A comparison of gene expression signatures from breast tumors and breast tissue derived cell lines

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Cell lines derived from human tumors have historically served as the primary experimental model system for exploration of tumor cell biology and pharmacology. Cell line studies, however, must be interpreted in the context of artifacts introduced by selection and establishment of cell lines in vitro. This complication has led to difficulty in the extrapolation of biology observed in cell lines to tumor biology in vivo. Modern genomic analysis tool like DNA microarrays and gene expression profiling now provide a platform for the systematic characterization and classification of both cell lines and tumor samples. Studies using clinical samples have begun to identify classes of tumors that appear both biologically and clinically unique as inferred from their distinctive patterns of expressed genes. In this review, we explore the relationships between patterns of gene expression in breast tumor derived cell lines to those from clinical tumor specimens. This analysis demonstrates that cell lines and tumor samples have distinctive gene expression patterns in common and underscores the need for careful assessment of the appropriateness of any given cell line as a model for a given tumor subtype.

1. Introduction

Oncologists rely upon clinical information, a morphologic assessment, and to a limited degree, immunohistochemical and molecular markers to classify malignancies into groups that have distinct clinical behavior. It is clear, however, that additional markers and/or technologies are needed for classifying tumors as current methods sometimes fail to accurately predict patient clinical course. In breast cancer for example, tens to hundreds of different genes/proteins have been shown to be of prognostic value; however, many of these markers co-vary, and hence, are not of independent prognostic value. In addition, progress in adopting these markers into clinical practice has been limited both by technical constraints in the number of markers that can be examined efficiently and by the difficulty in comparing and validating studies that use different reagents and clinical sample sets. In breast cancer, only three markers are typically scored for in the clinical setting which include the estrogen receptor (ER), the tyrosine kinase receptor ERBB2/HER2, and an assessment of tumor proliferation index (e.g. Ki-67 labeling fraction) [1].

The advent of modern genomic analysis tools, in particular DNA microarrays, has essentially created a new tool that is capable of collecting thousands of objective observations on clinical samples that can and are being used to characterize tumors and cell lines at a level of definition that was not possible even five years ago [2]. Many groups have begun to use microarrays to measure gene expression in hundreds of tumor samples with the expectation that the genomic scale measurement of gene expression will reveal a novel molecular based classification of malignant cells. In this review, we will first focus on the characterization of breast tissue and tumor derived cell lines using data obtained from cDNA microarrays. These data can be used to 1) identify which cell lines are the best models for different breast tumor subtypes, 2) define molecular signatures that distinguish the biology of different cell lines and tumor types, 3) identify new candidate markers for tumor diagnosis and classification, and 4) identify subtype specific targets for therapeutic intervention.
Table 1

| Cell line name      | Array # | Previous description of cell line and (source)                  |
|---------------------|---------|------------------------------------------------------------------|
| 184A1               | svcc38  | immortal derivative of 184Aa (M. Stampfer)                       |
| 184Aa               | svcc17  | primary HMEC strain Aa (M. Stampfer)                             |
| 184B5               | svcc40  | immortal derivative of 184Aa (M. Stampfer)                       |
| BT-474-ATCC         | svcc128 | ERBB2 and ER positive line (ATCC)                                |
| BT-474-Stanford     | ssvj107 | ERBB2 and ER positive line (Stanford)                            |
| BT-549              | svcc69  | papillary/ductal carcinoma derived (NCL)                         |
| Fibroblast-UTSW     | shav146 | hTERT immortalized stromal cell line (J. Shay/UTSW)              |
| HB2                 | svcc37  | SV40 immortalized breast epithelial line (H.S. Wiley)            |
| HCC-1937            | shaf046 | BRCA1 mutant carcinoma derived line (J. Shay/UTSW)               |
| HME31               | shat023 | primary HMEC strain 31 (J. Shay)                                 |
| HMEC-IFNo           | svcc500 | HMEC-C strain plus IFNα (Clonetics)                              |
| HMEC-C              | svcc94  | primary HMEC strain C (Clonetics)                                |
| HMEC-C CON          | svcc47  | HMEC-C strain 2 days at 100% confluence (Clonetics)             |
| HMS32               | shaf058 | primary breast stromal/fibroblast cell strain (J. Shay)          |
| Hs578T-ATCC         | shac095 | breast carcinosarcoma derived line (ATCC)                        |
| Hs578T-NCI          | svcc110 | breast carcinosarcoma derived line (NCI)                         |
| MCF-10A             | svv008  | non-tumorigenic breast epithelial cell line (ATCC)               |
| MCF-12A             | sham103 | non-tumorigenic breast epithelial cell line (ATCC)               |
| MCF7-NCI            | svcc1299| ER positive line isolated from a pleural effusion (NCI)          |
| MCF7-UCLA           | shat022 | ER positive line from a pleural effusion (UCLA)                  |
| MDA-MB-231-NCI      | svcc73  | ER negative line from a pleural effusion (NCI)                   |
| MDA-MB-231-UTSW     | shaf054 | ER negative line from a pleural effusion (J. Shay)               |
| SK-BR-3             | svcc15  | ERBB2 positive line from a pleural effusion (ATCC)               |
| T47D                | svcc71  | ER positive line from a pleural effusion (ATCC)                  |

2. Classification of breast cell lines

In general, cell lines established from breast tissues have been mainly characterized with respect to their expression of cytokeratins, the estrogen receptor (ER), and ERBB2/HER2 protein [3–5]. For some lines, the histology of xenografts has been compared to the pathology of their tumor of origin in order to confirm that the cell line has conserved features of its parental tumor. As part of our efforts to characterize the phenotypic diversity of human breast tumors, we have measured gene expression using cDNA microarrays in a number of breast tissue derived cell lines [6–8]. In this review, we have re-analyzed previously published data from thirteen cell lines and report new data from three additional independent lines. Furthermore, we explored cell line stability by measuring gene expression in the same cell line obtained from different sources and therefore propagated independently. Lastly, we included some instructive data derived from a normal mammary epithelial cell line treated with interferon, data derived from a confluent normal cell line culture, and a variant of a normal cell line immortalized in vitro. Gene expression in these cell lines was measured using spotted microarrays in comparison to a common reference sample in a manner that has been previously described and that allows all samples to be compared to one another [6,7].

One of the most striking findings from genomic studies of gene expression to date has been that tumors and/or cell lines have common characteristics of biological or clinical importance that can be identified in their patterns of expressed genes. Hierarchical clustering analysis has been used to identify systematic features in the patterns of variation of expression of genes across sample sets [9–12]. The functions of the known genes that are either relatively over-expressed or under-expressed in comparison between samples can give clues as to the differences in biology that are reflected in gene expression patterns [6–8,13–16]. In this breast cell line data set, we selected for analysis, the subset of genes that showed 1) a signal intensity of >70 arbitrary units in both the Cy3 and Cy5 channels, and 2) expression variation of at least 3-fold or more from average for that gene across the sample set in two or more of the 24 total experiments. This criterion selected 1287 genes out of the 8102 total original genes that were both well measured and changed in expression significantly between cell lines. All primary microarray data for the experiments presented here can be obtained from the Stanford Microarray Database at http://genome-www4.stanford.edu/MicroArray/SMD/, and all figures can be seen at our Supplementary Information website at http://genome-www4.stanford.edu/breast_cancer/cell_line_review2001/.

As can be seen in Fig. 1A, hierarchical clustering analysis divided the cell lines into three main dendrogram branches. The first branch on the far left (Red) contained all three of the primary Human
Mammary Epithelial Cell (HMEC) lines, all HMEC immortal derivatives and the non-tumorigenic MCF-12A [17] cell line ostensibly derived from breast epithelium. The center dendrogram branch (Orange) contained both normal fibroblast derived cell lines, a carcinosarcoma derived line (Hs578T [18]), a line ostensibly derived from breast epithelium (MCF-10A [19]), and two lines derived from breast carcinoma specimens (BT-549/Coutinho and Lasfargues 1978, and MDA-MB-231 [20]). The far right dendrogram branch (Blue) contained all cell lines that were thought to be derived from luminal epithelial cells including two estrogen receptor expressing cell lines (MCF-7 [21] and T47D [22]), two ERBB2 over-expressing lines (SK-BR-3/Trempe and Old 1970, and BT-474 [23]), the SV-40 transformed epithelial derived cell line HB2 [5], and the BRCA1 mutant cell line HCC-1937 that was originally isolated by A. Gazdar and colleagues [24]. The biological functions of the sets of genes that were differentially expressed across different “branches” of the cell line dendrogram (Fig. 1B–E) suggested that the gene expression patterns identified cell lines with features that could be related to different types of normal breast cells. The cell lines sorted into those that either expressed HMEC/basal-cell characteristics, those that expressed stromal/mesenchymal-cell-like characteristics or those that expressed luminal-cell characteristics. It should be emphasized that this is an interpretation of the gene expression patterns and that alternative interpretations of these gene expression patterns are possible.

3. Breast basal epithelial cell signature

The group containing all HMEC lines (Red dendrogram branch) was distinguished by the very high expression of a set of genes that contained many markers of normal breast basal-epithelial cells including keratins 5 and 17 (Fig. 1C) [3,4,25,26]. This set also included many genes whose roles in cell physiology distinguish basal from luminal epithelial cells including the production of basal lamina components and interactions with the extracellular matrix (e.g. gamma and alpha-laminin, collagen type-XVII, integrins alpha-3, alpha-6 and beta-4). The cultured basal-like cell lines expressed variable amounts of smooth-muscle-actin but much less relative to the other lines that expressed the “stromal cell” gene expression signature (Fig. 1D). Therefore, these cultured cells appeared to express some, but not all, of the features of so-called “myo-epithelial cells” which are mature smooth-muscle-actin expressing cells that have a functional contractile apparatus [3,5,27]. This “basal” pattern of gene expression was not restricted to HMEC in that this set of genes were moderately expressed in three other lines (MCF-10A, BT-549 and HB2) that also expressed strong stromal-like gene expression signatures and therefore, did not fall into this class by clustering analysis (see below). It should also be noted that even immortal HMEC derivatives, like 184B5 and 184A1, showed the dominant “basal” cell gene expression pattern and not other signature patterns, and hence, this pattern was not dramatically influenced by immortalization or transformation (see Supplementary Information Fig. 3 – http://genome-www.stanford.edu/breast_cell_line_review2001/).

4. Luminal epithelial cell signature

Approximately 60–70% of sporadic breast tumors are estrogen receptor positive and are believed to be derived from breast luminal epithelial cells [1], which can be distinguished by their expression of cytokeratins 8 and 18 and by their location and function in lining breast secretory-ducts. The in vitro study of this cell type has been complicated by the difficulty in maintaining primary cultures of normal estrogen-receptor-positive luminal cells for longer than a few population doublings [28]. Therefore, most in vitro studies on breast luminal epithelial cells have been performed on cell lines derived from primary breast tumors or pleural effusions from breast cancer patients.

The pattern of gene expression that distinguished the luminal-like signature was comprised of a set of genes that were nearly exclusively expressed in all of the luminal like lines while very low to absent levels were seen in all of the other tested lines (Fig. 1E). Contained within this set of genes were genes/proteins that have been previously used to distinguish luminal breast epithelial cells including the estrogen receptor and keratin 8 and 18 [3]. However, two distinct subtypes of cell lines that expressed luminal characteristics could be distinguished, 1) those that expressed high levels of the estrogen receptor and essentially lacked either stromal or basal gene expression signatures (e.g. MCF7, BT-474 and T47D), and 2) those that expressed little or no estrogen receptor but also expressed genes characteristic of the basal signature (HCC-1937 and HB2). SK-BR-3 was unique in this set of cell lines in that it expressed low levels of the estrogen receptor but a strong luminal gene expression signature without the expression of basal cell characteristics.
5. Mesenchymal/stromal cell signature

We have previously shown that a small, but significant, number of cell lines ostensibly of epithelial origins showed patterns of gene expression that were more consistent with characteristics expected of stromal cells (see [6] and http://genome-www.stanford.edu/nci60/). In order to further investigate the gene expression properties of these cell lines we compared them to two cell lines explicitly derived from breast stroma (HMS32 and Fibroblast-UTSW, both obtained from Jerry Shay/UTSW). The distinguishing gene expression signature for these strains/lines was comprised of the high expression of a number of genes with roles in remodeling of extracellular matrix including high expression of the genes encoding smooth muscle actin, vimentin, fibrillin, byglycan and collagen types I, III, V and VI (Fig. 1D), combined with the low expression of genes characteristic of epithelial cells (Fig. 1B). The cell lines contained within this branch of the dendrogram (Orange) were further subdivided into a branch that contained three similar lines that expressed the highest levels of this “stromal” gene expression signature and others that showed incrementally less expression of this set of genes. Consistent with the interpretation that this signature reflected expression of stromal cell physiology, this signature was the most strongly expressed in the carcinosarcoma derived line Hs578T and the two cell lines/strains established from fibroblasts, HMS32 (primary fibroblast strain with a finite lifespan) and Fibroblast-UTSW (telomerase immortalized breast derived fibroblast line). The remaining lines that clustered with these fibroblast-like lines, showed decreased overall expression of this set of genes with incrementally less expression in BT-549, less in MCF-10A, and finally, only a few stromal-signature genes expressed in the two independently propagated lines of MDA-MB-231 (Fig. 1D).

6. Common epithelial cell signature

In addition to the gene expression signatures that distinguished the three major branches of the cell line dendrogram, both the basal-epithelial-like cell lines and the luminal-epithelial-like cell lines expressed a set of genes that were absent in those lines expressing the stromal gene expression signature (Fig. 1B). This set was comprised of many genes that play roles in cell-to-cell contacts that seal the lumen or extracellular space in epithelial tissues (e.g. E-cadherin, plakoglobin and junctional-adhesion protein). This cluster likely distinguished genes involved in functions common between subtypes of epithelial cells, and therefore, comprised a molecular signature of epithelial cells (Fig. 1B).

7. Cell lines of ambiguous origins

Of particular interest were the breast derived cell lines that lacked expression of the common epithelial genes and expressed some characteristics of the “stromal” expression signature including BT-549, MCF-10A and MDA-MB-231. BT-549 showed strong expression of most of the genes in the stromal cluster (Fig. 1D) and lacked expression of the other signature expression patterns including the common epithelial cell pattern. This suggests that BT-549 may represents a myofibroblast-like line transformed in vitro or a line like Hs578T that was originally derived from a stromal-like tumor in vivo [18]. The MCF-10A [19] line is a well-studied breast model system that expressed characteristics of both the basal epithelial signature including keratin expression, as well as genes that comprised the stromal signature, but significantly, lacked expression of the common epithelial cell pattern. Perhaps the most enigmatic cell line was MDA-MB-231 that did not
show strong characteristics of any of the three signature patterns of expression (Fig. 1C–E) except for a fraction of genes that comprised the stromal cluster. This cell line has been previously shown to be similar to renal carcinoma derived cell lines in a separate study that compared cell lines derived from a diverse set of tumor types, and therefore, may represent a de-differentiated cell type that has lost expression of the signature of its tissue of origin [6]. Further gene expression studies on these cell lines, including studies of their responses to extracellular matrix stimuli, might distinguish their potential for differentiation into cells with a more clear relationship to their in vivo counterparts.

In a previous study, we shown that another cell line that has been used as a model of aggressive-metastatic-breast tumors, MDA-MB-435, showed a pattern of gene expression that was very similar to the pattern seen in seven independent melanoma derived cell lines (see http://genome-www.stanford.edu/nci60/images/figure-2c.html and [6]). This distinctive pattern included strong expression of many genes characteristic of melanocytes including tyrosinase, dopachrome tautomerase and S100-β and therefore suggested that the tested cell line was derived from a Melanoma and not from a breast carcinoma. A number of different samples of MDA-MB-435 derived from different sources showed a similar pattern suggesting that most, if not all, examples of this cell line are similarly misclassified (data not shown). These finding suggest that this cell line is not an appropriate model system for the study of breast carcinoma.

8. Breast tumor gene expression patterns

One of the most useful aspects of microarray technology is its utility in the study of gene expression patterns in clinical tumor specimens [7,13,15,29–32]. We have previously published a study of gene expression profiles of forty breast cancer patients that included twenty samples from patient’s tumors before and after a sixteen week course of doxorubicin chemotherapy [7]. In order to identify the best set of genes to use for tumor classification, we utilized a statistical approach to identify the subset of genes that showed significant variation in expression across different patients/tumors, but which varied little in expression within paired samples from the same tumor [7]; this set of genes, termed the “intrinsic” gene set, was enriched for those genes whose expression patterns were characteristic of each tumor as opposed to those that varied as a function of sampling error. An example of a gene that showed this “intrinsic” property was ERBB2/HER2, which was expressed at high levels in some tumors and not others (forty-fold difference within this sample set), but which was consistent in expression in comparison between multiple samples taken from the same tumor.

Hierarchical clustering analysis using this “intrinsic” gene set of 476 cDNA clones (including 426 different genes) resulted in a novel molecular classification of the tumor samples on the basis of gene expression patterns (see Fig. 2 of [7] at http://genome-www.stanford.edu/molecularportraits/images/figure2.html). Importantly, the tumor samples cluster dendrogram from this analysis showed that the repeat biopsies taken from the same patient (i.e. the 20 “before” and “after” doxorubicin sample pairs) were found to almost always be more similar to each other than either was to any of the other tumors tested (17/20 “before” and “after” pairs were paired and 2/2 tumor/lymph node metastasis pairs were paired). This implied that every tumor is unique and has a distinctive gene expression “signature” or “portrait”. These gene expression patterns distinguished four discrete tumor subtypes that, similar to the cell line studies, could be related to features of normal breast cell type in vivo. The classes of tumor subtypes identified were 1) a “luminal epithelial/ER+” subtype that was distinguished by high expression of a set of approximately twenty genes that included the ER gene and other genes known to be regulated by estrogen, 2) a normal breast-like group of samples that contained the three normal breast samples, a single fibroadenoma and 5 tumor samples, 3) a group of tumors most of which expressed high levels of the ERBB2/HER2 gene, and 4) a group of tumors that had gene expression patterns reminiscent of breast basal epithelial cells.

Building upon these studies, we recently reported gene expression patterns of an expanded set of 78 different breast tumors [33]. Cluster analysis of data derived from this larger tumor set re-identified the same four tumor subtypes in addition to one additional subclass of ER+ tumors, such that five subtypes were now distinguished. To explore whether this classification of breast tumors was clinically significant, we assigned each of the 51 tumors that comprised the cohort of “before” and “after” doxorubicin patients [34], to a class based upon its location within the tumor sample associated dendrogram and did Kaplan-Meier survival analysis (see http://genome-www.stanford.edu/mopo/clinical/ and [33]). We found that these subtypes, as defined by gene expression pat-
terns, had statistically significant different overall pa-

tient survival and relapse free survival characteristics.

The luminal/ER+ positive patients were sub-divided

to two classes of which a novel subtype was identi-
fied (Luminal B/C) that had a significantly worse out-
come when compared to the rest of the ER+ tumors
(Luminal A), which showed the most favorable out-
comes. Furthermore, the set of patients classified as
“basal-like” had outcomes as poor as those that over-
expressed ERBB2/HER2. The patterns of gene expres-
sion that were distinguished in these studies represent
novel molecular signatures of breast tumors that can
be used to 1) develop new clinical tests based upon
gene expression patterns to score for these subtypes,
2) identify candidate markers for diagnosis, 3) iden-
tify genes important for understanding the biology that
distinguishes basal and luminal epithelial cells, and 4) iden-
tify subtype specific targets for developing thera-
peutic interventions.

9. An integrated cell line and breast tumor analysis

The set of genes that defined epithelial cell character-
istics within the cell line panel was remarkably similar
to the set of genes that distinguished tumor subtypes
amongst the breast carcinomas. This suggested that
certain cell lines may be very good models for specific
subtypes of tumors. In order to more directly compare
contrast primary breast tumors and cell lines, we
created a single hierarchical clustering diagram using
the aforementioned “intrinsic” gene list and data from
16 cell lines discussed above and our previously pub-
lished study on 40 breast tumors and three normal breast
samples (Fig. 2, and see Supplementary Information
Fig. 4 for the complete cluster diagram – http://genome-
www.stanford.edu/breast

As expected, the results showed a similar grouping of
the tumors samples into at least four subtypes including
luminal/ER+ (dark blue), ERBB2/HER2+ (pink), normal
breast-like (green), and a basal-like classes (dark red).
The cell lines, regardless of their presumed cell-
type of origin, clustered together on a single large
dendrogram branch separate from all of the tumors,
however, they were also similarly subdivided into the
basal, luminal, and stromal-like groups described above
(Fig. 2).

The luminal/ER+ signature was most strongly ex-
presened in a large group of ER expressing tumors
and the subset of cell lines that expressed the lumi-
nal gene expression signature (blue dendrogram branch
and Fig. 2B). The pattern of expression of this set of
genes showed both quantitative and qualitative differ-
ences between the luminal/ER+ tumors in compari-
on to the luminal cell lines. These tumor samples
were comprised of at most 60–70% tumor cells, but
expressed stronger and more consistent levels of this
gene set when compared to the “pure” population of
cell derived from a single cell line. Given the diffi-
culty in establishing cultures from cells expressing lu-
minal characteristics, the loss of expression of these
genes may be related to the process of establishment
or maintenance of cell lines in vitro. It was interesting
that the ERBB2+ tumor subtype expressed fewer of
these genes than the luminal-like/ER+ tumors, and in
most cases, expression levels lower than the luminal
cell lines. These patterns were consistent with the no-
tion that the relative level of expression of this set of
genes may reflected the degree of luminal differentia-
tion of the tumor samples. Taken together, these re-
results suggested that the best models for ER+ luminal
epithelial cell derived tumors, choosing from the cell
lines tested here, are MCF7, T47D and BT-474, with
SK-BR-3 serving as a model of luminal-cell derived
ER-negative/ERBB2 positive tumors.

We have previously shown that most of the genes
highly expressed in a pattern similar to ERBB2 across
large sets of breast tumors are all part of the co-
amplified chromosomal region that contains ERBB2 [7,
35–38]. In this analysis, most of the tumors that ex-
pressed this gene expression signature as their pri-
mary distinguishing characteristic were clustered to-
gether and formed a discreet subtype (pink dendo-
Gram branch, and Fig. 2D). A few other tumors that
expressed the ERBB2 signature were present, however,
they also expressed either the luminal or normal breast
signatures and were clustered based upon those distin-
guishing characteristics. Both BT-474 and SK-BR-3
showed high level expression of the ERBB2 signature
and therefore are likely appropriate cell line models for
ER-positive and ER-negative, ERBB2 over-expressing
tumors, respectively.

The basal-like tumors were distinguished by strong
expression of keratin 5 and 17 relative to the luminal
cell-like tumors (Fig. 2 and [7]). The basal epithelial
gene expression signature expressed by HMEC lines
was in part expressed by both the basal-like tumors and
normal breast tissue, which appeared to be comprised
predominantly of basal epithelial cells (dark red den-
drogram branch and Fig. 2E). These results suggested
that the basal like cell lines/HMEC cultures and their
derivatives, are in general, appropriate model systems
Figure B: Heatmap representation of gene expression differences across various cell lines and tumor subtypes.

A: Taxonomy of breast tumors and cell lines.

B: Heatmap showing gene expression levels across different samples, with red indicating above average expression and green indicating below average expression.

Legend:
- **Black**: average expression
- **Red**: above average
- **Green**: below average

Gene List:
- complement component 4A AA664406
- KIAA0182 protein AI023801
- fatty acid amide hydrolase AA431988
- major histocompatibility complex, class I, A W92232
- ESTs AA029948
- B/K protein H23265
- cellular retinoic acid-binding protein 2 AA598508
- cellular retinoic acid-binding protein 2 AA036986
- claudin 7 AA487488
- tumor protein D52 AA459100
- hypothetical protein FLJ20150 W38021
- vav 3 oncogene W91879
- mucin 1, transmembrane AA486365
- KIAA1223 protein R72441
- nebulette N77806
- ESTS similar to transcription factor forkhead-like 7 FKHL7 AA495790
- ras homolog gene family, member B H88963
- cytochrome b-5 R91950
- annexin A9 N54608
- estrogen receptor 1 AA291702
- trefoil factor 3 intestinal N74131
- GATA-binding protein 3 H72474
- X-box binding protein 1 W90128
- hepatocyte nuclear factor 3, alpha T74639
- putative G protein-coupled receptor H50224
- selenium binding protein 1 T65736
- Homo sapiens cDNA FLJ11408 fis, clone HEMBA100082 H53479
- Human insulin-like growth factor binding protein 5 IGFBP5 T52829
- Human insulin-like growth factor binding protein 5 IGFBP5 AA054451
- Human insulin-like growth factor binding protein 5 IGFBP5 H08560
- ESTs, Weakly similar to macrophage lectin 2 [H.sapiens] N53421
- Homo sapiens cDNA FLJ12900 fis, clone NT2RP2004321 N69835
- Homo sapiens cDNA FLJ12900 fis, clone NT2RP2004321 R98407
- DKFZP586O1624 protein AA159578
- hepsin transmembrane protease, serine 1 H62162
- EST similar to transcription elongation factor TFIIS R09980
- trefoil factor 1 breast cancer, estrogen-inducible in R83377
- ESTs W87826
- cDNA: FLJ21918 fis, clone HEP04006 T72068
- serine protease inhibitor, Kunitz type, 2 AA031287
- cadherin 1, type 1, E-cadherin epithelial W86859
- cadherin 1, type 1, E-cadherin epithelial H97778
- junction plakoglobin R06417
- creatine kinase, mitochondrial 1 ubiquitous H43515
- nuclear factor I/A AA022462
- creatine kinase, mitochondrial 1 ubiquitous AA019332
- metargidin AA292676
- myeloid/lymphoid or mixed-lineage leukemia trithorax homolog AA454610
- zinc finger protein 144 Mel-18 AA464420
- cDNA DKFZp564O2364 from clone DKFZp564O2364 W90240
- matrix metalloproteinase 15 membrane-inserted AA443300
- SWI/SNF related, matrix associated, subfamily e, member 1 W51779
- ESTs T57034
- TNF receptor-associated factor 4 AA598826
- TGFB1-induced anti-apoptotic factor 1 AA446222
- hypothetical protein FLJ10700 W81185
- flotillin 2 R72913
- steroidogenic acute regulatory protein related AA504615
- growth factor receptor-bound protein 7 H53702
for the breast basal-like tumors. The absence of expression of any genes characteristic of the luminal signature suggests that HMEC cultures and their derivatives are NOT appropriate models of hormone responsive breast tumors, which comprise the majority of sporadic breast tumors [1]. HB2 and HCC-1937, as noted above, expressed genes characteristic of both the basal and luminal signatures, and therefore may be similar to basal-like tumors that often co-express cytokeratin 17 and 18 in vivo (M. van de Rijn, personal communication).

The cell lines that expressed the stromal cell signature (orange dendrogram branch), with the exception of MCF-10A, showed very few gene expression characteristics in common with any of the breast tumors, even those that were highly metastatic when sampled for microarray analysis. The cellular origins of these cell lines are still enigmatic and it is not clear which, if any, of these cell lines are appropriate models for breast carcinoma. It is interesting to note, however, that some of the cell lines contained within this cluster, in particular MDA-MB-231, are some of the most tumorigenic and aggressive in nude mouse xenograft models [39]. Most interesting amongst these cell lines was MCF-10A, which expressed some genes characteristic of basal like tumors including keratin expression, but lacked expression of the common epithelial signature. Interestingly, this cell line is not tumorigenic in xenograft models.

10. Summary

The advent of the DNA microarray technology has enabled researchers to measure genomic scale gene expression in human cancers and cell lines. The exploration of these gene expression patterns is challenging oncologists and pathologists to re-assess traditional classifications of cancer and incorporate molecular features into treatment regimens and drug development strategies. The great strength of cDNA microarray studies coupled to hierarchical clustering analysis is the ability to objectively identify sets of coordinately expressed genes and display the data in a format that a biologist can utilize to form hypotheses.

The data presented here, and in our previous studies, have shown that many different gene selection criteria, across different sets of breast tumors and cell lines, consistently identified similar sets of genes that can serve as markers for probing the biology of breast cancer [6, 7, 33]. The comparison of gene expression patterns between cell lines and tumors is dominated by differences related mostly to the proliferative index of the samples, with most cell lines growing at a much faster rate than in vivo tumor cells [6–8,13,15]). However, in the case of breast tumors, cell lines and tumors share many aspects of their gene expression patterns that can be related to the normal and pathological physiology that distinguishes breast cell types in vivo. These gene sets include 1) the basal epithelial cluster, 2) the luminal epithelial/ER+ cluster, 3) the ERBB2+ amplicon cluster, 4) the proliferation cluster, and 5) the interferon cluster [7,8]. Remarkably, the classes of tumors as defined by gene expression, in part, are consistent with current markers that are used for breast cancer stratification and prognostication (e.g. ER status, ERBB2 status, proliferative index) [33].

In addition to re-identifying and elucidating traditional classes of tumors, gene expression patterns are revealing novel subtypes of tumors that appear both biologically and clinically distinct. In the study by Sorlie et al., the class of tumors distinguished by expression of the basal epithelial signature, including expression of cytokeratins 5 and 17, showed an outcome as poor as ERBB2 over-expressing tumors and were as numerous. Similarly, this study also identified a sub-group of patients with ER-positive tumors, traditionally classified as having a good prognosis, that had very poor outcomes [33]. The similarities and differences between cell lines and tumors should allow a much more informed choice to be made about the appropriateness of any given cell line model for a particular aspect of tumor biology to be studied in vitro.

In addition to the dominant patterns of gene expression described in this review, there is tremendous additional variation in gene expression patterns in comparison between tumor subtypes (see Supplementary Information Figures at http://genome-www.stanford.edu/breast_cancer/cell_line_review2001/).
Genome expression studies of fifty to one hundred tumor specimens likely does not have the power to identify all of the inherent biologic diversity of tumors. It remains to be determined whether one, a few, or tens to hundreds of markers will be necessary to identify distinguishing characteristics of tumors in the clinical setting. It is likely that larger gene expression profiling studies, and/or large in situ or immunohistochemistry studies using candidate markers identified in microarray studies, will be necessary to distinguish all of the clinically relevant variation in biology that can be exploited to develop better patient management algorithms and targeted drug strategies.

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