Case Report

Symptomatic Apical Periodontitis of the Mandibular First Molar with the Accessory Canal in the Furcation Area Mimicking Furcation Perforation

Alexei Bolyachin, Zurab Khabadze, Oleg Mordanov, Magomed Gasbanov, and Takhir Teberdiev

1Iceberg Dental Trauma Center, Moscow, Russia
2RUDN University (People’s Friendship University of Russia), Moscow, Russia

Correspondence should be addressed to Oleg Mordanov; mordanov19@gmail.com

Received 1 August 2021; Revised 2 April 2022; Accepted 12 April 2022; Published 12 May 2022

Academic Editor: Konstantinos Michalakis

Copyright © 2022 Alexei Bolyachin et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Apical periodontitis frequently presents as a chronic disease. To arrive at a true diagnosis, in addition to the clinical examination, it is mandatory to undertake radiographic examinations and evaluate the clinical presentation. Knowledge of the root canal morphology is a prerequisite for effective nonsurgical endodontic treatment. The internal morphological features of the pulp chamber are variable and complex. This case report describes the treatment and outcome of symptomatic apical periodontitis of a mandibular first molar with the accessory (chamber) canal. The applied treatment fully contributed to the periapical lesion regression as shown in the four-year recall periapical radiography.

1. Introduction

Apical periodontitis is mainly a root canal infection, characterized by inflammation and destruction of periapical and periradicular tissues resulting from the interaction between microbial factors and the host immune response [1, 2]. Destruction of the periodontal ligament is triggered by degradation of the extracellular matrix by metalloproteinases (MMPs) [3] involving periradicular inflammation and bone destruction mediated by proinflammatory cytokines [4].

Normal oral microorganisms and bacteria may give rise to opportunistic pathogens if access to dental pulp tissues appears. Openings in the physical dentin barriers (enamel and cementum) by means of caries, cracks, or traumatic injuries create pathways for bacteria into the pulp chamber and root canal system consequently [5].

Communication between the pulp and the periodontal ligament may occur via the apical foramen and lateral and accessory canals in the apical and coronal parts of the root. Despite careful selection of cases and the use of correct procedures, endodontic treatment occasionally fails, and it can be due to the presence of communications between the pulp tissues and periodontal tissues [6].

Accessory canals result due to a localized failure in the formation of Hertwig’s sheath during the embryonic stages of tooth formation. This defect is probably due to the persistence of abnormally placed blood vessels reaching the pulp, which is more common in the furcation region [6].

These canals are also known as interradicular canals [7]. In 2018, Ahmed et al. [8] suggested a new accessory canal classification. The proposed classification provides an accurate description of the position and configuration of the accessory and chamber canals. By chamber canals, the authors mean small canals leaving the pulp chamber that (usually) communicates with the external surface of the root (including the furcation) (Figure 1).
The aim of this case report is to present the treatment and healing of apical chronic periodontitis with isolated furcation bone and apical bone destruction.

2. Case Report

A 27-year-old female patient with noncontributory general medical history presented and was referred to the dental practice in Moscow, Russia. The patient presented to the clinic with the complaint that included nonacute pain forcing on the tooth load during mastication from tooth # 46.

Pulp testing with carbon dioxide did not show any expressive reaction. Percussion was positive. Periodontal probing depth was 3 mm in average.

Periapical radiograph revealed a deep carious lesion in the distal surface of the tooth, communicating with the pulp chamber; there is presence of a radiolucency in the furcation region, not connected with the apical bone lesions present in both roots. No endodontic treatment had been previously performed (Figure 2).

The pulp chamber floor examination with the operative microscope (Carl Zeiss Meditec AG, Jena, Germany) with different zooming revealed no furcation, perforation, or fracture.

The following treatment strategy was applied. Coronal root canal portion was instrumented with Protaper SX (Dentsply Maillefer, Ballaigues, Switzerland) following a glide path development with C-Pilot (VDW, Munich, Germany) 6-15 ISO sizes. The shaping was provided with FlexMaster (Vereinigte, Dentalwerke, Munich, Germany) 15-35 ISO sizes, 4% tapered.

Irrigation protocol included the following: 3% NaOCl, 19% EDTA with the EndoActivator System (Dentsply Tulsa Dental Specialties, Tulsa, OK, USA). Pulp chamber irrigant solution was intensively activated with the ultrasound E4 tip, 3 cycles. Every cycle time was 30 seconds with 30% of ultrasound power in the general mode.

After two weeks, the medication was removed with the instrumentation with FlexMaster and irrigation with 3% NaOCl. As the patient was asymptomatic, the canals were dried and filled. The obturation was performed using continuous wave technique with gutta-percha and AH Plus (Dentsply International Inc., York, PA, USA) (Figure 3).

After obturation, the patient was referred to the restorative treatment. The tooth was restored with a lithium
disilicate full crown. The patient was recalled after 6, 12, 24, and 48 months (Figure 4). Total periapical and furcation healing was observed after 12 months and confirmed by the Cone Beam Computed Tomography (CBCT) scan 48 months after obturation (Figure 5).

3. Discussion

Different researches aimed at observing interradicular canals in lower molars have reported similar or different results, methodologies, terminology definition, and sample numbers [10].

Wolf et al. [10] observed 9 (7.7%) interradicular canals communicating the pulp chamber floor with the bifurcation area using dye in extracted teeth. The authors investigated the pulp chamber floor and the bifurcation area surface of 117 mandibular first molars. Access cavities were prepared; the pulp chambers were flooded with methylene blue and then centrifuged. An average of 4.2 (0.145 ± 0.03 mm thickness) slices per tooth were obtained by means of a diamond band saw. The presence of interradicular canals and diverticula was investigated using a light microscope (125x). Perlich et al. [11] found 3 canals (4.8%) using scanning electron microscopy and 22 canals (64.5%) using light microscopy in the pulp chamber floor of 62 human maxillary and mandibular molars. Chouchi et al. [12] studied accessory canals in 57 extracted permanent human teeth with micro-CT and revealed 7% (n = 4) and 21% (n = 12) furcation canals in the first and second mandibular molars, respectively.

Furcation perforation is an injury that leads to a communication between the root canal and the periodontal tissues or the oral cavity. It can be caused by resorption or dental caries or can have an iatrogenic cause including the misaligned use of rotary burs during access, preparation, and search for the root canals [13]. Furcation involvement refers to the condition when periodontal disease has caused bone resorption at the bifurcation or trifurcation of a multirooted tooth [14, 15]. Clinical examination with magnification, vital tests, and no previous endodontic treatment can help to exclude furcation impairment by periodontal disease.

Successful endodontic treatment depends on the thorough debridement of the entire root canal system. Difficult canal anatomy could be one of the causes of improper cleaning of the canal system, with the permanence of microorganisms [16, 17].

Irrigation of the root canals plays an important role in the disinfection process. From a microbiological standpoint, an irrigant should be able to demonstrate antimicrobial and antibiofilm activities and inactive endotoxins [18]. Activation of the irrigant helps to improve root canal cleanliness [19, 20]. Al-Jadaa et al. [21] demonstrated that ultrasonic irrigation causes a rise in the irrigant temperature in the accessory canals to 53.5 ± 2.7°C after the fifth minute, which could improve the irrigation effectiveness. No significant influence of accessory canal position or angulation was found.

Temporary restorative materials are often used during endodontic treatment to seal the root chamber between sessions or until a permanent restoration is placed. An ideal temporary restorative material should exhibit no leakage, good abrasion and compression resistance, lack of porosity, and lack of dimensional changes, and it must also be easily manipulated or removed while being effective in a moist environment [22]. PTFE is also an inexpensive material and previously used in dentistry in different fields. PTFE tape did not provide an avenue for bacterial contamination as temporary spacer materials [23]; however, PTFE retained by friction and may not be applicable in some other conditions, so it needs to be retained with stable material, such as the lite-cured composite resins [24]. E. faecalis grows on and penetrates through PTFE significantly more than it does with IRM (the most commonly used temporary restorations among specialists [24]), revealing its comparably reduced overall antimicrobial sealing ability when placed as the base part of temporary restorations [25]. Furthermore, research has shown that IRM has a poor marginal seal, performs poorly under stress, and exhibits bacterial penetration through the bulk of the material [26]; that is why composite
resin creates extra the protection layer from leakage. The use of the composite resin as a full external layer increases the cost of the endodontic treatment.

The “double seal” technique [9] involves placing composite resin as the deeper layer material inside the pulp chamber and access cavity. The IRM is then used as the outer layer which is exposed to loading and the oral cavity. This double layer functions in several ways: the outer layer of IRM is an antibacterial agent; the inner layer of composite resin prevents liquids from the oral cavity to reach the root canal system if it has been able to penetrate through the IRM margins. The white colour of the IRM is readily visible when the clinician needs to remove it. The IRM is also a cheap material that is easily and quickly mixed and placed in the tooth. It sets quickly, and therefore, there is no “waiting time” after placement before the rubber dam can be removed.

4. Conclusion

Activation of the irrigant associated with the instrumentation and the use of the intracanal medication could explain the repair of the furcation and periapical lesions after endodontic treatment.

Figure 5: CBCT scan of tooth # 46 in 48 months after obturation. Full bone recovery is noticed.

Conflicts of Interest

The authors declare no conflict of interests.

References

[1] P. N. Nair, “Pathogenesis of Apical Periodontitis and the Causes of Endodontic Failures,” Critical Reviews in Oral Biology and Medicine, vol. 15, no. 6, pp. 348–381, 2004.
[2] J. J. Segura-Egea, J. Martín-González, and L. Castellanos-Cosano, “Endodontic medicine: connections between apical periodontitis and systemic diseases,” International Endodontic Journal, vol. 48, no. 10, pp. 933–951, 2015.
[3] T. Sorsa, L. Tjäderhane, Y. T. Konttinen et al., “Matrix metalloproteinases: contribution to pathogenesis, diagnosis and treatment of periodontal inflammation,” Annals of Medicine, vol. 38, no. 5, pp. 306–321, 2006.
[4] N. Kawashima and P. Stashenko, “Expression of bone-resorptive and regulatory cytokines in murine periapical inflammation,” Archives of Oral Biology, vol. 44, no. 1, pp. 55–66, 1999.
[5] I. Prada, P. Micó-Muñoz, T. Giner-Lluesma, P. Micó-Martínez, N. Collado-Castellano, and A. Manzano-Saiz, “Influence of microbiology on endodontic failure. Literature review,”
[6] P. Poornima and V. V. Subba Reddy, “Comparison of digital radiography, decalcification, and histologic sectioning in the detection of accessory canals in furcation areas of human primary molars,” Journal of the Indian Society of Pedodontics and Preventive Dentistry, vol. 26, no. 2, pp. 49–52, 2008.

[7] A. L. Anderegg, D. Hajdarevic, and T. G. Wolf, "Interradicular Canals in 213 Mandibular and 235 Maxillary Molars by Means of Micro-computed Tomographic Analysis: An Ex Vivo Study," Journal of Endodontia, vol. 48, no. 2, pp. 234–239, 2022.

[8] H. M. A. Ahmed, P. Neelakantan, and P. M. H. Dummer, "A new system for classifying accessory canal morphology," International Endodontic Journal, vol. 51, no. 2, pp. 164–176, 2018.

[9] A. L. Jensen, P. V. Abbott, and S. J. Castro, "Interim and temporary restoration of teeth during endodontic treatment," Australian Dental Journal, vol. 52, 1 Suppl, pp. S83–S99, 2007.

[10] T. G. Wolf, S. Wentaschek, R. J. Wierichs, and B. Briseño-Marroquín, "Interradicular Root Canals in Mandibular First Molars: A Literature Review and Ex Vivo Study," Journal of Endodontia, vol. 45, no. 2, pp. 129–135, 2019.

[11] M. A. Perlich, A. Reader, and D. W. Foreman, "A scanning electron microscopic investigation of accessory foramen on the pulpal floor of human molars," Journal of Endodontia, vol. 7, no. 9, pp. 402–406, 1981.

[12] D. Chouchi, A. Berberoğlu, K. Orhan, I. Etikan, H. Tümer, and N. Başmış, "The location and incidence of patent accessory pulpal canals in permanent molars with periodontal lesion by using micro-computed tomography," Journal of Medical Imaging and Health Informatics, vol. 11, no. 1, pp. 85–88, 2021.

[13] S. D. Huamán, M. G. Brito Aragão, A. P. Dias Moreno et al., "Accuracy of conventional periapical radiography in diagnosing furcation repair after perforation treatment," Journal of Endodontia, vol. 46, no. 6, pp. 827–831, 2020.

[14] W. Zhang, K. Foss, and B. Y. Wang, "A retrospective study on molar furcation assessment via clinical detection, intraoral radiography and cone beam computed tomography," BMC Oral Health, vol. 18, no. 1, p. 75, 2018.

[15] E. S. Gusmão, A. C. Picarte, M. B. Ben Barbosa, C. K. Rösing, and R. Cimoes, "Correlation between clinical and radiographic findings on the occurrence of furcation involvement in patients with periodontitis," Indian Journal of Dental Research, vol. 25, no. 5, pp. 572–575, 2014.

[16] S. Gautam, S. R. Galgali, H. S. Sheethal, and N. S. Priya, "Pulpal changes associated with advanced periodontal disease: a histopathological study," Journal of Oral and Maxillofacial Pathology, vol. 21, no. 1, pp. 58–63, 2017.

[17] E. Weinberg, A. E. Pereda, S. Khurana, P. P. Lotlikar, C. Falcon, and C. Hirschberg, "Incidence of middle mesial canals based on distance between mesial canal orifices in mandibular molars: a clinical and cone-beam computed tomographic analysis," Journal of Endodontia, vol. 46, no. 1, pp. 40–43, 2020.

[18] P. R. Shenoi, E. S. Morey, C. S. Makade, M. K. Gunwal, R. T. Khode, and S. S. Wanmali, "In vitro evaluation of the antimicrobial efficacy of chitosan and other endodontic irrigants against Enterococcus faecalis," General Dentistry, vol. 64, no. 5, pp. 60–63, 2016.

[19] X. Guo, H. Miao, L. Li et al., "Efficacy of four different irrigation techniques combined with 60 °C 3% sodium hypochlorite and 17% EDTA in smear layer removal," BMC Oral Health, vol. 14, p. 114, 2014.

[20] C. Garcia-Romano, A. Zubizarreta-Macho, M. R. Baquero-Artigaño, and J. Ortega-Díaz, "An analysis of intracanal bacterial load before and after chemo-mechanical preparation: a comparative analysis of two irrigants and two activation techniques," Journal of Clinical and Experimental Dentistry, vol. 8, no. 1, pp. e9–e13, 2016.

[21] A. Al-Jadaa, F. Paqué, T. Attin, and M. Zehnder, "Necrotic pulp tissue dissolution by passive ultrasonic irrigation in simulated accessory canals: impact of canal location and angulation," International Endodontic Journal, vol. 42, no. 1, pp. 59–65, 2009.

[22] A. Ciftçi, D. A. Vardarlı, and I. S. Sönmez, "Coronal microleakage of four endodontic temporary restorative materials: an in vitro study," Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontics, vol. 108, no. 4, pp. e67–e70, 2009.

[23] A. P. Paranjpe, S. Jain, K. J. Alibhai, C. P. Wadhwani, R. P. Darvea, and J. D. Johnson, "In vitro microbiologic evaluation of PTFE and cotton as spacer materials," Quintessence International, vol. 43, no. 8, pp. 703–707, 2012.

[24] K. Olcay, L. Steier, H. Erdogan, and S. Belli, "Polytetrafluoroethylene tape as temporary restorative material: a fluid filtration study," Journal of Istanbul University Faculty of Dentistry, vol. 49, no. 3, pp. 17–22, 2015.

[25] M. Solomonov, D. H. Levy, A. Yaya, J. Ben Itzhak, and D. Polak, "Antimicrobial evaluation of polytetrafluoroethylene used as part of temporary restorations: an ex vivo study," Australian Endodontic Journal, vol. 48, no. 1, pp. 98–104, 2022.

[26] T. Alswayyd, R. Bin Nafesah, M. Alqutub et al., "Restorative treatment patterns after root canal obturation in public hospitals in Riyadh," Clinical, Cosmetic and Investigational Dentistry, vol. Volume 13, pp. 389–394, 2021.