TH17 cells in autoimmunity and immunodeficiency: protective or pathogenic?

Ashish K. Marwaha1,2, Nicole J. Leung1,2, Alicia N. McMurchy2,3 and Megan K. Levings2,3*

1 Department of Pathology and Laboratory Medicine, University of British Columbia, Vancouver, BC, Canada
2 Child and Family Research Institute, Vancouver, BC, Canada
3 Department of Surgery, University of British Columbia, Vancouver, BC, Canada

In 2005 a newly discovered T helper cell subset that secreted interleukin (IL)-17 became the center of attention in immunology. Initial studies painted Th17 cells as the culprit for destruction in many different autoimmune and auto-inflammatory diseases. Subsequently, the discovery of patients with primary immunodeficiencies in the IL-17 pathway taught us that Th17 cells have a critical role in defense against certain fungal and bacterial infections. Moreover, the paradoxical exacerbation of Crohn's disease in the clinical trials of a Secukinumab (AIN457), a fully human neutralizing antibody to IL-17A, has cast into doubt a universal pro-inflammatory and harmful role for Th17 cells. Evidence now suggests that depending on the environment Th17 cells can alter their differentiation program, ultimately giving rise to either protective or pro-inflammatory cells. In this review we will summarize the evidence from patients with immunodeficiencies, autoimmune, or auto-inflammatory diseases that teaches us how the pro-inflammatory versus protective function of Th17 cells varies within the context of different human diseases.

Keywords: Th17 cells, autoimmunity, T regulatory cells, immunodeficiency, inflammatory bowel disease, psoriasis, type 1 diabetes, secukinumab

INTRODUCTION

In rare cases, a mutation in an essential gene can disrupt immune homeostasis, leading to clinical immunodeficiency. More commonly, when individuals with a genetic predisposition are exposed to environmental triggers, a failure of immune homeostasis can lead to autoimmunity. In this review, we will discuss how parallel studies of immunodeficiencies and autoimmune diseases have advanced our knowledge of a CD4+ T cell lineage first characterized by production of IL-17A, Th17 cells.

Th17 cells were identified in 2005 (Harrington et al., 2005; Langrish et al., 2005; Park et al., 2005) and, as for other CD4+ T cell lineages, their development is controlled by a combination of transcription factors that initiate a program of transcription factor expression and epigenetic re-modeling (van der Gast et al., 2011). In humans, the cytokines which instruct Th17 cell lineage development likely include IL-6, IL-21, IL-23, and IL-1β (Acosta-Rodriguez et al., 2007a; Chen et al., 2007; Evans et al., 2007; Wilson et al., 2007; Liu and Rohowsky-Kochan, 2008), with a potential synergistic role for TGF-β (Manel et al., 2008; Volpe et al., 2008; Yang et al., 2008a) via its ability to suppress Th1 cell lineage commitment (Santarlasci et al., 2009). Cytokine-driven activation of the signal transducer and activator of transcription (STAT) 3 pathway is an essential step in Th17 cell differentiation (Holland et al., 2007; Yang et al., 2007; Ma et al., 2008), ultimately leading to expression of their lineage-defining transcription factor: retinoic orphan receptor (ROR)C2 (Acosta-Rodriguez et al., 2007a; Annunziato et al., 2007; Wilson et al., 2007; Manel et al., 2008; Crome et al., 2009). Although the IL-17 cytokine family includes six members (Kolls and Linden, 2004), Th17 cells are thought to only produce IL-17A and IL-17F, which are 55% identical (Kolls and Linden, 2004). IL-17A can combine with IL-17F to form a heterodimer and both can form homodimers (Wright et al., 2007).

Th17 cells have many phenotypic characteristics that distinguish them from other Th cell lineages. In addition to IL-17A and IL-17F, Th17 cells secrete other signature cytokines including IL-21 and IL-22 (Bending et al., 2011). They have also been reported to produce IFN-γ (Annunziato et al., 2007), IL-4 (Cosmi et al., 2010), IL-10 (McGeachy et al., 2007), IL-9 (Beriou et al., 2010), IL-26, CXCL8, and CCL20 (Boniface et al., 2008). They are poor producers of IL-2, which may result in their poor proliferative potential in vitro (Santarlasci et al., 2012). They constitutively co-express CCR4 and CCR6, but not CCRX (Acosta-Rodriguez et al., 2007b), and are derived from CD161+ precursors (Cosmi et al., 2008). The effects of Th17 cells on other Th cell lineages have recently been highlighted in many reviews (Annunziato and Romagnani, 2011; Gaffen, 2011; Gaffen et al., 2011; Ghoreschi et al., 2011; Milner, 2011; Pappu et al., 2011; Wilke et al., 2011).

Th17 cells initially developed a reputation as a destructive element in several diseases including multiple sclerosis (MS), rheumatoid arthritis (RA), psoriasis, and inflammatory bowel disease (IBD). In animal models this reputation, came from evidence that the lack of IL-17-producing cells ameliorates experimental autoimmune encephalitis (EAE) and collagen induced arthritis (CIA; Cua et al., 2003). In humans, the reputation was due to correlative data documenting an increase in IL-17-producing cells, particularly at sites of tissue inflammation (Wilke et al., 2011). However, these original conclusions were over-simplified...
investigated whether part of the cellular defect in IPEX patients et al., 2006; D’Hennezel et al., 2009), but recently Bacchetta et al. et al., 2010), resulting from mutations in including enteropathy, type 1 diabetes, and hyper-IgE (McMurchy linked syndrome (IPEX) is a triad of autoimmune syndromes and FOXP3 (Ayyoub et al., 2009; Beriou et al., 2009; Miyara et al., increasing evidence for the existence of cells that co-express IL-17 cells (McMurchy and Levings, unpublished data). Indeed there is directly interact via a DNA-independent mechanism, and dur-

Th17 cells have an essential role in protective immunity the specific pathogens C. albicans and S. aureus. In accordance with this conclusion, Sallusto et al. have characterized different subsets of human Th17 cells that can be differentiated in vitro with antigen specific stimulation by C. albicans and S. aureus (Zielinski et al., 2012).

Th17 cells have an intriguing close developmental link with FOXP3+ CD4+ regulatory T cells (Tregs). FOXP3 and RORC2 can directly interact via a DNA-independent mechanism, and during Th17 cell development FOXP3 is transiently expressed (Zhou et al., 2008). Moreover, upon activation fully differentiated human Th17 cells preferentially express FOXP3 in comparison to Th1 cells (McMurphy and Levings, unpublished data). Indeed there is increasing evidence for the existence of cells that co-express IL-17 and FOXP3 (Ayyoub et al., 2009; Beriou et al., 2009; Miyara et al., 2009; Voo et al., 2009; Kryczek et al., 2011; Ye et al., 2011).

Immune dysregulation, polyendocrinopathy, enteropathy X-linked syndrome (IPEX) is a triad of autoimmune syndromes including enteropathy, type 1 diabetes, and hyper-IgE (McMurphy et al., 2010), resulting from mutations in FOXP3. The cellular basis for this disease has been attributed to Treg dysfunction (Bacchetta et al., 2006; D’Hennezel et al., 2009), but recently Bacchetta et al. investigated whether part of the cellular defect in IPEX patients may not only relate to Treg dysfunction, but also to changes in Th17 cells. Indeed IPEX patients possessed an increased frequency of cells with a surface profile characteristic of Th17 cells (CD4+CCR6+CD161+), expressing RORC2, and producing IL-17 (Passerini et al., 2011). Interestingly, in patients with point mutations that do not abrogate FOXP3 expression, there was an increased frequency of FOXP3+ cells within the CD161+ Th17 cell gate. These data suggest that dysfunctional Tregs may prefer-

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and as discussed below in some diseases Th17 cells clearly have a protective role.

THE ROLE OF Th17 CELLS IN PRIMARY IMMUNODEFICIENCIES

Much of what we know about human Th17 cells comes from the study of a rare primary immunodeficiency called Hyper-immunoglobulin E (Job’s, syndrome). This disease is caused by mutations in STAT3 (Holland et al., 2007) but the underlying cellular basis for the characteristic phenotype of severe pneumonias, mucocutaneous candidiasis, and Staphylococcus aureus abscesses (Buckley et al., 1972; Grimbacher et al., 1999) remained unknown until several groups found that these patients lack Th17 cells in their peripheral blood (de Beaucoudrey et al., 2008; Ma et al., 2008; Milner et al., 2008; Renner et al., 2008). In addition, naïve Th cells from Job’s syndrome patients have low levels of RORC2 expression and cannot be differentiated into Th17 cells in vitro (de Beaucoudrey et al., 2008; Ma et al., 2008; Milner et al., 2008; Renner et al., 2008).

One complication when interpreting data from Job’s syndrome patients is that STAT3 is activated downstream of other cytokines, making it difficult to attribute a clinical phenotype to one pathway. Recently, other immunodeficiencies have been described which involve more specific defects in the IL-17 pathway. For example, two patients with chronic mucocutaneous candidiasis (CMC) disease, characterized by chronic or persistent infection with Candida albicans and S. aureus, were found to have an IL-17RA autosomal recessive deficiency or an IL-17F autosomal dominant deficiency (Puel et al., 2011). In addition, patients with a deficiency in the intracellular adaptor molecule CARD9, which is essential for dectin-1 signaling, also suffer from systemic Candidi
diagnosis and as discussed below in some diseases Th17 cells clearly have a protective role.

EVIDENCE FOR THE PRO-INFLAMMATORY ROLE OF Th17 IN HUMAN AUTOIMMUNITY

The first recognition of the importance of Th17 cells came from studies of EAE. The notion of EAE as a Th1-mediated disease was challenged when mice deficient in the p40 subunit of IL-12 were found to be resistant to EAE whereas mice deficient in the p35 subunit were actually more susceptible to disease (Becher et al., 2002). Cua et al. (2003) solved this paradox by using genetically deficient mice to show that IL-23p19 and IL-12p40, but not IL-12p35, were essential for EAE development. IL-23, which shares the IL-12p40 subunit with IL-12, was subsequently found to stabilize Th17 cells, and these cells were found to be the main contributing factor in EAE (Langrish et al., 2005). Subsequently, a correlation between Th17 cells and human autoimmunity was sought. Below we discuss the evidence for a pro-inflammatory role in autoimmunity and describe attempts to target this axis therapeutically.

Psoriasis is an auto-inflammatory skin disease characterized by recurrent demarcated red and scaly skin plaques. These plaques include infiltrating T cells (mainly Th cells) and dendritic cells in the dermis as well as cytotoxic T cells and neutrophil in the epidermis (Lowes et al., 2007). The resulting inflammatory process results in rapid keratinocyte proliferation, abnormal keratinocyte differentiation, and angiogenesis (Lowes et al., 2007). Initially, increased levels of IFN-γ, TNF-α, and IL-12 in the serum and lesions of psoriasis patients labeled this as a Th1-mediated disease (Di Cesare et al., 2009). However, RORC, IL-1β, IL-6, and IL-23 are also increased in psoriatic skin lesions (Di Cesare et al., 2009) leading to the possibility that Th17 and Th1 act in synergy to produce psoriatic inflammation.

Th17 cells are thought to be recruited to the skin by expression of CCL20, the ligand for CCR6, then locally stabilized by IL-17 and IL-23. IL-23, which shares the IL-12p40 subunit with IL-12, was subsequently found to stabilize Th17 cells, and these cells were found to be the main contributing factor in EAE (Langrish et al., 2005). Subsequently, a correlation between Th17 cells and human autoimmunity was sought. Below we discuss the evidence for a pro-inflammatory role in autoimmunity and describe attempts to target this axis therapeutically.

Th17 cells are thought to be recruited to the skin by expression of CCL20, the ligand for CCR6, then locally stabilized by IL-1 and IL-23. Since both IL-17 and IFN-γ cause keratinocytes and antigen presenting cells (APCs) to produce more IL-1, IL-23, and CCL20, a positive feedback loop causing keratinocyte proliferation is established (Kryczek et al., 2008; Zaba et al., 2009). Several monoclonal antibodies targeting TNF-α and the p40 subunit shared by IL-12 and IL-23 (Ustekinumab) have been approved for clinical use in psoriasis. Since IL-17 can act synergistically with TNF-α to induce keratinocytes to express inflammatory proteins (Chiricozzi et al., 2011), it is possible that anti-TNF-α acts in part by inhibiting...
Th17 cell-driven inflammation. Targeting IL-17 alone with Secukinumab (AIN457) or Ixekizumab, both fully human neutralizing antibodies to IL-17A, is also effective in psoriasis (Hueber et al., 2010; Leonardi et al., 2012), confirming that this is likely a major pathogenic cytokine in this skin disease. Since Th17 cells are not the sole producers of IL-17 (other possible sources of this cytokine in psoriatic plaques include γδ T cells (Cai et al., 2011), mast cells (Lin et al., 2011), neutrophils (Lin et al., 2011), and Tregs (Bovenschen et al., 2011)), whether or not Th17 cells are the major source of this cytokine in skin remains to be determined.

Another disease with strong links to Th17 cells is RA, a chronic autoimmune disease that leads to joint destruction. T cells infiltrating the synovial fluid of RA patients produce high amounts of IL-17A, IL-1β, and IL-6 (Cascao et al., 2010), especially during early disease and pre-treatment (Chabaud et al., 1999; Ziolkowska et al., 2000; Hwang and Kim, 2005; Leipe et al., 2010). Notably, the levels of IL-17 in the synovium correlate with joint damage, whereas those of IFN-γ correlate with protection (Kirkham et al., 2006). Recent evidence supports a role for IL-17F as well as IL-17A in RA, with the two related cytokines acting in synergy to induce other pro-inflammatory cytokines and chemokines in synoviocytes, myeloid cells, and synovial fibroblasts (Lundy et al., 2007; Tran et al., 2007; Hot and Miossec, 2011; van Hamborg et al., 2011). Hence, analogous to the process in psoriasis, a positive pro-inflammatory feedback loop encourages more Th17 differentiation and maintenance in the joint (reviewed in Sarkar and Fox, 2010). Direct clinical evidence for the role of IL-17 in RA comes from recent clinical trials which found that Secukinumab and another anti–IL-17A therapeutic known as LY2439821 significantly benefit these patients (Genovese et al., 2010; Morrison et al., 2011).

Multiple sclerosis is a neurological disease that results from auto-inflammatory damage to the myelin sheaths surrounding nerves in the brain and spinal cord. This disease has historically been associated with the discovery of Th17 cells since, as discussed above, they have a major pathogenic role in EAE (Bettelli et al., 2008; Dong, 2008; Dubin and Kolls, 2008; Weaver and Hatton, 2009). MS patients have increased IL-17 mRNA in their blood as well as cerebrospinal fluid (Matushevicius et al., 1999), and expression of miRNA326 in their peripheral blood mononuclear cells promotes Th17 cell differentiation and correlates with disease severity (Du et al., 2009). Blood-brain barrier endothelial cells layers are more permeable to in vitro polarized Th17 cells, especially if the monolayer is pre-treated with IL-17 or IL-22 (Kebir et al., 2007). These data led to the hypothesis that in MS Th17 cells weaken the blood-brain barrier and enable the migration of immune cells into the normally immune privileged sites within the central nervous system. If this is the case, then Th17 cells may have more of a facilitative than directly pathogenic role in the nervous system, distinct from their clear role in the positive feedback loop of inflammation in psoriasis and RA. Recruitment has begun for a phase II clinical trials of Secukinumab in patients with relapsing-remitting MS. This trial will provide significant insight into the question of whether IL-17 blockade in MS can induce a clinically relevant protective function.

Type 1 diabetes (T1D) is characterized by autoimmune destruction of pancreatic islet cells resulting in the loss of insulin production. Murine studies have yielded conflicting results on the role of Th17 in the NOD mouse model of T1D (Vukkadapu et al., 2005; Jain et al., 2008; Emamuallee et al., 2009; van den Brandt et al., 2010; Lee et al., 2011; Liu et al., 2011; Joseph et al., 2012), including a potential ability to convert into Th1 cells in vivo (Bending et al., 2009; Martin-Orozco et al., 2009). However, more recent data suggest some of this apparent plasticity could be related to the study of in vitro polarized Th17 cells, which are not sufficiently stabilized at the epigenetic level (Bending et al., 2011; Cohen et al., 2011). Patients with T1D, have an increase in circulating IL-17-producing cells (Honkanen et al., 2010; Marwaha et al., 2010; Hughson et al., 2011), including FOXP3 expressing Th17 cells (Marwaha et al., 2010), monocytes that secrete Th17 polarizing cytokines (Bradshaw et al., 2009), and islet-antigen specific Th17 cells (Arif et al., 2011). There is also evidence that pancreatic lymph nodes from T1D patients have an increase in Th17 cells (Ferraro et al., 2011) and that islets from T1D patients, who died close to diagnosis, express IL-17A, RORC, and IL-22 (Arif et al., 2011). Mechanistically IL-17 enhances IL-1β, IFN-γ, and TNF-α-induced apoptosis in human islets (Arif et al., 2011). We have also found that significantly elevated levels of CD8+IL-17+ cells are detectable in the peripheral blood of a large subset of patients with T1D at disease onset (Marwaha et al., 2010). In the context of mounting correlative evidence that IL-17-producing cells may be pathogenic in the early stages of T1D onset, clinical trials to test the effects of therapy with agents such as Secukinumab or Ustekinumab are warranted.

A PROTECTIVE ROLE FOR TH17 IN THE GUT

The success of Ustekinumab, a human IL-12/23 monoclonal antibody, in patients with moderate to severe Crohn’s disease held promise for the targeting of the IL-17 pathway to modulate this disease (Sandborn et al., 2008). However, the paradoxical exacerbation of Crohn’s disease in the clinical trial of a Secukinumab, cast into doubt the pro-inflammatory role of Th17 cells in the gut. Whilst IL-17 cell-producing cells are found in high numbers in inflamed mucosa in Crohn’s disease and Ulcerative colitis patients, more recent data demonstrate that characterization on the basis of IL-17 alone is insufficient to classify these cells as pathogenic. As described above, Th17 can co-secrete IFN-γ (Annunziato et al., 2007; Lee et al., 2009; Cosmi et al., 2011; Hirota et al., 2011) or co-express FOXP3 (Ayoub et al., 2009; Beriou et al., 2009; Miyara et al., 2009; Voo et al., 2009; Kryczek et al., 2011; Ye et al., 2011), indicating the existence of multiple subsets of Th17 cells with functional specialization (Figure 1).

In retrospect, data from mouse models of colitis need to warn as to the protective role for Th17 cells in Crohn’s disease. In the dextran sulfate sodium (DSS)-induced colitis model, administration of a neutralizing IL-17A antibody (Ogawa et al., 2004), deletion of IL-17A (Yang et al., 2008b), or of IL-22 (Zenenicz et al., 2008) all resulted in a worsening of the colitis. In contrast, IL-17F-deficient (Yang et al., 2008b) and IL-21-deficient mice (Fina et al., 2008) were protected against DSS induced colitis. These data suggest there are non-redundant roles of IL-17A versus F, and that, at least in the gut, IL-21 rather than IL-17A or IL-22 may be a primary Th17-derived pathogenic cytokine.

How could Th17-derived cytokines exert a protective functional role in the intestine? First, IL-17A improves barrier function.
The protective and pathogenic functions of Th17 cells. Depending on the local cytokine environment, different subsets of Th17 cells arise and mediate distinct effector function. In the presence of IL-23, Th17 cells seem to be pro-inflammatory and can either make IFN-γ themselves or work in concert with Th1 cells to drive a positive feedback pathway of tissue damage such as that seen in psoriasis or RA. In Crohn’s disease, Th17 cells can differentiate in the gut into protective or pathogenic Th17 cells. In the presence of TGF-β and IL-21, Th17 are re-programmed into “regulatory” Th17 cells which seem to protect from intestinal inflammation.

by strengthening tight junctions after inducing claudin and mucin expression (Kinugasa et al., 2000; Chen et al., 2003). Second, IL-22 improves barrier function by inducing epithelial cell proliferation (Brand et al., 2006) and enhancing goblet cell restoration and mucus production (Sugimoto et al., 2008). Also, a novel suppressive Th17 subset dubbed regulatory Th17 (rTh17) cells has recently been described. When Esplugues et al. (2011) used a CD3-antibody strategy to induce mucosal tolerance, Th17 cells were recruited to the gut but then re-programmed into suppressive, FOXP3-negative, rTh17 cells. The function of rTh17 cells depends on IL-10, TGF-β, and CTLA-4, and does not occur in CCR6-deficient mice where Th17 are not recruited to the gut. The latter data indicate that the mucosal immunity micro-environment is critical for the development of rTh17 cells.

CONCLUSION
Different flavors of Th17 differentiation, ranging from highly pro-inflammatory to suppressive, result from different cytokine micro-environments in various diseases. Th17 cells can no longer be identified solely on the production of IL-17A since the combination of co-secreted cytokines is key to defining their effector function. Moreover, IL-17 is not only produced by Th17 cells, and under certain conditions γδ T cells (Stark et al., 2005), CD8+ T cells (Shin et al., 1998; He et al., 2006), T follicular helper cells (Cua and Tato, 2010), Lymphoid Tissue induced (ITI) cells (Cupedo et al., 2009), and NKT cells (Michel et al., 2007; Lee et al., 2008; Rachitskaya et al., 2008), can all secrete IL-17. We must therefore start to redefine the partial role that Th17 cells play in IL-17-guided immune response. An additional consideration is their potential for plasticity and co-secretion of cytokines that define other Th cell lineages (e.g., IFN-γ), although this is more likely a transient rather than permanent change based on epigenetic analysis (Bending et al., 2011; Cohen et al., 2011). In summary, the notion that Th17 cells are purely pro-inflammatory cells is mistaken, rather these cells mediate a diverse set of responses in infection, autoimmunity, and immunodeficiency.

ACKNOWLEDGMENTS
The authors’ own work is supported by grants from the Canadian Institutes for Health Research and the Juvenile Diabetes Research Foundation. Megan K. Levings holds a Canada Research Chair in Transplantation. Alicia N. McMurchy is a Canada Vanier Scholar. Ashish K. Marwaha holds a Michael Smith Research Trainee award and a Radcliffe Travelling Fellowship at the University of Oxford.
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Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.