Quantitative progression of retinitis pigmentosa by optical coherence tomography angiography

Ruben Jauregui1,2,3, Karen Sophia Park1,2, Jimmy K. Duong4, Vinit B. Mahajan5 & Stephen H. Tsang1,2,6

Optical coherence tomography angiography (OCT-A) is a non-invasive alternative to fluorescein angiography that allows for the study of the retinal and choroidal vasculatures. In this retrospective cohort study of 28 patients with retinitis pigmentosa (RP), we used OCT-A to quantify changes in perfusion density, foveal avascular zone (FAZ) area, and choriocapillaris blood flow over time and correlated these variables with ellipsoid zone (EZ) line width and best-corrected visual acuity (BCVA). Perfusion density decreased by $2.42 \pm 0.62\%$ per year at the superior capillary plexus (SCP) ($P = 0.001$) and $2.41 \pm 0.76\%$ per year at the deep capillary plexus (DCP) ($P = 0.004$). FAZ area increased by $0.078 \pm 0.021$ mm$^2$ per year ($P = 0.001$) at the SCP and $0.152 \pm 0.039$ mm$^2$ per year ($P = 0.001$) at the DCP. No changes were observed in the choriocapillaris blood flow. EZ line width had the strongest correlation to perfusion density at the SCP ($r = 0.660$ and $0.635$, first and second visit, respectively, $P = 0.001$), while BCVA most strongly correlated with FAZ area at the SCP ($r = 0.679$ and $0.548$, $P = 0.001$ and $0.003$). Our results suggest that OCT-A is a useful tool for monitoring RP disease progression and may be used to measure retinal vascular parameters as outcomes in clinical trials.

Retinitis pigmentosa (RP) refers to a heterogeneous group of inherited rod-cone retinal dystrophies characterized by progressive visual field constriction and nyctalopia1–3. Its prevalence is estimated to be 1 in 4,000 people worldwide, while the vast majority of cases are inherited in an autosomal dominant, autosomal recessive, or X-linked manner1,11. With over 50 causative genes known to date, RP exhibits significant clinical and genetic heterogeneity, as a single mutation may cause a variety of clinical phenotypes and a variety of different mutations may cause the same syndrome4. The primary defect lies in the rod photoreceptors, which degenerate and lead to secondary cone cell death. Throughout the course of the disease, the retinal pigment epithelium and blood vessels are also affected, leading to additional clinical hallmarks of the disease such as attenuation of retinal vessels and intraretinal pigment migration.

With the advent of optical coherence tomography angiography (OCT-A), the study of the retinal and choroidal vasculatures has become more feasible. OCT-A serves as a non-invasive and ideal alternative to fluorescein angiography, as it not only is faster to obtain, but also avoids potential side effects of fluorescein angiography such as vomiting and hypersensitivity reactions5. OCT-A detects streaming blood flow and constructs an image of the retinal vasculature, allowing for the visualization of the superficial (SCP) and deep capillary plexus (DCP)5. The choriocapillaris is also visualized, but the small size and intersinusoidal spacing of its blood vessels cause the choriocapillaris to appear homogenous with bright areas representing blood flow6. The technology of OCT-A has been applied broadly to study vasculature changes in inherited retinal dystrophies, including RP, Stargardt disease, and choroideremia7–11.

It has previously been reported that perfusion density, defined as the total area of perfused vasculature per unit area in a region of measurement (also referred to as vessel density in some studies12), is decreased while the area of the foveal avascular zone (FAZ) is increased in patients with RP compared to controls5. This study aims...
to analyze and quantify changes in the retinal vasculature of patients with RP over time. In addition, we correlate these changes with the width of the ellipsoid zone (EZ) line, which is related to the size of a patient’s field of vision, and best-corrected visual acuity (BCVA). This work could not only have implications on the development of therapies for RP, but it could also establish the use of perfusion density and FAZ area as outcome measures for clinical trials and disease progression.

Results

Patients. In total, 28 patients (28 eyes) were analyzed for this study. Demographic characteristics of the patients are included in Table 1. The mean follow-up time was 1.3 ± 0.46 years. Complete descriptive statistics can be found in Supplementary Table S2.

Progression rates in the retinal and choroidal vasculatures. We observed a progression rate with quantitative OCT-A analysis in perfusion density at both the level of the SCP and DCP between the two visits. The perfusion density decreased over time at a rate of 2.42 ± 0.62% per year at the SCP (P = 0.001) and 2.41 ± 0.76% per year at the DCP (P = 0.004). We also observed a rate of progression in the FAZ area at both the SCP and DCP; at the SCP, the FAZ area increased at a rate of 0.078 ± 0.21 mm² per year (P = 0.001), while it increased at a rate of 0.152 ± 0.039 mm² per year at the DCP (P = 0.001). There was no significant progression rate observed for choriocapillaris blood flow (1.36 ± 1.23, P = 0.275). A rate of progression was also measured in the width of the EZ line, which decreased at a rate of 107.03 ± 13.67 μm per year (P < 0.001). BCVA decreased at a logMAR rate of 0.049 ± 0.021 per year (P = 0.026). These results are summarized in Table 2.

Vascular differences between the superior and deep capillary plexus in the retina. The mean perfusion density at the SCP was compared against the mean perfusion density at the DCP for both visits, and their respective progression rates were also compared. Similar comparisons were performed for the FAZ area. At both visit 1 and 2, the mean perfusion density at the SCP (33.4 ± 11.1% and 30.4 ± 11.1% for visit 1 and 2, respectively) was observed to be different from the mean perfusion density at the DCP (24.5 ± 9.69 and 21.5 ± 9.57% for visit 1 and 2, respectively) (P < 0.001 for both visits). Furthermore, at both visits, the mean FAZ area at the SCP (0.345 ± 0.226 mm² and 0.430 ± 0.292 mm² for visit 1 and 2, respectively) was observed to be different from the mean FAZ area at the DCP (0.784 ± 0.389 mm² and 0.944 ± 0.447 mm² for visit 1 and 2, respectively) (P < 0.001 for both visits). No difference was seen between the progression rate of perfusion density at the SCP (−2.42 ± 0.62%) and that at the DCP (−2.41 ± 0.76%) (P = 0.986). Similar results were observed between the progression rate of FAZ area at the SCP (0.078 ± 0.021 mm²) and that at the DCP (0.152 ± 0.039 mm²) (P = 0.053). These results are summarized in Table 2.

Correlations between vascular parameters in the retina and measures of visual function. EZ line width and BCVA were both correlated with perfusion density and FAZ area at the levels of the SCP and DCP as well as with choriocapillaris blood flow at both clinic visits (Table 3). EZ line width was observed to have a correlation with only perfusion density at the SCP and DCP, with the strongest correlation observed at the SCP (r = 0.660 and 0.635, P = 0.001 and 0.001 for the first and second visit, respectively). BCVA correlated with both perfusion density and FAZ area, but the strongest correlation was observed with FAZ area at the SCP (r = 0.679 and 0.548, P = 0.001 and 0.003 for the first and second visit, respectively). Choriocapillaris blood flow was not correlated with either EZ line width (r = −0.105 and −0.161, P = 0.594 and 0.413 for the first and second visit, respectively) or BCVA (r = −0.031 and 0.155, P = 0.876 and 0.432 for the first and second visit, respectively). These results are summarized in Table 3.

| Inheritance | Genes with disease-causing variants (No. of patients) |
|-------------|-----------------------------------------------------|
| ARRP        | FDE6A (2), USH2A (2), EYS (2), DHDDS (1), CERKL (1), KIZ (1), MERTK (1), TULP1 (1), Unknown (6) |
| ADRP        | RPI (3), RHO (2), KLLH7 (2), PRPF8 (1) |
| USH         | MYO7A (2), GPR98 (1) |

Table 1. Demographic and genetic characteristics of the retinitis pigmentosa patients. Data are summarized as mean ± standard deviation where appropriate. ARRP = autosomal recessive retinitis pigmentosa; ADRP = autosomal dominant retinitis pigmentosa; USH = Usher syndrome; CME = cystoid macular edema.
FAZ area at the DCP was significantly enlarged in RP patients. In our study, we found that at any given visit, the velocities have also been found to be lower in patients with RP compared to the control group. Recently, studies have confirmed by means of OCT-A that there are vascular alterations in patients with RP. OCT-A has ultimately allowed us to expand our knowledge regarding vascular changes in RP in a non-invasive manner.

Changes in the retinal vasculature and hemodynamics have long been associated with RP. By fundoscopy, attention of the retinal nerve fiber layer, is also reduced in RP patients as compared with normal controls. The advent of OCT-A has ultimately allowed us to expand our knowledge regarding vascular changes in RP in a non-invasive manner.

**Discussion**

Changes in the retinal vasculature and hemodynamics have long been associated with RP. By fundoscopy, attention of the retinal nerve fiber layer, is also reduced in RP patients as compared with normal controls. The advent of OCT-A has ultimately allowed us to expand our knowledge regarding vascular changes in RP in a non-invasive manner.

**Table 2.** Quantitative analyses of perfusion density, foveal avascular zone area, choriocapillaris blood flow, EZ line width, and BCVA at each visit and their yearly progression rate. Data are summarized as mean ± standard deviation where appropriate. FAZ = foveal avascular zone; SCP = superior capillary plexus; DCP = deep capillary plexus; EZ = ellipsoid zone; BCVA = best-corrected visual acuity; logMAR = logarithm of the minimal angle of resolution; *r* = Pearson correlation coefficient. *P*-values indicating statistical significance are italicized and bold.

| Variable                        | Visit 1                      | Visit 2                      | Visit 1                       | Visit 2                       | Visit 1                       | Visit 2                       | Visit 1                       | Visit 2                       | Visit 1                       | Visit 2                       | Visit 1                       | Visit 2                       | Visit 1                       | Visit 2                       |
|---------------------------------|------------------------------|------------------------------|-------------------------------|-------------------------------|-------------------------------|-------------------------------|-------------------------------|-------------------------------|-------------------------------|-------------------------------|-------------------------------|-------------------------------|-------------------------------|-------------------------------|
| Perfusion density, mean (%)     |                              |                              |                              |                              |                              |                              |                              |                              |                              |                              |                              |                              |                              |                              |
| SCP                             | 33.4 ± 11.1                  | 30.4 ± 11.1                  | 21.5 ± 9.57                  | −0.42 ± 0.62                  | 0.001                         | 0.986                         |                              |                              |                              |                              |                              |                              |                              |                              |
| DCP                             | 24.5 ± 9.69                  | 21.5 ± 9.57                  | −2.41 ± 0.76                  | 0.004                         |                              |                              |                              |                              |                              |                              |                              |                              |                              |
| FAZ area, mean (mm²)            |                              |                              |                              |                              |                              |                              |                              |                              |                              |                              |                              |                              |                              |                              |
| SCP                             | 0.345 ± 0.226                | 0.430 ± 0.292                | 0.944 ± 0.447                | 0.078 ± 0.021                 | 0.001                         | 0.053                         |                              |                              |                              |                              |                              |                              |                              |
| DCP                             | 0.784 ± 0.389                | 0.944 ± 0.447                | 0.152 ± 0.039                | 0.001                         |                              |                              |                              |                              |                              |                              |                              |                              |                              |
| Choriocapillaris blood flow     |                              |                              |                              |                              |                              |                              |                              |                              |                              |                              |                              |                              |                              |                              |
| Mean gray value                 | 99.5 ± 11.1                  | 98.8 ± 13.0                  | 1.36 ± 1.23                  | 0.275                         |                              |                              |                              |                              |                              |                              |                              |                              |                              |
| EZ line width, mean (μm)        | 2674.9 ± 1766.8              | 2533.2 ± 1755.9              | −107.03 ± 13.67              | <0.001                        |                              |                              |                              |                              |                              |                              |                              |                              |                              |
| BCVA, mean (logMAR)             | 0.28 ± 0.30                  | 0.33 ± 0.33                  | 0.049 ± 0.021                | 0.026                         |                              |                              |                              |                              |                              |                              |                              |                              |                              |

**Table 3.** Correlations of EZ line width and BCVA with perfusion density, foveal avascular zone areas, and choriocapillaris blood flow. FAZ = foveal avascular zone; SCP = superior capillary plexus; DCP = deep capillary plexus; EZ = ellipsoid zone; BCVA = best-corrected visual acuity; logMAR = logarithm of the minimal angle of resolution; *r* = Pearson correlation coefficient. *P*-values indicating statistical significance are italicized and bold.

| EZ line width (μm) | Perfusion density at SCP | FAZ area at SCP | Perfusion density at DCP | FAZ area at DCP | Choriocapillaris |
|--------------------|--------------------------|-----------------|--------------------------|-----------------|------------------|
|                    | Visit 1 | Visit 2 | Visit 1 | Visit 2 | Visit 1 | Visit 2 | Visit 1 | Visit 2 | Visit 1 | Visit 2 | Visit 1 | Visit 2 |
| *r*                |         |        |        |        |        |        |        |        |        |        |        |        |
|                   | 0.660   | 0.635  | −0.277 | −0.274 | 0.537  | 0.552  | 0.073  | 0.157  | −0.105 | −0.161 |
| *P*-value          | 0.991   | 0.001  | 0.001  | 0.001  | 0.003  | 0.002  | 0.713  | 0.424  | 0.594  | 0.413  |
| BCVA (logMAR)      |         |        |        |        |        |        |        |        |        |        |        |        |
| *r*                |         |        |        |        |        |        |        |        |        |        |        |        |
|                   | −0.434  | −0.403 | 0.679  | 0.548  | −0.472 | −0.374 | 0.383  | 0.272  | −0.031 | 0.155  |
| *P*-value          | 0.021   | 0.034  | 0.001  | 0.003  | 0.011  | 0.050  | 0.044  | 0.161  | 0.876  | 0.432  |
When analyzing blood flow at the choriocapillaris, no significant progression rate was observed between the two visits ($P = 0.275$). This result is in agreement with previous studies that have reported no difference in both the perfusion density of the choriocapillaris and the blood flow rate between RP patients and controls. A study by Li et al. compared the retinal and choroidal blood flow in RP mouse models; although blood flow was decreased in both retinal and choroidal circulation compared to controls, choroidal blood flow changes were seen much later than retinal blood changes. Results from our data and previous studies likewise suggest that changes in the choroidal vasculature occur later compared to those in the retinal vasculature. Thus, longer follow-up may be needed to observe changes in the choriocapillaris and choroidal blood flow in RP patients. Given that the primary lesion in RP exists in either photoreceptors or the RPE, choroid hemodynamics may become affected during the late stages of disease.

Various non-invasive imaging techniques are used to monitor the progression of RP. One common technique involves measurement of the EZ line width. The EZ line correlates with the patient's visual field boundaries. As such, multiple studies have used SD-OCT to visualize and measure EZ line width as a means of tracking disease progression. Another imaging modality used to measure RP progression is short-wavelength fundus autofluorescence, which often reveals a ring of hyperautofluorescence in patients with RP. This ring constricts over time, and the inner border has been found to correspond spatially with the EZ line. In this study, we observed an annual decline of $107.03 \pm 13.67 \mu m$ in EZ line width, a rate similar to what has previously been published. In correlating the EZ line width to the variables we analyzed via OCT-A, we found that EZ line width correlated only to perfusion density at both the SCP and DCP. EZ line width did not correlate with FAZ area or choriocapillaris blood flow. Grunwald et al. had suggested previously that degeneration of the highly metabolic, oxygen-consuming photoreceptors leads to increased levels of oxygen in the inner retina, causing a vasoconstriction regulatory response in the retinal vasculature. Further recent studies strengthen this hypothesis, as endothelin-1, a potent vasoconstrictor, has been shown to be significantly increased in the eyes of RP patients. We believe that as disease progresses and photoreceptors degenerate, the EZ line, which is formed mainly by photoreceptor mitochondria, decreases in length, leading to increased vasoconstriction in the retinal vessels. This change appears as decreased perfusion density on OCT-A.

We also correlated BCVA with the variables analyzed by OCT-A. We found that BCVA correlates the strongest with the FAZ area at the SCP ($r = 0.679$ and $0.548$, $P = 0.001$ and 0.003 for the first and second visit, respectively). This is similar to the results found in another study where OCT-A was used to estimate retinal blood flow in RP patients. The authors found that BCVA correlated the strongest with the FAZ area at the SCP and with the paravascular flow density in the DCP. In our study, we measured perfusion density in place of blood flow at the SCP and DCP. Of note, BCVA also correlated with perfusion density at both the SCP and DCP in our study. Nevertheless, the strongest correlation was observed with the FAZ area at the SCP. Other studies have also compared morphological vascular changes in RP and correlated them to functional parameters. In a study with RP patients by Toto et al., for example, macular capillary density was imaged with OCT-A and correlated with macular function as measured by multifocal electroretinogram (mERG). The authors found that the density of vessels in the choroid and retina was decreased, which correlated with a decline in ganglion cell complex layer thickness and macular function based on mERG.

Although there is no current treatment for RP, different treatment modalities such as gene therapy, neurotrophic growth factors, and retinal prostheses are being studied and have shown promising and encouraging results. Regardless of the treatment modality, however, a healthy vascular supply is needed to maintain and support the various retinal cells, including photoreceptors and cells of the inner retina. Any extent of degeneration of the retinal and choroidal vasculatures may limit the impact of these promising therapies. For this reason, it is important to study how these vasculatures change throughout the course of RP.

Limitations to this study include the relatively short length of follow-up and the inherent disadvantages of using OCT-A. The average length of time between visits in this study was 1.3 years; given the recent Food and Drug Administration (FDA) approval of OCT-A in 2015, it is challenging to gather patients with longer follow-up that qualify for the study. Furthermore, given that our study only includes two time points per patient, our data appears to indicate that the OCT-A parameters decline in a linear fashion. Future studies with longer follow-up and a greater number of time points should address whether these OCT-A parameters indeed decline linearly throughout time, as the inclusion of more time points may instead reveal that they in fact fit an exponential regression model. Finally, in order to produce an accurate analysis, only patients with high-quality OCT-A scans were analyzed. Due to the fact that the acquisition of high-quality OCT-A images is heavily dependent upon the patient’s ability to fixate, most patients with advanced RP were excluded from the study as they lack fixation due to poor vision. This limits the possibility of studying changes in the retinal and choroidal vasculature in patients with advanced RP, a stage where one might observe significant changes.

To our knowledge, our study is the first to follow RP progression through OCT-A over time. We observed that at the SCP perfusion density decreases at a rate of $2.42 \pm 0.62%$ per year, while the FAZ area increases at a rate of $0.078 \pm 0.021 \text{mm}^2$ per year. At the DCP, perfusion density decreases at a rate of $2.41 \pm 0.76%$ per year, while the FAZ area increases at a rate of $0.152 \pm 0.039 \text{mm}^2$ per year. In addition, we found that the EZ line width had a significant correlation with perfusion density at both the SCP and DCP, although it correlated the strongest at the SCP. Moreover, we found that BCVA correlated the strongest with the FAZ area at the SCP. This study is significant because we observe progressive vascular changes in the retina, which could have implications on emerging therapies for RP. Furthermore, perfusion density and FAZ area could be implemented as important variables and measures of progression in future clinical trials.

**Methods**

**Patients and Clinical Examination.** All study procedures were defined and informed patient consent for study participation was obtained as outlined by the protocol #AAAR0284 approved by the Institutional Review Board at Columbia University Medical Center. The study adhered to the tenets of the Declaration of Helsinki. None of the data presented in this study, including images and genetic testing results, is identifiable to individual
patients. A retrospective review of 100 patients with a clinical diagnosis of retinitis pigmentosa by an inherited retinal disease specialist (SHT) was conducted at the Department of Ophthalmology at Columbia University. The clinical diagnosis was made based on presenting symptoms, family history, fundus examination, and full-field electroretinography (ffERG) and subsequently supported by clinical imaging and/or genetic testing. The inclusion criteria for this study were the diagnosis of RP along with clear media and adequate fixation to allow for high-quality imaging. In addition, each patient was screened for a history of two visits in our office at least 6 months apart consisting of a complete ophthalmic examination by a retinal physician (SHT). Ophthalmic examinations included a slit-lamp and dilated funduscopic examination, best corrected visual acuity (BCVA), fundus autofluorescence (FAF, 488 nm excitation), spectral domain optical coherence tomography (SD-OCT), and OCT angiography (OCT-A). The exclusion criteria precluded patients affected by any other ocular disorder or an advanced form of RP. Furthermore, eyes with poor OCT-A images that exhibited a signal strength index lower than 7 out of 10 were excluded from analysis. One eye from each patient was chosen for analysis based on image quality and inclusion/exclusion criteria.

Clinical Characterization. Imaging across all modalities was conducted after pupil dilation (>7 mm) with phenylephrine hydrochloride (2.5%) and tropicamide (1%). Horizontal foveal SD-OCT scans measuring 9 mm and fundus autofluorescence (FAF, 488 nm excitation) were acquired with the Spectralis HRA + OCT (Heidelberg Engineering, Heidelberg, Germany). OCT-A 3 x 3 mm scans centered on the fovea were obtained using the Zeiss AngioPlex Cirrus HD-OCT 5000 (Zeiss Meditec, Inc, Dublin, California, USA). This instrument has an A-scan rate of 68 KHz scans and wavelength-scanning light of 840 nm. Each OCT-A contains 245 B-scans (each B-scan contains 245 A-scans). To image the motion of moving erythrocytes, 4 OCT scans are performed at the same location, assisted by eye tracking technology. Automated segmentation of full-thickness retina scans into superficial and deep inner retinal vascular (capillary) plexus (SCP and DCP, respectively) and choriocapillaris by the machine was performed. The SCP is bounded by the internal limiting membrane and the inner plexiform layer, containing vasculatures from the nerve fiber, ganglion cell, and inner plexiform layers. The DCP is bounded by the inner plexiform and outer plexiform layers, containing vasculatures from the inner nuclear and outer plexiform layers.
Image Analysis. The analysis of the OCT-A images was performed as previously described. All of the 3 × 3 mm OCT-A images were collected and exported from the system as a Joint Photographic Experts Group (JPEG) file into the National Institutes of Health Image 1.8.0 (National Institutes of Health, Bethesda, Maryland, USA) image-processing software. The image was converted from 8-bit into red green blue (RGB) color type and was subsequently split into the three channels. We chose the red channel as reference as previously performed. The adjust threshold tool was set to default and applied, with the dark-background option selected. This tool sets lower and upper threshold values (60–255 in this study) and segments the grayscale images into features of interest and background. After processing, the images were converted to RGB. The foveal avascular zone (FAZ) area was manually outlined using the free-hand selection tool, and its dimensions were expressed in millimeter squared using a method previously described. The FAZ area was colored in blue. Black pixels were considered vessels, white pixels were background, and blue pixels were the FAZ area. Perfusion density, defined as the total area of perfused vasculature per unit area in a region of measurement, was calculated as the ratio of black pixels to the total number of pixels, with the blue FAZ area pixels excluded. This method was used to calculate perfusion density in both the SCP and DCP (Fig. 1). To analyze the choriocapillaris, we used the mean gray value (MGV) of the unprocessed grayscale 8-bit OCT-A images of the choriocapillaris as an objective proxy for blood flow as previously described. A high degree of blood flow through the choriocapillaris is depicted by high MGV and image brightness. SD-OCT scans were acquired as a single horizontal slice through the fovea per patient, and measurement of the ellipsoid zone (EZ) line width from these scans was performed. Both perfusion density image analysis and EZ line width measurements were performed by two independent graders (RJ and KSP).

Statistical analyses. The statistical analyses were performed using the Stata 12.1 (StataCorp, College Station, Texas, USA) software. The Pearson correlation was calculated for the measurements of both independent graders (see Supplementary Table S1). Given the high correlation between the two graders, the average of the two values obtained from each of the graders was calculated and used for subsequent analysis. Statistical analysis included descriptive statistics for demographics, logMAR BCV A, EZ line width, and OCT-A measurements (perfusion density for the SCP and DCP, FAZ area for the SCP and DCP, and choriocapillaris blood flow) for both visits. Change over time, defined as the difference in values obtained between the follow-up and baseline visits divided by the length of follow-up, was calculated for these variables. One-sample Student’s t-test was used to determine whether the mean change-over-time variables were different from 0. A paired Student’s t-test was used to compare the mean values at each visit and the perfusion density and FAZ area progression rates between the SCP and DCP. Individual bivariate analyses were performed to correlate each OCT-A measurement value with the EZ line width and logMAR BCV A. In all of the statistical analyses, we defined statistical significance as a P-value of less than 0.05.

Data Availability Statement
The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

References
1. Hartong, D. T., Berson, E. L. & Dryja, T. P. Retinitis pigmentosa. *Lancet* **368**, 1795–1809, [https://doi.org/10.1016/S0140-6736(06)69740-7](https://doi.org/10.1016/S0140-6736(06)69740-7) (2006).
2. Hamel, C. Retinitis pigmentosa. *Orphanet journal of rare diseases* **1**, 40, [https://doi.org/10.1186/1750-1172-1-40](https://doi.org/10.1186/1750-1172-1-40) (2006).
3. Ferrari, S. et al. Retinitis pigmentosa: genes and disease mechanisms. *Curr Genomics* **12**, 238–249, [https://doi.org/10.2174/138920211793860107](https://doi.org/10.2174/138920211793860107) (2011).
4. Daiger, S. P., Bowe, S. J. & Sullivan, L. S. Perspective on genes and mutations causing retinitis pigmentosa. *Arch Ophthalmol* **125**, 151–158, [https://doi.org/10.1001/archophthalmol.125.2.151](https://doi.org/10.1001/archophthalmol.125.2.151) (2007).
5. Munk, M. R. et al. OCT-angiography: A qualitative and quantitative comparison of 4 OCT-A devices. *Plos One* **12**, e0177059, [https://doi.org/10.1371/journal.pone.0177059](https://doi.org/10.1371/journal.pone.0177059) (2017).
6. Al Sheikh, M. et al. Quantitative Features of the Choriocapillaris in Healthy Individuals Using Swept-Source Optical Coherence Tomography Angiography. *Ophthalmic Surg Lasers Imaging Retina* **48**, 623–631, [https://doi.org/10.3928/23258160-20170802-04](https://doi.org/10.3928/23258160-20170802-04) (2017).
7. Battaglia Parodi, M. et al. Vascular abnormalities in patients with Stargardt disease assessed with optical coherence tomography angiography. *Br J Ophthalmol* **101**, 780–785, [https://doi.org/10.1136/bjophthalmol-2016-308869](https://doi.org/10.1136/bjophthalmol-2016-308869) (2017).
8. Battaglia Parodi, M. et al. Vessel density analysis in patients with retinitis pigmentosa by means of optical coherence tomography angiography. *Br J Ophthalmol* **101**, 428–432, [https://doi.org/10.1136/bjophthalmol-2016-308925](https://doi.org/10.1136/bjophthalmol-2016-308925) (2017).
9. Samara, W. A. et al. Correlation of Foveal Avascular Zone Size with Foveal Morphology in Normal Eyes Using Optical Coherence Tomography Angiography. *Retina* **35**, 2188–2195, [https://doi.org/10.1097/IAE.0000000000000847](https://doi.org/10.1097/IAE.0000000000000847) (2015).
10. Iafe, N. A., Phaukkijwatana, N., Chen, X. & Sarraf, D. Retinal Capillary Density and Foveal Avascular Zone Area Are Age-Dependent: Quantitative Analysis Using Optical Coherence Tomography Angiography. *Invest Ophthalmol Vis Sci* **57**, 5780–5787, [https://doi.org/10.1167/iovs.16-20045](https://doi.org/10.1167/iovs.16-20045) (2016).
11. Sugahara, M. et al. Optical Coherence Tomography Angiography to Estimate Retinal Blood Flow in Eyes with Retinitis Pigmentosa. *Sci Rep* **7**, 46396, [https://doi.org/10.1038/srep46396](https://doi.org/10.1038/srep46396) (2017).
12. Pawzi, A. A. Consensus on Optical Coherence Tomographic Angiography Nomenclature: Do We Need to Develop and Learn a New Language? *JAMA Ophthalmol* **135**, 377–378, [https://doi.org/10.1001/jamaophthalmol.2017.0149](https://doi.org/10.1001/jamaophthalmol.2017.0149) (2017).
13. Li, Z. Y., Possin, D. E. & Milam, A. H. Histopathology of bone spicule pigmentation in retinitis pigmentosa. *Ophthalmology* **102**, 805–816 (1995).
14. Milam, A. H., Li, Z. Y. & Fariss, R. N. Histopathology of the human retina in retinitis pigmentosa. *Prog Retin Eye Res* **17**, 175–205 (1998).
15. Li, G. et al. Layer-specific blood-flow MRI of retinitis pigmentosa in RCS rats. *Exp Eye Res* **101**, 90–96, [https://doi.org/10.1016/j.exer.2012.06.006](https://doi.org/10.1016/j.exer.2012.06.006) (2012).
16. Muir, E. R., De La Garza, B. & Duong, T. Q. Blood flow and anatomical MRI in a mouse model of retinitis pigmentosa. *Magn Reson Med* **69**, 221–228, [https://doi.org/10.1002/mrm.24232](https://doi.org/10.1002/mrm.24232) (2013).
17. Beutelspacher, S. C. et al. Retinal blood flow velocity measured by retinal function imaging in retinitis pigmentosa. *Graefes Arch Clin Exp Ophthalmol* **249**, 1855–1858, [https://doi.org/10.1007/s00417-011-1757-y](https://doi.org/10.1007/s00417-011-1757-y) (2011).
18. Toto, L. et al. Macular Features in Retinitis Pigmentosa: Correlations Among Ganglion Cell Complex Thickness, Capillary Density, and Macular Function. *Invest Ophthalmol Vis Sci* 57, 6360–6366, https://doi.org/10.1167/iovs.16-20544 (2016).

19. Mastropasqua, R. et al. Radial Peripapillary Capillary Network in Patients with Retinitis Pigmentosa: An Optical Coherence Tomography Angiography Study. *Front Neurol* 8, 572, https://doi.org/10.3389/fneur.2017.00572 (2017).

20. Cabral, T. et al. Retrospective Analysis of Structural Disease Progression in Retinitis Pigmentosa Utilizing Multimodal Imaging. *Sci Rep* 7, 10473, https://doi.org/10.1038/s41598-017-10473-0 (2017).

21. Sujirakul, T. et al. Multimodal Imaging of Central Retinal Disease Progression in a 2-Year Mean Follow-up of Retinitis Pigmentosa. *Am J Ophthalmol* 160, 786–798 e784, https://doi.org/10.1016/j.ajo.2015.06.032 (2015).

22. Hood, D. C. et al. Method for deriving visual field boundaries from OCT scans of patients with retinitis pigmentosa. *Biomed Opt Express* 2, 1106–1114, https://doi.org/10.1364/BOE.2.001106 (2011).

23. Cai, C. X.; Locke, K. G.; Ramachandran, R.; Birch, D. G. & Hood, D. C. A comparison of progressive loss of the ellipsoid zone (EZ) band in autosomal dominant and X-linked retinitis pigmentosa. *Invest Ophthalmol Vis Sci* 55, 7417–7422, https://doi.org/10.1167/iovs.14-15013 (2014).

24. Duncker, T. et al. Quantitative fundus autofluorescence and optical coherence tomography in best vitelliform macular dystrophy. *Invest Ophthalmol Vis Sci* 55, 1471–1482, https://doi.org/10.1167/iovs.13-13834 (2014).

25. Duncker, T. et al. Comparison of near-infrared and short-wavelength autofluorescence in retinitis pigmentosa. *Invest Ophthalmol Vis Sci* 54, 385–391, https://doi.org/10.1167/iovs.12-11176 (2013).

26. Hood, D. C., Lazow, M. A., Locke, K. G., Greenstein, V. C. & Birch, D. G. The transition zone between healthy and diseased retina in patients with retinitis pigmentosa. *Invest Ophthalmol Vis Sci* 52, 101–108, https://doi.org/10.1167/iovs.10-5799 (2011).

27. Grunwald, J. E., Maguire, A. M. & Dupont, J. Retinal hemodynamics in retinitis pigmentosa. *Am J Ophthalmol* 122, 502–508 (1996).

28. Cellini, M., Strobel, E., Guizi, C. & Campos, E. C. ET-1 plasma levels and ocular blood flow in retinitis pigmentosa. *Can J Physiol Pharmacol* 88, 630–635, https://doi.org/10.1139/Y10-036 (2010).

29. Finzi, A., Cellini, M., Strobel, E. & Campos, E. C. ET-1 plasma levels, choroidal thickness and multifocal electroretinogram in retinitis pigmentosa. *Life Sci* 118, 386–390, https://doi.org/10.1016/j.lfs.2014.04.004 (2014).

30. Banbridge, J. W. et al. Long-term effect of gene therapy on Leber’s congenital amaurosis. *N Engl J Med* 372, 1887–1897, https://doi.org/10.1056/NEJMoa1414221 (2015).

31. MacLaren, R. E. et al. Retinal gene therapy in patients with choroideremia: initial findings from a phase 1/2 clinical trial. *Lancet* 383, 1129–1137, https://doi.org/10.1016/S0140-6736(13)62117-0 (2014).

32. Beltran, W. A. et al. Successful arrest of photoreceptor and vision loss expands the therapeutic window of retinal gene therapy to later stages of disease. *Proc Natl Acad Sci USA* 112, E5844–E5853, https://doi.org/10.1073/pnas.1309914112 (2015).

33. Lipinski, D. M. et al. CNTF Gene Therapy Confers Lifelong Neuroprotection in a Mouse Model of Human Retinitis Pigmentosa. *Mol Ther* 23, 1308–1319, https://doi.org/10.1038/mt.2015.68 (2015).

34. da Cruz, L. et al. Five-Year Safety and Performance Results from the Argus II Retinal Prosthesis System Clinical Trial. *Ophthalmology* 123, 2248–2254, https://doi.org/10.1016/j.ophtha.2016.06.049 (2016).

35. Campbell, J. P. et al. Detailed Vascular Anatomy of the Human Retina by Projection-Resolved Optical Coherence Tomography Angiography. *Sci Rep* 7, 42201, https://doi.org/10.1038/srep42201 (2017).

36. Corvi, F. et al. Reproducibility of Vessel Density, Fractal Dimension, and Foveal Avascular Zone Using Different Optical Coherence Tomography Angiography Devices. *Am J Ophthalmol* 186, 25–31, https://doi.org/10.1016/j.ajo.2017.11.011 (2018).

Acknowledgements

The Jonas Children’s Vision Care and Bernard & Shirlee Brown Glaucoma Laboratory are supported by the National Institutes of Health [P30EY019007, R01EY018213, R01EY026682, R21AG050437], National Cancer Institute Core [5P30CA013696], Foundation Fighting Blindness [TA-NMT-0116-0692-COLU], the Research to Prevent Blindness (RPB) Scientist-Award, unrestricted funds from RPB, New York, NY, USA. R.J. is supported by the RPB medical student eye research fellowship. S.H.T. is a member of the RD-CURE Consortium and is supported by Kobi and Nancy Karp, the Crowley Family Fund, the Rosenbaum Family Foundation, the Tistou and Charlotte Kerstan Foundation, the Schneeweiss Stem Cell Fund, New York State [C029572], and the Gebroe Family Foundation. The sponsor or funding organization had no role in the design or conduct of this research.

Author Contributions

R.J. conceived the study design. R.J. and K.S.P. performed the image analyses. J.K.D. performed the statistical analyses. R.J., K.S.P. and J.K.D. interpreted the data. R.J. and K.S.P. wrote the main manuscript text. V.B.M. and S.H.T. supervised the study and provided resources. All authors reviewed and approved the final version of the manuscript.

Additional Information

Supplementary information accompanies this paper at https://doi.org/10.1038/s41598-018-31488-1.

Competing Interests: The authors declare no competing interests.

Publisher’s note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article’s Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/.

© The Author(s) 2018