EFFECT OF ARTHROSPIRA PLATENSIS AS A FOOD SUPPLEMENT AGAINST SODIUM FLUORIDE-INDUCED INTOXICATION ON SOFT TISSUES OF MALE WISTAR ALBINO RATS

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Received: 01 June 2017, Revised and Accepted: 19 July 2017

ABSTRACT

Objective: The objective of the present study was to explore the in vitro antioxidant and effect of hydroalcoholic extract of Arthrospira platensis (ASP) against sodium fluoride (NaF)-induced toxic effects on soft tissues (heart, liver, and kidney).

Methods: In vitro antioxidant activity was assessed using 1,1-diphenyl-2-picrylhydrazyl radical scavenging, 2,2’-azino-bis(3-ethylbenzothiazoline-6-sulphonic acid) radical scavenging, metal chelation, total antioxidant and reducing power assays, and total flavonoid and phenol content. In this study, 36 male Wistar albino rats were divided into six groups of six animals each. Animals were served as Group I - normal control, Group II - toxic control, and Groups III, IV, and V - treatment groups, which received hydroalcoholic extract of ASP at doses of 100, 200, and 400 mg/kg body weight (p.o), respectively. Group VI served as plant control received hydroalcoholic extract of ASP at a dose of 400 mg/kg body weight (p.o). All groups except Groups I and VI received NaF (100 ppm) through drinking water for 30 days.

Results: Results showed that intoxication of NaF caused significant elevation of serum biomarkers of the heart, liver, and kidney and altered tissue oxidative stress markers’ levels. Administration of hydroalcoholic extract of ASP significantly normalized elevated serum levels of heart (p<0.001), liver (p<0.001, p<0.05), and kidney biomarkers (p<0.001). Decrease in lipid peroxidation and increase in reduced glutathione and catalase enzyme levels in a dose-dependent manner were observed in soft tissues (p<0.001, p<0.01, and p<0.05).

Conclusion: The study revealed that ASP has good antioxidant and mitigative action against NaF intoxication on soft tissues.

Keywords: Arthrospira platensis, Oxidative stress, Serum biomarkers, Sodium fluoride.

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INTRODUCTION

Fluoride ion is one of the highly reactive, strongly electropositive ions which belong to halogen group. It is derived from fluorigene which is not available in free form in the nature but chemically combines with other elements to exist as ionic forms such as sodium fluoride (NaF), hydrogen fluoride, and aluminum fluoride. It abundantly, hastily, and submissively crosses into to the intestinal mucosa to interfere various major metabolic processes of the living system after oral consumption through drinking water [1]. The natural source of fluoride ion is soil rock, and other chief sources include beverages, food, medicines, dust in air, and various other industrial processes [2].

 Globally, around 25 nations’ (200 millions of people) ground water is highly contaminated with fluoride and is at a risk of fluorosis-related health problems [3]. In India, approximately 150,000 villages (65 millions of people) are at risk of fluorosis of various levels and have been reported as “fluoride levels above the noticeable limits (1.5 ppm),” particularly Andhra Pradesh, Telangana, Rajasthan, and Gujarat states are highly prone to fluorosis and related problems [4].

Fluoride provokes excessive formation of free radicals and might cause the imbalance between the production and function of enzymatic and non-enzymatic antioxidant levels [5]. The success rate for prevention and mitigation of treatment of fluorosis was limited to very few physicochemical (ion exchange or adsorbent techniques) and therapeutic methods [6].

Earlier research reports strongly suggested that natural diet with a rich source of ascorbic acid, minerals, vitamins, and secondary metabolites effectively minimizes and recovers fluoride-induced health problems [7]. Arthrospira platensis (ASP) is widely and easily available edible material in the world. It is commonly called as super food which might be due to rich source of nutrients (proteins, vitamins, and minerals) and phytochemical constituents (alkaloids, phenols, and terpenoids). It was proved as antiviral and antimicrobial [8], hepatoprotective [9], nephroprotective [10], cardioprotective [11], antiobesity [12], and reduces dental plaque [13] edible material. Therefore, the purpose of present study is to find out the alleviatory effects of hydroalcoholic extract of ASP leaves against NaF-induced intoxication of soft tissues in male Wistar albino rats.

METHODS

Collection and authentication of plant material
ASP freeze-dried powder was purchased from Parry Neutraceuticals, Division of EID Parry (India) Ltd, Chennai, Tamil Nadu, India, and authentication was done by Dr. Sunita Garg, Chief scientist, Raw Material Herbarium and Museum, Delhi (RHMD), CSIR-NISCAIR; Voucher specimens were stored in the department of Pharmacology, CMR College of Pharmacy, Hyderabad, Telangana, India.

Preparation of the extract
The ASP powder (100 g) was mixed with water:alcohol (30:70) in a round bottom flask and left for 7 days at room temperature with occasional shaking. After 7 days, the mixture was filtered with Whatman filter paper no. 1. The filtrate was evaporated in vacuo using rotary flash evaporator. About 16% of semi-solid yield was obtained. The extract was stored at 4°C for further studies.

Experimental animals
Male Wistar albino rats (36) weighing in between 220 and 250 g were procured from Sai Thirumala Enterprises, Hyderabad, India. The animals were acclimatized for 10 days before starting the experiment.
Rat feed was provided with water ad libitum and maintained a photoperiod of 12 h light/dark cycle. The study was completed as per the guidelines of CPCSEA, Government of India, after approval from IAEC (IAEC no: CPCSEA/1657/IAEC/CMRCP/PhD-15/40).

**In vitro antioxidant studies**

The 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical scavenging activity, 2,2’-azino-bis(3-ethylbenzothiazoline-6-sulphonic acid) (ABTS) radical scavenging activity, metal chelating assay, total antioxidant activity, reducing power assay, total flavonoid content, and total phenol content were estimated as per standard protocols [14].

**Acute toxicity studies**

Acute toxicity study was performed according to the guidelines of OECD 425.

**Experimental design**

The dose of NaF was selected based on the previous research studies [15]. After 10 days of acclimatization period, the experimental animals were divided into six groups (n=6) as follows:

- **Group I**: The animals received drinking water for 30 days
- **Group II**: NaF (100 ppm) through drinking water for 30 days
- **Group III**: ASP extract at an oral dose of 100 mg/kg bw/day + NaF (100 ppm) through drinking water for 30 days
- **Group IV**: ASP extract at an oral dose of 200 mg/kg bw/day + NaF (100 ppm) through drinking water for 30 days
- **Group V**: ASP extract at an oral dose of 400 mg/kg bw/day + NaF (100 ppm) through drinking water for 30 days
- **Group VI**: Plant control - ASP extract at an oral dose of 400 mg/kg bw/day for 30 days (p.o) alone.

At the end of the study, animals were fasted overnight and blood samples were collected from retro-orbital plexus. Blood samples were allowed to clot for approximately 1 hr at room temperature and centrifuged at 2500 rpm for 15 minutes to obtain the serum, used for estimation of various biochemical parameters such as lactate dehydrogenase (LDH), creatine kinase-MB (CK-MB), glucose, magnesium, serum glutamic oxaloacetic transaminase (SGOT), serum glutamic-pyruvic transaminase (SGPT), total direct bilirubin, total protein, albumin, total cholesterol (TC), triglycerides (TGs), high-density lipoprotein cholesterol (HDL-C), blood urea nitrogen (BUN), uric acid, and creatinine were estimated using coral kits and semi auto analyzer (Inkarp ES-100). In vivo antioxidant markers such as lipid peroxidation, reduced glutathione, and catalase were estimated as per standard protocols [16-18].

**Statistical analysis**

The values were expressed as mean±standard error of mean. The statistical analysis was carried out by one-way analysis of variance followed by post hoc Dunnett’s test using GraphPad prism 5.0. The values were statistically significant at p<0.05.

**RESULTS**

**In vitro antioxidant activity**

The inhibitory concentration 50% (IC\textsubscript{50}) values for DPPH radical scavenging activity were found to be 6.8 and 213.42 µg/ml for the standard Vitamin C and ASP extract, respectively. The IC\textsubscript{50} values for ABTS radical scavenging activity were found to be 14.1 and 520 µg/ml for the standard Vitamin C and ASP extract, respectively. The IC\textsubscript{50} values for metal chelation assay were found to be 76.19 and 320 µg/ml for the standard Vitamin C and ASP extract, respectively. Total antioxidant activity was found to be 247.09±0.94 mg Vitamin C equivalents per mg of plant extract. The reducing power ability was found to be 82.67±6.12 μg Vitamin C equivalents per mg of plant extract. Total flavonoid content was found to be 60.67±0.08 μg quercetin equivalents per mg of plant extract, and total phenol content was found to be 110.00±0.02 μg gallic acid equivalents per mg of plant extract.

**Acute toxicity studies**

Acute toxicity studies of ASP showed well tolerance at 2000 mg/kg bw p.o. All the animals in the treatment groups were normal in alertness and behavior up to 72 hrs of post-administration. No mortality was observed until completion of the study. Therefore, 1/20, 1/10, and 1/5 of this test dose were selected as low (100 mg/kg), medium (200 mg/kg), and high (400 mg/kg) doses, respectively, for screening the protective effects against NaF-induced toxicity.

**Effect on serum biomarkers of the heart, liver, and kidney**

**Effect on CK-MB and LDH**

Fluoride toxicity also elevates the levels of CK-MB and LDH enzymes by increase the formation of superoxide anions and hydroxyl radicals which cause oxidative damage to the cell membrane of myocardium. In the present study, high intake of fluoride increased the blood serum levels of CK-MB and LDH of Group II (p<0.001) when compared to Group I indicating cardiac toxicity. The increase in the levels of CK-MB and LDH enzymes after NaF intoxication was agreed with the results of Ibrahim and Abdel-Daim, 2013 [19]. No significant variation was found in the serum levels of CK-MB and LDH levels in ASP control (Group VI) when compared to Group I. Marked reduction in the serum levels of CK-MB and LDH was observed in a dose-dependent manner (p<0.001) after treatment with ASP in Groups III-V when compared to the Group II. All the values are summarized in Table 1.

**Effect on glucose and magnesium levels**

Increased blood serum glucose and decreased magnesium levels indicate diabetic properties of NaF intoxication [20, 21]. Blood serum glucose and magnesium levels were presented in Table 2. Significant increases in serum glucose and decrease magnesium levels were observed in Group II (p<0.001) when compared to Group I. No significant variation was found in serum glucose and magnesium levels in Group VI when compared to the Group I. ASP treatment showed significant reduction in the serum glucose level and recovered magnesium levels in a dose-dependent manner (p<0.001, p<0.05) when compared to the Group II.

**Effect on lipid profile**

Significant changes occurred in lipid profile (TC, TG, and HDL-C) after chronic exposure with fluoride, and the results of this study matched with observations of the previous research report of Vasant et al., 2014 [22]. Increase in the serum cholesterol and TG levels and decrease in HDL-C were observed in Group II (p<0.001) when compared to Group I. Group VI showed decrease in the serum cholesterol and TGs levels (p<0.01 and p<0.05) and a significant increase in the levels of HDL-C (p<0.001). Treatment with ASP showed a significant decrease in the serum cholesterol and TG levels and increase in HDL-C level in a dose-dependent manner (Groups III-V) (p<0.001 and p<0.05). All the values are presented in Table 2.

**Effect on SGOT, SGPT, total and direct bilirubin, and total protein and albumin levels**

The earlier research studies are strongly reported that NaF intoxication enhanced lipid peroxidation and decreased levels of GSH and antioxidant enzymes which are directly reflected by altered serum liver biomarkers [1]. High intake of fluoride in Group II increased the blood serum glucose and decreased magnesium levels indicate diabetic properties of NaF intoxication [20, 21]. Blood serum glucose and magnesium levels were presented in Table 2. Significant increases in serum glucose and decrease magnesium levels were observed in Group II (p<0.001) when compared to Group I. No significant variation was found in serum glucose and magnesium levels in Group VI when compared to the Group I. ASP treatment showed significant reduction in the serum glucose level and recovered magnesium levels in a dose-dependent manner (p<0.001, p<0.05) when compared to the Group II.

| Name of the group | CK-MB (IU/L) | LDH (IU/L) |
|-------------------|--------------|------------|
| Group I           | 214.1±35.00  | 282.2±15.5 |
| Group II          | 1214.12.95   | 192.2±11.7 |
| Group III         | 110.0±28.6   | 1413.4±5.6 |
| Group IV          | 946.8±48.11  | 1021.7±7.0 |
| Group V           | 495.3±46.33  | 766.0±59.2 |
| Group VI          | 165.4±12.04  | 347.2±16.3 |

Values are represented as mean±standard error of mean. Statistical analysis performed using one-way analysis of variance followed by post hoc Dunnett’s test. *p<0.01 versus NaF control; **p<0.01 versus normal control. NaF: Sodium fluoride, ASP: Arthuspira platensis, CK-MB: Creatine kinase-MB, LDH: Lactate dehydrogenase.
The kidneys are the major excretory organs for the excretion of fluoride from the body. Numerous studies clearly showed that kidney has a close correlation between fluoride intake and renal injury [23]. Effect on serum BUN, uric acid, and creatinine levels were presented in Table 4. There was a significant increase in the serum BUN, uric acid, and creatinine in Group II (p<0.001) when compared to Group I. Group VI showed a significant increase in serum levels of BUN and uric acid and creatinine when compared to the Group I. Treatment with ASP showed significant recovery from lipid peroxidation and increased levels of total protein and albumin in a dose-dependent manner (p<0.001, p<0.01, and p<0.05) when compared to Group II. All the values are summarized in Table 3.

**Effect on oxidative stress markers of heart, kidney, and liver**

Oxidative stress is a condition that indicates the imbalance between the prooxidants and antioxidants leading to the chemical injury to lipids, proteins, and DNA [15]. Excessive intake of fluoride also causes oxidative stress by induce the production of free radicals and decrease in biological activities of endogenous enzymatic and non-enzymatic antioxidant levels which play a central role in the elimination of free radicals and maintenance of antioxidant homeostasis. In addition, disturbs the metabolism of nucleic acids suppress the immune system, oxidation of macromolecules, membrane phospholipids breakdown, lipid peroxidation, and apoptosis. In this study, the administration of NaF in Group II (p<0.001) resulted in increased lipid peroxidation level with decrease in reduced glutathione and catalase levels of the heart, liver, and kidney. These findings have coincided with the results of earlier research reports [5,24,25]. Decreased level of lipid peroxidation and increased level of reduced glutathione (p<0.001) were observed in Group VI when compared to the Group I. Treatment with ASP showed significant recovery from lipid peroxidation and increased levels of reduced glutathione and catalase in a dose-dependent manner (Groups III-V) (p<0.001, p<0.01, and p<0.05) when compared to the Group II. All the values are presented in Table 5.

CONCLUSION

The present study suggesting that ASP was effectively alleviating the NaF-induced toxicity on soft tissues - heart, liver, and kidney. Further detailed studies are required to understand the exact mechanism involved in that. However, the present study was very useful and plays a significant additive role for the development of specific antidote against fluoride-induced intoxication.

ACKNOWLEDGMENT

The authors are thankful to UGC, New Delhi, India, for research grants. We also gratefully acknowledge Dr. K. Abbulu, Principal, CMR College of Pharmacy, for his support in successful completion of the research work.

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**Table 2: Effect of ASP on serum glucose, magnesium, and lipid profile levels against NaF-induced hepatotoxicity**

| Name of the group | Glucose (mg/dl) | Magnesium (mEq/L) | Cholesterol (mg/dl) | TG (mg/dl) | HDL-C (mg/dl) |
|-------------------|-----------------|-------------------|---------------------|-----------|----------------|
| Group I           | 10.70±0.34      | 4.02±0.03         | 99.17±3.82          | 64.4±2.55 | 31.05±0.80     |
| Group II          | 131.5±0.88d     | 2.11±0.02         | 186.3±3.58          | 88.1±1.99 | 21.09±0.68     |
| Group III         | 118.0±0.52      | 3.18±0.15         | 169.5±2.28          | 81.0±1.63 | 25.40±0.89     |
| Group IV          | 105.0±0.90      | 3.59±0.12         | 140.3±0.03          | 69.0±2.35 | 31.79±1.43     |
| Group V           | 93.7±1.69a      | 3.84±0.17         | 115.6±5.50          | 65.3±2.25 | 49.7±3.25      |
| Group VI          | 88.0±2.79a      | 4.19±0.28         | 85.5±2.43           | 52.5±1.28 | 57.10±2.23     |

Values are represented as mean±standard error of mean, n=6 in each group. Statistical analysis performed using one-way analysis of variance followed by Dunnett’s multiple comparison test.∗ p<0.001 and † p<0.05 versus NaF control; ‡p<0.001 and § p<0.01 versus normal control. NaF: Sodium fluoride, ASP: *Arthrosira platensis*, TG: Triglycerides, HDL-C: High-density lipoprotein cholesterol.

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**Table 3: Effect of ASP on liver serum biomarkers against sodium fluoride-induced hepatotoxicity**

| Name of the group | SGOT (IU/ml) | SGPT (IU/ml) | Total bilirubin (mg/dl) | Direct bilirubin (mg/dl) | Total protein (mg/dl) | Albumin (g/dl) |
|-------------------|--------------|--------------|------------------------|-------------------------|----------------------|---------------|
| Group I           | 31.7±1.71    | 21.7±1.65    | 1.25±0.48              | 0.14±0.01               | 7.9±0.15             | 2.46±0.08     |
| Group II          | 100.9±6.48a  | 109.9±11.43a | 4.04±0.19a             | 0.30±0.02               | 6.39±0.18          | 2.09±0.07     |
| Group III         | 91.34±4.86c  | 94.6±6.03e   | 3.46±0.13c             | 0.28±0.01               | 7.26±0.13c          | 2.59±0.16     |
| Group IV          | 67.99±3.13d  | 68.02±3.73d  | 2.65±0.18d             | 0.21±0.08               | 8.49±0.23d         | 3.84±0.26d    |
| Group V           | 43.73±3.53c  | 47.94±3.07   | 1.74±0.27c             | 0.17±0.05               | 9.13±0.21d         | 4.86±0.10d    |
| Group VI          | 33.48±3.16   | 40.47±1.68   | 0.88±0.07              | 0.15±0.08               | 9.75±0.30c         | 4.78±0.26c    |

Values are represented as mean±standard error of mean, n=6 in each group. Statistical analysis performed using one-way analysis of variance followed by Dunnett’s multiple comparison test.∗ p<0.001 and † p<0.05 versus NaF control; ‡p<0.001 and § p<0.01 versus normal control. NaF: Sodium fluoride, ASP: *Arthrosira platensis*, SGOT: Serum glutamic oxaloacetic transaminase, SGPT: Serum glutamic-pyruvic transaminase.

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**Table 4: Effect of ASP on serum BUN, uric acid, and creatinine levels against NaF-induced nephrotoxicity**

| Name of the group | BUN (mg/dl) | Uric acid (mg/dl) | Creatinine (mg/dl) |
|-------------------|-------------|------------------|-------------------|
| Group I           | 6.207±0.48  | 1.01±0.73        | 0.62±0.05         |
| Group II          | 15.15±0.84d | 1.99±0.02        | 1.45±0.06†        |
| Group III         | 16.47±0.76d | 1.59±0.08        | 1.05±0.08ad       |
| Group IV          | 19.14±0.77d | 2.16±0.12        | 0.77±0.05         |
| Group V           | 23.73±0.91d | 2.67±0.19d       | 0.52±0.09         |
| Group VI          | 24.80±2.23d | 2.73±0.20d       | 0.54±0.04         |

Values are represented as mean±standard error of mean. Statistical analysis performed using one-way analysis of variance followed by Dunnett’s test.∗ p<0.001 and † p<0.05 versus NaF control; ‡p<0.001 and § p<0.01 versus normal control. NaF: Sodium fluoride, ASP: *Arthrosira platensis*, BUN: Blood urea nitrogen.
### Table 5: Effect of ASP on lipid peroxidation, reduced glutathione, and catalase against NaF-induced intoxication on soft tissues

| Name of the group | Lipid peroxidation (µMol/mg of tissue) | Reduced glutathione (µMol/mg of tissue) | Catalase (µMol/mg of tissue) |
|-------------------|---------------------------------------|----------------------------------------|----------------------------|
|                    | Heart | Liver | Kidney | Heart | Liver | Kidney | Heart | Liver | Kidney |
| Group I            |       |       |        |       |       |        |       |       |        |
|                    | 2.38±0.35 | 4.21±0.63 | 4.21±0.38 | 21.34±1.71 | 14.2±2.66 | 10.59±0.56 | 1.53±0.16 | 2.09±0.17 | 1.1±0.04 |
| Group II           | 11.0±0.79 | 10.2±0.50 | 10.12±1.12 | 8.237±0.85 | 6.01±1.09 | 4.16±0.75 | 0.165±0.02 | 0.31±0.02 | 0.28±0.01 |
| Group III          | 9.34±0.27 | 9.79±0.20 | 9.288±0.22 | 7.235±0.61 | 7.33±0.33 | 5.96±0.57 | 0.200±0.02 | 0.56±0.04 | 0.30±0.02 |
| Group IV           | 6.46±0.34 | 6.51±0.39 | 6.685±0.39 | 15.89±0.66 | 13.5±0.62 | 11.24±0.76 | 0.36±0.02 | 1.70±0.28 | 0.47±0.03 |
| Group V            | 5.15±0.29 | 4.65±0.14 | 3.81±0.21 | 21.56±0.90 | 18.0±0.76 | 15.53±0.58 | 0.92±0.06 | 2.9±0.12 | 0.7±0.04 |
| Group VI           | 2.83±0.30 | 3.56±0.32 | 3.30±0.27 | 28.81±1.77 | 19.7±0.76 | 17.75±0.75 | 1.47±0.19 | 3.29±0.37 | 1.6±0±0.05 |

Values are represented as mean ± standard error of mean, n=6 in each. Statistical analysis performed using one-way analysis of variance followed by post hoc Dunnett’s multiple comparison test. *p<0.001 and *p<0.01 versus NaF control; #p<0.05 (95%); ##p<0.01, and ###p<0.001 versus normal control. ASP: *Arthrospira platensis*, NaF: Sodium fluoride

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