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**Haemaphysalis hoodi** (Acari: Ixodidae) on a human from Yaoundé, Cameroon, and its molecular characterization

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**Abstract**

The genus *Haemaphysalis* Koch, 1844 (Acari: Ixodidae) is the second-largest genus, with more than 170 described species that primarily parasitize mammals and birds (Guglielmone et al. 2014, Guglielmone et al. 2020). *Haemaphysalis* species are three-host ticks, mainly distributed in southern and southeastern Asia and tropical Africa (Guglielmone et al. 2014). The present study identified a tick, *Haemaphysalis hoodi* Warburton & Nuttall, 1909, collected from a human in Yaoundé, Cameroon. This tick species feed on birds in sub-Saharan Africa. To the best of our knowledge, this is the second record of *H. hoodi* from humans. In addition, 16S ribosomal RNA and cytochrome oxidase I sequences were generated for this species for the first time. Screening pan-*Rickettsia*-PCR infection gave a negative result.

**Keywords** Human · Tick · *Haemaphysalis hoodi* · Cameroon

**Introduction**

The genus *Haemaphysalis* Koch, 1844 (Acari: Ixodidae) is the second-largest genus, with more than 170 described species (Guglielmone et al. 2020). *Haemaphysalis* species are three-host ticks that primarily parasitize mammals and birds (Guglielmone et al. 2014). Species from this genus are mainly distributed in southern and southeastern Asia and tropical Africa, some species are known from Australia, and only a few species occur in the Americas (Guglielmone et al. 2014). *Haemaphysalis* species are reservoirs and vectors of many pathogenic microorganisms of animals and humans. For instance, *Haemaphysalis leachi* (Audouin, 1826) transmits *Babesia rossi* in dogs (Kamani 2021), as well as *Rickettsia conorii*, which causes human tick-bite fever, and *Coxiella burnetii*, the causative agent of Q fever (Hoogstraal 1956). Limited studies have been performed on *Haemaphysalis* species from wildlife that focused on their diversity and role as potential vectors and reservoirs of pathogens.

Out of 47 described *Haemaphysalis* species endemic to the Afrotropic region, eleven species are known from Cameroon, namely *H. aciculifer* Warburton, 1913, *H. camicasi* Tomlinson & Apanaskevich 2019, *H. hoodi*...
Warburton & Nuttall, 1909. H. houyi Nuttall & Warburton, 1915, H. leachi, H. moreli Camicas et al. 1972, H. paraleachi Camicas et al. 1983, H. parmata Neumann, 1905, H. princeps Tomlinson & Apanaskevich 2019, H. punctaleachi Camicas et al. 1973, and H. tauffliebi Morel 1965 (Morel and Mouchet 1958; Morel 1965; Camicas et al. 1972, 1973, 1983; Hoogstraal and El Kammah 1972; Apanaskevich et al. 2007; Tomlinson and Apanaskevich 2019). The haemaphysalid subgenus Ornithophysalis Hoogstraal & Wassef, 1973, comprises 19 species divided into five structural-biological groups (Hoogstraal and Wassef 1973; Camicas et al. 1998). Many of the species have not been adequately studied structurally, biologically, or epidemiologically (Hoogstraal and Wassef 1973). Haemaphysalis houyi is one of the four species of the Haemaphysalis doenizti group, which also includes H. doenizti Warburton & Nuttall, 1909, H. phasiana Saito, Hoogstraal & Wassef, 1974, and H. madagascariensis Colas-Belcour & Millot, 1948 (Camicas et al. 1998). This species is broadly distributed in sub-Saharan Africa (Hoogstraal 1956; Hoogstraal and Wassef 1973). Adults, nymphs, and larvae of H. houyi feed primarily on various groups of birds, while records from mammals are rare (Guglielmone et al. 2014). In Cameroon, H. houyi was recorded from different ground-feeding bird species (Hoogstraal 1956; Santos Dias 1958). Here, we report for the first time a specimen of this species collected from a human in Yaoundé, Cameroon, and provide data on its mitochondrial (16S rRNA, cox I) genes.

Material and methods

In late October 2021, a light brown tick was removed manually from the shoulder of a woman in Nkozoa in the Mefou and Afamba Division of the center region of Cameroon (3°52′53.2″N 11°41′54.6″E). The collected tick was transferred to a 1.5 ml tube containing 600 µl absolute ethanol and sent to Bundeswehr Institute of Microbiology, Munich, Germany, for investigation. The tick specimen was first identified using morphological keys (Hoogstraal 1956), under a Keyence VHX-900F microscope (Itasca, IL, USA). DNA was extracted using the QIAamp mini DNA extraction kit (Qiagen, Hilden, Germany) according to the manufacturer’s instructions. As this is a rare tick species and no sequences from this species are available, 16S rRNA (Halos et al. 2004) and cox I (Apanaskevich et al. 2011) mitochondrial genes were sequenced, and the sequences obtained were edited and compared with the respective sequences deposited in GenBank using BLASTN and phylogenetic analysis. Sequences for each gene were aligned using MAFFT (Katoh and Standley 2013) with default parameters and phylogenetic analyses performed with IQ-Tree2 v1.6.12 (Minh et al. 2020). Optimal evolutionary models were calculated for each gene: 16S (K3Pu + F + I + G4) and Cox1 (TIM2 + F + I + G4). Nodal support was estimated using ultrafast bootstrap (n = 10,000) and the 50% consensus trees were reported. The partial sequences of the mitochondrial 16S rRNA and cox I genes generated in this study for H. houyi species have been deposited in GenBank under the accession numbers ON189038 and ON191014. Additionally, Rickettsia spp. screening was performed using a previously published real-time PCR assay targeting a part of the gltA gene (Wölfel et al. 2008).

Results and discussion

The tick was identified as a female Haemaphysalis houyi. The specific characteristics of the female include moderately dense punctations on scutum, broadly salient palpi, and absence of posterodorsal spur on palpal segment II (Fig. 1A,B) (Hoogstraal 1956; Morel 1965). Birds, especially ground feeder birds, are specific hosts for species within the Ornithophysalis subgenus; although some species parasitize birds and mammals, others only parasitize mammals (Hoogstraal 1956; Hoogstraal and Wassef 1973).

Haemaphysalis houyi is a very rare parasite of humans (Guglielmone and Robbins 2018). Adults of H. houyi have been found in three cases of human infestation in Ivory Coast although the exact localities were not reported (cited in Guglielmone et al. 2018). Our finding represents the second record of this tick species feeding on humans. Phylogenetic analysis indicated that the 16S rRNA sequence obtained (305 bp) from H. houyi group in a moderately supported clade with H. bancrofti, H. doenizti, and H. phasiana (Fig. 2). Of interest is that H. bancrofti is also part of this clade since it is classified in the subgenus Kaiseriana Dias, 1963, while H. houyi and H. phasiana are classified in the Ornithophysalis Hoogstraal and Wassef, 1973, subgenus (Hoogstraal and Wassef 1973). Similarly, in the cox I analysis (636 bp), H. houyi group in a well-supported clade with H. bancrofti, H. humerosa, and H. lagostrophi, the latter two species also belonging to the subgenus Ornithophysalis (Hoogstraal and Wassef 1973). While the overall support for the trees was weak, in each tree, several clades with good bootstrap support were obtained. No overwhelming support was found for any of the subgenera as monophyletic lineages. This may be due to limited phylogenetic signal due to the short sequences used in the analysis. It may, however, be noted that a recent analysis using 10 mitochondrial genes also resulted in a paraphyletic Haemaphysalis subgenus (Kelava et al. 2021).

As such, more studies that focus on molecular systematics of the Haemaphysalis subgenera are needed to ascertain the validity.
of various subgenera. Even so, both 16S rRNA and cox I indicate that H. hoodi presents a unique genetic signature compared to other sequences available in the database that shows a genetic relationship to other members of the Ornithophysalis subgenus.

Rickettsia spp. DNA was not amplified in the sample obtained from the H. hoodi female. It would have been expected to detect Rickettsia africae, which is responsible for the African tick-bite fever, mainly transmitted by Amblyomma species or Rickettsia aeschlimannii, a Hyalomma species related rickettsiae. Haemaphysalis species from Africa are not known as vectors for Rickettsia species. Haemaphysalis leachi was supposed to be a vector for Rickettsia conorii in southern Africa, but no isolates are available to confirm this. In Asia, especially in China, many Haemaphysalis

**Fig. 1** Haemaphysalis hoodi female collected from a human in Cameroon: A dorsal view, B ventral view

**Fig. 2** Maximum likelihood analysis of the 16S rRNA and cox I genes for the genus Haemaphysalis. Bootstrap support above 80% is indicated and the trees were rooted with Ixodes scapularis. The accession numbers used for the 16S rRNA and cox I genes are indicated behind the species names, respectively, and the tick sequenced in the current study is underlined. Subgenera are indicated in parentheses.

A) 16S rRNA

- Haemaphysalis aborensis KC170735 (Aborphysalis)
- Haemaphysalis formosensis JX572135 (Aborphysalis)
- Haemaphysalis punctata MT79994 (Aborphysalis)
- Haemaphysalis concinna KY546406 (Aborphysalis)
- Haemaphysalis flavus MG304958 (Aborphysalis)
- Haemaphysalis japonica NC037245 (Aborphysalis)
- Haemaphysalis megalaspidea AB61938 (Haemaphysalis)
- Haemaphysalis quinqueasianica MF632882 (Herpetobius)
- Haemaphysalis justakochi 1723288 (Goniocides)
- Haemaphysalis natalensis KY402315 (Herpetobius)
- Haemaphysalis japonica NP05996 (Goniocides)
- Haemaphysalis sopciopacenta MT027054 (Aborphysalis)
- Haemaphysalis wellingtoni MG194292 (Kaisertana)
- Haemaphysalis yensy AB819023 (Kaisertana)
- Haemaphysalis hispanicus ME307115 (Riphiptoma)
- Haemaphysalis para MT299714 (Segalia)
- Haemaphysalis salicata MT79994 (Herpetobius)
- Haemaphysalis orfki KU888549 (Riphiptoma)
- Haemaphysalis asiatica KC176734 (Riphiptoma)
- Haemaphysalis campimartia MT191710 (Haemaphysalis)
- Haemaphysalis pentalagi AB619199 (Haemaphysalis)
- Haemaphysalis leachi MM661151 (Riphiptoma)
- Haemaphysalis montgomeryi NC058313 (Segalia)
- Haemaphysalis elliptica HM069895 (Riphiptoma)
- Haemaphysalis mohucanen MN032115 (Riphiptoma)
- Haemaphysalis hisricus KC397865 (Kaisertana)
- Haemaphysalis bispinosa OM47850 (Kaisertana)
- Haemaphysalis intermedius MH04415 (Kaisertana)
- Haemaphysalis lagrangei KC170731 (Kaisertana)
- Haemaphysalis meshejimensis OK457150 (Kaisertana)
- Haemaphysalis longicornis NC037483 (Kaisertana)
- Haemaphysalis obesa KC170732 (Kaisertana)
- Haemaphysalis omniruphala OK457151 (Kaisertana)
- Haemaphysalis comigera AB619174 (Kaisertana)
- Haemaphysalis shigaoka KC170720 (Kaisertana)
- Haemaphysalis riphyptoma NC022330 (Allocreae)
- Haemaphysalis kitakai AB19902 (Allocreae)
- Haemaphysalis komini MZ054309 (Allocreae)
- Haemaphysalis hemorosa JX571318 (Ornithophysalis)
- Haemaphysalis aciulifor L034558 (Kaisertana)
- Haemaphysalis bancrofti NC041076 (Kaisertana)
- Haemaphysalis dornitzi JF03483 (Ornithophysalis)
- Haemaphysalis phasiana AB619220 (Ornithophysalis)
- Haemaphysalis houdi ON195038 (Ornithophysalis)

B) Cytochrome oxidase 1

- Haemaphysalis leechi MM662015 (Goniocides)
- Haemaphysalis formosensis JX572135 (Aborphysalis)
- Haemaphysalis quinqueasianica MF632882 (Herpetobius)
- Haemaphysalis punctata MT027054 (Aborphysalis)
- Haemaphysalis flava MG304958 (Aborphysalis)
- Haemaphysalis natalensis KY402315 (Herpetobius)
- Haemaphysalis japonica NC037245 (Aborphysalis)
- Haemaphysalis salicata MT79994 (Herpetobius)
- Haemaphysalis orfki KU888549 (Riphiptoma)
- Haemaphysalis asiatica KC176734 (Riphiptoma)
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- Haemaphysalis leechi MM661151 (Riphiptoma)
- Haemaphysalis montgomeryi NC058313 (Segalia)
- Haemaphysalis leachi MM661151 (Riphiptoma)
- Haemaphysalis longicornis NC037483 (Kaisertana)
- Haemaphysalis hystricus NC037483 (Kaisertana)
- Haemaphysalis ripphyptoma OK457150 (Kaisertana)
- Haemaphysalis hemorosa JX571318 (Ornithophysalis)
- Haemaphysalis leachi MM661151 (Riphiptoma)
- Haemaphysalis riphyprpha OK457151 (Kaisertana)
- Haemaphysalis houdi ON195038 (Ornithophysalis)
- Haemaphysalis leachi MM661151 (Riphiptoma)
- Haemaphysalis ripphyptoma OK457150 (Kaisertana)
- Haemaphysalis hemorosa JX571318 (Ornithophysalis)
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- Haemaphysalis ripphyptoma OK457150 (Kaisertana)
species are vectors for *Rickettsia* spp., e.g., *Rickettsia sibirica*, *Rickettsia heilongjiangensis*, and *Rickettsia japonica* (Raoult and Parola 2007).

**Author contribution** A.P. and M.K. collected and provided the tick for investigation. B.M. did the genetic analysis, sequences submission, and prepared Fig. 2. D.A.A. contributed to the morphological identification. L.C.D. did the morphological identification and the further lab work (DNA extraction and all tests) and prepared Fig. 1. A.P., B.M., D.A.A., and L.C.D. wrote the manuscript. All authors read the final version of the manuscript.

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**Data Availability** The sequences were submitted to GenBank under the following access numbers: ON189038, ON191014. The tick specimen is in the LCD collection at Bundeswehr Institute of Microbiology.

**Declarations**

**Competing interests** The authors declare no competing interests.

**Ethics approval** Not applicable.

**Consent to participate** Not applicable.

**Consent for publication** All authors gave their consent for publication.

**Conflict of interest** The authors declare no competing interests.

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