Serum Immunoglobulin M Concentration Is Positively Related to Metabolic Syndrome in an Adult Population: Tianjin Chronic Low-Grade Systemic Inflammation and Health (TCLSIH) Cohort Study

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Abstract

Persistent low-grade systemic inflammation has been increasingly recognized as a common pathological process, and an important contributing factor to cardiovascular diseases and its risk factor, metabolic syndrome. Immunoglobulin M is reactive to multiple autoantigens and is inferred to be important for autoimmunity, implying that immunoglobulin M may be a potential risk factor for metabolic syndrome. However, few epidemiological studies are available which are related to this potential link. Therefore, we designed a cross-sectional study of 9,379 subjects to evaluate the relationship between immunoglobulin M and metabolic syndrome in an adult population. Subjects who received health examinations were recruited from the Tianjin Medical University General Hospital-Health Management Center in Tianjin, China. Immunoglobulin M was determined with an immunonephelometric technique. Metabolic syndrome was defined according to the criteria of the American Heart Association scientific statements of 2009. Multiple logistic regression analysis was used to examine the relationships between the quartiles of immunoglobulin M and the prevalence of metabolic syndrome. After adjustment for covariates, the odds ratio of having metabolic syndrome in the fourth quartile compared with the first quartile of immunoglobulin M was 1.19 times for males (95% confidence interval, 1.002–1.41) and 1.39 times for females (95% confidence interval, 1.07–1.80). Immunoglobulin M levels also showed positive relationships with the ratio of elevated triglycerides and reduced high-density lipoprotein cholesterol in males. The study is the first to show that immunoglobulin M is independently related to metabolic syndrome and its individual components (elevated triglycerides and reduced high-density lipoprotein cholesterol) in males, whereas immunoglobulin M is independently related to metabolic syndrome in females but not to its individual components. Further studies are needed to explore the causality and the exact role of immunoglobulin M in metabolic syndrome.

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Introduction

Chronic diseases, such as cardiovascular diseases (CVD), cancer, and age-related diseases have long been considered among the most important global public health issues [1]. CVD are a group of disorders that affect the heart and blood vessels, and remain a major cause of mortality and morbidity worldwide [1]. Metabolic syndrome (MS) is a well-recognized risk factors for CVD, comprised of a constellation of physiological and biochemical abnormalities characterized by disturbances of glucose metabolism, hypertension, dyslipidaemia, and central obesity [2]. Clarifying the common pathological process of MS or CVD is a crucial step toward providing their early prevention and treatment. Persistent chronic low-grade systemic inflammation has been increasingly recognized as a common pathological process and an important contributing factor to MS or CVD [3–6].

Over the past few decades, there has been a steep increase in obesity throughout the world [7,8]. Obesity induces the development of MS [9]. With obesity, many immune cells infiltrate or populate in adipose tissue and promote chronic low-grade inflammation [10]. Furthermore, fat cells, particularly those in the visceral fat, are now considered an immune organ. These cells secrete numerous immune modulating molecules which directly contribute to the development of low-grade inflammation [11,12]. Obesity also influences specific immune responses mediated by the mechanisms of humoral immunity [13,14]. From the above, obesity is, due to innate immunity and/or humoral immune
responses that trigger autoantibody production, the most important risk factor for inducing a systemic inflammatory response.

On the other hand, Immunoglobulin M (IgM) is the first antibody to be produced during an immune response after an initial antigen encounter, and is the predominant isotype secreted in T-cell independent immune responses [15]. IgM has a low affinity for modified self-components [16]. An increased IgM concentration is reactive to a wide variety of autoantigens, and its levels are found markedly elevated in a series of autoimmune diseases [17]. It is therefore believed to be an important component in autoimmunity [17,18]. Because obesity induces the development of autoimmunity [13,14], and is a core factor of MS [19,20], it is hypothesized that IgM may be a crucial molecular link between the obesity-induced systemic inflammatory response and MS. However, few epidemiological studies have evaluated the relationships between IgM and MS among the general population [21]. Therefore, it is still unclear whether a higher level of serum IgM concentration is related to a higher prevalence of MS.

This cross-sectional study aimed to investigate how serum IgM concentration is related to the prevalence of MS in an adult population.

Materials and Methods

Participants

The Tianjin Chronic Low-grade Systemic Inflammation and Health (TCLSIH) Cohort Study is a large prospective dynamic cohort study focusing on the relationships between chronic low-grade systemic inflammation and the health status of a population living in Tianjin, China. Tianjin is a city of approximately 10.43 million inhabitants, located in the northeastern part of the North China Plain, facing the Bohai Sea [22]. Participants were recruited, while having their annual health examinations at the Tianjin Medical University General Hospital-Health Management Center, the largest and most comprehensive physical examination center in Tianjin.

This cross-sectional study used baseline data from the TCLSIH. During the research period there were 10,015 participants who had received health examinations including serum-immunological tests. We excluded participants who did not complete data collection on any components of MS (n = 41), body height and/or body weight measurements (n = 1), or those with a history of CVD (n = 513) or cancer (n = 81). Owing to these exclusions, the final cross-sectional study population comprised 9,379 participants (mean [standard deviation, SD] age: 46.6 [10.9] years, age range: 25–86 years; males, 60.5%). The blood sample was routinely drawn 12 ml of whole blood for 2 ml of plasma and 10 ml of serum from each subject. The protocol of this study was approved by the Institutional Review Board of the Tianjin Medical University and participants gave written informed consent prior to participation in the study.

Serum-immunological tests

Serum-immunological tests were measured as a health examination item. Serum levels of immunoglobulins (IgM, IgA, and IgG) were determined by the immunonephelometric technique using the automated IMMAGE 800 immunochemistry system (Beckman Coulter, Brea, CA, USA), and expressed as mg/dL. The detection limit of the assay was: IgM 4.2 mg/dL, IgA 6.7 mg/dL, IgG 33.3 mg/dL; the measurement range was: IgM, 4.2–14,400 mg/dL, IgA 6.7–25,200 mg/dL, IgG 33.3–21,600 mg/dL and the intra- and inter-assay coefficients of variation (CV) were less than 6% for three classes of immunoglobulins. The manufacturer indicates the following reference intervals for healthy adults: IgM 46–304 mg/dL, IgA 82–453 mg/dL, and IgG 751–1,560 mg/dL.

Assessment of MS and other variables

Waist circumference was measured at the umbilical level with participants standing and breathing normally. Blood pressure (BP) was measured twice from the upper left arm using a TM-2655P automatic device (A&D CO., Tokyo, Japan) after 5 minutes of rest in a seated position. The mean of these 2 measurements was taken as the BP value. Blood samples for the analysis of fasting blood sugar (FBS) and lipids were collected in siliconized vacuum plastic tubes. FBS was measured by the glucose oxidase method, triglycerides (TG) were measured by enzymatic methods, low-density lipoprotein cholesterol (LDL) was measured by the polyvinyl sulfinic acid precipitation method, and high-density lipoprotein cholesterol (HDL) was measured by the chemical precipitation method using reagents from Roche Diagnostics on an automatic biochemistry analyzer (Roche Cobas 8000 modular analyzer, Mannheim, Germany).

MS was defined in accordance with the criteria of the American Heart Association scientific statements of 2009 [23]. Participants were considered to have MS when they presented three or more of the following components: 1) elevated waist circumference for Chinese individuals (≥85 cm in males; ≥80 cm in females), 2) elevated TG (≥1.7 mmol/L), or drug treatment for elevated TG, 3) reduced HDL (<1.0 mmol/L in males; <1.3 mmol/L in females) or drug treatment for reduced HDL, 4) elevated blood pressure (SBP ≥130 mm Hg and/or DBP ≥85 mm Hg) or antihypertensive drug treatment, 5) elevated fasting glucose (≥5.56 mmol/L) or drug treatment for elevated glucose.

Assessment of other variables

Anthropometric parameters (height and body weight) were recorded using a standard protocol. Body mass index (BMI) was calculated as weight/height² (kg/m²). Sociodemographic variables, including gender and age, were also assessed. A detailed personal and family history of physical illness and current medications were noted from “yes” or “no” responses to relevant questions. Information on alcohol and tobacco use were obtained from a questionnaire survey.

Statistical analysis

All statistical analyses were performed using the Statistical Analysis System 9.3 edition for Windows (SAS Institute Inc., Cary, NC, USA). Because the prevalence of MS was significantly higher in males, and their IgM serum concentration was also significantly different (see the results section), males and females were analyzed separately in this study. Descriptive data are presented as the mean (range) for continuous variables, and as percentages for categorical variables. Differences in serum IgM levels and the MS prevalence between genders were examined by t-test or chi-squared test, respectively. Because the distribution of serum IgM levels was non-normal, the natural logarithm was applied to normalize the data before analysis of the t-test, and the descriptive data is presented as the geometric mean (95% confidence interval, CI) for it. For further analysis, the prevalence of MS was used as a dependent variable, and the quartiles of IgM as independent variables. For participant characteristics analysis, the differences among IgM categories were examined using analysis of variance (ANOVA) for continuous variables, and logistic regression analysis for proportional variables. Bonferroni-corrected P values were used for comparisons between IgM quartiles. Multiple logistic regression analysis was used to examine relationships between IgM categories
and the prevalence of MS after adjustment for covariates: age, sex, BMI, smoking status, drinking status, family history of CVD, hypertension, hyperlipidemia, or diabetes, and serum IgA and IgG concentrations. Odds ratio (OR) and a 95% CI were calculated. Moreover, a developed multivariable logistic model was obtained using stepwise variable selection methods applied to the variables according to quartiles of IgM. Mean age, BMI, waist circumferences, SBP, and TC, TG, LDL, SBP, and FBS were significantly lower across quartiles of IgM and MS (A, males; B, females). Furthermore, TC, TG, LDL, SBP, and DBP, and higher TG, IgA, and IgG. A lower proportion had a family history of CVD or diabetes (P for all trends ≤0.05). Otherwise, no significant difference was observed between different quartiles of IgM.

Table 2 presents characteristics of female participants according to quartiles of IgM. Mean age, BMI, waist circumferences, TC, TG, LDL, SBP, DBP, and FBS were significantly lower across IgM quartiles (P for trend <0.001). Mean IgG was significantly higher across IgM quartiles (P for trend <0.0001). The percentage of subjects with a family history of CVD, and who were current smokers were significantly lower in the highest IgM quartile (P for trend <0.01 and 0.001, respectively). No significant differences in HDL, drinking status, family history of hypertension, hyperlipidemia, or diabetes were observed across quartiles of IgM.

Table 3 shows the crude and adjusted relationships between quartiles of IgM and MS and its components in male participants. In the final multivariate models, the adjusted OR (95% CI) for MS across IgM quartiles were 1.00 (reference), 1.09 (0.92, 1.29), 1.07 (0.90, 1.26), and 1.19 (1.002, 1.41) (P for trend = 0.07), respectively. Similar results also were observed in females (Table 4). The figure 1 visualizes the relationship between the quartiles of IgM and MS (A, males; B, females). Furthermore, multivariable models were developed using stepwise variable selection. There are almost no changes for the adjusted ORs and 95% CI both in males and females.

Table 1. Male participant characteristics by quartiles of immunoglobulin M (n = 5,673)a.

| Quartiles of immunoglobulin M (range, mg/dL) | Level 1 (7.2–59.2) | Level 2 (59.3–80.6) | Level 3 (80.7–108.0) | Level 4 (109.0–208.0) | P for trendb |
|---------------------------------------------|--------------------|--------------------|---------------------|----------------------|-------------|
| (n = 1,422)                                 | (n = 1,415)        | (n = 1,406)        | (n = 1,430)         |                      |             |
| Age (y)                                     | 48.5 (27.0, 81.0)d | 45.7 (25.0, 85.0)d | 45.6 (26.0, 86.0)d  | 45.4 (25.0, 85.0)d  | <0.0001     |
| BMI (kg/m2)                                 | 26.3 (17.3, 45.2)  | 26.3 (16.5, 51.1)  | 26.0 (16.6, 44.6)   | 25.8 (15.4, 38.3)d  | <0.01       |
| Waist circumference (cm)                    | 91.5 (67.0, 125.0) | 91.1 (64.0, 133.0) | 90.2 (63.0, 135.0)d | 89.8 (63.0, 124.0)d | <0.001      |
| TC (mmol/L)                                 | 5.24 (2.72, 10.06) | 5.26 (2.97, 9.96)  | 5.26 (2.05, 10.42)  | 5.25 (2.99, 15.45)  | 0.55        |
| TG (mmol/L)                                 | 1.92 (0.41, 19.81) | 1.98 (0.32, 16.04) | 2.06 (0.37, 17.71)  | 2.20 (0.36, 28.35)d | 0.02        |
| LDL (mmol/L)                                | 3.15 (1.09, 7.06)  | 3.17 (0.81, 7.98)  | 3.17 (0.63, 8.00)   | 3.16 (1.19, 10.45)  | 0.51        |
| HDL (mmol/L)                                | 1.30 (0.30, 2.74)  | 1.27 (0.62, 2.75)  | 1.28 (0.51, 2.71)   | 1.26 (0.46, 2.60)d  | 0.08        |
| SBP (mmHg)                                  | 126.6 (90.0, 205.0)| 125.9 (90.0, 205.0)| 125.3 (85.0, 190.0) | 124.2 (80.0, 180.0)d| 0.03        |
| DBP (mmHg)                                  | 82.6 (50.0, 125.0) | 82.3 (50.0, 130.0) | 81.7 (50.0, 125.0)  | 81.2 (50.0, 125.0)d | 0.0504      |
| FBS (mmol/L)                                | 5.45 (3.60, 19.20) | 5.32 (3.60, 15.40) | 5.36 (3.50, 20.20)  | 5.28 (3.50, 19.10)d | 0.07        |
| IgA (mg/dL)                                 | 226.5 (81.6, 657.0)| 235.3 (51.8, 690.0)| 236.1 (41.2, 745.0) | 246.3 (67.7, 718.0)d| <0.01       |
| IgG (mg/dL)                                 | 1090.5 (517.0, 3330.0)| 1132.0 (420.0, 2340.0)d | 1145.0 (447.0, 2020.0)d | 1187.6 (55.4, 2330.0)d| <0.0001     |

aBMI, body mass index; TC, total cholesterol; TG, triglycerides; LDL, low density lipoprotein cholesterol; HDL, high-density lipoprotein-cholesterol; SBP, systolic blood pressure; DBP, diastolic blood pressure; FBS, fasting blood sugar; UA, uric acid; CVD, cardiovascular disease; Ig, immunoglobulin.
bAnalysis of variance or logistic regression analysis.
cMean (range) (all such values).
dSignificantly different from the lowest quartile of immunoglobulin M (Bonferroni correction): P < 0.05.

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Results

In this study, 60.5% of participants were males and 39.5% females, with mean ages (SD) of 46.3 (10.4) and 47.1 (11.6) years, respectively. The overall prevalence of MS was 34.9% (3,277 of 9,379). The prevalence of MS was significantly higher in males than in females (43.0% compared with 22.6%, P<0.0001). In contrast, the levels of IgM were significantly lower in males (geometric mean, [95% CI]: 80.4 [79.4–81.4] compared with 111.0 [109.3–112.7] mg/dL, P<0.0001).

Characteristics of male participants across quartiles of IgM are presented in Table 1. Compared with participants in the lowest quartile of IgM, participants in the upper three quartiles tended to be younger, have lower BMI, waist circumferences, SBP, and DBP, and higher TG, IgA, and IgG. A lower proportion had a family history of CVD or diabetes (P for all trends ≤0.05). Otherwise, no significant difference was observed between different quartiles of IgM.
In the MS components analysis, elevated TG was positively related to IgM quartiles (P for trend = 0.0001) in males, in the final model (Table 3). Although the difference was not statistically significant (P for trend = 0.09), the highest proportion of participants with reduced HDL was in the highest IgM quartile (OR [95% CI]: 1.22 [1.00, 1.49]) (Table 3). In contrast, no significant relations were found between serum immunoglobulin concentrations and other MS components in the final multivariate models in females.

### Discussion

In this cross-sectional study, we have investigated the relationships between levels of IgM concentration and MS in an adult population. This study is the first to show that the highest immunoglobulin M quartile is independently related to the highest prevalence of MS in both males and females. Furthermore, in males IgM levels were positively and independently related to the prevalence of elevated TG and reduced HDL.

No previous studies have indicated that serum IgM levels were positively and significantly related to MS among the general population. A small-scale cross-sectional study has investigated the relationships between serum immunoglobulin concentrations and MS among a population of 460 adults [21]. In contrast to our results, this study found no significant relationship between IgM and MS after adjustment for age, sex, smoking, and drinking status. Although the reason remains unclear, differences in adjustment factors and population size may partly explain the discrepancy. As in our male population, this study also found that IgM levels were related to elevated TG and reduced HDL. We therefore speculate that lipid metabolism disorder may be a key point in the link between serum IgM levels and MS among the general population. More interestingly, it was found that IgM levels were not related to elevated waist circumference, and were negatively related to BMI both in previous studies [21,24] and in our observations, suggesting that alteration in TG and HDL metabolism is more important in the relationships between IgM and MS than pure abdominal adiposity or increased BMI.

Several experimental studies have consistently pointed to serum fatty acid as an important inducing molecule of the innate immune system through activation of toll-like receptors (TLR) [25]. Recent studies have also suggested that fatty acid activates B cell TLR4, and that there is a requirement for B cell TLR stimulation by fatty acid for increased IgM [14, 26]. Because TG provides the source of fatty acid needed throughout body cells, and both are closely related, serum fatty acid concentration may help in the interpretation of our observations of the relationships between IgM and MS, TG, or HDL. However, due to our lack of data on serum fatty acid composition, we cannot adequately determine whether fatty acid is a more accurate molecule for explaining our observations of the relationships between IgM and MS, TG, or HDL.
### Table 3. Adjusted relationships of quartiles of immunoglobulin M to metabolic syndrome (males, n = 5,673).  

| Quartiles of immunoglobulin M (range, mg/dL) | Level 1 (7.2–59.2) | Level 2 (59.3–80.6) | Level 3 (80.7–108.0) | Level 4 (109.0–2080.0) | \( P \) for trend<sup>b</sup> |
|-------------------------------------------|-------------------|-------------------|-------------------|-------------------|---------------------|
| Level 1 (n = 1,422)                        | (n = 1,415)       | (n = 1,406)       | (n = 1,430)       |                   |                     |
| **No. of Metabolic syndrome (presence of any 3 of 5 risk factors)** |                   |                   |                   |                   |                     |
| Crude                                     | 618               | 584               | 605               | 618               |                     |
| Age-adjusted                               | Reference         | 0.97 (0.83, 1.12)<sup>c</sup> | 0.89 (0.76, 1.03) | 0.91 (0.79, 1.06) | 0.16               |
| Age- and BMI-adjusted                      | Reference         | 1.03 (0.89, 1.19) | 0.94 (0.81, 1.10) | 0.98 (0.84, 1.14) | 0.54               |
| Multiple adjusted<sup>d</sup>              | Reference         | 1.05 (0.89, 1.24) | 1.02 (0.87, 1.21) | 1.12 (0.95, 1.33) | 0.21               |
| Multiple adjusted<sup>e</sup>              | Reference         | 1.08 (0.91, 1.27) | 1.05 (0.89, 1.24) | 1.15 (0.98, 1.36) | 0.12               |
| **Metabolic syndrome components**          |                   |                   |                   |                   |                     |
| **No. of elevated waist circumference (≥85 cm in males; ≥80 cm in females)** | 1,104             | 1,030             | 1,037             |                   |                     |
| Crude                                     | Reference         | 0.92 (0.77, 1.10) | 0.71 (0.59, 0.84) | 0.68 (0.57, 0.81) | <0.0001            |
| Age-adjusted                               | Reference         | 1.00 (0.83, 1.20) | 0.77 (0.64, 0.92) | 0.75 (0.63, 0.89) | <0.0001            |
| Age- and BMI-adjusted                      | Reference         | 1.04 (0.80, 1.35) | 0.77 (0.59, 0.99) | 0.86 (0.67, 1.11) | 0.08               |
| Multiple adjusted<sup>d</sup>              | Reference         | 1.07 (0.82, 1.39) | 0.77 (0.60, 1.00) | 0.87 (0.67, 1.13) | 0.08               |
| Multiple adjusted<sup>e</sup>              | Reference         | 1.09 (0.84, 1.42) | 0.79 (0.61, 1.02) | 0.90 (0.69, 1.17) | 0.13               |
| **No. of elevated triglycerides (≥1.7 mmol/L)** | 640               | 676               | 665               | 703               |                     |
| Crude                                     | Reference         | 1.12 (0.96, 1.30) | 1.10 (0.95, 1.27) | 1.18 (1.02, 1.37) | 0.047              |
| Age-adjusted                               | Reference         | 1.10 (0.95, 1.28) | 1.08 (0.93, 1.26) | 1.17 (1.01, 1.35) | 0.07               |
| Age- and BMI-adjusted                      | Reference         | 1.12 (0.96, 1.31) | 1.15 (0.99, 1.35) | 1.29 (1.10, 1.50) | <0.01              |
| Multiple adjusted<sup>d</sup>              | Reference         | 1.14 (0.98, 1.33) | 1.19 (1.02, 1.39) | 1.32 (1.13, 1.54) | <0.001             |
| Multiple adjusted<sup>e</sup>              | Reference         | 1.16 (0.99, 1.36) | 1.22 (1.04, 1.42) | 1.38 (1.17, 1.61) | 0.0001             |
| **No. of reduced HDL (<1.0 mmol/L in males; <238 1.3 mmol/L in females)** | 265               | 245               | 281               |                   |                     |
| Crude                                     | Reference         | 1.15 (0.95, 1.39) | 1.05 (0.86, 1.28) | 1.22 (1.01, 1.47) | 0.10               |
| Age-adjusted                               | Reference         | 1.12 (0.92, 1.36) | 1.03 (0.84, 1.25) | 1.19 (0.98, 1.44) | 0.16               |
| Age- and BMI-adjusted                      | Reference         | 1.13 (0.93, 1.38) | 1.08 (0.88, 1.32) | 1.27 (1.05, 1.55) | 0.03               |
| Multiple adjusted<sup>d</sup>              | Reference         | 1.15 (0.94, 1.40) | 1.10 (0.90, 1.35) | 1.29 (1.06, 1.57) | 0.02               |
| Multiple adjusted<sup>e</sup>              | Reference         | 1.12 (0.92, 1.37) | 1.07 (0.88, 1.31) | 1.22 (1.00, 1.49) | 0.09               |
| **No. of elevated blood pressure (systolic: ≥130806 and/or diastolic: ≥85 mmHg)** | 756               | 718               | 704               |                   |                     |
| Crude                                     | Reference         | 0.88 (0.76, 1.02) | 0.80 (0.69, 0.93) | 0.74 (0.64, 0.86) | <0.0001            |
| Age-adjusted                               | Reference         | 0.98 (0.84, 1.14) | 0.89 (0.77, 1.04) | 0.84 (0.72, 0.97) | <0.01              |
| Age- and BMI-adjusted                      | Reference         | 0.99 (0.84, 1.16) | 0.94 (0.80, 1.10) | 0.90 (0.77, 1.06) | 0.15               |
| Multiple adjusted<sup>d</sup>              | Reference         | 0.99 (0.85, 1.17) | 0.94 (0.80, 1.10) | 0.92 (0.78, 1.07) | 0.21               |
| Multiple adjusted<sup>e</sup>              | Reference         | 0.98 (0.84, 1.16) | 0.93 (0.79, 1.09) | 0.90 (0.76, 1.05) | 0.13               |
| **No. of elevated fasting glucose (≥5.56 mmol/416 L)** | 371               | 369               | 365               |                   |                     |
| Crude                                     | Reference         | 0.86 (0.73, 1.01) | 0.86 (0.73, 1.02) | 0.83 (0.70, 0.98) | 0.04               |
| Age-adjusted                               | Reference         | 0.95 (0.80, 1.12) | 0.95 (0.81, 1.13) | 0.92 (0.78, 1.09) | 0.39               |
| Age- and BMI-adjusted                      | Reference         | 0.96 (0.81, 1.14) | 1.00 (0.84, 1.18) | 0.99 (0.83, 1.17) | 0.98               |
| Multiple adjusted<sup>d</sup>              | Reference         | 0.98 (0.83, 1.17) | 1.03 (0.86, 1.22) | 1.02 (0.86, 1.21) | 0.72               |
| Multiple adjusted<sup>e</sup>              | Reference         | 1.01 (0.85, 1.2)  | 1.06 (0.89, 1.26) | 1.07 (0.9, 1.28)  | 0.36               |

<sup>a</sup>BMI, body mass index; HDL, high-density lipoprotein-cholesterol.

<sup>b</sup>Multiple logistic regression analysis.

<sup>c</sup>Adjusted odds ratios (95% confidence interval) (all such values).

<sup>d</sup>Adjusted for age, BMI, smoking status, drinking status, and family history of cardiovascular disease, hypertension, hyperlipidemia, and diabetes.

<sup>e</sup>Adjusted for age, BMI, smoking status, drinking status, and family history of cardiovascular disease, hypertension, hyperlipidemia, and diabetes, and immunoglobulin A, and G.

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### Table 4. Adjusted relationships of quartiles of immunoglobulin M to metabolic syndrome (females, n = 3,706)

| Quartiles of immunoglobulin M (range, mg/dL) | Level 1 (7.2–80.8) | Level 2 (80.9–111.0) | Level 3 (112.0–152.0) | Level 4 (153.0–2480.0) | P for trend^[b] |
|---------------------------------------------|--------------------|----------------------|-----------------------|------------------------|---------------|
| (n = 927)                                   | (n = 942)          | (n = 900)            | (n = 937)             |                        |               |
| No. of Metabolic syndrome (presence of any 3 of 5 risk factors) |                    |                      |                       |                        |               |
| Crude                                       | Reference          | 0.81 (0.66, 1.00)^[c] | 0.61 (0.49, 0.76)      | 0.63 (0.51, 0.78)      | <0.0001       |
| Age-adjusted                                | Reference          | 1.05 (0.84, 1.32)    | 0.93 (0.73, 1.18)      | 1.05 (0.83, 1.33)      | 0.90          |
| Age- and BMI-adjusted                       | Reference          | 1.16 (0.91, 1.47)    | 1.14 (0.88, 1.48)      | 1.40 (1.08, 1.81)      | 0.02          |
| Multiple adjusted^[d]                       | Reference          | 1.15 (0.90, 1.47)    | 1.14 (0.88, 1.48)      | 1.40 (1.08, 1.81)      | 0.02          |
| Multiple adjusted^[e]                       | Reference          | 1.15 (0.90, 1.47)    | 1.14 (0.88, 1.48)      | 1.39 (1.07, 1.80)      | 0.02          |
| Metabolic syndrome components                |                    |                      |                       |                        |               |
| No. of elevated waist circumference          |                    |                      |                       |                        |               |
| (≥85 cm in males; ≥80 cm in females)        | 538                | 490                  | 368                   | 376                    | -             |
| Crude                                       | Reference          | 0.78 (0.65, 0.94)    | 0.50 (0.42, 0.60)      | 0.49 (0.40, 0.58)      | <0.0001       |
| Age-adjusted                                | Reference          | 0.99 (0.81, 1.21)    | 0.72 (0.59, 0.88)      | 0.75 (0.61, 0.91)      | <0.0001       |
| Age- and BMI-adjusted                       | Reference          | 1.22 (0.95, 1.57)    | 0.88 (0.68, 1.14)      | 1.07 (0.82, 1.39)      | 0.74          |
| Multiple adjusted^[d]                       | Reference          | 1.21 (0.94, 1.57)    | 0.88 (0.68, 1.14)      | 1.06 (0.82, 1.38)      | 0.71          |
| Multiple adjusted^[e]                       | Reference          | 1.22 (0.94, 1.57)    | 0.88 (0.68, 1.14)      | 1.07 (0.82, 1.39)      | 0.74          |
| No. of elevated triglycerides (≥1.7 mmol/L) |                    |                      |                       |                        |               |
| (≥1.3 mmol/L in females)                    | 255                | 209                  | 166                   | 191                    | -             |
| Crude                                       | Reference          | 0.75 (0.61, 0.93)    | 0.60 (0.48, 0.74)      | 0.68 (0.54, 0.84)      | <0.0001       |
| Age-adjusted                                | Reference          | 0.89 (0.71, 1.11)    | 0.79 (0.63, 1.00)      | 0.95 (0.76, 1.19)      | 0.59          |
| Age- and BMI-adjusted                       | Reference          | 0.93 (0.74, 1.16)    | 0.88 (0.70, 1.12)      | 1.11 (0.88, 1.40)      | 0.43          |
| Multiple adjusted^[d]                       | Reference          | 0.94 (0.75, 1.17)    | 0.90 (0.71, 1.14)      | 1.14 (0.90, 1.44)      | 0.30          |
| Multiple adjusted^[e]                       | Reference          | 0.94 (0.75, 1.18)    | 0.90 (0.71, 1.15)      | 1.15 (0.91, 1.46)      | 0.27          |
| No. of reduced HDL (≤1.0 mmol/L in males; ≤1.3 mmol/L in females) | 262                | 250                  | 229                   | 262                    | -             |
| Crude                                       | Reference          | 0.92 (0.75, 1.12)    | 0.87 (0.70, 1.07)      | 0.99 (0.81, 1.21)      | 0.89          |
| Age-adjusted                                | Reference          | 0.93 (0.76, 1.14)    | 0.89 (0.72, 1.10)      | 1.02 (0.83, 1.25)      | 0.84          |
| Age- and BMI-adjusted                       | Reference          | 0.98 (0.79, 1.21)    | 0.99 (0.79, 1.23)      | 1.16 (0.94, 1.44)      | 0.14          |
| Multiple adjusted^[d]                       | Reference          | 0.98 (0.79, 1.21)    | 0.99 (0.80, 1.23)      | 1.17 (0.94, 1.45)      | 0.13          |
| Multiple adjusted^[e]                       | Reference          | 0.97 (0.78, 1.20)    | 0.96 (0.77, 1.20)      | 1.11 (0.90, 1.38)      | 0.30          |
| No. of elevated blood pressure (systolic: ≥130 and/or diastolic: ≥85 mmHg) | 431 | 357 | 289 | 275 | - |
| Crude                                       | Reference          | 0.70 (0.58, 0.84)    | 0.54 (0.45, 0.66)      | 0.48 (0.40, 0.58)      | <0.0001       |
| Age-adjusted                                | Reference          | 0.95 (0.77, 1.18)    | 0.93 (0.75, 1.17)      | 0.89 (0.71, 1.11)      | 0.29          |
| Age- and BMI-adjusted                       | Reference          | 1.01 (0.81, 1.26)    | 1.05 (0.83, 1.31)      | 1.04 (0.83, 1.31)      | 0.69          |
| Multiple adjusted^[d]                       | Reference          | 1.01 (0.81, 1.26)    | 1.03 (0.82, 1.30)      | 1.03 (0.82, 1.30)      | 0.78          |
| Multiple adjusted^[e]                       | Reference          | 1.00 (0.80, 1.25)    | 1.02 (0.81, 1.28)      | 1.00 (0.79, 1.26)      | 0.97          |
| No. of elevated fasting glucose (≥5.56 mmol/L) | 195                | 141                  | 123                   | 122                    | -             |
| Crude                                       | Reference          | 0.66 (0.52, 0.84)    | 0.59 (0.46, 0.76)      | 0.56 (0.44, 0.72)      | <0.0001       |
| Age-adjusted                                | Reference          | 0.81 (0.63, 1.04)    | 0.86 (0.66, 1.11)      | 0.87 (0.67, 1.13)      | 0.40          |
| Age- and BMI-adjusted                       | Reference          | 0.84 (0.65, 1.08)    | 0.94 (0.72, 1.22)      | 0.98 (0.75, 1.27)      | 0.92          |
| Multiple adjusted^[d]                       | Reference          | 0.85 (0.66, 1.09)    | 0.95 (0.73, 1.24)      | 1.00 (0.77, 1.31)      | 0.77          |
| Multiple adjusted^[e]                       | Reference          | 0.85 (0.65, 1.09)    | 0.95 (0.73, 1.24)      | 0.99 (0.75, 1.29)      | 0.87          |

^[a]BMI, body mass index; HDL, high-density lipoprotein-cholesterol.
^[b]Multiple logistic regression analysis.
^[c]Adjusted odds ratios (95% confidence interval) (all such values).
^[d]Adjusted for age, BMI, smoking status, drinking status, and family history of cardiovascular disease, hypertension, hyperlipidemia, and diabetes.
^[e]Adjusted for age, BMI, smoking status, drinking status, and family history of cardiovascular disease, hypertension, hyperlipidemia, and diabetes, and immunoglobulin A, and G.

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observations. Further research is warranted to clarify the causality and precise mechanisms between IgM and lipid metabolism disorder, especially hypertriglyceridemia and/or elevated serum fatty acid composition.

In contrast to our observations, a previous study has shown that elevated levels of IgA, IgE, and IgG, but not IgM, are related to myocardial infarction and cardiac death in males with dyslipidemia [27]. It is noteworthy, however, that the baseline TG levels were similar between cases and controls in this case-control study (see textual Table 1). Because our results showed that IgM was strongly related to elevated TG, the difference in serum TG concentrations among groups might be a cause for the observational discrepancy.

Consistent with previous studies, our study also found that females have higher IgM levels than males [21]. Although the precise mechanism remains unclear, the stimulatory action of estrogens on B lymphocytes could be the cause [28]. It is interesting to note that the prevalence of MS is lower in females (it is also observed in other populations [29]), while IgM levels are higher, suggesting a gender-specific relationship between IgM and MS. Although the analysis of individual components of MS showed that serum IgM levels were mainly related to elevated TG in both males and females, the relationship is not statistically significant in females (females had the largest OR in the highest quartile as compared to other individual components of MS). Because TG levels are significantly higher in males (geometric mean, [95% CI]: 1.69 [1.66–1.71] compared with 1.16 [1.14–1.18] mmol/L, \( P < 0.0001 \)), differences in TG levels may partly explain these observations. Furthermore, despite being positively and significantly related to MS in females, IgM levels were not significantly related to any one of the individual components of MS.
MS. We speculate that in females, IgM may be strongly related to a constellation of risk factors rather than a simple risk factor.

A large number of studies have established that most autoimmune diseases occur significantly more frequently in females than males [30]. Several mechanisms such as X chromosome inactivation, the effects of sex hormones on immune function, fetal-maternal microchimerism, and redox state etc. have been proposed as explanations for this gender bias [30–32]. The present results showed that elevated levels of TG and low levels of HDL correlated with reduced levels of IgM in males, and that TG levels are significantly higher in males than females, implying the effect of TG on IgM may depend on TG levels, but not gender. Further studies are required to identify the hypothesis.

It is increasingly recognized that MS is a low-grade inflammatory condition and central abdominal fat is a contributing factor [33]. The molecular mechanisms that contribute to this inflammation are still quite unclear. Inflammation of central adipose tissue leads to adipokine production, followed by secretion of adipokines into the general circulation contributing to the overall inflammatory condition [34]. An increased accumulation of macrophages occurring in obese adipose tissue has emerged as a key process in metabolic inflammation [6]. The heterogeneity of adipose tissue macrophages and their physical and functional interactions with adipocytes, endothelial cells, and other immune cells within the adipose tissue microenvironment is recognized in playing a critical role in the development and progression of inflammation [6]. Furthermore, up-regulation of TLR4/nuclear factor-kappaB is considered to be major signaling pathways [34,35]. However, few studies have investigated the question of whether IgM has an influence on the pathological process of MS. The present study provides a novel viewpoint that IgM may be involved in the pathological process of MS.

The present study has two limitations. Firstly, because this is a cross-sectional study, further prospective studies and intervention trials should be undertaken to establish a causal relationship between IgM and MS. Secondly, although we adjusted for a considerable number of potentially confounding factors, we cannot exclude the possibility that MS is affected by other lifestyle variables which are intrinsically related to serum IgM concentration.

This large-scale epidemiological study has shown that the highest immunoglobulin M quartile is independently related to the highest prevalence of MS in males. But, IgM only was significantly related to elevated TG and reduced HDL, but not waist circumference, BP and FBS. This suggests that IgM may be involved in the pathological process of MS through lipid metabolism disorder. In females, serum IgM levels were independently related to a higher prevalence of MS, but not its components, suggesting IgM may be strongly related to a constellation of risk factors rather than a simple risk factor. To conclude, this study is the first to show that IgM is independently related to MS and its individual components (elevated triglycerides and reduced high-density lipoprotein cholesterol) in males, whereas IgM is independently related to MS in females but not to its individual components. The present study also implied that IgM might be a useful predictive factor for MS in an adult population. Further studies are needed to explore the causality and exact mechanisms of IgM in MS.

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Author Contributions

Conceived and designed the experiments: KS QZ KN. Performed the experiments: KS HD QZ CW YG HW LL QJ XW HS SS KN. Analyzed the data: KS HD QZ KN. Contributed reagents/materials/analysis tools: KS HD QZ KN. Wrote the paper: KS HD KN. Critical revision of the manuscript for important intellectual content: KS HD QZ KN. KS HD QZ KN. Contributed reagents/materials/analysis tools: KS HD QZ CW YG HW LL QJ XW HS SS KN. Analyzed the data: KS HD QZ KN. Contributed reagents/materials/analysis tools: KS HD QZ CW YG HW LL QJ XW HS SS KN. Contributed reagents/materials/analysis tools: KS HD QZ CW YG HW LL QJ XW HS SS KN.

References

1. Lozano R, Naghavi M, Foreman K, Lim S, Shibuya K, et al. (2012) Global and regional mortality from 235 causes of death for 20 age groups in 1990 and 2010: a systematic analysis for the Global Burden of Disease Study 2010. Lancet 380: 2095–2128.

2. Grundy SM (2008) Metabolic syndrome pandemic. Arterioscler Thromb Vasc Biol 28: 629–636.

3. Pearson TA, Mensah GA, Alexander RW, Anderson JL, Cannon RO 3rd, et al. (2003) Markers of inflammation and cardiovascular disease: application to clinical and public health practice: a statement for healthcare professionals from the Centers for Disease Control and Prevention and the American Heart Association. Circulation 107: 499–511.

4. Jenny NS (2012) Inflammation in aging: cause, effect, or both? Discov Med 13: 451–460.

5. Kamp DW, Shacter E, Weitzman EZ (2011) Chronic inflammation and cancer: the role of the mitochondria. Oncology (Williston Park) 25: 400–410, 413.

6. Grundy SM (2008) Metabolic syndrome pandemic. Arterioscler Thromb Vasc Biol 28: 629–636.

7. Nathan C (2008) Epidemic inflammation: pondering obesity. Mol Med 14: 485–492.

8. Calabro P, Golia E, Maddaloni V, Malvezzi M, Casillo B, et al. (2009) Adipose tissue-mediated inflammation: the missing link between obesity and cardiovascular disease? Inter J Mol Sci 10: 25–34.

9. Nikolopoulou A, Kadoglou NP (2012) Obesity and metabolic syndrome as related to cardiovascular disease. Expert Rev Cardiovasc Ther 10: 933–939.

10. Sun S, Ji Y, Kersten S, Qi L (2012) Mechanisms of inflammatory responses in obese adipose tissue. Annu Rev Nutr 32: 261–286.

11. Marti A, Marcos A, Martinez JA (2001) Obesity and immune function relationships. Obes Rev 2: 131–140.

12. Sell H, Eckel J (2010) Adipose tissue inflammation: novel insight into the role of macrophages and lymphocytes. Curr Opin Clin Nutr Metab Care 13: 366–370.

13. Winer DA, Winer S, Shen L, Waitha PP, Yauhla J, et al. (2011) B cells promote insulin resistance through modulation of T cells and production of pathogenic IgG antibodies. Nat Med 17: 610–617.

14. Arai S, Marhara N, Iwamura Y, Honda S, Nakashima K, et al. (2013) Obesity-associated autoantibody production requires AIM to retain the immunoglobulin M immune complex on follicular dendritic cells. Cell Rep 3: 1187–1198.

15. Ehrenstein MR, Notley CA (2010) The importance of natural IgM: scavenger, protector and regulator. Nat Rev Immunol 10: 775–781.

16. Vollmers HP, Brandlein S (2006) Natural IgM antibodies: the orphaned molecules in immune surveillance. Adv Drug Deliv Rev 58: 755–765.

17. Duarte-Rey C, Bogolan DP, Leung PS, Anaya JM, Gerwins M (2012) IgM predominance in autoimmune disease: genetics and gender. Autoimmun Rev 11: A404–412.

18. Marchalona JS, Schluter SF, Wilson L, Yocum DE, Boyer JT, et al. (1993) Natural human antibodies to synthetic peptide autoantigens: correlations with age and autoimmune disease. Gerontology 39: 63–79.

19. Licata G, Argano C, Di Chiara T, Parrinello G, Scaglione R (2006) Obesity: a main factor of metabolic syndrome? Fattura Med 48: 77–85.

20. Shirai K (2004) Obesity as the core of the metabolic syndrome and the management of coronary heart disease. Curr Med Res Opin 20: 295–304.

21. Gonzalez-Quintela A, Alende R, Gude F, Campos J, Rey J, et al. (2008) Serum levels of immunoglobulins (IgG, IgA, IgM) in a general adult population and their relationship with alcohol consumption, smoking and common metabolic abnormalities. Clin Exp Immunol 141: 52–69.

22. Tianjin Municipal Government (2007) About Tianjin. Available: http://www.tj.gov.cn/english/About_tianjin/Visiting_Tianjin/ Accessed 13 Aug 3.

23. Alberti KG, Eckel RH, Grundy SM, Zimmet PZ, Cleeman JI, et al. (2009) Harmonizing the metabolic syndrome: a joint interim statement of the International Diabetes Federation Task Force on Epidemiology and Prevention, National Heart, Lung, and Blood Institute; American Heart Association; World Heart Federation; International Atherosclerosis Society; and International Association for the Study of Obesity. Circulation 120: 1640–1645.
24. Marzullo P, Minocci A, Giarda P, Marconi C, Tagliaferri A, et al. (2013) Lymphocytes and immunoglobulin patterns across the threshold of severe obesity. Endocrine.

25. Xu H, Barnes GT, Yang Q, Tan G, Yang D, et al. (2003) Chronic inflammation in fat plays a crucial role in the development of obesity-related insulin resistance. J Clin Invest 112: 1821–1830.

26. Shi H, Kokoeva MV, Inouye K, Tzameli I, Yin H, et al. (2006) TLR4 links innate immunity and fatty acid-induced insulin resistance. J Clin Invest 116: 3015–3025.

27. Kovanen PT, Manttari M, Palosuo T, Manninen V, Aho K (1998) Prediction of myocardial infarction in dyslipidemic men by elevated levels of immunoglobulin classes A, E, and G, but not M. Arch Intern Med 158: 1434–1439.

28. Bouman A, Heineman MJ, Faas MM (2005) Sex hormones and the immune response in humans. Hum Reprod Update 11: 411–423.

29. Kolovou GD, Anagnostopoulou KK, Salpea KD, Mikhailidis DP (2007) The prevalence of metabolic syndrome in various populations. Am J Med Sci 333: 362–371.

30. Gleicher N, Barad DH (2007) Gender as risk factor for autoimmune diseases. J Autoimmun 28: 1–6.

31. Ortona E, Margutti P, Matarese P, Franco I, Malorni W (2008) Redox state, cell death and autoimmune diseases: a gender perspective. Autoimmun Rev 7: 579–584.

32. Knudsen GP (2009) Gender bias in autoimmune diseases: X chromosome inactivation in women with multiple sclerosis. J Neurol Sci 286: 43–46.

33. Maury E, Brichard SM (2010) Adipokine dysregulation, adipose tissue inflammation and metabolic syndrome. Mol Cell Endocrinol 314: 1–16.

34. Elks CM, Francis J (2010) Central adiposity, systemic inflammation, and the metabolic syndrome. Curr Hypertens Rep 12: 99–104.

35. Suganami T, Tanimoto-Koyama K, Nishida J, Itoh M, Yuan X, et al. (2007) Role of the Toll-like receptor 4/NF-kappaB pathway in saturated fatty acid-induced inflammatory changes in the interaction between adipocytes and macrophages. Arterioscler Thromb Vasc Biol 27: 84–91.