Introduction

Habitat change through natural or anthropogenic causes is implicated as the greatest threat to global biodiversity [1–3]. As habitats alter they become more or less favourable and the way organisms use space changes [4,5]. These shifts in the use of space may influence vulnerability to predators and alter the selective regime imposed on the community [6]. If this selection is directional then habitat change can influence the evolutionary trajectories of affected species by altering the distribution of traits in the surviving population [7].

Factors that influence an individual’s probability of survival depend upon the interaction between the predator and prey, and the context within which the interaction occurs. If the context makes the prey more vulnerable, then this context may be the primary source of inequality between prey individuals. For instance, patchy habitat features may make one prey more visible than another, or alternatively interactions between prey may decrease individual vigilance or place certain individuals in closer proximity to a foraging predator. Previous studies on a wide range of taxa have found prey selection regimes to be strongly influenced by habitat [6,8,9] and social environment [10,11]. The probability of loss is also affected by attributes of the prey (e.g. body size) that lead to differential performance in an interaction with a predator possessing an appropriate selection profile [12]. Little is known of how habitat affects the social interactions among prey individuals that shape differential vulnerability, and ultimately the selective nature of mortality.

Coral reefs are one of our most diverse ecosystems, but they are also under the greatest threat through changes in habitat composition as a result of carbon dioxide induced climate change [13,14]. Global sea surface temperatures are predicted to increase by 1.1°C and 6.4°C by 2100 depending upon various future CO₂ emissions scenarios [14]. Exposure of corals to temperatures that are elevated more than a few degrees above the long term average can lead to expulsion of their symbiotic zooxanthellae (known as bleaching) and may lead to death [15]. Bleaching has already substantially reduced global coral cover [13,16] and is predicted to intensify with warming temperatures [17].

This change in live coral cover is accompanied by a change in the fish community associated with coral, but little is known of the processes and mechanisms underlying these changes in community composition. While only 9–11% of fish species are coral dependent [18,19], a large proportion of the diverse tropical fish communities (upwards of 60%) rely on live hard coral for settlement [19,20] and show declines paralleling reductions in hard coral cover [19,21]. The death of corals caused by bleaching is often patchily distributed on scales within metres [22,23]; one colony will die, while another colony of the same species next to it will appear healthy. The loss of corals and patchiness of surviving colonies suggests that settling fish will become concentrated on the remaining live corals. Enhanced density dependence on the remaining live coral may lead to an accentuation of the importance of processes such as competition [24] in mediating local mortality levels and may alter the selective nature of this loss.
Selection for fishes with dispersive larvae is a critical period, as it is for all organisms with complex life histories such as amphibians and many invertebrates [25,26]. Mortality during settlement is both extremely high and often selective for individual attributes [10,27,28]. Size advantages are often important immediately upon settlement and can influence the suite of covarying traits that carryover to later life stages [29–31]. It is unclear how coral bleaching will influence the nature of selective mortality, or how characteristics of individuals at settlement may influence the distribution of traits among habitats.

The present paper explores the behavioural mechanism underlying mortality and selective loss of individuals from a coral habitat disturbed by bleaching. The study focuses on the ambon damselfish Pomacentrus amboinensis, who preferentially settle on live coral [20,32], but are found on a variety of habitats as juveniles and adults. Understanding how P. amboinensis responds to bleaching may elucidate the mechanism of change for the large number non-coral obligate species that show declines in response to coral degradation [21]. A series of three experiments address the questions: How site attached are juvenile fish when they settle to a patchily disturbed environment? Does high density increase the likelihood of individuals moving to a bleached patch and can fish size predict an individual’s likelihood of migration? How does habitat influence behaviour and what is the role of fish size? How does habitat influence fish mortality? Findings suggest that the way peers interact and use space is noticeably altered by habitat change and this directly influences not only mortality levels but the direction and intensity of size selective loss.

Materials and Methods

Study species

The damselfish Pomacentrus amboinensis is common on coral reefs of the Indo-Pacific. Males and females are strongly site attached and often live in a discrete group. P. amboinensis has a pelagic larval duration of 15–23 days and settles at 10.3–15.1 mm standard length with its juvenile body plan largely complete [33]. Although P. amboinensis settle to a wide variety of habitats on reefs of the northern Great Barrier Reef they are found in highest densities associated with mixed live coral, rubble and sand areas on the shallow reef base or reef slope. A tagging study of 295 newly settled fish on the continuous reef edge found that fish moved little over the first 3 months after settlement (mean = 0.63 m [34]). A recent study showed that P. amboinensis had newly metamorphosed in light traps displayed a dramatic and statistically significant preference for live coral (Pocillopora damicornis) over bleached, dead coral or sand habitats in selection trials conducted in 500 l circular tanks [35]. Studies of this species also suggest high levels of mortality are typical within the first 24 h on the reef, with values of up to 98% mortality being recorded (mean ~50%) [9,10]. Interestingly, despite this high mortality, McCormick and Gagliano (in press [36]) showed that for P. amboinensis there is a relationship between the size of the otolith at hatching and who changes sex to become the dominant male later in life; that is, early life history somehow pro-rates subsequent individual success. This link stresses the importance of carry-over effects between life stages to the fundamental population dynamics of this species.

Newly settled reef fishes tend to be site attached and are subject to an array of resident and transient predators [see 9,37]. Predators can be seen striking at and occasionally capturing recently settled and juvenile reef fishes during the summer recruitment period.

A previous study that undertook behavioural observations on 10 fish over 20 consecutive 1 min periods found that a 3 min observation period was sufficient to obtain a representative quantification of behaviour for P. amboinensis (Mero and McCormick unpublished data; also see [38]).

Ethics statement

This research was undertaken with approval of the James Cook University (JCU) animal ethics committee (JCU ethics permit: A112) under the JCU animal ethics guidelines.

Experimental design

The present study was conducted at the base of a shallow reef at Lizard Island (14°38’S, 145°28’E), northern Great Barrier Reef, Australia, during October to December 2008. In overview, the study involved three separate experiments. The first experiment measured size-related mortality trajectories on three different habitats. Large and small newly metamorphosed P. amboinensis were placed in pairs onto either live healthy Pocillopora damicornis (a bushy hard coral); thermally bleached P. damicornis (see protocol below); or dead P. damicornis. The dead coral was structurally intact and with small levels of algal growth and fouling invertebrates. Patches were spaced to preclude fish migration between patches and observations were conducted for up to 140 h. The second experiment examined movement between habitat patches and the influence of fish size. Here, the first experiment was repeated, but this time the three different habitat types were placed in close proximity to one another to allow fish migration. Fish were monitored for ~48 h and their behaviour was quantified in detail. The third experiment examined whether high densities of fish on small patches of live coral promoted movement to nearby bleached or dead patches, and whether that movement was related to fish size. This experiment had the same spatial design of clusters of three habitat types in close proximity, but placed 6 tagged fish of a size range on the live coral patch. Movement and limited aspects of behaviour were recorded for 48 h.

Experiment 1: Habitat and size related persistence.

Light traps were used to collect P. amboinensis at the end of their larval phase. Fish were placed into an aquarium with aerated flowing seawater. Fish were kept for 24 h and fed newly hatched Artemia sp. twice per day ad libitum to allow recovery from (or acclimation to) the stress of capture, prior to sizing and tagging. There was minimal mortality during this time. Individual fish were placed into clip-seal plastic bags containing aerated seawater and measured with digital calipers (±0.1 mm). Fish were paired, such that one individual (‘large’ individual) was 0.8–1.0 mm greater in standard length than the other (‘small’) [see supporting information Figure S1]. To enable individual identification fish were tagged through the plastic bag with either a red or blue subcutaneous fluorescent elastomer tattoo using a 27-gauge hypodermic needle (as per 39). This left a 1.5–2 mm long stripe of colour, which was visible under the scales. Colours were alternated between large and small fish among replicates to avoid the possible bias of predators selecting prey based on tag colour. Previous studies have found no evidence of this selection (T. Holmes unpublished data). Tagging with a single elastomer tattoo has been found to have no influence on the mortality or growth of this species [39]. Behaviour observations in the field previously [e.g. 9,10,12,40,41] and in the present study have indicated no size-dependent adverse affect of tagging.

Size-paired fish were transported to the study site in 2 l plastic bags of aerated seawater, and then released onto small patches (20×10×15 cm) of one of the three substrata: healthy P. damicornis (a bushy hard coral); thermally bleached P. damicornis (see below); dead P. damicornis. Habitat patches were about 4 m from the hard reef edge on sand and organized in a row 5 m apart to prevent migration between patches. All fish and mobile invertebrates were...
removed from the substrata prior to the commencement of a trial. A small wire cage (~30×30×30 cm, 6 mm mesh size) was placed over each patch for 30–40 min to allow the tagged fish to acclimate to their new surroundings while being protected from predators. Fish were released onto the reefs between 10:00 and 10:30 h. Survival of tagged fish was monitored 2–3 times per day (morning, mid-day, evening) by visual census (occasionally the mid-day census was not undertaken). When one or both individuals were missing the adjacent area (within 3 m of the release site) was searched to determine if the fish had simply migrated. During the study period, densities of up to six newly settled *P. amboinensis* were found to naturally occur on dead and live coral habitats of the size used in this study. Monitoring finished when both tagged individuals were lost from the sites, or was terminated due to the end of a field trip.

**Experiment 2: Inter-habitat movement.** Clusters of the three different habitat types were constructed to examine the importance of movement between patches and the social dynamics that may underlie mortality of individuals within a group. Clusters were composed of one small coral head (20×10×15 cm) of each of three coral habitat types 0.4 m from one another: healthy *Pocillopora damicornis*, bleached *P. damicornis*, and dead *P. damicornis*.

In a similar methodology to above, *P. amboinensis* caught in a light trap were kept for 24 h, measured with callipers, tagged with one of six different colours and size matched for a 0.8–1 mm difference in SL (as above). Pairs were then transported out to the field in a labeled plastic bag at 10:00–11:00 h. One pair was placed on each of the three substrata per cluster. Patches were visually obscured from one another by plastic barriers during a 30 min acclimation period, during which time fish were also enclosed within a 6 mm mesh cage to prevent predation (as above). After the acclimation period the barriers and then the cages were removed. Fifteen minutes after release the behaviour of all fish within the set of three patches was quantified in detail (3 min each; see below), and this was repeated over 48 h at ~16:00 and ~11:00 h. Mortality and movement between patches was also assessed at this time.

**Experiment 3: Migration at high densities.** To determine the likelihood of a fish migrating to a bleached or dead coral when the density of fish on a live coral patch is very high, fish were stocked in high density onto live coral patches and given the choice of dead or bleached coral patches nearby. The configuration and composition of patches was exactly the same as above (experiment 2), with the exception that only the healthy *Pocillopora* coral head was stocked with 6 newly settled *P. amboinensis*. Fish were once again tagged for individual recognition and they were measured as above prior to release. Acclimation procedure was the same as experiment 2 and the location of fish were monitored over 48 hours as above. Experiment 3 was conducted 2 weeks after experiment 2 and at the same location.

**Fish behaviour**

In experiment 2 the behaviour of fish on the three patch types within each cluster was assessed over 3 min periods. Behaviour of the fish was assessed by a scuba diver positioned 1.5 m away from the patch. A magnifying glass (×4) aided the assessment of bite rates and space use over the 3 min focal animal sampling period. Six aspects of activity and behaviour were assessed: a) total distance moved; b) distance ventured from the habitat patch (categorized as % of time spent within 0, 2, 5 or 10 cm away from the patch); c) height above substratum (categorized as % of the time spent within the bottom, middle or third of the patch); d) number of fin displays; e) the number of chases or bites; f) number of avoidance episodes in response to a conspecific; g) boldness (recorded as a variable on a scale from 0 to 3 at 0.5 increments, where: 0 is hiding in hole and seldom emerging; 1 is retreating to hole when scared and taking more than 3 sec to re-emerge, weakly or tentatively striking at food; 2 is shying to shelter of patch when scared but quickly emerging, purposeful strikes at food; and 3 is not hiding when scared, exploring around the coral patch, and striking aggressively at food). At the end of the 3 min observation period, the fish was approached with a finger and the fish's reaction and latency to emerge from shelter was taken into account in the assessment of boldness. Two additional variables were devised from these variables to summarise information and reduce the number of variables that were required in analyses. Relative height on the patch was summarized as a cumulative proportion of the time spent at varying heights over the 3 min observation period, with the top of the patch taken as height of 1, mid-patch a height of 0.5, and bottom a height of 0. An aggression index was also created by adding the number of displays to the product of three times the number of chases/bites and then subtracting the number of avoidance events. A weighting factor of three was used in conjunction with the chases/bites as the influence of this behaviour on the spatial distribution of the recipients appeared to be many times greater than their response to displays.

In experiment 3, the dominance status of the individuals within each pair was also categorised from the ~10 min observation period as dominant or subordinate, based on the number of displays, chases and avoidance events.

**Coral bleaching**

Bleaching (the loss of zooxanthellae) was induced by placing *Pocillopora damicornis* colonies in 500 l seawater aquaria and raising the temperature incrementally over 48 h from ambient (28°C) to a sustained maximum of 32°C for 9 to 10 days. Aquaria were constantly aerated, flow maintained using 2 1220 Lhr⁻¹ powerheads and heated with two 300 W aquarium baton heaters under very low light levels. After colonies were visibly bleached, water temperature was lowered to ambient incrementally over 48 h. This protocol resulted in a live coral with few or no zooxanthellae that would stay bleached in the field for up to 2 months. Many ended up regaining zooxanthellae over a 6 to 8 week period.

**Analyses**

To examine the differences in behaviour between large and small fish among the three habitat treatments repeated measures ANOVAs (RMANOVAs) were conducted. In these analyses fish size (large or small) was used as the variable on which the repeated measure was undertaken (effectively pairing the observations; [42]). As there are only two levels of size the assumption of sphericity was irrelevant. Assumptions of normality and homogeneity of variance was examined using residual analysis. Tukey's HSD tests were undertaken after RMANOVA to explore the nature of any significant differences found among more than 2 means. Trends were tested for data collected 15–30 min after cage removal. To determine whether the spacing pattern between large and small fish differed among habitats after cage removal, a one-factor ANOVA was undertaken to test the equality of distances between fish. To estimate this variable the difference between the distance from the patch for the largest fish was subtracted from the smallest fish. Data were log10(x+1) transformed.

Survival (up to 165 h) between large and small fish on the three substrata was compared using Survival Analysis (Statistica 8.0). Survival curves of each fish size and substrata were calculated and plotted using the Kaplan–Meier product–limit method. The
Kaplan-Meier method is a non-parametric estimator of survival that incorporates incomplete observations, such as those cases where censuses had to be terminated on trials prior to their completion due to time limitations of a field trip. Projected survival were compared between the three substrata using a Chi-square statistic, while differences in survival between large and small fish were compared using a Cox-F statistic.

**Results**

**Movement**

When two fish that differed by 0.8–1 mm SL were placed on each of the 63 habitat patches (representing 21 clusters of 3 habitats), 15.1% moved from their original placement to one of the other habitats in the cluster (i.e., 19 out of 126 fish moved). The original habitat they were on did not influence whether they moved or not, with equal numbers moving from each habitat (healthy coral, 14.2%; bleached coral, 16.7%; dead coral, 14.2%). There was no obvious preference for the habitat to which the 19 fish moved (7 moved to live coral; 7 to bleached coral; 5 to dead). Twelve out of 19 fish that moved habitats were the smaller, subordinate individual of the pair. All the smallest individuals (6 fish) within the cluster moved habitats more than once over the census period.

When the density of fish on live coral was boosted to the upper limit of natural densities (6 per patch) 12 out of 78 fish (15.4%) moved to the dead or bleached patch within the cluster; a similar figure to the frequency of movement when two fish were placed on each patch within a cluster (15.1% above). Seven of the 12 fish moved to bleached coral, and the remaining 5 went to the dead coral patch. All fish that moved were the smallest or second smallest fish allocated to the clusters.

**Behaviour**

The behaviour of large and small fish differed among the three habitats. There was a significant interaction between fish size and habitat that influenced their distance from the patch reefs 15–30 min after removal of the cage (RMANOVA: Treatment x Size, F_{2,60} = 3.288, p < 0.004; Treatment, F_{2,60} = 10.937, p < 0.001; Size, F_{2,60} = 75.984, p < 0.0001). Large fish were closer to shelter than the smaller fish on bleached and dead coral habitats (Tukey’s HSD tests), while on healthy corals both stayed close to shelter (Figure 1A). Large fish were significantly further from shelter on bleached coral, where both large and small fish were over 2 cm from the edge of the coral (Figure 1A).

There was also a significant interaction between fish size and habitat that influenced the relative height of large and small fish on the habitat patches (RMANOVA: Treatment x Size: F_{2,57} = 3.330, p = 0.04; Treatment, F_{2,57} = 12.420, p < 0.0001; Size, F_{1,57} = 45.875, p < 0.0001). Large fish were closer to the base of the patch than the smaller fish on bleached and dead coral habitats (Tukey’s HSD tests), while on healthy corals both stayed close to the base (Figure 1B).

The distance between smallest and largest fish 15 min after cage removal also differed among habitats (log_{10} (x+1) transformed; F_{2,50} = 6.174, p = 0.004), with the distance between fish within a pair being significantly smaller on healthy coral than on the other two habitats (as determined by Tukey’s HSD tests).

Large individuals of the pair were the most aggressive individuals (mean ± SEM: large 7.18 ± 0.92, small -1.51 ± 0.9; RMANOVA, Size: F_{1,57} = 30.296, p < 0.0001), regardless of habitat (RMANOVA, Treatment: F_{2,57} = 0.146, p = 0.864; Treatment x Size: F_{2,57} = 0.660, p = 0.521). Large individuals of the pair were also the boldest individuals (large 1.7 ± 0.09, small 1.43 ± 0.09; RMANOVA, Size: F_{1,56} = 5.498, p = 0.022), regardless of habitat (RMANOVA, Treatment: F_{2,56} = 1.705, p = 0.191; Treatment x Size: F_{2,56} = 0.103, p = 0.903).

The bite rates of large and small fish when in pairs differed among habitats (RMANOVA: Size x Habitat: F_{2,30} = 17.397, p < 0.0001; Treatment, F_{2,30} = 1.842, p = 0.168; Size, F_{1,30} = 135.381, p < 0.0001; Figure 2). Large and small fish displayed the same bite rates when on healthy coral, but small fish had the highest bite rates on bleached and dead coral (Figure 2). The largest difference between bite rates of large and small fish was found on bleached corals (Figure 2).

**Mortality**

Habitat type influenced levels of mortality of *P. amboinensis* (Chi-square = 12.671, df = 2, p = 0.0018; Figure 3A). Survival was highest on healthy coral with 32% surviving 70 h after settlement (Figure 3A). Survival was equally low on bleached and dead coral (8 and 14% respectively surviving 70 h; Figure 3A).

On healthy coral, small and large fish within the pairs survived equally well (Cox’s F-Test, F_{34,30} = 1.029, p = 0.471; Figure 3B). On bleached coral large fish survived better than small fish (Cox’s F-Test, F_{38,40} = 2.273, p < 0.0001; Figure 3C). Large fish also survived longer than small fish on dead coral skeletons (Cox’s F-Test, F_{30,40} = 2.2607, p < 0.0001; Figure 3D).

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**Figure 1. Influence of coral degradation on fish vulnerability.** Spatial patterns of large and small *Pomacentrus amboinensis* (large is 0.8 to 1.0 mm larger than small) placed in pairs (one large, one small) onto patches of three hard coral habitat types (healthy, bleached and dead *Pocillopora damicornis*). (A) Mean horizontal distance from the patch reef recorded 30 min after release, and (B) mean relative height on the patch reef (a value of 1 represents 100% of time spent on top of patch; 0, 100% time at the base). Letters above the bars represent Tukey’s HSD groups. Errors are standard errors. doi:10.1371/journal.pone.0007096.g001
Unfortunately for fish communities, the high water temperatures and light intensities that promote coral bleaching peak during the summer months [43], when the majority of fishes reproduce and settle. This means bleaching will most likely occur when many fishes are at their most vulnerable: when they are small, settling naive to reef based predators, and subject to high predation pressure. Findings of the present study suggest that the change from live coral, through bleached to algal covered dead coral, directly induced changes in the behaviour of individuals at settlement, which enhanced not only the intensity of mortality, but also altered the phenotypic selection on important traits such as fish size. Even when fishes are ecologically versatile and will use settlement substrata other than live coral, such as the present species that is known to settle onto rubble [20,32], the present study indicates that levels of mortality are likely to be higher than they would have been on live coral. The loss documented here was rapid, occurring hours after settlement, and therefore would be largely undetected by monitoring surveys designed to detect broad scale changes in community composition and relative abundance.

**Discussion**

Figure 2. Influence of coral degradation and fish size on foraging rates. Comparison of bite rates (per 3 min) of large and small newly-settled *Pomacentrus amboinensis* on three different habitat patch types. Errors are standard errors. doi:10.1371/journal.pone.0007096.g002

Figure 3. Comparison of fish size-specific survival on 3 coral habitats. Survival trajectories of *Pomacentrus amboinensis* regardless of size on three habitats (A), and then of large (triangles) and small (circles) fish that were settled onto (B) healthy live coral, (C) thermally bleached coral, and (D) dead coral. doi:10.1371/journal.pone.0007096.g003
Movement of fish was minimal after settlement even when distances were small and densities were close to the maximum that naturally occurred. Some individuals moved to explore other patches, but came back to their original patch. This stresses the importance of their initial choice in influencing their fate. High site fidelity in recently settled fishes is a common finding among bottom-associated coral reef fishes [44]. Evidence suggests that if fish are on corals that subsequently bleach they are unlikely to migrate to live coral, despite exhibiting higher survival on live coral. Bleached coral may act as an ecological trap [45] for species that are attracted to it, migrate to it, or end up on it through the bleaching process; it provides a settlement stimulus which is no longer optimal and does not support viable populations [46]. Interestingly, those fish that moved most often were the smallest individuals within the experimental groups that were subject to aggression from larger individuals. Larger fish, even when on bleached coral, defended their habitat patch and were less likely to move to a different habitat patch. This means that while small individuals may initially be exposed to higher predation by nature of their subordinate status, they may still have greater overall survival if they successfully move from bleached or dead coral, where survival was lowest in the longer term. The amount of movement of this kind was moderate (~15% of small fish) so could ameliorate some of the directional selection imposed on small fishes in degraded habitats.

Behaviour immediately after settlement differed depending upon whether the coral was in a healthy live state, bleached or dead. On bleached coral fish were higher from the base of the patch and further out from the coral than when on live coral. Fish on dead coral displayed a use of space intermediate between these two extremes. Large fish were closer to the base and in closer proximity to the coral than smaller fish. There are a number of possible non-exclusive reasons for these habitat-related differences in space use. Many marine predators have visual systems that rely on contrasting colouration to detect prey [47]. Yellow fish, such as *P. amboinensis*, will be effectively camouflaged if they can distance themselves from the monochrome white bleached background to be seen against the upwelling yellow/brown light from the sand [48,49] (Figure 4). A study of the spectral qualities of fish and their habitats found that yellow fish are effectively camouflaged against an average live coral reef because they have very similar colour reflectance [49]. Similar reflectance is emitted from yellow and brown sand on shallow reefs (Marshall pers. comm.). Since most predators on recruits are mid-water (e.g. sling-jaw wrasse, coral trout) and bottom dwelling (e.g. cods and lizardfish) species, this avoidance strategy is most effective when prey are close to the sand, within the shadow of the coral. This appears to be the strategy adopted by the largest, dominant fish on bleached coral. On live coral, yellow fish will be best camouflaged when close to the mottled colour of the live coral [48,49]: the position adopted by both large and small fish on live coral. There should be strong selective pressure to minimize risk of predation when most vulnerable, such as when newly settled. If behaviour alters distribution to maximize camouflage (minimize risk) against their background then it could be expected that fish with different colouration would have differing behaviour strategies to minimize predation risk by positioning themselves in the position of lowest average contrast [49]. This may mean that habitat-related rates of mortality and the nature of selection (directional, disruptive or stabilizing) will be species specific.

Alternatively, though not exclusive of the first argument, bleached and algal/invertebrate covered dead coral may emit olfactory cues that are repugnant or confusing to settling fish. A recent study using cafeteria-style choice trials that excluded visual stimuli and promoted olfactory signals found that three damselfish species consistently avoided bleached coral [35]. Late stage larval and newly metamorphosed fishes typically have well developed olfactory systems well before metamorphosis [50] and are known to use their sense of smell to navigate [51,52] and detect conspecifics or habitat during settlement [53,54]. During bleaching the symbiotic zooxanthellae die, become necrotic and are expelled [55]; a process that is likely to leave an olfactory signal that may be detectable by fishes for some time afterwards [56]. Whatever the reason, it is interesting that the large, dominant fish also move further from the bleached patch suggesting that something associated with the loss of zooxanthellae from the coral is causing the shift in behaviour.

This paper underscores the importance of behavioural interactions around the time of settlement in influencing levels of mortality on different coral habitats. Even though differences in the sizes of newly settled fish were small (~6–8% difference), the
larger fish of the pair was always dominant, exhibiting higher boldness and aggression regardless of their absolute size. The estimated distance between fish was higher on bleached and dead coral than on live coral. This suggests that large fish are either more aggressive on degraded corals or the signals produced by the dominant or received by the subordinate were more effective. Analysis showed no significant difference in aggression with habitat [57,58] since subtle differences in habitat influence predation risk [59] and the potential importance of specific areas as key refuge sites. This study is the first to underscore the important role played by behavioural interactions between cohort members immediately after settlement in driving the dynamics of post-settlement mortality, phenotypic selection profiles and post-settlement distribution patterns.

Acquisition of the spatial position of lowest risk by the large individuals in a pair traded-off immediately against feeding rate (and possibly growth) in the two degraded habitats. Large fish on dead and bleached coral stayed closer to the base of the reef and closer to shelter, and had a lower feeding rate than small fish that were positioned higher in the water column and further from the patch. Since planktivorous fishes usually feed on items well below their gape size, fish that are further into the current have access to more and larger food items than fish down-current. Other studies have found that planktivorous fish that were positioned higher and further out from their habitats had higher feeding rates and ingest higher quality food [60,61]. In these studies, this position was secured by dominant individuals, not subordinates. Large, dominant individuals often monopolise the best or greatest amount of prey in animals with strong dominance control (e.g., lions [62], chimpanzees [63], brown bears [64]). Obviously, since mortality is highest on small fish, the strategy of being exposed with a high feeding rate is suboptimal at this vulnerable life stage.

Hard coral loss through degradation, such as bleaching, disease or corallivore predation, leads to a widespread decline in the abundance and species diversity of fishes [65]. In a recent meta-analysis, Wilson et al. (2006 [21]) found that 62% of fish species examined declined following a 10% or greater loss in coral cover. Coral dwellers and feeders were most vulnerable, but many invertebrate feeders and planktivores also showed marked declines. Rapid changes are attributed to the negative impacts of coral degradation on settlement levels in fishes [19,66]. To compound matters, the present study found that mortality after settlement was much higher on non-live coral habitat patches, and movement to alternative habitat patches was limited even for an ecologically versatile species. An unknown in this dynamic landscape of population processes is how changes in phenotypic selection will impact the range of phenotypic and behavioural traits entering the reproductive life stages. Depending upon the consistency of this selective regime [67,68], which appears to be mediated by the behaviour of the prey, rather than that the selective profile of the predator, these shifts in the nature of mortality may have impacts on the fundamental links between life history stages and the evolution of life history strategies [69].

Supporting Information

Figure S1 Overall size frequency distributions of newly metamorphosed Pomacentrus ambonensis placed in pairs onto habitat patches to examine habitat-related size selection. Individuals with each pair differed in size by 0.8–1 mm standard length. Found at: doi:10.1371/journal.pone.0007096.s001 (3.48 MB TIF)

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Author Contributions

Conceived and designed the experiments: MIM. Performed the experiments: MIM. Analyzed the data: MIM. Contributed reagents/materials/analysis tools: MIM. Wrote the paper: MIM.

References

1. Brooks TM, Mittermeier RA, Mittermeier C, da Fonseca GAB, Rylands AB, et al. (2002) Habitat loss and extinction in the hotspots of biodiversity. Biol Conser 16: 999–923.
2. McKinney ML (2006) Urbanization as a major cause of biotic homogenization. Biol Conser 127: 247–260.
3. Hughes TP, Rodrigues MJ, Bellwood DR, Cuccarelli D, Hoegh-Guldberg O, et al. (2007) Phase shifts, herbivory, and the resilience of coral reefs to climate change. Current Biol 17: 360–365.
4. Brown DR, Sherry TW (2000) Alternative strategies of space use and response to resource change in a wintering migrant songbird. Behav Ecol 19: 1314–1325.
5. Long RA, Rachlow JL, Kie JG (2008) Effects of season and scale on response of elk and mule deer to habitat manipulation. J Wildl Manag 72: 1133–1142.
6. Evans KL, Gaston KJ, Sharp SP, McGowan A, Hatchwell BJ (2009) The effect of urbanisation on avian morphology and latitudinal gradients in body size. Oikos 118: 251–259.
7. McClure MM,Carlson SM, Beechie TJ, Pess GR, Jorgensen JC, et al. (2008) Evolutionary consequences of habitat loss for Pacific anadromous salmonids. Evol Appl 1: 300–319.
8. Raner CA, Yeh P, Eggert LS, Hunt KE, Woodruff DS, et al. (2004) Genetic and morphological evolution following a founder event in the dark-eyed junco, Junco hyemalis. Molec Ecol 13: 671–681.
9. Holmes TH, McCormick MI (2006) Location influences size-selective predation on newly-settled reef fish. Mar Ecol Prog Ser 317: 203–209.
10. McCormick MI, Meekan MG (2007) Social facilitation of selective mortality. Ecology 88: 1562–1570.
11. Scwambach R, Pouillon L (2009) Population density fluctuations change the selection gradient in Eurasian perch. Am Nat 173: 507–516.
12. Holmes TH, McCormick MI (2009) Influence of prey body characteristics and performance on predator selection. Oecologia 159: 401–413.
13. Hughes TP, Baird AH, Bellwood DR, Card M, Connolly SR, et al. (2003) Climate change, human impacts, and the resilience of coral reefs. Science 301: 929–933.
14. Munday PL, Jones GP, Pratchett MS, Williams AJ (2008) Climate change and the future for coral reef fishes. Fish Fish 9: 261–285.
15. Hoegh-Guldberg O (1999) Climate change, coral bleaching and the future of the world’s coral reefs. Mar Freshw Res 50: 839–866.
16. Gardner TA, Cote IM, Gill JA, Grant A, Watkinson AR (2003) Long-term region-wide declines in Caribbean corals. Science 301: 958–960.
17. Donner SD, Skirving WJ, Little CM, Oppenheimer M, Hoegh-Guldberg O (2005) Global assessment of coral bleaching and required rates of adaptation under climate change. Global Change Biol 11: 1–13.
18. Munday PL, Jones GP, Shemara M, Williams AJ, Goby G (2007) Vulnerability of fishes of the Great Barrier Reef to climate change: a vulnerability assessment. In: Johnson JE, Marshall PA, eds. Climate change and the Great Barrier Reef: Great Barrier Reef Marine Park Authority and Australian Greenhouse Office, Australia, pp 357–391.
19. Jones GP, McCormick MI, Smirnova M, Eagle Jv (2004) Coral decline threatens fish biodiversity in marine reserves. Proc National Acad Sci USA 101: 8251–8253.
20. Feary DA, Almany GR, McCormick MI, Jones GP (2007) Habitat choice, recruitment and the response of coral reef fishes to coral degradation. Oecologia 153: 727–737.
21. Wilson SK, Graham NAJ, Pratchett MS, Jones GP, Polunin NV (2006) Multiple disturbances and the global degradation of coral reefs: are reef fishes at risk or resilient? Global Change Biol 12: 2229–2234.
22. Marshall PA, Baird AH (2000) Bleaching of corals on the Great Barrier Reef: differential susceptibilities among taxa. Coral Reefs 19: 153–163.
23. Clark R, Jeffrey C, Woody K, Hillis-Starr Z, Monaco M (2009) Spatial and temporal patterns of coral bleaching around Buck Island Reef National Monument, St. Croix, U.S. Virgin Islands. Bull Mar Sci 84: 167–182.
24. Jones GP (1987) Competitive interactions among adults and juveniles in a coral reef fish. Ecology 154: 1547–1554.
25. Pechenik JA, Wendt DE, Jarrett JN (1998) Metamorphosis is not a new beginning. Larval experience influences juvenile performance. Biosci 48: 901–910.
26. Vonesh JR (2005) Sequential predator effects across three life stages of the African tree frog, Hyla erythraea. Oecologia 143: 280–290.
27. Almany GR, Webster MS (2006) The predation gauntlet: early post-settlement mortality in coral reef fishes. Coral Reefs 25: 19–22.
28. Hamilton SL, Regetz J, Warner RR (2008) Post-settlement survival linked to larval life in a marine fish. Proc Nat Acad Sci USA 105: 1561–1566.
29. Sponaugle S, Gerrod-Cobert K (2006) Environmental variability, early life-history traits, and survival of new coral reef fish recruits. Integr Comp Biol 46: 623–633.
30. Gagliano M, McCormick MI (2007) Compensating in the wild: is flexible growth the key to early juvenile survival? Oikos 116: 111–120.
31. Walker SPW, Ryen CA, McCormick MI (2007) Rapid larval growth promotes sex change and growth acceleration in a protogynous hermaphrodite, Pseudochromis aukle Jordan & Starks 1905. J Fish Biol 71: 1347–1357.
32. Öhman MC, Munday PL, Jones GP, Caley MJ (1998) Settlement strategies and distribution patterns of coral-reef fishes. J Exp Mar Biol Ecol 225: 219–236.
33. McCormick MI, Makey L, Dubour V (2002) Comparative study of metamorphosis in tropical reef fishes. Marine Biology 141: 841–853.
34. McCormick MI, Makey LJ (1997) Post-settlement survival in coral reef fishes: overlooked complexity in niche shifts. Mar Ecol Prog Ser 153: 247–257.
35. Moore JAY (2007) The influence of coral bleaching on early life-history processes of coral reef fishes. Honours thesis, School of Marine and Tropical Biology, James Cook University.
36. McCormick MI, Gagliano M (In press) Carry-over effects - the importance of a good start. Proc 11th Int Coral Reef Symp Session 10, Ft. Lauderdale, Florida, 7–11 July 2008.
37. Bevers JS, Jones GP (1997) Habitat complexity modifies the impact of piscivores on a coral reef fish population. Oecologia 114: 59–69.
38. Meekan MG, von Kuerthy C, McCormick MI (In press) Does behaviour mediate the costs and benefits of fast growth in fishes? Anim Behav.
39. Hoey AS, McCormick MI (2006) Effects of subcutaneous fluorescent tags on the growth and survival of a newly settled coral reef fish, Pomacentrus amboinensis (Pomacentridae). Proc 10th Int Coral Reefs Symp 420–425.
40. McCormick MI, Hoey AS (2004) Larval growth history determines juvenile growth and survival in a tropical marine fish. Oikos 106: 225–242.
41. Hoey A, McCormick MI (2004) Selective predation for low body condition at the larval-juvenile transition of a coral reef fish. Oecologia 139: 23–29.
42. Litell RC, Milliken GA, Stroup WW, Wolinder RD (1996) SAS system for mixed models. SAS Institute Inc., Cary, NC.
43. Berkelmans R (2009) Bleaching and mortality thresholds: how much is too much? Ecological Studies. Berlin: Springer-Verlag. pp 103–106.
44. Sale PF (1978) Reef fishes and other vertebrates: A comparison of social structures. In: Reese ES, Lighter FJ, eds. Contrasts in behaviour, adaptations in the aquatic and terrestrial environments. New York: John Wiley and Sons. pp 313–346.
45. Koko H, Sutherland WJ (2001) Ecological traps in changing environments: Ecological and evolutionary consequences of a behaviourally mediated Allee effect. Evol Ecol Res 3: 537–551.
46. F Cairn DA, McCormick MI, Jones GP (2009) Growth of reef fishes in response to live coral cover. J Exp Mar Biol Ecol (in press).
47. Lythgoe JN (1988) Light and vision in the aquatic environment. In: Atmaja J, Fay RR, Popper A, Tavolgan WN, eds. Sensory biology of aquatic animals: Springer-Verlag. pp 75–92.
48. Marshall NJ (2006) Communication and camouflage with the same 'bright' colours in reef fishes. Phases Trans R Soc Lond B 355: 1243–1248.
49. Marshall NJ, Jennings K, McFarland WN, Loew ER, Losey GS (2003) Visual biology of Hawaiian coral reef fishes: III environmental light and an integrated approach to the ecology of reef fish vision. Copeia: 467–480.
50. Wright KJ, Higgs DM, Belanger AJ, Les JMJ (2005) Auditory and olfactory abilities of pre-settlement larvae and post-settlement juveniles of a coral reef damselfish (Pisces: Pomacentridae). Mar Biol 147: 1425–1434.
51. Gerlach G, Atmaja J, Kingsford MJ, Black KP, Miller-Sims V (2007) Smelling home: can prevent dispersal of reef fish larvae. Proc Nat Acad Sci USA 104: 850–856.
52. Dixson DL, Jones GP, Munday PL, Planes S, Pratchett MS, et al. (2008) Coral reef fish smell flees to find island homes. Proc R Soc B 275: 2831–2837.
53. Sweatman H (1988) Field evidence that settling coral reef fish larvae detect resident fishes using dissolved chemical cues. J Exp Mar Biol Ecol 124: 163–174.
54. Arvedlund M, McCormick MI, Fautin DG, Bildsøe M (1999) Host recognition and possible imprinting in the anemonefish Amphiprion melanopus (Pisces: Pomacentridae). Mar Ecol Prog Ser 198: 207–218.
55. Strychar KB, Sammarco PW (2009) Exaptation in corals to high seawater temperatures: Low concentrations of apoptotic and necrotic cells in host coral tissue under bleaching conditions. J Exp Mar Biol Ecol 369: 31–42.
56. Noorem PW (1986) Origins of the freshwater attractant(s) of migrating elvers of the American eel, Anguilla rostrata. Environ Biol Fish 17: 185–200.
57. Rincon PA, Grossman GD (2001) Intraspecific aggression in rosy dace, a drift-feeding stream cyprinid. J Fish Biol 59: 968–986.
58. Lee JSF, Berejikian RA (2009) Structural complexity in relation to the habitat preferences, territoriality, and hatchery rearing of juvenile China rockfish, Sebastes nebulosus. Environ Biol Fish 84: 411–419.
59. Valeix M, Loveridge AJ, Chanmaile-Jammes S, Davidson Z, Murindagomo F, et al. (2009) Behavioral adjustments of African herbivores to predation risk by lions: Spatiotemporal variations influence habitat use. Ecology 90: 23–30.
60. Coates D (1980) Prey-size intake in humbug damselfish, Pomacentrus amboinensis (Pisces: Pomacentridæ). Mar Biol 147: 1425–1434.
61. Webster MS (2004) Density dependence via interspecific competition in a coral-reef fish. Ecology 85: 986–994.
62. Packer C, Pasie AE, Eberly LE (2001) Egalitarianism in female African lions. Science 291: 690–693.
63. Witting RM, Boesch C (2003) Food competition and linear dominance hierarchy among female chimpanzees of the Tai National Park. Int J Primatol 24: 847–867.
64. Grade SM, Quinn TP, Hilborn R, Hendry AP, Dickerson B (2004) Brown bears selectively kill salmon with higher energy content but only in habitats that facilitate choice. Oikos 104: 518–528.
65. Pratchett MS, Wilson SK, Graham NAJ, Munday PL, Jones GP, et al. (2009) Coral Bleaching and consequences for motile reef organisms: past, present and uncertain future effects. In: van Oppen MJH, Lough JM, eds. Coral Bleaching: Patterns, Processes, Causes and Consequences: Springer. pp 139–158.
66. Booth DJ, Beretta GA (2002) Changes in a fish assemblage after a coral bleaching event. Mar Ecol Prog Ser 245: 205–212.
67. Wallace B (1975) Hard and soft selection revisited. Evolution 29: 465–473.
68. Seches CJ, Hanski I (2006) Natural selection and population dynamics. Trends Ecol Evol 21: 341–347.
69. Podolsky RD, Morony AL (2006) Integrating function across marine life cycles. Integ Comp Biol 46: 577–586.