Article

Browning of Early and Late-Harvested ‘Empire’ Apples Affected by Cold Storage and 1-MCP

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Abstract: The effects of harvest time, 1-methylcyclopropene (1-MCP) and air storage time on the susceptibility of flesh browning in ‘Empire’ apples were studied during a seven-day shelf life period after air storage at 0.5 °C for seven months. Early- or late-harvested ‘Empire’ apples without 1-MCP increased production of ethylene, respiration rates and internal ethylene concentration during the shelf life. Respiration rates increased in the late-harvested fruit for the shelf life period with/without 1-MCP treatment. The 1-MCP-treated fruit was approximately 10 N firmer than fruit not treated with 1-MCP at an early harvest for the entire shelf life duration. Peroxidase activity and percent change in electrical conductivity in the flesh were elevated in late-harvested fruit for the shelf life duration, whereas polyphenol oxidase activities were found to be stimulated by 1-MCP treatment, regardless of harvest time. Late-harvested fruit treated with 1-MCP exhibited increased susceptibility to flesh browning during shelf life, mostly due to reduction of the antioxidant defense mechanism of the fruits to stress in extending storage life, increasing polyphenol oxidase (PPO) activity and electrolyte leakage rate.

Keywords: Malus × domestica; 1-MCP; browning disorder; polyphenol oxidase; peroxidase

1. Introduction

Internal flesh browning of ‘Empire’ apples (Malus sylvestris (L.) Mill Var. domestica Borkh.) is a physiological disorder known to occur intermittently during long-term storage that considerably reduces the postharvest quality of the fruit and renders it unacceptable in the market [1–5]. Biochemical browning in apples mostly results from the oxidation of phenolic compounds to o-quinones by polyphenol oxidase (PPO) leading to brown discoloration and synergistically reacting with peroxidase (POX) activity [5–8]. The oxidation of phenolic compounds in apple flesh is dependent upon numerous factors including a fruit’s nutritional status, flesh structure, the geographic location of orchards, irrigation systems, seasonal variability, tree age, harvest time, 1-MCP (1-methylcyclopropene) treatment, CO₂ and O₂ levels, storage temperature and duration and adoption or non-adoption of CA (controlled atmosphere) storage with low temperatures [9,10]. Therefore, understanding the relationship between phenolic enzyme activity and the factors affecting phenolic oxidation in ‘Empire’ apples during storage is important to control the risk of disorder development.

1-MCP is extensively used in the agricultural industry to maintain storage potential and quality in horticultural crops as an ethylene action inhibitor during long-term storage [11,12]. The benefit of 1-MCP for apples is extended shelf life of the fruits, while a disadvantage is unpredictable fruit browning in long-term storage. The 1-MCP and dynamic CA applications promoted internal browning in ‘Elstar’ apples due to CO₂ injury caused by high storage temperature [13]. PPO (peroxidase) or POX (polyphenol oxidase) antioxidants in controlled atmosphere-stored ‘Empire’ apples increased with 1-MCP treatment likely due to extended duration of fruit sensitivity to stress and lower energy.
levels, inducing the development of flesh browning [1,4,14]. There is little information available to date regarding the susceptibility of apples air-stored conventionally in developing countries. ‘Empire’ is particularly susceptible to browning on exposure to low O\textsubscript{2} and high CO\textsubscript{2} levels in the atmosphere [1,2,4,5], but the fruit’s resistance to browning in air and CA is largely dependent upon harvest time [3].

A late harvest increased incidences of flesh browning in CA-stored ‘Braeburn’ apples [9,15,16] and air-stored ‘Macoun’ apples [17] due to an observed increase in respiration rates, skin resistance to gas transport and internal CO\textsubscript{2} concentrations, causing CO\textsubscript{2} injury and low O\textsubscript{2} damage, which was not consistently observed for a core browning in air-stored ‘Empire’ apples [3]. Flesh browning in ‘Empire’ apples at early and late harvest does not develop until after six months of air-storage and its effect on shelf life has not been evaluated with regards to distribution time. Late-harvested ‘McIntosh’ apples with multiple 1-MCP treatments showed increased incidences of browning, which did not similarly occur on the treated ‘Empire’ fruit [18]. No information is yet available about the interaction between 1-MCP and storage time at harvest time. The aim of this research was to investigate the effects of harvest times, treatment of fruits with 1-MCP and length of time in air storage, all of which are the primary concerns of fruit storage management, on the susceptibility of flesh browning in ‘Empire’ apples exposed to room temperature after long-term cold storage.

2. Materials and Methods

2.1. Plant Material and Sampling

‘Empire’ apples (Malus sylvestris (L.) Mill Var. domestica Borkh.) at early and late stages of maturity were, respectively harvested once from mature trees grown at the Cornell University orchard located at Ithaca, USA. Early stages of fruit maturity indicated internal ethylene concentrations of less than 30 \mu\textsubscript{L} L\textsuperscript{−1} or greater for later stages of maturity, with a one-month interval between both maturities. ‘Empire’ apples from early and late stages of maturity were harvested from different locations on the trees in a season. Half of the sample of 600 ‘Empire’ apples were treated with or without 1 \mu\textsubscript{L} L\textsuperscript{−1} of 1-MCP after 1 d of overnight cooling. The 1-MCP treatments were applied following the method of Watkins et al. [19] using an activator solution added with SmartFresh tablets (AgroFresh, Philadelphia, PA, USA) released into a 4000 L plastic tent. After venting for 3 h and overnight cooling, 600-fruit were stored at low temperature (0.5 °C).

At each sampling point, five individual fruits from each treatment were sampled to analyze ethylene production, respiration rates, internal ethylene concentration (IEC) and flesh firmness at 0, 1, 3, 5 and 7 days shelf life after the 7-month storage period. After measuring IEC of each fruit, fruit flesh tissue was immediately put into liquid nitrogen and then stored at −80 °C to analyze peroxidase (PPO), polyphenol oxidase (PPO) and total phenolic concentrations.

Incidence of the disorder was visually assessed by the extent of flesh browning at each sampling point, expressed on a scale of 0 = 0% (no browning), 1 = 1–10% (slight browning), 2 = 11–25%, 3 = 26–75% and 4 = 76–100% (almost complete browning) from the cut surface of the flesh tissue.

2.2. Ethylene Production and Respiration

Each fruit was sealed in a 1 L plastic container for 1 h to analyze ethylene production, fruit respiration rate and IEC (internal ethylene concentration). One mL of gas from the plastic containers was used to measure the concentration of ethylene and IEC, using a Hewlett-Packard 5890 series II gas chromatograph (Hewlett-Packard Co., Wilmington, NC, USA) equipped with a flame ionization detector and an activated alumina SS column (Supelco, Philadelphia, PA, USA). Respiration rates were also measured by a Varian 3499 gas chromatograph (Series 580; Varian, Bridgewater, MA, USA) using gas from each sample.
Flesh firmness was measured on the opposite poles of each fruit using an EPT-1 pressure tester (Lake City Technical products, Lake City, AB, Canada) with an 11-mm-diameter probe, after the skin was thinly removed.

2.3. Enzyme Extraction and Assays

For each enzyme extraction, 3 g of flesh tissue sample was ground to fine powder in liquid nitrogen and put into 9 mL of extraction 200 mM phosphate buffer (pH 7.8) containing 2 mM of ethylenediaminetetraacetic acid (EDTA), 5% polyvinylpolypyrrolidone (PVPP) and 1 mM phenylmethanesulfonyl fluoride (PMSF). The mixture was then homogenized and centrifuged at 14,000×g for 30 min at 4 °C. The supernatant was divided into aliquots and kept at −80 °C to assay the enzymes and proteins.

POX and PPO activities were spectrophotometrically measured following the methods of Kochhar et al. [20] and Zhou et al. [21], respectively. In order to measure the activities of POX and PPO, 200 µL of extraction was added to each 2.8 mL assay solution, separately. The assay solution of POX was a 100 mM phosphate buffer (pH 6.8) containing 2.7 mM guaiacol and 4 mM H$_2$O$_2$. The assay solution of PPO was a 100 mM citrate–200 mM phosphate buffer (pH 5.0) containing 50 mM catechol. The activities of POX and PPO were assessed through changes in optical density (OD) at 470 and 420 nm mg$^{-1}$ of protein, respectively using a spectrophotometer (Beckman-Counter, DU 7400, Fullerton, CA, USA) [21]. Protein concentration was determined via the method of Bradford (1976) using bovine serum albumin as standard [22].

2.4. Total Phenolic Concentration

Ten g of frozen tissue from each fruit was extracted with 80% acetone to determine total phenolic concentrations, using a modified Folin–Ciocâlteu colorimetric method [4,23]. The absorbance was measured with a spectrophotometer (Beckman-Counter, DU 7400, Fullerton, CA, USA) and expressed as gallic acid equivalents based on fresh weight.

2.5. Leakage Assessment

Electric leakage was determined by measuring mesocarp tissue samples from apples stored at 0.5 °C in air storage for up to 7 months and evaluated for 7 d at 20 °C, which were then cut into discs with a cork borer (2 mm-thick and 12 mm in diameter) using a modified method by Barrett et al. [24]. Discs were washed with distilled water 3 times, incubated in 0.4 M mannitol for 30 min at 25 °C, autoclaved for 30 min at 121 °C and cooled to 25 °C. The leakage was then measured using a conductivity meter (Chemtrix Type 700, Hillsboro, OR, USA) after 30 min and calculated from the presence of total electrolytes.

2.6. Statistical Analysis

All data were subjected to ANOVA using Minitab software v. 15.1 (Minitab, Inc., State College, PA, USA). Means were separated using least significant differences (LSD) at $p = 0.05$. Data over time are shown as means ± standard errors.

3. Results

Ethylene production in the early harvested ‘Empire’ apples not treated with 1-MCP gradually increased over 4 d of shelf life at 20 °C after 7-months of cold storage and started to decrease from 5 days to 7 days, with very low levels observed for 1 MCP-treated fruit (Figure 1A). Initial amounts of ethylene produced in the late-harvested fruit was 3.3 ng kg s$^{-1}$ and 0.8 ng kg s$^{-1}$ for untreated- and 1-MCP-treated fruit, respectively, with continuously low levels observed for the 6 d of shelf life compared to the early harvested fruit (Figure 1B). The 1 MCP treatment resulted in lower respiration
rates in the early harvested fruit throughout a 7 days shelf-life (Figure 2A), but this effect was only slightly observed in late-harvested fruit. (Figure 2B).

**Figure 1.** Ethylene production of early (A) and late-harvested ‘Empire’ apples (B) and stored at 0.5 °C in air storage up to 7 months and evaluated for 7 d at 20 °C. Bars represent error of the means (SEM; n = 5), when larger than the dimension of the symbol. *, **, *** Adjacent to each data point for each storage time indicates significant differences as determined by Duncan’s multiple range test at p ≤ 0.05, 0.01 or 0.001, respectively. MCP-methylcyclopropene.

**Figure 2.** Respiration rate of early (A) and late-harvested ‘Empire’ apples (B) and stored at 0.5 °C in air storage up to 7 months and evaluated for 7 days at 20 °C. Bars represent error of the means (SEM; n = 5), when larger than the dimension of the symbol. *** Adjacent to each data point for each storage time indicates significant differences as determined by Duncan’s multiple range test at p ≤ 0.001; ns—not significantly different.

IEC (internal ethylene concentration) in the flesh ranged between 100–130 μL L⁻¹ for non-treated apples and less than 1.0 μL L⁻¹ for 1-MCP-treated apples at an early harvest until the end of a 7 days shelf life period after 7 months of cold storage (Figure 3A). The highest IEC of 192.1 μL L⁻¹ was observed at a 5 days shelf life for non-treated fruit, with 42.5 μL L⁻¹ at 7 days for 1-MCP-treated fruit at a late harvest. The 1-MCP treatment increased approximately 10 N above average firmness at an early harvest during shelf life compared to the non-treated fruit but did not affect the firmness retention in a late harvest (Figure 3C,D).
Figure 3. Internal ethylene concentration (IEC) (A, B) and flesh firmness (C, D) of early and late-harvested ‘Empire’ apples stored at 0.5 °C in air storage up to 7 months and evaluated for 7 days. Bars represent error of the means (SEM; n = 5), when larger than the dimension of the symbol. *, *** Adjacent to each data point for each storage time indicates significant differences as determined by Duncan’s multiple range test at \( p \leq 0.05 \) and 0.001, respectively; ns—not significantly different.

POX activities in the flesh were not affected by 1-MCP treatment at an early (\( p = 0.757; \) Figure 4A) and late harvest (\( p = 0.117; \) Figure 4B) but fluctuated at a late harvest. The 1-MCP treatment increased the flesh PPO activities at an early harvest (\( p = 0.001 \)) and increased the flesh PPO activity in late-harvested fruit (Figure 4C, D). Total phenolic concentrations in the flesh were not affected by 1-MCP treatment in early harvested fruit but affected by 1-MCP treatment in late-harvested fruit after 1 day and 5 days at 20 °C (Figure 4E, F). A late harvest increased the risk of the development of browning, in particular for 1-MCP treated fruit (Figure 4G, H).

CO\(_2\) concentrations significantly influenced all the POX and PPO activities and total phenolic contents in the 1-MCP-treated fruit, with a correlation observed between CO\(_2\) and POX and PPO activities in the early harvested fruit (Table 1).

Table 1. Correlation between CO\(_2\) and POX, polyphenol oxidase (PPO) and total phenolic contents of early and late-harvested ‘Empire’ apples with/without 1-MCP at 0.5 °C in air storage up to 7 months and evaluated for 7 days at 20 °C.

| Variables     | CO\(_2\)-POX |          | CO\(_2\)-PPO |          | CO\(_2\)-Phenol |          |
|---------------|--------------|----------|--------------|----------|----------------|----------|
|               | \( R^2 \)    | \( p \)-Value | \( R^2 \)    | \( p \)-Value | \( R^2 \)    | \( p \)-Value |
| Early harvest | 0.6077       | 0.0078   | 0.7061       | 0.0023   | 0.0051         | 0.9920   |
| Late harvest  | 0.0114       | 0.3627   | 0.0306       | 0.9754   | 0.0154         | 0.0103   |
| With MCP      | 0.5440       | 0.0149   | 0.4277       | 0.0402   | 0.7753         | 0.0008   |
| Without 1-MCP | 0.3850       | 0.0556   | 0.0650       | 0.4771   | 0.5329         | 0.0165   |
Figure 4. Peroxidase (POX) (A,B), PPO (C,D), total phenolic contents (E,F) and browning (G,H) of early and late-harvested ‘Empire’ apples and stored at 0.5 °C in air storage up to 7 months and evaluated for 7 days at 20 °C. Bars represent error of the means (SEM; n = 5), when larger than the dimension of the symbol. * Adjacent to each data point for each storage time indicates significant differences as determined by Duncan’s multiple range test at p ≤ 0.05; ns—not significantly different.
Percent change in electrical conductivity of the flesh ranged from 40% to 60% at an early harvest (Figure 5A) and from 50% to 80% at a late harvest (Figure 5B) for the duration of shelf life. The leakage was found to be slightly higher for fruit without 1-MCP at an early harvest and with 1-MCP at a late harvest after 7 days and 1 day at 20 °C, respectively.

![Figure 5.](image)

**Figure 5.** Leakage of early (A) and late-harvested ‘Empire’ apples (B) and stored at 0.5 °C in air storage up to 7 months and evaluated for 7 days at 20 °C. Bars represent error of the means (SEM; n = 5), when larger than the dimension of the symbol. * Adjacent to each data point each storage time indicates significant differences as determined by Duncan’s multiple range test at p ≤ 0.05; ns—not significantly different.

4. Discussion

The 1-MCP treatment delayed IEC and maintained high fruit firmness in ‘Empire’ apples during the seven-month cold storage period plus 7 days shelf life duration. However, incidences of flesh browning in late-harvested fruit were increased by 1-MCP treatment, which was known to risk CO₂ injury observed as internal browning in ‘Elstar’, ‘Jonagold’, ‘Gloster’, ‘Empire’ and ‘Braeburn’ apples [1,4,9,13] and as skin browning in ‘Granny Smith’ and ‘Golden Delicious’ [25,26]. Blocking the pathway of ethylene signaling in the 1-MCP-treated apples would have resulted in an extended period of insensitivity to ethylene and exposure to oxidative stress [1]. This may be associated with progressive loss of fluidity of the plasma membrane, cell wall modification and increase of skin resistance to gas diffusion, resulting in high CO₂ and low O₂-related stress [1,10,13,15,27–29]. Simultaneously, impermeable cell membranes altered energy levels of fruit metabolism, reduced the functioning of cellular membranes and facilitated the release of PPO activity, eventually leading to browning, which was shown in higher leakage in the late-harvested fruit treated with 1-MCP in this study.

The optimum window for harvest was very closely connected with susceptibility to flesh browning over time, which was dramatically alleviated from late-harvested ‘Empire’ apples stored at an air temperature of 0.5 °C in this study and at CA in the previous reports [3,30]. The 1-MCP-treatment of fruit may have resulted in higher tolerance of oxidative stress associated with concentrations of ethylene and free radical damage to proteins [8]. In our study, POX compound was found to increase in late-harvested fruit more sharply and would not be 1-MCP treatment dependent or at least not at this stage of fruit storage time. In addition, PPO activities increased in the late-harvested fruit treated with 1-MCP. Simultaneously, ‘Empire’ apples during the ripening period presumably experienced a decrease of gas diffusion rates across the cortical tissues of the fruit, signaling an initial symptom of browning, although the fruit density was lower than those of ‘Braeburn’, ‘Fuji’, ‘Gala’ and ‘Red Delicious’ apples [15].

Higher levels of phenolic compounds observed in the 1-MCP-treated fruit or late-harvested fruit may be associated with changes from the oxidizing enzymes of PPO and POX [4,13,31,32], which were
not found to affect the relationship between the compounds in this study. Interestingly, increased CO₂ concentrations in fruits treated with 1-MCP may be an important factor influencing the expression of POX and PPO in the flesh. Although 1-MCP treatment decreased CO₂ concentrations, the risk of CO₂ injury was increased in ‘Elstar’, ‘Jonagold’, ‘Gloster’ and ‘Empire’ apples [1,4,13]. Therefore, a strong reduction of concentrations of CO₂ could compensate for the negative effects of 1-MCP.

Daily variations incidences of browning during the seven-day shelf life were not likely linked with POX or PPO daily trends. Consequently, a slight increase in the susceptibility to browning was only identified with high PPO in the late-harvested fruit treated with 1-MCP, regardless of storage time. Browning incidence was primarily observed in the ‘Empire’ apples stored in CA with improper concentrations of O₂ or CO₂ [33,34]. This modified gas concentration in the fruit could be associated with a late harvest, increasing the percent change in electrical conductivity and disorder of the fruit treated with 1-MCP. Interestingly, fruit without 1-MCP treatment showed decreased browning rates for seven-day shelf life after a late harvest, which may be partially explained by the different sites on the tree used for fruit sampling as the fruits were hardly observed for browning symptoms in the higher branches.

5. Conclusions

There is little information about the 1-MCP mode of action under prolonged air-storage and shelf life simulation, affecting development of flesh browning in late-harvested ‘Empire’ apples. The 1-MCP treatment reduced the antioxidant defense mechanism of the fruits to stress in extending storage life, increasing PPO activity and electrolyte leakage rate. These factors could be evidently responsible for increasing browning symptoms of 1-MCP-treated ‘Empire’ apples under air-storage period for more than seven months. Further work should focus on extending experimental designs to include various harvest periods of the fruit variety for multiple years to identify an initiating signal for the development of browning through monitoring POX and other antioxidant activities.

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Conflicts of Interest: The authors declare no conflicts of interest.

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