Simultaneous Determination of Salbutamol Sulphate and Bromhexine Hydrochloride in Tablets by Reverse Phase Liquid Chromatography

P. N. S PAI*, G. K. RAO, M. S. MURTHY, A. AGARWAL AND S. PURANIK
Department of Quality Assurance, Al-Ameen College of Pharmacy, Bangalore-560 027, India

Pai et al.: RPLC Determination of Salbutamol and Bromhexine

A simple reverse phase liquid chromatographic method has been developed and subsequently validated for simultaneous determination of salbutamol sulphate and bromhexine hydrochloride. The separation was carried out using a mobile phase consisting of acetonitrile, methanol and phosphate buffer, pH 4 in the ratio 60:20:20 v/v. The column used was SS Wakosil-II C-18 with a flow rate of 1 ml/min and UV detection at 224 nm. The described method was linear over a concentration range of 10-110 µg/ml and 20-140 µg/ml for the assay of salbutamol sulphate and bromhexine hydrochloride, respectively. The mean recovery was found to be 95-105% for salbutamol sulphate and 96.2-102.1% for bromhexine hydrochloride when determined at five different levels.

Key words: Salbutamol sulphate, bromhexine hydrochloride, HPLC, UV detection

Salbutamol sulphate and bromhexine hydrochloride as components of a multi-ingredient formulation is useful in therapy of asthma. As found from the literature, salbutamol sulphate and bromhexine hydrochloride can be estimated individually by HPLC and spectrometric methods. Salbutamol sulphate and bromhexine hydrochloride has been reported to be estimated by spectrometric methods[1-8]. Salbutamol sulphate and bromhexine hydrochloride have been simultaneously determined by spectrometric[9] and HPLC methods[10]. The aim of the present work is to describe a liquid chromatographic procedure for the separation and simultaneous estimation of salbutamol sulphate and bromhexine hydrochloride in its formulation.

For the proposed method, acetonitrile HPLC grade, methanol HPLC grade, potassium dihydrogen phosphate, orthophosphoric acid and distilled water (Millipore) were used. The LC system consisted of Shimadzu LC-10AT pump, SS Wakosil-II C-18, 250×4.6 mm, 5 µm column, Rheodyne injector equipped with a 100 µl sample loop and UV Shimadzu SPD-10A VP detector, set at 224 nm. The output signal was monitored and integrated using Shimadzu CZ-RA software. Phosphate buffer pH 4, was prepared by dissolving 8.95 g of disodium hydrogen phosphate. 12 H2O and 3.40 g of potassium dihydrogen phosphate in 1000 ml of distilled water. The standard stock solution of salbutamol sulphate 1 mg/ml and bromhexine hydrochloride 1 mg/ml were prepared separately in mobile phase of acetonitrile, methanol and phosphate buffer, pH 4 in the ratio 60:20:20 v/v. The working standard solutions of salbutamol sulphate and bromhexine hydrochloride were prepared by diluting volumes ranging from 1 to 15 ml of standard stock solution of both the drugs separately to 100 ml with the mobile phase. The working standard solutions were injected into the chromatograph. The retention time for salbutamol sulphate and bromhexine hydrochloride at a flow rate of 1 ml/min were recorded as 4.2 and 6.3 min, respectively.

Analysis of marketed sample Mucolinc tablets (Cipla Ltd., Mumbai, India) of three different batches was carried out. Twenty tablets, each containing 2 mg salbutamol sulphate and 8 mg bromhexine hydrochloride were weighed. The tablets were crushed together in a mortar to a fine powder and an amount equivalent to 2 mg salbutamol sulphate and 8 mg bromhexine hydrochloride were weighed. The tablets were crushed together in a mortar to a fine powder and then volume made up with the mobile phase to obtain sample stock solution. The solution was filtered through 0.45 µ Whatman filter paper. The sample solutions were injected into the stabilized liquid chromatographic system. From the respective
peak areas obtained from the standard and sample chromatogram, the amount of salbutamol sulphate and bromhexine was calculated. The results of the analysis are tabulated in Table 1.

Accuracy of the method was checked by recovery studies, wherein sample was spiked with known quantity of standard drug of salbutamol sulphate and bromhexine hydrochloride at 5 different levels. The percentage recovery ranged from 95-105% for salbutamol sulphate and 96.2-102.1% for bromhexine hydrochloride. The precision of the method was studied by analysis of the mixture and expressed as percentage relative standard deviation, which was found to be 0.13% for salbutamol sulphate and 0.004% for bromhexine hydrochloride.

The linearity of the method was established by analysis of standard solution. The calibration curve was drawn by plotting the peak area versus concentration. The linearity range was found to be 10-110 µg/ml for salbutamol sulphate and 20-140 µg/ml for bromhexine hydrochloride. The specificity of the method was established by injecting placebo. No interference of the placebo was observed with the principal peaks. Ruggedness of the method was determined by carrying out the experiment on different instruments, by different chemists and on different days. The results showed that the method was rugged as percentage recovery was found to in the range of 95-100.5% for both of the drugs under study. The robustness of the method was determined by making slight changes in the chromatographic conditions. Buffer pH modification did not have any significant effect. The effect of organic strength on retention time was studied by small change in percentage polarity of the mobile phase system. It was found that even slight percentage change, up to 10% in ratio of mobile phase did not alter the position of the peaks.

The system suitability tests were carried out as per USP XXIV requirements. System suitability tests were carried out on freshly prepared standard stock solution of salbutamol sulphate and bromhexine and the parameters obtained with 100 µl injection volume. The number of theoretical plates for salbutamol sulphate and bromhexine hydrochloride was calculated as 19944 and 30337, respectively. The symmetry factor for salbutamol sulphate and bromhexine hydrochloride peak was found to be 1.08 and 1.11. The resolution between the two peaks was 1.85. The obtained results confirmed that the method is highly suitable for its intended purpose of separation of salbutamol sulphate and bromhexine hydrochloride and its simultaneous determination in tablet formulations.

In the reported method for estimation of salbutamol sulphate and bromhexine hydrochloride\textsuperscript{[10]} the retention time has been reported to be more than 7 min. The proposed method is specific, accurate, rugged, robust, and precise as found from the laboratory studies. The method when applied for the determination of salbutamol sulphate and bromhexine hydrochloride in combined dosage marketed formulation, gave results conforming to the label claim of the drug.

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**REFERENCES**

1. Bhatt KK, Shah SA, Pandya SS. Spectrophotometric determination of Salbutamol sulfate and its dosage forms. Indian Drugs 1999;36:524-8.
2. Talwar N, Singhai AK, Shakya AK, Saraf S, Jain NK. Difference spectrophotometric determination of Salbutamol sulfate in tablets. Indian Drugs 1991;28:244-5.
3. Naidu NV, Naidu DV, Rajeswari CV, Naidu PR. Simple spectrophotometric determination of Salbutamol sulfate in pharmaceutical formulations. Acta Chem Hung 1989;126:821-4.
4. Geeta N, Baggi TR. Improved spectrophotometric methods for the

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**TABLE 1: DETERMINATION OF SALBUTAMOL SULPHATE AND BROMHEXINE HYDROCHLORIDE CONTENT IN MUCOLINC TABLETS**

| Batch | Content of salbutamol sulfate (mg/tablet) | Amount of salbutamol sulfate found* (mg/tablet) | Content of bromhexine hydrochloride (mg/tablet) | Amount of bromhexine hydrochloride found* (mg/tablet) |
|-------|-------------------------------------------|-----------------------------------------------|-----------------------------------------------|--------------------------------------------------|
| Bt-1  | 2                                         | 1.95                                          | 8                                             | 7.96                                             |
| Bt-2  | 2                                         | 1.98                                          | 8                                             | 7.66                                             |
| Bt-3  | 2                                         | 1.97                                          | 8                                             | 7.81                                             |

*Each value is an average of six determinations.
5. Zarapkar SS, Rele RV, Doshi VJ. Simple spectrophotometric methods for the estimation of bromhexine hydrochloride. Indian Drugs 1988;26:38-41.
6. Santoro MI, Dos Santos MM, Magalhaes JF. Spectrophotometric determination of bromhexine hydrochloride in pharmaceutical preparations. J Anal Chem 1984;67:532-4.
7. Shingbal DM, Sardesai GD. Estimation of bromhexine hydrochloride in pharmaceutical preparations. Indian Drugs 1987;24:417-8.
8. Gala B, Gomez, Hens A, Bendito D. Direct kinetic determination of bromhexine hydrochloride in pharmaceutical formulations. Anal Lett 1993;26:2607-17.
9. Bhatia NM, Jain DK, Trivedi P. Simultaneous analysis of Salbutamol sulfate and bromhexine hydrochloride from solid dosage form using multiwavelength UV-Spectrophotometry. Indian Drugs 1998;35:566-9.
10. Rao GR, Raghuveer S, Khadgapathi P. High Performance liquid chromatographic determination of Salbutamol sulfate and bromhexine hydrochloride in its combined dosage forms. Indian Drugs 1987;25:15-20.

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**Catalase in Testes and Epididymidis of Wistar Rats Fed Zinc Deficient Diet**

S. BEDWAL, S. PRASAD, N. NAIR, M. R. SAINI1 AND R. S. BEDWAL*

Cell Biology Laboratory and 1Radiation Biology Laboratory, Department of Zoology, University of Rajasthan, Jaipur-302 004, India

Bedwal et al.: Catalase in testes and epididymidis of zinc deficient rats

Catalase activities have been evaluated in testes and caput and cauda epididymis of Wistar rats fed on zinc deficient diet for 2 and 4 weeks. The enzyme activity has been measured as chromic acetate formed by heating of dichromate (in acetic acid) in presence of H2O2 with perchromic acid as an unstable intermediate. Observed non-significant increase in catalase activity in testes as well as in caput and cauda epididymis of 2 weeks experiments has been related to low levels of H2O2 produced in two organs whereas significant (P<0.01/0.001) increase in catalase activity in 4-weeks experiments indicate for increased oxidative stress due to phagocytotic activity of Sertoli cells in testes and damaged spermatozoa in epididymis. Thus, zinc deficiency increases catalase activity in testes and epididymis.

**Key words:** Catalase, testes, epididymis, and zinc deficiency

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Zinc is indispensable element for growth, reproduction, development, differentiation, immune and antioxidant functions, gene expression, DNA synthesis, hormone synthesis, storage and release of neurotransmitters, memory, visual processes and apoptosis[1,2]. Cardinal symptoms of zinc deficiency are retarded growth and hypogonadism [2]. Oxidative stress has been suggested to be an early effect of zinc deficiency rather than a simple reflection of zinc deficiency-induced tissue pathology[3]. Reactive oxygen species (ROS) or free oxygen radical (FOR) are normally generated by Sertoli cells that cause alteration in cellular structures and induces morphological changes in spermatids during spermiogenesis[4] and controlled amount of ROS is essential for capacitation and acrosome reaction [5]. In fact, analysis of superoxide radical generated by rat epididymal spermatozoa has revealed a two component process involving leakage from sperm mitochondria at complex I and II and a plasma membrane NAD(P)H oxidoreductase whose activity is regulated by zinc [6,7]. In spite of these pivotal roles played by ROS in reproduction they have, in general, implicated in injuries of testes and spermatozoa leading to infertility. Catalase (CAT) has been reported from testis, seminal plasma, and spermatozoa and in peroxisomes of rodent Leydig cells[8]. A sharp eight-fold drop in CAT activity in rabbit testes from day 64 to 101 of age (this period corresponds to completion of Leydig cell maturation and on set of pre-pubertal spermatogenic cycle including spermatogenesis) suggests for a possible relationship between testosterone and catalase.