Identification of Prognosis-related RNA Binding Proteins to Reveal the Role of RNA Binding Proteins in the Progression and Prognosis of Colon Cancer

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Abstract

**Background:** RNA binding proteins (RBPs) are now under discussion as novel promising bio-markers for patients with colon cancer. The purpose of our study is to identify several RBPs related to the progression and prognosis of colon cancer, and to further investigate the mechanism of their influence on tumor progression.

**Methods:** The transcriptome data of colon cancer as well as clinical characteristics used in this study were downloaded from the The Cancer Genome Atlas (TCGA) database. Gene ontology (GO), Kyoto Encyclopedia of Genes and Genomes (KEGG) analysis and Gene set enrichment analysis (GSEA) were performed to elucidate the gene functions and relative pathways. Cox and lasso regression analysis were used to analyze the effect of immune genes on the prognosis of breast cancer. Immune risk scoring model was constructed based on the statistical correlation between hub immune genes and survival. Meanwhile, multivariate cox regression analysis was utilized to investigate whether the immune genes risk score model was an independent factor for predicting the prognosis of breast cancer. Nomogram was constructed to comprehensively predict the survival rate of breast cancer. P< 0.05 was considered to be statistically significant.

**Results:** The results of the difference analysis showed that 473 RBPs exhibited differential expression between normal and colon cancer tissues (p<0. 05). Univariate cox regression analysis revealed 25 RBPs statistically correlated with colon cancer related survival risk (P<0.05). In addition, a 10-RBPs based risk scoring model was constructed through multivariate cox regression analysis. KM curve indicated that patients in high-risk were associated with poor outcomes (p<0.001). ROC curve indicated that the immune risk score model was reliable in predicting survival risk (5-year OS, AUC=0.782). Our model showed satisfying AUC and survival correlation in the validation dataset (5-year OS AUC=0.744). Furthermore, multivariate cox regression analysis confirmed that the immune risk score model was an independent factor for predicting the prognosis of colon cancer. A nomogram was established to comprehensively predict the survival of colon cancer patients with the results of multivariate cox regression analysis. Finally, we found that 10 RBPs and risk scores were significantly associated with clinical factors and prognosis, and were involved in multiple oncogenic pathways.

**Conclusion:** Collectively, RBPs played an essential role in the progression and prognosis of colon cancer by regulating multiple biological pathways. Furthermore, RBPs risk score was an independent predictive factor of colon cancer, indicating a poor survival.

**Introduction**

Colon cancer, a major malignancy of the alimentary canal, ranked third among malignant tumors in terms of morbidity worldwide (1, 2). Relevant study revealed that more than 1 million people developed colon cancer each year, and the disease-specific mortality rate in developed countries was approximately 33% (3). Mortality of colon cancer are on the rise due to changes in diet and lifestyle (4, 5). Although colon
cancer treatment options (e.g. surgery, chemoradiotherapy, and immunotherapy) have been greatly improved, the 5-year survival rate remained as high as 50% (6).

RNA binding proteins (RBPs) are a variety of proteins that interact with RNA, which are widely expressed in cells (7–9). Through high-throughput screening, 1542 RBPs were identified, accounting for 7.5% of all protein coding genes (10). These RBPs affect posttranscriptional events and regulate physiological events of cells, thus involved in many biological processes, such as RNA splicing, mRNA stabilization and protein translation (7, 11). As RBPs play a variety of key functions in post transcriptional events, the changes of RBPs are related to the occurrence and development of many human diseases. However, the role of RBPs in the development of colorectal cancer remain unclear.

The Cancer Genome Atlas (TCGA) was regarded as the largest cancer database, containing samples of more than 20,000 primary cancers and normal matched samples of multiple cancer types. Therefore, we can investigate tumor gene data in greater depth with bioinformatics methods. Further, it can be linked to clinical data in order to obtain more valuable and meaningful results. Here, we described the expression and lineage of RBPs in colon cancer, and investigate the mechanisms of RBPs to the development and prognosis of colon cancer.

**Methods And Materials**

**Data acquisition**

We identified and downloaded the transcriptome data of patients with colon cancer from the TCGA database through the R package, including 41 cases of paracancerous normal tissue and 473 cases of tumor tissue. Further, relevant clinical information of 473 colon cancer patients were obtained such as age, gender, stage, tumor&Lymph node&metastasis stage, survival status and survival duration (Table 1). Finally, "Limma" package in R software was utilized to correct the transcriptome data we have downloaded.
Table 1
Clinical characteristics of included patients in the study.

| Variables | Total (n = 387) | Training cohort (n = 232) | Validation cohort (n = 155) |
|-----------|----------------|--------------------------|-----------------------------|
| Age (year) |                |                          |                             |
| < 60      | 91             | 53                       | 48                          |
| ≥ 60      | 286            | 179                      | 107                         |
| Gender    |                |                          |                             |
| FEMALE    | 184            | 109                      | 75                          |
| MALE      | 203            | 123                      | 80                          |
| Stage     |                |                          |                             |
| I         | 66             | 35                       | 31                          |
| II        | 158            | 100                      | 58                          |
| III       | 102            | 60                       | 42                          |
| IV        | 61             | 37                       | 24                          |
| T stage   |                |                          |                             |
| T1        | 8              | 5                        | 3                           |
| T2        | 65             | 32                       | 33                          |
| T3        | 168            | 172                      | 96                          |
| T4        | 46             | 23                       | 23                          |
| N stage   |                |                          |                             |
| N1        | 232            | 139                      | 93                          |
| N2        | 88             | 51                       | 37                          |
| N3        | 67             | 42                       | 25                          |
| M stage   |                |                          |                             |
| M0        | 326            | 195                      | 131                         |
| M1        | 61             | 37                       | 24                          |
| Survival  |                |                          |                             |
| Dead      | 73             | 46                       | 27                          |
| Alive     | 314            | 186                      | 128                         |
Gene function enrichment analysis

In order to explore the major biological process of selected hub-genes, methods were utilized to conduct the gene functional enrichment analyses including Kyoto Encyclopedia of Genes and Genomes (KEGG) and gene ontology (GO). We utilized Database for Annotation, Visualization, and Integrated Discovery (DAVID, https://david.ncifcrf.gov/) to identify enriched KEGG and GO themes.

Survival analysis and expression comparison of hub-genes

Clinical information for TCGA colon cancer including survival time, survival state, and TNM staging were also downloaded from the TCGA database (samples with missing information were excluded). Survival R package was applied in survival analyses for hub-genes. For the overall survival rates, the logrank test was used to detect significant differences. The results were visualized using Kaplan-Meier survival curves, and P-value < 0.05 was considered as statistically significant.

Gene set enrichment analysis

Gene enrichment analysis (GSEA) (version 3.0, the broad institute of MIT and Harvard, http://software.broadinstitute.org/gsea/downloads.jsp) was conducted between colon cancer and paracancerous normal tissues to study the biological characteristics of colon cancer. In detail, the “collapse data set to gene symbols” was set to false, the number of marks was set to 1000, the “permutation type” was set to phenotype, the “enrichment statistic” was set to weighted, and the Signal2Noise metric was used for ranking genes. High expression group was used as experimental group and low expression group was used as reference group. “c2.cp.kegg.v7.0.symbols.gmt” gene sets database was used for enrichment analysis. Gene set size > 500 and < 15, FDR < 0.25, and nominal P-value < 0.05 were regarded as the cut-off criteria.

Statistical analysis

All analyses were performed using R 3.6.1. All statistical tests were two-sided, and P value < 0.05 was considered statistically significant. Continuous variables that conformed to the normal distribution were compared with the use of independent t test for comparison between groups, while continuous variables with skewed distribution were compared with the Mann-Whitney U test. The correlation matrix was constructed by R software based on Pearson Correlation Coefficient. The relationship between immune cell infiltration and overall survival was analyzed through the Kaplan-Meier curve which was evaluated by log-rank test. Time-dependent ROC curves were used to analyze the sensitivity and specificity of the recurrence prediction model. The univariate and multivariate regression model was used to analyze the effects of individual variables on survival. The nomogram was constructed with the regression coefficients based on the cox analysis.

Results

Differential expression screening of colon cancer
The transcriptome data of 473 colon cancer cases and 41 adjacent normal tissues cases was obtained from TCGA database for differential expression analysis. A total of 473 RBPs were identified as differentially expressed RBPs (DERBPs) between breast cancer and normal tissues, including 321 up-regulated and 152 down regulated (p < 0.05, Fig. 1A, Table 2). The heatmap of the top 10 up-regulated and top 10 down regulated DERBPs was shown in Fig. 1B.
Table 2
Univariate cox regression analysis to screen RNA binding proteins associated with overall survival in colon cancer patients.

| Gene     | HR    | HR.95L  | HR.95H  | pvalue       |
|----------|-------|---------|---------|--------------|
| CELF4    | 15.87213555 | 4.608502177 | 54.66519864 | 1.18E-05     |
| LUZP4    | 344.8138082 | 15.22018153 | 781.1770317 | 0.000242467  |
| KHDC1L   | 3.309097051  | 1.701354981 | 6.436119103 | 0.000422443  |
| TDRD7    | 0.416408253  | 0.233832009 | 0.741540193 | 0.002924262  |
| NOP14    | 0.376647057  | 0.188141644 | 0.754022358 | 0.005830105  |
| TDRD6    | 0.063910916  | 0.009002748 | 0.453706475 | 0.00595612   |
| EEF1A2   | 1.369621708  | 1.090242892 | 1.720592389 | 0.006887091  |
| PPARGC1A | 0.579714951  | 0.380038455 | 0.884303731 | 0.011384635  |
| NOL3     | 1.670109583  | 1.115583263 | 2.500275965 | 0.012729851  |
| EXOG     | 0.292343173  | 0.109198587 | 0.782652346 | 0.014376367  |
| RIOK1    | 2.361681822  | 1.18371422 | 4.711898305 | 0.014746644  |
| CAPRIN2  | 1.666977568  | 1.087328425 | 2.55563466 | 0.019078017  |
| EIF2AK3  | 0.436146581  | 0.214390243 | 0.887351677 | 0.022034038  |
| PNLDC1   | 1.574919384  | 1.064983919 | 2.329022084 | 0.022884063  |
| PPARGC1B | 0.45676183  | 0.231685293 | 0.900494662 | 0.023660181  |
| ZNF385A  | 1.310439816  | 1.032491852 | 1.663211683 | 0.02625529   |
| AEN      | 1.694187437  | 1.059461667 | 2.70917878 | 0.027727458  |
| ERI1     | 0.589956581  | 0.368421266 | 0.94470325 | 0.028037187  |
| SRP14    | 1.93932163  | 1.07191063 | 3.508658541 | 0.028558768  |
| RRS1     | 1.620765905  | 1.039435069 | 2.527220985 | 0.033120985  |
| MAK16    | 0.583949222  | 0.353353075 | 0.965031066 | 0.035830825  |
| AFF3     | 3.343687337  | 1.053692809 | 10.61053554 | 0.040488238  |
| POP1     | 0.554888789  | 0.314556342 | 0.978840764 | 0.041970548  |
| ZC3H12C  | 0.624988088  | 0.392567184 | 0.995014676 | 0.047588085  |
| TRIM25   | 0.61232798  | 0.375283794 | 0.999098712 | 0.049579544  |

Functional annotation of these 473 DERBPs
In order to fully understand the biological attributes of these 473 DERBPs, we conducted Kyoto Encyclopedia of Genes and Genomes (KEGG) and gene ontology (GO) analysis. Based on the results of DAVID, the top three enriched GO terms for up-regulated genes and down-regulated genes were: ncRNA processing, ribosome biogenesis and rRNA metabolic process; RNA splicing, regulation of translation and regulation of cellular amide metabolic process, respectively (Fig. 2A). The top biological pathway enriched for up-regulated genes and down-regulated genes were: Ribosome biogenesis in eukaryotes, RNA transport and Spliceosome; Spliceosome, RNA transport and Influenza A pathway, respectively (Fig. 2B).

Establishment of immune prognosis model

For the purpose of revealing the relationship between these 473 DERBPs and overall survival, 25 prognostic DERBPs were identified by utilizing univariate Cox regression analyses (Fig. 3A, Table 2). TCGA colon cancer data were randomly divided into two sets (training set : validation set, 3 : 2). Then, multivariate cox regression analysis was applied to select the independent indicators for the overall survival based on training set and finally we got 10 DERBPs (PPARGC1A, ZNF385A, SRP14, RIOK1, ERI1, NOL3, RRS1, TDRD6, AEN and PNLDC1) for the construction of prognostic index (Fig. 3B, Table 3). After the construction of prognostic index, patients were separated into high risk and low risk (Fig. 3C, Fig. 3D). Heat map was utilized to visualize the difference of gene expression profile in low- and high- risk patients in breast cancer training set (Fig. 3E). The results from K-M analysis indicated that high risk patients had lower overall survival than low risk patients in both training group and validation group (P < 0.001) (Fig. 4A, Fig. 4B, Fig. 4C). The ROC curve revealed that the risk model had a good sensitivity and specificity in predicting survival risk (AUC = 0.782, AUC = 0.744 for 5 years overall survival in training and validation group, respectively) (Fig. 4D, Fig. 4E, Fig. 4F). To explore whether the constructed RBPs risk scoring model was independent form age, gender, stage, and other clinical pathological parameters, we performed an univariate and multivariate cox regression analysis for age, gender, stage, TNM and risk score. In univariate Cox model, age, pathological stage, pathological T, N, M stage and high risk score were associated with poor survival (Fig. 5A). In multivariate Cox model, only age, stage and risk score worked as independent predicted factors (Fig. 5B). To better predict the prognosis of breast cancer patients at three and five years post-surgery, we constructed a new nomogram from the variables associated with OS (age, pathological stage, TNM stage and risk score) (Fig. 5C, 5D, 5E).
Table 3
Multivariate cox regression analysis to establish RNA binding proteins risk prediction model.

| Gene     | Coef     | HR      | HR.95L  | HR.95H  | pvalue  |
|----------|----------|---------|---------|---------|---------|
| PPARGC1A | -0.43626957 | 0.646277076 | 0.368761746 | 1.132639335 | 0.127288514 |
| ZNF385A  | 0.347562389 | 1.415612627 | 0.979935812 | 2.044990177 | 0.064030703 |
| SRP14    | 0.801814972 | 2.22958389 | 0.974868312 | 5.099195716 | 0.057477623 |
| RIOK1    | 1.410546886 | 4.098196037 | 1.645449933 | 10.20706278 | 0.00244868 |
| ERI1     | -0.600993288 | 0.548266779 | 0.252246613 | 1.191676894 | 0.12920327 |
| NOL3     | 0.530884133 | 1.700435056 | 0.918066471 | 3.149531618 | 0.091385734 |
| RRS1     | 0.934795492 | 2.546692586 | 1.27947903 | 5.068971805 | 0.007774571 |
| TDRD6    | -2.621256381 | 0.072711452 | 0.004178531 | 1.265266376 | 0.07209323 |
| AEN      | 0.987247699 | 2.683837569 | 1.357006588 | 5.307994936 | 0.004549131 |
| PNLDC1   | 0.686749923 | 1.987246323 | 1.280883189 | 3.083144489 | 0.002179074 |

Clinical and prognostic correlation of 10 model genes and RBPs risk score

We further investigated the proportion of each model genes in different pathological stages. We demonstrated that SRP14, PPARGC1A and ERI1 were most significantly associated with development of colon cancer (Fig. 6). Regard to the immune genes risk score, a strong correlation with pathological stage, clinical N stage and clinical M stage was identified (Fig. 7).

Gene set enrichment analysis of risk scores

To explore the biological relevance of risk scores involved in progression of colon cancer, we performed a gene set enrichment analysis of risk scores based on the TCGA colon cancer cohort. GSEA analysis indicated high risk scores was associated with MYC_TARGETS_V2, UV_RESPONSE_UP and WNT_BETA_CATENIN_SIGNALING pathway (Fig. 8A, Fig. 8B, Fig. 8C). In addition, low risk scores was associated with PROTEIN_SECRETION pathway (Fig. 8D).

Discussion

Malignant tumors are characterized by uncontrolled cell growth, which is mainly due to the dysregulated expression of cancer driver genes that regulate cell proliferation (12). Posttranscriptional mechanisms can greatly influence gene expression patterns in cancer cells, in which RNA-binding proteins (RBPs) play key roles. They can interact with target mRNAs in a sequence-dependent and structure-dependent manner, and determine cellular behavior by manipulating the processing of these mRNAs (7). It has been reported that RBPs show dysregulated expression in various human cancers (13). However, little is currently
known about the expression patterns and roles of RBPs in colon cancer. Therefore, it was crucial to investigate the RBPs subsets for the evaluation of risk and tumor prognosis in colon cancer.

In our study, we conducted a comprehensive and detailed assessment of RBPs in breast cancer, based on the data from a large set of samples. All gene expression data and patients clinical characteristics information were downloaded from TCGA dataset. We analyzed the 1542 RBPs between colon cancer and normal tissues, eventually, we verified 473 DEIGs. Moreover, we identified and constructed a 10 hub RBPs risk score model for colon cancer via univariate and multivariate cox regression analysis, including PPARC1A, ZNF385A, SRP14, RIOK1, ERI1, NOL3, RRS1, TDRD6, AEN and PNLDC1. Furthermore, to investigate the prognostic value of the model, we performed the ROC curve and investigate the association between the model and clinical features. As expected, the high-risk group was correlated with worse overall survival and was inclined to have advanced stage and higher histological grade which might manifest poor outcome.

According to the result of the biological functions and pathway enrichment analysis of these differentially expressed RBPs, the upregulated RBPs were significantly enriched in ribosome biogenesis in eukaryotes, RNA transport and Spliceosome while downregulated differentially expressed RBPs were enriched in spliceosome, RNA transport and influenza A pathway. Recently, mechanistic data have emerged, suggesting a broader role for dysregulated ribosome biogenesis in the development and progression of most spontaneous cancers (14). Moreover, it is reported that RNA polymerases are consistently dysregulated in cancer, which is mostly mediated through upstream oncogenetic and tumor suppressive signaling pathways rather than through mutations (15). These results suggest that RBPs can affect the growth of tumor cells by regulating multiple biological pathways.

Several genes in the RBPs model had been investigated in human cancers. RIOK1 activates NF-KB signal transduction, which promotes cell cycle progression and tumor lung colonization in vivo. It is demonstrated that RIOK1 is overexpressed in different subtypes of human lung cancer and breast cancer, suggesting that RIOK1 is a potential therapeutic target, especially in Ras driven cancer (16). Ribosome biogenesis regulatory protein homolog (RRS1) is an important factor in ribosome biogenesis. At the molecular level, RRS1 silencing decreased the expression of M phase inducer phosphatase 3 (CDC25C), cyclin dependent kinase 1 (CDK1) and antigen Ki-67 (Ki67), and increased the protein levels of cyclin dependent kinase inhibitor 1 (CDKN1A) and tumor suppressor p53 (p53). In conclusion, RRS1 may promote the development of colon cancer. Therefore, targeting RRS1 may be a promising treatment strategy for CRC patients (17).

However, there were some limitations in our research. Firstly, the sample size in our study was small and a larger cohort and more abundant sequencing results were needed. Secondly, we only focused on the gene expression and gene mutation level, but ignored other events such as the gene methylation, and copy number amplification, which were also important in tumor progression. Finally, the effect of RBPs on the progression and prognosis of colorectal cancer need to be verified in vivo and in vitro.
In summary, our study sheds light on the utility of RBPs in the prognosis of colon cancer. The constructed RBPs risk scoring model is reliable in predicting the prognosis of colon cancer, and this risk scoring model is an independent influencing factor for the prognosis of colon cancer. With the rapid development of high-throughput technology, we have confidence to believe that our risk scoring model have great potential in clinical practice.

**Declarations**

**Conflicts of Interest:**

The authors have no conflicts of interest to declare.

**Ethical Statement:**

The authors are accountable for all aspects of the work (if applied, including full data access, integrity of the data and the accuracy of the data analysis) in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

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Not applicable.

**Authors contributions**

Qi Zou and Xiaoping Yang designed this work. Yue Ding and Dejun Wu performed the experiment and wrote the manuscript. Yuxiang Dong and Junyi Wang performed the bioinformatics analysis. Ping Liu, Wei Han and Zhijun Min performed the data review. All authors have read and approved the manuscript.

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**Figures**

**Figure 1**

Identification of DERBPs. (A) volcano plots of 473 DERBPs in colon cancer and normal tissues from TCGA database. (B) Heatmap plots of top 10 up-regulated and top 10 down-regulated DERBPs. The colors in the heatmaps from green to red represent expression level from low to high. The red dots in the volcano plots represent up-regulation, the green dots represent down-regulation and black dots represent genes without differential expression.
Figure 2

GO (A) and KEGG(B) enrichment analysis of DERBPs.
Figure 3

(A) Univariate survival analysis by cox proportional hazards models to select prognostic key RBPs. (B) Multivariate cox regression model for 10 prognostic RBPs used to construct RBPs risk score model. (C) Distribution of immune risk scores in breast cancer patients. (D) Distribution of survival status in colon cancer patients. (E) Distribution of specific risk factors in the high- and low-risk groups (divided by median value). (*P<0.05)
Figure 4

(A) Kaplan-Meier curve analysis of high-risk and low-risk patients in the training cohort. (B) Kaplan-Meier curve analysis of high-risk and low-risk patients in the testing cohort. (C) Kaplan-Meier curve analysis of high-risk and low-risk patients in the entire TCGA cohort. (D) Time-dependent ROC curve analysis of the training cohort. (E) Time dependent ROC curve analysis of the testing cohort. (F) Time-dependent ROC curve analysis of the entire TCGA cohort.
Figure 5

Cox's proportional hazard model of correlative factors in colon cancer patients. (A) Univariate COX regression analysis for seven clinicopathological parameters affecting the overall survival. (B) Multivariate COX regression analysis for seven clinicopathological parameters affecting the overall survival. (C) An established nomogram to predict breast cancer survival based on cox model. (D-E) Plots displaying the calibration of each model comparing predicted and actual 3- and 5-year overall survival.
Figure 6

Correlation analysis between TNM&Stage and 10 model genes in colon cancer cases. (A) Correlation analysis between tumor stage and 10 model genes expression in colon cancer cases. (B) Correlation analysis between node stage and 10 model genes expression in colon cancer cases. (C) Correlation analysis between metastasis stage and 10 model genes in colon cancer cases. (D) Correlation analysis between pathologic stage and 10 model genes expression in colon cancer cases.
Figure 7

Correlation between RBPs risk scores and various clinical factors. (A) Age. (B) Gender. (C) Stage. (D) T stage. (E) N stage. (F) M stage.
Figure 8

Gene set enrichment analysis of RBPs risk scores. (A-C) high risk scores. (D) low risk scores.