Differential Expression of \( V_n \) Gene Families in Peripheral B Cell Repertoires of Newborn or Adult Immunoglobulin H Chain Congenic Mice

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Summary

The pattern of \( V_n \) gene family expression in the primary B cell repertoire of the mouse is strain dependent. In C57B1/6 mice, the \( V_n \) J558 family is expressed by more than 45% of the cells, while the expression of \( V_n \) 7183, \( V_n \) Q52, and \( V_n \) 36-60 families together does not exceed 20%. In BALB/c mice, relative expression of \( V_n \) J558 is lower than 35%, while the sum of the other three families reaches 25%. To assess which genetic loci control strain-specific \( V_n \) gene family expression, we studied \( V_n \) gene family usage in splenic B cell repertoires of different congenic strains of mice. Changes in major histocompatibility complex or immunoglobulin (Ig) K light chain genes did not modify \( V_n \) gene family expression in adult mice. Differences at the IgH locus, however, modified \( V_n \) gene family usage. In 1-d-old mice, the strain-specific \( V_n \) gene family expression pattern is determined by the IgH haplotype. In adult mice, the \( V_n \) gene family expression pattern of resting B cells is independent of the IgH locus and follows the genetic background of the congenic strain, while it is determined by the IgH haplotype among Ig-secreting spleen cells. In F1(B6 × BALB/c) mice, each of the two spleen B cell populations, sorted on the basis of \( \mu \) heavy chain allotype expression, shows an independent \( V_n \) gene family expression pattern, determined by the IgH locus. The implications of these results in the control of \( V_n \) gene family expression, and in the selection of peripheral B cell repertoires are discussed.

Variable regions of Ig are generated by recombination of multiple \( V \) (D) and \( J \) genetic segments (1). Combinatorial association of these elements together with junctional diversity originate a potential B cell repertoire with more than \( 10^6 \) different Ig combining sites (2). In mice, \( H \) chain variable gene segments (\( V_n \))\(^1\) are clustered in subgroups or families of genes sharing more than 80% of nucleotide sequence homology (3). Twelve \( V_n \) gene families have been described with a chromosomal organization within the IgH locus which is conserved across species (4). The mouse strains C57Bl/6 and BALB/c carry different IgH haplotypes (5) of roughly the same genetic complexity (3-6), although the actual number of different active genes per \( V_n \) gene family, in each mouse strain, remains to be established (3, 7, 8).

\( V_n \) gene family usage does not follow genetic complexity, but varies with the mouse strain (9-11), and is influenced by genetic factors unlinked to the IgH locus (12). Within the same inbred strain, \( V_n \) gene family expression is genetically controlled during ontogeny (13-17) and lymphopoiesis (18, 19), varies with cellular environment (17), and is influenced by T-B cell interactions (20). Nevertheless, the strain-specific pattern of \( V_n \) gene family expression is stable for age-matched male or female individuals.

In a normal (nonimmunized) mouse, peripheral B cell repertoires are continuously renewed and selected (21). In fact, the number of newly formed B cells produced every day is estimated to be \( 2-5 \times 10^7 \) cells (22, 23). A part of these cells will integrate the peripheral B cell pool (available repertoire), which consists of \( <10^6 \) different clones of resting immunocompetent B cells (24). Moreover, only a fraction (1% or less) of all peripheral B cells are naturally Ig-secreting cells (actual repertoire) (25). Hence, the stable expression of \( V_n \) gene families in the context of this high turnover of B cells must result from the continuous selection of peripheral B cells leading to the strain-specific \( V_n \) gene family usage.

In this study, we investigated which genetic loci could control the strain-specific \( V_n \) gene family expression. We quantified \( V_n \) gene family expression, by in situ hybridization, both in neonatal and in adult peripheral B cell repertoires of different strains of mice congenic for the MHC, IgK \( L \) and \( H \)-chain loci. We observed that changes in IgH or IgK \( L \) chain loci did not modify \( V_n \) gene family expression of peripheral mature B cells. In IgH congenic mice, however, the \( V_n \) gene family usage among splenic resting B cells (available repertoire) changes from 1-d-old newborn, where it is determined by the IgH locus, to adult individuals, where it follows the genetic background of the congenic strain. We also found that \( V_n \) gene family expression among splenic Ig-
secretion of IgM secreting cells (actual repertoire) of adult IgH congenic mice is determined by the IgH locus, and thus differs from the adult repertoire. Finally, studies in F1(C57Bl/6 × BALB/c) mice, sorting adult splenocytes on the basis of their IgM allotype expression, showed that the Vµ gene family usage by each allotype sorted population is determined by the IgH locus.

Materials and Methods

Animals. Inbred strains of mice, raised in our colony at the Pasteur Institute, were used at either 1 d after birth or at 2–3 mo of age. The following strains of mice were studied: C57Bl/6 (IgHβ, H-2b), BALB/c (IgHβ, H-2a), Fl(C57Bl/6 × BALB/c), B6lghb1 (IgHβ, H-2b), CB20 (IgHβ, H-2a), BALB.B10 (IgHβ, H-2a), BALB.C3H (IgHβ, H-2a), and AKR (H-2d). The IgK L chain congenic strains used were: C.AKR and C.C58, bearing the K L-chain locus of AKR and C58, respectively, and in the BALB/C genetic background (26). For each experiment, pools of 2–3 age-matched animals were used.

Cells. Spleen cell suspensions from adult or newborn mice were prepared as described (18) and used for in situ hybridization, either ex vivo, to assay for IgM-secreting spleen cells, or after in vitro stimulation with LPS from Salmonella typhimurium at 25 µg/ml at 106/ml. It has been previously shown that LPS activation does not bias Vµ gene family expression (17). LPS-activated cells were harvested on day 3 of culture, cytopspun, and fixed onto slides as described in detail elsewhere (17). Sorted F1 spleen cells, when stimulated with LPS, were cultured for 4–5 d at 0.2 x 106/ml with 3 x 106/ml irradiated (100 rad) rat thymocytes. The Vµ gene family expression did not vary with the different protocols used for cell culture (not shown). Limiting dilution analysis of LPS-reactive cells was performed as described previously (27). The frequency of LPS-reactive B cells bearing each µ allotype was determined in sorted F1 B cells for quantitating the degree of purity of the sorted cell populations.

Flow Cytometry and Cell Sorting. Cells were stained with FITC-RS3.1 (anti-µ+), reference 28 and biotin-labeled MB86 (anti-µ-), reference 29) mAbs, followed by streptavidin-PE from Becton Dickinson & Co. (Grenoble, France). Dead cells were excluded from analysis by light-scatter gating. All analyses were performed on a FACStar Plus cell sorter (Becton Dickinson & Co., Sunnyvale, CA) interfaced to a Hewlett-Packard (Palo Alto, CA) computer. Cells were positively sorted for both IgM allotypes. The purity of each sample was controlled by limiting dilution analysis, and contamination never exceeded 4%.

Probes. The Cµ and Vµ gene probes were kind gifts of Drs. F. Alt, S. Riley, J. Kneary, U. Krawinkel, L. A. Reinginer, M. Sims, and R. Kofler. The Vµ gene family probes used (with the family name in parenthesis) were 81-X (Vµ 7183), 300-19 (Vµ 36-60), VNPB4 (Vµ 558), PBV14 (Vµ 606), MS1 (VGMAM3.8), V31 (V31/V, 36-09), CPI2 (Vµ, 11), and have been previously described (17). The VMS1 (VGMAM3.8) probe is a 830-bp HaelII fragment 98% homologous to VGMAM3.8, and the Vµ,10 (Vµ 10) probe is a 483-bp PsI fragment (31).

In Situ Hybridization. In situ hybridization was performed using a modification of the method described by Haase et al. (32), as detailed by Freitas et al. (17). cDNA probes were labeled by the random priming method (33) with 32P-dCTP (500 Ci/mmol, Amersham International, Amersham, Bucks, UK) and specific hybridization of probes with intracytoplasmic mRNA was revealed by autoradiography in a Kodak emulsion NTB2 (Eastman Kodak, Rochester, NY). Positive cells were covered by silver grains and analyzed by light microscopy.

Specificity of the Vµ Gene Family Probes. The family specificity of the Vµ gene family probes and experimental conditions were established by hybridization to cell lines expressing known Vµ of the different families (17). It must be noted that although cross-hybridization cannot be excluded between at least some members of the Vµ X24 and 7183 families, or the Vµ J558 and VGAM3.8 families, the lack of correlation in the expression of these families shows that the respective probes identify distinct cell populations (17, 20, 34). The absolute numbers of positive cells with all Vµ gene family probes correlated well with the numbers of cells positive with the Cµ probe, representing between 70% (in LPS-activated cells) and 100–120% (in non-LPS-stimulated spleen cells) of the latter, but showing that cells expressing isotypes other than µ are detected. Hence, Vµ gene family usage in the different cell populations studies was expressed as a relative distribution, calculated by the percentage of positive cells for each Vµ gene family out of the total number of positive cells detected with all Vµ gene family probes.

Detection Limits of in situ Hybridization. All Cµ and Vµ positive cells had the morphology of blasts or plasma cells, and the number of Cµ positive cells correlated well (r = 0.83–0.92) with the number of IgM-secreting cells, as determined by the protein A-plateau forming cell assay (17). It follows that our conditions of in situ hybridization exclusively score activated cells, with levels of mRNA severalfold higher than those of resting B cells. We could, therefore, discriminate Vµ gene family usage in the actual repertoire (by directly assaying IgM-secreting cells ex vivo) versus the available repertoire (by prior in vitro activation of resting B cells with a polyclonal mitogen).

Evaluation of the Number of Positive Cells. The number of Vµ positive cells were scored by counting between 30 and 100 microscopic fields (a total of 2,000–30,000 cells scored), as well as the mean number of total cells per field, from which the frequency of positive cells was calculated. The frequency of positive cells in splenic cell suspensions directly assayed ex vivo was 0.1–0.5%, while after LPS stimulation this frequency increased to 20–30% of total cells. A total of at least 50–200 (ex vivo) or 500–2,000 (after LPS stimulation) Vµ positive cells were counted. A cell was scored as positive if it contained 20 or more silver grains in a background with a mean grain number per cell <5. In most instances, positive cells were heavily labeled with an uncountable number of grains.

Statistical Analysis of the Results. Statistical analysis of the differences between cell populations were done using a one way ANOVA computer program (Unité d’Informatique Scientifique, Institut Pasteur, Paris, France).

Results

Peripheral B Cell Repertoires in C57Bl/6 and BALB/c Mice. Vµ gene family usage by LPS-activated splenocytes of adult C57Bl/6 and BALB/c mice is shown in Fig. 1. In BALB/c, the relative representation of the Vµ J558 family varies between 25 and 35%, and it never exceeds 35%. The Vµ 7183 and Q52 families are each expressed by about 10% of the splenocytes, while the Vµ 36–60 family is always used by 5% or more of the cells. In C57Bl/6, the Vµ J558 family is expressed by 45% or more of the splenic B cells. The Vµ 7183 and Q52 families are each used by about 7%, and the Vµ 36–60 family by <5% of the cells. A similar strain-
V. gene family usage among LPS-activated splenocytes (available repertoire) of adult C57Bl/6 and BALB/c mice. Results are expressed as the relative representation of each V. gene family among 11 families studied and represent the mean of 12 (C57Bl/6) or 26 (BALB/c) experiments. (Bars) SD. Ig-secreting spleen cells assayed ex vivo give similar strain-specific patterns (not shown).

Table 1. Relative Expression of the V. J558 Family by LPS-activated Splenic B Cells (available) from Newborn and Adult and Among Ig-secreting Spleen Cells (actual) from Adult IgH Congenic Mice.

|        | Newborn Available | Adult Available | Actual |
|--------|-------------------|-----------------|--------|
| BALB/c | 23.7 (4.6)        | 29.9 (2)        | 31.4 (2.3) |
| B6Igh | 32.3 (6)          | 47.2* (2.1)     | 32.6 (1.5) |
| C57Bl/6 | 53 (2.6)       | 52.8 (2.5)      | 63.8 (2.8) |
| CB20   | 59.7 (7.1)        | 35.3* (3.5)     | 54.9 (6.3) |

Adult or 1-d-old C57Bl/6, BALB/c, B6Igh, and CB20 spleen cells were analyzed by in situ hybridization with 11 V. gene family probes. The relative representation of the V. J558 family is shown. Adult spleen cells were either tested ex vivo (actual repertoire) or after LPS stimulation in vitro (available repertoire). Results shown represent the mean (SD) of five (CB20) or nine (B6Igh) different experiments and significant differences are given by one factor ANOVA treatment. The relative V. J558 family expression in the available repertoire of adult B6Igh differs significantly from BALB/c, but not from C57Bl/6 while, in the available repertoire of the newborn and in the adult actual repertoire, it is similar to BALB/c, and significantly different from C57Bl/6. Reciprocally, the relative expression of V. J558 family in the available repertoire of adult CB20 differs significantly from C57Bl/6, but not from BALB/c while, in the available repertoire of the newborn and in the adult actual repertoire, it is similar to C57Bl/6, but significantly different to BALB/c.

To investigate the genetic factors controlling V. gene family expression patterns, we analyzed several independent congenic strains of mice. To simplify our presentation, we shall restrict our analysis mainly to the V. J558 family, and refer to a C57Bl/6 or a BALB/c pattern of V. gene family usage, respectively, when >45 or <35% of the splenic B cells express the V. J558 family. In all experiments shown, however, the other ten V. gene families varied in agreement with the changes observed for the V. J558 family, after the strain-specific patterns described above.

Available Repertoire of LPS-activated Splenocytes in Adult Congenic Mice. The B6Igh and CB20 mouse strains are reciprocal congenics with C57Bl/6 and BALB/c strains for the IgH haplotype. As shown in Table 1, LPS-activated splenocytes of adult B6Igh mice show a relative expression of the VH J558 family that differs significantly from BALB/c (p <0.01), but not from C57Bl/6. Reciprocally, LPS-activated splenocytes from CB20 show a relative expression of VH J558 family that is significantly different from C57Bl/6 (p <0.05), but not from BALB/c. The V. gene family expression pattern in LPS-activated splenocytes (available repertoire) from adult IgH congenic mice segregates, therefore, independently of the IgH locus, and seems determined by the genetic background of the congenic strain.

In an attempt to identify the background genes that could play a role in the regulation of VH gene family expression, we have next investigated MHC and IgK L chain loci. We found that the representation of V. gene family among LPS-activated splenocytes of MHC congenic BALB.B10 and BALB.C3H strains, as well as in the IgK L chain congenic C.C58 and C.AKR strains, is identical to that observed in the BALB/c strain (Table 2). Similarly, V. gene family usage by Ig-secreting spleen cells did not differ in these congenic strains of mice. These results show that the strain-specific pattern of V. gene family expression is kept regardless of MHC and IgK L chain haplotypes.

Available Repertoire of LPS-activated Splenocytes in Newborn IgH Congenic Mice. In contrast to the adult, LPS-activated splenocytes in newborn B6Igh and CB20 congenic mice show a pattern of V. gene family expression identical to BALB/c and C57Bl/6, respectively (Table 1). The usage of the V. J558 family among LPS-activated splenocytes of newborn B6Igh differs significantly from that of the newborn C57Bl/6 (p <0.01), while it is similar to that of the newborn BALB/c. Reciprocally, the V. J558 family usage in spleen of newborn CB20 is significantly different from that of the newborn BALB/c (p <0.01).
LPS-activated splenocytes is determined by the IgH locus. The shift in the pattern of \( V_n \) gene family usage in the splenic available repertoire from newborn to adult IgH congenic mice suggests that V region selection operates in the establishment of the adult antibody repertoire (14–17).

**Actual Repertoire of Adult Splenic Plasmocytes in IgH Congenic Mice**. Analysis of spleen cells ex vivo allows assessment of \( V_n \) gene family expression among Ig-secreting splenocytes. Plasma cells in adult B6Igh^a^ and CB20 mouse strains show a pattern of \( V_n \) gene family expression determined by the IgH locus. Thus, the expression of the \( V_n \) J558 family in B6Igh^a^ Ig-secreting splenocytes is significantly different from C57BL/6 (\( p < 0.01 \)), but not from BALB/c plasma cells (Table 1). Reciprocally, expression of the \( V_n \) J558 among splenic plasma cells of CB20 mice is similar to C57BL/6, but differs significantly from BALB/c (\( p < 0.01 \)). These results show that the \( V_n \) gene family expression pattern among adult Ig-secreting splenocytes is determined by the IgH locus.

The \( V_n \) gene family expression patterns in newborn and adult mice of the congenic strains are summarized in Fig. 2. Splenic available repertoires of newborn and adult IgH congenic mice display different patterns of \( V_n \) gene family expression. Available and actual repertoires of adult IgH congenics differ, while newborn available and adult actual repertoires have similar patterns of \( V_n \) gene family expression. Available and Actual Repertoires in Newborn or Adult F1 Spleen Cells. We investigated the relative role of the IgH locus and of the genetic background on the pattern of \( V_n \) gene family expression in F1(C57BL/6 \( \times \) BALB/c) mice. In these mice, both C57BL/6 and BALB/c genetic backgrounds are present, but two subpopulations of B lymphocytes are available, each expressing \( V_n \) genes of either the BALB/c or the C57BL/6 IgH haplotype. We determined the \( V_n \) gene family usage, either ex vivo or after LPS stimulation, in FACS^®-isolated IgM^a^ or IgM^b^ splenic B cells from adult F1 mice. FACS^® profiles of F1-unseparated spleen cells and allelotype-sorted populations are shown in Fig. 3. The relative expression of the \( V_n \) J558 family expression by sorted and unseparated B cells from F1 mice is shown in Table 3. The \( V_n \) J558 usage by \( \mu^a \) cells differs significantly from \( \mu^a \) (\( p < 0.01 \)) and BALB/c (\( p < 0.01 \)), but not from C57BL/6 spleen cells. \( V_n \) J558 usage among \( \mu^b \) cells differs significantly from \( \mu^b \) (\( p < 0.01 \)) and C57BL/6 (\( p < 0.01 \)), but not from BALB/c splenocytes. Thus, each allelotype population of F1 mice follows an independent pattern of \( V_n \) gene family expression, both in resting or Ig-secreting cells. These results show that the B cell repertoires of \( \mu^a \) and \( \mu^b \) adult spleen cells in F1 mice are independent from one another.

**Table 2. Relative Expression of the \( V_n \) J558 Family by LPS-activated Splenic B Cells from Adult MHC and IgK L chain Congenic Mice**

| BALB/c (H-2d) | C57BL/6 (H-2b) | AKR (H-2k) | BALB.B10 (H-2b) | BALB.C3H (H-2k) | C.AKR | C.C58 |
|---------------|----------------|------------|-----------------|----------------|--------|--------|
| 29.9          | 52.8*          | 44.7*      | 33              | 36.5           | 31.2   | 26.8   |
| (0.8)         | (2.5)          | (4.2)      | (4)             | (2)            | (2.7)  | (2)    |

Spleen cells from adult BALB.B10 or BALB.C3H MHC congenic mice, C.AKR or C.C58 IgK L-chain congenic mice, or BALB/c, AKR, and C57BL/6 mice were used, either ex vivo or after in vitro LPS stimulation, to determine the relative expression of 11 \( V_n \) gene families by in situ hybridization. The relative expression of the \( V_n \) J558 family among LPS-activated spleen cells is shown. Results represent the mean (SD) of two (IgK L chain congenic mice) or three (MHC congenic mice) different experiments, and statistically different values are given by one factor ANOVA treatment. The relative representation of the \( V_n \) J558 family by LPS-activated splenocytes of the different congenic mouse strains differs significantly from both C57BL/6 and AKR, but is identical to BALB/c. Similar patterns were observed among Ig-secreting spleen cells (not shown).

\* \( p < 0.01 \)
Figure 3. Purification by cytofluorometry of IgM⁺ and IgM⁻ cells from adult F₁(C57Bl/6 × BALB/c) splenocytes. Adult spleen cells were labeled with anti-IgM⁺ and anti-IgM⁻ antibodies (see Materials and Methods), and sorted by gating each population as shown (a). Cytofluorometric reanalysis of the sorted cell populations gave the profiles shown (b and c). The purity of each sample, given by the percentage of the contaminant allotype, was controlled by limiting dilution analysis. For this experiment, in the IgM⁺ sorted population, the frequencies of LPS-reactive cells of the a and b allotypes were, respectively, 1:9 and 1:383 i.e., 2.6% contaminating IgM⁻ cells. In the IgM⁻ sorted population, the frequencies of LPS⁺ cells of the a and b allotypes were, respectively, 1:875 and 1:10 i.e., 1% contaminating IgM⁺ cells.

Discussion

The pattern of V₅ gene family usage in peripheral B cell repertoires of normal C57Bl/6 and BALB/c mice is strain specific. We have now studied peripheral B cell repertoires of newborn and adult mice from the respective IgH congenic strains, B6Igh¹ and CB20. We show here that the pattern of V₅ gene family usage in LPS-activated splenocytes (available repertoire) segregates with the IgH locus in newborns, but is determined by the genetic background in adult mice. These results partially confirm a previous report by Wu and Paige (12), studying V₅ gene family expression in LPS-induced B cell colonies from spleen cells of adult IgH congenic mouse. In these experiments, V₅ gene family usage
Table 3. Relative Expression of the \( V_n \) 558 Family by LPS-activated B Cells (available) from Newborn and Adult and by Ig-secreting Cells (actual) from Adult F1(C57BL/6 × BALB/c) Unseparated Spleen Cells or Adult IgM and IgM Sorted Spleen Cells.

|                | Newborn Available | Adult Available | Adult Actual |
|----------------|-------------------|-----------------|--------------|
| BALB/c         | 23.7 (4.6)        | 29.9 (0.8)      | 31.4 (2.3)   |
| F1, \( \mu^a \) sorted | 31* (2.2) | 37.5* (0.5)     |              |
| F1             | 48.8 (3.4)        | 35.1 (2.2)      | 43.3 (2)     |
| F1, \( \mu^b \) sorted | 64.7* (3.9) | 66.6* (0)       |              |
| C57BL/6        | 53 (2.6)          | 52.8 (2.5)      | 63.8 (2.8)   |

\( V_n \) gene family expression by LPS-activated and Ig-secreting cells was determined among either unseparated spleen cells from newborn and adult F1(C57BL/6 × BALB/c) mice, or among sorted IgM and IgM spleen cells from adult F1 mice. The relative representation of 11 \( V_n \) gene families was determined by in situ hybridization, and the relative expression of the \( V_n \) J558 family is shown. Values shown represent the mean (SD) of three (sorted cells) or five (unseparated cells) different experiments. The purity of each sorted sample was controlled by FACS analysis and limiting dilution analysis, and never exceeded 4% contamination. The relative \( V_n \) J558 family expression by \( \mu^a \) cells is similar to BALB/c, while significantly different from \( \mu^b \) and C57BL/6, both in LPS-activated and Ig-secreting cells. In \( \mu^b \) cells, the relative \( V_n \) J558 family expression, while similar to C57BL/6, differs significantly from \( \mu^a \) and BALB/c, both in LPS-activated and Ig-secreting spleen cells.

\( *p < 0.01 \)

was also determined by genes of the genetic background, and it was proposed that a locus, outside of the \( V_n \) region, regulates the strain-specific \( V_n \) gene family expression. We have now found that, in newborn IgH congenic mice, the strain-dependent \( V_n \) gene family expression is not regulated by a locus outside of the \( V_n \) region, but rather determined by the IgH locus itself. We conclude, from the shift of \( V_n \) gene family expression pattern in the available repertoire between newborn and adult IgH congenic mice, that the strain-specific \( V_n \) gene usage either is regulated by a locus outside of the IgH locus whose expression is developmentally controlled, or from cellular selection of the available repertoire during the development and establishment of the adult antibody repertoire. Since \( V_n \) gene family usage is not modified in MHC congenic mice, such cellular selection of the adult available repertoire would involve products of the genetic background other than MHC antigens.

The strain-dependent \( V_n \) gene family usage could also reflect a different \( V_n \) gene family genetic complexity in C57BL/6 and BALB/c strains, or in the respective IgH congenic strains. Although unlikely (34), the actual number of functional genes could differ among the IgH locus donor and its respective IgH congenic strain. The finding that, within the same IgH congenic strain, splenic B cells expressing the same IgH haplotype display different pattern of \( V_n \) gene family expression, depending upon the developmental stage of the mice, demonstrates that the \( V_n \) gene family usage does not only rely on genetic complexity.

In adult IgH congenic mice, while the pattern of \( V_n \) gene family expression in the available repertoire is determined by the genetic background of the congenic strain, it segregates with the IgH locus among Ig-secreting cells. The different patterns of \( V_n \) gene family expression observed between the available and actual repertoires of adult IgH congenic mice suggest that \( V_n \) gene family usage among natural splenic plasmocytes of a normal, nonimmunized mouse is not a random representation of the \( V_n \) gene families expressed by resting splenic B cells, and results from the activation and terminal differentiation of a selected population of B cells. Selection of the actual antibody repertoire was previously reported in Ig-transgenic mice (35), where it was found that the fraction of B cells expressing endogenous \( V_n \) gene families, which did not exceed 5% of the resting splenic B cells, represented over 50% of the Ig-secreting splenocytes.

The present results indicate that, in a nonimmunized mouse, available and actual B cell repertoires are generated through different mechanisms of cellular selection, acting, respectively, on the rate of accumulation and turnover of newly-formed B cells in the peripheral compartments of the immune system, and on the activation and terminal differentiation of immunocompetent B cells. It is interesting to note that \( V_n \) gene family usage by both neonatal resting and adult Ig-secreting B cells is determined by the IgH locus. This could suggest that selection for Ig secretion is stably kept from the newborn to the adult mice. We have previously shown, however, that the \( V_n \) gene family usage differs in neonatal resting and adult BALB/c mice, and that these changes are determined by exposure to external antigens, since they are not observed in mice kept under axenic conditions (36). On the other hand, resting nonactivated B lymphocytes have been shown to accumulate during the first 3 wk of age, in the spleen of normal mice (37). Our findings suggest that resting splenocytes could selectively accumulate according to their Ig variable regions through cellular interactions with their environment. The different environmental influences imposed by the C57BL/6 or BALB/c genetic backgrounds would dictate the differential selection of the available repertoires, in the adult IgH congenic mice, leading to the strain-specific \( V_n \) gene family usage. Alternatively, the differences of \( V_n \) gene family expression between available and actual repertoires of adult IgH congenic mice could result from different cellular origins of resting and Ig-secreting B cells (38).

We also investigated the relative role of the IgH locus and the genetic background on the pattern of \( V_n \) gene family usage in adult F1(C57BL/6 × BALB/c) mice. The question
was how, in F1 mice, the presence of both genetic backgrounds would determine the pattern of Vₜ gene family usage in each of the two B cell subpopulations expressing C57Bl/6 or BALB/c Vₜ genes. We found that the pattern of Vₜ gene family expression in each allotype population differs from unseparated B cells and is determined by the IgH locus, both in resting and Ig-secreting spleen cells. This finding suggests that the strain-specific Vₜ gene family expression of resting splenocytes, if regulated by genes located outside of the IgH locus, must be under control of a factor acting in cis. Such intracellular cis regulation of Vₜ gene family expression could result from differential recombinase activity in C57Bl/6 and BALB/c mice (39). Control of Ig gene rearrangements, however, cannot be the only mechanism involved in regulation of Vₜ gene family usage, since, in IgH congenic mice, resting B cells and Ig-secreting B cells, although expressing the same IgH haplotype, show different patterns of Vₜ gene family expression. Alternatively, in adult F1 mice, each allotype population could be preferentially selected by products of background genes from the parental origin of the IgM allotype expressed. The finding that the Vₜ gene family usage by μᵣ and μᵦ cells in the F₁ are independent suggests that V-region interactions of a functional immune network (35) would not operate in the regulation of the Vₜ gene family expression, unless these interactions distinguish between C57Bl/6 and BALB/c Ig variable regions. Kearney and colleagues (personal communication) have recently found that the frequency of V region interactions among neonatal IgM-secreting hybridomas is higher within the same mouse strain.

We conclude that different selective processes act on resting or naturally activated B cells of normal mice. The selection of the available repertoire in IgH congenic mice is responsible for the strain-specific Vₜ gene family usage. This selection occurs during ontogeny and involves background genes of the congenic strain, other than MHC. It is not clear, however, how this selection could differentiate between both alleles in F₁ mice.

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