Impact of cow strain and concentrate supplementation on grazing behaviour, milk yield and metabolic state of dairy cows in an organic pasture-based feeding system

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As ruminants are able to digest fibre efficiently and assuming that competition for feed v. food use would intensify in the future, cereals and other field crops should primarily be destined to cover the dietary needs of humans and monogastric animals such as poultry and pigs. Farming systems with a reduced or absent concentrate supplementation, as postulated by organic agriculture associations, require adapted dairy cows. The aim of this experiment was to examine the impact of concentrate supplementation on milk production, grazing and rumination behaviour, feed intake, physical activity and blood traits with two Holstein-Friesian cow strains and to conclude the consequences for sustainable and organic farming. The experiment was a cross-over study and took place on an organic farm in Switzerland. In all, 12 Swiss Holstein-Friesian (HCH) cows and 12 New Zealand Holstein-Friesian (HNZ) cows, which were paired according to lactation number, days in milk and age for primiparous cows, were used. All cows grazed full time and were supplemented either with 6 kg/day of a commercial, organic cereal-grain mix or received no supplement. After an adaptation period of 21 days, a measurement period of 7 days followed, where milk yield and composition, pasture dry matter intake estimated with the n-alkane double-indicator technique, physical activity based on pedometer measurements, grazing behaviour recorded by automatic jaw movement recorder and blood samples were investigated. Non-supplemented cows had a lower milk yield and supplemented HCH cows produced more milk than supplemented HNZ cows. Grazing time and physical activity were greater for non-supplemented cows. Supplementation had no effect on rumination behaviour, but HNZ cows spent longer ruminating compared with HCH cows. Pasture dry matter intake decreased with the concentrate supplementation. Results of blood analysis did not indicate a strong negative energy balance for either non-supplemented or supplemented cows. Minor differences between cow strains in this short-term study indicated that both cow strains are equally suited for an organic, pasture-based production system with no concentrate supplementation. Many factors such as milk yield potential, animal welfare and health, efficiency, grazing behaviour and social aspects influence the decision to supplement grazing dairy cows with concentrates.

Keywords: concentrate supplementation, organic farming, dairy cow, Holstein, pasture

Implications

In the future, competition between feed and food will increase. Ruminants like dairy cows are able to digest forage fibre efficiently. As concentrate supplementation is limited in organic dairy farming, restrictions may cause health problems as energy requirements for high-yielding dairy cows may not be met from forage-only rations. The aim of this study was to verify how supplementation changes the behaviour and production of grazing dairy cows under organic conditions. The two Holstein cow strains investigated in this short-term study are equally suited for an organic, pasture-based farming system with no concentrate supplementation.

Introduction

In Switzerland, the milk yield per cow has increased steadily over the last years (Bundesamt für Landwirtschaft, 2015), as has the cows' demand for nutrients and energy. This leads to the question of whether high-yielding dairy cows are still able to meet their energy requirements for efficient milk production on pasture-only diets. Farming systems with a reduced or...
absent concentrate supplementation, as postulated by organic agriculture associations, require adapted dairy cows. Studies suggest that high producing Holstein-Friesian (HF) cows are less suited for grazing systems, particularly under high stocking rates and seasonal calving systems and without concentrate supplementation (Kolver and Muller, 1998). In New Zealand, HF cows are bred for efficient pasture use with very little concentrate supplementation and seasonal calving (Washburn and Muller, 2014). They differ in BW, average body condition score (BCS) and milk yield from other HF cow strains (Piccand et al., 2013) and have a lower milk production response to concentrate supplementation compared with HF cow strains with a high genetic merit for milk production (Horan et al., 2005). In addition, they may be able to use pasture more efficiently for milk production (Macdonald et al., 2008) and therefore may be better suited for organic milk production systems. According to Penning and Rutter (2004), behaviour exhibited by animals is an indication of the relationship between their internal state (e.g. nutritional requirements, health) and their environment (e.g. sward state, supplementation and climate). Therefore, grazing behaviour such as grazing duration, herbage intake and intake rate may provide some evidence about well-suited dairy cows and should be examined in detail, as the efficient use of pasture is a priority. According to the knowledge of the authors, this is the first study that investigated the effect of concentrate supplementation on the eating and rumination behaviour, metabolic states and milk production of grazing dairy cows under organic farming conditions.

The aim of this study was to examine the milk yield and milk composition, grazing and rumination behaviour, physical activity and metabolic profile of grazing cows with and without concentrate supplementation under organic conditions. Furthermore, this study aimed to identify differences between two HF cow strains, especially on grazing and rumination behaviour, which could indicate a better suitability for pasture-based milk production systems with restricted concentrate supplementation for organic farming.

Material and methods

Animals and experimental design

The experiment was a 2 × 2 cross-over study with two concentrate levels and two cow strains. All experimental procedures were in accordance with the Swiss guidelines for animal welfare and were approved (no. 2012_10 FR) by the Animal Care Committee of the Canton of Fribourg, Switzerland. Before selecting the cows for the experiment, a medical examination was performed. The experiment consisted of two measurement periods, each consisting of a 21-day adaption period and a 7-day measurement period. For the flow of work and equipment reasons, the cows were equally divided into two consecutive data collection periods of 7 days/measurement period. The experiment took place on the organic farm ‘Ferme École de Sorens’, located 824 m above sea level in Sorens, Switzerland.

A total of 24 HF cows, including 12 Swiss Holstein-Friesian (HCH) cows and 12 HF cows of New Zealand origin (HNZ), were used for the experiment. In all, 14 of them were multiparous and 10 were primiparous. Matched pairs of HCH and HNZ cows were formed according to the number of lactation, days in milk (DIM) and age for primiparous cows. The average economic breeding value (ISEL; Swiss Holstein Breeding Association, Posieux, Switzerland) was 985 (SD 74) for HCH cows. The average economic breeding value for HNZ cows was 806 (SD 70), but this excluded four animals as no breeding value was available.

At the start of the first data collection period, HCH cows had an average number of lactations of 2.3 (SD 1.6), had been 109 (SD 17.9) DIM, had an average BW of 609 (SD 90.1) kg, a BCS of 2.4 (SD 0.28) and were producing 27.6 (SD 3.77) kg milk/day. The HNZ cows had an average number of lactations of 2.7 (SD 2.0), had been 114 (SD 16.7) DIM, had an average BW of 560 (SD 72.3) kg, a BCS of 2.9 (SD 0.24) and were producing 24.1 (SD 5.27) kg milk/day.

Grazing management, concentrate supplementation and weather conditions

The experiment was carried out in a rotational grazing system from 7 May to 8 July 2012. All 24 experimental cows were managed as a single group separated from the rest of the lactating herd and grazed on pasture from 0800 to 1400 h and from 1800 to 0430 h the following morning. Meanwhile, cows were milked and housed in a free-stall barn. Paddocks were rotationally grazed for 2 to 5 days based on decision rules considering sward height with a reference of 130-mm pre-grazing equivalent to an herbage mass of ~1000 kg dry matter (DM)/ha above 50 mm until a post-grazing sward surface height of 50 mm from ground level. The sward surface height was measured with a pasture meter (C-Dax pasture meter; C-Dax Ltd, Turitea, New Zealand) before cows entered a new paddock and after leaving the paddock. The average pre-grazing sward height was 129 (SD 16.4; n = 5) mm, corresponding to 1024 (SD 209.3) kg DM/ha above 50 mm in the first measurement period and 122 (SD 11.2; n = 6) mm, corresponding to 941 (SD 143.8) kg DM/ha for the second measurement period. The average post-grazing sward surface height was 57 (SD 6.6; n = 5) mm in the first measurement period and 71 (SD 19.2; n = 6) mm in the second measurement period. Herbage mass above 50 mm (kg DM/ha) was calculated according to: −624.5 + 12.8 × sward height (mm). This regression was calibrated for the pastures of the organic farm ‘Ferme École de Sorens’ (R² = 0.84; n = 89). The pastures were long established and composed predominantly of grasses (mainly Lolium perenne, Dactylis glomerata and Phleum pratense), but also of clover (mainly Trifolium repens) and herbs (mainly Taraxacum officinale). The pastures were fertilized once per year with 25 m³/ha of farm-produced manure (corresponding approximately to 80 kg N/ha, 22 kg P/ha and 108 kg K/ha). The chemical composition of the pasture during the measurement periods is presented in Table 1.

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During an adaptation period of 21 days before both measurement periods, a step-wise provision towards the targeted amount of concentrate (UFA 275 Bio; UFA AG, Herzogenbuchsee, Switzerland), 0 or 6 kg (as-fed basis), took place. The pelleted concentrate was offered to six HCH–HNZ cow pairs in two equal meals (3 kg at 0600 h and 3 kg at 1700 h) after milking in the free-stall barn using separate buckets for each cow. During the measurement period, all 6 kg of concentrate was ingested by all cows with no refusals. The other six HCH–HNZ cow pairs received no concentrate in addition to pasture. Fresh water was always available and a mineral block was available in the barn. The ambient outdoor temperature was recorded daily by the meteorological station in Grangeneuve (MeteoSchweiz, Station Grangeneuve, Switzerland), located about 15 km north of the experimental pastures. During the first measurement period, the average temperature was 16°C (minimum 13, maximum 19) and 19°C (minimum 15, maximum 24) in the second measurement period.

Data recording and sample collection
Milk yield (Flo-Master Pro; DeLaval AG, Sursee, Switzerland) was recorded daily and milk composition was analysed from a pooled sample of the morning and evening milk on days 1, 4 and 7 of each data collection period. Milk samples were preserved in tubes containing Broad Spectrum Microtabs II (Gerber Instruments AG, Effretikon, Switzerland) at 8°C. The BW was recorded twice daily after milking and BCS was assessed before each adaptation period and before and after each data collection period according to the five-point system of Edmonson et al. (1989).

To estimate individual feed intake on pasture, the n-alkane double-indicator technique was used (Mayes et al., 1986). Gelatin capsules (HGK 17–60 sl; Capsula GmbH, Ratingen, Germany), containing 0.5 g (weighing accuracy 0.001) alkane marker HC32 (dotriacontane, C32H66; Argenta Ltd, Auckland, New Zealand) on a carrier of dried fruit pomace, were administered manually with an applicator twice per day starting 6 days before the data collection periods. During the data collection period, a daily spot sample of faeces was taken from each cow with or without stimulus between 0700 and 0800 h. Samples were pooled by cow and collection period and stored at −20°C. Collection of pasture samples started and ended 1 day before the faeces sampling. Pasture sample collection was carried out as described by Graf et al. (2005). Daily samples were chopped and stored at −20°C until further analysis. Samples of concentrate were taken daily and pooled per data collection period.

Grazing and rumination behaviour was recorded automatically using a jaw movement recorder with a pressure sensor (Datenlogger MSR145; MSR Electronics GmbH, Hengart, Switzerland; Nydegger et al., 2011). The jaw movement frequency and amplitude were measured for 72 h. Data were evaluated with the software programs MSR-Reader (MSR 5.20.01) and MSR-Viewer (Viewer V2), as described in the MSR145 User Manual (MSR Electronics GmbH). The number of grazing and rumination bouts were counted manually from the evaluated output of the software.

Table 1 Average chemical composition of concentrate (n = 2) and pasture (n = 14) samples (mean ± SD).

| Items                              | Concentrate | Pasture |
|------------------------------------|-------------|---------|
| DM (g/kg of wet weight)            | 884 ± 1.8   | 174 ± 30.3 |
| Analysed nutrients and mineral composition (g/kg of DM) | | |
| OM                                 | 944 ± 0.3   | 894 ± 8.6 |
| CP                                 | 115 ± 1.3   | 174 ± 23.0 |
| Ether extract                       | 59 ± 0.1    | 47 ± 5.3 |
| Starch                             | 502 ± 0.7   | 47 ± 5.3 |
| ADF                                | 79 ± 2.2    | 249 ± 20.8 |
| NDF                                | 221 ± 5.7   | 397 ± 41.8 |
| Crude fibre                         | 55 ± 1.6    | 199 ± 25.4 |
| Ca                                 | 8.4 ± 0.11  | 8.1 ± 1.40 |
| P                                  | 6.2 ± 0.03  | 5.1 ± 0.46 |
| Mg                                 | 3.2 ± 0.01  | 2.1 ± 0.23 |
| Na                                 | 1.8 ± 0.03  | 0.2 ± 0.05 |
| K                                  | 7.9 ± 0.05  | 38 ± 3.4 |
| Calculated energy and protein supply1 per kg of DM | | |
| NEL (MJ)                           | 8.1 ± 0.02  | 6.3 ± 0.24 |
| APDE (g)                           | 79 ± 2.2    | 104 ± 5.1 |
| Analysed n-alkane contents (mg/kg of DM) | | |
| HC32                               | 0.5 ± 0.50  | 5.0 ± 0.66 |
| HC33                               | 2.6 ± 0.24  | 59 ± 10.0 |

**DM = dry matter; OM = organic matter; NEL = net energy for lactation; APDE = absorbable protein in the small intestine when rumen fermentable energy is limiting microbial protein synthesis in the rumen; HC32 = dotriacontane, C32H66; HC33 = tritriacontane, C33H68.**

1According to Agroscope (2013).
MSR-Viewer. A bout was defined as a sequence of a behaviour not interrupted by any other element of behaviour with a duration >4 min (Metz, 1975) and an inter-bout interval >7.5 min (Dado and Allen, 1994). Grazing bouts contain only those bouts during grazing on pasture not considering bouts when concentration was eaten in the barn.

Physical activity, including time spent standing, lying and walking was determined using the IceTag™ pedometer (IceRobotics Ltd, Edinburgh, Scotland, UK). The pedometer was attached to the right hind leg of the cow at the metatarsus level and recorded acceleration in three dimensions at 0.1 s intervals for 72 h. Using the software program IceTag-Analyser (V 4.005; IceRobotics Ltd), the data were downloaded and compiled over 60 s intervals. Walking was defined as >3 steps/min, as suggested by Thanner et al. (2014).

On days 4 and 5 of each data collection period, blood was collected at 0700 and 1400 h by puncture of the jugular vein, using the Vacuette® System (Greiner Bio-One GmbH, Kremsmünster, Austria). Plasma for the analysis of hormones was retrieved using Vacuette® EDTA tubes (Greiner Bio-One GmbH). After sampling, these tubes were cooled in ice water until they were centrifuged at room temperature (20°C) for 15 min at 3000×g. For analyses of blood metabolites and enzymes, Vacuette® serum tubes (Greiner Bio-One GmbH) were stored upside down for 1 h at room temperature (20°C) before centrifugation at 3000×g for 15 min and then at 4000×g for an additional 5 min (Thanner et al., 2014). The retrieved serum and plasma samples were stored at −20°C until they were analysed for hormones, metabolites and enzymes.

**Laboratory analysis**

Milk samples were analysed by IR spectrometry (CombiFoss FT+; Foss, Hillerød, Denmark) for contents of fat, protein and lactose (International Dairy Federation, 2000; method number 141C). Urea was determined with a differential pH analyser (Eurochem, Ardea, Italy) before and after hydrolysis with urease (International Dairy Federation, 2004; method number 195). For milk acetone determination, acetone and 3,5,3’-triiodothyronine (T3) and thyroxin (T4) were measured with urease (no. 61218; bioMérieux, Marcy-l’Etoile, France), aspartate aminotransferase (ASAT, no. 63212; bioMérieux), β-hydroxybutyrate (BHBA; no. RB1007; Randox Laboratories, Crumlin, UK), cholesterol (no. 61218; bioMérieux), creatinine (no. 11489291216; Roche Diagnostics), creatine kinase (CK, no. 61141; bioMérieux), γ-glutamyltransferase (GGT, no. 2016788; Roche Diagnostics), glutamate dehydrogenase (GLDH, no. 1929992; Roche Diagnostics), total protein (no. 1553836; Roche Diagnostics), urea (no. 61974, UV 250; bioMérieux), triglyceride (no. 61236; bioMérieux), non-esterified fatty acids (NEFA, no. FA 115; Randox Laboratories) and glucose (no. 1447513; Roche Diagnostics).

Plasma insulin and IGF-1 concentrations were quantified using radioimmunoassay as described by Vicari et al. (2008). Plasma insulin and IGF-1 concentrations were quantified using radioimmunoassay as described by Vicari et al. (2008).

Calculations and statistical analyses

Net energy for lactation (NEL) and the absorbable protein in the small intestine when rumen fermentable energy is limiting microbial protein synthesis in the rumen were calculated according to Agroscope (2013). The energy-corrected milk yield (ECM) was calculated based on a 4% fat, 3.2% protein and 4.8% lactose according to Agroscope (2013). Feed intake was calculated using the ratio of the n-alkanes HC32 and HC33 (tritriacontane, C33H66) were determined as described by Peiretti et al. (2006). The N content was determined using the Dumas method (Association of Official Analytical Chemists (AOAC), 1995) on a CN analyser (type FP-2000; Leco Instruments, St. Joseph, MI, USA) and then multiplied by 6.25 to determine the CP content. The ether extract was determined using the Soxtec Avanti 2050 apparatus (Foss, Hillerød, Denmark) for extraction following the guidelines of Verband Deutscher Landwirtschaftlicher Untersuchungs- und Forschungsanstalten 5.1.1. (1993). Acid-detergent fibre (procedure 973.18; AOAC, 1995) was determined with correction for residual ash obtained after incineration at 500°C for 1 h. Crude fibre was analysed in pasture and concentrate (procedure 978.10; AOAC, 1995) and NDF (Mertens, 2002) was evaluated with the addition of heat-stable amylase and sodium sulphite. Starch content was determined based on the polarimetric method (method 6493; International Organization for Standardization, 2000).

Metabolite concentrations and enzyme activities were determined using the following commercial test kits: albumin (no. 11970909; Roche Diagnostics, Rotkreuz, Switzerland), alkaline phosphatase (AP), alanine aminotransferase (ALAT), β-Mérieux, Marcy-l’Etoile, France), aspartate aminotransferase (ASAT, no. 63212; bioMérieux), γ-glutamyltransferase (GGT, no. 2016788; Roche Diagnostics), creatine kinase (CK), cholesterol (no. 61218; bioMérieux), creatinine (no. 11489291216; Roche Diagnostics), total protein (no. 1553836; Roche Diagnostics), urea (no. 61974, UV 250; bioMérieux), triglyceride (no. 61236; bioMérieux), non-esterified fatty acids (NEFA, no. FA 115; Randox Laboratories) and glucose (no. 1447513; Roche Diagnostics).

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considered, as a period of 1 week is too short to estimate these accurately:

\[ DMI_{\text{Conco}} = (0.293 \times BW^{0.75} + 3.14 \times ECM) / \text{NEL pasture}, \]

\[ DMI_{\text{Conc6}} = (0.293 \times BW^{0.75} + 3.14 \times ECM) / \text{NEL pasture} \]

where BW^{0.75} is the metabolic body size (kg BW to the power 0.75), ECM (kg/d) the energy-corrected milk yield, NEL Conc the NEL content of the concentrate (MJ/kg DM) and NEL pasture the NEL content of the pasture (MJ/kg DM).

The statistical analyses were carried out with SYSTAT 13 (Systat Software Inc., Chicago, IL, USA). Data for milk yield and composition, rumination and grazing behaviour, physical activity, feed intake and blood traits were collected over several days and averaged per cow and measurement period. They were analysed using following linear mixed model:

\[ Y_{ijklm} = \mu + \tau_i + \phi_j + P_k + (\tau \phi)_{ik} + (\tau p)_{ij} + P_l + K_m(P_l) + e_{ijklm} \]

where \( Y_{ijklm} \) is the response, \( \mu \) the least-squares mean, \( \tau_i \) the fixed effect of cow strain \( i \) (HCH, HNZ), \( \phi_j \) the fixed effect of the treatment \( j \) (Conc0, Conc6), \( P_k \) the fixed effect of the period \( k \) (period 1, period 2), \( (\tau \phi)_{ik} \) the effect of the interaction between cow strain \( i \) and period \( k \), \( (\tau p)_{ij} \) the effect of the interaction between cow strain \( i \) and treatment \( j \), \( P_l \) the random effect of cow pair \( l \) (1, …, 12), \( K_m \) the random effect of the cow \( m \) (1, …, 24) and \( e_{ijklm} \) the random error. Models of this type were recommended by Tempelman (2004) with variance components as variance–covariance structure for the repeated measurements. As there were only two periods no alternative variance–covariance structure was envisaged in this study. Not normally distributed data were either logarithmically transformed to fit with normal distribution (NEFA and creatinine kinase) or analysed using R (R Core Team, 2012) with permutation tests for linear models (lactose; Good, 2005). Data presented in the tables were back transformed.

Due to the high proportion of insulin ‘non-detects’ (i.e. values below 3 \( \mu \text{U/ml} \)), descriptive statistics was performed as described by Helsel (2012) using R (R Core Team, 2012). Inferential statistics were based on rank methods (Brunner et al., 2002). The model was of F1-LD-F2 type (one between-subject factor, two within-subject factors) and ANOVA-type test statistics (Brunner et al., 2002) were applied.

The effects were considered significant at \( P \leq 0.05 \). A value of 0.05 < \( P \leq 0.10 \) was considered a trend.

### Results

**Milk yield and milk composition and feed intake**

Non-supplemented cows produced less (\( P < 0.001 \)) milk and less (\( P < 0.001 \)) ECM compared with supplemented cows and HNZ cows had a lower (\( P = 0.04 \)) milk yield compared with HCH cows, but no difference between cow strains was observed for ECM (Table 2). An interaction (\( P = 0.02 \)) between cow strain and supplementation was reported for milk yield. The HNZ cows produced less (\( P = 0.04 \)) milk per kg concentrate than HCH cows.

### Table 2. Effect of concentrate supplementation and cow strain and their interactions on milk production performance and feed intake

| Items                        | Conc0    | Conc6    | SD | Treatment | Cow strain | Interaction |
|------------------------------|----------|----------|----|-----------|------------|-------------|
| Milk production performance  |          |          |    | HCH       | HNZ        |             |
| Milk yield (kg/d)            | 24.9\(^A\) | 23.0\(^B\) | 30.0\(^C\) | 26.0\(^D\) | 4.8        |             |
| Milk yield concentrate \(^1\) (kg/kg) | 0.84\(^a\) | 0.50\(^b\) | 0.30        |             |             |             |
| ECM \(^2\) (kg/d)           | 22.8\(^A\) | 22.6\(^A\) | 25.3\(^B\) | 23.9\(^B\) | 3.8        | <0.0001     |
| Fat (%)                      | 3.7\(^A\) | 4.0\(^B\) | 3.0\(^B\) | 3.4\(^B\) | 0.6        | <0.0001     |
| Protein (%)                  | 3.2\(^A\) | 3.4\(^A\) | 3.2\(^A\) | 3.5\(^B\) | 0.3        | 0.13        |
| Lactose \(^3\) (%)          | 4.4\(^A\) | 4.4\(^A\) | 4.5\(^A\) | 4.5\(^A\) | 0.3        | <0.01       |
| Acetone (mg/l)               | 2.87\(^A\) | 2.53\(^A\) | 1.52\(^B\) | 1.36\(^B\) | 0.79       | <0.001      |
| Urea (mg/l)                  | 239\(^A\) | 226\(^B\) | 178\(^B\) | 176\(^B\) | 30.0       | <0.001      |
| Feed intake                  |          |          |    | HCH       | HNZ        |             |
| Pasture DMI (kg DM/day)      | 12.5\(^A\) | 11.7\(^A\) | 9.2\(^B\) | 9.0\(^B\) | 1.7        | <0.001      |
| Total DMI (kg DM/day)        | 12.5\(^A\) | 11.7\(^A\) | 15.0\(^B\) | 14.3\(^B\) | 1.7        | <0.001      |
| Calculated pasture DMI \(^4\) (kg DM/day) | 17.3\(^A\) | 16.9\(^A\) | 12.0\(^B\) | 11.0\(^B\) | 2.4        | <0.001      |
| Calculated total DMI \(^5\) (kg DM/day) | 17.3 | 16.9 | 17.3 | 16.3 | 2.4 | 0.23 | 0.23 | 0.17 |

Conc0 = cows without concentrate supplementation; Conc6 = cows with concentrate supplementation; HCH = Swiss Holstein-Friesian; HNZ = New Zealand Holstein-Friesian; DMI = dry matter intake; DM = dry matter.

\(^A\)Means with different superscript letters within the same row differ (\( P < 0.01 \)).

\(^B\)Means with different superscript letters within the same row differ (\( P < 0.05 \)).

\(^1\)Milk yield per ingested concentrate (kg/kg).

\(^2\)ECM = energy-corrected milk yield (Agroscope, 2013).

\(^3\)Log_{10} transformed for statistical analyses.

\(^4\)According to Agroscope (2013) and Baker (2004) without BW changes and activity.

\(^5\)According to Agroscope (2013) and Baker (2004) without BW changes and activity plus 5.3 kg DM of concentrate.

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Grazing and rumination behaviour and physical activity

Grazing time, mastication (n) and mastication rate (n/min) were greater \((P<0.001)\) for non-supplemented cows compared with supplemented cows and no differences occurred between cow strains (Table 3). The number of grazing bouts was not affected by supplementation, but HNZ cows tended to have greater numbers compared with HCH cows. Non-supplemented cows had a greater \((P<0.001)\) duration of eating bouts compared with supplemented cows, but no difference between cow strains was reported.

Concentrate supplementation had no effect on traits describing rumination behaviour, but HNZ cows spent more \((P<0.005)\) time ruminating, had a greater \((P<0.005)\) number of mastication and a greater \((P<0.005)\) number of bolus compared with HCH cows. No difference between cow strains was observed for mastication rate, mastication per bolus and rumination bouts. New Zealand Holstein cows tended to have longer duration of rumination bouts compared with HCH cows.

Non-supplemented cows spent less \((P<0.001)\) time lying down, but stood, moved and walked more \((P<0.001)\). There was a trend recorded for HNZ to stand and move less and lie down more, but no difference was observed for walking. No significant interaction of concentrate supplementation and cow strain was observed.

Blood traits

Non-supplemented cows had lower \((P<0.01)\) serum glucose concentration but greater \((P<0.001)\) concentration of serum BHBA, NEFA and urea compared with supplemented cows (Table 4). Concentration of total protein in serum indicated a tendency to be affected by supplementation, with a greater concentration for non-supplemented cows. Concentrate supplementation had no effect on concentration of serum albumin, triglycerides, cholesterol and creatinine. The activity of creatinine kinase, GLDH and GGT was not affected by concentration but greater \((P<0.001)\).

### Table 3 Effect of concentrate supplementation and cow strain and their interaction on grazing and rumination behaviour and physical activity over 24 h

| Items                              | Conc0 | Conc6 | P-value |
|------------------------------------|-------|-------|---------|
|                                    | HCH   | HNZ   | SD      | Treatment | Cow strain | Interaction |
| Grazing behaviour over 24 h        |       |       |         |           |            |             |
| Time (min)                         | 547.1\(^a\) | 568.7\(^b\) | 442.9\(^b\) | 453.0\(^b\) | 43.9 | <0.001 | 0.20 | 0.63 |
| Mastications (n)                   | 41.232\(^a\) | 42.102\(^b\) | 32.070\(^a\) | 32.865\(^b\) | 3859 | <0.001 | 0.51 | 0.97 |
| Mastication rate (n/min)           | 75.3\(^a\) | 74.0\(^b\) | 72.5\(^b\) | 72.7\(^b\) | 4.0 | <0.001 | 0.73 | 0.06 |
| Grazing bouts (n)                  | 5.2 | 5.7 | 5.5 | 6.0 | 0.94 | 0.24 | 0.10 | 0.81 |
| Duration of grazing bouts (min)    | 110\(^a\) | 104\(^b\) | 81\(^b\) | 78\(^b\) | 18.8 | <0.001 | 0.42 | 0.77 |
| Ruminating behaviour over 24 h     |       |       |         |           |            |             |
| Time (min)                         | 381\(^a\) | 405\(^b\) | 398\(^b\) | 423\(^b\) | 40.8 | 0.15 | <0.05 | 0.99 |
| Mastications (n)                   | 27.661\(^a\) | 29.796\(^b\) | 28.888\(^a\) | 31.204\(^b\) | 3932 | 0.20 | 0.04 | 0.93 |
| Mastication rate (n/min)           | 72.4 | 73.4 | 72.7 | 73.7 | 5.3 | 0.47 | 0.48 | 0.92 |
| Ruminations bolus (n)              | 522\(^a\) | 578\(^b\) | 551\(^a\) | 597\(^b\) | 69.2 | 0.22 | 0.04 | 0.79 |
| Mastications bolus (n/boli)        | 51.5 | 53.8 | 54.9 | 53.4 | 5.0 | 0.26 | 0.83 | 0.17 |
| Ruminations bouts (n)              | 13.1 | 12.2 | 13.1 | 12.7 | 1.58 | 0.56 | 0.14 | 0.48 |
| Duration of rumination bouts (min) | 29 | 35 | 31 | 34 | 7.0 | 0.95 | 0.08 | 0.40 |
| Activity over 24 h                 |       |       |         |           |            |             |
| Lying (min)                        | 492\(^a\) | 541\(^a\) | 564\(^b\) | 590\(^b\) | 62 | <0.001 | 0.05 | 0.41 |
| Standing + moving (min)            | 949\(^a\) | 900\(^a\) | 877\(^b\) | 851\(^b\) | 62 | <0.001 | 0.05 | 0.41 |
| Walking (min)                      | 421\(^a\) | 419\(^a\) | 373\(^b\) | 362\(^b\) | 59 | <0.001 | 0.75 | 0.73 |

Conc0 = cows without concentrate supplementation; Conc6 = cows with concentrate supplementation; HCH = Swiss Holstein-Friesian; HNZ = New Zealand Holstein-Friesian.

\(^{a,b}\)Means with different superscript letters within the same row differ \((P<0.01)\).

\(^{a,b}\)Means with different superscript letters within the same row differ \((P<0.05)\).
Glucose (mmol/l) 3.15 A 3.31 B 3.25 C 3.46 D 0.18

BHBA (mmol/l) 0.91 A 0.82 B 0.68 B 0.69 B 0.17

NEFA (mmol/l) 0.12 A 0.14 B 0.08 B 0.09 B 0.05

Urea (mmol/l) 4.86 A 4.77 B 3.68 B 3.73 B 0.90

Total protein (g/l) 73.6 72.9 72.0 72.2 4.20

Albumin (g/l) 38.8 39.2 38.0 38.9 2.40

Triglycerides (mmol/l) 0.30 0.33 0.30 0.31 0.08

Cholesterol (mmol/l) 6.28 6.26 6.08 6.43 1.10

Albumin (g/l) 38.8 39.2 38.0 38.9 2.40

Total protein (g/l) 73.6 72.9 72.0 72.2 4.20

Urea (mmol/l) 4.86 A 4.77 B 3.68 B 3.73 B 0.90

Table 4: Effect of concentrate supplementation and cow strain and their interaction on blood metabolites, enzymes and hormones

| Items            | HCH0 | HNZ0 | HCH6 | HNZ6 | SD    | Treatment | Cow strain | Interaction |
|------------------|------|------|------|------|-------|-----------|------------|-------------|
| Glucose (mmol/l) | 3.15 | 3.31 | 3.25 | 3.46 | 0.18  | <0.01     | <0.01      | 0.54        |
| BHBA (mmol/l)    | 0.91 | 0.82 | 0.68 | 0.69 | 0.17  | <0.001    | 0.45       | 0.30        |
| NEFA (mmol/l)    | 0.12 | 0.14 | 0.08 | 0.09 | 0.05  | <0.001    | 0.48       | 0.62        |
| Urea (mmol/l)    | 4.86 | 4.77 | 3.68 | 3.73 | 0.90  | <0.001    | 0.95       | 0.72        |
| Total protein (g/l) | 73.6 | 72.9 | 72.0 | 72.2 | 4.20  | <0.10     | 0.88       | 0.48        |
| Albumin (g/l)    | 38.8 | 39.2 | 38.0 | 38.9 | 2.40  | 0.27      | 0.32       | 0.56        |
| Triglycerides (mmol/l) | 0.30 | 0.33 | 0.30 | 0.31 | 0.08  | <0.001    | 0.28       | 0.74        |
| Cholesterol (mmol/l) | 6.28 | 6.26 | 6.08 | 6.43 | 1.10  | 0.94      | 0.70       | 0.24        |
| Albumin (g/l)    | 38.8 | 39.2 | 38.0 | 38.9 | 2.40  | 0.66      | 0.07       | 0.19        |
| Total protein (g/l) | 73.6 | 72.9 | 72.0 | 72.2 | 4.20  | 0.26      | 0.13       | 0.79        |
| Urea (mmol/l)    | 4.86 | 4.77 | 3.68 | 3.73 | 0.90  | <0.001    | 0.17       | 0.86        |

Conc0 = cows without concentrate supplementation; Conc6 = cows with concentrate supplementation; HCH = Swiss Holstein-Friesian; HNZ = New Zealand Holstein-Friesian; NEFA = non-esterified fatty acids; GLDH = glutamate dehydrogenase; GGT = γ-glutamyltransferase; ASAT = aspartate aminotransferase; AP = alkaline phosphatase; ALAT = alanine aminotransferase; T3 = 3,5,3’-triiodothyronine; T4 = thyroxin.

1Means with different superscript letters within the same row differ (P < 0.01).

Effect of concentrate and consequences for pasture-based organic farming

In accordance with Bargo et al. (2003), milk yield and ECM increased for supplemented cows, but to a smaller extent. The observed interaction between concentrate supplementation and cow strain for milk production indicate the different genetic potential for milk production between the two cow strains (Bargo et al., 2002; Horan et al., 2005). Supplemented HCH cows responded to the extra energy...
supply with greater milk yield compared with HNZ cows, but ECM was similar.

The reduced milk fat content of supplemented cows is in agreement with other studies, when cows received >4 kg/day of concentrate (Bargo et al., 2002; Horan et al., 2005). Furthermore, the increased activity of plasma AP in supplemented cows may indicate acidotic stress. To ensure rumen health, rumination is a key factor as it influences salivation and therefore rumen buffering. In the current study, concentrate supplementation had no effect on rumination behaviour. It would have been expected that with increasing pasture DMI, time ruminating, rumination mastication and number of boli increased (Beauchemin and Rode, 1997; McCarthy et al., 2007b). The absence of differences in rumination mastication per bolus and number of boli may indicate a similar ease of bolus formation and swallowing for both supplemented and non-supplemented cows, although higher grain diets reflect a greater ease of bolus formation (Beauchemin and Rode, 1997). Rumination mastication per bolus was in the normal range. This poses the question of whether characteristics of rumination behaviour are a suitable indicator of sufficient fibre supply and therefore rumen health, at least in grazing dairy cows.

In agreement with other studies, supplementation caused a substantial reduction in grazing time (Bargo et al., 2002; McCarthy et al., 2007b) and therefore reduced grazing mastication and grazing mastication rate. This implies the lower motivation of supplemented cows to graze and is supported by the reduced duration of grazing bouts for supplemented cows, as grazing time or grazing time coupled with intake rate (bite rate and size) at the same BW might be indicators for the feeding drive (McCarthy et al., 2007b; Prendiville et al., 2010).

In accordance with other studies, pasture DMI decreased with concentrate supplementation, which is expressed as substitution rate (Bargo et al., 2002; McCarthy et al., 2007b). Both calculated and estimated (using n-alkanes) pasture DMI were lowered by concentrate supplementation. In contrast, total DMI of supplemented cows was greater or at least the same as the total DMI of non-supplemented cows. However, with reduced pasture DMI for supplemented cows, cheap pasture is substituted by expensive concentrate which is an economic aspect for the farmer. Furthermore, importing ingredients of a commercial organic concentrate mix disturbs the holistic approach of organic guidelines. In the current study, the energy:protein ratio may be balanced as concentrations of urea in blood and milk were in the normal range, with elevated values for non-supplemented cows. In organic farming, the risk of excessive protein intake from pasture might be reduced as the CP content of organic pasture is lower compared with conventional pasture (Spann et al., 2007).

Current results indicate a strong relationship between physical activity and grazing behaviour. As supplemented cows spent less time grazing, they spent less time standing and walking. In a pasture-based feeding system, where energy might be the first limiting nutrient, physical activity on pasture is an important factor to be considered. Grazing cows have a greater energy expenditure compared with cows fed indoors, as grazing cows take more steps, spend less time lying down and spend more time eating (Kaufmann et al., 2011). According to the Commonwealth Scientific and Industrial Research Organisation (2007), energy requirements for maintenance may increase in the range of 10% to 50% depending on grazing conditions, digestibility of pasture, distance walked, weather, topography and interactions between these factors. Thus, supplemented cows on pasture did not only ingest more energy, but also presented improved energy balance due to energy savings in relation to shorter grazing and physical activities.

A lack of energy in early lactation can cause metabolic problems such as ketosis. In the current study, the increased acetone concentration in milk for non-supplemented cows indicates a small risk of ketosis. This is confirmed by an increased concentration of BHBA and NEFA and decreased glucose and insulin concentration for non-supplemented cows. The increased blood glucose, insulin and IGF-1 concentrations for supplemented cows indicate an increased energy status, whereas increased concentration of blood NEFA and BHBA for non-supplemented cows suggest a lower energy status (Reist et al., 2002). As the concentration of T3 and T4 decreases with stronger negative energy balance (NEB) (Huszeniczcz et al., 2002), non-supplemented cows in current study had lower concentration of T3 and T4 and therefore had a lower energy status compared with supplemented cows. Furthermore, the increased activity of ALAT of non-supplemented cows indicates an increased use of amino acids as an energy source or for gluconeogenesis (Garber et al., 1976). The greater activity of ASAT of non-supplemented cows alone should be interpreted with caution. As the activities of GLDH, GGT and CK were not increased for non-supplemented cows, the occurrence of fatty liver syndrome in cows of this study is unlikely.

Results of blood traits indicate that non-supplemented cows were not in strong NEB. Supplementation may not be necessary to balance an organic pasture-based diet, at least for cows in mid-lactation and therefore more severe, but more flexible restrictions over the whole lactation for supplementation are favourable.

Finally, it can be stated that the effects of concentrate supplementation on milk yield, milk composition, grazing behaviour and intake are similar in the present organic study compared with the cited conventional studies.

Effect of cow strain and consequences for pasture-based organic farming

Another aspect of organic farming is the choice of the breed or strain. This implies the selection of cows adapted to low-input pasture or forage-based feeding system for organic dairy production. In the present study, the milk yield responses obtained for HNZ and HCH cows were similar to the results of Horan et al. (2005). Swiss Holstein cows reached almost the overall milk yield response of 1 kg milk/1 kg concentrate as published by Bargo et al. (2003), but not in ECM terms. The lower genetic potential for milk

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production of HNZ cows might be the reason for their lower response of 0.5 kg milk/kg of ingested concentrate, as pasture allowance and mass was the same for both strains. Cows with high genetic potential for milk yield have a greater milk yield response to concentrate supplementation (McCarthy et al., 2007b). This phenomenon might be attributed to greater nutrient partition to milk production in high genetic merit cows compared with lower genetic merit cows (Dillon et al., 2006). This is supported by the results of Rook (2000), with greater rumination mastication per day compared with HCH cows. Anatomical differences of the muzzle and incisor breadth might explain this (Rook, 2000). Prendiville et al. (2010) observed smaller bolus size for the smaller Jersey cows compared with HF cows, indicating that anatomical differences influence the pattern of bolus movement during rumination. Although greater chewing activity during grazing and rumination is associated with a greater salivary secretion and therefore a better digestibility (Domingue et al., 1991), HNZ cows could not benefit from longer rumination time in terms of milk production.

The trend for lower pasture DMI by HNZ cows and the lack of clear differences in grazing behaviour do not confirm a greater feeding drive for HNZ cows (McCarthy et al., 2007b). It could have been expected that HCH cows had a greater pasture DMI, as DMI and BW are positively correlated (Kertz et al., 1991), as BW is usually positively linked to rumen size and therefore intake capacity. Similar pasture DMI indicates a greater DMI per kg BW for HNZ cows compared with HCH cows. Furthermore, HNZ cows spent longer time ruminating. Digestion rate and discharging of the rumen might be increased as rumination determines digestion rate and therefore controls voluntary intake (Bae et al., 1983; Gregorini et al., 2012).

The substitution rate of pasture DMI is linked to milk response with a lower substitution rate for cows with high genetic potential for milk yield (Bargo et al., 2003). However, in the current study, substitution rate, estimated using n-alkane, was the same for both cow strains (0.5 kg/kg). The results are in accordance with missing differences for ECM.

Because no differences were observed between cow strains in blood concentration of insulin, NEFA and BHBA, which are indicators of the energy balance of dairy cows (Reist et al., 2002), no difference between cow strains are obvious in energy status. However, a trend for an increased concentration of creatinine in blood plasma for HCH cows may indicate a higher skeletal muscle breakdown and the mobilization of more protein as an energy source or for gluconeogenesis. In accordance with results from McCarthy et al. (2007a), HNZ cows had greater serum glucose concentration compared with HCH cows. The greater glucose concentration for HNZ cows during breeding season (60 to 150 DIM) represents an important source for energy for the ovary, as Forshell et al. (1991) reported an effect of glucose concentration on conception rates. Therefore greater glucose concentration might indicate greater conception rates which is in line with the differences in conception rates reported between HCH cows and HNZ cows (Piccard et al., 2013). The increase in ALAT activity is in accordance with the results of Thanner et al. (2014), which might suggest the elevated use of amino acids of the HNZ cows for purposes other than protein synthesis.

Minor differences between cow strains in the current study indicate that both cow strains are equally suited for an organic, pasture-based feeding system. However, the current study was a short-term experiment performed with a small number of animals after the peak of lactation without considering BCS losses, fertility and health traits during a whole lactation, which are important aspects for continuous and successful organic farming. Just choosing a different cow breed or cow strain may not ensure a well-working, low-input organic system. Careful selection of cows that have a high grazing drive ensures the efficient use of pasture for milk production, whereas cows that can deal with lower energy intake at the beginning of lactation and show properties of good health and fertility may be well-suited organic cows.

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