The Electropolishing of C-Steel in Orthophosphoric Acid Containing Methanolic Plant Extract

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Plant extracts have been regarded as “green” alternatives as additives for metal electropolishing improvement. Therefore, understanding the electrochemical properties and the reaction mechanisms of the electroactive compounds from the plant extracts is necessary to further explore the mechanism and application of the plant extract-based additives for metal dissolution. The C-steel electropolishing behavior in orthophosphoric acid using the galvanostatic polarization and weight loss methods was ascertained. This was inspected via anode potential-limiting current relationship measurement and comparison in a solution of regularly mounting concentrations (from 50 to 1800 ppm of methanolic marjoram, coriander seeds, chamomile, and guava leaves extract), and the influence of temperature on the dissolution kinetics was investigated. Surface morphologies, roughness, and reflection of investigated specimens were inspected with a scanning electron microscope (SEM), profilometry, and Vis-IR spectroscopy.

Addition of methanolic plant extract to the electropolishing solution results in a lower limiting current. Retardation percentage gained from mass loss measurement is comparable with those obtained from measurements of galvanostatic polarization. Addition of 500 ppm of marjoram, coriander, chamomile, and guava leaves increases the degree of surface brightness and reflectance to 64.9, 56.59, 27, and 24.5, respectively, relative to electropolishing electrolyte-free solution. The roughness (Ra) decreased from 2.7 μm to 0.52 μm without addition of any material. Ra values are 0.28, 0.23, 0.21, and 0.17 μm in the presence of guava leaves, chamomile, coriander seeds, and marjoram.

1. Introduction

The electropolishing (EP) method is an extremely proficient process for brightening and cleaning of metals and alloys. It has obtained great consideration owing to its realistic and educational curiosity. EP is achieved via metal surface anodic dissolution controlled in an appropriate electrolyte and can improve planarization efficacy of the metal surface [1, 2], where the peaks and valleys depth differences are reduced [3, 4]. It classically arises at the limiting current of mass transfer process, where the surface of metal becomes silky and even where the peaks and valleys depth differences are reduced [3, 4]. It classically arises at the limiting current of the mass transfer process, where the surface of metal becomes silky and even. Concurrently, throughout EP, several etched pits and defects over the metal surface can be formed owing to oxygen gases evolution adjoining to the anode surface at a potential elevated than the limiting current plateau [5, 6]. To reduce the incidence of surface defects formed on C-steel during EP, different additives were added to the polishing bath.

The progression is controlled via diffusion, dependent on the concentration incline forming a discriminating electrochemical dissolution and enhancing steel planarization efficiency, where the peaks and valleys depth variations are removed [7]. It characteristically happens at the limiting current of mass transfer at extreme anodic potential, since the metal surface is leveled. Concurrently, throughout the EP process, some blemishes and depths above the metal surface can be shaped owing to oxygen gas evolution greatly nearby to the surface at elevated electric potential [8, 9].

The polishing efficiency depends on the rate of Fe^{2+} ions mass transfer from the diffusion layer to solution bulk determined via the limiting current value. The mass transfer...
rate is a function of the relative ionic mobility, anode geometry, temperature, and electrolyte physical properties. Also, the concentration of the polishing bath electrolyte was examined [8].

C-steel is extensively used in all kinds of industry as an important construction material, and corrosion of C-steel is known to occur specially in acidic environment such as the cleaning, picking, oil well acidification, and descaling process. It is a major task to control the C-steel corrosion for both corrosion scientists and material technologists.

The employment of plant parts extractions for the embarrassment of the metal dissolution has some advantages over the use of some organic/inorganic inhibitors because they are nontoxic, cheap, and environmentally friendly. Consequently the goal of this research is to inspect the C-steel dissolution performance in the presence of methanolic plant extract which is a low-cost material, biodegradable, and environmentally friendly. The investigated compounds are of interest because of their safe use, also containing electronegative atoms such as O in their molecules, and being relatively cheap and biodegradable.

2. Procedures

2.1. Solution Preparation. 8M H₃PO₄ was prepared via analytical-grade H₃PO₄ (85% w/w) dilution with double distilled water with a measured resistivity > 18 MΩ·cm. Preparation of the natural products was by methanol. All chemicals were supplied by BDH Chemicals Ltd. All experiments were performed in 8M H₃PO₄-free solution and 8 M H₃PO₄ containing several concentrations of natural compounds (50–1800 ppm).

2.2. Material Sample Composition. The C-steel composition in weight percent was as follows: C: 0.2, Mn: 2.6, Si: 0.33, S: 0.04, P: 0.039, and Fe: balance.

2.3. Methanolic Plant Extracts Preparation. Marjoram, coriander seeds, chamomile, and guava leaves were purchased from the local market, washed with distilled water, air dried, and separately ground into a powder using a blender. 25 g of the powdered samples was soaked in 500 ml methanol for one week at room temperature. The mixtures were filtered to obtain sample extracts. This stock plant extract was kept in a refrigerator. The stock solution concentration was determined by evaporating a known volume of the filtrates to eliminate the methanol totally and weighing the residue.

2.4. Electrochemical Measurements. The cell used consists of a rectangular plexiglass container with a base of 15 × 5 and a height of 10 cm with carbon steel sheets electrodes of 10 cm height and 5 cm width. Electrode separation was 15 cm. 6 V D.C. power supply, a high impedance voltmeter, a variable resistance, and a multirange ammeter were connected in series with the cell as the electrical circuit. The temperature was adjusted via placing the cell in a thermostatic water bath at several temperatures, (20, 30, 40, and 50°C) ± 0.5°C.

2.5. Chemical Measurements. C-steel specimens used in mass loss procedures were mechanically cut into 2 × 2 × 0.3 cm³ pieces, then abraded with 180, 320, 800, and 1200 grades of emery papers, washed using deionized water, degreased with ethanol, and dried. The specimens were weighed before and after electropolishing galvanostatic measurements. Trials were performed in triplicate for reproducibility.

2.6. Surface Characterization

2.6.1. Scanning Electron Microscope. The analysis by using a SEM (JEOL JSM-5300 Scanning Electron Microscope) was carried out on the surface of C-steel samples with and without the optimum concentration of plant extract.

2.6.2. Profilometer Measurements. The quantitative surface roughness (Rₐ) was measured via stylus profilometers (Talysurf i60). The long cutoff wavelength sampling was 0.25 mm, while the short cutoff wavelength was 0.025 mm, respectively. The evaluation length was 5 mm (five sampling lengths). Approximately 150 measuring lines were used, and thus, the mean Rₐ values could be obtained.

2.6.3. Reflectance Measurements. A Shimadzu UV-3101 PC Spectrophotometer (UV-VIS-NIR) was used to gain specimens’ reflection spectra in the range of 300–1500 nm.

2.6.4. Gloss Value Measurements. Surface brightness degree was deliberated via the Brightness and Color Meter Model No. 68-50-00-0002.

2.7. Extract Characterization

2.7.1. Fourier Transform-Infrared Spectroscopy (FTIR). For FT-IR studies, a Bruker Tensor 37 Fourier transform infrared spectrophotometer instrument was used, and vacuum-dried plant extract liquid was used. The samples were measured by mixing with KBr. The spectra were measured in the range of 400–4000 cm⁻¹.

2.7.2. Gas Chromatography. GC analysis was carried out using a Gas Hewlett Packard HP-5890 series II with split/splitless injector and capillary column (30 m, 0.25 mm, 0.25 μm) fused with phenyl polysilphenylene siloxane. The injector and detector temperatures were set at 280 and 300°C, respectively, and the oven temperature was kept at 80°C for 1 min, which rose to 300°C at 20°C/min. Helium was used as carrier gas at a constant flow of 1.0 ml/min. A volume of 2 μl was injected in the splitless mode, and the purge time was 1 min.

2.7.3. Gas Chromatography-Mass Spectrometry (GC-MS) Analysis. The MS (Hewlett-Packard 5889 BMS Engine) with selected ion monitoring (SIM) was used; the mass
spectrometer was operated at 70 eV and scanned fragments from 50 to 650 m/z.

2.7.4. Identification of Components. The methodology carried out for identification of individual components of plant extract was based on chromatographic retention indices and by comparing the obtained mass spectra with those available in the NIST library.

3. Results and Discussion

3.1. Characterization of Methanolic Plant Extracts

3.1.1. Fourier Transform-Infrared Spectroscopy (FTIR). FTIR analysis of marjoram, coriander, chamomile, and guava leaves was performed, and the results are shown in Figure 1.

The broad peaks between 3386.04 cm⁻¹ to 3411.62 cm⁻¹ were assigned to the presence of a superficial absorbed water and stretching mode of (OH). The peaks at 1449, 1409.05, and 1405.54 cm⁻¹ correspond to bending vibration of aliphatic O-H and C-H. The peaks at 877.10, 864.79, 819.49, 819.38, 821.77, and 777.44 cm⁻¹ correspond to bending vibration of C-H. The peaks at 1694.23 cm⁻¹ are in proportion to stretching vibration of C=O of guava leaves extract. The peaks at 1602.59, 1607.82, 1610.60, and 1615.07 cm⁻¹ correspond to stretching vibration of C=C. The peaks at 1040.68 cm⁻¹ correspond to stretching vibration of CO-OCO of guava leaves extract. The peaks at 1054.06, 1063.11, 1071.04, 1262.96, 1281.31, and 1281.95 cm⁻¹ correspond to stretching vibration of C-O stretching.

The comparison between the absorbance bands observed by the FTIR spectra of four plant extracts before and after EP is shown in Figure 1. These absorbance bands sometimes appear, disappear, or shift after EP to indicate the formation of a C-steel plant extract complex or salt.

(1) Obviously in marjoram, a strong and broad peak of –OH stretching are observed at 3411.62 cm⁻¹ before EP and are also observed at 3409.83 cm⁻¹ after EP. The stretching peak of –C–H is shifted from 2934.31 to 2396.40 cm⁻¹. The stretching peak of C=O is shifted from 1607.82 cm⁻¹ to 1636.54 cm⁻¹. The stretching peak of C-O shifted from 1071.04 cm⁻¹ to 1023.81 cm⁻¹. Also, the bending peak of –C–H shifted from 776.70 cm⁻¹ to 503.89 and 420.22 cm⁻¹. In Guava leaves, the –C–H stretching peak is shifted from 2927.69 to 2391.11 cm⁻¹. The C=O and C-H bands’ stretching frequencies are shifted from 1694.35 to 1623.83 cm⁻¹. The C=O stretching peak disappeared after EP, C-O from 1283.31 to 1135.51 cm⁻¹, and CO-OCO from 1040.68 to 1029.04 cm⁻¹. Also, the bending peak of C-H disappeared after EP from 819.38 cm⁻¹ to 908.57 and from 768.88 to 712.95 and 524.97 cm⁻¹. In chamomile, a broad band at 3386.8 cm⁻¹ is attributed to –OH shifted to 3377.1 cm⁻¹. The C=O stretching peak is shifted from 2932.29 to 2352.07 and 2320.79 cm⁻¹, and the O-H bending peak at 1409.00 cm⁻¹ disappeared after EP. The C=C stretching peak shifted from 1610.60 to 1626.62 cm⁻¹, C-O from 1281.95 to 1257.72 and from 1063.11 to 1136.74 and 1033.80 cm⁻¹, and C=O located at 2120.72 cm⁻¹ disappeared after EP. Also, the bending peak of –C–H shifted from 777.44 cm⁻¹ to 908.28 and 523.36 cm⁻¹. In coriander, a strong –OH band at 3408.86 shifted to 3376.59 cm⁻¹, C=H stretching band at 2929.49 is shifted to 2350.69 cm⁻¹, C=C from 1602.59 to 1623.99 cm⁻¹, O-H bending peak at 1409.00 cm⁻¹ disappeared after EP, and C-O shifted from 1054.06 to 1029.38 and 1131.21 cm⁻¹. Also, the –C–H bending peak shifted from 877.10 cm⁻¹ to 907.33, 713.90 and 527.28 cm⁻¹.

The presence and shifted stretching peaks of the functional groups confirm the formation of active compound –Fe²⁺ complex on the metal surface. Hence, the FTIR spectra show that there is an interaction between the extract and the C-steel substrate which resulted in the retardation effect [10].

3.1.2. UV-Visible Spectroscopy. As seen in Figures 2(a)–2(d), the solution having Fe²⁺ ions exhibit two absorption bands at 209 and 305 nm in four plants’ extract; the guava leaves extract-containing solution shows two bands at 244 and 282 nm, and Fe²⁺ ions addition to guava leaves extract results in an intense absorption band at 323 nm indicative of Fe-guava leaves extract complex formation. The marjoram extract-containing solution shows two bands at 292 and 310 nm. Fe²⁺ ions addition to marjoram extract results in an intense absorption band at 325 nm indicative of Fe-marrjoram extract complex formation. The coriander seeds extract-containing solution shows two bands at 219 and 309 nm. Fe²⁺ ions addition to coriander seeds extract results in two absorption bands at 289 and 316 nm reflection of Fe-coriander extract complex formation. The solution containing chamomile extract shows only one band at 309 nm. Fe²⁺ ions addition to chamomile extract results in an intense absorption band at 323 nm investigative of Fe-chamomile extract complex formation.

Fe-plant extract may be as a stable complex adsorbed over the metal surface resulting in retardation influence, or it may be as a soluble complex leading to a catalytic effect. In systems that contain different chemical species, there is a possibility to form both of these two types of complexes. Therefore, the effect of plant extract on the steel dissolution behavior in 8 M H₂PO₄ could be interpreted on the basis of two aspects: the retardation influence of the stable complex and other catalytic effect of soluble metal complex. It appears that the retardation influence of the adsorbed metal complex is firstly predominant until a significant concentration where a certain leveling off value of their retardation efficiency is reached. On further increase of the amount of the plant extract, the accelerating effect of other soluble species appears, resulting in a decrease of the maximum retardation efficiency [11].

3.1.3. Gas Chromatography-Mass Spectrometry (GC-MS) Analysis. In the present investigation, 10 organic compounds from marjoram, coriander, chamomile, and guava leaves were identified through GC-MS analysis; Figure 3. The molecular formula, molecular weight, retention time (RT), and probability percentage are presented in Table 1. Among the total compounds, 1-phenanthrenemethanol, 1,2,3,4,4a,9,10,10a-octahydro-1,4a-dimethyl-7-(1-
Figure 1: Continued.
Figure 1: FTIR spectra of methanolic plant extract.

(a) H$_3$PO$_4$ + Fe$^{2+}$
1500 ppm Guava leaves + 8 M H$_3$PO$_4$
1500 ppm Guava leaves + 8 M H$_3$PO$_4$ + Fe$^{2+}$

(b) H$_3$PO$_4$ + Fe$^{2+}$
1500 ppm Marjoram + 8 M H$_3$PO$_4$
1500 ppm Marjoram + 8 M H$_3$PO$_4$ + Fe$^{2+}$

(c) H$_3$PO$_4$ + Fe$^{2+}$
1500 ppm Coriander seeds + 8 M H$_3$PO$_4$
1500 ppm Coriander seeds + 8 M H$_3$PO$_4$ + Fe$^{2+}$

(d) H$_3$PO$_4$ + Fe$^{2+}$
1500 ppm Chamomile + 8 M H$_3$PO$_4$
1500 ppm Chamomile + 8 M H$_3$PO$_4$ + Fe$^{2+}$

Figure 2: UV-visible spectrum of (a) guava leaves, (b) marjoram, (c) coriander seeds, and (d) chamomile.
Table 1: Some of the chemical components identified in methanolic extract by GC-MS.

| No | Identified compound                                      | Molecular formula | Molecular weight | RT   | Probability |
|----|----------------------------------------------------------|-------------------|------------------|------|-------------|
| Marjoram |                                                                      |                   |                  |      |             |
| 1   | cis-4-Methoxy thujane                                     | C_{11}H_{20}O     | 168              | 6.55 | 75.6%       |
| 2   | Cyclohexanol, 1-methyl-4-(1-methylethenyl)-, cis          | C_{10}H_{18}O     | 154              | 6.64 | 45.5%       |
| 3   | trans-4-Methoxy thujane                                   | C_{11}H_{20}O     | 168              | 6.96 | 38.3%       |
| 4   | Methyleugenol                                             | C_{11}H_{14}O     | 178              | 11.94| 64.2%       |
| 5   | 1H-cycloprop[e]azulen-7-ol, decahydro-1,1,7-trimethyl-4-methylene-, [1ar-(1aα,4αa,7β,7αβ,7βα)] | C_{13}H_{24}O     | 220              | 15.18| 12.3%       |
| 6   | α-Acorenol                                               | C_{15}H_{26}O     | 222              | 16.18| 7.30%       |
| 7   | Oxiraneundecanoic acid, 3-pentyl-, methyl ester, cis     | C_{19}H_{36}O     | 312              | 20.76| 12.9%       |
| 8   | n-Hexadecanoic acid                                      | C_{15}H_{32}O     | 256              | 21.49| 64.2%       |

Figure 3: GC-MS spectrum of of methanolic plant extract: (a) marjoram; (b) coriander seeds; (c) chamomile; and (d) guava leaves.
methylethyl)-, [1S-(1α,4aα,10αβ)] (RT 29.06) was the major compound identified in the marjoram extract, cis-vaccenic acid (RT 24.91 was the major compound identified in the coriander seed extract, anthracene, 1,2,3,4,5,6,7,8-octahydro-1-methyl C15H20 (RT 20.729) was the major compound identified in the chamomile seed extract, and diisooctyl phthalate (RT 31.43) was the major compound identified in the guava leaves extract.

3.2. Leveling Process. An atypical polarization curve is obtained for an electrolyte consisting of H₃PO₄ of concentrations ranged from 6 M to 14 M. Figure 4 is divided into three parts which are electrolytic etching, polishing, and (O₂) evolution that occurred consequently with the metal dissolution process. The influence of H₃PO₄ concentration on the value of \( I_L \) can be explicated based on the mass transfer equation [12, 13].

\[
I_L = \frac{nZFD_c}{\delta} \text{Fe}^{3+}.
\]  (1)

\( I_L \) = limiting current (A); \( Z \) = working area (cm²); \( n \) = charge of the ion involved; \( F \) = Faraday constant (As·mol⁻¹); \( D \) = diffusion coefficient of the rate-limiting species (cm²·s⁻¹); \( C_{Fe^{3+}} \) = saturation concentration of iron (mol·L⁻¹) in the solution for the salt film mechanism or bulk C of the acceptor species for acceptor mechanism; and \( d \) = thickness of the Nernst diffusion layer (cm).

Increasing orthophosphoric acid concentration leads to decrease in the saturation solubility of FePO₄, increase in viscosity (\( \mu \)) of the solution (which leads to reduce Fe⁺³ ions diffusivity (D) consistent with the Stokes–Einstein equation.

| No | Identified compound                                      | Molecular formula | Molecular weight | RT  | Probability |
|----|----------------------------------------------------------|-------------------|-----------------|-----|-------------|
| 9  | 9,12,15-Octadecatrienoic acid, (Z,Z,Z)                  | C₁₈H₃₀O₂          | 278             | 24.86 | 12.2%       |
| 10 | 1-Phenanthrenemethanol, 1,2,3,4,4a,9,10,10a-octahydro-1,4a-dimethyl-7-{(1-methylethyl)-, [1S-(1α,4aα,10αβ)]} | C₂₀H₃₀O           | 286             | 29.06 | 15.0%       |
| Coriander seeds                         |                  |                  |                 |     |             |
| 1  | 1,6-Octadien-3-ol, 3,7-dimethyl                         | C₁₀H₁₈O           | 154             | 6.52 | 76.1%       |
| 2  | Benzaldehyde, 4-(1-methylethyl)                         | C₁₀H₁₂O           | 148             | 9.21 | 33.9%       |
| 3  | p-Cymen-7-ol                                           | C₁₀H₁₄O           | 150             | 10.07| 58.6%       |
| 4  | Eugenol                                                 | C₁₀H₁₂O           | 164             | 11.24| 20.8%       |
| 5  | 3-Allyl-6-methoxyphenol                                 | C₁₀H₁₂O           | 164             | 11.32| 12.7%       |
| 6  | Caryophyllene oxide                                    | C₁₂H₂₀O           | 220             | 15.30| 28.7%       |
| 7  | n-Hexadecanoic acid                                    | C₁₈H₃₄O           | 256             | 21.47| 65.9%       |
| 8  | 6-Octadecenoic acid, methyl ester, (Z)                 | C₁₀H₁₈O           | 296             | 24.00| 12.3%       |
| 9  | cis-Vaccenic acid                                      | C₁₈H₃₄O           | 282             | 24.91| 15.1%       |
| 10 | Oleic Acid                                             | C₁₈H₃₄O           | 282             | 33.27| 14.5%       |
| Chamomile                               |                  |                  |                 |     |             |
| 1  | 2-Furanmethanol, tetrahydro-a,a,5-trimethyl-5-(4-methyl-3-cyclohexen-1-yl)-, [2S-[2α,5β (R*)]] | C₁₂H₂₀O           | 238             | 16.46| 53.4%       |
| 2  | 11,13-Dihydroxy-tetradec-5-ynoic acid, methyl ester     | C₁₂H₂₄O           | 270             | 17.83| 32.3%       |
| 3  | 2H-1-Benzopyran-2-one, 7-methoxy                       | C₁₂H₂₄O           | 176             | 18.01| 92.5%       |
| 4  | 2H-Pyran-3-ol, tetrahydro-2,2,6-trimethyl-6-(4-methyl-3-cyclohexen-1-yl)-, [3S-[3α,6α (R*)]] | C₁₂H₂₀O           | 238             | 18.16| 85.6%       |
| 5  | Limonen-6-ol, pivalate                                  | C₁₆H₃₂O           | 236             | 18.52| 11.0%       |
| 6  | 1,6-Dioxaspiro[4.4]non-3-ene, 2-(2,4-hexadiynylidene)   | C₁₄H₂₀O           | 200             | 20.50| 83.5%       |
| 7  | Anthracene, 1,2,3,4,5,6,7,8-octahydro-1-methyl          | C₁₃H₂₀O           | 200             | 20.73| 3.06%       |
| 8  | Formic acid, 3,7,11-trimethyl-1,6,10-dodecatrien-3-yl ester | C₁₄H₂₀O           | 250             | 21.07| 10.1%       |
| 9  | n-Hexadecanoic acid                                    | C₁₈H₃₄O           | 256             | 21.56| 52.1%       |
| 10 | 5-Hydroxymethyl-1,3,3-trimethyl-2-(3-methyl-buta-1,3-dienyl)-cyclopentanol | C₁₄H₂₀O           | 224             | 22.15| 4.00%       |
| Guava leaves                            |                  |                  |                 |     |             |
| 1  | Benzaldehyde, 4-(1-methylethyl)                         | C₁₀H₁₂O           | 148             | 9.18 | 41.3%       |
| 2  | Caryophyllene oxide                                    | C₁₀H₁₂O           | 220             | 15.31| 47.5%       |
| 3  | Tetracyclo[6.3.2.0(2,5).0(1,8)]tridecan-9-ol, 4,4-dimethyl | C₁₂H₂₄O           | 220             | 16.18| 51.6%       |
| 4  | Globulol                                                | C₁₂H₂₄O           | 222             | 16.51| 9.92%       |
| 5  | 9-Hexadecanoic acid                                    | C₁₄H₂₆O           | 254             | 16.67| 8.95%       |
| 6  | Isoaromadendrene epoxide                                | C₁₄H₂₆O           | 220             | 16.72| 12.7%       |
| 7  | 4,4,8-Trimethyltricyclo[6.3.1.0(1,5)]dodecane-2,9-diol  | C₁₄H₂₆O           | 238             | 20.53| 74.4%       |
| 8  | Hexadecanoic acid, methyl ester                         | C₁₄H₂₆O           | 270             | 20.74| 23.7%       |
| 10 | Diisooctyl phthalate                                   | C₂₄H₃₈O           | 390             | 31.34| 18.5%       |
Anodic dissolution of C-steel in H$_3$PO$_4$ shows a limiting current plateau. In the potential range below $I_L$, C-steel dissolution occupies concurrent Fe$^{2+}$ and Fe$^{3+}$ formation, Fe$^{3+}$ formation amount becoming significant at superior potential. In the potential range corresponding to the limiting current plateau, C-steel dissolution results in Fe$^{3+}$ formation only. At potentials elevated than $I_L$, O$_2$ evolution at C-steel anode dissolution will appear.

### 3.3. Electropolishing of C-Steel in H$_3$PO$_4$ with Natural Products

Figure 5 Demonstrates polarization curves for C-steel anodic dissolution in H$_3$PO$_4$ containing various concentrations of marjoram, coriander seeds, chamomile, and guava leaves methanolic extract, respectively. The current was plotted against the applied potential, and the linear portion, an extrapolation of the dissolution potential, gives the limiting current ($I_L$). $I_L$ values are given in Table 2.

Appearance of a limiting current plateau in the solution containing several extract concentrations shows that the extracts do not alter the diffusion-controlled mechanism for iron dissolution to a charge transfer mechanism. However, the plant extracts affect the $I_L$ by diverse extent reflecting their retardation influence which is dependant on the plant extract type.

If $I_L$(blank) is the limiting current in 8M H$_3$PO$_4$-free solution and $I_L$(extract) is the limiting current in 8M H$_3$PO$_4$ containing plant extract, then the inhibition efficiency percentage ($%IE$) can be calculated as follows:

$$%IE = \left(1 - \frac{I_L(\text{extract})}{I_L(\text{blank})}\right) \times 100.$$  \hspace{1cm} (2)
The anodic dissolution rate $I_p$ values and percentage inhibition efficiency ($%IE$) for the studied plant extract at a concentration range from 50 to 1800 and temperature range of 25–55°C with an increment of 10°C are given in Table 2. Table 2 shows that $%IE$ increases as the studied methanolic extracts increase and temperature decreases (except guava leaves). The $%IE$ of different methanolic extracts could be arranged in the order as follows: Marjoram > Coriander > Chamomile > Guava leaves. The order imitates the significant function played via the molecular size and the substituent group, as well as functional adsorption atom type, in the retardation processes.

The retardation efficiency of the studied plant extracts was found to be increasing from 50–1800 ppm extract...
concentration. A maximum of chamomile and guava leaves %IE has been exhibited at 1200 ppm. However, it is observed that the behavior is there after overturned with augment in the chamomile and guava leaves concentration (at 1500 and 1800 ppm).

They have an optimum concentration (1200) after which IL increases and %IE decreases. The increase in %IE could be featured to the chemical ingredients adsorption of methanolic extracts of the studied natural products.

The effect of chamomile and guava leaves on the anodic dissolution behavior of C-steel in 8 M H3PO4 could be interpreted on the basis of two aspects:

1. The retardation effect of the stable metal complex adsorbed over the metal surface
2. Catalytic effect of the soluble metal complex

Since chamomile and guava leaves extracts are rich in compounds such as alkaloids, flavonoids, and tannins, there is an opportunity to form both the complexes types. Consequently, their retardation influence could be explained on the basis of the two factors. It seems that inhibitive effect of the adsorbed metal complex is firstly major until optimum concentration where a confident leveling-off value of their retardation is reached. After a certain period, on further

| Table 2: Limiting current values for C-steel dissolution in 8 M H3PO4 acid in the absence and presence of natural extracts at several temperatures. |
|---------------------------------|
| Natural compound | Conc. (ppm) | 25°C | 35°C | 45°C | 55°C |
|-------------------|-------------|------|------|------|------|
|                   | IL (A) | IE% | IL (A) | IE% | IL (A) | IE% | IL (A) | IE% | IL (A) | IE% |
| Marjoram (Origanum majorana) | 0.00 | 0.500 | 0.560 | 0.670 | 0.750 |
| Coriander (Coriandrum sativum) seeds | 0.00 | 0.500 | 0.560 | 0.670 | 0.750 |
| Chamomile (Matricaria recutita) | 0.00 | 0.500 | 0.560 | 0.670 | 0.750 |
| Guava leaves (Psidium guajava L) | 0.00 | 0.500 | 0.560 | 0.670 | 0.750 |

| Natural compound | Conc. (ppm) | 25°C | 35°C | 45°C | 55°C |
|-------------------|-------------|------|------|------|------|
|                   | IL (A) | IE% | IL (A) | IE% | IL (A) | IE% | IL (A) | IE% | IL (A) | IE% |
| Marjoram (Origanum majorana) | 0.00 | 0.500 | 0.560 | 0.670 | 0.750 |
| Coriander (Coriandrum sativum) seeds | 0.00 | 0.500 | 0.560 | 0.670 | 0.750 |
| Chamomile (Matricaria recutita) | 0.00 | 0.500 | 0.560 | 0.670 | 0.750 |
| Guava leaves (Psidium guajava L) | 0.00 | 0.500 | 0.560 | 0.670 | 0.750 |

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increasing the plant extract amount, the accelerating effect of soluble metal complexes appears, leading to diminishing of the greatest retardation efficiency as in the case of chamomile and guava leaves [11, 16, 17].

On additional examination of Table 2, it is monitored that methanolic extract of marjoram leads to decrease in the anodic dissolution rate (Iₚ values) of C-steel and increase in the value of %IE up to 1800 ppm. The dissolution retardation can be featured to the main component adsorption of marjoram (cis- and tarns-sabien hydrate, thymol, and ethyl eugenol) on the C-steel surface. Due to the extract complex chemical composition, it is relatively tricky to allocate the retardation influence to a scruptulous component. However, communal influences of these constituent and others present in the extract cannot be ruled out. The structures of the compounds in Table 1 show that there is an abundance of O-H, C=O, C-O, and -CH₃ along with aromatic rings. This reality has been proved through the corresponding peaks in the FTIR (Figure 1). Therefore, it is reasonable to declare that the marjoram extract efficacy against C-steel dissolution belongs to the successful chemical constituent adsorption [18].

Methanolic extract of coriander seeds is composed of numerous naturally occurring compounds which may react with the iron, first dissolved from the metal surface, forming an organometal complex such as Fe-plant extract [Fe-PE] according to the following mechanism [19]

\[
\begin{align*}
Fe & \rightarrow Fe^{2+} + 2e^- \\
Fe^{2+} + \text{plant extract} & \rightarrow [\text{Fe-plant extract}]^{2+}
\end{align*}
\]

The principal active component of methanolic extract is reported to be linalool, and its structure is given in Table 1. The inhibition of methanolic extract of coriander seeds is primarily accredited to the linalool presence and other organic compounds (cumin aldehyde, cumin alcohol, eugenol, 3-allyl-6-methoxy phenol, and cryophllene oxide) in the extract. Thus, methanolic extract gets adsorbed on the C-steel surface, and a protective film is formed. This phenomenon might occur through the dipole interaction type between unshared electron pairs of oxygen atom or π-electrons (the extract aromatic ring constituents which considered a wealthy electron source) with the vacant, low energy d-orbitals of iron surface atom. Consistent with the literature molecule with OH groups, complexes with divalent ions may be willingly formed [20–22].

3.4. Electropolishing Rate Calculation. The electropolishing rate was computed according to the following equation:

\[
\text{EP rate} = \frac{[W \text{ before EP} - W \text{ after EP/area}]}{\text{Time}},
\] 

where \(W\) before EP is the weight of the test specimen before electropolishing and after surface preparation and \(W\) after EP is the weight after electropolishing. Area: 2 cm × 2 cm), and time is the electro-polishing time in seconds.

The electropolishing rate (EP rate) reduced with an augment in the methanolic extract concentration. Electropolishing rate measurements (Figure 6) confirmed the data obtained from potentiodynamic measurements, where addition of methanolic plant extract retards the dissolution rate and the retardation behavior amplifies via mounting methanolic plant extract concentration [23].

3.5. Kinetic Point of View for the Electropolishing Process. In the present study of C-steel electropolishing in \(H_3PO_4\) solutions, weight difference in the presence of different methanolic plant extracts at time \(t\) is elected, \(\Delta W\). When \(\log W\) was plotted against time (in min), a linear variation was observed with high correlation coefficient for marjoram, coriander, and chamomile except guava leaves, which corroborates a first-order reaction kinetics with respect to C-steel in \(H_3PO_4\) solutions in the presence of marjoram, coriander, and chamomile.

The first-order reaction kinetics equation is formulated as follows:

\[
\log \Delta W = \log \Delta W_o - kt,
\]

where \(\Delta W_o\) is the weight difference in the \(H_3PO_4\)-free solution, \(k\) is the rate constant, and \(t\) is the time. The rate constants, \(k\) values, obtained from the slopes of the plots in Figure 7 are presented in Table 3.

The results obtained reveal that the rate constant reduces with augment in plant extract concentration, and the half-life values, \(t^{1/2}\) (Table 3), of the metal in the test solutions were calculated using the following equation:

\[
t^{1/2} = \frac{0.693}{k},
\]

The half-life values were observed to rise with amplification in the concentration of the methanolic plant extract, demonstrating diminishing in the metal dissolution rate in the solutions with increase in the methanolic plant extract concentration [24]. Also, the rate constant values and the half-life values, \(t^{1/2}\), confirmed the result obtained from the
Figure 7: Variation of weight loss with time for mild steel (of a cross sectional area of 10 cm$^2$) in 8 M H$_3$PO$_4$ solutions containing methanolic extract: (a) marjoram, (b) coriander seeds, (c) chamomile, and (d) guava leaves at 25°C.

Table 3: Rate constant and half-life time values for mild steel coupons in 8.0 M H$_3$PO$_4$ solutions containing different concentrations of methanolic plant extract using the weight loss technique.

| Methanolic extract Conc.(ppm) | Marjoram | Coriander | Chamomile | Guava leaves |
|-------------------------------|-----------|-----------|-----------|--------------|
|                               | $k$, min$^{-1}$ | $t_{1/2}$ | $k$, min$^{-1}$ | $t_{1/2}$ | $k$, min$^{-1}$ | $t_{1/2}$ | $k$, min$^{-1}$ | $t_{1/2}$ |
| 200                           | 0.052     | 13.32     | 0.028     | 24.75      | 0.014     | 49.50     | 0.018     | 38.50       |
| 600                           | 0.047     | 14.74     | 0.026     | 26.65      | 0.0115    | 60.26     | 0.016     | 43.31       |
| 800                           | 0.034     | 20.38     | 0.025     | 27.72      | 0.006     | 115.50    | 0.013     | 53.30       |
| 1200                          | 0.023     | 30.13     | 0.023     | 30.13      | 0.004     | 173.25    | 0.006     | 115.50      |
| 1500                          | 0.011     | 63.00     | 0.022     | 31.50      | 0.0104    | 66.63     | 0.007     | 99.00       |
| 1800                          | 0.008     | 86.62     | 0.009     | 77.00      | 0.011     | 63.00     | 0.009     | 77.00       |
glavonstatic technique and confirmed via SEM image and roughness values.

3.6. Temperature Influence. The temperature was found to be an extremely significant parameter for electropolishing. The limiting current augments via mounting of temperature from 25°C to 55°C are demonstrated in Table 2 and Figure 8.

The current increase with temperature rising is in conformity with the authors’ observations [24]. This result may be related to equation (1) for the limiting current value. The temperature increase leads to amplification of the diffusion coefficient, $D$, of the rate limiting species values. The temperature is a function of $D$ according to equation (5).

$$D = D_0 \exp^{-\frac{Q_d}{RT}},$$

where $D_0$ is the temperature-independent pre-exponential, $Q_d$ = activation energy for diffusion, and $R$ is the gas constant ($8.314 J \cdot K^{-1} \cdot mol^{-1}$).

\begin{table}
\centering
\begin{tabular}{|c|c|c|}
\hline
IL (A) & Inhibition (%) \\
\hline
0.04 & 0.08 & 0.12 & 0.16 & 30 40 50 60 20 & Temp. (°C) & 80 85 90 95 \\
\hline
\end{tabular}
\caption{Inhibition of limiting current (IL) with different temperatures.}
\end{table}

\begin{figure}
\centering
\includegraphics[width=\textwidth]{figure8.png}
\caption{Variation of limiting current (IL) and % inhibition with temperature (°C) in the presence of 1800ppm of the four studied methanolic plant extracts.}
\end{figure}
The current variation with temperature may be attained via inserting equation (5) into (1):

\[ I_T = \frac{nZFD}{\delta}C_{Al^3+}exp^{\frac{Q_dRT}{RT}}. \] (7)

Taking the natural logarithm of both sides in equation (6) gives equation (7).

Equation (7) relates the bath temperature to the plateau current density (i.e., stable current density). Following this equation, it is obvious that the increase of the temperature would result in an exponential increase of the current density. However, it should be noticed that as the temperature rises, the viscosity of the diffusion layer on the anode surface is decreased, making it more difficult to maintain a viscous anode layer. This would inevitably affect the metal surface finish quality. In this sense, the temperature range to reach an optimum electropolishing result for a specific system should be carefully selected [25].

\[ \ln I_T = \ln \frac{nZFD}{\delta}C_{Al^3+}exp^{\frac{Q_dRT}{RT}}. \] (8)

Figure 9 demonstrates a curve of \( \ln I_T \) vs. \( T^{-1} \) in a 8 M \( \text{H}_3\text{PO}_4 \)-free solution and 8 M \( \text{H}_3\text{PO}_4 \) containing 1800 of the four studied methanolic plant extract. From Figure 7, assuming the activation energy would be allowed. In the current case, the activation energy (\( E_a \)) was approximately 11.33 kJ mol\(^{-1} \). The typical activation energy for the electrodissolution processes for C-steel in 8 M \( \text{H}_3\text{PO}_4 \) while in the presence of 1800 ppm of marjoram, coriander seeds, chamomile, and Guava leaves is 43.34, 32.23, 23.87, and 123.50 kJ mol\(^{-1} \), respectively. It was obvious that \( E_a \) amplified in the bath containing plant extract. The higher \( E_a \) values are an excellent confirmation for the strong retardation behavior of the four plant extract for the carbon steel electrodissolution [7, 26].

### 3.7. Surface Characterization

#### 3.7.1. Scanning Electron Microscopy

The C-steel surface morphology of samples in the 8 M \( \text{H}_3\text{PO}_4 \)-free solution and 8 M \( \text{H}_3\text{PO}_4 \) containing several concentrations of plant extract is revealed in Figure 10(a). The badly damaged and rough surface for raw the C-steel surface where there are severe damage, clear pits, and cavities. The surface heterogeneity is markedly observed. Inspections of the polished C-steel sample in the 8 M \( \text{H}_3\text{PO}_4 \) solution are shown in Figure 10(b); the surface morphology improved markedly, and there are fewer pits and cracks observed in the polished surface.

Figure 10(c) shows the C-steel surface after electropolishing in the 8 M \( \text{H}_3\text{PO}_4 \) solution containing 100 ppm of marjoram methanolic extract. It is significant to point out that when the plant extract is added to the solution, the C-steel surface morphology is fairly dissimilar from the previous one and the surfaces were smoother to some extent. On studying the influence of the four extracts, it was monitored that there is a gradual increase in surface quality and uniformity by increasing extract concentration from 100 to 500 ppm, and the surface is well polished, smooth, and completely uniform since grain boundaries represented in Figure 10(h) clearly became completely disappeared in the presence of high concentration of chamomile extract, where chamomile extract molecules are filling up the grain.

Figure 10(e) demonstrates SEM analysis of the C-steel after polishing in the 8 M \( \text{H}_3\text{PO}_4 \) solution containing 100 ppm coriander seeds extract. It can be observed that the surface has been moderately smooth in comparison with the 8 M \( \text{H}_3\text{PO}_4 \) without extracts sample, the dissimilarity becomes distinguished, mainly in orientation to the morphology obtained, and the grain boundaries are the main feature of the surface. But, the surface in the existence of 500 ppm coriander seeds extract (Figure 10(f)) was silky and minimal affected by the process of dissolution than the blank owing to the coriander seeds extract molecules adsorption on the C-steel surface and made a preservative layer on the surface. The formations of scattered pits are further diminished owing to involvement of coriander seeds extract molecules in pits and grain boundaries.

SEM C-steel surface images in the solution containing 100 ppm and 500 ppm chamomile extract are shown in Figures 10(g) and 10(h). The carbon steel surface is strongly influenced by the concentration of extract, it is monitored that there is a gradual increase in surface quality and uniformity by increasing extract concentration from 100 to 500 ppm, and the surface is well polished, smooth, and completely uniform since grain boundaries represented in Figure 10(h) clearly became completely disappeared in the presence of high concentration of chamomile extract, where chamomile extract molecules are filling up the grain.
Figure 10: C-steel surface morphology before and after electropolishing at 298 K in the absence and presence of natural extracts. (a) Raw C-steel sample before EP. (b) After EP without addition (blank). (c) After EP + 100 ppm guava leaves extract. (d) After EP + 500 ppm guava leaves extract. (e) After EP + 100 ppm chamomile extract. (f) After EP + 500 ppm chamomile extract. (g) After EP + 100 ppm coriander seeds extract. (h) After EP + 500 ppm coriander seeds extract. (i) After EP + 100 ppm marjoram extract. (j) After EP + 500 ppm marjoram extract.
boundaries, as obtained in Figure 10(g). This led to improvement in surface quality resulting a completely uniform and smooth surface.

The C-steel surface electropolished in the 8 M H₃PO₄ solution containing 100 ppm guava leaves extract (Figure 10(i)) is clear from any noticeable imperfections such as hollows and breaks, although some scratches are observable. In the presence of 500 ppm extract (Figure 10(j)), the C-steel cracks, heterogeneity, and pits are significantly reduced owing to the protective adsorption film formation offered by guava leaves extract, and clearly, a smooth and regular surface is noticed. The leveling and brightening influences were noticeably improved by the addition of Guava leaves extract where guava leaves extract is satisfying the channels and grain boundaries, and also, the etching effect is removed [27].

By comparing the effect of four extracts, it is noticed that the well-polished and uniform C-steel surface is obtained by the addition of 500 ppm marjoram extract where a completely smooth, even, and uniform surface is obtained, and this behavior confirmed the data obtained from the polarization and weight loss method since marjoram extract records highest % inhibition.

### 3.7.2. Surface Roughness.

The proliferation and morphology of C-steel in the 8 M H₃PO₄-free solution and 8 M H₃PO₄ containing several plant extract were examined. According to Table 4, surface roughness, Ra, peak to valley ratio, and Rq parameters for carbon steel samples on the 8 M H₃PO₄-free solution and 8 M H₃PO₄ containing several plant extract were represented. The same trend of Ra is observed for peak to valley ratio and Rq parameters.

Table 4: Measured roughness (Ra), Rq (RMS), Peak-valley ratio, and brightness of C-steel samples.

| Sample | EP conditions                  | Brightness (gloss value) | Peak-valley ratio (μm) | Ra (μm) | Rq (RMS) (μm) |
|--------|--------------------------------|--------------------------|------------------------|---------|---------------|
| A      | Before electropolishing        | 11.1                     | 4.8                    | 2.7     | 3.4           |
| B      | After EP without additives     | 16.0                     | 0.47                   | 0.52    | 0.66          |
| C      | After EP + 100 ppm guava leaves| 18.0                     | 0.40                   | 0.44    | 0.56          |
| D      | After EP + 500 ppm guava leaves| 24.5                     | 0.30                   | 0.28    | 0.32          |
| E      | After EP + 100 ppm chamomile   | 18.4                     | 0.36                   | 0.40    | 0.52          |
| F      | After EP + 500 ppm chamomile   | 27.0                     | 0.29                   | 0.23    | 0.23          |
| G      | After EP + 100 ppm coriander seeds | 19.2               | 0.34                   | 0.36    | 0.48          |
| H      | After EP + 500 ppm coriander seeds | 56.59           | 0.28                   | 0.21    | 0.22          |
| I      | After EP + 100 ppm marjoram    | 21.9                     | 0.31                   | 0.32    | 0.42          |
| J      | After EP + 500 ppm marjoram    | 64.9                     | 0.22                   | 0.17    | 0.20          |

Table 4: Measured roughness (Ra), Rq (RMS), Peak-valley ratio, and brightness of C-steel samples.

![Figure 11: Reflectance diagram in the absence and presence of methanolic plant extract at different concentrations.](image)
With respect to coriander seeds extract, Ra decreases from 2.7 μm to 0.36 μm and 0.21 μm for 100 and 500 ppm, respectively [28].

The roughness (Ra) value for carbon-steel specimen treated with 8 M H₃PO₄ solution containing 100 and 500 ppm of marjoram extract recorded 0.32 μm and 0.17 μm as a minimum Ra value obtained.

The extract molecules behavior can be attributed to the extract adsorption ability on the C-steel surface. It amplifies via growing extract concentration.

Roughness measurements confirm scanning electron microscope images and will also be confirmed in the following brightness measurements section.

3.7.3. Gloss Value (Brightness). The Vis-IR spectra of untreated specimen, C-steel electropolished in 8M H₃PO₄ (blank), and C-steel electropolished in 8M H₃PO₄ electrolyte containing 100 and 500 ppm of different plant extracts are shown in Figure 11.

It is seen that the electropolished specimens' reflectance is strongly affected by the type of plant extracts. It is clear that the reflectance of the specimen that is electro-polished at 8M H₃PO₄ increases compared with the untreated specimen, where the specular reflectance value before EP is 11.1 and after EP in 8M H₃PO₄ is 16. High reflection property is obtained when the different type of plant extracts is added to the electropolishing electrolyte. By addition of guava leaves extract, chamomile extract, coriander seeds extract, and marjoram extract, the degree of surface brightness and reflectance increased; Table 4.

In order to reduce the microirregularities and amplify the metal brightness, it is considered that it is essential to accomplish homogeneous formation and cations diffusion from the anode surface, on a microscale. It is credible that correct type and concentration of plant extract supply to control C-steel polishing in H₃PO₄ acid.

The adsorption enhancements and the normal anodic film action in improving the surface microstructure are credible from the four studied plant extracts.

4. Conclusions

(1) Addition of methanolic marjoram, coriander seeds, chamomile, and guava leaves extract to the electropolishing solution results in a lower limiting current.

(2) Qd increased in the presence of plant extract which is a good evidence of the strong retardation behavior of plant extract for the electrodissolution processes for C-steel.

(3) Activation parameters values confirm that the C-steel dissolution process was controlled by via diffusion and also it is endothermic in nature.

(4) According to SEM, UV-VIS-NIR spectra, and surface roughness easements, addition of moderate concentration (500 ppm) of methanolic plant extract to the electrolytic solution is highly effective to enhance C-steel surface quality.

(5) Reflection property is increased when the different types of methanolic plant extract are added to the electropolishing electrolyte. The addition of 500 ppm of marjoram coriander, chamomile, and guava leaves increased the degree of surface brightness and reflectance to 64.9, 56.59, 27, and 24.5, respectively, relative to electropolishing electrolyte-free solution 16.

(6) The roughness (Ra) decreased from 2.7 μm to 0.52 μm without addition of any material.

(7) Ra values are 0.28, 0.23, 0.21, and 0.17 μm in the presence of guava leaves, chamomile, coriander seeds, and marjoram [29].

Data Availability

The (limiting current table data and SEM data) data used to support the findings of this study are included within the article.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

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