Matrix Metalloproteinases in Cerebral Ischemia

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Matrix metalloproteinases (MMPs) are involved in the pathophysiology of several central nervous system diseases that share common pathogeneses, such as disruption of the blood-brain barrier (BBB), neuroinflammation, oxidative stress, and remodeling of the extracellular matrix (ECM). In early ischemic injury, MMPs participate in disruption of the BBB by digesting the basal lamina of capillaries and ECM, leading to vasogenic edema and hemorrhagic transformation. However, ECM degradation and remodeling are essential for tissue recovery, with MMPs having a key role as modulators of homeostasis between neuronal death and tissue regeneration. Thus, MMPs may be a double-edged sword that has a deleterious or beneficial role depending on the stage of brain injury.

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INTRODUCTION

Extracellular proteases comprise serine proteases, tissue-type and urokinase-type plasminogen activator (PA), and the families of matrix metalloproteinases (MMPs). MMPs are families of enzymes that require a zinc ion for catalytic activity. The role of MMPs has classically been described as strict regulators of extracellular matrix (ECM) remodeling in tissue morphogenesis and wound healing. MMPs regulate cellular activity in various ways, including ECM degradation, cell adhesion, proteolytic release of ECM-sequestrated molecules, and shedding of cell surface proteins that transduce signals from the extracellular environment (Table 1).1,2 Recently, a deleterious role of MMPs in several diseases of the nervous system has been widely studied. However, some beneficial functions of MMPs during the recovery phase after brain injury have also been suggested. This review focuses on the role of MMPs in the acute and recovery phases of cerebral ischemia.

STRUCTURE AND EXPRESSION OF MMPs

According to the basic protein domain and substrate preference, there are five major classes of MMPs: matrilysins, membrane-type MMPs (MT-MMPs), stromelysins, gelatinases, and other recently discovered MMP families.3 The protein structures of MMPs share a basic common pattern (Fig. 1) of propeptide and catalytic domains. However, they differ in more specific domains. The matrilysin MMP-7 has the simplest domain structure, with the propeptide domain and zinc containing catalytic domain, but it degrades most ECM components. The MT-MMPs (MMP-14, -15, and -16) have an
Table 1. Effects of MMPs in the central nervous system

| Beneficial effects          | Deleterious effects               |
|----------------------------|----------------------------------|
| Cell-fate specification    | Tumorigenesis                     |
| Angiogenesis               | Disruption of blood-brain barrier|
| Myelinogenesis             | Inflammation                      |
| Survival of cells          | Demyelination                     |
| Axonal growth              | Neuronal death                    |
| Signaling                  |                                   |
| Repair processes           |                                   |
| Migration of neuronal precursors |                        |
| Termination of inflammation|                                   |

Additional transmembrane domain, whereas the stromelysins (MMP-1, -3, and -10) have a hemopexin domain. The gelatinases (MMP-2 and -9) have a unique fibronectin-binding domain, allowing them to bind to basement membranes. Thus, gelatinases specifically attack type IV collagen, laminin, and fibronectin. The common propeptide domain maintains the latency state of the enzyme, with removal of the signal peptide of this domain activating enzymes. Activation can also occur through a change in configuration. Proteolytic removal or reconfiguration of the propeptide domain activates the enzymes. Exposure of the catalytic site is referred to as the “cysteine switch.” The various MMPs exist as secreted or membrane-bound enzymes that require conversion from zymogen to active forms through proteolytic processing, thus ensuring that MMP expression is tightly regulated at the transcriptional and posttranslational levels. Additionally, MMPs are also regulated extra-cellularly by proteins known as tissue inhibitors of metalloproteinases (TIMPs). The strict control of the activity of MMPs at multiple levels, such as transcription, posttranslational modification, and interaction with TIMPs is important, because MMPs play sophisticated roles and are extremely deleterious if they are erroneously activated.

Recent studies have revealed the involvement of MMPs with clinical vascular diseases (Table 2). Ischemic stroke is often associated with disruption of the blood-brain barrier (BBB), leading to vasogenic edema and hemorrhage. Numerous animal studies suggest that disruption of the BBB and hemorrhagic transformation in ischemic stroke results from MMP activation in pathologic conditions. A deleterious role of MMPs associated with BBB injury has been demonstrated by a reduced infarct size and less BBB damage after focal ischemia in MMP-9 knockout mice, and by both MMP activation and BBB injury being reduced in mice with superoxide dismutase overexpression. In nonhuman primates, early opening of the BBB after ischemic stroke correlates with elevated MMP-2, whereas hemorrhagic transformation is associated with elevated MMP-9.

INCREASED MMPs IN EXPERIMENTAL CEREBRAL ISCHEMIA

![MMP families](image)
| Disease                        | Target MMP          | Specimen            | Reference           |
|-------------------------------|---------------------|---------------------|---------------------|
| Carotid atherosclerosis       | MMP-1,2,3,9, TIMP-1 | Atherosclerotic plaque | Galis et al.        |
|                               | MMP-1               | Atherosclerotic plaque | Nikkari et al.     |
|                               | MMP-2,9, TIMP-1     | Atherosclerotic plaque, serum | Tziakas et al.    |
|                               | MMP-2,8,9           | Plasma               | Turu et al.         |
|                               | MMP-2, TIMP-1,2     | Serum                | Pawlak et al.       |
|                               | MMP-1,10            | Atherosclerotic plaque | Montero et al.      |
|                               | MMP-9               | Atherosclerotic plaque | Sigala et al.       |
| Ischemic heart disease        | MMP-9               | Serum                | Renko et al.        |
|                               | MMP-2,3,9           | Plasma               | Wu et al.           |
|                               | MMP-2,9             | Plasma               | Zeng et al.         |
|                               | MMP-3               | Serum                | Samnegard et al.    |
| Cerebral ischemia             | MMP-2,9             | Plasma               | Montaner et al.     |
|                               | MMP-9               | Plasma               | Castellanos et al.  |
|                               | MMP-2,9             | Plasma               | Montaner et al.     |
|                               | MMP-9               | Brain tissue         | Rosell et al.       |
| Intracranial hemorrhage       | MMP-9               | Plasma               | Castellanos et al.  |
|                               | MMP-2,9             | Plasma               | Montaner et al.     |
|                               | MMP-9               | Brain tissue         | Rosell et al.       |
|                               | MMP-2,3,9, TIMP-1,2 | Plasma              | Alvarez-Sabin et al.|
| Intracranial aneurysm         | MMP-9, TIMP         | Plasma               | Kim et al.          |
|                               | MMP-2,9             | Serum                | Todor et al.        |
|                               | MMP-9               | Serum                | McGirt et al.       |
| Vascular dementia             | MMP-9               | Brain tissue         | Asahina et al.      |
|                               | MMP-2,3,9           | Brain tissue         | Rosenberg et al.    |
|                               | MMP-2,9             | Cerebrospinal fluid | Adair et al.        |
| Alzheimer’s disease           | MMP-1               | Brain tissue         | Leake et al.        |
|                               | MMP-3               | Brain tissue         | Yoshiyama et al.    |
|                               | MMP-9               | Brain tissue         | Ashahina et al.     |
|                               | MMP-2,9, TIMP-1,2   | Plasma               | Lorenzl et al.      |
|                               | MMP-9, TIMP-1       | Plasma               | Inspector et al.    |

**CLINICAL IMPLICATION OF MMPs IN ISCHEMIC STROKE**

Gelatin zymography has been used to compare MMP activity between infarcted and matched noninfarcted cerebral tissue of acute or chronic ischemic stroke patients. MMP-9 activity was markedly elevated in the acute ischemic stroke tissue, whereas increases in MMP-2 activity were subtle at 2–5 days after ischemic stroke. An association between elevated MMPs and BBB disruption is also supported by the significant correlation between the level of MMP-9 and stroke severity being observed in stroke patients. The MMP-9 levels measured by ELISA after acute cardioembolic stroke and the final NIHSS score are closely related, and MMP-9 appears also to be especially associated with hemorrhagic transformation. One shortcoming of analyzing the correlation between biomarkers and the clinicoradiologic outcome is the uncertainty of the cause-
and-effect relationship. An increase in plasma MMP-9 levels may result from an acute-phase reaction or prior systemic causes. However, some studies have found that high plasma MMP-9 levels were related to hemorrhagic transformation, because increased MMP-9 levels were detected on admission in patients who subsequently developed hemorrhagic transformation in whom the initial stroke severity and final infarct volume were similar to those without hemorrhagic transformation. Interestingly, a correlation between the baseline MMP-9 level and hemorrhagic complications after thrombolysis treatment has been observed, with the baseline MMP-9 level being a possible predictor of hemorrhagic complications after tPA thrombolysis. Measurement of the pretreatment level of MMP-9 was suggested as a possible safety measure when selecting candidates for thrombolysis. These data suggest a role for MMP-2 and MMP-9 in both the acute and chronic states of cerebral ischemia, BBB disruption, and efficacy and complication rate of tPA treatment. Future studies should focus on the temporal profiles of activation, selective activation among MMPs families, and the source of released MMPs.

**tPA TREATMENT AND THE PA AXIS**

PA and MMPs are the major proteases that modulate the ECM in the brain. Emerging data show important linkages between tPA, MMPs, vasogenic edema, and hemorrhage after stroke. Thrombolysis using tPA is the only FDA-approved treatment modality in acute stroke patients. The beneficial effect of tPA is intravascular thrombolysis restoring the blood flow to the ischemic parenchymal tissue. However, when tPA gains access to the extravascular neural parenchyma through a compromised BBB, any interaction between the NMDA receptors and exogenous tPA is a potential threat to the neural parenchyma. In addition to the direct neurotoxicity of tPA, vasogenic edema or parenchymal hemorrhage induced by BBB disruption is another deleterious complication. MMP-9 may attack components of the basal lamina of cerebral vessels, and its activation can cause neurovascular damage of the BBB, leading to edema and hemorrhagic complications. The increased risk of edema and hemorrhagic complications in stroke patients following tPA therapy may therefore be due to tPA-induced upregulation of MMP-9. Plasmin is a potent protease that cleaves blood fibrin and activates other proteases, and thus is implicated in neurovascular and ECM degradation. Plasmin has an important role in the mechanism of tPA-induced hemorrhage by activating MMP-9 and MMP-2. Whilst tPA can resolve intravascular blood clots, it can also produce abnormally high levels of plasmin that can trigger excessive activation of MMPs, leading to uncontrollable BBB disruption and, finally, intracranial hemorrhagic complications. MMP-9 levels are increased by tPA after experimentally induced focal ischemic stroke, and MMP inhibitors reduce tPA-induced hemorrhagic transformation. These data suggest that the activation of MMP-9 by administered tPA induces ischemic injury in the neurovascular unit. Therefore, modulating tPA and MMP-9 activity may provide a new approach for reducing complications of tPA thrombolysis. Once the clinical effects of MMPs are determined, clinical trials to modulate their activity could be started. Coadministration of MMP inhibitor or TIMPs with thrombolytic agents could avoid BBB disruption or ECM degradation. However, in clinical settings, such a drug should be highly tissue specific and avoid disturbing the physiologic ECM-rebuilding properties. Simple downregulation of MMP levels could evoke other side effects. Many MMP inhibitors have been tested for their efficacy in preventing cancer and rheumatoid arthritis, but their side effects have so far precluded clinical trials. Thus, balanced modulation of MMP represents the major challenge for future stroke therapies.

**MMPs IN STROKE RECOVERY**

MMPs are known to be rapidly upregulated at 24–48 hours after ischemia, but little is known about their role during the subsequent recovery phase. The mechanism of stroke recovery mainly involves neuronal plasticity, neurogenesis, and angiogenesis. MMPs may be critical for this recovery process because they
can remodel components of the ECM. MMPs participate in dendritic and axonal extension as well as blood vessel formation. A recent study showed a beneficial role of MMP-9 in the recovery phase of experimental stroke, with surrogate markers of neuronal plasticity and vascular remodeling being correlated with MMP-9 signals in neurons and astrocytes in the peri-infarction area. Delayed inhibition of MMPs at 7 days poststroke worsened outcomes at 14 days by decreasing neurovascular remodeling, increasing infarction volumes, and blunting behavioral recovery. MMPs may in fact play beneficial roles during delayed stroke recovery, despite their deleterious effects in the acute phase of stroke. MMPs, and in particular MMP-9, have emerged as a target for stroke therapy in recent years. Pathophysiologic data from cell and animal models have been increasingly validated by the correlations between MMP-9 biomarkers and clinical outcomes in patients. Therefore, there is growing momentum toward the development of MMP inhibitors for stroke therapy. In contrast to the therapeutic potential of MMP inhibition for acute stroke therapy, approaches to modulating or even enhancing MMPs in the late stages of stroke may represent novel techniques for improving stroke recovery and enhancing the long-term functional outcome. However, future studies should focus on how to modulate such a delicate and orchestrated role of MMPs according to the different phases of stroke.

MMPs IN SUBCORTICAL ISCHEMIC VASCULAR DEMENTIA

The significance of the white matter lesions that appear in various neurodegenerative diseases is still debated. The pathophysiology of subcortical ischemic vascular dementia or CADASIL (cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy) is significantly associated with white matter lesions. Demyelination around affected subcortical blood vessels, activated microglia, and macrophages can also be found in white matter lesions. Brain tissue in patients with vascular dementia show fibrosis of small blood vessels with adjacent demyelination. Increased levels of several MMPs have also been observed in some patients with vascular dementia, with MMP-3 observed in reactive astrocytes and microglial cells. After controlling for the variation of MMP expression with age, the level of active MMP-9 in the cerebrospinal fluid or plasma of patients with vascular dementia still varies from that in Alzheimer’s disease and other neurodegenerative conditions. MMPs may play an important role in nearly all kinds of dementia, especially in vascular dementia, but whether they are more important for the disease progress or are only an epiphenomenon of tissue adjustment is not clear. The profile of MMPs may in the future be a helpful marker for differentiating between subtypes of dementia.

CONCLUSIONS

MMPs are implicated in all cerebrovascular diseases and play a deleterious role in the pathophysiology of BBB disruption, neuroinflammation, oxidative stress, and delayed neuronal cell death. MMPs also have a role in the subacute or recovery phase through glial scarring, neuronal cell migration, and brain tissue recovery. Therapeutic strategies designed to modulate the activity of MMPs should consider these pleiotropic roles, and hence oversimplified experimental approaches with MMP inhibitors are likely to produce disappointing results. Carefully considering the various effects of MMPs is a challenge for future studies.

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