STARCH DEGRADATION AND NUTRITION VALUE IMPROVEMENT IN CORN GRITS BY SOLID STATE FERMENTATION TECHNIQUE WITH CORIOLUS VERSICOLOR

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Submitted: November 29, 2009; Returned to authors for corrections: April 15, 2011; Approved: June 06, 2011.

ABSTRACT

The study was conducted to evaluate effect of Coriolus versicolor mycelia on degrading starch and improving nutrition value in corn grits through solid state fermentation technique. The results showed that using soybean meal as a nitrogen source, α-amylase secreted from C. versicolor expressed 407.25U/g of activity, leading to 45.15% of starch degraded. The activity grew with fermentation time until the 15th day, after that the amylase was deactivated rapidly. An orthogonal experiment designed for the study illustrated that degradation rate of starch in corn grits attained to maximum, 50.51%, when 100g of corn grits, added 16g of soybean meal, were fermented by C. versicolor for 12 days, in an initial pH 5.5. After fermenting, compared to the nonfermented control, contents of amino acids, total sugar, crude fat and crude protein were increased by 21.00%, 38.45%, 55.56%, 69.15% respectively. The significant improvement of nutrition value in corn grits is probably attributed to the intense metabolism of C. versicolor.

Key words: Coriolus versicolor, corn grits, starch, solid state fermentation, α-amylase

INTRODUCTION

Coriolus versicolor is a distinguished medicinal mushroom, owning functions on anti-age, immunoregulation (13), treating multiple hepatitis (4), inhibiting diverse cancer cells proliferation (7, 8, 10). It also has a certain efficacy on decreasing toxic actions and side effects from chemotherapeutic drugs, and treating AIDS (15, 16). At present, C. versicolor polysaccharide is used widely to cure hepatitis, and glycoprotein from its mycelia has been employed as an anticancer drug in Japan for decades. Besides, cases on turning it into food addictives and health-caring medicine are reported in domestic and abroad. But they are mainly operated by submerged liquid fermentation (SLF). Studying C. versicolor by solid state fermentation (SSF) has not been reported.

Compared with SLF, SSF expresses its advantages on simpler culture condition, better product properties, less capital investment, environmental pollution (14). It is commonly employed for production of multiple enzymes from microorganisms (1, 19), but seldom for improvement of food nutrition value.

Corn grits, being able to prevent hypertension and heart disease by micronutrients in them, show certain nutritional
advantages in cereal. But the high starch content could lead to a low absorption (18). *C. versicolor*, as a species of rot fungi, has a strong ability to decompose and reuse carbohydrate. Thus, we considered that nutrition of corn grits may be changed by cultivating *C. versicolor* mycelia in corn grits medium.

We describe here a series of experiments on SSF by *C. versicolor* in corn grits medium. We also analyze the activity of α-amylase secreted by *C. versicolor* and main nutritional components in the fermented product and the nonfermented control.

**MATERIALS AND METHODS**

**Fungus**

The strain Y 1 of *C. versicolor* was preserved in Mushroom Culture Collection, Department of Bioengineering, College of Life Science, South China Normal University. It was fostered in a PDA medium with the initial pH 6.5 and the cultivation temperature was in a range between 25 and 28 degrees centigrade.

**Solid state fermentation**

The basic medium included 100g dry corn grits which were moistened with 40mL of 1mg/mL KH₂PO₄ buffer (pH 6.5). The humidity of the medium was about 40%. Addition of different nitrogen source was different: the dry soybean meal was added 10g in 100g corn grits, tryptone 2g, NaNO₃ 2g, and (NH₄)₂SO₄ 2g. The solid medium was dispensed into erlenmeyer flasks (250mL capacity). After sterilization, each flask was inoculated with 10mL mycelia pellets liquid, and incubated at 25 degrees centigrade in the dark. The entire content of a flask was harvested after fermented some days, for analyzing α-amylase activity and the main nutritional components. Three replicates were prepared for each treatment. The control was non-inoculated.

**Enzyme extraction and assay**

Two grams of fermented product from a flask was mixed with deionized water and ground in an ice-cold mortar. After centrifuging the homogenate at 3000r/minute for 10 minutes, the supernatant was assayed for α-amylase (α-1,4-glucan-4-glucanohydrolase, EC 3.2.1.1) activity, using soluble starch as the substrate. A mixture of 1mL of the supernatant, 1mL of citric acid buffer (pH 5.6), and 2mL of 10g/L soluble starch was incubated at 40 degrees centigrade for 5 minutes. Then the reducing sugar released was determined by the method described by Chen (2), referring to a standard curve of maltose. One α-amylase unit (U) was defined as the amount of enzyme producing 1µmol maltose per minute at 40 degrees centigrade from soluble starch.

**Main nutritional components assay**

The samples were dried to constant weight at 75 degrees centigrade before main nutritional components assay. Starch content of the dried samples was determined by titration with potassium ferricyanide (9). The starch degradation rate was calculated according to the formula as follows:

\[
\text{Starch degradation rate (\%)} = \frac{(\text{starch content of control} - \text{starch content of fermented sample})}{\text{starch content of control}} \times 100
\]

Total sugar content was determined by phenol-sulphuric acid method (22), and crude fat content was by soxhlet extraction (9). Crude protein content was calculated from nitrogen content, determined by vario EL element analyzer (produced by Elementar Company, Germany). An amino acid analyzer (Hitachi Model 835-50) was used for amino acid determination.

Duncan’s multiple range test (3) was utilized to test significant differences among data of the treatment groups and the control at the 1% level of confidence.

**RESULTS**

**Effect of species of nitrogen source**

A suitable nitrogen source was chosen necessarily, because of its effect on α-amylase degrading starch (17).
According to Table 1, α-amylase of *C. versicolor* was more active in the media added organic nitrogen sources than control and inorganic ones. Specifically, the activity in the medium added soybean meal was superior to added tryptone. Because of that, the starch degradation rate was achieved 45.15%.

**Table 1.** Effect of species of nitrogen source on activity of α-amylase secreted by *C. versicolor* and degradation rate of starch in corn grits degrated by the α-amylase

| Nitrogen Source | α-amylase activity (U/g) | Degradation rate of starch (%) |
|-----------------|--------------------------|-------------------------------|
| Soybean meal    | 407.25±45.74A<sup>b</sup> | 45.15±0.08A                   |
| Tryptone        | 290.06±26.41B            | 44.84±0.02B                   |
| NaNO₃           | 163.10±11.77C            | 44.71±0.19B                   |
| (NH₄)₂SO₄       | 149.67±7.62C             | 44.13±0.06C                   |
| Control         | 143.56±13.20C            | 44.76±0.03B                   |

<sup>a</sup> One α-amylase unit (U) was defined as the amount of enzyme producing 1µmol maltose per minute at 40 degrees centigrade from soluble starch.

<sup>b</sup> Data in the same column followed by the same letter were not significantly different at the p<0.01 level according to Duncan’s multiple range test.

**Effect of fermentation time**

The α-amylase activity and starch content were determined after fermented 5, 10, 15, 20 and 25 days, respectively. Results (Table 2) showed that α-amylase activity was maximum at the 10<sup>th</sup> day, but not significantly different to the 15<sup>th</sup> day at the p<0.01 level. Starch degradation rate rose drastically with fermentation time in the first 15 days. But after that, the amylase deactivated rapidly, and the rate rose gently. It suggested that an efficient fermentation time was between or around 10 and 15 days.

**Table 2.** Effect of fermentation time on activity of α-amylase secreted by *C. versicolor* and degradation rate of starch in corn grits degrated by the α-amylase

| Fermentation time (day) | α-amylase activity (U/g) | Degradation rate of starch (%) |
|-------------------------|--------------------------|-------------------------------|
| 5                       | 339.97±6.00BC            | 45.32±0.06D                   |
| 10                      | 427.75±11.96A            | 46.18±0.08C                   |
| 15                      | 413.77±52.61AB           | 46.91±0.38B                   |
| 20                      | 332.99±43.11BC           | 47.31±0.10AB                  |
| 25                      | 259.35±15.41C            | 47.61±0.38A                   |

**Identification of optimum SSF condition**

An orthogonal experiment with 3 factors and 3 levels L₉(3³) was designed for studying connection among a number of factors and the influence degree of each factor systematically. According to Table 3, the theoretical optimum SSF condition was as following: 100g corn grits supplemented with 16g soybean meal, a fermentation time of 12 days, an initial pH of 5.5. According to range analysis, acting degree of three elements on *C. versicolor* degrading starch was different: soybean meal content > fermentation time > initial pH. At the p<0.01 level, soybean meal content was an extremely remarkable factor; at the p<0.05 level, fermentation time was a significant one. Orthogonal experiment No.8 was identical to the theoretical optimum SSF condition, and its degradation rate of starch reached maximum, 50.51%.
Table 3. Result of orthogonal experiment to identify optimum SSF condition by *C. versicolor*

| Experimental No. | Soybean meal (g/100g corn grits) | Fermentation time (day) | Initial pH | Degradation rate of starch (%) |
|------------------|----------------------------------|-------------------------|------------|-------------------------------|
| 1                | 4                                | 7                       | 5.5        | 47.82±0.10                    |
| 2                | 4                                | 12                      | 6.5        | 48.78±0.34                    |
| 3                | 4                                | 17                      | 7.5        | 48.32±0.13                    |
| 4                | 10                               | 7                       | 6.5        | 48.00±0.18                    |
| 5                | 10                               | 12                      | 7.5        | 49.30±0.67                    |
| 6                | 10                               | 17                      | 5.5        | 49.35±0.28                    |
| 7                | 16                               | 7                       | 7.5        | 49.14±0.14                    |
| 8                | 16                               | 12                      | 5.5        | 50.51±0.37                    |
| 9                | 16                               | 17                      | 6.5        | 49.81±0.25                    |
| R                | 1.513                            | 1.210                   | 0.364      |                               |

Effect on contents of amino acids in the protein

Fermented under the condition of orthogonal experiment No.8, each amino acid in the fermented product had an increase in quantity (Table 4). Specifically, glycine and serine were increased by over 50%. Quantity of total essential amino acids increased from 37.79 to 46.07mg/g; total amino acids from 96.79 to 117.12mg/g. But the ratios of essential amino acids to total amino acids in the product and the control were approximately the same. It indicated that SSF by *C. versicolor* might enhance contents of amino acids on the premise of maintaining the protein composition of corn grits.

Table 4. Amino acids contents of the fermented product and the nonfermented control (mg/g sample)

| Amino acid | Nonfermented | Fermented | Growth rate of amino acid (%) \(^a\) |
|------------|--------------|-----------|-----------------------------------|
| Asp        | 7.55         | 10.00     | 32.45                             |
| Thr        | 2.91         | 4.28      | 47.08                             |
| Ser        | 2.94         | 4.69      | 59.52                             |
| Glu        | 19.64        | 20.99     | 6.87                              |
| Pro        | 8.94         | 9.93      | 11.07                             |
| Gly        | 3.23         | 5.37      | 66.25                             |
| Ala        | 7.29         | 8.99      | 23.32                             |
| Cys        | 1.25         | 1.40      | 12.00                             |
| Val        | 5.63         | 7.97      | 41.56                             |
| Met        | 2.73         | 3.09      | 13.19                             |
| Ile        | 4.69         | 5.95      | 26.87                             |
| Leu        | 13.05        | 14.28     | 9.43                              |
| Tyr        | 1.72         | 2.12      | 23.26                             |
| Phe        | 5.75         | 7.13      | 24.00                             |
| Lys        | 3.03         | 3.37      | 11.22                             |
| His        | 2.33         | 2.62      | 12.45                             |
| Arg        | 4.11         | 4.94      | 20.19                             |
| EAA \(^b\) | 37.79        | 46.07     | 21.91                             |
| Total \(^c\) | 96.79    | 117.12    | 21.00                             |
| E/T \(^d\)  | 0.3904      | 0.3934    |                                   |

\(^a\) Growth rate of amino acid(%)=(amino acid content of the fermented product—amino acid content of the nonfermented control)/amino acid content of the nonfermented control×100

\(^b\) Quantity of total essential amino acids.

\(^c\) Quantity of total amino acids.

\(^d\) Ratio of essential amino acids to total amino acids.
Effect on contents of main nutritional components

Contents of main nutritional components in the fermented product (fermented according to orthogonal Experiment No.8) and the nonfermented control were compared in Table 5. After fermenting, total sugar, including starch and reducing sugar, rose from 54.1 to 74.9g in 100g substrate, even starch had been decreased by 50.51% (Table 3). The increase rates of crude fat and crude protein were also noticeable, reaching 55.56% and 69.15% respectively.

Table 5. Contents of main nutritional components of the fermented product and the nonfermented control

| Components   | Nonfermented (g/100g substrate) | Fermented (g/100g substrate) | Growth rate (%) |
|--------------|----------------------------------|------------------------------|-----------------|
| Total sugar  | 54.1                             | 74.9                         | 38.45           |
| Crude fat    | 2.7                              | 4.2                          | 55.56           |
| Crude protein| 9.4                              | 15.9                         | 69.15           |

*Growth rate(%)=(a certain component content of the fermented product— the component content of the nonfermented control)/the component content of the nonfermented control×100

DISCUSSION

Through SSF with C. versicolor, starch content of corn grits was reduced strikingly. It was probably partly because of using soybean meal as a nitrogen source. Soybean contained a certain deal of calcium ion which could activate and stabilize α-amylase (11, 20). It resulted in a better starch degradation than others. Besides, content of main nutritional components were increased dramatically. Particularly for the increase of protein content, after fermenting, the protein content exceeded several cereals, such as barley grain (10.2g/100g substrate), buckwheat flour (11.3), oat flour (13.7), wheat flour (15.7) (20). It could be involved in the metabolism of C. versicolor: starch of corn grits was degraded into maltose by catalysis of the α-amylase; then other amylases converted maltose into fungi polysaccharides, fat and/or carbon skeleton of amino acids (12). The starch degradation and the nutrition value improvement could lead to a great enhancement of digestibility and absorption rate (18).

Similarly, Hericium erinaceum, Ganaderma lucidum and Morchella esculenta have also been investigated the function on enhancing nutrition value of cornmeal by SSF. Both of them expressed similar results to C. versicolor (5, 6, 21). We consider that here the product by SSF is a nutrient combination of corn grits and C. versicolor. Cooperation between SSF and medicinal fungi may turn corn grits and other agricultural products into some kinds of new food owning new health-caring functions. As human being needs to upgrade nutrition value of food for better survive, it can be foreseen that researchers will increasingly focus on this topic and its further studies. In this case, a further investigation on health-caring functions of the fermented product is needed and its importance can readily been seen.

ACKNOWLEDGEMENTS

This work was supported by Guangzhou Science and Technology Key Project.

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