Subthalamic nucleus long-range synchronization—an independent hallmark of human Parkinson’s disease

Shay Moshel1,2,3,4,†, Reuben R. Shamir1,3†, Aeyal Raz2, Fernando R. de Noriega6, Renana Eitan7, Hagai Bergman1,2,3 and Zvi Israel6

1 Department of Medical Neurobiology, IMRIC, The Hebrew University-Hadassah Medical School, Jerusalem, Israel
2 The Interdisciplinary Center for Neural Computation, The Hebrew University, Jerusalem, Israel
3 The Edmond and Lily Safra Center for Brain Sciences, The Hebrew University, Jerusalem, Israel
4 The Research Laboratory of Brain Imaging and Stimulation, The Jerusalem Mental Health Center, Kfar-Shaul Etanim, Hebrew University-Hadassah Medical School, Jerusalem, Israel
5 Department of Anesthesiology, University of Wisconsin School of Medicine and Public Health, Madison, WI, USA
6 Department of Neurosurgery, Center for Functional and Restorative Neurosurgery, Hadassah University Hospital, Jerusalem, Israel
7 Department of Psychiatry, Hadassah University Hospital, Jerusalem, Israel

*Correspondence:
Shay Moshel, The Interdisciplinary Center for Neural Computation and the Edmond and Lily Safra Center for Brain Sciences and the Department of Medical Neurobiology, The Hebrew University - Hadassah Medical School, Building 3, PO Box 12272, Jerusalem 91120, Israel
e-mail: shaymoshel@gmail.com
†These authors have contributed equally to this work.

INTRODUCTION

The subthalamic nucleus (STN) plays a critical role in the control of basal ganglia activity (Kitai and Kita, 1987; Nambu et al., 2002). In Parkinson’s disease (PD), midbrain dopaminergic neurons degenerate, leading to a cascade of physiological changes that strongly affect the STN (Bergman et al., 1994; Hamani et al., 2004). Inactivation (Bergman et al., 1990; Aziz et al., 1991; Alvarez et al., 2009) and deep brain stimulation (DBS, Benazzouz et al., 1993; Pollak et al., 1993; Benabid et al., 1994; Weaver et al., 2009; Follett et al., 2010; Moro et al., 2010; Williams et al., 2010; Bronstein et al., 2011; Lhomme et al., 2012; Odekerken et al., 2013; Schuepbach et al., 2013) of the STN are highly effective in the management of advanced PD.

Neuronal oscillations, at the level of action-potential (spike) discharge (Rodríguez-Oroz et al., 2001; Kuhn et al., 2005; Moran et al., 2008; Zaidel et al., 2010; Guo et al., 2012; Lourens et al., 2013) and local field potential (Kuhn et al., 2009; Chen et al., 2010; Giannicola et al., 2010; Rosa et al., 2011) have been observed in physiological studies of the STN of PD patients undergoing DBS surgery. LFPs span the frequency range of 1–70 Hz (or 1–400 Hz, if one include the high gamma peaks reported at 65–90 Hz and 250–350 Hz (Danish et al., 2007), but see (Yuval-Greenberg et al., 2008) for possible confounding factors in the high frequency regime of LFP), whereas spikes have their maximal power around 1000 Hz. Thus, although LFP oscillations have been thought to imply spike synchronization (Brown and Williams, 2005; Hammond et al., 2007; de-Solages et al., 2011), they more likely represent sub-threshold phenomena such as synaptic activity (Belitski et al., 2010; Buzsaki et al., 2012) which is probably correlated with spike activity.

Conclusive evidence of the correlation (and causality) between neuronal oscillations and synchronization in the PD STN has

Keywords: Parkinson’s disease, subthalamic nucleus, deep brain stimulation, oscillations, synchronization
remained elusive. Physiological studies of neuronal synchronization in the STN of the MPTP primate model are not yet reported. Robust oscillatory synchronization patterns of STN spiking activity have been reported in the 6-hydroxydopamine rodent model of Parkinsonism (Machado et al., 2006; Mallet et al., 2008a,b; Lintas et al., 2012). In human PD patients, oscillatory synchronization of spiking activity has been reported in several studies (Levy et al., 2000, 2002a,b; Amirnovin et al., 2004; Weinberger et al., 2006; Hanson et al., 2012; Alavi et al., 2013; Lourens et al., 2013) but there have been no detailed descriptions of the dependence of the neuronal synchronization on the oscillatory activity or the spatial properties of the neuronal pairs (e.g., simultaneous recording of neurons from the oscillatory and non-oscillatory regions of the STN, see below).

Previous studies have shown that the STN of PD patients can be divided into a dorso-lateral oscillatory region (DLOR) and ventro-medial non-oscillatory region (VMNR) (Moran et al., 2008; Zaidel et al., 2010; Seifried et al., 2012; Guo et al., 2013). The first aim of this study was to explore the properties of neuronal (spike) synchronization of the STN of human PD patients, principally within and between the different STN domains. The second goal was to further explore the relationship between oscillations and synchronization phenomena in the neural activity of the STN.

To overcome the inherent technical difficulties of spike isolation (Joshua et al., 2007; Hill et al., 2011) and spike sorting (Lewicki, 1998) in the electrically noisy environment of the human operating room, and to increase the sensitivity of correlation analysis (Bedenbaugh and Gerstein, 1997; Gerstein, 2000) this study used the unresolved collective (multi-unit) spiking activity recorded by two different microelectrodes exploring the boundaries and the domains of the STN during DBS procedures. This enabled the exploration of the properties of long-range correlation in the STN, in contrast to correlation studies of the activity recorded by a single electrode (e.g., Moran et al., 2008) which can only reveal short range correlations.

**MATERIALS AND METHODS**

**PATIENTS AND SURGERY**

Simultaneous microelectrode recordings from two electrodes in patients with Parkinson’s disease (PD) undergoing surgery for subthalamic nucleus (STN) deep brain stimulation (DBS) were analyzed in this study. All patients met accepted criteria for STN DBS and signed informed consent for surgery. Microelectrode recording is performed to accurately localize STN borders and domains, in order to optimize the placement of the DBS electrode and thus enhance the therapeutic effects of the DBS procedure. The data collection was therefore done as part of our routine procedures and not part of a clinical trial. This study was authorized and approved by the Institutional Review Board of Hadassah University Hospital in accordance with the Helsinki Declaration (reference codes: 0545-08-HMO and HMO: 10-18.01.08).

Surgery was performed using a CRW stereotactic frame (Radionics, Burlington, MA, USA). STN target coordinates were chosen as a composite of the indirect anterior commissure-posterior commissure (AC-PC) atlas-based location and direct (1.5 or 3 Tesla) T2 magnetic resonance imaging (MRI), using Framelink 4 or 5 software (Medtronic, Minneapolis, USA). The recordings used in this study were made while the patients were awake without sedation. The patient’s level of awareness was continuously assessed clinically and, if drowsy, the patient was stimulated and awoken through conversation by a member of the surgical team. Data were obtained while the patients were off dopaminergic medication, which was stopped 12 h prior to surgery.

**MICROELECTRODE RECORDINGS**

Data were acquired with the MicroGuide system (Alpha-Omega Engineering, Nazareth, Israel). Neurophysiological activity was recorded using polyamide coated tungsten microelectrodes (Alpha Omega) with impedance mean ± standard deviation (SD) of 0.60 ± 0.11 MΩ (measured at 1 kHz at the beginning of each trajectory). The signal was amplified by 10,000, band-pass filtered from 250 to 6000 Hz using four-pole Butterworth filter hardware, and sampled at 48 kHz by a 12-bit A/D converter (using ±5 V input range). Local field potentials were not recorded due to constraints of electrical noise in the operating room.

Microelectrode recording was performed using two parallel microelectrodes starting 10 mm above the estimated center of the dorsolateral STN target, based on the pre-operative T2 MRI image. The two electrodes were simultaneously advanced, and therefore the distance between the two electrodes was fixed (2 mm) during all recordings. Trajectories followed a double-oblique approach (approximately 60° from the axial AC-PC plane and 15° from the mid-sagittal plane) toward the STN target. The angles of the trajectory were slightly modified to avoid the cortical sulci, the ventricles and major blood vessels as revealed by gadolinium-enhanced T1 MRI (Machado et al., 2006). The “central” electrode was directed at the center of the STN target, and an “anterior” (ventral) electrode was located 2 mm anterior to the central electrode. Typically, the electrodes were advanced in steps of ~100 μm between successive recordings sites within the STN. Only trajectories where both electrodes had passed through the STN for at least 4 mm were used in this study (yielding 72 trajectories of 2 electrodes from 57 PD patients undergoing bilateral STN deep brain stimulation surgery). After identification of the STN ventral border by the electro-physiologist, the STN and its sub-regions were automatically detected using the Hidden Markov model (HMM) method (Zaidel et al., 2009).

**DATABASE**

We studied 72 STN trajectories (each of 2 electrodes) from 57 PD patients, 40 males and 17 females, aged 58.9 ± 10.3 years (mean ± standard deviation, SD) and with disease duration of 10.3 ± 4.7 years (mean ± SD). The UPDRS motor part score, UPDRS III, was 49.2 ± 17.8 (mean ± SD) when assessed off dopamine replacement therapy before surgery. Patient details and clinical effects of the surgery are given at Table 1.

The minimal recording time duration of a STN pair to be included in this study was 5 s (analysis of the subset of recording with minimal recording duration of 10 s reveal similar results, data not shown). A total of 2390 multi-unit pair sites, in which both electrodes were judged to be inside the STN for the minimal
Table 1 | Summary of clinical data, before and after the DBS surgery, include medical treatment [total daily levodopa equivalent dose (LED)] and the motor part of patients rate scale for Parkinson; The Unified Parkinson’s Disease Rating Scale, part 3 (UPDRS III; maximum value is equal to 108).

| Patient number | Age (years) | Disease duration (years) | Gender | UPDRS III Before DBS (Med: On\Off) | UPDRS III After DBS: (Stim\Med: On\Off) | Medications (LED) daily doses |
|---------------|-------------|--------------------------|--------|-----------------------------------|------------------------------------------|-----------------------------|
|               |             |                          |        | On, Off                            | On, On, Off, Off, On, Off, Off, Months since DBS | Before STN DBS | After STN DBS | Months Since DBS |
| 1             | 62          | 8                        | M      | 10 39.5                            | 21 23 34 34 12                         | 400            | 1100         | 8             |
| 2             | 73          | 10                       | M      | 23 58.5                            | NA NA NA NA NA                         | 675            | 200          | NA            |
| 3             | 51          | 14                       | F      | 17 99                              | 2 16 9 46 13                           | 1700           | 875          | NA            |
| 4             | 68          | 9                        | M      | NA NA                              | NA NA NA NA NA                         | 1662.5         | 1246.9       | 6             |
| 5             | 31          | 13                       | F      | 19 62                              | 14 11 53 12                            | NA             | 0            | NA            |
| 6             | 58          | 22                       | M      | 22.5 54                            | 5 14 61 48 8                           | 2575           | 900          | 8             |
| 7             | 73          | 6                        | M      | 22 55                              | 5 18 NA NA 4                           | 1700           | 700          | NA            |
| 8             | 75          | 8                        | F      | 17 51                              | 17 21 24 37 8                          | 1400.1         | 89775        | 7             |
| 9             | 61          | 5                        | F      | 38 73                              | 1 10 47 8                             | 1250           | 125          | 5             |
| 10            | 56          | 8                        | M      | 29 70                              | NA NA NA NA NA                         | 875            | 250          | NA            |
| 11            | 63          | 12                       | F      | 19 49                              | 5 16 3                                | 890            | 459          | NA            |
| 12            | 49          | 10                       | M      | NA NA                              | NA NA NA NA NA                         | 900            | NA           | NA            |
| 13            | 64          | 11                       | F      | 3 21                               | 2 4 7 21 5                            | 1505           | 4375         | 4             |
| 14            | 61          | 6                        | M      | 32 60                              | 10 14 51 61 3                         | 700            | 200          | 2             |
| 15            | 39          | 10                       | F      | 16 47                              | 5 18 27 57 5                           | 512.5          | 3075         | 3             |
| 16            | 61          | 11                       | M      | 10 44                              | NA NA NA NA NA                         | 500            | 510          | NA            |
| 17            | 73          | 20                       | M      | 16 60                              | NA NA NA NA NA                         | 1496           | NA           | NA            |
| 18            | 56          | 13                       | F      | NA NA                              | NA NA NA NA NA                         | NA             | NA           | NA            |
| 19            | 62          | 17                       | M      | 22 72                              | NA NA NA NA NA                         | 1550           | 250          | NA            |
| 20            | 70          | 13                       | F      | 13 41                              | NA NA NA NA NA                         | 400            | NA           | NA            |
| 21            | 53          | 9                        | M      | 10 41                              | NA NA NA NA NA                         | 375            | NA           | NA            |
| 22            | 72          | 8                        | M      | 4 17                               | NA NA NA NA NA                         | 1125           | NA           | NA            |
| 23            | 50          | 8                        | F      | NA NA                              | NA NA NA NA NA                         | 550            | NA           | NA            |
| 24            | 62          | 5                        | M      | NA NA                              | NA NA NA NA NA                         | NA             | NA           | NA            |
| 25            | 58          | 7                        | M      | 14 42                              | NA NA NA NA NA                         | 300            | NA           | NA            |
| 26            | 63          | 12                       | M      | 19 56                              | NA NA NA NA NA                         | 1105           | NA           | NA            |
| 27            | 52          | 5                        | M      | 3 26                               | NA NA NA NA NA                         | 650            | NA           | NA            |
| 28            | 53          | 10                       | F      | 9 42                               | NA NA NA NA NA                         | 770            | NA           | NA            |
| 29            | 74          | 9                        | F      | 42 73                              | NA NA NA NA NA                         | 1250           | NA           | NA            |
| 30            | 57          | 7                        | M      | 37 78                              | NA NA NA NA NA                         | 1370           | NA           | NA            |

(Continued)
Table 1 | Continued

| Patient number | Age (years) | Disease duration (years) | Gender | UPDRS III Before DBS (Med: On/Off) | After DBS: (StimMed: On/Off) | Medications (LED) daily doses | Months since DBS |
|----------------|-------------|--------------------------|--------|--------------------------------|------------------------------|------------------------------|-----------------|
|                |             |                          |        | On | Off | On | On | Off | Off | Off | On | Off | Months | Before STN DBS | After STN DBS | Months | Since DBS |
| 31             | 67          | 9                        | M      | 14 | 34 | NA | NA | NA | NA | NA | Na | NA | 1000 | Na | Na | Na |
| 32             | 59          | 10                       | M      | 13 | 47 | NA | NA | NA | NA | NA | Na | NA | 1480 | Na | Na | Na |
| 33             | 56          | 8                        | M      | 22 | 69 | NA | NA | NA | NA | NA | Na | NA | 750  | Na | Na | Na |
| 34             | 53          | 8                        | M      | 32 | 49 | NA | NA | NA | NA | NA | Na | NA | 562.5| Na | Na | Na |
| 35             | 51          | NA                       | M      | 29 | 43 | NA | NA | NA | NA | NA | Na | NA | 200  | Na | Na | Na |
| 36             | 61          | 5                        | M      | 0  | 0  | NA | NA | NA | NA | NA | Na | NA | 850  | Na | Na | Na |
| 37             | 53          | 3                        | M      | 6  | 49 | NA | NA | NA | NA | NA | Na | NA | 750  | Na | Na | Na |
| 38             | NA          | 4                        | M      | 34 | 56 | NA | NA | NA | NA | NA | Na | NA | 600  | Na | Na | Na |
| 39             | 74          | 15                       | M      | 31 | 57 | NA | NA | NA | NA | NA | Na | NA | 750  | Na | Na | Na |
| 40             | 71          | 22                       | M      | 35 | 51 | NA | NA | NA | NA | NA | Na | NA | 2000 | Na | Na | Na |
| 41             | 66          | 3.5                      | F      | 16 | 46 | NA | NA | NA | NA | NA | Na | NA | 750  | Na | Na | Na |
| 42             | 66          | 10                       | M      | 11 | 63 | NA | NA | NA | NA | NA | Na | NA | 600  | Na | Na | Na |
| 43             | 71          | NA                       | F      | 6.5| 28.5| 15 | 23 | 56 | 59 | 28 | 1525| 100 | Na |
| 44             | 51          | 12                       | M      | NA | NA | 4  | 3  | 48 | 31 | 23 | 1025| 731.5| 10 |
| 45             | 46          | NA                       | M      | 1  | 25.5| NA | NA | NA | NA | NA | Na | NA | 900  | Na | Na | Na |
| 46             | 60          | 11                       | M      | 10.5| 42.5| 23 | 29 | 40 | 38 | 20 | 500 | 250 | Na |
| 47             | 59          | 6                        | M      | 15.5| 40.5| 23 | 29 | 40 | 38 | 20 | 500 | 250 | Na |
| 48             | 56          | 10                       | M      | 6.5| 37.5| 12 | 20 | 23 | 31 | 26 | 900 | 675 | Na |
| 49             | 72          | 16                       | M      | NA | NA | 4  | 3  | 48 | 31 | 23 | 1025| 731.5| 10 |
| 50             | 54          | 5                        | F      | 11 | 43.5| 10 | 14 | 60 | 22 | 23 | 850 | 600 | Na |
| 51             | 55          | NA                       | M      | 7.5| 36.5| NA | NA | NA | NA | NA | Na | NA | 1150 | 450 | Na |
| 52             | 47          | 8                        | M      | 7.5| 51  | NA | NA | NA | NA | NA | Na | NA | 1200 | 722.12| Na |
| 53             | 42          | 9                        | F      | 4.5| 43  | NA | NA | NA | NA | NA | Na | NA | 187  | 250 | Na |
| 54             | 29          | 9                        | M      | NA | NA | NA | NA | NA | NA | NA | Na | NA | 1200 | 612.5| Na |
| 55             | 69          | 12                       | M      | 7.5| 28.5| 12 | 13 | 27 | 21 | 14 | 1609.5| 768.13| Na |
| 56             | 53          | 20                       | M      | 39.5| 83  | NA | NA | NA | NA | NA | Na | NA | 2200 | 537.5| Na |
| 57             | 61          | 22                       | F      | 15.5| 56.5| NA | NA | NA | NA | NA | Na | NA | 1000 | 500 | 18 |
| Mean           | 58.9        | 10.3                     | M40/F17| 172| 49.2| 9.58| 15.7| 34 | 44 | 12.588| 1037.3| 506.48| 9.83 |
| STD            | 10.3        | 4.7                      |        | 11.1| 17.8| 6.75| 6.65| 18 | 15 | 8.3221| 516.56| 309.06| 7.6 |

Mean and STD are respectively for the mean and standard deviation of each column values.
duration, were studied. The same data base was used for the single site (oscillation) analysis, yielding 4780 single STN sites. Recording (and analysis) time duration of the STN pairs equaled 23.7 ± 25.3 s (mean ± SD).

**ANALYSIS OF SYNCHRONIZATION AND OSCILLATIONS**

All data analysis utilized custom-made MATLAB 7.10b (R2010b) routines. The local field potential frequency domain was filtered out by the recording apparatus. Burst frequencies below the range of the operating room band-pass filter (250–6000 Hz) could be detected using the rectified signal, which follows the envelope of multi-unit activity (Moran et al., 2008; Halliday and Farmer, 2010; Moran and Bar-Gad, 2010; Zaidel et al., 2010). The raw 250–6000 Hz analog signal was therefore rectified by the “absolute” operator and the global mean was subtracted. Thus, the resulting analysis represents only spike activity.

The average power spectrum density (PSD) at each site was calculated using Welch’s method with a 1.5 s Hamming window (50% overlap), after removing the local window mean, and with a 131,072 FFT points (nfft), yielding spectral resolution of 1/3 Hz \[f_{\text{res}} = \frac{\text{f}_\text{res}}{\text{nfft}}\], where \(f_s\) = sampling frequency and \(f_{\text{res}}\) is the spectral resolution. PSD amplitude is affected by the amplitude of the recorded neural activity, which is impacted by non-physiological factors such as the impedance of the electrode (Zaidel et al., 2010). To create homogenous PSD results for all recorded sites, the “relative” (normalized) power spectral density was calculated by dividing it by the total power of the signal between 0 and 3000 Hz. This relative, or normalized, power spectral density therefore estimates the spectral peak in relation to the other peaks in the spectrogram.

To compute coherence, the magnitude squared (MS-) coherence method (Kay, 1988; Miller and Sigvardt, 1998) was used. Welch’s method was utilized, with a 1.5 s Hamming window (50% overlap), after removing the local window mean and with a spectral resolution of 1/3 Hz (same conventions as for PSD). Coherence values are limited (by definition) between 0 and 1. All coherence averages were therefore calculated in Fisher’s transform domain (Miranda de Sa et al., 2009) and then reversed.

By definition, the removal of each window mean in the spectrum and the coherence analysis eliminate any power at 0 Hz (DC). We therefore start all the spectrum and coherence plot of this manuscript at 1 Hz.

A constant baseline level emerged in our coherence results (e.g., Figures 4A,B). This baseline probably resulted from the finite sampling of two “random noise” sources. To verify this, pairs of Gaussian random noise sources were simulated. The simulated data were subjected to the same filters and absolute operator as the real neuronal data and the same analysis tools. The magnitude of the coherence baseline dropped exponentially as time duration increased. Therefore, the baseline level seen in the STN coherence is most likely due to the finite (and relatively) short duration (mean = 23.7 s) of the recordings in human patients. The coherence functions were normalized by the subtraction of the average coherence of the randomly shuffled (10,000 times) pairs from the same STN domain. Note that the normalized coherence functions can therefore display negative coherence values.

**SYNCHRONIZATION AND OSCILLATION STRENGTH (SYNC AND OSCIL SCORES)**

Rosenberg and Halliday (Halliday et al., 1995, 2000; Farmer et al., 1997; Farmer, 1998) proposed a very useful method to estimate coherence significance. However, this method employs a threshold confidence level, and does not offer a quantitative measure of synchronization strength. Therefore, a Z-score like method (effective Z-score: \(Z^\ast\)) was devised to determine the synchronization and oscillation strength. The Z-score of a given parameter is defined as the number of standard deviations above (or below) the mean. In this case the parameter was the maximum value (peak value) of the smoothed PSD or coherence (see below). However, instead of using the standard deviation of the entire frequency range, a tail standard deviation (\(\sigma_{\text{tail}}\)) was defined in the frequency range of 35 to 70 Hz. In this range, no coherence or power spectrum phenomena were observed in our dataset (Figures 1D, 2A–C, 3A,B, 4B). To smooth the coherence, a simple moving average (SMA) was calculated, with a window size of 23 samples (7.67 Hz), and a delta of one sample (i.e., the frequency resolution of 1/3 Hz). The synchronization strength or score was defined as \(Z^\ast = \frac{\text{MAX}(\text{SMA}(C(f))) - \mu}{\sigma_{\text{tail}}}\). MAX(SMA(C(f))) is the maximum value of the coherence after smoothing with the moving average, and \(\mu\) is the coherence mean. To find the frequency in which the spectrum or the coherence achieved maximal change, the arg-max(SMA(C(f))) was calculated. The coherence \((C(f))\) maximal peaks were defined in the smoothed coherence function with minimal distances of 5 Hz between them. The search for the coherence peak was started at the lower frequency, and progressed to the largest value of the smoothed coherence function. All calculations (max, mean and arg-max) were performed in the frequency range between 1 and 70 Hz. Negative scores were found in a few cases, due to residual high power at the low (1–2 Hz) frequency range, and these were ignored.

To determine the oscillation strength, the same effective Z-score as for the synchronization was used and defined as the oscil score. The maximum value of the smoothed PSD (by a simple moving average, with window size of 23 samples, and delta of one sample, i.e., 1/3 Hz), and the tail standard deviation (\(\sigma_{\text{tail}}\)) in the frequency range of 35–70 Hz were calculated. The oscil score was defined as \(Z^\ast = \frac{\text{MAX}(|\text{PSD}(C(f))|) - \mu}{\sigma_{\text{tail}}}\). MAX(SMA(PSD(f))) is the maximum value of the PSD after smoothing with the moving average; \(\mu\) is the PSD mean.

To explore the relationship between oscillation and synchronization a statistical measure of the oscillation strength of the two oscillatory sites was used. The average PSD\(^\ast\) = (PSD1 + PSD2)/2 was calculated, where PSD1 and PSD2 were the power spectrum densities of each site in the neuronal pair, and the oscil score of PSD\(^\ast\) was calculated. Additionally, other estimates of oscil scores of the two sites were calculated as: \(\text{min}(\text{oscil}_1, \text{oscil}_2); \text{max}(\text{oscil}_1, \text{oscil}_2); \text{GeoMean}(\text{oscil}_1, \text{oscil}_2)\), and as the geometric mean of the two scores where oscil\(_1\) oscil\(_2\) were the oscil scores of each PSD site. The geometric mean was calculated as: oscil = \(\text{sign}(\text{oscil}_1 * \text{oscil}_2) * \text{GeoMean}(|\text{oscil}_1|, |\text{oscil}_2|)\), where sign was the sign operator of oscil\(_1\) and oscil\(_2\) product, in the case of negative values.
FIGURE 1 | An example of synchronous oscillations between two recording sites in the STN. (A) Raw analog data. Data is recorded by two microelectrodes separated horizontally by a distance of 2 mm, hardware band pass filtered between 250 and 6000 Hz, and digitally sampled at 48 KHz. The 1 s data in the rectangle is expanded in (B). (C) Power spectrum of the recording shown in (A). (D) Coherence function of the two STN sites. Oscillations scores = 31.57 and 17.13 respectively; Synchronization score = 4.89.

Synchronization or oscillations were defined to be significant when the scores reached the $Z^* \geq 2$ (i.e., the coherence or the PSD peak value was higher than 2 SD of the mean values of these functions).

COHERENCE CONFIDENCE LEVEL

To assess the validation of sync score the confidence level (CL) of the coherence analysis (Halliday et al., 1995) was used. We divided the microelectrode records of duration R into L non-overlapped disjoint segments of duration S ($R = L \times S$). The total spectrum was calculated using the average of the magnitude-squared (MS) of the discrete Fourier transform (periodogram), after removing the local mean in each segment S. Each segment contained $S = 2^{16}$ samples with a frequency resolution of 0.7336 Hz. Only complete segments were analyzed; data points at the end of the record that did not make a complete segment were not included in the analysis. The procedures were implemented using Neurospec free MATLAB toolbox: http://www.neurospec.org. To obtain the approximate confidence interval for 95% and 99% from the data points, the level thresholds: $CL_{95} = 1 - 0.05^{1/(L - 1)}$ and $CL_{99} = 1 - 0.01^{1/(L - 1)}$, respectively, were used. Figure 4B depicts examples of the relations between MS-coherence estimates ($Z$-scores) we used in the manuscript with coherence confidence levels of 95% and 99% respectively.

ASSESSING THE CAUSAL RELATIONS BETWEEN OSCILLATIONS AND SYNCHRONY

Spurious synchronization can arise from non-coupled oscillatory sites that oscillate in the same frequency bands (i.e., two atomic clocks might be synchronized due to their exact frequency although there is no physical coupling between them (Strogatz, 2003). To rule out this spurious oscillation-synchronization, the mean coherence of randomly shuffled pairs (10,000 times) was calculated for each category (all pairs, DLOR-DLOR, VMNR-VMNR, and DLOR-VMNR) of the STN. The shuffling was performed using the Mersenne Twister algorithm (Matsumoto and Nishimura, 1998) with a different seed number in each iteration.

RESULTS

SYNCHRONIZATION OCCURS ONLY BETWEEN DLOR PAIRS

Figure 1 show an example of synchronous oscillatory activity as recorded by two electrodes inserted into the STN of a PD patient during DBS procedures. The raw analog data is shown in two time scales in Figures 1A,B. The power spectrums and the coherence function of this recording are shown in Figures 1C,D, respectively. One can easily observe the synchronous oscillations in the beta range ($\sim$20 Hz) in this example.

To explore the properties of STN neuronal synchronization, STN spiking activity simultaneously recorded from two electrodes...
was analyzed (Figure 2). In total, 2390 multi-unit pairs along 72 STN trajectories (with >4 mm STN span in both electrodes) from 57 PD patients undergoing DBS surgery were included in the analysis.

Previous physiological studies of the basal ganglia in the rodent (Mallet et al., 2008a,b) and primate (Bergman et al., 1994; Nini et al., 1995; Raz et al., 1996, 2000, 2001; Goldberg et al., 2002) models of PD have indicated abnormal synchronicity of basal ganglia neurons as one of the major changes occurring in the network following dopamine depletion. Nevertheless, when the neuronal synchronization of simultaneously recorded STN sites (over the entire STN 2390 pairs) was measured, no distinguishable synchronization was found.

The STN can be spatially differentiated into sub-regions according to neural activity (Zaidel et al., 2010). Two areas could be robustly discriminated in our recording: the dorsolateral oscillatory region (DLOR, \( n = 1778 \) sites, Figure 3A) and the ventromedial non-oscillatory region (VMNR, \( n = 3002 \) sites, Figure 3B). Figures 3C,D show the distribution of the oscil scores in the DLOR and VMNR, respectively. As expected, significantly

![Figure 2](image-url)

**FIGURE 2 | Synchronization found only between sites in the DLOR. (A)** Substantial coherence at beta frequency range exists between sites within the DLOR (from the STN entrance to the DLOR/VMNR border). However, the relative coherence is around zero in the beta range for pairs recording from the VMNR (B), or between sites of the DLOR and VMNR (C). HMM was used to automatically define the DLOR-VMNR border. The first row is calculated using the HMM defined border, and lower rows calculated with a progressively increasing gap from the HMM border. \( N \) is the number of pairs, and the shaded areas represent the standard error of the mean (SEM) of the relative coherence function for each population. **(D)** Distribution of oscillations scores along the normalized DLOR depth. For each trajectory the DLOR length was normalized for 0-entry, 1-end of DLOR.
higher oscil score values were observed in the DLOR than in the VMNR.

The division of the STN into the DLOR and VMNR domains enabled testing of the synchronization of STN pairs from the same and different regions. Significant synchronization, mainly in the frequency band of 8–30 Hz was found, but only between pairs in the DLOR itself (DLOR-DLOR, $n = 615$ pairs, Figure 2A, upper subplot). This synchronization was not observed in pairs of electrodes at DLOR and VMNR ($n = 548$ pairs, Figure 2B, upper subplot) or in the VMNR ($n = 1227$ pairs, Figure 2C, upper subplot). This finding is consistent with previous multiple electrode studies of the human Parkinsonian STN (Levy et al., 2000, 2002a,b; Amirnovin et al., 2004; Weinberger et al., 2006, 2009; Alavi et al., 2013; Lourens et al., 2013) which reported coherence between STN oscillations in a small fraction of STN pairs. However, our findings indicated that the topographical location of the STN electrodes affected the probability of finding a correlation between STN sites, and coherence was only and robustly found between DLOR-DLOR multi-unit pairs.

Recent imaging studies (Lambert et al., 2012; Haynes and Haber, 2013) have clarified that the boundaries between the functional subdomains of the STN are fuzzy, and an overlap of motor and non-motor projections can be found in the transition areas between the STN domains. Therefore, the average coherence at the dorsolateral and ventromedial STN was tested with increasing gaps (0.5–2 mm) from the HMM borders. These results are shown in the lower five rows of Figure 2, and reveal a sharpening and increase of the average coherence peak in the STN DLOR when the gap is increased. Similarly, when the oscil scores are calculated along the normalized depth of the DLOR, a gradual decrease in the oscillatory scores is observed as the DLOR lower border is approached (Figure 2D).

The above results were obtained by averaging over pairs recorded for different durations. The average coherence results (Figure 2) was further compared to the average coherence results of the same pairs with homogenous intervals (only the first 10 s of each recording was included, and recordings with durations shorter than 10 s were excluded). Similar results (data not shown) were obtained.

**SYNCHRONIZATION vs. OSCILLATIONS IN THE DLOR AREA**

Next, correlation between the oscillations and synchronization in the STN was analyzed. The oscillation and synchronization strengths were calculated using the oscil and sync scores for each pair in the DLOR area. Figures 4A,B depict three examples of power spectrum and coherence function of STN activity with their relative oscil and sync scores, respectively. See also Figure 1 for an example of a simultaneous recording of two
sites in the STN, and their corresponding values of oscil and sync scores.

Figure 5B depicts the scatter plot of the oscil and sync scores for all DLOR pairs. Different indicators for the oscillation strength of pairs of STN sites were used: the minimal and maximal oscil score, the arithmetic average of the PSDs, and the geometric mean of oscil scores. In all cases, the scatter plot of the sync score vs. the oscil score of the pairs within the DLOR area (n = 615) indicated a significant correlation (r > 0.24, p < 0.001) between the synchronization and the oscillations. Here (Figure 5B) we show only the data for the arithmetic mean of the oscil scores.

The correlation between the oscillation and synchronization strength could imply that the synchronization pattern was dependent on the homogeneity of the neuronal oscillations within the DLOR. If the neural oscillations in different sites of the STN of a single patient have a very stable and equal frequency, the existence of synchronization may not be the result of physical coupling between the STN neurons. Therefore, the DLOR pairs of each trajectory were randomly shuffled and the synchronization between the shuffled (non-simultaneously recorded) pairs was re-quantified in each trajectory. After shuffling, the oscillations remained in the same frequency band, but the synchronization was no longer apparent. Figures 5A, C show the average coherence functions before and after shuffling of the DLOR-DLOR pairs (n = 615), respectively. Figures 6A, B depict the average sync and oscil scores before and after shuffling in the STN DLOR and the VMNR. As expected, shuffling had no significant effect on the oscil score in either area (oscillation is a property of a single element and therefore should not be affected by the shuffling procedure). However, the average sync score of the DLOR pairs, but not the VMNR pairs declined significantly after the shuffling procedure (Figure 6A). Finally, Figure 5D depicts the scatter plot of the sync and oscil scores of the shuffled pairs within the DLOR. The Spearman correlation between the sync score and oscil score dropped dramatically from r1 = 0.34 to r2 = 0.05 (p < 0.001 for the null assumption that r1 = r2). The mean PSD estimate for the average oscil score (as in Figure 5B) was used for this analysis. Similar results were obtained for the other indicators of oscillation strength of the STN pairs.

**COHERENCE IS MAINLY IN THE BETA FREQUENCY RANGE**

Next, the frequency value where each spectrum (Figure 7A) and coherence (Figure 7C) reached its maximal value was calculated. In both cases, a bi-modal distribution was observed, with a dominance of tremor frequency (3–7 Hz) and beta (12–30 Hz) oscillations, for the auto-spectrums and the coherence functions, respectively. Figures 7B, D show the scatterplot of the maximal oscil and sync scores, respectively, as a function of their frequency. While the oscil scores had similar values in the beta and the tremor range, the values of the maximal sync scores were much higher in the beta than in the tremor range. These results are in line with our previous primate studies (Raz et al., 2000) that revealed
FIGURE 7 | Dominance of coherence at the beta frequency range vs. tremor frequency and beta oscillations in the power spectrums in the STN DLOR. (A) Frequency distribution of the frequencies with maximal power in the power spectrums of STN DLOR recording sites. (B) Scatter plot of oscillations (oscil) scores vs. the frequency with maximal power. (C,D) show the frequency distribution and scatter plot, respectively, for the coherence, same conventions as in (A,B). $N = 1230$ single DLOR sites and 615 DLOR-DLOR pairs.

LACK OF POSITIVE CORRELATION BETWEEN THE STN OSCIL AND SYNC SCORES vs. PD SYMPTOMS

Previous studies have suggested that STN oscillations and synchronization are correlated with tremor in PD patients (Levy et al., 2002a). This would indicate that the STN synchronized oscillations are driven by the tremor (which may be generated by an independent neuronal loop). The above findings of robust synchronization in the beta range may suggest that STN synchronization is an independent phenomenon and not a mere reflection of neuronal oscillations at similar frequencies. However, we cannot rule out the possibility that the 10 Hz activity in this study is not tremor related and a harmonic feature (or n:m locking) of the tremor or of the neuronal oscillations at the tremor frequency.

THE STN SPIKING POPULATION ACTIVITY IS SYNCHRONIZED

In this study population spiking (multi-unit) activity was used as a measure of the spiking activity of the STN rather than the more classical parameter of single unit activity (Perkel et al., 1967; Abeles, 1982; Lemon, 1984; Eggermont, 1990). This was primarily for practical reasons. The goal of physiological recording in the operating room (OR) is to enable better identification of the borders of the subthalamic nucleus and its sub-regions. The electrode is therefore advanced in 100 μm steps rather than 2–5 μm steps as is customary in the research laboratory setup. The sampling duration at each step is also limited (Shamir et al., 2012) and the OR conditions often do not allow stable recordings (as compared to 30–90 min stable recordings in a research setting). On the other hand, the cross-correlation of composite spike trains derived from several un-discriminated cells recorded on a single electrode (multi-unit activity) enhances the sensitivity of correlation methods. First, the higher discharge rate of multi vs. single unit recording reduces the asymmetric sensitivity of correlation methods to excitation vs. inhibition (Aertsen and Gerstein, 1985). Second, multi-unit cross-correlation can be a more sensitive detector of a neuronal relationship than single-unit cross-correlation (Bedenbaugh and Gerstein, 1997). Thus, use of a multi-unit signal is warranted for both practical and theoretical
reasons. Furthermore, the use of signals recorded by two different electrodes in this study reveal the long range (2 mm) synchronization of STN DLOR. It is hoped that future studies of STN units using objective metrics for quantification of the quality of the unit isolation (Joshua et al., 2007; Hill et al., 2011) will shed more light on synchronization in the STN and other basal ganglia structures of human patients.

SYNCHRONIZATION ONLY OCCURS BETWEEN DLOR PAIRS IN THE STN

Early studies described neuronal synchronization in the STN as an epiphenomenon found mainly in patients presenting with tremor (Levy et al., 2000, 2002a). More recent studies (Hanson et al., 2012; Alavi et al., 2013; Lourens et al., 2013) have reported that synchronization can be found between some but not all STN pairs. On the other hand, beta-band LFP and spike oscillations have been described as a consistent feature of human PD in the dopamine depleted state (Brown and Williams, 2005; Foffani et al., 2005; Little et al., 2012). Moreover, many studies have documented the consistency of beta-band oscillations in both the spatial and temporal domains (Bronte-Stewart et al., 2009; de-Solages et al., 2010; Zaidel et al., 2010; Abosch et al., 2012; Little et al., 2012). In this study, synchronization within the Parkinsonian STN DLOR was indeed found to correlate with oscillations. However, the shuffling procedure revealed that STN synchronization was not due to independent oscillators with a similar oscillation frequency (Strogatz, 2003). If this had been the case, a significant synchronization should also have been observed between the shuffled (non-simultaneously recorded pairs of the same patient) DLOR-DLOR pairs. Thus, the synchronization of the simultaneously recorded STN pairs probably reflects the increased coupling between these neurons in the dopamine depleted state of Parkinson’s disease. This increased coupling is probably due to the increased efficacy of the common discharge rate and pattern, synchronization within the STN may be another pathophysiological marker of Parkinson’s disease. The potential consequences of synchronization (as opposed to other attributes like rate and pattern change) are probably mainly due to reduced information capacity of the basal ganglia neurons. However, the different pathological changes in the parkinsonian basal ganglia are probably not mutually exclusive. Synchronized oscillations have stronger effects than less synchronized oscillations and completely unsynchronized oscillations might have no effect on target neurons. Furthermore, future studies toward adaptive DBS (Rosin et al., 2011) should investigate which of the pathophysiological changes in the STN activity might be used as the optimal trigger for closed loop DBS.

ACKNOWLEDGMENTS

This work was supported partially by the Vorst Family Foundation for Parkinson Research, by the Simone and Bernard Guttmann chair of Brain Research, and the generous support of the Rosetrees and Dekker foundations (to Hagai Bergman) as well as by the ELSC post-doctorate fellowships to Shay Moshe and Reuben R. Shamir and the PATH and Bloom foundations of London to Zvi Israel. We would like to thank Prof. Y. Ritov for statistical advice and Dr. S. Freeman and E. Singer for help in scientific editing.

AUTHOR CONTRIBUTIONS

Shay Moshe and Reuben R. Shamir claim for equal contribution. Shay Moshe did the data analysis, and wrote the manuscript together with Hagai Bergman and Zvi Israel. Reuben R. Shamir handled the database and collected part of the data. AR initiated the study and helped in the collection of data. Hagai Bergman, Renana Eitan, Fernando R. de Noriega, and Zvi Israel collected the data. All authors discussed the results, reviewed the manuscript, and made their comments.

REFERENCES

Abeles, M. (1982). Local Cortical Circuits. Berlin; Heidelberg; New York: Springer-Verlag. doi: 10.1007/978-3-642-81708-3
Abosch, A., Lancitn, D., Onaran, L., Eberly, L., Spaniol, M., and Ince, N. F. (2012). Long-term recordings of local field potentials from implanted deep brain stimulation electrodes. Neurosurgery 71, 804–814. doi: 10.1227/NEU.0b013e3182676891
Aertsen, A. M., and Gerstein, G. L. (1985). Evaluation of neuronal connectivity: sensitivity of cross-correlation. Brain Res. 340, 341–354. doi: 10.1016/0006-8993(85)90933-X
Alavi, M., Dostrovsky, J. O., Hodaie, M., Lozano, A. M., and Hutchison, W. D. (2013). Spatial extent of beta oscillatory activity in and below the subthalamic nucleus and substantia nigra pars reticulata of Parkinson’s disease patients. Exp. Neurol. 245, 60–71. doi: 10.1016/j.expneurol.2012.09.021
Alvarez, I., Macias, R., Pavon, N., Lopez, G., Rodriguez-Oroz, M. C., Rodriguez, R., et al. (2009). Therapeutic efficacy of unilateral subthalamicotomy in Parkinson’s disease: results in 89 patients followed for up to 36 months. J. Neurol. Neurosurg. Psychiatry 80, 979–985. doi: 10.1136/jnnp.2008.154948
Amirnovin, R., Williams, Z. M., Cosgrove, G. R., and Eskandar, E. N. (2004). Visually guided movements suppress subthalamic oscillations in Parkinson’s disease patients. J. Neurosci. 24, 11302–11306. doi: 10.1523/JNEUROSCI.3242-04.2004
Bedenbaugh, P., and Gerstein, G. L. (1997). Multiunit normalized cross correlation differs from the average single-unit normalized correlation. *Neural Comput.* 9, 1265–1275. doi: 10.1162/neco.1997.9.6.1265

Beltitsi, A., Panzeri, S., Magri, C., Logothetis, N. K., and Kayser, C. (2010). Sensory information in local field potentials and spikes from visual and auditory cortices: time scales and frequency bands. *J. Comput. Neurosci.* 29, 533–545. doi: 10.1007/s10827-010-0230-y

Benabid, A. L., Pollak, P., Gross, C., Hoffmann, D., Benazzouz, A., Gao, D. M., et al. (1994). Acute and long-term effects of subthalamic nucleus stimulation in Parkinson's disease. *Stereact.* Funct. Neurosurg. 62, 76–84. doi: 10.1159/000086960

Benazzouz, A., Gross, C., Feger, J., Boraud, T., and Bioulac, B. (1993). Reversal of rigidity and improvement in motor performance by subthalamic high-frequency stimulation in MPTP-treated monkeys. *Eur. J. Neurosci.* 5, 382–389. doi: 10.1111/j.1460-9568.1993.tb00505.x

Bergman, H., Wichmann, T., and DeLong, M. R. (1990). Reversal of experimental parkinsonism by lesions of the subthalamic nucleus. *Science* 249, 1436–1438. doi: 10.1126/science.2402638

Bergman, H., Wichmann, T., Karmon, B., and DeLong, M. R. (1994). The primate subthalamic nucleus. II. Neuronal activity in the MPTP model of parkinsonism. *Brain* 117, 1597–1620. doi: 10.1093/brain/117.5.1597

Bronstein, J. M., Tagliati, M., Alterman, R. L., Lozano, A. M., Volkman, J., Stefani, A., et al. (2011). Deep brain stimulation for Parkinson disease: an expert consensus and review of key issues. *Arch. Neurol.* 68, 165. doi: 10.1001/archneur.2010.260

Bronte-Stewart, H., Barberini, C., Koop, M. M., Hill, B. C., Henderson, J. M., and Wingeier, B. (2009). The STN beta-band profile in Parkinson's disease is stationary and shows prolonged attenuation after deep brain stimulation. *Exp. Neurol.* 215, 20–28. doi: 10.1016/j.expneurol.2008.09.008

Brown, P., and Williams, D. (2005). Basal ganglia local field potential activity: Character and functional significance in the human. *Clin. Neurophysiol* 116, 2510–2519. doi: 10.1016/j.clinph.2005.05.009

Buzsaki, G., Anastassiou, C. A., and Koch, C. (2012). The origin of extracellular fields and currents - EEG, ECoG, LFP and spikes. *Nat. Rev. Neurosci.* 13, 407–420. doi: 10.1038/nrn3241

Chen, C. C., Hsu, Y. T., Chan, H. L., Chiou, S. M., Tu, P. H., Lee, S. T., et al. (2010). Complexity of subthalamic 13–35 Hz oscillatory activity directly correlates with clinical impairment in patients with Parkinson's disease. *Exp. Neurol.* 224, 234–240. doi: 10.1016/j.expneurol.2010.03.015

Danish, S. F., Moyer, J. T., Finkel, L. H., Baltuch, G. H., Jaggi, L. J., Priori, A., et al. (2007). High-frequency oscillations (>200 Hz) in the human non-parkinsonian subthalamic nucleus. *Brain Res. Bull.* 74, 84–90. doi: 10.1016/j.brainresbull.2007.05.006

de-Solages, C., Hill, B. C., Koop, M. M., Henderson, J. M., and Bronte-Stewart, H. (2010). Bilateral symmetry and coherence of subthalamic nuclei beta band activity in Parkinson's disease. *Exp. Neurol.* 221–226. doi: 10.1016/j.expneurol.2009.11.012

de-Solages, C., Hill, B. C., Yu, H., Henderson, J. M., and Bronte-Stewart, H. (2011). Maximal subthalamic beta hypersynchrony of the local field potential in Parkinson's disease is located in the central region of the nucleus. *J. Neurosci. Neurosurg. Psychiatry* 82, 1387–1389. doi: 10.1136/jnp.2010.223107

Eggermont, J. J. (1990). *The Correlative Brain. Theory and Experiment in Neuronal Interaction.* Berlin: Springer-Verlag. doi: 10.1007/978-3-642-51033-5

Farmer, S. F. (1995). A framework for the analysis of mixed time series/point process data–theory and application to the study of physiological tremor, single motor unit discharges and electromyograms. *Prog. Biophys. Mol. Biol.* 64, 257–278. doi: 10.1016/S0140-6736(99)00264-X

Goldberg, J. A., Arató, S., Boraud, T., Haber, S. N., Vaadia, E., and Bergman, H. (2002). Enhanced synchrony among primary motor cortex neurons in the MPTP primate model of Parkinson's disease. *J. Neurosci.* 22, 4639–4653.

Guo, S., Zhuang, P., Hallett, M., Zheng, Z., Zhang, Y., Li, J., et al. (2013). Subthalamic deep brain stimulation for Parkinson's disease: correlation between locations of oscillatory activity and optimal site of stimulation. *Parkinsonism Relat. Disord.* 19, 109–114. doi: 10.1016/j.parkreldis.2012.08.005

Halliday, D. M., and Farmer, S. F. (2010). On the need for rectification of surface EMG. *J. Neurophysiol.* 103, 3547–3549. doi: 10.1152/jn.00222.2010

Hamani, C., Saint-Cyr, J. A., Fraser, J., Kaplitt, M., and Lozano, A. M. (2004). The subthalamic nucleus in the context of movement disorders. *Brain* 127, 4–20. doi: 10.1093/brain/awt029

Hammond, C., Bergman, H., and Brown, P. (2007). Pathological synchronization in Parkinson's disease: networks, models and treatments. *Trends Neurosci.* 30, 357–364. doi: 10.1016/j.tins.2007.05.004

Hanson, T. L., Fuller, A. M., Lebedev, M. A., Turner, D. A., and Nicolelis, M. A. (2012). Subcortical neuronal ensembles: an analysis of motor task association, tremor, oscillations, and synchrony in human patients. *J. Neurosci.* 32, 8620–8632. doi: 10.1523/JNEUROSCI.0750-12.2012

Haynes, W. J., and Haber, S. N. (2013). The organization of prefrontal-subthalamic inputs in primates provides an anatomical substrate for both functional specificity and integration: implications for Basal Ganglia models and deep brain stimulation. *J. Neurosci.* 33, 4804–4814. doi: 10.1523/JNEUROSCI.4674-12.2013

Heimer, G., Bar-Gad, I., Goldberg, J. A., and Bergman, H. (2002). Dopamine replacement therapy reverses abnormal synchronization of pallidal neurons in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine primate model ofparkinsonism. *J. Neurosci.* 22, 7850–7855.

Follett, K. A., Weaver, F. M., Stern, M., Hur, K., Harris, C. L., Luo, P., et al. (2010). Pallidal versus subthalamic deep-brain stimulation for Parkinson's disease. *N. Engl. J. Med.* 362, 2077–2091. doi: 10.1056/NEJMoa1007083

Gerstein, G. L. (2000). Cross-correlation measures of unresolved multi-neuron recordings. *J. Neurosci. Methods* 100, 41–51. doi: 10.1016/S0165-0270(00)00226-0

Giannicola, G., Marceglia, S., Rossi, L., Mrakic-Sposta, S., Rampini, P., Tamma, F., et al. (2010). The effects of levodopa and ongoing deep brain stimulation on subthalamic beta oscillations in Parkinson's disease. *Exp. Neurol.* 226, 120–127. doi: 10.1016/j.expneurol.2010.08.011

Goldberg, J. A., Maraton, S., Boraud, T., Haber, S. N., Vaadia, E., and Bergman, H. (2002). Enhanced synchrony among primary motor cortex neurons in the MPTP primate model of Parkinson's disease. *J. Neurosci.* 22, 4639–4653.

Kuhn, A. A., Trottenberg, T., Kivi, A., Kupsch, A., Schneider, G. H., and Brown, P. (2005). The relationship between local field potential and neuronal discharge in...
the subthalamic nucleus of patients with Parkinson's disease. Exp. Neurol. 194, 212–220. doi: 10.1016/j.expneurol.2005.02.010
Kuhn, A. A., Tsui, A., Aizir, T., Ray, N., Brucke, C., Kupsch, A., et al. (2009). Pathological synchronisation in the subthalamic nucleus of patients with Parkinson's disease relates to both bradykinesia and rigidity. Exp. Neurol. 215, 380–387. doi: 10.1016/j.expneurol.2008.11.008
Lambert, C., Zrinzo, L., Nagy, Z., Lutti, A., Hariz, M., Foltynie, T., et al. (2012). Confirmation of functional zones within the human subthalamic nucleus: patterns of connectivity and sub-parcellation using diffusion weighted imaging. Neuroimage 60, 83–94. doi: 10.1016/j.neuroimage.2011.11.082
Lemon, R. (1984). Methods for Neuronal Recording in Conscious Animals. New York, NY: John Wiley & Sons.
Levy, R., Hutchison, W. D., Lozano, A. M., and Dostrovsky, J. O. (2002a). Synchronized neuronal discharge in the basal ganglia of parkinsonian patients is limited to oscillatory activity. J. Neurosci. 22, 2855–2861.
Levy, R., Ashby, P., Hutchison, W. D., Lang, A. E., Lozano, A. M., and Dostrovsky, J. O. (2002b). Dependence of subthalamic nucleus oscillations on movement and dopamine in Parkinson’s disease. Brain 125, 1196–1209. doi: 10.1093/brain/awf128
Levy, R., Hutchison, W. D., Lozano, A. M., and Dostrovsky, J. O. (2000). High-frequency synchronization of neuronal activity in the subthalamic nucleus of parkinsonian patients with limb tremor. J. Neurosci. 20, 7766–7775.
Lewicki, M. S. (1998). A review of methods for spike sorting: the detection and classification of neural action potentials. Network 9, R53–R87. doi: 10.1088/0954-898X/9/4/001
Lhommeé, E., Klinger, H., Thobois, S., Schmitt, E., Ardouin, C., Bichon, A., et al. (2012). Synchronized neuronal discharge in the basal ganglia of parkinsonian patients. Mov. Disord. 27, 841–853. doi: 10.1002/mds.25027-5
Linn, N., Moshel et al. Subthalamic synchronized oscillations in the primate basal ganglia. J. Neurophysiol. 74, 1800–1805.
Nambu, A. (2004). A new dynamic model of the cortico-basal ganglia loop. Prog. Brain Res. 143, 461–466. doi: 10.1016/S0079-1213(03)43043-4
Nambu, A., Tokuno, H., and Takada, M. (2002). Functional significance of the cortico-subthalamic-pallidal ‘hyperdirect’ pathway. Neuropsi. Rev. 43, 111–117. doi: 10.1016/S0168-0102(02)00027-5
Nini, A., Feingold, A., Slovin, H., and Bergman, H. (1995). Neurons in the globus pallidus do not show correlated activity in the normal monkey, but phase-locked oscillations appear in the MPTP model of parkinsonism. J. Neurophysiol. 74, 1800–1805.
Odekerken, V. I., van, L. T., Staal, M. J., Mosch, A., Hoffmann, C. F., Nijssen, P. C., et al. (2013). Subthalamic nucleus versus globus pallidus bilateral deep brain stimulation for advanced Parkinson’s disease (NSTAPs study): a randomised controlled trial. Lancet Neurol. 12, 37–44. doi: 10.1016/S1474-4422(12)70264-8
Parent, A., Sato, F., Wu, Y., Gauthier, J., Levesque, M., and Parent, M. (2000). Organization of the basal ganglia: the importance of axonal collateralization. Trends Neurosci. 23, S20–S27. doi: 10.1016/S1474-4422(00)00022-7
Parent, M., and Parent, A. (2007). The microcircuity of primate subthalamic nucleus. Parkinsonism Relat. Disord. 13(Suppl. 3), S92–S295. doi: 10.1016/j.parkreldis.2007.07.008
Perkel, D. H., Gerstein, G. L., and Moore, G. P. (1967). Neuronal spike trains and stochastic point processes. II. Simultaneous spike trains. Biophys. J. 7, 419–440. doi: 10.1016/S0006-3495(67)86597-4
Plenz, D., and Kitai, S. T. (1999). A basal ganglia pacemaker formed by the subthalamic nucleus and external globus pallidus. Nature 400, 677–682. doi: 10.1038/23281
Pollak, P., Benabid, A. L., Gross, C., Gao, D. M., Laurent, A., Benazzouz, A., et al. (1993). Effects of the stimulation of the subthalamic nucleus in Parkinson disease. Rev. Neurol. Paris 149, 175–176.
Raz, A., Feingold, A., Zelanskiaya, V., Vaadia, E., and Bergman, H. (1996). Neuronal synchronization of tonically active neurons in the striatum of normal and parkinsonian primates. J. Neurophysiol. 76, 2083–2088.
Raz, A., Frechter-Mazar, V., Feingold, A., Abeles, M., Vaadia, E., and Bergman, H. (2001). Activity of pallidal and striatal tonically active neurons is correlated in mptp-treated monkeys but not in normal monkeys. J. Neurosci. 21, RC128.
Raz, A., Vaadia, E., and Bergman, H. (2000). Firing patterns and correlations of spontaneous discharge of pallidal neurons in the normal and the tremulous 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine vervet model of parkinsonism. J. Neurosci. 20, 8559–8571.
Rodriguez-Oroz, M. C., Rodriguez, M., Guridi, J., Mewes, K., Chockkman, V., Vitek, J., et al. (2001). The subthalamic nucleus in Parkinson's disease: somatotopic organization and physiological characteristics. Brain Res. 913, 1777–1790. doi: 10.1016/j.brainres.2007.05.054
Rossi, M., Giannicola, G., Servello, D., Marceglia, S., Pacchetti, C., Porta, M., et al. (2011). Subthalamic local field beta oscillations during ongoing deep brain stimulation in Parkinson’s disease in hyperacute and chronic phases. Neurosignals 19, 151–162. doi: 10.1159/000328508
Rosin, B., Slovik, M., Mitelman, R., Rivlin-Etzion, M., Haber, S. N., Israel, Z., et al. (2011). Closed-loop deep brain stimulation is superior in ameliorating parkinsonism. Parkinson. 72, 370–384. doi: 10.1016/j.parkreldis.2011.08.023
Schuepbach, W. M., Rau, I., Knudsen, K., Volkman, J., Krack, P., Timmermann, L., et al. (2013). Neurostimulation for Parkinson’s disease with early motor complications. N. Engl. J. Med. 368, 610–622. doi: 10.1056/NEJMoai1201585
Seifried, C., Weise, L., Hartmann, R., Gasser, T., Baudrexel, S., Szelenyi, A., et al. (2012). Intraoperative microelectrode recording for the delineation of subthalamic nucleus topography in Parkinson’s disease. Brain Stimul. 5, 378–387. doi: 10.1016/j.brainstim.2011.06.002
Shamir, R. R., Zaidel, A., Joskowicz, L., Bergman, H., and Israel, Z. (2012). Microelectrode recording duration and spatial density determine the detection accuracy of the subthalamic nucleus. Stereotact. Funct. Neurosurg. 90, 325–334. doi: 10.1055/s-0033-1238252
Strogatz, S. (2003). Sync: The Emerging Science of Spontaneous Order. London: Penguin books.
Tachibana, Y., Iwamuro, H., Kita, H., Takada, M., and Nambu, A. (2011). Subthalamo-pallidal interactions underlying parkinsonian neuronal oscillations in the primate basal ganglia. Exp. J. Neurosci. 34, 1470–1484. doi: 10.1111/j.1460-9568.2011.07865.x
Weaver, F. M., Follett, K., Stern, M., Hur, K., Harris, C., Marks, W. J., Jr. et al. (2009). Bilateral deep brain stimulation vs best medical therapy for patients with Parkinson's disease. Mov. Disord. 22, 2855–2861.
advanced Parkinson disease: a randomized controlled trial. JAMA 301, 63–73. doi: 10.1001/jama.2008.929

Weinberger, M., Hutchison, W. D., Lozano, A. M., Hodaie, M., and Dostrovsky, J. O. (2009). Increased gamma oscillatory activity in the subthalamic nucleus during tremor in Parkinson’s disease patients. J. Neurophysiol. 101, 789–802. doi: 10.1152/jn.90837.2008

Weinberger, M., Mahant, N., Hutchison, W. D., Lozano, A. M., Moro, E., Hodaie, M., et al. (2006). Beta oscillatory activity in the subthalamic nucleus and its relation to dopaminergic response in Parkinson’s disease. J. Neurophysiol. 96, 3248–3256. doi: 10.1152/jn.00697.2006

Williams, A., Gill, S., Varma, T., Jenkinson, C., Quinn, N., Mitchell, R., et al. (2010). Deep brain stimulation plus best medical therapy versus best medical therapy alone for advanced Parkinson’s disease (PD SURG trial): a randomised, open-label trial. Lancet Neurol. 9, 581–591. doi: 10.1016/S1474-4422(10)70093-4

Yuval-Greenberg, S., Tomer, O., Keren, A. S., Nelken, I., and Deouell, L. Y. (2008). Transient induced gamma-band response in EEG as a manifestation of miniature saccades. Neuron 58, 429–441. doi: 10.1016/j.neuron.2008.03.027

Zaidel, A., Spivak, A., Grieb, B., Bergman, H., and Israel, Z. (2010). Subthalamic span of beta oscillations predicts deep brain stimulation efficacy for patients with Parkinson’s disease. Brain 133, 2007–2021. doi: 10.1093/brain/awq144

Zaidel, A., Spivak, A., Shpigelman, L., Bergman, H., and Israel, Z. (2009). Delimiting subterritories of the human subthalamic nucleus by means of microelectrode recordings and a Hidden Markov Model. Mov. Disord. 24, 1785–1793. doi: 10.1002/mds.22674

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 10 July 2013; accepted: 18 October 2013; published online: 19 November 2013.

Citation: Moshel S, Shamir RR, Raz A, de Noriega FR, Eitan R, Bergman H and Israel Z (2013) Subthalamic nucleus long-range synchronization—an independent hallmark of human Parkinson’s disease. Front. Syst. Neurosci. 7:79. doi: 10.3389/fnsys.2013.00079

This article was submitted to the journal Frontiers in Systems Neuroscience.

Copyright © 2013 Moshel, Shamir, Raz, de Noriega, Eitan, Bergman and Israel. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.