In vitro antimicrobial screening of isolated ethyl acetate fraction from Heterophragma adenophyllum leaves.

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ABSTRACT

The present study aimed to screen the in vitro antimicrobial activity of ethyl acetate fraction isolated from the leaves of Heterophragma adenophyllum against Escherichia coli, Salmonella enterica, Staphylococcus aureus, Bacillus anthracis and Klebsiella pneumonia broad spectrum microorganism. The study was carried out by using standard nutrient broth for microbes and streptomycin was taken as a positive control. Ethyl acetate fraction used as a test sample and DMSO as a negative control. From the results of the zone of inhibition it was concluded that the ethyl acetate fraction was possesses in vitro antimicrobial activity while results of minimum inhibitory concentration, it was revealed that all bacterial strains were sensitive towards selected fraction for the study.

Keywords: Antimicrobial, Heterophragma adenophyllum, ethyl acetate fraction.

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INTRODUCTION

The *Heterophragma adenophyllum* plant selected for the study was a traditional medicinal tree occurring in both tropical and subtropical regions of the world. In India, it is found in the forest of Maharashtra, Gujarat, Rajasthan, and Assam. *Heterophragma adenophyllum* areal part is important for the prevention and treatment of various diseases. In traditional medicine, the leaves are used for topical treatment of skin diseases. Fruits of *Heterophragma adenophyllum* were cooked and Flowers were consumed as fresh food. The tree is extensively used in traditional medicine. As an ingredient in massage oils, it is supposed to ease muscular tension sparingly cultivated as an ornamental tree. The wood is elastic and is used for making bows in Burma, and also for furniture (katsagon). Folk medicinal uses of *Heterophragma adenophyllum* roots in Piles, constipation and also prescribed as a drink in viper bite.\(^1\)\(^-\)\(^3\)

\(\alpha\)-Lapachone was previously isolated from the wood of the Bignoniaceae tree *Heterophragma adenophyllum*. A new symmetric naphthoquinone dimer, dilapachone, and a novel asymmetric naphthoquinone dimer, adenophyllone were isolated from the heartwood of *Heterophragma adenophyllum*. The aim of the present study was to evaluate the ethyl acetate fraction of the leaves extract for antimicrobial study.\(^4\)\(^-\)\(^6\)

MATERIALS AND METHOD

**Collection of plant material**

Leaves of *Heterophragma adenophyllum* was obtained and collected from Baroda, Gujarat during April-May, the voucher specimen was authenticated (authen.06/2012/botany) and deposited in pharmacognosy laboratory of Maliba Pharmacy College, Uka Tarsadia University, Bardoli, Gujarat, India.

**Preparation of extract**

Leaves were collected and washed with water to remove soil and straw from the base. The leaves were shade dried and coarsely powered for further process. The powdered leaves of *Heterophragma adenophyllum* were extracted with methanol by using hot percolation method.\(^7\)

The extract was oven dried at low temperature and fractionated with petroleum ether, ethyl acetate, and methanol. The ethyl acetate fraction was separated and evaporated to dryness. Dried ethyl acetate fraction treated with 05% aqueous KOH solution. Then separate the organic layer and treated with 10% aqueous hydrochloric acid, this acid-base reaction was repeated twice and collected organic layer. Finally, the organic layer washed with water and collected. The organic
layer was concentrated to 50 ml and centrifuge at 6000 rpm, the supernatant fluid was collected and evaporated to dryness. Dry residues suspended to DMSO for further use.

**Microorganism used**

The gram-positive organism *Staphylococcus aureus, Bacillus anthracis, Klebsiella pneumonia*, and gram-negative organism *Escherichia coli* and *Salmonella enterica* bacteria were used for *in vitro* antimicrobial study. All microbes were maintained in sterile conditions and grown on nutrient broth.

**Preparation of slandered bacterial suspension.**

Antimicrobial activity of petroleum ether fraction of *Heterophragma adenophyllum* was carried out by determining the zone of inhibition through agar well diffusion method and calculating minimum inhibitory concentration through micro dilution assay method. After growth, some colonies of microbes were selected and transferred aseptically into the tubes and centrifuge fully after adding sterile saline water. The bacterial suspension thus obtained were compared with the 1% McFerland standard. McFerland standard was checked by using a spectrophotometer with a 1-cm light path. The absorbance at the wavelength 600 nm was found to be 0.129, which is near to standard 0.123.

**Determination of zone of inhibition:**

Five sets of six sterile agar nutrient plates were taken for the study for the zone of inhibition of the five microbes. Agar plates were incubated with respective test organisms. Three holes of 6 mm diameter in the media of each plate were bored. One hole was filled with streptomycin solution of 50 µg/ml concentration as the positive control, another hole with 500 µg/ml concentration of ethyl acetate fraction solution as a test while the third hole was filled with DMSO as kept for negative control. Plates were then incubated at 37°C for 24 hrs. After incubation plates were examined for the presence of zone of inhibition.

**Determination of minimum inhibitory concentration by micro dilution assay method**

Six dilutions of the fraction ranging from 500 – 3.9 µg/ml were prepared using two-fold serial dilution method. Standardized inoculation of microorganisms of 1% McFerland standard turbidity prepared 1:1000 (10^5 CFU/ml) by adding sterile saline. Diluted sterile bacterial suspension and ethyl acetate fraction of *Heterophragma adenophyllum* were added into the wells of micro titer plates. streptomycin (50 µg/ml) was used as positive control and DMSO used as negative control while ethyl acetate fraction used as a test sample and incubated at 37°C for 24 hrs. Microbial growth was determined at an absorbance at 600 nm using RT-2100 micro plate reader. The MIC
values were taken as the lowest concentration of the fraction in the wells of the micro titer plates that shows no turbidity of the wells in the plates.

RESULTS AND DISCUSSION

Results of antimicrobial activity of ethyl acetate fraction of *Heterophragma adenophyllum* leaves extract by the agar-well diffusion method is shown in table 1. From the results of the zone of inhibition, it was revealed that the ethyl acetate fraction possesses an efficient and strong antimicrobial activity against both the gram-positive and gram-negative bacteria. Results of the antimicrobial activity of ethyl acetate fraction by micro dilution method are shown in table 2. From the results of the minimum inhibitory concentration (table 2), it was shown that the almost all broad spectrum tested microorganism was found sensitive towards the petroleum ether fraction of *Heterophragma adenophyllum* leaves extract. Phytochemical and preparative thin layer chromatography shows that various compounds of terpenoidal and steroidal nature were present in this fraction. Further studies were required for separation and isolation of active phytochemicals from ethyl acetate fraction of the leaves.

**Table 1: Antimicrobial activity of ethyl acetate fraction of *Heterophragma adenophyllum* leaves extract by agar well diffusion method.**

| Concentration | Zone of inhibition (mm) | E. coli, | S. enterica, | S. aureus, | B. anthracis | K. pneumonial |
|---------------|-------------------------|---------|-------------|-----------|-------------|--------------|
| Positive control 50 µg/ml | 34 ± 0.37 | 21 ± 0.31 | 29 ± 0.19 | 31 ± 0.43 | 26 ± 0.09 |
| Test Control 500 µg/ml | 24 ± 0.14 | 21 ± 0.32 | 22 ± 0.28 | 21 ± 0.24 | 18 ± 0.23 |
| Negative control | NA | NA | NA | NA | NA |

NA= No activity, all values are mean ± standard deviation, N=3 (experiment in triplicate)

**Table 2. Antimicrobial activity of ethyl acetate fraction of *Heterophragma adenophyllum* leaves extract by micro dilution assay method.**

| Minimum inhibitory concentration (µg/ml) | E. coli, | S. enterica, | S. aureus, | B. anthracis | K. pneumonial |
|----------------------------------------|---------|-------------|-----------|-------------|--------------|
| Positive control 0.353 | 0.294 | 0.191 | 0.281 | 0.213 |
| Test control 147 | 143 | 135 | 223 | 87.5 |
| Negative control | NA | NA | NA | NA |

NA= No activity

CONCLUSION:

The separated ethyl acetate fraction of *Heterophragma adenophyllum* leaves extract demonstrated broad-spectrum antimicrobial activity against both gram positive and gram negative bacteria. This activity of fraction may be possible due to bioactive phytochemicals are presence in the fraction. Bioactive compound from the fraction can be identified and isolated for further use in the
development of an antimicrobial formulation for the treatment of various infections. Thus the present study significantly proves that the isolated fraction from the ethyl acetate extract of the *Heterophragma adenophyllum* can be beneficial against antimicrobial agents.

REFERENCES:

1. Mohammed R, *et al.* An ethnomedicinal, pharmacological and phytochemical review of some bignoniaceae family plants and a description of bignoniaceae plants in folk medicinal uses in Bangladesh. Advances in natural and applied sciences, 2010: 236-253.

2. Amir RJ, Pahup S, Sonak, Satoshi T. Novel Naphthoquinones from Heterophragma adenophyllum. Helvetica Chemia Acta, 2004; 87(4): 820-824.

3. Tripetch K, Ryoji K, Kazuo Y. Lignan and phenylpropanoid glycosides from Fernandoa adenophylla. Phytochemistry, 2001; 57: 1245–1248.

4. Pahup S, Lalit P, Krishna J. Lapachol and other constituents from the bignoniaceae. Phytochemistry, 1972; 2: 149.

5. Petr B, Vojtech A, Ladislav H, Rene K. Noteworthy secondary metabolites naphthoquinones-their occurrence, pharmacological properties and analysis. Current Pharmaceutical Analysis, 2009; 5: 47-68.

6. Willium A, *et al.* US Patent. Patent no-US 7,749,544 B2. 2010 July 6.

7. Surana VS, *et al.*, Hepatoprotective effect of cassia tora seeds on experimental animal model. Am. J. PharmTech Res, 2012; 2(2): 301-309.

8. Salma Ahmed Mahmoud El Sawi *et al.* Composition of the non-polar extracts and antimicrobial activity of Chorisia insignis HBK. Leaves. Asian Pac J Trop Dis., 2014; 4(6): 473-479.

9. Jalalpure SS, Agarwal N, Patil MB, Chimkode R, Tripathi A. Antimicrobial and wound healing activites of leaves of Alternanthera sessilis Linn. Int J Green Pharmacy 2008; 2(3): 141-44.

10. Peruru D. *et al.* Isolation of eumelanin from sepia officinalis and investigation of its antimicrobial activity by ointment formulation. Int. J. Pharm. 2012; 2(2): 67-72.

11. Tamokou *et al.* Antioxidant and antimicrobial activities of ethyl acetate extract, fractions and compounds fromvstem bark of *Albizia adiantifolia* (Mimosoideae). BMC Complementary and Alternative Medicine, 2012; 12:99.

12. Singh B, Singh S: Antimicrobial activity of terpenoids from Trichodesma amplexicaule Roth. Phytother Res, 2003; 17:814–816.
13. Ushimaru, P.I. et al. Antibacterial activity of medicinal plant extracts. Brazilian Journal of Microbiology, 2007; 38:717-719.

14. Geone Maia Correa et al. Anti-inflammatory and antimicrobial activities of steroids and triterpenes isolated from aerial parts of Justicia acuminatissima. Int J Pharm Pharm Sci, 2014; 6(6):75-81.

15. B. Mahesh and S. Satish. Antimicrobial Activity of Some Important Medicinal Plant against Plant and Human Pathogens. World J. Agric. Sci., 2008; 4(S): 839-843.