Association of vitamin D receptor gene polymorphisms with gestational diabetes mellitus-a case control study in Wuhan, China

jianqiong liu
Maternal and Child Health Hospital of Hubei Province, Tongji Medical College, Huazhong University of Science and Technology

Qiong Dai
Maternal Child Health Hospital of Hubei Province, Tongji Medical College, Huazhong University of Science and Technology

Wei Li
Maternal and child health hospital of Hubei Province, Tongji Medical College, Huazhong University of Science and Technology

Yan Guo
Department of non-communicable chronic disease, Wuhan Centers for Disease Prevention and Control

Anna Dai
School of Basic Medical Sciences, Tongji Medical College, Huazhong University of Science and Technology

Yanqing Wang
Maternal and Child Health Hospital of Hubei Province, Tongji Medical College, Huazhong University of Science and Technology

Mengyao Deng
School of Medicine, Wuhan University of Science and Technology; Research Center for Health Promotion in Women, Youth and Children, Wuhan University of Science and Technology

Zhao Tang
School of Medicine, Wuhan University of Science and Technology; Research Center for Health Promotion in Women, Youth and Children, Wuhan University of Science and Technology

Lu She
School of Medicine, Wuhan University of Science and Technology; Research Center for Health Promotion in Women, Youth and Children, Wuhan University of Science and Technology

Xiaohong Chen
Maternal and Child Health Hospital of Hubei Province, Tongji Medical College, Huazhong University of Science and Technology

Mei Yang (✉ 644593079@qq.com)
Wuhan University of Science and Technology

https://orcid.org/0000-0003-4911-8205
Abstract

Background

Gestational diabetes mellitus (GDM) is extremely harmful to both the women and the fetuses. The association between the vitamin D receptor (VDR) gene and GDM has not been thoroughly investigated in Chinese pregnant women. Therefore, we aimed to determine whether VDR gene single nucleotide polymorphisms (SNPs) rs154410, rs7975232, rs731236, rs2228570 and rs739837 contribute to GDM risk in Wuhan, China. In addition, we aimed to explore their combined effect on the risk of GDM.

Methods

Pregnant women who had prenatal examination at 24 to 28 weeks’ gestation in our hospital were included in this case-control study. After exclusion, a total of 1684 pregnant women (826 GDM patients and 858 non-diabetic controls) were recruited. The clinical information and blood sample were collected by trained interviewers and nurses. Genotyping of candidate SNPs was conducted by the Sequenom MassARRAY platform. Statistical analyses such as t-test, chi-square test and logistic regression etc. were performed to the data with SPSS Software. Multifactor dimensionality reduction method was used to explore the gene-gene interactions on the risk of GDM.

Results

Differences in age, pre-pregnancy BMI, family history of diabetes and morning sickness between the case and control groups were statistically significant ($P<0.05$), whereas no significant differences were found in height, gravidity, parity, and age of menarche ($P>0.05$). There were no significant differences at allele and genotype distributions of the examined VDR gene SNPs ($P>0.05$). After adjusting by age, pre-pregnancy BMI, family history of diabetes and morning sickness, the results of logistic regression analysis showed no associations of the five SNPs with GDM in all the four genotype models ($P>0.05$). Furthermore, there were no gene-gene interactions on the GDM risk among the five examined VDR gene SNPs.

Conclusion

The VDR gene SNPs rs154410, rs7975232, rs731236, rs2228570 and rs739837 showed neither significant associations nor gene-gene interactions with GDM in Wuhan, China.

Background

Gestational diabetes mellitus (GDM) is defined as glucose intolerance with onset or first recognition during pregnancy [1]. The pooled prevalence of GDM which ranges from 5.4–14.8% depending on the
populations is increasing worldwide [2–5]. GDM is harmful to both the women and the fetuses. The primary four outcomes are macrosomia, primary cesarean delivery, clinical neonatal hypoglycemia, and fetal hyperinsulinemia. The secondary outcomes are premature delivery, shoulder dystocia or birth injury, need for intensive neonatal care, hyperbilirubinemia, and preeclampsia [6,7]. In severe cases, GDM can lead to prenatal death. Therefore, it is essential to identify potential risk factors of GDM for the health of women and children.

Although its exact etiology is unknown, genetic variations related to β-cell dysfunction and insulin resistance have been shown to contribute to the development of GDM [8,9]. Given the fact that women with a history of GDM are at an increased risk of developing type 2 diabetes (T2D) later in their lives [10] and women with a family history of diabetes may be predisposed to an increased risk of GDM [11], it is plausible to hypothesize that GDM may share the similar risk factors and genetic susceptibilities with T2D [12].

The vitamin D receptor (VDR) is a member of the large family of nuclear receptor transcription factors and specifically binds the micronutrient-derived hormone 1α,25(OH)₂D₃ [13]. The role for this receptor in T2D has been widely studied in recent years [14–17]. These findings have generated considerable interest in the association of VDR and GDM [18–22]. However, the conclusions were conflicting and the confounding factors and interactions between genetic polymorphisms were commonly neglected. Moreover, most analyses were limited by only examining one or two single nucleotide polymorphisms (SNPs). Therefore, according to genome-wide association studies of T2D, five SNPs rs154410, rs7975232, rs731236, rs2228570 and rs739837 were determined in the present case-control study along with their combined effects on the risk of GDM in Wuhan, China.

**Methods**

**Study population**

Pregnant women who had prenatal examination at the Obstetrics and Gynecology Clinic of Maternal and Child Hospital of Hubei Province from January 15, 2018 to March 31, 2019 were included in this case-control study. A total of 1684 pregnant women (826 GDM patients and 858 non-diabetic controls) were recruited in the study. The gestational age of participants, which was assessed from the date of the last menstrual period, was 24–28 weeks. A two-hour, 75g oral glucose tolerance test (OGTT) at 24 to 28 weeks’ gestation was performed for all participants, regardless of family history of diabetes or any other risk factors for GDM. The diagnosis of GDM was based upon the criteria of International Association of Diabetes and Pregnancy Study Groups (IADPSG): a fasting glucose ≥ 5.1 mmol/L (92 mg/dl), or a one-hour result of ≥ 10.0 mmol/L (180 mg/dl), or a two-hour result of ≥ 8.5 mmol/L (153 mg/dl) [7]. Exclusion criteria were: age<18 years; pre-gestational diabetes; multiple pregnancies; complicated pregnancy; chronic disease or any other medical condition that might affect glucose regulation. All subjects were unrelated Han Chinese and lived in Wuhan of Hubei Province, a central area of China.
Data Collection

A standard questionnaire was used by the trained interviewers to obtain information from all subjects regarding age, family history of diabetes, pregnant condition and other medical issues. Measurements of body weight and height were made for all subjects and body mass index (BMI) was calculated based on these measurements. Pre-pregnancy weight was obtained through medical records. The methods were carried out in accordance with the principles of the Declaration of Helsinki.

Selection and genotyping of SNPs

By tracking the literature, combined with genome-wide association studies of T2D and minor allele frequency (MAF)>0.05 reported in Chinese population, we selected five SNPs that were commonly investigated on the risk of GDM for assessment. These SNPs were rs1544410, rs731236, rs7975232, rs2228570 and rs739837. At recruitment, maternal blood samples in the fasted state (8 to 12h fast, no more than 12h) were collected by skilled nurses. After that, 2 ml blood were immediately placed on ice and separated into plasma and cells within 30 min, then distributed in aliquots and stored at -80 °C until analysis. Genomic DNA was isolated from 0.5 ml blood cells using the approved guideline of the Relax Gene Blood DNA System DP348 (Tiangen, China). Genotyping of candidate SNPs was conducted by the SequenomMassARRAY platform (Sequenom Inc., San Diego, CA, USA). For quality control, 5% of duplicate samples were independently reanalyzed in a blinded manner.

Statistical analysis

The differences among groups were compared by using unpaired Student’s t-test or analysis of variance (ANOVA). The differences in allele and genotype distribution as well as consistency of genotype distribution with Hardy-Weinberg equilibrium (HWE) were tested by using the chi-square test. Logistic regression was performed to evaluate the association of the genotypes and GDM risk. All P were two-sided and if below 0.05 the results were considered statistically significant. Analyses were conducted using SPSS Software, Version 24.0 for Windows (SPSS Inc., Chicago, IL, USA). Multifactor dimensionality reduction (MDR) [23] method was used to explore the gene-gene interactions on the risk of GDM.

Results

Clinical characteristics of subjects

The clinical characteristics of the study subjects were given in Table 1. The average age of GDM group and control group was 30.99±4.57 and 28.85±4.23 years, respectively. Differences in age, pre-pregnancy BMI, family history of diabetes and morning sickness between the case and control groups were statistically significant (P <0.05). The GDM patients had higher levels of age and pre-pregnancy BMI than the controls. No significant differences were found in height, gravidity, parity, and age of menarche between the GDM patients and controls (P >0.05).
### Table 1 Clinical characteristics of the subjects

|                      | GDM (n=826) | Non-GDM (n=858) | t/χ²       | P        |
|----------------------|-------------|-----------------|------------|----------|
| **Age**<sup>a</sup> (year) | 30.99±4.57  | 28.85±4.23      | 9.94       | <0.001   |
| **Height (cm)**      | 159.78±4.71 | 159.52±6.07     | 0.899      | 0.369    |
| **Pre-pregnancy BMI (kg/m<sup>2</sup>)** | 22.23±3.74  | 20.89±6.60      | 4.80       | <0.001   |
| **Family history of diabetes** |            |                 | 80.217     | <0.001   |
| No                   | 576(70.10)  | 742(87.90)      |            |          |
| Yes                  | 246(29.90)  | 102(12.10)      |            |          |
| **Gravidity**        |             |                 | 5.823      | 0.054    |
| 1                    | 289(35.90)  | 338(40.10)      |            |          |
| 2                    | 232(28.90)  | 254(30.10)      |            |          |
| ≥3                   | 283(35.20)  | 251(29.80)      |            |          |
| **Parity**           |             |                 | 1.114      | 0.291    |
| Nulliparous          | 484(58.70)  | 525(61.30)      |            |          |
| Multiparous          | 340(41.30)  | 332(38.70)      |            |          |
| **Age of menarche**<sup>b</sup> (year) | 13.40±1.40 | 13.34±1.31      | 0.947      | 0.344    |
| **Morning sickness** |             |                 | 37.420     | <0.001   |
| No                   | 302(38.00)  | 144(22.90)      |            |          |
| Yes                  | 493(62.00)  | 486(77.10)      |            |          |

Data were given as the mean ± SD or as n (%), with the significance of differences between groups evaluated using t-tests or the χ² test, respectively.

<sup>a</sup>Age refers to the age at which the participant was enrolled in the study.

<sup>b</sup>Age of menarche refers to the age at which the first menstruation took place.

Abbreviations: BMI, body mass index; GDM, gestational diabetes mellitus.

Bold represents significant P.

**Distribution of VDR gene SNPs**

The allele and genotype distributions of rs2228570, rs1544410, rs739837, rs731236 and rs7975232 among GDM patients and controls were shown in Table 2. There were no significant differences at allele
and genotype distributions of the five VDR gene SNPs ($P>0.05$). The distributions of five VDR gene SNPs in the control group were all in HWE ($P>0.05$).
| SNP       | GDM (n=826) | Non-GDM (n=858) | $\chi^2$ | $P$    | $\chi^2$ for HWE | $P$ for HWE |
|-----------|-------------|-----------------|---------|--------|------------------|-------------|
| rs2228570 |             |                 |         | 0.347  | 0.556            |             |
| G         | 801(49.20)  | 875(51.60)      | 1.899   | 0.168  |                  |             |
| A         | 827(50.80)  | 821(48.40)      |         |        |                  |             |
| GG        | 219(26.90)  | 230(27.10)      | 0.437   | 0.804  |                  |             |
| GA        | 389(47.80)  | 415(48.90)      |         |        |                  |             |
| AA        | 206(25.30)  | 203(23.90)      |         |        |                  |             |
| rs1544410 | 1.188       | 0.276           |         |        |                  |             |
| C         | 1567(95.80) | 1621(95.60)     | 0.084   | 0.772  |                  |             |
| T         | 69(4.20)    | 75(4.40)        |         |        |                  |             |
| CC        | 752(91.90)  | 776(91.50)      | -       | 0.933* |                  |             |
| CT        | 63(7.70)    | 69(8.10)        |         |        |                  |             |
| TT        | 3(0.40)     | 3(0.40)         |         |        |                  |             |
| rs739837  | 2.332       | 0.127           |         |        |                  |             |
| G         | 1152(70.60) | 1221(71.70)     | 0.538   | 0.463  |                  |             |
| T         | 480(29.40)  | 481(28.30)      |         |        |                  |             |
| GG        | 414(50.70)  | 447(52.50)      | 0.550   | 0.759  |                  |             |
| GT        | 324(39.70)  | 327(38.40)      |         |        |                  |             |
| TT        | 78(9.60)    | 77(9.10)        |         |        |                  |             |
| rs731236  | 3.630       | 0.057           |         |        |                  |             |
| A         | 1558(95.30) | 1623(95.60)     | 0.106   | 0.745  |                  |             |
| G         | 76(4.70)    | 75(4.40)        |         |        |                  |             |
| AA        | 745(91.20)  | 778(91.60)      | -       | 0.942* |                  |             |
| GA        | 68(8.30)    | 67(7.90)        |         |        |                  |             |
| GG        | 4(0.50)     | 4(0.50)         |         |        |                  |             |
| rs7975232 |             |                 | 1.501   | 0.221  |                  |             |
| C         | 1154(70.70) | 1220(71.90)     | 0.609   | 0.435  |                  |             |
Data were given as n (%).

Abbreviations: VDR, vitamin D receptor; SNPs, single nucleotide polymorphisms; GDM, gestational diabetes mellitus; HWE, Hardy-Weinberg Equilibrium.

-, no value because the frequency of CC, AA genotype was zero in this case; *, fisher’s exact test.

**Association between VDR gene SNPs and GDM**

The associations of these candidate VDR gene SNPs and GDM in different genotype models were shown in Table 3. The results showed that the associations of the five SNPs with GDM were not significant in different genotype models between cases and controls ($P>0.05$). To further evaluate the associations of these candidate SNPs and GDM, adjusted logistic regression analysis was also performed by age, pre-pregnancy BMI, family history of diabetes, morning sickness. The results showed that the associations of the five SNPs with GDM were still not significant in all the genotype models ($P>0.05$).
| Table 3 Association between VDR gene SNPs and the risk of GDM |
|-------------------------------------------------------------|
| **Crude OR (95% CI)** | **Crude P** | **Adjusted OR (95% CI)** | **Adjusted P** |
|------------------------|-------------|-------------------------|----------------|
| **rs2228570**          |             |                         |                |
| Co-dominant model      |             |                         |                |
| GG                    | 1(ref.)     | 1(ref.)                 |                |
| GA                    | 0.984(0.781-1.240) | 0.894       | 0.916(0.697-1.206) | 0.533       |
| AA                    | 1.066(0.815-1.393) | 0.641       | 2.397(0.728-1.379) | 0.989       |
| Dominant Model         |             |                         |                |
| GG                    | 1(ref.)     | 1(ref.)                 |                |
| GA+AA                 | 1.011(0.814-1.256) | 0.92        | 0.944(0.730-1.221) | 0.663       |
| Recessive Model        |             |                         |                |
| GG+GA                 | 1(ref.)     | 1(ref.)                 |                |
| AA                    | 1.077(0.861-1.346) | 0.517       | 1.059(0.811-1.384) | 0.672       |
| Over-dominant model    |             |                         |                |
| GG+AA                 | 1(ref.)     | 1(ref.)                 |                |
| GA                    | 0.955(0.788-1.158) | 0.639       | 0.915(0.728-1.152) | 0.451       |
| **rs1544410**          |             |                         |                |
| Co-dominant model      |             |                         |                |
| CC                    | 1(ref.)     | 1(ref.)                 |                |
| CT                    | 0.942(0.660-1.345) | 0.743       | 0.979(0.645-1.485) | 0.921       |
| TT                    | 1.032(0.208-5.129) | 0.969       | 0.636(0.127-3.194) | 0.583       |
| Dominant Model         |             |                         |                |
| CC                    | 1(ref.)     | 1(ref.)                 |                |
| CT+TT                 | 0.946(0.667-1.341) | 0.755       | 0.955(0.637-1.433) | 0.824       |
| Recessive Model        |             |                         |                |
| CC+CT                 | 1(ref.)     | 1(ref.)                 |                |
| TT                    | 1.037(0.209-5.152) | 0.965       | 0.637(0.127-3.198) | 0.584       |
| Over-dominant model    |             |                         |                |
| rs739837 | Co-dominant model |  |  |
| --- | --- | --- | --- |
| GG | 1(ref.) | 1(ref.) |  |
| GT | 1.070(0.873-1.311) | 0.516 | 1.102(0.863-1.408) | 0.435 |
| TT | 1.094(0.777-1.540) | 0.608 | 1.123(0.795-1.663) | 0.562 |
| rs731236 | Co-dominant model |  |  |
| AA | 1(ref.) | 1(ref.) |  |
| GA | 1.060(0.745-1.507) | 0.746 | 1.161(0.768-1.754) | 0.48 |
| GG | 1.044(0.260-4.191) | 0.951 | 0.489(0.108-2.209) | 0.352 |

|  |  |  |  |
| --- | --- | --- | --- |
| rs739837 | Dominant Model |  |  |
| GG | 1(ref.) | 1(ref.) |  |
| GT+TT | 1.074(0.887-1.302) | 0.464 | 1.107(0.880-1.392) | 0.386 |
| rs731236 | Dominant Model |  |  |
| AA | 1(ref.) | 1(ref.) |  |
| GA+GG | 1.059(0.752-1.492) | 0.743 | 1.098(0.736-1.637) | 0.648 |
| rs739837 | Recessive Model |  |  |
| GG+GT | 1(ref.) | 1(ref.) |  |
| TT | 1.062(0.763-1.479) | 0.72 | 1.078(0.738-1.574) | 0.698 |
| rs731236 | Recessive Model |  |  |
| AA+GA | 1(ref.) | 1(ref.) |  |
| GG | 1.039(0.259-4.170) | 0.957 | 0.483(0.107-2.180) | 0.344 |
| rs739837 | Over-dominant model |  |  |
| GG+GT | 1(ref.) | 1(ref.) |  |
|  |  |  |  |
| rs731236 | Over-dominant model |  |  |
| AA+GG | 1(ref.) | 1(ref.) |  |
### rs7975232

**Co-dominant model**

| Genotype | Sensitivity | Specificity | $\chi^2$ | $P$ | OR (95% CI) | Kappa |
|----------|-------------|-------------|----------|-----|-------------|-------|
| CC       | 1(ref.)     | 1(ref.)     |          |     |             |       |
| CA       | 1.062(0.866-1.301) | 0.565      |          |     | 1.095(0.857-1.398) | 0.468 |
| AA       | 1.118(0.791-1.580) | 0.527      |          |     | 1.187(0.796-1.769) | 0.401 |

**Dominant Model**

| Genotype | Sensitivity | Specificity | $\chi^2$ | $P$ | OR (95% CI) | Kappa |
|----------|-------------|-------------|----------|-----|-------------|-------|
| CC       | 1(ref.)     | 1(ref.)     |          |     |             |       |
| CA+AA    | 1.072(0.884-1.299) | 0.479      |          |     | 1.113(0.885-1.400) | 0.361 |

**Recessive Model**

| Genotype | Sensitivity | Specificity | $\chi^2$ | $P$ | OR (95% CI) | Kappa |
|----------|-------------|-------------|----------|-----|-------------|-------|
| CC+CA    | 1(ref.)     | 1(ref.)     |          |     |             |       |
| AA       | 1.090(0.780-1.523) | 0.614      |          |     | 1.142(0.776-1.680) | 0.5    |

**Over-dominant model**

| Genotype | Sensitivity | Specificity | $\chi^2$ | $P$ | OR (95% CI) | Kappa |
|----------|-------------|-------------|----------|-----|-------------|-------|
| CC+AA    | 1(ref.)     | 1(ref.)     |          |     |             |       |
| CA       | 1.044(0.857-1.271) | 0.668      |          |     | 1.065(0.841-1.349) | 0.599 |

Adjusted OR is adjusted for age, pre-pregnancy BMI, family history of diabetes, morning sickness.

Abbreviations: BMI: body mass index; OR: odds ratio; CI: confidence interval; ref: reference genotype

**Gene-gene interactions to GDM**

The analysis of gene-gene interactions indicated that both two-factor model and three-factor model had good cross-validation consistency at 9/10, but the test accuracy of the two-factor model (0.514) was higher than that of the three-factor model (0.511), so the best model was the two-factor gene-gene interaction model. However, as was shown in Table 4, there was no significance of the test set in the two-factor gene-gene interaction ($P>0.05$). Therefore, it could be speculated that there were no gene-gene interactions on the GDM risk among the five VDR gene SNPs.

### Table 4 Interaction of two-factor gene-gene model

|            | Sensitivity | Specificity | $\chi^2$ | $P$    | OR (95% CI)         | Kappa |
|------------|-------------|-------------|----------|--------|---------------------|-------|
| Training set | 0.369       | 0.669       | 6.920    | 0.009  | 1.323(1.074-1.630)  | 0.065 |
| Test set   | 0.377       | 0.652       | 0.154    | 0.695  | 1.133(0.607-2.116)  | 0.029 |
| Total set  | 0.412       | 0.652       | 7.202    | 0.007  | 1.308(1.075-1.591)  | 0.063 |
Discussion

In the present study, we analyzed the association of VDR gene SNPs rs154410, rs7975232, rs731236, rs2228570 and rs739837 with GDM in Wuhan, China. It was revealed that, VDR gene polymorphic markers were not found to be associated with GDM in central Chinese population. Furthermore, there were no gene-gene interactions on the GDM risk among the examined VDR gene SNPs.

The rs739837 SNP is located at the three-primer untranslated region (3′-UTR) of the VDR gene. This region does not affect amino acid sequence and is not likely to affect the function of the gene\[^{24}\]. However, the variant rs739837 might affect the expression of VDR gene by binding with microRNA\[^{25}\]. Several studies investigated the relationship between this locus and T2D and reported that rs739837 was associated with susceptibility to T2D\[^{15,17,24}\]. To date, two studies had studied the relationship between rs739837 and GDM. Shi et al. reported no relationship between the genotypic model of rs739837 and GDM, whereas Wang et al. found a statistical correlation between the rs739837 polymorphism and GDM risk\[^{26,27}\]. However, neither of the two studies had analyzed the combined effect with other VDR gene SNPs. As susceptibility was attributable not to a single polymorphism or allele, but rather to multiple polymorphisms\[^{28}\], we evaluated rs739837 and other four widely studied VDR gene SNPs and their gene-gene interactions on the risk of GDM. The result showed that there was no statistical correlation between rs739837 and GDM. Besides, no evidence was found at the gene-gene interactions. To our knowledge, this study is the first to investigate the combined effect between rs739837 and other VDR gene SNPs on the risk of GDM. The results need to be verified in future studies.

The rs1544410, rs7975232, and rs731236 SNPs are known as BsmI, ApaI and TaqI according to their restriction enzymes. A meta-analysis on the three polymorphisms with the risk of T2D produced negative results\[^{29}\]. As for the association between the three polymorphisms and GDM, the results were inconsistent\[^{18,19,22,30}\]. Our study reported a negative result, which was consistent with the finding of Apaydin et al.\[^{18}\] BsmI, Apal and TaqI are all located in the 3′-UTR and have been shown to be in strong linkage disequilibrium\[^{29}\]. Polymorphisms of TaqI, BsmI and Apal are probably non-functional because they are either located in intron (BsmI and Apal in intron 8), which will be removed during mRNA post transcriptional modification or result in no amino acid sequence change (TaqI in exon 9).

The rs2228570 is known as FokI according to its restriction enzyme. FokI polymorphism was linked to risk of GDM in Turkish women and Iranian population\[^{18,31}\]. However, studies in other countries could not establish association between FokI and GDM\[^{19,20}\]. In the present study, we reported no evidence of allelic or genotypic association of the FokI polymorphism with GDM in central Chinese population. The FokI is located at the 5′ end region of the VDR gene. It is reported as an independent marker of the VDR gene because it has not been shown to be in linkage disequilibrium with any other VDR polymorphisms\[^{31}\]. It produces either a 424 or a 427 amino acid VDR protein. These two isoforms are thus structurally
distinct, unlike those VDR gene that contain polymorphisms present in the 3′-portion of the gene that are either silent codon changes or are found in introns or in the 3′-untranslated regions \[^{32}\]. Even the relationship between the FokI polymorphism and T2D is still controversial \[^{14–16, 29}\], probably because the socio-demographic characteristics, experimental methods, and sample size are different in the studied populations.

Our study had several strengths. First, we employed MDR method to explore the gene-gene interactions on the GDM risk among the selected SNPs. The identification and characterization of gene-gene interactions had been limited mainly by a lack of powerful statistical methods and a lack of large sample size \[^{33}\]. To overcome these limitations, the MDR method was developed. It was used for detecting and characterizing high-order gene to gene interactions \[^{34}\] and was shown to have good power in relatively small case-control studies \[^{23, 35}\]. Second, we adjusted potential confounding factors such as age, pre-pregnancy BMI etc. to explore the association in different genotype models. At last, we used a relatively large sample size, which was able to provide enough statistical power.

However, there were also some limitations in this study. First, the level of plasma vitamin D was not measured in all subjects. Second, the information of environmental and lifestyle factors was lacked, which had been reported recently to be important determinants of GDM development \[^{36, 37}\]. Finally, there were some potential biases that came from the cross-sectional nature of the case–control study. Thus, cohort studies concerning the above-mentioned factors will be required in future to validate the findings of the study.

**Conclusions**

The VDR gene SNPs rs154410, rs7975232, rs731236, rs2228570 and rs739837 showed non-significant associations with GDM in central Chinese population. Furthermore, there were no gene-gene interactions on the GDM risk among these SNPs.

**Abbreviations**

GDM: gestational diabetes mellitus; T2D: type 2 diabetes; VDR: vitamin D receptor; SNPs: single nucleotide polymorphisms; OGTT: oral glucose tolerance test; IADPSG: International Association of Diabetes and Pregnancy Study Groups; BMI: body mass index; MAF: minor allele frequency; ANOVA: analysis of variance; HWE: Hardy-Weinberg Equilibrium; MDR: Multifactor dimensionality reduction; OR: odds ratio; CI: confidence interval; 3′-UTR: three-primer untranslated region

**Declarations**

**Ethics approval and consent to participate**
The study was approved by the institutional review board of Wuhan University of Science and Technology. All subjects provided written consents for participation.

Consent for publication

Not applicable

Availability of data and materials

The datasets used and analyzed during the current study are available from the corresponding author on reasonable request.

Competing interests

The authors declare that they have no competing interests.

Funding

This work was supported by The National Natural Science Fund of China (81703239), Chinese Center for Disease Control and Prevention (2018FYH014) and Health Commission of Hubei Province (WJ2018H0134, WJ2018H0145). The design of the study, analysis and interpretation of data were mainly supported by The National Natural Science Fund of China; the collection and analysis of blood samples was supported by Chinese Center for Disease Control and Prevention; the investigation to subjects and publication of manuscripts were supported by Health commission of Hubei Province.

Authors’ contributions

JL, QD contributed to the design of study, analysis and interpretation of data, and drafted the manuscript; WL, YG, AD, YW, MD, ZT and LS participated in the acquisition, analysis and interpretation of data; XC and MY involved in the design and coordination of the study and revised the manuscript critically for important intellectual content. All authors read and approved the final manuscript.

Acknowledgements

The authors appreciated all the study participants, hospital workers, research staff and students who participated in this work.

References

1. Metzger BE, Buchanan TA, Coustan DR, de Leiva A, Dunger DB, Hadden DR, Hod M, Kitzmiller JL, Kjos SL, Oats JN, et al: Summary and recommendations of the Fifth International Workshop-Conference
on Gestational Diabetes Mellitus. Diabetes Care 2007, 30 Suppl 2:S251-260.

2. Ferrara A. Increasing prevalence of gestational diabetes mellitus: a public health perspective. Diabetes Care. 2007;30(Suppl 2):141–6.

3. Gao C, Sun X, Lu L, Liu F, Yuan J. Prevalence of gestational diabetes mellitus in mainland China: a systematic review and meta-analysis. J Diabetes Investig. 2019;10(1):154–62.

4. Eades CE, Cameron DM, Evans JMM. Prevalence of gestational diabetes mellitus in Europe: a meta-analysis. Diabetes Res Clin Pract. 2017;129:173–81.

5. Mwanri AW, Kinabo J, Ramaiya K, Feskens EJ. Gestational diabetes mellitus in sub-Saharan Africa: systematic review and metaregression on prevalence and risk factors. Trop Med Int Health. 2015;20(8):983–1002.

6. Metzger BE, Lowe LP, Dyer AR, Trimble ER, Chaovarindr U, Coustan DR, Hadden DR, McCance DR, Hod M, McIntyre HD, et al. Hyperglycemia and adverse pregnancy outcomes. N Engl J Med. 2008;358(19):1991–2002.

7. Wendland EM, Torloni MR, Falavigna M, Trujillo J, Dode MA, Campos MA, Duncan BB, Schmidt MI. Gestational diabetes and pregnancy outcomes—a systematic review of the World Health Organization (WHO) and the International Association of Diabetes in Pregnancy Study Groups (IADPSG) diagnostic criteria. BMC Pregnancy Childbirth. 2012;12:23.

8. Kwak SH, Kim SH, Cho YM, Go MJ, Cho YS, Choi SH, Moon MK, Jung HS, Shin HD, Kang HM, et al. A genome-wide association study of gestational diabetes mellitus in Korean women. Diabetes. 2012;61(2):531–41.

9. Wu L, Cui L, Tam WH, Ma RC, Wang CC. Genetic variants associated with gestational diabetes mellitus: a meta-analysis and subgroup analysis. Scientific reports. 2016;6:30539.

10. Bellamy L, Casas JP, Hingorani AD, Williams D. Type 2 diabetes mellitus after gestational diabetes: a systematic review and meta-analysis. Lancet. 2009;373(9677):1773–9.

11. Williams MA, Qiu C, Dempsey JC, Luthy DA. Familial aggregation of type 2 diabetes and chronic hypertension in women with gestational diabetes mellitus. J Reprod Med. 2003;48(12):955–62.

12. Mao H, Li Q, Gao S. Meta-analysis of the relationship between common type 2 diabetes risk gene variants with gestational diabetes mellitus. PLoS One. 2012;7(9):e45882.

13. Tuoresmaki P, Vaisanen S, Neme A, Heikkinen S, Carlberg C. Patterns of genome-wide VDR locations. PLoS One. 2014;9(4):e96105.

14. Bid HK, Konwar R, Aggarwal CG, Gautam S, Saxena M, Nayak VL, Banerjee M: Vitamin D. receptor (FokI, BsmI and TaqI) gene polymorphisms and type 2 diabetes mellitus: a North Indian study. Indian J Med Sci. 2009;63(5):187–94.

15. Jia J, Ding H, Yang K, Mao L, Zhao H, Zhan Y, Shen C. Vitamin D receptor genetic polymorphism is significantly associated with risk of type 2 diabetes mellitus in Chinese Han population. Arch Med Res. 2015;46(7):572–9.
16. Malecki MT, Frey J, Moczulski D, Klupa T, Kozek E, Sieradzki J. Vitamin D receptor gene polymorphisms and association with type 2 diabetes mellitus in a Polish population. Exp Clin Endocrinol Diabetes. 2003;111(8):505–9.

17. Xu JR, Yang Y, Liu XM, Wang YJ. Association of VDR polymorphisms with type 2 diabetes mellitus in Chinese Han and Hui populations. Genet Mol Res. 2014;13(4):9588–98.

18. Apaydin M, Beysel S, Eyerici N, Pinarli FA, Ulubay M, Kizilgul M, Ozdemir O, Caliskan M, Cakal E. The VDR gene FokI polymorphism is associated with gestational diabetes mellitus in Turkish women. BMC Med Genet. 2019;20(1):82.

19. El-Beshbishy HA, Tawfeek MA, Taha IM, FadulElahi T, Shaheen AY, Bardi FA, Sultan II. Association of vitamin D receptor gene BsmI (A > G) and FokI (C > T) polymorphism in gestational diabetes among Saudi Women. Pak J Med Sci. 2015;31(6):1328–33.

20. Ghamdi, Al M. Association of vitamin D receptor gene polymorphisms with gestational diabetes in pregnant women. Wulfenia. 2014;21(9):7.

21. Jin L-P, Shi L-J, Zhang W-P. The correlation study on vitamin D receptor Apal gene polymorphism and gestational diabetes mellitus among pregnant women. Biomed Res. 2017;28(7):4.

22. Rahmannezhad G, Mashayekhi FJ, Goodarzi MT, Rezvanfar MR, Sadeghi A. Association between vitamin D receptor Apal and TaqI gene polymorphisms and gestational diabetes mellitus in an Iranian pregnant women population. Gene. 2016;581(1):43–7.

23. Hahn LW, Ritchie MD, Moore JH. Multifactor dimensionality reduction software for detecting gene-gene and gene-environment interactions. Bioinformatics. 2003;19(3):376–82.

24. Yu F, Wang C, Wang L, Jiang H, Ba Y, Cui L, Wang Y, Yu S, Li W. Study and evaluation the impact of vitamin D receptor variants on the risk of type 2 diabetes mellitus in Han Chinese. J Diabetes. 2017;9(3):275–84.

25. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. Cell. 2004;116(2):281–97.

26. Shi A, Wen J, Liu G, Liu H, Fu Z, Zhou J, Zhu Y, Liu Y, Guo X, Xu J. Genetic variants in vitamin D signaling pathways and risk of gestational diabetes mellitus. Oncotarget. 2016;7(42):67788–95.

27. Wang Y, Wang O, Li W, Ma L, Ping F, Chen L, Nie M. Variants in vitamin D binding protein gene are associated with gestational diabetes mellitus. Medicine. 2015;94(40):e1693.

28. Horikawa Y, Oda N, Cox NJ, Li X, Orho-Melander M, Hara M, Hinokio Y, Lindner TH, Mashima H, Schwarz PE, et al. Genetic variation in the gene encoding calpain-10 is associated with type 2 diabetes mellitus. Nat Genet. 2000;26(2):163–75.

29. Li L, Wu B, Liu JY, Yang LB. Vitamin D receptor gene polymorphisms and type 2 diabetes: a meta-analysis. Arch Med Res. 2013;44(3):235–41.

30. Zhu B, Huang K, Yan S, Hao J, Zhu P, Chen Y, Ye A, Tao F: VDR variants rather than early pregnancy vitamin D concentrations are associated with the risk of gestational diabetes: the Ma’anashan birth cohort (MABC) study. J Diabetes Res 2019, 2019:8313901.
31. Aslani S, Hossein-Nezhad A, Mirzaei K, Maghbooli Z, Afshar AN, Karimi F. VDR FokI polymorphism and its potential role in the pathogenesis of gestational diabetes mellitus and its complications. Gynecological endocrinology: the official journal of the International Society of Gynecological Endocrinology. 2011;27(12):1055–60.

32. Jurutka PW, Remus LS, Whitfield GK, Thompson PD, Hsieh JC, Zitzer H, Tavakkoli P, Galligan MA, Dang HT, Haussler CA, et al: The polymorphic N terminus in human vitamin D receptor isoforms influences transcriptional activity by modulating interaction with transcription factor IIβ. Molecular endocrinology (Baltimore, Md) 2000, 14(3):401–420.

33. Cho YM, Ritchie MD, Moore JH, Park JY, Lee KU, Shin HD, Lee HK, Park KS. Multifactor-dimensionality reduction shows a two-locus interaction associated with type 2 diabetes mellitus. Diabetologia. 2004;47(3):549–54.

34. Ritchie MD, Hahn LW, Roodi N, Bailey LR, Dupont WD, Parl FF, Moore JH. Multifactor-dimensionality reduction reveals high-order interactions among estrogen-metabolism genes in sporadic breast cancer. Am J Hum Genet. 2001;69(1):138–47.

35. Ritchie MD, Hahn LW, Moore JH. Power of multifactor dimensionality reduction for detecting gene-gene interactions in the presence of genotyping error, missing data, phenocopy, and genetic heterogeneity. Genet Epidemiol. 2003;24(2):150–7.

36. Carroll X, Liang X, Zhang W, Zhang W, Liu G, Turner N, Leeper-Woodford S. Socioeconomic, environmental and lifestyle factors associated with gestational diabetes mellitus: a matched case-control study in Beijing, China. Scientific reports. 2018;8(1):8103.

37. He JR, Yuan MY, Chen NN, Lu JH, Hu CY, Mai WB, Zhang RF, Pan YH, Qiu L, Wu YF, et al. Maternal dietary patterns and gestational diabetes mellitus: a large prospective cohort study in China. Br J Nutr. 2015;113(8):1292–300.