Low Serum Concentrations of Moxifloxacin, Prothionamide, and Cycloserine on Sputum Conversion in Multi-Drug Resistant TB

Seung Heon Lee,1 Kyung-Ah Seo,2 Young Min Lee,3 Hyun-Kyung Lee,3 Je Hyeong Kim,1 Chol Shin,1 Jong-Ryul Ghim,2 Jae-Gook Shin,2 and Dong Hyun Kim2

1Division of Pulmonary, Sleep, and Critical Care Medicine, Department of Internal Medicine, Korea University Ansan Hospital, Korea University College of Medicine, Ansan; 2Department of Pharmacology and Pharmacogenomics Research Center, Inje University Busan Paik Hospital, Inje University College of Medicine, Busan; 3Division of Pulmonary and Critical Care Medicine, Department of Internal Medicine, Inje University Busan Paik Hospital, Inje University College of Medicine, Busan, Korea.

Received: June 18, 2014
Revised: August 4, 2014
Accepted: August 28, 2014
Corresponding author: Dr. Dong Hyun Kim, Department of Pharmacology and Pharmacogenomics Research Center, Inje University College of Medicine, 75 Bokji-ro, Busanjin-gu, Busan 614-735, Korea. Tel: 82-51-890-6411, Fax: 82-51-893-1232 E-mail: dhkim@inje.ac.kr

- The authors have no financial conflicts of interest.

Purpose: Low serum concentrations of drugs used to treat multi-drug resistant tuberculosis (MDR-TB) have occasionally been associated with treatment failure. We determined the frequencies of low serum concentrations of anti-MDR-TB drugs, and assessed the effects of these concentrations on 2-month sputum conversion.

Materials and Methods: The serum levels of moxifloxacin (MF), prothionamide (PTH), and cycloserine (CS) were determined for 89 serum samples by high-pressure liquid chromatography-tandem mass spectrometry.

Results: Low serum concentrations of MF, PTH, and CS below the minimal levels of the normal ranges were 83.3% (20/24), 59.2% (29/49), and 71.2% (47/66), respectively. There were no significant differences between the 2-month sputum conversion group (n=25) and the 2-month sputum non-conversion group (n=4) in median drug concentrations (µg/mL) of MF (1.46 vs. 1.60), PTH (0.91 vs. 0.70), and CS (14.90 vs. 14.90). However, a poor compliance rate was significantly greater in the 2-month sputum non-conversion group (75.0%, 3/4) than in the 2-month sputum conversion group (0%, 0/25) (p=0.001). Conclusion: The frequency of low serum concentrations of anti-MDR-TB drugs was substantial and might not affect the 2-month sputum conversion rate. Larger prospective studies with timely sampling are needed to investigate the role of therapeutic drug monitoring in MDR-TB.

Key Words: Tuberculosis, multidrug resistance, moxifloxacin, prothionamide, cycloserine, drug monitoring

INTRODUCTION

Multi-drug resistant tuberculosis (MDR-TB), defined as TB that is resistant to both isoniazid and rifampin, is a serious global public health issue. In a 2010 prevalence survey, 5.4% of culture-positive TB cases included MDR strains.1 In Korea, TB has an intermediate incidence rate of 97/100000 in 2010,1 while the rate of MDR-TB in
new TB treatment is 2.7% and 14.7% in retreatment. The treatment success rate of MDR-TB patients in Korea was reported as 45.3–62.6%,5,14 which is similar to the global reported rate of 60% in 2009.5

Treatment of MDR-TB is difficult as second-line drugs are less potent and not as well tolerated as first-line drugs. Updated guidelines for MDR-TB management recommend that treatment regimens should include the fluoroquinolone, pyrazinamide, a parenteral agent such as kanamycin, ethionamide [or prothionamide (PTH)], and either cycloserine (CS) or 17 p-aminosalicylic acid if CS cannot be used.5 Moxifloxacin (MF) and levofloxacin are the two most commonly prescribed fluoroquinolones and have a pivotal role in the treatment of MDR-TB, although MF has better bactericidal activity than levofloxacin.6 Therefore, MF, PTH, and CS are key drugs for MDR-TB treatment, excluding injectable drugs and the first-line drug pyrazinamide. Moreover, in MDR-TB, the initial 2 months is a critical period and should be accompanied by 6 months of continued adjuvant therapy with an injectable drug after the patient first becomes and remains smear- or culture-negative.7 During the treatment of TB, positive cultures following the completion of 2 months of therapy is a marker of an increased relapse risk in drug-sensitive TB2 and a potential indicator of the development to MDR-TB.8

The relatively low rate of treatment success in MDR-TB may be associated with low serum concentrations of anti-MDR-TB drugs. Such low anti-TB drug concentrations have been associated with malabsorption, alcohol use, age, sex, hypoalbuminemia, weight-adjusted dose, and drug formulation.9-12 While therapeutic drug monitoring (TDM) is commonly used for many drugs, the role of TDM in the treatment of MDR-TB remains uncertain. However, TDM has been recommended by some clinicians as being particularly useful for more complicated patients, including those with more co-morbidities than ordinary patients responding to standard treatment.9 Furthermore, treatment failure or drug toxicities are ongoing concerns in some MDR-TB patients. Thus, dose adjustment using TDM with a prerequisite of appropriate timed sample11 seems to be a better treatment strategy than administering a standard fixed dose, especially in end-stage renal disease and liver disease.7 The need to monitor anti-TB drug levels in order to quickly identify treatment failure has been widely discussed in the literature. Previous studies have shown that low serum concentrations of anti-TB drugs are closely related to treatment failure in human immunodeficiency virus (HIV)-infected14 and non-HIV infected TB patients.15

The aim of the present study was to determine the frequencies of low serum concentrations of anti-MDR-TB drugs and to analyze the effects of these drug concentrations on the 2-month sputum conversion in MDR-TB patients.

MATERIALS AND METHODS

Subjects
The data for this study were retrospectively analyzed at Inje University Busan Paik Hospital. The study was approved by the Institutional Review Board of Inje University Busan Paik Hospital before serum sampling; written informed consent was obtained from all subjects to obtain blood samples.

This study included patients 18–75 years of age who had sputum-culture positive pulmonary TB caused by MDR-TB strains resistant to at least isoniazid and rifampicin based on in vitro drug susceptibility testing (DST) by Löwenstein-Jensen medium, using the absolute concentration method. Patients ultimately enrolled did not have gastroenteropathy on gastrofiberscopy and colonfiberscopy, neuropathy on nerve conduction tests, or renal insufficiency on serum urea nitrogen (BUN), creatinine, albumin, and electrolyte levels. DST results for the drugs prescribed for enrolled MDR-TB patients showed susceptibility to MF, PTH, and CS, except for one patient with extensively drug resistant (XDR)-TB who had a TB strain that was resistant to MF. Serum samples from enrolled MDR-TB patients that were not drawn within 2–6 h after drug ingestion were excluded from drug level measurement. During the initial visit, chest X-rays and computed tomography scans were obtained and interpreted separately by two board-certified experts. The radiologic severity of MDR-TB was estimated based on the recommendations of the National Tuberculosis Association of the United States.16

Serum sample preparation and modified directly-observed treatment
Venous blood was drawn 2–6 h after the ingestion of drugs from 29 MDR-TB patients during their monthly visit. Serum was separated by centrifugation at 1000×g for 10 min and stored at -80°C until analysis. Serum levels of the anti-MDR-TB drugs MF, PTH, and CS were obtained and measured for MF prescribed at 400 mg/day (n=29), PTH prescribed at 250 mg (n=15) or 500 mg (n=34) twice/day, and CS prescribed at 250 mg (n=49) or 500 mg (n=17) twice/
day. The number of samples per drug, which were collected between 2 and 3 hours post-dose, were 11 for MF, 18 for PTH, and 21 for CS.

TB nurses regularly monitored drug compliance and drug adverse events for the participating patients in this study. Monitored adverse drug reactions were categorized by neurologic abnormalities, gastrointestinal trouble, dermatologic abnormalities, and hepatotoxicity. Poor compliance was defined as treatment interruption for more than 20% of the prescribed doses.

**LC-MS/MS analysis**

Serum concentrations of MF, PTH, and CS were analyzed using a validated high-performance liquid chromatography-tandem mass spectrometry method (LC-MS/MS). Briefly, 180 μL of acetonitrile containing 1 μg/mL of MF-d₄ as an internal standard was added to a 20-μL serum sample aliquot. The supernatant was taken after centrifugation at 3000×g for 10 min, and a 1-μL aliquot was then injected into a PE SCIEX API 4000 LC-MS/MS system (Applied Biosystems, Foster City, CA, USA), equipped with an Agilent 1200 series high-performance liquid chromatography (HPLC) system (Agilent, Wilmington, DE, USA). The analytical column was an Atlantis dC18 column (150×2.0 mm, 3 μm; Waters, Milford, MA, USA) with a mobile phase consisting of 0.1% formic acid solution and acetonitrile. A gradient program was used for HPLC separation with a flow rate of 0.2 mL/min. The initial composition of acetonitrile was 30%, maintained for 2 min and then increased to 90% for 5.1 min, followed by re-equilibration to the initial condition for 8 min. The mass spectrometer with an electrospray source was run in the positive mode and m/z 402.0→384.0, 181.0→154.3, 103.0→58.0, and 406→388 monitored for MF, PTH, CS, and MF-d₄, respectively. The peak areas for all analytes were integrated automatically using Analyst software v. 1.4 (Applied Biosystems). The validation of the assay was performed according to the US Food and Drug Administration guidance for the validation of bioanalytical methods. Calibration curves in the range of 0.2–10.0 μg/mL for MF and PTH and 0.8–40.0 μg/mL for CS were established (r²=0.9996 for MF; r²=0.9980 for PTH; r²=0.9989 for CS). Three quality control samples (low, medium, and high) were used for the validation. The intra-day (n=5) and inter-day (n=5) precisions were less than 11.4%, and the accuracies were between 93.1% and 107.4% for the three anti-TB drugs. In addition, no matrix effects were observed near their elution times.

**References in serum levels of anti-MDR-TB drugs**

Serum levels of MF, PTH, and CS were measured 2–6 h after dosing, as described previously. Normal ranges were defined as 2.5–4.5 μg/mL for MF, 1–5 μg/mL for PTH, and 20–35 μg/mL for CS, based on published reference ranges.

Levels below normal ranges for each drug were defined as MF<2.5 μg/mL (400 mg daily oral dose), PTH<1 μg/mL (500 mg or 1000 mg daily oral dose), and CS<20 μg/mL (500 mg or 1000 mg daily oral dose).

**Sputum culture conversion**

We defined the critical time for effective MDR treatment as 2 months, based on previous clinical trials on MDR-TB treatment using a culture conversion rate at 2 months. Sputum AFB culture conversion was defined as two or more consecutive negative sputum cultures tested at least 4 weeks apart.

**Statistical analysis**

The median serum levels of each drug in the 2-month sputum conversion group and the 2-month sputum non-conversion group were compared using a two-tailed Mann-Whitney test. Fisher’s exact test was used to evaluate differences in the frequencies of low serum concentrations below the normal ranges for each anti-MDR-TB drug. Differences with a value of \( p < 0.05 \) were considered to be statistically significant. SPSS ver. 12.0 software (SPSS Inc., Chicago, IL, USA) software was used for all statistical analyses.

**RESULTS**

**Participant demographics**

Table 1 presents the baseline characteristics of the 29 enrolled MDR-TB patients. The median age was 47 years (range, 19–67 years), and 17 patients (58.6%) were male. A Bacillus Calmette-Guérin scar was present in 20 patients (69.0%). The median body mass index was 20.5 kg/m², and 5 patients (17.2%) had a familial TB history. Three patients had diabetes mellitus, although none had HIV disease. In chest radiographs, 16 patients had cavitary disease, and 16 had bilateral lung disease. Sputum non-conversion after 2 months of treatment occurred in 4 patients, and sputum conversion after 2 months of treatment occurred in 25 of 29 of the MDR patients. However, there were no significant differences in clinical characteristics except drug compliance between the 2-month conversion and 2-month non-conver-
sion groups (Table 2). Drug toxicity developed in some patients; this included gastrointestinal trouble (10 of 29), dermatologic abnormalities (3 of 29), and mild hepatotoxicity (3 of 29). A total of 89 serum samples were obtained. MF was prescribed at 400 mg/day, PTH at 250 mg (body weight<50 kg) or 500 mg (body weight≥50 kg) twice/day, and CS at 250 mg (body weight<50 kg) or 500 mg (body weight≥50 kg) twice/day.

**Serum moxifloxacin concentration**
The frequency of low serum MF concentrations below the minimal normal range was 83.3% (20/24). In the conversion group, the frequency of low serum MF concentrations was 85.0% (17/20), compared with 75.0% (3/4) in the non-conversion group, which was not a statistically significant difference (Fig. 1). The median serum concentration in the 2-month sputum conversion group was 1.46 μg/mL [interquartile range (IQR)=0.33–2.17; n=25], and in the 2-month sputum non-conversion group, the concentration was 1.60 μg/mL (IQR=0.93–2.54; n=4); this difference was also not statistically significant (p=0.394) (Table 2). The median concentration from the samples collected between 2 and 3 hours post-dose was 0.73 μg/mL (IQR=0.24–1.87; n=25) (Table 3).

**Serum prothionamide concentration**
The frequency of serum PTH concentrations below the minimal normal range was 59.2% (29/49). In the conversion group, the frequency of serum PTH concentrations below the minimal normal range was 57.8% (26/45) compared with 75.0% (3/4) in the non-conversion group; this difference was not statistically significant (Fig. 1). The median serum concentration in the 2-month sputum conversion group was 0.93 μg/mL (IQR=0.35–2.34; n=25), and in the 2-month sputum non-conversion group, the concentration was 0.70 μg/mL (IQR=0.24–1.28; n=4); this difference was also not statistically significant (p=0.597) (Table 2). The median concentration from the samples collected between 2 and 3 hours post-dose was 0.73 μg/mL (IQR=0.24–1.87; n=25) (Table 3).

**Table 1. Clinical Characteristics of MDR-TB Patients**

| Characteristics (n=29) | Number (range or %) |
|-----------------------|---------------------|
| Age, yrs, median (range) | 47 (19–67) |
| Male:female (%) | 17 (58.6):12 (41.4) |
| BMI, kg/m², median (IQR) | 20.5 (14.5–25.5) |
| Smoking history | 9 (31.0) |
| BCG scar | 20 (69.0) |
| Previous TB history (%) | 16 (55.1) |
| Familial TB history (%) | 5 (17.2) |
| Underlying disease (%) |  |
| DM | 3 (10.3) |
| Hypertension | 4 (13.8) |
| HIV | 0 (0) |
| Radiologic severity (%) |  |
| Cavity disease | 16 (55.2) |
| Bilateral disease | 16 (55.2) |
| Sputum non-conversion at 2 months | 4 (13.8) |
| Mean duration of follow-up (month) | 19.0±9.3 |

MDR-TB, multi-drug resistant tuberculosis; BMI, body mass index; IQR, interquartile range; BCG, Bacillus Calmette-Guérin; DM, diabetes mellitus; HIV, human immunodeficiency virus.

**Table 2. Differences in Sputum Conversion of MDR-TB Patients**

| Characteristics | Two-month conversion (n=25) | Two-month non-conversion (n=4) | p value |
|-----------------|-----------------------------|-------------------------------|--------|
| Serum drug level (median μg/mL, IQR) |  |
| MF | 1.46, 0.33–2.17 | 1.60, 0.93–2.54 | 0.394 |
| CS | 15.10, 8.23–21.65 | 14.90, 9.89–19.45 | 0.981 |
| PTH | 0.93, 0.35–2.34 | 0.70, 0.24–1.28 | 0.597 |
| No. of used TB drugs per patient during 2 months (median, IQR) | 5.0, 5.0–5.0 | 5.5, 5.0–6.75 | 0.216 |
| No. of resistant TB drugs on DST per patient (median, IQR) | 3.0 (2.5–4.5) | 6.0 (3.3–8.8) | 0.213 |
| Previous TB history (%) | 15/25 (60) | 1/4 (25) | 0.299 |
| Cavity disease (%) | 13/25 (52.0) | 3/4 (75) | 0.606 |
| Bilateral disease (%) | 13/25 (52.0) | 3/4 (75) | 0.606 |
| Poor compliance (%) | 0 (0) | 3/4 (75) | 0.001 |
| XDR (%) | 0 (0) | 1/4 (25) | 0.138 |
| Incidence of side-effects (%) | 2/25 (8) | 2/4 (50) | 0.080 |
| BMI, kg/m², median (IQR) | 20.1, 18.8–22.2 | 22.7, 20.8–25.1 | 0.061 |
| Treatment failure on last follow up | 0 (0) | 0 (0) | 1.000 |

MDR-TB, multi-drug resistant tuberculosis; IQR, interquartile range; MF, moxifloxacin; CS, cycloserine; PTH, prothionamide; DST, drug susceptibility test; XDR, extensively drug-resistant; BMI, body mass index.
concentration of PTH in patients taking 250 mg twice/day (median 0.13 g/mL; IQR 0.06–0.57) was significantly lower than in those taking 500 mg twice/day (median 1.27 g/mL; IQR 0.63–2.61; \(p\)=0.0006) (Fig. 1). The median concentrations for all samples according to doses are presented in Table 3.

**Serum cycloserine concentration**

The frequency of serum concentrations of CS below the minimal normal range was 71.2% (47/66). In the conversion group, the frequency of serum CS concentrations below the minimal normal range was 70.5% (43/61) compared with 80.0% (4/5) in the non-conversion group; this difference was not statistically significant (Fig. 1). The median serum concentration in the 2-month sputum conversion group was 15.10 μg/mL (IQR 8.23–21.65; n=25), and in the 2-month sputum non-conversion group, the concentration was 14.90 (IQR 9.89–19.45; n=4); this difference was also not statistically significant (\(p\)=0.981) (Table 2). The median concentration of CS in patients taking 250 mg twice/day (median 13.10 g/mL; IQR 6.80–18.25) was significantly lower than in those taking 500 mg twice/day (median 20.90 g/mL; IQR 17.40–29.80; \(p\)<0.0001) (Fig. 1). The median concentrations for all samples according to doses are presented in Table 3.

**Differences in sputum conversion after 2 months of treatment**

The number of TB drugs used during the initial 2 months of therapy did not differ between the sputum conversion (5.0±0.6) and non-conversion (5.8±1.0) groups. Furthermore, drug toxicity rates were not significantly different between the two groups [8% (2/25) vs. 50% (2/4); \(p\)=0.080]. The numbers of patients with a previous TB history [60% (15/25) vs. 25% (1/4); \(p\)=0.299] and the numbers of resistant TB drugs on DST per patient were also not significantly different between the two groups, respectively (3.0 drugs, IQR 2.5–4.5 vs. 6.0 drugs, IQR 3.3–8.8). However, the poor compliance rate was significantly greater in the 2-month sputum non-conversion group (75.0%, 3/4) than in the 2-month sputum conversion group (0%, 0/25; \(p\)=0.001) (Table 2).

---

**Table 3. Serum Drug Level of Each Drug According to Sampling Time and Dose**

| Drug   | Concentration (median µg/mL, IQR) | Normal range of concentration (µg/mL) |
|--------|-----------------------------------|--------------------------------------|
| MF 400 mg (2–3 hr), n=11 | 0.73, 0.24–1.87 | 0.24–1.87 |
| MF 400 mg (3–4 hr), n=5 | 0.60, 0.19–3.01 | 0.19–3.01 |
| MF 400 mg (4–6 hr), n=8 | 1.83, 0.61–2.41 | 0.61–2.41 |
| PTH 250 mg, n=15 | 0.19, 0.06–0.61 | 0.06–0.61 |
| PTH 500 mg, n=34 | 1.27, 0.67–2.53 | 0.67–2.53 |
| CS 250 mg, n=49 | 13.40, 6.74–18.30 | 6.74–18.30 |
| CS 500 mg, n=17 | 20.20, 16.60–29.60 | 16.60–29.60 |

MF, moxifloxacin; CS, cycloserine; PTH, prothionamide; IQR, interquartile range.

---

**Fig. 1. Serum concentrations of (A) moxifloxacin, (B) prothionamide, and (C) cycloserine in 29 MDR-TB patients. Hollow circles in (B and C) represent the assorted values marked in the conversion groups. The frequency of low serum concentrations of MF, PTH, and CS below the minimal normal range are described in the bottom line under each group. There were no differences between the 2-month sputum conversion group (n=25) and the 2-month sputum non-conversion group (n=4) in median drug concentration of MF, PTH, and CS. The median serum concentration in the 2-month sputum conversion group (n=25) and 2-month sputum non-conversion group (n=4) were as follows: (A) 1.46 µg/mL, IQR 0.33–2.17 vs. 1.60 µg/mL, IQR 0.93–2.54; (B) 0.90 µg/mL, IQR 0.35–2.34 vs. 0.70 µg/mL, IQR 0.24–1.26; and (C) 15.10 µg/mL, IQR 8.23–21.65 vs. 14.90 µg/mL, IQR 9.99–19.45. IQR, interquartile range; MDR-TB, multi-drug resistant tuberculosis; MF, moxifloxacin; PTH, prothionamide; CS, cycloserine.**
During the initial 2 months of treatment, a substantial number of MDR-TB patients did not achieve the minimal therapeutic levels of MF, PTH, and CS. However, these low drug concentrations did not affect sputum conversion after 2 months of treatment. Although the cost-effectiveness and clinical importance of TDM for anti-TB drugs has been uncertain, we nevertheless measured serum levels of secondary TB drugs, including MF, a promising new MDR-TB drug.

TDM intervention for conditions other than TB reduces adverse events, mortality rates, and length of stay. During the course of complicated MDR-TB treatment with second-line drugs, TDM allows the clinician to make an informed decision about poor drug absorption and to adjust the treatment accordingly in a timely manner. Moreover, TDM shortens the time of sputum conversion through adjusting drug dose and decreasing the adverse effects of high levels of anti-TB drugs. In diabetics, early TDM for TB patients can have a favorable time to sputum culture conversion. Considering that the liquid broth media culture system (MGIT tube; Beacon-Dickinson and Co., Sparks, MD, USA) can produce timely results in 2 weeks, TDM can affect changes in treatment regimens sufficiently.

Reports describing the serum levels of anti-TB drugs in MDR-TB patients are rare. However, the frequencies of low serum concentrations of the secondary anti-TB drugs PTH and CS have been reported to be 52% for PTH and 44% for CS. In our study, the frequencies of low serum concentrations of MF, PTH, and CS were 83.3%, 59.2%, and 71.2%, respectively. Interindividual variations in anti-TB drug concentrations can be explained by race, dosage, drug formulations, performance of directly observed treatment, and measurement methods, as well as inaccurate sampling times. In this study, drug dosage was considered a major factor governing the serum PTH and CS levels.

The compliance rate may be another critical factor affecting sputum conversion during the early period of treatment. Concomitant prescription of at least five anti-TB drugs and a relatively high incidence of secondary anti-TB drug-associated side effects can cause low compliance. In addition, drug concentration and pharmacokinetic variability of anti-TB drugs as well as drug compliance can influence treatment failure and drug resistance acquisition in TB patients, as TB has microbial, clinical, and programmatic causes. In our study, the incidence of side effects in the 2-month sputum non-conversion group (50%) was much higher than that in the 2-month sputum conversion group (8.0%). Our data suggest that anti-TB drug-related side effects may result in poor compliance, leading to sputum non-conversion. Therefore, TDM would be a useful tool for making sound clinical judgments, although case-holding strategies such as direct observation of treatment with anti-TB drugs seems essential for confident compliance and treatment success in terms of sputum conversion.

CS causes frequent central nervous system side effects, such as an inability to concentrate or lethargy; these appear even with serum concentrations at the low end of the normal range and may be aggravated at more elevated serum concentrations. In our study, depressive mood and agitation manifested in two patients receiving CS, and these side effects disappeared when CS was discontinued. CS concentrations at the time of adverse drug events were lower than the maximal normal range in both patients. Although gastrointestinal trouble (10/29), dermatologic abnormalities (3/29), and mild hepatotoxicity (3/29) were developed, we could not be certain that these adverse events were solely attributable to MF, PTH, or CS rather than background TB drugs.

This study had several limitations. First, the number of MDR-TB patients was not sufficient to make any definitive conclusions about differences in the 2-month serum conversion rate. Additionally, MDR-TB treatment efficacy was not fully evaluated, as the study considered only microbiological traits, human host characteristics, and environmental factors, as well as serum drug concentration. Second, the serum-sampling interval was relatively wide; thus, we could not assure the peak level of each anti-TB drug. Third, this was a retrospective study; therefore, we were unable to intervene in order to escalate drug doses or to measure serum drug levels. Fourth, although culture conversion after 2 months of treatment is an indicator of treatment effectiveness in drug-sensitive TB, there are no accumulated data for MDR-TB treatment that provide evidence for 2-month culture conversion as an indicator of treatment effectiveness in spite of several previously reported clinical studies. However, these results warrant a prospective detailed pharmacokinetic and pharmacodynamic study of secondary anti-TB drugs at multiple time points in relation to clinical outcomes, as well as an investigation of adverse drug events related to serum drug levels.

In conclusion, the incidences of low serum concentrations among secondary anti-TB drugs were 83.3% in MF, 59.2%...
in PTH, and 71.2% in CS. Although the clinical meaning of these results remains uncertain, they warrant a larger prospective study that includes scheduled multiple time samplings to determine the role of TDM in MDR-TB patients.

ACKNOWLEDGEMENTS

This work was supported in part by a grant funded by the Korea Centers for Disease and Prevention (R12-2007-023-00000-0) and in part by a National Research Foundation of Korea grant funded by the Korea government (R13-2007-023-00000-0).

REFERENCES

1. World Health Organization. Global tuberculosis control: WHO report 2011. Geneva: World Health Organization; 2011.
2. Bai GH, Park YK, Choi YW, Bai Ji, Kim HJ, Chang CL, et al. Trend of anti-tuberculosis drug resistance in Korea, 1994-2004. Int J Tuberc Lung Dis 2007;11:571-6.
3. Kim DH, Kim HJ, Park SK, Kong SJ, Kim YS, Kim TH, et al. Treatment outcomes and survival based on drug resistance patterns in multidrug-resistant tuberculosis. Am J Respir Crit Care Med 2010;182:113-9.
4. Kim HR, Hwang SS, Kim HJ, Lee SM, Yoo CG, Kim YW, et al. Impact of extensive drug resistance on treatment outcomes in non-HIV-infected patients with multidrug-resistant tuberculosis. Clin Infect Dis 2007;45:1290-5.
5. World Health Organization. Guidelines for the programmatic management of drug-resistant tuberculosis-2011 update. Geneva: World Health Organization; 2011.
6. Cremades R, Rodríguez JC, García-Pachón E, Galiana A, Ruiz-García M, López P, et al. Comparison of the bactericidal activity of various fluoroquinolones against Mycobacterium tuberculosis in an in vitro experimental model. J Antimicrob Chemother 2011;66:2281-3.
7. Blumberg HM, Burman WJ, Chaisson RE, Daley CL, Etkind SC, Friedman LN, et al. American Thoracic Society/Centers for Disease Control and Prevention/Infectious Diseases Society of America: treatment of tuberculosis. Am J Respir Crit Care Med 2003;167:603-62.
8. Glor MT, Skripconoka V, Sanchez-Garavito E, Xiao H, Cabrera-Rivero JL, Vargas-Vasquez DE, et al. Delamanid for multidrug-resistant pulmonary tuberculosis. N Engl J Med 2012;366:2151-60.
9. Peloquin CA. Therapeutic drug monitoring in the treatment of tuberculosis. Drugs 2002;62:2169-83.
10. McCuller H, Wash P, Burger A, Norman J, Folb PI, Smith P. Determinants of rifampin, isoniazid, pyrazinamide, and ethambutol pharmacokinetics in a cohort of tuberculosis patients. Antimicrob Agents Chemother 2006;50:1170-7.
11. Tappero JW, Bradford WZ, Agerton TB, Hopewell P, Reingold AL, Lockman S, et al. Serum concentrations of antimycobacterial drugs in patients with pulmonary tuberculosis in Botswana. Clin Infect Dis 2005;41:461-9.
12. van Crevel R, Alisjahbana B, de Lange WC, Borst F, Danusantoso H, van der Meer JW, et al. Low plasma concentrations of rifampicin in tuberculosis patients in Indonesia. Int J Tuberc Lung Dis 2002;6:497-502.
13. Li J, Buzynska JN, Lee YA, Berg D, Driver CR, Ridzon R, et al. Use of therapeutic drug monitoring for multidrug-resistant tuberculosis patients. Chest 2004;126:1770-6.
14. Weiner M, Benator D, Burman W, Peloquin CA, Khan A, Vernon A, et al. Association between acquired rifamycin resistance and the pharmacokinetics of rifabutin and isoniazid among patients with HIV and tuberculosis. Clin Infect Dis 2005;40:1481-91.
15. Kimerling ME, Phillips P, Patterson P, Hall M, Robinson CA, Dunlap NE. Low serum antimycobacterial drug levels in non-HIV-infected tuberculosis patients. Chest 1998;113:1178-83.
16. U.S. National Tuberculosis Association. Diagnostic standards and classification of tuberculosis. New York: National Tuberculosis Association; 1961.
17. Alffenaar JW, van Altena R, Bökererink HJ, Luijckx GJ, van Soolingen D, Aarnoutse RE, et al. Pharmacokinetics of moxifloxacin in cerebrospinal fluid and plasma in patients with tuberculous meningitis. Clin Infect Dis 2009;49:1080-2.
18. Schumacher GE, Barr JT. Economic and outcome issues for therapeutic drug monitoring in medicine. Ther Drug Monit 1998;20:539-42.
19. Heyssel SK, Moore JL, Staley D, Dodge D, Houpt ER. Early Therapeutic Drug Monitoring for Isoniazid and Rifampin among Diabetics with Newly Diagnosed Tuberculosis in Virginia, USA. Tuberculosis Res Treat 2013;2013:129723.
20. Pasipanodya JG, Srivastava S, Gumbo T. Meta-analysis of clinical determinants of rifampin, isoniazid, pyrazinamide, and ethambutol pharmacokinetics in medicine. Ther Drug Monit 1998;20:539-42.