Research Article

Modelling pH-Optimized Degradation of Microgel-Functionalized Polyesters

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We establish a novel mathematical model to describe and analyze pH levels in the vicinity of poly(N-vinylcaprolactam-co-acetoacetoxyethyl methacrylate-co-N-vinylimidazole) (VCL/AAEM/VIm) microgel-functionalized polymers during biodegradation. Biodegradable polymers, especially aliphatic polyesters (polylactide/polyglycolide/polycaprolactone homo- and copolymers), have a large range of medical applications including delivery systems, scaffolds, or stents for the treatment of cardiovascular diseases. Most of those applications are limited by the inherent drop of pH level during the degradation process. The combination of polymers with VCL/AAEM/VIm-microgels, which aims at stabilizing pH levels, is innovative and requires new mathematical models for the prediction of pH level evaluation. The mathematical model consists of a diffusion-reaction PDE system for the degradation including reaction rate equations and diffusion of acidic degradation products into the vicinity. A system of algebraic equations is coupled to the degradation model in order to describe the buffering action of the microgel. The model is validated against the experimental pH-monitored biodegradation of microgel-functionalized polymer foils and is available for the design of microgel-functionalized polymer components.

1. Introduction

Cardiovascular diseases are the number one cause of death worldwide, globally claiming 17 million lives each year and accounting for 29% of all deaths [1]. Through the introduction of minimally invasive procedures like percutaneous coronary intervention (PCI) and the use of intravascular stents, the treatment of many cardiovascular diseases such as coronary artery disease, the obstruction of a coronary artery due to development of atherosclerosis, has been revolutionized [2, 3].

Nevertheless, efficacy and safety of available stents are limited in part as suitable autologous tissue engineered stents are lacking [4]. Mostly, nondegradable synthetic materials are substituted to repair the injured cardiovascular tissue. Meant to prevail at the implantation site, these foreign and therefore inherently thrombogenic materials can be associated with several risks, including calcification and acute or late stent thrombosis, which require prevention by antiplatelet therapy [5].

In contrast, tissue engineered bioresorbable stents provide temporary scaffolding for the formation of autologous tissue with the capacity to regenerate and grow. After the degradation process of the stent, only the restored vessel remains which might reduce the risk of late stent thrombosis.

The most frequently used resorbable polymers for biomedical implants including bioresorbable stents are the aliphatic polyesters polylactic acid (PLA) and polyglycolic
acid (PGA) and their copolymers. The mechanism of poly-
ester degradation has been well investigated. Polymer chains are
degraded by hydrolytic scission of ester linkages in the
polymer backbone and thereby create carboxylic end groups.
These acidic carboxyl end groups diffuse into the vicinity of
the polymer and decrease the pHe level there [6–10].

This pHe drop is one of the main drawbacks of using
aliphatic polyesters for implants as a low pHe may cause tissue
reactions like inflammation [11–13].

Still, biodegradable stents have marked potential long-
term advantages [4, 14]. In order to overcome the disadvan-
tagewoacidicpHelevelsduringdegradationodbiodegradable
stents, we investigate the fabrication of polyactic acid (PLA)
fibers for cardiovascular stents with pHe-optimized degrada-
tion behavior using VCL/AAEM/VIm (8 mol%)-microgels as
insoluble buffers [15].

Microgels are defined as hydrogel particles with a dimen-
sion from 10 nanometers up to the micrometer range. Hydro-
gels in general are novel components that arouse special
interest in the biomedical field and nanotechnology [16, 17].

Depending on the molecular building units and the syn-
thesis procedure, it is possible to synthesize colloidal hydro-
gels with controlled particle size. Those can be defined as
polymer networks with smart properties such as water-
uptake capacity, degree of swelling, and responsiveness to
external stimuli. Furthermore, these nanoparticles show high
 colloidal stability, a well-defined structure, and a high sur-
face area. Through variation of the monomer type and/or
introduction of comonomers, responsiveness to temperature,
pHe, magnetic field, or light intensity can be tailored. This
property, in combination with the swelling features, makes
the utilization of microgels as biocompatible materials for
pharmaceutical applications possible [16, 18–23].

Monomers like N-isopropylacrylamide (NIPAAm) and
N-vinylcaprolactam (VCL) have been used for the synthesis
of thermoresponsive microgels because they can form water-
soluble polymers with lower critical solution temperature
(LCST) [18, 20, 24, 25]. Other interesting components such as
N-vinylimidazole (VIm) can be used to achieve pHe-
sensitivity due to its protonation/deprotonation process at
different pHe levels. Pich et al. [26] have successfully synthe-
sized VCL/VIm-based microgels, which show both tempera-
ture and pHe-responsiveness. In this study, the swelling could
be controlled by the VIm content in the microgel.

However, cardiovascular stents require for the degrada-
tion period to last over months, making their experimental
research as well as their design and optimization rather
time-intensive. This study aims at developing a mathematical
model able to predict the pHe level resulting from polymer
degradation and taking into account the buffering action of
eventually incorporated microgels. This mathematical model
will be validated against the experimental pHe-monitored
biodegradation of microgel-functionalized polymer foils and
will be available for the design of microgel-functionalized
polymer stents and other components.

Computational methods to model the degradation of
biodegradable polymers have been proposed by a quantity
of research teams. Siparsky et al. [27] described the kinetics
of hydrolysis reaction by rate equations and their analytic
solutions. Han [28] used Monte Carlo methods to predict
the degradation of PLA, PGA, and some copolymers. He
also investigated the influence of chemical buffers on the
degradation process and the pHe level. Wang et al. [29]
presented a reduced model for polymer degradation using
two rate equations for the concentrations of carboxylic end
groups and ester bonds in the PLA phase. Pan et al. [30]
cluded a model for chemical buffers in such a reduced
model for biodegradation. Moreover, rate equations for the
concentrations of different molecules sizes, which the poly-
mer undergoes in its degradation process, were built up and
analyzed by Lazzari et al. [31]. The buffering action of the
poly(N-vinylimidazole) hydrogel was modelled and analyzed
by Horta et al. [32, 33]. To the best of the authors’ knowledge,
so far no published model for the biodegradation of polymers
buffered with VCL/AAEM/VIm-microgels exists.

Investigating microgel-functionalized resorbable poly-
mers with pHe-regulatory potential, a recent study of Fehe´r
et al. [15] employs a publication-based keyword search strategy
to investigate the existing knowledge base underpinning the
topic. The authors find that only a marginal number of
studies is dedicated to investigate the pHe-regulative potential
of microgel containing degradable polymers. This clearly
demonstrates an underinvestigated topic. Given the poten-
tial scope of application, ranging from drug-coated, pHe-
regulating degradable textiles to stents, this paper is aligned
to the apparent research gap.

2. Materials and Methods

2.1. VCL/AAEM/VIm (8 mol%)-Microgel. The synthesis of
VCL/AAEM/VIm (8%)-microgel is well described in Fehe´r
et al. [15]. The description is not repeated here.

2.2. Polymer VCL/AAEM/VIm (8 mol%)-Microgel Foils.
Poly(L-lactide-co-glycolid) PLG 8523 by Purac and the
microgel are weighed separately. Both samples are then
filled up with a solvent in a way that the mass fraction of
polymer wpla and microgel wmic amount to 0.15, respectively.
The sealed samples are stirred at 250 rpm for 24 hours in
a magnetic stirrer. The resulting microgel and polymer
solutions are mixed thoroughly to ensure a homogeneous
solution. The solutions are evenly spread on a petri dish and
thereby infused to a foil. After 48 hours within the distractor
hood, the solvent is fully evaporated, allowing for the foil to
be peeled from the glass.

2.3. Degradation Experiment. The degradation experiments
are carried out according to ISO 13781 [34]. Each sample
contains 0.1 g of the produced foil and is supplemented with
distilled water (specific conductivity < 0.1 µS/cm at 25°C)
at the ratio of 1:100. The samples are prepared in 20 mL
glass containers and sealed during degradation periods. The
containers are only opened for pHe measurement.

Throughout the degradation process, the samples are
stored in an oven at 37°C. pHe measurement is conducted
daily within the first 20 days followed by weekly readings
for a total time period of at least 30 days. To determine
the pH level, the samples are withdrawn from the oven. Due to the high temperature sensitivity of the pH level, the samples are cooled down to ambient temperature (20°C) prior to the measurement. Each measurement is repeated twice and the three results are averaged. To increase the experiment’s significance, each sample is prepared twice and the three results are averaged. To increase the experiment’s significance, each sample is prepared twice and the three results are averaged.

2.4. Physical Model Reduction and Mathematical Modelling.
In order to analyze the pH level in a vicinity of a degrading polyester component, we predict the concentration of hydron [H⁺] resulting from dissociation of carboxyl groups of the degrading polymer. As the dissociation of the carboxyl groups COOH to COO⁻ and hydron H⁺ in aqueous solution takes place on a very short timescale, we make use of the equilibrium constant $K_{COOH} = [COO⁻][H⁺]/[COOH]$ for the dissociation reaction and insert it into the balance of mass and charge to calculate the concentration of hydron [H⁺]:

$$[H^+] = \frac{K_w}{[H^+]} + \frac{K_{COOH} [M]}{[H^+]} + K_{COOH}$$  \hspace{1cm} (1)

with the equilibrium constant for the dissociation of H₂O to OH⁻ and H⁺: $K_w = [H^+][OH^-]$. The brackets [X] mark the concentration of the corresponding species X. We used the fact that the concentration of oligomers [M] corresponds to the concentration of carboxyl groups [COOH]. The concentration of H₂O is not part of the equation as we assume it to be constant because the water penetration into the polymer is on a very short timescale.

The concentration of carboxyl groups increases with the progressing scission of polymer chains by hydrolysis reaction and is thus time dependent. Hydrolysis of polymers proceeds partly as spontaneous reaction and partly catalyzed by hydron H⁺. The reaction is called autocatalytic as the reaction products hydroxyl alcohol and carboxylic acid end groups—also referred to as oligomers [28]—dissociate and acidize the environment and as a result accelerate the rate of hydrolysis [27]. The hydrolysis reaction is explained in detail elsewhere [35].

We implement a phenomenological model for the degradation of biodegradable polymers initially introduced by Wang et al. [29]. As a result, we predict the evolution in time of oligomer concentration $[M]_{init}$ after degradation and before buffering as well as the evolution in time of concentration of ester bonds [E] inside the calculation area by two rate equations:

$$\frac{d[E]}{dt} = -k [E] - k_{cat} [E] [H^+]$$  \hspace{1cm} (2)

$$\frac{d[M]_{init}}{dt} = -\frac{d[E]}{dt}$$  \hspace{1cm} (3)

with the rate coefficients k and $k_{cat}$ for spontaneous and catalytic scission of ester bonds inside the polymer, respectively. Using the equilibrium constant $K_{COOH}$, we transform (2) to

$$\frac{d[E]}{dt} = -k [E] - k_{cat} \sqrt{K_{COOH}} [E] \sqrt{[M]}.$$  \hspace{1cm} (4)

[M] refers to the concentration of oligomers available for the autocatalytic scission.

After dissociation of oligomers, the hydrons are buffered by the VIm groups of the microgel. Microgel and polymer form a two-phase system. The hydrons diffuse inside the microgel and are bound to the VIm. The dissociation products COO⁻ bound to the oligomers will also diffuse inside the microgel and remain there undoubt due to electrostatic forces. The diffusion processes will continue until the thermodynamic equilibrium between the two phases is reached. Using the assumption that the buffer reaction is fast compared to the degradation processes, we use the equilibrium constant of the buffer reaction $K_p = [P][H^+] / [PH^-]$ with the concentration of free buffer molecules [P] and occupied buffer molecules [PH⁻], respectively, and find a system of algebraic equations describing the buffering action of the microgel. The derivation is explained in detail for poly(N-vinylimidazole) by Horta and Piérola [33]:

$$[COO^-]_{gel} = \frac{[H^+]_{gel}^3 + [H^+]_{gel}^2 (K_p + [P]_{init}) - [H^+]_{gel} \cdot K_w - K_p \cdot K_{cooh}}{[H^+]_{gel} \cdot ([H^+]_{gel} + K_p)}$$

$$[COO^-]_{gel} = -\left(\frac{[H^+]_{init} - K_w}{[H^+]_{init}}\right) \cdot \frac{V_{bath}}{V_{gel}} \cdot \left(\frac{[H^+]_{init} - \frac{K_w}{[H^+]_{init}}}{[H^+]_{gel} + K_{cooh}}\right)$$

$$[H^+] = \sqrt{[H^+]_{gel} [COO^-]_{gel} + K_{cooh}}$$

where $[X]_{gel}$ represents the concentration of the species X in the microgel phase, $[X]_{init}$ represents the concentration in the polymer before the buffering reaction, $V_{init/gel/bath}$ are the volumes occupied by the polymer without the gel, the volume of the microgel, and the volume of the polymer including the microgels, respectively, and $[P]_{init}$ is the initial concentration of buffer molecules in the system. Using (1), we calculate from $[H^+]$ the concentration of oligomers $[M]_{bath}$ after the buffering action of the microgel and complete the ODE system ((2) + (3)) by a rate equation for the concentration of oligomers $[M]$ after buffering which allows for diffusion of the oligomers:

$$\frac{d[M]}{dt} = \frac{[M]_{bath}}{[M]_{init}} \left(\frac{d[E]}{dt} + D_0 \Delta [M]\right)$$  \hspace{1cm} (6)

with $D_0$ being a constant diffusivity of oligomers.

The calculation area and initial and boundary conditions are chosen appropriate for the degradation experiment...
Table 1: Parameters used for simulation.

| Symbol     | Value       | Unit       | Description                                                                 | Source |
|------------|-------------|------------|------------------------------------------------------------------------------|--------|
| $K_{\text{COOH}}$ | $10^{-3.87}$ | mol/L      | Equilibrium constant of dissociation reaction of COOH at 37°C                | [28]   |
| $K_w$      | $1.8 \cdot 10^{-14}$ | (mol/L)$^2$ | Equilibrium constant of dissociation reaction of H$_2$O at 37°C             | [36]   |
| $k$        | 0.005       | 1/week     | Rate constant of spontaneous ester bond scission                             |        |
| $k_{\text{cat}} \cdot \sqrt{K_{\text{COOH}}}$ | $0.5 \cdot 10^{-5}$ | m$^2$/mol/week | Rate constant for autocatalyzed ester bond scission                          |        |
| $K_p$      | $1 \cdot 10^{-10}$ | mol/L      | Equilibrium constant for buffer reaction at 20°C                             |        |
| $[P]_0$    | 0.3487      | mol/L      | Initial buffer concentration                                                 |        |
| $V_{\text{gel}}/V_{\text{init}}$ | $9.42 \cdot 10^{-6}$ | — | Fraction of volumes occupied by gel and polymer                            | Derived from experimental procedure |
| $V_{\text{bath}}/V_{\text{init}}$ | $1 - V_{\text{gel}}/V_{\text{init}}$ | — | Fraction of volumes occupied by polymer + gel and polymer                   | —      |
| $D_{\text{0,p}}$ | $2 \cdot 10^{-9}$ | m$^2$/week | Diffusion constant of oligomer in the polymer                               | [28]   |
| $D_{\text{0,w}}$ | $2 \cdot 10^{-8}$ | m$^2$/week | Diffusion constant of oligomer in water                                      | [37]   |
| $d_p$      | 0.15        | mm         | Thickness of polymer foil                                                    | Derived from experimental procedure |
| $d_w$      | 19.6        | mm         | Thickness of water column above polymer foil                                 | Derived from experimental procedure |

As described above, a consequence of the typical extensions of a polymer foil, diffusion will be dominant in one dimension. Thus, we set up a 1D diffusion model and use $x$ as the coordinate in space. The calculation area consists of two components: the (microgel containing) polymer with the thickness $d_p$ and the surrounding water with the thickness $d_w$. At time $t = 0$, ester bonds have the initial concentration $\left[E\right]_0$ in the polymer foil ($0 \leq x \leq d_p$) and 0 in the water volume ($x > d_p$) and the initial concentrations of oligomers $\left[M\right]_{\text{init}}$ and $\left[M\right]$ are 0 everywhere:

$$\left[E\right](t = 0)|_{0 \leq x \leq d_p} = \left[E\right]_0,$$

$$\left[E\right](t = 0)|_{x > d_p} = 0,$$

$$\left[M\right](t = 0) = \left[M\right]_{\text{init}}(t = 0) = 0.$$

The boundaries at $x = 0$ and $x = d_w$ are isolated and do not allow for diffusion of oligomers. At the boundary $x = d_p$, continuity of oligomers is required:

$$\frac{d\left[M\right]}{dx} \bigg|_{x=0} = \frac{d\left[M\right]}{dx} \bigg|_{x=d_p} = 0,$$

$$\left[M\right](x = d_p) = \left[M\right](x = d_w).$$

A schematic representation of experimental set-up and the calculation area is depicted in Figure 1.

![Diagram of experimental set-up and calculation area](image-url)

The diffusivity of oligomers $D_{\text{0,p}}$ and $D_{\text{0,w}}$ is considered to be constant in the polymer and the water layer, respectively. In order to simulate the swiveling of the samples in the experimental procedure, we increase the diffusion constant $D_{\text{0,w}}$ in (6) by a factor of $10^3$ for a very short period on every simulated measurement event.

The parameters which have been used to carry out the simulation experiments, discussed in the following chapter, are listed in Table 1.

### 3. Results and Discussion

#### 3.1. Buffering Action of VCL/AAEM/VIm-Microgels

As the buffering model (5) was originally applied to a poly(N-vinylimidazole) gel [33], we investigate its appropriateness for the VCL/AAEM/VIm-microgels in a preliminary study.
Therefore, we adjust hydrochloric acid to different initial pH levels $pH_{init}$, add 0.1 g of VCL/AAEM/VIm-microgel, and measure the pH level of the common bath $pH_{bath}$. We calculate the resulting pH levels $pH_{bath}$ using (5) and the same initial pH levels as in the experiment. The simulation result in comparison to the experiment is plotted in Figure 2. We consider the unknown equilibrium constant for the buffer reaction $K_p$ and the amount of VIm molecules per microgel unit $\sigma$ as fitting parameters and use the withdrawn values of $K_p = 1 \cdot 10^{-10}$ mol/L and $\sigma = 1,522 \cdot 10^{20}$ 1/g for further simulation. The preliminary study shows a threshold behavior for the buffering effect. Initial pH levels above 4 ($pH_{init} \geq 4$) are buffered by VCL/AAEM/VIm-microgel to a neutral pH level. For initial pH levels below this value, the buffering action of VCL/AAEM/VIm-microgel in the used concentration is not sufficient to receive a neutral pH level.

Furthermore, we analyze the amount of VCL/AAEM/VIm-microgel necessary for successful buffering action to a neutral pH level. The result is shown in Figure 3. Initial pH levels of 3.75 and higher can be buffered to neutral level by a total amount of 0.005 g microgel (solid black line in Figure 3). This corresponds to a microgel mass fraction $w_{MG}$ of 0.05 in the polymer foils (for preparation, see above). A threshold below which very strong decay of pH level arises is seen at $pH_{init} = 1.78$ for the 0.005 g VCL/AAEM/VIm-microgel. Lower VCL/AAEM/VIm-microgel amounts (non-solid and gray lines in Figure 3) lead to higher threshold pH levels. Thus, the curves in Figure 3 are shifted to the right with decreasing VCL/AAEM/VIm-microgel amount. The threshold behavior is less distinct for lower microgel amounts. Hence, the curves in Figure 3 are also flattened with decreasing VCL/AAEM/VIm-microgel amount. Nearly no buffering action is predicted for a mass of $1 \cdot 10^{-6}$ g VCL/AAEM/VIm-microgel.

This analysis is valid for the ideal case when considering a homogenous VCL/AAEM/VIm-microgel distribution inside the polymer, where diffusion of oligomers to reach the VIm-molecules is not necessary and no barriers, for example, electrostatic reasons, occur.

### 3.2. Degradation Studies

The experimental results from the degradation studies of polymer foils with VCLVCL/AAEM/VIm-microgel mass fractions of $w_{MG} = 0.00$ and $w_{MG} = 0.05$, respectively, are depicted in Figure 4 via the mean and the standard deviation of the two measurement series (see above). Without microgel, a drop in pH level arises as expected leading to $pH = 5.4$ after 35 days. As the pH level is measured in the water component of the experimental setup ($d_p \leq x \leq d_w$ in Figure 1), the pH level in the core of the polymer foil ($x = 0$) will be lower. This lowest pH level is relevant to choosing the appropriate VCL/AAEM/VIm-microgel content.

We use the experimental results of polymer foils with a microgel mass fraction of $w_{MG} = 0.00$ to determine the rate constants in the degradation model (2) in order to find a good agreement between simulation and experimental measurements. The simulation result as average pH level in the water component ($d_p \leq x \leq d_w$ in Figure 1) is plotted as a solid line in Figure 4. The influence of the simulated swiveling process can be seen clearly as steps in the pH level curve. The obtained degradation rates as all other model parameters are listed in Table I.

The pH level in the center of the polymer foil ($x = 0$ in Figure 1) is evaluated and plotted over time in Figure 5. The pH level drops to 1.42 after 35 days using a mass fraction of $w_{MG} = 0.00$ (solid line). Applying a VCL/AAEM/VIm-microgel buffer with a mass fraction of $w_{MG} = 0.05$ to the resulting solution ($pH_{init} = 1.42$) will not be sufficient to buffer the pH to a neutral level but only to a pH level of approximately $pH_{bath} = 2$ according to Figure 3. Nevertheless, in the center of the foil after 35 days only a small decrease

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**Figure 2:** pH level ($pH_{bath}$) of different initial concentrations of hydrochloric acid ($pH_{init}$) after buffering with 0.1 g microgel.

**Figure 3:** Predicted pH-value ($pH_{bath}$) after buffering different initial pH levels ($pH_{init}$) with different constant microgel amount.
the reduction of overall degradation rate by incorporation of VCL/AAEM/VIm-microgel. Thus, a VCL/AAEM/VIm-microgel mass fraction of \( w_{\text{MG}} = 0.05 \) will possibly be sufficient to buffer the pH to a neutral level. Figure 4 confirms this expectation in theory and experiment. The predicted decrease of pH level in the water component is smaller than in the center of the foil and is in good accordance with the measurements.

In order to briefly demonstrate the influence of the experimental procedure on the development of pH level, three different swiveling procedures are simulated and the resulting average pH levels in the water phase are plotted in Figure 7 (no swiveling: dashed line, swiveling at discrete measurement events: solid line, and continuous swiveling: dotted line). The strong deviation in simulated pH levels shows the importance of standardized experimental procedures.

3.3. Limitations of the Study. The rate equation system for degradation of polymers ((1)–(3) and (6)) is a very reduced model. Any difference between scission of ester bonds at molecule ends and that of bonds in the middle of the polymer molecules is neglected as well as the effects of crystallinity, of diffusivity varying in time, and others. Nevertheless, a very similar model is already considered very useful for the prediction of size and shape effects on biodegradation [29]. One has to perform reference experiments without incorporation of VCL/AAEM/VIm-microgels to determine the rate coefficients of the presented degradation model. With these references our model can be used for investigating the pH levels in the vicinity of polymer components and for analyzing buffering behavior. The reference experiments will have different results using different polymers and therefore have to be performed for every material that is investigated.
The simulated model task consisting of a 1D calculation area is a simplified task and deviations from experimental measurements are expectable. It is adequate for a first benchmark of the model with experiments and appropriate for model development as very short computation time results from the simple task.

Model refinements and extensions are necessary in order to investigate the influence of microgel distribution inside the polymer and buffering reaction kinetics. For this purpose, new specific experiments have to be conducted.

We study the degradation behavior of polymer foils in a preclinical set-up according to ISO 13781. Any biological influences like enzymes, vascularization, buffering in body fluids, or patient individual factors cannot be investigated in degradation experiments of this kind. As can be seen from simulation results (Figure 7), conditions (e.g., flow of liquid medium) during degradation experiments have an important influence on the degradation results. Thus, the authors suggest building up a more complex test bench for degradation experiments simulating blood flow amongst others before beginning with clinical studies.

4. Conclusions

VCL/AAEM/VIm-microgel-functionalized fibers with pH-optimized degradation behavior are a promising approach for a wide range of medical applications. In particular, the treatment of many cardiovascular diseases such as coronary artery disease will benefit from biodegradable material for stents without the drawback of acidic pH levels as a consequence of degradation.

Within this study, a mathematical model is presented, which can deal as an important tool to design components with pH-optimized degradation behavior. Iterative design of suitable degradation behavior based on mathematical modelling is shown to exploit the potentials of the medical application and will save engineering costs and time for degradable polymer devices and can contribute to reducing animal studies.

The mathematical model consists of a reduced degradation model based on a rate equation system including diffusion of acidic degradation products into the vicinity of the component. This degradation model is coupled with a set of algebraic equations modelling the buffering action of VCL/AAEM/VIm-microgel. Both models (rate equations for degradation and algebraic buffering model) are evaluated separately in comparison to experimental measurements of pH level and show good agreement with them.

The pH level during the degradation of polymer foils is monitored during 35 days and serves as reference to fit the rate constants of the mathematical degradation model. Additionally, initial pH levels are buffered with a constant amount of VCL/AAEM/VIm-microgel to calibrate the model for the buffer reaction.

A VCL/AAEM/VIm-microgel mass fraction of \( w_{MG} = 0.05 \) is found to deliver a sufficient buffering action within at least 35 days of degradation of polymer foils. Simulation as well as experimental studies confirmed this buffering effect.

Competing Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

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