Polymorphisms of the cytomegalovirus glycoprotein B genotype in patients with Posner-Schlossman syndrome

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ABSTRACT

Aims The aim of this observational study was to report the distribution of glycoprotein B (gB) genotypes in the eyes of cytomegalovirus (CMV) positive patients with Posner-Schlossman syndrome (PSS), and to investigate their clinical characteristics and outcomes.

Methods We collected aqueous humour samples from 165 patients clinically diagnosed with PSS between 2017 and 2019. PCR was performed to analyse the CMV DNA and identify the gB genotypes in the samples. Clinical characteristics and responses to antiviral treatment were compared among patients with different gB genotypes.

Results CMV DNA was detected in 94 (56.97%) of the 165 aqueous humour specimens analysed. Owing to the quantity requirement for CMV gB genotype analysis, results could be obtained from only 14 specimens. CMV gB type 1 was detected in 11 samples (78.6%), whereas CMV gB type 3 was detected in three samples (21.4%). No other gB genotypes or mixed genotypes were detected. Overall, 9.1% (1/11) of the patients in the gB type 1 group and 66.7% (2/3) of the patients in the gB type 3 group had bilateral attacks (p=0.093). The concentration of anti-CMV immunoglobulin G (IgG) in the type 1 group was 0.94±0.79 s/co (ratio of aqueous humour CMV IgG/serum CMV IgG to aqueous humour albumin concentration/serum albumin concentration), whereas that in the type 3 group was 0.67±0.71 s/co.

Conclusion Genotype 1 was the most prevalent genotype in the aqueous humour of CMV-infected patients with PSS. Bilateral attack was predominant among patients with gB genotype 3. CMV gB gene may be related to the pathogenicity of CMV virus strain in patients with PSS.

INTRODUCTION

The Posner-Schlossman syndrome (PSS) is clinically characterised by recurring unilateral uveitis and elevated intraocular pressure (IOP).1 A retrospective study reported that the annual incidence of PSS is about 3.91 per 100 000 persons and is relatively high in China.2 Unlike glaucoma, the IOP of patients with PSS is usually within the normal range during the non-onset period; however, PSS is prone to relapse accompanied by inflammatory responses in the anterior segment.3 Although administration of corticosteroids is a traditional method of relieving inflammation, its side effects include more frequent relapses and the risk of developing corticosteroid dependence, which may cause steroid-induced cataract and steroid-induced glaucoma over time.4 Eventually, repeated elevated IOP can damage the optic nerve and induce severe visual dysfunction. To facilitate effective treatment of PSS, its aetiology and pathophysiological mechanism need to be elucidated. The aetiology of PSS has been examined in some studies, which indicated that PSS may be associated with cytomegalovirus (CMV) infection,5–7 gene susceptibility,8 inflammatory cytokines,9 10 and vascular endothelial dysfunction.11 Among these, CMV infection is considered a vital risk factor for PSS.5–7 12–17 Anti-CMV therapy has been used to treat patients with CMV-positive PSS.7 18 However, we found that in clinical practice, not all patients receive satisfactory curative effect after ganciclovir treatment, and their clinical manifestations are not always the same. The underlying causes of these irregularities were worth exploring. CMV shows wide genetic diversity. Envelope glycoproteins play an important role in host immune response and virus replication.19 Presumably, the variability of the genes encoding these proteins contributes to the virulence of the strain. CMV glycoprotein B (gB), encoded by the UL53 gene, is the major envelope glycoprotein of CMV. It is considered to play a vital role in viral entry, viral spreading between cells, and fusion of infected cells.20 CMV genotyping based on the gB nucleotide sequence has been performed to analyse infections caused by CMV. There were four main specific sequence variations of gB, named gB genotype 1, gB genotype 2, gB genotype 3 and gB genotype 4. Genotyping of these CMV genes determines the characteristics of CMV in each disease. However, due to the rarity of PSS and the relatively low viral load of CMV in aqueous humour, little is known about the genotypic composition of CMV in patients with PSS. Differences in the genotypes of patients with PSS may account for differences in characteristics. In this study, we determined the frequency distribution of the CMV gB genotype in the aqueous humour of patients with PSS and investigated the differences in the clinical characteristics of the patients according to their different CMV genotypes.
the tenets of the Declaration of Helsinki, revised in 2000. All participants provided written informed consent. The study was conducted at the EENT Hospital of Fudan University from July 2017 to May 2019.

Individuals diagnosed as PSS were enrolled. The diagnostic criteria for PSS were based on the following clinical manifestations during an attack: (1) recurrent mild inflammation in the anterior chamber; (2) characteristic keratic precipitates (KPs) and corneal oedema; (3) a history of transient elevated IOP; (4) no peripheral anterior or posterior synechiae; (5) no posterior inflammation and (6) open anterior angle. Exclusion criteria included previous antiviral treatment, intraocular surgery or penetrating ocular injury. General information on each patient, including sex, age and medical history, was collected. Glucocorticoid dependence was defined as the need for continuous use of glucocorticoids, and the aggravation or relapse of PSS-related inflammation when the dosage of the medication was reduced or the treatment was discontinued. Clinical data, including peak IOP, visual acuity, corneal endothelial cell density, KPs, Tyndall effect and vertical cup-disc ratio, were recorded. Relative corneal endothelial loss compared with the other eye was defined as the ratio of the number of corneal endothelial cells in the affected eye to that in the contralateral eye. Glaucomatous optic neuropathy is determined if any of these conditions are satisfied: (1) vertical cup-disc ratio ≥0.7, (2) retinal nerve fibre layer defects correspond with thinning width of rim or localised notches and (3) splinter haemorrhages. Using the Lens Opacities Classification System III, cataract was defined as meeting any of the following conditions: (1) nuclear opalescence ≥3.0, (2) cortical cataract ≥3.0 and (3) posterior subcapsular cataract ≥2.0. When the patient is examined with a slit lamp microscope, the abnormal enhancement of the light beam through the aqueous humour to greyish white is considered a positive Tyndall sign. After topical anaesthesia was administered, a 27-gauge needle was used to extract 100–150 µL of aqueous humour from the eyes of the patients under the magnified view of a slit-lamp biomicroscope, and the samples were immediately stored at −80°C. All patients with PSS were in the acute onset stage and received antiviral treatment (2% ganciclovir eye drops were positive for CMV DNA (56.97%) and gb was detected in 14 specimens, which were tested further to ascertain the distribution of the virus subtypes.

**CMV DNA sequence analysis**

Viral DNA was obtained from the sample with the QIAMP viral DNA extraction kit (Qiagen, Hilden, Germany), according to the instructions. DNA was eluted in the 100 µL buffer provided in the kit and stored at −40°C. Real-time PCR (RT-PCR) was used to detect CMV. The genes were amplified using the RT-PCR Kit (TaKaRa, Japan) and sequences were determined by sequencing the products. DNA Dynamo Sequence Analysis Software (Blue Tractor Software, Llanfairfechan, UK) and the Clustal W algorithm were used for sequence alignment and phylogenetic analysis. The clustering method used was the “unweighted pair group method with arithmetic mean”. Based on the results of other studies, the genotype pattern strains were categorised under UL55 (gb).

**Analysis of the concentration of antiviral immunoglobulin G in aqueous humour**

As reported in our previous study, 36 antiviral immunoglobulin (Ig) G in the aqueous humour and serum was detected using an ELISA kit (Virion/Serion, Germany). Scattering immunonephelometry was performed to examine albumin (Guosai Biotechnology Co, China). The level of CMV IgG in the aqueous humour was presented as a corrected ratio of aqueous humour CMV IgG/serum CMV IgG to aqueous humour albumin concentration/serum albumin concentration, abbreviated as s/co.

**RESULTS**

**Viral analysis of the aqueous humour samples of patients with PSS**

As shown in figure 1, a total of 165 aqueous humour specimens collected from patients with PSS were tested using RT-PCR analyses; 94 of the specimens were positive for CMV DNA (56.97%) and gb was detected in 14 specimens, which were tested further to ascertain the distribution of the virus subtypes.

**Distribution of the CMV gb genotype**

Phylogenetic analysis of the gb gene sequence was performed on 14 gb-positive samples according to the genotype sequence of the gb proteolytic site. CMV gb1 was detected in 11 specimens (78.6%) and CMV gb3 in three specimens (21.4%). No mixed infection with the gb1 and gb3 genotypes was detected. The concentration of anti-CMV IgG in the type 1 group and type 3 group was 0.94±0.79 s/co and 0.67±0.71 s/co, respectively.

**Demographics and clinical characteristics of patients in the gb-positive group**

There were more men among the 14 patients in the gb-positive group (85.7%, 12/14), as well as the gb1 genotype (90.9%, 10/11) and gb genotype 3 (66.7%, 2/3) groups, than women. As presented in table 1, patients infected with CMV gb1 seemed to be younger at the time of the clinic visit (41.55±11.25 vs 52.33±14.19) and at the time of the first onset of disease (37±12 vs 47±17) than patients infected with CMV gb3; the
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former also seemed to have a shorter course of disease than the latter (4.7±2.8 vs 5.7±4.5). In the genotype 1 group, only 9.1% (1/11) of the patients had bilateral attacks, whereas 66.7% (2/3) of the patients in the genotype 3 group had bilateral attacks (p=0.093). The frequencies of onset in the genotype 1 group and genotype 3 group were 1.5±0.8 per year and 1.7±0.6 per year, respectively. Patients in the genotype 1 group seemed to have a higher dependence on glucocorticoids (63.6%, 7/11) than patients in the genotype 3 group (33.3%, 1/3).

The clinical characteristics of the patients according to their gB genotype distributions are shown in table 2. There was no visible difference in visual acuity (logarithm of minimal angle resolution (LogMAR)), cup–disk ratio, peak IOP or corneal endothelial cell loss (%) between groups. The rates of occurrence of the Tyndall effect in the anterior chamber (3/11, 27.3% vs 2/3, 66.7%) and iris depigmentation (8/11, 72.3% vs 3/3, 100%) were higher in the CMV type 3 group than in the CMV type 1 group. Coin-shaped KPs (shown in figure 2) were observed in 100% of

### Table 1  
Details and characteristics of CMV-infected patients with PSS

| Case | Sex | Unilateral/Bilateral | Age at the clinic visit (years) | Age at the first onset (years) | Frequency of attack (n/year) | Peak IOP | GC dependence | Anti-HCMV IgG (s/co) | CMV (copies/mL) | Response to antiviral therapy | CMV genotype |
|------|-----|----------------------|---------------------------------|-------------------------------|-----------------------------|----------|----------------|-------------------|----------------|-----------------------------|--------------|
| 1    | Male| Uni/OS               | 54                              | 51                            | 46                          | 1.5      | 2213           | Complete control   | gB 1            |                             |              |
| 2    | Male| Uni/OO               | 32                              | 29                            | 45                          | 0.59     | 15,311         | Poor control       | gB 1            |                             |              |
| 3    | Male| Uni/OO               | 50                              | 49                            | 50                          | 1.06     | 10,965         | Poor control       | gB 1            |                             |              |
| 4    | Male| Uni/OO               | 42                              | 38                            | 50                          | 2.39     | 28,510         | Poor control       | gB 1            |                             |              |
| 5    | Male| Uni/OO               | 31                              | 25                            | 50                          | 0.34     | 473,151        | Complete control   | gB 1            |                             |              |
| 6    | Male| Bi/OOS               | 62                              | 56                            | 50                          | 0.76     | 2,585,235      | Complete control   | gB 1            |                             |              |
| 7    | Male| Uni/OO               | 51                              | 46                            | 60                          | 0.27     | 181,970        | Complete control   | gB 1            |                             |              |
| 8    | Female| Uni/OO               | 28                              | 25                            | 46                          | 0.28     | 142,889        | Complete control   | gB 1            |                             |              |
| 9    | Male| Uni/OO               | 40                              | 30                            | 41                          | 0.67     | 346,737        | Poor control       | gB 1            |                             |              |
| 10   | Male| Uni/OO               | 31                              | 22                            | 48                          | 2.29     | 307,256        | Poor control       | gB 1            |                             |              |
| 11   | Male| Uni/OO               | 36                              | 34                            | 43                          | 0.24     | 7,852,356      | Poor control       | gB 1            |                             |              |
| 12   | Female| Bi/OOD              | 37                              | 31                            | 45                          | 0.27     | 407,280        | Complete control   | gB 3            |                             |              |
| 13   | Male| Uni/OO               | 55                              | 45                            | 52                          | 0.24     | 301,995        | Poor control       | gB 3            |                             |              |
| 14   | Male| Bi/OOS               | 65                              | 64                            | 48                          | 1.49     | 38,238         | Poor control       | gB 3            |                             |              |

Bi, bilateral; CMV, cytomegalovirus; gB, glycoprotein B; GC, glucocorticoid; HCMV, human cytomegalovirus; IOP, intraocular pressure; OD, right eye; OS, left eye; PSS, Posner-Schlossman syndrome; Uni, unilateral.

### Table 2  
Comparison of the clinical manifestations of patients in various gB genotype groups

|                  | CMV gB genotype 1 | CMV gB genotype 3 |
|------------------|-------------------|-------------------|
|                  | Mean±SD | Range, median | Mean±SD | Range, median | P value |
| N (%)            | 11, 78.6% |          | 3, 21.4% |            |         |
| Male (n, %)      | 10, 90.9% |          | 2, 66.7% |          | 0.396   |
| Age at the clinic visit (years) | 41.55±11.25 | 28–62, 40 | 52.33±14.19 | 37–65, 55 |         |
| Age at the first onset (years) | 37±12 | 22–56, 34 | 47±17 | 31–64, 45 |         |
| Course of disease (years) | 4.7±2.8 | 1–10, 4 | 5.7±4.5 | 1–10, 6 |         |
| Bilateral (n, %) | 1, 9.1% |          | 2, 66.7% |          | 0.093   |
| Frequency of onset (n/year) | 1.5±0.8 | 1–3, 1 | 1.7±0.6 | 1–2, 2 | 0.209   |
| GC dependence (n, %) | 7, 63.6% |          | 1, 33.3% |          | 0.538   |
| Anti-HCMV IgG (s/co) | 0.94±0.79 | 0.24–2.39, 0.67 | 0.67±0.71 | 0.24–1.49, 0.27 |         |
| Response to antiviral therapy (n of Cc, %) | 5, 45.5% |          | 1, 33.3% |          | 1.000   |
| VA (LogMAR)      | 0.36±0.48 | 0.00–1.70, 0.22 | 0.23±0.32 | 0.00–0.6, 0.10 |         |
| Ratio of C/D     | 0.59±0.22 | 0.3–0.9, 0.5 | 0.47±0.12 | 0.4–0.6, 0.4 |         |
| Peak IOP        | 48±5 | 41–60, 48 | 48±3.5 | 45–52, 48 |         |
| Corneal endothelial cell loss (%) | 0.14±0.09 | −0.03–0.27, 0.12 | 0.08±0.09 | −0.02–0.14, 0.12 |         |
| KPs (n, %)       | Sheep-fat | 10, 90.9% |          | 2, 66.7% |          | 0.396   |
|                  | Coin-shaped | 5, 45.5% |          | 3, 100% |          | 0.209   |
|                  | Pigmented | 1, 9.1% |          | 2, 66.7% |          | 0.093   |
|                  | Tyndall effect (n, %) | 3, 27.3% |          | 2, 66.7% |          | 0.505   |
|                  | Iris depigmentation (n, %) | 8, 72.3% |          | 3, 100% |          | 1.000   |
|                  | Cataract (n, %) | 4, 36.4% |          | 2, 66.7% |          | 0.583   |
|                  | Glaucomatous optic neuropathy (n, %) | 8, 72.3% |          | 1, 33.3% |          | 0.505   |

Statistical analysis was performed using Fisher’s exact test.

CD, cup to disk ratio; gB, glycoprotein B; GC, glucocorticoid; HCMV, human cytomegalovirus; IgG, immunoglobulin G; IOP, intraocular pressure; KPs, keratic precipitates; LogMAR, logarithm of minimal angle resolution; n of Cc, number of patients with the ‘complete control’ outcome; VA, visual acuity.
and ocular fluid of patients with retinitis. The distribution of the gB genotype varies according to immune status and may type 2 is the most common gB genotype in the blood, urine retinitis in patients with AIDS. It has been found that genotype 2 is the most common gB genotype in the blood, urine and ocular fluid of patients with retinitis. The distribution of the gB genotype varies according to immune status and may thus influence prognosis. In previous studies, the genotype 1 and genotype 3 were found to be more prevalent in immunocompetent individuals and were more associated with non-fatal outcomes than other gB genotypes, especially genotype 2. The skewed genotype distribution caused differences in virulence, which might explain the differences in clinical manifestations of the fundus and anterior ganglia infection by CMV. In contrast, differences in gB genotypes may also result in different invasion pathways. Retinal infection by CMV is regarded as the result of haematogenous spread of the virus, while few patients with PSS show viremia by CMV. Thus, we speculate that the pathogenesis of CMV in the eyes of patients with PSS may be different from the direct blood invasion observed in the eyes of patients with retinitis.

In the present study, bilateral onset was predominant among patients infected with the genotype 3; this seemed to be different from the characteristics of most patients with PSS who are considered to be suffered unilateral onset. gB3 has been found to be more common among immunocompetent patients with congenital infections than among those with postnatal infections. Therefore, in addition, viruses with genotype 1 are considered to be weaker in lymphocyte tropism and virulence than those with genotype 2 and genotype 3. Therefore, we speculate that the clinical manifestations and prognoses of patients infected with genotype 1 CMV may be better than those of patients infected with other virus subtypes. We investigated whether there were other differences in the clinical characteristics of the two gB genotype groups, but no significant variations were found. Further studies with larger samples may be required to verify this finding.

The main limitation of this study is the small number of patients with genotype 3, which limited effective statistical analysis in this study. This is partly because of the generally low incidence of PSS, and partly because the viral load of CMV-infected patients with anterior uveitis is lower than that of patients with posterior uveitis or retinitis, which results in a lower acquired viral load after assay. In addition, the small quantity of aqueous humour samples (only 50–100 \( \mu \)L) yields a small number of available viruses.

In conclusion, this is the first report of the distribution of the gB genotypes of CMV in patients with PSS. CMV of genotype 1 was predominant in the aqueous fluid of patients with PSS. Patients infected with genotype 3 CMV mostly had bilateral attacks, but there was no significant difference in clinical characteristics between patients infected with different genotypes of CMV. The finding makes a significant contribution to the existing research and suggests CMV gB genotype may be an important target related to the virulence and pathogenicity of CMV strains in PSS.

**Clinical outcomes after topical ganciclovir treatment**

To evaluate the effect of antiviral therapy on PSS, we divided the outcomes into two categories, namely complete control and poor control. The outcomes were categorised based on records of IOP and inflammation within 1 year of treatment. When only antiviral therapy was performed, if IOP did not exceed 21 mm Hg and inflammation did not recur, the disease was considered to be under complete control; otherwise, it was considered to be poorly controlled. As shown in [table 2](#), in the gB genotype 1 group, 45.5% of the patients (5/11) showed a good response to the antiviral treatment and 4 patients underwent anti-glaucoma surgery within a year to reduce their IOP. 33.3% of the patients with the gB genotype 3 (1/3) who only received antiviral therapy had stable IOP and good control of inflammation for up to a year, whereas the remaining two patients needed to be treated with additional hormones and drugs to reduce IOP.

**DISCUSSION**

In this study, we determined the distribution of CMV gB genotypes in the aqueous humour of patients with PSS and investigated the differences in their clinical characteristics according to their different CMV genotypes. To the best of our knowledge, considering that PSS is generally considered as a rare condition and that the detection rate of CMV genome is relatively low in aqueous humour, the distribution of CMV gB genotypes in the aqueous humour of patients with PSS has not been published before. In addition, responses to antiviral treatment based on genotype distribution were also evaluated in this study.

In the present study, genotype 1 was the most prevalent genotype detected in the aqueous humour of patients with PSS. Only gB1 and gB3 were detected in this study; no other subtypes or mixed subtypes of genotype B were detected. A previous report of immunocompetent patients with CMV-related anterior uveitis showed that genotype 1 and genotype 3 were the common gB genotypes detected in the anterior chamber, a finding which is consistent with that of our study. However, the subset that causes inflammation in the anterior segment is distinct from the variants that cause posterior segment infection. In related studies of CMV infection in the posterior segment, gB genotype 2 was considered to be associated with human cytomegalovirus retinitis in patients with AIDS. It has been found that genotype 2 is the most common gB genotype in the blood, urine and ocular fluid of patients with retinitis. The distribution of the gB genotype varies according to immune status and may thus influence prognosis. In previous studies, the genotype 1 and genotype 3 were found to be more prevalent in immunocompetent individuals and were more associated with non-fatal outcomes than other gB genotypes, especially genotype 2. The skewed genotype distribution caused differences in virulence, which might explain the differences in clinical manifestations of the fundus and anterior ganglia infection by CMV. In contrast, differences in gB genotypes may also result in different invasion pathways. Retinal infection by CMV is regarded as the result of haematogenous spread of the virus, while few patients with PSS show viremia by CMV. Thus, we speculate that the pathogenesis of CMV in the eyes of patients with PSS may be different from the direct blood invasion observed in the eyes of patients with retinitis.

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**Contributors** RZ acquired and analysed the data. RZ wrote the first draft of the manuscript. ZW participated to extract DNA from the samples and helped with analysis. QS and XF participated in the collection and input of data. XS revised the manuscript. XK contributed to the experimental design and revised the manuscript.

**Funding** This work was supported by the Western Medicine Guidance Project of Shanghai Science and Technology Commission (grant number 19411961600), the Experimental Animal Research Project of Shanghai Science and Technology (grant number 201409006600), and the Double Excellent Project of Eye, Ear, Nose, and Throat Hospital (grant number SYB200303). The authors were funded by the Project of National Natural Science Foundation of China (grant numbers 81770922, 82070957, 81790641 and 81430007), the project of Shanghai Municipal Health Commission (grant number 201 740 204) and the Shanghai Science and Technology Innovation Project of the Shanghai Shenkang Hospital Development Centre (grant number SHDC12017x.18).

**Competing interests** None declared.
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Patient consent for publication Not required.
Provenance and peer review Not commissioned; externally peer reviewed.
Data availability statement Data are available upon reasonable request. Data are available upon reasonable request.

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