The eye of the Barbary sheep or aoudad (*Ammotragus lervia*): Reference values for selected ophthalmic diagnostic tests, morphologic and biometric observations

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**Abstract**

The purpose of this study was to describe the normal ocular anatomy and establish reference values for ophthalmic tests in the Barbary sheep or aoudad (*Ammotragus lervia*). Aoudad eyes are large and laterally positioned in the head with several specialized anatomic features attributed to evolutionary adaptations for grazing. Normal values for commonly used ophthalmic tests were established, Schirmer tear test (STT) - 27.22 ± 3.6 mm/min; Predominant ocular surface bacterial microbiota - *Staphylococcus* sp.; Corneal esthesiometry- 1.3 ± 0.4 cm; Intraocular pressure by rebound tonometry- 19.47 ± 3.9 mmHg; Corneal thickness- 630.07 ± 20.67 µm, B-mode ultrasonography of the globe- axial eye globe length 29.94 ± 0.96 mm, anterior chamber depth 5.03 ± 0.17 mm, lens thickness 9.4 ± 0.33 mm, vitreous chamber depth 14.1 ± 0.53 mm; Corneal diameter- horizontal corneal diameter 25.05 ± 2.18 mm, vertical corneal diameter 17.95 ± 1.68 mm; Horizontal palpebral fissure length- 34.8 ± 3.12 mm. Knowledge of these normal anatomic variations, biometric findings and normal parameters for ocular diagnostic tests may assist veterinary ophthalmologists in the diagnosis of ocular diseases in this and other similar species.

**Keywords**: Barbary sheep, Biometry, Ocular parameters, Wild caprid.

**Introduction**

The Barbary sheep or aoudad (*Ammotragus lervia*) is a species of wild caprid (goat-antelope), whose natural habitat includes northern Africa in Algeria, Tunisia, northern Chad, Egypt, Libya, northern Mali, Mauritania, Morocco, Niger and Sudan (west of the Nile, and in the Red Sea Hills east of the Nile). It is also known as waddan, arui, and arruis (Cassinello, 1998; Wacher et al., 2002; Cassinello et al., 2004). The binomial name *Ammotragus lervia* derives from the Greek ammos “sand”, referring to the sand-coloured coat and tragos (“goat”). The species name *lervia* derives from the wild sheep of northern Africa (Cassinello, 1998; Wacher et al., 2002). In its native distribution in northern Africa the aoudad was classified as a “vulnerable” species by the 2012 Red List of the International Union of Conservation of Nature (IUCN) due to natural habitat loss and poaching (Alados and Shackleton, 1997; Hilton-Taylor, 2000; Cassinello et al., 2008). It has, however, been successfully introduced to North America, Europe and elsewhere primarily for trophy-hunting purposes. These introduced populations contain a large number of individuals and are free-ranging, commonly competing with the native mammals for resources (Cassinello et al., 2008). The aoudad is a stocky, heavily built wild ruminant, with short legs and a rather long skull (Kingdon, 1997; Stuart and Stuart, 1997). Both sexes have horns that sweep backwards and outwards in an arch; those of the male are much thicker and reach up to 50 cm. Aoudads’ weight can vary from 40 to 140 kg. Males also differ from females by their significantly heavier weight, up to twice that of females (Kingdon, 1997), and the notably longer curtain of hair that hangs from the throat, chest and upper part of the forelegs (Kingdon, 1997; Stuart and Stuart, 1997; Cassinello, 1998). The coat is woolly during the winter, but mouls to a finer, sleek coat for the hot summer months. It has a sandy-brown color, darkening with age, with a slightly lighter underbelly and a darker line along the back (Kingdon, 1997; Stuart and Stuart, 1997). The eyes of the aoudad are bright and apparently large in relation to its body size, more consistent with a cervid- or antelocaprid-like morphology than a caprid one. Concerning aoudads in the scientific literature, hormonal parameters and studies about applied reproductive techniques have been published (Hamon and Heap, 1990; Crenshaw et al., 2000; Abäigar et al., 2012; Santiago-Moreno et al., 2013). Additionally, genetic studies (McLelland et al., 2005; Manca et al., 2006; Mereu et al., 2008), epidemiologic surveys and reports of specific infectious diseases (Yeruham et al., 2004; Candela et al., 2009; Pirastru et al., 2009; Portas et al., 2009; Münster et al., 2013; Morikawa et al., 2014) and parasites (Pence and...
Gray, 1981; Cho et al., 2006; Mayo et al., 2013) were investigated. An additional case report of pemphigus foliaceus has been published in this species (Brenner et al., 2009). However, no ophthalmic investigations or even reports of ocular diseases on this species were available, possibly because baseline values for diagnostic tests have not yet been established for aoudads. Knowledge of baseline values is essential for both making appropriate diagnoses and properly treating ocular diseases in zoo and exotic animals. Important parameters to be established in wild animals include tear production (Schirmer tear test, STT) and intraocular pressure (IOP), echobiometric findings as well as normal conjunctival bacterial microbiota (Prado et al., 2005; Kudirkienė et al., 2006; Montiani-Ferreira et al., 2006, 2008b; Martins et al., 2007; Wang et al., 2008; Ribeiro et al., 2009; Lima et al., 2010; Ghaffari et al., 2012). These normal ophthalmic parameters in domestic, exotic and zoo animals become important references for the veterinary clinician and other researchers once published. The purpose of this study was to describe normal ophthalmic parameters in aoudads, including morphological features, biometry of anatomical structures, corneal ultrasonic pachymetry, globe echobiometry, tear production (Schirmer’s tear test, STT), intraocular pressure (IOP), corneal sensitivity, bacterial conjunctival microbiota, and fundus photography.

**Materials and Methods**

All ophthalmic procedures using live aoudads were conducted in accordance with UFPR’s Animal Use Committee and with the ARVO Statement for the Use of Animals in Ophthalmic and Vision Research. Eighteen adult captive aoudads (11 males and 7 females) of different ages (varying from 1.5 to 7 years of age, mean 4 ± 2.0 years) belonging to Curitiba’s Zoo (Zoológico de Curitiba), Curitiba-PR, Brazil (25°25’S and 49°16’W) were captured for clinical evaluation as part of a health survey by the park authority (Fig. 1) during the winter of 2014 on three different occasions. A detailed ophthalmic evaluation including all the tests cited here was performed in this survey. Physical examinations, including a complete blood count panel, were performed before ocular examinations to exclude animals with indications of systemic disease. Aoudads with evidence of ocular or systemic diseases were excluded. Procedures and tests necessary to produce this work were split between the investigators. However, to avoid discrepancies related to inter-observer repeatability, the same person always performed the same ocular test on each occasion.

**Ophthalmic tests**

Clinical tests were performed while the aoudads were physically restrained by two experienced handlers using ropes, taking care to keep the animal comfortable. When the head was manually stabilized for taking measurements special attention was given to avoid applying pressure to the neck region with hands or ropes, to prevent iatrogenic alterations in IOP. The sequence of procedures performed in this study was, (i) ocular inspection (including photography), (ii) Schirmer tear test (STT), (iii) collection of material for bacterial culture analysis, (iv) corneal esthesiometry, (v) tonometry, (vi) central corneal thickness (CCT) measurement with an ultrasonic pachymeter, (vii) B-mode ultrasonography of the globe, (viii) funduscopy and lastly (ix) corneal and palpebral fissure measurements (Fig. 2).

**Ocular inspection**

A total of 36 eyes, from 18 healthy adult aoudads were selected and used in this investigation. The anterior ocular structures were evaluated using a Finoff transilluminator (3.5 V halogen fiber optic, Welch Allyn, Skaneateles Falls, NY, USA) and a slit lamp biomicroscope (Hawk Eye; Dioptrix, L’Union, France). The funduses were evaluated using an indirect ophthalmoscope (Heine Omega 180 Headworn Binocular Indirect Ophthalmoscope, Dover, NH) and photographed with a 7.2 megapixel reflex digital camera with a Carl ZeissTM lens and 12x of optical zoom (DSC-H5; Sony™, Minato, Tokyo, Japan) (Fig. 3a).

**Schirmer tear test**

Sterile standardized STT strips (Schering Plough Animal Health, Union, NJ, USA) were used to perform the Schirmer type I test (Fig. 2a), which measures the basal plus a portion of the reflex tear production.

**Microbiological analysis**

For the microbiological analysis, samples were obtained by carefully touching the conjunctival sac and ocular surface (cornea and bulbar conjunctiva) with a sterile cotton swab (Fig. 2e). No topical anesthetic was used with a 7.2 megapixel reflex digital camera with a Carl ZeissTM lens and 12x of optical zoom (DSC-H5; Sony™, Minato, Tokyo, Japan) (Fig. 3a).

**Fig. 1.** (a) Part of the group of aoudads (*Ammotragus lervia*) from Curitiba’s Zoo investigated in this study. The picture shows a mixed-aged group but only the adult animals were investigated. (b) A representative example of the general external appearance of the eye of the aoudad. True cilia (longer and thicker at the upper eyelid) are visible. Below the lower eyelid margin are two rows of sparsely distributed longer hairs (asterisk). Note in the anterior uvea the extensive iris collarette and the presence of an upper and a lower (more discrete) corpora nigra (arrows). The pupillary aperture shape was oval with the long axis horizontal. The limbus is relatively large and heavily pigmented.
used prior to sample collection as this may interfere with the growth of organisms (Mullin and Rubinfeld, 1997). Aerobic bacterial culture of the microorganisms was performed in BHI broth (brain–heart infusion), and on 5% sheep blood agar and MacConkey plates, which were incubated at 37°C in an aerobic environment for 24–48 h. The same bacterial growth media used in this research was also used elsewhere to establish normal conjunctival microbiota of the opossum, raccoon, ferret and chinchilla in other investigations (Pinard et al., 2002; Manca et al., 2006; Montiani-Ferreira et al., 2006, 2008b). Bacterial colonies were identified by Gram’s stain and standard procedures.

**Corneal esthesiometry**

For the normal corneal sensitivity analysis, all aoudads were manually restrained, and a Cochet-Bonnet esthesiometer (Luneau Ophthalmologie, Chartres Cedex, France) was used (Fig. 2b). This instrument contains an adjustable nylon filament with a defined diameter, length and surface (0.12 mm diameter, 60 mm length, and 0.0113 mm² surface), which was applied at different lengths to the center of the cornea. A stimulus produced by the instrument’s nylon monofilament that reaches the corneal touch threshold induces a corneal reflex, consisting of prompt eyelid closure, and discrete retraction of the globe. In this study only the center of the cornea was analyzed for corneal touch threshold, which was repeated five times using the same length of the nylon filament. The length of the nylon filament was then decreased at 5-mm increments until each aoudad responded with a corneal blink reflex. The corneal touch threshold was then quantified in millimeters length of the filament necessary to cause a blink reflex. The length of the filament, indicating a corresponding pressure at which the corneal blink reflex was positive, was deemed the central corneal sensitivity or central corneal touch threshold.

**Intraocular pressure**

Intraocular pressure (IOP) was measured in 36 eyes, using a veterinary rebound tonometer (Tonovet, Veterinary Division of S&V Technologies AG, Henningsdorf, Germany) (Fig. 2d) with the P setting, which was a preset for other animals except dogs and horses. Six measurements were taken and averaged by the tonometer’s internal software.

**Central corneal thickness**

Central corneal thickness (CCT) measurements were taken after the instillation of sterile topical anesthetic (proparacaine hydrochloride 0.5% ophthalmic solution USP; Alcon Laboratories, Forth Worth, TX, USA). CCT was measured using an ultrasonic pachymeter (Model 200P+; Micropach, Sonomed, Lake Success, NY, USA), with the speed of sound in the cornea preset at 1640 m/s (Fig. 2c).

**B-mode ultrasonographic biometry**

B-mode scan ultrasonography was performed using
a Sonix SP High Performance B-mode System (Ultrasonix, Richmond, BC, Canada). A drop of topical local anesthetic (0.5% proxymetacaine chlorohydrate, Anestalcon®, Alcon Laboratórios do Brasil, São Paulo, SP, Brazil) was instilled on each eye before ultrasonography. The B-scan 14-MHz probe was gently placed on the corneal surface perpendicular to the center of the cornea using ultrasonic transmission gel (Aquasonic-100; Parker Laboratories Inc., Fairfield, NJ, USA). Care was taken during probe placement to avoid corneal indentation. Reflected ultrasonic waves were captured. Optimal positioning was confirmed when the posterior wall of the eye globe could be clearly visualized on the B-scan ultrasonogram and the image appeared symmetrical and the reflections from the four principal landmarks (cornea, anterior lens surface, posterior lens surface and retinal surface) along the optic axis were perpendicular. The optimal image was frozen on the screen and then all echobiometric measurements were taken (Fig. 2f).

**Fundoscopy**

After B-mode ultrasonographic biometry the aoudads’ eyes were gently rinsed twice with 0.9% saline solution in order to remove the ultrasonic transmission gel. Subsequently the aoudad’s funduses were examined using an indirect ophthalmoscope (Heine Omega 180 Headworn Binocular Indirect Ophthalmoscope, Dover, NH) and photographed using the topical endoscopy fundus imaging technique (TEFIT) (Fig. 3a) or a slit lamp containing a built-in indirect ophthalmoscopy lens (Digital 1.0x Imaging Lens, Hawk Eye, Dioptrix, L’Union, France) (Fig. 3b). For the TEFIT procedure a rigid, 8-mm-diameter laparoscope with a 0 degree angle and a crescent-shape illumination tip (WeckTM, Pilling Weck, Markham, ON, Canada) was used. Both the rigid arthroscopy probe and the rigid laparoscope were connected to an adapter of a 7.2 megapixel reflex digital camera with a Carl ZeissTM lens and 12x of optical zoom (the same previously cited). The light source was a 175W xenon lamp (Karl StorzTM, Tuttinglen, Germany) linked to the arthroscopy probe and the rigid laparoscope by a flexible fiber optic cable. Pupillary dilation for fundoscopy and fundus photography was performed following instillation of the following eyedrops, tropicamide 1% and phenylephrine 10% (Frumtost, São Paulo, SP, Brazil) one drop of each in each eye, with approximate 3-min intervals, every 10 min three times.

**Corneal and palpebral fissure biometry**

Palpebral fissure length, vertical and horizontal corneal diameters were measured using a stainless steel caliper ruler with an LCD display and an accuracy of ±0.02 mm (Neiko Tools, Klamath Falls, OR, USA).

**Statistical analyses**

The obtained data were submitted to a Kolmogorov-Smirnov Goodness-of-Fit Test. Unpaired t-tests were used for data comparison between, right and left eyes and males and females. P-values < 0.05 were deemed significant. JMP (SAS Institute, Inc., Cary, NC, USA) software was used to perform both descriptive and inferential statistical analyses. Measurements are reported as mean ± standard deviation (SD).

**Results**

All continuous numeric data obtained for all ophthalmic tests in the population used in this investigation were normally distributed according to the Kolmogorov-Smirnov Goodness-of-Fit Test. Unpaired t-tests were used for data comparison between, right and left eyes and males and females. P-values < 0.05 were deemed significant. JMP (SAS Institute, Inc., Cary, NC, USA) software was used to perform both descriptive and inferential statistical analyses. Measurements are reported as mean ± standard deviation (SD).

| Ophthalmic Test or Parameter       | Unit | Mean     | Standard Deviation | 95% Confidence Interval |
|------------------------------------|------|----------|--------------------|-------------------------|
| Schirmer tear test                 | mm/min | 27.22    | 3.6                | 26.04-28.4              |
| Esthesiometry                      | cm   | 1.3      | 0.4                | 1.18-1.43               |
| Intraocular pressure               | mmHg | 19.47    | 3.9                | 18.2-20.74              |
| Central corneal thickness          | μm   | 630.07   | 20.67              | 623.32-636.82           |
| Axial globe length                 | mm   | 28.43    | 0.88               | 26.65-28.43             |
| Anterior chamber depth             | mm   | 5.03     | 0.17               | 4.7-5.4                 |
| Lens thickness                     | mm   | 9.4      | 0.33               | 8.73-10.06              |
| Vitreous chamber depth             | mm   | 14.1     | 0.53               | 12.93-15.06             |
| Palpebral fissure length           | mm   | 34.8     | 3.12               | 33.77-35.82             |
| Corneal horizontal length          | mm   | 25.05    | 2.18               | 24.34-25.77             |
| Corneal vertical length            | mm   | 17.95    | 1.68               | 17.40-18.50             |

Table 1. Results obtained for selected ophthalmic diagnostic tests and echobiometric findings for the aoudad (Ammotragus lervia) eye.
and more sparsely distributed. Aoudads have a third eyelid (nictitating membrane) which moves across the surface of the cornea from the nasal canthus to the temporal canthus. The leading edge of the third eyelid is pigmented. Above the upper eyelid margin and below the lower eyelid margins, two rows of modified-sparingly distributed longer hairs, resembling vibrissae, also called “tactile hair”, were found in all individuals (Fig. 1b). There were approximately 16 to 18 pairs located above and 6 to 8 pairs below the eye. The iris colors of individual animals varied from a yellowish-brown to a grayish-brown. The iris collarette showed no crypts of Fuchs visible, being somewhat flat (Fig. 1b). *Corpora nigra* were present at the ciliary margin (the peripheral border of the iris). The lower *corpora nigra* were considerably more discrete (Fig. 1b). The pupillary aperture shape was oval with the long axis being horizontal. The presence of corpora nigra makes the pupil gain a rectangular appearance when observed from a distance.

**Schirmer tear test (STT)**

No significant STT differences were determined between right and left eyes or between sexes. Mean STT results for both eyes was 27.22 ± 3.6 mm/min.

**Microbiological analysis**

Bacteria were isolated in microbiological samples from 33 out of 36 eyes. Five different genera of gram-positive bacteria species were identified. The genera of the isolates were, *Corynebacterium*, *Micrococcus*, *Bacillus*, *Streptococcus* and *Staphylococcus* sp. Four different genera of gram-negative bacteria were isolated. The genera of the isolates were, *Escherichia*, *Acinetobacter*, *Enterobacter* and *Citrobacter* sp. A single genus of bacteria was isolated from 11 eyes. Two genera of bacteria were isolated from 20 eyes. Three genera of bacteria were isolated from two eyes. *Staphylococcus* sp. were the most common bacteria isolated, being present in 13 eyes (prevalence of 36.1%). *Micrococcus* sp. and *Bacillus* sp. were the second most common bacteria isolated, being present in 9 eyes each (prevalence of 25%). Lastly, *Corynebacterium* sp. was present in 5 eyes (prevalence of 13.88%).

**Corneal esthesiometry**

There were no significant differences between males and females or between left and right eyes. The mean central corneal sensitivity was 1.3 ± 0.4 cm.

**Intraocular pressure (IOP)**

The mean value for IOP was 19.47 ± 3.9 mmHg. There was no significant difference in IOP between males and females and no significant differences between left and right eyes.

**Central corneal thickness (CCT)**

The mean CCT was 630.07 ± 20.67 μm. There was no significant difference in CCT between males and females and no significant differences between left and right eyes.

**B-mode ultrasonographic biometry**

No significant biometric differences were determined between right and left eyes or between sexes. The mean axial globe length was 29.94 ± 0.96 mm. Mean anterior chamber depth (axial anterior chamber length) was 5.03 ± 0.17 mm. Mean lens thickness (axial length) was 9.4 ± 0.33 mm. Mean vitreous chamber depth (axial chamber length) was 14.1 ± 0.53 mm.

**Fundus examination and fundus photography**

As viewed by the ophthalmoscope, it was possible to observe that the aoudad retina possess an extensive *tapetum lucidum* usually of a greenish-yellow to a yellowish-green color with a typical holangiotic retinal vascular pattern (Fig. 3). The *tapetum* has a granular or speckled appearance (Fig. 3). The optic disc was oval in shape and located just inferior to the inferior border of the *tapetum lucidum*. The major blood vessels of the retina radiate from the center of the optic nerve (Fig. 3). Blood vessels arising from the dorsal and ventral quadrants taper toward a region just above the inferior border of the *tapetum lucidum*. At this region no blood vessels are present and an imaginary line can be traced creating a streak where retinal blood vessels are rare or absent (Fig. 3a).

**Corneal and palpebral biometry**

The transition between cornea and the sclera (limbus) is relatively large and heavily pigmented and appears as a dense thick band (Fig. 1b). Mean horizontal corneal diameter (or width) of both eyes was 25.05 ± 2.18 mm and the mean vertical corneal diameter of both eyes was 17.95 ± 1.68 mm. The mean horizontal palpebral fissure length of both eyes was 34.8 ± 3.12 mm.

**Discussion**

This study established normal values and ranges of several ophthalmic and biometric measurements of the eyes of a group of clinically normal aoudads (*Ammotragus lervia*), which was previously unavailable in the scientific literature.

The eyes of the aoudad are relatively large for the size of its head and body, and are therefore prominent. For instance, Barbary sheep eyes are bigger than the ones of the normal goat or sheep and other wild same size animals belonging to the Order Artiodactyla. The eyelashes and eyelid vibrissae are long and add to the distinctive appearance. In other species already investigated, vibrissae are considered to be true sensory organs located in anatomical areas where protective reflexes are important such as around the eye, or where environmental light is limited (McGreevy, 2004). Aoudads have a fairly elongated head and their eyes are placed laterally and posteriorly. These features together are similar to the horse head morphology (McGreevy, 2004) and are probably evolutionary adaptations to prevent tall grass from obstructing the view when grazing in both species. The presence of an elongated horizontally oval pupil observed here in the aoudad but also in other ungulates...
such as horses (Murphy and Arkins, 2007), cows, sheep and goats (Walls, 1943) allows for wide lateral vision (Murphy and Arkins, 2007). This type of pupil alternatively called “rectangular” (Prince, 1956) is also present in the deer, camel and hyrax. Optical analyses show that this horizontal pupillary elongation expands the field of view horizontally allowing terrestrial prey animals to see objects near the ground both in front of and behind them (Sprague et al., 2013). Another evolutionary adaptation found in the eye of the aoudad is the corpora nigra, which are pigmented projections found on the upper and lower margins of the pupillary aperture. This anatomic structure already described in ungulates (Walls, 1943) is known to have many functions including contribution to pupillary constriction, prevention of actinic damage during grazing and possibly functions as an anti-glare device (Davidson, 1991). In the eye of the aoudad the upper corpora nigra is considerably larger than the lower ones. The authors believe that this feature accentuates information from the inferior visual field (Davidson, 1991). The horizontal palpebral fissure length of the aoudad (34.8 mm) is only a bit smaller than that of the cow (44.4 mm) and that of the horse (39.5 mm) (Wieser et al., 2013), which are both larger and heavier animals. It is however, considerably bigger than that reported for animals with similar sizes and weight such as sheep and goats (Walls, 1943), goats (Hughes and Whitteridge, 1973; Gonzalez-Soriano et al., 1997), giraffes (Coimbra et al., 2013), black rhinoceros (Pettigrew and Manger, 2008). The visual streak is not unique to ungulates. It has been found in other taxa including reptiles e.g. American garter snake (Wong, 1989), several species of birds e.g. Canada goose (Fernández-Juricic et al., 2011), ostrich (Boire et al., 2001), manx shearwater (Hayes et al., 1991), over 30 species of fish, and many other non-ungulate mammals including carnivores (spotted hyena Crocuta crocuta (Calderone et al., 2003)), aquatic mammals (common dolphin (Dral, 1983)), and marsupials (scrub wallaby (Tancred, 1981)). The visual streak determines visual acuity in a particular part of the visual field and its presence may have ecological correlations with habitat-type, anti-predator behaviors, and orientation behaviors (Johnson, 1901; Pumphrey, 1948; Luck, 1965; Hughes, 1977; Fernández-Juricic et al., 2011). The visual streak has been described to provide panoramic vision (Johnson, 1901; Vincent, 1912; Collin, 1999), and in combination with laterally placed eyes reduces the size of the blind area and offers a wide field of visual coverage (Hughes, 1977; Fernández-Juricic et al., 2011) thus reducing the need to sample visually by moving the eyes or head (Collin, 1999). Combined with the oval-shaped pupillary aperture, a visual streak in the aoudad would likely greatly enhance vision in the horizontal plane. The STT is considered the gold standard test used to diagnose keratoconjunctivitis sicca (KCS) in domestic and wild animals. It is therefore important to perform a STT in all aoudads with ocular disease to rule out KCS as a cause of chronic eye disease such as corneal ulcers, conjunctivitis, keratitis and ocular discharge (Brooks, 2010; Trbolova et al., 2012). Although there has been no report of KCS in aoudads to date, it likely occurs in the species as it is a common ocular disease in most animals and human beings. It may be that the disease is underreported in aoudads due to the lack of knowledge of normal values for this test. When comparing STT values found in available studies of other species of ruminants, STT results in aoudads are quite high, similar but even higher to those reported for humans and the area centralis in dogs and cats, where an increased density of retinal neurons affords higher visual acuity. Early ophthalmoscopic observations of visual streaks reported a band-like thickening across the retina (Chievitz, 1889, 1891; Slonaker, 1897), which has since been shown by microscopic examination to be a high density of retinal ganglion cells. Although further histologic characterization is required to be able to define a visual streak in the aoudad, it is likely that the fundic morphology described in the aoudad fundus represents a visual streak considering the presence of a streak in other previously studied ungulates: cattle (Hebel, 1976), sheep (Hebel, 1976; Shinozaki et al., 2010), goats (Hughes and Whitteridge, 1973; Gonzalez-Soriano et al., 1997), giraffes (Coimbra et al., 2013), black rhinoceros (Pettigrew and Manger, 2008). The visual streak is not unique to ungulates. 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Although there has been no report of KCS in aoudads to date, it likely occurs in the species as it is a common ocular disease in most animals and human beings. It may be that the disease is underreported in aoudads due to the lack of knowledge of normal values for this test. When comparing STT values found in available studies of other species of ruminants, STT results in aoudads are quite high, similar but even higher to those reported for...
sheep (26.40 ± 17.70 mm/min) (Wieser et al., 2013), llamas (17.3 ± 1.1 mm/min) (Trbolova et al., 2012), goats (14.50 ± 3.78 mm/min) (Wieser et al., 2013), and pigmy goats (15.8 ± 5.7 mm/min) (Broadwater et al., 2007). Normal STT results obtained for the Barbary sheep (27.22 ± 3.6) are quite high compared to other caprids (subfamily Caprinae) and other species of Artiodactyla. It is significantly higher than the one from llamas (P=0.0001), goats (P=0.0001) and pigmy goats (P=0.001). The observed STT mean value in Barbary sheep is higher than the one from sheep, however, due to the large variation reported for the sheep STT (SD = 17.70) (Wieser et al., 2013), the analysis showed that the difference is not statistically significantly.

Normal conjunctival bacterial microbiota has been studied in several wild mammals such as the opossum (Pinard et al., 2002), bison (Davidson et al., 1999), deer (Dubay et al., 2000), and elephant (Tuntivanich et al., 2002). In the vast majority of these reports, gram-positive bacteria were the most common isolates and the present report is no exception. Both pathogenic and nonpathogenic bacteria were found in this investigation. _Escherichia coli_, _Enterobacter_ sp and _Citrobacter_ sp were isolated from the eyes of aoudads in this study. The presence of these gram-negative bacteria suggests possible eye contamination with fecal material and/or may represent a transient agent of the conjunctiva. Nonetheless, _Escherichia coli_ was also isolated from normal conjunctival microbiota of dogs (Prado et al., 2005; Wang et al., 2008) and horses (Pisani et al., 1997; Andrew et al., 2003). _Enterobacter_ sp and _Citrobacter_ sp were isolated from the conjunctiva of clinically normal eyes of horses and human beings working as health professionals in a hospital environment (Pisani et al., 1997; Trindade et al., 2000). Additional studies are still necessary to try to determine whether or not some of these gram-negative bacteria are normal inhabitants of the aoudad’s ocular microbiota. The label pathogenic versus non-pathogenic is misleading because it is known that in some cases of bacterial conjunctivitis, a formerly nonpathogenic conjunctival bacterium can overgrow and cause an imbalance of the ocular surface microbiota population, becoming pathogenic (Samuelson, 1999). The Cochet–Bonnet esthesiometer estimates the degree of sensitivity of the cornea by evaluating the corneal touch threshold (Chan-Ling, 1989; Barrett et al., 1991). The mean corneal touch threshold obtained in this investigation was similar to that of the foal (1.4 cm) (Brooks et al., 2000), chinchilla (1.24 cm) (Lima et al., 2010), Guinea pig (1.35 cm) (Wieser et al., 2013) and rabbit (1.47 cm) (Wieser et al., 2013), demonstrating that aoudads possess a less sensitive cornea compared to other species such as the adult horse, cat and cow (Wieser et al., 2013). These results should be interpreted with caution because of the well-known low precision of the Cochet–Bonnet esthesiometer in the 0.5- to 2.0-cm filament length range (Wieser et al., 2013). The aoudad’s corneal sensitivity encountered in this investigation was exactly within that range. The pressure applied to the surface of the cornea by the examiner also can vary. It is known that these parameters affect this test results significantly (Boberg-Ans, 1956). In the present study, the temperature and humidity were not assessed in order to be able to correct the corneal sensitivity measurements with the nylon filament. Unfortunately, no formula or correction table exists at this time for the nylon filament currently used and the temperature or humidity conditions, which imposes a challenge for extrapolating corneal sensitivity data obtained with the Cochet-Bonnet esthesiometer. In light of all these possible variables and interferences produced by the examiner, some authors claim that a new esthesiometer, which can display the pressure applied to the surface of the cornea, should be created in order to make the measurement of the CTT more sensitive and comparisons between investigations more precise. Additionally, it might be worth considering a non-contact esthesiometer, since comparing to the Cochet-Bonnet esthesiometer the former allows for superior stimulus reproducibility and better control over stimulus characteristics, in addition to the ability for exploration of the response of all different types of neuro-receptors on the ocular surface (Golebiowski et al., 2011).

Tonometry is a fundamental part of a complete ophthalmic evaluation in any animal species. The main value of tonometry lies in the ability to detect pressure increases as an important clinical sign of glaucoma. However, a normal range of values for each species needs to be established. IOP measurements in the aoudads using the rebound tonometer resulted in means and ranges that were slightly higher than those reported for most other wild and domestic ungulates (Ofri et al., 2000, 2001; Willis et al., 2000). For instance, normal reported mean IOP for sheep was 16.36 ± 2.19 mmHg (Pigatto et al., 2011), which was significantly lower (P=0.0018) than that found in the aoudad (19.47 ± 3.9 mmHg). The aoudad’s IOP seems to be similar to other ungulates with higher IOP such as the zebra (Ofri et al., 1998) and dairy cattle (Gum et al., 1998). However, comparison is difficult since most of the normal ranges for IOP previously reported in ungulates were obtained with applanation tonometers, and some even with indentation tonometry (Ofri et al., 1998). Before comparing and extrapolating IOP data from one study to others, researchers need to make sure the tonometry method was the same. It was shown that the Tonovet rebound tonometer may significantly overestimate the IOP values compared to the applanation tonometer, at least in one study using normal Eurasian eagle owls (Jeong et al., 2007). Another study conversely showed
that results for the TonoVet-D calibration are similar to those obtained for dogs (Knollinger et al., 2005). Even though the rebound tonometer is tolerated well by most animal species because of its rapid and minimal stress-inducing method, another factor to be considered when establishing IOP in wild and exotic species is stress. It is known that IOP values increase if the animal is firmly restrained, particularly in wild animal species (Jeong et al., 2007). All animals examined in this study were physically restrained and thus it is possible that stress could have influenced our results, though care was taken to avoid neck pressure.

Ultrasonic corneal pachymetry is an accurate and reliable in vivo method to measure corneal thickness in animals and human beings (Korah et al., 2000). It was shown that ultrasonic pachymetry set at a standard velocity of 1636 m/s overestimates CCT as compared to optical coherence tomography (Alario and Pirie, 2014). However, correlation between the two mentioned modalities is excellent. Mean central corneal thickness (CCT) acquired with an ultrasonic pachymeter has been the subject of a number of reports investigating the cornea of human beings (Korah et al., 2000), several domestic (Stapleton and Peiffer, 1979; Gilger et al., 1991, 1998), exotic and wild animals (Montianni-Ferreira et al., 2006, 2008a, 2008b; Lima et al., 2010). In our investigation, mean CCT of the aoudad was not significantly different between males and females. The aoudad CCT is slightly thicker than adult dogs (598.54 μm) (Alario and Pirie, 2014) and slightly thinner than the horse (785.60 μm) (Plummer et al., 2003). It is similar to that of adult Saanen goats using a high-resolution 20-MHz A- and B-mode ultrasonography transducer (Ribeiro et al., 2009).

Echobiometric data of the globe obtained using A- and B-mode ultrasonography were reported in children (Kurtz et al., 2004) several domestic (Schiffer et al., 1982; Rogers et al., 1986; Cottrill et al., 1989; Gilger et al., 1998; Tuntivanich et al., 2002; Plummer et al., 2003; Ribeiro et al., 2009), exotic and wild mammal species (Fernandes et al., 2003; Hernandez-Guerra et al., 2007; Montianni-Ferreira et al., 2008a; Lima et al., 2010; Ruiz et al., 2015). The aoudad’s axial globe length, lens thickness, and chamber depths were not significantly different according to the eye (left or right) studied or gender. This lack of difference was also observed in dog eyes and eyes of most other wild and exotic animals studied using B-mode ultrasonography. The eye of the aoudad is large in both ways, absolutely and relative to its body size. The axial globe length found for adult aoudads is larger than that obtained in other large mammals including cadaveric eyes of Rambouillet sheep (El-maghrabmy et al., 1995), Ile de France Sheep (Brandao et al., 2004) and Saneen goats (Ribeiro et al., 2009). The dimension of the internal structures such as anterior chamber depth, lens thickness and vitreous chamber depth follow the same pattern, being all comparable but larger than the sheep (Brandao et al., 2004) and goat (Ribeiro et al., 2009). Only the bovine (Potter et al., 2008), buffalo (Bos bubalis) (Kassab, 2012; Assadnassab and Fartashvam, 2013) and the dromedary eye (Osuobeni and Hamidzada, 1999; Kassab, 2012) demonstrated similar echobiometric dimensions, with equivalent lens thickness and vitreous chamber depth even though these are considerable larger ungulates in terms of body size.

In conclusion, this study provides novel data for normal values and reference ranges for several ophthalmic tests and ocular biometric parameters in healthy aoudads. The eyes are large and laterally placed in the head with several anatomic features that are likely evolutionary adaptations for grazing, which was also previously observed in other prey species of ungulates, such as horses, sheep and cattle.

Often a complete ocular examination of zoo animals is not routinely performed (Townsend, 2010) due to limitations such as lack of appropriate instruments (ophthalmoscopes, tonometers), disposable diagnostic test material (such as STT strips, fluorescein strips and eyedrops) and proper facilities (safe, large dark rooms). Nevertheless, the results of this study may assist veterinarians and veterinary ophthalmologists in the diagnosis of ocular diseases in aoudads.

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**Conflict of interest**

The authors declare that there is no conflict of interest.

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