Chapter 7
Nicotinic Acetylcholine Receptors in Regulation of Pathology of Cerebrovascular Disorders

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Abstract Cerebrovascular disorders including ischemic stroke, intracerebral hemorrhage and subarachnoid hemorrhage are among the major clinical concerns for which effective therapies are poorly available. Accumulating lines of evidence indicate that drugs acting on nicotinic acetylcholine receptors (nAChRs) may provide therapeutic effects on these disorders, based on their neuroprotective and anti-inflammatory actions. For example, the cholinergic neurotransmission in the central nervous system via nAChRs may function as an endogenous neuroprotective system that prevents pathogenic events associated with ischemic stroke. On the other hand, exogenous administration of nicotine or nAChR agonists to experimental models of ischemic stroke has been reported to produce conflicting results (either protective or deleterious), which may be largely dependent on the different regimens of drug treatments. With regard to intracerebral hemorrhage, preclinical findings suggest that post-treatment with nAChR agonists is effective in alleviating brain tissue damage and neurological outcome. The beneficial actions of nAChR agonist have also been reported for an experimental model of subarachnoid hemorrhage, which should be confirmed by further investigations. Although smoking has been considered as an important risk factor for stroke episodes, specific targeting of the central nAChRs may prove to be an effective and novel strategy for the treatment of diverse types of cerebrovascular disorders.

Keywords Nicotine · Cerebral infarction · Hemorrhagic stroke · Neuroprotection · Neuroinflammation · Acetylcholinesterase inhibitor
7.1 Introduction

Acetylcholine (ACh) has a long history of biomedical research as it is the first compound that has been identified as a chemical neurotransmitter. In the periphery, motor neurons release ACh from their nerve terminals that innervate skeletal muscles and transmit signals via activation of muscle type nicotinic acetylcholine receptors (nAChRs) that are heteropentamers consisting of subunits named α1, β1, δ and ε. The cell bodies of sympathetic and parasympathetic postganglionic neurons express neuronal type of nAChRs composed of subunits distinct from those contained in muscle nAChRs, such as α3α5β4 and α3α5β2β4. These nAChRs expressed in autonomic ganglia receive cholinergic inputs from preganglionic neurons. Postganglionic neurons of the parasympathetic nervous system also utilize ACh as a neurotransmitter, and various peripheral tissues receive the signals from these neurons mainly via muscarinic ACh receptors.

Cholinergic neurotransmitter system is also present in the brain, and nAChRs as well as muscarinic receptors are widely distributed throughout the central nervous system (Taly et al. 2009). A notable feature of nAChR-mediated signal transmission in the central nervous system is that it may mediate various functions in addition to the role in fast neurotransmission. That is, stimulation of nAChRs in central neurons affords cytoprotective and survival-promoting effects, as discussed in other chapters as well as in the literatures elsewhere (Akaike et al. 2010; Mudo et al. 2007). In addition, nAChR stimulation may be able to limit inflammation in the brain under pathological conditions, by directly regulating the cells involved in inflammatory reactions such as brain-resident microglia and infiltrating monocytes/macrophages (Shytle et al. 2004; Wang et al. 2003). The principal nAChR subtypes expressed in the central nervous system are homopentamers consisting of α7 subunits and heteropentamers consisting of α4 and β2 subunits, although nAChRs containing other types of subunits such as α3, α6, β3 and β4 are also expressed in a brain region-specific manner (Taly et al. 2009; Zoli et al. 2015). Differences in subunit compositions between the peripheral nAChRs (either muscle type or neuronal type) and the central nAChRs may enable drug development specifically targeted for nAChRs in the brain, thereby avoiding adverse peripheral actions.

One of the major reasons for the central nAChRs being expected to serve as drug targets is that effective pharmacotherapies are not available at present for various kinds of central nervous system disorders associated with neurodegeneration. Among these disorders are cerebrovascular diseases resulting from the disruption of circulatory system that supplies blood flow to the brain. This chapter summarizes various preclinical and clinical findings related to the functions of nAChRs in regulating pathological events associated with cerebrovascular disorders, and discusses the potentials of nAChR-related drugs as novel therapeutics combating these disorders.
7.2 Overviews on Stroke Disorders

The term “stroke” represents diverse sets of disorders associated with disrupted cerebrovascular functions, which are classified into two major categories such as ischemic stroke and hemorrhagic stroke. Ischemic stroke is featured by interception of the blood flow in the brain tissues and, depending on the causes of interception, is further categorized into several different types including atherothrombotic brain infarction, cardiogenic embolism and lacunar stroke. In either case, the consequences of hypoperfusion are shortage of supplies of oxygen and glucose to the brain parenchymal cells. Because energy demands of neurons are met almost entirely by the aerobic metabolism of glucose, even short periods of ischemic episodes may result in severe and irreversible brain tissue damage accompanied by neuron loss. Rescuing neurons from cell death is extremely difficult in the ischemic core region where oxygen/glucose supply is almost totally abolished. In contrast, in the peri-infarct or “penumbra” region where oxygen/glucose supply is decreased but not abolished, neuronal cell death displays at least in part the features of programmed cell death that occurs with a certain delay from the onset of the insults. Therefore, neuroprotection in the penumbra after ischemic insults may be possible if appropriate treatment strategies are established (Catanese et al. 2017). Practically, however, only a few drugs effective in brain tissue preservation are available at present for the treatment of ischemic stroke (Table 7.1). A powerful approach is the reestablishment of blood flow by removal of the blood clot, and for this purpose, tissue plasminogen activator that promotes fibrinolysis is administered within 4.5 h after ischemic attack. Otherwise, edaravone is the sole choice of drugs in Japan intended for neuroprotection via its free radical-scavenging action.

The major types of hemorrhagic stroke are characterized by extravasation of blood into either brain parenchyma (intracerebral hemorrhage; ICH) or subarachnoid space (subarachnoid hemorrhage; SAH), and both cases are accompanied by poor prognosis. Hemorrhagic stroke has pathological features quite distinct from those of ischemic stroke. In ICH, tissue damage is caused primarily by the blood flowing out of ruptured vessels into the brain parenchyma. Biological actions of blood constituents (particularly, proteases involved in the coagulation cascade), in addition to the physical damage of blood mass (so called the “mass effect”) are the main contributors of pathogenic events in ICH (Qureshi et al. 2009). Currently, there are no established pharmacotherapies that are applicable after the occurrence of ICH events and are effective in neuroprotection (Katsuki 2010). Drugs such as glycerol and mannitol that manipulate plasma osmotic pressure and suppress brain edema formation may be available, but they still lack solid clinical evidence that justifies their application to ICH cases (Table 7.1). Clinical practice for SAH is in a similar situation to that for ICH, in the sense that direct neuroprotective pharmacotherapies are yet unavailable. Representative drugs used for the current treatment of SAH include fasudil, a Rho kinase inhibitor that prevents post-SAH vasospasm and ozagrel, an inhibitor of thromboxane A₂ synthase that prevents platelet aggregation.
Overall, although cerebrovascular disorders such as ischemic stroke, ICH and SAH are among the major issues of clinical concerns in the modern world, effective drug treatment strategies are, for the most part, far from established. With these situations under consideration, the following sections summarize the findings obtained from various kinds of efforts addressing the possibility of nAChRs as drug targets for the therapies of these disorders.

### 7.3 Ischemic Stroke and nAChRs

#### 7.3.1 Roles of Endogenous Cholinergic System in Regulation of Ischemic Injury

##### 7.3.1.1 Effects of nAChR Antagonists and Allosteric Modulators

Potential involvement of endogenous ACh in regulation of the pathogenic events in ischemic brain injury has been suggested by a study addressing the effects of nAChR antagonists in neonatal rats. In 7-day-old rats that underwent permanent occlusion of left common carotid artery, subcutaneous administration of subtype non-selective

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**Table 7.1** Current drug therapies applicable after cerebrovascular events

| Actions                  | Drug name                          | Grade | Time window     |
|--------------------------|------------------------------------|-------|-----------------|
| For ischemic stroke      |                                    |       |                 |
| Thrombolytic             | Tissue plasminogen activator       | A     | Within 4.5 h    |
|                          | Urokinase                          | B     | Within 6 h      |
| Anticoagulant            | Argatroban                         | B     | Within 48 h     |
|                          | Heparin                            | C1    | Within 48 h     |
| Anti-platelet            | Aspirin                            | A     | Within 48 h     |
|                          | Ozagrel                            | B     | Within 5 days   |
| Radical scavenger        | Edaravone                          | B     | Within 24 h     |
| Edema regulator          | Glycerol or mannitol               | C1    |                 |
| For ICH                  |                                    |       |                 |
| Anti-hypertensive        | Calcium blockers and others        | B or C1 |             |
| Edema regulator          | Glycerol or mannitol               | C1    |                 |
| Anti-fibrinolysis        | Tranexamic acid                    | C1    |                 |
| Capillary stabilizer     | Carbazochrome                      | C1    |                 |
| For SAH                  |                                    |       |                 |
| Anti-vasospasm           | Fasudil                            | A     |                 |
| Anti-platelet            | Ozagrel                            | A     |                 |
| Edema regulator          | Glycerol or mannitol               | C1    |                 |
| Anti-hypertensive        | Calcium blockers and others        | C1    |                 |

According to the Japanese Guidelines for the Management of Stroke 2015 by the Japan Stroke Society. Grade A, strongly recommended; Grade B, recommended; Grade C1, may be considered for application but scientific evidence is insufficient.
nAChR antagonist mecamylamine or specific α7 nAChR antagonist methyllycaconitine just before induction of 1-h hypoxia aggravated neural damage in the hippocampus. Mecamylamine also aggravated neural damage in the cerebral cortex (Furukawa et al. 2013). On the other hand, daily treatment with mecamylamine from 24 h after induction of 45-min transient global ischemia did not clearly affect the degree of cell death-related events such as the number of dead CA1 neurons and caspase-3 activity in the hippocampus in young adult mice (Ray et al. 2014).

PNU-120596 is a positive allosteric modulator of α7 nAChRs. The drug does not activate nAChRs by itself but inhibits desensitization and enhances activation of α7 nAChRs by orthosteric agonists. In addition, the ACh degradation product choline may produce agonistic activity at nAChRs in the presence of positive allosteric modulators. Indeed, Kalappa et al. (2013) demonstrated that choline delayed oxygen/glucose deprivation-induced injury of hippocampal CA1 neurons in vitro in the presence of PNU-120596, and PNU-120596 alone was not effective in the absence of choline in these experimental settings. The combined neuroprotective effect of choline and PNU-120596 was abolished by α7 nAChR antagonist methyllycaconitine. The neuroprotective potential of PNU-120596 was also confirmed in 90-min transient focal ischemia in rats, where 3-h pretreatment or 30-min posttreatment with the drug significantly reduced the infarct volume (Kalappa et al. 2013). A companion study by the same group reported that the effective time window of PNU-120596 administered intravenously was extended to 6 h after induction of middle cerebral artery occlusion (MCAO; Sun et al. 2013). These results strongly suggest that cholinergic neurotransmission functions as an endogenous neuroprotective system against ischemic brain injury.

7.3.1.2 Effects of Acetylcholinesterase Inhibitors

The concentration of ACh in the synaptic cleft is tightly regulated by acetylcholinesterase (AChE), as ACh released from the nerve terminals is promptly degraded by this hydrolyzing enzyme. Accordingly, application of AChE inhibitors is a conventional procedure to augment cholinergic neurotransmission. Several studies have addressed the effect of AChE inhibitors in rodent models of ischemic stroke. The earliest example is a study on donepezil, a centrally acting reversible inhibitor of AChE that is prescribed for amelioration of cognitive deficits in Alzheimer disease. An oral dose of donepezil (12 mg/kg) administered 2 h before or 1 h after induction of permanent MCAO in rats significantly reduced the infarct volume. The protective effect of donepezil was abolished by mecamylamine, suggesting the involvement of nAChR activation (Fujiki et al. 2005).

Another AChE inhibitor galantamine has also been reported to exhibit neuroprotective effects where the first administration was performed 24 h before or 3 h after induction of transient global ischemia in Mongolian gerbils by occlusion of the common carotid artery. The effects included an increase in surviving neurons, a decrease in terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling
(TUNEL)-positive cells/caspase-3-positive cells in hippocampal CA1 region and amelioration of spatial memory deficit, and these effects were attenuated by concurrent treatment with mecamylamine (Lorrio et al. 2007). A later examination using rat hippocampal slices subject to oxygen/glucose deprivation followed by reoxygenation revealed that a tyrosine kinase Janus kinase (Jak) 2 mediated the protective effect of galantamine by suppressing activation of nuclear factor (NF) -κB and induction of inducible nitric oxide synthase (iNOS) expression (Egea et al. 2012). Under similar experimental conditions in mouse hippocampal slices, 30-min pretreatment with nicotine (10–100 μM) reduced the extent of oxygen/glucose deprivation-induced cell injury via activation of α7 nAChRs (Egea et al. 2007).

Wang et al. (2008) reported the effect of huperzine A, yet another AChE inhibitor, in a rat model of transient focal cerebral ischemia. When administered at the onset or 6 h after transient MCAO, huperzine A restored regional cerebral blood flow, reduced infarct size and decreased neurological deficit score. Similar to the cases with galantamine, huperzine A inhibited NF-κB activation and decreased expression of proinflammatory factors such as tumor necrosis factor (TNF) -α, interleukin (IL) -1β, iNOS and cyclooxygenase (COX) -2. In addition, mecamylamine abolished the inhibitory effect of huperzine A on glial cell activation and partially reversed the reduction of infarct size by huperzine A.

Overall, these results obtained from the examinations on three different kinds of AChE inhibitors are consistent with the idea that endogenous ACh affords neuroprotective effects against ischemia and ischemia-reperfusion injury, by activating nAChRs (Fig. 7.1).

7.3.1.3 Effect of Cholinergic Neuronal Activity

Vagus nerve stimulation is known to produce several therapeutic effects, and in the case of ischemia/reperfusion injury in rats, a brief episode of vagus nerve stimulation ameliorates neurological deficits and reduces infarct volume (Sun et al. 2012). Jiang et al. (2014) addressed the mechanisms of the neuroprotective effects of vagus nerve stimulation. They confirmed that vagus nerve stimulation given at 30 min after transient focal ischemia was able to ameliorate neurological deficit and reduced the infarct volume at 24 h after reperfusion. They additionally found that the increase in the levels of several cytokines such as TNF-α, IL-1β and IL-6 in the peri-infarct region was significantly prevented by vagus nerve stimulation. Notably, vagus nerve stimulation prevented ischemia/reperfusion-induced loss of α7 nAChR expression in microglia in the penumbra region, and also reversed ischemia-induced decrease in the level of phosphorylated Akt. Although the evidence is at most indirect, these results imply the involvement of cholinergic neurotransmission pathway in the neuroprotective effect of vagus nerve stimulation.

As for microglial nAChRs, another line of findings in a positron emission tomography imaging study implicates α4β2 nAChRs in the pathology of cerebral ischemia.
Fig. 7.1  Enhancement of endogenous cholinergic signaling suppresses ischemia-induced pathological events. Stimulation of nicotinic acetylcholine receptor (nAChR) by endogenous acetylcholine (ACh) can be enhanced either by inhibitors of acetylcholinesterase (AChE) such as donepezil that promote accumulation of extracellular ACh, or by positive allosteric modulators (PAMs) of nAChRs such as PNU-120596 that increase the affinity and/or efficacy of ACh on nAChRs. PAMs may also allow choline to exert agonistic activity on nAChRs. Stimulation of nAChRs, particularly of α7 subtype, by these drugs leads to phosphorylation/activation of a tyrosine kinase Janus kinase 2 (Jak2). Although detailed mechanisms are yet to be identified, activated Jak2 may inhibit phosphorylation of inhibitor of κB (IκB) α, prevent ischemia-induced recruitment of nuclear factor κB (NF-κB; illustrated as subunits p65 and p50), and consequently, suppress NF-κB-dependent expression of pro-inflammatory factors (See Sects. 7.3.1.1 and 7.3.1.2 for details).

That is, α4β2 nAChR binding probed by $[^{18}F]$-fluoro-A85380 and $[^{11}C]$PK11195 in rat brains showed a transient increase that peaked at 7 days after MCAO (Martin et al. 2015). Increased expression of α4β2 nAChRs in microglia/macrophages and astrocytes was confirmed by immunohistochemical examinations. Although the functional significance of these changes in the expression of α4β2 nAChRs is unclear, this nAChR subtype in addition to α7 nAChRs may serve as a target for regulation of pathogenic events, particularly those involving inflammatory reactions, in ischemic brain injury.
7.3.2 Effects of nAChR Agonists on Ischemic Injury

7.3.2.1 Positive Findings

As discussed in the previous section (Sect. 7.3.1), stimulation of nAChRs appears to provide neuroprotective effects against ischemia-reperfusion injury. Indeed, many lines of evidence indicate that nicotine and other compounds with direct agonistic activity on nAChRs can provide beneficial effects on experimental model of ischemic stroke (Table 7.2). Kagitani et al. (2000) reported the effect of intravenous injection of nicotine in an intermittent transient ischemia model in rats. When rats with permanent ligation of bilateral vertebral arteries received transient and intermittent occlusion (two to three times for 2 min at 2-min intervals) of bilateral carotid arteries, the blood flow in the hippocampus was substantially and reversibly decreased, and delayed death of hippocampal CA1 neurons was induced. Administration of nicotine (30–100 μg/kg) at 5 min before occlusion slightly but significantly improved hippocampal blood flow during occlusion and increased the number of surviving CA1 neurons. Kagitani et al. (2000) assume that the neuroprotective effect of nicotine under these conditions is attributable to the vasodilative response induced by nAChR stimulation and resultant increase in regional blood flow in the hippocampus. On the other hand, the presence of nicotine during

**Table 7.2** Studies reporting beneficial effects of nicotine and nAChR agonists in experimental models of ischemia

| Conditions of drug treatment | References |
|-------------------------------|------------|
| Nicotine (30–100 μg/kg, i.v.) 5 min before transient ischemia in rats | Kagitani et al. (2000) |
| Nicotine (0.3–1.5 mg/kg, i.p.) 30 min before transient ischemia in Mongolian gerbils | Nanri et al. (1998b) |
| Nicotine hydrogen tartrate (1.2 mg/kg, i.p.) 2 h before transient ischemia in rats | Chen et al. (2013) |
| Nicotine (0.5 mg/kg, i.p.) 2, 6 or 12 h after transient ischemia followed by three times administration per day for 7 days | Guan et al. (2015) |
| Nicotine hydrogen tartrate (0.3 mg/kg, s.c.) twice daily for 12 days, from 2 days after focal devascularization | Gonzalez et al. (2006) |
| GTS-21 (1–5 mg/kg, i.p.) 30 min before, or GTS-21 (10 mg/kg, p.o.) twice daily for 2 weeks before, transient ischemia in Mongolian gerbils | Nanri et al. (1998b) |
| GTS-21 (1–10 mg/kg, p.o.) 24 h and 30 min before permanent ischemia in Mongolian gerbils | Nanri et al. (1998a) |
| PNU-282987 (10 mg/kg, i.p.) 60 min after thrombotic stroke in mice | Parada et al. (2013) |
| PHA-568487 (doses are not described, i.p.) immediately and 24 h after permanent ischemia in mice | Han et al. (2014b) |
| PHA-568487 (0.8 mg/kg, i.p.) 1 and 2 days after permanent ischemia in mice | Han et al. (2014a) |

See Sect. (7.3.2.1) in the main text for details of the findings in each study. i.p. intraperitoneal, i.v. intravenous, p.o. per oral, s.c. subcutaneous
ischemic episodes may produce direct protective effects on neuronal cells. For example, in rat primary cortical cultures, 4-h hypoxia induced neuronal cell death with features of apoptosis. Application of nicotine (10–100 μM) during hypoxia prevented cell death probably via activation of both α7 nAChRs and α4β2 nAChRs, as methyllycaconitine (α7 nAChR antagonist) and dihydro-β-erythroidine (α4β2 nAChR antagonist) blocked the effect of nicotine (Hejmadi et al. 2003).

GTS-21 is a synthetic nAChR agonist with fourfold higher affinity than nicotine for α7 nAChRs. The effect of GTS-21 and nicotine was tested on an ischemia-reperfusion model of Mongolian gerbils. When the drugs were injected intraperitoneally at 30 min before 3-min forebrain ischemia, both GTS-21 (1–5 mg/kg) and nicotine (0.3–1.5 mg/kg) improved performance of animals in passive avoidance and attenuated cell death in the hippocampal CA1 region. GTS-21 (10 mg/kg) was effective also when orally administered twice daily for 2 weeks prior to ischemia (Nanri et al. 1998b). Moreover, the same group examined the effect of GTS-21 in rats with permanent occlusion of bilateral common carotid arteries (Nanri et al. 1998a). In this study, GTS-21 (1 or 10 mg/kg) was administered orally 24 h and 30 min before induction of ischemia and then once daily for 2 months. GTS-21 significantly prevented ischemia-induced histopathological changes in the cerebral cortex and in the white matter, and improved the cognitive function of rats as assessed by radial maze learning performance.

Potential involvement of endocannabinoid system in the neuroprotective effect of nicotine has been proposed by a study by Chen et al. (2013). Intraperitoneal administration of nicotine hydrogen tartrate (1.2 mg/kg) transiently increased the tissue content of endocannabinoids 2-arachidonoylglycerol and anandamide in rats, which peaked at 2 h after administration. Nicotine also increased the protein expression level of cannabinoid CB1 receptor. Moreover, 2-h pretreatment with nicotine reduced the infarct volume and neurological deficit induced by 120-min MCAO, and these effects were significantly reversed by AM251, a CB1 receptor antagonist.

All of these findings mentioned above were obtained by pretreatment of nicotine or nAChR agonist. However, the pretreatment regimens of drugs are not very useful for combating stroke disorders, because in almost all clinical situations the drug therapy is available only after certain delay from the onset of acute stroke episodes. With these concerns, several studies have examined the effect of therapeutic treatment with nAChR agonists on ischemic injury and indeed reported positive results. In a study by Guan et al. (2015), transient forebrain ischemia was induced in rats by four-vessel occlusion. Nicotine (0.5 mg/kg) was administered intraperitoneally at 2, 6 or 12 h after induction of 15-min ischemia, and thereafter, three times per day for 7 days. Under these conditions, nicotine significantly increased the number of surviving CA1 neurons in the hippocampus. At the same time, nicotine was found to decrease the number of microglia and to prevent ischemia-induced up-regulation of mRNAs encoding TNF-α and IL-1β in the hippocampal CA1 region. In addition, nicotine inhibited proliferation of primary cultured microglia, and this effect was blocked by α7 nAChR antagonist α-bungarotoxin. These results suggest that attenuation of proliferation and activation of microglia contribute to the neuroprotective effect of the therapeutic treatment with nicotine.
Fig. 7.2  Specific α7 nAChR agonist may afford anti-inflammatory effects via expression of heme oxygenase-1 (HO-1) in the brains after ischemic insults. According to Parada et al. (2010, 2013), application of α7 nAChR agonist such as PNU-282987 activates Jak2 in microglia/macrophages, which leads to activation of PI3-kinase (PI3K)/Akt pathway. By the action of Akt, a transcription factor Nrf2 is liberated from Keap1, escapes from degradation and binds to antioxidant response element (ARE) of the promoter regions of genes encoding phase II detoxifying enzymes such as HO-1 and the catalytic subunit of glutamate cysteine ligase (GCL-c). HO-1 induced by these signaling pathways plays a critical role in inhibiting ischemia-associated pro-inflammatory responses such as the production of reactive oxygen species (ROS) and tumor necrosis factor-α (TNF-α)

Involvement of regulation of microglia in the therapeutic effect of α7 nAChR agonist has also been demonstrated in a photothrombotic ischemia model in mice (Parada et al. 2013). That is, intraperitoneal administration of PNU-282987 (10 mg/kg) at 60 min after induction of thrombotic stroke using Rose Bengal decreased the volume of cortical infarct and diminished neurological deficits. Notably, the protective effect of PNU-282987 was abolished by concomitant treatment with heme oxygenase (HO) inhibitor or by deletion of HO-1 gene. In hippocampal slice culture preparation, PNU-282987 induced HO-1 expression, prevented cell death and attenuated production of reactive oxygen species and TNF-α resulting from oxygen/glucose deprivation and reoxygenation. The protective effect of PNU-282987 in slice cultures was abolished by application of HO inhibitor or by depletion of microglia from the slices. These results propose HO-1 as a key mediator of the neuroprotective and anti-inflammatory effect of microglial α7 nAChR stimulation (Fig. 7.2).

Using another α7 nAChR agonist PHA-568487, Han et al. (2014b) examined the role of microglia/macrophage phenotypes. Mice were treated with PHA-568487 immediately and 24 h after permanent MCAO. The treatment significantly reduced the infarct volume and partially ameliorated behavioral performance. The peri-
infarct region of PHA-568487-treated mice contained a smaller number of pro-inflammatory microglia/macrophages (so called “M1” phenotype identified as CD11b+/Iba-1+) and a larger number of anti-inflammatory microglia/macrophages (so called “M2” phenotype identified as CD206+/Iba-1+), than that of saline-treated mice (Han et al. 2014b). Similar protective effect on neuron survival and regulatory effect on microglia/macrophage phenotypes were obtained when administration of PHA-568487 (0.8 mg/kg) was initiated at 1 day after permanent MCAO (Han et al. 2014a). These results suggest that the regulation of microglia/macrophage phenotypes underlies the neuroprotective effect of α7 nAChR agonist against ischemic injury.

The effect of repeated administration (twice daily for 12 days) of low dose nicotine (0.3 mg/kg as nicotine hydrogen tartrate) has been reported in a rat focal ischemia model prepared by unilateral devascularization in the motor cortex. The findings are notable in the sense that post-treatment started as late as 48 h after ischemic surgery was proven to be effective in promoting the recovery of motor performance. Interestingly, nicotine treatment increased the morphological complexity of dendritic processes of layer V pyramidal neurons of the cingulate cortex, both in control rats and ischemia-induced rats (Gonzalez et al. 2006). Therefore, nicotine-induced enhancement of dendritic growth of cortical neurons might facilitate the recovery of motor functions.

7.3.2.2 Negative Findings

Under several experimental conditions, nicotine has been reported to produce no effect or even worsen the outcome of ischemic events. For example, Lim et al. (2009) examined the effect of nicotine on rehabilitation following devascularization in the forelimb area of the motor cortex in rats. Oral administration of nicotine (0.3 mg/kg, twice daily for 3 weeks starting from 1 week before stroke induction) did not improve motor function recovery.

It should be noted that virtually all studies reporting the deleterious effect of nicotine on ischemic brain injury were aimed to reveal potential adverse effects of smoking on cerebrovascular disorders, and therefore that these studies generally employ long-term and/or continuous pretreatment regimen for nicotine administration (Table 7.3). An early study by Wang et al. (1997) examined the effect of nicotine (4.5 mg/kg/day) administered subcutaneously for 14 days by osmotic minipumps, prior to induction of ischemic insults in rats. Nicotine decreased the blood flow in the penumbra region during reperfusion after transient MCAO, worsened neurological deficit, and increased the injury and edema volume. Interestingly, nicotine-treated rats exhibited depletion of tissue plasminogen activator in cerebral capillaries, which might be causally relevant to the poor recovery of the blood flow after reperfusion and to the resultant exacerbation of brain tissue injury. In a more recent study, the effect of 4-week continuous treatment with nicotine (2 or 4 mg/kg/day, subcutaneously by osmotic pumps) prior to 2-h transient MCAO was examined (Li et al. 2016). Nicotine at both doses significantly increased the infarct size,
although the neurological deficit seems to be ameliorated by the treatments. Authors attributed the exacerbation of pathological changes by nicotine to enhanced oxidative stress, as the cerebral cortex and the cerebral arteries in nicotine-treated rats exhibited decreased expression levels of Mn superoxide dismutase and mitochondrial uncoupling protein-2, as compared to those in control rats.

Bradford et al. (2011) examined the effect of chronic nicotine treatment with special reference to inflammatory responses. In their study, 14-day pretreatment with nicotine (0.5–2 mg/kg/day, subcutaneously by osmotic pumps) dose-dependently increased the infarct volume and brain water content at 3 days after 30-min MCAO. In addition, nicotine given at 2 mg/kg/day worsened neurological outcome and decreased the survival rate of mice. They also demonstrated that 14-day treatment with nicotine (2 mg/kg/day), either alone or in combination with ischemia/reperfusion insults, markedly increased the expression of various cytokines (including TNF-α and IL-1β) and chemokines (including CCL2 and CXCL5) in brain tissue and isolated brain microvessels. Consistent with these observations, ischemia/reperfusion-induced infiltration of neutrophils and monocytes was augmented by 14-day pretreatment with nicotine.

In a study on permanent MCAO in mice, subcutaneous delivery of 4.5 mg/kg/day for 3 weeks has been shown to exacerbate brain edema (Paulson et al. 2010). Exacerbation of brain edema was also observed when nicotine was intraperitoneally administered 1 h before MCAO. Although the precise mechanisms of action of nicotine in these experimental settings have not been addressed, the effect might be relevant to nicotine-induced altered functions of Na⁺, K⁺, 2Cl⁻ cotransporters in the blood-brain barrier (Paulson et al. 2006).

| Table 7.3  | Studies reporting deleterious effects of nicotine in experimental models of ischemia |
|------------|----------------------------------------------------------------------------------|
| Conditions of drug treatment | References |
| Nicotine (4.5 mg/kg/day, s.c.) continuous infusion for 14 days prior to transient ischemia in rats | Wang et al. (1997) |
| Nicotine (2 and 4 mg/kg/day, s.c.) continuous infusion for 4 weeks prior to transient ischemia in rats | Li et al. (2016) |
| Nicotine (2 mg/kg/day, s.c.) continuous infusion for 14 days prior to transient ischemia in rats | Bradford et al. (2011) |
| Nicotine (187.5, 562.5 or 1125 μg/kg, i.p.) 1, 3 or 6 h, respectively, before permanent ischemia in mice | Paulson et al. (2010) |
| Nicotine (4.5 mg/kg/day, s.c.) continuous infusion for 1 or 3 weeks prior to permanent ischemia in mice | Paulson et al. (2010) |
| Nicotine (1 mg/kg, i.p.) daily injection for 15 days prior to transient ischemia in female rats | Raval et al. (2009) |
| Nicotine (4.5 mg/kg/day, s.c.) continuous infusion for 16 days prior to transient ischemia in female rats | Raval et al. (2011) |
| Nicotine (4 μg/kg/min, s.c. To dams) continuous infusion from day 4 of gestation to day 10 after birth in rats | Li et al. (2012) |

See Sect. 7.3.2.2 in the main text for details of the findings in each study

i.p. intraperitoneal, s.c. subcutaneous
Nicotine may produce harmful influences on ischemia-related pathological events. Long-term pretreatment with nicotine by continuous infusion is generally shown to aggravate the pathology of ischemic stroke in experimental animals. Multiple mechanisms are postulated to play important roles in these harmful consequences as summarized here (See Sect. 7.3.2.2 for details. SOD superoxide dismutase, t-PA tissue plasminogen activator, UCP-2 uncoupling protein-2)

Potential interaction of the effect of nicotine with that of estrogen has been proposed by studies using ovariectomized rats. Daily intraperitoneal injection of 1 mg/kg nicotine for 15 days to female rats with normal menstrual cycle accelerated neuron loss in the hippocampal CA1 region at 7 days after 10-min ligation of carotid artery. In addition, ovariectomy per se exacerbated ischemia-induced loss of CA1 neurons and occluded the effect of repeated nicotine treatment (Ravel et al. 2009). These results suggest that the neuroprotective effect of endogenous estrogen is cancelled by nicotine. Indeed, oral contraceptives that lowered the plasma level of 17β-estradiol exacerbated ischemia-induced loss of hippocampal CA1 neurons, synergistically with 16-day treatment of nicotine. Abolishment by nicotine of the neuroprotective effect of 17β-estradiol on oxygen/glucose deprivation-induced injury, as well as attenuation by nicotine of estrogen receptor-mediated recruitment of cyclic AMP response element binding protein, was also demonstrated in rat hippocampal slice cultures (Ravel et al. 2011).

Finally, neonatal male rats continuously exposed to nicotine during the perinatal period (from day 4 of gestation to day 10 after birth) exhibited larger infarct size after hypoxic-ischemic injury than control rats. The augmented pathology may be attributable to altered angiotensin II signaling resulting from decreased expression of angiotensin II AT2 receptors (Li et al. 2012).

Taken together, long-term continuous exposure to nicotine may aggravate the consequences of ischemic stroke, by altering expression and function of various cytoprotective proteins, pro-inflammatory factors, transporters and receptors in the brain (Fig. 7.3).
7.4 Hemorrhagic Stroke and nAChRs

7.4.1 Effects of nAChR Agonists on Intracerebral Hemorrhage (ICH)

ICH is initiated by the rupture of blood vessels within the brain parenchyma and bleeding from ruptured vessels. The mechanisms involved in brain tissue injury after ICH include several distinct features from those involved in ischemic brain injury. A notable feature of ICH is that blood constituents may exert specific biological actions on neurons and glial cells in the brain and make substantial contribution to the complicated pathogenic events. Thrombin, a serine protease that normally functions in the coagulation cascade, is a remarkable example because it can stimulate several members of protease-activated receptors (PARs) that are expressed in various cell types including neurons, astrocytes and microglia. Indeed, several lines of evidence indicate that thrombin may contribute to the pathogenesis of experimental ICH models. For example, a thrombin inhibitor argatroban reduces edema and inflammation associated with ICH in rats (Kitaoka et al. 2002; Nagatsuna et al. 2005).

To facilitate investigations on the pathogenic mechanisms of ICH and exploration of therapeutic drug candidates, we developed an in vitro neurodegeneration model of ICH, using organotypic brain slice cultures (Fujimoto et al. 2006). Coronal slices containing the cerebral cortex and the striatum were prepared from neonatal rats, and after cultivation for 9–11 days, they were treated with thrombin. Thrombin induced delayed neuronal cell death in the cerebral cortex and tissue shrinkage accompanied by cell death in the striatum. Results of various pharmacological interventions revealed the mechanisms involved in cell and tissue injuries in the cerebral cortex and the striatum. With regard to the tissue injury in the striatum, stimulation of PAR-1 and recruitment of mitogen-activated kinase (MAPK) families plays a critical role. In addition, thrombin-induced striatal tissue injury was abrogated by deletion of tissue microglia prior to thrombin treatment. Importantly, potential involvement of microglial MAPK activation in tissue injury was confirmed in a rat model of ICH induced in the striatum by collagenase injection (Ohnishi et al. 2007), which substantiates the validity of slice culture experiments as a model of ICH pathogenesis.

Then we examined the effect of nicotine on thrombin neurotoxicity in corticostriatal slice cultures (Ohnishi et al. 2009). In this set of experiments, cortico-striatal slices were prepared and cultured for 11 days in serum-containing medium, maintained for 1 day in serum-free medium, and then treated with 100 U/ml thrombin in serum-free medium for 3 days. When applied to slice cultures for the entire culture period of 15 days, nicotine (3–30 μM) attenuated thrombin-induced shrinkage of striatal tissue and prevented thrombin-induced neuron loss in the cerebral cortex. Thrombin-induced increase in the number of microglia in the cortex was also attenuated by nicotine in a concentration-dependent manner. The cytoprotective effect and anti-inflammatory effect of nicotine were largely abrogated by mecamylamine.
(non-selective nAChR antagonist), methyllycaconitine (α7 nAChR antagonist) and dihydro-β-erythroidine (α4β2 nAChR antagonist), suggesting the involvement of both of the major central nAChR subtypes.

With these lines of evidence in vitro, we moved onto investigations in vivo to address the effect of nicotine in a mouse model of ICH prepared by injection of collagenase (that disrupts vascular basement membrane) into the striatum. Based on the findings in slice culture preparations, we initially examined the effect of long-term pretreatment of nicotine (1 mg/kg, intraperitoneally once per day for 1 week) and obtained cytoprotective effect of nicotine on striatal neurons within the hematoma assessed at 3 days after induction of ICH (our unpublished observations). However, the long-term pretreatment regimen is unrealistic for ICH pharmacotherapy in clinical practice, so we next examined the effect of nicotine in a post-treatment regimen. ICH was induced by collagenase injection into the striatum, and nicotine tartrate dehydrate (1 and 2 mg/kg as nicotine free base) was first administered intraperitoneally 3 h after ICH induction. Nicotine treatment was repeated three times in total, at 24 h intervals. Under these conditions, nicotine (2 mg/kg) partially but significantly attenuated the decrease in the number of striatal neurons in the central region of hematoma at 3 days after ICH. ICH-induced increase in TUNEL-positive cells that may represent apoptotic cell death was also inhibited by nicotine treatment. Interestingly, nicotine decreased expression of a pro-apoptotic protein Bax in the striatum and consequently increased the relative expression level of Bcl-2, an anti-apoptotic protein. Therefore, counteraction of the apoptotic cell death events may underlie the neuroprotective effect of nicotine in ICH (Fig. 7.4). In the perihematoma region, the pathological features were accumulation of activated microglia/macrophages and increase in oxidative stress (as revealed by nitrotyrosine immunoreactivity), both of which were significantly attenuated by nicotine treatment. Moreover, nicotine at 2 mg/kg alleviated neurological deficits observed after ICH (Hijioka et al. 2011). When the effect of α7 nAChR agonist PNU-282987 (3 and 10 mg/kg) was tested in the same treatment regimen as nicotine, the drug showed neuroprotective and anti-inflammatory effects, whereas α4β2 nAChR agonist RJR-2403 showed no significant therapeutic effect (Hijioka et al. 2012). These results suggest that α7 nAChR is the major nAChR subtype responsible for the therapeutic effect of nicotine in ICH.

Consistent with our findings, Krafft et al. (2012) reported the therapeutic effect of PNU-282987 (12 mg/kg) and another α7 nAChR agonist PHA-543613 (4 or 12 mg/kg) in a mouse model of ICH based on autologous blood infusion into the striatum. In their study, agonists were administered intraperitoneally at 1 h after ICH surgery. These drug treatments improved behavioral outcome and prevented brain edema formation at 24 h after surgery. The effect of PHA-543613 was counteracted by a phosphoinositide 3-kinase (PI3K) inhibitor wortmannin. In addition, PHA-543613 increased the level of phosphorylated Akt and prevented ICH-induced activation of glycogen synthase kinase (GSK) -3β and caspase-3, which was also cancelled by wortmannin. These results indicate that the therapeutic effect of α7 nAChR stimulation is mediated by PI3K/Akt pathway leading to inactivation of
Fig. 7.4 nAChRs expressed in distinct types of cells may contribute to the therapeutic effects of nAChR agonists against brain tissue damage associated with intracerebral hemorrhage (ICH). Stimulation of nAChRs (probably α7 nAChRs) by nicotine in neurons results in decreased expression of a pro-apoptotic protein Bax, thereby inhibits ICH-induced apoptotic neuronal cell death (Hijioka et al. 2011, 2012). On the other hand, specific α7 nAChR agonists such as PNU-282987 and PHA-543613 may stimulate α7 nAChRs in endothelial cells, and recruit Jak2/PI3K/Akt pathway as in the case with Fig. 7.2. Activated Akt phosphorylates and inactivates glycogen synthase kinase-β (GSK-3β) and thereby prevents phosphorylation and degradation of β-catenin, a component of tight junctions. Stabilization of tight junctions may be relevant to the maintenance of integrity of the blood-brain barrier after hemorrhagic insults (Krafft et al. 2013).

GSK-3β and prevention of apoptotic cell death events. A follow-up study by the same group (Krafft et al. 2013) showed that PHA-543613-induced inactivation of GSK-3β led to stabilization of β-catenin and tight junction proteins such as claudin-3 and claudin-5, which may be relevant to the preservation of blood-brain barrier integrity and to the inhibition of edema formation by the drug (Fig. 7.4). A more recent study addressed the role of Jak – signal transducer and activator of transcription (STAT) signaling pathway in the effect of α7 nAChR stimulation (Krafft et al. 2017). That is, α7 nAChR agonist PHA-543613 augmented phosphorylation/activation of Jak2 and STAT3 in a mouse model of ICH by autologous blood infusion, and Jak2 inhibitor AG490 reversed the brain tissue-preserving effect of PHA-543613 in a rat model of ICH based on collagenase injection.

The main focus of the brain region in preclinical ICH studies has been the striatum (the caudate putamen), because the putamen is one of the most susceptible brain regions with regard to hypertension-associated ICH in humans, and also because this region is relatively easy of access by surgical operations in experimental animals. On the other hand, the relative incidence of non-hypertension-associated ICH that occurs in the cerebral cortex (cortical or lobar hemorrhage) has been
increasing in recent years, but therapeutic approaches for cortical hemorrhage have been poorly explored. We addressed whether nicotine can provide therapeutic effect in cortical hemorrhage in mice (Anan et al. 2017). Collagenase injection into the parietal cortex induced hemorrhage, which resulted in motor deficits in mice. Daily intraperitoneal administration of nicotine (1 and 2 mg/kg), with the first injection given at 3 h after ICH induction, significantly improved motor performance. Histochemical examinations on brain sections obtained 3 days after ICH demonstrated that nicotine (2 mg/kg) significantly increased the number of surviving neurons in the hematoma and decreased the number of activated microglia/macrophages in the peri-hematoma region. Notably, the neuroprotective effect of nicotine on cortical hemorrhage was blocked by antagonists at $\alpha_7$ nAChRs (methyllycaconitine) and $\alpha_4\beta_2$ nAChRs (dihydro-β-erythroidine). Therefore, the neuroprotective effect of nicotine in cortical hemorrhage may be mediated by both of these two receptor subtypes, unlike in the case with putaminal hemorrhage where $\alpha_7$ nAChR plays a dominant role.

7.4.2 Effects of nAChR Agonists on Subarachnoid Hemorrhage (SAH)

SAH is one of the major kinds of hemorrhagic stroke distinct from ICH. SAH is characterized by extravasation into the subarachnoid space, which mainly results from the rupture of intracranial aneurysm. At present, there is only one experimental study addressing the effect of nAChR agonist on the outcome of SAH. Duris et al. (2011) examined the effect of $\alpha_7$ nAChR agonist PNU-282987 on an endovascular perforation model of SAH in rats, by administering the drug intraperitoneally at 1 h after perforation surgery. They showed that PNU-282987 at 12 mg/kg ameliorated neurological function at 24 and 72 h after SAH induction, and prevented the increase in brain water content at 24 h. As in the case with ICH (Krafft et al. 2012), $\alpha_7$ nAChR stimulation enhanced phosphorylation/activation of Akt and prevented SAH-induced activation of caspase-3, which was abrogated by PI3K inhibitor wortmannin. Therefore, $\alpha_7$ nAChRs may serve as a promising drug target also for SAH therapy, although detailed pharmacological profiles of neuroprotection should be revealed by further investigations.

7.5 Nicotine, Smoking and Stroke: Potential Associations

As nicotine is the major pharmacoactive compound contained in tobacco that can directly modify the functions of the nervous system and the cardiovascular system, potential relationship between smoking and cerebrovascular disorders suggested by various epidemiological investigations may deserve considerations. It should be
noted, however, that tobacco smoke contains many (over 4000) kinds of chemicals and the adverse consequences of smoking are not necessarily and solely attributable to the biological actions of nicotine.

With regard to ICH, current smoking is among the modifiable risk factors (An et al. 2017; Ariesen et al. 2003), although smoking (both current use and history of use) may be more strongly associated with ischemic lacunar stroke than with ICH (Kaplan et al. 2014). Race/population-specific influences of smoking on ICH incidence have also been reported. A study by Faigle et al. (2016) showed that smoking was one of the predictors of mortality in white populations, but not black populations, of ICH patients. In addition, meta-analyses of the risk factors for ICH and ischemic stroke suggest that smoking is a risk factor more frequent in ischemic stroke in white population but not in Chinese population (Tsai et al. 2016). In any cases, whether or not nicotine is responsible for the increased risk of ICH associated with smoking is unknown.

Several reports addressed the association of smoking and nicotine exposure with the risk and the outcome of SAH. In a case-control study on population of 18–49 years of age, exposure to nicotine in pharmaceuticals as well as current smoking has been identified as a risk factor of SAH (Broderick et al. 2003). In addition, a retrospective analysis of the size of ruptured intracranial aneurysm of SAH patients demonstrated that the average maximal aneurysm diameter in patients with combined history of hypertension and smoking was significantly smaller than in patients with hypertension only. The latter results suggest that smoking lowers the threshold of aneurysm rupture in combination with hypertension. On the other hand, a systematic review on four independent studies concerning nicotine replacement therapy for tobacco smoking in SAH patients has concluded that the therapy in the first 21 days does not appear to be associated with increased risk of poor functional outcome, death and vasospasm (Turgeon et al. 2017). Actually, there are reports showing better functional outcome (Carandang et al. 2011) and lower risk of death (Seder et al. 2011) in patients with nicotine replacement therapy than those without the therapy. These results imply the beneficial effects of nicotine against SAH pathology in humans, although more detailed and well-designed studies with larger sample size are clearly required for drawing definitive conclusion.

Current smoking has long been known as a strong risk factor for the incidence of ischemic stroke (Hawkins et al. 2002). Increased number of cigarettes smoked per day has been associated with increased risk of ischemic stroke in a study on 15–49 year-old women (Bhat et al. 2008). With regard to the disease outcome, however, “smoker’s paradox” has been reported in ischemic stroke as in the case with myocardial infarction (Ovbiagele and Saver, 2005). For example, a study on 4305 cases of ischemic stroke patients, smoking was associated with lower all-cause inhospital mortality (Ali et al. 2013). In another study on ischemic stroke patients that received thrombolytic therapy by intravenous tissue plasminogen activator, smokers had better functional outcome in addition to better recanalization and reperfusion, as compared to non-smokers (Kufner et al. 2013). A recent investigation on the potential causes of smoker’s paradox showed that smokers experienced their first ever stroke 11 years younger than non-smokers, which might explain in part the
association between smoking and better functional outcome (Hussein et al. 2017). Notably, subgroup analysis in the same study detected favorable outcome of current smokers in women at or older than 65 years (Hussein et al. 2017).

Overall, although smoking may be considered as an important risk factor for the occurrence of stroke episodes, contribution of nicotine to the increased risk is unclear. On the other hand, several lines of evidence indicate that smoking or nicotine treatment (as the replacement therapy) might result in favorable outcome in several particular cases of stroke.

### 7.6 Conclusion and Future Perspectives

The functional roles of nAChRs in the central nervous system beyond neurotransmission have been increasingly recognized. Neuroprotective and anti-inflammatory effects of nicotine and other nAChR agonists are of particular interest for the development of novel pharmacotherapies against various kinds of disorders associated with neurodegeneration. This chapter summarized preclinical and clinical findings suggesting the relationship between nicotine/nAChRs and principal types of cerebrovascular disorders such as ischemic stroke, ICH and SAH.

Several lines of experimental approaches using in vitro culture system and in vivo disease models indicated the neuroprotective and anti-inflammatory roles of endogenous cholinergic system against pathogenic events in ischemic stroke. These include exacerbation of pathology by nAChR antagonists, as well as amelioration of pathology by AChE inhibitors and positive allosteric modulators of nAChRs. In contrast, studies addressing the effects of nAChR agonists on experimental ischemic injury models gave conflicting results, where some studies reported beneficial effects but others demonstrated deleterious effects. A possible cause of these discrepancies is the different procedures employed for drug administration. Namely, virtually all studies reporting the deleterious effects of nicotine performed continuous delivery by osmotic pumps or repeated daily administration of nicotine at relatively high doses. On the other hand, studies reporting beneficial effects of nicotine and nAChR agonists generally performed daily administration of the drugs for a few times or low-dose administration in the case of long-term treatment. The complex influences of nicotine and nAChR agonists on disease consequences of experimental ischemia models might be relevant to the complicated influences of smoking on the risk and the outcome of ischemic stroke patients that include “smoker’s paradox”.

The number of investigations on hemorrhagic stroke is much smaller than that on ischemic stroke, but distinct lines of experimental evidence are consistent with the idea that nAChRs may serve as a target for ICH therapy. Post-treatment of animal models of striatal (putaminal) hemorrhage with nicotine or α7 nAChR agonists ameliorates neurological outcome and alleviates various neuropathological changes. Involvement of α4β2 nAChRs in addition to α7 nAChRs in neuroprotection is suggested in the case of cortical (lobar) hemorrhage model. Although smoking has been
reported as a risk factor for mortality in particular populations of ICH patients, its relationship with nicotine is not evident. Accordingly, nicotine and subtype-specific nAChR agonists may be considered as novel therapeutic drugs for ICH. This proposal is particularly important because no effective pharmacotherapies for ICH directly aiming at brain tissue preservation have been established to date.

There is only one experimental study addressing the effect of nAChR agonist in SAH model, but according to the results, α7 nAChRs may serve as a promising target for therapy also for SAH. Interestingly, nicotine replacement therapy may be safely applicable to, and might produce beneficial outcome in, smokers after SAH.

An important concern at present is the shortage of information regarding the cellular and molecular mechanisms of actions of nicotine and nAChR agonists under stroke conditions, although a few studies have proposed potential involvement of several signaling pathways such as PI3K/Akt and Jak/STAT. Elucidation of the detailed mechanisms of neuroprotective and anti-inflammatory actions, in conjunction with determination of the ideal drug treatment regimens, may pave the way for developing novel strategies for treatment of distinct types of cerebrovascular disorders, by targeting nAChRs and nAChR-mediated neurotransmission.

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