An in vitro evaluation of surface roughness, color stability and bacterial accumulation of lithium disilicate ceramic after prophylactic periodontal treatment

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ABSTRACT

Objective: To evaluate the effect of scaling procedures using different ultrasonic tips on the surface roughness, color stability and bacterial accumulation of lithium disilicate ceramic. Material and Methods: Scaling procedure was carried out using ultrasonic scaler (Satalec, Acteon, North America) with stainless-steel tip (US), titanium tip (UT) and plastic tip (UP), on disc shaped lithium disilicate samples cemented into a cavity prepared onto the labial surface of freshly extracted bovine teeth (10 samples per group). The samples were stored in coffee solution in an incubator at 37°C for 12 days, which is equivalent to 1 year of coffee consumption. The surface roughness was measured before and after the scaling procedure using a profilometer and atomic force microscopy. The color parameters were measured before and after scaling and staining procedures using VITA Easyshade Advance 4.0 according to the CIE L*a*b* color order system. The samples were then incubated with Streptococcus mutans (S. mutans) suspension. After incubation, the plates with 30 to 300 typical colonies of S. mutans were counted in a colony counter and mean values of colony forming units were obtained (CFU/mL). Results: The titanium scaling tip showed a statistically significant higher mean values of change in surface roughness ΔRa and bacterial count than the plastic scaling tip. Color changes (ΔE) were not a statistically significant among the groups. The results showed a statistically significant positive (direct) correlation between surface roughness

RESUMO

Introdução: Avaliar o efeito de procedimentos de raspagem com diferentes pontas de ultrassom na rugosidade superficial, estabilidade de cor e acúmulo bacteriano em cerâmica de dissilicato de lítio após tratamento de profilaxia periodontal

Material e Métodos: O procedimento de raspagem foi realizado usando um aparelho de ultrassom (Satalec, Acteon, América do Norte) com ponta de aço inoxidável (US), ponta de titânio (UT) e ponta de plástico (UP), em amostras de dissilicato de lítio em forma de disco cimentadas em uma cavidade preparada na superfície vestibular de dentes bovinos recém-extraídos (10 amostras por grupo). As amostras foram armazenadas em solução de café em incubadora a 37°C por 12 dias, o que equivale a 1 ano de consumo de café. A rugosidade da superfície foi medida antes e após o procedimento de raspagem usando um perfilômetro e um microscópio de força atômica. Os parâmetros de cor foram medidos antes e depois dos procedimentos de raspagem e armazenagem no café usando VITA Easyshade Advance 4.0 de acordo com o sistema de ordem de cores CIE L*a*b* color order system. As amostras foram incubadas com suspensão de Streptococcus mutans (S. mutans) e depois incubadas em placa com 30 a 300 colônias típicas de S. mutans. Após a incubação, as placas com 30 a 300 colônias foram contadas em contador de colônias e obtidos os valores médios das unidades formadoras de colônias (UFC / mL). Resultados: A ponte de titânio mostrou valores estaticamente maiores de mudança na rugosidade da superfície ΔRa e contagem de bactérias do que a ponte de raspagem de plástico. A mudança de cor (ΔE) não foi estaticamente significativa entre os grupos. Os resultados mostraram uma correlação positiva (direta) estaticamente significativa entre rugosidade
INTRODUCTION

Ceramics are the material of choice for patients with high esthetic demands owing to their good physical and optical properties and to their ability to match natural dentition. The recent developments of ceramic systems and processing techniques enabled the treatment of teeth in both the anterior and posterior areas restoring form, function and esthetic excellence with metal-free restorations.

Each time esthetic procedures are considered, gingival health should also be evaluated. Initially, dental prophylaxis and periodontal treatment, including scaling and root planning, should be performed. Ultrasonic scaling have become the most widely used methods among dental surgeons and oral hygienists, due to the decreased time requirement and ease of application in comparison with hand instrumentation and various studies have confirmed that the two techniques yield similar results [1].

Surface roughness of the dental tissues is one of the most described alterations in the literature after periodontal instrumentation. The cumulative effect of minor substance removal per instrumentation performed over the years may lead to severe damage of the dental tissues and existing restorations over time which may increase surface roughness [2] and cause evident unesthetic color changes.

Unfortunately, there is limited information concerning the effects of periodontal instrumentation on lithium disilicate pressable ceramics. And hence the purpose of this study was to evaluate the effect of different scaling methods on surface roughness and color stability and bacterial accumulation of lithium disilicate ceramics.

MATERIALS AND METHODS

Forty bovine anterior teeth were collected. All soft tissue was removed until teeth were visually clean. The teeth were decapitated 2 mm below the cemento-enamel junction using tapered diamond stone with copious coolant to remove the root. The labial surfaces of the teeth were flattened using a cylinder diamond stone to obtain a flat area of 1x1 cm². The teeth were mounted in acrylic (Acrostone Dental Factory, Egypt) blocks using PVC tube of 1-inch inner diameter and 1.5 cm thickness. Then a cavity of 5 mm diameter and 0.5 mm depth was prepared and color change (p = 0.012) and also between surface roughness and bacterial count (p = 0.00). Conclusion: Within the limitations of this study, titanium scaling instruments cause irreversible surface alterations of lithium disilicate ceramics which was in direct correlation to the color changes and bacterial accumulation; therefore, dentists should proceed with caution when scaling lithium disilicate surfaces. The findings of the current study may indicate the need for instruments or equipment that can remove plaque and calculus without causing surface damage.

KEYWORDS
Surface properties; Color; Bacterial adhesion; Ultrasonics; Dental scaling; Ceramics.
on the flat surface of the bovine teeth using wheel diamond stone (Komet Dental, Gebr. Brasseler).

Forty disc shaped lithium disilicate (IPs e.max Press, Ivoclar Vivadent, Inc) samples of 0.5 mm thickness and 5 mm diameter were constructed according to the manufacturer's instructions. The fitting surfaces of the ceramic discs were etched using hydrofluoric acid (9.5 %) (BISCO, Inc, America) porcelain etchant followed by porcelain primer (Pre-Hydrolized Silane Primer) (BISCO, Inc, America) application according to the manufacturer's instructions. The cavities on the teeth surfaces were acid-etched using phosphoric acid (37 %) (Eco-Etch, Ivoclar Vivadent) followed by the bonding agent (TE-Econom Bond, Ivoclar Vivadent) according to the manufacturer's instructions. Finally, the discs were cemented into the cavities using Variolink N clear (Ivoclar Vivadent, Inc).

The samples were divided into four groups (10 samples per group); i) C: Control group (no scaling), ii) US: Ultrasonic stainless-steel tip (NSK), iii) UT: Ultrasonic titanium tip (Woodpecker), iv) UP: Ultrasonic plastic tip (NSK).

A specially designed scaling apparatus (Figure 1) was designed and fabricated to standardize the scaling procedure [3]. The samples were mounted onto one side of the double-pane balance and attached in place using screws. Scaling procedure was carried out using an ultrasonic scaler handpiece (Satalec, Acteon, North America) at intermediate power setting (level 5 of 14 grades). The ultrasonic scaling tips were angled 90° relative to the surface of sample. A constant force of 30 g was applied to the ultrasonic scalar tip by the vertical movement of a counterweighed balance. A standardized 5 mm horizontal movement and three consecutive cycles of 20 seconds each of the ultrasonic handpiece at a speed of 2 Hz was achieved and operated by the control box [3].

The surface roughness of samples was measured before and after the scaling procedure using the profilometer; Surface Roughness Tester TIME3202 (TR220) (Landmark Industrial Inc, USA), at cut off 0.25 mm, number of cuts 1 and range ± 40 μm. Measurements were made at three different regions (in the middle and sides) were evaluated in each sample to determine the surface roughness (Ra) values, and averaged to determine the mean values. Then the ceramic surface was analyzed using Atomic force microscopy using AutoProbe CP-Research (Thermomicroscope, Bruker Nano Inc., USA) operated in contact mode using nonconductive silicon nitride probe, at scan area of 25 μm, scan rate of 1 Hz and number of data points 256 x 256 m² was using proscan 1.8 software for controlling the scan parameters and IP 2.1 software for image analysis.

Staining procedure: Staining procedure was carried out using coffee solution (Nescafé Classic; Nestlé Egypt). The coffee solution was prepared according to the manufacturer's instructions by using 3.6 g of coffee and 300 mL of hot water. The solution was stirred for 10 minutes and passed through filter paper (Melitta; Melitta Haushaltsprodukte GmbH & Co Kg). In the interval between the 2 color measurements, all specimens were stored in coffee solution in an incubator (Model B 28, BINDER GmbH) at 37°C for 12 days, which is equivalent to 1 year of coffee consumption. [4,5] The solution was stirred every 12 ± 1 hours. After 12 days, the specimens were washed with tap water and dried with tissue paper.

The color parameters of each specimen were measured before and after the scaling
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and staining procedures using VITA Easyshade Advance 4.01 (VITA shade, VITA made, VITA) according to the CIE L*a*b* color order system.

Bacterial accumulation test: Initially, a standard suspension of S. mutans was prepared. As the S. mutans preferentially colonizes the pits and fissures of the occlusal surface of teeth, a swab was taken from an oral cavity of bad oral hygiene using sterile loop in laminar air flow, then it was spread on sheep blood agar and incubated for 24 hours at 37 °C in a CO2 chamber. After isolation, the cultured S. mutans was seeded onto a brain heart infusion (BHI) agar and incubated for 24 hours at 37 °C in a CO2 chamber. Then, the growth was suspended in sterile physiological solution [0.9% sodium chloride (NaCl)] to obtain a standard suspension containing 106 cells/ml. The number of cells in suspension was counted in a spectrophotometer. The parameters of optical density and wavelength used were, respectively, 0.620 and 398 nm. These parameters were previously established using a standard curve for CFU vs. absorbance.

Biofilm adhesion was performed in an aseptic environment. The broth used for adherence contains 20 g tripticase, 2 g NaCl, 3 g K2HPO4, 2 g KH2PO4, 1 g K2CO3, 120 mg MgSO4, 15 mg MnSO4, and 50 g C6H8O7 dissolved in 1000 mL of distilled water. The broth was sterilized by autoclaving it at 121 °C for 15 minutes [6]. The samples were placed in a sterile 24-well culture plate then 1.5 mL of broth and 0.1 mL of standardized S. mutans suspension were added to the samples. The plates were sealed and incubated at 37 °C for 24 hours in a CO2 chamber. Samples were then removed and washed twice with sterile physiological solution (0.9% NaCl) in order to remove loosely bound material. The samples were then placed in plates with 3 mL of sterile physiological solution (0.9% NaCl) and sonicated for 30 seconds to disperse the biofilms. The suspension obtained was diluted 100, 1000, and 10,000 times and aliquots of 0.1 mL were seeded in duplicate onto brain heart infusion (BHI) agar and incubated for 48 h at 37 °C in a CO2 chamber.

After incubation, the plates with 30 to 300 typical colonies of S. mutans were counted in a colony counter and mean values of colony forming units were obtained (CFU/mL).

RESULTS

Data was analyzed on an IBM® (IBM Corporation, NY, USA) personal computer, using Statistical Package for Special Science (SPSS)® (SPSS, Inc., an IBM Company) software computer program version 22. Data were described as mean ± standard deviation (SD) for quantitative (Numerical) variables. One-Way ANOVA followed by Tukey’s post-hoc test was used for comparison of quantitative variables among more than two independent groups. Dunnnett’s test was used to compare the control group with the other study groups. Correlation between continuous variables was performed using pearson’s correlation coefficient. The significance level was set at p ≤ 0.05.

The results showed that the mean values of ΔRa of the stainless-steel tip showed no statistically significant difference from that of the titanium and the plastic tips. While the titanium scaling tip showed a statistically significant higher mean values of ΔRa than the plastic scaling tip (Table I). AFM showed that scaling using ultrasonic stainless-steel tip clearly resulted in scraping of the ceramic surfaces and loss of their original texture, leading to increased surface roughness, they showed many shallow irregularities. Ultrasonic titanium tip showed more aggressive and deeper scratches than the ultrasonic stainless-steel tip. Scaling using plastic tip did not appear to markedly affect the ceramic surfaces, it showed the least surface alterations (Figure 2-5).

There was no statistically significant difference among mean values of ΔE of the three tip materials and the control group. However,
for the bacterial accumulation, the plastic groups showed the statistically significant least mean values of bacterial count. The stainless-steel and the titanium groups showed the statistically significant highest mean values of the bacterial count. All groups were statistically significant different from the control group.

**DISCUSSION**

Based on their strength, longevity, conservative nature, biocompatibility and esthetics, veneers have been considered one of the most viable treatment modalities since their introduction in 1983 [7]. Esthetic veneers in ceramic materials demonstrate excellent clinical performance and as materials and techniques have evolved, veneers have become one of the most predictable, most esthetic and least invasive modalities of treatment. Studies that followed ceramic veneers for up to 15 years reported total failures of only 7%, equaling approximately a 1% chance of failure per year [8,9].

However, the challenge with laminate veneers is to achieve ideal esthetics including color matching and subsequent color stability [10]. Although ceramics are considered to be color stable, discoloration of esthetic restorations may occur due to intrinsic or extrinsic factors. Intrinsic factors include changes within the material itself, while extrinsic factors involve adsorption or absorption of stains from the oral cavity. The smoothness of the surface of the restoration is one of the factors affecting extrinsic staining [11].

Periodontal prophylactic treatment is performed using supra- and subgingival scaling procedures to disrupt and thoroughly remove biofilm, calculus deposits, periodontal pathogens and deposits. Instrumentation options include hand scalers and ultrasonic scalers. Ultrasonic
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Scalers have a similar or greater efficiency than hand instruments in the removal of plaque and calculus on the surfaces of dental materials [12,13]. Accordingly, ultrasonic scalers have become an established tool for the removal of dental plaque and calculus. The cleaning procedures, especially using ultrasonic scaler, can increase surface roughness of dental restorations [14,15].

Recently, different types of nonmetallic instruments and ultrasonic tips such as rubber cups, plastic curettes, titanium curettes and air-power abrasive systems have been introduced as a substitute for implant maintenance to avoid alterations of the implant surfaces and they have been recommended for use in removing plaque from implants rather than metallic instruments [16-18]. These novel instruments may also be used with all-ceramic restorations in order to minimize the possible surface alterations that could result due to the use of conventional periodontal prophylactic instruments.

Accordingly, in the current study in order to determine which instrument is most appropriate for use on lithium disilicate surfaces; the effect of different scaling methods and instruments; (stainless-steel, titanium and plastic) on surface roughness and color stability of lithium disilicate ceramic was investigated.

The results of ΔRa readings indicated that scaling using ultrasonic titanium tip showed the highest mean value of ΔRa readings which was not statistically significantly different from the stainless-steel tip. However, it was statistically significantly different from the plastic tip. This may be attributed to the increased hardness of the titanium instruments (~751.9 MPa) compared to the stainless-steel instruments (~591.6 MPa)[19]. Ito A et al. (1991) reported that the Vickers hardness of titanium nitride was 1170 and Nukata K et al. (1995) reported it to be 1370 [20]. The results of the profilometry were in agreement with the observations made from the atomic force microscopy which showed that lithium disilicate surfaces treated by stainless-steel and titanium tips revealed multiple large scratches while the scaling method based on a plastic instruments caused the least alterations.

Concerning color stability, the color change of the control (ΔE = 2.638) and the plastic (ΔE = 2.765) groups recorded clinically acceptable value (ΔE > 3.5) while the titanium (ΔE = 4.431) and the stainless-steel (ΔE = 4.451) groups recorded clinically unacceptable values. These results were in accordance with the records of surface roughness indicating direct positive correlation between the surface roughness and the ΔE. This may be attributed to the removal of the surface glaze and the inferior ability of rough surface to reflect light.

In 1981, Obregon et al. [21] stated that surface gloss texture and roughness can alter the color perception of porcelain restorations. Also, Kim et al. (2003)[22] found that color differences due to the surface conditions were large enough to be visually perceivable when measured with spectrophotometer. These results are in accordance with Motro PF et al. (2012)[10] who demonstrated that surface roughness affected the stainability of the IPS e.max Ceram (Ivoclar Vivadent) feldspathic ceramic and Akar GC et al. (2014)[23] who reported statistically significant differences in the surface-finishing protocols applied for the Lithium disilicate ceramic systems and the Δ Ra, Δ E and translucency parameters. The smoothest surfaces and the lowest Δ E values were seen with the autoglazed method and the highest values were obtained with the polishing with adjustment kit plus diamond polishing paste method. Also, Gawriołek M et al. (2012)[11] reported that unpolished composite and ceramic specimens with a more developed surface have lower color stability.

Also, there was a direct positive correlation between bacterial accumulation and surface roughness. These observations may be attributable to the easier initial bacterial adhesion to and its more difficult removal from rough surfaces [24-26]. All groups showed statistically significant difference from the
The surface roughness of lithium disilicate ceramics can be increased due to prophylactic periodontal treatment. Plastic based periodontal instruments have the least statistically significant effect on lithium disilicate ceramic surface roughness. There is a positive correlation between surface roughness and color stability of lithium disilicate ceramics.

REFERENCES

1. Mourourzi P, Kouklousidou EA, Vassiliades L, Hovanlioglou-Antoniades M. Effects of sonic scaling on the roughness of dental restorative materials. J Oral Sci 2005;47:107-14.
2. Checkettts MR, Turkyilmaz L, Asar NV. An investigation of the effect of scaling-induced surface roughness on bacterial adhesion in common fixed dental restorative materials. J Prosthodont Dent 2014;112(5):565-70.
3. Flausino JS, Soares PB, Carvalho VC, Magalhães D, da Silva WM, Costa HL, et al. Biofilm formation on different materials for orthodontic treatment: analysis of surface characteristics. J Mater Sci 2014;49:6820-9.
4. Guler AU, Kurt S, Kulkun T. Effects of various finishing procedures on the surface characteristics of provisional restorative materials. J Prosthodont Dent 2005;95:465-8.
5. Guler AU, Yilmaz F, Kulkun T, Guler E, Kurt S. Effects of different dental instruments on the surface characteristics of provisional restorative materials. J Prosthodont Dent 2005;95:24-30.
6. Gybbons RJ, Nygaard M. Synthesis of insoluble dextran and its significance in the formation of gelatonous deposits by plaque-forming streptococci. Arch Oral Biol 1968;13:12-49.
7. McLaren EA, LeSage B. Feldspathic veneers: what are their indications? Compendium Contin Educ Dent. 2011;32(3):44-9.
8. Newsome P, Owen S. Ceramic veneers in general practice: part four; clinical procedures. Part 1: Treatment planning. International Dentistry SA 2008;10:1-6.
9. Friedeman MJ. A 15-year review of porcelain veneer failure—a clinician’s observations. Compendium Contin Educ Dent. 1998;19(6):625-30.
10. Rosenstitt M, Sawajranow A, Behr M, Kolbeck C, Preis V. Effect of tooth brush abrasion and thermo-mechanical loading on direct and indirect veneer restorations. Clin Oral Invest 2015;15:53-60.
11. Singh K, Suzawa S, Agnihotri Y, Sahoo S, Kumar P. Color stability of aesthetic restorative materials after exposure to commonly consumed beverages: a systematic review of literature. Eur J Prosthodont Restor Dent 2014;20(1):22.
12. Chen YL, Chang HH, Chang YC, Lin CP. Application and development of ultrasonics in dentistry. J Formos Med Assoc. 2013 Nov;112(11):659-65.
13. Petersilka GI, Fleming TF. Periodontal debridement with sonic and ultrasonic scalers. Periodontal Practice Today. 2004;14(1):353-62.
14. Ausschill TM, Amelieker NB, Brex M, Reich E, Sculean A, Nettusch L. The effect of dental restorative materials on dental biofilm. Eur J Oral Sci 2002;110:48-53.
15. Popat KC, Eltgroth M, Latempra T, Grimes CA, DeSai TA. Decreased Staphylococcus epidermis adhesion and increased osteoblast functionality on antibiotic-loaded titania nanotubes. Biomaterials 2007;28(32):4880-8.
16. Hallmon WW, Waldrop TC, Wade WH. A comparative study of the effects of metallic, nonmetallic, and ceramic instrumentation on titanium abutment surfaces. Int J Oral Maxillofac Implants 1996;11:196-100.
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17. Mengel R, Meer C, Flores-de-Jacoby L. The treatment of uncoated and titanium nitride-coated abutments with different instruments. Int J Oral Maxillofac Implants 2004;19:232-8.

18. Seol HW, Heo SJ, Koak JY, Kim SK, Baek SH, Lee SY. Surface alterations of several dental materials by a novel ultrasonic scaler tip. J Oral Maxillofac Implants 2012;27:801-10.

19. Steele JG, McCabe JF, Barnes IE. Properties of a titanium nitride coating for dental instruments. J Dent. 1991;19(4):226-9.

20. Tamura Y, Yajima T, Watanabe F, Un M, Kawasaki T. Mechanical properties of surface nitrided titanium for abrasion resistant implant materials. Materials Transactions 2002; 43(12):3043-51.

21. Obregon A, Goodkind RJ, Schwabacher WB. Effects of opaque and porcelain surface texture on the color of ceramic metal restorations. J Prosthet Dent 1981;46:330-40.

22. Kim LJ, Lee YK, Lim BS, Kim CW. Effect of surface topography on the color of dental porcelain. J Mater Sci Mater Med 2003;14:405-9.

23. Chen YL, Chang HH, Chiang YC, Lin CP. Application and development of ultrasonic in dentistry. J Formos Med Assoc 2013;112 (11): 659-65.

24. Baek SH, Shon WJ, Baek KS, Kurn KY, Lee WC, Park YS. Evaluation of the safety and efficiency of novel metallic ultrasonic scaler tip on titanium surfaces. Clin Oral Implants Res 2012;23(10):1269-74.

25. Meier R, Hauser-Gerspach L, Lüthy H, Meyer J. Adhesion of oral streptococci to all-ceramics dental restorative materials in vitro. J Mater Sci Mater Med 2006;17:329-41.

26. Teughels W, van Assche N, Sleipen L, Quirynen M. Effect of material characteristics and/or surface topography on biofilm development. Clin Oral Implant Res 2006;07:68-81.

27. Quirynen M, Bollen CM, Papaioannou W, Van Eldere J, van Steenberge D. The influence of titanium abutment surface roughness on plaque accumulation and gingivitis: short-term observations. Int J Oral Maxillofac Implants 1996;11:669-76.

28. Aykent F, Yonem I, Ozyesil AG, Guna SK, Arunduk MC, Oksan S. Effect of different finishing techniques for restorative materials on surface roughness and bacterial adhesion. J Prosthet Dent 2010;103:221-7.

29. Carlén A, Nikdel K, Wennerberg A, Holmberg K, Öllson J. Surface characteristics and in vitro biofilm formation on glass ionomer and composite resin. Biomaterials 2001;22:481-7.

30. Hahnel S, Rosentritt M, Handel G, bürgers R. Surface characterization of dental ceramics and initial streptococcal adhesion in vitro. Dent Mater 2009; 25(8):969-75.

31. Kawai K, Urano M, Ebita S. Effect of surface roughness of porcelain on adhesion of bacteria and their synthesizing glucans. J Prosthet Dent 2000;83:664-7.

32. Buergers R, Rosentritt M, Handel G. Bacterial adhesion of Streptococcus mutans to provisional fixed prosthodontic material. J Prosthet Dent 2007;98:461-9.

33. Milleding P, Gerdes S, Holmberg K, Karlsson S. Surface energy of non-corroded and corroded dental ceramic materials before and after contact with salivary proteins. Eur J Oral Sci 1999;107:364-92.

34. Quirynen M, Bollen CML. The influence of surface roughness and surface-free energy on supra- and subgingival plaque formation in man. A review of the literature. J Clin Periodontol 1995;22:1-14.

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