TEMPERATURE TOLERANCE AND OTHER PROPERTIES OF TWO ETHANOL PRODUCING SACCHAROMYCES CEREVISIAE STRAINS ISOLATED FROM COCONUT TODDY

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Abstract: Several yeast strains capable of fermenting glucose at 40°C were isolated from coconut toddy. Two strains, which gave high ethanol concentration at 40°C were identified as strains of Saccharomyces cerevisiae. Strain T-31 (NCYC 2401) at 40°C gave high alcohol concentration in both YPS and molasses media, 7.4% and 5.4% v/v respectively, while strain SK-33 (NCYC 2402) gave higher concentration of alcohol in synthetic medium, 8.4% v/v. The maximum growth temperature of both these strains was 45°C indicating their thermotolerant behaviour. Of these two strains, T-31 was found to be suitable for alcohol fermentation at high temperature (40°C) using molasses, based on parameters such as maximum CO₂ productivity (0.216 g h⁻¹), final ethanol concentration (7.4% v/v), yield efficiency (0.579) and minimal cell growth rate (0.029 cells ml⁻¹ h⁻¹).

Key Words: Coconut sap, coconut toddy, ethanol, Saccharomyces cerevisiae, temperature tolerance, yeast.

INTRODUCTION

The limited availability of world's crude oil and its constant price increase aroused renewed interest in the gasohol concept. Fuel ethanol has been produced in Brazil since 1933, while both Germany and Japan used it as a source of fuel during the Second World War. Production of fuel ethanol using industrial by-products such as molasses would be a promising endeavour for developing countries, which do not have any fossil fuels.

Molasses, which is a by-product of cane sugar industry, is mainly utilised in the production of ethanol in the distilleries of Sri Lanka. With the setting up of new sugar factories in the country, there is a considerable increase in the availability of sugarcane molasses. Nevertheless, these distilleries operate at low efficiencies (50 - 60%) compared to other distilleries in the world.¹ This is due to inefficiency in the method of distillation and unsuitability of the yeast strains for fermenting molasses under tropical environmental conditions. The compound effect of high ambient temperature in tropical countries and exothermic fermentation reaction elevate the temperature of the fermentation mash. High temperature usually inhibits the fermentation ability of the yeast strain used, as most of the conventional yeasts are temperature sensitive. To minimise this inhibition, a cooling system is used to maintain the temperature around 35°C or
below. However this process increases the capital and running costs. In view of this situation, tropical fermentation technology as regards ethanol fermentation by yeast, requires strains capable of fermenting sugars efficiently at high temperature of around 40°C. There are certain advantages in high temperature fermentation by yeast. They are, faster rates of substrate consumption and ethanol formation, facilitation of easy ethanol recovery and savings on cooling cost. As the fermentation is completed within 36 - 48 h, loss of ethanol due to evaporation is considerably less even at temperatures around 40°C. Therefore, use of high temperature tolerant yeast, in fermentation industry under tropical environmental conditions is cost effective. There are several reports on the isolation of thermotolerant strains belonging to different genera. The isolation of five thermotolerant strains of Kluyveromyces strains capable of growth and high-level ethanol production, on glucose and molasses fermentation, at temperatures in the range of 45-50°C had been reported. Kluyveromyces marxianus IMB 3 had produced 35 g/l of alcohol at 13 h following growth at 45°C on sucrose containing medium. Anderson et al. reported an efficient carbohydrate fermenting Kluyveromyces marxianus var. marxianus producing 6% w/v ethanol at 43°C in 24 h. Two Saccharomyces and one Candida strains reported by Hacking et al. were found to meet minimum commercial targets set at 8% (v/v) ethanol from 14% w/v glucose at 40°C. This paper describes some features of two thermotolerant ethanol producing Saccharomyces cerevisiae strains isolated from coconut sap.

**METHODS AND MATERIALS**

*Isolation of high-temperature yeast:* Enrichment cultures were prepared using 5 ml samples of toddy in 45 ml of 10% glucose and incubated at 40°C for 24 h. Subsequently the enriched cultures were used to prepare a series of dilution followed by plating 0.1 ml of each dilution on Yeast extract - Peptone - Dextrose - agar (YPD) medium composed of 2% dextrose, 2% peptone, 1% yeast extract, 2% agar and complete synthetic medium which has a composition similar to the basal medium described. To both these media 50 ppm of Streptomycin was added to inhibit bacterial growth. The plates were incubated at 40°C for 2-3 days. Morphologically different colonies that appeared on plates were picked up and purified by streaking on fresh YPD plates.

*Selection of yeast strains capable of fermenting at 40°C:* Fermentation tests were carried out with each of the strains isolated using broth cultures. First a seed culture was prepared using YPD broth at 30°C and the cell concentration of each of these cultures was monitored using a haemocytometer. For the fermentation test, 50ml amounts of YPD medium containing 15% (w/v) glucose were dispensed in 250-ml capacity Erlenmeyer flasks fitted with fermentation bungs. The fermentation bungs were filled with concentrated sulphuric acid, which permitted only carbon dioxide to evolve from the culture. Water vapour and vaporized
ethanol were trapped in the sulphuric acid. The flasks were inoculated with a known amount of cells (1x10^8 cells ml^{-1}), from the seed culture of each of the isolates and were weighed after inoculation and then incubated statically at 40°C for 72 h. The fermentation was monitored by measuring the daily carbon dioxide output as reflected by decrease in the weight of the whole culture. Twice the weight of carbon dioxide evolved is equivalent to the amount of sugar consumed by each yeast strain used in fermentation. Amount of sugar used up by each of the strains in fermentation was calculated and those strains that used up 50% or more of the initial sugar content were selected for further studies.

Fermentation tests were again done using Yeast extract - Peptone - Sucrose (YPS) medium and synthetic medium containing 20% (w/v) sucrose, at 40°C, to check the fermentation ability of the selected yeast strains with sucrose as the carbon source. For comparison, baker's yeast was also used in these experiments. The amount of carbon dioxide evolved (g) and the final ethanol concentration (% v/v) of the fermenting mash were measured. The final ethanol concentration was measured using an ebulliometer.

Methods of identification: For taxonomical identification of yeast, the methods described were followed.

Growth at different temperatures: The selected strains were tested for their ability to grow at different temperatures such as 40, 45, 47 and 50°C in YPD agar medium. In complete synthetic liquid medium, the growth rates of the selected strains at 40°C were compared with Baker's yeast by measuring the optical density at 660 nm, at intervals and then converted to cell density using a calibration curve.

Fermentation characteristics of selected yeast at different temperatures: Fermentation tests were carried out with the selected strains and Baker's yeast at different temperatures, 25, 30 and 40°C. As 40°C was found to be the optimum growth temperature of the yeast, it was selected to be the maximum temperature limit for fermentation. In both YPS and complete synthetic medium 20% (w/v) sucrose was used. The carbon dioxide evolution rate and final ethanol concentration were determined as described earlier.

Fermentation using molasses medium: Molasses was diluted three folds to give approximately 20% fermentable sugars and potassium phosphate (0.05%) and magnesium sulphate (0.5%) were added. Before autoclaving, the pH was adjusted to 4.8. Fermentation was carried out at 30 and 40°C. The amount of carbon dioxide evolved and the ethanol concentrations (% v/v) were determined as described earlier.
Selecting the best yeast strain for industrial alcohol production: For this purpose, the parameters, maximum productivity (based on carbon dioxide evolution), final alcohol concentration, ethanol yield ratio and maximum growth rate were used. Ethanol yield ratio was computed by dividing the actual ethanol yield by the theoretical ethanol yield on the basis of Gay-Lussac equation (i.e. 0.511 g ethanol g⁻¹ glucose, fructose or sucrose). The maximum growth rate was calculated from the exponential phase of the growth curve. Optical density was measured using a spectrophotometer at 660 nm and then converted to cell density using a calibration curve. Higher values for final ethanol concentration, carbon dioxide evolution rate and ethanol yield ratio have a positive influence on alcohol productivity while higher values for growth rate have a negative influence on alcohol productivity.⁹

RESULTS

Colonies, which had different morphology on dilution plates, were picked up and purified by streaking on fresh YPD plates. Fifteen stock cultures were prepared using YPD medium. Code numbers identified these.

In the fermentation experiments the loss in weight of the whole culture reflected the carbon dioxide productivity and twice that value indicated the amount of sugar consumed by the cell to produce alcohol according to Gay Lussac equation. Six strains capable of fermenting more than 50% of initial sucrose at 40°C were selected from the stock culture by preliminary fermentation tests. Out of the six strains, only two strains (SK-33 and T-31) gave high alcohol yield at 40°C with sucrose as the carbon source (Table 1).

Table 1: Fermentation screen of different strains of yeast at 40°C in YPS medium with 15% (w/v) sucrose.

| Strain No. | Carbon dioxide output g/100 ml | Sucrose consumed (g) | Alcohol % v/v |
|------------|-------------------------------|---------------------|---------------|
| SK- 41     | 3.5                           | 7.0                 | 4.4           |
| SK- 33     | 5.5                           | 11.0                | 6.9           |
| KT- 45     | 3.9                           | 7.8                 | 4.9           |
| T-31       | 5.3                           | 10.6                | 6.6           |
| KT- 34     | 4.0                           | 8.0                 | 5.0           |
| TO- 44     | 3.5                           | 7.0                 | 4.5           |
| Baker's yeast | 5.2                           | 10.4                | 6.2           |
The two strains, T-31 and SK-33 thus selected agreed well with the standard description for *Saccharomyces cerevisiae* (Tables 2 and 3). Full DNA fingerprinting analysis of the above two cultures showed that they are significantly different from other brewing strains held in the National Collection of Yeast Cultures (NCYC) at Norwich, UK. Therefore, the above cultures, T-31 & SK-33 were accessioned into NCYC after assigning the numbers NCYC 2401 and 2402 respectively.

Table 2: Morphological characters of yeast strains T-31 and SK-33.

| Morphology on YM medium | Strain T-31 | Strain SK-33 |
|-------------------------|------------|--------------|
| Broth                   |            |              |
| Cells after 48 h         | Oval (3-7)x (4-8) μ | Oval (3-7)x(4x8) μ |
| Culture after 21 d       | Cream non-flocculant deposit | Cream non-flocculant deposit |
| Agar                    |            |              |
| Cells after 48 h         | Oval (5-8)x(5-9) μ | Round to long oval (4-6)x(5-8) μ |
| Culture after 21 d       | Cream white, smooth slightly shiny | Cream white slightly shiny |
| Pseudo/true mycelium     |            |              |
| CMA Aerobic             | No mycelium | Well developed |
| Anaerobic               | No mycelium | No mycelium |
| PDA Aerobic             | No mycelium | Poorly developed |
| Anaerobic               | No mycelium | No mycelium |
| Balistospores           | Negative   | Negative     |
| Arthrospores            | Negative   | Negative     |
| Endospores              | Negative   | Negative     |
| Chlamydospores          | Negative   | Negative     |
| Sexual spores           |            |              |
| CMA                     | Negative   | Negative     |
| KAC & Gorodkowa         | 4 ascospores/ascus Round-oval | 3-4 ascospores/ascus Round |

YM - Malt Extract, Yeast extract; PDA - Potato Dextrose agar; CMA - Corn meal agar; KAC - potassium acetate agar.
Table 3: Physiological characters of the two yeast strains T-31 and SK-33.

|          | Ferment. | Assimi. | Ferment. | Assimi. |
|----------|----------|---------|----------|---------|
| D-Glucose| +        | +       | +        | +       |
| D-Galactose| +      | +       | +        | +       |
| Sucrose  | +        | +       | +        | +       |
| Maltose  | -        | -       | -        | -       |
| Cellobiose| -       | -       | -        | -       |
| Trehalose| -        | +(s)    | -        | -(s)    |
| Lactose  | -        | -       | -        | -       |
| Melibiose| -        | -       | -        | -       |
| Raffinose| +        | +       | +        | +       |
| Melzitose| -        | -       | -        | -       |
| Methyl-D  | +        | +       | +        | +       |
| glucopyranoside | | | | |
| Inulin   | -        | -       | -        | -       |
| Sol. Starch| -      | -       | -        | -       |
| Xylose   | -        | -       | -        | -       |
| L-Arabinose| -     | -       | -        | -       |
| D-Arabinose| -      | -       | -        | -       |
| D-Ribose | -        | -       | -        | -       |
| L-Rhamnose| -      | -       | -        | -       |
| Ethanol  | -        | +       | -        | +       |
Table 3 contd.

|                      | T-31 Assimil. | SK-33 Assimil. |
|----------------------|--------------|---------------|
| Glycerol             | -            | -             |
| Erythritol           | -            | -             |
| Ribitol              | -            | -             |
| Galcitol             | -            | -             |
| D-Mannitol           | -            | -             |
| Glucitol             | -            | -             |
| Salicilin            | -            | -             |
| Lactic acid          | +(w)         | +(w)          |
| Succinic acid        | -            | -             |
| Citric acid          | -            | -             |
| Myo-inositol         | -            | -             |
| D-Glucono 1,5 lactone| -            | -             |
| D-Glucosamine        | -            | -             |
| Methanol             | -            | -             |
| Xylitol              | -            | -             |
| Ammonium sulphate    | +            | +             |
| Potassium nitrate    | -            | -             |
| Ethyl amine          | -            | -             |
| Cadaverine           | -            | -             |
| L-Lysine             | -            | -             |
Table 3 contd.

| Vitamin free growth | T-31 | SK-33 |
|---------------------|------|-------|
| Cycloheximide       |      |       |
| (0.01% and 0.1%)    | -    | -     |
| 1% Acetic acid      |      |       |
| Acid production     | w    | w     |
| Growth at 40°C      |      |       |
| Arbutin hydrolysis  |      |       |
| Urease activity     |      |       |
| Salt tolerant 10%   | +    |       |

s = slow    w = weak

Growth at different temperatures: The maximum growth temperature of these two yeasts was 45°C. Both grew very well at 40°C (Table 4). No growth was observed at 47°C and 50°C. The growth rate of SK-33 in synthetic medium was higher while T-31 and Baker's yeast had a lower growth rate. The yeasts appear thermotolerant.

Table 4: Growth characteristics of selected yeast strains at different temperatures in YPD medium.

| Strain         | 40°C | 45°C | 47°C | 50°C |
|----------------|------|------|------|------|
| T-31           | +++  | ++   |      |      |
| SK-33          | +++  | ++   |      |      |
| Baker's yeast  | +++  | +    |      |      |

+++ very good, ++ good, + weak, - No growth
Alcohol fermentation at different temperatures: In fermentation tests at 30°C in YPS medium, SK-33 and T-31 gave more than 12% v/v alcohol, whereas Baker's yeast gave less alcohol 11.7% v/v. At 40°C in the same medium, strain T-31 performed better, giving an alcohol concentration around 7.4% v/v (Fig. 1). With increase of temperature from 30 to 40°C, the alcohol concentration obtained with this strain decreased by about 41%. In the synthetic medium with 20% sucrose at 30°C all strains gave higher alcohol concentration, more than 10% v/v and SK-33 gave highest concentration, 13.2% v/v. At 40°C also, the concentrations were higher in this medium with strain SK-33 giving the highest value (8.4% v/v, Fig. 2). In the synthetic medium with increase of temperature the % decrease in alcohol concentration was about 36%. The 'balance sheet' (Table 5) with respect to fate of sugar and production of ethanol in the two media at 40°C shows that the difference between the observed and calculated values for ethanol is very small. This confirmed that there is no great loss of ethanol due to evaporation at high temperature.

Table 5: 'Balance Sheet' regarding fate of sugar and production of ethanol in YPS and synthetic media at 40°C.

| Strain      | Amount of sugar used up (g) | % Alcohol (Theoretical) (v/v) | % Alcohol (Actual)(v/v) |
|-------------|-----------------------------|------------------------------|------------------------|
| Medium - YPS|                             |                              |                        |
| T -31       | 12.04                       | 7.68                         | 7.4                    |
| SK -33      | 10.61                       | 6.76                         | 6.7                    |
| Baker's yeast| 11.02                       | 7.03                         | 6.8                    |
| Medium - Synthetic|          |                              |                        |
| T - 31      | 13.07                       | 8.33                         | 8.2                    |
| SK - 33     | 13.27                       | 8.46                         | 8.4                    |
| Baker's yeast| 12.65                       | 8.06                         | 8.0                    |

YP's - Yeast extract, Peptone, Sucrose medium.
Figure 2: Time course of ethanol production in synthetic medium with 20% sucrose at 30 and 40°C.
Alcohol fermentation using molasses: In the molasses medium at 30 and at 40 °C, T-31 gave highest alcohol concentration of 11.9% v/v and 5.4% v/v respectively (Fig. 3). Baker's yeast in the same medium produced lower alcohol concentrations, 11.5%, 4.7% at 30 and 40°C respectively. The percentage decrease in the alcohol yield with increase of temperature, was less in the case of T-31 (54%) than in Baker's yeast (59%).

Figure 3: Time course of ethanol production in molasses medium at 30 and 40°C.
The values obtained for different parameters that were used in the selection of a suitable strain for industrial alcohol production are shown in Table 6. In strain T-31, all parameters that have a positive influence on alcohol production (i.e. CO₂ productivity, final ethanol concentration and ethanol yield ratio) gave higher values. Further, the same strain gave a lower value for maximum growth rate (0.029), which has a negative influence on alcohol production.

Table 6: Comparison of criteria used in evaluation of fermentation performance of selected yeast strains with Baker's yeast at 40°C in YPS medium.

| Strain     | Maximum carbon dioxide productivity (gh⁻¹) | Final ethanol conc. (v/v) | Ethanol yield efficiency | Growth rate (cells ml⁻¹ h⁻¹) |
|------------|-------------------------------------------|---------------------------|--------------------------|----------------------------|
| Baker’s yeast | 0.187                                      | 6.8                        | 0.532                    | 0.032                      |
| SK - 33    | 0.179                                      | 6.7                        | 0.524                    | 0.110                      |
| T - 31     | 0.216                                      | 7.4                        | 0.579                    | 0.029                      |

Yield efficiency = Actual yield/Theoretical yield = Actual yield/0.511.

**DISCUSSION**

Although several reports are available on the flora of fermenting coconut palm wine, no studies have been done on thermotolerant yeast strains. Temperature is a paramount important regulatory factor in alcohol fermentation, particularly in tropical countries. There are a number of studies on the thermotolerance of growth and fermentation of different yeast strains. However this is the first instance that apparently thermotolerant yeasts have been isolated from coconut toddy. The strain (T-31) *Saccharomyces cerevisiae* (NCYC 2401), isolated from coconut toddy gave 7.4% v/v alcohol within 24 to 36 h, in YPS medium, but the initial sugar concentration was higher 20% w/v. Two *Saccharomyces* and one *Candida* have been found to meet minimum commercial targets set at 8% (v/v) ethanol from 14% w/v glucose at 40°C.[4] There have been several studies on the thermotolerance of growth and fermentation of *S. cerevisiae* and *S. uvarum* strains and some results are contradictory. In general, the conclusion is that higher the fermentation temperature greater the inhibitory effect of ethanol on cells.[10,11] Some have reported increase in ethanol tolerance with increase of temperature.[12] It has also been reported that growth and fermentation can be lost in the range of 35-45°C.[13] In this study, at 40°C *Saccharomyces cerevisiae* (NCYC 2401) gave 7.4% and 5.4% v/v alcohol in YPS and molasses media respectively. When alcohol yields at high temperatures are compared with those
at lower temperatures, both these yeasts were found to give higher alcohol yields at low temperatures in all media tested. This decrease in alcohol yield at higher temperature could be due to the damaging effect of temperature on yeast cells. However, the results indicate that the yeast strains isolated from coconut toddy have the potential to produce high alcohol yields at fairly high temperatures and would be suitable to be used in molasses fermentation even without a sophisticated cooling system.

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