Effects of Ospemifene on Drug Metabolism Mediated by Cytochrome P450 Enzymes in Humans in Vitro and in Vivo

Miia Turpeinen 1,*, Jouko Uusitalo 2, Terhi Lehtinen 3, Marita Kailajärvi 3, Olavi Pelkonen 1, Jouni Vuorinen 4, Pasi Tapanainen 5, Camilla Stjernschantz 6, Risto Lammintausta 6 and Mika Scheinin 3

1 Department of Pharmacology and Toxicology, University of Oulu, Oulu 90230, Finland; E-Mail: olavi.pelkonen@oulu.fi
2 Technopolis Plc, Oulu 90590, Finland; E-Mail: jouko.uusitalo@technopolis.fi
3 Clinical Research Services Turku, Institute of Biomedicine, Turku 20520, Finland; E-Mails: terhi.lehtinen@fimea.fi (T.L.); marita.kailajarvi@tyks.fi (M.K.); mika.scheinin@utu.fi (M.S.)
4 Pharma Ltd., Turku 20520, Finland; E-Mail: jouni.vuorinen@4pharma.com
5 Vitabalans Ltd., Hämeenlinna 13500, Finland; E-Mail: pasi.tapanainen@vitabalans.fi
6 Hormos Medical Ltd., Turku 20520, Finland; E-Mails: camilla.stjernschantz@fimnet.fi (C.S.); risto.lammintausta@hormos.com (R.L.)

* Author to whom correspondence should be addressed; E-Mail: miia.turpeinen@oulu.fi; Tel.: +358-50-350-5857; Fax: +358-8-537-5347.

Received: 14 June 2013; in revised form: 27 June 2013 / Accepted: 28 June 2013 / Published: 5 July 2013

Abstract: The objective of these investigations was to determine the possible effects of the novel selective estrogen receptor modulator, ospemifene, on cytochrome P450 (CYP)-mediated drug metabolism. Ospemifene underwent testing for possible effects on CYP enzyme activity in human liver microsomes and in isolated human hepatocytes. Based on the results obtained in vitro, three Phase 1 crossover pharmacokinetic studies were conducted in healthy postmenopausal women to assess the in vivo effects of ospemifene on CYP-mediated drug metabolism. Ospemifene and its main metabolites 4-hydroxyospemifene and 4'-hydroxyospemifene weakly inhibited a number of CYPs (CYP2B6, CYP2C9, CYP2C19, CYP2C8, and CYP2D6) in vitro. However, only CYP2C9 activity was inhibited by 4-hydroxyospemifene at clinically relevant concentrations. Induction of CYPs by ospemifene in cultured human hepatocytes was 2.4-fold or less. The in vivo studies showed that ospemifene did not have significant effects on the areas under
the plasma concentration-time curves of the tested CYP substrates warfarin (CYP2C9),
bupropion (CYP2B6) and omeprazole (CYP2C19), demonstrating that pretreatment with
ospemifene did not alter their metabolism. Therefore, the risk that ospemifene will affect
the pharmacokinetics of drugs that are substrates for CYP enzymes is low.

Keywords: bupropion; CYP; omeprazole; selective estrogen receptor modulator; warfarin

1. Introduction

Ospemifene, a novel selective estrogen receptor modulator (SERM), was recently approved for the
treatment of vulvar and vaginal atrophy (VVA) in postmenopausal women [1,2]. Phase 2 and 3 clinical
trials have demonstrated the efficacy and safety of ospemifene in the treatment of VVA [1,3–6].

SERMs differ in their metabolism. Tamoxifen and toremifene are metabolized by cytochrome P450
(CYP) enzymes, whereas raloxifene is metabolized by glucuronide conjugation [7]. Ospemifene is
metabolized by the liver into two major metabolites, 4- and 4'-hydroxyospemifene, which represent
roughly about 70% and 7% of the total metabolites, respectively, and several minor metabolites [8].
Several cytochrome P450 enzymes, CYP2C9, CYP2C19, CYP2B6 and CYP3A4, contribute to
the formation of 4-hydroxyospemifene, whereas CYP3A4 is the principal enzyme producing
4'-hydroxyospemifene. Thus, several CYP enzymes are responsible for the hepatic clearance of
ospemifene. Renal clearance is a minor contributor to the overall clearance of ospemifene. Its average
plasma half-life is about 20 h.

The possible effects of ospemifene on CYP-mediated metabolism were investigated as part of
its preclinical and clinical development. This report presents the results of several in vitro and
in vivo studies that evaluated the interaction potential of ospemifene with regard to CYP-mediated
drug metabolism.

2. Results and Discussion

2.1. CYP Inhibition in Vitro

In vitro, ospemifene was a relatively potent inhibitor of CYP2B6 (IC_{50}, 7.8 µM) and CYP2C9 (IC_{50},
10 µM) a less potent inhibitor of CYP2C19, CYP2C8, CYP2D6 and CYP3A4 (Table 1). Further
studies indicated (results not shown) that ospemifene inhibition was competitive for CYP2B6 and
CYP2D6 and likely competitive for CYP2C8. Ospemifene inhibition of CYP2C9 was competitive up
to 10 µM, but a noncompetitive component emerged at higher concentrations. CYP2C19
inhibition was noncompetitive. At steady state, the mean C_{max} for ospemifene was found to be
785 ng/mL, C_{min} 94 ng/mL and C_{av} 227 ng/mL corresponding roughly 2 µM, 0.25 µM and 0.6 µM,
respectively [9].
Table 1. Comparison of inhibitory potencies of ospemifene, 4-hydroxyospemifene, and 4’-hydroxyospemifene as measured using the current N-in-one assay in human liver microsomes in vitro.

| Enzyme     | Model reaction (probe concentration) | IC50 (µM)   |
|------------|--------------------------------------|-------------|
|            |                                      | Ospemifene  | 4-Hydroxy- | 4’-Hydroxy- |
| CYP1A2     | Melatonin 6-hydroxylation (4 µM)     | >100        | >100        | >100        |
| CYP2A6     | Coumarin 7-hydroxylation (2 µM)      | >100        | >100        | >100        |
| CYP2B6     | Bupropion hydroxylation (1 µM)       | 7.8         | 26.5        | 15          |
| CYP2C8     | Amodiaquine desethylation (2 µM)     | 36.4        | 27.7        | 7           |
| CYP2C9     | Tolbutamide methylhydroxylation (4 µM)| 10          | 1.1         | 12          |
| CYP2C19    | Omeprazole 5-hydroxylation (2 µM)    | 22.5        | 15.3        | 50          |
| CYP2C19    | Omeprazole demethylation (2 µM)      | 47.2        | 19.7        | 50          |
| CYP2D6     | Dextromethorphan O-demethylation (0.2 µM)| 48.7      | 25.5        | 86          |
| CYP2E1     | Chlorzoxazone 6-hydroxylation (6 µM) | >100        | >100        | >100        |
| CYP3A4     | Midazolam 1’-hydroxylation (0.4 µM)  | >100        | >100        | 78          |
| CYP3A4     | Testosterone 6β-hydroxylation (1 µM) | >100        | >100        | >100        |
| CYP3A4     | Omeprazole sulfoxidation (2 µM)      | >100        | 35.9        | 85          |
| CYP3A4     | Omeprazole 3-hydroxylation (2 µM)    | 37.9        | 50.0        | 54          |

CYP = cytochrome P450; IC50 = half maximal inhibitory concentration.

4-Hydroxyospemifene most potently inhibited tolbutamide methylhydroxylase activity, i.e., CYP2C9 activity (IC50, 1.1 µM; Table 1). Inhibition of CYP2C19 activity was approximately an order of magnitude less potent. IC50 values for all other CYP-mediated enzyme activities were >25 µM, well above the clinical Cmax of 4-hydroxyospemifene (approximately 0.3 µM for repeated oral 60-mg dosing).

4’-Hydroxyospemifene inhibited amodiaquine desethylation, i.e., CYP2C8 activity, with the highest potency (IC50, 7 µM; Table 1). CYP2C9 and CYP2B6 were also inhibited by 4’-hydroxyospemifene, although with lesser potency. IC50 values for all other CYP-mediated reactions were >50 µM. The clinical Cmax of 4’-hydroxyospemifene is approximately 0.1 µM for repeated oral 60-mg dosing.

Taken together, in human liver microsomal preparations in vitro, ospemifene showed weak inhibition potential for CYP2C9, CYP2C19, CYP2C8, and CYP2D6. Ospemifene inhibited CYP2B6 metabolism with somewhat higher potency than the other CYP enzymes, but IC50 values were above its Cmax at therapeutic doses, indicating that ospemifene is unlikely to inhibit CYP-mediated drug metabolism. The potential of 4-hydroxyospemifene and 4’-hydroxyospemifene to inhibit CYP-mediated drug metabolism also appeared to be low. 4’-Hydroxyospemifene showed some potential for CYP2C8 inhibition, but after clinical doses of ospemifene, the Cmax of 4'-hydroxyospemifene at steady state is still much lower than its IC50 values. Conversely, 4-hydroxyospemifene demonstrated relatively high potency to inhibit CYP2C9 activity.

2.2. In Vitro CYP Induction in Human Hepatocytes

In isolated human hepatocytes, incubation with ospemifene (0.2, 2, and 20 µM) for 48 h did not cause any relevant changes in CYP1A2, CYP2C9, or CYP2C19 activity. The highest concentration (20 µM) of ospemifene was associated with approximately two-fold induction of CYP2B6 and
CYP3A4 activities. Three reference CYP inducers (omepazol, phenobarbital, or rifampicin) induced the expected approximately 1.3- to 41.6-fold increases in the respective CYP enzyme activities (data not shown).

2.3. Clinical Pharmacokinetics Studies

To assess the in vivo effects of ospemifene on CYP-mediated metabolism, three Phase 1 crossover pharmacokinetic studies were conducted using recommended probe drugs for CYP2C9 (warfarin), CYP2B6 (bupropion) and CYP2C19 (omepazol) [10,11].

Figure 1. Mean concentration-time curve of plasma S-warfarin after administration of 10 mg of warfarin with and without ospemifene pretreatment. Error bars are ±SE, N = 16.

Table 2. Mean (CV%) pharmacokinetic parameters of S-warfarin, R-warfarin, bupropion, and hydroxybupropion with and without ospemifene pretreatment.

| Parameter | S-Warfarin (N = 16) | R-Warfarin (N = 16) | Bupropion (N = 16) | Hydroxybupropion (N = 16) |
|-----------|---------------------|---------------------|--------------------|--------------------------|
| AUC∞ (µg h/mL) | 18.5 (29.9) | 19.2 (30.2) | 31.3 (21.7) | 32.8 (23.1) | 0.90 (32.3) | 0.75 (38.5) | 15.5 (29.4) | 15.2 (32.4) |
| AUCt (µg h/mL) | 17.2 (27.4) | 18.0 (28.3) | 26.2 (18.7) | 27.8 (19.6) | 0.85 (33.7) | 0.70 (38.5) | 14.3 (28.1) | 14.5 (31.7) |
| Cmax (ng/mL) | 497 (14.5) | 513 (13.3) | 518 (15.2) | 538 (15.4) | 74.9 (36.6) | 62.9 (40.4) | 398 (30.6) | 462 (29.0) |
| tmax (h) † | 4 (2–6) | 4 (2–8) | 4 (2–6) | 4 (2–8) | 3.5 (1–5) | 3.0 (1–5) | 6.0 (5–24) | 6.1 (5–8) |
| t1/2 (h) | 30.4 (13.2) | 29.2 (12.7) | 43.8 (17.1) | 42.7 (15.6) | 30.8 (29.4) | 29.5 (26.7) | 26.7 (18.8) | 21.2 (18.0) |
| CL/F (mL/h) | 587 (29.2) | 564 (27.5) | 334 (21.2) | 320 (23.0) | 185 (38.1) | 230 (38.1) | n.d. | n.d. |

† Median; AUC∞ = area under the plasma concentration-time curve extrapolated to infinity; AUCt = area under the plasma concentration-time curve from zero to the last sampling time point (120 or 96 h); CL/F = oral clearance; Cmax = maximal concentration; CV% = coefficient of variation; n.d. = not determined; OSP = ospemifene; tmax = time to reach Cmax; t1/2 = half-life.
2.3.1. Study 1: The Effect of Ospemifene on Warfarin Pharmacokinetics

Subjects ($N = 16$) were Finnish white females with a mean age of 61.7 years (range, 52–71 years). Mean plasma concentrations of $S$-warfarin over time were similar with and without ospemifene pretreatment (Figure 1). The ratio of geometric least square means of $\text{AUC}_\infty$ (90% CI) between pretreatment with ospemifene versus without was 0.963 (0.909–1.020), indicating that ospemifene did not interfere with $S$-warfarin metabolism. Secondary pharmacokinetic parameters were similar between the two treatments (Table 2).

2.3.2. Study 2: The Effect of Ospemifene on Bupropion Pharmacokinetics

Subjects ($N = 16$) were Finnish white females with a mean age of 61.9 years (range, 50–70 years). Mean plasma concentrations of hydroxybupropion over time were similar with and without ospemifene pretreatment (Figure 2A). The ratio of geometric least square means (90% CI) for hydroxybupropion $\text{AUC}_\infty$ with and without ospemifene pretreatment was 0.977 (0.921–1.036), indicating that ospemifene did not interfere with hydroxybupropion metabolism. Concentrations of bupropion in plasma (Figure 2B) were much smaller than those of hydroxybupropion. The mean $\text{AUC}_\infty$ of plasma bupropion was also similar with and without ospemifene pretreatment. Secondary pharmacokinetic parameters of hydroxybupropion and bupropion further indicated absence of a pharmacokinetic interaction (Table 2).

**Figure 2.** Mean concentration-time curves of plasma (a) hydroxybupropion and (b) bupropion after administration of 150 mg of bupropion with or without ospemifene pretreatment. Error bars are ±SE, $N = 16$.

2.3.3. Study 3: The Effect of Ospemifene on Omeprazole Pharmacokinetics

Subjects ($N = 14$) were Finnish white females with a mean age of 59.6 years (range, 50–67 years). Among the 12 subjects who completed the study, treatment compliance was 100%. Mean concentration-time curves for omeprazole, 5-hydroxyomeprazole, and omeprazole sulfone were similar with and without ospemifene pretreatment (Figure 3). The ratios of geometric least squares means with and without ospemifene treatment for omeprazole/5-hydroxyomeprazole and omeprazole/omeprazole sulfone based on 3-h plasma concentrations were close to one, indicating that ospemifene did not
influence the metabolic ratios of omeprazole and its two metabolites (Table 3). However, the 90% CIs were not entirely within the pre-specified 80% to 125% acceptance range, due to high variability, and thus, bioequivalence could not be concluded. In contrast, the corresponding ratios of geometric least square means based on AUC\(_{t}\), which captures the sampling time from 0 to 8 h, were close to one and had 90% CIs that were within the pre-specified acceptance range (Table 3). These results indicated bioequivalence (absence of a pharmacokinetic interaction) with and without ospemifene pretreatment.

**Figure 3.** Mean concentration-time curves of plasma (a) omeprazole; (b) 5-hydroxyomeprazole, and (c) omeprazole sulfone after administration of 20 mg of omeprazole with and without ospemifene pretreatment. Error bars are ±SE, \(N = 12\).
Table 3. Geometric mean (CV%) and equivalence statistics of the metabolic ratios of omeprazole and its metabolites (omeprazole sulfone and 5-hydroxyomeprazole) with and without ospemifene pretreatment.

| Metabolic ratio       | Ospemifene (−) (N = 12) | Ospemifene (+) (N = 12) | Ratio * (90% CI) (N = 12) |
|-----------------------|-------------------------|-------------------------|---------------------------|
| 3 h conc.             |                         |                         |                           |
| Omeprazole/5-hydroxy | 0.67 (78.4)             | 0.65 (98.7)             | (0.77–1.22)               |
| Omeprazole/           | 1.19 (52.7)             | 1.15 (100.9)            | 0.97                      |
| Omeprazole sulfone    |                         |                         |                           |
| 5-hydroxyomeprazole   | 0.78 (59.0)             | 0.76 (61.5)             | (0.88–1.08)               |
| Omeprazole/AUCₜ       | 1.36 (20.4)             | 1.37 (24.5)             | 1.00                      |
| Omeprazole sulfone    |                         |                         |                           |
| AUCₜ                  |                         |                         |                           |

* Ratio of ospemifene (+)/ospemifene (−). AUCₜ = area under the plasma concentration-time curve from 0 to 8 h; CI = confidence interval; conc = concentration.

2.3.4. Overall Safety

All treatments in the clinical studies (warfarin, bupropion, and omeprazole with and without ospemifene) were well tolerated. The most frequently reported treatment-emergent AE was headache. In the warfarin and bupropion studies, no subjects discontinued. In the omeprazole study, one subject discontinued before receiving study medication because of personal reasons; another subject received all doses of ospemifene in the first treatment period, but withdrew because of urinary tract infection, which was considered unrelated to study treatment.

The three clinical studies in healthy postmenopausal women demonstrated minimal drug-drug interactions of ospemifene with drug substrates (warfarin, bupropion, and omeprazole) metabolized by CYPs, with no clinically relevant changes in their pharmacokinetic parameters. Despite some in vitro interactions of ospemifene and its hydroxylated metabolites with CYP2B6-, CYP2C9-, CYP2C19-, and CYP3A4-mediated drug metabolism, the clinical study results indicated no effect of ospemifene on most pharmacokinetic parameters related to CYP2C19 and CYP3A4, and supported in vitro findings whereby ospemifene only weakly inhibited CYP2C19 activity and had no effect on CYP3A4 activity. Furthermore, in spite of the potency of 4-hydroxyospemifene to inhibit CYP2C9 in human liver microsomes in vitro, the in vivo data did not provide evidence of inhibition of CYP2C9 metabolism by ospemifene or its metabolites. Finally, there was no evidence that ospemifene would affect CYP2B6-mediated drug metabolism. These in vivo results are in agreement with the in vitro data demonstrating weak inhibition potential of ospemifene or its two major metabolites on CYP2B6 activity.

The results of these three clinical studies of ospemifene pretreatment on the pharmacokinetics of warfarin, bupropion, and omeprazole stand in partial contrast to the situation with other SERMs. Coadministration of tamoxifen with warfarin has led to bleeding complications, potentially due to inhibition by tamoxifen of CYP2C9-mediated metabolism of warfarin [12]. Toremifene is a weak inhibitor of CYP2C9, and it is recommended that patients receiving concomitant treatment with warfarin and other CYP2C9 substrates with a narrow therapeutic index should be carefully monitored [13]. Raloxifene is not metabolized by CYP enzymes [14] and thus is unlikely to have CYP-mediated effects on the pharmacokinetics of other drugs, although it did decrease the prothrombin time of patients on
warfarin by 10% [15]. As with ospemifene, clinically significant effects on the pharmacokinetics of bupropion and omeprazole have not been observed with tamoxifen, toremifene, and raloxifene.

Although ospemifene did not have clinically significant effects on the pharmacokinetics of the three probe drugs tested in the clinical studies described in this article, other crossover studies (reported separately) that examined the possible effects of CYP inhibitors and inducers on the pharmacokinetics of ospemifene (data on file) demonstrated that serum concentrations of ospemifene were reduced with rifampicin pretreatment, increased with ketoconazole or fluconazole pretreatment, and minimally changed with omeprazole pretreatment. Thus, strong CYP3A or CYP2C9 inducers such as rifampicin would be expected to decrease the exposure to ospemifene; coadministration of the CYP3A inhibitor ketoconazole with ospemifene should be undertaken with caution; and the potent CYP3A/CYP2C9/CYP2C19 inhibitor fluconazole should not be coadministered with ospemifene. The potent CYP2C19 inhibitor omeprazole is unlikely to affect the pharmacokinetics of ospemifene to a clinically relevant degree.

3. Experimental Section

3.1. In Vitro Metabolism

3.1.1. Reagents

Ospemifene and its two main metabolites, 4-hydroxyospemifene and 4'-hydroxyospemifene, were supplied by Hormos Medical Ltd. (Turku, Finland). All other reagents were obtained from commercial suppliers.

3.1.2. Human Liver Microsomal Preparations

Human liver microsomes were prepared and pooled from ten human livers (HL20–24,28–32) as previously described [16]. Human liver samples were obtained from the University Hospital of Oulu as surplus tissue from cadaver kidney transplant donors. The collection of surplus tissue was approved by the Ethics Committee of the Medical Faculty of the University of Oulu, Finland. All donors were Caucasians ranging in age from 21 to 62 years; four were female and six were male. Detailed information on each donor has been published previously [16]. Protein concentrations were measured using the Bradford method [17]. Each microsome preparation was characterized with CYP-specific substrates and reference inhibitors; only those microsomal preparations whose activities were specifically inhibited were included into the pool, which was used for the preliminary screening of inhibition by the N-in-one assay.

3.1.3. CYP Inhibition Assay

In vitro inhibition of CYP isoforms was first assessed using the N-in-one approach (also called the cocktail approach) [18,19]. CYP-selective substrates and reactions are listed in Table 1; concentrations were roughly at or below the respective Michaelis-Menten coefficients. Therefore, the IC_{50} values obtained for the tested inhibitors can be considered to approximate the results that would be derived from quantitative enzyme kinetic determinations. A Micromass LCT time-of-flight high-resolution mass spectrometer (Micromass Ltd., Manchester, UK) equipped with a Z-spray electrospray ionization
source was used for the monitoring of CYP-selective reactions. Integrated mass chromatogram peak areas were normalized versus internal standards before comparing metabolites in different preparations. More details of the assay are available in previous publications [18,19].

3.1.4. Determination of Inhibition Constants

IC\textsubscript{50} values for ospemifene and 4-hydroxyospemifene were determined graphically by linear regression analysis of the logarithmic plot of inhibitor concentration versus percentage of activity remaining after inhibition using Microcal Origin, version 4.10 (Microcal Software, Inc., Northampton, MA, USA). Enzyme activities were compared with control incubations.

The type of inhibition of ospemifene toward CYP2B6-, CYP2C9-, CYP2C19-, CYP2D6- and CYP2C8-associated activities was determined by measuring four appropriate concentrations of ospemifene (selected on the basis of the screening phase) at a minimum of three different concentrations of the respective CYP-specific substrate (\(K_m/2, K_m, \) and \(4 \times K_m\)). Dixon, Lineweaver-Burk, Hanes, and Hofstee plots were used for the analysis of the type of inhibition. Different plots were in agreement with each other.

3.1.5. Assessment of CYP Induction in Isolated Human Hepatocytes

Freshly isolated human hepatocytes were obtained from the UK Human Tissue Bank (Leicester, UK). Hepatocyte viability was assessed using the trypan blue exclusion test (73%–86%). Hepatocytes were diluted in pre-incubation medium (William’s Medium E (without phenol red) supplemented with 0.1 µM dexamethasone, 0.0001% insulin, 100 IU/mL penicillin, 100 µg/mL streptomycin, and 10% v/v fetal bovine serum or newborn calf serum) and incubated in a humidified incubator for 2 to 4 h at 37 °C in an atmosphere of 95% air/5% CO\textsubscript{2}. Pre-incubation medium was replaced with incubation medium (pre-incubation medium minus serum) and cells were incubated for an additional 24 to 36 h at 37 °C.

Next, fresh incubation media containing ospemifene at concentrations of 0.2, 2, or 20 µM was added to cell cultures and incubated for a further 48 h with medium changes at 24-h intervals. Omeprazole (CYP1A2 inducer; 30 µM), phenobarbital (CYP2B6 inducer; 1 mM) and rifampicin (CYP2C9, CYP2C19, and CYP3A4 inducer; 50 µM) dissolved in dimethyl sulfoxide (0.5% v/v final), served as reference inducers. At the end of this 48-h incubation period, cells were incubated for 1 h in media containing CYP substrates in various combinations: activities of 7-ethoxyresorufin \(O\)-deethylase (CYP1A2 probe; 5 µM), bupropion hydroxylase (CYP2B6 probe; 500 µM), diclofenac 4′-hydroxylase (CYP2C9 probe; 100 µM), \(S\)-mephenytoin hydroxylase (CYP2C19 probe; 150 µM), and \[^{14}\text{C}\]testosterone 6\(β\)-hydroxylase (CYP3A4 probe; 250 µM) were determined using fluorometric, liquid chromatography/mass spectrometry (LC/MS), or radiochromatographic methods according to established protocols.

3.2. In Vivo Drug Interaction Studies

All experiments with human subjects were based on voluntary informed consent and the clinical study protocols had appropriate ethical and regulatory approval. Subjects were asked to immediately report any significant adverse events (AEs) that occurred during the study; minor AEs were reported at clinic visits. AEs were coded and classified according to the Medical Dictionary for Regulatory Activities.
Studies 1 and 2 were open-label, two-period, single-center, Phase 1, balanced, crossover studies in healthy postmenopausal women. Subjects had not taken medications known to affect CYP2C9 or CYP3A4 metabolism within four weeks of the start date (study 1) or were not homozygous for the CYP2B*6 genotype (CYP2B*6 indicates the CYP2B6 variant that is associated with reduced activity; study 2). Study 3 was a Phase 1, open-label, single-center, balanced, two-period, crossover study in healthy postmenopausal women who were not homozygous for the CYP2C19*2 genotype (CYP2C19*2 indicates the CYP2C19 variant that is associated with reduced activity). In each study, equal numbers of subjects were assigned to the two different treatment sequences.

Study 1: In one of the study periods, the subjects were pretreated with once-daily oral ospemifene (60 mg) for 12 days, and were given a single dose of 10 mg of racemic warfarin and 10 mg of vitamin K after an overnight fast on the morning of study day 8. In the other study period, 10 mg of racemic warfarin and 10 mg of vitamin K were given without ospemifene pretreatment. The study periods were separated by ≥3 weeks. In both study periods, blood samples for determination of plasma warfarin concentrations were collected before and 2, 4, 6, 8, 12, 24, 48, 72, 96, and 120 h after warfarin administration.

Study 2: In one of the study periods, the subjects were pretreated with once-daily oral ospemifene (60 mg) for 8 days, and were given a single 150 mg dose of bupropion after an overnight fast on day 8. In the other study period, 150 mg of bupropion was given without ospemifene pretreatment. The study periods were separated by ≥14 days. In both study periods, blood samples for determination of plasma bupropion and hydroxybupropion concentrations were collected before and 1, 2, 3, 4, 5, 6, 8, 12, 24, 32, 48, 56, 72, and 96 h after bupropion administration.

Study 3: In one of the study periods, the subjects were pretreated with once-daily oral ospemifene (60 mg) for 7 days, and were given a single dose of 20 mg of omeprazole after an overnight fast on day 8. In the other study period, 20 mg of omeprazole was given without ospemifene pretreatment. The study periods were separated by ≥14 days. In both study periods, blood samples for determination of plasma omeprazole, 5-hydroxyomeprazole, and omeprazole sulfone concentrations were collected before and at 1, 2, 3, 4, 6, and 8 h after omeprazole administration.

Pharmacokinetic Parameters

In Studies 1 and 2, concentrations of S-warfarin, bupropion, and hydroxybupropion were determined using validated LC-MS/MS methods. Pharmacokinetic parameters of S-warfarin, bupropion, and hydroxybupropion were calculated from the concentration-time data of both treatment periods using the WinNonlin Professional software package (Pharsight Corporation, Mountain View, CA, USA) and noncompartmental methods. With the exception of $t_{\text{max}}$, which was analyzed using the nonparametric Wilcoxon’s test, pharmacokinetic parameters were analyzed after log-transformation using analysis of variance (ANOVA), with sequence, period, and treatment as fixed effects. The primary pharmacokinetic parameter was the mean area under the plasma concentration-time curve from time 0 to infinity (AUC$_\infty$) of S-warfarin (study 1) or hydroxybupropion (study 2). Bioinequivalence was indicated by 90% CIs for the ratios of AUC$_\infty$ that were outside the range of 0.80 to 1.25.

In Study 3, concentrations of omeprazole, 5-hydroxyomeprazole, and omeprazole sulfone were determined using a validated LC-MS/MS method. Pharmacokinetic parameters were analyzed after log-transformation using ANOVA, with sequence, period, and treatment as fixed effects. Primary
pharmacokinetic parameters included the metabolic ratios of omeprazole/5-hydroxyomeprazole and omeprazole/omeprazole sulfone, based on plasma concentrations at 3 h, which reflected CYP2C19 and CYP3A4 activities, respectively. The metabolic ratios based on AUC calculated from 0 to 8 h (AUCₜ) were the key secondary parameters. Bioinequivalence was indicated by 90% CIs for the ratios of primary and secondary parameters that were outside the range of 0.80 to 1.25.

4. Conclusions

Results from these studies indicate low potential for ospemifene to alter the metabolism or pharmacokinetics of drugs that are CYP substrates. There were no clinically relevant changes in pharmacokinetic parameters for warfarin, bupropion, or omeprazole. The results obtained in the preclinical in vitro assays were judged to demonstrate a relatively weak inhibitory potency of ospemifene and indeed, this expectation was confirmed by targeted clinical studies.

Acknowledgments

Editorial support was provided by Danielle Gross, and Michael Theisen, from Complete Publication Solutions, LLC; this support was funded by Shionogi Inc., which holds exclusive rights to develop ospemifene through a licensing agreement with QuatRx Pharmaceuticals.

Conflict of Interest

These studies were funded by Hormos Medical Ltd., a subsidiary of QuatRx Pharmaceuticals. M. Kailajärvi was an employee of Clinical Research Services Turku. C. Stjernschantz and R. Lammintausta are employees of Hormos Medical Ltd. M. Scheinin and O. Pelkonen have received departmental research funding from Hormos Medical Ltd.

References

1. Bachmann, G.A.; Komi, J.O.; The Ospemifene Study Group. Ospemifene effectively treats vulvovaginal atrophy in postmenopausal women: Results from a pivotal phase 3 study. Menopause 2010, 17, 480–486.
2. McCall, J.L.; DeGregorio, M.W. Pharmacologic evaluation of ospemifene. Expert Opin. Drug Metab. Toxicol. 2010, 6, 773–779.
3. Komi, J.; Lankinen, K.S.; Härkönen, P.; DeGregorio, M.W.; Voipio, S.; Kivinen, S.; Tuimala, R.; Vihtamäki, T.; Vihko, K.; Ylikorkala, O.; et al. Effects of ospemifene and raloxifene on hormonal status, lipids, genital tract, and tolerability in postmenopausal women. Menopause 2005, 12, 202–209.
4. Rutanen, E.M.; Heikkinen, J.; Halonen, K.; Komi, J.; Lammintausta, R.; Ylikorkala, O. Effects of ospemifene, a novel SERM, on hormones, genital tract, climacteric symptoms, and quality of life in postmenopausal women: A double-blind, randomized trial. Menopause 2003, 10, 433–439.
5. Simon, J.A.; Lin, V.H.; Radovich, C.; Bachmann, G.A.; The Ospemifene Study Group. One-year long-term safety extension study of ospemifene for the treatment of vulvar and vaginal atrophy in postmenopausal women with a uterus. Menopause 2013, 20, 418–427.
6. Portman, D.J.; Bachmann, G.A.; Simon, J.A.; The Ospemifene Study Group. Ospemifene, a novel selective estrogen receptor modulator for treating dyspareunia associated with postmenopausal vulvar and vaginal atrophy. *Menopause* 2013, 20, 623–630.

7. Morello, K.C.; Wurz, G.T.; DeGregorio, M.W. Pharmacokinetics of selective estrogen receptor modulators. *Clin. Pharmacokinet.* 2003, 42, 361–372.

8. Tolonen, A.; Koskimies, P.; Turpeinen, M.; Uusitalo, J.; Lammintausta, R.; Pelkonen, O. Ospemifene metabolism in humans *in vitro* and *in vivo*: Metabolite identification, quantitation, and CYP assignment of major hydroxylations. *Drug Metabol. Drug Interact.* 2013, 14, 1–9.

9. Koskimies, P.; Turunen, J.; Lammintausta, R.; Scheinin, M. Single-dose and steady-state pharmacokinetics of ospemifene, a selective estrogen receptor modulator, in postmenopausal women. *Int. J. Clin. Pharm.* 2013, Accepted for publication.

10. Pelkonen, O.; Turpeinen, M.; Hakkola, J.; Honkakoski, P.; Hukkanen, J.; Raunio, H. Inhibition and induction of human cytochrome P450 enzymes: Current status. *Arch. Toxicol.* 2008, 82, 667–715.

11. European Medicines Agency, Committee for Human Medicinal Products (CHMP). Guideline on the investigation of drug interactions, CPMP/EWP/560/95. Available online: http://www.emea.europa.eu/docs/en_GB/document_library/Scientific_guideline/2012/07/WC500129606.pdf (accessed on 26 June 2013).

12. Givens, C.B.; Bullock, L.N.; Franks, A.S. Safety of concomitant tamoxifen and warfarin. *Ann. Pharmacother.* 2009, 43, 1867–1871.

13. Fareston (toremifene citrate) Label 2011. GTx, Inc. Available online: http://www.accessdata.fda.gov/drugsatfda_docs/label/2011/020497s006lbl.pdf (accessed on 3 July 2013).

14. Evista (raloxifene hydrochloride) Label 2007. Eli Lilly & Company. Available online: http://www.accessdata.fda.gov/drugsatfda_docs/label/2007/022042lbl.pdf (accessed on 3 July 2013).

15. Miller, J.W.; Skerjanec, A.; Knadler, M.P.; Ghosh, A.; Allerheiligen, S.R. Divergent effects of raloxifene HCI on the pharmacokinetics and pharmacodynamics of warfarin. *Pharm. Res.* 2001, 18, 1024–1028.

16. Turpeinen, M.; Ghiciuc, C.; Opritoui, M.; Tursas, L.; Pelkonen, O.; Pasanen, M. Predictive value of animal models for human cytochrome P450 (CYP)-mediated metabolism: A comparative study *in vitro*. *Xenobiotica* 2007, 37, 1367–1377.

17. Bradford, M.M. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.* 1976, 72, 248–254.

18. Turpeinen, M.; Uusitalo, J.; Jalonen, J.; Pelkonen, O. Multiple P450 substrates in a single run: Rapid and comprehensive *in vitro* interaction assay. *Eur. J. Pharm. Sci.* 2005, 24, 123–132.

19. Tolonen, A.; Petsalo, A.; Turpeinen, M.; Uusitalo, J.; Pelkonen, O. *In vitro* interaction cocktail assay for nine major cytochrome P450 enzymes with 13 probe reactions and a single LC/MSMS run: Analytical validation and testing with monoclonal anti-CYP antibodies. *J. Mass. Spectrom.* 2007, 42, 960–966.

© 2013 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (http://creativecommons.org/licenses/by/3.0/).