MASTIC-G-POLY (ACRYLAMIDE): MICROWAVE-ASSISTED SYNTHESIS AND CHARACTERISATION

MOHIT KUMAR*, SURAJPAL

School of Pharmaceutical Sciences, Lovely Professional University Phagwara, Jalandhar, Punjab, India
Email: mohitmehta9@gmail.com

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ABSTRACT

Objective: The objective of the present investigation was the synthesis of grafted co-polymer gum mastic using acrylamide as the monomer.

Methods: 3-factor 3-level response surface Box-Behnken design, which requires 15 runs including three replicates of the central run, was used for the synthesis of graft copolymers of mastic gum with acrylamide using ceric ammonium nitrate as the free radical initiator. The critical synthesis and process parameters; CSPP (A = concentration of monomer, B = concentration of initiator and C= Temperature) to generate design space and optimize formulation with an aim to obtain critical quality attributes (CQA, Y1 =% Yield, Y2 =% Grafting, Y3 =% grafting efficiency).

Result: Formulation F14 having a maximum % yield of 75.89% with % grafting of 210% and % grafting efficiency 51.57% was selected as best.

Conclusion: The microwave-assisted grafted mastic gum was prepared successfully and optimized by using Box Behnken design.

Keywords: Mastic Gum, Microwave-assisted grafting, Acrylamide, Box Behnken design

INTRODUCTION

Polymeric materials are widely used in pharmaceutical formulations, especially for making modified release formulations. Their specialty of controlling drug release rate is because of their structure and composition. These are long-chain compounds formed by monomers. These chains can be linear, branched and cross-linked [1, 2].

Grafted co-polymer includes a previously formed polymer backbone onto which the other species of polymer chains, which are of varying chemical nature, are attached at different sites of the polymeric backbone. The connected side chains may be comprising of a monomeric unit or of a binate mix. The one which is having one monomer only is easier to synthesize and generally happens in a solitary step, nevertheless grafting in the case of binate blend requires to be done in continuous and stepwise addition of the monomers [3-5].

A grafted co-polymer shows macromolecular series with single or multiple types of block molecule series associated with core polymeric backbone chain as various side chains. Graft polymerization is one of the most convenient methods to use various natural polysaccharides in extended drug delivery systems [6, 7]. Nowadays in polymer science the synthesis and use of these grafted copolymers in various applications is a major area for researchers. Employment of a peculiar or selected functional groups into the polymer affects the various chemical, physical as well as rheological properties of the compound.

In comparison with other Asian countries, India is a rich source for these because of the inherent biodiversity [8-10].

The aim of the study was the development of a modified polymer from a natural gum by using a monomer and initiator.

MATERIALS AND METHODS

Materials

Mastic gum was obtained from Mastic grower association (Greece), acrylamide was purchased from Fisher Scientific Pvt. Ltd. (India), ceric ammonium nitrate was purchased from Qualikem Laboratories (India). Ethanol, petroleum ether and methanol were purchased from Merck, Mumbai, India.

Method

Grafting copolymerization of mastic gum

Mastic-g-poly(acrylamide) was prepared by redox initiator method. An accurately weighed 1.4 gm-2.8 gm of acrylamide was solubilized in 30 ml of ethanol in a beaker and in another beaker, 0.5 gm of fine powder of mastic gum was dissolved in 10 ml of ethanol. Both these above solutions were mixed and an initiator i.e. ceric ammonium nitrate was added. Above solutions were mixed and an initiator i.e. ceric ammonium nitrate was added.

The critical synthesis and process parameters; CSPP (A = concentration of monomer, B = concentration of initiator and C= Temperature) to generate design space and optimize formulation with an aim to obtain critical quality attributes (CQA, Y1 =% Yield, Y2 =% Grafting, Y3 =% grafting efficiency). Different concentration of three CSPP (as shown in table 1) viz., A = polyacrylamide with their low (1.4 g), medium (2.1 g) and high level (2.8 g) and B = ceric ammonium nitrate with their low (25 mg), medium (50 mg) and high level (75 mg) and Temperature low (40 °C), medium (50 °C) and high level (60 °C) were selected. Box behnken design was applied using design expert software [13, 14] (version 7.0, Stat-ease, Inc. USA) and the polynomial equation generated was given below:

\[ Y = b_0 + b_1A + b_2B + b_3C + b_4AB + b_5AC + b_6BC + b_7A^2 + b_8B^2 + b_9C^2 \]

Where Y is the dependent variable, \( b_0 \) is the arithmetic mean response of the 15 runs and \( b_1, b_2, b_3, b_4, b_5, b_6, b_7, b_8, b_9 \) is the estimated coefficient for the corresponding factor A, B and C, which represents the average results of changing one factor at a time from its low to high value. The interaction term (A x B, A x C and B x C) depicts the changes in the response when three factors are simultaneously changed. The polynomial terms A^2, B^2 and C^2 are included to investigate the quadratic model. The magnitude of coefficients in the polynomial equation has either a positive sign indicating a synergistic effect or a negative sign indicating an antagonistic effect. Best fitting experimental model (linear, two factor interaction and quadratic) was taken statistically on the basis of comparison of
several statistical parameters like coefficient of variation (CV), multiple correlation coefficient ($R^2$), adjusted multiple correlation coefficient (adjusted $R^2$), predicted residual sum of square and graphically by Contour Plot, 3D response surface plot provided by Design Expert software. The level of significance was considered at a p-value<0.05. The box Behnken design matrix is shown in table 2.

### Table 1: Levels of independent variables in box behnken design

| Factor | Name             | Units | Low level | Mid-level | High level |
|--------|------------------|-------|-----------|-----------|------------|
| A      | Acrylamide amount| gm    | 1.4       | 2.1       | 2.8        |
| B      | CAN amount       | mg    | 25        | 50        | 75         |
| C      | Temperature      | °C    | 40        | 50        | 60         |

FT-IR spectroscopy

The grafting on the polymer backbone was confirmed by FT-IR (Nicolet) and infrared spectra of native and grafted mastic gum and acrylamide were recorded in a range from 4000 to 500 cm$^{-1}$[15].

Differential scanning calorimetry

Differential scanning calorimetry (DSC) was used for screening. The specified samples were hermetically sealed in a flat-bottomed aluminum pan and heated in the differential scanning calorimeter (DSC Mettler Toledo, Mumbai, India) in an atmosphere of nitrogen and the rate of flow was 25 ml/min. A temperature range of 25 °C to 220 °C was used, and the heating rate was 10 °C/min [16, 17].

X-ray diffractometry

X-ray diffraction analysis of grafted and native mastic gum and acrylamide samples was done in powder form by means of X-ray diffractometer (Ultima-4, Rigaku Company, Japan) using K-beta filter, CuKa radiation, the voltage of 40 kV and a current of 30 mA. The scanning employed was over the 0.00 to 60.00 ° diffraction angles [18].

Scanning electron microscopy

Scanning electron micrographs of mastic Gum and grafted gum samples were taken by using a SEM (JEOL, JSM-6100) [19].

RESULTS AND DISCUSSION

Formulation and optimization of mastic-g-poly (acrylamide)

The dried mastic gum was used for the purpose of grafting of gum by following steps as shown in fig. 1.

In the present study, the optimization of microwave-assisted graft copolymerization of acrylamide on mastic gum has been explored by using 3-factor 3-level response surface Box-Behnken design. Levels of Independent Variables in Box Behnken Design are shown in table 1. The experimental design and result of experimental data are given in table 2.

### Table 2: Box behnken design matrix

| Formulation code | Acrylamide amount (gm) | Initiator amount (mg) | Temperature (°C) | % Yield | % Grafting | % Grafting efficiency |
|------------------|------------------------|-----------------------|------------------|---------|-----------|----------------------|
| F1               | 2.1                    | 50                    | 50               | 50.56  | 168       | 32.30                |
| F2               | 1.4                    | 75                    | 50               | 57.01  | 125.2     | 32.94                |
| F3               | 2.1                    | 50                    | 40               | 52.07  | 176       | 33.84                |
| F4               | 1.4                    | 50                    | 50               | 60.51  | 136       | 35.79                |
| F5               | 1.4                    | 25                    | 50               | 55.16  | 112.4     | 28.57                |
| F6               | 2.8                    | 50                    | 50               | 50.66  | 242       | 34.66                |
| F7               | 2.8                    | 50                    | 60               | 48.47  | 224.8     | 34.06                |
| F8               | 2.1                    | 75                    | 40               | 28.03  | 50        | 9.61                 |
| F9               | 2.1                    | 25                    | 60               | 55.24  | 192.8     | 37.07                |
| F10              | 2.1                    | 50                    | 50               | 52.05  | 125.2     | 32.94                |
| F11              | 2.1                    | 25                    | 40               | 22.09  | 16        | 3.07                 |
| F12              | 2.8                    | 50                    | 40               | 48.62  | 225.8     | 34.21                |
| F13              | 2.1                    | 75                    | 60               | 41.71  | 213.2     | 23.69                |
| F14              | 1.4                    | 50                    | 60               | 75.89  | 210       | 51.58                |
| F15              | 2.8                    | 25                    | 50               | 28.66  | 90.6      | 13.73                |

The model proposed the following equations for % Yield response, % Grafting response and % Grafting efficiency response respectively

\[
\%\text{Yield} (\%Y) = 52.66 - 90.1A + 593.5B + 5.85C + 5.84A - B - 3.8B - A + 2.54B + C + 12.35A^2 - 17.8B^2 - 59.8C^2
\]

\[
\%\text{Grafting} (\%G) = 178.93 + 24.95A + 36.92B + 19.52C + 34.65AB - 18.75AC + 15.82BC + 54.48A^2 - 90.36B^2 - 34.36C^2
\]

\[
\%\text{Grafting Efficiency} (\%GE) = 34.41 + 6.02A + 6.31B + 3.95C + 4.14AB - 3.98AC + 3.03BC + 10.01A^2 - 17.74B^2 - 6.31C^2
\]

Where A is the amount of acrylamide, B is the amount of initiator and C is temperature.

The above polynomial equations explain the positive or negative effects of various independent variables on % yield, % grafting and
The amount of acrylamide showed a positive effect on % grafting while the negative effect on %yield and % grafting efficiency. The amount of initiator (ceric ammonium nitrate) showed a positive effect on all three parameters. The temperature has also positive effect on all three parameters. The interaction effect A, B and B, C was found to be synergistic, while the effect A, C was found to be negative. Similarly, the effect A² was found to be more significant than B² and C². In the above polynomial equations, the magnitude of coefficients revealed that the concentration of initiator (B) had maximum effect on all the three parameters [13, 14].

The response surface plots show the effect of different independent variables on percentage yield, percentage grafting and percentage grafting efficiency (fig. 2 to fig. 10). It was observed that the concentration of acrylamide has no significant effect on percentage yield, percentage grafting and percentage grafting efficiency however, temperature and initiator concentration have a significant effect up to a level.

Fig. 2: 3D Surface response graph between acrylamide amount (A) and CAN amount (B) with % yield (Y₁)

Fig. 3: 3D surface response graph between acrylamide amount (A) and temperature (C) with % yield (Y₁)

Fig. 4: 3D surface response graph between initiator amount (B) and temperature (C) with % yield (Y₁)

Fig. 5: 3D surface response graph between acrylamide amount (A) and CAN amount (B) with % grafting (Y₂)

Fig. 6: 3D surface response graph between acrylamide amount (A) and temperature (C) with % grafting (Y₂)

Fig. 7: 3D Surface response graph between CAN amount (B) and temperature (C) with % grafting (Y₂)

Fig. 8: 3D surface response graph between acrylamide amount (A) and CAN amount (B) with % grafting efficiency (Y₃)
Formulation characterization

Optimized Batch (F14) of Acrylamide grafted mastic gum obtained from numerical optimization tool was subjected to FTIR, DSC, XRD and SEM studies.

The FTIR spectra (fig. 11) of mastic gum showed the characteristic peak of Mastic Gum. The spectrum showed a C-H stretch at 2926 cm⁻¹, C=O stretch at 1209 cm⁻¹, =C-H bend at 929 cm⁻¹. The FTIR spectrum of acrylamide is shown in fig. 12. The spectra of acrylamide presented absorption bands at 3340 cm⁻¹ and 3200 cm⁻¹ due to asymmetric and symmetric NH stretching of the NH group. CO stretching appeared at 1647 cm⁻¹. The spectra also showed a band at 1300 cm⁻¹ which can be attributed to CN stretching, while the CH stretching appeared at 1600 cm⁻¹. The FTIR spectrum of Mastic-g-polyacrylamide is shown in fig. 13. The FTIR spectra of grafted gum showed a broad absorption band at 2926 cm⁻¹ due to overlap of the OH stretching band of mastic gum and NH stretching 3391 cm⁻¹ band of acrylamide. Literature supports the above FTIR studies [20, 21].

Selection of optimized batch

The optimized batch of formulation was selected by using desirability, which was calculated by using box behnken design in design expert software. F14 was selected as an optimized formulation with % yield of 75.89744, % grafting of 210 and % grafting efficiency of 51.5789.
The fig. 14, fig. 15 and fig. 16 contains the DSC thermogram of Gum mastic, acrylamide and grafted mastic gum. An exothermic peak was observed at 254.01 °C of the grafted mastic gum. The increase in the glass transition of grafted gum is to be expected because the amine group in polyacrylamide chains grafted onto gum has more difficulty to interact. The GM-g-Polyacrylamide is expected to have longer acryl chains than Gum and acrylamide, thus more glass transition temperature [22, 23].
The XRD spectra (fig. 17) showed the amorphous nature of mastic gum as no characteristic peaks in the spectrum were observed while the diffractogram of acrylamide (fig. 18) showed the crystalline nature. The XRD spectrum of grafted gum (fig. 19) showed the characteristic peaks of acrylamide but with the decreased intensity, which confirmed the formation of graft copolymer [24, 25].

The SEM studies of Mastic Gum (fig. 20) and Mastic-g-Poly (Acrylamide) (fig. 21) showed a change in surface morphology after grafting. The more roughness and unevenness on the surface of grafted gum due to the deposition of acrylamide was clearly visible [26, 27].

CONCLUSION

Polymeric materials, because of its structure and composition, are generally used for making modified release formulations. A grafted
co-polymer shows macromolecular series with single or multiple types of block molecule series associated with core polymeric backbone chain as various side chains. Graft polymerization is a convenient way to use natural polysaccharides like gum mastic for making modified release formulation. Nowadays in polymer science the synthesis and use of these grafted copolymers in various applications is a major area for researchers, in summary, we prepared grafted co-polymer by redox initiator method, which can be used further for the sustained release formulation.

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AUTHORS CONTRIBUTIONS

Authors declare that the work done by the names mentioned in the article and all the liabilities and claims related to the content of the article will be borne by the authors.

CONFLICT OF INTERESTS

The authors report no conflict of interest. The authors alone are responsible for the content and writing of this article.

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