The Relationship Between Circulating Bone Morphogenetic Protein-4 and Inflammation Cytokines in Patients Undergoing Thoracic Surgery: A Prospective Randomized Study

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Background: Bone morphogenetic protein-4 (BMP4) has been identified as an inflammation regulator in the diseases of arteries and other organs. However, the relationship between circulating BMP4 and perioperative inflammation remains unclear.

Patients and Methods: Forty patients undergoing lobectomy were randomly allocated into the Control group (not receiving flurbiprofen) and the Flurb group (received 100mg flurbiprofen during surgery). Arterial blood was obtained before surgery (T1), at the end of surgery (T2), and 24 hours after surgery (T3) to test the plasma concentrations of BMP4, its antagonist Noggin, interleukin (IL)-1β, tumor necrosis factor-α (TNF-α), and IL-10. The relationship between BMP4 and other variables and the effects of flurbiprofen on BMP4 changes were investigated.

Results: A total of 35 patients were included. Circulating BMP4 was positively correlated with IL-1β (P<0.01, r=0.575) and TNF-α (P<0.01, r=0.491), negatively correlated with IL-10 (P<0.01, r=−0.675), but not correlated with Noggin. The plasma concentrations of BMP4, IL-1β, and TNF-α increased at T2 (P<0.01, compared with T1) and decreased at T3 (P<0.05, compared with T2). BMP4 concentrations at T3 were significantly higher than at T1 in the Control group (P<0.05), while showing no significant difference in the Flurb group. However, in the Flurb group, the relative changes of BMP4 and IL-1β at T2 and T3 were significantly lower than those in the Control group.

Conclusion: Circulating BMP4 was elevated during surgery and highly correlated with inflammation cytokines. The elevation of BMP4 and inflammatory cytokines could be alleviated by flurbiprofen, indicating that BMP4 may exert pro-inflammatory properties via cyclooxygenase-II signaling pathways.

Keywords: bone morphogenetic protein-4, Noggin, inflammation, non-steroidal anti-inflammatory drugs, perioperative, cyclooxygenase-II

Introduction

Surgical trauma and other invasive interventions (eg, mechanical ventilation, punctures, and blood transfusion) are unavoidable to trigger inflammation perioperatively.1 Current thinking suggests that inflammation can exert both beneficial and detrimental effects on the body, depending on the extent and duration of the inflammatory phase.2,3 Notably, when over-activated, inflammation can cause a series of severe clinical consequences, including sepsis, multiple organ...
dysfunction syndrome (MODS), and cancer recurrence, all of which can prove grave for patients.\textsuperscript{4} Thus, clarification of the clinical regulatory mechanism of inflammation is urgently required.

Bone morphogenetic protein-4 (BMP4) is a member of the transforming growth factor $\beta$ (TGF-$\beta$) superfamily. Previous studies have demonstrated the critical role that BMP4 plays in dictating tissue homeostasis and disease in adults post-injury.\textsuperscript{5–7} Crucially, BMP4 can be synthesized, stored, and released by the vascular endothelial cell,\textsuperscript{8} cardiomyocyte,\textsuperscript{9} and adipocyte.\textsuperscript{10} Recent experimental data shows that BMP4 expressed via the systemic arterial endothelium could induce endothelial dysfunction, promote leukocytes activation, and aggravate the severity of atherosclerosis.\textsuperscript{11} BMP4 expressed by cardiomyocytes following myocardial infarction could increase infarct size through upregulating pro-inflammatory factors, including interleukin (IL)-1$\beta$ and IL-9.\textsuperscript{12,13} Moreover, corroborating evidence\textsuperscript{11,14} further proves that BMP4 signaling is mainly linked to the induction of inflammatory nuclear factor-$\kappa$B (NF-$\kappa$B), nicotinamide adenine dinucleotide phosphate oxidase-1 (NOX1), and intracellular adhesions molecule-1 (ICAM-1), which are the key factors at the initial inflammation stage. All this evidence indicates that BMP4 elevation will exert pro-inflammatory properties at the lesion site of systemic arteries and the heart. In humans, however, the clinical relationship between circulating BMP4 and inflammation still remains unclear.

In the present study, we planned to enroll patients undergoing lobectomy and investigate their plasma concentrations of BMP4, its antagonist, and a series of canonical pro- and anti-inflammatory cytokines, including IL-1$\beta$, tumor necrosis factor-$\alpha$ (TNF-$\alpha$), and IL-10. Furthermore, we tested the effects of flurbiprofen – a non-steroidal anti-inflammatory drug – on the changes of the above variables. We chose Noggin as the antagonist for BMP4 because it has a high affinity to de-activate the BMP4 signaling\textsuperscript{15} and has been validated in previous studies.\textsuperscript{16,17} The results may help to heighten our understanding of the relationship between circulating BMP4 and inflammation, as well as assist in the identification of a new candidate for treating perioperative inflammation in the future.

Methods

The Second Xiangya Hospital’s Institutional Review Board (IRB #LYG20210111) approved this study, and written informed consent was obtained from all subjects participating in the trial. The trial was registered prior to patient enrollment at the Chinese Clinical Trial Registry (www.chictr.org.cn; ChiCTR2000038374; principal investigator: Lin Yang; date of registration: September 21st, 2020) and conducted in accordance with the principles of the Declaration of Helsinki.

Patients

Patients with an American Society of Anesthesiologists physical status of I or II undergoing lobectomy through thoracoscopic technique at the Second Xiangya Hospital, Central South University, were screened for study eligibility. Considering that circulating BMP4 may increase in diseases that include atherosclerosis, hypertension, myocardial infarction, diabetes, and obesity, the following patients were excluded: 1) patients with vascular diseases – especially vascular sclerosis or diagnosed plaque formation; 2) patients with cardiac, hepatic, and other vital organ diseases; 3) patients with hypertension and diabetes; 4) patients with a body mass index (BMI) above 30 kg/m$^2$; 5) patients with a history of drug abuse, receiving long-term steroid therapy, or allergic to non-steroidal anti-inflammatory drugs; 6) patients with an increased (above the upper normal limit) white blood cell count, neutrophil ratio, erythrocyte sedimentation rate, or procalcitonin before surgery; and 7) when the type of surgery changed, or the blood loss exceeded 1000mL during surgery.

Randomization and Allocation

Patients were randomly assigned to either the Control or Flurb group in a 1:1 ratio, with the random list generated using the Microsoft Excel’ RAND’ function. After randomization, a researcher who did not participate in the anesthesia management prepared ‘study drugs’ containing either lipid emulsion (10mL) or flurbiprofen (100mg, 10mL) outside the operating room. The anesthesiologists in charge of the whole anesthesia management were kept blind to the group allocation.

Anesthesia and Intervention

Routinely, after entering the operation room, each patient was monitored with an electrocardiograph, invasive arterial pressure, pulse oximetry, and Bispectral index (BIS). General anesthesia was induced by intravenous injections of midazolam 0.05mg/kg, etomidate 0.1mg/kg, sufentanil 0.5μg/kg, and cis-atracurium 0.1mg/kg. After the patients’ eyelid reflections disappeared and muscle relaxation was achieved, the same anesthesiologist performed double-lumen tracheal (DLT) intubation under the vision of a video laryngoscope and further confirmed the depth of
DLT by a bronchoscope. Anesthesia, analgesia, and muscle relaxation were maintained by continuous infusions of propofol 6–8mg/kg/h, remifentanil 0.3μg/kg/min, and intermittent injections of cis-atracurium. During the operation, end-expiratory carbon dioxide was adjusted between 35–45mmHg, and anesthesia depth was maintained between 40–50 of BIS values. The infusion of all anesthetics halted upon skin closure. Subsequently, 0.1μg/kg sufentanil was injected and a patient-controlled analgesia pump containing sufentanil 150μg and ondansetron 16mg in 100mL normal saline was connected intravenously to each patient to treat post-operative pain. All patients were transferred to a thoracic surgical intensive care unit postoperatively.

In the Flurb group, 100mg flurbiprofen was infused intravenously at the time of skin incision, within 30 minutes, while patients in the Control group were treated with an equal volume of lipid emulsion at the same time.

Data Collections
Arterial blood samples were drawn from each patient at three time points: T1 (before the surgery), T2 (the end of the surgery), and T3 (24 hours after the surgery). The blood samples were stored in tubes containing ethylene diamine tetraacetate acid and were further processed within 12 hours. Each sample was centrifuged for 15 min at 3000rpm to obtain the plasma. Then, the levels of BMP4 (ab231930, Abcam, Cambridge, UK), Noggin (JYM2457Hu, Wuhan, CHN), IL-1β (ab217608, Abcam, Cambridge, UK), TNF-α (ab181421, Abcam, Cambridge, UK), and IL-10 (ab185986, Abcam, Cambridge, UK) were determined using the enzyme-linked immunosorbent assays method (SimpleStep ELISA® kits, Abcam, Cambridge, UK) by a researcher who was blind to the group allocation.

Statistical Analysis
For the sample size calculation, we assumed there to be a correlation (|correlation coefficient|>0.3) between BMP4 and other inflammatory cytokines. We chose a correlation coefficient of 0.3 because it was the smallest value representing a non-negligible correlation, as suggested by a previous study.21 Using a smaller size of correlation in sample size calculation would require more patients, but it was a conservative way to avoid underpower. With a two-sided significance level of 0.05 and a power of 80%, 85 samples were required. Assuming a drop-out rate of approximately 25%, we planned to enroll 40 patients, with a sample size of 120 (40 patients*3 time points).

Continuous data were presented as median and interquartile range (IQR), while categorical data were presented as frequency and percentage. Two analyses were performed to investigate the relationship between BMP4 and inflammation (Figure 1). Firstly, we conducted pairwise comparisons among BMP4, Noggin, and inflammatory cytokines using Spearman’s rank correlation coefficient (r) based on samples from both groups. A |r| > 0.7 was identified as having a strong correlation; a |r| = 0.3–0.7 was identified as having low to moderate correlation; and a |r| < 0.3 was identified as having no correlation.21 Scatter plots and fitted linear regression lines were used to visualize the relationship between two variables. Secondly, we evaluated the effects of flurbiprofen on BMP4, Noggin, and other inflammatory cytokines. Their plasma concentrations at different time points were analyzed using the Kruskal–Wallis test, followed by Dunn’s multiple comparison test. Because the absolute values of BMP4, Noggin, and other inflammatory cytokines varied considerably, we further calculated the relative changes of variables to investigate the effects of

Figure 1 Flow chart.
Abbreviations: Flurb, flurbiprofen; BMP4, bone morphogenetic protein-4.
The plasma concentrations for all variables at different time points were presented in Table 2. The general information (including gender, age, and BMI) and surgical characteristics (including duration of surgery, blood loss, and fluid therapy) of patients from each group are presented in Table 1.

The correlations among plasma concentrations of BMP4, Noggin, and different inflammatory cytokines are shown in Figure 2. BMP4 was positively correlated with IL-1β (r=0.575, P<0.01) and TNF-α (r=0.491, P<0.01), but negatively correlated with IL-10 (r=-0.675, P<0.01). In contrast, Noggin did not show any correlation with either BMP4 (r=-0.03, P=0.73), IL-1β (r=-0.03, P=0.76), TNF-α (r=-0.05, P=0.58), or IL-10 (r=0.10, P=0.28).

The relative changes of BMP4, Noggin, IL-1β, TNF-α, and IL-10 at different time points were listed in Figure 3. Overall, the concentrations of BMP4, IL-1β, and IL-10 increased at T2 (<0.01, compared with T1, Figure 3), and IL-10 levels at T3 were significantly lower than at T1 (P<0.05, Figure 3). In contrast, in the Flurb group, concentrations of BMP4, IL-1β, TNF-α, and IL-10 at T1 and T3 did not show a significant difference (Figure 3). Finally, the Noggin concentration did not show any differences among three time points (Figure 3).

The relative changes of BMP4, Noggin, IL-1β, TNF-α, and IL-10 at different time points were listed in Table 3. In the Flurb group, the relative changes of BMP4 and IL-1β were significantly lower than those in the Control group at T2 and T3 (P<0.05). Otherwise, no significant differences were found between the two groups.

**Discussion**

This is the first human study investigating the relationship between circulating BMP4 and inflammation in perioperative settings. The results show some novel findings that have never been reported: 1) circulating BMP4 is positively correlated with pro-inflammatory cytokines (IL-1β and TNF-α), but negatively correlated with anti-inflammatory factor (IL-10), indicating that BMP4 is highly associated with inflammatory responses; 2) BMP4 increased at the end of surgery and decreased 24-hours after surgery, indicating that BMP4 could serve as a new inflammatory indicator during surgery; 3) the application of 100mg flurbiprofen during surgery decreased BMP4 and IL-1β levels, indicating that circulating BMP4 might regulate inflammation via the cyclooxygenase-II (COX-2) pathways.

BMP4 has now been identified as a novel inflammation regulator in multiple organs. Studies from the last two decades have shown conflicting results on whether BMP4 could act as a molecular switch that induces both pro-inflammatory and anti-inflammatory effects on special organs. Except for BMP4’s pro-inflammatory properties in systemic arteries and myocardium,11–13 the elevation of...
BMP4 levels protects pulmonary arteries from inflammatory injuries. Furthermore, BMP4 secreted by cancer cells will exert an immuno-suppressive effect via favoring anti-inflammatory M2 type macrophage nearby. One possible explanation for the discrepancy of BMP4’s effect on inflammation is due to the dynamic changes of BMPRII (one type of BMP4 receptors) expressions. Recent studies have revealed that pro-inflammatory cytokines could downregulate BMPRII expression in endothelium during the pathogenesis of atherosclerosis. Moreover, the knockdown of BMPRII was shown to switch BMP4’s effect from anti-inflammatory to pro-inflammatory. Above all, one can suggest that BMP4 plays a crucial role in the regulation of inflammation.

Although substantial evidence from cell and animal studies has revealed BMP4’s regulation on inflammation, scant data is available regarding the circulating BMP4 levels in humans. One research reported that patients’ BMP4 serum levels were elevated after myocardial infarction. The BMP4 elevation lasted for about 14 days and was correlated with left ventricular remodeling. Another study furtherly proved that BMP4 plasma levels in patients suffering from out-of-hospital cardiac arrest were significantly higher than patients with stable coronary artery disease or healthy volunteers, accompanied by increased mortality and unfavorable neurological outcomes. Remarkably, the serum from patients undergoing myocardial infarction and post-resuscitation was found to induce endothelial dysfunction via activating the BMP4 signaling pathway and to exert pro-inflammatory properties in vitro. All the evidence above suggests that circulating BMP4 in humans may play a key role in the process of tissue injury and repair.

To the best of our knowledge, the perioperative relationship between circulating BMP4 and inflammation has not been elucidated. In the present study, we have tested...
the plasma concentrations of BMP4 in each patient, finding that the baseline of patients suffering from lung cancer was mainly at 450–550 pg/mL, which was significantly higher than the reference values of circulating BMP4 (100–200 pg/mL) reported in healthy controls by previous researches.27–29 The possible reason can be attributed to the cancer secretion and systemic inflammatory status of the body. Moreover, we also detected the elevation of pro-inflammatory cytokines and the decline of anti-inflammatory cytokines both at the end of the surgery and 24 hours later, reflecting the initiation and long-lasting effect of inflammation caused by perioperative noxious stimuli. Simultaneously, we observed that plasma BMP4 levels were increased and maintained after surgery at a higher level, being strongly positively correlated with pro-inflammatory cytokines, including IL-1β and TNF-α, but negatively correlated with anti-inflammatory factor IL-10. All the results above indicate that BMP4 might serve as a sensitive pro-inflammatory indicator.

COX-2 is another known inflammatory regulator and could be inducible by pro-inflammatory cytokines.30 Previous studies have reported that BMP4 could induce COX2 upregulation in endothelial cells in vitro, and

| Table 2 Plasma Concentrations of BMP4, Noggin, and Inflammatory Cytokines Before Surgery (T1), at the End of the Surgery (T2), and 24 Hours After Surgery (T3) |
|-----------------|-------|-------|-------|-------|
| Control         | T1    | T2    | T3    | P-value |
| BMP4, pg/mL     | 414 (359–447) | 751 (705–819) | 532 (472–643) | <0.01 |
| IL-1β, pg/mL    | 142 (125–175)  | 317 (266–427)  | 211 (193–249)  | <0.01 |
| TNF-α, pg/mL    | 194 (151–212)  | 337 (267–396)  | 262 (219–305)  | <0.01 |
| IL-10, pg/mL    | 171 (156–195)  | 86 (81–117)    | 138 (128–144)  | <0.01 |
| Noggin, pg/mL   | 1542 (1254–1916) | 1398 (1197–1721) | 1333 (1182–1592) | 0.40 |

| Flurb           | T1    | T2    | T3    | P-value |
|-----------------|-------|-------|-------|-------|
| BMP4, pg/mL     | 510 (462–543) | 710 (649–833) | 506 (461–558) | <0.01 |
| IL-1β, pg/mL    | 196 (165–220)  | 319 (232–373)  | 220 (202–242)  | <0.01 |
| TNF-α, pg/mL    | 185 (167–240)  | 358 (257–422)  | 238 (219–296)  | <0.01 |
| IL-10, pg/mL    | 182 (164–200)  | 97 (81–105)    | 155 (131–165)  | <0.01 |
| Noggin, pg/mL   | 1749 (1101–2003) | 1482 (1217–1889) | 1408 (1169–1883) | 0.87 |

Notes: Data presented as median (interquartile range). P-values calculated based on the Kruskal–Wallis test.

Abbreviations: BMP4, plasma bone morphogenetic protein-4; IL-1β, interleukine-1β; TNF-α, tumor necrosis factor-α; IL-10, interleukine-10; Flurb, flurbiprofen.

Figure 3 Boxplots for the plasma concentrations of BMP4, Noggin, and inflammatory cytokines before surgery (T1), at the end of the surgery (T2), and 24 hours after surgery (T3). The results of the Dunn’s multiple comparison test are presented in each figure. *Indicate a P-value <0.05; **Indicate a P-value <0.01.

Abbreviations: BMP4, plasma bone morphogenetic protein-4; IL-1β, interleukine-1β; TNF-α, tumor necrosis factor-α; IL-10, interleukine-10; Flurb, flurbiprofen.
Table 3 The Relative Changes of Plasma Concentrations of BMP4, Noggin, and Inflammatory Cytokines at the End of the Surgery (T2) and 24 Hours After Surgery (T3)

|            | Time Point | Control (N=17) | Flurb (N=18) | P-value |
|------------|------------|----------------|--------------|---------|
| BMP4       | T2         | 1.9 (1.7–2.1)  | 1.4 (1.3–1.7) | 0.002   |
|            | T3         | 1.3 (1.1–1.6)  | 1.0 (0.9–1.1) | 0.002   |
| IL-1β      | T2         | 2.4 (1.9–2.8)  | 1.8 (1.4–2.0) | 0.006   |
|            | T3         | 1.6 (1.2–1.8)  | 1.1 (1.0–1.3) | 0.023   |
| TNF-α      | T2         | 1.8 (1.5–2.2)  | 1.6 (1.4–1.9) | 0.509   |
|            | T3         | 1.3 (1.1–1.6)  | 1.2 (1.0–1.5) | 0.621   |
| IL-10      | T2         | 0.6 (0.4–0.7)  | 0.5 (0.4–0.6) | 0.766   |
|            | T3         | 0.8 (0.7–0.9)  | 0.8 (0.7–0.9) | 0.597   |
| Noggin     | T2         | 1.1 (0.8–1.2)  | 1.1 (0.8–1.2) | 0.644   |
|            | T3         | 0.9 (0.7–1.1)  | 1.0 (0.6–1.4) | 0.597   |

Notes: Data presented as median (interquartile range). P-values calculated based on the Mann–Whitney U-test. The relative changes indicated the ratio of the plasma concentrations at different time points versus the concentration before surgery (T1).

Abbreviations: BMP4, bone morphogenetic protein 4; IL-1β, interleukine-1β; TNF-α, tumor necrosis factor-α; IL-10, interleukine-10; Flurb, flurbiprofen.

hypertensive human renal arteries show higher levels of COX-2 and BMP4, accompanied by endothelial dysfunction.\(^{19,31}\) Similarly, we also found that the elevation of BMP4 and pro-inflammatory factors 24 hours after surgery could be alleviated by the application of flurbiprofen, a non-steroidal anti-inflammatory drug. What’s more, flurbiprofen treatment decreased the relative changes of BMP4 and IL-1β from T2 to T1 and from T3 to T1. Thus, it prompted that circulating BMP4 might regulate inflammation clinically via COX-2 pathways.

Another key regulator of BMP4 signaling is Noggin, which directly binds with and de-activates BMP4 with high affinity.\(^{15}\) Strong evidence has already proven that Noggin application could exert anti-inflammatory properties by inhibiting NOX-1 upregulation and monocyte adhesion in the pathological process of BMP4-induced endothelial dysfunction\(^{19,32,33}\) and cardiomyocyte hypertrophy.\(^{34}\) In the present study, however, we found that endogenous Noggin levels stayed almost unchanged during surgery. The mechanism for such an approach remains unclear and whether Noggin can be a new therapeutic approach to treating inflammation needs further investigation.

Our study has several limitations. Firstly, we have only selected patients undergoing lobectomy, which may not be well representative of the whole general population. Secondly, we have calculated the planned sample size based on the correlations between BMP4 and other variables instead of the differences between Control and Flurb groups. Therefore, although we got some insignificant results regarding the relative changes of Noggin and other inflammatory cytokines between Control and Flurb groups, this might be due to inadequate sample size. Thirdly, we have not extended our investigation to define the effect of circulating BMP4 elevation on pulmonary arteries or tumors, considering that BMP4 would exert a diverse regulatory effect on inflammation in different organs. Future studies are required to determine the differential expressions of BMPRII at the lesion site, which might be important for determining the effects of BMP4.

Overall, the present study shows that circulating BMP4 is markedly elevated during surgery and highly correlated with inflammation cytokines. Moreover, the elevation of BMP4 and inflammatory cytokines could be alleviated by the applications of non-steroidal anti-inflammatory drugs, indicating that BMP4 may exert pro-inflammatory properties via the COX-2 signaling pathways.

Data Sharing Statement
All individual deidentified participant data and statistical analysis code that underlie the results reported in this article will be available after publication. Data can be requested for academic use by contacting the corresponding author (Lin Yang: linyang@csu.edu.cn).

Author Contributions
All authors contributed to data analysis, drafting or revising the article, gave final approval of the version to be published, agreed to the submitted journal, and agree to be accountable for all aspects of the work.

Disclosure
The authors report no conflicts of interest in this work.

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