Effectiveness of antioxidant nutraceuticals in attenuating canonical NF-κb signaling in human skeletal muscle resulting from exercise-induced inflammation and oxidative stress

Abstract

Oxidative stress is a process that produces cellular damage by oxidation of cellular components such as lipids, protein, and DNA. It is also a regulator of the acute phase inflammatory response, and also involves the production of reactive oxygen species (ROS) by mitochondrial respiratory activity and from the capillary endothelium in contracting skeletal muscles. Exercise-induced muscle damage is associated with an acute phase inflammatory response. This response results in the production of circulating pro-inflammatory cytokines such as tumor necrosis alpha (TNF-α) which is a mediator of muscle wasting and acts through the stimulation of nuclear factor kappa B (NF-κB). Skeletal muscle damage and injury becomes evident in many catabolic scenarios and is characterized by proteolysis and muscle wasting. Inflammation and oxidative stress are associated with skeletal muscle wasting in various disease states including cancer cachexia, muscular dystrophy, AIDS, sepsis, etc. Antioxidant supplementation has been suggested to attenuate exercise-induced oxidative stress and the subsequent activation of NF-κB through various mechanisms; however, there is still conflicting evidence and more research needs to be completed to better understand the role of NF-κB signaling in skeletal muscle. Antioxidant supplementation has also been shown to attenuate oxidative stress and the subsequent formation of reactive oxygen nitrogen species (ROS, RONS) in skeletal muscle. As a result, this area requires further research as this process has also been demonstrated to mediate NF-κB signaling. Prior evidence suggests that various nutraceutical supplements seem to play a beneficial role in maintaining muscle mass during periods of heightened inflammation and oxidative stress and possible muscle wasting.

Keywords: oxidative stress, inflammation, skeletal muscle, proteolysis, cytokine, antioxidant, nutraceutical

Abbreviations: ROS, reactive oxygen species; RONS, reactive oxygen nitrogen species; TNF, tumor necrosis factor; TNFR, tumor necrosis factor receptor; NF, nuclear factor; COPD, chronic obstructive pulmonary disease; RHD, rel homology domain; IL, interleukin; NIK, NF-κB inducing kinase; IKK, IkB kinase; BCL, b-cell lymphoma; MOD, minimal oligomerization domain; LPS, lipopolysaccharide; RIP, receptor interacting protein; TRAF, tnf-receptor associated factor; TRADD, tnf-associated death domain protein; c-IAP, cellular inhibitor of apoptosis protein; TAK1, tgf-β-activated kinase 1; LT-β, lymphotoxin β; BAFF, b-cell activating factor; CD, cluster of differentiating; MuRF1, murine ring finger-1; UPS, ubiquitin proteasome system; TWEAK, thf-like weak inducer of apoptosis; HSPs, heat shock proteins; NFAT, nuclear t factor of activated t cells

Introduction

Exercise preferentially biased with forced-lengthening (eccentric contractions) causes muscle damage, which is a manifestation of ultra-structural changes and protein degradation, and is associated with the development of an acute phase inflammatory response. This inflammatory response is a microvasculature-induced reaction characterized by the migration of serum proteins and leukocytes from the blood to the skeletal muscle. Involved in this migration are pro-inflammatory cytokines, most notably tumor necrosis factor alpha (TNF-α) which binds its trans-membrane receptor in skeletal muscle and activates a transcription factor known as nuclear factor kappa B (NF-κB). The activation of NF-κB has been found to be a critical factor involved in the multifaceted homeostasis of muscle. This becomes important as the development and growth of skeletal muscle requires a complicated series of events that are regulated through various pathways. The plasticity of skeletal muscle is important to regulate the need for anabolic and catabolic processes, and any perturbation of these processes can lead to various muscle disorders associated with wasting.

Furthermore, exercise-induced muscle damage is also associated with oxidative stress, a process that not only directly causes cellular damage by oxidation of cellular components such as lipids, protein, and DNA, but also acts as a regulator of the acute phase inflammatory response. Oxidative stress instigates the nuclear translocation of certain redox-sensitive transcription factors, pro-inflammatory cytokines, chemokines, and adhesion molecules. The resultant effect is phagocyte infiltration into skeletal muscle, thereby causing proteolysis, ultra-structural damage, and oxidative injury. During
Effectiveness of antioxidant nutraceuticals in attenuating canonical NF-κB signaling in human skeletal muscle resulting from exercise-induced inflammation and oxidative stress

Skeletal muscle damage and injury becomes evident in many catabolic scenarios and is characterized by proteolysis and muscle wasting. Inflammation and oxidative stress are associated with skeletal muscle wasting in various disease states including cancer cachexia, muscular dystrophy, AIDS, sepsis, diabetes, chronic obstructive pulmonary disease (COPD), and cystic fibrosis. Muscle wasting occurs as a result of a reduction in protein synthesis, an increase in protein degradation, or both. Various stimuli prompt skeletal muscle wasting through specific mechanisms and exercise-induced muscle damage is a fitting model for this process. While multiple signaling pathways are involved in the process of skeletal muscle wasting, recent evidence has suggested that NF-κB is one of the most important mechanisms because the activation of the NF-κB pathway stimulates muscle loss. The pro-inflammatory cytokine, TNF-α, is a mediator of muscle wasting and acts through the stimulation of NF-κB. Recent evidence has shown that modification of NF-κB activity through pharmacogenetics can actually prevent skeletal muscle loss in various diseases, injury, and unloading.

There is not an abundance of human studies demonstrating the role that antioxidant supplements may play in down-regulating NF-κB activity. However, enough evidence does exist suggesting a potential beneficial role for this type of nutraceutical supplementation. Therefore, it is feasible to review the potential for various antioxidant nutraceuticals on NF-κB activity, and the subsequent capacity they might possess in attenuating the effects of catabolic NF-κB activity, on oxidative stress and wasting in skeletal muscle. Therefore, the purpose of this brief review is to better understand the underlying mechanisms of NF-κB and its effects on oxidative stress and muscle wasting in humans, as well as possible antioxidant nutraceutical supplements that may attenuate these effects.

NF-κB signaling pathway

NF-κB is a ubiquitously expressed transcription factor that is considered vital to numerous cellular processes. It has been suggested that NF-κB directly modifies hundreds of gene products, including genes that encode cytokines, chemokines, cell adhesion molecules, growth factors, immunoregulatory receptors, acute-phase and stress response proteins, cell surface receptors, transcription factors, and several enzymes involved in protein degradation by the ubiquitin-proteasome system, as well as regulators of redox status, apoptosis, disuse atrophy, and host defense. This is summarized in Figure 1. The NF-κB family is comprised of five genes that code for protein subunits, RelA/p65, RelB, c-Rel, p50, and p52, which can be seen in Figure 2A. In humans, the proto-oncogene c-Rel is a protein that is encoded by the REL gene (v-rel avian reticuloendotheliosis viral oncogene homolog). The c-Rel protein is a member of the NF-κB family of transcription factors and contains a Rel homology domain (RHD) at its N-terminus and two C-terminal trans activation domains. As can be seen in Figure 2B, the IκB family consists of seven proteins including IκBα, IκBβ, IκBε, IκBγ, B-cell lymphoma 3-encoded protein (BCL-3), and precursor proteins p100 and p105. The most commonly described forms of NF-κB are the p50/p65 heterodimer and p50/p50 homodimer complexes, followed by p50/c-Rel and p52/RelB. Unlike the other genes, p50 and p52 genes lack transcriptional activation domains, which generates p50 and p52 homodimers that function as gene repressors by blocking DNA consensus sites, which can be seen in Figure 3.

Activation of the NF-κB complex occurs in response to various stimuli, including infection, exposure to pro-inflammatory cytokines, mitogens, growth factors, biomechanical stressors, and oxidative stressors. There are several pathways by which NF-κB can be activated following ligation of different receptor families, including tumor-necrosis factor receptor (TNFR) and interleukin-1 receptor (IL-1R). NF-κB complex activation also includes the activation of several intermediate kinases, including NF-κB inducing kinase (NIK). These intermediate kinases act as distinct signaling proteins that all unite on the IκB kinase (IKK) complex.
Effectiveness of antioxidant nutraceuticals in attenuating canonical NF-κB signaling in human skeletal muscle resulting from exercise-induced inflammation and oxidative stress

**Figure 2** In A, human NF-κB protein and in B, IκB proteins are illustrated. In A and B, the specific post-translational modifications of both are also illustrated. P, phosphorylation; Ac, acetylation; Ub, ubiquination; TAD, trans activation domain; RHD, rel homology domain; NES, nuclear export signal; NLS, nuclear localization signal; LZ, leucine zipper; DD, death domain; Ankyrin, ankyrin repeats; PEST, polypeptide sequences enriched in proline (P); glutamate (E); serine (S) and threonine (T).

**Figure 3** The NF-κB family members represented by the activating and repressor NF-κB heterodimers.

In regard to the activating complexes, these include the p50/p65, p50/c-Rel, and p52/Rel B heterodimers. Relative to the inhibiting complexes, these involve the p50/p50 and p52/p52 homodimer complexes. Contrary to the other genes, the p50 and p52 genes lack transcriptional activation domains, which generates p50 and p52 homodimers, and function as gene repressors by blocking DNA consensus sites.

**Figure 4** Activation of IκB/NF-κB pathway by a physiological induction with stimuli such as exercise. The contraction of skeletal muscle results in transient increases in cell stress, as evidenced by the accumulation of ROS, energy depletion, etc. that induces phosphorylation of IκBα kinase (IKK) κ. The activated IKKs phosphorylates the inhibitory peptide IκBα, thereby activating it for ubiquination and degradation by the proteasome. At this point, the freed NF-κB translocates to the nucleus and forms dimers to bind DNA and induce trans activation.

The IKK complex controls the breakdown of IκB proteins through regulation of the phosphorylation of serine 32 and 36 on IκBα, and serine 19 and 23 on IκBβ. This phosphorylation results in K48-linked polyubiquitination by the SCFβTrCP3 ubiquitin ligase complex on lysine 21 and 22 of IκBα, which is an ATP-dependent occurrence that quickly targets the breakdown of these proteins by the 26S proteasome. To fully understand the NF-κB pathway, it is necessary to understand the IKK complex and its role in NF-κB activation.

**IKK Complex**

The IKK complex is composed of two catalytic subunits IKKa/IKK1 and IKKβ/IKK2, and one non-catalytic regulatory subunit IKKy/NEMO (NF-κB essential modulator). IKKa and IKKβ maintain a strong sequence homology, especially in their catalytic region, but differ in their specific targets. Both kinases enhance NF-κB trans activation potential by phosphorylation of p65 at serine 536. However, IKKβ can further enhance NF-κB activity by targeting serine 468 of p65. Conversely, IKKa phosphorylates p100 to stimulate partial proteolysis and activation of the alternative NF-κB pathway. NEMO interacts with both IKKa and IKKβ, and associates with intermediates of upstream signaling.

IKK complex activation requires phosphorylation of either IKKa or IKKβ on two particular serine residues within the T loop of the catalytic domain of each kinase. Unfortunately, the kinase(s) that are responsible for T loop phosphorylation are not known. There are two possible mechanisms that have been suggested that may be responsible for phosphorylation. The first potential mechanism is that the IKK proteins phosphorylate each other by trans-autophosphorylation, which is facilitated by either conformational changes within the IKK complex or induced proximity through oligomerization of multiple core IKK complexes. The second possible mechanism could be the existence of separate IKK-kinases that target these residues with conformational changes within the complex exposing the T loop for phosphorylation. It may even be possible that both mechanisms may happen independently or together in a nature dependent manner of the particular upstream signaling pathway (Figure 5).
McKinley S, Willoughby DS. Effectiveness of antioxidant nutraceuticals in attenuating canonical NF-κb signaling in human skeletal muscle resulting from exercise-induced inflammation and oxidative stress. J Nutr Health Food Eng. 2014;1(6):214–221. DOI: 10.15406/jnhfe.2014.01.00033

Recent evidence has shown that both pathways can be activated in skeletal muscle, with either of these pathways capable of causing skeletal muscle wasting.15,16,17 The classical pathway is IKKβ- and IKKγ-dependent, with NF-κB activation occurring due to the degradation of IκB proteins. This pathway is activated in response to inflammatory factors, bacteria, oxidative stress, leukocyte adhesion molecules, and various pro-survival genes.1,2,12,24 Activators of this pathway include TNF-α, interleukin 1 (IL-1), and lipopolysaccharide (LPS), which triggers a receptor-mediated assembly of a multi-protein signaling complex, due to interaction with receptor interacting protein (RIP), that leads to activation of IKK. As a member of the TNF-receptor associated factor (TRAF) family, TRAF1-7, is at the forefront of this pathway and acts as a E3 ubiquitin ligase. TNF-α binds to its receptor, appropriately termed TNFR, which causes TNFR-associated death domain protein (TRADD), TRAF2, and receptor-interacting protein kinase (RIP) to be recruited. NF-κB activation by TRAF2, in association with cellular inhibitor of apoptosis protein (c-IAP) induces K-63 linked poly-ubiquitination of RIP, which activates TGF-β-activated kinase 1 (TAK1). TAK1 then directly phosphorylates IKK, which results in the degradation of IκB proteins, and subsequent activation of the classical p50/p65 complex.3

NF-κB can also be activated through its alternative pathway by lymphotixin β (LT-β), B-cell activating factor (BAFF), and cluster of differentiating 40 (CD40). Conversely to the classical pathway, the alternative pathway relies on IKKα and NF-κB-inducing kinase (NIK), as opposed to IKKβ and RIP.13 NIK phosphorylates IKKα, which results in activation and phosphorylation of IκB protein p100 on sites serine 866 and 870. Unlike other IκB members, p100 undergoes only partial proteolysis, resulting in a functional p52 subunit. This functional p52 subunit is then released and transported to the nucleus in complex with RelB. Figure 6 depicts the two pathways of NF-κB.

Role of NF-κB in skeletal muscle

It has been determined that murine C2C12 skeletal muscle cell lines contain p65/p50 in their nucleus, which is the most commonly studied form of NF-κB. In the myoblast cell line C2C12, NF-κB binds on κB sites of the cyclin D1 promoter. This then allows for
transcription regulation, leading into the S phase of the cell cycle. During myogenesis, binding activity of NF-κB on cyclin D1 is decreased. This suggests that NF-κB is a critical element regulating transition to the differentiation stage from the proliferation phase. During atrophic conditions, NF-κB binds on the promoter of murine ring-finger-1 (MuRF1), which is an E3 ubiquitin ligase. This increases expression of MuRF1, suggesting NF-κB regulates the ubiquitin-proteasome system (UPS), resulting in muscle wasting.

Another potential mechanism by which increased NF-κB activity leads to skeletal muscle wasting is the possibility of NF-κB increasing the expression of inflammation-related molecules, which either directly or indirectly enhances muscle wasting. Pro-inflammatory cytokines, including TNF-α, THF-like weak inducer of apoptosis (TWEAK), IL-1, and IL-6 are major inducers of muscle wasting. Evidence has shown that NF-κB regulates expression of cytokines, chemokines, and cell-adhesion molecules. It has also been determined that NF-κB inhibition promotes regeneration of skeletal muscle by limiting the inflammatory response.

**NF-κB activity induced by oxidative stress and inflammation**

Various physiological stressors such as disease and exercise, which causes muscle injury, are known to induce oxidative stress. After injury, infiltrating leukocytes exert antiseptic protection of muscle by releasing reactive oxygen species (ROS) through oxidative burst activation of NADPH oxidase. This process results in marked perturbations of redox status in muscle fibers. Consequently, pro-inflammatory cytokines, including TNF-α, are released by neutrophils and injured muscle fibers, thereby activating ROS-generating enzymes such as xanthine oxidase. When ROS production is greater than the capacity of the antioxidant defense system, an oxidative stress occurs resulting in many cellular contents suffering oxidation due to ROS attack and subsequent activation of NF-κB, activator protein 1 (AP-1), and forkhead box (FOXO) proteins. Additionally, increased levels of ROS results in oxidation of several proteins in the excitation-contraction mechanism, and the production of protein reactive carbonyl derivatives lead to a loss of catalytic activity and increased vulnerability to protein breakdown.

It is well known that cellular stress is an activator of the NF-κB classical pathway, specifically ROS formation. Sen et al., were the first to demonstrate that TNF-α-induced activation of NF-κB in L6 myocytes, which was augmented by conditions of oxidative stress. It was also determined that inhibition of NF-κB activity by the antioxidant pyrrolydine dithiocarbamate, coupled with an increase in ICAM-1 expression, suggested that the intracellular redox status was critical in activation of NF-κB, nuclear translocation, and subsequent gene transcription regulation of proteolytic genes. Li et al., also found that TNF-α led to ROS-mediated NF-κB activation, resulting in a decrease of total protein in skeletal muscle with a specific loss of myosin heavy chain. The enhanced muscle protein degradation was coupled with TNF-α-dependent stimulation of total ubiquitin conjugation of myotube proteins.

**Figure 7** Effects of ROS production on skeletal muscle signaling pathways. The release of ROS from mitochondria, induced by humoral mediators such as NF-κB, activator protein 1 (AP-1), and forkhead box (FOXO) proteins, or metabolic alterations results in the activation of various intracellular signaling pathways such as the proteasome system and mitogen activated protein kinase (MAPK) that collaboratively work to increase both proteolysis and cytotoxicity in the skeletal muscle.
Exercise-induced activation of NF-κB

Exercise causes mechanical and metabolic stresses on skeletal muscle, including an oxidative and inflammatory response. Various cell models have shown that increased ROS accumulation activates NF-κB. This has led to the hypothesis that exercise may also activate NF-κB. This hypothesis has gained support because several studies showed that, in addition to calcineurin-nuclear T factor of activated T cells (NFAT) signaling and heat shock proteins (HSPs), NF-κB can be also be activated with exercise. Ji et al. determined that rats exercised to exhaustion had increased levels of NF-κB activation when compared to control. Ho et al. also reported an increase of NF-κB activity by 50% I-3 three hours following a 60 minute treadmill running bout. As much as a 76% decrease in IKK phosphorylation was also seen. Another mechanism suggested due to exercise is that acute exercise may cause oxidation of glutathione in skeletal muscle, which results in increased oxidative stress and NF-κB activation. Evidence from this example suggests that regular exercise may increase glutathione levels in skeletal muscle, resulting in a down-regulation of NF-κB activity. However, there have been a few studies that have shown that exercise causes a down-regulation of NF-κB activity. While the majority of evidence favors exercise-induced NF-κB activity, more research needs to be completed to determine why there are some discrepancies in the research. The relationship between exercise, ROS, and RONS (reactive oxygen nitrogen species) can be seen in Figure 8.

Figure 8 Interactions between exercise, ROS/RONS, antioxidants, and cytokines. Exercise causes mechanical and metabolic stresses on skeletal muscle, thereby inducing an oxidative and inflammatory response. As a result, ROS accumulation occurs and activates NF-κB, in addition to calcineurin-nuclear T factor of activated T cells (NFAT) signaling and heat shock proteins (HSPs).

Antioxidant nutraceutical supplementation and NF-κB

Antioxidant supplementation has been suggested to attenuate exercise-induced oxidative stress. Redox regulation of cytokine production occurs at multiple levels, including the activation of oxidant-sensitive and redox-sensitive NF-κB. Anti-oxidant supplementation has been shown to attenuate NF-κB, activated by ROS associated with exercise-induced muscle damage and RONS. Aoi et al. demonstrated in rats that a 3-week diet high in α-tocopherol (500mg/kg) attenuated NF-κB activation and p65 nuclear translocation following treadmill running for 60min at a speed of 25m/min. This study showed there is a relationship between ROS and NF-κB and that supplementation of the antioxidant α-tocopherol was able to inhibit oxidative stress and NF-κB activation. Vassilakopoulos et al. investigated the effect of a multiple antioxidant supplement (vitamins E, A, C for 60days and N-acetyl cysteine for 3days) in healthy males on plasma cytokine concentrations following bicycle exercise for 45minutes at 70% of maximum oxygen consumption. They determined, compared to pre-supplementation, that antioxidant supplementation abolished the exercise-induced increase in circulating TNF-α. Michaelidis et al. showed that the thiol-based antioxidant N-acetyl cysteine provided orally to men at a dose of 200mg/kg/day for 8days effectively attenuated circulating pro-inflammatory cytokines and skeletal muscle NF-κB phosphorylation following muscle damaging exercise (300 eccentric contractions).

Conclusion

The information presented herein indicates that various antioxidant nutraceuticals can play a beneficial role in attenuating oxidative stress and the subsequent increase in circulating TNF-α and canonical NF-κB signaling occurring in response to exercise-induced muscle damage. Since the downstream consequences of exercise-muscle damage are similar to those associated with various pathologic conditions involving muscle wasting, the implications from this review are noteworthy. Inhibition of NF-κB pathway activity has been shown to attenuate muscle wasting through various mechanisms; however, there is still conflicting evidence and more research needs to be completed to fully elucidate the role of NF-κB signaling in skeletal muscle. Antioxidant supplementation has been shown to attenuate oxidative stress and the subsequent formation of ROS/RONS production in skeletal muscle. Furthermore, this area requires further research as this process has also been demonstrated to mediate NF-κB signaling. Research on NF-κB signaling in skeletal muscle is a rapidly growing area of interest, and advances in this area may affect the types of nutraceutical supplements ingested with the intent of maintaining muscle mass during periods of heightened inflammation and oxidative stress and possible muscle wasting.

Acknowledgments

This brief review project was not partially or fully sponsored by any external sponsoring agent, rather support was provided by the Department of Health, Human Performance, and Recreation of Baylor University.

Conflict of interest

Author declares that there is no conflict of interest.

References

1. Kerksick CM, Kreider RB, Willoughby DS. Intramuscular adaptations to eccentric exercise and antioxidant supplementation. Amino Acids. 2010;39(1):219–232.
2. Guo W, Kong E, Meydani M. Dietary polyphenols, inflammation, and cancer. Nutr Cancer. 2009;61(6):807–10.
3. Peterson JM, Bakkar N, Guttridge DC. NF-κB Signaling in Skeletal Muscle Health and Disease. Curr Top Dev Biol. 2011;96:85–119.
4. Evans WJ. Vitamin E, vitamin C, and exercise. Am J Clin Nutr. 2000;72(Suppl 2):647S–652S.
5. Li H, Malhotra S, Kumar A. Nuclear factor-kappa B signaling in skeletal muscle atrophy. J Mol Med (Berl). 2008;86(10):1113–1126.
Effectiveness of antioxidant nutraceuticals in attenuating canonical NF-κB signaling in human skeletal muscle resulting from exercise-induced inflammation and oxidative stress.

6. Li YP, Schwartz RJ, Waddell ID, et al. Skeletal muscle myocytes undergo protein loss and reactive oxygen-mediated NF-kappaB activation in response to tumor necrosis factor alpha. *FASEB J.* 1998;12(10):871–880.

7. Bhatnagar S, Panguluri SK, Gupta SK, et al. Tumor necrosis factor-alpha regulates distinct molecular pathways and gene networks in cultured skeletal muscle cells. *PLoS One.* 2010;5(10):e13262.

8. Dogra C, Changotta H, Wedhas N, et al. TNF-related weak inducer of apoptosis (TWEAK) is a potent skeletal muscle-wasting cytokine. *FASEB J.* 2007;21(8):1857–1869.

9. Reid MB, Li YP. Tumor necrosis factor-alpha and muscle wasting: a cellular perspective. *Respir Res.* 2001;2(5):269–272.

10. Jackman RW, Kandarian SC. The molecular basis of skeletal muscle atrophy. *Am J Physiol Cell Physiol.* 2004;287(4):C834–C843.

11. Ventadour S, Attiaux D. Mechanisms of skeletal muscle atrophy. *Curr Opin Rheumatol.* 2006;18(6):631–635.

12. Eley HL, Tisdale MJ. Skeletal muscle atrophy, a link between depression of protein synthesis and increase in degradation. *J Biol Chem.* 2007;282(10):7087–7097.

13. Kramer HF, Goodyear LJ. Exercise, MAPK, and NF-kappaB signaling in skeletal muscle. *J Appl Physiol.* 2007;103(1):388–395.

14. Acharya Y, Villalta SA, Bakkar N, et al. Interplay of IKK/NF-kappaB signaling in macrophages and myofibers promotes muscle degeneration in Duchenne muscular dystrophy. *J Clin Invest.* 2007;117(4):889–901.

15. Cai D, Frantz JD, Tawa NE, et al. IKKbeta/NF-kappaB activation causes severe muscle wasting in mice. *Cell.* 2004;119(2):285–298.

16. Hunter RB, Kandarian SC. Disruption of either the NFkb1 or the Rel3 gene inhibits skeletal muscle atrophy. *J Clin Invest.* 2004;114(10):1504–1511.

17. Mourikoti F, Kratsios P, Luede T, et al. Targeted ablation of IKK2 improves skeletal muscle strength, maintains mass, and promotes regeneration. *J Clin Invest.* 2006;116(11):2945–2954.

18. Sakkas GK, Schambelan M, Mulligan K. Can the use of creatine supplementation attenuate muscle loss in cachexia and wasting? *Curr Opin Clin Nutr Metab Care.* 2009;12(6):623–627.

19. Bonetto A, Penna F, Muscaritoli M, et al. Are antioxidants useful for treating skeletal muscle atrophy? *Free Radic Biol Med.* 2009;47(7):906–916.

20. Peake JM, Suzuki K, Coombes JS. The influence of antioxidant supplementation on markers of inflammation and the relationship to oxidative stress after exercise. *J Nutr Biochem.* 2007;18(6):357–371.

21. Ringeis R, Keller J, Eder K. Mechanisms underlying the anti-wasting effect of L-carnitine supplementation under pathologic conditions: evidence from experimental and clinical studies. *Eur J Nutr.* 2013;52(5):1421–1442.

22. Bakkar N, Guttridge DC. NF-kappaB signaling: a tale of two pathways in skeletal myogenesis. *Physiol Rev.* 2010;90(2):495–511.

23. Pahl HL. Activators and target genes of Rel/NF-kappaB transcription factors. *Oncogene.* 1999;18(49):6853–6866.

24. Solt LA, May MJ. The IkappaB kinase complex: master regulator of NF-kappaB signaling. *Immunol Rev.* 2008;221. DOI: 10.15406/jnhfe.2014.01.00033

25. Rudolph D, Yeh WC, Wakeham A, et al. Severe liver degeneration and lack of NF-kappaB activation in NEMO/IKKgamma-deficient mice. *Genes Dev.* 2000;14(7):854–862.

26. Yamaoka S, Courtois G, Bessia C, et al. Complementation cloning of NEMO, a component of the IkappaB kinase complex essential for NF-kappaB activation. *Cell.* 1998;93(7):1231–1240.

27. Delhase M, Hayakawa M, Chen Y, et al. Positive and negative regulation of IkappaB kinase activity through IKKbeta subunit phosphorylation. *Science.* 1999;284(5412):309–313.

28. May MJ, Marienfeld RB, Ghosh S. Characterization of the Ikappa B-kinase NEMO binding domain. *J Biol Chem.* 2002;277(48):45992–46000.

29. Schomer-Miller B, Higashimoto T, Lee YK, et al. Regulation of IkappaB kinase (IKK) complex by IKKgamma-dependent phosphorylation of the T-loop and C terminus of IKKbeta. *J Biol Chem.* 2006;281(22):15268–15276.

30. Hunter RB, Stevenson E, Koncarevic A, et al. Activation of an alternative NF-kappaB pathway in skeletal muscle during disuse atrophy. *FASEB J.* 2002;16(6):529–538.

31. Kumar A, Takada Y, Boriek AM, et al. Nuclear factor-kappaB: its role in health and disease. *J Mol Med (Berl).* 2004;82(7):434–448.

32. Hayden MS, West AP, Ghosh S. Snap Shot: NF-kappaB signaling pathways. *Cell.* 2006;127(6):1286–1287.

33. Nikolaidis M, Jamurtas A, Paschalis V, et al. The effect of muscle-damaging exercise on blood and skeletal muscle oxidative stress: magnitude and time-course considerations. *Sports Med.* 2008;38(7):579–606.

34. Ji LL, Gomez-Cabrera MC, Vina J. Role of nuclear factor kappaB and mitogen-activated protein kinase signaling in exercise-induced antioxidant enzyme adaptation. *Appl Physiol Nutr Metab.* 2007;32(5):930–935.

35. Smolka MB, Zoppi CC, Alves AA, et al. HSP72 as a complementary protection against oxidative stress induced by exercise in the soleus muscle of rats. *Am J Physiol Regul Integr Comp Physiol.* 2000;279(5):R1539–R1545.

36. Brotto MA, Nosek TM. Hydrogen peroxide disrupts Ca2+ release from the sarcoplasmic reticulum of rat skeletal muscle fibers. *J Appl Physiol.* (1955). 1996;81(2):731–737.

37. Radak Z, Kaneko T, Tahara S, et al. The effect of exercise training on oxidative damage of lipids, proteins, and DNA in rat skeletal muscle: evidence for beneficial outcomes. *Free Radic Biol Med.* 1999;27(1–2):69–74.

38. Sen CK, Khanna S, Reznick AZ, et al. Glutathione regulation of tumor necrosis factor-alpha-induced NF-kappaB activation. *FASEB J.* 2006;20(8):2356–2365.

39. Bar-Shai M, Carmeli E, Ljubicic P, et al. Exercise and immobilization in aging animals: the involvement of oxidative stress and NF-kappaB activation. *Free Radic Biol Med.* 2008;44(2):202–214.

40. Kamata H, Manabe T, Oka Si, et al. Hydrogen peroxide activates IkappaB kinases through phosphorylation of serine residues in the activation loops. *FEBS Lett.* 2002;519(1–3):231–237.

41. Muller JM, Rupe RA, Baueerle PA. Study of gene regulation by NF-kappa B and AP-1 in response to reactive oxygen intermediates. *Methods.* 1997;11(3):301–312.

42. Ho RC, Hirshman MF, Li Y, et al. Regulation of IkappaB kinase and NF-kappaB in contracting rat skeletal muscle. *Am J Physiol Cell Physiol.* 2005;289(4):C794–C801.

43. Hollander J, Fiebig R, Gore M, et al. Superoxide dismutase gene expression is activated by a single bout of exercise in rat skeletal muscle. *Pflugers Arch.* 2001;442(3):426–434.

Citation: McKinley S, Willoughby DS. Effectiveness of antioxidant nutraceuticals in attenuating canonical NF-κB signaling in human skeletal muscle resulting from exercise-induced inflammation and oxidative stress. *J Nutr Health Food Eng.* 2014;1(6):214–221. DOI: 10.15406/jnhfe.2014.01.00033
46. Ji LL, Gomez-Cabrera MC, Steinhafel N, et al. Acute exercise activates nuclear factor (NF)-κB signaling pathway in rat skeletal muscle. *FASEB J.* 2004;18(13):1499–1506.

47. Ji LL, Gomez-Cabrera MC, Vina J. Exercise and hormesis: activation of cellular antioxidant signaling pathway. *Ann N Y Acad Sci.* 2006;1067:425–435.

48. Spangenberg EE, Brown DA, Johnson MS, et al. Exercise increases SOCS-3 expression in rat skeletal muscle: potential relationship to IL-6 expression. *J Physiol.* 2006;572(Pt 3):839–848.

49. Sen CK. Glutathione homeostasis in response to exercise training and nutritional supplements. *Mol Cell Biochem.* 1999;196(1–2):31–42.

50. Durham WJ, Li YP, Gerken E, et al. Fatiguing exercise reduces DNA binding activity of NF-κB in skeletal muscle nuclei. *J Appl Physiol.* 1985;97(5):1740–1745.

51. Durham WJ, Arbogast S, Gerken E, et al. Progressive nuclear factor-κB activation resistant to inhibition by contraction and curcumin in mdx mice. *Muscle Nerve.* 2006;34(3):298–303.

52. Haddad JJ. Pharmaco-redox regulation of cytokine-related pathways: From receptor signaling to pharmacogenomics. *Free Radic Biol Med.* 2002;33(7):907–926.

53. Aoi W, Naito Y, Takanami Y, et al. Oxidative stress and delayed-onset muscle damage after exercise. *Free Radic Biol Med.* 2004;37(4):480–487.

54. Kosmidou I, Vassilakopoulos T, Xagorari A, et al. Production of interleukin-6 by skeletal myotubes: Role of reactive oxygen species. *Am J Respir Cell Mol Biol.* 2002;26(5):587–593.

55. Vassilakopoulos T, Karatza MH, Katsaounou P, et al. Antioxidants attenuate the plasma cytokine response to exercise in humans. *J Appl Physiol (1985).* 2003;94(3):1025–1032.

56. Michailidis Y, Karagounis LG, Terzis G, et al. Thio-based antioxidant supplementation alters human skeletal muscle signaling and attenuates its inflammatory response and recovery after intense exercise. *Am J Clin Nutr.* 2013;98(1):233–245.