Protective Effects of Epigallocatechin Gallate After UV Irradiation of Cultured Human Lens Epithelial Cells

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Purpose: To evaluate the protective effects of epigallocatechin gallate (EGCG) against UV irradiation of cultured human lens epithelial cells.

Methods: We irradiated cultured human lens epithelial cells with a 30-second pulse from a UV lamp with an irradiance of 0.6 mW/cm². Five minutes and 1 hour after UV irradiation, we administered 0, 5, 10, 15, 25, 50, or 100 μM EGCG. The cell number was measured with a microscopic counting chamber and cell viability was evaluated using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay.

Results: Compared to untreated cells, the total number of cultured human lens epithelial cells was markedly higher after UV irradiation. In a dose-dependent manner, viability was also higher in EGCG-treated cells.

Conclusions: EGCG increased the cell count and cell viability after UV irradiation of cultured human lens epithelial cells, indicating that EGCG can protect lens epithelium against UV damage.

Key Words: Cell viability, Cultured human lens epithelial cells, Epigallocatechin gallate (EGCG), 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay, UV irradiation

Ocular light injury is caused by several mechanisms, in particular, by short wavelength ultraviolet light. Ultraviolet light is divided into three categories: UVC (200-290 nm), UVB (290-320 nm) and UVA (320-400 nm). The ultraviolet light between 200 nm and 300 nm absorbs in the cornea, but ultraviolet light between 300 nm and 400 nm penetrates the cornea and is absorbed into the lens. The mechanistic cause of cataracts is complicated, but they can arise as a result of systemic diseases like diabetic mellitus, ultraviolet light, heat, hormone abnormalities, and smoking. Epidemiological reports and experiments indicate that ultraviolet light may be a primary cause of cataracts. The exact mechanism of UV-induced cataract formation is not fully understood. Cataract formation results from the accumulation of UV-induced DNA damage, changes in membrane transport and permeability, and changes in the biochemistry and physiology of the lens by reactive oxygen species such as hydrogen peroxide (H₂O₂), superoxide anion (O₂⁻), singlet oxygen (¹O₂), and hydroxyl radical (HO⁻). Flavonoids are a class of natural biological products that have evolved to protect plants from oxidative damage induced by chronic exposure to ultraviolet light. Flavonoids have many physiological health benefits, including protection from cardiovascular disease and cancer, and most of these beneficial effects are thought to stem from their potent antioxidant and free radical-scavenging properties, as well as their abilities to modulate many cellular enzyme functions.

The toxicities of most effective flavonoids are low. These include the dietary flavonoids such as fisetin, luteolin, quercetin, eriodictyol, baicalein, galangin, and EGCG, as well as synthetic flavonoids such as 3, 6-dihydroxy flavonol, and 3, 7-dihydroxy flavonol. (-)-epigallocatechin-3-gallate (EGCG) is the major polyphenolic constituent found in green tea. Several other polyphenolic compounds, known as catechins, are also found in green tea, though to a lesser degree. In addition to EGCG, the catechins include (-)-epicatechin-3-gallate (ECC), (-)-epigallocatechin (EGC), (-)-epicatechin (EC), and (+)-catechin. More than 50% of the tea catechin mass is composed of EGCG, and a vast body of scientific research suggests that EGCG (and the other catechins) are responsible for the majority of the potential health benefits attributed to green tea consumption. However, few studies have assessed the protective effects of EGCG after UV irradiation of human lens epithelial cells.

To evaluate the protective effects of EGCG against UV irradiation of cultured human lens epithelial cells we treated the cells with different concentration of EGCG. Five minutes...
and 1 hour after irradiation, we measured the effects on cell survival and viability. We choose these time points with future in vivo experiments in mind, in particular those that would evaluate green tea absorption and the protective effects of EGCG on the lens epithelium. Specifically, most people drink green tea indoors from 5 minutes to 1 hour after outdoor UV exposure. After another 24-hour incubation, the total cell count was measured and cell activity was evaluated using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay. The aim of this study was to determine whether EGCG can protect human lens epithelial cells from UV-induced damage, and to develop a new in vivo model of UV protection for human lenses using epigallocatechin gallate (EGCG).

**Materials and Methods**

1. **Cell culture**

   The human lens epithelial line CRL-1142 (HLE-B3) (ATCC, USA) was used for this study. These cells were cultured in Dulbecco's Modified Eagle's Medium (DMEM) (Sigma, St Louis, MO, USA), supplemented with 10% fetal bovine serum (FBS), 100 U/ml penicillin, 100 µg/ml streptomycin, and 25 µg/ml nystatin, and were cultured at 37°C in humidified atmosphere, containing 5% CO₂.

2. **Ultraviolet Irradiation**

   Human lens epithelial cells were irradiated for 30 seconds with an irradiance of 0.6 mW/cm² after removal of serum-free growth medium. UV irradiance was modulated by adjusting UV lamp (FS-20 T12-UVB, National Biological Corp.) output, which was calibrated with a UV sensor. The UVB spectrum was between 280 nm and 320 nm with peak irradiance at 312 nm. The total UVB exposure volume was 18 mJ/cm².

3. **Experimental Protocol and Groups**

   Human lens epithelial cells were cultured for 1 day and the experiments were performed on 3 groups: the normal control group, which was not exposed to UV lights (Group I); the second group exposed to UV lights without administration of EGCG (Sigma, St Louis, MO, USA) (Group II); and the third group exposed to UV lights with administration of EGCG (Sigma, St Louis, MO, USA) (Group III). Group III was treated with 5, 10, 15, 25, 50, or...
100 μM EGCG (Sigma, St Louis, MO, USA), 5 minutes or 1 hour after UV irradiation.

4. Cell count and MTT assay

24 hours after UV incubation, the cell count was measured using a microscopic counting chamber, and cell viability was evaluated using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay. In this assay, MTT is reduced to purple formazan in the mitochondria of living cells. A solubilization solution is added to dissolve the insoluble purple formazan product into a colored solution. The MTT assay was carried out using a standard protocol, and optical density was measured at 570 nm using a spectrophotometer.

5. Statistical analysis

To increase the reliability of the data, all experiments were repeated 5 times and average values were calculated. SPSS for Windows Version 10.1 was used to compute routine statistics. The data were analyzed for significance using repeated measures two-way ANOVA, followed by a Duncan’s multiple range test of post hoc tests, and were expressed as a mean percentage of the control value plus S.E.M. P values < 0.05 were considered significant.

Results

Epigallocatechin gallate (EGCG) protected cultured human lens epithelial cells from oxidative-stress-induced cytotoxicity with excellent efficacy. Compared with unirradiated cells (Fig. 1A), exposure to UV irradiation without administration of EGCG resulted in significant cell loss (Fig. 1B). With administration of 100 μM EGCG, both 5 minutes (Fig. 1C) and 1 hour (Fig. 1D) after UV irradiation, the cell count increased to those seen in unirradiated cells. The cell count of cultured human lens epithelial cells after UV irradiation markedly increased upon EGCG administration (Fig. 2). There was no significant relationship between the time to EGCG administration and cell loss (P=0.719). Cell viability after UV irradiation increased in a dose-dependent manner upon treatment with EGCG, as determined by the MTT assay (Fig. 3). Also, there were significant differences in viability between cells treated with 5 or 10 μM (P=0.000) and 10 or 15 μM EGCG (P=0.000). There was no significant difference between cells treated with 0 or 5 μM EGCG (P=0.233) or between cells treated with 15, 25, 50, or 100 μM EGCG (P=0.166). Compared with untreated cells, viability increased by approximately 24.8% or 21.5% in cells treated with 100 μM of EGCG, after 5 minutes or 1 hour after UV irradiation, respectively. There was no significance
between the time to the administration of EGCG and the survival rate of cells \((P=0.129)\).

**Discussion**

The exact mechanism of UV-induced cataract formation is not yet fully understood. However, UV-induced cataract formation does result from the accumulation of UV-induced DNA damage, changes in membrane transport and permeability, and changes in biochemical and physiology of lens epithelium by reactive oxygen species. Increasing interest in the health benefits of tea has led to the inclusion of tea extracts in dietary supplements and functional foods. While green tea contains a number of bioactive chemicals, it is particularly rich in catechins. Although a number of in vitro and in vivo studies have shown EGCG acts as an antioxidant and anti-apoptotic agent, the actual mechanism of its action as an antioxidant remains unclear. Importantly, total plasma antioxidant activity has been shown to increase following significant intake of green tea in laboratory studies. Moreover, an increase in plasma catalase activity and a decrease in nitric oxide levels occurred, suggesting that catechins have both direct antioxidant effects and indirect influences to increase the activity of other antioxidants or enzymes. It has been suggested that catechins, like EGCG, elicit antioxidant effects in a number of ways. However, few reports have demonstrated the protective effects of EGCG after UV irradiation on human lens epithelium. Therefore, we evaluated the effects of EGCG treatment on human lens epithelial cells after UV-induced damage. We also aimed to develop a new in vitro model of UV protection using epigallocatechin gallate (EGCG) for human lenses. In our study, the total cell count of cultured human lens epithelial cells after UV irradiation markedly increased upon EGCG treatment, compared with untreated cells. Also, cell viability markedly increased in a dose-dependent fashion upon EGCG administration, as determined by the MTT assay.

In conclusion, we have shown that EGCG increases cell count and cell viability after UV irradiation of cultured human lens epithelial cells, indicating the protective effects of EGCG against UV damage in cultured human lens epithelial cells. Future in vivo and in vitro studies examining mitochondria damage, nucleic acid damage, and phagocyte activity after UV exposure of human lens epithelial cells are needed.

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