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Adhesion of *Escherichia coli* under flow conditions reveals potential novel effects of FimH mutations

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Abstract FimH-mediated adhesion of *Escherichia coli* to bladder epithelium is a prerequisite for urinary tract infections. FimH is also essential for blood-borne bacterial dissemination, but the mechanisms are poorly understood. The purpose of this study was to assess the influence of different FimH mutations on bacterial adhesion using a novel adhesion assay, which models the physiological flow conditions bacteria are exposed to. We introduced 12 different point mutations in the mannose binding pocket of FimH in an *E. coli* strain expressing type 1 fimbriae only (MSC95-FimH). We compared the bacterial adhesion of each mutant across several commonly used adhesion assays, including agglutination of yeast, adhesion to mono- and tri-mannosylated substrates, and static adhesion to bladder epithelial and endothelial cells. We performed a comparison of these assays to a novel method that we developed to study bacterial adhesion to mammalian cells under flow conditions. We showed that *E. coli* MSC95-FimH adheres more efficiently to microvascular endothelium than to bladder epithelium, and that only endothelium supports adhesion at physiological shear stress. The results confirmed that mannose binding pocket mutations abrogated adhesion. We demonstrated that FimH residues E50 and T53 are crucial for adhesion under flow conditions. The coating of endothelial cells on biochips and modelling of physiological flow conditions enabled us to identify FimH residues crucial for adhesion. These results provide novel insights into screening methods to determine the effect of FimH mutants and potentially FimH antagonists.

Introduction Infection with *Escherichia coli* is the most frequent cause of septicaemia in humans and commonly originates from the urinary tract [1]. Uropathogenic *E. coli* (UPEC) adhere to bladder epithelium in a process mediated by type 1 fimbriae via FimH engaging uroplakin 1a on urothelium, leading to urinary tract infection [2, 3]. Subsequently, FimH promotes invasion and is critical for blood-borne dissemination to other tissues [4]. Thus, in neonatal meningitis, FimH is essential for the localisation of UPEC to brain microvascular endothelium and invasion of the meninges [5, 6]. This establishes the pathogenic importance of FimH-mediated adhesion beyond the urinary tract.

FimH is located at the tip of type 1 fimbriae expressed by Gram-negative pathogens, including *E. coli*, *Salmonella*
enterica and Klebsiella pneumoniae [7, 8]. It has two domains: an N-terminal lectin domain of FimH containing the mannose binding pocket (MBP) responsible for bacterial adhesion to cellular ligands and a C-terminal pilin domain that connects FimH to the fimbrial rod [9]. Introduction of shear stress after initial binding induces allosteric interactions between the lectin and pilin domains that increase the affinity of mannose for the MBP through a catch bond mechanism [10, 11].

Over the past decade, mutated FimH have been used extensively to probe the molecular basis for its binding to mannose, most commonly in studies performed under static conditions [7, 10] or bacterial adhesion to bladder carcinoma cell lines [13] as end points. These have provided considerable insights into the molecular basis for the MBP binding with MBP through a catch bond mechanism [10, 11].

Materials and methods

Chemicals

Alpha-D-mannopyranoside (mannoside), RNase B (triammannosylated-3 M) and bovine serum albumin (BSA) were from Sigma-Aldrich (St. Louis, MO, USA). D-Mannose-BSA (mono-mannose-1 M) (14 atom spacer) was from Dextra Laboratories (Reading, UK), and 0.05 % Trypsin-EDTA and HEPES were from Life Technologies (Carlsbad, CA, USA).

Antibodies

The following polyclonal antibodies were used for Western blot: uroplakin 1a (ABIN955479, 1:100, antibodies-online, Atlanta, GA, USA) and beta-actin (A2066, 1:500, Sigma-Aldrich). Secondary antibodies conjugated with alkaline phosphatase were from Promega (1:5000, Madison, WI, USA).

Cell lines

Human dermal microvascular endothelial cells (G1S1) and conditionally immortalised glomerular endothelial cells (GeNC) were cultured using standard validated methods [15, 16]. GeNC were propagated at 33 °C (proliferation phase) and differentiated at 37 °C for 5 days prior to each experiment [15]. The human transitional cell carcinoma cell lines 5637 (ATCC HTB-9) and HT-1376 [17] and SV40-transformed urothelial cell line SV-HUC [18] were all kind gifts from Michael Wirth (University of Vienna).

Bacterial strains and GFP labelling of bacteria

Escherichia coli were labelled with green fluorescence protein (GFP) using phase 1 transduction of gfp:bla from E. coli strain OS56 into the multiply disabled E. coli MSS28 (E. coli MG1655 Δfim Δflu) [19], resulting in strain E. coli MSC95. The gfp gene was inserted into the chromosome of MSS28 using the Lambda Red System with the lambda red proteins encoded on the plasmid pTP223, which includes a gene for tetracycline resistance [20, 21] (kindly provided by Antony Poteete). As the source of the drug resistance cassette, pKD4 carrying a kanamycin cassette was used [22].

Escherichia coli MSC95 completely devoid of all fimbriae was used as the FimH-negative control strain. Escherichia coli MSC95 expressing FimH (MSC95-FimH) was derived from E. coli PC31 fimH [23] located on the pMAS4 plasmid, together with pPKL115 carrying the entire fim gene cluster with a knock-out mutation in fimH [24] and used as standard FimH-bearing control strain.

Site-directed mutagenesis of fimH

Mutations were introduced into the fimH gene from E. coli PC31 carried by the pMAS4 plasmid [23] by site-directed mutagenesis using the Phusion Site-Directed Mutagenesis Kit (F-541, Thermo Scientific, Waltham, MA, USA), following the manufacturer’s instructions. Specific primers were designed for each desired point mutation. For the expression of type 1 fimbriae carrying the mutated FimH protein, plasmids carrying the mutated fimH gene were individually transformed into MSC95 containing the plasmid pPKL115 carrying the fim gene cluster with a knock-out mutation in fimH [24]. Recombinant strains were grown in LB medium supplemented with 10 μg/ml chloramphenicol and 100 μg/ml ampicillin.

Yeast agglutination assay

The ability of the recombinant fimH mutant strains to express a D-mannose binding phenotype was examined by yeast agglutination using an established method [25]. Briefly, 20 μl of 1 % (v/w) yeast in PBS were mixed with 20 μl of serial
dilutions (non-diluted up to 1:16) of bacterial suspensions in PBS (normalised to OD_{500}=0.3) of either MSC95-FimH or mutant strains on a microscopy slide, and the dilution at which agglutination occurred was recorded.

**Bacterial adhesion under static conditions**

GFP-expressing *E. coli* MSC95-FimH (4 × 10^6 CFU) were incubated with confluent mammalian cell lines in 12-well cell culture plates for 30 min on ice to prevent internalisation. The cultures were then washed three times with PBS to remove non-adherent bacteria before the mammalian cells were detached with trypsin-EDTA and the resulting single-cell suspensions were analysed by flow cytometry (LSRFortessa SORP, Becton Dickinson, San José, CA, USA). Bacterial adhesion was quantified from the intensity of the GFP signal from single endothelial or urothelial cells identified by forward/sideward scatter, thus excluding GFP signals associated with cell clusters and/or from free bacteria. The results were analysed with FlowJo (Tree Star, Inc., Ashland, OR, USA) and expressed as the adhesion index, defined as the percentage of GFP-positive *E. coli* bound to mammalian cells. The adhesion of mutant *E. coli* strains to the cell lines was expressed in the results as a percentage of adhesion of the MSC95-FimH parent strain. The mannose dependence of adhesion was assessed by suspending the *E. coli* in media containing 4 % mannose for 10 min on ice before the experiment.

**Bacterial adhesion under flow conditions**

Vena8 Fluoro+ biochips (Cellix, Dublin, Ireland) were coated overnight with either 200 μg/ml D-mannose-BSA (1 M), 100 μg/ml RNase B with high 3-mannose (3 M) residues or 2 % BSA alone at 4 °C and blocked prior to use with PBS + 0.2 % BSA. A total of 1 × 10^6 *E. coli* prepared as described above were added to the substrates. The biochips were set up and washed according to the manufacturer’s instructions using the VenaFlux Assay Software (Cellix). The specificity of binding was assessed by pre-incubating *E. coli* MSC95-FimH with 2 % mannose in PBS for 10 min prior to the experiment. Adhesion of bacteria under a shear stress of 1 dyne/cm² was recorded every second in phase contrast and the settings were equal in all conditions (exposure time 344 ms, magnification 32×) for 5 min. In some experiments (stop/flow), 1 dyne/cm² was exerted and paused for 5 min once bacteria were observed in the HPF, before 1 dyne/cm² shear stress was re-applied. This, however, induced some gaps between the cells; any *E. coli* adhering to this were excluded, as mentioned above. The total numbers of *E. coli* that were adherent were counted as above. *Escherichia coli* was considered adherent when they were adhering for at least five frames at the end of the 5 min. Excluding criteria were *E. coli* that adhered to any plastic surface.

**Transmission electron microscopy**

Transmission electron microscopy (TEM) was performed to confirm the expression of intact fimbriae on all mutant strains. Five independent TEM micrographs (final magnification 60,000×) were analysed from each mutant strain and were used to count the number of fimbriae along three circumferential areas of 500 nm each in which individual fimbriae were clearly distinguishable. The total number of fimbriae was then calculated from the circumferential outline of the bacteria (~4500 nm) using ImageJ.

**Structural modelling**

The crystal structure of FimH (PDB ID: 2VCO) [27] was uploaded in Visual Molecular Dynamics [28] (VMD, University of Illinois, Urbana–Champaign, IL, USA). The residues that we experimentally tested were highlighted.

**Statistical analyses**

All calculations were made using GraphPad Prism 5.0 (GraphPad Prism Software, La Jolla, CA, USA) and p-values <0.05 were considered significant. Absolute values for the number of adherent bacteria were summarised as means ± standard error of the mean (SEM) and the significance of
differences between them was assessed by Student’s t-test. Adhesion index assays were expressed as medians with interquartile ranges and illustrated using box and whisker plots. The overall significance was tested by Kruskal–Wallis followed by, when appropriate, individual two-tailed Mann–Whitney tests. In both cases, individual pairwise p-values were corrected for multiple comparisons using the Benjamini–Hochberg method, with a false discovery rate of α set to 0.05 [29]. For the correlation data, the average adhesion of the mutant strains was analysed; MSC95-FimH and MSC95 were excluded from the correlation calculations.

Results

Generation and characterisation of E. coli strains expressing mutated FimH

Using MSC95-FimH as the parent strain [23], we generated a panel of 14 GFP-tagged E. coli strains that uniquely express type 1 fimbriae with different alanine point mutations in the MBP of FimH (listed in Table 1). The residues that were selected for mutation directly interact with mannose (N46, D47, P49, D54, Q133, N135 and D140) and have been shown to

| Mutant strain | yeast agglutination | 1M (#) | 1M (norm %) | 3M (#) | 3M (norm %) |
|---------------|---------------------|--------|-------------|--------|-------------|
| MSC95-FimH    | 1:4                 | 21.7±2.4 | 100         | 20.6±2.0 | 100         |
| MSC95         | NA                  | 0      | 0           | 0      | 0           |
| N46A*         | NA                  | 0.3±0.3 | 1 (0-2)     | 6.0±1.0 | 11 (2-21)   |
| D47A*         | 1                   | 0      | 0           | 0      | 0           |
| P49A*         | NA                  | 0      | 0           | 0      | 0           |
| E50A          | 1                   | 6.4±2.5 | 34 (11-55)  | 8.2±1.7 | 56 (19-74)  |
| T53A          | 1:2                 | 14.1±4.4 | 58 (12-115) | 16.6±4.3 | 68 (23-85)  |
| D54A          | NA                  | 0      | 0           | 0      | 0           |
| Y55A          | 1:2                 | 8.7±3.1 | 17 (10-24)  | 12.3±4.4 | 38 (0-78)   |
| V56A          | 1:2                 | 11.3±4.0 | 44 (0-64)  | 9.5±0.9 | 65 (65-67)  |
| T57A          | 1                   | 4.0±1.7 | 15 (12-33)  | 7.8±3.1 | 14 (12-48)  |
| Q133A         | 1                   | 0      | 0           | 0.5±0.5 | 3 (0-6)     |
| N135A         | 1                   | 0      | 1 (0-2)     | 2.5±0.3 | 0           |
| D140A         | 1                   | 0.3±0.2 | 2 (0-4)     | 1.5±1.5 | 0           |

The ability of E. coli FimH mutants to adhere to mannose substrates under flow and to agglutinate yeast. Binding properties were comparable in all three assays; however, some mutants exhibited marked differences in their ability to bind either 1 M or 3 M. Titres for yeast agglutination was the highest dilution at which agglutination still occurred. The adhesion of mutant E. coli to 1 M and 3 M was assessed, and the raw numbers of binding E. coli were normalised (norm) against E. coli numbers obtained in the same experiment with MSC95-FimH. Data represent mean % (range: lower limit %–upper limit %). MBP: Mannose binding pocket; 1 M: D-mannose-BSA; 3 M: RNase containing 3-mannose residues; NA: no agglutination. An asterisk (*) indicates that the strains had dysmorphic fimbriae.
bind to bladder tissue [27, 30, 31]. Two other residues (E50 and T53) were characterised in their ability to agglutinate yeast [32] and were included to examine their influence on MBP–mannose interactions. Three further residues are located at the back of the MBP, but their influence on adhesion was only analysed regarding mannose binding (Y55 and T57 [32]) or not at all (V56). The use of this panel of FimH mutations allowed us to systematically analyse the influence of each residue on adhesion to mammalian cells and binding to mannosylated substrates, and relate our results to previous findings.

Since FimH mutations can disrupt fimbriogenesis [33], we assessed the integrity of the fimbriae of the mutant FimH strains by electron microscopy. Three strains had reduced numbers of dysmorphic fimbriae (N46A, D47A and P49A) and were excluded from further analysis. The remaining nine mutant strains had quantitatively and qualitatively normal fimbriae when compared to the parental MSC95-FimH strain (one-way ANOVA $p = 0.24$) (Fig. 1) and were used to analyse bacterial adhesion to yeast, to biochemical substrates and to mammalian cells.

**FimH-dependent bacterial adhesion to 1 M and 3 M**

First, we tested FimH function by the established yeast agglutination assay, which provides a swift but semi-quantitative assessment [12, 34]. While D54A did not bind yeast, weak agglutination was observed for E50A, T57A, Q133A and N135A. The mutants T53A, Y55A and V56A adhered yeast with moderate titres, when compared to the native FimH bearing strain MSC95-FimH (Table 1). MSC95 lacking fimbriae did not cause yeast agglutination. The mutants analysed here have different yeast agglutination patterns, but do not necessarily reflect in vivo conditions [35].

Next, we quantified FimH-mediated adherence to mannosylated substrates under flow conditions [10] that should better reflect physiological conditions in vivo. Binding to mono-mannosylated (1 M) proteins is crucial for uropathogenic *E. coli*, while binding to tri-mannosylated (3 M) proteins has been reported to be important for non-uropathogenic *E. coli* for colonisation elsewhere [36, 37]. The MSC95-FimH parental strain showed strong adherence to yeast and efficiently bound to the 1 M substrate, D-mannose-BSA (21.7 ± 2.4 bacteria/HPF), and to the 3 M substrate, RNase B (20.6 ± 2.0 bacteria/HPF). The MBP mutations showed abrogated adhesion to 1 M (range 0–58 %) and 3 M (range 0–68 %) compared to the parent strain MSC95-FimH (Fig. 2). The results of two tests correlated in five of the nine strains tested: E50A and T53A bound moderately to yeast and to both 1 M [34 % (11–55 %)] and 58 % (12–115 %) of MSC95-FimH binding, respectively]
and 3 M [56% (19–74%) and 68% (23–85%)] (Table 1). In contrast, D54A did not agglutinate yeast nor did it bind to 1 M or 3 M. Mutant D140 was able to weakly agglutinate yeast but did not bind to 1 M [2% (0–4%)] or 3 M (0%). Mutant Y55A agglutinated yeast but was severely impaired in binding to both 1 M [17% (10–24%)] and 3 M [38% (0–78%)] (Table 1). V56A, Q133A and N135A weakly agglutinated yeast but showed either moderate binding to 1 M [V56A: 44% (0–64%)] or none [Q133A: 0% and N135A: 1% (0–2%)], with similar binding to 3 M [65% (65–67%), 3% (0–6%) and 0%, respectively]. Mutant strains with dysmorphic fimbriae did not bind to 1 M or 3 M and did not agglutinate yeast (N46A, P49A) or only weakly (D47A) (Table 1).

**FimH-dependent bacterial adhesion to cells under static conditions**

We analysed the effect of different FimH mutations on adhesion to the relevant cell types by adapting the standard flow cytometry-based bacterial adhesion assay performed under static conditions [38]. FimH-dependent adhesion of GFP-tagged *E. coli* was quantified using an adhesion index calculated from the percentage of mammalian cells with adherent bacteria (Fig. 3a). Multiply-disabled MSC95-FimH adhered significantly better to endothelium derived from skin (G1S1) or glomeruli (GEnC) [G1S1; adhesion index 57.3%, interquartile range (IQR) 51.9–71.4; GEnC; 55.2% (IQR 40.2–63.9)] than to bladder urothelial cell lines, derived from normal bladder (SV-HUC; 37.3%; IQR 26.5–57.6, *p* < 0.001 to G1S1, *p* < 0.01 to GEnC) or bladder carcinomas (5637 cells; 11.6%; IQR 6.7–27.5, *p* < 0.01 to GEnC and HT-1376; 20.2%; IQR 16.3–22.4, *p* < 0.01 to GEnC). Adhesion was both FimH- and mannose-dependent because it did not occur in the absence of FimH expression and was abrogated by pre-incubation with 4% mannoside (Fig. 3b). Despite the strong expression of uroplakin 1a, the primary receptor for FimH on urothelium (Fig. S1), adhesion to urothelial...
cells was significantly less than to endothelial cells. To determine if the differences remained under physiological conditions, we analysed the adhesion of MSC95-FimH to urothelial and endothelial cells under flow.

**FimH-dependent bacterial adhesion under shear stress conditions**

Blood-borne bacteria encounter endothelium under shear stress, estimated to be around 1 dyne/cm² in both glomerular and dermal capillaries [39, 40]. At this rate of flow, MSC95-FimH adhered effectively to confluent monolayers of G1S1 and GEnC in microfluidic flow chambers (G1S1: 12.6 ± 4.4 bacteria/HPF, GEnC: 10.2 ± 2.7) (Fig. 4a, b). Adhesion occurred rapidly without the rolling behaviour that is characteristic of leukocyte adhesion and, once adherent, the bacteria did not detach. Again, adhesion was entirely FimH- and mannose-dependent (data not shown and Fig. 4a). Unexpectedly, MSC95-FimH did not adhere to any of the bladder cell lines under shear stress (Fig. 4a, b). To stimulate the conditions of bladder voiding, we allowed MSC95-FimH to adhere for 5 min under static conditions before applying flow (stop/flow assay). MSC95-FimH adhered effectively under static conditions and was not dislodged from either bladder or endothelial cells by subsequent flow conditions, up to 15 dyne/cm² (data not shown), but MSC95-FimH still adhered more efficiently to GEnC in this assay (GEnC: 38.0 bacteria/HPF; SV-HUC: 5.9; 5637 cells: 2.5; and HT-1376 cells: 4.4) (Fig. 4c).

**Adhesion of mutant FimH to mammalian cells under static and shear stress conditions**

We then measured the ability of the FimH mutant strains to adhere to SV-HUC and GEnC in our static adhesion assay. Seven of the strains (MSC95, D54A, Y55A, T57A, Q133A, N135A and D140A) adhered less well, between 0 and 31.3 % of MSC95-FimH (Fig. 5a). By contrast, alanine substitution of E50 and T53 had little or no effect (Fig. 5a) and adhesion to both cell lines was inhibited by mannoside, excluding the acquisition of novel mannos-
independent binding sites as an explanation (Fig. S2). The remaining mutant strain V56A showed partial [63 \% (55–71 \%)] adhesion compared to the MSC95-FimH to GEnC, but this was also not due to mannose-independent binding (Fig. 5aa and S2).

The effect of MBP mutations was even more pronounced under shear stress. Alanine substitution abrogated adhesion to GEnC in all mutant strains (Fig. 5b). Remarkably, the E50A and T53A mutants adhered normally under static conditions but had over 90 \% reduced adhesion under shear stress (0.6 ± 0.3 and 0.5 ± 0.2 bacteria/HPF, respectively) compared to 8.5 ± 1.9 of MSC95-FimH (both p < 0.001, Student’s t-test). Only V56A that retained 63 \% of its adhesive properties to GEnC under static conditions also adhered to these cells under flow [3.6 ± 0.5 bacteria/HPF (p = 0.08)] or 42 \% of MSC95-FimH (Fig. 5b).

MSC95-FimH did not adhere to SV-HUC under shear stress and, so, we used the stop/flow assay to compare the adhesion of the mutants. Again, most strains did not adhere in this condition (Fig. S3). E50A and T53A mutants that adhered normally under static conditions exhibited around 50 \% reduced adhesion (3.5 ± 1.5 and 6.0 ± 1.0 bacteria/HPF, respectively) compared to 8.8 ± 1.1 of MSC95-FimH. The only mutant strain not affected was V56A, which adhered more effectively than the parent strain (14.0 ± 1.0).
Correlation between FimH mutant strains analysed with different methods

The correlation of the FimH mutants (Fig. S4) between adhesion to 1 M and the static adhesion to cells was 0.96 for GEnC and 0.75 for SV-HUC (Spearman, $p = 0.0002$ and $p = 0.026$, respectively). Between static adhesion and to 3 M, the correlation was similar; 0.84 for GEnC and 0.64 for SV-HUC ($p = 0.006$ and $p = 0.066$, respectively). The correlation of the mutant strains between adhering under static or shear stress conditions to GEnC was 0.59 ($p = 0.097$). The correlation for the different mutants to GEnC under flow was equal for both 1 M and 3 M; 0.71 for 1 M and 0.72 for 3 M (both $p = 0.037$). The adhesion of the mutants to SV-HUC in the stop/flow assay correlated well with adhesion to GEnC under continuous flow; 0.76 ($p = 0.021$) but less with adhesion under static conditions to SV-HUC; 0.55 ($p = 0.133$). Finally, the correlation between adhesion to 1 M and 3 M was 0.90 ($p = 0.0020$). Thus, there is a strong correlation between the adhesion on mannosylated substrates and static adhesion to cells. However, the more stringent conditions of adhesion under flow revealed stronger effects of FimH mutations that were masked when adhesion was analysed under static conditions. The results demonstrate the critical influence of MBP residues on bacterial adhesion to endothelium under shear stress conditions, even if there is a high correlation between SV-HUC and GEnC; 0.91 ($p = 0.0013$) (Fig. S4). They also show that binding to mannosylated substrates is not a true reflection of the effect of FimH mutations. Therefore, an assay in which cells are used mimicking the physiological conditions encountered in vivo is more informative and provides additional insight into the adhesive properties of FimH variants.

Discussion

Type 1 fimbriated pathogens, like E. coli, adhere to a variety of biological surfaces, both epithelia and endothelia.
Accordingly, they have adapted by modifying the sequence and structure of adhesins like FimH to enable improved adhesion under different—static or shear stress—conditions [10, 41]. Our study explores the methodologies used to analyse FimH-mediated adhesion of E. coli using mutations in the MBP of FimH [31]. We first used yeast agglutination as an established screening assay [34] to test the functionality of MBP of FimH [31]. We measured the agglutination potential of these mutant strains and found that they correlated relatively well with other reports [31]. Minor differences could lie in the glycosylation of guinea pig red blood cells compared to yeast, as reported by others [47–49]. Regardless, the results with the remaining nine strains with normal fimbriae demonstrated that the mannose binding pocket is crucial for the adhesion of E. coli to endothelial cells [31]. Comparison of adhesion to endothelium under static and shear stress conditions revealed that E50 and T53 were essential under shear stress. This only partly correlated with binding to mannosylated substrates under identical assay conditions, demonstrating that there are important differences for the adhesion of isolated mannosylated substrates and mammalian cells. E50 has been demonstrated to not interact with the sugar ligand, but to stabilise R98, which is, thereby, oriented to interact with the ligand [50]. Due to its location on the backbone of the MBP, T53 interactions with 3-mannose are blocked by the side chains of I52 and D54. The effect of T53 mutations can, therefore, only be indirect through changes in its backbone energetic minimum, which could lead to gentle changes in the 3-mannose binding site [27, 50]. This supports a model in which these amino acid residues contribute to initial interaction between FimH and its cellular receptors, and that facilitates subsequent insertion of their terminal mannose residues into the MBP to establish firm adhesion. This would be consistent with the lack of reported mutations of these residues in clinical E. coli isolates, suggesting an important evolutionary conserved contribution to virulence [51].

The E. coli strain MSC95-FimH adhered efficiently to microvascular endothelium both under static conditions and shear stress, whereas adhesion to all three urothelial cell lines tested in static conditions was less efficient and appeared abrogated by flow. The observation that MSC95-FimH fails to adhere to bladder epithelial cells under flow is in apparent contrast to an earlier study [44]. However, the discrepancy probably relates to technical differences between the two studies and specifically to our use of an E. coli strain specifically engineered to express type 1 fimbriae exclusively, which enabled us to examine the unique effects of FimH.

In our experiments, differences between FimH-mediated adhesion to urothelial cells under static and shear stress conditions could be regarded, as the experimental results correlate stop/flow conditions that occur in the urinary bladder during voiding. Allowing MSC95-FimH to adhere under static conditions before applying shear stress resulted in an increase of bacteria bound to the non-neoplastic urothelial cell line SV-HUC...
and, to a lesser extent, to the neoplastic cell lines 5637 and HT1376 that were not dislodged by flow. Using this setup, we confirmed the reduced adhesion capabilities of E50A and T53A, although the effect was less obvious than during continuous flow. This supports the model in which these residues contribute to mannose binding. When mutated, FimH is limited in making contact with mannose residues under shear stress conditions.

In summary, we show that, by analysing bacterial adhesion under physiologically relevant conditions, novel effects of FimH mutations are observed. Our study thus highlights the fact that the results obtained from investigating FimH adhesion are dependent on the methodological differences of the assays used and the physiological situation they represent. The results with MSC95-FimH, expressing native or mutated variants of FimH, identified the importance of the MBP that critically influences the adhesion under flow, and provide novel insights into screening methods to determine the effect of FimH mutants and potentially FimH antagonists.

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Compliance with ethical standards

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Conflict of interest The authors declare no conflict of interest.

Ethics approval This article does not contain any studies with human participants or animals performed by any of the authors.

Informed consent This study did not involve human subjects or samples; therefore, no informed consent was required.

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