Review Article

Update on Berberine in Nonalcoholic Fatty Liver Disease

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Berberine (BBR), an active ingredient from nature plants, has demonstrated multiple biological activities and pharmacological effects in a series of metabolic diseases including nonalcoholic fatty liver disease (NAFLD). The recent literature points out that BBR may be a potential drug for NAFLD in both experimental models and clinical trials. This review highlights important discoveries of BBR in this increasing disease and addresses the relevant targets of BBR on NAFLD which links to insulin pathway, adenosine monophosphate-activated protein kinase (AMPK) signaling, gut environment, hepatic lipid transportation, among others. Developing nuanced understanding of the mechanisms will help to optimize more targeted and effective clinical application of BBR for NAFLD.

1. Introduction

As the global waistline continues to expand, metabolic abnormalities including obesity, type 2 diabetes, hypertension, and dyslipidemia, collectively termed the “metabolic syndrome,” are reaching epidemic proportions. The pathogenesis of these disease state is hypothesized to begin with abnormal accumulation of lipids in nonadipose tissues (steatosis), known as nonalcoholic fatty liver disease (NAFLD), a chronic condition that is currently the leading cause of referrals to hepatology clinics. Due to serious adverse effects and the limited effectiveness of currently available pharmacological therapies for “metabolic syndrome,” many research efforts have focused on the development of drugs from natural products.

Berberine (BBR, \( \text{C}_{20}\text{H}_{18}\text{NO}_4 \)) is an isoquinoline alkaloid of the protoberberine type, which presents in an array of plants, including Hydrastis canadensis (goldenseal), Coptis chinensis (Coptis or goldenthread), Berberis aquifolium (the Oregon grape), Berberis vulgaris (barberry), and Berberis aristata (tree turmeric) among others. The isoquinoline alkaloid drug belongs to the structural class of protoberberines which includes a quaternary base (Figure 1). There are many derivates and analogues available, such as berberine hydrochloride, berberine sulfate, and berberine citrate or phosphate, contributing to its multiple pharmacological and biochemical effects.

BBR is traditionally used as an antimicrobial and antiprotozoal drug, the antimicrobial activity against a variety of organisms, including bacteria, viruses, fungi, protozoans, helminths, and chlamydia, which have been applied in Chinese medicine for many decades. Recent researches have revealed novel pharmacological properties and multiple therapeutic applications, mainly concerning metabolic diseases, such as obesity and type 2 diabetes [1]. Kinetic study shows that BBR metabolites are widely distributed into various tissues, including liver, heart, kidney, spleen, lung, and even brain, with the liver being the most predominant organ, and average concentration of BBR in liver is approximately 70-fold greater than that in plasma [2]. Other dosing routes, such as femoral vein administration also identified the disposition of BBR in blood, liver, and bile fluid [3]. Additionally, BBR has longer half-life in liver than other tissues [4], suggesting liver as the main target organ of BBR.

In the past several decades, BBR’s action on glucose and lipid metabolic disorders has been widely studied; although the effects or mechanisms of BBR on NAFLD could not separate completely from other metabolic diseases, the role of BBR
2. Effect of BBR on Fatty Liver

BBR is reported to inhibit cholesterol and triglyceride synthesis in human hepatoma cell line (HepG2) cells and primary hepatocytes [5, 6], and treating rat hepatoma H4IIIE cells with BBR shows increased glucose consumption in dose-dependent manner [7]. In vivo data from various animal models also confirm BBR's beneficial role in preventing or treating NAFLD. Intraperitoneal injection of BBR for three weeks has been shown to alleviate hyperlipidemia and fatty liver in obese db/db and ob/ob mice [8]. In Zucker diabetic fatty rats, two-week treatment with a BBR-containing formula could attenuate fatty degeneration [9]. Treatment of hyperlipidemic hamsters with BBR strongly reduces fat storage in the liver [5]. As for mice with high-fat diet (HFD) induced fatty liver, sixteen weeks BBR supplement could alleviate hepatic steatosis and decrease liver lipid content by 14% [10]. This antisteatosis effect of BBR is also reported in diabetic hyperlipidemic rats, which demonstrates that BBR prevents the pathological progression of liver and reverts the increased hepatic triglyceride to near the control levels [11].

In addition, BBR further prevents the development of obesity and insulin resistance in HFD-fed rats [12], and BBR may also prevent liver fibrosis experimental models [13]. Clinical investigations showed that BBR supplement may reduce alanine and aspartate transaminase levels in patients with type 2 diabetes, indicating the restoration of liver function [14, 15]. Furthermore, BBR has been shown to reduce liver necrosis both in nonalcoholic steatosis and in steatosis due to hepatitis C infection [13]. In elderly hypercholesterolemic patients who were previously statin-intolerant, BBR demonstrates reduced cholesterolemia and plasma low density lipoprotein-cholesterol (LDL-c) levels [16].

3. Mechanisms of BBR in NAFLD

The precise mechanisms of the development of NAFLD or BBR improving fatty liver remain largely unclear. Defects in lipid metabolism pathways, insulin resistance, and inflammation are crucial players in the process of NAFLD. The recent literature points out that BBR may be integrated into lipid and glucose regulation, combating fatty liver and related syndromes, and the beneficial role of BBR on NAFLD might be achieved through multiple mechanisms.

3.1. BBR in Mediating Insulin Resistance. Insulin resistance plays a critical role in the pathogenesis of NAFLD [17]; hence, improving insulin sensitivity is of great importance in dealing with NAFLD. Evidence of BBR on insulin resistance has been elucidated in clinical trials as well as experimental animals and cells lines [14, 18, 19]. Although there is no definitive explanation of how BBR in regulating sensitivity, there are several encouraging observations revealing the possible mechanisms.

After meal, pancreatic islets secrete insulin, and insulin's presence at the cell surface is transduced to cytoplasmic and nuclear responses by tyrosine phosphorylation of insulin receptor substrates (IRSs). Nutrient-induced serine phosphorylation of IRS proteins is proved to be the counter-regulation of the signaling pathway, which blunts insulin action in stressed target tissues and stems the influx of nutrients into already overwhelmed cells [20]. Recently, low grade inflammation and endoplasmic reticulum (ER) stress have been proposed to be in close association with insulin resistance, activation of inflammatory pathways, such as Jun N-terminal kinases (JNKs), inhibitor of nuclear factor B (IκB) kinase β (IKKβ), inositol requiring enzyme 1 (IRE-1), could coverage [21–23].

BBR was reported to stimulate insulin secretion in HIT-T15 cells and pancreatic islets, which may have certain influence on its antidiabetic activity [24]. Interleukin-6 (IL-6) and tumor necrosis factor-α (TNF-α) production in HepG2 cells represent a state of inflammation and consequently impair insulin pathway, and treatment with BBR effectively inhibits IL-6 and TNFα production in a concentration-dependent manner, and improves insulin signaling cascade by modification of IRS-1 and Protein kinase B (Akt) Ser/Thr phosphorylation, indicating the insulin enhancing potential of BBR is through the anti-inflammatory activity [25]. Moreover, pretreating with BBR could block tunicamycin induced ER stress, coexisting with the inhibition of PKR-like eukaryotic initiation factor 2α kinase (PERK) and eukaryotic translational initiation factor 2α (eIF2α) phosphorylation, two protein markers of ER stress, and this results in increasing IRS-1 Tyr phosphorylation whereas decreasing its Ser (307) phosphorylation, thus improving insulin resistance under the condition [26]. Other mechanisms, including BBR-caused insulin receptor (InsR) promoter activation and InsR messenger RNA (mRNA) transcription, might also contribute to effect of BBR in regulating insulin sensitivity. In vitro studies further suggest that BBR-induced InsR gene exression depends on protein kinase C (PKC) activity [27], possibly depends on protein kinase C (PKC) activity in the liver, and BBR mediated PPARs restoration is observed to be in parallel with hepatic glycogen and triglyceride attenuation actions [11]. Our previous work also identified the direct action of BBR on IRS [28]. Taken together, these studies indicate that BBR might be of great insulin sensitive potential and an active player in the liver.
3.2. BBR in Regulating AMPK Pathway. Adenosine monophosphate-activated protein kinase (AMPK) is an attractive drug target that plays a key role in regulation of whole-body energy homeostasis. Increasing empirical evidence points towards AMPK activation as the target of BBR. In different animal models and various cell lines, phosphorylation of AMPK is identified, attributing to the hypoglycemic and hypolipidemic effect of BBR [5, 7, 29, 30]. To further verify this target, exposure cultured cells with Compound C, an AMPK inhibitor, BBR mediated effect is then abolished [8, 31], implying AMPK as a crucial player of BBR to dissipate stored fat and elevated glucose levels.

The mechanism of BBR on AMPK activation is still elusive, and studies suggest mitochondrial function as the key of this issue. BBR was used as cationic fluorescent probe for investigating energized state of liver mitochondria several decades ago, penetration of BBR into mitochondria could inhibit NAD-linked respiration, and the inhibition is totally dependent on the energization of the membrane [32]. Later, in vitro experiments provided detailed insight into the mechanisms of BBR in regulating this pathway. BBR inhibits glucose oxidation in mitochondria, which leads to an increase in the AMP/ATP (adenosine monophosphate/adenosine triphosphate) ratio in cells, thus accounting for AMPK activation [7, 19, 33, 34]. Other studies further identified complex I inhibition in the mitochondrial respiratory chain as the target of BBR, which blocks AMP conversion into ATP, allowing AMP accumulation in mitochondrial [19, 33, 34]. When elevated AMP binds to the subunit γ, the inhibitory domain of the α1 subunit is released from the kinase domain, results in an active conformation of AMPK [35].

The liver is a vital organ present in vertebrates and has a wide range of functions. Activation of hepatic AMPK leads to increased fatty acid oxidation and simultaneously inhibition of hepatic glucose production as well as lipogenesis and cholesterol synthesis. In vivo data from obese db/db and ob/ob mice indicate that BBR stimulates the expression of fatty acid oxidative genes, while suppresses that of lipogenic genes [8], and BBR also improves insulin resistance in nutrient stressed mice through activation of AMPK [36]. Hepatic sterol regulatory element-binding proteins (SREBPs), liver X receptor α (LXα) and peroxisome proliferator-activated receptor α (PPARα) transcriptional programs are also observed to be involved in the therapeutic mechanisms of BBR in type 2 diabetic hamsters [37], supporting the concept that BBR prevents dyslipidemia and fatty liver by directly promoting the activation of hepatic AMPK. As an alternative to animal studies, BBR hydrochloride regulates the transcription of hepatic genes that involve in glucose and fatty acid metabolism in vitro experiment using rat primary hepatocytes [6]. More importantly, AMPK specifically binds to and directly phosphorylates SREBP-1c and SREBP-2, and the Ser372 phosphorylation of SREBP-1c by AMPK could inhibit the proteolytic cleavage and nuclear translocation of SREBP-1c in hepatocytes, thereby preventing its autoregulation and transcription of target lipogenic genes [acetyl-CoA carboxylase 1 (ACCC), fatty acid synthase (FAS), and stearoyl CoA desaturase 1 (SCD1)] [38]. Indeed, BBR was reported to inhibit both SREBP1c and SREBP2 expression in hepatocytes [34, 39], quite consistent with its AMPK regulation mechanism, explaining the triglyceride and cholesterol lowering effect of the compound.

3.3. BBR in Modifying Gut Microenvironment. Roughly 75% of the blood entering the liver is venous blood from the portal vein, which is all from the digest system. Therefore, the liver gets “first pickings” of everything absorbed in the small intestine. While simple steatosis seems to be well tolerated and to have only mild consequences, a significant proportion of patients with NAFLD develop nonalcoholic steatohepatitis (NASH), a condition that may result in hepatic fibrosis, cirrhosis, and hepatocellular carcinoma [40, 41]. Although the circumstances that may lead to fatty liver progress remain largely unknown, components from the intestinal microflora may contribute to the regulation of proinflammatory processes in the liver that has been investigated in recent years [42].

Due to the low bioavailability and poor absorption through the gut wall, BBR might exert its effect in the intestinal tract before absorption. BBR has been shown to have significant antimicrobial activity against bacteria, fungi, parasites, worms, and viruses. In terms of bacteria, BBR has demonstrated highly significant activity against Staphylococcus, Streptococcus, Salmonella, Klebsiella, Clostridium, Pseudomonas, Proteus, Shigella, Vibrio, and Cryptococcus species [43]. BBR also exhibited effectiveness in combating enterotoxigenic Escherichia coli diarrhea [44]. Moreover, BBR inhibited the overgrowth organisms such as staphylococci and coliforms, while having no effect on indigenous lactobacilli and bifidobacteria.

Alternatively, BBR inhibits a wide range of intestinal microbes, modulates shift of the gut microbiota structure, and enriches some short-chain fatty acid (SCFA) producers in HFD-fed rats [12]. It has been shown that the antimicrobial activity of BBR can be mediated by inhibiting FtsZ (an essential cytoskeleton protein for bacteria cytokinesis) assembly and halting cell division of the bacteria [45]. Additionally, BBR-regulated reversion of inducible cyclooxygenase-2 (COX-2) might also contribute to this process [46]. Dysfunction of gut microbiota has more effective capacity to harvest energy from the diet, and when germ-free mice were transplanted with caecal microbiota from ob/ob mice, they developed obesity and insulin resistant within 2 weeks [47], thus the antimicrobial role of BBR might putatively affect energy absorption, which could partially explain its lipid-lowering action under nutrient oversupplying conditions.

BBR could regulate integrity of tight junction in cultured human Caco-2 cells [48], and similar effect was reflected in a mice model of endotoxemia; pretreatment with BBR attenuates disruption of tight junctions in intestinal epithelium in the animals [49], suggesting BBR's action in reducing epithelial gut permeability under pathogen stressed conditions, with the possible involvement of nuclear factor-kB (NF-kB) and myosin light chain kinase pathways [49, 50]. Therefore, BBR mediated intestinal barrier improvement could block the endotoxemia into circulation, thus reducing hepatic inflammation and preventing NAFLD progression.
3.4. BBR on Hepatic Lipid Secretion. Secretion of triglycerides as very low-density lipoprotein (VLDL) is an important aspect in maintaining hepatic lipid homeostasis. Hepatocytes have a unique ability to present triglycerides to the organelles where VLDL assembly takes place. Efficient assembly of apolipoprotein B (apoB) 100 with triglyceride and cholesterol into VLDL requires the activity of an ER-resident microsomal triglyceride transfer protein (MTTP), and loss-of-function mutations within the MTTP gene are the cause of the human abetalipoproteinemia, an autosomal recessive disease characterized by the total absence of triglyceride-rich lipoproteins in the plasma [51]. In an HFD-induced NAFLD rat model, Chang et al. [10] observed three increased DNA methylation sites in the MTTP promoter, which account for the reduced MTTP expression in the liver. BBR treatment could cause demethylation of the abnormal regions in MTTP promoter, counteracting HFD-induced MTTP dysregulation. BBR regulated restoration of MTTP expression and VLDL assembly further increase triglyceride secretion and alleviate fatty liver [10].

3.5. BBR on Cholesterol Metabolism. BBR has shown considerable impact on cholesterol metabolism, and preclinical and clinical studies both suggest lipid-lowering acting of the drug [52, 53]. The molecular mechanism has been proposed to be through stabilization of the low-density lipoprotein receptor (LDLR) messenger RNA, which led to upregulation of LDLR protein [54, 55]. Other studies illustrate that BBR-induced stabilization of LDLR mRNA is mediated by the extracellular signal-regulated kinase (ERK) signaling pathway through interactions of cis-regulatory sequences of 3′ untranslated region (UTR) and mRNA binding proteins that are downstream effectors of this signaling cascade [56]. Additionally, SREBPs also act as regulators of hepatic cholesterol levels and activate genes involved in the synthesis of cholesterol and free fatty acids [57]. SREBP cleavage-activating protein has a cholesterol-sensing domain that senses intracellular cholesterol levels and directs the activity of SREBPs. Therefore, the ability of BBR in suppressing SREBPs could be another mechanism for its role in cholesterol metabolism, further explain the beneficial effects on NAFLD.

3.6. Other Possible Mechanisms. BBR can downregulate hepatic expression of uncoupling protein-2 (UCP2) mRNA protein in NAFLD rats, promote the recovery of hepatocyte steatosis, and improve lipid metabolism disorder [58]. BBR also demonstrates to have antioxidative activities in cultured cells and animals, and this effect may help reducing reactive oxygen species (ROS) production in the liver [59, 60], thus preventing liver damage. Extrahepatic factors might also affect NAFLD pathology, and in adipocytes, studies illustrate that BBR inhibits adipogenesis in murine-derived 3T3-L1 preadipocytes and human white preadipocytes [61], while enhancing glucose and fatty acid uptake by muscle cells has been proved [62], BBR is proved to play a role in pancreatic islets as well [24]. Both intrahepatic and extrahepatic
mechanisms integrate into the BBR regulating action in combating metabolic diseases.

4. Prospects of BBR on NAFLD

There is a long history of safe usage of BBR in clinic, sporadic single case reports on the adverse effect of BBR include gastrointestinal side effects, allergic skin reaction and arrhythmia [63]. Even though some reports indicate that BBR could induce hepatoma cells apoptosis, the cytotoxic effects were absent in healthy hepatocytes [64, 65], and BBR actually shows antihepatotoxic action in a series of studies. BBR is investigated in rats with acetaminophen-induced hepatic damage, which shows decreased serum levels of alkaline phosphatase (ALP) and aminotransaminases, suggestive of hepatoprotection of the alkaloid [66]. In cultured rat hepatocytes, Hwang et al. [67] demonstrated bioactivity of BBR in protecting the cells against oxidativedamage.

However, despite the promising effects of BBR on animal models and cells, large clinical investigations are not available. Due to low bioavailability and poor absorption of BBR via oral administration [2], high dose oral administration usually causes gastrointestinal side effects, which greatly limit its clinical application. There are some reports, however, trying to explore new dosage forms of BBR to increase its bioavailability. Preclinical studies that use sodium caprate, one of the intestinal absorption enhancers, showed 1.5- to 2.3-fold increase of BBR bioavailability in different models [68, 69]. P-glycoprotein (P-gp) inhibitors, such as cyclosporine and verapamil, also illustrate marked increase of BBR absorption [70]. Though the beneficial action of BBR is obvious, further studies are still in need to optimize its clinical application in NAFLD.

5. Summary

NAFLD is the liver manifestation of obesity and the metabolic syndrome and is marked by lipid deposition and/or inflammation. In this review, we introduced the beneficial potential of a nature compound, BBR, in NAFLD and the possible mechanisms under the therapeutic actions (Figure 2). Though most evidence based on experimental studies, and clinical trials need to be further confirmed, we still hold the belief that BBR is a promising candidate in preventing and treating NAFLD in the future. However, more studies should be cautiously performed to clarify the mechanisms and optimize clinical application of the drug.

Abbreviations

- ACC1: Acetyl-CoA carboxylase 1
- ALP: Alkaline phosphatase
- AMP: Adenosine monophosphate
- AMPK: Adenosine monophosphate-activated protein kinase
- apoB: Apolipoprotein B
- BBR: Berberine
- COX-2: Cyclooxygenase-2
- eIF2α: Eukaryotic translational initiation factor 2α
- ER: Endoplasmic reticulum
- ERK: Extracellular signal-regulated kinase
- FAS: Fatty acid synthase
- HFD: High-fat diet
- IKKβ: Inhibitor of kappaB kinase β
- IL-6: Interleukin-6
- IRE-1: ER-to-nucleus signaling 1
- IRS: Insulin receptor substrate
- IC: Inhibitor of nuclear factor B
- JNK: Jun N-terminal kinases
- LDL-c: Low-density lipoprotein cholesterol
- LDLR: Low-density lipoprotein receptor
- LXα: Liver X receptor α
- mRNA: Messenger RNA
- MTTP: Microsomal triglyceride transfer protein
- NAFLD: Nonalcoholic fatty liver disease
- NASH: Nonalcoholic steatohepatitis
- NF-κB: Nuclear factor-κB
- PERK: PKR-like eukaryotic initiation factor 2α kinase
- P-gp: P-glycoprotein
- PKB, Akt: Protein kinase B
- PKC: Protein kinase C
- PPARs: Peroxisome proliferator-activated receptors
- ROS: Reactive oxygen species
- SCDI: Stearoyl CoA desaturase 1
- SCFA: Short-chain fatty acid
- Ser/Thr: Serine/threonine
- SREBPs: Sterol regulatory element-binding proteins
- t-BHP: Tert-butyl hydroperoxide
- TNF-α: Tumor necrosis factor-α
- UCP2: Uncoupling protein-2
- UTR: Untranslated region
- VLDL: Very low-density lipoprotein.

Conflict of Interests

The authors report no conflict of interests.

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References

[1] P. R. Vuddanda, S. Chakraborty, and S. Singh, “Berberine: a potential phytochemical with multispectrum therapeutic activities,” Expert Opinion on Investigational Drugs, vol. 19, no. 10, pp. 1297–1307, 2010.
[2] Y.-T. Liu, H.-P. Hao, H.-G. Xie et al., “Extensive intestinal first-pass elimination and predominant hepatic distribution of berberine explain its low plasma levels in rats,” Drug Metabolism and Disposition, vol. 38, no. 10, pp. 1779–1784, 2010.
[3] P.-L. Tsai and T.-H. Tsai, “Simultaneous determination of berberine in rat blood, liver and bile using microdialysis coupled to high-performance liquid chromatography,” Journal of Chromatography A, vol. 961, no. 1, pp. 125–130, 2002.

[4] Y. Liu, H. Hao, H. Xie, H. Lv, C. Liu, and G. Wang, “Oxidative demethylation and subsequent glucuronidation are the major metabolic pathways of berberine in rats,” Journal of Pharmaceutical Sciences, vol. 98, no. 11, pp. 4391–4401, 2009.

[5] J.-M. Brusq, N. Ancellin, P. Grondin et al., “Inhibition of lipid synthesis through activation of AMP kinase: an additional mechanism for the hypolipidemic effects of berberine,” Journal of Lipid Research, vol. 47, no. 6, pp. 1281–1288, 2006.

[6] Y. Ge, Y. Zhang, R. Li, W. Chen, Y. Li, and G. Chen, “Berberine regulated Gck, G6pc, Pck1 and srebp-1c expression and activated AMP-activated protein kinase in primary rat hepatocytes,” International Journal of Biological Sciences, vol. 7, no. 5, pp. 673–684, 2011.

[7] J. Yin, Z. Gao, D. Liu, Z. Liu, and J. Ye, “Berberine improves glucose metabolism through induction of glycolysis,” American Journal of Physiology, vol. 294, no. 1, pp. E148–E156, 2008.

[8] W. S. Kim, Y. S. Lee, S. H. Cha et al., “Berberine improves lipid dysregulation in obesity by controlling central and peripheral AMPK activity,” American Journal of Physiology, vol. 296, no. 4, pp. E812–E819, 2009.

[9] H.-L. Zhao, Y. Sui, C.-F. Qiao et al., “Sustained antidiabetic effects of a berberine-containing Chinese herbal medicine through regulation of hepatic gene expression,” Diabetes, vol. 61, no. 4, pp. 933–943, 2012.

[10] X. Chang, H. Yan, J. Fei et al., “Berberine reduces methylation of the MTTP promoter and alleviates fatty liver induced by a high-fat diet in rats,” Journal of Lipid Research, vol. 51, no. 9, pp. 2504–2515, 2010.

[11] J. Y. Zhou, S. W. Zhou, K. B. Zhang et al., “Chronic effects of berberine on blood, liver glucolipid metabolism and liver PPARs expression in diabetic hyperlipidemic rats,” Biological and Pharmaceutical Bulletin, vol. 31, no. 6, pp. 1169–1176, 2008.

[12] X. Zhang, Y. Zhao, M. Zhang et al., “Structural changes of gut microbiota during berberine-mediated prevention of obesity and insulin resistance in high-fat diet-fed rats,” PLoS ONE, vol. 7, no. 8, Article ID e42529, 2012.

[13] B.-J. Zhang, D. Xu, Y. Guo, J. Ping, L.-B. Chen, and H. Wang, “Protection by and anti-oxidant mechanism of berberine against rat liver fibrosis induced by multiple hepatic toxic factors,” Clinical and Experimental Pharmacology and Physiology, vol. 35, no. 3, pp. 303–309, 2008.

[14] F. Di Pierro, N. Villanova, F. Agostini et al., “Pilot study on the additive effects of berberine and oral type 2 diabetes agents for patients with suboptimal glycemic control,” Diabetes, Metabolic Syndrome and Obesity, vol. 5, pp. 213–217, 2012.

[15] H. Zhang, J. Wei, R. Xue et al., “Berberine lowers blood glucose in type 2 diabetes mellitus patients through increasing insulin receptor expression,” Metabolism, vol. 59, no. 2, pp. 285–292, 2010.

[16] G. Marazzi, L. Cacciotti, F. Pelliccia et al., “Long-term effects of nutraceuticals (berberine, red yeast rice, policosanol) in elderly hypercholesterolemic patients,” Advances in therapy, vol. 28, no. 12, pp. 1105–1113, 2010.

[17] S. F. Previs, D. J. Withers, J.-M. Ren, M. F. White, and G. I. Shulman, “Contrasting effects of IRS-1 versus IRS-2 gene disruption on carbohydrate and lipid metabolism in vivo,” Journal of Biological Chemistry, vol. 275, no. 50, pp. 38990–38994, 2000.

[18] Y. Chen, Y. Wang, J. Zhang et al., “Berberine improves glucose homeostasis in streptozotocin-induced diabetic rats in association with multiple factors of insulin resistance;” ISRN Endocrinology, vol. 2011, Article ID 519371, 8 pages, 2011.

[19] N. Turner, J.-Y. Li, A. Gosby et al., “Berberine and its more biologically available derivative, dihydroberberine, inhibit mitochondrial respiratory complex i: a mechanism for the action of berberine to activate amp-activated protein kinase and improve insulin action,” Diabetes, vol. 57, no. 5, pp. 1414–1418, 2008.

[20] J. I. Odegaard and A. Chawla, “Pleiotropic actions of insulin resistance and inflammation in metabolic homeostasis,” Science, vol. 339, no. 6166, pp. 172–177, 2013.

[21] M. Qatanani and M. A. Lazar, “Mechanisms of obesity-associated insulin resistance: many choices on the menu;” Genes and Development, vol. 21, no. 12, pp. 1443–1455, 2007.

[22] V. T. Samuel and G. I. Shulman, “Mechanisms for insulin resistance: common threads and missing links;” Cell, vol. 148, no. 5, pp. 852–871, 2012.

[23] G. S. Hotamisligil, “Endoplasmic reticulum stress and the inflammatory basis of metabolic disease;” Cell, vol. 140, no. 6, pp. 900–917, 2010.

[24] S.-H. Leng, F.-E. Lu, and L.-J. Xu, “Therapeutic effects of berberine in impaired glucose tolerance rats and its influence on insulin secretion,” Acta Pharmacologica Sinica, vol. 25, no. 4, pp. 496–502, 2004.

[25] T. Lou, Z. Zhang, Z. Xi et al., “Berberine inhibits inflammatory response and ameliorates insulin resistance in hepatocytes,” Inflammation, vol. 34, no. 6, pp. 659–667, 2011.

[26] Z.-S. Wang, F.-E. Lu, L.-J. Xu, and H. Dong, “Berberine reduces endoplasmic reticulum stress and improves insulin signal transduction in Hep G2 cells,” Acta Pharmacologica Sinica, vol. 31, no. 5, pp. 578–584, 2010.

[27] W.-J. Kong, H. Zhang, D.-Q. Song et al., “Berberine reduces insulin resistance through protein kinase C-dependent up-regulation of insulin receptor expression,” Metabolism, vol. 58, no. 1, pp. 109–119, 2009.

[28] L.-J. Xing, L. Zhang, T. L. Liu, Y.-Q. Hua, P.-Y. Zheng, and G. Ji, “Berberine reducing insulin resistance by up-regulating IRS-2 mRNA expression in nonalcoholic fatty liver disease (NAFLD) rat liver,” European Journal of Pharmacology, vol. 668, no. 3, pp. 467–471, 2011.

[29] Z. Cheng, T. Pang, M. Gu et al., “Berberine-stimulated glucose uptake in L6 myotubes involves both AMPK and p38 MAPK,” Biochimica et Biophysica Acta, vol. 1760, no. 11, pp. 1682–1689, 2006.

[30] Y. S. Lee, W. S. Kim, K. H. Kim et al., “Berberine, a natural plant product, activates AMP-activated protein kinase with beneficial metabolic effects in diabetic and insulin-resistant states,” Diabetes, vol. 55, no. 8, pp. 2256–2264, 2006.

[31] W. Chang, M. Zhang, J. Li et al., “Berberine improves insulin resistance in cardiomyocytes via activation of S'-adenosine monophosphate-activated protein kinase;” Metabolism, 2013.

[32] V. Mikes and L. S. Yaguzhinskij, “Interaction of fluorescent berberine alkyl derivatives with respiratory chain of rat liver mitochondria,” Journal of Bioenergetics and Biomembranes, vol. 17, no. 1, pp. 23–32, 1985.

[33] D.-Y. Lu, C.-H. Tang, Y.-H. Chen, and I.-H. Wei, “Berberine suppresses neuroinflammatory responses through AMP-activated protein kinase activation in BV-2 microglia,” Journal of Cellular Biochemistry, vol. 110, no. 3, pp. 697–705, 2010.
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[34] X. Xia, J. Yan, Y. Shen et al., “Berberine improves glucose metabolism in diabetic rats by inhibition of hepatic gluconeogenesis,” *PLoS ONE*, vol. 6, no. 2, Article ID e6566, 2011.

[35] L. H. Young, “A crystallized view of AMPK activation,” *Cell Metabolism*, vol. 10, no. 1, pp. 5–6, 2009.

[36] N. Shen, Y. Huan, and Z. F. Shen, “Berberine inhibits mouse insulin gene promoter through activation of AMP activated protein kinase and may exert beneficial effect on pancreatic beta-cell,” *European Journal of Pharmacology*, vol. 694, no. 1-3, pp. 120–126, 2012.

[37] X. Liu, G. Li, H. Zhu et al., “Beneficial effect of berberine on hepatic insulin resistance in diabetic hamsters possibly involves in SREBP1, LXRα and PPARα transcriptional programs,” *Endocrine Journal*, vol. 57, no. 10, pp. 881–893, 2010.

[38] Y. Li, S. Xu, M. M. Mihaylova et al., “AMPK phosphorylates and inhibits SREBP activity to attenuate hepatic steatosis and atherosclerosis in diet-induced insulin-resistant mice,” *Cell Metabolism*, vol. 13, no. 4, pp. 376–388, 2011.

[39] H. Li, B. Dong, S. W. Park, H.-S. Lee, W. Chen, and J. Liu, “Hepatocyte nuclear factor 1α plays a critical role in PCSK9 gene transcription and regulation by the natural hypocholesterolemic compound berberine,” *Journal of Biological Chemistry*, vol. 284, no. 42, pp. 28885–28895, 2009.

[40] E. Bugianesi, N. Leone, E. Vanni et al., “Expanding the natural history of nonalcoholic steatohepatitis: from cryptogenic cirrhosis to hepatocellular carcinoma,” *Gastroenterology*, vol. 123, no. 1, pp. 134–140, 2002.

[41] K. Rombouts and F. Marra, “Molecular mechanisms of hepatic fibrosis in non-alcoholic steatohepatitis,” *Dietetic Diseases*, vol. 28, no. 1, pp. 229–235, 2010.

[42] K. Miura, E. Seki, H. Ohnishi, and D. A. Brenner, “Role of toll-like receptors and their downstream molecules in the development of nonalcoholic fatty liver disease,” *Gastroenterology Research and Practice*, vol. 2010, Article ID 362847, 9 pages, 2010.

[43] Y. Yang, J. Qi, Q. Wang et al., “Berberine suppresses Th17 and dendritic cell responses,” *Investigative Ophthalmology & Visual Science*, vol. 54, no. 4, pp. 2516–2522, 2013.

[44] G. Derosa, A. D. Angelo, A. Bonaventura et al., “Effects of berberine on lipid profile in subjects with low cardiovascular risk,” *Expert Opinion on Biological Therapy*, vol. 13, no. 4, pp. 475–482, 2013.

[45] W. Xie, D. Gu, J. Li, K. Cui, and Y. Zhang, “Effects and action mechanisms of berberine and rhizoma coptidis on gut microbes and obesity in high-fat diet-fed C57BL/6j mice,” *PLoS ONE*, vol. 6, no. 9, Article ID e24520, 2011.

[46] A.-W. Feng, W. Gao, G.-R. Zhou et al., “Berberine ameliorates COX-2 expression in rat small intestinal mucosa partially through PPARy pathway during acute endotoxemia,” *International Immunopharmacology*, vol. 12, no. 1, pp. 182–188, 2012.

[47] F. Bäckhed, H. Ding, T. Wang et al., “The gut microbiota as an environmental factor that regulates fat storage,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 101, no. 44, pp. 15718–15723, 2004.

[48] L. Gu, N. Li, Q. Li et al., “The effect of berberine in vitro on tight junctions in human Caco-2 intestinal epithelial cells,” *Fitoterapia*, vol. 80, no. 4, pp. 241–248, 2009.

[49] L. Gu, N. Li, J. Gong, Q. Li, W. Zhu, and J. Li, “Berberine ameliorates intestinal epithelial tight-junction damage and down-regulates myosin light chain kinase pathways in a mouse model of endotoxemia,” *Journal of Infectious Diseases*, vol. 203, no. 11, pp. 1602–1612, 2011.

[50] H.-M. Li, Y.-Y. Wang, H.-D. Wang et al., “Berberine protects against lipopolysaccharide-induced intestinal injury in mice via alpha 2 adrenoceptor-independent mechanisms,” *Acta Pharmacologica Sinica*, vol. 32, no. 11, pp. 1364–1372, 2011.

[51] M. Sundaram and Z. Yao, “Recent progress in understanding protein and lipid factors affecting hepatic VLDL assembly and secretion,” *Nutrition and Metabolism*, vol. 7, article 35, 2010.

[52] H. Dong, Y. Zhao, L. Zhao et al., “The effects of berberine on blood lipids: a systemic review and meta-analysis of randomized controlled trials,” *Planta Medica*, vol. 79, no. 6, pp. 437–446, 2013.

[53] X. Xiao, Q. Zhang, K. Feng et al., “Berberine moderates glucose and lipid metabolism through multipathway mechanism,” *Evidence-Based Complementary and Alternative Medicine*, vol. 2011, Article ID 924851, 10 pages, 2011.

[54] W.-J. Kong, J. Wei, Z.-Y. Zuo et al., “Combination of simvastatin with berberine improves the lipid-lowering efficacy,” *Metabolism*, vol. 57, no. 8, pp. 1029–1037, 2008.

[55] Y. X. Wang, W. J. Kong, Y. H. Li et al., “Synthesis and structure-activity relationship of berberine analogues in LDLR up-regulation and AMPK activation,” *Bioorganic & Medicinal Chemistry*, vol. 20, no. 22, pp. 6552–6558, 2012.

[56] P. Abidi, Y. Zhou, J.-D. Jiang, and J. Liu, “Extracellular signal-regulated kinase-dependent stabilization of hepatic low-density lipoprotein receptor mRNA by herbal medicine berberine,” *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 25, no. 10, pp. 2170–2176, 2005.

[57] J. L. Goldstein, R. A. DeBoe-Boyd, and M. S. Brown, “Protein sensors for membrane sterols,” *Cell*, vol. 124, no. 1, pp. 35–36, 2006.

[58] Q.-H. Yang, S.-P. Hu, Y.-P. Zhang et al., “Effect of berberine on expressions of uncoupling protein-2 mRNA and protein in hepatic tissue of non-alcoholic fatty liver disease in rats,” *Chinese Journal of Integrative Medicine*, vol. 17, no. 3, pp. 205–211, 2011.

[59] T. Lao-ong, W. Chatuphonprasert, N. Nemoto et al., “Alteration of hepatic glutathione peroxidase and superoxide dismutase expression in streptozotocin-induced diabetic mice by berberine,” *Pharmacological Biology*, vol. 50, no. 8, pp. 1007–1012, 2012.

[60] X. Zhu, X. Guo, G. Mao et al., “Hepatoprotection of Berberine against hydrogen peroxide-induced apoptosis by upregulation of SirTuin 1,” *Phytotherapy Research*, vol. 27, no. 3, pp. 417–421, 2013.

[61] J. Yang, J. Yin, H. Gao et al., “Berberine improves insulin sensitivity by inhibiting fat store and adjusting adipokines profile in human preadipocytes and metabolic syndrome patients,” *Evidence-Based Complementary and Alternative Medicine*, vol. 2012, Article ID 363845, 9 pages, 2012.

[62] X. Ma, T. Egawa, H. Kimura et al., “Berberine-induced activation of 5′-adenosine monophosphate-activated protein kinase and glucose transport in rat skeletal muscles,” *Metabolism*, vol. 59, no. 11, pp. 1619–1627, 2010.

[63] Y.-C. Linn, J. Lu, L.-C. Lim et al., “Berberine-induced haemolysis of Rhizoma coptidis and cortex phellodendri against lipopolysaccharide-induced intestinal injury in mice via alpha 2 adrenoceptor-independent mechanisms,” *Acta Pharmacologica Sinica*, vol. 32, no. 11, pp. 1364–1372, 2011.
cytotoxicity in healthy Hepatocytes,” *PLoS ONE*, vol. 6, no. 6, Article ID e21416, 2011.

[66] K. H. Janbaz and A. H. Gilani, “Studies on preventive and curative effects of berberine on chemical-induced hepatotoxicity in rodents,” *Fitoterapia*, vol. 71, no. 1, pp. 25–33, 2000.

[67] J.-M. Hwang, C.-J. Wang, E.-P. Chou et al., “Inhibitory effect of berberine on tert-butyl hydroperoxide-induced oxidative damage in rat liver,” *Archives of Toxicology*, vol. 76, no. 11, pp. 664–670, 2002.

[68] M. Zhang, X. Lv, J. Li et al., “Sodium caprate augments the hypoglycemic effect of berberine via AMPK in inhibiting hepatic gluconeogenesis,” *Molecular and Cellular Endocrinology*, vol. 363, no. 1-2, pp. 122–130, 2012.

[69] X.-Y. Lv, J. Li, M. Zhang et al., “Enhancement of sodium caprate on intestine absorption and antidiabetic action of berberine,” *AAPS PharmSciTech*, vol. 11, no. 1, pp. 372–382, 2010.

[70] W. Zha, G. Wang, W. Xu et al., “Inhibition of P-glycoprotein by HIV protease inhibitors increases intracellular accumulation of berberine in murine and human macrophages,” *PLoS ONE*, vol. 8, no. 1, Article ID e54349, 2013.