Graphene-Encapsulated Silver Nanoparticles for Plasmonic Vapor Sensing

Gábor Piszter, György Molnár, András Pálinkás and Zoltán Osváth*

Centre for Energy Research, Institute of Technical Physics and Materials Science, 1121 Budapest, Hungary; gabor.piszter@ek-cer.hu (G.P.); gyorgy.molnar@ek-cer.hu (G.M.); andras.palinkas@ek-cer.hu (A.P.)
* Correspondence: zoltan.osvath@ek-cer.hu

Abstract: Graphene-covered silver nanoparticles were prepared directly on highly oriented pyrolytic graphite substrates and characterized by atomic force microscopy. UV–Vis reflectance spectroscopy was used to measure the shift in the local surface plasmon resonance (LSPR) upon exposure to acetone, ethanol, 2-propanol, toluene, and water vapor. The optical responses were found to be substance-specific, as also demonstrated by principal component analysis. Point defects were introduced in the structure of the graphene overlayer by O₂ plasma. The LSPR was affected by the plasma treatment, but it was completely recovered using subsequent annealing. It was found that the presence of defects increased the response for toluene and water while decreasing it for acetone.

Keywords: silver nanoparticles; graphene coating; LSPR-based sensing; VOC detection

1. Introduction

Noble metal nanoparticles (NPs) are widely used for chemical and biological sensing because of their local surface plasmon resonance (LSPR) [1,2] and surface-enhanced Raman scattering (SERS) [3,4] properties. The LSPR produces sharp spectral absorption, which can be used to detect changes in the molecular environment near the surfaces of NPs by spectral shift detection [5–7]. Due to their enhanced interaction with light, gold (Au) and silver (Ag) nanoparticles have been intensively studied as promising plasmonic sensing systems [8–14]. Their LSPR can be tuned by adjusting the size, shape, dispersion, and uniformity of the NPs, as well as the dielectric constant of the detection medium [15–23]. Ag nanostructures are often considered the best material for plasmonics, as there are no interband absorptions and only minimum loss at optical frequencies [24]. They have an inherently more intensive and sharper plasmonic spectrum than Au nanoparticles, enabling them to offer superior sensing functionality. Silver, however, has poor stability under ambient conditions and forms silver sulfide on its surface. This causes morphological changes in Ag NPs and a significant decrease in the optical properties [25]. Conserving the high LSPR intensity is critically important in potential sensor applications. A common method to protect Ag NPs from environmental effects is to form core–shell structures where the Ag surface is passivated with either organic or inorganic shells (for recent reviews, see Refs. [26,27]).

Graphene, as an atomically thin material with very low permeability [28–30], seems to be an ideal passivating coating to prevent the surface oxidation of Ag films and NPs [25,31,32]. It was demonstrated that graphene can improve the sensitivity of silver-based LSPR sensors and delay the oxidation process of Ag NPs effectively [33–36]. Nevertheless, it was also shown that oxygen can penetrate through the defects and grain boundaries of large-area graphene grown by chemical vapor deposition (CVD) [37].

The synthesis of Ag NPs can be accomplished using a variety of strategies [38] depending on the desired shape of the nanoparticles. Chemical reduction in a bottom-up approach is the most used method. Many factors influence the size and shape of the produced Ag NPs, including temperature, silver precursor concentration, the strength of the chemical...
interaction between the capping agent and the different silver crystallographic planes [39], and so on. In this work, we fabricate Ag NPs and graphene−silver nanoparticle hybrids directly on highly oriented pyrolytic graphite (HOPG) substrates using a straightforward approach described earlier [40]. Scanning electron microscopy (SEM), atomic force microscopy (AFM), and UV–Vis reflectance spectroscopy are used, respectively, to investigate the morphology, optical, and vapor sensing properties of the hybrid nanomaterial. We show that graphene-covered Ag NP samples display a pronounced optical response upon exposure to organic vapors (acetone, ethanol, 2-propanol, toluene, or water), which can be observed in the shift in the LSPR. This optical response to the volatile organic compounds (VOC) is proportional to the vapor concentration and is substance-specific. Furthermore, we show that the optical response can be tuned by introducing point defects into the structure of the graphene overlayer.

2. Experimental

2.1. Preparation of Graphene-Covered Silver Nanoparticles

Silver films of 7 nm nominal thickness were evaporated onto HOPG substrates at a background pressure of $5 \times 10^{-7}$ mbar and an evaporation rate of 0.1 nm s$^{-1}$. The deposited Ag films were covered with CVD graphene grown on copper foil (Graphenea), immediately after the opening of the evaporation chamber. We applied polymer tape as mechanical support to transfer large-area graphene samples. The copper foil was removed using an etchant composed of copper (II) chloride aqueous solution (20%) and hydrochloric acid (37%) in a 4:1 volume ratio. After rinsing and drying, the polymer tape with the graphene was pressed onto the evaporated Ag films. The tape was removed by lifting it with tweezers. To form nanoparticlest, both bare and graphene-covered Ag thin films were annealed at 400 °C under an inert gas (Ar) atmosphere for 1.5 h. The obtained samples (samples #1 and #2) were characterized by tapping-mode AFM on a MultiMode 8 system (Bruker FRANCE S.A.S., Strasbourg, Germany), as well as SEM using a Thermo Scientific Scios2 (Brno, Czech Republic) instrument. Point defects were introduced in the atomic structure of the graphene overlayer (sample #2) by 5 s of O$_2$ plasma treatment (Zepto cleaner, Diener electronic, Ebhausen, Germany) at 50 W, followed by annealing (400 °C) in Ar gas for 60 min.

2.2. UV–Vis Spectroscopy and Vapor Sensing Setup

Optical reflectance and vapor sensing measurements were carried out by mounting the samples in an airtight aluminum box covered with a fused silica window to provide UV transmission. The samples were illuminated using an Avantes AvaLight DH-S-BAL light source. The initial reflectance of the samples in synthetic air was measured with an Avantes HS 1024 × 122TEC spectrometer (Avantes BV, Apeldoorn, The Netherlands) by capturing the specularly reflected signal (measured under 15°). For vapor sensing experiments, five types of volatile vapors were passed through the cell’s gas inlet and vented through the outlet: acetone, ethanol, 2-propanol, toluene, and water (analytical-grade, VWR International Ltd., Radnor, PA, USA). Vapor concentration was adjusted by switching digital mass flow controllers (Aalborg DFC, Aalborg Instruments & Controls, Inc., Orangeburg, NY, USA) to pass synthetic air (Messer, 80% N$_2$, 20% O$_2$) and saturated volatile vapors through gas bubblers in the required ratio. During the measurements, a constant gas flow of 1000 mL/min was sustained through the cell. Vapor detection experiments were performed by varying the concentration and type of test vapor while tracking spectral variations over time. To purge the cell, a 20 s vapor flow at a given concentration was followed by a 60 s flow of synthetic air. In addition, before introducing the next vapor type, a 120 s purge was used to recover the initial reflectance of the sample.

3. Results and Discussion

Tapping-mode AFM and SEM were used to characterize the formed nanoparticles. The graphene-covered Ag nanostructures had elongated shapes (Figure 1a,b), with an average
height of 61 nm. Typical nanoparticles that are completely sandwiched between graphene and HOPG are shown in Figure 1c. Graphene wrinkles also developed near these NPs. In contrast, smaller NPs formed in areas with graphene discontinuities. These bare NPs had a rather disk-like shape, with an average height of 45 nm. The mean height distribution of both graphene-covered and bare Ag NPs is shown in Figure 1d.

The optical properties of such nanoparticle ensembles were studied by UV–Vis reflectance spectroscopy. For this, the samples were fixed inside an aluminum cell with a quartz window, as illustrated in Figure 2a, and a synthetic air atmosphere. Specular reflection was applied at 15° degrees between the two optical fibers used for illumination and light collection. Typical reflectance spectra are shown in Figure 2b, where the LSPR is observed as prominent minima near 400 nm. As a reference, we used a bare HOPG surface. In vapor sensing experiments, the spectral response was the shift in this sharp LSPR, and it was characterized by dividing the measured spectrum with the reference spectrum of Figure 2b, measured in synthetic air.
Note that the bare Ag NPs had a low LSPR intensity with a reflectance minimum at 390 nm, while the LSPR wavelength of graphene-covered Ag NPs was at 397 and 399 nm, for samples #1 and #2, respectively. The redshift in the spectra of graphene-covered nanoparticles, compared to the spectrum of bare Ag NPs, is related, on one hand, to the n-type doping of graphene by electrostatic contact with silver [41], and, on the other hand, the graphene-covered NPs are larger than the bare NPs (see Figure 1), resulting in a redshift [42]. Moreover, the observed redshift can be partly ascribed to the increased effective refractive index of the medium surrounding the NPs [34]. As the LSPR wavelength is sensitive to the composition of the nearby atmosphere, it can be efficiently used for the detection of VOC molecules adsorbed on the surface.

The adsorption of organic molecules to graphene is influenced by the presence of defects, as shown by recent calculations [43]. This, in turn, can affect the sensing properties of samples having a defected graphene overlayer. To investigate this, we introduced defects in sample #2 using O₂ plasma for 5 s. Such plasma treatment induced individual, point-like defects (vacancies) separated by several nanometers, as shown in Figure S1. Although the exposure to plasma was short, the nanoparticles were also affected. The reflectance measured after plasma treatment did not show a well-defined LSPR; it rather showed a significant decrease in the reflectance in the near-UV (Figure 3, spectrum in blue).

This abrupt change in the optical characteristics is ascribed to the silver reacting with oxygen, and the formation of AgO or Ag₂O phases [44]. These oxide phases were successfully decomposed by applying subsequent annealing of sample #2 at 400 °C in an inert atmosphere, which also resulted in the recovery of the metallic Ag NPs. This is expressed by the recovered resonance, as shown in Figure 3 (spectrum in red). Note that after annealing, the new LSPR was measured at a lower wavelength (385 nm). The applied annealing also stabilized the initially produced vacancy-type defects by dangling bond saturation either through the adsorption of molecules, or via the formation of non-hexagonal carbon rings (pentagons, heptagons, etc.) [45].
was proportional to the concentration for all VOCs. As an example, the data obtained for acetone were omitted from the analysis. The PCA score plots of ethanol, propanol, toluene, and water were plotted in Figure 5. Although both samples #1 and #2 showed similar responses, the reflectance change spectra were found to be unique for each VOC, when compared to each other. For example, in addition to the induced LSPR shift, the UV absorption of acetone and toluene vapors was also observed.

To investigate in more detail the VOC sensing data, principal component analysis (PCA) of the measured data sets was carried out and the resulting score plots were compared. As both samples showed a much higher response to acetone compared to the other four substances, and because the UV absorbance of this vapor was in the range of the LSPR, the data on acetone were omitted from the analysis. The PCA score plots of ethanol, propanol, toluene, and water are shown in Figure 6. The cumulated variance of the first three PCs was above 99% for both samples.
Here, each vapor had a unique trajectory. To facilitate the comparison, the data points were projected on the PC1–PC2 planes of the graphs. All trajectories originated from a common point, which was the data point of the initial synthetic air spectrum. Away from this, as the concentration of the vapors increased, the calculated points branched apart, resulting in a clear separation of the trajectories. This showed that the VOC detection by the graphene-covered Ag NPs was substance-specific. One can see on the PC1–PC2 projected plane that the third dimension is needed to properly separate the signal of ethanol and 2-propanol, as the trajectories are overlapping in the 2D projection. Furthermore, to observe the effect of defects in the graphene overlayer, we compared the maximal intensities of the peak (near the LSPR) in the reflectance change spectra to the maximal intensities obtained with the pristine graphene overlayer. These maximal peak intensities are plotted as a function of concentration in Figure 7, for each applied vapor.

![Figure 4](image1.png)

**Figure 4.** (a) LSPR shift of graphene-covered Ag NPs (sample #2) for different concentrations of ethanol vapor. The initial spectrum (0%) measured in synthetic air was used as a reference. (b) Reflectance change spectra, calculated from the same measurements (see main text).

![Figure 5](image2.png)

**Figure 5.** Reflectance change spectra of graphene-covered Ag NP samples when saturated vapors of acetone, ethanol, 2-propanol, toluene, or water are applied. The optical responses are substance-specific for both (a) as-prepared (#1) and (b) O2 plasma-treated (#2) samples.
Figure 6. PCA scores plots of ethanol, 2-propanol, toluene, and water, calculated for the two graphene-covered Ag NP samples. Slightly different vapor sensing properties can be seen in (a) sample #1 (pristine graphene overlayer), compared to (b) sample #2 (defected graphene overlayer). The cumulative variance of the first three PCs was above 99% for both samples.

Figure 7. Maximal peak intensities in the reflectance change spectra as a function of vapor concentration. For each applied vapor, the optical responses of samples #1 and #2 are compared. (a) Acetone, (b) ethanol, (c) 2-propanol, (d) toluene, and (e) water vapors were applied.

One can observe that there were differences in the maximal responses of the two samples, especially for acetone, water, and toluene. The defected graphene overlayer increased the sensitivity to water and toluene. This agrees with recent calculations showing that toluene [43] and water [47,48] adsorb better to graphene with defects. In comparison, exposure to ethanol and 2-propanol resulted in similar optical responses, while acetone...
even gave a reduced signal on defected graphene. To explain such changes in the sensitivity, one must consider the interaction between the VOC molecules, defected graphene, and the underlying Ag NPs as well, since possible charge redistribution effects can result in a slight, opposite shift in the LSPR, reducing the main effect of the refractive index change. This calls for theoretical calculations, which were beyond the scope of the current work. Nonetheless, as demonstrated in Figure 7, the sensitivity to certain vapors can be tuned with structural defects, which can be further explored in the development of selective VOC detectors.

4. Conclusions

We investigated the morphology, optical, and vapor sensing properties of graphene-covered Ag NPs elaborated directly on HOPG substrates. We showed that the prepared hybrid nanostructures displayed pronounced optical responses upon exposure to organic vapors (acetone, ethanol, 2-propanol, toluene, and water). The observed concentration-dependent shifts in the LSPR were substance-specific, as demonstrated by reflectance change spectra and PCA. We further showed that, by introducing structural defects in the graphene overlayer, the optical response of the graphene/Ag NPs increased for toluene and water, while it decreased for acetone detection. These findings can stimulate further research towards increasing the selectivity in VOC sensing.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/nano12142473/s1, Figure S1: STM image of a HOPG substrate exposed to 5 seconds of O$_2$ plasma, Figure S2: Reflectance of bare HOPG surface when different concentrations of ethanol vapor were applied.

Author Contributions: Conceptualization, Z.O.; Investigation, G.P., G.M. and A.P.; Resources, G.M.; Formal analysis, G.P. and A.P.; Funding acquisition, Z.O.; Visualization, G.P. and Z.O.; Writing—original draft, G.P. and Z.O.; Writing—review and editing, G.P., G.M., A.P. and Z.O. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the National Research, Development and Innovation Office (NKFIH) in Hungary, through Grant No. K-134258. Z.O. and G.P. acknowledge the János Bolyai Research Scholarship from the Hungarian Academy of Sciences.

Data Availability Statement: The data presented in this study are available on request from the corresponding author.

Conflicts of Interest: The authors declare no conflict of interest.

References
1. Anker, J.N.; Hall, W.P.; Lyandres, O.; Shah, N.C.; Zhao, J.; Van Duyne, R.P. Biosensing with plasmonic nanosensors. Nat. Mater. 2008, 7, 442–453. [CrossRef] [PubMed]
2. Mayer, K.M.; Hafner, J.H. Localized Surface Plasmon Resonance Sensors. Chem. Rev. 2011, 111, 3828–3857. [CrossRef] [PubMed]
3. Mosier-Boss, P.A. Review of SERS Substrates for Chemical Sensing. Nanomaterials 2017, 7, 142. [CrossRef]
4. Bora, T. Recent Developments on Metal Nanoparticles for SERS Applications. In Noble and Precious Metals—Properties, Nanoscale Effects and Applications; InTech: London, UK, 2018.
5. Cheng, C.-S.; Chen, Y.-Q.; Lu, C.-J. Organic vapour sensing using localized surface plasmon resonance spectrum of metallic nanoparticles self assemble monolayer. Talanta 2007, 73, 358–365. [CrossRef] [PubMed]
6. Bingham, J.M.; Anker, J.N.; Kreno, L.E.; Van Duyne, R.P. Gas Sensing with High-Resolution Localized Surface Plasmon Resonance Spectroscopy. J. Am. Chem. Soc. 2010, 132, 17358–17359. [CrossRef]
7. Kreno, L.E.; Hupp, J.T.; Van Duyne, R.P. Metal—Organic Framework Thin Film for Enhanced Localized Surface Plasmon Resonance Gas Sensing. Anal. Chem. 2010, 82, 8042–8046. [CrossRef]
8. Hossain, M.K.; Kitahama, Y.; Huang, G.G.; Han, X.; Ozaki, Y. Surface-enhanced Raman scattering: Realization of localized surface plasmon resonance using unique substrates and methods. Anal. Bioanal. Chem. 2009, 394, 1747–1760. [CrossRef]
9. Tabor, C.; Murali, R.; Mahmoud, M.; El-Sayed, M.A. On the Use of Plasmonic Nanoparticle Pairs As a Plasmon Ruler: The Dependence of the Near-Field Dipole Plasmon Coupling on Nanoparticle Size and Shape. J. Phys. Chem. A 2009, 113, 1946–1953. [CrossRef]
10. Ray, P.C. Size and Shape Dependent Second Order Nonlinear Optical Properties of Nanomaterials and Their Application in Biological and Chemical Sensing. Chem. Rev. 2010, 110, 5332–5365. [CrossRef]
11. Steinbrück, A.; Strani, O.; Csáki, A.; Fritzschke, W. Sensoric potential of gold–silver core–shell nanoparticles. *Anal. Bioanal. Chem.* 2011, 401, 1241–1249. [CrossRef]

12. Huang, H.; Huang, S.; Yuan, S.; Qu, C.; Chen, Y.; Xu, Z.; Liao, B.; Zeng, Y.; Chu, P.K. High-sensitivity biosensors fabricated by tailoring the localized surface plasmon resonance property of core–shell gold nanorods. *Anal. Chim. Acta* 2011, 683, 242–247. [CrossRef] [PubMed]

13. Lu, L.; Burkey, G.; Halaciuga, I.; Goia, D.V. Core–shell gold/silver nanoparticles: Synthesis and optical properties. *J. Colloid Interface Sci.* 2013, 392, 90–95. [CrossRef]

14. Guo, L.; Jackman, J.A.; Yang, H.-H.; Chen, P.; Cho, N.-J.; Kim, D.-H. Strategies for enhancing the sensitivity of plasmonic nanosensors. *Nano Today* 2015, 10, 213–239. [CrossRef]

15. Stockman, M.I. Nanoplasmonics: The physics behind the applications. *Phys. Today* 2011, 64, 39–44. [CrossRef]

16. Rycenga, M.; Cobley, C.M.; Zeng, J.; Li, W.; Moran, C.H.; Zhang, Q.; Qin, D.; Xia, Y. Controlling the Synthesis and Assembly of Silver Nanostructures for Plasmonic Applications. *Chem. Rev.* 2011, 111, 3669–3712. [CrossRef]

17. Shen, H. Plasmonic nanoparticles: Towards the fabrication of biosensors. *IOP Conf. Ser. Mater. Sci. Eng.* 2015, 87, 012009. [CrossRef]

18. Mortazavi, D.; Kouzani, A.Z.; Kaynak, A.; Duan, W. Nano-plasmonic biosensors: A review. In Proceedings of the 2011 IEEE/ICME International Conference on Complex Medical Engineering, Harbin, China, 22–25 May 2011; pp. 31–36.

19. Lee, K.S.; El-Sayed, M.A. Gold and silver nanoparticles in sensing and imaging: Sensitivity of plasmon response to size, shape, and metal composition. *J. Phys. Chem. B* 2006, 110, 19220–19225. [CrossRef] [PubMed]

20. Huang, X.; El-Sayed, M.A. Gold nanoparticles: Optical properties and implementations in cancer diagnosis and photothermal therapy. *J. Adv. Res.* 2010, 1, 13–28. [CrossRef]

21. Jain, P.K.; Lee, K.S.; El-Sayed, I.H.; El-Sayed, M.A. Calculated absorption and scattering properties of gold nanoparticles of different size, shape, and composition: Applications in biological imaging and biomedicine. *J. Phys. Chem. B* 2006, 110, 7238–7248. [CrossRef]

22. Scardaci, V.; PULVIRENTI, M.; Condorelli, M.; Compagnini, G. Monochromatic light driven synthesis and growth of flat silver nanoparticles and their plasmon sensitivity. *J. Mater. Chem. C* 2020, 8, 9734–9741. [CrossRef]

23. Condorelli, M.; Litti, L.; Pulvirenti, M.; Scardaci, V.; Meneghetti, M.; Compagnini, G. Silver nanoplates paved PMMA cuvettes as a cheap and re usable plasmonic sensing device. *Appl. Surf. Sci.* 2021, 566, 150701. [CrossRef]

24. Mori, T.; Mori, T.; Fujii, M.; Tominari, Y.; Otomo, A.; Yamaguchi, K. Optical Properties of Low-Loss Ag Films and Nanostructures on Transparent Substrates. *ACS Appl. Mater. Interfaces* 2018, 10, 8333–8340. [CrossRef] [PubMed]

25. Reed, J.C.; Zhu, H.; Zhu, A.Y.; Li, C.; Cubukcu, E. Graphene-enabled silver nanoantenna sensors. *Nano Lett.* 2012, 12, 4090–4094. [CrossRef]

26. Loiseau, A.; Asila, V.; Boitel-Aullen, G.; Lam, M.; Salmain, M.; Boujday, S. Silver-Based Plasmonic Nanostructures for and Their Use in Biosensing. * Biosensors* 2019, 9, 78. [CrossRef] [PubMed]

27. Kang, H.; Buchman, J.T.; Rodriguez, R.S.; Ring, H.L.; He, J.; Bantz, K.C.; Haynes, C.L. Stabilization of Silver and Gold Nanoparticles: Preservation and Improvement of Plasmonic Functionalities. *Chem. Rev.* 2019, 119, 664–699. [CrossRef] [PubMed]

28. Leenaerts, O.; Partoens, B.; Peeters, F.M. Graphene: A perfect nanoballoon. *Appl. Phys. Lett.* 2008, 93, 10–13. [CrossRef]

29. Bunch, J.S.; Verbridge, S.S.; Alden, J.S.; Van Der Zande, A.M.; Parpia, J.M.; Craighead, H.G.; McEuen, P.L. Impermeable atomic membranes from graphene sheets. *Carbon N. Y.* 2011, 49, 1241–1249. [CrossRef] [PubMed]

30. Mortazavi, D.; Kouzani, A.Z.; Kaynak, A.; Duan, W. Nano-plasmonic biosensors: A review. In Proceedings of the 2011 IEEE/ICME International Conference on Complex Medical Engineering, Harbin, China, 22–25 May 2011; pp. 31–36.

31. Lee, K.S.; El-Sayed, M.A. Gold and silver nanoparticles in sensing and imaging: Sensitivity of plasmon response to size, shape, and metal composition. *J. Phys. Chem. B* 2006, 110, 19220–19225. [CrossRef] [PubMed]

32. Huang, X.; El-Sayed, M.A. Gold nanoparticles: Optical properties and implementations in cancer diagnosis and photothermal therapy. *J. Adv. Res.* 2010, 1, 13–28. [CrossRef]

33. Jain, P.K.; Lee, K.S.; El-Sayed, I.H.; El-Sayed, M.A. Calculated absorption and scattering properties of gold nanoparticles of different size, shape, and composition: Applications in biological imaging and biomedicine. *J. Phys. Chem. B* 2006, 110, 7238–7248. [CrossRef]

34. Alharbi, R.; Irannejad, M.; Yavuz, M. A Short Review on the Role of the Metal-Graphene Hybrid Nanostructure in Promoting the localized Surface Plasmon Resonance Sensitivity. *Sensors* 2019, 19, 862–876. [CrossRef]

35. Rostami, S.; Mehdinia, A.; Niroumand, R.; Jabbari, A. Enhanced LSPR performance of graphene nanoborons-silver nanoparticles hybrid as a colorimetric sensor for sequential detection of dopamine and glutathione. *Anal. Chim. Acta* 2020, 1120, 11–23. [CrossRef] [PubMed]

36. El barghouti, M.; Akjoju, A.; Mir, A. Design of silver nanoparticles with graphene coatings layers used for LSPR biosensor applications. *Vaccine* 2020, 180, 109497. [CrossRef]

37. Schriver, M.; Regan, W.; Garnett, W.J.; Zaniiewski, A.M.; Crommie, M.F.; Zettl, A. Graphene as a Long-Term Metal Oxidation Barrier: Worse Than Nothing. *ACS Nano* 2013, 7, 5763–5768. [CrossRef] [PubMed]

38. Khodashenas, B.; Ghorbani, H.R. Synthesis of silver nanoparticles with different shapes. *Arab. J. Chem.* 2019, 12, 1823–1838. [CrossRef]

39. Wiley, B.; Sun, Y.; Mayers, B.; Xia, Y. Shape-Controlled Synthesis of Metal Nanostructures: The Case of Silver. *Chem. A Eur. J.* 2005, 11, 454–463. [CrossRef]
40. Osváth, Z.; Pálinkás, A.; Piszter, G.; Molnár, G. Synthesis and Characterization of Graphene–Silver Nanoparticle Hybrid Materials. *Materials* **2020**, *13*, 4660. [CrossRef]

41. Jimenez-Villacorta, F.; Climent-Pascual, E.; Ramirez-Jimenez, R.; Sanchez-Marcos, J.; Prieto, C.; De Andrés, A. Graphene–ultrasmall silver nanoparticle interactions and their effect on electronic transport and Raman enhancement. *Carbon* **2016**, *101*, 305–314. [CrossRef]

42. Yguerabide, J.; Yguerabide, E.E. Light-Scattering Submicroscopic Particles as Highly Fluorescent Analogs and Their Use as TracerLabels in Clinical and Biological Applications. *Anal. Biochem.* **1998**, *262*, 137–156. [CrossRef]

43. Anithaa, V.S.; Suresh, R.; Kuklin, A.V.; Vijayakumar, S. Adsorption of volatile organic compounds on pristine and defected nanographene. *Comput. Theor. Chem.* **2022**, *1211*, 113664. [CrossRef]

44. Al-Sarraj, A.; Saoud, K.M.; Elmel, A.; Mansour, S.; Haik, Y. Optoelectronic properties of highly porous silver oxide thin film. *SN Appl. Sci.* **2021**, *3*, 15. [CrossRef]

45. Osváth, Z.; Vértesy, G.; Tapasztó, L.; Wéber, F.; Horváth, Z.E.; Gyulai, J.; Biró, L.P. Atomically resolved STM images of carbon nanotube defects produced by Ar+ irradiation. *Phys. Rev. B* **2005**, *72*, 045429. [CrossRef]

46. Forker, R.; Gruenewald, M.; Fritz, T. Optical differential reflectance spectroscopy on thin molecular films. *Annu. Rep. Prog. Chem. Sect. C Phys. Chem.* **2012**, *108*, 34–68. [CrossRef]

47. Liang, Z.; Li, K.; Wang, Z.; Bu, Y.; Zhang, J. Adsorption and reaction mechanisms of single and double H2O molecules on graphene surfaces with defects: A density functional theory study. *Phys. Chem. Chem. Phys.* **2021**, *23*, 19071–19082. [CrossRef]

48. Carraro, G.; Savio, L.; Vattuone, L. Influence of Defects and Heteroatoms on the Chemical Properties of Supported Graphene Layers. *Coatings* **2022**, *12*, 397. [CrossRef]