Continuum Molecular Simulation of Large Conformational Changes during Ion–Channel Gating

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Abstract
A modeling framework was developed to simulate large and gradual conformational changes within a macromolecule (protein) when its low amplitude high frequency vibrations are not concerned. Governing equations were derived as alternative to Langevin and Smoluchowski equations and used to simulate gating conformational changes of the Kv7.1 ion-channel over the time scale of its gating process (tens of milliseconds). The alternative equations predict the statistical properties of the motion trajectories with good accuracy and do not require the force field to be constant over the diffusion length, as assumed in Langevin equation. The open probability of the ion–channel was determined considering cooperativity of four subunits and solving their concerted transition to the open state analytically. The simulated open probabilities for a series of voltage clamp tests produced current traces that were similar to experimentally recorded currents.

Introduction
Molecular dynamics (MD) simulations is a method used to model the molecular motion of proteins [1], including ion channels [2]. Simulations are conducted by solving the equations of motion for all atoms of a protein, starting from their known initial locations and assigned random initial velocities. The resultant motion is a high frequency, low amplitude vibration of the protein atoms. MD had limited success in predicting the large and gradual conformational changes that underlie the physiological function of many proteins (e.g. ion–channel gating). MD simulations are massive and usually can simulate up to 1 microsecond of protein dynamics and in some cases up to 1 milliseconds using special computer architectures [3], while the large conformational changes of ion–channels (and other proteins) occur over tens of milliseconds. Due to the very large number of degrees of freedom, it is impossible to analyze the motion in the entire configuration space. Therefore, MD generates a trajectory of conformational changes associated with assigned initial conditions for the atoms. To facilitate computations, MD simulations may be influenced to drag the structure toward preferred conformations by reducing the potential energy of those conformations [4,5]. Virtual increase of temperature to facilitate preferred conformations by reducing the potential energy of those simulations may be influenced to drag the structure toward conditions for the atoms. To facilitate computations, MD and gradual conformational changes that underlie the physiological function of many proteins (e.g. ion–channel gating). MD simulations are massive and usually can simulate up to 1 microsecond of protein dynamics and in some cases up to 1 milliseconds using special computer architectures [3], while the large conformational changes of ion–channels (and other proteins) occur over tens of milliseconds. Due to the very large number of degrees of freedom, it is impossible to analyze the motion in the entire configuration space. Therefore, MD generates a trajectory of conformational changes associated with assigned initial conditions for the atoms. To facilitate computations, MD simulations may be influenced to drag the structure toward preferred conformations by reducing the potential energy of those conformations [4,5]. Virtual increase of temperature to facilitate the passage of the structure through local minima has also been considered [6], as well as grouping atoms into coarse grains to reduce degrees of freedom [7]. With these manipulations, the motion trajectory samples a broader region in configuration space. The estimates for the potential of mean force [8], computed this way, rely on how accurate the motion trajectory represents the entire space of trajectories. Clearly, simulating with MD the microsecond dynamics of protein in a statistically meaningful manner is challenging, whereas simplified models allow one to study the millisecond dynamics in a computationally efficient way.

Large conformational changes usually involve gradual dislocations of protein segments which can be modeled with a limited set of degrees of freedom, \( x_i \). These degrees of freedom are usually translation and rotation of protein segments with reinforced secondary structure (e.g. helices). The proposed modeling framework of this paper simulates the protein dynamics within the entire configuration space of large conformational changes (all combinations of \( x_i \)), without explicitly simulating high frequency vibrations of single atoms and thermodynamic properties. It simulates the average conformational changes over \( \Delta t \), a time window sufficiently larger than the time between collisions. In this study we analyzed the molecular motion of particles under the influence of a conservative force field using a kinematic theory approach and derived the governing equations of the motion. These equations are more accurate alternatives to Langevin equation and Smoluchowski equation which have been used to model the gradual motion of proteins in a reduced configuration space, neglecting its atomic vibrations [9,10,11].

According to Newton's second law of motion, velocity of a particle (referred to as a target particle in this paper) on a molecular scale is determined by:

\[
\frac{m \, \ddot{v}_i(x_j)}{\dd t} = - \frac{\partial \Phi(x_j)}{\partial x_i} + \sum_k \delta(t - t_k)
\]

where \( m \) is the mass of the target particle, \( v_i \) is its velocity along coordinate \( x_i \), \( \Phi \) is the potential of any existing conservative force field, and the last term on the right represents the stochastic force...
of thermodynamic collisions on the particle. \( t_i \) is a time when another particle (referred to as a colliding particle in this paper) hits the target particle and \( \hat{c}_i \) is the impact momentum transferred to the target particle during the collision. The thermodynamic forces on the target particle have a non-zero mean that is proportional to the target particle velocity (in opposite direction). Therefore, Newton’s equation assumes the form of Langevin equation:

\[
\frac{m \ddot{v}_i(x_i)}{\partial t} = -\frac{\partial \Phi(x_i)}{\partial x_i} - \zeta v_i + f_s^i
\]

where \( f_s^i \) is a zero mean stochastic force and \(-\zeta v_i\) is a friction force that models the non-zero mean of the thermodynamic force.

However, modeling the non-zero mean of the stochastic force by the term \(-\zeta v_i\) alters the governing equation and consequently the stochastic properties of the resultant velocity. Assume the simplest case in the absence of a conservative force field. The actual motion of the target particle consists of constant velocity motions between any two consecutive impacts. At any collision incident, the velocity of the target particle changes abruptly by \( \Delta v_i = \hat{c}_i / m \) to a new random velocity. The expectation value of the velocity after the collision is a constant fraction of the velocity before the collision. But according to the Langevin equation, after any collision the expectation value of the velocity equals its velocity before the collision, and the magnitude of velocity decreases exponentially between collisions (rather than being constant). It means that Langevin equation overestimates the velocity magnitude right after a collision, and then compensates for this overestimation by reducing it toward zero between collisions. Therefore, the velocity ensembles that result from solving the Langevin equation are different from the actual velocity ensembles and consequently the Langevin equation may not be an accurate choice for determining the statistical properties of the motion (e.g. probability density function and autocorrelation of the velocity), or for estimating the motion trajectories when the stochastic motion is significant. An illustrative example of these Langevin equation properties is provided in the Text S1.

In Langevin model of motion, the stochastic force is assumed to be a zero mean Gaussian process with a Dirac delta autocorrelation function. Based on these assumptions, the variance of the stochastic force can be determined in terms of the variance of the velocity distribution using the fluctuation-dissipation theorem [12]. However, the prediction of Langevin equation for the velocity, in response to a stochastic force with a Gaussian distribution, does not have a Gaussian distribution (Figure S2 in Text S1); this contradicts the well known Boltzmann-Maxwell distribution (a Gaussian distribution) for the velocity (Figure S4 in Text S1). Compared to a Gaussian distribution with the same variance, the velocity distribution in Langevin equation has higher densities for velocity magnitudes in close vicinity of zero and for large velocity magnitudes (Figure S2 in Text S1). Because the friction term causes a decay of the velocity magnitude toward zero between the collisions (Figure S1 in Text S1), the probability of velocity magnitudes close to zero is increased (compared to the Gaussian distribution). And because the Langevin equation overestimates the velocity magnitude after a collision (Figure S1 compared with Figure S3) the probability of having large velocity magnitudes is increased as well. In addition, application of the Langevin equation is constrained to external force fields that are almost constant over the diffusion length.

Motivated by the need for a model that can accurately replicate the velocity distribution and does not constraint the conservative force field as in the Langevin equation, we introduce here a new modeling framework that is applicable to large, gradual conformational changes of a protein. We analyze the actual motion of a particle (protein segment) that undergoes multiple collisions in a probabilistic domain (a kinetic theory approach) and derive a governing equation for the average velocity. Unlike the Langevin equation, the governing equation of the average velocity does not have an inertial term. It should be emphasized that the inertial term is not neglected assuming a large friction coefficient (as assumed in the high friction limit of Langevin equation), it simply does not appear in the governing equation of the average motion after applying Newton’s law of motion for a system with multiple collisions. Because during the conformational changes of a protein segment the conservative force may vary significantly over the diffusion length, we do not consider it constant in our analysis and derive a more general equation for the effect of a conservative force on the motion trajectory and the probability distribution. The stochastic term in our equation appears as stochastic velocity (rather than force) and does not have a delta autocorrelation function. Note that the autocorrelation of the stochastic force is assumed to be a Dirac delta function in deriving the statistical properties of motion from Langevin equation and in deriving the Einstein-Smoluchowski relation between the friction coefficient and the diffusion constant. In reality, the autocorrelation decreases gradually over \( At \).

The newly developed modeling framework is used to simulate the conformational changes of the voltage sensor region (S1 to S4) of the Kv7.1 ion-channel during gating and the resultant open probability is compared to experimentally recorded macroscopic currents.

**Results**

We develop a model for the gradual motion of the helical transmembrane segments of an ion–channel protein. We use this model to simulate the motion trajectories and the transient probability distribution in the configuration space. Further, we use the results of these simulations to compute the single channel and macroscopic currents carried by the ion–channel. The model developed in this paper represents the stochastic motion of a particle (here a protein segment) on a molecular scale. It consists of two key sets of equations: equations (38)–(40) and equations (43), (47)–(49). The first set governs the average motion (over a time window) of the particle and can be used to simulate the motion trajectories. Compared to Langevin equation, a key feature of this equation set is that it accounts for potential fields that are not constant over the diffusion length by including the higher derivatives of the potential function. Additionally, it provides more accurate estimates for the stochastic properties of the motion. The second set of equations (derived based on the first set) governs the transient and steady state distribution of the particle in its configuration space. These equations also account for potentials that are not constant over the diffusion length.

For simulating the structural dynamics and electrophysiological function of the ion–channel, the first equation set is used to generate motion trajectories and from those the single channel current traces. The second equation set is used to compute the channel open probability and the macroscopic current through a large ensemble of ion channels.

**Equation of Motion for a Protein Segment**

The stochastic motion of a target particle (representing a protein segment) and colliding particles are considered in 3D Cartesian
space. Because of symmetry, the velocity of the colliding particles is uniformly distributed in all directions. Also, the probability that a point in space is occupied by a colliding particle is equal everywhere. Consider a target particle moving with a velocity \( v_0 \) in the space shown in Figure 1 where the \( z \) direction is chosen along the velocity. Assume an arbitrary location on the surface of the target particle that may be hit by a colliding particle. Panel A shows the impact point and the relative position of the target particle and the colliding particle. An impact occurs if the velocity of the colliding particle along the impact direction (dashed line) is larger than the velocity of the target particle along this direction: \( v_n > v_0 \cos \theta \). The tangential velocity of the colliding particle, \( v_t \), can have any value. For a collision at an opposite location (panel B), the condition of impact is: \( v_n < v_0 \cos \theta \). Therefore, along any impact direction the colliding particle may have any velocity. This means that the probability distribution of the impact direction, the colliding particle velocity and consequently the time and distance between the collisions are independent of the target particle velocity. The impact direction may be quantified by angles \( \theta \) (between 0 and \( \pi/2 \)) and \( \varphi \) (between 0 and \( 2\pi \)) in a spherical coordinate system. Particles with equal mass exchange their coordinate system. Particles with equal mass exchange their

\[
\begin{align*}
\langle w_z \rangle &= \frac{1}{2\pi} \int (v_n \cos \theta + v_0 \sin^2 \theta) \sin \theta g(v_n) d\theta d\varphi dv_n = \frac{2}{3} v_0 \\
\langle w_y \rangle &= \frac{1}{2\pi} \int (v_n \sin \theta - v_0 \sin \theta \cos \theta) \sin \varphi \sin \theta g(v_n) d\theta d\varphi dv_n = 0 \\
\langle w_z \rangle &= \frac{1}{2\pi} \int (v_n \sin \theta - v_0 \sin \theta \cos \theta) \cos \varphi \sin \theta g(v_n) d\theta d\varphi dv_n = 0
\end{align*}
\]

where \( g(v_n) \) is the probability density function of a 1D component of the velocity. It should be emphasized that \( \langle \vec{w} \rangle \) is independent of \( g(v_n) \). The expectation of the velocity in the \( z \) direction is non-zero, meaning that the stochastic velocity of a particle is not memory-less and therefore its autocorrelation function is not a delta function. However, this value vanishes after about ten impacts, implying that the autocorrelation approaches zero after a time required for about 10 impacts.

If there is no conservative force field acting on the particle, it will have a zero mean normally distributed stochastic velocity, \( v'_t \). The conservative force will add a deterministic velocity, \( v''_t \) to this zero mean stochastic velocity (causing the resultant stochastic velocity of the particle to have a non zero mean):

\[
v_t = v'_t + v''_t
\]

Assume that \( v'_n(k) \) is the velocity of the particle right before the \( k^{th} \) impact, \( v''_t(k) \) its velocity right after the \( k^{th} \) impact and \( t_k \) is the time interval between the \( k^{th} \) and \((k+1)^{th}\) impacts. Between the \( k^{th} \)

Figure 1. Collision between a colliding particle (light gray) and the target particle (dark gray) along an arbitrary direction of impact (dashed line). The colliding particle can have any velocity for an impact along any direction. Panel A shows a collision when the velocity of the colliding particle along the impact direction (\( v_n \)) is faster than the velocity of the target particle along this direction. In panel B, the velocity of the colliding particle is slower.

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and \((k+1)\text{th}\) impact, the particle accelerates under the influence of the conservative force field and as a result, its deterministic velocity increases by \(\Delta v_i^d(k)\) right before the \((k+1)\text{th}\) impact is:

\[
v_i^d(k+1) = v_i^d(k) + \Delta v_i^d(k)
\]

During an impact, two particles exchange some (or all, if they have equal masses) of their momentum in the direction of impact. Depending on the direction of impact and mass of the particles, the particle loses a fraction of its velocity and gains a fraction of the velocity of the colliding particle:

\[
v_i^c(k+1) = \gamma_k v_i^c(k) + \lambda_i v_n^c = v_i^c(k+1)
\]

where \(v_i^c(k+1)\) is the velocity of the particle immediately after the \((k+1)\text{th}\) impact, \(v_n^c\) is the normal component of the velocity of the colliding particle and \(\gamma_k\) is the fraction of particle velocity that is preserved during the impact. If there is no conservative force field, there would be no deterministic velocity and the velocity after the \((k+1)\text{th}\) impact is:

\[
v_i^c(k+1) = \gamma_k v_i^c(k) + \lambda_i v_n^c = v_i^c(k+1)
\]

From a statistical perspective, the stochastic velocity depends on the probability of collision, and on the probability distribution of the impact direction and the colliding particle velocity. These statistical parameters are independent of the velocity of the target particle and consequently its deterministic part. Therefore, the stochastic component of the velocity would be the same with or without conservative force field, and we may conclude that:

\[
v_i^s(k+1) = \gamma_k v_i^s(k) + \lambda_i v_n^s
\]

Note that \(\gamma\) is a random variable that obtains random values according to its probability distribution at each collision. The magnitude of \(\gamma\) is always less than 1. The deterministic velocity can be calculated as:

\[
v_i^d(k) = \sum_{j=0}^{k-1} \left(\prod_{j} \frac{\Delta t_j}{\Delta x_j}\right) \Delta v_i^d(j)
\]

where \(j\) refers to all collisions prior to the \(k\text{th}\) collision. The right side of equation (14) is a convergent series. Note that although we used the notion of infinity, in practice the series approaches a constant value if we calculate it up to a few preceding collisions (~10 for equal mass collisions).

To find the global velocity of the particle we average the velocity over a time interval \(\Delta t\). The time interval is chosen sufficiently long, such that the average of a stochastic parameter over \(\Delta t\) closely approximates its expectation value, and sufficiently short such that gradual conformational changes are small during \(\Delta t\). \(\bar{v}_i(t)\) is defined as the average of the particle velocity over this time window:

\[
\bar{v}_i(t) = \frac{1}{\Delta t} \int_{t-\frac{\Delta t}{2}}^{t+\frac{\Delta t}{2}} v_i(t') dt' = \frac{1}{\Delta t} \int_{t-\frac{\Delta t}{2}}^{t+\frac{\Delta t}{2}} v_i(t') dt' + \frac{1}{\Delta t} \int_{t-\frac{\Delta t}{2}}^{t+\frac{\Delta t}{2}} v_i^s(t') dt'
\]

We define \(\bar{v}_i^s(t)\) as the stochastic component of the average velocity. It is the average of the stochastic component of velocity, \(v_i^s(t)\):

\[
\bar{v}_i^s(t) \equiv \frac{1}{\Delta t} \int_{t-\frac{\Delta t}{2}}^{t+\frac{\Delta t}{2}} v_i^s(t') dt'
\]
Note that the motion is a multidimensional motion. Equation (20) integrates over the \( x_i \) direction, but the location of the particle \( x_j \) varies in all directions. Because \( \overline{v}^j \propto \overline{v}^i \) and \( \overline{v}^i \) is constant between impacts we may write:

\[
\frac{x_j(t) - x_j(t_k)}{x_j(t) - x_j(t_k)} = \frac{\Delta x_j(t) - \Delta x_j(k)}{\Delta x_j(t) - \Delta x_j(k)} = \overline{v}^j(k)
\]

(21)

and consequently:

\[
\overline{v}^i(k)\Delta x_j(t) = \overline{v}^j(k)\Delta x_j(k) + \overline{v}^i(k)(\Delta x_j(t) - \Delta x_j(k))
\]

(22)

where \( i \) and \( j \) refer to any two arbitrary degrees of freedom and \( \Delta x_j(k) \) refers to \( \Delta x_j(t_k) \). Substituting \( f \) with its Taylor expansion in equation (20) and substituting for \( x_j(t) \) from equation (22) we may write:

\[
\overline{v}^i(t) = \overline{v}^i(t_k) + \frac{1}{\overline{v}^i(k)}(\Delta x_j(t) - \Delta x_j(k))f_j + \frac{(\Delta x_j(t) - \Delta x_j(k))^2}{2\overline{v}^i(k)} \sum_j f_j^2_j \\
+ \frac{(\Delta x_j(t) - \Delta x_j(k))^2}{2\overline{v}^i(k)} \sum_j \overline{v}^j(k)\Delta x_j(k) + \frac{(\Delta x_j(t) - \Delta x_j(k))^2}{4\overline{v}^i(k)} \sum_j \overline{v}^j(k)\Delta x_j(k) + \frac{(\Delta x_j(t) - \Delta x_j(k))^2}{2} \sum_j \Delta x_j(k)\Delta x_j(k) + O(\Delta x^3)
\]

(23)

Using equation (23), the \( \Delta v^i(k) \) that is defined in equation (10) is:

\[
\Delta v^i(k) = \overline{v}^i(t_{k+1}) - \overline{v}^i(t_k) = \tau_s f_i + \frac{\tau_s^2}{2} \sum_j f_j^2_j \\
+ \frac{\tau_s^2}{6} \sum_j \overline{v}^j(k)\Delta x_j(k) + \frac{\tau_s^2}{2} \sum_j \overline{v}^j(k)\Delta x_j(k) + \frac{\tau_s^2}{12} \sum_j \Delta x_j(k)\Delta x_j(k) + O(\Delta x^3)
\]

(24)

And the average of the deterministic component of the velocity is:

\[
\overline{v}^i(t) = \frac{1}{\Delta t} \sum_{k=1}^{N} \int_{t_k}^{t_{k+1}} \overline{v}^i(t)\,dt = \frac{1}{\Delta t} \sum_{k=1}^{N} \overline{v}^i(t_k) \int_{t_k}^{t_{k+1}} \overline{v}^i(t)d\Delta x_j
\]

\[
= \frac{1}{\Delta t} \sum_{k=1}^{N} \{\tau_s \overline{v}^i(t_k) + \frac{\tau_s^2}{2} \sum_j f_j^2_j + \frac{\tau_s^2}{6} \sum_j \overline{v}^j(k)\Delta x_j(k) + \frac{\tau_s^2}{2} \sum_j \overline{v}^j(k)\Delta x_j(k) + \frac{\tau_s^2}{12} \sum_j \Delta x_j(k)\Delta x_j(k) + O(\Delta x^3)\}
\]

(25)

The expectation value of \( \overline{v}^i(k) \) can be determined using equations (14) and (24) as:

\[
\overline{\langle \overline{v}^i(k) \rangle} = \sum_{j=-\infty}^{k-1} \overline{\langle \overline{v}^i(j) \rangle} + \sum_{j=k}^{\infty} \overline{\langle \overline{v}^i(j) \rangle} = \overline{\langle \overline{v}^i(j) \rangle} \sum_{j=-\infty}^{k-1} \frac{\overline{\langle \overline{v}^i(j) \rangle}}{1 - \overline{\langle \overline{v}^i(j) \rangle}}
\]

(31)
It should be mentioned that because of the symmetry of the motion (location and velocity) in both the negative and positive directions, all the odd derivatives of $f_i$ will be eliminated. Combining equations (30), (31) and (32), the deterministic component of the velocity is:

$$\langle \Delta v_i^d(j) \rangle = \tilde{v}_i + \frac{1}{6} \sum_j \sigma^2 \frac{\partial^3 f_i}{\partial x_j^3} + \frac{1}{2} \sum_j \sigma \frac{\partial^2 f_i}{\partial x_j^2} + (\Delta x)^3$$

(32)

And the equation of motion for the average velocity is:

$$\dot{v}_i(t) = \tilde{v}_i - \frac{1}{\omega} \sum_j \frac{\partial^3 \Phi}{\partial x_i \partial x_j^2} + \frac{1}{\omega} \sum_j \frac{\partial^2 \Phi}{\partial x_i \partial x_j} \frac{\partial x_j}{\partial x_i} + \text{[higher order terms]}$$

(37)

In general, the characteristic parameters of the motion ($\eta, \gamma, \sigma$ and $\lambda$) are different for different degrees of freedom and we write equation (37) in a more general form:

$$\dot{v}_i(t) = \tilde{v}_i - \frac{1}{\zeta_i} \sum_j \frac{\partial^3 \Phi}{\partial x_i \partial x_j^2} \frac{\partial x_j}{\partial x_i} + \text{[higher order terms]}$$

(38)

where

$$\frac{1}{\zeta_i} = \frac{\tilde{v}_i(1+\gamma)}{2(1-\gamma)}$$

(39)
\[
\frac{1}{\omega_j} = \frac{\sigma_j^2 \tau_j}{12(1-\gamma_j)} + \frac{\hat{\gamma}_j \tau_j(1+\gamma_j)}{4(1-\gamma_j)} \tag{40}
\]

Probability Distribution of the Segment in Configuration Space

Time variation of the probability distribution is equal to the negative divergence of the probability flux. If the velocity field is a time invariant process, then the probability flux is the expectation value of the multiplication of the velocity field and the probability distribution. Therefore:

\[
\frac{\partial P}{\partial t} = - \frac{\partial}{\partial x_i} \langle v_i P \rangle \tag{41}
\]

For a stationary velocity field:

\[
\langle v_i P \rangle = \langle \bar{v}_i P \rangle \tag{42}
\]

Using equations (38) and (42) we may rewrite equation (41) as:

\[
\frac{\partial P}{\partial t} = \frac{\partial}{\partial x_i} \left( D_i \frac{\partial P}{\partial x_i} + \frac{P \partial \Phi}{\bar{v}_i \bar{v}_i} + P \sum_j \frac{1}{\omega_j \bar{v}_i \bar{v}_j} + [\text{higher order terms}] \right) \tag{43}
\]

The stochastic velocity leads to a diffusion (in the probability distribution) with a diffusion constant \(D_i\). At steady state, the time derivative of the probability distribution is zero:

\[
\frac{\partial P_{ss}}{\partial t} = - \frac{\partial}{\partial x_i} \left( D_i \frac{\partial P_{ss}}{\partial x_i} + \frac{P_{ss} \partial \Phi}{\bar{v}_i \bar{v}_i} + P_{ss} \sum_j \frac{1}{\omega_j \bar{v}_i \bar{v}_j} + [\text{higher order terms}] \right) = 0 \tag{44}
\]

This means that the total flux leaving a differential element in configuration space is zero. Imposing the principle of detailed balance (microscopic reversibility), probability flow in all directions of the configuration space should be zero (no circulation in probability space):

\[
\frac{\partial P_{ss}}{\partial x_i} = - \frac{P_{ss} \partial \Phi}{D_{\bar{v}_k} \bar{v}_k} - P_{ss} \sum_j \frac{1}{D_{v_k} \bar{v}_j \bar{v}_j} + [\text{higher order terms}] = 0 \tag{45}
\]

In order for \(P_{ss}\) to exist, its partial differentials in equation (45) must present a total differential, meaning that for any \(i\) and \(k:\)

\[
\frac{\partial}{\partial x_k} \left( \frac{P_{ss}}{D_{\bar{v}_k} \bar{v}_k} + P_{ss} \sum_j \frac{1}{D_{v_k} \bar{v}_j \bar{v}_j} + [\text{higher order terms}] \right) = \frac{\partial}{\partial x_k} \left( \frac{P_{ss}}{D_{\bar{v}_k} \bar{v}_k} + P_{ss} \sum_j \frac{1}{D_{v_k} \bar{v}_j \bar{v}_j} + [\text{higher order terms}] \right) \tag{46}
\]

Equation (41) is satisfied if and only if:

\[
D_{\bar{v}_i} \bar{v}_i = D_{v_k} \bar{v}_k = C_1 \quad ; \quad D_{v_i} \bar{v}_j = D_{v_k} \bar{v}_j \quad \text{and similarly for higher order terms} \quad \tag{47}
\]

where \(C_1\) and \(C_2\) are constants that vary with temperature. Therefore, the principle of detailed balance requires that the terms \(D_{\bar{v}_i} \bar{v}_i\) and \(D_{v_i} \bar{v}_j\) retain the same value for all \(i\). Under this condition the steady state distribution of probability has the following closed form:

\[
P_{ss} = A e^{-\frac{\Phi}{\alpha_t} - \sum_j \frac{1}{\alpha_j} \bar{v}_j \bar{v}_j} dV \tag{48}
\]

Simulation Results for Conformational Changes of the Kv7.1 Ion–Channel Voltage Sensor during Gating

**Degrees of Freedom.** A 3D structure of the Kv7.1 channel was derived previously [10,11] based on homology to Kv1.2 with a known crystal structure in the open state [13]. To simulate the protein structural dynamics, we first identify the significant degrees of freedom for large conformational changes.
during gating. A voltage dependent potassium channel consists of four similar polypeptides (subunits), each includes six transmembrane segments termed S1 to S6. S5 and S6 form the pore and S1–S4 form the voltage sensor that undergoes large conformational changes in response to variations in membrane potential. Voltage sensors of the four subunits are sufficiently distant so that their conformational changes can be assumed independent, except for the last transition to the open state that requires a cooperative transition of all four voltage sensors [14,15]. Within each voltage sensor, translation and rotation of S4 and S3 are associated with channel gating [16,17]. The relative location of S3 and S4 is constrained by the very short loop that connects them on one side and a salt bridge between S4 Histidine H240 and S3 Aspartic Acid D202 on the other side. Therefore, we assume that the significant degrees of freedom that can closely model the conformational changes during gating are translation and rotation of S4 and S3 as a single complex. Translation is perpendicular to the membrane surface, positive outward from the cell (\(z_{\text{direction}}\)); rotation is about an axis parallel to the first principal direction of the S4-S3 complex, located 5 Angstrom away from its center. The axes of rotation and translation were chosen to comply with the geometrical constraints imposed by the S4-S5 linker and to reduce the probability of a steric overlap of the S4-S3 complex with neighboring segments. Based on our recent loop closure technique [18], we found that the loop between the S3 and S2 segments is sufficiently large not to impose significant geometrical constraint. Based on experimental data [19,20], we allow S4-S3 to move outward up to 6 angstrom and inward up to 14 angstrom (in 0.2 angstrom steps) and rotate 1 radian in the clockwise and counter clockwise directions (in 0.05 radian steps).

**Energy Landscape.** Energy landscapes were constructed at multiple membrane potentials by computing the total electrostatic potential of a voltage sensor at all the conformations it assumes in the configuration space described above. The transmembrane potential adds a uniform electric field to the dielectric region (protein and membrane); it approaches zero outside this region. The potential energy of steric interactions was modeled with the Lennard–Jones potential [21]. Axis of rotation was adjusted by trial and error to eliminate any steric clashes between backbone atoms and also major clashes between side chain atoms. A penalty function was also applied for outward and inward motion of S4–S3 beyond 4 and 8 Angstroms respectively, to mimic the jump friction coefficients can be calibrated based on the time scale of the experimentally measured macroscopic current. Using the experimentally recorded currents, the friction coefficient was calibrated to \(\frac{\zeta_{\text{E0}}}{a^2} = 5 \times 10^{-3} \text{ kg} \cdot \text{Ang}^2 \cdot \text{s}^{-1}\) in \(z_{\text{direction}}\) and \(\zeta_\varphi = 12.5 \times 10^{-3} \text{ kg} \cdot \text{Ang}^2 / \text{s} \) in \(\varphi\) directions. The higher order frictions were neglected. Comparison of these friction coefficients requires transformation to a common dimension. An equivalent translational friction coefficient for rotation about an axis can be derived as:

\[\frac{\zeta_{\text{E0}}}{a^2} \approx \frac{\varphi}{a^2} \]  

(50)

Where \(a\) is the root mean square distance of the moving residues with respect to the axis of rotation, \(a\) is about 5 Angstrom in our simulation and we choose \(\zeta_\varphi = a^2 \zeta_{\text{E0}} = 25 \zeta_{\text{E0}}\), so that resistance to motion is similar for the two degrees of freedom.

Equation (43) was solved using the Finite Difference (FD) method to simulate the transition of probability distribution during step depolarization from resting potential of \(-100 \text{ mV}\) to a test potential of \(+60 \text{ mV}\) (activation test), and also during step repolarization from the depolarized potential of \(+60 \text{ mV}\) to the test potential of \(-100 \text{ mV}\) (deactivation test). These transitions and the resultant open probability were visualized for the activation and the deactivation tests in two supplemental movies (Movies S1 and Movie S2). By increasing the membrane potential from the resting potential of \(-100 \text{ mV}\) to the test potential of \(+60 \text{ mV}\), the transition starts with a rapid outward translation and counter clockwise rotation of the voltage sensors to an intermediate conformation, from which they gradually diffuse to a final conformation associated with an additional energy minimum at \(+60 \text{ mV}\) (Movie S1).

To check the accuracy of the FD scheme we computed the difference between 1 and the integral of probability distribution over the configuration space) during the transition:

\[e_1(t) = \int_{z_{\text{0}}}^{z_{\text{1}}} \int_{\varphi}^{\varphi_{\text{1}}} P_{\text{FD}}(z,\varphi,t)dzd\varphi - 1 \]  

(51)

Where \(P_{\text{FD}}(z,\varphi,t)\) is the FD solution of equation (43). \(e_1(t)\) was less than \(10^{-7} (<0.00001\% \text{ error})\). Additionally, we computed the difference between the probability distribution when the FD simulation approaches steady state (\(t=1000 \text{ ms}\)) and the probability distribution at steady state calculated analytically using equation (48):

\[e_2 = \int_{z_{\text{0}}}^{z_{\text{1}}} \int_{\varphi}^{\varphi_{\text{1}}} (P_{\text{FD}}(z,\varphi,t_f = 1000 \text{ ms}) - P_a(z,\varphi))dzd\varphi - 1 \]  

(52)
ε2 for the voltage clamp test from −100 mV to +60 mV was less than \(10^{-6}\) (less than 0.0001%). To ensure the accuracy of the FD solver, we refined the integration intervals. The FD solver maintained a relative error less than \(10^{-7}\) throughout the gating duration (Figure 5A).

Discretization of the configuration space was fine enough to capture all variations in energy smoothly; the maximum relative discretization errors over the energy landscape at all membrane potentials were less than 3% in \(z\) direction and less than 1% in \(\theta\) direction (Figure 5B).

**Activated and Resting States.** At the open state the S4–S3 complex is in an outward position, and during channel closing it moves inward between 7 and 13 Angstroms [19]. The probability distribution at +60 mV shows two high probability regions for voltage sensor conformations. Based on the above experimental observations, the region centered at \(z=2\) and \(\phi=0.2\) is the activated conformation, that at \(z=-5\) and \(\phi=0.6\) is the intermediate resting conformation and that at \(z=-10\) and \(\phi=0.7\) is the deep resting conformation. An outward conformation of the S4-S3 complex that allows channel opening is termed an activated conformation. The exact border between resting and activated conformations cannot be determined experimentally. We assume that the channel remains in the activated state within 2 Angstrom inward movement of S4-S3 relative to its position in the crystal structure [10,11,13] (\(z=0\) and \(\phi=0\) in configuration space is associated with the crystal structure conformation). The channel can transition to the open state only if the S4-S3 complexes of all four subunits are in the activated state. The permissive state is the channel state when all four subunits are in their activated states. The cooperative transitions of the channel from the permissive state to the open state and from the open state to the permissive state are modeled by Markovian transitions with transition rates \(a\) and \(b\), respectively.

Transitions to the open state reduce the concentration of voltage sensors in activated states (and consequently in resting states) at depolarized membrane potential due to flux of channels from the permissive state to the open state. Below we explain how to incorporate the cooperativity between the four subunits during transition to the open state in order to calculate the channel open probability. This extends the application of our previous

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Figure 4. Energy landscapes and their associated steady state distributions (in configuration space) for two different membrane potentials. A) Energy landscapes at two different membrane potentials \(V_m = -100\) mV (top) and +60 mV (bottom). Increasing membrane potential shifts minimum energy conformations of the S4–S3 complex toward more outward positions (right side of the energy landscape). Ellipses highlight the energy minima. Vertical black line is the border between resting and activated states. B) Steady state distribution of voltage sensors in the configuration space at a particular membrane potential; \(V_m = -100\) mV (top) and +60 mV (bottom). Channels mainly reside in minimum energy conformations (highlighted by the ellipses). Note that the brightness scale is logarithmic. White vertical line marks the border between resting and activated states. In response to a sudden change in membrane potential, voltage sensors distribution varies gradually toward the steady state distribution associated with the new membrane potential (Movies S1 and S2).

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formulation [15] to problems that simulate the conformational changes of the voltage sensor in continuum configuration space (rather than a discrete Markov model).

**Steady State Open Probability.** At steady state the net transition to and from the open state is zero and the probability distributions of the voltage sensors in configuration space are independent of each other. The open probability can be calculated as [15]:

\[
O_{ss} = \frac{z_{Ass}^4}{z_{Ass}^4 + \beta} = \frac{C_{Ass}^4}{C_{Ass}^4 + \frac{\beta}{z}}
\] (53)

Where \(C_{Ass}\) is the steady state probability of voltage sensors in activated conformations with no transitions to the open state:

\[
C_{Ass} = \int_{\text{Activated}} P_{\text{Ass}}(z,\varphi) \, dz \, d\varphi
\] (54)

The steady state open probability was calculated at all the test potentials assuming \(\beta/z = 0.02\). The resultant steady state open probabilities are plotted for different test potentials in **Figure 6**. The simulated curve has the typical S shape dependence on membrane potential. Note that there was no need to incorporate a voltage dependent transition between the permissive and open states to obtain this curve [14].

**Transient Open Probability.** We showed previously how to include the cooperativity of channel opening in computing channel open probability from the probability distribution of its subunits in a discrete Markov model [15]. An analogous approach is also applicable to a continuous distribution in the configuration space. Distribution of voltage sensors in the subunit configuration space changes during channel opening because of 1. redistribution of existing voltage sensors in the subunit configuration space in response to the altered (depolarized) membrane potential, 2. entrance or exit of subunits to or from the subunit configuration space via a net transient flux, \(F(t)\), from or to the open state. Therefore, the transient probability distribution in subunit configuration space, \(P_C\), is the sum of the probability distribution of the subunits that are initially in the subunit configuration space, and the probability distribution of the subunits that enter or exit the configuration space from or to the open state, \(P_E\):

\[
P_C(z,\varphi,t) = P_E(z,\varphi,t) + (1 - O_0)P_R(z,\varphi,t)
\] (55)

Where \(O_0\) is the initial open probability. \(P_E\) shows the distribution of the subunits that are initially in the configuration space (its integral over the subunit configuration space is set to 1). Therefore, probability distribution of the subunits that are initially in the configuration space among all the subunits (taking into account the subunits that are in open state as well) would be \((1 - O_0)P_R\), because only \((1 - O_0)\) of the subunits are initially in the subunit configuration space.

The transient flux can be determined using the transition rates \(z\) and \(\beta\):

![Figure 5. Finite Difference (FD) error in the numerical simulation. A) Time variations of the FD error for different integration steps. Each curve shows the integral over the configuration space of the difference between the solution with a given time step \((\Delta t)\) and the solution with the finest time step \((\Delta t_0 = 50\, \text{ns})\) as a function of time. FD simulations are reasonably accurate when they converge. \(\Delta t = 2.5\, \mu\text{s}\) was chosen in the simulations. B) Maximum relative discretization error in energy landscapes at different membrane potentials. doi:10.1371/journal.pone.0020186.g005](#)

![Figure 6. Steady state activation curve computed at several membrane potentials. Solid line shows the simulation results; dashed line is from experimentally measured currents. doi:10.1371/journal.pone.0020186.g006](#)
$F(t) = \beta O(t) - \alpha A(t)$  \hspace{1cm} (56)

where $A(t)$ is the probability of the channel occupying the permissive state. As a subunit can transition to the open conformation from any activated conformation, we express the channel open probability, $O(t)$, in terms of two activated-state probabilities, $U_A$ and $R_A$, defined as follows [15]:

$$O(t) = O_0 - \int_0^t F(\tau) d\tau$$  \hspace{1cm} (57)

$$F(t) + \int_0^t F(\tau) [\alpha U_A(t-\tau)^4 + \beta] d\tau = -\alpha (1 - O_0) R_A(t)^4 + \beta O_0$$  \hspace{1cm} (58)

$$U_A(t) = \iiint_{\text{Activated}} P_U(z,\phi,t) dz d\phi$$  \hspace{1cm} (59)

$$R_A(t) = \iiint_{\text{Activated}} P_R(z,\phi,t) dz d\phi$$  \hspace{1cm} (60)

Where $P_U$ is the solution of equation (43) (where there is no net flux to or from the open state) when all subunits are initially in activated conformations distributed proportionally to their final steady state distribution (no subunit is initially in resting conformation), and $U_A$ is probability of activated state under this condition. $R_A$ is the probability of activated state associated with the redistribution of the subunits that are initially in the subunit configuration space. Once the net flux, the open probability and the permissive state probability were computed using the above equations, flux from the open to the permissive state, $\alpha A(t)$, and from the permissive to the open state, $\beta O(t)$, can be determined.

Open probabilities for a series of voltage clamp tests from resting potential of $-100$ mV to test potentials of $-40$, $-20$, $0$, $20$, $40$ and $60$ mV were simulated and compared with experimentally recorded currents for the same protocol in Figure 7. The simulated transient open probabilities are similar to the experimentally recorded currents; both exhibit a sigmoidal shape (initial delay), biphasic activation (fast then slow) and slower activation at lower test potentials. In these simulations, the central difference method was used for FD simulations and the trapezoidal method was used to compute numerical integrations. The values of $\alpha$ and $\beta$ were calibrated to $0.2$ /ms and $0.004$ /ms. The current traces are similar to those calculated using Monte Carlo simulations and an intermediate Markov model in our previous study [10,11]. However, the different choice of configuration space in this study resulted in significantly lower energy barriers, so that scaling the membrane potential was not needed in the simulations presented here.

The trajectory of conformational changes within the configuration space can be estimated using equation (38). Figure 8 shows one motion trajectory of the average motion in configuration space. Although, the high frequency vibrations of the S4-S3 complex are averaged out, the motion trajectory of gradual conformational changes exhibits a stochastic behavior. Conformational changes of the Kv7.1 voltage sensor along this motion trajectory are animated [23] in Movie S3.

**Discussion**

Molecular motion of particles under the influence of a conservative force field was analyzed and equation (38) was derived for simulating the average motion trajectory. At the high friction limit, where “the effect of the Brownian forces on the velocity of the particle (friction term) is much larger than that of the external (conservative) force” [24], Langevin equation is reduced to a form similar to equation (38):

$$v_i = \frac{f_i^s}{k_i} - \frac{1}{\zeta_i} \partial \Phi(x_i)$$  \hspace{1cm} (61)

where $f_i^s$ represents the stochastic force associated with thermodynamic collisions. In equation (61) the stochastic velocity term, $\frac{f_i^s}{\zeta_i}$, is proportional to the stochastic force, $f_i^s$. However, from Newton’s second law of motion, velocity is proportional to the time integral of the force. This discrepancy is a consequence of eliminating the inertial term from the Langevin equation at the high friction limit.
dependent constant, $C$, are considered as the calibrating parameters. The temperature model. However, we chose $f$ freedom, as we had no reason to include anisotropy. This reduces that the friction coefficient is the same along all degrees of

Smoluchowski relation, derived for ideal gases. We also assumed

paper).

Because of steric clashes. The conformation at $z = 0$ and calibrated. Note that the position of the axis of rotation is restricted

resting conformations and the position of the axis of rotation were calibrated (along with $\phi$). For ion channels that undergo large conformational changes over $\Delta t$ this approximation could be inaccurate. In fact equations (38) and (43) enable the application of Brownian approach for studying the large and gradual conformational changes of proteins.

In our modeling approach, the friction coefficients, $\zeta_i$ and $\omega_\alpha$, are considered as the calibrating parameters. The temperature dependent constant, $C_1$, is also a calibrating parameter of the model. However, we chose $C_1 = kT$ based on the Einstein–Smoluchowski relation, derived for ideal gases. We also assumed that the friction coefficient is the same along all degrees of freedom, as we had no reason to include anisotropy. This reduces the calibrating parameters of the modeling framework to a single friction coefficient, $\zeta$ (i.e. $\zeta$ for the practical example of this paper).

For simulating the conformational changes of the Kv7.1 ion–channel during gating, the border between the activated and resting conformations and the position of the axis of rotation were calibrated. Note that the position of the axis of rotation is restricted because of steric clashes. The conformation at $z = 0$ and $\varphi = 0$ in configuration space is associated with the activated state (based on homology with Kv1.2 channel and its known crystal structure in the open state [13]). Channel transitions to the resting state occur by downward translation of S4 ($z<0$). The location where this state transition occurs (the border line in Figure 4) and the exact position of the axis of rotation were calibrated (along with $z$ and $\beta$, the transition rates between the activated state and the open state) to provide the best fit between the model prediction and the experimental steady state activation curve (Figure 6). Moving the border line shifts the steady state activation curve (about 10 mV for 1 Angstrom dislocation of the border line) but has negligible effect on its slope. Therefore, the slope of this curve can serve as a measure for evaluating the model prediction. This slope represents the sensitivity of the open probability to variations in the membrane potential and is replicated closely by the model. The dielectric constant of the lipid bilayer (as well as the protein) can affect the conservative force field and consequently the simulated open probability. Increasing the dielectric constants of the protein and the lipid bilayer to 8 (from 6) increases the slope of the steady state activation curve by about 6%, while decreasing it to 4 reduces this slope by about 12%.

The transient open probabilities in a series of voltage clamp tests (Figure 7) are also used to evaluate the predictive ability of the model. The friction coefficient $\zeta$ scales these curves in the time domain but does not change their shapes; it is calibrated to match the time course of activation of the simulated currents to that of the experimental currents.

A methodology for computing the energy landscape and relating it to the macroscopic current was developed in our laboratory and presented by Silva et al. [10]. In that approach, the macroscopic current was estimated using four identical Markov models representing the four channel subunits. Transition rates of the Markov model were assumed to be the reciprocal of the first passage time along an arbitrary path between the minima on the energy landscape. To overcome the complexity associated with the cooperativity between the four channel subunits, open probability was computed through Monte-Carlo simulations. In this paper we use a direct approach for relating the energy landscape to the channel open probability. We derive and solve the equation of motion to find the dynamics of subunit conformational changes in continuum configuration space. We then compute the open probability directly from the dynamics, without using an intermediate Markov model. The cooperativity among subunits is included analytically, avoiding the time consuming Monte-Carlo simulations.

Experimental measurements of the dynamics of gradual conformational changes are extremely difficult to perform, if not impossible. However, for the case of ion–channels these dynamics underly the dynamics of open probability, which can be recorded experimentally as channel conductance (current) under a variety of different conditions. Therefore, a close match between simulated and recorded open probability may be used as verification of appropriate simulations of conformational changes. Consistent with our previous studies [10,11,22], the energy landscape has two minima associated with two resting conformations: deep and intermediate. We envision that a more accurate match can be obtained by examining all non overlapping conformations using more than two degrees of freedom. Including more degrees of freedom can also provide more accurate estimates of the protein conformation at resting states [25].

Methods

The electrostatic energy of the protein was computed at different points of its configuration space using MATLAB codes. Protein and membrane were modeled as continuum dielectrics with dielectric constant of 6, consistent with the range of 2 to 25 determined for lipid bilayers [26,27,28] and a range of 2 to 20 determined for membrane proteins [29,30,31]. Intra- and extracellular electrolyte was modeled as a continuum with dielectric constant of infinity. Interfaces of membrane (and protein) with the abacus environment were modeled by parallel surfaces. The image
charge method was used to include the potential energy associated with the surface charge distribution caused by charged protein residues within the membrane [32]. Membrane thickness was assumed to be 30 Angstrom [33]. The protein was placed within the membrane so that the upper membrane surface is right above the E160 residue on the S2 segment. The surface charges were assumed to be distributed within 7 Angstrom from the interface (Debye-Huckel length) with a mean distance of 3 Angstrom. When a charged residue entered the electrolyte environment, the screening effect of this environment was considered by reducing its effective charge to zero within 3 Angstrom.

The governing partial differential equation of the motion (equation (43)) was solved using a central difference FD method and imposing natural boundary conditions. The FD was implemented using MATLAB programming environment and integrated over 50 ns time steps.

Supporting Information

Figure S1  Velocity trace of a particle computed using the Langevin equation during 10,000 impacts. Panel A) shows the entire trace and panel B) enlarges the small region marked by red ribbon in panel A). Dashed lines in panel B) mark the impact incidents. (TIF)

Figure S2 Probability density function of the velocity computed using Langevin model (blue curve) compared with its equivalent Maxwell-Boltzmann distribution (red curve). The expectation value of the velocity square is the same for both distributions. (TIF)

Figure S3 Velocity trace of a particle computed using the model developed in this paper during 10,000 impacts. Panel A) shows the entire trace and panel B) enlarges the small region marked by red ribbon in panel A). Dashed lines in panel B) mark the impact incidents. (TIF)

Figure S4 Probability density function of the velocity computed using the model developed in this paper (blue curve) compared with its equivalent Maxwell-Boltzmann distribution (red curve). The expectation value of the velocity square is the same for both distributions. (TIF)

Movie S1 Probability distribution of subunits in configuration space during gating, when the membrane potential increased to +60 mV from a resting potential of −100 mV. Vertical dashed line is the assumed border between resting and activated conformations. The white vertical curve shows the flux within configuration space across this border. The net flux from the permisive state to the open state is shown by the arrow exiting the subunit configuration space at the top right corner. Open state probability is shown (color scale) in the square on the upper right and plotted as a function of time in the right bottom panel. (WMV)

Movie S2 Probability distribution of subunits in configuration space during gating, when the membrane potential decreased to −100 mV from an activated potential of +60 mV. Vertical dashed line is the assumed border between resting and activated conformations. The white vertical curve shows the flux within configuration space across this border. The net flux from the open state to the permisive state is shown by the arrow exiting the open state toward the subunit configuration space at the top right corner. Open state probability is shown (color scale) in the square on the upper right and plotted as a function of time in the right bottom panel. (WMV)

Movie S3 A trajectory of conformational changes during gating, when the membrane potential is increased from a resting potential of −100 mV to a depolarized potential of +60 mV. The two orange helical segments are the S4–S3 complex. At −100 mV they are at their most downward location (deep resting). In response to increase of membrane potential to +60 mV they move upward (with slight rotation) to the intermediate resting state. They stay at that state for a while and then move upward along with a noticeable rotation to the activated state. Legend in upper right corner shows the voltage sensor state and in upper left corner the membrane potential. (WMV)

Text S1 Langevin equation prediction for the velocity distribution. The prediction of Langevin equation for the velocity, in response to a stochastic force with a Gaussian distribution, contradicts the Boltzmann–Maxwell distribution (a Gaussian distribution) for the velocity. Compared to a Gaussian distribution with the same variance, the velocity distribution in Langevin equation has higher densities for velocity magnitudes in close vicinity of zero and for large velocity magnitudes. (DOC)

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Author Contributions

Analyzed the data: AN. Contributed reagents/materials/analysis tools: YR. Wrote the paper: AN.

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