Nimodipine improves cortical efficiency during working memory in healthy subjects

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Abstract
The L-type calcium channel gene, CACNA1C, is a validated risk gene for schizophrenia and the target of calcium channel blockers. Carriers of the risk-associated genotype (rs1006737 A allele) have increased frontal cortical activity during working memory and higher CACNA1C mRNA expression in the prefrontal cortex. The aim of this study was to determine how the brain-penetrant calcium channel blocker, nimodipine, changes brain activity during working memory and other cognitive and emotional processes. We conducted a double-blind randomized cross-over pharmacoMRI study of a single 60 mg dose of oral nimodipine solution and matching placebo in healthy men, prospectively genotyped for rs1006737. With performance unchanged, nimodipine significantly decreased frontal cortical activity by 39.1% and parietal cortical activity by 42.8% during the N-back task (2-back > 0-back contrast; \( P_{FWE} < 0.05; n = 28 \)). Higher peripheral nimodipine concentrations were correlated with a greater decrease in activation in the frontal cortex. Carriers of the risk-associated allele, A \((n = 14)\), had a greater decrease in frontal cortical activation during working memory compared to non-risk allele carriers. No differences in brain activation were found between nimodipine and placebo for other tasks. Future studies should be conducted to test if the decreased cortical brain activity after nimodipine is associated with improved working memory performance in patients with schizophrenia, particularly those who carry the risk-associated genotype. Furthermore, changes in cortical activity during working memory may be a useful biomarker in future trials of L-type calcium channel blockers.

Introduction

Cognitive deficits are a core symptom of schizophrenia, but currently available antipsychotic drugs improve mainly positive symptoms (e.g. hallucinations and delusions), leaving patients with often debilitating cognitive dysfunction and negative symptoms (e.g. apathy, avolition, anhedonia). Efforts to develop cognitive-enhancing agents have been hindered, in part, by a lack of understanding of the neural systems underlying the cognitive deficits in schizophrenia, and therefore a lack of plausible drug targets for this clinical domain.

The CACNA1C gene (alpha 1C subunit of the L-type voltage-gated calcium channel) has been identified by several independent studies as a genome-wide associated risk gene for both schizophrenia and bipolar disorder. We previously reported that healthy adults who carry the CACNA1C SNP rs1006737 A allele, which is associated with risk for schizophrenia and bipolar disorder, had increased activation in the prefrontal cortex during the N-back working memory task, compared to non-risk genotype carriers. Increased activation of the prefrontal cortex during working memory without change in performance on the task (i.e. inefficient cortical engagement) has been previously reported in patients with schizophrenia and their healthy siblings (who are at increased genetic risk for schizophrenia, but without illness). We also found that mRNA expression of
CACNA1C is increased in the dorsolateral prefrontal cortex in human post-mortem brain samples that carry the risk-associated genotype. Yoshimizu and colleagues found that increased CACNA1C mRNA expression in carriers of the risk-associated genotype results in increased L-type voltage-gated calcium channel current density in induced human neurons. These data suggest that blocking the L-type calcium channel may improve cortical function in patients with schizophrenia.

The CACNA1C gene codes for the alpha 1C subunit of Cav1.2 channels and is the binding site for L-type calcium channel antagonists. Recently, Hayes et al. found that there were lower rates of psychiatric hospitalization and self-harm in patients with schizophrenia, bipolar disorder, and nonaffective psychosis who were taking LTCC antagonists. Nimodipine is a dihydropyridine LTCC antagonist used to treat cerebral vasospasm after subarachnoid hemorrhage and has been shown to be effective in treating mood symptoms in case reports and treatment trials of bipolar and unipolar depression. To date, there have been no clinical trials using nimodipine to treat schizophrenia. Nimodipine is highly lipophilic and readily crosses the blood-brain barrier, making it an ideal candidate to probe the effects of L-type calcium channel blockade on cortical activation during working memory.

We conducted the following pharmacoMRI study to evaluate the effects of nimodipine on brain activation during cognitive and emotional tasks in healthy subjects. Based on our previous imaging genetics study, we hypothesized that nimodipine would decrease frontal cortical activation during working memory in the context of fixed performance, and that this improvement in cortical physiological efficiency would be greatest in subjects carrying the CACNA1C risk-associated genotype.

Methods and materials

Subjects

All subjects provided written informed consent approved by the Johns Hopkins Institutional Review Board prior to study participation. Subjects were healthy, non-smoking, Caucasian, right-handed men 18–35 years old. Subjects were screened for current and past psychiatric illness using the Structured Clinical Interview for Diagnosis (SCID) of DSM-IV Disorders. Subjects completed a history and physical examination, biochemical and hematological laboratory screens, MRI safety screen, and screened for drugs of abuse including alcohol. Subjects were nonsmoking, as confirmed with urine nicotine and cotinine screening. Subjects were excluded for past or current neurological or psychiatric illness, uncontrolled medical disorder, hypotension or uncontrolled hypertension, or head trauma with loss of consciousness or evidence of functional impairment after head trauma.

Subjects did not take psychotropic medications within 3 months of the first study visit or any enzyme-inducing or inhibiting agent within 1 month of the study. All subjects were right-handed as determined by the Edinburgh Inventory. A blood sample was collected and processed using standardized methods to extract DNA. Targeted genotyping of rs1006737 was done using a standard allelic discrimination Taqman assay, as described in the supplement.

Study Design

The study was a randomized, double-blind, cross-over of a single 60 mg dose of oral nimodipine (20 mL of Nymalize®) and a matching placebo solution. An investigational pharmacist randomized subjects to receive either nimodipine or placebo for the first visit followed by the opposite treatment on the second visit. Visits were separated by a washout period (minimum 2-weeks). Nymalize and matching placebo were donated by Arbor Pharmaceuticals, LLC. Subjects were screened for drugs of abuse on the morning of each study visit and an intravenous catheter was placed in their left forearm for collection of multiple blood samples. Subjects completed the Profile of Mood States (POMS) at the screening visit and then upon study visit admission and 15, 90, and 240 min after drug administration on each study visit to assess subjects’ mood during the study and compare between treatments.

Nimodipine pharmacokinetics

Blood samples (10 mL) were taken just before and approximately 15, 30, 40, 50, 60, 70, 80, 90, 240 min after drug/placebo administration. Samples between 40 and 90 min were taken in between each of the fMRI tasks. Samples were processed using centrifugation to separate plasma, using standard techniques. Nimodipine was extracted from plasma via protein precipitation, and drug quantification was performed using a validated liquid chromatographic-mass spectrometric (LC-MS) method, with a lower limit of quantification of 0.2 ng/mL. Full method details are described in the supplement.

MRI acquisition

The first 15 subjects were scanned on the Siemens 3T Trio MRI and the last 15 subjects were scanned on the same magnet upgraded to the Siemens 3T Prisma MRI. All scanning parameters remained the same between the Trio and Prisma acquisitions. A structural MRI (T1-weighted 3D MPRAGE) was completed on each subject and reviewed by a neuroradiologist to ensure there were no structural abnormalities. The first functional MRI task was started ~30 min following drug/placebo administration. Subjects were trained or instructed on the fMRI tasks prior to scanning. Subjects completed the N-back
working memory task\textsuperscript{1,7}, a modified monetary incentive delay task\textsuperscript{20}, an emotional memory task\textsuperscript{1}, an emotional face matching task\textsuperscript{21}, and the flanker task\textsuperscript{22}, as previously described. For the functional MRI tasks, we acquired single-shot GRE echoplanar BOLD images using the Siemens 3D PACE (Prospective Acquisition CorRrection) MOCO (MOtion Correction) sequence\textsuperscript{23}. The PACE MOCO sequence, which prospectively corrects for motion during scan acquisition, eliminates subsequent need for realignment, thus minimizing the impact of motion on data quality.

N-back working memory task
The N-back working memory task was scanned second in the series of fMRI tasks, \textasciitilde 45 min to an hour after nimodipine/placebo administration, which corresponds with peak nimodipine levels\textsuperscript{17}. Neural activity was measured using blood oxygenation level dependent (BOLD) fMRI while subjects performed the N-back. Details of the N-back task have been previously described\textsuperscript{1,7}. Briefly, there were 4 blocks of the 2-back (i.e. working memory condition) alternating with a 0-back (i.e. control condition). During the 0-back blocks, the subject serially responds with the current digit presented (1,2,3,4 in a diamond shaped box). During the 2-back block, the subject responds with number presented 2 previous (“\( N \)” = 2). Subjects were trained on the N-back task until they could perform the 2-back condition above 80% accuracy twice in a row. BOLD fMRI parameters for the N-back task were the following: axial slices = 33, thickness = 3.0 mm, voxel size = 3.8 \( \times \) 3.8 \( \times \) 3.0 mm, TR = 2000 ms, TE = 28 msec, FOV = 240 mm, matrix = 64 \( \times \) 64.

Functional imaging analyses
Functional MR images were processed and analyzed using Statistical Parametric Mapping (SPM12) (http://www.fil.ion.ucl.ac.uk/spm) implemented in MATLAB (Mathworks Inc., Natick, MA, USA). (http://www.fil.ion.ucl.ac.uk/spm). For each task, functional scans (\( n = 124 \) for the N-back task) were acquired for each participant to measure the T2*-weighted BOLD signal. Dummy scans (\( n = 2–3 \)) were acquired at the beginning to allow for steady-state magnetization and discarded from subsequent analyses. Prospective motion-corrected images were spatially normalized to the first EPI scan, smoothed with a 10-mm full-width half-maximum (FWHM) Gaussian filter, and ratio normalized to the whole-brain global mean.

In the first-level analyses, for each task, linear contrasts were computed producing t-statistical parameter maps at each voxel. For example, t-statistical parameter maps were produced for the 2-back condition greater than 0-back condition contrast to isolate brain regions related to working memory. These statistical images were entered in a second-level model to identify significant activations within and between treatments and genotype groups.

The main effect of nimodipine on brain activity related to working memory (2-back > 0-back) was analyzed using a paired \( t \)-test (placebo versus nimodipine) and the contrast was thresholded at \( p_{\text{FWE}} < 0.05 \) and 10 contiguous voxels (spatial extent threshold). To determine the effect of rs1006737 genotype on the main effect of nimodipine, the Image Calculator (ImCalc facility) in SPM12 was used to create images of Placebo-Nimodipine for each subject. These images were then entered into a multiple linear regression (an additive genetic model using copies of \( A \) allele), and masked for the main effect of nimodipine on N-back task activation. The interaction of genotype*nimodipine was thresholded at \( p < 0.001 \) uncorrected and extent \( k = 10 \) voxels. First and second-level analyses were conducted for the other fMRI tasks, as previously described\textsuperscript{1,20–22}.

We conducted exploratory analyses to determine if there was a relationship between nimodipine concentrations, diastolic or systolic blood pressure, or heart rate, and the main effect of nimodipine on N-back working memory-related neural activation. A multiple linear regression was done for each covariate (regressed on images of Placebo-Nimodipine using ImCalc), masked for the main effect of nimodipine on N-back task activation, and thresholded at \( p < 0.001 \) uncorrected and extent \( k = 10 \) voxels for each analysis.

Physiological measurements
During fMRI scanning, a pulse oximeter was placed on their left middle finger and measured heart rate through a physiological signals acquisition unit (MP150, BioPac Systems Inc., Santa Barbara, CA) equipped with AcqKnowledge 3.7 software. Additionally, vital signs (heart rate and blood pressure) were measured outside of the MRI at baseline (study visit admission), just before the administration of nimodipine/placebo (pre-dose), 30 min after dosing (pre-MRI), just after the MRI, 1 h and 2 h after the MRI, and before discharge around 4 h after drug administration. Blood pressures and heart rates were compared between placebo and nimodipine treatments using paired t-tests. A Bonferroni corrected p-value of 0.01 was considered statistically different between treatments.

Cognitive performance data
Performance on the N-back task was assessed through a fiber optic response box, which recorded both percent correct (accuracy) and reaction time (RT). Reaction times faster than 100 milliseconds (ms) or slower than 1700 ms (300 ms before stimuli) were removed from the analyses to eliminate anticipatory responses to the stimuli. A paired t-test was used to compare accuracy and reaction
times between conditions (0-back and 2-back) and between treatments (nimodipine and placebo) using GraphPad Prism 8, for Windows, GraphPad Software, La Jolla California USA.

Mood assessment

Subjects completed the POMS at the screening visit, upon admission, and 15, 90, and 240 min after drug administration on each study visit to assess subjects’ mood during the study and compare between treatments, as described in more detail in the supplement.

Results

Thirty subjects completed the study. Subjects were between 19 and 33 years old (mean [SD] age, 25.8 [3.68]). One subject was missing from the N-back fMRI analyses because the response box failed during one visit. One subject was removed from the N-back fMRI analyses because he performed below 70% correct on the 2-back on both study visits. A total of 28 subjects were included in the N-back fMRI analyses, who performed >75% correct for the 2-back condition on both study visits. Of the 28 subjects, 14 were carriers of rs1006737 GG, 4 carried GA, and 10 carried the AA risk-associated genotype.

Effect of nimodipine on neural activation

Nimodipine significantly decreased cortical brain activity in the frontal and parietal cortices during the N-back working memory task (Fig. 1). Table 1 shows the clusters that are significantly decreased after nimodipine compared to placebo (threshold \( p_{FWE} < 0.05 \) and \( k = 10 \)). There were no brain regions that were significantly increased after nimodipine compared with placebo, even at a lower statistical threshold (\( p < 0.001 \) uncorrected, \( k = 10 \)). Because the MRI system was upgraded halfway through the study, we also analyzed the N-back fMRI data for the first half and the second half separately, which resulted in similar effects in the frontal and parietal cortical regions. There was no order effect, as we found no significant differences between visit 1 and visit 2. There were no significant differences in neural activation between nimodipine and placebo treatments during any of the other fMRI tasks, including: the modified monetary incentive delay task, which activates the striatum during encoding and retrieval of aversive scenes \( ^{2} \); an emotional memory task, which activates the prefrontal cortex, amygdala, and hippocampus during encoding and retrieval of aversive scenes \( ^{3} \); an emotional face matching task, which activates the amygdala \( ^{21} \); and the flanker cognitive control task, which activates the anterior cingulate cortex during response inhibition \( ^{22} \).

Nimodipine effect is concentration-dependent in the frontal cortex

Plasma nimodipine concentrations were significantly correlated with a greater decrease in neural activity after nimodipine (i.e. difference between placebo and nimodipine) in the inferior frontal cortex (peak voxel [54,17,−4], \( T = 5.37 \); linear regression with betas from peak: \( p = 1.87e-06, r^2 = 0.56, \) Fig. 2), with higher concentrations resulting in a greater decrease in frontal activation. Other regions where higher concentrations were associated with a greater decrease in neural activity, including the parietal cortex, can be found in Table 2. There were no brain regions where nimodipine concentrations were significantly inversely correlated with a decrease in brain activation after nimodipine compared with placebo (threshold \( p < 0.001 \) uncorrected, 10 voxels). There were also no brain regions where higher nimodipine concentrations were significantly correlated with increased neural activity. Concentrations were not different between CACNA1C genotype groups. Supplementary Fig. 1 shows individual nimodipine concentration-time profiles.

Interaction of CACNA1C genotype with nimodipine effect on neural activation

Carriers of the risk-associated allele, A, had a greater decrease in frontal cortical activation during working memory compared to non-risk allele carriers (linear regression thresholded at \( p < 0.001 \) uncorrected, \( k = 27 \); peak voxel: \( p_{FWE} = 0.046, T = 4.24, Z = 3.66, \) [39,8,23]; Fig. 3). While the effect of the risk-allele only trends towards significance after FWE correction, the interaction of genotype × drug is located in the peak cluster for the main effect of nimodipine on working memory. Subjects who carried the AA genotype had a 62.32% decrease in
BOLD activation after nimodipine compared to 37.42% for GA carriers and 16.05% for GG carriers (Fig. 3). There were no brain regions where G allele carriers had a significantly greater decrease than A allele carriers during working memory (threshold \( p < 0.001 \) uncorrected, 10 voxels).

**Working memory performance**

Subjects had higher accuracy on the 0-back (>99%) compared to the 2-back for both placebo and nimodipine treatments \( (p < 0.0001) \). Performance was not significantly different on the 2-back between placebo \( (91.6\%, \text{SD} = 8.07) \) and nimodipine \( (91.5\%, \text{SD} = 8.48) \) treatments \( (p = 0.90) \). Accuracy was not significantly different between genotype groups for any condition. Accuracy on the 2-back was not correlated with BOLD signal in the peak voxel \([21,2,59]\) of the main effect of nimodipine.

Subjects responded faster during the 2-back compared to the 0-back \((241 \text{ ms}, \text{SD} = 0.102, \text{vs.} 401 \text{ ms}, \text{SD} = 0.060)\) \( (241 [0.102] \text{ ms}, \text{vs.} 401 [0.060] \text{ ms for nimodipine}) \), likely because they were primed for the button press for the 2-back since they knew the number they would be pressing in advance. Reaction time was not significantly different between nimodipine and placebo treatments or between genotype groups for either condition \( (p > 0.05) \). Reaction time on the 2-back was not correlated with BOLD signal in the peak voxel \([21,2,59]\) of the main effect of nimodipine.

**Physiological measurements**

In general, peripheral blood pressure remained stable across the study visits (Supplementary Fig. 2). Diastolic blood pressure was 13.6%, SD = 8.901, lower 30 min after nimodipine administration (pre-MRI) as compared with placebo (paired \( t \)-test: \( t = 6.297, df = 29, p = 7.08e-07 \)), but was not significantly different at other time points. Systolic blood pressure was not significantly different between treatments at any time point. Changes in blood

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**Table 1** Brain regions with significantly decreased neural activity during working memory after nimodipine.

| pFWE peak voxel | \( T \)  | \( Z \)  | Voxel | Coordinates | Brain region               |
|-----------------|---------|---------|-------|-------------|-----------------------------|
| 0.001           | 7.48    | 5.46    | 88    | 21,2,59     | Superior frontal cortex     |
| 0.001           | 7.29    | 5.37    | 24    | −42,−1,29   | Precentral frontal cortex   |
| 0.001           | 7.28    | 5.37    | 31    | −24,−5,54   | Superior parietal cortex    |
| 0.005           | 6.69    | 5.09    | 37    | −48,−3,74   | Inferior parietal cortex    |
| 0.022           | 6.01    | 4.75    | 16    | 30,−5,835   | Superior parietal cortex    |

**Table 2** Higher nimodipine concentrations were associated with a greater decrease in neural activity after nimodipine.

| \( T \) | \( Z \) | Voxel | Coordinates | Brain region               |
|---------|---------|-------|-------------|-----------------------------|
| 5.37    | 4.28    | 24    | 54,17,−4    | Inferior frontal cortex     |
| 4.84    | 3.98    | 37    | −30,−61,65  | Superior parietal cortex    |
| 4.59    | 3.83    | 15    | 27,−73,53   | Superior parietal cortex    |
| 4.54    | 3.80    | 39    | 45,−40,50   | Supramarginal gyrus         |
| 4.42    | 3.72    | 11    | −48,−49,50  | Supramarginal gyrus         |
| 4.24    | 3.61    | 70    | 320,38      | Prefrontal cortex (SMA)     |
| 4.10    | 3.51    | 16    | 15,−61,62   | Superior parietal cortex    |

**Fig. 2** Higher nimodipine concentrations (measured at the time of N-back task) are associated with a greater decrease in frontal cortical activation during working memory (peak voxel \([54,17,−4]\)).
pressure were not different between genotype groups. Neural activity (BOLD signal) in the peak voxel of the main effect of nimodipine was not correlated with diastolic or systolic blood pressure.

Average heart rate increased 15.6%, SD = 10.5, 30 min after nimodipine administration, which corresponds with the decrease in blood pressure (Supplementary Fig. 3; \( p < 0.0001 \)). During the N-back task, heart rate was continuously higher after nimodipine compared with placebo (repeated-measures two-way ANOVA across all blocks \( F(1,22) = 6.458, \ p = 0.019, \ SD = 6.028; \) Supplementary Fig. 3). Heart rate was significantly higher during 2-back as compared to the 0-back during both placebo (paired \( t \)-test \( p = 0.0068 \)) and nimodipine (paired \( t \)-test \( p = 0.0003 \)) treatments. Heart rate was not correlated with BOLD signal in the peak voxel of the main effect of nimodipine. Change in heart rate was similar between genotype groups.

**Mood states**

There were no differences between nimodipine and placebo visits (at any time point) for any of the mood states, domain scores, or for the total mood disturbance score. Subjects reported not being able to tell which visit they received nimodipine.

**Discussion**

In healthy subjects, nimodipine administration significantly decreased neural activity in parietal and frontal cortices during the N-back working memory task, despite fixed performance on the task. This is a pattern of change that suggests that nimodipine improves the physiological efficiency of cortical processing of working memory information. There were no effects of blood pressure or heart rate on our pharmacoMRI results, supporting the notion that the present drug and genotype effects on neural activity (BOLD signal) are indicative of a central nervous system mechanism associated with increased risk for schizophrenia, which is defined operationally as a change in physiological activity without a change in performance. Nimodipine has shown pro-cognitive effects in several animal models of cognitive impairment, including improved spatial working memory performance in both young and aged rats, and improved associative learning in aging rabbits. Nimodipine also improved performance on a delayed-response memory task in aged nonhuman primates. Other animal studies have found that excessive calcium signaling can impair prefrontal cortical functioning, which then requires more extensive activation to compensate, which may explain the pro-cognitive effects of inhibiting calcium channel signaling. Research in nonhuman primates has shown that calcium channel expression in the prefrontal cortex, the specific target of nimodipine, may not be surprising that they would show the greatest effect in this region. However, evidence that CACNA1C risk-associated genotype predicts expression of this channel subunit is inconsistent. Since all genotype groups had a decrease in prefrontal cortical activity after nimodipine, and there was no difference in nimodipine’s effect in the parietal cortex between genotype groups, we would predict that nimodipine could improve performance in all patients who have a working memory deficit, not just those carrying the risk genotype.
actions in the prefrontal cortex can impair the information held in working memory through either neuronal hyperexcitability or calcium-cAMP signaling-dependent opening of potassium channels. In humans, chronic treatment with nimodipine (30 mg TID for 12 weeks) resulted in better cognitive performance in patients with dementia and cognitive impairment in an open-label trial and randomized, placebo-controlled, double-blind trials. Studies are needed to test the effects of nimodipine on working memory performance and other cognitive domains in patients with schizophrenia and related disorders, who have cognitive deficits and therefore do not perform as well on the N-back task. Future studies should test multiple doses and cognitive testing outside of the fMRI to determine the dose-response relationship and determine nimodipine's effects on other cognitive domains. Clinical studies of the cognitive effects of nimodipine should include functional MRI as a biomarker of treatment response, which would also provide insight into the mechanism behind nimodipine's pro-cognitive effects.

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Conflict of interest
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References
1. Bigos, K. L. et al. Genetic variation in CACNA1C affects brain circuitries related to mental illness. Arch. Gen. Psychiatry 67, 939–945 (2010).
2. Green, E. K. et al. The bipolar disorder risk allele at CACNA1C also confers risk of recurrent major depression and of schizophrenia. Mol. Psychiatry. 15, 1016–1022 (2009).
3. International Schizophrenia Consortium. Common polygenic variation contributes to risk of schizophrenia and bipolar disorder. Nature 460, 748–752 (2009).
4. Schizophrenia Working Group of the Psychiatric Genomics Consortium. Biological insights from 108 schizophrenia-associated genetic loci. Nature 511, 421–427 (2014).
5. Sklar, P. et al. Whole-genome association study of bipolar disorder. Mol. Psychiatry. 13, 558–569 (2008).
6. Ferrera, M. A. R. et al. Collaborative genome-wide association analysis supports a role for ANK3 and CACNA1C in bipolar disorder. Nat. Genet. 40, 1056–1058 (2008).
7. Callicott, J. H. et al. Abnormal MRI response of the dorsolateral prefrontal cortex in cognitively intact siblings of patients with schizophrenia. Am. J. Psychiatry 160, 709–719 (2003).
8. Hayes, J. F. et al. Association of hydroxymethyl glutaryl coenzyme a reductase inhibitors, Hype calcium channel antagonists, and bipolar states with rates of psychiatric hospitalization and self-harm in individuals with serious mental illness. JAMA Psychiatry. https://doi.org/10.1001/jamapsychiatry.2018.3907 (2019).
9. De Leon, O. A. Response to nimodipine in ultradian bipolar cycling after amygdalohippocampectomy. J. Clin. Psychopharmacol. 32, 146–148 (2012).
10. Waardenburg, J., Fritze, L., Van Calk, D., Berger, M. & Grunze, H. A calcium antagonist for the treatment of depressive episodes: single case reports. J. Psychiatr. Res. 29, 71–76 (1995).
11. Pazzaglia, P. et al. Nimodipine monotherapy and carbamazepine augmentation in patients with refractory recurrent affective illness. J. Clin. Psychopharmacol. 18, 404–413 (1998).
12. Pazzaglia, P. J., Post, R. M., Ketters, T. A., George, M. S. & Marangell, L. B. Preliminary controlled trial of nimodipine in ultra-rapid cycling affective dysregulation. Psychiatry Res. 49, 257–272 (1993).
13. Brunet, G. et al. Open trial of a calcium antagonist, nimodipine, in acute mania. Clin. Neuropharmacol. 13, 224–228 (1990).
14. Yingling, D. R., Utter, G., Vengalil, S. & Mason, B. Calcium channel blocker, nimodipine, for the treatment of bipolar disorder during pregnancy. Am. J. Obstet. Gynecol. 187, 1711–1712 (2002).
15. van den Kerkhoff, W. & Drewes, L. R. in Diagnosis and Treatment of Severe Dementia (eds Manfred Bergener & Barry Reisberg) 308-321 (Springer Berlin Heidelberg, 1989).
16. Nimboca [package insert], https://www.accessdata.fda.gov/drugsatfda_docs/label/2006s018869k0148blpdf.pdf
17. Nymalize [package insert] (Arbor Pharmaceuticals, LLC), https://www.accessdata.fda.gov/drugsatfda_docs/label/2013/203340lbl.pdf
18. First, M. B., Spitzer, R. L., Gibbon, M. & Williams, J. B. W. Structured Clinical Interview for DSM-IV Axis I Disorders – Patient Edition (SCID-I/P, Version 2.0); (Biometrics Research Department, New York State Psychiatric Institute, 1995).
19. Oldfield, R. C. The assessment and analysis of handedness: the Edinburgh inventory. Neuropsychologia 9, 97–113 (1971).
20. Kohli, A. et al. Using expectancy theory to quantitatively dissociate the neural representation of motivation from its interpersonal influences in the human brain: An fMRI study. Neuromage 178, 552–561 (2018).
21. Harri, A. R. et al. Serotonin transporter genetic variation and the response of the human amygdala [see comment]. Science 297, 400–403 (2002).
22. Sambataro, F. et al. Altered cerebral response during cognitive control: a potential indicator of genetic liability for schizophrenia. Neuropsychopharmacology 38, 846–853 (2013).
23. Lanka, P. & Deshpande, G. Resting state fMRI data from subjects scanned with the EPI-PACE (Echoplanar Imaging – Prospective Acquisition Correction) sequence. Data Brief 20, 2072–2075 (2018).
24. Plichta, M. M. et al. Test–retest reliability of evoked BOLD signals from a cognitive–emotive fMRI test battery. NeuroImage 60, 1746–1758 (2012).
25. Jaffe, A. E. et al. Developmental and genetic regulation of the human cortex transcriptome illuminate schizophrenia pathogenesis. Nat. Neurosci. 21, 1117–1125 (2018).
26. Vasgehi, G., Hajhashemi, V. & Rabbani, M. The effect of nimodipine on memory loss following naloxone-induced morphine withdrawal in object recognition. Res. Pharm. Sci. 9, 445–451 (2014).
27. Crestani, A. P. et al. Memory reconsolidation may be disrupted by a distractor stimulus presented during reactivation. Sci. Rep. 5, 13633 (2015).
28. Hopp, S. C. et al. Calcium dysregulation via L-type voltage-dependent calcium channels and ryanodine receptors underlies memory deficits and synaptic dysfunction during chronic neuroinflammation. J. Neuroinflamm. 12, 1–14 (2015).
29. De Oliveira Alvares, L. et al. Reactivation enables memory updating, precision-keeping and strengthening: exploring the possible biological roles of reconsolidation. Neuroscience 244, 42–48 (2013).
30. Fanelli, R. J., McCarthy, R. T. & Chisholm, J. Neuropharmacology of Nimodipine: From Single Channels to Behavior. Ann. N. Y. Acad. Sci. 747, 336–350 (1994).
31. Levere, T. E. & Walker, A. Old age and cognition: enhancement of recent memory in aged rats by the calcium channel blocker nimodipine. Neurobiol. Aging 13, 63–66 (1992).
32. Levy, A. et al. Nimodipine improves spatial working memory and elevates hippocampal acetylcholine in young rats. Pharmacol. Biochem. Behav. 39, 781–786 (1991).
33. Batuecas, A. et al. Effects of chronic nimodipine on working memory of old rats in relation to defects in synaptosomal calcium homeostasis. Eur. J. Pharmacol. 350, 141–150 (1998).
34. McMonagle-Strucko, K. & Fanelli, R. J. Enhanced acquisition of reversal training in a spatial learning task in rats treated with chronic nimodipine. Pharmacol. Biochem. Behav. 44, 827–835 (1993).
35. Hopp, S. C. et al. Differential rescue of spatial memory deficits in aged rats by L-type voltage-dependent calcium channel and ryanodine receptor antagonism. Neuroscience 280, 10–18 (2014).
36. Sandin, M., Jasmin, S. & Levere, T. E. Aging and cognition: facilitation of recent memory in aged nonhuman primates by nimodipine. Neurobiol. Aging 11, 573–575 (1990).
37. Arnsten, A. F. T. & Wang, M. Targeting prefrontal cortical systems for drug development: potential therapies for cognitive disorders. Annu Rev. Pharm. Toxicol. 56, 339–360 (2016).
38. Datta, D. & Arnsten, A. F. T. Unique molecular regulation of higher-order prefrontal cortical circuits: insights into the neurobiology of schizophrenia. ACS Chem. Neurosci. 9, 2127–2145 (2018).
39. Wang, W., Wang, L. N., Zhang, X. H., Ma, L. & Li, D. J. A nimodipine interventional study of patients with mild cognitive impairment. Zhonghua Nei Ke Za Zhi 45, 274–276 (2006).
40. Ban, T. A. et al. Nimodipine in the treatment of old age dementias. Prog. NeuroPsychopharmacol. Biol. Psychiatry 14, 525–551 (1990).
41. Tollefson, G. D. Short-term effects of the calcium channel blocker nimodipine (Bay-e-9736) in the management of primary degenerative dementia. Biol. Psychiatry. https://doi.org/10.1016/0006-3223(90)90050-C (1990).
42. Pantoni, L. et al. Efficacy and safety of nimodipine in subcortical vascular dementia. Stroke 36, 619 (2005).