An “acquired” hemoglobin J variant in a sickle cell disease patient

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Abstract: We report the case of a rare hemoglobin variant, “Hemoglobin J", discovered while performing hemoglobin electrophoresis following exchange transfusion of a sickle cell disease patient. It is usual practice in our institution to confirm the hemoglobin S level in sickle cell disease patients after red cell exchange. The patient had received 5 red cell units and the source of this variant was traced back to two of those units. Due to the uncertain clinical impact of this variant, and the lack of specific guidelines, the two donors were deferred from future donations to our institution.

Keywords: hemoglobin J, sickle cell disease, transfusion

Introduction
The clinical phenotype of sickle cell disease most commonly has one of the following genotypes: HbS/S, HbS/C or HbS/B thalassemia. Less commonly, the genotype may be double heterozygous traits of hemoglobin S with an uncommon hemoglobin trait such as Hb D, Hb O-Arab, or hemoglobin Lepore. The clinical impact of rare hemoglobin variants such as Hb J when combined with hemoglobin S disease (or trait) is not clear in the medical literature. Many sickle cell disease patients are on chronic transfusion to prevent stroke and other complications. Routine post-transfusion testing includes hemoglobin electrophoresis, mainly to assess the level of hemoglobin S. The goal is to maintain the patient hematocrit at 25% to 30% and the proportion of HbS below 30% (Elghetany and Davy 2001; Brecher 2005).

Case report
A 44-year-old African American male with history of sickle cell anemia presented to our institution for scheduled red blood cell exchange transfusion. He received 5 units of red blood cells matched for C, E, and K red cell phenotype.

Pre and post transfusion hemoglobin electrophoresis was performed and results revealed a new band on the electrophoretic gel, indicating the patient had “acquired” a new hemoglobin variant.

The variant band was seen in the location of Hb J (a fast migrating variant) on isoelectric focusing (IEF) and cellulose acetate. It migrated with “A” on citrate agar. The variant constituted approximately 15% of total hemoglobin.

Clinical follow up of the patient showed no complications and the clinical course was consistent with the patient’s underlying medical condition.

Retained segments from the donated units were obtained and hemoglobin electrophoresis revealed two of the suspected units having hemoglobin J. Those units were donated by two sisters.

Pathologic findings
The patient’s pre- and post-exchange hematological values are shown in Table 1. His pre- and post-transfusion hemoglobin electrophoresis was performed with the
following results: Cellulose acetate with a prominent Hb A band, smaller band in the S region, and a fast hemoglobin in the position of Hb J (Figure 1). The bands on citrate agar (acid) show prominent A and a smaller band located in the position of S (Figure 2). IEF was done on samples from the patient blood and the 5 donated units. The variant band appeared in three samples (the patient’s blood and two of the donated units), located in the position of Hb J (these bands were more prominent in the donor’s sample) (Figure 3).

The source of the variant band was apparently the 2 units of transfused blood. The donor center was notified and the transfused units were then traced to determined significant donor history, such as possible recalls of any other components.

**Discussion**

The human hemoglobin molecule is composed of polypeptide chains called globins, and iron-containing porphyrin rings termed heme. Every hemoglobin variant is composed of two alpha and two nonalpha globin chains, while the heme component is usually the same.

Hemoglobinopathies are a group of inherited mutations of the globin genes leading to qualitative and/or quantitative abnormalities of globin synthesis.

There are hundreds of different variants of hemoglobin caused by structural alteration of alpha, beta, or gamma globin chains varying from amino acids replacements, elongated deletions, insertions, or both deletions and insertions (Huisman et al 1998; Elghetany and Davy 2001).

Abnormalities of beta-chain or alpha-chain produce most of the clinically significant hemoglobinopathies. The status of zygosity also plays a very important role in the expression and detection of the disorder. In heterozygous variants the other normal allelic gene produces normal chains which may compensate for the defective gene. In the homozygous state, both allelic genes are affected which results in the production of a large amount of the variant (Scrivener et al 1985; Elghetany and Davy 2001).

**Table 1** Patient’s pre- and post-exchange CBC results

| Pre-transfusion | Post-transfusion | Reference ranges |
|----------------|------------------|------------------|
| RBC            | 2.87             | 3.29             | (3.9–5.3) M/cu mm |
| Hb             | 8.3              | 9.7              | (11.5–13.5) g/dL |
| Hct            | 24.4%            | 29.1%            | (34%–40%)         |
| RDW            | 23.9%            | 18.8%            | (10%–14.1%)       |

Abbreviations: Hb, hemoglobin; Hct, hematocrit; RBC, red blood cell; RDW, red cell distribution width.

**Figure 1**
Post-red cell exchange hemoglobin electrophoresis (cellulose acetate, pH 8.6) of the patient showing bands at positions Hb S, Hb A, and Hb J.
Abbreviation: Hb, hemoglobin.

**Figure 2**
Post-red cell exchange hemoglobin electrophoresis (citrate agar, pH 4.6) of the patient showing bands at positions Hb S and Hb A.
Abbreviation: Hb, hemoglobin.

**Figure 3**
Hemoglobin electrophoresis (isoelectric focusing method) showing a band of Hb J (arrowhead) in the patient (post-exchange) and the two red cell units from the implicated donors.
Abbreviation: Hb, hemoglobin.
The vast majority of hemoglobin variants are encountered in the heterozygous state and most individuals carrying these variants have a completely normal physical picture and normal hematological tests. In the US, most hemoglobin variants are diagnosed in early childhood. This is mainly a benefit of newborn screening which began in the 1980’s and is currently conducted in at least 44 states.

One of the most common hemoglobinopathies is sickle hemoglobin (Hb S), which is responsible for the sickle cell disease and trait. The molecular nature of this hemoglobin variant is a substitution of valine for glutamic acid at the sixth amino acid position in the beta globin gene, which results in decreased solubility of Hb, causing red cells to sickle (Suarez et al 1997; Rechavi et al 1986; Strobel et al 1987; Lambridis et al 1986; stidda et al 2002).

Most hemoglobinopathies are prevalent in tropical regions, perhaps due to the advantage of the heterozygote state in areas where malaria is endemic. Malaria parasites live inside red blood cells, but subtly disturb normal cellular function. In patients predisposed to rapid clearance of red blood cells, this may lead to early destruction of cells infected with the parasite and increased chance of survival for the carrier of the trait.

The prevalence of Hb S in the US is mainly in people from African or Hispanic descent. Sickle trait has a prevalence of 8%–10% among African Americans. Approximately 1/500 African Americans and 1/1000 Hispanics have sickle cell anemia.

The prevalence of hemoglobin variants among blood donors is unknown, as no hemoglobin electrophoresis is routinely performed on donated units.

We are aware of only a few articles in the literature describing hemoglobin variants acquired through transfusion (Gibaud et al 1974; Rechav et al 1986; Strobel et al 1987; Robertson et al 1997; Suarez et al 1999).

To our knowledge there is no reported case in the literature of transfusion-acquired hemoglobin J.

There are more than 50 hemoglobin J variants described in the literature. For example, Hb-J Capetown (alpha92(FG4)Arg->Gln), Hb-J Buda (alpha61(E10)Lys->Asn), Hb-J Chicago (beta76(E20)Ala->Asp), Hb-J Sardegna (alpha50(CE8)His->Asp), and Hb-J Toronto 9 alpha5(A3)Ala->Asp), etc. They all have an electrophoretic mobility “faster” than “A” on cellulose acetate (ie, close to the anode) in common. All are classified under “variants of the alpha- or beta-chains” (single or multiple base changes), or “hemoglobins with more than one amino acid substitution in the alpha chain” (eg, J-Singapore (alpha78(EF7)Asn->Asp) (Botha et al 1966; Lambridis et al 1986; stidda et al 2002).

One recently described hemoglobin J, found in a few members of an Italian family, named Hb J-Europa has a beta chain substitution (beta62(E6)Ala->Asp) (Huisman et al 1998).

Hemoglobin J (depending on its type) has different characteristics and functions. For example hemoglobin J Capetown (alpha2 92Gln beta 2), the most commonly seen Hb J variant (CGG->CAG), is associated in the heterozygous state with increased oxygen affinity and polycythemia. Those affected may also have a mild erythrocytosis and microcytosis (Botha et al 1966; Elghetany and Davy 2001). Other variants like Hb J Sardegna will show a completely unremarkable clinical picture in the heterozygote. Hemoglobin J Bankock (beta 56 Gly->Asp) and J Baltimore (beta 16 Gly->Asp) have been described in combination with sickle hemoglobin. These individuals of African descent were reportedly clinically asymptomatic, however, as a double heterozygote there may be the potential of sickle cell trait-like complications (Gellady and Schwartz 1973; Gunay et al 1974; Weatherall 1974).

In this particular case, review of the history of the two blood donors revealed them to be sisters with no significant past medical history (which would have deferred them as donors initially). These sisters, 81- and 83-year-old Caucasian women, from French, German, and English descent had a long history of blood donation (more than 15 donations each, starting in 1997). They were unaware of having this hemoglobin variant and were not aware of any abnormalities in their family (parents and 3 siblings).

There were no transfusion-associated adverse events reported in any of the recipients of their previously donated red blood cells, nor in the patient who received the current units. Further characterization of this hemoglobin variant was not indicated clinically, so molecular studies were not pursued.

The decision to defer the donors was made at the discretion of the medical director of our blood donor center. This decision is admittedly open for questions and debate as there are currently no specific guidelines or policies in the medical literature for accepting or deferring donors after discovering
a rare hemoglobin variant. We hope that this case report will stimulate discussions that will lead to specific guidelines.

Acknowledgment
The authors would like to thank Barbara Grooms for her administrative assistance in preparing this manuscript for publication.

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