Dear Editor,

Acute promyelocytic leukemia (APL) is one of the most incident hematologic neoplasms in the acute care setting.¹ Treatment of APL consists of oral all-trans retinoic acid (ATRA), which has been extensively studied since 1990.² The pharmacological target of ATRA is the Retinoic Acid Receptor-Alfa (RAR-α) in malignant promyelocytes where it promotes their differentiation into mature myeloid lineages. Nowadays, 90% of patients achieve complete remission status in one to three months. Depending on risk classification, ATRA can be used as oral monotherapy, or combined with arsenic trioxide or anthracyclines.²

Despite the high rates of disease remission, APL is a medical emergency, as it causes a life-threatening coagulation disorder and respiratory insufficiency. The latter may be a result of differentiation syndrome (DS), which comprises 30% of all causes of deaths in the APL population.³

When patients are diagnosed with DS, they are often intubated to restore ventilation parameters. They also require the placement of an enteral tube to receive nutrients and medications.

So, how can we keep administering ATRA to these patients? In other words, the risk of occupational exposure to ATRA does not allow the nursing staff to manipulate such drugs by puncturing the capsule. Moreover, extracting the oily content from the hard-coated jelly capsules may lead to significant ATRA losses.⁴

In our institution, we have been admitting from two to eight patients per year with APL/DS to the emergency room and intensive care units. Because of the aforementioned ATRA handling problems, we have seen dose administration delays, unaware nursing technicians trying to manipulate ATRA (some of them in the fertile age) and pharmacy staff recurrently asking how to prepare this drug without losses and unnecessary occupational exposure. Unfortunately, there is not enough published information to support any kind of initiative, possibly due to the quick clinical evolution of APL and relatively low incidence in comparison to other neoplasms.⁵

In one of the most recent cases that we accompanied, we standardized a protocol to administer ATRA to intubated patients. A 22-year-old woman was admitted to the emergency room with dyspnea, dry cough and 85% oxygen saturation. She was diagnosed with high-risk PML and induction chemotherapy was promptly initiated: 50 mg/m² daunorubicin plus ATRA 45 mg/m²/day. Two days later, her ventilatory parameters worsened due to DS, so she was intubated and an enteral tube was inserted. As DS is an ATRA-related life-threatening condition, the retinoid was suspended for two days until her respiratory parameters improved.

When ATRA was reintroduced, the problem of administering it by enteral tube arose, as her oral access was no longer available. After conducting a literature review, we suggest the following procedure (Figure 1):

- ATRA was dissolved in 10 mL of distilled water (45°C), and 5 mL of mineral oil was added to work as a lipophilic carrier. A 5-mL dead space was left inside a 20-mL syringe;
  - We found that ATRA could be dissolved at higher temperatures without affecting its molecular structure.⁶⁻⁷ Mineral oil was chosen because the patient was already receiving it due to constipation secondary to narcotic agents;
- The mixture was thoroughly shaken until the entire coat dissolved;
  - Other authors have suggested that vigorous shaking should be applied to dissolve ATRA inside the syringe, which was observed by our pharmacy staff.³ It took more than 5 min of shaking to dissolve the intact capsules completely;
  - We also observed that cold temperatures did not remodel the carbohydrate polymer-based coat, so the solution maintained its colloidal physico-chemical properties;
- All procedures were performed in a class II-B2 biological safety cabinet and the final pharmaceutical product was protected from light (Figure 1). The shelf life was set at 24 h (extemporaneous solution) although, according to other authors, this can be extended⁶;
We provided written and verbal instructions to nurses and physicians for clinical monitoring (liver enzymes and blood cell count, respectively for toxicity and efficacy) and safe ATRA handling (use of personal protection equipment and correct discard).

After seven days in the intensive care unit, the patient had improved clinically and she was weaned from mechanical ventilation and sedatives. She progressed to consolidation therapy, which consisted in mitoxantrone and oral ATRA, and was discharged from the oncology ward.

Due to the clinical benefits and physical and chemical stability, there were many advantages with the administration of ATRA dissolved in water through an enteral tube.

A Japanese group reported that a few patients who received ATRA by enteral tubes had significantly lower plasmatic levels of retinoic acid derivatives. However, two case reports discussed ATRA diluted in a manner similar to that described herein where patients achieved complete remission and it was suggested that the drug was absorbed in the gastrointestinal tract. It is worth remembering that, as a cytotoxic agent, nursing staff must not manipulate it without proper protection and materials and that uncoated ATRA is photosensitive. Thus, the disparities between the two aforementioned reports might be due to lack of protection against light, which explains why authors observed only physiological retinoid levels in their patients. The oral administration of ATRA as a single 80 mg dose (~45 mg/m² usual dosage) provides 347 ± 266 ng/mL of peak concentration in one to two hours. By looking at the pharmacokinetics graph of the authors, if an ATRA dose was entirely administered, we should observe at least a minimal peak concentration near drug administration, and logarithmic elimination of ATRA.

Lastly, our experience is not absent of limitations and successful cases of ATRA administration through enteral tubes should be monitored regarding safety (control of the hypercoagulation state in APL, risk of bleeding and hepatotoxicity) and dosing requirements as different pharmacokinetic parameters may influence clinical response. ATRA is the mainstay of treating APL patients. Considering the benefits of maintaining retinoic-based therapy, the dissolving of coated capsules remains a valuable option for patients without oral access.

Conflicts of interest

There are no conflicts of interest.

Acknowledgements

On behalf of all Intensive Care Unit staff, we would like to thank Dr. Hipólito Carraro Jr. for his knowledge and outstanding clinical support to pharmacists, residents and patients.

References

1. Degos L. All-trans retinoic acid (ATRA) therapeutical effect in acute promyelocytic leukemia. Biomed Pharmacother. 1992;46(5–7):201–9.
2. Ades L, Sanz MA, Chevrier S, Montesinos P, Chevallier P, Raffoux E, et al. Treatment of newly diagnosed acute promyelocytic leukemia (APL): a comparison of French-Belgian-Swiss and PETHEMA results. Blood. 2008;111(3):1078–84.

3. Rogers JE, Yang D. Differentiation syndrome in patients with acute promyelocytic leukemia. J Oncol Pharm Pract. 2012;18(1):109–14.

4. Vesnoid® (all-transretinoic acid) capsules prescribing information. Produtos Roche Químicos e Farmacêuticos S.A., Inc (2013) [cited 29.12.15]. Available from: http://www.anvisa.gov.br/datavisaprodut/frmVisualizarBula.asp?pNuTransacao=1023952013&pldAnexo=1891818.

5. Montesinos P, Bergua JM, Vellenga E, Rayón C, Parody R, de la Serna J, et al. Differentiation syndrome in patients with acute promyelocytic leukemia treated with all-trans retinoic acid and anthracycline chemotherapy: characteristics, outcome, and prognostic factors. Blood. 2009;113(4):775–83.

6. Tan X, Meltzer N, Lindenbaum S. Determination of the kinetics of degradation of 13-cis-retinoic acid and all-trans-retinoic acid in solution. J Pharm Biomed Anal. 1993;11(9):81722.

7. Gatti R, Goia MG, Cavrini V. Analysis and stability study of retinoids in pharmaceuticals by LC with fluorescence detection. J Pharm Biomed Anal. 2000;23(1):147–59.

8. Muindi JRF, Frankel SR, Huselton C, DeGrazia F, Garland WA, Young CW, et al. Clinical pharmacology of oral all-trans retinoic acid in patients with acute promyelocytic leukemia. Cancer Res. 1992;52(8):2138–42.

9. Bargetzi M, Tichelli A, Gratwohl A, Speck B. Oral all-transretinoic acid administration in intubated patients with acute promyelocytic leukemia. Schweiz Med Wochenschr. 1996;126(November (45)):1944–5.

10. Takitani K, Nakao Y, Kosaka Y, Inoue A, Kawakami C, Kuno T, et al. Low plasma level of all-trans retinoic acid after feeding tube administration for acute promyelocytic leukemia. Am J Hematol. 2004;76(1):97–8.

Lucas Miyake Okumura *, Patrícia Carvalho Baruel Okumura, Cleni Veroneze
Hospital de Clínicas de Porto Alegre (HCPA), Porto Alegre, RS, Brazil
* Corresponding author at: Clinical Pharmacy Division, Hospital de Clínicas de Porto Alegre (HCPA), R Ramiro Barcelos 2350, Bonfim, 90035-903 Porto Alegre, RS, Brazil.
E-mail address: okumura.lucas@gmail.com (L.M. Okumura).

Received 27 May 2016
Accepted 4 November 2016
Available online 23 December 2016

1516-8484/
© 2016 Associação Brasileira de Hematologia, Hemoterapia e Terapia Celular. Published by Elsevier Editora Ltda. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).
http://dx.doi.org/10.1016/j.bjhh.2016.11.001