Unique function words characterize genomic proteins

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Between 2009 and 2016 the number of protein sequences from known species increased 10-fold from 8 million to 85 million. About 80% of these sequences contain at least one region recognized by the conserved domain architecture retrieval tool (CDART) as a sequence motif. Motifs provide clues to biological function but CDART often matches the same region of a protein by two or more profiles. Such synonyms complicate estimates of functional complexity. We do full-linkage clustering of redundant profiles by finding maximum disjoint cliques. Each cluster is replaced by a single representative profile to give what we term a unique function word (UFW). From 2009 to 2016, the number of sequence profiles used by CDART increased by 80%; the number of UFWs increased more slowly by 30%, indicating that the number of UFWs may be saturating. The number of sequences matched by a single UFW (sequences with single domain architectures) increased as slowly as the number of different words, whereas the number of sequences matched by a combination of two or more UFWs in sequences with multiple domain architectures (MDAs) increased at the same rate as the total number of sequences. This combinatorial arrangement of a limited number of UFWs in MDAs accounts for the genomic diversity of protein sequences. Although eukaryotes and prokaryotes use very similar sets of “words” or UFWs (57% shared), the “sentences” (MDAs) are different (1.3% shared).

protein universe | genomic sequences | functional profiles | domain architecture | shared function

The size of protein sequence space as measured by the number of combinations of the 20 natural amino acids is essentially unlimited. Not every combination of amino acids can form a protein with a unique and stable fold and only a tiny part of full sequence space is occupied by protein sequences found in nature (1). Nevertheless, the number of protein sequences in the protein universe is still considerable (2, 3). Thanks to high-throughput sequencing (4), genomic sequences have been accumulating rapidly, leaving researchers struggling to comprehend the complexity of the almost 100 million nonredundant (NR) protein sequences known. Protein sequences, the building blocks of life, have been studied intensively. For some proteins, we know their structure, biological function, specificity, and reaction kinetics, but most of the vast protein sequence universe remains unexplored. Understanding how these proteins function and interact is indispensable for deciphering the language of life.

Given that functional and structural information on the protein sequence universe is so sparse, we need computational procedures to provide functional and/or structural information for unknown sequences. A powerful method for inferring function of a protein sequence is by relating it to similar proteins thereby allowing information on unknown family members to be deduced from known members. While such sequence clustering seems straightforward, it is complicated by the huge number of sequences and the difficulty of establishing a best way to match sequences. Consider the string of amino acids in a protein as analogous to the string of letters in an English sentence. For example, the two sentences “DOG IS BAD” and “BAD IS DOG” have very similar meanings and contain the same three words, but the sequences of letters themselves are different and cannot be made to match in more than 4 of 10 positions. This analogy suggests that the use of “words” is a better way to cluster protein sequences (5).

Words can be taken as the names of sequence profiles (6), which are derived from multiple sequence alignments to encapsulate information on amino acids occurring at each position along the sequence. Sequence profiles are often associated with a particular biological function giving them a meaning in analogy to words. The most common models for sequence profiles are hidden Markov models (HMMs) (7) and position-specific scoring matrices (PSSMs) (8). HMM-based profiles are used in PFAM (9) and SMART (10); PSSM-based profiles are used in CD (11) and PROSITE (12).

Sequence profiles from the different sources are used in two main databases: InterPro (13) and conserved domain architecture retrieval tool or CDART (14). Here we focus on the CDART resource (14) at the National Center for Biotechnology Information (NCBI), a PSSM-based database that includes profiles from seven different sources and matches sequences using RPS-BLAST (15), a variant of the widely used PSI-BLAST algorithm (16). Preliminary analysis of InterPro showed it less suitable than CDART for our purposes. In CDART, all profiles are matched by RPS-BLAST to all sequences using a consistent threshold, whereas in InterPro different thresholds as reported by each data source are used to match profiles to sequences. This lack of a consistent threshold makes it impossible to cluster profiles and so deduce a minimum set of unique function words as we do here. In both cases, the region of protein sequence matched by a profile is considered to be a domain.

In this study we have two classes of objects: sequences and profiles. The sequences and profiles are those matched every month or so to one another by CDART. This gives rise to domains, which are a length of a particular sequence matched to one or more profiles. The domains are named after the profile they match and this gives structural and functional information about the sequence. The patterns of domains along the polypeptide chain is called “domain architecture” and sequences can be characterized as having no domain architecture (dark matter), single (SDA), or multiple (MDA) domain architecture. The profiles are linked by strongly overlapping on the same region of sequence enabling them to be clustered by

Significance

The vast, mostly unknown protein universe can be explored by analyzing protein sequences as a string of domains. A broader coverage can be achieved when these domains, the essential blocks in protein evolution, are detected using sequence profiles. Using clustering to collapse redundant profiles into unique function words (UFWs), we find that over the years 2009–2016, the number of UFWs saturates while the number of sequences matched by a combination of two or more UFWs grows exponentially.

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clique-based full-linkage clustering. Each cluster can be represented by the name of just one profile in it, hence our unique functional words (UFWs).

In 2009 (17) we used CDART profiles to describe the organization of the protein universe in terms of the order of the domains in N- to C-terminal direction along the amino acid chain, a pattern that is termed “the domain architecture of a protein sequence.” Our study analyzed sequences in the protein universe in terms of families that have SDAs or MDAs. It found that MDAs grew very fast with number of added sequences while SDAs seemed to have saturated. In addition, there were known structures for a quarter of the single domain families and another quarter of sequences were dark matter (no profile matches). Inspired by our work, Chub et al. (18) confirmed a decline in the rate of discovery of new protein families. Another study (19) explored the success of structural genomics efforts in providing structural coverage of the protein universe and concluded that over the last 10 y, structural coverage at a residue level increased from 30% to 40%, with half of novel structures coming from the Structural Genomics Initiative.

Here we study how CDART has changed over 8 y since 2009. With our improved clique-based clustering, all members of a cluster are similar to one another and each cluster is represented by one unique function word, avoiding confusion caused by synonyms. The number of UFWs increases more slowly than the number of profiles. As almost all UFWs occur alone in at least one sequence, the number of SDAs is close to the number of UFWs. The trends predicted in 2009 remain valid: number of different MDAs containing two or more UFWs increase with time exponentially fast, whereas the number of SDAs containing one UFW increases very slowly. We further find that eukaryotes and prokaryotes use the same “words” (the SDAs) but combine them into different “sentences” (the MDAs).

Results

Finding UFWs. We use profiles as an indicator of function, assigning to a sequence matching a profile the properties of the particular profile. We use the matches between profiles and protein sequences provided by the CDART database. Because this database includes profiles from seven different sources, a particular region of a protein sequence can be matched by several profiles. Such redundancy hinders our attempt to comprehend the complexity of all of the proteins in the known protein universe. The matches in the CDART database offer a way to eliminate the redundancy: if two different profiles match the same region of sequence, then these profiles overlap and are synonymous names for the same function.

Fig. 1 illustrates how we eliminate redundant profiles by clustering. We judge whether two profiles overlap using the lengths of the profiles, and the length of the match (only a part of the profile may match the sequence). CDART provides two parameters to assess the strength of the match, the E value (Eval) and the bit score (Bscore). We tested values of both Eval and Bscore combined with values for PDLx, the maximum percent difference in profile lengths, and Fracx, the minimum fraction of profile length matched. We use Evalx = 0.001, PDLx = 10%, and Fracx = 0.9, but other values gave similar results.

When two profiles satisfy these conditions, they overlap and are considered to be linked, forming a network with possible cliques. In graph theory, a clique is a subset of vertices such that all vertices are connected to each other. Clique finding is a hard-to-solve nondeterministic polynomial time (NP)-complete problem (21). We clustered linked profiles by full-linkage clustering using a powerful and efficient algorithm to find disjoint maximum cliques of linked profiles (20).

In 2009 there were 27,038 profiles in CDART and 795 did not match any NR sequence; in 2016 there were 48,932 profiles in CDART and 426 had no match. Thus, the number of used profiles increased 85%, from 26,243 to 48,506. Increases in the number of UFWs were smaller going from 17,072 to 24,212, an increase of 42% (SI Appendix, Table S2). For our most recent CDART dataset (2016), clustering reduces the number of different profiles from 48,506 to 24,212, a reduction of 50%. Similar reductions are seen for all eight CDART releases analyzed here (SI Appendix, Figs. S1 and Table S2).

Combinatorial Growth of MDAs. The predicted massive combinatorial growth of MDAs and very slow growth of SDAs (17) is confirmed after 7 y and with 11 times more nonredundant sequence data. Fig. 2 shows that the number of MDAs is growing rapidly with time (or added sequence data) while the number of SDAs is almost constant and seems to have saturated. Between 2009 and 2016, the number of deposited sequences increased 11-fold (from...
and more dark matter sequences, 7.8 million to 85.3 million, the number of MDAs increased 8-fold (from 88,905 to 717,727), but the number of unique SDAs increased by only 1.42-fold (from 17,072 to 24,212). While the number of MDAs grows exponentially with time, the number of SDAs remains almost unchanged (Fig. 2). Interestingly, among 24,212 unique function words, 20,241 (83.6%) are shared between both types of families, whereas 3,625 (14.9%) are seen only in SDAs and very few UFWs (341 or 1.4%) are only seen in MDAs. The corresponding values for all CDART profiles are 41,269 (80.4%), 6,496 (16.3%), and 794 (1.4%) with a total of 48,506 profiles.

**Comparing Kingdoms of Life.** We analyzed the growth of SDAs and MDAs in the three main kingdoms of life (Fig. 3). In both prokaryotes and eukaryotes, the number of MDAs is much higher than the number of SDAs and these MDAs are growing fast, while SDAs are saturating. Eukaryote sequences are 2.7 times more likely to give rise to a new MDA than are prokaryote sequences. This is offset by the fact that prokaryote sequences are being determined at a much faster rate, especially since 2014 when prokaryote sequences are growing 2.5 times faster than eukaryote sequences. Overall, this means that the numbers of added MDAs in eukaryotes and prokaryotes are similar (8% higher in eukaryotes as 2.7/2.5 is 1.08).

Both prokaryotes and eukaryotes use about three-quarters of the UFWs: 74% (17,915 of 24,212) in prokaryotes and 68% (16,558 of 24,212) in eukaryotes. Less than half of the UFWs occur in both prokaryotes and eukaryotes (40%, 11,674 of 24,212). By contrast, eukaryotes use more of the MDAs than do prokaryotes; 61% (436,273 of 717,727) of the MDAs are seen in eukaryotes, only 40% (285,302 of 717,727) are seen in prokaryotes. Just 2% of MDAs (13,290 of 717,727) are shared between prokaryotes and eukaryotes, which supports the finding that domain combinations give rise to new functions (22–25). Another reason for more MDAs in eukaryotes could be to ensure that functional domains are coexpressed in different cell types of these multicellular organisms.

Unlike prokaryotes and eukaryotes, viruses do not have more MDAs than SDAs (Fig. 3C). There are 3,223 SDAs and 3,380 MDAs with UFWs and 4,445 and 3,739, respectively, with all profiles. Both SDAs and MDAs are growing at a comparable rate. Many viruses integrate their sequences into the genome of their host for replication (26) so it is not surprising viruses would not need many MDA architectures.

**Dark Matter Is Shrinking.** We define dark matter as those sequences that do not match any profile in CDART. In our previous work (17) we suggested that one of the reasons for the existence of dark matter sequences could be that labor-intensive discovery of new sequence profiles lagged behind the increase in the number of sequences and many sequence profiles remained to be discovered in the dark matter. Our present analysis of seven additional CDART releases reveals that with the discovery of new profiles, more of the older sequence is being annotated. This is confirmed by the drop of the overall fraction of dark matter (number of dark matter sequences/number of NR sequences) from 27.5% in 2009 to 19.9% in 2016 (Fig. 2F). While the number of sequences increased 11-fold from 7.8 million to 85.2 million in these 7 y, a significant decrease in dark matter was caused by the increase of UFWs from 17,072 to 24,212 (SI Appendix, Fig. S8). The fall in dark matter from 27.5% to 19.9% of all sequences means that over 6 million sequences are no longer classified as dark matter (7.6% of 85.2 million). Our previous analysis found that the dark matter contained equal numbers of prokaryote and eukaryote sequences (951,101 and 927,211, respectively), but there were more eukaryote residues. In 2016, we find there are more prokaryote than eukaryote dark matter sequences; of 16.9 million dark matter sequences, 11.4 million are prokaryotes and 4.7 million are eukaryotes. This is in accord with the number of prokaryote and eukaryote sequences in the entire NR database (63.5 million prokaryotes and 20.4 million eukaryotes). Despite having more sequences—and more dark matter sequences—fewer of the prokaryote sequences do not match a recognized profile (17.2%) than for the eukaryote sequences (20.4%). The majority of the virus sequences (93%) match a known sequence profile; their dark matter fraction is just 7%. Two recent studies of half a million Swiss-Prot sequences report similar results, with slightly higher fractions of eukaryotic sequences in the dark proteome (27, 28).

**Structural Coverage Is Increasing.** In 2009 we made a wild extrapolation concerning possible scenarios for the structural coverage of sequence families in 2050. Today with the benefit of many more data we can confirm some of those predictions. The NR sequence database has continued growing exponentially, doubling its size every 25 mo. Back in 2009, we presented two scenarios for the structural coverage: (i) Assuming continuous investment but no improvement in either experimental or computational methods, we predicted that the coverage by 2050 would reach 70%. This seems to be the right track (SI Appendix, Fig. S4, cyan line) with 26% of the families containing at least one known structure
in the Protein Data Bank (PDB) by July 2016. (ii) We also predicted that the structural coverage for structures not solved by the Protein Structural Genomics Initiative, referred to here as non-SG structures, would fall. We were unduly pessimistic. As shown in SI Appendix, Fig. S3, nonstructural genomics projects (no-SG) provided 65% of the 4,414 unique SDA structures solved during the last 6 y, whereas structural genomic (SG) projects provided just 35% of the unique SDA structures (SI Appendix, Fig. S3, yellow line). Although recent growth in unique coverage still benefits from structural genomics programs, the fraction of non-SG structures is rising faster than the SG structures (SI Appendix, Fig. S3, brown line). Lack of dependence of structural coverage on structural genomics programs is welcome as the NIH has phased out the program.

Common SDAs and Unique MDAs. While most SDAs are found in both prokaryotes and eukaryotes, the situation for MDAs is very different as clearly seen in the overlapping area of the Venn diagrams in Fig. 4. In 2016, a large fraction of SDAs (57% all profiles, 50% UFWs) are shared by eukaryotes and prokaryotes, while only a small percentage of MDAs (1.3% all profiles, 2.1% UFWs) are common to both kingdoms. This is not surprising since one would expect the more complex multidomain proteins to have more sophisticated functions and hence be more specific to a certain organism. As more sequences are identified, we expect that the overlap of SDAs and MDAs between the three kingdoms of life will maintain the trends seen in Fig. 4 and SI Appendix, Fig. S5. Specifically, we expect to see almost all of the SDAs shared between prokaryotes and eukaryotes. Interestingly, viruses show the same behavior as prokaryotes and eukaryotes: the majority (77%) of their SDAs overlap with SDAs in either prokaryotes, eukaryotes, or both, but much fewer (23%) of their MDAs are seen in other kingdoms.

Databases of Sequence Profiles. To keep up with the rapid growth in the number of protein sequences, frequent updates of sequence profile databases are essential. We analyzed how the profiles and unique profile datasets have changed with time (SI Appendix, Tables S6 and S7). The four main sources of profiles in CDART are PFAM, PRK, CD, and COG. In 2009, most of the profiles in CDART came from PFAM (32.4%). This percentage has remained almost unchanged (33.1% in 2016). The second most common source of profiles in CDART has been PRK, and its percentage has increased from 20.4% in 2009 to 23.3% in 2016. COG profiles represented 15.3% of CDART profiles in 2009 but dropped to 9.9% in 2016. The percentage of profiles coming from CD has increased from 12.5% in 2009 to 23.3% in 2016. We see similar trends when considering UFWs, but the contribution of PFAM is even more dominant. In 2009, 53.5% of the UFWs came from PFAM, and this percentage has remained almost unchanged at 54.1% in 2016. The changes for PRK, COG, and CD profiles are from 14.9% to 9.3%, from 18.4% to 10.1%, and from 8.4 to 10.2%, respectively.

Among all different profiles in CDART, PFAM and CD profiles seem to be updated more frequently than the others. Although the number of SMART profiles almost doubled since 2009, the SMART database seems to be updated less frequently (once a year) than the PFAM and CD databases. Note, the PSSM models we use are less well suited than HMMs to patterns containing insertions or deletions, of variable length, and with positional dependencies.

Our Clustering to Get UFWs Is Stable. In all eight independent analyses, clustering and the use of UFWs reduced the number of SDAs by approximately half but had a smaller (30%) effect on the number of MDAs probably because of the specific combinatorics (SI Appendix, Fig. S1). Puzzled by this difference, we simulated randomly sampled MDAs using the UFWs in our data. SI Appendix, Fig. S10 shows that simulated domain architecture plots are very similar to those of the real data and confirm that merging profiles into UFWs has a stronger effect on MDAs.

Fig. 3. Comparing growth of families in three kingdoms of life. The growth of SDA (squares) and MDA (triangles) families in prokaryotes (pink) and eukaryotes (green) shows the same trends seen for all species combined (Fig. 2). (A) MDAs are growing exponentially with time, whereas SDAs are saturating. In 2017, most of the MDAs are from eukaryotes (436,273 or 60.5% compared to 285,302 or 39.5% for prokaryotes). This is due to the faster growth of MDAs in eukaryotes during the last 2 y. The fractional coverage (defined as 1 – number dark matter sequences/number of sequences) is slightly higher in prokaryotes (82.8%) than in eukaryotes (79.5%). The majority of the virus sequences (92.9% or 1,041,468 of 1,126,166) have at least one match to a known profile so that the fractional coverage is very high at 93%, leaving just 7% dark matter. (B) The data for SDAs with an expanded y scale. About the same number of SDAs (18,514 or 76.4%) are seen in prokaryotes and in eukaryotes (17,287 or 71.4%). The corresponding values for all sequence profiles (dashed lines) are 560,577 or 59% and 390,640 or 41% for MDAs and 38,054 or 78.5% and 36,791 or 75.8% for SDAs. Saturation of SDAs in the three kingdoms is more evident when using unique profiles (solid lines) than original CDART profiles (dashed lines). Viruses (blue) have more SDA than MDAs, but this is difficult to see as both types of families occur much less often than in the other kingdoms of life. They are better seen in C with expanded y axis scale. In all panels, solid filled lines were obtained using unique CDART profiles, whereas dashed lines use all CDART profiles. E, eukaryotes; P, prokaryotes; and V, viruses.
Extrapolating the Sequence Growth. We attempted to answer the simple but important questions: What is the size of the protein universe? How many UFWs exist? How many unique sentences can be formed with these UFWs? We extrapolated the slowly growing number of SDAs as well as the exponentially growing number of sequences, all-length MDAs, and two-word MDAs over the next 50 y (Fig. 5). Assuming unchanged data trends, our data suggest that by 2066, the protein universe will contain 10\(^{15}\) nonredundant sequences. We estimate there will be between 30,000 and 50,000 SDAs, which combine to form 3 × 10\(^{12}\) MDAs (3 × 10\(^{11}\) of these composed by two SDAs).

Discussion

We characterized the protein universe—the collection of all proteins of every biological species that lives or has lived on earth—by classifying 85 million known sequences into families based on their having the same domain architectures. The two main types of domain architectures, SDAs and MDAs, are the main components of the language of life. Proteins speak using words (UFWs or SDAs) linked in an ordered way into sentences (MDAs). All 85 million NR sequences are characterized by just 24,212 UFWs and short sentences are made by combining these words. The number of UFWs in the language of proteins is saturating, and it looks like there are very few to be discovered. Structural coverage of the existing UFWs is high (27% of the UFWs have structures) and increasing rapidly (about 1% per year), thanks to equal contributions from both structural genomic projects and conventional crystallography. However, the number of sentences, which are mainly different combinations of words, is growing linearly with the number of sequences, but it too is expected to saturate. Although initial estimates of the protein universe size were much smaller (29, 30), fewer data were available.

These findings are consistent with the concept of protein evolution proceeding in large part by the creation of new proteins by new arrangements of existing protein domains to form novel multidomain proteins (25, 31). One can estimate the age of a domain family by finding the largest group of organisms within it (32). Different kingdoms of life share the same vocabulary, but use different sentences. Most words are used in both prokaryotes and eukaryotes, and this commonality increases with time, or number of sequences added. However, sentences made by combining words are much more organism specific and are rarely shared, with only 1.3% of the sentences used both in prokaryotes and eukaryotes. Sharing drops from 57% to 1.3% in going from SDAs to MDAs, which supports the finding that domain combinations give rise to new function (33–36). Remarkably, viruses share the majority (93%) of their words with either prokaryotes, eukaryotes, or both, but far fewer (7%) of their sentences are seen in other kingdoms. Viral proteins have a comparable number of single and multiple domain architectures. Many viruses integrate their genome into the host cell, which exempts them from carrying refined MDA machinery. On the other hand, for completing a successful
infection cycle, viruses must cope with the cell machinery for entry, replication, and translation while hiding from the host immune system so they still need some MDAs (37).

The dark matter of the protein universe are those sequences that do not have any match to a known sequence profile, or, following the language analogy, silent proteins. Although the absolute size of the dark matter in the protein universe has been growing with the number of deposited sequences in NR, its relative size is slowly decreasing. CDART and NR databases are well maintained and updated. Due to the admirable computational coverage of the protein sequence space, the unexplored regions of the protein sequence universe is slowly but consistently being reduced in its relative size. Although the protein universe has been increasing its size at a very fast pace, we still are orders of magnitude away from covering the entire feasible sequence space. Our insights about the protein universe are based on the sample of organisms that have been sequenced and we do not know how well this subset represents the protein universe. Still, the steady trends we have seen during a period when data increased 11-fold lend support to the views suggested here.

Conclusions

The functional language of proteins uses few words (fewer than 25,000 UFWs) and new words are added very slowly. Complexity of protein sequences comes for sentences made with these words. Different kingdoms of life use the same words but in different sentences. For UFWs, new sentences are being discovered rapidly with one per 196 added NR sequences in eukaryotes and one per 312 added NR sequences in prokaryotes. Corresponding values for all profiles are 156 and 227, respectively. Although our conclusions are based on the development of 7 y of protein sequencing data, we attempted to extrapolate these findings to millions of years of evolution across the entire protein repertoire; this could benefit from targeting sequencing efforts toward more diverse species.

Materials and Methods

We downloaded from ftp://ftp.ncbi.nih.gov/pub/mmdb/cdart/ the CDART data and from ftp://ftp.ncbi.nlm.nih.gov/blast/db/FASTA/nr.gz the NR sequences. PDB entries solved by structural genomics were downloaded from targetdb.rcsb.org/target_files and taxonomy from ftp://ftp.ncbi.nih.gov/pub/taxonomy/gi_taxid_prot.dmp.gz. We did our analysis on: (i) February 17, 2009; (ii) June 16, 2010; (iii) June 15, 2011; (iv) September 24, 2012; (v) January 9, 2013; (vi) January 13, 2014; (vii) January 1, 2015, and (viii) July 3, 2016. Each analysis used a different version of NR, CDART, PDB, and taxonomy files, downloaded on that day.

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