Molecular epidemiology of carbapenemase-producing Enterobacterales in Finland, 2012–2018

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Received: 3 February 2020 / Accepted: 30 March 2020 / Published online: 19 April 2020
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Abstract
Carbapenemase-producing Enterobacterales (CPE) pose an increasing threat to patient safety and healthcare systems globally. We present molecular epidemiology of CPE in Finland during 2012–2018 with detailed characteristics of CPE strains causing clusters during the same time period. All Finnish clinical microbiology laboratories send Enterobacterales isolates with reduced susceptibility to carbapenems or isolates producing carbapenemase to the reference laboratory for further characterization by whole genome sequencing (WGS). In total, 231 CPE strains from 202 patients were identified during 2012–2018. Of the strains, 59% were found by screening and 32% from clinical specimens, the latter were most commonly urine. Travel and/or hospitalization history abroad was reported for 108/171 strains (63%). The most common species were Klebsiella pneumoniae (45%), Escherichia coli (40%), and Citrobacter freundii (6%), and the most common carbapenemase genes blaNDM-like (35%), blaOXA-48-like (33%), and blaKPC-like (31%). During 2012–2018, the annual number of CPE strains increased from 9 to 70 and different sequence types from 7 to 33, and blaOXA-48-like genes became the most prevalent. Of the clusters, 3/8 were linked to traveling or hospitalization abroad and 5/8 were caused by K. pneumoniae clone clonal complex 258. Most of the clusters were caused by K. pneumoniae producing KPC. High variety among different sequence types indicates that majority of CPE cases detected in Finland are likely imported from foreign countries. Nearly one-third of the cases are not found by screening suggesting that there is hidden transmission occurring in the healthcare settings.

Keywords CPE · Enterobacterales · Finland · Whole genome sequencing · Molecular epidemiology

Introduction
Carbapenem-resistant Enterobacterales are one of the most significant increasing health threats globally according to the WHO [1]. In addition to carbapenemases genes, carbapenem-producing Enterobacterales (CPE) have typically collected other resistance genes and are often extensively drug-resistant or even pan-drug resistant, limiting treatment options [2] and leading subsequently to high fatality [3]. Transmission of CPE primarily occurs in hospitals and other healthcare facilities [4]. Transmission of the K. pneumoniae clonal complex (CC) 258 including, sequence types (ST) 258, 11, 340, and 512, has been shown to occur in European, US, and Israeli hospitals and long-term care facilities [5].

The CPE situation varies dramatically in different parts of the world, and also between European countries, from sporadic occurrence to endemic situation [6, 7]. Regions and countries are known to be endemic for a certain carbapenemase, for instance Klebsiella pneumoniae carbapenemase (KPC) in the
USA, Puerto Rico, Colombia, Brazil, Argentina, Greece, and Italy, New Delhi metallo-β-lactamase (NDM) in Indian subcontinent, China, Bangladesh, and Pakistan, and oxacillinase-48 (OXA-48) in Morocco, Turkey, and Malta [7].

CPE has been rare in Finland, as in the other Nordic countries [8]. Until 2013, the occurrence of CPE in Finland was sporadic and related to traveling, and transmission of CPE between two patients was suspected only once [9]. The first outbreak of colonization with KPC-producing K. pneumoniae (KPC-KP) strain ST512 affected nine patients in a primary care hospital in 2013 [10]. A regional outbreak, in which a KPC-KP ST512 strain spread from one hospital to four other healthcare facilities occurred during 2013–2018 and caused several clinical infections [11].

In this report, we present molecular epidemiology of CPE in Finland during 2012–2018 and detailed characterization of CPE strains causing clusters during the same time period.

Materials and methods

Surveillance and bacterial strains

In 2010, Finnish Institute for Health and Welfare (THL) gave guidelines to clinical microbiology laboratories to send CPE or carbapenem-resistant isolates for further characterization. From 2016, on CPE surveillance in Finland is based on communicable diseases act (1227/2016). All clinical microbiology laboratories electronically notify E. cloacae, E. coli, and K. pneumoniae isolates with reduced susceptibility to carbapenems to the National Infectious Disease Registry and send bacterial strains with carbapenemase gene to the Expert Microbiology Unit of the THL. Clinical laboratories also send other CPE species for further characterization. In addition to the patient’s identity information and demographics, the laboratories are requested to report specimen type (screening or clinical) and information on the patient’s preceding travel or hospitalization history abroad if known. According to the national guidelines for controlling multidrug-resistant (MDR) microbes, patients who have been hospitalized abroad in the preceding 12 months are placed on contact precautions upon admission and are screened for MDR bacteria, including CPE [12]. For this work, one isolate per patient per species per year was included, multiple isolates if the patient had isolates with different carbapenemase genes or sequence types (ST). Data on preceding hospitalization history abroad were completed from the local infection control nurses. Since 2015, all CPE isolates have routinely been sequenced using whole genome sequencing (WGS) and for this work, the older isolates (from years 2012 to 2014) were sequenced retrospectively.

Phenotypic and molecular analysis

The species identification was performed in clinical laboratories by matrix-assisted laser desorption/ionization time-of-flight (MALDI-TOF) mass spectrometry (VITEK MS, bioMerieux, Marcy-L’Etoile, France or Bruker Biotyper, Becton, Dickinson and Company, New Jersey, USA), and antimicrobial susceptibilities were assessed by disk diffusion method or by gradient minimum inhibitory concentration (MIC) determination test (Etest, bioMerieux, Marcy-L’Etoile, France) and interpreted according to clinical breakpoints as published by the European Committee on Antimicrobial Susceptibility Testing (EUCAST) versions 2.0-8.1, 2012-2018 [13]. Clinical laboratories use multiplex real-time PCR or other molecular amplification techniques for confirmation of carbapenemase genes in isolates with reduced susceptibility to any carbapenem, as directed in The Finnish national guideline [12]. National guideline screening breakpoints for carbapenems are the same for E. coli, E. cloacae, and K. pneumoniae as published in the EUCAST guideline [14] except for meropenem for which screening breakpoint is > 0.12 mg/L (zone diameter ≤ 22 mm). For other Enterobacterales, the breakpoints are the same as the EUCAST clinical breakpoints, ≥2 mg/L (zone diameter ≤ 22 mm).

Several different commercial or in-house molecular amplification techniques have been used during the study period. However, as based on the national guideline, these methods detect at least blαKPC, blαNDM, blαOXA-48, and Verona integron-encoded metallo-β-lactamase (blαVIM) genes [12].

WGS was performed on a MiSeq instrument (Illumina, San Diego, CA, USA) as previously described [11]. Analysis with Trimmomatic (version 0.33), fastQC (version 0.11.6), SRST2 version 0.2.0, and SeqSphere+ (Ridom GmbH, Münster, Germany) was accomplished as previously described [11]. Core genome (cg) MLST was performed to K. pneumoniae and E. coli using available cgMLST scheme from Ridom and C. freundii and K. oxytoca cgMLST schemes were made in house having 2007 and 2947 targets, respectively. A cut-off of 10 allele differences was used to define clusters in cgMLST analysis. This cut-off has been experimentally determined and used in similar studies previously [11, 15]. WGS data from each of the eight clusters are available from GenBank (Table 1).

Results

In total, 231 CPE strains obtained from 202 patients during 2012–2018 were included in the study: 57% (115/202) were from males and the median age of the patients was 56 years (range, 6 months–98 years). Of the strains, 59% (137/231) were found by screening, 32% (74/231) from clinical
specimens, and for 9% (20/231) the information was not available. Of the clinical specimens, 42 (57%) were from urine, 8 (11%) from wound or abscess, 7 (9%) from blood, and 5 (7%) from respiratory tract. Travel or hospitalization abroad was reported in 91 patients and travel data was not available for 53 patients, 58 patients had no travel or hospitalization abroad. The most common countries were India (n = 29 strains), Greece (n = 11), Thailand (n = 9), Spain (n = 7), and Turkey (n = 7). A total of 52 strains were imported after patient had been hospitalized abroad: 10 in India, 7 in Greece, 5 in Egypt, 4 in Spain, 3 in Turkey, 3 in Russia, 3 more in Europe, 16 outside Europe, and in one case, the country was unknown (Electronic Supplementary Material Table 1).

The annual number of CPE strains increased from 9 in 2012 to 70 in 2018 (Fig. 1), and the number of different STs increased from 7 in 2012 to 33 in 2018 (Fig. 2). K. pneumoniae was the dominant species during the study period, except for years 2016 and 2017 when E. coli was most common. E. cloacae was found consistently during the study period, and C. freundii only during the last 4 years (2015–2018).

The most common CPE species were K. pneumoniae (45%, 105/231), E. coli (40%, 92/231), C. freundii (6%, 14/231), and E. cloacae (4%, 9/231) (Table 1). Most prevalent carbapenemase genes were blaNDM-like (35%, 80/231), blaOXA-48-like (33%, 76/231), and blaKPC-like (31%, 71/231). Species where carbapenemase gene blaNDM-like was detected commonly were E. coli (n = 50) and K. pneumoniae (n = 24), blaOXA-48-like in E. coli (n = 39) and K. pneumoniae (n = 29), and blaKPC-like in K. pneumoniae (n = 55) and C. freundii (n = 9). Of the individual carbapenemase genes, blaKPC-3 was the most prevalent followed by blaOXA-48, blaNDM-5, and blaNDM-1, respectively, and these were found in several different species and STs. No plasmid mediated colistin resistance genes were found.

K. pneumoniae had 23 different STs among 105 strains and E. coli had 37 different STs among 92 strains. Among K. pneumoniae, the most prevalent STs were ST512 (n = 39), ST258 (n = 8), ST11 (n = 7), and ST395 (n = 7) and among E. coli, ST167 (n = 11), ST38 (n = 9), and ST405 (n = 9).

When the patient had travel or hospitalization history in a European region, the most common carbapenemase genes belonged to blaOXA-48-like (18/37), in South-East Asia, Western Pacific, and Africa regions, it belonged to blaNDM-like.

### Table 1: Clusters caused by carbapenemase-producing Enterobacterales in Finland, 2012–2018

| Year(s) | No. of strains | No. of patients | Species     | Gene       | Sequence type | Screening | Clinical Link abroad, country | GenBank accession number of strain belonging to cluster |
|---------|----------------|----------------|-------------|------------|--------------|-----------|-------------------------------|---------------------------------------------------|
| 2013    | 9              | 9              | K. pneumoniae | blaKPC-3   | ST512        | 8         | 1 No                          | SAMN14411195                                      |
| 2013–2018 | 26             | 23             | K. pneumoniae | blaKPC-3   | ST512        | 4         | 10 No                         | SAMN14411196                                      |
| 2015–2016 | 2              | 2              | K. pneumoniae | blaKPC-3   | ST512        | 1         | 0 Hospital transfer, Italy     | SAMN14411197                                      |
| 2016–2018 | 8              | 8              | C. freundii   | blaKPC-3   | ST18         | 2         | 6 No                          | SAMN14411198                                      |
| 2016–2018 | 3              | 3              | K. pneumoniae | blaOXA-48  | ST11/NF*     | 2         | 1 Yes, Colombia                | SAMN14411199                                      |
| 2018    | 2              | 2              | K. pneumoniae | blaOXA-48  | ST395        | 1         | 1 Hospitalization, Russia      | SAMN14411200                                      |
| 2018    | 2              | 2              | K. pneumoniae | blaOXA-48  | ST307        | 2         | 0 No                          | SAMN14411201                                      |
| 2018    | 2              | 2              | K. pneumoniae | blaOXA-48  | ST273        | 1         | 1 No                          | SAMN14411202                                      |

*For one strain completely identical ST was not found, there was one allele difference to ST11

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**Fig. 1** Annual number of carbapenemase-producing Enterobacterales species in Finland, 2012–2018
(34/40, 3/5, and 2/3 respectively), in Eastern Mediterranean region, it belonged to blaoxa-48-like or blandm-like (10/16 and 7/16 respectively), and in Americas, it belonged to blakpc-like (3/4).

Eight CPE clusters were detected during 2012–2018 (Table 1). The defined cut-off (10 allele differences) was used except in one case: one C. freundii isolate with 14 allele differences was defined belonging to the cluster since the patients were hospitalized in the same healthcare facility less than 8 months apart. Five of eight clusters were caused by K. pneumoniae strains belonging to the CC258. K. pneumoniae ST512 with blakpc-3 gene caused three clusters, two large ones with 9 and 23 patients, respectively and one small with two patients. C. freundii ST18 with blakpc-2 gene caused one cluster with 8 patients and K. pneumoniae ST11 with blakpc-3 gene one cluster with three patients. Furthermore, there have been three small clusters caused by K. pneumoniae strain having blaoxa-48 gene with different STs. In two clusters, CPE was found more often in clinical than in screening specimens and in four clusters, the first specimen was obtained on clinical basis. In three clusters, the link abroad was identified.

Discussion

Our study showed that CPE isolates are increasingly found in Finland and have caused several clusters and outbreaks during 2013–2018. The first K. pneumoniae producing KPC-2 strain was detected in Finland already in 2009 [16], and thereafter KPC has become one of the most dominant carbapenemases in Finland. KPC was common in K. pneumoniae and rare in E. coli in our material which is in line with data reported from Europe [8]. K. pneumoniae strains belonging to the clonal complex (CC) 258 are wide spread [5], and it has been shown that carbapenemase-producing K. pneumoniae strains are more eager to spread than other strains [4]. Noteworthy, despite that E. coli was the most prevalent species during 2 years of the study period and the second prevalent during the other years, no carbapenemase-producing E. coli clusters were detected in Finland. MLST results also showed that K. pneumoniae had fewer STs than E. coli, indicating more clonal population structure.

High number of C. freundii detected in our study was related to the cluster with eight cases. Germany has described one KPC-2 producing C. freundii outbreak with 6 cases [17]. These outbreaks indicate that also C. freundii with KPC can cause outbreaks and be a reservoir of CPE-genes.

We focused only on CPEs, we did not analyse carbapenem-resistant Enterobacterales (CRE). It is possible that there were CRE strains with carbapenemase genes that clinical laboratories did not detect by the molecular methods they had in use. However, based on the previous studies in Finland and other European countries, other carbapenemase genes than those recommended for testing in the national guideline are rare [9, 18]. It is also possible that there were Enterobacterales strains with carbapenemase genes which had MICs or inhibition zones for carbapenem antibiotic below screening breakpoints. Especially OXA-48-like enzymes are weak carbapenemases and the strains having blaoxa-48-like genes can have very low carbapenem MICs. However, also these kinds of strains seem to be very rare [8, 19, 20].

Simultaneously, as the number of strains increased annually, the number of different sequence types increased indicating that importation from different foreign countries played a crucial role in changing molecular epidemiology of CPE. Endemicity of certain CPE-genes can be seen in our material when examining travel destinations and hospitalization abroad in relation to genes imported. Also, wide selection of species, sequence types, and different CPE-genes detected support the hypothesis that most cases are imported, although direct links could not be found for all patients. The history of traveling abroad is not systematically collected for all patients admitted to Finnish healthcare facilities and this information was missing for 53/202 patients. We cannot exclude possible horizontal gene transfer between species, even though it seems more improbable than clonal spread [4].

However, a worrisome phenomenon is that CPE was initially often found from clinical specimens and from patients without direct link abroad indicating hidden local
transmissions. This means that there are unknown CPE cases and possible environmental sources challenging CPE control measures. Alarming was also that after detecting a cluster and in spite of infection control measures, onward transmission was not always successfully controlled. One reason for this might be that several Enterobacterales are known to survive for long periods in the hospital environment [21]. Therefore, we are currently updating national MDR control guidelines concerning CPE, terminal cleaning, and screening strategies.

In conclusion, CPE findings in Finland are increasing but still the majority are sporadic. The most common CPE so far was K. pneumoniae with KPC-3 gene which caused most of the CPE cases in outbreaks. Our CPE findings are similar to those reported by other Nordic countries [22, 23].

Acknowledgements The authors wish to acknowledge all Finnish clinical microbiology laboratories for participating, THL Expert Microbiology laboratory for technical assistance, and CSC – IT Center for Science, Finland, for providing computational resources.

Authors’ contributions K. Räisänen: whole genome sequencing, data analysis, preparation of tables and figures, manuscript writing. O. Lyytikäinen, J. Jalava: data analysis, manuscript writing. J. Kaaranen, E. Tarkka, B. Forsblom-Helander, J. O. Grönnroos, R. Vuontela: detection and characterization of bacterial isolates and cluster/outbreak investigation. D. Arifulla, E. Sarvikivi, S. Toura: data collection, cluster investigation.

Funding Information Open access funding provided by National Institute for Health and Welfare (THL).

Data availability The datasets generated during and/or analysed during the current study are available from the corresponding author on reasonable request.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

Ethics approval Collection of CPE strains is based on the communicable disease law. We do not contact patients, ethical permission is not needed.

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