Novel translational strategies in colorectal cancer research

Ignacio Gil-Bazo

Abstract
Defining translational research is still a complex task. In oncology, translational research implies using our basic knowledge learnt from in vitro and in vivo experiments to directly improve diagnostic tools and therapeutic approaches in cancer patients. Moreover, the better understanding of human cancer and its use to design more reliable tumor models and more accurate experimental systems also has to be considered a good example of translational research. The identification and characterization of new molecular markers and the discovery of novel targeted therapies are two main goals in colorectal cancer translational research. However, the straightforward translation of basic research findings, specifically into colorectal cancer treatment and vice versa is still underway. In the present paper, a summarized view of some of the new available approaches on colorectal cancer translational research is provided. Pros and cons are discussed for every approach exposed.

INTRODUCTION TO COLORECTAL CANCER
In the current century, despite the recent achievements in the treatment of advanced colorectal carcinoma (CRC), this tumor remains a major public health concern. In fact, it comprises the third most common cancer type to occur in men and women and was the second leading cause of death among cancer patients in the United States during 2006[1].

Different surgical approaches can guarantee low recurrence rates and high survival expectancy in stages I to III colon neoplasm patients[2]. Furthermore, adjuvant chemotherapy administration has been shown to effectively improve those rates[3]. However, the subset of stage II colon cancer patients to whom adjuvant therapy should be offered is still to be addressed[4]. In fact, different molecular pathology studies and genomic/proteomic investigations are working on that task[5].

In contrast, metastatic colorectal cancer is still far away from being a curable condition and the main goals in the treatment of stage IV colorectal cancer are to decrease tumor-related symptoms or, alternatively, to prolong symptom-free survival with tolerable toxicity[6,7]. However, the emergence of the highly selective therapeutic antibodies bevacizumab and cetuximab has definitely improved the survival of patients with metastatic CRC[8,9]. This fact has intensively boosted the search for other targeted therapies directed to other fundamental molecular checkpoints in colorectal tumorigenesis[10-12].

Thus, due to colorectal cancer clinical and economic relevance, its basic and clinical research has become one of the most funded among all tumor types in most developed countries. However, the straightforward translation of basic research findings into colorectal cancer therapies is still underway.

In the present paper, a summarized view of some of the new available approaches on colorectal cancer translational research is provided.
scientific discoveries must be translated into practical applications. Such discoveries typically start at “the bench” with basic research, in which scientists study disease at a molecular or cellular level\[^{[15-19]}\], and then move on to the clinical level, or the patient’s “bedside”\[^{[20-22]}\]. Scientists are increasingly aware that this bench-to-bedside approach to translational research should really be a two-way highway (Figure 1). Basic scientists provide clinicians with new tools to be used in patients and for assessment of their impact whereas physician-scientists formulate the clinically relevant questions to be tested by basic researchers in a better controlled and more simplified system. Actually, discoveries travel from the clinic to the laboratory in the form of clinical observations, human tissue, diagnostic images, and blood samples, which researchers use to further unlock the molecular and cellular features of cancer (Figure 1).

Often, translational research involves animal studies designed to mimic human conditions\[^{[21-24]}\]. Such studies are generally performed with the same care and scrutiny as the best-planned human clinical trials, and comprise a complex set of supporting laboratory techniques that aim to determine how and why the new diagnostic tool or therapy works or fails in these models. Translational research studies may involve many years of investigation on tools and techniques, to try to estimate how safe and how effective the new treatment or diagnostic procedure will be in human trials.

One of the main scopes of translational research in cancer implies the identification and characterization of molecular markers\[^{[25]}\]. These can be employed as diagnostic and prognostic tools but also for drug responsiveness assessment or even for targeted therapy design. Molecular markers of tumor responsiveness to drugs would help to select the patient populations that would most likely respond to the drug and identify therapeutic indications. Molecular markers of drug activity in normal tissue would allow pharmacodynamic monitoring of patients that could aid optimization of drug dosing and scheduling to maximize patient response\[^{[26]}\]. Furthermore, biological markers involved in tumor initiation and progression can be specifically targeted by new drugs such as therapeutic antibodies\[^{[27]}\] or anti-tumor vaccines\[^{[28]}\].

In fact, another main goal in cancer research is targeted therapy\[^{[22]}\]. Translational research is particularly feasible now because of the new understanding of what causes cancer in different individuals, which relates to different combinations of genetic events. This understanding has come primarily from the work of basic research scientists. Until fairly recently, the only effective armamentaria in cancer therapy were surgery, radiation therapy, and chemotherapy. These treatments generally affect neoplastic cells but also non-cancer tissues, leading to the often serious toxicity that characterizes most of traditional cancer treatments\[^{[29]}\]. While these standard therapies will continue to play an important role in the treatment of patients with cancer, they can be vastly aided in this process by targeted drugs, which literally target the aberrant molecular pathways that are actually involved in tumor initiation and progression. Therefore, specifically delivering the targeted drug to the malignant cell and its closest environment can significantly relieve cancer treatment related collateral effects\[^{[27]}\].

However, since extensive libraries of cytotoxic compounds are being developed for antitumor effect testing, it is becoming more and more common to find new therapies that are successfully developed, tested and commercialized against certain tumors but the ultimate molecular mechanisms involved in tumor response are not clearly known\[^{[29]}\]. In those cases, the translational process is rather directed from clinical findings to basic cellular and molecular experiments (from “the bedside” to “the bench”), trying to unravel the complex pathway in which the new compound is playing a definitive role and the specific target or group of them that results inhibited. Therefore, the bidirectional nature of translational research needs to be emphasized\[^{[29]}\].

IMPLEMENTING TRANSLATIONAL RESEARCH IN COLORECTAL CANCER

There is still a widening gap between basic research and clinical practice, particularly for colorectal cancer. This might be due to the genetic and molecular complexity of this tumor, the lack of the ideal in vivo model for colorectal cancer, and the difficulties found in reproducing animal results into clinical trials in patients.

The principal directions toward which translational research has spread and grown in colorectal cancer in recent years are genomics and proteomics, oncogenic pathways assessment and new targeted therapies discovery (Table 1).

Genomics and proteomics: Searching for new biomarkers and potential target genes
In the last years, there has been an increasingly high effort in the use of genome information in biomedical sciences. This genome information has greatly expanded the insight into the genetic basis of cancer, comprising one of the main fields of interest in translational cancer research. Traditional methods of identifying novel targets involved
in cancer progression were based on studies of individual genes. The following understanding, however, has also shown that gene analysis alone is not sufficient to explain why cancer appears and progresses[31].

Now, the use of DNA microarrays facilitates the analysis of the expression of thousands of genes at the same time and rapidly[32,33]. Microarray analysis has been used for gene expression analysis of different neoplasms[34,35], including CRC[36-39]. However, the application of DNA microarray technology for analysis of CRC is of limited value since it fails to offer direct protein expression measurements[36,38-39]. In addition, it is already known that important pathways in colon tumorigenesis are regulated at the posttranscriptional level where RNA expression data cannot offer any further information. In fact, due to the alternative splicing of both mRNA and proteins, combined with protein posttranslational modifications, one gene can encode a considerable protein population. Actually, the proteome comprises all proteins that result from the whole genome. In contrast to the genome, the proteome is rather a dynamic parameter constituted by proteins and reflects both the intrinsic genetic program of the cell as well as the impact of its surrounding environment.

However, only a few studies have looked for a further insight into the function and/or importance of individual genes and their application to the proteome research of a tumor. Some of these genes have been proposed as candidate cancer biomarkers[40-42]. More recently a number of proteomic studies have also addressed the identification of potential targets in CRC[44,45].

In the proteomics field, several different technical strategies have been developed and applied to CRC translational research over the last years. Each one has its own advantages and drawbacks that should be considered before deciding the experimental design[46].

The technique leading the field for a long time was the two-dimensional polyacrylamide gel electrophoresis (2D-PAGE)[40]. The 2D-PAGE is based on the separation of a gel of the protein content of a sample in two dimensions according to mass and charge. The gels are stained and spots in samples are compared among different gels. However, a number of serious disadvantages such as its lack of real high-throughput capability (one sample per gel) is responsible for having been replaced by more advanced and capable techniques. Similar to 2D-PAGE, the two-dimensional difference gel electrophoresis (DIGE)[44,46] strengthened the 2D platform by allowing the detection and quantization of differences between three samples resolved on the same gel, or across multiple gels, when linked by an internal standard. Again, it also is a low-throughput technology that does not permit the comparison of many samples in a feasible manner.

Other low-throughput proteomic techniques have recently evolved for cancer protein profiling such as liquid chromatography coupled to tandem mass spectrometry detection (LC-MS/MS)[49], isotope-coded affinity tag (ICAT)[50] and a variation of the latter, isotope tags for relative and absolute quantitation (iTRAQ)[51], (both consist of a differential tagging of proteins from samples that are compared using isotope-coded affinity tag in an iso-to-dilution mass spectrometry experiment).

A study conducted by Wu et al[52] has recently compared some of these diverse proteomic strategies (2D-DIGE, ICAT and iTRAQ) on HCT-116 colon epithelial cells concluding that regarding the number of peptides detected for each protein by each method, the global-tagging iTRAQ technique was more sensitive than the cysteine-specific ICAT method, which in turn was as sensitive as, if not more sensitive than, the 2D-DIGE technique.

Nevertheless, as aforementioned, one of the most important goals in protein profiling in oncology is the discovery of new biomarkers[53]. The use of molecular markers in translational research has expanded considerably during the last 3 decades, and this increased analysis of specific molecular changes has been associated with a concomitant decline in the use of more general and less specific histochemical stains and biochemical assays.

Some of the applications for molecular markers include diagnosis, early detection, and prognosis. Also, specific molecular markers are used to study the biology of the disease, to identify targets for novel therapies (e.g., use of Herceptin), and to aid the selection of specific therapies, as previously mentioned.

Therefore, cancer proteomic studies might identify disease-related biomarkers for early cancer diagnosis and new surrogate biomarkers for therapy efficacy and toxicity, but also for guidance of optimal anticancer drug combinations, enabling tailor-made therapy[54]. Furthermore, they could lead to new pharmacological targets. However, a crucial requisite for this purpose is to be able to perform a systematic analysis of a large number of proteins in an easy, reproducible, time-efficient and cost-effective way. High throughput technologies are therefore warranted.

Protein microarrays for instance[55], (targeted proteins bind to spotted probes on a “forward” microarray and specific probes bind to targeted proteins in spotted samples on a “reverse” microarray; bound proteins are detected by direct fluorescent labeling or by labeled secondary antibodies), provide a high throughput approach in terms of number of probes per “forward” array and

| Genomics          | Proteomics                        |
|-------------------|-----------------------------------|
| DNA microarrays   | 2D-PAGE                           |
| DNA microarrays   | DIGE                              |
| DNA microarrays   | LC-MS/MS                          |
| DNA microarrays   | ICAT                              |
| DNA microarrays   | iTRAQ                             |
| Oncogenic pathways| Preclinical models                 |
| AS-ODN            | Min mice                          |
| miRNAs            | Msh2, Msh4, Msh6 deficient mice    |
| siRNAs            | Apc163 8N mice                    |
| siRNAs            | Smad4/Apc mice                    |

Table 1 Translational research technologies in colorectal cancer

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number of samples per ‘reverse’ array with the advantage of previously knowing the biomarker identity. On the other hand, the synthesis of many different probes is necessary, the identity of biomarkers has to be known and cross-reactivity of probes along with possible impaired binding of proteins with post-translational modifications (PTM) exists.

In 2002, the Nobel committee acknowledged the advances in mass spectrometry of biopolymers with the recognition of the discovery of electrospray ionization (ESI) mass spectrometry\cite{56,57} and for the discovery of soft laser desorption (SLD) ionization, which led to the development of matrix-assisted laser desorption ionization (MALDI)\cite{58}. These discoveries for peptides, proteins and other macromolecules have been revolutionary, providing easy measurements of molecular weight with unprecedented accuracy. Because the dominant ions generated under SLD and MALDI conditions are singly charged, the technique is most often used in combination with a time-of-flight (TOF) analyzer to extend the m/\(z\) range to 100000 Da and beyond\cite{58}. MALDI-TOF technology is a highly capable tool allowing the measurement of up to 1536 samples per plate, also possessing access to PTM. On the negative side, this technique is unsuitable for high molecular weight proteins (>100 kDa) and sample fractioning is needed when measuring complex samples.

Surface-enhanced laser desorption ionization time-of-flight (SELDI-TOF) technology is a variant of MALDI-TOF in which a selected part of a protein mixture is bound to a specific chromatographic surface and the rest is washed away\cite{57}. Although SELDI-TOF technology only permits 96 samples to be tested by bioprocessor, fractioning of the sample is not necessary and direct application of the whole sample is possible. However, compared to MALDI-TOF, SELDI-TOF provides lower resolution and mass accuracy but requires smaller amounts of starting material. SELDI-TOF is also unsuitable for proteins heavier than 100 kDa.

SELDI-TOF is equally useful for the analysis of cell lysates from cell lines and tissues\cite{59}, however, in clinical practice its ultimate value derives from its application to easily accessible body fluids as serum or urine. In fact, in the last years several serum biomarker proteins have been identified through this technical approach\cite{60-62}.

In addition, low and high throughput techniques have been shown to be complementary and its combination can lead to a more efficient outcome\cite{60}.

In summary, compared to the genome, the proteome provides a more reliable picture of a biological status and is, thus, expected to be more useful than gene analysis for evaluating, for example, disease presence, progression and response to treatment.

A totally different approach for protein profiling has recently emerged in translational cancer research. To evaluate the clinical significance of newly detected potential cancer genes, it is usually required to examine a high number of well-characterized primary tumors. Using traditional methods of molecular pathology, this was a time consuming job that exploited precious tissue resources. However, a high throughput tissue analysis approach, [tissue microarray (TMA) technology], has been developed\cite{64-66}. Using this TMA technology, samples from up to 1000 different tumors are arrayed in one recipient paraffin block, sections of which can be used for all kinds of in situ analyses\cite{22,67}.

Sections from TMA blocks can then be utilized for the simultaneous analysis of DNA, RNA or protein tumor levels. TMA protein analysis has also been performed in CRC samples for prognostic evaluation\cite{68-72}. However, even though it has been suggested that minute arrayed tissue specimens are representative of their donor tissues, highly heterogeneous cancer types and low levels of protein expression could account for underestimating determined protein expression levels in certain tumors\cite{68-72}.

There are multiple different types of TMAs that can be utilized in cancer research including multi tumor arrays (containing different tumor types), tumor progression arrays (tumors of different stages) and prognostic arrays (tumors with clinical endpoints). The combination of multiple different TMAs allows a very quick but comprehensive characterization of biomarkers of interest.

Despite what proteomics have added to translational research in cancer, there are some novel approaches that combine the information provided by genomic and proteomic assays run in parallel in order to complement the translational impact of both procedures\cite{76}. This has also been applied to CRC profiling. Kwong et al\cite{76}, for instance, studied gene and protein expression performed in parallel across progressive stages of human CRC. For this purpose, they applied cDNA microarray and 2D-PAGE technologies in parallel to analyzed samples collected from 60 CRC cases at various stages of disease progression. Of 47 genes analyzed, 12 (26%) showed significant correlation between mRNA level and protein levels, suggesting that protein abundance is regulated at the transcriptional level. The remaining 31 genes showed either a non-significant correlation between mRNA and protein expression levels or, in 28% of the genes, a negative correlation. Therefore, the authors conclude that posttranscriptional mechanisms play an important role in the regulation of gene products activities in CRC, underline the importance of analyzing gene expression at multiple levels and claim that genomic and proteomic approaches actually complement each other.

In another recent study to identify new biomarkers, Madoz-Gurpide et al\cite{76} investigated the feasibility of expressing soluble proteins corresponding to up-regulated genes in surgically resected CRC samples. They used cDNA microarrays (CNIO Oncochip)\cite{77} to identify differentially expressed genes in malignant compared to normal samples isolated from 22 different CRC patients. After investigating different sources of cDNA clones for protein expression, from 29 selected genes, 21 different proteins were finally expressed soluble with, at least, one distinct fusion protein. Additionally, seven of these potential markers were tested for antibody production and/or validation, confirming six of them to be overexpressed in CRC tissues by immunoblotting and TMA analysis\cite{78}. Authors suggest that this kind of approach may provide relevant
biological information of the neoplastic processes and lead to a better characterization of potentially interesting markers in a quite straightforward way for early diagnosis or individualized prognosis assessment.

**Oncogenic pathways: Validating target candidates**

The previously reviewed development of genomics and proteomics in cancer research has yielded an uncountable number of new potential oncogenic mediators and checkpoints, in CRC, worth further investigating. These novel gene-depending elements, potential new targets for future drugs, are commonly involved in a variety of molecular pathways and their intimate upstream/downstream regulators as well as their crosstalk networks and functional relevance still need to be addressed.

Most widely used experimental methods for molecular pathway research in oncology are performed on fairly well-controlled *in vitro* systems. Recent cell biology achievements and discoveries however, have led to more reliable and physiologically relevant settings where observations on cell behavior and cell fate under particular conditions can be imported into *in vivo* experiments employing animal cancer models and even translating findings into new human therapeutic trials.

In the last few years, several approaches to find molecules able to inhibit the expression of genes (so-called gene-silencing molecules) involved in colorectal cancer progression and therapeutic resistance have been pursued. Sequence-specific gene suppression strategies using antisense oligonucleotides (AS-ODN), ribozymes and deoxyribozymes were initially described and developed. AS-ODN derivatives, depending on their type, recruit RNase H to cleave the target mRNA or inhibit translation by steric hindrance. Ribozymes though, directly bind to RNA via Watson-Crick base pairing and cleave the phosphodiester backbone of the RNA target by transesterification. Similarly, deoxyribozymes also bind to their RNA substrates via Watson-Crick base pairing and specifically cleave the target RNA.

Currently, in addition to their value in target validation studies, different AS-ODN strategies are under evaluation in phase II and III clinical trials, particularly in hematological malignancies, malignant melanoma and prostate cancer. However, consolidating AS-ODN as a broadly applicable functional genomic and therapeutic tool has proven difficult. For instance, difficulties in delivery of the AS-ODN into target tissues, instability of AS-ODN *in vivo*, poor oral availability, uncertainties about the precise mode of action, and toxic effects in animal and human studies have been argued. Moreover, a number of class effects are observed with AS-ODN that are unrelated to the specific targeted mRNA sequence. Acute effects include activation of the alternative complement pathway and inhibition of the intrinsic coagulation pathway. In fact, given repeated doses of AS-ODN to animals, accumulation of AS-ODN and/or metabolites occurs in the form of basophilic granules in various tissues, including the kidney, lymph nodes and liver. Although several approaches are known to overcome some of these difficulties, very few contributions have firmly supported the use of AS-ODN technology in CRC research.

But in the field of gene-silencing molecules, the most recent and fascinating tools discovered for studying gene regulation and gene expression control are microRNAs (miRNAs) and small interfering RNA (siRNAs). miRNAs and siRNAs are typically 21 to 25 nucleotide RNA molecules that induce gene silencing by RNA interference (RNAi) since the description of RNA interference (RNAi) in 1998, this gene-silencing technology has been developed into a widely used methodology in basic as well translational research. RNAi was originally discovered as a naturally occurring pathway in plants and invertebrates. Once long double-stranded RNA molecules are inserted into these organisms, they are processed by the endonuclease Dicer into siRNAs. These siRNAs are subsequently incorporated into the multicomponent RNA-induced silencing complex (RISC), which unwinds the duplex and uses the anti-sense strand as a guide to look for homologous mRNAs and degrade them, as previously reviewed by others. More strikingly, synthetic short siRNAs (20–25 bp) can be either delivered exogenously or expressed endogenously from RNA polymerase II or III promoters (in the form of siRNAs or short hairpin (sh)RNAs that are processed by Dicer into functional siRNAs) and used as a new powerful technology for achieving specific down-regulation of target mRNAs in mechanistic research or even therapeutic development in CRC.

**Testing targeted therapies: Preclinical modeling in colorectal cancer**

Once potential targets are discovered and their expression is successfully inhibited *in vitro*, the safety, efficacy and feasibility of their inhibitors need to be evaluated in animal models in which human disease can be faithfully reproduced. In fact, in the last years, the need of relevant *in vivo* models in colorectal cancer research has prompted many investigators to work on developing reliable, reproducible and human colorectal cancer-mimicking animal models.

However, in colorectal cancer, much has been learned from human inherited syndromes, such as familial adenomatous polyposis (FAP) and hereditary non-polyposis colorectal cancer (HNPCC). That knowledge in fact, has been translated into the design and development of CRC animal models.

Although several rat models have been created for the study of colorectal cancer, in this review, we will focus our attention on mouse models which have profusely evolved in the last few years because of their abundant genetic/genomic information, and easy mutagenesis using transgenic and gene knockout technology. Genetically engineered mice have become essential tools in both mechanistic studies and drug development in CRC, as previously reviewed by others. In fact, mice provide unique opportunities to define and identify genes that are involved in colorectal cancer progression.

The first mouse model obtained to carry a mutation in the adenomatous polyposis coli (APC) tumor suppressor gene was named multiple intestinal neoplasia (Min).
The Min mutation results in a truncated protein and induces the development of multiple intestinal adenomas (even more than one hundred) and a reduced lifespan of on average 150 d in heterozygous mice. Posterior models carrying mutations in different APC alleles have also been developed and each one possesses its own clinical manifestations. However, the majority of them shows small intestine adenomas and colonic tumors and distant metastases are rarely observed. Interestingly, it has been shown that different mutations in the APC gene, in Apc1638N mice for instance, confer distinct tumor susceptibility phenotypes and that fact resembles the heterogeneity observed in human FAP families. Other models of hereditary non-polyposis colorectal cancer (HNPPC) have been developed through the mutation of several mismatch repair genes. One representative example are Msh2 deficient mice that are fertile and develop normally, however, these animals develop T-cell lymphomas early in their life and die because of the disease. Msh2 deficient mice that survive more than 6 months develop gastrointestinal adenomas, carcinomas and skin tumors and can also be used for tumorigenesis studies.

Finally, other more recent models have also been developed to better study colorectal cancer. Smad4 heterozygous mice bearing Apc mutations present an enhanced progression and a more malignant phenotype. Other combinations responsible for increased gastrointestinal tumorigenesis are APC and oncogenic KRAS that seem to be synergistic in enhancing Wnt signaling.

**CONCLUSIONS**

Translational research is a key developing field in biomedical. The direct application of basic research findings to the patient's diagnosis and treatment is even more important in cancer. In addition, clinical observations can dramatically contribute to basic research improvement and relevant enhancement. Colorectal cancer, due to its epidemiological importance and economic impact, is one of the main entities in which translational research is a reality today.

However, there still is a long way to go until basic researchers and clinical investigators share information and work together in colorectal cancer research on a daily basis.

Several new technologies and tools have demonstrated a great value in cancer and are in fact responsible for the last crucial pieces of research work allowing a new conception of cancer diagnosis and treatment. Among them, the development of new biomarkers for colorectal cancer combining proteomics and genomics is especially relevant.

Also, anti-sense strategies have recently opened the path for new target-specific therapy development. These new therapeutic discoveries need to be tested in preclinical animal models.

Since extensive validation of the above mentioned research fields is necessary, adequate funding is required. This may imply some adjustments in the current funding policy because it involves non-innovative studies.

Furthermore, the pool of researchers/clinicians capable of performing translational research must be increased. Additionally, there should be an enhanced participation of patients in clinical trials and an optimization of the efficiency of these trials using validated surrogate markers. Only when these conditions are fulfilled the 'post-genomic' era of biomedical research will have unprecedented opportunities to innovate and improve therapy for cancer.

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