Effect of Epigallocatechin Gallate on Viability of Kudoa septempunctata

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Abstract: Kudoa septempunctata have been reported as a causative agent for acute transient gastrointestinal troubles after eating raw olive flounder (Paralichthys olivaceus). It raised public health concerns and quarantine control in several countries. Quantitative evaluation on viability of K. septempunctata is crucial to develop effective chemotherapeutics against it. A cytometry using fluorescent stains was employed to assess effect of three compounds on viability of K. septempunctata. Epigallocatechin gallate reduced markedly viability of K. septempunctata at 0.5 mM or more, and damaged K. septempunctata spores by producing cracks.

Key words: Kudoa septempunctata, olive flounder, flow cytometry, epigallocatechin gallate

Myxozoa (Cnidaria) include more than 2,100 species, most of which are coelozoic or histozoic parasites in fish [1]. Although most myxozoans do not cause a serious problem in fish, several species Enteroymyxus leei, E. scophthalmi, and Myxobolus cerebralis cause mass mortality [2-4], and Kudoa septempunctata, K. iwatai, K. hexapunctata, and Unicapsula seriola have been reported to cause fish-born illness in Korea and Japan [5-8]. In particular, K. septempunctata has been reported in olive flounder Paralichthys olivaceus, which is the most reared fish species in Korea. This species provoked a conflict in fish trade and economic loss in fish farming [9].

Based on the differences in fluorescent stains and morphological changes of the myxozoa, several studies have been conducted to find chemotherapeutic agents that affect the viability of K. septempunctata [10-12]. However, the quantification and determination of accurate effective doses of chemicals based on microscopic observation can lead to inconsistency and misinterpretations, particularly due to subjective visual criteria. As an alternative, flow cytometry allows rapid, accurate, and quantitative analysis on cell morphology and viability. Flow cytometry has been applied to determine viability of the protozoans such as Cryptosporidium parvum and Perkinsus marinus [13,14]. However, any assay on the viability of the myxozoans was performed using flow cytometry. Aim of present study is to assay on effect of three chemicals compounds to the viability of K. septempunctata spores using a simple and objective method of flow cytometry.

Olive flounders (n = 20) were obtained from an olive flounder farm that suffered annually from K. septempunctata infection. Squash preparations of the fish samples were examined microscopically to detect K. septempunctata spores. The spores were purified by a modified Percoll (Sigma-Aldrich, St. Louis, Missouri, USA) density-gradient centrifugation method, as previously described [15]. The K. septempunctata spores were exposed to three kinds of chemicals including amprolium hydrochloride (AH; Dr. Ehrenstorfer, Augsburg, Germany), chlorogenic acid (CA; Carl Roth, Karlsruhe, Germany), and epigallocatechin gallate (EGCG; Sigma-Aldrich, St. Louis, Missouri, USA), which have been reported to be effective against the myxosporidia [11,12]. Fifty microliters of chemicals (1 and 10 mM) were added to 450 µl of phosphate-buffered saline (PBS) containing 3.2 × 10³ spores, and the mixtures were incubated at room temperature (approximately 20-25°C) for 30 min. The spores were then stained with 50 µl of 20 µg/ml propidium iodide (PI) for 10 min. The proportion of dead and live spores was calculated to estimate viability based on 3,000 spores counted by the red fluorescence detector of the flow cytometer (CytoFlex, Beckman Coulter, Brea, California, USA).

To determine the criteria of viability of K. septempunctata, the spores (8.5 × 10³ spores/500 µl) were exposed to PBS, 1% for-
malin, and 0.01 and 0.1 mM final concentrations of EGCG. It determined that the PBS exposed group 100% viability while the 1% formalin exposed group has 0% viability. We investigated the effect of EGCG at concentrations, ranging from 0.01 to 1 mM. The flow cytometry assay was performed with 1.7 × 10^4 spores/500 µl exposed to 0.01, 0.05, 0.1, and 1 mM final concentrations of EGCG, and PBS. For scanning electron microscopy (SEM), the purified spores were divided into 3 groups that exposed to 0.1 and 1 mM EGCG, and PBS for 30 min, respectively. The spores were immediately rinsed twice with PBS and fixed in 2% glutaraldehyde in 0.1 M phosphate buffer (pH 7.4) at room temperature for 2 hr. The spores were post-fixed in 1% osmium tetroxide in the same buffer and dehydrated through 50, 70, 90, 95, and 100% ethanol each for 10 min. The spores were dried in an atmosphere and then dried using a CO₂ critical point dryer. Finally, the spores were sputter coated with platinum, and observed using a JSM-6700F SEM at 10 kV.

Since PI is permeable only through the membrane of dead K. septempunctata spores, intensity of the stain was counted as dead spores. The spores exposed to 0.1 and 1 mM of CA and AM were unstained with PI since there was no damage in the membrane (Fig 1A). On the other hand, the spores treated with 0.1 and 1 mM EGCG showed strong red fluorescence, indicating that the spores lost membrane integrity and were stained with PI (Fig 1A). The efficacy of EGCG was found at 0.1 mM, but not at 0.01 mM. The finding/efficacy was similar to that of 1% formalin (Fig. 1B). We observed that more than 90% of the spores were dead in EGCG concentrations ≥ 0.5 mM. The effect of EGCG on the viability of K. septempunctata was dependent on EGCG concentration (Fig. 2A-E). The evaluation of viability using flow cytometry was completed under 5 min.

It was reported several chemicals affecting the viability of K. septempunctata using approximately 100 spores with two kinds of fluorescent stains to identify the dead/live spores [11,12]. In our study, the viability of 5,000 spores was determined using one fluorescent stain. Ahn et al. [11] reported that AH had an anti-Kudoa effect at higher concentrations ≥ 1 mM and at longer treatment time (48 hr). The candidate chemicals should have anti-Kudoa effect at lower concentrations and in a shorter time since it will be used in fish farms. Ohnishi et al. [12] reported the anti-Kudoa effect of extract of coffee and tea, caffeine, and EGCG. The caffeine did not show any effect at 1 mM concentration, whereas 100% of K. septempunctata spores were inactivated at 0.1 mM EGCG. They proposed that the EGCG is an excellent candidate for the treatment and prevention of K. sep-

![Fig. 1. Assessment of viability of Kudoa septempunctata myxospores using flow cytometry and propidium iodide staining. (A) The myxospores were treated with PBS, and 0.1 and 1 mM of amprolium hydrochloride (AH), chlorogenic acid (CA), and epigallocatechin gallate (EGCG). (B) The myxospores were treated with PBS, 1% formalin, and 0.01 and 0.1 mM EGCG.](image-url)
Fig. 2. Assessment of viability of *K. septempunctata* myxospores depends on the concentration of epigallocatechin gallate. Myxospores were treated with (A) phosphate-buffered saline, and (B) 0.01, (C) 0.05, (D) 0.1, and (E) 1 mM epigallocatechin gallate.

Fig. 3. Scanning electron micrographs of the spores of *K. septempunctata* treated with epigallocatechin gallate. (A) The spores were treated with phosphate buffered saline, (B) 0.1 EGCG, and (C) 1 mM EGCG. (D-F) Magnification of the square regions. Arrow indicates cracks on the shell valve. Scale bar = 1 μm.
present in the spore and plasmodial membrane of *Kudoa* species [18,19]. In addition, the carbohydrate residues are participatory in the host-parasite relationships and immune responses in myxosporeans [19,20]. It was speculated that EGCG binds to carbohydrate or glycosylated components of the shell valve and damages integrity of the spores. However, further studies are needed to determine the type of cell wall constituents that EGCG binds to and the mechanism of crack formation on the shell valve of *K. septempunctata*.

Application of herbal remedies including EGCG were reported to control disease in the aquaculture, which had low toxicity and minimal environmental impacts [21]. EGCG inhibited grass carp reovirus, white spot syndrome virus, and *Vibrio alginolyticus* infection in fish, crab, and shrimp [22-24]. However, EGCG to crab caused the hemocyte damages and the administration of EGCG into mouse peritoneum produced severe hepatotoxicity [23,25]. We speculate that the sharp increase of EGCG concentration in plasma might cause adverse effects. It is suggested to administer EGCG to fish by adding to aquaculture feed.

In conclusion, EGCG markedly decreased viability of *K. septempunctata* spores and affected integrity of the shell valves of spores. This finding will contribute to the development of chemotherapeutic agents against the infection of *K. septempunctata* on olive flounders. In addition, flow cytometry could be used to evaluate and quantify the efficacy of chemicals on other myxosporeans. Further studies need to determine an optimal mixing ratio of EGCG to the feed by identifying pharmacokinetics and toxicity of EGCG after oral administering to fish.

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**CONFLICT OF INTEREST**

The authors declare no conflict of interest related to this study.

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