 Genetic Analysis and QTL Detection on Fiber Traits Using Two Recombinant Inbred Lines and Their Backcross Populations in Upland Cotton

Lianguang Shang,* Yumei Wang,† Xiaocui Wang,* Fang Liu,* Abdugheni Abduweli,* Shihu Cai,* Yuhua Li,* Lingling Ma,* Kunbo Wang,‡ and Jinping Hua*,1

*Department of Plant Genetics and Breeding/Key Laboratory of Crop Heterosis and Utilization of Ministry of Education/ Beijing Key Laboratory of Crop Genetic Improvement, China Agricultural University, Beijing 100193, China, †Institute of Cash Crops, Hubei Academy of Agricultural Sciences, Wuhan 430064, China, and ‡Institute of Cotton Research, Chinese Academy of Agricultural Sciences/State Key Laboratory of Cotton Biology, Anyang 455000, Henan, China

ABSTRACT Cotton fiber, a raw natural fiber material, is widely used in the textile industry. Understanding the genetic mechanism of fiber traits is helpful for fiber quality improvement. In the present study, the genetic basis of fiber quality traits was explored using two recombinant inbred lines (RILs) and corresponding backcross (BC) populations under multiple environments in Upland cotton based on marker analysis. In backcross populations, no significant correlation was observed between marker heterozygosity and fiber quality performance and it suggested that heterozygosity was not always necessarily advantageous for the high fiber quality. In two hybrids, 111 quantitative trait loci (QTL) for fiber quality were detected using composite interval mapping, in which 62 new stable QTL were simultaneously identified in more than one environment or population. QTL detected at the single-locus level mainly showed additive effect. In addition, a total of 286 digenic interactions (E-QTL) and their environmental interactions [QTL × environment interactions (QEs)] were detected for fiber quality traits by inclusive composite interval mapping. QE effects should be considered in molecular marker-assisted selection breeding. On average, the E-QTL explained a larger proportion of the phenotypic variation than the main-effect QTL did. It is concluded that the additive effect of single-locus and epistasis with few detectable main effects play an important role in controlling fiber quality traits in Upland cotton.

KEYWORDS fiber quality QTL recombinant inbred line backcross population Upland cotton

Cotton is an important cash crop providing most of the natural fiber for the textile industry. Upland cotton (Gossypium hirsutum L.) accounts for 95% of the total production of cotton fiber in the world (Chen et al. 2007). Cotton fiber is widely employed in the textile industry. With the development of spinning technology, the improvement of fiber quality is becoming highly crucial in Upland cotton (Kohel et al. 2001). However, a major problem for cotton breeding is that fiber quality has a negative genetic correlation with cotton yield. The advent of molecular markers made it possible for cotton breeders to improve yield and fiber quality traits in cotton (Paterson et al. 1988). Previous studies have shown that fiber quality traits were quantitative traits and were affected by genetic background and the specific growing environment. These studies also observed that epistatic effects and genotype × environment (GE) interaction effects played an important role in the genetic basis of fiber traits (Paterson et al. 2003; Shen et al. 2006). Understanding of the genetic basis of both yield and fiber quality traits is vital for cotton breeding.

Detection of stable quantitative trait loci (QTL) for fiber quality is essential for developing cotton cultivars with superior fiber quality using molecular marker-assisted selection (MAS) strategy. A large number of studies on mapping main agronomic characters can be found in the CottonGen database (Yu et al. 2014). Hundreds of QTL conditioning...
fibre traits were obtained based on intraspecific genetic populations in Upland cotton (Said et al. 2015). These fibre QTL identified using low density genetic maps have very limited value in MAS programs. Fortunately, publically available sequences of the D9 genome of G. raimondii, A2 genome of G. arboreum, AD genome of G. hirsutum, and AD2 genome of G. barbadense provide an opportunity to improve the density of intraspecific genetic maps (Paterson et al. 2012; Wang et al. 2012; F. Li et al. 2014, 2015; Yuan et al. 2015; T. Zhang et al. 2015). Recently, a population of 178 recombinant inbred lines (RILs) was developed from a cross between G. hirsutum acc ‘DH962’ and G. hirsutum cv ‘jimian5’. A total of 644 polymorphic loci were employed to construct a genetic linkage map, and a total of 64 QTL associated with fibre qualities were identified in seven environments (Wang et al. 2015a). Z. Zhang et al. (2015) developed a population of 196 RILs from a cross between ‘0-153’ and ‘sGK9708’ and detected a total of 37 QTL for fibre quality traits on chromosome 25, of which 17 were stably identified under at least two environments. Wang et al. (2015b) constructed an intraspecific genetic map of Upland cotton containing 1013 loci by developing markers using parental restriction site-associated DNA sequencing. And 27 new QTL for yield and fibre quality were identified, suggesting that the efficiency of QTL identification is greatly improved by the increase in genetic map density. Islam et al. (2016) validated three QTL regions controlling three fibre quality traits and further fine-mapped with 27 new single nucleotide polymorphism markers. The limitations of traditional breeding can be alleviated using an Upland cotton intraspecific high density genetic map to identify fibre QTL under multiple environments and further to conduct molecular marker-assisted breeding.

In our previous studies, two RIL populations and two corresponding backcross populations were applied to elucidate the genetic basis of oil content, seed index, and yield heterosis in Upland cotton (Shang et al. 2016a,b). In our laboratory, 39 QTL for fibre quality were identified in three generations (four environments) of F2, F2:3, and F2:4 populations derived from cross ‘GX1135’ × ‘GX100-2’ (Liang et al. 2013). Recently, 20 QTL associated with fibre quality traits were detected using 581 loci and a separate RIL population derived from a cross ‘GX1135’ × ‘GX100-2’. Among 20 QTL, four QTL were again detected, verifying the previous results in F2, F2:3, and F2:4 populations (Shang et al. 2015a). In the present research, two RIL populations and their backcross progeny derived from two hybrids were used to further detect the stable QTL for fibre quality traits in Upland cotton. Previous reported results showed that most of the QTL controlling fibre quality traits were identified only in single mapping populations, such as F2:3 (Mei et al. 2004; Liang et al. 2013) and RIL (Wang et al. 2015a; Shang et al. 2015a; Z. Zhang et al. 2015). There are few reports available of fibre quality QTL using multiple genetic populations in multiple environments. The objective of the present study is to detect stable QTL and conduct the genetic analysis of fibre quality traits by the single-locus and two-locus analysis using two RIL populations and their backcross progeny in Upland cotton. This study will provide new insights into the genetic basis of fibre quality and make contributions to improve fibre quality in Upland cotton.

MATERIALS AND METHODS

Plant materials and population construction

As described previously (Shang et al. 2016a,b), two hybrids were used in the present research. One is ‘Xinza 1’ (Liang et al. 2013, 2015; Shang et al. 2015a,b, 2016c; hereinafter referred to as the ‘XZV hybrid’), derived from ‘GX1135’ × ‘GX100-2’. The other one has a common female parent with ‘Xinza 1’, derived from ‘GX1135’ × ‘VGX100-2’ (Shang et al. 2016a,b; hereinafter referred to as the ‘XZ hybrid’).

In total, four populations were employed: (1) the RIL population of the XZ hybrid; (2) another RIL population (RILV) from the XZV hybrid; (3) a backcross population (BC) of the XZ hybrid (the BC population included 177 BCF1 hybrids, and each BCF1 hybrid was from a cross where one F9 RIL was used as the female parent and the common parent GX1135 was used as the male parent, respectively); and (4) another backcross population (BCV) of the XZV hybrid. One hundred and eighty BCF1 hybrids were developed from crosses between RILs from the F9 RILV population used as the female parent and the common parent GX1135 used as the male parent, respectively (Shang et al. 2016a,b).

For ease of description, we refer to the RIL(V) in BC(V) population as the RIL(V)’ population, respectively. In the BC(V) population, six-row plots were set which included the BCF1 hybrids RIL(V)’ × GX1135 in the middle, and its corresponding female RIL(V)’ and the recurrent parent, GX1135. Each line in the RIL(V)’ population was used as the female parent in the BC(V) population and was the same as that in the RIL(V) population. In BCF1 population experiments of population 3 (BC) and population 4 (BCV), each plot consisted of two rows of the female RIL(V)’, BCF1 hybrids and GX1135, respectively.

In addition, two special plots, each consisting of two rows of the XZ hybrid F1, and its parents, respectively, were used as controls in the experiment of population 2 and 4, and each plot consisted of the XZV hybrid F1, and its parents.

Field trials and phenotypic evaluation

The parents together with the four populations were evaluated in three environments in China (Shang et al. 2015a): E1 – Quzhou Experimental Station of the China Agricultural University, Handan, Hebei Province; E2 – Guoxin Seed Company, Limited, Cangzhou, Hebei Province; and E3 – Xiangyang Academy of Agricultural Sciences, Xiangyang, Hubei Province. The field planting followed a randomized complete block design with duplicate at each location in 2012. Two-row plots were 80 cm and 50 cm row spacing alternately for the experiment in E1 and E2, and two-row plots in E3 were 100 cm and 80 cm row spacing alternately. The length of plots was 4 m in E1, and 3 m in E2 and E3. In the experiment for populations 1 and 2, two-row plots with each line were used. In the experiment for populations 3 and 4, six-row plots with each plot consisting of two rows of BCF1, hybrid [RIL(V) × GX1135], and two for each of the corresponding parents: the female RIL(V)’ and GX1135. Field management followed the local conventional standard field practices (Shang et al. 2016a,b).

Thirty open bolls from each plot were sampled by hand in three sites, respectively. Fiber quality traits were measured with an HVI 900 instrument (USTER HVISPECTRUM, SPINLAB) at the Cotton Fiber Quality Inspection and Test Center of the Ministry of Agriculture (Anyang, China). The fiber quality traits included 2.5% fiber span length (mm), fiber uniformity (%), fiber strength (cN/tex), fiber elongation, and micronaire.

Genotype analysis

Extraction of individual genomic DNA and population genotype analysis were carried out following the methods of Liang et al. (2013). A total of 48,836 pairs of SSR primer were employed to screen polymorphic loci between three parents (Shang et al. 2016a). The SSR primers newly added in this study included the EST-SSR primers, named as CAU primers, developed from the salt-tolerance EST sequences, SWU and PGML primers developed from the G. raimondii genome sequence, and ICR primers developed from the G. arboreum genome sequence. In total, 653 polymorphic loci for the XZ hybrid and 400 polymorphic
loci for the XZV hybrid were used to conduct genotype analysis of two RIL populations (Shang et al. 2016a,b). The genotype for each BCF1 was deduced on the basis of the RIL genotype used as the parent for the cross.

### Data analysis

A basic statistical analysis was implemented using the software SPSS version 19.0 (SPSS, Chicago). The software MAPMAKER 3.0 was employed to construct a genetic linkage map with the Kosambi function (Lander et al. 1987). For the RIL(V) and RIL(V)F1 populations, the means from two replications were used as raw data at each location. For each of the BC(V)F1 populations, the means of the BC(V)F1 population were used independently as raw data at three locations. Single-locus QTL was conducted using composite interval mapping by the software WinQTL Cartographer 2.5 in RIL(V), RIL(V), and BC(V)F1 data (Zeng 1994; Wang et al. 2005). A LOD threshold of 3.0 was used to declare suggestive QTL, whereas the same QTL in another environment or population with LOD of at least 2.0 was considered to be a common QTL (Shang et al. 2015a). The graphic representation of the linkage group was created using the software Map Chart 2.2 (Voorrips 2002). QTL nomenclature used in rice was employed in the present study (McCouch et al. 1991). Two-locus analysis that tests the main-effect QTL (M-QTL), and digenic epistatic QTL (E-QTL) and their environmental interactions (QTL × environment, QE), was conducted using the software ICIMapping 4.0 (www.isbreeding.net). LOD thresholds were respectively set at 2.5 and 5.0 for declaring the presence of M-QTL, E-QTL, and their QEs (S. Li et al. 2015).

### Data availability

All raw data are available as Supporting Information Table S9 and Table S10, which include genotypes and traits of two hybrids.

## RESULTS

### Performance of fiber quality traits

The means of phenotypic data for fiber quality traits of two hybrids in three environments are shown in Table 1. Not all the fiber quality traits possessed higher phenotype values in heterozygotes (BCF1s) than in respective homozygotes (RILs). Most of the extreme lines in the RIL (V) populations exceeded those of BC(V)F1 populations under different environments. The analysis of variance in RIL(V) and BC(V) populations was conducted and significant genotypic variances and environment variances for most of the fiber quality traits were found in four populations (Table 2). Significant variations for fiber quality traits are observed in two hybrids. Skewness and kurtosis values were calculated, and results showed that all fiber quality traits fit a normal distribution in two RIL and two backcross populations of two hybrids (data not shown). The BCF1 population had higher means for most of the fiber quality traits than the RIL population in the two hybrids. These results suggested that fiber quality traits of RIL(V) and BC(V) populations were highly variable and conducive for QTL analysis.

### Correlation analysis among fiber quality traits

Correlation analysis was carried out using the mean values of three environments, respectively (Table 3). The majority of fiber quality traits were significantly associated with each other in two hybrids. Fiber length is significantly positively correlated with fiber strength and fiber elongation, but was negatively correlated with micronaire. Fiber strength was significantly positively correlated with fiber elongation, but it was negatively correlated with micronaire. Fiber uniformity was significantly positively correlated with fiber elongation. These results were consistent with a previous report (Shang et al. 2015a). The correlation coefficients among the mean values of RILs and their BCF1s for fiber quality traits are shown in Supplemental Material, Table S1. Most of the fiber trait values of the RILs and that of their BCF1s showed significant positive correlation. Population performance of the BCF1 for most of the fiber traits was largely determined by performance of the parental RIL.

### QTL analysis for fiber quality traits at the single-locus level

Two genetic linkage maps were previously constructed based on the polymorphic loci identified in two hybrids (Shang et al. 2016a,b). For the XZ hybrid, the genetic map with 623 loci spanned 3889.9 cM. For the XZV hybrid, the genetic map with 308 loci spanned 3048.4 cM. QTL detected using composite interval mapping for fiber quality traits in XZ and XZV hybrids are shown in Table S2 and Figure S1. A total of 71 and 40 QTL were detected for fiber quality traits in RIL(V), RIL(V), and BC(V)F1 data sets of XZ and XZV hybrids in three environments, respectively. The genetic effect identified in the RIL population is generally larger than in the backcross population. These results suggested that heterozygosity was not always necessarily advantageous for the expression of the fiber quality traits.

In the XZ hybrid, a total of 17 QTL were detected for fiber length (FL) in three data sets, among which 13, 12, and 9 QTL were identified for FL

## Table 1 Summary statistics on fiber quality traits in two hybrids

| Trait               | Mean RIL | Mean BC | Min RIL | Min BC | Max RIL | Max BC | Parents | $\varphi_0$ | $\alpha$ |
|---------------------|----------|---------|---------|--------|---------|--------|---------|-------------|----------|
| **XZ hybrid**       |          |         |         |        |         |        |         |             |          |
| Fiber length (mm)   | 28.48    | 28.52   | 25.96   | 26.74  | 31.01   | 30.49  | 28.72   | 27.78       |          |
| Fiber uniformity    | 84.63    | 84.88   | 81.82   | 82.22  | 86.78   | 87.08  | 84.28   | 85.00       |          |
| Fiber strength (cN/tex) | 29.12   | 29.29   | 25.55   | 27.13  | 32.38   | 31.78  | 29.75   | 27.90       |          |
| Fiber elongation    | 6.91     | 6.77    | 6.43    | 6.50   | 7.28    | 7.00   | 6.83    | 6.68        |          |
| Micronaire          | 4.61     | 4.68    | 3.59    | 4.08   | 5.42    | 5.25   | 4.76    | 4.74        |          |
| **XZV hybrid**      |          |         |         |        |         |        |         |             |          |
| Fiber length (mm)   | 28.07    | 28.60   | 24.81   | 26.09  | 31.86   | 31.38  | 28.72   | 28.47       |          |
| Fiber uniformity    | 84.45    | 85.25   | 81.40   | 82.02  | 86.50   | 87.25  | 84.28   | 83.97       |          |
| Fiber strength (cN/tex) | 29.75   | 30.18   | 25.62   | 27.33  | 33.80   | 32.77  | 29.75   | 30.14       |          |
| Fiber elongation    | 6.72     | 6.67    | 6.30    | 6.40   | 7.10    | 6.93   | 6.83    | 6.73        |          |
| Micronaire          | 4.39     | 4.28    | 3.27    | 3.64   | 5.31    | 4.86   | 4.76    | 4.11        |          |
Table 2 The results of analysis of variance (ANOVA) of fiber quality traits

| Trait              | Source of Variation | RIL       | BCF1      | RILV      | BCVF1     |
|--------------------|---------------------|-----------|-----------|-----------|-----------|
| Fiber length       | G                   | 4.43**    | 1.73**    | 9.44**    | 2.71**    |
|                    | E                   | 1259.77** | 998.88**  | 686.68**  | 719.22**  |
|                    | G × E               | 0.88**    | 0.72      | 1.19**    | 1.00      |
|                    | e                   | 0.67      | 0.82      | 0.90      | 0.91      |
| Fiber uniformity   | G                   | 1.91*     | 1.85*     | 2.65**    | 1.44      |
|                    | E                   | 967.44**  | 839.95**  | 1084.62** | 838.20**  |
|                    | G × E               | 1.61      | 1.44      | 1.47      | 1.79      |
|                    | e                   | 1.46      | 1.45      | 1.39      | 1.71      |
| Fiber strength     | G                   | 6.26**    | 2.16**    | 10.76**   | 3.36**    |
|                    | E                   | 775.17**  | 119.33**  | 38.46**   | 65.35**   |
|                    | G × E               | 1.33      | 1.25      | 2.58**    | 1.43      |
|                    | e                   | 1.22      | 1.37      | 1.67      | 1.57      |
| Fiber elongation   | G                   | 0.07**    | 0.02      | 0.08**    | 0.02      |
|                    | E                   | 0.09*     | 6.03**    | 1.21**    | 21.12**   |
|                    | G × E               | 0.03*     | 0.02      | 0.04*     | 0.02      |
|                    | e                   | 0.02      | 0.03      | 0.03      | 0.02      |
| Micronaire         | G                   | 0.44**    | 0.15**    | 0.57**    | 0.19**    |
|                    | E                   | 35.80**   | 66.45**   | 59.14**   | 100.40**  |
|                    | G × E               | 0.11**    | 0.07      | 0.10      | 0.08      |
|                    | e                   | 0.07      | 0.06      | 0.08      | 0.07      |

* P = 0.05, **P = 0.01. G, genotype; E, environment; e, error MS, mean square.

Table 3 Correlations between fiber quality traits of RIL and backcross populations in two hybrids

| Trait          | Fiber Length | Fiber Uniformity | Fiber Strength | Fiber Elongation |
|----------------|--------------|------------------|----------------|-----------------|
|                | Env.         | RIL   | BC   | RIL   | BC   | RIL   | BC   | RIL   | BC   |
| XZ hybrid      |              |       |      |       |      |       |      |       |      |
| Fiber uniformity | E1           | 0.27** | 0.28** |       |      |       |      |       |      |
|                | E2           | 0.05   | 0.04  |       |      |       |      |       |      |
|                | E3           | 0.15   | -0.05 |       |      |       |      |       |      |
| Fiber strength | E1           | 0.64** | 0.51** | 0.21** | 0.21** |       |      |       |      |
|                | E2           | 0.76** | 0.58  | 0.10  | 0.05  |       |      |       |      |
|                | E3           | 0.58** | 0.38** | 0.32** | 0.17** |       |      |       |      |
| Fiber elongation | E1         | 0.60** | 0.57** | 0.15** | 0.63** | 0.64** |       |       |      |
|                | E2           |       |      |       |      |       |      |       |      |
|                | E3           | 0.52** | 0.43** | 0.17** | 0.18** | 0.44** | 0.59** |       |      |
| Micronaire     | E1           | -0.27** | -0.09 | 0.33** | 0.11 | -0.22** | -0.25** | 0.04 | 0.07 |
|                | E2           | -0.25** | -0.12 | -0.04 | 0.13 | -0.15** | -0.13 | —     | —    |
|                | E3           | -0.48** | -0.34** | -0.09 | 0.1 | -0.34** | -0.16** | 0.03 | 0.08 |
| XZV hybrid     |              |       |      |       |      |       |      |       |      |
| Fiber uniformity | E1          | 0.30** | 0.37** |       |      |       |      |       |      |
|                | E2           | 0.17   | 0     |       |      |       |      |       |      |
|                | E3           | 0.12   | -0.01 |       |      |       |      |       |      |
| Fiber strength | E1           | 0.65** | 0.40** | 0.42** | 0.33** |       |      |       |      |
|                | E2           | 0.73   | 0.65  | 0.31  | 0.14  |       |      |       |      |
|                | E3           | 0.53** | 0.32** | 0.27** | 0.14  |       |      |       |      |
| Fiber elongation | E1         | 0.68** | 0.49** | 0.33** | 0.29** | 0.77** | 0.78** |       |      |
|                | E2           |       |      |       |      |       |      |       |      |
|                | E3           | 0.49** | -0.58** | 0.11  | -0.02 | 0.60** | 0.43** |       |      |
| Micronaire     | E1           | -0.31** | -0.05 | 0.12  | 0.23** | -0.07  | 0.08  | 0.16* | 0.18* |
|                | E2           | -0.18  | -0.18 | -0.02 | -0.03 | -0.12  | -0.16 | —     | —    |
|                | E3           | -0.57** | -0.34** | 0     | 0.11  | -0.33** | -0.40** | 0.01 | 0.04 |

* and ** indicate that the correlation is significant at 0.05 and 0.01 probability levels, respectively. — indicates missing data. Env., environment; E1, Handan; E2, Cangzhou; E3, Xiangyang.
populations. Two QTL, qFL-Chr2-2 and qFL-Chr14-1, with the same genetic direction were identified simultaneously in three environments and populations.

For fiber uniformity (FU), in the XZ hybrid, a total of six QTL were resolved explaining from 4.33% to 14.18% of phenotypic variance (PV). Two QTL were identified in more than two environments or populations. In the XZV hybrid, three QTL were detected and one QTL was identified in two populations.

For fiber strength (FS), in the XZ hybrid, a total of 15 QTL were detected in three data sets, among which 11, eight, and six QTL were respectively identified in the RIL’s, RILVs, and BCVF1 hybrids data. Eleven QTL were identified in more than two environments or populations. In the XZV hybrid, a total of eight QTL were detected, among which two, six, and five QTL were identified in the RILV’s, RILVs, and BCVF1 hybrids data, respectively. Four QTL were identified in more than two environments or populations, in which qFS-Chr26-1 with negative genetic effect was stably expressed in multitude environments and populations.

For fiber elongation (FE), in the XZ hybrid, a total of 11 QTL were detected in three data sets, among which five, four, and six QTL were identified in the RIL’s, RILVs, and BCVF1 data, respectively. Three QTL were identified in more than two environments or populations. In the XZV hybrid, a total of six QTL were detected, among which three, five, and one QTL were respectively identified in the RILV’s, RILVs, and BCVF1 data. Two QTL were identified in more than two environments or populations.

For fiber micronaire (FM), in the XZ hybrid, a total of 21 QTL were resolved explaining from 3.76% to 25.93% of PV. Eleven QTL were identified in more than two environments or populations, among which two stable, QTL qFM-Chr2-1 and qFM-Chr19-1, were simultaneously observed in two environments and three populations. In the XZV hybrid, 13 QTL were detected and seven QTL were identified in more than two environments or populations, among which one major QTL, qFM-Chr14-1, was observed in all of the environments and populations explaining from 3.97% to 14.06% of PV.

QTL and QE interactions resolved by two-locus analyses
A total of 98 and 88 M-QTL and QEs were respectively detected by inclusive composite interval mapping (ICIM) in five fiber quality traits of XZ and XZV hybrids (Table 4, Table S3, and Table S4). In the XZ hybrid, a total of 65 and 33 M-QTL and QEs were detected in the RILs and BCVF1 hybrids data, respectively. On average, M-QTL explained 2.35% and 2.14% of the phenotype variation, and the QE explained 0.74% and 0.86% of the phenotype variation in the RILs and BCVF1 hybrids data, respectively. In the XZV hybrid, a total of 56 and 32 M-QTL and QEs were detected in the RILVs and BCVF1 hybrids, respectively. On average, M-QTL explained 2.73% and 1.77% of the phenotype variation, and the QE explained 0.43% and 0.92% of the phenotype variation in the RILVs and BCVF1 hybrids data, respectively.

In total, 157 and 129 E-QTL and QEs were detected by ICIM in five fiber quality data sets of XZ and XZV hybrids, respectively (Table 4, Table S5, and Table S6). In the XZ hybrid, a total of 90 and 67 E-QTL and QEs were detected in the RILs and BCVF1 hybrids data, respectively. On average, E-QTL explained 3.40% and 2.25% of the phenotype variation, and the QE explained 0.61% and 1.90% of the phenotype variation in the RILs and BCVF1 hybrids data, respectively. In the XZV hybrid, a total of 101 and 28 E-QTL and QEs were detected in the RILVs and BCVF1 hybrids data, respectively. On average, E-QTL explained 4.04% and 2.85% of the phenotype variation, and the QE explained 0.38% and 0.64% of the phenotype variation in the RILVs and BCVF1 hybrids data, respectively.

DISCUSSION

Advantage of permanent RIL and BC1 populations design
Fiber quality traits are quantitatively inherited, and the QTL detected tend to vary under different environments. In order to identify more stable and convincing QTL in more than one environment, permanent populations such as RILs are required. Permanent populations possessing heterozygotes could be applied to study the genetic basis of complex traits using QTL mapping strategy. For example, BC1 populations based on RIL population were previously constructed and used to conduct QTL analysis referring to complex agronomic traits (Mei et al. 2005; You et al. 2006, Jiang et al. 2014; Shang et al. 2016). In this study, we constructed the BC1 population using the RIL population to identify QTL contributing to fiber quality traits and estimate their genetic effects in Upland cotton. Several QTL that were not identified in the RIL population can be detected in the BC1 population, for example,

| Table 4 Summary of M-QTL and E-QTL detected controlling fiber quality traits by ICIM in two hybrids |
|---------------------------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|
| Trait                          | M-QTL           | E-QTL           | RIL             | BCF1            | RIL             | BCF1            |
|--------------------------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|
|                                | n   | P(A)  | P(AE) | n   | P(A)  | P(AE) | n   | P(A)  | P(AE) | n   | P(A)  | P(AE) |
| Fiber length                   | 12  | 2.67  | 0.58  | 8   | 2.17  | 0.72  | 18  | 2.94  | 0.13  | 7   | 2.18  | 0.59  |
| Fiber uniformity               | 8   | 1.06  | 1.20  | 8   | 1.35  | 1.12  | 6   | 0.31  | 1.74  |
| Fiber strength                 | 21  | 2.41  | 0.41  | 7   | 2.20  | 0.76  | 9   | 3.20  | 0.42  | 5   | 2.83  | 0.56  |
| Fiber elongation               | 8   | 2.80  | 1.17  | 2   | 3.17  | 0.86  | 6   | 2.17  | 0.90  |
| Micronaire                     | 16  | 2.79  | 0.36  | 7   | 1.80  | 0.87  | 23  | 2.61  | 0.25  | 14  | 1.78  | 0.78  |
| Mean                           | 13  | 2.35  | 0.75  | 6.6 | 2.14  | 0.86  | 11.2| 2.73  | 0.43  | 6.4 | 1.77  | 0.92  |
|                                | n   | P(AA) | P(AAE)| n   | P(AA) | P(AAE)| n   | P(AA) | P(AAE)| n   | P(AA) | P(AAE)|
| Fiber length                   | 20  | 3.38  | 0.32  | 16  | 2.97  | 1.23  | 34  | 4.13  | 0.17  | 13  | 2.89  | 0.60  |
| Fiber uniformity               | 6   | 2.28  | 1.53  | 5   | 1.76  | 1.74  | 6   | 2.69  | 0.90  |
| Fiber strength                 | 14  | 2.98  | 0.32  | 7   | 1.98  | 1.31  | 25  | 3.30  | 0.42  | 7   | 3.13  | 0.47  |
| Fiber elongation               | 6   | 4.65  | 0.46  | 1   | 1.82  | 4.27  | 1   | 6.43  | 0.24  |
| Micronaire                     | 44  | 3.69  | 0.46  | 38  | 2.74  | 0.96  | 35  | 3.66  | 0.15  | 8   | 2.52  | 0.85  |
| Mean                           | 18  | 3.40  | 0.61  | 13.4| 2.25  | 1.90  | 20.2| 4.04  | 0.38  | 5.6 | 2.85  | 0.64  |

n, the number of QTL identified. P (in %) was the mean of trait PVs explained by a single M-QTL or E-QTL.
qFL-Chr2-2 was identified only in the BCF1 population in the XZ hybrid. Moreover, the QTL identified using the RIL population could be verified using the BCF1 population in the current study. For instance, the QTL qFM-Chr19-1 identified in RIL populations was confirmed in the BCF1 population in both environments again in the XZ hybrid. Furthermore, the QTL detected in the XZ hybrid could be verified in the XZV hybrid in that two hybrids shared one parent (Shang et al. 2016b). The advantages of the permanent BC1 population are that it can be repeatedly made by RILs and the genotype of the BC1 population can be easily deduced by the genotype of the RIL population. In addition, the QTL actions can be inferred by comparing the genetic effects of RIL, BC1 performance, and midparental heterosis (Mei et al. 2005). However, the disadvantage of the BC1 population is that only half of the possible heterozygous loci are available (Radoev et al. 2008). It may be the reason why some QTL with dominant effect are omitted using BCF1 data rather than F2 population data.

**Consensus QTL and improvement of fiber quality**

Two RIL populations and two corresponding backcross populations were developed and used to detect stable QTL in the present research. In a previous study, 19 QTL including eight for FL, three for FS, four for FE, and four for FM were detected in the RIL population derived from Upland cotton cross ‘GX1135’ × ‘GX100-2’ (Table S7). These 19 QTL for fiber traits detected previously were once again identified in the current study. In XZ and XZV hybrids, 62 stable QTL were identified in more than one environment or population. A stable QTL, qFS-Chr26-1, flanked by HAU1571 and PGML2562 was detected in RILV, RILV, and BCV populations in three environments, and this QTL could contribute to 6.07–15.03% of the phenotypic variation. Another stable QTL, qFM-Chr14-1, was detected in RILV, RILV, and BCV populations, explaining the phenotypic variation by 3.97–14.06%, respectively. The single-locus QTL identified by composite interval mapping and the M-QTL from ICIM were compared. A total of 59 common QTL were found, and common QTL are shown as blue figures in Table S2. In addition, QTL mapping for fiber quality using the overall means across three environments (joint analysis) were conducted, and a total of 53 QTL for fiber quality traits were identified (Table S8). Of 53 QTL, 44 QTL detected using joint analysis were the same as the QTL with a single environment. These novel stable QTL for fiber quality traits identified using multiple populations and environments will be helpful to improve fiber quality in the future. We compared the QTL detected in the current study with QTL identified in other studies, however the same QTL were not found. It is difficult to search for common QTL, because the genetic map, population types, population structure, and environmental conditions vary and affect the comparison of common QTL (Shang et al. 2015a).

The result of two hybrids can be compared and verified using the common markers located in two connected genetic maps. Two consistent QTL were acquired in both hybrid populations. A good example is the QTL qFL-Chr2-1 for FL on chromosome 2 in the XZV hybrid; furthermore, we also identified another two QTL, qFL-Chr2-1 and qFL-Chr2-2, for FL in the XZ hybrid. These two QTL identified in the XZ hybrid were located in the same marker region as mentioned above and had a narrower interval of flanking markers than that in the XZV hybrid. Therefore, using connected genetic populations may have increased the number of common QTL identified and improved the accuracy of QTL location (Blanc et al. 2006).

It is difficult to simultaneously improve yield and fiber quality in Upland cotton breeding programs (Liang et al. 2013). In the present study, QTL that were located on chromosome five possessed multiple effects on yield and fiber quality. Two stable QTL with flanking markers between SWU20917 and NAU6240 associated with FL and FS shared a common marker interval with the QTL referring to the boll weight and lint percentage (Shang et al. 2016b). These stable QTL simultaneously controlled yield and fiber quality with a large contribution to the phenotypic variation and provided valuable information for pyramiding elite genes of yield and fiber quality traits. Further study should be conducted to prove the effects of these QTL in the improvement of yield and fiber quality in Upland cotton.

**Genetic basis of fiber quality traits and QTL × environment interaction**

The genetic basis of fiber quality traits is explored using two connected RIL populations and two corresponding BCF1 populations at the single-locus and two-locus levels in Upland cotton. The number and phenotypic variation of QTL identified using composite interval mapping in the RIL population were collectively larger than those QTL detected in the BCF1 population in the XZ and XZV hybrids. This suggested that the QTL controlling fiber quality traits mainly showed an additive effect at the single-locus level.

At the two-locus level, lots of digenic interactions and QEs resolved by ICIM were acquired in two hybrids. The epistasis detected had been previously classified into three types: (I) two loci with significant M-QTL, (II) one locus with M-QTL and another locus without significant M-QTL, and (III) two loci without significant M-QTL (Li et al. 2001; Shang et al. 2016d). In the XZ and XZV hybrids, we found that 39 of 301 (12.96%) epistatic interactions were type II, and the remaining 262 (87.04%) were type III for fiber quality traits. No type I interactions were acquired (Table 5). The fact that the digenic interaction mainly included loci without detectable M-QTL further confirmed the importance of epistasis in Upland cotton breeding (Shen et al. 2006). In addition, the number of E-QTL and the mean of PVs explained by E-QTL for most of the fiber traits are much greater than that for M-QTL in two hybrids (Table 4). These results revealed that epistasis played an important role not only in the variation of the performance of the RIL(V) population but also in the expression of hybrids in the BC (V) population. These results are in agreement with previous results which indicated that additive effect and epistatic interaction are common and important for fiber quality traits in Upland cotton. Selection

| Type of Epistasis | I | II | III | Sum |
|------------------|---|----|-----|-----|
| Fiber length     |  RIL | 0 | 0 | 16 | 31 |
|                  |  BC  | 0 | 0 | 16 | 16 |
| Fiber uniformity |  RIL | 0 | 0 | 5  | 6  |
|                  |  BC  | 0 | 0 | 5  | 5  |
| Fiber strength   |  RIL | 0 | 2 | 12 | 23 |
|                  |  BC  | 0 | 3 | 13 | 16 |
| Fiber elongation |  RIL | 0 | 0 | 5  | 5  |
|                  |  BC  | 0 | 0 | 1  | 1  |
| Micronaire       |  RIL | 0 | 8 | 36 | 54 |
|                  |  BC  | 0 | 4 | 34 | 38 |
| Sum              |  0  | 0 | 19| 147| 155|

Type of epistasis: (I) two loci with main-effect QTL, (II) a locus with main-effect QTL and a locus without significant main-effect QTL, and (III) two loci without significant main-effect QTL. Sum, total number of epistatic interactions.
for improving trait values should pay attention to the best multilocus combinations and major loci (Shen et al. 2006).

The genotype × environment interactions accounted for a small proportion of the mean phenotypic variation for fiber traits (Table 4). The QTL for fiber quality in the BC(V) population was more sensitive to the environment than that in the RIL(V) population as shown by the mean phenotypic variation explained by QEs. Particularly environmental conditions were important in the expression of fiber quality, especially for Upland cotton hybrid. The results suggest that trials in duplicate environments are a prerequisite for evaluating fiber quality traits (Shang et al. 2016a). The environmental factors in QTL associated with fiber quality and epistasis, and effect of QEs should be considered in MAS breeding (Xing et al. 2002; Shang et al. 2016b).

Overall, the single-locus with additive effect and epistasis with few detectable main effects play an important role in controlling expression of fiber quality in Upland cotton.

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