Density-dependent variability in an eruptive bark beetle and its value in predicting outbreaks

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Abstract. Several species of aggressive bark beetle in the genus Dendroctonus (Coleoptera: Curculionidae) undergo large fluctuations in population density with distinct outbreak and non-outbreak phases. We investigated attributes we hypothesized as subject to density-dependent variation (in particular, those likely to express phenotypic plasticity) in the southern pine beetle (SPB), Dendroctonus frontalis, as possible indicators of population fluctuations. These traits were morphology (body size and hindwing shape) and the sex ratio of trap-captured, dispersing SPB populations. We compared attributes of beetles from locations that ranged from having zero to high numbers (>1500) of SPB infestations (spots) at the county level for two summers. Southern pine beetle were obtained from six states in the southeastern USA and had been collected during a springtime, region-wide trapping survey used for forecasting outbreaks annually. Although we detected an expected but weak sexual dimorphism in both size and shape-related traits, no morphological differences were found between SPB collected from traps in counties with low or high densities of spots (≤10 or >10 per county, respectively). We found no relationship between numbers of SPB spots per county and trapped sex ratios in 2016, but we observed a strong trend in 2017, with about three times higher proportions of females trapped in counties with low compared with high numbers of spots. This implies that one or more known or possible factors influencing trapped sex ratios (e.g., disparities between the sexes in their responses to semiochemicals or in their propensity to disperse) can be density-dependent. Including trap-captured sex ratios in prediction models may improve current forecasting of SPB outbreaks in the southeastern USA, informing more timely and effective management of one of the most economically and ecologically important beetle species of this region.

Key words: bark beetles; density dependence; forest pests; geometric morphometrics; hindwing morphology; phenotypic plasticity; sex ratio; sexual dimorphism; southern pine beetle.

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INTRODUCTION

Population densities can alter availability of resources such as food and breeding substrate and influence a range of other pressures, and adaptive density-dependent phenotypic plasticity of relevant traits can offer a selective advantage under either low or high densities (Simpson
et al. 2005). Phenotypic plasticity can increase an organism’s fitness through adaptive modifications of morphological, behavioral, and other characteristics to address the shifting demands of variable environments (Stearns 1989, Leclaire and Brandl 1994). Phenotypic plasticity is more likely to evolve in species with fluctuating populations (Svanbäck et al. 2009), and consequently, density-dependent phenotypic plasticity occurs in eruptive pest insect species (Anstey et al. 2009, Whitman and Agrawal 2009). Non-adaptive plasticity in traits may also occur in response to density (Stearns 1989).

The dispersal capacity of individual insects is substantially determined by their morphology and behavioral traits, whereas changes in population density can create pressures for dispersal (Travis et al. 1999). Plasticity in dispersal-associated traits can be advantageous, as investment of energy and resources in dispersal can cause a reciprocal loss in reproductive output (Simmons and Thomas 2004, Homburg et al. 2013, Fraser et al. 2014). Morphological traits associated with flight capacity, and therefore dispersal ability, include body size and mass, wing loading (the ratio between body mass or size and wing area), and wing shape (Angelo and Slansky 1984, Arribas et al. 2012, Hassall 2015, Kalberer and Kölliker 2017). Insect species with density-dependent polyphenism in traits associated with flight capacity have been identified in several orders of insects, including Diptera (BitnerMathé and Klaczko 1999), Hemiptera (Fujisaki 1986, Mori and Nakasuji 1991), Hymenoptera (Kölliker-Ott et al. 2003), Lepidoptera (Angelo and Slansky 1984), and Orthoptera (Nishide and Tanaka 2013). Within Coleoptera, species that have been shown to exhibit differences in wing morphology associated with population phases include Callosobruchus maculatus Fabricius (Utida 1972), Hylobius abietis (L.) (Tan et al. 2010), Oreina cacalaiæ (Schrank) (Kalberer and Kölliker 2017), and Diabrotica virgifera virgifera LeConte (Mikac et al. 2019). Evidence for density-dependent plasticity has been reported in bark beetles (Coleoptera: Curculionidae: Scolytinae) (Bentz et al. 2011, Klutsch et al. 2020), and differing behavioral responses to tree defenses are known to occur in bark beetles from either outbreak or endemic populations (Lorio 1986, Wallin and Raffa 2004, Boone et al. 2011). Bark beetle behavioral responses to semiochemicals may exhibit plasticity depending on local population densities (Sullivan et al. 2011, Klutsch et al. 2020), and in the case of mountain pine beetle (Dendroctonus ponderosae Hopkins), individuals are less deterred by host pine defenses when at high densities (Wallin and Raffa 2004). However, the density dependence of flight-associated traits has received limited investigation in bark beetles (Sallé et al. 2005, Jones et al. 2019, Shegelski et al. 2019).

Certain species of bark beetles produce population eruptions that result in devastating, widespread mortality of their host trees (Turchin et al. 1991, Birt 2011, Jarvis and Kulakowski 2015). Colonization of healthy, high-quality hosts by aggressive bark beetles occurs predominantly in association with these eruptions, when beetles are present in sufficient numbers to mass attack such hosts and overcome their defenses (Raffa and Berryman 1983, Martinson et al. 2013). Rates of reproduction are high on these high-quality hosts, which can contribute to rapid population growth (Martinson et al. 2013). However, at low population densities, bark beetles are restricted to hosts with compromised defenses that are typically widely dispersed and low-quality (Clarke 2012, Raffa et al. 2016). Hence, alternative phenotypes (e.g., plasticity in wing morphology, body size, and responses to semiochemicals) may be better adapted for survival and reproduction under the specific conditions that result from either high or low population densities. For example, when beetle populations are low, individuals may have to fly longer distances to locate rarer and widely distributed hosts rendered susceptible by microclimate, disease, or localized disturbances (Raffa and Berryman 1980). During such periods, beetles possessing a wing size and wing loading that confers greater flight capacity should be at an advantage (Kausrud et al. 2012, Brown et al. 2017, Jones et al. 2019, Shegelski et al. 2019). Likewise, responses to semiochemicals may differ during outbreak and non-outbreak periods (Miller et al. 2005). Differing concentrations and combinations of semiochemicals may signal a host’s susceptibility to colonization (Erbilgin et al. 2003, Klutsch et al. 2020), which differs between periods of high or low density of aggressive bark beetle species.

Southern pine beetle (SPB), Dendroctonus frontalis Zimmermann, is an aggressive bark beetle
This native pest has caused significant destruction of pine trees in the southeastern USA, particularly on loblolly \( (\text{Pinus taeda} \ (L.) \) and shortleaf pine \( (\text{P. echinata} \ 	ext{Mill.}) \), causing over a billion dollars in economic losses over the past three decades (Clarke and Nowak 2009, Pye et al. 2011). In recent years, SPB has expanded its range northward, taking advantage of warmer winter temperatures and attacking novel \( \text{Pinus} \) spp. hosts in the northeastern USA (Lesk et al. 2017, Dodds et al. 2018). Regional SPB outbreaks in the southern USA occur periodically but are largely unpredictable (Pye et al. 2011, Weed et al. 2017), although factors including weather, predation, and host availability have been extensively investigated as possible drivers of population cycles (Beal 1933, Turchin et al. 1991, 1999, Friedenberg et al. 2008, Clarke and Nowak 2009, Nowak et al. 2015, Reeve 2018).

To address this unpredictability, a model for same-year forecasting of SPB outbreaks was developed that utilizes data from a network of pheromone-baited traps deployed each spring throughout the southeastern USA (Billings 1988, 2011). More recent, ongoing investigations have resulted in the development of a new, updated model for predicting SPB outbreak risk based on current year SPB trap catches, spot abundances the year before last, and the previous year’s trap catches of the primary predator of SPB, \( \text{Thanasi-} \) \( \text{mus dubius} \) (Fabricius) \( \text{(Coleoptera: Cleridae)} \) (Aoki 2017). However, predictions for the data in this study (2016 and 2017) were based on the methodology outlined below. Data on previous year spot abundances and trap catch data of both SPB and its predator \( T. \) \( \text{dubius} \) were entered into the model to generate a rough prediction of spots per county for the coming summer and predict if levels of SPB are increasing, static, or declining (Billings and Upton 2010). Advance warning of an outbreak provides forest managers additional time for preparations such as scheduling of spot detection flights and acquisition of resources necessary for anticipated management efforts. Between 1999 and 2005 in 12 surveyed states, the mean prediction accuracy for outbreak occurrence in a particular county was 82%, and for change in infestation, abundance was 74% (Billings and Upton 2010). However, forecast reliability was low when SPB populations were increasing or high, which is also when accurate foreknowledge is most critical to resource protection. This insufficiency has spurred recent methodology changes aimed at improving efforts to monitor SPB populations, and these include adoption of a revised lure in the monitoring traps (Sullivan et al. 2011, 2016) and a longer trapping interval. We posit that monitoring of density-correlated traits of beetles in the trap catches may offer a novel basis for detecting and forecasting SPB population eruptions, potentially augmenting or superseding the data currently used for predicting outbreaks.

Bark beetle sex ratio is a trait subject to variability that is potentially influenced by or correlated with population densities (Lobinger 1996, James et al. 2016). Although the sex ratio of emergent brood SPB has been reported to be approximately 1:1 (Stephen 2011), sex ratios of SPB caught in pheromone-baited traps are typically male-biased and vary considerably (Sullivan 2005, Staeben et al. 2015, Sullivan et al. 2016), and may deviate substantially from the sex ratio of adults in the local population (Hughes 1976, Moser and Browne 1978, Cronin et al. 2000). This disparity is apparently due at least in part to behavioral differences between the sexes in response to the aggregation attractant (Hughes 1976, Moser and Browne 1978, McCarty et al. 1980). One of the authors (BTS) has observed that sex ratios of SPB in pheromone-baited traps vary between years and over relatively long distances, indicating trap-captured sex ratio may be a population-associated trait with interannual fluctuations possibly linked to density. Further, sex ratios of trapped SPB are more male-biased inside SPB infestations than in the surrounding forest (Sullivan et al. 2011), implying that a correlation exists between trapped sex ratio and local abundance of colonized trees.

The objective of our study was to identify traits of SPB trap catches—beyond the number of trapped beetles—that are correlated to population
densities, as these might have value for predicting outbreaks. We investigated the quantitative associations between the summertime number of spots per county and two qualities of catches in the spring SPB trapping survey: (1) beetle morphology (body size and hindwing size and shape), and (2) sex ratios. These qualities were chosen because they are easily quantifiable and are likely to be plastic and influenced by population density.

**METHODS**

**Data**

*Trapping survey.*—For this study, we examined SPB specimens collected during the annual springtime SPB trapping survey coordinated by the USDA Forest Service Forest Health Protection (FHP). The survey uses 12-unit Lindgren multiple-funnel traps (Lindgren 1983) baited with devices releasing the SPB pheromone component frontalin and the pine odors alpha- and beta-pinene (Billings and Upton 2010, Billings 2011). Starting in 2017, the survey lure also included the pheromone component endo-brevicomin, which increases attractiveness of the lure by approximately an order of magnitude (Sullivan and Mori 2009, Sullivan et al. 2011).

Traps were deployed in hardwood inclusions within mature pine stands and at least ~10 m from any live pines to avoid initiating an SPB attack (Billings and Upton 2010). Traps used for insect collection in our study were dispersed within boundaries of National Forests, and individual traps (3–6 per National Forest district with minimum 1.6-km spacing) were at approximately the same locations during both years of our study. Trapping was initiated at each location at the start of blooming for eastern redbud (*Cercis canadensis* L.), and catch collections were made on a weekly basis for 4–6 weeks. A total of 79,982 SPB specimens were collected, identified, sexed (according to Wood 1982), and counted.

*Selection of SPB adults.*—We used specimens collected in Alabama, Georgia, Mississippi, North Carolina, South Carolina, and Tennessee during 2016 and 2017 to assess morphological variation in SPB (Table 1). We sampled five beetles, at a minimum, from each trap within each year; if fewer than five SPB were collected in a trap, we excluded the trap from the study due to insufficient sample size. We randomly sampled beetles until two of each sex were obtained for each trap/year; the fifth beetle was randomly selected regardless of sex. In traps with fewer than two female SPB, we sampled additional males to obtain five total specimens. For assessment of morphological variation in SPB populations throughout the southeastern USA, we sampled 148 male and 86 female SPB from the 2016 trap catches, and 135 male and 102 female SPB from the 2017 catches (Table 1).

*Number of SPB spots per county.*—We obtained data on the SPB spots that occurred during the summer through fall of 2016 and 2017 from FHP, which documents the total number and locations of multiple-tree infestations (≥10 trees) at the county level at the end of each calendar year. Southern pine beetle spots are initially detected through repeated aerial surveys that begin in early summer, are identified aerially as clusters of fading or dead foliage in the forest canopy, and are recorded as infestations only after confirmation through ground-checks. We used the number of spots per county as the dependent variable in our analyses to be consistent with SPB outbreak risk prediction and preventative management efforts which are carried out at county level. The SPB specimens we examined were collected from 31 counties in 2016 and 34 counties in 2017. For the counties where we collected SPB samples for our study, the number of SPB spots per county ranged from 0 (numerous counties) to 277 (Franklin County, Mississippi, USA) spots in 2016, and from 0 (numerous

| State       | 2016 | 2017 |
|-------------|------|------|
| Alabama     | 23   | 15   |
| Georgia     | 19   | 27   |
| Mississippi | 28   | 33   |
| North Carolina | 11 | 16  |
| South Carolina | 4  | 9   |
| Tennessee   | 1    | 2    |
| Total       | 86   | 102  |

Table 1. Number of southern pine beetles sampled for morphological analysis in 2016 (*n* = 234) and 2017 (*n* = 237) from six states and 31 (2016) and 34 (2017) counties in the southeastern USA.
hurdle model features a count data model counts (Mullahy 1986, Zeileis et al. 2008). The a censored count distribution, for modeling zero a hurdle component, either a binomial model or negative binomial, for modeling positive counts and cated count component, typically Poisson or negative hurdle (i.e., counties with zero SPB spots; Mullahy 1986, Chen et al. 2016). The negative binomial distribution for \( y|x \) can be parameterized using the following probability density function (Cameron and Trivedi 2005, Zeileis et al. 2008): 

\[
f(y|x, \beta, \gamma) = \begin{cases} 
  f_{\text{zero}}(0|z, \gamma) \\
  (1 - f_{\text{zero}}(0|z, \gamma)) \times f_{\text{count}}(y|x, \beta) / (1 - f_{\text{count}}(0|x, \beta)) & \text{if } y > 0 
\end{cases}
\]

where the parameters \( \beta, \gamma \), and any additional dispersion parameters \( \theta \) (for negative binomial distributions) are estimated using a maximum likelihood procedure. The hurdle modeling framework is more flexible than zero-inflated models, as the count component is only used where the hurdle for modeling the occurrence of zeros is exceeded (i.e., only one source of zeros) (Zeileis et al. 2008, Chen et al. 2016). Due to overdispersion of the residuals for spots/county for both years, a truncated negative binomial model provided the best fit for modeling the positive counts (i.e., counties with \( \geq 1 \) SPB spot), and a binomial model with a logit link for the absence or presence of spots was used for the hurdle (i.e., counties with zero SPB spots; Mullahy 1986, Chen et al. 2016). The negative binomial distribution for \( y|x \) can be parameterized using the following probability density function (Cameron and Trivedi 2005, Zeileis et al. 2008): 

\[
f(y|x, \beta, \gamma) = \frac{\Gamma(y+\theta)}{\Gamma(\theta) \cdot y!} \frac{\mu^y \times \theta^\theta}{(\mu+\theta)^{y+\theta}}
\]

where \( \mu \) is the mean, \( \theta \) is the shape (dispersion) parameter, \( \Gamma(\cdot) \) is the gamma function, and maximum likelihood is used to estimate all parameters.

**SPB sex ratio**

SPB spots per county.—Due to the non-normal distribution of the residuals of spots/county, we used hurdle models (Eqs. 1, 2) to assess whether trapped SPB sex ratio was a predictor of the number of SPB spots per county during 2016 or 2017.

**SPB caught per trap per day.**—We used linear regressions to determine whether SPB sex ratio was a predictor of the mean number of SPB caught per trap per day (i.e., trap catch rates). Linear models take the following form (Seal 1967):

\[
y_i = \beta_0 + \beta_1 x_{i1} + \cdots + \beta_p x_{ip} + \epsilon_i
\]

where \( y_i \) is the response variable (SPB/trap/day), \( \beta_0 \) is the intercept, \( \beta \)'s are the slope coefficients, \( x \) values are the predictor variable(s) (sex ratio), and \( \epsilon_i \) is the error term. The residuals of SPB/trap/day were non-normally distributed for both years; therefore, we applied a log base 10 transformation to improve the fit of these models (Barrera-Gómez and Basagaña 2015). We ran analyses in R 1.0.136 using the pscl, readxl, and stat packages (Zeileis et al. 2008, Wickham and Bryan 2017, R Development Core Team 2019).

**SPB morphological trait variation**

We used measurements of three different morphological characteristics (length of right elytron,
length of right hind femur, Fig. 1a; and length of right hindwing, Fig. 1b) for assessing the overall size of each specimen. Pronotum width has previously been used as a single measurement for assessing overall size of Dendroctonus spp. (Armendáriz-Toledano et al. 2014a, Esch et al. 2016), but there is no literature demonstrating that this is a good proxy for overall body size (Arribas et al. 2012, Knapp and Knappová 2013, Kalberer and Kolliker 2017). Additionally, including measurements of multiple features from each individual can reduce the influence of allometry and provide a more robust assessment of overall body size (Knapp and Knappová 2013). We measured these features with an ocular micrometer to the nearest 0.1 μm and converted to mm for analyses.

We averaged SPB size measurements at the trap-level prior to analyses (n = 48 traps in 2016; n = 47 traps in 2017). We analyzed the two years of data separately due to the addition of the pheromone component endo-brevicom in to the trapping survey lure in 2017, as this has been shown to increase lure attractiveness (Sullivan and Mori 2009, Sullivan 2016) and could potentially influence which SPB phenotypes are attracted. We used Pearson’s correlation coefficients to evaluate allometry of trap-level elytra length, hind femur length, and hindwing length for both years of samples (Pearson 1895). We ran hurdle models (Eq. 1) to assess whether any of the average trap-level SPB size attributes were predictors of spots/county. To examine the relationships between body size traits and catch rates, we used linear models (Eq. 3) to regress log-transformed SPB/trap/day against the mean trap-level size measurements.

To account for any sexual size dimorphism that could bias our assessment of differences in morphological traits with sexes combined, we also averaged individual body size measurements by sex for each trap. We modeled relationships between male and female size measurements and spots/county using hurdle models (Eq. 1), and between male and female size measurements and SPB/trap/day using linear models (Eq. 3; 2016 n = 48 for males, n = 42 for females; 2017 n = 47 for males, n = 46 for females; numbers of replicates varied between sexes because some traps had no female SPB).

To test for sexual dimorphism, we regressed size measurements of individual beetles (n = 234 in 2016; n = 237 in 2017) against sex; all residuals were normally distributed, and thus, linear regression (Eq. 3) was appropriate to model the body size variables. We used dummy coding to include SPB sex as a predictor in these models (i.e., 1 and 0 were assigned for male and female, respectively; Cohen et al. 2003). Individual SPB were the unit of replication for these analyses, as trap-level means of size variables could not be used due to missing data (i.e., several traps had no females). We ran all analyses in R 1.0.136, and used the ggpurbr, readxl, pscl, and stat packages (Zeileis
et al. 2008, Wickham and Bryan 2017, Kassambara 2018, R Development Core Team 2019).

**Hindwing landmark and semilandmark acquisition**

We utilized landmark-based geometric morphometrics (GM) to quantify variation in hindwing shape among our specimens (Bookstein 1986, Webster and Sheets 2010, Zelditch et al. 2012). The GM technique summarizes shape as a configuration of landmarks (LMs; discrete, homologous points that can be identified in every specimen) that are captured in an image and plotted on Cartesian axes. Geometric morphometrics is a commonly used tool for detecting and assessing subtle morphological differences among individuals and populations (Bookstein 1986, Webster and Sheets 2010). The membranous wings in insects (the hindwings in Coleoptera) are particularly well suited for GM; their two-dimensional shape reduces possible distortions due to angle and depth-of-field that are possible with 3-dimensional subjects when photographed (Webster and Sheets 2010), and they possess numerous suitable LMs at the terminations and intersections of veins. Membranous wings have been shown to be a particularly useful feature for identifying morphological variation among insect populations (Oguz et al. 2017, Wilk-da-Silva et al. 2018).

We made slide mounts with the right hindwings by compressing them under a coverslip following immersion in 95% ethanol. We took micrographs with a Leica S6D digital camera, and the total hindwing lengths (to the nearest 0.001 mm) were measured using the Leica software’s measurement tool (Leica Microsystems, Wetzlar, Germany; Fig. 1b). In studies on hindwing morphology in Coleoptera, LMs have typically been chosen along the costal, radial, medial, and cubitus veins (Bai et al. 2011, Benítez et al. 2014, Su et al. 2015). We selected ten type I LMs (i.e., occurring at the discrete juxtaposition of features; in this study, points of termination or intersection along the medial, cubitus, and radial veins; LM1-9; Fig. 2a), and one type II LM (i.e., geometric constructs including minima/maxima of curves; LM10 was the most distal point of the wing margin; Fig. 2a; Zelditch et al. 2012).

We also assigned semilandmarks (SLMs) in order to provide a shape analysis of the curvature along the outline of the wing (Zelditch et al. 2012, Armendáriz-Toledano et al. 2014b). SLMs are points placed at evenly distributed increments along a curve or outline of a given feature and can be incorporated into a landmark-based analysis to provide a more robust characterization of shape (Webster and Sheets 2010, Zelditch et al. 2012). We used MakeFan8 (Sheets 2014) to superimpose over each micrograph a grid of 10 evenly spaced, parallel lines (a comb) oriented perpendicular to a line between LM1 and LM10. We assigned 16 SLMs where the gridlines intersected the anterior and posterior margins of the wings, beginning near the base of the costa and ending near the
terminus of the cubitus 1 vein (Fig. 2b; Hopkins 1909, Sheets 2014). We randomly ordered the hindwing images to reduce bias and then manually assigned the locations of both LMs and SLMs on each digital image to obtain their Cartesian coordinates using tpsDig2 version 2.30 software (Rohlf 2017a).

Analysis of hindwing shape.—To remove non-shape variation (i.e., size, position, and rotation of the hindwings in the micrographs) associated with the raw LM data, we ran a full Procrustes superimposition on these coordinates (Webster and Sheets 2010, Klingenberg 2011, Zelditch et al. 2012, Mikac et al. 2013). We generated a covariance matrix from this data set (Klingenberg 2011, Klingenberg et al. 2012) and derived a correlation matrix for multivariate analyses to account for heterogeneity in the variances. We ran a principal component analysis (PCA) on the Procrustes coordinates of 27 LMs and SLMs to examine overall patterns of wing shape variation in the sampled SPB populations for 2016 (n = 234) and 2017 (n = 237; Klingenberg et al. 2012). Additionally, we ran a discriminant function analysis (DFA) using the hindwing LMs and SLMs to determine whether these data could partition sampled SPB into low spot density (≤10 spots/county) and high spot density (>10 spots/county) groups based on hindwing shape (Konigsberg and Frankenberg 2018). The DFA used Fisher’s cutoff point (i.e., a score of zero) to separate discriminant scores into two groups corresponding to spot density, and Hotelling’s $T^2$ statistic was used to test for significant differences in discriminant scores (Lachenbruch 1967, Klingenberg 2011, Mikac et al. 2013). We performed a permutation test with 10,000 iterations to test the null hypothesis of equal group means, and the reliability of low spot density–high spot density discrimination was determined using leave-one-out cross-validation (Lachenbruch 1967). We applied an identical DFA and error rate estimation to test for the presence of sexual dimorphism in wing shape.

To test for allometry between hindwing shape and length, we ran a multivariate regression of shape against the centroid size (the square root of the sum of squared distances of landmarks from the object centroid) of each hindwing (Klingenberg et al. 2012, Mikac et al. 2013). Scores associated with changes in hindwing shape (i.e., direction of the shape change vectors) were the dependent variable, and centroid size was the independent variable (Drake and Klingenberg 2008, Klingenberg et al. 2012).

We used individual SPB hindwing centroid size as a metric of overall hindwing size to assess whether this attribute was a predictor of spots/county using hurdle models (Eq. 1). We used log-transformed centroid size to address test assumptions. To determine whether hindwing centroid size differed between male and female SPB, we ran linear regressions (Eq. 3) using sex as the predictor. Hindwing photos (with LMs and SLMs assigned) were consolidated into data-sets using tpsUtil32 version 1.74 software (Rohlf 2017b), geometric morphometric and multivariate analyses (including Procrustes superimposition and DFA) were performed in MorphoJ version 1.06d (Klingenberg 2014), and all models and PCAs were executed in R 1.0.136 using the pscl, readxl, and stat packages (Zeileis et al. 2008, Wickham and Bryan 2017, R Development Core Team 2019).

RESULTS

Sex ratio in trap catches

The trap-level sex ratios (female counts/male counts) ranged from 0 to 0.64 in 2016, and from 0.012 to 0.75 in 2017. In 2016, we found no relationship between the captured sex ratio and spots/county (Table 2). However, in 2017, there was a strong negative relationship between trapped sex ratio and SPB spots/county for both components of the hurdle model; traps that had a higher proportion of female SPB were associated with lower numbers of spots ($P < 0.001$, Table 2; Fig. 3). Similarly, trap-captured sex ratio was not a good predictor of SPB/trap/day in 2016 (Table 3). However, sex ratio increased as the mean number of SPB/trap/day decreased in 2017 (linear model, $F_{1,45} = 7.14$, $P = 0.01$, Table 3; Fig. 4).

Size variation

In both 2016 and 2017, we found strong positive Pearson correlations among hindwing length, elytra length, and femur length (for all three pairwise correlations, $P < 0.001$; Table 4). Mean trap-level elytra length, hindwing length,
and hind femur length were not significant predictors of spots/county in 2016 or 2017 either with data for sexes combined or analyzed separately (Tables 2, 5). Similarly, for both years, linear models did not show a relationship between the number of SPB/trap/day and the mean trap-level elytra length, hindwing length, or hind femur length with data for the sexes combined or analyzed separately (Table 3).

We found differences in elytra length between the sexes in both 2016 ($F_{1,232}= 6.40, P = 0.01$) and 2017 ($F_{1,235} = 19.01, P < 0.001$). On average, male elytra were 97.3% as long as female elytra in 2016 (male $1.735 \pm 0.012$ mm, female $1.783 \pm 0.017$ mm; mean ± SEM), and 96.2% as long as female elytra in 2017 (male $1.728 \pm 0.010$ mm, female $1.796 \pm 0.012$ mm), respectively. In 2016, we found no relationship between hindwing length and sex ($P = 0.34$). However, in 2017, hindwing lengths differed between the sexes ($F_{1,235} = 5.79, P = 0.02$); male hindwings were 97.9% as long as female hindwings (male $3.641 \pm 0.020$ mm, female $3.717 \pm 0.026$ mm). We did not detect any differences in hind femur lengths between males and females in 2016 ($P = 0.20$) or 2017 ($P = 0.10$).

**Hindwing shape variation**

In the 2016 PCA on LMs and SLMs, the first five principal components explained 68.7% of the overall variance in SPB hindwing shape. Plots of the first two components displayed nearly complete overlap of specimens from low spot density (≤10 spots/county) and high spot density (>10 spots/county) counties, indicating the absence of a strong association between hindwing shape and infestation abundances (Appendix S1: Fig. S1). The DFA revealed that there were differences in average shape between SPB captured in low and high spot density areas in 2016 ($T^2 = 122.0, P = 0.001$; Fig. 5a), with hindwings of SPB trapped in low spot density counties being slightly wider and longer. However, a relatively small

| Independent variable | Year | Model component | Estimated coefficient† | z | P       |
|----------------------|------|-----------------|------------------------|---|---------|
| Sex ratio§           | 2016 | Positive counts| −5.528                 | −0.664 | 0.51 |
|                      |      | Zero counts     | −3.431                 | −1.368 | 0.17 |
|                      | 2017 | Positive counts| −12.385                | −4.321 | <0.001 |
|                      |      | Zero counts     | −14.911                | −3.514 | <0.001 |
| Elytra length        | 2016 | Positive counts| −10.610                | −1.529 | 0.13 |
|                      |      | Zero counts     | 2.885                  | 3.684  | 0.43 |
|                      | 2017 | Positive counts| −0.175                 | −0.036 | 0.97 |
|                      |      | Zero counts     | −4.899                 | −0.918 | 0.36 |
| Hindwing length      | 2016 | Positive counts| −6.106                 | −1.208 | 0.23 |
|                      |      | Zero counts     | 1.480                  | 0.809  | 0.42 |
|                      | 2017 | Positive counts| −1.464                 | −0.636 | 0.52 |
|                      |      | Zero counts     | −1.405                 | −0.600 | 0.55 |
| Femur length         | 2016 | Positive counts| −11.324                | −0.798 | 0.43 |
|                      |      | Zero counts     | 3.765                  | 0.399  | 0.69 |
|                      | 2017 | Positive counts| −6.494                 | −0.529 | 0.60 |
|                      |      | Zero counts     | −14.285                | −0.988 | 0.32 |
| Centroid size¶       | 2016 | Positive counts| −5.851                 | −1.999 | 0.05 |
|                      |      | Zero counts     | 1.153                  | 0.686  | 0.49 |
|                      | 2017 | Positive counts| −1.582                 | −0.790 | 0.43 |
|                      |      | Zero counts     | −1.660                 | −0.846 | 0.40 |

*Note:* Significant results are shown in bold.

† Trap-level data.

‡ Estimated coefficients are the beta parameters from the count model (positive counts) and the hurdle model (zero counts) components, respectively.

§ Female counts/male counts.

¶ The square root of the sum of squared distances of hindwing landmarks from the wing centroid; it is a measure of overall wing size. Data were log-transformed.
Procrustes distance (0.00633) between the average hindwing shapes for the two groups indicated minimal overall shape variation corresponding to infestation density, and leave-one-out cross-validation of the DFA resulted in the misclassification of 36% (56 of 155) of low spot density SPB as high, and 44% (35 of 79) of high spot density SPB as low (Fig. 5b).

The DFA analyzing hindwing shape by sex in 2016 showed there were differences in average shape between female and male SPB ($T^2 = 138.4, P < 0.001$; Fig. 5c), with female hindwings being slightly longer and wider. However, the small Procrustes distance (0.00822) between average hindwing shapes for female and male SPB suggests minimal sexual dimorphism in this feature. Leave-one-out cross-validation misclassified 39.5% (34 of 86) of female SPB as males, and 33% (49 of 148) of male SPB as females (Fig. 5d).

In the 2017 PCA, the first five principal components explained 69% of the overall variation in hindwing shape. As in 2016, graphical assessment of the principal components indicated extensive overlap in the hindwing shapes of SPB collected from counties with high and low spot densities (Appendix S1: Fig. S2). The DFA revealed differences in average hindwing shape between SPB captured in low and high spot density counties in 2017 ($T^2 = 171, P < 0.001$; Fig. 6a), with SPB trapped in low spot density counties having slightly wider and longer hindwings; however, the Procrustes distance (0.00323) between the average shapes was small. Cross-validation resulted in the misclassification of 29.5% (38 of 129) of low spot density SPB as high, and of 33% (36 of 108) of high spot density SPB as low (Fig. 6b).

The DFA assessing hindwing shape by sex in 2017 showed that female SPB hindwings were slightly longer and wider than male hindwings ($T^2 = 131.2, P < 0.001$; Fig. 6c), although the Procrustes distance (0.00669) between the average shapes was small. Cross-validation misclassified 35% (36 of 102) of female SPB as males, and 37% (50 of 135) of male SPB as females (Fig. 6d).

For 2016, multivariate regression analysis assessing allometry within the hindwing data revealed that centroid size explained 25.2% of the total variation in hindwing shape ($R^2 = 0.25$; Fig. 3).

| Independent variable† | Year | $F$   | Total df | $P$   |
|-----------------------|------|-------|----------|-------|
| Sex ratio             | 2016 | 0.855 | 46       | 0.36  |
|                       | 2017 | 7.135 | 45       | 0.01  |
| Elytra length         | 2016 | 2.050 | 46       | 0.16  |
|                       | 2017 | 0.253 | 45       | 0.62  |
| Elytra length (male)  | 2016 | 3.396 | 46       | 0.07  |
|                       | 2017 | 1.642 | 45       | 0.21  |
| Elytra length (female)| 2016 | 0.659 | 40       | 0.42  |
|                       | 2017 | 0.0002 | 44    | 0.99  |
| Hindwing length       | 2016 | 0.173 | 46       | 0.68  |
|                       | 2017 | 1.087 | 45       | 0.30  |
| Hindwing length (male)| 2016 | 0.594 | 46       | 0.44  |
|                       | 2017 | 2.60  | 45       | 0.11  |
| Hindwing length (female)| 2016 | 0.054 | 40       | 0.82  |
|                      | 2017 | 0.152 | 44       | 0.70  |
| Femur length          | 2016 | 1.775 | 46       | 0.19  |
|                       | 2017 | 3.188 | 45       | 0.08  |
| Femur length (male)   | 2016 | 0.254 | 46       | 0.62  |
|                       | 2017 | 2.911 | 46       | 0.09  |
| Femur length (female) | 2016 | 0.449 | 40       | 0.51  |
|                      | 2017 | 0.216 | 44       | 0.64  |

Notes: SPB, southern pine beetle. Significant results are shown in bold ($\alpha \leq 0.05$).
† Data log-transformed for analyses.
‡ Morphological measurements averaged at the trap level; sex ratio has trap as unit of replication.
The linear regression of centroid size on SPB sex did not show a difference in hindwing centroid sizes between males and females in 2016 ($P = 0.32$). Using the hurdle model with positive spot counts (i.e., for $y > 0$; Eq. 1), we detected a marginally significant relationship between spots/county and hindwing centroid size, with SPB having slightly larger centroid sizes in counties with lower numbers of spots ($P = 0.05$; Table 2), but this relationship was not significant ($P = 0.49$) for counties with zero spots (i.e., for $y = 0$; Eq. 1).

For 2017, hindwing centroid size explained 35.2% of the total variation in hindwing shape ($R^2 = 0.35$; $P < 0.001$). Contrary to 2016, we found differences in centroid size between the sexes in 2017 ($F_{1,235} = 5.44$, $P = 0.02$): hindwing centroid size for males was 98% as large as female centroid size (male: 1802.6 ± 10.1, female: 1840.1 ± 12.8). The hurdle model showed no relationship between SPB spots/county and hindwing centroid size in 2017 (Table 2).

**DISCUSSION**

Relationship between SPB trait variation and the number of SPB spots per county

To our knowledge, there are no studies comparing multiple phenotypic traits of SPB adults collected from areas with differing levels of SPB damage, although variation has been observed in attributes of SPB and other *Dendroctonus* species that vary with population density (e.g., female SPB body size, oviposition rate, gallery length and density, and development time; Bentz et al. 2011, Stephen 2011, Foelker and Hofstetter 2014). However, we did not find compelling evidence for variation in trapped SPB body size (elytra length, hind femur length, or hindwing length) that was associated with the number of spots in surrounding forests. The size of our data set may have limited our ability to detect density-dependent variation. We had only two years of SPB trapping survey data available, and because the trap lure components differed between 2016 and 2017, we were unable to integrate morphological trait data and infestation numbers from these two years. However, the very small differences in size parameters associated with areas with high and low spot densities do not suggest that these differences are biologically relevant or adaptive.

**Relationship between SPB trait variation and SPB caught per trap per day**

The number of SPB captured in baited funnel traps has been shown to be significantly but inconsistently correlated with other indicators of local population density such as proximity of attacked trees (Sullivan et al. 2011; also recently demonstrated in the congeneric *D. ponderosae*, Klutsch et al. 2020). Raw SPB catches are broadly correlated with spot abundances and used currently as one indicator for forecasting SPB damage (Billings and Upton 2010). Intuitively,
springtime trap catches should be associated with SPB damage potential, since a threshold density of dispersing, pheromone-responding SPB is necessary for beetles to organize mass attacks to overcome host defenses and initiate spots (Stephen 2011, Schowalter 2012). However, we did not find a relationship between SPB body size attributes and trap catch numbers, providing corroborating evidence that beetle size is not a useful indicator of beetle population density and damage potential.

Relationship between hindwing shape and the number of SPB spots per county

Although we found in 2016 that SPB hindwing centroid size was correlated with spot numbers for counties that had at least one spot (Table 2), this relationship was only marginally significant and did not hold when counties with zero spots were included in the analysis. Combined with the absence of any trend in 2017, these data imply that wing centroid size would not be useful in predicting outbreaks and is similar to our

Table 5. Estimated coefficients, z values, and P values from the truncated negative binomial model (counties with ≥1 spot) and the binomial model with logit link (counties with zero spots) for hurdle models with morphological characters of trapped southern pine beetle as the independent variable and spots per county as the dependent variable for sexes analyzed separately.

| Independent variable† | Year | Model component | Estimated coefficient | z   | P§ |
|-----------------------|------|-----------------|----------------------|-----|-----|
| Elytra length (male)  | 2016 | Positive counts | −7.500               | −1.191 | 0.23  |
|                       |      | Zero counts     | −0.605               | −0.181 | 0.86  |
|                       | 2017 | Positive counts | −1.459               | −0.331 | 0.74  |
|                       |      | Zero counts     | −3.186               | −0.726 | 0.47  |
| Elytra length (female)| 2016 | Positive counts | −5.123               | −1.144 | 0.25  |
|                       |      | Zero counts     | −8.220               | 1.718  | 0.09  |
|                       | 2017 | Positive counts | 1.980                | 0.833  | 0.40  |
|                       |      | Zero counts     | −2.641               | −0.870 | 0.38  |
| Hindwing length (male)| 2016 | Positive counts | −4.186               | −1.109 | 0.27  |
|                       |      | Zero counts     | 0.003                | 0.002  | 0.99  |
|                       | 2017 | Positive counts | −3.246               | −1.351 | 0.18  |
|                       |      | Zero counts     | −0.415               | −0.201 | 0.84  |
| Hindwing length (female)| 2016 | Positive counts | −4.068               | −0.913 | 0.36  |
|                       |      | Zero counts     | 2.467                | 1.728  | 0.08  |
|                       | 2017 | Positive counts | 0.728                | 0.686  | 0.49  |
|                       |      | Zero counts     | −1.316               | −0.925 | 0.35  |
| Femur length (male)   | 2016 | Positive counts | −10.447              | −0.744 | 0.46  |
|                       |      | Zero counts     | 0.252                | 0.032  | 0.97  |
|                       | 2017 | Positive counts | −9.055               | −0.823 | 0.41  |
|                       |      | Zero counts     | −13.687              | −1.064 | 0.29  |
| Femur length (female) | 2016 | Positive counts | −4.671               | −0.438 | 0.66  |
|                       |      | Zero counts     | 8.801                | 1.240  | 0.21  |
|                       | 2017 | Positive counts | 3.010                | 0.525  | 0.60  |
| Centroid size (male)  | 2016 | Positive counts | −4.710               | −1.274 | 0.20  |
|                       |      | Zero counts     | −0.449               | −0.189 | 0.85  |
|                       | 2017 | Positive counts | −3.539               | −1.137 | 0.26  |
|                       |      | Zero counts     | 0.311                | 0.117  | 0.91  |
| Centroid size (female)| 2016 | Positive counts | −8.426               | −1.392 | 0.16  |
|                       |      | Zero counts     | 2.733                | 1.138  | 0.25  |
|                       | 2017 | Positive counts | 0.482                | 0.170  | 0.86  |
|                       |      | Zero counts     | −4.459               | −1.450 | 0.15  |

† Trap-level data for both sexes.
‡ None of the models were statistically significant (α ≤ 0.05).
§ The square root of the sum of squared distances of hindwing landmarks from the wing centroid; it is a measure of overall wing size. Data were log-transformed.
other size measurements in this respect. There were subtle average hindwing shape differences between SPB trapped in low vs. high spot density counties in 2016 and 2017, with hindwings of SPB from low spot density counties being slightly wider and more elongated. However, cross-validation indicated that hindwing shape scores were not a reliable metric for distinguishing SPB captured from low vs. high spot density counties in either year (63% and 70.5% accuracy in classifying low spot density SPB, and 56% and 67% accuracy in classifying high spot density SPB in 2016 and 2017, respectively; Figs. 5b, 6b). It is important to note that to assess differences in hindwing shape using DFA, the trapped SPB samples were partitioned into two, somewhat arbitrary groups (counties with >10 or ≤10 spots). However, the lack of clustering of beetles into distinct groups in the PCA indicates the absence of polymorphism (Appendix S1: Figs. S1, S2), and the strong overlap of clusters for our two spot density classes suggests that alternative partitioning of spot densities would likely not have permitted stronger group separation. Additionally, the
small difference in average shape and significant overlap in hindwing shape scores between SPB captured in lower vs. higher spot density locations (Figs. 5a, 6a) suggests that this phenotypic trait would have minimal if any utility in detecting shifts in population density.

Sexual dimorphism

Our results provide support for the existence of sexual size dimorphism in SPB, with female beetles being slightly larger than males (longer elytra in both years; longer hindwings and larger hindwing centroid sizes in 2017). Results from the DFA assessing hindwing shape variation also showed subtle differences between male and female SPB, with female hindwings being slightly more elongated and wider in both years as compared to males. However, the cross-validation on the hindwing shape discrimination indicated that shape was not a consistent indicator of sex (60.5% and 65% accuracy in classifying female SPB, and 67% and 63% accuracy in classifying male SPB in 2016 and 2017, respectively; Figs. 5d, 6d). Previous studies have shown that
female beetles, on average, are larger than males for several species in Dendroctonus (Bentz et al. 2011, Graf et al. 2012, Foelker and Hofstetter 2014, Lachowsky and Reid 2014, Liu et al. 2017). For example, there may be greater selective pressures on females than males for larger size, as the females initiate attacks on a new host and need adequate resources to locate the best environment for brood feeding and development (Elkin and Reid 2005, Bentz et al. 2011). The successful detection of expected sexual dimorphism with our data indicated the suitability of our morphometric procedures for exploring density-dependent variation in morphology. Furthermore, the very small differences between the sexes in our examined morphological traits suggest that it was unlikely that sex was a confounding factor in detecting morphological correlates of spot densities.

Captured SPB sex ratio

For one year of trapping data (2017), female SPB were trapped in substantially higher proportions in counties with lower infestation densities; the average trap-captured sex ratio was 0.294 in low spot density counties (≤10 spots/county) vs. 0.097 in high spot density counties (>10 spots/county). This trend was absent in 2016. The major procedural difference between years was the addition of endo-brevicomin to the trapping survey lure in 2017. Although endo-brevicomin has not influenced captured sex ratios of SPB in previous tests (i.e., it enhances attraction of both sexes similarly; Sullivan and Mori 2009, Sullivan 2016), it is possible that an undiscovered, density-dependent sexual dimorphism in effects of endo-brevicomin exists. Additionally, the average number of SPB spots per county was substantially higher in 2017 (~211 spots/county) than 2016 (~45 spots/county), and it is possible that a trend in trap-captured sex ratios was not detectable in 2016 due to an insufficient quantity of data from counties with high spot densities. Other studies have indicated an association between sex ratios in traps and local population densities in Dendroctonus. Catches in traps located inside of active SPB infestations (where beetle densities were ostensibly high) were substantially more male-biased than catches in traps deployed 100 and 200 m away among uninfested trees (Sullivan et al. 2011). In one of two years and with specific lure combinations, higher population densities of D. ponderosae were associated with a higher proportion of males in traps (Klutsch et al. 2020), as was observed with SPB in this study.

In 2017, we found that the proportion of males in traps increased with increasing trap catch rates (Fig. 4). This relationship is consistent with the correlation between spot abundance and trapped sex ratios observed in 2017 (Table 2, Fig. 3), since trap catches of SPB and spot abundances are known to be correlated, as discussed previously. Density dependence in sex ratios of trap-captured SPB could result from changes in the sex ratio within the dispersing population or in the sex-specific propensities of beetles to be caught in traps; however, these possibilities cannot be distinguished with our data. Sex ratios of emerging brood adult and reemerging parent adult SPB have not been found to differ significantly from 1:1 (Cooper and Stephen 1978, Coulson et al. 1979, Cook and Hain 1985). This suggests that skewed sex ratios in brood production or sexual differences in mortality rates in the host could not be factors contributing to strongly skewed sex ratios in the dispersing population or in trap catches. During outbreaks, traps are competing with a greater number of sources of natural attractant (i.e., mass-attacked trees), and traps and attacked trees likely differ in their relative attractiveness to the sexes. Male and female SPB differ in their responses to some semiochemicals. Two male-produced compounds in SPB, verbenone and acetophenone, reduce attraction of males more than females (Salom et al. 1992, Sullivan 2005), and likely other such semiochemicals exist. The odor blends of trap lures and trees are substantially different (Pureswaran and Sullivan 2012). Hence, if females have a stronger attraction than males to mass-attacked trees relative to traps, the higher numbers of these trees associated with outbreaks could produce male-skewed trap catches. Additionally, a semiochemical background in the environment can alter SPB responses to baited traps (B. T. Sullivan, unpublished data), and increased background of SPB semiochemicals within outbreak areas could affect male and female responses to trap lures differently.

Applications of findings for management and monitoring of SPB

The trend we documented in 2017, with lower proportions of female SPB in traps occurring in
counties that experienced higher numbers of beetle infestations, has important implications for managing SPB and predicting outbreaks. Inclusion of trap-captured sex ratios in the total data set currently used for outbreak forecasting (i.e., SPB trap catches, T. dubius catches, and spot densities of previous years) could contribute to the accuracy of these forecasts at local and regional levels. This potential will be assessed with trial collections of sex ratio data during the annual survey for the next several years. If implemented as part of the survey, collection of sex ratio data would likely require a more efficient means of sexing large numbers of insects than inspection of individuals under a dissecting microscope. However, relatively affordable, high-throughput molecular techniques such as qPCR could be used to determine sex ratios of beetles within homogenates of catch samples by quantifying proportions of sex-linked genes (Belousova et al. 2019). Hence, it may potentially be feasible to incorporating sex ratio data in SPB outbreak forecasting within the ongoing trap catch analyses, and these data may potentially contribute to longer-term sustainability and conservation of resources in the southeastern USA.

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**LITERATURE CITED**

Angelo, M. J., and F. Slansky. 1984. Body-building by insects: trade-offs in resource allocation with particular reference to migratory species. Florida Entomologist 67:22–41.

Anstey, M. L., S. M. Rogers, S. R. Ott, M. Burrows, and S. J. Simpson. 2009. Serotonin mediates behavioral gregarization underlying swarm formation in desert locusts. Science 323:627–630.

Aoki, C. F. 2017. Forest risk and irruptive insect pests: ecology for management in changing times. Dissertation. Dartmouth College, Hanover, New Hampshire, USA.

Armendáriz-Toledano, F., A. Niño, B. T. Sullivan, J. Macías-Sámano, J. Víctor, S. R. Clarke, and G. Zúñiga. 2014a. Two species within Dendroctonus frontalis (Coleoptera: Curculionidae): evidence from morphological, karyological, molecular, and crossing studies. Annals of the Entomological Society of America 107:11–27.

Armendáriz-Toledano, F., A. Niño, J. Macías-Sámano, and G. Zúñiga. 2014b. Review of the geographical distribution of Dendroctonus ritis (Curculionidae: Scolytinae) based on the geometric morphometrics of the seminal rod. Annals of the Entomological Society of America 107:748–755.

Arribas, P., J. Velasco, P. Abellán, D. Sánchez-Fernández, C. Andújar, P. Calosi, A. Millán, I. Ribera, and D. T. Bilton. 2012. Dispersal ability rather than ecological tolerance drives differences in range size between lentic and lotic water beetles. Journal of Biogeography 39:984–994.

Bai, M., E. McCullough, K.-Q. Song, W.-G. Liu, and X.-K. Yang. 2011. Evolutionary constraints in hind wing shape in Chinese dung beetles (Coleoptera: Scarabaeinae). PLOS ONE 6:1–12.

Barrera-Gómez, J., and X. Basagaña. 2015. Models with transformed variables: interpretation and software. Epidemiology 26:16–17.

Beal, J. A. 1933. Temperature extremes as a factor in the biology of southern pine beetle. Journal of Forestry 31:329–336.

Belousova, I., N. Ershov, S. Pavlushin, Y. Ilinsky, and V. Martemyanov. 2019. Molecular sexing of Lepidoptera. Journal of Insect Physiology 114:53–56.

Benítez, H. A., D. Lemic, R. Bažok, C. M. Gallardo-Araya, and K. M. Mikac. 2014. Evolutionary directional asymmetry and shape variation in Diabrotica virgifera virgifera (Coleoptera: Chrysomelidae): an example using hind wings. Biological Journal of the Linnean Society 111:110–118.
Bentz, B. J., R. R. Bracewell, K. E. Mock, and M. E. Pfrender. 2011. Genetic architecture and phenotypic plasticity of thermally-regulated traits in an eruptive species, Dendroctonus ponderosae. Evolutionary Ecology 25:1269–1288.

Billings, R. F. 1988. Forecasting southern pine beetle infestation trends with pheromone traps. Pages 295–306 in Integrated Control of Scolytid Bark Beetles: Proceedings of IUFRO Working Party and 17th International Congress of Entomology Symposium. Virginia Polytechnic Institute and State University, Blacksburg, Virginia, USA.

Billings, R. F. 2011. Aerial detection, ground evaluation, and monitoring of the southern pine beetle: state perspectives. Pages 129–139 in R. N. Coulson and K. D. Klepzig, editors. Southern pine beetle II. U.S. Department of Agriculture Forest Service, Southern Research Station, Asheville, North Carolina, USA.

Billings, R. F., and W. W. Upton. 2010. A methodology for assessing annual risk of southern pine beetle outbreaks across the southern region using pheromone traps. Pages 73–85 in Advances in threat assessment and their application to forest and rangeland management. General Technical Report PNW-GTR-802. U.S. Department of Agriculture Forest Service, Portland, Oregon, USA.

Birt, A. 2011. Population dynamics of southern pine beetle in forest landscapes. Pages 91–108 in R. N. Coulson and K. D. Klepzig, editors. Southern pine beetle II. U.S. Department of Agriculture Forest Service, Southern Research Station, Asheville, North Carolina, USA.

BittnerMathé, B. C., and L. B. Klaczko. 1999. Plasticity of Drosophila melanogaster wing morphology: effects of sex, temperature and density. Genetica 105:203.

Bookstein, F. L. 1986. Size and shape spaces for landmark data in two dimensions. Statistical Science 1:181–242.

Boone, C. K., B. H. Aukema, J. Bohlmann, A. L. Car- roll, and K. F. Raffa. 2011. Efficacy of tree defense physiology varies with bark beetle population density: a basis for positive feedback in eruptive species. Canadian Journal of Forest Resources 41:1174–1188.

Brown, S., V. Soroker, and G. Ribak. 2017. Effect of larval growth on adult body mass and long-distance flight endurance in a wood-boring beetle: Do smaller beetles fly better? Journal of Insect Physiology 98:327–335.

Cameron, A. C., and P. K. Trivedi. 2005. Microecometrics: methods and applications. Cambridge University Press, Cambridge, UK.

Chen, F., X. Ma, S. Chen, and L. Yang. 2016. Crash frequency analysis using hurdle models with random effects considering short-term panel data. International Journal of Environmental Research and Public Health 13:1–11.

Clarke, S. R. 2012. Implications of population phases on the integrated pest management of the southern pine beetle, Dendroctonus frontalis. Journal of Integrated Pest Management 3:1–9.

Clarke, S. R., and J. T. Nowak. 2009. Southern pine beetle. Forest Insect and Disease Leaflet No. 49. U.S. Department of Agriculture Forest Service, Washington, D.C., USA.

Cohen, J., P. Cohen, S. G. West, and L. S. Aiken. 2003. Applied multiple regression/correlation analysis for the behavioral sciences. Third edition. Lawrence Erlbaum Associates, Mahwah, New Jersey, USA.

Cook, S. P., and F. P. Hain. 1985. Comparison of loblolly and shortleaf pine bolts as hosts of the southern pine beetle, Dendroctonus frontalis Zimmerman (Coleoptera: Scolytidae). Environmental Entomology 14:332–335.

Cooper, M. E., and F. M. Stephen. 1978. Parent adult reemergence in southern pine beetle populations. Environmental Entomology 7:574–577.

Coulson, R. N., W. S. Fargo, P. E. Pulley, D. N. Pope, J. L. Foltz, and A. M. Bunting. 1979. Spatial and temporal patterns of emergence for within-tree populations of Dendroctonus frontalis (Coleoptera: Scolytidae). Canadian Entomologist 111:273–287.

Cronin, J. T., J. L. Hayes, and P. Turchin. 2000. Evaluation of traps used to monitor southern pine beetle aerial populations and sex ratios. Agricultural and Forest Entomology 2:69–76.

Dodds, K. J., C. F. Aoki, A. Arango-Velez, J. Cancel- liere, A. W. D’Amato, M. F. DiGirolomo, and R. J. Rabaglia. 2018. Expansion of southern pine beetle into northeastern forests: management and impact of a primary bark beetle in a new region. Journal of Forestry 116:178–191.

Drake, A. G., and C. P. Klingen berg. 2008. The pace of morphological change: historical transformation of skull shape in St Bernard dogs. Proceedings of the Royal Society of London B: Biological Sciences 275:71–76.

Elkin, C. M., and M. L. Reid. 2005. Low energy reserves and energy allocation decisions affect reproduction by mountain pine beetles, Dendro- tonus ponderosae. Functional Ecology 19:102–109.

Erbilgin, N., J. S. Powell, and K. F. Raffa. 2003. Effect of varying monoterpenic concentrations on the response of Ips pini (Coleoptera: Scolytidae) to its aggregation pheromone: implications for pest management and ecology of bark beetles. Agricultural and Forest Entomology 5:269–274.

Esch, E. D., D. W. Langor, and J. R. Spence. 2016. Gallery success, brood production, and condition of
mountain pine beetles (Coleoptera: Curculionidae) reared in whitebark and lodgepole pine from Alberta, Canada. Canadian Journal of Forest Research 46:557–563.

Foelker, C. J., and R. W. Hofstetter. 2014. Heritability, fecundity, and sexual size dimorphism in four species of bark beetles (Coleoptera: Curculionidae: Scolytinae). Annals of the Entomological Society of America 107:143–151.

Fraser, C. I., O. Brahy, P. Mardulyn, L. Dohet, F. Mayer, and J.-C. Grégoire. 2014. Flying the nest: Male dispersal and multiple paternity enables extrafamilial matings for the invasive bark beetle Dendroctonus micans. Heredity 113:327–333.

Friedenberg, N. A., S. Sarkar, N. Kouchoukos, R. F. Billings, and M. P. Ayres. 2008. Temperature extremes, density dependence, and southern pine beetle (Coleoptera: Curculionidae) population dynamics in east Texas. Environmental Entomology 37:650–659.

Fujisaki, K. 1986. Reproductive properties of the oriental chinch bug, Cavelierius saccharivorus Okajima (Heteroptera: Lygaeidae), in relation to its wing polymorphism. Researches on Population Ecology 28:43–52.

Graf, M., M. L. Reid, B. H. Aukema, and B. S. Lindgren. 2012. Association of tree diameter with body size and lipid content of mountain pine beetles. Canadian Entomologist 144:467–477.

Hain, F. P., A. J. Duehl, M. J. Gardner, and T. L. Payne. 2011. Natural history of the southern pine beetle. Pages 13–24 in R. N. Coulson and K. D. Klepzig, editors. Southern pine beetle II. U.S. Department of Agriculture Forest Service, Southern Research Station, Asheville, North Carolina, USA.

Hassall, C. 2015. Strong geographical variation in wing aspect ratio of a damselfly, Calopteryx maculata (Odonata: Zygoptera). PeerJ 3:e1219.

Homburg, K., A. Schuldt, C. Drees, and T. Assmann. 2013. Broad-scale geographic patterns in body size and hind wing development of western Palearctic carabid beetles (Coleoptera: Carabidae). Ecography 36:166–177.

Hopkins, A. D. 1909. Contributions toward a monograph of the Scolytid beetles. Technical Series 17: pt. 1–2. U.S. Bureau of Entomology, Washington, D.C., USA.

Hughes, P. R. 1976. Response of female southern pine beetles to the aggregation pheromone frontal. Zeitschrift für Angewandte Entomologie 81:463–466.

James, P. M., J. K. Janes, A. D. Roe, and B. J. Cooke. 2016. Modeling landscape-level spatial variation in sex ratio skew in the mountain pine beetle (Coleoptera: Curculionidae). Environmental Entomology 45:790–801.

Jarvis, D. S., and D. Kulakowski. 2015. Long-term history and synchrony of mountain pine beetle outbreaks in lodgepole pine forests. Journal of Biogeography 42:1029–1039.

Johnson, P. C., J. E. Coster, and Z. L. Florence. 1982. Behavioral and genetic diversity during dispersal: analysis of a polymorphic esterase locus in southern pine beetle, Dendroctonus frontalis. Environmental Entomology 11:1014–1018.

Jones, K. L., V. A. Shegelski, N. G. Marculis, A. N. Wijerathns, and M. L. Evenden. 2019. Factors influencing dispersal by flight in bark beetles (Coleoptera: Curculionidae: Scolytinae): from genes to landscapes. Canadian Journal of Forest Research 49:1024–1041.

Kalberer, N. M., and M. Kölliker. 2017. Flight dimorphism is related to survival, reproduction and mating success in the leaf beetle Oreina cacaliae. Ecological Entomology 42:355–363.

Kassambara, A. 2018. ggpubr: ’ggplot2’ Based publication ready plots. R package version 0.1.8. https://cran.r-project.org/web/packages/ggpubr/readme/README.html

Kausrud, K., B. Økland, O. Skarpaas, J.-C. Grégoire, N. Erbilgin, and N. C. Stenseth. 2012. Population dynamics in changing environments: the case of an eruptive forest pest species. Biological Reviews 87:34–51.

Klingenberg, C. P. 2011. MorphoJ: an integrated software package for geometric morphometrics. Molecular Ecology Resources 11:353–357.

Klingenberg, C. P. 2014. MorphoJ [Integrated computer software]. Apache License, Version 2.0. The University of Manchester, Manchester, UK.

Klingenberg, C. P., S. Duttkse, S. Whelan, and M. Kim. 2012. Developmental plasticity, morphological variation and evolvability: a multilevel analysis of morphometric integration in the shape of compound leaves. Journal of Evolutionary Biology 25:115–129.

Klutsch, J. G., G. Classens, C. Whitehouse, J. F. Cahill Jr, and N. Erbilgin. 2020. Density-dependent responses of mountain pine beetle to its pheromones and host volatiles in naïve lodgepole pine stands. Forest Ecology and Management 472. https://doi.org/10.1016/j.foreco.2020.118257.

Knapp, M., and J. Knappová. 2013. Measurement of body condition in a common carabid beetle, Pecicus cupreus: a comparison of fresh weight, dry weight, and fat content. Journal of Insect Science 13:1–10.

Kölliker-Ott, U. M., M. W. Blows, and A. A. Hoffmann. 2003. Are wing size, wing shape and asymmetry related to field fitness of Trichogramma egg parasites? Oikos 100:563–573.
Konigsberg, L. W., and S. R. Frankenberg. 2018. Typi-
cality and predictive distributions in discriminant
function analysis. Human Biology 90:31–44.
Lachenbruch, P. A. 1967. An almost unbiased method
of obtaining confidence intervals for the probabil-
ity of misclassification in discriminant analysis.
Biometrics 23:639–645.
Lachowsky, L. E., and M. L. Reid. 2014. Developmental
mortality increases sex-ratio bias of a size-dimorphic
bark beetle. Ecological Entomology 39:300–308.
Leclaire, M., and R. Brandl. 1994. Phenotypic plasticity
and nutrition in a phytophagous insect: consequences
of colonizing a new host. Oecologia 100:379–385.
Lesk, C., E. Coffel, A. W. Damato, K. Dodds, and R.
Horton. 2017. Threats to North American forests
from southern pine beetle with warming winters.
Nature Climate Change 7:713–717.
Lindgren, B. S. 1983. A multiple funnel trap for Scoly-
tid beetles (Coleoptera). Canadian Entomologist
115:299–302.
Liu, Z., B. Xu, Y. Guo, K. F. Raffa, and J. Sun. 2017. Gal-
lery and acoustic traits related to female body size
mediate male mate choice in a bark beetle. Animal
Behaviour 125:41–50.
Lobinger, G. 1996. Variations in sex ratio during an
outbreak of Ips typographus (Col., Scolytidae) in
Southern Bavaria. Umweltschutz 69:51–53.
Lorio Jr., P. L. 1986. Growth–differentiation balance: a
basis for understanding southern pine beetle-tree
interactions. Forest Ecology and Management
14:259–273.
Martinson, S. J., T. Tlioja, B. T. Sullivan, R. F. Billings,
and M. P. Ayres. 2013. Alternate attractors in the
population dynamics of a tree-killing bark beetle.
Population Ecology 55:95–106.
McCarty, F. A., P. M. Billings, J. V. Richerson, T. L.
Payne, and L. J. Edson. 1980. Response of the
southern pine beetle to behavioral chemicals in the
laboratory. Journal of the Georgia Entomological
Society 15:307–317.
Mikac, K. M., J. Douglas, and J. L. Spencer. 2013. Wing
shape and size of western corn rootworm (Coleop-
tera: Chrysomelidae) is related to sex and resis-
tance to soybean-maize crop rotation. Journal of
Economic Entomology 106:1517–1524.
Mikac, K. M., D. Lemic, H. A. Benitez, and R. Bažok.
2019. Changes in corn rootworm wing morphology
are related to resistance development. Journal of
Pest Science 92:443–451.
Miller, D. R., B. S. Lindgren, and J. H. Borden. 2005.
Dose-dependent pheromone responses of moun-
tain pine beetle in stands of lodgepole pine. Envi-
rmental Entomology 34:1019–1027.
Mori, K., and F. Nakasuji. 1991. Effects of day length
and density on development and wing form of the
small brown planthopper, Laodelphax striatellus
(Hemiptera: Delphacidae). Applied Entomology
and Zoology 26:557–561.
Moser, J. C., and L. E. Browne. 1978. A nondestruc-
tive trap for Dendroctonus frontalis Zimmerman (Coleop-
tera: Scolytidae). Journal of Chemical Ecology 4:1–7.
Mullahy, J. 1986. Specification and testing of some
modified count data models. Journal of Economet-
rics 33:341–365.
Nishide, Y., and S. Tanaka. 2013. Wing dimorphism in
the migratory locust, Locusta migratoria: differentia-
tion of wing morph and phase polyphenism. Ento-
mological Science 16:421–431.
Nowak, J. T., J. R. Meeker, D. R. Coyle, C. A. Steiner,
and C. Brownie. 2015. Southern pine beetle infesta-
tions in relation to forest stand conditions, previ-
sous thinning, and prescribed burning: evaluation of
the southern pine beetle prevention program.
Journal of Forestry 113:454–462.
Oguz, G., O. E. Kasap, and B. Alten. 2017. Wing mor-
phology variations in a natural population of Phle-
botomolus tobbi Adler and Theodor 1930. Journal of
Vector Ecology 42:223–232.
Pearson, K. 1895. Notes on regression and inheritance
in the case of two parents. Proceedings of the Royal
Society of London B: Biological Sciences 58:240–242.
Price, T. S., C. Doggett, J. M. Pye, and T. P. Holmes, edi-
tors. 1992. A history of southern pine beetle out-
breaks in the southeastern United States. Sponsored
by: Southern Forest Insect Work Conference. The
Georgia Forestry Commission, Macon, Georgia,
USA.
Pureswaran, D. S., and B. T. Sullivan. 2012. Semio-
chemical emission from individual galleries of the
southern pine beetle, (Coleoptera: Curculionidae:
Scolytinae), attacking standing trees. Journal of
Economic Entomology 105:140–148.
Pye, J. M., T. P. Holmes, J. P. Prestemon, and D. N.
Wear. 2011. Economic impacts of the southern pine
beetle. Pages 213–222 in R. N. Coulson and K. D.
Klepzig, editors. Southern pine beetle II. U.S.
Department of Agriculture Forest Service, South-
ern Research Station, Asheville, North Carolina,
USA.
R Development Core Team. 2019. R: A language and
environment for statistical computing. R Founda-
tion for Statistical Computing, Vienna, Austria.
Raffa, K. F., M. N. Andersson, and F. Schlyter. 2016.
Host selection by bark beetles: playing the Odds in
a high-stakes game. Pages 1–74 in Advances in
insect physiology. University of Oxford, Academic
Press, London, UK.
Raffa, K. F., and A. A. Berryman. 1980. Flight responses
and host selection by bark beetles. Pages 213–233
in Proceedings of the 2nd IUFRO Conference.
Raffa, K. F., and A. A. Berryman. 1983. The role of host plant resistance in the colonization behavior and ecology of bark beetles (Coleoptera: Scolytidae). Ecological Monographs 53:27–49.

Reeve, J. D. 2018. Synchrony, weather, and cycles in southern pine beetle (Coleoptera: Curculionidae). Environmental Entomology 47:19–25.

Rohlf, F. J. 2017a. tpsDIGw32 [Computer software] Version 2.30. State University of New York at Stony Brook, Stony Brook, New York, USA.

Rohlf, F. J. 2017b. tpsUtil32 [Computer software] Version 1.74. Morphometrics: State University of New York at Stony Brook, Stony Brook, New York, USA.

Salom, S. M., R. F. Billings, W. W. Upton, M. J. Dalusky, D. M. Grosman, T. L. Payne, C. W. Berisford, and T. N. Shaver. 1992. Effect of verbenone enantiomers and racemic endo-brevicomin on response of Dendroctonus frontalis (Coleoptera, Scolytidae) to attractant-baited traps. Canadian Journal of Forest Research 22:925–931.

Simmons, A. D., and C. D. Thomas. 2004. Changes in dispersal during species’ range expansions. American Naturalist 164:378–395.

Simpson, S. J., G. A. Sword, and A. De Loof. 2005. Advances, controversies and consensus in locust phase polyphenism research. Journal of Orthoptera Research 14:213–222.

Staeben, J. C., B. T. Sullivan, J. T. Nowak, and K. J. K. Gandhi. 2015. Enantiospecific responses of southern pine beetle (Dendroctonus frontalis) and its clerid predator, Thanasimus dubius, to α-pinene. Chemoecology 25:73–83.

Stearns, S. C. 1989. The evolutionary significance of phenotypic plasticity. BioScience 39:436–445.

Stephen, F. M. 2011. Southern pine beetle population dynamics in trees. Pages 51–74 in R. N. Coulson and K. D. Klepzig, editors. Southern pine beetle II. U.S. Department of Agriculture Forest Service, Southern Research Station, Asheville, North Carolina, USA.

Su, J., K. Guan, J. Wang, and Y. Yang. 2015. Significance of hind wing morphology in distinguishing genera and species of cantharid beetles with a geometric morphometric analysis. ZooKeys 502:11–25.

Sullivan, B. T. 2005. Electrophysiological and behavioral responses of Dendroctonus frontalis (Coleoptera: Curculionidae) to volatiles isolated from conspecifics. Journal of Economic Entomology 98:2067–2078.

Sullivan, B. T. 2016. Semiochemicals in the natural history of southern pine beetle Dendroctonus frontalis Zimmermann and their role in pest management. Pages 129–193 in Advances in insect physiology. University of Oxford, Academic Press, London, UK.

Sullivan, B. T., C. Brownie, and J. P. Barrett. 2016. Intra-annual variation in responses by flying southern pine beetles (Coleoptera: Curculionidae: Scolytinae) to pheromone component endo-brevicomin. Journal of Economic Entomology 109:1720–1728.

Sullivan, B. T., M. J. Dalusky, K. Mori, and C. Brownie. 2011. Variable responses by southern pine beetle, Dendroctonus frontalis Zimmermann, to the pheromone component endo-brevicomin: influence of enantiomer composition, release rate, and proximity to infestations. Journal of Chemical Ecology 37:403–411.

Sullivan, B. T., and K. Mori. 2009. Spatial displacement of release point can enhance activity of an attractant pheromone synergest of a bark beetle. Journal of Chemical Ecology 35:1222–1233.

Svanbäck, R., M. Pineda-Krch, and M. Doebeli. 2009. Fluctuating population dynamics promotes the evolution of phenotypic plasticity. American Naturalist 174:176–189.

Tan, J. Y., D. Wainhouse, K. R. Day, and G. Morgan. 2010. Flight ability and reproductive development in newly emerged pine weevil Hylobius abietis and the potential effects of climate change. Agricultural and Forest Entomology 12:427–434.

Thatcher, R. C., J. L. Searcy, J. E. Coster, and G. D. Her tel, editors. 1980. The southern pine beetle. Application Program, Science and Education Administration, U.S. Department of Agriculture Forest Service, Pineville, Louisiana, USA.
Travis, J. M., D. J. Murrell, and C. Dytham. 1999. The evolution of density-dependent dispersal. Proceedings of the Royal Society of London B: Biological Sciences 266:1837–1842.

Turchin, P., P. L. Lorio Jr., A. D. Taylor, and R. F. Billings. 1991. Why do populations of southern pine beetle (Coleoptera: Scolytidae) fluctuate? Environmental Entomology 20:401–409.

Turchin, P., A. D. Taylor, and J. D. Reeve. 1999. Dynamic role of predators in population cycles of a forest insect: an experimental test. American Association for the Advancement of Science 285:1068–1071.

Utida, S. 1972. Density dependent polymorphism in the adult of Callosobruchus maculatus (Coleoptera, Bruchidae). Journal of Stored Products Research 8:111–125.

Wallin, K. F., and K. F. Raffa. 2004. Feedback between individual host selection behavior and population dynamics in an eruptive herbivore. Ecological Monographs 74:101–116.

Webster, M., and H. D. Sheets. 2010. A practical introduction to landmark-based geometric morphometrics. Pages 163–188 in Quantitative methods in paleobiology. The Paleontological Society Papers. Volume 16. Cambridge University Press, Cambridge, UK.

Weed, A. S., M. P. Ayres, A. M. Liebhold, and R. F. Billings. 2017. Spatio-temporal dynamics of a tree-killer beetle and its predator. Ecography 40:221–234.

Whitman, D., and A. Agrawal. 2009. What is phenotypic plasticity and why is it important? Pages 1–63 in Phenotypic plasticity of insects: Mechanisms and consequences. CRC Press, Boca Raton, Florida, USA.

Wickham, H., and J. Bryan. 2017. readxl: Read Excel Files. R package version 1.0.0. https://cran.r-project.org/web/packages/readxl/readxl.pdf

Wilk-da-Silva, R., M. M. Cavalcanti de Souza Leal Diniz, M. T. Marrelli, and A. B. Bruno Wilke. 2018. Wing morphometric variability in Aedes aegypti (Diptera: Culicidae) from different urban built environments. Parasites & Vectors 11:1–9.

Wood, S. L. 1982. The bark and ambrosia beetles of North and Central America (Coleoptera: Scolytidae), a taxonomic monograph. Brigham Young University Press, Provo, Utah, USA.

Zeileis, A., C. Kleiber, and S. Jackman. 2008. Regression models for count data in R. Journal of Statistical Software 27. http://www.jstatsoft.org/v27/i08/

Zelditch, M. L., D. L. Swiderski, and H. D. Sheets. 2012. Geometric morphometrics for biologists: a primer. Second edition. Academic Press, Waltham, Massachusetts, USA.

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