Genetic Polymorphisms in Genes Involved in the Type I Interferon System (IFIH1/MDA-5, TNFAIP3/A20, and STAT4): Association with SLE Risk in Egyptian Children and Adolescents

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Purpose: Systemic lupus erythematosus (SLE) is a multifactorial autoimmune inflammatory disease that is influenced by both genetic and environmental factors and associated with dysregulation of type I interferon (INF) response. This study aimed to investigate the effects of single nucleotide polymorphisms (SNPs) of the IFIH1, TNFAIP3, and STAT4 genes in the type I INF system on SLE risk in Egyptian children and adolescents.

Patients and Methods: We recruited 94 SLE individuals and 94 healthy subjects. SNPs of IFIH1 rs3747517 C/T, TNFAIP3 rs610604 G/T, and STAT4 rs7574865 G/T were evaluated using TaqMan™ SNP genotyping assay.

Results: Individuals with the TT, CT+TT genotypes, and T allele of rs3747517 in the IFIH1 gene were protective for SLE patients (OR = 0.429, 95% CI = 0.191–0.962, P = 0.040), (OR = 0.685, 95% CI = 0.477–0.984, P = 0.041), and (OR = 0.705, 95% CI = 0.527–0.944, P = 0.019), respectively. Also, individuals with the TT, GT+TT genotypes, and T allele of rs7574865 in the STAT4 gene were associated with SLE risk (OR = 3.945, 95% CI = 1.303–11.947, P = 0.015), (OR = 1.536, 95% CI = 1.058–2.231, P = 0.024), and (OR = 1.522, 95% CI = 1.113–2.082, P = 0.009), respectively. In the case of TNFAIP3 rs610604, no significant association of genotypes or alleles with SLE were detected, while the three SNPs did not show any significant association with the SLE clinical or laboratory features.

Conclusion: Our findings indicated that rs3747517 in IFIH1 was protective for SLE in Egyptian children and adolescents. Moreover, rs7574865 in STAT4 not rs610604 in TNFAIP3 was associated with SLE risk.

Keywords: systemic lupus erythematosus, type I interferon system, autoimmune disease, genetic polymorphism

Introduction

SLE is a multifactorial autoimmune disease involving genetic and environmental factors, which is characterized by pathogenic autoantibodies, chronic inflammation, and dysregulation in type I IFN pathway function.1,2 Type I interferons (IFNs) are innate immune response mediators that enhance the adaptive immune response by acting on dendritic cells, T and B cells, and natural killer cells in both direct and indirect ways.3 Genetic differences in the type I IFN pathway genes represented a common genetic factor of SLE.4

The interferon-induced Helicase C domain 1 (IFIH1, also known as melanoma differentiation-associated protein 5, MDA5) gene, is found on chromosome 2p24.3.
The IFIH1 gene is a cytosolic RNA-responsive sensor that causes type I INF to progress by inducing INF regulatory factors 3,7 and the proinflammatory transcription factor nuclear factor-kB (NF-kB).\textsuperscript{5–7} Furthermore, NF-kB activation is a key factor in the pathogenesis of many inflammatory diseases.\textsuperscript{8} The tumor necrosis factor-alpha-induced protein 3 (TNFAIP3, also known as A20) gene, the ubiquitination enzyme, is located on the 6q23 chromosome. TNFAIP3 has emerged as a central negative regulator of the NF-kB signaling pathway and inflammation.\textsuperscript{9} The signal transducer and activator of transcription 4 (STAT4) gene have a key role in the activation of dendritic cells and macrophages, as well as the signal transductions within activated peripheral blood monocytes.\textsuperscript{10} It is located on the 2q32.3 chromosome.\textsuperscript{11}

SNPs are the most common form of genetic variation in the human genome, and they can change the function of a gene if they are present.\textsuperscript{12} Polymorphisms in genes encoding proteins that are main components of the type I IFN pathway, such as IFIH1, TNFAIP3, and STAT4, have also been related to the risk of developing autoimmune disorders, including SLE.\textsuperscript{2} Here, we conducted the present study to clarify the hypothesis that SNPs in IFIH1, TNFAIP3, and STAT4 may affect the susceptibility of patients with SLE.

**Patients and Methods**

**Patients**

This research included 94 Egyptian pediatric and adolescent participants (82 females and 12 males with an average age of 13.7 years) and 94 controls (80 females and 14 males with the mean age of 13.5 years). All patients with SLE were diagnosed using the American College of Rheumatology (ACR) guidelines from 1997.\textsuperscript{13} Systemic lupus erythematosus Disease Activity Index (SLEDAI) was measured by the Systemic Lupus International Collaborating Clinics/American College of Rheumatology (SLICC/ACR) damage index (SDI).\textsuperscript{14} Renal biopsy grades of SLE patients with nephritis according to the International Society of Nephrology/Renal Pathology Society (ISN/RPS) 2003 classification of lupus nephritis, as well as SLE demographic data and clinical features, were obtained from medical records filled in the Medical Archive Service and Health Information (Figures 1 and 2). All participants were enrolled between May 2018 and August 2020 from the Outpatient Clinic of pediatric nephrology (Mansoura University Children’s Hospital, Faculty of Medicine, Egypt). Each control was a healthy individual, did not suffer from any systemic disease, and had no family relation with SLE patients. The protocol was approved by the Institutional Research Board (IRB) of the Faculty of Medicine (Mansoura University, Egypt), and written informed parental consent, and child assent from participants was obtained. The study was conducted at the Laboratory according to the guidelines of the Declaration of Helsinki.

**Routine Laboratory Tests**

We determined serum Anti-Nuclear Antibodies (ANA) by using enzyme-linked immunosorbent assay (ELISA) technique (Euroimmun, Germany) and Anti-ds-DNA by

![Figure 1](https://doi.org/10.2147/JIR.S309008)

**Figure 1** Renal biopsy classes (stage I, II, III, IV and V) in lupus nephritis cases.
ELISA technique (INOVA Diagnostic, USA). We assessed serum complement levels (C3 and C4) with a turbidimetric assay (DIALAB GmbH, Austria). Estimation of Calcium and creatinine by using computerized DIALAB, Austria, and Phosphorous using SPINREACT, Spain. Assays had been accomplished according to the manufacturer’s instructions.

DNA Isolation and Genotyping
Genomic DNA was extracted from the peripheral blood using QIAamp DNA Blood Kit (Qiagen, Germany). Genotyping of the IFIH1 (ID: C__25984418_10 for rs3747517 C/T), TNFAIP3 (ID: C____884105_20 for rs610604 G/T) and STAT4 (ID: C__29882391_10 for rs7574865 G/T) variants were performed using the TaqMan® genotyping assays (Thermo Fisher Scientific Inc., Waltham, MA, USA). PCR reaction was conducted on Artus Rotor-Gene Qiagen fast real-time PCR System (Applied Biosystems, software 2.1.0).

Statistical Analysis
The Statistical Package for Social Science (IBM Corp., 2017) was used to analyze the data. Version 25.0 of IBM SPSS Statistics for Windows (Armonk, NY: IBM Corp.). The statistical significance of the difference in parametric data between the two groups was determined using the Student’s T-test. Kruskal Wallis test was used for differences between nonparametric numerical data among three groups. The relationship between qualitative variables was determined using the Chi-square test or Fisher’s exact test.

The Goodness-of-Fit test was used to determine deviations from Hardy–Weinberg equilibrium expectancy. Moreover, Logistic regression analysis was used for the prediction of risk factors, using generalized linear models. P-values less than 0.05 were considered significant.

Results
Demographic Manifestations of the Study Population
Demographics and basic information for SLE patients and controls are summarized in Table 1, Figure 3.

Association of IFIH1 rs3747517 C/T, TNFAIP3 rs610604 G/T, and STAT4 rs7574865 G/T Polymorphisms with the Risk of SLE
Full genotypes and alleles data were corresponded to Hardy-Weinberg equilibrium (HWE) in patients and controls; Table 2. The IFIH1 rs3747517 “TT”, “CT+TT” genotypes and “T” allele were significantly lowered in patient group compared with control subjects {(OR = 0.429, 95% CI = 0.191–0.962, P = 0.040), (OR = 0.685, 95% CI = 0.477–0.984, P = 0.041) and (OR = 0.705, 95% CI = 0.527–0.944, P = 0.019)}, respectively; Table 2. However, no significant evidence was detected in “CT” genotype (P=0.108); Table 2.

As shown in Table 2, no significant difference in TNFAIP3 rs610604 genotype and allele distributions were found between patients and controls (P>0.05).
We also determined no association of rs610604 SNP with SLE under the combined model (“GT+TT”, $P = 0.205$).

The frequencies of the “TT”, “GT+TT” genotypes and “T” allele of rs7574865 in STAT4 gene were significantly increased in SLE cases compared with control group (OR= 3.945; 95% CI, 1.303–11.947, $p = 0.015$), (OR= 1.536; 95% CI, 1.058–2.231, $p = 0.024$) and (OR= 1.522; 95% CI, 1.113–2.082, $p = 0.009$), respectively; Table 2.
Association of IFIH1 rs3747517, TNFAIP3 rs610604, and STAT4 rs7574865 SNPs with Clinical and Laboratory Features in SLE Cases

Regression analysis showed no potential genetic association between the IFIH1, TNFAIP3, and STAT4 with clinical and laboratory features (P> 0.05; Table 3). However, Genotypes “CT+TT” in IFIH1 were significantly associated with the lower of SLE severity in univariate analyses and multivariate analysis {(OR =0.685, 95% CI: 0.477–0.984, P=0.041) and (OR =0.638, 95% CI: 0.440–0.924, P=0.018)}, respectively; Table 4. Furthermore, STAT4 “GT+TT” genotype was significantly associated with the risk of SLE in univariate and multivariate analysis {(OR =1.536, 95% CI: 1.058–2.231 P=0.024) and (OR =1.644, 95% CI: 1.122–2.408, P=0.011)}, respectively; Table 4. While no significant association was detected in the case of genotype “GT+TT” in TNFAIP3 (P=0.205; Table 4).

Discussion

SLE is a multifaceted autoimmune inflammatory disorder with several clinical features including immunologic defects, blood cell abnormalities, arthritis, kidney disorder, and skin disease.

Similarly, immunological, hematological, renal, and cutaneous disorders were predominated in SLE cases in our research. In the present study, we investigated the association between IFIH1 rs3747517, TNFAIP3 rs610604, and STAT4 rs7574865 SNPs and SLE in Egyptian pediatric and adolescents. Type I IFNs play a key role in the pathogenesis and severity of autoimmune diseases including, SLE. Previous studies have related to the IFIH1, TNFAIP3, and STAT4 polymorphisms with a variety of inflammatory and autoimmune disorders such as multiple sclerosis, type 1 diabetes, psoriasis, inflammatory diseases, and SLE.

IFIH1 is found in various immune system cells and is involved in autoimmune disorders such as SLE. IFIH1 increased IFN production by activation of NF-kB, IFN-regulatory factors 3 and 7. For the first time, we confirmed that IFIH1 rs3747517 plays a beneficial role in Egyptian children and adolescents with SLE. Our results indicated that SLE patients carrying IFIH1 “TT”, “CT+TT” genotypes, and “T” allele have a potential effect on lowering the risk of SLE. Therefore, we suggested that a protective role of the IFIH1 rs3747517 gene in SLE cases. Similarly, the IFIH1 rs3747517 polymorphism was found to be protective against SLE by Wang et al (P=0.025). A haplotype analysis with IFIH1 rs78456138 and rs3747517 revealed a protective effect against SLE development in the same study. However, “CT” and “CC+CT/TT” genotypes at IFIH1 rs3747517 were linked to an

Figure 3 SLE features.
Abbreviations: SLE, systemic lupus erythematosus; ANA, anti-nuclear antibodies.
increased risk of SLE in the Chinese Han population. On the other hand, Van Eyck et al found no association between the IFIH1 rs3747517 polymorphism and SLE susceptibility in Brazil in a previous study. Moreover, ethnic differences in the allele distributions of rs3747517 have been observed among European, Asian, and African individuals. Also, we failed to detect any significant association between polymorphism of rs3747517 in IFIH1 and other clinical or laboratory investigations of SLE, Table 3. Our results reinforced a previous study that found no association between the IFIH1 rs3747517 SNP and positive anti-dsDNA in SLE patients.

The A20 encoded by TNFAIP3 has emerged as a key negative regulator of nuclear factor-KB-mediated inflammatory signaling. Multiple SNPs in the promoter, intronic regions, and coding exons of the TNFAIP3 gene have been discovered, and they may play a role in inflammatory and autoimmune diseases including SLE.
| Clinical/Laboratory Features | IFIH1 rs3747517 C/T | P | TNFAIP3 rs610604 G/T | P | STAT4 rs7547865 G/T | P |
|-----------------------------|--------------------|---|----------------------|---|----------------------|---|
|                            | CC N=58            | CT N=33 | TT N=3               |   | GG N=10              | GT N=46 | TT N=38               |   | GG N=51              | GT N=35 | TT N=8               |   |
| Cutaneous                   | 27 (46.6%)         | 22 (66.7%) | 2 (66.7%)            | 0.157 | 6 (60%)              | 27 (58.7%) | 18 (47.4%)            | 0.699 | 27 (52.9%) | 18 (51.4%) | 6 (75.0%)            | 0.781 |
| Ulcers                      | 3 (5.2%)           | 6 (18.2%) | 1 (33.3%)            | 0.375 | 2 (20%)              | 4 (8.7%) | 4 (10.5%)            | 0.340 | 5 (9.8%) | 4 (11.4%) | 1 (12.5%)            | 0.775 |
| Alopecia                    | 14 (24.1%)         | 14 (42.4%) | 0 (0.0%)             | 0.131 | 2 (20%)              | 16 (34.8%) | 10 (26.3%)            | 0.467 | 16 (31.4%) | 9 (25.7%) | 3 (37.5%)            | 0.714 |
| Arthritis                   | 16 (27.6%)         | 9 (27.3%) | 1 (33.3%)            | 0.984 | 5 (50%)              | 15 (32.6%) | 6 (15.8%)            | 0.111 | 16 (31.4%) | 8 (22.9%) | 2 (25.0%)            | 0.380 |
| Serositis                   | 2 (3.4%)           | 1 (3.0%) | 0 (0.0%)             | 0.857 | 0 (0.0%)             | 0 (0.0%) | 3 (7.9%)             | 1.000 | 2 (3.9%) | 0 (0.0%) | 1 (12.5%)            | 0.661 |
| Neurologic                  | 7 (12.1%)          | 3 (9.1%) | 0 (0.0%)             | 0.566 | 0 (0.0%)             | 7 (15.2%) | 3 (7.9%)             | 0.593 | 7 (13.7%) | 3 (8.6%) | 0 (0.0%)            | 0.290 |
| Hematological disorder      | 40 (69.0%)         | 27 (81.8%) | 3 (100%)             | 0.118 | 6 (60%)              | 34 (73.9%) | 30 (78.9%)            | 0.284 | 37 (72.5%) | 27 (77.1%) | 6 (75.0%)            | 0.656 |
| Renal biopsy stages I       | 1 (50%)            | 1 (50%) | 0 (0.0%)             | 0.400 | 1 (50%)              | 1 (50%) | 0 (0.0%)            | 0.191 | 0 (0.0%) | 2 (100%) | 0 (0.0%)            | 0.140 |
| II                          | 7 (77%)            | 1 (11.1%) | 1 (11.1%)            | 7 (77.8%) | 2 (22.2%)            | 0 (0.0%) | 16 (55.2%)            | 9 (31%) | 4 (13.8%) | 1 (5%) | 0 (0.0%)            | 0.020 |
| III                         | 17 (58.6%)         | 12 (41.4%) | 0 (0.0%)            | 2 (6.90%) | 14 (48.3%) | 13 (44.8%) | 8 (40%)            | 4 (100%) | 0 (0.0%) | 0 (0.0%) | 0.020 |
| IV                          | 10 (50%)           | 9 (45%) | 1 (5%)              | 5 (25%) | 7 (35%) | 8 (40%) | 4 (100%) | 0 (0.0%) | 0 (0.0%) | 0 (0.0%) | 0.020 |
| V                           | 3 (75%)            | 1 (25%) | 0 (0.0%)            | 1 (25%) | 2 (50%) | 1 (25%) | 1 (25%) | 1 (25%) | 1 (25%) | 1 (25%) | 0.020 |
| Non proliferative           | 4 (66.7%)          | 2 (33.3%) | 0 (0.0%)             | 0.701 | 2 (33.3%) | 3 (50%) | 1 (16.7%)            | 0.192 | 4 (66.7%) | 2 (33.3%) | 0 (0.0%)            | 0.535 |
| Proliferative               | 34 (58.6%)         | 22 (37.9%) | 2 (3.4%)            | 7 (12.1%) | 28 (48.3%) | 23 (39.7%) | 31 (53.4%) | 20 (34.5%) | 7 (12.1%) | 0.020 |
| SLEDAI (medium (range))     | 8 (0–19)           | 11 (5–9) | 4 (4–6)             | 0.177 | 6.5 (0–12) | 10 (4–19) | 8 (0–19)            | 0.177 | 8 (0–19) | 10 (0–19) | 9.5 (6–17)            | 0.382 |
| Active SLE                  | 28 (48.3%)         | 19 (57.6%) | 0 (0.0%)            | 0.671 | 4 (40%) | 27 (58.7%) | 16 (42.1%) | 0.504 | 25 (49%) | 18 (51.4%) | 4 (50%)            | 0.836 |
| Anti-ds-DNA                 | 37 (63.8%)         | 26 (78.8%) | 2 (66.7%)            | 0.152 | 9 (90%) | 29 (63.0%) | 27 (71.1%) | 0.128 | 37 (72.5%) | 23 (65.7%) | 5 (62.5%)            | 0.438 |
| ANA                         | 52 (89.7%)         | 31 (93.9%) | 3 (100%)            | 0.416 | 9 (90%) | 43 (93.5%) | 34 (89.5%) | 0.860 | 47 (92.2%) | 32 (91.4%) | 7 (87.5%)            | 0.801 |
| Hypocomplementemia          | 56 (96.6%)         | 32 (97.0%) | 3 (100%)            | 0.857 | 9 (90%) | 45 (97.8%) | 37 (97.4%) | 0.257 | 49 (96.1%) | 35 (100%) | 7 (87.5%)            | 0.661 |

**Notes:** Values in parentheses are percentages or data are presented as percentages and medium (range); P>0.05 is statistically non-significant. Test, Chi-square for categorical data, Kruskal Wallis test for numerical data.

**Abbreviations:** N, number in each group; SLE, systemic lupus erythematosus; SLEDAI, systemic lupus erythematosus Disease Activity Index; ANA, anti-nuclear Antibodies; Anti-ds-DNA, anti-double-stranded DNA; IFIH1, interferon-induced helicase C domain 1; TNFAIP3, tumor necrosis factor alpha-induced protein 3; STAT4, signal transducer and activator of transcription 4.
a few studies in the literature investigated the TNFAIP3 rs610604 polymorphism in SLE. Our study found the frequencies of genotype and allele in rs610604 were higher in SLE patients than the healthy control group but no statistically significant differences between the two studied groups as cleared in Table 2. This observation confirmed no association between examined rs610604 SNP in TNFAIP3 and SLE risk in Egyptian populations. In agreement with a previous study reported by Lodolce et al, genetic polymorphism in TNFAIP3 rs610604 was not linked to SLE susceptibility in African American SLE patients.\(^27\)

STAT proteins are involved in the signaling of many cytokines, hormones, and growth factors that regulate key cellular processes such as survival, proliferation, and differentiation, and are needed for IFN-dependent gene expression.\(^28\) In this study, we assessed the role of the STAT4 rs7574865 variant in SLE susceptibility. It is well documented that STAT4 can be activated by type I IFNs and STAT4 genetic variants have been linked to the development of SLE in several populations.\(^28,29\) The STAT4 gene’s SNP rs7574865 in the third intron\(^30\) is one of the most commonly studied polymorphisms\(^31\) and has been linked to the severity of SLE, with conflicting results.\(^32\) In the present study, it was found that in the Egyptian pediatric and adolescents, STAT4 rs7574865 G/T genotype displayed an increased risk of SLE and this result confirmed an association of the polymorphism with SLE. We found individuals with TT, GT+TT genotypes, and T allele were significant positive risk and severe factors for SLE. Although GT genotype is more frequent in SLE patients than in controls, no significance was detected (\(P>0.05\); Table 2). Our genetic data agree with the meta-analysis performed by Wang et al, which revealed that STAT4 rs7574865 G/T SNP is linked to the severity of SLE in Argentinians, Colombians, Mexicans, Japanese, Chinese, and Iranians.\(^32\) Our findings confirm and extend the results of the previous study in Egyptian cohorts with adult-onset SLE.\(^33\) On the other hand, the study performed by Zervou et al found that there was no significant association in Turkish cohorts.\(^34\) This may be attributed to disparities between racial and ethnic groups.

Although, our study demonstrated that the STAT4 rs7574865 associate with the SLE risk, but we did not confirm any association between rs7574865 polymorphism and SLE clinical or laboratory parameters; Table 3. Similarly, Mirazkemi et al\(^35\) and Yin et al\(^35\) found no connection between the same SNP in the STAT4 gene and clinical manifestations of SLE in their studies. Our results were contradicted with Yang et al, where the STAT4 rs7574865 variant was linked to positive anti-dsDNA antibodies\(^36\) and Kadota et al, the rs7574865 T allele in the STAT4 gene was linked to cutaneous manifestation such as malar rash in SLE patients.\(^37\) Furthermore, the rs3747517 “CT+TT” genotype was linked to SLE protection, while the rs7574865 “GT+TT” genotype was associated with disease risk in both univariate and multivariate tests; Table 4. SNPs in IFIH1 and STAT4 were considered to be independent predictors of SLE in our present study. These discrepancies between our genetic data and previous studies may be explained by both genetic and environmental influences, attributed to race/ethnic diversity, disease duration as well as variations in sample size and studied age.

Our research has a few limitations. Since the sample size of the patients examined was so small, large sample studies are needed. Besides, the patients enrolled in this study were all from the Egyptian Delta area. We were unable to obtain cases from other regions to include in our study to investigate the effect of people’s residence on their outcomes. Also, we only look at one genetic variation per gene. To understand the function of these genes in the severity of SLE, further research into more variants in the IFIH1, TNFAIP3, and STAT4 genes is needed.
Conclusion
SLE is triggered by a complex interaction of genetic and environmental factors, which is well understood. The IFIH1 rs3747517 polymorphism was found to have a protective effect in Egyptian children and adolescents with SLE in our research. Also, STAT4 rs7574865, not TNFAIP3 rs610604 SNPs, was linked to SLE risk. Furthermore, our study found no evidence of a correlation between the three SNPs studied and any of the clinical or laboratory features of SLE. However, further research is required to validate our findings.

Abbreviations
SLE, systemic lupus erythematosus; INF, interferon; SNPs, single nucleotide polymorphisms; IFIH1, interferon-induced Helicase C domain 1; TNFAIP3, tumor necrosis factor alpha-induced protein 3; STAT4, signal transducer and activator of transcription 4; MDA5, melanoma differentiation-associated protein 5; NF-kB, transcription factor nuclear factor-kB; SLICC/ACR, Systemic Lupus International Collaborating Clinics/American College of Rheumatology; SLEDAI, systemic lupus erythematosus Disease Activity Index; IRB, Institutional Research Board; ISN/RPS, International Society of Nephrology/Renal Pathology Society; ANA, Anti-Nuclear Antibodies; Anti-ds-DNA, Anti-double stranded DNA; ELISA, enzyme-linked immunosorbent assay; C3 & C4, complement components 3 and 4.

Data Sharing Statement
All data detected or analyzed during this current study are involved in this published work.

Ethics Approval and Consent to Participate
The study protocol was approved by the Institutional Research Board (IRB) of the Faculty of Medicine of Mansoura University. Written informed parental consent, and child assent from participants was obtained before any research procedures were conducted. This study was carried out under the guidelines of the Declaration of Helsinki.

Author Contributions
All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis, and interpretation, or in all these areas; took part in drafting, revising, or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

Disclosure
The authors report no conflicts of interest in this work.

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