Cloning and Characterization of 5E6(Ly-49C), A Receptor Molecule Expressed on a Subset of Murine Natural Killer Cells

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Summary

5E6 is a cell surface molecule expressed on a subpopulation of murine natural killer (NK) cells that are involved in the specific rejection of H-2^d or H-2^f (hemopoietic histocompatibility determinant) bone marrow cell grafts. Here, we isolated and cloned the gene encoding 5E6 and determined the nucleotide sequence of the cDNA. 5E6 is nearly identical to Ly-49C; the deduced amino acid sequence reveals a polypeptide of 266 amino acids with a molecular weight of 31,284 that contains multiple cysteine residues to explain its disulfide-linked homodimer structure and five potential N-linked glycosylation sites. 5E6 is a type II integral membrane protein with an extracellular carbohydrate recognition domain characteristic of C-type (Ca^{2+}-dependent) animal lectins. Chromosomal mapping indicates that 5E6 is located within the NK gene complex on chromosome 6. The sequence of 5E6 mRNA and the degree of glycosylation of 5E6 protein are under genetic control. Immunoprecipitation before removal of N-linked sugars reveals different size molecules. There are several nucleotide differences among BALB/c, B6, and NZB mRNAs; however, none of them would be expected to affect N-glycosylation. Of particular interest are two findings: (a) BALB/c, B6, and (BALB/c × B6)F1 5E6 reduced molecules are 65, 54, and 54 kD, and (b) the cDNA sequence of (BALB/c × B6)F1 is identical to B6. Thus, there appears to be allelic exclusion of 5E6 expression that may be related to the ability of F1 hybrid mice to reject parental H-2^d bone marrow cell grafts.

NK cells comprise a small population of lymphocytes that have a large granular morphology and are distinct from mature T and B cells and from cells of the myeloid lineage (1, 2). NK cells are defined as a functionally important immune effector population that display “natural” cytolytic activity against certain tumor cells and virally infected cells in a manner that is not MHC restricted (3, 4). NK cells also mediate the rejection of incompatible bone marrow cell allografts in lethally irradiated mice, possibly by the specific recognition of recessively inherited hemopoietic histocompatibility 1 (Hh-1)^1 antigens, a phenomenon known as “hybrid resistance” (5–7). The mechanism by which NK cells recognize bone marrow cells in a highly specific manner and mediate hybrid resistance is not known. However, it has been postulated that specific receptors are present on subsets of NK cells that allow for this recognition. Indeed, NK-specific molecules have been reported in humans (8–12), rats (13, 14), and mice (15–22).

Recently, several murine NK cell receptor-like molecules have been cloned and categorized into two genetically linked families termed the NKR-P1 and Ly-49 gene families. These gene families share some structural features and are located in close proximity on mouse chromosome 6 in an area that has been termed the NK gene complex (23, 24). These two families of molecules have been suggested to deliver opposing signals, with NKR-P1 molecules activating natural killer and Ly-49 molecules delivering inhibitory signals to NK cells when they encounter certain MHC class I molecules on target cells (23, 25).

Abbreviations used in this paper: A-LAK, adherent lymphokine-activated killer (cells); DGGE, denaturing gradient gel electrophoresis; Hh-1, hemopoietic histocompatibility 1; RI, recombinant inbred; UT, un-translated.
The mAb SW5E6 has been described as binding to a population of ~50% of murine NK cells of most strains tested. Previous experiments in this laboratory have shown that treatment of recipient mice with mAb SW5E6 eliminates the ability to reject H-2d but not H-2b bone marrow cell grafts (26, 27). In this study, we report the cloning, chromosomal mapping, and molecular characteristics of the 5E6 gene, as well as its unique expression in different strains of mice.

Materials and Methods

Cell Purification and Cell Culture. Murine cells were cultured in RPMI 1640 (GIBCO BRL, Gaithersburg, MD) supplemented with 10% FCS (Hyclone Laboratories, Logan, UT), 2 mM L-glutamine, 100 U/ml penicillin, 100 μg/ml streptomycin (Irvine Scientific, Santa Ana, CA), 1 mM sodium pyruvate, and 0.1 mM nonessential amino acids (GIBCO BRL) (complete RPMI). COS-7 cells have been described (28) and were cultured in DMEM (GIBCO BRL) with the same supplements (complete DMEM). T lymphokine-activated killer (A-LAK) cells were prepared as previously described (29) with minor modifications. Briefly, A-LAK cells were generated by culturing nylon wool nonadherent spleen cells in T25 flasks (Corning Inc., Corning, NY) in RPMI 1640 complete media (RPMI 1640 [GIBCO BRL] supplemented with 10% FCS [Hyclone Laboratories], 2 mM L-glutamine, 100 U/ml penicillin, 100 μg/ml streptomycin [Irvine Scientific], 1 mM sodium pyruvate, and 0.1 mM nonessential amino acids [GIBCO BRL]) containing 22.5 μM β-mercaptoethanol (Sigma Immunochemicals, St. Louis, MO), 1 μg/ml indomethacin (Sigma), 50 μg/ml gentamicin (GIBCO BRL), and 500 or 1,000 U/ml human rIL-2 (a kind gift from the Cetus Corporation, Emeryville, CA) (LAK culturing media) for 3 d at 37°C and 5% CO2. On day 3 of culture, the nonadherent cells were removed by gentle washing with warm complete RPMI. The adherent cells were further cultured and fed every 2–3 d with LAK culturing media, maintaining IL-2 at a concentration of 500 or 1,000 U/ml. Cells were harvested at 8–9 d of culture. LAK cells from C.B-17 scid/scid bone marrow were prepared in the same manner, with the exception of the unnecessary step of removing the nonadherent T cells on day 3.

Construction of cDNA Library. Total cytoplasmic RNA was prepared from day 9 C.B-17 scid/scid LAK cells by the acid-guanidine-phenol-chloroform method (30). Poly (A)+ RNA was prepared by oligo (dT)-cellulose chromatography using an mRNA separator system for cDNA synthesis and cloning (BILL-Life Technologies, UT) and 157 bp 3′ UT sequences, was amplified using the following primers: 5′-pGACTAGTT-GCATTCTGATGTCGTCGTCGGCGCCCGGCGCATCAGTCATCAGTTCGAC-3′ (first primer) and 5′-pGACTAGTT-GCATTCTGATGTCGTCGTCGGCGCCCGGCGCATCAGTCATCAGTTCGAC-3′ (second primer). Reverse transcription PCR amplification of the product (1/1) was used for TA cloning according to the instructions provided by the supplier (InVitrogen). The cloned genes were sequenced by using specific primers based on the sequence of 5E6 from BALB/c using the Sequenase system. The PCR mixture contained 1 μM of each primer, 200 μM dNTPs, 1 x PCR buffer, 1 U Ampli Taq DNA polymerase, 5 μl of cDNA reaction product and 2 mM magnesium chloride in a reaction volume of 50 μl. PCR conditions were 94°C 4 min, 35 cycles of 94°C 1 min, 55°C 1 min, 72°C 1 min followed by extension of 72°C for 7 min. 5 μl of the PCR products were run on 1.2% agarose gel and visualized by staining with ethidium bromide. A portion of the product (1 μ1) was used for TA cloning according to the instructions provided by the supplier (InVitrogen). The cloned genes were sequenced by using specific primers based on the sequence of 5E6 from BALB/c using the Sequenase system.

Surface iodination, Immunoprecipitation, Removal of N-linked Carbohydrates, and SDS-PAGE. Transiently transfected COS-7 cells
or A-LAK cells from various strains of mice were labeled with Na23H1 (Amersham Corp., Arlington Heights, IL) using lactoperoxidase as described (39). Cells were washed three times in PBS and lysed in 50 mM Tris, pH 8.0, 1% NP-40 (Sigma) containing 1 mM EDTA, 0.5 μM leupeptin, 0.7 μM pepstatin, and 150 μg/ml PMSF (Boehringer Mannheim Biochemicals, Indianapolis, IN) for 10 min on ice. The nuclei were removed by centrifugation at 12,000 g for 10 min. The lyastes were precleared with rabbit anti-mouse IgG (Jackson ImmunoResearch Laboratories) coated protein A-Sepharose (Pharmacia Fine Chemicals, Piscataway, NJ) for 30 min at 4°C while rocking. Precleared lysates were incubated with mAb SW5E6 or isotype control mAb 22B5 for 2 h at 4°C while rocking. Protein A-Sepharose coated with rabbit anti-mouse IgG was added for an additional 45 min at 4°C while rocking. The beads were recovered by centrifugation and washed three times with NET gel (50 mM Tris-HCl, pH 7.5, 150 mM NaCl, 0.1% NP-40, 1 mM EDTA, 0.025% gelatin, 0.02% sodium azide). The bound proteins were eluted by boiling for 5 min in SDS sample buffer for SDS-PAGE analysis or in antigen elution buffer (100 mM Tris-HCl, pH 7.5, 1% SDS, 1% β-mercaptoethanol) for removal of N-linked carbohydrates. Endoglycosidase F digestion was performed by removing the supernatant from the beads and adding endoglycosidase F buffer (0.1 M potassium phosphate, pH 6.1, 1% Triton X-100, 0.1% SDS, 45 mM EDTA, 1% β-mercaptoethanol) to a final volume of 1 ml. 1 U of endoglycosidase F/N-glycosidase F (Boehringer Mannheim Biochemicals) was added and the samples were allowed to digest overnight at 37°C. The protein was precipitated with TCA and washed three times with ice cold acetone followed by addition of 1 x SDS sample buffer and boiling for 5 min. Samples were analyzed on 12 or 7.5% slab gels according to the method of Laemmli (40). Gels were fixed, dried, and exposed to Hyperfilm-MP (Amersham Corp.) at -70°C.

Genetic Mapping of 5E6(Ly-49C). Mapping of 5E6(Ly-49C) was performed using a panel of 26 genomic DNAs representing the complete C57BL/6 X DBA/2 (B x D) recombinant inbred strain collection. Candidate primer sequences for the amplification of genomic fragments from 5E6(Ly-49C) were initially identified using the computer program Primer (kindly provided by Eric Lander, Massachusetts Institute of Technology, Cambridge, MA) to analyze the cDNA sequence of 5E6(Ly-49C). The primer sequences ultimately selected were: 5'TTCCCTGATTAATTTTCCAACC and 3'TCTTCTTCTTGTAAAGACTTCCCG. These sequences were from the 3' end of 5E6(Ly-49C) and amplified a 278-bp genomic fragment that had minimal homology with sequences in Ly-49. Polymorphisms between C57BL/6 and DBA/2 in this amplified fragment were identified by automated nucleotide sequence analysis using an automated DNA sequencer (model 373; Applied Biosystems, Inc., Foster City, CA) as previously described (41). Allelic forms of the amplified fragment were subsequently distinguished by denaturing gradient gel electrophoresis (DGGE) as previously described (42). Preliminary analysis using perpendicular gradients established that optimal variation in DGGE mobility occurred in a denaturing gradient of 30-70% for 150 μg/ml PMSF (Boehringer Mannheim Biochemicals, Indianapolis, IN) was put through a "hot start" of 75°C for 15 min followed by cooling to 20°C. DNA (100 ng) and Amersham Hot Tub Taq polymerase (1.25 U) were then added to each reaction mix. PCR conditions were 94°C 5 min, 35 cycles: 94°C 1 min, 62°C 1 min, and 72°C 1 min, followed by extension at 72°C for 7 min and storage at 4°C. A portion of the PCR product (9 μl of the 50 μl total) of each sample was electrophoresed on a 2% agarose gel and visualized by staining with ethidium bromide.

Results

Isolation and Characterization of 5E6 cDNA Clone. A cDNA library was constructed in the mammalian expression vector pME18S using mRNA isolated from IL-2-propagated C.B-17 (scid/scid) bone marrow cells. The library was transfected into COS-7 cells and these cells were then screened for surface expression of the 5E6 molecule using the transient expression-immunoselection procedure of Seed and Aruffo (33). After three rounds of transfection and selection by panning using the mAb SW5E6, 18 bacterial colonies were isolated at random for further analysis. Three of these plasmids contained inserts of ~1.2 kb and each of these clones directed the surface expression of 5E6 protein on transfected COS-7 cells as determined by staining with the 5E6 mAb and flow cytometry analysis (Fig. 1). Each of these three clones were identical at the 3' end and differed only in the length of the 5' untranslated region. The nucleotide sequence of the largest clone is shown in Fig. 2. Comparison of the predicted amino acid sequence of 5E6 to the databanks revealed 64% identity to the Ly-49A molecule (75% identity at the nucleotide level) (data not shown). The 5E6 polypeptide also contains an extracellular carbohydrate recognition domain characteristic of Ca2+-dependent lectins (45) and members of the Ly-49 gene family (23, 24). Comparison of the carbohydrate recognition domains of 5E6 and several other known C-type lectins shows that multiple residues are conserved and that 5E6 is a member of this group of molecules. The 5E6 cDNA was found to be identical to a previously reported molecule, Ly-49C (17), with the following exceptions. The 5E6 cDNA is 84 nucleotides longer at the 5' UT sequence and has a 78-bp deletion at nucleotide 995 in the 3' untranslated region corresponding to nucleotides 912-989 of the reported Ly-49C sequence. There are also two nucleotide differences at 279 and 280. We report a cysteine followed by an adenine, while Ly-49C is reported to have an adenine followed by a thymidine. This results in a change from asparagine in the 5E6 peptide to tyrosine in the Ly-49C peptide (data not shown). Comparison of amino acid sequence of 5E6 from different strains of mice is shown in Fig. 3.

The 5E6 cDNA Reveals Significant Polymorphism. Southern blots of HindIII-, BamHI-, and EcoRI-digested DNA from
Figure 1. Cell surface expression of 5E6 on transiently transfected COS-7 cells. COS-7 cells were stained with biotin-conjugated anti-5E6 mAb followed by PE-conjugated streptavidin (filled histograms) or with secondary reagent only (streptavidin-PE) (open histograms). Analysis was on a FACScan flow cytometer. (A) Untransfected COS-7 cells. (B) COS-7 cells transfected with pME18S vector only. (C) COS-7 cells transfected with 5E6 cDNA clone number 16. The two other positive CDNA clones showed similar staining patterns.

NZB, C3H, 129/J, DBA/2, BALB/c, and C57BL/6 liver hybridized to the full-length 5E6 cDNA revealed the presence of multiple bands (data not shown). Previous reports have shown that the closely related gene, Ly-49A, divides most common laboratory mouse strains into five polymorphic patterns (46). Our limited analysis shows that the 5E6 gene corresponds to the same pattern, with BALB/c, C3H, and DBA/2 having identical RFLP patterns and C57BL/6, NZB, and 129/J each having unique RFLP patterns. These results are consistent with the hypothesis that there are multiple members of the Ly-49 gene family, and indeed, several new molecules have recently been identified that have significant homology to Ly-49A (47).

The 5E6 Gene Maps to the NK Complex on Mouse Chromosome 6. PCR analysis using primers specific for 5E6 was performed using DNA from a panel of mouse–hamster somatic cell hybrids (48). Genetic marker analysis revealed that each hybrid had randomly segregated mouse chromosomes and that the panel as a whole was informative for assigning genes to any mouse chromosome (49). Products were obtained from the parental mouse cell DNA and from only those hybrids containing mouse chromosome 6 (data not shown). The percent discordancy for each chromosome using 17 different hybrid cell lines indicated that chromosome 6 had a 0% discordancy. These results place the 5E6 gene on chromosome 6 with a high degree of probability. Independently, the chromosomal location of 5E6(Ly-49C) was determined using the B × D recombinant inbred strain panel. Polymorphisms distinguishing the 5E6(Ly-49C) alleles present in the C57BL/6 and DBA/2 parental strains were sought in the

![Figure 2. Nucleotide sequence of the 5E6 cDNA insert and predicted amino acid sequence of the 5E6 peptide. The predicted transmembrane region is double underlined. Five potential N-linked glycosylation sites are underlined. The polyadenylation signal is in bold letters and underlined. The nucleotide sequence is numbered on the left side and the amino acid sequence is numbered on the right side. Cysteine residues are highlighted. These sequence data are available from GenBank under accession number U09739.](image-url)
sequence variations resulted in a readily detectable variation in mobility in denaturing gradient gel electrophoresis (DGGE), and this assay was used to determine the 5E6(Ly-49C) genotypes of all 26 (B × D) recombinant inbred (RI) strains.

The strain distribution pattern obtained for 5E6(Ly-49C) was identical to that reported for Ly-49A, Prp, and Ly-55 for 25 of the 26 RI strains, thus mapping 5E6(Ly-49C) to chromosome 6 in close proximity to Ly-49A and Ly-55. Representative results of the DGGE assay for 5E6(Ly-49C) polymorphisms between C57BL/6 and DBA/2 are illustrated in Fig. 4 A, which also presents the typing results for the single RI strain defining recombination between 5E6(Ly-49C) and Ly-49A (B × D No. 22). B × D No. 22 was previously reported to carry C57BL/6-derived alleles of Ly-49A (46). Our results with 5E6(Ly-49C) typing clearly indicate that B × D No. 22 carries a DBA/2-derived allele. This DGGE typing result for B × D No. 22 was confirmed by nucleotide sequence analysis (data not shown).

These results position 5E6(Ly-49C) within the current composite map for chromosome 6 as illustrated in Fig. 4 B. The loci listed on the left are the six reference loci for chromosome 6 defined in the current Chromosome 6 Committee Report (50). The (B × D) strain distribution pattern (SDP) for 5E6(Ly-49C) places it in close proximity with Ly-49A, Ly-55, and Prp, all of which have identical SDPs in the current reference data set for the (B × D) RI strains. Since Ly-55 and Ly-49A are placed 2 cm telomeric to Prp, in the current composite map of chromosome 6, this would tentatively place 5E6(Ly-49C) telomeric to Ly-55 and Ly-49A.

Analysis of the 5E6 Protein in Different Strains of Mice. The mAb SW5E6 has previously been shown to detect an apparent disulfide linked homodimer with ~54-kD subunits on the surface of C57BL/6 and NZB NK cells (26). In this study, initial immunoprecipitations using anti-5E6 antibody revealed that on BALB/c NK cells 5E6 is present as a homodimer with ~65 kD subunits, while on transfected COS-7 cells, 5E6 appears as a homodimer with ~54 kD subunits, as was seen in previous studies with C57BL/6 NK cells (Fig. 5). When N-linked carbohydrates were removed, both species appeared as ~31-kD proteins, indicating that the molecular weight difference can be attributed to a

Figure 3. Comparison of amino acid sequences of 5E6 from different strains of mice. The amino acid residues different from BALB/c only are shown. The potential N-linked glycosylation sites (Asn-X-Thr/Ser) are boxed.

Figure 4. (A) Representative data for the genetic typing of 5E6(Ly-49C) in the BXD RI strain panel using DGGE. The region of the denaturant gradient included within the area of the gel displayed is shown at the top and the genomic origins of the fragments electrophoresed are shown on the left. (B) Representation of the genetic map of chromosome 6 showing the location of 5E6(Ly-49C). Loci listed on the left are reference loci for chromosome 6 (50), with their respective loci in cM. An expanded view of the region containing 5E6(Ly-49C) is shown on the right. All distances between loci in cM are from the current composite map for chromosome 6 (50).
difference in glycosylation. When 5E6 antigen precipitated from BALB/c (H-2a) and C57BL/6 (H-2b) NK cell extracts were compared, it was discovered that a difference in molecular mass indeed exists between these two strains with BALB/c having ~65-kD subunits and C57BL/6 having ~54 kD subunits (Fig. 6). This difference can also be attributed to glycosylation since both appear as ~31-kD proteins upon removal of N-linked carbohydrates. We also compared 5E6 immunoprecipitates from BALB.B (H-2b on BALB/c background) and BALB/c x C57BL/6 (CB6F1), and it was observed that the 5E6 protein on CB6F1 NK cells exists as the smaller form (~54 kD), while on BALB.B NK cells, 5E6 is an intermediate size of ~60 kD (Fig. 7). To determine whether the different levels of glycosylation result from differences in the primary structure of the molecule, we amplified the coding region of 5E6 from different strains of mice. Comparison of the amino acid sequence of 5E6 (Fig. 3) revealed that there is no change in the number of potential N-linked glycosylation sites. Other factor(s) might therefore contribute to the different level of glycosylation of the molecule. The implications of these varying sizes of 5E6 will be discussed later.

Discussion

In the present study, we describe the molecular characterization of the gene encoding 5E6. The 5E6 molecule is expressed on the surface of murine NK cells as a disulfide-linked homodimer. Spleen cells of most mouse strains express 5E6 on ~40–50% of their NK cells, with the exception of 129/J, C57BR, and RII mice. We have previously established that depletion of the 5E6 § subset of NK cells in vivo abrogates the ability of recipient mice to reject bone marrow cell grafts of H-2d and H-2b origin. These studies suggested that the 5E6 § subset of NK cells, perhaps the 5E6 molecule itself, may be a receptor for Hh-1 determinant 2 (26, 27). We have now cloned the 5E6 cDNA using the mammalian expression-immunoselection procedure described by Seed and Aruffo (33) using a cDNA library made from IL-2-propagated NK cells from C.B-17 scid/scid bone marrow. The 5E6 gene was mapped to mouse chromosome 6 by two independent methods. It was also found to be genetically linked to NK1.1 and Ly-49, which are present in that portion of mouse chromosome 6 termed the NK gene complex (23).

A single transcript for the 5E6 gene 1.3 kb in length was present in total RNA isolated from A-LAK cells from the strains C57BL/6, BALB/c, DBA/2, C3H, and NZB but not from 129/J and RII strains (data not shown). No detectable transcript was found in RNA from several normal tissues isolated from BALB/c mice. The mAb SW5E6 was made by immunizing 129/J mice with purified NK cells from C57BL/6 mice, therefore, it was expected that 129/J would not express 5E6. The significance of the lack of expression
in the strain RI1 is not known at this time. It is interesting that while 5E6+ cells apparently recognize and reject H-2d/Hh-1d bone marrow cell grafts, they are not deleted during development in H-2d homozygous mice as evidenced by the presence of message in BALB/c and DBA/2 NK cells. It is also interesting to note that injection of SW5E6 antibodies had a minimal effect on the ability of H-2d/Hh-1d-type mice to clear radiolabeled YAC-1 tumor cells, but had a much more pronounced effect on animals with other H-2/Hh-1 types (26).

It appears as if the cytolytic function of 5E6+ cells against YAC-1 targets is somehow downregulated in H-2d/Hh-1d mice. It is possible that the 5E6+ subset of NK cells may have multiple functions in different strains of mice. This point will be mentioned later.

The molecular weight of the 5E6 protein varied significantly depending on the strain of mice from which the NK cells were derived. Thus far, our analysis has revealed three different sizes of the 5E6 protein: 54 kD, present on C57BL/6 and CB6F1 mice; 60 kD, present on BALB/c mice; and 65 kD, present on BALB/c mice. When N-linked carbohydrates were removed, each of these had an identical molecular mass of 31 kD, indicating that the difference in size between the strains is attributed to a varying amount of glycosylation. One possibility is that the differences in the size of glycosylated molecule could be caused by the differences in the primary structure of the molecule in different strains of mice. However, as shown in Fig. 3, there is no difference in the number of potential N-linked glycosylation sites. Thus, other factor(s) might contribute to the level of glycosylation of the molecule in different strains of mice. Comparison of 5E6 from CB6F1 showed that it is identical to the B6 allele (Fig. 3).

To determine whether F1 mice express any BALB/c allele, we amplified by reverse transcription PCR a 184-bp \textit{s} region that contains two restriction sites (BgII and MboI) in the BALB/c allele, but are absent in the B6 allele. Ethidium bromide staining of the PCR products and their digests with BgII and MboI showed similar pattern for B6 and CB6F1 (data not shown). Thus it appears that the CB6F1 NK cells contain only the B6 form of 5E6 mRNA. However, we cannot absolutely rule out that minute quantities of the BALB/c form of 5E6 mRNA are not present in CB6F1 NK cells because very low quantities of DNA cannot be visualized by ethidium bromide staining. It is noteworthy that both CB6F1 and C57BL/6 mice are able to reject BALB/c (H-2^d/Hh-1^d) bone marrow cell grafts. The expression of only the C57BL/6 form of 5E6 in the CB6F1 could help explain some aspects of hybrid resistance. The differences seen in the sizes of BALB/c, BALB.B, and C57BL/6 5E6 molecules (65, 60, and 54 kD, respectively) indicates that a factor associated with the H-2 complex is important, but there are obviously other factors not associated with the H-2 complex that allow for a variance in the glycosylation of 5E6. It is entirely possible that the degree of glycosylation present on the 5E6 protein is a determining factor in the recognition of alloantigens on bone marrow stem cells. It remains to be seen what exact effect the difference in glycosylation of the 5E6 protein has on NK cell function.

The Ly-49A molecule is the only other well-characterized member of this multigene family. It is unknown if Ly-49 exhibits a similar pattern of glycosylation in different mice strains. Ly-49A+ cells have been shown to receive a negative signal when H-2d class I molecules are encountered, thus preventing the lysis of these targets (24, 51). Similar results have been obtained using an in vitro assay for hybrid resistance (Yu, Y. Y. L., J. Roland, V. Kumar, and M. Bennett, unpublished data). These data suggest that the 5E6 molecule on CB6F1 NK cells may receive negative signals from H-2^d (probably H-2^K^d) but not H-2^d targets. It was noted that CB6F1 5E6+ NK cells could not lyse C57BL/6 (H-2^d) or B10.D2(R107) (H-2^K^d/Ps^K^d/D^d1^d) T cell Con A blasts unless the effector cells were pretreated with P(ab')_2 anti-5E6 mAb. This data argues that negative signals received from MHC class I molecules, rather than positive signals received from putative Hh-1 determinants, mediate the presence or absence of "hybrid resistance," at least in experiments using the in vitro system. However, Yu et al. also observed that BALB/c (H-2^d) Con A blasts, which are resistant to lysis by 5E6+ syngeneic NK cells, become susceptible to lysis when anti-5E6 mAb is present, as if the 5E6 molecule can also recognize H-2a/Hh-1a-type 5E6(Ly-49C)–
transfected cells were recently shown to bind to various class I molecules (52). It is conceivable that the 5E6 molecule may function as an activating molecule when H-2\textsuperscript{a} targets are encountered by H-2\textsuperscript{a} or H-2\textsuperscript{ad} NK cells, but as a negative signaling receptor with respect to H-2\textsuperscript{b} targets. This provides an attractive explanation for the differences seen in the glycosylation of 5E6 in H-2\textsuperscript{d} homozygous versus other strains of mice.

It is also noteworthy that NK cells of NZB (H-2\textsuperscript{a}) origin express the smaller (54 kDa subunit) form of 5E6, as opposed to BALB/c (H-2\textsuperscript{b}), which expresses 5E6 with 65-kDa subunits (data not shown). This is particularly interesting in view of the fact that treatment of recipient C57BL/6 mice with anti-5E6 mAb had no effect on their ability to reject NZB bone marrow cells (53). Therefore, we cannot be absolutely certain that NZB bone marrow cells express Hh-1 determinant.

Indeed, one study maps the gene for hybrid resistance to H-2 b or H-2 bxa NK cells, but as a negative signal of 5E6 in H-2 a homozygous versus other strains of mice.

Another recent report (Murphy, W. J., A. Raziuddin, L. Mason, V. Kumar, M. Bennett, and D. L. Longo, manuscript submitted for publication) demonstrate that 5E6\textsuperscript{+} and 5E6\textsuperscript{-} subsets of NK cells can function differently in the regulation of hematopoiesis in different strains of mice. They observed that in H-2\textsuperscript{d} mice, 5E6\textsuperscript{+} cells produce more GM-CSF than IFN-\gamma and stimulate hematopoiesis, while 5E6\textsuperscript{-} cells produce more INF-\gamma than GM-CSF and inhibit hematopoiesis. In contrast the reverse is true in H-2\textsuperscript{b} mice: 5E6\textsuperscript{+} cells make more INF-\gamma than GM-CSF and inhibit hematopoiesis, while 5E6\textsuperscript{-} cells make more GM-CSF than INF-\gamma and stimulate hematopoiesis. These differences in cytokine secretion patterns in 5E6\textsuperscript{+} cells of H-2\textsuperscript{a} and H-2\textsuperscript{b} strains of mice may also be related in some manner to the variation we report in the glycosylation of 5E6.

Note added in proof: Further PCR analysis has detected low levels of the BALB/c form of 5E6 mRNA in CB6FI NK cells.

The authors would like to thank Thomas Moore for flow cytometry analysis.

This work was supported by grants CA-36921, CA-36922, AI-20451, and CA-09082 from the National Institutes of Health, and by a grant from the Texas Department of Ladies’ Auxiliary, Veterans of Foreign Wars.

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Received for publication 4 August 1994 and in revised form 23 February 1995.

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