Follicular lymphoma patients with KIR2DL2 and KIR3DL1 and their ligands (HLA-C1 and HLA-Bw4) show improved outcome when receiving rituximab

Amy K. Erbe, Wei Wang, Lakesha Carmichael, Anna Hoefges, Bartosz Grzywacz, Patrick K. Reville, Erik A. Ranheim, Jacquelyn A. Hank, KyungMann Kim, Songwon Seo, Eneida A. Mendonca, Yiqiang Song, Vaishalee P. Kenkre, Fangxin Hong, Randy D. Gascoyne, Elisabeth Paietta, Sandra J. Horning, Jeffrey S. Miller, Brad Kahl and Paul M. Sondel

Abstract

Background: The ECOG-ACRIN Cancer Research Group evaluated rituximab treatment schedules for patients with newly-diagnosed low-tumor-burden follicular-lymphoma (FL). All patients received 4-weekly rituximab treatments as induction therapy. Clinically-responding patients were randomized to receive rituximab every 13 weeks ("maintenance") vs. no additional rituximab until progression ("non-maintenance"). Based on "time-to-rituximab-failure (TTRF)", the study-committee reported there was no overall-benefit for maintenance rituximab in this setting. Tumor-reactive mAbs, like rituximab, trigger natural killer (NK) cells. NK-cell responses are regulated, in part, by interactions between killer immunoglobulin-like receptors (KIRs) on NK cells and their interactions with KIR-ligands. In a separate study of children with neuroblastoma treated with a different mAb, we found certain KIR/KIR-ligand genotypes associated with improved outcome. Here, we assessed whether a subset of FL patients show improved outcome from the maintenance rituximab based on these same KIR/KIR-ligand genotypes.

Methods: Genotypes for KIR/KIR-ligand were determined and assessed for associations with outcome [duration of response, TTRF and % tumor shrinkage] as a post-hoc analysis of this phase III trial. Our primary objective was to assess specific KIR/KIR-ligand genotype associations, followed by separate prespecified KIR/KIR-ligand genotype associations in follow-up analyses. Statistical analyses for association of genotype with clinical outcome included: Log-rank tests and Cox proportional hazards regression models to assess duration of response and TTRF; analysis of variance (ANOVA) was used for assessment of % tumor shrinkage.

Results: We found that patients inheriting KIR2DL2 and its ligand (HLA-C1) along with KIR3DL1 and its ligand (HLA-Bw4) had improved outcome over patients without this genotype. In addition, patients with KIR2DL2 and HLA-C1 along with KIR3DL1 and HLA-Bw4 also showed improved duration of response and tumor shrinkage if they received maintenance, while patients without this genotype showed no such improvement when receiving maintenance.

(Continued on next page)
Conclusions: The data presented here indicate that a subset of FL patients, identified by certain KIRs/KIR-ligands, have improved outcome and may benefit from additional rituximab treatment. Taken together, this suggests that the efficacy of tumor-reactive mAb treatment for some patients is influenced by KIRs on NK cells. However, prior to considering these genotypes in a clinically-actionable manner, these findings need independent validation in other studies.

Keywords: KIR, HLA, Follicular lymphoma, NK cells, Rituximab, Immunotherapy, Monoclonal antibody, ADCC, CD20, MHC class I

Background
The most common form of indolent lymphoma is follicular lymphomas (FL). The use of rituximab to treat FL has markedly transformed the care of these patients [1, 2]. For patients with low-tumor burden FL, a maintenance rituximab strategy has been shown to improve the progression-free survival following induction with either chemotherapy or rituximab [3–5]. Yet, whether other clinical outcome parameters could benefit from the continual rituximab treatment schedule (maintenance rituximab vs. a close “watch and wait” approach) was unclear [6]. Additionally, concerns related to the added healthcare expenditures required for a maintenance treatment schedule suggested that it may be more cost-efficient to treat with rituximab on a less frequent basis [6–8]. As such, the ECOG-ACRIN Cancer Research Group (ECOG-ACRIN) conducted and reported results from a phase III clinical trial (E4402) to determine the optimal rituximab dosing strategy for patients with low-tumor burden FL [6]. In this report of the E4402 trial, Kahl and colleagues concluded that maintenance rituximab treatment (continual doses of rituximab every 13 weeks) provided no benefit in the time to rituximab failure for this population of low-tumor burden FL patients, compared to a non-maintenance rituximab treatment schedule (an additional course of 4 weekly doses of rituximab only upon disease progression) [6]. The purpose of this present study was to determine if inherited genotypic variances in genes that influence immune function, and potentially rituximab’s antitumor effects, may identify subpopulations of patients that differ in their outcome following maintenance vs. non-maintenance rituximab schedules [8, 9].

In a separate study of FL patients, patients with lower NK cell counts had inferior clinical prognosis [10]. This result suggests that NK cell count may be used as a prognostic biomarker for FL patients, and that treatments designed to activate NK cells might potentially be beneficial [10]. NK cells contribute to the anti-tumor effects of rituximab via antibody-dependent cell-mediated cytotoxicity (ADCC), and several studies have assessed NK cell specific immunogenetic factors that may be predictive of response to rituximab treatment in FL patients [11–15]. We hypothesize that NK cell specific immunogenetic factors influence the clinical outcome following rituximab treatment for some FL patients, and that the maintenance rituximab treatment schedule differentially impacts clinical outcome dependent upon individual genotypic differences.

NK cell activation is based on the balance of inhibitory and activating signals transmitted by receptors on NK cells. One class of these receptors is killer-cell immunoglobulin-like receptors (KIRs); some of these interact with certain class I HLA molecules (which can function as KIR ligands) to modulate NK cell responses [16, 17]. The interactions between endogenous KIRs and KIR-ligands modulate NK cell function and immunotherapeutic responses [12, 18–21]. During NK cell maturation, NK cells lacking inhibitory KIRs specific for self-HLA class I become less potent than NK cells expressing one or more inhibitory receptors for self-HLA class I through a process termed licensing [22, 23]. Yet, these same inhibitory KIRs can suppress mature NK cells through specific interactions with the class I HLA molecules that function as their ligands when expressed by tumors and other nucleated cells.

Prior clinical studies have reported associations between KIR/KIR-ligand genotypes and patient clinical response in various immunotherapeutic settings that likely involve NK cells [12, 18, 20, 24–29]. In a separate randomized clinical trial of patients with neuroblastoma, we have shown that patients with certain KIR/KIR-ligand genotypes benefited from treatment with mAb-based immunotherapy [dinutuximab (anti-GD2 mAb) + GM-CSF + interleukin-2] while patients with the opposing KIR/KIR-ligand genotype were not clinically influenced by the immunotherapy treatment [30]. In addition, we found that patients that did not receive immunotherapy responded differently dependent upon which KIR/KIR-ligand genotypes they had, suggesting that NK cells influence outcome even in the absence of immunotherapy [30]. In this study, we assessed whether the presence of certain KIR/KIR-ligand genotypes might affect whether the use of maintenance rituximab influences outcome. Separately, we also investigated whether certain KIR/
KIR-ligand genotypes influenced the clinical outcome based on the rituximab treatment schedule (i.e., how KIR/KIR-ligand genotype might affect clinical outcome within the maintenance or non-maintenance treatment schedules). The primary endpoint in the E4402 clinical trial was the time to rituximab failure (TTRF). In this present study, we evaluated associations of KIR/KIR-ligand genotype with TTRF, using determinants based on biological causes (detailed in the statistical section of Methods). We also assessed two additional biologically relevant clinical parameters: duration of response and % tumor shrinkage. In the current report, we found that for subsets of patients with certain KIR/KIR-ligand genotypes (but not for others), clinical outcome (TTRF) was improved by the maintenance rituximab treatment schedule.

Methods
Clinical trial and clinical samples
The phase III ECOG-ACRIN E4402 clinical trial (ClinicalTrials.gov #NCT00075946) evaluated the efficacy of single agent, rituximab therapy for adults with low-tumor burden indolent lymphoma. Clinical results from this study have been reported elsewhere [6]. A total of 408 patients with follicular lymphoma were entered, 289 of which responded and were randomized to maintenance vs non-maintenance therapy with rituximab. Disease measurements were obtained every 13 weeks [6]. Of the 408 patients, 213 patients had evaluable DNA and clinical data; of the 289 responding randomized patients, 159 had evaluable DNA and clinical data for this study. Clinical and demographic data for the 213 patients and the 159 randomized patients for which DNA and clinical data were evaluable (non-maintenance n = 80 and maintenance n = 79) are included in Additional file 1: Table S1, along with comparative data for the 289 responding patients reported on in the primary clinical report. The clinical trial was conducted in accordance with the Helsinki Declaration of 1975.

Genotyping
KIR gene status was determined by a real-time PCR technique [31, 32]. The genotypes of KIR-ligand (HLA-C1, HLA-C2, HLA-Bw4) were performed in a blinded fashion, and determined by PCR-SSP using the KIR HLA Ligand SSP typing kit (Olerup) with GoTaq DNA Polymerase (Promega). Additional genotyping details can be found in the supplemental methods section.

Data management
Study data (genotyping data from our lab) were entered into and managed using the REDCap system hosted at the University of Wisconsin-Madison. REDCap (Research Electronic Data Capture) is a secure, HIPAA-compatible, web-based application designed to support data capture for research studies, providing: 1) an intuitive interface for validated data entry; 2) audit trails for tracking data manipulation and export procedures; and 3) procedures for importing data from external sources [33]. The clinical outcome data from the ECOG-ACRIN study database (which is HIPAA compliant) in Excel were merged with the genotyping data in REDCap to create a SAS dataset for analysis.

KIR/KIR-ligand interaction analysis
Individuals that have all KIR-ligands present for the inhibitory KIRs they possess were defined as having a “KIR-ligands present” genotype. Individuals that lack any KIR-ligand for any one of the KIR genes they possess were defined as having a “KIR-ligand missing” genotype [18, 20, 29, 30]. Detailed descriptions of these genotypes can be found in Additional file 1: Table S2.

Statistical methods
The primary objective was to evaluate the association of clinical outcome with treatment regimen and KIR-ligand status (all KIR-ligands present compared with KIR-ligands missing). Other analyses were exploratory, but the KIR/KIR-L genotype combinations evaluated here were performed based on associations with outcome for similar KIR/KIR-L genotype combinations in a prior study of neuroblastoma patients treated with a separate mAb [30]. Thus, statistical analyses were performed, and p values are reported, without any adjustment for multiplicity of testing. Only randomized patients were included in the analyses. The post-hoc analysis of the clinical outcomes from this phase III trial that were assessed included the duration of response (n = 155), the time to rituximab failure (TTRF) (n = 159), and % tumor shrinkage (n = 139). For each of the clinical parameters assessed, Additional file 1: Table S3 includes the mean/median response data with 95% confidence intervals. The duration of response was defined as the time from randomization (following an initial response to the induction rituximab treatment) to documented first disease progression. The TTRF was defined as the time from randomization to treatment failure, as reported by Kahl et al. [6]. Treatment failure was defined as whichever came first of: 1) the time at which patients no longer responded to rituximab [disease progression for patients receiving maintenance; no response to retreatment rituximab or time to progression [< 26 weeks from day 1 of last rituximab for patients in the non-maintenance treatment schedule], 2) the time at which an alternative therapy was initiated, or 3) the time at which patients were determined unable to complete their assigned rituximab schedule [6]. For all TTRF analyses reported here, treatment failures that were considered non-biological (largely the
The decision to change to alternate therapy by the patient or physician) were censored, and as presented in the supplemental materials in the initial clinical report [6].

The % tumor shrinkage was defined as the % change in tumor size from the baseline measurement at the time of randomization (measured 13 weeks after initiating the induction rituximab treatment) to the smallest tumor size obtained post-randomization, only including those who had a partial response 13 weeks after initiating the induction rituximab (n = 139). While all evaluable randomized patients could be analyzed for duration of response, tumor shrinkage following randomization could not be calculated for 16 patients (10 in non-maintenance and 6 in maintenance) that had achieved a complete response with a tumor measurement of zero at week 13 of induction, as it was impossible to measure any further shrinkage after their complete response measured at week 13. Changes in tumor size were represented using box plots (described in detail in supplemental materials).

Log-rank tests and Cox proportional hazards regression models were used to compare the duration of response and TTRF by treatment and genotype combinations. For the % tumor shrinkage, analysis of variance (ANOVA) was used. For our analyses, the associations between outcome and KIR/KIR-ligand were evaluated using Cox regression models with treatment group and KIR/KIR-ligand genotype as the main effects. In addition, we evaluated for possible interaction effects between treatment schedule and KIR/KIR-ligand genotype on outcome. For the analyses assessing associations of outcome with specific KIRs and their ligands (as in Tables 1 and 2), we set a minimal p-value of p < 0.100 in the interaction analysis in order to subsequently perform association comparisons directly between individual genotype groups and outcome. Statistical analyses were performed using SAS v9.4 (SAS Institute, Cary, NC).

**Results**

KIR ligand missing status does not significantly influence TTRF, duration of response or tumor shrinkage

In some previous studies, patients with at least one KIR-ligand missing ("KIR-ligand missing") had improved clinical outcome as compared to those with all KIR-ligands present ("KIR-ligands present") when treated with NK-based immunotherapy [18, 25, 26, 29]. The genotypes used to define KIR-ligands present vs. KIR-ligand missing are detailed in Additional file1: Table S2. Based on the findings from these prior studies, we hypothesized that FL patients that received rituximab therapy that had the KIR-ligand missing genotype would have improved outcome as compared to those patients with KIR-ligands present. However, here we found no significant association between TTRF with KIR-ligands present vs. KIR-ligand missing status (Fig. 1a) amongst either those receiving maintenance or those receiving non-maintenance.

### Table 1 Interaction analyses for individual KIR and KIR ligand genotypes with TTRF

| Line | Treatment | Genotype Group | Number of Events/n | 4 yr Fail Rate (95% CI) | Interaction p-value |
|------|-----------|----------------|--------------------|------------------------|---------------------|
| 1    | Maintenance | KIR2DL1+/C2+ | 21/52              | 38 (25–54)             | 0.524               |
| 2    | Maintenance | not KIR2DL1+/C2+ | 7/28             | 30 (15–56)             |                     |
| 3    | Non-Maintenance | KIR2DL1+/C2+ | 18/49              | 38 (26–55)             |                     |
| 4    | Non-Maintenance | not KIR2DL1+/C2+ | 6/30             | 21 (9–43)              |                     |
| 5    | Maintenance | KIR2DL2+/C1+ | 11/36              | 32 (18–53)             | 0.547               |
| 6    | Maintenance | not KIR2DL2+/C1+ | 17/44             | 38 (25–56)             |                     |
| 7    | Non-Maintenance | KIR2DL2+/C1+ | 6/28               | 24 (10–49)             |                     |
| 8    | Non-Maintenance | not KIR2DL2+/C1+ | 18/51             | 36 (24–52)             |                     |
| 9    | Maintenance | KIR2DL3+/C2+ | 17/56              | 33 (21–50)             | 0.982               |
| 10   | Maintenance | not KIR2DL3+/C2+ | 11/24             | 41 (23–64)             |                     |
| 11   | Non-Maintenance | KIR2DL3+/C2+ | 16/60              | 27 (17–43)             |                     |
| 12   | Non-Maintenance | not KIR2DL3+/C2+ | 8/19              | 45 (25–70)             |                     |
| 13   | Maintenance | KIR3DL1+/Bw4+ | 20/54              | 38 (25–55)             | 0.055               |
| 14   | Maintenance | not KIR3DL1+/Bw4+ | 8/26             | 29 (15–52)             |                     |
| 15   | Non-Maintenance | KIR3DL1+/Bw4+ | 13/50              | 25 (15–41)             |                     |
| 16   | Non-Maintenance | not KIR3DL1+/Bw4+ | 11/29             | 43 (25–66)             |                     |

*95% Confidence interval; lines 13-14 (bolded text) had a p-value <0.100 and were analyzed further for associations with outcome*
Similarly, KIR-ligands present vs. KIR-ligand missing status did not influence duration of response (Fig. 1b) or tumor shrinkage (Fig. 1c) for either treatment regimen. In addition, the treatment regimen (maintenance vs. non-maintenance) did not influence TTRF for patients with either KIR-ligands present or KIR-ligand missing (Fig. 1a), consistent with the data for the overall group of patients from the initial clinical report [6]. In contrast, amongst patients with KIR ligands present, those receiving maintenance showed significantly increased duration of response (Fig. 1b) and tumor shrinkage (Fig. 1c), and those with KIR ligands missing that received maintenance also showed increased duration of response (Fig. 1b).

**Specific individual inhibitory KIR/KIR-ligand combinations:**

The presence of KIR3DL1 and its HLA-Bw4 ligand, but not absence of KIR3DL1 and HLA-Bw4, are associated with improved outcome for patients receiving rituximab maintenance treatment

We hypothesized that patients with all KIR-ligands present have NK cells that are more inhibited due to the presence of KIR-ligands for all of the inhibitory KIRs inherited [18, 20]. Yet, there is also the concept that those patients with KIR-ligands present have more licensed NK cells that may be able to better lyse rituximab-treated tumor cells [22, 34, 35]. We further assessed whether there was a differential influence on outcome in this clinical trial dependent upon the presence or absence of specific inhibitory KIRs/KIR-ligands.

HLA-C alleles can be divided based on their KIR binding as HLA-C1 or HLA-C2 containing epitopes [19, 36]. As C1 and C2 are alleles, every individual will genotype as either HLA-C1/C1, HLA-C1/C2, or HLA-C2/C2. KIR2DL1 recognizes HLA-C2 as its ligand. Thus individuals with both KIR2DL1 and HLA-C2 (designated as KIR2DL1+/C2+) include those that have KIR2DL1+ and also have HLA-C2+ (HLA-C genotype of either C1/C2 or C2/C2). All other possible genotypes regarding KIR2DL1 and HLA-C are designated as not KIR2DL1+/C2+ and include the following genotypes: (KIR2DL1+/C2-, KIR2DL1+/C2+, or KIR2DL1-/C2-) described in detail in Additional file 1: Table S4. Similarly, KIR2DL3+/C1+ individuals have KIR2DL3+ with HLA-C1+ (HLA-C genotype of either C1/C1 or C1/C2); all other possible genotypes of KIR2DL2 and HLA-C are designated as not KIR2DL2+/C1+, as detailed in Additional file 1: Table S4. Similarly, KIR2DL3+/C1+ individuals have KIR2DL3+ with HLA-C1+ (HLA-C genotype of either C1/C1 or C1/C2); all other possible genotypes of KIR3DL1 and HLA-Bw4 are designated as not KIR3DL1+/Bw4+ (Additional file 1: Table S4).

To assess whether the effect of individual KIR/KIR-ligand pairs may be influenced by treatment regimen, we first did an interaction analysis between outcome for each treatment regimen and genotype status for individual KIR/KIR-ligand pairs (Table 1). Since TTRF was the primary endpoint in the E4402 clinical trial [6], for the initial interaction analysis, we used TTRF as our primary endpoint. Only the genotypes determined by KIR3DL1/ Bw4 showed an interaction \( p \)-value \( \leq 0.100 \) with

| Line | Treatment | Genotype Group | Number of Events/n | 4 yr Fail Rate (95% CI) \( \% \) | Interaction \( p \)-value |
|------|-----------|----------------|-------------------|---------------------------|-------------------|
| 1    | Maintenance | KIR2DL1+/C2+ and KIR3DL1+/Bw4+ | 17/44 | 37 (24–54) | 0.694 |
| 2    | Maintenance | not KIR2DL1+/C2+ and KIR3DL1+/Bw4+ | 11/36 | 34 (19–55) | \- |
| 3    | Non-Maintenance | KIR2DL1+/C2+ and KIR3DL1+/Bw4+ | 10/32 | 33 (18–54) | \- |
| 4    | Non-Maintenance | not KIR2DL1+/C2+ and KIR3DL1+/Bw4+ | 14/47 | 31 (19–48) | \- |
| 5    | Maintenance | Group 1: KIR2DL2+/C1+ and KIR3DL1+/Bw4+ | 9/22 | 45 (24–72) | 0.068 |
| 6    | Maintenance | Group 2: not KIR2DL2+/C1+ and KIR3DL1+/Bw4+ | 19/58 | 32 (21–47) | \- |
| 7    | Non-Maintenance | Group 1: KIR2DL2+/C1+ and KIR3DL1+/Bw4+ | 4/23 | 17 (6–44) | \- |
| 8    | Non-Maintenance | Group 2: not KIR2DL2+/C1+ and KIR3DL1+/Bw4+ | 20/56 | 38 (26–54) | \- |
| 9    | Maintenance | KIR2DL3+/C1+ and KIR3DL1+/Bw4+ | 12/35 | 38 (26–54) | 0.447 |
| 10   | Maintenance | not KIR2DL3+/C1+ and KIR3DL1+/Bw4+ | 16/45 | 34 (21–51) | \- |
| 11   | Non-Maintenance | KIR2DL3+/C1+ and KIR3DL1+/Bw4+ | 10/37 | 25 (13–44) | \- |
| 12   | Non-Maintenance | not KIR2DL3+/C1+ and KIR3DL1+/Bw4+ | 14/42 | 38 (24–57) | \- |

\( \% \) confidence interval; lines 5-8 (bolded text) had a \( p \)-value \( < 0.100 \) and were analyzed further for associations with outcome.

\( a \)
treatment regimen (corresponding to lines 13–16 in Table 1). This p-value of 0.055 justified further analyses of associations of outcome (shown in Fig. 2) with these 4 individual KIR/KIR-ligand groups.

For patients that were KIR3DL1+/Bw4+, those that received the maintenance regimen showed significantly improved duration of response (p < 0.001, Fig. 2b) and % tumor shrinkage (p = 0.04, Fig. 2c) vs. those that did not receive maintenance. There was no significant improvement in duration of response or tumor shrinkage associated with maintenance treatment vs. non-maintenance for the patients that were not KIR3DL1+/Bw4+. These results suggest that the maintenance regimen improved clinical outcome for KIR3DL1+/Bw4+ patients.

Additionally, amongst patients receiving maintenance rituximab, KIR3DL1+/Bw4+ patients had improved outcome compared to those that were not KIR3DL1+/Bw4+ (p < 0.05 for TTRF, Fig. 2a; p = 0.01 for duration of response, Fig. 2b; and a trend of p = 0.07 for tumor shrinkage, Fig. 2c).

Inhibitory KIR2DL2/C1+ interactions in combination with KIR3DL1/KIR-ligand interactions improve outcome for patients receiving rituximab maintenance treatment

While the KIR-ligands present genotype (as shown in Fig. 1) considers all three KIR-ligands (HLA-C1, HLA-C2 and HLA-Bw4) equally contributing to the licensing or inhibition of NK cells, (described in
Additional file 1: Table S2), we assessed whether certain combinations of KIR2DL1, KIR2DL2 or KIR2DL3 and their KIR-ligands together with KIR3DL1 and its KIR-ligand would affect patients' response to rituximab differently. We and others have found that a subset of patients, based on the presence of KIR2DL1, KIR2DL2 or KIR2DL3 and their respective ligands together with the presence of KIR3DL1 and its ligand, benefited from anti-GD2-based treatment while the converse genotype did not [38, 39]. As mentioned above, HLA-C1 and HLA-C2 are alleles, and thus everyone will have either HLA-C1 and/or HLA-C2. With this in mind, since every individual will have an interaction of inhibitory KIR2DL1, KIR2DL2 or KIR2DL3 with their HLA-C ligands, we assessed the effects of the presence or absence of KIR3DL1 and its ligand on outcome in combination with the various possible combinations of KIR2DL1, KIR2DL2 or KIR2DL3 with their ligands.

The following dual combinations of inhibitory KIRs were compared:

KIR2DL1+/C2+/KIR3DL1+/Bw4+ vs. not KIR2DL1+/C2+/KIR3DL1+/Bw4+; KIR2DL2+/C1+/KIR3DL1+/Bw4+ vs. not KIR2DL2+/C1+/KIR3DL1+/Bw4+ and KIR2DL3+/C1+/KIR3DL1+/Bw4+ vs. not KIR2DL3+/C1+/KIR3DL1+/Bw4. These double-inhibitory KIR/KIR-ligand genotype combinations are described in further detail in the text.
detail in Additional file 1: Table S5. For these “double” combinations, the only interaction comparison that met our predefined statistical cutoff of a p-value ≤ 0.100 was the comparison of KIR2DL2+/C1+/KIR3DL1+/Bw4+ vs. not KIR2DL2+/C1+/KIR3DL1+/Bw4+, herein referred to as “Group 1” vs. “Group 2”, respectively (as shown in Table 2, lines 5–9, p = 0.068).

Subgroup comparisons were done for Group 1 vs. Group 2 treated with maintenance vs. non-maintenance rituximab (Fig. 3). Group 1 patients treated with maintenance vs. non-maintenance showed a trend toward improved TTRF (p = 0.10, Fig. 3a), improved duration of response (p < 0.001, Fig. 3b), and a trend toward improved % tumor shrinkage (p = 0.08, Fig. 3c). Conversely, those in Group 2 did not even have a trend toward improved clinical outcome for any of these 3 parameters if treated with maintenance vs. non-maintenance rituximab (Fig. 3a, b and c). In addition, amongst patients that were treated with maintenance rituximab, Group 1 had improved clinical outcome as compared to those in Group 2 (p = 0.04 for TTRF, Fig. 3a; p = 0.006 for duration of response, Fig. 3b; p = 0.08 for tumor shrinkage, Fig. 3c).

**Discussion**

In this study, we found no influence of the KIR-ligands present vs. KIR-ligand missing genotypes on clinical outcome. However, we did find that for some FL patients, treatment with maintenance rituximab improved their clinical outcome based on their KIR/KIR-ligand genotype. Namely, the 28% (45 of 159) of this patient population with genotypes in Group 1 (Fig. 3a) appear to have improved outcome (significant for duration of response and tumor shrinkage and a trend for TTRF) from maintenance vs. non-maintenance rituximab. In contrast, for those remaining 72% of patients with the Group 2 genotype, we found insufficient evidence of an improvement in outcome if treated with maintenance compared to the non-maintenance rituximab treatment schedule [18, 20]. We recently found a similar result for these same patients treated for relapsed FL, and also found that the persistent tumor was more likely to have missing KIR-ligands (p = 0.018, Fig. 3a).
genotypes in a separate randomized study of anti-GD2-based mAb immunotherapy for children with high-risk neuroblastoma. Namely, neuroblastoma patients with the Group 1 genotype appear to have improved outcome from anti-GD2-based immunotherapy (vs. no immunotherapy), while for those patients with the Group 2 genotype, no improvement in outcome from the anti-GD2-based immunotherapy (vs. no immunotherapy) was observed [38]. These similar results regarding improved outcome with the immunotherapy regimens for patients with the Group 1 genotype in these two separate randomized studies, which include two different patient populations (adult vs. pediatric patients), two different disease types (FL vs. neuroblastoma), and two different anti-tumor-reactive mAb-based immunotherapies (rituximab/anti-CD20 vs. dinutuximab/anti-GD2), provides some degree of validation for this finding. In addition to the Group 1 genotype, we recently reported similar findings for patients with certain HLA-Bw4 isoforms from these same two clinical studies [40]. In that report, we evaluated the role of three separate HLA-Bw4 isoforms (HLA-A-Bw4, HLA-B-Bw4-I80 and HLA-B-Bw4-T80). In both clinical trials, patients with KIR3DL1+/A-Bw4+ or with KIR3DL1+/B-Bw4-T80 + genotypes randomized to neuroblastoma immunotherapy or FL maintenance immunotherapy had better outcome versus those randomized to no immunotherapy for neuroblastoma or non-maintenance for FL. In contrast, for those with KIR3DL1+/B-Bw4-I80+, there was no evidence of a difference in outcome between immunotherapy vs. no-immunotherapy [40]. These combined results suggest that the correlation between these KIR/KIR-ligand genotypes and outcome may serve as a biomarker for identifying those that might benefit from this type of immunotherapy using tumor-reactive mAb. Furthermore, these results suggest similar analyses of KIR/KIR-ligand genotype and immunotherapy should be pursued for other clinical trials in other diseases that utilize other tumor-reactive mAbs to see if these findings we have noted for rituximab in FL and dinutuximab in neuroblastoma might extend to other cancers treated with other tumor-reactive mAb [40].

In addition, in this study, amongst patients that received rituximab maintenance, those that are KIR3DL1+/Bw4+ showed significantly improved TTRF, duration of response and tumor shrinkage than for those not KIR3DL1+/Bw4+. Similarly, amongst patients that received rituximab maintenance, those that are Group-1 (KIR2DL2+/C2+ and KIR3DL1+/Bw4+) showed improved TTRF, duration of response and tumor shrinkage than those that are Group-2 (not KIR2DL2+/C2+ and KIR3DL1+/Bw4+). These demonstrate an association (likely via ADCC) of KIR/KIR-ligand genotype on the outcome of rituximab maintenance therapy.

Several studies have found associations of patient response to treatment with KIR2DL2 status, with or without its ligand, HLA-C1 [41–43]. In a study of neuroblastoma patients mentioned previously, we found that patients that had both KIR2DL2+/C1+ [KIR2DL2+ and HLA-C1+ (C1/C1 or C1/C2)] and KIR3DL1+/Bw4+ (i.e. Group 1) had improved clinical outcome if treated with monoclonal antibody-based immunotherapy compared to no-immunotherapy [38].

In addition to assessing the influence of HLA-Bw4 on clinical outcome, Du and colleagues found that FL patients that were missing both HLA-C2 and HLA-Bw4 and were treated with either rituximab and galaximib, or rituximab and epratuzumab, had longer duration of response than those individuals that possessed both of those KIR-ligands [12]. That analysis focused on the subsets of patients with both HLA-C2 and HLA-Bw4 or missing both HLA-C2 or HLA-Bw4, but excluded the subsets with only one or the other of these combinations. In our analysis here of FL patients treated with maintenance or non-maintenance rituximab, we included all patients in each comparative analysis, and also considered the inhibitory KIR gene status, when evaluating KIR2DL1, KIR2DL2 or KIR2DL3, with their respective ligands with KIR3DL1 and its HLA-Bw4 ligand. With respect to HLA-C status and HLA-Bw4 status, following assessment of all possible KIR2DL1, KIR2DL2 or KIR2DL3, with their respective ligands (HLA-C1 or HLA-C2) and with KIR3DL1 and its ligand (HLA-Bw4), our analysis suggests patients that are both KIR2DL2+/C1+ and KIR3DL1+/Bw4+ (Group 1) had improved clinical outcome if treated with rituximab maintenance compared with the non-maintenance treatment schedule. However, as the subpopulations of patients with varying KIRs and KIR-ligands studied in each report are different, and the exact therapy we used (rituximab maintenance) was different than that used by Du et al. (rituximab and galaximib, or rituximab and epratuzumab), we cannot actually determine whether our results are discordant from those of Du et al.

Kahl and colleagues used TTRF as their primary endpoint in the primary clinical report of this trial, as it may provide more value to understanding of the clinical benefit of rituximab than using duration of response as an endpoint [6, 8]. Rituximab has limited side effects compared with cytotoxic chemotherapeutic agents, which have more adverse side effects [8], and the TTRF can reflect when progressive or unresponsive disease requires initiation of cytotoxic chemotherapy. If a maintenance schedule can, for some patients, delay the need for cytotoxic therapy, this could provide clinical benefit [44, 45]. In our analyses of the TTRF parameter shown in this report, we considered only those failure events that were considered to be biologically relevant. In the
primary study report, Kahl et al. noted no difference in time to rituximab failure between the maintenance and non-maintenance schedules when all patients were evaluated independent of genotype [6]. However, in this report, by assessing different genotype groupings of KIRs/KIR-ligands, we were able to identify a set of patients (Group 1: KIR2DL2+/C1+/KIR3DL1+/Bw4+), reflecting 28% of this patient population, that may have prolonged TTRF if treated with maintenance rituximab as compared to non-maintenance. This finding provides some evidence that rituximab maintenance may still provide a clinically meaningful benefit for a subgroup of patients based on their KIR/KIR-ligand genotypes.

Most NK cells express an array of both inhibitory and activating receptors; NK cell activation reflects the balance of both activating and inhibitory signals. The clinical data and associations presented in this report suggest that there are in vivo interactions that were simultaneously influenced by inhibitory KIRs and their ligands, and activating signaling through stimulation with mAb (rituximab), which appeared to also reflect influences on NK cell licensing [35]. While this report focuses on the associations of inhibitory-KIR/KIR-ligand genotype with clinical outcome for this clinical trial, patients from this same clinical trial were also evaluated for other KIR-related genotype associations. We also evaluated associations of KIR haplotype (A or B) with clinical outcome, but found no associations with clinical outcome (data not shown) [24]. In addition, for this same ECOG-ACRIN study, Kenkra et al. assessed genotypes for high and low affinity allelic variants of the activating Fc gamma receptors (FCGR), FCGR3A and FCGR2A, and found no associations with clinical outcome [46].

Conclusions
In summary, the associations of outcome and KIR/KIR-ligand genotypes presented here demonstrate some role for KIR recognition of KIR-ligands in the in vivo response to rituximab therapy in at least some patients with FL. In particular, the 28% of patients in this trial that have the Group 1 genotype had an association of clinical outcome benefit with administration of the rituximab maintenance treatment schedule. Whether the statistically significant benefit in outcome for this subset shown here merits prospective testing (via genotyping of all patients to identify this group for further analyses of maintenance treatment) requires consideration of medical as well as cost-related issues. Furthermore, as the associations observed here are based on relatively small numbers of patients in the individual comparative groups, separate validation may be helpful before considering whether this KIR/KIR-ligand genotyping should be used prospectively for treatment assignment decisions. In addition, based on the findings described here, as well as our similar findings using other monoclonal antibodies [30], it may be of interest to study the impact of KIR and KIR-ligands with the newer anti-CD20 antibodies, obinutuzumab and ofatumumab, which have shown efficacy in treatment of indolent non-Hodgkin lymphomas [47, 48]. Finally, the associations reported here likely reflect the regulation of NK cells mediating in vivo ADCC as a result of the rituximab therapy, consistent with other preclinical and clinical data. Thus these data further support the roles that KIR and KIR-ligands play, for at least some patients, in the setting of certain types of cancer immunotherapy.

Additional file

Additional file 1: Table S1. Description of KIR/KIR-ligand genotypes included in analyses for Fig. 1. Table S2. Description of KIR/KIR-ligand genotypes included in analyses for Table 1 and Fig. 2. Table S3. Detailed statistical information for data pertaining to Figs. 1 and 2. Table S4. Detailed statistical information for data pertaining to Figs. 1 and 3. Table S5. Demographic and clinical characteristics of ECOG patients with FL that were randomly assigned and genotyped. (DOCX 531 kb)

Abbreviations
ECOG-ACRIN: ECOG-ACRIN Cancer Research Group; FCGR: Fc gamma receptors; FL: Follicular lymphoma; Group 1: KIR2DL2+/C1+/KIR3DL1+/Bw4+; Group 2: not KIR2DL2+/C1+/KIR3DL1+/Bw4+; KIR: Killer immunoglobulin-like receptors; Maintenance: Rituximab every 13 weeks; NK Cells: Natural Killer cells; Non-Maintenance: No additional rituximab until progression; TTRF: Time to Rituximab Failure

Acknowledgements
The authors thank the medical and nursing staff that participated in the care of patients in this study, and especially all patients who participated in this study. Thanks also to Dr. David Evans for helpful discussions and editing, and to Robert Gordon for his assistance in figure editing.

Funding
This research was supported by Hyundai Hope on Wheels Grant; Midwest Athletes Against Childhood Cancer; Stand Up 2 Cancer; The St. Baldrick’s Foundation; American Association for Cancer Research; University of Wisconsin-Madison Carbone Cancer Center; Forward Lymphoma Fund; ASH Trainee Research Award; by the ECOG-ACRIN Cancer Research Group (Peter J. O’Dwyer, MD and Mitchell D. Schnall, MD, PhD, Group Co-Chairs) and supported in part by Public Health Service Grants CA180820, CA180794, CA180799, CA180816, CA180859, CA180833 and CA197078, from the National Cancer Institute; the National Institutes of Health and the Department of Health and Human Services. Its content is solely the responsibility of the authors and does not necessarily represent the official views of the National Cancer Institute.

Availability of data and materials
The datasets used and/or analysed during the current study are available from the corresponding author upon reasonable request.

Authors’ contributions
AK and WW genotyped the patients, interpreted the patient data regarding the clinical associations with outcome, and were major contributors to the writing of the manuscript. LC and KK performed statistical analysis and inference of clinical outcome in association with genotype status and were major contributors to the writing of the manuscript. BG participated in the design of the study; AH, PKR and JAH assisted in interpretation of the patient data regarding the clinical associations with outcome and were contributors in writing of the manuscript. EAM and YS created the database for genotype
entry and were contributors in writing of the manuscript. EAR served as the clinical pathologist for the study and evaluated patient specimens and was a contributor to the writing of the manuscript. SS performed statistical associations of clinical outcome with genotype status and was a contributor in writing of the manuscript. VPK, RDG, EP and SJH were involved in clinical data collection and were contributors in writing of the manuscript. ISM was a major contributor to interpretation of the data regarding the clinical associations with outcome, and was contributor to the writing of the manuscript. FH led the statistical analyses of the clinical data set, interpreted the patient data regarding the clinical associations with outcome, and was a contributor to the writing of the manuscript. BK led the clinical data portion of this data set, interpreted the patient data regarding the clinical associations with outcome, and was a major contributor to the writing of the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate
As approved in the initial clinical report (6), all patients signed approved consent forms. These associative analyses reported in this manuscript were deemed by our institutional review board as exempt from human subjects research.

Consent for publication
Not Applicable.

Competing interests
Dr. Brad Kahl discloses consulting and research funding from Roche/Genentech. Dr. Randy Gascoyne discloses consulting for Roche/Genentech. All other authors declare no competing financial interests.

Publisher’s Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Author details
1Department of Human Oncology, University of Wisconsin, Madison, WI, USA. 2Department of Biostatistics and Medical Informatics, University of Wisconsin, Madison, Madison, WI, USA. 3Department of Laboratory Medicine and Pathology, University of Minnesota, Minneapolis, MN, USA. 4Department of Medicine, University of Wisconsin, Madison, WI, USA. 5Department of Biostatistics, Harvard University, Dana Farber Cancer Institute, Boston, MA, USA. 6Department of Pathology and Laboratory Medicine, Centre for Lymphoid Cancer, British Columbia Cancer Agency, Vancouver, British Columbia, Canada. 7Montefiore Medical Center-North Division, Albert Einstein College of Medicine, Bronx, NY, USA. 8Department of Medicine, Stanford University, Stanford, CA, USA. 9Department of Medicine, University of Minnesota, Minneapolis, MN, USA. 10Department of Medicine, Washington University School of Medicine, St. Louis, MO, USA. 11Department of Pediatrics, University of Wisconsin, Madison, WI, USA. 12University of Wisconsin-Madison, 1111 Highland Avenue, 4159 WIMR Bldg, Madison, WI 53705, USA.

Received: 7 November 2018 Accepted: 20 February 2019
Published online: 12 March 2019

References
1. Izutsu K. Treatment of follicular lymphoma. J Clin Exp Hematop. 2014;54(1):31–7.
2. Kahl BS, Yang DT. Follicular lymphoma: evolving therapeutic strategies. Blood. 2016;127(17):2055–63.
3. Ghielmini M, et al. Prolonged treatment with rituximab in patients with follicular lymphoma significantly increases event-free survival and response duration compared with the standard weekly x 4 schedule. Blood. 2004;103(12):4416–23.
4. Hainsworth JD, et al. Rituximab as first-line and maintenance therapy for patients with indolent non-Hodgkin’s lymphoma. J Clin Oncol. 2002;20(4):2461–7.
5. Martielli G, et al. Long-term follow-up of patients with follicular lymphoma receiving single-agent rituximab at two different schedules in trial SAKK 35/98. J Clin Oncol. 2010;28(29):4480–4.
6. Kahl BS, et al. Rituximab extended schedule or re-treatment trial for low-tumor burden follicular lymphoma: eastern cooperative oncology group protocol e4402. J Clin Oncol. 2014;32(28):3096–102.
7. Hornberger J, et al. Cost-effectiveness of rituximab as maintenance therapy in patients with follicular non-Hodgkin lymphoma after responding to first-line rituximab plus chemotherapy. Leuk Lymphoma. 2012;53(12):2371–7.
8. Barr PM. Rituximab maintenance in follicular lymphoma. Clin Adv Hematol Oncol. 2015;13(3):158–61. 26352422.
9. Jacobson CA, Freedman AS. Is observation dead in follicular lymphoma? Still appropriate. J Natl Compr Cancer Netw. 2015;13(3):367–70.
10. He L, et al. Low natural killer (NK) cell counts in peripheral blood adversely affect clinical outcome of patients with follicular lymphoma. Blood Cancer J. 2016;6(8):e457.
11. Carton G, et al. Therapeutic activity of humanized anti-CD20 monoclonal antibody and polymorphism in IgG fc receptor FcgammaRlla gene. Blood. 2002;99(3):754–8.
12. Du J, et al. CALGB 150905 (Alliance): rituximab broadens the anti-lymphoma response by activating unlicensed NK cells. Cancer Immunol Res. 2014;2(9):878–89.
13. Weng WK, Levy R. Two immunoglobulin G fragment C receptor polymorphisms independently predict response to rituximab in patients with follicular lymphoma. J Clin Oncol. 2003;21(21):3940–7.
14. Herberman RB, Reynolds CW, Oortado JR. Mechanism of cytotoxicity by natural killer (NK) cells. Annu Rev Immunol. 1986;4:651–80.
15. Lanier LL. NK Cell Recognition. Annu Rev Immunol. 2005;23(1):225–74.
16. Boyington JC, Sun PD. A structural perspective on MH-C class I recognition by killer cell immunoglobulin-like receptors. Mol Immunol. 2002;38(14):1007–21.
17. Vilches C, Parham P. KIR: diverse, rapidly evolving receptors of innate and adaptive immunity. Annu Rev Immunol. 2002;20:217–51.
18. Delgado DC, et al. Genotypes of NK cell KIR receptors, their ligands, and fc receptors in the response of neuroblastoma patients to Hui14.18-112 immunotherapy. Cancer Res. 2010;70(23):9554–61.
19. Moesta AK, et al. Synergistic polymorphism at two positions distal to the ligand-binding site makes KIR2DL2 a stronger receptor for HLA-C than KIR2DL3. J Immunol. 2008;180(6):3969–79.
20. Tarek N, et al. Unlicensed NK cells target neuroblastoma following anti-CD2 antibody treatment. J Clin Invest. 2012;122(9):3626–70.
21. Wilson MJ, et al. Plasticity in the organization and sequences of human KIR/LILT gene families. Proc Natl Acad Sci. 2000;97(9):4778–83.
22. Kim S, et al. Licensing of natural killer cells by host major histocompatibility complex I molecules. Nature. 2005;436(7051):709–13.
23. Joncker NT, et al. NK cell responsiveness is tuned commensurate with the number of inhibitory receptors for self-MHC class I: the rheostat model. J Immunol. 2009;182(8):4572–80.
24. Cooley S, et al. Donor selection for natural killer cell receptor genes leads to superior survival after unrelated transplantation for acute myelogenous leukemia. Blood. 2010;116(14):2411–9.
25. Cooley S, et al. Donor killer cell Ig-like receptor B haplotypes, recipient HLA-C, and HLA-C mismatch enhance the clinical benefit of unrelated transplantation for acute myelogenous leukemia. J Immunol. 2014;192(10):4592–600.
26. Venstrom JM, et al. HLA-C–dependent prevention of leukemia relapse by donor ActivatingKIRDS1. N Engl J Med. 2012;367(9):805–16.
27. Leung W, et al. Inhibitory KIR–HLA–receptor–ligand mismatch in autologous haematopoietic stem cell transplantation for solid tumour and lymphoma. Br J Cancer. 2007;97(4):539–42.
28. Ruggeri L, et al. Donor natural killer cell allorrecognition of missing self in haploidentical hematopoietic transplantation for acute myeloid leukemia: challenging its predictive value. Blood. 2007;110(1):433–40.
29. Cheung NK, et al. Murine anti-CD2 monoclonal antibody 3F8 combined with granulocyte-macrophage colony-stimulating factor and 13-cis-retinoic acid in high-risk patients with stage 4 neuroblastoma in first remission. J Clin Oncol. 2012;30(26):3264–70.
30. Erbe AK, et al. Neuroblastoma Patients’ KIR and KIR-ligand genotypes influence clinical outcome for Dinutuximab-based immunotherapy: a report from the Children’s oncology group. Clin Cancer Res. 2018;24(1):189–96.
31. Alves LGT, Rajalingam R, Canavez F. A novel real-time PCR method for KIR genotyping. Tissue Antigens. 2009;73(2):188–91.
32. Vilches C, et al. Facilitation of KIR genotyping by a PCR-SSP method that amplifies short DNA fragments. Tissue Antigens. 2007;70(5):415–22.
33. Harris PA, et al. Research electronic data capture (REDCap)—a metadata-driven methodology and workflow process for providing translational research informatics support. J Biomed Inform. 2009;42(2):377–81.

34. Jonsson AH, Yokoyama WM. Natural killer cell tolerance licensing and other mechanisms. Adv Immunol. 2009;101:27–79.

35. Yokoyama WM, Kim S. Licensing of natural killer cells by self-major histocompatibility complex class I. Immunol Rev. 2006;214:143–54.

36. Almeida CR, et al. Human NK cells differ more in their KIR2DL1-dependent thresholds for HLA-Cw6-mediated inhibition than in their maximal killing capacity. PLoS One. 2011;6(9):e24927.

37. Gumperz, J.E., et al., Conserved and variable residues within the Bw4 motif of HLA-B make separable contributions to recognition by the NK11 killer cell-inhibitory receptor. J Immunol, 1997. 158(11): p. 5237–5241. PMID: 9164941.

38. Erbe A, et al. Impact of KIR/KIR ligand genotype for neuroblastoma patients in a Phase 3 COG Immunotherapy Trial, in AACR Annual Meeting (abstract). Washington, D.C. Cancer Research; 2017.

39. Siebert N, et al. Neuroblastoma patients with high-affinity FCGR3A, −3A and stimulatory KIR 2DS2 treated by long-term infusion of anti-GD2 antibody ch14.18/CHO show higher ADCC levels and improved event-free survival. Oncol Immunol. 2016;5(11):e1235108.

40. Erbe AK, et al. HLA-Bw4-I-80 isoform differentially influences clinical outcome as compared to HLA-Bw4-T-80 and HLA-A-Bw4 isoforms in rituximab or Dinutuximab-based Cancer immunotherapy. Front Immunol. 2017;8:675.

41. Middleton D, et al. Killer immunoglobulin-like receptors (KIR2DL2 and/or KIR2DS2) in presence of their ligand (HLA-C1 group) protect against chronic myeloid leukaemia. Tissue Antigens. 2009;73(6):553–60.

42. Wisniewski A, et al. KIR2DL2/S2 and HLA-C CYC1 genotype is associated with better response to treatment and prolonged survival of patients with non-small cell lung cancer in a polish Caucasian population. Hum Immunol. 2012;73(9):927–31.

43. Velarde-de la Cruz EE, et al. KIR2DL2 and KIR2DS2 as genetic markers to the methotrexate response in rheumatoid arthritis patients. Immunopharmacol Immunotoxicol. 2016;38(4):303–9.

44. Webster K, Cella D. Quality of life in patients with low-grade non-Hodgkin’s lymphoma. Oncology (Williston Park). 1998;12(5):697–714 discussion 714, 717, 721.

45. Arcaini L, Merli M. Rituximab maintenance in follicular lymphoma patients. World J Clin Oncol. 2011;2(7):281–8.

46. Kenkre VP, et al. Fc gamma receptor 3A and 2A polymorphisms do not predict response to rituximab in follicular lymphoma. Clin Cancer Res. 2016; 22(4):821–6.

47. Marcus R, et al. Obinutuzumab for the first-line treatment of follicular lymphoma. N Engl J Med. 2017;377(14):1331–44.

48. Hillmen P, et al. Chlorambucil plus ofatumumab versus chlorambucil alone in previously untreated patients with chronic lymphocytic leukaemia (COMPLEMENT 1): a randomised, multicentre, open-label phase 3 trial. Lancet. 2015;385(9980):1873–83.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions