Research Article

Evaluation of Preemergence Herbicide Programs for Control of Protoporphyrinogen Oxidase-Resistant *Amaranthus palmeri* in Soybean

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Protoporphyrinogen oxidase- (PPO-) resistant *Amaranthus palmeri* (S.) Wats. (Palmer amaranth) was confirmed in Arkansas in 2015. Field trials were conducted in Crawfordsville, Gregory, and Marion, Arkansas in 2016, and Crawfordsville and Marion in 2017, assessing PPO-resistant Palmer amaranth control options in *Glycine max* (L.) Merr. (soybean). Twelve trials consisted of 26 preemergence (PRE) treatments, evaluated for Palmer amaranth control and density reduction at 28 days after treatment (DAT). Treatments that consisted of PPO- or acetolactate synthase- (ALS-) inhibiting herbicides such as flumioxazin (72 g ai ha$^{-1}$) or sulfentrazone + cloransulam (195 g ha$^{-1}$ + 25 g ha$^{-1}$) controlled Palmer amaranth < 60%. At 28 DAT, treatments including mixtures of a very-long-chain fatty acid (VLCFA) plus the photosystem II- (PSII-) inhibiting herbicide metribuzin provided increased control over single herbicide sites of action (SOA) or herbicides mixtures to which Palmer amaranth was resistant. Pyroxasulfone + metribuzin (149 g ha$^{-1}$ + 314 g ha$^{-1}$) controlled Palmer amaranth 91% control across twelve trials at 28 DAT. S-metolachlor alone did not provide consistent, acceptable control of PPO-resistant Palmer amaranth (55–77%); subsequent research has determined that these populations are resistant to S-metolachlor. A minimum of two effective herbicides should be included in soybean PRE programs for control of PPO-resistant Palmer amaranth.

1. Introduction

Over-reliance on a single herbicide site of action (SOA) perpetuates the evolution and spread of herbicide-resistant biotypes [1–3]. In *Amaranthus palmeri* (S.) Wats. (Palmer amaranth), the repeated use of acetolactate synthase- (ALS-) inhibiting herbicides and glyphosate was selected for Palmer amaranth with mechanisms of resistance to one or both of these SOA [4–6]. Subsequently, growers shifted to PRE followed by residual postemergence (POST) weed management programs [7, 8].

After the identification of GR-resistant Palmer amaranth, use of PPO-inhibiting herbicides increased, offering both residual and foliar control of GR-resistant Palmer amaranth [9, 10]. PPO inhibitors offered both versatile and reliable control of Palmer amaranth [11, 12]. Fomesafen, a PPO-inhibiting herbicide in the diphenylether family, use increased from 2 to 16% of all US soybean acreage after confirmation of GR-resistant Palmer amaranth [7, 8], with the heaviest reliance on this herbicide in the Midouth. Only two herbicides were used on more acreage in the US, sulfentrazone (17%), which is another PPO-inhibiting herbicide, and glyphosate (85%) [8].

Season-long *Amaranthus* control is usually greater with the use of PRE herbicides. Hoffner et al. [13] reported that S-metolachlor and fomesafen PRE followed by glufosinate POST controlled Palmer amaranth better than sequential applications of glufosinate. In a similar experiment, at-harvest *Amaranthus rudis* (Moq.) Sauer (waterhemp) control with one glufosinate application was improved 22% when following flumioxazin PRE [14]. These findings lead to the conclusion that inclusion of PRE and residual POST herbicides increase season-long *Amaranthus* control, even when utilizing an effective,
nonresidual POST herbicide. Modeled resistance risk in Palmer amaranth was found to be reduced when effective PRE herbicides were included with POST programs [15]. Reduced resistance risk, improved season-long control, and confirmed ALS- and GR-resistant Palmer amaranth made residual herbicides, especially PPO inhibitors, a necessary component of successful Palmer amaranth weed control programs in soybean [6, 15].

Palmer amaranth resistance to PPO inhibitors was first reported in 2015 [1, 16]. Previously, waterhemp had been the focus of PPO resistance research, with seven states having confirmed PPO-resistant biotypes [1]. Susceptible Palmer amaranth PPO amino acid sequences were identical to susceptible waterhemp before the ΔG210 deletion occurred. Palmer amaranth, like waterhemp, remained absent of the nucleotide polymorphism seen in the *Amaranthus acanthochiton* at the replacement codon [17]. Riggins and Tranel [17] identified that the repeat motif in Palmer amaranth was identical to waterhemp for the PPX2 gene, which was confirmed as a mechanism for PPO resistance in waterhemp. Subsequently, this was shown to confer resistance to PPO inhibitors in Palmer amaranth as well. Known to hybridize, it is possible that the dioecious *Amaranthus* species crossed resulting in the transfer of the ΔG210 deletion to Palmer amaranth [16, 18, 19]. Two additional PPO resistance-conferring amino acid substitutions, Arg-128-Gly and Arg-128-Met, in Palmer amaranth were reported [20, 21]. Each of these mutations has since been determined to confer cross-resistance to other PPO-inhibiting herbicides [22–24].

Field trials conducted in several locations showed that PRE applications of PPO-inhibiting herbicides remained an effective control option for PPO-resistant waterhemp [24, 25]. In these field trials, PPO-resistant waterhemp was confirmed resistant not only to the diphenylether chemistry, but to all tested PPO-inhibiting herbicides [10, 25]. Wuerffel et al. [10] and Tranel et al. [3] noted the potential dangers for PPO-resistant waterhemp if PRE use continued on this biotype. As shown in a study by Wuerffel et al. [10], adding an effective herbicide such as *S*-metolachlor can help delay resistance selection by decreasing the number of resistant plants that survive. Preemergence applications of PPO-inhibiting herbicides provided some control of resistant waterhemp, but the addition of another effective SOA with residual activity often resulted in improved and more consistent control across environments [10, 24, 25].

As early as 1998, waterhemp control with sulfentrazone and flumioxazin was improved when co-applied with another effective residual herbicide such as metribuzin [9, 11]. Evidence supporting the benefit of PPO inhibitors, even after the confirmation of PPO-resistant *Amaranthus*, could lead to continued utilization of this chemistry if they are applied along with other effective herbicides.

With widespread ALS- and PPO-resistant Palmer amaranth, control using PRE herbicide programs once deemed effective needed to be reevaluated [4, 5, 16, 23]. The objective of this research was to evaluate the effectiveness of PRE-applied herbicide programs for control of PPO-resistant Palmer amaranth.

### 2. Materials and Methods

Field experiments were conducted on commercial farms near Marion, AR, on a Dubbs silt loam (fine-silty, mixed, active, thermic Typic Haplustalfs) in 2016 and 2017; Gregory, AR, on a Wiville sandy loam (fine-loamy, siliceous, active, thermic Typic Hapludalfs) in 2016; and Crawfordsville, AR, on both a Forestdale silt loam (fine, smectitic, thermic Typic Endoaqualfs) in 2016 and a Dundee silty loam (fine-silty, mixed, active, thermic Typic Endoaqualfs) in 2016 and 2017. The soil near Marion had a pH of 5.8 with 1.6% organic matter and the soil for both fields near Crawfordsville had a pH of 5.28 and 5.34 with an organic matter content of 1.8 and 1.95%. Plots consisted of 4 single rows, spaced 97 cm apart at a density of 370,500 seeds ha⁻¹ in Marion, rows, spaced 19 cm apart, at 449,540 seeds ha⁻¹ in Gregory, and rows, spaced 97 cm apart, at 345,800 seeds ha⁻¹ in Crawfordsville (Table 1). Trials were set up with 9.1 m long by 3.9 m wide plots in Marion, 6.1 m long by 2.3 m wide plots in Gregory, and 7.6 m long by 3.9 m wide plots in Crawfordsville. Group IV soybean varieties were used for this experiment and are listed along with planting dates in Table 1. Herbicide applications were made at 140 L ha⁻¹ with a four-nozzle boom attached to a CO₂-pressurized backpack sprayer in Crawfordsville and 112 L ha⁻¹ using a Bowman Mudmaster™ multiboom system in Marion and Gregory. All herbicides were applied with 110015 TeeJet® air induction extended range nozzles at a speed of 4.8 kph. All locations were absent of irrigation, requiring precipitation for PRE treatment activation. Listed in Table 2, precipitation gathered from on-site weather stations for each site-year shows at least 1.72 cm of rainfall within the first 14 days after treatment (DAT).

Twenty-five PRE programs were evaluated in soybean for Palmer amaranth control, not including the nontreated control. PRE programs consisted of single active ingredients and herbicide mixtures with more than one SOA. Preemergence treatments focused on four distinct SOAs: ALS-, PPO-, very-long-chain fatty acid- (VLCFA-), and PSII-inhibiting herbicides. Table 3 is a comprehensive list of treatment composition, herbicide group, and rate of each herbicide. Visible estimates of Palmer amaranth control and Palmer amaranth density in each plot were recorded 28 DAT, targeting evaluations before a typical POST application timing. Ratings were taken on a 0 to 100% scale, with 0% representing no control and 100% indicating complete weed mortality [26]. To determine Palmer amaranth density, two separate counts per 0.5 m² were counted in the center two rows of the plots. Palmer amaranth density on a m² basis was then converted to a percentage relative to counts in the nontreated control. Palmer amaranth densities were not recorded at the Gregory location, hence the exclusion of this site in Table 3.

Data were analyzed using JMP Genomics 8 (SAS Institute Inc., SAS Campus Drive, Cary, North Carolina 27513). This experiment was set up as a randomized complete block design with four replications, utilizing data from 10 trials in 2016, and 2 in 2017. Experiments were analyzed across location and separately by year using a fit model and
beta distribution. Replication was considered a random
effect in each individual model, and treatment was designed
as a fixed effect in each model. Box and whisker plots were
also included to provide treatment variation, which dem-
onstrate both outliers and the range of control of individual
treatments (Figures 1 and 2). Means and separation are
included, which were derived from a Tukey’s HSD of visible
control ratings. As explained in each figure, a sample size of
32 points for each treatment was included in 2016, and 8 in
2017. Data from the nontreated were excluded and not used
in analyses. Where appropriate, data were separated using
Tukey’s HSD

3. Results and Discussion

There was a minimum of 1.72 cm of precipitation within the
first 14 DAT at all environments (Table 2). Experiments at
each location were conducted using common producer
methods, with a considerable difference in seeding rate and
row-spacing (Table 1). Gregory, Marion, and Crawfordsville
Palmer amaranth populations were all previously confirmed
to be PPO-, ALS-, and GR-resistant [1, 16, 22, 23]. The
mechanisms of PPO resistance were confirmed as follows for
the three locations: Crawfordsville (AG210), Gregory
(∆G210; Arg-128), and Marion (Arg-128).

3.1. Palmer Amaranth Response in 2016. The nontreated
plots at Marion and Crawfordsville in 2016 averaged 221
Palmer amaranth plants m⁻² at 28 DAT (data not shown).
For all herbicide treatments at 28 DAT, Palmer amaranth
density reduction ranged from 30 to 95% (Table 3). Sul-
fentrazone + cloransulam (196 + 25 g ha⁻¹) reduced Palmer
amaranth density the least, and saflufenacil + dimethena-
mid-p + pyroxasulfone + metribuzin (25 + 219 + 149 + 314
gha⁻¹) reduced Palmer amaranth density the most. The low
Palmer amaranth reduction with PPO-inhibiting herbicides
was apparent. For example, flumioxazin (72 g ha⁻¹) alone
reduced Palmer amaranth density 35% at 28 DAT. Fur-
thermore, because of confirmed ALS resistance at these sites,
PRE treatments that consisted only of a PPO or ALS in-
hibitor reduced Palmer amaranth an average of 55% (Ta-
ble 3). The only herbicide tank-mix of an ALS- plus PPO-
inhibiting herbicide that reduced Palmer amaranth density

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**Table 1:** Locations, soybean varieties, planting dates, row spacings, and seeding rates for 12 trials conducted in Arkansas, USA, in 2016 and 2017.

| Location     | Variety            | Technology       | Planting date | Row spacing (cm) | Seeding rate (1,000 seed ha⁻¹) |
|--------------|--------------------|------------------|---------------|------------------|--------------------------------|
| Crawfordsville | AG 47 × 6          | Roundup ready    | May 11, 2016  | 97               | 346                            |
|              | AG 47 × 6          | Roundup ready 2 xtend | May 11, 2016  | 97               | 346                            |
|              | HBK 4950 LL        | LibertyLink      | May 11, 2016  | 97               | 346                            |
|              | 43R15Y9            | Enlist           | June 10, 2016 | 97               | 346                            |
|              | P49T09 BR          | LibertyLink      | May 22, 2017  | 97               | 346                            |
| Gregory      | AG 4633            | Roundup ready    | May 5, 2016   | 19               | 450                            |
|              | DG 4957 LL         | LibertyLink      | May 5, 2016   | 19               | 450                            |
|              | 43R15Y9            | Enlist           | May 5, 2016   | 19               | 450                            |
| Marion       | AG 4632            | Roundup ready    | May 12, 2016  | 97               | 370                            |
|              | AG 47 × 6          | Roundup ready 2 xtend | May 12, 2016  | 97               | 370                            |
|              | P49T31 LL          | LibertyLink      | May 12, 2016  | 97               | 370                            |
|              | HBK 4953 LL        | LibertyLink      | May 10, 2017  | 97               | 370                            |

**Table 2:** Accumulated rainfall following PRE herbicide application from 12 trials conducted near Crawfordsville, Gregory, and Marion, Arkansas, in 2016 and 2017a.

| Rainfallb | 2016 |            | 2017 |            |
|-----------|------|------------|------|------------|
| DAT       | Crawfordsville | Gregory | Marion | Crawfordsville | Marion |
| 7         | 0.28 | 3.91       | 1.47 | 0.58       | 3.15 | 1.57 |
| 14        | 3.63 | 0          | 0.25 | 3.53       | 3.53 | 1.57 |
| 21        | 4.90 | 3.68       | 3.96 | 1.80       | 0    | 3.45 |
| 28        | 0.28 | 2.59       | 4.75 | 0.18       | 3.15 | 1.19 |
| 35        | 3.91 | 2.06       | 0.25 | 3.91       | 6.80 | 0   |
| 42        | 0    | 0.18       | 0    | 0          | 0.23 | 3.18 |

**Abbreviations:** AG, Asgrow, Bayer CropScience, Research Triangle Park, NC 27709; DG, Delta Grow, Delta Grow Seed, England, AR 72046; HBK, Hornbeck, Hornbeck Seed Company, De Witt, AR 72042; P, Pioneer, DowDupont Midland, MI 48674; 43R15Y9, Enlist soybean variety designation, Corteva, Midland, MI 48674.

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a Abbreviation: DAT, days after treatment. b Values represent cumulative precipitation from previous timing. c Two columns are presented for Crawfordsville in 2016 as three experiments were established on May 12th, 2016, represented by the first column, and one was established on June 10th, 2016, represented by the second column.
Table 3: Palmer amaranth density reduction following PRE herbicide application in soybean from 12 trials conducted near Crawfordsville and Marion, Arkansas, in 2016 and 2017<sup>a,b,c</sup>.

| Herbicide                                      | Rate g ai ha<sup>-1</sup> | WSSA group | 28 DAT Density<sup>d</sup> | 2016 | 2017 |
|------------------------------------------------|---------------------------|------------|----------------------------|------|------|
| Saflufenacil + dimethenamid-p + pyroxasulfone + metribuzin | 25                        | 14         | 95 a                        | 98 a |      |
|                                                 | 191                       | 15         | 93 a                        | 98 a |      |
|                                                 | 314                       | 15         | 92 a                        | 85 ab|      |
| Flumioxazin + pyroxasulfone                     | 88                        | 14         | 90 a                        | 98 a |      |
|                                                 | 111                       | 15         | 90 a                        | 98 a |      |
| S-metolachlor + metribuzin                      | 1,105                     | 15         | 88 ab                       | 67 cd|      |
| Flumioxazin + metribuzin + chlorimuron-ethyl + pyroxasulfone | 250                       | 5          | 91 a                        | 97 a |      |
|                                                 | 22                        | 2          |                             |      |      |
|                                                 | 90                        | 15         |                             |      |      |
| Pyroxasulfone + metribuzin                      | 149                       | 15         | 90 a                        | 98 a |      |
|                                                 | 314                       | 5          |                             |      |      |
| Metribuzin + chlorimuron-ethyl + S-metolachlor  | 88                        | 14         | 90 a                        | 96 a |      |
|                                                 | 17                        | 2          |                             |      |      |
| Metribuzin + chlorimuron-ethyl + S-metolachlor  | 1,105                     | 15         | 88 ab                       | 67 cd|      |
| S-metolachlor + metribuzin                      | 45                        | 2          |                             |      |      |
|                                                 | 1,070                     | 15         |                             |      |      |
| Flumioxazin + pyroxasulfone + chlorimuron-ethyl | 98                        | 15         | 86 ab                       | 98 a |      |
|                                                 | 21                        | 2          |                             |      |      |
| Metribuzin + chlorimuron-ethyl + pyroxasulfone  | 45                        | 2          | 85 ab                       | 92 ab|      |
|                                                 | 90                        | 15         |                             |      |      |
| Flumioxazin + cloransulam                       | 106                       | 14         | 84 a-c                      | 88 ab|      |
|                                                 | 35                        | 2          |                             |      |      |
| Flumioxazin + metribuzin + chlorimuron-ethyl    | 250                       | 5          | 80 a-c                      | 92 ab|      |
|                                                 | 22                        | 2          |                             |      |      |
| Sulfentrazone + S-metolachlor                   | 134                       | 14         | 78 a-c                      | 86 ab|      |
|                                                 | 1,210                     | 15         |                             |      |      |
| Flumioxazin + cloransulam                       | 71                        | 14         | 74 a-c                      | 79 a-c|      |
|                                                 | 24                        | 2          |                             |      |      |
|                                                 | 76                        | 14         |                             |      |      |
| Flumioxazin + chlorimuron-ethyl + thifensulfuron + pyroxasulfone | 13                        | 2          | 73 a-c                      | 93 ab|      |
|                                                 | 90                        | 15         |                             |      |      |
| Metribuzin                                      | 420                       | 5          | 73 a-c                      | 74 a-c|      |
|                                                 | 72                        | 14         |                             |      |      |
| Flumioxazin + chlorimuron-ethyl + thifensulfuron + pyroxasulfone | 23                        | 2          | 67 a-c                      | 94 ab|      |
|                                                 | 7                         | 2          |                             |      |      |
| Flumioxazin + chlorimuron-ethyl + thifensulfuron | 13                        | 2          | 61 a-c                      | 73 a-c|      |
|                                                 | 4                         | 2          |                             |      |      |
| Flumioxazin + chlorimuron-ethyl                 | 63                        | 14         | 56 a-c                      | 77 a-c|      |
| S-metolachlor                                   | 22                        | 2          |                             |      |      |
| Flumioxazin + chlorimuron-ethyl + thifensulfuron | 1,064                     | 15         | 55 a-c                      | 79 a-c|      |
|                                                 | 72                        | 14         |                             |      |      |
| Flumioxazin + chlorimuron-ethyl + thifensulfuron | 23                        | 2          | 53 a-c                      | 87 ab|      |
80% was flumioxazin + cloransulam (106 + 35 g ha\(^{-1}\)) at 84%. Although reliance on these SOAs alone results in poor Palmer amaranth control, the combination of PPO and ALS inhibitors with other effective herbicides is still a viable option. This is demonstrated by treatments such as flumioxazin + pyroxasulfone (88 + 111 g ha\(^{-1}\)), which reduced Palmer amaranth density 93% at 28 DAT.

Preemergence treatments that include both Group 5 and 15 herbicides reduced Palmer amaranth density an average of 90% (Table 3). Tank-mixtures containing Group 5 and 15 herbicides control Palmer amaranth at an average of 87% across all mixtures over both years, which was achieved when mixing these two herbicide groups as shown in Figure 1. Comparison of treatment means for the box and whisker plots can be derived from an ANOVA mentioned previously (\(P < 0.0001\)). Herbicide treatments that consisted of only one herbicide, regardless of SOA, controlled Palmer amaranth <68%. This result was not surprising because in previous greenhouse research on some of these populations, the single SOA herbicide treatments had low efficacy on these multiple-herbicide-resistant Palmer amaranth populations [22]. Surprisingly, S-metolachlor (1064 g ha\(^{-1}\)), controlled Palmer amaranth an average of 63%, had an extremely wide range of control at 28 DAT (Figure 1). Poor control with S-metolachlor is also reflected by the 55% density reduction (Table 3). Just recently, S-metolachlor-

### Table 3: Continued.

| Herbicide + + | Rate | WSSA group | 28 DAT Density\(^d\) |
|----------------|------|------------|---------------------|
|                | g ai ha\(^{-1}\) | 2016 | 2017 |
| Sulfentrazone + metribuzin | 188 | 14 | 51 | a-c | 77 | a-c |
| Sulfentrazone + cloransulam | 130 | 14 | 47 | a-c | 31 | d |
| Flumioxazin | 17 | 2 | 35 | bc | 77 | a-c |
| Sulfentrazone + cloransulam | 196 | 14 | 30 | c | 54 | cd |

\(a\) Abbreviations: DAT, days after treatment; WSSA, Weed Science Society of America. \(b\) Means within a column followed by the same lowercase letter are not different based on Tukey’s HSD (\(P > 0.05\)). \(c\) Average Palmer amaranth density for the nontreated control was 217 per m\(^2\) in 2016 and 37 per m\(^2\) in 2017. \(d\) Gregory not included as density data are not available.
resistant Palmer amaranth have been confirmed at the Marion and Crawfordsville sites [1, 27], but no resistance testing or confirmation has occurred for the Gregory population.

3.2. Palmer Amaranth Response in 2017. The two trials conducted in 2017 focused exclusively on the Marion and Crawfordsville sites. The nontreated plots across Marion and Crawfordsville in 2017 averaged 75 Palmer amaranth per m$^{-2}$ at 28 DAT (data not shown). The likely reason for the lower density in 2017 is that Palmer amaranth seed production was not allowed at these sites in 2016. Improved herbicide efficacy in the second year can be attributed, in part, to lower Palmer amaranth density; trends in herbicide efficacy were consistent between years. Palmer amaranth density reduction ranged from 31 to 98% in 2017 (Table 3). In 2017, two PRE treatments with ALS- and PPO-inhibiting herbicides, flumioxazin + cloransulam (106 + 35 g ha$^{-1}$) and flumioxazin + chlorimuron-ethyl + thifensulfuron (72 + 23 + 7.11 g ha$^{-1}$), controlled ALS- and PPO-resistant Palmer amaranth 88 and 87%, respectively. In 2017, density reduction at 28 DAT shows that reliance on PPO or ALS inhibitors alone would often result in poor control of these Palmer amaranth populations, especially without the presence of a Group 5 or 15 herbicide. Box and whisker plots, with corresponding means and separation, for 28 DAT in 2017 show less variability in the majority of treatments versus 2016 (Figure 2). The more consistent response of herbicides among replications and the two locations in 2017 is in part why years were analyzed separately for visible estimates of control, as the $P$ value for the year effect was 0.0016 when included in the ANOVA model. There was no significant effect for site during this analysis for 2016 or 2017.

At 28 DAT, metribuzin controlled PPO-resistant Palmer amaranth 71%. Visible estimates of Palmer amaranth control for S-metolachlor, while higher than in 2016, averaged only 77%, with a relatively high amount of variation across plots and locations (Figure 2). This finding leads to the hypothesis that control of these Palmer amaranth populations with S-metolachlor alone could be extremely variable depending on year and location, which may contribute to the evolution of S-metolachlor-resistant Palmer amaranth. However, including this herbicide as part of an effective PRE program still has benefit in that it provides some Palmer amaranth control as well as other weeds such as barnyard grass *(Echinochloa crus-galli* L. Beauv.), the most common grass weed of soybean in this region.

3.3. Practical Implications. Reliance on PPO- and ALS-inhibiting herbicides for PRE control of glyphosate-, ALS-, and PPO-resistant Palmer amaranth provided less than acceptable control at 28 DAT and will put added pressure on POST herbicides for acceptable weed control. It was evident that PRE programs including metribuzin and a Group 15 herbicide can successfully control PPO-resistant Palmer amaranth for the first four weeks after soybean planting. However, reliance on S-metolachlor or metribuzin alone failed to provide acceptable control, hence the need for two or more PRE herbicides. For both 2016 and 2017, treatments containing pyroxasulfone never provided <80% control and averaged 89% control across all site-years. This indicates pyroxasulfone remains one of the few proven, effective herbicides for control of PPO-resistant Palmer amaranth. S-metolachlor-resistant Palmer amaranth, now confirmed in Crawfordsville and Marion, Arkansas, has further limited...
potential weed control options in soybean. Although S-metolachlor can be used in combination with other effective herbicides, it should be noted that reduced efficacy is likely with the confirmation of S-metolachlor-resistant Palmer amaranth in 2019 [27]. Also, it should be noted that there is a slight reduction in sensitivity of the Marion and Crawfordsville accessions to pyroxasulfone [27] and continued reliance on this Group 15 herbicide could increase the selection intensity for Group 15 resistance in these fields. While flumioxazin was not deemed effective alone (67 to 81%), the data suggested it does have value in a tank-mix PRE application for control of PPO-resistant Palmer amaranth. The coapplication of two or more effective SOAs is recommended for control of Palmer amaranth.

Data Availability

The underlying data for this experiment set are held on a device accessible by the corresponding author.

Disclosure

This experiment is part of thesis work by the corresponding author and is available at https://scholarworks.uark.edu/etd/3395/.

Conflicts of Interest

The authors declare no conflicts of interest.

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