Clinical significance of mismatch repair gene expression in sporadic colorectal cancer

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Abstract. Mismatch repair (MMR) genes play an important role in the occurrence and development of sporadic colorectal cancer; however, the effect of MMR genes on clinicopathological features and prognosis remains unclear. The aim of the present study was to observe the clinical significance of MMR gene expression in sporadic colorectal cancer. Clinicopathological data and postoperative samples from 404 patients with sporadic colorectal cancer were obtained from the Affiliated Tumor Hospital of Xinjiang Medical University. The immunohistochemistry PV-9000 two-step method was performed to measure the protein expression of human mutL homolog 1 (hMLH1), human mutS homolog (hMSH) 2, human postmeiotic segregation increased 2 (hPMS2) and hMSH6. Differences in clinicopathological features, family history and survival time subsequent to surgery between groups with normal and aberrant MMR protein (MMRP) expression were compared. A total of 27.23% of all patients showed aberrant nuclear staining of MMRP. Among the patients with aberrant MMRP expression, a higher proportion of patients showed aberrant expression of more than one type of MMRP than aberrant expression of only one type of MMRP. Aberrant expression of hMLH1/hPMS2 was most commonly observed (29/404). In addition, aberrant MMRP expression in colorectal cancer was indicated predominantly in the right hemicolon. Histological type primarily showed mucinous adenocarcinoma. In addition, with increasing body mass index (BMI), the MMRP deficiency rate was also shown to increase gradually. There was a close association between MMRP expression deficiency and family history of cancer (P<0.05). For TNM stage III patients, the Kaplan-Meier survival curve showed that the aberrant MMRP expression group had a three-year disease-free survival (DFS) rate of 66.67%, which was longer than the DFS rate of the normal group (55.41%), with no statistical difference (P>0.05). In conclusion, the immunohistochemistry PV-9000 two-step method can be used to measure MMRP expression in colorectal cancer. Aberrant MMRP expression is closely correlated with tumor location, histological type, BMI and tumor family history in sporadic colorectal cancer. Aberrant MMRP expression may have an effect on the prognosis of stage III patients.

Introduction

The occurrence and development of colorectal cancer is a complicated multi-step process, which involves numerous factors and genes. A number of tumor-related events are involved in this process, including oncogene activation, tumor suppressor gene inactivation, mismatch repair (MMR) gene mutations and gene promoter hypermethylation (1,2). Since the identification of MMR genes, studies have investigated the association between the aberrant expression of MMR genes and hereditary nonpolyposis colorectal cancer (HNPPC) or sporadic colorectal cancer (3-5). A number of studies have found that aberrant MMR gene expression plays an important role in the occurrence of colorectal cancer (6,7). At present, numerous genes are known to be involved in the MMR process, including human mutL homolog 1 (hMLH1), human mutS homolog (hMSH) 2, hMSH6, human postmeiotic segregation increased (hPMS) 1, hPMS2, hMSH3 and hMSH5. The protein products of MMRP expression are enzymes that can repair mismatched base groups in the DNA replication process in order to maintain the fidelity of DNA replication.

At present, there are numerous studies investigating the pathogenesis of HNPPC (8,9); however, fewer studies have investigated the role of MMR gene mutations in sporadic colorectal cancer and microsatellite instability (MSI). A previous study found that ~15% of sporadic colorectal cancer cases exhibit a similar pathogenesis to HNPPC (10). However, the contribution of MMR gene mutation to the pathogenesis

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of these two types of colorectal cancer is considered to be different. It has been indicated in a number of previous studies (11,12) that MMR gene mutations can result in tumorigenesis through two mechanisms. Firstly, simple sequence repeats can cause homologous genetic recombination in the DNA replication process. Consequently, variations in the sequence containing the simple sequence repeats increase DNA MSI in tumor cells. Secondly, aberrant MMR gene expression can result in the accelerated accumulation of gene mutations in proto-oncogenes and cancer suppressor genes. Consequently, this can affect the proliferation regulation of normal cells. In recent years, studies have focused on four types of MMR genes, MLH1, MSH2, MSH6 and PSM2 (13-15).

In excess of 90% of patients with HNPCC have DNA with high MSI (MSI-H), suggesting that the occurrence of HNPCC is associated with the functional loss of cell MMR. Therefore, DNA MSI can be regarded as a reliable indicator to measure the function of cell MMR (16). A typical characteristic of MMR gene mutation is MSI expression. The microsatellite sequence mutation rate due to MMR of tumor cells is 100-1,000 fold higher than that of normal cells. Furthermore, the MSI in colorectal tumors caused by aberrant MMR gene expression is ~15% (17). Therefore, detecting MSI is of high value. MSI may be used as a positive prognostic factor for sporadic colorectal cancer (18), but also as a negative forecasting sign for fluorouracil-based chemotherapy (19,20).

Informed consent was obtained from all the patients prior to inclusion in the study and this study was approved by the Medical Ethics Committee of the Affiliated Tumor Hospital of Xinjiang Medical University (no. W201302).

**Immunohistochemistry.** The neutral formalin-fixed (concentration, 40 g/l), paraffin-embedded specimens were serially sectioned (5-μm thickness), and a PV-9000 two-step method was performed using mouse anti-human monoclonal antibodies against MLH1, MSH2, MSH6 and PSM2 (Beijing Zhongshan Golden Bridge Biotechnology Co., Ltd., Beijing, China) as primary antibodies with a working concentration of 1:150. A universal two-step method (horseradish peroxidase) detection kit (Fujian Maixin Biological Products Co., Ltd., Fuzhou, China) was utilized. Phosphate-buffered saline was used instead of primary antibodies as a negative control, while normal colorectal mucosa and/or infiltrating lymphocytes were used as a positive control. Positive expression of MLH1 and MSH2 was observed in the nucleus. The results were analyzed in accordance with the method previously described by Plevová et al (22), in which the number of microscopic tumor cells showing positive nuclear staining was combined with the staining intensity and percentage of positive cells to determine the positive expression levels. A total of five high-power fields were selected from each sample using a light microscope and 100 cells were counted in each field. The grading of staining intensity was as follows: no staining, 0 points; light yellow, 1 point; yellow, 2 points; and brown, 3 points. The grading of the percentage of positive cells was as follows: No positive cells, 0 points; ≤10%, 1 point; 11-50%, 2 points; 51-75%, 3 points; and >75%, 4 points. If the result obtained by multiplying the two scores above was ≥2 points, the case was considered to have positive expression; however, if the score was <2 points, the case was considered to have negative expression. The positive control was positive nuclei of normal colorectal mucosa and/or infiltrating lymphocytes. However, a negative result was judged in the case of positive nuclear expression in the positive control and missing staining in the tumor cell nuclei.

**Materials and methods**

**Patients and clinicopathological parameters.** Clinicopathological data and postoperative samples from 404 patients with sporadic colorectal cancer were collected from the Tumor Hospital of Xinjiang Medical University (Urumqi, China). The aims of the study were to detect MMR protein (MMRP) expression using immunohistochemistry, in order to elucidate how aberrant MMRP expression was distributed in Chinese patients with sporadic colorectal cancer, and to analyze the association between aberrant MMRP expression and clinicopathological features, in order to investigate their prognostic effect.

**Statistical analysis.** Univariate analysis was performed using the χ² test. Multivariate correlation analysis was performed using the logistic regression test. Disease-free survival (DFS) was analyzed using the Kaplan-Meier method and the Log-rank test was used for comparison between groups. The statistical analysis was performed using SPSS for Windows version 18 (SPSS Inc., Chicago, IL, USA). The Fisher's exact test from the statistical package STATA 9.0 (Stata Corp., College Station, TX, USA) was used for the calculations. P<0.05 was considered to indicate a statistically significant difference.

**Results**

**Immunohistochemical results of MMRP expression.** In 404 cases with sporadic colorectal cancer, 110 (27.23%) patients showed aberrant nuclear staining for MMRP. For patients with only one type of aberrant expression, hMLH1 expression was absent in 17 cases, hMSH2 in 9 cases, hPSM2 in 7 cases and hMSH6 in 5 cases. In patients with more than one type of aberrant MMRP expression, the protein expression of hMLH1/hMSH2/hPSM2/hMSH6 was absent in 3 cases, hMLH1/hMSH2/hMSH6 in 3 cases, hMSH2/hPSM2/hMSH6...
in 4 cases, hMLH1/hPMS2 in 29 cases, hMSH2/hMSH6 in 17 cases, hMLH1/hMSH2 in 7 cases, hMLH1/hMSH6 in 5 cases and hMSH2/hPMS2 in 4 cases. The highest frequency of aberrant MMRP expression was for hMLH1/hPMS2 (Fig. 1).

Univariate analysis between MMRP expression and clinicopathological parameters. Using univariate analysis, aberrant MMRP expression in colorectal cancer was found to be closely associated with tumor location, histological type, BMI and family history of cancer, and this was statistically significant (P<0.05; Table I).

Multivariate analysis between MMRP expression and clinicopathological parameters. Using logistic regression analysis, independent risk factors for aberrant MMR were identified; these included tumor location, histological type, BMI and family history of cancer (P<0.05; Table II).

Survival analysis of normal and aberrant MMRP expression groups. A total of 104 cases with stage III colorectal cancer were randomly selected from 404 cases and followed up for >3 years. A total of 5 cases were lost midway through the follow-up: Three patients succumbed (two due to other diseases and one due to a traffic accident), one patient moved abroad, leading to a loss of contact, and one patient quit the study midway due to mental disease. In total, 74 cases had normal MMRP expression, while 30 cases exhibited aberrant MMR expression. The three-year overall survival was 70.19%, and 31 cases succumbed from distant metastasis or local recurrence. However, the three-year DFS rate was 58.65%, of which 20 cases showed aberrant MMRP expression. The three-year overall survival was 70.19%, and 31 cases succumbed from distant metastasis or local recurrence. However, the three-year DFS rate was 58.65%, of which 20 cases showed aberrant MMRP expression. The Kaplan-Meier survival curve showed that the aberrant expression group had a three-year DFS rate of 66.67%, which was higher than the three-year DFS rate of the normal group (55.41%). However, no statistical difference was found using the Log-Rank test (P>0.05; Fig. 2).

Figure 1. Immunohistochemical staining showing normal and aberrant MMRP expression (magnification, x100). (A) hMLH1, (B) hMSH2, (C) hPMS2 and (D) hMSH6. (Aa-Da) Normal immunohistochemical staining of (Aa) hMLH1, (Ba) hMSH2, (Ca) hPMS2 and (Da) hMSH6. Normal nuclear staining of the MMRPs can be observed not only in stromal cells, but also in epithelial tumor cells, showing a brownish accumulation of dye in the nucleus. (Ab-Db) Aberrant staining of (Aa) hMLH1, (Ba) hMSH2, (Ca) hPMS2 and (Da) hMSH6. Aberrant nuclear staining of the MMRPs can only be observed in stromal cells, not in epithelial tumor cells. MMRP, mismatch repair protein; hMLH1, human mutL homolog 1; hMSH, human mutS homolog; hPMS2, human postmeiotic segregation increased 2.
Table I. Univariate analysis between MMRP expression and clinicopathological parameters.

| Clinicopathological index | Normal MMRP, n=294 | Aberrant MMRP, n=110 | Total, n=404 | $\chi^2$ | P-value |
|----------------------------|-------------------|----------------------|--------------|---------|---------|
| Age (years)                |                   |                      |              | 0.274   | 0.601   |
| <50                        | 88                | 30                   | 118          |         |         |
| ≥50                        | 206               | 80                   | 286          |         |         |
| Gender                     |                   |                      |              | 0.335   | 0.563   |
| Male                       | 167               | 66                   | 233          |         |         |
| Female                     | 127               | 44                   | 171          |         |         |
| Nationality                |                   |                      |              | 3.907   | 0.272   |
| Han                        | 219               | 73                   | 292          |         |         |
| Uyghur                     | 38                | 16                   | 54           |         |         |
| Hui                        | 21                | 12                   | 33           |         |         |
| Others                     | 14                | 9                    | 25           |         |         |
| BMI (kg/m$^2$)             |                   |                      |              | 7.911   | 0.048   |
| <18.5                      | 20                | 4                    | 24           |         |         |
| 18.5-23.99                 | 133               | 37                   | 170          |         |         |
| 24-27.99                   | 107               | 49                   | 156          |         |         |
| ≥28                        | 34                | 20                   | 54           |         |         |
| Anemia                     |                   |                      |              | 1.238   | 0.266   |
| Yes                        | 83                | 25                   | 108          |         |         |
| No                         | 211               | 85                   | 296          |         |         |
| Tumor size (cm)            |                   |                      |              | 1.258   | 0.533   |
| <4                         | 68                | 25                   | 93           |         |         |
| 4-6                        | 156               | 53                   | 209          |         |         |
| ≥6                         | 70                | 32                   | 102          |         |         |
| Tissue type                |                   |                      |              | 7.226   | 0.007   |
| Glandular                  | 244               | 78                   | 322          |         |         |
| Mucous gland/signet cell   | 50                | 32                   | 82           |         |         |
| Differentiation degree     |                   |                      |              | 8.119   | 0.004   |
| Well/moderately            | 200               | 58                   | 258          |         |         |
| Poorly                     | 94                | 52                   | 146          |         |         |
| Tumor general type         |                   |                      |              | 0.257   | 0.880   |
| Ulcerative                 | 196               | 76                   | 272          |         |         |
| Mass                       | 85                | 29                   | 114          |         |         |
| Infiltrative               | 13                | 5                    | 18           |         |         |
| TNM staging                |                   |                      |              | 1.061   | 0.786   |
| I                          | 21                | 8                    | 29           |         |         |
| II                         | 104               | 33                   | 137          |         |         |
| III                        | 144               | 59                   | 203          |         |         |
| IV                         | 25                | 10                   | 35           |         |         |
| Tumor location             |                   |                      |              | 11.607  | 0.003   |
| Rectal                     | 159               | 46                   | 203          |         |         |
| Left hemicolon             | 86                | 28                   | 114          |         |         |
| Right hemicolon            | 51                | 36                   | 87           |         |         |
| Familial cancer history    |                   |                      |              | 7.510   | 0.023   |
| Colorectal cancer          | 20                | 16                   | 36           |         |         |
| Others                     | 45                | 21                   | 66           |         |         |
| No                         | 229               | 73                   | 302          |         |         |

MMRP, mismatch repair protein; BMI, body mass index.
Discussion

To date, a number of previous studies have confirmed that immunohistochemistry is a reliable method for MMR gene analysis (23-26). The method has been utilized in the majority of hospitals and research institutions, and has been shown to be cost effective, stable and with a high sensitivity (77 -100%) and specificity (98‑100%) (23,24). As a result, the immunohistochemical method has been suggested as the preferred method for MMR gene mutation analysis (25,26). The immunohistochemistry PV-9000 two-step method is an enzymatic biotin method. Monovalent Fab fragments of second antibody molecules polymerize with enzymes instead of the traditional method of secondary and tertiary antibodies. Consequently, the antigen-antibody binding signal is directly amplified. Compared with the traditional streptavidin-peroxidase three-step method, the PV-9000 two-step method is simple, fast and sensitive. In addition, it avoids background staining due to a lack of biotin. Thus, the immunohistochemistry PV-9000 two-step method is often used in clinical practice. In the present study, the immunohistochemistry PV-9000 two-step method was performed to measure the expression levels of hMLH1, hMSH2, hPMS2 and hMSH6 in 404 postoperative pathological specimens.

Numerous studies have investigated MLH1 and MSH2 expression (27,28); however, fewer studies have investigated MSH6 and PSM2. It has been shown that the rate of aberrant MLH1 expression is higher than that of MSH2 (23). This may be due to the inactivation of the MLH1 gene in somatic cells (29). CpG islands within the MLH1 gene promoter region are hypermethylated. This methylation causes barriers against gene transcription and translation, resulting in aberrant MLH1 expression. Aberrant MLH1/PSM2 expression is the most common type of aberrant MMR gene expression due to the high frequency of MLH1 methylation and easy heterodimer formation. Correspondingly, aberrant PSM2 expression becomes relatively higher (30,31). In the present study, only one type of aberrant MMRP expression was observed in 38 cases (9.4%). Aberrant expression of hMLH1/hPSM2 showed the highest rate (26.36%), while the rate of aberrant hMSH2/hMSH6 expression was the second highest (15.45%). The results from this study were consistent with those from a previous study by Molaei et al (32).

A number of previous studies (33,34) have demonstrated that aberrant MMR is associated with certain clinicopathological features. This association has an important role in the clinical diagnosis and treatment of colorectal cancer. In the present study, cases where the tumor was in the right hemi-colon or the tissue type was mucous gland or signet ring cell carcinoma were found to have a higher incidence of aberrant MMRP expression, which is consistent with the results from previous studies (23,35). This may be due to the fact that aberrant MMRP expression is closely associated with MSI-H. The clinicopathological features of right hemicolon tumors or mucinous adenocarcinomas include MSI-H (36-38). In the present study, no difference was observed between rectal and left hemicolon tumors with regard to MMRP expression.

| Variable                        | B-value | OR  | Lower bound | Upper bound | P-value |
|---------------------------------|---------|-----|-------------|-------------|---------|
| BMI (kg/m²) ≤24 vs. ≥24         | 0.341   | 0.711| 0.529       | 0.956       | 0.024   |
| Histological type               |         |     |             |             |         |
| Glandular vs. mucous gland/signet cell | 0.609 | 1.838| 1.072       | 3.152       | 0.027   |
| Tumor location                  |         |     |             |             |         |
| Left hemicolon/rectal vs. right hemicolon | 0.761 | 0.467| 0.278       | 0.784       | 0.004   |
| Family history of cancer        |         |     |             |             |         |
| Positive vs. negative           | 0.413   | 1.511| 1.075       | 2.124       | 0.017   |

MMRP, mismatch repair protein; BMI, body mass index; OR, odds ratio.
suggesting that the aberrant MMR expression pathways exhibit consistency. However, a statistical difference was observed between left and right hemicolon carcinomas, suggesting that a higher incidence of gene promoter hypermethylation may occur in the right hemicolon tissues, leading to the occurrence of MSI.

In the present study it was demonstrated that the rate of aberrant MMRP expression was not associated with age at diagnosis, gender, nationality, anemia, tumor size or TNM staging (P>0.05). The association between anemia and MMRP expression has, to this date, been unclear. The rate of aberrant MMRP expression in the anemia group (23.15%) was lower than that in the normal hemoglobin group (28.72%); however, this difference was not statistically significant (P>0.05). Tumors with aberrant MMRP expression were mostly located in right hemicolon, the clinical manifestations of which showed a higher risk of anemia. Further studies are required to elucidate the specific association between these factors.

With improvements in living standards, dietary structure has also been changing. The dietary habit of consuming more meat and less fiber has caused an increasing incidence of overweight and obese individuals. An increasing number of studies are focusing on the association between BMI and colorectal cancer. Several studies have shown a close correlation between increasing BMI and risk factors of colorectal cancer (39,40). However, with the exception of the study by van Duijnhoven et al (41), which described certain aspects of the association between BMI and MMR gene expression, studies focusing on the association between BMI and MMR gene expression are relatively rare. In the present study, increasing BMI was significantly correlated with aberrant MMRP expression. In the study by Botma et al (42), it was revealed that BMI had a close correlation with colorectal adenomas; however, the study subjects were all male. In the study by Win et al (43), manifested BMI was reported to be a potential risk factor for individuals in early adulthood carrying MMR gene mutations. Therefore, previous study results suggest that MMR gene mutation occurs in the early pathogenetic stage of colorectal cancer. Being overweight or obese may be independent risk factors of aberrant MMR gene expression. However, further studies are required to investigate the underlying mechanism, as this has yet to be elucidated.

Studies investigating whether the pathogenesis of sporadic colorectal cancer in patients with a tumor familial history is the same as that of HNPCC are rare. Germline MMR gene mutations have been identified as the molecular genetic basis underlying HNPPC. By contrast, mutations in the adenomatous polyposis coli gene are believed to comprise the molecular genetic basis underlying familial adenomatous polyposis and the majority of sporadic colorectal cancer cases. Sporadic colorectal cancer additionally exhibits a polygenic and multi-stage process of tumor formation, which includes activating mutations in adenoma-carcinoma sequences in oncogenes and inactivating mutations in tumor suppressor genes (44,45). In the present study, the rate of aberrant MMRP expression in the group with a family history of cancer (36.27%) was higher than that in the group without a family history of cancer (24.17%), with a statistically significant difference (P<0.05). Therefore, cancer family history was correlated with aberrant MMR expression.

A number of studies have revealed that patients with a positive MSI in colorectal cancer show a more favorable prognosis (46,47); however, the mechanism associated with this remains unclear. Popat et al (18) reported that, although colorectal cancer with MSI-H had numerous features associated with a poor prognosis, MSI-H was also associated with a relatively good prognosis due to increased inflammatory cell infiltration. In addition, Sargent et al (20) revealed that cancer with MSI-H was not sensitive to 5-FU-based chemotherapy. However, as to whether it is associated with MMR gene mutations, a number of studies (48,49) have produced affirmative results. In the present study, 104 patients with stage III colorectal cancer were followed up for >3 years. Survival analysis showed that the three-year DFS of the aberrant MMRP expression group was higher than that of the normal expression group. However, no statistically significant difference was identified between the groups (P>0.05). This may be due to the fact that patients with aberrant MMRP expression had a higher MSI, which, according to the above studies, was a good prognostic factor.

In conclusion, the immunohistochemistry PV-9000 two-step method can be feasibly used to detect the MMRP expression level in sporadic colorectal cancer. MMRP expression is closely associated with tumor location, histological type, differentiation degree, BMI and a family history of cancer, respectively. MMRP expression level may be a promising prognostic factor. Therefore, MMR plays a significant role in the occurrence and development of colorectal cancer; further studies are required to explore its detailed mechanism.

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