Unraveling the Hidden Heterogeneities of Breast Cancer Based on Functional miRNA Cluster

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Abstract

It has become increasingly clear that the current taxonomy of clinical phenotypes is mixed with molecular heterogeneity, which potentially affects the treatment effect for involved patients. Defining the hidden molecular-distinct diseases using modern large-scale genomic approaches is therefore useful for refining clinical practice and improving intervention strategies. Given that microRNA expression profiling has provided a powerful way to dissect hidden genetic heterogeneity for complex diseases, the aim of the study was to develop a bioinformatics approach that identifies microRNA features leading to the hidden subtyping of complex clinical phenotypes. The basic strategy of the proposed method was to identify optimal miRNA clusters by iteratively partitioning the sample and feature space using the two-ways super-paramagnetic clustering technique. We evaluated the obtained optimal miRNA cluster by determining the consistency of co-expression and the chromosome location among the within-cluster microRNAs, and concluded that the optimal miRNA cluster could lead to a natural partition of disease samples. We applied the proposed method to a publicly available microarray dataset of breast cancer patients that have notoriously heterogeneous phenotypes. We obtained a feature subset of 13 microRNAs that could classify the 71 breast cancer patients into five subtypes with significantly different five-year overall survival rates (45%, 82.4%, 70.6%, 100% and 60% respectively; p = 0.008). By building a multivariate Cox proportional-hazards prediction model for the feature subset, we identified has-miR-146b as one of the most significant predictors (p = 0.045; hazard ratios = 0.39). The proposed algorithm is a promising computational strategy for dissecting hidden genetic heterogeneity for complex diseases, and will be of value for improving cancer diagnosis and treatment.

Background

The patients with similar prognosis often respond differently to the same treatment. One explanation for this is that disease with similar phenotypes may have different genetic causes, a phenomenon of genetic heterogeneity. Such genetic heterogeneity presents a significant challenge for modern clinical practice and biomedical research on common human diseases. MicroRNAs are a class of small noncoding RNAs that regulate translation of protein coding mRNAs by translational inhibition or cleavage of the mRNA transcripts. In humans, miRNAs are approximately 22-nucleotide-long single-stranded RNAs[1]. They play a key role in the post-transcriptional regulation of up to 30% of protein-coding genes, and have a profound impact on many cellular processes during development and adult life.

MicroRNA/miRNA expression profiles of tumour samples have recently been shown to provide phenotypic signatures for specific types of cancer [2-6], making it potentially useful in tackling the heterogeneity issues for complex human diseases. Recent studies have used DNA microarrays to study breast cancer, and have shown that it was possible to identify subgroups of patients in terms of different survival courses by gene expression profiles [7]. Blenkiron et al. [8] analysed the expression of miRNAs in breast tumour samples using a bead-based flow-cytometric profiling method. To our knowledge, this was the first integrated analysis of miRNA expression, mRNA expression and genomic changes in breast cancer. They showed that miRNAs could act as molecular signature to distinguish the subtypes of breast cancer, which would be unlikely to be discovered by traditional clinical approaches. They further identified distinctive microRNA signatures that correlated with cytogenetic and molecular subtypes of breast cancer. However, most methods that aimed to identify clinically relevant subtypes using microRNAs usually employed unsupervised learning techniques, such as hierarchical clustering, which would be of limited use when the disease heterogeneity results from only a small subset of the miRNAs participating in a particular cellular process. In these cases, the “signal” or relevant miRNAs may be overwhelmed by the “noise” generated by the vast majority of unrelated data.

In this study, we aimed to identify a subset of miRNAs that could dissect breast cancer patients with different survival outcomes. We employed a coupled two-way clustering algorithm.
(CTWC) [9–11] to iteratively partition breast cancer samples and microRNA sets. The partition was done by a super-paramagnetic clustering (SPC) algorithm [12] that can automatically determine the number of partitions, and generate robust stable phenotypic partitions and the significant feature signatures (miRNA subsets). For identifying the optimal miRNA cluster, we developed a functional consistency index to evaluate the functional consistency of the significant miRNA subsets. The index considers not only expression correlation but also chromosome distance between miRNAs within a significant miRNA subset. The index was developed based on the following considerations. With this measurement, the optimal miRNA cluster is expected to not only dissect breast cancer samples efficiently, but also function in a coherent manner. Finally we applied the proposed method to a publicly available breast cancer microRNA expression profiling dataset, and then employed the Kaplan-Meier survival analysis [13] and multivariate Cox proportional-hazards prediction modelling [14] to determine the differential survival outcomes of new subtypes.

Materials and Methods

Dataset Preprocessing

A miRNA expression data (GSE7842) was download from the Gene Expression Omnibus (GEO)[http://www.ncbi.nlm.nih.gov/geo/]. The raw dataset consisted of the expression profiles of 309 miRNAs in 137 samples in which 119 passed quality control (93 primary human breast tumours, 21 cell lines and five normal breast samples). The original breast cancer study used a single sample predictor (SSP) to assign individual samples to one of five breast tumour subtypes: Luminal A (ER+, PR+, HER2–), Luminal B (ER+, PR+, HER2+), Basal-like (ER–, PR–, HER2–), HER2+ (ER–, PR–, HER2+) and Normal breast-like [8]. For heterogeneity analysis, we used the subset of the data that consists of the expression profiles of 133 miRNAs, which express in 71 human breast cancers with specific clinical information (Menopause stage, Size, Grade, Total nodes, NPI (Nottingham Prognostic Index), Survival time, Dead, DFI and Age etc.) for subsequent analysis.

Coupled two-way clustering

The applied CTWC algorithm is a heuristic and iterative method, and is implemented by a stand-alone package Super-Paramagnetic Clustering was used as the underlying clustering tool to partition the whole dataset into subsets of miRNA and samples iteratively until significant partitions (submatrices) are obtained. A detailed description of the SPC and CTWC algorithm can be found in [15]. For a microRNA expression profile matrix M, we denoted the initial sample set as S, and the microRNA set mGi. Clustering microRNA set mGi on the basis of their expression levels over the set of samples S was referred to the process in an operation denoted by mGi(S). Similarly defined, S(mGi) described the process of clustering S using all microRNA of mGi. We employed CTWC for identifying significant miRNA subsets in the breast cancer dataset. Specifically, first, we clustered all samples using all miRNAs to identify stable sample partitions, and clustered all miRNAs using all samples to identify stable miRNA subsets. Then, we clustered the miRNA gained in the previous step using the newly defined sample partitions (including all samples) to find the responsible miRNA subsets of high discriminating power. Finally, we clustered each sample partition again using each miRNA subset with high discriminating power. In the searching process, we explored the cluster depth for both dimensions of samples and miRNAs. The cluster depth selected was based on the empirical judgement on whether the clinical samples could be well separated using the candidate miRNA subset(s).

Evaluation of a miRNA subset using the functional consistency score

We obtained many high-correlation sample partitions and miRNA subsets by CTWC. To evaluate the power of dissecting tumour subtypes, we developed a functional consistency score that evaluates the biological significance of a miRNA subset. The steps for computing a functional consistency score of miRNA subsets mGi(i = 1, 2, · · · ,N) were as follows:

1) Correlation coefficient (CC). For each mGi, we computed the correlation coefficient

\[ mC(mGi) = \text{mean} \sum_{j=1}^{T} CC(mj,mj) \]

where CC(mj,mj) is the Pearson correlation coefficient between microRNA mj and mj in mGi, and T is the number of microRNA in mGi.

2) Computing the chromosome cluster. Based on the chromosome location of each miRNA in human via MiRGen [16], we grouped human miRNAs into different clusters by requiring that in each cluster the maximum distance between any two miRNAs be less than 50K bp, and obtained 51 intergenic or gene-resident spatial clusters in which 38 overlap with the 133 miRNAs in this study. For each pair P(mj,mj), we map the pair P(mj,mj) to 51 chromosome clusters. Then, we computed the number of the pair P(mj,mj) that overlaps the 51 chromosome clusters N(mj,mj). The chromosome consistency score of mGi is then defined as:

\[ CCS(mGi) = \frac{\sum_{j=1}^{T} N(mj,mj)}{\sum_{j=1}^{T} P(mj,mj)} \]

where T is the total number of microRNA in mGi.

3) Functional consistency. For each mGi,

\[ FC(mGi) = mC(mGi) + CCS(mGi), (i = 1, 2, · · · , N) \]

A higher FC(mGi) corresponds to a higher degree of functional consistency among the miRNAs involved in mGi.

Survival analysis

To verify the clinical significance of the identified hidden breast cancer subtypes, we plotted their survival curves by Kaplan–Meier product-limit method, and assessed the differences between the survival curves of breast cancer patients belonging to different subtypes by log-rank test. The multivariate Cox proportional-hazards model was used to predict the overall survival time, and to determine the significance (at significant level p<0.05) of the effects if the miRNAs is included in the identified miRNA subset(s) on the patient’s survival months. Wald Chi-square test was used to determine the significance of each predictor’s hazard toward the survival time.
Predicting the targets of optimized miRNA subset

To further understand the function and dissect the involved pathways in the optimized miRNAs, we predicted the targets of the optimized miRNA subset \( mG_4 \) using mirGen [16] which includes miRanda, PicTar and TargetScanS prediction programs. This is based on Sethupathy et al. study [17], which found that the intersection of the predictions from prediction program could achieve both high sensitivity and specificity. In addition, we also considered the expression of predicted target genes of miRNAs. Those genes with anti-correlated expression of the miRNAs were filtered and mapped to KEGG in order to study the underlying related pathways.

Computational algorithms

The flowchart of the proposed method was graphically depicted by Figure 1. The programming codes for computing a function consistency score are available upon request to the authors. The hierarchical dendrogram resulted from the coupled two-way clustering was plotted by Treeview[18,19].

Results

Selection of miRNA subset that partitions breast cancer samples with highest functional consistency score

We employed CTWC algorithm to search for significant miRNA subsets that could distinguish breast cancer subtypes. CTWC identified many highly correlated miRNA subsets during the recursive partitioning of samples and miRNAs. In this study we aimed to identify the partition of breast cancers [8]. With CTWC, we identified 75 stable and significant miRNA subsets at the cluster depth of one (\( mG_2,mG_3,mG_4,mG_5,mG_6 \)) (Table 1). Then, we determined the function consistency scores of the 75 stable miRNA subsets separately.

We computed and sorted the correlation coefficient of 75 miRNA subsets. The CC (the average Pearson correlation coefficient across all pairs of miRNAs in the cluster, see Methods for details) of the top 5 subsets (\( mG_3,mG_4,mG_5,mG_6,mG_7 \)) are: 0.5632\( \times \)0.769, 0.55772, 0.50603, 0.48114, 0.47562, respectively. \( mG_4 \) has the highest CC. The CC of pairwise miRNAs in \( mG_4 \) is shown in Table S1.

For 5 microRNA subsets with higher CC, we mapped all combinations of microRNAs pairs in each microRNA subset into 51 different chromosome clusters based on mirGen, and then computed a chromosome consistency score for each miRNA subset. We found that score of \( mG_4 \) was 2/78 = 0.0256. The other 4 microRNA subsets have no miRNAs pairs belonging to any chromosome cluster. Based on the correlation coefficient and chromosome cluster of miRNA subsets, we found the has \( mG_4 \) the highest functional consistency score with 0.5908.

Among the 13 miRNAs in \( mG_4 \), miR-221/222 negatively regulates estrogen receptor alpha, and is associated with tamoxifen resistance in breast cancer [20]. The expression of miR-206 is down-regulated in estrogen receptor alpha-positive human breast cancer [21]. Recent data [22] indicated that the pattern of expression of miR-146a and miR-146b was similar, suggesting that their target genes might be coregulated, although they may be located on different chromosomes. Furthermore, the expression of miR-146a/b is high in those samples that have been classified as Basal-like. In the Basal-like cell lines with the highest miR-146a/b expression level, the amount of BRCA1 was particularly low. Further analysis revealed that the expression levels of miR-146b was significantly elevated only compared to Luminal B and Basal-like subtype. In addition, miR-143/145 microRNAs were repressed in Basal-like compared to Luminal subtype [23]. Lu et al [24] demonstrated that the expression level of miR-155 was inversely correlated with estrogen receptor. Our results are consistent with the recent discoveries.

Clustering breast cancer samples using the selected miRNA subsets

We applied \( mG_4 \) using SPC to cluster 71 breast samples from Blenkiron et al. using SPC [8]. Here, the Euclidean distance and Pearson’s correlation coefficient were used as the sample and the miRNA expression similarity measures, respectively. We successfully partitioned the 71 samples into five subtypes (Figure 2).

To verify the clinical significance of the identified hidden breast cancer subtypes, we plotted the survival curves by Kaplan-Meier product-limit method, and assessed the differences between the survival curves of breast cancer patients with different subtypes by a log-rank test (Figure 3). The 5 year survival rates for five subtypes were 45%, 82.4%, 70.6%, 100% and 60% (\( p = 0.008 \)), respectively.

In order to develop a compact model for clinical use, we further identified miRNAs that contributed mostly to the high prediction power. Multivariate Cox proportional-hazards model was used to analyze miRNAs in \( mG_4 \). To reduce the number of variables to be modeled, we applied the stepwise variable selection option (with the same inclusion and exclusion \( p \) value of 0.05) for the multivariate Cox proportional-hazards regression model. We found one predictor (miRNAs) in \( mG_4 \) (Table 2). miR-146b was...
selected to be the significant prognostic predictor because of its importance involved in the underlying pathogenic mechanisms for breast cancer. We also used the Cox model to select significant predictor for ER+ and ER-. In surprise, the miR-146b was also found to be the best predictor ($p = 0.038$).

Predicting the function microRNAs in $mG_4$

We used the Target-Function-Expression Module of miRGen to predict the targets of each microRNA in $mG_4$ and obtained miRNA-mRNA pairs. In addition, The corresponding mRNA expression profiling of 71 samples and 19,061 mRNAs (http://www.ebi.ac.uk/arrayexpress/experiments/E-UCON-1/) was downloaded and 5228 pre-proceeded mRNAs [8] was selected after normalization. The Pearson’s Correlation coefficient of each miRNA-mRNA pair was computed. Then, anti-association (CC $<-0.4$) pairs were regarded as reliability. Here, we only provided the results of miR-146b targets. The number of predicted miRNA-mRNA pairs by miRGen is 1,311, among which 664 have information of mRNA expression. 76 miRNA-mRNA pairs were anti-correlation in expression with CC $< -0.4$ in Table S2. In these mRNAs, we also mapped all reliable targets to KEGG by DAVID database [25], and found that the target genes of miR-146b mainly take part in MAPK signaling pathway, Toll-like receptor signaling pathway and Pathway in cancer. The targets of other miRNAs in $mG_4$, such as miR-146a, miR-221 and miR-222, were also found to be involved in Toll-like receptor signaling pathway, indicating that this pathway may be associated with the subtyping of breast cancer.

Discussion and Conclusions

Increasing evidence have suggested that miRNAs are involved in cancer development through regulating distinct biological processes, including cellular growth and proliferation, cellular movement and migration, extra cellular matrix degradation. Though the expression level of miRNAs is generally low in cancer, their unique profiles may have significant clinical outcomes, especially for the phenotypes of cancer. Recently, many studies have shown that the abnormal expression of miRNAs is correlated with human breast cancer. In this study we showed that the two-way clustering algorithm could result in an improved prognostic accuracy over the breast cancer patients’ survival profiles.

Table 1. miRNA clusters using CTWC

| Objects Clusters |
|------------------|
| mG6(S1) mG7 mG8 mG9 mG10 |
| mG6(S2) mG11 mG12 mG13 mG14 |
| mG6(S3) mG15 mG16 mG17 mG18 |
| mG6(S4) mG19 mG20 mG21 mG22 |
| mG6(S5) mG23 mG24 mG25 mG26 |
| mG6(S6) mG27 mG28 mG29 mG30 |
| mG6(S7) mG31 mG32 mG33 mG34 |
| mG6(S8) mG35 mG36 mG37 mG38 |
| mG6(S9) mG39 mG40 mG41 mG42 |
| mG6(S10) mG43 mG44 mG45 mG46 |
| mG6(S11) mG47 mG48 mG49 mG50 |
| mG6(S12) mG51 mG52 mG53 mG54 |
| mG6(S13) mG55 mG56 mG57 mG58 |
| mG6(S14) mG59 mG60 mG61 mG62 |
| mG6(S15) mG63 mG64 mG65 mG66 |
| mG6(S16) mG67 mG68 mG69 mG70 |
| mG6(S17) mG71 mG72 mG73 mG74 |
| mG6(S18) mG75 mG76 mG77 mG78 |

Figure 2. The five partitions of breast cancer were identified using $mG_4$ as the disease feature set in the breast cancer dataset. In the figure, each microRNA corresponds to a row, and each breast cancer sample corresponds to column. The 71 breast cancer samples were divided into five subtypes (Subtype 1, Subtype 2, Subtype 3, Subtype 4 and Subtype 5). Red areas indicate increased expression, and green areas decreased expression. Each column represents a single breast cancer sample, and each row represents a single microRNA. The dendrogram at the top shows the degree to which each breast cancer subtype is related to the others with respect to microRNA expression.

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Computational discoveries of the hidden subtypes for a complex disease have to be verified by some means, e.g., a functional assay using bioinformatics approaches or a clinical validation using epidemiological approaches such as survival analysis. In unsupervised clustering analysis, however, identifying the best subset for dissecting clinically heterogeneous disease can be a challenging task as no cross-validation can be done internally. The underlying assumption for a clustering algorithm is that microRNA with similar expression patterns and adjacent chromosome location are more likely to have a similar biological function(s). However, a clustering algorithm itself does not provide proof of the best grouping of microRNA in terms of biological functions. Thus, the biological interpretation of the disease clustering results relies heavily on the expert knowledge which often may be subjective [26]. Therefore, in this study, we designed a functional consistency score for evaluating a candidate miRNA subset in terms of functional consistency. In terms of the better-characterized functionality of subset mG4 and based on the significantly different survival results for the patients defined by the newly defined subtypes, the applied two-way clustering algorithm has been demonstrated to be a feasible and promising toolbox for peeling off molecular heterogeneities of complex human diseases.

Many methods use all the microRNAs on chips or a large number of microRNAs to predict patient survival. Since the vast majority of the microRNAs in a given dataset are irrelevant to the survival of the studied patients, this may reduce the prediction accuracy of the model because of the added noise. Hence, McLachlan et al. [27] proposed a mixture model-based approach to the clustering of microarray expression data. In this study, we applied an integrative approach that combines a SPC-based two-way clustering with a functional consensus to identify the functionally sounding and the most compact subset of microRNAs underlying the phenotypic partitions of patients.

Application of the proposed approach to breast cancer datasets led to identification of microRNA subsets, and further multivariate Cox proportional-hazards modeling defined microRNA-146b as one significant predictor for the survival of the breast cancer patients in the dataset. The individual miRNAs have only limited impact on their targets and multiple miRNAs are needed to drastically reduce transcription levels of target. We also found the targets of serval microRNAs in the optimal miRNA subset involved similar pathways. Overall, our results demonstrated that the proposed approach is highly promising for peeling off the hidden genetic heterogeneity based on modern omics data, and may lead to an improved diagnosis and treatment of cancers.

### Supporting Information

**Table S1** The correlation coefficient of among miRNAs in mG4.

| Variable | Estimated coefficient | Wald \(x^2\) | p value | Hazard ratio \((95\% CI)\) |
|----------|-----------------------|-------------|---------|--------------------------|
| hsa-miR-146b | -3.232 | 4.021 | 0.045 | 0.039\((0.002–0.930)\) |

**Table S2** The anti-association miRNA-mRNA of among miR-146b.

| Variable | Estimated coefficient | Wald \(x^2\) | p value | Hazard ratio \((95\% CI)\) |
|----------|-----------------------|-------------|---------|--------------------------|

Figure 3. Survival curves for five subtypes of the breast cancer patients in the breast cancer dataset.
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Author Contributions

Conceived and designed the experiments: LL YC LP. Performed the experiments: LL WM. Analyzed the data: LL JZ. Contributed reagents/materials/analysis tools: CL FW ZK. Wrote the paper: LL LP.

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