Quantitative assessment of vascular density in diabetic retinopathy subtypes with optical coherence tomography angiography

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Abstract

Background: Quantitative assessment of vascular density (VD) of retinal and choriocapillaris (CC) in various stages of diabetic retinopathy (DR) using spectral domain optical coherence tomography angiography (SD OCTA).

Methods: 188 eyes of 97 participants were recruited in this cross-sectional study. The macular OCTA (3x3mm) scan was performed and the computer algorithm assessed VD at the level of superficial capillary plexus (SCP), deep capillary plexus (DCP) and CC.

Results: All measured parameters were decreased in retinal VD at the more extreme stages of DR, with the exception of SCP foveal VD. There was a constant pattern of decrease in VD of CC from normal cases to cases of NDR and NPDR and then a slight increase occurred in the PDR stage but never touching the normal quantities. Age, fasting blood sugar, and years of diabetes mellitus were correlated with reduced VD in different segments. Multivariate linear regression analysis showed that best-corrected visual acuity (BCVA) was positively correlated with parafoveal VD at SCP and VD of foveal area at CC. VD of all subfields of macular area except foveal DCP VD showed reduced levels in diabetic macular edema (DME) patients compared to those without DME.

Conclusions: The findings of the study endorse retina VD changes as a potential biomarker for DR development before retinopathy becomes clinically evident. It seems that parafoveal VD of SCP and foveal VD of CC are good biomarkers to predict VA in the diabetic patients.

Keywords: Capillary plexus, Choriocapillaris, Diabetes, Diabetic retinopathy, Optical coherence tomography angiography, Vascular density

Background

Diabetic retinopathy (DR) is a degenerative neurovascular disease caused by activation of multifactorial pathologic mechanisms, which leads to microvascular abnormalities including microaneurysm development, capillary non-perfusion, vascular leakage, and neovascularization [1–3]. Microvascular changes and destructions, as the loss of pericytes and endothelial cells and capillary leaks and occlusions also occur in earlier DR stages [4–6]. The resulting ischemia induces upregulation of angiogenic signaling molecules, including vascular endothelial growth factor (VEGF) and erythropoietin [7], which increases vascular permeability and ultimately fosters proliferative diabetic retinopathy (PDR) [8].
Fluorescein angiography (FA) and indocyanine green angiography (ICGA) are the gold standard imaging modalities for imaging of microvascular abnormalities of retina and choroid in DR for years. However, a number of limitations, including their invasive nature, the risk of allergic reactions in patients with iodine or seafood allergy or idiosyncratic reaction, the cost, the long duration of imaging and the occasional shortage of fluorescein and ICG dyes, have raised the clinical need for a shift to newer imaging techniques. For these reasons, FA and ICGA are not routinely performed in the early stages of DR for the assessment of the retinal and choroidal vasculatures [9].

New noninvasive imaging modalities (as OCTA) may provide valuable information about microvascular changes, the perfusion status of the retina and the likelihood of retinopathy progression during the various stages of DR. OCTA uses the motion contrast provided by flowing erythrocytes to allow dye-free and volumetric visualisation of the retinal and choroidal vasculatures at micron-scale resolutions [10–13]. This technology uses split-spectrum amplitude-decorrelation angiography (SSADA) algorithm to detect motion of erythrocytes in the capillaries [14]. OCTA delivers depth-resolved retinal vascular structure images, making it possible to differentiate superficial and deep capillary plexuses (SCP and DCP) of retina and also choriocapillaris (CC) [11, 15, 16]. Previous qualitative studies in DR have shown that OCTA is able to delineate retinal capillary nonperfusion with higher resolution than conventional FA [17–19].

Previous studies have shown that vascular density (VD) of parafovea in SCP and DCP decreases with significant FAZ enlargement in diabetic patients with DR, even in those with no diabetic retinopathy (NDR), compared to healthy subjects [20, 21]. There were no significant differences in the retinal thickness between control subjects and patients with NDR at the same time. Significantly reduced VD in the SCP and DCP in mild non-proliferative DR (NPDR) in comparison to control subjects has also been observed [22]. It seems that retinal vascular alterations precede structural changes in the retina. This may highly suggest a causal role of circulatory deficit in the development of DR. [20] Kim et al. have detected qualitatively decreasing capillary density, branching complexity, and progressively increasing average vascular caliber in eyes at different stages of DR. [23] Overall, depending on the method used and methodological differences and metabolic status of the patients studied, conflicting results of the blood flow in diabetic retinal vessels have also been reported both to decrease [24–26] and to increase [27–30].

In this cohort, the aim is the quantitative measurement of VD at foveal and parafoveal area (as a marker of macular perfusion) using OCTA in subtypes of DR including those with NDR, NPDR, and proliferative diabetic retinopathy (PDR) as a continuum, and to compare these findings with each other and the normal population. In addition, we evaluate the correlation of VD as an independent predictor of BCVA in diabetic patients.

**Methods**

This prospective cross sectional study was performed between January 2015 and December 2019 at Farabi Eye Hospital, a tertiary university eye center in Tehran, Iran. The institutional ethics committee of Tehran University of Medical Sciences approved the research protocol and a written informed consent was obtained from the participants. The study adhered to the tenets of the Declaration of Helsinki. More than 10 eligible patients did not provide informed consent and were excluded from the study.

Naïve diabetic patients with a history of more than 10 years have been recruited and the control patients have been selected from healthy volunteers. Inclusion criteria were best-corrected visual acuity (BCVA) of 20/20 for normal cases and refractive error between −3 and +1 D spherical equivalent in all groups. The normal volunteers had to have no ocular and systemic disease. Exclusion criteria were significant media opacity preventing high-quality imaging, motion and blinking artifact on the images, poor quality images, previous focal or panretinal laser photocoagulation, intravitreal anti-VEGF, steroid and/or potentially retinotoxic or neurotoxic drugs consumption, optic neuropathy, any previous ocular and macular disease, previous surgeries other than uncomplicated phacoemulsification (more than 3 months), any inflammatory diseases or active or recent infectious disease (ocular and/or systemic), immunosuppressive drugs or biologic therapies, pregnancy, and uncontrolled hypertension.

Demographic characteristics and relevant laboratory tests such as fasting blood sugar (FBS) and total serum cholesterol level and presence of hypertension were documented. Hemoglobin A1C levels were not checked for all the cases. Best-corrected visual acuity (BCVA) was measured on a Snellen chart and expressed as the logMAR.

Subjects underwent thorough ophthalmic exam including slit lamp biomicroscopy and fundus examination. Intraocular pressure (IOP) was measured with Goldman applanation tonometry. Diabetic patients have been diagnosed based on the criteria of the American Diabetes Association and all were under treatment for diabetes. The diabetic patients were classified into three groups ranked in ascending order of DR severity: NDR, NPDR, and PDR, based on early treatment for diabetic retinopathy study (ETDRS) classification [3, 31].
Acquisition of the images
Clinical examination and SD-OCT imagings were performed at the same day between 8:00 am and 2:00 pm. Two professional image readers (FG, SB) checked and assessed all of the OCT images. AngloVue OCTA imaging (RTVue XR Avanti; Optovue, USA- version: 2016.1.0.23- beta) using SSADA algorithm was performed [32, 33]. This instrument performs 70,000 A-scans per second (840-nm) to capture OCTA images of horizontal and vertical B-scans in transverse dimension to provide a 3 × 3 mm (304 × 304 pixels) image centered at the fovea. Scans with low quality (i.e., presence of blink or motion artifact) were repeated until good quality scans were achieved. Automated segmentation was utilized for defining SCP, DCP, and CC. The SCP was defined as area between 3 μm below to internal limiting membrane (ILM) and 15 μm below internal plexiform layer (IPL). The DCP was considered to be between 15 μm and 71 μm below IPL.

Automatic segmentation was fine-tuned manually where appropriate. Each macular OCT-A layer was subdivided into nine areas of interest—whole image (WI), fovea, parafovea, superior hemifield (SH), inferior hemifield (IH), temporal (T), superior (S), nasal (N), and inferior (I) for quantitative measurements of the vascular density of SCP, DCP, and CC. The foveal region was outlined as a central circle with a 120-pixel (1.2 mm) diameter, and the parafoveal region was delineated as a ring, by 91 pixels wide, surrounding the foveal region [34]. To calculate VD, the AngloVue Analytics software extracts a binary image of the blood vessels from the gray scale OCTA image, and then calculates the percentage of pixels occupied by blood vessels in the defined region [35]. Diabetic macular edema (DME) was defined as the central macular thickness (CMT) of more than 300 μm.

Statistical methods
All quantitative variables were reported as mean with standard deviation after confirming normality of distribution with the Kolmogorov–Smirnov test. Non-normal distributed parameters are reported by median with the range. All statistical analyses were performed using statistical software (SPSS software Version 21; SPSS, Inc., Chicago, IL, USA). Kruskal–Wallis test and one-way analysis of variance (ANOVA) were performed for nonparametric and parametric comparison. Mann Whitney U test and post-hoc analysis (dunnett’s test) were used to compare choroidal thicknesses between groups. In this study collinearity for different variables was checked. P values less than 0.05 were considered statistically significant.

Results

General characteristics
A total of 188 eyes of 97 participants with the mean age of 56.5 ± 8.9 years (range: 25–80) were analyzed. Of these 41 (42.3%) were male and 56 (57.7%) were female. BCVA was significantly lower in the patients with PDR and NPDR in comparison with NDR and normal subjects.

Mean FBS was 209.85 ± 85.6 mg/dl in the diabetic patients. Mean diabetes mellitus (DM) duration in diabetic patients was 12.7 ± 6.3 years. The study included 40 eyes in the control group and 148 eyes in the diabetic group. Based on the DR severity scale, the diabetic group had 39 (26.4%) eyes with NDR, 41 (27.7%) eyes with mild to moderate NPDR, 25 (16.9%) eyes with severe NPDR, 26 (17.6%) with early PDR and 17 (11.5%) eyes with high-risk characteristic PDR. Diabetic macular edema (DME) was present in 22 (34.4%) of NPDR and 16 (35.6%) of PDR groups. Table 1 presents the baseline characteristics of the participants.

The difference in BCVA (decimal), sex and age were statistically significant. Spherical equivalent, axial length or intraocular pressure did not vary significantly in the groups (p > 0.05). Hyperlipidemia and hypertension were more prevalent in diabetic groups than the control group (p < 0.001). The hypertension was under control in all groups.

Vascular density
Tables 2, 3, and 4 show VD in SCP, DCP, and CC, respectively. Figure 1 illustrates the distribution of grid based VD in the SCP, DCP, and CC in a radar plot. Statistically significant differences were observed in VD at various subsegments among normal control subjects and different stages of DR. All the VD amounts in SCP and DCP had abnormal distribution in the Kolmogorov–Smirnov test. The trend toward lower amount of VD (median) from normal subjects to PDR patients in macular area and all various subsegments was notable in both SCP and DCP. There was continuous and significant decrease from the normal cases to the NDR group in both SCP and DCP (Tables 2 and 3). The comparative tests (Mann-Whitney U test) showed that in all studied subsegments the decrease in the amounts by stepping from one stage to the others was statistically significant (P < 0.05). The exception was the foveal VD that was not significantly changed when comparing NDR with NPDR and NPDR with PDR in both SCP and DCP.

We have seen a trend of fluctuation in the VD of SCP and DCP in the whole image and foveal region, but a steady slight decrease from NL to the PDR level is evident, which is more pronounced in the NPDR stage in the parafoveal region. (Fig. 2, Table 5).

In the CC the amounts of VD were normally distributed and the amount of the changes were statistically
Table 1 Demographic characteristics of normal subjects and subtypes of diabetic retinopathy (N = 188)

| Groups                     | NL (40 eyes) (M ± SD) | NDR (39 eyes) (M ± SD) | NPDR (64 eyes) (M ± SD) | PDR (45 eyes) (M ± SD) | P-value |
|----------------------------|-----------------------|------------------------|-------------------------|------------------------|---------|
| Age (Y)                    | 50 ± 0.72             | 59.28 ± 1.02           | 58.31 ± 1.14            | 59.03 ± 1.68           | 0.003   |
| (%) OD                     | 20 (50)               | 19 (48.7)              | 33 (51.6)               | 25 (55.6)              | 0.929   |
| Sex-male (%)               | 24 (60)               | 15 (38.5)              | 25 (39.1)               | 20 (44.4)              | 0.156   |
| BCVA (decimal)             | 0.93 ± 0.01           | 0.84 ± 0.02            | 0.65 ± 0.02             | 0.62 ± 0.04            | < 0.001 |
| The last FBS (mg/dl)       | –                     | 158.46 ± 5.72          | 189.78 ± 8.38           | 202.12 ± 7.30          | < 0.001 |
| Duration of diabetes mellitus (Y) | –                     | 12.38 ± 1.02          | 12.96 ± 0.78            | 16.54 ± 1.1            | < 0.001 |
| Hypertension number (%)    | 2 (5%)                | 16 (41%)               | 27 (42%)                | 17 (51.5%)             | < 0.001 |
| Hyperlipidemia number (%)  | 0 (5%)                | 15 (38.5%)             | 32 (50%)                | 16 (48.5%)             | < 0.001 |

**BCVA** Best corrected visual acuity, **DME** Diabetic macular edema, **NDR** No diabetic retinopathy, **NL** Normal, **NPDR** Nonproliferative diabetic retinopathy, **PDR** Proliferative diabetic retinopathy

significant (ANOVA) (Table 4). Post-hoc analysis (Tuckey test) revealed no statistically significant changes between normal cases and the patients with NDR (P > 0.05). The VD of CC under fovea and all studied subsegments increased in NDR cases and then in NPDR and PDR cases decreased to less than those in normal cases. This comparison with NDR showed significant changes in the NPDR group in all subsegments.

**Correlations of VD**

In order to determine the association of VD and other variables univariate analysis was performed for diabetic patients. Age was negatively correlated with parafoveal VD at SCP (r = -0.189, P = 0.015).

Systolic blood pressure was weakly inversely correlated with WI-MVD in CC (r = -0.155, P = 0.034) but not with VD in other segments. FBS was inversely correlated with WI-VD in SCP, DCP (r = -0.50, P < 0.001; r = -0.460, P < 0.001; and r = -0.222, P = 0.002; respectively) and foveal VD in SCP, DCP (r = -0.264, P < 0.001; r = -0.371, P < 0.001; respectively) but had weak negative correlation with foveal VD at CC layer (r = -0.141, P = 0.056).

DM duration in patients was also a significant factor in association with VD. It had inverse correlation with WI-VD in SCP, DCP, and CC (r = -0.354, P < 0.001; r = -0.352, P < 0.001; r = -0.106, P = 0.002; respectively). It was slightly correlated only with foveal VD in DCP (r = -0.263, P < 0.001).

Partial correlation after adjusting for the effect of age and sex showed that in diabetic patient BCVA was not correlated with the foveal VD at SCP. There was a positive correlation between BCVA and WI-VD in SCP, DCP, and CC (r = 0.497, P < 0.001; r = 0.505, P < 0.001, r = 0.233, P = 0.0021; respectively), foveal VD in DCP, and CC (r = 0.171, P = 0.023; r = 0.354, P < 0.001; respectively) and parafoveal VD in SCP, DCP, and CC (r = 0.542, P < 0.001; r = 0.520, P < 0.001, r = 0.354, P < 0.001; respectively).

In linear regression analysis, it seems that in diabetic patients after adjusting for sex, age, duration of DM, presence of DME, blood pressure and FBS in the model, among foveal VD in 3 layers, only VD of CC was significantly correlated with BCVA (B: 0.40, P < 0.001). This test for WI-VD in all 3 layers showed correlation with VD of SCP (B: 0.26, P = 0.052). In the same model for parafoveal VD, only SCP showed the correlation (B: 0.37, P = 0.007). Applying two significant factors, considering the collinearity, the test showed that irrespective to the staging of the DR, superficial parafoveal VD (B: 0.35, P < 0.001) and VD of foveal CC (B: 0.26, P = 0.001) had the most effect on the BCVA. Adding the staging of the DR (B: -0.25, P = 0.007), showed similar results.

**DME**

In diabetes categories, we observed that all sub-fields had reduced VD in DME patients (all P < 0.05), except for foveal DCP VD (P = 0.716). These were far less than the amounts in the healthy control eyes (P < 0.05).

**Discussion**

This study, using the commercially available OCTA method, showed lower levels of VD in the macular region and all different sub-segments of SCP and DCP in diabetic patients compared to normal controls. With the exception of SCP foveal VD and DCP WI-VD, there was a steady and substantial reduction of VD in SCP and DCP from normal cases to NDR, NDR to NPDR, and NPDR to PDR groups. In the CC, the VD decreased significantly in NPDR stage and then rose marginally (not statistically significant) in the PDR stage but never reached the normal quantities. BCVA was specifically associated with parafoveal SCP and fovea CC VD. In DME patients, VD of all subfields with the exception of foveal DCP was reduced levels when compared with those without DME.
VD in retinal microvessels

The current study showed lower macular SCP and DCP VD levels in patients with diabetes relative to standard controls. Though methodologically different, previous studies have shown decreased parafoveal VD in DR without considering the DR staging [18, 36–39]. But our results for fovea and WI-VD in SCP and DCP are different from their results. Some qualitative or quantitative studies showed increased non-perfused areas from normal cases to the higher stages of the DR. [18, 35, 37, 38, 40]

After adjusting for hypertension, age, sex, and duration of DM our research showed that SCP and DCP VD in control eyes were higher than that in NDR group. Nesper et al. obtained the same result by adjusting to the hypertension. This finding supports VD modifications to be a potential biomarker of DR development.

The retinal vascular changes in diabetic patients including leukostasis and sluggish circulation [41–43], excitotoxic damage to glial cells [44], impaired neurovascular coupling and hypoperfusion [45] and endothelial
cell and pericytes loss, may trigger capillary occlusion and hypoxia at some point [39, 40]. This indicates that slower capillary blood flow below the lowest detection level for retinal vasculature in OCTA will result in undetectable vessels in the earlier phase of diabetic retinopathy as NDR.

Unlike our result, Nesper et al. showed the strongest correlation between the DCP vessels density and with DR severity [35]. Some qualitative studies have shown a more defective VD of DCP in NDR [20], unlike our quantitative findings of a consistent decrease of VD in both SCP and DCP. Previous reports have found microaneurysms to be present in a larger extent in DCP than in the superficial plexus [41, 42, 46, 47].

The contradictory findings found in the studies may be due to different imaging and measurement techniques, methodological differences, the various phases of the disease being investigated and the varying length of DM in the studies.

### Table 3: Vascular density in deep capillary network (DCN) in normal subjects and subtypes of diabetic retinopathy (N = 188)

| Groups | VD/Reference group | NL (40 eyes) Median (range) (%) | NDR (39eyes) Median (range) (%) | NPDR (64eyes) Median (range) (%) | PDR (33eyes) Median (range) (%) | P-value Kruskal-Wallis test |
|--------|--------------------|---------------------------------|---------------------------------|---------------------------------|---------------------------------|--------------------------|
| Whole image | Normal | 60.42 (47.00–64.55) | 58.05 (51.34–60.64) | 53.84 (41.34–60.67) | 51.24 (42.50–57.58) | < 0.001 |
| Fovea | Normal | 31.24 (20.30–42.32) | 27.72 (20.28–35.46) | 27.14 (15.48–40.50) | 24.94 (12.19–39.30) | < 0.001 |
| Superficial hemi | Normal | 63.45 (50.10–67.22) | 61.02 (52.40–64.34) | 56.14 (44.02–62.61) | 55.33 (41.91–61.91) | < 0.001 |
| Infrared hemi | Normal | 62.91 (49.50–66.68) | 60.51 (52.17–63.94) | 55.96 (40.75–63.36) | 54.07 (43.65–60.35) | < 0.001 |
| Temporal | Normal | 61.49 (48.50–64.82) | 61.04 (49.90–63.36) | 53.33 (32.32–61.74) | 53.31 (42.70–60.56) | < 0.001 |
| Superior | Normal | 64.82 (51.20–69.15) | 62.09 (53.30–65.34) | 58.62 (45.35–63.41) | 55.60 (42.14–60.70) | < 0.001 |
| Nasal | Normal | 62.56 (52.90–67.02) | 59.17 (52.40–62.53) | 55.64 (35.39–63.44) | 53.48 (39.92–59.70) | < 0.001 |
| Inferior | Normal | 63.76 (48.20–67.86) | 62.09 (52.23–65.59) | 56.76 (43.45–64.21) | 54.06 (44.73–61.92) | < 0.001 |

NDR: No diabetic retinopathy, NL: Normal, NPDR: Nonproliferative diabetic retinopathy, PDR: Proliferative diabetic retinopathy
*Mann-Whitney test
VD in CC

Some studies described diabetic choroidopathy in histology, electron microscopy, and FA and ICGA studies in the earlier stages of DR. [44, 48–51] Another study showed significant decrease in posterior ciliary arteries flow, using Color Doppler imaging, in patients with NPDR [20].

This study provides new information on CC alterations that occur in the course of DR stages from NDR to PDR. Consistent with our series, Forte et al. showed that in CC, the VD was not more different in type 1 and type 2 DM with NDR than controls in different subsegments [36].

The VD of CC was substantially reduced in NPDR compared to NDR in all subsegments and remained unchanged from NPDR to PDR (Fig. 2, Table 3). The severity of VD changes in SCP and DCP was apparently correlated with CC alteration (P > 0.05).

In vivo study on GK rats (a rat model of Type 2 diabetes) showed that, the VD of CC was reduced. This finding is consistent with the CC degeneration found in humans with T2D, either by post-mortem or OCT-angiography (OCTA) specimens [52–54].

Pericyte loss via vascular endothelial receptor 1 (VEGF R1), nitric oxide and reactive oxygen species identified in GK rats [55], triggered by hypoxia and/or inflammation [56], appears to be causing capillary degeneration and subsequent thinning during NPDR phase. This may contribute to a subsequent ischemic cascade involving CC, RPE and the outer retina propagating diabetic retinopathy.

Table 4 Choriocapillaris (CC) vascular density in different study groups (ANOVA) normal participants and diabetic patients. Post Hoc analysis of subfields choroidal thickness of the studied participants (N = 188)

| Groups                  | NL (40 eyes) | NDR (39eyes) | NPDR (64eyes) | PDR (33eyes) | P-value |
|-------------------------|--------------|--------------|---------------|--------------|---------|
|                         | (M ± SD)     | (M ± SD)     | (M ± SD)      | (M ± SD)     | ANOVA   |
| Whole image             | 69.82 ± 4.02 | 70.01 ± 2.90 | 67.06 ± 5.52  | 67.25 ± 4.66 | <0.001  |
| Normal                  | 0.949*       | <0.001*      | <0.001*       | <0.001*      | 0.950   |
| NDR                     | 0.996        | <0.001*      | 0.001*        | 0.001*       | 1.000   |
| NPDR                    | 0.980*       | <0.001*      | <0.001*       | 0.008*       | 0.798   |
| Fovea                   | 69.36 ± 4.54 | 69.52 ± 2.97 | 66.37 ± 5.89  | 67.05 ± 4.88 | <0.001  |
| Normal                  | 0.980*       | <0.001*      | <0.001*       | 0.002*       | 0.798   |
| NDR                     | 0.996        | <0.001*      | 0.001*        | 0.008*       | <0.001  |
| Superior hemi           | 69.92 ± 4.12 | 69.72 ± 2.94 | 66.74 ± 5.74  | 67.39 ± 4.25 | <0.001  |
| Normal                  | 0.914*       | <0.001*      | <0.001*       | 0.011*       | 0.437   |
| NDR                     | 0.904*       | <0.001*      | 0.004*        | 0.002*       | 0.798   |
| NPDR                    | 0.991*       | <0.001*      | <0.001*       | 0.002*       | 0.798   |
| Inferior hemi           | 69.70 ± 4.06 | 70.28 ± 3.12 | 67.16 ± 5.58  | 67.70 ± 5.18 | <0.001  |
| Normal                  | 0.993*       | <0.001*      | <0.001*       | 0.004*       | <0.001  |
| NDR                     | 0.993*       | <0.001*      | <0.001*       | 0.002*       | <0.001  |
| NPDR                    | 0.991*       | <0.001*      | <0.001*       | 0.002*       | <0.001  |
| Temporal                | 69.25 ± 5.79 | 69.66 ± 3.46 | 66.66 ± 6.23  | 67.88 ± 6.13 | <0.001  |
| Normal                  | 1.000*       | 0.0002*      | 0.020*        | 0.018*       | 0.964   |
| NDR                     | 0.980*       | 0.0002*      | 0.018*        | 0.964*       | 0.964   |
| NPDR                    | 0.991*       | 0.0002*      | 0.018*        | 0.964*       | 0.964   |
| Superior                | 69.19 ± 4.52 | 69.19 ± 3.15 | 66.66 ± 6.14  | 66.92 ± 4.91 | <0.001  |
| Normal                  | 0.984*       | <0.001*      | 0.001*        | 0.107*       | 0.488   |
| NDR                     | 0.880*       | <0.001*      | <0.001*       | 0.107*       | 0.488   |
| NPDR                    | 0.980*       | <0.001*      | <0.001*       | 0.107*       | 0.488   |
| Nasal                   | 69.34 ± 4.56 | 68.78 ± 3.63 | 65.84 ± 7.10  | 66.92 ± 4.33 | <0.001  |
| Normal                  | 0.785*       | <0.001*      | 0.025*        | 0.236*       | 0.382   |
| NDR                     | 0.785*       | <0.001*      | 0.001*        | 0.236*       | 0.382   |
| NPDR                    | 0.893*       | <0.001*      | <0.001*       | 0.236*       | 0.382   |
| Inferior                | 69.54 ± 4.32 | 70.90 ± 3.01 | 66.78 ± 6.38  | 67.33 ± 5.34 | <0.001  |
| Normal                  | 0.983*       | <0.001*      | 0.001*        | 0.001*       | 0.999   |
| NDR                     | 0.983*       | <0.001*      | <0.001*       | 0.001*       | 0.999   |
| NPDR                    | 0.983*       | <0.001*      | <0.001*       | 0.001*       | 0.999   |

NDR: No diabetic retinopathy, NL: Normal, NPDR: Nonproliferative diabetic retinopathy, PDR: Proliferative diabetic retinopathy
*Tuckey HSD test
Fig. 1 The radar diagram of choroidal thickness and choroidal volume map showing variation of the amounts taken by OCT in grid based vascular density of parafoveal area for comparison of diabetic patients in variant stages with normal eyes. *NDR: No diabetic retinopathy, NL: Normal, NPDR: Nonproliferative diabetic retinopathy, PDR: Proliferative diabetic retinopathy. ** I: Inferior, IN: inferonasal, N: Nasal, SN: Superonasal, S: superior, ST: Superotemporal, T: Temporal
Correlation of VD and visual acuity

After adjusting to several conflicting factors, this study found that the vision was directly associated with parafoveal VD at SCP and foveal VD at CC. The result was independent of the stage of the DR. Dissimilar to our result, previous researches showed that VD of DCP has more impact on the BCVA than SCP [57].

In diabetic patients, the reasons for reduced VA include macular ischemia [58], photoreceptor dysfunction [59] and accumulated subfoveal hard exudates [60]. There is a correlation between retinal nonperfusion
and the integrity of the photoreceptors [35, 61]. There was a strong correlation between external limiting membrane (ELM) and ellipsoid zone (EZ) integrity in DME patients [62].

Using oxygen-sensitive microelectrodes and oximetry, Linsenmeier and Zhang discovered that the choroidal and retinal circulation could supply the metabolic demand of photoreceptors in various proportions under both dark and light conditions [63]. It is important to know that, while most of the outer retina including photoreceptor is nourished with choroidal circulation, the contribution of the retinal circulation to photoreceptor metabolism is modest [63]. They calculated that the photoreceptor demand, met by the retinal circulation, was different in dark and lighted conditions (10 to 18% in darkness) [63].

Nesper et al. observed changes in capillary dropout areas in the DCP, indicating that DCP integrity is required for the metabolism of the photoreceptor [64].

Recently, by using spectral-domain OCT (SD-OCT), the concept of disorganization of retinal inner layers (DRIL) has been described to characterize retinal thinning with the loss of identifiable borders between retinal cellular layers in the background of capillary non-perfusion [65, 66]. Areas of DRIL were significantly correlated with disturbance of the photoreceptor (P = 0.035) relative to adjacent DRIL-free areas [66].

The decrease in VD in the SCP was significantly associated with the decrease in inner retinal thickness in DR, a notorious finding in mild DR, which may contribute to DRIL in the case of more serious capillary dropout [65]. Conversely, there was no association between the decrease in VD in the DCP and the thickness of the inner nuclear / outer plexiform layer (INL / OPL). The explanation for this disparity, also seen in healthy eyes, is unknown [66–70]. May be the SCP is much important for the inner layer nourishment. Moein et al. reported capillary flow impairment in the SCP on OCTA may be a predictor of visual acuity in patients with DRIL [71]. It is believed that the SCP, whose vascular supply is mainly arterial [72], has a standardized measurement, and is better to reveal ischemia data than the DCP [73].

**DME**

Macular edema can develop at any stage of DR. We found that in all subfields, the diabetic patients with DME had less VD than the patients without DME, except for the foveal VD at DCP. Tang et al. did not detect any statistically significant correlation between OCTA metrics and the existence of DME on SCP, incompatible with our findings [74]. Lee et al. have recently shown that DME eyes only have lower VD at DCP relative to DME-free eyes [75].

A limitation of this study was a modest age difference between control group and other groups, although in all stages of the analytic part of the study, the age matching was performed. Some baseline data as smoking and type of prescription have not been considered in the investigation, which may influence our conclusions. The small field of OCTA imaging and the use of both eyes in the analysis were other limitations. The automated programmed algorithm in the AngioVue system only segments two retinal capillary plexuses: the SCP and the DCP, and the middle capillary plexus (MCP) that is defined by swept source OCT could not be calculated.

Despite previous similar study [35], the advantage of present study was the inclusion of treatment naïve cases and an almost large sample size as well as the use of statistical models with adjustments for covariates.

**Conclusions**

In assumption, our study showed that VD in OCTA is correlated significantly and linearly with disease severity in eyes with DR. The results support VD of retinal vessels and CC to be a potential surrogate for DR before clinical signs development. We introduced parafoveal VD of SCP and foveal VD of CC as biomarkers to predict the VA loss in the diabetic patient. As a noninvasive

| Percentage of changes | NDR–NL/NL (%) | NPDR–NDR/NPDR (%) | PDR–NPDR/NPDR (%) |
|-----------------------|---------------|-------------------|-------------------|
| SCP Whole image       | −5.1          | −7.3              | −5.4              |
| SCP Fovea             | −9.9          | 3.0               | −11.4             |
| SCP Para-fovea        | −5.1          | −8.3              | −5.6              |
| DCP Whole image       | −3.8          | 1.2               | −12.9             |
| DCP Fovea             | −11.2         | −2.2              | −8.1              |
| DCP Para-fovea        | −3.3          | −7.1              | −4.1              |
| CC Whole image        | 0.3           | −4.1              | 0.3               |
| CC Fovea              | 0.1           | −4.5              | 1.0               |
| CC Para-fovea         | 0.1           | −4.5              | 1.1               |

NDR No diabetic retinopathy, NL Normal, NPDR Nonproliferative diabetic retinopathy, PDR Proliferative diabetic retinopathy
and rapidly acquired technique, OCTA may be a tool for early detection of microvascular abnormalities in the retina and choroid, elucidating the pathogenesis of retinopathy, and treatment response monitoring in patients with diabetes.

**Abbreviations**
- CC: choriocapillaris; CMT: central macular thickness; DR: diabetic retinopathy; DME: diabetic macular edema; MCP: middle capillary plexus; NPDR: nonproliferative diabetic retinopathy; OCTA: optical coherence tomography angiography; PDR: proliferative diabetic retinopathy; SD OCTA: spectral domain optical coherence tomography angiography; DCP: deep capillary plexus; SCP: superficial capillary plexus; VD: vascular density; VEGF: vascular endothelial growth factor; Wi: whole image

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**Authors’ contributions**
AB, SB, AG, and FG collected the data. FG and SS analyzed and interpreted the data. FG and KF were major contributors in writing the manuscript. All authors read, commented and approved the final manuscript.

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**Availability of data and materials**
The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

**Ethics approval and consent to participate**
Ethics approval was obtained from the Tehran University of Medical Sciences Institutional Review Boards and written consent form was taken from all participants.

**Consent for publication**
Not applicable.

**Competing interests**
The authors declare that they have no competing interests.

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**References**
1. Rama M, Pradeepa R. Diabetic retinopathy: an Indian perspective. Indian J Med Res. 2007;125:297.
2. Zeng J, Chen B. Epigenetic mechanisms in the pathogenesis of diabetic retinopathy. Ophthalmologica. 2016;232:1–9.
3. ETDGR. Fundus photographic risk factors for progression of diabetic retinopathy: ETDRS report number 12. Ophthalmology. 1991;98:823–833.
4. Hammes H-P. Pericytes and the pathogenesis of diabetic retinopathy. Horm Metab Res. 2005;37:39–43.
5. Makita S, Hong Y, Yamani M, Yatagai T, Yaruno Y. Optical coherence angiography. Opt Express. 2006;14:7821–40.
6. Durham JT, Herman IM. Microvascular modifications in diabetic retinopathy. Curr Diab Rep. 2011;11:253–64.
7. Antontetti DA, Klein R, Gardner TW. Diabetic retinopathy. N Engl J Med. 2012;366:1227–39.
8. Garner M. Histopathology of diabetic retinopathy in man. Eye. 1993;7:250–3.
9. Yannuzzi LA, Rohrer KT, Tindel LJ, Sobel RS, Costanza MA, Shields W, Zang E. Fluorescein angiography complication survey. Ophthalmology. 1989;96:611–7.
10. Fingler J, Schwartz D, Yang CH, Fraser SE. Mobility and transverse flow visualization using phase variance contrast with spectral domain optical coherence tomography. Opt Express. 2007;15:12636–53.
11. Jia Y, Tan O, Tokayer J, Wang Y, Liu J, Kaur MS, Subhash H, Fujimoto JG, Hornegger J, Huang D. Split-spectrum amplitude decorrelation angiography with optical coherence tomography. Opt Express. 2012;20:7410–25.
12. Mariampillai A, Standidge B, Montyak EH, Khurana M, Munce NR, Leung MK, Jiang J, Jiang J, Cable A, Wilson BC, Vittin IA, Yang YK. Speckle variance detection of microvasculature using swept-source optical coherence tomography. Opt Lett. 2008;33:1530–2.
13. Matsunaga D, Yi J, Puliafito C, Kashani AH. OCT angiography in healthy human subjects. Ophthalmic Surg Lasers Imaging Retina. 2014;45:510–5.
14. Spaide RF, Klancnik JM, Cooney MJ. Retinal vascular layers imaged by fluorescein angiography and optical coherence tomography angiography. JAMA ophthalmology. 2015;133:45–50.
15. Huang Y, Zhang Q, Thorell MR, An L, Dubin MK, Laron M, Sharma U, Gregori G, Rosenfeld PJ, Wang RK. Swept-source OCT angiography of the retinal vasculature using intensity differentiation-based optical microangiography algorithms. Ophthalmic Surg Lasers Imaging Retina. 2014;45:382–9.
16. Gao SS, Jia Y, Zhang M, Su JP, Liu G, Hwang TS, Bailey ST, Huang D. Optical coherence tomography angiography. Invest Ophthalmol Vis Sc. 2016;57: OCT27–36.
17. Freiberg FJ, Pflaum W, Mors J, Wirth MA, Becker MO, Michels S. Optical coherence tomography angiography of the foveal avascular zone in diabetic retinopathy. Graefes Arch Clin Exp Ophthalmol. 2016;254:1051–8.
18. Ishibazawa A, Nagaoka T, Takahashi A, Omae T, Tani S, Sagawa K, Yokota H, Yoshida A. Optical coherence tomography angiography in diabetic retinopathy: a prospective pilot study. Am J Ophthalmol. 2015;160:35–44.
19. Couturier A, Mane V, Bonnin S, Erginay A, Massin P, Gaudric A, Tadayoni R. Capillary plexus anomalies in diabetic retinopathy on optical coherence tomography angiography. Retina. 2015;35:284–91.
20. Dimitrova G, Chihara E, Takahashi H, Amano H, Okazaki K. Quantitative retinal optical coherence tomography angiography in patients with diabetes without diabetic retinopathy. Invest Ophthalmol Vis Sc. 2017;58:190–6.
21. Al-Shiekh M, Akih M, Pflaum M, Sadda SR. Swept-source OCT angiography imaging of the Foveal avascular zone and macular capillary network density in diabetic retinopathy. Invest Ophthalmol Vis Sc. 2016;57:3907–13.
22. Agemy SA, Sciprierna N, Shah CM, Chui T, Garcia PM, Lee JG, Gentile RC, Hsiao YS, Zhou Q, Ko T, Rosen RB. RETINAL VASCULAR PERFUSION DENSITY MAPPING USING OPTICAL COHERENCE TOMOGRAPHY ANGIOGRAPHY IN NORMALS AND DIABETIC RETINOPATHY PATIENTS. Retina. 2015;35:2533–63.
23. Kim AY, Chu Z, Shahidzadeh A, Wang RK, Puliafito CA, Kashani AH. Quantifying microvascular density and morphology in diabetic retinopathy using spectral-domain optical coherence tomography angiography. Invest Ophthalmol Vis Sc. 2016;57:362–70.
24. Arend O, Wolf S, Jung F, Bertam B, Pöstgörs H, Toonen H, Reim M. Retinal microcirculation in patients with diabetes mellitus. dynamic and morphological analysis of perifoveal capillary network. Br J Ophthalmol. 1991;75:514–8.
25. Rimmer T, F felon TJ, Kohner EM. Long-term follow-up of retinal blood flow in diabetes using the blue light entoptic phenomenon. Br J Ophthalmol. 1989(73):1–5.
26. Dimitrova G, Kato S, Yamashita H, Tamaki Y, Nagahara M, Fukushima H, Kitano S. Relation between retinobulbar circulation and progression of diabetic retinopathy. Br J Ophthalmol. 2003;87:622–5.
27. Grunwald JE, Riva CE, Sinclair SH, Baine J, Brucker AJ. Total retinal volumetric blood flow rate in diabetic patients with poor glycaemic control. Invest Ophthalmol Vis Sc. 1992;33:356–63.
28. Burgansky-Eliash Z, Barak A, Barash H, Nelson DA, Pupko O, Lowenstein A, Grinvald A, Rubinstein A. Increased retinal blood flow velocity in patients with early diabetes mellitus. Retina. 2012;32:1126–34.
29. Pemp B, Poliska E, Garhofer G, Mayerle-Eder M, Kautzky-Willer A, Schmetterer L. Retinal blood flow in type 1 diabetic patients with no or mild diabetic retinopathy during euglycemic clamp. Diabetes Care. 2010;33:2038–42.
30. Patel V, Rassam S, Newson R, Wiek J, Kohner E. Retinal blood flow in diabetic retinopathy. BMJ. 1992;305:678–83.
31. Fluorescein angiographic risk factors for progression of diabetic retinopathy. ETDRS report number 13. Early Treatment Diabetic Retinopathy Study Research Group. Ophthalmology. 1991;98(Suppl):834–840.
32. Savastano MC, Lumbroso B, Rispoli M. In vivo characterization of retinal microaneurysms using optical coherence tomography angiography. Retina. 2015;35(3):2196–203.
33. Spadé RF, Klanicnik JM Jr, Cooney MJ. Retinal vascular layers imaged by fluorescein angiography and optical coherence tomography angiography. JAMA Ophthalmol. 2015 Jan;133(1):45–50.
34. Shahlaee A, Pefkianaki M, Hsu J, Ho AC. Measurement of foveal avascular zone dimensions and its reliability in healthy eyes using optical coherence tomography angiography. Am J Ophthalmol. 2016;161:50–55.e51.
35. Nesper PL, Roberts PK, Onishi AC, Choi H, Liu L, Jampol LM, Fawzi AA. Quantifying microvascular abnormalities with increasing severity of diabetic retinopathy using optical coherence tomography angiography. Invest Ophthalmol Vis Sci. 2017;58:BIO307–15.
36. Forte R, Haulani H, Jürgens I. QUANTITATIVE AND QUALITATIVE ANALYSIS OF THE THREE CAPILLARY PLEXUSES AND ChORIOCAPILLARIS IN PATIENTS WITH TYPE 1 AND TYPE 2 DIABETES MELLITUS WITHOUT CLINICAL SIGNS OF DIABETIC RETINOPATHY: a prospective pilot study. Retina. 2020;40:333–44.
37. Hwang TS, Gao SS, Liu L, Lauker AK, Bailey ST, Flaxel CJ, Wilson DJ, Huang D, Jia Y. Automated quantification of capillary nonperfusion using optical coherence tomography angiography in diabetic retinopathy. JAMA Ophthalmol. 2016;134:637–73.
38. Salz DA, de Caro TE, Adhi M, Moulit E, Choi W, Baumrul CR, Witkin AJ, Duker JS, Fujimoto JG, Waheed NK. Select features of diabetic retinopathy on swept-source optical coherence tomographic angiography compared with fluorescein angiography and normal eyes. JAMA Ophthalmol. 2016;134:644–50.
39. Ciloglu EUF, Sukgen EA, Kokjul Y. Evaluation of Foveal avascular zone and capillary plexus in diabetic patients by optical coherence tomography angiography. Korean J Ophthalmol. 2019;33:359–65.
40. de Caro TE, Bordi Filho MA, Adhi M, Branchini L, Salz DA, et al. Detection of microvascular changes in eyes of patients with diabetes but not clinical diabetic retinopathy using optical coherence tomography angiography. Retina. 2015;35:2364–70.
41. Hasegawa N, Nozaki M, Takase N, Yoshida M, Ogura Y. New insights into microaneurysms in the deep capillary plexus detected by optical coherence tomography angiography in diabetic macular edema. Invest Ophthalmol Vis Sci. 2016;57:OC348–55.
42. Ishibazawa A, Nishijima K, Kita M, Oh H, Tissukawa A, Yoshimura N. Association between foveal photoreceptor status and visual acuity after resolution of diabetic macular edema. Retina. 2018;38:2277–45.
43. Jahn CE, VSK T, Richter J, Boller J, Kron M. Improvement of visual acuity in eyes with diabetic macular edema after treatment with para plana vitrectomy. Graefes Arch Clin Exp Ophthalmol. 2004;242:325–30.
44. Otani T, Kishi S. Tomographic findings of foveal hard exudates in diabetic macular edema. Am J Ophthalmol. 2001;131:50–4.
45. Catnici F, Nesper PL, Fawzi AA. Deep retinal capillary nonperfusion is associated with photoreceptor disruption in diabetic macular ischemia. Am J Ophthalmol. 2016;168:129–38.
46. Shen Y, Liu K, Xu X. Correlation between visual function and photoreceptor integrity in diabetic macular ischemia: spectral-domain optical coherence tomography. Curr Eye Res. 2016;41:391–9.
47. Linsenmeier RA, Zhang HF. Retinal oxygen: from animals to humans. Prog Retin Eye Res. 2017;58:115–51.
48. Nesper PL, Scarchini F, Fawzi AA. Adaptive optics reveals photoreceptor abnormalities in diabetic macular ischemia. PLoS One. 2017;12:e0169926.
49. Sun JK, Lin MM, Lammier J, Prager S, Sarangi R, Silva PS, Aiello LP. Disorganization of the retinal inner layer-ers as a predictor of visual acuity in eyes with center-involved diabetic macular edema. JAMA Ophthalmol. 2014;132:1309–16.
50. Onishi AC, Ashraf M, Soetinko BT, Fawzi AA. Multilevel ischemia in disorganization of the retinal inner layers on projection-resolved optical coherence tomography angiography. Retina. 2019;39(8):1588–94.
51. Lavin C, Couturier A, Erginay A, Dupas B, Tadayoni R, Gaudric A. Reduced vessel density in the superficial and deep plexuses in diabetic retinopathy is associated with structural changes in corresponding retinal layers. PLoS One. 2019;14:e0216164.
52. Cao D, Yang D, Huang Z, Zeng Y, Wang J, Hu Y, Zhang L. Optical coherence tomography angiography discerns preclinical diabetic retinopathy in eyes of patients with type 2 diabetes without clinical diabetic retinopathy. Acta Diabetol. 2018;55:669–77.
53. Scarcini F, Jampol LM, Linsenmeier RA, Fawzi AA. Association of Diabetic Macular Nonperfusion with Outer Retinal Disruption on optical coherence tomography. JAMA Ophthalmol. 2015;133:1306–44.
54. Joklth KA, Sesi CA, de Castro VM, Davila JR, Anand R, Khan SM, Fatibn M, Jackson JR, Johnson CA, Gardner TW. Disorganization of retinal inner layers (DRII) and Neuroretinal dysfunction in early diabetic retinopathy. Invest Ophthalmol Vis Sci. 2018;59:5481–6.
55. Moein HR, Novais EA, Rebhun CB, Cole ED, Louzada RN, Witkin AJ, Baumal CR, Duker JS, Waheed NK. Optical coherence tomography angiography to detect macular capillary ischemia in patients with inner retinal changes after resolved diabetic macular edema. Retina. 2018;38:2277–84.
56. Gass JD. Stereoscopic atlas of macular diseases. St.Louis: Mosby; 1997.
57. Durbin MK, An L, Shemonski ND, Soares M, Santos T, Lopes M, Lopes M, Neves C, Cunha-Vaz J. Quantification of retinal microvascular density in optical coherence Tomo- graphie angiography images in diabetic retinopathy. JAMA Ophthalmol. 2017;135:370–5.
58. Tong FY, Ng DS, Lam A, Lok F, Wong R, Chan C, Mohammed S, Feng A, Lok J, Tso L, Tafir, M, Wong TY, Tham CC, Cheung CY. Determinants of
quantitative optical coherence tomography angiography metrics in patients with diabetes. Sci Rep. 2017;31:2575.

75. Lee J, Moon BG, Cho AR, Yoon YH. Optical coherence tomography angiography of DME and its association with anti-VEGF treatment response. Ophthalmology. 2016;123:2368–75.

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