Targeting Lyn Kinase in Chorea-Acanthocytosis: A Translational Treatment Approach in a Rare Disease

Kevin Peikert 1,2*, Hannes Glaß 1,2, Enrica Federti 3, Alessandro Matte 3, Lisann Pelzl 4,5, Katja Akgün 1, Tjalf Ziemssen 1*, Rainer Ordemann 6, Florian Lang 4, The Network for Translational Research for Neuroacanthocytosis Patients 4, Lucia De Franceschi 3,*,# and Andreas Hermann 1,2,7,8,9,*,#

Abstract: Chorea-acanthocytosis (ChAc) is a neurodegenerative disease caused by mutations in the VPS13A gene. It is characterized by several neurological symptoms and the appearance of acanthocytes. Elevated tyrosine kinase Lyn activity has been recently identified as one of the key pathophysiological mechanisms in this disease, and therefore represents a promising drug target. Methods: We evaluated an individual off-label treatment with the tyrosine kinase inhibitor dasatinib (100 mg/d, 25.8–50.4 weeks) of three ChAc patients. Alongside thorough safety monitoring, we assessed motor and non-motor scales (e.g., MDS-UPDRS, UHDRS, quality of life) as well as routine and experimental laboratory parameters (e.g., serum neurofilament, Lyn kinase activity, actin cytoskeleton in red blood cells). Results: Dasatinib appeared to be reasonably safe. The clinical parameters remained stable without significant improvement or deterioration. Regain of deep tendon reflexes was observed in one patient. Creatine kinase, serum neurofilament levels, and acanthocyte count did not reveal consistent effects. However, a reduction of initially elevated Lyn kinase activity and accumulated autophagy markers, as well as a partial restoration of the actin cytoskeleton, was found in red blood cells. Conclusions: We report on the first treatment approach with disease-modifying intention in ChAc. The experimental parameters indicate target engagement in red blood cells, while clinical effects on the central nervous system could not be proven within a rather short treatment time. Limited knowledge on the natural history of ChAc and the lack of appropriate biomarkers remain major barriers for “clinical trial readiness”. We suggest a panel of outcome parameters for future clinical trials in ChAc.

Keywords: ChAc; neuroacanthocytosis; off-label; dasatinib; TKI
1. Introduction

Chorea-acanthocytosis (ChAc) is a rare neurodegenerative disease of the early adulthood which is characterized by a large spectrum of neurological symptoms and the presence of acanthocytes [1–4]. The autosomal-recessive condition is caused by mutations in the VPS13A gene leading to loss of function of the respective encoded protein “chorein” [5–8]. A disease-modifying therapy is not available yet. Hence, treatment options of the devastating disease remain purely symptomatic [9], even though it causes considerable morbidity, markedly reduced life-span, and severely affects self-determined living.

The clinical phenotype of ChAc is highly heterogeneous. As patients often present with movement disorders like chorea, Parkinsonism, and/or dystonia [2,10,11], ChAc belongs to the group of Huntington’s disease phenocopies [12]. Furthermore, dysarthria and dysphagia, peripheral neuropathy, epilepsy, or cognitive impairment may occur, while tongue and lip biting, self-mutilating behavior, feeding dystonia, or head drops are more specific signs of ChAc [1,2,10,11]. Red blood cell (RBC) acanthocytosis, elevated creatine kinase (CK), and serum neurofilament (sNfL) levels are common laboratory findings [1,2,10,13]. In correlation with the clinical manifestations, loss of striatal medium spiny neurons and distinct cortical neurodegeneration represent the main histopathological characteristics [14,15]. Epidemiological estimations suggest a prevalence of around 1000–5000 cases worldwide [1].

While there is growing evidence that members of the VPS13 protein family are involved in the non-vesicular transport of phospholipids [16–19], the precise function of these proteins in humans remains incompletely understood. It is most likely that impaired lipid transfer, and consequently, disturbed organelle lipid homeostasis, contributes to neuronal dysfunction in this disease [20]. So far, VPS13A has been implicated in a variety of important cellular processes, e.g., regulation of cytoskeletal architecture, exocytosis, autophagy, Na+/K+ pump capacity, and Ca\(^{2+}\) homeostasis, and therefore, overall cell survival [4,21–26]. Hence, lack of functional VPS13A leads to impaired cellular homeostasis, particularly resulting in acanthocytosis and neurodegeneration. We recently identified two mechanisms that are considered to be key drivers of ChAc pathophysiology due to VPS13A deficiency: decreased phosphoinositide-3-kinase signaling and increased activity of Src family tyrosine kinase Lyn (for review, see [4]).

In previous studies, we found that ChAc RBCs and induced pluripotent stem cell (iPSC)-derived nerve cells are involved in the non-vesicular transport of phospholipids [16–19], the precise function of these proteins in humans remains incompletely understood. It is most likely that impaired lipid transfer, and consequently, disturbed organelle lipid homeostasis, contributes to neuronal dysfunction in this disease [20]. So far, VPS13A has been implicated in a variety of important cellular processes, e.g., regulation of cytoskeletal architecture, exocytosis, autophagy, Na+/K+ pump capacity, and Ca\(^{2+}\) homeostasis, and therefore, overall cell survival [4,21–26]. Hence, lack of functional VPS13A leads to impaired cellular homeostasis, particularly resulting in acanthocytosis and neurodegeneration. We recently identified two mechanisms that are considered to be key drivers of ChAc pathophysiology due to VPS13A deficiency: decreased phosphoinositide-3-kinase signaling and increased activity of Src family tyrosine kinase Lyn (for review, see [4]).

In previous studies, we found that ChAc RBCs and induced pluripotent stem cell (iPSC)-derived nerve cells are characterized by the accumulation of active Lyn: hyperactive Lyn kinase hyperphosphorylates membrane proteins in RBCs, e.g., band 3, which are involved in anchoring the membrane to the cytoskeletal network. This results in mechanical instability of the membrane [22,27]. Accumulation of active Lyn was also found to be related to impairment of autophagy in erythrocytes [22]. Consistent with that, perturbations of autophagy processes have been reported in in vitro cell models that are defective for VPS13A [17,28]. In a previous study, we showed that in vitro treatment of ChAc RBCs with Src family kinase inhibitors reduces Lyn kinase activity, improves autophagy, and restores the morphological phenotype [22]. Moreover, Src family kinase inhibition also attenuated pathologically enhanced synaptic transmission in striatal medium spiny neurons derived from patient-specific iPSCs [29]. These findings strongly implicate that Lyn kinase should be considered as a promising potential druggable target in ChAc. FDA-approved specific inhibitors of Src family kinases with a reasonable benefit-risk profile, such as the tyrosine kinase inhibitor (TKI) dasatinib, were successfully established in the treatment of chronic myeloid leukemia [30]. Dasatinib was previously shown to cross the blood–brain barrier in humans [31]. Therefore, these inhibitors are ideal candidates for “repurposing” strategies in this context. TKIs have been evaluated in a variety of other neurodegenerative diseases, e.g., Parkinson’s disease (nilotinib), Alzheimer’s disease (e.g., nilotinib, saracatinib), or Amyotrophic Lateral Sclerosis (masitinib), each with a different pathophysiological rationale [30,32–36].

Here, we evaluate a translational off-label treatment with dasatinib (100 mg/d) in three ChAc patients. To our knowledge, this is the first implemented potentially disease-
modifying approach in this ultra-rare disease. Alongside thorough safety monitoring, we regularly performed assessments of treatment efficacy. Therefore, we designed a novel ChAc-related panel of read out parameters, also challenging current concepts of disease markers.

2. Materials and Methods

2.1. Patients and Dasatinib Treatment

We treated three ChAc patients (P1–3) off-label with the FDA-approved tyrosine kinase inhibitor (TKI) dasatinib (DRKS00023177). Patient characteristics are shown in Table 1 and reflect the variety of ChAc phenotypes. Diagnosis was based on clinical manifestations, the absence of chorein in Western blot analysis, and genetic testing [37]. All patients gave their informed consent for off-label treatment with dasatinib, including the risk of possible life-threatening adverse reactions, as well as for video documentation and publication of the data. Patients and healthy control blood donors were enrolled in ongoing studies on the pathogenesis and natural history of neurodegenerative diseases approved by the institutional review board of the Technische Universität Dresden, Germany (EK 45022009, EK 78022015, EK517122019). The standard dose of 100 mg dasatinib per day was administered orally.

Table 1. Demographic characteristics, main clinical observations, and adverse reactions/events in chorea-acanthocytosis patients treated with dasatinib.

|                      | P1          | P2          | P3          |
|----------------------|-------------|-------------|-------------|
| **Sex**              | male        | male        | female      |
| **Age at beginning of treatment (Y)** | 27          | 32          | 49          |
| **Age at onset (Y)** | – symptoms  | – 14        | – 18        | – 23        | – 23        | – 21        | – 39        |
| **Current clinical presentation** | refractory epilepsy, cognitive impairment, mild chorea | refractory epilepsy, muscle atrophy, areflexia, mild chorea, cognitive impairment | marked parkinsonism, epilepsy, cognitive impairment, dystonia, areflexia |
| **Laboratory findings** | elevated levels of CK, AST, ALT, cTroponin T, LDH, acanthocytosis | elevated levels of CK, AST, ALT, cTroponin T, LDH, acanthocytosis | elevated levels of CK, cTroponin T, LDH, lowered level of haptoglobin, acanthocytosis |
| **Time on dasatinib treatment (weeks)** | 50.4        | 48.7        | 25.8        |
| **Comedication**     | Lacosamide 2 × 300 mg, Mirtazapine 1 × 15 mg, Vitamin D and Calcium, PRN: Lorazepam/Midazolam | Lacosamide 2 × 300 mg, Zonisamide 2 × 200 mg, PRN: Lorazepam/Midazolam | Levetiracetam 2 × 2000 mg, Valproate 2 × 1000 mg, Clobazam 2 × 5 mg, Zonisamide 2 × 100 mg, Vitamin D |
| **Main clinical observations during dasatinib treatment** | improvement of short-term memory reported by the caregivers | reappearance of deep tendon reflexes, variation of seizure frequency as known/usual before treatment | variation of seizure frequency as known/usual before treatment |
Table 1. Cont.

| P1 | P2 | P3 |
|----|----|----|
| Adverse reactions or events | • mild intermittent diarrhea and abdominal pain | • mild intermittent diarrhea and abdominal pain | • Acne and discrete alopecia increased frequency of defecation |

PRN—pro re nata; Y—years.

2.2. Evaluation of Dasatinib Treatment

Evaluation of the off-label treatment primarily included monitoring of potential adverse reactions or events, as well as outcome assessments in the context of routine care adjusted to the clinical phenotype (UHDRS, MDS-UPDRS, seizure frequency, blood acanthocyte, and CK level). Evaluation was performed initially every 2 weeks, after 8 weeks of treatment monthly, and after 6 months of treatment every 2 months. At every visit, each patient was assessed with a clinical examination and medical history (including the caregivers’ observations), the Unified Huntington’s Disease Rating Scale Total Motor Score (UHDRS-TMS), Total Functional Capacity (UHDRS-TFC), and Functional Assessment Scale (UHDRS-FA), as well as the Movement Disorders Society Unified Parkinson Disease Rating Scale (MDS-UPDRS) parts I-III and the Clinical Global Impression (CGI) scale by a specialist experienced in the care of patients with ChAc and other movement disorders. Patients were asked to complete the McGill Quality of Life Single-Item Scale (McGill-QoL; range 0–10) and the Schedule for the Evaluation of Individual Quality of Life-Direct Weighting (SEIQoL) questionnaires. Routine electroencephalography was performed. Seizure frequency was defined as number of seizures within the preceding month. At baseline, after 2 months, and as indicated, patients underwent electrocardiography, echocardiography, and abdominal ultrasound for safety monitoring. Routine laboratory chemistry/hematology tests were also performed on blood samples obtained by venipuncture.

2.3. Serum Neurofilament Light Chain Quantification

Serum samples were stored at −20 °C directly after collection, since the neurofilament light chain is stable during freezing process [38,39]. The sNfl measurement was performed using the Advantage NF-Light singleplex Kit and prepared as defined in the manufacturer’s instructions (Quartered, Lexington, MA, Datasheet Quanterix: Simoa™ NF-Light Advantage Kit) as previously described [40,41] with the single molecule array (SIMOA) analysis. Both the mean intra-assay coefficient of variation of duplicates and the mean inter-assay coefficient of variation were <10%.

2.4. Immunoblot Analysis

Additional EDTA-blood samples (ChAc patients and healthy control donors) were shipped to Verona, Italy at 4 °C and processed immediately after arrival. Lyn activity was determined by Western blot analysis using an anti-phospho-Lyn (Y396) antibody, as described previously [27]. We evaluated the amount of ULK1 and p62, known markers of autophagy in RBCs, as reported previously [22,42,43].

2.5. Immunofluorescence

Additional EDTA-blood samples (ChAc patients and healthy control donors) were shipped to Tübingen, Germany at 4 °C and processed immediately after arrival. The erythrocytes were stained with an anti-β-Actin-FITC-conjugated antibody (1:50; biorbyt) and Phallolidin-eFluor660 (1:100, eBioscience) to detect filamentous actin (F-actin) as described previously [25,44]. Confocal microscopy was performed with a Zeiss LSM 5 EXCITE confocal laser-scanning module (Carl Zeiss). The images were analyzed with the software of the instrument.
2.6. Osmotic Fragility of Red Blood Cells

In previous reports, we showed that red cell osmotic fragility is increased in patients with ChAc compared to healthy controls [22,27,45,46]. We evaluated osmotic fragility in EDTA blood using a single osmotic point at 158 mOsm. Erythrocytes from healthy controls were always analyzed in the same experiments with ChAc RBCs.

2.7. Statistical Analysis

Data were analyzed with GraphPad Prism 5 software. Statistical analysis was made by analysis of variance (ANOVA), and \( p < 0.05 \) was considered as statistically significant.

3. Results

3.1. Dasatinib Treatment Was Safe

Overall, dasatinib treatment was safe in all three patients (Table 1). No severe adverse reactions or events were reported. Because of discrete acne and mild alopecia as well as increased defection frequency, P3 and her caregivers asked for the discontinuation of treatment after 25.8 weeks. Otherwise, P1 and P2 reported irregular defection including episodes of mild diarrhea and abdominal pain, but they both stayed on dasatinib treatment for 48.7 and 50.4 weeks, respectively.

We observed a reversible decrease of RBC level and hematocrit, but no manifest anemia in all three patients in the first weeks after treatment initiation (Figure 1A,B). No major change in haptoglobin, a marker of hemolysis, was detected during dasatinib administration (Figure 1C); the mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH), and red blood cell distribution width (RDW) increased slightly under medication, especially in P3 (Figure 1D–F). In the first week of treatment, we observed a transient thrombocytopenia (minimum 107 GPl/L in P1). No relevant neutropenia <1.5 GPl/L nor thrombopenia <100 GPl/L occurred. After discontinuation of dasatinib, there was a drop in hemoglobin and haptoglobin, suggesting a transient hemolytic episode in P1 and P2 (Figure 1B,C). Taken together, these recorded mild symptoms and laboratory abnormalities are known adverse reactions/events of dasatinib, which did not require us to discontinue the treatment [47]. Dasatinib was withdrawn after 50.4 (P1) and 48.7 (P2) weeks for re-evaluation purposes as predetermined with the health insurance.

Figure 1. Graphs (A–F) represent hematology parameters. Patients 1–3 (P1–3), PRE represents baseline value before initiation of dasatinib treatment, dotted vertical lines represent individual end of treatment. Mean corpuscular volume (MCV); mean corpuscular hemoglobin (MCH); red blood cell distribution width (RDW).
3.2. Disease Progression Remained Stable during Dasatinib Treatment in ChAc Patients

During dasatinib treatment, neither significant clinical improvement nor deterioration of neurological manifestations was observed in the three ChAc patients. Caregivers of P1 reported an improvement of short-term memory. In P2, who was initially noted to have areflexia, the re-appearance of patellar (after 2 weeks) and bicep (after 4 weeks) deep tendon reflexes was a striking observation (Table 1).

Quality of life improved for a short time period after initiation of treatment (Figure 2A,B), but dropped to baseline level before treatment was withdrawn, most likely representing a placebo effect. Seizure frequency (Figure 2C) fluctuated widely and was associated with physical and mental stress in P1 and P2. Unified Huntington’s Disease Rating-Scale (UHDRS) and Movement Disorders Society Unified Parkinson Disease Rating Scale (MDS-UPDRS) scores remained stable without clinically relevant progression (Figure 2D-I). Electroencephalograms at baseline and under treatment revealed no significant changes during treatment.

Figure 2. Graphs represent (A) Schedule for the Evaluation of Individual Quality of Life-Direct Weighting (SEIQoL); (B) McGill Quality of Life Single-Item Scale (McGill); (C) seizure frequency per month; (D) Unified Huntington’s Disease Rating-Scale Total Functional Capacity (UHDRS TFC); (E) UHDRS Functional Assessment Scale (UHDRS FA); (F) UHDRS Total Motor Scale (UHDRS TMS); (G) Movement Disorders Society Unified Parkinson’s Disease Rating Scale part I (UPDRS I); (H) MDS-UPDRS part II (UPDRS II); and (I) MDS-UPDRS part III (UPDRS III). Patients 1–3 (P1–3), PRE represents baseline value before initiation of dasatinib treatment, dotted vertical lines represent individual end of treatment.

3.3. Identification of Robust Biomarkers Is Still an Unmet Need in Chorea-Acanthocytosis

We next investigated whether dasatinib treatment alters current biomarker candidates of the disease. Of those, the amount of acanthocytes showed severe variations. The acanthocyte level of P1 (range 21.4%–42.0%, at baseline 35.2%) and P2 (range 19.4%–44.6%, at baseline 24.2%) varied markedly during the treatment without a clear trend. P3 (range
12.7%–28.9%, at baseline 13.5%) showed a trend to increased acanthocyte levels under treatment and subsequent decrease after dasatinib withdrawal (Figure 3A). However, limitations of the accuracy of the acanthocyte testing (manual counting) and pre-analytical variations (transport time to the laboratory, etc.) have to be taken into account just like the intra-individual variations on the amount of acanthocytes in ChAc.

Elevated creatine kinase (CK) persisted in P1 and P2 throughout the study, but increased slightly in P3 (Figure 3B). As CK markedly fluctuated due to athletic activity and tonic-clonic seizure activity, it was not a robust clinical marker of clinical severity in ChAc. Alanine aminotransferase (ALAT) and aspartate aminotransferase (ASAT) levels correlated with CK, while gamma-glutamyltransferase (GGT) and bilirubin remained rather stable within the physiological range, suggesting a likely muscular origin of the increased ASAT and ALAT activities (Figure 3C,D).

Neurofilament light chain (Nfl) was evaluated here as a biomarker for neuroaxonal damage [13,48]. As shown in Figure 3E, sNfl varied through the observation period (16.7–22.7 pg/mL in P1; 15.5–23.9 pg/mL in P2; 14.6–20.7 pg/mL in P3). While sNfl concentrations showed a trend to decrease under treatment in P2, the levels of P1 and P3 increased. The concentration of other peripheral markers for neuronal destruction, S100 and neuron-specific enolase (NSE), were subject to fluctuation without an evident relevant trend (Figure 3F).

### 3.4. Dasatinib Treatment Prevented Accumulation of Lyn and Autophagy-Related Proteins and Improved Features of ChAc RBCs

In RBCs from all three ChAc patients, dasatinib markedly reduced the amount of initially increased active Lyn (phospho-Lyn) to levels lower than in the healthy controls (Figure 4A–C). Active Lyn abundance progressively increased after dasatinib was withdrawn. It is of note that in P1 and P2 (Figure 4A,B) we observed a slight and temporary increase in Lyn activation after 12 and 14 weeks of treatment, respectively, to values similar to those of controls, but still lower compared to baseline.
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Figure 4. Graphs represent red cell features in ChAc patients. (A–C) Total Lyn was immunoprecipitated from blood samples and detected with an antibody against active Lyn (phospho-Lyn 396) or total Lyn (Wb: Western blot). Initially overactive Lyn was suppressed by dasatinib treatment in all three patients. Lyn activity increased after dasatinib withdrawal. (D–F) Western blot (Wb) analysis of Ulk1 and p62 in red blood cells (RBCs) and densitometric analyses of the immunoblot bands. (G) Confocal microscopy of microfilament actin-fluorescence and transmission light microscopy of RBCs from patient 2 (P2) and the healthy control. Anti-β-Actin-FITC-conjugated antibody-stained total actin and Phalloidin-stained filamentous actin (F-actin). (H) Phalloidin was used for F-actin staining. Scale bar: 10 μm. (I) F-actin staining intensity in P1–P3. Shown are the mean values of intensity ± SEM from n = 3 independent experiments. One-way ANOVA/Bonferroni’s multiple comparison test (p < 0.05). (J–L) Osmotic fragility of RBCs is shown as percentage of RBC lysis at a single osmotic point at 158 mOsm in P1–P3. Patients 1–3 (P1–3), PRE: represents baseline value before initiation of dasatinib treatment, dotted vertical lines represent individual end of treatment.
Autophagy initiator ULK1 markedly decreased after 4 weeks of treatment in all patients (Figure 4D–F) and increased after dasatinib withdrawal. The accumulation of the late-phase autophagy marker p62 markedly decreased between 14 and 24 weeks of treatment in P1 (Figure 4D), and between 12 and 22 weeks of dasatinib in P2 (Figure 4E). In contrast, p62 accumulation in P3 RBCs was only temporarily reduced versus baseline (Figure 4F) and re-accumulation along with active Lyn before the scheduled interruption of dasatinib treatment was consistent with possibly uncompliant dosing.

We revealed a severe loss of cortical actin in RBCs from P2 and P3 as compared to the healthy controls at baseline, whereas the cortical actin network was less impaired in P1 (Figure 4G,H). Consistent with the findings described above, we observed a slight but significant increase of F-actin staining in RBCs from P2 and P3 (Figure 4H,I) associated with improvement/normalization of osmotic fragility (Figure 4J–L). After withdrawal of dasatinib, osmotic fragility again progressively deteriorated (Figure 4J–L).

4. Discussion

Chorea-acanthocytosis is a devastating neurodegenerative multi-system disorder without any established disease-modifying treatment. In our previous work, we showed that elevated Lyn kinase activity represents a pathophysiological hallmark of the disease [22,27,29]. FDA-approved TKIs like dasatinib with specificity to the Lyn kinase are therefore promising candidates for a potential disease-modifying treatment. Herein, we report on three phenotypically different ChAc patients who were prescribed dasatinib and monitored for potential adverse reactions or events [47] and therapy response.

Dasatinib successfully engaged the target Lyn kinase with subsequent normalization of ChAc RBC features, but neither showed positive nor negative effects on central nervous system parameters in the three ChAc patients. Oral administration of 100 mg dasatinib once a day appeared to be reasonably safe in subjects with ChAc. Only rather mild adverse reactions including gastrointestinal and cutaneous problems occurred, and hematological adverse reactions were transient and well tolerated.

Dasatinib sufficiently inhibited Lyn kinase at least in RBCs. In all patients, the phospho-Lyn/total Lyn ratio in erythrocytes was pathologically increased at baseline, consistent with our previous findings [22,27]. The ratio markedly decreased under dasatinib treatment. Moreover, Lyn kinase inhibition improved autophagy as supported by the reduction of accumulated autophagy-related proteins [22], and led to a rearrangement of the initially severely impaired cortical filamentous actin network. Together with the reduction of the osmotic fragility, these findings implicate improved mechanical properties of ChAc RBC membranes in dasatinib-treated subjects.

On the other hand, clinical motor and non-motor parameters could not show a clear treatment effect. Considering the rather slow progression rate of ChAc, however, the absence of an immediate short-term improvement with such a disease-modifying approach is not surprising. Since limited knowledge regarding the natural history and the heterogeneity of the disease course complicates the interpretation of these data, it remains uncertain if the lack of evident clinical progression in all three patients can be attributed to positive treatment effects. Most notable was the regain of deep tendon reflexes in P2, which might point to a neuromuscular (peripheral) treatment response. However, the variation of deep tendon reflexes in principle limits the significance of this finding and more objective parameters such as nerve conduction velocity studies (including H-reflex) should be performed in future studies. sNfL levels in cerebrospinal fluid or in serum reflects neuroaxonal damage both of the central and the peripheral nervous system [48] and is known to be elevated in ChAc [13]. Under dasatinib treatment, there was a trend towards decreased sNfL concentration in the patient who regained deep tendon reflexes; however, there was an increase in the other two patients. It is not clear if this course is due to the therapy, the natural history of the disease or if it reflects intraindividual variations. Therefore, sNfL and other brain damage markers needs to be further established as biomarker for ChAc by long-term profiling of the individual levels.
One main finding of the study is that currently believed evident read-out parameters for ChAc (e.g., acanthocyte count, elevated CK, seizure frequency) which were hypothesized to show possible short time effects all failed to be useful in a stringent longitudinal follow-up of these patients. Particularly CK turned out to be subject of pronounced fluctuations due to athletic activity and bilateral convulsive seizures. Also, the amount of acanthocytes could not be used as parameter to evaluate the drug effect due to limitations of the accuracy of the acanthocyte testing and significant intraindividual variations on the amount of acanthocytes in ChAc. Thus, all these parameters are not as robust as expected, which—combined with the slow progression rate of the disease—makes it currently difficult to perform clinical studies in this population of patients. With respect to the limited knowledge of the natural history, defining appropriate read-out parameters remains challenging. This is still, however, a classical translational roadblock in ultra-rare diseases in general. Larger observational studies, such as worldwide case registers like the Neuroacanthocytosis database, a submodule of the European Huntington’s Disease Network (https://www.euro-hd.net/html/na/submodule), need to be further supported and enlarged to establish robust biomarkers and to enable “clinical trial readiness”. Our study represents an additional contribution to achieve this objective, as we suggest a novel panel of outcome parameters for future clinical studies and for cohort characterization in ChAc.

Further open questions relate to the duration and time point of the treatment. It can be assumed that disease modification is more effective in the early stages of disease. However, all of our patients already experienced the first symptoms 9 to 28 years prior to dasatinib therapy, which might limit its effects. Considering the rather low progression rate of ChAc, longer treatment periods might be necessary to achieve further results.

5. Conclusions

In summary, this is the first report on a potentially disease-modifying treatment approach in ChAc. Our observations during an off-label treatment suggest that dasatinib is safe and may successfully target pathologically accumulated Lyn kinase, at least in RBCs. However, only a very small number of patients were treated over a rather short time period, which are two major limitations of this study. Nevertheless, our observations warrant moving forward with the evaluation of Lyn kinase inhibitor effects in larger studies. This study also reveals the need for further investigations of the natural history and robust biomarkers of the disease.

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Institutional Review Board Statement: Patients and healthy control blood donors were enrolled in ongoing studies on the pathogenesis and natural history of neurodegenerative diseases approved by the institutional review board of the Technische Universität Dresden, Germany (EK 45022009, EK 78022015, EK517122019).

Informed Consent Statement: All patients gave their informed consent for off-label treatment with dasatinib, including the risk of possible life-threatening adverse reactions, as well as for video documentation and publication of the data.

Data Availability Statement: The data presented in this study are available within this article.
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Abbreviations

Chorea-acanthocytosis (ChAc); Red blood cell (RBC); Creatine kinase (CK); Serum neurofilament (sNFL); Induced pluripotent stem cell (iPSC); Tyrosine kinase inhibitor (TKI); Patients 1–3 (P1–3); Unified Huntington’s Disease Rating Scale Total Motor Score (UHDRS-TMS), Total Functional Capacity (UHDRS-FC) and Functional Assessment Scale (UHDRS-FA); Movement Disorders Society Unified Parkinson Disease Rating Scale (MDS-UPDRS); Clinical Global Impression (GCI); McGill Quality of Life Single Item Scale (McGill-QoL); Schedule for the Evaluation of Individual Quality of Life-Direct Weighting (SEIQoL); Single molecule array (SIMOA); Mean corpuscular volume (MCV); Mean corpuscular hemoglobin (MCH); Red blood cell distribution width (RDW); Alanine aminotransferase (ALAT); Aspartate aminotransferase (ASAT); Gamma-glutamyltransferase (GGT); Neurofilament light chain (Nfl); Neuron-specific enolase (NSE).

Appendix A

Members of The Network for Translational Research for Neuroacanthocytosis Patients (coordinator: Andreas Hermann):

Ezio Bonifacio (Center for Regenerative Therapies Dresden (CRTD), Technische Universität Dresden, Dresden, Germany), Giel Bosman (Department of Biochemistry, Radboud University Medical Center, Nijmegen, The Netherlands), Adrian Danek (Neurologische Klinik, Klinikum der Universität, Ludwig-Maximilians-Universität München, Munich, Germany), Peter Claus (Institute of Neuroanatomy and Cell Biology, Hannover Medical School, Hannover, Germany and Center for Systems Neuroscience (ZSN) Hannover, Hannover, Germany), Jochen Guck (Biotechnology Center, Technische Universität Dresden, Dresden, Germany), Ulrich Salzer (Center for Medical Biochemistry, Max Perutz Labs, Medical University of Vienna, Austria).
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