Intra-Regional Glu-GABA vs Inter-Regional Glu-Glu Imbalance: A 1H-MRS Study of the Neurochemistry of Auditory Verbal Hallucinations in Schizophrenia

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Glutamate (Glu), gamma amino butyric acid (GABA), and excitatory/inhibitory (E/I) imbalance have inconsistently been implicated in the etiology of schizophrenia. Elevated Glu levels in language regions have been suggested to mediate auditory verbal hallucinations (AVH), the same regions previously associated with neuronal hyperactivity during AVHs. It is, however, not known whether alterations in Glu levels are accompanied by corresponding GABA alterations, nor is it known if Glu levels are affected in brain regions with known neuronal hypoactivity. Using magnetic resonance spectroscopy (MRS), we measured Glx (Glu+glutamine) and GABA+ levels in the anterior cingulate cortex (ACC), left and right superior temporal gyrus (STG), and left inferior frontal gyrus (IFG), in a sample of 77 schizophrenia patients and 77 healthy controls. Two MRS-protocols were used. Results showed a marginally significant positive correlation in the left STG between Glx and AVHs, whereas a significant negative correlation was found in the ACC. In addition, high-hallucinating patients as a group showed decreased ACC and increased left STG Glx levels compared to low-hallucinating patients, with the healthy controls in between the 2 hallucinating groups. No significant differences were found for GABA+ levels. It is discussed that reduced ACC Glx levels reflect an inability of AVH patients to cognitively inhibit their “voices” through neuronal hypoactivity, which in turn originates from increased left STG Glu levels and neuronal hyperactivity. A revised E/I-imbalance model is proposed where Glu-Glu imbalance between brain regions is emphasized rather than Glu-GABA imbalance within regions, for the understanding of the underlying neurochemistry of AVHs.

Key words: Magnetic resonance imaging (MRI)/MR spectroscopy (MRS)/schizophrenia/hallucinations/the neurochemistry of auditory verbal hallucinations/glutamate/GABA/Glx/excitatory/inhibitory (E/I) imbalance model

Introduction

Auditory verbal hallucinations (AVHs) is a key symptom in schizophrenia, and refers to auditory experiences of “hearing a voice,” in the absence of an external auditory source. Neuronal underpinnings of AVHs are not fully understood, although imbalance between excitatory and inhibitory influences (E/I imbalance) in the brain have been proposed as potential mechanisms (ref. for a review), and appear to exist on multiple levels including large-scale cognitive networks (between regions) and small-scale neuronal circuits (within regions). For large-scale cognitive networks, evidence suggests E/I imbalance between language and cognitive control regions in schizophrenia patients. fMRI meta-analyses have suggested (hyper-)activation during AVHs in a bilateral frontal-temporal network including speech perception (superior temporal gyrus [STG]) and speech production (Broca’s area in inferior frontal gyri [IFG]) areas. Furthermore, a consistent finding in schizophrenia patients is hypoactivity in prefrontal cortex (eg, anterior cingulate cortex, ACC). These findings go along with findings of impaired cognitive control/executive functions and regulation of attention in these patients. Based on findings from behavioral and neuroimaging studies, Hugdahl proposed a 2-fold model of the pathophysiology underlying AVHs involving neuronal hyper-excitation in temporal lobe.
speech perception regions leading to the perception of a “voice” in a bottom-up way, and frontal lobe top-down failure in suppressing the bottom-up input (c.f.11,12). The aim of the present study was to investigate any underlying neurotransmitter excitatory-inhibitory correspondence to the bottom-up and top-down imbalance found at the cognitive/behavioral level of explanation (cf.13).

Dysfunction of glutamatergic and/or gamma amino butyric acid (GABA)-ergic neurotransmitter systems is implicated in the etiology of schizophrenia, and could result in regional imbalances of excitation and inhibition. Dysfunction of glutamatergic transmission has been attributed to hypo-function of the N-methyl-D-aspartate (NMDA)-type glutamate (Glu) receptor, and NMDA receptor dysfunction on GABAergic neurons could lead to disinhibition and increase of Glu release.14 Administration of the NMDA antagonist ketamine to healthy volunteers has produced symptoms comparable to that of schizophrenia patients15-17 and exacerbated psychotic symptoms (including auditory hallucinations) in schizophrenia patients.17,18 In support of deficits in GABAergic neurotransmission is the consistent post-mortem finding of lowered expression of glutamatergic acid decarboxylase 67 (GAD67) mRNA (eg,19-21), which codes for a protein involved in the conversion of Glu to GABA (for a review see ref.22). In vivo studies of altered Glu and GABA levels are further needed in confirming a Glu-GABA imbalance hypothesis of schizophrenia.

Glu and GABA concentration levels can be measured in vivo by hydrogen magnetic resonance spectroscopy (1H-MRS). MRS takes advantage of the magnetic properties of the hydrogen proton. The surroundings of the hydrogen proton(s), ie, the molecule in which the proton is bound, affect its magnetic properties.23 As a result, it is possible to identify signals from different molecules. Multiple studies have investigated Glu and GABA concentrations in schizophrenia patients applying MRS, although results are inconsistent. A recent meta-analysis of Glu24 concluded of generally elevated Glu, glutamine (Gln), or of the composite signal, Glx, in sub-cortical regions in schizophrenia patients. For the ACC, significantly elevated Glx levels were found only for a high-risk population. A recent meta-analysis on GABA25 concluded with no overall difference between patients and controls in spite of single studies confirming low GABA levels in patients in the ACC.26,27 One reason for inconsistent results in MRS studies might be different symptom profiles in patients across samples.24

Only a few studies have investigated Glu in relation to auditory hallucinations. Elevated Glu levels have been found associated with auditory hallucination severity in the left STG,28 and left IFG.29,30 Whether these hallucination-related Glu alterations are accompanied by GABAergic changes within the same regions—as predicted by the E/I imbalance model—has not yet been investigated. It should also be noted that both left STG1,4 and left IFG4 have been related to neuronal hyperactivity during hallucinations in schizophrenia patients. Whether similar relationships exist between AVH and Glu in regions related to hypo-activity is unknown. In fact, one might consider that there also exist Glu-Glu imbalances and/or GABA-GABA imbalances between regions associated with respectively hyper- and hypo-activity, but this remains to be tested.

The current study, therefore, aimed to explore whether Glu-GABA imbalances are associated with severity of AVHs in schizophrenia patients in language (left and right STG, and left IFG) and executive (ACC) regions. The left and right STG and ACC regions were examined with Protocol 1 (n = 39); and the left IFG and STG were examined with Protocol 2 (n = 38). The 2 MR protocols used did not differ with regard to the aims of the study. We hypothesized that relationships between AVH and Glu and GABA levels would be region-specific, such that positive Glu and negative GABA correlations with AVH would be seen in the STG and IFG regions, reflecting increased excitation (hyperactivity) in the language regions, while the reverse relationship should be found for the ACC region reflecting decreased excitation (hypoactivity) in executive/cognitive control regions.

Methods

Participants

Seventy-seven schizophrenia patients (mean age 29.83, SD 11.48) and 77 matched controls (mean age 30.23, SD 10.23) underwent MRS scanning (some tested repeatedly at 2 different time points, see table 1 for sample details). The patients were recruited via the Bergen Psychosis Study 2. Two MRS data collection protocols, including different voxel placements, were used. One protocol (hereafter referred to as Protocol 1) included MRS recordings from the left STG, right STG, and ACC and was used for 39 patients and 39 controls. The second protocol (hereafter referred to as Protocol 2) included MRS recordings from the left STG and left IFG and was used for 38 patients and 38 controls (see figure 1 for voxel placement). Data from the 2 protocols were pooled in the analyses.

The controls were individually matched to the patients with regard to sex and age (within ±3 y from their respective controls), except for 9 patients who were outside of the 3 years range (4–7 y). Patients and controls were in addition matched in terms of scanning protocol, and handedness, with the exception of ambidextrous patients who were matched with right-handed controls. All patients that were on medication used second-generation antipsychotic medication while some, in addition, used first-generation antipsychotics (mean defined daily dose (DDD):1.03, SD 0.62). A few patients also used anti-depressants (n = 10), mood stabilizers (n = 2), opioids (n = 1), benzodiazepines (n = 18), anticholinergic (n = 5), or ADHD medication (n = 1). All patients were diagnosed with schizophrenia spectrum disorder according to the ICD-10 diagnostic
Table 1. Regional Concentration Means and SDs of Glx, Glutamate, Glutamine, GABA (Institutional Units), and Data Quality Parameters of the Patients and Controls That Were Included in Data Analysis

|       | Patients |          |            |         |          |
|--------|----------|----------|------------|---------|----------|
|        | Mis/ Excl. | Mean     | SD         | Mis/ Excl. | Mean     | SD     |
| LSTG P |                        | 15.73    | 4.10       | 15.87    | 4.19      |
| Glx    | 92 (76)   | 13.23    | 2.13       | 60       | 13.61    | 2.0   |
| Glutamate | 81 (67)      | 3.60    | 2.54       | 60       | 4.06    | 2.63  |
| Glutamine | 81 (67)      | 39.55   | 12.17      | 77       | 40.81   | 12.28 |
| SNR    | 92 (76)   | 0.07     | 0.02       | 77       | 0.07    | 0.02  |
| FWHM   | 92 (76)   | 4.95     | 0.97       | 75       | 4.82    | 1.08  |
| MP     | 83 (68)   | 21.51    | 5.94       | 75       | 20.53   | 6.36  |
| GABA   | 83 (68)   | 0.07     | 0.02       | 75       | 0.07    | 0.02  |
| SNR    | 83 (68)   | 0.07     | 0.02       | 75       | 0.07    | 0.02  |
| LIFG P |                        | 16.93    | 2.86       | 38       | 16.18   | 3.23  |
| Glx    | 48 (37)   | 14.28    | 1.88       | 34       | 14.35   | 1.8   |
| Glutamate | 46 (36)      | 3.12    | 1.64       | 34       | 2.75    | 1.5   |
| Glutamine | 46 (36)      | 42.48   | 9.02       | 38       | 47.29   | 6.79  |
| SNR    | 48 (37)   | 0.06     | 0.01       | 38       | 0.05    | 0.01  |
| FWHM   | 48 (37)   | 27.68    | 3.93       | 38       | 28.45   | 4.16  |
| SNR    | 47 (37)   | 0.06     | 0.02       | 38       | 0.06    | 0.02  |
| MP     | 47 (37)   | 20.18    | 3.33       | 37       | 19.83   | 3.59  |
| GABA   | 47 (37)   | 17.09    | 1.80       | 33       | 16.55   | 1.69  |
| SNR    | 46 (37)   | 4.22     | 2.39       | 33       | 4.31    | 2.28  |
| FWHM   | 46 (37)   | 40.51    | 7.26       | 37       | 41.46   | 6.03  |
| SNR    | 43 (38)   | 0.06     | 0.02       | 37       | 0.06    | 0.01  |
| MP     | 43 (38)   | 4.44     | 0.83       | 38       | 4.59    | 0.82  |
| GABA   | 42 (37)   | 18.41    | 6.15       | 38       | 19.05   | 5.67  |
| SNR    | 42 (37)   | 0.05     | 0.01       | 38       | 0.06    | 0.01  |
| MP     | 42 (37)   | 22.30    | 3.25       | 38       | 21.14   | 4.31  |
| GABA   | 42 (37)   | 16.40    | 1.96       | 35       | 15.34   | 2.51  |
| SNR    | 40 (35)   | 6.95     | 3.49       | 35       | 7.08    | 3.66  |
| SNR    | 40 (35)   | 33.71    | 9.51       | 38       | 31.76   | 8.13  |
| MP     | 37 (32)   | 14.05    | 4.93       | 37       | 14.22   | 4.84  |
| SNR    | 37 (32)   | 0.08     | 0.03       | 37       | 0.09    | 0.03  |

Note: LSTG, left superior temporal gyrus; LIFG, left inferior frontal gyrus; ACC, anterior cingulate cortex; RSTG, right superior temporal gyrus; P, PRESS sequence; MP, MEGA-PRESS sequence; Mis, missing spectra; Excl, excluded spectra; SNR, signal-to-noise ratio; FWHM, full-width at half maximum (linewidth). N refers to number of observations (as some patients were tested repeatedly at 2 sessions), and in parentheses the number of unique subjects. Number of participants differ across regions and groups (patients and controls) due to missing or excluded (low quality) spectra for single measurements. N also differ between Glx and glutamate/glutamine due to cases where the composite signal was inseparable.

Mean illness duration for the patient group was 4.34 years, SD 7.68. Global severity of symptoms in the patient group as assessed by the Positive and Negative Syndrome Scale (PANSS) total score were 62.48, SD 17.63 (Positive-total 15.53, SD 5.51; Negative-total 15.03, SD 5.02; General-total 31.91, SD 9.70). The study was approved by the Regional Committee for Medical Research Ethics at the University of Bergen (REK no 2010–3387), and conducted according to the Declaration of Helsinki. All participants received oral and written information about the study before signing a written consent form.

Hallucinations were assessed with the PANSS interview. All PANSS-raters were trained and certified by the PANSS Institute, and satisfactory inter-rater reliability was documented. Severity of auditory hallucinations were scored from the P3 item on the PANSS positive sub-scale, ranging from 1 to 7 (1 = absent, 7 = extreme). Although the PANSS questionnaire does not explicitly distinguish between modalities, hallucinations in the auditory domain (as “voice hearing”) is most common, and the P3 item is hereafter referred to as AVH. The PANSS scores were obtained on the same day as the MRS measurements.

**MR Image Acquisitions**

Imaging data were acquired with a 3T GE-SignaHDx MRI scanner. See table 2 for MR acquisition parameters. Unsuppressed water reference spectra (8 repetitions)
were acquired automatically after the acquisition of water-suppressed metabolite spectra in both protocols. A scanner upgrade was conducted between data collection of Protocol 1 and 2, including a change of head-coil from 8 to 32 channels.

Data Analysis

MRS data from the PRESS sequence were analyzed using the LCModel version 6.3-1J,\textsuperscript{33} with the standard basis-set incorporating components from 15 metabolites (Alanine, Aspartate, Creatine, $\gamma$-amino-butyric acid, Glucose, Gln, Glu, Glycerophosphorylcholine, Phosphorylcholine, Lactate, myo-inositol, $N$-acetylaspartate, $N$-acetylaspartylglutamate, scyllo-inositol, and Taurine). Basic processing of MEGA-PRESS data was achieved using in-house scripts to perform coil combination, phase correction, alignment, and residual water subtraction before subtracting edit-OFF from edit-ON parts to yield the final difference spectrum for each acquisition. This spectrum was then quantified in LCModel with the mega-press-3 spectra type, using a simulated basis set\textsuperscript{34} with Kaiser coupling constants.\textsuperscript{35} Metabolite estimates were scaled to an internal water reference, accounting for differing water concentration in the different tissue classes (GM, WM, CSF), partial volume effects, and metabolite relaxation times and differing water relaxation times between the tissue classes.\textsuperscript{36} Tissue content within the MRS voxel was estimated from the T1-image using the segmentation tool of the Statistical Parametric Mapping (SPM8) software (www.fil.ion.ucl.ac.uk/spm). Individual spectra were subject to quality control, which led to the rejection of inadequate spectra (see Table 1 for excluded spectra, and spectral quality parameters, and supplementary figure S1 for spectrum examples and information on quality control procedure). For further details on analysis procedure, see ref.\textsuperscript{37} Glx and GABA+ (GABA including macromolecules) levels were used in statistical analyses.

Statistical Analysis

Glx and GABA+ values were subjected to statistical analysis using Linear mixed models available in the SPSS software package (https://www.ibm.com/analytics/).

Table 2. MR Acquisition Parameters for Protocol 1 and 2

| N | Regions          | Sequence   | TR  | TE  | TI  | FOV | Rep   | Voxel size (m$^3$) |
|---|------------------|------------|-----|-----|-----|-----|-------|-------------------|
|   |                  |            |     |     |     |     |       |                   |
| Protocol 1 | 39 | T1 MRS | Whole brain | 3DSPGR | 7.74 | 2.9  | 500  | 260  | 1 $\times$ 1 $\times$ 1 |
|            |     |         | LSTG,RSTG, ACC | PRESS | 1500 | 35   | 128  | STG: 24 $\times$ 40 $\times$ 30 |
|            |     |         |           |       |      |      |       |       | ACC: 40 $\times$ 25 |
| Protocol 2 | 38 | T1 MRS | Whole brain | MEGA-PRESS | 1500 | 68   | 128  | 24 $\times$ 30 $\times$ 31 |
|            |     |         | LSTG, LIFG | PRESS | 6.8  | 2.95 | 450  | 256  | LIFG: 24 $\times$ 38 $\times$ 28 |

Note: LSTG, left superior temporal gyrus; LIFG, left inferior frontal gyrus; ACC, anterior cingulate cortex; RSTG, right superior temporal gyrus; PRESS, point-resolved spectroscopy sequence; MEGA-PRESS, Mescher-Garwood PRESS; TR, repetition time; TE, echo time; TI, inversion time; FOV, field of view; Rep, repetitions. The repetitions for the MEGA-PRESS acquisition refer to the number of edit ON/OFF pairs acquired.
A VH score was entered as a regressor variable, while par-
smodels, Region was entered as a repeated fixed factor, and
between regional metabolites and severity of A VHs. In these
run for patients only in order to test the linear relationship
models with Glx and GABA+ as dependent variables were
which resulted in 3 levels for the Group factor (A VH+, A VH-,
Fisher's LSD post hoc test was used to fol-
 cant differences between all 4 regions (all $P < .001$), with
$\text{F}(3,305.5) = 0.88$, $P = .26$), and interaction-effect of Group × Region
showed a significant difference ($P = .04$). There was also a nonsignificant trend for the AVH- sub-group to show higher Glx levels compared to the controls ($P = .07$).

The next set of analyses involved patients only and
and assessed the linear relationship between AVH and Glx. Again, there was a significant main-effect of Region, ($F(1,171.78) = 20.56$, $P < .001$), while the main-effect of AVH was nonsignificant ($F(1,125.4) = 0.05$, $P = .83$, respectively). However, the interaction of Region and AVH
severity was significant ($F(3,170.1) = 4.38$, $P = .005$). Post hoc analyses showed that the interaction was driven by a marginally significant positive correlation between Glx
levels in the left STG and P3 score ($P = .054$, $B = 0.46$), and a significant negative correlation between Glx and the P3 score in the ACC ($P = .04$, $B = 0.56$; see figure 3).

AVHs were not significantly correlated with Glx levels in the right STG ($P = .93$, $B = -0.14$) or in the left IFG ($P = .79$, $B = -0.06$).

Total negative symptom score was not found related to Glx.

GABA+$A$ trend of a negative linear relationship between total
symptom score and GABA+ was found ($F(1,84.99) = 3.5$, $P = .065$) across regions.

Discussion

The current study investigated the neurochemical under-
pinnings of AVH by addressing Glu-GABA imbalances
within selected brain regions, and Glu-Glu and GABA-GABA imbalances between regions previously associated with hyper- and hypo-activity during AVH experience. No overall differences between patients and controls were found. However, significant differences were found between patient groups, where AVH+ patients showed lower ACC and higher left STG Glx levels relative to the AVH- group (figure 2). The control group showed Glx levels which were in between those of the AVH+ and AVH- patients. These findings were confirmed by correlation analysis, where an interaction was found between brain region and AVH severity. The interaction was driven by a negative relationship between ACC Glx and the AVH score, whereas a positive relationship was shown between left STG Glx and the AVH score (figure 3).

**Glu Levels: STG and IFG Voxels**

As hypothesized, Glx in the left STG was positively associated with AVH severity. Although the correlation was marginally significant, previous findings by Hugdahl et al. and Curčić-Blake et al. suggest that an a priori hypothesis could be formulated regarding the direction of the correlation. Therefore, a one-sided test could have been argued for and would, in this case, shown a significant effect.

In this sense, the present results extend the findings by Hugdahl et al. on a larger sample. The results are also in line with previous fMRI findings showing increased activation in the left STG during AVH experiences. This is not surprising when seen in the light of functional MRS studies that show significant increase in Glx levels during stimulus processing and task execution. In this sense, left STG Glx levels might cause neuronal hyper-excitation that initiate the perceptual experience of “hearing a voice.” In spite of a positive association between left STG Glx and AVH severity, the patients did not show overall higher Glx levels as compared to controls. One could, therefore, speculate that the increased neuronal activity associated with AVHs elevates Glx to “normal levels” corresponding to that of healthy individuals. This is further supported by the finding of lowest Glx levels in AVH+ group, also lower than the controls. It should, however, be noted that neuronal activity was not measured. We can therefore not rule out that Glx would rather be associated with hypo-activity due to neuronal loss as a consequence of neurotoxic effects of Glx. However, since AVHs have previously been associated with left STG hyperactivity, we find the results of a positive relationship between Glx and AVHs to be unlikely if Glx would cause hypo-activity.

Interestingly, the positive association between Glx and AVH experiences was restricted to the left STG, suggesting a laterality effect implicating more the left than the right hemisphere. This result is in line with several fMRI meta-analyses showing left, but not right, STG activations in patients with AVHs. Since the left hemisphere is the dominant hemisphere for language processing and since AVH are typically experienced as “voices,” an increase in Glx levels in left STG only is a feasible finding.

There was a lack of significant findings for the left IFG, which contradicts 2 previous MRS studies: Hugdahl et al. and Curčić-Blake et al. both found lower Glx levels in patients relative to controls, and higher Glx levels in hallucinating relative to non-hallucinating patients. Several methodological differences between the studies could explain the discrepancies in results, such as voxel placement (more medial and anterior in former studies), or sample characteristics such as lower P3 scores in Hugdahl et al., and AVH assessed as a trait measure in

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**Fig. 2.** Glutamate and GABA concentration levels are shown on the y-axes for healthy controls, low-hallucinating patients (AVH-), and high-hallucinating patients (AVH+) in the 4 brain regions. Error bars indicate standard error. * indicates significant post hoc comparisons at $P < .05$, conducted to explore the significant interaction of Group and Region. STG, superior temporal gyrus; IFG, inferior frontal gyrus; ACC, anterior cingulate cortex; IU, institutional units.
Ćurčić-Blake et al. as opposed to a state measure in the current study.

**Glx Levels: ACC Voxel**

As hypothesized, ACC Glx levels were found to be negatively correlated with AVHs. This could be related to neuronal hypo-activity and reduced executive and cognitive control functions that has been observed in hallucinating schizophrenia patients. A possible interpretation is that patients with low Glx levels lack the cognitive resources necessary to inhibit the “voices” originating from the left STG, due to top-down hypo- and bottom-up hyper-excitation. Interestingly, no association was found between total negative symptom score and Glx levels in ACC, nor in any of the other regions, highlighting the specificity of AVHs symptoms in predicting Glx concentrations.

The finding of no overall ACC Glx level difference between patients and controls adds to previous inconsistent
findings, with some studies showing higher Glx levels in patients relative to controls, other showing lower levels, and others again reporting no difference. However, when taking AVH into account, a significant group-effect appeared, driven by differences between the 2 patient sub-groups (AVH+ and AVH-). There was also a trend towards higher Glx levels in the AVH- group compared to the control group.

Thus, as for both the ACC and left STG regions, the healthy controls showed Glx levels in between the levels for the AVH+ and AVH- sub-groups. The ACC results, therefore, suggest that one reason for inconsistencies in previous studies when comparing schizophrenia patients and controls could, at least partly, be due to inter-individual heterogeneity in symptom severity, in particular, severity of AVHs. AVH should, therefore, be reported or controlled for in future MRS studies.

**GABA+ Levels: All Voxels**

GABA+ levels were not found to differ between patients and controls, nor were GABA+ levels associated with AVHs. Previous GABA MRS studies have shown inconsistent results, in spite of consistent postmortem findings of reduced GAD67 mRNA production. The current study showed that even when inter-individual and inter-regional differences were taken into account, the measurements failed to show GABAergic alterations in the patients. de Jonge et al suggested that other subclasses of GABAergic neurons might compensate for the reduced GABA production in parvalbumin-neurons, or could be stimulated by increased glutamergic activity. This would be a plausible explanation for the left STG findings where Glx was elevated in AVH+ patients.

**Intra-Regional vs Inter-Regional E/I-Imbalances**

The current study investigated the underlying neurochemistry in terms of the E/I imbalance model for AVHs. While the results could not unambiguously confirm intra-regional Glu-GABA imbalances due to the lack of significant GABA findings, we found an inter-regional Glu-Glu imbalance associated with AVH severity. We thus propose a revised and extended E/I-imbalance model where Glu-Glu imbalance between brain regions may be more relevant than Glu-GABA imbalance within regions, for the understanding of the underlying neurochemistry of AVHs.

The suggested region-specificity regarding the relation between Glx and AVH give rise to questions of underlying mechanisms. It is increasingly accepted that NMDA receptor deficiency should result in elevated Glu and reduced GABA production in schizophrenia patients. If NMDA receptor deficiency, as eg, indicated by compensatory up-regulation of NMDA receptor subunits, underpin the results of the current study, they are likely to be region-specific, and Glx alterations seen in other regions secondary. One possibility is that dysfunction of NMDA-receptors in the left STG causes neuronal hyperactivity and elevated Glx levels, leading to the perceptual experience of “hearing a voice.” In a non-dysfunctional brain, the ACC would be similarly activated to suppress or inhibit the “voices” to enter awareness. In the hallucinating brain, this top-down regulation might be upset because of low ACC Glx levels, together with, or as a consequence of, “strong” bottom-up perceptual impulses, resulting in AVH. It should, however, be remembered that MRS do not measure NMDA receptor function per se, and therefore further studies applying, eg, PET-MR, may be instrumental in disentangle cause and effect with regard to the underlying neurochemistry of AVH. Another limitation of the current study is that different numbers of patients were assessed in the 2 regions, and therefore, it remains undetermined whether the regional alterations co-exist in the same patients. With these limitations in mind, the findings may nevertheless point in new directions for the development of symptom-specific targets for pharmacological treatment interventions.

**Supplementary Material**

Supplementary material is available at Schizophrenia Bulletin online.

**Figure S1:** Examples of PRESS and MEGA-PRESS spectra from one patient and one control subject in each of the four regions. Abbreviations: STG=superior temporal gyrus, IFG=inferior frontal gyrus, ACC=anterior cingulate cortex. All spectra were subjected to quality control, before included in data analysis. A local algorithm assigned a quality score to each spectra, considering the following factors: spectral linewidth (FWHM), signal-to-noise ratio (SNR) and CRLB %SD of key metabolites in the spectrum, the magnitude of aberrant features in the individual metabolite spectrum relative to the group mean, and the magnitude of features in the residuals after fitting. This quality score does not impose strict rejection criteria (which in some instances could risk introducing systematic biases), but rather flags aspects of spectra of possible concern to guide visual inspection, after which spectra deemed to be of insufficient quality for meaningful assessment were rejected.

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