Ginsenoside Rb1 improves intestinal aging via regulating the expression of sirtuins in the intestinal epithelium and modulating the gut microbiota of mice

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Intestinal aging seriously affects the absorption of nutrients of the aged people. Ginsenoside Rb1 (GRb1) which has multiple functions on treating gastrointestinal disorders is one of the important ingredients from Ginseng, the famous herb in tradition Chinese medicine. However, it is still unclear if GRb1 could improve intestinal aging. To investigate the function and mechanism of GRb1 on improving intestinal aging, GRb1 was administrated to 104-week-old C57BL/6 mice for 6 weeks. The jejunum, colon and feces were collected for morphology, histology, gene expression and gut microbiota tests using H&E staining, X-gal staining, qPCR, Western blot, immunofluorescence staining, and 16S rDNA sequencing technologies. The numbers of cells reduced and the accumulation of senescent cells increased in the intestinal crypts of old mice, and administration of GRb1 could reverse them. The protein levels of CLDN 2, 3, 7, and 15 were all decreased in the jejunum of old mice, and administration of GRb1 could significantly increase them. The expression levels of Tert, Lgr5, mKi67, and c-Myc were all significantly reduced in the small intestines of old mice, and GRb1 significantly increased them at transcriptional or posttranscriptional levels. The protein levels of SIRT1, SIRT3, SIRT4, and SIRT7 were all significantly increased in the jejunum and colon of old mice, and administration of GRb1 could reverse them. The numbers of bacteria were reduced in the feces of old mice, and administration of GRb1 could reverse them. The results of this study showed that administration of GRb1 could improve intestinal aging via regulating the expression of sirtuins in the intestinal epithelium and modulating the gut microbiota of mice.
and SIRT6 were all reduced in the jejunum of old mice, and GRβ1 could increase the protein levels of them. The 16S rDNA sequencing results demonstrated the dysbiosis of the gut microbiota of old mice, and GRβ1 changed the composition and functions of the gut microbiota in the old mice. In conclusion, GRβ1 could improve the intestinal aging via regulating the expression of Sirtuins family and modulating the gut microbiota in the aged mice.

**KEYWORDS**
Ginsenoside Rb1, gut microbiota, intestinal aging, sirtuin, intestinal integrity

### Introduction

The growing of aging societies is one of the major challenges for today’s medical science (Friedrich, 2019). The nutrients absorption ability of the intestines becomes impaired with age (Pénzes, 1984) and causes the vulnerability to disease and the physical weakness of the elderly peoples (Ben Othman et al., 2020). It was also found that the morphology of jejunum changed in old rats (Hassan et al., 2017). Therefore, it would be meaningful to develop drugs for improving intestinal aging.

Ginsenoside Rb1 (GRβ1) is the important ingredient from Panax ginseng Meyer which is the famous herb in traditional Chinese medicine (Lin et al., 2022). The Panax ginseng has been widely used to treat many kinds of disease. Recent study showed that the doxorubicin-induced early cancer therapeutics-related cardiac dysfunction and early decline in left ventricular ejection fraction in breast cancer patients can be protected through prophylactic Panax ginseng supplementation (Hamidian et al., 2022). The lifespan of Drosophila is extended with the treatment of total ginsenosides (TGGR), the main active components in Panax ginseng (Zhao et al., 2022b). Many types of ginsenosides have been demonstrated to have neuroprotective effects (Zhao et al., 2022a). There are around 200 ginsenosides have been detected from ginseng and GRβ1 is one type of major ginsenosides (Zhao et al., 2022a; Hyun et al., 2022).

GRβ1 has been reported to have multiple functions in various diseases. It can be used to treat obesity, hyperglycemia and diabetes through multi-targets (Zhou et al., 2019; Xiong et al., 2010). GRβ1 also can ameliorate diabetic kidney podocyte injury via inhibiting the activity of aldose reductase (He et al., 2022). It was also found that GRβ1 can reduce the myocardial ischemia/reperfusion injury via inhibiting cardiomyocyte autophagy through the PI3K/AKT/mTOR pathway (Qin et al., 2021). GRβ1 also has anti-aging effect (Cheng et al., 2005), but the related mechanism is unclear. GRβ1 can be used to treat many kinds of gastrointestinal disorders. It improves colitis in mice via alleviating endoplasmic reticulum (ER) stress through activating Hrd1 signaling pathway (Dong et al., 2021). GRβ1 also can reduce ischemia/reperfusion-induced intestinal injury via activating PI3K/AKT/Nrf2 pathway (Chen et al., 2019). It was also found that GRβ1 can promote the intestinal epithelial would healing of rats via activating ERK and Rho signaling (Toyokawa et al., 2019). GRβ1 can protect the peritoneal air exposure caused intestinal mucosa damage in rats (Zhou et al., 2016). However, it is still unclear if GRβ1 can improve intestinal aging.

There are many genes have been reported to be related to the aging of intestines and other tissues. Stem cell exhaustion is one of the hallmarks of aging (López-Otín et al., 2013). Lgr5 is the mark gene of intestinal stem cells (Lei et al., 2012; Baghdadi et al., 2022). Telomerase plays important role in the intestinal stem cells and TERT is the important telomerase subunit (Hoffmeyer et al., 2012). Sirtuins, including Sirt1-7 in mammals, have been demonstrated to play important roles in maintaining the longevity of various tissues (Gámez-García and Vazquez, 2021; Yang et al., 2021a; Watroba and Szkiewicz, 2021). Hence, it would be very meaningful to explore if GRβ1 could regulate the expression of these genes in the intestines of aging mice.

Many studies have demonstrated the changes of the composition and functions of gut microbiota with aging (Ishaq et al., 2021; Niu et al., 2021; Ruiz-Gonzalez et al., 2022). It was reported that specific bacterial community pattern and signature taxa are related to longevity of people (Ren et al., 2021). The dysbiosis of gut microbiota is also associated with age-related disorders (Sharma, 2022). Relationships between gut microbiota and age-related macular degeneration have been found (Lima-Fontes et al., 2021). Gut microbiota-derived pro-inflammatory neurotoxins have been detected in brain cells and tissues of aged people with Alzheimer’s disease (Lukiw et al., 2021; Zhao et al., 2021). Gut microbiota dysbiosis has also been found to promote the age-related atrial fibrillation via activating NLRP3-inflammasome (Zhang et al., 2021). GRβ1 can improve glucose and lipid metabolic disorders through regulating gut microbiota of high fat diet induced obesity mice (Yang et al., 2021b; Bai et al., 2021). GRβ1 also can be converted into compound K by the gut microbiota to prevent inflammatory-associated colorectal cancer (Yao et al., 2018). However, it still needs to explore whether GRβ1 could improve intestinal aging via modulating gut microbiota.

In the present study, we reported the function and mechanisms of GRβ1 on improving the intestinal aging of old mice. Our work encouraged the exploration of drugs for prevention and treatment of age-related diseases.
Materials and methods

Mice

All animal experimental procedures were approved by the Experimental Animal Ethics Committee of Guangdong Pharmaceutical University. Female C57BL/6 mice (5-week-old) purchased from Hunan Lex Jingda Laboratory Animal Co., Ltd. (Changsha, Hunan Province, China), were housed in the specific pathogen-free (SPF) animal facility, at 25°C, 60%–65% humidity, 12 h light-dark cycle, with free access to water and food. At the age of 104-week-old, the mice were randomly divided into three groups, 10 mice in each group. The Old + GRb1 group was administrated with GRb1 (50mg/kg; Meilunbio, Dalian, China; MB6856-1) intragastrically once a day. The GRb1 was diluted in 0.5% CarboxyMethylCellulose-Na (CMC-Na) (Tianjin Zhiyuan Chemical Reagent Co., Ltd., Tianjin, China). The Old group was administrated with the corresponding volume of 0.5% CMC-Na intragastrically once a day. Resveratrol (RSV; Meilunbio, Dalian, China; MB5267-1) was used as the positive drug. The Old + RSV group was intragastrically administrated with RSV (50 mg/kg) diluted in 0.5% CMC-Na once a day. The 8-week-old mice in Youth group was used as control, and they were also administrated with the corresponding volume of 0.5% CMC-Na intragastrically once a day. After 6 weeks of administration, the intestines were collected (Figure 1A).

H&E staining and X-gal staining

The H&E staining was performed as previously (Lei et al., 2021b). Briefly, intestinal tissues were fixed in 4% paraformaldehyde at 4°C for overnight, then dehydrated, embedded in paraffin and sectioned. 4-µm-thick sections were stained with hematoxylin (H9627, Sigma-Aldrich) for 3 min, and then followed with eosin (E4009, Sigma-Aldrich) for 20 s at room temperature.

For X-gal staining, intestinal tissues were embedded in optimal cutting temperature compound (OCT) (Sakura Finetek) and sectioned. 7-µm-thick frozen sections were stained according to the manufacturer’s protocols for Senescence Detection Kit (Abcam, ab65351).

Images for H&E staining and X-gal staining were got using the Olympus DP74 microscope.

Immunofluorescence staining

The immunofluorescence staining was performed as previously (Lei et al., 2021b). The intestinal tissues were fixed in 4% paraformaldehyde at 4°C for overnight, then dehydrated, embedded in OCT compound and sectioned. 7-µm-thick frozen sections were first boiled in 10 mM citric acid (Merck) at pH 6.0 for 5 min, then exposed in goat serum blocking buffer (ZSGB-BIO, ZLI-9056) to block nonspecific sites for 1h at room temperature, following incubated with primary antibodies in blocking buffer at 4°C for overnight, and then with secondary antibodies for 1h at room temperature. The primary and
secondary antibodies were listed in Supplementary Table 1. Images were got by using Olympus confocal microscope.

**qRT-PCR**

Total RNA was extracted from each jejunal and colonal tissue using Trizol reagent (T9108, Takara Bio, Inc.), then subjected to reverse transcription via the PrimeScript™ RT Reagent kit (RR047A, Takara Bio, Inc.) at 37°C for 15 min and then 85°C for 5 s. The qPCR was conducted through the SYBR Premix Ex Taq kit (RR820A, Takara Bio, Inc.) via the LightCycler 480II System (Roche, Inc.). The processes of cycling were: 95°C for 30 s; followed 40 cycles of 95°C for 5 s, then 60°C for 20 s and 65°C for 15 s. Mouse GAPDH was used as the internal reference. All primers were listed in Supplementary Table 2.

**Western blot**

Jejunal and colonal tissues of mice were lysed using the Radio-Immunoprecipitation Assay lysis buffer (MA0151, Dalian Meilun Biotechnology co., Ltd., Dalian, China), centrifuged at 13,680 x g, 4°C, for 30 min, then the supernatant was collected. Protein concentration was measured by the BCA kit (P0011, Beyotime, Shanghai, China). Equal amounts of protein (40 μg) were separated through the SDS-PAGE, subsequently transferred to a PVDF membrane. The PVDF membrane was blocked using the 5% skimmed milk (0040895, Biosharp, Hefei, China) in TBST buffer at room temperature for 1 h, incubated with primary antibodies in 4°C for overnight, and then incubated with HRP (horseradish peroxidase)-labeled secondary antibodies, the signals were detected via the enhanced chemiluminescence reagent. The primary and secondary antibodies were listed in Supplementary Table 3. The quantification of western blot bands was analyzed using the Lane 1d software (version 5.1.0.0; SageCreation).

**The 16S rRNA gene analysis**

Fecal samples were quickly collected and frozen in the liquid nitrogen and stored at −80°C. The extraction of fecal bacterial DNA, PCR amplification of 16S rRNA genes, sequencing, and analysis were performed by the Gene Denovo Biotechnology Company (Guangzhou, China). The experimental procedures were performed as previously (Lei et al., 2021a).

**Statistical analysis**

Statistical differences were determined via the SPSS software (version 25.0; IBM Corp.). Mean ± SE was used to express data. One-way ANOVA was performed between two groups. p-value<0.05 was considered to be significant.

**Results**

**GRb1 improved the aging state of intestines of old mice**

After 6 weeks of the administration of GRb1 or RSV, 7 (70%) mice survived in each of Old and Old + GRb1 groups, 9 (90%) mice survived in Old + RSV group, and all the mice survived in the Youth group. The intestines of old mice were significantly longer than the Youth group, and they are shorter but not significant in mice of the Old + GRb1 group compared to the Old group (Figures 1B,C). The numbers of cells in crypts of jejunum from old mice decreased compared to the Youth group, and it was increased after administration of GRb1 or RSV (Figure 1D). The numbers of cells in crypts of the colon of old mice were also lower than that of yang mice, and the administration of GRb1 or RSV could also improve it (Figure 1E).

The increase of cellular senescence is another hallmark of aging (López-Otín et al., 2013). Therefore, senescence-associated beta-galactosidase (X-gal) staining was next performed. The accumulation of senescent cells increased in crypts of jejunum from the Old group compared to young mice, and GRb1 or RSV could reduce them (Figure 1F; Supplementary Figure 1A). The senescence-associated signal was stronger in the colon of old mice than the Youth group, and it became weak and reduced after administration of GRb1 or RSV (Figure 1G; Supplementary Figure 1B). The intestinal stem and progenitor cells are localized in the crypts of the intestines. So, the increase of the numbers of the X-gal stained cells in the crypts of the intestines indicated the aging of the intestinal stem and progenitor cells of the old mice. Hence, the administration of GRb1 or RSV could improve the aging of the intestinal stem and progenitor cells of these mice. These results demonstrated that GRb1 could improve the aging state of intestines from old mice.

**GRb1 improved the intestinal integrity of old mice**

The increase of the permeability of the intestinal barrier has been reported in both aged human and animals (Tran and Greenwood-Van Meerveld, 2013; Parrish, 2017; Li et al., 2021), indicating the impaired intestinal integrity with aging. Hence, the protein levels of CLDN 1, 2, 3, 7, and 15 which are abundant components of tight junctions (TJs) in the intestinal epithelium (Lei et al., 2012; Lei et al., 2020) were first checked. CLDN 3, 7, and 15 were all significantly reduced in the jejunum of Old group compared to young mice, and the administration of
FIGURE 2
GRb1 improved the expression and localization of junctional proteins in the intestines of old mice. (A,B) Images of Western blot bands of CLDN 1, 2, 3, 7, 15, and EpCAM in the (A) jejunum and (B) colon. (C,D) Representative images of immunofluorescence staining with antibodies to CLDN 7 of frozen sections of (C) jejunum and (D) colon. (E,F) Representative images of immunofluorescence staining with antibodies to EpCAM of frozen sections of (E) jejunum and (F) colon. Scale bar, 50 μm.
GRb1 increased the expression of them (Figures 2A; Supplementary Figures 2C–E). CLDN 2 was also reduced in the jejunum of old mice, and it was also increased after GRb1 administration, although these changes were not significant (Figure 2A; Supplementary Figure 2B). RSV also could improve the expression of CLDN 3, 7, and 15, but the level of CLDN 2 had no significant change in the Old + RSV group (Figures 2A; Supplementary Figures 2B–E). Immunofluorescence staining results showed that the localization of CLDN 7 was still normal in the jejunum from Old group, but the expression level of it was significantly lower in Old group than the Youth, Old + GRb1 and Old + RSV groups (Figure 2C). The protein level of CLDN 1 had no significant difference in the jejunum of mice among the Youth, Old, Old + GRb1 and Old + RSV groups (Figure 2A; Supplementary Figure 2A). The protein level of EpCAM which is essential to maintain the functional tight junctions in the intestinal epithelium via recruiting proteins of Claudins (Lei et al., 2012; Wu et al., 2013) was significantly lower in the jejunum of Old group than the Youth group, and administration of GRb1 could not improve it (Figure 2A; Supplementary Figure 2F). The administration of RSV could significantly increase the protein level of EpCAM in the jejunum of old mice (Figure 2A; Supplementary Figure 2F). However, the localization of EpCAM had no significant difference in the jejunum of mice among the four groups (Figure 2E).

CLDN 15 was also lower in the colon of Old group than the Youth group, although the decrease was not significant (Figure 2B; Supplementary Figure 3E). The administration of GRb1 or RSV could significantly increase the protein level of CLDN 15 in the colon of old mice (Figure 2B; Supplementary Figure 3E). CLDN 1 and 2 were all increased in the colon of Old group compared to the Youth group, and CLDN 2 was significantly increased in the Old + GRb1 and Old + RSV groups (Figure 2B; Supplementary Figures 3A,B). CLDN 3 and 7 were all significantly increased in the colon of old mice compared to the Youth group, and RSV could also increase them in the colon of old mice but not significantly (Figure 2B; Supplementary Figures 3C, D). The immunofluorescence staining results confirmed that the localization of CLDN 7 had no significant difference in the colon among the four groups (Figure 2D). The expression and localization of EpCAM had no significant difference in the colon among the Youth, Old, Old + GRb1 and Old + RSV groups (Figures 2B,F; Supplementary Figure 3F). These results demonstrated that GRb1 could improve the integrity of intestinal epithelium of old mice.

**GRb1 improved the function of intestinal stem and progenitor cells of old mice**

Tert was significantly reduced in the jejunum of old mice at both mRNA and protein levels compared to the Youth group (Figures 3A,B; Supplementary Figure 4A,B). The administration of GRb1 or RSV could not change the transcription of Tert in the jejunum of old mice (Figure 3A). However, both GRb1 and RSV could evidently increase the reduced TERT protein in the jejunum of old mice (Figure 3B; Supplementary Figure 4A). The protein level of TERT was also significantly lower in the colon of Old group than the Youth group, but GRb1 and RSV could not improve it (Figure 3C; Supplementary Figure 4B). The transcriptional level of Lgr5 was significantly reduced in the jejunum of old mice compared to the young mice, and it was increased in the Old + GRb1 group although the increase was not significant (p = 0.061) (Figure 3A). RSV could not increase the mRNA level of Lgr5 in the jejunum of old mice (Figure 3A). The transcriptional levels of other intestinal stem cell related genes, including Olfn4, Ascl2, Rnf43, and Sp5, showed no significant difference in the jejunum from Old and Youth groups (Figure 3A). However, RSV could increase Ascl2 and Sp5 in the jejunum of old mice (Figure 3A).

The proliferative ability of intestinal stem and progenitor cells was checked via testing the expression of mKi67 in the intestines of mice. Compared to the young mice, the mRNA level of mKi67 was significantly reduced in the jejunum of the Old group, but GRb1 or RSV could not improve it (Figure 3A). The protein level of Ki67 was also significantly decreased in the jejunum of the Old group compared to the Youth group, and GRb1 could evidently increase it (Figure 3D; Supplementary Figure 4C). GRb1 increased the numbers of Ki67 positive cells in crypts of jejunum of the old mice (Supplementary Figure 5A). The administration of GRb1 also increased the reduced Ki67 protein in the colon of old mice, although the increase was not significant (p = 0.06) (Figure 3E; Supplementary Figures 4E, 5B). The protein level of c-Myc which is responsible for the transcription of pro-proliferative genes (Ruan et al., 2021) was significantly reduced in the jejunum of old mice, and GRb1 could significantly improve it (Figure 3D; Supplementary Figure 4D). The protein level of c-Myc was evidently higher in the colon of old mice than the Youth group, and it was decreased in the colon of Old + GRb1 and Old + RSV groups but not significantly (Figure 3E; Supplementary Figure 4F). These results indicated that GRb1 could improve the function of intestinal stem and progenitor cells.

**GRb1 regulated the expression of sirtuins in the intestines of old mice**

The mRNA levels of SirT4 and SirT6 were all significantly decreased in the jejunum of old mice compared to the Youth group, and GRb1 or RSV could significantly increase the transcription of SirT6 but not SirT4 in old mice (Figure 4A). There was no significant difference of the transcriptional levels of SirT1, SirT2, SirT3, SirT5, and SirT7 between young and old mice (Figure 4A). However, the mRNA levels of SirT2 and SirT7 were all
FIGURE 3
GRb1 was effective to improve the function of intestinal stem cells of old mice. (A) Relative mRNA expression levels of Tert, Lgr5, miK67, Olfm4, Ascl2, Rnf43, and Sp5 in the small intestines. (B,C) Images of western blot bands of TERT in the (B) jejunum and (C) colon. (D,E) Western blot results of Ki67 and c-Myc from the (D) jejunum and (E) colon. *p < 0.05, ***p < 0.001, compared with the Youth group; **p < 0.01, compared with the Old group.
significantly increased in the jejunum of Old + RSV group compared to the Old group (Figure 4A). The protein levels of SIRT1, SIRT3, SIRT5, and SIRT6 were all lower in the jejunum of old mice than the Youth group, and GRb1 could rescue SIRT1 and SIRT6 in the jejunum of old mice (Figure 4B; Supplementary Figures 6A, C, E, F). The administration of GRb1 or RSV could also increase the expression of SIRT3 and SIRT7 in the jejunum of old mice, but the increase was not significant (Figure 4B; Supplementary Figures 6C, G). SIRT2 and SIRT4 were significantly increased in the jejunum of Old mice, but the protein level of SIRT4 was significantly reduced after administration of GRb1 or RSV (Figure 4B; Supplementary Figures 6B, D). However, the administration of GRb1 or RSV could not reduce the protein level of SIRT2 in the jejunum of old mice (Figure 4B; Supplementary Figure 6B).

The protein levels of SIRT1 and SIRT7 showed no significant difference among the Youth, Old, Old + GRb1 and Old + RSV groups (Figure 4C; Supplementary Figures 7A, G). The protein levels of SIRT2, SIRT3, SIRT4, SIRT5, and SIRT6 were all significantly increased in the colon of old mice compared to the Youth group, and administration of GRb1 or RSV could reduce SIRT2 in the colon of old mice (Figure 4C; Supplementary Figures 7B–F). These results indicated that GRb1 might improve the aging of intestines via regulating the expression sirtuins at both transcriptional and post-transcriptional levels.

**GRb1 changed the composition and function of gut microbiota of old mice**

The 16S rRNA gene sequence was performed to analyze the composition and functions of the gut microbiota in mice (https://www.ncbi.nlm.nih.gov/sra/PRJNA856886). The Shannon rarefaction curves for every group had reached the saturated platform (Figure 5A), and the principle coordinates analysis (PCoA) showed that the Youth and the Old groups could be clearly distinguished (Figure 5B). Analysis of similarity (ANOSIM) showed that the rank of the Old group was lower than the Youth group, and the rank of the Old + GRb1 and Old + RSV groups was higher than the Old group (Figure 5C). At the phylum level, the abundance of *Firmicutes* and *Tenericutes* was significantly increased in the Old group compared to the Youth group, and the abundance of *Firmicutes* was significantly reduced after administration of GRb1 (Figures 5D, E). The abundance of *Bacteroidetes* and *Verrucomicrobia* was significantly reduced in the Old group compared to the Youth group (Figures 5D, E). The abundance of *Proteobacteria* was significantly increased in the Old + GRb1 group compared to the Old group (Figures 5D, E). LEFse analysis showed there were 79 bacterial taxa differed in abundance between the Youth and Old groups, with 32 predominant for the Youth group and 47 predominant for the Old group (Supplementary Figures 8A, B). There were 57 bacterial taxa differed in abundance between the Old + GRb1 group and the Old group, with 21 predominant for the Old + GRb1 group and 36 predominant for the Old group (Supplementary Figures 8A, B). There were 24 bacterial taxa differed in abundance between the Old + RSV group and the Old group, with 12 predominant for the Old + RSV group and 12 predominant for the Old group (Supplementary Figures 10A, B). Compared to the Old group, there were three bacterial taxa predominant in all the three groups of the Youth, Old +
FIGURE 5
GRβ1 changed the relative abundance and functions of gut microbiota of old mice. (A) Shannon rarefaction curves for each group. (B) The PCoA analysis of the gut microbiota. (C) Analysis of similarity (ANOSIM) of the gut microbiota. (D) Relative abundance of the gut microbiota at phylum levels in mice. Different colors illustrated different flora. (E) Bar chart of proportional abundance of the gut microbiota at phylum levels in mice. (F) KEGG analysis showed the top 20 altered pathways of the gut microbiota. *p < 0.05, **p < 0.01, ***p < 0.001, compared with the Youth group; #p < 0.05, ##p < 0.01, compared with the Old group. PCoA, Principle coordinates.
GRb1 and Old + RSV, including Class Actinobacteria, Order Corynebacteriales and Family Corynebacteriaceae, and the Family Corynebacteriaceae belongs to the Order Corynebacteriales, the Order Corynebacteriales belongs to the Class Actinobacteria (Supplementary Figures 8A,B; Supplementary Figures 9A,B; Supplementary Figures 10A,B).

The top 20 altered pathways in the Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analysis were shown in Figure 5F. All the 20 pathways increased in the Old group, and GRb1 reduced four of them including “Biosynthesis of vancomycin group antibiotics,” “Pentose phosphate pathway,” “Streptomycin biosynthesis” and “Secondary bile acid biosynthesis,” although RSV could not reduce them. The other 16 pathways all increased after administration of GRb1 or RSV. These results indicated that the composition and functions of gut microbiota changed in old mice, and GRb1 might improve the intestinal aging partly through regulating the gut microbiota in old mice.

**Discussion**

We uncovered a new role of GRb1 on improving the intestinal aging of old mice (Figure 6). First, administration of GRb1 could increase the numbers of cells and reduce the accumulation of senescent cells in crypts of both small and large intestines from old mice. Then, GRb1 could improve the integrity of the intestinal epithelium via increasing the protein levels of the intestinal abundant Claudins in the intestinal epithelium of old mice. GRb1 could improve the function of intestinal stem and progenitor cells via upregulating the expression of Tert, Lgr5, mKi67, and c-Myc at transcriptional or posttranscriptional level in the small intestines of old mice. Then, it was demonstrated that GRb1 might improve intestinal aging through modulating the expression of members of Sirtuin family at both transcriptional and posttranscriptional levels in the intestines of old mice. Finally, 16S rDNA sequence results showed that
GRb1 could modulate the composition and functions of gut microbiota in old mice, and it might be one of the mechanisms of GRb1 on improving intestinal aging of old mice.

Intestinal barrier defects are one of the hallmarks of intestinal aging (Arnold et al., 2021). It was reported that the serum LPS level is significantly higher in old mice than the young control mice (Shin et al., 2020), indicating the gut leaky of the old mice. In the present study, CLDN 2, 3, 7 and 15 all decreased in the small intestines of old mice and CLDN 15 also decreased in the large intestines of old mice. We speculated that the reduction of these intestinal abundance Claudins might be the important reason for the defects of the intestinal barrier of the old mice. Tight junction proteins, such as ZO-1, Occludin and CLDN 1, has also been found reduced in the ileum of aged rats (Ren et al., 2018). CLDN 2 and 15 have been reported to have important functions on regulating the paracellular flow of Na⁺ from the intestinal submucosa to dominate the absorption of glucose, amino acids and fats (Tamura et al., 2011; Wada et al., 2013). Therefore, the decrease of CLDN 2 and 15 in the small intestines of old mice might affect the absorption of nutrients. GRb1 might promote the nutrients absorption of aged mice via increasing the levels of CLDN 2 and 15 in the small intestines of them.

Previous study showed that GRb1 can promote the differentiation of muscle stem cells (Go et al., 2020). Neural stem cells in rats of Alzheimer’s disease models are also improved by GRb1 (Zhao et al., 2018). In the present study, GRb1 could improve the function of intestinal stem and progenitor cells via upregulating the expression of Tert, Lgr5, mKi67, and c-Myc in the small intestines of old mice. Tert has been confirmed to specifically express in the intestinal stem cells (Breault et al., 2008; Itzkovitz et al., 2011; Montgomery et al., 2011; Muñoz et al., 2012). Overexpression of TERT improves the fitness of intestinal barriers and produces a system delay in aging of mice (Tomás-Loba et al., 2008). GRb1 enhanced the protein level of TERT in the small intestines of old mice indicating its effects on anti-aging of intestinal stem cells. Ki67 has been used as the cell proliferation marker in both normal and cancer tissues (Chakrabortysabong et al., 2021; Silva et al., 2022). The increase of Ki67 in both small and large intestines of old mice after administration of GRb1 demonstrated that the number of proliferative cells increased in the intestinal crypts of them. We speculated the increase of the proliferative cells should be the direct mechanism on the increase of cells in crypts of intestines of the GRb1 treated mice.

Members of sirtuin family play the key role in aging and age-related disease (Kaitsuka et al., 2021). In the present study, GRb1 could increase the protein levels of SIRT1, SIRT3, SIRT6, and SIRT7 in the small intestines of old mice. SIRT1 becomes a target for the prevention and treatment of age-related cardiovascular and cerebrovascular diseases since it has been confirmed to have important function on preventing vascular aging (Begum et al., 2021). Recent study reported that LARP7 can ameliorate cellular senescence and aging through enhancing the activity of SIRT1 (Yan et al., 2021). The increase of the expression or activity of SIRT3 can extend the life span of human (Silaghi et al., 2021; Rose et al., 2003). Recently, it was found that reduced SIRT3 abundance in mice can exacerbate age-related periodontal disease (Chen et al., 2021). The level and activation of SIRT6 have been found to be reduced in the aging brain (Stein et al., 2021). The overexpression of SIRT6 can extend the life span of both mice and Drosophila melanogaster (Reichman et al., 2021; Taylor et al., 2022). SIRT7 has been found to antagonize stem cell aging via stabilizing heterochromatin (Sun and Dang, 2020; Bi et al., 2020). Therefore, the upregulation of SIRTs should be considered as one of the important mechanisms on improving the small intestinal aging of old mice.

In the present study, the composition and functions of gut microbiota changed in the old mice after administration of GRb1. At the phylum level of gut microbiota, the ratio of Bacteroidetes/Firmicutes decreased in the Old group compared to the Youth group, and administration of GRb1 could improve it. Many studies confirmed the decrease of the ratio of Bacteroidetes/Firmicutes in ob/ob mice compared with normal control mice (Turnbaugh et al., 2006; Abenavoli et al., 2019). The dysbiosis of the gut microbiota can increase the intestinal permeability (Zhang et al., 2010). Therefore, GRb1 might enhance the integrity of the intestinal epithelium via improving the dysbiosis of the gut microbiota in old mice. Compared to the Old group, the Class Actinobacteria was predominant in the Youth, Old + GRb1 and Old + RSV groups. Actinobacteria have been confirmed to be the biosynthetic factories which produce various bioactive metabolites, and many of these bioactive metabolites can be developed as drugs for human (Azman et al., 2019; Hussain et al., 2020; Jose et al., 2021). The pathways for “Valine, leucine and isoleucine biosynthesis” and “Lysine biosynthesis” significantly increased in old mice after administration with GRb1. Lysine, valine, leucine and isoleucine are essential amino acids for human, so the increase of the biosynthesis of them should be good for the health of the old mice. Hence, we speculated that the regulating of the gut microbiota might be another important mechanism of GRb1 on improving the intestinal aging of the old mice.

**Conclusion**

In conclusion, GRb1 could improve the intestinal aging via regulating the expression of members of Sirtuin family in the intestinal epithelium at transcriptional or posttranscriptional levels and modulating the composition and functions of gut microbiota in the old mice (Figure 6).
Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found below: https://www.ncbi.nlm.nih.gov/sra/PRJNA856886.

Ethics statement

The animal study was reviewed and approved by the Experimental Animal Ethics Committee of Guangdong Pharmaceutical University.

Author contributions

Conceptualization, ZL and YY; methodology, LC, KL, and HR; validation, LC, QH. TL and YY; formal analysis, YY, SY, and QS; investigation, ZL, LC, and QH; resources, JG; data curation, ZL and JG; project administration, ZL and JG; funding acquisition, YY, ZL, and JG. All authors have read and agreed to the published version of the manuscript.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fphar.2022.991597/full#supplementary-material

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