Optimization of Biogas Production in Dry Anaerobic Digestion of Swine Manure by the Use of Alkalinity Index to Monitor a Prototype Cylindrical Digester

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Abstract Anaerobic digestion (AD) is one of the best alternative sustainable technologies for energy production and recovery from organic solid wastes. Up to now dry AD has been commercialized in the treatment of municipal solid wastes. Little information could be found on the practical application of dry AD to manure wastes or waste activated sludge. This study aimed at testing the feasibility of using alkalinity to manage dry AD system for swine manure treatment and clarify its effect on the stability and efficiency of the newly-developed prototype cylindrical digester system. A prototype cylindrical digester with a diameter of 40 mm and a volume of 1.3 liters was designed and fabricated. It was operated under mesophilic conditions (38°C). The alkalinity of manure was increased by 3000 g/L (R1) and 6000 g/L (R2) by adding sodium bicarbonate with the raw swine manure as the control (R0). Results showed that R1 and R2 maintained a relatively higher level of alkalinity during the whole operation compared to the control (R0). Only one peak appeared in biogas production for the control reactor (R0) which almost ceased on day 12, whereas R1 and R2 exhibited two biogas peaks. The 30 days’ biogas yield for R2 was 276.6 ml/g- VSadded while R1 was 204.8 ml/g- VSadded which corresponds to an increase by 2.7- and 1.7- fold respectively as compared to the control (R0). 2.2- and 4.1-fold increase in methane production was achieved in R1 and R2 respectively as compared to R0. This difference is most probably attributable to the high alkalinity in R1 and R2 that stabilized the digestion process and minimized the influence of pH variations on methanogenesis.

Keywords: alkalinity, dry anaerobic digestion, prototype cylindrical digester, swine manure

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1. Introduction

Kenya, a sub-Saharan African country, has a national electrification level of 36% and thus only about 12.6% of the rural households have access to electricity [1]. Nationally, 69% of all households use firewood for cooking [2]. Inadequate power supply capacity due to rise in demand for electricity, which is growing faster than the ability to install additional generation plants is one of the greatest challenges. Furthermore, over-dependence on hydro-power, which exposes the country to power rationing due to extreme weather conditions that result in drought worsens this situation. The need for alternative energy resources is therefore imperative.

Dry Anaerobic Digestion (AD) has attracted increasingly extensive attentions in the studies of biogas fermentation with advantages of water-saving, higher volumetric organic loading rate, higher biogasification performance, smaller reactor capacity requirement, less energy used for heating, easier handle ability of digestate, and higher energy recovery as compared to wet AD [2,3]. This process is more feasible to a wide range of organic wastes including waste water sludge from industries with the recovery of renewable energy and reduction in pollution load [4]. The process also results in a lower production of leachate and easy handlings of digested residues that can further be treated by aerobic composting processes and used as organic fertilizer [5]. In this regard, dry AD is a remarkable method that could offer potential by-products such as fertilizer for the large population of farmers as well as energy generation. It is the most promising technology based on its characteristics and advantages. Over the past 20 years, different commercialized dry AD systems have been developed and marketed by different companies in Europe. The Kompogas, Valorga and Dranco dry AD systems are the most prevalent and have been mainly used for commercially processing of municipal solid waste, kitchen waste, or yard waste [6]. However, up to now there is no commercial dry AD system for treatment of manure waste.

Alkalinity is often referred to as buffer capacity in AD, which is the equilibrium of carbon dioxide and bicarbonate...
ions that provide resistance to significant and rapid changes in pH. The buffering capacity is therefore proportional to the concentration of bicarbonate and is a reliable method of measuring digester imbalance. Increasing a low buffering capacity is best accomplished by reducing the Organic loading Rate (OLR), although a more rapid approach is the addition of strong bases or carbonate salts to remove carbon dioxide from the gas space and convert it to bicarbonate, or alternatively bicarbonate can be added directly. A more sensitive parameter for monitoring digesters and measuring process stability is the VFA/alkalinity ratio: when this ratio is less than 0.35 - 0.40 (equiv. acetic acid/equiv. CaCO₃), the process is considered to be operating favorably without acidification risk [7].

[8] found out that the use of anaerobic baffled reactor as pretreatment prevented alkalinity shocks and acidity, avoiding sudden variation in pH, which positively assisted the maintenance of microbial activity, especially methanogenic archaea, due to the system buffering. The buffering conditions had positive influence on biogas production. [9] and [10] proposed the control of AD based on alkalinity titration. Automatic control of anaerobic digestion based on alkalimetric measurements was proposed by [11] and [12]. Similarly to VFA measurements, the utility of alkalinity as a control parameter in anaerobic digesters has not been established.

More rapid techniques are needed to avoid significant process deterioration and failure of dry AD process. Previously, researchers have put efforts to develop better indicators for process monitoring. An ideal indicator would be easy to measure, available on a real-time and perhaps on-line basis, and, of course, would have intrinsic meaning as it must reflect the current metabolic status of the system. Feasible and relevant indicators of the AD process condition have been argued for decades [13]. Little systematic information could be found about the management of dry AD systems.

According to previous research, dry AD has been commercialized in the treatment of municipal solid waste. Little information could be found on the practical application of dry AD to manure wastes or waste activated sludge, possibly due to their high nitrogen content which easily brings about high concentration of ammonia and large fluctuation in system pH resulting in low methane production or even AD failure. Moreover, little information is also available on how to use the alkalinity index to control and manage dry AD. Thus the objective of this study was to test the feasibility of using alkalinity to manage dry AD system for swine manure treatment and to clarify its effect on the stability and efficiency of a newly-developed prototype cylindrical digester system.

2. Materials and Methods

2.1. Swine Manure

The swine manure used in this study was obtained from a pig farm located in Tsukuba, Ibaraki, Japan. The pigs were raised in traditional pig houses paved with chopped straw bedding material to absorb urine. The manure was therefore collected in solid state, thoroughly mixed to achieve homogeneity and stored at 4°C before the experiments. The main characteristics of the swine manure are shown in Table 1.

| Parameters | Average value |
|------------|---------------|
| TS         | 27.9 %        |
| VS         | 217.0 g/L     |
| Alkalinity | 11300.0 mg/L  |
| pH         | 7.0           |
| Total VFAs | 21.6 mgC/g-VS |

TS - Total solids, VS - Volatile solids, VFAs - Volatile fatty acids

**Table 1. Characteristics of swine manure used in this study**

2.2. Experimental Setup

The digester used in this study was a 1.3-L clear PVC cylindrical pipe with a working volume of 1 L. It consisted of a sampling outlet at the center, a gas outlet port, digestate outlet and feed inlet as shown in Figure 1. A water bath constantly maintained at mesophilic temperatures of about 38°C was placed besides the reactor. This water was pumped round the digester to maintain a constant temperature in the digester. The gas outlet port on the digester was connected to a water displacement gas collection system consisting of sodium bicarbonate (NaHCO₃) solution and a graduated cylinder for measuring gas volume. All the outlet and the inlet ports were maintained at gas tight condition to ensure anaerobic conditions within the digester.

2.3. Experimental Design

The digester was subjected to several tests to ensure that it was gas tight. The TS of swine manure was adjusted to about 15% before dosing into the digester. The swine manure was then loaded into the digester until the digester was full. The initial total alkalinity, pH VS, TS and VFAs were determined on the day when the experiment began. The experiments were divided into 3 groups. The first experiment (R₀), i.e. the control experiment, had swine manure directly fed into the digester with its alkalinity unadjusted. In the second experiment (R₁),
The initial alkalinity of swine manure was increased by 3000 g/L before feeding into the digester. And the third experiment (R2) had the initial alkalinity increased by 6000 g/L before feeding into the digester. Digestate was sampled out after every 2 days with its alkalinity, pH, VS, TS and VFAs being determined. The amount of biogas produced was determined at an interval of 24 hours with its methane yield checked. The experiments were terminated when no significant gas production was detected. The efficiency of the systems was determined and compared at operation duration of 30 days.

2.4. Analytical Methods

Total solids (TS) and volatile solids (VS) were determined using standard techniques [14]. Total alkalinity was determined by titration method to an end point pH of 4.3. Biogas production was measured by water displacement method. The volume of biogas was directly read from the graduated cylinder. pH was measured directly in the digestate using a semi-solid pH meter (Testo 206, Germany). Gas composition was determined using a gas chromatography (GC-8A, SHIMAZU, Japan) equipped with a thermal conductivity detector (80°C) and a Porapak Q column (60°C). Nitrogen (N2) was used as the carrier gas.

For VFAs analysis, 4 g of well mixed sample was diluted with 40 ml of deionized water. The mixture was centrifuged at 9900 rpm and 4°C for 20 min after being shaken well. The resultant supernatant was filtrated through filter membranes with a pore size of 0.22 µm. 3% phosphoric acid solution was added to the filtrate to acidify samples at a volume ratio of 1:9. Gas chromatograph (GC-8A, Shimadzu) equipped with Unisole F-200 30/60 column and flame ionization detector (FID) was used for quantification of VFAs, including HAc, HPr, i-HBu, n-HBu, i -HVa and n -HVa. 1µl prepared samples were injected to GC-FID directly with a retention time of 12 min. Nitrogen was used as carrier gas. The temperature of injection port and column were maintained at 180°C and 150°C, respectively.

3. Results and Discussions

3.1. Alkalinity and pH Variation

A high alkalinity trend was observed in R1 and R2 when compared to the control (R0) as shown in Figure 2. This was probably because NaHCO3 was added in R1 and R2 which boosted the total alkalinity. There were two alkalinity peaks in R1 and R2 that almost corresponded to the respective daily biogas production peaks. In the first 30 days, R2 had the highest average alkalinity of 18,090.9 mg/L while R1 and R0 had 15,196.4 mg/L and 11,628.6 mg/L respectively. In R1 and R2, alkalinity was observed to increase up to the biogas peak and later reduced after the peak possibly due to utilization of VFAs to form methane gas that reduced the acid levels in the reactor. Also, the production of ammonia might elevate the system pH.

In all the three reactors, pH decreased during the first 4 days of operation as shown in Figure 3. We can easily understand that the pHs decreased first due to the hydrolysis and acidification process, and then increased to a relatively stable value during the methanogenesis process. With a low buffer capacity, the pH of R0 decreased dramatically to less than 6.0 and then increased slightly to 6.3 however, it couldn’t rise more. This may be attributed to the production and accumulation of VFAs that increased the acid levels in the digester which inhibited the activity of methanogenic bacteria. In R1 and R2, the pH was able to rise again to an average of 7.1 and 6.8 respectively due to high buffer capacity that was able to neutralize the acids produced. According to [5] maximal biogas yield in anaerobic digestion was found to be in the pH range of 6.5 - 7.5. The average pHs in R1 (6.8) and R2 (7.1) were detected to be able to maintain within this pH range and hence maximum yield of biogas as compared to R0 (6.3). From these results, it can be easily seen that increase in alkalinity is beneficial to maintain stable pH within the optimum condition for biogas production, even in dry AD process.

3.2. Biogas Production

R1 and R2 had two biogas peaks observed on day 10, 26 and day 11, 20 respectively as shown in Figure 4. The 30 days’ biogas yield for R2 was 276.6 ml/g-VSadded while R1 was 204.8 ml/g-VSadded which corresponds to an increase by 2.7- and 1.7-fold respectively as compared to the control R0 (74.8 ml/g-VSadded). This was possibly due to high alkalinity in R1 and R2 that was able to maintain the pH within optimum range in the digester which favored methanogenesis process. R0 had only one biogas peak on day 5 and methanogenesis stopped on day 12, probably due to its
low buffering capacity and hence inhibition from the VFAs accumulation which increased the acid levels thus lower pH in the digester. Within 30 days, R₁ and R₂ produced 22.8 and 31.9 liters of biogas which corresponds to 1.6- and 2.7-fold increase in biogas production respectively as compared to R₀ (8.7 liters). These findings are consistent with [15] who obtained the highest biogas production from pH 7.0 reactor in comparison to other different pH reactors. On day 30, biogas production had stopped in R₂ while there was still biogas produced in R₁ as shown in Figure 5, therefore R₂ had a shorter biogasification period than R₁.

3.3. Methane production

R₂ had the highest methane yield of 230.6 ml/g-VS added, as compared to R₁ (151.0 ml/g-VS added) and R₀ (45.1 ml/g-VS added) respectively for the first 30 days. R₁ and R₂ produced 16.8 and 26.6 liters of methane; this corresponds to 2.2- and 4.1-fold increase in methane production respectively as compared to R₀ (5.3 liters) within the first 30 days. In Figure 6, R₀ was observed to be stabilized at around 5 liters which was the lowest as compared to R₁ and R₂. R₁ had not yet stabilized due to its continuous production of methane after the 30th day. R₂ stabilized at around 26.6 liters which was the highest achieved in these experiments.

There was a significant increase in average methane content from control R₀ (60.3%) to R₁ (73.7%) and R₂ (83.4%) after initial alkalinity of swine manure being elevated (shown in Figure 7). Results from this work agrees with [16] who found out that CH₄ content significantly increase to 74% under alkaline condition as compared to other conditions.

3.4. Volatile Solids Removal

One of the most useful parameters for evaluating the efficiency of anaerobic digestion is the reduction in VS. VS are decreased due to solubilization of particulate organics for biogas production. 2.2- and 0.9-fold reductions in VS were achieved in R₂ and R₁ respectively as compared to R₀. This is shown in Figure 8. R₂ had the highest VS conversion efficiency among the 3 tested reactors.

This may be due to the high alkaline environment in R₂ that facilitated optimum condition for degradation and solubilization of the organics which enhanced biogas production. Initially, R₀, R₁ and R₂ had a VS concentration of 116.65 g/L, 111.36g/L and 115.41 g/L, respectively. On the 30th day, R₀, R₁ and R₂ had a VS concentration of 92.82 g/L, 65.10 g/L and 40.34g/L respectively. R₂ had the lowest VS content in the final effluent after 30 days’ digestion period whereas R₀ had the highest VS content.
3.5. Profiles of Volatile Fatty Acids

Initially in R₀, R₁ and R₂, VFAs increased during the acidogenesis phase as shown in Figure 9. In R₁, total VFAs increased to 53.71 mgC/g-VS whereas in R₂ it increased to 42.91 mgC/g-VS on the 9th day. Later on, the total VFAs decreased to 3.90 mgC/g-VS in R₁ and 3.72 mgC/g-VS in R₂ on the 30th day. This decrease is probably attributable to the utilization of VFAs for methane production in the subsequent methanogenesis phase. The high buffering capacity in R₁ and R₂ provided an optimum environment for the methanogenesis process. In R₀, total VFAs did not subsequently decrease after the continuous increase to 53.63 mgC/g-VS on the 20th day, this therefore caused its accumulation and hence inhibition on the methanogenesis process due to acidic environment.

From the VFAs profiles in Figure 10, accumulation of propionic acid (HPr) was observed in R₀ which could be associated with the slow HPr degradation. HPr degradation depends on the activities of propionic acid oxidizing bacteria (POB) which are slow-growth microorganisms and can be sensitive to reactor operating conditions [17,18]. [19] reported that pH was an important parameter affecting growth of a mixed POB culture and the optimal pH range for its growth was 6.80–8.50. R₀ with low buffering capacity and an average pH of 6.3 was outside the optimal range as compared to 6.8 and 7.1 for R₁ and R₂ respectively. This probably led to low degradation of HPr that increased its accumulation further lowering reactor pH and hence inhibiting the methanogenesis process. These results are consistent with [20] and [21] who found that accumulation of VFAs especially HPr led to failure in AD systems.

4. Conclusions

Energy is the heart of most critical economic, environmental and developmental issues facing developing countries today. The current patterns of energy production and consumption in Kenya are unsustainable and threaten the environment on both local and global scales. Biogas production through AD provides a sustainable source of energy and the bio-slurry produced enriches the soil for agricultural activities. It also provides an opportunity to treat and re-utilize organic wastes and reduces land use problems associated with the disposal of organic waste. This study investigated the effect of different levels of alkalinity on the efficiency of dry anaerobic digestion process of swine manure.

The following major conclusions could be arrived at.

1. A high alkalinity trend was observed in R₁ and R₂ when compared to the control (R₀). In the first 30 days, R₂ had the highest average alkalinity of 18,090.9 mg/L while R₁ and R₀ had 15,196.4 mg/L and 11,628.6 mg/L respectively.

2. R₂ (+6000mg/L) exhibited a stable digestion process as compared to R₁ (+3000mg/L) and R₀ (control). The 30 days’ biogas yield for R₂ was 276.6 ml/g-VS added while R₁ was 204.8 ml/g-VS added which corresponds to an increase by 2.7- and 1.7-fold respectively as compared to the control R₀(74.8 ml/g-VS added). R₁ and R₂ produced 16.8 and 26.6 liters of methane; this corresponds to 2.2- and 4.1-fold increase in methane production respectively as compared to R₀(5.3 liters) within the first 30 days.

3. A high rate of VS removal was detected in R₂ which had the highest VS conversion efficiency among the 3 tested reactors. 2.2- and 0.9-fold reductions in VS were achieved in R₂ and R₁ respectively as compared to R₀. This may be due to the high alkaline environment in R₂ that facilitated optimum condition for degradation and solubilization of the organics which enhanced biogas production.

4. VFAs accumulation especially HPr is one of the major causes of AD system failure. R₀ achieved an average pH of 6.3 that was outside the optimal range for HPr degradation as compared to 6.8 and 7.1 in R₁ and R₂ respectively. This probably led to low degradation of HPr that increased its accumulation. The accumulation of VFAs further lowered reactor pH and hence inhibited the
methanogenesis process as observed from the low biogas yield in $R_0$.

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