Luseogliflozin reduces epicardial fat accumulation in patients with type 2 diabetes: a pilot study

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Abstract

Background: Accumulation of epicardial fat (EF) is associated with increased cardio-metabolic risks and coronary events, independently of traditional cardiovascular risk factors. Therefore, the reduction of EF volume (EFV) may be associated with reduced cardio-metabolic risks and future cardiovascular events. Sodium-glucose co-transporter-2 (SGLT2) inhibitors reduce body fat including visceral fat and cardiovascular events in patients with type 2 diabetes. However, it has still been unknown whether SGLT2 inhibitors can reduce EFV.

Methods: Type 2 diabetic patients with HbA1c 6.5–9.0% and body mass index (BMI, kg/m²) ≥ 25.0 were enrolled in this single arm pilot study. Participants were administered luseogliflozin 2.5 mg daily and the dosage was tolerated to be increased up to 5.0 mg daily. EFV [median (interquartile range), cm³] was measured by magnetic resonance imaging. Primary endpoint was the decrease in EFV at 12 weeks. Visceral fat area (VFA, cm²) and liver attenuation index (LAI) measured by the abdominal computed tomography, and skeletal muscle index (SMI) and body fat (%) measured by the whole body dual-energy X-ray absorptiometry were also determined at baseline and at 12 weeks.

Results: Nineteen patients (mean age: 55 ± 12 years; 26% female) completed this study. Luseogliflozin treatment significantly reduced EFV at 12 weeks [117 (96–136) to 111 (88–134), p = 0.048]. The body weight, BMI, systolic and diastolic blood pressure, HbA1c, fasting plasma glucose, insulin, homeostasis model assessment-insulin resistance (HOMA-IR), triglycerides, SMI, and body fat were significantly reduced by luseogliflozin at 12 weeks. The reduction of EFV was significantly correlated with the reduction of C-reactive protein (r = 0.493, p = 0.019). Neither VFA nor LAI were significantly reduced by the luseogliflozin treatment. No severe adverse events were observed.

Conclusions: Our data suggest that luseogliflozin could reduce the EFV in parallel with the improvement of systemic micro-inflammation and the reduction of body weight in Japanese patients with type 2 diabetes. The reduction of muscle mass after the administration of SGLT2 inhibitors may require a particular attention.

Trial registration umin.ac.jp, UMIN000019072

Keywords: Luseogliflozin, Epicardial fat, Type 2 diabetes
Background
Epicardial fat (EF), the local adipose tissue surrounding the heart, located between the myocardium and the visceral pericardium, has been recognized as one of the phenotypes of ectopic fat accumulation [1, 2]. EF accumulation is thought to be involved in the progression of coronary atherosclerosis, through the secretion of bioactive inflammatory cytokines and adipokines [3–6]. It has recently been reported that EF volume (EFV) is associated with non-calcified coronary plaque [7–9]. In addition, EFV is associated with cardio-metabolic risks including insulin resistance [10, 11] and type 2 diabetes [12] and both fatal and nonfatal coronary events, independently of traditional cardiovascular risk factors [9]. It is therefore conceivable that the reduction of EFV could be associated with reduced cardiac metabolic risks and future cardiovascular events.

EFV was reported to be correlated with obesity [13]. In addition to obesity, diabetes was also associated with EFV [14]. Therefore, obese patients with type 2 diabetes have an extremely high risk for developing ectopic adi- 

EFV was reported to be correlated with obesity [13]. In addition to obesity, diabetes was also associated with EFV [14]. Therefore, obese patients with type 2 diabetes have an extremely high risk for developing ectopic adiposity [15]. In diabetic patients, EFV was associated with dyslipidemia [16] and endothelial dysfunction [17], compared to non-diabetic subjects. Furthermore, a recent study demonstrated that diabetes, even in the absence of obesity, is associated with an increased myocardial triglyceride content, increased hepatic triglyceride content, and impaired myocardial energetics [18]. The authors also demonstrate that the degree of both hepatic and epicardial fat accumulations are associated with a cardiac contractile dysfunction in the patients with diabetes. Given these findings, the evaluation of EFV may be important to predict future coronary events, especially in obese patients with diabetes.

Sodium-glucose co-transporter-2 (SGLT2) inhibitors lower the blood glucose level in patients with type 2 diabetes by decreasing the renal glucose reabsorption [19] and its improvement of glycemic control is parallel with the reduction of body weight and visceral fat [20, 21]. Recently, it has been revealed from the EMPA-REG outcome study that empagliflozin significantly reduces the composite outcome of death from cardiovascular causes, nonfatal myocardial infarction, or nonfatal stroke (3-point MACE) in patients with type 2 diabetes and high cardiovascular risks [22]. Although it has still been elusive which mechanisms are involved in the favorable effects of empagliflozin on cardiovascular events, it is expected that the reduction of EFV in addition to the reduction of body weight, may be associated with the improvement of CV outcomes. Therefore, we hypothesized that the administration of SGLT2 inhibitors could reduce the EFV in patients with type 2 diabetes, and its reduction could be correlated with the changes in insulin sensitivity, body fat, and secretion of adipokines or cytokines, all of which are associated with cardiovascular events. A pooled analysis of four 52-week Phase III trials of luseogliflozin in Japanese patients with type 2 diabetes has recently showed that luseogliflozin is especially beneficial in patients with a higher body mass index (BMI) in terms of metabolic abnormalities, including obesity, hyperinsulinemia, and hypertension [23]. It is also reported that luseogliflozin can reduce a waist circumference, a surrogate marker for visceral adiposity, with a significant reduction of HbA1c among Japanese patients with type 2 diabetes [24]. Given these findings, we have focused on the potential of luseogliflozin to improve the abnormal regional fat distribution, especially an excessive accumulation of EFV and we conducted this small-scale pilot study to investigate whether luseogliflozin could reduce the EFV in Japanese patients with type 2 diabetes.

Methods
Study design
This was a 12-week single-arm pilot study to assess the effects of luseogliflozin on EFV in patients with type 2 diabetes. This study was undertaken in accordance with the principles of the Declaration of Helsinki and has been approved by the ethical committee of Tokyo Medical and Dental University (No. 2102). This study was registered at the UMIN Clinical Trial Registry (UMIN000019072). All patients provided written informed consent before participation.

Subjects
We screened patients with type 2 diabetes aged older than 20 years who regularly visited the Tokyo Medical and Dental University Hospital. The inclusion criteria were as the followings: (1) type 2 diabetes diagnosed according to the criteria of the Japan Diabetes Society (JDS) [25], (2) a BMI equal to or greater than 25 kg/m², (3) HbA1c levels between 6.5 and 9.0%. Exclusion criteria were: (1) type 1 diabetes, (2) any history of taking SGLT2 inhibitors, (3) insulin treatment, (4) severe renal or liver disease, (5) malignant neoplasm, (6) pregnant women, (7) any history of acute coronary syndrome or stroke within 3 months prior to the enrollment, (8) patients whose treatment for diabetes were changed 3 months prior to baseline evaluation, (9) those who had contraindications for magnetic resonance imaging (MRI).

Intervention
Luseogliflozin was administered from 2.5 mg once daily after breakfast and the dosage was allowed to be increased up to 5.0 mg if HbA1c kept more than 7.0%
by 2.5 mg of luseogliflozin. During the 12 weeks, anti-hyperglycemic agents other than luseogliflozin were unchanged, except when unacceptable hyperglycemia, hypoglycemia, or adverse events (AEs) occurred. The diet/exercise therapy and the combination anti-hyperglycemic agents were to be continued unchanged from the baseline until the end of the study.

Clinical and biochemical analysis
Blood samples were collected at an overnight fasting state. HbA1c was measured using the latex agglutination method. HbA1c levels were expressed in accordance with the National Glycohemoglobin Standardization Programs recommended by the Japanese Diabetes Society [25]. The insulin sensitivity was assessed by a homeostasis model assessment as an index of insulin resistance (HOMA-IR) in patients whose fasting plasma glucose levels were less than 7.8 mmol/l. The systolic and diastolic blood pressures (SBP and DBP) were measured in a sitting position after at least 5 min rest, using an electronic sphygmmomanometer (ES-H55, Terumo Inc., Tokyo, Japan). The BMI was calculated as the weight divided by the square of the height (kg/m²). Routine tests included alanine transaminase (ALT), aspartate transaminase (AST), gamma-glutamyl transeptidase (γ-GTP), high-density lipoprotein (HDL) and low-density lipoprotein (LDL) cholesterol, triglycerides, uric acid, C-reactive protein, hemoglobin, and hematocrit. All tests were determined using standard laboratory procedures. Urinary albumin and creatinine excretion were measured in a spot urine collection by a turbidimetric immunoassay and enzymatic method. The ratio (ACR, mg/g) was used for the assessment of albuminuria. The GFR was calculated using the following equation for the Japanese [26]; GFR = 194 × SCr⁻¹.⁰⁹⁴ × age⁻⁰.₂₈⁷ [(if female) × 0.739], where SCr (mg/dl) is measured by an enzymatic method. Adiponectin, leptin and interleukin 6 (IL-6) were measured using latex turbidimetric immunoassay, double-antibody radioimmunoassay, and chemiluminescent enzyme immunoassay, respectively.

Quantification of abdominal adiposity, hepatic fat accumulation by CT
Both visceral fat area (VFA) and subcutaneous fat area (SFA) were measured at the level of umbilicus by the abdominal computed tomography (CT) examination (Aquilion PRIME, Toshiba Medical Systems, Tochigi, Japan) as described previously [27]. The hepatic fat accumulation was also determined by the liver attenuation index (LAI) in the CT examination as described previously [28]. The LAI was calculated as follows: the average attenuation value of the liver (eight points) divided by the average attenuation value of the spleen (three points).

Body composition measured by DXA
The regional fat and fat-free mass were measured using the whole body DXA (Lunar idDXA, GE Healthcare, Madison, WI, USA) as described previously [27]. The skeletal muscle index (SMI) was calculated as the appendicular non-fat mass divided by the square of the height (kg/m²). The body fat (%) was calculated as the whole body fat mass divided by the body weight.

Quantification of epicardial fat volume by MRI
The EFV was measured using 1.5T-MRI system (Achieva Dual, Philips Healthcare, Netherland or Titan, Toshiba Medical Systems, Japan) with a 32-channel commercially available cardiac coil and was assessed using a modified fat excitation based on the whole-heart coronary MR angiography sequence [29]. A patient-specific acquisition window was set during either systole or diastole, depending on the phase of minimal motion of the right coronary artery. Typical parameters of the sequence included repetition time, 3.2 ms; echo time, 1.6 ms; flip angle, 15°; sensitivity-encoding factor, 2.5; field of view, 330 × 330 × 128 mm; acquisition matrix, 248 × 238; acquired spatial resolution, 1.3 × 1.4 × 1.6 mm, and reconstructed voxel size, 0.64 × 0.64 × 0.8 mm. All MR mages were transferred to a workstation (Ziostation 2, Japan). Semiautomatic analyses of EFV were performed by an experienced operator blinded to the clinical information.

Study endpoints
The primary endpoint of this study was the decrease in EFV at 12 weeks. The secondary endpoints were the losses in the following variables at 12 weeks: VFA, SFA, SMI, LAI, HbA1c, HOMA-IR, blood pressure, lipid profile, albuminuria, C-reactive protein (CRP), adiponectin, leptin, and IL-6. We further determined the impact of luseogliflozin on the EFV and VFA levels normalized by the body surface area (BSA) using the ratios of the EFV and VFA divided by the BSA (EFV/BSA ratio and VFA/BSA ratio).

Statistical analysis
Statistical analysis was carried out using a IBM SPSS version 21.0 statistical package (IBM Corp. Released 2012. IBM SPSS Statistics for Windows, Version 21.0. Armonk, NY: IBM Corp.). Data are presented as the mean ± standard deviation (SD) or standard error of mean (SE), median with interquartile range (IQR), or percent as appropriate according to the data distribution. Pearson product-moment correlation analysis was used to investigate the correlation of EFV with the markers for body composition and cardio-metabolic risks.
The primary endpoint was analyzed using the Wilcoxon signed-rank test. The secondary endpoints were also analyzed by paired t test or Wilcoxon signed-rank test. p values <0.05 were considered to be statistically significant.

**Results**

**Demographics**

Twenty patients were recruited and inform consent was obtained from all subjects. At baseline, one patient was not eligible for the study because the HbA1c level was less than 6.5% and finally 19 patients (95.0%) completed the study. Table 1 presents the baseline demographic data and Table 2 shows the medications at baseline. Regarding the anti-diabetic medications, the patients were treated primarily with metformin and/or dipeptidyl peptidase 4 (DPP4) inhibitors. Approximately one-third of the participants were receiving calcium channel blockers, angiotensin receptor blockers, and anti-platelet agents and statins were prescribed to more than 50% of the patients.

**Correlation of EFV with markers for body composition and cardio-metabolic risks**

As shown in Table 3, the EFV was significantly correlated with the BMI, VFA, total fat mass, and HOMA-IR and tended to be correlated with the SFA. In contrast, no significant association was observed between the EFV and the markers for cardio-metabolic risks, hepatic steatosis, adipokines, and SMI.

**Efficacy and safety**

As shown in Table 1, luseogliflozin treatment significantly reduced the EFV at 12 weeks; whereas, the hepatic fat accumulation (LAI) kept unchanged by luseogliflozin. Among markers for body composition, body fat (%), total fat and non-fat mass, non-fat mass in the upper and the lower extremities, and the appendicular SMI, all of which were measured using the whole body DXA, were significantly decreased by luseogliflozin. In contrast, neither VFA nor SFA, measured by abdominal CT were unchanged by luseogliflozin at 12 weeks. When EFV and VFA were normalized by the BSA (EFV/BSA ratio and VFA/BSA ratio), the significant impact of luseogliflozin on the decline of EFV was slightly attenuated [58.4 (50.7–70.0) cm³/m² at baseline to 57.2 (46.5–68.5) cm³/m² at 12 weeks, p = 0.064] and luseogliflozin did not reduce the VFA/BSA ratio [89.1 (61.4–106.2) cm²/m² at baseline to 86.7 (70.1–103.5) cm²/m² at 12 weeks, p = 0.619]. Among the cardio-metabolic risk factors, fasting plasma glucose, insulin, and HbA1c levels were significantly reduced by luseogliflozin. The insulin sensitivity, assessed by HOMA-IR, was significantly improved

| Table 1 Clinical characteristics at baseline and at 12 weeks after the administration of luseogliflozin in patients with type 2 diabetes |
|-----------------|-----------------|-----------------|
|                | Baseline        | 12 weeks        |
| Age (years)    | 55 ± 12         |                 |
| Gender (% female) | 26             |                 |
| SBP (mmHg)     | 137 ± 14        | 128 ± 15        |
| DBP (mmHg)     | 83 ± 7          | 80 ± 7          |
| Body weight (kg) | 82.2 ± 11.4    | 79.5 ± 11.5     |
| Body mass index (kg/m²) | 28.7 ± 2.7  | 27.8 ± 2.9     |
| Waist circumference (cm) | 101 ± 8   | 99 ± 8         |
| HbA1c (mmol/mol) | 57.9 ± 5.4     | 50.9 ± 4.5     |
| HbA1c (%)      | 7.5 ± 0.7       | 6.8 ± 0.6       |
| Plasma glucose (mmol/l) | 7.6 ± 2.2     | 6.8 ± 1.6       |
| Insulin (μU/ml) | 8.9 (5.3–13.6) | 8.1 (4.7–11.7) |
| HOMA-IR (N=15) | 3.6 (1.5–4.8)   | 2.6 (1.3–3.8)   |
| Triglycerides (mmol/l) | 1.67 (1.15–2.85)| 1.10 (0.93–2.39)|
| HDL cholesterol (mmol/l) | 1.38 ± 0.36  | 1.36 ± 0.33   |
| LDL cholesterol (mmol/l) | 2.89 ± 0.51  | 3.02 ± 0.66    |
| AST (U/l)      | 24 (18–40)      | 23 (17–28)      |
| ALT (U/l)      | 19 (17–43)      | 23 (18–30)      |
| γ-GTP (U/l)    | 35 (23–80)      | 28 (19–59)      |
| Estimated GFR (ml/min/1.73 m²) | 69.7 ± 13.8 | 68.1 ± 13.3 |
| Log urinary ACR (mg/g) | 1.62 ± 0.55 | 1.56 ± 0.48 |
| Uric acid (μmol/l) | 351 ± 64       | 326 ± 91       |
| Log CRP (mg/l) | −0.01 ± 0.45    | −0.10 ± 0.33    |
| Hemoglobin (g/dl) | 141 ± 3.3      | 153 ± 1.3      |
| Hematocrit (%) | 43.4 ± 3.1      | 45.7 ± 3.7     |
| Adiponectin (μg/ml) | 8.8 ± 3.3    | 8.5 ± 3.4     |
| Leptin (ng/ml) | 13.9 ± 6.1      | 13.1 ± 6.8     |
| Log Interleukin 6 (pg/ml) | 0.30 ± 0.23   | 0.24 ± 0.26   |
| EFV (cm³)      | 117 (96–136)    | 111 (88–134)    |
| VFA (cm²)      | 167 (134–201)   | 151 (124–209)   |
| SFA (cm²)      | 226 (184–273)   | 218 (178–240)   |
| Liver attenuation index | 1.03 ± 0.21  | 1.04 ± 0.22    |
| Body fat (%)   | 36.9 ± 5.6      | 36.2 ± 6.0     |
| Non-fat mass in upper extremities (kg) | 5.7 ± 1.3    | 5.5 ± 1.3     |
| Non-fat mass in lower extremities (kg) | 16.8 ± 3.3   | 16.3 ± 3.4    |
| Android (kg)  | 3.4 ± 1.4       | 2.9 ± 0.8       |
| Gynoid (kg)   | 4.6 ± 1.8       | 3.8 ± 1.0       |
| Total fat mass (kg) | 29.3 ± 5.9    | 27.9 ± 6.3     |
| Total non-fat mass (kg) | 50.4 ± 8.4  | 49.3 ± 8.4    |
| Skeletal muscle index | 7.81 ± 1.09  | 7.58 ± 1.11    |

ACR albumin-to-creatinine ratio, ALT alanine transaminase, AST aspartate transaminase, CRP C-reactive protein, DBP diastolic blood pressure, EFV epicardial fat volume, GFR glomerular filtration ratio, GTP glutamyl transpeptidase, HDL high-density lipoprotein, HOMA-IR homeostasis model assessment as an index of insulin resistance, LDL low-density lipoprotein, SBP systolic blood pressure, SFA subcutaneous fat area, VFA visceral fat area
in patients whose fasting plasma glucose level was less than 7.8 mmol/l (N = 15). Significant reductions of both the systolic and diastolic blood pressures were observed by luseogliflozin. The triglycerides level was significantly reduced by luseogliflozin, whereas the HDL cholesterol level was unchanged. The adiponectin, leptin, and IL-6 levels were unchanged by luseogliflozin. No severe AEs including severe hypoglycemic episodes were observed during the study period.

Correlation of changes in EFV, LAI, VFA, SFA with those in markers for body composition and cardio-metabolic risks by luseogliflozin

Table 4 shows the correlation of the changes in EFV, LAI, VFA, and SFA with those in the markers for body composition, cardio-metabolic risks, and adipokines. The loss of EFV was significantly correlated with the reductions in both BMI and log CRP (Table 4). In contrast, the increase in LAI was significantly correlated with the decreases in VFA, body fat (%), and log CRP. Among adipokines, the decrease in leptin was significantly correlated with the decrease in VFA as was the increase in adiponectin with the reduction in SFA. However, neither the decrease in EFV nor the increase in LAI showed a significant association with the changes in adipokines.

Discussion

This single-arm pilot study demonstrates that luseogliflozin treatment significantly reduces EFV in patients with type 2 diabetes and the reduction of EFV is associated with the improvement of micro-inflammation and the reduction of body weight. To the best of our knowledge, this is the first study to examine the effect of SGLT2 inhibitors on EFV in patients with type 2 diabetes and the finding suggests the possibility that SGLT2 inhibitors may reduce cardiovascular, especially coronary artery events partly by reducing the EFV in patients with type 2 diabetes.

EF accumulation has recently been reported to be associated with a non-calcified coronary plaque [7–9], cardio-metabolic risks including insulin resistance [10, 11] and type 2 diabetes [12] and fatal and nonfatal coronary events, independently of traditional cardiovascular risk factors [9]. These observations imply the presence of mechanisms that can account for the increased risks of coronary artery disease by an increase in EFV. Adipokines and inflammatory cytokines secreted from EF are thought to physiologically regulate heart vessels, via paracrine and vasocrine mechanisms [30] and EF is recognized as an energy reservoir for resident cardiomyocytes [31]. It is therefore conceivable that the reduction of EFV could be associated with reduced cardio-metabolic risks, presumably leading to the prevention of
future cardiovascular events, via the reduction of proinflammatory adipokines. In this study we were unable to see the significant changes in the levels of adipokines by luseogliflozin treatment. It may be due to the relatively small reduction of body fat including EFV by luseogliflozin. Finally, luseogliflozin may have less of an impact on the circulatory levels of adipokines.

When considering the correlation between cardiometabolic risks and EFV at baseline, the EFV was significantly correlated with the BMI, VFA, and HOMA-IR in this study. EF embryologically derives from the splanchnopleuric mesoderm [32], sharing an embryologic origin with the intra-abdominal visceral adipose tissue [33, 34]. Therefore, it may be acceptable that patients with a high EFV are at a high risk for the increased visceral adiposity, leading to the peripheral insulin resistance. Indeed, EFV was reported to be strongly correlated with VFA [35]. Given these findings, we believe that the statistical significance of EFV with other markers for adiposity or insulin resistance in this study can be considered as moderate (Pearson’s correlation coefficient were approximately 0.5), although the sample size of this study was small. In contrast, luseogliflozin treatment significantly reduced EFV but not VFA in the present study even though some previous studies demonstrated that SGLT2 inhibitors can reduce visceral adiposity [36, 37]. This unexpected result might be partially explained by the small sample size of this study. Additionally, it has been reported that the reduction of EFV was not correlated with that of VFA in severely obese patients with a rapid weight loss by bariatric surgery [38]. Therefore, further large-scale clinical studies are needed to elucidate the association between the reduction of EFV and that of VFA in case obese patients with diabetes lose weight. We also revealed that the reduction of EFV is associated with the reduction of CRP. Systemic micro-inflammation causes insulin resistance and is located in the central of the initiation and the propagation of atherosclerosis [39]. It is therefore possible that luseogliflozin could reduce the risks for future cardiovascular events partially by the attenuation of systemic micro-inflammation.

There is growing evidence that diabetic treatment can reduce the EFV in patients with diabetes. Sack et al. firstly showed that pioglitazone decreases a genetic expression of proinflammatory and anti-inflammatory cytokines in EF in patients with type 2 diabetes and coronary

### Table 4 Correlations of the changes between markers for body composition and the markers for cardio-metabolic risks in patients with type 2 diabetes

| Changes in body composition markers | Delta EFV |  | Delta LAI |  | Delta VFA |  | Delta SFA |  |
|-----------------------------------|----------|---|-----------|---|-----------|---|-----------|---|
| Body weight (kg)                  | 0.301    | 0.113 | −0.616    | 0.003 | 0.686     | 0.001 | 0.250     | 0.159 |
| Body mass index (kg/m²)           | 0.387    | 0.046 | −0.531    | 0.012 | 0.719     | <0.001 | 0.292     | 0.120 |
| EFV                              | NA       | 0.154 | 0.264     | −0.237 | 0.164     | −0.075 | 0.168     | 0.245 |
| LAI                              | 0.154    | 0.264 | NA        | −0.391 | 0.049     | −0.168 | 0.245     | 0.130 |
| VFA (cm²)                        | −0.237   | 0.164 | −0.391    | 0.049 | NA        | 0.272 | 0.130     | 0.130 |
| SFA (cm²)                        | −0.075   | 0.379 | −0.168    | 0.245 | 0.023     | 0.463 | NA        | 0.130 |
| Body fat (%)                     | 0.103    | 0.342 | −0.468    | 0.025 | 0.296     | 0.117 | −0.149    | 0.277 |
| Total fat mass (kg)              | 0.166    | 0.255 | −0.723    | <0.001 | 0.284     | 0.127 | 0.082     | 0.374 |
| Total non-fat mass (kg)          | −0.063   | 0.402 | 0.224     | 0.186 | −0.255    | 0.154 | −0.082    | 0.374 |
| Skeletal muscle index            | 0.187    | 0.229 | −0.154    | 0.271 | 0.143     | 0.285 | 0.350     | 0.047 |

Changes in cardio-metabolic markers

| HbA1c (%)                         | 0.256    | 0.152 | −0.150    | 0.277 | 0.073     | 0.387 | 0.282     | 0.128 |
| HOMA-IR                           | −0.271   | 0.155 | 0.315     | 0.118 | 0.306     | 0.117 | 0.094     | 0.364 |
| ALT (U/l)                         | −0.044   | 0.490 | −0.170    | 0.251 | 0.249     | 0.159 | −0.203    | 0.210 |
| Log CRP (mg/l)                    | 0.493    | 0.019 | −0.453    | 0.029 | −0.039    | 0.438 | 0.302     | 0.112 |
| Log ACR (mg/g)                    | −0.069   | 0.393 | 0.042     | 0.434 | 0.333     | 0.048 | −0.069    | 0.393 |
| Adiponectin (μg/ml)               | −0.228   | 0.189 | 0.154     | 0.264 | −0.167    | 0.261 | −0.571    | 0.008 |
| Leptin (ng/ml)                    | 0.146    | 0.295 | 0.115     | 0.330 | 0.378     | 0.044 | −0.138    | 0.305 |
| Log interleukin 6 (pg/ml)         | −0.411   | 0.064 | −0.318    | 0.115 | 0.296     | 0.142 | 0.007     | 0.490 |

ACR albumin-to-creatinine ratio, ALT alanine transaminase, CRP C-reactive protein, DBP diastolic blood pressure, EFV epicardial fat volume, HDL high-density lipoprotein, HOMA-IR homeostasis model assessment as an index of insulin resistance, LAI liver attenuation index, SBP systolic blood pressure, SFA subcutaneous fat area, VFA visceral fat area.
artery disease [40]. More recently, Lima-Martínez et al. reported that the addition of sitagliptin produced a significant and rapid reduction of EFV in overweight/obese patients with type 2 diabetes who were inadequately controlled on metformin monotherapy [41]. Iacobellis et al. also has demonstrated that liraglutide, glucagon-like peptide 1 receptor agonist, has a similar effect of reducing the cardiac fat independently of the loss of body weight [42]. Therefore, we believe that the findings in the present study expand evidence of therapeutic approach to preventing coronary artery disease by diabetic treatment aimed at the reduction of EFV.

In contrast to the favorable effects of luseogliflozin on the reduction of EFV and the improvement of glycemic control, we need to address the significant reduction of non-fat mass and SMI by luseogliflozin in this study. It is reasonable that SGLT2 inhibitors can be administered in patients who are expected to show the decrease in both visceral and ectopic fat including EFV and no loss of the muscle mass by these drugs. Unfortunately, luseogliflozin reduced SMI in all patients in this study. Therefore we were unable to clarify the clinical features of patients who can be safely administered luseogliflozin from the perspective of muscle mass. A decrease in skeletal muscle mass is a component of sarcopenia which is defined as the progressive loss of muscle mass and function with aging [43, 44]. Diabetes is a strong risk factor for both the decrease in muscle mass and the loss of muscle function [45, 46]. Therefore, to avoid the unfavorable decrease in muscle mass, the administration of SGLT2 inhibitors should be carefully considered in patients with diabetes who are at a high risk for sarcopenia, especially in elderly patients with diabetes. Resistance training is reported to be associated with the attenuation of a rapid decline of muscle mass in elderly patients with diabetes [47]. It is therefore expected in the future studies that the combination of SGLT2 inhibitors and resistance training could be feasible and effective to preserve the muscle mass and function in patients with diabetes.

Potential limitations of our study are as follows: (1) this study is a single-arm pilot study and the sample size is small. Therefore, larger randomized control trials are needed; (2) no information is available regarding the diet and exercise which could influence not only the glycemic control but also the changes in body composition including EFV; (3) it is to be elucidated whether the findings in this study could be observed in non-obese patients with type 2 diabetes; (4) it is important to investigate whether the reduction of EFV could be associated with the improvement of cardiovascular function; however, we were unable to examine the change of parameters related to cardiovascular function, such as diastolic function or pulse wave velocity in this study; (5) we need to mention the absolute reduction of EFV by luseogliflozin. Luseogliflozin reduced EFV by 6 cm³ (5.1% reduction relative to the baseline value) in this study. EFV has been reported to be robustly reduced from 137 ± 37 cm³ to 98 ± 25 cm³ with decreases in BMI (43.1 ± 4.5 kg/m² to 32.3 ± 4.0 kg/m²) or VFA (190 ± 83 cm² to 107 ± 44 cm²) by bariatric surgery in severely obese patients [38]. Therefore, in order to avoid an exaggerated expectation about the clinical use of luseogliflozin, we would like to emphasize that luseogliflozin may have a significant but relatively weak impact on the reduction of EFV as well as that of VFA or body weight compared to bariatric surgery.

In summary, our data suggest that luseogliflozin could reduce EFV in parallel with the improvement of systemic micro-inflammation and the reduction of body weight in Japanese patients with type 2 diabetes. It may be necessary to pay close attention to the potential of SGLT2 inhibitors to decrease the skeletal muscle mass.

Abbreviations

ACEIs: angiotensin converting enzyme inhibitors; ACR: albumin-to-creatinine ratio; ALT: alanine aminotransferase; ARBs: angiotensin receptor blockers; AST: aspartate aminotransferase; BMI: body mass index; CCB: calcium channel blocker; CI: confidence interval; CRP: C-reactive protein; CT: computed tomography; CVD: cardiovascular disease; DBP: diastolic blood pressure; DPP4: dipeptidyl peptidase 4; EF: epicardial fat; EFV: epicardial fat volume; eGFR: estimated glomerular filtration rate; γ-GTP: γ-glutamyl transpeptidase; Glis: glucagon-like peptide 1; HDL: high-density lipoprotein; HOMA-IR: homeostasis model assessment-insulin resistance; IL-6: interleukin 6; IRB: institutional review board; LAI: liver-spleen-attenuation index; LDL: low-density lipoprotein; MRI: magnetic resonance imaging; NAFLD: non-alcoholic fatty liver disease; SFA: subcutaneous fat area; SGLT2: sodium-glucose cotransporter 2; SBP: systolic blood pressure; SMI: skeletal muscle index; TG: triglyceride; TZDs: thiazolidinediones; VFA: visceral fat area.

Authors’ contributions

RB designed the study, researched data, and wrote and edited the manuscript. RM, IM, TY and YO contributed to intellectual discussion and reviewed and edited the manuscript. YS, MA, TF, TT, YN, MM, HI and KH researched data. As the corresponding author and guarantor of this manuscript, RB is the guarantor of this work and, as such, had full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis. All authors read and approved the final manuscript.

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Competing interests
The authors declare that they have no competing interests.

Availability of data and materials
Not applicable. The conclusions of the manuscript are based on relevant data sets available in the manuscript.

Ethics approval and consent to participate, consent for publication
The Institutional Review Board (IRB) at Tokyo Medical and Dental University approved the study (No. 2102), and all patients provided written informed consent before participation.

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