Introduction

The prevention and management of exacerbations are main objectives of chronic obstructive pulmonary disease (COPD) treatment. Each new exacerbation is harmful for the patient for diverse reasons: it increases in itself the risk of future exacerbations, deteriorates the quality of life, accelerates the deterioration of lung function and increases the risk of hospitalization and death. Its prevention is, therefore, a central aspect of the management of these patients. There are various pharmacological and non-pharmacological strategies aimed at both the control and prevention of COPD exacerbations. Although airway inflammation is one of the significant contributors to symptoms and exacerbations, current COPD guidelines do not consider the evaluation of the type of bronchitis or other complex pathophysiological processes involved in its genesis. That leads to generalized management strategies, which are
often suboptimal. Although "endotyping" is recommended for "individualized" care of COPD exacerbations, this is not often practiced.

We present the following three cases to illustrate the limitations of current guidelines and common clinical practice in most outpatient clinics across the world.

1. A 67-year-old male with a past smoking history of 21 years, moderate airflow obstruction (forced expiratory volume in 1 second [FEV1] of 61% predicted), and recurrent exacerbations (two in the last 12 months): He is on fluticasone/salmeterol 1,000 μg/100 μg daily and tiotropium 18 mcg daily. After his first exacerbation, his FEV1 decreased to 44% predicted and subsequently worsened to 33% predicted after the second exacerbation. Current guidelines would suggest that both exacerbations “be treated with more bronchodilators,” and perhaps with a “short burst of prednisone” and a "broad-spectrum antibiotic"4, and perhaps adding long-term macrolide or a phosphodiesterase 4 inhibitor4,5.

2. A 57-year-old male, current smoker with a history of 15 pack-years: He reports productive cough, and in increase in wheeze and exertional dyspnea. His FEV1/forced vital capacity (FVC) is 2.8 L/4.4 L (ratio of 63%) and improves to 2.9 L/4.2 L post bronchodilator, which is consistent with mild to moderate airflow obstruction (FEV1, of 78% predicted). Chest X-ray is normal. His current treatment includes salbutamol as needed, which he uses about 2 to 4 times a day. Current guidelines would suggest that he be commenced on a combination of a long-acting beta-2 agonist (with or without a long-acting anti-cholinergic inhaler)4.

3. An 81-year-old male, with a 34 years history of smoking: His previous medical history includes glaucoma, benign prostate hyperplasia, diabetes and coronary artery disease. He presents with exertional breathlessness and cough and has had two exacerbations within the last year. His pre-bronchodilator FEV1/FVC is 0.9 L/4.4 L and postbronchodilator is 1.0 L/4.5 L, which are 29% and 90% predicted, respectively. Total lung capacity is 122%, residual volume is 160%, and KCO is 30% predicted. Arterial blood gases show a PCO2 of 58 mm Hg, PO2 of 64 mm Hg and pH of 7.38. Right ventricular systolic pressure is 40 mm Hg. Computed tomography of the thorax reveals heterogenous centrilobular emphysema. Current treatment is budesonide/formoterol (200 μg/6 μg) 2 puffs twice daily, terbutaline as needed, furosemide and ramipril. Current guidelines would suggest adding a long-acting anti-cholinergic inhaler or alternatively switching to a single combination inhaler.

Current COPD Guidelines on Treatment and Prevention of Acute Exacerbations

Current recommendations are largely focused on decreasing exacerbations and improving symptoms by optimizing the use of bronchodilators. It is known that both long-acting beta agonists (LABA) and long-acting anti-cholinergics (LAAC) can reduce the rate of exacerbations in patients with COPD. Furthermore, the current literature supports that combined therapy (LABA/LAAC) is superior to monotherapy4 and to LABA/inhaled corticosteroid (ICS)7 combination, although its effect does not reach the sum of both8. More recently, attempts have been made to demonstrate that triple inhalation therapy is even more effective than the LABA/LAAC combination; but the results have been variable. A large study of more than 10,000 patients, which compared triple therapy against two types of dual therapy (LAAC/LABA and LABA/ICS), showed lower rates of moderate and severe exacerbations in the first group9. A similar effect is observed when comparing single-inhaler triple combination with single-inhaler dual bronchodilator combination, with a 15% reduction in the risk of moderate to severe exacerbations10. The latter was not reflected in a subsequent trial performed in a "real-world clinical setting" where there were no differences between the groups treated with triple therapy and LAAC/LABA. However, when analyzing by subgroups, it was observed that among patients with blood eosinophilia >6% and in those with two or more previous exacerbations, there was a statistically significant benefit (hazard ratio [HR] of 0.66 and 0.83, respectively)11. Therefore, based on the current evidence, there is an inclination to add ICS to dual bronchodilator therapy in patients with moderate to very-severe stable COPD with a high risk of exacerbations, especially in those with serum eosinophils higher than 300 cells/μL12. A summary of the current principal studies which compare dual and triple inhalers' effect on FEV1, and exacerbation rates is shown in Table 1.

Despite the availability of these pharmacologic interventions, about 30% of patients with COPD have frequent exacerbations1, which entails high healthcare costs. As part of the numerous efforts aimed at trying to reduce the rate of hospitalizations in these patients, comprehensive care management programs (CCMP) have been implemented, including strategies such as patient education, implementation of action plans, and serial telephone evaluations by a trained team. Unfortunately, the application of these strategies has not been consistently shown a reduction in the risk of hospitalization, but rather, in some cases they have increased it. This was shown in a trial performed in 426 COPD patients who were assigned to a CCMP versus conventional care. The study was terminated before enrolment was completed, given higher risks of hospitalization and mortality in the intervention group (HR of 1.13 and 3.0, respectively)12. Furthermore, there was no improvement in quality of life outcomes with these interventions13. These results contrast with other studies in which these strategies were proven effective to reduce COPD-related admissions, although the studied populations were significantly different (e.g., patients with other comorbidities were excluded)14. Another interesting explanation for this phenom-
Sputum, COPD, and Exacerbation Phenotype

Classically, an acute exacerbation of COPD (AECOPD) is defined as respiratory symptoms that worsen beyond the normal day-to-day variability requiring additional therapy. However, this definition of an AECOPD may be overlapping with many concurrent pathologies such as worsening left or right heart function, respiratory or metabolic acidosis, with or without bronchitis. The pillars of AECOPD management according to most guidelines include bronchodilation, systemic corticosteroids, and antibiotics, without in depth characterization of the bronchitic component. The non-specific manner of AECOPD treatment with the above strategy does not adequately target the key pathology involved while subjecting the patient to additional side effects. A potential resolution to overcome this challenge is to assess the sputum characteristics at baseline and at each exacerbation for COPD patients and to guide treatment based on these objective measures.

Sputum cytology has already been well established, particularly in asthma. In brief, it involves sputum being induced from the lower respiratory tract using increasing concentrations of nebulized saline. Total cell count (TCC) and viability is assessed following processing, and a cytospin slide is made for differential cell count. This protocol has been validated with well-established normal limits described in literature (Table 2). Apprehension regarding the bronchoconstrictive properties of nebulized saline is unfounded as several studies have demonstrated the safety of sputum induction in both stable and exacerbating COPD. If the FEV₁ is too low for hypertonic saline induction, sputum can be obtained with a modified protocol using normal saline. Spontaneously expectorated sputum is comparable to induced sputum in terms of cell differential and can be used for clinical purposes. When assessed during an exacerbation, the sputum cytology can allow a more discriminatory approach in therapy.

Using sputum analysis, it is possible to characterize the patient’s airway inflammation at baseline and at each exacerbation, as there is good evidence that they may be discordant. A comparison of airway characteristics of COPD patients during stability and exacerbation were studied in a retrospective cross-sectional survey and showed that neutrophilic bronchitis is much more prominent during exacerbations compared to baseline. In a similar study that included a subpopulation of COPD patients who had successive sputum analysis during convalescence and exacerbations showed that there was poor correlation between the baseline and exacerbation airway cytology. Furthermore, 85.2% of patients had subsequent exacerbations that differed in bronchitic subtype from previous exacerbations that differed in bronchitic subtype from previous exacerbations.

### Table 1. Principal studies comparing dual to triple therapy effect on reducing the risk of AECOPD

| Study                        | Clinical scenario                                  | RR moderate or severe AECOPD | Δ in FEV₁ (mL) |
|------------------------------|---------------------------------------------------|------------------------------|---------------|
| IND/GLY vs. SAL/FP           | Severe to very severe COPD with high risk of AECOPD | 0.78 (95% CI, 0.7 to 0.86; p<0.001) | +63           |
| TIO/placebo vs. TIO/SAL/FP   | Moderate to severe COPD with high risk of AECOPD   | 0.98 (95% CI, 0.872 to 1.088; p=0.71) | N/A           |
| FF/UMEC/VI vs. FF/VI         | Moderate to severe COPD with high risk of AECOPD   | 0.85 (95% CI, 0.8 to 0.9; p<0.001) | +97           |
| FF/UMEC/VI vs. UMEC/VI       | Moderate to severe COPD with high risk of AECOPD   | 0.75 (95% CI, 0.7 to 0.81; p<0.001) | +54           |
| BDP/FOR/GLY vs. IND/GLY     | Severe to very severe COPD with high risk of AECOPD | 0.848 95% (95% CI, 0.723 to 0.995; p=0.043) | +20           |
| TIO/SAL vs. FP/SAL/TIO      | Severe to very severe COPD with high risk of AECOPD | 1.06 (95% CI, 0.94 to 1.19) | −43           |
| IND/GLY vs. FP/SAL/TIO      | Moderate to severe COPD without high risk of AECOPD | 1.08 (95% CI, 0.83 to 1.40) | −26           |

AECOPD: acute exacerbations of COPD; RR: rate ratio; FEV₁: forced expiratory volume in one second; IND: indacaterol; GLY: glycopyrronium; SAL: salmeterol; FP: fluticasone propionate; COPD: chronic obstructive pulmonary disease; CI: confidence interval; N/A: non-available; TIO: tiotropium; FF: fluticasone furoate; UMEC: umeclidinium; VI: vilanterol; BDP: beclometasone dipropionate; FOR: formoterol fumarate.

### Table 2. Normal values for total and differential cell counts in healthy adults

|                    | Mean     | Median | 2SD* | 90th percentile |
|--------------------|----------|--------|------|-----------------|
| Total cell count   | 4.1      | 2.4    | 13.8 | 9.7             |
| Eosinophils (%)    | 0.4      | 0.0    | 2.2  | 1.1             |
| Neutrophils (%)    | 37.5     | 36.7   | 77.7 | 64.4            |
| Macrophages        | 58.8     | 60.8   | 100  | 86.1            |

Data source: Belda et al.®. *Two standard deviations.
baseline or even a previous exacerbation.\textsuperscript{30} Taken together, this suggests that exacerbations are not simply a worsening of the underlying inflammation and emphasizes the importance of sputum analysis at every exacerbation to tease out these variations that can lead to change in therapeutic strategy for each episode. The approach of combined antibiotics and corticosteroids is only appropriate in 2.5\textendash{}8\% of exacerbations as defined by the presence of mixed granulocytic bronchitis. Hence, by characterizing the luminal inflammation using sputum examination, the most appropriate and effective therapeutic strategy can be put in place and reduce the likelihood of adverse drug events and economic burden for the patient and the health care system, respectively.\textsuperscript{31,32}

**Eosinophils in COPD: Actor or Spectator?**

A proportion of COPD patients have evidence of eosinophilic bronchitis either during exacerbation or at baseline. Sputum eosinophilia is found in 10\%\textendash{}40\% of patients with COPD\textsuperscript{33} and has been associated with severity of the disease, exacerbation frequency and degree of emphysema on quantitative imaging.\textsuperscript{14,35} AECOPD with viral infections, in particular, demonstrate increased eosinophilic activity as evidenced by a significant increase in the presence of soluble eosinophil cationic protein in sputum.\textsuperscript{34} The presence of eosinophils in sputum also predicts response to corticosteroids, both systemic and inhaled.\textsuperscript{46} In randomized control trials, controlling luminal eosinophils using corticosteroids have been shown to reduce severe AECOPD.\textsuperscript{35}

Although it is more impractical to obtain, sputum eosinophil counts cannot be replaced by blood eosinophils as a biomarker. In a study investigating the blood eosinophil count that correlates with exacerbations, a threshold of 300 cells/μL or more was found to predict an increased risk of AECOPD.\textsuperscript{36} However, when directly compared with sputum, circulatory eosinophil counts correlates poorly with luminal eosinophil counts.\textsuperscript{31,40} The role of circulatory eosinophils in airway disease remains questionable as well. When analyzing the effect of blocking eosinophil recruitment from circulation into target tissues, paradoxical elevation of blood eosinophil count was seen in clinical trials using anti–interleukin 13 (IL-13) monoclonal antibodies (MABs) for the treatment of asthma.\textsuperscript{41} Despite this elevation, patients in the treatment arm demonstrated improved lung function. One can argue that this lack of correlation or direct pathological role is arbitrary and, if blood eosinophil counts can predict response to therapy, then it remains a valid biomarker. However, this stance may be flawed when looking closer at anti–interleukin 5 (IL-5) drug trials in COPD and comparing them to those done in asthma.

The use of anti-eosinophil therapy targeting the IL-5 pathway in asthma have shown overall improvement in asthma-related quality of life, reduced corticosteroid use and improvement in lung function. Importantly, it demonstrated dose related reduction in exacerbation frequency\textsuperscript{42-45}. The same cannot be said of COPD trials using the same molecules. A study that combined the results of two phase 3 clinical trials where COPD patients with an eosinophilic phenotype were given low and high dose mepolizumab (100 mg in METREX, 100 mg, and 300 mg in METRO). This study concluded that 100 mg mepolizumab is effective at reducing exacerbation frequency among this population. However, it appears that while the lowest dose of mepolizumab appeared to be effective at reducing exacerbation frequency (p=0.04), a three-fold higher dose paradoxically did not (p=0.14)\textsuperscript{46} which opposes the data seen in asthma.\textsuperscript{47} Moreover, the exclusion of subjects with asthma in this study was based on self-report and may have contaminated the sample. In this study, eosinophil phenotype was determined by blood eosinophil levels of >150 cells/μL at time of screening or >300 cells/μL during the previous year. These thresholds may not be specific to eosinophilic patients as studies on normal leukocyte differentials done in the 1970s and 1980s among healthy volunteers has established a 95\% normal range for blood eosinophils of 0 to 700 cells/μL with a median of 150 cells/μL.\textsuperscript{47,48} To resolve these methodological issues, the diagnosis of asthma need to be rigorously excluded and eosinophilia be identified at the tissue level by sputum analysis. When these amendments in patient selection are applied, mepolizumab only depletes sputum and blood eosinophils in COPD without an effect on exacerbation rate.\textsuperscript{49} Similarly, benralizumab, an anti-IL-5 receptor MAB, also failed to demonstrate reduction in exacerbation regardless if selecting for patients based on elevated circulatory\textsuperscript{39} or sputum\textsuperscript{40} eosinophilia. Thus, these negative studies have led to the speculation that eosinophils, whether circulatory or luminal, predict steroid responsiveness and disease severity but is otherwise not directly involved in COPD. Perhaps eosinophilia is only a marker that rises and falls in concordance with an another, unknown process that is key in COPD pathobiology are not intrinsically contributory themselves.

More recent studies regarding the role of eosinophils in COPD have revealed evidence that challenges the spectator theory. In murine models, lung matrix metalloproteinase 12 (MMP-12), a key mediator in emphysematous alveolar destruction produced by macrophages, was found to be elevated.\textsuperscript{55} The levels of MMP-12 correlated with the presence of activated eosinophils and their subsequent IL-13 release. In these experiments, the eosinophils were preferentially activated in vitro by IL-33, and not IL-5. This finding suggests that, not only do eosinophils contribute to the pathophysiology of COPD, the luminal eosinophils in COPD have different biology compared to those found in asthma and have entirely different downstream effects. Thus, this emphasizes the importance in controlling eosinophilic inflammation in COPD not only during exacerbations but also while stable, as ongoing eosinophilia may continue to propagate tissue damage even if...
noxious inhalants have been removed.

While these studies regarding the role of eosinophils represent important breakthroughs in biology, the true impact of the research is still yet to be seen in clinical decision making and, in translation, to patient care. When following currently available guidelines, the use of ICS is not recommended as part of first line therapy. The current guideline recommendations regarding ICS addition are based on blood eosinophil counts, as opposed to sputum. As mentioned above, there is an inclination to add ICS in patients with at least moderate COPD but this may be putting some at increased risk of infections. At the same time, as blood and sputum eosinophils do not correlate well, another proportion of patients who would benefit from ICS may be overlooked when following these principles. Among those without severe disease or who are not considered at “high risk” based on spirometry and previous frequency of exacerbation, and not type of exacerbation, the initiation of ICS is only recommended after a minimum of 12 months of persistent symptoms and reduced health status despite LABA and LAAC dual therapy. However, up to 18% of COPD patients have eosinophilic bronchitis at baseline and may not necessarily fall under the “high risk” profile. Given the new evidence of the role of eosinophils in COPD, rigorous adherence of guidelines without sputum analysis may result in both over- and under-treatment of a large number of patients.

**Beyond Cells: Other Uses for Sputum**

The utility of sputum is not limited to just cytology but can also be used to assess for other comorbid conditions that may contribute to a patient's symptoms. The detection of lipids in sputum macrophages can be used as a non-invasive marker for gastroesophageal reflux. Using a reproducible method of indexing the degree of lipid staining using oil red O in macrophages, an index of seven or more correlated with the gold standard 24-hour ambulatory esophageal pH recording, and was highly specific and sensitive.

Hemosiderin-laden macrophages (HLM) can also be detected on induced sputum as a maker of pulmonary capillary leakage from left ventricular dysfunction. This has been previously well described on bronchoalveolar lavage (BAL) fluid but acquisition of BAL fluid is an inefficient way of establishing left ventricular dysfunction. In a prospective cross-sectional pilot study, 46 dyspneic patients and nine healthy controls underwent echocardiography and sputum induction within 72 hours of each other. The presence of HLM proportion directly correlated with decrease cardiac function and was found to be a sensitive and specific maker for left ventricular dysfunction. This was further validated among patients who were presenting with acute respiratory symptoms as well where HLM were significantly elevated in those whose dyspnea was of cardiac as opposed to pulmonary origin. Persistent elevations of sputum HLM may also increase the likelihood of infectious AECOPD, which is further explored in the next section.

Apart from using sputum for immediate patient management decisions, other non-cellular components can be quantified to better understand the biology of airway diseases. The fluid phase of sputum has several utilities including the measurement of soluble cellular by-products. Such measurements include downstream secretion products of various leukocytes, as a surrogate marker of activity, as well as various cytokines, chemokines, and adhesion molecules that are key in supporting local inflammation. More recently, autoantibodies have been detected in sputum, which may provide further insight on the contribution of local immunity to underlying pulmonary conditions. While systemic immunoglobulin deficiency is a contributor to infectious exacerbations, as discussed below, local immunoglobulin levels also appear to impact exacerbation frequency and lung function in COPD.

Bacterial migration across the epithelium in small airways is poorly inhibited where there is local deficiency of secretory IgA. This promotes further inflammation and remodeling, even with smoking cessation. Thus far, these assessments have been limited to research use but, may one day be employed clinically to further fine tune management strategies and personalized medicine.

**What Causes Infective Exacerbations?**

The most common causes of COPD exacerbations are lower respiratory tract infections, although other factors such as environmental pollution and exposure to fine particulate matter may also contribute. Whilst viruses are the most frequent causative microbe, bacterial colonization also plays a significant role in infective exacerbations. Why some individuals with COPD are more susceptible than others to develop infective exacerbations is still an unresolved question. Though some explanations for this propensity are already well established, such as structural alterations and immune dysregulations secondary to smoking, there are still novel mechanism which remain poorly understood. Several biomarkers have been linked to the predisposition to respiratory infections in these patients, including quantification of multiple proteins, specific cells, metabolites in exhaled air, images and even genetic predictors. Recurrent respiratory tract infection may also occur because of local humoral deficiency. We recently measured a complete immunoglobulin profile in asthmatics with recurrent respiratory tract infection and found that there was a relative deficiency of sputum IgA compared to healthy controls.

It has been observed that the development of these infections could be correlated with changes in the composition of the pulmonary microbiome through increased inflammation of the airways. A study that followed 87 subjects and
analyzed their pulmonary microbiota with molecular biology techniques showed that both the diversity and the proportion of different families of bacteria varied within the same individual throughout periods of stability versus exacerbation. Interestingly, in acute respiratory episodes, individuals tended to carry more abundantly Proteobacteria (*Moraxella catarrhalis* and *Haemophilus influenzae*) than Firmicutes (*Streptococcus pneumoniae*). This relationship was reversed during periods of stability. Likewise, it was observed that the treatment with oral corticosteroids was related to lower diversity and a change in the microbiome composition of the investigated subjects. These alterations were maintained even six weeks after the episodes were treated.

Among the genetic factors involved with an increased risk of respiratory infections, those related to iron metabolism stand out. As an essential nutrient, iron plays a central role in bacterial survival and multiplication. Furthermore, in states of chronic inflammation, hepcidin-mediated intracellular iron overload could potentially cause immune cell dysfunction. Locally, the iron overload at the respiratory system can be measured by staining hemosiderin in sputum macrophages, which can undoubtedly be an interesting biomarker that could predict higher infectious risk in these patients. A retrospective study showed a correlation between a higher percentage of HLM (hemosiderin index) and a higher frequency of infectious exacerbations in the previous two years. Interestingly, it was also shown that higher local levels of interleukin 6, the cytokine responsible for promoting hepcidin release, correlated with a higher hemosiderin index. Excess iron in pulmonary macrophages also appears to be a predisposing factor for infective exacerbation when studied prospectively, with those with a higher sputum hemosiderin index having a significantly shorter time to infective exacerbation (unpublished data).

**Management of AECOPD: St. Joseph’s Strategy and Resolving Our Cases**

Our local approach to AECOPD could be summarized (Figure 1) as follows.

First, we try to identify the cause of the symptoms, whether it is worsening of airflow limitation, bronchitis, respiratory or metabolic acidosis, left or right ventricular dysfunction, or a combination of these. A general approach to this can be easily made through simple tests available in the emergency department, including spirometry, chest X-ray, and arterial blood gases. Treatment is initiated according to guidelines, with bronchodilators, systemic steroids and antibiotics if they are indicated. Secondly, as soon as feasible to obtain, a sputum sample is collected, and treatment is adjusted accordingly. If sputum shows eosinophilic bronchitis, steroids are continued.
or increased. Neutrophilic bronchitis (neutrophils ≥64.4% with a TCC greater than 25×10⁶/g cells) suggests an underlying bacterial infection and is thus treated with antibiotics. Notably, even when mixed granulocytic bronchitis (≥2% eosinophils and ≥65% neutrophils) has been associated with poorer clinical outcomes⁶⁶, its physiopathology has not been completely understood and thus is managed with both antibiotics and steroids⁵⁶. Normal cell counts would prompt discontinuation of both systemic steroids and antibiotics. If we observe a high hemosiderin index, left ventricle function will be further assessed. Additionally, aspiration would be suspected if there is a high lipid index, and speech and language pathologist evaluation would be requested.

Within 2 weeks of discharge, patients are seen in a specialized post-discharge COPD clinic, where we utilize a similar clinical phenotyping approach in combination with a comprehensive care management program. Patients are seen again at 8- and 16-weeks post-discharge with each visit accompanied by clinical assessment, including complete blood count (for peripheral eosinophilia), blood gases (for persistent hypercapnic respiratory failure), spirometry, and spontaneous sputum cytometry and culture. These measurements allow for the individualized prescription of inhalers, to optimize airflow and suppress persistent inflammation. CCMP is delivered by a trained nurse and includes case management, regular telephone follow-up, inhaler technique assessment and education during clinic visits, and referral to appropriate services (e.g., smoking cessation clinic). Patients can contact the clinic nurse if they develop worsening symptoms or require clarification regarding their treatment. Combining these two seemingly disparate interventions accounts for the medical (e.g., uncontrolled bronchitis) and non-medical (e.g., poor adherence) contributors to uncontrolled disease. Retrospective study of this combined clinic intervention has demonstrated a significant reduction in healthcare utilization, with sputum cytometry influencing treatment decisions in many cases (unpublished data). A clinical trial of this intervention is planned to rigorously test its efficacy and cost-effectiveness.

1. How would this be applied to the proposed cases?

At his first exacerbation, his sputum showed 23.4×10⁶/g total cells, with 0.2% eosinophils and 88% neutrophils. He was prescribed an antibiotic course and short-acting beta-agonist in addition to continuing his maintenance inhalers. Symptoms resolved in six days. At his second exacerbation, sputum TCC was 7.3×10⁶/g with 22% eosinophils and 46% neutrophils, for which he received a course of prednisone. This example outlines how two exacerbations could be related to different underlying mechanisms and therefore, should be treated differently.

According to current guidelines, this patient would be categorized as mild COPD and thus should receive either LABA or LAAC. However, his sputum test showed a TCC of 6.2×10⁶/g, with 57% neutrophils and 11% eosinophils. Though this patient would benefit from ICS, his assigned disease severity would at most prompt the addition of another long-acting bronchodilator, thus leaving his eosinophilic bronchitis untreated. It is relevant to note that unpublished data from our clinical experiences has shown that after bronchitis treatment, whether neutrophilic or eosinophilic, a percentage of previous "COPD" patients will no longer meet the spirometric diagnostic criteria, suggesting that unresolved bronchitis may lead to overdiagnosis.

This patient’s investigations showed a sputum TCC of 64.8×10⁶/g cells with 82% neutrophils, and 9% eosinophils. His sputum hemosiderin index was 13%. His bloodwork revealed a chloride of 84 mEq/L, and potassium of 3.1 mEq/L. In this case, given severe COPD with a high risk of exacerbations and uncontrolled eosinophilic bronchitis, chronic treatment should be adjusted to higher doses of ICS, and low prednisone maintenance dose and long-term macrolides should be considered. Ventricle offload treatment should be implemented if cardiac involvement is demonstrated by clinical exam and echocardiogram, and electrocardiographic abnormalities may be corrected, including spironolactone, chloride, and potassium replacement. Acetazolamide, which is often prescribed, should only be added if the patient has enough respiratory muscle reserve to manage higher respiratory rates. Finally, BiPAP support could be incorporated if it is appropriate to do so.

2. Key points

- COPD exacerbations are common even in patients treated according to current guidelines. These events are associated with worsening in lung function, worse symptom control, higher mortality rates and an elevated economic burden.
- Current treatments aimed to reduce AECOPD rates include both pharmacologic and non-pharmacologic approaches. Some strategies, like CCMP, have failed to improve health indicators in these patients, and actually, have been associated with higher hospitalization and mortality risk.
- Sputum assessment, while stable and during exacerbation, is essential for phenotyping and for guidance of therapeutic strategies. It is a safe and rapid method of evaluation and will likely broaden beyond cytology and provide insights regarding other causes of respiratory symptoms.
- The main cause of COPD exacerbations are acute respiratory infections. There are several alterations which underlie higher susceptibility to them, including structural changes, immune impairment, and novel mechanisms that are still poorly understood. Some of them, like iron overload, have been investigated and could be considerate as target checkpoints for future treatment strategies.
- Our local approach to AECOPD include the assessment of the bronchitic component and is focused on airway inflammetry. This personalized strategy has been proven effec-
tive to decrease exacerbations rates.

Authors’ Contributions

Conceptualization: Nair P. Writing - original draft preparation: Venegas C, Zhao N. Writing - review and editing: Ho T, Nair P.

Conflicts of Interest

CV, NZ, and TH do not have any conflicts of interest to disclose. PN has received honoraria and grant supports from Teva, Sanofi, AZ, Novartis, Roche, BI, Merck, Equillium, and Methapharm.

Acknowledgments

Professor Nair is supported by the Frederick E Hargreave Teva Innovation chair in Airway Diseases.

References

1. Hurst JR, Vestbo J, Anzueto A, Locantore N, Mullerova H, Tal-Singer R, et al. Susceptibility to exacerbation in chronic obstructive pulmonary disease. N Engl J Med 2010;363:1128-38.
2. Soler-Cataluna JJ, Martinez-Garcia MA, Roman Sanchez P, Salcedo E, Navarro M, Ochando R. Severe acute exacerbations and mortality in patients with chronic obstructive pulmonary disease. Thorax 2005;60:925-31.
3. Woodruff PG, Agusti A, Roche N, Singh D, Martinez FJ. Current concepts in targeting chronic obstructive pulmonary disease pharmacotherapy: making progress towards personalised management. Lancet 2015;385:1789-98.
4. Singh D, Agusti A, Anzueto A, Barnes PJ, Bourbeau J, Celli BR, et al. Global strategy for the diagnosis, management, and prevention of chronic obstructive lung disease: the GOLD science committee report 2019. Eur Respir J 2019;53:1900164.
5. Bourbeau J, Bhutani M, Hernandez P, Aaron SD, Balter M, Beauchesne ME, et al. Canadian Thoracic Society clinical practice guideline on pharmacotherapy in patients with COPD: 2019 update of evidence. Can J Respir Crit Care Sleep Med 2019;3:210-32.
6. Wedzicha JA, Decramer M, Ficker JH, Niewoehner DE, Sandstrom T, Taylor AF, et al. Analysis of chronic obstructive pulmonary disease exacerbations with the dual bronchodilator QVA149 compared with glycopyrrolate and tiotropium (SPARK): a randomised, double-blind, parallel-group study. Lancet Respir Med 2013;1:199-209.
7. Wedzicha JA, Banerji D, Chapman KR, Vestbo J, Roche N, Ayers RT, et al. Indacaterol-glycopyrronium versus salmeterol-fluticasone for COPD. N Engl J Med 2016;374:2222-34.
8. Farne HA, Cates CJ. Long-acting beta2-agonist in addition to tiotropium versus either tiotropium or long-acting beta2-agonist alone for chronic obstructive pulmonary disease. Cochrane Database Syst Rev 2015;(10):CD008989.
9. Lipson DA, Barnhart E, Brealey N, Brooks J, Criner GJ, Day NC, et al. Once-daily single-inhaler triple versus dual therapy in patients with COPD. N Engl J Med 2018;378:1671-80.
10. Papi A, Vestbo J, Fabbri L, Corradi M, Prunier H, Cohuet G, et al. Extrafine inhaled triple therapy versus dual bronchodilator therapy in chronic obstructive pulmonary disease (TRIBUTE): a double-blind, parallel group, randomised controlled trial. Lancet 2018;391:1076-84.
11. Sissa S, Dell’Aniello S, Ernst P. Comparative effects of LAMA-LABA-ICS vs LAMA-LABA for COPD: cohort study in real-world clinical practice. Chest 2020;157:846-55.
12. Fan VS, Gaziano JM, Lew R, Bourbeau J, Adams SG, Leatherman S, et al. A comprehensive care management program to prevent chronic obstructive pulmonary disease hospitalizations: a randomized, controlled trial. Ann Intern Med 2012;156:673-83.
13. Aboumatar H, Naqibuddin M, Chung S, Chaudhry H, Kim SW, Saunders J, et al. Effect of a hospital-initiated program combining transitional care and long-term self-management support on outcomes of patients hospitalized with chronic obstructive pulmonary disease: a randomized clinical trial. JAMA 2019;322:1371-80.
14. Zwerink M, Brusse-Keizer M, van der Valk PD, Zielhuis GA, Munninkhof EM, van der Palen J, et al. Self management for patients with chronic obstructive pulmonary disease. Cochrane Database Syst Rev 2014;(3):CD002990.
15. Ho T, Dasgupta A, Hargreave FE, Nair P. The use of cellular and molecular biomarkers to manage COPD exacerbations. Expert Rev Respir Med 2017;11:403-11.
16. Pizzichini E, Pizzichini MM, Efthimiadis A, Evans S, Morris MM, Squillace D, et al. Indices of airway inflammation in induced sputum: reproducibility and validity of cell and fluid-phase measurements. Am J Respir Crit Care Med 1996:154(2 Pt 1):308-17.
17. Pizzichini MM, Popov TA, Efthimiadis A, Hussack P, Evans S, Pizzichini E, et al. Spontaneous and induced sputum to measure indices of airway inflammation in asthma. Am J Respir Crit Care Med 1996;154(4 Pt 1):866-9.
18. Iredale MJ, Wanklyn SA, Phillips IP, Krausz T, Ind PW. Non-invasive assessment of bronchial inflammation in asthma: no correlation between eosinophilia of induced sputum and bronchial responsiveness to inhaled hypertonic saline. Clin Exp Allergy 1994;24:940-5.
19. Pavord ID, Pizzichini MM, Pizzichini E, Hargreave FE. The use of induced sputum to investigate airway inflammation. Thorax 1997;52:498-501.
20. Belda J, Leigh R, Parameswaran K, O’Byrne PM, Sears MR,
Hargreave FE. Induced sputum cell counts in healthy adults. Am J Respir Crit Care Med 2000;161(2 Pt 1):475-8.

21. Wilson AM, Leigh R, Hargreave FE, Pizzichini MM, Pizzichini E. Safety of sputum induction in moderate-to-severe smoking-related chronic obstructive pulmonary disease. COPD 2006;3:89-93.

22. Gao P, Gibson PG, Zhang J, He X, Hao Y, Li P, et al. The safety of sputum induction in adults with acute exacerbation of COPD. Clin Respir J 2013;7:101-9.

23. Vlachos-Mayer H, Leigh R, Sharon RF, Hussack P, Hargreave FE. Success and safety of sputum induction in the clinical setting. Eur Respir J 2000;16:997-1000.

24. Bhownik A, Seemungal TA, Sapsford RJ, Devalia JL, Wedzicha JA. Comparison of spontaneous and induced sputum for investigation of airway inflammation in chronic obstructive pulmonary disease. Thorax 1998;53:953-6.

25. Siva R, Green RH, Brightling CE, Shelley M, Hargadon B, McKenna S, et al. Eosinophilic airway inflammation and exacerbations of COPD: a randomised controlled trial. Eur Respir J 2007;29:506-13.

26. D’Silva L, Hassan N, Wang HY, Kjarsgaard M, Efthimiadis A, Hargreave FE, et al. Heterogeneity of bronchitis in airway diseases in tertiary care clinical practice. Can Respir J 2011;18:144-8.

27. Aaron SD, Vandemheen KL, Fergusson D, Maltais F, Bourbeau J, Goldstein R, et al. Tiotropium in combination with placebo, salmeterol, or fluticasone-salmeterol for treatment of chronic obstructive pulmonary disease: a randomized trial. Ann Intern Med 2007;146:545-55.

28. Magnussen H, Disse B, Rodriguez-Roisin R, Kirsten A, Watz H, Tetzlaff K, et al. Withdrawal of inhaled glucocorticoids and exacerbations of COPD. N Engl J Med 2014;371:285-94.

29. Chapman KR, Hurst JR, Frew SM, Larbig M, Fogel R, Guerin T, et al. Long-term triple therapy de-escalation to indacaterol/glycopyrronium in patients with chronic obstructive pulmonary disease (SUNSET): a randomized, double-blind, triple-dummy clinical trial. Am J Respir Crit Care Med 2018;198:329-39.

30. Wang H, Dasgupta A, Lee KA, Cook RJ, Nair P. Changing pattern of sputum cell counts during successive exacerbations of chronic obstructive pulmonary disease. COPD 2015;12:628-35.

31. D’Silva L, Gafni A, Thahane L, Jayaram L, Hassack P, Hargreave FE, et al. Cost analysis of monitoring asthma treatment using sputum cell counts. Can Respir J 2008;15:370-4.

32. Mittmann N, Kuramoto L, Seung SJ, Haddon JM, Bradley-Kennedy C, Fitzgerald JM. The cost of moderate and severe COPD exacerbations to the Canadian healthcare system. Respir Med 2008;102:113-21.

33. George L, Brightling CE. Eosinophilic airway inflammation: role in asthma and chronic obstructive pulmonary disease. Ther Adv Chronic Dis 2016;7:34-51.

34. Hastie AT, Martinez FJ, Curtis JL, Doerschuk CM, Hansel NN, Christenson S, et al. Association of sputum and blood eosinophil concentrations with clinical measures of COPD severity: an analysis of the SPIROMICS cohort. Lancet Respir Med 2017;5:956-67.

35. Tashkin DP, Miravitlles M, Celli BR, Metzdorf N, Mueller A, Halpin DM, et al. Concomitant inhaled corticosteroid use and the risk of pneumonia in COPD: a matched-subgroup post hoc analysis of the UPLIFT(R) trial. Respir Res 2018;19:196.

36. Papi A, Bellettato CM, Bracci G, Romagnoli M, Casolari P, Caramori G, et al. Infections and airway inflammation in chronic obstructive pulmonary disease severe exacerbations. Am J Respir Crit Care Med 2006;173:1114-21.

37. Pizzichini E, Pizzichini MM, Gibson P, Parameswaran K, Gleich GJ, Berman L, et al. Sputum eosinophilia predicts benefit from prednisone in smokers with chronic obstructive bronchitis. Am J Respir Crit Care Med 1998;158(5 Pt 1):1511-7.

38. Leigh R, Pizzichini MM, Morris MM, Maltais F, Hargreave FE, Pizzichini E. Stable COPD: predicting benefit from high-dose inhaled corticosteroid treatment. Eur Respir J 2006;27:964-71.

39. Yun JH, Lamb A, Chase R, Singh D, Parker MM, Saferali A, et al. Blood eosinophil count thresholds and exacerbations in patients with chronic obstructive pulmonary disease. J Allergy Clin Immunol 2018;141:2037-47.

40. Turato G, Semenzato U, Bazzan E, Biondini D, Tine M, Torrecilla N, et al. Blood eosinophilia neither reflects tissue eosinophils nor worsens clinical outcomes in chronic obstructive pulmonary disease. Am J Respir Crit Care Med 2018;197:1216-9.

41. Corren J, Lemanske RF, Hanania NA, Korenblat PE, Parsey MV, Arron JR, et al. Lebrikizumab treatment in adults with asthma. N Engl J Med 2011;365:1088-98.

42. Hamblly N, Nair P. Monoclonal antibodies for the treatment of refractory asthma. Curr Opin Pulm Med 2014;20:87-94.

43. Padvard ID, Korn S, Howarth P, Bleecker ER, Buli R, Keene ON, et al. Mepolizumab for severe eosinophilic asthma (DREAM): a multicentre, double-blind, placebo-controlled trial. Lancet 2012;380:651-9.

44. Bleecker ER, FitzGerald JM, Chanez P, Papi A, Weinstein SF, Barker F, et al. Efficacy and safety of benralizumab for patients with severe asthma uncontrolled with high-dosage inhaled corticosteroids and long-acting beta2-agonists (SIROCCO): a randomised, multicentre, placebo-controlled phase 3 trial. Lancet 2016;388:2115-27.

45. Mukherjee M, Sehni R, Nair P. Anti-IL5 therapy for asthma and beyond. World Allergy Organ J 2014;7:32.

46. Padvard ID, Chanez P, Criner GJ, Kersijens HAM, Korn S, Lugo-go N, et al. Mepolizumab for eosinophilic chronic obstructive pulmonary disease. N Engl J Med 2017;377:1613-29.

47. Orfanakis NG, Ostlund RE, Bishop CR, Athens JW. Normal blood leukocyte concentration values. Am J Clin Pathol 1970;53:647-51.

48. Juul S, Pliskin JS, Fineberg HV. Variation and information in white blood cell differential counts. Med Decis Making 1984;4:69-80.

49. Dasgupta A, Kjarsgaard M, Capaldi D, Radford K, Aleman F, Boylan C, et al. A pilot randomised clinical trial of mepo-
lizumab in COPD with eosinophilic bronchitis. Eur Respir J 2017;49:1602-46.

50. Criner GJ, Celli BR, Brightling CE, Agusti A, Papi A, Singh D, et al. Benralizumab for the prevention of COPD Exacerbations. N Engl J Med 2019;381:1023-34.

51. Brightling CE, Bleeker ER, Panettieri RA Jr, Bafadhel M, She D, Ward CK, et al. Benralizumab for chronic obstructive pulmonary disease and sputum eosinophilia: a randomised, double-blind, placebo-controlled, phase 2a study. Lancet Respir Med 2014;2:891-901.

52. Doyle AD, Mukherjee M, LeSuer WE, Bittner TB, Pasha SM, Frere JJ, et al. Eosinophil-derived IL-13 promotes emphysema. Eur Respir J 2019;53:1801291.

53. Calverley PM, Anderson JA, Celli B, Ferguson GT, Jenkins C, Jones PW, et al. Sputum autoantibodies in patients with severe eosinophilic asthma. J Allergy Clin Immunol 2018;141:1269-79.

54. Parameswaran K, Anvari M, Efthimiadis A, Kamada D, Hargrave FE, Allen CJ. Lipid-laden macrophages in induced sputum are a marker of oropharyngeal reflux and possible gastric aspiration. Eur Respir J 2000;16:1119-22.

55. Grebski E, Hess T, Hold G, Speich R, Russi E. Diagnostic value of hemosiderin-containing macrophages in bronchoalveolar lavage. Chest 1992;102:1794-9.

56. Danel C, Israel-Biet D, Costabel U, Rossi GA, Wallaert B. The clinical role of BAL in pulmonary haemorrhages. Eur Respir J 1990;3:951-2, 961-9.

57. Leigh R, Sharon RF, Efthimiadis A, Hargreave FE, Kitching AD. Diagnosis of left-ventricular dysfunction from induced sputum. Lancet 1999;354:833-4.

58. Bellido-Casado J, Belda J, Bayes-Genis A, Margarit G, Lopez L, Casan P, et al. Hemosiderin-laden macrophages count in sputum examination. Lancet 1999;354:833-4.

59. Kurashima K, Mukaida N, Fujimura M, Schroder JM, Matsuda T, Matsushima K. Increase of chemokine levels in sputum during exacerbation of acute asthma attacks. J Leukoc Biol 1996;59:313-6.

60. Louis R, Shutte J, Biagi S, Stanciu L, Marrelli F, Tenor H, et al. Cell infiltration, ICAM-1 expression, and eosinophil chemotactic activity in asthmatic sputum. Am J Respir Crit Care Med 1997;155:466-72.

61. Mukherjee M, Thomas SR, Radford K, Dvorkin-Gheva A, Davydchenko S, Kjarsgaard M, et al. Sputum antineutrophil cytoplasmatic antibodies in serum antineutrophil cytoplasmatic antibody-negative eosinophilic granulomatosis with polyangiitis. Am J Respir Crit Care Med 2019;199:158-70.

62. Mukherjee M, Bulir DC, Radford K, Kjarsgaard M, Huang CM, Jacobsen EA, et al. Sputum autoantibodies in patients with severe eosinophilic asthma. J Allergy Clin Immunol 2018;141:1269-79.

63. Polosukhin VV, Richmond BW, Du RH, Cates JM, Wu P, Nian H, et al. Secretory IgA deficiency in individual small airways is associated with persistent inflammation and remodeling. Am J Respir Crit Care Med 2017;195:1010-21.

64. Woodhead M, Blasi F, Ewig S, Garau J, Huchon G, Ieven M, et al. Guidelines for the management of adult lower respiratory tract infections: full version. Clin Microbiol Infect 2011;17 Suppl 6:E1-59.

65. Liu S, Zhou Y, Liu S, Chen X, Zou W, Zhao D, et al. Association between exposure to ambient particulate matter and chronic obstructive pulmonary disease: results from a cross-sectional study in China. Thorax 2017;72:788-95.

66. Sethi S, Murphy TF. Bacterial infection in chronic obstructive pulmonary disease in 2000: a state-of-the-art review. Clin Microbiol Rev 2001;14:336-63.

67. Leitao Filho FS, Ra SW, Mattman A, Schellenberg RS, Criner GJ, Woodruff PG, et al. Serum IgG subclass levels and risk of exacerbations and hospitalizations in patients with COPD. Respir Res 2018;19:30.

68. Ishii T, Kida K. Predictors of chronic obstructive pulmonary disease exacerbations. Curr Opin Pulm Med 2014;20:138-45.

69. Ho T, Al-Selahi E, Mukherjee M, Huang C, Radford K, Kjarsgaard M, et al. Clinical role of BAL in pulmonary haemorrhages. Eur Respir J 2014;52:2813-23.

70. Wang Z, Bafadhel M, Haldar K, Spivak A, Mayhew D, Miller BE, et al. Lung microbiome dynamics in COPD exacerbations. Am J Respir Crit Care Med 2016;194:1082-92.

71. Wang Z, Bafadhel M, Haldar K, Spivak A, Mayhew D, Miller BE, et al. Lung microbiome dynamics in COPD exacerbations. Am J Respir Crit Care Med 2016;194:1082-92.

72. Verrills NM, Irwin JA, He XY, Wood LG, Powell H, Simpson JL, et al. Identification of novel diagnostic biomarkers for asthma and chronic obstructive pulmonary disease. Am J Respir Crit Care Med 2011;183:1633-43.

73. Zhou H, Kobzik L. Effect of concentrated ambient particles on macrophage phagocytosis and killing of Streptococcus pneumoniae. Am J Respir Crit Care Med 2007;176:460-5.

74. Winter WE, Bazydlo LA, Harris NS. The molecular biology of human iron metabolism. Lab Med 2014;45:92-102.

75. Mohan S, Ho T, Kjarsgaard M, Radford K, Borhan AS, Thabane L, et al. Hemosiderin in sputum macrophages may predict infective exacerbations of chronic obstructive pulmonary disease: a retrospective observational study. BMC Pulm Med 2017;17:60.

76. Gao P, Zhang J, He X, Hao Y, Wang K, Gibson PG. Sputum inflammatory cell-based classification of patients with acute exacerbation of chronic obstructive pulmonary disease. PLoS One 2013;8:e57678.