Mechanisms and Advances in Anti-Ovarian Cancer with Natural Plants Component

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Abstract: Ovarian cancer ranks seventh in the most common malignant tumors among female disease, which seriously threatens female reproductive health. It is characterized by hidden pathogenesis, missed diagnosis, high reoccurrence rate, and poor prognosis. In clinic, the first-line treatment prioritized debulking surgery with paclitaxel-based chemotherapy. The harsh truth is that female patients are prone to relapse due to the dissemination of tumor cells and drug resistance. In these circumstances, the development of new therapy strategies combined with traditional approaches is conductive to improving the quality of treatment. Among numerous drug resources, botanical compounds have unique advantages due to their potentials in multitarget functions, long application history, and wide availability. Previous studies have revealed the therapeutic effects of bioactive plant components in ovarian cancer. These natural ingredients act as part of the initial treatment or an auxiliary option for maintenance therapy, further reducing the tumor and metastatic burden. In this review, we summarized the functions and mechanisms of natural botanical components applied in human ovarian cancer. We focused on the molecular mechanisms of cell apoptosis, autophagy, RNA and DNA lesion, ROS damage, and the multiple-drug resistance. We aim to provide a theoretical reference for in-depth drug research so as to manage ovarian cancer better in clinic.

Keywords: ovarian cancer; natural plant products; molecular mechanisms; bioactive compounds; traditional Chinese medicine

1. Cognitive Status of Ovarian Cancer

Ovarian cancer is the seventh most common malignant tumor cancer and the fifth leading cause of death among female reproductive diseases [1]. The number of cases diagnosed is steadily increasing with the higher life expectancy, while wildly recognized preventative measures or effective treatments have not kept up yet. The disease is not specifically diagnosed until an advanced stage, causing a relatively high morbidity and mortality. Correspondingly, symptoms progress insidiously as tumorigenesis, including pelvis pain and lower abdomen pain, vaginal bleeding during menstrual periods, unregular bleeding after menopause, bloating, and changes in urinary or bowel habits. In some cases, the primary cancer cells attack adjacent tissue and spread to distant organs. The metastasis of ovarian cancerous cells in the peritoneal cavity actually move for a long time without obvious symptoms, and the routes of dissemination of ovarian cancer include intracavitary implantation, hematogenous metastasis, and lymphatic metastasis [2]. On the one hand, its threatening aggressiveness relies on the rapid dissemination of cancer cells to adjacent tissues, such as the peritoneum, the omentum, and abdominal organs. On the other
hand, the development of drug resistance to chemotherapeutic agents cannot be neglected. Histopathologically, nearly 90% of ovarian cancer originates from the ovarian surface epithelium (OSE), with the rest being classified as serous, mucinous, endometrioid, transitional, and clear cell carcinomas [3,4]. It is a heterogeneous disease involved with inherited mutations in susceptibility genes, such as p53 tumor suppressor gene, ERBB2, and PIK3CA oncogenes, etc. Approximately 10% of ovarian cancer occurs in women carrying BRCA1 or BRCA2 mutations [5,6], causing some ovarian cancers clustering in families. Moreover, various classic signaling pathways participate in oncogenesis. For instance, Wnt/β-catenin target genes mediate cancer initiation and progression. PI3K/AKT/mTOR plays a regulatory role in cell survival, growth, and proliferation [7,8]. Notch signal pathway is associated with cell proliferation, migration, cell stemness, and chemoresistance [9]. Hyperactivation of Hedgehog signal transduction also participates in chemotherapy-resistant phenotype, because the overexpression of transcription factor is connected to the regulation of ABCB1 and ABCG2 gene expression in ovarian cancer [10].

2. Current Treatments for Ovarian Cancer

At present, clinic treatment prioritized debulking surgery along with platinum-based chemotherapy, for example, cyclophosphamide and doxorubicin hydrochloride. The standard chemotherapy regimens are taxanes including paclitaxel and docetaxel and platinum agents such as carboplatin and cisplatin [11]. In combination with paclitaxel and carboplatin, humanized monoclonal antibody bevacizumab is administered for patients with advanced ovarian cancers. The harsh truth is that quite a number of patients who received and responded to chemotherapy are prone to develop chemotherapy resistance. Moreover, the cumulative toxic effects included but were not limited to nephrotoxicity, neurotoxicity, ototoxicity with cisplatin, and myelosuppression with carboplatin [12]. Although early treatment is effective for most patients, relapse is prone to occur within a median of 16 months in advanced-stage patients [13]. In particular, the genetic alterations lead to increasing repairs of DNA lesions, derangements of intracellular signal transduction, and drug metabolic inactivation. The success of complete remission relied on surgical skills accompanied with the gene signature of the tumor [14]. Patients who carried BRCA1 or BRCA2 germline mutations are recommended for olaparib administration as a poly ADP-ribose polymerase (PARP) inhibitor [15]. It is also applied for relapsed and subsequent platinum-based chemotherapy.

However, once it relapses, the interval between subsequent treatments declines steadily due to rapid tumor progression and chemotherapy resistance. Novel options rather than only traditional chemotherapy are needed to improve the survival quality of patients for systemic treatment. Among the potential drug resources, botanical components are natural antioxidant constituents due to their long history in ethnopharmacology. Conventionally, plant-derived natural products are considered to be auxiliary nutritional supplementary. Meanwhile, as part of an initial treatment or an option for maintenance therapy, the effective components from natural plants could improve multiple-drug sensitivity, further reducing the tumor and metastatic burden of ovarian cancer. In traditional Chinese medicine, triptolide is a herb-derived antineoplastic agent, which has been proved to selectively kill both p53 mutated and p53 wild-type ovarian cancerous cells [16]. Emodin is a natural anthraquinone extracted from traditional Chinese herbs, which was identified to inhibit the growth of human ovarian carcinomas [17]. Potential herbal remedies have advantages in wide availability, few side effects, and are generally well tolerated. From experimental findings to clinical applications, classic herbal extracts have been broadly used for management in ovarian cancer.

3. Mechanisms with Natural Plants Compound in Ovarian Cancer Cells

3.1. Cytotoxic Effect

Cryptotanshinone, tanshinone-I, and tanshinone-IIA are natural components mainly isolated from the roots of Salvia miltiorrhiza Bunge, which is named Danshen in traditional
Chinese herbs [18]. TNF-related apoptosis-inducing ligand (TRAIL) is known as ‘death receptors’ in the TNF superfamily, which initiates the cell death progression by binding to its associated receptor [19,20]. In TRAIL-resistant cells A549 and human ovarian cancer cells TOV-21G, exposure with three tanshinones effectively enhanced the functionality of TRAIL, thus reducing cell viability and inhibiting the colony formation capacity. Especially, tanshinone-IIA had the most potential among all for displaying an IC$_{50}$ of 2.00 ± 0.36 µM and 2.75 ± 0.23 µM in both A549 and TOV-21G cell lines [21]. In a dose-dependent pattern, TII-A enhanced JNK-mediated signaling transduction. TII-A augmented the expression levels of DR5 and mRNA, leading to TRAIL-sensitive cytotoxicity [22]. Kadsuphilactone B is a nortriterpenoid compound derived from traditional Chinese herb Schisandra chinensis (Turcz) Baill. In ovarian cancer cells A2780 and Ishikawa, it exhibited a cytotoxic effect with IC50 values below 25 µM. In a dose-dependent pattern, kadsuphilactone B promoted the stimulations of caspase-3/8/9, MAPKs, cleaved PARP, and alterations in Bcl-2 family proteins levels [23]. Methyl lucidone (ML) is an herbal component contained in the dried fruits of L. erythrocarpa Makino. In SKOV-3 and OVCAR-8 cell lines, ML exhibited a cytotoxic effect by displaying the IC$_{50}$ of 48.8–60.7 and 33.3–54.7 µM. Treatment with ML induced obvious cellular apoptosis and morphological changes. ML not only stimulated the activation of cleaved caspase-3/9 and PARP but also released cytochrome c in mitochondria. In addition, ML diminished the protein expression levels of Bcl-2 family, including Bcl-2 and Bcl-xL [24].

3.2. Inhibit Proliferation and Promote Apoptosis

Apoptosis is a conserved form of programmed cell death required for homeostasis, whereas in noncontrolled situations could be a major cause for tumorigenesis [25]. Zeylenone is a naturally occurring cyclohexene oxide extracted from the leaves of Uvaria grandiflora Roxb. of the family Annonaceae [26,27]. In SKOV3 cells incubated with zeylenone at different concentrations (2.5, 5, and 10 µmol/L for 24 h), Janus family of tyrosine kinase (p-JAK), and signal transducer with activator of transcription (p-STAT) expression levels were reduced variably. In a dose-dependent pattern, zeylenone increased the loss of MMP, apoptosis-inducing factor (AIF), and cytochrome c. Apart from that, it increased caspase-3, Fas, Fasl, and Bax in both the mRNA and protein expression levels but decreased Bcl-2 expression level [28].

Berbamine is a plant-derived component isolated from the traditional Chinese medicine Berberis amurensis, which is already applied for leukemia treatment in clinic. In ovarian cancer cells SKOV-3, berbamine upregulated the cleaved caspase-3, cleaved caspase-9, and Bax in protein expression levels, while it downregulated the antiapoptotic protein level of Bcl-2 family. Berbamine inhibited cell proliferation and promoted apoptosis by partially suppressing the Wnt/β-catenin signaling transduction [29]. Pinus massoniana bark proanthocyanidins (PMBP) is a bioactive compound derived from natural root bark of Pinus massoniana. In A-2780 and OV2008, PMBP downregulated the expression level of Bcl-2 family and increased the levels of Caspase-3/9. PMBP significantly repressed MMP-9, ERK1/2, and p38 MAPK expression, thus blocking the activity of NF-κB. Furthermore, it triggered activations of mitochondria-associated apoptosis through the loss of mitochondrial membrane potential [30]. Sanguin H-6 is a natural component derived from red raspberry fruits. In A2780 cells incubated with Sanguin H-6 at concentrations of 20/40 µM, the early apoptotic percentages differently exhibited at 35.39 and 41.76. Sanguin H-6 triggered the stimulation of MAPK p38 through a caspase-8-dependent BID cleavage signaling pathway. Furthermore, Sanguin H-6 promoted the antiproliferative effect by activations of caspasases. It is also observed a severe morphological change and more cleaved PARP, which is known as the biomarker of apoptosis [31].

3.3. Suppress Cell Migration and Invasion

The epithelial cells transdifferentiated into motile mesenchymal cells, which is known as epithelial-mesenchymal transition (EMT) [32]. EMT is a self-regulated process in phys-
iological conditions, whereas EMT might step to cancer in pathological situations [33]. Tetramethylpyrazine (TMP) is a widely used medicinal compound extracted from Ligusticum Wallichil Franch. In SKOV-3 and OVCAR-3 cells, TMP significantly increased the expression level of miR-211, which could be partially abrogated by the knockdown of miR-211 [34]. TMP inhibited the cell viability, migration, and invasion ability of both cell lines in a dose-dependent pattern. Treatment with TMP at various concentrations (25–100 µg/mL) in SKOV-3 for 24 h exhibited the downregulation of IL-8 accompanied with the suppression of cell migration [35].

Dihydroartemisinin (DHA) extracted from the traditional Chinese herb Artemisia annua is widely used for malaria treatment. Unusual activation of the Hedgehog (Hh) signaling transduction is generally associated with human ovarian cancer tumorigenesis. In ovarian cancer cells, DHA obviously inhibited cell malignant behaviors and also augmented cell apoptosis. By contrast, DHA had few effects in human ovarian surface epithelial cells (HOSEPICs) in vitro. Specifically, DHA suppressed the Hh signaling pathway by inhibiting its agonist purmorphamine; in addition, DHA enhanced the Hh signaling pathway inhibitor GANT61 [36]. Emodin is a natural anthraquinone contained in various Chinese medicine herbal preparation herbs, for instance, Rheum palmatum, Polygonum cuspidatum, and Polygonum multiflorum [37]. In A2780 and SKOV-3 cells, it has been proven that emodin specially suppressed the abilities including cell proliferation, invasion, and migration. In a dose-dependent pattern, emodin treatment led to upregulation of epithelial markers, such as E-cadherin and Claudin. It also resulted in downregulation of mesenchymal markers, such as N-cadherin, Vimentin, and transcription factor Slug. Moreover, emodin diminished ILK, p-GSK-3β, β-catenin, and Slug expression levels, producing similar effects of siRNA transfection. These results could be exemplified by the following emodin incubation. It revealed that emodin repressed the EMT of ovarian cancer cells through ILK/GSK-3β/Slug signaling transduction [38].

3.4. Reactive Oxygen Species (ROS) Damage

From inflammation to cancer, excessive oxidative stress is deemed as a regulatory factor in multiple diseases. Although partial signaling transduction still need to be explored, clear molecular mechanisms for diseases progression have been elucidated. The association from excessive reactive oxygen species (ROS) damage to tumorigenesis implied the importance of antioxidant therapy [39]. Sulforaphane (SFN) is a phytochemical component contained in cruciferous plants such as broccoli. In cell lines OVCAR3, OVCAR4, OVCAR5, and SKOV3, SFN significantly promoted apoptosis at various concentrations. On the other hand, xenograft experiments in vivo elucidated that SFN effectively suppressed tumor growth through targeting tumor-related signaling transduction, while SFN did not produce consequential levels of ROS in cells. Moreover, a thiol-oxidizing agent named diamide enhanced the effect present by SFN, indicating protein thiols oxidation and/or cellular redox status changes may be an underlying mechanism [40].

Quercetin is a prominent dietary flavonoid, which ubiquitously presents as an antioxidant in onion, milk thistle, and French maritime pine bark [41]. In SKOV3 and OVCAR3 cell lines, quercetin increased apoptosis through the stimulation of transcription factor CCAAT enhancer-binding protein homologous protein (CHOP). The expression level of death receptor DR5 was induced by CHOP, followed with ROS-mediated endoplasmic reticulum-stress damage. In the human SKOV-3 xenograft model, quercetin inhibited tumor growth mediated by TRAIL sensitization. It was synergistically accompanied by the induction of apoptosis and the activation of caspase-3 [42]. Formononetin (FMN) is an isoflavone contained in red clovers and soy, which exhibits antioxidant and anticancer capacities in various cell types. In ES2 and OV90 cell lines, FMN augmented the loss of mitochondrial membrane potential as well as generation ROS. Additionally, FMN suppressed cell proliferation and induced apoptosis via interfering with sub G0/G1 cell cycle phase arrest, accompanied by decreasing phosphorylation levels of ERK1/2, P90RSK, P70S6K, and AKT and increasing the phosphorylation level of P38 protein. In both cell types, inco-
bation of FMN simultaneous with pharmacological inhibitors LY294002/U0126 resulted in additional antiproliferative outcomes. [43].

3.5. Trigger Cell Cycle Arrest

Although ovarian cancer possesses its complexity and idiopathy, one contributing factor propelled the tumorigenesis is deregulated cell cycles [44]. To unravel the dynamics of cell cycle arrest, dysregulation presents a better understanding in human cancer cell lines. Cucurbitacin-A, isolated from *Momordica charantia* L. (M. charantia), has been identified with anticancer and anti-inflammatory activities [45]. In SKOV3 cell line, cucurbitacin-A exhibited an IC50 of 40 µM and triggered the G2/M checkpoint in cell cycle arrest. It caused DNA lesions and prompted mitochondrial membrane potential loss (MMP) through ROS-mediated alterations. Moreover, it considerably suppressed the expression levels of several proteins of PI3K/Akt/mTOR signaling transduction [46]. Asiatic acid is a natural triterpenoid component enriched in the tropical medicinal herb Centella asiatica. In SKOV3 and OVCAR-3 cell lines, incubation with Asiatic acid (10–100 µg/mL for 48/72 h) significantly downregulated the phosphorylation levels of PI3K, AKT, and mTOR. Both cell lines treated with Asiatic acid exhibited an increasing 7–10-fold apoptosis at G0/G1 phase cell cycle arrest. The colony formation of cells was remarkably reduced by 25–30% at a specific concentration of 10 µg/mL. Satisfyingly, the ovarian cancer cells viability was reduced by 50%, and little effect was observed on human ovarian epithelial cells [47].

Amentoflavone (AF) is one of the natural herbal bioflavonoids extracted from *Selaginella tamariscina* [48]. In SKOV3 and OVCAR-3 cell lines, AF downregulated the expression levels of S-phase kinase protein 2 (Skp2), in which progression was regulated through ROS/AMPK/mTOR signaling pathway. In a dose-dependent pattern, AF incubation for 48 h triggered G1/G0 phase cell cycle arrest. In a xenograft animal model, AF could also significantly induced apoptosis in vivo [49]. Proanthocyanidins (BLPs) is isolated from Chinese bayberry leaves (Myrica rubra Sieb. et Zucc.). In chemotherapy-resistant OVCAR-3 spheroid cells, BLPs could attenuate chemotherapy-resistant traits. BLPs showed repressive effects on cell growth and cancer stemness characteristics. BLPs inhibited the abilities of cell viability, sphere and colony formation, and expression of stemness-related proteins. Specifically, BLPs targeted the Wnt/β-catenin signaling pathway by decreasing the expression of β-catenin, cyclin D1, and c-Myc and induced G1 phase cell cycle arrest, further inhibiting the capacity of self-renewal [50]. The traditional Chinese herbs Scutellaria baicalensis (SB) exhibited significant activities in treating hepatitis and diarrhea. Fritillaria cirrosa (FC) is also broadly used in respiratory diseases for a long history. In ovarian cancer cells OVCA 420 and endometrial cancer cell OVCA 429, cotreatment with either herb interfered with G0/G1 phase cell cycle arrest. Both of SB- and FC-induced p27 expression, caused the activation of caspase-3, along with downregulation of cyclin D1 and cyclin D3. Furthermore, both herbs abrogated the activation of NF-κB and downregulated metastasis-promoting proteins [51].

3.6. Induce Autophagic Cell Death

Autophagy, unlike the apoptosis mentioned above, is relatively a conserved pathway. Materials in cytoplasm are transmitted to lysosomes for degradation by targeted enzymes. It is activated by quite a lot of stimuli, including hypoxia, protein aggregates, and oxidative stress in response to drugs and radiation [52]. Tanshinone I (Tan-I), an essential fat-soluble monomer component, is derived from Chinese folk herbs DanShen. In cell lines A2780 and ID-8, incubation with Tan-I (0, 1.2, 2.4, 4.8, and 9.6 µg/mL for 24 h) augmented the protein levels of apoptosis-associated protein Caspase-3 and autophagic-associated proteins, for instance Beclin1, ATG7, p62, and LC3II/LC3I. The PI3K/AKT/mTOR signaling transduction were also upregulated simultaneously. The autophagic vacuoles were observed, which demonstrated that Tan-I promoted apoptosis, activated the cleavage of caspase-3, and induced autophagy in human ovarian cancer [53]. Grifolin is one of the secondary metabolic byproducts contained in the polypore mushroom Albatrellus, which
had potential in treating nasopharyngeal carcinoma. In both cell lines A2780 and SKOV3, grifolin induced autophagic cell death and inhibited cell proliferation. Grifolin treatment also upregulated expression levels of autophagy markers, for instance LC3B, Atg7, and Beclin-1. It downregulated the expression level of P62 and suppressed the proteins of p-Akt, p-mTOR, p-p70S6K, and p-4E-BP1. In addition, grifolin regulated the AKT/mTOR/S6K signaling transduction in autophagy of human ovarian cancer [54].

Genistein is a phenolic compound abundant in soy-based products, which possess the characteristics of antitumor in various cancer types [55]. In cisplatin sensitive and resistant (A2780/CaOV3, ES-2) ovarian cancer cells, different concentrations of genistein treatment (0.1–10 µM for 24–96 h) led to both cell apoptosis and autophagy. Genistein caused caspase-independent cell autophagy in a dose-dependent manner. Genistein also regulated substrate for glucose uptake, oxidative phosphorylation, and fatty acid synthesis. In addition, genistein downregulated the expression level of phosphorylated Akt, which involved in mechanisms of limited glucose utilization [56]. Isoliquiritigenin (ISL) is a flavonoid component enriched in the licorice plants. In OVCAR-5 and ES-2 cell lines, ISL significantly suppressed the cell viability in a both concentration- and time-dependent pattern. ISL promoted the expression levels of apoptosis and autophagy proteins, including cleaved PARP, cleaved caspase-3, Bax/Bcl-2 ratio, LC3B-II, and Beclin-1. Moreover, ISL triggered the G2/M phase cell cycle arrest [57].

3.7. Inhibit Angiogenesis

Chinese bayberry leaves proanthocyanidin (BLP) is a major effective compound isolated from Chinese bayberry. In ovarian cell line IOSE-364, BLP exhibited inhibitory effects of angiogenesis. It triggered G1 phase cell cycle arrest by repressing the protein levels of c-Myc, cyclin D1, and CDK4. In cisplatin resistant A2780 and CP70 cells, BLP especially attenuated the ability of wound healing. Furthermore, BLP reduced the expression level of HIF-1α. It decreased the level of angiogenesis-associated biomarker vascular endothelial growth factor (VEGF). BLP also suppressed ROS generation and regulated Akt/mTOR signaling transduction in human ovarian cancer [58]. Six natural bioactive flavonoids, including apigenin, taxifolin, luteolin, quercetin, genistein, and kaempferol, are all water-soluble vitamins. In OVCAR-3 cell line, six flavonoids suppressed the expression of VEGF in a dose-dependent manner. In addition, six plant-derived flavonoids inhibited the ovarian cancer cell proliferation [59].

Baicalin and its aglycone baicalein are specially enriched in traditional Chinese herbs Scutellaria [60]. In OVCAR-3 and CP-70 cell lines, either baicalin or baicalein reduced cell viability with LD_{50} at a median of 50 and 32.5 µM. In normal ovarian cell line IOSE-364, both natural components exerted less inhibitory effects. Baicalin and baicalein suppressed cancer promoting genes such as VEGF, HIF-1α, c-Myc, and NF-kB at different concentrations, thus inhibiting the cell viability. Specifically, baicalein achieved the same effect at a lower concentration in both ovarian cancer cell lines [61]. Harmine (HM) is a small-molecular β-carbolinujhe alkaloid present in medicinal plants, which has been applied in folk medicine for antitumor therapy. In SKOV-3 cell line, HM remarkably downregulated the cell proliferation in a dose-dependent pattern. It also suppressed the cell proliferation and migration regulated by epidermal growth factor (EGF). Furthermore, HM inhibited the concentrations of ERK1/2 and CREB in both the basal level and EGF-induced phosphorylation level. HM significantly downregulated the expression level of VEGF and MMP-2/9 [62]. Cranberry proanthocyanidin-1(PAC-1) is abundant in cranberry, which targets ovarian cancer viability due to its cytotoxic and angiogenesis properties. In chemotherapy-resistant SKOV-3 cell lines, PAC-1 blocked G2/M phase cell cycle progression and increased ROS generation in a dose-dependent pattern. It induced apoptosis in both intrinsic and extrinsic pathway. In human umbilical vein endothelial cell line (HUVEC), PAC-1 suppressed the cell viability significantly. Moreover, PAC-1 interfered with the phosphorylation level of VEGF-stimulated receptor, thus inhibiting the formation of endothelial tube [63].
3.8. Interfere with RNA Expression

The discovery of aberrant RNAs play important oncogenic roles in various human cancers. These RNAs and their connections with gene expressions may provide the new sight to the detection, diagnosis, and targeted therapeutics for human ovarian cancer. Astragalus polysaccharide (APS) is a bioactive component isolated from a major traditional herb Astragalus membranaceus, which is named HuangQi in Chinese folk medicine. In OV-90 and SKOV-3 cell lines, APS repressed the cell proliferation and induced apoptosis. F-box and WD-40 domain protein 7 (FBXW7) is a star tumor inhibitor, which translation process is targeted and suppressed by miR-27a directly. In both cell lines, miR-27a upregulation obviously reversed the inhibitory effects exhibited by APS, such as antiproliferation and proapoptotic. It produced the same as the effects of knockdown of FBXW7 by si-FBXW7 [64]. Tanshinone IIA (Tan-IIA) is a natural plant-derived component enriched in Chinese herbal medicine DanShen. In the A2780 xenograft model, Tan-IIA suppressed tumor growth significantly. In ID-8 and A2780 cell lines, Tan-IIA induced apoptosis and promoted antiangiogenesis in a dose-dependent pattern. Tan-IIA also triggered G2/M phase cell cycle arrest. It downregulated expression level of Bcl-2 and upregulated level of Bax. Tan-IIA also decreased VEGF and cyclooxygenase-2 (COX2) mRNA expressions to promote antiangiogenesis [65].

3.9. Promote DNA Damage Response

Platinum-based drugs, radiation, and many other chemotherapeutic agents could induce DNA damages directly or indirectly. The ability of ovarian cancer cells to repair DNA lesions partly determined the efficiency of chemotherapeutic drugs [66]. Sideroxylin is a natural bioactive flavone contained in Callistemon lanceolatus leaves, which exhibits anti-inflammatory activity against the pathogen S. aureus. In ES2 and OV90 cell lines, sideroxylin significantly suppressed cell proliferation and induced apoptosis. Sideroxylin at different concentrations stimulated the phosphorylation levels of proteins including ERK1/2, JNK, P38, and MAPK. Furthermore, it caused DNA fragmentation and depolarization of mitochondrial membrane. It increased more lipid peroxidation and the generation of ROS through PI3K/MAPK signal transduction [67]. Epigallocatechin gallate (EGCG) is one of the major components in green tea. Sulforaphane (SFN) is extracted from cruciferous vegetables. In paclitaxel-sensitive and -resistant ovarian cancer cell lines (SKOV3TR-ip1/SKOV3-ip2), SFN suppressed cell viability in a time- and dose-dependent pattern. Furthermore, EGCG potentiates the outcomes triggered by SFN. The cotreatment resulted in both G2/M along with S phase cell cycle arrest and decreased Bcl-2 protein levels. Treatment combined with both components in SKOV3TR-cells increased apoptosis and reduced hTERT concentration, which acted as major regulatory subunit in telomerase. Therefore, EGCG and SFN treatment promoted DNA damage response especially in paclitaxel-resistant ovarian cancer cell lines [68]. Berberine is an alkaloid component extracted from various herbal plants, which was proved to inhibit cell proliferation. In three cell lines A2780, HEY, and HO8910, berberine triggered DNA damages and repressed homologous recombination repair (HRR). PARP1 is a protein that senses the oxidative DNA-strand damages and coordinates their restorations. Correspondingly, berberine hyperactivated PARP1 in the A2780 as well as HO8910 cell lines. In coincubation of berberine and PARP inhibitor niraparib, tumor progression was remarkably diminished in vivo. Thus, berberine sensitized ovarian cancer cells to PARP inhibition by causing oxidative DNA damages and repressing HRR [69]. The detail effects and various cellular signaling pathways were summarized in Table 1 and Figure 1.
Table 1. The antitumor effects of natural plants components in ovarian cancer.

| Compound                        | Classification       | Model                  | Effect                                                                 | Mechanism                                                                                     | Reference |
|---------------------------------|----------------------|------------------------|-----------------------------------------------------------------------|-----------------------------------------------------------------------------------------------|-----------|
| Tanshinones                     | Terpenoid/Abietane   | A-549, TOV-21G         | Cytotoxic action, ROS/JNK-CHOP, PI3K/AKT/mTOR signaling pathway       | Reduce cell viability, inhibit colony formation capacity                                      | [21,22,53]|
| Kadsuphilactone B               | Terpenoid/Triterpene | A2780, Ishikawa cells  | Cytotoxic action                                                      | Stimulate the activity of caspase-3/9 and MAPKs                                               | [23]      |
| Methyl lucidone (ML)            | Oxo Steroid          | OVCAR-8, SKOV-3        | Cytotoxic action                                                      | Induce cellular apoptosis, stimulate the cleavage of caspase-3/9                               | [24]      |
| Zeylenone (Zey)                 | Alicyclic            | SKOV-3                 | JAK2/STAT3 signaling pathway                                         | Reduce p-JAK and p-STAT expression level                                                       | [28]      |
| Berbamine (BBM)                 | Alkaloid/Benzylisoquinoline | SKOV-3          | Wnt/β-catenin signaling pathway                                      | Increase activity of caspase-3/9, Bax and decrease Bcl-2 levels                              | [29]      |
| Pinus massoniana bark proanthocyanidins (PMBP) | Phenol/Tannin | A-2780                 | NF-κB signaling pathway                                              | Activate mitochondria-associated apoptosis                                                    | [30]      |
| Sanguin H-6 (SH-6)              | Phenol/Tannin        | A-2780                 | MAPK signaling pathway                                               | Increase p15/BID level, activate MAPKs (p38)                                                    | [31]      |
| Tetramethylpyrazine (TMP)       | Alkaloid/Pyrazine    | SKOV-3, OVCAR-3, A-2780 xenograft model | Suppress cell migration and invasion, reverse multiple-drug resistance (MDR) | Increase miR-211, diminish expression of proliferation and angiogenesis markers             | [34,35,70]|
| Dihydroartemisinin (DHA)        | Terpenoid/Artemisinin | HOSEPICs              | Hedgehog signaling pathway                                           | Diminish the levels of ILK, p-GSK-3β, β-catenin, and Slug, repress EMT                        | [36]      |
| Emodin                          | Quinone/Anthraquinones | A-2780, SKOV-3        | ILK/GSK-3β/Slug signaling pathway                                    | Inhibit cell malignant behaviors and augment apoptosis                                         | [38]      |
| Sulforaphane (SFN)              | Ester/Isothiocyanate | Xenograft model, SKOV-3-ip1, SKOV3TR-ip2 | Promote DNA and ROS damage                                            | Upregulate CHOP-induced DR5 expression following ROS damage                                    | [40]      |
| Quercetin                       | Flavonoid/Flavonol   | SKOV-3, xenograft model | ROS-JNK-CHOP signaling pathway                                       | Upregulate caspase 3/9 and bax/bcl-2, interfere with sub G0/G1 phase arrest, increase P38 phosphorylation DNA damage, ROS damage, MMP alterations, G2/M checkpoint | [42]      |
| Formononetin (FMN)              | Flavonoid/Isoflavone | ES2, OV-90             | ERK1/2 signaling pathway                                             | Downregulate phosphorylation levels of PI3K, Akt, and mTOR                                   | [43]      |
| Cucurbitacin-A                  | Terpenoid            | SKOV-3                 | Trigger cell cycle arrest                                            | Downregulate the expression of β-catenin, induce G1 cell cycle arrest, downregulate HIF-1α and VEGF  | [46]      |
| Asiatic acid                    | Terpenoid/Triterpene | SKOV-3, OVCAR-3        | PI3K/Akt/mTOR signaling pathway                                      | Activation of caspase-3 along with downregulation of cyclins D1 and D3                      | [47]      |
| Amentoflavone (AF)              | Flavonoid            | Xenograft model        | ROS/AMPK/mTOR signaling pathway                                      | Upregulate autophagy markers LC3B, Atg7, Beclin-1, downregulate P62                           | [49]      |
| Proanthocyanidins (BLPs)        | Phenol/Tannin        | A-2780, CP70, OVCAR-3 SP stem cell | Wnt/β-catenin, AKT/mTOR/p70S6K/4E-BP-1 signaling pathway             | Reduce the expression of β-catenin, induce G1 cell cycle arrest, downregulate HIF-1α and VEGF  | [50,58]   |
| Scutellaria baicalensis (SB) and Fritillaria cirrhosa (FC) | Polysaccharides, Phenols, etc. | OVCA-420, OVCA-429 | NF-κB signaling pathway                                              | Upregulate autophagy markers LC3B, Atg7, Beclin-1, downregulate P62                           | [51]      |
| Grifolin                        | Terpenoid            | A-2780, SKOV-3         | AKT/mTOR/S6K signaling pathway                                       |                                                                                               | [54]      |
Table 1. Cont.

| Compound          | Classification | Model                          | Effect          | Mechanism                                                                 | Reference |
|-------------------|----------------|--------------------------------|-----------------|---------------------------------------------------------------------------|-----------|
| Genistein         | Terpenoid/sesquiterpenol | OVCAR-3, SKOV-3 | Inhibit VEGF levels | Reduce CD3 levels and restore NKT-cell functions, Increase cleaved PARP, caspase-3 and Bax/Bcl-2 ratio; trigger G2/M cell cycle phase | [56,71]   |
| Isoliquiritigenin (ISL) | Ketone | OVCAR5, ES-2 | Suppress cell viability | Downregulate VEGF in dose-dependent manner; Suppress cancer promoting genes including VEGF, HIF-1α, cMy-c, and NF-kB | [57]      |
| Flavonoids        | Flavonoids     | OVCAR-3                        | Inhibit angiogenesis | Increase cleaved PARP, caspase-3 and Bax/Bcl-2 ratio; trigger G2/M cell cycle phase | [58]      |
| Baicalin and baicalein | Flavonoid     | OVCAR-3, CP-70 | Inhibit angiogenesis | Increase cleaved PARP, caspase-3 and Bax/Bcl-2 ratio; trigger G2/M cell cycle phase | [59]      |
| Harmine (HM)      | Alkaloid       | SKOV-3                         | Inhibit cell proliferation | Increase cleaved PARP, caspase-3 and Bax/Bcl-2 ratio; trigger G2/M cell cycle phase | [60]      |
| Cranberry proanthocyanidin-1 (PAC-1) | Phenol/Tannin | Chemotherapy-resistant SKOV-3, human umbilical vein endothelial cells (HUVEC) | Increase ROS generation, induce apoptosis | Increase cleaved PARP, caspase-3 and Bax/Bcl-2 ratio; trigger G2/M cell cycle phase | [61]      |
| Astragalus polysaccharide (APS) | Polysaccharide | OV-90, SKOV-3 | Inhibit proliferation and promote apoptosis | Increase cleaved PARP, caspase-3 and Bax/Bcl-2 ratio; trigger G2/M cell cycle phase | [62]      |
| Sideroxylin       | Flavonoid      | ES2, OV-90                     | ERK1/2 signaling pathway | Increase cleaved PARP, caspase-3 and Bax/Bcl-2 ratio; trigger G2/M cell cycle phase | [63]      |
| Epigallocatechin gallate (EGCG) | Flavonoid | SKOV3-ip1, SKOV3TR-ip2 | Promote DNA damage response | Increase cleaved PARP, caspase-3 and Bax/Bcl-2 ratio; trigger G2/M cell cycle phase | [64]      |
| Berberine         | Alkaloid       | A2780, HEY, HO8910             | Promote DNA damage response | Increase cleaved PARP, caspase-3 and Bax/Bcl-2 ratio; trigger G2/M cell cycle phase | [65]      |
| Carotenoids       | Terpenoid/Tetraterpenoid | Epidemiological statistics | Inhibit VEGF levels | Increase cleaved PARP, caspase-3 and Bax/Bcl-2 ratio; trigger G2/M cell cycle phase | [66]      |
| Lycopene          | Terpenoid      | OV-MZ-6                        | Diminish tumor load | Increase cleaved PARP, caspase-3 and Bax/Bcl-2 ratio; trigger G2/M cell cycle phase | [67]      |
| Deoxyschizandrin  | Phenol/Tannin  | A-2780                        | Increase ROS production | Increase cleaved PARP, caspase-3 and Bax/Bcl-2 ratio; trigger G2/M cell cycle phase | [68]      |
| Curcumin          | Phenol         | SKOV3-ip1, HeyA8               | Reduce mean tumor growth | Increase cleaved PARP, caspase-3 and Bax/Bcl-2 ratio; trigger G2/M cell cycle phase | [69]      |
| Procyanidin (GSP) | Phenol/Tannin  | A-2780, SKOV-3                 | NF-κB and MAPK/ERK pathway | Increase cleaved PARP, caspase-3 and Bax/Bcl-2 ratio; trigger G2/M cell cycle phase | [70]      |
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Epigallocatechin gallate (EGCG) Flavonoid SKOV3-ip1, SKOV3TR-ip2 Promote DNA damage response Suppress cell viability in time- and dose-dependent manner, reduce the expression of hTERT and Bcl-2 [68]

Berberine Alkaloid A2780, HEY, HO8910 Promote DNA damage response Trigger oxidative DNA damages [69]

Carotenoids Terpenoid/Tetra terpenoid Epidemiological statistics Inhibit VEGF levels Stimulate the activities of lymphocytes, macrophages, and cytotoxic T-cells [72]

Lycopene Terpenoid OV-MZ-6 Diminish tumor load Decrease expression of MMP9, ILK and EMT biomarkers [73]

Deoxyschizandrin Phenol/Tannin A-2780 Increase ROS production Inhibit macrophages, M2 phenotype markers CD163 and CD209 [74]

Curcumin Phenol SKOV3-ip1, HeyA8 Reduce mean tumor growth Decrease microvessel density, increase tumor cell apoptosis [75]

Procyanidin (GSP) Phenol/Tannin A-2780, SKOV-3 NF-κB and MAPK/ERK pathway Augment cytotoxicity of paclitaxel and adriamycin [76]

Figure 1. The various cellular signaling pathways of natural plants components in ovarian cancer.

4. Mechanisms of Botanical Components Improve Drug Sensitivity

4.1. Modulate Immune Cell Responses

Considerable progress in elucidating how cancers escape from destructive immunity has been made, yet measures to solve tumor evade have not kept up [77]. As previous studies have illustrated, the activations of natural killer T (NKT) cells are inhibited immune responses and antitumor activities. As mentioned above, genistein induced ovarian cancer cells in autophagic. In OVCAR-3 and SKOV-3 cell lines, treatment with bevacizumab and genistein caused downregulation in ganglioside GD3 levels and repair of NKT-cell functions. Coincubation with bevacizumab and genistein also suppressed the VEGF, which was associated with ovarian cancer cells [71]. Natural herbal carotenoids, such as β-carotene, lutein, zeaxanthin, and curcumin, contained in retinoids-rich vegetables and fruits. In both a time- and dose-dependent pattern, carotenoids stimulated the activities of immune cells including lymphocytes, macrophages, and cytotoxic T-cells. Epidemiological statistics proved that supplementation with high carotenoid intake concomitants with a decreased risk of ovarian cancers [72].

4.2. Regulate Immune Molecular Expression Level

It has been shown that immune cells would alter functional phenotypes when cancer cells signal diversely. The immune molecules are a key factor in the tumor microenvironment. Lycopene, a bioactive natural compound in tomato, possessed the highest antioxidant properties among carotenoids [78]. In both ovarian cancer-bearing mice and established ovarian cancer cell models, lycopene effectively diminished the tumor load. In OV-MZ-6 cell lines, lycopene decreased expression levels of ITGA5, ITGB1, MMP9, FAK, ILK, and EMT biomarkers. It downregulated protein expressions of integrin α5 and activations of MAPK. Moreover, lycopene enhanced antitumorigenic concomitantly with antimetastatic effects of paclitaxel and carboplatin [73]. Deoxyschizandrin is an active lignin ingredient abundant in Schisandra Chinensis (Turcz.) Baill and Schisandra berries have been proven to possess antioxidant effect in previous studies [79]. In A-2780 cell lines, deoxyschizandrin arrested the G0/G1 phase cell cycle and interfered with the cyclin E

Figure 1. The various cellular signaling pathways of natural plants components in ovarian cancer.
expression level. It increased the production of ROS and decreased the activation of Akt. It also inhibited macrophages, which were activated by the ovarian cancer cells. Moreover, deoxyschizandrin inhibited the expression level of the M2 phenotype markers CD163 and CD209 in tumor-associated macrophages (TAMs). The tumor-promoting factors MMP-9, RANTES, and VEGF were significantly enhanced in TAMs, while the expression and production levels of these immune molecules were all suppressed by deoxyschizandrin effectively [74].

4.3. Reverse Multiple-Drug Resistance (MDR)

Drug resistance is a troublesome phenomenon when proceeding the treatment of classical platinum drugs in ovarian cancer. Accordingly, there are two mechanisms of chemoresistance categories classified by most studies, including de novo (intrinsic) and acquired (extrinsic). However, the detailed reasons have not been illustrated comprehensively [80]. Curcumin is often named ‘curry powder’ in daily life, which is popularly in Indian spice for cooking. It is mainly derived from plant curcuma longa and well known as turmeric in constituent folk medicine [81]. In two in vivo models of SKOV3ip1 and HeyA8, curcumin led to a 49% and 55% reduction in average tumor growth compared with controls. When coincubated along with docetaxel, the number ranked at 96% and 77% significantly. In multidrug-resistant cell line HeyA8-MDR, curcumin alone and in combination with docetaxel exhibited 47% and 58% average reductions in tumor growth. In addition, both of the treatments reduced cell proliferation, microvessel density, and increased tumor cell apoptosis [75]. Tetramethylpyrazine (TMP), enriched in natural herb Ligusticum hort, was identified to act as a multilink role in the anticancer process. In A-2780 and SKOV-3 cell lines, the antitumor effects of paclitaxel were augmented by TMP. Treatment of TMP and paclitaxel inhibited signal transduction of ERK1/2 and Akt, promoted cancer cell apoptosis, and suppressed angiogenesis. In A-2780 xenograft mouse models, TMP potentiated the decrease in the tumor burden induced by paclitaxel. Moreover, TMP partially decreased the drug toxicity of paclitaxel [70]. Procyanidin (GSP) is a natural effective polyphenol compound abundant in grape seed [79]. In both A-2780 and paclitaxel-resistant A-2780/T cell lines, GSP significantly augmented the cytotoxicity of paclitaxel and adriamycin. P-glycoprotein (P-gp) is a protein related to the phenotype of MDR. GSP reversed ovarian cancer MDR by suppressing the transcription and expression of P-gp. Moreover, it downregulated the activation of NF-κB and MAPK/ERK pathway [76]. The molecular mechanisms are shown in Figure 2.

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**Figure 2.** The role of molecular mechanisms in ovarian cancer.
5. Antitumor Effects and Chemical Structure Relationship in Natural Plants Compounds

In treating ovarian cancer, although some natural bioactive components have already been applied clinically, the widespread adoption of them is not quite optimistic for several parameters. It is dragged down by weak solubility, uncontrolled stability, and poor bioavailability. The nonspecific distribution, short retention time, and rapid rate of elimination are also limiting factors [82]. Accordingly, investigating the structure generalities and specialties of effective components might be the breakthrough in ovarian cancer treatment. Based on the preliminary analysis of natural bioactive compounds in antiovarian cancer, we summarized several potential target structures including flavonoids, phenols, alkaloids, terpenoids, quinones, and esters. Kadsurilactone B, cucurbitacin-A, grifolin, and lycopene are classified as terpenoids, which are familiar in daily diet tastes. Terpenoids possess the same precursor as phytocannabinoids, which is a star agent for cancer pain alleviation [83]. The synergistic relationships of the terpenoids and other drugs used in ovarian cancer indicated that some natural bioactive components might also retain unknown effective properties in human body, for the shared structure determine analogous functions. The chemical structures of effective compounds hinted that most of them contain the exact or potential polyphenol structure and alkaloids containing nitrogen. The structural formula of formononetin, genistein, emodin, proanthocyanidins, and other flavonoids carry with arrangements of polyphenol structure in meta-, ortho-, and para-position. In addition, curcumin contains a benzene ring connected with methylol groups, which probably lead to hydrolytic reactions and phenolic hydroxyl formations. Owing to the structural commonality of natural phenolic components, tannin, quinones, and flavonoids might play similar roles in tumor prevention and carcinogen metabolism regulation [84]. Quercetin, sideroxylin, formononetin, amentoflavone, and epigallocatechin gallate (EGCG) are included in flavonoids, which shared a basic structure with vitamin E. Sharing a three-ring structure, flavonoids can be classified into subgroups based on various substitutions in different positions. For instance, quercetin and sideroxylin are flavonols, formononetin belongs to isoflavonoids, amentoflavone is from polyflavonoids, and EGCG is included in proanthocyanidins. Each subgroup possesses a broad spectrum of in vitro activities including antioxidant, antitumor proliferative, and the characteristics to regulate enzymatic signaling pathways [85]. Moreover, the B ring of the flavonols owns a unique hydroxylation pattern, which reinforces the inhibition of mast cell secretion [86]. These findings implied that substitutions of the main structure should also be highlighted because they could offer distinct pharmacological functions and complementary roles. The substitutions in certain positions or the linkage among different parts could change the particular properties. The biomedical potential of natural compounds synchronously fluctuated with the reaction systems, because the human internal environment is extremely sophisticated [87]. Apart from that, the very existence of isomers should also be taken into consideration in natural phytochemicals [88,89].

Compared with multiple cellular studies demonstrating how natural plants compounds are applied in ovarian cancer, only a few mature animal models, such as MC38 murine models and xenograft models, have been explored in the present studies [90,91]. Therefore, the lack of large clinical approved trials is understandable. In these circumstances, deciphering the structure–activity relationship (SAR) of natural components may contribute to understanding the detailed pharmacology and pharmacocchemistry, further identifying and mapping the cellular mechanisms. This could be the pointcuts of enhancing the bioavailability in human bodies, which can be applied to develop targeted therapeutic drugs and reduce side effects. Still, whether and how the antitumor potential is related to chemical structures remain to be explored, as well as how these structures may yet fulfill their unique attributes. In pharmacokinetics and pharmacological properties, a promising solution is to conjugate effective constituents together with carriers. These unique carriers are designed to target tumor tissue or immune cells specifically, such as silver nanoparticles, liposomes, polymers, micelles, polysaccharides, etc. [92]. Take polymer-based vesicular delivery platforms for example: they are macromolecular in order
to recognize the physiology traits of tumors and raise up intratumoral bioavailability [93]. As for polysaccharides carriers, β-d-glucans have served as immune-adjuvants due to their modulatory properties in macrophages [94]. In clinical application, several natural herbs have been approved to treat ovarian cancer. The seed of brucea javanica oil has already been used in vivo, and its oil microemulsion and loaded liposomes forms are explored as improved formulations [95]. The encapsulated nanoparticles of curcumin enhanced the properties of drug release in the delivery system [96]. Nanocarrier quercetin validated its potential in various capsule forms, namely, liposomes, antibody-conjugated micelles, and silver nanoparticles [97]. The milk exosomes augmented the concentration of berry Anthos in ovarian cancer patients [98]. There is a brand new co-delivery system that introduced both cisplatin and other drugs, which synergistically reversed cisplatin resistance and reduced side effects [99]. Taken together, these up-to-date strategies offered insight into the wild application of phytochemicals synergistic treatment in ovarian cancer.

6. Conclusions and Perspectives

Although first-line conventional chemotherapy is widely accepted to exert positive initial effects in many patients with ovarian cancer, drug resistance often hampers the following effective treatments. Natural botanical components have been exploited to apply for the prevention and adjuvant chemotherapy of ovarian cancer, which can specially improve the life quality of patients. As previous studies have demonstrated, numerous natural components are less toxic to healthy adjacent tissue and could mitigate tumor progression. The novel combination therapy strategy with natural drugs enhances the efficacy of chemotherapy and reduces toxic and side effects, thus prolonging the survival period of postoperative patients. The present review summarizes the antitumor mechanisms of plant-derived active ingredients in ovarian cancer, primarily their roles in the induction of apoptosis and autophagy, stimulation of ROS damage, inhibition of angiogenesis and metastasis, and reversing the MDR phenotype. In these circumstances, based on the natural new approaches in the form of nanostructures, chemical adjustment and synthetic production still merit further investigations [100]. Afterall, the underlying mechanisms of natural plant components for ovarian cancer are relatively complicated. Previous studies have paid attention to several classic signaling pathways, and only a few in vitro models have been explored. To comprehensively and systematically profile each accurate component, deeper research including pharmacology is warranted. However, there are still major drawbacks in current domestic and international research: (i) limited bioavailability and poor stability in vivo; (ii) the current mechanisms focused on certain parts of signaling pathways and lacked comprehensive molecular mechanisms; (iii) experimental animal model studies are not broadly applied; and (iv) lack of large, randomized, placebo-controlled clinical trials. Collectively, a breakthrough in these studies would significantly contribute to a better understanding of ovarian cancer pathologies, providing a theoretical basis for in-depth research and a reference for neoadjuvant chemotherapy in clinical application [101].

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