Effect of Breed and Gender on Meat Quality of M. longissimus thoracis et lumborum Muscle from Crossbred Beef Bulls and Steers

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Abstract: The objective of this study was to determine whether sire breed and/or castration had an effect on meat quality of M. longissimus thoracis et lumborum (LTL) muscle from crossbred bulls and steers and to investigate the relationship amongst the traits examined. Warner–Bratzler shear force (WBSF), intramuscular fat (IMF)%, cook-loss%, drip-loss%, colour (L*, a*, b*) and ultimate pH (upH) were determined in the LTL muscle from eight beef sire breeds representative of the Irish herd (Aberdeen Angus, Belgian Blue, Charolais, Hereford, Limousin, Parthenaise, Salers and Simmental). The results indicate that IMF%, cook-loss% and drip-loss% were associated with breed (p < 0.05); while WBSF, IMF% and cook-loss% differ between genders (p < 0.05). Steer LTL had a greater IMF% and exhibited reduced WBSF and cook-loss% in comparison to the bull LTL (p < 0.05). This study provides greater insight into how quality traits in beef are influenced by breed and gender and will support the industry to produce beef with consistent eating quality.

Keywords: beef quality; castration; breed; shear force; intramuscular fat

1. Introduction

Factors such as breed, gender, age of animal at slaughter, diet and feeding regime can influence muscle characteristics, which in turn affect meat quality [1–3]. Meat quality attributes such as tenderness, colour, flavour, juiciness, and water-holding capacity (WHC)-related traits—cook-loss% and drip-loss%—influence consumer satisfaction [4,5]. The relative importance and value of particular meat quality traits vary according to the type of meat product being produced and marketed and also the end target consumer of the product; e.g., tenderness is more important for beef meat than sheep meat [5]. As progress is made on the development of quality traits, their relative value is also altered, which has potential to impact on their prioritisation in animal breeding programmes [6]. In order to meet the demand for the high-quality product anticipated by consumers, beef producers must focus on improving the quality in addition to quantity [7]. This has increased the focus of both industry and academia on husbandry and breeding strategies aimed at improving meat eating quality traits [5].

Growth rate, carcass yield, feed efficiency and carbon efficiency are positively influenced in bulls (intact adult males) compared to steers (castrated adult males) [8,9]. Carcass fat is, however, a limiting factor for bull beef production [10]. Gender has been associated with many aspects of meat quality and has been proposed to favourably influence fat deposition and tenderness [11]. Steers are commonly
used to produce the highest quality beef that is distinguished from beef attained from bulls and obtains a premium price in restaurants and markets in developed nations [11].

Breed also influences meat quality. Individual cattle breeds have been developed through extensive long-term selection for specific production attributes, such as increased growth rate, carcass conformation and intramuscular fat (IMF) [12,13]. Early maturing breeds such as Angus and Hereford have higher levels of IMF (and associated traits tenderness and flavour) in comparison to late maturing continental breeds [14–17] and this is reflected in the price [18].

Pre-slaughter handling and subsequent post-mortem processing play a major role in the final quality attributes of meat. Factors such as feed withdrawal [19], transport time [11] and stress during transport [20] can have a negative impact on subsequent meat quality. Bulls are more sensitive than steers to all these factors due to their sexual maturity and greater aggression in the lairage. This can lead to higher ultimate pH measurements and unfavourable meat quality [11,21,22].

Gaining insights into how meat quality traits are affected by animal breed and gender could inform pre-slaughter handling practises and post-mortem technologies aimed at maximising quality [22]. Furthermore, it could allow meat processors to optimise meat management systems based on specific quality traits (due to the animal’s breed or gender) [23–25]. Therefore, the objective of this study was to determine whether sire breed and/or castration had an effect on the meat quality of M. longissimus thoracis et lumborum (LTL) muscle from crossbred bulls and steers and to investigate the relationship amongst the traits examined.

2. Materials and Methods

2.1. Animals and Sample Preparation

Crossbred bull and steer progeny were obtained and reared under the same feeding and environmental conditions by the Irish Cattle Breeders Federation Tully Progeny Test Centre (Tully, Kildare, Republic of Ireland). Bull and steer progeny examined in this study were bred from crossbred commercial suckler dams artificially inseminated by elite Irish beef breed bulls. Animals were acclimatised for approximately 30 days before starting a 90-day testing period. Bulls were offered an ad-libitum concentrate diet with 3 kilograms (kg) of fresh hay, while steers were offered 8 kg of concentrates and 5 kg of hay on a fresh-weight basis per head per day. Hay was offered to support the healthy functioning of the rumen and to reflect an Irish commercial high concentrate-based dietary regimen. All animals were finished to a specified carcass conformation and fat score range. For bulls this was U− to E+ conformation score, 3− to 5+ fat score and 678 kg live weight (±58 kg); for steers this was R− to E= conformation score, 2+ to 5+ fat score and 637 kg live weight (±64 kg). Bulls were slaughtered at approximately 487 days old (±24 days), while steers were slaughtered at approximately 634 days old (±52 days). Eight beef breeds, with numbers representative of that in the Irish herd, were included as part of this study as follows: Aberdeen Angus (AA; bull n = 36, steer n = 28), Belgian Blue (BB; bull n = 67, steer n = 10), Charolais (CH; bull n = 127, steer n = 41), Hereford (HE; bull n = 2, steer n = 11), Limousin (LM; bull n = 234, steer n = 62), Parthenaise (PT; bull n = 11, steer = 4), Salers (SA; bull n = 25, steer n = 16) and Simmental (SI; bull n = 63, steer n = 16). Animals were slaughtered in batches of approximately 50, between February 2014 and May 2017 in a commercial plant by electrical stunning (50 Hz) followed by exsanguination from the jugular vein. Between 40–60 min post exsanguination, carcasses were split in half then chilled for 24 h at 2 °C. Twelve steaks with a thickness of 2.54 cm were removed sequentially from the right-side LTL 48 h post-mortem starting at the rump end and vacuum packaged. Steaks were labelled 1–12 according to the trait being measured in order to ensure the analysis was conducted in a consistent location within the LTL muscle. Steaks were frozen at −20 °C after 2 or 14 days of ageing at 4 °C, dependent on the trait being determined.
2.2. Warner–Bratzler Shear Force and Cook-Loss%

For the determination of cook-loss%, 14-day aged steaks were frozen at −20 °C until analysis then thawed within unsealed plastic vacuum bags in a circulating water bath at room temperature (20 °C). Steaks were trimmed of external fat, blotted lightly with tissue paper to remove moisture and weighed. Steaks were immersed in a water bath for cooking (Grant Instruments Ltd., Royston, England) at 72 °C until an internal core temperature of 70 °C was reached using a temperature probe (Eirelec Ltd., Dublin, Ireland). Samples were cooled to room temperature, blotted lightly with tissue paper and weight was recorded. Samples were then placed within new unsealed vacuum bags and left to temper overnight in a fridge at 4 °C. Cook-loss was expressed as a percentage of the raw weight of the steak as follows:

\[
\text{Cook-loss} \, (\%) = \frac{\text{raw weight} - \text{cooked weight}}{\text{raw weight}} \times 100 \quad (1)
\]

Following cook-loss determination, the tempered steaks were used for Warner–Bratzler shear force analysis according to a modified version of American Meat Science Association guidelines [26]. Seven cores per steak were removed for analysis using a 1.27-cm core and sheared perpendicular to the fibre direction using the Instron 4464 Universal testing machine (Instron Ltd., Buckinghamshire, UK), with a load cell of 500 N and a cross head speed of 50 mm/min, and analysed using Bluehill®2 Software (Instron Ltd., Buckinghamshire, UK). The maximum peak force recorded during analysis was reported as Newton (N) shear force. The highest and lowest measurements were excluded with the average of the remaining 5 cores recorded as the result to reduce standard deviation.

2.3. Intramuscular Fat%

IMF% was determined on 2-day aged steaks using the Smart System-5 microwave moisture drying oven and NMR Smart-Trac rapid fat analyser (CEM Corporation, Matthews, NC, USA) using AOAC Official Method 985.14 [27]. In brief, steaks previously frozen at −20 °C were thawed within unsealed plastic vacuum bags in a circulating water bath at room temperature (20 °C). Once thawed, the steaks were trimmed of external fat, cut into cubes approximately 2.5 × 2.5 cm and placed into a RobotCoupe R2 blender and homogenised to a fine consistency. Two grams of homogenised meat free of connective tissue was then placed within the Smart-Trac for analysis.

2.4. Ultimate pH

Ultimate pH (upH) measurements were collected from carcasses 48 h post-mortem by placing a calibrated pH meter (Hanna HI 9125 pH meter, Woonsocket, RI, USA) within the loin between the 12th and 13th rib avoiding bone and connective tissue. Calibration of the pH electrode was performed with standardized buffers (pH 4.0 and 7.0).

2.5. Drip-Loss%

Drip-loss% was analysed according to the procedure of Honikel and Hamm [28]. From each steak (2-day aged samples, assigned for drip-loss analysis), a piece was removed (2.5 cm in thickness, 7.5 cm in length and 5 cm in width) avoiding connective tissue and large areas of fat not representative of the sample. Samples for drip-loss weighed approximately 100 ± 5 g and were lightly blotted with tissue paper, weighed, then suspended by string and an unfolded paperclip (formed into a hook shape) within an expanded clear plastic bag, with care taken to ensure the sample did not come into contact with the bag. The samples were suspended in a chill room at 4 °C for 96 h, after which, the surface was lightly blotted with a tissue and re-weighed. Drip-loss was expressed as a percentage of the original weight of the steak as follows:

\[
\text{Drip-loss} \, (\%) = \frac{\text{initial weight} - \text{final weight}}{\text{initial weight}} \times 100. \quad (2)
\]
2.6. Colour

Colour measurements were performed using the HunterLab UltraScan Pro CIE L∗a∗b∗ system with a dual beam xenon flash spectrophotometer (Hunter Associates Laboratory, Inc., Reston, VA, USA). CIE L∗ (lightness), a∗ (redness), b∗ (yellowness) values were recorded. The illuminant (D65, 10°) consisted of an 8° viewing angle and a 9.9-mm port size. Calibration was carried out using a white standard tile (L = 100) and light trap (L = 0). The white tile was covered in cling film prior to calibration to prevent any effect on the colour reading. Steaks (2 d post-mortem) were wrapped in cling film and allowed to bloom for 1 h prior to measurement. Three measurements were taken in three separate locations on each steak, avoiding intramuscular fat and connective tissue.

2.7. Statistical Analysis

Statistical analysis was performed by two-way Analysis of Variance (ANOVA) using Tukey–Kramer adjusted generalised linear model (GLM) procedures of Statistical analysis Software (SAS) 9.4 (SAS Institute, Cary, NC, USA). Pearson Correlations between beef LTL quality attributes were calculated using the CORR procedure in SAS. Differences were considered significant at the p < 0.05 level.

3. Results

3.1. Correlations between Traits

Pearson correlations between eight beef quality traits examined as part of this study are presented in Table 1. Cook loss, drip loss, pH and colour traits showed a number of correlations with each other. WBSF and IMF were negatively correlated, with higher fat-content meat being associated with lower shear force. Cook loss was also linked to both fat content and shear force.

|               | IMF (%) | upH   | Cook-Loss (%) | Drip-Loss (%) | L*    | a*    | b*    |
|---------------|---------|-------|---------------|---------------|-------|-------|-------|
| WBSF          | −0.26 *** | −0.05 | 0.19 ***      | −0.16 ***     | 0.06  | −0.15 *** | −0.05 |
| IMF (%)       | −0.015  | −0.22 *** | −0.08        | −0.07         | 0.1 ** | 0.05  |
| upH           | −0.15 *** | 0.03  | −0.01         | −0.01         | −0.01 | −0.01 |
| Cook-loss (%) | 0.06    | 0.18 *** | 0.23 ***      | 0.16 ***      | 0.04  | 0.13 ** |
| Drip-loss (%) | 0.23 *** | 0.42 *** | 0.83 ***      | 0.83 ***      |       |       |

* p < 0.05, ** p < 0.01, *** p < 0.001; WBSF—Warner–Bratzler shear force; IMF—Intramuscular fat; upH—Ultimate pH.

3.2. Effect of Breed and Gender on Meat Quality

The effects of breed and gender on LTL muscle quality traits from eight beef breeds are presented in Table 2. Gender had a significant effect on WBSF values, with bull LTL samples having higher shear force than that of steer LTL. However, no effect was observed on WBSF values for breed.

IMF was associated with both breed and gender, with LTL from steers containing almost twice the IMF% of bull LTL (2.85 and 1.27%, respectively; Table 2). AA sired LTL samples had the highest levels of IMF in the current dataset (2.78%) with BB and PT (1.12%) sired progeny, the lowest. Cook-loss was significant for both breed and gender effects, with LM-sired progeny having the lowest cook-loss values (29.09%). Bull LTL cook-loss% was higher than that of steers (30.4 and 29.25%, respectively). Drip-loss values only tended to be higher in steers than in bulls (3.41 and 2.73%); however, sire breed had an effect on drip-loss% values. The AA sire progeny had the lowest drip-loss% measured (2.15%), with BB having the highest (4.11%).
Table 2. Effect of breed and gender on quality traits of *M. longissimus thoracis et lumborum* (least squares mean and standard error).

| Trait | Breed | Gender | p-Value |
|-------|-------|--------|---------|
|       | AA    | HE     | LM      | CH     | SA     | SI     | BB     | PT     | Bull   | Steer  | Breed | Gender |
| n     | 64    | 13     | 296     | 168    | 41     | 79     | 77     | 15     | 565    | 188    | -     | -     |
| WBSF  | 41.59 (1.2) | 36.96 (3.8) | 42.69 (0.7) | 40.62 (0.9) | 42.57 (1.6) | 44.37 (1.4) | 42.55 (1.7) | 40.97 (2.9) | 44.54 (1.2) | 37.81 (1.1) | 0.3122 | 0.0001 |
| IMF (%) | 2.78 (0.2) b | 2.16 (0.5) ab | 2.13 (0.1) ab | 2.05 (0.1) ab | 2.41 (0.2) ab | 2.13 (0.3) ab | 1.7 (0.2) a | 1.12 (0.4) a | 1.27 (0.1) | 2.85 (0.1) | 0.009 | 0.0001 |
| upH   | 5.55 (0.02) | 5.33 (0.05) | 5.55 (0.01) | 5.54 (0.01) | 5.56 (0.02) | 5.6 (0.03) | 5.54 (0.04) | 5.37 (0.01) | 5.54 (0.01) | 0.6594 | 0.286 |
| Cook-loss (%) | 30.15 (0.4) ab | 29.09 (1.1) ab | 29.09 (0.2) a | 29.66 (0.2) ab | 29.33 (0.5) ab | 30.59 (0.4) b | 29.36 (0.5) ab | 31.07 (0.8) b | 30.4 (0.3) | 29.25 (0.3) | 0.0032 | 0.005 |
| Drip-loss (%) | 2.15 (0.3) a | 2.5 (0.5) ab | 2.97 (0.1) e | 3.22 (0.1) f | 2.72 (0.2) bcd | 2.52 (0.3) abc | 4.37 (0.3) ef | 4.11 (0.5) ef | 2.73 (0.2) | 3.41 (0.2) | 0.0009 | 0.0950 |
| L*    | 41.89 (0.5) | 41.68 (1.1) | 42.66 (0.2) | 42.52 (0.3) | 41.75 (0.5) | 42.27 (0.7) | 42.14 (0.5) | 42.11 (0.9) | 42.33 (0.3) | 41.93 (0.3) | 0.5777 | 0.3769 |
| a*    | 14.37 (0.3) | 14.15 (0.7) | 14.31 (0.1) | 14.56 (0.2) | 14.63 (0.3) | 14.27 (0.4) | 13.61 (0.3) | 14.17 (0.5) | 14.01 (0.2) | 14.52 (0.2) | 0.2665 | 0.0635 |
| b*    | 11.15 (0.3) | 10.67 (0.6) | 11.41 (0.1) | 11.6 (0.1) | 11.49 (0.3) | 11.12 (0.4) | 10.82 (0.3) | 11.31 (0.5) | 11.33 (0.2) | 11.06 (0.2) | 0.2902 | 0.3130 |

n—number of animals in each breed; AA—Angus; BB—Belgium Blue; CH—Charolais; HE—Hereford; LM—Limousin; PT—Parthenaise; SA—Salers; SI—Simmental; ()—Standard error; WBSF—Warner–Bratzler shear force; IMF—Intramuscular fat; upH—Ultimate pH; a,b,c,d,e,f,g Within a row values not sharing a common superscript are significantly different for breed.
4. Discussion

This study illustrates the significant effect castration has on important technological beef quality traits, such as increasing IMF%, reducing WBSF and improving cook-loss%. Moreover, the quality attributes, IMF, cook-loss and drip-loss, varied significantly according to sire breed. However, very little variability exists for texture, as reflected by WBSF scores, and sire breed was not determined to be a significant factor for this trait.

Interesting correlations amongst the traits examined as part of our study were found. Notably, the three traits affected by gender, WBSF, IMF% and cook-loss%, were all significantly correlated to each other. In this study, traits relating to different aspects of meat quality show trends that were associated with each other, pointing to particular animals having an overall higher quality, relative to others within the study. For example, animals with higher IMF% and redness values had lower WBSF scores and reduced cook-loss%, which could be considered to be a combination of trait values associated with good overall meat quality. Monteiro et al. [29] also reported a positive relationship between WBSF and cook-loss%. They concluded that cook-loss% influenced WBSF more than any other physiochemical trait and was the reason for shear force variation. The multifactorial relationship amongst these traits suggests that selecting one of these correlated traits for improvement may have a beneficial effect on other meat quality attributes. The two water-holding capacity related traits (cook-loss% and drip-loss%) were not correlated with each other. The two traits capture different aspects of fluid loss in processing. Drip-loss is reflective of exudate, consisting mainly of water and proteins, whereas cook-loss may also be associated with glycogen potential and additionally melting fat during the thermal processing [30]. These traits have been well studied in pork [31]. Interestingly, a comparable lack of correlation between these traits is present in beef in the current study.

The highest scoring sire breed for IMF% was the early-maturing AA sire offspring, which was significantly different from the two lowest scoring sire breeds for this trait, i.e., the late maturing continental BB and PT sire progeny. In a comprehensive study by Gagaoua et al. [1], breed had a significant influence on IMF%, with AA bulls having twice the IMF% relative to continental breed bulls examined in their study. Differences in IMF% values between breeds is mostly attributed to genetics, with early maturing breeds (such as AA and HE) having higher fat deposition than continental breed animals [16]. Steers had greater than twice the IMF% relative to bulls, consistent with Moran et al. [10] and Nian et al. [32] who reported that gender was significant for IMF. The difference between IMF% values between bulls and steers is attributed to the removal of the testes in steers, which arrests sexual maturation leading to reductions in growth rate and muscular development and increases fat deposition and accelerates the fattening period [33]. Other hormones involved in muscle and adipose tissue metabolism include leptin, growth hormone, insulin, cortisol, insulin-like growth factor 1, thyroxin and triiodothyronine [34–36]. Testosterone binds to receptors within the muscle, increasing the incorporation of amino acids into protein and increasing the capacity for muscular development and growth rate [37]. This results in an increase in muscle mass without increases in IMF% [37]. Greater IMF values in steers are attributed to the diminished physiological effects of this androgen, reducing plasma lipids, increasing lipolysis by adipocytes and stimulating androgen receptors [38,39]. The castration of bulls is directly involved in the upregulation of the lipogenic gene expression of fatty acid synthase (FASN) and acetyl-CoA carboxylase (ACC); furthermore, it also downregulates the lipolytic gene expression of monoglyceride lipase (MGL) and adipose triglyceride lipase (ATGL) [40]. Hence, castration contributes to improved IMF deposition mediated through increased lipid uptake and lipogenesis and decreased lipolysis [40].

WBSF values exhibited a gender effect but not a breed effect, with steers exhibiting a WBSF value approximately 7 N lower than bulls, in agreement with Nian et al. [32]. A gender effect for WBSF between bulls and steers was also found by Mberema et al. [41]. However, they observed 14-day aged bull LTL to be more tender than steer and heifer LTL. The results of the current study contrast with Moran et al. [10] who found no effect on WBSF values between bulls and steers. Gerrard et al. [42] reported that any difference in WBSF values explained by gender is not observed following 13 days
of ageing. However, the current study aged samples for 14 days and an effect was still observed. Other factors that contribute to the textural toughness of beef (namely sarcomere length and collagen) were not assessed but may play a role in the differences observed between genders, with bulls having shorter sarcomeres and higher levels of collagen within muscle [32]. Various studies on the use of late maturing beef bulls have also reported that breed has no significant effect on instrumental tenderness [13,43,44]. With regards to breed effect and shear force, Marino et al. [45] reported significance when comparing three cattle breeds contrasting in traditional breed purpose (Friesian dairy bulls; Romagnola crossbred beef bulls; Podolian bulls—indigenous to southern Italy), with the indigenous Podolian bulls significantly tougher than their counterparts. This is in contrast to the findings of the current study as sire breed had no effect on shear force values.

In the current study, cook-loss% is associated with both breed and gender. The breed effect was observed between LM and SI sire breeds. This is in agreement with Chambaz et al. [14] who also observed a significant difference in cook-loss between these two breeds. IMF is one of the factors associated with cook-loss; as IMF% increases, cook-loss% values decrease [46]. Findings relating breed to cook-loss are sometimes conflicting. Mandell et al. [47] examined HE and SI breeds for cook-loss% and found no significance; however, when comparing AA, BB and LM bulls for cook-loss, Cuvelier et al. [2] found AA bulls to be significantly different. Steers had lower cook-loss% than the bulls examined which is in agreement with numerous studies [10,32,47]. The differences observed between bulls and steers may be attributed to the larger muscle fibre diameter of bulls, induced by androgens, with muscles of increased cross-sectional area exhibiting greater cook-loss [9,48]. In contrast, Knight et al. [49] found no gender effect between bulls and steers for cook-loss%. In our study, drip-loss% was significant in yield associated with hanging. Similar trends regarding drip-loss values and breed effect were shown by Cuvelier et al. [2] when studying BB, LM and AA breeds, and Chambaz et al. [14] when studying AA, LM, SI and CH breeds. Cuvelier et al. [2] suggested that higher drip-loss% values in larger animal breeds (such as BB and PT, the highest scoring for this trait) may be due to their higher meat:water content. The lower collagen content of double muscle animals may also contribute to increased drip-loss, as WHC is known to increase with increasing amounts of connective tissue [50]. Breeds exhibiting lower drip-loss values have the added economic benefit of less product weight loss, leading to higher financial gain on carcass and primal cuts.

Lightness ($L^*$), redness ($a^*$) and yellowness ($b^*$) values were not influenced by breed or gender in our study. This is consistent with Moran et al. [10] who did not see any difference when comparing colour values between bulls and steers. Nian et al. [32] also found no significant effect for gender on $L^*$ values; however, in that study, steers had significantly higher $a^*$ and $b^*$ values in comparison to bulls, which is in contrast with the current findings. Papaleo Mazzucco et al. [16] found breed to have no effect on $L^*$, $a^*$ or $b^*$ values when comparing AA and HE steers. However, Cuvelier et al. [2] found breed to influence both $L^*$ and $a^*$ values when comparing AA, BB and LM bulls; with BB bulls having lighter and less red $LTL$, and AA bulls having the darkest and reddest $LTL$ ($b^*$ values were not reported).

5. Conclusions

Sire breed had a significant effect on three beef quality traits analysed, IMF%, cook-loss% and drip-loss%, which were also correlated to each other. With respect to breed, Aberdeen Angus sired progeny had the highest IMF% and the lowest drip-loss%, Limousin sired offspring had the lowest cook-loss%, while Belgian Blue and Parthenaise sired progeny scored the highest for drip-loss%. Castration significantly impacted three of the beef quality traits analysed: WBSF, IMF% and cook-loss%. In comparison to bulls, steers had higher IMF% and reduced WBSF and cook-loss%, implying steer
beef to be more tender and juicy, with more favourable IMF%. This study supports the hypothesis that breed and gender influence meat quality traits.

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References
1. Gagaoua, M.; Terlouw, E.M.C.; Micol, D.; Hocquette, J.F.; Moloney, A.P.; Nuernberg, K.; Picard, B. Sensory quality of meat from eight different types of cattle in relation with their biochemical characteristics. J. Integr. Agr. 2016, 15, 1550–1563. [CrossRef]
2. Cuvelier, C.; Clinquart, A.; Hocquette, J.F.; Cabaraux, J.F.; Dufrasne, I.; Istasse, L.; Horlick, J.L. Comparison of composition and quality traits of meat from young finishing bulls from Belgian Blue, Limousin and Aberdeen Angus breeds. Meat Sci. 2006, 74, 522–531. [CrossRef]
3. Mateescu, R.G. Genetics of meat quality. In The Genetics of Cattle, 2nd ed.; Garrick, D., Ruvinsky, A., Eds.; CABI: Oxfordshire, UK, 2015; pp. 544–562.
4. Dransfield, E.; Martin, J.F.; Bauchart, D.; Abouelkaram, S.; Lepetit, J.; Culioli, J.; Picard, B. Meat quality and composition of three muscles from French cull cows and young bulls. Anim. Sci. 2003, 76, 387–399. [CrossRef]
5. Hopkins, D.L.; Mortimer, S.I. Effect of genotype, gender and age on sheep meat quality and a case study illustrating integration of knowledge. Meat Sci. 2014, 98, 544–555. [CrossRef] [PubMed]
6. Thompson, J.M. The effects of marbling on flavour and juiciness scores of cooked beef, after adjusting to a constant tenderness. Aust. J. Exp. Agric. 2004, 44, 645–652. [CrossRef]
7. Melucci, L.M.; Panarace, M.; Feula, P.; Villarreal, E.L.; Grigioni, G.; Carduza, F.; Miquel, M.C. Genetic and management factors affecting beef quality in grazing Hereford steers. Meat Sci. 2012, 92, 768–774. [CrossRef] [PubMed]
8. O’Riordan, E.; Crosson, P.; McGee, M. Finishing male cattle from the beef suckler herd. Irish Grassl. Assoc. J. 2011, 45, 131–146.
9. Seideman, S.C.; Cross, H.R. Utilization of the Intact Male for Red Meat Production: A Review. J. Anim. Sci. 1982, 55, 826–840. [CrossRef]
10. Moran, L.; O’Sullivan, M.G.; Kerry, J.P.; Picard, B.; McGee, M.; O’Riordan, E.G.; Moloney, A.P. Effect of a grazing period prior to finishing on a high concentrate diet on meat quality from bulls and steers. Meat Sci. 2017, 125, 76–83. [CrossRef]
11. Guerrero, A.; Valero, M.V.; Campo, M.M.; Sañudo, C. Some factors that affect ruminant meat quality: From the farm to the fork. Review. Acta Sci. 2013, 335–347. [CrossRef]
12. Park, S.J.; Beak, S.H.; Jung, D.J.S.; Kim, S.Y.; Jeong, I.H.; Piao, M.Y.; Baik, M. Genetic, management, and nutritional factors affecting intramuscular fat deposition in beef cattle—A review. Asian-Australas. J. Anim. Sci. 2018, 31, 1043–1061. [CrossRef] [PubMed]
13. Xie, X.; Meng, Q.; Cui, Z.; Ren, L. Effect of cattle breed on meat quality, muscle fiber characteristics, lipid oxidation and fatty acids in China. Asian-Australas. J. Anim. Sci. 2012, 25, 824–831. [CrossRef]
14. Chambaz, A.; Scheeder, M.R.L.; Kreuzer, M.; Dufey, P.A. Meat quality of Angus, Simmental, Charolais and Limousin steers compared at the same intramuscular fat content. Meat Sci. 2003, 63, 491–500. [CrossRef]
15. Coleman, L.W.; Hickson, R.E.; Schreurs, N.M.; Martin, N.P.; Kenyon, P.R.; Lopez-Villalobos, N.; Morris, S.T. Carcass characteristics and meat quality of Hereford sired steers born to beef-cross-dairy and Angus breeding cows. Meat Sci. 2016, 121, 403–408. [CrossRef]
16. Papaleo Mazzucco, J.; Goszczyński, D.E.; Ripoli, M.V.; Melucci, L.M.; Pardo, A.M.; Colatto, E.; Villarreal, E.L. Growth, carcass and meat quality traits in beef from Angus, Hereford and cross-breed grazing steers, and their association with SNPs in genes related to fat deposition metabolism. *Meat Sci.* 2016, 114, 121–129. [CrossRef] [PubMed]

17. Samoottkwam, K.; Jaturasitha, S.; Tipnate, B.; Warithitham, A.; Wicke, M.; Kreuzer, M. Effect of Improving Lamphun Cattle with Black Angus on Carcass and Meat Quality. *Agric. Agric. Sci. Procedia* 2015, 5, 145–150. [CrossRef]

18. Hanrahan, K. The Significance of Beef. In *Teagasc Beef Manual*; Available online: https://www.teagasc.ie/media/website/publications/2016/Beef-Manual-Section1.pdf (accessed on 10 December 2018).

19. Ilian, M.A.; Morton, J.D.; Bekhit, A.E.D.; Roberts, N.; Palmer, B.; Sorimachi, H.; Bickerstaffe, R. Effect of preslaughter feed withdrawal period on longissimus tenderness and the expression of calpains in the ovine. *J. Agric. Food Chem.* 2001, 49, 1990–1998. [CrossRef] [PubMed]

20. Warner, R.D.; Greenwood, P.L.; Pethick, D.W.; Ferguson, D.M. Genetic and environmental effects on meat quality. *Meat Sci.* 2010, 86, 171–183. [CrossRef]

21. Troy, D.J.; Kerry, J.P. Consumer perception and the role of science in the meat industry. *Meat Sci.* 2010, 86, 214–226. [CrossRef]

22. Troy, D.J.; Ojha, K.S.; Kerry, J.P.; Tiwari, B.K. Sustainable and consumer-friendly emerging technologies for application within the meat industry: An overview. *Meat Sci.* 2016, 120, 2–9. [CrossRef]

23. McCarthy, S.N.; Henchion, M.; White, A.; Brandon, K.; Allen, P. Evaluation of beef eating quality by Irish consumers. *Meat Sci.* 2017, 132, 118–124. [CrossRef]

24. Merlino, V.M.; Borra, D.; Gengenti, V.; Dal Vecchio, A.; Massaglia, S. Beef meat preferences of consumers from Northwest Italy: Analysis of choice attributes. *Meat Sci.* 2018, 143, 119–128. [CrossRef] [PubMed]

25. Ngapo, T.M.; Braña Varela, D.; Rubio Lozano, M.S. Mexican consumers at the point of meat purchase. Beef choice. *Meat Sci.* 2017, 134, 34–43. [CrossRef]

26. American Meat Science Association. Research Guidelines for Cookery, Sensory Evaluation, and Instrumental Tenderness Measurements of Meat. Available online: https://www.meatscience.org/docs/default-source/ publications-resources/amsa-sensory-and-tenderness-evaluation-guidelines/research-guide/2015-amsa-sensory-guidelines-1-0.pdf?sfvrsn=6 (accessed on 14 January 2019).

27. AOAC 985.14. Moisture in meat and poultry products-rapid microwave drying method. In *Official Methods of Analysis of AOAC International*, 15th ed.; Cuniff, P., Ed.; AOAC: Arlington, VA, USA, 1991.

28. Honikel, K.; Hamm, R. Measurement of water holding capacity and juiciness. In *Advances in Meat Research*, 9th ed.; Pearson, A.M., Dutson, T.R., Eds.; Blackie Academic and Professional: London, UK, 1994; pp. 125–161.

29. Monteiro, A.C.G.; Gomes, E.; Barreto, A.S.; Silva, M.F.; Fontes, M.A.; Bessa, R.J.B.; Lemos, J.P.C. Eating quality of 'Vitela Tradicional do Montado’-PGI veal and Mertolenga-PDO veal and beef. *Meat Sci.* 2013, 94, 63–68. [CrossRef] [PubMed]

30. Ryan, M.T.; Hamill, R.M.; O’Halloran, A.M.; Davey, G.C.; Mc Bryan, J.; Mullen, A.M.; McGee, C.; Gisbert, M.; Southwood, O.I.; Sweeney, T. SNP variation in the promoter of the PRKAG3 gene and association with meat quality traits in pigs. *BMC Gen. 2012, 13, 66.* [CrossRef]

31. Huff-Lonergan, E.; Baas, T.J.; Malek, M.; Dekkers, J.C.M.; Prusa, K.; Rothschild, M.F. Correlations among selected pork quality traits. *J. Anim. Sci.* 2001, 80, 617–627. [CrossRef]

32. Nian, Y.; Allen, P.; Harrison, S.M.; Kerry, J.P. Effect of castration and carcass suspension method on the quality and fatty acid profile of beef from male dairy cattle. *J. Sci. Food Agric.* 2018, 98, 4339–4350. [CrossRef]

33. Eichhorn, J.; Bailey, C.; Blomquist, G. Fatty acid composition of muscle and adipose tissue from crossbred bulls and steers. *J. Anim. Sci.* 1985, 61, 892–904. [CrossRef]

34. Ronge, H.; Blum, J. Original article growth factor I during growth. *Reprod. Nutr. Dev.* 1989, 29, 105–111. [CrossRef] [PubMed]

35. Dais, M.; Pirotte, C.; Bister, J.L.; Wergifosse, F.; Cuvelier, C.; Cabaraux, J.F.; Paquay, R. Relationship between leptin content, metabolic hormones and fat deposition in three beef cattle breeds. *Vet. J.* 2008, 177, 273–278. [CrossRef]

36. Davis, M.; Boyles, S.; Moeller, S.; Simmen, R.C. Genetic Parameter Estimates for Serum Insulin-Like Growth Factor I Concentration and Performance Traits in Angus Beef Cattle. *Am. Soc. Anim. Sci.* 2003, 81, 2164–2170. [CrossRef] [PubMed]
37. Venkata Reddy, B.; Sivakumar, A.S.; Jeong, D.W.; Woo, Y.B.; Park, S.J.; Lee, S.Y.; Hwang, I. Beef quality traits of heifer in comparison with steer, bull and cow at various feeding environments. *Anim. Sci. J.* **2015**, *86*, 1–16. [CrossRef]

38. Lee, H.K.; Lee, J.K.; Cho, B. The Role of Androgen in the Adipose Tissue of Males. *World J. Mens. Health*. **2013**, *31*, 136. [CrossRef]

39. Xu, X.; De Pergola, G.; Björntorp, P. The Effects of Androgens on the Regulation of Lipolysis in Adipose Precursor Cells. *Endocrinology* **1990**, *126*, 1229–1234. [CrossRef]

40. Bong, J.J.; Jeong, J.Y.; Rajasekar, P.; Cho, Y.M.; Kwon, E.G.; Kim, H.C.; Baik, M. Differential expression of genes associated with lipid metabolism in longissimus dorsi of Korean bulls and steers. *Meat Sci.* **2012**, *91*, 284–293. [CrossRef]

41. Mberema, C.H.H.; Lietz, G.; Kyriazakis, I.; Sparagano, O.A.E. The effects of gender and muscle type on the mRNA levels of the calpain proteolytic system and beef tenderness during post-mortem aging. *Livest. Sci.* **2016**, *185*, 123–130. [CrossRef]

42. Gerrard, D.E.; Jones, S.J.; Aberle, E.D.; Lemenager, R.P.; Diekman, M.A.; Judge, M.D. Collagen Stability, Testosterone Secretion and Meat Tenderness in Growing Bulls and Steers. *J. Anim. Sci.* **1987**, *65*, 1236–1242. [CrossRef] [PubMed]

43. Avilés, C.; Martínez, A.L.; Domenech, V.; Peña, F. Effect of feeding system and breed on growth performance, and carcass and meat quality traits in two continental beef breeds. *Meat Sci.* **2015**, *107*, 94–103. [CrossRef]

44. Peña, F.; Avilés, C.; Domenech, V.; González, A.; Martínez, A.; Molina, A. Effects of stress by unfamiliar sounds on carcass and meat traits in bulls from three continental beef cattle breeds at different ageing times. *Meat Sci.* **2014**, *98*, 718–725. [CrossRef]

45. Marino, R.; Albenzio, M.; Della Malva, A.; Santillo, A.; Loizzo, P.; Sevi, A. Proteolytic pattern of myofibrillar protein and meat tenderness as affected by breed and aging time. *Meat Sci.* **2013**, *95*, 281–287. [CrossRef] [PubMed]

46. Ozawa, S.; Mitsuhashi, T.; Mitsumoto, M.; Matsumoto, S.; Itoh, N.; Itagaki, K.; Dohgo, T. The characteristics of muscle fiber types of longissimus thoracis muscle and their influences on the quantity and quality of meat from Japanese Black steers. *Meat Sci.* **2000**, *54*, 65–70. [CrossRef]

47. Mandell, I.B.; Gullett, E.A.; Wilton, J.W.; Kemp, R.A.; Allen, O.B. Effects of gender and breed on carcass traits, chemical composition, and palatability attributes in Hereford and Simmental bulls and steers. *Livest. Prod. Sci.* **1997**, *49*, 235–248. [CrossRef]

48. Waritthitham, A.; Lambertz, C.; Langholz, H.J.; Wicke, M.; Gauly, M. Muscle fiber characteristics and their relationship to water holding capacity of longissimus dorsi muscle in brahman and charolais crossbred bulls. *Asian-Australas. J. Anim. Sci.* **2010**, *23*, 665–671. [CrossRef]

49. Knight, T.W.; Cosgrove, G.P.; Death, A.F.; Anderson, C.B. Effect of interval from castration of bulls to slaughter on carcass characteristics and meat quality. *N. Z. J. Agric. Res.* **1999**, *42*, 269–277. [CrossRef]

50. Raes, K.; Balcaen, A.; Dirinck, P.; De Winne, A.; Claey s, E.; Demeyer, D.; De Smet, S. Meat quality, fatty acid composition and flavour analysis in Belgian retail beef. *Meat Sci.* **2003**, *65*, 1237–1246. [CrossRef]

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