Astrocyte elevated gene-1 (AEG-1) induces epithelial-mesenchymal transition in lung cancer through activating Wnt/β-catenin signaling

Welling He¹, Shanyang He²†, Zuo Wang³, Hongwei Shen², Wenfeng Fang⁴, Yang Zhang⁵, Wei Qian⁵, Millicent Lin⁶, Jinglun Yuan⁶, Jinyang Wang⁷, Wenhua Huang⁷, Liantang Wang³ and Zunfu Ke³*

Abstract

Background: Non-small cell lung cancer (NSCLC) is a highly metastatic cancer with limited therapeutic options, so development of novel therapies that target NSCLC is needed. During the early stage of metastasis, the cancer cells undergo an epithelial-mesenchymal transition (EMT), a phase in which Wnt/β-catenin signaling is known to be involved. Simultaneously, AEG-1 has been demonstrated to activate Wnt-mediated signaling in some malignant tumors.

Methods: Human NSCLC cell lines and xenograft of NSCLC cells in nude mice were used to investigate the effects of AEG-1 on EMT. EMT or Wnt/β-catenin pathway-related proteins were characterized by western blot, immunofluorescence and immunohistochemistry.

Results: In the present study, we demonstrated that astrocyte elevated gene-1 (AEG-1) ectopic overexpression promoted EMT, which resulted from the down-regulation of E-cadherin and up-regulation of Vimentin in lung cancer cell lines and clinical lung cancer specimens. Using an orthotopic xenograft-mouse model, we also observed that AEG-1 overexpression in human carcinoma cells led to the development of multiple lymph node metastases and elevated mesenchymal markers such as Vimentin, which is a characteristic of cells in EMT. Furthermore, AEG-1 functioned as a critical protein in the regulation of EMT by directly targeting multiple positive regulators of the Wnt/β-catenin signaling cascade, including GSK-3β and CKIδ. Notably, overexpression of AEG-1 in metastatic cancer tissues was closely associated with poor survival of NSCLC patients.

Conclusions: These results reveal the critical role of AEG-1 in EMT and suggest that AEG-1 may be a prognostic biomarker and its targeted inhibition may be utilized as a novel therapy for NSCLC.

Keywords: AEG-1, Epithelial-mesenchymal transition, Non-small cell lung cancer, Wnt, β-catenin

Background

Lung cancer is the most common malignant tumor in the world, and the leading cause of cancer-related death in human beings [1]. Despite the achievements made in diagnosis and treatment in the recent years, the prognosis of lung cancer patients is still poor and their overall 5-year survival rate is 15% [2]. Although the clinical stage at diagnosis is the key prognostic determinant for lung cancer survival [3], considerable variability in reoccurrence and survival is commonly observed in patients with a similar stage. Thus, the initial diagnosis is extremely important because it could reduce the mortality rate for lung cancer patients [4].

The progress of cancer metastasis depends on the unique mechanisms of cancer cells evading from the primary tissue and spreading into surrounding tissues. Molecular reprogramming, as a part of the epithelial–mesenchymal transition (EMT), is considered to be a crucial step in the metastasis process of most carcinomas [5]. During metastatic progression, EMT drives primary epithelial-like tumour cells to acquire invasive potential, such as increased motility and mesenchymal characteristics, triggering dissemination from the tumor and infiltration into
the tumor vessel. Then, the EMT-driven cells circulating in the blood flow redifferentiate into primary status via MET during colonization and growth at distant metastatic sites [6,7]. Because of EMT’s role in the metastatic process, controlling EMT progress and progression in tumors is now thought to be a promising strategy to inhibit metastasis and to prolong cancer patients’ survival.

Astrocyte-elevated gene-1 (AEG-1), also known as LYRIC (lysine-rich CEACAM1) or metadherin, is originally induced in primary human fetal astrocytes [8]. Recently, numerous reports demonstrated that AEG-1 might play a pivotal role in the pathogenesis, progression, invasion, metastasis and overall patient survival in diverse human cancers [9-12]. This evidence indicates that the upregulation of AEG-1 contributes to malignant progression [13]. Furthermore, AEG-1 overexpression can facilitate migration and invasion of human glioma cells [14], as well as activate Wnt/β-catenin signaling via ERK42/44 activation [11]. Although AEG-1 is an oncogene that has been implicated in pathways critical to lung cancer carcinogenesis [15], AEG-1 was also found to control the expression of E-cadherin and Vimentin [16]. The above findings suggest that AEG-1 may mediate the metastasis of lung carcinoma through the regulation of EMT.

In this study, we concentrated on elucidating the role of AEG-1 in EMT of NSCLC. We demonstrated that up-regulation of AEG-1 was significantly associated with lymph node metastasis and EMT status of NSCLC. We further investigated that AEG-1 could activate Wnt/β-catenin signaling by inducing GSK-3β (glycogensynthasekinase 3β) phosphorylation via CKIδ (casein kinase 1δ), consequently enhancing EMT status.

**Methods**

**Cell culture and tissue specimen selection**

Lung cancer cell lines, including NCI-H226, NCI-H460, L-78, A549 and Slu-01, were maintained in Dulbecco’s modified Eagle’s medium (DMEM; Invitrogen, USA) supplemented with 10% fetal bovine serum (HyClone, Logan, UT). AEG-1 overexpression plasmid pcDNA3.1-AEG-1, β-catenin overexpression plasmid pcDNA3.1-β-catenin, AEG-1 siRNA and CKIδ siRNA (RiboBio, China) were transiently transfected using Lipofectamine 2000 (Invitrogen, USA).

A total of 210 cases from 2000 to 2005 coded as “lung cancer” were collected consecutively from the pathology archives of the Affiliated First Hospital, Sun Yat-sen University. The medical ethics committee of Sun Yat-sen University approved the present retrieval of cancer specimens and the connection with clinical data from our institute.

**Migration assay**

Invasive ability was measured by using 24-well BioCoat cell culture inserts (Costar, New York, NY, USA) with an 8-μm-porosity polyethylene terephthalate membrane coated with Matrigel Basement Membrane Matrix (Cultrex, MD, USA). At the end of the assay, cells that did not migrate or invade through the pores were removed with a cotton swab. The invasion ability was determined by counting the cells that migrated to the lower side of the filter.

**Western blot and immunofluorescence**

Western blot was carried out as described earlier [17]. Blotted membranes were incubated with the antibodies for AEG-1(Invitrogen, USA), Twist 1, E-cadherin, Vimennt, β-catenin, p-GSK-3β(Ser-9), GSK-3β, CKIδ and GAPDH (Abcam, Cambridge, UK) in 5% milk/TFBS (tris-buffered saline Tween-20). For immunofluorescence microscopy, cells grown on chamber slides were probed with AEG-1 E-cadherin, Vimennt and β-catenin. The fluorescein isothiocyanate (FITC)-conjugated or rhoda-mine-conjugated anti-IgG was purchased from Molecular Probes. Cells were visualized in an Olympus BX51 fluorescence microscope (Olympus, Tokyo, Japan).

**Total RNA extraction and real-time RT-PCR**

Total RNA was extracted using the RNAeasy kit (Qiagen, USA). The amplification was carried out in a total volume of 20 μL containing LightCycler FastStart DNA Master SYBR green I (Roche, USA). Ct value (initial amplification cycle) of each standard dilution was plotted against standard cDNA copy numbers. On the basis of the standard curves for each gene, the sample cDNA copy number was calculated according to the sample Ct value. Standard curves and PCR results were analyzed using ABI7000 software (Applied Biosystems, Foster City, CA, USA). Primers were β-catenin: (sense) 5′GTTCGGTTCCGGCTGTTA 3′, (antisense)5′TTTCTCCCTTGGCCCATC 3′ and AEG-1: (sense) 5′CGAGAAGCCCAAACCAAAATG 3′, (antisense) 5′TGGTGCGTCTTTGCTGT 3′. β-actin (primers: sense 5′ GCATGTTGCTAGAAGAGATTC 3′, antisense 5′ TCAGGCTTGGTCAAGC 3′) was used as an internal control.

**Immunoprecipitation**

For immunoprecipitation, all of the procedures were done at 4°C. Transfected Slu-01 cells were washed twice with cold PBS and rinsed in 1.5 mL of cold lysis buffer for 20 min on ice. After preclearing, 1 mg of total protein was incubated with antibody, AEG-1-GSK-3β, or CKIδ. An equal concentration of sheep (Upstate Cell Signalling Solutions), mouse, or rabbit (Vector laboratories) immunoglobulin was used as controls. The immunocomplexes were subjected to Western blot analysis according to the manufacturer’s protocol.
Luciferase reporter gene assay
For the reporter gene assay, cells seeded in 24-well plates were transfected with the firefly luciferase reporter gene construct (TOP or FOP; 200 ng), and 1 ng of pRL-SV40 Renilla luciferase (as an internal control). Cell extracts were prepared 24 hours after transfection, and luciferase activity was measured using the Dual-Luciferase Reporter Assay System (Promega, USA).

Analysis of the Wnt signaling pathway
Wnt-3a-conditioned medium (Wnt-3a-CM) was produced from L cells transfected with pGKWnt-3a. The medium was centrifuged at 1,000 g for 15 min and filtered through a nitrocellulose membrane. Then, cells were treated with Wnt-3a CM for 24 hours, and Wnt signaling was monitored by various assays, including Western blotting and luciferase reporter gene assays.

Immunohistochemical staining and evaluation
Sections (4 \( \mu \text{m} \)) of formalin-fixed, paraffin-embedded tissues were made using a rotary microtome (Leica, Wetzlar, Germany) and labeled with anti-AEG-1 (Abcam, Cambridge, UK), anti-E-cadherin (Abcam, Cambridge, UK) and anti-Vimentin (Abcam, Cambridge, UK) primary antibodies. We used the known positive slice in the SP kit (Maxim-Bio, Fuzhou, China) as a positive control. The number of immunopositive cells was semiquantitatively estimated. The staining index was calculated using Aperio Image-Scope software (Aperio Technologies).

In vivo orthotopic xenograft studies in athymic nude mice
Male nude mice (about 8 weeks of age) were anesthetized with sodium pentobarbital (50 mg/kg) in a sterile environment. A small skin incision to the right chest wall was made approximately 5 mm to the tail side of the scapula. Then, Slu-01 (5 × 10^6) or Slu-01/AEG-1-expressing cells (5 × 10^6; Slu-01 cells stably transfected with the human AEG-1 complementary DNA) were implanted into the mice. The skin incision was sutured using metallic clips, which were removed on day 16 after the operation. Different time after inoculation, the mice were killed, tumors were weighed and measured, and tumor tissues were fixed in 10% neutral buffered formalin for the immunohistochemical study. For H&E staining, deparaffinized tissue sections were stained with Mayer hematoxylin and eosin solution. Tumor growth and local metastasis were monitored by an IVIS Imaging System (Xenogen). Images and bioluminescent signals were analyzed using Living Image and Xenogen software. All experimental projects were approved by the medical ethics committee of Sun Yat-sen University.

Statistical analysis
All above experiments were performed at least three times. Statistical analysis was carried out using software SPSS (version 16.0; SPSS, Chicago, IL, USA). Unpaired two-tailed Student's \( t \)-test was used to determine the statistical relevance between groups. Survival curves were plotted using the Kaplan-Meier method and compared with the log-rank test. ROC curve analysis was conducted to determine the cutoff point of high or low AEG-1 level and EMT status. Values of \( P < 0.05 \) were considered statistically significant.

Results
AEG-1 is closely correlated with EMT status in vitro
To investigate the role of AEG-1 expression in lung cancer, we comparatively analyzed AEG-1 protein profiles in lung cancer cell lines with different metastatic ability. As shown in Figure 1A, Western blot analysis revealed that AEG-1 protein was highly expressed in NCI-H226 cells (from lung squamous cell carcinoma with high metastatic ability), whereas Slu-01 cells (from lung adenocarcinoma with low metastatic ability) had undetectable AEG-1 protein expression. In cell lines (with middle metastatic ability) such as NCI-H460, L-78 and A549, the expression levels of AEG-1 protein were significantly lower than that of NCI-H226 cells, but higher than that of Slu-01 cells. We also showed that NCI-H226 cells expressed high levels of Twist1, Vimentin and E-cadherin, but low level of E-cadherin, while Slu-01 cells displayed the opposite expression pattern (Figure 1B and C). These results indicated that AEG-1 might be associated with the metastasis process of lung cancer.

In addition, of particular note was the fact that AEG-1 could regulate EMT. EMT may aberrantly take place in epithelial neoplasms, leading to the loss of cell polarity, cell-to-cell contact and enhanced cell motility. At the early metastatic stage of tumors, EMT is characterized by the loss of E-cadherin expression, an increase in motility, invasive potential and mesenchymal characteristics such as Vimentin. These phenotypic changes were also observed in NCI-H226 cells transfected with AEG-1 siRNA, which displayed a clear morphological transition from spindle-like fibroblastic (vector control) to cobblestone-like cells with well-organized cell contact and polarity (Figure 1D). The transfection of AEG-1 siRNA resulted in an increase of E-cadherin and a decrease of Vimentin expression in NCI-H226 cells (Figure 1B). On the contrary, AEG-1 overexpression in Slu-01 cells led to a spindle- or star-like morphology in the culture media, as well as a decrease of E-cadherin and an increase of Vimentin expression (Figure 1B and D). These results strongly suggest that AEG-1 may promote a transition from epithelial to mesenchymal phenotype. Matrigel-coated transwell assay also
showed that AEG-1 overexpression could significantly enhance cell motility in vitro (Figure 1E).

AEG-1 promotes β-catenin nuclear translocation and Wnt/β-catenin signaling mediates AEG-1–induced EMT

Based on the critical role of the Wnt/β-catenin pathway in metastasis, we then explored whether AEG-1 activates Wnt/β-catenin signaling and if the Wnt/β-catenin pathway mediates AEG-1–induced EMT. In the canonical Wnt/β-catenin pathway, the hallmark of Wnt signaling activation is β-catenin’s nuclear translocation, where it forms a complex with a specific T-cell factor/lymphoid enhancer factor (Tcf/Lef) [18]. After up-regulating AEG-1 expression in Slu-01 cells with pcDNA3.1-AEG-1, we observed a substantial accumulation of β-catenin in the nucleus, suggesting that AEG-1 might contribute to the activation of Wnt signaling (Figure 2A). However, the total β-catenin mRNA level did not change significantly after AEG-1 overexpression in Slu-01 cells (Figure 2B). As expected, luciferase assays also demonstrated that AEG-1 overexpression noticeably increased the transcriptional activity of β-catenin/TCF in Slu-01 cells, as determined by the β-catenin reporter system (TOP/FOP) (Figure 2C). In contrast, transfection of AEG-1 siRNA could reduce the β-catenin/TCF transcriptional activity in NCI-H226 cells (Figure 2C).

If we treated NCI-H226 cells transfected with AEG-1 siRNA with pcDNA3.1-β-catenin, it could restore the EMT status of NCI-H226 cells, as determined by EMT-related marker expression and morphology (Figure 2D and E). Furthermore, elevated expression of β-catenin protein levels in NCI-H226/AEG-1-siRNA cells induced EMT in a dose-dependent manner (Figure 2E and F). However, the opposite phenomenon appeared in the Slu-01/AEG-1 cells with corresponding morphology and EMT-related marker changes (Figure 2D and F).
AEG-1 interacts with Gsk-3β and CKIδ to activate Wnt/β-catenin

We then investigated the molecular mechanism by which AEG-1 activates Wnt/β-catenin signaling. In the absence of Wnt signaling, cytoplasmic β-catenin undergoes sequential phosphorylation, first at Ser45 (β-catenin) by casein kinase I (CKI) and then at Ser33,37/Thr41 by glycogen synthase kinase (GSK)-3β, leading to targeted ubiquitination through E3 ubiquitin ligase. In Slu-01 cells transfected with pcDNA3.1-AEG-1, immunoprecipitation experiments and Western blot analysis revealed that AEG-1 appeared to directly associate with GSK-3β and promote its phosphorylation at Ser9 (Figure 3A). In addition, co-immunoprecipitation results showed that AEG-1 could form a complex with both GSK-3β and CKIδ (Figure 3B). Moreover, after Slu-01/AEG-1 cells were treated with CKIδ-siRNA, CKIδ-siRNA treatment abolished AEG-1-mediated phosphorylation of GSK-3β at Ser9 and EMT (Figure 3C).

AEG-1 promotes Wnt/β-catenin-mediated EMT through inactivating GSK-3β

Wnt/β-catenin signaling has been demonstrated to participate in the EMT process during embryonic development...
and cancer progression; however, the involvement of AEG-1 in Wnt/β-catenin-mediated EMT has not been completely defined. To address this question, we tested whether manipulating AEG-1 levels in various cell lines would be able to convert the mesenchymal phenotypes. Whereas Wnt-3a only slightly induced EMT in NCI-H226, AEG-1 depletion notably elicited a change in NCI-H226 cells from the mesenchymal phenotype to an epithelial phenotype as manifested by increased expression of the epithelial marker E-cadherin concomitant with a down-regulation of the mesenchymal marker Vimentin (Figure 4A).

Similarly, the knockdown of AEG-1 also resulted in the decrease of the p-GSK-3β level and reduced β-catenin/TCF transcriptional activity (Figure 4B). In contrast, restoring AEG-1 expression in Slu-01 cells (AEG-1-negative cell) reinforced Wnt/β-catenin-induced EMT and led to the increase of the p-GSK-3β level and β-catenin/TCF transcriptional activity (Figure 4C and D), strongly suggesting that AEG-1 is a promoter of Wnt/β-catenin-mediated EMT.

**AEG-1 increases distant metastasis in vivo by the regulation of EMT**

Because Slu-01 cells are of low metastatic potential and decreased AEG-1 expression, and show EMT inhibition status (Figure 1), we then observed the prometastatic trait of AEG-1 up-regulation in Slu-01 cells versus its corresponding vector control cells using an orthotopic mouse model. Stable luciferase activity ensured that every group had an equal level of AEG-1 expression before the injection of Slu-01/AEG-1 cells. Bioluminescent imaging (BLI) was utilized to monitor tumor growth and the onset of metastases dynamically. Strikingly, mice injected with Slu-01/AEG-1 cells displayed multiple distant metastatic lesions at various sites, whereas less metastasis lesions were found in mice injected with control Slu-01 cells.

**Figure 3** AEG-1 was associated with CKIδ and modulated the GSK-3β/β-catenin signaling pathway. (A) Slu-01 cells stably overexpressing AEG-1 were established. GSK-3β was immunoprecipitated from cell lysates, and its expression was confirmed by immunoblotting with the indicated antibodies. (B) CKIδ was critical for AEG-1-mediated regulation of GSK-3β/β-catenin signaling and EMT. AEG-1 complexes were associated with both GSK-3β and CKIδ. (C) CKIδ played a role in AEG-mediated regulation of EMT and Ser-9 phosphorylation of GSK-3β. Slu-01 cells transfected with pcDNA3.1-AEG-1 were co-transfected with CKIδ-specific siRNA. Cell lysates were then subjected to Western blotting.
Our data also showed that Slu-01/AEG-1 xenotransplants approximately generated a 4-fold increase in the number of distant metastases than that of vector control cells (Figure 5B), which was verified by H&E staining (Figure 5C). To further validate the fact that AEG-1 enhanced metastasis in vivo by regulating EMT status, immunohistochemistry (IHC) was applied to detect the expression characteristics of EMT-related molecular markers. Immunohistochemistry (IHC) revealed that the majority of tumor cells in Slu-01/AEG-1 xenotransplants strongly expressed Vimentin, but exhibited weak staining of E-cadherin (Figure 5C).

**AEG-1 promotes metastasis in lung cancer patients**

To further understand the clinical relevance of the above findings, we examined the relationship between AEG-1 expression and EMT markers in lung cancer patients. Patients from different clinical stages were first divided into two groups according to H&E staining (Figure 6A) and positron emission tomography/computed tomography (PET/CT) (Figure 6B): the primary site of cancer with metastasis and the primary site of non-metastasizing cancer, respectively. Based on the TNM (Tumor node metastasis) staging system, we selected six patients from stage I and stage IV. As shown in Figure 6C, the expression levels of AEG-1 were significantly elevated in patients with distant metastasis, compared to that in primary tumors without detectable distant metastasis. Furthermore, up-regulation of AEG-1, Vimentin, p-GSK-β, and β-catenin levels, as well as suppression of E-cadherin, were clearly observed in tissues from patients with distant metastasis (Figure 6C). In all six examined samples, there was a significantly positive correlation between the levels of AEG-1 and Vimentin and an inverse correlation between the levels of AEG-1 and E-cadherin. These data indicate that AEG-1 plays a pivotal role in lung cancer EMT and metastasis in vivo, which is consistent with our in vitro data from various cancer cell lines.

**Prognostic value of AEG-1 and EMT status in lung cancer patients**

To explore the prognostic value of AEG-1 in patients, we used the Kaplan-Meier method to evaluate the relationship between the survival curve and AEG-1 expression, as well as EMT status. Survival analysis data indicated a significantly inverse correlation between AEG-1 protein expression level and the overall survival time (p < 0.001), clearly disclosing that higher levels of AEG-1 expression...
were associated with shorter survival time. As shown in Figure 7A, the cumulative 5-year survival rate was 37.8% (95% CI: 25.8%–49.8%) in the AEG-1 low expression group, whereas it was only 5.3% (95% CI: 4.2%–6.4%) in the AEG-1 high expression group.

In addition, when we combined the expression status of AEG-1 and EMT, the difference of overall survival rate between AEG-1(+)/EMT(−) and AEG-1(+)/EMT(+) was significant (p < 0.001, Figure 7B), and it was similar to the result between AEG-1(−)/EMT(−) and AEG-1

---

**Figure 5** AEG-1 promoted tumor metastasis in vivo. (A) Representative BLI images of mice bearing Slu-01/AEG-1-expressing tumors with metastatic lesions. Mice (n = 15) were imaged six weeks later to determine local tumor growth and metastasis. (B) Number of metastatic nodules or distant metastasis in individual dead mouse bearing con or Slu-01/AEG-1-expressing tumors. (C) AEG-1 overexpression in Slu-01 cells promoted EMT in athymic nude mice in vivo. H&E staining showed primary tumors without detectable metastasis in control mice and the lymph node metastases in mice bearing Slu-01/AEG-1-expressing tumors two weeks after injection (magnification, ×200). IHC showed that up-regulation of AEG-1 resulted in an increased in the expression of Vimentin and weak E-cadherin staining (magnification × 200).
(−)/EMT(+) (p < 0.001, Figure 7C). Simultaneously, AEG-1 level and EMT status, and their combined status were further analyzed by the receiver operating characteristic (ROC) method to assess their predictive value for death. As shown in Figure 7D, the combined status of AEG-1 and EMT predicted death with better performance (p < 0.001). The area under the curve for both AEG-1 and EMT status was 0.8394 (95% CI: 0.7361–0.9428), which was larger than that of AEG-1 or EMT status, with areas under the curve of 0.7248 (95% CI: 0.6504–0.7992) and 0.7145 (95% CI: 0.6002–0.8287), respectively (both p < 0.001) (Figure 7D).

To further illustrate the clinical significance of the above findings in human lung cancer, AEG-1 mRNA expression was examined in 53 lung cancer tissue specimens. Patients were first divided into two groups: those with distant metastasis and those without metastasis during the follow-up period, respectively. As shown in Figure 7E, AEG-1 mRNA expression level was significantly up-regulated in 23 patients with distant metastasis, compared to that of 30 patients without detectable distant metastasis. The above results suggest a strong correlation between AEG-1 and distant metastasis.

Discussion
Cancer metastasis is a complex, multistep process involving the escape of neoplastic cells from a primary tumor (local invasion), the intravasation into the systemic circulation, the establishment of micrometastases, and ultimately, the outgrowth of macroscopic secondary tumors [19]. Metastasis is the leading cause of cancer-related deaths worldwide, particularly in NSCLC. Thus, there is an urgent need for the identification of metastatic factors and understanding of the molecular mechanisms underlying NSCLC. AEG-1 is a novel oncoprotein essential for malignant progression in various types of human cancers [9,13,20-23]. Brown et al. pointed out that AEG-1 expression in HEK293T cells enhanced lung localization of the cells. In addition, the knockdown of AEG-1 or anti-AEG-1 antibody inhibited lung metastasis of 4T1 cells [24]. However, the role of AEG-1 in mediating lung cancer metastasis remains unknown. In our present study, Western blot analysis showed that AEG-1 levels were strikingly up-regulated in the pleura-metastatic derivatives of NCI-H226 lung cancer cell lines. Moreover, AEG-1 evidently induced nonmetastatic Slu-01 cells to invade and metastasize in vitro and promoted a dramatic increase in
the incidence of lymph node metastases in vivo. In addition, AEG-1 is significantly correlated with clinical stage, including stages of lymph node spread (an early stage of metastasis) and distant metastasis in breast cancer [21]. In hepatocellular carcinoma cells, expression of AEG-1 gradually increases from stage I to IV [9]. Our analysis also demonstrated that AEG-1 overexpression was closely correlated with metastatic recurrence in lung cancer patients. Thus, the above evidence provides new insight on the function of AEG-1 as a clinically relevant promoter of tumor metastasis.

EMT, a process by which epithelial cells acquire characteristics of mesenchymal cells, is largely thought to play an important role in invasion and metastasis [25]. In hepatocellular carcinoma cells, expression of AEG-1 gradually increases from stage I to IV [9]. Our analysis also demonstrated that AEG-1 overexpression was closely correlated with metastatic recurrence in lung cancer patients. Thus, the above evidence provides new insight on the function of AEG-1 as a clinically relevant promoter of tumor metastasis.

EMT, a process by which epithelial cells acquire characteristics of mesenchymal cells, is largely thought to play an important role in invasion and metastasis [25]. During EMT, epithelial cells lose their cell polarity and molecular characteristics, but gain migratory and invasive properties [26]. For example, cells undergoing EMT typically show both an increase in protein abundances of Vimentin and a decrease in E-cadherin [27]. These particular phenotypic changes were also observed in Slu-01 cells, which exhibited an obvious morphological transition from a rounded or cobblestone-shaped, epithelial-like morphology to spindle-shaped fibroblast with the loss of its cell polarity, cell–cell adhesive interactions and junctions when transfected with pcDNA3.1-AEG-1. Moreover, AEG-1 could up-regulate Vimentin and down-regulate E-cadherin expression levels in Slu-01 cells. In contrast, when endogenous AEG-1 expression was knocked down in NCl-H226 cells, EMT was clearly reversed. Furthermore, AEG-1 can enhance the Twist 1 expression, which is a potential EMT regulator. The association between EMT and cancer progression has been revealed in several types of cancer [28,29]. More importantly, a conversion from E-cadherin to N-cadherin showed strong and significant associations with prostate cancer progression [30]. However, less research has been done on the role of EMT in lung cancer. We further assessed the relationship between AEG-1 expression and EMT-related markers in lung cancer patients. The suppression of E-cadherin, as well as increase of Vimentin, AEG-1 and p-GSK-3β, was clearly observed in tissues from metastatic lymph nodes. In the orthotopic lung cancer animal model, mice bearing
Slu-01/AEG-1 cells also showed a significant increase in the number of lymph node metastases where cancer tissues clearly exhibited mesenchymal characteristics. These results strongly suggest that EMT should play a major role in the AEG-1-mediated metastasis of NSCLC.

Molecular mechanisms of EMT in lung cancer have been the most investigated fields. AEG-1 was found to control the expression level of Vimentin and E-cadherin. In some cancers, a mechanism involving the AEG-1-Vimentin interaction has been reported [16,31]. Recently, the role of Vimentin in EMT has also been reported in breast cancer cell lines, which was attributed to a mechanism involving the activation of AEG-1 [32]. In this study, we detected a typical EMT process induced by AEG-1 in NSCLC cells. However, EMT is regulated by various cell signaling pathways that originate from the tumor stroma, including TGF-β [33], Wnt/β-catenin, Hedgehog [35], Notch [36] and Ras-MAPK pathways [37]. Among these pathways, aberrant activation of Wnt/β-catenin signaling has been found in a wide range of cancers, especially in NSCLC [38]. Moreover, Wnt/β-catenin activation may induce EMT through its downstream targets: Twist, Snail and Slug. Previous studies have shown that Wnt/β-catenin signaling participates in EMT in numerous cancers; however, the phenotypes and downstream molecular events are fairly different, reflecting the dependence on cellular context and tissue specificity [39]. Our current data shows that AEG-1 can promote the accumulation of nuclear β-catenin. The activities of β-catenin luciferase reporter constructs were significantly decreased by AEG-1 siRNA in NCI-H226 cells. Furthermore, β-catenin could reverse AEG-1-siRNA’s effect on EMT. Consistent with the previous reports, we also found that both activation of β-catenin and promotion of EMT can result from up-regulation of AEG-1, further supporting the notion that the β-catenin-mediated pro-EMT function of AEG-1 up-regulation might also contribute to AEG-1–induced metastasis in lung cancer. These data indicate that AEG-1 is a key promotor of EMT through activating Wnt/β-catenin signaling.

Wnt/β-catenin signaling pathway has been widely implicated as the regulator of cell invasion and migration in cancers [40,41]. β-catenin is a main downstream effector of the canonical Wnt signaling pathway, which has a dual role in EMT: it not only enhances cell-cell adhesion by associating with cadherin complexes in adherent junctions of cell membrane, but also functions as a transcriptional co-activator by interacting with TCF transcription factor complexes in the nucleus [42]. Yoo et al. reported that AEG-1 can activate the canonical Wnt signaling pathway [9]. Other studies demonstrated that many growth factors, such as insulin growth factor, transforming growth factor-β, and epidermal growth factor, could increase β-catenin accumulation through Ser-9 phosphorylation of GSK-3β [43]. Our analysis also revealed that Ser-9 phosphorylation of GSK-3β was involved in the stability and transcriptional activity of AEG-1-mediated β-catenin. Furthermore, the physical interaction between AEG-1 and GSK-3β facilitates GSK-3β inactivation through Ser-9 dephosphorylation, which increases nuclear β-catenin accumulation and transcriptional activity. These results revealed the potent promoting function of AEG-1 on Wnt/β-catenin signaling. However, since AEG-1 is not a phosphatase, the molecular mechanism of GSK-3β inactivation may be mediated by a separate phosphatase associated with this complex. Recent studies have also implicated CKI as a positive regulator of β-catenin signaling, which phosphorylates several components of the β-catenin degradation complex in vitro such as GSK-3β [44]. In our study, coimmunoprecipitation data indicated that AEG-1 could form a complex with GSK-3δ and CKIδ. Thus, identification of small molecules that could perturb the interaction between AEG-1 and its partners, resulting the inhibition of AEG-1 function, might be a rational and effective way of target therapy of NSCLC.

In recent years, the prognostic value of AEG-1 has been widely confirmed in various cancers [45,46], and its tumor-promoting role has also been manifested. Our data provided evidence that high expression of AEG-1 was closely correlated with poor prognosis and lower patient survival rate. We concluded that the combined detection of the AEG-1 level and EMT status showed more significant prognostic value, suggesting that they may be regarded as correlative predictive factors for death in lung cancer patients. From the results of Kaplan-Meier analysis, we can conclude that AEG-1 is a reliable prognostic factor of the overall survival; moreover, AEG-1 combining with EMT status is able to more accurately predict the probability of death in lung cancer patients.

**Conclusions**

In summary, this study first delineates the functional role of AEG-1 in EMT and metastasis of NSCLC, and demonstrates how AEG-1 underlies the onset of EMT and aggressive metastasis of lung cancer by activating Wnt/β-catenin signaling. These findings also uncover a novel molecular mechanism that maintains the constitutive activation of the Wnt/β-catenin signaling by AEG-1, and AEG-1 may prove to be clinically useful for developing a new prognostic biomarker and therapeutic target for lung cancer.

**Competing interest**

The authors declare that they have no competing interest.

**Authors’ contributions**

Conception and design: ZK, LW and WH. Development of methodology: WH and SH. Acquisition of data: ZW and HS. Analysis and interpretation of data: ZW and WF. Writing, review and/or revision of the manuscript: ZK, YZ and JY.
Administrative, technical, or material support: WQ, ML, and JW. Study supervision: ZK. All authors read and approved the final manuscript.

Acknowledgements

This work was financially supported by the National Natural Science Foundation of China (No. 30006505/H1615, 81372501/H1615, 81172322/H1615 and 81172564/H1625), the Guangdong Natural Science Foundation (No. S2012100008378, S2013100115327, 2013B011801026), and the Introduced Major Research and Development Project Funded by Fujian Province.

Author details

1 Department of Gastrointestinal Surgery, Guangzhou 510080, Province Guangdong, Peoples' Republic of China. 2 Gynecology, and the First Affiliated Hospital of Sun Yat-sen University, Guangzhou 510080, Province Guangdong, Peoples' Republic of China. 3 Department of Pathology, The First Affiliated Hospital, Sun Yat-Sen University, 58 Zhongshan Road II, Guangzhou, Guangdong 510080, Peoples' Republic of China. 4 Department of Oncology, Sun Yat-sen University Cancer Center, Guangzhou 510060, Province Guangdong, Peoples' Republic of China. 5 College of Engineering, University of Texas, El Paso 500 West University Avenue, El Paso, TX 79968, USA. 6 Department of Molecular and Medical Pharmacology, University of California, Los Angeles, 570 Westwood Plaza, Los Angeles, CA 90095-1770, USA. 7 Department of Anatomy, School of Basic Medical Science, Southern Medical University, Guangzhou, Guangdong 510515, Peoples' Republic of China.

Received: 3 September 2014 Accepted: 24 February 2015

Published online: 08 March 2015

References

1. Siegel R, Naishadham D, Jemal A. Cancer statistics, 2012. CA Cancer J Clin. 2012;62:10–29.

2. Cho J. The international association for the study of lung cancer-the lung cancer staging project: better data, better decisions, better outcomes. Hawaii Med J. 2006;67:220–2.

3. Miradraee S, Owal A, Alizadeh Y, Caoulo A, van Jr. Beek E. The 7th lung cancer TNM classification and staging system: review of the changes and implications. World J Radiol. 2012;4:128–34.

4. Couzin-Frankel J. Clinical trials: experimental cancer therapies move to the front line. Science. 2012;335:282–3.

5. Christophor G. New signals from the invasive front. Nature. 2006;441:444–7.

6. Bonnomet A, Syne L, Brysse A, Feyereisen E, Thompson EW, Noel A, et al. A novel way of activating the nuclear factor kappaB pathway by astrocyte elevated gene-1. Cancer Res. 2011;71:7539–43.

7. Lee TK, Poon RT, Yuen AP, Ling MT, Kowk WK, Wang XH, et al. Twist overexpression correlates with hepatocellular carcinoma metastasis through induction of epithelial-mesenchymal transition. Cancer Res. 2006;66:323.

8. Yang S, Kang Y, Yang L, Song L, Li L, Liu Z, et al. The role of astrocyte elevated gene-1 in human nonsmall cell lung cancer A549 cells. Anticancer Drugs. 2013;24:494–503.

9. Ke Z, Zhang X, Ma L, Wang L. Expression of DPC4/Smad4 in non-small cell lung carcinoma and its relationship with angiogenesis. Neoplasma. 2008;55:323–9.

10. Logan CY, Nusse R. The Wnt signaling pathway in development and disease. Annu Rev Cell Dev Bio. 2004;20:781–810.

11. Chaffer CL, Weinberg RA. A perspective on cancer cell metastasis. Science. 2011;331:594–505.

12. Emdad L, Shima H, Uraoki S, Kawamoto X, Hirata H, Tanaka Y, et al. Knockdown of astrocyte-elevated gene-1 inhibits prostate cancer progression through upregulation of FOXO3a activity. Oncogene. 2007;26:7647–55.

13. U. J. Zhang N, Song L, Yao LT, Jiang LL, Gong L, et al. Astrocyte elevated gene-1 is a novel prognostic marker for breast cancer progression and overall patient survival. Clin Cancer Res. 2008;14:3315–9.

14. Li J, Yang L, Song L, Xiong H, Wang L, Yan X, et al. Astrocyte elevated gene-1 is a proliferation promoter in breast cancer via suppressing transcriptional factor FOXO1. Oncogene. 2009;28:3188–96.

15. Hui AB, Bruce JP, Alagez NM, Shi W, Yue S, Perez-Ordonez B, et al. Significance of dysregulated mediator and microRNA-375 in head and neck cancer. Clin Cancer Res. 2011;17:7539–50.

16. Brown DM, Rusolati E. Metadherin, a cell surface protein in breast tumors that mediate lung metastasis. Cancer Cell. 2004;5:365–74.

17. Weinberg RA. Mechanisms of malignant progression. Carcinogenesis. 2008;29:1092–5.

18. Boyer B, Vaillet A, Edme N. Induction and regulation of epithelial-mesenchymal transitions. Biochem Pharmacol. 2003;65:1091–9.

19. Weber CE, Li NY, Wal PK, Xuo PC. Epithelial-mesenchymal transition, TGF-β, and osteopontin in wound healing and tissue remodeling after injury. J Burn Care Res. 2012;33:311–8.

20. Hugo H, Achain MLD, BLT, Tawrence MG, Clements JA, Williams ED, et al. Epithelial–mesenchymal and mesenchymal–epithelial transitions in carcinoma progression. J Cell Physiol. 2007;213:374–83.

21. Lee TK, Poon RT, Yuen AP, Ling MT, Kowk WK, Wang XH, et al. Twist overexpression correlates with hepatocellular carcinoma metastasis through induction of epithelial-mesenchymal transition. Cancer Res. 2006;66:3239–66.

22. Gravdal K, Halvorsen OJ, Haukaas SA, Asken LA. A switch from E-cadherin to N-cadherin expression indicates epithelial to mesenchymal transition and is of strong and independent importance for the progress of prostate cancer. Clin Cancer Res. 2007;13:7902–11.

23. Lee, IJ, Zhang X, Ma L, Cai Z, Cimk F, Wu W, et al. AEG-1 participates in TGF-beta1-induced EMT through p38 MAPK activation. Cell Biol Int. 2013;37:1016–21.

24. Tan L, Kang Y. Pleiotropic roles of AEG-1/MTDH/LYRIC in breast cancer. Adv Cancer Res. 2013;120:113–34.

25. Valcourt U, Kovacevic P, Nihm I, Heldin CH, Mouratas A. The Smad signaling pathway supports transcriptional reprogramming during epithelial-mesenchymal cell transition. Mol Biol Cell. 2005;16:1807–2002.

26. Shin SY, Roth O, Zebisch A, Choo SM, Kolch W, Cho KH. Functional roles of multiple feedback loops in extracellular signal-regulated kinase and Wnt signaling pathways that regulate epithelial-mesenchymal transition. Cancer Res. 2010;70:6715–24.

27. Karhadkar SS, Boia GS, Abdallah N, Dharma S, Gardner D, Maitra A, et al. Hedgehog signalling in prostate regeneration, neoplasia and metastasis. Nature. 2004;431:707–12.

28. Timmerman LA, Grego-Bessa J, Raya A, Bertran E, Perez-Pomares JM, Diez J, et al. TGF-beta and the epithelial-mesenchymal transition. J Pathol. 2000;191:1987–93.

29. Shin SY, Roth O, Zebisch A, Choo SM, Kolch W, Cho KH. Functional roles of multiple feedback loops in extracellular signal-regulated kinase and Wnt signaling pathways that regulate epithelial-mesenchymal transition. Cancer Res. 2010;70:6715–24.
40. Chien AJ, Conrad WH, Moon RT. A Wnt survival guide: from flies to human disease. J Invest Dermatol. 2009;129:1614–27.
41. Giles RH, van Es JH, Clevers H. Caught up in a Wnt storm: Wnt signaling in cancer. Biochim Biophys Acta. 2003;1653:1–24.
42. Polette M, Mestdagh M, Bindels S, Nawrocki-Raby B, Hunziker W, Foidart JM, et al. Beta-catenin and ZO-1: shuttle molecules involved in tumor invasion-associated epithelial-mesenchymal transition processes. Cells Tissues Organs. 2007;185:61–5.
43. Kitazawa M, Cheng D, Tsukamoto MR, Koke MA, Wes PD, Vaselevko V, et al. Blocking IL-1 signaling rescues cognition, attenuates tau pathology, and restores neuronal -catenin pathway function in an Alzheimer’s disease model. J Immunol. 2011;187:6539–49.
44. Gao ZH, Seeling JM, Hill A, Yochum A, Virshup DM. Casein kinase I phosphorylates and destabilizes the beta-catenin degradation complex. Proc Natl Acad Sci U S A. 2002;99:1182–7.
45. Ke ZF, He S, Li S, Luo D, Feng C, Zhou W. Expression characteristics of astrocyte elevated gene-1 (AEG-1) in tongue carcinoma and its correlation with poor prognosis. Cancer Epidemiol. 2013;37:179–85.
46. Song L, Li W, Zhang H, Liao W, Dai T, Yu C, et al. Over-expression of AEG-1 significantly associates with tumour aggressiveness and poor prognosis in human non-small cell lung cancer. J Pathol. 2009;219:317–26.