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Mucor irregularis-associated cutaneous mucormycosis: Case report and review

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1. Introduction

Cutaneous localizations are frequently observed in healthcare-associated mucormycosis [1]. In this setting, highly immunosuppressed patients such as those with hematological malignancy or transplantation are particularly at risk. In one study, mucormycosis accounted for 2.3% of invasive fungal diseases in solid organ transplantation (SOT), with a mortality rate often approaching 50% [2]. While the highest incidence rate was observed in liver transplant recipients, the estimated mucormycosis incidence in kidney transplant recipients ranged from 0.2% to 1.2% [3]. In a large cohort of patients with invasive fungal diseases from 23 US transplant centers, cutaneous localization accounted for 13% of the 28 mucormycosis cases [2]. The diagnosis of mucormycosis is often delayed in SOT [4], but this is highly dependent on the infected site. However, even though skin localizations can be easily documented by skin biopsies, species identification remains difficult. In fact, in healthcare-associated mucormycosis, more than 30% of Mucorales species responsible for cutaneous involvement were unidentified [1].

We report a subacute cutaneous mucormycosis in a kidney transplant recipient due to a very rare Mucorale species. It is possible that the infection was healthcare-associated. To our knowledge, this is the first reported case of Mucor irregularis infection in a solid organ transplant recipient. However, the management of mucormycosis in the SOT population differs from treatment in hematological patients, and the management of this specific Mucorale species is discussed.

2. Case

A 69 year-old woman was admitted in March 2012 for subcutaneous nodular lesions on the back of the left hand and left elbow (Fig. 1). She had chronic renal failure secondary to a hemolytic uremic syndrome and had received two kidney transplants from cadaveric donors in 1987 and 1993. Her baseline creatinine serum level was 150 μmol/L. Her current immunosuppressive therapy was cyclosporine 50 mg b.i.d., azathioprine 50 mg/d and prednisolone 5 mg/d. One year earlier, she was diagnosed with an epidermoid carcinoma on the right leg, which was treated with radiation therapy. She also had a history of gout.
At the end of January 2012, she had corticosteroid infiltrations on the left radiocarpal and right metacarpus-phalangeal joints for a gout flare-up. Three weeks later, five-millimeter white subcutaneous nodules appeared, first on the left hand and then on the left elbow. The patient did not report previous injury to the left arm. She mentioned that she applied a moisturizer on both arms daily, but did not recall any contact with plants or animals.

On admission (day 0), the hand nodules had become ulcerated and purulent (Fig. 1A). Overall, the lesions were painless and the patient had no fever. No sign of arthritis or neurological abnormality were noted. Blood test results showed 6000 leukocytes/mm³, 800 lymphocytes/mm³, 200,000 platelets/mm³, hemoglobin 10.8 g/dL, C-reactive protein 6 mg/L, creatinine 195 μmol/L, glucose 16.4 mmol/L, with HbA1c 8.1%, iron level 14 μmol/L, ferritin 774 μg/L, and LDH 548 UI/L. A skin biopsy was performed and histopathologic findings showed non-caseating eosinophilic necrosis, granulomatous histiocytic infiltrate, and multinucleate giant cells in the dermis and subcutis. Necrotic areas were surrounded by large, irregular, non-septate, wide-angle branching hyphae on periodic acid-Schiff and Grocott stainings (Fig. 2).

Cultures of biopsy specimens on Sabouraud chloramphenicol agar slants (BioRad Laboratories, Marnes-La-Coquette, France) incubated at 30 °C and 35 °C were positive (day +5) with profuse fluffy whitish colonies presenting a lemon-yellow reverse. The identification was performed by sequencing of the ITS1-5.8S-ITS2 region of the ribosomal DNA using the universal primers ITS1-ITS4 [5]. The nucleotide sequence (deposited in Genbank under accession number KJ477286) had 100% identity over 451 bp for M. irregularis (formerly Rhizomucor variabilis var. variabilis [6]), compared to the nucleotidic sequences of strains CMFCCC B 50 m and CBS 654.78 published under GenBank accession numbers JX976252 and JX976261 [7]. Minimal inhibitory concentrations, measured by the E-test agar diffusion method (bioMérieux SA, Marcy l’Etoile, France), were respectively 0.094, 12, 8 and > 32 μg/ml to amphotericin B (Amb), itraconazole, posaconazole and voriconazole, respectively.

To assess whether mucormycosis was localized to the skin or disseminated, we performed sinus, chest and abdominal computed tomography, as well as plain films of the left arm, and a cerebral magnetic resonance. All were normal. Cyclosporine and prednisolone were tapered and azathioprine was discontinued. Concurrent type 2 diabetes was balanced with oral medication. Liposomal amphotericin B (L-AmB) was immediately introduced at 3 mg/kg/day, and then increased at 5 mg/kg/day (day +9) due to progression of the lesions. During the course of therapy, the patient showed signs of L-AmB nephrotoxicity with serum creatinine increasing up to 380 μmol/L and respiratory distress due to pulmonary edema. Dialysis was performed twice. Given the severe nephrotoxicity to L-AmB, the drug was discontinued (day +21).

The patient received a total of 1.1 g of L-AmB over 3 weeks. Extensive surgical debridement of the left hand and elbow was performed on day +13 (Fig. 1B). Progressive local improvement was noted in the following weeks. One month post-surgery (day +30), a skin graft was successfully performed. No mucormycosis relapse was noted 2 years after treatment discontinuation.

Suspecting a healthcare-associated infection, we attempted to look the presence of fungi in the corticosteroid vial, but it unfortunately could not be analyzed. However, no mucormycosis in patients receiving steroids infiltration was reported to the French National Institute for Health Surveillance (InVS) during the same period. In addition, the patient’s skin moisturizer was also cultured on Sabouraud agar slants but no fungus was isolated.

3. Discussion

In 1999, Voigt et al. reported that R. variabilis was misplaced in the Rhizomucor clade and would be better positioned within...
Mucor spp. [8], and its name was changed to M. irregularis. This species was not reported in the recent French retrospective cohort of patients with mucormycosis [4]. However, it has occasionally been responsible for cutaneous mucormycosis in China [9–11], India [12,13], Japan [14], USA [15,16] and Australia [17]. The 12 documented cases are presented in Table 1. Infections were all proven by skin or mucosal biopsies, with DNA sequencing of cultured isolates for cases published since 2009. This fungus exhibits a specific cutaneous tropism involving the uncovered skin. Chronic lesions lasting from several months to years have been described in immunocompetent patients [9–13]. Although the incubation period is unknown, the infection seems to develop within few days in neutropenic patients [15,16], and rapidly in the incubation period is unknown, the infection seems to develop within few days in neutropenic patients [15,16], and rapidly in other immunocompromised hosts [14]. Histopathological examination shows inflammatory granuloma in the dermis of patients with chronic infections [10–13], whereas it reveals dermis abscesses without granuloma in more acute forms [15]. Although cutaneous mucormycosis due to other species of Mucorales often causes disseminated disease, M. irregularis infection can be localized, as in this case. Nevertheless, progressive tissue invasion and dissemination should be assessed, given the vascular tropism of these aggressive fungi, especially in immunocompromised hosts. Liposomal amphotericin B, ≥ 5 mg/kg/day, is the optimal treatment of these destructive mycoses and monotherapy is recommended as first line treatment [18]. Posaconazole can be used as maintenance therapy, but some Mucorales species exhibit high MIC values to posaconazole. M. irregularis posaconazole MIC values are only rarely reported in the literature, but some authors reported MIC ranging from 0.25 to 2 μg/ml using the microdilution reference method [12,19]. Caspofungin alone has no activity against Mucorales. However, it already showed a good synergistic activity in vitro in combination with amphotericin B [20]. In vivo positive effect of Caspofungin was seldom observed and always in combination with other antifungal drugs. In this case, liposomal amphotericin B had to be stopped because of nephrotoxicity and the other therapeutic option, posaconazole, could not be used due to high MIC values.

Surgery probably had a major impact on the favorable outcome in this case, since amphotericin B could not be prolonged more than one month and limited cumulative dose of 1.1 g was administered. Surgery has not been systematically performed for M. irregularis infections in the literature (Table 1). The need for surgery seems to mainly depend on lesion extension and patient’s overall condition. Hematological conditions, such as thrombocytope尼亚 and neutropenia often increase risks associated with invasive procedures. Thus, neither of the two neutropenic cases had surgery to control the infection [15,16]. Extensive tissue debridement could be necessary to avoid progression, decrease fungal load, and allow for better diffusion of antifungal drugs through non-necrotic areas, especially in solid organ transplant recipients.

Table 1: Proven cases of Mucor irregularis published in the literature.

| Country | Age of cases | Gender | Localization | Comorbidities | Disease duration | Treatment | Duration | Outcome | Ref. |
|---------|--------------|--------|--------------|---------------|-----------------|-----------|----------|---------|------|
| India 18 | M | Rhinofacial | None | Type 2 diabetes, bladder cancer, rheumatoid arthritis, steroids | 12 years | Debridement; systemic fluconazole | 60 days | Improved | [12] |
| Japan 78 | M | Legs | None | Type 2 diabetes, bladder cancer, rheumatoid arthritis, steroids | 3 months | Debridement + L-AmB 5 mg/kg/day | 12 weeks | Cured at EOT; no further follow-up | [14] |
| China 37 | F | Face, sinus | None | | 10 years | d-AmB total dose 1.5 g | 3 months | Cured at 3-month follow up | [14] |
| 37 | F | Hand | None | | 18 years | Ketoconazole | 2 months | ND | Recurrence | [10] |
| 54 | F | Face | None | | 16 years | Flucnozole | 2 months | ND | Recurrence | [10] |
| 33 | M | Arm | None | | 7 years | Itraconazole + terbinafine; d-AmB total dose 42.3 mg/kg | ND | Cured at EOT | [10] |
| 5 | F | Face | None | | 7 months | Itraconazole | ND | Cured at EOT | [10] |
| 40 | F | Cheek | None | | 9 years | Itraconazole | ND | Cured at EOT | [10] |
| 57 | M | Rhinofacial | None | | 3 years | Itraconazole | 3 months | Cured at EOT; no further follow-up | [9] |
| 35 | M | Rhinofacial | None | | | L-AmB + terbinafine | 2 months | Cured at 1-year follow up | [11] |
| USA 57 | M | Arms | Rheumatoid arthritis, acute myeloid leukemia, neutropenia, steroids | Few days | d-AmB total dose 770 mg ABL total dose 63.9 g | 10 days | Cured at EOT; died from leukemia | [15] |
| USA 14 | F | Palate | Hematopoietic stem cell transplant recipient | 2 weeks | Posaconazole + Caspofungin | 2 weeks | Improved after 4 weeks of treatment; died from Hormographiella aspergillata infection | [16] |

Notes: The following abbreviations were used: d-AmB: amphotericin B deoxycholate; L-AmB: liposomal AmB; ABLC: amphotericin B lipid complex; EOT: end of therapy; ND: no data.

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Inappropriate skin disinfection prior to the injection could also have contributed to the infection. Other potential environmental sources such as water, with which the breached skin could have been in contact with, were not investigated. Water is indeed a potential reservoir for fungi that infection control specialists and clinicians should not underestimate [17]. This case also highlights the limitations associated with investigating an isolated case of suspected healthcare-associated invasive mucormycosis.

**Conflict of interest statement**

BR has received travel grants from Gilead Sciences and MSD. OL is a consultant for Gilead Sciences, and has received grants or speaker’s fees from MSD, Roche, Astellas, Gilead Sciences and Pfizer. MEB has received travel grants or speaker’s fees from Ademtech, MSD, Gilead Sciences, Astellas, and Pfizer. Other authors declared no conflict of interest.

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