Augmented mast cell infiltration and microvessel density in prostate cancer

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Introduction

Prostate cancer is the most threatening and very often fatal malignancy in elderly men. There are over 540,000 new cases diagnosed each year worldwide [1]. The mortality rate associated with prostate carcinoma ranks second in Poland and first in the USA [2]. Searching for new methods of cancer therapy and prognosis is a key task for many researchers around the world.

The classic, widely used prognostic factors for prostate cancer include the prostate-specific antigen (PSA) levels in serum, histological grading – Gleason score – and clinical staging, using the TNM (tumour-node-metastasis) system. Some investigations, carried out during recent years, have taken into account other important prognostic factors, such as mast cell infiltration and their influence on tumour angiogenesis [3–6].

Neoplastic proliferation is almost always accompanied by inflammatory infiltrates: neutrophils, lymphocytes, macrophages and mast cells. The mast cells play an important role in growth regulation of many tumours. One of these tumours is prostate cancer, but the role of mast cells in this particular tumour is a rather controversial issue. Mast cells belong to inflammatory regulators. Inflammation is a defence line against tumour, and inflammatory infiltrates may lead to better outcomes in tumour-affected patients. On the other hand, mast cells produce many bioactive mediators and growth factors, including fibroblasts and angiogenic growth factors, such as FGF-2 (fibroblast growth factor-2), VEGF (vascular endothelial growth factor), TGF-β (transforming growth factor-β) and IL-8 (interleukin-8) [3]. These mediators play a crucial role in promoting tumour angiogenesis and metastasis, and thus are associated with poor prognosis for affected patients. Also mast cells produce proteolytic enzymes, e.g., specific tryptase and matrix metalloproteinases (MMP-2, MMP-9) [4], which may facilitate cancer stromal invasion.

The angiogenic potential of mast cells, indicated in some studies [3–9], is reflected by increased microvessel density in prostate cancer.

In view of the above-mentioned observations, the aim of our study was to compare mast cell infiltration and microvessel density, as expressed by CD-34 immunoreactivity, in prostate cancer formation and in benign prostate hyperplasia. We also attempted to find possible relationships among mast cell infiltration and microvessel density, Gleason score and serum PSA levels.

Material and methods

Patients

The investigation was confined to evaluations of materials from prostate needle biopsies, carried out in 26 patients, diagnosed at the Provincial Hospital in Skierniewice, Poland, during the years 2008–2010. In all the cases, elevated serum PSA levels indicated the need of biopsy. The mean ± SD (standard deviation) age of the patients was 69.95 ± 6.22 years, PSA levels varied from 4.03 ng/ml to 100 ng/ml, and the mean PSA level was 37.47 ± 32.32. Gleason score varied from 4 to 9. All the patients of that group were diagnosed...
with prostate cancer, using light microscopy and the standard haematoxylin-eosin staining method. Fourteen specimens of prostate gland, diagnosed as benign hyperplasia, were used as controls. In all the control patients, serum PSA levels of PSA were within normal limits (from 0.9 ng/ml to 3.9 ng/ml, mean 2.2 ±18).

Immunohistochemistry

Paraffin-embedded tissue sections were mounted onto Super-Frost slides, deparaffinised, treated in a microwave oven with a solution of TRS (Target Retrieval Solution, pH 6.0, Dao) for 30 minutes (2 × 6 minutes 360 W, 2 × 5 minutes 180 W, 2 × 4 minutes 90 W) and transferred to distilled water. Endogenous peroxidase activity was blocked by 0.3% hydrogen peroxide in distilled water for 30 minutes and then the sections were rinsed with Tris-buffered saline (TBS, Dako, Denmark) and incubated for 30 minutes with primary mouse monoclonal antibodies against mast cell tryptase (Dako; clone AA1, dilution 1 : 50) and CD34 antigen (Dako; clone QBend10, dilution 1 : 25). Immunoreactive proteins were visualized using an EnVision-HRP kit (Dako, Carpenteria, CA, USA), according to the manufacturer’s instructions. Visualisation was performed by incubation of the sections in a solution of 3,3’-diaminobenzidine (DakoCytona tion, Denmark). After washing, the sections were counter-stained with haematoxylin and coverslipped. A negative control was processed for each antibody and for each sample. Negative controls were carried out by incubation in the absence of the primary antibody and always yielded negative results.

Morphometry

Histological morphometry was performed by means of an image analysis system, including a PC with a Pentagram graphical tablet, an Indeo Fast card (frame grabber, true-colour, real-time), produced by Indeo (Taiwan), and a Panasonic colour TV camera (Japan), coupled with a Carl Zeiss microscope (Germany). On that hardware the (MultiScan 8.08 program of Computer Scanning Systems, Poland) was installed to calculate the number of objects (semiautomatic function). The colour microscopic images were serially saved in the computer memory, followed by quantitative examinations. The numbers of tryptase-positive mast cells and of CD34-positive vessels were determined by counting all the positive cells and vessels (semiautomatic function) in a sequence of ten consecutive computer images of 400 × high power fields – 0.0047 mm² each. Only vessels with an evident lumen were counted. Mast cells were scored positive when displayed cytoplasmic granules stained positively with anti-tryptase monoclonal antibody. The results were expressed as the mean number of immunopositive cells or vessels per mm².

Table 1. Quantitative data of mast cell infiltrates and microvessel density in prostate cancer and controls

| Number of cases | Controls (n = 14) | Prostate cancer (n = 26) | P value |
|-----------------|-----------------|------------------|--------|
| Mast cells (per 1 mm²) | 72.82 ±55.6 | 123.73 ±82.32 | < 0.05 |
| Microvessel density* | 67.72 ±28.5 | 179.43 ±72.52 | < 0.001 |

*Microvessel density as mean number of CD34-positive vessels per 1 mm²

Statistical methods

All the obtained values were expressed as means ± SD. Differences between groups were tested by the Mann-Whitney U test. Correlation coefficients were calculated by Spearman’s method. Results were considered statistically significant if p < 0.05.

Results

Table 1 presents quantitative data of mast cell infiltration and of microvessel density for the patients with prostate cancer and for the controls. Regarding the former, both mast cell infiltrates and microvessel density (MVD) were significantly increased, when compared to respective values in the control patients (Figs. 1–4). Furthermore, our correlative study (see Table 2) revealed significant positive correlations between the mean number of mast cells and microvessel density both in the group with prostate cancer and in the control group. In the patients with prostate cancer, significant positive correlations were observed between the mean number of tryptase-positive cells and Gleason score, as well as between microvessel density and Gleason score. The correlations between the number of mast cells and PSA levels and between microvessel densities and PSA levels were positive but they did not reach statistical significance. In the controls, those correlations were weak and not significant, either.

Discussion

In our investigations, we tried to unveil the role of mast cell infiltration in the progression of prostate cancer and its associations with classic prognostic factors, such as Gleason score and serum PSA levels. An increase of mast cell infiltration and microvessel density was observed in the patients with prostate cancer, as compared to those with benign hyperplasia. That finding suggests a stimulating role of mast cells in tumourigenesis, resulting from the angiogenic activity of mast cells. Enhanced angiogenesis is associated with degradation of mast cells, which releases biological mediators, including the regulators of vascular functions, such as FGF-2, VEGF and TGF-β [10, 11]. The formation of new vasculature is a crucial process for tumour growth and progression. One of the major regulators in this process is VEGF.

Table 2. Correlations between selected parameters in prostate cancer and controls

| Correlation : between | Controls (n = 14) | Prostate cancer (n = 26) |
|-----------------------|-----------------|------------------|
| Mast cells and microvessel density | r = 0.54, p < 0.05 | r = 0.64, p < 0.005 |
| Mast cells and PSA level | r = 0.24, p = 0.42 (NS) | r = 0.35, p = 0.079 (NS) |
| Mast cells and Gleason score | – | r = 0.63, p < 0.001 |
| Microvessel density and PSA level | r = 0.14, p = 0.63 (NS) | r = 0.19, p = 0.35 (NS) |
| Microvessel density and Gleason score | – | r = 0.48, p < 0.02 |

NS – not significant
secreted, among others, by mast cells [8, 11]. Vascular endothelial growth factor is a homodimeric glycoprotein, taking part in many normal processes, such as embryonic development, somatic growth and wound healing. Vascular endothelial growth factor stimulates mainly the growth of endothelial cells but also endothelial cell mitogenesis and cell migration [12, 13]. However, VEGF is not the only angiogenesis-stimulating factor. Mast cells secrete chymase and tryptase and these proteins act as degradation factors for extracellular matrix. Moreover, a mast cell also produces metalloproteinases, such as MMP-2 and MMP-9, which demonstrate pro-angiogenic potential by their ability to break down extracellular matrix [10, 14, 15]. Histamine, one of the main and characteristic products of mast cells, increases permeability of capillaries.

Mast cell products not only support the progression of prostate cancer but their role is also observed in other tumours, e.g., colorectal cancer [16, 17], renal cell carcinoma [18] or pulmonary cancer [19]. Moreover, the mediators released by degranulated mast cells promote metastasis of these malignant tumours. The proteolytic enzymes – tryptase and metalloproteinases (MMP-2 and MMP-9) – promote cancer cell invasion into stromal tissues by degrading the extracellular matrix [14, 15]. All of the above-mentioned mechanisms may suggest a tumour promoting role of mast cells. Tumour growth is promoted by multiple mechanisms with angiogenesis being one of the major ones. This process can be evaluated by assessing microvessel density, taking into account the immunoreactivity of endothelial cells with CD34 antibody. In our study, microvessel density in prostate cancer correlated positively with the intensity of mast cell infiltration, as well as with Gleason score. The correlation between mast cell infiltration and microvessel density is widely reported [3–10]. In our study, MVD was observed as 67.72 CD34 positive vessels per 1 mm² in benign lesions and 179.43 vessels in prostate cancer. The mast cell density was 72.82 and 123.73, respectively. Similar observations were made by Nonomura [5], who found a positive correlation between the number of mast cells around cancer foci and Gleason score, also with clinical outcome. Nassin [4] stated that angiogenesis, expressed as a microvessel count, is augmented in prostate cancer, as compared to benign hyperplasia. He also found a positive correlation between MVD and PSA serum levels, indicating a higher risk of cancer recurrence in postoperative patients with high MVD
values. It is noteworthy that, in our study, this correlation was weak and not significant. Similar findings were reported by a number of authors [6, 11, 20]. Ahmadi [21] found differences in microvessel density between low grade (109 vessels/mm²) and high-grade prostate cancer (168 vessels/mm²). Such relationships between the number of mast cells and MVD are observed not only in prostate cancer. Tana et al. [18] found this positive correlation in renal cancer. Moreover, these relationships were found in colorectal cancer, breast cancer and oral squamous cell carcinoma. The opposite opinions are not very strong. Mohseni et al. [22] found no associations between mast cell density, angiogenesis and clinicopathological parameters of renal cell carcinoma. Also, no relationship between MVD and Gleason score was found in prostate cancer by Motta [23].

Johannson et al. presented an interesting observation in 2010 [3]. They found a difference in the functionality of mast cells within the tumour, compared to that in the peritumoural region. The mast cells, located within the tumour, inhibited angiogenesis, while peritumoural mast cells promoted it, stimulating tumour growth. Those findings correlated with patients’ clinical outcomes and treatment responses. Johannson’s observations corresponded to the dual role of mast cells in tumourigenesis. They act as “pro and against” cells, simultaneously promoting and inhibiting tumour development, the effects possibly depending on their location, i.e., either intra- or peritumoural. Similar observations were reported by Dyduch et al. [7]. The anti-tumour function of mastocytes was proven by Aydin et al. [24], Fleischmann et al. [25] and Pitzoni et al. [26]. Also Dyduch et al. in another study [27] observed a smaller count of mast cells in melanomas of the skin than in benign nevi. These authors indicate an ability of mast cells to induce fibrotic processes, which confirms their role in triggering inflammatory processes. Mast cells secrete many bioactive mediators, such as histamine, prostaglandins, cytokines or leukotriene C4, which take part in the immunological host-defence reaction against tumour growth [28]. No evidence was found in our study for mast cell infiltration in prostate cancer. It should be noted, however, that our biopsy specimens were rather small and contained mainly prostate cancer material, while materials from the peritumoural region were scarce and sporadic. This fact did limit the evaluation of the role of mast cells in the peritumoural area.

In conclusion, our investigations may suggest a promoter function of mast cells in prostate cancer formation and development, whereas no evidence was found for their opposite activity. As the concept of dual function of tryptase-positive cells is still a fairly interesting issue, it needs further studies on larger post-operative specimens to get a better understanding of neoplastic processes in the prostate gland, while also searching for improved and more effective therapy protocols for patients suffering from prostate cancer.

The authors declare no conflict of interest.
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