Intensity-Modulated Radiotherapy (IMRT) vs Helical Tomotherapy (HT) in Concurrent Chemoradiotherapy (CRT) for Patients with Anal Canal Carcinoma (ACC): an analysis of dose distribution and toxicities

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Abstract

Purpose: Intensity-modulated radiotherapy (IMRT) and helical tomotherapy (HT) have been adopted for radiotherapy treatment of anal canal carcinoma (ACC) due to better conformality, dose homogeneity and normal-tissue sparing compared to 3D-CRT. To date, only one published study compares dosimetric parameters of IMRT vs HT in ACC, but there are no published data comparing toxicities. Our objectives were to compare dosimetry and toxicities between these modalities.

Methods and materials: This is a retrospective study of 35 ACC patients treated with radical chemoradiotherapy at two tertiary cancer institutions from 2008–2010. The use of IMRT vs HT was primarily based on center availability. The majority of patients received fluorouracil (5-FU) and 1–2 cycles of mitomycin C (MMC); 2 received 5-FU and cisplatin. Primary tumor and elective nodes were prescribed to ≥54 Gy and ≥45 Gy, respectively. Patients were grouped into two cohorts: IMRT vs HT. The primary endpoint was a dosimetric comparison between the cohorts; the secondary endpoint was comparison of toxicities.

Results: 18 patients were treated with IMRT and 17 with HT. Most IMRT patients received 5-FU and 1 MMC cycle, while most HT patients received 2 MMC cycles (p < 0.01), based on center policy. HT achieved more homogenous coverage of the primary tumor (HT homogeneity and uniformity index 0.14 and 1.02 vs 0.29 and 1.06 for IMRT, p = 0.01 and p < 0.01). Elective nodal coverage did not differ. IMRT achieved better bladder, femoral head and peritoneal space sparing (V30 and V40, p ≤ 0.01), and lower mean skin dose (p < 0.01). HT delivered lower bone marrow (V10, p < 0.01) and external genitalia dose (V20 and V30, p < 0.01). Grade 2+ hematological and non-hematological toxicities were similar. Febrile neutropenia and unscheduled treatment breaks did not differ (both p = 0.13), nor did 3-year overall and disease-free survival (p = 0.13, p = 0.68).

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Conclusions: Chemoradiotherapy treatment of ACC using IMRT vs HT results in differences in dose homogeneity and normal-tissue sparing, but no significant differences in toxicities.

Keywords: Anal cancer, Tomotherapy, Intensity modulated radiotherapy, Dosimetry, Toxicities

Introduction
Since the 1980s, standard management of anal canal carcinoma (ACC) has been definitive chemoradiation therapy (CRT), with salvage abdomino-perineal resection (APR) for those who fail CRT [1-3]. Although CRT has been demonstrated in several randomized controlled trials as effective treatment for ACC, it is associated with significant toxicities [4-6]. Treatment breaks in up to 40-50% of patients have been reported due to hematological, dermatological and gastrointestinal toxicities [7,8]. Efforts have been made to reduce toxicities through newer chemotherapy regimens and radiotherapy (RT) techniques [9-11]. Development of more conformal RT techniques has reduced normal tissue toxicity and the need for unintended treatment breaks [11].

RTOG 0529 demonstrated significantly lower hematological, gastrointestinal and dermatological toxicities with intensity-modulated radiotherapy (IMRT) compared to conventional 2D-planning in ACC treatment [12]. Helical tomotherapy (HT) delivery, a newer RT technique, has shown improved target conformity, dose homogeneity and normal-tissue sparing compared to IMRT in other tumor sites [12,13].

In Alberta, use of IMRT vs HT is dependent on center availability and preference. To date, only one published study compares dosimetric parameters of IMRT vs HT in ACC, but there are no published data comparing toxicities [14]. Our objectives were to compare dosimetry and toxicities between these modalities.

Materials and methods
Patient population
This retrospective study included ACC patients treated with definitive CRT between 2008–2010 at two provincial tertiary cancer centers (Tom Baker Cancer Center (TBCC) and Cross Cancer Institute (CCI)). Approval for this study was obtained from the University of Calgary Conjoint Health Research Ethics board.

Patients were included if they were ≥18 years of age, had a histologic diagnosis of ACC, no other active malignancies, and were treated with curative-intent CRT with a primary planning target volume (PTVprimary) dose of 54–55.4 Gy. All patients were treated with IMRT or HT, and chemotherapy consisting of 2 cycles of 5-FU and 1–2 cycles of mitomycin C (MMC) or cisplatin. Patients who had metastatic disease, received PTVprimary dose <54 or >55.4 Gy, RT alone, and RT treatment techniques other than IMRT or HT were excluded.

Pre-treatment evaluation of all patients included tumor biopsy, clinical examination, baseline complete blood count (CBC), and computed tomography (CT) abdomen and pelvis. Tumor size was based on clinical exam (when documented) or imaging. Weekly CBC and toxicities (skin, gastrointestinal, genitourinary) while on treatment were graded using the RTOG acute scoring index [15]. All blood counts were retrieved from the provincial clinical database (Alberta Netcare) during CRT and up to four weeks post last chemotherapy cycle. Hematological nadirs were recorded and analyzed.

Volume definitions
Treatment plans were evaluated on an Eclipse workstation (Eclipse™ v8.9, Varian Medical Systems, Palo Alto, CA) without change to original planning target volumes delineated by treating physicians. Gross tumor volume (GTV), clinical target volume (CTV), planning target volume (PTV) were contoured, following the RTOG 0529 protocol, with deviations, if necessary, based on clinical judgment [11]. CTVprimary included the primary tumor and involved lymph nodes >1 cm identified on CT imaging and/or endoscopic ultrasound, plus a margin. CTVnodes included regional lymph nodes at risk including perirectal, internal iliac, external iliac, obturator, presacral and inguinal lymph nodes. PTVprimary and PTVnode were generated with a uniform 1 cm margin around the CTVprimary and CTVnode, respectively.

Organs at risk (OARs) included the bladder, peritoneal cavity, femoral heads, external genitalia (vulva in women, penis and scrotum in men), skin and bone marrow. These volumes were contoured on the existing plans. OAR volume definitions were based on the RTOG 0529 protocol except for small bowel and iliac crests [11]. The peritoneal cavity included large and small bowel from the L4/5 junction to the level of the bladder dome, with exclusion of named structures. In lieu of iliac crests, bone marrow was delineated, consisting of the L5 vertebra, sacrum, and bilateral iliac crests. This was felt to better reflect bone marrow dose from SPECT bone marrow imaging and IMRT bone marrow sparing studies [16,17]. Skin was generated as a 5 mm thick layer of tissue within the body contour, excluding PTV.

Plan evaluation
The homogeneity of each plan with respect to PTVprimary and PTVnode was evaluated by the homogeneity (HI) and...
### Table 1 Baseline characteristics of ACC patients treated with chemoradiation by treatment cohort

| Characteristic                          | IMRT (N = 18) n (%) | HT (N = 17) n (%) | P-value* |
|----------------------------------------|---------------------|-------------------|----------|
| Treatment center (TBCC/CCI)            | 18 (100%)/0 (0%)    | 0 (0%)/17 (100%)  | <0.001   |
| Age, y (median(range))                 | 61 (45.1, 85.1)     | 52 (34.8, 69.7)   | 0.0045   |
| Gender, (male/female)                  | 6 (33.3%)/12 (66.7%)| 4 (23.5%)/13 (76.5%)| 0.71     |
| Smoker                                 | 4 (22.2%)           | 11 (64.7%)        | 0.02     |
| ECOG status                            |                     |                   |          |
| ECOG 0                                 | 4 (22.2%)           | 7 (41.2%)         | 0.29     |
| ECOG ≥1                                | 14 (77.8%)          | 10 (58.8%)        |          |
| Histology                              |                     |                   |          |
| Squamous                               | 17 (94.4%)          | 17 (100%)         | 1.00     |
| Other                                  | 1 (5.6%)            | 0 (0%)            |          |
| AJCC T-stage                           |                     |                   |          |
| 1                                      | 1 (5.6%)            | 0 (0%)            | 1.00     |
| 2                                      | 8 (44.4%)           | 7 (41.2%)         |          |
| 3                                      | 7 (38.9%)           | 8 (47.1%)         |          |
| 4                                      | 2 (11.1%)           | 2 (11.8%)         |          |
| AJCC N-stage                           |                     |                   |          |
| N0                                     | 14 (77.8%)          | 11 (64.7%)        | 1.00     |
| N1-3                                   | 4 (23.2%)           | 6 (36.3%)         |          |
| Pretreatment Blood Counts              |                     |                   |          |
| (median(range))                        |                     |                   |          |
| Hb (g/dL)                              | 124.5 (101, 156)    | 141 (101, 163)    | 0.10     |
| WBC (x10^9/L)                          | 7.8 (5.1, 14.3)     | 8.6 (5.2, 15.5)   | 0.70     |
| Neutrophil (x10^9/L)                   | 5.7 (2.8, 10.3)     | 6.3 (2.7, 11.3)   | 0.60     |
| Platelet (x10^9/L)                     | 287.5 (146, 525)    | 287 (167, 368)    | 0.96     |
| RT dose to Primary Tumor, Gy           | 54 (54, 55.4)       | 54 (54, 54)       | 0.04     |
| (median(range))                        |                     |                   |          |
| Chemotherapy                           |                     |                   |          |
| Cisplatin + 5FU                        | 2 (11.1%)           | 0 (0%)            | <0.001   |
| MMC 1 cycle + 5FU                      | 16 (88.9%)          | 1 (5.9%)          |          |
| MMC 2 cycles + 5FU                     | 0 (0%)              | 16 (94.1%)        |          |

*Fisher exact testing used where any cell n < 5, otherwise Chi-square testing used.

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**Figure 1** Dose distribution for ACC treated with IMRT vs HT.
### Table 2 Dosimetric coverage of treatment volumes and OARs by treatment cohort

| Parameter                  | IMRT Median (range), Mean | HT Median (range), Mean | P-value* |
|----------------------------|---------------------------|-------------------------|----------|
| **PTV primary**            |                           |                         |          |
| HI                         | 0.28 (0.10, 1.09), 0.51   | 0.14 (0.05, 0.36), 0.18 | 0.015    |
| UI                         | 1.06 (1.04, 1.52), 1.09   | 1.03 (1.01, 1.07), 1.03 | <0.001   |
| **PTV node**               |                           |                         |          |
| HI                         | 0.42 (0.14, 1.25), 0.48   | 0.55 (0.33, 0.79), 0.54 | 0.06     |
| UI                         | 1.12 (1.04, 1.52), 1.16   | 1.17 (1.09, 1.21), 1.16 | 0.15     |
| **Bladder**                |                           |                         |          |
| V30                        | 94 (51, 100), 88.8        | 100 (94, 100), 99       | 0.009    |
| V40                        | 56.5 (13, 92), 55.5       | 96 (44, 100), 90.9      | <0.001   |
| V50                        | 14 (0.9, 54), 17.5        | 28 (0.2, 35), 21        | 0.37     |
| Mean (Gy)                  | 40.9 (32.4, 49.5), 41.3   | 47 (38.3, 49.2), 46.5   | 0.002    |
| Median (Gy)                | 41.6 (30.3, 51.1), 41.5   | 47.1 (37.1, 49.0), 46.5 | 0.002    |
| **Bone Marrow**            |                           |                         |          |
| V10                        | 82.5 (47, 98), 78.6       | 67 (63, 75), 67.4       | 0.007    |
| V20                        | 62.5 (22, 88), 60.9       | 56 (51, 60), 56         | 0.08     |
| Mean (Gy)                  | 27.6 (12.4, 37.4), 27.2   | 26.2 (23.1, 28.5), 26.3 | 0.39     |
| Median (Gy)                | 29.4 (9.1, 44.3), 28.9    | 28.3 (21.1, 33.4), 28.3 | 0.81     |
| **Femoral heads**          |                           |                         |          |
| V30                        | 68.5 (15, 99), 63.7       | 91 (76, 100), 90.3      | <0.001   |
| V40                        | 14.5 (0, 50), 15.3        | 47 (27, 67), 50         | <0.001   |
| V50                        | 0 (0, 14), 1.2            | 0 (0, 26), 3.1          | 0.94     |
| Mean (Gy)                  | 32 (23.5, 41.9), 31.5     | 38.9 (35.4, 44.1), 39.1 | <0.001   |
| Median (Gy)                | 32.7 (23.6, 40.2), 31.6   | 39.3 (35.2, 43.2), 39.8 | <0.001   |
| **Peritoneal Cavity**      |                           |                         |          |
| V30                        | 26.5 (0.1, 66), 30.7      | 46 (6, 76), 44.6        | 0.01     |
| V40                        | 11 (0.9, 53), 17.2        | 34 (2, 66), 32.5        | 0.003    |
| V45                        | 4.5 (0, 44), 9.2          | 22 (0.5, 57), 23.4      | 0.002    |
| V50                        | 0.9 (0, 17), 1.7          | 2 (0, 27), 4.1          | 0.01     |
| Mean (Gy)                  | 20.2 (4.7, 36.4), 20.4    | 26.3 (9, 40.6), 25.6    | 0.04     |
| Median (Gy)                | 19.7 (2.3, 41.8), 19.3    | 26 (4.1, 45.7), 25.4    | 0.09     |
| **External Genitalia**     |                           |                         |          |
| V20                        | 98.5 (62, 100), 91        | 73 (48, 98), 73.7       | <0.001   |
| V30                        | 64 (19, 99), 65           | 25 (6, 58), 27.1        | <0.001   |
| V40                        | 27 (0, 86), 25.7          | 5 (0, 48), 9.8          | 0.02     |
| V50                        | 2 (0, 44), 7.2            | 0.9 (0, 39), 4.4        | 0.11     |
| Mean (Gy)                  | 33.6 (21.8, 47.6), 33.6   | 25.1 (18, 38.9), 26     | <0.001   |
| Median (Gy)                | 33.3 (23.9, 48.9), 33.6   | 23.4 (18.9, 35.2), 24   | <0.001   |
| **Skin**                   |                           |                         |          |
| Mean (Gy)                  | 17 (1.4, 21.3), 16        | 23.1 (11.2, 26.8), 22   | <0.001   |
| Median (Gg)                | 15.9 (11.9, 19.2), 15.9   | 20.4 (6, 23.4), 19.1    | <0.001   |

*Mann–Whitney test reported.
uniformity index (UI). HI was defined as the difference between $D_{\text{max}}$ and $D_{\text{min}}$ divided by the prescription dose, while UI was $D_{5}/D_{95}$. A value approaching 0 for HI and unity for UI indicates optimal dose homogeneity [18,19]. For OARs, median and mean dose were reported. In addition, V30, V40, and V50 were recorded for the femoral heads, bladder and peritoneal cavity. For external genitalia, V20, V30, and V40 were recorded while V10 and V20 were recorded for bone marrow.

**Statistical analysis**

Patients were classified into two treatment cohorts: IMRT vs HT. A dosimetric comparison of plan homogeneity and OARs was performed. Disease-free survival (DFS), overall survival (OS), colostomy-free survival (CFS) and acute hematological and non-hematological toxicities were analyzed.

DFS was the interval between diagnosis and evidence of local, regional or metastatic failure, second primary, death or last follow-up for patients who did not fail. Local failure was evidence of persistent local disease or local recurrence. Regional failure was persistence, appearance or recurrence of regional nodal disease. Patients with persistent disease were considered as failing on the day of their first follow-up post CRT or date of biopsy-proven persistent disease (when available). Failure for OS was death due to any cause and was measured from diagnosis to the date of death or last follow-up. CFS was the interval between diagnosis and date of colostomy, including diverting colostomy and colostomy from salvage APR, or last follow-up for those not requiring a colostomy.

**Results**

**Demographics**

72 patients were diagnosed with ACC in Alberta between 2008–2010. Thirty-seven patients were excluded from the analysis (13 treated with a technique other than IMRT or HT, 7 received RT alone, 1 was treated with surgery alone, 1 had metastatic disease at diagnosis, 13 received a dose PTV$_{\text{primary}}$ <54 Gy or >55.4 Gy, and 2 with missing data). Of the remaining 35 patients, 18 patients were treated with IMRT (all treated at TBCC) and 17 with HT (all treated at CCI). Patients treated with HT were treated on TomoTherapy® Hi-Art® system, version 2.2.4.1 (Accuray, Inc, Sunnyvale, CA). Patients treated with IMRT were treated on Clinac 21EX, Clinac IX, or Triology (Varian Medical, Palo Alto, CA). Patient, tumor and treatment characteristics are summarized in Table 1.

Both groups were balanced in regards to performance status, histology, T stage, N stage, and pre-treatment hematological parameters. The IMRT group had slightly older patients ($p = 0.0045$) and fewer smokers ($p = 0.02$). The median RT dose was the same between the groups, but dose was more variable in the IMRT group. Chemotherapy regimen was significantly different, with 16 patients

### Table 3 Grade 2+ toxicities in ACC patients treated with chemoradiation by treatment cohort

| Toxicity                              | Total (N = 35) | IMRT (N = 18) | HT (N = 17) | P-value* |
|---------------------------------------|---------------|---------------|-------------|----------|
|                                       | n (%)         | n (%)         | n (%)       |          |
| Grade 2+ Hematologic Toxicities†      |               |               |             |          |
| Leukopenia                            | 27 (77.1)     | 13 (72.2)     | 14 (82.4)   | 0.69     |
| Neutropenia                           | 22 (62.9)     | 10 (55.6)     | 12 (70.6)   | 0.36     |
| Thrombocytopenia                      | 13 (37.1)     | 6 (33.3)      | 7 (41.2)    | 0.63     |
| Anemia                                | 11 (31.4)     | 5 (27.8)      | 6 (35.3)    | 0.63     |
| Febrile neutropenia requiring hospitalization | 5 (14.3) | 1 (5.6) | 4 (23.5) | 0.18     |
| Grade 2+ Non-Hematologic Toxicities†  |               |               |             |          |
| Skin                                  | 29 (82.9)     | 14 (77.8)     | 15 (88.2)   | 0.66     |
| Upper GI                              | 9 (25.7)      | 6 (33.3)      | 3 (17.7)    | 0.44     |
| Lower GI                              | 18 (51.4)     | 12 (66.7)     | 6 (35.3)    | 0.06     |
| GU                                    | 3 (8.6)       | 2 (11.1)      | 1 (5.9)     | 1.00     |
| Unscheduled treatment break           | 8 (22.9)      | 6 (33.3)      | 2 (11.8)    | 0.23     |

*Fisher exact testing used where any cell n < 5, otherwise Chi-square testing used.
†RTOG acute radiation morbidity scoring criteria [17].
in the IMRT group receiving 1 MMC cycle with 5-FU and 16 patients in the HT group receiving 2 MMC cycles with 5-FU (p < 0.001).

Dosimetric outcomes
Figure 1 shows a typical dose distribution on axial imaging at the level of PTV\textsubscript{primary} for IMRT and HT techniques. Table 2 details the dosimetric coverage of treatment volumes and OARs by cohort. The HT group achieved more homogenous PTV\textsubscript{primary} coverage compared to IMRT (HI p = 0.015, UI p < 0.001). PTV\textsubscript{nodes} coverage did not differ between the techniques, although HI approached significance (p = 0.06). IMRT achieved better bladder, peritoneal space and femoral head sparing (V30 and V40, p ≤ 0.01). Median and mean skin dose was also lower with IMRT (p < 0.001). HT delivered lower dose to bone marrow (V10, p < 0.01) and external genitalia (V20 and V30, p < 0.001). A dosimetry model was constructed containing the factors of HI and UI for PTV\textsubscript{primary} and HI for PTV\textsubscript{nodes}. No factor was significant for either outcome on multivariate analysis.

Toxicities
Acute hematological and non-hematological toxicities are presented in Table 3. The most common toxicities were leukopenia and skin reaction. Grade 2+ hematological and non-hematological toxicities were similar between the groups. There were no significant differences in lower or upper gastrointestinal, genitourinary and skin toxicities. Additionally, febrile neutropenia and unscheduled treatment breaks did not differ between the two groups.

Overall survival, disease-free survival and colostomy-free survival
Median follow-up for survivors at the time of analysis was 23.7 months. Estimated 3-year OS (IMRT 87.1% vs HT 100%, p = 0.13, Figure 2A) and DFS (IMRT 32.9% vs HT 51.8%, p = 0.68, Figure 2B) by Kaplan Meier analysis were similar between the two groups. Estimated 3-year CFS was also similar (IMRT 57.3% vs HT 70.2%, p = 0.29, Figure 2C).

Multivariate analysis was not performed on OS, due to the small number of events. Multivariate analysis for DFS and CFS outcomes was limited to 3 factors per model, accounting for the number of events. For each outcome, a demographics model consisting of age, smoking status, and treatment group was constructed. No factor was significant for either outcome.

Discussion
Advances in CRT for ACC have focused on toxicity reduction by alterations in chemotherapy regimens and radiotherapy delivery [9-11]. Large retrospective studies have shown promising results with cisplatin-based regimens compared to standard MMC treatment [20,21]. However, two recent phase III trials, RTOG 98–11 and ACTII, failed to demonstrate superiority of cisplatin to MMC for ACC treatment [9,10]. As such, ACC treatment with MMC and 5-FU remains standard of care.

Development of more conformal RT techniques has resulted in reduced normal-tissue toxicity. RTOG 0529 established superiority of IMRT to conventional 2D-planning in ACC treatment, with significant reduction in
grade 2+ hematological, and grade 3+ gastrointestinal and dermatological toxicities using IMRT [11]. Newer RT techniques, such as HT, have shown improved target conformity, dose homogeneity and normal-tissue sparing compared to IMRT [12,13]. To date, there is only one study by Joseph et al. comparing dosimetry between HT and IMRT in ACC treatment [14]. Our current retrospective study serves as the first study comparing toxicities in ACC treatment between these two techniques.

Seventy-two patients were diagnosed with ACC between our study period, but less than half of the patients were included in the current analysis. The majority of exclusions were based on RT technique and PTV primary dose outside of 54–55.4 Gy, in an effort to maintain consistency in dose and treatment volumes. Chemotherapy was significantly different, with the majority of IMRT patients receiving 1 MMC cycle, versus 2 cycles in the HT group. Historically, patients treated at the center where HT is available have been treated with 2 MMC cycles due to physician preference, versus 1 MMC cycle at the center where only IMRT is available.

Similar to the dosimetric study by Joseph et al., HT in our study achieved superior target conformity compared to IMRT for PTV primary coverage [14]. OAR dose constraints in our study reflect those in RTOG 0529, with the exception of peritoneal cavity and bone marrow [11]. In both the current study and the previous study by Joseph et al., IMRT achieved better sparing of bilateral femoral heads, while HT delivered lower dose to external genitalia. However, in our study, IMRT also achieved lower bladder, peritoneal cavity and skin doses, while Joseph et al. reported improvement in sparing of bladder and peritoneal cavity with HT. In addition, our study demonstrated better bone marrow and external genitalia sparing with HT [14].

Differences in dose distribution and sparing of OARs between the two studies may be secondary to differences in calculation algorithms and study design. Although we found a significant difference between skin dose for IMRT and HT, it is known that there are large uncertainties in calculated skin dose for both the Eclipse analytic anisotropic algorithm and HT dose calculation algorithms [22,23]. Furthermore, the planning study by Joseph et al. generated HT and IMRT plans on the same patients, thereby minimizing anatomical variations between the two groups. There was also strict adherence to contouring and planning guidelines [14]. Our study, on the other hand, is retrospective with a larger sample of patients, and reflects real practice in a more generalized population.

Despite significant differences in OAR doses and chemotherapy regimens, acute hematological and non-hematological toxicities were similar between the groups. Three-year OS, DFS and CFS Kaplan Meier estimates were also similar. A recent retrospective study by our group suggests reduction of grade 3+ hematological and skin toxicities with 1 MMC cycle versus 2 cycles in ACC treatment, without compromise to OS, CFS and DFS [24]. Potential differences in acute toxicities secondary to different chemotherapy regimens may have been mitigated by radiotherapy technique. Further studies with more balanced groups in regards to chemotherapy may assist in confirming this hypothesis and our current results.

This study has inherent limitations of a retrospective study from a single health authority. Despite utilizing a standardized toxicity grading scale, potential subjectivity exists in retrospectively grading toxicities. In effort to obtain consistency, a single reviewer graded all toxicities.

Conclusions
In our analysis of IMRT vs HT in ACC, differences in dose homogeneity and normal-tissue sparing were observed, but there were no significant differences in toxicities. Further investigation to increase cohort patient numbers should be performed, to definitively determine the impact of these techniques on toxicities and outcomes.

Consent
Waiver of consent was granted by the local research ethics board, thus individual patient consents were not obtained. Patient identifiers were not used.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
RY contributed to the design of the study, acquisition of data, interpretation of data, and drafted the manuscript. YM contributed to the analysis and interpretation of data, and helped draft the manuscript. HW contributed to the acquisition and interpretation of data, and assisted with drafting the manuscript. DG and BW contributed to the interpretation of data. JX contributed to the conception and design of this study and interpretation of data. CD contributed to the conception and design of this study, interpretation of data and assisted with drafting the manuscript. All authors read and approved the final manuscript.

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References
1. Nigro ND, Vaitkevicius VK, Buroker T, Bradley GT, Considine B. Combined therapy for cancer of the anal canal. Dis Colon Rectum. 1981;24:73–5.
2. Nigro ND, Seydel HG, Considine B, Vatkevicius VK, Leichman L, Kinzie JJ. Combined preoperative radiation and chemotherapy for squamous cell carcinoma of the anal canal. Cancer. 1983;51:1826–9.

3. Leichman L, Nigro N, Vatkevicius VK, Considine B, Buroker T, Bradley G, et al. Cancer of the anal canal. Model for preoperative adjuvant combined modality therapy. Am J Med. 1985;78:211–5.

4. Northover J, Gynne-Jones R, Sebag-Montefiore D, James R, Meadows H, Wan S, et al. Chemoradiation for the treatment of epidermoid anal cancer: 13-year follow-up of the first randomised UKCCCR Anal Cancer Trial (ACT II). Br J Cancer. 2010;102:1123–8.

5. Bartelink H, Roelofsen F, Eschwege F, Rougier P, Botzen JF, Gonzalez DG, et al. Concomitant radiotherapy and chemotherapy is superior to radiotherapy alone in the treatment of locally advanced anal cancer: results of a phase III randomized trial of European Organization for Research and Treatment of Cancer Radiotherapy and Gastrointestinal Cooperative Groups. J Clin Oncol. 1997;15:2040–9.

6. Flam M, John M, Pajak TF, Petrelli N, Myerson R, Doggett S, et al. Role of mitomycin in combination with fluorouracil and radiotherapy, and of salvage chemoradiation in the definitive nonsurgical treatment of epidermoid carcinoma of the anal canal: results of a phase III randomized intergroup study. J Clin Oncol. 1996;14:2527–39.

7. Vuong T, Devic S, Belliveau P, Muanza T, Hegyi G. Contribution of conformal therapy in the treatment of anal canal carcinoma with combined chemoradiotherapy: results of a phase II study. Int J Radiat Oncol Biol Phys. 2003;56:623–31.

8. Salama JK, Moll LK, Schornas DA, Miller RC, Devisetty K, Jani AB, et al. Concurrent chemotherapy and intensity-modulated radiation therapy for anal cancer patients: a multicenter experience. J Clin Oncol. 2007;25:4581–6.

9. Gunderson LL, Winter KA, Ajanja JA, Pedersen JE, Moughan J, Benson AB, et al. Long-term update of U.S. Intergroup RTOG 98–11 phase III trial for anal carcinoma: survival, relapse and colostomy failure with concurrent chemoradiation involving fluorouracil/mitomycin versus fluorouracil/cisplatin. J Clin Oncol. 2012;30:4344–51.

10. James RD, Gynne-Jones R, Meadows HM, Cunningham D, Myntt AS, Saunders MP, et al. Mitomycin or cisplatin chemoradiation with or without maintenance chemotherapy for treatment of squamous-cell carcinoma of the anus (ACT II): a randomized, phase 3, open-label, 2×2 factorial trial. Lancet. 2013;381:23–4.

11. Kacznik LA, Winter K, Myerson RJ, Goodyear MD, Willis J, Estabhann J, et al. RTOG 0529: a phase 2 evaluation of dose-painted intensity-modulated radiation therapy in combination with 5-fluorouracil and mitomycin-c for the reduction of acute morbidity in carcinoma of the anal canal. Int J Radiat Oncol Biol Phys. 2013;86:273–33.

12. Lian J, Mackenzie M, Joseph K, Perez N, Dundas G, Urtasun R, et al. Assessment if extended-field radiotherapy for stage IIIC endometrial cancer using three-dimensional conformal radiotherapy, intensity-modulated radiotherapy, and helical tomotherapy. Int J Radiat Oncol Biol Phys. 2008;70:935–43.

13. Skórska M, Piotrowski T, Kazimierska J, Adamik A. A dosimetric comparison of IMRT versus helical tomotherapy for brain tumors. Fizyka Medycyna. 2014;29:219–31.

14. Joseph K, Syne A, Small C, Warkentin H, Quon H, Ghosh S, et al. A treatment planning study comparing helical tomotherapy with intensity-modulated radiotherapy for the treatment of anal cancer. Radiother Oncol. 2010;94:50–6.

15. Cox JD, Stetz J, Pajak TF. Toxicity criteria for the radiation therapy oncology group (RTOG) and the European Organization for research and treatment of cancer (EORTC). Int J Radiat Oncol Biol Phys. 1995;31:1341–46.

16. Roeseke JC, Lujan A, Reba RC, Penney BC, Diane Yamada S, Mundt AJ. Incorporation of SPECT bone marrow imaging into intensity-modulated whole-pelvis radiation therapy treatment planning for gynecologic malignancies. Radiother Oncol. 2005;77:11–7.

17. Liang Y, Bydder M, Yashar CM, Rose BS, Cornell M, Hoh CK, et al. Prospective Study of functional bone marrow-sparing intensity-modulated radiation therapy with concurrent chemotherapy for pelvic malignancies. Int J Radiat Oncol Biol Phys. 2013;85:406–14.

18. Wang X, Zhang X, Dong L, Liu H, Gillin M, Ahamad A, et al. Effectiveness of noncoplanar IMRT planning using a parallelized multiresolution beam angle optimization method for paranasal sinus carcinoma. Int J Radiat Oncol Biol Phys. 2005;63:594–601.