Case Report

Skull base osteomyelitis by *Pandoraea apista*: An unusual pathogen at unusual location – A case report

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### ABSTRACT

**Background:** *Pandoraea apista* is predominantly recovered from the respiratory tract of patients with cystic fibrosis (CF). Authors report first case of central nervous system infection by *P. apista* in the form of skull base osteomyelitis.

**Case Description:** A 67-year-old male presented with complaints of earache and hearing deficit for few months. The radiology was suggestive of skull base osteomyelitis and polypoidal soft tissue extending from the middle cranial fossa to the infratemporal fossa. The sample from the targeted area revealed *P. apista* on matrix-assisted laser desorption ionization-time-of-flight mass spectrometry. With adequate antibiotic therapy, there was clinicoradiologic improvement.

*P. apista* is an infection exclusively seen in pulmonary infection in patients with CF. We identified its intracranial involvement in a patient for the 1st time in the literature. The serendipitous diagnosis needs evaluation on specific PCR and matrix-assisted laser desorption spectrometry. The treatment with antibiotics provides a definite cure.

**Conclusion:** We report a rare opportunistic infection with central nervous system involvement which can be cured by accurate diagnosis and appropriate antibiotic treatment.

**Keywords:** Cystic fibrosis, Infection, Matrix-assisted laser desorption ionization-time-of-flight mass spectrometry, *Pandoraea apista*, PCR

### INTRODUCTION

*Pandoraea* genus is a relatively new genus first described by Coenye *et al.* in 2000. The name *Pandoraea* was given to this genus to refer to Pandora’s box (from Greek mythology which was considered as the origin of diseases to the mankind), because of surprising diversity amongst the members of this genus. The authors report first ever case of *Pandoraea* pathogen with a central nervous system involvement.

### CASE REPORT

A 67-year-old male presented with complaint of severe pain in the left ear along with decreased hearing for few months. Examination revealed a bulge in the external auditory canal making the...
tympanic membrane visualization impossible. The patient had nonserviceable conductive and sensorineural hearing loss in the left ear. His systemic and other neurological examination was nonremarkable. CT scan revealed poorly defined heterogeneous enhancing soft-tissue mass involving external auditory canal, pterygoid space, masticator space, nasopharynx, and infratemporal fossa. CT scan of skull base revealed polypoidal soft tissue occluding the left external auditory canal [Figure 1a] having contiguous extension inferiorly and medially along nasopharynx and infratemporal fossa. The muscles of mastication showed heterogeneous enhancement with diffuse increase in bulk [Figure 1b]. There was associated erosion of the bones forming the base of skull [Figure 1c and d].

The positron emission tomography scan neither revealed any other pathology nor showed any metabolic activity. Considering clinical and radiologic considerations, the possibilities of skull base osteomyelitis or a neoplastic lesion were considered. Biopsy from nasopharynx was inconclusive while CT-guided FNAC from skull base was sent for bacterial culture. The sample was inoculated on blood agar and MacConkey agar and incubated at 37°C overnight under aerobic conditions as a part of routine processing for aerobic isolates. The culture revealed monomicrobial growth of a nonlactose fermenting colony on MacConkey agar which was identified as *Pandoraea apista* on matrix-assisted laser desorption ionization-time-of-flight mass spectrometry (MALDI-TOF, MS, Bruker Daltonik GmbH, Germany) using the MALDI BioTyper database. The patient was then kept on ciprofloxacin 750 mg twice a day for 1-month duration. This led to relief from pain and subsequent imaging showed gradual resolution of lesion over many moths with disappearance of the bulge of the ear canal. Follow-up CT and MRI performed after 1 year showed near-complete resolution of the lesion [Figure 2].

**DISCUSSION**

At present, *Pandoraea* genus consists of 10 exclusive human clinical species. These include *P. apista, Pandoraea pulmonicola, Pandoraea sputorum, Pandoraea anapnoica, Pandoraea anhela, Pandoraea bronchicola, Pandoraea captiosa, Pandoraea morbifera, Pandoraea nosoerga*, and *Pandoraea pneumonica*. Nonclinical species are mainly isolated from soil, water, animal feces, and powdered milk. These are environmental organisms found in water, soil, and on plants. Because of their ability to survive in aqueous environments, these organisms have become particularly problematic as opportunistic nosocomial pathogens in health-care settings.

**Pathogenesis and virulence factor**

Human *Pandoraea* species are primarily isolated from respiratory secretions of patients of cystic fibrosis (CF).
Seldom, from non-CF patients, has also been reported [Table 1]. The growth of organism from blood and urine samples indicates their potential to invade lung epithelial cells. However, the ability to invade lung epithelial cells is not common and is unlikely to be the major virulence factor of these strains.[2]

All *Pandoraea* species stimulate release of pro-inflammatory cytokines, particularly IL-6 and IL-8. Upregulation of these two interleukins is the primary virulence factor which is largely responsible for the chronic lung inflammation. Chronic colonization of *Pandoraea* further leads to *in vivo* point mutations and strain evolution overtime.[2,9]

### Table 1: Reports of *Pandoraea* infection with antibiotic susceptibility in non-CF patients.

| Reference | Species | Strain | Country | Source | Age/Sex | Underlying condition/diagnosis | Sensitive to | Resistant to |
|-----------|---------|--------|---------|--------|---------|--------------------------------|--------------|-------------|
| Coenye *et al.*,[3] 2000 | *P. norimbergensis* | LMG 13019 | Belgium | Blood | NA | NA | NA | NA |
| Daneshvar *et al.*,[6] 2001 | *P. norimbergensis* | LMG 16603 | Sweden | BAL | NA | NA | Imipenem, Ampicillin, amoxicillin-clavulanate, cefotaxime, cefazolin, meropenem, piperacillin, gentamycin | |
| | *P. apista* | G3307 | USA | Blood | 66 yr/F | COPD | Ampicillin, Amikacin, tetracycline, ciprofloxacin |
| | *P. apista* | G3308 | USA | BAL | 75 yr/F | NA | Imipenem |
| | *P. pnomenusa* | G5056 | USA | Blood | 46 yr/M | Septicaemia | Imipenem, tetracycline |
| | *P. pnomenusa* | G7835 | USA | Blood | 76 yr/M | Bacteraemia | Imipenem | |

(Contd...)
| Reference       | Species          | Strain | Country | Source                | Age/Sex | Underlying condition/diagnosis                          | Sensitive to                                                                 | Resistant to                                                                 |
|-----------------|------------------|--------|---------|-----------------------|---------|----------------------------------------------------------|------------------------------------------------------------------------------|--------------------------------------------------------------------------------|
| Stryjewski et al.,[14] 2003 | *P. pnomenusa*   | G8107  | USA     | BLOOD                 | 49/M    | NA                                                       | Imipenem                                                                    | Ampicillin, amoxicillin-clavulanate, amikacin, gentamycin, cefotaxime, cefazolin, meropenem, piperacillin, ciprofloxacin |
| Monzón et al.,[15] 2018 | *P. sputorum*   | NA     | Spain   | Blood and BAL         | 30 yr/M | lung transplant for pulmonary sarcoidosis                | Imipenem                                                                    | Ampicillin, aminoglycosides, cefazidine, ciprofloxacin, piperacillin-tazobactam, cotrimoxazole |
| Monzón et al.,[15] 2018 | *P. sputorum*   | G5084  | USA     | Maxillary sinus       | NA/F    | NA                                                       | Imipenem                                                                    | Ampicillin, aminoglycosides, aminoglycosides, aztreonam, meropenem, piperacillin, ciprofloxacin |
| Falces-Romero et al.,[7] 2016 | *P. pnomenusa*   | G9805  | USA     | Sputum                | 71 yr/M | Pneumonia                                                | None                                                                        | Ampicillin, minocycline, and Imipenem                                                                 |
| Monzón et al.,[15] 2018 | *P. sputorum*   | NA     | Spain   | Blood                 | 10 months/M | PRE-B ALL                                        | Imipenem                                                                    | Minocycline and Imipenem                                                                 |
| Gao et al.,[9] 2018 | *Pandoraea species* | NA     | China   | Blood                 | 23 days/M | Neonatal jaundice                                        | Imipenem, cotrimoxazole, ampicillin-sulbactam, tetracycline                  | Amikacin, gentamycin, aminoglycosides, aztreonam, meropenem, piperacillin, ciprofloxacin |

*Table 1: (Continued)*
In CF, lung infection is polymicrobial in nature with *Pseudomonas aeruginosa* as the most commonly colonizing pathogen. *P. aeruginosa* and *Burkholderia multivorans* have growth suppressing effects on *Pandoraea* species, that is why *Pandoraea* is more virulent in the absence of co-colonization with these pathogens and is more likely to cause bacteremia in non-CF patients than CF patients. Therefore, younger patients who had higher pathogen diversity showed lower incidence of permanent lung damage than older patients. Plasmids of nonclinical strain contain more virulence genes than clinical strains and if transmitted into clinical environment they may prove pathogenic. Antibiotic resistance genes were present in plasmids of nonclinical strains only and aid them in acclimatizing to their environment.

**Diagnosis**

Routine diagnostic tests are not sufficient for the diagnosis of *Pandoraea* species and molecular studies are required for their identification. 16S rRNA, *gyr B, gltB* gene sequencing, and genus-specific PCR all allow identification of *Pandoraea* species from its closely associated genera, but high level of identity among its species requires further analysis to identify individual members of *Pandoraea* species.

Recently, the use of MALDI-TOF, MS has shown good results. MALDI-TOF MS designates an organic matrix compound, which helps in the soft desorption and ionization of highly abundant bacterial or fungal proteins using laser energy. Subsequently, accurate identification relies on the availability of well-curated, extensive databases preferably encompassing the full spectrum of microbes encountered in clinical specimens.

With its use, early identification of the bacteria is possible. The use of MALDI-TOF MS for the routine identification of most microbes has largely replaced conventional biochemical and phenotypic tests in clinical laboratories, since it is 1.45 days faster, less expensive, and does not involve a labor-intensive protocol. Imaging may help in mapping out the disease process, however, as this is the 1st time imaging of this entity has ever been presented, it is near impossible to detail the specifics of the lesions on CT or MRI. Nevertheless, in the index case, the skull base lesion with its characteristic heterogeneous enhancement amidst the native structures around the nasopharynx and bone destruction pattern resembled an inflammatory process more than a neoplastic one.

**Antibiotic susceptibility**

Multidrug resistance is because of enzyme production and efflux pumps. Oxacillinase-62 plays an important role against imipenem resistance. All show resistance to meropenem, aminoglycosides, quinolones, and most β-lactam antibiotics while they are susceptible to imipenem, tetracycline, and trimethoprim-sulfamethoxazole. Its greatly variable antibiotic susceptibility necessitates accurate diagnosis and antibiotic sensitivity for its treatment.

There is no report so far for the intracranial invasion of *P. apista*. Authors present the very first case of this pathogen showing involvement of central nervous system. We are not able to identify the source of this pathogen in the patient. There is no case reported so far from our region. Surprisingly, the patient has no host factor to suggest any predicament or susceptibility for this extremely rare pathogen. The identification of the pathogen in our case is a serendipity.
and we could manage the case only after going through the available literature.

**CONCLUSION**

*Pandoraea* is responsible for opportunistic pulmonary infections predominantly affecting CF patients. Although it can also affect non-CF patients, in whom it may be more virulent because of absence of polymicrobial environment. It requires molecular analysis for its accurate diagnosis. Because of its multidrug resistance, antibiotic sensitivity is necessary for the treatment. This pathogen may present with primary involvement of central nervous system and a high index of suspicion is necessary for its diagnosis and management.

**Ethical approval**

All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki Declaration and its later amendments or comparable ethical standards.

**Declaration of patient consent**

The authors certify that they have obtained all appropriate patient consent.

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**Conflicts of interest**

There are no conflicts of interest.

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