**Lycium intricatum** Boiss.: An unexploited and rich source of unsaturated fatty acids, 4-desmethylsterols and other valuable phytochemicals

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**Abstract**

**Background:** *Lycium intricatum* Boiss., a Solanaceous shrubbery is used in Tunisia as a windbreak and medicinal plant. However, it is considered as underexploited species despite its high potential to serve as a source with economic and nutritional value. To date only limited information about its phytochemistry, especially of its oil has been published. This work provides data on fatty acids, phytosterols and vitamin D composition of *L. intricatum* seed oil. It opens up new possibilities of developing *L. intricatum* as a new crop that contains phytochemicals with high added value little influenced by selection or commercial breeding.

**Findings:** The composition of fatty acids, phytosterols and vitamin D in *L. intricatum* seed oil was assessed by GC-FID. The main fatty acids of *L. intricatum* seed oil were linoleic acid (49.47 %), palmitoleic acid (27.96 %) and erucic acid (13.62 %). Palmitic acid was present at low percentage (0.63 %). The content of unsaturated fatty acids was high as 94.04 %. The sterolic fraction was composed of stigmasterol (18.56 mg/100 g), β-sitosterol (13.04 mg/100 g). *L. intricatum* oil is an oily matrix that contains hydrocarbons, mainly squalene (63.36 mg/100 g) and two triterpenic alcohol erythrodiol (80.36 mg/100 g) and uvaol (24.06 mg/100 g). provitamin D was present in high quantity (8.12 mg/100 g).

**Conclusions:** From these results it has been shown that *L. intricatum* seeds have great potential as a source of fatty acids and phytosterols for natural health products.

**Keywords:** *Lycium intricatum*, Seed oil, Fatty acids, Phytosterols, Vitamin D

**Additional non-English language abstract**

La composition en acides gras et en phytosterols des graines de *Lycium intricatum* Boiss. (Famille des Solanacées), de haute valeur nutritionnelle, a été étudiée moyennant la chromatographie en phase gazeuse (GC-FID). L’huile des graines de *L. intricatum* est caractérisée par des fortes teneurs en acide linoléique (49,47 %), acide palmitoleique (27,96 %) et acide erucique (13,62 %). Les acides gras insaturés présentent 94,04 % de la composition totale en acides gras. La fraction stérolique est composée du stigmasterol (18,56 mg/100 g) et du β-sitosterol (13,04 mg/100 g). L’huile de *L. intricatum* est principalement composée par des hydrocarbures tels que le squalène (63,36 mg/100 g) et deux alcool triterpeniques qui sont l’erythrodiol (80,36 mg/100 g) et l’uvaol (24,06 mg/100 g). La provitamine D présente une teneure de 8,12 mg/100 g.

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Background

The fatty acid composition of oils from vegetable sources varies depending on plant origin, genetic factors, ripening grade of fruits and specific climatic conditions. In addition to fatty acids, vegetable oils contain phytosterols which are divided into three main classes: 4-desmethylsterols (sterols), 4-monomethylsterol and the 4,40-dimethylsterol [1].

The unsaponifiable fraction of vegetable oils contains a variety of bioactive substances, which include sterols, hydrocarbons, tocopherols, terpenes and others. These minor compounds are more characteristic of each fat and oil [2, 3]. Moreover, it has been reported that phytosterols, which represent the predominant portion of unsaponifiable matter, have multifunctional properties, including anti-inflammatory, antitumor, hypercholesterolemia, antifungal and antibacterial activities [4–6].

The genus Lycium (Solanaeae family) has been identified as a rich source of polysaccharidic, proteins and particularly glycopeptides, which are responsible for many health related benefits of this plant. Lycium sp. contains 18 different amino-acids, including eight essential amino-acids. The genus includes more than 70 species growing in temperate to subtropical parts of North and South America, Southern Africa, Eurasia, and Australia [7]. Lycium sp. is well known as a traditional herbal medicine and functional food. Among the chemical constituents of Lycium fruits, the most well researched components are anthocyanins and flavonoids [8]. Recent studies indicate that extracts from some Lycium species possess a range of biological activities, including effects on ageing, neuroprotection, anti-fatigue/endurance, glucose control in diabetics, and antioxidant and anti-tumour properties [9, 10].

In Tunisia, four Lycium species have been identified: Lycium europaeum L., L. halimifolium Mill., L. arabicum Boiss., and L. intricatum Boiss. [11]. L. intricatum Boiss. is a common fleshy-fruited, thorny shrub up to 3 min height, typical of sub-humid and semi arid bioclimatic zones in Tunisia. It produces berries that are red when ripe. It is used as a hedge and as wind break plant. In addition the dry powder of its fruit was used to protect from eye diseases. However, in Tunisia L. intricatum is considered as underexploited specie despite its high potential to serve as source with economic and nutritional value. An improved knowledge about its chemical composition and biological activities would contribute to the use of this natural resource as a source of phytochemicals as well as to agronomic and economic advancement.

To the best of our knowledge, although the potential beneficial effects of L. intricatum were obvious, there are no report in the literature concerning its fatty acids and phytosterols composition. The aim of this study was to determine for the first time the fatty acid composition and phytosterol content of L. intricatum seed oil.

Results and discussion

Fatty acids profiles by GC

The yield of seed oil of L. intricatum aerial parts was 20 % (±3). Seed oil yield (w/w) as calculated on the basis of dry matter weight. A total of six different fatty acids were identified (Table 1 and Fig. 1). In L. intricatum oil, linoleic acid was the dominating fatty acid with an exceptional level, up to 49.47 % followed by palmitoleic acid (27.96 %) and erucic acid (13.62 %). Linoleic acid is an essential fatty acid and a precursor of arachidonic acid biosynthesis, the substrate for eicosanoid synthesis. According to Keys et al. [12], linoleic acid has hypocholesterolemia effects.

The content of unsaturated fatty acids was 94.04 %. Our results are not in agreement with those published by Altintas [13] for L. barbarum oil: hexadecanoic acid (47.5 %), linoleic acid (9.1 %), myristic acid (4.2 %) and ethylhexadecanoate (4 %). However, saturated fatty acids (SFA) fraction was characterized by a lower level (5.93 %). Recently, it was proven by clinical evidence that PUFAs are able to alleviate symptoms of certain diseases such as coronary heart disease, stroke and rheumatoid arthritis [14]. Also, linoleic acid has beneficial properties for skin, and for this purpose it is used by the cosmetics products industry [15]. These results bring attention to the possible use of cactus seed oil as a natural source of PUFAs for nutritional, industrial or pharmaceutical purpose. Indeed, different means are used to increase, directly or indirectly, the human consumption of PUFAs [16].

Oleic acid and linoleic acid exhibited contrasting accumulation patterns in L. intricatum fruit. The percentage of linoleic acid was 49.47 % whilst the oleic acid percentage 2.99 %. This is probably the consequence of an alteration of the desaturation step from oleic acid to linolenic acid, which is mediated by specific olate desaturase enzymes. This is in agreement with the results obtained by

Table 1 Fatty acid composition (in %) of L. intricatum seeds

| Fatty acid       | Percentage (%) |
|------------------|----------------|
| Myristic acid    | C14:0          | 5.3            |
| Palmitic acid    | C16:0          | 0.63           |
| Palmitoleic acid | C16:1          | 27.96          |
| Oleic acid       | C18:1          | 2.99           |
| Linoleic acid    | C18:2          | 49.47          |
| Erucic acid      | C22:1          | 13.62          |
| Saturated fatty  |                | 5.93           |
| Monounsaturated  |                | 44.57          |
| Polyunsaturated  |                | 49.47          |
Guoliang [17]. However, *L. intricatum* and *L. barbarum* present a disadvantage to fast oxidation due to their richness in polyunsaturated fatty acids. On the other hand, the higher content of unsaturated fatty acids (94.04 % *L. intricatum* and 86.5 % *L. barbarum*) which allow them to act as antiatherogenic and hypocholesterolemia agents in these two oils which are a good sources of omega 6 with antiallergic and anti-inflammatory properties.

**Phytosterol analysis by GC**

**Phytosterol (4-desmethylsterol) content**

The sterolic fraction was composed by stigmasterol (18.56 mg/100 g), β-sitosterol (13.04 mg/100 g), and ergosterol (8.12 mg/100 g). *L. intricatum* oil contained a higher amount of sterols 39.72 mg/100 g. Similar values were published by Potterat [10] for the sterol content in seed oil of two Goji species (*L. barbarum* and *L. chinense*). Recently, sterols have been added to vegetable oils as an example of a successful functional food [18]. This type of product is now available and has been scientifically proven to lower blood LDL. Cholesterol by around 10-15 % as part of a healthy diet [19].

In oil, Provitamin D was represented only by ergosterol. The Vitamin D level was higher in *L. intricatum* oil 8.12 mg/100 g. The high level of Vitamin D, detected in the oil, may contribute to great stability toward oxidation. Ergosterol is the end product of the sterol biosynthetic pathway and is the major sterol in yeasts. Like cholesterol in mammalian cells, it is responsible for membrane fluidity and permeability [20]. Previous works have been reported a role for ergosterol in physiological functions, such as membrane permeability, resistance to drugs, protein transport to the plasma membrane, sporulation and endocytosis [21–23].

A relatively high content of triterpenes such as erythrodiol (80.36 mg/100 g), uvaol (24.06 mg/100 g) and hydrocarbons, mainly squalene (63 mg/100 g) were detected in the seed oils of *L. intricatum* (Table 2 and Fig. 2). Das [24] reported that squalene, a hydrocarbon of lipid composition, exerted antioxidant effects used as food supplement and a vaccine additive.

A similar trend was also observed with hydrocarbons, mainly squalene; triterpenes such as uvaol, erythrodiol in Virgin olive oil [25], Vuorio et al. [26] and Rajaratnam et al. [27] have been found elevated ratios of serum squalene to cholesterol, leading them to propose that reduced cholesterol synthesis may be related to coronary atherosclerosis [28]. In fact, squalene administration modulates lesion development in a sex-specific manner and that it could be used as a safe alternative to correct hepatic steatosis and atherosclerosis, particularly in males [29].

Uvaol and erythrodiol exhibited antioxidant properties against lipid peroxidation in vitro, and also reduced the generation of hydrogen peroxide by stimulated macrophages in a dose-dependent manner [30–32]. Allouche et al. have shown that uvaol and erythrodiol significantly decreased thrombin formation [30], and they also inhibit cell proliferation in a dose- and time-dependent manner [32]. These compounds have interesting therapeutic potential as cardiovascular drugs.

*L. intricatum* oil is an oily matrix that contains hydrocarbons, mainly squalene; triterpenes such as uvaol, and erythrodiol, phytosterols, and a wide range of phenolic compounds comprising polyphenols, flavonoids, and anthocyanins. The 4-desmethylsterois are the final

**Table 2** Sterols (4-desmethylsterols) composition (mg/100 g of oil) of *L. intricatum* seed

| Phytosterols (mg/100 g) |  
|-------------------------|
| Provitamin D (Ergosterol) | 8.12 |
| Stigmasterol | 18.56 |
| β-Sitosterol | 13.04 |
| **Total** | **39.72** |
| Squalene (mg/100 g) | 63.36 |
| Erythrodiol | 80.36 |
| Uvaol | 24.06 |
products of the phytosterols biosynthesis; this category corresponds to plant sterols accumulated mainly in the fruit and seeds. These compounds display anti-inflammatory properties and have long been considered to be the main active principle of *Lycium* sp [33].

**Conclusions**
This study shows that *Lycium intricatum* seed oil was found to possess 94.04 % PUFA. Linoleic acid was the dominating fatty acid with an exceptional level, up to 49 %. Oil contained a higher amount of sterols. These results bring attention to the possible use of *Lycium* seed oil as a natural source of PUFA for nutritional, industrial or pharmaceutical purpose. *L. intricatum* oil is an oily matrix that contains hydrocarbons, mainly squalene, and triterpenes such as uvaol and erythrodiol. This study helps to develop *L. intricatum* as a new crop for oil production.

**Methods**

**Chemicals**
Solvents of analytical (n-hexane, petroleum ether, chloroform, EtOH and MeOH) grade were purchased from Carlo Erba Reactif-CDS (Val de Reuil, France). Sodium sulfate (Na₂SO₄), sodium methylete (CH₃ONa), potassium hydroxide (KOH), chlorhydric acid (HCl) were obtained from Merck (Darmstadt, Germany). The standards α-cholestanol, and 3,3-bis(4-hydroxyphényl)-1-(3H)-monobenzofuranone (φφ) were sourced from Sigma–Aldrich (St. Louis, MO).

**Plant material**
While there are no signs of toxicity of this plant, Tunisian peoples collect its fruits only in the full maturity stage to avoid risk of toxicity. So, fruits of *L. intricatum* were collected in May 2013, from the region of Sidi Thabet, area of Ariana (Northern Tunisia, latitude 36°54'45.25"N, longitude 10°06'02.10"E, altitude 30 m) at full maturity stage. Seeds (3 × 25 g) were carefully separated from fruits prior to extraction.

**Lipid extraction**
Oil was extracted by a soxhlet extractor for 6 h using n-hexane, chloroform and petroleum ether as solvent. The solvent was evaporated under reduced pressure, using a rotary evaporator at 40 °C. The oil content was determined as the difference in weight of dried *L. intricatum* seed sample before and after the extraction [34]. Oil was weighed and stored at −20 °C. All the analyses were conducted in triplicate.

**Fatty acid methyl ester (FAME) preparation and gas chromatography analysis**
In 25 mL round bottom flask, oil samples (3 g) were added to 3 mL methylete sodium solution with φφ. Mixture was refluxed for 10 min, and 3 mL of MeOH solution of hydrochloric acid (a mixture of hydrochloric acid gas and anhydrous methanol or, acetyl chloride and anhydrous methanol) were added to φφ discoloration. The mixture was refluxed for 10 min and then cooled at room temperature.
Hexane (8 mL) and water (10 mL) were added and the organic phase was recovered, dried over anhydrous sodium sulphate and filtered for subsequent GC analysis. FAMEs were analyzed by gas chromatography (GC) using an Agilent 6890 chromatograph series using an Innowax capillary column with the following characteristics: length, 50 m; internal diameter, 0.30 mm; film thickness 0.25 μM. The carrier gas was helium, at a flow through the column of 1 mL/min. The injector temperature was maintained at 230 °C and the flame-ionisation detector (FID) at 250 °C. The oven temperature was 165 °C. The fatty acids were identified by comparison of their retention times with those of standards (COI/T.20/Doc. n° 24).

Saponification
Unsaponifiable lipids were determined by saponifying 5 g of lipid extracts with 50 mL ethanolic KOH 2 N (w/v) mixed with both 0.5 mL α-cholestanol solution (internal standard: 0.2 % (w/v)) and heating at 60 °C for 1.30 h. After cooling, 50 mL of H₂O was added and the unsaponifiable matter was extracted four times with 80 mL ethyl ether. The combined ether extract was washed with 50 mL of H₂O. The ether extracted was dried over anhydrous Na₂SO₄ and evaporated. The dry residues were dissolved in chloroform for TLC analysis.

Thin-layer chromatography
The unsaponifiable matter was separated into subfractions on preparative silica gel thin-layer plates (silica gel 60 G F254), using 1-dimensional TLC with hexane-Et₂O (65:35 V/V) as the developing solvent. The unsaponifiable (0.3 mL CHCl₃) containing 1 % (w/w) of α-cholesterol as the internal standard for 4-desmethylsterols was applied on the silica gel plates in 3 cm bands. To correctly identify the sterols bands, a reference sample of purified sterol (α-cholesterol) were applied on the left side of the TLC plates. After development the plate was sprayed with 2,7'-dichlorofluorescein and viewed under UV light. On the basis of the reference spot, the sterols band was identified. The band corresponding to 4-desmethylsterols was scraped off separately and was extracted three times with CHCl₃-Et₂O (1:1), filtered to remove the residual silica, dried in a rotary evaporator and stored at 10 °C for further analysis.

GC analysis
Each sterol fraction was silylated with 100 μL of Bis (trimethylsilyl) trifluoroacetamide + 1 % trimethylchlorosilane agent at 6 °C for 30 min. Phytosterols content was determined using gas chromatography. An Agilent 6890 N capillary gas chromatograph equipped with an automatic split/splitless injection (1 μL), flame-ionization detector and HP-5 MS column (30 m × 0.25 mm L.D., 0.25 μm film thickness) was used. Operating conditions were: injection temperature, 260 °C; detector temperature, 320 °C, oven temperature 140 °C to 300 °C, at 10 °C/min, holding at 300 °C for 14 min, helium as carrier gas (1.9 mL/min). Identification and quantification were based on external standards using ergosterol, stigmastanol and β-sitosterol from Sigma-Aldrich Inc (St. Louis, MO) [35].

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
AB and AB have carried out the experimental part such as selection of plant material, injections and identification of fatty acids and phytosterols. Béjaoui evaluated the results and corrected the manuscript for publication. All authors read and approved the final manuscript.

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