Quality Matters as Much as Quantity of Skeletal Muscle: Clinical Implications of Myosteatosis in Cardiometabolic Health

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Although age-related changes in skeletal muscles are closely associated with decreases in muscle strength and functional decline, their associations with cardiometabolic diseases in the literature are inconsistent. Such inconsistency could be explained by the fact that muscle quality—which is closely associated with fatty infiltration of the muscle (i.e., myosteatosis)—is as important as muscle quantity in cardiometabolic health. However, muscle quality has been less explored compared with muscle mass. Moreover, the standard definition of myosteatosis and its assessment methods have not been established yet. Recently, some techniques using single axial computed tomography (CT) images have been introduced and utilized in many studies, as the mass and quality of abdominal muscles could be measured opportunistically on abdominal CT scans obtained during routine clinical care. Yet, the mechanisms by which myosteatosis affect metabolic and cardiovascular health remain largely unknown. In this review, we explore the recent advances in the assessment of myosteatosis and its changes associated with aging. We also review the recent literature on the clinical implication of myosteatosis by focusing on metabolic and cardiovascular diseases. Finally, we discuss the challenges and unanswered questions that need addressing to set myosteatosis as a therapeutic target for the prevention or treatment of cardiometabolic diseases.

Keywords: Muscle, skeletal; Myosteatosis; Cardiometabolic health

INTRODUCTION

Skeletal muscle is the largest organ in our body, comprising 30% to 40% of the total body mass, and thus plays an important role in health and aging. Although the loss of muscle mass is closely associated with the decline in muscle strength in aging,
the decline of muscle strength is known to be much more rapid than the concomitant loss of muscle mass [1]. In addition, sarcopenia is now considered a muscle disease (muscle failure), with low muscle strength overtaking the role of low muscle mass as a principal determinant [2]. Meanwhile, there has been much controversy about the associations between skeletal muscle mass and clinical outcomes such as metabolic health, subclinical or clinical cardiovascular disease, and mortality; such dissonance could be explained by the deterioration of skeletal muscle quality associated with myosteatosis, which may induce proinflammatory changes in adipose tissues infiltrating the muscles [3]. Therefore, myosteatosis has emerged as an important concept in the field of sarcopenia in both clinical practice and research. However, there is a lack of international agreements on its standard definition, method for assessment, and clinical outcomes including cardiometabolic diseases.

In this review, we will discuss the recent development in the assessment methods and clinical implications of myosteatosis. The various assessment methods for myosteatosis will be compared and a novel index of good quality muscle for representing myosteatosis will be introduced. This review also includes age-related changes of muscle components in men and women to gain a better understanding of the aging of skeletal muscles. Then, we will review the clinical studies about the association between muscle quality or myosteatosis and various clinical outcomes including metabolic health, cardiovascular disease, muscular and mobility dysfunction, and mortality. At the end of the review, we will briefly explore the areas that should be investigated in future research including potential mechanisms and intervention methods for myosteatosis.

**COMPONENTS OF MYOSTEATOSIS**

The term “myosteatosis” has been traditionally used to describe three different adipose depots found in the skeletal muscle including: (1) intermuscular adipose tissue, the extracellular adipose tissue found beneath the fascia and in-between muscle groups; (2) intramuscular adipose tissue, the extracellular adipose tissue found within individual muscle fascicles; and (3) intramyocellular lipids (IMCL) [4]. These are illustrated in detail in Fig. 1 [5].

**ASSESSMENT OF MYOSTEATOSIS**

Although muscle biopsy is considered to be a standard method for the assessment of myosteatosis, its invasiveness hinders a widespread application in clinical practice [6]. Therefore,
Measuring techniques such as computed tomography (CT) or magnetic resonance imaging offer several advantages including application in a larger sample and possibility for repeated measurements. CT measurements will be mainly discussed in this review because it is the most commonly used method in current studies.

**Measurement of the mean attenuation density (Hounsfield unit) and intermuscular adipose tissue by CT scan**

Many researchers have sought to develop a proper muscle quality index for the assessment of myosteatosis with CT measurements [7-9], most of which have used the average muscle density (Hounsfield unit [HU]) [6,10-15] and/or inter and intramuscular adipose tissue area (IMAT) [16-20] measured by CT scan. The mean attenuation of muscles measured by non-enhanced CT reflects the lipid content determined by biochemical and histological analysis, such that a lower mean muscle attenuation reflects a higher lipid content [6]. However, many clinical studies used enhanced CT scans or did not disclose CT contrast enhancement, even though muscle attenuation on CT images is influenced by contrast enhancement [7]. Therefore, the average muscle density has some limitations for the comparison of data among different institutions using various different CT protocols.

IMAT as a muscle quality index has been measured in the mid-thigh [17,18] or abdominal muscles [19,20] in previous studies [16]. This includes the visible storage of lipids in adipocytes located between the muscle fibers (also referred to as intramuscular fat) and between muscle groups (intermuscular fat) [16,21]. IMAT is known to be positively associated with insulin resistance and an increased risk of developing type 2 diabetes [12,13,22], loss of strength and mobility with aging [14,23], and poor oncologic outcomes [9]. However, the proportion of IMAT is only approximately 8% of total muscle area and is thus too small to effectively assess the entire trunk or the thigh muscle [24]. In another study, the proportion of IMAT in men and women was 3.3% and 4.8% of total trunk muscle area, respectively [22]. Thus, IMAT could have led to an underestimation of the differences in myosteatosis between individuals.

**Different qualities of skeletal muscle areas by CT scan on L3 vertebral level**

Several studies showed that a single scan at the level of L3 is the best compromise method for assessing the total tissue volumes of skeletal muscle [25] and has the benefit of being less susceptible to measurement errors that will be averaged out [26]. In addition, the revised European Working Group on Sarcopenia in Older People (EWGSOP) also stated in 2018 that the total abdominal muscle measurement at the level of L3 vertebral body might be the preferred option [2], as it could be measured opportunistically without additional cost or radiation exposure by using the clinical abdominal CT scans obtained during routine care. Maltais et al. [27] reported that the trunk muscle measurements showed good associations with body composition and insulin resistance, similar to the association shown in the mid-thigh muscles; the author also suggested that trunk muscles could offer an alternative for the assessment of muscle quality comparable to additional mid-thigh CT scans.

The total muscle area measured by CT scan can be divided into the areas of good quality healthy muscle (normal attenuation muscle area [NAMA]) and the areas of poor quality unhealthy muscle (low attenuation muscle area [LAMA]) [8,16,24]. NAMA represents an area where little fatty infiltration occurs, and LAMA is an area where high levels of adipocytes and intramyocellular fat are found within muscle fibers and myocytes, and thus shows a decreased density on CT scans [8,16,24]. LAMA, which is mainly measured in the mid-thigh level, was shown to be related to deficits in physical functioning, altered metabolism, and poor physical and metabolic prognosis in previous studies [16,24,28]. LAMA and/or NAMA in trunk muscles were associated with glucose tolerance [27] and metabolic syndrome [29].

The total abdominal muscle area (TAMA) in axial CT images can be segmented into three areas according to the CT density [16] and colored differently for clear visual representation (Fig. 2) [22]. Fig. 3 shows the histograms of each variable. While NAMA shows a normal distribution, IMAT and LAMA, which have high fat contents, show a left-side skewed distribution.

**NAMA/TAMA index calculated by CT measurements**

The NAMA/TAMA index, which is calculated by dividing NAMA by TAMA and multiplying by 100, was newly proposed by Kim et al. [22]. It is known that contrast-enhancement affects the skeletal muscle density (HU), but not skeletal muscle mass or area [30,31]. The influence of CT contrast enhancement could be negligible for NAMA/TAMA index, because it is a ratio of two measurements taken in the same condition [22]. Thus, the NAMA/TAMA index might be a stable and representative index of good-quality muscle for the assessment of myosteatosis, which is a critical feature for clinical use.

**Other methods for the assessment of myosteatosis**

1H Magnetic resonance spectroscopy (MRS) is a spectroscopic technique that identifies the proton resonances of the methylene...
(CH$_2$) and methyl (CH$_3$) groups of extramyocellular lipid (EMCL) stored in adipocytes and IMCL stored in spherical droplets in the myoplasm [32,33]. MRS may be advantageous over CT because it distinguishes between IMCL and EMCL and does not involve the hazard of radiation. The ability to distinguish the fat storage inside compared with outside the muscle may be an important advantage, because recent MRS studies have suggested that IMCL rather than EMCL influences insulin resistance [33]. One study showed that IMCL stores, rather than EMCL stores, measured by MRS better reflect muscle attenuation as measured by CT especially in the soleus but not in the tibialis anterior muscle [34]. However, while CT scans can be obtained opportunistically in clinical practice, MRS is only suitable for researchers and not for clinical use due to its high cost.

Fig. 2. Muscle quality map derived from abdominal computed tomography (CT) using an automated artificial intelligence software. The total abdominal muscle area (TAMA) includes all muscles on the selected axial images (i.e., psoas, paraspinal, transversus abdominis, rectus abdominis, quadratus lumborum, and internal and external obliques). The TAMA is segmented into three areas according to the CT density [16]: (1) inter and intramuscular adipose tissue area (IMAT; −190 to −30 Hounsfield unit [HU], yellow), (2) normal attenuation muscle area (NAMA; 30 to 150 HU, red), and (3) low attenuation muscle area (LAMA; −29 to 29 HU, sky blue). The skeletal muscle area (SMA, −29 to 150) refers to the combined area of NAMA and LAMA. Reprinted from Kim et al. [22], with permission from Elsevier.

Fig. 3. Histogram of computed tomography measurements in a Korean population of general health check-up participants (n = 12,697 men and 7,967 women, age 20 to 88 years). TAMA, total abdominal muscle area; SMA, skeletal muscle area; NAMA, normal attenuation muscle area; IMAT, inter and intramuscular adipose tissue area; LAMA, low attenuation muscle area.
Peripheral quantitative computed tomography (pQCT) had been used in several population studies [35,36], and has shown benefits of lower cost, lower radiation dose, and portability; however, pQCT has limitations in terms of the area of measurement such as lower leg or arm, and cannot clearly differentiate the individual muscle groups [4]. Qualitative ultrasound could measure the muscle thickness and echogenicity that reflect myosteatosis [37]. It has many benefits such as low cost, no radiation hazard, and portability, but has limitations such as inter-machinery and inter-operator variability and lack of a standardized approach for assessing myosteatosis [4]. Therefore, further studies are needed to set up the measurement standard for research and clinical use. The comparisons of various modalities for myosteatosis assessment are summarized in Table 1.

The D3-creatine dilution method has been introduced as a direct and accurate measurement of the total body creatine pool size and skeletal muscle mass [38]. Recent studies reported that in older men, muscle mass assessed by D3-creatine dilution is more strongly related to physical performance and adverse health outcomes than lean mass measured by dual-energy X-ray absorptiometry (DXA) [39,40]. Thus, D3-creatine-measured muscle mass may reflect net muscle mass excluding infiltrating fat.

### AGE-RELATED DISTRIBUTION AND PREVALENCE OF MYOSTEATOSIS

**Age-related distributions of different CT measurements**

In addition to the loss of muscle mass and strength, aging also leads to the redistribution of adipose tissue, through which subcutaneous adipose tissue relocates to more detrimental locations (e.g., intramuscular and intermuscular adipose tissues and fatty liver) [16]. Previous longitudinal studies [41,42] have shown that IMAT increases according to age. Especially, Delmonico et al. [42] reported that aging is associated with an increase in IMAT regardless of changes in weight; the authors also reported that in older adults, the degree of the loss of muscle strength is greater than that of muscle mass loss, which suggests a decrease in muscle quality. To date, however, age-related distribution or changes in different quality muscles such as NAMA or LAMA have rarely been reported because most studies focused on the myosteatosis itself. One study showed a negative association between the NAMAs and age [27], and a recent study [22] reported that NAMA decreased and LAMA and IMAT increased according to increasing age in both sexes despite showing different peak age groups in terms of total skeletal muscle area in men and women (Fig. 4).

**Diagnostic cutoff and prevalence of myosteatosis using CT indices**

Unlike the cutoff points for sarcopenia, the cutoff points for myosteatosis were mainly studied in oncologic fields [9], most of which used the cutoff points of muscle attenuation <41 HU with BMI <25 kg/m² or muscle attenuation <33 HU with BMI ≥25 kg/m² [7,9]. However, importantly, there is a need for the standardization of CT-derived diagnostic thresholds for muscle mass and myosteatosis for use in various types of patients in addition to cancer patients [7]. Recently, by using data from a large Korean population of healthy individuals, Kim et al. [22] reported the reference values of NAMA, LAMA, IMAT, and

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**Table 1. Modalities for Assessing Myosteatosis**

|                      | Computed tomography (CT) | Magnetic resonance spectroscopy | Peripheral quantitative CT | Qualitative ultrasound |
|----------------------|--------------------------|--------------------------------|---------------------------|------------------------|
| Muscle measurement   | Density (HU), NAMA, LAMA, IMAT | IMCL, EMCL                      | Density (HU), IMAT         | Muscle thickness, echogenicity |
| Accessibility        | Low                      | Very limited                    | Portable                   | Portable               |
| Ionizing radiation   | ++                       | –                               | +                          | –                      |
| Reproducibility      | High                     | High                            | High                       | Low                    |
| Cost                 | High                     | Very high                       | Low                        | Low                    |
| Additional advantages| Can be obtained opportunistically in clinical practice | Can measure intramyocellular lipid | Usefull in epidemiologic studies |
| Additional disadvantages| Can only be used for research | Axial sites cannot be measured | Measurements are not standardized |

HU, Hounsfield unit; NAMA, normal attenuation muscle area; LAMA, low attenuation muscle area; IMAT, inter and intramuscular adipose tissue area; IMCL, intramyocellular lipid; EMCL, extramyocellular lipid.
their indices, and skeletal muscle area and TAMA attenuation (HU) measured by CT scans and suggested the cutoff points for diagnosing myosteatosis based on T-scores. The suggested cutoff points of the NAMA/TAMA index were 73% for class I myosteatosis (−2.0 < T-score < −1.0) and 66% for class II myosteatosis (T-score < −2.0) in both men and women. Further studies in other populations are necessary to establish standardized diagnostic criteria for myosteatosis.

The overall prevalences of myosteatosis defined by the NAMA/TAMA index in men and women were 16.6% and 21.6% for class I and 8.8% and 20.5% for class II, respectively; notably, the prevalence of class II myosteatosis was increased with age, reaching up to 17.4% in men and 45% in women over 60 years of age [22]. Particularly, women showed a markedly decreased value of the NAMA/TAMA index (Fig. 5) and an increased prevalence of myosteatosis after 50 years of age (Fig. 6) [22]. These findings suggest that myosteatosis progresses rapidly in postmenopausal women.

ASSOCIATION OF MYOSTEATOSIS WITH CLINICAL OUTCOMES

Myosteatosis could explain the controversial effects of skeletal muscle mass on metabolic health and type 2 diabetes

While many studies showed that high lean body mass or skeletal muscle mass was associated with high levels of insulin sensitivity and/or metabolic health [43,44], many other studies also convincingly reported that high lean body mass was associated with a metabolically unhealthy phenotype [45-49]. Recently, Kim et al. [50] published a cross-sectional study in 23,311 individuals aged 20 years or older who underwent abdominal CT scans during routine health check-ups. In that study, the metabolically unhealthy phenotype was defined as having two or more components of metabolic syndrome or the presence of hypertension or diabetes. They reported that the total muscle mass was not significantly different according to the metabolic phenotype or even lower in metabolically healthy phenotypes than in metabolically unhealthy phenotypes. However, compared with those with metabolically unhealthy phenotype, the individuals with the metabolically healthy phenotype had a significantly higher proportion of good quality muscle as represented by the NAMA/TAMA index.

In addition, studies on the association between the prevalence or incidence of type 2 diabetes and the quantity of skeletal muscles measured by CT scan at the lumbar vertebrae also showed inconsistent results [43,51,52]. A recent study on myosteatosis of skeletal muscle showed that patients with type 2 diabetes had higher total and low attenuation muscle (fatty muscle) areas but lower values of NAMA (muscle with little fat) and NAMA/TAMA index [53].

These results suggested that in addition to muscle mass, the
The degree of myosteatosis should be considered in the comparison of skeletal muscle between metabolically healthy and unhealthy phenotypes as well as the comparison between patients with type 2 diabetes and those without.

**Myosteatosis could explain the controversial effects of skeletal muscle mass on cardiovascular disease**

Although several studies showed that decreased muscle mass is a risk factor for coronary atherosclerosis [54,55], the muscle mass was usually adjusted with BMI or body weight and limited by the use of DXA or bio-impedence analysis in most studies. However, several studies using CT scans have shown inconsistent results according to different CT measurements such as muscle density, IMAT, or skeletal muscle area only. A 7.2-year follow-up longitudinal study showed that greater skeletal muscle fat infiltration measured by the calf muscle density using CT scan was associated with higher all-cause and cardiovascular mortality in older men [35]. The Coronary Artery Risk Development in Young Adults (CARDIA) study showed that higher intermuscular adipose volume (IMAT, –190 to –30 HU) measured by abdominal CT was significantly associated with coronary artery calcification [20]. However, in another study that analyzed participants from the Multi-Ethnic Study of Atherosclerosis (MESA), greater abdominal muscle area (0 to 100 HU) measured by CT was significantly associated with a more harmful coronary artery calcification profile, which was contrary to their hypothesis [56].

Recently, a cross-sectional study on 4,068 individuals without cardiovascular disease reported that those with a higher NAMA/TAMA index had a higher prevalence of myosteatosis, particularly in older men.
TAMA index showed favorable metabolic characteristics including lower blood pressure, glucose, homeostatic model assessment of insulin resistance (HOMA-IR), and visceral fat area as well as a lower prevalence of significant coronary artery calcification score adjusted for these confounding factors [57]. Therefore, the study suggested that good skeletal muscle quality itself is an important protective factor for subclinical coronary atherosclerosis [57]. Fig. 7 shows the clinical profiles of representative cases with different degrees of myosteatosis. Further longitudinal studies on cardiovascular morbidities and mortality are warranted to establish the roles of skeletal muscle quality and skeletal muscle mass.

Myosteatosis and nonalcoholic fatty liver disease

In obesity, surplus lipids accumulate in non-adipose tissues, such as the liver and skeletal muscle [58]. Furthermore, the gradual disruption of cellular and molecular mechanisms of the adipose-muscle-liver axis results in tissue injuries in muscles (myosteatosis or sarcopenia) and liver (nonalcoholic fatty liver disease [NAFLD]) [5]. Therefore, the association between NAFLD and skeletal muscle dysfunction is being increasingly recognized. While many studies have focused on the relationship between NAFLD and sarcopenia or sarcopenic obesity, only few studies have analyzed the relationship between myosteatosis and NAFLD. Recently, the UK-Biobank study reported that the amount of muscle fat was significantly elevated in those with NAFLD compared with those without [59]. In addition, when compared with those with low muscle volume alone, NAFLD patients with adverse muscle composition (defined as the presence of low muscle volume and high muscle fat infiltration) had a higher prevalence of type 2 diabetes, coronary heart disease, and poor physical function [59]. In a preliminary cross-sectional analysis including 13,452 individuals from author’s study group, we found that a higher proportion of good quality muscle was strongly associated with lower risks of NAFLD as assessed by ultrasonography and liver fibrosis using the NAFLD fibrosis score and fibrosis-4 index scores. It was also reported that muscle fat content, but not muscle mass, is strongly and independently associated with nonalcoholic steatohepatitis in patients with morbid obesity [60]. Taken together, these reports suggest that myosteatosis and nonalcoholic fatty liver disease are closely interrelated.
Suggested Areas of Future Research

Apart from the issues discussed above, there are still gaps in our knowledge about myosteatosis. A profound understanding of the pathophysiological mechanisms of myosteatosis and targeted intervention studies are needed to develop effective therapeutic strategies for the prevention and treatment of myosteatosis to improve clinical outcomes.

Mechanisms of myosteatosis and its relation to insulin resistance

The mechanisms of myosteatosis are yet to be fully elucidated. A common explanation is that during weight gain and with aging, adipocytes may meet their capacity for fat storage, which increases the ectopic storage of fat around and within non-adipose tissues and organs such as skeletal muscle, liver, and pancreas [65].

For intramyocellular fat accumulation, decreased fat oxidation due to mitochondrial dysfunction such as low basal adenosine triphosphate leads to impaired fatty acid metabolism, which subsequently results in increased intramyocellular fat content [66,67]. The mechanisms linking the accumulation of fat within skeletal muscle with insulin resistance and type 2 diabetes are still unclear, but some have proposed that the accumulation of IMCL may impair the insulin receptor substrate 1/phosphatidylinositol-3-kinase pathway and the growth-factor-regulated protein kinase B pathway of insulin signaling [68]. Others have suggested that increased accumulation of lipid intermediates (e.g., diacylglycerol, long-chain fatty acyl-coenzyme A species, ceramides, and oxidized lipid mediators) due to increased accumulation of fat in the myocytes may be responsible for the suppression of insulin signaling [69].

For IMAT, there are many unanswered questions regarding the etiology, including its cellular origins and how it accumulates with age. However, several insightful studies [70,71] reported that fibroblast/adipocyte progenitors (FAPs) may be precursor cells considering their presence in muscle tissues and their ability to differentiate into adipocytes in response to sedentary lifestyle, low physical activity, or pathological conditions (e.g., Duchenne muscular dystrophy). During homeostasis, muscle lineage cells interact with FAPs and maintain them in an undifferentiated state. During tissue regeneration, FAPs remain in an undifferentiated state but proliferate and produce signals, possibly those including interleukin 6, that stimulate the differentiation of satellite cells, restoring muscle mass. During degeneration, the inhibition of FAP differentiation is blocked and...
FAPs differentiate into adipocytes and fibroblasts, which may inhibit the activation of muscle progenitors and block the restoration of muscle mass [70]. Recently, the fibroblast growth factor-2-dependent pathway was reported to be activated in aged skeletal muscle and to promote the differentiation of FAPs and formation of IMAT [72].

Although IMAT is closely related to insulin resistance, the exact mechanisms explaining this relationship are unclear as well. However, one possible mechanism linking intermuscular fat with type 2 diabetes is the impaired secretion of adipokines [73], as one study showed that increased accumulation of intermuscular fat may modulate the nutritive blood flow to the muscle and thus contribute to insulin resistance by impairing insulin action and insulin diffusion capacity [74].

Several studies suggested that skeletal muscle metabolism is regulated by the intrinsic circadian clocks [75] and that circadian misalignment results in disturbed energy metabolism and insulin resistance in skeletal muscles [76]. Further research is needed on the role of skeletal muscle circadian rhythms in the pathogenesis of myosteatosis.

**Intervention studies for myosteatosis**

Many previous intervention studies have focused on increasing muscle mass, but fewer studies have addressed issues on improving muscle quality. Recently, a meta-analysis [77] including 12 randomized controlled trials reported that a significant reduction in muscle lipid infiltration and an increase in muscle radiation attenuation were observed in the exercise group compared with the control group. However, the sample size was small in most studies (eight to 51 participants) and the duration of supervised physical exercise intervention (mean, 23 weeks; interquartile range, 12 to 36) and session duration (average, 37 minutes; interquartile range, 23 to 49) were somewhat short. Programs including aerobic [78,79] or resistance training [80] alone seemed to moderately increase the muscle radiation attenuation, and the combination of both exercise training modalities might be an optimal strategy [28,81]. One study reported that exercise showed preferential reductions in IMAT and visceral fat volumes, although both calorie restriction and exercise-induced weight loss decreased these fat depots [78]. Long-term longitudinal studies are needed not only to develop effective interventional protocols for improving myosteatosis but also to establish whether the reversal of myosteatosis can lead to clinically meaningful improvement in health outcomes.

Pharmacological interventions for delaying or mitigating myosteatosis are still largely unexplored. Pioglitazone with lifestyle weight loss intervention was shown to reduce visceral adipose tissue, but it was not effective in improving myosteatosis [82]. A few intervention studies to date have reported some benefits on body composition using myostatin inhibitors [83] or testosterone [84], but the effects on myosteatosis have not been explored yet.

There are additional issues that need to be addressed for effective intervention studies. Most of the studies so far have targeted older people and few have involved younger individuals. In order to perform well-designed controlled trials, the development and validation of standardized diagnostic methods and cutoff values of myosteatosis are essential. In addition, the relevant threshold for changes in these measures associated with clinically meaningful outcomes needs to be determined.

**CONCLUSIONS**

There is a growing body of evidence showing that myosteatosis is associated with aging, various metabolic diseases, and poor clinical outcomes. Therefore, assessment of myosteatosis in addition to muscle mass is expected to help guide treatment choices and monitor the response to the treatment of sarcopenia. However, there is no universal consensus on the standard definition and assessment methods of myosteatosis for routine clinical practice. Assessment of myosteatosis by CT scan on the lumbar vertebral level and calculation of the NAMA/TAMA index seem to be useful methodologies for evaluating muscle quality related to various clinical outcomes including metabolic and cardiovascular disease. New measurement tools that can be easily used in clinical practice are necessary to identify at-risk individuals and monitor the effectiveness of treatment options.

Importantly, targeted studies on the mechanisms of the development of myosteatosis and its association with aging or disease state along with intervention strategies for preventing or mitigating myosteatosis are warranted. Furthermore, large prospective intervention trials for improving muscle quality are also needed to provide evidence-based recommendations to promote healthy aging.

**CONFLICTS OF INTEREST**

No potential conflict of interest relevant to this article was reported.
ACKNOWLEDGMENTS

We thank our study group (prof. Sung-Jin Bae, Eun Hee Kim, and Min Jung Lee) for their co-works. We also thank Dr. Joon Seo Lim from the Scientific Publications Team at Asan Medical Center for his editorial assistance in preparing this manuscript.

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