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A Helicobacter pylori Homolog of Eukaryotic Flotillin Is Involved in Cholesterol Accumulation, Epithelial Cell Responses and Host Colonization

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The human pathogen Helicobacter pylori acquires cholesterol from membrane raft domains in eukaryotic cells, commonly known as “lipid rafts.” Incorporation of this cholesterol into the H. pylori cell membrane allows the bacterium to avoid clearance by the host immune system and to resist the effects of antibiotics and antimicrobial peptides. The presence of cholesterol in H. pylori bacteria suggested that this pathogen may have cholesterol-enriched domains within its membrane. Consistent with this suggestion, we identified a hypothetical H. pylori protein (HP0248) with homology to the flotillin proteins normally found in the cholesterol-enriched domains of eukaryotic cells. As shown for eukaryotic flotillin proteins, HP0248 was detected in detergent-resistant membrane fractions of H. pylori. Importantly, H. pylori HP0248 mutants contained lower levels of cholesterol than wild-type bacteria (P < 0.01). HP0248 mutant bacteria also exhibited defects in type IV secretion functions, as indicated by reduced IL-8 responses and CagA translocation in epithelial cells (P < 0.05), and were less able to establish a chronic infection in mice than wild-type bacteria (P < 0.05). Thus, we have identified an H. pylori flotillin protein and shown its importance for bacterial virulence. Taken together, the data demonstrate important roles for H. pylori flotillin in host-pathogen interactions. We propose that H. pylori flotillin may be required for the organization of virulence proteins into membrane raft-like structures in this pathogen.

Keywords: Helicobacter pylori, lipid rafts, membrane raft domains, cholesterol, bacterial flotillins, type IV secretion system, CagA

INTRODUCTION

The human gastric pathogen Helicobacter pylori induces chronic gastric inflammation that usually remains asymptomatic. In 10–20% of infections, however, individuals develop either peptic ulceration or gastric cancer (The EUROGAST Study Group, 1993). These severe forms of disease are more commonly associated with infection by H. pylori strains which harbor a cag pathogenicity
island (cagPAI), encoding a type IV secretion system (T4SS) (Montecucco and Rappuoli, 2001). The H. pylori T4SS system mediates the induction of pro-inflammatory (e.g., interleukin-8, IL-8) responses (Viala et al., 2004) and a cell scattering or so-called “ hummingbird” phenotype in epithelial cells (Segal et al., 1999). These responses are mediated by the T4SS-dependent delivery of cell wall peptidoglycan (Viala et al., 2004) and the effector protein, CagA (Odenbreit et al., 2000), respectively. In contrast, the H. pylori T4SS appears to be dispensable for the induction of cytokine responses in macrophages and monocytes (Maeda et al., 2001; Gobert et al., 2004; Koch et al., 2016).

H. pylori T4SS functionality depends on cholesterol-rich microdomains in the plasma membrane of epithelial cells (Lai et al., 2008; Hutton et al., 2010). These microdomains are known as membrane rafts, also commonly referred to as lipid rafts. Interestingly, cholesterol is an important factor for H. pylori chemotaxis and adherence (Anborg et al., 1992). H. pylori has a specific affinity for cholesterol (Trampenau and Muller, 2003) and is able to grow in cholesterol-supplemented media (Testerman et al., 2001). This is consistent with the fact that H. pylori does not appear to carry cholesterol biosynthesis genes critical for de novo sterol synthesis (Testerman et al., 2001) and must obtain the cholesterol from an exogenous source. Indeed, H. pylori is able to up-regulate cholesterol gene expression in gastric epithelial cells in vitro (Guillemin et al., 2002), suggesting one mechanism by which the bacterium may ensure an abundance of cholesterol is present in its environment. H. pylori can acquire cholesterol from membrane raft domains in host cells for incorporation into its own membrane (Wunder et al., 2006). Once incorporated, cholesterol is α-glucosylated by a cholesterol α-glucosyltransferase (Wunder et al., 2006), resulting in glycolipid forms called cholesterol glucosides. This α-glucosylation of cholesterol allows H. pylori to escape phagocytosis, T-cell activation and bacterial clearance in vivo (Wunder et al., 2006), thereby providing a novel mechanism for persistence within the host.

Cholesterol is an indispensable constituent of the plasma membrane and is required for many functions in eukaryotic cells, including cell viability, proliferation (Golusko and Nowicki, 2005), and for the formation of membrane rafts (Simons and Ikonen, 1997). Membrane rafts control numerous protein–protein and lipid–protein interactions at the cell surface and have been implicated in protein sorting, membrane trafficking, cholesterol metabolism, and signal transduction events (Simons and Toomre, 2000; Manes et al., 2003). Prokaryotes may also contain membrane domains with the characteristic structural and functional features of membrane rafts (Lopez and Kolter, 2010). The membrane raft domains in bacteria are likely to harbor and organize proteins involved in small molecule translocation, protein secretion and signal transduction. These membrane raft-like domains have been identified in the human pathogen Borrelia burgdorferi and are thought to contribute to the pathogenesis of Lyme disease (Larocca et al., 2010; Toledo et al., 2014).

Eukaryotic membrane rafts typically contain many proteins, including a prominent raft-associated protein called flotillin, also known as reggie (Simons and Toomre, 2000). There are two known flotillin proteins: flotillin-1 (reggie-2) and flotillin-2 (reggie-1), both of which associate with membrane rafts (Lang et al., 1998). Flotillin-1 is involved in a variety of cellular processes, including vesicle trafficking, cytoskeletal rearrangement, and signal transduction (Langhorst et al., 2005). Flotillin proteins also play key roles in cell–cell adhesion (Otto and Nichols, 2011), clathrin-independent endocytosis (Otto and Nichols, 2011), and the uptake of dietary cholesterol via vesicular endocytosis (Ge et al., 2011).

Flotillins belong to the Stomatlin, Prohibitin, Flotillin, and YuaG/C (SPFH) protein superfamily, whose members share an SPFH domain at their N-terminus. These proteins are highly conserved across human and animal species and also exist in some bacteria, plants and fungi (Langhorst et al., 2005). Bioinformatic analyses indicate that most bacterial genomes encode proteins with similarity to Flotillin-1 (Lopez and Kolter, 2010). The best characterized of these bacterial flotillins is the YuaG protein of the gut commensal, Bacillus subtilis. This flotillin homolog was reported to play roles in a diverse range of cellular functions, including cell division, the maintenance of bacterial shape, and sporulation (Donovan and Bramkamp, 2009; Bach and Bramkamp, 2013; Mieliich-Suss et al., 2013). It has been suggested that bacterial flotillins, such as YuaG, may be involved in membrane order or organization and are likely to guide the recruitment of specific proteins to defined areas within the membrane (Bach and Bramkamp, 2013). Consistent with this suggestion, B. subtilis YuaG forms punctate staining patterns along the cell membranes of the bacterium (Donovan and Bramkamp, 2009; Lopez and Kolter, 2010) and interacts with proteins involved in various functions, including protein secretion, cell wall metabolism and signaling processes (Bach and Bramkamp, 2013). Although a flotillin homolog has been cloned from the human pathogen, Staphylococcus aureus (Lopez and Kolter, 2010), the role(s) of flotillin proteins in bacterial pathogenesis have yet to be investigated.

Herein, we present the first characterization of a flotillin-like protein in the virulence of a human pathogen, H. pylori. This protein (HP0248) was shown to be involved in the accumulation of cholesterol within H. pylori cell membranes. Importantly, we show that H. pylori HP0248 mutants were affected in their abilities to induce T4SS-dependent responses in gastric epithelial cells and to establish chronic infection in mice. Collectively, the data demonstrate that the H. pylori flotillin-like protein, HP0248, is involved in the accumulation of bacterial membrane cholesterol, thereby contributing to H. pylori pathogenesis.

**MATERIALS AND METHODS**

**Bacterial Strains, Media, and Culture Conditions**

H. pylori 251 and 26695 are cagPAI+/T4SS+ laboratory strains (Viala et al., 2004), whereas SS1 (cagPAI+/T4SS−) and X47-2AL (cagPAI+/T4SS−) are mouse colonizing strains (Grubman et al., 2010). The H. pylori 251ΔcagA mutant was described previously (Hutton et al., 2010). H. pylori strains were routinely cultured on either blood agar or brain heart infusion.
broth (BHI; Oxoid) containing a modified Skirrow’s selective supplement (comprising vancomycin, 10 µg/ml; polymyxin B, 25 ng/ml; trimethoprim, 5 µg/ml; and amphotericin B, 2.5 µg/ml), according to standard procedures (Ferrero et al., 1998). Escherichia coli BL21 was propagated on Luria-Bertani (LB) agar or broth with the appropriate antibiotic. H. pylori was incubated with epithelial cells at a multiplicity of infection (MOI) of 10 (Hutton et al., 2010). Viable counts of H. pylori were determined by serial dilution and plating. The cholesterol content of the bacteria was determined using the Amplex red cholesterol detection kit, according to the manufacturer’s instructions (Molecular Probes, OR, USA).

**Cell Culture**

Human gastric adenocarcinoma cells (AGS) and murine macrophages (RAW 264.7) were routinely cultured in RPMI medium, supplemented with 10% FCS, 50 units/ml penicillin, 50 µg/ml streptomycin and 1% (v/v) L-glutamine (all reagents from Life Technologies, CA, USA). Cells were seeded at 1 × 10^5 cells/ml and incubated at 37°C in 5% CO2. For co-culture assays, cells were serum-starved overnight in antibiotic-free RPMI medium, then washed 2–3 times in antibiotic-free RPMI medium prior to the addition of H. pylori bacteria.

**In vitro Adherence Assay**

Bacterial adherence was assessed using modifications to a previously described method (Chionh et al., 2009). Briefly, AGS cells were seeded at 1 × 10^4 cells in duplicate sets of triplicate wells in 96-well plates (Falcon) and incubated for 24 h at 37°C in 5% CO2. Cells were serum-starved overnight and co-cultured with H. pylori bacteria for 6 h. One set of the duplicate wells was washed three times with PBS to remove any unattached bacteria. Both sets of wells were fixed for 20 min in a final concentration of 4% paraformaldehyde and then blocked in 1% BSA in PBS for 30 min at room temperature. Cells were incubated for 1 h at room temperature with rabbit anti-H. pylori antiserum (diluted 1:500 in PBS; Ferrero et al., 1994). A horseradish peroxidase (HRP)-conjugated goat anti-rabbit antibody (diluted 1:2,000 in PBS; Dako, Glostrup, Denmark) was then added, followed by color development with 3,3′,5,5′-tetramethylbenzidine (TMB) (Pierce, Thermoscientific, Rockford, IL, USA). The reaction was stopped by adding 0.5 M H2SO4. Absorbance was read at 450 nm and the percentages of adherent bacteria were calculated by dividing the average OD of the washed set of samples by the average OD of the unwashed set.

**Cytokine Responses**

Serum-starved cells were stimulated with either live H. pylori (MOI = 10) or bacterial lysates (20 µg protein/ml), prepared by freeze-thawing, and incubated at 55°C for 20 min. Following incubation for 1 h, the culture medium was replaced and the cells washed 2–3 times to remove bacteria. Cells were then placed in fresh antibiotic-free medium and incubated a further 5 or 23 h, respectively. IL-8 and IL-6 levels in culture supernatants were determined by sandwich ELISA, according to the manufacturer's instructions (BD Biosciences).

**Isolation of H. pylori Detergent-Resistant Membranes (DRMs)**

H. pylori DRMs were isolated using modifications to a previously described method (Lopez and Kolter, 2010). Briefly, H. pylori bacteria (~1–5 × 10^5 bacteria) were pelleted after 16 h of growth in BHI liquid culture medium, washed three times with 20 mM Tris-HCl (pH 7.5) and resuspended in 20 mM Tris-HCl containing protease inhibitors. Bacterial cells were sheared using a French press and cell debris was eliminated by centrifugation. Membrane fractions were precipitated from supernatants by ultracentrifugation (40,000 × g for 30 min at 4°C) using a Sorvall RC90 ultracentrifuge (Kendro, NC). Proteins that associated with hydrophobic DRM fractions were separated from hydrophilic DSM fractions by phase separation, using the CellLytic MEM protein extraction kit (Sigma) (Lopez and Kolter, 2010).

**Cloning the SPFH Domain of HP0248**

The Gateway® Cloning System (Life Technologies) was used to generate an expression vector for the production of a His-tag-labeled 163 amino acid region internal to the predicted SPFH domain of H. pylori HP0248 (deduced total length 362 amino acids). Primers MH9 and MH10 (Supplementary Table 1) were used to amplify a 487 bp fragment, containing attB1 and attB2 sites, from the HP0248 gene of H. pylori 26695. The resulting PCR product was cloned into the entry clone, pDONR 221. The PCR amplicon was then recombined into the destination vector pDEST17 using LR clonase. Final destination plasmids were confirmed by sequence analysis using the T7 promoter and terminator primers. The pDEST17 expression vector was transformed into E. coli BL21 cells for expression.

**Expression and Purification of Recombinant HP0248**

The SPFH domain of H. pylori HP0248 was expressed in E. coli using standard techniques (see Supplementary Methods for details). Briefly, expression was induced by the addition of 0.4 mM isopropyl β-D-1-thiogalactopyranoside (IPTG; Promega) and the bacterial suspensions pelleted, lysed by both sonication and 0.1 mg/ml lysozyme (Sigma) treatment. The insoluble pellets were sonicated and the proteins solubilized using 6 M guanidine hydrochloride (Amresco, Ohio, USA). The solubilized proteins were loaded onto a HisTrap HP column (GE Healthcare, Uppsala, Sweden) and recombinant HP0248 purified by immobilized metal ion affinity chromatography. Polyclonal antibodies to HP0248 were generated by administering the purified recombinant protein to a New Zealand White rabbit (Walter and Eliza Hall Institute of Medical Research Antibody Facility; Bundoora, Melbourne, Australia). This anti-HP0248 serum (diluted 1:1,000) was used in Western blot analyses.

**SDS-Page and Western Blotting**

Whole cell lysates, DRM or DSM fractions were resuspended in solubilization buffer, separated in either 12.5% (v/v) acrylamide or 4–12% (v/v) NuPAGE® Bis-tris (LifeTechnologies) gels and transferred to nitrocellulose. H. pylori proteins were reacted with rabbit anti-HP0248 serum (diluted 1:1,000) or anti-UreA
antibody (1:5,000; Ferrero et al., 1994), followed by addition of a goat anti-rabbit-HRP conjugated antibody (1:1,000 dilution; Dako). Antibody-antibody complexes were detected using ECL detection reagent (Pierce).

H. pylori Mutagenesis
Mutagenesis was performed using the Gateway® Cloning System (Life Technologies). Primer combinations and DNA fragments for cloning were as follows: MH1 and MH3 (Supplementary Table 1), a 299 bp fragment from the 5’ end of the hp0248 of H. pylori 26695; GmB4r and GmB3r, the gentamicin resistance cassette (GmR) from the pUC1813apra vector (Bury-Mone et al., 2003); and MH4 and MH2, a 368 bp fragment from the 3’ end of the hp0248 gene. PCR products were cloned into three entry clones and integration confirmed by sequence analysis using the M13 primer pair (Supplementary Table 1). Recombination was performed in the destination vector pDEST17 using LR clonase and the final destination plasmid confirmed by sequencing using the T7 promoter and terminator primers (Supplementary Table 1). H. pylori ΔHP0248 mutants were created by natural transformation and selected on horse blood agar (HBA) containing apramycin (30 µg/mL; Sigma) (Grubman et al., 2010). Apramycin-resistant transformants in each strain of H. pylori were verified by sequencing using primers MH5, MH6, and GmFwd or GmRvs (Supplementary Table 1). Mutant bacteria were characterized by a 263-amino acid deletion in HP0248. This truncated form of HP0248 included a 185-amino acid deletion within the 231 amino acid SPFH domain.

Complementation of H. pylori HP0248 Mutants
Primer combinations and DNA fragments for cloning were as follows: LT8F and LT8R (Supplementary Table 1), a 301 bp fragment from the 5’ end of the H. pylori 26695 rdxA locus; LT9F and LT9R, a 191 bp fragment, encompassing the H. pylori 26695 ureA promoter sequence; MH11 and MH12, H. pylori hp0248 from H. pylori 26695; and LT10F and LT10R, the 3’ end of rdxA. The four PCR products were cloned into the pDEST17 destination vector and confirmed as above. H. pylori ΔHP0248 mutants, in which an exogenous copy of hp0248 was inserted into the rdxA locus (Smeets et al., 2000), were selected by natural transformation and cultured on HBA plates supplemented with metronidazole (8–12 µg/mL; Sigma). Mutants were verified by sequencing using the primers AG1F, MH5, MH6, and MH13R (Supplementary Table 1).

Densitometric Analysis of HP0248 Abundance in DRM and DSM Fractions
Western blots of DRM and DSM fractions from H. pylori 251 WT, ΔFLOT, or FLOT (FLOT+) strains were analyzed using ImageJ software. The percentage area for HP0248 in each fraction was quantified relative to that for the non-specific protein band in the corresponding fraction. All values were then normalized to the wild-type DRM fraction.

Proteomic Analysis of H. pylori DRM and DSM Fractions
Coomassie-stained protein bands in DRM samples from H. pylori 251 WT were excised from preparative SDS-PAGE gels (4–12% NuPAGE® Bis-tris; Life Technologies) and trypsin digested prior to identification by Matrix Assisted Laser Desorption/Ionization Time-of-Flight (MALDI-TOF) and Liquid Chromatography/Mass Spectrometry (LC-MS/MS). For MALDI-TOF, digested samples were co-spotted onto the MALDI target plate and analyzed on an Applied Biosystems (Foster City, CA, USA) 4700 Proteomics Analyser MALDI TOF/TOF in reflectron mode (see Supplementary Methods for details). Data were searched against an in-house database compiled from H. pylori genomes downloaded from the ExPASy FTP site (ftp.expasy.org) using the MASCOT search engine (version 1.9, Matrix Science Inc., London, UK) with all taxonomy selected. The following search parameters were used: missed cleavages, 1; peptide mass tolerance, ± 50 ppm; peptide fragment tolerance, ±0.1 Da; peptide charge, 1+; fixed modifications, carbamidomethyl; variable modification, oxidation (Met), and the top five matches reported. Scores were considered significant when above the MASCOT-generated probability-based Mowse score minimum threshold.

LC-MS/MS was performed using an HCT ULTRA ion trap mass spectrometer (Bruker Daltonics, Bremen, Germany), coupled online with an RSLC nano HPLC (Ultimate 3000, Dionex Corporation, SunnyBrook, CA, USA; see Supplementary Methods for details). Data from LC-MS/MS analysis were exported in the Mascot generic file format (*.mgf) and searched against an in-house database, as above. The following search parameters were used: enzyme specificity, trypsin; missed cleavages, 1; peptide mass tolerance, ±0.6 Da; peptide fragment tolerance, ±0.3 Da; peptide charge, 2+ and 3+; fixed modifications, carbamidomethyl; variable modification, oxidation (Met).

To identify proteins in DRM and DSM fractions from H. pylori WT, ΔFLOT, and ΔFLOT (FLOT+) bacteria, sections of SDS-PAGE gel corresponding to a molecular weight of ~40 kDa were excised. These sections of gel (~1 cm high and spanning the width of the gel) were sliced into six pieces and each trypsin digested prior to analysis. In this case, LC-MS/MS was performed using the QExactive mass spectrometer (Thermo Scientific, Bremen, Germany) coupled online with an RSLC nano HPLC (Ultimate 3000, Thermo Scientific, Bremen, Germany; see Supplementary Methods for details). For each sample, pooled data from six gel slices were exported in the Mascot generic file format (*.mgf) and analyzed as above.

Quantitation of Cell Scattering Responses by High Throughput Analysis
AGS cells were seeded on coverslips at 1 × 10⁵ cells/ml and incubated for 24 h at 37°C in 5% CO₂. Cells were then serum-starved overnight prior to co-culture with H. pylori for 6 h. Immunofluorescence staining of cellular actin using Alexa...
Fluor® 488 phalloidin (1:40 dilution; Molecular Probes) was performed as described previously (Hutton et al., 2010). For staining of bacteria, co-cultured cells or H. pylori that had been air dried and fixed onto microscope slides (Polysine™, Menzel-Glaser, Braunschweig, Germany) were incubated with rabbit anti-H. pylori sera (diluted 1:5,000 in PBS) for 1 h at room temperature, followed by incubation with an anti-rabbit Alexa Fluor® 568 conjugated antibody (1:1,000 dilution; Molecular Probes). The stained bacteria were examined using a Nikon C1 confocal microscope. Actin staining in cells was viewed using the Cellomics ArrayScan VTI HCS Reader (Thermo Scientific), capturing 20 fields per well with the 20 x objective.

Quantification of CagA Translocation by Immunofluorescence
AGS cells were seeded in μ-slide eight well-chambers (Ibidi) at 3 × 10⁴ cells/ml and incubated for 24 h at 37°C in 5% CO₂. Cells were then serum-starved overnight prior to co-culture with H. pylori for 6 h. Cells were subsequently fixed using 4% paraformaldehyde and their nuclei stained with DAPI (diluted 1:1000 in PBS; Molecular Probes). Extracellular CagA was detected using a rabbit anti-CagA (b-300) antibody (1:100; sc-25766, Santa-Cruz Biotechnology), followed by incubation with an anti-rabbit Alexa Fluor 488 conjugated antibody (1:1,000 dilution; Molecular Probes). Cells were then re-fixed and permeabilized using 1% Triton-X. Intracellular CagA was probed using the same rabbit anti-CagA antibody, followed by incubation with an anti-rabbit Alexa Fluor 647 conjugated antibody (1:1,000 dilution; Molecular Probes). Cells were imaged with the same background intensity settings on a Deltavision API wide-field microscope (60 x objective). Image analysis was performed using ImageJ, where relative intensities of A₆₄⁷/A₄₈₈ were quantified for five cells per field viewed (five fields viewed per sample; n = 2 experiments).

Mouse Infection
Animal handling and experimentation was performed in accordance with Victorian State Government regulations and approved by the Monash University Animal Ethics Committee (ethics no. MMCA 2010/18). H. pylori suspensions for mouse inoculation were prepared by harvesting bacteria from HBA plates using BHI broth (Ferrero et al., 1998). Six- to eight-week-old female specific pathogen/Helicobacter-free C57BL/6 mice were each intrastracically administered a single 100-μl aliquot of the inoculating suspension (10⁷ cfu/mouse) using polyethylene catheters (Ferrero et al., 1998). The presence of H. pylori infection in mice was determined after 30 days of infection by quantitative culture as described previously (Ferrero et al., 1998).

Statistical Analysis
Data were analyzed using the Student’s t-test, Mann-Whitney U-test or ANOVA, as appropriate. Differences in data values were considered significant at a P < 0.05.

RESULTS
H. pylori Has a Flotillin-Like Protein (HP0248) That Partitions to Membrane Raft-Like Domains in Its Cell Membrane
Studies have recently described the presence of membrane raft-like domains within the cell membranes of a pathogenic bacterium, B. burgdorferi, suggesting that membrane rafts may be conserved amongst both prokaryotic and eukaryotic organisms (Larocca et al., 2010; Toledo et al., 2014). Consistent with this suggestion, the commensal bacterium S. subtilis was reported to contain a protein, YuaG, with homology to flotillins, a family of membrane raft-associated proteins (Donovan and Bramkamp, 2009). A flotillin-like protein was identified in the bacterial species, S. aureus, but this protein was not characterized (Lopez and Kolter, 2010). Given that H. pylori cell membranes are highly enriched in cholesterol, a feature of membrane rafts, we speculated that H. pylori may have a flotillin-like protein. To address this question, we used the primary amino acid sequence of B. subtilis YuaG (Donovan and Bramkamp, 2009; Lopez and Kolter, 2010) to perform BLAST analyses on the order Campylobacterales. We first identified homologs in Helicobacter hepaticus (HH0856) and several Campylobacter spp. These proteins were, in turn, used to identify the presence of a hypothetical protein, HP0248, in the H. pylori 26695 strain. HP0248 and its predicted protein in this H. pylori strain have accession numbers NC_000915.1 (257084-258172) and NP_207046.1, respectively. We identified a 231 amino acid region in H. pylori HP0248, which when subjected to CLUSTALW analysis, exhibited 14.7% identity and 42.7% similarity with the SPFH domain of human flotillin-1 (Supplementary Figure 1) (Langhorst et al., 2005). The SPFH domain of H. pylori HP0248 shares a similar level of identity with that of human flotillin-2 (data not shown). Homologs of HP0248 are present in other H. pylori strains (e.g., J99, B128, G27), as well as in various human and animal Helicobacter spp e.g., Helicobacter acinonychis, Helicobacter felis, Helicobacter mustelae (data not shown). This highlights the likely importance of HP0248 in Helicobacter biology. On the basis of the findings, we have named the H. pylori HP0248 protein, “flotillin-like protein” or “FLOT.”

In order to characterize the biological functions of the H. pylori flotillin-like protein, we generated Δhp0248 deletion mutants (ΔFLOT) in H. pylori 26695 and 251, both of which have functional T4SSs (Viala et al., 2004), as well as in two strains that do not i.e., X47-2AL and S51 (Grubman et al., 2010). We also generated a complemented mutant (ΔFLOT (FLOT+)) in the 251 strain. Western blot analyses using a polyclonal rabbit antibody raised against the HP0248 SPFH domain confirmed the production of HP0248 in H. pylori 251 wild-type (WT) and ΔFLOT (FLOT+) strains, but not in a ΔFLOT mutant (Figure 1A).

To determine the cellular localization of H. pylori HP0248, whole bacteria were fractionated into cytoplasmic, inner and outer membrane compartments (Voss et al., 2014). By Western blotting, HP0248 was shown to be present in the outer but not inner membrane (Figure 1B). The H. pylori urease A subunit (UreA), which is present in both cytoplasmic and membrane
fractions (Phadnis et al., 1996), was used as a sample loading control. Given that flotillins normally partition to membrane rafts, we next sought to determine whether H. pylori HP0248 might also localize to membrane raft-like structures within the bacterial membrane. To do this, we exploited the known property of raft-associated proteins to partition to detergent-resistant membrane (DRM) fractions. This approach has been used previously to isolate hydrophobic proteins from membrane rafts in other bacteria (Larocca et al., 2010; Lopez and Kolter, 2010). As shown in Figure 1C, DRM fractions of H. pylori membranes exhibited markedly different SDS-PAGE profiles to those of detergent-sensitive membrane (DSM) fractions, consistent with the separation of hydrophobic raft-associated proteins from hydrophilic proteins. Importantly, HP0248 was detected in the DRM fractions of WT and ΔFLOT bacteria, but not in those of ΔFLOT bacteria (Figures 1D,E). The presence of HP0248 in the DSM of the ΔFLOT (FLOT+) mutant can be attributed to over-expression of this protein in the strain due to the use of the strong ureA promoter for complementation (Grubman et al., 2010). Using two different biochemical approaches, we showed that the H. pylori FLOT protein associates with the bacterial membrane.

**Proteomic Analyses Identify H. pylori HP0248 within DRM Fractions**

Proteomic analyses of the total DRM fractions of WT bacteria identified 29 putative and five hypothetical proteins (Table 1). Amongst the putative proteins, 19 were proven experimentally to associate with membranes (Baik et al., 2004; Carlsohn et al., 2006), with an additional four predicted by the PSORTb program to be membrane-associated (Table 1, Figure 1B). Although the six remaining putative proteins are defined as cytoplasmic (Table 1, Supplementary Table 2), five of these have actually been shown to associate with H. pylori membranes i.e., HP0072, HP0073, HP0248, HP1462, HP1563. Amongst these predicted cytoplasmic proteins are HP0248 and UreA (HP0073) (Figure 1B, Phadnis et al., 1996). Similar findings were reported in a proteomic study on B. burgdorferi DRMs in which the majority (63%) of the proteins found in those fractions were predicted to be associated with the bacterial membrane, however,
To directly compare the protein composition of DRM and DSM fractions, we performed LC/MS-MS analysis on DRM and DSM fractions from *H. pylori* WT, ΔFLOT and ΔFLOT (FLOT+) bacteria. Proteins in the molecular weight range of HP0248 i.e., 40 kDa were selectively analyzed by LC-MS/MS using the QExactive mass spectrometer. This analysis yielded a larger number of protein “hits” than that acquired on the HCT ULTRA ion trap mass spectrometer (Table 1). This is most likely due to the greater level of sensitivity of the former. Most importantly, however, HP0248 was again detected in the DRM but not DSM fractions of WT and ΔFLOT (FLOT+) bacteria (Supplementary Table 3). Additionally, 50 proteins were identified within the DRM fractions from all three *H. pylori* strains (Supplementary Figure 2), of which 31 were not detected in any of the DSM fractions (Supplementary Tables 3, 4). The latter were considered to be putative membrane raft-associated proteins. In addition to HP0248, several outer membrane proteins (OMPs) were found to partition to the DRM fractions (Table 1, Supplementary Tables 3, 4), suggesting that proteins involved in host-pathogen interactions may selectively localize to the membrane raft-like structures in *H. pylori* membranes. Taken together, the data demonstrate that *H. pylori* HP0248 preferentially associates with a fraction enriched in membrane rafts.

**H. pylori** HP0248 Is Important for Cholesterol Accumulation and Induction of IL-8 Responses

The *H. pylori* 251 ΔFLOT and ΔFLOT (FLOT+) strains grew as well as the WT strain in BHI broth medium supplemented or not with cholesterol (data not shown). Importantly, however, the *H. pylori* 251 ΔFLOT mutant strain possessed significantly lower cholesterol levels, when compared with WT bacteria (Figure 2A; P < 0.01). Similar findings were obtained for ΔFLOT mutants on the *H. pylori* 26695, X47-2AL, and SS1 backgrounds, with these mutants consistently displaying 40% less cholesterol than the respective WT strains (data not shown). Finally, the *H. pylori* 251 ΔFLOT (FLOT+) complemented strain exhibited WT levels of cholesterol (Figure 2A), thereby confirming the important function of HP0248 in *H. pylori* cholesterol accumulation.

Given the importance of *H. pylori*-associated cholesterol for the induction of cellular responses, we next examined interactions between *H. pylori* WT, ΔFLOT, or ΔFLOT (FLOT+) bacteria and host epithelial cells. As shown in Figure 2B, *H. pylori* ΔFLOT 26695 and 251 mutants adhered as well to AGS cells as the corresponding WT strains, suggesting that adhesion occurs independently of the flotillin-like protein. In contrast, *H. pylori* ΔFLOT mutant strains on both 26695 and 251 backgrounds induced significantly lower IL-8 responses, when compared with WT strains (Figure 2C; P < 0.05 and P < 0.001, respectively). ΔFLOT (FLOT+) bacteria induced significantly increased IL-8 responses when compared with ΔFLOT organisms, however, these responses were only partially restored with respect to those induced by WT bacteria (Figure 2C; P < 0.001 and P < 0.01, respectively). As observed for cholesterol-enriched and cholesterol-depleted bacteria, *H. pylori* ΔFLOT mutants induced similar IL-6 and TNF-α responses in macrophages as WT organisms (Figure 2D; data not shown). Cumulatively, these data indicate that *H. pylori* HP0248 is involved in cholesterol accumulation in the bacterial cell membrane and induction of IL-8 responses in epithelial cells. We speculate that the absence of HP0248 may destabilize membrane raft domains in the *H. pylori* cell membrane, thereby affecting T4SS-dependent functions.

### Table 1 | Proteins identified in DRM fractions of *H. pylori* 251 WT bacteria.

| HP number | Protein | Predicted protein location |
|-----------|---------|---------------------------|
| HP0025    | OMP2   | Membrane-associatedh,c    |
| HP0072    | Urease subunit beta (UreB) | Cytoplasmich,c           |
| HP0073    | Urease subunit alpha (UreA) | Cytoplasmich,c           |
| HP0097    | Hypothetical protein | Unknown                   |
| HP0127    | OMP4 (HorB) | Membrane-associatedh,c   |
| HP0130    | Hypothetical protein | Unknown                   |
| HP0185    | Hypothetical protein | Cytoplasmic               |
| HP0229    | OMP6 (HopA) | Membrane-associatedh,c   |
| HP0248    | Flotillin-like protein | Cytoplasmic               |
| HP0252    | OMP7 (HopF) | Membrane-associatedh     |
| HP0317    | OMP9 (HopU or BabC) | Membrane-associatedh      |
| HP0377    | Thioldisulphide interchange protein (DistE) | Membrane-associatedh      |
| HP0472    | OMP11   | Membrane-associatedh,c    |
| HP0488    | OMP (HorC) | Periplasmic               |
| HP0606    | Membrane fusion protein (MtrC) | Membrane-associatedh,c   |
| HP0671    | OMP14 (HorF) | Membrane-associatedh,c   |
| HP0706    | OMP15 (HopE) | Membrane-associatedh,c   |
| HP0797    | Lipoprotein (HpaA) | Membrane-associatedh      |
| HP0896    | OMP19 (BabB or HopT) | Membrane-associatedh      |
| HP0912    | OMP20 (HopC, AlpA) | Membrane-associatedh,c   |
| HP0913    | OMP21 (HopB, AlpB) | Membrane-associatedh,c   |
| HP1069    | ATP-dependent zinc metalloprotease | Membrane-associatedh     |
| HP1125    | OMP18 (Peptidoglycan-associated lipoprotein precursor) | Membrane-associatedh     |
| HP1132    | ATP synthase F1, subunit beta | Cytoplasmic               |
| HP1173    | Hypothetical protein | Cytoplasmic               |
| HP1177    | OMP27 (HopQ) | Membrane-associatedh,c   |
| HP1243    | OMP28 (HopS, BabA) | Membrane-associatedh      |
| HP1295    | OMP30 (HorL) | Membrane-associatedh      |
| HP1462    | Secreted protein involved in motility | Cytoplasmich             |
| HP1463    | Hypothetical protein | Unknown                   |
| HP1469    | OMP31 (HopV) | Membrane-associatedh,c   |
| HP1488    | 36 kDa antigen | Membrane-associatedh      |
| HP1540    | Ubiquinol cytochrome c oxidoreductase | Membrane-associatedh      |
| HP1563    | Alkyl hydroperoxide reductase (TisA) | Cytoplasmic               |

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a Determined using the PSortb prediction program (http://www.psort.org/psortb/).
b Membrane association proved experimentally (Carlsohn et al., 2006).
c Membrane association proved experimentally (Bask et al., 2004).
**H. pylori** HP0248 Is Important for T4SS-Induced Cell Scattering Responses and CagA Translocation in Epithelial Cells

In addition to the induction of IL-8 production in epithelial cells, the *H. pylori* T4SS mediates delivery of the effector molecule, CagA, which causes rearrangement of the host cell actin cytoskeleton and results in cell scattering (Segal et al., 1999; Odenbreit et al., 2000; Backert et al., 2001). To further investigate the importance of *H. pylori* HP0248 in T4SS-dependent functions, we co-cultured AGS cells with *H. pylori* WT, ΔFLOT, or ΔFLOT (FLOT+) bacteria, and used a high throughput imaging technique to quantify the proportions of cells displaying a cell scattering response (Figure 3A). Consistent with the IL-8 data (Figure 2C), AGS cells that had been co-cultured with an *H. pylori* ΔFLOT mutant strain displayed weaker cell scattering responses, when compared with cells co-cultured with either the WT or complemented mutant strains (Figure 3B; *P* < 0.05 and *P* > 0.05, respectively).

We also confirmed that the reduced levels of cell scattering induced in AGS cells by *H. pylori* ΔFLOT bacteria were due to reduced translocation of the CagA effector protein, a functional read-out of T4SS activity. Using an immunofluorescence-based technique to detect both extracellular and intracellular CagA, we observed that the levels of intracellular CagA were significantly reduced in AGS cells that had been co-cultured with *H. pylori* ΔFLOT or ΔcagA bacteria, when compared with cells co-cultured with either WT or ΔFLOT (FLOT+) bacteria (Figure 4; *P* < 0.02 and *P* < 0.008, respectively). Together, the data show that *H. pylori* ΔFLOT bacteria are significantly affected in their ability to induce CagA-dependent cell scattering and IL-8 responses in gastric epithelial cells, thereby confirming the importance of the *H. pylori* flotillin-like protein, HP0248, in T4SS functionality.

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**FIGURE 2** | The *H. pylori* flotillin-like protein, HP0248, is important for cholesterol accumulation and induction of cytokine responses in epithelial cells but not macrophages. (A) Levels of cholesterol in *H. pylori* 251 WT, ΔFLOT, and ΔFLOT (FLOT+) bacteria, as measured by the Amplex Red cholesterol detection kit. Data have been normalized to those for the WT strain and thus no error bars can be presented for this group. (B) Adhesion assays were performed on AGS gastric epithelial cells that had been co-cultured for 6 h with *H. pylori* WT 26695 or 251 strains, or the corresponding ΔFLOT mutants. Data are expressed as the % of bacteria that adhered. The results represent the average of three independent experiments. (C) AGS cell responses to *H. pylori* WT, ΔFLOT, or FLOT (FLOT+) bacteria. Non-stimulated cells (NS) served as negative controls. The supernatants were analyzed by ELISA for the detection of IL-8 at 24 h post-stimulation. (D) RAW 264.7 cell responses to *H. pylori* 251 WT, ΔFLOT, or FLOT (FLOT+) bacteria. The supernatants were analyzed by ELISA for IL-6, at 6 and 16 h post-stimulation. Data correspond to the mean ± S.E.M. (determined in triplicate) and are pooled from at least three independent experiments. Data were analyzed using the one-way ANOVA (A,C,D) or Mann-Whitney test (B) *P* < 0.05; **P** < 0.01; ***P** < 0.001; ΔP > 0.05.
**H. pylori** HP0248 Is Important for Establishment of a Chronic Infection in Mice

Finally, mouse infection studies were performed with *H. pylori* \( \Delta FLO T \) mutants that had been generated in the SS1 and X47-2AL mouse-colonizing strains (Grubman et al., 2010). A dramatic
and highly significant effect on colonization was observed for *H. pylori* SS1 ∆FLOT mutants, with no bacteria cultured from the gastric tissues of these mice at 30 days post-infection (Figure 5; *P < 0.0001*). Although we were unable to complement the FLOT mutation in *H. pylori* SS1 (data not shown), we were able to do so in another mouse-colonizing strain, *H. pylori* X47-2AL. Bacterial loads in X47-2AL FLOT-infected mice were significantly reduced with respect to those in WT-infected mice, albeit less strikingly than for SS1 (Figure 5; *P = 0.039*). Additionally, mice that were infected with the X47-2AL FLOT (FLOT+) strain exhibited significantly higher bacterial loads than animals infected with the ∆FLOT mutant (*P = 0.01*), but similar loads to those infected with the WT strain (*P = 0.549*). The fact that the FLOT mutation had a more dramatic effect on colonization in SS1 than in X47-2AL is similar to the findings of a previous study (Grubman et al., 2010), indicating likely strain-dependent differences. Taken together, the data demonstrate that HP0248 plays a role in chronic colonization of the mouse gastric mucosa.

**DISCUSSION**

It has been known for some time that *H. pylori* obtains host cell-derived cholesterol to generate cholesteryl glucosides which the bacterium incorporates into its own membrane (Ansorg et al., 1992; Hirai et al., 1995). These cholesteryl glucosides allow *H. pylori* to evade phagocytosis, the activation of a T-cell response and thus bacterial clearance (Wunder et al., 2006). It is becoming apparent that some bacteria (Mcgee et al., 2011; Trainor et al., 2011) use cholesteryl glucosides as an additional virulence factor (Figure 5). Interestingly, although the *H. pylori* FLOT-like protein, HP0248, is required for mouse colonization, *C57BL/6* mice were inoculated with *H. pylori* by intragastric gavage. Quantitative culture assays were performed at 30 days post-infection. Data are presented as Log10 colony forming units (CFU)/g stomach, with each point representing the mean of two estimations for a single mouse. Horizontal bars represent the geometric means of viable cell count determinations. The limit of detection for the assay is approximately 1,000 CFU/gram. Data for *H. pylori* SS1 are from three independent experiments, whereas those for X47-2AL are from 2 (WT, ∆FLOT) and 1 experiment (∆FLOT/FLOT+), respectively. Data were analyzed using the Mann-Whitney test *P < 0.05; ****P < 0.0001; ∆P > 0.05.*

![Figure 5](image)

It is well-established that cholesterol accumulates within membrane raft microdomains of eukaryotic cells. An important property of membrane rafts is that they can selectively include or exclude proteins to control interactions at the cell surface (Manes et al., 2003). It is becoming apparent that some bacteria possess domains analogous to eukaryotic membrane rafts in their outer membranes (Larocca et al., 2010; Lopez and Kolter, 2010). Additionally, it has been reported that homologs of the archetypal membrane raft molecule, flotillin, can be found in certain bacteria (Donovan and Bramkamp, 2009; Lopez and Kolter, 2010). For the first time, we have characterized a flotillin-like protein in a human bacterial pathogen and demonstrated its importance in pathogenesis. This protein, encoded by the *hp0248* gene in *H. pylori* 26695, was originally annotated as a putative flotillin-encoding gene, however, no evidence for this conclusion was provided (Bramkamp and Lopez, 2015). Interestingly, although we used the amino acid sequence of the *B. subtilis* flotillin-like protein YuaG (also known as FlOT) to identify *H. pylori* HP0248, it was suggested that HP0248 was likely to be more similar to the *B. subtilis* flotillin-like protein YqfA (or FloA) (Bramkamp and Lopez, 2015). It is therefore possible that in common with *B. subtilis* and other bacteria, *H. pylori* has two flotillin-like proteins, a fact that may explain the significant but relatively modest differences observed between the *H. pylori* WT and ∆FLOT mutants observed here. CLUSTALW analysis of *H. pylori* HP0248 revealed a highly conserved region encompassing the putative C-terminal flotillin domain, which is involved in the formation of membrane microdomains.
HP0248 (Supplementary Figure 1) showed that it has a predicted SPFH domain, characteristic of flotillin family members. By homologous recombination, we were able to generate *H. pylori* HP0248 mutants lacking all but 47 amino acids in the N-terminal region of the SPFH domain. In contrast, and despite two different mutagenesis approaches, all attempts to generate *H. pylori* HP0248 mutants lacking the entire SPFH domain were unsuccessful (data not shown). The precise reason for this observation remains unclear. One possibility is that the downstream gene, *hp0249*, which was used as a homology arm for recombination, may be an essential gene for *H. pylori*. The start codon of *hp0249* is situated only seven base pairs from the stop codon of *hp0248* and the two genes are likely to be part of the same transcriptional unit. Thus, any perturbation in *hp0249* expression may be lethal for the bacterium.

*H. pylori* can incorporate cholesterol into its cell membrane (Wunder et al., 2006) and thus it appeared likely that the bacterium may have membrane raft-like structures in its outer membrane. Based on the known property of membrane raft components to partition to DRM fractions, our data indicate that *H. pylori* HP0248 is a membrane raft-associated protein (Figure 1, Supplementary Tables 3, 4). Bioinformatic analyses suggest that this protein does not have a signal sequence but does have the predicted transmembrane region (data not shown) required for the association of SPFH proteins with the bacterial membrane (Bramkamp and Lopez, 2015). Proteomic analyses of the DRM preparations (Table 1, Supplementary Tables 3, 4) also revealed a selective enrichment of OMPs, including several of which are important for adhesion e.g., BabA, HopB, HopC, HopQ. In *S. aureus*, DRMs were reported to contain proteins required for biofilm formation, signaling, attachment, and virulence (Lopez and Kolter, 2010). Similarly, selective packaging of outer surface lipoproteins into membrane rafts has been reported in *B. burgdorferi* (Toledo et al., 2014). We hypothesize that specific proteins, required for *H. pylori* interactions with target cells and/or colonization, may compartmentalize within the membrane raft domains of this pathogen.

Eukaryotic flotillin is found within membrane raft domains and is important for various cellular functions (Simons and Ikonen, 1997). We therefore speculated whether the flotillin-like protein, HP0248, renamed here as FLOT, may hold a similar importance for *H. pylori* physiology. Indeed, *H. pylori* ΔFLOT mutants displayed ~40% less cholesterol in their membranes (Figure 2A), thus indicating that the protein is involved in cholesterol accumulation by the bacterium. Although a cholesterol receptor has yet to be identified in *H. pylori*, previous experimental evidence suggested that cholesterol uptake in the bacterium is protein-mediated (Trampenau and Muller, 2003). It is therefore possible that *H. pylori* FLOT may be one protein required for the uptake of cholesterol within the *H. pylori* cell membrane. We hypothesize that through its stabilization of membrane raft domains, *H. pylori* FLOT may be important for cholesterol accumulation within its cell membrane. In turn, these domains may be required for the proper formation and functioning of the T4SS apparatus. Consistent with this hypothesis, we demonstrated that T4SS-dependent functions, as determined by cell scattering responses and CagA translocation, were significantly affected in *H. pylori* ΔFLOT mutants (Figures 3, 4, respectively). Furthermore, we showed that these mutants were also significantly affected in their ability to colonize mice in a bacterial strain-dependent manner (Figure 5). Similar strain-dependent effects on colonization have been reported for other *H. pylori* mutants (Grubman et al., 2010) and are most likely reflective of the enormous genetic diversity in this bacterial species (Falush et al., 2003).

In conclusion, the flotillin-like protein, HP0248 appears to be critical for *H. pylori* pathogenesis, as not only is this protein important for the induction of T4SS-mediated host cell responses, but also for colonization. Although we have shown that HP0248 is involved in the optimal delivery of CagA into host cells and complete induction of IL-8 immune responses, it would be of interest to directly show whether T4SS-dependent delivery of peptidoglycan to host cells is reduced in the absence of HP0248. Furthermore, it remains to be elucidated whether the reduced amounts of cholesterol in the *H. pylori* ΔFLOT bacteria, or a combination of reduced cholesterol and the absence of FLOT, account for the lower host cell responses and decreased colonization levels observed. The difficulty in separating the contribution of cholesterol and FLOT to the phenotypes observed during this study has raised further questions that will form the basis of future investigations.

Toledo et al., 2014 (Figure 5). Similar strain-dependent effects on colonization have been reported for other *H. pylori* mutants (Grubman et al., 2010) and are most likely reflective of the enormous genetic diversity in this bacterial species (Falush et al., 2003).

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**AUTHOR CONTRIBUTIONS**

MH and KD performed and analyzed experiments and drafted initial versions of the manuscript. AR designed and constructed suicide vectors for construction of mutants and analyzed the SPFH domain of HP0248. LW, SM, and BC assisted with acquisition and interpretation of data. LT assisted with the construction of vectors for expression of FLOT protein and generation of FLOT mutants. DS performed and analyzed all proteomic analyses. MK assisted with the design of experiments.
RF conceived and coordinated the study and drafted the final version of the manuscript. All authors reviewed the results and approved the final version of the manuscript.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: http://journal.frontiersin.org/article/10.3389/fcimb.2017.00219/full#supplementary-material

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