Research Article

Modified Carbon Nanotube Paste Electrode for Voltammetric Determination of Carbidopa, Folic Acid, and Tryptophan

Sakineh Esfandiari Baghbamidi,1 Hadi Beitollahi,2 Hassan Karimi-Maleh,3 Somayeh Soltani-Nejad,1 Vahhab Soltani-Nejad,4 and Sara Roodsaz 4

1 Islamic Azad University, Bandar Abbas Branch, Bandar Abbas 7915893144, Iran
2 Environment Department, Research Institute of Environmental Sciences, International Center for Science, High Technology & Environmental Sciences, Kerman 763133131, Iran
3 Department of Chemistry, Science and Research Branch, Islamic Azad University, Mazandaran 4816819195, Iran
4 Department of Chemistry, Islamic Azad University, North Tehran Branch, Tehran 1913674711, Iran

Correspondence should be addressed to Hadi Beitollahi, h.beitollahi@yahoo.com

Received 28 November 2011; Accepted 23 January 2012

1. Introduction

Electrochemical detection of analyte is a very elegant method in analytical chemistry [1]. The interest in developing electrochemical-sensing devices for use in environmental monitoring, clinical assays, or process control is growing rapidly. Electrochemical sensors satisfy many of the requirements for such tasks particularly owing to their inherent specificity, rapid response, sensitivity, and simplicity of preparation for the determination of organic molecules, including drugs and related molecules in pharmaceutical dosage forms and biological fluids [2, 3]. Carbon electrodes, especially glassy and paste electrodes, are widely used in electrochemical investigations [4–8].

Electrochemical sensors based on carbon nanotubes (CNTs) represent a new and interesting alternative for quantification of different analytes. There are reports on the synthesis of multiwalled carbon nanotubes (MWCNTs) [9] and single-walled carbon nanotubes (SWCNTs) [10]. These materials have attracted enormous interest because of their unique structural, mechanical, electronic, and chemical properties. Some of these properties include high chemical and thermal stability, high elasticity, high tensile strength and, in some instances, metallic conductivity. The subtle electronic properties suggest that CNTs have the capability of promoting electron transfer reactions and improving sensitivity in electrochemistry, and thus they are widely used as electrodes [11–13]. CNT modified electrodes have been proved to have excellent electroanalytical properties, such as wide potential window, low background current, low detection limits, high sensitivities, reduction of over potentials, and resistance to surface fouling. There are reports that reveal that CNT modified electrodes have shown electrocatalytic behavior with excellent performance in the study of a number of biological species [14–18].

Drug analysis is one of the important tools for drug quality control. Therefore, the development of simple, sensitive, rapid, and reliable method for the determination of drug is of great importance [19–21].
Parkinson’s disease victims show a significant depletion of dopamine in the brain. Since this neurotransmitter can not cross the blood-brain barrier into the central nervous system and it can not be employed to restore its normal level, levodopa (LD) (a precursor of dopamine) has been successfully used and is the most widely prescribed drug for the treatment of such patients [22, 23]. After its administration, LD is converted into dopamine via an enzymatic reaction catalyzed by dopa-decarboxylase. However, since the metabolism of LD is also extracerebral, several side effects of systemic dopamine can arise if LD is administered in high dosages. In order to achieve a better therapeutic effect and lower toxicity, carbodiopa (CD) is administered in association with LD in pharmaceutical preparations, which contain 10–25% CD [24]. This catecholamine acts as an inhibitor for the decarboxylase activity. Hence, a combination of LD with CD leads to a control of the dopamine concentration at suitable levels, reducing the side effects and improving the efficiency of the therapy. Accordingly, the development of an analytical method is very important to control the content of these catecholamines in pharmaceuticals. Different techniques such as spectrophotometry, H-point standard addition, flow-batch, synchronous fluorescence spectrometry, and high-performance liquid chromatography have been employed for the determination of CD in pharmaceutical formulations [25–29]. Long analysis times, the use of organic solvents, and high costs are some of the drawbacks associated with these techniques. Voltammetry is considered as an important electrochemical technique utilized in electroanalytical chemistry because it provides low cost, sensitivity, precision, accuracy, simplicity, and rapidity [30, 31].

Folic acid (FA) is a water-soluble vitamin and can act as coenzyme in the transfer and utilization of one-carbon groups and in the regeneration of methionine from homocysteine [32]. Deficiency of FA is a common cause of anaemia, and it is thought to increase the likelihood of heart attack and stroke. Many studies suggest that diminished folate status is associated with enhanced carcinogenesis as FA with vitamin B12 participates in the nucleotide synthesis, cell division, and gene expression [33]. Periconceptual supplementation of FA has been demonstrated to reduce significantly the incidence and reoccurrence of neural tube defects, such as spina bifida of women [34]. In January 1998, the US Food and Drug Administration introduced mandatory fortification of cereal grain products with FA at a concentration of 140 mg/100 g [35]. In the UK, the Department of Health proposed fortification of flour with FA at 240 mg/100 g [36]. Numerous methods for the measurement of FA are available. As FA is an electroactive component, some electrochemical methods have been reported for its determination. Comparing with other technologies, electrochemical method is more desirable because of its convenience and low cost [37–41].

Tryptophan (2-amino-3-(1H-indol-3-yl)-propionicpropionic acid, TRP) is an essential amino acid for humans and a precursor for serotonin (a neurotransmitter), melatonin (a neurohormone), and niacin. It has been implicated as a possible cause of schizophrenia in people who cannot metabolize it properly. This compound is sometimes added to dietary, food products, and pharmaceutical formulas due to its scarce presence in vegetables [42]. Therefore, simple, sensitive and less expensive detection of TRP is of great interest. Therefore, various methods have been reported for the determination of TRP. Concentration of amino acids in biological samples is low; therefore it is necessary to use a highly sensitive method that provides determination of these analytes at subordinate concentrations. The electrochemical analytical technique is an attractive method due to its simplicity, low expense, high sensitivity, and possibility of miniaturization [43–46].

In the present work, we describe the preparation of a new electrode composed of CNPE modified with ferrocene dicarboxylic acid (FCDCNPE) and investigate its performance for the electrocatalytic determination of CD in aqueous solutions. We also evaluate the analytical performance of the modified electrode for quantification of CD in the presence of FA and TRP.

2. Experimental

2.1. Apparatus and Chemicals. The electrochemical measurements were performed with an Autolab potentiostat/galvanostat (PGSTAT 302 N, Eco Chemie, the Netherlands). The experimental conditions were controlled with General Purpose Electrochemical System (GPES) software. A conventional three-electrode cell was used at 25 ± 1°C. An Ag/AgCl/KCl (3.0 M) electrode, a platinum wire, and the FCDCNPE were used as the reference, auxiliary, and working electrodes, respectively. A Metrohm 691 pH/ion meter was used for pH measurements.

All solutions were freshly prepared with double distilled water. CD, FA, TRP, and all other reagents were of analytical grade from Merck (Darmstadt, Germany). Graphite powder and paraffin oil (DC 350, density = 0.88 g cm⁻³) as the binding agent (both from Merck) were used for preparing the pastes. Multiwalled carbon nanotubes (purity more than 95%) with o.d. between 10 and 20 nm, i.d. between 5 and 10 nm, and tube length from 0.5 to 200 μm were provided from Nanostructured & Amorphous Materials, Inc. The buffer solutions were prepared from orthophosphoric acid and its salts in the pH range of 2.0–11.0.

2.2. Preparation of the Electrodes. The FCDCNPEs were prepared by hand mixing 0.01 g of FCD with 0.89 g graphite powder and 0.1 g CNTs with a mortar and pestle. Then, ~0.7 mL of paraffin oil was added to the above mixture and mixed for 20 min until a uniformly wetted paste was obtained. The paste was then packed into the end of a glass tube (ca. 3.4 mm i.d. and 15 cm long). A copper wire inserted into the carbon paste provided the electrical contact. When necessary, a new surface was obtained by pushing an excess of the paste out of the tube and polishing with a weighing paper.

For comparison, FCD modified CPE electrode (FCDCP) without CNTs, CNTs paste electrode (CNPE) without FCD, and unmodified CPE in the absence of both FCD and CNTs were also prepared in the same way.
2.3. Procedure of Urine Samples Preparation. Urine samples were stored in a refrigerator immediately after collection. Ten milliliters of the sample was centrifuged for 15 min at 2000 rpm. The supernatant was filtered out using a 0.45 μm filter. Then, a different volume of the solution was transferred into a 10 mL volumetric flask and diluted to the mark with phosphate buffer (pH 5.0). The diluted urine sample was spiked with different amounts of CD, FA, and TRP.

3. Results and Discussion

3.1. Electrochemical Behavior of FCDCNPE. We have previously shown that a carbon paste electrode spiked with FCD is constructed by the incorporation of FCD in a graphite powder-paraffin oil matrix [47]. The experimental results show that well-defined and reproducible anodic and cathodic peaks were related to ferrocene dicarboxylic acid/ferricenium dicarboxylic acid (Fc/Fc+) redox system, which show a quasireversible behavior in an aqueous medium [48]. The electrode capability for the generation of a reproducible surface was examined by cyclic voltammetric data obtained in optimum solution, pH 5.0, from five separately prepared FCDCNPEs (Table 1). The calculated RSD for various parameters was accepted as the criterion for a satisfactory surface reproducibility (about 1–4%), which is virtually the same as that expected for the renewal or ordinary carbon paste surface [7, 12]. However, we regenerated the surface of FCDCNPE before each experiment according to our previous result [47].

In addition, the long-term stability of the FCDCNPE was tested over a three-week period. When CVs were recorded after the modified electrode was stored in atmosphere at room temperature, the peak potential for CD oxidation was unchanged and the current signals showed less than 2.5% decrease relative to the initial response. The antifouling properties of the modified electrode toward CD oxidation and its oxidation products were investigated by recording the cyclic voltammograms of the modified electrode before and after use in the presence of CD. Cyclic voltammograms were recorded in the presence of CD after they have cycled the potential 20 times at a scan rate of 20 mV s⁻¹. The peak potentials were unchanged and the currents decreased by less than 2.1%. Therefore, at the surface of FCDCNPE, not only does the sensitivity increase, but the fouling effect of the analyte and its oxidation product also decreases.

3.2. Influence of pH. The electrochemical behavior of CD is dependent on the pH value of the aqueous solution, whereas the electrochemical properties of Fc/Fc⁺ redox couple are independent of pH. Therefore, pH optimization of the solution seems to be necessary in order to obtain the electrocatalytic oxidation of CD. Thus, the electrochemical behavior of CD was studied in 0.1 M phosphate-buffered solutions (PBSs) in different pH values (2.0 < pH < 11.0) at the surface of FCDCNPE by cyclic voltammetry. It was found that the electrocatalytic oxidation of CD at the surface of FCDCNPE was more favored under acidic conditions than in neutral or basic medium. This appears as a gradual growth in the anodic peak current and a simultaneous decrease in the cathodic peak current in the cyclic voltammograms drawn at the surface of FCDCNPE. The variation of Ipa versus the variation of pH was studied. Results showed that the anodic peak current and the shifted potential value for electrooxidation of CD are high at pH 5.0. Thus, the pH 5.0 was chosen as the optimum pH for electrocatalysis of CD oxidation at the surface of FCDCNPE.

Table 1: Cyclic voltammetric data obtained for constructed FCDCNPE in 0.1 M PBS (pH 5.0) at 10 mV s⁻¹.

| pH  | Epa (V)  | Ep (V)  | E1/2 (V) | ΔEp (V) | Ipa (μA) | Ip (μA) |
|-----|----------|---------|----------|---------|----------|---------|
| 4.5 | 0.4 ± 1.2 | 0.4 ± 1.3 | 0.4 ± 1.1 | 0.1 ± 1.2 | 1.0 ± 1.6 | 0.5 ± 1.8 |

a Versus Ag/AgCl/KCl (3.0 M) as reference electrode.

All the “±” values are RSD% (n = 5).

3.3. Electrocatalytic Oxidation of CD at an FCDCNPE. Figure 1 depicts the CV responses for the electrochemical oxidation of 0.3 mM CD at unmodified CPE (curve b), CNPE (curve d), FCDCPE (curve e), and FCDCNPE (curve f). As it is seen, while the anodic peak potential for CD oxidation at the CNPE and unmodified CPE is 830 and 890 mV, respectively, the corresponding potential at FCDCNPE and FCDCPE is ~500 mV. These results indicate that the peak potential for CD oxidation at the FCDCNPE and FCDCPE electrodes shifts by ~330 and 390 mV toward negative values compared to CNPE and unmodified CPE, respectively. However, FCDCNPE shows much higher anodic peak current for the oxidation of CD compared to FCDCPE, indicating that the combination of CNTs and the mediator (FCD) has significantly improved the performance of the electrode toward CD oxidation. In fact, FCDCNPE in the absence of CD exhibited a well-behaved redox reaction (Figure 1, curve c) in 0.1 M PBS (pH 5.0). However, there was a drastic increase in the anodic peak current in the presence of 0.3 mM CD (curve f), which can be related to the strong electrocatalytic effect of the FCDCNPE towards this compound [48].

The effect of scan rate on the electrocatalytic oxidation of CD at the FCDCNPE was investigated by linear sweep voltammetry (LSV) (Figure 2). Also, a plot of peak height (Ipa) versus the square root of scan rate (ν^1/2) was found to be linear in the range of 2–20 mV s⁻¹, suggesting that, at sufficient overpotential, the process is diffusion rather than surface controlled (Figure 2 (a)). A plot of the scan rate-normalized current (Ipa/ν^1/2) versus scan rate (Figure 2 (b)) exhibits the characteristic shape typical of an EC process [48].

Figure 3 shows a Tafel plot that was drawn from points of the Tafel region of the linear sweep voltammogram. The Tafel slope of 83.1 mV obtained in this case agrees well with the involvement of one electron in the rate determining step of the electrode process, assuming a charge transfer coefficient of α = 0.29.

3.4. Chronoamperometric Measurements. Chronoamperometric measurements of CD at FCDCNPE were carried out by setting the working electrode potential at 0.55 V versus Ag/AgCl/KCl (3.0 M) for the various concentrations of CD.
Figure 1: CVs of (a) unmodified CPE in 0.1 M PBS (pH 5.0), (b) unmodified CPE in 0.3 mM CD, (c) FCDCNPE in 0.1 M PBS, (d) CNPE in 0.3 mM CD, (e) FCDCPE in 0.3 mM CD, and (f) FCDCNPE in 0.3 mM CD. In all cases the scan rate was 20 mV s$^{-1}$.

Figure 2: Linear sweep voltammograms of FCDCNPE in 0.1 M PBS (pH 5.0) containing 0.35 $\mu$M CD at various scan rates; numbers 1–7 correspond to 2, 4, 6, 8, 10, 15, and 20 mV s$^{-1}$, respectively. Insets: variation of (a) anodic peak current versus $v^{1/2}$; (b) normalized current ($I_p/v^{1/2}$) versus $v$.

in PBS (pH 5.0) (Figure 4). For an electroactive material (CD in this case) with a diffusion coefficient of $D$, the current observed for the electrochemical reaction at the mass transport limited condition is described by the Cottrell equation [48]. Experimental plots of $I$ versus $t^{-1/2}$ were employed, with the best fits for different concentrations of CD (Figure 4(a)). The slopes of the resulting straight lines were then plotted versus CD concentration (Figure 4(b)). From the resulting slope and the Cottrell equation the mean value of the $D$ was found to be $5.4 \times 10^{-6}$ cm$^2$/s.

Also, double potential step chronocoulometry, as well as other electrochemical methods, was employed for the investigation of electrode processes at FCDCNPE (Figure 4(c)). As can be seen, the forward and backward potential
3.5. Electrocatalytic Determination of CD. SWV method (with initial potential = 0.1 V, end potential = 0.6 V, step potential = 0.0195 V, amplitude = 0.045 V, and frequency = 10 Hz) was used to determine the concentration of CD. The plot of peak current versus CD concentration consisted of two linear segments with slopes of 0.0643 and 0.0138 μA μM⁻¹ in the concentration ranges of 0.1 to 80.0 μM and 80.0 to 750.0 μM, respectively. The decrease in sensitivity (slope) of the second linear segment is likely due to kinetic limitation. The detection limit (3σ) of CD was found to be 65.0 nM. These values are comparable with values reported by other research groups for electrocatalytic oxidation of CD at the surface of chemically modified electrodes by other mediators (see Table 2).

3.6. Simultaneous Determination of CD, FA, and TRP. To our knowledge, there is no report on the simultaneous determination of CD, FA, and TRP using FCDCNPE. Therefore, the main objective of this study was to detect CD, FA, and TRP simultaneously using FCDCNPE. This was performed by simultaneously changing the concentrations of CD, FA, and TRP and recording the SWVs (with initial potential = 0.1 V, end potential = 1.2 V, step potential = 0.0195 V, amplitude = 0.045 V, and frequency = 10 Hz). The voltammetric results showed well-defined anodic peaks at potentials of 470, 740, and 1030 mV, corresponding to the oxidation of CD, FA, and TRP, respectively, indicating that simultaneous determination of these compounds is feasible at the FCDCNPE as shown in Figure 5.

3.7. Real Sample Analysis. To evaluate the applicability of the proposed method to real samples, it was applied to the determination of CD, FA, and TRP in urine samples. The CD, FA, and TRP contents were measured after sample preparation using the standard addition method. The results are given in Table 3.

4. Conclusions

In this study, a modified carbon nanotube paste electrode was fabricated for the voltammetric determination of CD. The enhancement in the oxidation current of CD and the negative shift of peak potential of CD at the modified electrode was observed. This newly developed method is sensitive, convenient, rapid, and suitable for determining CD. The modified electrode greatly catalyzed the electrooxidation reactions of CD, FA, and TRP, improving their oxidation peak separation. Thus, the large peak separations between CD, FA, and TRP allow their simultaneous analysis through...
square wave voltammetry technique. The proposed method could be applied to the determination of CD, FA and TRP in urine sample with quite promising results.

References

[1] H. Beitollahi, H. Karimi-Maleh, and H. Khabazzadeh, “Nanomolar and selective determination of epinephrine in the presence of norepinephrine using carbon paste electrode modified with carbon nanotubes and novel 2-(4-oxo-3-phenyl-3,4-dihydroquinazinyl)-N'-phenyl-hydrazinecarbothioamide,” Analytical Chemistry, vol. 80, no. 24, pp. 9848–9851, 2008.

[2] T. H. Tsai, S. H. Wang, and S. M. Chen, “Electrodeposited indigotetrasulfonate film onto glutaraldehyde-cross-linked poly-1-lysine modified glassy carbon electrode for detection of dissolved oxygen,” Journal of Electroanalytical Chemistry, vol. 659, no. 1, pp. 69–75, 2011.

[3] R. Sivasubramanian and M. V. Sangaranarayanan, “Detection of lead ions in picomolar concentration range using underpotential deposition on silver nanoparticles-deposited glassy carbon electrodes,” Talanta, vol. 85, no. 4, pp. 2142–2147, 2011.

[4] J. B. Raoof, R. Ojani, H. Beitollahi, and R. Hosseinzadeh, “Electrocatalytic determination of ascorbic acid at the surface of 2,7-bis(ferrocenyl ethyl)fluoren-9-one modified carbon paste electrode,” Electroanalysis, vol. 18, no. 12, pp. 1193–1201, 2006.

[5] S. Sabahat, N. K. Janjua, M. Brust, and Z. Akhter, “Electrochemical fabrication of self assembled monolayer using ferrocene-functionalized gold nanoparticles on glassy carbon electrode,” Electrochimica Acta, vol. 56, no. 20, pp. 7092–7096, 2011.

[6] Q. Thuan Tran, J. F. Bergamini, C. Mangeney, C. Lagrost, and P. Pellon, “Grafting of borane-protected aliphatic and aromatic aminophosphine ligands to glassy carbon electrodes,” Electrochemistry Communications, vol. 13, no. 8, pp. 844–847, 2011.

[7] H. Beitollahi, J. B. Raoof, and R. Hosseinzadeh, “Fabrication of a nanostructure-based electrochemical sensor for simultaneous determination of N-acetylcysteine and acetaminophen,” Talanta, vol. 85, pp. 2128–2134, 2011.

[8] X. Liu, L. Luo, Y. Ding, and D. Ye, “Poly-glutamic acid modified carbon nanotube-doped carbon paste electrode for sensitive detection of L-tryptophan,” Bioelectrochemistry, vol. 82, pp. 38–45, 2011.

[9] S. Iijima, “Helical microtubules of graphitic carbon,” Nature, vol. 354, no. 6348, pp. 56–58, 1991.

[10] S. Iijima and T. Ichihashi, “Single-shell carbon nanotubes of 1-nm diameter,” Nature, vol. 363, no. 6439, pp. 603–605, 1993.

[11] G. Liu, H. Chen, H. Peng et al., “A carbon nanotube-based high-sensitivity electrochemical immunosensor for rapid and portable detection of clenbuterol,” Biosensors and Bioelectronics, vol. 28, no. 1, pp. 308–313, 2011.

[12] H. Beitollahi and I. Sheikhshoaei, “Selective voltammetric determination of norepinephrine in the presence of acetylmethionine and folic acid at a modified carbon nanotube paste electrodes,” Journal of Electroanalytical Chemistry, vol. 661, pp. 336–342, 2012.

[13] M. Noroozifar, M. Khorasani-Motlagh, and A. Taheri, “Determination of cyanide in wastewaters using modified glassy carbon electrode with immobilized silver hexacyanoferrate nanoparticles on multiwall carbon nanotube,” Journal of Hazardous Materials, vol. 185, no. 1, pp. 255–261, 2011.

[14] H. Beitollahi and I. Sheikhshoaie, “Electrocatalytic oxidation and determination of epinephrine in the presence of uric acid and folic acid at multiwalled carbon nanotubes/molybdenum(VI) complex modified carbon paste electrode,” Analytical Methods, vol. 3, pp. 1810–1814, 2011.

[15] R. Ahmad Dar, P. Kumar Brahmans, S. Tiwari, and K. Sadashiv Pitre, “Adsorptive stripping voltammetric determination of...
podophyllotoxin, an antitumour herbal drug, at multi-walled carbon nanotube paste electrode,” Journal of Applied Electrochemistry, vol. 41, pp. 1311–1321, 2011.

[16] H. Beitollahi and I. Sheikhshoaie, “Novel nanostructure-based electrochemical sensor for simultaneous determination of dopamine and acetaminophen,” Material Science and Engineering C, vol. 32, pp. 375–380, 2012.

[17] L. Zheng and J. F. Song, “Electrocatalytic oxidation of hydroxy-ylate at Ni(II)-morin complex modified carbon nanotube paste electrode,” Journal of Applied Electrochemistry, vol. 41, no. 1, pp. 63–70, 2011.

[18] J. Tashkhourian, S. Javadi, and F. Nami Ana, “Anodic stripping voltammetric determination of silver ion at a carbon paste electrode modified with carbon nanotubes,” Microchimica Acta, vol. 173, no. 1–2, pp. 79–84, 2011.

[19] H. Beitollahi and I. Sheikhshoaie, “Electrocatalytic and simultaneous determination of isoproterenol, uric acid and folic acid at molybdenum (VI) complex-carbon nanotube paste electrode,” Electrochimica Acta, vol. 56, pp. 10259–10263, 2011.

[20] A. A. Ensafi and H. Karimi-Maleh, ”Modified multilwall carbon nanotubes paste electrode as a sensor for simultaneous determination of 6-thioguanine and folic acid using ferrocenedicarboxylic acid as a mediator,” Journal of Electroanalytical Chemistry, vol. 640, no. 1–2, pp. 75–83, 2010.

[21] I. Santos da Silva, M. F. Antão Araújo, H. A. Ferreira et al., “Quantification of N-acetylcysteine in pharmaceuticals using cobalt phthalocyanine modified graphite electrodes,” Talanta, vol. 83, pp. 1701–1706, 2011.

[22] H. V. Barnes, Clinical Medicine, Year Book Medical, New York, NY, USA, 1988.

[23] P. Gomes and P. Soares-da-Silva, “Interaction between L-Dopa and 3-O-methyl-L-Dopa for transport in immortalised rat capillary cerebral endothelial cells,” Neuropharmacology, vol. 38, no. 9, pp. 1371–1380, 1999.

[24] J. E. F. Reynolds, Ed., Martindale, The Extra Pharmacopoeia, The Pharmaceutical Press, London, UK, 1993.

[25] M. Chamsaz, A. Safavi, and J. Fadaee, “Simultaneous kinetic-spectrophotometric determination of carbidopa, levodopa and methyldopa in the presence of citrate with the aid of multivariate calibration and artificial neural networks,” Analytica Chimica Acta, vol. 603, no. 2, pp. 140–146, 2007.

[26] A. Safavi and M. Tohidi, “Simultaneous kinetic determination of levodopa and carbidopa by H-point standard addition method,” Journal of Pharmaceutical and Biomedical Analysis, vol. 44, no. 1, pp. 313–318, 2007.

[27] M. Grünhut, M. E. Centurion, W. D. Fragoso, L. F. Almeida, M. C. U. de Araújo, and B. S. F. Band, “Flow-batch technique for the simultaneous enzymatic determination of levodopa and carbidopa in pharmaceuticals using pls and successive projections algorithm,” Talanta, vol. 75, no. 4, pp. 950–958, 2008.

[28] W. H. Kim, M. M. Karim, and S. H. Lee, “Simultaneous determination of levodopa and carbidopa by synchronous fluorescence spectrometry using double scans,” Analytica Chimica Acta, vol. 619, no. 1, pp. 2–7, 2008.

[29] M. Karimi, J. L. Carl, S. Loftin, and J. S. Perlmutter, “Modified high-performance liquid chromatography with electrochemical detection method for plasma measurement of levodopa, 3-O-methyldopa, dopamine, carbidopa and 3,4-dihydroxyphenyl acetic acid,” Journal of Chromatography B, vol. 836, no. 1–2, pp. 120–123, 2006.

[30] H. Yaghoubian, H. Karimi-Maleh, M. A. Khalilzadeh, and F. Karimi, “Electrochemical detection of carbidopa using a ferrocene-modified carbon nanotube paste electrode,” Journal of the Serbian Chemical Society, vol. 74, no. 12, pp. 1443–1453, 2009.

[31] R. E. Sabzi, A. Hasanazadeh, K. Ghasemlu, and P. Heravi, “Preparation and characterization of carbon paste electrode modified with tin and hexacyanoferrate ions,” Journal of the Serbian Chemical Society, vol. 72, no. 10, pp. 993–1002, 2007.

[32] C. M. Pfeiffer, Z. Fazili, L. McCoy, M. Zhang, and E. W. Gunter, “Determination of folate vitamers in human serum by stable-isotope-dilution tandem mass spectrometry and comparison with radioassay and microbiologic assay,” Clinical Chemistry, vol. 50, no. 2, pp. 423–432, 2004.

[33] D. Hoegger, P. Morier, C. Vollet, D. Heini, E. Reynold, and J. S. Rossier, “Disposable microfluidic ELISA for the rapid determination of folic acid content in food products,” Analytical and Bioanalytical Chemistry, vol. 387, no. 1, pp. 267–275, 2007.

[34] E. Gujska and A. Kunczewicz, “Determination of folate in some cereals and commercial cereal-grain products consumed in Poland using trienzyme extraction and high-performance liquid chromatography methods,” European Food Research and Technology, vol. 221, no. 1–2, pp. 208–213, 2005.

[35] Food and Drug Administration, Food Standards: Amendment of Standards of Identity for Enriched Grain Products to Require Addition of Folic Acid, vol. 61, Food and Drug Administration, Rockville, Md, USA, 1996.

[36] A. J. A. Wright, P. M. Finglas, and S. Southon, “Proposed mandatory fortification of the UK diet with folic acid: have potential risks been underestimated?” Trends in Food Science and Technology, vol. 12, no. 9, pp. 313–321, 2001.

[37] H. Beitollahi, J. B. Raoof, H. Karimi-Maleh, and R. Hossein-zadeh, “Electrochemical behavior of isoproterenol in the presence of uric acid and folic acid at a carbon-paste electrode modified with 2,7-bis(ferrocenyl ethyl)/fluoren-9-one and carbon nanotubes,” Journal of Solid State Electrochemistry, vol. 16, no. 4, pp. 1701–1707, 2012.

[38] B. B. Prasad, R. Madhuri, M. Prasad Tiwari, and P. S. Sharma, “Electrochemical sensor for folic acid based on a hyperbranched molecularly imprinted polymer-immobilized solgel-modified pencil graphite electrode,” Sensors and Actuators, B, vol. 146, no. 1, pp. 321–330, 2010.

[39] H. Beitollahi, J. B. Raoof, and R. Hossein-zadeh, “Application of a carbon-paste electrode modified with 2,7-(ferrocenyl ethyl)/fluoren-9-one and carbon nanotubes for voltammetric determination of levodopa in the presence of uric acid and folic acid,” Electroanalysis, vol. 23, no. 8, pp. 1934–1940, 2011.

[40] L. Bandzuchová, R. Šelešovská, T. Navrátil, and J. Chylková, “Electrochemical behavior of folic acid on mercury meniscus modified silver solid amalgam electrode,” Electrochimica Acta, vol. 56, no. 5, pp. 2411–2419, 2011.

[41] H. Beitollahi, J. B. Raoof, and R. Hossein-zadeh, “Electroanalytical and simultaneous determination of 6-thioguanine in the presence of uric acid and folic acid using a modified carbon paste electrode,” Analytical Sciences, vol. 27, pp. 991–997, 2011.

[42] Z. D. Chen, J. X. Wei, W. C. Wang, and Y. Kong, “Separation of tryptophan enantiomers with molecularly imprinted polymeric electrode column,” Chinese Chemical Letters, vol. 21, no. 3, pp. 353–356, 2010.

[43] J. Xu, Y. Yuan, W. Li, P. Deng, and J. Deng, “Carbon paste electrode modified with a binuclear manganese complex as a sensitive voltammetric sensor for tryptophan,” Microchimica Acta, vol. 174, pp. 239–245, 2011.
[44] X. Liu, L. Luo, Y. Ding, and D. Ye, “Poly-glutamic acid modified carbon nanotube-doped carbon paste electrode for sensitive detection of L-tryptophan,” Bioelectrochemistry, vol. 82, no. 1, pp. 38–45, 2011.

[45] P. Prabhua, R. S. Babua, and S. S. Narayanan, “Electrocatalytic oxidation of L-tryptophan using copper hexacyanoferrate film modified gold nanoparticle graphite-wax electrode,” Colloids and Surfaces B, vol. 87, pp. 103–108, 2011.

[46] S. Shahrokhian and M. Bayat, “Pyrolytic graphite electrode modified with a thin film of a graphite/diamond nano-mixture for highly sensitive voltammetric determination of tryptophan and 5-hydroxytryptophan,” Microchimica Acta, vol. 174, pp. 361–366, 2011.

[47] J. B. Raoof, R. Ojani, and H. Beitollahi, “L-cysteine voltammetry at a carbon paste electrode bulk-modified with ferrocenecarboxylic acid,” Electroanalysis, vol. 19, no. 17, pp. 1822–1830, 2007.

[48] A. J. Bard and L. R. Faulkner, in Electrochemical Methods Fundamentals and Application, John Wiley & Sons, New York, NY, USA, 2nd edition, 2001.

[49] M. Mazloum-Ardakani, B. Ganjipour, H. Beitollahi et al., “Simultaneous determination of levodopa, carbidopa and tryptophan using nanostructured electrochemical sensor based on novel hydroquinone and carbon nanotubes: application to the analysis of some real samples,” Electrochimica Acta, vol. 56, pp. 9113–9120, 2011.

[50] H. M. Moghaddam, “Electrocatalytic determination of carbidopa and acetaminophen using a modified carbon nanotube paste electrode,” International Journal of Electrochemical Sciences, vol. 6, pp. 6557–6566, 2011.