Evaluation of in vitro antioxidant, anticancer activities and molecular docking studies of *Capparis zeylanica* Linn. leaves

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Abstract

Background: *Capparis zeylanica* Linn. leaf extract was subjected to phytochemical screening for the determination of antioxidant and anticancer activity on (MCF-7) human breast cancer cells. The phytoconstituents previously determined were subjected to molecular docking studies against human epidermal growth factor receptor 2 (HER2) protein as a target receptor to support antioxidant and anticancer activities.

Results: Powdered plant leaves were extracted by maceration method using ethyl acetate, chloroform, methanol, ethanol and distilled water. Preliminary phytochemical evaluation and total phenolic and flavonoid content of the extract were evaluated using biochemical tests. Total antioxidant capacity of the extract was evaluated using different assays. Anticancer potential of methanolic and ethanolic extracts was studied on human breast cancer cells. Molecular docking studies were performed to evaluate the binding interactions of phytoconstituents on HER2 protein using AutoDock Vina. Phytochemical evaluation confirmed the presence of saponins, flavonoids, tannins, phenols, carbohydrates and proteins. Ethanolic extract showed a maximum total phenolic and flavonoid content in support with antioxidant and anticancer activities. The ethanolic leaf extract showed 66.63% cell growth inhibition against MCF-7 cells. Molecular docking studies revealed the highest binding affinity (−8.4 Kcal/mol) of α-amyrin followed by quercetin and β-carotene. Glucocapparin, syringic acid, vanillic acid and p-coumaric acid showed almost a similar binding affinity to the amino acid residues of HER2 protein as compared to 5-FU.

Conclusion: *C. zeylanica* leaf extract showed the presence of phenolic and flavonoid constituents responsible for antioxidant and in vitro anticancer activities. Molecular docking studies showed the binding affinity of phytoconstituents on targeted HER2 protein.

Keywords: In vitro anticancer, *Capparis zeylanica*, HER2 protein, Flavonoids, Phenolics, MCF-7 cells

Background

Cancer is recognized as a leading cause of death. Many cancer treatments such as chemotherapy, surgery and radiotherapy are available to treat cancer, although severe side effects remain a concern. Cancer is usually associated with accumulation of mass of cells resulted from poor signal transduction across pathways due to overexpression of epidermal growth factor receptors. Breast cancer generally occurs in women and rarely in men. As per Globocan 2018, breast cancer stands second of all cancers for a cause of death. The international agency for research on cancer released the estimates in 2018 on the global burden of cancer. The global burden was raised to 18.1 million new cases and 9.6 million deaths in 2018 [1]. In India, more than 11.5 million cases were detected with all types of cancers. Out of which, 14% deaths were associated with breast cancer. Since then, newer techniques in detection and treatments were...
developed, such as chemotherapy, radiation therapy and surgery. But these were contributed with severe side effects. Chemotherapy is the treatment of choice in most of the cancer cases rather than radiation and surgical operations [2, 3]. Medicinal plants have always remained a choice of treatment in many diseased conditions with the availability of isolated novel phytoconstituents with minimum side effects. According to the World Health Organization reports, 252 drugs were approved for cancer treatment, out of them 11% drugs were of plant origin [4].

*C. zeylanica* Linn., family Capparidaceae, is widely distributed throughout India, China, Nepal, Bangladesh, Malaysia and Pakistan. The leaf extracts are suggested for the treatment of dysentery, diabetes and rheumatism [5]. Pharmacologically, ethanol and methanol extracts of roots of *C. zeylanica* showed promising in vitro antioxidant activities [6]. The leaves of *C. zeylanica* exhibit immunostimulant [7] antidepressant [8] and antimicrobial [9] activities.

The free radicals are reactive oxygen species responsible for the cause of cancer [10]. Antioxidants are the chemical constituents which either delay or prevent the oxidation process of free radicals in the body. The present study was aimed at evaluation of anticancer potential of *C. zeylanica* Linn. leaf extract. The preliminary screening of phytoconstituents responsible for antioxidant activity was done based on the total phenolic and flavonoid content of the extract. The plant was not researched out for the anticancer activity in human breast cancer cells. Thus, an attempt was made to evaluate antioxidant activity by 2,2-diphenyl-1-picrylhydrazyl (DPPH) assay and anticancer activity of *C. zeylanica* Linn. leaf extract against MCF-7 cells. The molecular docking studies of phytoconstituents of *C. zeylanica* Linn. on human epidermal growth factor receptor (HER2) protein revealed the binding affinity with the amino acids within proximity of active sites of HER2 protein [11].

**Methods**

**Materials**

Aluminium chloride, ethyl acetate, chloroform, ethanol and methanol were purchased from Lobachemie. Butylatedhydroxytoluene (BHT), Trolox, gallic acid and quercetin were purchased from Sigma-Aldrich. 2,2-Diphenyl-1-picrylhydrazyl (DPPH), molybdate reagent, dimethyl sulfoxide (DMSO), Dulbecco’s modified Eagle’s medium (DMEM), actinomycotic 100X solution and ascorbic acid were purchased from Thermo Fisher Scientific.

**Collection and authentication of the plant part**

The plant *C. zeylanica* was collected from the rural area in Kolhapur district, Maharashtra, India. The plant was identified and authenticated (MB/RW/02/013) by Dr. M. Y. Bachulkar, Ex-Principal and Taxonomist, Department of Botany, Shri Vijaysinha Yadav Arts and Science College, Kolhapur, Maharashtra. A fresh leaves were collected, washed and dried in shade for 2–3 weeks. Dried leaves were blended to fine powder and stored in airtight container and used for further studies.

**Preparation of crude extract**

A 50 g of powdered crude drug was subjected to extraction by maceration method using ethyl acetate, chloroform, ethanol, methanol and aqueous solvents for 6–8 days to extract soluble compounds [12, 13]. Filtered the liquid using filter paper and concentrated the extracts. Concentrated extracts were collected and stored at 4 °C until further use.

**Phytochemical analysis**

The preliminary phytochemical evaluation of the extracts was done using different biochemical tests as per reported method [14]. Further, ethanolic and methanolic extracts of *C. zeylanica* were selected for further studies based on the qualitative analysis from five extracts.

**Estimation of total phenolic content**

The total phenolic content of *C. zeylanica* was estimated by the Folin-Ciocalteu reagent method [15] with slight modification. Briefly, 200 μl of methanolic and ethanolic extracts were added in equal volume of Folin-Ciocalteu reagent (tenfold diluted) and incubated for 10 min. Then, 1.25 mL of aqueous sodium carbonate was added to neutralize the mixture, incubated for 45 min at 37 °C with intermittent shaking to generate colour. Distilled water was used as a blank. The extracts were analysed for total phenolic content by measuring the blue colour at 765 nm using UV-Vis spectrophotometer (UV-2700i/2600i, Shimadzu, Japan) in triplicate. Gallic acid was used as a reference standard for calibrating the method of analysis and to construct the calibration curve. The total phenolic content of the extract was calculated from the linear equation of calibration curve of gallic acid, in terms of milligram per gram of gallic acid equivalent (GAE) of dry extract.

**Estimation of total flavonoid content**

The total flavonoid content of *C. zeylanica* was estimated as per the procedure reported [16]. Briefly, 0.5 mL of methanolic and ethanolic extracts were mixed with equal volume of 2% AlCl3 and incubated at room temperature for about 1 h. Evaluation of extract samples was carried out at a final concentration of 1 mg/mL. Quercetin was used to construct the calibration curve, and the total flavonoid content was calculated as milligram per gram quercetin equivalent (QE). The results
were carried out in triplicate by measuring absorbance at 420 nm. The procedure was repeated as above for both the extracts.

**Determination of total antioxidant capacity (TAC)**
The total antioxidant capacity of *C. zeylanica* leaf extract was determined by the phosphomolybdenum assay using the method described by an earlier report [17]. Briefly, 1.0 mL of different concentrations of methanolic and ethanolic extracts (100 to 500 μg/mL) was mixed with 3.0 mL phosphomolybdenum reagent containing 28 mM sodium phosphate and 4 mM ammonium molybdate in 0.6 M sulphuric acid in capped test tubes. Distilled water was added to make the final volume to 5 mL. The test tubes containing mixture was incubated for 90 min in a water bath set at 95 °C in dark. After incubation, the mixture was cooled at room temperature and absorbance of resulting solution was measured in triplicate at 695 nm against ethanol and methanol as blank using UV-visible spectrophotometer. The TAC results were expressed as Trolox equivalents (mg TE/g of dry extract). Butylatedhydroxytoluene was used as reference controls.

**DPPH scavenging assay**
The free radical scavenging activity of *C. zeylanica* leaf extract was estimated by the DPPH as per earlier reports [18], with slight modifications. One milliliter of different concentrations of methanolic and ethanolic extracts ranging from 100 to 500 μg/mL was added in each test tube containing 2 mL 1.0 mmol/L DPPH solution. Ascorbic acid solution was prepared separately with the same procedure as a standard. All test tubes were incubated at room temperature for 30 min in dark. The absorbance was measured at 517 nm. The higher is the free radical scavenging activity. The percentage free radical scavenging activity was calculated using the following equation:

\[
\text{DPPH scavenging activity (\%)} = \frac{A_c - A_t}{A_c} \times 100
\]

where \(A_c\) is the absorbance of the control (1 mL of ethanol with 1 mL of DPPH solution) and \(A_t\) is the absorbance of the test sample. The results were analysed in triplicate.

**Cytotoxicity study**
Human breast cancer cell line, MCF-7, and human breast cell line, MCF10A, were procured from the National Centre for Cell Science, Pune, India. The cells were subcultured in Dulbecco’s modified Eagle medium (DMEM) with low glucose having phenol red supplemented with 10% fetal bovine serum and 1% penicillin/streptomycin solution to minimize the microbial contamination. The cells were seeded at a density of 5 × 10^3 cells/well in a 96-well flat bottom microplate and maintained at 37 °C in 5% CO₂ incubator with 95% humidity overnight prior to the experiment. The cells were treated with different concentrations (500, 250, 125, 62.5, 32.75 μg/mL) of methanolic and ethanolic extracts by serial dilution method. Initially, cells were diluted with serum-free medium to achieve twice the desired maximum test concentration. The cells were incubated for another 48 h, and cytotoxicity was evaluated using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay [19, 20]. MTT assay was initiated with 5 × 10^3 cells/well, 20 μL of the MTT staining solution (5 mg/mL) in phosphate buffer solution was added to each well and plate was incubated at 37 °C for 4 h. After incubation, the medium was removed and subsequently added with 100 μL DMSO in each well to dissolve the formed formazan crystals. The absorbance was measured at 570 nm using micro plate reader (BioRad, India) in triplicate [21, 22]. The percentage cell inhibition was calculated using equation as.

\[
\text{Surviving cells (\%)} = \frac{\text{Mean OD of the test extract}}{\text{Mean OD of negative control}} \times 100
\]

\[
\text{Inhibiting cells (\%)} = 100 - \text{Surviving cells}
\]

The inhibitory cells (%) = 100 - Surviving cells

The inhibitory concentration (IC₅₀) value for the extracts was determined based on the concentration of extract inhibiting the cells by 50% of its initial cell population.

**Molecular docking studies**
The molecular docking studies were performed to determine the chemical interactions of the phytoconstituents of plant *C. zeylanica* Linn. with targeted protein responsible for breast cancer. Molecular docking was initiated by drawing 3D structures of phytoconstituents, considered as ligands and prepared, energy minimized. The target protein HER2 was downloaded from RCSB (PDB id: 1N8Z). The protein was prepared and refined using MMCF-2 forcefield. All the heteroatoms were removed from crystal structure of protein to make all the complex receptors free of ligand before docking using PyMOL-2.3.4 software. AutoDock Tools (version-4.2.6) software was used for the preparation and optimization of protein and ligand molecules. Water molecules were removed, and polar hydrogens and Kollman charges were added in the protein molecule. Since ligands were small molecules, Gasteiger charge was added. The binding site of the protein was selected by selecting a grid box with
dimensions size \((X = 8.820, Y = 91.432, Z = 127.425)\) and docked using AutoDock vina (version 1.1.2) \[23\]. All the AutoDock vina docking runs were performed in Intel Centrino Core2Duo CPU @ 2.20 GHz of IBM system origin, with 4 GB DDR2 RAM. AutoDock vina was compiled and run under Microsoft Windows 10 operating system. The results were analysed on the basis of binding affinity of ligand with the protein and refined mean standard deviation. Images of protein-ligand binding confirmations were visualized and processed using Discovery studio visualizer 2020 software. 2D structures of constituents were drawn using Chemdraw Professional 16.0.

**Statistical analysis**
The data were expressed as mean ± standard deviation of results obtained from three independent experiments. \(P \leq 0.05\) was considered significant for all the represented results. All the graphs were plotted using OriginPro 9.1 software.

**Results**

**Total yield of crude extract**
The percentage yield of crude extracts obtained from \(C.\) \(zeylanica\) leaves using different solvents like ethyl acetate, chloroform, ethanol, methanol and water was 15.12%, 8.62%, 17.33%, 21.15% and 14.75%, respectively, by maceration method.

**Phytochemical analysis**
Preliminary phytochemical evaluation of \(C.\) \(zeylanica\) leaf extract (ethyl acetate, chloroform, ethanol, methanol and aqueous) showed the presence of a variety of phytochemical constituents. Proteins were found to be present in all five extracts while saponins, flavonoids, tannins, phenolic compounds and carbohydrates were found in ethyl acetate, ethanol, methanol and aqueous extracts. Alkaloids, fats and oils were found in chloroform extract (Table 1). Based on total percentage yield, polarity, solvent compatibility and phytochemical evaluations, ethanolic and methanolic extracts were selected for the further studies.

**Estimation of total phenolic content**
The total phenolic content of ethanolic and methanolic extracts of \(C.\) \(zeylanica\) leaves was estimated by Folin-Ciocalteu reagent method and expressed as milligram per gram gallic acid equivalent (GAE). The total phenol content obtained in the ethanolic extract was 59.13 ± 1.45 mg/g GAE and that in methanolic extract was 48.43 ± 0.15 mg/g GAE. All the values were calculated using linear regression equation of gallic acid (Fig. 1).

| Chemical constituents | Test/reagent | EAE | CHE | EE | ME | AQE |
|-----------------------|--------------|-----|-----|----|----|-----|
| Saponins              | Foam test    | ++  | --  | ++ | ++ | ++  |
|                       | Haemolytic test | --  | ++  | ++ | ++ | ++  |
| Alkaloids             | Dragendorff’s test | --  | ++  | -- | -- | --  |
|                       | Wagner’s test | --  | ++  | -- | -- | --  |
|                       | Hager’s test  | --  | ++  | -- | -- | --  |
|                       | Mayer’s test  | --  | ++  | -- | -- | --  |
| Flavonoids            | Shinoda test | ++  | --  | ++ | ++ | ++  |
|                       | Lead acetate test | ++  | --  | -- | ++ | ++  |
|                       | \(H_2\)SO_4 test | --  | --  | ++ | ++ | ++  |
| Tannins and phenols   | Bromine water test | ++  | --  | ++ | ++ | --  |
|                       | Lead acetate test | ++  | --  | ++ | ++ | ++  |
|                       | \(HNO_3\) test | ++  | --  | ++ | ++ | ++  |
|                       | Dilute iodine solution test | --  | --  | ++ | -- | --  |
| Carbohydrates         | Molisch test | ++  | --  | ++ | ++ | ++  |
|                       | Fehling’s test | ++  | --  | ++ | ++ | ++  |
|                       | Barfoed’s test | ++  | --  | ++ | ++ | --  |
| Steroids              | Salkowski test | --  | ++  | -- | -- | --  |
| Proteins              | Million’s test | ++  | ++  | ++ | ++ | ++  |
|                       | Biuret test   | ++  | ++  | ++ | ++ | ++  |
| Fats and oils         | Filter paper test | --  | ++  | -- | -- | --  |

++ present, -- absent, EAE Ethyl acetate extract, CHE Chloroform extract, EE Ethanol extract, ME Methanol extract, AQE Aqueous extract
Fig. 1 Calibration curve for determination of total phenolic content using gallic acid in methanolic and ethanolic extracts of *C. zeylanica* leaves.

Fig. 2 Calibration curve for determination of total flavonoid content using quercetin, in methanolic and ethanolic extracts of *C. zeylanica* leaves.
Estimation of total flavonoid content
The total flavonoid content present in the ethanolic extract of *C. zeylanica* leaves was 36.87 ± 0.45 mg/g QE while methanolic leaf extract was found to exhibit 32.25 ± 0.45 mg/g QE calculated using linear regression equation of quercetin (Fig. 2).

Determination of total antioxidant capacity
Total antioxidant capacity of the ethanolic and methanolic extracts of *C. zeylanica* leaves were determined by the phosphomolybdenum assay using BHT as a reference standard. The results were expressed as concentration of extracts against TAC mg TE/g of dry extract. The reduction of phosphate-molybdenum (VI) to phosphate-molybdenum (V) on reacting with phytochemicals from extracts to produce green molybdenum complex was correlated to the amount of TAC mg TE/g of dry extract. From (Fig. 2), the methanolic extract (97.3 ± 1.4 mg TE/g) showed comparatively low antioxidant capacity than the ethanolic extract (123.6 ± 3.4 mg TE/g) and reference standard BHT (162.4 ± 4.2 mg TE/g) at 500 μg/mL. The ethanolic extract showed significant TAC in comparison to BHT which depicted greater antioxidant potential among other extracts due to excess phenolic compounds. The extracts were found to show increase in mg TE/g of dry extract with an increase in the concentrations of methanolic and ethanolic extracts (Fig. 3).

DPPH scavenging assay
DPPH is a free radical at room temperature and becomes a stable diamagnetic molecule on reaction with electron or hydrogen radical. The lower the absorbance of reaction mixture, the higher is the free radical scavenging activity. In the present study, methanolic and ethanolic extracts of *C. zeylanica* leaves were subjected to DPPH radical scavenging assay and the antioxidant activity of extracts were compared with standard ascorbic acid. The ethanolic extract was found to have comparatively high free radical scavenging activity than methanolic extract (Fig. 4).

Cytotoxicity study
Cytotoxicity of extracts of *C. zeylanica* leaves on MCF-7 cells was determined by MTT assay. MTT assay measures the reduction of yellow-coloured MTT by mitochondrial succinate dehydrogenase. In the present work, MTT was reduced to insoluble and dark, purple-coloured formazan with the phytoconstituents present in *C. zeylanica* leaves extracts. As the reduction of MTT can occur only in metabolically active cells, the level of activity was considered as an indicator of the cell viability. The result of MTT assay revealed that the ethanolic and methanolic extracts of *C. zeylanica* leaves decreased the percentage of cell viability. The extracts showed cytotoxic effect in the form of cell growth inhibition against MCF-7 cells. In case of ethanolic extract,
Fig. 4 Inhibition of DPPH by methanolic and ethanolic extracts of *C. zeylanica* leaves. The results were expressed as mean ± standard deviation.

Fig. 5 % Cell growth inhibition of MCF-7 cell lines by methanolic and ethanolic extracts of *C. zeylanica* leaves. The results were expressed as mean ± standard deviation.
# Molecular docking studies of phytoconstituents found in *C. zeylanica* Linn. leaves

| Sr. no. | Compound                      | Binding affinity (Kcal/mol) | Root mean standard deviation (RMSD) l. b. | Root mean standard deviation (RMSD) u. b. | Amino acid residues                                                                 | Hydrogen bonds |
|---------|-------------------------------|-----------------------------|------------------------------------------|------------------------------------------|-----------------------------------------------------------------------------------|---------------|
| 1       | α-Amyrin                      | − 8.4                       | 0.000                                    | 0.000                                    | Ile-413(c)                                                                         |               |
| 2       | β-Carotene                    | − 6.9                       | 0.000                                    | 0.000                                    | Ile-413(c), Arg-332(c)                                                            |               |
| 3       | β-Sitosterol                  | − 6.6                       | 0.000                                    | 0.000                                    | Val-3(c), Ser-239(c), Val-274(c), Pro-278(c), Phe-269(c), Thr-281(c), His-468(c) | 1             |
| 4       | E-Octadec-7-en-5-ynoic acid   | − 4.0                       | 0.000                                    | 0.000                                    | Phe-555(c), Cys-562(c), Lys-569(c)                                                 | 2             |
| 5       | Ferulic acid                  | − 5.3                       | 0.000                                    | 0.000                                    | Ser-435(c), Arg-437(c), Arg-465(c), Val-507(c)                                     | 1             |
| 6       | Glucocapparin                 | − 5.7                       | 2.378                                    | 3.550                                    | Arg-12(c), Leu-328(c), Arg-329(c), Thr-387(c), His-415(c)                           | 5             |
| 7       | Linoleic acid                 | − 3.8                       | 0.000                                    | 0.000                                    | Ala-61(b), Arg-62(b)                                                              | 1             |
| 8       | Malvalic acid                 | − 3.3                       | 0.000                                    | 0.000                                    | Cys-316(c), Arg-318(c), Val-319(c), Tyr-321(c)                                    |               |
| 9       | p-Coumaric acid               | − 5.3                       | 2.748                                    | 6.719                                    | Thr-5(c), Asn-380(c)                                                              | 2             |
| 10      | p-Hydroxybezoic acid          | − 4.9                       | 0.000                                    | 0.000                                    | Pro-278(c), Tyr-279(c), Asn-380(c), Tyr-281(c)                                    | 2             |
| 11      | Quercetin                     | − 7.7                       | 0.000                                    | 0.000                                    | Thr-5(c), Pro-278(c), Tyr-279(c)                                                  | 2             |
| 12      | Ricinolic acid                | − 3.8                       | 0.000                                    | 0.000                                    | Leu-146(c), Lys-148(c), Thr-160(c), Ile-162(c)                                    | 1             |
| 13      | Sterculic acid                | − 3.2                       | 0.000                                    | 0.000                                    | Pro-529(c), Arg-530(c), Pro-547(c), Ala-566(c), His-567(c)                        | 1             |
| 14      | Syringic acid                 | − 5.8                       | 0.000                                    | 0.000                                    | Val-3(c), Thr-5(c), Phe-269(c), Pro-278(c), Tyr-281(c)                             | 2             |
| 15      | Thioglucoside                 | − 5.7                       | 0.000                                    | 0.000                                    | Val-3(c), Val-274(c), Pro-278(c), Asn-280(c), Ser-441(c), Asn-466(c)               | 2             |
increase in the cell growth inhibition was observed with an increase in concentration and showed significant cell growth inhibition, 66.63% at 500 μg/mL, while methanolic extract showed a slightly less activity in comparison to ethanolic extract (Fig. 5). However, both the extracts showed non-cytotoxic effect against normal cells (MCF-10A). The IC₅₀ value of methanolic and ethanolic extracts was found to be 233.4 μg/mL and 199.0 μg/mL, respectively, which indicated a promising anticancer activity against MCF-7 cells.

Molecular docking studies
Molecular docking studies were performed to determine the binding interactions of phytochemical constituents of C. zeylanica extracts with HER2 using software AutoDock vina (version-1.1.2). HER2 receptor is concerned with positive breast cancer. All the phytochemical constituents were docked with HER2 along with 5-fluorouracil (Table 3) to determine comparative binding interactions [24]. The results were presented in the various docking parameters (Table 2). From the results, binding affinity was found to be significant for α-amyrin, β-carotene and quercetin (−8.4 to −6.9 Kcal/mol), Glucocapparin, quercetin, syringic acid, vanillic acid and p-Coumaric acid (Table 3) showed maximum hydrogen bonds (≥2) while 5-FU showed 4 H bonds. The stable confirmations of all the ligands with HER2 protein with corresponding bond lengths were represented in (Fig. 6, Table 2).

Discussion
Cancer is recognized globally as a life-threatening disease, and breast cancer is the most common cause of death in women [1]. In the present study, methanolic and ethanolic extracts of C. zeylanica leaves were found to have shown significant anticancer activity against human breast cancer cells. Preliminary phytochemical evaluation of ethyl acetate, ethanol, methanol and aqueous extracts indicated the presence of saponins, flavonoids, tannins and phenolic compounds and carbohydrates and proteins. Methanolic extract exhibited the presence of alkaloids, steroids, fats and oils (Table

| Sr. no. | Compound     | Binding affinity (Kcal/mol) | Root mean standard deviation (RMSD) l. b. | Amino acid residues     | Hydrogen bonds |
|--------|--------------|-----------------------------|-------------------------------------------|-------------------------|----------------|
| 16     | Vanillic acid| − 5.0                       | 0.000                                     |                         | Arg-434(c), Ser-435(c), Cys-509(c) | 3              |
| 17     | 5-FU         | − 5.5                       | 0.000                                     |                         | Arg-434(c), Thr-504(c), Cys-506(c), Cys-509(c) | 4              |

| Sr. no. | Name of the constituent      |
|---------|------------------------------|
| 1       | α-Amyrin                     |
| 2       | β-Carotene                   |
| 3       | Quercetin                    |
| 4       | Glucocapparin                |
| 5       | Syringic acid                |
| 6       | Vanillic acid                |
| 7       | P-Coumaric acid              |
| 8       | 5-FU (reference)             |
Fig. 6  

(a) Crystal structure of extracellular domain of human HER2 (PDB id.: 1N8Z); Molecular interactions of the 2D and 3D confirmations of selected ligands showing significant binding affinity and hydrogen bond interactions (≥ 2).  

(b) Glucocapparin, (c) p-Coumaric acid, (d) Quercetin, (e) Syringic acid, (f) Vanillic acid, (g) 5-FU complexed with Herceptin Fab
The phytochemical constituents in leaf extract of *C. zeylanica* were studied to exhibit various medicinal activities such as for the treatment of dysentery, curing cough and cold; as a rubefacient; as counterirritant and as antidiarrheal [5]. So far, phenolic- and flavonoid-containing compounds have shown antimicrobial activity in various cancer cells [25, 26]. Estimation of total phenolic content and total flavonoid content provides measurement of total antioxidant capacity and antioxidant activity which in turn is correlated with anticancer potential of the plant. The total phenolic content in Ethanolic extract and methanolic extract of *C. zeylanica* leaves were 59.13 ± 1.45 mg/g GAE and 48.43 ± 0.15 mg/g GAE, respectively, which indicate the plant-rich source of phenolic compounds. Further, the total flavonoid content in ethanolic extract and methanolic extract of *C. zeylanica* leaves were 36.87 ± 0.45 mg/g QE and 32.25 ± 0.45 mg/g QE, respectively. The presence of phenolic and flavonoid compounds was responsible for antioxidant activity of the plant [25, 26].

Reactive free radicals in physiology of living cells play an important role in triggering diseased conditions under stressed conditions. As the free radicals are considered toxic in biological system, the effect of free radical scavenging activities of plant extracts are necessarily determined. In the present study, the plant extracts were evaluated for total antioxidant capacity and antioxidant activity using phosphomolybdenum assay and DPPH assay. In the present study, DPPH assay served a significant indicator for potential antioxidant activity based on % inhibition of DPPH (Fig. 4). The graph plotted between concentration and percentage inhibition of DPPH showed increase in antioxidant potential of extracts with an increasing concentration of extracts under study. Ethanolic extract showed comparatively high percentage inhibition than methanolic extract.

MTT assay was performed to screen the in vitro cytotoxicity of *C. zeylanica* leaves’ extract against MCF-7 cells [19, 20]. MTT assay measures the detection of reduced, insoluble, dark purple-coloured formazan solution. For the assay, different concentrations of extract showed decrease in cell viability with the increase in the concentration of extracts. The percentage cell growth inhibition was markedly increased from 5.84% at 32.75 μg/mL to 66.63% at 500 μg/mL ofethanolic extract which was comparatively higher than methanolic extract. The IC50 value of ethanolic extract of *C. zeylanica* leaves against MCF-7 cells was 199.0 μg/mL. However, no any cytotoxicity was observed for extracts of plants against normal breast cells (MCF10A).

Molecular docking studies revealed docking interactions of phytochemical constituents of *C. zeylanica* extracts with HER2 protein. From the results, it was found that α-amyrin showed the highest binding affinity (− 8.4 Kcal/mol) followed by quercetin and β-carotene in comparison with 5-FU. However, other constituents like glucocapparin, p-Coumaric acid, syringic acid and vanillic acid showed (≥ 2) hydrogen bonds to the amino acid residues of HER2 protein. On the basis of above results, it has been found that the ethanolic extract of *C. zeylanica* leaves has significant antioxidant and cytotoxic potential against cancer cells which was in support with the docking analysis.

**Conclusion**

The ethanolic and methanolic extracts of *C. zeylanica* leaves showed the presence of phenolic and flavonoid contents with significant antioxidant and cytotoxic effects. The molecular docking studies on HER2 protein revealed the binding interactions of phenolic and flavonoid compounds of the extracts with target protein and supported the results of antioxidant and anticancer activities. In addition, further research is required to explore the scientific evidences to support the anticancer activity of extracts of *C. zeylanica* leaves by performing in vivo studies.

**Abbreviations**

*C. zeylanica*: Capparis zeylanica; HER2: Human epidermal growth factor receptor; BHT: Butylatedhydroxytoluene; DPPH: 2,2-Diphenyl-1-picrylhydrazyl; DMSO: Dimethyl sulfoxide; DMEM: Dulbecco’s modified eagle’s medium; AlCL3: Aluminium chloride; MTT: 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide; IC50: Inhibitory concentration; RCSB: Research collaboratory for structural bioinformatics; PDB: Protein data bank

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**Plant authentication**

The plant was identified and authenticated (MB/RW/02/013) by Dr. M. Y. Bachulkar, Ex-Principal and Taxonomist, Department of Botany, Shri Vijaysinha Vidyapeeth College of Pharmacy, Kolhapur, Maharashtra, India, for providing the experimental work done, interpretation of the data, and critical revision of the manuscript for intellectual content. RJJ contributed in the design of the work, analysis and interpretation of the data and critical revision of the manuscript for intellectual content. RPD took part in the design of the work, drafting of the work, and critical revision of the manuscript for intellectual content. The authors have read and approved the final manuscript.

**Authors’ contributions**

RAW did the conception of the work, design of the work, experimental work done, interpretation of the data and drafting of the work. RJJ contributed in the design of the work, analysis and interpretation of the data and critical revision of the manuscript for intellectual content. RPD took part in the design of the work, drafting of the work, analysis and interpretation of the data and critical revision of the manuscript for intellectual content. NSL helped in the experimental work done, interpretation of the data, drafting of the work and visualization of images. The authors have read and approved the final manuscript.

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**Availability of data and materials**

The datasets used and/or analysed during the current study will be available from the corresponding author on reasonable request.
Declarations

Ethics approval and consent of participate
Not applicable

Consent for publication
Not applicable

Competing interests
The authors declare that they have no competing interest.

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