Determination of *Salmonella* in egg shell and egg content in some selected areas of Bangladesh

Akter S¹, Ferdowshi Z¹, Islam MN¹,², Prodhan MAM¹ and Chowdhury MYE¹*

¹Chittagong Veterinary and Animal Sciences University, Bangladesh ²Massey University, New Zealand

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**ABSTRACT**

Salmonellosis is one of the most common and widely distributed food-borne diseases. It constitutes a major public health burden and represents a significant cost in many countries. *Salmonella* are known for its wide range host and can cause clinical diseases in some hosts while in others, can be asymptomatic. Poultry and eggs are considered as major sources for different pathogenic *Salmonella* serotypes. Eggs produced locally under the small scale layer farm may present a hazard to consumers which may increase the spread of *Salmonella* in the environment. To investigate the occurrence of *Salmonella*, a total of 72 samples were taken from 6 poultry farm in some selected areas of Bangladesh. Sampling program was executed between October and December, 2013 and samples were tested using standard laboratory methods. Data was collected through direct interview and structured questionnaire. Our study shows that, true prevalence of *Salmonella* in egg shell and egg contents were 0.093% and 0.068% respectively. The highest apparent prevalence in Udayan poultry farm (50%) and lowest in Liza poultry farm (16.67%). The average frequency of egg shell contamination is positively correlated with re-use of egg tray without disinfection. To the best of our knowledge, this is the first investigation on *Salmonella spp.* in selected local egg production farms in Chittagong. Further detail study is highly recommended.

**Keywords:** Salmonella, small scale layer farm, table egg, food borne diseases, public health.

**INTRODUCTION**

Poultry products especially the eggs and egg products plays vital nutritive role in human health specifically for developing country like Bangladesh [¹][²]. Eggs are enriched with protein, minerals, fat and different vitamins like vitamin B₁₂ [³]. However, consuming inaccurately treated eggs and egg products can causes food borne diseases like salmonellosis [⁴]. *Salmonella* is a major food-borne pathogen distributed worldwide and contaminated poultry products [⁵]. In fact, Salmonellosis is considered as major public health burden in developing countries and important food borne pathogen in developed countries [⁶][⁷]. Broad range of *Salmonella* like *Salmonella choleraesuis*, *S. enterica*, *S bongori*, *S. typhi*, *S paratyphi* and *S. typhimurium* causes gastrointestinal (GIT) and typhoid fever. Among them, *S. enterica* subspecies *enteric* is responsible for more than 99% infection in man and animal [⁸]. Most of the infections of *Salmonella* are zoonotic in nature except *S. typhi* and *S. paratyphi*. The non typhoidial salmonellosis has been increased dramatically in past 10 years with predominant serotypes of *S. enteric* serotype *enteritidis* and *S. typhimurium* [⁹][¹⁰].

*Salmonella spp.* contaminating food and food products can causes Salmonellosis. Animal originated foods like poultry, poultry products and raw eggs are often contaminated by different *Salmonella spp.*. However, other sources of exposure include water, vegetables, fruits, pet and domestic animal handling and person-to-person when hand-mouth contact occurs without proper personal hygiene. Human cases of salmonellosis caused by *S. enteritidis* increased recently due to ingestion of poultry products specifically eggs [¹⁰]. Additionally, presence of *Salmonella spp.* in egg shell also possesses a considerable public health hazards and economic losses in poultry industry. Contamination of egg by *Salmonella spp.* may cause at any stage of production like collection, transportation or marketing through vertical or horizontal transmission. Importantly, reusable egg tray is a potential source for contaminating egg shell by salmonella in developing country like Bangladesh. Small scale layer farms are the major source of eggs in Bangladesh. The egg consumption is considerably increased in Bangladesh in past two decade due to promotion of egg as an ideal food by GO and NGO. So it could be acted as a potential vehicle of salmonella transmission in human but it is unlikely to develop surveillance program in Bangladesh like other develop country. Concern authorities are not aware of salmonella at farm level due to constrain of resource and facilities. The actual data on prevalence of salmonellosis in eggs and egg products is poorly documented that prone to zoonotic threat. In addition, small scale commercial farm is predominant in Bangladesh with minimum bio-security practices unlike other large scale commercial production system; enhance the chance of infection to the birds. The data on prevalence and rate of infection in eggs and egg products in Bangladesh is limited. Furthermore, in Bangladesh, there are no directives to control the process of egg production [¹¹] or limited study to evaluate the quality

* Corresponding author: dalim20003@gmail.com
of eggs in Bangladesh. This investigative work is proposed to address this issue and was focus on table egg produced in local farms to determine the apparent and true prevalence of egg contamination by *Salmonella spp.* with the associated factors.

**MATERIALS AND METHODS**

**Study design and data collection**

Small scale commercial layer farm (*n*=6) of Chittagong (*n*=3) and Noakhali (*n*=3) were selected purposively for the study. The Jafar Poultry Farm (farm 1), Hoque Poultry Farm (farm 2) and Kachwya Poultry Farm (farm 3) is located in Moddhomchorkakra, Charparbotipur and Bagtara village of Companygonj upazilla respectively. The Udayan Poultry Farm (farm 4), Islam Poultry Farm (farm 5) and Liza Poultry Farm (farm 6) is located in Satkaniya, potiya and chandanish of Chittagong districts. The landscape characteristics of this study were both high and lowland where farming is major way of livelihood. A cross sectional study was conducted in selected farms for the investigation of *Salmonella spp.* and associated factors for its prevalence in table eggs of small scale layer farms. The studied farm 1, 2, 3, have 4000, 2000 and 2500 birds of Noakhali and farm 4, 5, 6 have 1800, 7000 and 4000 birds of Chittagong. The studied farm 1, 2 and 6 having Isa brown strain while 3, 4 and 5 were rearing Hisex brown strain of layer bird. The study was conducted between October and December 2013. A structured record keeping sheet was developed, validated and used to collect the necessary information. The questionnaire contained closed, semi closed and open ended question. The questionnaire was grouped on: 1) basic information related to farm identity, farm composition and bird demography, 2) farm management system related to biosecurity level of farm and 3) egg collection, preservation and marketing procedure. All information was collected by face to face interview to the farm owner, manager or attendance as well as by physical examination.

**Source population and sample collection:**

Only egg laying flocks of each farm were used to develop the sampling frame where smallest unit consist of 2000 birds. Initially, 12 fresh eggs were collected from the selected farms and transported to microbiology laboratory at Chittagong Veterinary and Animal Sciences University (CVASU). Obtained samples were transferred carefully with a layer of sterile cotton for avoiding the breakage of egg. Each egg was given a unique identification number according to the farm identity and strain of the farm. The samples were preserved in 4°C until processing.

**Table 1: Result of sample culture on XLD, SS agar and TSI for Salmonella isolation and identification.**

| Farms   | Egg samples | Positive samples in different media | Microscopic features |
|---------|-------------|-------------------------------------|---------------------|
|         |             | XLD | SS | TSI | XLD | SS | TSI |
| F1      | 12          | 8   | 6  | 6   | 4   | 6  | 4   | Gram-negative, pink colored, small rod |
| F2      | 12          | 0   | 4  | 2   | 2   | 4  | 2   |
| F3      | 12          | 4   | 6  | 4   | 6   | 6  | 6   |
| F4      | 12          | 8   | 6  | 6   | 6   | 2  | 2   |
| F5      | 12          | 6   | 8  | 6   | 4   | 6  | 4   |
| F6      | 12          | 4   | 4  | 4   | 4   | 2  | 4   |
| Total   | 72          | 30  | 34 | 28  | 26  | 26 | 22  |

F1= Jafar poultry, F2= Hoque poultry, F3= Kachwya poultry, F4= Udayan poultry, F5 = Islam poultry and F6 = Liza poultry

On SS agar salmonella colonies were blackish and in XLD agar, the colonies appeared as black centered because of H2S production (Figure 1).

![Figure 1. Black centered colony in XLD agar suspected to *Salmonella spp.*](image)

*Salmonella spp.* and associated factors for its prevalence in table eggs of small scale layer farms. The studied farm 1, 2, 3, have 4000, 2000 and 2500 birds of Noakhali and farm 4, 5, 6 have 1800, 7000 and 4000 birds of Chittagong. The studied farm 1, 2
Nutrient agar (Oxoid Ltd., $P^H$: 6.2±0.0) was used as primary enrichment media. The Xylose Lysine Deoxycholate (XLD) agar (Oxoid Ltd., $P^H$: 7.4±0.2), Salmonella-Shigella (SS) agar (Merck, $P^H$: 6.9±0.2) and TSI agar (Oxoid Ltd., $P^H$: 7.2±0.2) were also used as selective media for the isolation of Salmonella spp. All the samples were subjected to the laboratory evaluation. The swabbing techniques were used to detect the Salmonella spp. in egg shell. The inner masses were inoculated in media for the detection of salmonella in egg inner mass as described previously \cite{12}. For the preparation of inoculums of egg shell surface, a sterile cotton swab wetted in sterilized normal saline solution (NSS) was used for surface swabbing and it was re-immersed into the same tube having 10 ml NSS. The surfaces of each of the egg were primarily disinfected with 70% ethanol and then the eggs were broken to collect the inner content swab. Finally, the content thoroughly mixed for approximately 1 minute and centrifuged for preparation of inoculum of inner content.

**Isolation and identification of Salmonella spp.**

1ml of prepared inoculum from egg shell and egg inner mass was inoculated in screw cap test tube containing nutrient broth and incubated for 24 hours (h) at 37°C. After incubation a loopful of culture was streaked on both XLD and SS agar and incubated at 37°C for 24 h. The colonies with black center in XLD and blackish growth in SS were considered as presumptive Salmonella spp. Suspected colonies were taken over a slide and allowed it to air dry followed by fixation in light flame. Gram’s staining was performed as per procedures described by Merchant and Packer (1969) \cite{13} and observed under microscope at 100x magnification for bacterial characterization. For further confirmation, the colonies were subjected to the biochemical analysis by TSI slant by stabbing the butt down to the bottom, and then streaked over the surface of the slant. The TSI slant was incubated overnight at temperature of 37°C. The positive results for salmonella were detected based on the properties.

**Data analysis**

Data obtained was imported to the Microsoft Office Excel-2007 and transferred to the software STATA/IC-11 for analysis. Descriptive statistics and associated factors were correlated with high frequency of egg contamination within farm level through descriptive statistics.

**RESULTS**

**Confirmation of Salmonella spp. by cultural and biochemical characteristics**

Among farms, the highest egg shell contamination by salmonella recorded in farm 1 and 4 (n=8) and none from farm 2 in XLD. Similarly, in the case of inner content of egg, farm 3 and 4 (n=3) showed highest number of positive and lowest from farm 2. In case of SS agar highest egg shell contamination was recorded in farm 5 (n=8) and lowest (n=2) in farm 2 and 6. In case of inner content, farm 1, 3 and 5 showing higher (n=6) than others. The positive samples from XLD and SS agar were further analyzed in TSI slant for final confirmation. Among all positive cases of 6 studied farm, a total number of 28 eggs shell and 22 (n=72) egg inner content found positive to the Salmonella spp. in TSI slant media (Table 1).

**Table 2. True and apparent prevalence of Salmonella spp in egg samples**

| Farm | Total population | Average egg production n/ day | Sample collected | AP (ES)% | TP (EC)% | TP (EC)% | Average TP (EC)% | Averag e TP(ES)% |
|------|-----------------|-------------------------------|-----------------|----------|----------|----------|-----------------|----------------|
| F1   | 3811            | 3650                          | 12              | 50%      | 33.33%   | 0.079%   | 0.052%          |                |
| F2   | 4700            | 4500                          | 12              | 16.66%   | 16.66%   | 0.02%    | 0.021%          |                |
| F3   | 2382            | 2000                          | 12              | 33.33%   | 50%      | 0.08%    | 0.126%          | 0.093%         |
| F4   | 1846            | 1750                          | 12              | 50%      | 16.66%   | 0.16%    | 0.054%          | 0.068%         |
| F5   | 1603            | 1500                          | 12              | 50%      | 33.33%   | 0.19%    | 0.125%          |                |
| F6   | 6635            | 5800                          | 12              | 33.33%   | 33.33%   | 0.03%    | 0.03%           |                |

AP=Apparent prevalence, TP= True prevalence, ES=Egg shell, EC=Egg content, F1= Jafar poultry, F2= Hoque poultry, F3= Kachuya poultry, F4= Udayan poultry, F5 = Islam poultry and F6 = Liza poultry

**Table 3 Salmonella positive% within different strain**

| Strain   | Salmonella positive (Egg shell) | p-value | Salmonella positive (Egg content) | p-value |
|----------|---------------------------------|---------|----------------------------------|---------|
| Isa brown| 66.67%                          | 0.273*  | 66.67%                           | 0.406*  |
| Hisex brown| 100%                            |         | 33.33%                           |         |

*NS= non-significant
Determination of apparent and true prevalence of Salmonella spp

Individual and overall salmonella positive percentages in egg of studied farm were also determined on the basis of biochemical test. The overall apparent prevalence of salmonella in table egg collected from study area was 38.8% in egg shell and 30.5% in egg inner content while in individual farm level, highest prevalence was 50% in Jafar, Islam and Udayan Poultry Farm. On the contrary, lowest prevalence was observed in Hoque Poultry of 16.66%. Others, 33.3% prevalence was observed in both Liza and Kachuya Poultry Farm. In the case of strain variation, among the positive samples we found that, Hisex brown showed 100% in egg shell and 33.33% in egg content were salmonella positive. On the other hand, Isa brown is showing similar percentages of salmonella contamination in both egg shell and inner content of the eggs (66.67%). There were no statistically significance among the strain variation (Table 3).

Factor associated with the presence of Salmonella spp. in egg shells and egg contents

The table 4 showing that, the overall management system of Liza Poultry farm was better than the other as they follow the strict bio-security measurement. However, the management system of other farm is more or less similar except Islam Poultry which was better than the rest of farms except Liza farm. For collection and transportation of egg, all poultry farms were using the tray without disinfection except Liza and Islam Poultry Farm, who were regularly using disinfectant before using the tray. The vehicle of egg transportation was usually used without any disinfection except Liza Poultry Farm.

DISCUSSION

Egg and egg products is vital human dietary content due to lower cost. But improperly handled egg could be source of public health hazard. Foods of animal origin, especially poultry and poultry products, including eggs, have been consistently implicated in sporadic cases and outbreaks of human salmonellosis. Several studies referring the salmonella contamination in both inner mass and outer shell of eggs responsible for human infection. The aim of this study was to determine the presence of Salmonella spp. in eggs in selected local farms where the true and apparent prevalence of salmonella in table egg from both egg shell and inner contents were determined. The true prevalence of egg shell contamination by salmonella was 0.093% and egg inner content was 0.068%. The incidence levels of S. enteritidis in egg shell reported earlier were variable. In Spain, around 0.8 to 1% salmonella contaminated eggs were reported while in United Kingdom, prevalence was from zero to 2%.

Table: 4 Associated factor for infection of Salmonella spp. in farm level

| Farm management factor | F1 | F2 | F3 | F4 | F5 | F6 |
|------------------------|----|----|----|----|----|----|
| Pullets reared on floor| Y  | N  | Y  | Y  | N  | N  |
| Feed contains animal products| Y  | Y  | Y  | Y  | Y  | Y  |
| Water chlorinated | N   | Y  | N  | N  | Y  | Y  |
| Visitors allowed (no business) | Y  | N  | N  | Y  | N  | N  |
| Proper Manure handling | N   | Y  | N  | Y  | Y  | Y  |
| Cleaning and disinfecting between flocks | ST | Y  | Y  | N  | ST | Y  |
| Wash and fumigate | Y   | Y  | Y  | Y  | Y  | Y  |
| Reuse of egg trey | Y   | Y  | Y  | Y  | Y  | Y  |
| Washing and disinfecting the egg trey | N   | Y  | ST | N  | Y  | Y  |
| Vehicle disinfected | N   | Y  | N  | ST | N  | ST |
| Storage room disinfection | N   | Y  | N  | N  | N  | N  |

F1= Jafar poultry, F2= Hoque poultry, F3= Kachuya poultry, F4= Udayan poultry, F5 = Islam poultry and F6 = Liza poultry and Y=yes, N= No and ST= Some times

The prevalence of salmonella in egg shell and egg content from bulk egg processing plant is also reported 0.5-3.7% in United States of America and 3.7% in United Kingdom, prevalence was from zero to 1%. Other study in Belgium, New Zealand, Australia, and Canada reported a range of 2-13% salmonella infection in large sampling frame as part of public health surveillance system. A study on table egg in Dhaka city showed overall 14-17% inner egg and 18-31% egg shell contamination with Salmonella spp. Another study in Kuhlania city found 8% eggs were contaminated by Salmonella spp. In which 3% S. typhi and rest of S. enterica. The findings is very close to our apparent prevalence in farm level that is...
30-35%. Our study found higher contamination in egg shell than the inner content by *Salmonella* spp. Another study in northern part of India reported that lower frequency in egg shell (1-2%) but higher frequency in egg content (8%) contamination [17]. The collected samples were from retail and wholesale market of that study. Our study was supported by another study done in India, where they reported that, higher incidence on egg shell surface than the internal contents. In our study, true prevalence in individual farm level showed highest in farm 1 of both in egg shell (0.079%) and egg inner content (0.052%) and lowest (0.03%) in farm 6 ranges from 0.23-1.5%. The result is little lower than one study [23] while similar to another [38]. In this study, the apparent prevalence showed highest in farm 1 and farm 4 (50%). The apparent prevalence is higher than the study conducted in Pakistan (35%) [38], India [17] (28%). This variation may be caused due to small sampling strategy of our study or sensitivity of diagnosing tools used for the detection of salmonella [33]. Alternatively, it could be higher for the farm level prevalence of salmonellosis in birds. Eggs from known infected flocks of commercial layer farm could be expected to more prevalence of salmonella contamination. The overall prevalence of salmonellosis in commercial layer farm in Bangladesh is ranged from 8-24% [38]. So the current study findings of 33% are more or less consistent to the farm level infection. The previously reported Salmonellosis in a farm or known infected farm’s egg showed higher prevalence in *Salmonella* contamination [23]. Three of our studied farm having salmonellosis during the study period which causes highest level of contamination in inner egg mass. Various study on epidemiological risk factor in egg contamination suggested that regular vaccination could decrease the level of contamination in farm by *Salmonella* spp. [33, 34]. Our studied farm 2 and farm 6 were practicing regular vaccination against salmonella. It could be the reason of lower prevalence than other in egg content. Small scale layer farm with low biosecurity measurement in Bangladesh may causing higher incidence of salmonellosa in farm level resulting higher rate of egg contamination [32, 35]. In this study, low bio secured farm showed higher frequency of egg contamination. The Jafar poultry, where the visitors are allowed and improper handling of manure having highest rate of egg contamination than others. The farmers in the study area used to buy fish meal or other feed ingredients from the local markets where birds and eggs of different farms are also selling. The same vehicles are using for transportation of birds, eggs and feeds between the farms and the markets, and in most cases, these vehicles remain contaminated with faces, and non-disinfected. Different degrees of fecal contaminations of vehicles and frequencies of market visits may have role in higher frequency of salmonella infection in eggs [32]. In our study, higher frequency of egg shell contamination shows in farm 1 and farm 4 where same vehicle are using for transportation of feed as well as egg marketing. The farms are not concern about disinfecting there vehicle during transportation of egg in the market. Reuse of egg tray for the collection and transportation of egg in developing countries like Bangladesh have influence in contamination of egg [30]. But regular disinfection of egg collecting tray can reduce the risk of contamination [36].

CONCLUSION

The present study focused on the identification of *Salmonella* spp. in 72 samples from different farm level of Chittagong and Noakhali districts of Bangladesh. The overall prevalence of egg inner content contamination is 38.8%. The egg is considered as ideal food and major source of protein in Bangladesh. But the contamination of *Salmonella* spp. can causes major public health burden by consuming raw or under cooked eggs and egg products. In addition, the contamination of egg by salmonella involves major health expenses in developing country. So it is necessary to monitor the infection level in marketing channel of egg and egg products.

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