Assessment of the effect of age on macular layer thickness in a healthy Chinese cohort using spectral-domain optical coherence tomography

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Abstract

Background: To determine the effect of age on the thickness of individual retinal and choroidal vascular layers in the macula in an ophthalmologically healthy Chinese cohort by using spectral-domain optical coherence tomography (SD-OCT).

Methods: In all, 525 health eyes of 525 subjects were examined with SD-OCT. The instrument automatically obtained the regional retinal thickness of 8 layers. Subfoveal choroidal vascular layers' thickness was measured using enhanced depth imaging mode. The correlation of age with layer thickness measurements was determined.

Results: No age-associated variation was found on retinal thickness (RT) in the fovea; however, the foveal thickness of outer nuclear layer (ONL), retinal pigment epithelium (RPE) and vascular sublayers of the choroid decreased significantly with aging in this area (P < 0.05, respectively). Significant age-related reduction was seen in RT in the pericentral and peripheral rings (P < 0.05, respectively). The significant variation in thinning of the ganglion cell layer, inner plexiform layer, and ONL with aging is thought to be the main determinant of these results (P < 0.05, respectively). On the contrary, the RPE layout showed age-related thickening (P < 0.05, respectively) in the pericentral and peripheral regions.

Conclusions: The thickness of individual layers of the macula may be determinants of the age-related variations observed in the ophthalmologically healthy Chinese cohort, as assessed by SD-OCT examination.

Keywords: SD-OCT, Aging, Retina, Choroid, Layer thickness, Macula

Background

Detailed assessment of the macular area is critical in the diagnosis and management of a variety of ocular diseases. Traditional investigations such as fundus photography and fluorescein angiography can only provide qualitative and prospective information, therefore being subjective and relatively insensitive to small changes of the macula and unable to provide any cross-sectional or thickness-related data. The introduction of optical coherence tomography (OCT) has made it possible to noninvasively quantify macular structures in vivo with high resolution [1, 2]. In addition, because OCT is easy to use, ensures patient comfort, and is economical, it has become an important diagnostic tool for fundus diseases.

Spectral domain-OCT (SD-OCT) is an advanced modification of time-domain OCT that provides better reproducibility for image acquisition, high-resolution three-dimensional images, and volumetric analyses [3, 4]. Techniques such as enhanced-depth imaging (EDI) permit improved analysis of the living choroid [5]. In addition, advances in layer segmentation algorithms have facilitated the automatic measurement of the thickness of individual retinal layers [6, 7].

Thickness measurement of the macula using SD-OCT has been shown to play an important role in understanding of the anatomy of individual macular layers, each of which has its own normal three-dimensional shape and may be affected in various ways by different diseases. Several studies
have investigated morphological abnormalities of the macula in some ocular diseases by using SD-OCT. Macular thickening due to fluid accumulation is found in diabetic retinopathy and central serous chorioretinopathy (CSCR) [8–10]. The visual acuity of center-involved diabetic macular edema or CSCR eyes may be dependent on the disorganization of the retinal inner layers or the outer nuclear layer (ONL) in the fovea [8, 9]. Macular morphology is also an important parameter for monitoring and staging of glaucoma or age-related macular degeneration (AMD) [11, 12]. Moreover, clinically detected morphologic changes of different retinal layers were identified in many systemic diseases such as multiple sclerosis, [13] Parkinson’s disease, [14] Alzheimer's disease, [15] and diabetes mellitus with preclinical retinopathy [16]. Therefore, measuring macular thickness by OCT is a powerful tool for physicians to evaluate progression of certain diseases, especially those that involve certain layers.

Recently, SD-OCT was used to study normal retinal and choroid thickness among subjects of different ethnicities, gender, and ages [17–20]. Age-related reduction in macular thickness was shown in a Caucasian and a Japanese population [18, 21]. However, the aforementioned reports were insufficient to facilitate the detailed analysis of the structure of specific retinal and choroidal layers. Moreover, to our best knowledge, there is no normative database available for the thickness of individual macular layers in the Chinese population.

Therefore, in this study, we used SD-OCT to measure the total retinal thickness (RT), the thickness of individual retinal layers of the macula that were divided into nine sectors, and the subfoveal choroidal thickness (SFCT) including vascular sublayers in 525 ophthalmologically healthy eyes in order to evaluate the effect of age on normal mean regional retinal and subfoveal choroidal layers on the macula.

**Methods**

**Subjects**

In this prospective observational study, self-reported, ophthalmologically healthy subjects of Chinese ethnicity aged ≥20 years were randomly recruited from May 2015 to December 2016. The study adhered to the tenets of the Declaration of Helsinki and was approved by the Ethics Committee of Qilu Hospital. Written consent was obtained from each subject.

All participants underwent a comprehensive ophthalmologic examination including best corrected visual acuity (BCVA), refraction, slit-lamp biomicroscopy, intraocular pressure (IOP) measurement by Goldmann applanation tonometry, and fundus photography obtained by two trained ophthalmologists. The inclusion criteria were as follows: BCVA ≥20/25 Snellen (0.1 LogMAR), spherical equivalent refractive error not exceeding ±6.0 diopters, IOP < 21 mm Hg, no history of any ocular abnormalities other than mild to moderate cataracts, no family history of glaucoma, and no systemic diseases such as hypertension, diabetes, or any other autoimmune or infectious diseases. One eye of each participant was randomly selected for OCT examination with the pupil dilated using 0.1% tropicamide.

**Optical coherence tomography and layer segmentation**

OCT measurements were performed with the Heidelberg Spectralis OCT (Heidelberg Engineering, Heidelberg, Germany). The instrument incorporates a real-time eye-tracking system that combines a confocal scanning laser ophthalmoscope and SD-OCT scanners to adjust for eye motion. The experienced operators performed all OCT scans under the same intensity of dim room lighting. If any scan was of insufficient quality, it was immediately repeated and reviewed until the image was satisfactory.

The macula was segmented into three concentric circles with diameters of 1 mm, 3 mm, and 6 mm, which were termed as the fovea, pericentral ring, and peripheral ring, respectively (Fig. 1a). Furthermore, the pericentral and peripheral rings were equally divided into four regions: superior, nasal, inferior, and temporal, according to the Early Treatment Diabetic Retinopathy Study (ETDRS). In all, 9 sectors were involved in the macular area (Fig. 1b and c). Each SD-OCT image was analyzed using an image segmentation algorithm, and thickness profiles of RT and eight individual retinal layers were automatically generated by the Spectralis OCT software (Fig. 2). The distance from the internal limiting membrane to the outer border of Bruch’s membrane or external limiting membrane was taken as the RT or inner retinal thickness (IRT), and the individual retinal layers were identified as follows (from inner to outer surface): retinal nerve fiber layer (RNFL), ganglion cell layer (GCL), inner plexiform layer (IPL), inner nuclear layer (INL), outer plexiform layer (OPL), ONL, photoreceptor layer with retinal pigment epithelium (PRL + RPE), and the RPE alone.

SFCT was determined from images acquired by the Heidelberg Spectralis OCT device with enabled EDI mode and analyzed with the OCT-supplied software (Fig. 3). High-quality horizontal and vertical line scans centered on the fovea were obtained. In the fovea, the SFCT was manually measured from the hyperreflective line of the Bruch’s membrane to the innermost surface of the chorioido-scleral interface [5]. The thickness of Haller’s layer was measured from the inner border of the chorioido-scleral interface junction to the innermost point of the selected large choroidal vessel that was located close to the chorioido-scleral border and within the closest proximity to the locations of the choroidal thickness measurement lines. The difference of these measurements was considered as the depth of the choriocapillaris/Sattler’s layer [10]. Means were calculated as the average thicknesses measured from horizontal and vertical sections.
Statistical analysis

All data were described as mean ± standard deviation (SD) where applicable. Statistical analyses were performed with commercial statistical software (IBM SPSS Statistics 21; SPSS Inc., Chicago, IL). The partial correlation test was used to determine the effect of age on individual layers’ thicknesses with spherical equivalent and IOP as confounders that were known to influence OCT thickness measurements [17, 22]. Finally, simple linear regression analysis was performed for the layer whose thickness correlated significantly with age. P values < 0.05 were considered statistically significant.

Fig. 1 Early Treatment Diabetic Retinopathy Study (ETDRS) grid. a Delineation of the nine macular sectors, according to the ETDRS, within which we measured macular layer thickness. b Nine ETDRS sectors in right eye. c Nine ETDRS sectors in left eye.

Fig. 2 An automated method (with manual correction) was used to segment retinal boundaries in each of the averaged B-scans in the spectral-domain optical coherence tomography examination. The individual retinal layers were identified as follows (from inner to outer surface): (Layer 1) Retinal nerve fiber layer (RNFL), (Layer 2) Ganglion cell layer (GCL), (Layer 3) Inner plexiform layer (IPL), (Layer 4) Inner nuclear layer (INL), (Layer 5) Outer plexiform layer (OPL), (Layer 6) Outer nuclear layer (ONL), (Layer 7) Photoreceptor layers (PRL), (Layer 8) Retinal pigment epithelium (RPE). Abbreviations: ILM: internal limiting membrane, BM: Bruch’s membrane, ELM: external limiting membrane.
Results
The study included 525 ophthalmologic healthy eyes of 525 subjects ranging in age from 20 to 87 years (mean age, 44.82 ± 17.74 years). Demographic and ocular features of the study population are presented in Table 1.

The mean thickness of RT and eight individual retinal layers in 9 macular EDTRS sectors of all participants are presented in Appendix: Table 6. IRT was excluded owing to similar results as that of RT (data not shown). After adjusting for spherical equivalence and IOP, no significant correlation was found on foveal RT ($P = 0.54$) (Table 2). In the fovea, the ONL and RPE correlated negatively with age (Correlation = $−0.15$, $P < 0.01$; Correlation = $−0.09$, $P = 0.03$, respectively) (Table 2); however, the RNFL, INL, and OPL correlated positively with age (Correlation = $0.13$, $0.30$, $0.10$, respectively; all $P < 0.05$) (Table 2). Regression analysis indicated an increase for the RNFL boundary as well as the INL boundary and a loss for the RPE boundary with increasing age (Beta = $0.13$, $0.10$, $−0.14$, respectively; all $P < 0.05$) (Table 2).

As shown in Table 3, the total SFCT, thickness of the large choroidal vessel layer (Haller’s layer), choriocapillaris layer and Sattler’s layer (medium choroidal vessel layer) in the fovea showed significant negative correlation with age (Correlation = $−0.55$, $−0.42$, $−0.46$, respectively; all $P < 0.05$). In addition, our study found that SFCT and the thickness of choroidal vascular sublayers decreased linearly with age (Beta = $−0.61$, $−0.47$, $−0.52$, respectively) (all $P < 0.05$).

Significant age-related reductions were seen for the RT, GCL, and IPL in the pericentral and peripheral rings (all $P < 0.05$; Tables 4 and 5). Moreover, the OPL of the temporal sector, ONL except the temporal sector in the pericentral ring (both $P < 0.05$; Table 4), RNFL of both superior and inferior sectors, INL except the superior sector, ONL of all sectors, and PRL + RPE of the inferior sector in the periperal ring (all $P < 0.05$; Table 5) showed significant decreases with respect to age. However, significant age-related increase was demonstrated in the RNFL of the temporal sector, INL and OPL of the nasal sector, RPE of all sectors in the pericentral ring (all $P < 0.05$; Table 4), RNFL of the temporal sector, OPL of the nasal sector, and RPE of the superior and temporal sectors in the peripheral ring (all $P < 0.05$; Table 5).

Discussion
In this study, consistent with previous reports, [23–25] no significant correlation was found between age and foveal RT. However, the ONL, RPE and choroid vascular sublayers in this region showed significant age-related thinning, accompanied with age-related thickness of RNFL, INL, and OPL. To our best knowledge, we believe this is the first study to report the detailed age-related changes of foveal microstructure. On comparing with total thickness,

Table 1 Demographic and Ocular Features of Included Subjects

| Age groups (y) | Number | Men/Women (ratio) | Mean Refractive Error (diopters) | Mean Intraocular Pressure (mm Hg) | Mean Age (y) |
|----------------|--------|------------------|-------------------------------|---------------------------------|-------------|
| 20–29          | 176    | 90/86(1.05)      | $−1.93 ± 1.71$                | 14.1 ± 2.4                      | 25.71 ± 1.52|
| 30–39          | 50     | 27/23(1.17)      | $−0.74 ± 1.19$                | 14.3 ± 2.1                      | 35.38 ± 3.13|
| 40–49          | 108    | 68/40(1.7)       | $−0.36 ± 1.29$                | 13.8 ± 2.3                      | 46.07 ± 2.33|
| 50–59          | 79     | 41/38(1.08)      | $−0.54 ± 1.54$                | 146 ± 2.1                       | 53.90 ± 2.51|
| 60–69          | 60     | 32/28(1.14)      | 0.08 ± 0.59                   | 14.5 ± 2.6                      | 64.53 ± 3.12|
| 70+            | 52     | 27/25(1.08)      | $−0.04 ± 0.91$                | 14.2 ± 2.0                      | 79.40 ± 5.25|
| Total          | 525    | 285/240(1.19)    | $−0.87 ± 1.60$                | 14.3 ± 2.3                      | 44.82 ± 17.74|

Mean refers to mean ± standard deviation
assessment of macular layers provides a higher diagnostic power. The most significant observation herein was the age-related thinning of foveal ONL, RPE and choroid measurements even in ophthalmologically healthy subjects, which could be a potential anatomic predisposing factor for monitoring the age-related diseases in this eye region. The atrophy of RPE and choroid layer in the central retina is a feature of early/intermediate AMD, the incidence of which is increased with age. A 32% loss in the RPE/PRL thickness and a 22% loss in ONL thickness were found over the drusen as compared to the adjacent drusen-free regions in AMD patients [26]. The choriocapillaris degenerates in early stages of AMD, before loss of photoreceptor cells or RPE [12]. Although AMD is a complicated process that involves both age-related change and tissue damage caused by multiple stresses, age plays the most important role [27]. Functionally normal RPE and choroidal vasculature play a critical role in maintaining retinal health. Thinner RPE and choroid layer thickness may be anatomic features lead to increased risk in AMD. Our results showed that assessment of the foveal layer thickness with OCT in ophthalmologically healthy aged subjects’ eyes may lead to early identification and treatment of AMD. Moreover, further investigations are needed on the mechanism of age-related variations of the ONL, RPE, RNFL, INL, and OPL in the fovea.

We have observed significant age-associated reductions of RT in the pericentral and peripheral rings that were distinct from the foveal results; these results were consistent with previous studies [19, 28, 29]. Notably, age-related changes of GCL, IPL, and ONL in the region likely contribute to this result. Parikh et al. [30] reported that age was related to the loss of neurons or glial cells in the inner retina, which may be responsible for the SD-OCT–examination outcome in this area.

Several studies have shown that assessment of GCL thickness could be a surrogate method to evaluate glaucomatous damage [31]. In this study, the observed age-related variation in GCL thickness in this ophthalmologically healthy cohort is a reminder that GCL thinning requires more accurate quantification before widespread adoption as a surrogate for glaucoma assessment.

The thickness of RPE in the pericentral and peripheral regions was significantly increased with aging. Many pathological changes led to the thickening of the RPE, which included the density of residual bodies and accumulation of lipofuscin, accumulation of basal deposits on or within the Bruch’s membrane, formation of drusen, and thickening of the Bruch’s membrane [32]. The age-related variation of RPE in the macular region requires future investigation.

This study has some limitations. The small sample size might have introduced some bias. Our data are limited to Chinese ethnicity and need to be tested in other ethnic groups in the future.

**Table 2** Correlations of Age with Regional Retinal Thickness of Foveal Layers

| Retinal Layer | RT | RNFL | GCL | IPL | INL | OPL | ONL | PRL + RPE | RPE |
|---------------|----|------|-----|-----|-----|-----|-----|-----------|-----|
| **P Value**  | 0.54 | < 0.01<sup>b</sup> | 0.36 | 0.10 | < 0.01<sup>c</sup> | 0.02<sup>c</sup> | < 0.01<sup>c</sup> | 0.77 | 0.03<sup>c</sup> |
| **Correlation** | 0.03 | 0.13 | − 0.04 | 0.07 | 0.30 | 0.10 | − 0.15 | − 0.01 | − 0.09 |
| **P Value**  | 0.28 | < 0.01<sup>c</sup> | 0.15 | 0.25 | < 0.01<sup>c</sup> | 0.63 | 0.07 | 0.68 | < 0.01<sup>c</sup> |
| **Beta**     | 0.05 | 0.13 | − 0.06 | 0.05 | 0.10 | 0.02 | − 0.08 | − 0.02 | − 0.14 |

*Abbreviations: RT Retinal thickness, RNFL Retinal nerve fiber layer, GCL Ganglion cell layer, IPL Inner plexiform layer, INL Inner nuclear layer, OPL Outer plexiform layer, ONL Outer nuclear layer, PRL Photoreceptor layer, RPE Retinal pigment epithelium, IOP Intraocular pressure |

<sup>a</sup>Partial correlation coefficient after adjusting for spherical equivalent and IOP

<sup>b</sup>Simple linear regression analysis

<sup>c</sup>Statistically significant

**Table 3** Correlations of Age with Thickness of Suboveal Choroidal Layers

| Suboveal Choroidal Layer | Thickness (mean ± SD, μm) | P Value<sup>a</sup> | Correlation<sup>a</sup> | P Value<sup>b</sup> | Beta<sup>a</sup> |
|--------------------------|--------------------------|---------------------|---------------------|-------------------|------------------|
| Total Choroidal Thickness | 225.02 ± 35.71           | < 0.01<sup>b</sup> | − 0.55              | < 0.01<sup>b</sup> | − 0.61           |
| Haller’s Layer Thickness  | 157.62 ± 26.57           | < 0.01<sup>b</sup> | − 0.42              | < 0.01<sup>b</sup> | − 0.47           |
| Choriocapillaris /Sattler’s layer Thickness | 67.41 ± 17.83 | < 0.01<sup>b</sup> | − 0.46              | < 0.01<sup>b</sup> | − 0.52           |

<sup>a</sup>Partial correlation coefficient after adjusting for spherical equivalent and IOP

<sup>b</sup>Statistically significant

<sup>c</sup>Simple linear regression analysis
### Table 4: Correlations of Age with Thickness of Macular Retinal Layers in Sectors of the Pericentral ring

| Retinal Layer | Pericentral Superior | Pericentral Nasal | Pericentral Inferior | Pericentral Temporal |
|---------------|----------------------|-------------------|----------------------|----------------------|
|               | $P$ Value$^a$ | Correlation$^a$ | $P$ Value$^b$ | Beta$^b$ | $P$ Value$^a$ | Correlation$^a$ | $P$ Value$^b$ | Beta$^b$ | $P$ Value$^a$ | Correlation$^a$ | $P$ Value$^b$ | Beta$^b$ | $P$ Value$^a$ | Correlation$^a$ | $P$ Value$^b$ | Beta$^b$ |
| RT            | $< 0.01^c$ | $-0.27$ | $< 0.01^c$ | $-0.24$ | $< 0.01^c$ | $-0.21$ | $< 0.01^c$ | $-0.16$ | $< 0.01^c$ | $-0.26$ | $< 0.01^c$ | $-0.23$ | $< 0.01^c$ | $-0.25$ | $< 0.01^c$ | $-0.19$ |
| RNFL          | $0.72$ | $-0.02$ | $0.51$ | $-0.03$ | $0.74$ | $0.01$ | $0.66$ | $0.02$ | $0.17$ | $-0.06$ | $0.27$ | $-0.05$ | $< 0.01^c$ | $0.24$ | $< 0.01^c$ | $0.31$ |
| GCL           | $< 0.01^c$ | $-0.31$ | $< 0.01^c$ | $-0.32$ | $< 0.01^c$ | $-0.24$ | $< 0.01^c$ | $-0.24$ | $< 0.01^c$ | $-0.32$ | $< 0.01^c$ | $-0.34$ | $< 0.01^c$ | $-0.35$ | $< 0.01^c$ | $-0.37$ |
| IPL           | $< 0.01^c$ | $-0.37$ | $< 0.01^c$ | $-0.39$ | $< 0.01^c$ | $-0.33$ | $< 0.01^c$ | $-0.33$ | $< 0.01^c$ | $-0.36$ | $< 0.01^c$ | $-0.37$ | $< 0.01^c$ | $-0.28$ | $< 0.01^c$ | $-0.29$ |
| INL           | $0.60$ | $-0.02$ | $0.20$ | $0.06$ | $< 0.01^c$ | $0.18$ | $< 0.01^c$ | $0.27$ | $0.29$ | $-0.05$ | $0.75$ | $-0.01$ | $0.08$ | $-0.08$ | $100$ | $0.00$ |
| OPL           | $0.63$ | $0.02$ | $0.08$ | $0.08$ | $< 0.01^c$ | $0.18$ | $< 0.01^c$ | $0.19$ | $0.49$ | $0.03$ | $0.85$ | $-0.01$ | $< 0.01^c$ | $-0.13$ | $< 0.01^c$ | $-0.19$ |
| ONL           | $< 0.01^c$ | $-0.13$ | $< 0.01^c$ | $-0.14$ | $< 0.01^c$ | $-0.28$ | $< 0.01^c$ | $-0.26$ | $0.01^c$ | $-0.11$ | $0.36$ | $-0.04$ | $0.27$ | $-0.05$ | $0.39$ | $0.04$ |
| PRL + RPE     | $0.16$ | $0.06$ | $0.31$ | $0.01$ | $0.91$ | $-0.01$ | $0.64$ | $0.02$ | $0.76$ | $0.01$ | $0.40$ | $0.04$ | $0.18$ | $0.06$ | $0.32$ | $0.09$ |
| RPE           | $< 0.01^c$ | $0.19$ | $< 0.01^c$ | $0.23$ | $0.01^c$ | $0.12$ | $0.01^c$ | $0.11$ | $< 0.01^c$ | $0.18$ | $< 0.01^c$ | $0.23$ | $< 0.01^c$ | $0.20$ | $< 0.01^c$ | $0.21$ |

**Abbreviations:** RT Retinal thickness, RNFL Retinal nerve fiber layer, GCL Ganglion cell layer, IPL Inner plexiform layer, INL Inner nuclear layer, OPL Outer plexiform layer, ONL Outer nuclear layer, PRL Photoreceptor layer, RPE Retinal pigment epithelium; IOP, Intraocular pressure

$^a$Partial correlation coefficient after adjusting for spherical equivalent and IOP

$^b$Simple linear regression analysis

$^c$Statistically significant
Table 5 Correlations of Age with Thickness of Macular Retinal Layers in Sectors of the Peripheral ring

| Retinal Layer | Pericentral Superior | Pericentral Nasal | Pericentral Inferior | Pericentral Temporal |
|---------------|----------------------|-------------------|----------------------|----------------------|
|               | P Value\(^a\)       | Correlation\(^a\) | P Value\(^b\)       | Beta\(^b\)          |
| RT            | <0.01\(^c\)          | −0.37             | <0.01\(^c\)          | −0.37                |
| RNFL          | 0.01\(^c\)           | −0.11             | <0.01\(^c\)          | −0.18                |
| GCL           | <0.01\(^c\)          | −0.39             | <0.01\(^c\)          | −0.37                |
| IPL           | <0.01\(^c\)          | −0.40             | <0.01\(^c\)          | −0.40                |
| INL           | 0.08                 | −0.16             | 0.01\(^c\)           | −0.11                |
| OPL           | 0.33                 | 0.04              | 0.02\(^c\)           | 0.11                 |
| ONL           | <0.01\(^c\)          | −0.28             | <0.01\(^c\)          | −0.29                |
| PRL + RPE     | 0.87                 | −0.01             | 0.59                 | 0.02                 |
| RPE           | 0.04\(^c\)           | 0.09              | 0.16                 | 0.45                 |

Abbreviations: RT Retinal thickness, RNFL Retinal nerve fiber layer, GCL Ganglion cell layer, IPL Inner plexiform layer, INL Inner nuclear layer, OPL Outer plexiform layer, ONL Outer nuclear layer, PRL Photoreceptor layer, RPE Retinal pigment epithelium, IOP Intraocular pressure

\(^a\)Partial correlation coefficient after adjusting for spherical equivalent and IOP

\(^b\)Simple linear regression analysis

\(^c\)Statistically significant
Appendix

Table 6 Mean Thickness of Retina and eight individual retinal layers in nine Macular Sectors

| Retinal Layer | fovea | Pericentral Superior | Pericentral Nasal | Pericentral Inferior | Pericentral Temporal | Peripheral Superior | Peripheral Nasal | Peripheral Inferior | Peripheral Temporal |
|---------------|-------|----------------------|-------------------|---------------------|--------------------|--------------------|-----------------|-------------------|--------------------|
| RT            | 257.04±18.85 | 339.37±17.56 | 339.58±17.22 | 335.10±16.97 | 324.92±15.82 | 300.39±15.80 | 317.43±21.96 | 287.16±15.33 | 283.77±15.41 |
| RNFL          | 10.78±2.24  | 23.44±3.68 | 20.11±2.32 | 24.50±3.10 | 16.82±3.13 | 37.52±5.14 | 46.12±6.85 | 38.93±5.70 | 19.21±8.37 |
| GCL           | 12.31±3.20  | 52.03±4.76 | 50.12±6.04 | 51.33±5.33 | 46.66±5.51 | 36.58±3.88 | 40.48±4.26 | 34.03±3.69 | 36.92±4.74 |
| IPL           | 18.28±2.85  | 41.18±3.82 | 42.34±3.84 | 40.90±3.67 | 41.01±3.64 | 29.67±2.96 | 31.20±3.19 | 27.81±3.06 | 32.61±3.00 |
| INL           | 15.27±4.63  | 39.75±4.47 | 38.48±4.09 | 40.46±4.23 | 36.48±3.84 | 32.33±3.19 | 35.07±3.03 | 32.09±3.34 | 33.90±2.63 |
| OPL           | 23.48±6.08  | 32.86±8.44 | 31.87±8.27 | 36.17±9.90 | 31.14±6.02 | 26.15±2.91 | 28.20±3.47 | 27.02±3.12 | 26.37±2.58 |
| ONL           | 88.44±11.40 | 67.48±12.72 | 72.52±11.54 | 60.45±12.68 | 70.02±9.31 | 58.15±7.73 | 55.87±7.57 | 49.58±7.16 | 55.80±6.33 |
| PRL+RPE       | 90.19±4.40  | 82.74±8.44 | 83.94±11.54 | 60.45±12.68 | 70.02±9.31 | 58.15±7.73 | 55.87±7.57 | 49.58±7.16 | 55.80±6.33 |
| RPE           | 17.5±2.20   | 16.16±1.89 | 16.30±1.99 | 15.36±1.76 | 15.10±1.60 | 14.29±1.37 | 14.18±1.47 | 13.70±1.76 | 13.50±1.38 |

Values are mean ± SD (µm). RT, retinal thickness; RNFL, retinal nerve fiber layer; GCL, ganglion cell layer; IPL, inner plexiform layer; INL, inner nuclear layer; OPL, outer plexiform layer; ONL, outer nuclear layer; PRL, photoreceptor layer; RPE, retinal pigment epithelium.

Abbreviations

AMD: Age-related macular degeneration; BCVA: Best corrected visual acuity; BM: Bruch’s membrane; CSCR: Central serous choriororetinopathy; EDIRI: Enhanced-depth imaging; ELM: External limiting membrane; ETDRS: Early Treatment Diabetic Retinopathy Study; GCL: Ganglion cell layer; ILM: Internal limiting membrane; INL: Inner nuclear layer; IOP: Intracocular pressure; IPL: Inner plexiform layer; RT: Retinal thickness; SD-OCT: Spectral-domain optical coherence tomography; ONL: Outer nuclear layer; OPL: Outer plexiform layer; PRL: Photoreceptor layer; RNFL: Retinal nerve fiber layer; RPE: Retinal pigment epithelium.

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Availability of data and materials

The datasets used and analysed during the current study are available from the corresponding author on reasonable request.

Authors’ contributions

YQ, QX, YL and YC conceived of and designed the experimental protocol. QX and YL collected the data. QX, YL and YC were involved in the analysis. YQ, QX, YL and YC read and approved the final manuscript.

Ethics approval and consent to participate

The study adhered to the tenets of the Declaration of Helsinki and was approved by the Ethics Committee of Qilu Hospital. Written consent was obtained from each participant.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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