Chromosome band 11q23 deletion predicts poor prognosis in bone marrow metastatic neuroblastoma patients without MYCN amplification

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Abstract

Background: Interphase fluorescence in situ hybridization (FISH) of bone marrow cells has been confirmed to be a direct and valid method to assess the v-myc avian myelocytomatosis viral oncogene neuroblastoma derived homolog (MYCN) amplification in patients with bone marrow metastatic neuroblastoma. MYCN amplification alone, however, is insufficient for pretreatment risk stratification. Chromosome band 11q23 deletion has recently been included in the risk stratification of neuroblastoma. In the present study, we aimed to evaluate the biological characteristics and prognostic impact of 11q23 deletion and MYCN amplification in patients with bone marrow metastatic neuroblastoma.

Methods: We analyzed the MYCN and 11q23 statuses of 101 patients with bone marrow metastatic neuroblastoma using interphase FISH of bone marrow cells. We specifically compared the biological characteristics and prognostic impact of both aberrations.

Results: MYCN amplification and 11q23 deletion were seen in 12 (11.9%) and 40 (39.6%) patients. The two markers were mutually exclusive. MYCN amplification occurred mainly in patients with high lactate dehydrogenase (LDH) and high neuron-specific enolase (NSE) levels (both \( P < 0.001 \)), and MYCN-amplified patients had more events (tumor relapse, progression, or death) than MYCN-normal patients (\( P = 0.004 \)). 11q23 deletion was associated only with age (\( P = 0.001 \)). Patients with MYCN amplification had poorer outcomes than those with normal MYCN (3-year event-free survival [EFS] rate: 8.3 ± 8.0% vs. 43.8 ± 8.5%, \( P = 0.001 \); 3-year overall survival [OS] rate: 10.4 ± 7.9% vs. 63.5% ± 5.7%, \( P < 0.001 \)). 11q23 deletion reflected a poor prognosis only for patients with normal MYCN (3-year EFS rate: 34.3 ± 9.5% vs. 53.4 ± 10.3%, \( P = 0.037 \); 3-year OS rate: 42.9 ± 10.4% vs. 75.9% ± 6.1%, \( P = 0.048 \)). Those with both MYCN amplification and 11q23 deletion had the worst outcome (\( P < 0.001 \)).

Conclusions: Chromosome band 11q23 deletion predicts poor prognosis only in bone marrow metastatic neuroblastoma patients without MYCN amplification. Combined assessment of the two markers was much superior to single-marker assessment in recognizing the patients at a high risk of disease progression.

Keywords: Neuroblastoma, MYCN amplification, 11q23 deletion, Fluorescence in situ hybridization, Bone marrow metastasis, Event-free survival, Overall survival, Neuron-specific enolase, Lactate dehydrogenase

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Background
Neuroblastoma is the most common extracranial malignancy in children with remarkable heterogeneity in clinical behaviors. Neuroblastoma accounts for 10%–15% of all pediatric cancer-related deaths [1, 2]. Although some of the tumors can regress spontaneously, most patients’ diseases progress aggressively despite multimodality treatment, which may include chemotherapy, radiotherapy, surgical resection, hematopoietic stem cell transplantation, and immunotherapy [3, 4]. In the clinic, half of neuroblastoma patients present with metastasis at diagnosis, and less than 50% survive 5 years [5, 6].

Recently, many prognostic variables have been used for risk stratification to predict the survival of patients with neuroblastoma. Among them, genetic features such as the status of the v-myc avian myelocytomatosis viral oncogene neuroblastoma derived homolog (MYCN) gene [7], 11q23 allele status [8], and tumor ploidy [9, 10] are the most significant and clinically relevant factors. However, for pediatric patients with metastatic neuroblastoma who need chemotherapy prior to surgery, timely identification of genetic aberrations is not possible because of the difficulty in obtaining tumor biopsies. In addition, the results of genetic aberration identification may be inaccurate after the patients have received chemotherapy, leading to insufficient dosage and period of treatment [11, 12].

For more than two decades, a number of genetic aberrations closely associated with outcome have been identified in neuroblastoma; for the MYCN gene, these include 17q gain, loss of 3p, and 11q deletion [13]. Among them, 11q deletion is the most frequent, occurring in approximately 40% of primary neuroblastomas [14], and is associated with poor prognosis [15, 16]. However, the clinical significance of 11q deletion is unclear in neuroblastoma metastatic to bone marrow. What’s more, 11q deletion is not routinely examined when genome-wide methods are used to evaluate genetic aberrations [17]. 11q23 is the most commonly deleted 11q region, and interphase fluorescence in situ hybridization (FISH) has been previously confirmed as a reliable method for detecting MYCN status in bone marrow metastases of neuroblastoma [18, 19]. Therefore, in this study, we used FISH to simultaneously detect MYCN and 11q23 deletion in bone marrow cells in order to evaluate the biological characteristics and survival associated with these two markers.

Materials and methods
Patients and treatment protocols
Consecutive children with neuroblastoma newly diagnosed at the Hematology Oncology Center, Beijing Children’s Hospital, Capital Medical University between January 2015 and December 2016 were selected. All patients were diagnosed according to the International Neuroblastoma Staging System [20]. The criteria for patient inclusion were ≥ 20% neuroblastoma cells in the bone marrow samples and treatment according to the BCH-NB-2007 protocol (based on the Hong Kong neuroblastoma protocol) [21].

The BCH-NB-2007 protocol for newly diagnosed high-risk neuroblastoma was comprised of 7 cycles of intensive chemotherapy: CAV regimen [cyclophosphamide (70 mg/kg) on days 1–2, adriamycin (25 mg/m²) on days 1–3, and vincristine (0.033 mg/kg)] on days 1–3 for cycles 1, 2, 4, and 6; CVP regimen [cisplatin (50 mg/m²) on days 1–4 and VP16 (200 mg/m²) on days 1–3] for cycles 3, 5, and 7. Primary tumor resection was performed after 4 cycles of chemotherapy; peripheral blood stem cells (PBSCs) were then harvested for possible autologous bone marrow transplantation. Autologous PBSC transplantation (PBSCT) was performed after 7 cycles of chemotherapy. The patients received local irradiation (20–25 Gy) at the site of primary tumor as indicated [22]. At 4–5 weeks after autologous PBSCT, they received maintenance chemotherapy with cis-retinoic acid (160 mg/m²) for 14 days alternating with 14 days off for 9–12 months.

The treatment protocol for low-/intermediate-risk neuroblastoma was comprised of 2–3 cycles of CBVP regimen [carboplatin (200 mg/m²) and etoposide (150 mg/m²)] and CADO regimen [vincristine (1.5 mg/m²) and adriamycin (25 mg/m²)] used alternately. Primary tumor resection was usually performed after chemotherapy. If surgery was performed before chemotherapy, 2–3 cycles of chemotherapy for low-risk patients and 4–6 cycles for intermediate-risk patients were needed.

All patients were treated and evaluated according to the BCH-NB-2007 protocol. After the treatment, the patients were followed-up every 3 months mainly by outpatient follow-up visits (bone marrow examination and computed tomography or magnetic resonance imaging) and telephone interview. The final date for data collection was August 31, 2018. This study was approved by the Beijing Children’s Hospital Institutional Ethics Committee. Informed consent was obtained from the parents or their guardians in accordance with the Declaration of Helsinki.

Bone marrow sample processing
Bone marrow samples of all patients obtained by bone marrow puncture were collected in heparin-coated tubes at diagnosis. The sample was immediately shaken gently after collection to avoid bone marrow agglutination. Then, the cells were cultured in 8 mL RPMI-1640 (Hyclone, Beijing, China) and 2 mL fetal bovine serum medium (Invitrogen, Carlsbad, CA, USA) for 24 h. The cell density was (1 – 3) × 10⁶/mL, and the total amount
of cells was about \((1 - 3) \times 10^7\). Colchicine was added at 2 h before termination of the culture. The cells were then treated with 0.075 mol/L KCl for 30 min and fixed twice in a 3:1 mixture of methanol:acetic acid. Finally, the cell suspension was stored in a fresh fixative solution at 4 °C. Bone marrow biopsies were prepared on wax blocks for morphological examination.

**FISH analysis of bone marrow cells**

The FISH technique was performed as previously reported [17]. The statuses of MYCN and 11q were determined with DNA probes from Vysis [N-MYC(2p24)/CEP2(2p11.1-q11.1) Dual Color Probe and LSI MLL Dual Color, Break Apart Rearrangement Probe] (cat. No. 7)72-01 and 8L57-20, Abbott Laboratories, Abbott Park, IL, USA). A Leica DM6000B microscope (Leica Microsystems GmbH, Wetzlar, Germany) was used to take fluorescence images. According to the recommendations of the European Neuroblastoma Quality Assessment group [23, 24], MYCN amplification was defined as a > fourfold increase of MYCN signals in relation to the number of chromosome 2, and 11q23 deletion was defined as only one fusion signal displayed.

**Karyotype analysis of bone marrow cells**

Chromosome karyotype analysis of bone marrow cells was performed on G-banded preparations. At least 20 metaphases were analyzed in detail. The karyotype results were interpreted according to the International System for Human Cytogenomic Nomenclature guidelines (2016) [25]. The clones containing at least 2 cells with the same additional chromosome or structural abnormality or at least 3 cells with the same missing chromosome were defined as abnormal clones.

**Morphologic analysis and diagnostic biomarker detection**

Microscopic examination of bone marrow biopsies to determine the presence of neuroblastoma cells was done by at least 2 independent laboratory experts. Serum tumor markers such as lactate dehydrogenase (LDH) and neuron specific enolase (NSE) levels were detected by radioimmunoassay and full-automatic biochemical analysis at the time of diagnosis in all patients.

**Statistical analysis**

All statistical analyses were performed using the SPSS software, version 16.0 (SPSS Inc., Chicago, IL, USA). Comparisons of patients with or without 11q23 deletion and MYCN amplification and the association of clinical characteristics were evaluated using the \(\chi^2\) test. Event-free survival (EFS) was defined from the date of diagnosis to the date of death due to any reason or the last contact with patients in continuous CR. Kaplan–Meier survival analysis and log-rank test were used to determine the differences in EFS and OS in all patients by MYCN and 11q23 statuses. Multivariate backward Cox regression analysis model was built which included only those variables reaching a \(P\) value < 0.05 in the respective univariate analysis. All tests with a \(P<0.05\) in two-sided distributions were considered statistically significant.

**Results**

**Clinical characteristics of 101 patients with bone marrow metastatic neuroblastoma**

Among the 161 patients with newly diagnosed neuroblastoma, 45 had no bone marrow metastasis, 13 gave up treatment after diagnosis, and 2 were lost to follow-up. A total of 101 patients with stage IV neuroblastoma were thereby analyzed, including 57 males and 44 females in a ratio of 1.3:1. The patients were aged 7–105 months (median, 36 months), and 85 (84.2%) were older than 18 months. The median white blood cell (WBC) count was \(5.6 \times 10^9/L\) (range \(1.7–42.5 \times 10^9/L\)) with 60 (59.4%) patients having normal WBC. Only 8 (7.9%) patients had hemoglobin >120 g/L. The rest had different degrees of anemia, but most cases were mild. The median hemoglobin level was 94 g/L (range 58–135 g/L). The amount of platelets ranged from \(37 \times 10^9/L\) to \(622 \times 10^9/L\) (median, \(275 \times 10^9/L\)). Almost all patients (92.1%) were grouped as having high-risk neuroblastoma. The primary tumor sites were the abdomen for 88 patients, thorax for 12 patients, and neck for 1 patient. 11q23 deletion was seen in 40 (39.6%) patients, and MYCN amplification in 12 (11.9%). In terms of clinical serum tumor markers, only 22 (21.8%) patients had LDH levels ≥ 1500 IU/L, and 41 (40.6%) had NSE levels ≥ 370 ng/mL.

Each patient received a total of 7 cycles of chemotherapy except for those who died during therapy. During follow-up, 39 (38.6%) patients died, and 13 (12.9%) suffered from tumor relapse or progression; the other 49 (48.5%) had stable disease. Among the patients who died, 11 died of tumor progression during intensive chemotherapy, 24 died of tumor progression during maintenance treatment, and 4 died of tumor recurrence after the completion of chemotherapy. Of note, 4 of the 24 patients who died during maintenance treatment had intracranial metastasis. The main clinical characteristics of the 101 patients are shown in Table 1.

**Genetic markers of MYCN and 11q23 in bone marrow cells**

In all patients, interphase FISH detected the MYCN gene and 11q23 status of bone marrow cells. Chromosome
karyotype analysis was performed in 55 patients. MYCN amplification was seen in 12 (11.9%) patients, with > 30 gene copies/cell (Fig. 1a). MYCN gain was seen in 21 (20.8%) patients, with 2–9 signal copies. Among the 101 patients, 40 (39.6%) had 11q23 deletion (Fig. 1b). Only 2 (2.0%) patients had both MYCN amplification and 11q23 deletion. MYCN amplification and 11q23 deletion were not associated with higher percentages of tumor cells in bone marrow or more severe hematopoietic suppression (both \( P > 0.05 \)).

**Relationship of MYCN and 11q23 statuses with clinicobiological characteristics of patients with bone marrow metastatic neuroblastoma**

We observed no association of MYCN amplification with age, sex, or primary tumor site (Table 2). However, the primary tumor site of all patients with MYCN amplification was in the abdomen. We found that MYCN amplification mainly occurred in those with high LDH and NSE levels (both \( P < 0.001 \)), and more events were observed in patients with MYCN amplification than in those with normal MYCN (\( P = 0.004 \)). Unlike the MYCN gene, 11q23 deletion was associated with age (\( P = 0.024 \)). We found 38 (95.0%) cases of 11q23 deletion occurred in children older than 18 months. The median age at diagnosis was 41 months for patients with 11q23 deletion, but 24.5 months for those with MYCN amplification. Although 24 (60.0%) patients with 11q23 deletion had events occurred, no significant difference was observed between patients with 11q23 deletion and normal 11q23 (\( P = 0.118 \)).

**Impact of MYCN and 11q23 on prognosis of bone marrow metastatic neuroblastoma**

The 3-year EFS and OS rates of the 101 patients were 45.9 ± 5.5% and 59.1 ± 5.2%, with a median follow-up time of 23 months (range, 4–44 months). The estimated 3-year EFS and OS rates of patients with MYCN amplification were obviously lower than those of patients with normal MYCN (8.3 ± 8.0% vs. 43.8 ± 8.5% and 10.4 ± 9.7% vs. 63.5% ± 5.7%, both \( P < 0.001 \)) (Fig. 2a, b). The difference in survival was not significant between patients with and without 11q23 deletion (Fig. 2c, d). The median EFS of patients with and without MYCN amplification was 12.5 and 25 months, respectively. The median EFS of patients with and without 11q23 deletion was 20.5 and 26 months, respectively.

To better understand the effect of 11q23 deletion on prognosis of bone marrow metastatic neuroblastoma, we performed further survival analysis of the 89 patients without MYCN amplification. We surprisingly found that 11q23 deletion was a powerful prognostic factor if the influence of the MYCN gene was excluded. The

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**Table 1 Baseline characteristics of the 101 patients with bone marrow metastatic neuroblastoma**

| Characteristic                  | Total [cases (%)] |
|--------------------------------|-------------------|
| Age (months)                   |                   |
| < 18                           | 16 (15.8)         |
| ≥ 18                           | 85 (84.2)         |
| Sex                            |                   |
| Male                           | 57 (56.4)         |
| Female                         | 44 (43.6)         |
| WBC (x 10^9/L)                 |                   |
| > 10                           | 17 (16.8)         |
| 4–10                           | 60 (59.4)         |
| < 4                            | 24 (23.8)         |
| Hemoglobin (g/L)               |                   |
| > 120                          | 8 (7.9)           |
| 90–120                         | 59 (58.4)         |
| 60–89                          | 33 (32.7)         |
| < 60                           | 1 (1.0)           |
| Platelet (x 10^9/L)            |                   |
| > 300                          | 47 (46.5)         |
| 100–300                        | 48 (47.5)         |
| < 100                          | 6 (6.0)           |
| Risk stratification            |                   |
| Low-risk                       | 1 (1.0)           |
| Intermediate-risk              | 7 (6.9)           |
| High-risk                      | 93 (92.1)         |
| Primary tumor site             |                   |
| Abdomen                        | 88 (87.1)         |
| Thorax                         | 12 (11.9)         |
| Neck                           | 1 (1.0)           |
| 11q23 deletion                 |                   |
| Yes                            | 40 (39.6)         |
| No                             | 61 (60.4)         |
| MYCN amplification             |                   |
| Yes                            | 12 (11.9)         |
| No                             | 89 (88.1)         |
| LDH (IU/L)                     |                   |
| < 1500                         | 79 (78.2)         |
| ≥ 1500                         | 22 (21.8)         |
| NSE (ng/mL)                    |                   |
| < 370                          | 60 (59.4)         |
| ≥ 370                          | 41 (40.6)         |
| Event                          |                   |
| No event                       | 49 (48.5)         |
| Relapse/progression            | 13 (12.9)         |
| Death                          | 39 (38.6)         |

WBC white blood cell, MYCN the v-my c avian myelocytomatosis viral oncogene neuroblastoma derived homolog, LDH lactate dehydrogenase, NSE neuron-specific enolase
Fig. 1 Typical fluorescence in situ hybridization (FISH) images of bone marrow cells from patients with bone marrow metastatic neuroblastoma. a The status of v-myc avian myelocytomatosis viral oncogene neuroblastoma derived homolog (MYCN) amplification was detected using a dual-color probe. Green signals represent the specific probe for MYCN, and red signals stand for centromeric chromosome 2 probes. MYCN signals show more than 10 copies within the nuclei. b The status of 11q23 deletion was also detected using a dual-color probe. Normal cells show a two orange/green fusion signal pattern. In a cell with 11q23 deletion, the pattern is one orange/green fusion signal.

Table 2 Clinical characteristics of bone marrow metastatic neuroblastoma patients according to MYCN and 11q23 statuses

| Variable          | MYCN status [cases (%)] | P value* | 11q23 status [cases (%)] | P value* |
|-------------------|-------------------------|----------|--------------------------|----------|
|                   | Amplified | Normal |               | Deleted | Normal |               |                |
| Total             | 12 | 89 |               | 40 | 61 |               |                |
| Sex               |          |       |               |          |       |               |                |
| Male              | 7 (58.3) | 50 (56.2) | 0.536 | 24 (60.0) | 33 (54.1) | 0.682 |
| Female            | 5 (41.7) | 39 (43.8) |       | 16 (40.0) | 28 (45.9) |       |
| Age (months)      |          |       |               |          |       |               |                |
| < 18              | 4 (33.3) | 12 (13.5) | 0.095 | 2 (5.0) | 14 (23.0) | 0.024 |
| ≥ 18              | 8 (66.7) | 77 (86.5) |       | 38 (95.0) | 47 (77.0) |       |
| Primary tumor site|          |       |               |          |       |               |                |
| Abdomen           | 12 (100.0) | 76 (85.4) | 0.436 | 33 (82.5) | 55 (90.2) | 0.339 |
| Thorax            | 0 (0) | 12 (13.5) |       | 6 (15.0) | 6 (9.8) |       |
| Neck              | 0 (0) | 1 (1.1) |       | 1 (2.5) | 0 (0) |       |
| LDH (IU/L)        |          |       |               |          |       |               |                |
| < 1500            | 1 (8.3) | 78 (87.6) | <0.001 | 32 (80.0) | 47 (77.0) | 0.808 |
| ≥ 1500            | 11 (91.7) | 11 (12.4) |       | 8 (20.0) | 14 (23.0) |       |
| NSE (ng/mL)       |          |       |               |          |       |               |                |
| < 370             | 1 (8.3) | 59 (66.3) | <0.001 | 21 (52.5) | 39 (63.9) | 0.302 |
| ≥ 370             | 11 (91.7) | 30 (33.7) |       | 19 (47.5) | 22 (36.1) |       |
| Event             |          |       |               |          |       |               |                |
| No                | 1 (8.3) | 48 (53.9) | 0.004 | 16 (40.0) | 33 (54.1) | 0.118 |
| Yes               | 11 (91.7) | 41 (46.1) |       | 24 (60.0) | 28 (45.9) |       |

MYCN, the v-myc avian myelocytomatosis viral oncogene neuroblastoma derived homolog, LDH, lactate dehydrogenase, NSE, neuron-specific enolase

* The χ² test was used for statistical analysis
Fig. 2 The prognostic significance of MYCN amplification and 11q23 deletion in patients with bone marrow metastatic neuroblastoma. 

a, b Event-free survival (EFS) and overall survival (OS) of the 101 patients stratified by MYCN gene. 

c, d EFS and OS of the 101 patients stratified by 11q23 status. 

e, f EFS and OS of the 89 patients without MYCN amplification stratified by 11q23 status. 

g, h EFS and OS of the 101 patients stratified by combined assessment of MYCN and 11q23 statuses.
Table 3 Univariate and multivariate analysis of event-free survival (EFS) of children with bone marrow metastatic neuroblastoma

| Variable                        | Total patients (n = 101) | Patients without MYCN amplification (n = 89) |
|---------------------------------|--------------------------|---------------------------------------------|
|                                 | Univariate | Multivariate | Univariate | Multivariate |
|                                 | P          | HR 95% CI    | P          | HR 95% CI    |
| MYCN status (amplification vs. normal) | 0.001      | 2.399 1.113–5.169 | 0.032      | –            |
| LDH (≥ 1500 vs. < 1500 IU/L)    | 0.001      | 1.543 0.622–3.83 | 0.351      | 0.040 1.488 0.564–3.926 |
| NSE (≥ 370 vs. < 370 ng/mL)     | 0.001      | 1.829 0.972–3.44 | 0.068      | 0.025 2.016 1.074–3.784 |
| 11q23 status (deletion vs. normal) | 0.153      | –           | –          | 0.037 1.674 0.869–3.225 |

*Univariate analysis for EFS was performed using the Kaplan–Meier log-rank test. All factors with P < 0.05 in the univariate analysis are shown.

Discussion

In recent years, the pretreatment risk stratification for neuroblastoma patients has become increasingly necessary [26, 27]. Currently, the identified biological factors closely related to prognosis include age, histology, ploidy, MYCN gene and specific segmental chromosomal aberrations [28, 29]. In the present study, we evaluated the prognostic value of 11q23 deletion and the prognostic factor in multivariate analysis only for patients without MYCN amplification, but it was not an independent prognostic factor in the multivariate analysis.

It was reported that 11q23 deletion may be involved in the dysregulation of important genes, including tumor suppressor genes and oncogenes [32]. One candidate
gene is forkhead box R1 (FOXRI), which is located in 11q23.3. 11q23.3 deletion results in either mixed-lineage leukemia (MLL) or platelet activating factor acetylhydrolase 1b catalytic subunit 2 (PAFAH1B2) fusing to the proximal side of FOXRI. The new fusion transcripts stimulate the overexpression of FOXRI. Functional analyses have shown that FOXRI overexpression could promote cell proliferation and may be a tumor-driving event [33]. Another important tumor suppressor gene is h2a histone family member X (H2AFX), which resides in the 11q23.2-q23.3 region. H2AFX participates in DNA double-strand breaks (DSBs) through phosphorylation. The loss of H2AFX promotes tumorigenesis due to defective DSB repair, increased radiation sensitivity, and genomic instability [10]. It was reported that more chromosomal breaks were seen in neuroblastoma patients with 11q23 deletion, which may be explained by genomic instability related to the loss of H2AFX gene [34]. This hypothesis also implied that 11q23 deletion occurred early in tumorogenesis. However, in the present study, the median age at diagnosis was 41 months for patients with 11q23 deletion and 24.5 months for those with MYCN amplification. The definitive role of 11q23 deletion in tumorigenesis of neuroblastoma still needs to be elucidated.

The present study showed that a large number of patients with MYCN amplification or 11q23 deletion had unfavorable prognosis. The survival rate was higher in patients with 11q23 deletion than in those with MYCN amplification. However, OS has been reported to be similar for patients with the two aberrations after 8 years of follow-up [10]. We further evaluated the prognostic value of combined assessment of MYCN and 11q23 statuses and found that the combined assessment was much superior to single-marker assessment in recognizing the patients at a high risk of disease progression.

The present study investigated the outcomes of bone marrow metastatic neuroblastoma patients with 11q23 deletion and of the rare patients with both MYCN amplification and 11q23 deletion. However, there were some limitations of this study. First, it is unclear how the results would be used to affect treatment. Second, the cases of both MYCN amplification and 11q23 deletion were limited. In our future study, we will focus on how further subdivision of these patients would alter treatment.

Conclusions
In the present study, we found that patients with bone marrow metastatic neuroblastoma, 11q23 deletion predicted poor prognosis only in those without MYCN amplification. Combined assessment of the two markers was superior to single-marker assessment in identifying patients at a high risk of disease progression.
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