Inhibition of breast cancer growth and metastasis by a biomimetic peptide

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Metastasis is the main cause of mortality in cancer patients. Though there are many anti-cancer drugs targeting primary tumor growth, anti-metastatic agents are rarely developed. Angiogenesis and lymphangiogenesis are crucial for cancer progression, particularly, lymphangiogenesis is pivotal for metastasis in breast cancer. Here we report that a novel collagen IV derived biomimetic peptide inhibits breast cancer growth and metastasis by blocking angiogenesis and lymphangiogenesis. The peptide inhibits blood and lymphatic endothelial cell viability, migration, adhesion, and tube formation by targeting IGF1R and Met signals. The peptide blocks MDA-MB-231 tumor growth by inhibiting tumor angiogenesis in vivo. Moreover, the peptide inhibits lymphangiogenesis in primary tumors. MDA-MB-231 tumor conditioned media (TCM) was employed to accelerate spontaneous metastasis in tumor xenografts, and the anti-metastatic activity of the peptide was tested in this model. The peptide prevents metastasis to the lungs and lymph nodes by inhibiting TCM-induced lymphangiogenesis and angiogenesis in the pre-metastatic organs. In summary, a novel biomimetic peptide inhibits breast cancer growth and metastasis by blocking angiogenesis and lymphangiogenesis in the pre-metastatic organs as well as primary tumors.

Breast cancer is the most frequently diagnosed malignancy among women in the United States1. Although anti-angiogenic agents have been used for treating breast cancer in the clinic, overall survival in advanced breast cancer patients has not been substantially improved2 possibly because breast cancer metastasis occurs preferentially via lymphatic dissemination compared to hematogenous spread3. Thus, there is an unmet need to develop therapeutic agents to block lymphangiogenesis as well as angiogenesis to efficiently block breast tumor growth and metastasis.

We have identified several endogenous peptides to inhibit lymphangiogenesis by using bioinformatics-aided methodologies and in vitro and animal experiments4–7. Lee et al. have reported that short peptides derived from somatotropin domain-containing proteins inhibit lymphatic endothelial cell (LEC) proliferation, migration, adhesion, and tube formation4. Lee et al. have also documented that a 14-mer peptide derived from transmembrane protein 45A (TMEM45A), a member of the somatotropin family of proteins, exhibits potent anti-lymphangiogenic activity in breast tumors xenografts and in tumor-conditioned lymph nodes4. Koskimaki et al. showed synergy between the collagen IV derived mimetic peptide SP2012 and the TMEM45A peptide as lymphangiogenesis inhibitors6. This collagen IV derived peptide has also shown potent anti-angiogenic activity in earlier studies6,8. When tested separately, the collagen IV peptide SP2012 was significantly more potent than TMEM45A peptide in inhibition of angiogenesis; its activity was comparable to the TMEM45A peptide in inhibition of lymphangiogenesis8. Based on these findings, SP2012 was further optimized to be soluble, easy to synthesize and stable by using bioinformatics and amino acid mutation methodologies9,10. Here we report the in vitro and in vivo activities of the optimized collagen IV biomimetic peptide SP2043, whose amino acid sequence is LRRFSTAPFAFIDINDVINF. This peptide has activity against lymphangiogenesis and angiogenesis, and inhibits the growth of a primary triple negative breast cancer (TNBC) tumor xenograft and its metastasis to the lungs and lymph nodes by blocking lymphangiogenesis and angiogenesis in pre-metastatic organs as well as primary tumors. For the metastasis study we employ a tumor-conditioned media (TCM) mediated spontaneous metastasis model developed previously in our laboratory11,12, which shows accelerated formation of thoracic metastases within one month after orthotopic tumor inoculation into the inguinal mammary fat pad. The use of TCM mimics secretion of pro-metastatic factors by the primary tumor in patients.
**Results**

SP2043 blocks viability, adhesion, migration and tube formation of lymphatic and blood endothelial cells. The effect of SP2043 on cell viability, adhesion, migration, and tube formation of human umbilical vein endothelial cells (HUVEC) was tested (Fig. 1a–e). The WST-1 reagent was used to determine the effect of SP2043 on the viability of HUVEC. SP2043 inhibits HUVEC viability with an IC$_{50}$ value of $7.0 \pm 2.1 \mu$M (Fig. 1a). The real-time cell analysis

![Graph showing HUVEC viability](image)

**Figure 1| In vitro activity of SP2043 on HUVEC, MEC, and LEC.** SP2043 blocks angiogenesis and lymphangiogenesis in vitro. The controls for cell viability, migration, adhesion, and tube formation assays correspond to cell phenotypes in normal endothelial growth media (EGM-2 in the HUVEC experiment, or EGM-2MV in LEC and MEC experiments). (a) Inhibition of HUVEC viability. (b) Inhibition of HUVEC adhesion. (c) Inhibition of HUVEC migration. (d) Inhibition of HUVEC tube formation at 18 h. Scale bars represent 200 μm. (e) Quantification of (d). HUVEC tube formation was quantified by using Angiogenesis Analyzer for ImageJ (NIH). The process is described in the Supplementary Fig. S8 online and Methods. Briefly, the original images (3 images per group) were transformed to the skeletonized binary tree images. These were analyzed and all the values were normalized with the analyzed area. Number of nodes (Nb nodes), Nb junctions, Nb segments, Nb branches, total length (Tot length), Tot branching length, and Tot segment length were obtained (*$P < 0.05$). (f) Inhibition of MEC tube formation at 18 h. Scale bars represent 200 μm. (g) Inhibition of LEC tube formation at 18 h. Scale bars represent 200 μm. Data (a,b,e) are reported as mean ± s.e.m.
(RTCA) system was employed to measure HUVEC adhesion and migration. The RTCA system (ACEA) measures electrical impedance. Gold electrodes embedded on the bottom of a 96-well plate to measure adhesion or on the underside of the top chamber of a Boyden-chamber-like 2-chamber setup to measure migration register the number of cells or the extent of contact between the cells and the electrodes as a change in impedance. This change is converted to a cell index that is a direct measure of the amount of adhesion or the number of migrated cells. SP2043 inhibits HUVEC adhesion (IC50 value of 0.93 ± 0.56 μM) and migration (IC50 value less than 6.3 μM) (Fig. 1b,c). HUVEC tube formation is also blocked by SP2043 (Fig. 1d). Tube formation was analyzed by using Angiogenesis Analyzer for ImageJ (NIH). Number of nodes (Nb nodes), Number of junctions (Nb junctions), Number of segments (Nb segments), Number of branches (Nb branches), total length (Tot length), Tot branching length, and Tot segment length were obtained and normalized. The pixel number of the analyzed area is taken to be 100,000. From the analysis, we showed that SP2043 treatment inhibits HUVEC tube formation (Fig. 1e). HUVEC are routinely used in angiogenesis in vitro assays, but due to their venous origin they may not adequately represent tumor microvessels. For this reason we tested SP2043 on microvascular endothelial cells (MEC) which are derived from capillaries: SP2043 blocks MEC tube formation and adhesion (Fig. 1f and Supplementary Fig. S1 online). SP2043 also inhibits the adhesion and tube formation of lymphatic endothelial cells (LEC) (Fig. 1g and Supplementary Fig. S1 online). Scrambled-SP2043 (Scram-2043; seq: LRRFSTAPFAFIPEAK-VNF) was prepared by replacing “DIND” from the SP2043 (seq: LRRFSTAPFAFIDVDNFIF) with “PEAK” (Supplementary Fig. S2 online). The detailed rationale for the preparation of this scrambled peptide is described in the Methods (‘Peptide synthesis and handling’). Scram-2043 inhibits MEC adhesion and migration very poorly compared to SP2043 showing that the scrambled peptide has lost most of activity compared to SP2043, and that the activity of SP2043 is amino acid sequence specific (Supplementary Fig. S2 online).

SP2043 blocks IGF1R and Met signals in lymphatic and blood endothelial cells. We previously found that the collagen IV biomimetic peptide SP2012, which is an earlier analog of SP2043, inhibits VEGFR2 signaling pathways in blood endothelial cells by interacting with β1 integrin15–18. As the β1 integrin can associate with diverse receptor tyrosine kinase receptors, including VEGFR2, Met, IGF1R, and EGFR in diverse cell types, here we expand its molecular targets15–16. We showed that EGF-induced phosphorylation of EGFR and its downstream signals (MAPK and Akt) are not blocked by SP2043 (Supplementary Fig. S3 online). Therefore, the effect of SP2043 on the IGF1R and Met signaling pathways in endothelial cells were further studied (Fig. 2). In HGF-induced MEC, SP2043 blocks the levels of phospho-Met (Y1234/1235) and phospho-Gab1 (Y307) and phospho-MAPK (T202/Y204) (Fig. 2a). In IGF1 induced MEC, SP2043 inhibits the level of phospho-IGF1R (Y1135/1136) and inhibits phosphorylation of MAPK and Akt (S473) (Fig. 2b). HGF induces the phosphorylation of Met and Akt in LEC. These phosphorylations are blocked by SP2043 in a dose dependent manner (Fig. 2c). SP2043 also blocks the phosphorylation of IGF1R, MAPK induced by IGF treatment of LEC (Fig. 2d).

CD58, CD155, and ADAM17 dissociate from IGF1R and Met receptor complexes upon SP2043 treatment. To understand the multimodal inhibition of the IGF1R and Met signals by the SP2043 peptide, first the protein components of the IGF1R or Met receptor complexes induced by IGF1 or HGF were determined. Next the proteins that were dissociated from the IGF1R or Met receptor complexes after SP2043 treatment were identified. In IGF1 induced MEC, 11 proteins are found to be part of the receptor complex with IGF1R (CD58, CD155, CD31, CD36, CD44, CD147, RECK, ADAM17, Galectin-3, Integrin β1, and Integrin β2). After SP2043 treatment, 8 of these proteins (CD31, CD147, Integrin β1, RECK, Galectin-3, CD58, CD155, ADAM17) are found to dissociate from the receptor complex (Fig. 3a). In HGF-induced MEC, 8 proteins are found to be part of the receptor complex with Met (CD58, CD155, TIMP3, CD36, CD154, CD44, ADAM17, Integrin β2). After SP2043 treatment, 7 of these proteins (CD58, CD155, TIMP3, CD36, CD154, CD44, ADAM17) are found to dissociate from the receptor complex (Fig. 3b). Taken together, CD58, CD155, and ADAM17 are targeted by SP2043 and dissociated from both the IGF1R and Met receptor complexes (Fig. 3c).

SP2043 inhibits viability of MDA-MB-231, MCF-7, and SUM-149 breast cancer cells. The roles of IGF1R, Met, CD58, CD155, and ADAM17 in cancer progression have been studied19–21. Given the role of the IGF and HGF signaling pathways in breast cancer cells22–25 and the result of inhibition of IGF1R and Met signals by the peptide and the dissociation of CD58, CD155, and ADAM17 from the IGF1R and Met receptor complexes, we hypothesized that the SP2043 peptide influences breast cancer cells as well. The potential anti-proliferative potential of SP2043 on breast cancer cells was tested on two triple-negative breast cancer (TNBC) cell lines (MDA-MB-231 and SUM-149), and one estrogen receptor-positive (ER +) breast cancer cell line (MCF-7). The peptide inhibits viability of MDA-MB-231 (IC50 value of 22.1 ± 5.2 μM), MCF-7 (IC50 value of 37.9 ± 4.9 μM), and SUM-149 cells (IC50 value of 11.8 ± 4.3 μM) (Supplementary Fig. S7 online).

SP2043 inhibits MDA-MB-231 tumor growth. Given the inhibition of angiogenesis and lymphangiogenesis by SP2043 (Fig. 1), and the inhibition of breast cancer cell viability (Supplementary Fig. S7 online), we evaluated it for efficacy in animal models of TNBC. MDA-MB-231 tumor xenografts were established in athymic nude mice as previously described26–29. After tumors reached a volume of about 100 mm3, SP2043 (10, 20 mg/kg) was administered i.p. every day for up to 33 days. SP2043 significantly inhibits tumor growth: 77.8 and 90.7% inhibition by 10 and 20 mg/kg of SP2043, respectively at day 33 (Fig. 4a). The mouse body weights are statistically identical among those three groups showing no severe toxicity of SP2043 (Supplementary Fig. S4 online). Additional western blot assays show that HGF induces phosphory-Met (Y1234/1235) in MDA-MB-231 cells, and SP2043 treatment inhibits this phosphorylation in vitro as in blood and lymphatic endothelial cells (Supplementary Fig. S7 online). To determine if this effect of SP2043 could be detected in vivo, we stained tumor samples from the control and the SP2043 treated groups with anti-anti-phospho-Met (p-Met) antibody and anti-mouse CD31 antibody. Control tumors show ubiquitous p-Met around the mCD31 signal (possibly blood and lymphatic vessels) as well as in cancer cells and other stromal cells (DAPI stained). SP2043 inhibits the amount of p-Met (Supplementary Fig. S7 online). Next, we assessed anti-angiogenic and anti-lymphangiogenic activity of the peptide in the tumors (Fig. 4b–e). Blood vessel (lectin-positive) and lymphatic vessel (LYVE-1 positive) formation is potently inhibited by the peptide (Fig. 4b). Enlarged images from the peri- and intratumoral areas also show the reduction of blood and lymphatic vasculature in the presence of the peptide (Fig. 4c). Pixel based quantification show inhibition of angiogenesis by 57.8% and lymphangiogenesis by 69.5% by SP2043 (Fig. 4d,e).

SP2043 inhibits MDA-MB-231 tumor metastasis in the TCM induced spontaneous metastasis model. We have previously reported that tumor-conditioned media (TCM) pre-treatment followed by tumor inoculation facilitated spontaneous metastasis by influencing lymphangiogenesis and angiogenesis in the primary tumors and pre-metastatic organs30–32. We also previously showed that TCM-induced lymphatic and blood vessel formation in the
The inhibition of HGF and IGF1 signals by SP2043. (a) SP2043 blocks HGF-induced phospho-Met (Y1234/1235/1349), phospho-Gab1 (Y307), and phospho-MAPK (T202/Y204) in MEC. (b) SP2043 blocks IGF1 induced phosphorylation of IGF1R (Y1135/1136), MAPK and Akt (S473) in MEC. (c) SP2043 blocks HGF-induced phosphorylation of Met (Y1234/1235), Akt, and MAPK in LEC. (d) SP2043 also blocks IGF1 induced phosphorylation of IGF1R (Y1135/1136) and MAPK in LEC. All right panels represent GAPDH normalized western intensities. Original gel images of data (a,b,c,d) are presented in Supplementary Fig. S6 online.

brachial lymph nodes was inhibited by treating animals with an anti-lymphangiogenic and anti-angiogenic peptide from the transmembrane 45A (TMEM45A) protein. We hypothesized that SP2043 would show similar inhibitory effects because of its anti-angiogenic and anti-lymphangiogenic properties, preventing the formation of the pre-metastatic niche and subsequent metastasis to these organs. SP2043 was evaluated in the TCM-induced metastasis model (Supplementary Fig. S5 online). Fifty μl of TCM was subcutaneously administered together with or without SP2043 (10 mg/kg, i.p.) for two weeks, after which luciferase-expressing MDA-MB-231 (luc-MDA-MB-231) tumors were established in the inguinal mammary fat pads (Supplementary Fig. S5 online). The primary tumor size is not influenced by either peptide or TCM pre-treatment, as the luc-MDA-MB-231 cancer cells were inoculated after completing the TCM and peptide pre-treatment (Supplementary Fig. S5 online). However, these treatments do significantly
influence metastatic progression. By week 5, 3 of the 10 mice in the SFM treated group have metastases, in the TCM treated group 9 of the 10 mice have metastases (90%), but in the SP2043 treatment group there are fewer than 50% of metastases compared to the positive control group (Fig. 5a). At week 5, the brachial lymph nodes, lungs, livers, brains, humerus/ulna, and femur/tibia were harvested, bathed in D-luciferin for 5–10 min and imaged using the IVIS imager (Fig. 5b). The LN, lungs, and livers from TCM treated animals show large numbers of metastases while the same tissues from peptide treatment animals have fewer metastases. Since the tumors were inoculated in the inguinal mammary fat pads, only the LN and lung metastasis can be considered to be a result of distant metastasis in the model we used (Supplementary Fig. S5 online). Liver metastasis is a result of adjacent metastasis, as we showed previously in Lee et al11. In that study, inguinal tumor inoculation resulted in abdominal metastasis and the liver and the intestine were the major organs involved in this adjacent metastasis. Luciferase-mediated photon flux from these organs shows that SP2043 significantly blocks lung and LN metastasis (Fig. 5e). Antibodies for human vimentin, a marker for metastatic cancer cells26, were used to detect metastases in the LN (Fig. 5e,f). TCM-treated positive LN show macro-metastases (>2 mm), compared to SP2043 treated LN (Fig. 5e). Importantly, the metastases are highly colocalized with the lymphatic endothelium in the LN (Fig. 5f). The enlarged image shows the association of the invading breast cancer cells (red) with the lymphatic vessels (green) in the LN (Fig. 5f), demonstrating that the anti-lymphangiogenic activity of SP2043 inhibited metastasis.

**Figure 3** | IGF1R and Met binding receptor proteins. We identified the IGF1R and Met binding receptor proteins that are induced to make a complex by IGF1 and HGF, as well as those that are disassociated by SP2043 treatment. (a) Eleven receptor proteins in the newly formed receptor complex with IGF1R (CD58, CD155, CD31, CD36, CD44, CD147, RECK, ADAM17, Galectin-3, Integrin β1, Integrin β2) in response to IGF-1 treatment. After SP2043 treatment, 8 proteins (CD31, CD147, Integrin β1, RECK, Galectin-3, CD58, CD155, ADAM17) were dissociated from the receptor complex. (b) Eight proteins formed a complex with Met (CD58, CD155, TIMP3, CD36, CD154, CD44, ADAM17, Integrin β2) in response to HGF treatment. After SP2043 treatment, 7 proteins (CD58, CD155, TIMP3, CD36, CD154, CD44, ADAM17) were dissociated. (c) CD58, CD155, ADAM17 were targeted by SP2043 to dissociate from the receptor complex shown in a Venn diagram.

**Discussion**

This study shows that the collagen IV derived biomimetic peptide SP2043 inhibits triple negative breast cancer (TNBC) growth and metastasis. The main mechanism by which SP2043 manifests its anti-cancer effects is by blocking IGF1R and Met signaling in lymphatic and blood endothelial cells. This results in blockage of lymphangiogenesis and angiogenesis in breast cancer primary tumors and in pre-metastatic lungs and lymph nodes, thus inhibiting primary tumor growth and metastasis. Our novel peptide SP2043 (amino acid sequence: LRRFSTAPFADINF) was designed by replacing some natural or synthetic amino acids from our previous collagen IV peptide SP2012 (LRRFSMTMPMF-Aba-NINNV-Abu-NF) for which in vitro and in vivo studies from our laboratory were published.
earlier. These modifications delete the synthetic amino acid Abu (2-aminobutyric acid) and alkyl chain M (methionine), and replace the hydrophobic N (asparagine) with negatively charged D (aspartic acid) for greater water solubility, making the peptide agent translatable and easy to synthesize for the next scale-up step. These modifications were performed based on previous expertise in SAR (Structure & Activity Relationship) analysis in the laboratory.

SP2043 targets both lymphangiogenesis and angiogenesis. Though anti-angiogenic therapies inhibit primary tumor growth, patients suffer from metastasis facilitated by tumor lymphatic vessels which current anti-angiogenic agents, including anti-VEGF agents, are not able to inhibit. In breast cancer, tumor lymphatic vessels are the preferential routes of tumor dissemination. FDA-approved anti-lymphangiogenic therapeutic agents have not been developed to date, although a number of anti-angiogenic therapeutic agents are in the clinic and in development. The SP2043 peptide exhibits both anti-lymphangiogenic and anti-angiogenic activity by targeting two different receptor tyrosine kinases: IGF1R and Met in lymphatic and blood endothelial cells (Fig. 2). IGF1-PI3K/Akt signaling has been studied in skin cancer progression. IGF1 regulates the expression of the pituitary tumor transforming gene (PTTG), an oncogene in breast tumor. Met is also associated with cancer growth, resistance and metastasis. Physiological Met signaling regulates angiogenesis and lymphangiogenesis in ductal carcinoma in situ, thus implicating Met signaling in breast diseases. HGF is a well-known lymphangiogenic...
Figure 5 | Inhibition of tumor-conditioned media induced spontaneous metastasis by SP2043. Tumor-conditioned media (TCM) pre-treatment followed by tumor inoculation induce accelerated metastasis formation. SP2043 was administered (10 mg/kg, i.p.) during the TCM induction phase and the resulting effect on metastasis was observed for 5 weeks. (a) Number of mice with thoracic metastasis determined every week. (b) Number of organs with metastases ex vivo at week 5. (c) Mean photon flux from the LN and lungs. Luciferase expressed by MDA-MB-231 cells emit luminescent signal in the presence of D-luciferin (*P < 0.05, **P < 0.01). (d) Representative images of the harvested organs. (e) LN metastases. Anti-human vimentin antibodies detect metastatic human breast cancer cells in the LN. LN from animals treated with TCM showed macro-metastases (>2 mm); SP2043 treatment potently prevents LN metastasis. Scale bars represent 1,000 μm. (f) Colocalization of the metastases and lymphatic vessels in the LN. Scale bars in the whole images represent 1,000 μm. Enlarged images also showed this colocalization. The scale bar in the enlarged image represents 200 μm. Data (c) are reported as mean ± s.e.m.
Figure 6 | SP2043 inhibits tumor-conditioned media induced lymphangiogenesis and angiogenesis within pre-metastatic organs. (a) Activity of SP2043 in LEC tube formation induced by TCM at 18 h. SFM was used as a negative control. Scale bars represent 200 μm. (b) From the metastasis models in Fig. 5, we selected SFM-, TCM-, and SP2043-treated LN and lungs without metastases. These selected organs were stained with anti-mouse LYVE-1 and anti-mouse lectin. TCM treated LN showed enhanced lymphangiogenesis and angiogenesis, but SP2043 treatment inhibited them (upper). SP2043 inhibited the increased lymphangiogenesis seen in the lungs of animals treated with TCM. Angiogenesis, however, was not significantly influenced by either TCM or SP2043 (lower). Scale bars represent 1,000 μm. (c) Quantification of (b) upper panels (*P < 0.05). (d) Quantification of (b) lower panels (*P < 0.05). Data (c,d) are reported as mean ± s.e.m.
genic factor\textsuperscript{22}, thus the HGF-Met axis has been recognized to have a role in lymphogenous metastasis. For example, intratumoral Met expression is associated with VEGFC expression, lymphangiogenesis, and LN metastasis\textsuperscript{35}. Recently, Met mediated tumor drug resistance has been reported in several types of cancer\textsuperscript{24-26}. Therefore, the multimodal activity against different receptor tyrosine kinases would be a promising way to overcome tumor-drug resistance that usually arise after using mono-targeted therapies.

We previously reported molecular target data with an earlier version of the peptide, SP2012. In that study, we showed that \( \beta_1 \) integrin is a key target of SP2012 by using a receptor pull-down assay. SP2012 binds to \( \beta_1 \) integrin in cell lysates from HUVEC; however, SP2012 does not bind to cell lysates from HUVEC in which \( \beta_1 \) integrin is knocked down by siRNA\textsuperscript{2}. Integrins communicate with multiple receptor-tyrosine kinases (RTKs) and can be targeted for cancer angiogenesis/lymphangiogenesis\textsuperscript{29-41}. This communication is achieved by the formation of a receptor complex that includes integrins and the extracellular parts of particular growth factor receptors as well as other co-receptors. If the peptide binds to domains of integrins resulting in a loosening of the crucial binding pockets or in critical structural changes, the integrins may not appropriately interact with the RTKs and fail to promote signaling from the RTKs. SP2043 differs from SP2012 by 6 amino acids, making SP2043 easier to manufacture and more translational for use in humans. A combination of co-immunoprecipitation and reverse western assays detecting diverse soluble receptors or factors showed that SP2043 treatment disrupts the receptor complexes formed by IGF1R and Met in the presence of IGF1 (Fig. 3). One interesting point is that the \( \beta_1 \) integrin, which is a target of SP2012\textsuperscript{6,8}, is found to be a co-receptor of only IGF1R upon IGF1 activation, and not in a complex with Met after HGF induction in endothelial cells. Based on these data, we conclude that SP2043 could target more than one receptor in addition to the \( \beta_1 \) integrin, including binding to a new unknown receptor that regulates IGF1R and Met signaling. In the current manuscript, however, we focused on the potent anti-metastatic effects of SP2043 in breast cancer models in vivo and identified IGF1R and Met on lymphatic and blood endothelial cells as potential targets. Identification of other possible targets of SP2043 will be the focus of future studies.

The co-receptor data additionally revealed that CD58, CD155, and ADAM17 are completely eliminated after SP2043 treatment from IGF1R and Met co-receptor complexes in the presence of IGF1 and HGF (Fig. 3c). We hypothesize that ADAM17 could be an important target of the SP2043 peptide, as ADAM17 has been widely studied in angiogenesis and cancers\textsuperscript{32,43}, and specifically in breast cancer\textsuperscript{44-46}. Although CD58 and CD155 have not been directly implicated in angiogenesis and lymphangiogenesis, CD58 has been reported to promote self-renewal of tumor initiating cells\textsuperscript{19}, and CD155 is known to induce tumor cell invasion and migration\textsuperscript{20}. Hence CD58 and CD155 could be novel targets of the peptide to directly inhibit breast cancer growth and metastasis. Studies to measure SP2043 peptide binding to/interacting with these receptors and functional assays of angiogenesis, lymphangiogenesis, and tumorigenesis with neutralizing antibodies against these receptors remain subjects for future investigations. This report focuses on the discovery of the dual inhibitory activity of the peptide to efficiently block lymphangiogenesis and angiogenesis, as well as breast cancer growth and metastasis.

SP2043 dramatically inhibits the growth of MDA-MB-231 tumor xenografts (Fig. 4a). The blockade of tumor growth is a result of both the anti-angiogenic (Fig. 4b-d) and possibly anti-tumorigenic activities of SP2043 (Supplementary Fig. S7 online). The peptide also dramatically blocks peri-tumoral lymphangiogenesis in the tumor stroma (Fig. 4e). This is noteworthy for the prevention of metastasis, since the peritumoral lymphatics serve as the initial entry point into the lymphatic system for the dissemination of tumor cells for lymphatic metastasis\textsuperscript{47}. The SP2043 treatment regimen (10, 20 mg/kg/day, i.p. for a month) described in the paper was designed to initially evaluate the therapeutic efficacy of SP2043 in breast tumor models, but this may not be the final formulation of the drug. Most peptide agents may not be sufficiently stable in vivo, thus they need to be modified with non-natural amino acids or be encapsulated in appropriate drug carriers to extend their half-life\textsuperscript{48,49}. We have designed PLGA [Poly(D,L-lactide-co-glycolide)] based nanoparticles to load a Serpin derived anti-angiogenic peptide, and the formulation has been tested in mouse models for age-related macular degeneration (AMD)\textsuperscript{50}. The encapsulated peptide formulation, which is a more practical solution in the clinic, showed potent therapeutic effects in vivo.

As conventional spontaneous metastasis models take more than 2 months to show significant metastasis, tumor-conditioned media (TCM) induced metastasis models were employed (Supplementary Fig. S5 online)\textsuperscript{11,12}. The TCM injection mimics the release of tumor secretions from the primary tumor containing pro-metastatic factors in patients and conditions pre-metastatic organs to prepare them for tumor metastasis. In the orthotopic tumor xenografts established in inguinal mammary fat pads dramatic distal metastasis occurs within 4–5 weeks\textsuperscript{11}. SP2043 (10 mg/kg) was administered i.p. only during the 2 week TCM induction phase; it was not administered after tumor inoculation (Supplementary Fig. S5 online). The experiment was designed in this way to determine the efficacy of the peptide in inhibition of metastatic dissemination and colonization by specifically disrupting pre-metastatic niche formation before tumor inoculation. If the peptide was injected after tumor inoculation, the peptide would significantly inhibit tumor growth as well making it difficult to interpret any decrease in metastases. The decrease could have been a result of mechanism based lymphatic and blood vessel targeting or just from the relatively smaller size of the tumors\textsuperscript{50}. In this experiment the tumor sizes are identical within all the experimental groups (Supplementary Fig. S5 online). This temporal treatment with SP2043 inhibits only metastatic dissemination and colonization (Fig. 5). LN metastasis is highly correlated with lymphatic vessel density (Fig. 5f) and SP2043 treatment dramatically inhibits lymphangiogenesis and metastasis in the LN of animals treated with TCM (Fig. 6b). Angiogenesis is also crucial for metastatic colonization in the LN as the invading tumor cells require new blood vessels to survive and make colonies at the new location\textsuperscript{51}. Hence, TCM-induced LN angiogenesis can facilitate tumor metastasis. SP2043 prevents TCM-induced angiogenesis in LN and thus inhibits the growth of the metastases there (Fig. 6c). The lungs are well vascularized organs and so we could not see any dramatic changes in pulmonary angiogenesis after treating with either TCM or SP2043. However, TCM induced lymphatic expansion is significant in the lungs and SP2043 treatment inhibits it (Fig. 6d). Determination of the roles of the lymphatic vessels in the metastatic lungs remains for future studies.

The TCM-induced metastasis model and the anti-metastatic activity of our peptide provide clinical insights and a rationale for using the peptide agent to prevent metastasis in cancer patients in both the adjuvant and neoadjuvant settings. In the neoadjuvant setting, where the tumor is not excised during chemotherapy, the stressed tumor may be secreting factors to promote angiogenesis and lymphangiogenesis at distant sites to allow it to move to a more favorable site where the local tumor microenvironment may protect it. Similarly in the adjuvant setting, although the tumor has been excised, the long-lived factors secreted by the tumor prior to excision could also participate in promoting metastasis by, for example, conditioning the pre-metastatic organs facilitating the circulating tumor cells (CTCs) to invade the organs and colonize them. If the patients are treated with the peptide possibly in combination with chemotherapy, CTC proliferation and survival and pre-metastatic niche formation would be inhibited. As a result, secondary recur-
rence of the tumor, including metastases and primary tumor regrowth may be more effectively prevented.

In summary, the anti-lymphangiogenic and anti-angiogenic peptide SP2043 inhibits breast cancer growth and metastasis. As angiogenesis and lymphangiogenesis are central to metastatic cell dissemination and metastatic disease is the primary cause of death in cancer, this peptide could be a potential treatment for metastatic disease.

**Methods**

**Peptide synthesis and handling.** SP2043 and scrambled-SP2043 peptides were produced by New England Peptide (Gardner, MA) by using a solid-phase synthesis technique. HPLC and M5 analyses of the peptides were provided by the manufacturer to indicate a purity of more than 95%. Peptides were solubilized in 5% DMSO in PBS and the final concentration of DMSO was less than 0.3% in all the experiments. In our previous study, Rosca et al showed that the NINNV sequence in our collagen IV and the final concentration of DMSO was less than 0.3% in all the experiments. In our technique. HPLC and MS analyses of the peptides were provided by the manufacturer.

**Methods**

**Cell viability assay.** Viability assays using the WST-1 (Roche, Indianapolis, IN) reagent was performed on endothelial cells and breast cancer cells as previously described. Briefly, cancer cells (1,500/well in RPMI-1640) or endothelial cells (2,000/well in EGM) were plated in 96-well plates and incubated overnight. Normal media was replaced with the media with or without peptides and the plates were incubated for 72 h. The controls for cell viability assays correspond to cell phenotypes in normal endothelial cells (EC-2) in the HUVEC (E-plate, EGM-2) or in the xenograft model (tetraploid/AngioXenograft, EGM-2 and EGM-3 experiments) without peptide treatment. Media was removed and WST-1 solution (1:100 diluted in serum-free media) was added. After incubation for 4 h, the absorbance at 450 nm was measured by the Victor V plate reader (Perkin Elmer). IC50 values of the peptide were calculated by using GraphPad Prism 5 (GraphPad Software Inc.).

**Cell migration and adhesion assays.** Cell migration and adhesion assays on endothelial cells were performed by using the Real-Time Cell Analysis (RTCA) system (ACEA) as described previously. The ACEA RTCA system measures electrical impedance in real time that is expressed as cell index. When cells are in contact with the gold electrodes, electrical impedance increases. The resulting change in cell index indicates the extent of cell migration from the top compartment to the bottom compartment of a CIM plate, or of cell adhesion to the bottom of the well in an E-plate. In migration, the membrane of the top chamber of a CIM plate was coated with fibronectin. 180 µL of EGM-2 (HUVEC) or EGM-2M (LEC or MEC) was added to the bottom chambers, as a chemoattractant. The two chambers were assembled together, 30 µL of serum free media was added to the top chamber and the assembled plate was incubated in a tissue culture incubator at 37°C for 1 h. The equilibrated plate was removed from the incubator and 100 µL of the trypsized cells (45,000 HUVEC/well; 30,000 MEC/well; 120,000 LEC/well; 200,000) without or with peptides were added to the top chamber. After 30 min incubation at room temperature, the stabilized chamber was loaded in the RTCA machine and the cell index was measured continuously for 20 h. Cell indices at 20 h were selected for analysis. In adhesion assays, 100 µL of 2% concentrated peptide solutions were added to the appropriate E-plates. LEC or MEC (150,000 cells/well) or HUVEC (100 µL of EGM-2M (LEC or MEC) or EGM-2 (HUVEC) media) were added next to each well diluting the peptides to the appropriate final concentrations. After equilibrating at room temperature for 30 min, the E-plate was loaded into the RTCA personal system. Cell indices at 3 h were analyzed.

**Tube formation assay.** Tube formation assays were performed as described previously. 50 µL of Matrigel was loaded in each well of the 96-well plate and the plate was incubated at 37°C for 30 min. Trypsinized HUVEC, LEC, MEC were mixed with the peptide making the appropriate cell density (15,000 cells/well) and peptide concentrations. 100 µL of cell and peptide mixture was added on top of the gel in the 96-well plate. The plates were incubated for 24 h in a CO2 incubator and the formation of the capillary-like tubes was observed after 18 h using a Nikon microscope. We quantified tube formation by using Angiogeny Transporter for ImageJ (NIH). The process was described in the Supplementary Fig. S8 online. Briefly, the original jpeg images (three images per group) were transformed to binary images. The binary images were treated with the “fill hole” method to minimize shadow effects. The prepared images were skeletonized. The skeletonized binary tree was analyzed and all the values were normalized with the analysis area, making the pixel number of the area equal to 100,000 in all the groups. Among several categories of quantification, we chose number of nodes (Nb nodes), Nb junctions, Nb segments, and Nb branches. Also we considered total length (Total length), Tot branching length, and Tot segment length.

**Western blot assay.** MEC, LEC and MDA-MB-231 cells were plated on 100 mm tissue culture dishes at 1,000,000 cells/dish. After allowing a day for cell attachment, the cells were starved in serum-free media (SMF) for 24 h, after which SP2043 was added for 90 min. Hepatocyte Growth Factor (HGF) (50 ng/ml) or Insulin-like Growth Factor (IGF1) (100 ng/ml) was added and the incubation was continued for 10–30 min at 37°C. Generation of cell lysates, SDS-PAGE and westerns were done as described in our previous study. The primary antibodies that we used include p-Met (Y1234/1235), p-Met (Y1349), p-Akt (S473), Akt, p-MAPK (T202/204), MAPK, p-IGF1R (Y1135/1136), IGFIR, and GAPDH (all from Cell Signaling). All the original images of immunoblot analyses are presented in Supplementary Fig. S6 online.

**Identification of IGFIR/Met binding proteins.** We hypothesized that the SP2043 peptide binds to unknown receptor proteins on endothelial cells and interrupts both IGFIR and Met. Experiments were designed to determine which receptor proteins make a complex with IGFIR or Met in the presence of IGF1 or HGF and which of these co-receptor proteins are dissociated from IGFIR or Met after treating with SP2043. MEC were plated on tissue culture–treated 100 mm dishes at 1,000,000 cells/dish in EGM-2M. After allowing a day for cell attachment, the cells were starved in SMF for 24 h, after which SP2043 (30 µM) was added for 90 min. HGF (100 ng/ml) or IGF1 (100 ng/ml) was added and the incubation was continued for 60 min at 37°C. Treatments were stopped by adding cold PBS and cell lysates were prepared using Pierce IP lysis buffer (Thermo Scientific) with 100 µl/ml protease inhibitor (Sigma) and 100 µl/ml phosphatase inhibitor (Sigma). The cell lysates were incubated with the anti-IGFIR or anti-Met antibody (1 : 50, Cell Signaling) overnight, after which protein A/G PLUS agarose (Santa Cruz) was added and incubated for 3 h to pull down the antibody–receptor complex. After removing the supernatant and washing with PBS, the PBS-rinsed pellets were treated with glycine (pH 2, 10.0 ml) for 2 h to dissociate all the proteins bound to the beads. The beads were then removed by centrifugation. To allow the proteins to recover their structures and to maintain normal pH, 0.5 ml of Tris (pH 8) was added. The mixture was dialyzed against Amicon Ultra 3K filter units (Millipore) to remove glycine and Tris. The resulting mixture was analyzed using reverse antibody arrays for human soluble receptors (Proteome profiler human cytokine array kit, R&D systems) followed by the product manual.

**MDA-MB-231 tumor xenografts.** MDA-MB-231 tumor xenografts were established as described previously. SP2043 (10 or 20 mg/kg) or vehicle was injected daily intraperitoneally (i.p.), and the administration was continued for 28 days. The tumor size and mouse body weight were measured every four days. The tumor volume was calculated by using the formula: \( V = 0.52 \times (\text{long axis} \times \text{short axis}) \) of the tumor.

**TCM-induced spontaneous metastasis models.** The tumor-conditioned media (TCM) induced spontaneous metastasis model that we previously developed was used to accomplish accelerated metastasis and to evaluate the anti-metastatic effects of SP2043. MDA-MB-231-luc-D3H2LN cells were treated with either TCM or SFM for 24 h, after which SP2043 (30 µM) was added for 90 min. HGF (100 ng/ml) or IGF1 (100 ng/ml) was added and the incubation was continued for 60 min at 37°C. Treatments were stopped by adding cold PBS and cell lysates were prepared using Pierce IP lysis buffer (Thermo Scientific) with 1000 µl/ml protease inhibitor (Sigma) and 1000 µl/ml phosphatase inhibitor (Sigma). The cell lysates were incubated with the anti-IGFIR or anti-Met antibody (1 : 50, Cell Signaling) overnight, after which protein A/G PLUS agarose (Santa Cruz) was added and incubated for 3 h to pull down the antibody–receptor complex. After removing the supernatant and washing with PBS, the PBS-rinsed pellets were treated with glycine (pH 2, 10.0 ml) for 2 h to dissociate all the proteins bound to the beads. The beads were then removed by centrifugation. To allow the proteins to recover their structures and to maintain normal pH, 0.5 ml of Tris (pH 8) was added. The mixture was dialyzed against Amicon Ultra 3K filter units (Millipore) to remove glycine and Tris. The resulting mixture was analyzed using reverse antibody arrays for human soluble receptors (Proteome profiler human cytokine array kit, R&D systems) followed by the product manual.

**Immunoﬂuorescence.** Immunoﬂuorescence was performed following the protocol described previously. Primary antibodies that we used include rabbit anti-mouse LYVE-1 antibody (1 : 200, AngioTech), rat anti-mouse CD31 antibody (1 : 100, BD Pharmingen), rabbit anti-phospho-Met (1 : 400, Cell Signaling), mouse anti-human PCNA (1 : 100, BD Pharmingen), mouse anti-human vimentin (1 : 50, Santa Cruz), and goat anti-mouse lectin FITC (1 : 100, Sigma). Secondary antibodies include: FITC-conjugated goat anti-rat, FITC-conjugated goat anti-rabbit, rhodamine-conjugated goat anti-mouse, and goat anti-mouse lectin FITC (1 : 100, Sigma). Signal detection was performed using the Alexa Fluor 488 goat anti-rabbit (1 : 500, all from Jackson ImmunoResearch). Fluorescent signals were visualized and digital images were obtained using the LSM-510 confocal microscope (Carl Zeiss). We quantified the images using ImageJ (NIH, Bethesda, MD), measuring...
Statistical analysis. Error bars correspond to SEM, unless otherwise stated. Differences between the control and the peptide treated group are regarded as significant when P is less than 0.05 using the Student’s t-test.

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Author contributions

E.L., N.B.P. and A.S.P. conceived the work. E.L. performed most of the experiments. S.J.L. performed western blot assays with blood and lymphatic endothelial cells. J.E.K. performed HUVEC viability, migration and adhesion assays. Z.H. performed western blot assays with breast cancer cells. E.L., S.J.L., J.E.K., Z.H., N.B.P. and A.S.P. interpreted the experimental data. E.L. wrote the manuscript, and E.L., N.B.P. and A.S.P. edited the manuscript.

Additional information

Supplementary information accompanies this paper at http://www.nature.com/scientificreports

Competing financial interests: Yes there is potential Competing Interest. ASP is a co-founder and serves as the CSO of AsclepiX Therapeutics, LLC; the terms of this arrangement are being managed by the Johns Hopkins University in accordance with its conflict of interest policies.

How to cite this article: Lee, E. et al. Inhibition of breast cancer growth and metastasis by a biomimetic peptide. Sci. Rep. 4, 7139; DOI:10.1038/srep07139 (2014).

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