EFFECTIVENESS OF PRE PRP INJECTION AND POST ELEVATION FLAP AT EXTENDED RANDOM FLAP RAT SKIN

Thomas Eduardus Sudrajat Wahyu Nugroho, Sitti Rizaliyana, David S Perdanakusuma
Department of Plastic Reconstrutive and Aesthetic Surgery, Faculty of Medicine, Universitas Airlangga, Dr. Soetomo General Hospital, Surabaya, Indonesia

ABSTRACT

Closure of the defect with a random skin flap is constrained by the extent of the defect area. Several studies have shown the benefits of administration of PRP (Platelet Rich Plasma) in random skin flap. The purpose of this study is to compare the effect of PRP injection given before flap elevation, after flap elevation and control on the extended random skin flap procedures in rats. This was an experimental study with randomized posttest-only control group design (n=27, divided into 3 treatment groups) that compare the effectiveness of PRP injection 24 hours prior to the elevation of the flap, after the elevation of the flap, and control in the extended random skin flap in rats. Random skin flap is made in the ratio 1:5 on the rat skin. Measurement of viable area were observed on days 1, 7 and 14. There were significantly increased viability of random skin flap on the group with PRP injection 24 hours prior flap elevation compared to other group of treatment. The average of viability on day first 39%±13%; 42%±34%; 62%±14%. On day 7th 24%±13%; 36%±26%; 62%±12%. On day 14th 16%±15%; 28%±22%; 60%±11%. Injection of PRP on extended random skin flap on rat 24 hours before flap elevation increase viability of the flap compared to control group and group which receive PRP injection on extended random flap on rat skin after elevation of the flap. Viability increased due to angiogenesis stimulation after PRP injection.

Keywords: Platelet Rich Plasma; flap survival; extended random skin flap

INTRODUCTION

In recent years, surgical procedures that leave extensive defects that are difficult to cover primarily is increasing. The extensive defect causes exposure of important structures such as blood vessels, tendons and bones, so that reconstructive surgery for the closure of the defect with the flap also has a significant increase. In Dr. Soetomo General Hospital, the use of random skin flaps to close defects has become a routine procedure.
Platelet Rich Plasma (PRP) is a product of blood plasma, rich in platelets and a kind of natural source of growth factors and fibrinogen which is thought to accelerate healing of soft tissue (Carlson et al 2002). Kim et al in 2013 proved that injecting PRP on axial flaps can increase the success of the flap (reducing necrotic area) in rats. Giving injection of PRP in the skin flap can increase the success rate through angiogenic and arteriogenic mechanisms (Kim et al 2013). In this study the researchers wanted to find out the effectiveness of PRP in extended random skin flap, which was pre-injected and post-action flap elevation in increasing the success of extended random skin flap in rats.

MATERIALS AND METHODS

Preparation of Platelet-Rich Plasma

All of the procedures were approved by the Ethics Committee of the Faculty of Veterinary Medicine, Universitas Airlangga. Winstar male adult males rats (200-300 grams) were given ketamine. After sedation, intracardiac blood was taken in 3 rats. Rat blood was then processed into PRP. PRP was made from 3 ml of aspiration of rat intracardiac blood mixed with 0.5 ml of anticoagulant in the form of a solution of Anticoagulant Citrate Dextrose (ACD) through twice intracardiac blood transfusion. The first centrifuge, which is a hard spin with a 4000 rpm centrifuge for 10 minutes, separates intracardiac blood into three layers from top to bottom, namely the plasma layer, buffy coat layer and erythrocyte layer. For soft spin centrifuging, the top two layers, the plasma layer and buffy coat, are separated into another tube and then centrifuged at a speed of 2000 rpm for 5 minutes. The second centrifuge separates into two layers, namely the platelet-rich plasma layer below and the plasma layer with a slightly platelet count above. The top layer is taken by leaving a layer of platelet concentrate in the plasma. This concentrate is then mixed with 0.1 ml of calcium chloride (CaCl2) 10% to activate platelets (De Rossi et al 2009). This PRP is then immediately injected into the flap. Pre-elevation PRP injection flap is given as much as 0.2 cc in total done 24 hours beforehand at 5 points with a distance of 1cm from the proximal flap.

Extended random skin flap

In this study rats were given a premedicated drug with atropine 0.02 mg/kgBW and acipromasin 0.1mg/kgBW then given ketamine (Ketalar®, Parke Davis) at a dose of 0.5-1 mg/kgBW, after which an aseptic procedure was performed/antiseptic in the back region of rats that have been given PRP before, or in the back region of rats that have not been given PRP. Rats will get 3 treatments (given PRP before elevation flap, PRP after elevation flap, and control) and specimen collection and examination will be carried out with Visitrek® (day 21), so that a total of 27 rats are needed.

Area of viable flap

The area of the viable is seen regularly. Namely at 1, 7 and 14 days post elevation using a Visistrek® tool. The viable area of the flap is marked by the skin color of the rats that fits the surrounding tissue, warm touch, <2 seconds CRT. In this study it was measured as a percentage of the total flap area, this was to avoid any bias due to the flap contraction factor. Data from the study were obtained from measurements of extended random skin flap and the area that was still viable. Measurements were carried out by researchers with transparent markers and paper, the data obtained were then measured in extent by Visitrek®. The data obtained were analyzed using the Kolmogorov-Smirnov One-Sample method and the research data were analyzed using Independence Sample T-Test with SPSS 17.0 software.

On day 0 the elevation of skin flaps was carried out on the backs of rats measuring 1cm x 5cm in the K and P1 groups, in P2 group PRP injections were carried out in the area where the skin flap elevation would be carried out. The evaporated skin flap is sewn back into the skin to prevent a change in position, put vaseline paper under the flap to prevent imbibition of nutrients from below.

Measurement of necrotic area was carried out on days 1, 7 and 14. Measurements were made by means of the necrotic area being copied on transparent paper, measuring using a digital visitrak. The data obtained from the results of this study are in the form of data on the area of the viable flap on male back rats (Rattus norvegicus strain Wistar). Data on viability of mouse skin flaps is data with a ratio scale.

RESULTS

The average area of the viable area in the control group is 39% with a standard deviation of 13% on the first day, on the 7th day with an average of 24% with a standard deviation of 13%, while on the 14th day with an average of 16% and a standard deviation of 15%. In the pre-elevation group PRP injection treatment group the mean value on the first day of observation was 62% with a standard deviation of 14%, on the 7th day with a mean of 62% and a standard intersection of 12% while on the 14th day with an average of 60% and standard deviation 11%.
Table 1. The average of viable area

| Treatment | Without PRP (%) | PRP Post Elevation (%) | PRP Pre Elevation (%) | p   |
|-----------|----------------|------------------------|-----------------------|-----|
| Viability Flap Day-1 | 39 ± 13 | 42 ± 34 | 62 ± 14 | 0.054 |
| Viability Flap Day-7 | 24 ± 13 | 36 ± 26 | 62 ± 12 | 0.000 |
| Viability Flap Day-14 | 16 ± 15 | 28 ± 22 | 60 ± 11 | 0.000 |

Table 2. Paired-sample statistics

| Paired Samples Statistics | Mean (%) | Mean Difference (%) | p value |
|---------------------------|----------|---------------------|--------|
| Pair 1                    | Without PRP D-1 | 38.6807 | 14.32738 | 0.009 |
|                           | Without PRP D-7 | 24.3534 |                |        |
| Pair 2                    | Without PRP D-1 | 38.6807 | 22.37103 | 0.000 |
|                           | Without PRP D-14 | 16.3097 |                |        |
| Pair 3                    | Without PRP D-7 | 24.3534 | 08.04365 | 0.016 |
|                           | Without PRP D-14 | 16.3097 |                |        |
| Pair 4                    | PRP Post Elevation D-1 | 41.9643 | 6.30581 | 0.343 |
|                           | PRP Post Elevation D-7 | 35.6585 |                |        |
| Pair 5                    | PRP Post Elevation D-1 | 41.9643 | 13.45194 | 0.118 |
|                           | PRP Post Elevation D-14 | 28.5124 |                |        |
| Pair 6                    | PRP Post Elevation D-7 | 35.6585 | 7.14613 | 0.070 |
|                           | PRP Pre Elevation D-1 | 28.5124 |                |        |
| Pair 7                    | PRP Pre Elevation D-7 | 62.3805 | 0.03603 | 0.986 |
|                           | PRP Pre Elevation D-14 | 62.3445 |                |        |
| Pair 8                    | PRP Pre Elevation D-7 | 62.3805 | 1.79181 | 0.568 |
|                           | PRP Pre Elevation D-14 | 60.5887 |                |        |
| Pair 9                    | PRP Pre Elevation D-7 | 62.3445 | 1.75578 | 0.311 |
|                           | PRP Pre Elevation D-14 | 60.5887 |                |        |

In the group with injection of PRP after the rat skin flap elevation, the average value on the first day was 42% with a standard deviation of 34%. On the 7th day the average is 36% and the standard deviation is 26%. On the 14th day the average is 28% and the standard deviation is 22%. The mean area of viable area in the untreated (control) group decreased by 68% (in the first to last day's observations) as the observation days increased, with differences that were statistically significant. The average viable area of the total flap area on the 1st day observation was 39%, on the 7th day observation at 24%, and on the 14th day observation 16% with p <0.05.

The mean area of viable area in the group given injection of PRP after flap elevation decreased to 31% (in the first to last day's observations) with increasing observation days, although no statistically significant differences were found. The average ratio of the viable area to the total area of the flap on the 1st day observation was 41%, on the 7th day observation at 35%, and on the 14th day observation at 28% with a p value >0.05.

The mean value of the ratio of the viable area to the total flap area in the group given injection of PRP 24 hours before flap elevation decreased the area of viable area by 3% (from the first to the last observation) as the observation day increased, although no statistically significant difference was found. The average ratio of the area of the viable area to the total area of the flap on the 1st day observation was 62%, on the 7th day observation at 62%, and on the 14th day observation at 60% with a p value >0.05.

The mean ratio of the area of viable area to the total area of the flap on the first day post flap elevation of the control group was 38%, in the flap pre-elevation PRP injection group at 62% and in the PRP injection group post flap elevation was 42%. With the Viability of the...
first day flap in the PRP group the pre elevation was significantly greater than the group without PRP (p=0.026).

The mean ratio of the area of viable area to total flap area on the 7th day post flap elevation of the control group was 24%, in the flap pre-elevation PRP injection group at 62% and in the PRP injection group post flap elevation was 35%. With the viability of the flap in the pre-elevation PRP group it was significantly greater than the group without PRP (p=0.000). And the seventh day viability of the pre-elevation PRP group was significantly greater than the post elevation PRP group (p=0.003).

The ratio of the area of viable to total flap area on the 14th day post flap elevation of the control group was 16%, in the pre-elevation flap injection group of 60% and in the PRP injection group post flap elevation was 28%. With flap viability in the pre-elevation PRP group it was significantly greater than the group without PRP (p=0.000). Viability of the pre-elevation PRP flap group was significantly greater than the post elevation PRP group (p=0.000).

DISCUSSION

In this study we used homologous PRP from rats which were sacrificed for blood sampling and processed into PRP. With this PRP retrieval method, we get enough PRP to inject all tested animals, without causing tissue reactions. Researchers chose to use homologous PRP because the use of autologous PRP is technically impossible to do for small animals because they do not have enough blood, so we mix blood obtained from 4 different rats and prepare PRP from the blood (Orhan et al 2017).

There was a significant decrease in the percentage of viable area in the control group, but no significant difference was found in the group who received the PRP injection before or after the flap elevation in the first, 7th and 14th observations. In the control group there was a 68% reduction in the percentage of the viable area, in the group receiving the PRP injection before the flap elevation was 3% and in the group receiving the PRP after the flap elevation of 31%.

Table 3. Independent sample t-test

|               | Mean (%) | p value (%) | Mean Difference (%) |
|---------------|----------|-------------|---------------------|
| Without PRP D-1 | 38.6807  | 0.747       | -3.28359            |
| PRP Post Elevation D-1 | 41.9643  |             |                     |
| PRP Pre Elevation D-1 | 62.3805  | 0.026       | -3.28359            |
| PRP Post Elevation D-1 | 41.9643  |             |                     |
| PRP Pre Elevation D-1 | 62.3805  | 0.052       | -20.41618           |
| Without PRP D-7 | 24.3534  | 0.171       | -11.30516           |
| PRP Post Elevation D-7 | 35.6585  |             |                     |
| PRP Pre Elevation D-7 | 24.3534  |             |                     |
| PRP Post Elevation D-7 | 62.3445  | 0.000       | -11.30516           |
| PRP Pre Elevation D-7 | 35.6585  |             |                     |
| PRP Pre Elevation D-7 | 62.3445  | 0.003       | -26.68597           |
| Without PRP D-14 | 16.3097  | 0.115       | -12.20268           |
| PRP Post Elevation D-14 | 28.5124  |             |                     |
| PRP Pre Elevation D-14 | 16.3097  |             |                     |
| PRP Post Elevation D-14 | 60.5887  | 0.000       | -12.20268           |
| PRP Pre Elevation D-14 | 28.5124  |             |                     |
| PRP Pre Elevation D-14 | 60.5887  | 0.000       | -32.07632           |

There was a significant difference in the percentage of viable area to the total area of skin flap that was significant between the groups who received injection of PRP before the flap elevation with the control group and the group who received injection of PRP after flap elevation at 7th and 14th day observations. Provision of
PRP before flap elevation increased the ratio of the viable area to the total flap area as much as 258% to the control group at the 7th day observation. Provision of PRP before flap elevation increased the ratio of the viable area to the total flap area as much as 177% for the group receiving the PRP post elevation flap at the 7th day observation. Provision of PRP before flap elevation increased the ratio of the viable area to the total flap area as much as 375% to the control group at the 14th day observation. Provision of PRP before flap elevation increased the ratio of the viable area to the total flap area as much as 218% to the group that received the PRP post elevation flap at the 14th day observation.

In this study, no optimal dose and platelet-rich plasma concentrations were needed to improve the surviving flap area. For further use in clinical practice, different doses and concentrations of PRP may be needed depending on the type and size of the flap. Further research is needed to determine the optimal dose needed to increase the viability of the flap. This is due to an increase in angiogenesis in the group that received the PRP injection on the flap. Kim et al (2013) found that the area of the flap that received injection of PRP had better viability than the control group, whereas microscopic examination showed an increase in mature blood vessel density (Kim et al 2013).

Platelet-rich plasma, which has a platelet concentration of at least 1,000,000 platelets/mm3 in 5 mL plasma, is strongly correlated with an increase in flap healing (Marx et al 2001). PRP contains a variety of growth factors such as Platelet Derived Growth Factor (PDGF), Transforming Growth Factor-β (TGF-β), Vascular Endothelial Growth Factor (VEGF), Endothelial Growth Factor (EGF), Insulin-like Growth Factor (IGF1), Endothelial Cell Growth Factor (ECGF) (Alcántara et al 2018). Growth factors released by PRP can increase epithelialization, amount of collagen, wound strength, epidermal regeneration, spur angiogenesis, accelerate homeostasis, therefore the use of PRP can increase the viability of skin flaps (Karayannopoulou et al 2015, Li et al 2012, Takikawa et al 2011).

Li (2012) found a significant increase in PDGF expression after 8 hours, and lasted from 3 to 7 days. The release of PDGF into the skin flap can have a chemotactic effect on monocytes, neutrophils, fibroblasts, mesenchymal stem cells, and osteoblasts. PDGF is also a strong mitogen for fibroblasts and smooth muscle cells and is involved in the wound healing phase (i.e., angiogenesis, fibrous tissue formation, and reepithelialization) (Li et al 2012).

The study by Li (2012) showed that VEGF mRNA expression increased in the group given PRP injection after 8 hours to the next 72 hours. Platelet activation induces VEGF release, an angiogenesis mediator that stimulates endothelial cell proliferation (Kliche et al 2001). VEGF is a permeability factor of blood vessels released from the wound epithelium and extracellular matrix by proteases from endothelial cells, stimulates endothelial cell proliferation and increases vascular permeability. This affects extravasation of plasma proteins and creates a temporary support structure where activated endothelial cells, leukocytes and epithelial cells can migrate (Shweiki et al 1992).

PRP contains more VEGF and induces VEGF expression from endothelial cells, increases microvessel density and prevents tissue necrosis from the initial phase. To fully understand how angiogenesis occurs in skin flaps with platelet-rich plasma treatment, further analysis can be done by microscopic examination and to see blood flow using Doppler laser or flow chart speckle (Wang et al 2016).

**CONCLUSION**

PRP injection for pre-elevation flap can increase the viability of extended random skin flaps. There was a significant difference in flap viability between the treatment groups receiving the pre-elevated flap injection of PRP, and the group receiving the PRP injection post elevation flap and the control group. Pre-elevated PRP injection of flap can increase the viable area to 375% compared to the control. Pre-elevation PRP flap injection can increase the area of viable area by up to 218% compared to the group that received post elevation injection of PRP. Post-elevation PRP injection of the flap had a 175% greater area of viable area compared to the control group, but did not differ statistically. Microscopic examination can be done to see the increase in blood vessel density (angiogenesis), amount of collagen, epidermal regeneration in the administration of PRP on skin flaps and examination of levels of growth factors (VEGF, TGF-β, PDGF) to see the effect of PRP injection on skin flaps. So that it is expected to be done in patients with vascular disorders and comorbid diseases that affect the quality of blood vessels, so that the success rate of skin flap action can be increased.

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