Detection of Porcine Cysticercosis in Nagaland of North East, India

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A B S T R A C T

A total of 360 pig carcasses were examined through meat inspection in different market places of Dimapur and Kohima district of Nagaland, India in the month of March, 2019. Out of which, 6 (1.67%) were found positive for porcine cysticercosis with visible cysts. The serum samples were collected from 300 pigs and out of which 10 (3.00%) serum samples were found positive for *Cysticercus cellulosae* antibody. Polymerase chain reaction (PCR) assay was performed to confirm *Cysticercus cellulosae* and to validate the results of meat inspection. Oligonucleotide primers targeting against the large subunit rRNA gene (TBR primers) of *Taenia solium* were used in this study. On reactivity in PCR test, the TBR primers yielded products of 286bp in cysticercosis positive cases.

Keywords
Pig, Nagaland, Cysticercus cellulosae, PCR, Taenia solium

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Introduction

*Cysticercus cellulosae*, the metacestode stage of *Taenia solium* is an underrated and a neglected zoonotic disease involving pig and man (Willingham *et al.*, 2008). Porcine cysticercosis plays a crucial role in the transmission and maintenance of human taeniosis and cysticercosis (Pinto *et al.*, 2000). In Indian context, meat consumption is greatly influenced by culture, traditions, customs and taboos especially in the rural societies. In north eastern states of India pork is considered as a traditional food item (Anonymous. 2012) and has much higher pork consumption than the other parts of the country. The demand for pork was increasing along with prices in both Assam and Nagaland according to the traders. Thus, accurate inspection of pork is very important.

Although the pork consumption is highest in North East India but the scientific procedure of diagnosis and meat inspection is still lacking. While meat inspection is the preferred diagnostic tool to detect heavily infected carcasses but it is not reliable in detecting lightly infected carcasses (Cai *et al.*, 2006). So detection of antibodies by ELISA (Ab-ELISA) can be used as an alternative tool for the diagnosis of porcine cysticercosis (Deckers *et al.*, 2008).
Moreover, meat inspection requires expertise of the meat inspector; otherwise cysticerci may be confused with *T. hydatigena* cysticerci (Kedra *et al.*, 2001), hydatid cysts (Deplazes *et al.*, 2005) and left over of muscle fasciae (Geysen *et al.*, 2007). To overcome such type of difficulties application of ELISA and molecular tools like Polymerase chain reaction (PCR) can be effective in diagnosis and validation of meat inspection results.

So considering the public health significance of the disease, the purpose of this study was to evaluate the performance of ELISA for ante-mortem diagnosis in pigs and estimating the prevalence of porcine cysticercosis in slaughtered pig through meat inspection and PCR for validation of results.

**Materials and Methods**

**Study area**

The study was carried out between January 2019 to February 2019 in Dimapur and Kohima district of Nagaland, India. Three blocks from each district were selected for collection of sera sample and for carcass examination.

**Sample collection**

**Collection of blood samples**

A total of 300 blood samples were collected from the anterior vena cava of pigs by using BDV accutainer® needles (gauge 19) and BDV acutainer® plain tubes (10 ml). The blood samples were kept standing in an ice box at +4°C to ensure no haemolysis occurred while in the field.

At the laboratory, blood was centrifuged to separate serum from blood clot. Serum was harvested into barcoded 2 ml vials that were stored at −20°C until processing.

**Carcass Inspection**

A total of 360 pig carcasses were examined as per the standard method [10] for the presence of *Cysticercus cellulosae* from different market places of Dimapur and Kohima district. Majority of the pigs were brought from Uttar Pradesh, India to Dimapur and distributed to the rest of the districts of Nagaland on weekly basis (Fig. 1). Fifty gram (g) of tissue each from brain, tongue, liver and skeletal muscles of infected pig carcasses were brought to the laboratory in ice-box

**Serological tests**

ELISA was performed as per the manufacturer’s protocol of RIDASCREEN® *Taenia solium* IgG kit (RBiopharm AG, Germany). The absorbance was read at 450 nm with an ELISA reader (Lab systems Multiskan Plus, Thermo Fisher Scientific, USA). Samples having percent positivity value 0.50 or above (%P ≥ 0.05) were categorized as positive and below 0.50 as negative.

**Extraction of DNA from cysts**

Extraction of DNA from cysts/suspected lesion was made possible using a commercially available QIAamp tissue kit (QIAGEN, Hilden, Germany) according to the manufacturer’s instructions (Kolesarova *et al.*, 2012). Briefly, 200 μl of cyst/lesion homogenate, 20 μl of proteinase K stock solution, and 200 μl of lysing buffer were pipetted into 1.5 ml Eppendorf tube. The mixture was incubated at 37 °C for 1 h and then at 70 °C for 30 min before the addition of 200 μl of absolute alcohol and mixing by vortexing. The mixture was then transferred to the QIAamp spin column placed in a clean 2 ml collection tube and centrifuged at 8000 RPM in MiniSpin centrifuge (Eppendorf, Wesseling-Berzdorf, Germany) for 1 min at
room temperature. The QIAamp spin column was washed twice with 500 μl of the washing buffers by spinning for 1 min. The QIAamp spin column was placed in a clean 1.5 ml eppendorf tube and the DNA was eluted with 100 μl of elution buffer preheated at 70 °C. Maximum DNA yield was obtained by spinning at 12,000 rpm for 1 min at room temperature. From the suspended nucleic acid 5 μl was used in the PCR amplification. The extracted DNA was quantified using spectrophotometer at 260 nm wave length

**Oligonucleotide primers**

The oligonucleotide primers specific to *T. solium* were adopted from already published sequences. The primers TBR3 (5′-GGC TTG TTT GAATGG TTT GAC G-3) and TBR-6 (5′-GCT ACTACA CCT AAA TTC TAA CC-3) against large subunit rRNA gene (Jardim et al., 2006).

**PCR amplification and detection of PCR product**

The PCR reaction was performed in a thermal cycler (Eppendorf, Hamburg, Germany) in 20 μL volume containing 2μl DNA sample (100 ng/ μl), 1 μl (10 pmol) of each forward and reverse primer, 10 μL of DreamTaq Green PCR Master Mix (2X) (Thermo Scientific™, USA) and 7 μL of nuclease free water (Thermo Scientific™, USA). A total of 40 PCR cycles were run with the following conditions: one initial denaturation cycle at 94°C for 3 min, followed by 40 repeated cycles with temperatures at 94°C for 30 s (denaturation), 59°C for 30 s (annealing, specific for primers) and 72°C for 1 min. After the final cycle, the preparations were kept at 72°C for 5 min for final elongation, and the PCR products were stored at 4°C in thermal cycler for further use (Sreedevi et al., 2012). Five microliters of PCR amplicons was analyzed on ethidium bromide stained 2% agarose gel (In Genius Gel documentation system, Syngene, UK). The sizes and quantities of PCR products were verified by comparison with a 100 bp plus quantitative ladder (Thermo Scientific™, USA).

**Results and Discussion**

Overall sero-prevalence of porcine cysticercosis in Dimapur and Kohimadistict was record as3.00 % (Table 2). Highersero-prevalence of porcine cysticercosis was recorded in Dimapur (3.33 %) compared to Kohima district (2.67%) of Nagaland. These finding were in close conformity of Barua et al., (2018) where they recorded sero-prevalence of porcine cysticercosis in 4 states of North EastIndiaas 2.72 %.

This might be because, in our study area, the vast majority of pigs are confined in pens, even among smallholders, compared to more extensive (i.e., free roaming and scavenging) pig husbandry practices which may predominate in the other study settings.

All the 360 pigs inspected at slaughter were from different market places of Dimapur and Kohima district of Nagaland, India (Fig. 2 & Fig. 3). The overall prevalence of porcine cysticercosis based on post-mortem inspection was 1.67% (Table 1). The prevalence was higher in Dimapur (2.22 %) compared to Kohima district (1.67%) of Nagaland.

Previously, Sarma (1977) recorded 6.64 % positive cysticercosis from greater Guwahati of Assam. Plain (1991) recorded highest infection 11.90 % infection from North Eastern region of Assam. Borkataki et al., (2012) conduct study in three districts of Assam for a period of one year and found 93 pigs (9.50%) positive for cysticercosis out of 978 pigs. Although the previous workers recorded higher prevalence of porcine cysticercosis, Barua et al., (2018) recorded
prevalence as 0.92% in 4 states of North East India. This might be due to importing of scavenging pigs from other parts of India as reported by butchers.

**Table 1** Prevalence of porcine cysticercosis pigs of Dimapur and Kohima district of Nagaland

| District | Study area | Animals inspected(No.) | Cyst positive pig (%) |
|----------|------------|------------------------|-----------------------|
| Dimapur  | Medziphema | 60                     | 1 (1.67%)             |
|          | Khubboto   | 60                     | 2 (3.33%)             |
|          | Nihokhu    | 60                     | 1 (1.67%)             |
|          | **Total**  | 180                    | **4 (2.22%)**         |
| Kohima   | Sechu-Zubza| 60                     | 1 (1.67%)             |
|          | Kohima     | 60                     | 0 (0.00%)             |
|          | Zakhama    | 60                     | 1 (1.67%)             |
|          | **Total**  | 180                    | 2 (1.11%)             |
|          | **Overall Total** | 360 | **6 (1.67%)** |

**Table 2** Sero-Prevalence of porcine cysticercosis in Dimapur and Kohima district of Nagaland

| District | Study area     | No. of Pig Serum | Sero Positive (%) |
|----------|----------------|------------------|-------------------|
| Dimapur  | Medziphema     | 50               | 2 (4%)            |
|          | Khubboto       | 50               | 2 (4%)            |
|          | Nihokhu        | 50               | 1 (2%)            |
|          | **Total**      | 150              | **5 (3.33%)**     |
| Kohima   | Sechu-Zubza    | 50               | 2 (4%)            |
|          | Kohima         | 50               | 1 (2%)            |
|          | Zakhama        | 50               | 1 (2%)            |
|          | **Total**      | 150              | **4 (2.67%)**     |
|          | **Overall Total** | 300 | **9 (3%)**     |

**Fig.1** Unloading of pigs in Dimapur brought from Uttar Pradesh, India
Fig.2 Cysts of *Cysticercus cellulosae* in skeletal muscle of pig carcass

![Image of cysts](image1.jpg)

Fig.3 Cysts of *Cysticercus cellulosae* collected from tissue samples

![Image of cysts](image2.jpg)

Fig.4 PCR assay with TBR primers to detect *Cysticercus cellulosae* from pig carcasses. Lane L: 100-bp DNA ladder; Lane 1 to 4: DNA samples extracted from cysticercosis positive pigs. Lane P: Positive control and Lane N: Negative control

![PCR gel](image3.jpg)
Identification of *T. Solium* cysticerci from the infected pig carcasses and suspected carcasses was based on amplification of large subunit rRNA gene (TBR) gene with a product size of 286 bp. All the positive cases show positive amplification of TRB gene (Fig. 4). Dalmasso *et al.*, (2004) extracted DNA from degenerated and calcified cysts/lesions and found positive amplification for TRB gene. Lino Junior (2004) also reported the importance of PCR test with TBR primers as a reliable method for detection of cysticerci in tissues from human autopsies that are in advanced evaluative stages.

From the above study it was found that porcine cysticercosis is still a major public health concern. However there is less information regarding the prevalence of this disease has been recorded from North Eastern states of India. Therefore, an extensive study regarding the prevalence, transmission, risk factors and prevention of this neglected zoonotic disease is utmost concern.

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**References**

Anonymous, 2012. Pork meat in India. www.g-e-f a.de/fileadmin/termine.../Pork_Meat_in_India_-_Report.pdf.
Barua A., Raj H., Goswami C., Sonowal D. And Rajkhowa U., 2018. Prevalence of Porcine Cysticercosis in Four States of North East India. *International Journal of Livestock Research*, 8(10): 212-218.
Cai X.P., Zheng Y.D., Luo X.N., Jing Z.Z., Hu Z.M. and Lu C.P., 2006. Immunodiagnosis of cysticercosis in China. *The Journal of Applied Research*, 6(1): 69-76.
Dalmasso A., Fontanella E., Piatti P., Civera T., Rosati S., Bottero M.T., 2004. A multiplex PCR assay for the identification of animal species in feedstuffs. *Molecular and Cellular Probes*, 18: 81–87.
Deckers, N., Kanobana, K., Silva, M., Gonzalez, A.E., Garcia, H.H., Gilman, R.H. and Dorny, P., 2008. Serological responses in porcine cysticercosis: a link with the parasitological outcome infect. *Int. J. Parasitol.*, 38: 1191–1198.
Deplazes P., Grimm F., Sydler T., Tanner I. and Kapel C.M.O., 2005. Experimental alveolar echinococcosis in pigs, lesion development and serological follow up. *Veterinary Parasitology*, 130: 213-222.
Geysen D., Kanobana K., Victor B., Rodriguez-Hidalgo R., De Borchgrave J. and Brandt J., 2007. Validation of meat inspection results for *Taenia saginata* cysticercosis by PCR restriction fragment length polymorphism. *Journal of Food Protection*, 70(1): 236-240.
Jardim E.A.G.V., Linhares G.F.C., Torres F.A.G., Araujo J.L.B. and Barbosa S.M., 2006. Diferenciaciones specifica entre *Taenia saginata* e *Taenia solium* poresao de PCR e duplex PCR. Specific discrimination between *Taeniasaginata* and *Taenia solium* by one step PCR assay and duplex PCR. *Ciencia Rural, Santa Maria*, 36(1):166-172.
Kedra A.H., Tkach V.V., Swiderski Z. and Pawowski Z., 2001. Intraspecific variability among NADH dehydrogenase subunit 1sequences of *Taenia hydatigena*. *Parasitology International*, 50: 145-148.
Kolesarova M., Herich R., Levkut Jr. M., Curlik J. and Levkut M., 2012. Suitability of different tissue fixatives for subsequent PCR analysis of *Cysticercusovis*. *Helminthologia*, 49(2): 67-70.

Lino Junior R.S., 2004. Cysticercosis diagnostic methods in autopsies. *Revista do Instituto de Medicina Tropical de São Paulo*, 46(3): 138.

Pinto P.S.A., Vaz A.J., Germano P.M.L. and Nakamura P.M., 2000. ELISA test for the diagnosis of cisticercosis in pigs using antigens of *Taenia solium* and *Taenia crassiceps* cysticerci. *Revista do Instituto de Medicina Tropical de São Paulo*, 42(2): 71-79.

Sreedevi C., Hafeez M., Kumar P.A., Rayulu V.C., Subramanyam K.V. and Sudhakar K., 2012. PCR test for detecting *Taenia solium* cysticercosis in pig carcasses. *Tropical Animal Health Production*, 44: 95-99.

Willingham III A.L., Harrison L.J., Fevre E.M. and Parkhouse M.E., 2008. Inaugural meeting of the cisticercosis working group in Europe. *EID J Home*, 14(12).

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