Human non-Hodgkin's malignant lymphomas serially transplanted in nude mice conditioned with whole-body irradiation

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Summary Direct transplantation of non-Hodgkin's malignant lymphoma into athymic nude mice was successfully achieved after whole-body irradiation (5 Gy). Twenty-seven per cent (6/22) of transplanted lymphomas were established as nude mouse lines. The successful lines were derived solely from the patients with diffuse lymphoma who showed advanced clinical stage, high LDH value, large mass and poor prognosis. The histological, immunophenotypic and chromosomal characteristics of the nude mouse lines were compared with those of the original lymphomas, and the proliferative characteristics of the lines were examined. The transplanted lymphomas substantially retained the characteristics of the original lymphomas, and could be useful in biological, oncological and therapeutic studies of human malignant lymphoma.

Materials and methods

Animals and irradiation

Nude mice (BALB/c nu/nu) are propagated in our institute and maintained under specific pathogen-free (SPF) conditions. Male nude mice (6–12 weeks old) were used as recipients. The mice were given 5 Gy whole body irradiation from a $^{137}$Cs source and kept under SPF conditions immediately before transplantation.

Clinical materials and methods of implantation

At diagnostic biopsy, fresh specimens of lymph nodes or tumours were excised aseptically from 21 patients (three follicular lymphoma, 17 diffuse lymphoma and one mycosis fungoides). Tumour specimens were cut into small pieces with scissors in a Petri dish containing Ham's F-10 medium, antibiotics and 10% calf serum. About 8 mm cubes of tumours thus prepared were implanted subcutaneously with a trocar. The implants were completed within 4 h after surgical excision. Lymphoma cells in the pleural effusion of one patient with lymphoblastic lymphoma were collected by centrifugation. The pellet (containing $2.2 \times 10^8$ cells) was also implanted subcutaneously.

Tumour take and serial transplantation

When implanted tumours began growing and attained 15–20 mm in diameter, mice were killed by cervical dislocation and their tumours were excised. Then the tumour pieces were passaged into at least three new mice. On every passage, this procedure was repeated for the establishment of the tumour line. A nude mouse line of more than five passages was defined as an established one. If tumour take was not observed within 90 days after implantation, it was regarded as unsuccessful. Autopsy was performed on all killed mice to check for metastasis.

Histopathological studies

The specimens of original and nude mouse tumours stained with Haematoxylin and Eosin were subjected to histopathological analysis according to the Working Formulation of non-Hodgkin's lymphomas (National Cancer Institute, 1982).

Immunophenotypic studies

Clinical and nude mouse line materials were minced and pipetted to prepare cell suspensions. Each suspension was placed on a Ficoll-Hypaque gradient (Nycomed AS, Oslo, Norway) and centrifuged. The pure lymphoma cells thus separated were used for the immunophenotypic studies. By applying the conventional sheep red cell rosetting technique, cells from clinical specimens were examined for T-cell markers.

Various monoclonal antibodies including OKla-I (Ortho, Raritan, NJ), LeuHLA-DR (Becton Dickinson, Sunnyvale, CA), B1 (CD20), B2 (CD21), B4 (CD19), J5 (CD10) (Coulter, Hialeah, FL) and so on conjugated with fluorescein isothiocyanate (FITC) were used to characterise the immunological phenotype in detail by flow cytometry (Ortho).

Frozen or paraffin-embedded specimens or tumour cell implants were subjected to immunohistological studies by using the 'ABC' immunoperoxidase method, as previously described by Hsu et al. (1981). To detect heavy and light chains of immunoglobulin on the cell surface and in the cytoplasm, rabbit antiserum (Ortho) were employed. A series of anti-T cell monoclonal antibodies such as Leu-1 (CD5), Leu-2a (CD8), Leu-3a (CD4) (Becton Dickinson) and OKT6 (CD1) (Ortho) were also used to study in detail the materials derived from T cell lymphoma and the nude mouse counterpart.

Chromosome analysis

The cells from six established nude mouse tumours were subjected to chromosomal analysis. The details of methods

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were previously described by Minamihisamatsu et al. (1986). Briefly, approximately 1–2 x 10^6 cells taken from exponentially growing tumours were inoculated at 37°C for 40 h in RPMI-1640 medium containing 20% fetal calf serum in the presence of pokeweed mitogen. Colcemid (0.01 μg ml^-1) was added for the last 12 h of the culture period. Thereafter, the cells were subjected to hypotonic treatment in 0.075 M KCl, expanded on the slide and then fixed in methanol/acetic acid (3:1). The slides were stained for Q-banding with a quinacrine mustard solution and for G-banding with Giemsa solution. Thirty metaphase cells were selected per case and analysed.

**Tumour growth and cell cycle parameters**

To obtain the growth curve, three perpendicular diameters of implanted tumours were measured weekly with a sliding calliper after every passage. Tumour volume (V) was calculated as a hemiellipsoid form according to V = 1/3 x length x width x height and plotted on semi-log graph paper against time. Volume doubling time (VDT) was measured from the exponential part of the growth curve. Single cell suspensions were obtained from a whole tumour by mincing it and pipetting the paste in the complete medium. A part of them was stained with 0.4% erythrocyte B to identify dead cells. For the determination of cell cycle parameters, another cell suspension was centrifuged on a Ficoll-Hypaque gradient. The separated cells were collected and adjusted to a cell number of 2 x 10^6. The cells were washed twice with chilled 0.1% sodium citrate and directly stained with chilled 50 μg ml^-1 propidium iodide (Sigma, St Louis, MO) dissolved in a solution of 0.1% sodium citrate and 0.1% Triton X-100 (Sigma). Subsequently the cells were incubated with 250 μg ml^-1 RNase (RNase A, Sigma) at 37°C for 30 min and were kept refrigerated for at least 2 h at 4°C. The cells were resuspended by gentle pipetting and subjected to flow cytometry (Epics C, Coulter). The percentages of cells in the G1, S and G2+M phases were calculated by assuming a Gaussian distribution (Krishan, 1975; Vindelov, 1977; Baisch et al., 1982).

Established tumour fragments in a medium containing 10% dimethylsulphoxide (DMSO) were cryopreserved in liquid nitrogen.

**Results**

Tumours or cells of 22 patients with non-Hodgkin's lymphomas were transplanted into nude mice. Tumour growth occurred in seven cases. One tumour was lost in first passage, but six tumours were serially transplanted and attained more than five passages, so that the six tumour lines (27%) were successfully established.

Table I shows the characteristics of the patients and their lymphomas from which the nude tumour lines were established. Immunological studies on clinical specimens revealed that four lymphomas showed B cell markers and one showed T cell markers. The E rosette setting was not applied to the remaining one, and immunoperoxidase studies of a paraffin-embedded specimen failed to find heavy and light chains.

To clarify further the nature of the six nude mouse lines, we compared the clinical data on the patients from whom nude mouse tumour lines were successfully established with the data on the others (Table II). Implanted follicular lymphomas showed no positive tumour take, while six nude mouse lines were established from 19 diffuse lymphomas.

With regard to B or T cell markers of diffuse lymphoma, the success rate in establishing tumour lines was 25% (1/4) for T lymphoma and 31% (5/16) for B lymphoma. Although T lymphomas were too small in number to be significant, it seems that nude mouse lines tend to be more easily established from B lymphoma than from T lymphoma.

On the other hand, it is noteworthy that the six established tumour lines were preferentially derived from the patients who were characterised by advanced clinical stage, high LDH value, large mass and poor response to chemotherapy (or short survival).

Table III summarises the cell markers of the six nude mouse lines. All of the B lymphoma lines showed immunoglobulins consisting of IgG (λ) and 34M (κ), positive B1, B20 and B4 (CD19) as pan-B markers, negative B2, B21 indicating limited B cell differentiation, and positive 1a-antigen OIA-1+ and or LeuHLA DR+. One T lymphoma line showed positive Leu-1 (CD5) and Leu-4 (CD3) as pan-T cell markers, Leu-2a (CD8) as a cytotoxic suppressor T cell marker, Leu-3a (CD4) as a helper/inducer T cell marker and OKT6 (CD1) indicating an origin from cortical thymocytes.

Compared with the clinical specimens from two B lymphomas showing negative immunoglobulin, the nude mouse counterparts began to express IgG (λ) clearly on the surface membrane and in the cytoplasm after establishment.

Tumour cells in the nude mouse lines showed normal diploid or near diploid karyotype. These results were obtained from 27–28 cells out of 30 examined. As shown in Table IV, the number of chromosomes was 44, 46, 49, 50 (one case each) and 47 (two cases), indicating human origin and clonal nature of the established lymphoma cells. The abnormalities included appearance of marker chromosomes, missing chromosomes and structural rearrangements, which were frequently observed in chromosomes 8, 12, 14 and 18. Among them, the alterations seen in chromosomes 14 (14q+ or 14q–) and 1 (1–1) are noteworthy (Figures 1, 2 and 3). The tumour growth curve of the six nude mouse tumour lines was approximated by a logistic function with some lag time. The lag time and volume doubling time shortened during the first to third passages. The stable VDT is given in Table V. The fraction of dead cells ranged from 44 to 14%. The cell populations participating in tumour growth were analysed by the flow-cytometric method. The average percentages of cells in G1, S and G2+M were 56.6 ± 4.0, 31.1 ± 3.1 and 12.3 ± 2.3 respectively. The high percentages of S and G2+M phases imply a high growth fraction in the nude mouse tumour lines.

**Discussion**

Compared with common cancers, leukaemia and lymphoma are most difficult to transplant into nude mice (Nara & Miyamoto, 1982). The poor transplantability is supposed to
be owing to the rich surface antigens of the neoplasms which induce a thymus-independent immune reaction. Various attempts to increase the take rate of lymphoma have been made by modifying the methods of heterotransplantation, e.g. by using an intracranial implant (Epstein et al., 1976), premedication with immunosuppressive drugs (Kopper et al., 1980; Habu et al., 1981), or anti-lymphocyte serum (Ohshgi et al., 1980) and whole-body irradiation with or without splenectomy (Watanabe et al., 1980; Morgan et al., 1978). Athymic and asplenic mice (Lasat mice) and newborn or young nude mice were used instead for the same purpose (Gershwin et al., 1978; Prehn & Outzen, 1977; Hanna & Fidler, 1981). The take rate from established haematological neoplasms in culture has been increased, but few successes in direct transplantation have been reported (Nilsson et al. 1977; Sordillo et al., 1983). In contrast, we succeeded in the establishment of six lymphoma lines from 22 cases (27%) after direct heterotransplantation into nude mice. We used 5 Gy whole-body gamma-ray irradiation for preconditioning of the nude mice. In the subsequent passages at least three mice were used, and the same preconditioning dose was given every time. The average percentage serial take was more than 77% throughout the passages. Among our cases no follicular lymphoma gave a positive take, while diffuse lymphoma showed a 32% establishment rate. This is much higher than that of Watanabe et al. (1980) who succeeded in establishing only two lines out of 13 diffuse lymphomas. This difference may be partly due to our procedures at subsequent passages. Lymphomas of high grade malignancy have been reported to show higher transplantability than those of low grade malignancy (Delso et al., 1980). Our histopathological results seem to support this. In addition, it was clearly observed that the established nude lines were preferentially derived from the primaries showing advanced clinical stage, high LDH value, large mass and poor prognosis.

All of the B cell nude mouse lymphomas were negative for

### Table II Relationship between patient’s clinical data and success or failure in establishing a nude mouse tumour line

| Pathology (working formulation) | Successful | Unsuccessful |
|---------------------------------|------------|-------------|
| Low grade | FSC (follicular small cleaved) | 0 | 1 |
| | FM (follicular mixed) | 0 | 2 |
| Intermediate grade | DSC (diffuse small cleaved) | 3 | 2 |
| | DM (diffuse mixed) | 1 | 1 |
| | DL (diffuse large) | 1 | 9 |
| High grade | LBL (lymphoblastic) | 1 | 0 |
| | MISC (miscellaneous) | 0 | 1 |
| Cell origin | B cell (16 cases) | 5° | 11 |
| | T cell (4 cases) | 1 | 3 |
| | non-T, non-B (2 cases) | 0 | 2 |
| Clinical stage | I = II | 0 | 5 |
| | III | 1 | 6 |
| | IV | 5 | 5 |
| Total | 6 (27%) | 16 |
| In diffuse lymphoma (19 cases) | LDH (on admission) > 400 units | 5 | 3 |
| | with large mass | 6 | 5 |
| | initial CR | 1 | 9 |
| Total | 6 | 13 |

*In one case there was no immunoglobulin in the primary tumour cells, but cells of the nude mouse line showed cytoplasmic IgG; *including mycosis fungoides which developed to cutaneous T cell lymphoma; *Normal LDH value in our hospital is under 400U (Cabaud-Wroblewski’s unit); *Our standard therapy was CHOP chemotherapy with or without local radiotherapy. Complete remission (CR) was judged by the complete disappearance of enlarged lymph nodes, palpable mass, or leukaemic lymphoma cells from peripheral blood and bone marrow, and clinical symptoms.

### Table III Immunophenotypic characteristics of the nude mouse lines

| E-rosetting | Immunoperoxidase |
|-------------|-----------------|
| Case 1 | sm/cigG (i) +, B1 (CD20) +, B2 (CD21) —, B4 (CD19) +, HLA-DR +, J5 (CD10) — |
| Case 2 | sm/cigM (k) +, B1 (CD20) +, B2 (CD21) —, B4 (CD19) +, OKJ4a +, J5 (CD10) — |
| Case 3 | clgM (k) +, B1 (CD20) +, B2 (CD21) —, B4 (CD19) +, OKJ4a +, J5 (CD10) — |
| Case 4 | OKT6 (CD1), Leu4 (CD3) +, Leu3a (CD4) +, Leu1 (CD5) +, Leu2a (CD8) + |
| Case 5 | sm/cigM (k) +, B1 (CD20) +, B2 (CD21) —, B4 (CD19) +, HLA-DR +, J5 (CD10) — |
| Case 6 | sm/cigG (i) +, B1 (CD20) +, B2 (CD21) —, B4 (CD19) +, OKJ4a +, J5 (CD10) — |

Positivity was judged as + for more than 70% cells, ± for 40–70% and — for less than 40%. Abbreviations: CD, cluster designation; sm/cig, surface membrane and cytoplasmic immunoglobulin; k, kappa; l, lambda.

### Table IV Cytogenetic characters of the nude mouse lines

| Passage | Cell karyotype |
|---------|---------------|
| Case 1 | p-26 49, X, −Y, −4, +7, +12, +13, +mar1, +mar2, 2q−, 4q+ |
| Case 2 | p-18 50, XY, −1, +1p−, +1q−, +18, +18, +mar. 2q+, 7p−, 12q+ |
| Case 3 | p-26 47, XX, −1, +1p−, +1q−, −10, +mar. 1p+, 6q−, 7p+, 11q−, 12q+, 14q+ |
| Case 4 | p-5 46, XX, 6q−, 7p+, 9q−, 14q− |
| Case 5 | p-17 44, X, −X, −8, −13, +mar, 6q−, 10p+, 14q+, iso(18q) |
| Case 6 | p-3 47, XX, −5, −6, −12, +mar1, +mar2, +mar3, +mar4, 7p− |

### Table V Growth characteristics of the nude mouse lines

| Volume doubling time (VDT, days) | % dead cell fractions | Mean % cells in the cell cycle (± s.d.) |
|-------------------------------|----------------------|---------------------------------------|
| Passage | G1 phase | S phase | G2 + M phase |
| Case 1 | 27 | 5.8 | 44.2 | 57.5 ± 2.6 | 28.8 ± 1.7 | 13.7 ± 3.2 |
| Case 2 | 20 | 8.1 | 42.6 | 63.5 ± 1.2 | 25.3 ± 1.7 | 11.2 ± 0.5 |
| Case 3 | 30 | 23.8 | 57.3 ± 6.7 | 34.4 ± 7.9 | 8.3 ± 1.3 |
| Case 4 | 17 | 3.9 | 51.6 ± 6.9 | 32.8 ± 3.2 | 15.6 ± 4.6 |
| Case 5 | 23 | 7.1 | 52.9 ± 4.4 | 33.0 ± 2.4 | 14.0 ± 4.5 |
| Case 6 | 4 | 7.8 | 57.0 ± 1.5 | 32.3 ± 2.1 | 10.7 ± 3.2 |
| Mean | 6.0 ± 2.0 | 29.2 | 56.0 ± 4.0 | 31.3 ± 3.1 | 12.3 ± 2.3 |

*The value of dead cell fraction was judged from % positive erythroblastic staining cells in cell suspensions which had been prepared from exponentially growing tumours.
B2 marker, but expressed immunoglobulin on the cell surface and in the cytoplasm, suggesting the terminal stage of B cell differentiation (Anderson et al., 1984). As mentioned in the Results section, two of the original lymphomas were not demonstrated to have immunoglobulin, but the established nude mouse counterparts expressed IgG (λ), indicating the activation of B cell differentiation including the change in heavy chain isotype. On the other hand, compared with the original cells, the T cell nude mouse lymphoma showed no changes in histology or immunological phenotype as far as we examined. The presence of the cellular antigens CD1, CD4, CD5 and CD8 supported the view that this line retained the fundamental characteristics of common thymocyte stage (Reinherz & Schlossman, 1980; Weiss et al., 1986).

As shown in Table IV, the karyotypes of all nude mouse lines were diploid, which indicated the retention of fundamental features of the human chromosome. Some nude tumour lines showed chromosomal changes such as 14q+ abnormality, which is thought to be specific for the development of B-cell lymphoma (Yunis, 1983; Levine et al., 1983), but it could not be determined whether the other abnormalities were specific for B-cell lymphoma or accidental alterations occurring during the serial passages.

The growth kinetics studies on the six nude mouse tumour lines revealed that they were characterised by short VDT with a high growth fraction. The results seem to reflect the high malignancy indicated in the clinical data of the patients with the original lymphoma. In a previous study, we measured the growth parameters of Burkitt lymphoma grown in nude mice in detail. The cell cycle time of the tumour was determined to be 66 h (100%), consisting of 31 h (54%) of G1, 28 h (36%) of S and 7 h (10%) of G2+M (Miymoto & Terashima, 1986). The numbers in parenthesis represent the fraction of cells at each cell cycle, and are similar to those of our nude mouse lines, as shown in Table V. It follows that the other growth parameters may be similar to those of Burkitt lymphoma. Based on the similarity, it can be inferred that almost all the lymphomas were proliferating with a VDT of less than one week.

As indicated in Table II, the original lymphomas were poorly responsive to chemotherapy, including CHOP and regional radiation. Experimental chemotherapy of these nude mouse lymphomas would demonstrate whether the resistance is caused by the rapid cell proliferation or is intrinsic.

The established lymphoma lines reported here are expected to be useful in biological and oncological studies of malignant lymphoma in the future.

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Figure 1 Cell karyotype of case 2.

Figure 2 Cell karyotype of case 3.

Figure 3 Cell karyotype of case 5.
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