Short Lag Times for Invasive Tropical Plants: Evidence from Experimental Plantings in Hawai’i

Curtis C. Daehler

Department of Botany, University of Hawai’i at Manoa, Honolulu, Hawaii, United States of America

Abstract

Background: The lag time of an invasion is the delay between arrival of an introduced species and its successful spread in a new area. To date, most estimates of lag times for plants have been indirect or anecdotal, and these estimates suggest that plant invasions are often characterized by lag times of 50 years or more. No general estimates are available of lag times for tropical plant invasions. Historical plantings and documentation were used to directly estimate lag times for tropical plant invasions in Hawai’i.

Methodology/Principal Findings: Historical planting records for the Lyon Arboretum dating back to 1920 were examined to identify plants that have since become invasive pests in the Hawaiian Islands. Annual reports describing escape from plantings were then used to determine the lag times between initial plantings and earliest recorded spread of the successful invaders. Among 23 species that eventually became invasive pests, the average lag time between introduction and first evidence of spread was 14 years for woody plants and 5 years for herbaceous plants.

Conclusions/Significance: These direct estimates of lag times are as much as an order of magnitude shorter than previous, indirect estimates, which were mainly based on temperate plants. Tropical invaders may have much shorter lag times than temperate species. A lack of direct and deliberate observations may have also inflated many previous lag time estimates. Although there have been documented cases of long lag times due to delayed arrival of a mutualist or environmental changes over time, this study suggests that most successful invasions are likely to begin shortly after arrival of the plant in a suitable habitat, at least in tropical environments. Short lag times suggest that controlled field trials may be a practical element of risk assessment for plant introductions.

Introduction

During biological invasions, lag times occur when there is a delay between the time of a species’ introduction to a new region and the time when it begins to spread or invade. Lag times could be caused by ecological barriers, such as delayed arrival of a required mutualist [1]. Evolutionary factors, such as time required for recombination and adaptation, have been suggested as potential explanation for lag times [2]. Demographic factors such as Allee effects may also be important [3]. Time lags of more than 50 years between first introduction and subsequent invasion are often assumed to be common in plants [4]. In a statistical analysis estimating lag times for woody plants introduced to Europe, the average time between introduction and first spread (escape from cultivation) was estimated at 170 years for trees and 131 years for shrubs [5]. These lag times were inferred indirectly, based on the time difference between when the plants appeared on planting lists and when they appeared in a published flora or sometimes in unpublished floristic data. The use of heterogeneous historical information presents some limitations: actual dates of introductions are sometimes not known and correspondence between recorded dates of naturalization and actual dates of naturalization or start of spread are typically not known. Furthermore, estimates of lag times are coarse-scale when several decades or more pass in the publication of subsequent floras, and floras vary in comprehensiveness and criteria for including species. An analysis of lag times in Australian cultivated plants using planting lists to indicate date of arrival and herbarium specimens as evidence of naturalization estimated a mode time of 149 years between introduction and naturalization [6]. Analyses of herbarium records [7] can provide valuable information about lag times, but they suffer from limitations because many herbarium specimens are collected opportunistically, potentially long after first naturalization, and the collected plant’s status (i.e. planted or naturalized) is not always clear.

The common presumption of long lag times for invasions [8] requires testing through direct and frequent observations of the behavior of many introduced species over time. Wangen and Webster [9] point out that the invasion process can include multiple lag phases. For example, a patchy landscape and a long time to invader maturity can cause sporadic rates of spread long after an invasion has started. This study focuses on the lag time between introduction and the start of spread when plantings have been deliberately made in the immediate vicinity of a semi-natural environment. I used historic planting records from the Harold L. Lyon Arboretum (Honolulu, Hawai’i) and documentation of
spread from these plantings to directly determine lag times between planting and the start of spread among 23 plants that subsequently became invasive pests in the Hawaiian Islands.

**Results**

Among hundreds of introduced species that were deliberately planted at the Harold L. Lyon Arboretum, 23 species have become recognized as invasive pests in Hawai‘i (Table 1). For these invaders, Lyon's historical records allowed determination of the lag time between planting (Figure 1) and first evidence of spread (Figure 2). Most of the invasive plants (91%) were woody species, with an average lag time of 14 years (range 2–22 years). Among the two herbaceous invaders, the average time lag was 5 years (range 4–6 years). No invaders started to spread between 23 and 89 years after planting (Figure 2). This retrospective analysis does not allow detection of lag times greater than 89 years, but the lack of any invasive species that started to spread in a 66-year period (Figure 2) suggests that most invasions can be expected to start within a decade or two of first planting in disturbed habitats that are in the vicinity of appropriate natural or semi-natural vegetation. Based on field observations (Table 1), the woody invaders require 2–10 years to reach reproductive maturity in Hawai‘i, while the herbaceous invaders require 1–3 years; therefore, all of the invaders began to spread within the first few years after reaching maturity.

**Discussion**

Previously reported lag times [5,6] were an order of magnitude greater than those reported here. Findings reported here are from a tropical environment, while most prior estimates of time lags have come from temperate environments. It is possible that lag times are shorter in tropical environments where plants can grow year round, and where narrower temperature extremes impose fewer restrictions on establishment. Kowarik [5] noted that many of the species with long lag times in Europe were adapted to warmer climates and their recent spread may have been associated with warming of parts of Europe. In contrast, temperature is unlikely to restrict spread of most tropical invaders. Differences in methodology and circumstances could also explain the shorter lag times reported here. In this study, the initial planting occurred within disturbed but naturally recovering vegetation, in the immediate vicinity of natural or semi-natural environments. In contrast, when date of first introduction is recorded in gardens or urban areas, these locations may be isolated from appropriate natural habitats, hindering spread into natural areas and resulting in long lag times.

A third consideration is that previous estimates of lag times may have been inflated because they were not determined by direct or systematic observations. Naturalization dates determined from herbarium specimens often over-estimate time to naturalization because herbarium specimens are usually not collected in a systematic or regular fashion. Dates of introduction are subject to

| Invader          | Life form | Year first planted | Year first recorded escaping | Lag (years) | Years from planting to first reproduction 2 |
|------------------|-----------|--------------------|-------------------------------|-------------|-------------------------------------------|
| Acacia confusa   | woody     | 1922               | 1938                          | 16          | 2–4                                       |
| Angiopteris evecta | herb    | 1935               | 1939                          | 4           | 2–4                                       |
| Ardisia elliptica Thunb. | woody | 1920               | 1938                          | 18          | 2–3                                       |
| Ardisia vires Kurz | woody | 1979               | 1981                          | 2           | 2–3                                       |
| Cecropia obtusifolia Bertol. | woody | 1921               | 1932                          | 11          | 3                                         |
| Chrysophyllum oliviforme L. | woody | 1921               | 1943                          | 22          | 3–5                                       |
| Cinnamomum verum J. Presl | woody | 1921               | 1939                          | 18          | 3–5                                       |
| Citreolum pudicum L. | woody | 1931               | 1937                          | 6           | 2–3                                       |
| Clerodendrum macrostegium Schauer | woody | 1981               | 1983                          | 2           | 2–3                                       |
| Clusia rosea Jacq. | woody | 1927               | 1943                          | 16          | 4–8                                       |
| Falcataria moluccana (Miquel) Barneby & Grimes | woody | 1920               | 1934                          | 14          | 4–8                                       |
| Heliconia laterifolia Bert. | herb | 1931               | 1937                          | 6           | 2                                         |
| Leptospermum polygallofolium Salisb. | woody | 1923               | 1932                          | 9           | 2–4                                       |
| Macaranga mappa (L.) Muell.-Arg. | woody | 1922               | 1941                          | 19          | 2–4                                       |
| Macaranga tanarius (L.) Muell.-Arg. | woody | 1922               | 1939                          | 17          | 2–4                                       |
| Prunus grisea Kalkman | woody | 1985               | 1989                          | 4           | 3–4                                       |
| Psidium cattleisum Sabine | woody | 1920               | 1938                          | 18          | 3–4                                       |
| Scheflera actinophylcta (Endl.) H.A.T. Harms | woody | 1920               | 1937                          | 17          | 3–4                                       |
| Spathodea campanulata Beauv. | woody | 1920               | 1938                          | 18          | 3–4                                       |
| Sphaeropentis cooperi (Hook. ex F. Muell.) Domin | woody | 1927               | 1934                          | 7           | 2–3                                       |
| Syzygium cumini (L.) Skeels | woody | 1923               | 1943                          | 20          | 3–4                                       |
| Toona ciliata Roem. | woody | 1921               | 1941                          | 20          | 5–10                                      |
| Trema orientalis (L.) Blume | woody | 1923               | 1938                          | 15          | 4–8                                       |

1So far, recorded only as a pest on the arboretum grounds.
2Based on personal observations in Hawai‘i by the author.

doi:10.1371/journal.pone.0004462.t001

Lag Times for Tropical Plants

PLoS ONE | www.plosone.org 2 February 2009 | Volume 4 | Issue 2 | e4462
similar errors, but in some cases they are known more precisely. Analyses of planting lists, floras and herbarium records suggest that the likelihood of detection outside of cultivation, naturalization, and/or area of spread of introduced species increase with residence time [10] and planting frequency [11]. This general pattern is true by necessity, since it is impossible for naturalization or spread to occur instantaneously due to the time required to reach reproductive maturity. But such a pattern is also to be expected due to the nature of herbarium collections: species that have been in residence for a longer period of time, or that have been planted at a greater frequency (more points of naturalization) are more likely to be collected as herbarium specimens. These facts complicate estimates of time lags based on herbarium specimens. It is rare to find studies recording direct field observations of invaders during the early naturalization process, but when such observations have been made, lag times between introductions and the start of spread of an invader have often been short [12–14]. Similarly, in the present study, deliberate and frequent searches were made to document spread, and much shorter lag times for plant invasions are evident.

The lag time at the start on an invasion is sometimes defined in mathematical terms as the time required for an invader to reach exponential growth or spread after the first planting or spontaneous

---

**Figure 1.** Cecropia obtusifolia (A) and Falcataria moluccana (B) are examples of early plantings in the Manoa Valley that became invasive. Natural or semi-natural vegetation is visible in the background of both photos, less than 500 m from these plantings. Photos: E. Caum, 31 December 1922.
doi:10.1371/journal.pone.0004462.g001

**Figure 2.** Frequency distribution for time lags between first planting and start of spread in Manoa Valley (Honolulu, Hawai‘i) for plant species that became invasive. Records extend for 89 years.
doi:10.1371/journal.pone.0004462.g002
occurrence in an area [15]. To determine the lag time using this approach requires analysis of records over many time intervals [15,16], whereas the present study focused simply on the start of spread for species that became invasive pests. Defining lag time as the transition time to exponential growth is theoretically appealing; however, this lag time is difficult to measure because the exponential growth curve is expected to be exceedingly shallow at the start of exponential growth or spread [17, p. 178], and conclusions are likely to be sensitive to statistical methodologies and assumptions about observation intensity over time. In one of the only experimental studies to examine lag times in this way, Memmott et al. [18] released insect populations of different sizes and deliberately monitored their growth and spread over time. They found that although the smaller populations took longer to reach a given size (an apparent “lag”), the growth of all populations was entirely consistent with exponential growth. There were no unexplained lags. Additional detailed studies of this type are certainly needed. In plants, which commonly require several years to reach reproductive age, heterogeneous rates of exponential expansion are expected at least during the first few generations after arrival [9], so it is important to start with an appropriate null model for exponential growth before seeking alternate explanations for apparent lags.

Although in some cases long lag times are possible due to delayed introduction of a required mutualist [19], change in climate [20], or imposition of a new disturbance regime [21], the results reported here indicate that in all cases, species that became invasive pests began spreading within a few years after the initial plantings reached maturity in disturbed but naturally recovering vegetation. This finding, although from a single tropical region and involving a limited set of species, suggests that carefully controlled field trials [22–24] or “in ground” evaluation [25] could become a practical element of risk assessment for plant invaders. In Hawai‘i, many invaders could have been identified prior to becoming pests, through their spread within a few years after maturation. A field trial approach is not a substitute for pre-entry risk assessment [26,27]; rather, it could be most useful and feasible for potentially high-value plant introductions when conventional pre-entry risk assessments are indecisive.

Botanic gardens of all types have great potential to contribute to our understanding of plant invasions, as recently shown by Dawson et al. [28]. Botanical garden records provide a valuable resource documenting planting dates and locations, while observations of spread and plant limitations can be easily made for large numbers of plant species. Closer collaboration between ecologists and botanical garden staff would likely result in new insights into invasion ecology while also yielding new approaches and successes in plant conservation.

Materials and Methods

Background and Historic Records

Beginning in 1919, Harold Lyon, working under the auspices of the Hawaiian Sugar Planters Association (HSPA), directed the experimental planting of hundreds of introduced species in the Manoa Valley, near Honolulu, Hawai‘i [29]. The planting area consisted of 111 hectares of degraded tropical or sub-tropical wet forest and former sugar cane fields. Lyon’s objective was to identify species that could be used for reforestation, allowing restoration of degraded watersheds. He procured plant materials from around the world. Typically, 6–30 individuals of each species were planted as rows, blocks or as scattered individuals. Weeds were initially cleared from around the plantings but persistent clearing or cultivation was not practiced due to limited availability of labor, and perhaps because Lyon wanted to see how these plants would grow on their own. Subsequently, Lyon monitored the plantings and searched for evidence of their spread across the valley. He was particularly interested in identifying species that showed evidence of naturalization. From 1921 to 1950, Lyon and grounds manager Ed Caum produced annual reports to the HSPA detailing their findings and reporting annually on species that were escaping from plantings across the valley. Shortly thereafter, the site came under the management of the University of Hawai‘i and eventually it was formally named the Harold L. Lyon Arboretum [29]. Following Lyon’s observations, arboretum staff continued to make observations on naturalizing species, recording many of these observations in a database that is used to manage the arboretum grounds. In 2006, Dachler and Baker [30] made systematic surveys in order to identify additional naturalizing species. Only species that were deliberately introduced to the Manoa Valley by Lyon or by subsequent Lyon Arboretum staff were used for analyses because the precise dates of first planting for these species are known.

Invasive Pest Species

Invasive pest species were independently defined as species meeting at least one of the following two criteria, 1) species that have been listed as significant invaders in Hawai‘i based on information collected by Smith [31] or 2) species that have become troublesome weeds in the management of the Lyon Arboretum, as documented by efforts made by the staff to control these species [30]. Both of these criteria consider spread and perceived or actual negative impacts in defining invaders.

Lag Times

Among the species that were deliberately planted at the Lyon Arboretum and that subsequently where classified as invasive pests, I used Lyon’s annual reports to HSPA along with notes recorded in the Lyon Arboretum database to determine the time between first planting and first spread in Manoa Valley, as evidenced by naturalized seedlings. Immature plantings are incapable of producing seeds, and this will impose a minimum lag time on each species. To estimate this minimum time, I have reported personal observations on the time required for these species to reach first reproduction in Hawai‘i.

Acknowledgments

I thank Ray Baker for introducing me to historical records relating to plantings at the Harold L. Lyon Arboretum and for discussions of plant behavior at the Arboretum. Ray Baker, Petr Pyšek, Marcel Rejmánek and Dennis Hansen made helpful comments on an earlier version of this manuscript.

Author Contributions

Conceived and designed the experiments: CCD. Performed the experiments: CCD. Analyzed the data: CCD. Wrote the paper: CCD.

References

1. McKey D (1989) Population biology of figs: Applications for conservation. Experientia 45: 661–673.
2. Crooks J, Soule ME (1999) Lag times in population explosions of invasive species: causes and implications. In: Sandlund OT, Schei PJ, Viken A, eds. Invasive species and biodiversity management. Dordrecht: Kluwer. pp 103–125.
3. Taylor CM, Hastings A (2005) Allee effects in biological invasions. Ecology Letters 8: 985–990.
4. Hobbs RJ, Humphries SE (1995) An integrated approach to the ecology and management of plant invasions. Conservation Biology 9: 761–770.
5. Kowarik I (1995) Time lags in biological invasions with regard to the success and failure of alien species. In: Pyšek P, Prach K, Rejmaněk M, Wadle M, eds. Plant Invasions, general aspects and special problems. Amsterdam, The Netherlands: SPB Academic Publishers. pp 15–38.

6. Caley P, Groves RH, Barker R (2008) Estimating the invasion success of introduced plants. Diversity and Distributions 14: 196–203.

7. Barney JN (2006) North American history of two invasive plant species: phytogeographic distribution, dispersal vectors, and multiple introductions. Biological Invasions 8: 703–717.

8. Crooks JA (2005) Lag times and exotic species: The ecology and management of biological invasions in slow-motion. Ecocience 12: 316–329.

9. Wangen SR, Webster CR (2006) Potential for multiple lag phases during biotic invasions: reconstructing an invasion of the exotic tree Acer platanoides. Journal of Applied Ecology 43: 238–268.

10. Pyšek P, Jaroslík V (2005) Residence time determines the distribution of alien plants. In: Inderjit, ed. Invasive Plants: Ecological and Agricultural Aspects. Switzerland: Birkhäuser Verlag. pp 77–96.

11. Dehnen-Schmutz K, Touza J, Perrings C, Williamson M (2007) A century of the ornamental plant trade and its impact on invasion success. Diversity and Distributions 13: 527–534.

12. Broncano MJ, Vila M, Roada M (2005) Evidence of Pseudotsuga menziesii naturalization in montane Mediterranean forests. Forest Ecology and Management 211: 257–263.

13. Burdon JJ, Chilvers GA (1977) Preliminary studies on a native Australian Eucalypt forest invaded by exotic pines. Oecologia 31: 1–12.

14. Huang H, Zhang L (2007) A study of the population dynamics of Spartina alterniflora at Jiuhansha shoals, Shanghai, China. Ecological Engineering 29: 164–172.

15. Pyšek P, Prach K (1995) Plant invasions and the role of riparian habitats - a comparison of four species alien to central Europe. Journal of Biogeography 20: 413–420.

16. Pyšek P, Prach K (1995) Invasion dynamics of Impatiens glandulifera: A century of spreading reconstructed. Biological Conservation 74: 41–48.

17. Lockwood J, Hoopes M, Marchetti M (2006) Invasion Ecology. Oxford, UK: Blackwell Publishing. 280 p.

18. Memmott J, Craze PG, Harman HM, Syrett P, Fossey SV (2005) The effect of propagule size on the invasion of an alien insect. Journal of Animal Ecology 74: 50–62.

19. Richardson DM, Allsopp N, D’Antonio CM, Milton SJ, Rejmaněk M (2000) Plant invasions: the role of mutualisms. Biological Reviews 75: 63–93.

20. Walther G-R, Post E, Convey P, Menzel A, Parmesan C, et al. (2002) Ecological responses to recent climate change. Nature (London) 416: 389–395.

21. Beauchamp VB, Stromberg JC (2007) Flow regulation of the Verde River, Arizona encourages Tamarix recruitment but has minimal effect on Populus and Salix stand density. Wetlands 27: 381–389.

22. Patterson RR, Mack RN (2008) Potential distribution of the invasive tree Tsinca aethiops (Euphorbiaceae) in the United States: evaluating CLIMEX predictions with field trials. Global Change Biology 14: 813–826.

23. Mack RN (1996) Predicting the identity and fate of plant invaders: emergent and emerging approaches. Biological Conservation 78: 107–121.

24. Pierson EA, Mack RN (1996) The population biology of Bromus tectorum in forests distinguishing the opportunity for dispersal from environmental restriction. Oecologia (Berlin) 104: 519–525.

25. Jefferson L, Havens K, Ault J (2004) Implementing invasive screening procedures: The Chicago Botanic Garden model. Weed Technology. pp 1434–1440.

26. Daehler CC, Denslow JS, Anari S, Kuo H (2004) A risk assessment system for screening out invasive pest plants from Hawai‘i and other Pacific Islands. Conservation Biology 18: 360–368.

27. Virtue JG, Spencer RD, Weiss JE, Reichard SE (2008) Australia’s Botanic Gardens weed risk assessment procedure. Plant Protection Quarterly 23: 166–178.

28. Dawson W, Madshwa AS, Burdlem D, Hulme PE (2008) Assessing the risks of plant invasions arising from collections in tropical botanical gardens. Biodiversity and Conservation 17: 1979–1993.

29. Hartt CE, ed. Harold Lloyd Lyon, Hawaiian sugar botanist. Honolulu: Harold L. Lyon Arboretum, University of Hawaii. 110 p.

30. Daehler CC, Baker RF (2006) New records of naturalized and naturalizing plants around Lyon Arboretum, Mānoa Valley, O‘ahu. Bishop Museum Occasional Papers 86: 5–18.

31. Smith CW (1998) Pest Plants of Hawaiian Native Ecosystems. http://www.botany.hawaii.edu/faculty/cw_smith/aliens.htm. Accessed: 2008-11-07. (Archived by WebCite at http://www.webcitation.org/5cA1bfsxM).