The Effect of Fermentation and Drying Processes on Oil Absorption Capacity of Palm Fruit Mesocarp Fibre Flour

I INTRODUCTION

Literatures have [1,2] reported that palm mesocarp fibre (PMF) are solid wastes generated from separating mesocarp fibre from other solid wastes obtained from processing oil palm fruits into crude palm oil (CPO). Small and medium scale (SMS) farmers are responsible for the processing of most oil palm fruits into CPO [3]. In Nigeria, individual household processors of CPO use the PMF as fuel for cooking and in steam boilers at the small and medium scale mills. Palm mesocarp fibre has also been used for bio-composite production [4]. The PMF can serve as an attractive feed stock for dietary fibre because it is produced in large quantities from a renewable source [5] The PMF is also a lignocellulosic biomass that can be used as raw materials for fermentation processes [6].

Fermentation simply can be defined as a metabolic process concerned with the release of energy by cells for the conversion of carbohydrate into an acid or alcohol by microorganisms. Under favourable conditions fermentation can occur naturally in different foods materials and had been used for production of wine and beer [7]. According to [8] a fermenter is the main equipment used for fermentation. The fermenter is used to obtain a desired product under a controlled environment for the growth of a microorganism or a mixture of microorganisms. Microorganisms operate efficiently at certain temperature and pH conditions, an effective fermenter should provide these conditions while the rate of evaporation losses should not be in excess [7].

Oil absorption capacity (OAC) is associated with the chemical composition of plant polysaccharides [6]. Surface properties and hydrophobic nature of the plant material are the characteristics that determine the OAC of foods [20]. Flavour and mouth feel are the product parameters greatly affected by OAC [24].

II MATERIALS AND METHODS

A. Preparation of Palm Mesocarp Fibre

Two kilograms (2kg) of fresh PMF from the tenera specie were obtained from processors of oil palm fruits in Isuochi, Umunneochi LGA, Abia State of Nigeria. The PMF samples were divided into two equal parts. One portion (1kg) was fermented in an open vat for 7 days. The fermented PMF was dried using an oven Fermented and dried PMF samples were processed according to the experimental runs in Table 1 generated using Central Composite Design (CCD) method found within the Response Surface Design (RSD) using [12]. The other portion (1000g) which served as the control sample was oven dried to 14% moisture content. The dried fermented and control samples were ground, hermetically sealed, and stored at room temperature.

The substrate solution used for the fermentation process was prepared using the method adapted by [9] at the experimental runs shown in Table 1. Glucose solution used as the substrate solution was prepared by dissolving 100g of glucose in one litre of distilled water. To adjust the solution to be more acidic concentrated HCl was added into it.

B. Determination of the Oil Absorption Capacity

The OAC was determined by the method adopted in [13]. One gramme (1g) of the fermented PMF samples was refined groundnut oil (10ml) to form a solution. The solution was stirred for 30 seconds. The fermented PMF samples stood for 30 minutes at 30°C in a water bath and centrifuged at 300rpm.
for 20 minutes. The supernatant volume value was noted. The supernatant volume value was used to calculate the WAC and OAC.

### TABLE 1: EXPERIMENTAL RUNS FOR FERMENTATION AND DRYING PROCESSES

| Run | A  | B  | C  | D  | E  | F  |
|-----|----|----|----|----|----|----|
| 1   | 50 | 6  | 6  | 40 | 30 | 40 |
| 2   | 50 | 4  | 4  | 40 | 30 | 40 |
| 3   | 40 | 5  | 5  | 60 | 45 | 30 |
| 4   | 40 | 5  | 5  | 12 | 45 | 30 |
| 5   | 50 | 4  | 6  | 80 | 60 | 20 |
| 6   | 63 | 5  | 5  | 60 | 45 | 30 |
| 7   | 40 | 5  | 5  | 60 | 45 | 6  |
| 8   | 30 | 6  | 4  | 40 | 30 | 40 |
| 9   | 40 | 7  | 5  | 60 | 45 | 30 |
| 10  | 30 | 6  | 6  | 80 | 30 | 40 |
| 11  | 40 | 5  | 5  | 60 | 9  | 30 |
| 12  | 30 | 4  | 6  | 80 | 60 | 40 |
| 13  | 40 | 5  | 5  | 60 | 45 | 30 |
| 14  | 30 | 6  | 4  | 40 | 60 | 20 |
| 15  | 40 | 5  | 5  | 60 | 45 | 30 |
| 16  | 30 | 4  | 4  | 80 | 60 | 20 |
| 17  | 30 | 4  | 6  | 80 | 30 | 20 |
| 18  | 30 | 4  | 6  | 40 | 60 | 20 |
| 19  | 50 | 6  | 6  | 80 | 30 | 20 |
| 20  | 40 | 2  | 5  | 60 | 45 | 30 |
| 21  | 50 | 6  | 4  | 80 | 30 | 40 |
| 22  | 40 | 5  | 5  | 60 | 45 | 53 |
| 23  | 16 | 5  | 5  | 60 | 45 | 30 |
| 24  | 30 | 6  | 6  | 80 | 60 | 20 |
| 25  | 40 | 5  | 5  | 60 | 80 | 30 |
| 26  | 40 | 5  | 5  | 60 | 45 | 30 |
| 27  | 40 | 5  | 2  | 60 | 45 | 30 |
| 28  | 30 | 6  | 6  | 40 | 30 | 20 |
| 29  | 40 | 5  | 5  | 60 | 45 | 30 |
| 30  | 30 | 6  | 4  | 80 | 60 | 40 |
| 31  | 50 | 4  | 6  | 80 | 30 | 40 |
| 32  | 50 | 6  | 4  | 40 | 30 | 20 |
| 33  | 50 | 4  | 4  | 40 | 60 | 20 |
| 34  | 50 | 6  | 6  | 80 | 60 | 40 |
| 35  | 30 | 4  | 6  | 60 | 30 | 20 |
| 36  | 40 | 5  | 5  | 60 | 45 | 30 |
| 37  | 40 | 5  | 5  | 60 | 45 | 30 |
| 38  | 30 | 4  | 4  | 40 | 60 | 40 |
| 39  | 30 | 4  | 6  | 40 | 30 | 40 |
| 40  | 50 | 6  | 4  | 80 | 60 | 20 |
| 41  | 30 | 6  | 6  | 40 | 60 | 40 |

A=Fermentation Temperature (°C), B= Fermentation pH, C=Fermentation Time (days), D= Fermentation Mass (g), E= Drying Temperature (°C), F= Drying Time (minutes)

D. Proximate Composition of PMF Flour

Proximate composition analysis for samples was conducted following the [10] method. The components analysed were moisture content; crude lipid; crude fibre; crude protein and ash. The results obtained were in triplicates and the mean values were used for the analysis.

III RESULTS AND DISCUSSIONS

A. Effect of Fermentation Temperature on Oil Absorption Capacity

The result of the effect of fermentation temperature OAC in this study is illustrated in Figure 1. The highest fermentation temperature of 40°C was obtained in this study. The result obtained in the study could be linked to microbial activity which is at their best at temperatures within and slightly above room temperature. [11] reported that fermentation process can occur at different temperatures depending on the environmental conditions. The temperature range according to [14] where the highest rates of fermentation occur is from 30°C to 45°C. The finding of this study is within the proximity of fermentation temperature range as observed by [15]. [14] also reported that at fermentation temperature above 50°C stress is induced on microbial activity in a multi microorganism environment.

![Figure 1: Effect of fermentation temperature on oil absorption capacity](image)

B. Effect of Fermentation pH on Oil Absorption Capacity

The result obtained in the study from the effect of fermentation pH from 2.0 to 7.0 is shown in Figure 2. Figure 2 indicates that the pH value of 5 gave the highest value for OAC. The OAC values increased between the pH of 2.0 to 5.0 and decreased thereafter. Microbial activity is a function of the acidity or
alkalinity of the reaction medium. [15] and [9] reported that as the pH of a reaction medium moves from acidic to alkaline the microbial enzymes that catalyse the metabolism of glucose loses their ability to function efficiently. The trend observed from the result of this study could be because enzymes that facilitate the metabolism of glucose functions are known to be more effectively in an acidic condition [14]. The result of this study shows close agreement to [9] that reported the pH of 4.0 to 5.0 as the optimum pH range for any fermentation processes.

C. Effect of Fermentation Time on Oil Absorption Capacity

The result of the effect of fermentation time on OAC obtained in this study is shown in Figure 3. The result of the study indicated that the OAC value increased from 40% (2 days) to 51% (5 days) and then decreased to 45% (7 days). The highest value for OAC was recorded after 5 days of fermentation. The result obtained from the study indicates that the 5th day marked the apex of the exponential growth rate when there was little or no stress on microbial activity due to unfavourable conditions of competing reactions for substrates and production of undesirable products [17]. The result of this study is closely related to [18] who noted 5 days as the optimal number of days required for a fermentation process.

D. Effect of Fermentation Mass on Oil Absorption Capacity

Results presented in Figure 4 from the study reveals that fermentation mass of 60g gave the highest value for OAC. The result obtained in this study indicates that below and above 60g fermentation mass there was a reduced percent yield of OAC. [16] and [9] observed that during a fermentation process the feedstock mass of the material to be fermented determines the outcome of the yield result. The pattern of result observed in the study could be attributed to the fact that microbial activity during the fermentation process was overwhelmed by the either high or low substrate concentration. High fermentation masses beyond the optimum fermentation mass also showed reduced values for OAC. This is an indication that the microbial activity was overwhelmed by reduced substrate concentration. The optimum fermentation mass pattern in this study shows proximity with the works of [16] and [11] in their study of bioethanol yield from biomass.

E. Effect of Drying Temperature on Oil Absorption Capacity

The highest values for OAC were obtained in this study at a drying temperature of 45°C as shown Figure 5. This pattern of result could be attributed to the increase in the soluble dietary fibre and change in the starch and amino acid content of the PMF. The drying temperature of 45°C obtained in this study is within the proximity of temperature range observed by [19] and [20] in their study of blueberry liquids and chia seeds respectively. It also agrees with work of [21] which reported that heat processing led to an increase in non-polar amino acid and soluble dietary fibre thus increasing the presence of hydrophilic starch content.

F. Effect Drying Time on Oil Absorption Capacity

The effect of drying time on OAC in this study is as presented in Figures 6. The result of the study shows that the highest values for OAC were obtained at a drying time of 30 minutes. According to [22] the effectiveness and efficiency of a drying process is dependent on processing time. The results obtained from this study can be ascribed to the quality of starch in the PMF and the retention of moisture with more hydrophobic groups within the matrix of the PMF being analysed. The result of this work shows an agreement to [23]; [11] and [16] that reported similar drying processing time for agro-waste fibrous materials.
A decrease from 17.60% (unfermented PMF) to 15.72% (fermented PMF) for crude fibre was observed in this study. The study’s result for the crude fibre can be credited to the degradation of hemicellulose, cellulose, and lignin into soluble dietary fibre fraction during fermentation. This trend of result agrees with [25] Lario et al. (2004) that reported similar results from processing of agro-waste fibrous materials.

The crude fat from this study decreased from 30.17% to 26.20%. The observed decrease can also be attributed to the presence of more hydrophilic amino acids and the fact that fermentation produces mainly short-chain fatty acids that are easily utilised by lipolytic microorganisms. [15], noted that fermentation resulted in the use of short chain fatty acids as feedstock by lipolytic microorganisms.

There was an increase in the crude carbohydrate and crude protein from 25.14% to 26.12% and .35% to 4.84% respectively. The incremental trend for crude carbohydrate and crude protein as reported in this study can be attributed to the degradation of the starch in the fibrous material and increase in the microbial load respectively. This finding agrees with range of values in the work of [1] who reported crude carbohydrate and crude protein values of 25% and 3% respectively for PMF.

There were no significant differences in ash content from the study. The major minerals in the PMF as observed in the study is phosphorous and calcium. The ranges of values for these minerals are similar to those reported also in [1].

### IV CONCLUSION

The result from the study indicated that the use of fermentation and drying processes modified the structure of PMF flour. Fermentation and drying processes resulted in the production of PMF flour with good oil absorption capacity. The PMF flour showed decreases in crude fat and crude fibre while there were increases in moisture content, crude protein and crude carbohydrate. The ash content of the PMF flour was constant. The PMF flour produced from the study thus could serve as raw materials for food product formulations.

The moisture content of food products increased from 16.19% (unfermented PMF) to 19.40% (fermented PMF). The moisture content of food products determined how long they store at room temperature [20]. The obtained result can be associated with the conversion of more of the fibre content of the PMF into soluble dietary fibre and the drying processing method used for drying the PMF flour. According to [24] fermentation increases the soluble dietary fibre content of food material.

### TABLE 2: PROXIMATE COMPOSITION OF THE PMF FLOUR

| Constituent       | unfermented | Fermented Oil Palm Mesocarp Fibre |
|-------------------|-------------|----------------------------------|
|                   | Day 1       | Day 2       | Day 3       | Day 4       | Day 5       | Day 6       |
| Moisture Content  | 16.19±0.31a | 17.67±0.13a | 19.3±0.12b  | 19.4±.18a   | 19.4±0.18b  | 19.4±.18b   |
| Crude Carbohydrate (%) | 25.14±2.11a | 24.69±1.38a | 25.40±1.77a | 25.79±2.06a | 26.12±3.13a | 26.12±3.13a |
| Crude Protein (%)  | 3.35±0.14a  | 3.29±0.11ab | 4.03±0.17b  | 4.43±0.26c  | 4.84±0.34c  | 4.84±0.34c  |
| Crude Fibre (%)    | 17.60±0.02a | 17.28±0.04a | 16.53±0.24a | 16.12±0.05c | 15.72±0.07c | 15.72±0.07c |
| Crude Fat (%)      | 30.17±0.03a | 29.63±0.16a | 27.40±0.15ab| 26.60±0.06b | 26.20±0.06b | 26.20±0.06b |
| Ash (%)            | 7.54±0.11a  | 7.41±0.23a  | 7.25±0.04a  | 7.25±0.04b  | 7.25±0.04b  | 7.25±0.04b  |

Means ± SE (N = 3) means in the column with same superscript differ not significantly (p<0.05)
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