The Progeroid Phenotype of Ku80 Deficiency Is Dominant over DNA-PKCS Deficiency

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Abstract

Ku80 and DNA-PKCS are both involved in the repair of double strand DNA breaks via the nonhomologous end joining (NHEJ) pathway. While ku80−/− mice exhibit a severely reduced lifespan and size, this phenotype is less pronounced in dna-pkcs−/− mice. However, these observations are based on independent studies with varying genetic backgrounds. Here, we generated ku80−/−, dna-pkcs−/− and double knock out mice in a C57Bl6/J*FVB F1 hybrid background and compared their lifespan, end of life pathology and mutation frequency in liver and spleen using a lacZ reporter. Our data confirm that inactivation of Ku80 and DNA-PKCS causes reduced lifespan and bodyweights, which is most severe in ku80−/− mice. All mutant mice exhibited a strong increase in lymphoma incidence as well as other aging-related pathology (skin epidermal and adnexal atrophy, trabecular bone reduction, kidney tubular anisokaryosis, and cortical and medullar atrophy) and severe lymphoid depletion. LacZ mutation frequency analysis did not show strong differences in mutation frequencies between knock out and wild type mice. The ku80−/− mice had the most severe phenotype and the Ku80-mutation was dominant over the DNA-PKCS-mutation. Presumably, the more severe degenerative effect of Ku80 inactivation on lifespan compared to DNA-PKCS inactivation is caused by additional functions of Ku80 or activity of free Ku70 since both Ku80 and DNA-PKCS are essential for NHEJ.

Introduction

DNA damage is known to be involved in tumorigenesis and aging. There are multiple DNA repair pathways that specialize in repairing a specific DNA lesion. One such pathway is nonhomologous end joining (NHEJ), which plays an import role in the repair of double strand breaks (DSBs). Ku80 and DNA-PKCS are both components of the NHEJ pathway. Ku80 forms a heterodimer with Ku70, known as KU, which binds to DNA ends at a DSB [1]. DNA-dependent protein kinase (DNA-PK) is a holo enzyme formed by a complex between Ku and DNA-dependent protein kinase catalytic subunit (DNA-PKcs). DNA-PKcs, together with Artemis, Xrcc4-DNA ligase and Xrcc4-like factor, process DNA overhangs and ligation [2–4]. Cells with deletion of any of these NHEJ components show severe combined immunodeficiency (SCID) due to defects in assembling variable (diverse) joining (V(D)J) segments of antigens, genetic instability and hypersensitivity to DSB inducing agents [5–9]. However, there is some phenotypic variation. For instance, deletion of Ku70 and Ku80 results in reduced size, severely decreased lifespan, neuronal apoptosis and accelerated aging.

Strikingly, in previous experiments using mixed backgrounds, these mice seem to be protected against tumor development, although they show a low level of thymic lymphomas [10,11]. In contrast, such a phenotype is less pronounced in DNA-PKCS knock out mice. Compared to the ku70−/− and ku80−/− mice, the early aging phenotype for the dna-pkcs−/− mice is less severe and best observed with shortened telomeres [12–14]. Yet, these studies were performed in different genetic backgrounds in different labs. Therefore, phenotypic differences could be the result of inconsistent genetic backgrounds and environments. On the other hand, some phenotype differences can be explained by additional functions of the deleted NHEJ components or that a non-deleted NHEJ component is deleterious in absence of the deleted protein. Some phenotype differences can be explained by additional functions of the deleted NHEJ components or that a non-deleted NHEJ component is deleterious in absence of the deleted protein. Some phenotype differences can be explained by additional functions of the deleted NHEJ components or that a non-deleted NHEJ component is deleterious in absence of the deleted protein. Some phenotype differences can be explained by additional functions of the deleted NHEJ components or that a non-deleted NHEJ component is deleterious in absence of the deleted protein. Such deleterious effects have been observed previously [15,16]. For instance, DNA ligase IV deficiency is lethal in presence of Ku80 but lethality is rescued by deletion of Ku80 [16]. Furthermore, the prenatal lethal phenotype of Xrcc4 deficient mice is partly rescued by p53 knock out [15], that negates neuronal apoptosis.

Here, we generated cohorts of ku80−/−, dna-pkcs−/−, double knock out and wild type mice in an identical genetic background.
and compared lifespan, development of body weight, end-of-life pathology and accumulation of genetic insults (LacZ mutant frequencies in liver and spleen). We find that ku80<sup>-/-</sup> mice exhibit a more severe phenotype than dna-pk<sub>cs</sub><sup>−/−</sup> mice while the double mutant mice are as severe as the ku80<sup>-/-</sup>−/− mice. These observations suggest that Ku80 has additional functions that do not require DNA-PK<sub>cs</sub>, or that free Ku70 has a deleterious affect in the absence of Ku80. Both possibilities are in line with our previous reports that show Ku70 and Ku80 have functions independent of the Ku heterodimer [17].

**Results**

Previous reports suggest that ku80<sup>-/-</sup> mutant mice exhibit a more severe phenotype than dna-pk<sub>cs</sub><sup>−/−</sup> mice. This was not predicted since both Ku80 and DNA-PK<sub>cs</sub> are essential for NHEJ. Thus, the phenotypic differences could be due to divergent function or divergent genetic background and environment. Alternatively, one of the components could be toxic in the absence of the deleted protein. To better understand the reason for these different phenotypes we generated ku80<sup>-/-</sup>−/− and dna-pk<sub>cs</sub><sup>-/−</sup>−/− double mutant cohorts in the same genetic background using double heterozygous breeders (ku80<sup>+</sup>/− dna-pk<sub>cs</sub><sup>+</sup>/−) as shown in figure 1. The breeding pairs were composed of C57Bl6/J-pUR288 males and FVB females so the cohorts were F1 brothers and sisters raised in the same cages. Thus, all cohorts are controlled for genetic background and environment so any phenotypic difference raised in the same cages. Thus, all cohorts were F1 brothers and sisters.

During the longevity study, all mice were weighed every two weeks. As can be seen in figure 2, all mutant mice were smaller compared to wild type mice. ku80<sup>-/-</sup>−/− and double knock out mice weighted between 10 and 20 grams, while dna-pk<sub>cs</sub><sup>-/−</sup>−/− mice weighed between 15 and 35 grams and were therefore considerably heavier than ku80<sup>-/-</sup>−/− and double knock out mice. Due to differences in body size, organ weights differed significantly between the different genotypes (Figure S1). However, after correction for body weight, most organ weights were comparable.

The exception is the testis, which was significantly heavier in dna-pk<sub>cs</sub><sup>-/−</sup>−/− compared to the other knock out mice. Marginal significant differences were observed for males in relative organ weight for kidney, liver, spleen and heart. The wild type cohort was terminated after the final mutant mouse died since their life span appeared much longer; therefore, no end of life organ weight data were collected for this cohort. Thus, deletion of either Ku80 or DNA-PK<sub>cs</sub> reduced body weight with Ku80-deletion being more severe and dominant to DNA-PK<sub>cs</sub>-deletion.

All mutant mice displayed a severely reduced lifespan compared to wild type mice (Fig. 3). Previously, we showed that dna-pk<sub>cs</sub><sup>-/−</sup>−/− mice were longer-lived than ku80<sup>-/-</sup>−/− mice for both males and females (p < 0.01 and p = 8.7*10−5 respectively) [17]. Here we also show that the double knock out mice had the same short life span as ku80<sup>-/-</sup>−/− mice (Fig. 3). Thus, deletion of either Ku80 or DNA-PK<sub>cs</sub> reduced life span with Ku80-deletion being more severe and dominant to DNA-PK<sub>cs</sub>-deletion.

We screened for aging pathology in a selected set of animals (n = 4–5). Results are shown in table 1. Evidence of aging was found in the kidney. Mild increased renal tubular anisokaryosis was observed in all knock out models. Moderate to severe renal tubulonephrosis was present in all mutant cohorts compared to wild type animals. Analysis of skin revealed moderate to severe epidermal and adnexal atrophy. Dermis and epidermis thickness was reduced in all mutant mice. Finally, all mutant mice showed mild to moderate reduction in trabecular bone thickness, which is indicative of osteopenia. There were no differences between genotypes in anisokaryosis or lipofuscin accumulation in liver, which was previously observed in aged wild type mice [18]. Thus, deletion of either Ku80 or DNA-PK<sub>cs</sub> caused an early onset of many but not all aging characteristics previously reported for wild type mice.

Mutant cohorts showed a high tumor incidence at the time of death compared to age-matched controls (table 2). Most tumors were observed in the thymus followed by liver tumors. Pathological examination showed that tumors were almost exclusively

**Figure 1. Breeding strategy to generate the knockout cohorts.** Ku80<sup>-/-</sup> and dna-pk<sub>cs</sub><sup>−/−</sup> animals are backcrossed for multiple generations to C57Bl6/J-pUR288 (left of dashed line) and FVB (right of dashed line) background. Double heterozygous knock out C57Bl6/J male mice are crossed with double heterozygous knock out FVB female mice. Using this strategy all four desired cohorts are generated as F1 hybrids (grey boxes). doi:10.1371/journal.pone.0093568.g001
lymphomas. CD3 staining showed lymphomas to be of T-cell origin (three mice analyzed for each genotype). There were also rare hepatic adenomas (n = 6), a hepatic carcinoma (n = 1) and a bronchiolo-alveolar carcinoma (n = 1), which might be caused by the C57Bl6/J background according to the Mouse Tumor Database (http://tumor.informatics.jax.org/mtbwi/index.do).

Lymphomas were also present in other organs than thymus with a high frequency in liver, kidney, spleen and lymph nodes. Lymphoid tissues of mutant animals, not affected by lymphomas, showed severe lymphoid depletion (table 1) as expected due to the failure to complete V(D)J recombination. This was observed in spleen and mesenteral lymph nodes and was severe in most mutant animals, with exception of female dna-pkcs<sup>−/−</sup> mice, which showed a mild to moderate lymphoid depletion in mesenterial lymph nodes. Animals affected by lymphoid depletion also showed an increased myeloid/erythroid ratio. Although tumor incidence was slightly higher in ku80<sup>−/−</sup> mice compared to dna-pkcs<sup>−/−</sup> and the highest in double knock out mice, this did not reach statistical significance. No synergistic effects of DNA-PKCS deletion in addition to Ku80 deletion were found nor did deletion of one protein ameliorate the phenotype for deletion of the other protein.

Since Ku80 and DNA-PKCS are DNA damage repair proteins, it would be expected that deletion of these proteins affect mutant frequency. In order to test the LacZ mutant frequencies in liver and spleen, 5 ku80<sup>−/−</sup>, dna-pkcs<sup>−/−</sup> and double knock males were killed at 38.7, 41.3 and 33.7 weeks, which corresponds approximately with their median lifespan (figure 3). Because the wild type cohort was not maintained beyond median age, we selected the 5 oldest wild type males available at the same observation time (mean age is 37.3 weeks). No significant differences in mutant frequency were observed between genotypes in both liver as spleen (figure 4). We did observe a trend towards increased mutant frequency in double knock out compared to wild type mice in spleen, but this did not reach statistical significance (p = 0.07).

Discussion

Previously, it was reported that Ku80 inactivation resulted in reduced cancer and accelerated aging [12,14,19] while a similar but less pronounced phenotype was observed with DNA-PKCS inactivation [12–14]. This phenotypic variance is unexpected since both Ku80 and DNA-PKCS are essential for NHEJ. Therefore, these different phenotypes could be due to differences in genetic background and/or environment. However, our experiment presented here does not support either possibility since we controlled for both genetic background and environment but still observed a disparity in phenotypes. Alternatively, ku80<sup>−/−</sup> mice could exhibit a more severe phenotype than dna-pkcs<sup>−/−</sup> mice, if DNA-PKCS is toxic in the absence of Ku80. There is evidence for this possibility since Ku80-deletion rescued embryonic lethality for DNA ligase IV-null mice showing that Ku80 was toxic in the absence of DNA ligase IV. However, deleting DNA-PKCS did not ameliorate the Ku80-mutant phenotype. Instead the double-mutant mice exhibited the same severe phenotype as the Ku80-mutant mice. Therefore, our experiment suggests that Ku80 has extra-NHEJ function or that free Ku70 has a toxic activity in the absence of Ku80.

Our previously published data supports both possibilities. First we found that Ku80 has extra-NHEJ activity in mice by comparing p53-mutant mice deleted for Ku80 or Ku70 or both.
For this experiment, we found that ku70<sup>−/−</sup> p53<sup>−/−</sup> mice lived longer than ku80<sup>−/−</sup> mice. This lifespan extension required Ku80 since the triple-mutant mice had the same lifespan as the ku80<sup>−/−</sup> p53<sup>−/−</sup> mice. The ku70<sup>−/−</sup> p53<sup>−/−</sup> mice lived longer because they had a lower incidence of pro-B cell lymphoma that was restored with deletion of Ku80. Therefore, these experiments suggest that Ku80 has extra-NHEJ activity. In addition, we showed that free Ku70 and free Ku80 bind to apurinic/apyrimidinic (AP) sites and that free Ku70 inhibits AP endonuclease 1 [17]. Another group also showed that Ku and DNA-PKCS inhibited AP site cleavage by APE1 [21]. Thus, Ku80 and Ku70 have activity that is separable from the Ku heterodimer and NHEJ that could exacerbate the Ku80 mutant phenotype and account for the phenotypic disparity between deleting Ku80 and DNA-PKCS.

Ku80 as well as DNA-PK<sub>CS</sub> inactivation accelerated aging. These characteristics included skin atrophy, femur osteopenia, renal tubular anisokaryosis, and cortical and medullary atrophy. Accelerated aging was observed previously in Ku80 and DNA-PK<sub>CS</sub> negative mice in different genetic backgrounds and environments suggesting that early aging is not sensitive to genetic and environmental changes [10,22] [19].

Deletion of either Ku80 or DNA-PK<sub>CS</sub> also increased cancer risk. Pathological examination revealed that tumors were predominantly lymphomas. Most lymphomas were observed in the thymus. Lymphomas in non-lymphoid tissues (e.g. liver and lung) are presumably metastases.

By contrast to this report, our prior reports show that ku80<sup>−/−</sup> and ka70<sup>−/−</sup> mice exhibited low levels of cancer [10,11]. Furthermore, deleting Ku80 in a familial adenomatous polyposis mouse model (APC<sub>MIN</sub>) reduced intestinal tumors and increased life span [23]. We proposed that Ku80-deletion reduced cancer levels due to constitutive activation of the p53 DNA damage response and possibly other responses [24]. In support, deleting p53 greatly enhanced pro-B cell lymphomas and medulloblastoma in ku80<sup>−/−</sup> and ku70<sup>−/−</sup> mice [20,25,26]. Thus, difference in cancer incidences between our prior reports and this report suggest genetic background influence cancer incidence in the ku80<sup>−/−</sup> mice. This is possible since the former reports described mice derived from a 129*C57Bl6/J hybrid background while the current report describes mice derived from a C57Bl6/J*FVB F1 hybrid background.

Furthermore, we observed an increased myeloid/erythroid ratio that suggests true myeloid hyperplasia. However, complete blood counts are needed to confirm this. By contrast, we observed severe lymphoid depletion in all knock out cohorts. Lymphoid depletion is a known phenotype in SCID mice, which have a defective V(D)J recombination [27,28]. Since ku80<sup>−/−</sup> and dna-pk<sub>cs</sub><sup>−/−</sup> are SCID [9,22], it is not surprising that lymphoid depletion was found in the knock out cohorts. Therefore, this element of the phenotype is not age-related. The increased myeloid/erythroid ratio could be the result of a compensatory response to this lymphoid depletion, triggering myeloid hyperplasia and lymphoma development.

Since Ku80 and DNA-PK<sub>CS</sub> mutant mice have a defective DSB repair system, one expects accumulation of DNA damage. However, LacZ mutant frequency analysis could not confirm the previously observed increased mutant frequency in spleen [29]. Although our data do show a weak trend into the same direction,
### Table 1. Histopathology non-neoplastic lesions.

| Lesion                          | Median pathology score (min – max) | Wild type |
|---------------------------------|------------------------------------|-----------|
|                                 | ku80<sup>−/−</sup> | dna-pkcs<sup>−/−</sup> | kl-df-<sup>−/−</sup> | Male (41w) | Female (42w) | Male (45w) | Female (45w) | Male (42w) | Female (40w) | Male (37w) | Female (40w) |
| Liver anisokaryosis             | 2 (1-3) | 1.5 (1-3) | 3 (1-3) | 2* (2-3) | 2 (2-3) | 2.5 (1-3) | 1 (1-2) | 1 (1-2) |
| Liver lipofuscin                | 1 (1-1) | 1 (1-1) | 1 (1-1) | 1 (1-2) | 1 (1-1) | 1 (1-1) | 1 (1-1) | 1 (1-1) |
| Renal tubular anisokaryosis     | 2 (1-3) | 2 (1-3)* | 2 (1-3) | 2 (2-3) | 2* (2-3) | 2 (1-3) | 1 (1-2) | 1 (1-1) |
| Renal tubulonephrosis           | 3*** (1-3) | 3*** (2-4) | 3*** (1-3) | 4*** (2-4) | 2*** (1-3) | 3*** (1-3) | 0 (0-0) | 0 (0-0) |
| Epidermal atrophy              | 2** (1-3) | 2* (1-3) | 3* (2-4) | 3* (2-3) | 3** (1-4) | 3* (2-3) | 0 (0-0) | 0 (0-0) |
| Adnexal atrophy                | 1** (1-2) | 1* (0-1) | 2.5** (1-4) | 3*** (1-3) | 2* (0-4) | 2** (1-3) | 0 (0-0) | 0 (0-0) |
| Lymphoid depletion spleen       | 4* (3-4) | 4* (3-5) | 3* (3-3) | 3* (3-3) | 4* (3-5) | 3** (2-4) | 0 (0-0) | 0 (0-0) |
| Lymphoid depletion mes ln       | 4* (4-5) | 3.5** (2-5) | 2* (1-2) | 1* (1-2) | 4** (4-4) | 4** (4-4) | 0 (0-0) | 0 (0-0) |
| Tissue thickness in μm (St. dev.) |                  |                  |                  |                  |                  |                  |                  |                  |
| Dermis and epidermis           | 253.8* (56.9) | 151.3* (11.7) | 222.0** (38.3) | 172.8 (58.7) | 252.0* (99.0) | 159.0 (86.3) | 374.3 (50.6) | 247.3 (43.4) |
| Trabecular bone                | 130.8* (36.7) | 131.0* (38.2) | 124.5* (18.8) | 139.8* (25.5) | 138.4 (43.1) | 118.8** (10.6) | 1840 (19.4) | 1853 (22.4) |

4–5 animals examined per group. All selected mutant mice are at end of life, wild type mice are aged matched to mutant mice.

Mean age in weeks is shown (w).

*St. ln: mesenterial lymph node.

**<sup>−/−</sup>; <sup>dna-pk</sup><sup>−/−</sup>.

St. Dev.: Standard deviation.

<sup>p</sup>-values based on comparison with wild type animals.

* <sup>p</sup>< 0.05.

** <sup>p</sup>< 0.01.

*** <sup>p</sup>< 0.001.

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| Organ       | Tumor       | ku80<sup>-/-</sup> Male (n = 25) | Female (n = 15) | dna-pk<sub>cs</sub> <sup>-/-</sup> Male (n = 27) | Female (n = 10) | k<sup>-/-</sup>d<sup>-/-</sup> Male (n = 24) | Female (n = 21) |
|-------------|-------------|----------------------------------|-----------------|-----------------------------------------------|----------------|-----------------------------------------------|----------------|
| Liver       | lymphoma    | 9                                | 5               | 4                                             | 7              | 5                                             | 2              |
|             | adenoma     | 1                                | 1               |                                               |                |                                               |                |
|             | carcinoma   |                                  |                 |                                               |                | 3                                             | 1              |
| Lung        | lymphoma    | 1                                |                 |                                               |                | 1                                             |                |
|             | carcinoma   |                                  |                 |                                               |                |                                               |                |
| Pancreas    | lymphoma    |                                  |                 |                                               |                |                                               |                |
| Kidney      | lymphoma    | 5                                | 2               | 3                                             | 6              | 4                                             | 5              |
| Thymus      | lymphoma<sup>*</sup> | 23                            | 14              | 21                                           | 9              | 19                                           | 15             |
| ovary/uterus| lymphoma    | 1                                |                 |                                               |                |                                               |                |
| Spleen      | lymphoma    | 4                                |                 | 2                                             | 3              | 3                                             |                |
| mes. lm. nd.| lymphoma    | 3                                |                 | 5                                             | 5              | 3                                             |                |
| ax. lm. nd. | lymphoma    | 3                                |                 | 5                                             | 5              | 5                                             |                |
| Skin        | lymphoma    | 1                                |                 |                                               |                |                                               |                |
| Testis      | lymphoma    | 1                                |                 |                                               |                |                                               |                |
| bone marrow<sup>**</sup> | lymphoma | 2                                | 1               | 2                                             | 2              | 1                                             | 1              |
| pituitary gland | lymphoma | 1                                |                 |                                               |                |                                               |                |
| Tumor incidence (%) | 67 | 39 | 55 | 24 | 75 | 51 |

Mentioned n-numbers are tumor bearing animals available for microscopical examination.

k<sup>-/-</sup>d<sup>-/-</sup>; ku80<sup>-/-</sup>; dna-pk<sub>cs</sub> <sup>-/-</sup>

mes. lm. nd.: mesenterial lymph node.

ax. lm. nd.: axillary lymph node.

<sup>**</sup>—5 thymus tumor preparations were microscopically examined per group and found to be all lymphomas. All other macroscopic neoplastic lesions observed in thymus are assumed to be lymphomas as well.

<sup>**</sup> analyzed in femur.

Tumor incidence based on all animals from longevity cohort, including those not microscopically examined.

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Mutation of Ku80 is Dominant over DNA-PKcs.
Ku80. extra-NHEJ function or that free Ku70 is toxic in the absence of
observation is consistent with the possibility that Ku80 has an
deletion was more severe than DNA-PKCS –deletion. This
deletion does not further enhance these characteristics. Yet, Ku80–
strongly increased incidence of lymphomas and that simultaneous
background resulted in a accelerated aging phenotype and a
deviation. (A) Mutant frequency in liver. (B) Mutant frequency in spleen.

Figure 4. LacZ mutant frequency. Error bars represent standard
deviation. (A) Mutant frequency in liver. (B) Mutant frequency in spleen.
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variation is too large and effect sizes too small to achieve statistical
significance. Previously we had to separate the small mutations
from the big mutations before a difference could be seen since the
former was reduced while the latter was increased.

In conclusion, we have shown that deletion of the NHEJ
components Ku80 and DNA-PKCS in a C57Bl6/J*FVB F1 hybrid
background resulted in a accelerated aging phenotype and a
strongly increased incidence of lymphomas and that simultaneous
deletion does not further enhance these characteristics. Yet, Ku80–
deletion was more severe than DNA-PKCS –deletion. This
observation is consistent with the possibility that Ku80 has an
extra-NHEJ function or that free Ku70 is toxic in the absence of
Ku80.

Materials and Methods

Ethics Statement

All animal work was approved by the ethics committee of the
National Institutes for Public Health and the Environment (RIVM), Antonie van Leeuwenhoeklaan, Bilthoven, The Netherlands, IACUC protocol #99047x.

Mice breeding

Ku80 null mice are not viable in a pure C57BL6/J background
[30]. Therefore, we generated F1 hybrid animals with a C57Bl6/
J*FVB background. This resulted in viable offspring and
prevented C57BL6/J related ulcerative dermatitis [31]. ku80–/–
mice [9] and dna-pkcs–/– mice [32] were imported in our SPF
facility, rederived and back crossed to C57Bl6/Jco (Charles
River, France) and FVB/NHanHsd (Harlan, Germany) using a
speed congenics approach [33]. Knock out mice in C57Bl6/Jco
background were maintained by backcrossing with C57Bl6/J-
pUR288 (LacZ locus integrated at chromosome 13 [34])
nerating heterozygous knock out animals which are homozygous
for pUR288. Knock out mice on FVB background were
maintained by backcrossing with pure FVB wild type animals.

Cohorts with single and double knock out animals were generated
using double heterozygous knock out breeders (ku80–/–; dna-pkcs–/–)

All male breeders were on C57Bl6/J-pUR288 and all female
breeders on FVB background. Using this breeding strategy, all
genotypes could be generated from the same breeding colony and
all experimental mice were C57Bl6/J*FVB F1 hybrids carrying
one LacZ allele. The breeding scheme is depicted in figure 1.

This study was carried out in strict accordance with institutional
guidelines and regulations. All animal work was approved by the
ethics committee of the National Institutes for Public Health and the
Environment (RIVM), Antonie van Leeuwenhoeklaan, Bilthoven, The Netherlands, IACUC protocol #99047x. These
were survival studies; therefore, mice were monitored every day
without intervention. Moribund mice were sacrificed with
ketamine/xylazine anesthesia followed by cervical dislocation
and all efforts were made to minimize suffering and discomfort.
Criteria for moribund were >15% weight loss within 2 weeks, not
responsive to touch, prominent appearance of ribs, spine and hips,
hunch body position, matted fur, or a visible tumor.

Study setup

Each cohort consisted of 45 males and 45 females. All animals
were maintained under specific pathogen free conditions and were
fed ad libitum using CRM pelleted maintenance diet (Special Diet
Services, UK). A 12 hr/12 hr dark/light cycle was maintained
and temperature was 20°C. Animals were maintained until death
or moribund. Moribund animals were euthanized by exsanguination.
and major organs were isolated and partly stored in
formaldehyde and partly snap frozen in liquid nitrogen. Results
from ku80–/– and dna-pkcs–/– longevity cohorts were also used in
a study comparing Ku70 and Ku80 function [17]. In addition, 5
male animals of all genotypes were euthanized at approximately
median lifespan for LacZ mutant frequency analysis.

Pathology

Pathology lesions in mutant animals were compared with those
in age matched wild type littermates. Representative sections from
the liver, kidney, thymus, spleen, axillary and mesenteric lymph
nodes, femur, vertebra, uterus, ovaries, testes, and accessory male
genital glands were processed (n = 4–5 per genotype), stained with
Hematoxylin and Eosin, and microscopically examined for the
presence of histopathologic lesions. All tumors macroscopically
identified during necropsy were also prepared for histopathologic
evaluation. Severity score of all recorded lesions was semi-
quantitatively assessed. Scores were given as absent (0), subtle (1),
mild (2), moderate (3), severe (4), and massive (5) for each criteria.

Digital images from the femur cortical bone at mid-shaft area, and
skin were taken for morphometric analysis using Labsense image
analysis software (Olympus). The thickness of femur cortical bone
thickness, and skin thickness (dermis and epidermis with exclusion of subcutaneous fat) were measured using arbitrary line option.

**CD3 staining**

After deparaffinization, rehydration and citrate buffer pre-treatment, sections were incubated with rabbit polyclonal antibody to the human CD3 molecule (DAKO Corporation- Code-Nr. A 0452), diluted 1:250 in 10% normal goat serum, for 30 minutes at room temperature. The remainder of the procedure was accomplished according to the manufacturer’s instructions. Normal goat serum was substituted for the primary antibody as a negative control. Mouse lymphoid tissue was used as a positive control.

**LacZ mutant frequency analysis**

LacZ mutant frequency was assessed in liver and spleen using the pUR208(lacZ) reporter. This assay has been described previously [34]. In short, total DNA was extracted from liver and spleen using a phenol/chloroform/isomyl alcohol extraction (25:24:1) and digested with the endonuclease HindIII in the presence of magnetic beads, coated with lacI-lacZ fusion protein. After digestion, magnetic beads were washed and DNA fragments eluted using isopropyl-L-thio-B-D-galactopyranoside. DNA fragments were circularized with T4 DNA ligase and transformed into E. Coli (lacZ, galE-) bacteria. Of the transformed bacteria, one thousandth was plated on X-gal plates and the remainder on selective p-gal plates. The mutant frequency was defined as the amount of colonies on the selective plate divided by the amount of colonies on the X-gal plate multiplied by 1000 (dilution factor).

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