Constraints on barium isotope fractionation during 
aragonite precipitation by corals

CHLOÉ PRETET*, KIRSTEN VAN ZUILEN†*, THOMAS F. NÄGLER†, STÉPHANIE REYNAUD‡, 
MICHAEL E. BÖTTLER§ and ELIAS SAMANKASSOU*

*Department of Earth Sciences, University of Geneva, Rue des Marachets 13, CH-1205, Geneva, Switzerland (E-mails: chloepretet@no-log.org; Elias.Samankassou@unige.ch)
†Institute of Geological Sciences, Bälterstrasse 1+3, CH-3012, Bern, Switzerland
‡Centre Scientifique de Monaco, 8 quai Antoine 1er, 98000, Monaco
§Geochemistry and Stable Isotope Biogeochemistry Group, Marine Geology Department, Leibniz Institute for Baltic Sea Research (IOW), Seestr.15, D-18119, Warnemünde, Germany

INTRODUCTION

The earth alkaline element barium (Ba) has attracted significant attention in the biogeochemistry community. In the photic zone of marine surface waters, Ba is incorporated in or adsorbed on organic matter (Sternberg et al., 2005) and planktonic carbonate tests, causing depletion of dissolved Ba (Cardinal et al., 2005) and resulting in a nutrient-like distribution in the water column (Wolgemuth & Broecker, 1970; Chan et al., 1976; Church, 1979; Monnin et al., 1999; Chester, 2009). Therefore, variation in the accumulation rates of Ba in marine sediments is thought to be indicative of variations in marine biological productivity through time (Schmitz, 1987; François et al., 1995; Paytan et al., 1996; McManus et al., 1999; Martínez-Ruiz et al., 2000; Babu et al., 2002; Paytan & Griffith, 2007; Hull & Norris, 2011). Furthermore, Ba/Ca ratios in the shells of foraminifers are used to reconstruct past ocean alkalinity and circulation patterns (Lea et al., 1989; Hall & Chan, 2004; Weldeab et al., 2007a) or act as a

ABSTRACT

We present a barium (Ba) isotope fractionation study of marine biogenic carbonates (aragonitic corals). The major aim is to provide first constraints on the Ba isotope fractionation between modern surface seawater and coral skeleton. Mediterranean surface seawater was found to be enriched in the heavy Ba isotopes compared to previously reported values for marine open ocean authigenic and terrestrial minerals. In aquarium experiments with a continuous supply of Mediterranean surface water, the Ba isotopic composition of the bulk sample originating from cultured, aragonitic scleractinian corals ($\delta^{137/134}$Ba between +0.16 ± 0.12‰ and +0.41 ± 0.12‰) were isotopically identical or lighter than that of the ambient Mediterranean surface seawater ($\delta^{137/134}$Ba = +0.42 ± 0.07‰, 2SD), which corresponds to an empirical maximum value of Ba isotope fractionation of $\Delta^{137/134}$Ba_{coral-seawater} = −0.26 ± 0.14‰ at 25°C. This maximum Ba isotope fractionation is close and identical in direction to previous results from inorganic precipitation experiments with aragonite-structured pure BaCO$_3$ (witherite). The variability in measured Ba concentrations of the cultured corals is at odds with a uniform distribution coefficient, $D_{\text{Ba(Ca)}}$, thus indicating stronger vital effects on isotope than element discrimination. This observation supports the hypothesis that the Ba isotopic compositions of these corals do not result from simple equilibrium between the skeleton and the bulk seawater. Complementary coral samples from natural settings (tropical shallow-water corals from the Bahamas and Florida and cold-water corals from the Norwegian continental shelf) show an even wider range in $\delta^{137/134}$Ba values (+0.14 ± 0.08 to +0.77 ± 0.11‰), most probably due to additional spatial and/ or temporal seawater heterogeneity, as indicated by recent publications.
continental run-off proxy in coastal waters (Weldeab et al., 2007b; Sprovieri et al., 2008).

The marine Ba cycle, however, is not fully understood (Dymond et al., 1992; Paytan & Kastner, 1996; Dickens et al., 2003). Different Ba sources to the ocean and their relative contributions in the past are still difficult to quantify. If different Ba sources could be distinguished based on their respective isotopic composition, the relative contribution of these sources to the marine Ba pool could be approached by mass balance models. Further, transport, dissolution-reprecipitation or vital processes may induce resolvable Ba isotope fractionation in sediments and biogenic carbonate. Thus, the analysis of the Ba isotopic composition of different Ba sources in seawater and marine sediments bears the potential to improve the understanding of the Ba cycle.

Eugster et al. (1969) measured various meteorites, reagents Ba and one terrestrial standard and concluded that there is no anomaly present in Ba isotope abundances in excess of 1 ‰. Subsequently, only a very limited number of studies have dealt with Ba isotopes in natural terrestrial samples (von Allmen et al., 2010; Miyazaki et al., 2014; Horner et al., 2015; Nan et al., 2015; Cao et al., 2016), or experimental carbonate and sulphate precipitates (von Allmen et al., 2010; Böttcher et al., 2012). Von Allmen et al. (2010) found that synthetic wetherite (BaCO₃) and barite (BaSO₄) are generally depleted in the heavy Ba isotopes down to −0.3 ‰ in δ¹³⁷/¹³⁴Baolid-liquid. The calculated total amounts of Ba precipitated in those experiments were below 5%, that is, the results were not significantly influenced by Rayleigh effects. Terrestrial Ba gangue minerals (four barites, one norsethite [BaMg(CO₃)₂]) were all close to standard solutions. In contrast, a δ¹³⁷/¹³⁴Ba value of −0.5 ‰ was found in a diagenetic barite sample from ODP Leg 207 that formed under reducing diagenetic conditions. It is only recently that modern seawater data have been published for Ba. Open ocean water profiles in the South Atlantic (Horner et al., 2015) and the East China Sea (Cao et al., 2016) showed consistently a general decrease in δ³⁵⁷/³⁴Ba of about 0.2 to 0.3 ‰ with water depth. The actual depth in which the transition to lower δ³⁵⁷/³⁴Ba values starts and the Ba isotopic composition of (sub)surface waters, however, are variable. The latter varies between 0.45 ‰ in δ³⁵⁷/³⁴Ba in the South Atlantic (Horner et al., 2015) and 0.8 ‰ in the East China Sea (Cao et al., 2016), recalculated relative to the Fluka Ba(NO₃)₂ standard used here (see ‘MC-ICP-MS analysis’ below).

In this study, the Ba isotope composition of cultured coral skeletons is investigated and compared to the composition of the ambient seawater. Corals were chosen because they are abundant in the photic zone where Ba is depleted. Given the observed fractionated Ba isotope values in (sub)surface waters (Horner et al., 2015; Cao et al., 2016) and Ba depletion related to microorganism activity, corals may record changes in productivity and seawater heterogeneities with respect to their Ba isotopic compositions. In order to use coral skeletons as archives, however, the fractionation factor between carbonate and ambient seawater must be known. Next to isotope effects due to carbonate precipitation (von Allmen et al., 2010), vital effects accompanying coral growth may contribute to the overall Ba isotope fractionation in an analogous manner to that observed in the Ca-isotope system. For example, Inoue et al. (2015) found significant difference in ∆⁴⁰⁴⁴Ca(solid-solution) in cultured Porites australiensis compared to published data from inorganic aragonite precipitates, what these authors attributed to coral-specific bio-mineralization processes. In contrast, vital effects on stable Sr isotope ratios are ambiguous. Constant ∆⁸⁸/⁸⁶Sr values of −0.2‰ were reported for various samples of Lophelia pertusa derived from different locations along the European continental margins (Raddatz et al., 2013). Fietzke & Eisenhauer (2006), on the other hand, reported temperature-dependent strontium isotope fractionation during calcium carbonate precipitation. Natural coral samples from Pavona clavus yielded a steeper temperature gradient than inorganically precipitated aragonite.

Here, the experimental setup with different coral species (Fig. 1) and constant ambient conditions allowed the
number of factors involved to be reduced and, in particular, the Ba isotopic composition of the ambient water to be measured. Thus, the magnitude and direction of Ba isotope fractionation during coral growth can be investigated. In addition, corals from different natural settings (tropical shallow-water corals: Bahamas/Florida; deep, cold-water corals: Norwegian continental shelf, Fig. 2) were analysed to get a first indication of Ba isotope fractionation in natural habitats. Further, a set of five Mediterranean surface water samples taken over a distance of >2200 km east to west provides a first glimpse of surface water homogeneity at this scale (Fig. 3). The purpose of this study is to evaluate (i) if Ba isotope fractionation in corals is on the same order as that of abiogenic precipitation or if significant vital effects are observable; and (ii) if corals from natural settings show larger variations than explainable by precipitation, that is, are there indications for isotopically heterogeneous seawater.

![Corals from natural habitats](image)

**Fig. 2.** Corals from natural habitats. The bulk of the samples pictured was powdered for analysis. (A and B) *Favia fragum*; (C) *Agaricia agaricites*; (D) *Montastraea cavernosa*; (E) *Lophelia pertusa*. 
MATERIAL AND METHODS

Coral skeleton and seawater samples, experimental approaches and sampling environments

Characteristics of the coral samples are summarized in Tables 1 and 2. Two sets of samples consisting of tropical scleractinian corals (cultured and from natural settings), a standardized coral powder and a cold-water scleractinian coral were analysed.

The first sample set is composed of *Porites* sp., *Acropora* sp., *Stylophora* sp. and *Montipora* sp. cultured under constantly monitored conditions at the Centre Scientifique de Monaco. The aquaria were supplied with Mediterranean seawater pumped from a site situated at 50 m depth and 30 m off the coastline (Centre Scientifique de Monaco, Fig. 3). The salinity was monitored using a conductivity meter (Mettler LF 196) and was found to stay almost constant at 38. Seawater was maintained at a temperature of 25°C using a temperature controller (EW, PC 902/T). Metal halide lamps (Philips HPIT, 400 W) provided irradiance of 204 μmol m⁻² s⁻¹ on a 12 : 12 photoperiod. The seawater was continuously aerated with ambient air. A water sample was collected in the aquarium, representing the ambient seawater in which the cultured corals grew (‘Monaco seawater’). Prior to grinding of the coral tips in an agate mortar, coral skeletons were bleached for 24 h with NaClO (10%), ultrasonic-cleaned and rinsed with Milli-Q water in order to remove organic matter.

The second sample set comprises sub-Recent tropical corals *Montastraea cavernosa*, collected off Mayaguana Island (Bahamas, Fig. 3; 22°25.2'47"N, 73°8.99'W) in 2008, and *Favia fragum* and *Agaricia agaricites* from the Florida Keys (Florida, USA; 24°44'23-94"N, 80°58'51-54"W) collected in 2004. The samples were ultrasonic-cleaned in Milli-Q water and powdered in an agate mortar prior to analysis. Unlike the other samples, the coral skeletons from the Bahamas and Florida are not pristine as their aragonite content ranges from 79½ to 89½% (Table 2), that is, up to 20% of aragonite is recrystallized into secondary calcite. In addition, a standardized coral powder (JCp-1, Geological Survey of Japan) was analysed. JCp-1 is used as an international reference material in coral skeleton studies (Okai et al., 2002; Inoue et al., 2004). This sample comes from a colony of *Porites* sp. off Japan and consists of 100% aragonite.

A sub-Recent *Lophelia pertusa* from the Norwegian continental shelf collected at a depth of around 200 m (Freiwald et al., 2002) represents a cold-water scleractinian coral specimen. Prior to analysis, the coral sample was ultrasonic-cleaned in Milli-Q to remove attached sedimentary particles. The aragonitic skeleton of cold-water corals like *L. pertusa* may be coated post-mortem by a ferromanganese oxyhydroxide crust (Lomitschka & Mangini, 1999) as is the case for our sample (Fig. 2E). To test the potential influence of this coating on Ba isotope data, a portion of the sample was treated with 0.1M HCl to remove the coating (Copard et al., 2010). Barium contents were very similar prior to (15 ± 0.7 mg kg⁻¹, Loph-1) and after this procedure (13 ± 0.6 mg kg⁻¹,
Table 2. Sample characteristics and geochemical data set

| Location                  | Nature     | Species/name                  | Ba (mg kg⁻¹) | Ba/Ca (µmol mol⁻¹) | D (Ba/Ca) /Ca₀ | n | C (µg) | BarCa (µmol mol⁻¹) | Data source |
|---------------------------|------------|-------------------------------|--------------|--------------------|----------------|---|--------|-------------------|-------------|
| CSM Tropical coral        | Cultured corals | Acropora sp. (1) | 5.98E+04 | 0.34              | 5.39E+04 | 100 | 0.17 | 3 379 700            | 1.4         |
|                           |            | Acropora sp. (2)                  | 6.41E+04 | 0.36              | 6.40E+04 | 100 | 0.21 | 3 379 700            | 1.5         |
|                           |            | Meritipora sp.                  | 5.98E+05 | 0.22              | 5.98E+05 | 100 | 0.09 | 3 383 900            | 1.5         |
|                           |            | Porites sp.                     | 1.98E+05 | 0.41              | 1.98E+05 | 100 | 0.12 | 3 377 800            | 1.4         |
|                           |            | Sphenopora sp.                  | 5.98E+04 | 0.16              | 5.98E+04 | 100 | 0.08 | 3 377 800            | 1.4         |
|                           | Natural settings | Rival flagum (1) | 2.36E+05 | 0.55              | 2.36E+05 | 85  | 0.08 | 3 200 m depth        | 8           |
|                           |            | Rival flagum (2)                | 5.7        | 0.56              | 5.7        | 89  | 0.08 | 3 200 m depth        | 8           |
|                           |            | Favia fragum (1)                | 1.2E+04   | 0.04              | 1.2E+04   | 1   | 0.08 | 3 200 m depth        | 6           |
|                           |            | Favia fragum (2)                | 5.6        | 0.04              | 5.6        | 1   | 0.08 | 3 200 m depth        | 6           |
|                           |            | Agaricia agaricites             | 3.14E+04  | 0.08              | 3.14E+04  | 1   | 0.08 | 3 200 m depth        | 6           |
|                           |            | Montipora sp.                   | 2.16E+05  | 0.46              | 2.16E+05  | 100 | 0.16 | 3 377 800            | 1.4         |
|                           |            | Montipora sp.                   | 5.64E+05  | 0.29              | 5.64E+05  | 100 | 0.08 | 3 200 m depth        | 1.4         |
|                           |            | Stylophora CSN                  | 1.98E+05  | 0.16              | 1.98E+05  | 100 | 0.08 | 3 200 m depth        | 1.4         |
| Florida                   | Natural settings | Rival flagum (1) | 2.36E+05 | 0.55              | 2.36E+05 | 85  | 0.08 | 3 200 m depth        | 8           |
|                           |            | Rival flagum (2)                | 5.7        | 0.56              | 5.7        | 89  | 0.08 | 3 200 m depth        | 8           |
|                           |            | Favia fragum (1)                | 1.2E+04   | 0.04              | 1.2E+04   | 1   | 0.08 | 3 200 m depth        | 6           |
|                           |            | Favia fragum (2)                | 5.6        | 0.04              | 5.6        | 1   | 0.08 | 3 200 m depth        | 6           |
|                           |            | Agaricia agaricites             | 3.14E+04  | 0.08              | 3.14E+04  | 1   | 0.08 | 3 200 m depth        | 6           |
|                           |            | Montipora sp.                   | 2.16E+05  | 0.46              | 2.16E+05  | 100 | 0.16 | 3 377 800            | 1.4         |
|                           |            | Montipora sp.                   | 5.64E+05  | 0.29              | 5.64E+05  | 100 | 0.08 | 3 200 m depth        | 1.4         |
|                           |            | Stylophora CSN                  | 1.98E+05  | 0.16              | 1.98E+05  | 100 | 0.08 | 3 200 m depth        | 1.4         |
|                           | Florida    | Favia fragum (1)                | 2.36E+05  | 0.55              | 2.36E+05  | 85  | 0.08 | 3 200 m depth        | 8           |
|                           |            | Favia fragum (2)                | 5.7        | 0.56              | 5.7        | 89  | 0.08 | 3 200 m depth        | 8           |
|                           |            | Favia fragum (1)                | 1.2E+04   | 0.04              | 1.2E+04   | 1   | 0.08 | 3 200 m depth        | 6           |
|                           |            | Favia fragum (2)                | 5.6        | 0.04              | 5.6        | 1   | 0.08 | 3 200 m depth        | 6           |
|                           |            | Agaricia agaricites             | 3.14E+04  | 0.08              | 3.14E+04  | 1   | 0.08 | 3 200 m depth        | 6           |
|                           |            | Montipora sp.                   | 2.16E+05  | 0.46              | 2.16E+05  | 100 | 0.16 | 3 377 800            | 1.4         |
|                           |            | Montipora sp.                   | 5.64E+05  | 0.29              | 5.64E+05  | 100 | 0.08 | 3 200 m depth        | 1.4         |
|                           |            | Stylophora CSN                  | 1.98E+05  | 0.16              | 1.98E+05  | 100 | 0.08 | 3 200 m depth        | 1.4         |

A set of four surface seawater samples was taken on a transect through the Mediterranean Sea (Fig. 3) on-board R/V Maria S. Merian (Cruise MSM33) in November, 2013. Continuously supplied surface water from about 5 m water depth (MSM on-board sampling system) was sampled with 60 cm³ plastic syringes, filtered through 0.2 µm syringe filters (Minisart, Sartorius) into pre-conditioned polyethylene bottles, and acidified to final 1 vol % HNO₃ (Winde et al., 2014) according to established procedures of filtering alkaline seawater through the membrane filters for earth alkaline element analyses. Seawater salinity and temperature data as well as geographical site coordinates were obtained from the standard MSM board equipment.

Sample preparation and analytical procedures

The analytical procedure of Ba extraction for isotope ratio measurements was conducted using PFA labware in a class 100 laminar flow hood. All acids were distilled before application. A detailed description of the chromatographic purification of Ba from carbonate matrices can be found in Pretet (2013). In brief, several grams of bulk coral material (see Figs 1 and 2) were powdered and homogenized. Aliquots comprising 2 µg of Ba – corresponding to 40 to 400 mg per analysis – were first mixed with an appropriate amount of ¹³⁸Ba,¹³⁹Ba double-spike, following the procedure reported in von Allmen et al. (2010). The spiked carbonate samples were then digested in 5 ml of 6-4 M HCl at 90°C for 12 h. After digestion, the samples were dried down and then redissolved in 0.6 ml of 2-5 M HCl to be loaded onto the column. The column chemistry was performed in PFA micro-columns (4-0 mm ID; Savillex, Eden Prairie, MN, USA) equipped with 4 mm PFA frits (30 µm pore size) and filled with 1 cm³ of Dowex® 50WX8 cation-exchange resin (200 to 400 mesh). First, the resin was cleaned with 10 ml of 6-4 M HCl and conditioned with 4 ml of 2-5 M HCl. Sample loading and removal of the matrix were conducted with 8-9 ml of 2-5 M HCl in total. Finally, Ba was eluted in 6 ml of 6-4 M HCl and taken to dryness.

A modified chromatographic procedure was applied for Ba separation from seawater, consisting of two cation-exchange columns. Before purification, 15 ml of Mediterranean seawater, corresponding to ca 100 ng Ba, were

Loph-2). Moreover, the two aliquots have identical ¹³⁷/¹³⁸Ba values (Table 2). We therefore conclude that coating by ferromanganese oxyhydroxides has a negligible influence on the Ba isotope composition of the coral skeleton analysed in this study. A previous study has shown that dissolved Mn does not affect the measured Ba isotope signal (Böttcher et al., 2012).

...
double-spiked and evaporated at 100°C. This aliquot size provides enough Ba for one isotope measurement, while minimizing the amount of matrix elements. The precipitated salt was then equilibrated with 0.5 ml of 2.5 M HCl for about 10 h prior to ion-exchange purification. Remaining salt that did not re-dissolve was removed by centrifugation and discarded. Barium recovery from the evaporates was found to be quantitative, that is, less than 1% of total Ba was lost. In a first column, we removed the largest fraction of major elements. For this we used Spectrum® PP Columns 104704 (Spectrum Chromatography, Houston, TX, USA) filled with 2 cm³ of Dowex® 50WX8 cation-exchange resin. Cleaning and conditioning of the resin was done with 20 ml of 6-4 M HCl and 7 ml of 2-5 M HCl, respectively. The samples were loaded in 0.5 ml of 2-5 M HCl, followed by matrix removal with 5-5 ml of 2-5 M HCl and 2 ml of 6-4 M HCl. Barium was collected in 15 ml of 6-4 M HCl. The Ba yield of this first column was ca 90%. Removal of Na, Mg and K was close to 100%, whereas ca 15% of Ca and ca 55% of Sr were eluted together with Ba. Therefore, Ba was further purified from the remaining matrix in a second step, following the procedure outlined above for carbonate samples. The total Ba yield of our chromatographic separation from seawater was ca 80%. To test for potential analytical isotope fractionation, four aliquots of one seawater sample were spiked at different, successive analytical steps. They gave identical results of the coral standard JCp-1 with other studies. Two replicate measurements of JCp-1 resulted in a maximum of five standards (one after each three samples) was measured per analytical session. The δ137/134Ba values of the samples reduced by the double-spike routine were further corrected with the average value of the standard solution measurements during each session. The long-term (2011 to 2013) intermediate precision of the reference solution was ± 0.10‰ (2sd, n = 270). To assess the precision of coral sample analyses, the pooled standard deviation (2sp) was calculated, giving an average deviation of all repeated samples of ± 0.11‰ (2sp). The intermediate precision of four Mediterranean water samples analysed in 2015 was <0.08‰ due to improved analytical conditions.

Accuracy of the analyses can be assessed by comparison of the coral standard JCp-1 with other studies. Two replicate measurements of JCp-1 resulted in a δ137/134Ba value of 0.16 ± 0.01‰. Horner et al. (2015) measured a δ136/134Ba value of 0.29 ± 0.03‰ relative to NIST SRM 3104a. Assuming mass-dependent behaviour, this value can be recalculated to δ137/134Ba = 0.22‰. Furthermore, we can relate this result to Fluka Ba(NO₃)₂ by using δ137/134BaIAEA-CO-9 = 0.017 ± 0.049‰ (Nan et al., 2015) and δ137/134BaOIAEA-CO-9 - Fluka = −0.03 ± 0.06‰ (von Allmen et al., 2010). The recalculated δ137/134Ba of JCp-1 from Horner et al. (2015) is 0.19‰ relative to Fluka Ba(NO₃)₂, which is within analytical uncertainty identical to the value of 0.16‰ measured here. Note that all data from Horner et al. (2015) cited here were recalculated accordingly. Similarly, data from Cao et al. (2016), that are originally given relative to a Ba(NO₃)₂ solution from Inorganic Ventures, were recalculated using IAEA-SO-5 (δ137/134BaIAEA-SO-5 = 0.12 ± 0.10‰) and δ137/134BaOIAEA-CO-9-Fluka = 0.02 ± 0.05‰ (von Allmen et al., 2010). The Ba concentration of JCp-1 was measured to be 9 mg kg⁻¹, which is in good agreement with results of Okai et al. (2002) (10.5 ± 0.3 mg kg⁻¹ ± 3% RSD) and Inoue et al. (2004) (8 mg kg⁻¹ ± 3% RSD).

**MC-ICP-MS analysis**

Barium isotope analyses were performed using a double focusing Nu Instruments® ‘Nu plasma’ multi-collector ICP-MS at the Institute of Geological Sciences, Bern. Samples were introduced via a PFA nebulizer connected to an ESI® Apex-Q. The measurements were controlled for isobaric interferences of tellurium (Te), affecting mass 130Ba, and of xenon (Xe) on 130Ba, 132Ba and 134Ba. No Te was detected during the measurements. Signal intensities of 130Xe, 132Xe and 134Xe were calculated based on the 131Xe beam and Xe isotope ratios taken from IUPAC (de Laeter et al., 2003), which were exponentially corrected for instrumental isotope fractionation.

Data acquisition included three to five blocks of 10 cycles of 10 seconds, depending on the available quantity of sample Ba. Peak centring was performed prior to each block. The isotopic values were expressed relative to a Ba nitrate ICP-OES standard solution from Fluka-Aldrich® as (expressed in %):
RESULTS

Barium concentrations

Barium concentrations in skeletal material are commonly normalized to calcium (Ca) and reported as Ba/Ca ratios. A distribution coefficient $D_{(\text{Ba/Ca})}$ is then applied to derive the respective composition of seawater from the analysed skeleton (Lea & Spero, 1994; Böttcher & Dietzel, 2010), which is defined according to the following equation:

$$\frac{(\text{Ba}/\text{Ca})_{\text{carbonate}}}{(\text{Ba}/\text{Ca})_{\text{solution}}} = D_{(\text{Ba/Ca})}$$

However, as $(\text{Ba/Ca})_{\text{solution}}$ is constant for the cultured corals, $(\text{Ba}/\text{Ca})_{\text{carbonate}}$ ratios bear the same information on relative variations as $D_{(\text{Ba/Ca})}$. Furthermore, Ba concentrations vary by one order of magnitude in the corals, while the range of Ca concentrations is smaller than 2% (Table 2). Thus, in the data set of cultured corals, Ba concentrations basically carry the same information as Ba/Ca ratios or $D_{(\text{Ba/Ca})}$.

Mediterranean seawater barium isotope composition

Sampling of the seawater supplied for the aquarium experiments showed a Ba isotopic composition of $0^{+38}_{-0.07}$ in $\delta^{137/134}$Ba (2sd, Table 2). In order to estimate potential variations in modern surface seawater, a further set of four samples from Mediterranean surface waters were analysed for their Ba isotope composition (Table 3). Results range from $0.38$ to $0.46_{-0.07}$ with individual uncertainties of $\leq 0.07$ (2sd; Fig. 4). These data indicate an essentially homogeneous $\delta^{137/134}$Ba for Mediterranean surface waters. The Ba isotope composition of deeper water masses in the Mediterranean may differ from those of the surface waters and are the subject of further investigations.

Barium isotope composition of coral skeleton

The $\delta^{137/134}$Ba values of scleractinian coral skeletons, comprising different coral families and genera, range

| Sample          | Location                     | Sampling date (2013) | Salinity (S) | $T$ (°C) | Ba (nmol kg$^{-1}$) | $\delta^{137/134}$Ba$_{\text{SM}}$ | 2sd | n  |
|-----------------|------------------------------|----------------------|--------------|----------|-------------------|-----------------------------------|-----|----|
| MSM_033_685, #2 | 36°15'972”N, 3°8’892’W      | November 2nd         | 36.7         | 19-14    | 43.0              | 0.38                              | 0.06 | 2  |
| MSM_033_688, #5 | 37°1’283”N, 2°25’914”E      | November 3rd         | 36.7         | 21-8     | 42.0              | 0.45                              | 0.07 | 2  |
| MSM_033_690, #7 | 37°31’494”N, 7°49’722”E     | November 4th         | 37.8         | 22-6     | 50.0              | 0.46                              | 0.08*| 1  |
| MSM_033_707, #17| 34°33’3758”N, 21°33’537”E   | November 5th         | 39.4         | 22.5     | 54.0              | 0.40                              | 0.03 | 2  |
| Aqueduct        | Monaco CSM                  |                      | 38.0         | 25.0     | 58.0              | 0.42                              | 0.18 | 5  |
| Average         | Mediterranean               |                      | 49.3         |          |                  | 0.42                              | 0.07 |     |

*Intermediate precision of the reference solution during analytical period: $<0.08$ (cf. Text).
Lophelia pertusa coral

Here, significant differences exist between the cold-water D(Ba/Ca) values of 1\textsuperscript{et al.} pioneering work of Lea and the tropical corals (Florida and the Bahamas; different coral species and locations. The D(Ba/Ca) values of cultured corals have identical (\(\delta^{137/134}\text{Ba}\)) values and Ba concentration in the coral data (Fig. 6). Also, the values are independent from the systematic position of the respective genera and species (cf. Table 1).

The range in \(\delta^{137/134}\text{Ba}\) of the analysed natural corals is much larger (0-14 to 0.56, with one specimen yielding 0.77 ± 0.11\textsuperscript{oo} Table 2) than that of the cultured species. Here, significant differences exist between the cold-water coral Lophelia pertusa (\(\delta^{137/134}\text{Ba} = 0.18 \pm 0.08\textsuperscript{oo}\)) and the tropical corals (Florida and the Bahamas; A. agaricites, F. fragum and M. cavernosa).

**INTERPRETATION AND DISCUSSION**

**Barium concentrations and empirical distribution coefficients**

The D\(_{(\text{Ba/Ca})}\) values of cultured corals vary between 1.4 and 9.8 (Table 2). Most values are in agreement with the pioneering work of Lea \textit{et al.} (1989), who found average D\(_{(\text{Ba/Ca})}\) values of 1.41 ± 0.14 and 1.27 ± 0.03 for two different coral species and locations. The D\(_{(\text{Ba/Ca})}\) values for aragonite that was abiogenetically precipitated under laboratory conditions were found to be in the range of about 1.0 to 2.6, and to be inversely correlated with temperature (Dietzel \textit{et al.}, 2004; Gaetani & Cohen, 2006).

Thus, estimated D\(_{(\text{Ba/Ca})}\) values of all but one cultured coral are close to the range of experimental results for abiogenic Ba incorporation into aragonite at high precipitation rates. However, the pronounced D\(_{(\text{Ba/Ca})}\) of one aquarium coral of 9.8 is too high to be explained as dominated by abiogenic Ba incorporation and cannot be reconciled by a unique D\(_{(\text{Ba/Ca})}\) value. Thus, it appears reasonable to consider additional vital effects, as suggested previously by Gaetani & Cohen (2006), impacting internal pool sizes or crystallization kinetics beyond the scale considered in experiments so fare.

The Ba concentrations of the corals from natural settings presented here (5 to 24 mg kg\(^{-1}\)) largely overlap with the cultured corals (8 to 57 mg kg\(^{-1}\)).

**Coral skeleton Ba isotope fractionation**

Barium isotope fractionation between seawater and coral skeleton is derived from the data of cultured corals (Table 2). As \(\delta^{137/134}\text{Ba}_{\text{seawater}}\), Ba stable isotope fractionation during biogenic Ba uptake seems to be similar to isotope effects during precipitation of aragonite-structured BaCO\(_3\) (wetherite) (von Allmen \textit{et al.}, 2010). It is further comparable to isotope fractionation of Sr (Krabbenhoff \textit{et al.}, 2009), Mg (Yoshimura \textit{et al.}, 2011), and Ca (Böhm \textit{et al.}, 2006) in coral skeletons (see introduction). The experiments performed by von Allmen \textit{et al.} (2010) suggest that different supersaturation degrees, and associated precipitation rates, influenced the reaction and the enrichment of light isotopes in the precipitates. The maximal observed Ba isotope fractionation between seawater and cultured corals (\(-0.26\textsuperscript{oo}\) in \(\Delta^{137/134}\text{Ba}_{\text{seawater}}\)) is still consistent both in direction and magnitude with previous precipitation experiments (von Allmen \textit{et al.}, 2010; Böttcher \textit{et al.}, 2012). A similar fractionation factor was approximated by Horner \textit{et al.} (2015), whose minimum estimate of \(\Delta^{137/134}\text{Ba}_{\text{seawater}}\) corresponds to a \(\Delta^{137/134}\text{Ba}_{\text{seawater}}\) of \(-0.21\textsuperscript{oo}\).

Substantial differences in \(\delta^{137/134}\text{Ba}\) values for corals from the same environment, that is, the aquarium, cannot be attributed to simple, quasi abiogenic precipitation processes in isotope exchange equilibrium with bulk (aquarium) water, as already indicated by the variable D\(_{(\text{Ba/Ca})}\) values. A simple first order model for this observation is that variable fractions of the given amount of seawater were precipitated during the bio-mineralization process. The question whether skeletogenesis in corals includes an open or semi-closed calcification compartment, is still a matter of intense debate (Böhm \textit{et al.}, 2006; Cohen &
Holcomb, 2009; Cohen & Gaetani, 2010; Allemand et al., 2011; Gaetani et al., 2011; Tambutte et al., 2011; Gagnon et al., 2012). A semi-closed calcification compartment might provide the restricted reservoir needed for a Rayleigh-type Ba isotope fractionation. Taken at face value, the variations in δ137/134Ba of the cultured corals could reflect a range of 25 to 99% of Ba from a given amount of seawater precipitated in a Rayleigh-type fractionation process, using a model with a Mediterranean δ137/134Ba_seawater value as initial seawater value (0.42 ± 0.07‰, 2sd) and a theoretical Δ137/134Ba_precipitate-fluid of −0.3‰ (von Allmen et al., 2010). However, a semi-closed calcification compartment is just one hypothesis to explain isotope fractionation in corals. For example, Inoue et al. (2015) reported combined δ44/40Ca and Sr/Ca of Porites australiensis samples from multiple culture experiments. They conclude that their results are not compatible with a Rayleigh-type fractionation directly from a fluid, which is seawater-like in terms of δ44/40Ca and Sr/Ca. Therefore, Inoue et al. (2015) favoured Ca-isotope fractionation in corals to be the result of Ca transmembrane transport (Tambutte et al., 1996). Although these data give indications of isotope fractionation in corals also for Ba, the fractionation is comparatively small given the associated uncertainties and, thus, do not allow us to make conclusions on the competing hypotheses on bio-mineralization.

Diagenetic recrystallization

Following the experimentally derived Ba isotope fractionation factor during carbonate precipitation and the results of our coral culture experiments, the δ137/134Ba value of a bulk coral is not higher than that of the ambient seawater, unless secondary processes contribute to the overall isotope fractionation. This finding is consistent with results for Sr or Ca (see introduction). In the coral samples from Florida and the Bahamas, up to 20% of the aragonite is recrystallized into calcite (Table 2), a process that may change the Ba isotopic compositions significantly. However, only very low amounts of Ba can be incorporated into the lattice of secondary calcite during recrystallization (Böttcher, 1997; Böttcher & Dietzel, 2010) or nano-scale dissolution-reprecipitation. Accordingly, calcitic fossils show D_Ba/Ca values consistently <1 (e.g. foraminifers (Lea & Spero, 1994; Hönsch et al., 2011) and calcitic bivalve Mytilus eludes (Gillikin et al. 2006)). Pingitore & Eastman (1985) investigated the Ba partitioning during recrystallization of coral aragonite to calcite. They compared altered and unaltered corals from the Pleistocene reef terraces exposed on Barbados, West Indies, and reported a Ba decrease from typical ranges of 8 to 15 mg kg$^{-1}$ in aragonite to 1 to 3 mg kg$^{-1}$ in secondary calcite. Therefore, the bulk carbonate δ137/134Ba should remain dominated by the composition of the original aragonite fraction. This phenomenon can be illustrated by a simple mass balance model. To increase an aragonitic skeleton δ137/134Ba value of 0-4 (Mediterranean-like seawater) to 0-8 (like A. agaricites from the Florida setting), considering a 12% fraction of recrystallized calcite (as observed), and a Ba concentration five times lower in calcite than in primary aragonite (based on Pingitore & Eastman, 1985), the calcite fraction would have to carry a δ137/134Ba value as high as +16‰. Therefore, partial diagenetic carbonate recrystallization is deemed unlikely to alter the primary Ba isotopic composition of a coral significantly.

Heterogeneity of the Ba isotopic composition of seawater

Precipitation of carbonate causes Ba isotope fractionation towards lower δ137/134Ba values in a solid. Currently no data are available that would indicate a process that drives the Ba isotopic composition of a precipitate to δ137/134Ba values above the corresponding dissolved Ba. This holds generally also true for isotope systems of other Earth alkaline elements (Nielsen et al., 2012). Observed exceptions are Ca-isotope fractionation between aqueous solutions and inorganic solids that trap the hydrated ion in the lattice (Colla et al., 2013) and the equilibrium Mg isotope fractionation factor between inorganic hydrous magnesium sulphate and aqueous MgSO$_4$ solutions (e.g. epsomite MgSO$_4$·7H$_2$O, Li et al., 2011).

The δ137/134Ba value of A. agaricites from Florida (0.77 ± 0.11‰) is significantly higher than that of Mediterranean seawater (0.42 ± 0.07‰). Given the above outlined conditions, carbonates of A. agaricites cannot have been precipitated from seawater with a Ba isotopic composition identical to that of Mediterranean surface waters. Indeed, recent studies indicate heterogeneous Ba isotopic compositions of surface ocean waters, ranging in δ137/134Ba from 0-45 to 0-8‰ (Horner et al., 2015; Cao et al., 2016). Similarly, the vertical seawater heterogeneity, with shifts in δ137/134Ba of about 0-2 to 0-3‰ (Horner et al., 2015; Cao et al., 2016), could be an explanation for the lower δ137/134Ba values of the cold-water coral L. pertusa of 0-18 ± 0.08‰. We thus tentatively interpret the variations in the Ba isotopic composition of the investigated natural corals as indicators of spatial and/or temporal differences in the barium isotope composition of seawater.

CONCLUSIONS

Culture experiments at the Centre Scientifique de Monaco allowed for the first time the investigation of stable Ba
isotope fractionation between seawater and coral skeletons under controlled standard state conditions (25°C, 1 atm total pressure).

The seawater sample of the supply for the aquarium experiments and a set of four additional surface water samples from the Mediterranean Sea, collected over a distance of >2200 km from east to west, support a homogeneous Ba isotopic composition for Mediterranean surface waters with an average $\delta^{137/134}$Ba value of 0.42 ± 0.07‰.

The cultured corals show either no Ba isotope fractionation relative to ambient seawater or slight enrichment in the lighter Ba isotopes ($A^{137/134}$Ba_{coral-seawater} between −0.01‰ and −0.26‰). The direction and magnitude of Ba isotope fractionation during coral growth is comparable with fractionation during precipitation of inorganic carbonates observed by a previous experimental study. The observed range of $D_{(\text{Ba/Ca})}$ values in the cultured corals in part exceed previously reported Ba distribution in abiotic experiments, and indicate additional vital effects leading to reservoir effects or crystallization velocities beyond the conditions of previous experimental calibrations. However, no correlation between $D_{(\text{Ba/Ca})}$ and the Ba isotopic composition of the corals is found in the growth experiments of this study. Instead, isotope fractionation might be governed by the efficiency of Ba incorporation into the coral skeleton. The number of investigated samples and the analytical resolution, however, do not allow further conclusions to be drawn.

One naturally grown tropical coral from Florida (A. agaricites) exhibits a $\delta^{137/134}$Ba value significantly higher than that of Mediterranean surface water, suggesting heterogeneous Ba isotopic compositions of surface seawaters, which is consistent with recently published data by Horner et al. (2015) and Cao et al. (2016). While the Ba isotope system is still in its infancy, these findings provide support to the use of coral Ba isotopes as a palaeo-proxy for water masses and the productivity related isotope composition they carried.

ACKNOWLEDGEMENTS

This research was supported by the Swiss National Science Foundation (SNF) through grants 20MA21-115944, 200020_140618 and 200021_140223. The 2 first grants were in the frame of the European Science Foundation (ESF) EUROCORES Program EuroMARC. MEB wishes to thank Deutsche Forschungsgemeinschaft and Leibniz IOW for support of sampling during research cruise MSM33 in the Mediterranean. Thanks are due to A. Freiwald (Wilhelmshaven, Germany) for providing sample of L. pertusa. We are grateful to two anonymous referees and journal editors Adrian Immenhauser and Peter Swart for constructive comments and suggestions for improvement.

The data will be available after publication on the PANGEA repository (http://www.pangaea.de).

References

Allemand, D., Tambutté, E., Zoccola, D. and Tambutté, S. (2011) Coral calcification, cells to reefs. In: Coral Reefs: An Ecosystem in Transition (Eds Z. Dubinsky and N. Stambler), pp. 119–150. Springer, Dordrecht.

von Allmen, K., Böttcher, M.E., Samankassou, E. and Nägler, T. (2010) Barium isotope fractionation in the global barium cycle: first evidence from barium minerals and precipitation experiments. Chem. Geol., 277, 70–77.

Babu, C., Brumsack, H.-J., Schnetger, B. and Böttcher, M.E. (2002) Barium as a productivity proxy in continental margin sediments: a study from the eastern Arabian Sea. Mar. Geol., 184, 189–206.

Bohm, F., Gussone, N., Eisenhauer, A., Dullo, W.-C., Reynaud, S. and Paytan, A. (2006) Calcium isotope fractionation in modern scleractinian corals. Geochim. Cosmochim. Acta, 70, 4452–4462.

Böttcher, M.E. (1997) Comment on: ‘Solid solution partitioning of Sr$$^\text{2+}$$ and Ba$$^\text{2+}$$, and Cd$$^\text{2+}$$ to calcite’ by A.J. Tessoriero and J.F. Panckow. Geochim. Cosmochim. Acta, 61, 661–662.

Böttcher, M.E. and Dietzel, M. (2010) Metal-ion partitioning during low-temperature precipitation and dissolution of anhydrous carbonates and sulfates. EMU Notes Mineral., 10, 139–187.

Böttcher, M.E., Geprägs, P., Neubert, N., von Allmen, K., Pretet, C., Samankassou, E. and Nägler, T.F. (2012) Barium isotope fractionation during experimental formation of the double carbonate BaMn(CO$_3$)$_2$ at ambient temperature. Isot. Environ. Health Stud., 48, 457–463.

Cao, Z., Siebert, C., Hathorne, E.C., Dai, M. and Frank, M. (2016) Constraining the oceanic barium cycle with stable barium isotopes. Earth Planet. Sci. Lett., 434, 1–9.

Cardinal, D., Savoye, N., Trull, T.W., André, L., Kopczynska, E.E. and Dehairs, F. (2005) Variations of carbon remineralisation in the Southern Ocean illustrated by the Ba (xs) proxy. Deep Sea Res. Part I, 52, 355–370.

Chan, L.H., Edmond, J.M., Stallard, R.F., Broecker, W.S., Chung, Y.C., Weiss, R.F. and Ku, T.L. (1976) Radium and barium at GEOSECS stations in the Atlantic and Pacific. Earth Planet. Sci. Lett., 32, 258–267.

Chester, R. (2009) Marine Geochemistry. Wiley, Oxford, 506 pp.

Church, T.M. (1979) Marine barite. In: Marine Minerals (Ed. R.G. Burns), Rev. Mineral., 6, pp., 175–210.

Cohen, A.L. and Gaetani, G.A. (2010) Ion partitioning and the geochemistry of coral skeletons: solving the mystery of the vital effect. EMU Notes Mineral., 11, 377–397.

Cohen, A. and Holcomb, M. (2009) Why corals care about ocean acidification? Uncovering the mechanism. Oceanography, 22, 118–127.
Colla, C.A., Wimpenny, J., Yin, Q.-Z., Rustad, J.R. and Casey, W.H. (2013) Calcium-isotope fractionation between solution and solids with six, seven or eight oxygens bound to Ca(II). Geochim. Cosmochim. Acta, 121, 363–373. doi: 10.1016/j.gca.2013.07.041.

Copard, K., Colin, C., Douville, E., Freiwalder, A., Gudmundsson, G., De Mol, B. and Frank, N. (2010) Nd isotopes in deep-sea corals in the North-eastern Atlantic. Quatern. Sci. Rev., 29, 2499–2508.

Dickens, G.R., Fewless, T., Thomas, E. and Bralower, T.J. (2003) Excess barite accumulation during the Paleocene-Eocene Thermal Maximum: massive input of dissolved barium from seafloor gas hydrate reservoirs. Geol. Soc. Am. Spec. Pap., 369, 11–23.

Dietzel, M., Gussone, N. and Eisenhauer, A. (2004) Co-precipitation of Sr²⁺ and Ba²⁺ with aragonite by membrane diffusion of CO₂ between 10 and 50 degrees C. Geochim. Cosmochim. Acta, 68, 363–370.

Dymond, J., Suess, E. and Lyle, M. (1992) Barium in deep-sea sediment: a geochemical proxy for paleoproductivity. Paleocenography, 7, 163–181.

Eugster, O., Tera, F. and Wasserburg, G.J. (1969) Isotopic analyses of barium in meteorites and in terrestrial samples. J. Geophys. Res., 74, 3897–3908.

Fietzke, J. and Eisenhauer, A. (2006) Determination of temperature-dependent stable strontium isotope (⁸⁸Sr/⁸⁶Sr) fractionation via bracketing standard MC-ICP-MS. Geochim. Geosyst., 7, Q08009. doi:10.1029/2006GC001243.

Francois, R., Honjo, S., Manganini, S.J. and Ravizza, G.E. (1995) Biogenic barium fluxes to the deep sea: implications for paleoproductivity reconstruction. Global Biogeochem. Cycles, 9, 289–303.

Freiwalder, A., Hühnerbach, V., Lindberg, B., Wilson, J.B. and Campbell, J. (2002) The Sula reef complex, Norwegian shelf. Facies, 47, 179–200.

Gaetani, G.A. and Cohen, A.L. (2006) Element partitioning during precipitation of aragonite from seawater: a framework for understanding paleoproxies. Geochim. Cosmochim. Acta, 70, 4617–4634.

Gaetani, G.A., Cohen, A.L., Wang, Z. and Crusius, J. (2011) Rayleigh-based, multi-element coral thermometry: a biominalerization approach to developing climate proxies. Geochim. Cosmochim. Acta, 75, 1920–1932.

Gagnon, A.C., Adkins, J.F. and Erez, J. (2012) Seawater transport during coral biomineralization. Earth Planet. Sci. Lett., 329–330, 150–161.

Giillinik, D.P., Dehairs, F., Lorrain, A., Steenmans, D., Baeyens, W. and Andrie, L. (2006) Barium uptake into the shells of the common mussel (Mytilus edulis) and the potential for estuarine paleo-chemistry reconstruction. Geochimica et Cosmochimica Acta, 70, 395–407.

Hall, J.M. and Chan, L.H. (2004) Ba/Ca in benthic foraminifera: thermocline and middepth circulation in the North Atlantic during the last glaciation. Paleocenography, 19, PA4018.

Hönisch, B., Allen, K.A., Russell, A.D., Eggoins, S.M., Bijma, J., Spero, H.J., Lea, D.W. and Yu, J. (2011) Planktic foraminifers as recorders of seawater Ba/Ca. Mar. Micropaleontol., 79, 52–57.

Horner, T.J., Kinsley, C.W. and Nielsen, S.G. (2015) Barium-isotopic fractionation in seawater mediated by barite cycling and oceanic circulation. Earth Planet. Sci. Lett., 430, 511–522.

Hull, P.M. and Norris, R.D. (2011) Diverse patterns of ocean export productivity change across the Cretaceous-Paleogene boundary: new insights from biogenic barium. Paleocenography, 26, PA3205.

Inoue, M., Nohara, M., Okai, T., Suzuki, A. and Kawahata, H. (2004) Concentrations of trace elements in carbonate reference materials coral JCP-1 and giant clam ICT-1 by inductively coupled plasma-mass spectrometry. Geostand. Geoanal. Res., 28, 411–416.

Inoue, M., Gussone, N., Koga, Y., Iwase, A., Suzuki, A., Kazuhiho Sakai, K. and Kawahata, H. (2015) Controlling factors of Ca isotope fractionation in scleractinian corals evaluated by temperature, pH and light controlled culture experiments. Geochim. Cosmochim. Acta, 167, 80–92.

Krabbenhøft, A., Fietzke, J., Eisenhauer, A., Liebetrau, V., Böhm, F. and Vollstaedt, H. (2009) Determination of radiogenic and stable strontium isotope ratios (⁸⁷Sr/⁸⁶Sr, ⁸⁸Sr/⁸⁶Sr) by thermal ionization mass spectrometry applying an ⁸⁷Sr/⁸⁶Sr double spike. J. Anal. At. Spectrom., 24, 1267–1271.

de Laeter, J.R., Böhlike, J.K., De Bievre, P., Hidaka, H., Peiser, H.S., Rosman, K.J.R. and Taylor, P.D.P. (2003) Atomic weights of the elements. Review 2000 (IUPAC Technical Report). Pure Appl. Chem., 75, 683–800.

Lea, D.W. and Spero, H.J. (1994) Assessing the reliability of paleochemical tracers – barium uptake in the shells of planktonic foraminifera. Paleocenography, 9, 445–452.

Lea, D.W., Shen, G.T. and Boyle, E.A. (1989) Coralline barium records temporal variability in equatorial Pacific upwelling. Nature, 340, 373–376.

Li, W., Beard, B.L. and Johnson, C.M. (2011) Exchange and fractionation of Mg isotopes between epsomite and saturated MgSO₄ solution. Geochim. Cosmochim. Acta, 75, 1814–1828.

Lomitschka, M. and Mangini, A. (1999) Precise Th/U-dating of small and heavily coated samples of deep sea corals. Earth Planet. Sci. Lett., 170, 391–401.

Martinez-Ruiz, F., Kastner, M., Paytan, A., Ortega-Huertas, M. and Bernasconi, S.M. (2000) Geochemical evidence for enhanced productivity during S1 sapropel deposition in the eastern Mediterranean. Paleocenography, 15, 200–209.

McManus, J., Berelson, W.M., Hammond, D.E. and Klinkhammer, G.P. (1999) Barium cycling in the North Pacific: implications for the utility of Ba as a
paleoproductivity and paleoalkalinity proxy.
Paleoceanography, 14, 53–61.

Miyazaki, T., Kimura, J.-I. and Chang, Q. (2014) Analysis of stable isotope ratios of Ba by double-spike standard–sample bracketing using multiple-collector inductively coupled plasma mass spectrometry. J. Anal. At. Spectrom., 29, 483–490.

Monnin, C., Jeandel, C., Cattaldo, T. and Dehairs, F. (1999) The marine barite saturation state of the world’s oceans. Mar. Chem., 65, 253–261.

Nan, X., Wu, F., Zhang, Z., Hou, Z., Huang, F. and Yu, H. (2015) High-precision barium isotope measurements by MC-ICP-MS. J. Anal. At. Spectrom., 30, 2307–2315.

Nielsen, L., Druhan, J., Yang, W., Brown, S. and DePaolo, D. (2012) Calcium isotopes as tracers of biogeochemical processes. In: Handbook of Environmental Isotope Geochemistry. Advances in Isotope Geochemistry (Ed. M. Baskaran), pp. 105–124. Springer, Berlin, Heidelberg.

Okai, T., Suzuki, A., Kawahata, H., Terashima, S. and Imai, N. (2002) Preparation of a new geological survey of Japan geochemical reference material: coral JICp-1. Geostandards Newslett., 26, 95–99.

Paytan, A. and Griffith, E.M. (2007) Marine barite: recorder of variations in ocean export productivity. Deep Sea Res. Part II, 54, 687–705.

Paytan, A. and Kastner, M. (1996) Benthic Ba fluxes in the central Equatorial Pacific, implications for the oceanic Ba cycle. Earth Planet. Sci. Lett., 142, 439–450.

Paytan, A., Kastner, M. and Chavez, F.P. (1996) Glacial to Interglacial fluctuations in productivity in the equatorial Pacific as indicated by marine barite. Science, 274, 1355–1357.

Pingitore, N.E. and Eastman, M.P. (1985) Barium partitioning during the transformation of corals from aragonite to calcite. Chem. Geol., 48, 183–187.

Pretet, C. (2013) Non-Traditional Isotopes (Barium and Calcium) and Elemental Ratios in Scleractinian Coral Skeleton: New Look into Geochemical Cycles, Environmental Proxies and Bio-Calcification Processes. PhD Thesis, Terre et Environnement, University of Geneva. Available at: http://archive-ouverte.unige.ch/unige:33163

Raddatz, J., Liebetrau, V., Ruggeberg, A., Hathorne, E., Krabbenhöft, A., Eisenhauer, A., Böhm, F., Vollstaedt, H., Fietzke, J., López Correman, M., Freiwald, A. and Dullo, W.-C. (2013) Stable Sr-isotope, Sr/Ca, Mg/Ca, Li/Ca and Mg/Li ratios in the scleractinian cold-water coral Lophelia pertusa. Chem. Geol., 352, 143–152.

Schmitz, B. (1987) Barium, equatorial high productivity, and the northward wandering of the Indian continent. Paleoceanography, 2, 63–77.

Sprovieri, M., d’Alcala, M.R., Manta, D.S., Bellanca, A., Neri, R., Lirer, F., Taberner, C., Pueyo, J.J. and Sammartino, S. (2008) Ba/Ca evolution in water masses of the Mediterranean late Neogene. Paleoceanography, 23, PA3205.

Sternberg, E., Tang, D., Ho, T.-Y., Jeandel, C. and Morel, F.M.M. (2005) Barium uptake and adsorption in diatoms. Geochemica et Cosmochimica Acta, 69, 2745–2752.

Tambutté, E., Allemand, D., Mueller, E. and Jaubert, J. (1996) A compartmental approach to the mechanism of calcification in hermatypic corals. J. Exp. Mar. Biol. Ecol., 199, 1029–1041.

Tambutté, S., Holcomb, M., Ferrier-Pagès, C., Reynaud, S., Tambutté, E., Zoccola, D. and Allemand, D. (2011) Coral biomineralization: from the gene to the environment. J. Exp. Mar. Biol. Ecol., 408, 58–78.

Veron, J.E.N., Odorico, D.M., Chen, C.A. and Miller, D.J. (1996) Reassessing evolutionary relationships of scleractinian corals. Coral Reefs, 15, 1–9.

Weldeab, S., Lea, D.W., Schneider, R.R. and Andersen, N. (2007a) 155,000 years of West African monsoon and ocean thermal evolution. Science, 316, 1303–1307.

Weldeab, S., Lea, D.W., Schneider, R.R. and Andersen, N. (2007b) Centennial scale climate instabilities in a wet early Holocene West African monsoon. Geophys. Res. Lett., 34, L24702.

Winde, V., Böttcher, M.E., Escher, P., Böning, P., Beck, M., Liebezeit, G. and Schneider, B. (2014) Tidal and spatial variations of D13C and aquatic chemistry in a temperate tidal basin during winter time. J. Mar. Syst., 129, 396–404.

Wolgemuth, K. and Broecker, W.S. (1970) Barium in seawater. Earth Planet. Sci. Lett., 8, 372–378.

Yoshimura, T., Tamimizu, M., Inoue, M., Suzuki, A., Iwasaki, N. and Kawahata, H. (2011) Mg isotope fractionation in biogenic carbonates of deep-sea coral, benthic foraminifera, and hermatypic coral. Anal. Bioanal. Chem., 401, 2755–2769.