Fullerol Nanocatalysis and Trimodal Surface Plasmon Resonance for the Determination of Isocarbophos

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Fullerol (C₆₀OH) has been shown to catalyze the trisodium citrate (TSC)–silver nitrate reaction to generate Ag nanoparticles (AgNPs). These AgNPs exhibit significant nanoplasmic surface-enhanced Raman scattering (SERS), resonance Rayleigh scattering (RRS), and absorption (Abs). When an aptamer (Apt) adsorbs on the C₆₀OH surface, catalysis is inhibited, and the intensities of SERS, RRS, and Abs decrease. In the presence of isocarbophos (IPS), Apt forms a stable complex (Apt-IPS) and releases C₆₀OH. As a result, SERS, RRS, and Abs intensities increase with increasing IPS concentration. Accordingly, a new SERS, RRS, and Abs trimodal method using Apt-labeled fullerol was established for the determination of IPS. Of the three spectral methods, SERS was the most sensitive, while the Abs method was the most cost-effective.

Keywords: isocarbophos, aptamer, fullerol nanocatalysis, SERS, RRS, Abs

INTRODUCTION

Surface plasmon resonance (SPR) is an optical phenomenon caused by the oscillation of free electrons in a metal surface layer produced by incident light (Jackman et al., 2017). Because of their exponentially larger surface areas, metal nanoparticles exhibit enhanced SPR and produce a more sensitive response. With the development of nanomaterial fabrication technologies, SPR has been used increasingly for the analysis of metal nanomaterials (Ye et al., 2016; Ouyang et al., 2017). Surface-enhanced Raman spectroscopy (SERS), in which the SPR effect is amplified by substances adsorbed on the nanoparticle surface, is an increasingly popular direct application of nanoscale plasma detection (Alvarez-Puebla and Liz-Marzan, 2012). Nanomaterials, especially those comprising noble metals and carbon nanomaterials, have novel spectral, electric, magnetic, thermal, and chemical properties (Gao et al., 2007; Kotov, 2010; Wei and Wang, 2013). Because noble metal nanomaterials, such as AuNPs and AgNPs, possess both catalytic activity and SERS activity, they have drawn attention (Jiang et al., 2008, 2010a,b; Liang et al., 2011, 2015; Yao et al., 2013). Carbon-based nanomaterials have abundant conjugated π bonds (C=C), which are characterized by high electron density, delocalization, and electron transfer ability (Krätschmer et al., 1990; Zhang et al., 2011; Zhao et al., 2016; Zhou et al., 2016; Justino et al., 2017) and have potential as green catalysts. Fullerene is of particular interest as a promising carbon nanomaterial, and it has been widely used in solar energy conversion materials and catalysis (Zhao et al., 2016; Cai et al., 2017) since it was successfully prepared. C₆₀ is stable and possesses good electron-transfer ability, due to the highly delocalized conjugated system consisting of 30 C=C bonds (Starodubtseva et al., 2008; Zhang et al., 2016). However, C₆₀ is a hydrophobic nanomaterial; it has very low
aqueous solubility and easily forms aggregates in water, which restricts its applicability (Jafvert and Kulkarni, 2008). Modification (such as carboxylation and hydroxylation) of the C$_{60}$ surface enhances its water solubility and expands the range of possible applications (Mohan et al., 1998; Niu et al., 2011; Li et al., 2013; Lu et al., 2013; Hang et al., 2014; Lanzellotto et al., 2014; Cao et al., 2016; Xu et al., 2016; Najafi, 2017). Lanzellotto et al. (2014) constructed a Trametes versicolor laccase biosensor on Au-AuNC$_{60}$OH. Fullerol enhanced the electron transfer between the active site of the enzyme and the electrode surface, leading to improved electrochemical biosensor performance. Tea polyphenols in beer were detected in the range of 0.03–0.30 mmol/L, with a limit of detection of 6 µmol/L.

Isocarbophos (IPS) is a fast-acting insecticide and acaricide that can cause poisoning by the esophagus, skin, and respiratory tract; all these have acute toxicity and cause cancer (Yamashita et al., 1997). The widespread uses of organic phosphorous insecticides indicate the extensive availability and potential for accidental and intentional human exposure (El-Behissy et al., 2001). Therefore, a rapid and accurate analytical method for the estimation of IPS is required. The main methods for IPS detection include chromatography (Huang et al., 2002; Yao et al., 2015; Li et al., 2017), chemiluminescence (Chen et al., 2012), and electrochemical methods (Yan et al., 2012). In recent years, new methods, such as highly selective aptamers and catalytic techniques, have been used to detect IPS (Pang et al., 2014; Zhang et al., 2014; Chen et al., 2017). In this paper, we propose nanocatalytic SPR spectroscopy for IPS detection, combining the favorable electron-transfer capabilities and catalytic behavior of fullerol to catalyze the sodium citrate–silver nitrate reaction and generate SPR on silver nanoparticles. The inhibitory effect on the catalytic reaction of the aptamer has been studied. Isocarbophos was selected as the target for a nanocatalytic SPR spectroscopy method. To the best of our knowledge, this is the first report describing the use of SPR (SERS, RRS, and Abs) combined with aptamer-labeled C$_{60}$OH and AgNP (generated from the trisodium citrate–silver nitrate catalytic reaction) for the detection of IPS.

**MATERIALS AND METHODS**

**Apparatus**

The following instrumentation was used: a DXR SmartRaman spectrometer (Thermo Company, USA) with a 633-nm laser at 3 mW power, a Cary Eclipse fluorescence spectrophotometer (Varian Company, USA), a TU-1901 double-beam UV-Visible spectrophotometer (Beijing General Instrument Co., LTD., China), and a FEI Quanta 200 FEG field-emission scanning electron microscope (Field Electron and Ion Company, Holland).

**Reagents**

Aptamer (Apt) sequence of 5′-3′ AGC TTG CTF CAG CGA TTC TTG ATC GGC ACA GAG CT [Sangon Biotech (Shanghai) Co., Ltd., China], 0.01 mol/L silver nitrate (Sinopharm Chemical Reagent Co. Ltd., China), 0.2 g/L fullerene(C60), 0.04 g/L fullerol(C$_{60}$OH), 0.1 g/L graphene oxide(GO) (Nanjing XFNANO Materials Tech Co., Ltd, China), 0.1 mol/L trisodium citrate (TSC) (Xilong Scientific Co., Ltd., China), 10.3 mol/L Victoria Blue B (VBB) solution, Victoria 4R (VB4R) solution, rhodamine 5 (Rh5), rhodamine 6G (Rh6G, Sinopharm Chemical Reagent Co., Ltd., China), isocarbophos (Beijing Century OuKe Biological Technology Co., Ltd., China), profenofos (Sinopharm Chemical Reagent Co., Ltd., China), and glyphosate (I&K Scientific Ltd., China) were prepared. All reagents were analytically pure, and water was double-distilled.

To prepare fullerene (Andrievsky et al., 1995), 0.02 g fullerene was dissolved in 20 mL methylbenzene to give a bright purple solution. Double-distilled water (100 mL) was added, and the solution was sonicated until the toluene was completely volatilized. The solution changed to a dark-yellow suspension, and 0.2 g/L of fullerene sol was obtained.

Hydroxylated fullerene was prepared, referring to Li et al. (1998): 1 mL 0.2 g/L fullerene sol, 10 µL 30% H$_2$O$_2$ solution, and 100 µL 1 mol/L NaOH were mixed and reacted at room temperature. Then, 98 µL 1 mol/L hydrochloric acid was added to adjust pH to 7.5 and diluted 5 mL with water to obtain 0.04 g/L C$_{60}$OH solution.

**Procedure**

Apt (20 µL of a 1.5 µmol/L solution), a certain amount of IPS, and 10 µL of 0.04 g/L fullerol solution were added to a 5-mL graduated tube, mixed well, and allowed to react for 10 min. Next, 200 µL 0.01 mol/L AgNO$_3$ and 70 µL 0.1 mol/L trisodium citrate were added and diluted to 1.5 mL. The mixture was heated for 21 min to 85°C in a water bath, then cooled with ice water. Next, 50 µL of 1.0 × 10$^{-5}$ mol/L VBB and 40 µL of 1 mol/L NaCl were added and mixed well. SERS spectra were recorded using a Raman spectrometer. SERS intensity of the reaction solution at 1614 cm$^{-1}$ ($I_{1614cm^{-1}}$) and the blank solution without IPS ($I_0$) were recorded. The value of $\Delta I = I_{1614cm^{-1}} - I_0$ was calculated.

**RESULTS AND DISCUSSION**

**Principle**

C$_{60}$(OH)$_n$ is a good electron acceptor (Samal and Sahoo, 1997). It transfers electrons from a donor to an acceptor, thereby facilitating, or catalyzing, the reaction. The silver nitrate–trisodium citrate reaction does not occur in solution because of the effective collision between silver ions and citrate. When C$_{60}$(OH)$_n$ is added, silver ions and citrate adsorb onto its surface, allowing electrons to transfer from citrate to silver ions, which leads to the generation of silver, 1,3-acetonedicarboxylic acid, and CO$_2$. Silver assembles as yellow Ag nanoparticles (Figure 1). Fullerol has abundant hydroxyl groups that can form hydrogen bonds with water; it combines better with silver ions and citrate for more efficient electron transfer. Thus, fullerol has enhanced catalytic ability compared to fullerenes lacking the hydroxyl group. An aptamer coating on the fullerol surface hinders the interaction between C$_{60}$(OH)$_n$ and citrate and silver ions, such that C$_{60}$(OH)$_n$ catalytic activity is inhibited. In the presence of IPS specific to the aptamer, C$_{60}$(OH)$_n$ is once again exposed to the reaction system and its catalytic activity is recovered. The amount of Ag nanoparticles generated increases linearly with IPS concentration. Using this relationship, a method to...
detect IPS using the SPR absorption spectrum, RRS, and SERS was developed.

**SERS Spectra**

In this study, fullerol exhibited increased catalytic activity compared to C₆₀. Fullerol has –OH groups with excellent water solubility that increase its catalytic activity compared to C₆₀. Thus, fullerols were prepared by H₂O₂ oxidization using a previously published procedure (Li et al., 1998). Hydroxyl content in fullerol increases with increasing H₂O₂ (Figure S1), as does the catalytic action of silver nitrate–trisodium citrate; a 70-mmol/L H₂O₂ solution was selected to obtain highly catalytic fullerol (C₆₀OH). VBB, VB₄R, RhS, Rh₆G, and RhB were used as signal molecules; their strongest SERS peaks occurred at 1614, 1385, 1361, 1362, and 1508 cm⁻¹, respectively (Figure S2). The SERS intensities of VBB and VB₄R were stronger than those of the others; VBB was chosen for further study. The catalytic activities of C₆₀OH, C₆₀, GO, and AgNP were investigated (Figure 2A and Figure S3). C₆₀OH exhibited the highest catalytic activity and was chosen for use. In the presence of Apt, which coats the fullerol surface and isolates it from the reactants, fullerol catalytic activity is suppressed and decreased SERS intensity is observed (Figure 2B). When added to the system, IPS conjugates to Apt, releasing fullerol and restoring its catalytic activity. As the IPS concentration increases, the amount of released fullerol increases and more AgNP is produced as well; thus, SERS intensity increases linearly with IPS concentration (Figure 2C and Figure S4).

**RRS and Absorption Spectra**

In a water bath at 85°C, fullerol and other nanoparticles catalyze the reaction of silver nitrate and trisodium citrate to generate AgNP, which exhibits two strong RRS peaks at 360 and 550 nm (Figure 3 and Figure S5A) and a strong surface plasma resonance (SPR) absorption peak at 410 nm (Figure 4 and Figure S6A). The RRS peak at 550 nm is characteristic of AgNP, and the intensity of ΔI₅₅₀nm and ΔΔA₄₁₀nm increase linearly with the amount of nanocatalyst. When Apt coats the nanocatalyst surface, it isolates the nanocatalyst from the system and inhibits its catalytic activity, leading to decreased ΔI₅₅₀nm and ΔΔA₄₁₀nm.
FIGURE 2 | SERS spectra. (A) C₆₀OH–AgNO₃–TSC catalytic system. (0, 2.67, 5.33, 13.33, 26.67, 53.33, 133.33, 266.67, 533.33 µg/L C₆₀OH+ + 1.33 mmol/L AgNO₃ + 4.67 mmol/L TSC + 85°C + 21 min + 3.33 × 10⁻⁷ mol/L VBB + 0.02 mol/L NaCl. (B) Apt–C₆₀OH–AgNO₃–TSC inhibitory catalytic system. (0, 1.03, 2.07, 5.17, 10.33, 15.5, 20.67 nmol/L Apt + 266.67 µg/L C₆₀OH+ + 21 min + 1.33 mmol/L AgNO₃ + 4.67 mmol/L TSC + 85°C + 3.33 × 10⁻⁷ mol/L VBB + 0.02 mol/L NaCl. (C) Apt–C₆₀OH–AgNO₃–TSC–IPS detection system. 20.67 nmol/L Apt + 266.67 µg/L C₆₀OH+ (0, 0.02, 0.05, 0.21, 0.52, 2.07, 5.17 µg/L) IPS + 1.33 mmol/L AgNO₃ + 4.67 mmol/L TSC + 85°C + 21 min + 3.33 × 10⁻⁷ mol/L VBB + 0.02 mol/L NaCl.
values. In the presence of IPS, which conjugates specifically with Apt, fullerol is released and its catalytic activity recovers. As IPS concentration increases, the amount of released fullerol increases, as does the amount of AgNP generated; consequently, $\Delta A_{550\text{nm}}$ and $\Delta A_{410\text{nm}}$ intensities increase linearly with IPS concentration (Figures 3B, 4B and Figures S5B, S6B).

The Catalytic Effect of C$_{60}$OH and Inhibition of the Aptamer

In the absence of the catalyst, AgNO$_3$ does not react readily with trisodium citrate. However, in the presence of a fullerol nanocatalyst, silver ions and citrate adsorb to the fullerol surface by interface free energy. As shown in Figure S7, the intensity of RRS for fullerol in aqueous solution is considerably lower than that of fullerene, suggesting that the fullerol particle size is less than that of fullerene. This may be responsible for the reduced catalytic activity of fullerenes compared to fullerol. In addition, silver ions and citrate adsorb to the surface more readily and electron transfer between the silver and citrate ions occurs more efficiently. That is, smaller particles demonstrate greater catalytic efficiency. As shown in Table 1, the slope of the C$_{60}$OH catalytic system is about 50 times that of C$_{60}$. In addition, the catalytic effect of AgNP on this reaction was studied. As shown in Figure S8 and Table 1, AgNP is an effective catalyst even with AgNP concentrations as low as 13.33 nmol/L. AgNP participates in autocatalysis, strengthening the catalytic effect. Furthermore, fullerene was hydroxylated using H$_2$O$_2$, according to a previously published procedure [42] and its catalytic activity was determined. Fullerol exhibited enhanced catalytic activity (compared to fullerene), and its catalytic activity increased with increasing hydroxylation (Table 1). This suggests that improved solubility of fullerol in water would increase its ability to bind with ions, thereby enhancing catalysis. When Apt is added, the intensity of RRS increases, as shown in Figure S7C, indicating

![Image of RRS spectra](image-url)
that Apt coats the nanocatalyst surface and blocks the adsorption of silver and citrate ions to the nanocatalyst, inhibiting its catalytic activity. It is worth mentioning that the catalytic activity of fullerol is suppressed by Apt (Table 1). This is likely due to its smaller size; the hydroxyl group of fullerol produced a better combination of hydroxyl and –COOH, –NH2.

**Scanning Electron Microscopy (SEM)**

The reaction solution was prepared and diluted 10 times. A 10 µL sample solution was dropped onto a silicon wafer and allowed to dry naturally, then scanning electron microscopy (SEM) was performed. As shown in Figure 5, in the absence of IPS, few AgNPs are detected in the reaction solution, with a mean grain size of 20 nm (Figure 5A). Upon addition of IPS, the catalyst recovered catalytic activity; a large amount of AgNP was generated and formed aggregates with a mean grain size of 40 nm (Figure 5B).

**Optimization of Catalysis Conditions**

The effect of reagent concentration on the determination was studied. When AgNO3 and TSC concentrations were 1.33 and 4.67 mmol/L, respectively, the SERS value was at its maximum. Thus, 1.33 mmol/L AgNO3 and 4.67 mmol/L TSC were chosen as the optimal concentrations (Figures S9, S10). The effects of reaction temperature and time were tested as well; 85°C and 21 min resulted in the maximum value for ΔI (Figures S11, S12). The effects of VBB, VB4R, RhB, RhS, and Rh6G concentrations on ΔI were considered. Maximum ΔI was observed at VBB, VB4R, RhB, RhS, and Rh6G concentrations of 3.33 × 10−7 mol/L, 1 × 10−6 mol/L, 1 × 10−5 mol/L, 1.67 × 10−6 mol/L, and 1 × 10−6 mol/L, respectively. Among these, VBB was selected because of its lower detection limit (Figure S13). At an Apt concentration of 20.67 mmol/L, the ΔI value reached a maximum value and thus was chosen for use (Figure S14). Binding times for the aptamers with fullerol were tested. A maximum value of ΔI was reached and maintained at 8 min; thus, it was chosen as the optimal binding time (Figure S15).

**Working Curve**

Using the optimal conditions described in section Optimization of Catalysis Conditions, working curves were prepared for IPS concentration at the corresponding ΔI<sub>1614cm</sub>−1, ΔI<sub>550mm</sub>, and ΔA<sub>410mm</sub> values for SERS, RRS, and Abs, respectively (Figure 6 and Figures S16–S19). Analytical characteristics are listed in Table 2. SERS exhibited the best performance, with a maximum slope of 492.83, and a limit of detection of 8.2 ng/L; RRS and 

![Image](https://via.placeholder.com/150)

**FIGURE 4** | Absorption spectra. (A) C6OH–AgNO3–TSC catalytic system, (0, 5.33, 13.33, 26.67, 53.33, 133.33, 266.67, 533.33 µg/L) C6OH + 1.33 mmol/L AgNO3 + 4.67 mmol/L TSC + 85°C + 21 min +3.33×10−7 mol/L VBB+0.02 mol/L NaCl. (B) Apt-C6OH–AgNO3–TSC–IPS detection system, 20.67 mmol/L Apt + 266.67 µg/L C6OH + (0, 0.1, 0.21, 0.52, 1.03, 2.07, 5.17 µg/L) IPS + 1.33 mmol/L AgNO3+4.67 mmol/L TSC + 85°C + 21 min +3.33×10−7 mol/L VBB+0.02 mol/L NaCl.

**TABLE 1** | The catalytic effect of various catalyst and the inhibiting effect of Apt.

| System | Linear equation | Linearity range | Correlation coefficient (R²) |
|--------|-----------------|-----------------|-------------------------------|
| C6O    | ΔI<sub>1614cm</sub>−1 = 0.24 C + 155.77 | 133.33–1333.33 µg/L | 0.9954                       |
| C6OH   | ΔI<sub>1614cm</sub>−1 = 9.33 C + 127.81 | 13.33–533.33 µg/L | 0.9947                       |
| C6OHf  | ΔI<sub>1614cm</sub>−1 = 9.33 C + 127.81 | 13.33–533.33 µg/L | 0.9947                       |
| AgNP   | ΔI<sub>1614cm</sub>−1 = 11.36 C + 50.21 | 1–359.56 µg/L | 0.9979                       |
| Apt-C6OH | ΔI<sub>1614cm</sub>−1 = 132.8 C + 157.15 | 1.03–20.67 nmol/L | 0.9797                       |
| Apt-C6O | ΔI<sub>1614cm</sub>−1 = 21.41 C − 9.85 | 5.17–51.67 nmol/L | 0.9913                       |
was the next most effective method. However, the Abs method is inexpensive, convenient, and aligns with national standards such that it could be used for on-site tests. Fullerol, because of its small size, higher surface electronic density, and ability to bind with silver and citrate ions, displays enhanced catalytic activity and a greater sensitivity for IPS detection compared

Table 2 | Analytical characteristics of the aptamer adjust catalysis-Ag nano plasma SERS for the determination of IPS.

| Test method | System | Working curve | Linearly range | Limit of detection | Coefficient (R²) |
|-------------|--------|---------------|----------------|-------------------|-----------------|
| SERS        | C₆₀OH-VBB | ΔI₁₁₁₄cm⁻¹ = 492.83 C + 29.74 | 0.02–5.17 μg/L | 8.2 ng/L | 0.9982 |
|             | C₆₀OH-VB4R | ΔI₁₁₁₄cm⁻¹ = 364.83 C + 35.84 | 0.02–5.17 μg/L | 9.1 ng/L | 0.9945 |
|             | C₆₀OH-RhS  | ΔI₁₃₆cm⁻¹ = 301.08 C + 62.38 | 0.02–5.17 μg/L | 8.7 ng/L | 0.9904 |
|             | C₆₀OH-Rh6G | ΔI₁₃₆cm⁻¹ = 292.33 C + 60.7 | 0.02–5.17 μg/L | 10.3 ng/L | 0.9906 |
|             | C₆₀OH-RhB  | ΔI₁₃₆cm⁻¹ = 203.21 C + 23.26 | 0.02–5.17 μg/L | 10.1 ng/L | 0.9946 |
|             | C₆₀OH-VBB  | ΔI₁₃₆cm⁻¹ = 78.41 C + 44.2 | 0.21–15.5 μg/L | 0.03 μg/L | 0.9954 |
|             | C₆₀OH-VB4R | ΔI₁₃₆cm⁻¹ = 80.67 C + 64.34 | 0.52–15.5 μg/L | 0.2 μg/L | 0.9816 |
|             | C₆₀OH-RhS  | ΔI₁₃₆cm⁻¹ = 361.28 C + 69.57 | 0.02–5.17 μg/L | 10.2 ng/L | 0.9824 |
|             | C₆₀OH-Rh6G | ΔI₁₃₆cm⁻¹ = 183.95 C + 46.36 | 0.1–5.17 μg/L | 0.02 μg/L | 0.9828 |
|             | C₆₀OH-RhB  | ΔI₁₃₆cm⁻¹ = 0.27 C + 0.07 | 0.52–15.5 μg/L | 0.03 μg/L | 0.9845 |
|             | C₆₀OH-VB4R | ΔI₁₃₆cm⁻¹ = 0.05 C – 0.0014 | 0.52–15.5 μg/L | 0.04 μg/L | 0.9917 |
TABLE 3 | Sample analysis results (n = 5).

| Sample          | Found  | Added  | Found  | Recovery/% |
|-----------------|--------|--------|--------|------------|
| Pond water      | –      | 2 µg/L | 2.01 µg/L | 100.5%     |
| Lijiang River   | –      | 2 µg/L | 1.95 µg/L | 97.5%      |
| Cropland 1      | –      | 2 µg/L | 2.03 µg/L | 101.5%     |
| Cropland 2      | 0.22 µg/L | 2 µg/L | 2.28 µg/L | 103%       |
| Grape           | –      | 2 µg/L | 1.86 µg/L | 93%        |
| Orange          | –      | 2 µg/L | 1.92 µg/L | 96%        |
| Chinese cabbage | –      | 2 µg/L | 2.01 µg/L | 100.5%     |
| Pond water      | –      | 2 µg/L | 2.01 µg/L | 100.5%     |

IPS was added to each sample, and recoveries of 93–101.5% were obtained (Table 3).

CONCLUSIONS

C₆₀OH is an effective catalyst to generate yellow AgNP via the AgNO₃-trisodium citrate reaction. The generated AgNPs exhibit a strong plasma resonance effect that increases linearly with catalyst amount at a certain concentration. RRS spectra demonstrate that the abundant hydroxyl groups of C₆₀OH increase its hydrophilicity and its ability to bind silver and citrate ions, resulting in increased catalytic activity compared to C₆₀. When C₆₀OH is coated with an aptamer, silver ions cannot bind to C₆₀OH, and catalytic activity is suppressed. Conversely, when isocarbophos conjugates with the specific aptamer, C₆₀OH is released and catalytic activity is recovered. SPR (Abs, RRS, and SERS) intensities increased linearly with increasing IPS concentration. Thus, aptamer binding and nanocatalysis combine with SPR to provide a sensitive, selective, simple, and rapid method for the determination of IPS.

DATA AVAILABILITY STATEMENT

All datasets presented in this study are included in the article/Supplementary Material.

AUTHOR CONTRIBUTIONS

AL and ZJ conceived and designed the experiments. HO performed the experiments, analyzed the data, and wrote the paper. All authors contributed to the article and approved the submitted version.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fchem.2020.00673/full#supplementary-material

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**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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