Low-dose 2-Deoxy Glucose Stabilises Tolerogenic Dendritic Cells and Generates Potent in vivo Immunosuppressive Effects

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Supplementary Information
Supp. Fig. 1 Gating strategy for GM-CSF BMDC cultured cells. BMDC were depleted of Lin⁺ cells (CD4, CD8, CD45R/B220 and MHC II) and cultured in GM-CSF (10 ng/ml) for 6 days with medium exchange every two days. Loosely adherent cell clusters were harvested, depleted of Gr-1⁺ cells and resuspended in complete medium (cRPMI). A sample of 1x10⁶ cells was then prepared for flow cytometry (see Methods) and analysed on an LSR II cytometer (1x10⁵ events recorded). The gating strategy was as follows: (a) discrimination of leukocytes by forward scatter vs. side scatter; (b) single cells were then selected using a forward scatter height (FSC-H) vs. forward scatter area (FSC-A) pulse geometry gate to discriminate between clumps and single cells; (c) live/dead discrimination was achieved using eFluor 455UV fixable viability dye, which stains dead cells, hence the gated negative population corresponds to live cells; (d) population of interest (CD11b⁺CD11c⁺) was identified using CD11b vs. CD11c staining within the live-single cell-leukocyte gate; and (e) bar chart of the frequency of CD11b⁺CD11c⁺ (black bar) and CD11b⁻/negCD11c⁻neg (white bar) within the live population.
Supp. Fig. 2 Tolerogenic DC (tolDC) and 2-DG-treated DC (2-DGtolDC) express low levels of CD86; 2-DGtolDC fail to upregulate CD86 when stimulated with Mtb.

BMDC were prepared as previously described. Flow cytometry was performed to assess CD86 expression on CD11b<sup>hi</sup>CD11c<sup>hi</sup> cells (for gating strategy see Supp. Fig. 1): (a) percent CD86+ cells within the CD11b<sup>hi</sup>CD11c<sup>hi</sup> BMDC population; (b) upper panel: representative histograms showing CD86 expression in CD11b<sup>hi</sup>CD11c<sup>hi</sup> cells treated with or without 2.5 mM 2-DG, and/or stimulated with Mtb; and (c) BMtolDC were cultured as described and treated with 2-DG (2.5 mM) (top panels), and/or stimulated with 15 µg/ml heat inactivated mycobacterial extract, Mtb (bottom panels): representative flow cytometry dot plots of the percentage of CD86+ cells in the CD11b<sup>hi</sup>CD11c<sup>hi</sup> population; error bars denote Standard Error of the Mean (SEM), n=3, p value: *<0.05
Supp. Fig. 3 Expression of progenitor markers CX3CR1 and c-kit by GM-CSF-cultured bone marrow cells. BMDC were prepared as described in Methods and flow cytometry performed to differentiate CD11b<sup>hi</sup>CD11c<sup>hi</sup> cells from CD11b<sup>lo/neg</sup>CD11c<sup>neg</sup> cells (see legend to Supp. Fig 1): (a) CD11b<sup>hi</sup>CD11c<sup>hi</sup> (presumed DC) expressed moderate levels of CX3CR1 (~40 %) while CD11b<sup>lo/neg</sup>CD11c<sup>neg</sup> express high levels of CX3CR1 (~80 %). (b) CD11b<sup>hi</sup>CD11c<sup>hi</sup> and CD11b<sup>lo/neg</sup>CD11c<sup>neg</sup> CX3CR1 MFI. (c) 2-DG treatment slightly reduced the overall percentage of CD11b<sup>hi</sup>CD11c<sup>hi</sup> in the BMDC population while increasing the percentage of CX3CR1<sup>+</sup> cells as shown in representative flow cytometry plots with heat map analysis of CX3CR1 and c-kit expression on non-2-DG-treated DC and 2-DGtolDC. There was minimal c-kit expression with or without 2-DG treatment. Warm colours denote higher relative expression, while cool colours denote lower expression levels.
BMDC cultured with 2.5 mM 2-DG in the presence of glucose do not undergo apoptosis. BMDC were prepared as described in Methods and on d6 were re-plated in glucose-rich (11 mM) cRPMI media with 2-DG at various concentrations (1, 2.5 or 10 mM 2-DG). Cells were harvested and assessed for apoptosis by flow cytometry using Annexin V staining (see Methods): (a) early apoptosis (Annexin V+ 7AAD- cells) in 2-DG-treated cells, either stimulated with Mtb extract (15 µg/ml) (white bars), or unstimulated (black bars). In glucose rich media there was no evidence of 2-DG-induced cell death up to a concentration of 2.5 mM. At 10 mM, apoptosis levels increased significantly (p < 0.05); Mtb stimulation had no additional effect; (b) early apoptosis in glucose-free medium; significant levels of cell death were observed particularly when challenged with Mtb antigen; (c) and (d) levels of late apoptosis in BMDC (Annexin V+ 7AAD+ cells) were unchanged in the presence of 2-DG both in glucose-rich (11 mM) and glucose-free media with and without Mtb stimulation. Error bars denote Standard Error of the Mean (SEM); n=3, p values: *<0.05, **<0.01, ***<0.001
Supp. Fig. 5 Summary of findings (refer to next page for caption).

Steady-state aerobic and anaerobic glucose processing.

Substrate competition (Glu-1, hexokinase) lowers overall metabolic rate.

Predominantly anaerobic metabolism with low glucose availability.

Predominantly oxidative metabolism with high glucose availability (glycogen).
Supp. Fig. 5 Summary of findings.

Blue panel: Untreated CD11b⁺ CD11c⁺ Zbtb46⁺ MHCI⁻ tolerogenic DC (tolDC) undergo baseline aerobic and anaerobic glucose metabolism at a steady-state (“resting”) rate. In the presence of glucose Glut-1 allows for glucose uptake as is required to meet metabolic demand. SIRP-1α is constitutively activated, i.e. phosphorylated. Untreated tolerogenic DC are unable to halt spontaneous experimental autoimmune uveitis (EAU) in an induced animal model.

Red panel: 2-deoxy glucose (2-DG) and glucose compete for uptake through Glut-1 and for intracellular phosphorylation by hexokinase to allow for downstream aerobic/anaerobic substrate processing. This results in an overall reduced metabolic rate. 2-DG stabilises DC in a tolerogenic state, enabling them to prevent EAU progression in vivo.

Grey panel: Extracellular noxae such as mycobacterium toxin (Mtb) or lipopolysaccharide (LPS) in the presence of glucose induce a metabolic stress response, pushing glucose metabolism towards fermentation (i.e. lactate production along with high glucose flux resulting in low substrate availability). This leads to DC activation/maturation (mDC) with increased MHC II surface expression and loss of SIRP-1α phosphorylation. Signalling through NFκB p65 with downstream pro-inflammatory cytokine production aggravates the pro-inflammatory state.

Yellow panel: In the presence of both 2-DG and glucose, extracellular Mtb or LPS are unable to activate tolDC. 2-DG exerts a stabilising effect on tolDC, reflected by increased SIRP-1α phosphorylation and signalling through NFκB p100, resulting in an anti-inflammatory state with decreased IL-12 and increased IL-10 expression. These stable tolDC show a high metabolic rate (predominantly aerobic/OXPHOS) with high glucose availability and ample glycogen storage.

Abbreviations:
Glu, glucose
2-DG, 2-deoxy glucose
Glut-1, glucose transporter 1
G6P/2-DG6P, glucose-6-phosphate/2-deoxy glucose-6-phosphate
TCR, T cell receptor
LDH, lactate dehydrogenase
LPS, bacterial lipopolysaccharide
Mtb, heat inactivated mycobacterial toxin
OXPHOS, oxidative phosphorylation
TCA, tricarboxylic acid cycle (Krebs cycle)
tolDC/mDC, tolerogenic dendritic cells/mature dendritic cells
EAU, induced experimental autoimmune uveoretinitis