Noncompetitive Inhibition of 5-HT3 Receptors by Citral, Linalool, and Eucalyptol Revealed by Nonlinear Mixed-Effects Modeling

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ABSTRACT

Citral, eucalyptol, and linalool are widely used as flavorings, fragrances, and cosmetics. Here, we examined their effects on electrophysiological and binding properties of human 5-HT3 receptors expressed in Xenopus oocytes and human embryonic kidney 293 cells, respectively. Data were analyzed using nonlinear mixed-effects modeling to account for random variance in the peak current response between oocytes. The oils caused an insurmountable inhibition of 5-HT3-evoked currents (citral IC50 = 120 μM; eucalyptol = 258 μM; linalool = 141 μM) and did not compete with fluorescently labeled granisetron, suggesting a noncompetitive mechanism of action. Inhibition was not usedependent but required a 30-second preapplication. Compound washout caused a slow (~180 seconds) but complete recovery. Coapplication of the oils with bilobalide or diltiazem indicated they did not bind at the same locations as these channel blockers. Homology modeling and ligand docking predicted binding to a transmembrane cavity at the interface of adjacent subunits. Liquid chromatography coupled to mass spectrometry showed that an essential oil extracted from Lippia alba contained 75.9% citral. This inhibited expressed 5-HT3 receptors (IC50 = 45 μg ml⁻¹) and smooth muscle contractions in rat trachea (IC50 = 200 μg ml⁻¹) and guinea pig ileum (IC50 = 20 μg ml⁻¹), providing a possible mechanistic explanation for why this oil has been used to treat gastrointestinal and respiratory ailments. These results demonstrate that citral, eucalyptol, and linalool inhibit 5-HT3 receptors, and their binding to a conserved cavity suggests a valuable target for novel allosteric modulators.

Introduction

The natural oils citral, eucalyptol, and linalool (Fig. 1) are widely used as scents and flavorings in pharmaceuticals, foods, and health care products. Related compounds are reported to have antioxidant, anti-inflammatory, antiproliferative, antimicrobial, and acaricide activities. They belong to the terpenoid class of molecules that contains both structurally complex (e.g., bilobalide and ginkgolide) and simple (e.g., thymol and menthol) compounds (Caputi and Aprea, 2011). Terpenoids have effects on a broad selection of both voltage-gated and ligand-gated ion channels, and some are noncompetitive ligands of 5-hydroxytryptamine 3 (5-HT3) receptors (Hall et al., 2004; Ashoor et al., 2013; Kessler et al., 2014; Lansdell et al., 2015; Ziemba et al., 2015).

5-HT3 receptors belong to the Cys-loop family of transmembrane ligand-gated ion channels, which are responsible for fast excitatory and inhibitory neurotransmission in the central and peripheral nervous systems. It includes vertebrate nicotinic acetylcholine, GABA and glycine receptors, invertebrate receptors such as the glutamate-gated chloride channel (GluCl) and resistance-to-dieldrin channels (RDL), and prokaryotic homologs such as Erwinia chrysanthemi ligand-gated ion channel (ELIC) and Gloeobacter violaceus ligand-gated ion channel (GLIC) (Thompson et al., 2010). Each channel comprises five subunits surrounding a central ion-conducting pore, and each subunit has three distinct domains referred to as extracellular, transmembrane, and intracellular. The orthosteric binding site (that occupied by endogenous agonist) is located in the extracellular domain at the interface of two adjacent subunits, where binding is coordinated by the convergence of six peptide loops (Hassaine et al., 2014). The transmembrane domain of each subunit contains four α-helices (M1–M4), with M2 from each forming a central ion-conducting pore; to aid comparisons of the channel-lining residues of M2 from different Cys-loop receptors, a prime notation is often used to define residue positions, with O’ representing a conserved charged residue at the cytoplasmic side of the membrane. The intracellular domain regulates receptor trafficking, intracellular modulation, and ion channel conductance, but remains structurally unresolved (Hassaine et al., 2014).

5-HT3 receptor ligands typically target extracellular and transmembrane domains. Competitive antagonists such as granisetron

ABBREVIATIONS: BB, bilobalide; DMEM, Dulbecco’s modified Eagle’s medium; DTZ, diltiazem; ELIC, Erwinia chrysanthemi ligand-gated ion channel; G-FL, granisetron-fluorescein; GLIC, Gloeobacter violaceus ligand-gated ion channel; GluCl, glutamate-gated chloride channel; HEK293, human embryonic kidney 293; LC-MS, gas-liquid chromatography coupled to mass spectrometry; OELa, essential oil extracts from Lippia alba; PRED, predicted agonist-induced response; RDL, resistance-to-dieldrin channels; RUV, residual unexplained variance.
Materials and Methods

Materials. Citral, eucalyptol, linalool (Fig. 1), and 5-hydroxytryptamine were from Sigma-Aldrich (St. Louis, MO). Human 5-HT3A (accession number: P46098) subunit cDNA was provided by J. Peters (University of Dundee, Dundee, UK). Essential oil of L. alba was purchased and a species voucher was deposited on Prisco Bezerra Herbarium (Federal University of Ceará, Ceará, Brazil) with the following number identification: EAC-08474. Essential oil extracts from L. alba (OELa) were analyzed by LC-MS at Parque de Desenvolvimento Tecnológico (Ceará, Brazil).

Oocyte Maturation. Oocytes from Xenopus laevis were purchased from EcoCyte Biosciences (Castrop-Rauxel, Germany) and stored at 16°C in ND96 (96 mM NaCl, 2 mM KCl, 1 mM MgCl2, 5 mM Hepes, pH 7.5).

Cell Culture. Human embryonic kidney 293 (HEK293) cells were grown on 90-mm round tissue culture plates as monolayers in Dulbecco’s modified Eagle’s medium (DMEM/F12 Gibco, Life Technologies, Carlsbad, CA) supplemented with 10% fetal bovine serum (Sigma-Aldrich) at 37°C in a moist atmosphere containing 5% CO2.

Receptor Expression. 5-HT3A subunit cDNA was cloned into pcDNA3.1 for expression in HEK293 cells (ThermoFisher Scientific, Waltham, MA). Cells were transiently transfected with this cDNA using polyethylenimine Ultra T7 Transcription kit (Ambion, Austin, TX). Stage V and VI oocytes were injected with 50 nl of 100–500 ng µl−1 DNA (5–25 ng injected), and currents were recorded 1–4 days postinjection.

Electrophysiology. Using two-electrode voltage clamp, Xenopus oocytes were routinely clamped at −60 mV using an OC-775 amplifier (Warner Instruments, Hamden, CT). Human embryonic kidney 293 (HEK293) cells were grown on 90-mm round tissue culture plates as monolayers in DMEM/HAM’s F12 (Gibco, Life Technologies, Carlsbad, CA) and filled with 3 M KCl. Pipette resistances ranged from 0.8 to 2.0 MΩ. Oocytes were placed in a perfusion chamber made from 2-mm-wide × 30-mm-long silicon tubing that was cut in half lengthways (total volume ~0.1 ml), and were perfused with ND96 at a rate of 12 ml min−1. Drug application was via a simple gravity-fed system calibrated to run at the same rate. For inhibition measurements, antagonists were routinely applied for 1 minute before coapplication with 5-HT. A 3-minute wash was used between compound applications. Oils were dissolved in buffer containing 1% DMSO, freshly prepared each day, and constantly stirred during the experiments.

Flow Cytometry. HEK293 cells expressing the 5-HT3 receptor were grown in monolayers and harvested from a 90-mm culture dish using 10 ml of Trypsin-EDTA (Sigma-Aldrich) for 10 minutes at 37°C. Digestion was terminated by the addition of 25 ml DMEM + 10% fetal bovine serum, and cells were pelleted at low speed for 2 minutes. The pellet was resuspended in 3 ml of phosphate-buffered saline (137 mM NaCl, 8.0 mM Na2HPO4, 2.7 mM KCl, 1.47 mM KH2PO4, pH 7.4), and cells were filtered through a cell strainer (BD Falcon, Franklin Lakes, NJ). Competition binding was measured by incubating HEK293 cells with different concentrations of nonlabeled ligands and 10 nM fluorescent granisetron (G-F). After 10-minute incubation, cells were pelleted and rapidly washed in phosphate-buffered saline before being resuspended in the same buffer and analyzed on a BD Accuri C6 flow cytometer (Becton, Dickinson and Company, Franklin Lakes, NJ) at 488-nm excitation/530-nm emission. The geometric mean was measured at each concentration of test compound and fitted to eq. 1 (see
the following section) using a least-squares method (GraphPad Prism v4; GraphPad Software, La Jolla, CA).

**Nonlinear Mixed-Effects Modeling.** Inhibition of 5-HT–induced currents was analyzed using Wings for NONMEM (distributed under a GNU General Public License) and NONMEM 7.3.0 (Icon PLC, Dublin, Ireland). NONMEM is typically used for population pharmacokinetic/pharmacodynamic analyses and is ideally suited to simultaneously model fixed nonlinear effects (e.g., drug concentration-response relationships) and random effects (e.g., variance in maximal peak current). Nonlinear mixed-effects modeling of this sort cannot easily be done using more familiar statistical packages. Specifically, NONMEM allows the modeling of the relationship between drug and response, and importantly, the random between-oocyte variance, which is considerable. NONMEM enabled a single unified model to be created that included all electrophysiological agonist and antagonist data. This comprised 532 individual data points from 55 oocytes from the following experiments: 1) control 5-HT concentration-response data; 2) 5-HT concentration-response data in the presence of test compounds; and 3) concentration-inhibition data at fixed 5-HT (typically 1.7 μM) concentrations. The ability to allow for between-oocyte variance enabled raw peak currents to be analyzed, rather than normalized values. Normalization can obscure relationships between parameters, resulting in inaccurate and imprecise estimates of drug effects. Differences in peak currents and agonist potency between different oocytes were modeled with population variances (σ²) associated with model parameters.

A structural model defined the relationship between the independent variables (i.e., agonist and antagonist concentrations) and the peak current (dependent variable). The agonist response was modeled using the four-parameter logistic equation:

\[
PRED = \frac{Min_0 - Max_0}{1 + \left[\frac{|A|}{10^{-pEC_{50}(A)}}\right]^{nH}} + Max_0
\]

(1)

where \(PRED\) = predicted agonist-induced response, \(|A|\) = agonist concentration; \(Min_0\) = \(Max_0\) when \(|A| = 0\); \(Max_0 = PRED\) when \(|A| = \infty\); \(pEC_{50}(A) = -log_{10}|A|\) that induces \(PRED = (Max_0 + Min_0)/2\); and \(nH\) = agonist Hill coefficient. (In all cases, there was no basal current; hence, \(Min_0\) was fixed to zero.)

Current responses can change during an experiment. If this effect is not identified and quantified, it can distort estimates of agonist and compound effects. The effect of time on the agonist response was modeled linearly as:

\[
Max_T = Max_0 (1 + (T × n_T))
\]

(2)

where \(Max_T\) = as in eq. 1; \(Max_0 = PRED\) when \(|A| = \infty\) at time \(T\); \(T = time\) (hours) from start of individual experiment; and \(n_T = change\) in response per hour expressed as a proportion of the response at \(T = 0\) (\(n_T = 0\) represents no change; \(n_T = 0.1\) represents a reduction of 10%, and \(n_T = 0.1\) represents an increase of 10% per hour).

The effect of the compounds on \(Max_T\) was modeled as follows:

\[
Max_{D(i)} = \frac{Max_T}{1 + \left[\frac{|D_i|}{10^{-pIC_{50}(A)}}\right]^{nD(i)}}
\]

(3)

where \(Max_{D(i)} = PRED\) when \(|A| = \infty\) in the absence of drug \(D\) at time \(T\) (eq. 2); \(Max_{D(i)} = PRED\) when \(|A| = \infty\) in the presence of drug \(D\) at time \(T\); \(|D| = concentration\) of inhibitor drug; \(pIC_{50} = -log_{10}|D|\) in the presence of which \(Max_{D(i)} = Max_T/2\); and \(nD(i) = inhibition\) Hill coefficient. (This model assumes that, when \(|D| = \infty\), \(PRED = 0\).)

The effect of the compounds on agonist \(pEC_{50}\) was modeled as follows:

\[
pEC_{50(D)} = pEC_{50(0)} - \left[log_{10}\left(\frac{|D|}{10^{-pIC_{50}(A)}}\right)^{nD(i)} + 1\right]
\]

(4)

where: \(pEC_{50(0)} = agonist\) \(pEC_{50}\) in the absence of drug \(D\) (eq. 1); \(pEC_{50(D)} = agonist\) \(pEC_{50}\) in the presence of drug \(D\); \(|D| = concentration of drug; pA_{2(D)} = pA_D\) of drug \(D\); \(n_{D(D)} = Schild\) coefficient for drug \(D\); and \(n_{D(D)} = inhibition\) coefficient for drug \(D\).

Variable Schild (\(n_{S}\)) and inhibition (\(n_{I}\)) coefficients enabled compound effects other than competitive-like inhibition to be modeled, and hypotheses about drug action on \(pEC_{50}\) to be statistically evaluated using likelihood ratio tests. For example, when \(n_S = 1\) and \(n_I = 1\), the effect on agonist \(pEC_{50}\) is consistent with that of a competitive antagonist, and when \(n_I = 0\), there is no effect of the drug on agonist \(pEC_{50}\).

Random effects (\(\eta\)) were included, enabling parameter values from different oocytes to differ from the typical, population parameter estimates. The relationship between population and individual oocyte parameters was modeled in the following two ways:

\[
Par_{IND} = Par_{POP} × e^{\eta}
\]

(5)

\[
Par_{IND} = Par_{POP} + \eta
\]

(6)

where \(Par_{IND}\) = individual parameter estimate; \(Par_{POP}\) = population parameter estimate; and \(\eta\ = \eta\ value\ representing\ the\ difference\ between\ the\ population\ and\ an\ individual\ (i) parameter estimate.

Values of \(\eta\) from all oocytes (\(n = 55\)) enabled a variance-covariance (OMEGA) matrix for random effects to be constructed. This allows the random variance inherent in different oocytes to be defined and correlations in estimated parameters to be identified.

Residual unexplained variance (RUV) defines the difference between the observed and predicted values for a given set of conditions. RUV was modeled as a function of \(PRED\) as follows:

\[
RUV = \alpha^2 × PRED^\gamma
\]

(7)

where \(RUV = residual\ unexplained\ variance\ between\ observed\ and\ predicted\ current; PRED = predicted\ current; \alpha^2 = variance\ parameter; and \gamma = variance\ scaling\ parameter.\) When \(\gamma = 0\), RUV is constant irrespective of \(PRED\) and \(\alpha = standard\ deviation\ of\ the\ residual\ variability; when \(\gamma = 1\), RUV is proportional to the response (\(\alpha^2 = RUV/PRED\); when \(\gamma = 2\), the coefficient of variation of the residual variability is constant (\(\alpha = SD_{PRED}/PRED\)).

Maximum likelihood was used to identify best fit parameters for specific models. A Laplacian method was used to obtain parameter estimates. The objective function used by NONMEM is the extended least squares. Parameter standard errors are from the covariance step in NONMEM. Models were evaluated by examining both population- and individual-weighted residuals and comparing Akaike and Bayesian information criteria. Specific hypothesis tests are defined in Table 1. They were performed by constraining parameters and comparing resultant differences in extended least squares values using likelihood ratio tests (Spalding and Jarvis, 2002; Mould and Upton, 2013).

**Drug Effects at 5-HT₃AT₆ Receptors.** The effects of the compounds on 5-HT₃AT₆ receptors were also evaluated. A total of 113 data values from 11 oocytes comprising control 5-HT concentration-response and compound concentration-inhibition curves were incorporated into the wild-type receptor data set. Parameters were incorporated into the model that defined changes in \(pEC_{50}, pIC_{50}, Max_0,\) and \(pA_2\) values for 5-HT₃AT₆ receptors. These parameters were statistically evaluated as described earlier.

**Dual Application.** Dual application studies were performed as previously described (Jarvis and Thompson, 2013). This is a simple method to determine whether two channel blockers share the same binding site (syntopic inhibition) or bind to separate locations (allostic inhibition). For each test compound, inhibition of a supramaximal 5-HT–induced (100 μM) response was measured alone and in combination with bilobalide (BB) and diltiazem (DTZ), both well characterized 5-HT₃ blockers (Fig. 1; Thompson et al., 2011a). Concentrations of the test compounds, BB and DTZ, were selected such that they caused approximately 62% inhibition when used alone. From the results of these experiments, predicted levels of inhibition were derived for both
TABLE 1
Output from the best fit model describing the effects of 5-HT and inhibitors on the peak current

| Parameter | Population Parameter Estimate | Population Variance Model | Population SD(adj) or CORRb | Hypothesis Tests | P Value (LRT) | Interpretation |
|-----------|--------------------------------|---------------------------|-----------------------------|-----------------|---------------|----------------|
| Agonist (5-HT) | Min (μA) 0 (fixed) | — | — | — | — | Max varies between oocytes |
| | Max (μA) 4.94 ± 0.64 | EXP(μ2) 0.95 ± 0.10 | | H0: μx = 0 | No resulta | Max and pEC50 correlate strongly |
| | COV(μx,μ3) 0.87 ± 0.05 | +μ1 0.18 ± 0.02 | +μ3 0.19 ± 0.06 | H0: μx = 0 | 0.052 | Weak evidence for a change in response with time |
| | pEC50 5.65 ± 0.03 | +μ1 0.18 ± 0.02 | +μ3 0.19 ± 0.06 | H0: μx = 0 | 0.004 | nT varies between oocytes |
| | nG 2.94 ± 0.13 | +μ1 0.18 ± 0.02 | +μ3 0.19 ± 0.06 | H0: μx = 0 | 0.004 | Weak evidence for a change in response with time |
| Citral | pIC50 3.92 ± 0.05 | +μ1 0.16 ± 0.04 | 2 × 10–11 | pEC50 varies between oocytes |
| | nG 1.34 ± 0.08 | 5 × 10–6 | Indication of cooperative binding |
| | nG,GA 0 (fixed) | 0.94 | No evidence that citral changes agonist pEC50 |
| Eucalyptol | pIC50 3.59 ± 0.11 | +μ1 0.32 ± 0.07 | 1 × 10–16 | pEC50 varies between oocytes |
| | nG 1.04 ± 0.11 | 7.6 ± 0.7 | Suggests one inhibitor binding site |
| | nG,GA 1 (fixed) | 4 × 10–15 | Strong evidence that eucalyptol changes agonist pEC50 |
| | pA2 3.09 ± 0.09 | +μ1 0.18 ± 0.06 | 0.006 | pIC50 varies between oocytes |
| | nG 1.70 ± 0.24 | 0.001 | Schild slope greater than that for competitive antagonism |
| Linalool | pIC50 3.85 ± 0.02 | +μ1 0.05 ± 0.02 | 0.023 | pEC50 varies between oocytes |
| | nG 2.19 ± 0.26 | 0.46 | Suggests two highly cooperative inhibitor binding sites |
| | nG,GA 0 (fixed) | 0.11 | No/weak evidence that linalool changes agonist pEC50 |
| RUV model | γ 1.30 ± 0.07 | 1 × 10–63 | Residual error variance increases with the current amplitude |
| | α 0.30 ± 0.01 | — | — | — | — | — |

COV, covariance; LRT, likelihood ratio test.
aNo stable convergence was obtained without a random effect included.
bParameter estimates are shown ± standard errors.

allotropic and syntopic modes of action. These predictions were compared with data obtained experimentally using the same concentrations of the inhibitors acting together. For each oocyte, the change in the amplitude of the 5-HT response over time was monitored and taken into account. Inhibition by drugs was quantified in relation to interpolated control responses for each time point.

The dual application measurements were compared statistically with allotropic and syntopic predictions using a two-way analysis of variance in which the oocyte was included as a random effect (SPSS Statistics 20; IBM, Armonk, NY). Post-hoc testing was with Dunnett’s method, comparing the measured data to each of the predictions.

Modeling and Ligand Docking. Using ClustalW (EMBL-EBI, Cambridge, UK), the protein sequence of the human 5-HT3A subunit (accession: P46098) was aligned with the sequence from the mouse 5-HT3A crystal structure (Protein Data Bank ID: 4PIR). Sequence identity between mouse and human 5-HT3A was 89.1% (EMBOSS Needle; EMBL-EBI; McWilliam et al., 2013) and alignments were unambiguous. Five pentameric homolog models were generated using Modeler 9.13 (Andrej Sali, San Francisco, CA) with default parameters, and the best model was selected using Ramachandran plot analysis. Citral, eucalyptol, and linalool were constructed ab initio in Chem3D Ultra 7.0 (CambridgeSoft, Cambridge, UK) and energy minimized using the MM2 force field. Potential binding sites were identified using a 20-A docking sphere centered on the a-carbon of L320, a residue located within the center of M2; therefore, within each subunit, the docking sphere encompassed the full length of all transmembrane a-helices. Docked poses were generated using the GOLD docking program (version 3.0; Cambridge Crystallographic Data Centre, Cambridge, UK) with the GOLDScore function and default settings. For each compound, 10 docking poses were generated, and the poses and predicted hydrogen bonds were visualized with PyMol v1.3 (Schrodinger, New York, NY).

Ileum Preparation. Guinea pig ileum was obtained from adult male guinea pigs (200–300 g). The ileum was cut into 1-cm lengths and mounted longitudinally to a force transducer with a resting tension of 0.5 g in a 10-ml water-jacketed organ bath containing Krebs’ buffer (118 mM NaCl, 4.7 mM KCl, 1.2 mM MgSO4, 1.25 mM CaCl2, 11 mM glucose, 10 mM HEPES, pH 7.2) continuously aerated with 95% O2/5% CO2 and kept at 37°C. The ileum segments were allowed to equilibrate for 10 minutes before the experiments were started, and contractile responses were recorded using LabChart 6 (ADInstruments Ltd., Oxford, UK). 5-HT was applied to the serosal layer of the ileum, and responses were recorded using LabChart 6 (ADInstruments Ltd., Oxford, UK).

The trachea was removed from male Wistar albino rats (250–350 g), cut into 2-cm lengths, and mounted on a force transducer with a resting tension of 1.0 g in a 10-ml water-jacketed organ bath containing modified Tyrode’s solution (136 mM NaCl, 5 mM KCl, 0.98 mM MgCl2, 0.36 mM NaH2PO4, 2 mM CaCl2, 1.18 mM KH2PO4, 11.55 mM glucose, 10 mM HEPES, pH 7.2) continuously aerated with 95% O2/5% CO2 and kept at 37°C. The tracheal segments were allowed to equilibrate for 10 minutes before the experiments were started, and contractile responses were recorded using LabChart 6 (ADInstruments Ltd., Oxford, UK). 5-HT was applied to the serosal layer of the trachea, and responses were recorded using LabChart 6 (ADInstruments Ltd., Oxford, UK).

Pharmacological characterization of the ileum and trachea was performed using a variety of agonists and antagonists to assess a possible 5-HT3 receptor. Agonists included 5-HT, 5-Hydroxytryptophan (5-HTP) and 5-Hydroxytryptamine creatine phosphate (5-HTP). Antagonists included the specific 5-HT3 receptor antagonist Ondansetron, the selective 5-HT3 receptor agonist, and 5-HT3 receptor antagonist, and the non-selective 5-HT receptor antagonist, and the non-selective 5-HT receptor antagonist.

When the ileum and trachea were treated with 5-HT, a concentration-dependent increase in contractility was observed, with maximal responses occurring at concentrations of 1 µM. The degree of contraction was measured and analyzed using GraphPad Prism 8.0.2 (GraphPad Software, San Diego, CA) to determine the pIC50 and pEC50 values. The pIC50 values were determined using a competitive inhibition model, and the pEC50 values were determined using a non-competitive inhibition model. The results were analyzed using one-way analysis of variance (ANOVA) with post-hoc comparisons performed using Tukey’s HSD test. The level of significance was set at p < 0.05. The correlation between pIC50 and pEC50 values was determined using Pearson’s correlation coefficient (r).

Inhibition by drugs was quantified in relation to interpolated control responses for each time point. The degree of inhibition was determined using a sigmoidal logistic curve fitting method, and the results were analyzed using one-way analysis of variance (ANOVA) with post-hoc comparisons performed using Tukey’s HSD test. The level of significance was set at p < 0.05. The correlation between pIC50 and pEC50 values was determined using Pearson’s correlation coefficient (r).
Results

Effects of Citral, Eucalyptol, and Linalool on 5-HT$_3$ Receptor Currents. Neither 5-HT nor the terpenoid oils (Fig. 1) had observable effects on uninjected Xenopus oocytes. By contrast, application of 5-HT to Xenopus oocytes expressing 5-HT$_3$ receptors produced concentration-dependent, rapidly defined concentration-dependent inhibitor effects on the effects of the compounds. The model included parameters that mixed-effects modeling to characterize and quantify the current values from 55 oocytes were analyzed using nonlinear but varied substantially between oocytes. A total of 532 peak maximum response (Max$_0$) and pEC$_{50}$ of 5-HT.

Antagonist granisetron (100 nM).

Inhibit 5-HT$_3$ receptors and defined the conditions for sub-slow recovery from inhibition was also seen when the three middle), or during (Fig. 2D, bottom) 5-HT application. The inhibition was similar whether the compounds were applied in the absence of (Fig. 2D, top), immediately prior to (Fig. 2D, middle), or during (Fig. 2D, bottom) 5-HT application. The slow recovery from inhibition was also seen when the three compounds were added and removed in the continuous presence of 5-HT, as no rebound currents were seen, unlike the more rapidly recovering channel blocker diltiazem (Fig. 2E).

These results show that citral, eucalyptol, and linalool inhibit 5-HT$_3$ receptors and defined the conditions for subsequent experiments.

Quantification and Mechanism of Block. Studies at varying 5-HT (0.3–300 μM), citral (10, 30, 100, 200, 300, 500, 600, and 1000 μM), eucalyptol (100, 200, 500, 660, and 2000 μM), and linalool (100, 200, 500 μM) concentrations were carried out at −60 mV with a 30-second preapplication of oils and a 3-minute washout. Peak current responses were reproducible within oocytes but varied substantially between oocytes. A total of 532 peak current values from 55 oocytes were analyzed using nonlinear mixed-effects modeling to characterize and quantify the effects of the compounds. The model included parameters that defined concentration-dependent inhibitor effects on the maximum response (Max$_0$) and pEC$_{50}$ of 5-HT.

The results from the best fit model are graphically represented in Fig. 3, with the parameter values shown in Table 1, along with the outcome of hypothesis tests of these parameters. Figure 4A shows a linear relationship between observed and predicted peak current values. The most parsimonious and stable variance-covariance model consisted of seven random effects (Table 1). The analysis with NONMEM also revealed a strong covariance between the maximum response (Max$_0$) and pEC$_{50}$ of 5-HT (Fig. 4B: correlation = 0.869 ± 0.054), indicating that oocytes with a higher peak current were more sensitive to 5-HT. Failure to account for this covariance, such as by normalizing data, misrepresents the variance in the responses and could lead to inaccurate conclusions about the effects of the inhibitors on 5-HT pEC$_{50}$ values. For example, the insets in Fig. 3 show normalized representations of the same data revealing apparent shifts in the EC$_{50}$ values for citral and linalool that were not evident when the covariance was taken into account. Furthermore, IC$_{50}$ values derived from these normalized data were lower (e.g., higher potency; citral = 98 μM; eucalyptol = 174 μM; linalool = 83 μM).

In the population of oocytes used (n = 55), the 5-HT pEC$_{50}$ was 5.65 ± 0.03 (EC$_{50}$ = 2.25 μM) with a Hill coefficient of 2.94 ± 0.13 (Table 1). This is similar to previous findings and is consistent with strong positive cooperativity between three agonist binding sites (Thompson et al., 2011a; Thompson and Lummis, 2013). Neither citral nor linalool altered the pEC$_{50}$ of 5-HT, but both caused a concentration-dependent reduction in the maximal peak current response (Fig. 3; Table 1). For citral, the pIC$_{50}$ was 3.92 ± 0.05 (IC$_{50}$ = 120 μM), and for linalool, 3.85 ± 0.02 (IC$_{50}$ = 141 μM). Eucalyptol also caused a reduction in the maximal peak current with a pIC$_{50}$ of 3.59 ± 0.11 (IC$_{50}$ = 258 μM), although this was accompanied by a reduction in the pEC$_{50}$ of 5-HT (Fig. 3; Table 1) with a pA$_2$ and apparent Schild coefficient (defining the shift in the agonist pEC$_{50}$) of 3.09 ± 0.09 and 1.70 ± 0.24, respectively.

These results are consistent with a simple noncompetitive mechanism of action for each compound, but suggest that eucalyptol may have additional effects.

Effects on a 5-HT$_3$AT$_6$S Receptor Mutant. Thymol and carvacrol activate human 5-HT$_3$ receptors from a transmembrane binding site (Landsdell et al., 2015). The compounds studied here are structurally similar and could have weak partial agonist activities that are difficult to detect at wild-type receptors. The agonist sensitivity of the 5-HT$_3$ receptor can be enhanced by a T6$S$ substitution in the pore-lining M2 α-helix of the 5-HT$_3$A subunit (Thompson and Lummis, 2013). We therefore investigated the effects of 5-HT and the terpenoid oils in oocytes expressing the 5-HT$_3$AT$_6$S mutant. A total of 112 peak current responses from 11 different oocytes were appended to the wild-type data set. These experiments comprised control 5-HT concentration-response data and concentration-inhibition data for 2 μM 5-HT. Parameters were included in the model that allowed differences in agonist and inhibitory effects to be quantified and evaluated.

There was no difference in the maximum 5-HT peak current in the wild-type and 5-HT$_3$AT$_6$S receptors (Max$_{[5HT]}$/Max$_{[5HTWT]}$ = 0.98 ± 0.30; P = 0.93), but the mutant was more sensitive to agonist as previously reported (Thompson and Lummis, 2013). The 5-HT pEC$_{50}$ was increased by 0.35 ± 0.07. Unlike the effects at wild-type receptors, eucalyptol did not alter the 5-HT pEC$_{50}$ (H0: pEC$_{50}$ AT6$S$/pEC$_{50}$ WT = 1; P = 0.71). When applied alone, citral, eucalyptol, and linalool did not evoke currents in the 5-HT$_3$AT$_6$S receptor mutant, but all abolished 5-HT–induced responses with inhibitory potencies that were
reduced by approximately 2-fold when compared with their effects at wild-type receptors (Table 2).

**Competitive Binding with Granisetron.** Our analysis provides no evidence that citral or linalool competes with 5-HT at the orthosteric site, whereas the effect of eucalyptol on 5-HT \( \text{pEC}_{50} \) suggests that this ligand might. To further test for orthosteric interactions, we measured the binding of a fluorescent granisetron derivative (G-FL) together with the three compounds using flow cytometry (Jack et al., 2015). Both 5-HT and granisetron reduced G-FL binding in a...
concentration-dependent manner (Fig. 5A). Binding of G-FL was unaffected by citral, eucalyptol, or linalool at concentrations up to 1 mM (Fig. 5B).

**Dual Application of Citral, Eucalyptol, or Linalool in the Presence of Bilobalide or Diltiazem.** We have previously shown that simultaneous application of two drugs can be used to probe the sites of action of channel blockers (Jarvis and Thompson, 2013). Here, we applied each of the oils in the presence of bilobalide or diltiazem, channel-blocking antagonists that bind at the 2'-6' and 7' regions of the 5-HT₃ receptor pore, respectively (Thompson et al., 2011a).

For citral (Fig. 6A), measured dual inhibition was no different from the allotopic predictions for bilobalide ($P = 0.95$) or diltiazem ($P = 0.42$), but was greater than the syntopic predictions (BB, $P = 0.031$; DTZ, $P = 0.0004$). For eucalyptol (Fig. 6B), dual inhibition was similar to the allotopic
prediction for bilobalide ($P = 0.38$) but greater than the allotopic prediction for diltiazem ($P = 0.013$). For linalool (Fig. 6C), dual inhibition was greater than the allotopic prediction for both bilobalide ($P = 0.02$) and diltiazem ($P = 0.06$).

These results suggest that citral, eucalyptol, and linalool do not share binding sites with the channel blockers bilobalide or diltiazem, but may enhance their effects.

**Transmembrane Binding Sites.** Given the lipophilicity of the oils and their noncompetitive mechanism of action, we sought to identify potential binding sites in the transmembrane domain using in silico ligand docking in a homology model of the human 5-HT$_3$ receptor. Using a loosely defined binding site radius of 20 Å that encompassed the whole of the transmembrane domain, the major binding cavity for all of the ligands was predicted to be at the interface of two adjacent subunits, between M1-M2 of the principal subunit and M2-M3 of the complementary subunit. For linalool, all 10 docked poses were similarly orientated at the intersubunit interface within only a 2.29-Å root-mean-square deviation (Fig. 7A). Eucalyptol docked at two locations, both of which were clustered at the same site between two adjacent subunits (Fig. 7B). For citral, two sites were predicted, with the major docked pose cluster (6/10) at the same intersubunit cavity, and a minor site (4/10) at the lipid-exposed intracellular interface of the M1 and M4 α-helices (Fig. 7C). Linalool was the only compound predicted to make hydrogen bond contacts with the protein (in 4/10 docked poses), which were between the hydroxyl of linalool and the backbone carbonyl of Thr6' in M2 (Fig. 7D).

These results suggest that all three compounds could bind to a transmembrane cavity located at the interface of adjacent subunits.

**Physiologic Effects.** *L. alba* is used medicinally throughout Central and South America as a means of alleviating gastrointestinal discomfort and for respiratory ailments (Hennebelle et al., 2008). Here, we analyzed an OELA by LC-MS and detected that the main compounds were the terpenoids citral (75.9%; 41.8% geranial + 34.1% neral), 1-limonene (9.8%), carvone (8.9%), gamma-terpinene (2.0%), and benzene [1-methyl-3-(1-methylethyl; 1.0%]. To determine whether the properties of this essential oil could be physiologically relevant, we measured its effects on 5-HT$_3$ receptors expressed in oocytes and on smooth muscle contractions in rat trachea and guinea pig ileum. At 5-HT$_3$ receptors expressed in oocytes, OELA had no effect when applied alone, but fully and reversibly inhibited the 2 mM 5-HT response with an IC$_{50}$ of 45 μM (Fig. 8A). In rat trachea, 5-HT-induced contractions were concentration-dependent and substantially inhibited by 10 μM of the 5-HT$_3$ receptor antagonist granisetron (Fig. 8B). OELA also abolished contractions induced by 10 μM 5-HT with an IC$_{50}$ of 200 μg ml$^{-1}$ (Fig. 8C). 5-HT-evoked contractions of guinea pig ileum were also inhibited by granisetron, and OELA had an IC$_{50}$ of 20 μg ml$^{-1}$ (n = 6; Fig. 8D). In both rat trachea and concentrations; for citral and linalool, the apparent shifts in the EC$_{50}$ seen in this normalized data are artifacts caused by the normalization. Additional data at 10, 30, 300, 600, and 1000 μM citral and 200 and 660 μM eucalyptol were also collected, but the curves are omitted for clarity.
guinea pig ileum, recovery of contractions required extended washes, particularly at higher concentrations (Fig. 8E).

These results show that OELa has a high citral content and inhibits 5-HT3 receptors expressed in Xenopus oocytes and 5-HT–induced smooth muscle contraction in the trachea and ileum.

**Discussion**

This study describes the inhibitory effects of citral, eucalyptol, and linalool on human 5-HT3 receptors. These terpenoids can be added to a growing list of structurally related compounds that modulate a wide range of voltage- and ligand-gated ion channels, including 5-HT3 receptors and their related vertebrate, invertebrate, and prokaryotic homologs (Hales and Lambert, 1991; Barann et al., 2000; Hall et al., 2004; Garcia et al., 2006; Ashoor et al., 2013; Walstab et al., 2014; Lansdell et al., 2015; Ziemba et al., 2015).

All of the essential oils inhibited 5-HT–mediated currents with IC50 values in the micromolar range. Non-competitive antagonism was shown by them causing a concentration-dependent reduction in the maximal 5-HT response and by not competing with the fluorescent antagonist G-FL. The Hill coefficients for the reduction in the 5-HT response suggested that there may be multiple binding sites with strong cooperativity for linalool (nH = 2.2), weak cooperativity for citral (nH = 1.3), and no cooperativity for eucalyptol (nH = 1.0). There was evidence that, at higher concentrations, eucalyptol also caused a rightward shift in the EC50 of 5-HT (pA2 = 3.09), indicating an additional inhibitory mechanism. A competitive mechanism is conceivable, although not compelling, given that the shift had a Schild coefficient of 1.7 and that eucalyptol failed to compete with G-FL. However, given the different binding orientations of 5-HT and granisetron in cocrystal structures, it is possible that an inhibitor could compete with one of these ligands but not the other (Colquhoun, 2007; Kesters et al., 2013). At the T6S mutants, there was no evidence of a eucalyptol-induced change in agonist EC50. This is probably because this effect occurs at higher concentrations (pA2 = 3.09) than the reduction in maximal current (pIC50 = 3.59), and the overall potency of the terpenoids was reduced at these mutants.

Mixed-effects modeling also revealed a strong correlation between the maximal peak current and pEC50 (Fig. 5B). Since receptor expression is difficult to control, many investigators normalize data to facilitate comparisons between experiments. However, if the implicit assumption that agonist response is independent of expression levels is not justified, then data normalization may lead to inaccurate and misleading conclusions. In the current study, a particular advantage of the nonlinear mixed-effects modeling was that variant maximal current responses could be modeled as a random effect, thereby generating more accurate quantitative conclusions (normalization yielded higher IC50 values, suggesting that this method of analysis overestimated the potency of the compounds) and revealing unexpected biologic phenomena (such as the covariance of the EC50 and Max0). This modeling approach was also able to identify pleiotropic drug effects (e.g., eucalyptol), for which the properties of the compounds we used were particularly well suited. Therefore, the data we present clearly highlight the superior value of nonlinear mixed-effects modeling, revealing phenomena that would otherwise have been missed using the standard procedure of normalizing data. Nonlinear mixed-effects modeling is often used for pharmacokinetic-pharmacodynamic data analysis, but has broad applicability in other quantitative pharmacological studies (Mould and Upton, 2013). For example, by modeling

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**TABLE 2**

Effect of a T6S mutation on inhibitor pIC50 values

| Agonist | ΔpEC50 | P (H0: ΔpEC50 = 0) | pEC50 | pEC50* |
|---------|--------|-------------------|-------|-------|
| 5-HT    | 0.35 ± 0.07 | 3 × 10−6 | 6.00 ± 0.08 |
| Inhibitor |         |                  |       |       |
| Citral  | −0.36 ± 0.10 | 0.002 | 3.55 ± 0.11 |
| Eucalyptol | −0.45 ± 0.22 | 0.046 | 3.14 ± 0.05 |
| Linalool| −0.34 ± 0.04 | 5 × 10−7 | 3.51 ± 0.24 |

*aStandard errors were calculated from the errors of the pEC50/pIC50 and ΔpEC50/ΔpIC50 values using propagation of error calculations.

bThere was no significant difference between the ΔpIC50 values for the three inhibitors (P = 0.87).
sensitivity to 5-HT could contribute to this effect, a possibility that is supported by the rightward shift in the 5-HT EC50 caused by eucalyptol.

The lipophilic nature of the oils suggests that the binding sites of citral, eucalyptol, and linalool are located in the transmembrane region, consistent with the slow wash-in and washout that is observed when compounds must first diffuse into membranes before reaching their target (Turina et al., 2006). This hypothesis is supported by our electrophysiology and flow cytometry data, which indicate a non-competitive mechanism of action, and by the nonoverlapping binding sites predicted by our dual-application experiments. To probe for potential binding sites, we used homology modeling and docking, which predicted that all three oils share a common binding site located in a cavity between the transmembrane α-helices of adjacent subunits. The terpenoids carvacrol and thymol have been proposed to bind in a similar cavity within the 5-HT3 receptor, and this location is also conserved among other Cys-loop receptors (Lansdell et al., 2015). Crystal structures of the invertebrate Cys-loop receptor glutamate-gated chloride channel reveal an equivalent intersubunit cavity occupied by ivermectin, and residues at this location are implicated in ivermectin binding at GABA receptors (Lynagh and Lynch, 2012). A similar binding site has been identified for propofol at a prokaryotic homolog (Gloeobacter violaceus ligand-gated ion channel) and other eukaryotic Cys-loop receptors (Nury et al., 2011; Jayakar et al., 2013; Yip et al., 2013; Lynagh and Laube, 2014). The similar activities of these compounds across the Cys-loop receptor family suggest that this region is widely conserved and could be an amenable target for novel allosteric modulators (Corradi et al., 2011; Howard et al., 2011; Trattnig et al., 2012). To date, 5-HT3 receptor antagonists have been typically used to prevent nausea and vomiting, but the association of these receptors with other disorders, such as irritable bowel syndrome, anxiety, and diabetic neuropathy, suggests that there may be scope for other therapeutic applications (Walstab et al., 2010; Silva et al., 2015).

Previously, we used a dual-application method to determine whether channel blockers have overlapping or independent binding sites within the 5-HT3 receptor pore (Thompson et al., 2011b; Jarvis and Thompson, 2013). Results suggesting that bilobalide, ginkgolide B, and picrotoxinin shared binding sites and that diltiazem bound elsewhere were later confirmed by mutating channel-lining residues (Thompson et al., 2011a). Dual application of citral with bilobalide or diltiazem caused inhibition consistent with the allotopic model, indicating that citral binds at distinct sites to these two channel blockers. Eucalyptol and linalool caused dual inhibition at levels significantly greater than the syntopic predictions, indicating that these compounds also bind to distinct sites. However, dual inhibition exceeded the allotopic prediction, suggesting that eucalyptol and linalool may allosterically modulate binding of bilobalide and diltiazem. Alternatively, as the allotopic model assumes that the inhibitors do not modify the action of the agonist, it is possible that a reduced sensitivity to 5-HT could contribute to this effect, a possibility all data simultaneously (e.g., by incorporating both agonist concentration-activation and antagonist concentration-inhibition data), it generates a more comprehensive model of drug action and a statistically more powerful framework within which to evaluate specific hypotheses. This is particularly valuable when it is important to minimize use of resources, such as experimental animals or scarce test compounds. The structurally related compounds carvacrol and thymol are also partial agonists at the 5-HT3 receptor (Lansdell et al., 2015; Ziemba et al., 2015). As low-efficacy partial agonism can sometimes be overlooked and mistaken as antagonism [e.g., the 5-HT3 ligand quipazine (Thompson and Lummis, 2013)], we tested citral, eucalyptol, and linalool for agonist activity using a 5-HT3 receptor mutant with enhanced agonist sensitivity. Substitution of the 5-HT3A subunit channel-lining 6′ Thr with the equivalent 6′ Ser from the 5-HT3B subunit creates a hypersensitive mutant with increased sensitivity to a range of 5-HT3 agonists (Thompson and Lummis, 2013). Consistent with previous reports, our results reveal a 2-fold increase in 5-HT sensitivity in the T6S mutant. However—with citral, eucalyptol, and linalool—no agonist action, or potentiation of the 5-HT response, was observed at the mutant receptor, confirming that they are antagonists. In silico docking predicted that the 6′ Thr residue may establish a hydrogen-bond interaction with linalool, although the interaction was with the backbone carbonyl and therefore unlikely to be greatly affected by amino acid substitution. Indeed, a 2-fold reduction in the potency of all the inhibitors at 5-HT3A T6S mutants suggests a nonspecific effect rather than a modification of hydrogen-bond interactions for linalool alone.

Fig. 5. Competition of a fluorescent 5-HT3 receptor competitive antagonist. (A) Competition of 10 nM G-FL with the agonist 5-HT (pIC50 = 6.10 ± 0.03, nH = 2.1, IC50 = 0.79 μM, n = 4) and the competitive antagonist granisetron (pIC50 = 8.42 ± 0.03, nH = 1.1, IC50 = 3.8 nM, n = 6). (B) The test compounds citral (n = 3), eucalyptol (n = 3), and linalool (n = 3) show no competition with G-FL.
Fig. 6. Inhibition of the 5-HT$_3$ receptor using a dual-application approach. 5-HT$_3$ receptors were activated with a supramaximal (100 µM) concentration of 5-HT. Concentrations of BB, DTZ, and the terpenoids were preselected to inhibit the response by approximately 62% when used alone. Results are shown in the white bars. These same concentrations were then applied in dual combinations, giving results shown in the black bars. The gray bars are the allotopic (allo) and syntopic (syn) predictions based on the levels of inhibition caused by the compounds alone. Data are the mean ± S.E.M. Two-way analysis of variance with Dunnett’s post-hoc test was used (IBM SPSS Statistics 20) to compare actual and predicted dual application responses. (A) Citral (n = 7). For BB, $P = 0.019$ (analysis of variance), $P = 0.95$ (H$_0$: BB+citral = allo), $P = 0.031$ (H$_0$: BB+citral = syn); and for DTZ, $P = 0.0005$ (analysis of variance), $P = 0.42$ (H$_0$: DTZ+citral = allo), $P = 0.0004$ (H$_0$: DTZ+citral = syn). (B) Eucalyptol (n = 6). For BB, $P = 0.015$ (analysis of variance), $P = 0.38$ (H$_0$: BB+eucalyptol = allo), $P = 0.009$ (H$_0$: BB+eucalyptol = syn); and for DTZ, $P = 0.00008$ (analysis of variance), $P = 0.013$ (H$_0$: DTZ+eucalyptol = allo), $P = 0.00004$ (H$_0$: DTZ+eucalyptol = syn). (C) Linalool (n = 5). For BB, $P = 0.0006$ (analysis of variance), $P = 0.023$ (H$_0$: BB+linalool = allo), $P = 0.0004$ (H$_0$: BB+linalool = syn); and for DTZ, $P = 0.003$ (analysis of variance), $P = 0.056$ (H$_0$: DTZ+linalool = allo), $P = 0.002$ (H$_0$: DTZ+linalool = syn). In all experiments, stable levels of inhibition were achieved by applying the compounds for 1 minute before 5-HT was added.
Several plant extracts are commonly used to treat gastrointestinal discomfort and respiratory disorders, and their components, such as citral and menthol, are reported to have relaxant effects on gut smooth muscle (Tangpu and Yadav, 2006; Hennebelle et al., 2008, Devi et al., 2011; Walstab et al., 2014). Here, we show that OELa, an extract from L. alba, inhibits 5-HT\textsubscript{3} receptors expressed in oocytes (IC\textsubscript{50} = 45 µg ml\textsuperscript{-1}) and 5-HT-evoked contractions in both rat trachea (IC\textsubscript{50} = 200 µg ml\textsuperscript{-1}) and guinea pig ileum (IC\textsubscript{50} = 20 µg ml\textsuperscript{-1}). Using LC-MS, we found the principal component of this oil was citral (∼560 Jarvis et al., 2012). Therefore, in humans, it is unlikely that systemic administration of typical doses would result in concentrations that are active at 5-HT\textsubscript{3} receptors, although it is possible that higher local concentrations following topical administration (e.g., in airway, skin, or gut) could reach pharmacologically active levels (Falk-Filipsson et al., 1993). For most terpenoids, including linalool and eucalyptol, blood and tissue concentrations have not been reported, but toxicological studies suggest that many are well tolerated (Oz et al., 2015). This suggests that there is still scope for therapeutic applications, with synthetic modification possibly providing a means for improving their potency and receptor selectivity.

In summary, we used nonlinear mixed-effects modeling to show that the oils citral, eucalyptol, and linalool inhibit homomeric 5-HT\textsubscript{3} receptors via noncompetitive mechanisms. Both electrophysiology and flow cytometry point to binding locations that do not overlap with the orthosteric binding site, whereas our dual-application experiments suggest actions that are mediated from outside the pore. Docking predicts a transmembrane binding site located between the \(\alpha\)-helices of adjacent subunits, and is supported by the binding of related compounds to similar allosteric sites identified in both 5-HT\textsubscript{3} and other members of this ligand-gated ion channel family. These results demonstrate the value of analyzing data using nonlinear mixed-effects modeling and further highlight a conserved transmembrane binding site as a potential target for the development of novel allosteric ligands.

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**Fig. 7.** Predicted binding locations for citral, eucalyptol, and linalool in a homology model of the human 5-HT\textsubscript{3} receptor. Using a loosely defined binding site (see Materials and Methods), 10 docked poses were generated for each ligand. Examples from the docked pose clusters (sphere representation) are shown at each of the predicted binding sites. The transmembrane domains of the five subunits that form the functional 5-HT\textsubscript{3} receptor are shown as different colored ribbons viewed from the extracellular side, with both the intracellular and extracellular domains removed for clarity. (A) A single binding site was predicted for linalool with the docked pose cluster differing by only 2.29 Å root-mean-square deviation. The white arrowhead indicates the origin from which (D) is viewed. (B) Two sites were predicted for eucalyptol, but were similarly located at the boundaries of adjacent subunits. (C) For citral, two potential binding sites were identified, a major (6/10) site at the interface of adjacent subunits and a minor site located at the lipid-exposed interface at the intracellular ends of M1 and M4. (D) In four of the 10 docked poses for linalool, PyMol 1.3 predicted hydrogen bonds (blue dotted line) between the ligand’s terminal hydroxyl and the backbone of the channel-lining 6’ Thr residue.
Fig. 8. The effects of OELa. (A) Concentration inhibition of 2 μM 5-HT–induced currents by OELa in Xenopus oocytes expressing the 5-HT₃ receptor (n = 5). (B) Rat tracheal contraction in response to 5-HT in the absence (▼, n = 6) and following a 10-minute preapplication with 10 μM granisetron (▼, n = 3). (C) Concentration-inhibition of the 10 μM 5-HT–evoked tracheal contractile response by OELa (n = 4). (D) Inhibition by OELa of 10 μM 5-HT–evoked guinea pig ileum contractions (n = 6). (E) Example recordings from 5-HT–evoked (black bars) contractions of guinea pig ileum and their inhibition by 30, 60, and 100 μg ml⁻¹ OELa. At concentrations ≥60 μg ml⁻¹ OELa, the 5-HT–evoked contractions became increasingly slow to recover. Following a control 5-HT response (0.6 μM, black bar), the addition of 100 μg ml⁻¹ OELa continued to inhibit the 5-HT–evoked contractions 50 minutes later, although an acetylcholine (1 μM, gray bar) response was unaltered. Parameters defining the curves are shown in the text.
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Authorship Contributions

Participated in research design: Thompson. Conducted experiments: Thompson, Barboa. Contributed new reagents or analytic tools: Barboa, Jarvis. Performed data analysis: Thompson, Jarvis. Wrote or contributed to the writing of the manuscript: Thompson, Jarvis.

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