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Can polyoxometalates (POMs) prevent of coronavirus 2019-nCoV cell entry? Interaction of POMs with TMPRSS2 and spike receptor domain complexed with ACE2 (ACE2-RBD): Virtual screening approaches

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ABSTRACT

The unexpected appearance and global spread of COVID-19 create significant difficulties for healthcare systems and present an unusual challenge for the fast discovery of medicines to combat this fatal disease. Screening metallodrugs libraries from the medicinal inorganic chemistry society may expand the studied 'chemical space' and improve the probability of discovering effective anti-COVID drugs, including polyoxometalates. POMs are an oxygen-rich family of inorganic cluster systems that have previously been tested for antiviral action against different types of viruses. Human angiotensin-converting enzyme 2 (ACE2), human transmembrane protease serine 2 (TMPRSS2), and the SARS-CoV-2 spike glycoprotein are required for host cell-mediated viral entrance. Targeting these proteins demonstrates potential possibilities for preventing infections and transmissions in the initial stage. As a result, POMs with known antiviral effects were investigated for this purpose using molecular docking and dynamic simulations. This research shows that POMs can prevent SARS-CoV-2 from entering cells by blocking TMPRSS2, which SARS-CoV-2 uses for spike glycoprotein priming. They may also engage with ACE2 and the spike glycoprotein and disrupt their binding by blocking the active sites. We think that a thorough investigation of POMs as possible anti-COVID-19 drugs will provide significant opportunities.

1. Introduction

A novel coronavirus, named as severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is the latest pandemic in the series of other infectious diseases that poses an unprecedented challenge for the rapid explore of drugs against this deadly virus. Iran, which reported two fatalities due COVID-19 50 days following China on Feb 18, 2020, is among the nations dealing with the highest number of instances of COVID-19 infections and consequent deaths [1]. Even with the promising results of vaccines developed in the world, it is well recognized that the need for additional modalities is essential, due to the sheer enormity of the problem [2]. Therefore, both experimental and computational approaches are employed to investigate appropriate drugs from the library of FDA-approved drugs against this deadly virus. SARS-CoV-2 is an RNA virus that encapsulated inside a membrane envelope which has proteins like spikes that sticking out from its surface. These proteins are surface exposed and are involved in virus entry into the host cells [3]. The spike glycoprotein of SARS-CoV-2 contains two subunits: S1 subunit that contains the receptor binding domain (RBD) [4], and the S2 subunit (carryout the union of viral and host cell membrane). These subunits are responsible for binding to the host cell angiotensin-converting enzyme 2 peptidase domain (PD) and ensuring membrane fusion with the host cell, respectively [5]. ACE2 is an enzyme that is found on the outer membranes of the intestines, lung cells, kidneys, arteries and heart and is a primary target for CoVs. Interaction of surface spike glycoproteins of CoVs with this enzyme facilitates the virus entry into the host cell [6]. At first, the spike glycoprotein of SARS-CoV-2 is cleaved by host cell transmembrane protease serine 2 (TMPRSS2) [7,8], then ACE2 is hijacked by cleaved spike glycoprotein as an entry point to host cell [9]. This process provides the entry of the viral RNA genome into the host cell and increases the SARS-CoV-2 chance of human to human transmission [10]. Alternative therapeutic options must be investigated when a novel severe disease emerges for which there are no viable medical therapies, such as COVID-19. Metals and metallic compounds have long fascinated physicians for their intriguing, almost ‘magic’ characteristics and have therefore played a key role in the pioneering days of advanced

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pharmacology beginning in the late 19th to early 20th century. Notably, over the last 40 years, attention in metal-based drugs has grown significantly, due mainly to the enormous successful treatment of cisplatin, which the FDA first approved in 1978, and has greatly contributed to a fairly large and active scientific community working in the field of inorganic medicinal chemistry. The distinct chemical and biological characteristics of different metal centers, in certain instances, non-physiological metals, can be used for medicinal purposes [11,12]. POMs are inorganic metal-oxide assemblies presenting great interest properties in medicine including, a wide range of possible structures, metal-combination and size, as well as large redox chemistry and high solubility in water and the low toxicity toward the human body. Therefore, POMs were examined to produce inorganic-drugs against bacteria, tumor cells, and viruses. Computational chemistry is a helpful way to illuminate POM complex chemistry and its structure [13]. Molecular docking and molecular dynamics simulation are two major computational methods that provide great applicability in pharmaceutical research industries in search of novel drugs in a short duration time [14]. POMs were reported to have anti-tumor, anti-bacterial and anti-viral activities. In the antibacterial and antiviral actions, suppression of translation/transcription processes and inhibition of viral binding to the host cell and/or penetration have been observed. There is no organized study on the inhibition of the coronavirus by POMs to the best of our knowledge. The primary aim of this research is to investigate the ability of POMs to engage with and block the cleavage function of TMPRSS2, which may prevent CoVs from binding and fusion to cells, thus preventing cell infection as well as viral multiplication. We examined POMs further using in silico calculations to see how effective they were in inhibiting spike glycoprotein binding to the ACE2 receptor and, as a result, inhibiting COVID-19 progression in an infected host. Our group thinks that extensive investigation of POMs as possible anti-COVID-19 drugs will provide significant possibilities; grounds for this belief are provided below.

2. Methods

2.1. Molecular docking studies

2.1.1. Receptors and ligand preparation

In this section, we selected an α-Keggin structure of POMs [SiW12O40]−4 to perform molecular docking simulations and evaluate the binding affinity and interaction of the POMs to TMPRSS2 and SARS-CoV-2 chimeric receptor-binding domains complexed with ACE2 (ACE2-RBD). Such computational study helps us to assess the potential of POMs in preventing SARS-CoV-2 cell entry. AutoDock Vina [15] with MGL tools 1.5.4 was used to perform blind docking calculations. AutoDock Vina runs faster than AutoDock software and makes more precise docking calculations [16]. The TMPRSS2 (PDB ID: 2OQ5) and ACE2-RBD (PDB ID: 6LZG) were obtained from the Protein Data Bank, and inhibitors were selected and removed. AutoDock Tools (v.1.5.4) was used to prepare the receptors. MGL Tools (v.1.5.4) combined non-polar hydrogen atoms and added Kollman charged atoms to protein crystal structures. TMPRSS2 and ACE2-RBD coordinate files were then saved in PDBQT format. The 2OQ5 was surrounded in a 52 × 46 × 44 Å box direction with a grid spacing of 1.00 Å and grid set centers of −1.87, 18.03, 17.47 Å, while the 6LZG was surrounded in a 64 × 74 × 110 box direction with a grid spacing of 1.00 Å and grid set centers of −25.41, 18.43, and −6.37. The structures of POM (PDB ID: 6Y7O) were obtained from protein data bank and inhibitors were selected and removed. AutoDock Tools (v.1.5.4) was used to prepare the receptors. MGL Tools (v.1.5.4) combined non-polar hydrogen atoms and added Kollman charged atoms to protein crystal structures. TMPRSS2 and ACE2-RBD coordinate files were then saved in PDBQT format. The 2OQ5 was surrounded in a 52 × 46 × 44 Å box direction with a grid spacing of 1.00 Å and grid set centers of −1.87, 18.03, 17.47 Å, while the 6LZG was surrounded in a 64 × 74 × 110 box direction with a grid spacing of 1.00 Å and grid set centers of −25.41, 18.43, and −6.37. The structures of POM (PDB ID: 6Y7O) were obtained from protein data bank. MGL Tools (v.1.5.4) was used to select the POM in PDBQT format, and docked results were visualized through the BIOVIA Discovery Studio software. The following are the methods for achieving rigid docking of POM. Initially, during the simulation process, the rigid form of POM is accurately preserved [17]. Secondly, to improve electrostatic interaction, MULLIKEN charges are allocated to each atom within POM [17]. Lastly, for tungsten and silicon, efficient “dummy” atoms are employed. In this study, iodine is used in place of tungsten, and carbon is used in place of silicon. The atomic replacement method’s concept is described in Hill and colleagues’ paper [18]. Optimization of the POM was performed using Gaussian 09 software package at the semi-empirical PM6 method [19,20]. We hypothesized that POMs with antiviral properties may disrupt the cell entry of CoVs and consequently prevent viral replication, transmissibility, and pathogenicity. Hence, we examined the virtual interaction of the POMs to the TMPRSS2 and ACE2-RBD.

2.2. Molecular dynamics (MD) simulations

The molecular dynamic simulations of POM were investigated by using GROMACS software in the presence of TMPRSS2 and spike receptor domain complexed with ACE2 proteins. Classical MD simulations [21,22] were conducted using the CHARMM27 force field [23,24]. TIP3P water [25] was used to solve the free protein and protein-ligand complexes in the cubic box with periodic boundary conditions in three directions. The solutes were positioned in the box’s center, with a minimum distance of 1.0 nm between the solutes’ surface and the box. Na+ ions were added to the system to neutralize the charge. The systems were balanced at 300 K and 1.0 bar after energy minimization through the steepest descent method. The Parrinello-Rahman barostat was used to maintain a pressure of 1.0 bar, and a temperature of 300 K was maintained using a modified Berendsen thermostat. The LINCS algorithm enabled the computation of bond lengths, while the particle-mesh Ewald scheme (PME) was used to compute long-range electrostatic forces (grid spacing 0.16 nm) [26]. The short-range nonbonded interactions were computed using cutoff ratios of 1.0 nm for van der Waals and Coulomb potentials. Finally, a 50 ns MD simulation with a time step of 2 fs was performed with random generation of velocities through a Maxwell distribution.

2.3. Interaction analysis by MM-PBSA binding energy

Using the Molecular Mechanics Poisson Boltzmann Surface Area (MM-PBSA) technique, the binding free energy of protein-ligand systems was calculated. This in silico method was a combined energy system described by the binding free energy, composed of electrostatic, SASA, van der Waals, and polar solvation energies. The MM-PBSA binding free energies were calculated using the GROMACS script g.mmpbsa [27]. Using the MM-PBSA method, the following equation was employed to determine the binding free energy of the interacting proteins:

\[ \Delta G_{\text{binding}} = \Delta G_{\text{complex}} - (\Delta G_{\text{receptor}} + \Delta G_{\text{ligand}}) \]

\[ \Delta G_{\text{binding}} \] denotes the total binding energy of the protein-ligand complexes; \( \Delta G_{\text{receptor}} \) and \( \Delta G_{\text{ligand}} \) denote the binding energy of the free receptor and unbound ligand, respectively.

3. Results

3.1. Molecular docking simulation

While significant efforts are being made to develop drugs and vaccines against COVID-19, only a few available therapeutic agents are currently available. In this regard, molecular knowledge of the virus increases quickly, and potential druggable targets are being discovered and profiled [28]. Several inorganic medicines are still used in medical care today for a few particular applications in which they serve important and irreplaceable functions, combining exceptional effectiveness with tolerable toxicity [29,30]. The most notable example is the widespread utilization of cisplatin as well as its analogs in chemotherapy; despite their significant systemic toxicity, platinum medications are believed to be included in approximately 50% of present chemotherapeutic regimens for treatment for cancer [30]. Because of the high severity of cancer, their pertinent toxicity could be tolerated on the
Polyoxometalates (POMs) are oxygen rich class of inorganic cluster systems that the antiviral activity of them has previously been examined against various types of viruses. These negatively charged clusters with three-dimensional structures perform remarkable biological activities, with low toxicity and high efficacy. POMs were reported to have in vitro antiviral activities against different virus types. many polyoxometalates, in particular, $K_2[PTi_2W_9O_{37}]_2\cdot 6H_2O$ (PM-19), $[Pr_{10}W_{38}O_{162}]^2-$H$_2$O (PM-523), $K_2H_2[GeTi_2W_9O_{37}]_2$ (16H$_2$O (PM-504), $[Me_2NH_2]_3\left[\left\{SiNb_7W_{67}O_{226}\right\}_2\right]$ (JM2820), $K_2[Ce(SiW_9O_{39})_2]26H_2O$ (JM1590), $K_2[BGa\left(H_2O\right)\left(SiW_9O_{39}\right)]15H_2O$ (JM2766), $K_2[P_3W_{12}(NbO)\O_4]$, $K_2[Fe_2(W_5O_{26})\cdot(PW_{6}O_{27})_2]$ (HS-058), and $K_2[P_2W_7NbO_{22}]$ demonstrated antiviral activity against influenza virus, HSV-1, HSV-2, HIV-1, and HBV. This kind of POM localizes on the cell surface, and nonspecifically masked cellular receptors for viral entry [32]. The mechanism of action of POM-4960 is inhibition of virus attachment and subsequently penetration [33]. POM with Keggin-type structures was shown to have activity against human immuno-deficiency virus types 1 and 2 [31,34,38], hepatitis simplex viruses [39], thymidine kinase deficient herpes simplex virus [38], togaviruses [31], measles virus [40], HCMV [38], parainfluenza virus [40], rhabdovirus [31], RSV [38,40,41] influenza viruses [40,41] arenaviruses [31] and other retroviruses [31]. The surface of the cell membrane is surrounded by negatively charged polymers. The virus is electrostatically attracted to the cellular membranes and uses the HA molecule to bind to a particular receptor [42]. Negatively charged POMs are believed to be disrupting the mechanism and binding the virions to their receptors non-specifically [33,43]. POMs with vanadium or titanium were shown to inhibit several RNA viruses, including Paramyxoviridae (respiratory syncytial virus), Flaviviridae (Dengue virus), Lentiviridae (human immunodeficiency virus type 1) and Orthomyxoviridae (influenza virus type A) [44]. The suppression of virus-cell interaction and syncytium formation between HIV-infected cells has been ascribed to the process of anti-HIV suppression for most POM compounds [31,37,45]. Similar chemicals blocked the binding of an HIV-specific gp120 monoclonal antibody to the gp120 protein [31], and the anti-HIV action was ascribed to POM binding of gp120, which prevented virus-mediated fusion and cell-to-cell transmission. Envelope glycoprotein GP120 is a glycoprotein exposed on the surface of the HIV envelope and is required for viral attachment to particular cell surface receptors and virus entry into cells. PM-1001 was shown to significantly inhibit the binding of viral gp120 antibodies [46]. The enveloped virus such as Ebola, HIV-1, Influenza and the SARS-CoV-2 enter to the cells through their spike-like glycoprotein [47]. One effective approach to fight SARS-CoV-2 infection is to prevent the virus from entering host cells by inhibiting the molecular machinery involved in this key phase [48-51]. Pradhan et al. [52] investigated the 2019-nCoV spike glycoprotein and discovered many intriguing results. This group discovered four separate inserts in the 2019-nCoV spike glycoprotein, which have not been found in any other COVID-19 case reported to date, and all four inserts in the

| No. | Amino acid | Amino acid atom | POM atom | Distance | Nature of interaction |
|-----|------------|-----------------|----------|----------|----------------------|
| 1   | ARG41      | H-Donor         | O(H-)    | 2.23     | Hydrogen Bond        |
| 2   | ARG41      | H-Donor         | O(H-)    | 2.48     | Hydrogen Bond        |
| 3   | ARG41      | H-Donor         | O(H-)    | 2.21     | Hydrogen Bond        |
| 4   | HIS57      | H-Donor         | O(H-)    | 2.43     | Hydrogen Bond        |
| 5   | THR62      | H-Donor         | O(H-)    | 2.09     | Hydrogen Bond        |
| 6   | HIS96      | H-Donor         | O(H-)    | 3.07     | Hydrogen Bond        |
| 7   | SER195     | H-Donor         | O(H-)    | 2.23     | Hydrogen Bond        |
| 8   | HIS57      | H-Donor         | O(H-)    | 2.93     | Hydrogen Bond        |
| 9   | THR61      | H-Donor         | O(H-)    | 3.12     | Hydrogen Bond        |
| 10  | HIS57      | Pi-Orbitals     | H(O-Donor) | 3.94 | Hydrogen Bond        |
Predicted bonds between interacting atoms of POM and ACE2-RBD.

| S. No. | Amino acid | Amino acid atom | POM atom | Distance | Nature of interaction |
|--------|------------|-----------------|----------|----------|----------------------|
| 1      | ASP30      | Negative         | W(Positive) | 4.31     | Electrostatic         |
| 2      | B:         | Negative         | W(Positive) | 5.59     | Electrostatic         |
| 3      | A:         | Negative         | W(Positive) | 5.02     | Electrostatic         |
| 4      | B:         | Negative         | W(Positive) | 4.44     | Electrostatic         |
| 5      | B:         | Negative         | W(Positive) | 5.52     | Electrostatic         |
| 6      | A:         | H-Acceptor       | O(H-Donor) | 3.24     | Hydrogen Bond         |
| 7      | A:         | H-Acceptor       | O(H-Donor) | 3.22     | Hydrogen Bond         |
| 8      | A:         | H-Acceptor       | O(H-Donor) | 3.01     | Hydrogen Bond         |
| 9      | A:         | H-Acceptor       | O(H-Donor) | 3.07     | Hydrogen Bond         |
| 10     | A:         | H-Donor          | O(H-Acceptor) | 2.67   | Hydrogen Bond         |
| 11     | A:         | H-Donor          | O(H-Acceptor) | 2.49   | Hydrogen Bond         |
| 12     | A:         | H-Donor          | O(H-Acceptor) | 2.62   | Hydrogen Bond         |
| 13     | A:         | H-Donor          | O(H-Acceptor) | 2.53   | Hydrogen Bond         |
| 14     | A:         | H-Donor          | O(H-Acceptor) | 2.45   | Hydrogen Bond         |
| 15     | A:         | H-Donor          | O(H-Acceptor) | 2.36   | Hydrogen Bond         |
| 16     | A:         | H-Donor          | O(H-Acceptor) | 2.10   | Hydrogen Bond         |
| 17     | A:         | H-Donor          | O(H-Acceptor) | 2.56   | Hydrogen Bond         |
| 18     | A:         | H-Donor          | O(H-Acceptor) | 2.58   | Hydrogen Bond         |
| 19     | A:         | H-Donor          | O(H-Acceptor) | 1.89   | Hydrogen Bond         |
| 20     | A:         | H-Donor          | O(H-Acceptor) | 2.15   | Hydrogen Bond         |
| 21     | B:         | H-Donor          | O(H-Acceptor) | 3.09   | Hydrogen Bond         |
| 22     | B:         | H-Donor          | O(H-Acceptor) | 2.86   | Hydrogen Bond         |
| 23     | A:         | H-Donor          | O(H-Acceptor) | 3.45   | Hydrogen Bond         |
| 24     | B:         | H-Donor          | O(H-Acceptor) | 3.30   | Hydrogen Bond         |

values of binding free energy authenticate the strong interaction of [SiW12O40]−4 with TMPRSS2, and ACE2-RBD. In other words, [SiW12O40]−4 interacts efficiently with TMPRSS2, and ACE2-RBD and it makes a stable complex with them. The nature of interactions, atoms involved in bonding with POM and bond lengths are shown in Tables 1 and 2. The amino acid residues of TMPRSS2 and ACE2-RBD that interacted with POM are demonstrated in Figs. 1 and 2. The POM + ACE2-RBD forms hydrogen bonds with twelve amino acids, which are Asp30, Ala387, Asn33, His34, Arg393, and Pro389, from ACE2 and Glu406, Arg403, Arg408, Glu409, Lys417 and Gly416 of the spike glycoprotein (RBD). Also, for the POM + ACE2-RBD system, there are electrostatic interactions between Asp30 from ACE2 and Asp30, Asp405, and Glu416 of the spike glycoprotein (RBD) and the POM. Six amino acids Arg41, His57, Thr62, His96, Ser195, and Thr61 are involved in the formation of hydrogen bonding between the POM and TMPRSS2. As can be seen, [SiW12O40]−4 has stronger interactions and makes a more stable complex with ACE2-RBD compared to TMPRSS2as revealed by its lower value of binding free energy. The crucial step for the fusion of SARS-CoV-2 and host cell membranes is the activation and cleavage of spike glycoprotein of SARS-CoV-2 by host protease TMPRSS2 [55,56]. Therefore, as reported already, we can inhibit cell entry of SARS-CoV by blocking the activity of TMPRSS2 [57]. The association of POMs with TMPRSS2 may restrict the functionality of the S1/S2 site in the spike glycoprotein [58], limiting the fusion of viral and human cellular membrane, and suggests that POMs may serve as SARS-CoV-2 cell entry inhibitors. Lung with the massive surface area and the vast distribution of ACE2 in human alveolar epithelial cells is the vulnerable target organ for SARS-CoV. Thus, the investigation of compounds that interact with ACE2, and hinder viral entry into the cells by blocking this receptor and halting transmissibility and pathogenicity is an important field for research. The docking method showed that the POM might prevent SARS-CoV-2/APE-2 interaction and viral entry because it had a high binding affinity at the ACE-2–RBD complex interface. Overall, computational studies indicate that POMs may limit CoV cell entry by decreasing the TMPRSS2 used by SARS-CoV-2 for spike glycoprotein priming and interrupting the interaction of SARS-CoV-2 spike protein with the human ACE-2 receptor. However, their systemic effects should be further examined in suitable ex vivo human organ culture or organoids, animal models, or clinical trials.

3.2. Molecular dynamics simulations

3.2.1. Radius of gyration (Rg)

The radius of gyration (Rg) of both TMPRSS2-POM and ACE2-RBD-POM complexes quantifies the molecule’s overall extension during a 50ns MD run (Figs. 3 and 4). A low Rg value demonstrates better structural entirety and folding treatment [59]. Throughout the 50ns MD simulation, two complexes maintain a stable mean Rg of 1.65 nm for 2OQ5-POM and 3.12 nm for 6LZG-POM. Likewise, the obtained Rg value for the unbound 2OQ5 and 6LZG was 1.66 and 3.13 nm. This further shows that the proteins gained more stability upon the binding of the ligand. During simulation, a slight enhancement in the Rg value of the 6LZG-POM complex was observed, indicating its structural integrity. The MD simulation results entirely support that POM forms stable complexes with 2OQ5 and 6LZG, indicating its inhibitory properties for the TMPRSS2 and ACE2-RBD receptor, respectively [60].

3.2.2. RMSD

RMSD analysis revealed insights and structural changes in the protein that confirm the protein’s stability and equilibrium during simulation. The RMSD plot of the backbone atoms for the 2OQ5, 2OQ5-POM and 6LZG, 6LZG-POM is shown in Figs. 5A and 6A, respectively. RMSD was calculated for the unbound proteins and proteins-POM structures that converged during the 50ns MD simulation. The average RMSD of 2OQ5-POM was found to be 0.37 Å, and for the unbound 2OQ5 was 0.39 Å. Likewise, the obtained average RMSD for 6LZG-POM and 6LZG was 0.36 Å, and 0.37, respectively. If the RMSD value is less than 1.5 Å, it is considered to be good and acceptable. But, with the value of more than 3 Å for RMSD, it is clearly rejected. RMSD values are used to find the stability of the receptors with and without ligands and also to study the conformational changes of the receptors [61]. The low RMSD values indicated that POM was stable in MD simulations with proteins [62].

3.2.3. RMSF

RMSF analysis was used to determine the flexibility of the total protein concerning its average structure. Low RMSF values demonstrated narrowed movements, whereas high RMSF values demonstrated increased flexibility [63]. Ligand binding poses energy, and interaction has a direct correlation with residual fluctuation (RMSF) values. The RMSF plots for 2OQ5, 2OQ5-POM and 6LZG, 6LZG-POM are shown in Figs. 5B and 6B, respectively. The RMSF values for the 2OQ5-POM and 6LZG-POM complexes were extremely low; as a result, they exhibited minimal movement, indicating that both complexes were stable. RMSF values were 0.30 and 0.29 Å on average for 2OQ5 and 2OQ5-POM during a 50 ns simulation. Likewise, the RMSF values for 6LZG and 6LZG-POM were 0.29 and 0.29 Å. During the simulation, it was observed that Lys187 in the loop region of 2OQ5 was more fluctuated than those in the alpha-helix and beta-sheet regions. This indicated that the protein...
remained stable throughout the 50ns simulation period [64]. Except for the loop region, the residues 102 and 562 in the alpha-helix region of 6LZG exhibited significant variation up to 0.40 Å and 0.45 Å, respectively. Overall, the RMSF values for both proteins indicated that the complexes 2OQ5-POM and 6LZG-POM were stable [65].

3.2.4. Hydrogen bond interactions

Hydrogen bond formation plays an important role in the stabilization of protein-ligand complex structure by minimizing the energy of the system. The hydrogen bond interactions of the complexes were calculated to validate the affinity of the POM to inhibit the proteins. Protein-ligand hydrogen bonding pattern were studied in bound POM with TMPRSS2 and ACE2-RBD. Figs. 7 and 8 show the number of hydrogen bonds.
bonds versus time during the simulation. The average H-bonding in 2OQ5-POM and 6LZG-POM was 9.05 and 17.41 during the 50 ns simulation. Overall, the H-bonding patterns in 2OQ5-POM and 6LZG-POM interactions showed an energetically favorable and stable complex formation [66].

3.3. MM-PBSA binding free energy

The average free binding energy of the 2OQ5-POM and 6LZG-POM was computed by a python script MmPbSaStat.py (Table 3). We computed the average free binding energy and its standard deviation/error of the files, which were obtained from g_mmpbsa. The binding free energy can acceptably illustrate the durability of the linking ligand receptor, which is an essential parameter of evaluation in drug discovery. The lesser the binding energy, the better is the binding of the ligand and protein [67]. Except for the polar solvation energy, the favorable bonds versus time during the simulation. The average H-bonding in 2OQ5-POM and 6LZG-POM was 9.05 and 17.41 during the 50 ns simulation. Overall, the H-bonding patterns in 2OQ5-POM and 6LZG-POM interactions showed an energetically favorable and stable complex formation [66].

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This study found that POMs may prevent COVs from entering the cells by blocking the host cell serine protease TMRPSS2, which SARS-CoV-2 uses for spike glycoprotein priming. They may also engage with cells by blocking the host cell serine protease TMPRSS2, which SARS-CoV-2 uses for spike glycoprotein priming. Because of the wide range of antiviral action, and based on our computational results, it is evident to consider POMs as a great source of new medicinally helpful drugs; consequently, we highly suggest that POMs be included as much as possible in new drug discovery screening programs. This may be especially important in the pursuit of new medications for treating COVID-19 illness, a severe and quickly spreading disease for which no medical treatments are currently available and are desperately required.

4. Conclusions

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