ABSTRACT

Breast carcinomas are a heterogeneous group of malignancy, having variable clinical outcomes depending on their cellular and molecular properties. Tall cell carcinoma with reverse polarity (TCCRP) is a recently described rare entity, which shares morphological features with tall cell variant of papillary thyroid carcinoma but has a distinct morphological, immunohistochemical, and molecular profile. We describe a case of a 40-year-old female patient, who presented with lump in the breast. The patient underwent lumpectomy and was diagnosed as tall cell carcinoma with reverse polarity. Immunohistochemistry and bi-directional Sanger sequencing for IDH2 mutation were used for diagnosis. Tall cell carcinoma with reverse polarity is a rare and newly described entity with characteristic morphological and molecular findings, which carries an excellent prognosis.

Keywords: Breast carcinoma, Tall cell carcinoma, Reverse polarity, Solid papillary, IDH2

INTRODUCTION

Tall cell carcinoma with reverse polarity (TCCRP) is a rare type of invasive breast carcinoma with characteristic immuno-morphologic and molecular findings and is often difficult to diagnose. Eusebi et al., in 2003, reported a rare variety of breast carcinoma having features similar to papillary thyroid carcinoma described as “Breast tumor resembling the tall cell variant of papillary thyroid carcinoma” (1). These tumors were also called breast tumor resembling the tall cell variant of papillary thyroid carcinoma; solid papillary breast carcinoma resembling tall cell variant of papillary thyroid carcinoma; solid papillary carcinoma with reverse polarity (1–5). These tumors were consistently immuno-negative for TTF-1, thyroglobulin and HBME1, and there was no associated thyroid malignancy. Chiang et al. described 13 more cases and also identified the IDH2 hotspot mutations at R172 in these tumors, which has become a defining feature of this entity (4). We are presenting a case report of this rare entity in a 40-year-old female, currently named “Tall cell carcinoma with reverse polarity” by the World Health Organization (6) and there are only scattered case reports and very few case series published in the world literature to the best of our knowledge.
and follicular structures with colloid-like eosinophilic secretion were also noted. Mitosis was sparse (1-2 per 10 high power field). No necrosis, lymphovascular emboli, or perineural invasion were noted (Figure 1A-F).

Immunohistochemistry was performed using an automated stainer (Ventana Benchmark XT system) with UltraView Universal DAB detection. The antibodies used were GATA3, GCDFP-15, estrogen receptor (ER), progesterone receptor (PgR), Androgen receptor (AR), HER2/neu, synaptophysin, chromogranin A, Bcl2, EMA, CK 5/6, p63, thyroid transcription factor 1 (TTF-1), inhibin, and Ki67. Antibody details are shown in Table I. The neoplastic cells were immuno-positive for GATA3, CK5/6, EMA and calretinin with patchy and weak expression for ER and PgR. AR showed heterogenous staining pattern with peripheral accentuation in the tumor lobules (Figure 2A-I), which has

**Figure 1:** Histomorphological features of Tall cell carcinoma with reverse polarity (TCCRP).

A) Tumor composed of compact solid nests in a background of fibrotic stroma, (H&E; x40).

B) Nest and papillae are lined by tall columnar epithelial cells, nuclei exhibiting reverse polarity (arrow) (H&E; x100).

C) Tumor cells have eosinophilic granular cytoplasm, oval nuclei with chromatin clearing, and intranuclear pseudo-inclusion (Inset show intranuclear pseudo inclusion) (H&E; x100).

D) Nuclei exhibit crowding, overlapping and nuclear groove (Inset show nuclear groove) (H&E; x400).

E) Follicular structures with eosinophilic/amphophilic colloid-like secretions (H&E; x100).

F) Fibrovascular core with foamy macrophages (H&E x100).
not been described earlier in the literature. While neoplastic cells were immuno-negative for HER2/neu, synaptophysin, chromogranin, Bcl2, GCDFP-15, TTF1, and inhibin, the Ki67 proliferation index was 5%. No staining for p63 was seen around the lobules. Immunohistochemical features of the tumors are illustrated in Figure 2.

DNA was isolated from formalin-fixed paraffin-embedded blocks and targeted sequencing for the commonly mutated site of IDH1 (R132) and IDH2 (R172) was performed through a bi-directional Sanger sequencing method on the amplified template. The tumor tissue showed a mutation in the IDH2 gene (DNA description: c.A514>G; Protein description: p.Arg172Gly – R172G). DNA sequence chromatogram for the IDH2 gene is shown in Figure 3. Based on the characteristic immuno-morphologic and molecular features, a diagnosis of tall cell carcinoma with reverse polarity was made. Following lumpectomy, completion mastectomy with axillary lymph node dissection was performed owing to the positive surgical margin. Grossly and microscopically, no residual tumor was identified and all nine axillary lymph nodes were free of tumor. PET-CT scan revealed no evidence of metastasis. No further treatment was given owing to the indolent behavior of the tumor, and the patient is under observation and is currently disease-free for 6 months post-surgery. Written informed consent for publication was taken from the patient.

DISCUSSION

Tall cell carcinoma with reverse polarity is a recently described invasive breast carcinoma, included in the 5th edition of WHO classification of breast tumors as a slow-growing tumor with favorable prognosis (1-6). These tumors most commonly occur in women around the age of 65 years (range 52 to 75 years), with characteristic immuno-morphologic features. Molecular studies have shown IDH2 mutation in almost all of these tumors, which has become a defining feature. Morphologically, they form well-circumscribed masses with cells arranged in compact solid nests in a background of fibrotic stroma, having thin fibrovascular cores that recapitulates a solid-papillary pattern, and sometimes the cores contain foamy macrophages. Papillae are lined by tall columnar cells having abundant eosinophilic cytoplasm, bland oval to round nuclei exhibiting nuclear grooves and intranuclear cytoplasmic inclusions (1,3). The most characteristic feature is apical location of the nuclei resulting in reverse polarity. Other features include low mitotic count and absent myoepithelial cells at the periphery and within the papillary fronds. Psammoma bodies, granular calcification, and follicular structures with eosinophilic/amphophilic colloid-like secretions are sometimes noted (7). Areas of necrosis, lymphovascular emboli, or peri-neural invasions are usually not seen (8). The present case showed all the

Table I: Details of primary antibodies.

| Anti-sera  | Source | Clone  | Dilution |
|------------|--------|--------|----------|
| AR         | BioGenex | F39.4.1 | 1:20     |
| Bcl-2      | Bio SB  | BSB-5  | 1:200    |
| Calretinin | BioGenex | 2E7    | 1:30     |
| CK5/6      | Bio SB  | B5-16B4| 1:150    |
| Chromogranin | Bio SB | LK2H10 | 1:4000   |
| EMA        | BioGenex | E29    | 1:120    |
| ER         | Roche  | SP1    | Pre-diluted |
| GATA3      | Bio SB  | L50-823| 1:100    |
| GCDFP-15   | BioGenex | EP95  | 1:50     |
| HER2       | Roche  | 4B5    | Pre-diluted |
| Inhibin    | BioGenex | R1    | 1:30     |
| Ki67       | BioGenex | MIB-1 | 1:50     |
| P63        | Roche  | 1E2    | Pre-diluted |
| Synaptophysin | BioGenex | SMP 88| 1:150    |
| TTF-1      | BioGenex | SP141 | 1:80     |

AR: Androgen receptor, CK: Cytokeratin, EMA: Epithelial membrane antigen, ER: Estrogen receptor, GATA3: GATA binding protein-3, GCDFP-15: Gross cystic disease fluid protein 15, HER2: human epidermal growth factor 2, PgR: progesterone receptor, TTF-1: Thyroid transcription factor 1.
Figure 2: A) Neoplastic cells show weak and patchy expression for ER (IHC; x100) and B) PgR (IHC; x400). C) AR showed heterogenous staining pattern with intense staining at the peripheral of the tumor (IHC; x100). D) Neoplastic cells are positive for CK5/6 (IHC; x40), E) Calretinin (IHC; x40), F) GATA-3 (IHC; x40), G) with luminal expression of EMA (IHC; x400). H) Low Ki67 proliferation (IHC; x100) and I) negative for TTF-1 (IHC; x40).

Figure 3: DNA sequence chromatogram showing mutation in IDH2 gene at R172.
above characteristic morphological features. These tumors show weak or absent ER, PgR, AR expression, and are almost always negative for HER2/neu. Although morphological features of TCCRP resemble papillary thyroid carcinoma, they are negative for TTF-1, thyroglobulin and HBME 1 (9, 10), and the breast origin of the neoplasm is confirmed by the positive staining for markers of mammary differentiation like GATA3, mammaglobin and GCDFP-15 (7,8,10,11). Myoepithelial markers p63, SMMHC, and calponin are absent within and around the lesion, suggestive of invasive disease. Neoplastic cells are also immuno-positive for CK5/6, and focally for EMA, CEA, and bcl-2, and exhibit a low Ki67 proliferation index ranging from 1 to 5% (3,5,8). All TCCRP also show diffuse and strong expression calretinin. The immunohistochemical profile in the present case was similar to the cases described in the literature, with strong expression of GATA3, CK5/6, calretinin, and weak expression of hormone receptors, and AR showed a heterogenous staining pattern with peripheral accentuation in the tumor lobules (Figure 2C), which has not been described earlier in the literature. Ki67 proliferation was low (5%). In addition, the neoplastic cells showed patchy expression of GCDFP15 and luminal expression of EMA (12). Chiang et al. performed molecular analysis on 13 cases of TCCRP using whole-exome and targeted sequencing and found that 77% of cases harbored IDH2 mutation at R172 (4). Alsadoun et al., after genetic and transcriptomic profiling of this tumor using whole-exome analysis, showed that the IDH2 gene harbors a hotspot mutation at the level of arginine 172 (R172), which was not present in any other breast carcinoma, in 78% of TCCRP (7). Although IDH2 mutations are commonly identified in many tumors such as secondary gliomas, myeloid malignancy, cholangiocarcinoma, microsatellite stable colorectal cancer, chondrosarcoma etc., they are infrequent among breast tumors but are typically seen in TCCRP (7, 13-17). Pareja et al. demonstrated that immunohistochemistry using a monoclonal antibody against IDH2 R172S (Clone - 11C8B1) is sensitive and specific for TCCRP harboring the IDH2 R172 hotspot mutation (18). Genetic alterations prevalent in papillary thyroid carcinoma like RET/PTC rearrangements and BRAF mutations (18,19) are almost always absent in TCCRP. Bi-directional Sanger sequencing on the paraffin block showed IDH2 hotspot mutations at R172 in the present case, which is a characteristic and unique molecular finding of this tumor. TCCRP has an indolent clinical behavior with favorable outcomes with surgery being the mainstay of treatment (3,4,8,20). It has the potential for recurrence in case of incomplete excision of the tumor; thus complete excision with adequate margin is the optimal treatment for TCCRP (3). However, two instances of metastasis have been reported in literature: Cameselle-Teijeiro et al. reported one case with bone metastasis and Foschini et al. reported one case with intramammary lymph node metastasis (5,8).

TCCRP should be differentiated from other papillary lesions of the breast like encapsulated or solid papillary carcinoma and invasive micropapillary carcinoma as it shares papillary architecture with lack of myoepithelial cells. TCCRP can be differentiated from these entities by the characteristic morphology of cells with reverse polarity and immuno-positivity for CK5/6, calretinin and negative or weak expression of hormonal receptors. Intraductal papilloma with usual ductal hyperplasia shares nuclear groove and pseudo-inclusion with TCCRP but can be differentiated by the presence of myoepithelial cell layer in papilloma and usual ductal hyperplasia, while TCCRP lacks a layer of myoepithelial cells. In conclusion, TCCRP is a rare and newly described entity with characteristic morphological findings that can mimic other papillary lesions of the breast. IDH2 hotspot mutation is a distinctive and characteristic finding in TCCRP. A high index of suspicion is necessary to avoid misdiagnosis of TCCRP as other papillary breast lesions or metastasis from the thyroid, and accurate diagnosis of this entity is necessary as it carries an excellent prognosis.

CONFLICT of INTEREST
None of the authors have any competing interests.

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AUTHORSHIP CONTRIBUTIONS
Concept: CSP, Design: CSP, Data collection or processing: MJ, Analysis or Interpretation: CSP, GVPO, Literature search: MJ, Writing: MJ, CS, Approval: MJ, CS.

REFERENCES
1. Eusebi V, Damiani S, Ellis I, Azzopardi J, Rosai J. Breast tumor resembling the tall cell variant of papillary thyroid carcinoma. The American Journal of Surgical Pathology. 2003;27:1114-8.
2. Tosi A, Ragazzi M, Ascoli S, Del Vecchio M, Cavalieri M, Eusebi LH, Foschini MP. Breast tumor resembling the tall cell variant of papillary thyroid carcinoma: Report of 4 cases with evidence of malignant potential. International Journal of Surgical Pathology. 2007;15:14-9.

3. Bhargava R, Florea A, Pelmus M, Jones M, Bonaventura M, Wald A, Nikiforova M. Breast tumor resembling tall cell variant of papillary thyroid carcinoma. American Journal of Clinical Pathology. 2017;147:399-410.

4. Chiang S, Weigelt B, Wen H, Pareja F, Raghavendra A, Martelotto L, Burke KA, Basili T, Li A, Geyer FC, Piscuoglio S, Ng C, Jungbluth A, Balss J, Pschuch S, Baker GM, Cole KS, Von Deimling A, Batten JM, Marotti JD, Soh H, McCalip BL, Serrano J, Lim RS, Sziopikou KP, Lu S, Liu X, Hammon T, Brogi E, Snuderl M, Iafra AS, Reis-Filho JS, Schnitt SJ. IDH2 mutations define a unique subtype of breast cancer with altered nuclear polarity. Cancer Research. 2016;76:7118-29.

5. Camuselle-Teijeiro J, Abdulkader I, Barreiro-Morandera F, Ruiz-Ponte C, Reyes-Santias R, Chavez E, Sobrinho-Simoes M. Breast tumor resembling the tall cell variant of papillary thyroid carcinoma: A case report. International Journal of Surgical Pathology. 2006;14:79-84.

6. Yang WT, Bu H, Foschini MP, Schnitt SJ. Tall cell carcinoma with reversed polarity. In: Allison KH, Brogi E, Ellis IO, Fox SB, Morris EA, Sahin A, editors. WHO Classification of Tumours. Breast tumours. Lyon (France): International Agency for Research on Cancer; 2019. 153-4.

7. Alsadoun N, MacGrogan G, Lacroix-Triki M, Bedegdian I, Koeb M, El Alam E, Medioni D, Parent M, Wuthier P, Robert I, Boidot R, Arnould L. Solid papillary carcinoma with reverse polarity of the breast harbors specific morphologic, immunohistochemical and molecular profile in comparison with other benign or malignant papillary lesions of the breast: A comparative study of 9 additional cases. Modern Pathology. 2018;31:1367-80.

8. Foschini M, Ascoli S, Foreid S, Cserni G, Ellis I, Eusebi V, Rosai J. Solid papillary breast carcinomas resembling the tall cell variant of papillary thyroid neoplasms: Applications in the diagnosis of tall cell carcinoma. The American Journal of Surgical Pathology. 2017;41:887-95.

9. Toss M, Billingham K, Egbuniwe I, Moreno F, Abass A, Rakha E. Breast tumours resembling the tall cell variant of thyroid papillary carcinoma: Are they part of the papillary carcinoma spectrum or a distinct entity? Pathobiology. 2018;86:83-91.

10. Pitino A, Squillaci S, Spairani C, Rassu PC, Cosimi MF. Tall cell variant of papillary breast carcinoma: An additional case with review of the literature. Pathologica. 2017;109:162-7.

11. Xiao Xian T, Baohua Y, Xiaoli X, Yifan C, Xiaoyu T, Hongfeng L, Rui B, Xiagie S, Ruohong S, Wentao Y. Characterisation of GATA3 expression in invasive breast cancer: Differences in histological subtypes and immunohistochemically defined molecular subtypes. Journal of Clinical Pathology. 2017;70:926-34.

12. Darb-Esfahani S, von Minckwitz G, Denkert C, Ataseven B, Göbel B, Mehta K, Kalteke G, Rüdiger T, Pflitzer B, Kittel K, Fiedler B, Baumann K, Moll R, Dietel M, Eidtmann H, Thomassen C, Loibl S. Gross cystic disease fluid protein 15 (GCDPF-15) expression in breast cancer subtypes. BMC Cancer. 2014;14:546.

13. Cohen A, Holmen S, Colman H. IDH1 and IDH2 mutations in gliomas. current neurology and neurosience reports. 2013;13:345.

14. Green C, Evans C, Zhao L, Hills R, Burnett A, Linch D, Gale RE. The prognostic significance of IDH2 mutations in AML depends on the location of the mutation. Blood. 2011;118:409-12.

15. Borger D, Tanabe K, Fan K, Lopez H, Fantin V, Straley K, Schenkein DP, Hezel AF, Ancukiewicz M, Liebman HM, Kwak EL, Clark JW, Ryan DP, Deshpande V, Dias-Santagata D, Ellisen LW, Zhu AX, Iafra AS, Freid PK. Frequent mutation of isocitrate dehydrogenase (IDH1) and IDH2 in cholangiocarcinoma identified through broad-based tumor genotyping. The Oncologist. 2011;17:72-79.

16. Whitehall V, Dumenil T, McKeone D, Bond C, Bettington M, Buttenshaw R, Bowdler L, Montgomery G, Wockner L, Leggett B. Isocitrate dehydrogenase 1R132C mutation occurs exclusively in microsatellite stable colorectal cancers with the CpG island methylator phenotype. Epigenetics. 2014;9:1454-60.

17. Amery M, Bacci K, Maggiani F, Damato S, Halai D, Berisha F, Pollock R, O'donnell P, Grigoriadis A, Dass T, Eskandarpour M, Presneau N, Bogendoom PC, Futeat A, Tirabosco R, Flanagan AM. IDH1 and IDH2 mutations are frequent events in central chondrosarcoma and central and periosteal chondromas but not in other mesenchymal tumours. The Journal of Pathology. 2011;224:334-43.

18. Pareja F, da Silva EM, Frosina D, Geyer FC, Lozada JR, Basili T, Co Cruz Paula A, Zhong E, Derakshshan F, Dal'fonso T, Wen HY, Giri DD, Hayes MM, Kriangs K, Bhargava R, Palazzo JP, Rakha EA, Hoda SA, Sanders ME, Collins LC, Schnitt SJ, Chen Y, Weigelt B, Jungbluth AA, Reis-Filho JS, Brogi E. Immunohistochemical analysis of IDH2 R172 hotspot mutations in breast papillary neoplasms: Applications in the diagnosis of tall cell carcinoma with reverse polarity. Mod Pathol. 2020;33:1056-64.

19. Basolo F, Giannini R, Monaco C, Melillo R, Carlongagno F, Pancrazi M, Salvatore G, Chiaipetta G, Pacini F, Elisei R, Miccoli P, Pinchera A, Fusco A, Santoro M. Potent mitogenicity of the RET/PTC3 oncogene correlates with its prevalence in tall-cell variant of papillary thyroid carcinoma. The American Journal of Pathology. 2002;160:247-54.

20. Czarniecka A, Oczko-Wojciechowska M, Barczyński M. BRAF V600E mutation in prognostication of papillary thyroid cancer (PTC) recurrence. Gland Surgery. 2016;5:495-505.