Stem Cells and Cloning: Advances and Applications

Autologous bone marrow cell therapy for peripheral arterial disease

C Botti
C Maione
A Coppola
V Sica
G Cobellis

Department of General Pathology, Second University of Naples, Naples, Italy

Correspondence: Gilda Cobellis
Department of General Pathology, Second University of Naples, Via Luigi De Crecchio 7, 80138 Napoli, Italy
Tel +39 081 566 5681
Fax +39 081 292 399
Email g.cobellis@unina2.it

Abstract: Inadequate blood supply to tissues caused by obstruction of arterioles and/or capillaries results in ischemic injuries – these injuries can range from mild (eg, leg ischemia) to severe conditions (eg, myocardial infarction, stroke). Surgical and/or endovascular procedures provide cutting-edge treatment for patients with vascular disorders; however, a high percentage of patients are currently not treatable, owing to high operative risk or unfavorable vascular involvement. Therapeutic angiogenesis has recently emerged as a promising new therapy, promoting the formation of new blood vessels by the introduction of bone marrow–derived stem and progenitor cells. These cells participate in the development of new blood vessels, the enlargement of existing blood vessels, and sprouting new capillaries from existing blood vessels, providing evidence of the therapeutic utility of these cells in ischemic tissues. In this review, the authors describe peripheral arterial disease, an ischemic condition affecting the lower extremities, summarizing different aspects of vascular regeneration and discussing which and how stem cells restore the blood flow. The authors also present an overview of encouraging results from early-phase clinical trials using stem cells to treat peripheral arterial disease. The authors believe that additional research initiatives should be undertaken to better identify the nature of stem cells and that an intensive cooperation between laboratory and clinical investigators is needed to optimize the design of cell therapy trials and to maximize their scientific rigor. Only this will allow the results of these investigations to develop best clinical practices. Additionally, although a number of stem cell therapies exist, many treatments are performed outside international and national regulations and many clinical trials have been not registered on databases such as ClinicalTrials.gov or EudraCT. Therefore, more rigorous clinical trials are required to confirm the first hopeful results and to address the challenging issues.

Keywords: adult stem cells, critical limb ischemia, bone marrow transplantation, therapeutic angiogenesis

What is peripheral arterial disease?

Peripheral arterial disease (PAD) is a common circulatory problem in which narrowed arteries reduce blood flow to the limbs, especially the legs. The most common causes of PAD are atherosclerosis obliterans (ASO) and thromboangiitis obliterans (TAO).1

Two major classification systems are currently used to evaluate the spectrum of symptoms: (1) the Fontaine classification, not used in everyday clinical practice but useful for research purposes, and (2) the Rutherford classification, more commonly cited in recent publications in the field of vascular medicine (Table 1). The American College of Cardiology/American Heart Association 2005 guidelines noted the usefulness of the Rutherford classification for standardized communication between clinicians.1
### Table 1 Two classifications of peripheral arterial disease (PAD): Fontaine and Rutherford

| PAD | Pathophysiology | Fontaine | Rutherford |
|-----|-----------------|----------|------------|
|     |                 | Stage    | Clinical   | Grade | Category | Clinical |
| Fortuitous discovery of aortic and iliac calcifications | ATS plaques | I | Asymptomatic | 0 | 0 | Asymptomatic |
|     | Plaques at risk (inflammation of ATS plaques) | | | | | |
|     | Atherothrombosis | | | | | |
| ACD > 200 m | Discrepancy between oxygen demand and arterial supply | IIa | Intermittent claudication | I | 1 | Intermittent claudication |
| Recovery time < 2 minutes | Higher discrepancy between oxygen demand and arterial supply | IIb | Moderate or severe claudication | I | 2 | Moderate claudication |
| ACD ≤ 200 m | Higher discrepancy between oxygen demand and arterial supply | I | Severe claudication | I | 3 | Severe claudication |
| Recovery time > 2 minutes | Acidosis | | | | | |
| Ischemic rest pain | Severe skin hypoxia and acidosis | III | Ischemic rest pain | II | 4 | Ischemic rest pain |
| Necrosis | Severe skin hypoxia and acidosis | IV | Critical limb ischemia | III | 5 | Critical limb ischemia |
| Gangrene | Severe skin hypoxia and acidosis | | | | | |
|     | Infection | | | | | |
|     | Tissue loss and gangrene | | | | | |

Abbreviations: ACD, absolute claudication distance; ATS, atherosclerotic.

Disease staging and classification systems are important for clinical management of these patients. Based on the severity of symptoms, usually two distinct clinical presentations are distinguished in PAD patients: (1) intermittent claudication, characterized by intermittent pain in leg muscles when the person walks, and (2) critical limb ischemia (CLI), a more severe form of PAD, characterized by pain at rest, nonhealing wounds, and gangrene. After 1 year, 30% of patients with CLI will lose their leg and 25% will die.2

The incidence of CLI in Western societies is approximately 220 new cases per million people per year, and, with an aging population, the population at risk is expected to increase because of persistent rates of tobacco abuse and an increase in diabetes.2 Fifty percent of diabetics (7% of the world population in 2010) suffer from PAD, which may lead to amputation due to CLI.3 Moreover, smoking, hypertension, dyslipidemia, a sedentary lifestyle, and a genetic predisposition all contribute to the development of PAD.4,5

### Current treatments for PAD

Revascularization, either surgical or endovascular, is the gold standard treatment for patients with severe PAD. However, despite advances in surgical and endovascular techniques,6 more than 30% of patients do not qualify as candidates for revascularization because of excessive operative risk or adverse vascular involvement. Furthermore, the presence of extensive atherosclerotic plaques in the tibial and/or peroneal arteries renders revascularization unsuccessful. These patients are left to medical therapy, which may only slow disease progression, and the only remaining alternative for relief of rest pain or gangrene is amputation of the affected leg.

An estimated 120–500 amputations are performed per million people per year, and one-quarter of these patients require long-term institutional care or professional assistance at home.2 Medical therapy is limited to antithrombotic therapy,7 the prostaglandin analogue iloprost,8 or recently to cilostazol. Cilostazol has been found to be effective for the treatment of intermittent claudication. This compound has several beneficial effects on platelet aggregation, serum lipids, and endothelial cells (ECs), but how these might relate to improvements in walking is not entirely understood.9

Thus, there is a critical need to develop novel strategies to promote neovascularization in patients with CLI who are not candidates for conventional treatments.

In 1997, Asahara et al10 made a big step forward when they identified a class of bone marrow–derived circulating endothelial progenitor cells (EPCs) that contribute to angiogenesis and/or vasculogenesis in ischemic tissues.11 Since then, several studies have reported the capability of stem and progenitor cells to promote neovascularization, reducing ischemic damages.12,13 Encouraging results of several clinical studies have rapidly demonstrated the beneficial effects of autologous stem cell transplantation in patients affected by CLI. Clinical improvements were observed in objective and subjective measurements of perfusion (ie, transcutaneous oxygen tension [TcPO2] and laser Doppler flowmetry [LDF]), pain reduction, increased pain-free total walking distance, and decreased rate of amputation.3,14–69
What is vascular regeneration?

Vascular regeneration involves the restoration of normal vascular function and structure and the growth of new blood vessels. This includes a plethora of processes, such as the distribution of blood flow via the formation of collateral networks; the response of newly generated vessels to hemodynamic, humoral, and local tissue factors; the modulation of the immune response and the trafficking of circulating cells; and the permeation of nutrients and macromolecules through the microvasculature, which can in turn have trophic effects on blood fluidity and hemostasis.9 Vascular regeneration is also important in a variety of processes: during embryonic organogenesis and organ growth in born individuals, in the course of restoration of blood supply to ischemic tissues, and in the establishment of blood supply to tumours.70

Neovascularization involves the growth of new structures from preexisting vessels by migration, proliferation, and differentiation of progenitor cells and the interplay between growth factors and cytokines. The process of neovascularization comprises three distinct phenomena: (1) vasculogenesis, (2) angiogenesis, and (3) arteriogenesis.70

The essential mechanism responsible for new blood vessel formation in adults is based on neoangiogenesis. During angiogenesis, ECs present in vessel walls are activated in response to various stimuli and begin to release various growth factors and cytokines. The process of neovascularization comprises three distinct phenomena: (1) vasculogenesis, (2) angiogenesis, and (3) arteriogenesis.70

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What are the features of these cells?

MSCs are a subset of cells that express on their surface specific molecules such as CD73, CD90, and CD105; MSCs also express CD54/CD102, CD166, and CD49 (alpha integrin), which regulate cell-to-cell interactions, and they do not express any hematopoietic and/or EPC surface markers.76 MSCs can be found in many fetal and adult tissues and are generally isolated from bone marrow, adipose tissue, umbilical cord blood, and compact bone. Furthermore, MSCs are able to migrate to and home to injured sites, where they act by differentiating into specific cells and by secreting trophic factors, which activate paracrine signaling.77 Moreover, these cells interact with the immune system, particularly modulating the immune response, apparently by inhibiting tumor necrosis factor-alpha (TNFa) and interferon-gamma (IFN-γ) and by increasing interleukin 10 (IL-10).79 This unique immunomodulatory property makes these cells suitable for both autologous and heterologous transplants, since they avoid and/or actively suppress eventual rejection of transplants.79

MSCs display a great therapeutic potential because of their capability to differentiate into muscle, neural precursors, cardiomyocytes, and perivascular cells. Perivascular cells (herein referred to as pericytes) are critical cells in vascular biology. Pericytes typically express alpha smooth muscle actin (α-SMA), platelet-derived growth factor receptor beta
(PDGF-β), and nerve/glial antigen-2 (NG-2) proteoglycan. They are branched cells embedded within the basement membrane of capillaries and postcapillary venules, stabilizing the vessel wall. Pericytes are considered cells that control EC proliferation and migration, and thereby also the growth of new capillaries. In turn, ECs stimulate expansion and activation of the pericyte precursor cell population. The balance between ECs and pericytes is highly controlled by a series of signaling pathway mechanisms operating in an autocrine and/or a paracrine manner. In pathological conditions in which angiogenic activity is impaired, pericytes and ECs could be partly responsible for abnormalities in blood vessels.

EPCs are adult hemangioblast-derived cells characterized by the expression of CD34, VEGF receptor 2, and CD133. These markers are expressed by precursor cells, but not by differentiated ones. In fact, as the hemangioblasts differentiate to become ECs, they downregulate the hematopoietic stem cell marker CD133 expression. EPCs can be isolated from human peripheral or umbilical cord blood and can also be found in bone marrow niches. EPCs have shown in vitro all the functional properties of ECs; moreover, EPCs have direct angiogenic action, supporting angiogenesis through their ability to secrete paracrine mediators. In this respect, several studies have shown that these cells release interleukins, growth factors, and chemokines that altogether regulate CD14-positive cells, accelerate vascular network formation, and enhance healing processes.

Adult stem cells with angiogenic potential such as EPCs and MSCs will stimulate the production of new blood vessels, as shown in Figure 1.

**Cell therapy in PAD: clinical results**

Promotion of collateral vessel formation and angiogenesis in PAD patients is an important therapeutic strategy to minimize tissue injury associated with severe ischemia. The Therapeutic Angiogenesis using Cell Transplantation trial was the first report on the use of bone marrow–derived mononuclear cells in the treatment of PAD. Starting from this, the search of the literature yielded a total of 57 early-phase clinical trials for a total of 1997 enrolled patients. The safety and feasibility of autologous cell transplantation has been reported in 1667 treated patients (Tables 2 and 3). Among these, a total of 303 diabetic patients with CLI and foot ulcers underwent cell therapy. The degree of ischemia varied throughout the groups, ranging from Rutherford category 4/Fontaine stage III through to severe CLI classified as Rutherford category 6/Fontaine stage IV.

Only a minority of trials (n = 13) included appropriate controls (Table 4). In these studies, the follow-up of the untreated or placebo group did not differ from that observed

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**Figure 1** Schematic representation of neoangiogenesis promoted by circulating and bone marrow–resident stem cells.

**Notes:** Ischemia induces production of growth factors, cytokines, and hormones, which promotes proliferation, differentiation, and mobilization of mesenchymal stem cells (MSCs) and endothelial progenitor cells (EPCs) to form new vessels. In addition, the growth factors can stimulate EPCs sprouting from preexisting blood vessels.

**Abbreviations:** G-CSF, granulocyte colony-stimulating factor; IL, interleukin; NO, nitric oxide; VEGF, vascular endothelial growth factor.
Table 2  Clinical trials with cell therapy in peripheral arterial disease (PAD)

| Published study | Delivery route | Condition (n) | Patients (n) | Cell type | Follow-up time | Improved functional outcomes |
|-----------------|----------------|---------------|--------------|-----------|----------------|-----------------------------|
| Tateishi-Yuyama et al14 | IM | PAD, DM | 45 | BMCs or PB-MNCs | 4 and 24 weeks | ABI, TcPO2, rest pain, pain-free walking time |
| Esato et al15 | IA | PAD | 8 | BMSCs | N/D | Ulceration healing |
| Saigawa et al16 | IM | PAD, DM | 8 | BMSCs | 4 weeks | ABI, TcPO2 |
| Higashi et al17 | IM | PAD | 7 | BMSCs or BM-MNCs | 4 and 24 weeks | ABI, TcPO2, pain-free walking time |
| Miyamoto et al18 | IM | CLI | 12 | BMSCs and EPCs | N/D | ABI, pain-free walking time |
| Huang et al19 | IM | PAD | 5 | PB-MNCs | 3 months | ABI, LDF |
| Kawamura et al20 | IM | PAD, CLI | 30 | PB-MNCs | N/D | T°C |
| Lenk et al21 | IA | CLI | 7 | PB-MNCs | 20 weeks | ABI, TcPO2, rest pain, pain-free walking time |
| Huang et al22 | IM | CLI, DM | 28 | MPB-MNCs | 3 months | ABI, pain, ulcers |
| Ishida et al23 | IM | PAD | 6 | MPB-MNCs | 4 and 24 weeks | ABI, ulcers |
| Durdu et al24 | IM | PAD | 28 | BM-MNCs | 3 and 6 months | ABI, rest pain, pain-free walking time |
| Koshikawa et al25 | IM | PAD | 7 | BM-MNCs | 6 months | ABI, pain, ulcers |
| Ara et al26 | IM | PAD | 25 | BMSCs | 1 month | ABI, TcPO2 |
| Miyamoto et al27 | IM | PAD, CLI | 8 | BM-MNCs | 4 weeks, 4 and 7 months, and 1 year | Rest pain, ulcers |
| Kawamura et al28 | IM | CLI | 92 | BM-MNCs | 6 weeks | Limb salvage, VEGF serum level |
| Bartsch et al29 | IM and IA | PAD, CLI | 13 | BMSCs | 2 and 13 months | ABI, pain-free walking distance |
| Huang et al30 | IM | PAD | 150 | BM-MNCs or MPB-MNCs | 12 weeks | ABI, rest pain |
| Kajiguchi et al31 | IM | CLI | 7 | BM-MNCs or PB-MNCs | 1 month | ABI |
| Hernández et al32 | IM | CLI, DM | 12 | BM-MNCs | 24 months | ABI, SaO2, pain-free walking time, rest pain |
| Saito et al33 | IM | PAD | 7 | BM-MNCs | N/D | TcPO2 |
| Matoba et al34 | IM | PAD | 115 | BM-MNCs | 3 years | Pain, ulcers, pain-free walking distance |
| Napoli et al35 | IA | PAD | 18 | BM-MNCs | 3, 6, 12, and 18 months | ABI, ulcers, pain-free walking distance |
| Gu et al36 | IM or IA | PAD, DM | 32 | BM-MNCs | 4 weeks | ABI, TcPO2, limb salvage |
| Choehola et al37 | IA | PAD, DM | 24 | BM-MNCs | 1 year | Limb salvage, wound healing |
| Wester et al38 | IM | CLI | 8 | BMSCs | 4 and 8 months | ABI, TcPO2, ulcers |
| Van Tongeren et al39 | IM and IA | CLI | 27 | BMSCs | 6 and 12 months | Pain-free walking distance, ABI, pain reduction |
| De Vriese et al40 | IM | CLI | 16 | BM-MNCs | 12 weeks | TcPO2, pain reduction |
| Cobellis et al41 | IA | PAD | 10 | BM-MNCs | 12 months | ABI, pain-free walking distance |
| Motukuru et al42 | IM | PAD | 38 | BM-MNCs | 6 months | Ulcer healing, limb salvage, ABI, TcPO2 |
| Amann et al43 | IM | CLI | 45 | BM-MNCs | 3 months | Limb salvage, ABI, TcPO2, pain-free walking distance |
| Amann et al44 | IM | PAD | 51 | BM-MNCs | 6 months | Limb salvage, ABI, TcPO2, pain-free walking distance |
| Capiod et al45 | IM | CLI | 24 | BM-MNCs or PB-MNCs | N/D | No clinical evaluation reported |
| Franz et al46 | IM and IA | PAD | 9 | BM-MNCs | 2 weeks and 3 months | ABI, pain reduction, ulcers, limb salvage |
| Franz et al47 | IM and IA | PAD | 20 | BM-MNCs | 3 months | ABI, pain reduction, ulcers, limb salvage |
| Zafarghandi et al48 | IM | CLI | 50 | BM-MNCs and G-CSF | 4 and 24 weeks | ABI, pain-free walking distance |
| Procházka et al49 | IM | CLI, DF | 37 | BMSCs | 3 months | LDF, ABI, TcPO2, limb salvage |
| Procházka et al50 | IM | PAD | 96 | BMSCs | 4 months | ABI, limb salvage |
| Lara-Hernández et al51 | IM | CLI | 28 | Mobilized EPCs | 14 and 18 months and 1 year | ABI, limb salvage |
| Iso et al52 | IM | CLI | 13 | BM-MSCs | 4 months | TcPO2, rest pain |

(Continued)
in several large population studies.²,³,³⁻⁷ In addition, the Edinburgh Artery Study defined the prevalence of asymptomatic and symptomatic PAD and related comorbidities in the general population.⁵⁷ Therefore, since the progression of disease is well defined in CLI patients, the lack of an untreated or a placebo group – even if scientifically compelling – cannot diminish the significance of the studies.

Two sources of cells were used in these trials: (1) bone marrow aspiration (n = 46) and (2) apheresis of peripheral blood with or without GSF stimulation (n = 11). The route of cell administration was intramuscular in 39 trials,

### Table 2 (Continued)

| Published study          | Delivery route | Condition | Patients (n) | Cell type     | Follow-up time | Improved functional outcomes                                                                 |
|--------------------------|---------------|-----------|--------------|---------------|----------------|---------------------------------------------------------------------------------------------|
| Sprengers et al⁵²         | IA            | CLI       | 120          | BM-MNCs       | 6 months       | Limb salvage, ulcers, rest pain, ABI, TcPO₂                                               |
| Murphy et al⁵¹            | IM            | CLI       | 29           | BM-MNCs       | 1 year         | First-toe pressure and toe-brachial index increase, perfusion index by computed tomography, rest pain |
| Walter et al⁵⁴            | IA            | CLI       | 40           | BM-MNCs       | 3 months       | ABI, ulcer healing, rest pain                                                               |
| Iafrati et al⁵⁵           | IM            | CLI, DM   | 48           | BMSCs         | 12 weeks       | Limb salvage, pain, ABI, Rutherford classification, quality of life                         |
| Idei et al⁵⁶              | IM            | CLI       | 97           | BM-MNCs       | 56 months      | ABI, TcPO₂, pain, amputation-free survival rate                                             |
| Ruiz-Salmeron et al⁵⁷     | IA            | CLI       | 20           | BM-MNCs       | 3 and 12 months| Rutherford classification, diabetic wound scales, ABI, mortality rate                        |
| Lu et al⁵⁸                | IM            | CLI, DM   | 82           | BM-MNCs, BMSCs| 6 and 24 weeks | Pain-free walking time, ABI, TcPO₂, ulcers, limb salvage                                    |
| Gabr et al⁵⁹              | IM            | CLI       | 20           | BM-MNCs       | 3 months       | Walking distance, rest pain, skin condition, ABI                                           |
| Benoit et al⁶⁰            | IM            | CLI       | 48           | BMSCs         | 6 months       | Limb salvage, Rutherford classification, ABI, pain                                          |
| Powell et al⁶¹            | IM            | CLI       | 86           | BMSCs         | 6 and 12 months| Limb salvage, TTF, wound healing                                                             |
| Perin et al⁶²              | IM            | CLI       | 21           | BM-MNCs       | 6 and 12 weeks | Rutherford classification, ABI, TcPO₂, quality of life, pain                                |
| Smadja et al⁶³            | IM            | CLI       | 11           | BM-MNCs       | 6 and 12 months| TcPO₂, wound healing, limb salvage                                                         |
| Powell et al⁶⁴            | IM            | CLI       | 77           | Multicellular therapy | 12 months | TTF, limb salvage                                                                            |
| Klenyan et al⁶⁵           | IM or IA      | CLI       | 41           | BMSCs         | 6 months       | Rutherford classification, ABI, TcPO₂, pain, limb salvage, wound healing                    |
| Schiavetta et al⁶⁶        | IA            | CLI, DM   | 60           | BM-MNCs       | 12 months      | LDF, TcPO₂, limb salvage                                                                    |

**Abbreviations:** ABI, ankle-brachial index; BMSCs, bone marrow stem cells; BM-MNCs, bone marrow–derived mononuclear cells; CLI, critical limb ischemia; DF, diabetic foot; DM, diabetes mellitus; EPCs, endothelial progenitor cells; G-CSF, granulocyte colony-stimulating factor; IA, intra-arterial injection; IM, intramuscular injection; LDF, laser Doppler flowmetry; MPB-MNCs, mobilized peripheral blood–derived mononuclear cells; N/D, not determined; PB-MNCs, peripheral blood–derived mononuclear cells; SaO₂, arterial oxygen saturation; T°C, temperature expressed in degrees Celsius; TcPO₂, transcutaneous oxygen tension; TTF, time to first occurrence of treatment failure; VEGF, vascular endothelial growth factor.

### Table 3 Clinical trials with intralesional administration of stem cells in foot ulcers

| Published study          | Condition   | Patients (n) | Cell type | Follow-up time | Improved functional outcomes                                                                 |
|--------------------------|-------------|--------------|-----------|----------------|---------------------------------------------------------------------------------------------|
| Vojtassák et al⁶⁷         | DF          | 1            | BMSCs     | 29 days        | Wound size and ulcer healing                                                                  |
| Dash et al⁶⁸              | DF ulcers, PAD | 24          | BMSCs     | 3 months       | Wound size and pain-free walking distance                                                     |
| Subrammaniyan et al⁶⁹     | CLI         | 6            | BM-MNCs   | 6 months       | ABI and pain-free walking distance, limb salvage, ulcer healing, and rest pain               |

**Abbreviations:** ABI, ankle-brachial index; BM-MNCs, bone marrow–derived mononuclear cells; BMSCs, bone marrow stem cells; CLI, critical limb ischemia; DF, diabetic foot; PAD, peripheral arterial disease.
intramuscular in nine trials, and combined intra-arterial plus intramuscular in four trials (Table 2). Furthermore, two studies compared the therapeutic effects of intramuscular or intra-arterial delivery of bone marrow cells in patients with lower limb ischemia, showing similar beneficial results.64

To prevent clot formation, harvested cells were collected in the presence of anticoagulant.3,14-69 Intramuscular administration is usually performed through multiple injections at the level of limb muscles, while intra-arterial infusion is usually performed via classic femoral access. Three studies reported the use of intralesional administration of bone marrow–derived stem cells in 31 diabetic patients with foot ulcers, showing encouraging results (Table 3).

In general, bone marrow aspiration was well tolerated, and the most frequent adverse reaction was local pain or mild anemia. However, serious adverse reactions such as angina with ST segment depression were observed in a small number of patients.65

The average follow-up of these clinical studies was 8.4 ± 9.55 months. Considering all studies, the reported outcomes for therapeutic efficacy of cell therapy involved the ankle-brachial index, TcPO₂, LDF, pain-free walking distance, ulcer healing, and amputation-free survival. In all studies, symptoms improved after the procedure, as evidenced by clinical evaluation, relief of rest pain, and improvement by at least one level in Rutherford and Fontaine classifications. Furthermore, autologous cell therapy promoted amputation-free survival with an average of 7.8 months and promoted complete wound healing within 3 months in most patients with ulcers prior to bone marrow stem cell transplantation, in comparison with the natural history of PAD patients. Therefore, autologous transplantation of bone marrow–derived cells significantly improved both the objective and the subjective endpoints.

**Conclusion**

Herein, the authors provide the most comprehensive review of cell therapy trials describing the background and first results of stem and progenitor cell therapy in patients with CLI who are not suitable for revascularization. Both the principle, as far as it is understood, and the methods are described. Compelling evidence suggests that stem cell therapy may become a useful adjunct to the current treatment options. Because of poor prognosis and the increasing number of patients, there is a need for new therapeutic methods.

About 1997 patients without revascularization options were enrolled in these trials and 1667 patients were treated. Cell therapy significantly improved functional outcomes such as ankle-brachial index, TcPO₂ or LDF, rest pain, pain-free walking distance, ulcer healing, and limb salvage. Although it is generally agreed that controlled trials yield more reliable results, the authors also included noncontrolled studies, which are the majority of published reports. The authors believe the main reason for this majority is that the authorized studies have chosen to treat end-stage patients, without other therapeutic options. The procedures are generally safe and well tolerated. Reported deaths were expected, given the severe underlying disease, and could not be directly attributed to cell therapy.

Challenges in this new therapeutic option still include open questions regarding cell number, phenotype, processing, route of optimal delivery, and frequency of application. The number of injected cells ranged from 4 × 10⁶ to 10⁷ for bone marrow cells and from 7 × 10⁶ to 3 × 10⁷ for peripheral blood–derived mononuclear cells, with positive effects on blood perfusion, even when low cells were used. Nevertheless, no correlation study between clinical response and cell number has been performed so far, and no proven correlation exists between the phenotype of used cells and efficacy of neoangiogenesis. Answering these two points is critical to understanding which and how many cells are needed to obtain a clinical response.

The question of optimal delivery route remains open. The rationale behind the intramuscular injection is to generate a reservoir of cells near the ischemic area, which can be recruited by active paracrine mechanisms. The intra-arterial injection relies on the fact that the blood flow transports cells up to the ischemia site; however, it is not known how many cells are able to leave the blood stream to reach the ischemic area. Again, no correlation

**Table 4 Controlled clinical trials with cell therapy in peripheral arterial disease**

| Published study | Year | Patients Total (N) | Treated (n) | Control (n) |
|-----------------|------|-------------------|-------------|-------------|
| Powell et al84  | 2012 | 72                | 48          | 24          |
| Benoit et al60  | 2011 | 48                | 34          | 14          |
| Lu et al48      | 2011 | 82                | 41          | 41          |
| Powell et al51  | 2011 | 46                | 32          | 14          |
| Idei et al54    | 2011 | 97                | 51          | 46          |
| Iafra et al55   | 2011 | 48                | 34          | 14          |
| Walter et al54  | 2011 | 40                | 19          | 21          |
| Procházka et al3 | 2010 | 96                | 42          | 54          |
| Sprengers et al52 | 2010 | 110               | 55          | 55          |
| Cobellis et al53 | 2008 | 19                | 10          | 9           |
| Bartsch et al59 | 2007 | 25                | 13          | 12          |
| Arai et al10     | 2006 | 25                | 13          | 12          |
| Huang et al52   | 2005 | 28                | 14          | 14          |
study between the two routes of administration has been performed, although the present trend is for intramuscular administration. It has been reported that the combination of both routes (intramuscular plus intra-arterial) has given substantial improvements in clinical outcomes, but this must be confirmed in exhaustive experiments in preclinical models.

In summary, over the past 10 years there has been considerable interest in stem cells, including extensive clinical activity involving stem cells. Unfortunately, the rationale for the clinical application of adult stem cells, particularly in regenerative medicine, has lagged behind initial laboratory observations. At this point, the authors believe that additional research initiatives should be undertaken to better identify the nature of stem cells and that an intensive cooperation between laboratory and clinical investigators is needed to optimize the design of cell therapy trials and to maximize their scientific rigor. Only this will allow the results of these investigations to develop best clinical practices.

Additionally, although a number of stem cell therapies exist, many treatments are performed outside international and national regulations and many clinical trials have been not registered on databases such as ClinicalTrials.gov or EudraCT. Therefore, more rigorous clinical trials are required to confirm the first hopeful results and to address the challenging issues.

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