were got at surgery for primary EOC in patients treated at the Department of Obstetrics and Gynecology of the Ludwig-Maximilians-University Munich between 1990 and 2002. Specimens were assessed by two gynecological pathologists according to the criteria of the FIGO and the World Health Organization (WHO). Follow up data, which were received from the Munich Cancer Registry, and patients’ characteristics are listed in Table 1.

Sera of 111 patients, who underwent surgery at the Department of Obstetrics and Gynecology of Ludwig-Maximilians-University Munich between 2002 and 2005, were collected before surgery. Histological diagnoses (Table 1 B, benign ovarian diseases, n = 73 and EOC, n = 38) were made by gynecological pathologists. Written informed consent was obtained from all patients before surgery. Benign ovarian diseases were set up of cystic lesions (n = 21; serous cysts, mucinous cysts, follicle cysts, inclusion cyst, corpus luteum cysts), inflammatory diseases (n = 18; endometriosis cysts, sactosalpinx) and benign tumors (n = 34; serous and mucinous cystadenofibroma, fibroma, teratoma). Sera of EOC patients (n = 38) were analyzed in parallel.

Immunohistochemistry
Tissue specimens were dewaxed in xylol and endogenous peroxidase was quenched with 3% hydrogen peroxide for 20 min. Following epitope retrieval, slides were blocked and antibody staining was performed using the Vectastain elite kit (Vector Laboratories, Burlingame, USA) according to the manufacturer’s protocol. For details on blocking and staining procedure see Table 2. Finally, slides were stained with diaminobenzidine (Dako, Hamburg, Germany), counterstained in Mayer’s acidic hematoxylin and cover slipped with Consul Mount (Thermo Shandon, Pittsburgh, PA). Isotype matched mouse IgG instead of the primary antibody was used as a negative control and tissue positive controls are listed in Table 2. The signal was quantified using a semi quantitative method [16] by two examiners. At a glance the immuno-reactive (IR)-score quantifies staining intensity (1 = low, 2 = moderate, 3 = strong) and percentage of stained cells (0 = no, 1 = less than 10%, 2 = 10% - 50%, 3 = 51% - 80%, 4 = 81% - 100% stained cells). Multiplication of these values results in the IR-score ranging from 0 (negative) to 12 (strongly positive). Antibody staining used to calculate Spearman’s rho are either published [17,18] (LHCGR, FSHR, hCG) or are in press (PankoMabGEXTM).

Analysis of Gd serum levels
Sera were collected prior to surgery and stored at -80°C. Analysis was performed using a competitive enzyme immuno assay (EIA) principle. Briefly, plates were coated with antibodies (Table 2), which were rose against a peptide sequence localized in the C-terminal

| Stage | Patients (n) |
|-------|-------------|
| I     | 34          |
| II    | 10          |
| III   | 100         |
| IV    | 3           |
| n.a.  | 5           |
| Grade |             |
| G1    | 37          |
| G2    | 50          |
| G3    | 53          |
| n.a.  | 12          |
| deaths| 102         |
| cystic lesions | 21 |
| inflammatory diseases | 18 |
| benign tumors | 34 |
| malignant tumors | 38 |
| total | 111         |

n.a. = data on grade/FIGO stage were not available.
(GdQ13) or N-terminal (GdN20) end of the Gd protein core, over night at RT and blocked in blocking solu-
tion for 30 min. Following washes with PBS plates were incubated with biotinylated Gd standard peptides
(GdN20 peptid 6 ng/ml; GdQ13 peptid 1.5 ng/ml) for
1 hour followed by washes in PBS. Serum was diluted
1/2 in PBS and applied onto the plate for 1 hour to
perform the competition step. After washes with PBS
the plate was incubated with POD streptavidine for
2 min before the reaction was stopped with 1 N sulphuric
acid. The readout was performed using an ELISA Reader
Dynex MRX (Dynex Technologies, Chantilly, VA). Amni-
otic fluid or just blocking solution was used as positive
and negative controls, respectively.

Statistical analysis
Data were analysed employing the SPSS (v19, IBM,
Armonk, New York) statistic software for MS windows.
Statistical significance for pair wise comparisons of
unlinked non parametric values was determined by
Mann–Whitney-U test whereas differences among three
or more groups were tested using Kruskal-Wallis one-
way analysis of variance by ranks, respectively. Survival
and recurrence free survival was plotted in accordance
with Kaplan-Meier survival analysis. Correlations were
assessed using Spearman’s rho and gamma correlation
coefficient for two independent variables. Statistical sig-
nificance for all tests was assumed for p < 0.05 and data
are presented as mean ± standard error.

Results
Patients’ characteristics and Glycodelin expression
34 patients presented with early disease in stage I (FIGO I).
10 patients had FIGO stage II and 103 patients underwent
surgery because of suspected ovarian cancer involving the
peritoneal cavity (100 patients FIGO stage III and 3
patients stage IV). All patients who suffered from EOC
staged FIGO II - IV received carboplatin and paclitaxel as
chemotherapy after surgery. Histological 53 tumors were
assigned with WHO grade G3, 50 with G2 and another 37
were classified as G1. Mean follow-up time was 11.03 ±
0.58 years and mean overall survival 7.12 ± 0.63 years. 27
relapses and 102 deaths were documented.

Gd was detected by three different antibodies (Figure 1),
which recognize distinct peptide epitopes of Gd or tar-
get GdA specifically. Both peptide antibodies showed
strong to moderate staining (mean IRS GdQ13 = 5.35 ±
0.18; mean IRS GdC15 = 5.86 ± 0.20) which was equally
distributed within the four EOC subtypes examined
(Figure 1 B, C, D). However GdA staining in general
was much less intense (mean IRS = 1.31 ± 0.13) and
showed significantly different staining within subtypes
(p = 0.004) and highest expression (p = 0.007) in mucin-
osous (mean IRS = 3.33 ± 0.89) compared to serous, clear
cell and endometrioid carcinomas (Figure 1 A, D).
Interestingly, Gd staining (IRS > 0) determined by the
peptide specific antibodies (GdC15, GdQ13) was found
in 100% of the tissue sections examined, while GdA sig-
nal was just detected in 58.6%.

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Correlation of Glycodelin with tumor biomarkers, clinicopathological criteria and patients’ prognosis

For the peptide specific antisera GdC15 and GdQ13 a strong correlation in between the two (Table 3) was observed (p < 0.001). Expression of GdC15, GdQ13 and GdA epitopes rose with positivity for PankoMabGEX™ (GdC15, p = 0.007; GdQ13, p = 0.010; GdA, p = 0.004), which recognizes a special tumour-associated epitope on MUC1, TA-MUC1, that either localizes to the cell membrane (mPankoMabGEX™) or cytoplasm (cPankoMabGEX™). Furthermore GdA staining correlated with gonadotropin receptor (FSHR, p = 0.031; LHCGFR, p = 0.048) and hCG (p = 0.027) expression. Statistical analysis revealed no correlation of either GdA or GdQ13 antibodies staining with known prognostic markers in EOC, namely histological grade or FIGO stage. GdC15 was positively correlated to FIGO stage (p = 0.048).

Glycodelin A predicts prognosis in EOC patients

Whether Gd protein expression is useful to predict patients’ prognosis was analyzed by drawing Kaplan-Meier plots. Low expression - in respect of median expression IR-scores - was defined as IRS (GdA) ≤ 1; IRS (GdQ13) ≤ 4; IRS (GdC15) ≤ 4. In Kaplan-Meier analysis patients with GdA-positive advanced EOC (stage III and IV) had a significantly poorer overall-survival (log rank, p = 0.014, Fig. 2 A) and five-year survival (log rank, p = 0.009) than patients with GdA-negative tumors. In these patients with advanced stage ovarian cancer GdA positivity predicted significantly shorter recurrence free survival (log rank, p = 0.038) when compared to GdA negative tumors (Figure 2 B). In stage I and II no such difference could be shown. Further, GdA expression was analyzed in advanced staged patients whose survival after first diagnosis was longer than ten years (n = 7). Interestingly, none of

### Table 3: Correlations between GdC15, GdQ13 and GdA; Correlations between GdC15, GdQ13 and GdA were assessed using Spearman’s rho test for two independent variables

|          | GdA | GlycC15 | GlycQ13 | LHCGFR | FSHR | hCG | mPankoMabGEX | cPankoMabGEX |
|----------|-----|---------|---------|--------|------|-----|--------------|--------------|
| Spearman’s rho GdA | 1.000 | -0.142 | -0.054 | 0.162 | 0.178 | 0.187 | 0.045 | 0.241** |
| Sig. (2-tailed) | — | 0.081 | 0.508 | 0.048 | 0.031 | 0.027 | 0.596 | 0.040 |
| GlycC15 | Correlation Coefficient | -0.142 | 1.000 | 0.456** | -0.027 | 0.155 | -0.045 | 0.228** | -0.094 |
| Sig. (2-tailed) | 0.081 | <.001 | .740 | 0.061 | 0.598 | 0.007 | .270 |
| GlycQ13 | Correlation Coefficient | -0.054 | 0.456** | 1.000 | 0.118 | 0.080 | 0.047 | 0.217** | -0.128 |
| Sig. (2-tailed) | 0.508 | <.001 | — | 0.152 | 0.338 | 0.585 | 0.010 | .131 |

A correlation coefficient is presented and statistical significance for all tests was assumed for p < 0.05. * represents p < 0.05, ** represents p < 0.01.
these cases was found to be positive for GdA. Neither GdC15 nor GdQ13 were related to patients’ prognosis regarding overall survival or recurrence free survival, respectively.

**Serum Glycodelin concentrations**

Sera (n = 111) came from patients either presenting with benign (n = 73) or malignant (n = 38) ovarian diseases. Ovarian cancer patients mostly presented at stage III (FIGO I: 16.0%, FIGO II: 12.0%, FIGO III: 56.0% and FIGO IV: 16.0%). Benign neoplasias were further sub classified into simple ovarian cysts (n = 21), endometriosis and inflammatory diseases (n = 18) and benign ovarian tumors (n = 34). Serum Gd concentrations were determined by EIA using the two peptide antibodies GdN20 and GdQ13 (Figure 3 A, B) for targeting the N - and C-term of Gd, respectively. Gd was detected in 100% (GdN20) and 99.1% (GdQ13) of all sera. Significantly higher Gd concentrations (Figure 3 A, B) were found in sera collected from patients with benign ovarian tumors when compared to those with ovarian cancer (GdN20: p = 0.003, 10.84 ± 0.92 ng/ml vs. 15.81 ± 1.50 ng/ml and GdQ13: p = 0.001, 3.53 ± 0.40 ng/ml vs. 6.00 ± 0.56 ng/ml). GdQ13 (Figure 3 C) and GdN20 turned out to sensitively distinguish sera of patients with benign ovarian tumors with acceptable specificity (Figure 3 C; GdQ13: AUC = 0.729 ± 0.59, p = 0.001, 95% CI: 0.61 - 0.85; GdN20: AUC = 0.705 ± 0.06, p = 0.003, 95% CI: 0.58 - 0.83). No
significant differences in Gd serum concentration were observed between patients presenting with malignant ovarian cancer, inflammatory ovarian diseases or ovarian cysts.

Discussion

This study found that GdA independently predicts unfavorable prognosis in advanced stage ovarian cancer patients. Gd has already been published to correlate with less favorable clinical presentation in breast [19] and lung cancer [20], though controversial results were observed in ovarian cancer patients [21]. However a common problem in research on Gd is that most commercially available antisera were raised against peptide epitopes. Since gender specific glycosylation pattern is responsible for its tumor promoting activity. Therefore GdA might be an interesting marker to discriminate between high risk and low risk patients in advanced ovarian cancer (stage III and IV). A former study showed that Gd is also secreted to ascites fluid of ovarian cancer patients and that this Gd has significant differences in its structure of sialyl Lewis-type oligosaccharides compared to GdA. Additionally, ascites Gd inhibits IL-2 stimulated proliferation of peripheral blood leucocytes and inhibits adhesion of SLeX-positive cells to E-selectin. Therefore, Gd could act as an inhibitor of lymphocyte activation and/or adhesion in ovarian cancer [23].

As several studies proofed the immune suppressive effect of GdA [6,11], we suppose that GdA interferes with host anti tumor immune defense in EOC as well. Besides, it is widely accepted, that EOC has the ability to escape immune rejection by creating an immune suppressive environment [24]. Thus expression of the immunomodulatory GdA in ovarian carcinomas might contribute to permit immunological destruction of malignant cells and therefore might be related to tumor aggressiveness and poor clinical outcome.

Furthermore GdA staining correlated with gonadotropin receptor (FSHR and LHCGR) and with hCG expression itself (Table 3). There are only few studies on human chorionic gonadotropin and its receptor expression in ovarian cancer tissue [25,26]. In a recent study we found an opposed prognostic value of LHCGR and FSHR in ovarian cancer [17] in almost the same panel of patients as described within this study. GdA and hCG stimulate each other in a positive feedback mechanism in the placental decidual/trophoblast cell system as described earlier [27,28]. Results obtained within this study seem to confirm former results of GdA - hCG interaction. Gd expression showed also a positive correlation with a tumour-associated epitope of MUC1 (TA-MUC1), which was stained by the Pankoord GEXTM antibody. This epitope was found to be steroid hormone receptor dependent in breast cancer [29]. Because it is known that Gd expression is also regulated by steroid hormones [30], there could be a common regulation pathway for both Gd and TA-MUC1.

The use of Gd as a serum biomarker in ovarian cancer patients has been controversially discussed [31,32]. As Gd might interfere with tumor development, we decided to examine precursor lesions as well, which turned out to be characterized by elevated serum Gd levels. Interestingly in sera collected from ovarian cancer patients Gd levels were not significantly altered compared to cystic or inflammatory diseases. Benign ovarian tumors are part of a multistep process leading to low grade invasive ovarian cancer [33]. Gd mediated immune attenuation has been described during pregnancy [34-36] as well as in inflammatory processes [37]. Thus, besides of its pathophysiological role, Gd might be an interesting to detect benign or inflammatory changes in the ovaries, but the detection of GdA with its unique glycosylation may be a more suitable serum marker for ovarian cancer patients.

Conclusion

GdA is related to tumor aggressiveness and poor clinical outcome in advanced ovarian cancer. GdA might be an interesting marker to discriminate between high risk and low risk patients in stage III and IV ovarian cancer.

Competing interests

There is no financial or personal interest in relation to the work described.

Authors' contributions

CS, SH and ML have made substantial contributions to conception, design and acquisition of data. DM has made substantial contributions to analysis and interpretation of data, and has been involved in drafting the manuscript and revising it critically for important intellectual content. KF and UJ have given final approval of the version to be published. In addition, DM and UJ have made substantial contributions to conception and design of the study. All the authors have read and approved the manuscript for publication.

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