Male Central Hypogonadism in Paediatrics – the Relevance of Follicle-stimulating Hormone and Sertoli Cell Markers

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The definition of male hypogonadism, used in adult endocrinology, is not fully applicable to paediatrics. A clear understanding of the developmental physiology of the hypothalamic-pituitary-testicular axis is essential for the comprehension of the pathogenesis of hypogonadal states in boys and for the establishment of adequate definitions and classifications in paediatric ages. This is particularly true for central hypogonadism, usually called hypogonadotropic in adults. Because childhood is a period characterised by a physiological state of low gonadotropin and testosterone production, these markers of hypogonadism, typically used in adult endocrinology, are uninformative in the child. This review is focused on the physiological importance of prepubertal Sertoli cell markers – anti-Müllerian hormone (AMH) and inhibit B – and of the intratesticular actions of follicle-stimulating hormone (FSH) and testosterone during early infancy and the first stages of pubertal development. We discuss the role of FSH in regulating the proliferation of Sertoli cells – the main determinant of prepubertal testicular volume – and the secretion of AMH and inhibit B. We also address how intratesticular testosterone concentrations have different effects on the seminiferous tubule function in early infancy and during pubertal development.

Key words
Micropenis, microorchidism, cryptorchidism, puberty, hCG, human chorionic gonadotropin, recombinant human FSH

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In the adult male, hypogonadism is usually defined by the failure of the testis to produce physiological concentrations of testosterone and/or a normal number of spermatozoa. When due to abnormalities of the hypothalamic-pituitary axis, the condition is referred to as hypogonadotropic or central hypogonadism, whereas those due to testicular defects are called hypergonadotropic or primary hypogonadism. However, these definitions are inadequate for most paediatric conditions. Indeed, during the largest part of infancy and childhood, the hypogonadotropic axis is quiescent and the testes produce neither spermatozoa nor detectable amounts of testosterone. A more adapted definition and classification of male hypogonadism is based on the developmental physiology of the gonadal axis. Accordingly, the definition should be extended to any decreased testicular function as compared to what is expected for age, involving impaired hormone secretion by Leydig cells (i.e., androgens, insulin-like 3 [INSL3]), Sertoli cells (i.e., anti-Müllerian hormone [AMH], inhibit B) and/or a disorder of spermatogenesis. Moreover, it should be emphasised that primary hypogonadism rarely presents with elevated gonadotropin levels1 and even anorchidism might present with normal gonadotropin levels in about 30–60% of the cases during childhood, indicating that the term hypergonadotropic hypogonadism may be misleading in paediatrics. Additionally, since circulating gonadotropin concentrations are low in the normal prepubertal boy, it is difficult to demonstrate the existence of levels below normal, thus making the term hypergonadotropic hypogonadism also inadequate.

A clear understanding of the developmental physiology of the hypothalamic-pituitary-testicular axis is essential for the comprehension of the pathogenesis of hypogonadal states in paediatric ages (Figure 1). Under hypothalamic control, the gonadotrophs secrete luteinizing hormone (LH) and follicle-stimulating hormone (FSH) during the second and third trimesters of foetal life, and also during the first 3–6 months after birth – a period usually referred to as mini-puberty. LH induces Leydig cell activation, resulting in testosterone production at levels that are similar to those of the adult. Androgens are responsible for testicular descent and penile growth. FSH induces Sertoli cell proliferation2 and since Sertoli cells represent the largest part of testicular mass before puberty, FSH is the main factor regulating testicular volume (Figure 2). Sertoli cells produce peptide hormones, such as inhibit B and AMH. Although their basal production is gonadotropin-independent, there is clinical3,4 and experimental evidence5 that FSH increases Sertoli cell secretion of inhibit B and AMH.

Around birth, there is a transient decrease in gonadotropin and testicular hormone levels, but from the second week of postnatal life, all hormone levels increase and remain high for 3–6 months.
Thereafter and during the rest of infancy and childhood, LH and testosterone decrease to very low or undetectable levels. FSH and inhibin B also decrease but remain clearly detectable. Conversely, AMH peaks in the first year and remains high during childhood until it decreases in puberty inhibited by testosterone. In spite of the high levels of testosterone that seminiferous cords are exposed to during the foetal and postnatal periods, no signs of maturation are observed in Sertoli and germ cells. This is explained by the fact that Sertoli cells are physiologically insensitive to testosterone because the androgen receptor is not expressed in Sertoli cells until the second year of postnatal life (Figure 1). In fact, when elevated androgen levels abnormally persist beyond the age of 1 year (e.g. in boys with central precocious puberty), isolated and it is known as isolated hypogonadotropic hypogonadism, 22 resulting in response to AMH, mo = months; Trim = trimester of foetal life; WD & EG diff = differentiation of Wolffian ducts (anlagen of the epididymis, vas deferens and seminal vesicles) and external genitalia, occurring in response to testosterone. Modified with permission from Grinspon et al., 2014.

The onset of puberty is characterised by the increase in gonadotropin pulse frequency and amplitude. FSH further boosts testicular volume to >4 cc (Figure 2). LH reawakens Leydig cell activity, leading initially to an increase in intratesticular testosterone concentration which provokes Sertoli cell maturation: they stop dividing and decrease in number. From a clinical standpoint, when testicular volume reaches 4 cc the boy is said to have started pubertal development (Tanner stage two). This is due mainly to Sertoli cell proliferation in response to increased FSH activity. Concomitantly, LH induces Leydig cell testosterone secretion, resulting in an increase in intratesticular testosterone concentration. This leads to Sertoli cell maturation (reflected by a decrease in AMH secretion and an increase in inhibin B during Tanner stages two to three) and the development of full adult spermatogenesis, the main responsible for testicular volume growth between pubertal Tanner stages two to five. Subsequently, testosterone also increases in serum (Tanner stages three to five). AMH = anti-Müllerian hormone; FSH = follicle-stimulating hormone; LH = luteinizing hormone; MD regr = regression of Müllerian ducts (anlagen of the uterus and Fallopian tubes) occurring in response to AMH; mo = months; Trim = trimester of foetal life; WD & EG diff = differentiation of Wolffian ducts (anlagen of the epididymis, vas deferens and seminal vesicles) and external genitalia, occurring in response to testosterone. Modified with permission from Edelsztein et al., 2016.

During infancy and childhood, there is little change in testicular volume, which depends mainly on Sertoli cell number. From a clinical standpoint, when testicular volume reaches 4 cc the boy is said to have started pubertal development (Tanner stage two). This is due mainly to Sertoli cell proliferation in response to increased FSH activity. Concomitantly, LH induces Leydig cell testosterone secretion, resulting in an increase in intratesticular testosterone concentration. This leads to Sertoli cell maturation (reflected by a decrease in AMH secretion and an increase in inhibin B during Tanner stages two to three) and the development of full adult spermatogenesis, the main responsible for testicular volume growth between pubertal Tanner stages two to five. Subsequently, testosterone also increases in serum (Tanner stages three to five). AMH = anti-Müllerian hormone; FSH = follicle-stimulating hormone; LH = luteinizing hormone; MD regr = regression of Müllerian ducts (anlagen of the uterus and Fallopian tubes) occurring in response to AMH; mo = months; Trim = trimester of foetal life; WD & EG diff = differentiation of Wolffian ducts (anlagen of the epididymis, vas deferens and seminal vesicles) and external genitalia, occurring in response to testosterone. Modified with permission from Edelsztein et al., 2016.

**Figure 1: Serum levels of reproductive axis hormones in the male**

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**Figure 2: Schematic representation of the association between testicular volume and hormonal changes during postnatal development**

**Central hypogonadism in boys**

**Aetiology**

**Congenital central hypogonadism**

Congenital central (“hypogonadotropic”) hypogonadism reflects the failure of the pituitary to produce sufficient amounts of gonadotropins. Most frequently, it is caused by deficient production, secretion or action of gonadotropin-releasing hormone (GnRH). The condition can be isolated and it is known as isolated hypogonadotropic hypogonadism, 22 or associated to other pituitary hormone deficiencies.23 Several gene defects have been described as the cause of gonadotropin insufficiency. The pathogenesis may be related to: 1) altered GnRH neuron formation in the olfactory placode or its migration to the hypothalamus (usually associated with hypnosmia/anosmia and known as Kallmann syndrome), 2) abnormal GnRH production due to defects in regulatory factors or in the GnRH gene, and 3) defective action in the gonadotroph owing to abnormalities in the GnRH receptor or its transduction pathway.23–25 Extremely rare are mutations in the genes encoding the beta subunits of LH or FSH.26 Finally, genes associated with defective formation of the hypothalamic-pituitary region have been described in a low proportion of cases with multiple pituitary hormone deficiency.23

**Acquired central hypogonadism**

Of all paediatric and pubertal forms of central hypogonadism, approximately 20% are of functional origin. In these cases, a chronic illness or an underlying stressor causes the suppression of gonadotropin secretion and the condition can be reverted when the chronic illness is adequately treated. Conversely, permanent forms are due to conditions affecting the hypothalamic-pituitary area, e.g. high radiation doses, tumours, infections, surgery, etc.
Clinical presentation

Neonatal and infancy

The insufficiency of LH and testosterone during the second half of foetal life usually results in micropenis, cryptorchidism and hypoplastic scrotum. On the other hand, FSH insufficiency leads to microacidosis due to Sertoli cell hypoplasia. Midline defects and signs like hypoglycaemia or jaundice can orientate to multiple pituitary hormone deficiencies. Unfortunately, the insufficient attention given to cardinal features, such as micropenis, cryptorchidism, cleft lip/palate, deafness, coloboma, etc., in the newborn or infant results in missing the window of opportunity for the diagnosis during the postnatal activation period.

Prepubertal age

Since changes in testicular volume and penile size are minimal during childhood, acquired central hypogonadism established in this period of life may go underdiagnosed even if the underlying aetiology is evident, e.g. tumours or surgery of the central nervous system.

Pubertal age

Absence of pubertal signs is the main feature of complete forms of central hypogonadism, whereas delayed onset or progression, or even arrest of pubertal development, may be indicative of partial forms. At this age, smell testing can be helpful if a congenital form (Kallmann syndrome) is suspected.

Hormonal laboratory testing

Gonadotropins and testosterone

The window of opportunity for confirming central hypogonadism using gonadotropin and testosterone measurements in term newborns and infants resides between the third week and the third month. Indeed, in the first 2 weeks, gonadotropins and testosterone may still be low and only starting to increase after the perinatal nadir in normal newborns, thus hormone values should be interpreted cautiously in order to avoid false positive diagnoses. Similarly, from the fourth to sixth month onwards, gonadotropins and testosterone may already be decreasing physiologically. During the post-natal active period, or mini-puberty, low LH and testosterone have high positive predictive values for central hypogonadism.28,29

If the diagnosis was not certified before the sixth month of age, LH and testosterone levels are no longer informative until pubertal age. Although FSH is usually neglected in the study of male central hypogonadism, circulating levels never drop to undetectable during childhood and can therefore be more informative than LH. Indeed, in a series of boys presenting without spontaneous pubertal development, basal FSH had a higher predictive value than LH for the diagnosis of central hypogonadism.6,9

Sertoli cell markers

As already mentioned, AMH and inhibin B are Sertoli cell products, whose basal production is independent of gonadotropins. Nevertheless, FSH is capable of increasing inhibin B41 and AMH39,40 testicular secretion, thus indicating that they both are good markers of FSH action on the testis.41 The increased AMH testicular output in response to FSH in prepubertal male patients reflects two effects of FSH on immature Sertoli cells. On one hand, FSH induces Sertoli cell proliferation, which explains that FSH treatment in newborns or infants causes an increase in testicular volume.42 AMH levels reflect the amount of functional Sertoli cells in different conditions.43–46 On the other hand, using two experimental models, knockout mice null for the gene encoding the FSH beta subunit47 and the prepubertal Sertoli cell lineage SMAAT1 expressing AMH,48 we showed that FSH upregulates the AMH promoter activity signalling through the cyclic AMP-mediated pathway, mainly via protein kinase A activity.49–51 The aforementioned regulation of Sertoli cells by FSH during foetal life and the neonatal period explains the findings of microacidosis and low serum AMH and inhibin B in boys with congenital central hypogonadism.4,9,20,23,52

Treatment

Testosterone

In patients with congenital central hypogonadism, the presence of micropenis may prompt testosterone treatment. When the diagnosis is made at birth – or more rarely during childhood – depot testosterone (testosterone enanthate or cypionate) at 25–50 mg/dose intramuscular (IM) every 4 weeks for 3 consecutive months may be sufficient to normalise penile size without provoking an advancement of bone age.13 In older boys with central hypogonadism reaching the age of puberty, steroid replacement is necessary for the development of secondary sex characteristics, the acquisition of full bone mass and the optimisation of height velocity, as well as for avoiding psychosocial distress. Depot testosterone is given at 50 mg IM each month for 6–12 months, with a subsequently progressive dose escalation until a full dose of 250 mg/month in 2–3 years. The aim is to mimic normal pubertal tempo. When full pubertal development is attained, treatment should be switched to follow standards for adults.1 Alternative approaches include oral androgens such as testosterone undecanoate43 and transdermal gels.51,52

Gonadotropins

Exogenous testosterone supplementation aims to attain physiological circulating androgen levels. However, physiological intratesticular testosterone concentration is never reached with these treatments, whose disadvantages are that Sertoli cells neither proliferate nor mature and, subsequently, adult spermatogenesis does not develop. In fact, testicular volume remains small (<4 cc) in patients receiving only testosterone treatment.

Treatment with pulsatile GnRH or combined gonadotropins is well established in adult patients with central hypogonadism seeking fertility. Conversely, limited information is available on the use of gonadotropins in paediatric patients.43 Observational studies on infants with congenital central hypogonadism suggest that the postnatal peak in reproductive hormones plays an important role in genital development.44 Therefore, various authors have proposed the use of gonadotropins during infancy, instead of testosterone, to mimic mini-puberty in patients with congenital central hypogonadism. A case report describes an infant treated from the age of 7.9 months with recombinant human (rh) LH (subcutaneous injections 20 IU twice a week) and FSH (subcutaneous injections 2.5 IU/kg twice a week).53 Testicular volume increased 170%, reflecting FSH action on Sertoli cells.

A case series reports eight patients aged 0.25–11 months who received continuous infusion of rhLH and rhFSH with insulin pumps at a daily rate of 50 and 75–150 IU, respectively.54 Testicular hormones reached “mini-puberty” levels, complete testes descent occurred in six patients and partial descent in two, and testes and penis reached normal dimensions in all cases. Similar results were observed in patients receiving continuous subcutaneous infusion of rh gonadotropins.55 Even though long-term follow-up is needed to fully evaluate the outcomes of this therapeutic approach, these results suggest that subcutaneous gonadotropin infusion is able to induce not only Sertoli cell proliferation and function but also testis descent in patients with congenital central hypogonadism, thus avoiding surgical treatment.45

Only one study reports prepubertal treatment with rhFSH of three boys with central hypogonadism.4 In this case, the rationale was that
FSH would be capable of inducing Sertoli cell multiplication before they mature at puberty. The boys were given rhFSH subcutaneously (1.5 IU/kg) three times a week for 12 months. Treatment induced testicular growth and inhibin B production, reflecting stimulation of immature Sertoli-cell function. These findings suggest that FSH treatment in childhood is capable of inducing Sertoli-cell proliferation, which may result in an increased sperm-producing capacity in adulthood.

Finally, several publications exist on combined recombinant gonadotropin treatment for the induction of puberty in boys with central hypogonadism. Some of them used the standard protocols for adults initiating with human chorionic gonadotropin (hCG) and then adding rhFSH and showed acceptable results. Moreover, in studies using initial priming with rhFSH prior to induction of puberty with the combination of hFSH and hCG, rhFSH induced prepubertal testicular growth and increased serum levels of AMH (Figure 3) and inhibin B levels (Figure 4), confirming the importance of FSH action on immature Sertoli cells. The addition of hCG resulted in a progressive increase in testosterone production, Sertoli cell maturation suggested by an important decrease in serum AMH and acquisition of sperm production. Also, the increase of inhibin B proved useful for monitoring spermatogenic development. More recently, an open-label randomised study in hypogonadal males aged >18 years showed better results in men receiving rhFSH pre-treatment in terms of spermatogenic development.

**Concluding remarks**

While LH and testosterone levels have been used for years in the diagnosis of central hypogonadism at pubertal age and adulthood, in paediatric patients they are useful only during the early post-natal activation period. During childhood, low levels of the Sertoli cell markers AMH and inhibin B support the diagnosis of central hypogonadism. Furthermore, although testosterone replacement therapies have been used for many years with satisfactory results on the development of secondary sex characteristics in adolescents with central hypogonadism, more recently the attention has been driven to the physiological importance of the intratesticular actions of FSH and testosterone during early infancy and the first stages of pubertal development.

The limited number of studies available to date demonstrate that a sequential treatment approach with rhFSH priming to induce Sertoli cell proliferation, before adding hCG or rhLH to provoke an increase in intratesticular testosterone concentration and Sertoli cell maturation, shows encouraging results in terms of induction of testicular growth and sperm production. To definitively prove the superiority of this approach, larger randomised prospective studies are necessary.
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1. Bhasin S, Brito JB, Cunningham GR, et al. Testosterone therapy in men with hypogonadism: an endocrine society clinical practice guideline. J Clin Endocrinol Metab. 2016;101:1-30.

2. Rey RA, Grinspon RP, Gottlieb S, et al. Male hypogonadism: an extended classification based on a developmental, endocrine physiology based approach. Andrology. 2013;1:3-16.

3. Grinspon RP, Gottlieb S, Bedecarras P, Rey RA. Anti-Müllerian hormone and testicular function in prepubertal boys with cryophthalmic. Front Endocrinol (Lausanne). 2018;9:182.

4. Grinspon RP, Raposo MG, Bedecarras E, et al. Gonadotrophin secretion pattern in anorches boys from birth to pubertal age: pathological and diagnostic usefulness. Clin Endocrinol. 2008;69:1248-55.

5. Rey RA. Puberty: true and puberity: differences in testicular function. Ann Endocrinol (Paris). 2014;75:58-63.

6. Kuhl Hanneli, S. Sankampli L, Dunkel L. Activation of the hypothalamic-pituitary-gonadal axis in infancy, mini-puberty. Horm Res Paediatr. 2014;82:73-80.

7. Drife M. The role of follicle-stimulating hormone in controlling Sertoli cell proliferation in testes of rat fetuses. Endocrinology. 1984;115:1248-55.

8. Raivio T, Toppari J, Peerheisterpaa A, et al. Treatment of prepubertal gonadotrophin-deficient boys with recombinant human follicle-stimulating hormone. Lancet. 1997;350:263-4.

9. Lambert AS, Bougnères P. Growth and descent of the testes: a developmental and molecular genetic approach. J Clin Endocrinol Metab. 2010;95:5357–64.

10. Al-Attar L, Noël K, Dutertre M, et al. Hormonal and cellular mechanisms of anti-Müllerian hormone action in the postnatal mouse. J Clin Endocrinol Metab. 2012;97:3533–43.

11. Rey RA, Grinspon RP, Gottlieb S, et al. Male central precocious puberty: serum profiles of anti-Müllerian hormone in children. J Clin Endocrinol Metab. 2011;96:2899–907.

12. Grinspon RP, Raposo MG, Gottlieb S, et al. Anti-Müllerian hormone production through Sertoli cell proliferation and a nonclassical cyclic adenine 3′-monophosphate-mediated activation of the AMH gene. Mol Endocrinol. 2002;17:550–61.

13. Berga SL, Timms I, Cardozo D, et al. Longitudinal studies of sexual maturity and testicular function: a developmental study of healthy boys. J Pediatr Endocrinol Metab. 2016;29:295–305.

14. Andersson AM, Toppari J, Haavisto AM, et al. Longitudinal reproductive hormone profiles in infants: peak of inhibin B levels in pubertal boys exceeds levels in adult men. J Clin Endocrinol Metab. 1998;83:675–81.

15. Grinspon RP, Bedecarras E, Raposo MG, et al. Early onset of primary hypogonadism revealed by serum anti-Müllerian hormone determination during infancy and childhood in boys with Klinefelter syndrome. J Pediatr. 2011;158:2487–91.

16. Alexiadis I, Mattheus T, Kamitsuka R, et al. Anti-Müllerian hormone and testosterone serum levels are inversely related during normal and precocious pubertal development. J Clin Endocrinol Metab. 1997;82:1202–6.

17. Chemes HE, Rey RA, Nistal M, et al. Physiological androgen insensitivity of the foetal, neonatal, and early infantile testes is explained by the antagonism of the androgen receptor expression gene in mammalian and nonmammalian species. DNA Cell Biol. 2004;23:572–85.

18. Lasala C, Schingel PF, Araúcho N, et al. SOX9 and SP1 are involved in cyclic AMP-mediated upregulation of anti-Müllerian gene expression in the testicular prepubertal Sertoli cell SMA1. Am J Physiol Endocrinol Metab. 2011;301:E539–47.

19. Grinspon RP, Rey RA. Anti-Müllerian hormone and Sertoli cell function in paediatric male hypogonadism. Horm Res Paediatr. 2010;73:11–9.

20. Bin-Abbas B, Contie F, Grumbach M, Kaplan S. Congenital hypogonadotropic hypogonadism and microcephaly: Effect of hypogonadotropic hypogonadism on adult size–Why the sex difference is not indicated. J Pediatr. 1999;134:579–83.

21. Lawitz EG, Hagen CP, Meintz MG, et al. Evaluation of a Danish boys with delayed puberty: diagnostic use of a new puberty nomogram and effects of oral testosterone therapy. J Clin Endocrinol Metab. 2015;100:1376–85.

22. Chomia L, Papuciu C, Fintiri D, Cappa M. Use of testosterone gel compared to intramuscular formulation for puberty induction in males with constitutional delay of growth and puberty: a preliminary study. J Endocrinol Invest. 2018;41:259–63.

23. Contreras MF, Rašarangie M, Prasad K, et al. Transdermal testosterone gel for induction and continuation of puberty in adolescent boys with hypogonadal dysfunction. Pediatr Endocrinol Rev. 2017;15:105–9.

24. Ulla Ayugarte A, Lira-Abalbiso S. Clinical applications of anti-Müllerian hormone in the male. Prog Mol Biol Transl Sci. 2016;143:121–74.

25. Main KM, Schmidt M, Skakkebaek NE. A possible role for reproductive hormones in newborn boys: progressive hypogonadism without the postnatal testosterone peak. J Clin Endocrinol Metab. 2000;85:4950–7.

26. Main KM, Schmidt M, Toppari J, Skakkebaek NE. Early postnatal treatment of hypogonadotropic hypogonadism with recombinant human FSH and LH. Eur J Endocrinol. 2002;146:75–9.

27. Stoupia A, Samara-Boustani D, Flechter I, et al. Efficiency and safety of continuous subcutaneous infusion of recombinant human growth hormone for congenital micrognathia during early infancy. Horm Res Paediatr. 2017;87:103–10.

28. Bouwetter C, Malone L, Bouguerd J, et al. Neonatal gonadotropin therapy in congenital hypogonadotropic hypogonadism. J Pediatr Endocrinol Metab. 2012;25(1–2):172–82.

29. Rohayem J, Hauffa BP, Zacharin M, et al. Testicular growth and spermatogenesis: new goals for pubertal hormone replacement in boys with hypogonadotropic hypogonadism—a multicentre prospective study of hCG/FSH treatment outcomes during adolescence. Clin Endocrinol (Oxf). 2017;86:75–87.

30. Zacharin M, Sabin MA, Naer V, Dabaghian P. Addition of recombinant follicle-stimulating hormone to human chorionic gonadotropin treatment in adolescents and young adults with hypogonadotropic hypogonadism promotes normal testicular growth and may promote early spermatogenesis. Fertil Steril. 2012;98:836–42.

31. Stalvi T, Wåkå M, AM, Dunkel L. Treatment of gonadotrophin-deficient boys with recombinant human FSH: long-term observation and outcome. Eur J Endocrinol. 2007;156:105–11.

32. O’Shaughnessy AA, Sykes J, Hayes FJ, et al. Trial of recombinant follicle-stimulating hormone pretreatment for GnRH-induced fertility in patients with congenital hypogonadotropic hypogonadism. J Clin Endocrinol Metab. 2013;98:E1790–5.