IgM-Positive Tubulointerstitial Nephritis Associated With Asymptomatic Primary Biliary Cirrhosis

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INTRODUCTION

Tubulointerstitial nephritis (TIN) is an inflammatory disorder in renal tubules and interstitium without the involvement of glomerular lesions.¹ TIN is often induced by drugs and infections, but idiopathic or systemic disease–related forms have also been observed.²

Primary biliary cirrhosis (PBC) is an immune-mediated chronic and progressive cholestatic liver disease.³ Laboratory data show increased serum values of alkaline phosphatase, γ-glutamyl transpeptidase, and IgM. The hallmark of the disease is the circulating antimitochondrial antibody (AMA). PBC is often associated with extrahepatic autoimmune disorders, such as Sjögren syndrome, thyroid disorders, systemic sclerosis, and rheumatoid arthritis.⁴

Although distal renal tubular acidosis (RTA) has been observed in 33% to 60% of PBC patients,⁵,⁶ TIN and subsequent Fanconi syndrome are relatively rare.⁷ We report herein a case of TIN associated with asymptomatic PBC, which was diagnosed after 4 years of treatment for TIN and Fanconi syndrome.

CASE REPORT

A 57-year-old woman who had not had any abnormality previously was found to have proteinuria on a health checkup 7 years earlier; however, she did not visit a doctor. She developed general malaise and saw her general practitioner 4 years ago. Because urinary examinations revealed proteinuria and glycosuria, she was referred to our hospital for further evaluation. Her height was 160 cm, and her body weight was 59 kg. Her blood pressure was 114/83 mm Hg, and her pulse was 72/min. Physical examination revealed no abnormality except for mild bilateral lower-leg edema. Goiter was not palpable. Laboratory data are listed in Table 1. Urinary examinations showed no cylinders, and pan-aminooaciduria was detected. Serum concentrations of iron, copper, magnesium, cadmium, aluminum, chromium, and cobalt were within normal ranges. The Fishberg concentration test showed decreased renal concentration capacity, and a bicarbonate ion tolerance test showed decreased tubular transport maximum for HCO³⁻. The percent tubular reabsorption of phosphate (%TRP) was low at 63.8% (reference value 85%—95%). There were no signs of osteomalacia on bone scintigraphy. There were no abnormalities of bone marrow puncture. She was clinically diagnosed with Fanconi syndrome and distal renal tubular acidosis.

The first renal biopsy showed 8 instances of global sclerosis among 15 glomeruli. There were no cases of mesangial proliferation, extracapillary proliferative lesions, or adhesion. Severe lymphocyte infiltration in the tubulointerstitial area was noted, accompanied by tubular atrophy (Figure 1a, upper-left panel). She was diagnosed with idiopathic TIN and treated with 50 mg of prednisolone daily. Her serum Cr level temporarily improved from 1.7 to 1.15 mg/dl, but it increased to 1.46 mg/dl during tapering of prednisolone. Urinary β₂-mg increased from 16,000 to 47,456 μg/l, and serum IgM increased from 133 to 959 mg/dl. She was admitted to our hospital again for evaluation of her renal dysfunction. Laboratory data at the second admission showed hypophosphatemia and a high titer of antimitochondrial M2 antibody with normal liver enzymes (Table 1). The second renal biopsy showed global sclerosis in 26 of 51 glomeruli. Severe
lymphocyte infiltration in the tubulointerstitial area was not improved (Figure 1a, lower-left panel). IgM-positive cells were more predominantly observed than IgG-positive cells (Figure 1a). A liver biopsy revealed epithelioid granuloma formation and inflammatory cell infiltration surrounding the bile ducts, which was compatible with PBC (Figure 1b). IgM-positive cells were abundant, whereas IgG-positive cells were scarce in the liver (Figure 1b). To further define the types of cells, immunohistochemical staining with cluster of differentiation (CD)3, a marker for T lymphocytes, CD20, a marker for B lymphocytes, and CD138, a marker for plasma cells, was performed. CD3-positive cells were mixed with CD20- or CD138-positive cells in the tubulointerstitial area of the first renal biopsy, and CD138-positive cells were more abundant than CD20-positive cells (Figure 2, upper panels). On the second renal biopsy, CD3-positive cells were more dominantly observed than CD20- or CD138-positive cells in the tubulointerstitial area (Figure 2, middle panels). On the liver biopsy, CD3-positive cells were mixed with CD20- or CD138-positive cells in the inflammatory cell infiltrates (Figure 2, lower panels).

Because the serum Cr, IgM, and urinary β2-mg levels increased during prednisolone tapering, i.v. administration of 500 mg of methylprednisolone per day for 3 consecutive days, followed by 10 mg of prednisolone daily, was initiated. The patient’s serum Cr, IgM, and urinary β2-mg levels improved, and, at the time of writing, her renal function has been stable without recurrence for 2 years.

**DISCUSSION**

We report herein a patient treated for TIN and Fanconi syndrome who was diagnosed with asymptomatic PBC 4 years later. The antinuclear antibody titer was low, and the serum concentrations of various metal ions were normal at the first admission. As she was not taking any medicines and had no infections, she was diagnosed with idiopathic TIN. Hematological disorders were suspected based on her high serum IgM level, but there were no obvious findings from a bone marrow puncture. Because the serum IgM level improved after the administration of prednisolone, no further investigation was performed. During prednisolone tapering, the patient’s serum Cr level as well as her serum IgM or urinary β2-mg levels increased again. Antimitochondrial M2 antibody was positive at the second admission, and a liver biopsy confirmed the diagnosis of PBC. Severe lymphocyte infiltration in the tubulointerstitial area was not improved at the second renal biopsy.
We searched for articles written in English in PubMed with the terms “renal tubular acidosis primary biliary cirrhosis” or “tubulointerstitial nephritis primary biliary cirrhosis” or “Fanconi syndrome primary biliary cirrhosis.” Among the 13 articles found that described TIN with PBC, we excluded 4 because a renal biopsy was not performed in 2 articles and the other 2 articles described cases that were complicated with Sjögren syndrome. We included 2 cases from an article of a 13-case series that was published recently because the 2 cases were PBC without Sjögren’s syndrome. There were 16 similar cases, including our own.

Figure 1. (a) Renal histology findings on the first and the second admissions. Periodic acid–Schiff (PAS) staining showed severe lymphocyte infiltration in the tubulointerstitial area accompanied by tubular atrophy at the first admission, and the infiltration in the tubulointerstitial area was not improved at the second admission. IgM-positive cells were dominantly observed than IgG-positive cells in the tubulointerstitial area. Bars = 100 μm. (b) Hepatic histology findings at the second admission. Hematoxylin and eosin (HE) staining of a liver biopsy specimen showed inflammatory cell infiltration surrounding the bile ducts at a low magnification and epithelioid granuloma formation at a high magnification. Although there were many IgM-positive cells, IgG-positive cells were scarce. Bars = 100 μm.
At the first admission, CD3-positive cells were mixed with CD20- or CD138-positive cells in the tubulointerstitial area. CD138-positive cells were more abundant in the tubulointerstitial area than CD20-positive cells. At the second admission, CD3-positive cells were more dominantly observed than CD20- or CD138-positive cells in the tubulointerstitial area. In the liver, CD3-positive cells were mixed with CD20- or CD138-positive cells in the inflammatory cell infiltrates. Bars = 100 μm.

Table 2. Summary of the reported cases of tubulointerstitial nephritis with primary biliary cirrhosis

| Age/sex | U-β2mg (µg/d) | U-NAG (UI) | Urine pH | sCr (mg/dl) | AST (UI) | ALT (UI) | ALP (UI) | γGTP (UI) | IgM (mg/dl) | AMA | M2A | FS | OM |
|---------|-----------------|-------------|----------|-------------|-----------|----------|----------|------------|-------------|-----|-----|----|----|
| Mcdougall et al.1 1987 | 50/F | NA | NA | 2 | NA | 117 | NA | 630 | 800 | NA | x | x |
| Kodama et al.6 1996 | 36/F | NA | NA | 0.8 | 19 | 14 | 83 | 32 | 1260 | 640 | (--) | o | x |
| Lino et al.10 2005, pat 1 | 51/F | 25 mg/24 h | NA | 6.5 | 1.8 | 41 | 67 | 107 | 53 | NA | NA | 800 | o | o |
| Lino et al.10 2005, pat 2 | 68/F | 57 mg/24 h | NA | 6 | 1.3 | 25 | 34 | 276 | 51 | NA | NA | 640 | o | o |
| Terrier et al.11 2008 | 42/F | 263,000 | NA | NA | 1.3 | 3N | 2N | 4.5N | 3.5N | 358 | NA | 200 | o | o |
| Komatsu et al.10 2010, pat 1 | 36/F | 10,595 | NA | 7.3 | 0.8 | 19 | 14 | 83 | 32 | 1260 | 640 | (--) | o | x |
| Komatsu et al.10 2010, pat 2 | 66/F | 84,400 | 21.1 | 6.3 | 1.5 | 20 | 10 | 290 | 13 | 663 | 80 | 178 | o | x |
| Komatsu et al.10 2010, pat 3 | 77/F | 15,586 | 2.5 | 7 | 1.4 | 32 | 19 | 152 | 37 | 74 | 20 | 40 | x | x |
| Komatsu et al.10 2010, pat 4 | 52/F | 60,000 | 7.4 | 7.5 | 1.5 | 25 | 19 | 812 | 24 | 665 | 320 | 157 | o | x |
| Bonsel et al.12 2012 | 46/F | NA | NA | 2.3 | NA | NA | 209 | 252 | NA | (+) | (+) | x | x |
| Iwakura et al.13 2013 | 46/F | 20,975 | 11.1 | NA | 1.1 | 38 | 50 | 694 | 190 | 966 | 160 | 143 | x | x |
| Rasoulofdegan et al.14 2014 | 28/F | NA | NA | 2 | 185 | 138 | NA | 698 | NA | (--) | 19.5 | x | x |
| Yonemochi et al.15 2015 | 49/F | NA | NA | 6 | 1.4 | 23 | 24 | 663 | 18 | 1084 | NA | 10 | o | o |
| Takahashi et al.16 2017, pat 12 | 44/F | 69,359 | NA | NA | 1.4 | NA | NA | 1559 | 56 | 1284 | NA | 562 | o | x |
| Takahashi et al.16 2017, pat 13 | 56/F | 66,987 | NA | 2.7 | NA | NA | 806 | 178 | 2045 | (--) | 10 | o | x |
| Present case | 57/F | 47,456 | 6.8 | 8 | 1.5 | 19 | 18 | 283 | 31 | 959 | 40 | 400 | o | x |

ALP, alkaline phosphatase; ALT, alanine transaminase; AMA, anti-mitochondrial antibody; AST, aspartate transaminase; F, female; FS, Fanconi syndrome; γGTP, γ-glutamyl transpeptidase; M, male; M2A, anti-mitochondrial M2 antibody; NA, not available; OM, osteomalacia; pat, patient; sCr, serum creatinine; U-β2mg, urinary β2-microglobulin; U-NAG, urinary N-acetyl-β-D-glucosaminidase; o, yes; x, no.
All 16 patients were female, and 11 were diagnosed with Fanconi syndrome. Of the 16 patients, 12 did not have osteomalacia. The long-term use of corticosteroids in patients with PBC is contraindicated because of the progression of osteoporosis. However, treatment with corticosteroids was successful when the patients had TIN, and the common dose was 0.5 to 1 mg/kg per day (Table 3). We started treating our patient with 0.8 mg/kg per day of prednisolone. Although TIN temporarily improved after the treatment, it deteriorated during prednisolone tapering. Relapse of TIN after prednisolone tapering seemed to be rare, as only 1 case has been reported (by Macdougall et al.) aside from the present case (Table 3). Because we judged TIN still to be active based on the findings of the second renal biopsy, we performed methylprednisolone pulse therapy, and the TIN improved. To our knowledge, this is the first reported case of TIN with PBC in a patient who was successfully treated with steroid pulse therapy. Bisphosphonates were used to prevent osteoporosis, and the bone density of the lumbar spine was 129% compared to the average in women of the same age.

Takahashi et al. recently reported a disease group of TIN with high serum IgM levels. The investigators observed CD138- and IgM—double-positive plasma cells in the tubulointerstitial area. In the present case, IgM-positive cells were more dominantly observed than IgG-positive cells in both the kidney and the liver. Although we were unable to perform double staining with CD138 and IgM, CD138-positive cells were also predominantly observed in the first renal biopsy, and we surmised that some of these cells were positive for IgM.

In conclusion, we diagnosed a rare case of IgM-positive TIN associated with asymptomatic PBC.

DISCLOSURE
MI received departmental scholarship funds of $1,000,000 yen per year for direct expenses from Merck Sharp & Dohme K.K., Bristol-Myers Squibb, BIOTORONIK JAPAN, Inc., Astellas Pharma Inc., Shionogi & Co., Ltd., and Otsuka Pharmaceutical Co., Ltd., in 2015, Merck Sharp & Dohme K.K., Astellas Pharma Inc., Takeda Pharmaceutical Company, Ltd., Daiichi Sankyo Company, Ltd., and Otsuka Pharmaceutical Co., Ltd., in 2016, and Merck Sharp & Dohme K.K., Shionogi & Co., Otsuka Pharmaceutical Co., Ltd., Takeda Pharmaceutical Company, Ltd., and Daiichi Sankyo Company, Ltd., in 2017. The companies associated with this funding were not involved in the manuscript preparation or the decision to publish the present case report. All the other authors declared no competing interests.

AUTHOR CONTRIBUTIONS
SM and TM took clinical care of the patient and participated in the acquisition of clinical data. SM, KK, TM, EL, and YK carried out analysis of patient’s clinical course and data interpretation. SM and KK wrote a draft of the manuscript and TM, EL, YK, YHo, KO, YHi, TI, and MI revised it critically. All authors read and approved the final manuscript.

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