[11C]-Methionine PET for Identification of Pediatric High-Grade Glioma

Recurrence

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ABSTRACT

Rationale: Differentiating tumor recurrence or progression from pseudoprogression during surveillance of pediatric high-grade gliomas (PHGGs) using magnetic resonance imaging (MRI), the primary imaging modality for evaluation of brain tumors, can be challenging. The aim of this study was to evaluate whether [11]C-methionine positron emission tomography ([11C]MET-PET), a molecular imaging technique that detects functionally active tumors, is useful for further evaluating MRI changes concerning for tumor recurrence during routine surveillance.

Methods: We evaluated 27 lesions in 26 patients with new or worsening MRI abnormalities, where PHGG tumor recurrence was of concern during follow-up visits with [11C]MET-PET. We performed quantitative and qualitative assessments of both [11C]MET-PET and MRI data to predict the presence of tumor recurrence. Further, to assess for an association with overall survival we plotted the time from development of the imaging changes against survival.

Results: Qualitative evaluation of [11C]MET-PET achieved 100% sensitivity, 60% specificity, and 93% accuracy to correctly predict the presence of tumors in 27 new or worsening MRI abnormalities. Qualitative MRI evaluation achieved sensitivity ranging from 86% to 95%, specificity ranging from 40% to 60%, and accuracy ranging from 85% to 89%. The interobserver agreement for [11C]MET-PET assessment was 100%, whereas the interobserver agreement was only 50% for MRI (P = <0.01). Quantitative MRI and [11C]MET-PET evaluation using receiver operating characteristics demonstrated higher specificity (80%) than qualitative evaluations (40-60%). Postcontrast enhancement volume, metabolic tumor volume, tumor-to-brain ratio and presence of tumor as determined by consensus MRI assessment were inversely associated with overall survival.

Conclusion: [11C]MET-PET has slightly higher sensitivity, and accuracy for correctly predicting presence of tumor recurrence, with excellent interobserver agreement, than does MRI. Quantitative [11C]MET-PET can also predict overall survival. These findings suggest [11C]MET-PET can be useful for further evaluation of MRI changes during surveillance of previously treated PHGG.

Key Words: MRI; [11C]MET-PET; pediatric high-grade glioma; pseudoprogression; recurrence.
INTRODUCTION

It has only recently been discovered that pediatric high-grade gliomas (PHGGs) are biologically distinct from adult high-grade gliomas (1). However, this new knowledge has not yet changed diagnoses, classifications, World Health Organization (WHO) grading, or treatment of PHGGs (2). PHGGs in children older than 3 years are treated with a combination of maximal safe surgical resection, radiation therapy with or without adjuvant chemotherapy, and subsequent continued chemotherapy, similar to the treatment regimen for adult high-grade gliomas (3-5). Despite this aggressive therapy, outcomes in young children are dismal, with a local 1-year failure-free survival rate of 60% (6), suggesting that recurrence is common. Accurate diagnosis of tumor recurrence is important because a) the median overall survival (OS) of recurrent PHGGs is 4 to 7 months (7) and b) treatment of pseudoprogression is different from that of tumor recurrence. However, the diagnosis of recurrence is not always straightforward with magnetic resonance imaging (MRI), which is the clinical standard of care test for assessing responses to treatments. Indeed, treatment-related effects including pseudoprogression frequently mimics tumor recurrence, thereby leading to misdiagnosis and incorrect management (8,9).

Pseudoprogression is characterized by temporary enlargement and increased enhancement of clinical target volumes with MRI (10) and occurs in up to 20% of patients treated with radiation therapy and adjuvant chemotherapy (11). The incidence of pseudoprogression following initial therapy of PHGGs is similar to the incidence in adults following treatment of high grade gliomas (12). Tumor recurrence is also characterized by enlargement of tumor volume with increased enhancement making the distinction challenging (13-15). Many advanced MRI techniques have been extensively studied to differentiate treatment-related effects from true tumor progression with variable benefits (16-19). Positron emission tomography with various radiotracers has been studied to distinguish true tumor progression from
pseudoprogression (17, 20-24). Of the many PET radiotracers used to evaluate tumor recurrence, in adults, study results using amino acid PET tracers (i.e., [11C]-methionine ([11C]MET), [18F] Fluoroethyl tyrosine (FET) and [18F] - Dihydroxyphenylalanine (F-DOPA)) suggests that that a reduction of amino acid uptake and/or a decrease of the metabolically active tumor volume is a sign of treatment response associated with long-term outcome (25). Response assessment in Neuro-Oncology working group and European Association for Neuro-Oncology now suggest that [18F]-FET may facilitate the diagnosis of pseudoprogression in glioblastoma patients within the first 12 weeks following completion of chemoradiotherapy (25). [11C]MET, a true amino acid PET tracer with properties very similar to 18FET PET, has recently been shown to differentiate true tumor progression from treatment-related effects than other PET tracers in adults, with a sensitivity and specificity of 91.2% and 87.5%, respectively (26). Although the utility of [11C]MET-PET for evaluating nonenhancing PHGGs has been investigated (27), it has not been systematically investigated to evaluate tumor recurrence in PHGGs.

Here, we evaluated whether [11C]MET-PET can be useful for the identification of tumor recurrence in previously treated PHGGs. Specifically, we compared the accuracy of [11C]MET-PET for predicting the presence of tumors when recurrence is suspected with that of MRI. We also compared the interobserver agreement of [11C]MET-PET and MRI to determine whether [11C]MET-PET imaging adds value to conventional MRI and whether [11C]MET-PET or MRI can predict overall survival (OS).

MATERIALS AND METHODS

Study subjects

We retrospectively included all subjects with PHGGs who were enrolled in the ongoing “Methionine PET/CT studies in patients with cancer” clinical trial (NCT00840047) at St. Jude Children’s Research Hospital (St. Jude) since 2009. This study was approved by the St. Jude Institutional Review Board and each subject, or parent or legal guardian signed informed consent to participate. The inclusion criteria for this study were as follows: (1)
previously treated WHO grade III or IV PHGGs that demonstrated worsening or new imaging abnormalities on fluid-attenuated inversion recovery (FLAIR) sequences, or on postcontrast T1-weighted sequences, or on both the sequences during routine surveillance MRIs, in comparison with the MRI findings from the baseline or from the best response; (2) [11C]MET-PET scans obtained within 3 weeks of the surveillance MRI scans; and (3) definitive diagnosis of tumor recurrence was established within 8 weeks of either the MRI surveillance scan or [11C]MET-PET scan.

**Imaging acquisition**

**[11C]MET-PET**

[11C]MET was prepared as previously described (28). [11C]MET-PET imaging occurred after at least 4 hours of fasting. Each subject received intravenous injections of 740 MBq (20 mCi) of [11C]MET per 1.7 m² of body surface area (maximum prescribed dose, 740 MBq). Transmission computed tomography (CT) images for attenuation correction and lesion localization and PET images were acquired approximately 5–15 (8.7 ± 3.3) minutes after [11C]MET injection with a GE Healthcare Discovery 690 PET/CT scanner or GE Discovery LS PET/CT scanner (GE Healthcare) using these parameters: field-of-view = 30 cm; matrix = 192 x 192; recon method = VUE point HD; quantification method = SharpIR; Filter cutoff = 5.0 mm; subsets = 34; iterations = 4; z-axis filter = standard. Q.Clear-350-SharpIR quantification method was used only in one subject. The CT acquisition parameters are as follows: 0.5 cm slice thickness, 0.8 s tube rotation, 1.5 cm/rotation table speed, 1.5:1 pitch, 120 kV, and 90 mA with dose modulation. PET images were acquired in 3D mode for 15 min. Data were reconstructed into multiplanar cross-sectional images with standard vendor-supplied software and displayed on a nuclear medicine workstation (Hermes Medical Systems, Inc.) for analysis.

**MRI**
The following sequences were acquired with a 1.5 Avanto magnet or 3T TrioTim, Skyra, or Prisma magnet (Siemens Medical Solutions) with 0.1 mmol/Kg intravenous gadobutrol (Gadavist, Bayer Health Care): 3D magnetization-prepared rapid gradient echo (1 mm³ isotropic acquisition, 1590 ms repetition time [TR], 2.7 ms echo time [TE], 900 ms inversion time [TI], and 15° flip angle [FA]); 2D transverse T1 fast low angle shot (4-mm slice thickness [ST], no gap, 259 ms TR, 2.46 ms TE, and 70° FA); 2D transverse diffusion-weighted sequence and postcontrast 2D transverse T1 flash low angle shot (parameters identical with those of precontrast axial 2D T1); 2D transverse T2 TSE (4mm ST, no gap, 4810 ms TR, 87 ms TE, and 180° FA); 2D transverse T2 FLAIR (4-mm ST, no gap, 10 000 ms TR, 106 ms TE, 2600 ms TI, and 130° FA); and 3D sagittal T1 (parameters identical with those of precontrast sagittal 3D T1). Apparent diffusion coefficient maps were calculated from the diffusion images with the vendor provided software (Syngo, Siemens Healthcare).

**Qualitative image analysis**

**MRI**

Each surveillance MRI was evaluated four times. The first evaluation was performed during generation of the clinical report by one of the neuroradiologists assigned to the clinical service. The second evaluation was performed by a single neuroradiologist (NDS, observer 1) with twelve years experience evaluating response assessments of pediatric brain tumors. The third evaluation was performed by a single neuroradiologist (SNH, observer 2) with eight years experience evaluating response assessments of pediatric brain tumors. Both observers were blinded to the [11C]MET-PET findings and did not have access to any clinical information or any imaging studies obtained after the index surveillance MRI. The fourth evaluation consisted of a consensus evaluation by observers 1 and 2. New or worsening MRI abnormalities were subjectively categorized as definitely tumor (score = 1), definitely not tumor (score = 2), or indeterminate (score = 3). The consensus readings were also scored with the same 1–3 scale. If a discrepancy in opinion occurred between two observers, the reading was scored as 3. The first rating from neuroradiologists on clinical duties was scored with the same scale on the basis of the clinical reports. Diffusion and apparent diffusion coefficient maps were used together for subjective evaluation only.
[11C]MET-PET

[11C]MET-PET images were independently reviewed by 2 observers, 1 with 15 years experience (BLS) and the other with 2 years’ experience (AKB) in molecular imaging for assessments of treatment response in pediatric brain tumors. The observers were provided the location of the MRI abnormality with access to the MRI images. The [11C]MET-PET images were rated qualitatively on a 4-point scale relative to frontal white matter (in all of the included subjects at least some component of the frontal lobe white matter was free of tumor) : 0: no detectable uptake, or 1: mild uptake but less than contralateral frontal lobe white matter, or 2: mild uptake similar to the contralateral frontal lobe white matter, or 3: uptake greater than frontal lobe white matter. Finally, the results of visual assessment were consolidated into just two groups. The first group was “no or same or lower uptake than the reference region (grades 0, 1, and 2) and the second group was “higher uptake than the reference region (grade 3)”.

Quantitative imaging analysis

Worsening or new imaging abnormalities on postcontrast T2 FLAIR and T1-weighted sequences were manually segmented by using Vitrea Advanced Visualization (Vital Images) software. Three patients had subtle enhancement on T1-weighted sequences, and their T1-weighted regions of interest were drawn on the delta T1 images (precontrast T1-weighted images were subtracted on a voxel-by-voxel basis from the postcontrast T1-weighted images).

Standardized uptake values (SUV) for the [11C]MET-PET images were calculated using Hermes software. After co-registration of the PET data set with FLAIR and/or postcontrast T1 weighted MRI sequences, regions of interest were manually drawn either around the areas of abnormal [11C]MET uptake or around the MRI abnormality. In addition, quantitative tumor metrics (metabolic tumor volume, MTV; tumor to brain ratio, TBR) were calculated as suggested by Laws et al (29). However, instead of using a crescentic ROI, we used a 1.0 cm³ sphere to calculate SUVmean of the contralateral normal prefrontal lobe cortex and juxtacortical white matter as suggested by Hotta et al (22) for consistency. Briefly, was calculated using. The three-dimensional metabolic
tumor volume with an SUV >1.3-times greater than normal brain cortex (obtained on the prior step) was automatically contoured using Hermes software, which automatically calculated the SUVmax, and SUVmean of the tumor. Tumor-to-brain ratios (TBR: TBRmax) were then manually calculated by dividing the tumor SUVMax with the SUVmean of the contralateral normal frontal lobe cortex. TBRmean was manually calculated by dividing the tumor SUVmean with the SUVmean of the contralateral normal frontal lobe cortex. In lesions with SUV of <1.3 times the contralateral frontal lobe, a volume of interest (VOI) was manually drawn on the FLAIR abnormal areas and agreed upon by both nuclear medicine physicians and then the VOIs were copied to the PET images. The SUVmax of the VOIs were automatically calculated by the software. The TBR was then calculated as described above.

**Final outcomes**

The final outcomes of the lesions evaluated with MRI and [11C]MET-PET were determined with the following methods: (1) response assessment in neuro-oncology criteria applied to imaging and clinical findings; (2) biopsies; or (3) follow-up imaging and clinical course. Tumor was defined as present in the evaluated lesions if the lesions were treated as progressive disease (defined by RANO), a predominant tumor was evident via biopsy, progressive worsening was evident by follow-up MRI within 8 weeks of the surveillance MRI or [11C]MET-PET scan, or the subject died of tumor progression without any other identifiable cause. Because all of the evaluated lesions were included at recurrence, OS was calculated from the date of diagnosis of recurrent tumor or pseudoprogression.

**Statistical analysis**

MRI and [11C]MET-PET readings were defined as true positives when definitely tumor scores correctly identified the final outcome and as false positives when definitely tumors scores differed from the final outcome. Ratings were defined as true negatives when definitely not tumor scores correctly identified the final outcome and as false negatives when definitely not tumor scores differed from the final outcome. Sensitivity and specificity
were calculated by standard statistical definitions. Accuracy was defined as the proportion of true positives and true negatives in all of the scans. Interobserver agreement between different MRI and [11C]MET-PET observers was calculated with Cohen κ values, which were interpreted as previously indicated (31). Log-rank tests were used to assess the association of subjective [11C]MET-PET and MRI findings with OS. By using optimal cutoff values, Kaplan–Meier curves were generated for MRI parameters (T1-enhancing volumes, FLAIR volumes), and PET parameter (SUVmax) to test whether these measurements from quantitative imaging analysis were associated with OS.

The sensitivity and specificity of metabolic tumor volume, TBR, T1 and FLAIR volumes and SUVmax using optimal cutoff values for predicting final outcomes were evaluated. We used the optimized cutoff values to categorize these imaging features, and log-rank tests were performed to test whether each of these features is associated with OS values, which calculated from the time of the MRI and [11C]MET-PET scans to the death of the subjects, or the date of the last follow-up up for alive subjects. The 95% confidence intervals for all diagnostic accuracy measures were calculated using bias-corrected bootstrap methods with resampling. All the statistical analyses were done using R Statistical Software.

RESULTS

We used May 2020 as the cutoff date for our analysis and found 27 patients who matched our inclusion criteria. We excluded 1 patient with L-2-hydroxyglutaric aciduria because differentiating tumor tissue from healthy brain was challenging because of diffuse brain signal abnormalities in the entire brain due to this condition. Of the remaining 26 patients, 27 tumors (1 patient had a left frontal lobe recurrence that was treated and evaluated similarly to the original tumor in the cerebellum) were included in the analysis. Details of patient demographics and tumors are shown in Table 1 and Supplemental Table 1. The details of the previous treatment, tumor location, and genetic alterations are included in Supplemental Table 2.
Qualitative MRI and [11C]MET-PET interpretations for predicting final outcomes

The final outcomes in 5 of the 27 lesions evaluated were no tumor present (i.e., pseudoprogression) and presence of tumor (i.e., tumor progression) in the remaining 22 lesions. The final outcomes were confirmed by follow-up MRI in 16 cases, by biopsy in 4, and by RANO criteria in 7.

The sensitivity, specificity, and accuracy of correctly predicting the presence of tumors from MRI by observer 1 were 86% (95% CI: 64% - 96%), 80% (95% CI: 0 - 100%), and 85% (95% CI: 63% - 93%), respectively, and 95% (95% CI: 73% - 100%), 40% (95% CI: 0-100%), and 85% (95% CI: 63% - 93%), respectively, for observer 2. The interobserver agreement was fair (Cohen κ = 0.49; P = 0<.001). The sensitivity, specificity and accuracy for correctly predicting the presence of tumors by consensus readings were 95% (95% CI: 71% - 100%), 60% (95% CI: 0 - 100%), and 89% (95% CI: 67% - 93%), respectively. The details are summarized in Table 2.

The sensitivity, specificity, and accuracy for correctly predicting the presence of tumors with [11C]MET-PET scans were 100% (95% CI: NA), 60% (95% CI: 0 - 100%), and 93% (95% CI: 70% - 96%), respectively, and the interobserver agreement was 100% (Cohen κ = 1). Positive [11C]MET-PET readings had higher sensitivity, specificity, and accuracy for correctly predicting presence of tumors than did individual MRI readings. [11C]MET-PET also had higher sensitivity, and accuracy for correctly predicting the presence of tumors than did the consensus MRI readings. The consensus MRI and [11C]MET-PET readings were concordant in 88.9% of cases and discordant in 11.1%. In one subject, there was significant discrepancy between the MRI abnormality and PET abnormality, in which there were considerably surgery-related MRI abnormalities as the scans were obtained 21 days after surgery. (Fig 1).

We tested the accuracy between MRI observer 1, MRI observer 2, MRI consensus reads, and [11C]MET-PET reads in pairs with McNemar tests. There were no significant differences for any pair in the comparisons. In five of the 27 lesions, a discrepancy occurred between MRI observer 1, MRI observer 2, or the consensus MRI read for correctly predicting the final outcome, but [11C]MET-PET correctly predicted the final outcomes in all of these cases. The final outcomes of three of these five lesions were presence of tumor, and the final outcomes of
two of these lesions were pseudoprogression. Only 1 case was indecisive for tumor versus nontumor treatment-related changes in the consensus MRI interpretation but was correctly predicted by the [11C]MET-PET evaluation (Fig 2).

**Quantitative imaging parameters from both [11C]MET-PET and MRI for predicting final outcomes**

The ROC curves of the SUVmax, metabolic tumor volume, TBRmax, TBRmean, T1 contrast enhancing tumor volume, and abnormal tumor volume by FLAIR were assessed for their ability to predict the final outcomes. ROC analysis yielded an optimal cutoff value of 3.3 for SUVmax to differentiate between the presence and absence of tumors 60% sensitivity (95% CI: 36% - 78%), 100% specificity (95% CI: NA), and 67% accuracy (95% CI: (44% - 81%); 0.98 cm³ for metabolic tumor volume with a sensitivity of 90% (95% CI: 69%-100%) and specificity of 80% (95% CI: 0% - 100%), and accuracy 89% (95% CI: 64% - 96%); 1.82 for TBRmax with a sensitivity of 77% (95% CI: 55% - 91%) and specificity of 100% (95% CI: NA), accuracy 81% (95% CI: 59% - 89%); 1.4 for TBRmean with a sensitivity of 72% (95% CI: 50% - 88%), specificity of 40% (95% CI: (0% - 100%), and accuracy 67% (95% CI: (44% - 78%)); 86% (95% CI: 63% - 93%), respectively, for abnormal FLAIR volumes of 13.76 cm³. The details are summarized in Table 2.

**Quantitative MRI and [11C]MET-PET interpretations associated with overall survival**

We used the optimized cutoff values to categorize imaging features, including the T1-enhancing volume, FLAIR volume, SUVmax, metabolic tumor volume, and TBR. T1 contrast enhancing tumor volume, metabolic tumor volume, TBR were significant by themselves for predicting final outcome. However, association of final outcome with the quantitative imaging parameters were not significant when tested with multivariable analysis. Log-rank tests were performed to test whether these imaging features are associated
with OS. Using the above mentioned ROC-determined cut-off values, we found that there were that overall survival is significantly associated with metabolic tumor volume (p=0.0074), TBRmax (p=0.027), and T1 volume (p=0.016) (Fig. 3, 4). However, the SUVmax, TBRmean, and FLAIR volume did not show a significant association with OS.

DISCUSSION

Differentiating true tumor progression from treatment-related effects can be challenging because of overlapping features (11,19,25). Many advanced MRI techniques and molecular imaging techniques have been studied to address this challenge (19,25). Recent evidence suggests that amino acid PET tracers (i.e., [18]F- dihydroxyphenylalanine (F-DOPA)-PET and [18]F-Fluoroethyl Tyrosine (FET)-PET) can assist conventional MRI at correctly identifying surgical margin and distinguishing between tumoral and nontumoral changes (15,33-36). [11C]MET-PET, in particular, has shown substantial promise (37-40), but these studies were only performed in adults, and many included metastatic non-primary CNS tumors. Therefore, we explored the role of [11C]MET-PET in evaluating only recurrent PHGG.

The [11C]MET uptake is directly related to L-type amino acid transporter 1 (LAT1) expression (41); high [11C]MET uptake characteristically occurs in tumors with a high degree of neoangiogenesis and cellular proliferation (8,41). Previous studies have demonstrated high sensitivity and specificity of [11C]MET-PET for diagnosing high-grade tumors (8,42). In our study we found that the sensitivity and accuracy of [11C] MET-PET for correctly differentiating true tumor progression from treatment-related effects are 100% and 93% respectively, compared to the reported 70-80% sensitivity and 75% accuracy in previous studies (37,38,40). This difference may be due to the heterogeneous samples in the previous studies, which included both metastases and gliomas that were treated with different radiation doses and
chemotherapy regimens. However, the sensitivity and specificity of the [11C]MET-PET for differentiating tumor progression from treatment related effects in our study is similar to the results of the study done by Dunkle et al (43). Our study also demonstrated higher specificity of the quantitative PET evaluation in comparison to qualitative evaluation. This is in contrast to the study done by Minamimoto et al. (37), which reported no significant difference between qualitative and quantitative [11C]MET-PET evaluations for assessment of tumor progression. More recently, study done by Marner et al has also demonstrated high specificity and accuracy of [18F]FET PET for differentiating tumor from non-tumor lesions (44).

Qualitative interpretation of MRI findings is the standard of care for follow-up of high-grade gliomas after treatment (19). Unlike qualitative [11C]MET-PET assessments, qualitative interpretation of MRI findings involves careful evaluation of many different MRI sequences that exploit the different magnetic properties of tissues and changes in these magnetic properties with MRI contrast compounds. This multifactorial evaluation process inherently leads to interpretation bias, as we observed in our study. The sensitivity, specificity, and accuracy of the 2 MRI observers in our study significantly differed, although both observers had expertise in evaluating pediatric brain tumors for ten years or more. Such interpretation bias influences the diagnostic performance of MRI; indeed, we found that the consensus MRI interpretation performed significantly better, similar to that of [11C]MET-PET, than did the individual MRI readings. Because consensus MRI interpretations by multiple neuroradiologists are not practical in routine clinical practice, the addition of [11C]MET-PET imaging of suspicious MRI findings adds value to the overall care of patients with PHGG.

Our study demonstrated significant association of metabolic tumor volume, and TBRmax with OS, as previously described (45,46). Additionally, post contrast T1 volume was also significantly associated with OS, similar to multiple prior studies (47,48).
Our study includes limitations. The sample size was small but relatively large, considering the rarity of this tumor. As this study was initiated in 2009, the acquisition time of our PET scan is set to be 15 minutes, instead of currently recommended 20 minutes. In addition, the criteria for performing [11C]MET-PET on the included patients were based on a high clinical suspicion for recurrence or high likelihood of tumor recurrence on MRI findings. Consequently, there was a high pretest probability of the MRI abnormalities representing tumor recurrence, thereby introducing selection bias. A larger prospective multi-institutional study with regularly scheduled [11C]MET-PET scans might alleviate such selection bias. These studies should be sufficiently powered to examine whether [11C]MET-PET SUVmax cutoff values and qualitative interpretations can quantitatively predict final outcomes. However, due to the short half-life of carbon-11 (~20 min), [11C]MET is currently available only at institutions with access to a cyclotron facility, such a study would need to be restricted to centers with [11C]MET synthesizing capability or institutions able to refer patients with suspicious findings on MRI to a center with [11C]MET synthesizing capability. To mitigate this problem, FET-PET with longer half-life is increasingly used in assessment of gliomas in many countries (49-52).

**CONCLUSION**

Our study shows that [11C]MET-PET has slightly higher sensitivity, specificity, and accuracy for correctly predicting the presence of tumor recurrence than does MRI when new or worsening imaging abnormalities are detected during surveillance imaging of previously treated PHGG. The interobserver agreement of interpretation for [11C]MET-PET findings was excellent and better than that of MRI. Our study also shows that quantitative [11C]MET-PET and MRI can also predict overall survival. These findings indicate that [11C]MET-PET imaging may add value for predicting PHGG recurrence. However, the results from this small cohort should be validated in larger prospective, preferably multi-institutional studies.
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KEY POINTS:

QUESTION: How does the diagnostic performance of 11C-methionine PET compare with that of MRI for predicting tumors in lesions suspicious for recurrence during follow-up of pediatric high-grade gliomas?

ANSWER: [11C]MET-PET has 100% sensitivity, 60% specificity, and 93% accuracy for correctly predicting the presence of tumors in new or worsening MRI abnormalities suspicious for tumors, in contrast with 95%, 60%, and 89%, respectively, for qualitative MRI interpretation. The interobserver agreement for [11C]MET-PET was higher than that for MRI.

IMPLICATIONS FOR PATIENT CARE: We found that 11C-methionine PET is a complementary modality to MRI for evaluating lesions suspicious of recurrence in previously treated pediatric high-grade glioma.
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Figure 1. A. Post contrast coronal T1 weighted image demonstrates nodular enhancement (arrow) at the superior surgical margin. Images (B and C correspond to the axial plane through this enhancement, whereas images C and D correspond to the axial plane demarcated by the *. T2 weighted image through the level of the basal ganglia shows a large cystic resection cavity in the right temporal lobe (white arrow). There is ill-defined T2 abnormality at the posterior aspect of the resection cavity (white arrow). B. Axial T2 FLAIR image obtained through the level of the nodular enhancement as seen on image A demonstrates areas of heterogeneously hyperintense tissue at the medial (white arrow) and posterior (arrowhead) surgical margin. C. Axial reconstruction of the MET-PET images through this level shows two foci of tracer uptake at the medial (arrowhead) and posterior (arrow) surgical margin. D. Axial T2 FLAIR image obtained through a plane inferior to the plane of image B and C demonstrates a relatively large areas of heterogeneously hyperintense tissue at the posterior surgical margin.
(white arrow). **E.** Axial reconstruction of the MET-PET images through this level shows no MET uptake at the posterior surgical margin (arrow). There is minimum uptake at the anteromedial surgical margin (arrowhead). Of note, this area was not included in the metabolic tumor volume due to low SUV (lower than 1.3 times the contralateral frontal lobe cortex).
Figure 2. A. Axial T2 FLAIR weighted image through the level of midbrain shows a large cystic resection cavity in the left temporal lobe (white arrow). There is ill-defined T2 abnormality at the medial aspect of the resection cavity (black arrow). No obvious abnormality is noted posterior and lateral to the resection cavity (arrowhead). B. Axial post contrast T1-weighted image through the same level better shows focal area of contrast enhancement (black arrow). This enhancing focus has been followed up since prior treatment. Subtle contrast enhancement, a new finding compared to the previous MRIs, is noted posterior and lateral to
the resection cavity (arrowhead). **C.** Axial reconstruction of the [11C]MET-PET images through the same level shows intense MET uptake posterior and lateral to the resection cavity (arrowhead) corresponding to the new subtle T1 enhancement. **D.** Postcontrast T1-MET-PET fused image also shows the MET abnormality corresponds to the new subtle enhancement at the posterior and lateral aspect of the resection cavity (arrowhead).
FIGURE 3. Kaplan–Meier curves demonstrating the overall survival probability of subjects according to MET-PET quantitative metrics. The $P$-values of the log-rank tests of the Kaplan–Meier curves for (A) metabolic tumor volume, and (B) TBRmax.
FIGURE 4. Kaplan–Meier curves demonstrating the overall survival probability of subjects according to quantitative MRI metrics. The $P$-values of the log-rank tests of the Kaplan–Meier curves for (A) Post contrast T1 volume and (B) FLAIR volume.
Graphical Abstract
**TABLE 1 Demographics of Patients Included in Study (n = 27)**

| Patient characteristics                  | No. patients |
|-----------------------------------------|--------------|
| Diagnosis                               |              |
| Glioblastoma                            | 17           |
| WHO grade III astrocytoma               | 5            |
| High-grade neuroepithelial tumor        | 2            |
| High-grade glioma                       | 2            |
| Anaplastic pleomorphic xanthoastrocytoma| 1            |
| Age (years)*                            |              |
| 0–5                                     | 4            |
| 6–10                                    | 2            |
| 11–15                                   | 8            |
| 16–20                                   | 8            |
| 20–25                                   | 4            |
| Sex                                     |              |
| Male                                    | 16           |
| Female                                  | 10           |
| Patient status                          |              |
| Deceased                                | 22           |
| Alive                                   | 4            |

*Age at the time of PET imaging.
Table 2. Diagnostic accuracy for tumor detection

|                      | Qualitative MRI reading | Qualitative PET reading | T1-enhancing volume | FLAIR volume | SUVmax | MTV | TBRmax | TBRmean |
|----------------------|-------------------------|--------------------------|---------------------|-------------|--------|-----|--------|---------|
| Sensitivity [CI]     | 0.95[0.71-1]            | 1[na]                    | 0.73[0.50-0.88]     | 0.86[0.64-0.96] | 0.60[0.36-0.78] | 0.90[0.69-1] | 0.77[0.55-0.91] | 0.72[0.50-0.88] |
| Specificity [CI]     | 0.60[0-1]               | 0.60[0-1]                | 0.80[0-1]           | 0.80[0-1]   | 1[na]  | 0.80[0-1] | 1[na]  | 0.40[0-1] |
| Accuracy [CI]        | 0.89[0.67-0.93]         | 0.93[0.7-0.96]           | 0.74[0.52-0.85]     | 0.85[0.63-0.93] | 0.67[0.44-0.81] | 0.89[0.64-0.96] | 0.81[0.59-0.89] | 0.67[0.44-0.78] |

Abbreviations: CI, Confidence interval; MRI, magnetic resonance imaging; MTV, metabolic tumor volume; PET, [11C]MET-PET; TBRmax, Tumor SUVmax to contralateral normal brain SUVmean; TBRmean, Tumor SUVmean to contralateral normal brain SUVmean
### Supplemental Table 1. Detailed list of the qualitative and quantitative analyses of MET PET and MRI.

| Lesions Evaluated | Age | Time since RT | PET Observer 1* | PET Observer 2 | SUVmax | TBRmax | TBR mean | MTV (cm³) | Clin MR Observer** | MRI Observer 1 | MRI Observer 2 | Consensus MR Grading | GTV FLAIR (cm³) | GTV T1 (cm³) | Final outcome | OS (in days) | Diagnosis by | Tumor type |
|-------------------|-----|----------------|-----------------|---------------|--------|--------|---------|----------|-------------------|---------------|---------------|---------------------|----------------|----------------|---------------|-------------|--------------|------------|
| 1                 | 24  | >4 weeks       | 1               | 1             | 2      | 1.50   | 1.35    | 1.05     | 1                 | 2             | 1             | 1                   | 33.07          | 0.9           | 1             | 97          | MRI          | GB         |
| 2                 | 25  | >4 weeks       | 1               | 1             | 3.5    | 1.60   | 1.4     | 0.98     | 1                 | 2             | 1             | 1                   | 25.33          | 0.54          | 1             | Alive       | MRI          | AA         |
| 3*                | 16  | >4 weeks       | 1               | 1             | 8.3    | 3.39   | 1.99    | 53.1     | 1                 | 1             | 1             | 1                   | 34.74          | 1.76          | 1             | 59          | MRI          | GB         |
| 4*                | 15  | na             | 2               | 2             | 1.9    | 1.46   | 0.65    | 0        | 3                 | 2             | 2             | 3                   | 4.52           | 1.74          | 2             | 179         | MRI          | GB         |
| 5                 | 9   | >4 weeks       | 1               | 1             | 2.7    | 1.88   | 1.49    | 10.53    | 1                 | 1             | 1             | 1                   | 15.72          | 2.86          | 1             | 89          | Death        | HGG        |
| 6                 | 19  | >4 weeks       | 1               | 1             | 2.4    | 1.79   | 1.43    | 3.9      | 1                 | 1             | 1             | 1                   | 86.28          | 2.45          | 1             | 287         | Biopsy       | GB         |
| 7                 | 25  | >4 weeks       | 1               | 1             | 2.6    | 1.82   | 1.5     | 1.25     | 1                 | 1             | 1             | 1                   | 1.83           | 2.4           | 1             | 286         | MRI          | GB         |
| 8                 | 18  | >4 weeks       | 1               | 1             | 6.2    | 3.13   | 1.71    | 39.48    | 1                 | 1             | 1             | 1                   | 110.3          | 2.71          | 1             | 313         | RANO         | AA         |
| 9                 | 23  | >4 weeks       | 1               | 1             | 5.9    | 4.03   | 1.97    | 80.61    | 1                 | 1             | 1             | 1                   | 226.6          | 2.67          | 1             | 881         | RANO         | OA         |
| 10                | 5   | >4 weeks       | 2               | 2             | 0.9    | 0.65   | 0.63    | 0        | 3                 | 2             | 3             | 2                   | 2.38           | 0             | 2             | Alive       | MRI          | AA         |
| 11                | 5   | >4 weeks       | 1               | 1             | 2.7    | 2.64   | 1.5     | 8.04     | 1                 | 1             | 1             | 1                   | 5.38           | 3.81          | 1             | 151         | MRI          | GB         |
| 12                | 17  | >4 weeks       | 1               | 1             | 6.7    | 3.59   | 1.67    | 76.33    | 1                 | 1             | 1             | 1                   | 79.11          | 58.33         | 1             | 58          | MRI          | GB         |
| 13                | 7   | >4 weeks       | 1               | 1             | 0.9    | 1.17   | 0.56    | 0        | 1                 | 1             | 1             | 1                   | 25.2           | 0.4           | 1             | 391         | Biopsy       | GB         |
| 14                | 16  | >4 weeks       | 1               | 1             | 4.3    | 2.75   | 1.58    | 55.92    | 3                 | 1             | 1             | 1                   | 58.12          | 28.19         | 1             | 411         | Biopsy       | GB         |
| 15                | 13  | >4 weeks       | 1               | 1             | 11.4   | 4.72   | 2.23    | 12.69    | 1                 | 1             | 1             | 1                   | 24.57          | 5.6           | 1             | 531         | RANO         | HG PIXA    |
| 16                | 12  | >4 weeks       | 1               | 1             | 5.8    | 3.06   | 1.62    | 17.96    | 1                 | 1             | 1             | 1                   | 120.4          | 99.31         | 1             | 195         | MRI          | HGG nos    |
| 17                | 14  | >4 weeks       | 2               | 2             | 2      | 0.36   | 0.83    | 0        | 2                 | 2             | 2             | 2                   | 9.44           | 1.04          | 2             | Alive       | MRI          | HGNET      |
| 18                | 2   | >4 weeks       | 1               | 1             | 2.1    | 2.21   | 1.53    | 9.5      | 1                 | 1             | 1             | 1                   | 53.14          | 7.52          | 1             | 314         | MRI          | GB         |
| 19                | 12  | >4 weeks       | 1               | 1             | 2.9    | 2.60   | 1.54    | 5.13     | 1                 | 1             | 1             | 1                   | 87.97          | 11.92         | 1             | 650         | RANO         | GB         |
| 20                | 5   | **            | 1               | 1             | 0.9    | 1.81   | 1.28    | 11.27    | 1                 | 1             | 1             | 1                   | 69.03          | 68.92         | 2             | 1146        | Biopsy       | HGNET      |
| 21                | 13  | >4 weeks       | 1               | 1             | 2.9    | 1.47   | 1.37    | 0.18     | 2                 | 2             | 2             | 2                   | 4.76           | 2.12          | 2             | Alive       | MRI          | GB         |
| 22                | 19  | >4 weeks       | 1               | 1             | 8.7    | 4.17   | 1.88    | 6.5      | 2                 | 3             | 2             | 2                   | 13.76          | 1.84          | 1             | 630         | MRI          | GB         |
| 23                | 13  | **            | 1               | 1             | 9      | 6.24   | 2.01    | 136.8    | 2                 | 1             | 1             | 1                   | 176.8          | 93.03         | 1             | 134         | MRI          | GB         |
| 24                | 12  | >4 weeks       | 1               | 1             | 3.1    | 2.26   | 1.73    | 26.27    | 3                 | 1             | 1             | 1                   | 63.88          | 10.05         | 1             | 192         | MRI          | GB         |
| 25                | 18  | >4 weeks       | 1               | 1             | 5.8    | 3.54   | 1.99    | 133.3    | 1                 | 1             | 1             | 1                   | 113            | 1.77          | 1             | UK, lost follow-up | RANO       | AA         |
| 26                | 17  | <4 weeks       | 1               | 1             | 2.7    | 1.87   | 1.28    | 31.09    | 1                 | 1             | 1             | 1                   | 48.8           | 21.28         | 1             | 431         | RANO         | GB         |
| 27                | 13  | <4 weeks       | 1               | 1             | 6.6    | 4.22   | 1.86    | 289      | 1                 | 1             | 1             | 1                   | 133.7          | 3.37          | 1             | 178         | MRI          | GB         |
* Denotes the same patient. ** Denotes imaging abnormality between surgery and initiation of radiation therapy. ^ PET Observation scale: 1= no or same or lower uptake than the reference region [contra-lateral frontal lobe white matter], 2= higher uptake than the reference region (contra-lateral frontal lobe white matter). ^^ Clinical MRI Observation scale: 1= definitely tumor, 2= definitely not tumor, 3= neither 1 nor 2. ^^^ OS is calculated from the date of the PET scan. If the first follow up magnetic resonance imaging obtained with a gap of at least 4 weeks. If progressive disease was made at the first follow-up, the date of progression of was back dated to the index MRI. If the response was anything other than progressive disease, longitudinal MRI was assessed until a final outcome was decided. !! The subject was lost to follow-up.

Acronyms: AA, anaplastic astrocytoma; FLAIR, Fluid attenuation inversion recovery; GB, glioblastoma; GTV, gross tumor volume; HGG, high grade glioma; HGGNET, high grade neuroepithelial tumor; MRI, magnetic resonance imaging; MTV, Metabolic Tumor volume; OA, grade III oligoastrocytoma; OS, overall survival; PET, positron emission tomography; PXA, pleomorphic xanthoastrocytoma; RANO, response assessment of neurooncologist; SUVmax, maximum standard uptake value; TBRmax, Tumor SUVmax-to Brain SUVmean ratio; TBRmean, Tumor SUVmean-to Brain SUVmean ratio; UK= unknown.
| Lesion       | Tissue Diagnosis | Location | Treatment at Diagnosis                                                                 | Location of new abnormality        | Molecular markers                  |
|-------------|-----------------|----------|----------------------------------------------------------------------------------------|------------------------------------|------------------------------------|
| 1           | Glioblastoma    | R temporal lobe | Surgery + RT with TMZ + Follow-up TMZ                                                  | R temporal lobe                    | P53 mutation                       |
| 2           | AA              | R temporal lobe | Surgery + RT with TMZ + Follow-up TMZ and erlotinib                                     | R temporal lobe                    | 19q13 deletion, IDH1 mutation      |
| 3           | Glioblastoma    | Cerebellum | Surgery + RT + Follow-up pembrolizumab                                                  | Cerebellum                         | Alterations in PMS2, PIK3CA, FGFR1, NF1, EZH2 |
| 4           | Glioblastoma    | L frontal lobe | No treatment offered                                                                   | L frontal lobe                     | FGFR1 mutation, PIK3CA mutation    |
| 5           | HGNET           | Cerebellum | Surgery + RT + Follow-up COPE                                                           | Cerebellum                         | na                                 |
| 6           | Glioblastoma    | R frontal lobe | Surgery + RT + Follow-up pembrolizumab + 1 recurrence treated with                     | R frontal lobe                    | P53 mutation                       |
|              |                 |           | Surgery + RT with TMZ + Follow-up TMZ + LOM                                              |                                    |                                    |
| 7           | Glioblastoma    | L occipital lobe | Surgery + RT with TMZ + Follow-up TMZ + LOM, Multiple recurrences                      | corpus callosum                    | P53 mutation                       |
| 8           | AA              | Cerebellum | Surgery + RT + Follow-up TMZ + Lomustine                                                 | Cerebellum                         | P53 mutation                       |
| 9           | AA              | Frontal lobes | Surgery + RT + Follow-up erlotinib; Multiple recurrences                               | L & R Frontal lobes                | na                                 |
| 10          | AA              | L parietal lobe | Surgery + SJY07 chemotherapy                                                            | R frontal lobe                     | na                                 |
| 11          | Glioblastoma    | Cerebellum | Surgery + RT with TMZ + Follow-up TMZ and Bev                                            | Cerebellum                         | P53 mutation                       |
| 12          | Glioblastoma    | R parietooccipital lobes | Surgery + RT with Vornostat+ Follow-up TMZ + Bev                                     | Both Frontal                       | CDKN2A, p53                        |
| 13          | AA              | Thalamus | Biopsy + RT with TMZ + Follow-up with Etoposide                                          | Thalamus                           | P53 mutation                       |
| 14          | Glioblastoma    | R parietal region | Surgery + RT with TMZ + Follow-up TMZ                                                   | R parietal region                  | P53 mutation, ATRX                 |
| 15          | AA              | L occipital lobe | Surgery + RT with TMZ + Multiple relapse and multiple therapy                          | L occipital lobe                   | BRAFV600E                          |
| 16          | HG/G A NOS      | R thalamus | Biopsy + Carbo + VCR + Multiple recurrences                                             | R thalamus                         | H3.3 K27M                          |
| 17          | HGNET           | R parietal lobe | Surgery + RT + Follow-up COPE                                                           | R parietal lobe                    | H3.3 K27M                          |
| 18          | Glioblastoma    | L frontal lobe | Surgery + SJY07 chemotherapy                                                             | L frontotemporal lobe              | BCOR alteration                    |
| 19          | Glioblastoma    | R frontal lobe | Surgery + RT with TMZ + Follow-up TMZ + CCNU + multiple recurrences                    | R frontal lobe                     | P53 mutation                       |
| 20          | HGNET           | R frontal, temporal, and parietal lobes | Surgery + RT with TMZ + Follow-up TMZ + CCNU, multiple recurrences                        | R frontal, temporal, and parietal lobes | BCOR mutation                       |
| 21          | Glioblastoma    | Cerebellum | Surgery + RT with TMZ + Follow-up TMZ + CCNU                                            | Cerebellum                         | ATRX                               |
| 22          | Glioblastoma    | L temporal lobe | Surgery + RT + Follow up with ribociclib and trametinib                                | L temporal lobe                    | P53 mutation, ATRX, 1q gain, CDKN2A |
| 23          | Glioblastoma    | R frontal lobe | Surgery                                                                               | R frontal lobe                     | BRAFV600E                          |
| 24          | Glioblastoma    | Pons       | Biopsy + RT                                                                            | Pons                               | H3.3 K27M                          |
| 25          | HG/G A NOS      | L frontal lobe | Surgery + RT with TMZ + Follow-up TMZ                                                   | L frontal lobe                     | H3F3A p.G34R, mutations in TP53 and ATRX |
| 26          | Glioblastoma    | L temporal lobe | Surgery + RT with TMZ + Follow-up TMZ                                                  | L temporal lobe                    | P53 mutation                       |
| 27          | Glioblastoma    | L parietal lobe | Surgery + RT with veliparib                                                             | L parietal lobe                    | H3F3A p.G34R                       |

Acronyms: AA, anaplastic astrocytoma; Bev, bevacizumab; carbo, carboplatin; CCNU, Lomustine; COPE, cyclophosphamide, vincristine, cisplatin and etoposide; GTV, gross tumor volume; HGG, High grade glioma; HGNET, high grade neuroepithelial tumor; L, left; LOM, lomustine; na, not known; NOS: Not otherwise specified; OA, grade III oligoastrocytoma; r, right; RT, radiation therapy; SJY07 chemotherapy, cyclophosphamide in combination with topotecan; TMZ, temozolomide; VCR, vincristine.