In Situ Molecular-Level Observation of Methanol Catalysis at the Water–Graphite Interface

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Supporting Information

ABSTRACT: Methanol occupies a central role in chemical synthesis and is considered an ideal candidate for cleaner fuel storage and transportation. It can be catalyzed from water and volatile organic compounds, such as carbon dioxide, thereby offering an attractive solution for reducing carbon emissions. However, molecular-level experimental observations of the catalytic process are scarce, and most existing catalysts tend to rely on empirically optimized, expensive, and complex nanocomposite materials. This lack of molecular-level insights has precluded the development of simpler, more cost-effective alternatives. Here, we show that graphite immersed in ultrapure water is able to spontaneously catalyze methanol from volatile organic compounds in ambient conditions. Using single-molecule resolution atomic force microscopy (AFM) in liquid, we directly observe the formation and evolution of methanol–water nanostructures at the surface of graphite. These molecularly ordered structures nucleate near catalytically active surface features, such as atomic step edges, and grow progressively as further methanol is being formed. Complementary nuclear magnetic resonance analysis of the liquid confirms the formation of methanol and quantifies its concentration. We also show that electric fields significantly enhance the catalysis rate, even when as small as that induced by the natural surface potential of the silicon AFM tip. These findings could have a significant impact on the development of organic catalysts and on the function of nanoscale carbon devices.

KEYWORDS: methanol catalysis, water, highly oriented pyrolytic graphite, atomic force microscopy, nuclear magnetic resonance, electric fields

INTRODUCTION

The conversion of unwanted volatile organics, such as carbon dioxide, to methanol is of high interest given the pressing need for alternative energy sources to fossil fuel and the significant potential to reduce carbon emissions.3 Methanol also functions as an important platform molecule for chemical synthesis and offers an ideal solution for cleaner energy storage and transportation.3 At the present time, methanol is catalyzed on an industrial scale, but usually at high temperatures and pressures and relying on catalysts made of complex composite materials typically comprising active metal nanoparticles in an oxide support.5–7 Given the complexity of these composites, their catalytic behavior is still not fully understood although the synergy between the constituent components has been shown to be one of the key elements.8 Significantly, composites tend to require a specific nanoscale arrangement, making them expensive and highly sensitive to even slight structural changes. There is hence a strong need for simpler and cheaper alternatives that can be easily sourced and replaced. Organic materials, such as graphite derivatives, present obvious candidates as alternative catalysts.9 The use of graphitic materials in science and technology has grown dramatically over the last decade owing to graphene’s extraordinary electronic and physical properties. Current applications range from electrochemical devices and fuel cells10 to energy storage,11 photovoltaics,12 and the development of materials exhibiting unique mechanical properties.13 Pure graphene is hydrophobic, and the ambient humidity can influence the behavior of devices14,15 due to the formation of a nanoscopic water layer on all exposed surfaces. The hydrophilic derivative of graphene, graphene oxide (GrO), is obtained by the replacement of single carbon atoms in graphene sheets with oxygen containing functional groups, such as epoxy and methoxy groups. GrO retains some of the unique electronic properties of graphene, but the presence of hydrophilic groups makes it soluble in water, opening new avenues for applications in water filtration and ion sieving16 along with molecular sensing.17 Recent results suggest that GrO can act as a photocatalyst for the conversion of water and carbon dioxide to methanol.18 This behavior is attributed to the presence of the hydrophilic functional groups that stretch...
and then 4 min elapses between the subsequent frames.

Once stable, the molecular details of the growing reaction, the nucleating structures are seen predominantly at the water at the surface of HOPG. Consistent to a catalytic process with single-molecule resolution atomic force microscopy in liquid. This includes the resulting self-assembly of water molecules at the HOPG interface. The white scale bars are 100 nm in (a) and 1 nm in (b). The purple color scale bar represents a topographic variation of 20 Å in (a) and 1 Å nm in (b). The blue scale bar represents a phase variation of 20° in (a) and 10° in (b). In (a) the time lapse between the first and second frames is 1 min and then 4 min elapses between the subsequent frames.

Figure 1. High-resolution amplitude modulation AFM imaging of HOPG immersed in initially ultrapure water. (a) A solid-like patch formed by the self-assembly of molecules (dashed white outline) nucleates from an atomic step at the HOPG surface (dashed black line). The molecular self-assembly is observed here in situ as it progressively grows across the HOPG surface over a period of 9 min, with the patch edges moving away from the step. Rowlike structures with a periodicity of 4.30 ± 0.28 nm as visible within the patch. (b) Sub-nanometer imaging of other structures reveals detailed features (0.79 ± 0.08 nm periodicity, red arrows) perpendicular to the main rows (periodicity 2.45 ± 0.08 nm, white arrow). The exact molecular arrangement is not known, but strongly reminiscent of the alternated water–methanol nanoribbons recently reported by our group. The white scale bars are 100 nm in (a) and 1 nm in (b). The purple color scale bar represents a topographic variation of 20 Å in (a) and 1 Å nm in (b). The blue scale bar represents a phase variation of 20° in (a) and 10° in (b). In (a) the time lapse between the first and second frames is 1 min and then 4 min elapses between the subsequent frames.

The surface of bulk graphite presents singularities at exposed atomic steps and edges. These singularities have long been known to make the edges of graphite electrochemically active. Recent studies have also demonstrated that graphite’s basal plane, previously considered electrochemically inert, has an activity comparable to that of noble metal electrodes, such as platinum. These findings suggest that graphite may offer a suitable alternative to metal electrodes given the fact that it can be readily immersed in aqueous solutions, unlike graphene.

The bandgap of GrO, hence allowing the photogenerated electrons and holes to serve as oxidation and reduction sites. Such catalytic effects have never been observed for pure graphene where the regular bandgap and the absence of chemical singularities do not favor localized electrons.

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■ RESULTS AND DISCUSSION

Here, we demonstrate catalytic production of methanol at the surface of immersed highly orientated pyrolytic graphite (HOPG) in ambient conditions. The process occurs spontaneously with the thermal energy available, but is stimulated in the presence of an applied electric field. We quantify the amount of methanol produced using 1H NMR spectroscopy of the resulting solution and follow in situ the process with single-molecule resolution atomic force microscopy (AFM) in liquid. This includes the resulting self-assembly of water and alcohol molecules at the HOPG–water interface. Representative examples of this self-assembly can be seen in Figure 1a, where a molecularly structured patch is slowly growing in a system that initially consisted only of ultrapure water at the surface of HOPG. Consistent to a catalytic reaction, the nucleating structures are seen predominantly at the more electrochemically active edges of HOPG and are first observed after scanning for some time, typically more than an hour. Once stable, the molecular details of the growing domains can be imaged with AFM, often revealing assemblies involving units comparable to the size of water and short alcohol molecules (Figure 1b). Pure liquid water itself cannot form long-lived structures on HOPG at room temperature, indicating that the observed patch must contain molecules formed in situ.

A recent study by our group has shown that water and methanol can self-assemble into stable, layered structures at the interface with HOPG. Combining high-resolution AFM with computational simulations, these structures were shown to be stabilized by a group effect through an extended network of hydrogen bonds involving both water and alcohol molecules. The relatively weak interaction between the solvent and the graphite surface enabled a high degree of polymorphism in the molecular arrangement of the self-assembled structures. The formation of rowlike features with periodicities up to 6 nm was the most commonly observed supramolecular assembly. The formation of the patterns did not appear to be influenced by the scanning AFM tip, but was instead driven by the underlying graphite with the rows orientation in registry with the graphite lattice. Here, the molecular assemblies developing at the interface with HOPG in ultrapure water (Figure 1) are consistent to those reported for methanol–water mixtures suggesting that HOPG-induced catalysis of water into methanol is occurring.

To independently confirm the formation of methanol, we conducted NMR analysis on the solution in direct contact with the HOPG before and after nucleation was observed. Practically, the measurement is challenging because it requires observing small quantities of methanol (typically submillimolar) in the presence of a signal (water) that is 5 orders of magnitude larger. Therefore, it was necessary to suppress the water signal very efficiently to allow for unambiguous identification of the methanol produced. This was achieved with the recently reported Robust-5 pulse sequence (see
Experimental Section for details). The results, presented in Figure 2, compare three sets of measurements: (i) ultrapure water placed for 5 s in contact with the surface of HOPG, (ii) ultrapure water placed for 2 h in contact with the surface of HOPG, and (iii) ultrapure water placed for 2 h in contact with the surface of HOPG while applying a DC potential of +1 V to the HOPG. The peak at 3.21 ± 0.01 ppm is associated to the presence of methanol, and the tracer peak (DMSO, peak just below 3 ppm) was used to adjust the relative magnitude of the curves. The determined concentrations of methanol are 0.116 ± 0.010 μM (i), 0.000 ± 0.010 μM (ii), and 0.295 ± 0.010 μM (iii). For comparison with samples (i–iii), spectra were also collected in a 1 μM solution of methanol (b) and in ultrapure water exposed to air for 2 h (c). All samples were prepared in identical conditions (see methods for experimental details).

Figure 2. 1H NMR analysis for the methanol (MeOH) content of the ultrapure water solution after catalysis. (a) Spectra quantitatively comparing: (i) ultrapure water placed for 5 s in contact with the surface of HOPG, (ii) ultrapure water placed for 2 h in contact with the surface of HOPG, and (iii) ultrapure water placed for 2 h in contact with the surface of HOPG while applying a DC potential of +1 V to the HOPG. The peak at 3.21 ± 0.01 ppm is associated to the presence of methanol, and the tracer peak (DMSO, peak just below 3 ppm) was used to adjust the relative magnitude of the curves. The determined concentrations of methanol are 0.116 ± 0.010 μM (i), 0.000 ± 0.010 μM (ii), and 0.295 ± 0.010 μM (iii). For comparison with samples (i–iii), spectra were also collected in a 1 μM solution of methanol (b) and in ultrapure water exposed to air for 2 h (c). All samples were prepared in identical conditions (see methods for experimental details).

water placed for 5 s in contact with the surface of HOPG, (ii) ultrapure water placed for 2 h in contact with the surface of HOPG, and (iii) ultrapure water placed for 2 h in contact with the surface of HOPG while applying a direct current (DC) potential of +1 V to the HOPG with respect to a platinum electrode placed directly in the water. The value of the applied voltage was selected to avoid any chemical modification of the surface.24 All of the samples were collected in identical conditions, at room temperature, in contact with air and over HOPG previously heated above 120 °C to evaporate any historical contaminant (see Experimental Section).25 Methylsulfonylmethane (DMSO₂) was used as a tracer (2 μM) in each sample to allow for quantification of the methanol detected.

Sample (i) acts as an immediate control for possible contaminants since no supramolecular structures could be observed by AFM on such short time scales. We chose a 5 s water–HOPG contact time because this is the shortest time scale in which the sample could be prepared. No detectable level of methanol is expected and this is indeed confirmed (Figure 2a,i) demonstrating that the methanol present in the solution originates from catalytic activity and is not due to any form of external contamination.

Sample (ii) represents the shortest time scale typically necessary for the nucleation of supramolecular structures at the HOPG–water interfaces, as observed with AFM. A distinctive methanol peak is present (Figure 2a,ii), confirming catalysis of water to methanol at the surface of HOPG immersed in water in ambient conditions. Comparing the area of the methanol peak with that of the DMSO₂ indicates a methanol concentration of 0.116 ± 0.010 μM in the solution. This is an underestimate since it excludes any methanol that remained at the HOPG surface after the solution was removed for analysis. Indeed, a significantly higher alcohol concentration is expected to remain at the interface with the hydrophobic HOPG surface where alcohol preferentially resides.26,27 The present result shows that even at low concentrations, methanol can have a profound effect on the behavior of the interfacial liquid, as visible in Figure 1.

Sample (iii) serves two purposes. First, the observed enhancement of the catalytic activity under an electrical potential further confirms the expected electrocatalytic activity of the HOPG both at atomic steps and edges where the existence of additional functional groups19 may serve as oxidizing and reduction sites, similar to hydrophilic groups in GrO, and in the basal plane where fast electron transfer under applied fields is expected.20 When a potential of +1.00 ± 0.01 V was applied for 2 h, 0.295 ± 0.010 μM of methanol was detected (Figure 2a,iii), more than twice the amount formed without the electric potential. Second, results from this sample suggest that the scanning AFM tip is likely to have an influence on the catalytic activity of the HOPG surface. Since catalysis can occur spontaneously at room temperature and is enhanced by the presence of an electric field, the silicon oxide AFM tip may also enhance the process. Silicon oxide tips, such as those used in the present study, develop a negative surface potential of typically −60 mV28,29 when immersed in ultrapure water (pH of 5.8 in our experimental conditions), and the highly curved apex (<10 nm) can significantly enhance the resulting local electric field. Consistently, when water is placed in contact with the HOPG surface for several hours prior to imaging, ordered structures do not appear immediately at the start of imaging, but instead seem to be stimulated by the presence of the scanning tip.

Overall, the NMR results confirm spontaneous methanol catalysis. The process is enhanced by the presence of an electric field, potentially even as small as that induced by an immersed AFM tip at the interface with HOPG. Composite nanomaterials involving oxides are widely used in methanol catalysis and could play a significant role here. We therefore decided to further investigate the effect of small voltages (comparable in magnitude to the surface potential of typical oxides) on the formation of interfacial structures. Since most of the alcohol produced resides at the interface with the HOPG, even small electric potentials could potentially have a significant impact. Here, we compared the evolution of samples exposed to 50 ± 1 mV or in open circuit for 24 h. In both cases, it was necessary to dewet the HOPG surface to transfer the preconditioned sample to the AFM chamber, thereby leaving a thin interfacial liquid layer containing the produced methanol. More ultrapure water is then added, and the imaging starts within minutes. This procedure makes it difficult to rule out any disassembly/reassembly of the
interfacial structures during the transfer, but the differences between the two samples are obvious nonetheless (Figure 3).

Figure 3a shows that no features other than the characteristic graphite steps are initially visible in the absence of electrical preconditioning. After 1 h of continuous imaging, small raised patches about 100 nm in diameter begin to nucleate near step edges (Figure 3b). The patches exhibit supramolecular row patterns, comparable to those shown in Figure 1a. The assembly of these rows proceeds at a slow rate, here $0.14 \pm 0.10$ nm/s for the patch in Figure 3b, suggesting that growth is limited by the rate of methanol catalysis. Indeed, previous work with water—methanol mixtures upward of 5% volume concentration reported a growth of structures more than an order of magnitude faster than visible here. Additionally, structures never fully cover the surface over the time scale of the experiment (several hours) with the largest patch observed exhibiting a diameter of $\sim 1.2 \mu m$.

In contrast, when a 50 mV external electric field is applied for 24 h prior to imaging, unstructured patches form a monolayer on the HOPG surface and can immediately be observed (Figure 3c). The patches rapidly develop into ordered structures resembling those of the methanol—water monolayers. Significantly, within an hour of imaging the structures have almost completely covered the surface of the graphite (Figure 3d), at times forming multiple layers (see Figure S1 in the Supporting Information). Although there are still nucleation sites near surface features, such as in Figure 3c, we also observe nucleation deep into the basal plane (see Figure S2 in the Supporting Information) indicating that the electrical preconditioning has caused electrocatalysis across all of the HOPG, instead of being limited to just surface singularities. These results all support the existence of a significantly larger quantity of methanol at the interface when compared to the sample prepared without any applied electric field. Furthermore, the hour long delay in nucleation between Figure 3a,b despite the 24 h of contact beforehand is consistent to the idea that while there is initially methanol present in the solution (as shown in the NMR data, Figure 2a,ii), the local concentration is initially too low for nucleation, but the presence of the charged tip with a surface potential comparable to the electric field applied in (c) and (d). The white scale bars are 500 nm in the main images and 10 nm in the insets. The color scale bar represents a height variation of 1.5 nm in the main images and 1.2 nm in the insets.
carbon in our experiment and is responsible for the slight acidity of ultrapure water in ambient conditions (pH 5.8). Catalytic conversion of carbon dioxide and water into methanol has been previously reported for GrO\textsuperscript{18} where it is stimulated by white light (photocatalysis). Here, we did not observe any appreciable catalytic increase under illumination (see Figure S3 in the Supporting Information). However, experiments run in a sealed atmosphere for 2 h (comparable in duration to sample (ii) in Figure 2) revealed no detectable concentration of methanol upon subsequent NMR analysis of the liquid (see Figure S4 in the Supporting Information), supporting the hypothesis of carbon dioxide as a reagent. We note that a degree of atmospheric contamination of the HOPG\textsuperscript{25,30} is expected despite all of the steps taken to minimize contamination. However, the consistent and reproducible trends observed indicate that possible contaminants do not dominate the results. Furthermore, the typical HOPG contaminants\textsuperscript{25,30} are molecules far larger than methanol and so would not interfere with the AFM observations of its catalytic production.

The exact molecular mechanism underlying the catalytic process cannot be deduced from the present results alone, partly because the exact chemical details of the graphite are not known; surface groups at the edge plane\textsuperscript{33} could significantly alter graphite’s catalytic behavior by modifying the local electron-accepting/donating abilities and inducing charge delocalization that would in turn impact chemisorption.\textsuperscript{9} Our results suggest that the methanol production is caused by multiple factors, all involving the HOPG surface, that are difficult to disentangle. The AFM results provide consistent evidence of the AFM tip influencing the catalysis, likely through a tip field effect\textsuperscript{34} electrochemical reaction when in proximity to the HOPG surface (Figure 3). However, catalysis also occurs in the absence of an applied electric potential (Figure 2), through a mechanism dominated by step edges. Oxygen containing functional groups unique to multilayered graphite could also be at play, inducing absorption via bonded and nonbonded interactions with the liquid molecules and serving as active sites.\textsuperscript{33} In any case, the catalytic activity benefits from a positive polarization of the HOPG under an external electrical potential. This could be due to both a further enhancement of the chemical reaction at the step edges or to induced electron transfer occurring elsewhere. Indeed, recent studies have shown that doped graphene is able to reduce carbon dioxide in ambient conditions, when submitted to an electric potential.\textsuperscript{33}

When considering practical applications, the catalytic process reported here is far from optimized. It occurs slowly and is limited by the thermal energy available. In ambient conditions, we estimate a catalytic production rate of 4.6 mg h\textsuperscript{−1} m\textsuperscript{−2}, far smaller than the best reported catalysis rates.\textsuperscript{34} However, since singularities in the potential landscape, such as atomic step edges or the proximity of an AFM tip, can significantly enhance the rate of catalysis, there is much scope for improvement. Additionally, the relatively low cost of graphite, its outstanding stability, and the fact that material of technically lower quality (more defects) is catalytically more efficient should enable device geometries that maximize the catalytically active area without significant challenges.

From an electrochemical perspective, our results suggest that a graphite electrode immersed into an aqueous solution progressively develops an interfacial "passivation" layer formed by a solid self-assembled layer of water and methanol molecules produced in situ. Although relatively easy to destroy, this layer reforms spontaneously and may significantly affect interfacial processes, such as charge exchange\textsuperscript{35} and molecular adsorption,\textsuperscript{36} as well as the catalysis of other molecules.\textsuperscript{33} Interestingly, this result also suggests that local probe investigations of graphitic materials in aqueous solution may be prone to tip-induced catalysis effects.

CONCLUSIONS

In conclusion, our combined AFM and NMR results consistently show that graphite is able to spontaneously catalyse methanol at room temperature. The amount of methanol produced is relatively modest, and catalysis appears to occur almost exclusively at surface singularities on the graphite, such as atomic steps and in the absence of any external input of energy. The underlying molecular mechanism remains unclear due to uncertainties over the chemical composition of the graphite, but applying an external electrical potential across the interface considerably enhances the catalysis rate, even when due to the surface potential of nanoobjects located near the interface. We believe that our findings could have a significant impact on the development and understanding of novel carbon-based catalytic materials as well as devices highly sensitive to interfacial liquids.

EXPERIMENTAL SECTION

Sample Preparation. All of the solutions were prepared with ultrapure water (AnalaR NORMAPUR ISO 3696 Grade 3, VWR Chemicals, Leicestershire, U.K.) and high-performance liquid chromatography-grade methanol with a purity of ≥99% (Sigma-Aldrich, Dorset, U.K.). In a typical experiment, a liquid droplet (≈200 μL) of water was deposited on a freshly cleaved HOPG substrate (SPI supplies, West Chester, PA) mounted on a stainless steel disk using silver paint (Ted Pella Inc, Redding, CA). In all cases, the HOPG was baked to >120 °C for 15 min to remove any contaminants\textsuperscript{8} before depositing the droplet. For the experiments in Figures 2 and 3, the droplet was then left for a set period (5 s to 24 h) inside a partially sealed glass container at room temperature (20 ± 1 °C). The container was thoroughly cleaned with ultrapure water beforehand and protected from the light throughout the incubation. The same procedure was used for the electric field experiments, except for a platinum wire (Sigma-Aldrich) immersed in the droplet. The wire and HOPG sample were connected to a DC power supply (Ami-TTi, Cambridgeshire, U.K.) with a positive voltage applied to the HOPG with respect to the platinum.

NMR. After the determined incubation period, the droplet was pipetted from the HOPG into a clean NMR tube. The solution was then diluted with deuterium oxide (purity 99.9%, Cambridge Isotope Laboratories, Inc., MA) as needed and the DMSO\textsubscript{2} (Sigma-Aldrich, Dorset, U.K.) tracer added before conducting the measurement. The intense water signal was attenuated using the Robust-5 pulse sequence using a Varian (CA) 600 MHz spectrometer equipped with an Agilent (CA) probe able to deliver a maximum pulsed field gradient of 62 G/ cm. Scans (9248) were collected, each comprising 32 728 complex data points and a spectral width of 10 kHz. The repetition time was 3.6 s, of which 1.6 s comprised the acquisition time. The W5 interpulse delay was set to 287 μs. Rectangular 1 ms pulsed field gradients were used with a strength of G = 4.8 G/cm. The gradient stabilization delay was 1 ms. The error associated to estimating the quantity of methanol produced is dominated by the error in measuring the volume of liquid for NMR analysis, leading to an overall uncertainty on the concentration of ±0.01 μM. The contribution to this error from the NMR measurement itself is negligible.

AFM. Imaging was conducted in amplitude modulation mode using a commercial Cypher ES AFM (Asylum Research, Santa Barbara) equipped with temperature control and photothermal drive.
The cantilevers (Arrow UHF-AUD, Nanoworld, Neuchatel, Switzerland) exhibit a nominal spring constant of ~1.95 nN/nm (from thermal spectrum calibration) and a resonance frequency of ~430 kHz in liquid. The cantilevers were cleaned by immersion in ultrapure water before imaging. All parts of the AFM in direct and indirect contact with the solution (cantilever holder, imaging chamber) were thoroughly cleaned with ultrapure water prior to imaging. After washing, the stage was heated to 105 °C for 20 min to evaporate possible substances from previous experiments. To nucleate the structures, all of the samples were imaged at 40 °C, although all of the preconditioning occurred at room temperature.

**ASSOCIATED CONTENT**

Supporting Information
The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acsami.8b12113.

Additional AFM images showing multilayer growth (Figure S1); nucleation points far away from the step edges after the application of small electric fields (Figure S2); and control experiments testing the effect of light on the catalysis rate (Figure S3); additional NMR spectra showing that no catalysis occurs in a sealed environment (Figure S4) (PDF)

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Author Contributions
W.F., H.K., and K.V. designed all of the experiments. W.F. conducted all of the experiments except the NMR spectroscopy. W.F. prepared the samples for the NMR spectroscopy. J.A.A. designed and conducted the NMR measurements and assisted with their analysis. All authors were involved in the writing of the paper and subsequent commenting.

Notes
The authors declare no competing financial interest.

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