Response of seaward-migrating European eel (Anguilla anguilla) to manipulated flow fields

Adam T. Piper, Costantino Manes, Fabio Siniscalchi, Andrea Marion, Rosalind M. Wright and Paul S. Kemp

1International Centre for Ecohydraulics Research, Faculty of Engineering and the Environment, University of Southampton, Southampton SO17 1BJ, UK
2Department of Industrial Engineering, University of Padua, via Marzolo 9, Padova 35131, Italy
3Environment Agency, Rivers House, Thresherfords Business Park, Inworth Road, Feering CO5 9SE, UK

Anthropogenic structures (e.g. weirs and dams) fragment river networks and restrict the movement of migratory fish. Poor understanding of behavioural response to hydrodynamic cues at structures currently limits the development of effective barrier mitigation measures. This study aimed to assess the effect of flow constriction and associated flow patterns on eel behaviour during downstream migration. In a field experiment, we tracked the movements of 40 tagged adult European eels (Anguilla anguilla) through the forebay of a redundant hydropower intake under two manipulated hydrodynamic treatments. Interrogation of fish trajectories in relation to measured and modelled water velocities provided new insights into behaviour, fundamental for developing passage technologies for this endangered species. Eels rarely followed direct routes through the site. Initially, fish aligned with streamlines near the channel banks and approached the intake semi-passively. A switch to more energetically costly avoidance behaviours occurred on encountering constricted flow, prior to physical contact with structures. Under high water velocity gradients, fish then tended to escape rapidly back upstream, whereas exploratory ‘search’ behaviour was common when acceleration was low. This study highlights the importance of hydrodynamics in informing eel behaviour. This offers potential to develop behavioural guidance, improve fish passage solutions and enhance traditional physical screening.

1. Introduction

Globally, freshwater ecosystems are the most anthropogenically impacted, in part due to a loss of connectivity caused by infrastructure such as weirs, dams and other impediments [1–3]. In-channel structures may inhibit or prevent the movement of aquatic biota [4], causing population decline, or even extirpation [5]. For fish, physical barriers obstruct dispersal and migration between habitats required for different ontogenetic stages, and thus disrupt the life cycle [6,7]. River infrastructure, such as hydropower and pumping facilities, can also cause direct injury and mortality to fish that pass through them due to blade strike, cavitation and grinding [8,9]. Further, migratory delay at structures may increase susceptibility to predation, parasites and infectious diseases, and impose energetic costs [7,10].

Despite centuries of efforts to restore and maintain connectivity for fish (typically by providing fish passes), effective solutions remain elusive under many scenarios [11–13]. The development of effective fish passage depends on fundamental knowledge of swimming capabilities, which has received much attention [14], with a historical bias towards salmonids [15,16]. However, this must be combined with an understanding of behavioural response to environmental stimuli [4,17], both those that attract and repel fish [18]. This knowledge is currently lacking for many species [12,18] and there is insufficient understanding of the
transferability of salmonid research to other fish. As fish move with the flow during their downstream migration, it is expected that behavioural response will be more influential than swimming capability, compared with upstream-moving migrants, for which both components play an important role [18].

Fish gain information about their spatial location from multiple stimuli [19]. The discriminability of a specific stimulus and the subsequent response elicited is dependent on both its absolute and relative magnitude in comparison with background noise [20]. Further, discriminability differs among species [9] and ontogenetic stage [7], and with motivation [21], behavioural bias [20], prior experience, learning and habituation [22]. In the complex environments encountered at river infrastructure, hydrodynamic factors probably constitute the dominant cues that inform fine-scale navigation and route selection [23,24].

On a broad scale, the high proportion of river flow diverted through water intakes (such as at hydropower plants or other abstraction points) presents a strong directional cue to downstream-migrating fish that encounter them [15,25]. Fish react to localized changes in flow field characteristics, including turbulence [26,27] and spatial velocity gradient [28], using the lateral line to detect flow strength and direction [29,30] and the otolith of the inner ear to detect whole-body acceleration, deceleration and gravitation [31,32]. The rapid acceleration of flow at constrictions such as at intake channels and downstream fish passage facilities (hereafter referred to as bypasses) can elicit rejection behaviour among downstream-migrating juvenile salmonids [33–35]. Hydrodynamics may also explain observed rejection at river structures for other species [36–38], although understanding is limited for non-salmonids [12,39].

The severe decline of the critically endangered European eel (Anguilla anguilla) has in part been attributed to delayed or blocked seaward migration of escaping adults (silver eels) at river infrastructure [40]. Eels suffer high rates of injury and mortality at pumps and hydropower turbines (typically 15–38% per turbine encountered [41,42]), and are susceptible to impingement at exclusion screens [37,43]. For the few downstream passage solutions trialled for eels, effectiveness is highly variable but generally low [44–46]. Adult eels tend to follow routes of bulk flow [36,47], but on encountering structures display exploratory behaviour and make multiple approaches before passing [46,48,49]. The resolution at which both hydrodynamics and fish migratory paths have been quantified in the field is generally insufficient to determine the relative roles of localized variation in the flow field and physical contact with structures in eliciting specific behaviours [45,49,50]. In common with other diadromous fish species, there has been a historical focus on physical as opposed to behavioural exclusion or guidance for eels. Flume-based studies report eel rejection behaviour after contact with structures such as screens [51,52], leading to the view that, compared with salmonids, adult eels are less sensitive to changes in velocity [18]. This thigmotactic propensity of eels increases the probability of impingement and injury at screens, emphasizing the urgent need to find alternative mitigation solutions.

Given the likely role of riverine barriers in the decline of the European eel and our current lack of knowledge about their response at structures, this study aimed to assess the effect of flow fields on the behaviour of adults during downstream migration. Using acoustic telemetry that enabled near-continuous tracking of fish at sub-metre accuracy, combined with three-dimensional hydrodynamic measurement techniques and computational fluid dynamics (CFD) modelling, a field experiment was conducted to quantify eel responses to manipulated flow fields under two treatments: (1) unrestricted flow with low water acceleration (unrestricted low, UL), and (2) constricted flow with high water acceleration (constricted high, CH). We quantified: (i) behavioural response to flow fields to investigate how hydrodynamics influence eel behaviour, and (ii) the impact of flow fields on swim path characteristics including eel route choice, residence time and track length as indicators of passage efficiency and energetic cost.

2. Material and methods

(a) Site description and experimental set-up

The study was conducted in the forebay upstream of a redundant hydropower (RHP) facility at Longham on the River Stour, Dorset, UK (50° 28.26′ N, 1° 44.16.57′ W). The forebay channel narrows from 17.0 to 12.2 m width, at which point flow is diverted down an intake channel (7.6 m, width) oriented 90° to the forebay (figure 1). The RHP originally housed two turbines but ceased operating during the 1970s.

The intake channel was manipulated to generate two hydrodynamic treatments: (1) UL and (2) CH. A sloping bar rack (7.6 m width, 55° angle, 58 mm vertical bar spacing) extended the full depth of the water column at the RHP intake. In the UL treatment, flow through the bar rack was relatively uniform across the full width of the intake channel. In the CH treatment, the flow was constricted by 66% by wooden boards placed on the upstream face of the bar rack to leave a full-depth opening in the centre channel. Undershoot sluice gates 5 m downstream of the bar rack were manipulated to ensure equal flow passed under both treatments (6.28 ± 0.2 m s⁻¹). Natural fluctuations in water level were controlled by diverting flow via a radial weir directly upstream of the study site.

Hydrodynamics in the two treatments were quantified using water velocity and bathymetry measurements collected with a raft-mounted downward-looking acoustic Doppler current profiler (ADCP; RiverSurveyor ADP M9, SonTek, San Diego, CA, USA), and used to inform and calibrate a two-dimensional CFD model of water velocity within the study site. The ADCP was pulled along tensioned guide wires at eight transect locations (figure 1), repeated before each trial. Data were visually inspected using RiverSurveyor Live v. 3.01 and exported to MATLAB (R2010a, Mathworks, Natick, MA, USA), and used to determine and calibrate a two-dimensional CFD model of water velocity within the study site. The ADCP was pulled along tensioned guide wires at eight transect locations (figure 1), repeated before each trial. Data were visually inspected using RiverSurveyor Live v. 3.01 and exported to MATLAB (R2010a, Mathworks, Natick, MA, USA), and used to inform and calibrate a two-dimensional CFD model of water velocity within the study site. The ADCP was pulled along tensioned guide wires at eight transect locations (figure 1), repeated before each trial. Data were visually inspected using RiverSurveyor Live v. 3.01 and exported to MATLAB (R2010a, Mathworks, Natick, MA, USA), and used to inform and calibrate a two-dimensional CFD model of water velocity within the study site. The ADCP was pulled along tensioned guide wires at eight transect locations (figure 1), repeated before each trial. Data were visually inspected using RiverSurveyor Live v. 3.01 and exported to MATLAB (R2010a, Mathworks, Natick, MA, USA), and used to inform and calibrate a two-dimensional CFD model of water velocity within the study site. The ADCP was pulled along tensioned guide wires at eight transect locations (figure 1), repeated before each trial. Data were visually inspected using RiverSurveyor Live v. 3.01 and exported to MATLAB (R2010a, Mathworks, Natick, MA, USA), and used to inform and calibrate a two-dimensional CFD model of water velocity within the study site. The ADCP was pulled along tensioned guide wires at eight transect locations (figure 1), repeated before each trial. Data were visually inspected using RiverSurveyor Live v. 3.01 and exported to MATLAB (R2010a, Mathworks, Natick, MA, USA), and used to inform and calibrate a two-dimensional CFD model of water velocity within the study site. The ADCP was pulled along tensioned guide wires at eight transect locations (figure 1), repeated before each trial. Data were visually inspected using RiverSurveyor Live v. 3.01 and exported to MATLAB (R2010a, Mathworks, Natick, MA, USA), and used to inform and calibrate a two-dimensional CFD model of water velocity within the study site. The ADCP was pulled along tensioned guide wires at eight transect locations (figure 1), repeated before each trial. Data were visually inspected using RiverSurveyor Live v. 3.01 and exported to MATLAB (R2010a, Mathworks, Natick, MA, USA), and used to inform and calibrate a two-dimensional CFD model of water velocity within the study site. The ADCP was pulled along tensioned guide wires at eight transect locations (figure 1), repeated before each trial. Data were visually inspected using RiverSurveyor Live v. 3.01 and exported to MATLAB (R2010a, Mathworks, Natick, MA, USA), and used to inform and calibrate a two-dimensional CFD model of water velocity within the study site. The ADCP was pulled along tensioned guide wires at eight transect locations (figure 1), repeated before each trial. Data were visually inspected using RiverSurveyor Live v. 3.01 and exported to MATLAB (R2010a, Mathworks, Natick, MA, USA), and used to inform and calibrate a two-dimensional CFD model of water velocity within the study site. The ADCP was pulled along tensioned guide wires at eight transect locations (figure 1), repeated before each trial. Data were visually inspected using RiverSurveyor Live v. 3.01 and exported to MATLAB (R2010a, Mathworks, Natick, MA, USA), and used to inform and calibrate a two-dimensional CFD model of water velocity within the study site. The ADCP was pulled along tensioned guide wires at eight transect locations (figure 1), repeated before each trial. Data were visually inspected using RiverSurveyor Live v. 3.01 and exported to MATLAB (R2010a, Mathworks, Natick, MA, USA), and used to inform and calibrate a two-dimensional CFD model of water velocity within the study site.
anaesthetized (benzocaine 0.2 g l⁻¹) and transferred to the holding barrel (220 l) for a maximum of 8 h before being individually released into the study site.

Fish were maintained in within-river perforated plastic holding barrels at a rate of one per barrel 3 m upstream of the site and held for 10–12 h. The barrel was tethered in the channel centre to reduce bias in route choice and the lid removed at 20.00 (in darkness) from the bank using a rope and pulley system to minimize disturbance and to enable the eels to leave volitionally.

Five eels were released and tracked through the site per trial (four replicates, yielding 20 eels per treatment). Test treatments were alternated to reduce temporal bias (eight trial nights over a 16 night period). Range-testing using known tag locations demonstrated a minimum accuracy and precision of less than 0.5 m within the hydrophone array. Tag detection at both PIT antennae was consistent (98% and more than 99% at antennae 1 and 2, respectively) for depths of less than 0.2 m.

(b) Fish telemetry

Actively migrating adult eels (n = 40) were tracked using acoustic and passive integrated transponder (PIT) telemetry. Eight hydrophones (300 kHz) around the perimeter of the study site and a receiver (HTI Model 290, Hydroacoustic Technology Inc., Seattle, WA, USA) logged all acoustic tag detections. Due to the shallow water depths, it was not possible to accurately determine tag position in the z-dimension. PIT telemetry was used to quantify swim depth at two swim-over antennae and receiver stations (Model LF-HDX-RFID Oregon, Portland, OR, USA), which covered the full channel width in the forebay (14 m length, 0.5 m width) and intake channel (7.5 m length, 0.5 m width) (figure 1).

On the night preceding each trial, actively migrating silver eels were captured at a rack trap downstream of the RHP facility. Fish were maintained in within-river perforated plastic holding barrels (220 l) for a maximum of 8 h before being individually anaesthetized (benzocaine 0.2 g l⁻¹), weighed (wet weight, W, g) and measured (total length, TL, mm). The length of the left pectoral fin (mm) and maximum vertical and horizontal left eye diameter (mm) were used to determine the degree of sexual maturation or ‘silvering’. The first five eels considered migratory (following methods of Pankhurst [55] and Durif et al. [56]) were selected for tagging each night. An acoustic (HTI model 795G, 11 mm diameter, 25 mm length, 4.5 g mass in air, 300 kHz, 0.7–1.3 s transmission rate) and PIT (Texas Instruments, HDX, 3.65 mm diameter, 32 mm length, 0.8 g mass in air) tag were surgically implanted into the peritoneal cavity of each individual following methods similar to Baras & Jeandrain [57] and carried out under a UK Home Office licence. Tagged eels ranged from 639 to 921 mm TL, 566 to 1207 g W, with mean Ocular Index 9.1 (range 7.5–13.5) and mean Fin Index 4.9 (range 4.3–5.8).

After recovery, eels were transferred to a perforated holding barrel 3 m upstream of the site and held for 10–12 h. The barrel was tethered in the channel centre to reduce bias in route choice and the lid removed at 20.00 (in darkness) from the bank using a rope and pulley system to minimize disturbance and to enable the eels to leave volitionally.

Five eels were released and tracked through the site per trial (four replicates, yielding 20 eels per treatment). Test treatments were alternated to reduce temporal bias (eight trial nights over a 16 night period). Range-testing using known tag locations demonstrated a minimum accuracy and precision of less than 0.5 m within the hydrophone array. Tag detection at both PIT antennae was consistent (98% and more than 99% at antennae 1 and 2, respectively) for depths of less than 0.2 m.

(c) Data analysis

Acoustic tag detections were processed using MARKTag v. 5 and ACOUSTICTag v. 5 (Hydroacoustic Technology Inc.), and PIT data were examined for detections when eel tracks intersected antenna locations.

Figure 1. Bathymetry within the forebay and intake channel at a RHP facility on the River Stour, Dorset, UK, during the study period, November 2010. Black lines indicate transects (1–8) along which water velocities were recorded; red lines indicate PIT antennae (I and II). X denotes the fish release point.
To determine whether treatment induced a behavioural response, tracks were overlaid on maps of flow streamlines in MATLAB. A theoretical boundary was imposed at the point where streamlines began to distort upstream of the bend leading to the intake channel (flow distortion boundary). The distance between two adjacent streamlines was set to 1 m (at the entrance), which is comparable with the uncertainty of the telemetry positioning. Tracks were visually assessed and a set of numerical rules devised to determine when trajectories deviated from the streamlines. These deviations, termed 'behavioural switch points', were defined as the first point at which a downstream-moving fish exhibited a turn angle of 90–180° (i.e. deviated from the predominate flow direction) and proceeded in the new direction for a minimum of 3 m. Mann–Whitney U-tests were used to test for a treatment effect on water velocity and acceleration at the point of behavioural switch.

Based on assessment of trajectories immediately after a behavioural switch, individuals were assigned to one of two categories:

Rejection: when downstream-moving fish abruptly switched from negative to positive rheotaxis and moved in a counter streamwise direction for a distance greater than 3 m.

Exploratory behaviour: when downstream-moving fish switched from negative rheotaxis to exhibit lateral movements of greater than 3 m length perpendicular to streamwise flow and encompassing more than two turns.

To quantify the effect of behaviours on the speed and efficiency of migration through the site, the following metrics were calculated for each fish: residence time (duration between first and last detection before passage through the bar rack), mean speed over ground (m s⁻¹) and track length (m). Movement metrics among treatment groups were compared using t-test and Mann–Whitney U-test where the assumptions of parametric analysis were not met. Where trajectories were aligned with streamlines, the velocity of fish over ground (m s⁻¹) was calculated using the difference in fish position every 5 s compared with mean water velocity in the streamline (m s⁻¹). As the study focus was primarily on fish behaviour during movement through the site, near stationary points (values below 0.02 m s⁻¹) were eliminated from the dataset.

Trajectory analyses were carried out using a combination of ArcMap (v. 10, ESRI, Redlands, CA, USA), Geospatial Modelling Environment v. 6.0 [58] and MATLAB. R v. 3.0.0 [59] was used for all statistical analyses.

3. Results

Of the 40 fish released under the two treatments, three swam upstream shortly after release and did not re-enter the study area; these were omitted. The remaining 37 individuals passed through the RHP intake. There was no indication from swim tracks that eels were impinged on the bar rack during passage (i.e. were not stationary at this structure) under either treatment. Residence time was highly variable and ranged from 2.9 to 58.7 min (median, 10.25 min) across all fish, with no treatment effect (Mann–Whitney U = 1.0, p = 0.33).

Upstream of the flow distortion boundary, the majority of fish (73%, 27 out of 37) under both treatments followed trajectories reasonably well aligned with streamlines. The preferred routes were along both sides of the channel (figure 3). The mean ratio of eel ground speed to water velocity in streamlines was 0.78 (± 0.49 s.d.) in this upstream part of the domain.

As downstream-moving individuals approached the 90° bend at the entrance to the intake channel, trajectories generally became more erratic and switches in behaviour were apparent for 35 out of the 37 eels that reached this point. The two fish that did not exhibit a behavioural switch (both in UL treatment) followed relatively direct routes through the site. The majority of fish that responded did so in the intake channel (80% and 95%
for UL and CH treatments, respectively). Four switches occurred more than 2 m from the intake channel so were deemed not to be influenced by hydrodynamic treatment and were therefore excluded from further analysis.

In the CH treatment, switch points were distributed throughout the intake channel and area immediately upstream, whereas under the UL treatment they tended to be concentrated within a narrow band across the channel width (figure 4). Mean depth-averaged flow velocities at the points of behavioural switch ranged from 0.034 to 0.72 and from 0.14 to 0.67 m s\(^{-2}\) for UL and CH treatments, respectively.

The median depth-averaged water velocity at the point of switch was higher in the UL compared with the CH treatment (0.67 and 0.57 m s\(^{-2}\), respectively; Mann–Whitney \(U = 2.68, p = 0.006\)). Velocity acceleration at the point of switch ranged from 0.001 to 0.051 and from 0.002 to 0.083 for UL and CH treatments, respectively, and did not vary among treatments (Mann–Whitney \(U = 1.28, p = 0.21\)).

Overall, rejection dominated behaviour immediately following a switch, apparent in 71% of fish. Treatment influenced post-switch behaviour as all fish exhibited rejection (figure 5a) under CH, whereas exploratory behaviour (figure 5b) was observed in 75% of individuals under UL treatment. Nearly all (91%) of the fish that rejected did so multiple times, up to a maximum of four occasions. Second points of rejection occurred closer to the bar rack than the first in 70% of cases.

Individuals in CH swam greater distances after behavioural switch than in the UL treatment (\(t_{29} = 2.23, p = 0.03\), mean 89.3 \(\pm\) 38.7 m and 54.3 \(\pm\) 44.8 m (\(\pm\) s.d.), respectively), and a higher proportion of the post-switch tracks occurred outside of the intake channel (median proportion 68%, range 21–85%; Mann–Whitney \(U = 3.93, p < 0.001\)). Conversely, in the UL treatment, fish rarely left the intake channel after switch (median proportion 10%, range 0–68%). Mean eel ground speed (m s\(^{-1}\) over ground, unadjusted for water velocity) compared before and after a change in behaviour also revealed a treatment effect after switch (\(t_{29} = 2.88, p = 0.007\)), but not before (\(t_{29} = 1.67, p = 0.11\)). Ground speed was higher post-switch under CH (0.28 \(\pm\) 0.05 m s\(^{-1}\), median \(\pm\) s.d.) than for the UL treatment (0.23 \(\pm\) 0.05 m s\(^{-1}\), median \(\pm\) s.d.).

During downstream movements, 84% of eels were detected within 20 cm of the channel bed at the PIT antennae (I and II, figure 1), whereas upstream-moving fish were less frequently detected (56%), which may suggest a reduced tendency for benthic orientation after rejection.

4. Discussion

Manipulation of flow fields clearly influenced the behaviour of downstream-moving adult eels. On encountering flow acceleration eels displayed erratic behaviour, and the magnitude of response was positively related to maximum water velocity.
behaviour denotes fish movement with a biased correlated random walk, downstream in the direction of flow (B1). On approaching flow accelerations or decelerations, the fish exhibits one of two responses (determined by thresholds governed by recent past experience); either it orients swimming in the direction leading to faster water to facilitate obstacle and turbulence avoidance (B2), or a repulsive escape response is elicited in which the fish temporarily abandons downstream migration and swims upstream (B3). The fourth behaviour regulates fish response to pressure, and thus dictates swim depth. Incorporating all four behaviours provided the best fit between simulated fish and actual swim paths of juvenile salmonid smolts (*Oncorhynchus* sp.) descending complex flow fields at Lower Granite Dam, Snake River, USA [64].

The tendency for eels to align with streamlines in the upstream part of the forebay suggested advective behaviour (i.e. semi-passive drift with the local flow), which is broadly supported by the correspondence between eel ground speed and mean streamline velocity. Downstream movement was predominantly benthic-oriented, as observed in previous studies [45,46,48]. Therefore, the lower than 1 : 1 ratio of ground speed to water velocity probably reflects the lower velocities eels would have experienced near the channel bed, relative to modelled depth-averaged values. This resembled the B1 behaviour described by the Goodwin *et al.* [64] framework. However, in this study, the eels predominantly followed routes parallel to streamlines located close to the banks of the channel. Given their thigmotactic nature [51], it is surprising that individual eels seldom came into contact with the channel banks. Instead, our findings suggest that proximity to (rather than contact with) structures was used to some benefit. For eels, which undergo migration during dark and often turbid conditions which reduce visual cues, proximity to lateral boundaries may be an important navigational cue [65]. Fish are able to detect the hydraulic signatures created by structures through the mechanosensory system [23,30] and learn that near-field hydraulic patterns provide information on the environment beyond their sensory range [23]. For example, frictional resistance, resulting in decreasing average velocities towards the channel bed and banks, can be distinguished from form resistance induced by in-channel objects (e.g. rocks and woody debris), where water velocities increase due to reduced area, and increased travel distance, of flow around the object [23,24]. It was not clear what hydraulic signatures were detected by eels to identify the lateral banks. There was no apparent difference in characteristics between the streamlines eels descended, near the lateral boundaries, and those in the centre of the channel. It is plausible that such hydraulic signatures could be identified by using three-dimensional numerical models because they provide information on flow features induced by lateral boundaries (e.g. secondary currents) that cannot be captured by two-dimensional models such as the one used in this study.

The response of eels on encountering velocity acceleration depended on both the novelty and strength of the transition stimuli. When approaching rapid velocity acceleration, most fish responded by rejecting upstream, a behaviour that closely corresponded to B3 in the Goodwin *et al.* framework [64]. This is postulated to occur when the relative change in acceleration exceeds a threshold intensity, causing the fish to swim in the opposite direction to the principal velocity vector and return upstream [64]. The greater the magnitude of acceleration, the faster the subsequent swim speed during eel response. A similar relationship has been observed and flow acceleration. When flow was constricted, eels exhibited rejection as opposed to exploratory behaviour, which was observed predominantly under unrestricted conditions.

The response to the abrupt hydrodynamic transitions described may suggest the avoidance of hazardous areas that could cause damage or disorientation [25,33,35]. Other studies have demonstrated a rejection response exhibited by fish on encountering velocity gradients (e.g. juvenile Atlantic salmon [33] and juvenile Pacific salmon [34]). Although most eel rejections were several metres upstream of the bar rack, a small proportion (5%) could have been associated with physical contact at this structure. Findings differ from previous studies under both laboratory [51,52,60] and field conditions [46], which report that the majority of eels did not respond until making contact with structures. Although pre-contact rejection has been previously documented in the field [61,62], in this study both the close proximity to the intake at which the behavioural switch occurred and the positive relationship between magnitude of response and velocity acceleration provide evidence of the link between hydrodynamic stimuli and avoidance by eels.

In common with salmonids, eel behaviour during downstream migration was influenced by flow acceleration. To advance fish passage research, it is important that common concepts and approaches are adopted to aid the transfer of knowledge about different species and study systems [4,63]. A conceptual framework to understand and predict fish movement patterns in relation to complex flow fields around river structures is provided by Goodwin *et al.* [64]. In a model that describes four mutually exclusive downstream behavioural states (B1–4) used to govern the movements of simulated fish, individuals adjust swim orientation and speed in response to local water acceleration and pressure (depth). The first
for salmon smolts rejecting decelerating flow [28]. When approaching a less abrupt acceleration transition, eels ventured closer to the intake, and therefore experienced higher velocities before switching to slower exploratory behaviour. This broadly conforms to similar milling/exploratory behaviour in salmonids [66], expressed as recursive cycles between behaviours B3 and B1–2 in modelled fish [64]. Eels are influenced more by thigmotactic cues than salmonids [67], thus observed exploratory behaviour may have been both hydraulically and thigmotactically mediated. The propensity of eels to explore their environment has been associated with active searching for a way through (or alternative route past) screens and bar racks at hydropower and pumping facilities [46,61]. In this study, eels generally restricted exploration to the area within the intake channel, yet were rarely detected to contact the bar rack.

After their first response to velocity acceleration in the intake channel, individuals appeared to become somewhat habituated to the transition and more likely to pass through the same region on subsequent encounters. For a fish to detect change in a stimulus relative to the background noise it is acclimated to, the stimulus must exceed a threshold value (termed ‘just notable difference’). Therefore, the response is dependent on exposure history [24,68]. Such adaptive behaviour enables animals to repeatedly test their environment and adjust their risk of exposure to potentially harmful elements based on prior experience.

The importance of hydrodynamics in influencing eel behaviour has significant implications for progressing guidance and passage technologies for this threatened species. Eel bypasses should be designed to avoid abrupt velocity acceleration at the entrance, as is currently advised for salmonids [15,69], with the aim to minimize rejection. Conversely, avoidance behaviours present an opportunity to guide eels away from dangerous areas and towards safe passage routes. There is clear potential for hydrodynamic-based guidance to enhance the effectiveness of traditional physical screens that can be expensive to install and maintain, reduce power generation or pumping efficiency, and towards safe passage routes. There is clear potential for hydrodynamic-based guidance to enhance the effectiveness of traditional physical screens that can be expensive to install and maintain, reduce power generation or pumping efficiency, and towards safe passage routes. There is clear potential for hydrodynamic-based guidance to enhance the effectiveness of traditional physical screens that can be expensive to install and maintain, reduce power generation or pumping efficiency, and towards safe passage routes. There is clear potential for hydrodynamic-based guidance to enhance the effectiveness of traditional physical screens that can be expensive to install and maintain, reduce power generation or pumping efficiency, and towards safe passage routes. There is clear potential for hydrodynamic-based guidance to enhance the effectiveness of traditional physical screens that can be expensive to install and maintain, reduce power generation or pumping efficiency, and towards safe passage routes.

The increased resolution afforded by the fish-positioning telemetry and flow-mapping techniques employed in this study has challenged historical perceptions about eel behaviour derived from more coarse-scale investigations. These advances represent an important step forward in the drive to develop effective guidance and passage solutions for this species at anthropogenic barriers. Combining fine-scale fish movement data with empirically informed hydrodynamic models offers great potential to further our limited understanding of fish behaviour in relation to the complex hydrodynamic environments encountered at river infrastructure.

Ethics. Fish tagging was carried out in compliance with UK Home Office regulations which include an ethical review process.

Data accessibility. Fish trajectory data file available at Dryad: doi:10.5061/dryad.c77jn.

Authors’ contributions. A.T.P.c. conceived of and designed the study, conducted data collection, conducted analysis of fish data, contributed to analysis of hydrodynamic data, and drafted the manuscript. C.M., F.S. and A.M. conducted analysis and modelling of hydrodynamic data, and helped draft the manuscript. R.M.W. participated in the design of the study and helped draft the manuscript. P.S.K. coordinated the study, participated in the design of the study, contributed to data collection and analysis, and helped draft the manuscript.

Competing interests. We declare we have no competing interests.

Funding. This study was joint-funded by the University of Southampton and the Environment Agency, UK. Funding is also acknowledged from the EPSRC Doctoral Training Grant awarded to the University of Southampton (EPS/P505119/1).

Acknowledgements. The authors thank Sembcorp Bournemouth Water, Paula Rosewarne, Alan Piper, Roger Castle and Jim Davis for their assistance. Thanks also to two anonymous reviewers for their valuable suggestions to improve the manuscript.

References

1. Malmqvist B, Rundle S. 2002 Threats to the running water ecosystems of the world. Environ. Conserv. 29, 134 – 153. (doi:10.1017/S0376892902000097)

2. Vorissmarty CI et al. 2010 Global threats to human security and river biodiversity. Nature 467, 555 – 561. (doi:10.1038/nature09440)

3. Strayer DL, Dudgeon D. 2010 Freshwater biodiversity conservation: recent progress and future challenges. J. N. Am. Benthol. Soc. 29, 344 – 358. (doi:10.1899/08-171.1)

4. Kemp PS. 2012 Bridging the gap between fish behaviour, performance and hydrodynamics: an ecohydraulics approach to fish passage research. River Res. Appl. 28, 403 – 406. (doi:10.1002/rra.1599)

5. Nilsson C, Reidy CA, Dynesius M, Revenga C. 2005 Fragmentation and flow regulation of the world’s large river systems. Science 308, 405 – 408. (doi:10.1126/science.1107887)

6. Northcote T. 1998 Migratory behaviour of fish and its significance to movement through riverine fish passage facilities. In Fish migration and fish bypasses (eds M. Jungwirth, S. Schmutz, S. Weiss), pp. 3 – 18. Vienna, Austria: Wiley-Blackwell.

7. Lucas M, Baras E. 2001 Migration of freshwater fishes. Oxford, UK: Blackwell Science.

8. Turrensey A, Clough S, Hanson K, Ramsay R, McEwan D. 2000 Risk assessment for fish passage through small, low-head turbines. Harwell, UK: Atomic Energy Research Establishment, Energy Technology Support Unit, New and Renewable Energy Programme.

9. Schilt CR. 2007 Developing fish passage and protection at hydropower dams. Appl. Anim. Behav. Sci. 104, 295 – 325. (doi:10.1016/j.applanim.2006.09.004)

10. Garcia De Leaniz C. 2008 Weir removal in salmonid streams: implications, challenges and practicalities. Hydrobiology 609, 83 – 96. (doi:10.1007/s10750-008-9397-x)

11. Cooke S, Bunt C, Hamilton S, Jennings C, Pearson M, Cooperman M, Markle D. 2005 Threats, conservation strategies, and prognosis for suckers (Catostomidae) in North America: insights from regional case studies of a diverse family of
non-game fishes. Biol. Conserv. 121, 317 – 331. (doi:10.1016/j.biocon.2004.05.013)

12. Roscoe DW, Hinch SG. 2010 Effectiveness monitoring of fish passage facilities: historical trends, geographic patterns and future directions. Fish Fish. 11, 12 – 33. (doi:10.1111/j.1467-2979.2009.00333.x)

13. Noonan MJ, Grant JW A, Jackson CD. 2012 A quantitative assessment of fish passage efficiency. Fish Fish. 13, 450 – 464. (doi:10.1111/j.1467-2979.2011.00445.x)

14. Anderson JJ. 1988 Diverting migrating fish past turbines. Northwest Environ. J. 4, 109 – 128.

15. Clay CH. 1995 Design of fishways and other fish facilities, 2nd edn. Boca Raton, FL: Lewis Publishers.

16. Kalmijn A. 1989 Functional evolution of lateral line and inner ear sensory systems. In The mechanosensory lateral line: neurobiology and evolution (eds S Goumbs, P Gomer, H Munz), pp. 187 – 215. New York, NY: Springer.

17. Castro-Santos T, Cotel A, Webb P. 2009 Fishway Design of fishways and other fish passage structures. In Developing realistic fish passage criteria: an integrated approach. In Symposium on fish migration and fish bypasses (eds M Jungwirth, S Schmutz, S Weiss), pp. 315 – 328. Oxford, UK: Fishing News Books.

18. Kemp PS, Anderson JJ, Vowles AS. 2012 Quantifying fish passage in a historical context. Ecol. Eng. 48, 8 – 18. (doi:10.1016/j.ecoleng.2011.07.004)

19. Castro-Santos T, Cotel A, Webb P. 2009 Fishway evaluations for better bioengineering: an integrative approach. In Challenges for diadromous fishes in a dynamic global environment. American Fisheries Society Symposium 69 (eds A Haro, C Moffit, M Dadswell), pp. 557 – 575. Bethesda, MD: American Fisheries Society.

20. Williams JS, Armstrong G, Katopodis C, Larinier M, Travade F. 2012 Thinking like a fish: a key ingredient for development of effective fish passage facilities at river obstructions. River Res. Appl. 28, 407 – 417. (doi:10.1002/rra.1551)

21. Colgan P. 1993 The motivational basis of fish behaviour. In Behaviour of teleost fishes, 2nd edn. Boca Raton, FL: Lewis Publishers.

22. Colgan P. 1993 The motivational basis of fish behaviour. In Behaviour of teleost fishes, 2nd edn. Boca Raton, FL: Lewis Publishers.

23. Goodwin A, Nestler JM, Anderson JJ, Webber L. 2012 Characterizing the swimming of Atlantic salmon smolts using high-speed video analysis. Ecol. Freshwat. Fish 19, 355 – 371. (doi:10.1111/j.1467-2979.2011.00459.x)

24. Kemp PS, Anderson JJ, Vowles AS. 2012 Quantifying behaviour of migratory fish: application of signal detection theory to fisheries engineering. Ecol. Eng. 41, 22 – 31. (doi:10.1016/j.ecoleng.2011.12.013)

25. Odling-Smee L, Braithwaite V A. 2003 The role of fish behaviour in fishway design. Fish Fish. 4, 235 – 246. (doi:10.1046/j.1467-2979.2003.00127.x)

26. Goodwin A, Nestler JM, Anderson JJ, Webber L. 2012 A new tool to forecast fish movement and passage. Hydro. Res. 29, 3 – 8.

27. Nestler JM, Goodwin RA, Smith DL, Anderson JJ, Li S. 2008 Optimum fish passage and guidance designs are based in the hydrogeomorphology of natural rivers. River Res. Appl. 24, 148 – 168. (doi:10.1002/rra.1056)

28. Kubota CC. 2001 Turbulent attraction flows for guiding juvenile salmonids at dams. In Behavioral technologies for fish guidance: American Fisheries Society Symposium 26 (ed. CC Cubanto), pp. 57 – 77. Bethesda, MD: American Fisheries Society.

29. Liao JC. 2007 A review of fish swimming mechanics and behaviour in altered flows. Phil. Trans. R. Soc. B 362, 1973 – 1993. (doi:10.1098/rstb.2007.2082)

30. Liao JC, Beal DN, Lauer GV, Triantafyllou MS. 2003 The Caiman gait: novel body kinematics of rainbow trout swimming in a vortex street. J. Exp. Biol. 206, 1059 – 1073. (doi:10.1242/jeb.00209)

31. Enders EC, Gesell MH, Anderson JJ, Williams JG. 2012 Effects of decelerating and accelerating flows on juvenile salmonid behavior. Trans. Am. Fish. Soc. 141, 357 – 364. (doi:10.1080/00288390.2012.646404)

32. Voigt R, Carton AG, Montgomery JC. 2000 Responses of anterior lateral line afferent neurons to water flow. J. Exp. Biol. 203, 2495 – 2502.

33. Montgomery J, Carton G, Voigt R, Baker C, Diebel C. 2000 Sensory processing of water currents by fishes. Phil. Trans. R. Soc. Lond. B 355, 1325 – 1327. (doi:10.1098/rstb.2000.0693)

34. Kalmijn A. 1989 Functional evolution of lateral line and inner ear sensory systems. In The mechanosensory lateral line: neurobiology and evolution (eds S Goumbs, P Gomer, H Munz), pp. 187 – 215. New York, NY: Springer.

35. Kemp PS, Gesell MH, Williams JG. 2008 Response of downstream migrant juvenile Pacific salmonids to accelerating flow and overhead cover. Hydrobiologia 609, 205 – 217. (doi:10.1007/s10750-008-9412-2)

36. Enders EC, Gesell MH, Williams JG. 2009 Development of successful fish passage structures for downstream migrants requires knowledge of their behavioural response to accelerating flow. Can. J. Fish. Aquat. Sci. 66, 2109 – 2117. (doi:10.1139/f09-141)

37. Jansen HM, Winter HV, Bruijs MCM, Polman HJG. 2007 Just go with the flow? Route selection and mortality during downstream migration of silver eels in relation to river discharge. ICES J. Mar. Sci. 64, 1437 – 1443. (doi:10.1016/j.icesjms.2013)

38. Calles O, Olsson IC, Camoglio C, Kemp PS, Blunden L, Schmitz M, Greenberg LA. 2010 Size-dependent mortality of migratory silver eels at a hydropower plant, and implications for escapement to the sea. Freshwat. Biol. 55, 2167 – 2180. (doi:10.1111/j.1365-2427.2010.02459.x)

39. Monik B, Weaver D, Thompson C, Osiander F. 1989 Effects of flow and weir design on the passage behavior of American shad and salmonids in a fishway in the middle reaches of the Delaware River. N. Am. J. Fish. Manage. 9, 60 – 67. (doi:10.1577/1548-8675(1989)009<0060:efawd>2.3.co;2)

40. Calles O, Greenberg LA. 2005 Evaluation of nature-like fishways for re-establishing connectivity in fragmented salmonid populations in the River Eman. River Res. Appl. 21, 951 – 960. (doi:10.1002/rra.865)

41. ICES. 2011 Report of the 2011 Session of the Joint EIFAC/ICES Working Group on Eels. Lisbon, Portugal. ICES.
53. Dinehart R, Burau J. 2005 Repeated surveys by acoustic Doppler current profiler for flow and sediment dynamics in a tidal river. J. Hydro. 314, 1 – 21. (doi:10.1016/j.jhydrol.2005.03.019)

54. Galland J-C, Goutal N, Hervouet J-M. 1991 TELEMAC: a new numerical model for solving shallow water equations. Adv. Water Res. 14, 138 – 148. (doi:10.1016/0309-1708(91)90006-A)

55. Pankhurst NW. 1982 Relation of visual changes to the onset of sexual maturation in the European eel Anguilla anguilla L. J. Fish Biol. 21, 127 – 140. (doi:10.1111/j.1095-8649.1982.tb03994.x)

56. Durif CM, Ginneken V, Dufour S, Müller T, Elie P. 2009 Seasonal evolution and individual differences in silverying eels from different locations. In Spawning migration of the European eel (eds G van den Thillart, S Dufour, JC Rankin), pp. 13 – 38. Dordrecht, The Netherlands: Springer.

57. Baras E, Jeandrain D. 1998 Evaluation of surgery procedures for tagging eel Anguilla anguilla with biotelemetry transmitters. Hydrobiologia 371 – 372, 107 – 111. (doi:10.1023/A:10170900117425)

58. Beyer HL. 2012 Geospatial Modelling Environment (v. 0.6.0.0). See http://www.spatialecology.com/gme.

59. R core team. 2013 R: a language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical computing.

60. Amanal SV, Winchell FC, McMahon BJ, Dixon DA. 2003 Evaluation of angled bar racks and louvers for guiding silver phase American eels. In Biology, Management, and Protection of Catadromous Eels Symposium 33, St Louis, Missouri, 2 – 22 August 2000 (ed. D Dixon), pp. 367 – 376. Bethesda, MD: American Fisheries Society.

61. Keeken OA, Viscount D, Winter HV. 2011 Behaviour of eels around a fish exclusion system with strobe lights at pumping station Ijmuiden. DIDSON measurements. Wageningen, The Netherlands: Institute for Marine Resources and Ecosystem Studies (IMARES).

62. Holzner M. 2000 Neue Versuche zur Schaden minimierung bei der Aalabwanderung. Paper presented at Voordracht SVK Fischereitagung, Künzell, 1 – 2 March.

63. Nestler JM, Goodwin RA, David L Mathematical and conceptual framework for ecohydraulics. In Hydroecology and ecohydrology: past, present and future (eds PJ Wood, DM Hannah, J Sadler), pp. 205 – 224. Chichester, UK: John Wiley and Sons.

64. Goodwin RA et al. 2014 Fish navigation of large dams emerges from their modulation of flow field experience. Proc. Natl Acad. Sci. USA 111, 5277 – 5282. (doi:10.1073/pnas.1311874111)

65. Montgomery J, Goembs S, Halstead M. 1995 Biology of the mechanosensory lateral line in fishes. Rev. Fish Biol. Fish. 5, 399 – 416. (doi:10.1007/BF01103813)

66. Svendsen JC, Aarestrup K, Malte H, Thygesen Uh, Baktorf H, Koed A, Deacon MS, Fiona Cubitt K, Scott McKinley R. 2011 Linking individual behaviour and migration success in Salmo salar smolts approaching a water withdrawal site: implications for management. Aquat. Living Resour. 24, 201 – 209. (doi:10.1051/alr/2011121)

67. Russon IJ, Kemp PS. 2011 Advancing provision of multi-species fish passage: behaviour of adult European eel (Anguilla anguilla) and brown trout (Salmo trutta) in response to accelerating flow. Ecol. Eng. 37, 2018 – 2024. (doi:10.1016/j.ecoleng.2011.08.005)

68. Weber EH. 1846 Der tastiss und das gemeingefühl. In Handwörterbuch der Physiologie: Band 3, Abt. 2 (ed. R Wagner), pp. 481 – 588. Braunschweig, Germany: Vieweg.

69. O'Keefe N, Turnpenny AWH. 2005 Screening for intake and outfalls: a best practice guide. Science Report SC030231. Bristol, UK: Environment Agency.

70. Bates D, VanDerwalker J. 1964 Exploratory experiments on the deflection of juvenile salmon by means of water and air jets. Fish Passage Res. Program Rev. Progress 3, 1 – 6.

71. Muir WD, Williams JG. 2012 Improving connectivity between freshwater and marine environments for salmon migrating through the lower Snake and Columbia River hydropower system. Ecol. Eng. 48, 19 – 24. (doi:10.1016/j.ecoleng.2011.06.034)