Insights into modification of lignocellulosic fillers with isophorone diisocyanate: structure, thermal stability and volatile organic compounds emission assessment

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Received: 21 December 2019 / Published online: 4 October 2020 © The Author(s) 2020

Abstract
This study presents an analysis of the structure and properties of different types of lignocellulosic fillers modified by isophorone diisocyanate (IPDI) to provide insights into the possibility of their application to the manufacturing of wood polymer composites (WPCs). Moreover, it deals with the environmental aspects of modified fillers, by assessment of volatile organic compounds (VOCs) emitted during modification, as well as from final products. Three types of commercially available lignocellulosic fillers were modified with different content of IPDI (from 1 to 15 wt%) using the internal mixer. The main compounds detected in the air during modifications were terpenes and terpenoids. No IPDI was detected, which is very beneficial considering its toxicity. On the other hand, IPDI was emitted from modified fillers at a slightly elevated temperature of 40 °C, which was associated with a significant rise in its vapor pressure. Successful modification of fillers was confirmed by FTIR spectroscopy and thermogravimetric analysis. Performed modifications of lignocellulosic fillers with IPDI had a relatively small impact on their particle size and color properties. Such an effect can be considered beneficial for the mechanical performance and appearance of WPCs, which could be potentially manufactured using IPDI-modified lignocellulosic fillers.

1 Introduction
One of the biggest challenges in the manufacturing of wood polymer composites (WPCs) is the improvement of interfacial interactions between hydrophilic lignocellulosic fillers and often hydrophobic polymer matrix, which are crucial for the performance of WPCs. Numerous ways to enhance these interactions have been developed, which often include different filler treatment methods, such as silanization, mercerization, acetylation, maleic anhydride treatment, esterification and etherification, isocyanate grafting, as well as a modification with different surfactants and plasticizers (Błędzki et al. 2008; Colom et al. 2003). Such treatments can significantly improve the characteristics of obtained WPCs. However, they may cause burdens for the environment, for example, due to the use of organic solvents or emission of chemicals during modification and storage of modified fillers (Dányádi et al. 2010). Unfortunately, this issue is usually omitted in the literature.

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Among the applied methods of chemical modification of lignocellulosic fillers, one that is commonly considered as toxic and non-environmentally friendly, is the isocyanate treatment. Isocyanates are very interesting for such applications because they can create stable urethane bonds with hydroxyls present on the surface of lignocellulosic fillers, but also with functional groups present in chains of various polymers such as polyurethanes, poly(e-caprolactone) or other polar polymers. Because of the high reactivity of free isocyanate groups, they are categorized as irritant, toxic, and hazardous towards humans and the environment (Musk et al. 1988). However, it is only related to their unbonded form, when the isocyanate group is bonded and reacted with other functional groups, for example, hydroxyl or amine groups, no toxic effects are observed. Therefore, it is possible to use polyurethane materials in the manufacturing of biomedical grade articles, such as vascular catheters, blood bags, or implants (Zdrahala and Zdrahala 1999). A similar case is observed for the introduction of isocyanate-modified fillers into WPCs, when the toxic effect should be eliminated (or at least significantly reduced), due to the excess of hydroxyl groups of fillers. Moreover, they are already commonly used in the plastics industry for a very long time. Hence, they are well known and analyzed. As a group, isocyanates have been repeatedly proven as excellent coupling agents for polyolefin-based biocomposites, providing properties superior to those compatibilized with anhydrides or silanes (Ashori and Nourbakhsh 2009; Geng et al. 2005).

One of the first works related to filler modification with isocyanates was published about 20 years ago by the research group of Maldas et al. (Maldas et al. 1988, 1989; Raj et al. 1989, 1990), who investigated the modification of wood sawdust and various types of pulps from paper industry with isocyanates was published about 20 years ago by the research group of Maldas et al. (Maldas et al. 1988, 1989; Raj et al. 1989, 1990), who investigated the modification of wood sawdust and various types of pulps from paper industry with isocyanates and their further impact on potential applications.

### 2 Experimental

#### 2.1 Materials

Three types of commercially available Arbocel® fillers from JRS J. Rettenmaier & Söhne GmbH (Germany) were used in the present study: B400, CW630PU, and UFC100. In Table 1, the properties of the applied fillers are listed and Fig. 1 shows microscopic images of the applied fillers.

Fillers were modified with isophorone diisocyanate characterized by a content of isocyanate groups of 37.5%. IPDI was acquired from Merck Millipore.

#### 2.2 Modification of fillers

Fillers were modified using GMF 106/2 Brabender batch mixer at room temperature (varied from 21.1 to 23.1 °C) and rotor speed of 100 rpm. Room temperature was applied

| Filler type | Properties of applied Arbocel® fillers |
|-------------|----------------------------------------|
| B400        | CW630PU | UFC100 |
| Whitehead, %| 86      | 60      | 85      |
| Average particle length, μm | 900      | 40      | 8       |
| Average particle thickness, μm | 20      | 20      | 2       |
| Aspect ratio L/D | 45      | 2       | 4       |
| Bulk density, g/l | 30      | 205     | 160     |
| Moisture content, wt% | 6.01    | 6.81    | 4.84    |
to reduce the possible emissions from an applied diisocyanate, which at 20 °C shows relatively low volatility—vapor pressure equals $4.0 \times 10^{-2}$ Pa. At higher temperatures, its volatility significantly increases. When the temperature is elevated to 50 °C, the vapor pressure is increased 22.5 times. A proper amount of fillers was placed in an internal mixer (amount varied for different fillers, which was associated with different densities) with a calculated amount of isocyanate—0; 1.0; 2.5; 5.0; 10.0 or 15.0 wt% respectively to the mass of fillers. Mixing was performed for 5 min. Then, the samples were put in zipper storage bags.

### 2.3 Measurements and analyte sampling equipment

The air quality screening studies performed in an enclosed area were carried out with the use of Radiello® diffusive passive samplers (Fondazione Salvatore Maugeri, Padova, Italy). The application of a passive sampling technique facilitates collection of the analytes from the gaseous phase without any additional equipment such as aspirators, pumps, or additional gas wires, which allows performing the research without disturbing the regular use of defined enclosed area or working place. Detailed information about the characteristics and technical parameters of the Radiello® diffusive passive sampler is listed elsewhere (Marć et al. 2014a, b; Plaisance et al. 2008). In brief, the exposure time of applied passive samplers in enclosed space was set up to 4 h. The short sampling period was associated mainly with the indoor environment conditions (the possibility of high or very high content level of organic compounds in gaseous phase due to the specific character of the indoor sampling area that might “overload” the sorption medium installed inside the diffusive porous polyethylene membrane). Each time, two independent Radiello® passive samplers were installed, at a distance of more than 1 m to ensure optimal conditions for collecting analytes from the gaseous phase (adequate air circulation around the passive sampler, as well as avoiding the phenomenon of competitiveness between installed passive samplers). The passive samplers were installed very close to the workplace and in close vicinity to the materials preparation area. During the passive sampling working period, the temperature inside the studied area was constantly monitored and was in the range from 21.1 up to 23.1 °C. After the sampling period, the cylindrical tubes with sorption medium (Carbotrap 4) were removed from the diffusive membrane and enclosed in the glass containers. Next, the sorption medium with adsorbed analytes was transported to the laboratory and stored at a temperature of 4 °C no longer than 12 h after sampling. The identification of the main volatile organic compounds in the air of the enclosed space was performed with the use of thermal desorption technique (Unity v.2, Markes International Ltd., Pontyclun, UK), connected with gas chromatography (Agilent Technologies 6890) combined with a mass spectrometer (5873 Network Mass Selective Detector, Agilent Technologies). The cylindrical containers filled with sorption medium were desorbed at a temperature of 290 °C for 15 min. During this process, the analytes were transported to the microtrap. Then, the analytes were liberated from the microtrap (ballistic heating up to 300 °C and maintained for 5 min) and transported by the helium flow (1.5 mL min$^{-1}$) to the GC capillary column (Agilent 122-5563, J&W DB-5MS, 60 m × 0.25 mm × 1 µm). The GC oven temperature program was as follows: 50 °C for 1 min, then raised at a rate of 10 °C min$^{-1}$ up to 280 °C and maintained for 10 min. The temperature of the TD-GC transfer line was set to 150 °C, and the temperature of the GC–MS transfer line was 280 °C. The employed mass spectrometer was working in a SCAN mode. Detailed information about the principals and characteristic of the thermal desorption technique is presented elsewhere (Król et al. 2012; Zabiegała et al. 2007). To eliminate signals associated with other processes occurring in the laboratory hall and select only signals related to the performed modification, as well as to ensure the reliability of obtained results, the “background” was also analyzed. Additionally,
before each sampling period, the sorption medium placed in the cylindrical tube was conditioned for 30 min at a temperature of 300 °C.

The emission studies (expressed by TVOCs parameter) were performed using the microscale stationary emission chamber system (Markes International Micro-Chamber/Thermal Extractor M-CTE250) consisting of four stainless steel chambers with the internal volume of 114 cm³. Detailed information about the characteristic of the stationary emission chamber system is described elsewhere (Marć and Zabiegała 2017; Marć et al. 2017). In brief, the sample of modified lignocellulosic fillers (mass of a sample—1.073 ± 0.059 g) was firstly placed on a Petri glass dish and then installed inside the chamber to prevent chamber contamination with small particles of solid samples. Next, the chambers were tightly closed, and at the head of the cover, the stainless steel tube filled with sorption medium Tenax TA was installed to collect the analytes emitted from the surface of the studied samples to the gaseous phase. Then, the sampling conditions were set: samples seasoning time—20 min; inert gas (nitrogen) flow rate—15 mL min⁻¹; seasoning temperature—40 °C. After the sampling process, the stainless steel tubes were removed and transferred directly to the thermal desorber (Mareks Int, Unity 2) connected with gas chromatograph (Agilent Technologies 7820A) combined with flame ionization detector. The stainless steel tubes filled with Tenax TA were desorbed at a temperature of 285 °C for 12 min. During this process, the analytes were transported to the microtrap. Then, similar to the TD-GC–MS system, the analytes were liberated from the microtrap (ballistic heating up to 300 °C and maintained for 5 min) and transported by the helium flow (2.0 mL min⁻¹) to the GC capillary column (J&W, DB-1, 30 m × 0.32 mm × 5 µm). The GC oven temperature program was as follows: 45 °C for 1 min, then raised at a rate of 15 °C min⁻¹ up to 120 °C maintained for 2 min, and then raised at a rate of 10 °C up to 250 °C and held for 5 min. The temperature of the TD-GC transfer line was set to 160 °C, and the FID temperature was 250 °C. Before every sampling period, the chambers with the glass Petri dish inlet were conditioned at a temperature of 100 °C for 30 min, and the blank sample (background value) was measured to ensure the reliability of the obtained results. Additionally, before each sampling period, the Tenax TA stainless steel tubes were conditioned for 30 min at a temperature of 300 °C. The most important information about the calibration process of the mentioned TD-GC-FID system and the assessment of the basic validation parameters is described in detail in previous papers (Marć and Zabiegała 2017; Marć et al. 2017). The values of the TVOC parameter were assessed based on the toluene equivalent. The recovery values of toluene from the applied Tenax TA sorption resin ranged from 95 up to 105%. As for the LOD, the value of this parameter was estimated based on the characteristic of the prepared calibration curve. The calculated value of LOD for toluene was 0.040 ng, and the LOQ value (3 × LOD) was 0.12 ng.

The particle size distribution of modified lignocellulosic fillers was characterized using a laser particle sizer Fritsch ANALYSETTE 22 apparatus operated in the range of 0.08–2000 µm.

Scanning electron microscopy (SEM) was applied in order to evaluate the changes on the fillers’ surface as a result of performed modifications. The scanning electron microscope (SEM)—model MIRA3—from Tescan (Brno, Czech Republic), was used to assess the structure of the external and internal surfaces of the rotationally molded products. The structures of the surfaces of the rotationally molded samples were assessed at an accelerating voltage of 1 kV.

The chemical structure of lignocellulosic fillers’ samples was determined using Fourier transform infrared spectroscopy (FTIR) analysis performed by a Nicolet Spectrometer IR200 from Thermo Scientific (USA). The device had ATR attachment with a diamond crystal. Measurements were taken with 1 cm⁻¹ resolution in the range from 4000 to 400 cm⁻¹ and 64 scans.

The color of ground organic powders was evaluated according to the Commission Internationale de l’Eclairage (CIE) through L*a*b* coordinates (International Commission on Illumination 1978). In this system, L* is the color lightness (L* = 0 for black and L* = 100 for white), a* is the green (−)/red (+) axis, and b* is the blue (−)/yellow (+) axis. Thirty tests of each sample were done and used for the determination of arithmetic mean values. The color was determined by optical spectroscopy using HunterLab Miniscan MS/S-4000S spectrophotometer, placed additionally in a specially designed light trap chamber. The total color difference parameter (ΔE*) was calculated according to the following formulation (1) (Bociąga and Trzaskalska 2016):

\[
\Delta E^* = \left(\frac{\Delta L^*}{2} + (\Delta a^*)^2 + (\Delta b^*)^2\right)^{0.5}
\]

The thermal analysis was performed using the TG 209 F3 apparatus from Netzsch (Germany). Samples of fillers weighing approximately 10 mg were placed in a ceramic dish. The study was conducted in an inert gas atmosphere—nitrogen in the range from 30 to 900 °C with a temperature increase rate of 10 °C min⁻¹. Two specimens were analyzed for each sample.

### 3 Results and discussion

#### 3.1 VOCs emission during filler modification

Determination of volatile organic compounds emissions during various chemical processes is essential for human safety, though not very often investigated and described in
the literature. To develop a sustainable process and fully evaluate its benefits and drawbacks for its optimization, it is crucial to determine its impact on the environment. Therefore, during the conducted modification of lignocellulosic fillers, passive samplers were used to collect generated VOCs and subsequently analyze them with chromatographic

| Compound     | Formula | Chemical structure | Flash point, °C | NFPA 704 codes | GHS pictograms |
|--------------|---------|--------------------|-----------------|-----------------|----------------|
| Octanal      | C₈H₁₆O  |                    | 51              | 2 0 0           |                |
| o-Cymene     | C₁₀H₁₄  |                    | 50              | 2 1 0           |                |
| m-Cymene     | C₁₀H₁₄  |                    | 47              | 2 1 0           |                |
| p-Cymene     | C₁₀H₁₄  |                    | 47              | 2 1 0           |                |
| Myrtenal     | C₁₀H₁₄O |                    | 78              | 2 1 0           |                |
| Verbenone    | C₁₀H₁₄O |                    | 85              | 2 2 0           |                |
| α-Pinene     | C₁₀H₁₆  |                    | 33              | 3 1 0           |                |
| β-Pinene     | C₁₀H₁₆  |                    | 36              | 3 1 0           |                |
| Camphene     | C₁₀H₁₆  |                    | 34              | 3 2 1           |                |
| 2-Carene     | C₁₀H₁₆  |                    | 38              | 2 2 0           |                |
| 3-Carene     | C₁₀H₁₆  |                    | 46              | 2 2 0           |                |
techniques. In Table 2, the main compounds generated during processing are summarized.

For all compounds, temperatures of flash point, as well as their codes and pictograms are presented according to NFPA 704: Standard System for the Identification of the Hazards of Materials for Emergency Response and Globally Harmonized System of Classification and Labelling of Chemicals, which are commonly used for classification of chemicals by the risk they pose towards human health and safety (European Parliament and the Council 2008; NFPA 2017).

NFPA 704 system is commonly known as “safety square” or “fire diamond” and is applied to quickly and easily identify the risks and select the proper procedure for handling chemicals. It is developed by National Fire Protection Association from the United States and assesses flammability, health, and instability of chemicals on a scale from 0 (minimal hazard) to 4 (severe hazard), also providing information about special hazards related to the material.

Globally Harmonized System of Classification and Labelling of Chemicals is an internationally agreed standard, which includes criteria for testing hazards and harmonized safety data sheets providing information about chemicals. It uses a combination of hazard statements and pictograms describing the physical, health and environmental properties of materials and hazards they pose to the environment.

During the modification of lignocellulosic fillers, the presence of terpenes and terpenoids was detected. The majority of them is not only considered as potentially hazardous to human health (causing skin and eye irritation or acting as allergens), but they are a serious environmental threat because of their flammability (Dev 1989). According to literature data, increasing the content of terpenes in various species of wood implicates higher fire threat (Ormeño et al. 2009). Therefore, their emission during modification of lignocellulosic materials, especially when performed on an industrial scale, should be monitored in order to prevent fire accidents.

It can be seen that almost all detected compounds are labeled as flammable according to the GHS system, and their flammability is rated as 2 or 3 on the NFPA scale, which indicates that they must be moderately heated or exposed to relatively high ambient temperature in order for ignition to occur. Rating 2 is related to flash points between 37.8 and 93.3 °C while rating 3 indicates a flash point between 22.8 and 37.8 °C (NFPA 2017). It means that during the modification of lignocellulosic materials, serious precautions have to be taken. Depending on the actual value of ambient temperature, which is associated with the latitude and season, even the processing of cellulose materials without external heating may be considered as severe fire threat, therefore the surrounding has to be secured.

It also is very important that isocyanates were not detected during modification. The presence of a moderate amount of other chemicals, for example BTEX hydrocarbons, styrene, or chlorinated hydrocarbons was noted. However, it was ascribed to their presence in the atmosphere in

| Compound    | Chemical Formula | NFPA Rating | Codes | Pictograms |
|-------------|------------------|-------------|-------|------------|
| 4-Carene    | C₁₀H₁₆            | 38          | 2 2   | 0          |
| α-Phellandrene | C₁₀H₁₆       | 46          | 2 1   | 0          |
| Terpinene   | C₁₀H₁₆            | 37          | 3 2   | 2          |
| Alloocimene | C₁₀H₁₆            | 69          | 2 1   | 0          |
| Limonene    | C₁₀H₁₆            | 50          | 2 3   | 0          |
| Camphor     | C₁₀H₁₆O           | 54          | 2 2   | 0          |
the laboratory during modification associated with storage and processing of various polymer materials, such as rubbers, polyurethanes or polyolefins. Detected compounds are listed in Table 3. The strongest signals were observed for toluene and tetrachloroethylene, which are used as solvent, but also associated with the major activities in the laboratory hall—manufacturing of rubbers and flame retarded polyurethane foams.

### Table 3 Compounds detected in the air during a background check of the laboratory hall

| Detected compound       | Origin                              | References |
|------------------------|-------------------------------------|------------|
| **Hydrocarbons**        |                                     |            |
| $n$-Pentane             | Polyurethane foams                  | Piszczyk et al. (2014) |
| Hexane                  | Acrylic plastics, Polypropylene     | Willoughby et al. (2003), Wolkoff (1988) |
| Methylcyclopentane      | Polypropylene                       | Reingruber et al. (2011) |
| Methylcyclohexane       | Polypropylene                       | Reingruber et al. (2011) |
| Benzene                 | SBR rubber                          | Yu and Crump (1998) |
| 2-Methylheptane         | Polypropylene                       | Willoughby et al. (2003) |
| Toluene                 | SBR rubber, Solvent                 | Yu and Crump (1998) |
| Ethylbenzene            | Polystyrene                         | Curran et al. (2016) |
| Xylenes                 | Polyester resins, SBR rubber, Solvent | Curran et al. (2016), Yu and Crump (1998) |
| Styrene                 | Polystyrene, Polyester resins, SBR rubber | Curran et al. (2016), Yu and Crump (1998) |
| Undecane                | Polyethylene, Polypropylene         | Lattuati-Derieux et al. (2013) |
| Dodecane                | Polyethylene, Polypropylene, Natural rubber | Curran et al. (2016), Nielsen et al. (1994) |
| Naphthalene             | Poly(vinyl chloride)                | Yu and Crump (1998) |
| Tridecane               | Poly(vinyl chloride), Polyethylene  | Yu and Crump (1998) |
| 3-Methyl tridecane      | Polyester resins, Polyethylene, Polypropylene | Lattuati-Derieux et al. (2013), Yu and Crump (1998) |
| Tetradecane             | Polyester resins, Polyethylene, Polypropylene | Lattuati-Derieux et al. (2013), Yu and Crump (1998) |
| 3-Methyl pentadecane    | Polyester resins, Polyethylene, Polypropylene | Lattuati-Derieux et al. (2013), Yu and Crump (1998) |
| **Chlorinated hydrocarbons** | Flame retarded polyurethane foams, Solvent | Hillier et al. (2003) |
| Methylene chloride      | Flame retarded polyurethane foams, Solvent | Hillier et al. (2003) |
| Trichloromethane        | Solvent                             | Stringer and Johnston (2001) |
| 1,2-Dichloroethane      | Solvent                             | Tsai et al. (2009) |
| Trichloroethylene       | Solvent                             | Stringer and Johnston (2001) |
| Tetrachloroethylene     | Flame retarded polyurethane foams, Solvent | Hillier et al. (2003) |
| Chlorobenzene           | Poly(ethylene terephthalate)        | Curran et al. (2016) |
| **Ketones**             |                                     |            |
| Acetone                 | Poly(ethylene terephthalate), Solvent | Curran et al. (2016) |
| 2-Heptanone             | Polyethylene                        | Villberg and Veijanen (2001) |
| **Aldehydes**           |                                     |            |
| Pentanal                | Natural rubber, Polyethylene        | Curran et al. (2016), Hoven et al. (2003), Villberg and Veijanen (2001) |
| Hexanal                 | Natural rubber, Polyethylene        | Curran et al. (2016), Hoven et al. (2003), Villberg and Veijanen (2001) |
| Heptanal                | Polyethylene                        | Villberg and Veijanen (2001) |
| Benzaldehyde            | Polyester resins                    | Curran et al. (2016) |
| **Others**              |                                     |            |
| Acetic acid             | Natural rubber, Polyethylene        | Curran et al. (2016), Hoven et al. (2003) |
| Ethyl acetate           | Solvent                             | Riemenschnieder and Bolt (2005) |
| Dimethylformamide       | Solvent                             | De Vasconcelos et al. (2001) |
| Benzothiazole           | Rubbers—vulcanization accelerators  | Reemtsma et al. (1995) |
| Diethyl phthalate       | Various plastics—plasticizer        | Hauser et al. (2004) |

### 3.2 VOCs emission from modified lignocellulosic fillers

Figure 2 shows total VOCs emission from various Arboce™ fillers depending on IPDI content. The analysis was performed at 40 °C. Clearly, the volume of emission strongly depends on the type and size of filler (900, 40, and 8 mm, respectively, for B400, CW630PU, and UFC100).
Qualitatively, very similar compounds were detected for all types of fillers, so in Fig. 3, an exemplary chromatogram obtained for CW630PU modified with 15 wt% of IPDI is presented. Traces of limonene (retention time of 13.53 min) were detected. However, the main compound emitted from modified fillers was by far IPDI (retention times of 21.65 and 21.97 min), whose emission strongly depends on temperature (vapor pressure at 50 °C is more than 30 times higher than at 20 °C). Therefore, it was detected despite its absence during modification (European Aliphatic Isocyanates Producers Association 2013). It accounted for around 41, 60, and 63% of the area below the peaks present on the chromatograms obtained for B400, CW630PU, and UFC100 fillers, respectively. Moreover, the presence of a moderate amount of other chemicals, for example BTEX hydrocarbons or solvents (e.g., acetone at 4.58 min, butanone at 5.64 min, or butanone oxime at 8.59 min) was noted. It was ascribed to their presence in the atmosphere in the laboratory during modification associated with the processing of various polymer materials.

### 3.3 Particle size distribution

In Fig. 4, the plots are showing the particle size distribution of modified fillers as a function of IPDI content. The analysis was performed for samples without IPDI and for those containing 1, 5, and 15 wt% of modifier. In the case of B400 fillers, the determination of particle size for the highest content of IPDI was impossible because of the very strong agglomeration of fillers after modification. Samples named 0% on the graphs were also mechanically treated in a Brabender mixer. Therefore, their average particle size differs from the values provided by the producer (see Table 1). It can be noted that the addition of isocyanate increased the average particle size, which is associated with the generation of urethane bonds, linking separate fillers. Such effect was expected due to the high reactivity of isocyanate functional groups of IPDI and the multiplicity of hydroxyl groups on the surface of lignocellulosic fillers. Nevertheless, in the case of CW630PU and UFC100 fillers, modification with an IPDI content not exceeding 5 wt% did not show a very significant impact on the particle size distribution. For B400 samples, the effect was more noticeable because the average particle size was increased from 278 to 344 μm. Nevertheless, no significant agglomeration was noted. Such an effect should be considered as very beneficial for further manufacturing of polymer composites because the size of filler has a noticeable influence on their performance.

### 3.4 Microscopic analysis of modified fillers

Figure 5 shows SEM images of the investigated fillers before and after the modification with IPDI. It can be clearly seen that the performed treatment resulted in the changes of the
fillers’ surface. After treatment, the surface became rougher, which is indicated by the circles in the SEM images. Such an effect can be associated with the chemical reactions occurring between diisocyanate modifier and hydroxyl groups present on the surface of fillers. As a result, diisocyanate was deposited on the surface and might also cause sticking of the smaller particles of cellulosic fillers. Nevertheless, microscopic analysis confirmed a rather low agglomeration effect, as indicated by the results of particle size analysis. Nevertheless, some changes in the aspect ratio of fillers may be noted, especially in case of CW630PU and UFC100 fillers. For relatively long fibers of B400 filler, the rise of particle size was rather related to the entanglements of fibers. In case of other, less fibrous, fillers, the drop of aspect ratio, especially for 15 wt% content of IPDI, was related to sticking of the particles to each other. It can be seen in Fig. 5f and i. Such an effect only slightly affected the average diameter, but particles changed their form from flake-like to more spherical shape.

3.5 Chemical structure

Figure 6 shows the spectra of three types of fillers modified with 15 wt% of IPDI. It can be seen that all spectra show an appearance typical for lignocellulosic materials. Signals between 3340 and 3355 cm\(^{-1}\) are characteristic for stretching vibrations of hydroxyl groups present on the surface of fillers. They all show relatively high intensity after modification, which is related to their overlapping with signals associated with stretching vibrations of N–H bonds, which are generated during the modification with isocyanate. In the range of 2880–2960 cm\(^{-1}\), peaks characteristic for symmetric and asymmetric stretching vibrations of C–H bonds can be noticed. Broad signals between 2250 and 2340 cm\(^{-1}\) were ascribed to the asymmetric stretching vibrations of free isocyanate groups, which was related to the performed modifications. Prominent peaks around 1740 cm\(^{-1}\) were attributed to the vibrations of carbonyl group present in urethane linkage between filler and IPDI. The presence of these peaks confirms the successful modification of lignocellulosic fillers. Other very noticeable peaks were observed in the range of 1010–1050 cm\(^{-1}\) and were characteristic for the vibrations of C–O bonds present in the structure of cellulose. Multiple signals noted in the range of 1100–1450 cm\(^{-1}\) were associated with the bending vibrations of C–H bonds, stretching of various C–O–C groups, and C–C bonds. The presence of a large amount of small signals in this range is typical for lignocellulosic fillers (Barczewski et al. 2018; Matykiewicz et al. 2019).

Figure 7 presents spectra of UFC100 fillers modified with varying amounts of isophorone diisocyanate. It can be seen that an increase in the modifier content results in a noticeable rise of particular signals associated with the presence of urethane linkages between fillers and isocyanate. The most significant impact of modifier was noted for signals around 2250–2340 and 1740 cm\(^{-1}\), related to
the unbonded isocyanate groups and C = O stretching in urethane groups. Similar effects were observed by other researchers for the isocyanate modification of cellulose fillers (Botaro and Gandini 1998; Siqueira et al. 2010). Moreover, a slight rise in the intensity and changes in the shape of the broad signal around 3350 cm\(^{-1}\) were noted.
which was associated with the reaction between hydroxyls present on the surface of fillers and isocyanate groups of the modifier. As mentioned above, it resulted in the generation of urethane groups, which also contain N–H bonds. Signals characteristic for them are practically overlapping with those of hydroxyl groups, hence only slight changes were noted (Gao et al. 2005).

### 3.6 Color assessment

In Table 4, the color parameters of the prepared samples determined during spectrophotometric analysis are presented. The color of materials was evaluated using CIELab color space and converted to popular Adobe RGB space. Moreover, values of total color difference parameter (ΔE*) associated with performed modifications, related to unmodified, reference samples, are presented, as well as colors of obtained materials.

It can be seen that applied modifications of lignocellulosic fillers with IPDI caused some changes in L* and b* parameters. The decrease in the lightness of fillers was associated with the increase in their average particle size. Such a phenomenon is commonly known and widely described in the literature by various researchers (Ahmed et al. 2014, 2015, 2016). Rise of b* parameter is related to

| Table 4 | Color properties of modified lignocellulosic fillers |
| --- | --- |
| **Filler type** | IPDI content, wt% | L* | a* | b* | ΔE* | R | G | B | Color |
| B400 | 0.0 | 93.55 | -0.62 | 4.45 | - | 237.8 | 235.9 | 227.5 |
| | 1.0 | 93.14 | -0.63 | 4.63 | 0.45 | 236.7 | 234.7 | 226.0 |
| | 2.5 | 93.46 | -0.69 | 4.52 | 0.12 | 237.6 | 235.6 | 227.1 |
| | 5.0 | 92.65 | -0.62 | 4.86 | 0.99 | 235.4 | 232.3 | 224.1 |
| | 10.0 | 91.56 | -0.63 | 5.47 | 2.24 | 232.6 | 230.0 | 219.8 |
| | 15.0 | 91.63 | -0.63 | 5.67 | 2.27 | 232.9 | 230.2 | 219.7 |
| CW630PU | 0.0 | 85.38 | 2.01 | 15.30 | - | 223.2 | 209.9 | 184.4 |
| | 1.0 | 85.17 | 2.04 | 15.17 | 0.25 | 222.5 | 209.3 | 184.1 |
| | 2.5 | 84.32 | 2.25 | 15.88 | 1.23 | 220.7 | 206.7 | 180.5 |
| | 5.0 | 83.84 | 2.31 | 15.93 | 1.69 | 219.4 | 205.3 | 179.1 |
| | 10.0 | 82.31 | 2.72 | 17.22 | 3.69 | 216.0 | 200.7 | 172.6 |
| | 15.0 | 81.28 | 3.05 | 18.24 | 5.15 | 213.9 | 197.5 | 168.0 |
| UFC100 | 0.0 | 96.46 | -0.04 | 1.91 | - | 245.6 | 244.3 | 240.8 |
| | 1.0 | 96.41 | -0.05 | 1.87 | 0.06 | 245.4 | 244.2 | 240.7 |
| | 2.5 | 96.31 | -0.04 | 1.95 | 0.16 | 245.2 | 243.9 | 240.3 |
| | 5.0 | 96.28 | -0.05 | 1.97 | 0.19 | 245.1 | 243.8 | 240.2 |
| | 10.0 | 96.12 | -0.01 | 2.54 | 0.72 | 245.0 | 243.3 | 238.6 |
| | 15.0 | 96.40 | -0.03 | 2.32 | 0.41 | 245.7 | 244.1 | 239.9 |
the yellowing of cellulose, as a result of mechanical and chemical treatment (Ahn et al. 2019).

However, overall, it can be seen that the performed modifications of fillers did not cause significant differences in their color, which was expressed by relatively low values of ΔE* parameter. Only for B400 and CW630PU fillers modified with higher contents of IPDI, the values were exceeding 2.0, indicating medium color variations, recognizable by the inexperienced observer, while distinct and significant color variations were noted only for CW630PU fillers (Bociąga and Trzaskalska 2016). Such information is very important for the potential industrial

Table 5 Moisture content of modified lignocellulosic fillers

| Filler type | IPDI content, wt% | Moisture content, wt% | Filler type | IPDI content, wt% | Moisture content, wt% | Filler type | IPDI content, wt% | Moisture content, wt% |
|-------------|-------------------|-----------------------|-------------|-------------------|-----------------------|-------------|-------------------|-----------------------|
| B400        | 0.0               | 6.28                  | CW630PU     | 0.0               | 7.10                  | UFC100      | 0.0               | 4.87                  |
|             | 1.0               | 6.18                  |             | 1.0               | 6.98                  |             | 1.0               | 4.49                  |
|             | 2.5               | 5.94                  |             | 2.5               | 6.76                  |             | 2.5               | 4.53                  |
|             | 5.0               | 5.46                  |             | 5.0               | 6.45                  |             | 5.0               | 4.36                  |
|             | 10.0              | 5.33                  |             | 10.0              | 6.44                  |             | 10.0              | 4.31                  |
|             | 15.0              | 4.82                  |             | 15.0              | 6.28                  |             | 15.0              | 4.18                  |
application of investigated modification process. The appearance of wood polymer composites is often crucial for the final recipients of the products, and it is significantly affected by the appearance of introduced fillers,

### 3.7 Thermal stability

Figure 8 presents the results of the thermogravimetric analysis of modified lignocellulosic fillers. It can be seen that thermal stability strongly depends on the type of filler. Obviously, the first step of decomposition, below 100 °C, is related to the moisture content of particular samples, whose values are presented in Table 5. Its varying values can be associated with differences in the chemical composition, as well as particle size, hence surface area of fillers. All those factors are significantly affecting the water absorption of lignocellulosic fillers. A drop in moisture content with increasing amount of applied modifier is associated with changes in the chemical structure of the surface and reduced polarity compared to unmodified samples.

Generally, it can be seen that B400 and UFC100 samples show quite similar behavior, while more significant differences are observed for CW630PU filler. Such effect is associated with the cellulose content, which for B400 and UFC100 filler is significantly higher than for CW630PU. All lignocellulosic fillers show quite a similar course of decomposition, and differences are associated mainly with moisture content. CW630PU fillers, despite the lowest thermal stability (highest moisture content), show the highest content of residue after analysis. Such an effect can be attributed to the lignin content, which decomposes quite slowly and produces a high yield of char (Brebu and Vasile 2010). Moreover, differential thermogravimetric (DTG) curves for lignin are rather flat, with no sharp peaks comparing to cellulose and hemicellulose, which can be confirmed by the presented results (Yang et al. 2006).

With the increase in IPDI content, slight changes in thermal decomposition are observed (Figs. 8 and 9). Especially in the case of the UFC100 sample, an additional peak is noted at DTG curves around 305 °C. Such effect is associated with thermal degradation of urethane bonds generated as a result of isocyanate groups of modifiers with hydroxyls present on the surface of lignocellulosic fillers (Hejna et al. 2017). Moreover, char residue generated during degradation of urethane bonds, containing noticeable amounts of nitrogen, may partly act as an inhibitor of further degradation. Hence, peaks on DTG curves are slightly shifted towards higher temperatures.

### 4 Conclusion

Structure and properties of three types of lignocellulosic fillers modified with isophorone diisocyanate were investigated to determine its potential for application as a dispersed phase for wood polymer composites. Fillers were modified with different contents of IPDI (from 1 to 15 wt%) using the internal mixer. The emission of volatile organic compounds during modification was analyzed. The main detected compounds were terpenes and terpenoids, which, despite their lower harmfulness compared to IPDI, show irritating and allergic character. Moreover, they are characterized by relatively low values of flash point, even below 40 °C. Therefore, precautions need to be taken during modifications of lignocellulosic fillers.

At a slightly elevated temperature of 40 °C, IPDI was detected among VOCs emitted from modified fillers (vapor pressure of IPDI significantly increases with temperature). It suggests that part of the applied modifier was only deposited on the surface of the filler, which was confirmed by FTIR analysis, indicating the presence of free isocyanate groups in the structure of modified fillers. On the other hand, the generation of urethane bonds on the surface of fillers was noted, confirming successful modification.

Regarding particle size, IPDI-treatment increased this parameter. For B400 filler, the addition of 1 and 5 wt% of modifier caused an increase from 278 μm to 344 and 363 μm, respectively. For other types of fillers, such contents of IPDI did not cause significant effects. Increasing the amount of modifier to 15 wt% resulted in the rise of particle size from 30 to 49 μm and from 8 to 25 μm, for CW630PU and UFC100 fillers, respectively. Such an effect was associated with slight agglomeration of particles. For the color of the lignocellulosic fillers, the effect of modification also was not very significant. Color differences, which could be considered as recognizable by the inexperienced observer were noted only for B400 and CW630PU fillers modified
with 10 or 15 wt% of IPDI. Moreover, for CW630PU filler modified with 15 wt%, color changes could be categorized as distinct and significant.

We believe that the presented paper provided valuable insights into the issue of isocyanate modifications of lignocellulosic fillers and what is important, in contrast to the majority of the papers, not only focused on properties of modified fillers, but also their impact on human health and the surrounding environment.

Acknowledgements This work was supported by the National Science Centre (NCN, Poland) in the frame of SONATINA 2 project 2018/28/C/ST8/00187 – Structure and properties of lignocellulosic fillers modified in situ during reactive extrusion. The Authors would also like to thank dr. Adam Piasecki from the Institute of Materials Engineering of Poznan University of Technology for the valuable help with the investigation of the microscopic structure of modified fillers.

Compliance with ethical standards

Conflict of interest All author(s) declare that they have no conflict no interest.

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