Individualized mechanical ventilation in a shared ventilator setting: limits, safety and technical details.

Michiel Stiers (✉ michielstiers@hotmail.com)  
St-Dimpna Geel  https://orcid.org/0000-0002-9175-8890

Matthias Mergeay  
St-Dimpna Geel

Hannah Pinson  
Vrije Universiteit Brussel

Luc Janssen  
St-Dimpna Geel

Evy Voets  
St-Dimpna Geel

Harald De Cauwer  
St-Dimpna Geel

Tom Schepens  
Antwerp University Hospital

Research Article

Keywords: COVID-19, ARDS, bench testing, individualized split ventilation

Posted Date: May 27th, 2020

DOI: https://doi.org/10.21203/rs.3.rs-31765/v1

License: ☀️ ☀️ This work is licensed under a Creative Commons Attribution 4.0 International License.  
Read Full License

Version of Record: A version of this preprint was published at Journal of Clinical Monitoring and Computing on October 6th, 2020. See the published version at https://doi.org/10.1007/s10877-020-00596-7.
Abstract

Background- The COVID-19 pandemic has resulted in an increased need for ventilators. The potential to ventilate more than one patient with a single ventilator, a so-called split ventilator setup, provides an emergency solution. Our hypothesis is that ventilation can be individualized by adding a flow restrictor to limit tidal volumes, add PEEP, titrate FiO 2 and monitor ventilation. This way we could ensure optimization of patient safety and clinical applicability. We performed bench testing to test our hypothesis and identify limitations.

Methods- We performed a bench testing in two lungs: 1) determine lung compliance 2) determine volume, plateau pressure and PEEP, 3) illustrate individualization of airway pressures and tidal volume with a flow restrictor, 4a) illustrate that PEEP can be applied and individualized 4b) create and measure intrinsic PEEP 4c-d) determine PEEP as a function of flow restriction, 5) individualization of FiO 2.

Results- The lung compliance varied between 13 and 27 mL/cmH 2 O. Set ventilator settings could be applied and measured. Extrinsic PEEP can be applied except for settings with a large expiratory time. Volume and pressure regulation is possible between 70-39% flow restrictor valve closure. Flow restriction in the tested circuit had no effect on the other circuit or on intrinsic PEEP. FiO 2 could be modulated individually between 0.21 and 0.8 by gradually adjusting the additional flow, and minimal affecting FiO 2 in the other circuit.

Conclusions- Tidal volumes, PEEP and FiO2 can be individualized and monitored in a bench testing of a split ventilator. In vivo research is needed to further explore the clinical limitations and outcomes, making implementation possible as a last resort ventilation strategy.

Introduction

The COVID-19 pandemic has resulted in an increased need for ventilators, as the amount of ventilated patients in some areas in the world have greatly surpassed the existing capacity.[1] It is unclear whether the demand can be met by increasing ventilator production, and local rapid surges in critically ill COVID-19 patients can result in local shortage. Furthermore, there is uncertainty whether developing nations will have the same resources to increase their ventilator capacity.

The potential to ventilate more than one patient with a single ventilator, a so-called split ventilator setup, has been proposed previously.[2-7] In its simplest form, the connected patients each get the exact same delivered pressures and inspiratory oxygen fraction. Even though this is a valuable idea in a crisis setting, and likely yields reasonable results in healthy lungs, multiple elements make this setup unlikely to be successful in ventilated patients with COVID-19 pneumonia. Furthermore, a split ventilator setup is strongly discouraged by a recent joint statement.[8]

Lung injury in COVID-19 often results in acute lung injury or acute respiratory distress syndrome (ARDS). [9] Mechanical ventilation in ARDS ideally needs to be individualized to the patient to prevent ventilator-
induced lung injury.[10] This includes titration of positive end-expiratory pressure (PEEP), limiting tidal volumes, and limiting inspiratory pressures. Patients would need to be matched so that the set respiratory support is suited for both patients, but this is not a feasible approach.

To facilitate a maximal degree of individualized respiratory support, we explored several adaptations to a split ventilator setup. We discuss the adaptations that can be made to individualize the respiratory support as much as technically possible, and discuss the remaining limitations.

We aimed to provide a technical overview of the last resort ventilation strategy: splitting the circuit to share a ventilator among two patients. It’s our hypothesis that it can be individualized by adding a flow restrictor to limit tidal volumes, adding PEEP, regulating FiO2 and monitor ventilation mechanics.[11] In this way we could ensure optimization of patient safety and clinical applicability. We performed bench testing to test our hypothesis and identify limitations.

**Methods**

**Basic shared ventilator setup and settings**

The design of a shared ventilator setup has been published before.[2] Briefly, the inspiratory limb of the ventilation circuit is split with an y-piece, with one subsequent limb going to each patient. The expiratory limbs are joined together again with an y-piece prior to their connection to the ventilator. To prevent flow from one patient to the other, and thus avoid cross-contamination, one-way valves can be added to the expiratory limbs of the circuit.[11]

The only way to safely ventilate patients in a basic shared ventilator setup is by delivering pressure-controlled ventilation. With volume-controlled ventilation, a sudden increase in airway resistance in one patient will result in large increases in delivered tidal volume for the other patient. Inspiratory time should be set, not variable.

Patients should be placed close together with the ventilator in between them, so that no additional deadspace (tubing) needs to be added. The one-way valves also reduce dead space and thereby prevent CO2 rebreathing.

In most modes of ventilation, the patient has the potential to interact with the ventilator. This should be avoided, by deep sedation or neuromuscular blockade. Ideally, inspiratory effort could be monitored to assure absence of spontaneous breathing activity. Another safety measure is that the threshold for the inspiratory trigger should be set as high as possible, so inspiratory effort does not lead to triggering the ventilator. Finally, should some effort still occur, the patient is more protected in the suggested pressure-controlled ventilation mode, avoiding tidal volume ‘steal’ from the other patient.

**Supplemental ventilator circuit components to individualize and monitor ventilation**
Even though the use of a shared ventilator without circuit modifications could be life-saving in some settings, there are a number of potential modifications to this setup that could make its applicability in clinical practice more feasible.

Monitoring respiratory dynamics becomes both more difficult and more important in a shared ventilator setup. A readily available pressure transducer, commonly used for invasive blood pressure monitoring, can be installed on each ventilator limb, with a fluid/water interface. This can be connected to the patient's monitor, thus displaying an additional specific curve for airway pressure. This pressure monitor can then also be used to measure intrinsic PEEP during an expiratory hold maneuver. Additionally, spirometry can be added as well, to measure individual tidal volumes, as the ventilator's output can only display the sum for both patients. Capnometry should be measured and displayed per patient.

- Limiting individual tidal volumes

The delivered tidal volume per connected patient will be a result of their individual lung compliance. This is the reason why ‘pairing’ of patients by lung compliance and size will need to be done prior to connecting a ventilator. However, this complicates the practical applicability of this setup, as patients would need to be shifted around, and possibly re-arranged when one would deteriorate or improve.

To overcome this issue, we can restrict the flow to a patient by partial obstruction of the inspiratory limb of the ventilator's circuit. (supplemental Fig 1) By this dynamic flow restriction, we can limit the tidal volume that is going to any patient, thus effectively removing the need to pair patients. The restriction needs to be done for the patient with the highest lung compliance.

The tidal volumes per patient can then be measured or calculated, and the pressure on the ventilator set so that the desired tidal volume is achieved for the patient with the least compliant lungs.[12] Subsequently, by restricting the flow to the patient with the more compliant lungs, the tidal volume is limited for that person not to exceed a set level.

- Setting individual PEEP levels

One of the major setbacks of using a single ventilator is that the PEEP level for both patients would need to be the same, whereas titration of PEEP is an important element of ventilator management in ARDS.[13, 14] This problem can be overcome by introducing a PEEP-valve in the circuit. When this valve is placed in-line, the released gas is returned to the ventilator, and the measured expired volume is still accurate. The PEEP valve can be placed on the expiratory limb of the circuit for the patient that needs the highest PEEP of both connected patients, the PEEP level for the other patient can then be set by the ventilator.

- FiO2

The inspiratory oxygen fraction (FiO2) remains a setting that is shared when you split a ventilator. As far as we know, no studies have yet tested the potential to individualize FiO2 in a shared ventilator setup. Our
hypothesis was that we could increase FiO2 in a patient's ventilator circuit by injecting a variable flow of 100% oxygen into the inspiratory limb of that patient. Figure 5 demonstrates this concept.

**Pitfalls and safety**

- **Intrinsic PEEP and need for long inspiratory times**

As we use pressure-controlled ventilation, it is important to make sure that the set airway pressure results in the desired alveolar pressure. In a dynamic situation, i.e. not during an inspiratory breath hold, an indication that this is actually present in patients is by creating a stable plateau pressure with a no-flow end-inspiratory result. Consequently, longer inspiratory times can be useful to achieve this stabilized distributed tidal volume. In ARDS, this could result in higher levels of intrinsic PEEP, and dynamic airway obstruction and expiratory flow limitation frequently result in air trapping.[15] Our bench setup was designed to see whether we could demonstrate the presence of intrinsic PEEP in the individual patient in a dynamic situation.

- **Recommended alarm settings and bias flow and estimation of compressible gas volume in circuit.**

Alarm settings are described in the ‘Columbia protocol’, developed in New York.[16] As stated in this protocol, the ventilator may misestimate compressible gas volume in circuit. A mismatch of approximately 80mL is reported for both tidal volumes and minute volume. Therefore ventilation should be checked by blood gas analysis.

- **Adding deadspace in acid-base differences between the two patients**

When a large difference in acid-base status is present between the two ventilated patients (one patient acidotic and one alkalotic), restoring one will aggravate the other. This can in theory be overcome by adding deadspace for the patient with a respiratory alkalosis, so that increased minute ventilation does not lead to increased alkalosis.

**Experimental setup**

We performed a bench testing with five experiments to: 1) determine lung compliance as function of the ventilator settings 2) determine volume, plateau pressure and PEEP in two lungs without added in-line PEEP or flow restriction 3) illustrate individualization of airway pressures and tidal volume with a flow restrictor, 4a) illustrate that PEEP can be applied and individualized 4b) create and measure intrinsic PEEP 4c) determine PEEP as a function of flow restriction 4d) and create PEEP with an additional in-line PEEP module 5) test our hypothesis that FiO2 can be individualized.

Several ventilator settings were applied (see Table 1). Two Dräger test lungs were used: lung 1 was a standard and another test lung was made less compliant, lung 2. The flow restrictor we used was a diaphragm valve with markings per quarter to adjust the flow. (See supplemental material S1)
balloon was ventilated in a pressure controlled mode, inspiratory pressure was always 20cm H$_2$O above PEEP.

The ventilators used in our experiments are a Puritan Bennett 980 Series and an AS3/ADU anesthesia machine, all with standard 22mm tubes. The use of two different types of ventilators was due to clinical needs.

Pressure transducers were obtained from the transpac disposable pressure transducer and connected to a monitor. We used an additional monitor to display the pressure curves in mmHg from our transducer from circuit A and B. The transducers were equilibrated, prior to each experiment. We used a Datex Ohmeda gas analyzer spirometry module for an AS3/ADU anesthesia machine, with oximetry. To introduce expiratory obstruction in experiment three we used a gradual clamp.

**Different bench tests**

1) Determining the compliance as function of the ventilator setting

The lung compliance was measured for our test lung as function of the different ventilator settings with the anesthesia ventilator in a normal circuit of 1 test lung.

2) Determining volume, plateau pressure and PEEP in two lungs whitout in-line PEEP or flow restriction.

Using the anesthesia ventilator. (Fig1) the volumes were measured via our spirometry module on the inspiratory limb of circuit A and B and were displayed respectively on the anesthesia machine in mL for A and on the monitor in mL for B. The plateau pressure and total PEEP were displayed on and could be read from the monitor in mmHg, for the different ventilator settings.

3) Measurement of the volumes and plateau pressures as a function of the flow restrictor.

Using the same set-up as in experiment 2, the flow restrictor was added on the inspiratory limb of circuit A and adjusted from totally closed per quarter to fully open. (Fig 2) We measured the volume and plateau pressure as described above using ventilator settings 1 and 5.

4a) illustrate that PEEP can be applied and individualized

Using the same set-up as in experiment 2, we applied the in-line PEEP 7.5 cmH2O in circuit A as a variable. (Fig 3). The total PEEP was displayed on and could be read from the pressure curve on the monitor in mmHg. The experiment was done with ventilator settings 1 and 5 and with ventilator settings 1 and 5 without extrinsic PEEP. We did three runs with the in-line PEEP as a variable: 1) no in-line PEEP, 2) no in-line PEEP and 8cmH2O extrinsic PEEP on the ventilator and 3) with in-line PEEP 7.5cmH2O in circuit A.

4b) Applying intrinsic PEEP and measurement of the total PEEP to determine intrinsic PEEP.
For this experiment we used the Bennett ventilator, with an in-line PEEP 7.5cmH2O on circuit A. (Fig 4a) Obstruction on the expiratory limb from circuit A was simulated with a clamp adjusted from fully open to totally closed, to induce intrinsic PEEP. The total PEEP was measured, by performing an expiratory hold maneuver, as a function of flow obstruction in the expiratory limb. The intrinsic PEEP was calculated in cmH2O as follows: (measured total PEEP in mmHg *1.36)-[(extrinsic PEEP from ventilator in cmH2O)+(7.5cmH2O in-line PEEP)]. The experiment was done with the different ventilator settings.

4c) Measurement of the total PEEP on the ventrilo, in circuit A and B and as function of the flow restriction to determine intrinsic PEEP in both circuits.

We used the same set-up as experiment 2, the flow restrictor was added on the inspiratory limb of circuit A and adjusted from totally closed per quarter to fully open. (Fig 5a) We measured the total PEEP as described above using ventilator settings 1 and 5.

4d) Experiment 4c with an additional in-line PEEP.

We used the same set-up as experiment 4c with an additional in-line PEEP. (Fig 6a)

5) Measurement of the FiO2 % in circuit A and B as a function of added flow in circuit A.

We used the anesthesia ventilator in the standard set-up with test lung 1 on both circuits. (Fig 7a) We added a laterally inserted varying flow (C) in two different positions and measured the volume and FiO2 at points A and B in the circuits of the same name. The additional flow was inserted at two points on the inspiratory limb of circuit A. Point 1, just after the splitting of the inspiratory limbs on circuit A and point 2 just before the entrance of the lung on circuit A. Ventilator setting 1 was used to perform the test with a flow of 15L/min and FiO2 of 0.21. We waited one minute per measurement to record the values.

Results

1) Determining the compliance as function of the ventilator setting

The lung compliance for our test lung varied between 13 mL/cmH2O (setting 5) and 27 mL/cmH2O (setting 3) for the different ventilator settings.

2) Determining volume, plateau pressure and total PEEP in two lungs whitout in-line PEEP or flow restriction.

In both circuits we see almost equal values per ventilator setting for plateau pressures, total PEEP and volume. The set extrinsic (applied) PEEP is approximated for all settings, except for setting 4 with a larger difference. This setting has a large expiratory time of 5.6 seconds. We observed the same with the plateau pressures, for all ventilator settings is this the inspiratory pressure of 20cmH2O above the set extrinsic (applied) PEEP.

3) Measurement of the volumes and plateau pressures as a function of the flow restrictor
With the flow restrictor that we have used, there is a possibility to regulate volume and pressure between 70% and 39% valve closure in ventilator setting 1 and between 70%-48% in ventilator setting 5. (Fig 2b)

The volume curve in ventilator setting 1 has a gradual increase in volume. Ventilator setting 5 has a steep increase and yields lower volumes with an open valve as a result of higher PEEP.

The same results were obtained when applying flow restriction in circuit A or circuit B. We noticed that any change made with the flow restrictor in the tested circuit had no effect on the other circuit, in terms of volumes or plateau pressures.

4a) illustrating that PEEP can be applied and individualized

In-line PEEP was only applied in circuit A. Any change made with the in-line PEEP in the tested circuit A had no effect on circuit B in terms of plateau pressure, total PEEP and volume. PEEP can be applied in a circuit and individualized without affecting the other circuit. (Table 2) The ventilator used for this experiment had a minimum extrinsic PEEP of 3cmH2O.

Parallel to the findings from experiment 2, we were not able to reach the set peep for high extrinsic PEEP settings. Adding an in-line PEEP of 7.5 cmH2O with the ventilator set at zero PEEP gives a net resulting PEEP of 10cmH2O. This is explained because of the minimum extrinsic PEEP of 3cmH2O given by the ventilator in any circumstance. Adding an in-line PEEP of 7.5cmH2O is cumulative for ventilator setting 1, with low extrinsic PEEP. This cumulative effect is not measured with setting 5, with a high set extrinsic PEEP (20.4cmH2O). These findings illustrate that applying additional in-line extrinsic PEEP is possible and results in cumulative PEEP values when ventilator PEEP is set low.

An extrinsic PEEP of 8cmH2O applied by the ventilator resulted in a tidal volume of 480 mL. However, when we apply this extrinsic PEEP via an in-line PEEP of 7.5cm H2O, with the ventilator set at zero PEEP, we only see a tidal volume of 370ml. When we combine extrinsic PEEP from the ventilator 5cmH2O and in-line PEEP 7.5cmH2O, we measure a tidal volume of 250mL and a total PEEP of 13.6cmH2O. When applying extrinsic PEEP via an in-line PEEP, there are lower tidal volumes measured in that circuit. As illustrated in experiment 2, ventilator 5 has lower tidal volumes. However, we also observed with this setting that an in-line PEEP causes a lower tidal volume than if we applied this extrinsic PEEP via the ventilator.

4b) Applying intrinsic PEEP and measurement of the total PEEP to determine intrinsic PEEP in two lungs with different compliance.

Fig 4b shows the measured total PEEP and corresponding intrinsic PEEP as function of the obstruction in the expiratory limb. Circuit A has an extrinsic PEEP of 7.5cm H2O from the in-line PEEP, and ventilator setting 2 of 5 cmH2O. Theoretically, this means an extrinsic PEEP in circuit A of 12.5 cmH2O. We illustrated the possibility to induce intrinsic PEEP by applying flow restriction on the expiratory limb and measure total PEEP with calculation of the intrinsic PEEP with the theoretically extrinsic PEEP.
4c and d) Measurement of the total PEEP on the ventilator, in circuit A and B and as function of the flow restriction to determine intrinsic PEEP in both circuits.

In experiment 3 we illustrated that pressures can be regulated by a flow restrictor. When plateau pressures are registered from a valve closure of 70%, total PEEP can be measured. (Fig 5b and 6b) In experiment 4b we illustrated that intrinsic PEEP can be applied and calculated from the measured total PEEP. When we calculated the intrinsic PEEP for ventilator settings 1 and 5 in circuit A and B, it was negative. We have already described the discrepancy between high applied extrinsic PEEP and the effectively measured PEEP. Therefore, we calculated the intrinsic PEEP for ventilator setting 5 with both the theoretical and the measured extrinsic PEEP (experiment 4a), however both turned out to be negative.

In addition, we placed an in-line PEEP in the expiratory limb, but no intrinsic PEEP could be demonstrated.

5) Measurement of the FiO2 % in circuit A and B as a function of added flow in circuit A.

During this experiment we noticed a variable FiO2 over time at point A and B. Therefore we measured the lower and upper limit, and calculated the median. We plotted this median for point A and B for position 1 and 2 as a function of the added flow. (FIG 7b)

Our hypothesis stated that we can increase the FiO2 in a circuit by adding an additional flow of 100% into the inspiratory limb of that circuit. With ventilator setting 1, a flow of 15 L/min and 21% FiO2 we were able to adapt FiO2 in circuit A and minimally affecting the FiO2 in the other circuit. (Fig 7b) FiO2 can be individually adjusted between 0.21 and 0.80 by gradually adjusting the additional flow. The position in the circuit of the added flow had no influence on the measurements.

Discussion

We aimed to individualize ventilation parameters in a split ventilator setup by adding a flow restrictor to limit tidal volumes, adding PEEP, in-line O2 and monitor ventilation mechanics. This bench study was designed to test the limitations of this setup.

We illustrated that with a flow restrictor we can adapt tidal volumes and pressure in a circuit. In clinical situations this would be the circuit of the patient with the most compliant lung. We used a diaphragm valve and noticed a short adaptation interval. This needs further improvement to maximize the range of adjustments, to optimize individualized ventilation. Flow restriction in one circuit has no effect on tidal volumes or airway pressure in the other circuit.

We illustrated in experiment 3 that intrinsic PEEP can be applied by a flow restriction in the expiratory limb and can be measured. Individualization of PEEP was possible as well. Applying in-line PEEP of 7.5 cmH2O with ventilator settings with low extrinsic PEEP (settings 1-3, 6.8 cmH2O) has a cumulative course, while additional in-line PEEP with high extrinsic PEEP (settings 4-5, 20.4 cmH2O) has no cumulative course, but presumably sigmoidal. Applying PEEP in one circuit has no effect on the other
circuit. We observed lower tidal volumes at high added extrinsic in-line PEEP settings, applying PEEP is at the expense of ventilation. Augmenting the driving pressure is needed to keep tidal volumes the same. Combining the flow restrictor and in-line PEEP valve in one part of the circuit seems unlikely to be a realistic clinical scenario, as the patient with the highest compliance will get the flow restrictor, but probably also needs the lowest PEEP.

We tested the ventilator settings with high extrinsic PEEP to measure intrinsic PEEP as function of flow restriction and found that we could not measure intrinsic PEEP. Because we suspect a sigmoidal curve for additional in-line PEEP with high extrinsic PEEP ventilator settings, we both tested with calculated and measured extrinsic PEEP to measure intrinsic PEEP. However, we were unable to demonstrate intrinsic PEEP for ventilator settings 4 and 5 for both.

One of the reasons why patients still needed to be ‘paired’ was on the basis of needed FiO$_2$. We illustrated that by an additional side stream of O$_2$ the FiO$_2$ can be adapted with minimal impact on the other circuit. We noticed a varying FiO$_2$ with a minimum and maximum value within the time of observing, for the various additional flows. The fluctuation of the measured FiO$_2$ is probably due to incomplete mixing of the added O$_2$ within the inspiratory limb of the circuit during standstill of the gases in expiratory phase. Pressure build-up could be suspected when laterally inserting additional O$_2$ flow behind the flow restrictor (C2) when closed or partially closed. Therefore we recommend to do this before the flow restrictor (C1).

Our tests have several limitations. The bench testing was performed with material available in a small regional hospital with changing clinical needs, not with material from a professional lab. This implies possible test errors, which would also be present in a normal clinical environment. Pressures in cmH$_2$O were displayed on the ventilator and in mmHg on the monitor, original data was converted to cmH$_2$O, which makes this setup more difficult to use. However, this demonstrates broad clinical applicability, also in non-academic centers.

The Joint Statement on Multiple Patients Per Ventilator states that with the current available resources and knowledge, it is impossible to share ventilators.[8] During the introduction and with the performed bench testing we provide answers to uncertainties that are discussed in the Joint Statement [8, 17]. Likewise, the FDA recently the FDA approved a Y-connector intended to facilitate ventilator sharing, under an FDA device Emergency Use Authorization, but still needed patients pairing without individualization.

To our knowledge no other research groups has described an experiment to create and measure intrinsic PEEP, nor are there any reports known to us about individualization of FiO$_2$.

This bench testing was a proof of concept that individualized mechanical ventilation in a shared ventilator setting is feasible with readily available equipment in a regional hospital. Further research should be carried out into the individualization of FiO$_2$ and the effect of additional PEEP and tidal volumes; an in vivo testing in an animal model is the next step.
Declarations

Acknowledgments

We would like to thank Mr. Luc Geens from our technical staff for the technical support and Ir. Philippe Caers and Ir. Dirk Wenmakers for the technical support and advice. We would like to thank Mr. Lucas Caesens for the artwork of our set-up figures.

Funding: The authors received no specific funding for this work. H.P. acknowledges a fellowship from the Research Foundation Flanders (FWO-Vlaanderen) under Grant No. 11A6819N.

Conflicts of interest/Competing interests: Conflict of Interest: The authors declare that they have no conflict of interest.

Authors’ contributions

All authors contributed to the study conception and design. Material preparation, data collection and analysis were performed by Michiel Stiers, Matthias Mergeay, Hannah Pinson, Luc Janssen and Tom Schepens. The first draft of the manuscript was written by Michiel Stiers and Tom Schepens and all authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.

References

1. Coronavirus disease 2019 (COVID-19) pandemic: increased transmission in the EU/EEA and the UK – seventh update, 25 March 2020. Stockholm: ECDC; 2020.

2. Neyman, G. and C.B. Irvin, A single ventilator for multiple simulated patients to meet disaster surge. Acad Emerg Med, 2006. 13(11): p. 1246-9.

3. Branson, R.D. and L. Rubinson, A single ventilator for multiple simulated patients to meet disaster surge. Acad Emerg Med, 2006. 13(12): p. 1352-3; author reply 1353-4.

4. Branson, R.D. and L. Rubinson, One ventilator multiple patients—what the data really supports. Resuscitation, 2008. 79(1): p. 171-2; author reply 172-3.

5. Paladino, L., et al., Increasing ventilator surge capacity in disasters: ventilation of four adult-human-sized sheep on a single ventilator with a modified circuit. Resuscitation, 2008. 77(1): p. 121-6.

6. Paladino, L., et al., Reply to Letter: One ventilator multiple patients—What the data really supports. Resuscitation, 2008. 79(1): p. 172-173.

7. Branson, R.D., et al., Use of a single ventilator to support 4 patients: laboratory evaluation of a limited concept. Respir Care, 2012. 57(3): p. 399-403.

8. The Society of Critical Care Medicine (SCCM), A.A.f.R.C.A., et al. Joint Statement on Multiple Patients Per Ventilator. 2020
9. Gattinoni, L., et al., *Covid-19 Does Not Lead to a "Typical" Acute Respiratory Distress Syndrome*. Am J Respir Crit Care Med, 2020.

10. Henderson, W.R., et al., *Fifty Years of Research in ARDS. Respiratory Mechanics in Acute Respiratory Distress Syndrome*. Am J Respir Crit Care Med, 2017. 196(7): p. 822-833.

11. Pinson, H., et al. *A better way of connecting multiple patients to a single ventilator*. [website] 2020; Available from: https://medium.com/@pinsonhannah/a-better-way-of-connecting-multiple-patients-to-a-single-ventilator-fa9cf42679c6.

12. Plummer, A.R., et al., *The BathRC model: a method to estimate flow restrictor size for dual ventilation of dissimilar patients* medRxiv 2020.04.12.20062497; doi: https://doi.org/10.1101/2020.04.12.20062497.

13. Sahetya, S.K., E.C. Goligher, and R.G. Brower, *Fifty Years of Research in ARDS. Setting Positive End-Expiratory Pressure in Acute Respiratory Distress Syndrome*. Am J Respir Crit Care Med, 2017. 195(11): p. 1429-1438.

14. Sahetya, S.K., J. Mancebo, and R.G. Brower, *Fifty Years of Research in ARDS. Vt Selection in Acute Respiratory Distress Syndrome*. Am J Respir Crit Care Med, 2017. 196(12): p. 1519-1525.

15. Aguirre-Bermeo, H., et al., *End-inspiratory pause prolongation in acute respiratory distress syndrome patients: effects on gas exchange and mechanics*. Ann Intensive Care, 2016. 6(1): p. 81.

16. Beitler, J.R.M., MPH1, et al. *Ventilator Sharing Protocol: Dual-Patient Ventilation with a Single Mechanical Ventilator for Use during Critical Ventilator Shortages*

17. Farkas, J. *PulmCrit wee – Why the SCCM/AARC/ASA/APSF/AACN/CHEST joint statement on split ventilators is wrong*. 2020; Available from: https://emcrit.org/pulmcrit/pulmcrit-wee-why-the-sccm-aarc-asa-apsf-aacn-chest-joint-statement-on-split-ventilators-is-wrong/.

### Tables

| Setting | PEEP (cmH2O) | I:E | Respiratory rate |
|---------|--------------|-----|-----------------|
| 1       | 5            | 1:2 | 14              |
| 2       | 5            | 1:1 | 30              |
| 3       | 5            | 1:1 | 8               |
| 4       | 15           | 1:3 | 8               |
| 5       | 15           | 1:3 | 30              |

Table 1 Different ventilator settings used in the bench testing. All settings are in pressure cycled ventilation with an inspiratory pressure of 20cmH2O.
Table 2. Illustration that PEEP can be applied and individualized. Experiment set-up is illustrated in supplemental figure 3. The in-line PEEP in the expiratory tract of circuit A was the variable in this experiment, the Vent ePEEP represents the asked PEEP from the ventilator. PEEPex is the theoretical ePEEP from the ventilator and in-line PEEP. PEEPtot is the measured PEEP in circuit A. PEEPint is the calculated intrinsic PEEP.

Supplemental Figure Legend

Supplemental figure 1. Diaphragm valve we used in the experiments.

Figures
Figure 1

Determining volume, plateau pressure and PEEP in two lungs without in-line PEEP or flow restriction. The Heat and Moisture Exchanger Filter (HMEF) is indicated and the pressure and spirometry module is colored yellow.
Figure 2

Measurement of the volumes and plateau pressures as a function of the flow restrictor. A) Set-up used for this experiment with the flow restrictor on the inspiratory limb of tract A. B) Tidal volume in red and plateau pressure in blue as function of the valve closure for setting 1 (circles) and setting 5 (triangles).
Figure 3

Illustration that PEEP can be applied and individualized. The in-line PEEP in the expiratory tract of circuit A was the variable in this experiment. The Heat and Moisture Exchanger Filter (HMEF) is indicated and the pressure and spirometry module is colored yellow.
**Figure 4**

Applying intrinsic PEEP and measurement of the total PEEP to determine intrinsic PEEP. A) Set-up used for this experiment with the flow restrictor and in-line PEEP on the expiratory limb of trajectory A to induce intrinsic PEEP. B) Intrinsic PEEP as function of the applied flow restriction in the expiratory limb for the different ventilator settings, displayed with their expiratory time.

**Figure 5**

Measurement of the total PEEP on the ventilator, in circuit A and B and as function of the flow restriction to determine intrinsic PEEP in both circuits. A) Set-up used for this experiment with the flow restrictor on the inspiratory limb of trajectory A. B) Total PEEP as function of valve closure for ventilator settings 1 and 5 displayed with their inspiratory times.
Figure 6

Measurement of the total PEEP on the ventilator, in circuit A and B and as function of the flow restriction with an additional in-line PEEP to determine intrinsic PEEP in both circuits. A) Set-up used for this experiment with the flow restrictor on the inspiratory limb and the in-line PEEP on the expiratory limb of circuit A. B) Total PEEP as function of valve closure for ventilator settings 2 and 5 displayed with their inspiratory times.

Figure 7

Measurement of the FiO2 % in circuit A and B as a function of added flow in circuit A. A) Set-up used for this experiment with the laterally inserted varying flow (C) in two different positions C1 and C2. B) FiO2% as function of the inserted flow on position 1 (blue) and 2 (red).
Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- Appendixsupplementaldigitalcontent.docx