Global Consensus Sequence Development and Analysis of Dengue NS3 Conserved Domains

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Abstract

The dengue virus (DENV) genome encodes 10 different genes including the NS3 gene, which has a protease and helicase domain used in virus replication. This domain is a potential target for antiviral agents against dengue. Due to a high mutation rate, DENV is classified into four major serotypes (DENV1–DENV4). This study was designed to perform conservancy analysis of all four serotypes by drawing a consensus sequence for each serotype and then drawing a global consensus sequence to study conserved residues in all four serotypes. A total of 127 NS3 sequences belonging to all four serotypes were retrieved and aligned using multiple alignment feature of CLC Workbench and were subjected to phylogenetic tree construction. Conservancy analysis of NS3 revealed conserved peptides with active site residues that can be important in developing antiviral agents against dengue virus. Among conserved residues, residues G142, Ser144, and G145 (catalytic pocket residues), A219, D220, and D221 (divalent cations binding residues), and His56, Asp79, Ser144, 146 were highly conserved among all the serotypes. Residues from L138 to L149 and from L226 to L245 were also considerably conserved in all serotypes, while lysine141 mutated to serine in serotype 3. A total of 14 peptides from the conserved regions of DENV NS3 protein were identified, which may be helpful to develop peptide inhibitors. The DENV NS3 phylogenetic tree showed the evolutionary relationship among all four serotypes, and all serotypes of dengue were found to have evolved from the dengue 4 serotype. Because of its high variability, DENV has become a global health concern. It is important to study residues that are present in protease, helicase, the catalytic pocket Mg$^{2+}$ binding site, and the AAA domain. This study revealed peptides with active site residues that are highly conserved among all four serotypes. These regions of the NS3 sequence may be helpful in developing antiviral agents.

Key words: consensus sequence; dengue virus; dengue virus serotypes; NS3 protease/helicase

Introduction

Dengue infection has become a major health problem in more than 100 countries in Africa, Asia, America, the Western Pacific, and the Eastern Mediterranean. In the last decade, dengue infection has caused many endemics in Pakistan and thus has become a major health issue in Pakistan. Dengue infection is caused by a member of the Flaviviridae family known as dengue virus (DENV), which has four different serotypes (DENV1–DENV4). From a prevalence analysis of dengue infections in Pakistan during the last decade, it was found that DENV2 serotype is most prevalent serotype circulating in Pakistan. Dengue infection results in two types of infections ranging from a dengue fever to a more severe infection that can cause dengue hemorrhagic fever and dengue shock syndrome. Dengue fever is an infectious disease transmitted by mosquitoes (Aedes aegypti and Aedes albopictus).

Dengue infection is most common in tropical and subtropical areas of the world especially during rainy seasons, which provide the best conditions for mosquito breeding. According to an estimate by the World Health Organization, approximately 50–100 million infections occur each year, causing 500,000 cases of dengue hemorrhagic fever and 22,000 deaths.3–5 The mode of DENV transmission makes an attractive target to study DENV propagation.6 Dengue virus is an enveloped, 11-kilobase-long RNA positive-strand virus with three structural proteins, C (capsid), M (membrane), and E (envelop), and seven nonstructural proteins, NS1, NS2A, NS2B, NS3, NS4A, NS4B, and NS5, that are important in viral pathology.2,9,10 Among the nonstructural proteins, NS3 has enzymatic reactions that are important for viral replication. NS3 has two domains, the N-terminal protease domain, which is important for the proteolysis of polyprotein, and the C-terminus helicase domain.11 The NS3 N terminal

ABBREVIATIONS: C, capsid; DENV, dengue virus; E, envelop; M, membrane; NTPase, nucleoside triphosphatase; UPGMA, unweighted pair group method with arithmetic mean.
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domain is about 180 residues long with a triad of active residues, histidine, aspartate, and serine, that form a catalytic triad that actively participates in the catalysis process of the enzyme. The C-terminal domain consists of conserved motifs that are found in several nucleoside triphosphatases (NTPases) and the DEXH family of RNA helicase12,13 and help to increase the overall length of NS3, which results in enhanced enzymatic activity. It has also been reported that NS3 has DNA unwinding activity. Despite the high mutation rate of DENV, some residues that actively participate in viral replication remain conserved. These conserved residues can be important in developing specific antiviral agents and inhibitors against DENVs. In this study, a novel approach is used to produce a consensus sequence of NS3, which will be useful in designing peptides that may help in finding a possible cure for dengue infection. The present study was designed to draw a global consensus sequence of the NS3 protein of DENV, study the DENV NS3 conserved domain function, and draw a phylogenetic tree.

Methods

Drawing a consensus sequence of DENV NS3

A total of 127 sequences of dengue NS3 protein were retrieved from the National Center for Biotechnology Information (NCBI).14 Consensus sequences of dengue NS3 proteins for each serotype were drawn using multiple sequence analysis feature of CLC Workbench, a software that is use to analyze and visualize sequence comparisons.15 Thirty sequences of serotype 1 (DENV1) reported from Cuba, Indonesia, Japan, United States, Thailand, and China were used to construct the serotype 1 consensus sequence. Thirty-three sequences of serotype 2 (DENV2) from China, United States, Taiwan, and Singapore were used to construct the serotype 2 consensus sequence. Thirty-one sequences of serotype 3 from the United States, Brazil, Singapore, and French West Indies, were used to construct the serotype 3 consensus sequence. Thirty-one sequences of serotype 4 (DENV4) retrieved from NCBI and originating in Taiwan, Indonesia, United States, China, Thailand, and Brazil were used to produce the consensus sequence of DENV4.

Peptides designing for potential peptide vaccine

The consensus sequences of all four serotypes (DENV1–DENV4) were drawn in CLC Workbench and aligned to get the global consensus sequence. The consensus sequence was used to study variations in different motifs and domains of the DENV NS3 region. Short peptides from the highly conserved regions of the DENV NS3 protein were selected from the consensus sequence analysis; these peptides would be the best targets to be tested as a potential peptide vaccine.

Phylogenetic tree analysis

To draw a phylogenetic tree of the DENV NS3 gene belonging to different serotypes, we used 125 sequences; 30 sequences from DENV1, 33 from DENV2, 31 from DENV3, and 31 from DENV4. All 125 sequences were first aligned in CLC Workbench, and the aligned file was then subjected to the unweighted pair group method with arithmetic mean (UPGMA) method to draw a phylogenetic tree with boot strap value 100. The UPGMA method is based on pairwise similarity/dissimilarity distance matrix.16

FIG. 1. Multiple sequence alignment of consensus sequences of dengue NS3 belonging to all serotypes (DENV1–DENV4). The global consensus sequence is shown at the base. Conserved residues are shown with their corresponding symbols while the highly variable amino acids are denoted by an “x” symbol. DENV, dengue virus.
Results

NS3 protein has two domains (serine protease and NTPase helicase) and is considered the second largest protein of DENV genome. NS3 has become an important drug target because both of its domains are involved in viral replication. The NS3 consensus sequence of each DENV serotype was drawn in CLC Workbench, and the sequences were aligned to study the residues that were highly conserved among all the serotypes. Figure 1 shows the alignment of the consensus sequence of all four DENV serotypes. The alignment of all the consensus sequences will help us to study the strongly conserved regions of the DENV NS3 protein. Short peptides of 9 to 18 amino acids were designed from the highly conserved regions of the DENV NS3 consensus protein sequences; the sequence and position of these peptides are shown in Table 1. These positions are highly conserved and may serve as the targets to design peptide vaccines or site-specific inhibitors. A phylogenetic tree of 127 DENV NS3 sequences belonging to serotypes DENV1–DENV4, reported from all over the world, was constructed using the UPGMA method in CLC Workbench as shown in Figure 2. A default value of 100 was used in bootstrap analysis, and the values are present at each branch. Sequences from different serotypes are clustered together. The tree shows that the different DENV serotypes have evolved from the serotype DENV4.

Discussion

Dengue infection has become a global threat to human health. Dengue virus has four serotypes, which hinders development of a successful vaccine. NS3 domains, because of their conservancy, are considered as promising targets for drug development. NS3 has three conserved domains: peptidase S7 family domain, helicase C-terminus conserved domain, and flavivirus DEAD domain. Residues 1–180 of the NS3 protease domain forms two β-barrels, and between these β-barrels there are three catalytic sites (His51, Asp75, and Ser135). All the remaining residues (180–618) of NS3 forms three subdomains (I, II, III) of the helicase domain. The helicase domain belongs to the P-loop NTPase family. The helicase domain family is found in other helicase-related proteins and is restricted to DEAD/H helicase.

The consensus sequence alignment shows that His56, Asp79, Ser144, and Ser146 are highly conserved among all serotypes. Site-directed mutagenesis has revealed the catalytic role of these triad residues (His, Asp, Ser). Thus, it can be inferred that this conserved triad is involved in a catalysis mechanism among serine proteases. The consensus sequence alignment shows that L138 to L149 are highly conserved among all serotypes, while L141 has mutated to a serine in serotype 3.

This peptide fragment encodes the peptidase S7 superfamily, which helps in processing of the NS3 poly-protein precursor into mature proteins; this region also encodes trypsin-like serine protease. Consensus sequence analysis shows that G142, Ser144, and G145 are highly conserved and form the oxyanion hole. Mutation in residues (Y159 and G160) to alanine can completely abrogate the enzyme activity. Any impairment in helicase activity halts DENV replication. The consensus sequence analysis shows that the string of amino acids from I349 to R357 encodes a part of the helicase DEAD-box domain involved in nuclear transcription, nucleic acid unwinding, pre-mRNA splicing, nucleocytoplasmic transport, ribosome biogenesis, translation, RNA decay, and organelle gene expression. In addition, it also contains a putative Mg2+ binding and ATP binding site. The tripeptide string of A219, A220, and A221 is involved in DNA transportation. The consensus sequence alignment shows that the peptide string from 465 to 473 (QRRGRXGRN) is the part of the helicase domain mutation in R467 and R469 that will reduce the activity of helicase. In this study, we have drawn a phylogenetic tree of 127 DENV NS3 sequences reported from different countries. The tree was constructed by the UPGMA method as shown in Figure 2. The tree shows that

| Position | Peptides | Domain encoding |
|----------|----------|-----------------|
| 55–66    | FHTMWHVRTGAV | Peptidase S7 |
| 136–148  | LDKFPGTSGPI  | Peptidase S7 |
| 157–165  | GLYNGNVVT    | No putative conserved domain |
| 226–245  | LRLILAPTRVVAEMEEL | P-loop NTPase superfamily |
| 266–281  | IVDMCHAFFTIMRLLS | P-loop NTPase superfamily |
| 305–314  | AARGYISTRV  | No putative conserved domain |
| 316–330  | MGEAAIFMTATPPG | AAA-domain |
| 349–357  | IPERSWNNSG  | DEAD-box helicase |
| 366–376  | GKTWVFVPISIK | DEADc domain |
| 415–427  | VVTTDISEMGANF | No putative conserved domain |
| 446–456  | DGPERVILGP  | No putative conserved domain |
| 466–474  | QRRGRXGRN   | Helicase domain |
| 496–508  | AHWTEAKMLLDNI | C-terminus domain |
| 546–556  | LMRRGDLVWL  | No putative conserved domain |
the serotype DENV4 occupies the root of the tree, and the first serotype to evolve from DENV4 was DENV2. DENV2 bifurcates into two wings; from one wing, DENV3 evolved, and from the second, DENV2. Regato et al. drew a phylogenetic tree of 79 DENV NS5 gene partial sequences reported from all over the world and reported that different DENV serotypes have evolved from DENV4. Our study suggests that there are certain stretches of amino acids that take part in binding with divalent cations, pre mRNA splicing, ATPase binding, unwinding of dsRNA by helicase activity, and viral replication that are highly conserved and can be used as a potential target for the development of antiviral agents. Phylogenetic analysis suggests that different DENV serotypes have evolved from the serotype DENV4.

Disclosure Statement
No competing financial interests exist.

References
1. Idrees S, Ashfaq UA. A brief review on dengue molecular virology, diagnosis, treatment and prevalence in Pakistan. Genet Vaccines Ther. 2012;10:6.
2. Idrees S, Ashfaq UA, Khaliq S. RNAi: antiviral therapy against dengue virus. Asian Pac J Trop Med. 2013;3:232–236.
3. Almas A, Parkash O, Akhter J. Clinical factors associated with mortality in dengue infection at a tertiary care center. Southeast Asian J Trop Med Public Health. 2010;41:333–340.
4. Centers for Disease Control and Prevention. Dengue homepage. Available at www.cdc.gov/dengue/epidemiology/index.html (accessed February 7, 2013).
5. World Health Organization. Impact of dengue. Available at www.who.int/csr/disease/dengue/impact/en (accessed February 7, 2013).
6. Shah M, Wadood A, Rahman Z, Husnain T. Interaction and inhibition of dengue envelope glycoprotein with mammalian receptor DC-sign, an in-silico approach. PLoS ONE. 2013;8:e59211.
7. Rodenhuis-Zybert IA, Wilschut J, Smit JM. Dengue virus life cycle: viral and host factors modulating infectivity. Cell Mol Life Sci. 2010;67:2773–2786.
8. Seema, Jain SK. Molecular mechanism of pathogenesis of dengue virus: entry and fusion with target cell. Indian J Clin Biochem. 2005;20:92–103.
9. Murthy HM, Clum S, Padmanabhan R. Dengue virus NS3 serine protease. Crystal structure and insights into interaction of the active site with substrates by molecular modeling and structural analysis of mutational effects. J Biol Chem. 1999;274:5573–5580.
10. Yang CC, Hsieh YC, Lee SJ, et al. Novel dengue virus-specific NS3B/NS3 protease inhibitor, BP2109, discovered by a high-throughput screening assay. Antimicrob Agents Chemother. 2011;55:229–238.
11. Luo DH, Xu T, Hunke C, et al. Crystal structure of the NS3 protease-helicase from dengue virus. J Virol. 2008;82:173.
12. Gorbatenya AE, Kooin EV, Donchenko AP, et al. Two related superfamilies of putative helicases involved in replication, recombination, repair and expression of DNA and RNA genomes. Nucl Acids Res. 1989;17:4713–4730.
13. Kadare G, Haenmi AL. Virus-encoded RNA helicases. J Virol. 1997;71:2583–2590.
14. Lipman D, Altschul S, Landsman D, et al. National Center for Biotechnology Information. Bethesda, MD, 1988.
15. CLC Genomic Workbench. Home page. Available at www.clcbio.com/products/clc-genomics-workbench/ (accessed January 13, 2013).
16. Edwards RJ, Parker J. UPGMA worked Example. [http://www.southampton.ac.uk/~re1u06/teaching/upgma/] (accessed April 1, 2013).
17. Padmanabhan N. NIH-developed candidate dengue vaccine shows promise in early-stage trial. Bethesda, MD, 2013.
18. Luo D, Xu T, Hunke C, et al. Crystal structure of the NS3 protease-helicase from dengue virus. J Virol. 2008;82:173–183.
19. Perera R, Kuhn RJ. Structural proteomics of dengue virus. Curr Opin Microbiol. 2008;11:369–377.
20. Lee LC, Lee YL, Leu RJ, et al. Functional role of catalytic triad and oxyanion hole-forming residues on enzyme activity of Escherichia coli thioesterase I/protease I/phospholipase L1. Biochem J. 2006;397:69–76.
21. Salaemae W, Junaid M, Angsuthanasombat C, et al. Structure-guided mutagenesis of active site residues in the dengue virus two-component protease NS2B-NS3. J Biomed Sci. 2010;17:68.
22. Walker JE, Saraste M, Runswick MJ, et al. Distantly related sequences in the alpha- and beta-subunits of ATP synthase, myosin, kinases and other ATP-requiring enzymes and a common nucleotide binding fold. EMBO J. 1982;1:945–951.
23. Singleton MR, Wigley DB. Modularity and specialization in superfamilies 1 and 2 helicases. J Bacteriol. 2002;184:1819–1826.
24. Matusan AE, Pryor MJ, Davidson AD, et al. Mutagenesis of the Dengue virus type 2 NS3 protein within and outside helicase motifs: effects on enzyme activity and virus replication. J Virol. 2001;75:9633–9643.
25. Frick DN, Banik S, Rypma RS. Role of divalent metal cations in ATP hydrolysis catalyzed by the hepatitis C virus NS3 helicase: magnesium provides a bridge for ATP to fuel unwinding. J Mol Biol. 2007;365:1017–1032.
26. Cabezon E, de la Cruz F. TrwB: an F(1)-ATPase-like molecular motor involved in DNA transport during bacterial conjugation. Res Microbiol. 2006;157:299–305.
27. Matusan AE, Pryor MJ, Davidson AD, et al. Mutagenesis of the dengue virus type 2 NS3 protein within and outside helicase motifs: effects on enzyme activity and virus replication. J Virol. 2001;75:9633–9643.
28. Regato M, Recarey R, Moratorio G, et al. Phylogenetic analysis of the NS5 gene of dengue viruses isolated in Ecuador. Virus Res. 2008;132:197–200.

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