The possible protective effects of dipyridamole on ischemic reperfusion injury of priapism

Ersagun Karaguzel 1, Cemil Bayraktar 1, Omer Kutlu 1, Esin Yulug 2, Ahmet Mentese 3, Ali Ertan Okatan 1, Fatih Colak 1, Serap Ozer 3, Ilke O.Kazaz 1

1 Department of Urology, Faculty of Medicine, Karadeniz Technical University, Trabzon, Turkey; 2 Department of Histology and Embryology, Karadeniz Technical University, Faculty of Medicine, Trabzon, Turkey; 3 Department of Medical Biochemistry, Karadeniz Technical University, Faculty of Medicine, Trabzon, Turkey

ABSTRACT

Purpose: To investigate the protective effects against ischemia reperfusion injury of dipyridamole in a model of induced priapism in rats.

Materials and Methods: Twenty-four male Sprague-Dawley rats were divided into four groups, control, P/R, P/R+DMSO and P/R+D. 3ml blood specimens were collected from vena cava inferior in order to determine serum MDA, IMA, TAS, TOS and OSI values, and penile tissue was taken for histopathological examination in control group. Priapism was induced in P/R group. After 1h, priapism was concluded and 30 min reperfusion was performed. In P/R+DMSO group 1ml/kg DMSO was administered intraperitoneally 30 min before reperfusion, while in P/R+D group 10mg/kg dipyridamole was administered intraperitoneally 30 min before reperfusion. Blood and penis specimens were collected after the end of 30 min reperfusion period. Sinusoidal area (µm²), tears in tunica albuginea and injury parameters in sinusoidal endothelium of penis were investigated.

Results: Histopathological examination revealed no significant changes in term of sinusoidal area. A decrease in tears was observed in P/R+D group compared to P/R group (p<0.05). Endothelial injury decreased in P/R+D group compared to P/R group (p>0.05). There were no significant differences in MDA and IMA values between groups. A significant increase in TOS and OSI values was observed in P/R+D group compared to P/R group. A significant decrease in TAS levels was observed in P/R+D group compared to the P/R group.

Conclusions: The administration of dipyridamole before reperfusion in ischemic priapism model has a potential protective effect against histopathological injury of the penis.

INTRODUCTION

Priapism is defined as complete or partial penile tumescence exceeding 4 h in length (1, 2). There are three types of priapism, ischemic (low flow, veno-occlusive), non-ischemic (high flow, arterial) and stuttering (repeating). Ischemic priapism is the most common form and represents a urological emergency (3). In low-flow priapism, a reduced venous flow in cavernous tissues, hypoxia related to venous stasis and histopathological changes associated with acidosis are observed. Ir-
reversible corporal tissue necrosis and fibrotic tissue formation may occur if ischemia exceeds 4-6 h (4, 5).

Ischemia-reperfusion (I/R) injury occurs with termination of priapism (6). Although reperfusion is a necessary mechanism for the protection of ischemic tissue, reperfusion itself initiates a pathophysiological process that leads to injury. With I/R injury neutrophils, inflammatory cytokines and adhesion molecules with increased thrombogenicity are activated, and free oxygen radicals develop with the release of massive intracellular Ca (7, 8). Oxidative stress damages the cell membrane and has damaging effects on tissue by leading to permanent cellular injury.

Dipyridamole is a platelet inhibitor with antithrombocytic effects. It is a cyclic guanosine monophosphate (cGMP)-dependent phosphodiesterase and adenosine carrier inhibitor with antioxidant properties that prevents the formation of reactive oxygen radicals in endothelial cells and thrombocytes (9, 10). In addition to antiaggregant effects, it also possesses anti-inflammatory and neuroprotective effects. Experimental studies have shown that dipyridamole reduces I/R injury in various organs and tissues, such as the testis, liver and myocardium (11-13).

The purpose of this study was to determine the protective effects of dipyridamole against I/R injury in an induced experimental model of acute ischemic priapism in rats.

**MATERIALS AND METHODS**

All animal experiments were performed following Karadeniz Technical University Animal Care and Ethics Committee approval, in compliance with the principles of laboratory animal care (National Institutes of Health publication no. 85-23, revised 1985). Twenty-four adult (approximately 4 months) male Sprague Dawley rats weighing 400-450 gr were used in the study. Rats were placed in cages 1 week before the study for adaptation and given standard rat chow and tap water. Rats were numbered and randomized by lots. Throughout the experiment all animals were kept under standard room temperature (23±2°C), lighting (12 h light/12 h dark) and humidity conditions (50%±10).

**Experimental design and groups**

Anesthesia in all surgical procedures was established with 80 mg/kg ketamine and 10 mg/kg xylazine. The priapism model was induced using the vacuum and constrictor band previously described in the literature (14). A 50 cc irrigation injector was attached to the base of the flaccid rat penis and used like a vacuum device. A con- striction band, which was cut from a 16 Fr foley catheter in 2 mm slices, was attached to the tip of the vacuum erection device before vacuum was applied to the penis. Erection was induced by placing the tip of the syringe to the base of the rat penis and withdrawing it to create a 20 cc negative pressure. Once sufficient erection had been induced, the constriction band was removed from the syringe and attached to the base of the penis (Figure 1). The powdered dipyridamole (Koçak Farma, Turkey) used in the experiment was dissolved in DMSO (dimethyl sulfoxide) at 10mg/ml. This was injected intraperitoneally at a dose of 10 mg/kg into rats weighing 400-450 gr. Drug dose

**Figure 1 - Priapism model induced in the rat penis using a constrictor band.**
was determined based on the report demonstrated that dipyridamole protects rat testis against testicular I/R injury (11).

**Control Group:** Three-milliliter blood specimens were collected from the vena cava inferior (VCI) in order to determine basal serum malondialdehyde (MDA), serum ischemia modified albumin (IMA), serum total antioxidant status (TAS), serum total oxidant status (TOS) and serum oxidative stress index (OSI) values. Penectomy was performed and tissues were placed in a 10% formaldehyde solution for basal histopathological examination of penile tissues.

**Priapism/Reperfusion (P/R) Group:** Thirty minute reperfusion was established by removing the constrictor band on the penis following 1 h priapism.

**Priapism/Reperfusion+DMSO (P/R+DMSO) Group:** The same surgical procedure was performed as in the P/R group. This group received the same volume of DMSO solution as the P/R+D group without dipyridamole intraperitoneally, ½ h before reperfusion (Figure-2).

**Priapism/Reperfusion+Dipyridamole (P/R+D) Group:** The same surgical procedure was performed as in the P/R group. Half an hour before reperfusion, 10 mg/kg dipyridamole were administered intraperitoneally.

Following 30 min reperfusion, 3ml blood specimens were collected from all animals in order to determine serum MDA, IMA, TAS, TOS and OSI values. Penectomy was performed on all animals at the end of the experiment and penises were placed in 10% formaldehyde solution for histopathological examination of penile tissues.

**Histopathological analysis**

Penis tissue was extracted from rats from all groups at the end of the study. Penis tissues, taken from approximately the same areas, were kept for 72 h in 10% formaldehyde solution for histopathological examination. Tissue samples were passed through 70%, 90%, 96% and 100% alcohol series for dehydration. They were then passed through xylene solution. They were then again washed in water, and finally dried after being passed through alcohol.

**Masson’s Trichrome Staining:** Following deparaffinization, sections were stained with Masson’s trichrome for detailed histological examination and for better investigation of the cavernous structure (15).

Preparates were evaluated by an experienced and blinded histologist using a light microscope (Olympus BX-51, Olympus Optical Co., Tokyo, Japan). General histological architecture was evaluated. Pathological findings (tunica albuginea tear, endothelial injury) were evaluated semi-quantitatively on a scale of 0 to 3 (0: None, 1: Mild, 2: Moderate, 3: Severe). Groups were also examined using a light microscope at a magnification of 200X on Analysis 5 Research (Olympus Soft Imaging Solutions, Münster, Germany) software for measurement of cavernous tissue sinusoidal area. Means were calculated for sinusoidal area measurements in five different areas for each group.
**BIOCHEMICAL ANALYSIS**

**Serum MDA Activity Assay**

The method described by Yagi was used to determine lipid peroxidation in rat serum samples in the form of MDA concentration (16). Tetramethoxypropane was employed as a standard, and MDA levels were expressed as nmol/mL.

**Measurement of Ischemia-Modified Albumin (IMA)**

Assessment of reduced cobalt to albumin binding capacity (IMA level) was performed based on the colorimetric method described by Bar-Or et al. (17). Briefly, 200 µL of rat serum was added to 50 µL of 0.1% cobalt chloride (Sigma, CoCl₂.6H₂O) in H₂O. The solution was gently shaken and then left for 10 minutes to ensure sufficient cobalt albumin binding. Fifty microliters of dithiothreitol (DTT) (Sigma, USA 1.5 mg/ml H₂O) was added as a colorizing agent. After 2 min the reaction was quenched with the addition of 1.0 mL of 0.9% NaCl. Colorimetric control specimens were prepared for preoperative and postoperative serum samples with 50 µL of distilled water replacing 50 µL of 1.5 mg/ml DTT. Specimen absorbencies were analyzed at 470 nm on a spectrophotometer (Shimadzu UV1601, Ausburn, Australia). The color of the specimens with DTT was compared against the colorimetric control tubes. The results were expressed as absorbance units (ABSUs).

**Measurement of Total Oxidant Status (TOS)**

TOS levels were determined using a method previously described by Erel (18). Serum TOS levels were calculated in µmol H₂O₂ equivalent/L.

**Total antioxidant status levels**

Total antioxidant status levels were calculated based on the method described by Erel (19) and expressed as mmol Trolox equivalent/L.

**Calculation of oxidative stress index (OSI)**

OSI was calculated as the TOS:TAS ratio. TAS units in the form of mmol Trolox equivalent/L were converted to µmol Trolox equivalent/L, with OSI being determined using the formula

\[ OSI = \frac{[\text{TOS}, \text{µmol H}_2\text{O}_2 \text{equivalent/L}]}{[\text{TAS}, \text{µmol Trolox equivalent/L}]} \times 100 \]

**Statistical analysis**

Kruskal-Wallis analysis of variance (Bonferroni-corrected Mann Whitney U test) was used to compare the groups. P<0.05 was regarded as statistically significant.

**RESULTS**

**Histopathological Findings**

Common parameters in all tissues, sinusoidal area (µm²), tears in the tunica albuginea and injury in the sinusoidal endothelium were investigated. Assessment was performed semi-quantitatively, with findings being scored between 0 and 3 (0: None, 1: Mild, 2: Moderate, 3: Severe). Distribution of histopathological parameters in all groups is shown in Table 1. No significant difference was determined between the groups in terms of sinusoidal area (µm²) (p>0.05). Significantly more tears in the tunica albuginea were determined in the P/R group compared to the control group (p<0.05). Comparing the P/R+D and P/R groups, dipyridamole resulted in significantly fewer tears in the

| Groups                | Control     | P/R        | P/R+DMSO   | P/R+D     | p     |
|-----------------------|-------------|------------|------------|-----------|-------|
| Sinusoidal area (µm²) | 3550±1096   | 4959±1187  | 4349±600.6 | 3550±393.9| 0.0312|
| Tunica albuginea tear | 0.333±0.5164| 2.5±0.5477 | 0.833±0.4082| 1.167±0.4082| 0.0005|
| Endothelial injury    | 0.166±0.4082| 2.33±0.5164| 0.5±0.5477 | 1.5±0.5477| 0.0005|
tunica albuginea (p<0.05). In terms of endothelial injury, significantly more damage was determined in the P/R group compared to the control group (p<0.05). Comparing the P/R+D and P/R groups, dipyridamole caused a decrease in endothelial injury, but the difference was not statistically significant (p>0.05).

A normal corpus cavernosum and corpus spongiosum appearance was observed in the control group. Normal morphology was also observed in the surrounding tunica albuginea and the external surrounding superficial fascia. Histological data for vascular spaces and the surrounding smooth muscle layer were normal (Figure-3A). In the P/R group, tears in the tunica albuginea surrounding the corpus spongiosum and connective tissue irregularities were present. The urethral epithelium had a normal appearance. Damage was observed in the endothelial cells lining vascular spaces in the corpus cavernosum. Sinusoidal areas had decreased in size (Figure-3B). In the P/R+DMSO group, a normal appearance was observed in the urethral structure and the tunica albuginea around the corpus spongiosum and corpus cavernosum. The vascular endothelium around the corpus cavernosum had a normal morphology (Figure-3C). In the P/R+D group, the tunica albuginea surrounding the corpus spongiosum and urethral epithelium had a normal appearance. Although occasional damage was observed in the endothelial cells around the vascular spaces in the corpus spongiosum, a close to normal appearance was observed in most of the area (Figure-3D).

Biochemical Findings
Distribution of biochemical parameters in all groups is shown in Table-2. Although an increase was observed in MDA values in the P/R group, no statistically significant difference in MDA was observed among the groups (p>0.05). Although an increase in serum IMA levels was observed in rats in the P/R group, no significant improvement in IMA levels was observed in the P/R+D group (p>0.05). A significant increase in TOS values was observed in the P/R+D group compared to the P/R group (p<0.05). A significant increase in terms

Figure 3 - Histopathological examination of the penis from each experimental group (Masson’s trichrome staining x200). A: Group 1, B: Group 2, C: Group 3, D: Group 4.
of TAS was observed in the P/R group compared to the control group (p<0.05), while a decrease was observed in the P/R+D group compared to the P/R group (p>0.05). OSI values were similar in the control and P/R groups (p>0.05), while an increase was observed in the P/R+D group compared to the P/R group (p<0.05).

**DISCUSSION**

Priapism may be defined as a prolonged erection due to disturbance of the mechanisms that control penile tumescence, rigidity and flaccidity (21). Ischemic priapism refers to a painful and continuing rigid erection characterized by little or no cavernous blood flow. Hypoxia, hypercarbia and acidosis develops in cavernous blood gas (22).

Ultrastructural changes in smooth muscle occur after 12 h in ischemic priapism, focal necrosis after 24 hs and finally necrosis and transformation of fibroblast-like cells after 48 h. If priapism is untreated or treated late (>24 hs) necrosis in cavernous smooth muscle, irreversible corporal fibrosis and permanent erectile dysfunction occur (4). The most significant determining factor in the prevention of tissue damage after ischemic priapism is duration of ischemia, which is directly correlated with reperfusion injury. Several studies have investigated I/R injury in different tissues (6, 11, 23). The purpose of this study was to investigate early histological and biochemical changes and the probable protective effects against I/R injury of dipyridamole before the emergence of irreversible damage following 1 h acute ischemic priapism in a model of ischemic priapism experimentally induced in rats.

Dipyridamole is a platelet inhibitor that increases cAMP levels by inhibiting the enzyme phosphodiesterase in platelets. It blocks re-absorption of adenosine by cells and increases interstitial adenosine concentrations (24, 25). It causes vasodilation by increasing adenosine formation, and probably improves tissue perfusion through a combination of its antiplatelet and vasodilator effects (10). Various studies have investigated the effects of dipyridamole on I/R injury. In a study of the antioxidant properties of dipyridamole, Vargas et al. reported that dipyridamole scavenges reactive oxygen radicals (ROS) released by human polymorphonuclear leukocytes (26). Iuliano et al. reported that dipyridamole possesses antioxidant properties (27). An experimental study determined that dipyridamole protects liver cells against warm I/R injury (28). Karagüzel et al. reported that the use of dipyridamole before testicular reperfusion in testicular torsion protects the testis against the long-term effects of I/R injury (11). The intraperitoneal 10 mg/kg dose of dipyridamole used in that study of testicular torsion was also employed in our priapism model which is another model of I/R injury. It may be possible for further studies to use different doses of dipyridamole to show increased effectiveness.

Very few experimental drug studies have investigated protective efficacy against I/R injury occurring in priapism (14, 29). Ours is the first study in the literature to investigate the effect of dipyridamole in an experimental priapism model. Although dipyridamole is a molecule that is safely used in various indications because of its anti-

| Table 2 - Groups’ biochemical data. |
|-------------------------------------|
| Groups               | Control | P/R    | P/R+DMSO | P/R+D | p       |
|----------------------|---------|--------|----------|-------|---------|
| TOS (µmol H₂O₂ Eq/L) | 11.06±2.224 | 10.08±3.029 | 22.06±7.188 | 19.44±4.715 | 0.0008 |
| TAS (mmol/L)         | 2.40±0.1754   | 2.80±0.140 | 2.74±0.161 | 2.49±0.209  | 0.0075 |
| OSI                  | 0.46±0.102     | 0.35±0.104 | 0.79±0.226 | 0.79±0.233  | 0.0012 |
| Serum MDA (nmol/L)  | 0.307±0.044     | 0.3865±0.088 | 0.3515±0.089 | 0.3803±0.061 | 0.2245 |
| IMA (ABSU/gr)        | 0.739±0.051     | 0.757±0.011 | 0.8015±0.036 | 0.7315±0.044 | 0.0452 |
Protective effects of DiPyriDamole on Priapism

-aggregating properties in humans, there are no studies concerning its use in humans with clinical priapism. There is a strong probability that as a result of this and future studies, dipyridamole will be used to prevent potential I/R injury in priapism, a new indication in humans, through investigation of the most appropriate and effective dosage.

Although histopathological evaluation of tunica albuginea and endothelial injury parameters in this study revealed ischemic changes, no change in favor of ischemia was determined in the levels of serum IMA and MDA and other biochemical parameters. Although a decrease was determined in TAS and IMA values in the group receiving dipyridamole compared to the P/R group, this was not statistically significant. Biochemical data not exhibiting significant differences in ischemic priapism, a compartment syndrome analogue, may be attributed to the 30 min reperfusion time we used being too short for ischemic tissue products to enter the circulation with reperfusion and establish a systemic effect. A longer reperfusion process is needed for penile ischemic products to exhibit systemic effects and for significant changes to be identified in ischemic parameters in serum. This study represents the first investigation in a priapism model of IMA, a biochemical parameter that has been investigated and found to be of value in such different models of I/R injury as testicular torsion (30, 31).

The effects of I/R injury occurring in priapism and the probable protective effects of dipyridamole were investigated through histological examination of the sinusoidal area, tears in the tunica albuginea and damage parameters in the sinusoidal endothelium. Examination of I/R injury in the P/R group revealed tears in the tunica albuginea surrounding the penis corpus spongiosum and irregularities in connective tissue, injury to the endothelial cells around the vascular spaces in the penis corpus cavernosum and contractions in sinusoidal areas. Examination of the penises of rats given dipyridamole revealed improvement in tunica albuginea and endothelial injury and showed that dipyridamole, which has antioxidant and anti-inflammatory properties, exhibits histopathological protective effects in I/R injury caused by priapism.

A certain amount of I/R injury will develop in penis corpus cavernosum tissue in patients with priapism due to various etiological causes, even if treated in the early period. Further studies are needed on the subject of the clinical outcomes that I/R may cause in the long-term. This study represents one of the rare examples of medical therapies that can be used in order to reduce I/R injury that may occur in patients treated in the early period.

The main limitation of this study is that the length of ischemia in the model of priapism used was limited to 1 hour. More significant results in terms of biochemical data may be obtained with a longer duration of ischemia. Another limitation is the difficulty in applying the animal model used in this study to humans and in determining appropriate dose selection.

CONCLUSIONS

In conclusion, this experimental study shows that the application of dipyridamole before reperfusion in a model of ischemic priapism has potential protective effects against histopathological injury that may develop in the penis. Further and more comprehensive studies are now needed in order to prevent I/R injury in ischemic priapism.

CONFLICT OF INTEREST

None declared.

REFERENCES

1. Burnett AL, Bivalacqua TJ. Priapism: new concepts in medical and surgical management. Urol Clin North Am. 2011; 38:185-94.
2. Montague DK, Jarow J, Broderick GA, Dmochowski RR, Heaton JP, Lue TF, et al. American Urological Association guideline on the management of priapism. J Urol. 2003; 170:1318-24.
3. Eland IA, van der Lei J, Stricker BH, Sturkenboom MJ. Incidence of priapism in the general population. Urology. 2001; 57:970-2.
4. Costa WS, Felix B, Cavalcanti AG, Medeiros J Jr, Sampaio FJ. Structural analysis of the corpora cavernosa in patients with ischaemic priapism. BJU Int. 2010; 105:838-41.
5. Muneer A, Minhas S, Freeman A, Kumar P, Ralph DJ. Investigating the effects of high-dose phenylephrine in the management of prolonged ischaemic priapism. J Sex Med. 2008; 5:2152-9.
6. Munarriz R, Park K, Huang YH, Saenz de Tejada I, Moreland RB, Goldstein I, et al. Reperfusion of ischemic corporal tissue: physiologic and biochemical changes in an animal model of ischemic priapism. Urology. 2003; 62:760-4.
7. Cuzzocrea S, Riley DP, Caputi AP, Salvemini D. Antioxidant therapy: a new pharmacological approach in shock, inflammation, and ischemia/reperfusion injury. Pharmacol Rev. 2001; 53:135-59.
8. Lysiak JJ, Nguyen QA, Kirby JL, Turner TT. Ischemia-reperfusion of the murine testis stimulates the expression of proinflammatory cytokines and activation of c-jun N-terminal kinase in a pathway to E-selectin expression. Biol Reprod. 2012; 69:202-10.
9. Chakrabarti S, Blair P, Wu C, Freedman JE. Redox state of dipyridamole is a critical determinant for its beneficial antioxidant and antiinflammatory effects. J Cardiovasc Pharmacol. 2007; 50:449-57.
10. Chakrabarti S, Vitseva O, Iyu D, Varghese S, Freedman JE. The effect of dipyridamole on vascular cell-derived reactive oxygen species. J Pharmacol Exp Ther. 2005; 315:494-500.
11. Karagüzel E, Kutlu Ö, Yuluğ E, Mungan S, Kazaz İO, Tok DS, et al. Comparison of the protective effect of dipyridamole and acetylsalicylic acid on long-term histologic damage in a rat model of testicular ischemia-reperfusion injury. J Pediatr Surg. 2012; 47:1716-23.
12. Hong Y, Liao WS, Mo RX. [Dipyridamole preconditioning protects against ischemia/reperfusion injury of rat liver]. Zhongguo Wei Zhong Bing Ji Jiu Yi Xue. 2006; 18:425-7.
13. Figueredo VM, Diamond I, Zhou HZ, Camacho SA. Chronic dipyridamole therapy produces sustained protection against cardiac ischemia-reperfusion injury. Am J Physiol. 1999; 277:H2091-7.
14. Sanli O, Armagan A, Kandirali E, Ozerman B, Ahmedov I, Solakoglu S, et al. TGF-beta1 neutralizing antibodies decrease the fibrotic effects of ischemic priapism. Int J Impot Res. 2004; 16:492-7.
15. Bradbury P, Rae K. Connective tissues and stains, in Bancroft JD and Stevens A (eds): Theory and Practice of Histological Techniques Churchill Livingstone Company, 1996; pp. 129.
16. Yagi K. Lipid peroxides and related radicals in clinical medicine, in Armstrong D (ed): Free Radicals in Diagnostic Medicine. New York, Plenum Press, 1994; pp. 1-15.
17. Bar-Or D, Lau E, Winkler JV. A novel assay for cobalt-albumin binding and its potential as a marker for myocardial ischemia-a preliminary report. J Emerg Med. 2000; 19:311-5.
18. Erel O. A new automated colorimetric method for measuring total oxidant status. Clin Biochem. 2005; 38:1103-11.
19. Erel O. A novel automated method to measure total antioxidant response against potent free radical reactions. Clin Biochem. 2004; 37:112-9.
20. Aycicek A, Erel O, Kocyigit A. Decreased total antioxidant capacity and increased oxidative stress in passive smoker infants and their mothers. Pediatr Int. 2005; 47:635-9.
21. Broderick GA, Kadioglu A, Bivalacqua TJ, Ghanem H, Nehra A, Shamlov R. Priapism: pathogenesis, epidemiology, and management. J Sex Med. 2010; 7:476-500.
22. Broderick GA, Harkaway R. Pharmacologic erection: time-dependent changes in the corporal environment. Int J Impot Res. 1994; 6:9-16.
23. Mark LA, Robinson AV, Schulak JA. Inhibition of nitric oxide synthase reduces renal ischemia/reperfusion injury. J Surg Res. 2005; 129:236-41.
24. Harker LA, Kadatz RA. Mechanism of action of dipyridamole. Thromb Res Suppl. 1983; 4:39-46.
25. Schaper W. Dipyridamole, an underestimated vascular protective drug. Cardiovasc Drugs Ther. 2005; 19:357-63.
26. Vargas F, Rivas C, Díaz Y, Contreras N, Silva A, Ejeda LE, et al. Antioxidant properties of dipyridamole as assessed by chemiluminescence. Pharmacaz. 2003; 58:817-23.
27. Iuliano L, Ghiselli A, Alessandri C, Bonavita MS, Violi F. Superoxide anion scavenging property of dipyridamole. Thromb Haemost. 1989; 61:149.
28. Taniguchi M, Magata S, Suzuki T, Shimamura T, Jin MB, Iida J, et al. Dipyridamole protects the liver against warm ischemia and reperfusion injury. J Am Coll Surg. 2004; 198:758-69.
29. Uluocak N, Atilgan D, Erdemir F, Parlaktas BS, Yasar A, Erkorkmaz U, et al. An animal model of ischemic priapism and the effects of melatonin on antioxidant enzymes and oxidative injury parameters in rat penis. Int Urol Nephrol. 2010; 42:889-95.
30. Kutlu O, Mentese A, Turkmken S, Turedi S, Gunduz A, Yulug E, et al. Investigation of the possibility of using ischemia-modified albumin in testicular torsion: an experimental study. Fertil Steril. 2011; 95:1333-7.
31. Turkmken S, Mentese A, Karagüzel E, Karaca Y, Kucuk A, Uzun A, et al. A comparison of the effects of N-acetylcysteine and ethyl pyruvate on experimental testicular ischemia-reperfusion injury. Fertil Steril. 2012; 98:626-31.

Correspondence address:
Ersagun Karagüzel, MD
Karadeniz Technical University, School of Medicine
Department of Urology
61080, Trabzon, Turkey
Telephone: +90 462 377-5908
E-mail: ersagunkaraguzel@gmail.com