Introduction
Platelet-rich plasma (PRP) is defined as a portion of the plasma fraction of autologous blood having a platelet concentration above baseline [1]. It is an emerging treatment in the modern health sector known as ‘orthobiologics’. The goal of this discipline is to enhance the body’s innate ability to repair and regenerate. PRP therapy has lately gained a lot of attention as a safe, non-surgical, biological treatment of osteoarthritis and musculoskeletal repair [2]. The global market for PRP was valued at $45 million in 2009, and is projected to be worth more than $120 million by 2016 [3]. That being said, presently there is a lack of data available to truly confirm that PRP works as postulated. Future large randomized controlled trials (RCTs) are needed to assess its efficacy. Thus, PRP should be used with great caution at this time.

Clinical relevance - the ‘need’
Musculoskeletal conditions have an enormous and growing impact worldwide [4]. The World Health Organization has acknowledged that musculoskeletal injuries affect hundreds of millions of people worldwide and are the most common cause of severe long-term pain and physical disability [5].

Soft tissue injuries that include ligament and tendons represent up to 45% of all musculoskeletal injuries [6,7]. Chronic and degenerative musculoskeletal conditions can be challenging to treat and can have a protracted course compromising an otherwise active lifestyle. Many of these patients with sports- or overuse-related injuries might require a surgical intervention and protracted rehabilitation and yet have unclear outcomes. Additionally, there is still a dearth of treatments that could effectively enhance repair in the face of these chronic conditions. It is in this setting that pre-clinical studies and case reports suggesting improvement of osteoarthritis symptoms with PRP treatment have generated interest in its application for articular cartilage repair. If proven effective, this would constitute a breakthrough non-operative treatment option for osteoarthritis.

Platelet physiology and function
A typical blood specimen comprises 93% red blood cells, 6% platelets, and 1% white blood cells [8]. Platelets were
first seen in the blood by French physician Alfred Donné in 1842 [9]. These are small discoid cells with a life span of about 7 to 10 days. Following injury that causes bleeding, platelets are activated and aggregate together to release their granules containing growth factors that stimulate the inflammatory cascade and healing process. Platelets are responsible for hemostasis, construction of new connective tissue and revascularization and most of the research over the past century has been focused on this primary function [10]. Only in the past two decades have we learned that platelet activation in the body releases healing proteins called growth factors [11]. There are numerous growth factors with diverse functions, but cumulatively they may accelerate tissue and wound healing [12].

The ideology behind PRP treatment is the reversal of the red blood cell:platelet ratio by decreasing red blood cells to 5% (which are less useful in the healing process) and more importantly concentrating platelets containing a powerful congregation of growth factors to 94%. A normal platelet count in a healthy individual is between 150,000 and 450,000 cells per microliter of blood. Platelet concentrations of less than $1,000 \times 10^6$/ml were not reliable for enhancing wound healing [1], and most studies have suggested that tissue reparative efficacy with PRP can be expected with a minimum increase of five times the normal concentration of platelets (approximately 1 million platelets/μl) [13], whereas much higher concentrations did not show further enhancement of wound healing. The ideal concentration remains to be defined. The broad variability in platelet concentrating equipment and techniques used [14] in different studies may alter platelet degranulation characteristics that could affect clinical outcomes [15-17], making interpretation of the results challenging.

**Growth factors in platelet-rich plasma**

The notable components of PRP presented in Table 1 include transforming growth factor (TGF)-β, platelet-derived growth factors (PDGF-AB and PDGF-BB), insulin-like growth factor (IGF), vascular endothelial growth factors (VEGFs), epidermal growth factor (EGFs) and fibroblast growth factor (FGF)-2 [15,18,19]. TGF-β1 and PDGF stimulate proliferation of mesenchymal cells. TGF-β1 also stimulates extracellular matrix production, including collagen. Principally, these factors stabilize the damaged tissue during initial stages of tissue repair, and direct the local mesenchymal and epithelial cells to migrate, divide, and increase collagen and matrix synthesis, ultimately leading to fibrous connective tissue and scar formation [20]. VEGF and FGF-2 are important for stimulating new blood vessel formation to bring nutrients and progenitor cells to the injury site; however, additional factors are also required for neo-vascularization. PRP is postulated to improve the early healing of tendon defects by over-expression of IGF-1 [21]. The 70 amino acid polypeptide hormone IGF is a normal component of the plasma and is transported by IGF-binding proteins [22-24]. IGF-1 storage in platelets is unclear, with few proteomic studies reporting it to be absent and most literature detecting IGF-1 in platelets [19,25,26]; however, most studies have detected IGF-1 in PRP [27-29]. For PRP’s role in multiple healing pathways, it deserves due consideration as an adjunctive therapy for specific applications.

**Clinical application for platelet-rich plasma**

Autologous PRP injections were reportedly first used in 1987 in an open heart surgery [30]. Over 20 years ago, PRP was used in the dental field for promoting accelerated wound healing in cancer patients following jaw reconstruction. Physicians have used PRP to aid bone healing after spinal injury and soft tissue recovery following plastic surgery. PRP therapy gained broad popularity in early 2009, when it was reported that two of the Pittsburgh Steelers received PRP for their ankle injuries before their triumph at the Super Bowl. Due to the media attention, PRP became an accepted though unproven treatment for sports-related injuries [31]. Currently, PRP injections are being used in various applications, including orthopaedics, cardiovascular surgery, cosmetics, facio-maxillary surgery and urology [10]. As a result, multiple studies are now underway to understand PRP’s mechanism of action, refine the treatment, and formally demonstrate efficacy in placebo-controlled trials.

**Platelet-rich plasma treatment - what does it entail?**

The simplicity of PRP application is defined by three steps (Figure 1). For most orthopedic use, which currently happens to be in an outpatient setting, the physician harvests the venous blood and transfers it to the centrifuge. Once the blood is in the centrifuge, processing usually takes between 5 and 20 minutes, and a sterile barrier may be necessary, depending on automation and centrifuge processing protocols. The current recommendations state that the platelet concentration should be raised between four and six times above the baseline concentration. After the centrifuge processes, the physician extracts PRP according to device instructions. The platelets collected in PRP are often activated by the addition of thrombin and calcium chloride, which induce the release of these factors from the alpha granules. With wider acceptance of PRP treatment, its application is selectively being adapted for use in the operating room while surgery is in progress.

Research is ongoing to determine the best concentration, preparation and timing of the injections. We know
of several techniques for PRP preparation (Figure 2), with multiple commercially available products (Table 2); however, their application has been confusing because each technique leads to a different product with potentially dissimilar biology and unknown relative efficacy.

Variations in platelet-rich plasma preparations

The importance of having such a classification is to better understand the relative success or failure of different PRP preparations. Additionally, this can establish an objective approach for future development and research.

Although the evidence is anecdotal, platelet-leukocyte-rich plasma (PLRP) deserves a notable mention as a novel concept. PLRP enriched in growth factors and antimicrobial proteins has been used to address infection concerns in healing injuries associated with soft tissue defects. Cieslik-Bielecka and colleagues [32] were the first to report the application of PLRP in an infected high-energy soft tissue injury and indicated that the volume and concentration of platelets and leukocytes were adequate to induce the healing process despite concurrent infection.

Proposed mechanisms of action

Most of the published literature on the efficacy of PRP in treating orthopedic conditions, ranging from acute rotator cuff tears to chronic achilles tendinitis, inherently lack accurate reporting of the specific formulation used. The variability in the composition and preparation methods poses a predicament in making true comparisons between different studies and their relative effectiveness.

### Table 1. Summary of growth factors contained in platelet-rich plasma [64,77,78]

| Growth factor | Function |
|--------------|----------|
| Transforming growth factor-β (TGF-β) | Stimulates undifferentiated mesenchymal cell proliferation<br>Regulates endothelial, fibroblastic, and osteoblastic mitogenesis<br>Regulates collagen synthesis and collagenase secretion<br>Regulates mitogenic effects of other growth factors<br>Stimulates endothelial chemotaxis and angiogenesis<br>Inhibits macrophage and lymphocyte proliferation |
| Fibroblast growth factor (FGF) | Promotes growth and differentiation of chondrocytes and osteoblasts<br>Mitogenic for mesenchymal cells, chondrocytes, and osteoblasts |
| Platelet-derived growth factor a and b (PDGF) | Mitogenic for mesenchymal cells and osteoblasts<br>Stimulates chemotaxis and mitogenesis in fibroblast, glial, or smooth muscle cells<br>Regulates collagenase secretion and collagen synthesis<br>Stimulates macrophage and neutrophil chemotaxis |
| Epidermal growth factor (EGF) | Stimulates endothelial chemotaxis or angiogenesis<br>Regulates collagenase secretion<br>Stimulates epithelial or mesenchymal mitogenesis |
| Vascular endothelial growth factor (VEGF) | Increases angiogenesis and vessel permeability<br>Stimulates mitogenesis for endothelial cells |
| Connective tissue growth factor (CTGF) | Promotes angiogenesis<br>Cartilage regeneration<br>Fibrosis and platelet adhesion |
| Insulin like growth factor (ILGF 1 and 2) | Chemotactic for fibroblasts and stimulates protein synthesis<br>Enhances bone formation |
| Platelet factor 4 (PF-4) | Stimulate the initial influx of neutrophils into wounds<br>Chemo-attractant for fibroblasts |
| Interleukin 8 (IL-8) | Pro-inflammatory mediator<br>Recruitment of inflammatory cells |
| Keratinocyte growth factor (KGF) | Promote endothelial cell growth, migration, adhesion and survival<br>Angiogenesis |
effectiveness. Principally, application of an autologous plasma rich in growth factors is considered to be beneficial in restoring connective tissues because of the slow delivery of growth factors from harvested platelets that have been activated by endogenous thrombin promoted by the addition of calcium chloride [33]. Admittedly, the precise mechanisms of action of PRP are poorly understood.

**Basic science of platelet-rich plasma therapy**

**In vitro studies**

**Bone formation**

PRP and its soluble fraction stimulate the osteoblastic differentiation of myoblasts and osteoblastic cells in three-dimensional cultures in the presence of bone morphogenetic protein (BMP)-2, BMP-4, BMP-6 or BMP-7 [34]. Moreover, heparin-binding fractions obtained from serum also stimulated osteoblastic differentiation in the presence of BMP-4. These results suggested that platelets contain not only growth factors for proliferation but also novel potentiator(s) for BMP-dependent osteoblastic differentiation.

**Osteoarthritis**

Platelet released growth factors (PRGFs) regulate endogenous hyaluronic acid (HA) synthesis, thereby protecting the cartilage and lubricating the joint [35]. PRGF enhances the secretion of HA and induces hepatocyte growth factor (HGF) production by synovial fibroblasts isolated from arthritic patients [36]. Platelets pre-incubated in an acidic environment (pH 5.0) induced the highest degree of fibroblast proliferation and the concentration of PDGF in the different treated lysates [37]. Intra-articular administration of PDGF might be beneficial in restoring HA concentration and switching angiogenesis to a more balanced status but does not halt the effects of IL-1β on synovial cells.

**Tendon healing**

In 1994, Packer and colleagues [38] described an *in vitro* model to study fibroblast activity during tendon healing. Later, Bernard-Beauvois and colleagues [39] were the first to culture and characterize the rabbit tenocytes *in vitro*, and Schulze-Tanzil and colleagues [40] established a three-dimensional high-density culture system for cultivation of human tenocytes for tissue engineering.

An *in vitro* study suggested that administering autologous platelet-rich clots might be beneficial in the treatment of tendon injuries by inducing cell proliferation and promoting the synthesis of angiogenic factors during the healing process [41]. Cultured tendon cells synthesize VEGF and HGF in the presence of growth factors from PRP clots, with the amounts synthesized being significantly higher with supernatants from platelet-rich clots than supernatants from a platelet-poor clot. Also notable was that the mitogenic activity of the supernatants was not decreased by the thrombin inhibitor hirudin. In human tenocyte cultures, not only platelet-rich clot releasate, but also platelet-poor clot releasate, stimulated cell proliferation and total collagen production. The former, but not the latter, slightly increases the expression of matrix-degrading enzymes (matrix metalloproteinases (MMPs) 1, 3 and 13) and endogenous growth factors (TGF-β and VEGF-a) [42].

**Animal studies**

**Distraction osteogenesis**

In the rabbit bone defect/lengthening model, local injection of PRP enhanced bone consolidation during the consolidation phase; however, in the early stage of distraction osteogenesis, the combined use of decalcified bone matrix along with PRP did not reveal any additional benefit [43].

**Muscle healing**

Muscle healing is an intricate and dynamic process aimed at restoration of anatomic continuity and function of the injured muscle [18]. Local delivery of PRP has been found to shorten the recovery time after a muscle strain injury in a small-animal model [44].

**Osteochondral defect repair**

Sun and colleagues [45] in their rabbit model of osteochondral defect found improved healing as assessed by macroscopic examination, micro-computed tomography,
Table 2. Classification of different platelet concentrates into four broad categories depending on leukocyte and fibrin content alone [14]

| Category                          | Example                                                                 |
|-----------------------------------|------------------------------------------------------------------------|
| Pure platelet rich plasma         | Cell separator PRP, Vivostat PRF or Anitua’s PRGF                      |
| Leukocyte and platelet rich plasma | Curasan, Regen, Plateletx, SmartPReP, PCCS, Magellan or GPS PRP         |
| Pure platelet rich fibrin         | Fibrinet                                                               |
| Leukocyte and platelet rich fibrin| Choukroun’s PRF                                                        |

GPS, gravitational platelet separation; PCCS, platelet concentrate collection system; PRP, platelet rich fibrin; PRGF, platelet released growth factor; PRP, platelet-rich plasma.

and histological evaluation at 4 and 12 weeks with PRP in poly-lactic-glycolic acid.

**Tendon healing**

In a rabbit patellar tendon defect model, a single application of PRP was found to improve tendon defect healing by over-expression of IGF-1 [21]. Another study with equine tendons cultured in 100% PRP demonstrated enhanced gene expression of the matrix molecules collagen type I (COL1A1), collagen type III (COL3A1), cartilage oligomeric matrix protein (COMP) without an associated increase in the catabolic molecules MMP-3 and MMP-13 [46]. Some studies implicate PRP in enhancing neovascularization that further accelerates the healing process and promotes better histological quality scar tissue formation [47]. Lyras and colleagues [48] found a significant improvement in the mechanical properties of the regen-erated tendon in the PRP-treated group at 14 days, returning to baseline at 28 days, concluding that PRP has a strong effect in the early phases of tendon healing.

**Human clinical studies - applications in different pathological conditions**

The use of platelet rich plasma is still a relatively new treatment for orthopedic patients, although physicians are increasingly using PRP as an alternative to medication and surgery. Although much of the literature has a low evidence level - that is, case-control studies, case series and anecdotal case reports - there are a handful of RCTs on this subject. Pooled results from six randomized trials from different clinical applications of PRP in a recent meta-analysis [49] found no significant benefit up to 24 months with the use of PRP. The observed trend towards benefit with PRP use still remains questionable.

**Articular cartilage repair**

Sanchez and colleagues [33] describe a case report where autologous plasma rich in growth factors was injected into the area between the defect and the fixed fragment after the arthroscopic fixation of a large (>2 cm) loose chondral body from the medial femoral condyle in the knee of an adolescent soccer player. Despite the poor prognosis, complete articular cartilage healing was reported to be accelerated and functional outcome was remarkable, allowing a quick return to symptom-free athletic activity.

**Achilles tendon repair**

A case-control (level 3 evidence) study comparing open suture repair in conjunction with a preparation rich in growth factors (PRGF) in six athletes and retrospective comparison with six matched athletes who had a conventional surgical procedure for a complete Achilles tendon tear reported that athletes receiving PRGF recovered their range of motion earlier, showed no wound complication, and took less time to resume gentle running and training activities [50]. The cross-sectional area of the PRGF-treated tendons increased less, and TGF-β1, PDGF-αβ, VEGF, EGF and HGF significantly correlated with the number of platelets, suggesting that operative management of tendons combined with the application of autologous PRGF promoted enhanced healing and functional recovery.

On the contrary, a RCT of 30 patients with surgical repair of Achilles tendon reported no additional benefit of PRP over standard treatment [51]. The Achilles Tendon Total Rupture Score was lower in the PRP group, possibly suggesting a detrimental effect. A variation in elasticity modulus was observed, although it was unclear how early biomechanics could be connected to late clinical results. In another independent RCT of patients with chronic Achilles tendinopathy, a PRP injection was no better in improving pain and activity compared to saline injection [52].

**Anterior cruciate ligament repair**

A human study examined the effects of local platelet gel application on graft healing in bone tunnels and the ligamentization process after anterior cruciate ligament (ACL) reconstruction [53]. Locally applied platelet gel enhanced early revascularization of the graft in the osteoligamentous interface zone after ACL reconstruction. After 4 to 6 weeks, the platelet gel-treated group demonstrated a significantly higher level of vascularization in the osteoligamentous interface, with no evidence of revascularization in the intra-articular part of the graft in either group.

Vogrin and colleagues [54] in their study evaluating the effects of local application of platelet-leukocyte gel on knee stability after ACL reconstruction surgery reported significantly better anteroposterior knee stability at 6 months with PRP treatment.
Arthroplasty
A retrospective case control study has described the effect of autologous platelet gel applied to exposed tissues, synovium and the lining of the wound at closure following total knee arthroplasty [55]. Interestingly, the patients receiving platelet gel during surgery had less postoperative blood loss as measured by differences in preoperative and postoperative hemoglobin on day 3, lesser narcotic requirement and a higher range of motion prior to discharge than their control counterparts. Researchers concluded that the application of autologous platelet gel might lead to improved hemostasis, better pain control and a shortened hospital stay.

Everts and colleagues [56] obtained similar results in their study, where autologous platelet gel and fibrin sealant application during unilateral total knee arthroplasty was found to help reduce the peri-operative incidence of blood transfusions.

Epicondylitis
A Dutch RCT comparing the effectiveness of leukocyte-enriched PRP to standard corticosteroid treatment for lateral epicondylitis found the former reduced visual analog scale pain scores and Disabilities of the Arm, Shoulder and Hand (DASH) outcome scores by 25% [57]. When baseline visual analog scale and DASH scores were compared with the scores at 2-year follow-up, both groups significantly improved across time. However, the DASH scores of the corticosteroid group returned to baseline levels, while those of the PRP group remained significantly improved. Furthermore, there were no adverse effects related to the use of PRP in 51 patients.

Another independent study found a single platelet-rich plasma injection improved pain and function scores in 29 patients who failed to improve with 6 months of corticosteroid treatment [58].

Osteoarthritis
A study of 261 patients with symptomatic osteoarthritis of the knee, following intra-articular infiltration of PRGF, reported improvement in the function and quality of life measured by osteoarthritis-specific and general clinical assessment instruments at 6-month follow-up [59]. These findings suggest PRGF is a potential therapy for osteoarthritis. Filardo and colleagues [60] in a study of 91 patients observed that treatment with PRP injections reduced pain and improved knee function and quality of life with short-term efficacy; the greatest effect was observed at 12 months but diminished by 24 months.

Rotator cuff repair
Rendelli and colleagues [61] in their prospective, randomized, controlled, double blind study of 53 patients with complete rotator cuff tear observed a lower pain score at 3, 7, 14 and 30 days after surgery with the local application of autologous PRP. Additionally, the Simple Shoulder Test, University of California Los Angeles (UCLA), Constant scores and strength in external rotation as measured by a dynamometer were significantly higher in the treatment group than the control group at 3 months after surgery. However, no long-term noticeable differences were noted after 6 months. Follow-up magnetic resonance imaging showed no significant difference in the healing rates of the rotator cuff tear in both groups. The long-term results of subgroups of grade 1 and 2 tears suggest that PRP positively affected cuff rotator healing. On the contrary, Castricini and colleagues [62] in their RCT did not find the use of autologous platelet-rich fibrin matrix beneficial for augmentation of a double-row repair of small or medium rotator cuff tears. These results, however, are applicable only for small and medium rotator cuff tears, with an undetermined possibility of benefit of platelet-rich fibrin matrix for large and massive rotator cuff tears.

Sub-acromial decompression
A study of 40 patients evaluating the effect of platelet-leukocyte gel produced from PLRP on the postoperative recovery of patients undergoing open sub-acromial decompression observed faster recovery, earlier return to daily activities and decreased pain medication requirement in the platelet-leukocyte gel-treated group [63].

Safety profile
Since PRP is prepared from autologous blood, theoretically there are minimal risks for disease transmission, immunogenic reactions or cancer [64]. Based on the pioneering and long-term clinical experience of the oral-maxillary field with PRP, and thousands of patients being treated so far, the use of PRP is considered to be safe [65,66]. In the musculoskeletal field, although no long-term outcome studies with PRP exist, a large number of patients have been treated worldwide. Wang-Saegusa and colleagues [59] in their study of over 800 patients reported no adverse effects following injection of plasma rich in growth factors (PRGF) into the knee joint at 6 months.

Adverse effects are rare but, as with any injection, there is always a small risk of injection site morbidity, infection or injury to nerves or blood vessels. Scar tissue formation and calcification at the injection site have been reported [10]. Hypersensitivity to the bovine thrombin used for activation was concerning and is therefore avoided in modern techniques. Rarely, development of antibodies against clotting factors V and IX leading to life-threatening coagulopathies have been reported [67-69]. To date, there is no compelling evidence of any systemic effects of a local PRP injection. Some probable arguments
for these considerations include the limited need of PRP injections in clinics (as PRP is not chronically administered) and the short in vivo half-lives and local bioavailability of growth factors produced by PRP. In our review of literature we did not come across RCTs that have comprehensively evaluated the safety of PRP treatment.

Practical aspects and issues

Physicians thinking about using PRP in their practices should consider the time commitment it takes to learn and employ the technique, insurance coverage, informed consent and sporting regulations.

Role of insurance companies - billing and reimbursement

A logistical problem with providing PRP injections is that most insurance plans do not reimburse for this procedure. The cost of single treatment ranges from $200 to $500. The Current Procedural Terminology code for PRP injection, which was implemented in 1 July 2010, is 0232T (injection(s), platelet-rich plasma, any tissue, including image guidance, harvesting, and preparation when performed). This is a temporary code used for emerging technologies, services, and procedures that allows data collection to be used to document widespread use for Food and Drug Administration approval. This code also includes imaging guidance, harvesting, and preparation; therefore, these aspects of the procedure cannot be billed separately. The placement/injection of the cells into the operative site is an inclusive component of the operative procedure performed and not separately reported. Category III code 0232T can only be reported if the PRP injection is performed on a nonsurgical site or when performed unrelated to the surgical site.

Anti-doping regulations and platelet-rich plasma

The World Anti-Doping Agency (WADA) is an international independent agency that regulates dopingsfree sport by enforcing the Anti-Doping Code. WADA monitors the current literature, conducts comprehensive consultations and, upon agreement by expert panels, releases an annual list of prohibited substances. PRP, after being mentioned in its list of prohibited items for the first time in 2010 [70], was subsequently deleted from the 2011 Prohibited List [71]. Initially, only intramuscular PRP injections were prohibited, whereas all other routes of administration, such as intra-articular or intra- or peri-tendinous, were permitted and only required a declaration of use. This restriction was due to the concern of the WADA List Expert Group that growth factors contained in PRP may stimulate muscle satellite cells and increase muscle size and strength beyond normal healing. Specific purified or recombinant growth factors (for example, IGF-1, VEGF, PDGF) are explicitly prohibited except when part of platelet-derived preparations from the centrifugation of autologous whole blood.

The different PRP formulations and treatments existing nowadays have not been found to increase muscle growth beyond return to a normal physiological state. Some animal studies do show faster muscle regeneration and recovery to full function following experimentally induced injury, but no enhancement of performance beyond normal has been reported [6,72,73]. It has now generally been agreed by the sporting regulation bodies that the use of PRP injections for therapeutic purposes only does not violate the spirit of sport. Hence, PRP use is currently permissible by all routes of administration. WADA continues to review PRP usage in the light of new scientific information as it becomes available.

Limitations

The composition of PRP varies from patient to patient and may also vary with the device used to prepare it, the time and method of storage until used and whether it interacts with other biologics or materials. Due to these inherent inconsistencies, the current use of PRP in orthopedics is patient-driven and based on anecdotal reports of its use in elite athletes. Platelet rich plasma may enhance soft-tissue repair, especially for tendons, though it may inhibit bone formation [74]. Some physicians use PRP as a way to provide and/or promote growth factors and cytokines during tissue repair. PRP can lead to fibrous connective tissue and scar formation. Additionally, PRP is not osteoinductive. This field is still blurred by controversial results from different studies, and a definite direction remains elusive [75]. The American Academy of Orthopaedic Surgeons, at their 2011 PRP Forum, concluded that ‘PRP is an option that yet remains unproven’ [76].

Conclusion

PRP was a major initial step that led researchers to consider bone marrow aspirates and stem cells in the growing field of ‘orthobiologics’. Available data suggest that PRP may be helpful in enhancing soft-tissue repair, particularly for tendon and wound healing. The clinical application of PRP in arthritis and bone repair remains controversial. PRP is most helpful in chronic tendinopathies, such as tennis elbow and patellar tendonitis, as well as certain revision surgeries with compromised healing milieu.

The role of PRP in the directed stimulation of musculoskeletal and mesenchymal stem cells is being widely investigated and shows promise as a short-term use agent that may assist in the simultaneous healing of several musculoskeletal tissues after trauma or elective surgery. PRP is not uniformly successful as an adjuvant to bone grafting procedures. The potential role of PRP in
healing musculoskeletal injuries, especially in elite athletes, is an exciting frontier that may eventually lead to superior therapies, but a healthy amount of caution should be exercised until clinical evidence is established. Continued research is required to optimize its preparation and use during surgery, to evaluate relative effectiveness of various techniques and to determine the best ways to use it to improve healing.

This article is part of the series on Cutting edge topics and advances in orthopaedic research, edited by Edward Schwarz and Jay Lieberman. Other articles in this series can be found at http://arthritis-research.com/series/orthopaedics

Abbreviations
ACL, anterior cruciate ligament; BMP, bone morphogenetic protein; DASH, Disabilities of the Arm, Shoulder and Hand; EGF, epidermal growth factor; FGF, fibroblast growth factor; HA, hyaluronic acid; HGF, hepatocyte growth factor; IGF, insulin-like growth factor; IL, interleukin; MMP, matrix metalloproteinase; PDGF, platelet-derived growth factor; PLP, platelet-leukocyte-rich plasma; PRG, platelet released growth factor; RBP, platelet-rich plasma; RCT, randomized controlled trial; TGF, transforming growth factor; VEGF, vascular endothelial growth factor; WADA, World Anti-Doping Association.

Competing interests
The authors declare that they have no competing interests. No funding was received for preparation of this manuscript.

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