Live-imaging of astrocyte morphogenesis and function in zebrafish neural circuits

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How astrocytes grow and integrate into neural circuits remains poorly defined. Zebrafish are well suited for such investigations, but bona fide astrocytes have not been described in this system. Here we characterize a zebrafish cell type that is remarkably similar to mammalian astrocytes that derive from radial glial cells and elaborate processes to establish their territories at early larval stages. Zebrafish astrocytes associate closely with synapses, tile with one another and express markers, including Glast and glutamine synthetase. Once integrated into circuits, they exhibit whole-cell and microdomain Ca²⁺ transients, which are sensitive to norepinephrine. Finally, using a cell-specific CRISPR–Cas9 approach, we demonstrate that fgfr3 and fgfr4 are required for vertebrate astrocyte morphogenesis. This work provides the first visualization of astrocyte morphogenesis from stem cell to post-mitotic astrocyte in vivo, identifies a role for Fgf receptors in vertebrate astrocytes and establishes zebrafish as a valuable new model system to study astrocyte biology in vivo.

Glia are critical regulators of nervous system development and function. Glia constitute at least half of the cells in the human brain, and astrocytes are among the most abundant glial cell type in the mammalian central nervous system (CNS). Astrocytes are unusually elaborate cells, and much of their functional associations depend on their highly branched morphology. They extend many fine cellular processes to interact closely with synapses, neuronal cell bodies, axons, blood vessels and other glial cells in the CNS. Through these associations, they fulfill diverse functions to support and enhance neuronal activity, maintain CNS homeostasis and modulate neural circuits. However, it remains poorly understood how astrocytes are specified, how they develop their intricate morphological associations, how astrocyte–neuron interactions influence in vivo functional roles and how diverse their functions might be in different brain regions. Given the growing number of studies demonstrating direct roles for astrocytes in regulating neural circuit function, it is of particular interest to understand how astrocyte–synapse interactions modulate synaptic and circuit function. Astrocyte dysfunction has also been implicated in an array of neurological diseases, although the mechanisms by which changes in astrocyte physiology lead to disease require considerable further study.

Most understanding of astrocyte biology derives from investigations of rodent models, where the molecular and morphological features of astrocytes are well described. There appears to be a striking conservation of astrocyte biology across diverse species, including mouse and Drosophila, suggesting that this is an ancient CNS cell type. Curiously, evidence that zebrafish have bona fide astrocytes has, to date, been lacking. Radial glial cells (RGCs), the precursor of astrocytes in mammals, have instead been proposed in zebrafish to functionally substitute for astrocytes. In zebrafish, RGCs serve as neural progenitor cells throughout life; they typically exhibit bipolar shape with long processes spanning the entire parenchyma; and recent studies argue that a subset indeed plays important roles in modulating zebrafish behavior.

Here we report the discovery and characterization of a cell type in the zebrafish CNS that is remarkably similar to mammalian astrocytes. By generating a stable transgenic line that labels the membrane and nuclei of all Glast+ cells (glutamate aspartate transporter) cells, we found cells with dense cellular processes in both the brain and spinal cord of zebrafish larvae. Using a collection of new tools, we examined the cells with single-cell resolution and demonstrate that they elaborate a dense meshwork of fine cellular processes, morphologically similar to astrocytes in Drosophila and mammals. Time-lapse in vivo confocal microscopy showed that these cells begin to transform from RGCs into astrocyte-like cells at 2 d post-fertilization (dpf) in the spinal cord and display dynamic process elaboration over the course of development. These cells exhibit several additional defining characteristics of mammalian astrocytes, including expression of glutamine synthetase (GS), close association with synapses and astrocyte–astrocyte tiling behavior. By performing in vivo Ca²⁺ imaging, we found that these cells exhibit spontaneous microdomain Ca²⁺ transients in the fine processes with dynamics similar to Ca²⁺ transients in awake behaving mice, and they respond to norepinephrine (NE) application. To begin exploring the molecular basis of astrocyte morphogenesis and function in vertebrates, we developed a cell-specific CRISPR–Cas9 approach that allows rapid disruption of genes of interest in zebrafish astrocytes. Using this approach, we show that Fgf receptors (fgfr3 and fgfr4) are required for astrocyte morphogenesis. Our work establishes zebrafish as a new model system to explore astrocyte development and function, including the first opportunity to live-image astrocyte morphogenesis from birth to maturity in vivo, and provides new insights into the molecular and cellular mechanisms regulating astrocyte development and growth.

Results

Scl1a3b/Glast-expressing cells display dense meshwork morphology in the zebrafish larval brain and spinal cord. With the aim of identifying astrocyte-like glia in zebrafish, we first performed a targeted whole-mount in situ hybridization screen at 6 dpf to examine the expression of marker genes known to be highly enriched in both human and mouse astrocytes. We found that the glutamate transporters skl1a2b (EAAT2a/GLT-1) and skl1a3b...
(EAT1b/Glast) and GABA transporter slc6a11b (GAT-3) are expressed in the zebrafish larval CNS at 6 dpf (Extended Data Fig. 1a). We further characterized their expression patterns at different developmental stages and observed that all three of these genes could be detected at 1 dpf in restricted CNS regions (Extended Data Fig. 1b). At 3 dpf, they express more broadly in the CNS with some overlapping and unique expression patterns (Extended Data Fig. 1b), and brain expression was maintained through later larval stages (Extended Data Fig. 1c). To explore the nature of these potential astrocyte-like cells further, we generated a stable transgenic line, Tg[slc1a3b:myrGFP-P2A-H2AmCherry], in which membrane-targeted myristoyl-GFP (myrGFP) and the nuclear marker H2AmCherry were expressed under the control of the slc1a3b/Glast promoter (Fig. 1a). We observed strong expression of myrGFP and H2AmCherry markers throughout the CNS in 6-dpf larvae (Fig. 1b) and a dense meshwork of myrGFP-labeled cellular processes in brain regions that likely house the synaptic neuropil (Fig. 1c). In the spinal cord, we observed similar complex cellular processes in lateral regions (Fig. 1d–f), the position of which overlapped with the synapse-rich lateral neuropil. Given that astrocytes in flies and mammals intimately associate with synapses12,22, we take the spatial organization of these fine membranes near CNS synapses as further support for the notion that these cells are astrocyte-like cells in zebrafish.

To enable analysis of these Glast+ cells at single-cell resolution, we injected slc1a3b:myrGFP-P2A-H2AmCherry DNA constructs into one-celled zygotes to sparsely label individual cells. With this approach, we could identify RGCs that exhibit bipolar morphology as well as ependymal cells in the CNS ventricular zone (Extended Data Fig. 2). In zebrafish, recent dye-filling studies marked glial cells termed ‘radial astrocytes’ in the medulla oblongata with long processes that ramify at distal ends2. We first tested whether we could detect these cells in the hindbrain using our reporter construct. At 6 dpf, we found labeled cells in the hindbrain with nuclei at the midline and one main process extended laterally with bushy branches (Fig. 1e), similar to the previously reported radial astrocytes2. We noticed that myrGFP is highly expressed in the cerebellum in the transgenic line (Fig. 1c), consistent with the enrichment of Glast expression in mouse cerebellum22. By sparse labeling, we identified RGCs that exhibit bipolar morphology, as well as ependymal cells in the CNS ventricular zone (Extended Data Fig. 2). In zebrafish, recent dye-filling studies marked glial cells termed ‘radial astrocytes’ in the medulla oblongata with long processes that ramify at distal ends2. We first tested whether we could detect these cells in the hindbrain using our reporter construct. At 6 dpf, we found labeled cells in the hindbrain with nuclei at the midline and one main process extended laterally with bushy branches (Fig. 1e), similar to the previously reported radial astrocytes2. We noticed that myrGFP is highly expressed in the cerebellum in the transgenic line (Fig. 1c), consistent with the enrichment of Glast expression in mouse cerebellum22. By sparse labeling, we detected cells with somata sitting ventrally and extending a dense meshwork of processes toward the pial surface in the cerebellum at 6 dpf (Fig. 1f), which are similar to Bergmann glia, a specialized subset of astrocytes, in mammals22. We next analyzed the spinal cord with single-cell clones and found a cell type elaborating membrane projections with increasing density of fine processes laterally (Fig. 1g). In contrast to the radial astrocytes that show thin long processes with bushy ends, these cells began to ramify complex processes in close proximity to their somata. Collectively, these data indicate that there are astrocyte-like cells in the zebrafish larval CNS, with morphology highly similar to astrocytes characterized in Drosophila and mammals. Hereafter, we refer to these cells as zebrafish spinal cord astrocytes.

Zebrafish spinal cord astrocytes dynamically elaborate processes and establish unique spatial domains at early larval stages. As mammalian astrocytes are derived from RGCs during postnatal development, we sought to test whether zebrafish astrocytes derive from RGCs in the spinal cord. Transparent zebrafish larvae offer the unique opportunity to live-image the morphogenesis of single RGCs and astrocytes from birth through late larval stages in vivo. At 1 dpf, clones sparsely labeled with slc1a3b:myrGFP-P2A-H2AmCherry had a radial glial-like morphology, with the nuclear marker H2AmCherry positioned close to the ventricular midline and a long main process extended toward the lateral pial surface. By time-lapse confocal imaging, we found that most of the analyzed cells at 1 dpf differentiated into neurons, consistent with the role of RGCs serving as neural progenitors during early neurogenesis22. The myrGFP/H2AmCherry markers subsequently diminished in the labeled neurons, likely owing to the silencing of slc1a3b/Glast promoter in the neural lineage. However, starting at 2 dpf, we observed individual cells exhibiting dynamic cellular process that became increasingly elaborate over developmental time toward the lateral synaptic neuropil regions of the spinal cord (Fig. 2a,d and Supplementary Video 1). We tracked the dynamics of astrocyte growth by repeatedly imaging individually labeled cells from 2 through 9 dpf in intact larvae (n = 40–59 individual cell clones; n = 20–25 fish analyzed at each stage). We quantified the spatial territory of individual cells and found that astrocytes in the developing spinal cord increased in size to rapidly establish their overall spatial domains between 2 and 4 dpf (1,333 ± 669.5 µm² at 2 dpf versus 2,629 ± 813.7 µm² at 4 dpf; P < 0.0001, one-way analysis of variance (ANOVA) with Tukey’s post hoc test) and then appeared to maintain individual territories thereafter (Fig. 2b,c). Astrocyte expansive growth is, therefore, most robust between 2 and 4 dpf, with subsequent growth not leading to significant expanded domain size (Fig. 2d) but perhaps elaboration of more intimate contacts with synapses, other glia and the vasculature during this later time window.

Spinal cord astrocytes express additional astrocyte markers, elaborate fine processes during synaptic formation and tile with other astrocytes. We next sought to determine whether zebrafish astrocytes share additional cardinal features with mammalian astrocytes. Previous studies in mammals showed that GS is specifically enriched in astrocytes27. We, therefore, performed immunostaining experiments on Tg[slc1a3b:myrGFP-P2A-H2AmCherry] transgenic larvae with an anti-GS antibody. We found that GS was localized in the spinal cord cell somata as well as in the dense processes, which were co-labeled by the astrocytic myrGFP and H2AmCherry markers (Fig. 3a), suggesting that GS is present in these astrocytes. In addition, we compared the expression of slc1a3b with other common mammalian astrocyte markers, gfap and kcnj10a/kir4.1, by in situ hybridization experiments. We found that, in the 3-dpf spinal cord, expression of slc1a3b and kcnj10a were spatially restricted to the ventricular zone, and kcnj10a expression seemed to mostly overlap with myrGFP-labeled astrocytes in the spinal cord (Extended Data Fig. 3a,b). In contrast, we observed that gfap was expressed more broadly throughout the spinal cord compared to slc1a3b or kcnj10a (Extended Data Fig. 3a). Moreover, we detected diminished expression of gfap in 3-dpf spinal larval regions, in contrast to the expression of slc1a3b (Extended Data Fig. 3a,c), suggesting spatiotemporally different regulation of these two genes across the CNS. Together, these data indicate that slc1a3b-labeled astrocytes also express kcnj10a and gfap, with gfap likely expressed in other cell types as well, which is consistent with recent single-cell RNA-sequencing studies in zebrafish29. In the developing and adult CNS, astrocyte processes enwrap synaptic structures and regulate synapse plasticity22,29,30. To determine whether zebrafish astrocytes also associate closely with synapses during development, we examined the spatiotemporal correlation between our transgenic markers and the presynaptic marker synaptic vesicle glycoprotein 2A (SV2). This was performed in the developing spinal cord at 2–4 dpf and 6 dpf, stages at which astrocyte morphologies are rapidly growing or relatively stable, respectively. We found that anti-SV2 staining in the spinal cord increased dramatically from 2 to 6 dpf, and this was highly correlated with the growth of astrocyte processes labeled by myrGFP (Fig. 3b). At 6 dpf, when we examined this relationship more closely with high-resolution microscopy, we found that myrGFP-labeled astrocyte processes were indeed in close apposition to SV2-labeled synaptic structures throughout the spinal cord (Fig. 3c). Taken together, these data indicate that astrocyte growth in the spinal cord is tightly associated with synapse formation in zebrafish, and...
The image contains a page from a scientific paper discussing astrocyte processes in zebrafish. The text reads:

Astrocyte processes cover the entire synaptic neuropil. In the developing mammalian CNS, several studies have demonstrated that astrocyte-derived signals are important to promote synaptogenesis29,31,32. Thus, our data suggest that astrocyte processes need to be proximal to synaptic structures to fulfill their function, consistent with mammalian studies.

Mammalian astrocytes tile with one another to minimize overlap with other astrocytes and ensure full coverage of CNS neuropil33. To test whether zebrafish astrocytes exhibit tiling behavior, we injected two different membrane-labeled DNA constructs driven by the slc1a3b/Glast promoter (slc1a3b:myrGFP and slc1a3b:mCD8mCherry) into one-celled zygotes and looked for labeled neighboring clones that expressed either myrGFP or mCD8mCherry at 6 dpf. We found that, indeed, the slc1a3b:myrGFP-expressing clone is closely apposed to the slc1a3b:mCD8mCherry-expressing clone with limited overlap (Fig. 3d and Supplementary Video 2), suggesting that zebrafish astrocytes tile with each other and likely occupy unique territories.

Zebrafish astrocytes exhibit spontaneous microdomain Ca\(^{2+}\) transients and respond to NE. In the CNS, individual astrocytes interact with thousands of synapses; they also intimately contact neuronal cell bodies, blood vessels and other glial cells. Previous studies have argued that astrocytes can form functionally independent compartments that adapt to local demands and exhibit spontaneous microdomain Ca\(^{2+}\) activities15. To determine whether zebrafish astrocytes are capable of increasing Ca\(^{2+}\) transients locally in the fine processes, and the dynamics of such transitions.
events, we generated a transgenic line, \( \text{Tg[slc1a3b:myrGCaMP6s]} \), in which a membrane-targeted myristoyl-Ca\(^{2+}\) indicator GCaMP6s (myrGCaMP6s) is expressed under the control of the \( \text{slc1a3b}/\text{Glast} \) promoter. By performing time-lapse confocal imaging with this transgenic line at 6 dpf in the larval spinal cord, we observed robust microdomain Ca\(^{2+}\) transients in astrocyte fine processes (Fig. 4a and Supplementary Video 3). We analyzed the Ca\(^{2+}\) events with automated Astrocyte Quantitative Analysis (AQuA) software\(^{34}\) (Fig. 4a–c) and found that microdomain Ca\(^{2+}\) events exhibit diverse ranges of activities: individual events vary in domain area size, amplitude and duration (Fig. 4d–f; \( n = 1,239 \) events, \( n = 6 \) fish analyzed). On average, these events have slow kinetics (5.84 ± 3.81 s), similar to those found in mouse astrocytes\(^{34–36}\).

In mammals, a startle stimulus or direct application of NE has been shown to promote microdomain Ca\(^{2+}\) events\(^{37}\). To test whether zebrafish astrocytes are sensitive to NE signaling, we performed live Ca\(^{2+}\) imaging in the intact larval spinal cord with the treatment of NE or dimethylsulfoxide (DMSO) control at 6 dpf (Supplementary

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**Fig. 2 | Zebrafish spinal cord astrocytes show dynamic cellular process elaboration and establish individual cell territories between 2 and 4 dpf.**

- **a.** Time-lapse still images of an \( \text{slc1a3b:myrGFP-P2A-H2AmCherry} \)-expressing astrocyte in the spinal cord between 2 and 3 dpf. Scale bar, 20 \( \mu \)m. See also Supplementary Video 1.
- **b.** Representative images show the same astrocyte at different developmental stages in the spinal cord. MAX projection, lateral view. Scale bar, 20 \( \mu \)m.
- **c.** Quantification of individual spinal cord astrocyte cell territory between 2 and 9 dpf. ****\( P \leq 0.0001 \); NS, not significant; one-way ANOVA with Tukey’s post hoc test. \( n \), number of fish analyzed. Error bars represent mean values ± s.d.
- **d.** Schematic showing the developmental stages important for spinal cord astrocyte growth.
Videos 4 and 5) (DMSO, n = 8 fish; NE, n = 9 fish). Using AQuA for quantification, we found a significant increase in microdomain Ca\textsuperscript{2+} events in NE-treated spinal cord astrocytes compared to DMSO controls (DMSO, 4.68 ± 1.47 events per minute; NE, 18.80 ± 3.39 events per minute in normalized 1,000 \(\mu\)m\(^2\) area; \(P = 0.0023\), two-tailed unpaired \(t\)-test) (Fig. 4g,h). The average duration of Ca\textsuperscript{2+} transients was increased in NE-treated astrocytes (DMSO, 5.46 ± 4.21 s; NE, 7.42 ± 5.69 s; \(P < 0.0001\), two-tailed unpaired \(t\)-test) (Fig. 4i–k), whereas the average microdomain area appeared unaltered (DMSO, 7.81 ± 0.65 \(\mu\)m\(^2\); NE, 6.81 ± 0.24 \(\mu\)m\(^2\); \(P = 0.0837\), two-tailed unpaired \(t\)-test), and the average amplitude was slightly decreased (DMSO, 0.39 ± 0.08 \(\Delta F/F\); NE, 0.34 ± 0.09 \(\Delta F/F\); \(P < 0.0001\), two-tailed unpaired \(t\)-test). To test whether the response of spinal cord astrocytes to NE signaling requires neuronal activity, we injected tetrodotoxin (TTX) into the yolk of 6-dpf Tg[slc1a3b:myrGCaMP6s] larvae to block voltage-gated sodium channels\(^{38}\). This injection paralyzed the larvae, indicating the efficacy of TTX administration, and injected larvae were treated with NE or DMSO before Ca\textsuperscript{2+} imaging (Extended Data Fig. 4a). We found that TTX treatment had no effect on the increased number of NE-induced microdomain Ca\textsuperscript{2+} events in spinal cord astrocytes (TTX + DMSO, 5.31 ± 0.88 events per minute; TTX + NE, 14.01 ± 2.86 events per minute in normalized 1,000 \(\mu\)m\(^2\) area; \(P = 0.0094\), two-tailed unpaired \(t\)-test) or the increased duration of Ca\textsuperscript{2+} transients in NE-treated astrocytes (TTX + DMSO, 5.15 ± 4.11 s; TTX + NE, 7.62 ± 6.97 s; \(P < 0.0001\), two-tailed unpaired \(t\)-test) (Extended Data Fig. 4b,c).
As a recent study demonstrated that radial astrocytes in zebrafish hindbrain can respond to NE activation to elicit cytosolic Ca\(^{2+}\) events, we also performed Ca\(^{2+}\) imaging experiments in the hindbrain region to test whether mammalian Ca\(^{2+}\) activity can be influenced in radial astrocytes by NE signaling. Consistent with our data in the spinal cord, we observed a similar increase in NE-induced microdomain Ca\(^{2+}\) events in the hindbrain radial astrocyte fine processes (Extended Data Fig. 5 and Supplementary Videos 6 and 7) (DMSO, \(n=8\) fish; NE, \(n=8\) fish). Together, these results show that zebrafish astrocytes exhibit Ca\(^{2+}\) signaling dynamics that are remarkably similar to those in awake behaving mice, and that, in mice, NE promotes microdomain Ca\(^{2+}\) events and enhances the duration of individual events.

A cell-specific CRISPR–Cas9 approach to study astrocyte gene function in vivo. Our data indicate that zebrafish have astrocytes that are morphologically, molecularly and functionally similar to mammalian astrocytes. Given that little is still known about astrocyte development and function in vivo, we aimed to develop a rapid assay in zebrafish to genetically manipulate candidate genes in an astrocyte-specific manner. To this end, we adapted a tissue-specific CRISPR–Cas9 system along with the sparse labeling approach to inactivate genes and assay their function in individual astrocyte clones in the \(F_0\) generation. Briefly, we generated a DNA construct (\(slc1a3b:Cas9;U6:sgrRNA\)) to express \(sgrRNAs\) ubiquitously while driving Cas9 expression under the \(slc1a3b\) promoter. We co-injected our \(slc1a3b:myrGFP-P2A-H2AmCherry\) construct together with \(slc1a3b:Cas9;U6:sgrRNA\) DNA to target genes of interest into zygotes and carried out mosaic analyses of \(myrGFP/H2AmCherry\)-labeled cells in the injected \(F_0\) larvae (Fig. 5a). Owing to the high efficacy of CRISPR–Cas9 in zebrafish, gene function can routinely be studied in the \(F_0\) generation.

As a proof of principle, we decided to test whether Fgf receptors play a role in astrocyte morphogenesis in zebrafish. In \(D.\) Drosophila, astrocyte depletion of Fgf receptor Heartless (Htl) has been shown to lead to strong defects in astrocyte morphogenesis in the developing CNS\(^{17}\). We sought to test whether the function of Fgf signaling is conserved in vertebrate astrocyte development. In zebrafish, there are five homologous genes encoding Fgfr, which became stellate-like astrocytes\(^{42}\). Similarly, in zebrafish, RGCs have become stellate-like astrocytes\(^{42}\). In contrast to mammals, zebrafish RGCs persist in most brain regions, which allowed us to develop a cell-specific CRISPR–Cas9 approach, we demonstrated that \(fgfr3\) and \(fgfr4\) are required in spinal cord astrocytes for proper morphogenesis. This work establishes zebrafish as a powerful new model to study astrocytes, which should allow for rapid forward genetic screening to identify novel genes critical for many aspects of astrocyte development and function.

Discussion

Zebrafish represent an excellent vertebrate model system to study neurodevelopment in vivo. The transparency of zebrafish embryos and young larvae makes them accessible to long-term cell fate tracing in intact animals and to imaging of molecular and cellular behaviors by time-lapse microscopy, but bona fide astrocytes have not been identified in this model system. Here we report a previously undescribed glial cell type in zebrafish with several defining characteristics of mammalian astrocytes, including their intricate bushy morphology, expression of astrocyte markers Glut and GS, close association with synapses, tiling behavior and dynamic global and microdomain Ca\(^{2+}\) transients. Most of the growth of these cells occurs during 2–4 dpf in the spinal cord, and, using a cell-specific CRISPR–Cas9 approach, we demonstrated that \(fgfr3\) and \(fgfr4\) are required in spinal cord astrocytes for proper morphogenesis. This work establishes zebrafish as a powerful new model to study astrocytes, which should allow for rapid forward genetic screening to identify novel genes critical for many aspects of astrocyte development and function.

In mammals, astrocytes are derived from RGCs, which serve as neural progenitors during early brain development\(^{41}\). By late neurogenesis, most RGCs retrace their cell bodies from the ventricles and become stellate-like astrocytes\(^{42}\). Similarly, in zebrafish, RGCs have been characterized in various CNS regions during development\(^{25, 41}\). However, in contrast to mammals, zebrafish RGCs persist in most regions of the adult CNS and are thought to be responsible for the impressive CNS regenerative capacity observed in this species\(^{10, 41}\).
**Figure 1: AQUA Imaging of Neuronal Circuitry**

*a* Tg[slc1a3b:myrGCaMP6s] in the AQUA imaging system. Scale bar, 200 µm.

*b* Area size (µm², log²) vs. fraction of total events.

*c* Amplitude (ΔF/F) vs. fraction of total events.

*d* Duration (s) vs. fraction of total events.

*e* Area size (µm², log²) vs. fraction of total events.

*f* Amplitude (ΔF/F) vs. fraction of total events.

*g* DMSO and NE images showing reporter activity.

*h* Events per minute (normalized to 1,000 µm²).

*i* Area (µm², log²).

*j* Amplitude (ΔF/F).

*k* Duration (s).

**Legend**

- **DMSO** (n = 8)
- **NE** (n = 9)

Significance levels:

- **EE** p < 0.01
- **EE** p < 0.001
- **EE** p < 0.0001

**Note:** The images and graphs illustrate the quantification of neuronal activity in the AQUA imaging system, comparing DMSO and NE conditions.
It has remained controversial whether zebrafish have true astrocytes that have not yet been described or if zebrafish RGCs might perform necessary functions of astrocytes. Our work demonstrates that the zebrafish CNS indeed houses a population of astrocytes very similar to those in mammals and Drosophila. This provides further support for the notion that astrocytes are an ancient, well-conserved CNS cell type. We also noted that spinal cord astrocytes and radial astrocytes in the hindbrain exhibit morphological differences, with hindbrain radial astrocytes maintaining a long main process between the cell body and the dense branches. However, given the similarities in molecular markers and responses to NE signaling, it is possible that spinal cord astrocytes and hindbrain radial astrocytes represent same cell type or closely related cell types in different CNS areas, whereby surrounding cells or structural constraints might play a role in regulating their morphogenesis. In the zebrafish spinal cord, using several new markers, we were able to live-image astrocyte morphogenesis from birth through late larval stages as they establish their unique spatial domains. We found that

**Fig. 5 | Cell-specific inactivation of fgfr3 and fgfr4 disturbs spinal cord astrocyte morphogenesis.**

- **a**, Schematic of the slc1a3b:nlsCas9nls;U6-sgRNA DNA construct used for gene inactivation and the experimental design.
- **b**, Representative images showing the three main classes of labeled clones observed at 6 dpf after injections and the quantifications of each fgfr gene-inactivated cell in comparison with the control sgRNA group. Scale bar, 20 μm.
- **c**, Representative images and Imaris-generated three-dimensional models of control, fgfr3 and fgfr4-disrupted astrocytes. Scale bar, 20 μm.
- **d**, Quantification of individual astrocyte volumes in control, fgfr3 and fgfr4-disrupted cells in the 6-dpf spinal cord. Error bars represent mean values ± s.d. Control, n = 33 cells in 18 fish analyzed; fgfr3, n = 68 cells in 33 fish analyzed; fgfr4, n = 21 cells in 12 fish analyzed. *P < 0.05. For control versus fgfr3, P = 0.0131; for control versus fgfr4, P = 0.0379. Two-tailed unpaired t-test.
astrocytes show dynamic extension and retraction of cellular processes, rapidly occupy their final spatial domains and elaborate fine processes concomitant to the onset of synapse formation.

Understanding how astrocytes elaborate their remarkable morphologies, and how much of this process is guided intrinsically versus extrinsically, will be essential to understand their functions. Many of the key proposed functions for astrocytes require specialized cell–cell associations, such as endfeet on the vasculature or associations with synaptic elements. In mammals, the intimate association of astrocytes with neuropil is critical for synaptogenesis during development[13,19,20,42] and also later in mature circuits for neurotransmitter clearance and modulation of synaptic activity[9,40]. Although the precise roles for astrocyte Ca\(^{2+}\) signaling remain a point of controversy, microdomain Ca\(^{2+}\) transients have been implicated in local neural circuit control in multiple species[12-14]. Here we showed that zebrafish astrocyte Ca\(^{2+}\) transients are enhanced by NE stimulation (Fig. 4 and Extended Data Figs. 4 and 5). Our data, therefore, strongly suggest that zebrafish astrocytes function similarly to mammalian astrocytes with respect to Ca\(^{2+}\) signaling dynamics. Given the genetic tractability of zebrafish, along with our ability to live-image signaling in intact animals, a deep analysis of the mechanistic basis of Ca\(^{2+}\) signaling in zebrafish neural circuit function should provide exciting new insights into their function.

Little is understood about the signaling pathways that drive astrocyte morphogenesis and association with other CNS cell types. We have shown that astrocytes are derived from RGCs in the zebrafish spinal cord and, over 2–4 d, expand to occupy their unique spatial domains and then tile with one another (Fig. 2 and Supplementary Video 1). By developing a cell-specific CRISPR-Cas9 approach, we discovered that inactivating fgfr3 and fgfr4 in astrocytes abolished normal process infiltration in zebrafish (Fig. 5). In Drosophila, the fgfr homolog htl is critical for astrocyte growth and interaction of astrocyte processes with synapse-rich neuropil. Htl loss resulted in astrocytes with a similar less-intricate morphology that failed to elaborate fine processes in the synaptic neuropil[15]. In mammals, Fgf signaling has been shown to play a role in neural progenitors to switch cell fate from neurons to astrocytes[16]. In addition, Fgfr3 has been found to be highly expressed in both human and mouse astrocytes[12,13,14]. However, whether Fgf signaling plays a role and which Fgf receptors (Fgfr1–4) are responsible for astrocyte morphogenesis in vertebrates remain unclear. Our data support the notion that Fgf signaling is a conserved feature of astrocyte morphogenesis in vertebrates and particularly of Fgfr3 and Fgfr4 in the spinal cord.

Astrocytes are integral components of neural circuits, modulating neuronal function in a variety of ways, but we still lack basic knowledge of astrocyte development and function in vivo. Studying astrocyte biology in vivo is crucial, as astrocytes radically transform their phenotypes when placed in cell culture. This work establishes zebrafish as a new system in which to explore astrocyte biology. Zebrafish astrocytes develop over the course of only a few days; one can exploit the battery of genetic tools available in zebrafish to explore gene function; and imaging and manipulation can be performed with single-cell resolution, through the entirety of astrocyte development. As whole-brain imaging is becoming quite common in zebrafish, even in animals executing simple behaviors, our work lays the foundation for future studies to link specific changes in astrocyte signaling with changes in neural circuit function or behavior.

Online content
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Methods
Zebrafish husbandry and maintenance. Wild-type zebrafish (AB strain), Tg(slca1a3:myrGCaMP6-P2A-H2AmCherry) and Tg(slca1a3b:myrGCaMP6s) transgenic lines were used in this study. All zebrafish experiments and procedures were performed in compliance with institutional animal rights for testing and research at Oregon Health & Science University (OHSU). Experiments were approved by the Institutional Animal Care and Use Committee of OHSU. Zebrafish larvae and young fish were nurtured using rotifer suspension and dry food (Gemma 150). Adult fish were maintained and fed with a combination of brine shrimp, rotifer suspension and dry food (Gemma 300).

Plasmids and injections. To generate the slc1a3b:myrGFP-P2A-H2AmCherry reporter construct, the ~9.5-kb DNA sequence upstream of zebrafish slc1a3b translational start site was amplified from genomic DNA and subcloned into Tol2 vector pT2A_min. The sequence encoding myrGFP-P2A-H2AmCherry was PCR-amplified in two separate fragments and subsequently inserted in the 3′ end of the slc1a3b promoter. For the slc1a3b:EmbDmCherry construct, slc1a3b promoter and mCD8mCherry sequences were subcloned into Gateway vectors to generate p5E-slc1a3b and pME-mCD8mCherry, and Gateway LR reactions were performed to produce slc1a3b:EmbDmCherry in the pDestTol2CG2 backbone. The slc1a3b:myrGCaMP6s construct was generated by replacing the myrGFP sequence in the slc1a3b:myrGFP-P2A-H2AmCherry construct with myrGCaMP6s using PCR. Tissue-specific CRISPR-Cas9 backbone vector pDestTol2CG2-U6-RnaG and pME-Cas9 were purchased from Addgene. The sgRNAs targeting individual fgfr genes were designed using the online CRISPR tool CHOPCHOP (9). Validated sgRNAs were then cloned into the pDestTol2CG2-U6-RnaG vector, and Gateway LR reactions were performed to recombine slc1a3b promoter and Cas9 sequences. All constructs generated in this study were confirmed by Sanger sequencing (GENEWIZ) and are available upon request.

For mosaic analyses of individual cells, DNA constructs were injected into one-cell zygotes as follows: 20–30 pg of slc1a3b:myrGFP-P2A-H2AmCherry, 2A-H2AmCherry, or slc1a3b:mCD8mCherry or 10–20 pg of slc1a3b:EmbDmCherry in embryo medium was used. For TTX injection experiments, 6-dpf Tg(slca1a3b:myrGCaMP6s) larval embryos were injected with 1 nl of 0.5 mM TTX into the yolk, and 10 min were allowed to elapse to confirm that the larvae were paralyzed. After 10 min and confirmation of paralysis, DMSO or NE was applied to the embryo medium, and larvae were incubated for 20–30 min before Ca2+ imaging, as described above.

Immunofluorescence staining. Zebrafish embryos and larvae were fixed in 4% PFA/1x PBS at 4 °C overnight and then incubated with 150 mM Tris-HCl, pH 9.0, at 70 °C for 15 min for antigen retrieval (20). Samples were then permeabilized with 100% acetone at −20 °C for 20 min, and antibody staining was performed via standard procedures (21). The following primary antibodies were used: chicken α-GFP (Abcam, 1:1,000), rabbit α-RFP (Rockland, 1:1,000), mouse α-SV2 (DSHB, 1:200) and mouse α-GS (Sigma-Alrich, 1:1 prediluted). The following secondary antibodies were used: Alexa 488 (Jackson ImmunoResearch, 1:250), Rhodamine Red-X α-rabbit (Jackson ImmunoResearch, 1:250) and Dylight 649 α-mouse (Vector Laboratories, 1:250). Stained samples were mounted in Vectashield antifade mounting medium (Vector Laboratories) for confocal analysis.

Quantification and statistical analysis. Astrocyte cell territories or volumes were analyzed using Slidebook 6.0 or Imaris 8 (Bitplane) software. Ca2+ imaging data were analyzed with ImagJ 1.52p and AQUA 1.0 software, as described previously (22). All statistical analyses were performed with GraphPad Prism 8 software. When comparing two groups, unpaired two-tailed Student’s t-tests were used. One-way ANOVA was performed for multiple groups followed by Tukey’s multiple comparisons test. See figure legends and text for specific statistical analyses used. No statistical methods were used to predetermine sample sizes, but our sample sizes are similar to those reported in previous publications (22,23). Data distribution was assumed to be normal, and zebrafish embryos and larvae were randomly allocated to groups. Experiments were not performed blinded to the conditions of the experiments; data analyses were performed blinded to the scorer or did not require manual scoring.

Reporting summary. Further information on research design is available in the Life Sciences Reporting Summary linked to this article.

Data availability
The data that support the findings of this study are available in the manuscript or the Supplementary Information. All reagents and additional data from this study are available upon reasonable request. Source data are provided with this paper.

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Author contributions
J.C., M.R.F. and K.R.M. conceived the project. J.C. carried out experiments and data analyses. K.E.P. provided support for the AQUA software analyses. All authors contributed to the final version of the manuscript.
**Competing interests**
The authors declare no competing interests.

**Additional information**
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Extended Data Fig. 1 | Whole mount in situ hybridization of mammalian astrocyte markers in different stage zebrafish larvae. a, 6 dpf expression patterns of slc1a2a/EAAT2b, slc1a2b/EAAT2a, slc1a3a/Glasta, slc1a3b/Glastb, slc6a11a/GAT-3a, slc6a11b/GAT-3b, aldh1l1, and aqp4. b, Expression patterns of slc1a2b/EAAT2a, slc1a3b/Glastb, and slc6a11b/GAT-3b at 1 dpf, 3 dpf, and 6 dpf in lateral and dorsal view. c, Expression patterns of slc1a2b/EAAT2a, slc1a3b/Glastb, slc6a11b/GAT-3b, aldh1l1, and aqp4 in 14 dpf dissected brains. Scale bar, 200 μm. All images are representative of three or four independent repeats.
Extended Data Fig. 2 | slc1a3b:myrGFP-P2A-H2AmCherry-labeled RGCs and ependymal cells in 6 dpf zebrafish larvae. **a, b,** Representative images show RGCs (**a**) and ependymal cells (**b**) in zebrafish spinal cord labeled by the slc1a3b:myrGFP-P2A-H2AmCherry DNA construct. Scale bar, 20 μm. Representative images from three independent repeats.
Extended Data Fig. 3 | In situ hybridization of slc1a3b, kcnj10a/Kir4.1, and gfap in 3 dpf larvae. **a**, Representative images show the comparison of slc1a3b, kcnj10a/Kir4.1, and gfap in the spinal cord. Dash lines mark the outline of spinal cord. Images are representative of N=3-4 fish larvae. Scale bars represent 500 μm for left panel, 200 μm for middle panel, and 20 μm for right panel, respectively. **b**, RNAScope in situ hybridization of kcnj10a in Tg[slc1a3b:myrGFP-P2A-H2AmCherry] fish spinal cord at 3 dpf. Single z-plane, dorsal view. Scale bar, 20 μm. Representative images from N=3 fish larvae. **c**, Double staining in situ hybridization of slc1a3b (purple) and gfap (red) at 3 dpf. Scale bar, 500 μm. Representative images from N=6 fish larvae.
Extended Data Fig. 4 | NE-induced Microdomain Ca\(^{2+}\) transients in spinal cord astrocytes are not driven by neuronal activity. a, Schematic overview of the TTX injection experiments. b-e, Comparisons of average microdomain Ca\(^{2+}\) events frequencies (b), area sizes (c), amplitudes (d), and durations (e) in DMSO control and NE-treated fish following TTX injections. Error bars represent Mean values +/- SD. **, p<0.01; ****, p<0.0001. p=0.0094 (b), p=1.07x10^{-7} (c), p<1.0x10^{-15} (d and e). Two-tailed unpaired t test. N, number of fish analyzed.
Extended Data Fig. 5 | Microdomain Ca\textsuperscript{2+} transients in the hindbrain radial astrocytes are sensitive to NE treatment. 

**a, b**, AQuA-detected Ca\textsuperscript{2+} events in DMSO control versus NE-treated Tg[slc1a3b:myrGCaMP6s] fish hindbrain radial astrocytes, and corresponding 20 individual ΔF/F traces. Scale bar, 20 μm. Dashed lines mark the regions representing the fine cellular processes of radial astrocytes that were analyzed. See also Supplementary Videos 6 and 7.

**c-f**, Quantifications of average microdomain Ca\textsuperscript{2+} events frequency, area size, amplitude, and duration in DMSO control and NE-treated fish hindbrain regions. Error bars represent mean values ±/− SD. *, p<0.05; **, p<0.01; ****, p<0.0001. p=0.0136 (**), p=4.43x10\textsuperscript{−9} (**), p=6.55x10\textsuperscript{−6} (**), p=0.0058 (**). Two-tailed unpaired t test. N, number of fish analyzed.
Extended Data Fig. 6 | Designed sgRNAs targeting fgfr1-4 are effective in disrupting corresponding genes. a, Genotyping PCR results show that the co-injections of individual sgRNAs together with Cas9 protein led to the disruptions of endogenous restriction enzyme sites in contrast to uninjected controls. Two independent sgRNAs were tested except for fgfr4. NA, not available due to high toxicity.
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| Sample size | No statistical methods were used to predetermine sample sizes, but our sample sizes were similar to those reported in previous studies (Ackerman et al., 2015; Herbert et al., 2017; Baraban et al., 2018). |
| Data exclusions | No data was excluded from the analysis. |
| Replication | All data were confirmed with at least 2 experimental repeats, and similar results were concluded. |
| Randomization | Fish embryos and larvae were randomly allocated into groups for all experiments. |
| Blinding | Experiments were not performed blind to the conditions of the experiments, data analyses were performed blinded to the scorer or did not require manual scoring. |

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**Antibodies**

Antibodies used:

- Chicken anti-GFP (Abcam, ab13970), rabbit anti-RFP (Rockland, 600-401-379), mouse anti-SV2 (DSHB, AB 2315387), mouse anti-GS (Sigma, 389M), Alexa 488 anti-chicken(Jackson ImmunoResearch, 703-545-155), Rhodamine Red-X -rabbit (Jackson ImmunoResearch, 111-295-003), and Dylight 649 anti-mouse (Vector Laboratories, DI-2649), anti-Digoxigenin-AP (Sigma, 11093274910), anti-Fluorescein-AP (Sigma, 11426338910)

Validation:

- Chicken anti-GFP and rabbit anti-RFP antibodies has been used in zebrafish for immunostaining (Lessen et al., 2017); mouse anti-SV2 antibody has been used in zebrafish for immunostaining (Turner et al., 2014); For mouse anti-GS antibody, same clone antibody (GS-6)from Chemicon (MAB302) has been used in zebrafish for immunostaining (Grupp et al., 2010).

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**Laboratory animals**

- Zebrafish at 1-14 days post-fertilization were used for all experiments in this study. Sex could not be determined at this stage of zebrafish.

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- No wild animals were used in this study.

**Field-collected samples**

- No field-collected samples were used in this study.

**Ethics oversight**

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