Electrocatalysis of Lindane Using Antimony Oxide Nanoparticles Based-SWCNT/PANI Nanocomposites

Kgotla K. Masibi1,2, Omolola E. Fayemi1,2, Abolanle S. Adekunle1,2,3, El-Sayed M. Sherif4,5 and Eno E. Ebenso1,2*

1 Department of Chemistry, Faculty of Natural and Agricultural Sciences, North-West University, Mmabatho, South Africa, 2 Material Science Innovation and Modelling Research Focus Area, Faculty of Natural and Agricultural Sciences, North-West University, Mmabatho, South Africa, 3 Department of Chemistry, Obafemi Awolowo University, Ile-Ife, Nigeria, 4 Center of Excellence for Research in Engineering Materials, King Saud University, Al-Riyadh, Saudi Arabia, 5 Electrochemistry and Corrosion Laboratory, Department of Physical Chemistry, National Research Centre, Cairo, Egypt

This work describes the chemical synthesis of antimony oxide nanoparticles (AONPs), polyaniline (PANI), acid functionalized single-walled carbon nanotubes (fSWCNTs), and the nanocomposite (AONP-PANI-SWCNT) as catalyst for the trace detection of lindane. Successful synthesis of the nanomaterials was confirmed by Fourier transform infrared (FT-IR) spectroscopy, ultraviolet-visible (UV-Vis) spectroscopy, x-ray diffraction (XRD) spectroscopy, and scanning electron microscopy (SEM). Cyclic voltammetry (CV) and electrochemical impedance spectroscopy (EIS) were used for investigating the electrochemical behavior of the modified electrodes in the ferrocyanide/ferricyanide ([Fe(CN)6]4−/[Fe(CN)6]3−) redox probe. GCE-AONP-PANI-SWCNT exhibited faster electron transport properties as well as higher electroactivity as compared to bare-GCE, GCE-AONPs, GCE-PANI, and GCE-SWCNT electrodes. Electrocatalytic studies further showed that GCE-AONP-PANI-SWCNT modified electrode was stable (after 20 scans) with only a small current drop in lindane (0.57%). The GCE-AONP-PANI-SWCNT electrode with low detection limit of 2.01 nM performed better toward the detection of lindane as compared to other studies in literature. The GCE-AONP-PANI-SWCNT electrode is highly selective toward the detection of lindane in the presence of various organic and inorganic interfering species. Real sample analysis of river water and tap water samples using the developed sensor gave satisfactory percentage recoveries therefore confirming the potential of the proposed sensor for practical application.

Keywords: polyaniline, single-walled carbon nanotubes, antimony oxides nanoparticles, lindane, square-wave voltammetry, cyclic voltammetry

INTRODUCTION

Organochlorine pesticides (OCPs) are chlorinated hydrocarbons emanating from persistent organic pollutants (POPs) used extensively from early 1900 in agriculture and mosquito control. Like POPs they accumulate in the environment and are very persistent (Jones and de Voogt, 1999; Chrysikou et al., 2008; El-Shahawi et al., 2010; Van Dyk and Pletschke, 2011; Salihovic et al., 2012; Gill et al., 2013). Examples of these POPs are
dichlorodiphenyltrichloroethane, dieldrin, mirex, kepone, lindane, alachlor, methoxychlor, endosulfan, chlordane, dicofol, heptachlor, toxaphene, and benzene hexachloride (Jones and de Voogt, 1999; Abdollahi et al., 2004; Chrysikou et al., 2008; El-Shahawi et al., 2010; Van Dyk and Pletschke, 2011). Lindane (γ-hexachlorocyclohexane (γ-HCH)) is an organochlorine pesticide with properties such as molecular formula C₈H₈Cl₆, molecular weight of 290.85 g/mol, and melting point of 112–113°C. Physically, lindane appears as a colorless-to-white powder with a musty odor (Abdollahi et al., 2004; Prathap and Srivastava, 2013; Anirudhan and Alexander, 2015). Lindane was globally banned for agricultural use, however the pesticide is still widely used in the production of shampoos, lotions to treat head lice, and scabies (Blair et al., 1998; Prathap and Srivastava, 2013). The accumulation of lindane has been detected globally in human breast milk, human blood, and adipose tissue (Prathap and Srivastava, 2013), which have resulted to a number of adverse human health effects. These health effects include neurological disorders such as seizures, convulsions, vertigo as well as the elevated risks of liver and breast cancer. Most of the reported health effects have been related to agricultural use and chronic occupational exposures (Blair et al., 1998; Wilson and Tisdell, 2001; Prathap and Srivastava, 2013; Anirudhan and Alexander, 2015).

Sophisticated analytical techniques have been reported for quantitative detection of these pesticides. These methods include; gas chromatography (Sen et al., 2011), gas chromatographic-mass spectrometry (Rodrigues et al., 2007), colorimetric methods (Lichtenstein et al., 1956), thin layer chromatography (Norton and Westöö, 1968), and high-performance liquid chromatography (Vassilikis et al., 1998). The analytical techniques are known to be very expensive and complex, requiring highly skilled personnel and time consuming sample preparations, they are also not suitable for onsite monitoring (Prathap et al., 2015, 2016; Fayemi et al., 2016). Therefore, electrochemical sensors are emerging as a technology of choice for the electrochemical detection of important biological and environmental analytes (Fayemi et al., 2016; Prathap et al., 2016; Zhuang et al., 2016). Electrochemical sensors are known to possess a vast number of advantages which include high sensitivity, rapid response, convenience in operation, and low fabrication costs (Prathap et al., 2016; Zhuanga et al., 2017; Chena et al., 2018). Different materials have been employed for the fabrication of these sensors and an example of such materials is carbon nanotubes. Carbon nanotubes (CNTs) possess unique properties such as high surface area to volume ratio, tensile strength, chemical stability, good electrical conductivity, and high adsorption capacity. CNTs have evolved as a good supporting material for surface modification of electrodes (Vairavapandian et al., 2008; Adekunle et al., 2010a, 2011b; Cesario et al., 2013). There are reports on the use of single-walled carbon nanotubes (SWCNTs) as a supporting material for surface modification of electrodes in electrochemical studies of some important analytes. Literature studies include nitrite electrochemical sensor based on Prussian blue/single-walled carbon nanotubes modified pyrolytic graphite electrode (Adekunle et al., 2011b), electrocatalytic detection of dopamine at single-walled carbon nanotubes functionalized with iron(III)oxide nanoparticles (Adekunle et al., 2010a) and probing the electrochemical behavior of SWCNT–cobalt nanoparticles and their electrocatalytic activities toward the detection of nitrite at acidic and physiological pH conditions (Adekunle et al., 2010b). Conducting polymers such as polyaniline (PANI) are of great importance due to their unique properties such as good environmental stability, ease of synthesis, oxidation or protonation, adjustable electro-optical properties (Yu et al., 2008; Tovide et al., 2014), and wide application potential (Trčová and Stejskal, 2011). PANI is known to have found potential applications in various areas like batteries (Dalui et al., 2008), separation devices (Sairam et al., 2006), light-emitting diodes (Prathap and Srivasta, 2011), electro rheological material (Zhang et al., 2002) and molecular sensors (Rahman et al., 2008). There is currently an interest in the use of metal oxide-CNTs nanocomposites, and metal oxide-polymer composites in electrochemistry to improve the performance of electrochemical detection of biological and environmental analytes. Also, owing to their intrinsic properties such as negative overpotential to hydrogen evolution, large scan potential range, antimony (Sb) based electrodes have received growing attention in electrochemistry (Prathap and Srivasta, 2011; Tovide et al., 2014). Despite these characteristics there are few number of reports describing the use of electrodes based on antimony and its oxides.

Thus, the novelty in this work is to explore the synergistic electrical properties and sensitivity of a nanocomposite based electrode materials made from carbon nanotubes (CNT), polyaniline (PANI) and antimony oxide (AO) toward nano detection of lindane and related pesticides in river and tap water samples. To the best of our knowledge, this study represents the first time a highly selective GCE-AONP-PANI-SWCNT electrode architecture for lindane detection is described. The study also provides information on the mechanism of electron transport between the surface active material and the underlying electrode or the analyte in solution for enhanced electrochemical response. This study therefore describes successful synthesis of GCE-AONPS-PANI-SWCNT electrode and its application for the electrochemical detection of lindane in river and tap water samples.

**METHODS**

**Materials, Reagents, and Instruments**

Lindane (C₈H₈Cl₆), pristine single-walled carbon nanotubes (SWCNT; 90% purity, 0.7–1.1 nm), tetrabutylammonium bromide (TBAB), and N,N-Dimethylformamide (DMF) were purchased from Sigma-Aldrich. Methanol, nitric acid (HNO₃), sulphuric acid (H₂SO₄), antimony chloride (SbCl₃), polyvinyl alcohol (PVA), sodium hydroxide (NaOH), hydrochloric acid (HCl), aniline (99%), ammonium peroxydisulfate (APS) [(NH₄)₂S₂O₈], sodium phosphate monobasic (NaH₂PO₄), sodium phosphate dibasic (Na₂HPO₄) and ethanol were of analytical grade and obtained from Merck chemicals. A 3 mm glassy carbon electrode (GCE) was used as the working electrode. Alumina micro powder (0.3 μm alumina slurries) and polishing

Frontiers in Chemistry | www.frontiersin.org 2 September 2018 | Volume 6 | Article 423
pads were used for polishing the working electrode. Phosphate buffer solution (PBS) of 0.1 M at pH 7 was prepared by using suitable amounts of NaH$_2$PO$_4$ and Na$_2$HPO$_4$ and then adjusted with NaOH. During electrochemical experiment purging of the prepared solutions was done by using nitrogen for oxygen elimination and also to prevent any form of oxidation. The electrochemical workstation used is the Autolab Potentiostat PGSTAT 302 with GPES software version 4.9. UV-vis and FTIR characterization were carried out using Agilent Technology, UV-1901, and Cary 600 series FTIR spectrometer, USA, respectively. Scanning electron microscope and X-ray diffraction spectrophotometer were used for morphological characterization of the nanomaterials.

### Synthesis of Antimony Oxide Nanoparticles (AONPs) and Polyaniline (PANI)

About 228 mg of SbCl$_3$ was dissolved in 100 mL of 1 M HCl, then 3 g of PVA was added. The mixture was dispersed by bath sonication for 15 min. Approximately 12 mL of 10 M NaOH was added slowly to the mixture until the mixture color changed to transparent-pale yellow. The mixture was then refluxed for 1 h and the color became more intense. The mixture was heated at 350°C for another 1 h to obtain a dry powder samples (Zhang et al., 2013). PANI was synthesized using a reported method by Kavitha et al. (2013). Quantitative 50 mL of 1 M HCl and 2 mL aniline were stirred together in 250 mL flask on a magnetic stirrer. Concurrently, 50 mL of 1 M HCl and 5 g of APS aqueous solution were also stirred together. This solution was added dropwise to the former solution and polymerized at a temperature of 70°C for 10 h to complete the reaction. The obtained precipitate was filtered and washed with 1 M HCl followed by distilled water until a clear filtrate was achieved. The final residue was dried for 24 h in the oven at 60°C.

### Acid Treatment of SWCNT

About 40 mg pristine SWCNTs were first dispersed in 40 mL DMF for 40 min at 40°C. The resulting dispersion was then filtered and washed with 10 mL methanol followed by deionized water. The solid was transferred into a glass vial containing 20 mL of 8 M HNO$_3$ and sonicated for 30 min at 40°C followed by the addition of 40 mL deionized water. After filtration the recovered solid was washed with deionized water until the filtrate was neutral and finally washed with 10 mL of methanol and 10 mL of DMF. Lastly the solid was dispersed in 20 mL DMF by bath sonication for 60 min at 40°C, it was filtered and dried to get powder form of fSWCNTs (Tchoul et al., 2007).

### Preparation of Nanocomposites

Approximately 2 mg of fSWCNT suspended in 2 mL DMF was mixed with 2 mg each of PANI and AONPs. The mixture was stirred for 48 h at room temperature. The formed AONP-PANI-fSWCNT nanocomposites was dried at 25°C overnight for the solvent to evaporate (Fayemi et al., 2015).

### Electrode Modification and Characterization of the Electrode

The electrode was modified by the drop-cast method (Adekunle et al., 2011a; Silwana et al., 2015). GCE surface was cleaned first by a gentle polish with 0.3 µm alumina slurries on a polishing pad followed by a gentle rinse with distilled water. The electrode was then sonicated in ethanol for 5 min and again in distilled water for another 5 min. About 5 mg each of the prepared nanomaterials, AONPs, PANI, fSWCNTs, and AONP-PANI-fSWCNT were suspended in 1 mL DMF. Each suspension was then dispersed by ultrasonic vibration for 30 min, and a 20 µL aliquot of this dispersion was dropped onto the GCE surface and dried at 50°C to obtain the modified electrodes. Fourier transform infrared spectroscopy (FT-IR), ultraviolet-visible (UV-Vis) spectrophotometry, X-ray diffraction (XRD) spectroscopy, and scanning electron microscopy (SEM) were used for the structural and morphological characterization of the synthesized nanomaterials.

### Electrochemical Studies

Electrochemical experiments were carried out to establish the successful modification of the electrodes, electron transport and electrocatalytic properties of the bare and modified GCE. Bare or modified glassy carbon electrode (GCE) disk (d = 3.0 mm in Teflon) was used as the working electrode, platinum disk as counter electrode and Ag/AgCl, KCl (sat’d) as reference electrode. A bench top pH/ISE ORION meter, model 420A, was used for pH measurements. All solutions were de-aerated by bubbling with nitrogen prior to each electrochemical experiment. Experiments were performed at 25 ± 1°C. Electrochemical experiments were carried out using an AUTOLAB Potentiostat PGSTAT 302 (Eco Chemie, Utrecht, and The Netherlands) driven by the GPES software version 4.9. Electrochemical impedance spectroscopy (EIS) measurements were performed with an AUTOLAB Frequency Response Analyser (FRA) software between 0.1 Hz and 100 kHz using a 5 mV rms sinusoidal modulation with a solution of 5 mM mixture of K$_4$Fe(CN)$_6$ and K$_3$Fe(CN)$_6$ (1:1) prepared with pH 7.0 phosphate buffer solution (PBS). CV experiments were carried out by running the bare and modified GCE in 0.1 M buffer solution (PBS, pH 7.0) alone, 5 mM [Fe(CN)$_6$]$_{4−/3−}$ solution prepared in 0.1 M PBS and 9 µM of lindane prepared in 60:40 methanol/water containing 0.05 M TBAB as supporting electrolyte.

### RESULTS AND DISCUSSION

#### Characterization of Synthesized Nanomaterials

##### Fourier Transform Infrared Spectroscopy

Figure 1 depicts comparative FT-IR spectra of (a) AONPs, (b) PANI, (c) fSWCNTs and (d) AONP-PANI-fSWCNT. From the AONPs spectrum (Figure 1A) the peak at 3727 cm$^{-1}$ was attributed to the stretching and bending vibrations of OH group in water. The peak at 733 cm$^{-1}$ was assigned to the Sb-O-Sb vibrations, whilst the small absorption band at 601 cm$^{-1}$ was attributed to the metal oxygen stretching vibration (Zhang et al.,...
FIGURE 1  | FT-IR spectra of (A) AONPs, (B) PANI, (C) fSWCNTs, and (D) AONP/PANI/SWCNT.

The PANI spectrum (Figure 1B) shows characteristic peaks at ~878, 1422, 1575, and 3019 cm⁻¹, respectively. The sharp peaks observed at 1575 and 1422 cm⁻¹ were due to the C=N and C=C stretching vibrations of quinoid and benzenoid rings, respectively (Cesarino et al., 2011; Konyushenko et al., 2011; Abdolahi et al., 2012; Olad et al., 2012; Kavitha et al., 2013; Prathap et al., 2013). The peak at 878 cm⁻¹ can be assigned to the C-H out of plane bending (Abdolahi et al., 2012; Olad et al., 2012; Prathap et al., 2013). The broad peak at ~3019 cm⁻¹ represents the N-H stretching vibrations (Olad et al., 2012).

Figure 1C depicts FT-IR spectrum of acid functionalized single-walled carbon nanotubes and from the spectrum, characteristic peaks were observed at 2930, 2340, 1698, 1104, and 820 cm⁻¹. The peaks at 2930 and 2340 cm⁻¹ observed in the high frequency region were attributed to the symmetric mode and anti-symmetric mode of CH₂ (Tchoul et al., 2007; Helali et al., 2016). The peak observed at 1698 cm⁻¹ was assigned to the carbonyl stretching vibration and the peaks at 2930 and 2340 cm⁻¹ were due to the symmetric and anti-symmetric mode of CH₂ (Helali et al., 2016).

Ultraviolet-Visible Spectroscopy

Figure 2 shows the comparative UV-vis spectra of (a) AONPs, (b) PANI, (c) fSWCNTs and (d) AONP-PANI-SWCNTs. AONPs spectrum. A sharp peak at about 260 nm is characteristic of antimony oxide nanoparticles formation Figure 2D (Khalef, 2013). In Figure 2B peaks at 341 nm was attributed to the SWCNT defects or residual carbon impurities or both (Helali et al., 2016).
\(\pi-\pi^*\)benzenoid transition while 650 nm was assigned to the quinoid transition in PANI (Konyushenko et al., 2011; Prathap and Srivasta, 2011). The absorption spectrum of the fSWCNTs (Figure 2C) showed a significant loss of the van Hove band structure typical for pristine SWCNTs and usually centered around 650 nm and 900 nm (Chiang et al., 2001; Peng et al., 2003; Price and Tour, 2006). Figure 2D shows the spectrum of AONP-PANI-SWCNTs composite with characteristic peaks of the AONPs at around 260 nm, and quinoid transition peak of PANI around 650 nm, which are indicative of the presence of AONPs and PANI in the nanocomposite.

**X-Ray Diffraction Spectroscopy**

Figure 3 depicts x-ray diffraction pattern of (a) AONPs, (b) PANI, (c) fSWCNTs, and (d) AONP-PANI-SWCNT. Antimony oxide nanoparticles are generally known to exist in a wide range of compositions and are also known to display interconvertible polymorphism. The two most common forms are cubic and orthorhombic consisting of \(\text{Sb}_4\text{O}_6\) molecules and chains of \(\text{SbO}_3\) trigonal pyramids (Khalef, 2013). From the AONPs spectrum (Figure 3A), the sharp and intense peak observed at \(2\Theta = 37.5^\circ\) corresponds to the (2 0 0) plane indicating preferred crystallographic orientation similar to the sample prepared in iso-propional (Naidu et al., 2009). Similar increase in intensity for (1 1 0) and (2 0 0) was also reported (Deng et al., 2006), confirming successful oxidation of Sb metal to form nanorods like \(\text{Sb}_2\text{O}_3\). From the literature it was reported that PANI prepared at pH 7 exhibited high crystallinity with the XRD pattern showing an intense peak at \(2\Theta = 6.4^\circ\) as shown in the XRD pattern obtained for PANI in Figure 3B. Other peaks observed were at \(2\Theta = 18.5^\circ, 19.8^\circ, 23.4^\circ, 25.5^\circ, 26.4^\circ,\) and \(28.4^\circ\) confirming the high order structure of PANI, however broad peaks at \(20.4^\circ\) and \(25.5^\circ\) suggests semi-crystallinity and amorphous nature of the PANI with an average particle size of about 3.8 nm (Prathap and Srivasta, 2011). The XRD spectrum of fSWCNTs is depicted by Figure 3C and a broad peak at \(2\Theta = 28^\circ\) was observed. The peak is a typical pattern of an amorphous structure (Ng et al., 2005), numerous other x-ray studies on carbon materials also suggested that the peak is due to the (0 0 2) plane of the turbo-stratic graphitic layer arising from the
acid treatment degradation of SWNTs. The peak at $2\theta = 43^\circ$ was attributed to the presence of (1 0 0) peak in graphite. The XRD spectrum of AONP-PANI-SWCNT composite is shown in Figure 3D, the intense peak at $2\theta = 37.5^\circ$ corresponding to the (2 0 0) plane of AONPs confirms composite formation with well crystallized AONPs. The broad peak at $2\theta = 20.4^\circ$ was attributed to the amorphous structure of fSWCNTs as well as semi-crystallinity of PANI.

**Scanning Electron Microscopy**

The morphology of the synthesized nanomaterials were depicted by their respective SEM images. Figure 4 shows the SEM images of (a) AONPs, (b) PANI, (c) fSWCNTs and (d) AONP-PANI-SWCNT. The SEM image in Figure 4A showed that the AONPs appeared unevenly distributed and clustered randomly; the image also depicts compacted flaky platelets. The SEM image for PANI (Figure 4B) showed porous morphology with evenly distributed agglomerations. Figure 4C represents the SEM image for fSWCNTs showing unevenly distributed and tangled porous tube-like structures. The SEM image for the composite AONP-PANI-SWCNT (Figure 4D) shows a characteristic morphology for antimony oxide nanoparticles with random clusters but here the clusters are tangled by the SWCNTs resembling a web like assembly.

**Electrochemical Characterization**

**Cyclic Voltammetric Experiments**

Investigation of the electrochemical properties of the bare and modified electrodes were carried out by using cyclic voltammetry in 0.1 M PBS (pH 7.0), then in 5 mM $\text{[Fe(CN)}_6\text{]}^{3-}/\text{4-}$ solution at a scan rate of 25 mVs$^{-1}$. Figure 5A represents the comparative CVs for the modified electrodes in buffer solution. From the voltammograms, it was observed that the electrodes GCE-fSWCNTs and GCE-AONPs-PANI-SWCNT exhibited higher current responses. This may be due to the influence of SWCNTs which are known to enhance the electronic communication between the electrode and the electrolyte solution, thereby acting as “Coulombic Islands” (electron transfer stations) and tunneling electrons between the electrode surface and redox
species (Liu et al., 2012). A similar electrochemical behavior was further observed in the $[\text{Fe(CN)}_6]^{3^-/4^-}$ solution. As shown in Figure 5B, the cyclic voltammetric responses of the electrodes exhibited a pair of well-defined redox peaks characteristic of one-electron transfer redox process in the region $-0.2$ to $0.4$ V typical of $[\text{Fe(CN)}_6]^{3^-/4^-}$ redox process (Adekunle et al., 2015). The surface coverage ($\Gamma$) (Shap et al., 1979; Fu et al., 2009) for each electrode was calculated to be 65.8, 53.3, 34.9, 6.45, and 8.80 nmol.cm$^{-2}$ for GCE-AONP-PANI-SWCNT, GCE-fSWCNTs, GCE-PANI, GCE-AONPs, and bare GCE, respectively. Table 1 summarizes the electrochemical data and various electrochemical parameters obtained for the electrodes in 5 mM $[\text{Fe(CN)}_6]^{3^-/4^-}$ redox probe such as ratio of anodic and cathodic peak current ($I_p/I_{pc}$), surface coverage ($\Gamma$/mol.cm$^{-2}$) and peak-to-peak separation ($\Delta E_p$/V). The $I_p/I_{pc}$ values obtained are closer to 1 indicating that the couple is reversible (Bard and Faulkner, 2001), and the peak-to-peak separation ($\Delta E_p$) values indicate that the electron transport was fastest at the GCE-fSWCNTs electrode (90.3 mV/s) and slowest at the GCE-AONPs electrode (178.3 mV/s). The overpotential was relatively small at GCE-AONP-PANI-SWCNT nanocomposite electrode because the higher current response recorded at the electrode (anode: 451 $\mu$A; cathode: 499 $\mu$A), was greater than that at GCE-fSWCNTs (anode: 365 $\mu$A; cathode: 422 $\mu$A).

**Electrochemical Impedance Spectroscopy**

EIS was carried out to investigate the electron transfer properties of the various electrodes in $[\text{Fe(CN)}_6]^{4^-/[\text{Fe(CN)}_6]^{3^-}$ solution. The experiments were conducted at frequency range of 0.1–100 kHz. Figure 6 depicts the Nyquist plots (a) and Bode plots (b) obtained for the bare-GCE, GCE-AONPs, GCE-PANI, GCE-fSWCNTs, and GCE-AONP-PANI-SWCNT electrodes, whilst Figures 6C,D represent the circuit model used to fit the EIS data. The circuit represented by Figure 6C was used to fit the data of the bare-GCE, GCE-AONPs and GCE-PANI and circuit represented by Figure 6D was used to fit the data for GCE-fSWCNTs and GCE-AONP-PANI-SWCNT. The circuits parameters were described as $R_s$, $R_{ct}$, $C_{dl}$, and $W$ representing the electrolyte resistance, charge transfer resistance, double layer capacitance, and Warburg impedance, respectively (Lisdat and Shäfer, 2008; Adekunle et al., 2011b).

Table 2 summarizes the EIS data obtained for the various modified electrodes. It was observed from the results that
FIGURE 5 | CVs of modified electrodes in (A) 0.1 M PBS solution (B) 5 mM [Fe(CN)₆]³⁻/⁴⁻ solution at a scan rate of 25 mVs⁻¹.

TABLE 1 | Summary of cyclic voltammetric parameters obtained for bare and modified electrodes in 5 mM [Fe(CN)₆]³⁻/[Fe(CN)₆]⁴⁻ solution prepared in 0.1 M PBS.

| Electrode                        | Ipₐ/µA | Ipᶜ/µA | Ipᵃ/Ipᶜ | ι/mol·cm⁻² | Eₚa/mV | Eₚc/mV | ΔEp/mV |
|---------------------------------|--------|--------|---------|------------|--------|--------|--------|
| GCE                             | 60.4   | 61.7   | 0.98    | 8.80       | 271.1  | 161.3  | 109.8  |
| GCE-AONPs                       | 44.2   | 49.9   | 0.89    | 6.45       | 337.1  | 158.8  | 178.3  |
| GCE-PANI                        | 239    | 199    | 1.20    | 34.9       | 278.5  | 161.3  | 117.2  |
| GCE-fSWCNTs                     | 365    | 422    | 0.86    | 53.3       | 251.6  | 161.3  | 90.3   |
| GCE-AONP-PANI-SWCNT             | 451    | 499    | 0.90    | 65.8       | 278.5  | 161.3  | 117.2  |

GCE-fSWCNT and GCE-AONP-PANI-SWCNT electrodes had the lowest Rct values (Table 2) than the bare GCE, GCE-AONPs and GCE-PANI indicating that modification particularly with the f-SWCNT on the GCE significantly increased the conductivity of the electrode and enhanced charge transfer.

Electrocatalytic Studies of Lindane

Electrocatalytic Reduction of Lindane

The electrochemical behavior of 9 µM lindane at a potential of −0.2 to −1.7 V and a scan rate of 25 mVs⁻¹ using CV technique is represented in Figure 7. Figure 7A shows the comparative cyclic voltammograms obtained for GCE, GCE-AONPs, GCE-PANI, GCE-fSWCNTs, and GCE-AONP-PANI-SWCNT electrodes. The obtained reduction peaks with no anodic peaks suggests that the electrocatalysis of lindane is an irreversible reaction and similar results have been reported (Kumaravel et al., 2013; Kaur et al., 2015; Fayemi et al., 2016; Otles, 2016). From the voltammogram in Figure 7A, the reduction potential for lindane was observed at −1.14, −1.00, −0.93, and −0.88 V at the bare GCE, GCE-PANI, GCE-AONP, and GCE-AONP-PANI-SWCNT electrodes, respectively. There was no observable peak for lindane at the GCE-fSWCNT modified electrode probably due to the capacitive current of the electrode. However, the introduction of the AONP in GCE-AONP-PANI-SWCNT electrode increased the electrode conductivity and catalysis with well pronounced and observable lindane reduction peak as compared to SWCNT or PANI modified electrodes alone. Generally, the modified electrodes shifted to a more positive potential as compared to that of the bare GCE. It was interesting to note that the reduction current for GCE-AONP-PANI-SWCNT composite modified electrode significantly increased by over 6-folds as compared to that of the bare GCE. This may be due to the high electrocatalytic activity and strong adsorption capabilities of the nanomaterial on the modified electrodes and its effective interaction with the analyte (Zhang and Fang, 2010).

Since the GCE-AONP-PANI-SWCNT modified electrode proved superior to the other electrodes investigated, its stability and resistance toward fouling effect as a result of lindane reduction product was further explored by CV using 20 scans. Figure 7B presents the CV obtained in 9 µM lindane at a scan rate of 25 mVs⁻¹. Lindane reduction current was stable after 20 scans with a small current decrease (0.57 %) suggesting good stability of the GCE-AONP-PANI-SWCNT modified electrode in lindane.

Effect of Scan Rate on Lindane Reduction

The effect of scan rate on lindane reduction current was carried out by using CV experiments in the presence of
9 µM lindane. **Figure 8A** presents the cyclic voltammograms obtained for GCE-AONP-PANI-SWCNT modified electrode. It was observed that as the scan rate increases there was simultaneous increase in the peak current which suggests diffusion controlled process (Siswana et al., 2006; Hegde et al., 2008; Kumaravel et al., 2013). **Figure 8B** shows the plot of current (I) against square root of scan rate (v$^{1/2}$) with a linear regression equation $I = -1.4147v^{1/2} + 1.4058$ and a correlation coefficient $R^2 = 0.9851$. A non-zero intercept value can be attributed to process of adsorption after diffusion (Prathap et al., 2015). It was proposed that the mechanism of lindane reduction continues by dissociative electron transfer (DET) leading to the scission of the carbon-chlorine bond. The mechanism involves the formation of an intermediate radical and an anion as shown by **Scheme 1**, or in a concerted mechanism yielding directly a radical and an anion shown by **Scheme 2** (Issié et al., 2008; Kaur et al., 2015).

The repeatability study on the modified AONP-PANI-SWCNT glassy carbon electrode was carried out in the presence of 9 µM lindane for a period of 30 successive days under the same condition. The electrode was rinsed in distilled water after every run. A 5% increase in reduction peak current was observed on the second day while after the 30th day, a drop in current by about 50% was observed as shown in **Figure 8C** suggesting fouling (electrode poisoning) effect. This suggests that the catalytic property of the modified electrode could only last for about 15 days with relatively little drop in current response (ca 20%). However, the composite modified electrode showed no significant drop in lindane reduction current when reused after storage in phosphate buffer solution (pH 7.0) in
Table 2 | Summary of the impedance data obtained for the modified electrodes in 5 mM \([\text{Fe(CN)}_6^{3-}/4^-]\).

| Electrode        | \(R_s\) (\(\Omega\) cm\(^2\)) | \(R_{ct}\) (\(\Omega\) cm\(^2\)) | \(Q\) (\(\times 10^{-3}\)) | \(W\) (\(\Omega\) cm\(^2\)) (\(\times 10^{-3}\)) | \(C_{dl}\) (nF) | Chi-square (\(\times 10^{-3}\)) |
|------------------|-------------------------------|-------------------------------|----------------------|---------------------------------|-------------|----------------------|
| GCE              | 108.3                         | 362                           | 0.0497               | 1.616                           | –           | 2.667                |
|                  | (0.456)                       | (0.76)                        | (3.508)              | (0.822)                         |             |                      |
| GCE-PANI         | 106.5                         | 371                           | 0.1103               | 1.38                            | –           | 34.116               |
|                  | (1.543)                       | (5.138)                       | (11.93)              | (4.632)                         |             |                      |
| GCE-AONPs        | 97.5                          | 27.40                         | 0.0226               | 1.344                           | –           | 123.64               |
|                  | (1.746)                       | (2.439)                       | (1.069)              | (11.746)                        |             |                      |
| GCE-ISWCNTs      | 15.87                         | 25.01                         | 7.692                | –                               | 125.7       | 9.5302               |
|                  | (0.0)                         | (0.934)                       | (1.782)              | (0.406)                         |             |                      |
| GCE-AONP-PANI-SWCNT | 60.7                       | 46.6                          | 7.79                 | –                               | 271.6       | 32.658               |
|                  | (7.85)                        | (9.963)                       | (1.181)              | (0.521)                         |             |                      |

The values in parenthesis represent the percentage error of the data fitting.

Figure 7 | (A) CVs of the GCE modified electrodes in the presence of 9 \(\mu\)M lindane at a scan rate of 25 mVs\(^{-1}\). (B) CVs (20 scans) of GCE-AONP-PANI-SWCNT modified electrode in the presence of 9 \(\mu\)M lindane at a scan rate of 25 mVs\(^{-1}\).

a refrigerator (at 4°C) for 4 weeks. Therefore, the current is reproducible.

Electroanalysis of Lindane

Square-wave voltammetry (SWV) experiments were carried out in various concentrations of lindane prepared in 0.05 M TBAB. Figure 9A shows the square-wave voltammograms obtained for the GCE-AONP-PANI-SWCNT modified electrode with varying concentrations of lindane. Figure 9B presents plot of current response versus lindane concentration with a linear regression equation \(I_p = 202.5 \text{[lindane]} + 80.997\) and correlation coefficient \(R^2 = 0.9903\). It was observed from Figure 9A that the lindane reduction peak current simultaneously increased with increase in lindane concentration and the potential gradually shifted negatively with increasing lindane concentration. In Figure 9B the linear response obtained was satisfactory over the range of lindane concentrations 0.0–18.8 nM. The standard deviation error value from the linear graph (Figure 9B) was found to be 0.21. The detection limit (LoD) for lindane at GCE-AONP-PANI-SWCNT modified electrode was calculated by using the relationship \(3.3 \delta/m\), where \(\delta\) is the relative standard deviation of the intercept of the y-coordinates and \(m\) is the slope of the same line (Adekunle et al., 2010a). The LoD, limit of quantification (LoQ) and sensitivity of the GCE-AONP-PANI-SWCNT electrode toward lindane were determined to be 2.01, 6.09 nM, and 202.5 \(\mu\)A/\(\mu\)M, respectively. The LoD and sensitivity reported here were found to be better compared to those reported at NiCo\(_2\)O\(_4\) modified GCE (3.6 \(\mu\)M, 0.2 \(\mu\)A/\(\mu\)M) (Prathap and Srivastava, 2013), cellulose acetate modified GCE (37 \(\mu\)M) (Kumaravel et al., 2013), Nylon6,6/MWCNT/Fe\(_3\)O\(_4\) modified GCE (32 nM) (Fayemi et al., 2016) and CuO/MnO\(_2\) hierarchical nano-microstructures modified electrode (4.8 nM, 0.022 \(\mu\)A/\(\mu\)M) (Prathap et al., 2015). This suggests that the AONP-PANI-SWCNT modified GCE has excellent and improved electrocatalytic activity for the electro-oxidation of lindane. Table 3 summarizes the comparison
FIGURE 8 | (A) CVs of GCE-AONP-PANI-SWCNT electrode in the presence of 9 µM lindane prepared in supporting electrolyte containing 0.05 M TBAB at a scan rate of 25–1,000 mVs\(^{-1}\). (B) Plot of current (I) against square root of scan rate (\(\sqrt{v}\)) for GCE-AONP-PANI-SWCNT modified electrode in lindane, and (C) Repeatability studies of GCE-AONP-PANI-SWCNT electrode in the presence of 9 µM for 30 successive days.

of analytical performance of AONP-PANI-SWCNT modified GCE electrode with other sensors reported for electrocatalytic detection of lindane.

**Lindane Interference Studies**

Interference studies were carried out to investigate the selectivity of the developed sensor toward lindane determination using continuous chronoamperometry (CA) and CV. The selection of interfering species for this study was based on the historical use of lindane both in industry and agriculture sector and also the fact that these species may be present in water or soil from industrial areas and agricultural land (Kumaravel et al., 2013; Fayemi et al., 2016; Prathap et al., 2016). The selected species which may potentially interfere with lindane determination include electroactive organic compounds such as cyclohexane (C\(_6\)H\(_{12}\)), benzene (C\(_6\)H\(_6\)), phenol (C\(_6\)H\(_5\)OH) and some inorganic cations such as calcium (Ca\(^{2+}\)), potassium (K\(^+\)), magnesium (Mg\(^{2+}\)), and iron (Fe\(^{2+}\)). The chronoamperometric response at the GCE-AONP-PANI-SWCNT modified electrode with addition of 1 mM each of lindane, and the interfering species at a fixed potential of 0.2 V (graph not shown) shows that the addition of organic compounds such as C\(_6\)H\(_{12}\), C\(_6\)H\(_6\), and C\(_6\)H\(_5\)OH did not interfere with the current response of lindane. Presence of phenol lead to the process of hydroxylation in the reaction mixture and hence the removal or substitution of chloride from chlorobenzene (lindane) with hydroxyl ions, resulting in oxidation of lindane. Likewise, in the presence of inorganic interfering species such as Ca\(^{2+}\), K\(^+\), Mg\(^{2+}\), and Fe\(^{2+}\) there was no interference with lindane signal. The results obtained indicated that the GCE-AONP-PANI-SWCNT
modified electrode can be used for selective determination of lindane in the presence of organic and inorganic molecules. Figure 10 shows the percentage current response of lindane reduction peak in the presence of the interfering species at GCE-AONP-PANI-SWCNT modified electrode using CV at a scan rate of 25 mVs⁻¹. The results obtained showed that

![Scheme 1](image1.png)

SCHEME 1 | Stepwise mechanism of lindane reduction.

![Scheme 2](image2.png)

SCHEME 2 | Concerted mechanism of lindane reduction.

![Figure 9](image3.png)

FIGURE 9 | (A) SWV of GCE-AONP-PANI-SWCNT electrode in 0.0 nM–18.8 nM lindane. (B) Plot of current response versus lindane concentration and error bars represent standard deviations of three independent measurements.

![Table 3](image4.png)

TABLE 3 | Comparison of analytical performance of AONP-PANI-SWCNT modified GCE electrode with other non-enzymatic sensors reported for electrocatalytic detection of lindane.

| Composite electrode                  | LoD (µM) | Linear range (µM) | Sensitivity (µA/µM) | References                  |
|--------------------------------------|----------|-------------------|---------------------|-----------------------------|
| NiCo₂O₄/GCE                          | 3.6      | 10–170            | 0.2                 | Prathap et al., 2013        |
| CA/GCE                               | 37       | 50–1000           | –                   | Kumaravel et al., 2013     |
| Nylon6,6/MWCNT/Fe₃O₄/GCE             | 32 × 10⁻³| 9.9–5.0           | –                   | Fayemi et al., 2016         |
| CuO/MnO₂/GCE                         | 4.8 × 10⁻³| 0.0–700          | 0.022               | Prathap et al., 2015        |
| αMnO₂-NW/GCE                         | 114 × 10⁻³| 1.1–510          | –                   | Prathap et al., 2016        |
| AgNPs(5%)-PANI-Nano-ZSM-5/GCE        | 5 × 10⁻³ | 0.01–90           | 1.24 mA/µM         | Kaur et al., 2015           |
| GCE-AONP-PANI-SWCNT                  | 2.01 × 10⁻³| 0.0 × 10⁻³ – 18.8 × 10⁻³| 202.5 µA/µM        | Present study                |

GCE, Glassy carbon electrode; CA, Cellulose acetate; MWCNT, Multi-walled carbon nanotube; NW, Nanowires; Nano-ZSM-5, Nanocrystalline Zeolite Socony Mobil-5; PANI, Polyaniline.
FIGURE 10 | Current response of lindane reduction signal as a percentage in the presence of interfering species (benzene, cyclohexane, phenol, Ca$^{2+}$, Fe$^{2+}$, K$^+$, and Mg$^{2+}$) at GCE-AONP-PANI-SWCNT modified electrode by cyclic voltammetry.

the GCE-AONP-PANI-SWCNT modified electrode had anti-interference behavior toward the detection of lindane in the presence of interfering species (benzene, cyclohexane, phenol, Ca$^{2+}$, Fe$^{2+}$, K$^+$, and Mg$^{2+}$) with an average of only 12.1% current drop on the reduction signal of lindane.

Lindane Real Sample Analysis
Real sample analysis was carried out to establish the practical application of the GCE-AONP-PANI-SWCNT modified electrode toward lindane detection in both river water and tap water samples. The river water samples were collected from upper crocodile sub—catchment of Crocodile River in Rustenburg of the North West province, South Africa. This river water system is known to receive runoff from agricultural land as well as effluents from the industrial mines around the area. The samples were collected from a point in the river with GPS coordinates: S 025° 40’ 09.3 E 027° 47” 31.7. The river water was used without further purification. Lindane/water samples with concentrations ranging from 0.5–100 µM were prepared by using 60:40 methanol/river-water containing 0.05 M TBAB and CV studies were carried out to determine the concentration of lindane in the samples. Table 4 presents the results obtained and it was observed that recoveries of lindane at lower concentrations of added lindane was higher suggesting the presence of lindane in the river water samples; however at higher added concentrations of lindane recoveries were lower. This observation may be due to the sensor’s kinetic range.

The tap water used for the study was obtained from the local municipality water supply and was used without further purification. Concentration of prepared Lindane samples added ranges from 10 to 50 µM. The sensor yielded good recoveries (99.9% average) at lower concentrations of lindane when compared to higher concentrations (86.5% average).

Table 4 | Determination of Lindane concentration in river water and tap water samples using CV method.

| Sample     | Compound | Added (µM) | Found$^a$ (µM) | Recovery (%) | RSD (%) |
|------------|----------|------------|----------------|--------------|---------|
| River water| Lindane  | 0.5        | 1.0            | 200          | 14.4    |
|            |          | 1          | 1.6            | 160          | 10.1    |
|            |          | 20         | 18.7           | 94           | 12.9    |
|            |          | 100        | 100.2          | 100.2        | 0.35    |
| Tap Water  | Lindane  | 10         | 9.87           | 98.7         | 3.51    |
|            |          | 20         | 20.3           | 101          | 2.99    |
|            |          | 40         | 33.6           | 84           | 3.88    |
|            |          | 50         | 44.3           | 89           | 4.01    |

$^a$Average value of three determinations.

CONCLUSIONS
This paper describes the successful fabrication of a sensor for the electrochemical detection of lindane. Electrocatalytic reduction of lindane was carried out using CV technique and the highest current response was recorded at the GCE-AONP-PANI-SWCNT electrode with reduction peak at $-0.95 \, \text{V}$. Stability study (20 scans) toward lindane was carried out using GCE-AONP-PANI-SWCNT electrode and the observed current drop between the 1st and 20th scan was only 0.57%. This suggested that the fabricated GCE-AONP-PANI-SWCNT sensor was stable and resistant to electrode fouling effects. The effect of scan rate was investigated using CV experiment and the plot of reduction peak currents versus square root of scan rate gave a linear relationship where reduction current increased simultaneously as the scan rate increased suggesting that lindane reduction was a diffusion controlled process. The results recorded for the LoD, LoQ, and sensitivity of the GCE-AONP-PANI-SWCNT electrode toward lindane were better compared with those reported in literature. The proposed sensor further demonstrated good selectivity and anti-interference behavior toward the detection of lindane in the presence of interfering species. Real sample analysis to establish the analytical potential of the developed GCE-AONP-PANI-SWCNT sensor was carried out using river and tap water samples. The percentage recoveries obtained at the fabricated sensor indicated its potential for the determination of lindane in river and tap water samples.

AUTHOR CONTRIBUTIONS
EE and AA conceptualized and designed the work and were part of the manuscript write-up. KM, OF, and E-SS carried out the experiments, interpreted some of the results and were also involved in the manuscript preparation. All the authors reviewed the manuscript and have agreed to its publication.
ACKNOWLEDGMENTS

AA thanks the North-West University for post-doctoral fellowship and Obafemi Awolowo University, Nigeria for the research leave visit. EE acknowledges the National Research Foundation of South Africa for Incentive funding for Rated Researchers. This project received support from the King Saud University, Deanship of Scientific Research, College of Engineering Research Center.

REFERENCES

Abdolahi, A., Hamzah, E., Ibrahim, Z., and Hashim, S. (2012). Synthesis of uniform polyaniline nanofibers through interfacial polymerization. Materials (Basel), 5, 1487–1494. doi: 10.3390/ma5081487

Abdolahi, M., Ranjbar, A., Shadnia, S., Nikfar, S., and Rezaie, A. (2004). Pesticides and oxidative stress: a review. Med. Sci. Monit. 10, RA141–RA147.

Adekunle, A. S., Agboola, B. O., Pillay, J., and Ozoemena, K. I. (2010a). Electrochemical detection of dopamine at single-walled carbon nanotubes–iron (III) oxide nanoparticles platform. Sensors Actuat. B Chem. 148, 93–102. doi: 10.1016/j.snb.2010.03.088

Adekunle, A. S., Ayenimo, J. G., Fang, X., Doherty, W. O., Arotiba, O. A., and Mamba, B. B. (2011b). Nitrite electrochemical sensor based on polyaniline nanofibers through interfacial polymerization. Int. J. Electrochem. Sci. 6, 4388–4403.

Adekunle, A. S., Pillay, J., and Ozoemena, K. I. (2010b). Probing the electrochemical behaviour of SWCNT–cobalt nanoparticles and their electrocatalytic activities towards the detection of nitrite at acidic and physiological pH conditions. Electrochim. Acta 55, 4319–4327. doi: 10.1016/j.electacta.2009.02.102

Anirudhan, T. S., and Alexander, S. (2015). Design and fabrication of molecularly imprinted polymer-based potentiometric sensor from the surface modified multi-walled carbon nanotube for the determination of lindane (γ-hexachlorocyclohexane), an organochlorine pesticide. Biosens. Bioelectron. 64, 586–593. doi: 10.1016/j.bios.2014.09.074

Bard, A. J., and Faulkner, L. R. (2001). Electrochemical Methods: Fundamentals and Applications. New York, NY: John Wiley and Sons.

Blair, A., Cantor, K. P., and Zahm, S. H. (1998). Non-Hodgkin’s lymphoma and agricultural use of the insecticide lindane. Am. J. Ind. Med. 33, 82–87.

Cesarino, I., Moraes, F. C., and Machado, S. A. S. (2011). A biosensor based on polyaniline-carbon nanotube core-shell for electrochemical detection of pesticides. Electroanalysis 23, 2586–2593. doi: 10.1002/elan.201100161

Cesarino, L., Cesarino, V., and Lanza, M. R. V. (2013). Carbon nanotubes modified with antiomy nanoparticles in a paraffin composite electrode: simultaneous determination of sulfamethoxazole and trimethoprim. Sensors Actuat. B Chem. 188, 1293–1299. doi: 10.1016/j.snb.2013.08.047

Chena, D., Zhuang, X., Zhai, J., Zhenga, Y., Luan, H., and Chena, L. (2018). Preparation of highly sensitive Pt nanoparticles-carbon quantum dots/ionic liquid functionalized graphene oxide nanocomposites and application for H2O2 detection. Sens. Actuat. B Chem. 250–1506. doi: 10.1016/j.snb.2017.08.156

Chiang, I. W., Brinson, B. E., Smalley, R. E., Margrave, J. L., and Hauge, R. H. (2001). Purification and characterization of Single-Wall Carbon Nanotubes (SWNTs) obtained from the gas-phase decomposition of CO (HiPco Process). J. Phys. Chem. B 105, 1157–1161. doi: 10.1021/jp003453v

Chrysikou, L., Gemenetzis, P., Kouras, A., Manoli, E., Terzi, E., and Samara, C. (2008). Distribution of persistent organic pollutants, polycyclic aromatic hydrocarbons and trace elements in soil and vegetation following a large scale landfill fire in northern Greece. Environ. Int. 34, 210–225. doi: 10.1016/j.envint.2007.08.007

Dalui, B., Basumallick, I. N., and Ghosh, S. (2008). Zinc-polyaniline rechargeable battery assembled with aqueous electrolyte. Ind. J. Chem. Technol. 15, 576–580.

Deng, Z., Fang, F., Chen, D., Meng, X., Cao, L., and Zou, B. (2006). A simple solution route to single-crystalline Sb2O3 nanowires with rectangular cross sections. J. Phys. Chem. B 110, 18225–18230. doi: 10.1021/jp063748y

El-Shahawi, M. S., Hamza, A., Bashammakh, A. S. A., and Al-Saggaf, W. T. (2010). An overview on the accumulation, distribution, transformations, toxicity and analytical methods for the monitoring of persistent organic pollutants. Talanta 80, 1587–1597. doi: 10.1016/j.talanta.2009.09.035

Fayemi, O. E., Adekunle, A. S., and Ebenso, E. E. (2015). Metal oxide nanoparticles/multi-walled carbon nanotube nanocomposite modified electrode for the detection of dopamine. J. Biosens. Bioelectron. 6, 190–194.

Fayemi, O. E., Adekunle, A. S., and Ebenso, E. E. (2016). A sensor for the determination of lindane using PANI/Zn, Fe(III) oxides and nylon 6,6/MWCNT/Zn, Fe(III) oxides nanofibers modified glassy carbon electrode. J. Nanomater. 2016, 78–88. doi: 10.1155/2016/4094730

Fu, C., Yang, W., Chen, X., and Evans, G. D. (2009). Direct electrochemistry of glucose oxidase on a graphite nanosheet–nafion composite film modified electrode. Electrochem. Commun. 9, 1007–1010. doi: 10.1016/j.elecom.2009.02.042

Gill, S., Bowers, W. J., Nakai, J. S., Yagminas, A., Mueller, R., and Pulido, O. (2013). EffectsOf environmentally relevant mixtures of persistent organic pollutants on the developmental neurobiology in rats. Toxicol. Pathol. 41, 38–47. doi: 10.1177/0192623312451370

Hegde, R. N., Swamy, B. K. E., Sherigara, B. S., and Nandibewoor, S. T. (2008). Electro-oxidation of atenolol at a glassy carbon electrode. Int. J. Electrochem. Sci. 3, 302–314.

Helali, S., Bohl, N., Mostafa, H. M. A., Zina, H. B., Al-Hartimi, O. A., and Abdelghani, A. (2016). Electrical impedance spectroscopy using single wall carbon nanotubes carboxodic acid functionalized: detection of copper in Tabuk-Kingdom of Saudi Arabia water. J. Nanomed. Nanotechnol. 7:396. doi: 10.1155/2016/342312

Isse, A. A., Gottardelo, S., Durante, C., and Gennaro, A. (2008). Dissociative electron transfer to organic chlorides: electrocatalysis at metal cathodes. Phys. Chem. Chem. Phys. 10, 2409–2416. doi: 10.1039/b719936h

Jones, K. C., and de Voogt, P. (1999). Persistent organic pollutants (POPs): state of the science. Environ. Pollut. 100, 209–221. doi: 10.1016/S0269-7491(99)00986-6

Kaur, B., Srivastava, R., and Satpathi, B. (2015). Silver nanoparticle decorated polyaniline–zeolite nanocomposite material based non-enzymatic electrochemical sensor for nanomolar detection of lindane. RSC Adv. 5, 57657–57665. doi: 10.1039/C5RA09461E

Kavita, B., Kurma, K. S., and Narsimlu, N. (2013). Synthesis and characterization of polyaniline nano-fibers. Ind. J. Pure Appl. Phys. 51, 207–209.

Kaviyarasu, K., Sajan, D., and Devarajan, P. A. (2013). A rapid and versatile method for solvothermal synthesis of Sb2O3 nanocrystals under mild conditions. Appl. Nanosci. 3, 529–533. doi: 10.1007/s13204-012-0156-y

Khalef, W. K. (2013). Synthesis of antiomy oxide nanoparticles by pulsed laser ablationin wet media. Iqaluj. J. Appl. Phys. 9, 5–13.

Konyushenko, E. N., Reynard, S., Pellerin, V., Trchové, M., Stejskal, J., and Supurina, I. (2011). Polyaniline prepared in ethylene glycol...
Electrocatalysis of Lindane

Kumaravel, A., Vincent, S., and Chandrasekaran, S. (2013). Development of an electroanalytical sensor for α-hexachlorocyclohexane based on a cellulose acetate modified glassy carbon electrode. Anal. Methods 5, 931–938. doi: 10.1039/c2ay26119g

Lichtenstein, E. P., Beck, S. D., and Schulz, K. R. (1956). Colorimetric determination of lindane in soils and crops. J. Agric. Food Chem. 4, 936–944. doi: 10.1021/jf60069a002

Lisdat, F., and Shafer, D. (2008). The use of electrochemical impedance spectroscopy forbiosensing. Anal. Bioanal. Chem. 391, 1555–1567. doi: 10.1007/s00216-008-1970-7

Liu, G., Wang, S., Liu, J., and Song, D. (2012). An electrochemical immunosensor based on chemically assembled of vertically aligned carbon nanotubes on carbon substrates for direct detection of the pesticide endosulfan in environmental water. Anal. Chem. 84, 3921–3928. doi: 10.1021/ac20275ap

Naidu, B. S., Pandey, M., Sudarsan, V., Vatsa, R. K., and Tewari, R. (2008). Electrochemical determination of some chlorinated pesticides in vegetable oils, margarine, butter, milk, eggs, meat, and fish by gas chromatography and thin-layer chromatography. J. Agric. Food Chem. 56, 107–115. doi: 10.1021/jf702178p

Norén, K., and Westöö, G. (1968). Determination of some chlorinated pesticides of an electroanalytical sensor for α-hexachlorocyclohexane based on cellulose acetate modified glassy carbon electrode. Anal. Chem. Acta 51, 23–28. doi: 10.1016/0003-2670(68)80076-5

Prathap, A. M. U., Sun, S., Wei, C., and Xu, Z. J. (2015). A novel α-hexachlorocyclohexane based on a carbon nanotube hybrid with nanoscale construction for enhanced sensitivity. J. Electroanal. Chem. 771, 155–162. doi: 10.1016/j.jelechem.2015.06.043

Prathap, A. M. U., Sun, S., and Xu, Z. J. (2016). Functionalization of single-walled carbon nanotubes with amino-functional silicon–carbon–nitrogen–phosphorus recognition materials. New J. Chem. 40, 7479–7513. doi: 10.1039/c6nj00814a
Zhang, Z., Guo, L., and Wang, W. (2001). Synthesis and characterization of antimony oxide nanoparticles. *J. Mater. Res.* 16, 803–805. doi: 10.1557/JMR.2001.0096

Zhuang, X., Tian, C., Luan, F., Wu, X., and Chen, L. (2016). One-step electrochemical fabrication of a nickel oxide nanoparticle/polyaniline nanowire/graphene oxide hybrid on a glassy carbon electrode for use as a non-enzymatic glucose biosensor. *RSC Adv.* 6, 92541–92546. doi: 10.1039/C6RA14970G

Zhuang, X., Chena, D., Wanga, S., Liua, H., and Chena, L. (2017). Manganese dioxide nanosheet-decorated ionic liquid-functionalized graphene for electrochemical theophylline biosensing. *Sens. Actuat. B* 185–191. doi: 10.1016/j.snb.2017.05.049

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Masibi, Fayemi, Adekunle, Sherif and Ebenso. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.