Introduction

Placental glucose transfer is essential to sustain fetal development (1). Insufficient glucose transfer may result in intrauterine growth restriction (IUGR) while too much glucose is associated with fetal macrosomia. Such conditions can further lead to complications at birth and an increased burden of chronic diseases in adulthood (1). The fetal glucose level varies with the gestational age and the
maternal glucose level. In addition, diabetes and gestational diabetes have been associated with elevated maternal glucose levels and risk of birth complications. Whereas the glucose tolerance test screens for gestational diabetes (2), it does not provide information about placental glucose transfer. Routine ultrasound exams are not are not able to characterize maternal-fetal glucose relationships. Although PET can map tracer distribution across the placenta and in fetal organs in experimental animals, it is not feasible in humans due to the ionizing radiation (3). Hence, it is urgent to develop a safe non-invasive means to assess placental glucose transport.

Advanced MRI techniques are being investigated for placental imaging. For example, placental perfusion can be measured by dynamic contrast-enhanced (DCE) MRI (4,5), arterial spin labeling (ASL) (6) and intravoxel-incoherent motion (IVIM) MRI (7-9). In addition, placental oxygenation can be assessed using T2 and T2* mapping (10,11) and BOLD MRI (12-14). In addition, MR spectroscopy (MRS) has be used to study the placenta (15-17) but its spatial resolution is limited. Recently, chemical exchange saturation transfer (CEST) MRI has been shown to be glucose sensitive via the exchange between bulk water and the glucose hydroxyl protons, dubbed GlucoCEST MRI (18,19). Dynamic glucose enhanced (DGE) MRI provides time-resolved glucose measurement in brain tumors at high field (20,21), which has been extended for head and neck cancer imaging at 3.0 Tesla (22). Recently, Wu et al. demonstrated DGE MRI at 11.7 Tesla in a mouse model of intrauterine inflammation (IUI), which is known to result in acute placental injury (23). It is necessary to note that most of these DGE experiments were performed with intravenous (IV) glucose administration.

Although it is advantageous to perform CEST MRI at high field (24), it is important to develop GlucoCEST MRI at 3.0 Tesla, particularly for the pregnant population. For patient acceptance, getting MRI scan after an oral glucose drink is far preferable to an IV load. Oral glucose tolerance test is commonly used during pregnancy to screen for gestational diabetes. To address these two issues, our study here optimized continuous wave (CW) GlucoCEST echo planar imaging (EPI) at 3.0 Tesla (25). We tested GlucoCEST in ex vivo placentas and characterized the relationship between GlucoCEST and placental vascular density. We also demonstrated GlucoCEST MRI following the glucose tolerance test and established the feasibility of placental DGE as an ancillary exam.

Methods

The study has been approved by the Institutional Review Board (IRB) at the Boston Children’s Hospital and Massachusetts General Hospital.

Glucose phantom study

D-glucose was dissolved in PBS at concentrations of 4, 8, 12 and 20 mM with pH titrated to 7.4. The solution was transferred into separate 50 mL Falcon tubes, and positioned in a cylindrical phantom holder.

Ex vivo placental tissue study

Fresh placentas were obtained from uncomplicated term deliveries within 6 h. Cannulation was performed on the surface fetal chorionic arteries of the placenta specimen. The lobules were flushed with normal saline and 5,000 IU/L heparin (NSH) until the venous return was clear. Subsequently, NSH supplemented with D-glucose (5, 10, 20 mM) and 1 mL/L yellow India ink (Cancer Diagnostics, Inc.) were perfused in separate placenta lobules of a single placenta. After the perfusion, the area around the perfused lobule was clamped, sutured, and fit into a cylindrical plastic container of 10 cm diameter. Four placenta sections were prepared and stacked in the container, which was then filled with agar gel (1%) (Figure 1). The sample was maintained at room temperature before imaging. Hematoxylin and Eosin histology was obtained after MR imaging and examined by an experienced placenta pathologist.

In vivo study

The GlucoCEST scans were performed in pregnant mothers (Table 1). After a glucose drink (TrutolTM) in a sitting position with a dose of 50 gm, equivalent to glucose tolerance test, subjects re-entered the scanner while keeping the same maternal position for the dynamic GlucoCEST scan. T2*-weighted scans were performed simultaneously.

Data acquisition

All studies were performed on a 3.0 Tesla Skyra (Siemens Healthcare, Erlangen, Germany). While phantom and ex vivo tissue was scanned using a 32-channel array head coil, in vivo subjects were scanned using a combined
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18-channel body and 12-channel spine receive arrays. CEST imaging was performed with single-shot gradient echo EPI (26) with a CW RF irradiation of 1 second. For the phantom study, we repeated CEST MRI measurements under B1 fields of 1, 1.5, 2 and 3 µT, and flip angle (FA) of 20, 45, 70 and 90 degrees. Z-spectra were obtained with RF saturation frequency from –5 to 5 ppm with intervals of 0.25 ppm. Imaging matrix 48 × 48; in-plane resolution 3 × 3 mm², slice thickness 10 mm; TR/TE =4 s/17 ms, FA=90°, BW =2.3 kHz/px, averages =2. WASSR scan was performed with the same FOV and resolution as CEST scan at B1 =0.5 µT.

For the ex vivo experiment, double spin echo MRI was performed for T2 imaging (TR =1 s, FA=90 degree, TE1/TE2 =8.5/50 ms). In addition, T1 was calculated based on images acquired with a double FA gradient echo sequence (TR/TE =15/1.7 ms, FA1/FA2 =5/26 degrees). Phantom and ex vivo experiments were performed at room temperature (25°C).

For in vivo study, all scans were performed free-breathing. The baseline (before glucose drink) CEST image was collected with a CW CEST MRI (RF saturation time =1 s, B1 =1.5 µT). Upon mother re-entering the scanner, the dynamic CEST scan was performed with saturation frequency sweep densely between 0.75 and 4 ppm with intervals of 0.25 ppm, and sparsely between –4 to 0 with intervals of 2.0 ppm. Each dynamic scan has a total of 17 frequency offsets plus two reference scans without on resonance saturation, which adds up to 1 min 16 s for each dynamic scan. At the end of the scan, a full CEST z-spectrum was acquired. A field map was collected with a double gradient echo MRI.

**Data processing**

B0 map was obtained by Lorentzian fitting of z-spectrum, followed by Water saturation shift referencing (WASSR) correction of z-spectrum (27). ROIs of placental perfusion regions (and phantom tubes) were manually outlined and magnetization transfer asymmetry (MTRasym) [defined as Msat(–) – Mctrl and Msat(+) are saturated scans with RF irradiation applied on negative and positive ppms, respectively] was plotted for each ROI. The GlucoCEST enhancement (GCE) is defined as the change in the integral of MTRasym between 0.75 and 4 ppm. The DGE is defined as the change in integral of Z spectra between 0.75 and 4 ppm with reference to the Z spectrum of the first time point. The reference scans of each dynamic CEST acquisition, which are T2*-weighted EPI images, were used to obtain the dynamic change of R2*, as ΔR2* = log(S/S0, baseline)/TE. Therefore R2* changes were recorded concurrently as GCE changes.

**Figure 1** Sample configuration of the placental tissue study. (A) The fetal surface of the placenta, yellow lines indicate where the placenta was sectioned after perfusion. Each segment and its glucose concentration were labeled; (B) illustration of perfused placental tissue positioned in a cylindrical container.

**Table 1** Subject demographics

| Subject | Gestational age | Position        | Fast (h) |
|---------|----------------|-----------------|----------|
| 1       | 33 weeks       | Left lateral    | 2        |
| 2       | 35 weeks       | Left lateral    | 2        |
| 3       | 30 weeks + 5 d | Left lateral    | 2        |
| 4       | 34 weeks + 6 d | Right lateral   | 2        |
| 5       | 35 weeks       | Left lateral    | 2        |

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Nevertheless, villitis (45x121), perfused with 10 mM glucose was found to have chronic (45x135), with the glucose perfusion concentration (45x148), tissue (30) perfused "non-perfused placental tissue showed higher GlucoCEST (45x162), placenta is reported to be around 4 mM (10% v/v) (28) may be due to blood sugar in the fetal vasculature (45x270). This observation is also consistent with previous work that compared GCE effect in "glucose perfused" placenta and "saline perfused" placenta (29). Glucose-perfused placental tissue showed higher GlucoCEST effect than the non-perfused control placental tissue (45x377), the GCE peaked when $B_1=2 \, \mu T$ (25) and this finding was used as a reference in optimization placental tissue GlucoCEST MRI.

In the ex vivo placenta, the sub-regions of perfused tissue were identified based on the T2* weighted image (Figure 3A). CEST effect in the non-perfused tissue may be due to blood sugar in the fetal vasculature (10% v/v) (28). The normal serum glucose level in the placenta is reported to be around 4 mM in vivo (29). Glucose-perfused placental tissue showed higher GlucoCEST effect than the non-perfused control placental tissue (45x377), this observation is also consistent with previous work that compared GCE effect in "glucose perfused", "non-perfused", and "saline perfused" placenta tissue (30). However, the GlucoCEST MRI effect did not scale linearly with the glucose perfusion concentration (45x377), Notably, the section of placenta tissue perfused with 10 mM glucose was found to have chronic villitis (45x385), which may explain its lower GCE. Nevertheless, the tissue perfused by 20 mM glucose showed only slightly higher GCE than that of 5 mM glucose may be due to normal variations in packing density of villi and the number of vessels/capillaries per villi, which could be a major confounding factor when inferring glucose content from the GCE intensity.

The in vivo experiments were performed in five volunteers whose gestational age ranged from 30 to 35 weeks. The duration of the dynamic scans varied for each subject, from 10 to 18 min, due to the different time needed to re-position the imaging field of view to match the baseline scan, while the maximum stay in the scanner for each subject was fixed. Nonetheless, the dynamic scans were kept within 30 min of the glucose drink. Example of the z-spectra of one subject is shown in Figure 4A. The baseline scans that were taken before glucose drink resulted in sizable variations of the GCE (−0.8 to 1.8) among individuals. Given the subject position changes between the baseline and the dynamic scans, the DGE of glucose was reported in reference to the first dynamic scan (Figure 4B). And the tendency of DGE increase during the first 30 min within glucose drink is consistent for all five subjects. Further, the $\Delta R^2*$ was also assessed by dynamic T2* weighted images in reference to the first time point after the glucose drink (Figure 4C). The time series of $\Delta R^2*$ and the time series DGE exhibit similarities in most subjects, however, they do not have significant correlations.
Discussion

Our study investigated GlucoCEST MRI at 3.0 Tesla and preliminarily tested DGE MRI in human placenta following oral glucose tolerance test. The DGE signal increase was observed for each individual within 30 min after the glucose drink. We have demonstrated the feasibility of GlucoCEST as a novel imaging test for characterizing placental glucose transport.

The ex vivo study demonstrated that GCE is promising for detection of placental pathologies. Glucose perfused tissue exhibited higher GCE than the non-perfused tissue. Although we did not show a simple linear relationship between GCE and glucose content, the results could be partially explained by the histological findings of variable tissue perfusion possibly affected by localized placental pathology. The vessel perfusion and vascular density in the placenta tissue directly affect the volume fraction of fetal blood in a single imaging voxel and thus affect the magnitude of GCE effect. Because the maternal/fetal glucose gradient is only 1.22 mM (fetal 3.48–3.87 and...
maternal 4.4–4.7 mM) (29), the glucose concentration is largely homogenous across the placenta, and therefore GCE might be sensitive to regional pathologies such as infarction and chronic villitis.

The assessment of placental glucose is somewhat confounded by several known factors that may affect the GlucoCEST MRI effect, such as field inhomogeneity and motion (31, 32) and factors that are specific to placenta structure, such as volume fractions of fetal and maternal blood. For example, the absolute GCE effect is sensitive to the B₁ field. In the human placenta in vivo (~20 cm in diameter), the B₁ field may vary up to 40%, which could translate to 20% difference in GCE according to previous analysis of CEST effect (25). In addition, the blood sugar level varies throughout the day, whereas the normal range of fasting blood sugar level (3.9–5.5 mM) is close to that of prediabetes (5.5–7 mM) and diabetes (more than 7 mM) (33).

In light of the technical challenges, further development of GCE is needed before it can reliably characterize placental glucose transport. Although the time to peak generally occurs within 2–4 min after IV glucose infusion (21), the time to peak is highly variable following oral glucose drink (34). Despite these challenges, the in vivo DGE showed a consistent trend in all subjects during glucose challenge, which is very encouraging.

It has been shown that apart from the CEST effect, glucose also reduces the water T2 relaxation due to chemical exchange, which would broaden the water saturation curve (35, 36). T2 change of the glucose perfused ex vivo tissue is consistent with the GCE effect, though may be confounded by blood wash out. The in vivo experiments show the ΔR₂* increases that are largely similar to the DGE signal, which is confirmative of the glucose enhancement effect.

There are several limitations of the current study, for

Figure 4 In vivo DGE MRI following oral glucose tolerance drink. (A) Example of Z-spectra in dynamic scan. (B) the DGE MRI of all subjects with reference to the first time point of the dynamic scan; (C) the dynamic R₂* changes of all subjects with reference to the first time point of the dynamic scan. Data (lines) were smoothed with a moving average filter of five points, whereas each data point (circles) were shown as original without any smoothing.
example, the protocol of glucose drink required the subject to sit up and drink the glucose in order to prevent reflux, thus causing inconsistency between baseline measurements and the dynamic measurements. It was also challenging to get a blood sugar level validation, since even a simple serum glucose measure with a finger prick requires the patient to be out of the scan room and sitting down comfortably, which takes about 5–10 min. Thus, the values would not be matched in time. Finally, fetal organs such as fetal livers would be of interest to observe, but accurate assessment of glucose change was challenging due to the fetal motion. In addition, motion caused by maternal free breathing, fetal motion, Braxton-hicks contractions, as well as blood perfusion change induced by increased blood sugar level, needs to be considered in future studies. Quantitative validation of the dynamic GCE signal may also require iv administration for a more rapid bolus, so as to enable correlation between serum glucose and each dynamic scan. Here, we chose to do a simple noninvasive feasibly study as the more invasive study design would need to be justified by a successful feasibility study.

Conclusions

Our study demonstrated the feasibility of DGE MRI in the human placenta following an oral glucose load at 3.0 Tesla. We found that in vivo DGE showed a consistent trend in all subjects during glucose challenge.

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Footnote

Conflicts of Interest: The authors have no conflicts of interest to declare.

Ethical Statement: The study was approved by the Institutional Review Board at the Boston Children’s Hospital and Massachusetts General Hospital.

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