Generalised Leukaemic Gingival Enlargement: a Case Report

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ABSTRACT

Background: Acute myeloblastic leukaemia is a malignant bone marrow neoplasm of myeloid precursors of white blood cells. Due to its high morbidity rate, early diagnosis and appropriate medical therapy is essential.

Methods: The article highlights normal blood alterations like anaemia, thrombocytopenia, leukocytosis and advanced diagnostic aids like flow cytometry, special staining as a diagnostic modality as well as for prognostic information in acute leukaemia, particularly as a tool for assigning lineage and facilitating further pathologic classification which may be helpful in influencing treatment strategies.

Results: On clinical examination the case presented with features of inflammatory gingival enlargement with presence of local deposits and calculus. Routine blood examination anaemia, thrombocytopenia, leukocytosis with haemoglobin 5.6 gm% and total leucocyte count of 1,12,000 / cu mm suggestive of leukaemia. Myeloperoxidase and leukocyte nonspecific esterase (NSE) special stain were used which showed presence of myeloblasts in the peripheral smear suggestive of acute myelocytic leukaemia. Flow cytometry were done which further helped in interpretation of these cells which showed to be strongly positive for CD45, CD13, CD14, and anti HLADR and moderately positive for CD4, CD34 and Anti MPO confirming to be case of AML-M4 with 57.73% gating.

Conclusions: Fact that gingival alterations are sometimes the first manifestations of the disease implies that dental professionals must be sufficiently familiarized with the clinical manifestations of systemic diseases. The timely referral by the general dentist for a suspicious lesion provided an early diagnosis and early intervention reducing the patient morbidity.

Keywords: acute myeloid leukemia; flow cytometry; gingival hyperplasia.

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INTRODUCTION

Leukaemia is a hematologic disorder resulting from the proliferation of a clone of abnormal hematopoietic cells with impaired differentiation, regulation, and programmed cell death. Leukaemic cell multiplication at the expense of normal hematopoietic cells lines causes marrow failure, depressed blood cell count, and death as a result of infection, bleeding, or both [1]. The oral manifestations of leukaemia include gingival enlargement, oral ulcerations, gingival bleeding, petechia and mucosal pallor. Oral lesions occur in both acute and chronic form of all types of leukaemias; myeloid, lymphoid and monocytic. However, the oral manifestations are far more common in the acute stages of the disease and are most common in monocytic leukaemia [2].

Gingival enlargement because of infiltration of premature leukocytes in leukaemia is well documented in literature and is one of the most common symptoms leading to the diagnosis of leukaemia that directs the patients to seek dental consultation [3]. In this paper, we report a case of a patient who reported for bleeding gums to our department and was diagnosed with acute myeloid leukaemia incidentally on routine blood investigations followed by flow cytometry for staging of leukaemia.

CASE DESCRIPTION AND RESULTS

A 32 years old male patient reported to the Division of Periodontics, Armed Forces Medical College, Pune, India with the chief complaint of bleeding gum from last five months. He was apparently normal five months before and gradually developed pain and bleeding on brushing. On eliciting the medical history, patient was hospitalized six months before for breathlessness and enterocolitis. Patient also gave a history of mild weight loss and loss of appetite from last few months. General physical examination revealed that he was moderately built, poorly nourished had signs of anaemia and skin looked pale but did not have signs of cyanosis and jaundice. On intraoral examination generalised gingival enlargement was noticed (Figure 1). Gingival was bluish in colour with presence of ecchymosis in the floor of the mouth (Figure 2). His oral hygiene was poor with presence of local factors. On palpation, gingiva was soft and oedematous without stippling and was tender on palpation. Based on systemic and intraoral examination patient was advised to undergo routine blood investigations in the Department of Pathology and was also advised to report back with all old blood and hospitalization reports. He was diagnosed with acute myeloid leukaemia (AML-M4) a day later by peripheral smear examination by special staining (Figures 3 and 4) and flow cytometry. His old reports of hospitalization were checked which revealed that he had hepatomegaly and fluid filled oedematous bowel loops suggestive of enterocolitis. His comparative blood profile and normal blood count is given in Table 1.

Flow cytometry was done with forward scattering (FSC) versus side scattering (SSC) in which 57.73% cells were gated and the cells included all atypical cells. Further interpretation of these cells revealed strongly positive for CD45, CD13, CD14, and anti HLADR and moderately positive for CD4, CD34 and Anti MPO (Figure 5). These cells were negative for CD10, CD79a, CD7 and CD20 giving an impression of AML-M4. The patient was referred to haematooncologist for further management and chemotherapy but the patient expired four days later due to multi organ failure and secondary infection.

DISCUSSION

Leukaemia is classified as lymphocytic or myelocytic according to the lineage of white blood cells (WBC)
involved; a subgroup of the myelocytic leukaemias is monocytic leukaemia. According to their evolution, leukaemias can be acute, which is rapidly fatal; sub-acute; or chronic. In acute leukaemia, the primitive “blast” cells are released into the peripheral circulation, whereas in chronic leukaemia, the abnormal cells tend to be more mature with normal morphologic characteristics and function when released into the circulation [1]. The two most commonly used classification schemata for AML are World Health Organization (WHO) system and French-American-British (FAB) classification system. WHO classify AML into four subtypes as; M1 (acute myeloblastic leukaemia), M2 (acute myeloblastic leukaemia with maturation), M3 (acute promyelocytic leukaemia), M4 (acute myelo-monocytic leukaemia), M5 (acute monocytic leukaemia), M6 (acute erythroidleukaemia), and M7 (acute megakaryoblastic leukaemia). The morphologic subtype of AML also include a rare type not included in FAB system as ninth subtype, M8, as acute basophilic leukaemia [5].

Patients with AML generally present with symptoms related to complications of pancytopenia (anaemia, neutropenia, and thrombocytopenia) including weakness, and easy fatigue, infections of variable severity, and/or hemorrhagic findings such as gingival bleeding, ecchymoses, epistaxis or menorrhagia. In some cases atypical features like chin numbness and tooth pain has also been reported [6]. The expression of these signs is more common in acute and sub-acute

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**Table 1.** Comparison of blood indices

|                      | Initial blood picture (old hospitalization report) | Present blood picture (in department) | Normal value               |
|----------------------|--------------------------------------------------|--------------------------------------|-----------------------------|
| **Haemoglobin**      | 6.9 gm%                                         | 5.6 gm%                              | 11.5 - 16.5 gm%             |
| **RBC**              | 3.17 million / cu mm                            | 1.41 million / cu mm                 | 3.5 - 6.0 million / cu mm   |
| **Platelets**        | 75,000 / cu mm                                  | 25,000 / cu mm                       | 1,50,000 - 4,50,000 / cu mm |
| **TLC**              | 98,000 / cu mm                                  | 1,12,000 / cu mm                     | 4,000 - 11,000 / cu mm      |
| **DLC**              |                                                  |                                      |                             |
| Neutrophils 0.4%     |                                                  |                                      |                             |
| Lymphocytes 10%      |                                                  |                                      |                             |
| Neutrophils 15%      |                                                  |                                      |                             |
| Neutrophils 15%      |                                                  |                                      |                             |
| Eosinophil 4%        |                                                  |                                      |                             |
| Myeloblasts 25%      |                                                  |                                      |                             |
| Basophil 0%          |                                                  |                                      |                             |
| Monoblasts 15%       |                                                  |                                      |                             |
| Lymphocytes 77%      |                                                  |                                      |                             |
| Promonocytes 22%     |                                                  |                                      |                             |
| Myelocytes 4%        |                                                  |                                      |                             |
| Monocytes 20%        |                                                  |                                      |                             |
| Myelocytes 0.1%      |                                                  |                                      |                             |
| Promyelocytes 0.1%   |                                                  |                                      |                             |
| Metamyelo + band forms 0.2% |                                  |                                      |                             |

RBC = Red blood corpuscles; TLC = Total leukocyte count; DLC = Differential leukocyte count.

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**Figure 3.** Peripheral smear MPO Positive (Myeloperoxidase) myeloid cells.

**Figure 4.** NSE Positive (Leukocyte nonspecific esterase) monocytoid cells.
Figure 5. Flow cytometry (Positive for CD34, CD45, CD13, CD33, CD14, CD4, Anti-HLA DR, CD11 and anti MPO).
forms of leukaemia than in chronic forms. Oral manifestations in patient with leukaemia have been described in all subtypes of AML, chronic myeloid leukaemia (CML), acute lymphocytic leukaemia (ALL), and chronic lymphocytic leukaemia (CLL). Gingival infiltration represents a 5% frequency as the initial presenting complication of AML. Gingival infiltration of leukemic cells is most commonly seen in acute monocytic leukaemia (M5) and acute myelomonocytic leukaemia (M4). The proposed hypothesis for gingival involvement is considered due to its microanatomy and expression of endothelial adhesion molecules which enhances infiltration of leukocytes [7]. Dreizen et al. [8] in an observational study have reported gingival involvement in 66.7% of 1076 cases of AML M5 cases and 18.5% in M4.

Leukaemia cell gingival infiltrate is not observed in edentulous individuals, suggesting that local irritation and trauma associated with the presence of teeth may play a role in the pathogenesis of this abnormality [6,7]. The case described here had adequate local factors which are the predisposing factor for gingival enlargement. The specific type of leukaemia must be investigated in order to provide the best treatment and most accurate prognosis. Special leukaemia stains help to distinguish one cell type from another. Commonly used special staining methods are sudan black, periodic acid-schiff, terminal deoxynucleotidyl transferase stain, leukocyte alkaline phosphatase, tartrate-resistant acid phosphatase stain, myeloperoxidase (MPO), leukocyte specific and nonspecific esterase [9,10]. In our case myeloperoxidase and leukocyte nonspecific esterase (NSE) special stain were used which showed presence of myeloblasts in the peripheral smear suggestive of acute myelocytic leukaemia.

Flow cytometry works on the principle that when a LASER beam is passed through a particle like a cell in a hydrodynamically focused stream, the light will cause scattering and this fluorescent light is picked by an electronic detector which analyze and determines the physical and chemical nature of the particle or cell. The data so generated by flow cytometry are plotted in single or two dimension dot plots depending on the fluorescence intensity. The plot can be sequentially separated creating a series of subset extractions called “Gates” [10]. In our case we have used flow cytometry for the determination of diagnosis, prognosis and progression of the disease. The report of flow cytometry has confirmed the present case to be a case of AML M4 as the cells were strongly positive for CD 45, CD13, CD14 and anti HLADR.

CONCLUSIONS

The mechanisms underlying AML and the reasons for difficulties of treating patients with acute myeloblastic leukaemia have only partly been unravelled. This case reminds that dentist and physicians should be aware of the importance of recognizing oral manifestations of systemic diseases as oral cavity is considered to be the mirror of systemic health. A patient seeking dental treatment for bleeding gums was incidentally diagnosed as leukaemia based on thorough dental and medical history followed by definitive diagnosis by peripheral staining and flow cytometry as presented in this case. Therefore, the dentist may be the first person to diagnose such cases. The timely referral by the general dentist for a suspicious lesion provided an early diagnosis and early intervention reducing the patient morbidity. Therefore the role of dental practitioner in diagnosing a case of leukaemia for better management and prognosis cannot be overemphasized.

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