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IL-10 from CD4$^+$CD25$^-$Foxp3$^-$CD127$^-$ Adaptive Regulatory T Cells Modulates Parasite Clearance and Pathology during Malaria Infection

Kevin N. Couper¹, Daniel G. Blount¹, Mark S. Wilson², Julius C. Hafalla¹, Yasmine Belkaid², Masahito Kamanaka³, Richard A. Flavell³, J. Brian de Souza¹,⁴, Eleanor M. Riley¹*  

¹Immunology Unit, Department of Infectious and Tropical Diseases, London School of Hygiene and Tropical Medicine, London, United Kingdom, ²Laboratory of Parasitic Diseases, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda, Maryland, United States of America, ³Section of Immunobiology, Yale University School of Medicine, New Haven, Connecticut, United States of America, ⁴Department of Immunology and Molecular Pathology, University College London Medical School, London, United Kingdom

Abstract

The outcome of malaria infection is determined, in part, by the balance of pro-inflammatory and regulatory immune responses. Failure to develop an effective pro-inflammatory response can lead to unrestricted parasite replication, whereas failure to regulate this response leads to the development of severe immunopathology. IL-10 and TGF-β are known to be important components of the regulatory response, but the cellular source of these cytokines is still unknown. Here we have examined the role of natural and adaptive regulatory T cells in the control of malaria infection and find that classical CD4$^+$CD25$^+$ (and Foxp3$^+$) regulatory T cells do not significantly influence the outcome of infections with the lethal (17XL) strain of Plasmodium yoelii (PyL). In contrast, we find that adaptive IL-10-producing, CD4$^+$ T cells (which are CD25$^-$, Foxp3$^-$, and CD127$^-$ and do not produce Th1, Th2, or Th17 associated cytokines) that are generated during both PyL and non-lethal P. yoelii 17X (PyNL) infections are able to down-regulate pro-inflammatory responses and impede parasite clearance. In summary, we have identified a population of induced Foxp3$^-$ regulatory (Tr1) T cells, characterised by production of IL-10 and down regulation of IL-7R, that modulates the inflammatory response to malaria.

Introduction

The erythrocytic stage of malaria infection is characterised by the development of strong pro-inflammatory immune responses which, although required to control parasite replication and promote clearance of infected erythrocytes, must be tightly regulated to prevent the immune-mediated pathology which is integral to the development of the severe complications of infection in humans and in a number of well-characterised animal models [1–3]. Previous studies have highlighted important components of the regulatory response, but the cellular source of these cytokines is still unknown. Here we have examined the role of natural and adaptive regulatory T cells in the control of malaria infection and find that classical CD4$^+$CD25$^+$ (and Foxp3$^+$) regulatory T cells do not significantly influence the outcome of infections with the lethal (17XL) strain of Plasmodium yoelii (PyL). In contrast, we find that adaptive IL-10-producing, CD4$^+$ T cells (which are CD25$^-$, Foxp3$^-$, and CD127$^-$ and do not produce Th1, Th2, or Th17 associated cytokines) that are generated during both PyL and non-lethal P. yoelii 17X (PyNL) infections are able to down-regulate pro-inflammatory responses and impede parasite clearance. In summary, we have identified a population of induced Foxp3$^-$ regulatory (Tr1) T cells, characterised by production of IL-10 and down regulation of IL-7R, that modulates the inflammatory response to malaria.

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* E-mail: eleanor.riley@lshtm.ac.uk

Current address: School of Pharmacy, University of Nottingham, University Park, Nottingham, United Kingdom
and high levels of CD25) have since been shown to regulate the immune response in a number of infections including *Leishmania spp* infections, *Mycobacterium tuberculosis* and helminth infections [15–18], mediating their effects either via direct cell contact or by release of cytokines. However, it is now becoming apparent that both adaptive (Foxp3⁺) regulatory T cell populations and classical T-bet expressing Th1 cells also play crucial immunoregulatory roles during infection and mediate their effects through secretion of IL-10 [19–21].

In this study we have examined the generation and function of both nTreg and adaptive IL-10-secreting T cells during malaria infection. We observe equivalent expansion of natural Foxp3⁺ regulatory T cells during both lethal and non-lethal *P. yoelii* infections but, using either anti-CD25 treatment or adoptive transfer of purified CD25⁺/Foxp3⁺ nTreg or CD25⁻/Foxp3⁻ non-Treg T cell populations, we find no role for nTreg during PyL infection. Conversely, we demonstrate that populations of adaptive regulatory CD4⁺ T cells, that are CD25⁺, Foxp3⁻ and CD127⁻, which do not make IFN-γ, IL-4 or IL-17, develop during both PyL and PyNL infections. These cells inhibit parasite clearance but, importantly, also prevent the development of pathology via production of IL-10. These data are consistent with the notion that whilst endogenous populations of nTreg may be sufficient to prevent immune-mediated pathology during chronic infections which induce rather modest inflammatory responses, such as avirulent leishmanias, tuberculosis or helminth infections, rapid induction of distinct populations of adaptive/Th1 CD4⁺ T cells producing IL-10 may be required to counter the powerful inflammatory signals provided by virulent, rapidly replicating pathogens.

**Results**

**Course of infection with PyL and PyNL in C57BL/6 mice**

In accordance with previous observations [5,22], infection of C57BL/6 mice with 10⁴ *P. yoelii* 17XL (PyL) parasites was associated with a rapid onset of fulminant parasitaemia (approaching 100% by day 7 pi) that was universally fatal (Figure 1A, B). In contrast, infection with 10⁴ *P. yoelii* 17X (NL) (PyNL) parasites led to a more gradual increase in parasitaemia with peak parasitaemia of approx. 30% on day 14 pi, before the infection eventually resolved. Significant differences in malaria-induced anaemia were also evident between lethal and non-lethal infections, with more rapid onset and increased severity of anaemia occurring in PyL-infected mice compared with PyNL-infected mice (Figure 1C).
Similar expansion and activation of CD4+Foxp3+ regulatory T cells during PyL and PyNL infection

We have previously reported that simultaneous neutralisation of TGF-β and blockade of IL-10 signalling allows a proportion of PyL-infected mice to resolve their infections and survive [5], suggesting that active immune regulation/immune suppression occurs during PyL infection that inhibits optimal parasite control. In agreement with these observations, Kobayashi et al [7] have reported that IL-10 is produced very early during PyL (but not during PyNL) infection and Perry et al [23] have reported a switch from IL-12 (at day 3 pi) to IL-10 (at day 17 pi) production by splenic dendritic cells during the course of a non-lethal Py infection. These data are consistent with the hypothesis that protective pro-inflammatory responses develop during the acute phase of PyNL infection that limit parasite numbers, whereas an early anti-inflammatory cytokine response during the acute phase of PyL infection inhibits the development of protective immune responses. As CD4+CD25+ regulatory T cells (nTreg) have been reported to regulate immunity in a number of auto-immune and infectious diseases [14–18] and can exert their regulatory role through secretion of IL-10 and/or TGF-β we investigated, using intracellular staining for Foxp3 as well as transgenic Foxp3-GFP reporter mice [24], whether nTreg activation is correlated with the virulence of PyL infection.

CD4+Foxp3+ cells, the expression levels of Foxp3 and the ratios of CD4+Foxp3+ (nTreg) to CD4+Foxp3− (non-regulatory T cells) were assessed over the first 7 days pi (Figure 2B-D). In accordance with previous observations [23] a significant increase in the numbers of splenic CD4+Foxp3+ nTreg was observed during the first 5 days of PyL infection (Figure 2B) and this was accompanied by increased levels (MFI) of Foxp3 expression (Figure 2C) and a transient increase (on day 3pi) in the nTreg/non-Treg ratio (Figure 2D). However, almost identical changes in nTreg numbers and Foxp3 expression levels were observed in mice infected with PyNL, and there were no significant differences in any nTreg parameter between PyL-infected and PyNL-infected mice at any time up to 7 days pi, after which the PyL-infected mice succumbed to their infections.

Similar results were obtained with Foxp3-GFP reporter mice [24]. Importantly, the course of PyL and PyNL infections were equivalent in Foxp3-GFP mice and C57BL/6 mice (data not shown). A representative plot showing Foxp3-GFP expression in infected and uninfected mice is shown in Figure 2E. Numbers of splenic GFP+ (Foxp3+) CD4+ regulatory T cells on day 5 post infection. 3–5 mice per group. Results are representative of 3 independent experiments. Symbols represent significant differences (p<0.05) between groups: ≠ PyL vs PyNL; * PyL vs uninfected; ~ PyNL vs uninfected. doi:10.1371/journal.ppat.1000004.g002

Figure 2. Expansion and activation of natural Foxp3+ regulatory T cell populations during PyL and PyNL infections. Numbers and activation status of Foxp3+ regulatory T cells were determined on various days post-infection with PyL or PyNL in (A–D) WT mice or (E–F) Foxp3-GFP transgenic mice. (A) Representative dot plot of intracellular staining in CD4+ T cells. On selected days post-infection with PyL or PyNL, splenic CD4+ lymphocytes were analysed for (B) numbers of Foxp3+ cells,(C) intensity (MFI) of Foxp3 staining and (D) ratio of Foxp3+ to Foxp3− cells. (E) Representative dot plots showing GFP (Foxp3) expression in CD4+ splenocytes from 7 day PyL and PyNL-infected animals and uninfected controls. (F) Numbers of splenic GFP+ (Foxp3+) CD4+ regulatory T cells on day 5 post infection. 3–5 mice per group. Results are representative of 3 independent experiments. Symbols represent significant differences (p<0.05) between groups: ≠ PyL vs PyNL; * PyL vs uninfected; ~ PyNL vs uninfected.
Neither CD25<sup>hi</sup> nor Foxp3<sup>+</sup> endogenous regulatory CD4<sup>+</sup> T cells contribute to the virulence of lethal P. yoelii infection

The similarity of the nTreg response during PyL and PyNL infections suggested that, in our hands, suppression of effector cell responses by nTreg was unlikely to explain the highly virulent nature of PyL infections. However, to formally test the role of nTreg, mice were treated with a cocktail of anti-CD25 antibodies (previously shown to give optimal depletion of CD4<sup>+</sup>CD25<sup>hi</sup>-Foxp3<sup>+</sup> cells; 25) 3 days prior to infection with PyL (Figure 3). As previously reported [25], the 7D4 (IgM, anti-CD25) antibody substantially reduced the proportion of splenic CD25<sup>+</sup> CD4 cells within 3 days (i.e. day of infection) but CD25<sup>+</sup> cells recovered to normal levels by day 4 pi (results not shown) and 7D4 treatment had no significant effect on the frequency of CD4<sup>+</sup>Foxp3<sup>+</sup>ve cells (results not shown). In contrast, PC61 (IgG anti-CD25) given in combination with 7D4 induced an approximately 50% reduction in the frequency of both CD25<sup>hi</sup> and Foxp3<sup>+</sup> cells that was sustained throughout the 7 day infection period [25]. Nonetheless, neither 7D4 treatment nor combined 7D4 + PC61 treatment significantly altered the course of parasitaemia, anaemia or survival of PyL infection in C57BL/6 mice (Figure 3A–C). As these observations contradict those of a similar published study [13] we considered whether some effect of natural T reg might be being masked by the rapidly ascending parasitaemia and early mortality associated with infection with 10<sup>4</sup> PyL parasites. We therefore repeated the anti-CD25 antibody treatment in C57BL/6 mice infected with either a 10 fold lower dose of PyL parasites (10<sup>3</sup> PyL) or with 10<sup>4</sup> PyNL parasites. However, anti-CD25 antibody treatment did not alter the outcome of either of these infections.
in the CD25

CD4

actually indistinguishable in RAG

2

infection with the same kinetics as WT mice (compare Figure 3E, 

CD4

2

control (unreconstituted) RAG

2

or BALB/c mice. 

anti-parasitic immunity during PyL infection in either C57BL/6 

infections, since IL-10 mRNA is significantly upregulated in 

spleen CD4+ cells on days 5 and 7 post-infection compared with 

cells from uninfected mice (Figure 4A). Furthermore, CD4+ 

cells (and potentially B cells) may be the major source of IL-10 during 

infection since plasma IL-10 does not increase above baseline 

levels in RAG−/− mice (Figure 4B, C) except on day 3 pi of PyL 

infection.

It has been reported that regulatory T cell responses are more 
effective at limiting pro-inflammatory responses in BALB/c mice 
than in C57BL/6 mice [26]. Therefore, to determine whether 
mouse strain influences the outcome of anti-CD25 treatment 
during PyL infection, we repeated the CD25-depletion 

settings in BALB/c mice and compared our depletion strategies 

single dose of 7D4 or 7D4+PC61 given 3 days prior to infection) 

with a strategy previously shown to affect PyL infection [13], 
namely repeated injections of 7D4 antibody on days −3, −1 and 5 

relative to PyL infection. Repeated administration of 7D4 did not 

increase either the duration of CD25+ T cell depletion or the 

extent of depletion of CD4+Foxp3+ve cells compared to the other 
treatment regimes (Figure S3). Consistent with this, repeated 
administration of 7D4 did not alter the course of PyL infection 

compared with single 7D4 administration or combined 7D4 and 

PC61 administration (Figure S3), and none of our CD25-depletion 

regimes had any effect on PyL infection in BALB/c mice (Figure 

S3). 

It is becoming increasingly evident that anti-CD25 antibody 
treatment is not a specific or robust strategy to examine the 
importance of natural regulatory T cells during inflammatory 
episodes [25,27–29]. CD25 expression is not limited to nTreg 
[24]. Moreover, depending on the precise protocol used, a variable 
but significant proportion of Foxp3+ cells escape depletion by anti-
CD25 antibody. We therefore compared the outcome of PyL 
infection in RAG−/− mice reconstituted or not with purified 
naive CD4+CD25− (putative effector) T cells or a 10:1 ratio of 
effector (CD4+CD25+) to nTreg (CD4+CD25−) cells. Furthermore, 
as nTreg can down-regulate NK cell responses [30], and as 
NK cells have previously been reported to play a protective role 
during malaria infection [31–33], we adoptively transferred 
CD4+CD25− (nTreg) cells in the absence of CD4+CD25− (effector) cells, to determine whether nTreg modulate innate 
immune responses during malaria infection. The proportion of 
Foxp3+ cells fell from 10–15% in unsorted CD4+ T cells to 1–2% 
in the CD25−CD4+ population, whereas CD25+ cells were highly 
enriched for Foxp3+ cells (70–80%; Figure 3D). 

In accordance with our previous studies [22], we found that 
control (unreconstituted) RAG−/− mice succumbed to PyL 
infection with the same kinetics as WT mice (compare Figure 3E, 
F with Figure 1). Furthermore, the course of infection was 
virtually indistinguishable in RAG−/− mice reconstituted with 
CD4+CD25−, CD4+CD25+ or a 10:1 ratio of CD4+CD25− / 
CD4+CD25+ T cells (Figure 3E, F). Thus, using two independent 
models of nTreg depletion, we have found no significant role for 
natural CD4+CD25−Foxp3+ regulatory T cells in suppression of 
anti-parasitic immunity during PyL infection in either C57BL/6 
or BALB/c mice.

PyL- and PyNL-induce IL-10 production from CD4+, 

CD25−, Foxp3+ T cells that express low levels of IL-7Rα 

Having found no evidence that nTreg influence the outcome of 
PyL infection we next investigated the possibility that IL-10 
producing CD4+ T cells (“adaptive” Treg or Tr1 cells) might be 
duced during PyL and/or PyNL infection that regulate parasite 
killing and/or pathology. 

Expression of IL-10 mRNA was determined by real time PCR 
in purified splenic CD4+ T cells obtained on days 1, 3, 5 and 7 
post-infection from wild type (WT) C57/BL6 mice and plasma 
levels of IL-10 were determined by ELISA on days 1, 3, 5 and 7 pi 
from WT and RAG−/− mice. We find that CD4+ T cells are a 
significant source of IL-10 by day 5 of both PyL and PyNL 

infections, since IL-10 mRNA is significantly upregulated in 
splenic CD4+ cells on days 5 and 7 post-infection compared with 
cells from uninfected mice (Figure 4A). Furthermore, CD4+ T cells 
(and potentially B cells) may be the major source of IL-10 during 
infection since plasma IL-10 does not increase above baseline 
levels in RAG−/− mice (Figure 4B, C) except on day 3 pi of PyL 

infection.

Figure 4. CD4+ T cells are a major source of IL-10 during both 

PyL and PyNL infection. (A) On selected days post-infection, CD4+ T 
cells were purified from PyL and PyNL infected mice by MACS sorting 
and levels of IL-10 mRNA were determined by real time PCR (Taqman) 
relative to the house keeping gene GAPDH. The results are shown as 
the fold change in expression relative to uninfected naïve CD4+ T cells. 
In separate experiments the levels of IL-10 in the plasma of WT and 
RAG−/− mice were determined by ELISA on selected days of (B) PyL 
and (C) PyNL infection. Groups consisted of 3–5 mice and the results are 
representative of 2 separate experiments. For Taqman analysis, purified 
cells from several mice in each group were pooled; no significant 
differences between groups were identified. * indicates significant 
differences (p<0.05) between WT and RAG−/− infected mice. 
doi:10.1371/journal.ppat.1000004.g004

(Figures S1 and S2) suggesting that natural T reg cells do not 
markedly influence P. yoelii infections in C57/BL6 mice.
To more accurately determine the cellular source of IL-10 during *P. yoelii* infection, splenocytes from IL-10-GFP reporter mice [21] were examined for expression of GFP and various cell surface markers on selected days after PyL or PyNL infection (Figure 5A–C). In both infections, from day 5 onwards, the vast majority of the IL-10+ cells were CD4+ lymphocytes. At no point during either PyL or PyNL infection did we observe significant IL-10 production by myeloid (CD11b+), lymphoid dendritic cells (CD11c+) or macrophages (F4-80+) (results not shown). IL-10 production by CD19+ B cells was observed, on day 7 post-infection, only during PyL but not PyNL infection (results not shown). Moreover, IL-10 producing non-CD4+ T cells produced only low quantities of IL-10, whereas CD4+ T cells were heterogeneous in their ability to produce IL-10 (Figure 5A).

Since it is not possible to stain for intranuclear Foxp3 without quenching the fluorescence of GFP, IL-10/GFP+ CD4+ T cells were analysed for expression of CD25 and Foxp3, or CD69, CD11c, CD62L or CD127. (D) The frequency and number of GFP+ (IL-10)+ CD4+ T cells was calculated 7 days post-infection in infected or uninfected mice. Groups consisted of 3–5 mice and the results are representative of 2 independent experiments.

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**Figure 5.** CD127low Foxp32 CD4+ T cells that do not constitutively express CD25 are the major source of IL-10 during *P. yoelii* infection. Transgenic, IL-10-GFP knockin tiger mice were infected with PyL or PyNL. (A) Splenic lymphocytes from infected or uninfected control mice were analysed for expression of CD4+ and GFP. (B,C) 7 days post infection, splenic CD4+ T cells were analysed for (B) expression of CD25 and Foxp3, or (C) GFP (IL-10) and CD25, CD69, CD11c, CD62L or CD127. (D) The frequency and number of GFP+ (IL-10)+ CD4+ T cells was calculated 7 days post infection in infected or uninfected mice. Groups consisted of 3–5 mice and the results are representative of 2 independent experiments.
CD4\textsuperscript{+} T cells that produce IL-10 during malaria infection are not classical nTreg, Th1, Th2, or Th17 cells

We have shown that CD4\textsuperscript{+} T cells are the primary source of IL-10 during malaria infection, and that these cells do not express CD25, suggesting that they may not be conventional nTreg cells. Since IL-10 can be produced by various effector CD4\textsuperscript{+} T cell subsets (including Th1, Th2 and Th17 cells), as well as specialised regulatory populations such as Tr1 [19, 20 34–36], we examined the expression of Th1, Th2 and Th17 lineage-associated cytokines in IL-10-producing (GFP\textsuperscript{+}) and IL-10-GFP\textsuperscript{−} CD4\textsuperscript{+} T cells purified from IL-10-GFP reporter mice on day 7 of infection. As seen previously (Figure 5), GFP expression was similar in CD4\textsuperscript{+} T cells isolated from PyL and PyNL infected mice (Figure 6A). As expected, IL-10 mRNA was expressed at much higher levels in GFP\textsuperscript{+} than in GFP\textsuperscript{−} cells but cells isolated from PyL and PyNL infected animals expressed similar levels of IL-10 mRNA (Figure 6B). Importantly, Foxp3 mRNA was not upregulated in IL-10-GFP\textsuperscript{+} cells isolated during either PyL or PyNL infection, confirming that the IL-10-producing CD4\textsuperscript{+} T cells that develop during \textit{P. yoelii} infection are neither natural nor induced Foxp3\textsuperscript{+} regulatory T cells. Moreover, GFP\textsuperscript{−} cells did not express significant amounts of mRNA for IFN-\textgreek{g}, IL-4 or IL-17, thus distinguishing them from classical Th1, Th2 and Th17 cells. Although IL-10-GFP\textsuperscript{−} cells expressed IL-13 mRNA, levels were comparable to those seen in GFP\textsuperscript{−} cells indicating that IL-10 producing cells did not preferentially co-produce IL-13. Thus, the IL-10-producing CD4\textsuperscript{+} T cells induced during \textit{P. yoelii} infection fit the definition [35] of adaptive, Tr1, regulatory T cells.

IL-10 from CD4\textsuperscript{+} T cells inhibits parasite killing during infection

To determine whether IL-10 production from T cells is functionally important during Py infection, we first compared the course of PyL and PyNL infection in IL-10\textsuperscript{−/−} and WT mice (Figure 7). PyNL infection was significantly attenuated - with significant reductions in parasitaemia and anaemia in IL-10\textsuperscript{−/−} mice compared with WT mice (Figure 7A–D), although the IL-10\textsuperscript{−/−} mice did lose significantly more weight than age-matched WT mice (Figure 7C). Furthermore, approx 30\% (6/21 mice) of IL-10\textsuperscript{−/−} (but not WT) mice infected with 10\textsuperscript{4} PyL pRBC were able to control their infections and survived (Figure 7E–H), with parasitaemia declining from a peak of approx 45\% on day 6pi. Moreover, IL-10\textsuperscript{−/−} (but, again, not WT) mice given a low dose PyL infection (10\textsuperscript{3} pRBC) were fully able to control parasitaemia and 100\% of the mice survived (Figure 7I–L).

Taken together, these data indicate that IL-10 suppresses immune effector mechanisms which would otherwise be able to control low dose PyL infections. Since this IL-10 emanates...
principally from CD4+ T cells (Figure 5) we hypothesised that IL-10-deficient CD4+ T cells may promote more effective parasitemia control than WT CD4+ T cells. To test this, purified naive WT or IL-10−/− CD4+ T cells were adoptively transferred into RAG−/− mice which were then infected with PyNL or PyL parasites.

PyNL-infected RAG−/− mice that had received IL-10−/− CD4+ T cells developed significantly lower parasitemia than those which had received WT CD4+ T cells (Figure 8A). Although both groups developed similar levels of anaemia, mice that received IL-10−/− T cells lost significantly more weight and succumbed to infection more rapidly than mice that received WT CD4+ T cells (Figure 8B-D). Exacerbation of disease despite improved parasitemia control in mice receiving IL-10−/− CD4+ T cells was associated with more extensive proliferation of the adoptively transferred T cells (IL-10−/− T cells comprised >30% of total splenic leucocytes compared with <10% for transferred WT cells), higher concentrations of circulating IFN-γ and lower plasma concentrations of IL-10 (data not shown). These data are consistent with the conclusion that recipients of IL-10−/− CD4+ T cells died of immunopathology whilst recipients of WT CD4+ T cells eventually died because they were unable to fully resolve their infections.

By contrast, RAG−/− mice that had received IL-10−/− CD4+ T cells were somewhat better able to control infections with 103 (B-E-H) or 104 (8I–L) PyL infections than were mice receiving WT CD4+ T cells; a proportion of mice receiving IL-10−/− T cells were able to control their infections, although failure to fully eliminate parasites eventually led to death from anaemia. Thus, IL-10 derived from CD4+ T cells significantly modulates the outcome of both PyL and PyNL infection.

IL-10 protects against immune pathology during P. yoelii infection

It has previously been shown that IL-10−/− mice succumb to normally avirulent P. chabaudi chabaudi infections despite comparable or more effective control of malaria parasitemia compared to WT mice [9]. The increased susceptibility of IL-10−/− mouse is due to elevated plasma concentrations of IFN-γ and TNF-α [10] and survival of IL-10−/− mice following malaria infection can be enhanced by treatment with anti-TNF-α [10]. Whilst there was no marked difference in mortality between P. yoelii-infected IL-10−/− and WT mice, IL-10−/− mice (and RAG−/− mice reconstituted with IL-10−/− T cells) lost significantly more weight than mice reconstituted with WT T cells during
PyNL infection, indicative of more severe morbidity (Figure 7C, 8C).

Histopathological examination of infected animals did not reveal any liver or lung damage 3 days post-infection (data not shown) but revealed significantly more hepatic cellular changes including periportal inflammation, necrosis and bridging necrosis in IL-10<sup>2/2</sup> mice than in WT mice on days 7 and 14 post-infection (Figure 9A) and this was significantly more severe in PyL-infected than PyNL-infected animals on day 7 post-infection. We also found that by day 25 of PyNL infection, RAG<sup>2/2</sup> recipients of IL-10<sup>2/2</sup> CD4<sup>+</sup>T cells had developed significantly more severe hepatic periportal inflammation and necrosis (including bridging necrosis) than RAG<sup>2/2</sup> recipients of WT CD4<sup>+</sup>T cells (Figure 9B). Thus, T cell derived IL-10, although negatively regulating parasite killing, is protective during malaria infection by preventing the onset of immunopathology.

**Discussion**

It is well established, in a variety of infections, that regulatory cytokines both ameliorate immunopathology and delay pathogen clearance [5,8,9-11,37-42]. Manipulation of these cytokines by vaccination or immunotherapy, to simultaneously enhance pathogen clearance and limit the associated pathology, requires a better understanding of their cellular sources and mechanisms of induction. Important roles have been demonstrated for both IL-10 [6-11] and TGF-β [4,5] in modulating the outcome of murine malaria infections, and observational data strongly suggests that they play a similar role in human infections [43-45]. Recently, endogenous or natural, CD25<sup>hi</sup>, Foxp3<sup>+</sup>CD4<sup>+</sup>T cells (nTreg) have been implicated as major regulators of malarial pathology [13,46] but their mechanisms of action remain undefined. Attempting to elucidate the role of nTreg in murine Plasmodium yoelii infections, we were surprised to find no role for these cells in regulating the outcome of either high dose (10<sup>8</sup>) or lower dose (10<sup>5</sup>) lethal (Py17XL; PyL) or non-lethal (Py17X; PyNL) infection in either C57BL/6 or BALB/c mice. In contrast, we find that adaptive, IL-10-producing CD4<sup>+</sup>Tr1 cells (CD25<sup>+</sup>, Foxp3<sup>+</sup>, CD127<sup>+</sup>, IFN-γ<sup>-</sup>, IL-4<sup>-</sup> and IL-17<sup>-</sup>), are generated during both PyL and PyNL infections and are associated with down-regulation of pro-inflammatory responses, moderation of both morbidity and mortality and failure to clear parasites. Crucially, we were able to demonstrate a causal relationship between these various observations by showing that IL-10<sup>2/2</sup> CD4<sup>+</sup>T cells adoptively transferred into RAG<sup>2/2</sup> mice provided more effective parasite control than did WT CD4<sup>+</sup>T cells, but at the cost of more severe pathology.
IL-10 components of the normal intestinal flora of conventionally commensal organisms between mice in different laboratories. Results may be differences in prior exposure to pathogens or on cerebral pathology [51]. One explanation for these inconsistent pathology in normally resistant BALB/c mice [50] or has no effect effector T cell responses and increases the severity of brain cerebral pathology in C57BL/6 mice [49], or enhances infection in C57BL/6 mice we were unable to reproduce the previously reported to ameliorate PyL infection [13], in both C57BL/6 and BALB/c mice we were unable to reproduce the published observations. Anti-CD25 treatment failed to ameliorate PyL infection initiated by a 10 fold lower dose of parasites, discounting the possibility that the virulence of high dose PyL infection masks regulatory activity of nTreg that would otherwise be evident during a less virulent infection. Furthermore, adoptive transfer of CD25+ and/or CD25− CD4+ T cells into RAG−/− mice also failed to reveal any role for CD25+ Foxp3+ cells in this infection. The discrepancy between our findings and those of other labs is reminiscent of the disparate results obtained for P. berghei ANKA infection which report that depletion of CD25+ regulatory T cells either facilitates parasite control and prevents the onset of the cerebral pathology infection in C57BL/6 mice [49], or enhances effector T cell responses and increases the severity of brain pathology in normally resistant BALB/c mice [50] or has no effect on cerebral pathology [51]. One explanation for these inconsistent results may be differences in prior exposure to pathogens or commensal organisms between mice in different laboratories. Components of the normal intestinal flora of conventionally housed animals are essential for development of intestinal nTreg [52] and nTreg development is facilitated by the presence of covert infections such as Helicobacter hepaticus [33]. Depletion of nTreg by anti-CD25 treatment in such mice may lead to more profound alteration in the effector / regulatory cell balance than in mice (such as those used in our studies) raised in low-infection environments.

Although, we could show no role for nTreg in acute PyL and PyNL infection, we have shown that the adaptive IL-10-producing regulatory T cells that develop during P. yoelii infection hinder parasite control but simultaneously limit disease severity. In contrast to recent studies describing a role for IL-10 producing Th1 cells in Toxoplasma gondii [20] and Leishmania spp [19,54,55] infections, the adaptive IL-10 producing Tregs we describe do not co-express IFN-γ or other effector cytokines and better fit the definition of Tr1 cells. Nevertheless in several virulent protozoal infections (PyL, Toxoplasma gondii, Leishmania major SD and L. donovani), adaptive IL-10-producing CD4+ T cells are required to regulate the fulminant Th1-effector responses that are induced whereas classical (Foxp3+) Treg appear to be sufficient to regulate the effector response to a less virulent (healing) strain of L. major [15]. Given that naive CD4+ T cells can develop into adaptive Treg after interaction with IL-10-producing dendritic cells expressing low levels of co-stimulatory molecules [56–59], it is possible that the induction of Treg during P. yoelii infection is linked to the modulation of macrophage and dendritic cell function that occurs in response to prolonged toll-like receptor signalling [60]. Alternatively, parasite-induced TGF-β [40,60], IL-6 [44] and/or IL-27 may synergise to promote production of IL-10 by Th1, Th2, Th17 and Tr1 cells [34,36,61,62].

We have not yet definitively identified the mechanism by which IL-10 suppresses parasite clearance but given our recent findings that control of the acute phase of P. yoelii parasitaemia is critically dependent on macrophages [22], it is likely that T cell-derived IL-10 acts directly on macrophages to inhibit their anti-parasitic mechanisms. It is also possible that, as in mycobacterial infections, adaptive Treg induce an autocrine signalling loop in which macrophages both secrete and respond to IL-10 with consequent down regulation of effector function and pathology via a STAT-3–dependent pathway [65–66].

In summary, we have demonstrated that adaptive, but not natural, regulatory T cells control parasite numbers during PyL and PyNL infections whilst also limiting the onset of immunopathology. These cells are characterised by lack of expression of CD25 and Foxp3, down-regulation of CD127 and production of IL-10 but not IFN-γ, IL-4 or IL-17. Taken together with our data highlighting the importance of macrophages in the control of malaria infection [22], these findings identify an important pathway of adaptive, T cell-mediated control of innate immune responses. Further studies are required to identify the pathways leading to induction of this important regulatory cell population.

**Materials and Methods**

**Mice and parasites**

C57BL/6, Foxp3-GFP (F2: 129/C57BL/6; from A. Rudensky, University of Washington, 24), C57BL/6 RAG-1−/−, C57BL/6 IL-10−/− and BALB/c mice were bred in-house or purchased from Harlan and maintained under barrier conditions at LSHTM. IL-10-GFP reporter mice [21] were maintained under barrier conditions at the National Institutes of Health. Cryopreserved Plasmodium yoelii 17X (non lethal; PyNL) and P. yoelii 17XL (lethal; PyL) parasites were passaged once through mice before being used in experimental animals.
Unless stated otherwise, male or female mice, 7–9 weeks of age, were infected intravenously with $1 \times 10^5$ or $1 \times 10^4$ parasitised red blood cells (pRBC). Parasitaemia was determined daily by examination of Giemsa-stained thin smears of tail blood for the first seven days of infection and every second day thereafter. On every second day, mice were weighed and RBCs were counted using an automated haemoanalyser (Beckman Coulter). Plasma was stored (at −20°C) for cytokine quantification. On selected days post-infection, mice were sacrificed and spleens were removed. Single spleen cell suspensions were prepared by homogenisation through a 70 µm cell strainer (BD Biosciences) and live cells enumerated by trypan blue exclusion.

Cell sorting

CD4+ T cells were positively selected using anti-CD4- conjugated midMACS beads (Miltenyi Biotec) according to the manufacturer’s instructions and the purity of eluted cells was checked by flow cytometry. In some experiments, the CD4+ cells were labelled with anti-mouse CD4 (GK1.5; Rat IgG2b; E-bioscience) and anti-mouse CD25 (PC61; Rat IgG1; E-bioscience) fluorochrome-labelled antibodies and sorted, using a BD FACSVantage SE, into CD4+CD25+ and CD4+CD25− populations. In separate experiments CD4+ T cells, isolated from IL-10-GFP reporter mice [21] on day 7 of infection, were labelled with anti-mouse CD4 (GK1.5; Rat IgG2b; E-bioscience) and IL-10 producing (GFP+) and non-IL-10 producing (GFP−) CD4+ T cells were purified by flow cytometric cell sorting.

Real time PCR

IL-10, Foxp3, IFN-γ, IL-4, IL-13 and IL-17A mRNA were quantified by Taqman (ABI, Warrington, UK). RNA was extracted (RNAasy) and DNase1 treated prior to cDNA synthesis. cDNA expression for each sample was standardised using the housekeeping gene GAPDH. Cycling conditions were: initialisation 2 min at 50°C and 10 min at 95°C followed by 40 cycles of 15 sec at 95°C and 1 min at 60°C.

Primer sequences:

IL-10 Forward AGA GCC AGA TTA TCT CTT TCT ACC TCA G
Reverse CCT TTT TCG CCT TGC TGT TG
IFN-γ Forward AGA GCC AAG AGA AAG GGA
Reverse CGA AGT GGG AGT TAC ACA
IL-4 Forward ACG AGG TCA CAG GAG AGG GGA
Reverse AGC CCT ACA GAG GAC CTC ACT C
IL-13 Forward CCT CTG ACC CTA GAG GAG CTG CCT ATAT
Reverse CG TGT GCA CAG GGG GAG TCT
IL-17A Forward TGG TGAAG TCA ACC CTC AA AGTC
Reverse AGG GATAC TCT ATAC CAG GTCT TCT ATT

Cytokine ELISA

Rat anti-mouse IL-10 (JES5-2A5; Rat IgG1; Mabtech, Sweden) or rat anti-mouse interferon (IFN)-γ (AN-18; Rat IgG1; eBioscience) antibodies were used as capture antibodies, diluted in 0.5 M Tris-HCl, pH 8.9 buffer. Biotinylated rat anti-mouse IL-10 MAb (JES5-16E3; pH 8.9 buffer, Biotinylated rat anti-mouse IL-10 MAb (JES5-16E3; pH 8.9 buffer) or rat anti-mouse IFN-γ MAb (R4-6A2; Rat IgG1; Mabtech) were used as detecting antibodies and were visualised using streptavidin-alkaline phosphatase (eBioscience) and p-nitrophenyl phosphate (Sigma Aldrich, UK). Absorbance was read at 405 nm using a MRX TG II microplate reader (Dynex Technologies Ltd., UK).

Flow cytometry

For flow cytometric analysis, cells were surface stained with anti-mouse CD4 (RM4-5; Rat IgG2a; BD Biosciences), anti-mouse CD25 (PC61; Rat IgG1; eBioscience), anti-mouse CD69 (H1.2F3; Armenian Hamster IgG; eBioscience), anti-mouse CD62L (MEL-14; Rat IgG2a; eBioscience) or anti-mouse CD127 (AV34; Rat IgG2a; eBioscience). Intracellular Foxp3 staining using anti-mouse Foxp3 (FJK-16s; Rat IgG2a; eBioscience) was performed by permeabilising cells with 0.1% Saponin/PBS. Cells were concurrently incubated with anti-mouse CD16/32 (Fc block) when staining with all conjugated antibodies. Flow cytometric acquisition was performed using a FACS Calibur (BD Immunocytometry Systems, USA) and all analysis was performed using Flowjo software (Treestar Inc., OR, USA).

Histopathology

Liver and lung tissues were fixed in 10% Formalin-saline. Fixed tissues were paraffin embedded and stained with haematoxylin and eosin. Slides were microscopically examined at 20X magnification.

Statistical analysis

Statistical significance was determined using Student’s t test, unless otherwise stated, with P<0.05 taken as indicating a significant difference.

Supporting Information

Figure S1 Anti-CD25 antibody administration does not affect the outcome of low dose PyL infection. C57BL/6 mice were treated with either a single dose of 0.75 mg 7D4, or with 0.25 mg 7D4 combined with 0.75 mg PC61, 3 days prior to infection with 103 PyL parasites. The course of PyL in anti-CD25 antibody-treated mice and control mice was then followed by monitoring (A) parasitaemia, (B) anaemia and (C) mortality. * indicates significant differences (P<0.05) between 7D4 and 7D4 + PC61 treated mice. No significant differences were observed between the other groups of mice. Groups consisted of 4 mice and the results are representative of 2 independent experiments. Found at: doi:10.1371/journal.ppat.1000004.s001 (0.02 MB TIF)

Figure S2 Anti-CD25 antibody administration does not affect the outcome of PyNL infection. C57BL/6 mice were treated with either a single dose of 1 mg PC61 7 days prior to infection with PyNL (Expt 1) or with a single dose of 0.75 mg 7D4 or 0.25 mg 7D4 combined with 0.75 mg PC61 on the day of PyNL infection (Expt 2). The course of PyNL in anti-CD25 antibody-treated mice and control mice was then followed by monitoring (A, D) parasitaemia, (B, E) anaemia and (C, F) weight loss. No mortality occurred in any of the groups during either experiment. * indicates significant differences (P<0.05) between PC61 treated and control mice. No significant differences were observed between 7D4, 7D4 + PC61 treated mice and control mice. Groups consisted of 3–5 mice per group. Found at: doi:10.1371/journal.ppat.1000004.s002 (0.02 MB TIF)

Figure S3 Anti-CD25 antibody administration does not affect the outcome of PyL infection in BALB/c mice. - BALB/c mice were treated with either a single dose of 0.75 mg 7D4, or with 0.25 mg 7D4 combined with 0.75 mg PC61, 3 days prior to infection with 104 PyL pRBC or with 3 repeated injections of 0.5 mg 7D4 on days −3, −1 and +5 relative to PyL infection. The efficiency of depletion of CD25+ and Foxp3+ CD4+ T cells was determined in peripheral venous blood by measuring the expression of (A) CD25 and (B) Foxp3 on CD4+ T
cells. The course of PyL in anti-CD25 antibody-treated mice and control mice was then followed by monitoring (G) parasitaemia, (D) anaemia and (E) mortality. Symbols represent significant differences (p < 0.05) between groups: *7D4 vs PBS, ∆7D4/PC61 vs PBS, +7D4/PC61 vs 7D4 x3, #7D4 vs 7D4/PC61. Groups consisted of 5 mice.

Found at: doi:10.1371/journal.ppat.1000004.s003 (0.15 MB TIF)

References

1. Artavanis-Tsakonas S, Konig R, Riley EM (2003) The war between the malaria parasite and the immune system: immunoregulation and immunopathology. Clin Exp Immunol 133: 145–152.

2. Engwaerda C, Belnoue E, Gruner AC, Renia L (2005) Experimental models of cerebral malaria. Curr Top Microbiol Immunol 297: 103–143.

3. Omer FM, Kurtzhals JA, Riley EM (2000) Maintaining the immunological balance in parasitic infections: a role for TGF-beta? Parasitol Today 16: 18–23.

4. Omer FM, Riley EM (1998) Transforming growth factor beta production is inversely correlated with severity of murine malaria infection. J Exp Med 180: 1119–1125.

5. Omer FM, de Souza JB, Riley EM (2003) Differential induction of TGF-beta regulates proinflammatory cytokine production and determines the outcome of lethal and nonlethal Plasmodium yoelii infections. J Immunol 171: 5430–5436.

6. Omer FM, Kurtzhals JA, Hao C, Maizel A, Renia L (2000) Effects of in vivo administration of anti-IL-10 or anti-IFN-gamma monoclonal antibody on the host defense mechanism against Plasmodium yoelii yoelii infection. J Vet Med Sci 62: 383–387.

7. Kobayashi F, Mori T, Masui T, Fujino T, Watanabe Y, et al. (1996) Production of interleukin 10 during malaria caused by lethal and nonlethal variants of Plasmodium yoelii yoelii. Parasitology Research 82: 383–391.

8. Kossodo S, Monsio C, Julliard P, Velu T, Goldman M, et al. (1997) Interleukin-10 modulates susceptibility in experimental cerebral malaria. Immunology 91: 536–540.

9. Li C, Corraliza I, Langhorne J (1999) A defect in interleukin-10 leads to fatal cerebral malaria in mice. Infection Immunity 67: 4435–4442.

10. Li C, Sanni LA, Omer F, Riley E, Langhorne J (2003) Pathology of Plasmodium chabaudi chabaudi: differential susceptibility of gene-targeted mice deficient in transforming growth factor beta antibodies. Infect Immun 71: 4850–4856.

11. Linke A, Kuhn R, Muller W, Honavar N, Li C, et al. (1996) Plasmodium chabaudi chabaudi: differential susceptibility of gene-targeted mice deficient in IL-10 to an erythrocytic-stage infection. Exp Parasitol 84: 253–263.

12. Wu Y, Wang QH, Zheng L, Feng H, Liu J, et al. (2006) Plasmodium yoelii Distinct CD4(+)/CD25(+) regulatory T cell responses during the early stages of infection in susceptible and resistant mice. Exp Parasitol.

13. Hisaeda H, Maeoka Y, Iwakawa D, Okada H, Himeono K, et al. (2004) Escape of malaria parasites from host immunity requires CD4(+) CD25(+) regulatory T cells. Nat Med 10: 29–30.

14. Sakaguchi S (2005) Naturally arising Foxp3-expressing CD25(+)/CD4(+) regulatory T cells in immunological tolerance to self and non-self. Nat Immunol 6: 345–352.

15. Belkaid Y, Piccirillo CA, Mendez S, Shevach EM, Sacks DL (2002) CD4(+)/CD25(+) regulatory T cells control Leishmania major persistence and immunity. Nature 420: 502–507.

16. Taylor MD, LeGoiff T, Hariri A, Malone E, Allen JE, et al. (2005) Removal of regulatory T cells in experimental autoimmune encephalomyelitis reverses tissue reverses hyperactivity and leads to flarial parasite clearance in vivo. J Immunol 174: 4924–4933.

17. Kursar M, Koch M, MITTRUCKER HW, NOUAILLES G, BONFANTE K, et al. (2007) Cutting Edge: Regulatory T Cells Prevent Efficient Clearance of Mycobacterium tuberculosis. J Immunol 178: 2601–2603.

18. Belkaid Y, Rouse BT (2005) Natural regulatory T cells in infectious disease. Nat Immunol 6: 353–360.

19. Anderson CF, Ofukka M, Kuchroo VJ, Sachs D (2007) CD4(+)/CD25(+)Foxp3(-) Th1 cells are the source of IL-10-mediated immune suppression in chronic cutaneous leishmaniasis. J Exp Med 204: 285–297.

20. Jankovic D, Kullberg MC, Feng CG, Goldszmid RS, Collazo CM, et al. (2007) Macrophage-mediated but gamma interferon-independent innate immune responses control the primary wave of Plasmodium yoelii parasitemia. Infect Immun 75: 5806–5810.

21. Kamanaka M, Kim ST, Wan YY, Sutterwala FS, Lara-Tegero M, et al. (2006) Expression of Interleukin-10 in Intestinal Lymphocytes Detected by an Intraepithelial CD4(+)Foxp3(-) Regulatory T Cell Subset. J Immunol 177: 5374–5380.

22. Cooper KN, Blount DG, Haffalla JC, van Rooijen N, de Souza JB, et al. (2007) Macrophage-mediated but gamma interferon-independent innate immune responses control the primary wave of Plasmodium yoelii parasitemia. Infect Immun 75: 5806–5810.

23. Perry JA, Oliver CS, Burnett RC, Avery AC (2005) Cutting Edge: The Acquisition of TLR Tolerance during Malaria Infection Impacts T Cell Activation. J Immunol 174: 5921–5925.

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Author Contributions

Conceived and designed the experiments: KC JS ER. Performed the experiments: KC DB MW. Analyzed the data: KC DB MW. Contributed reagents/materials/analysis tools: YB MK RF. Wrote the paper: KC ER.
44. Walther M, Woodruff J, Edle F, Jeffries D, Toungren JE, et al. (2006) Innate immune responses to human malaria: heterogeneous cytokine responses to blood-stage Plasmodium falciparum correlate with parasitological and clinical outcomes. J Immunol 177: 5736–5745.

45. Dodoo D, Omer FM, Todd J, Akamori BD, Koram KA, et al. (2002) Absolute levels and ratios of proinflammatory and anti-inflammatory cytokine production in vitro predict clinical immunity to Plasmodium falciparum malaria. J Infect Dis 185: 971–978.

46. Hisaeda H, Hamano S, Mitoma-Obata C, Tetsutani K, Imai T, et al. (2005) Absolute T cell derived IL-10 regulates immunity to malaria. Am J Pathol.

47. Seddiki N, Santner-Nanan B, Martinson J, Zaunders J, Sasson S, et al. (2006) CD127 expression inversely correlates with FoxP3 and suppressive function of human regulatory and activated T cells. J Exp Med 203: 1701–1711.

48. Liu W, Putnam AL, Xu-Yu Z, Soot GL, Lee MK, et al. (2006) CD127 expression inversely correlates with FoxP3 and suppressive function of human CD4+ T reg cells. J Exp Med 203: 1693–1700.

49. Amante FH, Stanley AC, Randall LM, Zhou Y, Haque A, et al. (2007) A Role of Natural Regulatory T Cells in the Pathogenesis of Experimental Cerebral Malaria. Am J Pathol.

50. Nie CQ, Bernard NJ, Schofield L, Hansen DS (2007) CD4+ Regulatory T Cells Suppress CD4+ T-Cell Function and Inhibit the Development of Plasmodium berghei-Specific TH1 Responses Involved in Cerebral Malaria Pathogenesis. Infect Immun 75: 2275–2282.

51. Walther M, Woodruff J, Edle F, Jeffries D, Toungren JE, et al. (2006) Innate immune responses to human malaria: heterogeneous cytokine responses to blood-stage Plasmodium falciparum correlate with parasitological and clinical outcomes. J Immunol 177: 5736–5745.

52. Strauch UG, Obermeier F, Grunwald N, Gurster S, Dunger N, et al. (2005) Influence of intestinal bacteria on induction of regulatory T cells: lessons from a transfer model of colitis. Gut 54: 1546–1552.

53. Murray PJ, de Souza JB, Corran PH, Sultan AA, Riley EM (1997) T cell-derived interleukin 10 secreted by interleukin 27-stimulated T cells. J Exp Med 174: 2934–2941.

54. Nylen S, Maurya R, Eidsmo L, Manandhar KD, Sundar S, et al. (2007) Splenic accumulation of IL-10 mRNA in T cells distinct from CD4+CD25+ Foxp3+ regulatory T cells in human visceral leishmaniasis. J Exp Med 204: 805–817.

55. Anderson CF, Mendez S, Sacks DL (2005) Nonhealing infection despite Th1 polarization produced by a strain of Leishmania major in C57BL/6 mice. J Immunol 174: 2934–2941.