The safety of isotretinoin treatment in patients with bone fractures

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Abstract
Isotretinoin is widely applicable in dermatology, although it may develop severe side effects in the skeletal system. An intention of this review was to establish the safety of oral isotretinoin in patients with bone fractures. Both MEDLINE/Pubmed and SCOPUS databases were searched to investigate the influence of isotretinoin on the skeletal system. The drug shows a strong osteoporotic activity in rats whereas this effect is milder in humans. Biochemical markers of bone turnover remain unchanged except for serum calcium in patients receiving a high dose of isotretinoin. An excessive intake of vitamin A may impair functioning of vitamin D especially in people with a vitamin D deficiency, therefore a similar side effect may also occur in patients on isotretinoin treatment. We suggest reducing the use of isotretinoin after bone injury or continuing the treatment at low dosing with a concomitant correction of vitamin D and calcium status.

Key words: isotretinoin, retinoids, bone fracture, bone healing.

Introduction
Isotretinoin has been the most effective drug for severe acne since its approval by the US Food and Drug Administration in May 1982. It belongs to retinoids which are derivatives of vitamin A. The drug targets all four major factors involved in the pathogenesis of acne, such as sebaceous gland hyperplasia with hyperseborrhea, follicular hyperkeratinization, hypercolonization of the pilosebaceous unit with Propionibacterium acnes and inflammation [1, 2].

There are at least several adverse effects which can preclude the use of isotretinoin and teratogenicity is the most important systemic toxicity. A fetal exposure to isotretinoin can contribute to severe skeletal defects in the baby [3]. A postnatal exposure of children to the drug may develop a premature closure of the lower-extremity growth plates with a later reduction in adult stature [4]. There is also a risk of diffuse skeletal hyperostosis in patients treated with isotretinoin [5]. On the other hand, isotretinoin was supposed to exert osteoporotic activity in bones [6, 7]. Overall, bone turnover is known to be altered by both vitamin A and its derivatives, which affect functioning of osteoclasts and osteoblasts [8, 9].

Undoubtedly, isotretinoin may affect different healing processes. Recently, a detrimental influence of isotretinoin on tissue regeneration after surgical procedures was shown to be mostly overestimated [10–12]. The management of patients on isotretinoin who experienced bone fracture remains however still unclear.

Aim
The aim of this review was to establish the safety of oral isotretinoin in patients with bone fracture.

Material and methods
Both Medline/Pubmed and SCOPUS databases were searched to investigate the influence of isotretinoin on the skeletal system. Medical Subject Headings were applied in various combinations: isotretinoin, retinoids, all-trans retinoic acid, bone and bones, bone fractures, fracture healing, and bone density. Initially 833 records were found, but only 62 items were included in the final review. The authors analyzed only English papers published since 1995, but earlier manuscripts were cited when they had historic significance. Data were extracted with a focus on bone mineral density and metabolism, vitamin D.
and calcium status, bone repair, activity of osteoblasts and osteoclasts. Animal studies other than rodent ones were not analyzed. There were 11 human and 2 rat studies included in the major analysis of bone mineral density (BMD) and biochemical changes. Bone mineral density was analyzed only for the hip and the lumbar spine, but here rat studies were excluded due to distinct techniques of BMD measurement and short treatment protocol. Biochemical changes were analyzed in both human and rat studies. Three human and two rat case reports were extracted to present the only papers available in the English literature showing response of bones after an injury.

Results and discussion

Osteoporotic effect of isotretinoin

A high dietary intake of vitamin A was demonstrated to develop bone thinning in rats [8]. An excessive dietary intake of vitamin A is known to increase a risk of osteoporotic bone fractures in humans [13–17]. Presumptively, isotretinoin exerts a negative impact on bone health (Table 1) and one should also expect a decrease of BMD in patients receiving oral isotretinoin. However, a significant change of BMD was seen neither at the lumbar spine nor at the hip in a cumulative number of 67 patients from three clinical studies [18–20]. The only study which showed a significant decrease of BMD at the hip on isotretinoin treatment was presented by Saadi et al., who however enrolled ten Emirati women only [21]. Another slight decrease in BMD at the hip was observed by Leachman et al. [22] and DiGiovanna et al. [6] but only when the density was measured at the Ward’s triangle [22]. This area is currently not advisable for interpretation of BMD due to a large influence of positional changes [23], however one must expect an increase in femoral BMD rather than its anyway slight decrease in adolescents. A greater predisposition to a bone loss was seen in those patients with the lowest values of BMD before starting isotretinoin. Pretreatment evaluation of bone density seems to be therefore reasonable management before starting isotretinoin to identify those patients at greater risk of drug-induced osteoporosis [22]. Undoubtedly, this risk is much greater in females due to a concomitant use of oral contraceptives, which are known to have an unfavorable impact on the skeletal system [24, 25].

Four studies which analyzed a change of BMD at the lumbar spine in patients receiving oral isotretinoin showed its slight increase (1.1–3.0%) in contrast to the reported decrease at long bones [6, 22, 26, 27]. This change remained however in a normal range for young adoles-

| Authors        | Study group | Control group | Treatment protocol, birth control in females, method of BMD analysis (device) | Mean change of BMD | Other BMD measurements |
|---------------|-------------|---------------|-----------------------------------------------------------------------------|-------------------|------------------------|
| Tekin et al. [20] | 15 M, 21 F | 16 M, 20 F     | – ISO until 120 mg/kg of the cumulative dose                                  | NS                | NS                     |
| Kindmark et al. [18] | 9 M, 2 F  | N/A           | – ISO 0.71–0.88 mg/kg daily for 6 months                                     | –                 | NS                     |
| Margolis et al. [19] | 6 M, 14 F | N/A           | – ISO until 122 mg/kg of the cumulative dose                                  | NS                | NS                     |
| Leachman et al. [22] | 18 M | 14 M          | – ISO 1 mg/kg daily for 6 months – 4 persons showing the greatest decrease in BMD at the Ward’s triangle had lowest baseline densities at the spine | NS*               | 1.1% ↑ (p = 0.02) **4.4% ↓ (p = 0.03) at the Ward’s triangle |
| Saadi et al. [21] | 10 F       | N/A           | – ISO until 80.8 ±28.8 mg/kg – Two methods of contraception required         | −5.3 ±1.9% ↓ (p = 0.002) | NS                     |
| DiGiovanna et al. [6] | 136 M, 81 F | N/A | – ISO 1 mg/kg daily until 8821 ±2328 mg of the cumulative dose | NS**              | 1.403 ±2.479% ↑ (p < 0.00001) **1.423 ±4.212% ↓ (p < 0.00001) at the Ward’s triangle |
| Hoover et al. [26] | 289 M, 69 F | N/A | – ISO 0.5 mg/kg (first 4 weeks), 1 mg/kg (16 weeks) – Oral contraceptives allowed | –     | 1.7–2.0% ↑ (p < 0.0001) |
| Kocijancic [27] | 15 M       | N/A           | – ISO average total dose 0.4 mg/kg daily for 6 months                       | –                 | 3.0% ↑                |
| Erdogan et al. [28] | 14 M, 7 F | 10 M, 11 F   | – ISO until 120 mg/kg of the cumulative dose                                | NS                | NS                     |

ISO – isotretinoin, M – males, F – females, NS – non-significant, BMD – bone mineral density, N/A – not available.
Biochemical changes following exposure to isotretinoin (7 human studies and 2 rat study)

Table 2. Biochemical changes following exposure to isotretinoin (7 human studies and 2 rat study)

| Authors          | Vitamin D metabolites | Markers of bone formation | Markers of bone resorption | Other markers of bone turnover |
|------------------|-----------------------|---------------------------|--------------------------|-----------------------------|
|                  | 25(OH)D3 | 1,25(OH)2D3 | OC | PICP | BAP | Urine DPD/serum PYD | Urine OHP | TRACP | Urine NTx | PTH | Serum calcium | Serum phosphate |
| Tekin et al. [20] | –        | –         | ↑ (NS) | –         | –         | ↑ (NS) | –         | –         | ↑ (NS) | ↓ (NS) | ↓ (NS) |
| Leachman et al. [22] | ↑ (p = 0.02) | ↑ (NS) | –         | –         | –         | –         | –         | –         | ↑ (NS) | ↓ (NS) | ↑ (NS) |
| Kindmark et al. [18] | –         | –         | –         | ↓*         | –         | ↓ (NS) | –         | –         | ↑*         | ↓*         | –         |
| Margolis et al. [19] | ↑ (NS) | ↓ (p = 0.02) | ↑ (NS) | –         | –         | –         | –         | ↑ (NS) | –         | ↑ (NS) | ↓ (NS) |
| Saadi et al. [21] | NS | –         | –         | NS | –         | –         | ↑ (p < 0.001) | NS | ↓ (p = 0.04) | NS |
| Ertugrul et al. [31] | ↓ (p < 0.0001) | ↑ (p < 0.005) | –         | –         | ↑ (p < 0.0001) | –         | –         | ↑ (p < 0.005) | ↓ (p < 0.05) | ↑ (NS) |
| Trifirò and Norbiato [32] | NS | NS | NS | NS | –         | –         | –         | ↓ (p < 0.05) | NS | –         | – |
| Hotchkiss et al. [30] rat study | – | – | – | – | ↑** (p < 0.05) | NS change | – | – | – | ↑*** (p < 0.05) | – |
| Bergoli et al. [42] rat study | – | – | – | – | – | – | – | ↓**** (p < 0.05) | – | – | – |

NS = non-significant, OC = osteocalcin, PICP = procollagen type I carboxyl-terminal propeptide, BAP = bone alkaline phosphatase, PYD = pyridinoline, DPD = deoxypyridinoline, OHP = hydroxyproline, TRACP = tartrate-resistant acid phosphatase, PTH = parathyroid hormone, NTx = N-terminal telopeptide of type I collagen.

*Transient change only within first 5 days, then normalized. **Significant for all-trans retinoic acid (ATRA), but not for isotretinoin. ***Significant only for a high dose of isotretinoin, but not for ATRA; ****Significant at days 21, 28 and 90 of isotretinoin administration.

Adenoids who typically manifest an increase in bone density 0.5–7.0% yearly [27]. Undoubtedly, one must look for artifacts such as confounding extraosseous calcification when assessing BMD at the lumbar spine in the anteroposterior view. Secondary adverse events such as longitudinal ligament calcification can lead to a paradoxical increase in BMD, requiring a correction of measurements at this area [18, 23, 27, 28].

Although human studies revealed only minimal influence of isotretinoin on bone density, animal studies demonstrated a more severe detrimental influence of retinoids on bone health. Recently, rats receiving intragastrically tazarotene (0.075 mg/kg) in sesame oil (4 ml/kg) daily were demonstrated to have worse bone mechanical properties than control animals. Both mean Young’s modulus and the stiffness were significantly impaired at the femur in tazarotene-treated rats suggesting osteoporotic nature of the drug [29]. Previously, Hotchkiss et al. also demonstrated an unfavorable effect of retinoids in long bones. Both isotretinoin given in a daily dose of 7.5 mg/kg b.w. or 30 mg/kg b.w and ATRA in a daily dose of 10 or 15 mg/kg b.w did reduce significantly bone mineral content and density at the femur. Femoral shaft diameter and bone marrow cavity were markedly smaller than in control subjects, although this detrimental effect was not seen in rats receiving low doses of isotretinoin. Histomorphometry showed a greater loss of both cortical and trabecular bone in animals treated with ATRA than in ones receiving isotretinoin. An inhibition of cortical bone expansion by retinoids is a unique effect since most of other drugs affect primarily remodeling of the trabecular bone. A progressively poorer quality of bones reduces their resistance to bending therefore predisposing to spontaneous long bone fractures that were reported in rats treated with ATRA [30].

Retinoids and bone metabolism

An analysis of biochemical markers of bone turnover after an exposure to isotretinoin showed mostly insignificant or only transient disturbances (Table 2). Two human studies showed a significant increase in serum parathyroid hormone, although in one of them (Kindmark et al.) this change was only transient [18, 31]. Bone alkaline phosphatase, a marker of bone formation was significantly increased in only one (Ertugrul et al.) of four human studies [20, 21, 31, 32] and in one rat study (Hotchkiss et al.) [30]. Other markers of bone formation,
ostecalcin and procollagen type I carboxyl-terminal propeptide were rather transiently decreased within the first 5 days of therapy with isotretinoin in a study of Kindmark et al. [18].

Enhanced bone resorption was manifested by a significant increase in serum tartrate-resistant acid phosphatase in the study of Saadi et al. [21], whereas other markers (serum pyridinoline, urine deoxypyridinoline, urine hydroxyproline) were elevated insignificantly [19, 20, 22, 29]. Surprisingly, Trifiro and Norbiato found even a decrease in urine N-terminal telopeptide of type I collagen (NTx) level on isotretinoin treatment. NTx level correlates however with a degradation of type I collagen distributed both in the skeletal system and in the skin, thus different burdens of the skin may destroy collagen bundles with a consequent release of type I collagen degradation products. Treatment with isotretinoin probably limits this catabolism therefore decreasing the urine NTx level [32, 33].

Previously, hypercalcemia was postulated to have a dose-limiting toxic effect in patients receiving oral isotretinoin [34, 35] and ATRA [36–38]. This complication was also observed after an excessive use of over-the-counter supplements or enteral feeding formula containing vitamin A, especially in case of concomitant renal disease [39–41]. Only one (Hotchkiss et al.) from those reviewed studies showed a significant elevation of serum calcium, however exclusively in rats receiving a high-dose of isotretinoin [18, 20–22, 30, 31, 42]. Duncan et al. proposed controlling periodically serum calcium in those patients receiving a daily dose of isotretinoin greater than 1.3 mg/kg [43]. None of reviewed human studies showed any significant change of serum phosphate [20–22, 31].

Retinoids and vitamin D

A strong detrimental influence of vitamin A on bone mineralization in a low vitamin D supply was previously observed in rats [44]. An attenuation of vitamin D-dependent oral calcium absorption by an intense vitamin A supplementation was reported in both rats [44] and humans [45]. A Korean cross-sectional study showed however that an excessive intake of vitamin A had a negative impact on bone health only in a population with a vitamin D deficiency (< 50 nmol/l of 25(OH)D) [46]. One must expect that optimal BMD may depend on the favorable ratio of vitamin A intake to vitamin D repletion or optimal ratio of serum retinol to 25-hydroxyvitamin D [47, 48].

The vitamin D deficiency was also reported to be a condition predisposing to increased bone resorption and decreased BMD in patients receiving isotretinoin [21]. There is a need for vitamin D supplementation in children with ichthyotic disorders receiving oral acitretin, although a defective epidermal synthesis of vitamin D should be theoretically improved by oral retinoids [49]. A measurement of serum 25-hydroxyvitamin D and correction of vitamin D deficiency appear to be advisable before starting isotretinoin treatment for acne [47]. The National Osteoporosis Foundation recommends 400–800 international units (IU) of vitamin D daily for most individuals aged under 50 and a need of 800–1,000 IU daily in most adults aged 50 and older. A supplementation of vitamin D at intakes over 4000 IU daily was previously shown to be safe management which effectively raises 25(OH)D to high-normal concentrations in most of adults [50].

Retinoids and osteoblasts

Normal osteoblast differentiation requires an up-regulation of Cyp26b1, the major enzyme responsible for degradation of retinoids [8]. An exposure to ATRA (pan-RAR agonist) delays both differentiation and mineralization of osteoprogenitor cells into mature ones whereas a blockage of RAR receptors accelerates differentiation of early osteoprogenitors. A cessation of ATRA treatment is required to achieve a full expression of osteoblastic genes in osteogenic conditions. However, this inhibitory effect of ATRA appears to be long lasting and even after stopping a treatment, osteoprogenitor cells cannot retain a full differentiation capacity [51].

Retinoids downregulate expression of Phex, a protein which is necessary for mineralization of osteoblasts. A decrease in proteins Runx2 and Sp7 (Osterix) impairs mineralization and differentiation of osteoblasts. Both human osteoblasts and a murine preosteoblastic cell line have drastically reduced expression of alkaline phosphatase and osteocalcin, markers of osteoblast maturation, when treated with retinoids [8, 18].

Kneissel et al. showed that osteoblast performance following an exposure to retinoids depends on the cellular system. Retinoids manifest neither stimulatory nor inhibitory effect on pre-osteoblastic cell line MC3T3-E1, but they easily inhibit mouse primary calvarial osteoblasts [52]. Surprisingly, de Oliveira et al. demonstrated distinct observations. Isotretinoin at a daily dose of 7.5 mg/kg (an equivalent to the standard dose administered to humans in the treatment of severe acne – 1 mg/kg/day) did accelerate bone repair in rat calvaria, although this increase was not statistically significant [53].

Retinoids and osteoclasts

Retinoids were found to exert an inhibitory effect on osteoclastogenesis via RAR-gamma signaling [9]. There is a repression of RANKL stimulation with an ongoing decrease in c-Fos signaling pathway, therefore preventing from ongoing activation of NFAT [54]. NFAT promotes normal differentiation of osteoclasts from their precursor cells whereas an activation of RAR receptors delays this process [9]. The inhibitory effect starts already from early hematopoietic precursor cells located in bone marrow but retinoids can prevent from differentiation of those cells into mature osteoclasts. This effect occurs at a low concentration of retinoids when they do not even
Skeletal effects of retinoids appear to depend on bone compartment. There is enhanced subperiosteal bone resorption in rats treated with retinoids, but this activity is reduced at the proximity to the bone marrow [52]. Presumptively, this increased bone resorption along the subperiosteal surface may explain bone loss under a treatment with retinoids [30]. Bone formation is decreased at the subperiosteal envelope, but it remains unaffected in the trabecular bone compartment. As a consequence, there is thinning of long bones due to the loss of cortical bone while bone length remains unaffected [52]. A similar compartment-dependent effect is also observed in rats with an excessive oral intake of vitamin A, which develops a significant reduction in microvessels at the endosteal site therefore leading to bone marrow hypoxia. This condition activates osteoblasts at the endosteal/marrow compartment, which increases endosteal mineralization. An inverse tendency shows however osteoclasts which disappear at the endosteal compartment whereas their periosteal number starts to increase. Although hypervitaminotic A bones get thinner due to the increased periosteal osteoclastic activity, their stiffness is preserved since there is increased endosteal mineral deposition [55].

Retinoids and bone repair

There is a paucity of papers which report bone healing process after surgical interventions or injuries on isotretinoin treatment (Table 3). The maxillofacial area seems to respond with enhancement of new bone formation after exposure to isotretinoin. Soon after marketing of isotretinoin, Novick et al. reported bilateral nasal bone osteophytosis in a healthy woman who had undergone uneventful rhinoplasty 12 years earlier [56]. Much later, Allen and Rhee observed nasal tip complications (nasal tip asymmetry, alar collapse, prominence of composite graft cartilage) which developed within 6 months on isotretinoin treatment in three patients who had undergone rhinoplasty 2 years before [57]. Presumptively, this could be a superficial manifestation of diffuse skeletal hyperostosis or optionally, some periosteal tags (having osteogenic capacity) were stimulated by isotretinoin to undergo ossification [56]. Bergoli et al. observed also a stimulation of alveolar repair after tooth extraction in rats receiving isotretinoin. A new bone formation was greater and sockets were filled with compact bone earlier than control animals [42].

Human studies did not show any significant influence of systemic isotretinoin on healing after tooth extraction. Sharma et al. observed postoperative alveolar osteitis only in three of 26 subjects treated with isotretinoin who underwent surgical extraction of wisdom teeth but without long-term complications [58]. Molecular analysis showed however decreased activity of retinaldehyde dehydrogenase-2 (enzyme which produces endogenous retinoids in body tissue) in adult molars [59]. An inhibition of retinoic acid receptor γ function was found to promote endochondral bone formation. Mice deprived of RAR-γ-signaling manifest a larger volume of cartilaginous tissue than wild type mice in a tibial bone defect model. An appearance of cartilaginous tissue is followed by a replacement with mineralized materials much higher than in control mice [60].

Conclusions

Isotretinoin was demonstrated to exert osteoporotic activity only in rat studies whereas this influence was not

**Table 3. Studies reporting postsurgical skeletal effects under isotretinoin treatment**

| Authors            | Type of surgery | Subjects | Control subjects | Time to isotretinoin treatment operation | Skeletal effect                                                                 |
|--------------------|-----------------|----------|------------------|-----------------------------------------|--------------------------------------------------------------------------------|
| Novick et al. [56] | Rhinoplasty     | 1 F      | N/A              | 12 years after the procedure            | Nasal bone osteophytes noticeable after 5 weeks of ISO treatment               |
| Allen and Rhee [57]| Septorhinoplasty| 2 F, 1 M | N/A              | 2 years after (1st case), 7 months after (2nd case), 1 year after (3rd case) | Nasal tip asymmetry, alar collapse, prominence of composite graft cartilage, nasal tip bossae |
| Sharma et al. [58] | Third molar surgery | 26 | N/A | Current therapy with ISO or within 3 months of its discontinuation | Dry socket only in 3 patients but without long-term complications |
| de Oliveira et al. [53] rat study | 2-mm cavity drilled in rat calvaria | 18 | 15 | 18 experimental rats receiving ISO daily for 30 days prior to the surgical procedure | Acceleration of new bone formation in rat calvaria by ISO |
| Bergoli et al. [42] rat study | Tooth extraction in rats | 20 | 12 | 20 experimental rats receiving daily isotretinoin for 30 days prior to the surgical procedure | Acceleration of alveolar repair by ISO |

ISO – isotretinoin, M – males, F – females, N/A – not available.
striking in humans. Undoubtedly, daily doses on a body weight in rats were much higher than the ones applicable for human patients. An intention was to generate quickly a concentration of the drug which would be achieved following an oral exposure to 0.5–1 mg/kg daily of isotretinoin for dermal conditions. Even here, an equivalent of a human daily dose of 1 mg/kg for isotretinoin did not develop marked changes in rat bones.

A change of bone turnover markers appears to be nothing but transient following an exposure to retinoids. The only mineral component which should be controlled is serum calcium in those patients receiving a daily dose of isotretinoin greater than 1.3 mg/kg. Undoubtedly, a correction of vitamin D deficiency appears to be advisable management before starting isotretinoin to maintain a concentration of 25(OH)D$_3$ on the moderate level of 50–75 nmol/L.

Several experimental studies showed however an unfavorable influence of retinoids on osteoblasts and osteoclasts functioning. An upregulation of enzymes responsible for degradation of retinoid in bone tissue was evidenced to be necessary to achieve a full mineralization of osteoblasts. Retinoids were shown to be also negative regulators of osteoclastogenesis and this effect starts already from early hematopoietic precursors. Overall, those findings confirm unfavorable influence of retinoids on bones since preserved activity of both cellular components is required to restore the original bone structure post fracture [61, 62].

Most of reviewed studies allow for only indirect conclusions regarding safety of isotretinoin treatment in patients who experienced bone fractures. Reports showing a direct influence of the drug on outcomes of surgical procedures in the skeletal system or bone healing process. The available ones show a facilitation of bone healing at the skull and hip fractures among postmenopausal women. JAMA 2002; 334: 1349-58.

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