Pomelo peel oil alleviates cerebral NLRP3 inflammasome activation in a cardiopulmonary resuscitation rat model

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Abstract. The NLR family pyrin domain-containing 3 (NLRP3) inflammasome, which is composed of NLRP3, apoptosis-associated speck-like protein containing a CARD (ASC) and pro-caspase-1 protein complexes, is activated by the reactive oxygen species (ROS) that are associated with ischemia-reperfusion (I/R) and are involved in brain damage. Pomelo peel oil (PPO) exhibits antioxidant activity. However, it is unclear whether PPO is able to attenuate NLRP3 inflammasome-induced inflammation and pyroptosis. Healthy male Sprague-Dawley rats were subjected to 7 min of cardiac arrest via trans-esophageal electrical stimulation, followed by cardiopulmonary resuscitation (CPR). The rats were then treated with PPO prior to reperfusion for 24 h. Hematoxylin and eosin staining was used to evaluate brain tissue and cell damage. In the brain tissues, reactive oxygen species (ROS) were assayed, immunofluorescence was used to analyze the expression of NLRP3 and western blotting was performed to determine the expression levels of neuroenolase (NSE), NF-xB, interleukin-1 (IL-1β), gasdermin D (GSDMD) and the NLRP3 inflammasome. Treatment of the rats with PPO significantly decreased the pathological damage of the brain tissue and reduced the expression of NSE, production of ROS and secretion of NF-xB, NLRP3, IL-1β and GSDMD. In conclusion, these results demonstrate the ability of PPO to protect the brain against I/R injury in rats after CPR by a mechanism involving inhibition of the inflammation and pyroptosis mediated by NLRP3 inflammasome activation.

Introduction

Cardiac arrest (CA) and subsequent cardiopulmonary resuscitation (CPR) induces systemic organ tissue ischemia/reperfusion (I/R) injury, particularly cerebral I/R injury (CIRI), which directly affects the prognosis and quality of life of patients. Numerous pathophysiological mechanisms of I/R injury, including oxidative stress, amino acid toxicity, energy metabolism, calcium overload, apoptosis and autophagy, have been targeted by interventions for brain protection (1-3). In addition, the pathological effects of NLR family pyrin domain-containing 3 (NLRP3) inflammasome-induced inflammation and pyroptosis exacerbate CIRI (4). During I/R, excessive quantities of oxygen free radicals attack tissues or cells and induce damage-associated molecular patterns that are recognized by specific pattern recognition receptors, thereby resulting in caspase-1 precursor (pro-caspase-1) activation and cell death; this is the classical pathway underlying caspase-1-dependent pyroptosis (5). NLRP3 accompanied by pyroptosis is widely observed in neurons, microglia and astrocytes and is associated with neurological diseases (6). When damage-associated molecular signaling occurs, NLRP3 interacts with apoptosis-associated speck-like protein containing a CARD (ASC) and recruits pro-caspase-1. Subsequently, pro-caspase-1 is cleaved to form mature caspase-1p20, which cleaves pro-interleukin (IL)-1β and gasdermin D (GSDMD) to form mature IL-1βp17 and the GSDMD-N domain (GSDMD-N). The GSDMD-N protein has been demonstrated to execute pyroptosis (7). The GSDMD-N domain targets the plasma membrane and forms a pore of diameter 10-14 nm that allows mature IL-1βp17 to leak out and ions and water to enter, which causes inflammation and cell pyroptosis (8-11). Therefore, the inhibition of NLRP3 inflammasome activation by antioxidants may be clinically useful for the treatment of I/R injury.

A previous study has demonstrated that, in most organs, including the brain, heart, liver, lungs and intestines, activation of the NLRP3 inflammasome aggravates CIRI (12). Although NLRP3-mediated CIRI has been reported in the middle
cerebral artery occlusion (MCAO) model (1), it remains unclear whether the global CIRI following CA/CPR, which causes more extensive critical brain damage than MCAO, is associated with the NLRP3 inflammasome and pyroptosis. Pomelo (Citrus maxima), which is a citrus fruit belonging to the genus Rutaceae, has high nutritional value (13). Traditional Chinese medicine considers pomelo to be beneficial for the prophylaxis and treatment of nervous system disorders, peroxidation damage, cardiovascular disease, bruising, hyperlipidemia, wounds, acne, osteoarthritis and fatigue (14–18). Pomelo peel contains numerous beneficial chemicals, including flavonoids, coumarins, essential oils and limonoids. The essential oils of pomelo peel, extracted using a cold-press method, were found to contain 94.15% terpenes, comprising 55.92% limonene, 31.17% β-myrcene, 3.16% β-pinene, 1.42% ocimene and 1.24% β-copaene, when analyzed using gas chromatography-mass spectrometry (GC-MS) (19). Pomelo peel oils (PPOs) have been demonstrated to exhibit strong antioxidant, anti-inflammatory, antiviral and antibacterial activities (20–22). Therefore, we hypothesize that PPO may inhibit the NLRP3 inflammasome-associated inflammation and pyroptosis induced by CIRI. In the present study, the chemical composition of PPO was analyzed. In addition, the effects of PPO on reactive oxygen species (ROS) and the NLRP3 inflammasome, GSMDL, IL-1β and NF-kB expression were examined in a rat model of CA/CPR in order to elucidate the potential neuroprotective effect of PPO.

Materials and methods

PPO extract. Fresh and mature pomelo peels [C. maxima (Burm.) Merr. cv. Shatian Yu, identified by the Institute of Botany, Guangxi Zhuang Autonomous Region, Chinese Academy of Science; specimen ID: (W.Y.Wang:201909001)] were obtained from the village of Shatian in Rongxia, Guangxi, China. The steam distillation method was used to extract the PPO (23). The outer layer of the pomelo peel was washed and cut into ~5x5-mm pieces. The pieces of peel were combined with distilled water (1:2) and put in a distiller, where they were boiled for 1 h. The PPO was separated from the water using a cooling condenser and stored at -4°C.

GC-MS analysis. The compounds in the PPO were qualitatively analyzed using GC-MS (TRACE™ 1300; Thermo Fisher Scientific, Inc.). Helium was injected as a carrier gas at a flow rate of 1 ml/min. The temperature was increased from 45 to 250°C according to the procedure. The heating program is maintained at 45°C for 1 min, then at 10°C/min to 165°C for 2 min, then at 1.5°C/min to 180°C for 2 min and finally at 10°C/min to 250°C for 2 min. The temperatures of the injector and detector were set at 250°C. Mass spectra were scanned from m/z 41-400 amu. The electron impact ionization energy was 70 eV. Identification of the detected compounds was performed by comparing the mass spectra with published data. Samples were identified using the Retention Time Locked database (NIST MS Search 2.3; http://www.Inchi-trust.org/download/105/licence.pdf) with Deconvolution Reporting Software (MSD ChemStation F. 01.03.2357; 1989-2015; Agilent Technologies, Inc.).

CA/CPR animal model. In total, 60 male Sprague-Dawley (SD) rats (body weight, 220-250 g; age 7-8 weeks) were provided by the Experimental Animal Center of Guangxi Medical University (License number SYXK Gui 2014-0003). All animals were handled in accordance with the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health (24). The study was approved by the Animal Care and Use Committee of Guangxi Medical University. The rats were raised at 22±2°C, humidity 50-60%, 12-h light/dark cycle, were free to eat and drink and fasted 12 h before the operation. Prior to modeling, the rats were anesthetized with an intraperitoneal injection of 2% pentobarbital sodium (30 mg/kg). A PE50 tube was inserted into the left femoral artery and connected to a physiological recorder (BL-420E; Chengdu TME Technology Co., Ltd.) via a pressure converter to monitor the blood pressure. Another PE50 tube was inserted into the femoral vein for the injection of drugs.

The CA model was established according to the method reported by Chen et al (25). A temporary pacemaker electrode (Chengdu TME Technology Co., Ltd.) was inserted into the esophagus of the rat at a depth of ~7 cm. Ventricular fibrillation was induced by the application of 12 V of direct current for 1 min. The criteria used to define CA were as follows: ECG exhibiting ventricular fibrillation and a mean arterial pressure of <20 mmHg. Following 7 min of CA, CPR was initiated using a frequency of mechanical chest compression of 180/min, with a depth of 25-30% of the anterior posterior diameter of the chest. Ventilator-assisted ventilation (DH-150; Zhejiang University Medical Instrument Co., Ltd.; https://med4868.yixie8.com/) was applied immediately through endotracheal intubation after 1 min of CPR (tidal volume, 6 ml/kg; respiration rate, 70 breaths/min; positive end expiratory pressure, 0 cmH₂O). In addition, 20 mg/kg epinephrine was administered through the PE50 tube in the femoral vein. The restoration of spontaneous circulation (ROSC) standard was as follows: Supraventricular rhythm (sinus, atrial and borderline heart rhythm) accompanied by a mean arterial pressure of >60 mmHg lasting for >1 min. Surviving rats were randomly divided into five groups: NS group (n=8; 0.9% physiological saline 200 µl), glycerin group (Gly group; n=8; 10% glycerin 200 µl), PPO-low group (PPO-L group; n=8; 10 mg/kg PPO), PPO-medium group (PPO-M group; n=8; 20 mg/kg PPO) and PPO-high group (PPO-H; n=8; 40 mg/kg PPO). The mass of 1 ml PPO, weighed using an electronic balance (BSA223S; Sartorius AG), was 800 mg. In each group treated with PPO, the PPO was combined with 10% glycerol to a total volume of 200 µl. Following 1 min of ROSC, the rats were intravenously injected with the respective treatment. In addition, 8 rats were randomly selected as the sham group, which underwent exposure of the left femoral artery and femoral vein followed by vascular ligation without CA/CPR. All procedures were performed by two skilled operators.

Preparation of brain tissues. All rats were anesthetized using pentobarbital (30 mg/kg) by intraperitoneal injection 24 h post reperfusion. Three rats in each group were perfused with 4% paraformaldehyde for hematoxylin and eosin (H&E) staining and immunofluorescence experiments. Cerebral cortices were immediately harvested from the remaining 5 rats in each group and stored at -80°C for subsequent western blot analysis.
**Results**

**Chemical composition of PPO.** The extraction yield of PPO was 0.2%. The PPO was obtained as a faint yellow, transparent liquid. The total ion chromatogram obtained from the GC-MS analysis of the PPO is shown in Fig. 1. A total of 17 compounds were detected (Table I). The most abundant compound in the PPO was limonene at 88.683%, followed by nootkatone (5.732%) and myrcene (1.027%; Table I).

**Antioxidant activities of PPO.** The antioxidant activities of PPO in the cerebral cortex tissues of the CIRI model rats are compared with those in the NS and Gly groups. However, in the PPO-H group, the PPO treatment significantly decreased the levels of ROS compared with those in the NS and Gly groups.
PPO ameliorates cerebral cell morphological changes and reduces the expression of NSE. To investigate the effects of PPO on CIRI, cerebral cell morphology was evaluated with H&E staining (Fig. 2A). In the sham group, the cell structures were complete and the tissue structure was tightly packed, with uniform staining and clearly visible nuclei. By contrast, the cerebral tissues from the NS and Gly groups exhibited fewer intact cells; cells were disordered, with intense staining and vacuoles clearly evident. The tissues treated with PPO exhibited improved cell morphology compared with those in the NS and Gly groups. NSE has been widely studied as a brain injury marker (21,26). Therefore, the expression of NSE was analyzed by western blotting (Fig. 2C). Compared with that in the sham group, the expression of NSE was significantly increased in the NS, Gly, PPO-L and PPO-M groups (Fig. 2D). However, the expression of NSE in the PPO-H group was significantly lower than that in the NS, Gly and PPO-L groups (Fig. 2D). These results indicate that PPO ameliorates cerebral injury in CA/CPR model rats.

PPO inhibits the expression of NF-κB. When NF-κBp105 is lysed, it forms NF-κBp50 and thereby serves pro-inflammatory functions (27). No significant differences in the expression of NF-κBp105 were detected among the groups (Fig. 2E and F). However, in comparison with the sham group, the NS and Gly groups exhibited significant increases in NF-κBp50 and phosphorylated NF-κBp105 (NF-κBp-p105) levels (Fig. 2G-I). Following treatment with PPO, the levels of NF-κBp50 and NF-κBp-p105 decreased in a dose-dependent manner, and the level of NF-κBp-p105 was significantly decreased in the PPO-H group compared with the NS and Gly groups (Fig. 2H). The NF-κBp-p105/NF-κBp105 ratio also exhibited a significant reduction in the PPO-H group compared with the NS and Gly groups after PPO treatment (Fig. 2I). These results suggest that PPO partially inhibits NF-κB-mediated inflammation.

PPO decreases the expression of NLRP3 in the cerebral cortex. To determine whether PPO is able to inhibit NLRP3...
activation in the microglia, the expression and co-expression of NLRP3 and IBA-1 were detected using immunofluorescence double staining. The expression of NLRP3 and IBA-1, and the co-expression of NLRP3 + IBA-1 increased markedly in the microglia following CR/P (Fig. 3A). The expression of NLRP3, IBA-1 and NLRP3 + IBA-1 was significantly increased in the NS group. After treatment with PPO, the expression levels of NLRP3, IBA-1 and NLRP3 + IBA-1 in the PPO-H group were significantly lower than those in the NS group, indicating that activation of microglia was inhibited (Fig. 3B). The results indicate that PPO decreased the expression of NLRP3 and activation of the microglia.

**PPO downregulates the expression of pyroptosis-associated proteins NLRP3, ASC, caspase-1, IL-1β and GSDMD.** To investigate the effects of PPO on the inflammatory response and pyroptosis-associated proteins in the CA/P (CR/P) model rats, the levels of NLRP3, ASC, caspase-1, IL-1β and GSDMD were analyzed using western blotting (Fig. 4). Pro-caspase-1, pro-IL-1β and GSDMD were activated to form caspase-1p20, IL-1βp17 and GSDMD-N, respectively. The expression levels of NLRP3, ASC, caspase-1p20, IL-1βp17 and GSDMD-N in the NS group were significantly higher than those in the sham group. Following treatment with PPO, the levels of these proteins were significantly lower than those in the NS group. These data indicate that PPO reduced the pyroptosis-associated protein cascade.

**Discussion**

In the present study, PPO was obtained from fresh and mature Shatian pomelo peels. The main ingredient of PPO was found to be limonene (88.683%), followed by nootkatone (5.732%) and myrcene (1.027%). The results of the in vivo experiments revealed that PPO reduced the production of ROS induced by global CIRI in rats following CA/CPR and ameliorated the pathological injury of the cerebral cortex. The effects of PPO on the NLRP3 inflammasome response and pyroptosis were also demonstrated in this model. Furthermore, PPO downregulated pyroptosis-associated protein levels.

The proportions of limonene and other components detected in the present study differ from those in other studies (19,28); this may be due to different extraction methods, growth environments, isolation conditions and provenance. However, consistent with other studies (19,29), limonene was the main component of the PPO extracted via distillation in the present study. PPO has been reported to contain 94.15% terpenes (19), with structures that include phenolic hydroxyl groups and unsaturated double bonds, thereby providing electrons that are able to reduce free radicals to form less active substances and chelate metal ions to prevent the generation of free radicals (30). PPO has also been shown to exert an antioxidant effect against superoxide anion radical formation (19). Previous studies have demonstrated that the overproduction of ROS is associated with CIRI (31,32), which is consistent with the results of the present study. Furthermore, the results indicate that PPO inhibited the production of ROS associated with global CIRI. It has been reported that limonene has antioxidant activities (33). Therefore, we hypothesize that the antioxidant activity of PPO is associated with the limonene it contains.

The generation of excessive ROS leads to cellular damage and triggers the activation of microglia and immune pathways (34). The present study demonstrated that CIRI upregulated the expression of NLRP3 in the microglia, and treatment with PPO reduced CIRI-induced activation of the microglia and the expression of NLRP3. The microglia are

### Table I. Chemical components of pomelo peel oil.

| No. | Compound                | Molecular formula | Retention time (min) | Peak area (%) |
|-----|-------------------------|-------------------|----------------------|--------------|
| 1   | α-pinene                | C_{10}H_{16}       | 5.309                | 0.127        |
| 2   | Myrcene                 | C_{10}H_{16}       | 6.677                | 1.027        |
| 3   | Limonene                | C_{10}H_{16}       | 8.079                | 88.683       |
| 4   | γ-terpinene             | C_{10}H_{16}       | 10.356               | 0.250        |
| 5   | Limonene oxide          | C_{10}H_{18}O     | 11.207               | 0.270        |
| 6   | α-terpineol             | C_{10}H_{18}O     | 14.223               | 0.149        |
| 7   | Carvone                 | C_{10}H_{18}O     | 15.273               | 0.403        |
| 8   | Geranyl acetate         | C_{12}H_{20}O_2   | 15.837               | 0.234        |
| 9   | Carophyllene            | C_{15}H_{24}      | 16.332               | 0.505        |
| 10  | α-caryophyllene         | C_{15}H_{24}      | 21.530               | 0.193        |
| 11  | α-gurjunene             | C_{15}H_{24}      | 22.393               | 0.351        |
| 12  | 8-cedren-13-ol          | C_{15}H_{20}O     | 23.939               | 0.183        |
| 13  | Nerolidol               | C_{15}H_{20}O     | 29.380               | 0.208        |
| 14  | Globulol                | C_{15}H_{20}O     | 31.674               | 0.184        |
| 15  | Nootkatone              | C_{15}H_{20}O     | 35.392               | 5.732        |
| 16  | Osthole                 | C_{15}H_{18}O_1   | 41.095               | 0.196        |
| 17  | Eicosane                | C_{15}H_{24}      | 43.278               | 0.197        |
| -   | Total                   | -                 | -                    | 98.89        |
Figure 2. PPO improves cell morphology, reduces ROS levels and attenuates the expression of NSE and NF-κB in a cardiopulmonary resuscitation rat model. (A) Hematoxylin and eosin staining of brain tissue from rats in the (A-a) sham, (A-b) NS, (A-c) Gly, (A-d) PPO-L, (A-e) PPO-M and (A-f) PPO-H groups. A large number of cerebral cortex cells with normal morphology were observed in the sham group (indicated by black arrows). However, numerous abnormal cerebral cortex cells exhibiting nuclear pyknosis, intense staining, vacuolation, swelling and necrosis (indicated by red triangles) were visible in the NS and Gly groups. Cell morphology was ameliorated in the PPO groups. Magnification, x400. (B) Antioxidant activity of PPO determined by an ROS assay. Representative western blots of NSE (C) and (D) quantified results. (E) Representative western blots of NF-κB and (F-I) quantified results. Band intensities were quantified using densitometric software. The band intensities of (F) NF-κBp105, (G) NF-κBp50 and (H) NF-κBp-p105 were normalized to that of GAPDH, and (I) the NF-κBp-p105/NF-κBp105 ratio was calculated. Data are presented as the mean ± SEM of two independent experiments (n=3). *P<0.05, **P<0.01 and ***P<0.001 vs. the sham group; #P<0.05 and ##P<0.01 vs. the NS group; &P<0.05 and &&P<0.01 vs. the Gly group; @@P<0.01 vs. the PPO-L group (n=5). PPO, pomelo peel oil; ROS, reactive oxygen species; NSE, neuroenolase; NS, physiological saline; Gly, glycerin; PPO-L, 10 mg/kg PPO; PPO-M, 20 mg/kg PPO; PPO-H, 40 mg/kg PPO; NF-κBp-p105, phosphorylated NF-κBp105.
the resident immune cells in the brain. Following activation, microglia switch their phenotypes between two polarizations, M1 and M2, which can release large quantities of pro-inflammatory and anti-inflammatory cytokines, respectively. In a previous study, NLRP3 immunoreactivity was detected in the IBA-1-labeled microglia of mice with CIRI following transient MCAO, which induced the overproduction of the M1 microglia-regulated pro-inflammatory cytokine IL-1β (35). Therefore, the phenotypic changes occurring in the microglia following activation require identification.

NLRP3, a specific pattern recognition receptor, initiates cell death processes (pyroptosis) as soon as it is activated by factors induced by pathological damage. It promotes the pyroptosis-associated protein cascade and causes inflammation and pyroptosis (36). The examination of factors in this cascade in the present study revealed that the upregulation of NLRP3, ASC, caspase-1p20, IL-1βp17 and GSDMD-N induced in rats by CA/CPR was attenuated by PPO treatment, suggesting that it targets NLRP3 inflammasome induced inflammatory responses and pyroptosis, and serves as an intervention against CIRI.

The control of NLRP3 activity can be divided into two processes, namely priming and activation. Priming prepares NLRP3 for subsequent activation (37) and may occur by an ROS-mediated non-transcriptional pathway (38,39). ROS released into the cytoplasm by damaged mitochondria also promote activation of the NLRP3 inflammasome (40,41). Since excess ROS production occurs during CIRI, antioxidants are able block pyroptosis-associated NLRP3 inflammasomes. Therefore, we hypothesize that the antioxidant activity of PPO resulted in the levels of NLRP3 inflammasome being low in the early stages of CIRI.

IL-1β is produced during CIRI; it induces cerebrovascular endothelial cells to express adhesion molecules, which mediate

Figure 3. PPO decreases the expression of NLRP3 in the microglia. (A) Immunofluorescence staining of rat brain slices. Green staining indicates NLRP3 protein expression, red staining indicates IBA-1 protein expression and DAPI staining indicates nuclei (blue). Magnification, x400 and x2,000. (B) Column charts showing the (B-a) NLRP3-positive area (%), (B-b) IBA-1-positive area (%) and (B-c) the amount of NLRP3 expression co-localized with IBA-1-positive microglia in each group quantified using image analysis software. All data are presented as the mean ± SEM (n=3). *P<0.05 and **P<0.01 vs. the sham group; #P<0.05 and ##P<0.01 vs. the NS group; &&P<0.01 vs. the Gly group. PPO, pomelo peel oil; NLRP3, NLR family pyrin domain-containing 3; IBA-1, allograft inflammatory factor 1; Gly, glycerin; PPO-L, 10 mg/kg PPO; PPO-M, 20 mg/kg PPO; PPO-H, 40 mg/kg PPO.
interaction between the endothelial cells and leukocytes, thereby promoting the infiltration of leukocytes into the brain tissues. Leukocyte infiltration activates microglial cells, which causes them to secrete inflammatory factors and aggravates brain injury. In addition, leukocytes adhere and accumulate in microvessels, causing blockages and reducing cerebral blood flow, which also aggravates brain injury. These pathological changes occur at a late stage after CIRI (42‑45). We hypothesize that the antioxidant activity of PPO reduces the secretion of IL‑1β from cells and thus attenuates the IL‑1β‑mediated inflammatory response.

An increase in ROS levels can stimulate the phosphorylation of IkB by casein kinase II or tyrosine kinases, and thereby induce NF‑κB gene transcription (46). NF‑κB mediates the priming of NLRP3 via the transcriptional pathway (39). NF‑κBp105/p50 is a member of the NF‑κB family, and NF‑κBp50 is produced from NF‑κBp105 by ubiquitin‑dependent limited proteolysis (27). The present study found that PPO significantly downregulated
the phosphorylation of NF-κB. The phosphorylation of NF-κBp105 directly affects the formation of NF-κBp50 (47). NF-κBp50 has been demonstrated to be involved in pyroptosis as an inflammasome promoter (48). Furthermore, a recent study indicated that the brain damage in MCAO rat models of CIRI may be associated with increased levels of NF-κBp50 (49). The results of the present study are consistent with this, and showed that the levels of NF-κBp50 increased in a global CIRI model. PPO downregulated the formation of NF-κBp50 in a dose-dependent manner; the antipyroptotic properties of PPO may be partly due to its antioxidant activity. Since significant differences were found, the results suggest that the transcriptional pathway of NF-κB-mediated NLRP3 activation may be one of the pathway in the present model.

In conclusion, the present study indicated that the NLRP3 inflammasome, which is associated with pyroptosis, is involved in CA/CPR-induced CIRI. Furthermore, PPO treatment downregulated the expression of multiple factors in the NLRP3 inflammasome and ameliorated brain cell death. PPO, as a mixture, may inhibit the inflammation and pyroptosis caused by activation of the NLRP3 inflammasome through its antioxidant capacity. However, further research is required to identify the active components and the underlying mechanism.

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Availability of data and materials

All data generated and/or analyzed during this study are included in this published article.

Authors' contributions

XSZ and LX contributed equally to this work, in terms of developing the concept and study design, acquiring, analyzing and interpreting the data, authenticate the raw data and drafting the manuscript. WYW, GYZ and XYT performed the experiments and analyzed the data. MHC designed and led the study. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The study was approved by the Animal Care and Use Committee of Guangxi Medical University (Nanning, China).

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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