Selenium is an essential immunonutrient which holds the human’s metabolic activity with its chemical bonds. The organic forms of selenium naturally present in human body are selenocysteine and selenoproteins. These forms have a unique way of synthesis and translational coding. Selenoproteins act as antioxidant warriors for thyroid regulation, male-fertility enhancement, and anti-inflammatory actions. They also participate indirectly in the mechanism of wound healing as oxidative stress reducers. Glutathione peroxidase (GPX) is the major selenoprotein present in the human body, which assists in the control of excessive production of free radical at the site of inflammation. Other than GPX, other selenoproteins include selenoprotein-S that regulates the inflammatory cytokines and selenoprotein-P that serves as an inducer of homeostasis. Previously, reports were mainly focused on the cellular and molecular mechanism of wound healing with reference to various animal models and cell lines. In this review, the role of selenium and its possible routes in translational decoding of selenocysteine, synthesis of selenoproteins, systemic action of selenoproteins and their indirect assimilation in the process of wound healing are explained in detail. Some of the selenium containing compounds which can acts as cancer preventive and therapeutics are also discussed. These compounds directly or indirectly exhibit antioxidant properties which can sustain the intracellular redox status and these activities protect the healthy cells from reactive oxygen species induced oxidative damage. Although the review covers the importance of selenium/selenoproteins in wound healing process, still some unresolved mystery persists which may be resolved in near future.
Keywords Selenium · Selenoproteins · Wound healing · Selenium drugs · Gpxs · SEPP1 · SEPS1 · ROS

Introduction

Selenium is an essential trace element which possesses the symbol Se with atomic number 34 in the periodic table. Selenium was discovered by Swedish chemist Jons Jacob Berzelius in the year 1817 and it was named after the ancient Greek word ‘Selene’ which refers to Moon. Selenium is a reactive nonmetal or metalloid located in P Block, Period IV and Group 16 of the Mendeleev’s periodic table of elements. Selenium originates from Sulfide ore which partially replaces the Sulphur in certain reactions since it possesses similar physical and chemical properties of Sulphur (Meija et al. 2016). Selenium is an amorphous brick red powder and when it reaches its melting point, it tends to turn into its black vitreous form. It has six different isotopes with varied stabilization states, half-life and mode of decay. Selenium exists in two different forms, namely organic and inorganic. Organic forms of selenium are present as selenocysteine and selenomethionine in human body. Inorganic forms such as selenite and selenate get accumulated in plants through soil. Selenium has the ability to combine with other minerals and elements of Sulphide, Copper, Silver, Nickel and Lead. Selenium holds the 67th rank for the most abundant element on the Earth’s crust (USEPA 2014).

Selenium is the most required micronutrient for human beings and animals. Inorganic selenium is mostly accumulated in plants via Sulphur assimilation pathway, whereas animals and humans consume these sources later as vegetables, meats and dietary supplements. The level of Se concentration in plants differs by the type of soil, level of accumulation capacity, pH, salinity, organic matter, redox reactions, and type of plant species. Excessive accumulation of selenium in plants leads to toxicity (Drahoňovský et al. 2014). The selenium content in soil differs globally and when it is compared with other nations, the Indian and European soils possess less selenium (Rayman 2008). Daily intake of selenium is highly recommended for maintaining the natural metabolism and homeostasis in the human body.
The intake dosage of selenium is determined as 55 μg and 70 μg per day for adult males and females, respectively. Selenium consumption is mostly dependent on the food contents and dietary supplements (Navarro-Alarcon and Cabrera-Vique 2008).

Seafoods and meats are first to be placed in the selenium rich foods. The other selenium rich foods include muscle meats, cereals, grains and dairy products. The amount of selenium in drinking water is not nutritionally significant in most geographic regions. As a result, selenium concentrations in water cannot be considered as selenium supplementation. However, selenium concentration in soil has a reduced effect in animal products than in plant-based foods because animals maintain predictable tissue concentrations of selenium through homeostatic mechanisms (Chun et al. 2009).

Selenium is an essential component for many enzymes’ activities. It has been widely experimented for its crucial role as an antioxidant and in immunological cycles, where it can be incorporated non-specifically with the amino acid Methionine in body proteins. Skeletal muscle is the major site of selenium storage, accounting for approximately 28–46% of the total selenium pool (Pascual and Aranda 2013). Selenium has several promising roles in the human body such as antioxidant, anti-inflammatory, anti-mutagenic, anti-carcinogenic, antiviral, antibacterial and antifungal effects (Rayman 2000; Cihalova et al. 2015; Shakibaie et al. 2015; Malhotra et al. 2016; Peng et al. 2016; Stolzoff and Webster 2016; Maiyo and Singh 2017).

Selenium is the integral compound of selenoproteins in the human body; the selenoprotein family consists of 25 eukaryotic genes where 25 are human genes and 24 are mouse genes. All these proteins have selenocysteine residue in their primary predefined structure (Kryukov et al. 2003). Selenium has a specific protein synthesis pathway where its codes for the 21st amino acid called UGA Codon; this process is called as Translational Decoding (Bubenik et al. 2014). These selenoproteins are responsible for function and regulation of thyroid hormones, regulators of growth, development and differentiation, inhibitors of non-specific immune response, neutralisers of inflammatory, chemotactic and phagocytic responses like anti-male sterility, reproduction and oxidative stress influencer (Schomburg 2012; Dentice et al. 2013; Pascual and Aranda 2013; Effraimidis and Wiersinga 2014; Zubair et al. 2015; Chan et al. 2016). Role of selenium in human body is important and crucial in stabilising and neutralising the body metabolism. During wound healing process, certain selenoproteins like GPX-1, GPX-4, selenoprotein S and selenoprotein P combine to perform various reactions such as antioxidant activities, inhibition of inflammatory cytokines and elimination of Peroxynitrate (a super radical ion) in inflammatory phase (Lei et al. 2009; Cox et al. 2013; Talbi et al. 2019).

Wound healing is a complex process with many cascades of events involving different phases, such as (1) haemostasis or blood clotting, (2) inflammation, (3) proliferation and (4) remodelling or maturation. A wound is defined as disruption or damage to the normal cells which changes the anatomical structure and function. Wounds can be classified according to their shape, depth and infection (Coupand 1992). The Centre for disease control and prevention (CDC) categorized the surgical wounds into three types by clean, contaminated, and infected. Primarily the wound is classified into three types (1) acute (2) chronic and (3) complicated. Acute wounds heal themselves in accordance with proper duration and follows orderly healing pathway; the duration of healing normally ranges either from 5 to 10 days nor within a month. Chronic wounds are wounds that cannot be restored normally by healing pathway; they have irregular duration and prolonged process like infection, necrosis, hypoxia, and higher amount of inflammatory cytokine release. The combination of tissue disintegration and infection is termed to be complicated wound. For example: tumor cell in a complicated region such as brain, oesophagus, uterus, etc. (Lazarus et al. 1994; Bischoff et al. 1999).

Skin is the first line of defense which covers the entire body, safeguarding it from the external environment. The skin protects the body from pathogens, harmful rays, foreign invaders, and oxidative stress. Apart from that, it also possesses functions like maintaining the body temperature, immunological reactions, regulations of specific metabolic reactions, and many signalling activities. An injury starts with tissue disintegration followed by bleeding which helps to flush out the blood containing microbes and foreign particles. Immediately the clotting factors trigger the phases of coagulation and haemostasis which leads to platelet aggregation and fibrinogen formation (Martin et al. 1997; Sen 2009). The inflammatory phase which deals with cellular and vascular responses usually starts within few minutes of the injury and lasts up to 24 h. In migration phase, the epithelial cells and fibroblasts migrate towards the injured area to replace the disintegrated, damaged cells and tissues. On day 3, the proliferation phase starts with granulation for tissue formation, synthesis of lymphatic vessels, synthesis of collagen by fibroblasts, and maximum formation of blood vessels and tissues. The final phase of wound healing is said to be maturation or remodelling which involves the regeneration of cellular connective tissues and strengthening of the new epithelium. This prolonged process may take months to 2 years to recover fully (Velnar et al. 2009). As discussed above, certain selenoproteins act as antioxidants as well as inducers during specific phases of wound healing process. In this review, we try to elucidate some of the possible routes in which selenoproteins participate in wound healing mechanism.
Selenium uptake and accumulation by plants

Selenium is one of the important elements needed for plant growth and development. The major physiological role of selenium in plant is to prevent oxidative stress, phyto remediation, biofortification, etc. Plants are classified into two major types based on the level of selenium accumulation. Certain plants grown in selenium enriched soil tend to accumulate high levels of the element; they are called as hyper accumulators or Se-tolerant plants. The plants that do not accumulate selenium are known as hypo accumulators or Se-sensitive plants (Zhu et al. 2009; Yu and Gu 2013; Alfthan et al. 2015; El-Ramady et al. 2015). The major plant species that are hyper accumulators come under the families of Brassicaceae, Fabaceae and Asteraceae (Läuchli 1993). These hyper accumulator plants store organic forms of selenium such as methyl-SeCysteine and selenocysteine, whereas hypo accumulators store inorganic selenium (De Souza et al. 1998). The physiological cycle of plant uptake and accumulation of selenium is clearly shown in Fig. 1.

Selenium competes with Sulphur through Sulphur transport channels by a process called Sulphur assimilation pathway which leads to the formation of two major components mainly selenocysteine, selenomethionine and other organic selenium derivatives (Stadtman 1990). The major events of physiological process are conversion of selenate to selenite, selenide followed by selenocysteine. Some species of plants metabolise selenomethionine into volatile dimethylselenide (DMSe) which helps to reduce toxicity (Terry et al. 2000). With the help of Sec selenocysteine Lyase, the selenocysteine gets converted to elemental selenium and Alanine (Domokos-Szabolcsy et al. 2012). Methylation process takes place when selenocysteine gets converted into Se-Cystathionine by Cystathionine gamma synthase and Ortho-Phospho Hemoserine (OPH) coupling. This helps in the Se detoxification process (Neuhierl and Böck 1996).

Selenium in foods

Selenium has several chemical derivatives such as selenomethionine, selenocysteine, selenate and selenide. These are the major sources of dietary selenium of which the most widely

Fig. 1  Selenium uptake and accumulation in plants (ST Sulfur transport channels, $SeO_3$, selenate, $APSe$ adenosine phosphoselenate, $PAPSe$ phospho adenosine phospho selenite, $OAS$ O-acetyl serine, $OPH$ O-phospho hemoserine, $DMSe$ dimethylselenide; products are shown in purple color. Metabolites are shown in orange color)
consumed is selenomethionine. The major administration of selenium supplementation is via food. The level of selenium in each diet depends upon the plant uptake level. Hyperaccumulator plants contain higher amount of selenium concentration and vice versa (Rayman 2008). Foods that contribute to major selenium sources include cereals, breads, millets, wheat, nuts, meats, eggs, etc. Mushrooms, Brazil nuts and Broccoli are determined to have higher amount of selenium accumulation (Hart et al. 2011; Banuelos et al. 2013). The selenium concentrations in various foods are detailed in Table 1.

Approximately 20 μg per day is minimal RDA of selenium for adults to prevent Keshan disease. Keshan disease is a dilated cardiomyopathy (abnormality of the heart muscle) which was first found in “Keshan County” of North East China Provinces. This disease is characterised by multifocal myocardial necrosis and fibrosis which leads to cardiogenic shock and congestive heart failure. The normal intake of selenium should be about 50–55 μg per day. The dietary recommendations and regulations may differ across geographical locations, pH, humidity, climatic changes, and soil (Moore et al. 2000; Fairweather-Tait et al. 2010). Vegetables contain higher amount of selenomethionine, selenate, and selenite than selenocysteine. Selenomethionine is the major selenium source for humans and animals. Selenomethionine is synthesised through plants at the maximal concentration (Rayman 2002; Thomson 2004; Cubadda et al. 2010; Yasin et al. 2015). The consumption and upper level tolerance of selenium are elaborated in Tables 2 and 3.

### Table 1 Selenium Concentrations in various food items

| Sources           | Selenium concentration (mg/kg) | Major selenium species | References         |
|-------------------|-------------------------------|------------------------|--------------------|
| Brazil nuts       | 0.03–515                      | SeMet                  | Hart et al. (2011) |
| Bread             | 0.01–30                       | SeMet/selenate         |                    |
| Cereals           | 0.02–35                       | SeMet/selenate         |                    |
| Rice              | 0.05–0.08                     | SeMet                  |                    |
| Onions            | 0.02–0.05                     | SeMet/Sec              |                    |
| Garlic            | 0.05–1.0                      | SeMet/Sec              |                    |
| Broccoli          | 0.5–1.0                       | SeMet/selenate         |                    |
| Potatoes          | 0.12                          | SeMet                  |                    |
| Lentils           | 0.24–0.36                     | SeMet/selenate         |                    |
| Meat and meat products |                   |                        |                    |
| Beef              | 0.42–0.142                    | SeMet                  | Banuelos et al. (2013) |
| Chicken           | 0.081–0.142                   | SeMet/Sec              |                    |
| Fish              | 0.1–5.0                       | SeMet/selenite/selenate|                    |
| Eggs              | 3–25                          | SeMet/Sec              |                    |
| Pork              | 0.032–0.198                   | SeMet/selenate         |                    |
| Lamb              | 0.033–0.260                   | SeMet                  |                    |
| Milk and dairy products |                  |                        |                    |
| Yeast             | 0.6–15                        | SeMet                  |                    |

### Table 2 Nominal intake of selenium μg/per day (Vinceti et al. 2001)

| Age               | Male (μg) | Female (μg) | Pregnancy (μg) | Lactation (μg) |
|-------------------|-----------|-------------|----------------|----------------|
| Birth–6 months    | 15        | 15          | –              | –              |
| 7–12 months       | 20        | 20          | –              | –              |
| 1–3 years         | 20        | 20          | –              | –              |
| 4–8 years         | 30        | 30          | –              | –              |
| 9–13 years        | 40        | 40          | –              | –              |
| 14–18 years       | 55        | 55          | 60             | 70             |
| 19–50 years       | 55        | 55          | 60             | 70             |
| 51 + years        | 55        | 55          | –              | –              |

### Selenium supplements

Because of inadequacy of selenium in soil, it is sometimes necessary for humans to take selenium supplementation. Selenium is an excellent micronutrient that is acquired through food sources like nuts, breads, cereals, meat, fish, milk, and dairy products. These supplements act as antioxidants by its radical scavenging activities and reduce the oxidative damages in different internal organs (Rahmanto and Davies 2012). Reports elucidate the selenium supplementation and its effects on HIV, Crohn’s disease, Thyroid diseases, and cardiovascular diseases (Kuroki et al. 2003; Stone et al. 2010; Wu et al. 2015; Benstoem et al. 2015). The major forms of selenium supplements are Sodium selenite.
(inorganic), selenomethionine (organic), and selenium rich yeasts.

### Initial metabolism of selenium in human

Selenomethionine is the principal chemical form of dietary selenium and its consumed widely (Diamond et al. 1993). The initial metabolism starts by ingesting the selenomethionine (SeMet) via food or supplements. The SeMet absorbed through intestinal transport channels, reaches the Methionine pool. The major function of Methionine pool is to store selenomethionine and simultaneously incorporate them into proteins replacing the position of Methionine and forming selenomethionine. One of the reports reveals that “the ratio of 1 selenium atom (SeMet) per 8000 Methionine residues is equivalent to 1 selenomethionine per 1100 albumin molecules” (Burk 2001). The selenomethionine travels through Methionine cycle and Transsulphuration pathways (mostly in liver). The selenomethionine undergoes transsulphuration to produce selenocysteine and further with the help of Sec Lyase it tends to form selenide (Martin and Hurlbut 1976; Esaki et al. 1982). During dietary intake, the consumption of selenocysteine is comparatively low than selenomethionine resulting in highly reactive selenocysteine (Sec). These are maintained at low concentration whereas the less reactive selenomethionine (SeMet) is highly metabolised (Suzuki 2005). In liver, methyl selenol is produced as a result of α-lyase activity, and this methyl selenol in turn gets demethylated to form selenide (Suzuki et al. 2006). The major reported biological function of selenomethionine is to produce derivatives of selenium as well as to store selenomethionine for further synthesis of selenoproteins. The final step in the metabolism of selenium is the conversion of SeMet to Sec through Transsulphuration pathway followed by biosynthesis of selenoproteins (Labunskyy et al. 2014). The pathway for initial metabolism of selenium is partially explained in Fig. 2.

### Biosynthesis of selenoproteins

In the year 1985, the cloning of GPX1 leads to the discovery of UGA which is a stop codon for Sec during regular translational process (Chambers et al. 1986). Sec is a recently explored eukaryotic amino acid which directly encodes into proteins forming the 21st amino acid. The mechanism of translational decoding or otherwise called as biosynthesis of selenoproteins is shown in Fig. 3.

During translational decoding, UGA codon is one among the three stop codons which works for mRNA in which tRNA for selenocysteine (tRNA^Sec) recognizes and attaches to the respective codon (Buettner et al. 1999). Helper amino acid Serine conjugates with tRNA^Sec by seryl-tRNA synthetase which is further phosphorylated into phosphoserine by an enzyme phosphoseryl-tRNA kinase (Ganyc et al. 2006). After the initial metabolism of selenium, the dietary selenium gets phosphorylated by selenophosphate synthase 2 (SPS2) which is then conjugated with phosphoserine by selenocysteine synthase to produce selenocysteine (Chen and Berry 2003). All eukaryotic selenoprotein genes require selenocysteine Insertion Sequence (SECIS) element in 3′ UTR of the mRNA for recoding of UGA stop codon for Sec insertion. The unique stem loop structure of SECIS is the binding site of selenocysteine incorporation sequence binding protein 2 (SPB2) (Böck et al. 1991). The RNA binding domain of SPB2 (SECIS binding protein 2) belongs to L7Ae riboprotein family. SPB2 is a complex protein containing minimum of five isoforms which splices alternatively around 17–19 possible exons (Papp et al. 2008). The location and functions of different selenoproteins is detailed in Table 4. Other factors include specialized elongation factor (EFsec) that gets bound to tRNA^Sec for an extra protein binding support system (Squires and Berry 2008). The assembly of certain factors and cofactors on selenoprotein mRNA leads to the decoding of UGA into Sec via translational decoding process (De Jesus et al. 2006). Oxidation of SPB2 in nucleus gets reduced by nuclear specific GPX and Thioredoxin reductases (TRXR) proteins followed by CRM-1 dependent nuclear transport (Sneddon et al. 2003). Cofactors which contribute during the synthesis of selenoproteins are SECp43 as tRNA^Sec binding protein and the

### Table 3  Tolerable intake levels of selenium μg/per day (Vinceti et al. 2001)

| Age            | Male (μg) | Female (μg) | Pregnancy (μg) | Lactation (μg) |
|----------------|-----------|-------------|----------------|----------------|
| Birth–6 months | 45        | 45          | –              | –              |
| 7–12 months    | 60        | 60          | –              | –              |
| 1–3 years      | 90        | 90          | –              | –              |
| 4–8 years      | 150       | 150         | –              | –              |
| 9–13 years     | 280       | 280         | –              | –              |
| 14–18 years    | 400       | 400         | 400            | 400            |
| 19+ years      | 400       | 400         | 400            | 400            |
chaperone protein which binds to SECIS element and plays a role in protein translation (Shen et al. 2006). Another SECIS element binding protein is said to be nuclease sensitive element-binding protein 1 (NSEP1) which binds during the synthesis of selenoproteins via ribosomal pathway (Allmang et al. 2009). Translational decoding of selenoproteins gives idea on the importance of selenium in human metabolism and health. The selenoproteins synthesis processes such as translational decoding of selenocysteine, locations and their transport are shown in Figs. 3 and 4.

Role of Glutathione peroxidases—a type of selenoprotein in wound healing

When an injury occurs, the platelets start to aggregate and form thrombi at the wounded area. Meanwhile, the immune cells release the pro-inflammatory cytokines and neutrophils which produce huge amounts of reactive oxidative species (ROS) like hydrogen peroxidases (HP) and lipid peroxidases (LP). Reactive oxygen species (ROS) are the substances that are formed as a natural byproduct and have important roles in cell signalling and homeostasis. However, during times of environmental stress (e.g., UV or heat exposure), ROS levels can increase dramatically which may result in significant damage to cell structures and cumulatively—this is known as oxidative stress. During inflammation, Matrix Metalloproteinase (MMP), HP, metal scavengers and LP are released, resulting in higher expression levels of NOX2 gene in plasma membrane of cells and NOX2 gets activated during phagocytosis leading to production of superoxide radicals (Sen and Roy 2008; Gill and Parks 2008).

Glutathione peroxidases (GPXs) of selenoprotein family play a major role in the wound healing process. Glutathione peroxidases (GPXs) are known as the family of antioxidants. There are eight types of GPXs with diverse characteristics and properties such as chemical forms, location, functions and modes of action (Brigelius-Flohe and Maiorino 2013). Out of eight isoforms of GPXs, five have a SeCys residue in their side chains which helps to catalyse the hydrogen peroxidases, lipid peroxidases and heavy metals. Glutathione (GSH) reduces disulphide bonds formed within cytoplasmic proteins to cysteine by serving as an electron donor. In the process, glutathione is converted to its oxidised form, glutathione disulfide (GSGS), also called l-(-)-glutathione (Takeda 2015). GPXs detoxify the hydrogen peroxide into...
peroxidases to avoid Fenton’s reaction or Haber–Weiss reaction (several metals have a special oxygen transfer property which improve the use of hydrogen peroxide and some metals have a strong catalytic power to generate highly reactive hydroxyl radicals). It has been observed that during inflammatory process a strong mRNA expression of GPX1 was found which helps in the wound healing (Bedard and Krause 2007; Steiling et al. 1999). Later researchers state that there is a decreased level of GPX1 during early wounded phase resulting in the reduction of its activity in wounded control and immune-compromised rats (Iuchi et al. 2010). It has been reported that alkylation or nitric oxide dependent oxidation of selenocysteine results in reduction of GPXs activity (Gupta et al. 2002). Excess of hydrogen peroxide and super oxide dismutase (SOD) causes the selenocysteine to get converted into dehydrolalanine, which results in irreversible inactivation and degradation (You et al. 2010). The study clearly revealed that the protein levels of GPX1 are increased between 3 and 7 days after skin injury (Woo et al. 2010). Availability of selenium restricts the abundance of GPX proteins. Trauma patients with skin wound healing disorder present with reduced serum selenium status. Therefore, lower selenium restricts the abundance of GPX2 protein; providing patients with selenium supplementation helps to improve GPX2 protein level (Blass et al. 2013). Appropriate mechanism responsible for this needs to be further experimented. Selenium can be used as therapeutic agents for the treatment of wound healing disorders. The functions of various selenoproteins during the wound healing process are shown in Fig. 5.

**Role of selenoprotein S in wound healing**

During inflammatory phase in wound healing process, soluble factors such as chemokines and cytokines are released, which can phagocyte the debris, bacteria, and damaged tissues. Recent studies suggested that selenoprotein S is plays a crucial role in inflammatory phase. Selenoprotein S (otherwise named as Sel S, SELENOS, TANIS, SEPS1 and VIMP) is a transmembrane protein found in Endoplasmic Reticulum (ER) and plasma membranes. Their major
Table 4 Various functions and health effects of selenoproteins

| S.no | Selenoproteins (GPxs) | Location | Functions | Health effects | References |
|------|-----------------------|----------|-----------|----------------|------------|
| 1    | Glutathione peroxidase-1 (GPx-1) | Highly expressed at erythrocytes, liver, lungs and kidneys | Antioxidant activity, prevents viral mutations by reducing the retroviral virulence capacity | Deficiency causes cardiomyopathy, autism, blood pressure, vascular disease, Keshan disease, cancers such as lung, prostate, bladder and primary liver damages | Reeves and Hoffmann (2009) |
| 2    | Glutathione peroxidase-2 (GPx-2) | Present predominantly in gastrointestinal tissues and Human liver | Antioxidant activity, protects against Oxidative damages, maintains intestinal mucosal integrity, anti-apoptotic function in colon | Oxidative stress, free radical sedimentation | Florian et al. (2010) |
| 3    | Glutathione peroxidase-3 (GPx-3) | Predominantly in extracellular fluid and plasma. It is also found in liver, breasts, heart, kidneys, GI tract, placenta and male reproductive system | Reduce lipid hydro peroxides, antioxidant in plasma region, protection of thyroid gland from hydrogen peroxide in thyrocytes | Thyroid cancer and ischaemic stroke, oxidative stress | Yant et al. (2003); Schmutzler et al. (2007) |
| 4    | Glutathione peroxidase-4 (GPx-4) | Highly expressed in Testes. In the cells of cytosol, mitochondria and nucleus | Antioxidant protects brain membranes from peroxidaive degradation, conversion of cholesterol and cholesterol esters to less toxic derivatives. Essential for sperm motility and viability | Colorectal cancer, prostate cancer | Maiorino et al. (2003); Papp et al. (2010) |
| 5    | Glutathione peroxidase-5 (GPx-5) | Embryo and olfactory epithelium | Not known | Not known | Ashton et al. (2009) |
| 6    | Glutathione peroxidase-6 (GPx-6) | Found only in human | Not known | Not known | Brigelius-Flohé et al. (2012) |
| 7    | Glutathione peroxidase-7 (GPx-7) | Lumen of the endoplasmic reticulum | Inverse relationship between GPx-7 and proliferation of cancer cells | Not known | Köhrle et al. (2000) |
| 8    | Glutathione peroxidase-8 (GPx-8) | Membrane protein of endoplasmic reticulum | Involves in protein folding, antioxidant | Not known | Florian et al. (2010) |

Idodothyronine deiodinases (DIOs)

| S.no | Idodothyronine deiodinases-1 (DIO1) | Predominantly in liver, kidney, thyroid and brown fat | Production of active T3 cell hormones in thyroid and peripheral tissues. It converts inactive thyroxin into active 3', 3"-5' triiodothyronine | Loss of appetite, muscle strength and free IGF-1 concentration | Tinggi (2008); Schomburg and Köhrle (2008) |
|------|-----------------------------------|--------------------------------------------------|-----------------------------------------------------------------|-------------------------------------------------|------------------------------------------|
| 10   | Idodothyronine deiodinases-2 (DIO2) | Highly expressed in Central nervous system, Brown adipose tissue and skeletal muscle, pituitary, heart | T3 production in peripheral tissues. Activation of thyroid hormones | Diabetes type-2, reduction in bone mineral density, mental retardation, Osteoarthritis | Lu and Holmgren (2009) |
| 11   | Idodothyronine deiodinases-3 (DIO3) | Present in placenta, uterus, fetal, skin, cerebral cortex and CNS | Prevents high exposure of foetus towards T3 cells. Deactivation of thyroid hormones | Osteoarthritis | Rasmussen et al. (2011) |
| S.no | Selenoproteins | Location | Functions | Health effects | References |
|------|----------------|----------|-----------|----------------|------------|
| **Thioredoxin reductases (TrxR)** | | | | | |
| 12 | Thioredoxin reductases 1 (TrxR₁) | Intracellular content in cytosol and nucleus | Antioxidant activity, reduction of Thioredoxin, controls transcription factors, apoptosis and cell proliferation | Mutations can occur during DNA synthesis, colorectal adenoma and lateral sclerosis | Reeves and Hoffmann (2009) |
| 13 | Thioredoxin reductases 2 (TrxR₂) | Wide spread and highly expressed in mitochondria | Cell growth factor in DNA synthesis and inhibition of apoptosis | Gastric cancer and gastric ulcers | Ashton et al. (2009) |
| 14 | Thioredoxin reductases 3 (TrxR₃) | Specifically on testes | Not known | Not known | Ashton et al. (2009) |
| **Other selenoproteins** | | | | | |
| 15 | Selenoprotein P (SEPP1) | Extracellular glycoprotein predominantly found in plasma, highly expressed in brain, liver and testes | Transportation of selenium to tissues, regulator of homeostasis, antioxidant functions, contains 10 selenocysteine residues | Infertility in male, prostate cancer, abnormal kidney movements, spontaneous seizures in mice | Mostert et al. (2001); Schomburg et al. (2004); Burk and Hill (2009) |
| 16 | Selenoprotein S (SEPS1) | Predominantly in endoplasmic reticulum and plasma membrane | Regulate inflammation, it deletes the misfolded proteins in endoplasmic reticulum, induce ER stress apoptosis | High risk of CHD, pre-eclampsia, ischaemic stroke. Cancers such as colorectal and gastric | Meyer et al. 2012; Alanne et al. (2007) |
| 17 | Selenoprotein N (SelN) | A trans membrane glycoprotein related to ER | Redox signaling, muscle development, calcium haemostasis | Yet to be discovered | Arbogast and Ferreiro (2010) |
| 18 | Selenoprotein W (SelW) | Skeletal muscle, colon, heart and prostate | Antioxidant in human lungs, calcium binding | Not known | Yao et al. (2013) |
| 19 | Selenoprotein K (SelK) | Immune cells and spleen | Antioxidant activity | Not known | Liu et al. (2012) |
| 20 | Selenoprotein H (SelH) | Brain, nucleus and muscle cells | Gene regulation of glutathione | Not known | Mehta et al. (2013) |
| 21 | Selenoprotein R (SelR) | Kidney and liver cells | Antioxidant, protein repair, methionine metabolism | Not known | Davis et al. (2012) |
| 22 | Selenoprotein M (SelM) | ER and neuronal cells | Antioxidant activity | Not known | Yao et al. (2013) |
| 23 | 15Kd selenoprotein (SEP15) | ER | Affects glycoprotein folding | Cancers prostate and lung | Tsuji et al. (2012) |
| 24 | Selenophosphate synthase 2 (SPS-2) | Kidney, liver | Synthesis of selenophosphates for selenocystine biosynthesis | Not known | Hart et al. (2011) |
activity includes (1) removing the misfolded proteins from ER lumen (2) protecting the cells from oxidative damage and (3) participating in ER-stress induced apoptosis. Selenoprotein S was initially derived from diabetic rats during the observational studies on fasting diabetic assay (Walder et al. 2002; Ye et al. 2004; Gao et al. 2004, 2007). This study also reveals that Sel S interacts with Serum Amyloid A (SAA), an acute phase inflammatory response protein and provides a mechanistic link between Type-2 diabetes, inflammation and cardio vascular disorders. Other reports state that Endoplasmic Reticulum stress induces the Sel S expression by ER stress response element via nuclear factor (NF)-κb pathway (Gao et al. 2006). The depletion of Sel S by siRNA increases the release of inflammatory cytokines IL-6 and TNF-α, concluding that Sel S may regulate the cytokine production in macrophages and plays a major role in controlling the inflammatory responses. A particular polymorphism of Sel S was proven to be responsible for increased plasma levels of the inflammatory cytokines. Sel S gene has been linked to variant markers of inflammation and one among the particular variant is 105G-A (Curran et al. 2005). The role of Sel S in the inflammatory response may be correlated with the inflammation phase of the wound healing mechanism. But still these assumptions can be solved in future with more experimental evidences. The assumptions are shown in Fig. 5.

**Role of other selenoproteins in wound healing**

**Thioredoxin reductases**

There are three Thioredoxin Reductases (TR1, TR2, and TR3). The major functions of TRs are to control the redox signalling and antioxidant activities (Ducros and Favier 2004). Thioredoxin system plays an important role in the regulation of gene expression via redox control of transcription factors including NF-kB, Ref-1, AP-1, P-53, Glucocorticoid receptor and apoptosis regulating kinase (ASK-1) which results in immune responses and other major activities (Selenius et al. 2010). They have the ability to reduce the ascorbyl free radicals through recycling the ascorbate. Humans lack the capacity to synthesise ascorbic acid which is an important antioxidant in the protection of cells from oxidative stress and inflammatory responses (Mustacich and Powis 2000). Thus, the selenoprotein derivative such as Thioredoxin reductase plays important role during oxidative stress and inflammation in wound healing process.

**Selenoprotein P**

Selenoprotein P (SEPP 1, Sel P) is the only selenoprotein which contains 10 selenocysteine residues (Olson et al. 2005). It is a glycosylated protein where there are 3
occupied N-glycosylation site and 1 occupied O-glycosylation site (Suzuki et al. 2010a, b). Sel P is mainly produced in liver and then transported to plasma. Sel P is released into the blood stream and is responsible for the distribution of selenium to other organs and tissues (Burk and Hill 2005). With these evidences we assume that selenoprotein P acts as a selenium transport system and also selenoprotein P is responsible for maintaining the haemostasis throughout the human body during the normal and disturbed metabolism (e.g. during wound healing process) (Schomburg et al. 2003). Focusing on the relationship between selenoprotein P and wound healing mechanism requires further studies.

**Selenium drugs**

Selenium is an important micronutrient for the human body that is especially received via food supplement (Lau et al. 2017). Very small amounts of selenium are required for maintenance of ideal health as selenium is a constituent of the selenoproteins that contribute to a vast array of cellular functional processes (Wu et al. 2015). The functions like redox regulations, immunity and inflammatory reactions, cardiovascular and reproductive well-being, brain activities and repairs, thyroid hormone control factors and carbohydrate regulations too (Forootanfar et al. 2014; Mistry et al. 2012). Selenium deficiency is related to several human illnesses with numerous levels of diseases (Vinceti et al. 2017). Excessive selenium intake can be toxic and might lead to selenosis condition (Sutter et al. 2008). Currently, the recommended dietary allowance of selenium for adults is set at 55 µg (0.7 µmol)/day. Individuals with day by day selenium intake much less than ~ 15 µg appear to be susceptible to selenium deficiency-associated illnesses, while folks who consume over 400 µg/day are prone to selenium toxicity, although a few researches have proven that safe ranges of selenium intake may be a whole lot decrease than anticipated (Vinceti et al. 2001).
Selenium containing compounds for cancer treatment

The relationship between selenium and cancer was explored during the middle of the nineteenth century, which then out-stretched the interest in selenium supplements to cancer prevention and therapy. The selenium containing compounds can be grouped into three main types (1) inorganic, (2) organic (natural and synthetic) and selenium nanoparticles (SeNPs) (recently added) which has exhibited anticancer ability.

Generally it is shown that selenium compounds exhibit anticancer capability primarily through their direct or indirect antioxidant properties that sustain the intracellular redox status and these actions guards the healthy cells from reactive oxygen species induced oxidative damage (Rahmanto and Davies 2012). Reactive oxygen species are free radicals generated during normal bio-physiological functions with unpaired electrons and there is evidence that excessive reactive oxygen species promotes carcinogenesis via raised oxidative stress and DNA mutation (Prasad et al. 2017). Despite the links between reactive oxygen species and cancer formation but still ideal (usually low) levels of reactive oxygen species are actually helpful as they play important roles in regulating many biological functions. Some enzymes and cells intentionally produce superoxide radicals to destroy the attacking pathogens (Georgieva et al. 2017). Reactive oxygen species also terminate the damaged cells by stimulating cellular senescence and apoptosis (Valko et al. 2006).

Research on inorganic and organic forms of selenium compounds have metabolized differently and have diverse mechanisms of action towards cancer cells. Organic selenium compounds are better retained and utilised than inorganic forms though the both are readily absorbed in the human body (Fairweather-Tai et al. 2010). Numerous publications with biochemical, epidemiological, clinical, and animal studies supports the cancer prevention ability of inorganic and organic selenium compounds (Rayman 2005; Papp et al. 2007; Jackson and Combs 2008; Hatfield et al. 2009; Fernandes and Gandin 2015; Brigelius-Flohé and Flohé 2017). Conversely, selenium compounds toxicity risks have also been recorded. It has been found that inorganic selenium compounds shows higher genotoxic stress, which may have lower therapeutic value, higher toxicity and an increased threat on cancer proliferation. Contrarily, organic selenium compounds retain significant anti-tumor activity along with increased ability to prevent metastasis, have fewer side effects and lower systemic effects. Organic selenium compounds comprise a vast group of chemically diverse nucleophilic molecules. In reality, the toxic effects of selenium compounds are determined by multiple factors, its forms and dosage exposure being two of the most important parameters (Misra et al. 2015).

Inorganic selenium compounds

Commonly studied inorganic selenium compounds include the sodium selenite and sodium selenate for the cancer prevention and treatment. Depending on their valence states of these compounds, the functions and its effects differ. Different concentrations of sodium selenate (5, 10, 30, 50 µM for 48 h) and sodium selenite (0.1, 0.25, 0.5 µM for 48 h) were tested for their capability to sensitize human oral squamous carcinoma (KB) cells which were resistant to chemotherapeutic drug vincristine (KBV20C). On observation they found that they are able to sensitize KBV20C to the similar level as the sensitive parent KB cells. By arresting the cell cycle at G2-phase and activating apoptotic pathways, Sodium selenite compound are able to show higher sensitizing effect on the KBV20C cells (Choi et al. 2015).

The most studied inorganic selenium compounds are sodium selenite which exhibits outstanding chemo-preventive and anticancer features (Kieliszek et al. 2017). The compound produced noticeable anti-proliferative activities against three oral cancer cell lines (HSC-3, HSC-4, and SAS) at a concentration of 5–100 µM on treatment for 2–5 days (Endo et al. 2017). Sodium selenite was found to be efficiently inhibiting the cell proliferation of several types of cancer cells even the deadliest cancer in worldwide, the lung cancer cells too (Tan et al. 2018). Among different human cancer cell lines tested, lung cancer cells, in general, appeared to be especially sensitive to selenite (Berthier et al. 2017; Lipinski 2017; Chen et al. 2018a, b). Previous reports showed that selenite cytotoxicity at concentration of 5 µM for treatment of 5 h was correlated with selenium uptake of three lung cancer cell lines (H157, H611, and U2020) and it results indicated that at high concentrations (> 1 mM) of selenate was found to be non-toxic for these cell lines (Olm et al. 2009). Selenite played a key role in natural killer (NK) cell-based anticancer immunotherapy where it could raise the susceptibility of cancer cells to CD94/NK group 2A-positive NK cells, and it showed positive clinical applications to lung cancer patients (Enqvist et al. 2011). On reduction of sodium selenite to hydrogen selenide which is the common intermediate of dietary selenium metabolism might activate the apoptosis of cancer cells (HepG2, HeLa, and MCF-7 cells) through accumulation in mitochondria, this might then lead cell death (Hu et al. 2018). In general most of the inorganic selenium compounds exhibit higher genotoxic stress, which account for lower therapeutic value, higher systemic toxicity and an increased risk of tumour burden.

Organic selenium compounds

In the field of cancer research organic selenium compounds drew more attention than inorganic selenium compounds
because of their lesser toxicity risk and have capability to bring substantial anticancer activity (Ronai et al. 1995; Jariwalla et al. 2009). Some of the organic selenium compounds like selenomethionine derivatives (selenomethionine, selenocystine, Methylseleno-cysteine, selenodiglutathione), Methylseleninic acid, Ebselen, Ethaselen, Diselenolides, selenazofurin, selenocyanates, selenoesters, selenium-non-steroidal anti-inflammatory drug (Se-NSAID), selenoureas, selenocarbonyl derivatives are explained below with their functions and structures in Table 5 (Gandin et al. 2018). These compounds display anticancer and chemo-preventive activity by their different mechanisms of action like reducing the oxidative stress, inducing apoptotic events and enhancing the chemotherapeutic activity. Organic selenium compounds have all capabilities to be used as anti-neoplastic agents against solid tumours (Alvarez-Perez et al. 2018). Organic selenium compounds largely preferred in cancer therapy because of its anti-necrotic and pro-apoptotic features since necrosis in cancer cells is related to host inflammatory response and may lead to treatment complications (Ali et al. 2018).

Natural organo-selenium compounds

Selenomethionine  Selenomethionine has displayed prominent cytotoxicity in many cancer cells like lung, breast, colorectal, prostate, melanoma and this was achieved at higher concentrations from medium to high µmolar. Selenomethionine induced apoptosis by numerous pathways like the activation of caspases, p53, ER stress, HDAC inhibition, altered expression of Bclxl, Bax, Bad and Bim, decreased cyclooxygenase-2 expression and decreased glutathione peroxidase activity (Baines et al. 2002; Yang et al. 2009). Furthermore, selenomethionine on combination with ionizing radiation exhibited enhanced selectivity towards NCI-H460 and H1299 lung cancer cells, whereas having slight effect on WI-38 human diploid lung fibroblasts (Shin et al. 2007). Reports on low systemic toxicity and high tumor selectivity of selenomethionine compound in a mouse xenograft model of colorectal carcinoma (SW480) (Suzuki et al. 2010a, b).

Selenocystine  It is a diselenide oxidation product formed from the amino acid selenocysteine. By low-molecular weight thiols and disulphide reductases this selenocystine being efficiently reduced to selenocysteine which is of highly redox active in nature. Their cytotoxic effects were found to be in the low to medium µm range for lung, breast, cervical, liver cancer cells and also in melanoma cells. Cytotoxicity of selenocystine is due to increased ROS production which induces DNA damage and leads to mitochondrial mediated apoptosis associated with p53 phosphorylation. In addition to these cytotoxic mechanisms selenocystine exhibited the extrinsic/death-receptor pathway in melanoma cells (Chen and Wong 2008). In another study selenocystine treatment in cervical cancer cells has been related with two different types of cell death via apoptotic and paraptotic-like associated with ER stress which leads to induction of the unfolded protein response (UPR) (Wallenberg et al. 2014). Selenocystine inhibited tumor growth in a mouse xenograft model of melanoma with no signs of systemic toxicity. Further, selenocystine and auranofin exhibited a synergistic effect on inducing apoptosis in a mouse xenograft model of lung cancer (Fan et al. 2014). Selenocystine on administration in humans for acute and chronic myeloid leukemia treatment worked remarkably, particularly effective against immature leucocytes with respect to mature leucocytes and also without bringing any significant effect on the bone marrow (Weisberger and Suhrland 1956).

Methylselenocysteine  It is derivative of mono-methylated selenoaminoacid which is converted to methylselenol by activity of selenocysteine Se-conjugated β-lyases (Suzukiet al. 2007). Methylselenocysteine efficacy is completely dependent upon the production of the active metabolite, methylselenol which varies across cells, tissues and organs. The in vitro cytotoxicity of this compound has been found out to be in micromolar (µm) range for many human cancer cell lines which includes colon, breast, lung and oral squamous cells. Furthermore, Methylselenocysteine treatment in vitro has also shown to decrease vascular endothelial growth factor expression (Suzuki et al. 2010a, b). Moreover, the mechanisms by which Methylselenocysteine treatment causing cell death has been associated with role for caspase-dependent pathway and to extent of mitochondrial signalling involvement in this process still remains unclear (Kim et al. 2001; Suzuki et al. 2008; Schröterová et al. 2009). Methylselenocysteine treatment in mice bearing FaDu human head and neck squamous cell carcinoma xenografts has caused series of promising antiangiogenic effects, tumor growth inhibition, enhanced doxorubicin delivery associated with decreased microvessel density, improved vascular maturation, better vessel functionality and decreased vascular permeability (Bhattacharya et al. 2008). Similarly, a synergistic effect on tumor growth inhibition by Methylselenocysteine and tamoxifen was detected in a mouse xenograft model of MCF-7 breast cancer (Li et al. 2009). In another study a similar synergistic effect of Methylselenocysteine and irinotecan was witnessed in mouse xenograft models of colon carcinoma and squamous cell carcinoma of the head and neck. Likewise this chemotherapeutic synergy effects was also apparent in both sensitive (HCT-8 and FaDu) and drug-resistant (HT-29 and A253) xenografts (Cao et al. 2004).

Selenodiglutathione  During the reduction of selenite by glutathione, the selenodiglutathione metabolite is formed.
| S.no | Name of the compounds | Structure | Types of cancer/cell lines | Dosage concentration in micromolar (µM) | Apoptosis mechanism | References |
|------|-----------------------|-----------|---------------------------|----------------------------------------|---------------------|------------|
|      |                       |           |                           |                                        |                     |            |
| 1)   | Sodium selenite       | ![Sodium selenite structure](image) | 1) HSC-3, HSC-4, and SAS 5–100 µM for 2–5 days | Pronounced anti-proliferative effect of selenite against three oral cancer cell lines | Endo et al. (2017) |
| 2)   | Sodium selenate       | ![Sodium selenate structure](image) | 2) Lung cancer cell lines (H157, H611, and U2020) 5 µM for 5 h | Played a role in natural killer (NK) cell-based anticancer immunotherapy where it could increase the susceptibility of cancer cells to CD94/NK group 2A-positive NK cells | Olm et al. (2009) |
| 3)   | Selenomethionine      | ![Selenomethionine structure](image) | 3) HepG2, HeLa, and MCF-7 cells | Apoptosis of cancer cells occurs by accumulation in mitochondria which would subsequently damage mitochondrial function, structure and lead to cell death | Hu et al. (2018) |
|      |                       |           |                           |                                        |                     |            |
| 1)   | Selenomethionine      | ![Selenomethionine structure](image) | 1) Lung cancer cells -NCI-H460 and H1299 10–1000 µM | SeMet used in combination with ionizing radiation enhanced treatment | Yang et al. (2009) |
| 2)   | Mouse xenograft model | ![Mouse xenograft model structure](image) | 2) Mouse xenograft model of colorectal carcinoma (SW480) 10–1000 µM | Low systemic toxicity and high tumor selectivity has been reported | Shin et al. (2007) |
| 3)   | Colon cancer          | ![Colon cancer structure](image) | 3) Colon cancer 100–1000 µM | Participation of decreased COX-2 expression | Baines et al. (2002) |
| 4)   | Breast and prostate cancer | ![Breast and prostate cancer structure](image) | 4) Breast and prostate cancer 10–1000 µM | Cell death is facilitated by caspases and ER stress | Suzuki et al. (2010a) |
| S.no | Name of the compounds | Structure | Types of cancer/cell lines | Dosage concentration in micromolar (µM) | Apoptosis mechanism | References |
|------|-----------------------|-----------|----------------------------|------------------------------------------|---------------------|------------|
| 4    | Selenocystine         | ![Structure](image1.png) | 1) Cervical cancer | 100–1000 µM | Paraptotic-like mediated by ER stress and UPR | Wallenberg et al. (2014) |
|      |                       |           | 2) Nasopharyngeal, Liver, Lung and Melanoma | 1–20 µM | Cell death is facilitated by caspases, mitochondrial dysfunction/signalling and PARP cleavage | Poerschke and Moos (2011) |
|      |                       |           | 3) Acute promyelocytic leukemia, | 10–100 µM | | |
|      |                       |           | 4) Colon cancer | 1–100 µM | | |
| 5    | Methylselenocysteine  | ![Structure](image2.png) | Breast cancer | 100–1000 µM | Cell death is facilitated by caspases, ER stress, mitochondrial dysfunction/signalling and PARP cleavage | Suzuki et al. (2010b) |
|      |                       |           | Colorectal cancer | 10–1000 µM | | |
|      |                       |           | Lung cancer | 10–1000 µM | | |
|      |                       |           | Oral Squamous | 10–1000 µM | | |
| 6    | Selenodiglutathione  | ![Structure](image3.png) | Acute myeloid leukaemia | 1–20 µM | Cell death is facilitated by ROS production and oxidative damage | Wallenberg et al. (2014); Tobe et al. (2015) |
|      |                       |           | Breast cancer | 1–20 µM | | |
|      |                       |           | Cervical cancer | 1–20 µM | | |
|      |                       |           | Lymphoma | 1–20 µM | | |
|      |                       |           | Oral cancer | 0.1–2 µM | | |
### Table 5 (continued)

| S.no | Name of the compounds | Structure | Types of cancer/cell lines | Dosage concentration in micromolar (µM) | Apoptosis mechanism | References |
|------|------------------------|-----------|----------------------------|------------------------------------------|---------------------|------------|
| 7    | Methylseleninic acid    | ![Structure](image1) | Lung cancer                  | 10–100 µM                               | Cell death is facilitated by caspases, ER stress, UPR, mitochondrial dysfunction/signalling and PARP cleavage | Poerschke and Moos (2011); Wang et al. (2014); Shigemi et al. (2017) |
|      |                        |           | Pancreatic cancer           | 0.1–2 µM                                |                     |            |
|      |                        |           | Lymphoma                    | 1–100 µM                                |                     |            |
| 8    | Ebselen                | ![Structure](image2) | Bone marrow cancer           | 10–100 µM                               | Cell death is mediated by mitochondrial signaling. Acts as a substrate for thioredoxin reductase and rapidly oxidizes thioredoxin, leading to oxidative stress | Zhang et al. (2014) |
|      |                        |           | myeloma                     |                                          |                     |            |
| 9    | Ethaselen              | ![Structure](image3) | Tongue cancer                | 1–20 µM                                 | Cell death is mediated by thioredoxin reductase inhibition and subsequent oxidative stress | Xing et al. (2008); Wang et al. (2012) |
|      |                        |           | cervical, gastric, liver, lung cancers | 0.1–2 µM                                |                     |            |
| 10   | Diselenides            | ![Structure](image4) | Colon cancer cells           | 1–100 µM                                | Cytotoxicity by cell-cycle arrest and caspase pathway | Nedel et al. (2012) |
| 11   | Selenazofurin          | ![Structure](image5) | MCF-7 human breast carcinoma cells | 0.1–2 µM                                | Pro-apoptotic activity and higher growth inhibition | Zhou et al. (2015) |
|      |                        |           | Bladder, cervical, colon, leukemia, lymphoma, kidney | 0.1–2 µM in blood cell tumours, 1–20 µM in solid tumours | Act through non-competitive inhibition of inosine monophosphate dehydrogenase, thus limiting de novo guanine nucleotide biosynthesis | Franchetti et al. (1997) |
| S.no | Name of the compounds | Structure | Types of cancer/cell lines | Dosage concentration in micromolar (µM) | Apoptosis mechanism | References |
|------|-----------------------|-----------|---------------------------|----------------------------------------|---------------------|------------|
| 12   | Selenocyantes         | ![Structure](image1) | CCRF-CEM, HT-29, HTB-54, PC-3 and, MCF-7 cell lines | 1–100 µM | Increase in compound lipophilicity and thereby increasing the alky chain length was consistent in cells associated to a caspase-dependent apoptotic cell death through the induction of p53, Bax and suppression of Bcl-2 | Romano et al. (2015) |
| 13   | Selenoesters          | ![Structure](image2) | Prostate, breast, lung and colon cancer cell lines | Below 0.1 µM | Acting as GSH depleting agents | Dominguez-Alvarez et al. (2014) |
|      |                       |           | Human hepatoma cells | Ability to act as strong inhibitor of cellular efflux pump P-gp |                       |            |
|      |                       |           | In mouse MDR T-lymphoma cells and in human colon cancer cells |                       |                       |            |
| 14   | Se-NSAID              | ![Structure](image3) | Breast, colorectal, melanoma and pancreatic cancers | 1–100 µM | Apoptosis mediated by caspases, PARP cleavage and ROS production. Involvement of COX-2 and PI3K/AKT inhibition | Gowda et al. (2013); Plano et al. (2016) |
| 15   | Selenoureas           | ![Structure](image4) | Colon cancer cell models | Below 10 µM | Activation of caspase-dependent pathways and inhibition of antiapoptotic proteins | Alcolea et al. (2016) |
| 16   | Selenocarbonyl derivatives | ![Structure](image5) | MCF-7 cells | 1–100 µM | Arresting the cell cycle and caspase pathway | Martins et al. (2015) |

Dosage concentration (in vitro dose for 48–72 h IC₅₀)—very low (0.1–2 µM), low (1–20 µM), low to medium (1–100 µM), medium (10–100 µM), medium to high (10–1000 µM), high (100–1000 µM)
Through glutaredoxin and thioredoxin systems activity, selenodiglutathione is reduced to hydrogen selenide by glutathione. This pro-oxidant play a central role in redox cycling with glutathione and producing superoxides, hydrogen peroxide which further results in ROS generation (Björnstedt et al. 1992; Wallenberg et al. 2010). Selenodiglutathione showed apoptotic effect in the low µm concentrations towards different cancer cells like breast, ovarian, cervical, lymphoma, promyelotic leukemia and oral squamous (Wu et al. 1995; Last et al. 2006; Tobe et al. 2015). Recently it has also been shown that selenodiglutathione exhibited different intracellular targets, expression patterns and mechanism of cytotoxicity when compared to selenite treatment (Wallenberg et al. 2014).

**Synthetic organoselenium compounds**

Till the year 1990, the chemistry of organoselenium compounds was hardly established in contrast with that of organosulfur compounds, mainly because of the high uncertainty and harmfulness of numerous Se-containing compounds. But lately due to developments in the field of synthesis and reactivity of organoselenium compounds as well as the discovery on the essential role of selenium and selenoproteins in cancer has enormously motivated for the intense development of new therapeutics in medicine.

**Methylseleninic acid**

The compound was synthesized on oxidative decomposition of methylselenocysteine and it is most simple organic selenium compounds with therapeutic properties. They are able to kill cancer cells in different cell lines through the activation several caspase pathways (3, 7, 8 and 9), ER stress, UPR induction, cytochrome c release and PARP cleavage (Jiang et al. 2001, 2002; Singh et al. 2008; Shigemi et al. 2017). They induce apoptosis specifically in p53 wild-type, p53-mutant, p53-null cells and therefore representing a p53-independent cancer cell death pathway (Zu et al. 2006). In vivo and in vitro studies have shown that Methylseleninic acid has been excellent anticancer agent against a range of cancer models like lung, breast, melanoma and prostate cancers (Plano et al. 2007). In a recent study, Methylseleninic acid has exhibited substantial cytotoxic effects toward monocytes leukemia cells (THP1) when likened with the healthy peripheral blood mononuclear (PBM) cells. In dose-dependent manner at various concentrations (2.5, 5, and 15 µM for 48 h), Methylseleninic acid showed improved anticancer activity against the malignant THP1 cells (Lobb et al. 2018).

**Ebselen**

Ebselen [2-phenyl-1,2-benziselenazol-3(2H)-one (PZ-51)] was synthesized by Lesser and Weiss in 1924 and measured immaterial pharmacologically for 60 years. Later Helmut Sies gave a new life by highlighting the abilities of this molecule to mimic the activity of glutathione peroxidase enzyme (Muller et al. 1984; Zhao et al. 2002). From that time surplus of studies has been carried out on the pharmacological and biochemical properties of Ebselen (Azad and Tomar 2014). Recent reports on various applications of Ebselen include anticancer, antimalarial antitubercular and neuroprotection agent (Parnham and Sies 2013).

Ebselen is an effective source of thioredoxin reductase, and which stimulates the thioredoxin to get rapid oxidation. Anti-oxidative properties of this compound have been extensively studied, but only few studies on ebselen highlights about the antiproliferative and anticancer properties through ROS production (Zhang et al. 2014). Ebselen might also alter the mitochondrial function by Bax activation and these properties leads to inhibit Quiescin sulphydryl oxidase 1, an enzyme that helps in promoting the growth and invasion of tumor cells by altering the extracellular matrix composition (Hanavan et al. 2015). There has been report that on daily oral treatment with ebselen has resulted in a 58% reduction in tumor growth in mice bearing human pancreatic tumor xenografts. Major antitumor proficiency was attained by combining the ebselen with gamma radiation which induces apoptosis and as well helps in modulating the response of pro- and anti-inflammatory cytokines (Thabet and Moustafa 2017).

New cyclo-hexyl ring containing ebselen analogues has been synthesized which showed higher redox properties and also reducing cell growth in both MCF-7 and DU145 cancer cell lines with IC50 values that were within the low micromolar range (Pacula et al. 2017). Sequences of benzoselenazole-stilebene hybrids were synthesized by combing ebselen and reservatrol. These were designed to develop more multi-target redox-active agents. Certainly upon their biological assessments, it showed that all the developed test compounds acted as antiproliferative agents in the human cancer cell lines Bel-7402, A549, HeLa, and MCF-7 and also proven to be TrxRd inhibitors. Furthermore, mechanistic studies indicated that these compounds increase the intracellular ROS which implements apoptosis in cancer cells (Yan et al. 2015).

**Ethaselen**

Ethaselen (1,2-[bis(1,2-benziselenazolene-3(2H)-ketone)ethane) has been widely studied in a number of cell lines (Zhao et al. 2006) and it is found to target and inhibit the central antioxidant and redox regulatory enzyme,
thioredoxin reductase, which has been the promising target for any anticancer drugs (Urig and Becker 2006). Ethaselen, through thioredoxin reductase inhibitory effect arrest the cell growth S phase and thereby the proliferation of tumor cells was inhibited (Shi et al. 2003). Concurrently, ethaselen established promising outcomes in in vivo experiments, with tumor inhibitory effects in the range of 40–80% in prostate, tongue and liver cancer models (Xing et al. 2008; Tan et al. 2009; Wang et al. 2012). Ethaselen in combination with cisplatin (cis-diamine dichloroplatinum II, DDP) in a lung xenograft mouse model showed significant synergistic reduction of tumor size, with no evident signs of systemic or organ toxicity (Tan et al. 2010). Combination of cisplatin and ethaselen produced similar results in a leukemic cisplatin resistant cell line with substantial synergistic effects. Ethaselen with Sunitinib (multitargeted tyrosine kinase inhibitor) combination has exhibited similar synergistic effects against spreading of colorectal cancer cells (Zheng et al. 2016). In spite of ethaselen encouraging anticancer activity, its solubility in physiological media is not ideal, thus preventing its application without appropriate formulation. Copolymer micelles have helped in increasing ethaselen water solubility which led to enhanced antitumor activity and attribute to massive accumulation at the tumor site (Liu et al. 2010). Currently ethaselen in Phase I clinical trials in patients with non-small cell carcinoma (NSLCLC). Tolerable and safe dose in the Ia/b phase was found to be 1200 mg/day (ClinicalTrials.gov identifier: NCT02166242).

**Diselenides**

Diselenides are characterized by the presence of a Se–Se bond largely used in organic synthesis because of their accessibility through a number of different synthetic protocols and relative stability. Diphenyldiselenide exhibited substantial cytotoxicity against neuroblastoma cells and it is effectively carried out through ERK1/2 mediated apoptosis (Posser et al. 2011). Substituted diaryldiselenides (3-(trifluoromethyl)-diphenyldiselenide and 4-methoxy-diphenyl diselenide) have capability to induce cytotoxicity by cell-cycle arrest in human colon cancer cells and triggering the caspase-dependent or independent pathways (Nedel et al. 2012). Diphenyldiselenide can perform as a substrate for thioredoxin reductase which gives foundation for the progress the novel diselenides as prospective anticancer agents (De Freitas and Rocha 2011). Later diverse sequence of diselenides containing bioactive ligands (e.g., naphthalene, cyclic imides) or medically relevant heterocycles (e.g., thiazolidinone, tetrazole, pyrazole and thiazolopyrimidine) are developed and few showed cytotoxicity at sub-micromolar concentrations against many cancer cell types (Shaaban et al. 2015; Abbas et al. 2015). These compounds induce cytotoxicity by down regulation of Bcl-2, Ki-67 expression level and also by activated caspase-8 (Shaaban et al. 2016).

**Selenazofurin**

Selenazoles are characterized by five-membered rings with the presence of selenium and one methylene group replaced by nitrogen. Selenazofurin are the most studied compound belonging to this class of molecules. In 1983 selenazofurin (2-β-n-ribofuranosylselenazole-4-carboxamide) was synthesized, which is the Se analog of tiazofurin (Srivastava and Robins 1983). It was shown initially that this compound possesses a noticeable antitumor activity against murine melanoma cancers in in vitro and animal models. It is also likely to exhibit anticancer activity against human promyelocytic leukemia cells in vitro at nanomolar levels (Lucas et al. 1983; Boritzki et al. 1985). Later there are reports on selenazofurin dinucleotides analogues which are found to be more effective than the mononucleotide precursor (Gebeylehu et al. 1985). A new analog called selenophenfurin was developed by replacing selenazole ring with selenophene heterocycle. They are found to be effective against leukemia, lymphoma, and solid tumor cell lines at concentrations alike that of selenazofurin (Franchetti et al. 1997). Selenazofurin as well as its derivatives get metabolized to the selenazole-4-carboxamide-adenine dinucleotides which can perform as effective non-competitive inhibitors of de novo guanine nucleotide biosynthesis like 5-isoquinoline monophosphate dehydrogenase (IMPD) a rate-limiting enzyme.

**Selenocyanates**

Selenocyanates have established their efficacy in the prevention and treatment of variety of cancers both in vitro and in vivo. In specific, phenylalkyl selenocyanates revealed to be effective in in vitro conditions for various cancer cell lines like melanoma, glioblastoma, sarcoma, prostate, breast, and colon. The same compound in in vivo condition was capable of reducing the tumour growth significantly in a melanoma xenograft model, without causing any systemic toxicity or affecting the blood related parameters. The compound was able to affect cancer cell redox state by assessing its cycle in the presence of GSH forming nitric oxide, superoxide and other ROS (Crampsie et al. 2012). There are reports additionally that they effects specific phase II detoxifying and antioxidant enzymes like glutathione-S-transferase, superoxide dismutase and catalase and also selenoenzymes like thioredoxin reductase and glutathione peroxidase (Das et al. 2013). These mechanistic conditions in cells lead to caspase-dependent apoptotic cell death via induction of p53, Bax and suppression of Bcl-2 (Chakraborthy et al. 2015). The compound 2-[5-(selenocyanato-penty1]-7-amino benzo[de] isoquinoline-1,3-dione which is the combined form of
diphenyl-methylselenocyanate and naphthalimide were found to possess effective therapeutic efficacy than the classical chemotherapeutic drugs like cisplatin and cyclophosphamide. Report on the adverse and toxicity effect exhibited by the classical chemotherapeutic drugs has been reduced by this compound (Ghosh et al. 2015). The synthetic compound 1,4-phenylenebis (methylene) selenocyanate (p-XSC) has been found to be effective in vitro even at low micromolar concentrations against androgen responsive and androgen unresponsive human prostate cancers. They does the activity by inducing cell cycle arrest in the G1 phase, stimulating JNK, p38, ERKs 1&2 and Akt cancer cell signalling and preventing the m-TOR downstream effectors (Facompre et al. 2012).

A series of selenocyanate derivatives was developed by combining an indole-heterocycle microtubule inhibitor with a selenocyanate moiety. They found to be very promising in vitro antitumor activity even in low µM concentrations against colon, breast and lung cancers and against melanoma. They do this apoptotic activity through mechanism encompassing both microtubule and Akt inhibition (Krishnegowda et al. 2011). A compound selenocyanate containing triterpenes based on homobetulin scaffold was developed and showed antiproliferative activity in the low µM range against non-tumor cells (Sidoryk et al. 2016).

Selenoesters

Primarily the selenoesters were synthesized on the reason that they can be subjected to hydrolysis or enzymatic reduction in cells which leads to the generation of redox active compounds. These compounds could be readily helps in triggering apoptosis. Many alkyl and aryl selenoesters exhibited readily noticeable cytotoxic activity even in nano-molar concentrations against prostate, breast, lung and colon cancer cell lines (Dominguez-Alvarez et al. 2014).

Among sugar containing selenoesters, xylitol and sucrose based selenious ester have been produced. The xylitol-seleno ester has revealed apoptotic conditions in human hepatoma cells in a dose-dependent manner by substituting GSH depleting agents (Guo et al. 2013a, b; Wu et al. 2016). Unfortunately this cytostatic effect was not detected in normal human hepatic cells. Likewise, the sucrose-seleno ester was able to inhibit the propagation of many cancer cell lines in a dose-dependent manner without affecting the normal human liver cells. Further sucrose-seleno ester showed acute toxicity in mice with very high median lethal dose compared to sodium selenite (290.0 and 13.1 ppm, respectively). This paves the way for a further preclinical development of this type of selenocompounds (Guo et al. 2013a, b).

Selenium-nonsteroidal anti-inflammatory drug (Se-NSAID)

A nonsteroidal anti-inflammatory drug (NSAID) namely sulindac on combination with selenium compound have inhibited intestinal tumours. They reduced the tumours by 52% and tumour multiplicities of 80%. Mainly this happened due to expression of p27 and p53 as well as JNK1 phosphorylation (Bi et al. 2013). In another study Se-containing selenocoxib derivative was established. They have the capability to decrease the cell count in melanoma cells by arresting the cell cycle growth phase of G0-G1. This derivative retains the mechanism of COX-2 inhibitory and PI3K/Akt inhibitory activity which ultimately promotes apoptosis (Gowda et al. 2013). Another most effective compound called SeCN-aspirin analog was developed. They found extremely selective toward cancer cells and cause cell cycle arrest in G1 and G2/M phases. This SeCN-aspirin analog induce apoptosis by activating caspase 3/7 and PARP cleavage in colorectal cancer cells (Plano et al. 2016).

Selenoureas

Selenourea derivatives along with their corresponding thio-urea analogues were synthesized. They were assessed for their antiproliferative effects in many human cancer cell lines. The study exhibited higher potency with IC₅₀ values below 10 µM in all cell lines tested and then further evaluated their capability with different colon cancer cell models. The compound assists the activation of caspase-dependent pathways and also helps in inhibition of cancer causing proteins. Nearly 17 ferrocenyl-selenoureas have been developed and they were studied for their DNA binding properties, antioxidant activity, cytotoxicity in neuroblastoma cell lines like MYCN2, SK-N-SH, liver hepatocarcinoma cell line Hepa1c1c7, and breast cancer cell line MCF-7. Most of the compounds exhibited cytotoxicity towards cancer cell lines (90%) with low µM concentrations (Hussain et al. 2015). Another selenourea derivative was tested in vivo with prostate cancer mouse xenograft model and it suppressed tumor growth around 80% without any troublesome effects (Ibanez et al. 2012).

Selenocarbonyl derivatives

Several carbonyl-containing compounds have been chemically transformed into selone analogue derivatives and were tested for their biological activities in different models. The selone analogue of chrysin and tetramethylquercetin has been synthesized and this compound exhibited better antioxidant activity, cytotoxicity and in specific the IC₅₀ in MCF-7 cells was lowered (Martins et al. 2015). Selenohydantoin another compound was evaluated in vitro against many
cancer cell lines and they showed excellent anti-proliferative activities in low μM concentrations (Ivanenkov et al. 2016).

Selenium nanoparticles for cancer treatment

Currently the selenium nanoparticles have gained much interest as potential novel therapeutic mediators, because of their excessive biocompatibility, constancy and distinct selectivity. They are generally synthesized by three major routes (1) chemical based (2) biological and (3) physical (Maiyo and Singh 2017). Between them, chemical synthesis is considered the most common method to prepare selenium nanoparticles. In chemical synthesis, selenium in the form of selenite, selenious acid, or selenium dioxide were frequently used as precursors, reducing agents like ascorbic acid and glutathione are employed and stabilizing agents like chitosan and pectin helps in the selenium nanoparticles formation and maintenance (Qiu et al. 2018; Zhai et al. 2017). Synthesized selenium nanoparticles in the sodium selenite/glutathione (Na₂SeO₃–GSH) redox system are inclined to gather in cancer cells and to present stronger pro-oxidant activity (Zhao et al. 2018a, b). For optimize their function, the chemically synthesized selenium nanoparticles are usually decorated with other bioactive molecules which can enhance the therapeutic effects upon certain types of cancers when compared with non-decorated selenium nanoparticles. Apart from direct therapeutic effects, chemically altered selenium nanoparticles can act as vehicles that endow the carried objects with favourable properties like low toxicity, high efficacy and tumor targeting. Markedly, chemically synthesized selenium nanoparticles were also observed as diagnostic agents, imaging mediators, and radio-sensitizers (Yang et al. 2017; Chen et al. 2018a, b; Zhao et al. 2018a, b; Abd-Rabou et al. 2019).

On comparison with chemically synthesized selenium nanoparticles, biologically synthesized nanoparticles appear to be eco-friendly, biocompatible, sustainable, economical and safer too (Wadhwani et al. 2016). These nanoparticles are synthesized using extracts or media from plant sources, bacteria, fungi and other organisms. The synthesized selenium nanoparticles exhibit varied biological activities like preventing DNA damage by UVB-induced and inhibiting the proliferation of cancer cells (Prasad et al. 2013). Until now, bacteria have been the most important source of biologically synthesized nanoparticles. Bacterial source which helps in the fabrication of selenium nanoparticles includes Bacillus licheniformis JS2, Ochrobactrum sp. MPV1, Streptomyces minitisceroticus M10A62 and Acinetobacter sp. SW30 (Tugarova and Kamnev 2017; Ramya et al. 2015). Synthesis of selenium nanoparticles using bacteria is done by culturing the strains in the growth media containin sodium selenite at a concentration range from 0.5–2 mM. Under the stress condition of sodium selenite concentration (0.5 or 2 mM), these Ochrobactrum sp. MPV1 bacteria are capable of converting selenite to elemental Se by which selenium nanoparticles synthesis occur intra-cellularly and rather their medical applications remains unexplored (Zonaro et al. 2017).

In another study selenium nanoparticles synthesized from Bacillus licheniformis strain JS2 have the capability to initiate necroptosis in PC-3 cell by the ROS-mediated activation controlled via Receptor-interacting protein kinase 1 (RIP1 kinase) (Saratate et al. 2018; Sonkusre and Cameotra 2017).

Biological synthesis of selenium nanoparticles using Plant sources is another interesting feature. By using simple refluxing method uniform size (3–18 nm) and shaped, selenium nanoballs were synthesized from dried Vitis vinifera (raisin) extracts and selenous acid (Sharma et al. 2014). These selenium nanoparticles synthesized from variety of plants sources has enormous potential in future though the exact mechanism of synthesis and their therapeutic applications remains mostly underexplored.

Synthesis of selenium nanoparticles using physical methods have been carried using pulsed laser ablation and gamma radiation (Quintana et al. 2002). Previous study showed that pure selenium nanoparticles synthesized by pulsed laser ablation in liquids were able to disturb the biofilm formation of a human pathogen Candida albicans, which highlights its potential for medical application. But still the application of these selenium nanoparticles synthesized by physical methods for cancer treatment and prevention is still undeveloped, which may be due to the equipment requirements (Guisbiers et al. 2017).

Selenium nanoparticles have great prospective not only in cancer treatment but also as diagnostic/imaging agents and more. Selenium nanoparticles exhibited anticancer effects in a series of cancers like hepatocarcinoma, breast cancer, colon adenocarcinoma, lymphoma, esophageal cancer, prostate cancer, ovarian cancer, and glioma (Maiyo and Singh 2017). Further studies and clinical trials are required to explain the possible applications of selenium nanoparticles in cancer treatment.

Conclusion and future perspectives

For the past few decades, research on the elemental selenium has reached its remarkable heights. Particularly its role in the synthesis of various types of selenoproteins and many regulatory metabolic pathways starting from the plant source to human’s mechanistic approach. Throughout the years, many novel selenoproteins have been identified and annotated its importance in various aspects. This review is mainly focused on some of the functions of selenoproteins and their part in wound healing mechanism. Some of the predictable or possible routes like translational decoding of selenocysteine, biosynthesis of selenoproteins, systemic
action of selenoproteins and their direct or indirect assimilation of selenoproteins in the process of wound healing are discussed. In near future those predicted routes can be experimentally proved and may help in resolve the mystery about the role of selenium in wound healing process. Selenium compounds have been reasonably beneficial in cancer treatment and diagnosis. Recent day’s selenium nanoparticle has attracted considerable attention against various types of cancers. They will be playing critical role in cancer prevention and therapy in the near future. Many new selenium drugs pertaining to inflammations and other deficiencies/diseases may also arise to prove the importance of selenium to the medical and pharmaceutical world.

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**Compliance with ethical standards**

**Conflict of interest** The authors declare that they have no competing interest.

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