Influence of Bacterial Attachment on Arsenic Bioleaching from Mine Tailings: Dependency on the Ratio of Bacteria-Solid Substrate

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Abstract

The present study investigates the bioleaching efficiencies of arsenic via contact and non-contact mechanisms. The attachment of Acidithiobacillus ferroxidans was restricted by a partition system comprising a semi-permeable membrane with a molecular weight cutoff of 12–14 kDa. The results were compared for two arsenic concentrations in the system (1.0% and 0.5% w/v) to maintain a homogeneous system. The overall bacterial performance was monitored by comparing total arsenic and...
iron concentrations, Fe ion speciation, pH, and solution redox potentials in flask bioleaching experiments over a period of 10 d. Our results indicated that bacterial attachment could increase arsenic extraction efficiency from 20.0% to 44.9% at 1.0% solid concentrations. These findings suggest that the bacterial contact mechanism greatly influences arsenic bioleaching from mine tailings. Therefore, systems involving two-step or non-contact bioleaching are less effective than those involving one-step or contact bioleaching for the efficient extraction of arsenic from mine tailings.

Key words: Bioleaching, arsenic removal, Acidithiobacillus ferrooxidans, solid concentration, bacterial attachment

1. Introduction

South Korea has >1000 abandoned mines with stockpiled mine tailings that are rich in heavy metals, including arsenic\(^1\). These tailings represent not only an environmental threat but also an economic opportunity, as arsenic is in demand in various industries.

Arsenopyrite mine tailings are the fine-ground residual wastes that remain after beneficiation of a mineral ore. Often, these residual wastes are rich in toxic metals like arsenic (As) and long-term weathering of mine tailings containing arsenic can negatively affect the environment and thus can present a critical risk in terms of arsenic poisoning. To lower the negative environmental impacts of arsenic or other heavy metals, alternative methods to stabilize metal species are required. Solubilization of arsenic from these residual wastes by bioleaching can be environment friendly, low cost and less labor-intensive process\(^2\). In recent years, bio-leaching has made significant progress in solubilization of metal from low grade resources and secondary wastes\(^3,4\). Bioleaching processes exploit the ability of microorganisms to solubilize metals by catalyzing reduction-oxidation reactions for a simplified metallurgical process\(^5\). Acidithiobacillus ferrooxidans is among one of the most prominent microorganisms in the biological leaching of (heavy)-metals\(^6,7\). These microorganisms predominantly oxidize ferrous (Fe\(^{2+}\)) to ferric (Fe\(^{3+}\)) ions under aerobic conditions. The strong oxidizing environment resulted from the production of Fe\(^{3+}\) ions in addition to acidic conditions that favor metal solubilization\(^8-11\). The bacteria interact with the mineral substrates by direct contact and non-contact mechanisms, which sometimes cooperate together\(^2,12-14\). In the contact mechanism, bacteria attach to the substrate surface (e.g., mine tailings) for the solubilization of metal\(^15,16\). In the non-contact mechanism, planktonic bacteria oxidize Fe\(^{2+}\) ions to Fe\(^{3+}\) ions in the solution medium. The resulting Fe\(^{3+}\) becomes the oxidizing agent that releases the desired metal ions from its substrate\(^15,16\). Following these simple mechanisms, different metals can be solubilize from several types of substrates (e.g., contaminated soils, low-grade resources, residual wastes) as long as Fe\(^{2+}\) is available for bacterial oxidation.

Up to now, no detailed work is observed on the importance of bacterial contact and non-contact mechanism for solubilization of arsenic from indigenous mine tailing. Therefore, to investigate the possibility of improving the arsenic solubilization efficiency of bioleaching, we studied the influences of the bacterial attachment and solid concentration on the arsenic solubilization from mine tailings. The contact and non-contact mechanisms were conducted by installing a partition system with a semi-permeable membrane, which prevents bacterial attachment to the mine tailings’ surface. The experiments were conducted with two solid concentrations (0.5% and 1.0% w/v) with constant temperature, shaking speed, and particle size. These findings will better elucidate the influences of each of the bioleaching mechanisms on arsenic solubilization efficiency from mine tailings and will give an insight into appropriate methodologies development for increasing metal bio-extraction efficiency.
2. Material and methods

2.1. Mine tailings

An oxidized mine tailings sample was obtained from the Janggun mine (from a Pb, Zn, and Ag-mining site in Bonghwa, Gyeongsang, South Korea)\(^7\). The sample was kindly provided by the Mine Reclamation Corporation. The particle sizes of the mine tailings in the bioleaching tests ranged from 63 to 74 μm (measured by a sieving technique) to determine contact and non-contact mechanisms except size effect. The chemical composition and mineralogy of the mine tailings were determined by inductively coupled plasma (ICP, Optima 7300DV, PerkinElmer) and X-ray diffraction (XRD, Bruker D8 HRXRD, Germany) analyses, respectively. The ICP analysis confirmed the high As content in the mine tailings (53200 mg/kg; see Table 1). Most of the As-associated mineral existed as arsenopyrite (FeAsS) (XRD analysis; see Fig. 1). Additionally, the Fe content in the mine tailing was determined to be 183364 mg/kg, and the presence of pyrite (FeS\(_2\)) was confirmed. FeS\(_2\) is a recognized suitable source of ferrous ions for Acidithiobacillus sp. during bioleaching processes\(^{17,19}\).

| Materials | \(d_{50}\) (μm)\(^a\) | \(\rho_0\) (g cm\(^{-3}\)) | Specific surface area (m\(^2\) g\(^{-1}\))\(^b\) | Chemical composition (mg/kg) |
|-----------|-----------------|-----------------|-----------------|-----------------|
| Mine tailing | 73 | 2.65 | 13 | As | Cd | Cu | Zn | Pb | Fe |
| | | | | 53200 | 154 | 4966 | 19461 | 9808 | 183364 |

\(^a\)Mean particle size measured by laser-diffraction scattering (Accusizer 780/SIS, PSS).

\(^b\)Specific surface area calculated by the BET model using nitrogen adsorption branch (ASAP2020, Micromeritics).

![XRD patterns of the raw mine tailings. It was determined that the raw mine tailings mainly consisted in quartz (squares), arsenopyrite (circles), and pyrite (diamonds).](image)

Fig. 1. XRD patterns of the raw mine tailings. It was determined that the raw mine tailings mainly consisted in quartz (squares), arsenopyrite (circles), and pyrite (diamonds).

2.2. Bacterial selection and culture

Acidithiobacillus ferrooxidans (KCTC 4515) was obtained from the Korea Research Institute of Bioscience and Biotechnology, South Korea. The culture medium for the bacteria was prepared with reference to DSMZ medium 882\(^20\). The initial pH value was adjusted to 1.8 with 10 N H\(_2\)SO\(_4\) (Fisher Scientific), and the speed and temperature of the shaker (SIF-5000R, JEIO TECH, Seoul, Republic of Korea) was fixed at 150 rpm and 30 °C, respectively. The bacterial concentration in the bioleaching medium was determined by a Burker–Turk cell counting chamber (Marienfeld Laboratory Glassware, Germany) under phase-contrast microscopy (ODEO-2003 Triple, IPONACOLOGY). In all bioleaching experiments, the initial bacterial concentration was adjusted to 1.2 × 10\(^8\) cells mL\(^{-1}\).

2.3. Bioleaching tests

Prior to the bioleaching tests, bacterial cultures of A. ferrooxidans were grown to reach stationary phase (i.e., 48 h). Experiments were carried out in 500 mL beakers at solid mine tailing concentrations of either 0.5% or 1.0%, 30 °C, and 150 rpm. To study the effects of bacterial attachment, A. ferrooxidans was separated from the mine tailings by a partition system\(^{21,22}\) consisted of a semi-permeable membrane with a molecular weight cutoff of...
(MWCO) of 12–14 kDa (Spectra/Por® 4 membrane, Spectrum® laboratories, Inc. CA. USA). The partition system setup (Fig. 2) was designed as proposed by Silva et al. (2015). Metal ions were confirmed to pass freely through the membrane, eliminating any possible negative effects on the bioleaching results.

Inside the membrane of the partition system, we pipetted 10 mL of DSMZ medium 882 containing either 1.5 or 3.0 g of solid (corresponding to solid concentrations of 0.5 and 1.0%, respectively). The chamber outside the membrane was seeded with 30 mL of bacterial suspension and 290 mL of DSMZ medium 882. In the non-partitioned system, 30 mL of bacterial suspension and either 1.5 or 3.0 g of mine tailings were added to 300 mL of DSMZ medium 882. The oxidation/reduction potential (ORP), pH, and metal (As and Fe) concentrations were measured daily. After measuring the solution pH (ORION 4STARS, Thermo) and ORP (Hanna Instruments, model 2211), a 2.0-mL aliquot was sampled from the leachate and filtered through a 0.45-μm nylon syringe filter (Corning Incorporated, Corning, Germany). The As and Fe concentrations in the aliquots were determined by ICP analysis. Additionally, the Fe$^{2+}$ concentration was analyzed by the o-phenanthroline method, and the Fe$^{3+}$ concentration was calculated as the difference between the total Fe and Fe$^{2+}$ contents.

To compare the total Fe and Fe$^{3+}$ concentration dynamics between the contact and non-contact systems, we re-expressed the total Fe and Fe$^{3+}$ concentrations as Eqs. (1) and (2), respectively. Here, $f_{T,Fe}$ denotes the relative total Fe concentration at a certain reaction time $t$, defined as the total Fe concentration $C_{T,Fe}$ at time $(t)$ divided by the maximum total Fe concentration $C_{Fe} (Max)$ over the entire reaction time. Similarly, $f_{Fe}^{3+}$ denotes the relative Fe$^{3+}$ concentration at reaction time $t$ and is defined as the Fe$^{3+}$ concentration $C_{Fe}^{3+}$ at time $(t)$ divided by $C_{T,Fe} (t)$.

$$f_{T,Fe} = \frac{C_{T,Fe} (t)}{C_{Fe} (Max)}$$ (1)

$$f_{Fe}^{3+} = \frac{C_{Fe}^{3+} (t)}{C_{T,Fe} (t)}$$ (2)

2.4. X-ray diffraction (XRD)

The XRD analysis revealed the mineral forms present in the mine tailings before and after bioleaching tests. The raw mine tailings were ground in a mortar. After the bioleaching experiments, the precipitates were dried at 80 °C and ground for analysis. Powder XRD patterns were collected within the 2θ range of 5°–70° in steps of 0.01°, with a counting time of 1 s step$^{-1}$. The XRD instrument was a Bruker D8 HRXRD (Germany) with CuKα radiation ($\lambda = 0.154606$ nm, 40 kV, 40 mA) and a SolX detector. The spectra were evaluated using the Diffracplus EVA package.
2.5. Bacterial attachment test

The bacterial attachment test determined the extent of initial bacterial attachment to the mine tailings. The attachment test was carried out under the same conditions as the bioleaching process (pH = 1.8, temperature = 30 °C, shaking speed = 150 rpm, initial cell concentration = 1.2 × 10⁸ cells mL⁻¹), but without nutrient supplement (as cell growth was undesired) to prevent bacterial growth over time. Note that several studies reported that initial cell adhesion to substrates normally govern biofilm formation[24-26]. The solution ionic strength, which is a recognized critical parameter for cell adhesion, was adjusted with NaCl to 320 mM (the ionic strength of the culture medium for bioleaching). The attachment test was conducted in a 50 mL centrifuge tube (BD Falcon™, USA) with a shaker (WiseShake SHR-1D, Witeg, Germany). The number of cells attached to the mine tailings was determined as the difference between the initial cell concentration and the cell concentration at specific time intervals (0, 40, and 80 min). The maximum time (80 min) is the equilibrium time of cell adhesion, as determined in previous studies[27,28]. The cell number was determined by a Burker–Turk counting chamber. All tests were triplicated (at least) to ensure reproducibility.

2.6. Statistical analysis

In the figures, mean data are presented with error bars indicating one standard deviation. Differences between mean values were analyzed using Student’s t-test and were considered statistically significant when \( P < 0.05 \).

3. Results and discussion

3.1. Arsenic extraction in contact and non-contact systems

Arsenic extraction in non-contact system can represent chemical extractions facilitated by oxidizing agents produced biologically in a two-step bioleaching system without any contact to mine tailing. Solubilization of metals by oxidizing agents produced biologically, like 

\[
\text{MS} + 2\text{Fe}^{3+} \rightarrow \text{M}^{2+} + \text{S}^{0} + 2\text{Fe}^{2+}
\]  

Fig. 3 shows the effect of non-contact mechanism when comparing arsenic extraction in systems with 0.5 and 1.0% solid concentration. Higher arsenic extraction was observed with lower solid concentration.

3.2. Effects of contact and non-contact mechanism at 0.5% solids

The arsenic solubilization for both contact vs. non-contact system at solid concentrations of 0.5% show overlapping behaviors. Both system’s trends increased linearly, exceeding 60% during the first 7 days of experiments and reached a plateau for the subsequent days (see Fig. 3). The pH profile for both systems also show overlapping behaviors (see Fig. 4a) increasing from 1.8 to 2.2 in the first two days and then decrease to 2.0 after 4 days and remained relatively constant afterwards. Similarly, the redox potential profile (See Fig. 4b) also show similar behaviour for both systems from 580 mV to 675 mV in first 3 days of experiment and then stayed constant until the end of the experiment.
As observed in Fig. 5a & b, the Fe speciation behaved similarly for both contact vs. non-contact systems throughout the bioleaching experiment. Fig. 5a shows a rapid decrease in Fe$^{2+}$ concentration after the first day of experiment indicating its conversion to Fe$^{3+}$ ions as expressed by equation 4:

$$4\text{Fe}^{2+} + \text{O}_2 + 4\text{H}^+ \rightarrow A_\text{f} 4\text{Fe}^{3+} + 2\text{H}_2\text{O} \quad (4)$$

In addition, the $f_{\text{f,Fe}}$ (Fig. 5b) in both systems rapidly increased throughout the first 24 h of reaction, suggesting a release of Fe from mine tailings as is also reflected by the total Fe trend in Fig. 5a. After the sharp increase in the first 24h, $f_{\text{f,Fe}}$ remained relatively stable with a slight decrease at the end of the bioleaching experiment. Contrarily, $f_{\text{f,Fe}^{3+}}$ slowly increased during the first four days of reaction suggesting a slow oxidation of Fe$^{2+}$ to Fe$^{3+}$ by bacterial activity. Nevertheless, stable oxidations rates (i.e., above 650 mV vs Ag/AgCl) were observed after the fifth day of reaction (Fig. 4b), together with a depletion of Fe$^{2+}$ in the system (Fig. 5a) suggesting a higher bacterial activity in both systems.

Thus, we conclude that bacterial attachment do not show significant improvement in arsenic solubilization at solid concentrations of 0.5% solids. Similar Fe concentration trends also suggest that attached bacteria did not increase Fe solubilization from the FeS$_2$ and FeAsS contained in mine tailings. We believe that even if bacteria attached to the surface of mine tailings the amount of 0.5% solid is not enough to sustain a significant increase in the efficiency of arsenic extraction.
3.3. Effects of contact and non-contact mechanism at 1.0% solids

Contact mechanism studied with 1.0% solid concentration showed 75% increase in the As extraction in 10 day bioleaching period (Fig. 3). Interestingly, during the first 5 days, the arsenic extraction in contact system increased linearly reaching values close to 40%, whereas the arsenic extraction was negligible in the non-contact system. However, in the subsequent days the arsenic extraction levelled off while the non-contact system increased linearly reaching a maximum of 19% by the end of the experiment.

Figs. 6a, 6b, 7a, and 7b represent pH profile, the ORP, the changes in the Fe speciation and relative concentrations of the total Fe and Fe\(^{3+}\) species in contact and non-contact system at 1.0% solid concentration. The pH increased from 1.8 to 2.3 within the first day of bioleaching and then stayed constant over the remaining test period in both systems. The fluctuations in pH measurement among contact vs. non-contact system were within experimental error. Likewise, the ORP followed similar trend (an increase in ORP within first day of bioleaching) and then stayed constant throughout the rest of the experiment reaching different magnitudes 660 mV vs. 640 mV in contact vs. non-contact system, respectively. Higher redox potentials correspond to stronger oxidizing environments. Therefore, a higher ORP in contact system provide indirect evidence of different rates of Fe\(^{2+}\) to Fe\(^{3+}\) oxidation among systems (Equation (4)). Similarly, a higher oxidation rate of Fe\(^{2+}\) to Fe\(^{3+}\) suggest a higher bacterial activity, as it is expressed by equation 4\(^{29}\). At the end of the experiment, the difference in ORP trends was found to be statistically significant (\(P < 0.05\)) suggesting that Fe\(^{3+}\) was produced at higher rate when there was no barrier between bacteria and mineral substrate.

Contrarily, the total Fe and Fe\(^{3+}\) concentrations in both systems decreased with bioleaching time (Fig. 7a). Notably, the Fe\(^{2+}\) concentration decreased under the detection limits. Thus, it is understood as an “instantaneous” oxidation from Fe\(^{2+}\) to Fe\(^{3+}\) due to bacterial activity in both systems. In contact system, within 1-5 days, total Fe and Fe\(^{3+}\) decreased slightly from 9 g/l to 8 g/l, indicating the availability of Fe\(^{3+}\) to leach arsenic in the solution. This may explain a greater extraction of arsenic within 1-5 days in contact system (See Fig. 3). In next 6-10 days, total Fe and Fe\(^{3+}\) decreased by 50% in contact system (i.e., from 8 g/l to 4 g/l) indicating less availability of Fe\(^{3+}\) in solution. Therefore, it decreased the extraction of arsenic in the solution and reached the plateau observed (see Fig. 3). In contrast, in the non-contact system the total Fe and Fe\(^{3+}\) decreased steadily from 10 g/l to 2 g/l thus showing reduced extraction efficiency.

At pH values exceeding 2.2, the decrease in total Fe in bioleaching system has been attributed to the pre-
precipitation of Fe$^{3+}$ as insoluble Fe$^{3+}$ species like jarosites and iron hydroxides$^{21,30,31}$. The precipitation largely depends on the Fe$^{3+}$ concentration, solution pH, ORP, and solid concentration$^{7,30}$. Therefore, a decreased in total Fe observed at 1.0% w/v solid concentration likely represents the precipitate formation that reduces the Fe availability in solution. This was more prominent in non-contact than contact system. Importantly, Fe$^{3+}$ can also precipitate with As-associated minerals, possibly reducing the As concentration in solution$^{13,32}$. Jarosite formation was confirmed by XRD analysis (Fig. 8) that can form a passivation layer on tailing sample and inhibiting the interaction between the metals and the oxidizing agents thus decrease bioleaching of arsenic$^{13}$. The effect of decreasing the availability of Fe$^{2+}$ ions on the bacterial population was confirmed in Fig. 9a showing lower bacterial concentrations in non-contact system compared to contact system.

Initial bacterial attachment to the mine tailing surface was confirmed by a cell adhesion test conducted during the first 80 min of reaction. Approximately $2.45 \times 10^{10}$ cells were attached to each gram of mine tailings (Fig. 9b) accounting to around 88% of the initial cell concentration.

By confirming bacterial attachment, altogether with an improvement of arsenic extraction in contact system, we conclude that non-contact mechanism negatively affects the extraction of arsenic from mine tailings. Decrease in bioleaching efficiency at high solid concentrations can be due to less tolerance of bacterial culture with tailing, oxygen deficiency in liquid media, and toxic effect of heavy metals. However, all of the drawbacks of systems at high concentration can be overcome by bacterial adaptations to high solid concentrations.
4. Conclusions

Current work can be a fundamental step to better understand environment aspects around indigenous mine sites of Korea and to reduce detrimental effect of tailing by proper management prior to dumping/discarding. The bioleaching efficiency of arsenic was less in non-contact system compared to contact system at 1.0% solid concentration. While at 0.5% solid concentration the effect of contact and non-contact mechanism was not so prominent. Interestingly, at 1.0% of concentration solid concentration, the arsenic extraction increased by 75% with contact mechanism. Therefore, we conclude that processes that base the metal extraction solely on oxidizing agents produced biologically on independent reactors, as the so-called two-step or non-contact bioleaching process, are not suitable for the extraction of arsenic from mine tailings. Bacterial adaptations are recommended to further improve overall arsenic extraction efficiency at high solid concentrations.

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References

1. Silva, Rene A, Danilo Borja, Gukhwa Hwang, et al., 2017: Analysis of the Effects of Natural Organic Matter in Zinc Beneficiation, Journal of Cleaner Production, 168, pp.814-22.
2. Holmes, David S., 2008: Review of International Biometallurgy Symposium, Frankfurt, 2007, Hydrometallurgy, 92(1-2), pp.69-72.
3. Borja Danilo, Nguyen Kim Anh, Silva R. A., et al., 2016: Experiences and Future Challenges of Bioleaching Research in South Korea, Minerals, 6(4), p.128.
4. Yin Shenghua, Leiming Wang, Eugie Kabwe, et al., 2018: Copper Bioleaching in China: Review and Prospect, Minerals, 8(2), p.32.
5. Watling, H.R., 2006: The Bioleaching of Sulphide Minerals with Emphasis on Copper Sulphides — A Review, Hydrometallurgy, 84(1-2), pp.81-108
6. Donati, Edgardo R, and Wolfgang Sand, 2007: Microbial Processing of Metal Sulfides. Springer.
7. Park, Jeonghyun, Yosep Han, Eunseong Lee, et al., 2014: Bioleaching of Highly Concentrated Arsenic Mine Tailings by Acidithiobacillus Ferrooxidans, Separation and Purification Technology, 133, pp.291-96.
8. Dastidar, Manisha G, Anushree Malik, and Pradip K Roychoudhury, 2000: Biodesulphurization of Indian (Assam) Coal Using Thiobacillus Ferroxidans (ATCC 13984), Energy Conversion and Management, 41(4), pp.375-88.
9. d’Hugues, P., P. Cezac, T. Cabral, et al. 1997: Bioleaching of a Cobaltiferous Pyrite: A Continuous Laboratory-Scale Study at High Solids Concentration, Minerals Engineering, 10(5), pp.507-27.
10. Mohseni, S., A. Marzban, S. Sepehr, et al., 2011: Investigation of Some Heavy Metals Toxicity for Indigenous Acidithiobacillus Ferroxidans Isolated from Sarcheshmeh Copper Mine, Jundishapur Journal of Microbiology, 4(3), pp.159-66.
11. Muñoz, J.A., F. González, M.L. Blázquez, et al., 1995: A
Study of the Bioleaching of a Spanish Uranium Ore. Part I: A Review of the Bacterial Leaching in the Treatment of Uranium Ores, Hydrometallurgy, 38(1), pp.39-57.

12. Bayard, Rémy, Vincent Chatain, Céline Gachet, et al., 2006: Mobilisation of Arsenic from a Mining Soil in Batch Slurry Experiments under Bio-Oxidative Conditions, Water Research 40(6), pp.1240-48.

13. Lee, Eunseong, Yosep Han, Jeonghyun Park, et al., 2015: Bioleaching of Arsenic from Highly Contaminated Mine Tailings Using Acidithiobacillus Thiooxidans, Journal of Environmental Management, 147, pp.124-31.

14. Lilova, K., and D. Karamanev, 2005: Direct Oxidation of Copper Sulfide by a Biofilm of Acidithiobacillus Ferrooxidans, Hydrometallurgy, 80(3), pp.147-54.

15. Fowler, T A, and F K Crundwell, 1998: Leaching of Zinc Sulfide by Thiobacillus Ferrooxidans: Experiments with a Controlled Redox Potential Indicate No Direct Bacterial Mechanism, Applied and Environmental Microbiology, 64(10), pp.3570-75.

16. Xin, B P, D Zhang, X Zhang, et al., 2009: Bioleaching Mechanism of Co and Li from Spent Lithium-Ion Battery by the Mixed Culture of Acidophilic Sulfur-Oxidizing and Iron-Oxidizing Bacteria, Bioresource Technology, 100(24), pp.6163-69.

17. Bryan, C G, E L Watkin, T J McCredden, et al. 2015: The Use of Pyrite as a Source of Lixiviant in the Bioleaching of Electronic Waste, Hydrometallurgy, 152, pp.33-43.

18. Bas, Ahmet Deniz, Haci Deveci, and Ersin Y Yazici, 2013: Bioleaching of Copper from Low Grade Scrap TV Circuit Boards Using Mesophilic Bacteria, Hydrometallurgy, 138, pp.65-70.

19. Ilyas, S, M A Anwar, S B Niazi, et al., 2007: Bioleaching of Metals from Electronic Scrap by Moderately Thermophilic Acidophilic Bacteria, Hydrometallurgy, 88(1-4), pp.180-88.

20. Hong, Jeongsk, Rene A Silva, Jeonghyun Park, et al., 2016: Adaptation of a Mixed Culture of Acidophiles for a Tank Biooxidation of Refractory Gold Concentrates Containing a High Concentration of Arsenic, Journal of Bioscience and Bioengineering, 121(5), pp.536-42.

21. Silva, Rene A, Jeonghyun Park, Eunseong Lee, et al., 2015: Influence of Bacterial Adhesion on Copper Extraction from Printed Circuit Boards, Separation and Purification Technology, 143, pp.169-76.

22. Chen, Y M, A H Lu, Y Li, et al., 2011: Naturally Occurring Sphalerite As a Novel Cost-Effective Photocatalyst for Bacterial Disinfection under Visible Light, Environmental Science & Technology, 45(13), pp.5689-95.

23. Koch, S, G Ackermann, and S Uhlig, 1989: Extraction-Spectrophotometric Determination of Iron(II) in the Presence of Iron(III) with 1,10-Phenanthroline, Zeitschrift Fur Chemie, 29(8), 298.

24. Jerez, Carlos A, 1997: Molecular Methods for the Identification and Enumeration of Bioleaching Microorganisms, In Biomining, 281-97. Berlin, Heidelberg: Springer Berlin Heidelberg.

25. Rohwerder, T, T Gehrke, K Kinzler, et al., 2003: Bioleaching Review Part A: Progress in Bioleaching: Fundamentals and Mechanisms of Bacterial Metal Sulfide Oxidation, Applied Microbiology and Biotechnology, 63(3), pp.239-48.

26. Vu, Barbara, Miao Chen, Russell Crawford, et al., 2009: Bacterial Extracellular Polysaccharides Involved in Biofilm Formation, Molecules, 14(7), pp.2535-54.

27. Kim, Hyunjung N, Yongsuk Hong, Ilkeun Lee, et al., 2009: Surface Characteristics and Adhesion Behavior of Escherichia Coli O157: H7: Role of Extracellular Macromolecules, Biomacromolecules, 10(9), pp.2556-64.

28. Kim, H N, S L Walker, and S A Bradford, 2010: Coupled Factors Influencing the Transport and Retention of Cryptosporidium Parvum Oocysts in Saturated Porous Media, Water Research, 44(4), pp.1213-23.

29. Sand, W, T Gehrke, P G Joza, et al., 2001: (Bio) Chemistry of Bacterial Leaching - Direct vs. Indirect Bioleaching, Hydrometallurgy, 59(2-3), pp.159-75.

30. Daoud, J, and D Karamanev, 2006: Formation of Jarosite during Fe2+ Oxidation by Acidithiobacillus Ferrooxidans, Minerals Engineering, 19(9), pp.960-67.

31. Jensen, A B, and C Webb, 1995: Ferrous Sulfate Oxidation Using Thiobacillus-Ferrooxidans - a Review, Process Biochemistry, 30(3), pp.225-36.

32. Guo, Zhaohui, Yi Cheng, et al., 2010: Effects of PH, Pulp Density and Particle Size on Solubilization of Metals from a Pb/Zn Smelting Slag Using Indigenous Moderate Thermophilic Bacteria, Hydrometallurgy, 104(1), pp.25-31.

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