Variation of Macro- and Microelements, and Trace Metals in Spring Wheat Genetic Resources in Siberia

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Abstract: Western Siberia is one of the major spring wheat regions of Russia, cultivating over 7 Mha. The objective of the study was to evaluate the variation of macro- and microelements, and of trace metals in four distinct groups of genetic resources: primary synthetics from CIMMYT (37 entries), primary synthetics from Japan (8), US hard red spring wheat cultivars (14), and material from the Kazakhstan–Siberian Network on Spring Wheat Improvement (KASIB) (74). The experiment was conducted at Omsk State Agrarian University, using a random complete block design with four replicates in 2017 and 2018. Concentrations of 15 elements were included in the analysis: macroelements, Ca, K, Mg, P, and S; microelements, Fe, Cu, Mn, and Zn; toxic trace elements, Cd, Co, Ni; and trace elements, Mo, Rb, and Sr. Protein content was found to be positively correlated with the concentrations of 11 of the elements in one or both years. Multiple regression was used to adjust the concentration of each element, based on significant correlations with agronomic traits and macroelements. All 15 elements were evaluated for their suitability for genetic enhancement, considering phenotypic variation, their share of the genetic component in this variation, as well as the dependence of the element concentration on other traits. Three trace elements (Sr, Mo, and Co) were identified as traits that were relatively easy to enhance genetically. The CIMMYT and Japanese synthetics had significantly higher concentrations of K and Sr, compared to the local check. The Japanese synthetics also had the highest concentrations of Ca, S, Cd, and Mo. The US cultivars had concentrations of Ca as high as the Japanese synthetics, and the highest concentrations of Mg and Fe. KASIB’s germplasm had near-average values for most elements. Superior germplasm, with high macro- and microelement concentrations and low trace-element concentrations, was found in all groups of material included.

Keywords: biofortification; cereals; nutritional quality; protein concentration

1. Introduction

Spring wheat is an important crop in Russia, with an annual cropping area of 11–12 Mha. The main spring wheat-production belt stretches from the mid-Volga region across the southern Ural mountains to Western Siberia. This is a short-season crop grown from May...
to August in an extensive, rainfed cropping system dominated by cereals, and occasionally rotated with oilseed and legume crops. Morgounov et al. [1] compared the effects of climate change on spring wheat production in Eurasia (Russia and Kazakhstan) and North America. The grain yields and production gains in North America were almost twice that in Eurasia, owing to better environmental conditions, including precipitation, as well as application of more advanced and diverse production technologies based on conservation agriculture. Wheat produced in Russia is traded both regionally and internationally. According to the FAO (www.fao.org/faostat, accessed on 1 January 2019), Russia exported 43.9 Mt of wheat grain in 2018. Therefore, grain quality, including health benefits or hazards, is important for global food security and safety.

The ionome has been defined as the mineral element composition of an organism, and represents the inorganic component of cellular and organismal systems [2]. Therefore, ionomics involves the quantitative measurement of the element composition of living organisms, and of changes in this composition in response to environment, growth stage, and genotype. Minerals comprising wheat grain can be divided into three groups: macroelements (Ca, K, Mg, P, Cl, and S) that are important for starch and protein formation; toxic heavy metals (arsenic (As), cadmium (Cd), chromium (Cr), and lead (Pb)), normally regulated not to exceed certain concentrations; and microelements essential for plants and humans (selenium (Se), boron (B), Mn, Cu, Fe, Mo, and Zn), which can also be harmful when exceeding certain concentrations. All five microelements, along with Ca and iodine, were selected as candidates for biofortification to improve nutritional value of crops, including wheat [3].

Wheat biofortification has been successfully applied for increasing Zn content in grain with commercial cultivars being grown in India and Pakistan [4,5]. A nutrition study with preschool children and women showed that consumption of high Zn-biofortified wheat prevented morbidity [6]. The concept of biofortification has not been applied in wheat breeding in Russia, and only a few studies have assessed the mineral composition of wheat cultivars grown in Russia. Bityutskii et al. [7] studied around 30 registered wheat cultivars and found variation in the microelement concentrations of grains (µg/g), Fe 15–22, Zn 14–21, and Mn 2.4–4.1. Morgounov et al. [8] determined 15 macro- and microelement concentrations in 49 genotypes grown across six sites in Russia and Kazakhstan in two years. Several cultivars (Element-22, Lutescens-3-04-21-11, and Silach) were identified as having high grain yields, relatively high protein content, and high concentrations of P, S, Mn, Cu, and Zn, singly or in combination. The above studies suggest that it could be possible to develop a varietal development program for improving mineral nutrients. In particular, the presence of multiple elements in combination in the same genotype indicates a positive association among them. A positive correlation between two elements could be exploited in a breeding program more efficiently, and could simultaneously improve them, using a selection index [9].

The success of wheat biofortification at CIMMYT was largely based on the utilization of genetic resources with high concentrations of nutritionally valuable elements [10]. Significant genetic variation for grain Zn and Fe was found in landraces and ancestors of common wheat, such as Aegilops tauschii, Triticum turgidum ssp. dicoccoides, T. turgidum ssp. Dicoccum, and T. aestivum ssp. spelta. Einkorn (Triticum monococcum) was later added to this list [11]. This variation has been introduced into high-yielding germplasm through conventional breeding and marker-assisted breeding [5].

Western Siberia (Kurgan, Tuymen, Omsk, Novosibirsk, and Altai regions) grows around 7 Mha of spring wheat with an average grain yield of 2 t/ha. Drought is the most common abiotic stress, and leaf and stem rust are the main diseases affecting the crop. Spring wheat breeding programs in the region are united through the Kazakhstan–Siberian Network on Spring Wheat Improvement (KASIB). This network was established in 2000 with the objective of exchanging germplasm and conducting cooperative multilocational testing to characterize advanced breeding lines and new cultivars. The majority of the cultivars grown in the region represent tall, daylength-sensitive material with good drought
tolerance and suitable bread-making quality [12]. The studies of KASIB germplasm demonstrated limited genetic diversity for resistance to stem [13] and leaf rust [14]. Overall, cultivars grown in the region have similar genetic makeup and high phenotypic similarity.

In order to expand spring wheat’s diversity, a panel of genetic resources (OMON-GAI (Omsk Observation Nursery–Genetically Associated Improvement)) was assembled at Omsk State Agrarian University, the KASIB network coordinator for Russia. The panel included primary synthetics from CIMMYT and Japan, cultivars from USA, Omsk cultivars developed by the university, and Omsk Agrarian Research Center and the KASIB network’s germplasm. The panel was phenotyped for common agronomic traits in 2017–2020 and genotyped using the genotyping-by-sequencing method, resulting in over 46,000 SNPs (single nucleotide polymorphism). The genetic diversity study clearly separated all material into three groups: CIMMYT synthetics, Japanese synthetics, and a combined group of bread wheat germplasm from KASIB and the USA [15]. A genome-wide association study (GWAS) was conducted on yield and 26 yield-related traits, disease resistance, and grain quality traits [16]. The study identified 243 significant marker–trait associations for 35 traits that explained up to 25% of the phenotypic variance, with the most significant of these having already been used in the marker-assisted breeding at the university.

Ionome phenotyping of the OMON-GAI panel for 23 elements was performed using grain from the 2017 and 2018 seasons. The objective of the study was to evaluate the variation of macro- and microelements, and trace metals in the different groups of genetic resources, identify the relationship between the agronomic and quality traits and element concentrations, select superior genotypes, and develop approaches to be used for breeding to enhance the element composition of wheat cultivars.

2. Results
2.1. Variation for Agronomic Traits

The analysis of variance of the number of days to heading, TKW, grain yield, and protein content data demonstrated the high significance of genotypes, years, and their interaction, except for the effect of years on TKW (Table S1). The weather conditions during the spring wheat growing season in 2017 were characterized by air temperatures close to the long-term average (17.3 °C in May–August versus 17.0 °C) and a moderate moisture stress, with precipitation of 163 mm during May–August, compared to the 238 mm long-term average. The 2018 growing season was cooler (15.2 °C in May–August) with 270 mm of rainfall. Leaf and stem rust affected susceptible genotypes with up to 30–40% severity in 2017 and up to 60–80% in 2018. These pathogens certainly affected the grain yield. The average yield of the whole panel was 320 g/m² in 2017 and 395 g/m² in 2018.

The relative performance of the different groups of genetic resources across the two years is presented in Figure 1 and for individual years in Table S2. The number of days to heading varied from 35 (USA group) to 47 days (Japanese synthetics). The differences between the three KASIB groups were 36.3 days for the early group, 39.5 for the intermediate, and 42.5 for the late group. The highest grain yield was recorded for the three KASIB groups (440–471 g/m²), followed by the US cultivars (320 g/m²), the CIMMYT synthetics (236 g/m²), and the Japanese synthetics (104 g/m²). The early maturing check Pamyati Azieva demonstrated a grain yield of 399 g/m², and the intermediate-maturing check Serebristaya, 470 g/m². TKW was in a range of 44.0–45.7 g for all groups, though the US cultivars had smaller grain (36.9 g). The highest protein content was recorded for the Japan synthetics (20%), followed by the US cultivars (17.9%), the CIMMYT synthetics (16.7%), and the KASIB groups (15.9–16.4%). Overall, the research panel used in the study was highly heterogeneous and contrasting, especially for vegetative period and grain yield.
2.2. Adjustment of Grain Element Concentration

The correlation analysis was conducted to find the relationship between the concentrations of the 15 elements and agronomic traits (grain yield, protein content, and TKW) (Table 1). The analysis was performed separately for each year. Protein content was found to be positively correlated with the concentrations of 11 of the elements in one or both years. Only Ca, K, Co, and Rb were not correlated with protein content. However, the correlation coefficients between protein content and the elements ranged from 0.2 to 0.4, indicating weak relationships. Mg concentration was significantly correlated with 10 of the elements, with correlation coefficients exceeding 0.6 for P, S, Cu, and Mn. P and S concentrations were also positively correlated with microelements (Cu, Mn, and Zn) and toxic trace elements (Cd, Ni, and Mo). Among the macroelements, variation in Ca and K concentrations was the least correlated with any agronomic trait or other element. Co, Rb, and Fe (2018) concentrations were not correlated with any trait.

Table 1. Correlation coefficients between agronomic traits and element concentrations in grain, 2017–2018.

| Element | Year  | Yield | PC    | TKW   | Ca   | K    | Mg   | P    | S     |
|---------|-------|-------|-------|-------|------|------|------|------|-------|
| Ca      | 2017  | −0.11 | 0.06  | −0.14 | −    | 0.16 | 0.15 | 0.01 | 0.09  |
|         | 2018  | −0.14 | 0.06  | −0.15 | −    | −0.03| 0.22 *| 0.03 | 0.11  |
| K       | 2017  | −0.13 | 0.03  | 0.10  | 0.16 | −    | −0.07| −0.07| 0.30 *|
|         | 2018  | −0.19 | 0.16  | 0.17  | −0.03| −    | 0.07 | 0.07 | 0.09  |
| Mg      | 2017  | −0.24 *| 0.43 *| −0.07 | 0.15 | −0.07| −    | 0.79 *| 0.60 *|
|         | 2018  | −0.19 | 0.29 *| −0.06 | 0.22 *| 0.07 | −    | 0.76 *| 0.55 *|
| P       | 2017  | −0.14 | 0.32 *| 0.08  | 0.01 | 0.20 *| 0.79 *| −    | 0.68 *|
|         | 2018  | −0.22 *| 0.30 *| 0.05  | 0.03 | 0.48 *| 0.76 *| −    | 0.20 *|
All significant correlations presented in Table 1 were used for adjustment of element concentrations through multiple regression. The basic statistical parameters of the original and adjusted values are presented in Table 2. The adjusted data had the same means, but the differences between the minimum and maximum values were reduced. This resulted in the reduction of the coefficient of variation (CV) for all elements by 0.2–6.4%. A factorial ANOVA (genotype x year) was performed for each element, using original and adjusted data (Table S2). The level of significance of the main factors (genotype and year), using original and adjusted values, was identical. However, for five of the elements (Ca, Mg, P, S, and Cu) the interaction of genotypes x year was not significant \((p > 0.05)\) using the original values, although it was significant when adjusted values were used in the ANOVA. This indicates that adjustment contributed to the higher capacity to distinguish significance of these important interactions. Overall, the adjustment of element concentrations using multiple regression was well justified and provided a more balanced approach for comparison of genetic resources and individual genotypes.

The broad sense \(H^2\) was calculated for each year using original and adjusted values (Table 2). The adjustment slightly increased \(H^2\) for Mg and P and decreased for S and Cu, in both years. The average \(H^2\) across all elements was 0.59 for the original data and 0.58 for the adjusted data. The \(H^2\) calculated based on a factorial ANOVA was similar for K, Mg, P, Cu, Mn, Ni, and Mo for original and adjusted values. Slightly different \(H^2\) estimates were found for Ca (0.73 for the original data vs. 0.62 for the adjusted), S (0.81 vs. 0.62), Fe (0.58 vs. 0.47), Zn (0.27 vs. 0.37), Cd (0.76 vs. 0.67), and Sr (0.87 vs. 0.77).

| Element | Year | Yield | PC   | TKW  | Ca   | K    | Mg   | P    | S    |
|---------|------|-------|------|------|------|------|------|------|------|
| S       | 2017 | −0.23 * | 0.45 * | 0.13 | 0.09 | 0.30 * | 0.60 * | 0.68 * | -    |
|         | 2018 | −0.34 * | 0.47 * | 0.12 | 0.11 | 0.09 | 0.55 * | 0.20 * | -    |
| Cu      | 2017 | −0.14 | 0.28 * | −0.02 | −0.03 | 0.10 | 0.62 * | 0.68 * | 0.54 * |
|         | 2018 | −0.21 * | 0.25 * | 0.01 | 0.08 | 0.21 * | 0.60 * | 0.60 * | 0.47 * |
| Fe      | 2017 | −0.10 | 0.23 * | 0.29 * | −0.01 | −0.14 | 0.23 * | −0.06 | −0.09 |
|         | 2018 | −0.13 | 0.17 | −0.14 | −0.03 | −0.12 | 0.05 | −0.04 | −0.10 |
| Mn      | 2017 | −0.13 | 0.35 * | −0.02 | 0.11 | −0.10 | 0.68 * | 0.57 * | 0.49 * |
|         | 2018 | −0.09 | 0.05 | 0.09 | 0.22 * | −0.10 | 0.63 * | 0.46 * | 0.38 * |
| Zn      | 2017 | −0.03 | 0.20 * | 0.04 | −0.08 | 0.02 | 0.53 * | 0.73 * | 0.48 * |
|         | 2018 | −0.30 * | 0.26 * | 0.02 | 0.09 | 0.27 * | 0.49 * | 0.63 * | 0.50 * |
| Cd      | 2017 | −0.20 * | 0.29 * | 0.01 | 0.00 | −0.02 | 0.46 * | 0.44 * | 0.49 * |
|         | 2018 | −0.22 * | 0.38 * | 0.10 | 0.09 | 0.05 | 0.37 * | 0.32 * | 0.40 * |
| Co      | 2017 | 0.01 | 0.07 | −0.19 | 0.12 | 0.14 | 0.17 | 0.14 | 0.04 |
|         | 2018 | 0.01 | 0.05 | 0.04 | 0.19 | 0.12 | 0.11 | 0.15 | 0.15 |
| Ni      | 2017 | −0.08 | 0.15 | −0.01 | −0.01 | 0.09 | 0.25 * | 0.25 * | 0.23 * |
|         | 2018 | −0.24 * | 0.36 * | 0.01 | −0.03 | 0.14 | 0.23 * | 0.28 * | 0.31 * |
| Mo      | 2017 | −0.07 | 0.20 * | 0.05 | 0.01 | 0.17 | 0.12 | 0.20 * | 0.24 * |
|         | 2018 | −0.02 | 0.15 | 0.18 | 0.05 | 0.27 * | 0.21 * | 0.33 * | 0.26 * |
| Rb      | 2017 | −0.05 | 0.06 | 0.16 | −0.02 | 0.12 | 0.04 | 0.08 | 0.07 |
|         | 2018 | −0.18 | 0.12 | 0.01 | 0.07 | 0.18 | −0.01 | 0.00 | 0.16 |
| Sr      | 2017 | −0.34 * | 0.22 * | −0.08 | 0.60 * | 0.11 | 0.17 | −0.00 | 0.16 |
|         | 2018 | −0.34 * | 0.31 * | −0.22 * | 0.65 * | 0.04 | 0.17 | 0.04 | 0.16 |

Number of significant correlations 11 18 3 4 6 16 14 15

*—significant at \(p > 0.05\).
| Element-Year | Data       | Adjustment Variables | Mean  | Min  | Max  | CV, % | H² (Original–Adjusted) | r |  |
|--------------|------------|----------------------|-------|------|------|-------|-------------------------|--|--|
|              |            |                      | Macrolelements |      |      |      |       |                         |   |  |
| Ca-2017      | Original   | -                    | 362   | 237 | 518  | 15.1  | 0.59                    | - |  |
| Ca-2018      | Original   | -                    | 397   | 266 | 531  | 13.8  | 0.49                    | 0.98 |  |
|              | Adjusted   | Mg                   | 266   | 273 | 523  | 13.6  | 0.50                    | - |  |
| K-2017       | Original   | -                    | 3652  | 2654| 5453 | 10.2  | 0.67                    | 0.99 |  |
| K-2018       | Original   | -                    | 3642  | 2298| 4992 | 15.7  | 0.79                    | - |  |
| Mg-2017      | Original   | -                    | 1208  | 922 | 1532 | 9.1   | 0.60                    | 0.56 |  |
|              | Adjusted   | YLD, PC, P, S        | 922   | 997 | 1400 | 5.4   | 0.62                    | - |  |
| Mg-2018      | Original   | -                    | 1225  | 926 | 1524 | 8.5   | 0.58                    | 0.66 |  |
|              | Adjusted   | PC, Ca, P, S         | 926   | 1007| 1461 | 5.6   | 0.69                    | - |  |
| P-2017       | Original   | -                    | 5180  | 3906| 6701 | 10.2  | 0.43                    | 0.54 |  |
| P-2018       | Original   | -                    | 4699  | 3428| 6993 | 12.3  | 0.57                    | 0.48 |  |
| S-2017       | Original   | -                    | 2059  | 1710| 2682 | 9.5   | 0.69                    | 0.60 |  |
| S-2018       | Original   | -                    | 2050  | 1762| 2433 | 6.1   | 0.66                    | 0.64 |  |
| Microelements |            |                      |       |     |      |       |                         |   |  |
| Cu-2017      | Original   | -                    | 4.66  | 3.09| 7.08 | 13.8  | 0.54                    | 0.76 |  |
|              | Adjusted   | PC, Mg, P, S         | 3.09  | 3.53| 6.29 | 10.4  | 0.63                    | - |  |
| Cu-2018      | Original   | -                    | 3.72  | 2.30| 6.06 | 15.3  | 0.57                    | 0.74 |  |
|              | Adjusted   | YLD, PC, Mg, P, S    | 2.30  | 2.67| 5.22 | 11.3  | 0.55                    | - |  |
| Fe-2017      | Original   | -                    | 37.7  | 21.1| 53.6 | 14.2  | 0.50                    | 0.90 |  |
|              | Adjusted   | PC, Mg               | 21.1  | 21.4| 51.8 | 12.9  | 0.40                    | - |  |
| Fe-2018      | Original   | -                    | 35.8  | 22.5| 47.2 | 12.9  | 0.25                    | - |  |
| Mn-2017      | Original   | -                    | 43.1  | 31.9| 60.0 | 11.2  | 0.58                    | 0.72 |  |
|              | Adjusted   | PC, Mg, P, S         | 31.9  | 31.0| 52.4 | 8.2   | 0.58                    | - |  |
| Mn-2018      | Original   | -                    | 44.8  | 31.9| 59.1 | 11.7  | 0.63                    | 0.81 |  |
|              | Adjusted   | Mg, P, S             | 31.9  | 32.6| 52.4 | 8.2   | 0.70                    | - |  |
| Zn-2017      | Original   | -                    | 53.3  | 31.1| 77.7 | 15.6  | 0.16                    | 0.74 |  |
|              | Adjusted   | PC, Mg, P, S         | 31.1  | 40.0| 69.3 | 10.9  | 0.27                    | - |  |
| Zn-2018      | Original   | -                    | 41.8  | 23.9| 76.3 | 19.1  | 0.42                    | 0.67 |  |
|              | Adjusted   | PC, K, Mg, P, S      | 23.9  | 26.4| 62.0 | 13.1  | 0.39                    | - |  |
| Toxic trace elements |            |                      |       |     |      |       |                         |   |  |
| Cd-2017      | Original   | -                    | 0.044 | 0.019| 0.117| 35.8  | 0.66                    | 0.75 |  |
|              | Adjusted   | YLD, PC, Mg, P, S    | 0.019 | 0.024| 0.099| 27.7  | 0.57                    | - |  |
| Cd-2018      | Original   | -                    | 0.031 | 0.011| 0.088| 35.6  | 0.68                    | 0.77 |  |
|              | Adjusted   | YLD, PC, Mg, P, S    | 0.011 | 0.013| 0.071| 27.7  | 0.55                    | - |  |
| Co-2017      | Original   | -                    | 0.010 | 0.006| 0.021| 27.0  | 0.72                    | - |  |
| Co-2018      | Original   | -                    | 0.009 | 0.006| 0.017| 17.1  | 0.27                    | - |  |
| Ni-2017      | Original   | -                    | 0.212 | 0.113| 0.356| 23.6  | 0.44                    | 0.94 |  |
|              | Adjusted   | Mg, P, S             | 0.113 | 0.113| 0.360| 22.4  | 0.44                    | - |  |
Table 2. Cont.

| Element-Year | Data  | Adjustment Variables | Mean   | Min   | Max   | CV, % | H² | r (Original–Adjusted) |
|--------------|-------|----------------------|--------|-------|-------|-------|----|----------------------|
| Ni-2018      | Original | -                     | 0.148  | 0.078 | 0.312 | 26.6  | 0.57 | 0.85                 |
|              | Adjusted | YLD, PC, Mg, P, S    |        | 0.075 | 0.224 | 22.6  | 0.47 |                     |
| Trace elements|       |                       |        |       |       |       |     |                     |
| Mo-2017      | Original | -                     | 0.347  | 0.209 | 0.579 | 22.4  | 0.71 | 0.94                 |
|              | Adjusted | PC, P, S              | 0.208  | 0.195 | 0.592 | 21.4  | 0.74 |                     |
| Mo-2018      | Original | -                     | 0.308  | 0.183 | 0.554 | 22.6  | 0.69 | 0.91                 |
|              | Adjusted | K, Mg, P, S           | 0.195  | 0.183 | 0.592 | 24.0  | 0.78 |                     |
| Rb-2017      | Original | -                     | 4.35   | 2.29  | 6.75  | 21.7  | 0.25 | -                    |
| Rb-2018      | Original | -                     | 3.51   | 2.20  | 5.34  | 18.9  | 0.25 | -                    |
| Sr-2017      | Original | -                     | 2.09   | 0.914 | 3.627 | 30.3  | 0.81 | 0.91                 |
|              | Adjusted | YLD, PC, TKW          |        | 0.909 | 3.628 | 27.7  | 0.77 |                     |
| Sr-2018      | Original | -                     | 2.42   | 1.175 | 4.307 | 27.2  | 0.75 | 0.82                 |
|              | Adjusted | YLD, PC, TKW          | 1.366  | 4.053 | 22.2  | 0.63  |     |                     |

1 YLD, yield; PC, protein content; TKW, 1000 kernel weight.

2.3. Variation for Element Concentrations

Phosphorus had the highest concentration of all the elements in wheat grain, at 5180 and 4699 µg/g in 2017 and 2018, respectively (Table 2). The other macroelement concentrations (µg/g) in decreasing order were K (3642–3652), followed by S (2050–2059), Mg (1208–1225), and Ca (362–397). The variation between years was limited for K, S, and Mg, but exceeded 10% for P and Ca. The ranking of the macroelements for grain concentration coefficient of phenotypic variation, based on the original values, was Ca (14.4%), followed by K (12.9%), P (11.3%), S (9.5%), and Mg (8.8%). Among the microelements, Zn had the highest concentration (µg/g) in the grain (48.8–53.3 over the two years), followed by Mn (43.1–44.8), Fe (35.8–37.7), and Cu (3.72–4.66). The ranking of the microelements for grain concentration coefficient of phenotypic variation, based on the original values, was Zn (17.3%) followed by Cu (14.5%), Fe (13.6%), and Mn (11.5%).

Among the three toxic trace elements, Ni had the highest concentrations, at 0.212 and 0.148 µg/g in 2017 and 2018, respectively. Ni was also a highly variable element, both within and between years, with CV exceeding 23.6%. Cd and Co had only low concentrations (<0.044 µg/g) that were highly variable (CV 17.1–35.8%). The three remaining trace elements (Mo, Rb, and Sr) were also characterized by low concentrations (0.31–4.35 µg/g) and high variability (CV 18.9–30.3%).

2.4. Elements’ Suitability for Genetic Enhancement

The progress for wheat genetic enhancement to increase or decrease the concentration of a certain element depends on several factors: the degree of phenotypic variation within the germplasm, the share of the genetic component in this variation, as well as the dependence of the element concentration on other traits. Table 3 summarizes three main criteria characterizing suitability of elements for genetic enhancement. The coefficient of phenotypic variation varied from 8.8% (Mg) to 35.7% (Cd). It is assumed that higher variation provides the opportunity for selection. Therefore, the element with the highest variation was ranked 1 and the lowest, 15. H² calculated based on a factorial ANOVA (genotype x year) of the original data varied from 0.27 (Zn) to 0.87 (Sr). Similar to CV, the highest value of H² was ranked 1 and the lowest, 15. The ideal breeding trait may have a variation independent of other traits.
Table 3. Parameters characterizing elements’ suitability for genetic enhancement.

| Element | Coefficient of Phenotypic Variation for 2017–2018 Means | Number of Significant Correlations with Agronomic Traits and Other Elements | H² Based on ANOVA of Original Data | Overall Sum of Ranks |
|---------|--------------------------------------------------------|--------------------------------------------------------------------------------|----------------------------------|---------------------|
|         | % | Rank | Value | Rank | Value | Rank | Value | Rank |
| Ca      | 14.5 | 9 | 1 | 3.5 | 0.73 | 5 | 17.5 | 4 |
| K       | 13.0 | 11 | 1 | 3.5 | 0.61 | 8 | 22.5 | 7 |
| Mg      | 8.8 | 15 | 8 | 9.5 | 0.67 | 6 | 30.5 | 12 |
| P       | 11.3 | 13 | 9 | 12.5 | 0.60 | 10 | 34.5 | 14 |
| S       | 9.6 | 14 | 9 | 12.5 | 0.81 | 3 | 28.5 | 11 |
| Cu      | 14.6 | 8 | 10 | 11 | 0.61 | 9 | 28.0 | 10 |
| Fe      | 13.6 | 10 | 2 | 5 | 0.58 | 11 | 26.0 | 9 |
| Mn      | 11.5 | 12 | 7 | 7 | 0.55 | 12 | 31.0 | 13 |
| Zn      | 17.4 | 7 | 10 | 14 | 0.27 | 15 | 36.0 | 15 |
| Cd      | 35.7 | 1 | 10 | 14 | 0.76 | 4 | 19.0 | 5 |
| Co      | 22.1 | 5 | 0 | 1.5 | 0.65 | 7 | 13.5 | 3 |
| Ni      | 25.1 | 3 | 8 | 9.5 | 0.49 | 13 | 25.5 | 8 |
| Mo      | 23.2 | 4 | 7 | 7 | 0.84 | 2 | 13.0 | 2 |
| Rb      | 20.3 | 6 | 0 | 1.5 | 0.48 | 14 | 21.5 | 6 |
| Sr      | 28.8 | 2 | 7 | 7 | 0.87 | 1 | 10.0 | 1 |

For each element, Table 3 provides the number of significant correlation coefficients with three agronomic traits (grain yield, protein content, and TKW) and macroelements, based on the data in Table 1. The concentrations of Co and Rb did not correlate with any trait or element. They were ranked the highest (rank 1.5), while Cu, Zn, and Cd had 10 significant correlations, being ranked the lowest (rank 14). The last column in Table 3 presents the sum of three ranks, which were again ranked from lowest (better suited for genetic enhancement) to highest. Three trace elements (Sr, Mo, and Co) were the highest ranked, representing breeding traits that are relatively easy to improve through breeding. These were followed by Ca, Cd, Rb, and K. The important biofortification elements Fe and Cu were ranked 9 and 10, respectively. Mn and Zn were among the lowest ranked, representing traits that are difficult to improve genetically.

2.5. Genetic Resources Characterization for Grain Ionome

The original and adjusted average concentrations across 2017 and 2018 for different groups of genetic resources for the macroelements Fe, Zn, and Cd, are presented in Figure 2, and for all elements for each year independently in Table S3. There is a clear difference between the original and adjusted values. For Mg, P, S, and Zn, the Japanese synthetics were clearly superior when the original data were used. However, the adjusted means demonstrated different performance, comparable to other groups of genetic resources.
Taking into account the adjusted values, the CIMMYT and Japanese synthetics had significantly higher concentrations of K (9.2–10.1% higher) and Sr (9.4–16.4% higher), compared to local Check-1 (Pamyati Azieva). The Japanese synthetics also had the highest concentrations of Ca (4.9% higher, compared to local Check-1), S (3.7%), Cd (38.1%), and Mo (4.7%). The US cultivars had concentrations of Ca as high as the Japanese synthetics, and the highest concentrations of Mg (6.9% higher than Check-1) and Fe (5.4% higher). This group was also characterized by low concentrations of K (13.8% lower, compared
to Check-1) and Mo 11.8% lower. The KASIB germplasm had near-average values for most elements. However, there were differences between the early, intermediate, and late maturity groups. For Mg and Fe, there were decreasing concentrations from early to intermediate to late material of 1.5–2%. For S and Zn, concentrations increased in the later-maturing germplasm, also by 1.5–2%.

2.6. Superior Germplasm

All of the germplasm was ranked for concentration of each element, based on mean adjusted values for 2017–2018 (Table S4). Superior genotype performance was defined as being in the top 15 entries, based on the highest concentration of macro- and microelements. For the trace elements, including toxic metals, favorable performance was considered as being in the bottom 15 entries, based on the lowest concentrations. The germplasm was also ranked for grain yield and protein content. Table 4 presents the 10 highest-yielding genotypes and 20 entries with favorable concentrations for at least three elements. The five highest-yielding genotypes had high concentrations of Ca, including cv. Silach, with a high concentration of Mg and low Rb; Element 22, with high concentrations of P and S, but low Ni; and breeding line Lutestsens 1296, with high K and low Cd concentrations. The second-highest yielding genotype Lutestsens 15-14 was characterized by high concentrations of the essential microelements Mn and Zn. As expected, all top-yielding genotypes belonged to the KASIB group of germplasm.

Table 4. Genotypes with the highest grain yield, highest concentrations of macro- and microelements, and lowest concentrations of trace elements.

| Entry | Genotype            | Group * | Highest/Lowest Elements | Grain Yield g/m² | Protein Content % | Rank |
|-------|---------------------|---------|-------------------------|------------------|------------------|------|
| -     | Pamyati Azieva (Check-1) | -       | -                       | 399              | -                | 16.5 |
| -     | Serebristaya (Check-2) | -       | -                       | 471              | -                | 14.9 |
| 151   | Lutestsens KS 963    | K-E     | Ca, Mg                  | 572              | 1                | 17.2 |
| 112   | Lutestsens 15-14     | K-L     | Mn, Zn                  | 557              | 2                | 16.5 |
| 91    | Lutestsens 7-04-4    | K-I     | -                       | 541              | 3                | 17.5 |
| 164   | Silach               | K-L     | Ca, Mg, Rb              | 541              | 4                | 16.7 |
| 94    | Element 22           | K-L     | Ca, P, S, Ni            | 535              | 5                | 17.0 |
| 152   | Lutestsens 1296      | K-I     | Ca, K, Cd               | 521              | 6                | 15.6 |
| 157   | OmGAU-100            | K-L     | Ca, P, Cu               | 518              | 7                | 16.4 |
| 116   | Uralosibirskaya      | K-I     | -                       | 515              | 8                | 17.6 |
| 85    | Lutescens 310-00-1   | K-I     | P, Rb                   | 514              | 9                | 17.5 |
| 89    | Aestivum 947         | K-I     | Ni                      | 514              | 10               | 15.8 |
| 156   | Novosibirskaya 41    | K-E     | P, Cu, Cd, Ni           | 482              | 23               | 19.0 |
| 143   | Lutestsens 1103      | K-I     | Ca, Mg, Mn, Ni, Mo      | 469              | 30               | 16.1 |
| 125   | Stepnaya 253         | K-I     | Ca, Fe, Zn              | 468              | 31               | 14.1 |
| 114   | OmGAU-90             | K-I     | Ca, Mg, S               | 466              | 33               | 15.7 |
| 132   | Lutestsens 248-01    | K-I     | Mn, Zn, Mo, Sr          | 394              | 58               | 15.9 |
| 66    | RBOT USA             | USA     | Cu, Fe, Rb              | 386              | 63               | 18.4 |
| 87    | L 485                | K-E     | Ca, P, Mn               | 378              | 66               | 17.0 |
| 103   | Lutestsens 15-12     | K-I     | Cu, Co, Ni              | 372              | 71               | 17.6 |
Table 4. Cont.

| Entry | Genotype | Group * | Highest/Lowest Elements | Grain Yield g/m² | Protein Content % | Rank |
|-------|----------|---------|-------------------------|------------------|------------------|------|
| 73    | Freyr    | USA     | Mg, Zn, Ni, Mo, Rb      | 354              | 19.0             | 75   |
| 61    | Pandur/Ae. Squarrosa(409) | S-M | K, S, Cu, Co         | 330              | 17.4             | 79   |
| 78    | Alpine   | USA     | Mg, Fe, Zn, Rb        | 321              | 17.9             | 81   |
| 13    | Ukr-Od 1530.94/Ae. Squarrosa(392) | S-M | Fe, Mn, Sr           | 313              | 17.2             | 82   |
| 77    | Brennan  | USA     | Mg, Fe, Mn            | 297              | 18.7             | 87   |
| 36    | Aisberg/Ae. Squarrosa (369)/Demir | S-M | Cu, Zn, Cd, Co, Mo   | 273              | 16.3             | 94   |
| 3     | Ukr-Od 952.92/Ae. Squarrosa(1031) | S-M | Cd, Co, Rb           | 255              | 17.0             | 100  |
| 12    | Aisberg/Ae. Squarrosa(511) | S-M | Ca, S, Cu, Mo        | 245              | 16.8             | 103  |
| 6     | Ukr-Od 1530.94/Ae. Squarrosa(458) | S-M | P, Cd, Co            | 213              | 17.7             | 115  |
| 57    | Ukr-Od 1530.94/Ae. Squarrosa(392) | S-M | K, P, S, Zn, Cd, Mo  | 209              | 18.2             | 116  |
| 16    | Ukr-Od 1530.94/Ae. Squarrosa(458) | S-M | Ca, Cd, Rb           | 206              | 17.2             | 118  |
| 22    | Langdon/IG 48042 | S-J | Ca, S, Fe            | 141              | 20.7             | 125  |
| 51    | Langdon/IG 131606 | S-J | Ca, Co, Ni           | 97               | 20.6             | 130  |

LSD 0.05 15 - 0.3 -

* K—KASIB group; E—early; I—intermediate; L—late; S—synthetics; M—Mexico; J—Japan.

The genotype with favorable concentrations of six elements was Ukr-Od 1530.94/Aegilops squarrosa(392) (high K, P, S, and Zn, and low Cd and Mo). Superior performance for five elements was found for Lutestsens 1103 (Ca, Mg, Mn, Ni, and Mo), Freyr, USA (Mg, Zn, Ni, Mo, and Rb), and Aisberg/Ae. squarrosa(369)/Demir (Cu, Zn, Cd, Co, and Mo). High Fe and Zn were found for cvs Stepnaya 253 and Alpine. Several genotypes demonstrated low concentrations of at least two trace elements: Novosibirskaya 41 (Cd and Ni), Lutesens 1103 (Ni and Mo), Lutesens 248-01 (Mo and Sr), Lutesens 15-12 and Langdon/IG 131606 (Co and Ni), Freyr (Ni, Mo, and Rb), Aisberg/Ae. squarrosa(369)/Demir and Ukr-Od 952.92/Ae. squarrosa(1031) (Cd, Co, and Rb), Ukr-Od 1530.94/Ae. squarrosa(458) (Cd and Co), Ukr-Od 1530.94/Ae. squarrosa(392) (Cd and Mo), and Ukr-Od 1530.94/Ae. squarrosa(458) (Cd and Rb). Overall, there was a diversity of germplasm with high macro- and micronutrient concentrations and low trace element concentrations within the material studied. The superior genotypes were found in all groups of material, including KASIB, US cultivars, and primary synthetics.

3. Discussion

The growing concern and interest in healthy food, as well as the strategies to combat malnutrition, have resulted in a greater research and development priority to improve the nutritional quality of wheat grain [17]. Recent reviews summarized the achievements and challenges of biofortification for essential elements, including Cu, Fe, Se, and Zn [18], Zn [19], Fe [20], and Se [21]. There is general agreement that agronomic approaches and genetic improvement need to be combined to achieve the best results and to raise the concentrations to target levels. There is evidence from experiments with Fe [22] and Zn [6] that biofortified foods have significantly improved in nutritional value in human diets. The
concept of biofortification of food crops or cereals has yet to attract sufficient attention in Russia, either in research or in the plant-breeding community. Recent reviews by Loskutov and Khlestkina [23] and Shelenga et al. [24], and the study of Bityutskii [7] demonstrated that there is an understanding of the importance of the nutritional value of wheat, barley, and oats, taking into consideration concentrations of essential microelements. However, this has not yet been converted into integrated biofortification programs for wheat and other staple crops.

Two wheat-grain ionome studies recently conducted in Russia and Kazakhstan by our research group provided important information on the element composition of wheat grain and laid the foundation for the development of targeted programs to optimize the concentration of macro- and microelements, and trace metals. Abugalieva et al. [25] evaluated the concentrations of macroelements, toxic heavy metals, and microelements in 179 wheat grain samples collected in 2017 and 2018 from production fields across northern Kazakhstan and the Omsk region of Russia. The concentrations of essential microelements were similar to wheat grain produced in other countries, with exception of Zn. The concentrations of this important element in Omsk and East Kazakhstan were 50 µg/g above the values targeted by the Harvest Plus biofortification program. Even with the losses of Zn during milling, the grain from these regions could be particularly beneficial for human health. In the second study, a KASIB trial from six locations in Kazakhstan and Russia in 2017–2018 was used for grain ionomics analysis to evaluate genotype ×

environment interaction [8], as the effect of year was the least important. For several elements (P, S, Cu, Mn, and Mo), the effect of the site was 2–3 times higher, compared to the effect of the genotype. The effects of the genotype and site were similar for Ca, Mg, Fe, Cd, and Sr concentrations. That study recommended establishment of a modern biofortification breeding program using phenotyping and genomic tools, and effectively using the multilocalional KASIB network.

The current ionomics study is the third in this series addressing the specific issue of variation of element concentrations in diverse spring wheat genetic resources, evaluated over two years under a typical production system in Western Siberia. The study revealed the advantage of the ionomics approach, when all important grain macro- and microelements, and trace metals are phenotyped to analyze their relationships and to evaluate germplasm in an integrative manner. This study proved the importance of protein content as a key trait affecting the concentration of almost all grain elements, reported previously in Fatyukha et al. [26]. Macroelements Mg, P, and S were also significantly correlated with other elements. The share of the seed-coat increases in smaller grains normally results in higher protein content and some elements’ concentrations. However, this relationship was not well pronounced in this study. The grain yield in this diverse germplasm varied fourfold between the different groups, and this trait also significantly correlated with the concentrations of a number of elements. Considering the diversity of the germplasm and the high variation for agronomic traits, the concentrations of all elements were adjusted based on the correlations. Similar adjustments were made by Fatyukha et al. [26], using protein content and P concentration as variables. The adjustments made in this study were well justified and allowed more precise evaluation of genetic resources.

Primary synthetic wheat developed from crosses of durum wheat with Ae. tauschii has been reported as a source of high concentrations of microelements, including Fe and Zn [27,28]. In the current study, based on original values, primary synthetics from Japan also demonstrated high concentrations of a number of elements, including Ca, Mg, P, S, Fe, and Zn. However, after adjustment using multiple regression, the synthetic wheat germplasm largely lost its advantage. Superior germplasm combining high concentrations of macro- and microelements, and low concentrations of trace elements was identified in all germplasm groups, including the KASIB material, the US cultivars, and both synthetics groups. The genotypes with favorable concentrations of five and six elements were also identified in all germplasm groups. Previous study of KASIB trials across six sites [8] included around 40 entries, which were also included in the present study. Cvs Silach
and Novosibirskaya 41, identified as superior for agronomic and ionomic traits in a multilocational trial, also demonstrated superior performance in this study, validating the earlier results.

The crossing strategy to incorporate and combine optimal concentrations of a wide range of elements depends on the nature of the germplasm. Synthetic wheat with a low yield and a number of undesirable traits, such as spike threshability, requires a top-and-back crossing scheme to transfer useful traits, while maintaining and improving grain yield. Several synthetics from the current study possess resistance to leaf, stem rust, and powdery mildew [29], making them attractive as parental material. Disease resistance, short stature, and earliness were additional positive traits of the US cultivars for improvement of Siberian wheat for the ionome profile. Simple crosses and development of a large population may be sufficient to combine positive traits of KASIB and US materials. However, the back-and-top crosses with local material may also be efficiently used. A crossing program within the KASIB breeding network would be straightforward, based on simple crosses and consequent selection.

Genetic gains in utilization of genetic resources for practical breeding will largely depend on the traits’ heritability and phenotyping precision. The KASIB multilocational study established the broad-sense heritability values for macroelements: Mg (0.59) > Ca (0.50) > K (0.44) > P (0.30) > S (0.20), and for microelements: Zn (0.44) > Mn (0.41) > Cu (0.40) > Fe (0.38) [8]. In the current study, the $H^2$ values were higher, due to the utilization of one location, and different for macroelements: S (0.81) > Ca (0.73) > Mg (0.67) > K (0.61) > P (0.60) and microelements: Cu (0.61) > Fe (0.58) > Mn (0.55) > Zn (0.27). This difference is explained by the diversity of the material used and the testing site’s genotype x environment interaction. However, for trace elements, the ranking in the two studies was almost identical. Considering $H^2$, the coefficient of phenotypic variation, and correlations with other traits, trace elements seem to be easier ionomic targets for genomic enhancement. At the same time, important elements, such as Zn, Fe, and Cd, are more influenced by environment and relations with other traits. Consequently, their improvement requires high throughput and precision phenotyping. Determination of grain protein content as a key variable affecting element concentrations will be an important component of a successful selection program.

The levels of heritability for elements varied from low to high, suggesting that breeding improvement for certain elements, such as Sr, could be more effective, compared to Zn. Further, there were positive correlations between certain elements. Positively correlated traits with high heritability could be improved more efficiently by selecting for only one of the correlated traits. For improving elements with low heritability, low selection pressure would be effective. For the elements with no, low, or negative correlations, some kind of intuitive index selection could be used that would allow balancing moderate defects in one trait with obvious gain in others [30] (Simmonds, 1981).

The utilization of genomics greatly enhances breeding efficiency and genetic gains [31]. The material used in this study was subjected to a GWAS analysis, and a number of marker–trait associations were identified for agronomic traits, including protein content [16]. However, grain element composition was not included in that analysis. CIMMYT synthetic wheat used in the current study was assayed for concentrations of 10 minerals in the USA as part of a larger synthetics panel [32]. Multitraits and stable marker–trait associations were identified, and the 13 top synthetic lines were recommended for higher concentrations of beneficial grain minerals (Cu, Fe, Mg, Mn, Ni, and Zn). The next logical step following the current study is to evaluate the effects of functional markers and undertake a GWAS analysis to identify marker–trait associations, validate them against previously published data, and make recommendations for use in practical breeding.

Grain element composition is affected by molecular homeostasis, physiological and biochemical alterations, and intracellular compartmentalization [33]. The current study did not attempt to analyze physiological and biochemical pathways affecting concentrations of elements in wheat grain. However, wheat physiology is an important component of
an integrated program to increase the concentrations of beneficial elements and decrease 
the amounts of toxic metals in the grain. The current ionome study contributes to the 
development of wheat biofortification programs in Russia to develop healthy grain for 
domestic and international markets.

4. Materials and Methods

4.1. Panel Composition and Evaluation

The panel in 2017 and 2018 comprised 135 entries, including two checks, as listed in 
Table S5. The research material included 37 primary synthetics from CIMMYT, developed 
through crosses between Ukrainian winter durum wheat cultivars and several accessions 
of *Aegilops tauschii* from its gene bank. The development of the synthetics through targeted 
selection under abiotic and biotic stresses was described by Morgounov et al. [34]. Eight 
primary synthetics developed by Kyoto University in Japan [35] comprised the second 
group. The US cultivars (14 in total) included hard red spring wheat entries, primarily from 
the University of Minnesota and from Syngenta. Material from KASIB was represented 
by new cultivars and breeding lines, and was divided into early maturing (15 entries), 
intermediate (42 entries), and late-maturing (17 entries) groups. The two checks were 
widely grown spring wheat cultivars in the Omsk region, Pamyati Azieva (Check-1) and 
Serebristaya (Check-2), representing early and intermediate maturity groups, respectively. 
The main contributors of the KASIB germplasm were Omsk State Agrarian University with 
17 entries, and Omsk Agrarian Research Center with 14 entries. The panel also included 
17 cultivars and breeding lines from Kazakhstan.

The trial was planted in the experimental field of Omsk State Agrarian University 
(55.0404° N; 73.3604° E) as a randomized complete block design, with plots of 1 m² and 
four replicates. The soil of the experimental field was meadow chernozem, with 5% organic 
matter content and an average availability of NPK. The preceding crop was black fallow. 
Spring soil preparation comprised harrowing in early May, followed by shallow cultivation 
and harrowing in mid-May. Planting took place between May 15 and May 20, in both years. 
The trials were harvested in the first week of September. Neither fertilizer nor fungicides 
were applied. Weeds were controlled by the application of common herbicide after the 
tillering stage in mid-June. The field observations included agronomic traits, including 
heading dates, disease evaluations, yield, and yield components. The CIMMYT Wheat 
Physiology Manual [36] was used as a guide for germplasm evaluations for all traits and 
diseases. The protein content in the grain was determined using Infratec FOSS 1841.

4.2. Grain Ionomics Analysis

The grain sample (10 gr) for analysis was taken after manual harvesting of the plots 
and machine threshing. The sample was random from a bag of threshed and cleaned seeds. 
One sample was taken from each replication. All the collected samples were shipped to 
the UK and stored in a dry (<60% humidity) and cool (18–20 °C) storage facility. The 
samples were once again cleaned prior to ionomics analysis and the grain moisture content 
was determined. The ionomic analyses were performed with a phenotyping platform at 
the University of Nottingham (UK) through the European Plant Phenotyping Network, 
a research infrastructure project funded by the Horizon 2020 Program of the EU, which 
offered researchers access to a wide range of state-of-the-art plant-phenotyping facilities. 
The analyses were performed using a state-of-the-art PerkinElmer NexION 2000 inductively 
coupled plasma mass spectrometer (ICP-MS).

The samples were prepared for the ICP-MS analysis in the adjoining high-throughput 
preparation laboratory. Wheat grains were transferred into the Pyrex test tubes, weighted, 
and initially predigested with 1 mL concentrated trace-metal-grade nitric acid Primar Plus 
(Fisher Chemicals, Hampton, VA, USA) spiked with 20 μg/L of indium internal standard, 
for approximately 20 h at room temperature. Indium was added to the nitric acid as 
an internal standard for assessing errors in dilution, variations in sample introduction, 
and plasma stability in the ICP-MS instrument. After the predigestion step, samples
were transferred into DigiPREP MS dry block heaters (SCP Science, QC, Canada; QMX Laboratories, Thaxted, UK) and digested for 4 h at 115 °C. After cooling down, 1 mL of trace-metal-grade hydrogen peroxide (Primar, Fisher Chemicals, Hampton, VA, USA) was added to the tubes, and samples were digested in the dry block heaters for an additional 2 h at 115 °C, and then diluted to 10 mL with 18.2 MΩ cm Milli-Q Direct water (Merck Millipore, Burlington, MA, USA).

Five replicate analyses were conducted for each sample from each replication, and the mean value represented the sample’s final readings. The ionomics results were obtained for 23 elements: macroelements, Ca, K, Mg, P, and S; microelements, B, Fe, Cu, Na, Mn, and Zn; toxic trace elements, As, Cd, Cr, Ni, Pb, and Se; and trace elements, Li, Mo, Rb, and Sr. The concentrations were either at trace levels or below the limit of quantification for B, Na, As, Cr, Pb, Se, Li, and Ti. Therefore, these elements were excluded from the analyses. For the remaining 15 elements, all of the concentrations were normalized to the weight of the samples and expressed as μg/g of dry weight.

4.3. Data Processing and Statistical Analysis

Correlations between individual element concentrations and the other variables, viz., grain yield, 1000 kernel weight (TKW), protein content, and macroelement concentrations, were calculated using Microsoft Excel. The correlation analysis results (Table 1) were used to adjust the original element concentration values using multiple regression on the following traits: grain yield, protein content, TKW, and concentrations of Ca, K, Mg, P, and S. The concentration of each element was adjusted only for the traits with significant correlations. Some elements (Co and Rb) did not correlate with any variable and, therefore, no adjustments were made. Some elements (Ca, K, and Fe) did not correlate with other traits in one year but correlated in another year, and adjustments were made only for the year with significant correlations. For all other elements, adjustments were made in both years and the number of variables in the regression varied from one to five.

A factorial ANOVA (genotype x year) was used for statistical analysis for all agronomic traits and for each element independently, using R version 3.4 [37]. Broad-sense heritability (H²) was estimated for each element in individual trials (for each year separately) based on the ANOVA results. All analyses were conducted separately for the original and adjusted values.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/plants11020149/s1, Table S1: ANOVA F-value probability of the effects of genotypes, year, and their interactions for days to heading, grain yield, protein content, TKW, and element composition, Table S2: Variation for agronomic traits and elemental grain concentration in genetic resource groups in 2017–18, Table S3: Variation for agronomic traits and elemental grain concentration in genetic resource groups in 2017–18, Table S4: Variation in grain yield, protein content, 1000 kernel weight, and elemental composition of different groups of germplasm, Omsk, 2017–2018, Table S5: Wheat genotypes used in the study.

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References

1. Morgounov, A.; Sonder, K.; Abugalieva, A.; Bhaduria, V.; Cuthbert, R.D.; Shamanin, V.; Zelenskiy, Y.; DePauw, R.M. Effect of climate change on spring wheat yields in North America and Eurasia in 1981–2015 and implications for breeding. *PLoS ONE* **2018**, *13*, 1–16. [CrossRef] [PubMed]

2. Salt, D.E.; Baxter, I.; Lahner, B. Ionomics and the study of the plant ionome. *Annu. Rev. Plant Biol.* **2008**, *59*, 709–733. [CrossRef] [PubMed]

3. White, P.J.; Broadley, M.R. Biofortification of crops with seven mineral elements often lacking in human diets—Iron, zinc, copper, calcium, magnesium, selenium and iodine. *New Phytol.* **2009**, *182*, 49–84. [CrossRef]

4. Bhatt, R.; Hossain, A.; Sharma, P. Zinc biofortification as an innovative technology to alleviate the zinc deficiency in human health: A review. *Open Agric.* **2020**, *5*, 176–187. [CrossRef]

5. Velu, G.; Singh, R.P.; Joshi, A.K. A Decade of Progress on Genetic Enhancement of Grain Zinc and Iron in CIMMYT Wheat Germplasm. *In Wheat and Barley Grain Biofortification; Om Gupta, P., Pandey, V., Narwal, S., Sharma, P., Ram, S., Singh, G.P., Eds.; Woodhead Publishing: Sawston, UK*, 2020; pp. 129–138.

6. Szazawal, S.; Dhingra, U.; Dhingra, P.; Dutta, A.; Deb, S.; Kumar, J.; Devi, P.; Prakash, A. Efficacy of high zinc biofortified wheat in improvement of micronutrient status, and prevention of morbidity among preschool children and women—A double masked, randomized, controlled trial. *Nutr. J.* **2018**, *17*, 8. [CrossRef] [PubMed]

7. Bityutskii, N.; Yakkonen, K.; Loskutov, I. Content of iron, zinc and manganese in grains of *Triticum aestivum*, *Secale cereale*, *Hordeum vulgare* and *Avena sativa* cultivars registered in Russia. *Genet. Resour. Crop Evol.* **2017**, *64*, 1955–1961. [CrossRef]

8. Morgounov, A.; Savin, T.; Flis, P.; Babkenov, A.; Chudinov, V.; Kazak, A.; Koksel, H.; Likhenko, I.; Sharma, R.; Sheleeva, T.; et al. Effects of environments and cultivars on grain iron content of spring wheat grown in Kazakhstan and Russia. *Crop Pasture Sci.* **2021**, accepted, early view on-line.

9. Sharma, R.C.; Duveiller, E. Selection Index for Improving Helminthosporium Leaf Blight Resistance, Maturity, and Kernel Weight in Spring Wheat. *Crop Sci.* **2003**, *43*, 2031–2036. [CrossRef]

10. Guzman, C.; Medina-Larqué, A.S.; Velu, G.; González-Santoyo, H.; Singh, R.P.; Huerta-Espino, J.; Ortiz-Monasterio, I.; Peña, R.J. Use of wheat genetic resources to develop biofortified wheat with enhanced grain zinc and iron concentrations and desirable processing quality. *J. Cereal Sci.* **2014**, *60*, 617–622. [CrossRef]

11. Velu, G.; Crespo, H.L.; Guzman, C.; Huerta, J.; Payne, T.; Singh, R.P. Assessing genetic diversity to breed competitive biofortified wheat with enhanced grain Zn and Fe concentrations. *Front Plant Sci.* **2019**, *9*, 1971. [CrossRef]

12. Trethowan, R.M.; Morgounov, A.; He, Z.; De Pauw, R.; Crossa, J.; Warburton, M.; Baytasov, A.; Zhang, C.; Mergoum, M.; Alvarado, G. The global adaptation of bread wheat at high latitudes. *Euphytica* **2006**, *152*, 303–316. [CrossRef]

13. Shamanin, V.; Salina, E.; Wanyera, R.; Zelenskiy, Y.; Olivera, P.; Morgounov, A. Genetic diversity of spring wheat from Kazakhstan and Russia for resistance to stem rust Ug99. *Euphytica* **2016**, *212*, 287–296. [CrossRef]

14. Morgounov, A.; Pozherukova, V.; Kolmer, J.; Gultyaeva, E.; Abugalieva, A.; Chudinov, V.; Kuzmin, O.; Rasheed, A.; Rysmbeotov, A.; Shepelev, S.; et al. Genetic basis of spring wheat resistance to leaf rust (Puccinia triticina) in Kazakhstan and Russia. *Euphytica* **2020**, *216*, 1–15. [CrossRef]

15. Bhatta, M.; Shamanin, V.; Shepelev, S.; Baenziger, P.S.; Pozherukova, V.; Pototskaya, I.; Morgounov, A. Genetic diversity and population structure analysis of synthetic and bred wheat accessions in Western Siberia. *J. Appl. Genet.* **2019**, *60*, 283–289. [CrossRef] [PubMed]

16. Bhatta, M.; Shamanin, V.; Shepelev, S.; Baenziger, P.S.; Pozherukova, V.; Pototskaya, I.; Morgounov, A. Marker- Trait Associations for Enhancing Agronomic Performance, Disease Resistance, and Grain Quality in Synthetic and Bred Wheat Accessions in Western Siberia. *G3* **2019**, *9*, 4209–4222. [CrossRef]

17. Hazard, B.; Trafford, K.; Lovegrove, A.; Griffiths, S.; Uauy, C.; Shewry, P. Strategies to improve wheat for human health. *Nat. Food* **2020**, *1*, 475–480. [CrossRef]

18. El-Ramady, H.; Abdalla, N.; Elbasioni, H.; Elbehiry, F.; Elshakhawy, T.; Omara, A.E.-D.; Amer, M.; Bayoumi, Y.; Shalaby, T.A.; Eid, Y.; et al. Nano-biofortification of different crops to immune against COVID-19: A review. *Ecotoxicol. Environ. Saf.* **2021**, *222*, 112500. [CrossRef]

19. Prabhaj, S.; Skalicky, M.; Maitra, S.; Bhadra, P.; Shankar, T.; Breistic, M.; Hejnak, V.; Vachova, P.; Hossain, A. Zinc Biofortification in Food Crops Could Alleviate the Zinc Malnutrition in Human Health. *Molecules* **2021**, *26*, 3509. [CrossRef]

20. Shi, Y.; Li, J.; Sun, Z. Success to iron biofortification of wheat grain by combining both plant and microbial genetics. *Rhizosphere* **2020**, *15*, 100218. [CrossRef]

21. Galić, L.; Vinković, T.; Ravnjak, B.; Lončarić, Z. Agronomic Biofortification of Significant Cereal Crops with Selenium—A Review. *Agronomy* **2021**, *11*, 1015. [CrossRef]

22. Finkelstein, J.L.; Haas, J.D.; Mehta, S. Iron-biofortified staple food crops for improving iron status: A review of the current evidence. *Curr. Opin. Biotechnol.* **2017**, *44*, 138–145. [CrossRef] [PubMed]
23. Loskutov, I.G.; Khlestkina, E.K. Wheat, Barley, and Oat Breeding for Health Benefit Components in Grain. *Plants* 2021, 10, 86. [CrossRef]
24. Shelenga, T.; Kerv, Y.; Perchuk, I.; Solovyeva, A.E.; Khlestkina, E.; Loskutov, I.G.; Konarev, A. The Potential of Small Grains Crops in Enhancing Biofortification Breeding Strategies for Human Health Benefit. *Agronomy* 2021, 11, 1420. [CrossRef]
25. Abugalieva, A.I.; Flis, P.; Shamanin, V.P.; Savin, T.; Morgounov, A. Ionomics analysis of spring wheat grain produced in Kazakhstan and Russia. *Comm. Soil. Sci. Plant Anal.* 2021, 52, 704–7011. [CrossRef]
26. Fatiukha, A.; Klymiuk, V.; Peleg, Z.; Saranga, Y.; Cakmak, I.; Krugman, T.; Korol, A.B.; Fahima, T. Variation in phosphorus and sulfur content shapes the genetic architecture and phenotypic associations within the wheat grain ionome. *Plant J.* 2019, 101, 555–572. [CrossRef]
27. Aberkane, H.; Payne, T.; Kishi, M.; Smale, M.; Amri, A.; Jamora, N. Transferring diversity of goat grass to farmers’ fields through the development of synthetic hexaploid wheat. *Food Secur.* 2020, 12, 1017–1033. [CrossRef]
28. Gupta, P.K.; Balyan, H.S.; Sharma, S.; Kumar, R. Biofortification and bioavailability of Zn, Fe and Se in wheat: Present status and future prospects. *Theor. Appl. Genet.* 2020, 134, 1–35. [CrossRef]
29. Shamanin, V.; Shepelev, S.; Pozherukova, V.; Guityaeva, E.; Kolomiets, T.; Pakhulkova, E.; Morgounov, A. Primary hexaploid synthetics: Novel sources of wheat disease resistance. *Crop Prot.* 2019, 121, 7–10. [CrossRef]
30. Simmonds, N.W. *Principles of Crop Improvement*, 2nd ed.; Longman Group: New York, NY, USA, 1981.
31. Crossa, J.; Fritsche-Neto, R.; Montesinos-Lopez, O.A.; Costa-Neto, G.; Dreisigacker, S.; Montesinos-Lopez, A.; Bentley, A.R. The Modern Plant Breeding Triangle: Optimizing the Use of Genomics, Phenomics, and Enviromics Data. *Front. Plant Sci.* 2021, 12. [CrossRef]
32. Bhatta, M.; Baenziger, P.S.; Waters, B.M.; Poudel, R.; Belamkar, V.; Poland, J.; Morgounov, A. Genome-Wide Association Study Reveals Novel Genomic Regions Associated with 10 Grain Minerals in Synthetic Hexaploid Wheat. *Int. J. Mol. Sci.* 2018, 19, 3237. [CrossRef] [PubMed]
33. Rai, P.K.; Lee, S.S.; Zhang, M.; Tsang, Y.F.; Kim, K.-H. Heavy metals in food crops: Health risks, fate, mechanisms, and management. *Environ. Int.* 2019, 125, 365–385. [CrossRef] [PubMed]
34. Morgounov, A.; Abugalieva, A.; Akan, K.; Akin, B.; Baenziger, S.; Bhatta, M.; Dababat, A.A.; Demir, L.; Dutbayev, Y.; El Bouhssini, M.; et al. High-yielding winter synthetic hexaploid wheats resistant to multiple diseases and pests. *Plant Genet. Resour.* 2017, 16, 273–278. [CrossRef]
35. Matsuoka, Y.; Takumi, S.; Kawahara, T. Natural variation for fertile triploid F1 hybrid formation in allohexaploid wheat speciation. *Appl. Genet.* 2007, 115, 509–518. [CrossRef] [PubMed]
36. Pask, A.J.D.; Pietragalla, J.; Mullan, D.M.; Reynolds, M.P. *Physiological Breeding II: A Field Guide to Wheat Phenotyping*; CIMMYT: Veracruz, Mexico, 2012.
37. R Core Team. *R: A Language and Environment for Statistical Computing*; R Foundation for Statistical Computing: Vienna, Austria, 2017.