Cleavage by Granzyme B Is Strongly Predictive of Autoantigen Status: Implications for Initiation of Autoimmunity

By Livia Casciola-Rosen,* Felipe Andrade,† Danielle Ulanet,‡ Wes Bang Wong,‡ and Antony Rosen†§

Summary

Systemic autoimmune diseases are a genetically complex, heterogeneous group of disorders in which the immune system targets a diverse but highly specific group of intracellular autoantigens. The molecules targeted are not unified by common structure, function, or distribution in control cells but become clustered and concentrated in surface blebs when cells undergo apoptosis. We show here that the majority of autoantigens targeted across the spectrum of human systemic autoimmune diseases are efficiently cleaved by granzyme B in vitro and during cytotoxic lymphocyte granule-induced death, generating unique fragments not observed during any other form of apoptosis. These molecules are not cleaved by caspase-8, although this protease has a very similar specificity to granzyme B. The granzyme B cleavage sites in autoantigens contain amino acids in the P$_2$ and P$_3$ positions that are preferred by granzyme B but are not tolerated by caspase-8. In contrast to autoantigens, nonautoantigens are either not cleaved by granzyme B or are cleaved to generate fragments identical to those formed in other forms of apoptosis. The striking ability of granzyme B to generate unique fragments is therefore an exclusive property of autoantigens and unifies the majority of molecules targeted in this spectrum of diseases. These results focus attention on the role of the cytotoxic lymphocyte granule-induced death pathway in the initiation and propagation of systemic autoimmunity.

Key words: apoptosis • cytotoxic T lymphocyte • protease • caspase

The highly specific autoantibody response in systemic autoimmune diseases generally predicts the biologic phenotype of the disease, making autoantibodies diagnostically useful (1, 2). Although molecules targeted by the immune system in these diseases are exceptionally diverse in terms of structure, function, and subcellular distribution in healthy cells, they are strikingly redistributed in apoptotic cells, becoming clustered and concentrated in two populations of surface structures on the dying cell (3). It has been proposed that cleavage of proteins might play a role in selecting molecules as autoantigens, perhaps by cleavage-induced revelation of cryptic epitopes (4, 5). In this regard, several of the clustered antigens are substrates for caspases during apoptosis. These include poly (ADP-ribose) polymerase (PARP),¹ U 1-70kD, the catalytic subunit of DNA-dependent protein kinase (DNA-PK$_c$), the nuclear mitotic apparatus protein (NuMA), nucleolus organizer region (NOR)-90, fodorin, topoisomerase I, signal recognition particle (SRP)-72, and lamin B (6–10). However, the autoantigen fragments produced by caspases during all forms of apoptotic death described to date (including thymocyte apoptosis) are all identical, making it likely that caspase-derived fragments have previously been generated during apoptosis and tolerized centrally and peripherally (11, 12). Furthermore, there are several autoantigens (fibrillarin, centromere protein (CENP)-B, Ku, Ro, and La) that are not cleaved by caspases during apoptosis, indicating that susceptibility to cleavage by caspases is not a general feature of autoantigens (for review see reference 13).

Granzyme B is a serine protease found in the cytoplasmic granules of CTLs and NK cells that plays an important role in inducing apoptotic changes in target cells during granule exocytosis-induced cytotoxicity (14–16). This function is achieved partly by catalyzing the cleavage and activation of several caspases, as well as through caspase-independent pathways (17–20). We recently demonstrated that gran-
zyme B (which shares with caspases a requirement for aspartic acid in the substrate P₁ position) efficiently cleaves three caspase-3 substrates (DNA-PK₂, NuMA, and PARP), generating novel fragments not generated during any other form of cell death (21). To determine whether the generation of unique autoantigen fragments by granzyme B was a universal feature of autoantigens, we assessed whether a wide range of autoantigens was susceptible to cleavage by granzyme B in vitro and in vivo. These studies demonstrate that despite their diverse structure, distribution, and function, the majority of autoantigens in systemic autoimmune diseases are efficiently cleaved by granzyme B, generating unique fragments. In contrast, granzyme B either does not cleave or does not generate unique fragments in all of the nonautoantigen molecules that we tested. Granzyme B cleavage sites in autoantigens were defined; in all cases, the tetrapeptide sequence immediately adjacent to the cleavage site was highly conserved. Cleavage by granzyme B therefore (a) unites these otherwise unrelated molecules and (b) generates unique fragments of these antigens, strongly suggesting that this protease plays a mechanistic role in selecting the molecules against which autoimmune responses are initiated. These results highlight a potential role for the CTL granule-induced death pathway in initiation and propagation of autoimmunity.

Materials and Methods

Isolation of YT cell granule contents (GC) and purification of granzyme B was as described (21). Purified caspase-3 and -8 and CrmA (cytokine response modifier 1) were gifts from Nanny Thornberry (Merck Research Labs, Rahway, NJ). cDNAs for CENP-B, fibrillarin, topoisomerase I, and post meiotic segregation (PM/S1) were gifts from Drs. Ann Pluta (University of Maryland, Baltimore, MD), John Aris (University of Florida, Gainesville, FL), Barbara White (University of Maryland, Baltimore, MD), and Bert Vogelstein (Johns Hopkins University), respectively. Autoantibodies to PMS1 and PMS2 were gifts from Drs. Ann Pluta (University of Maryland, Baltimore, MD), and Bert Vogelstein (Johns Hopkins University), respectively. Salmon sperm recognizing ribosomal protein P, Ku-80, or mAb to Ki-67 (Sigma Chemical Co.). Proteins were detected using horseradish peroxidase–labeled secondary antibodies and chemiluminescence (Pierce Chemical Co.).

In Vitro Cleavage of Endogenous A Uantoigens in Hela Cells. Hela lysates were prepared as described (23), except that DTT was omitted from the lysis buffer. To inactivate endogenous caspase activities, 1 mM IAA was added to the lysates (see Fig. 9, all lanes except 3 and 6) and incubated for 15 min at 4°C. 5 mM DTT was then added in the absence or presence of 50 nM caspase-8, and the reactions were incubated for 1 h at 37°C. Before using the “IAA poisoning protocol” in the caspase-8 experiments, the validity of this experimental approach to inactivate endogenous caspases present in lysates while still permitting exogenously added caspases to cleave was tested. As PARP and U1-70kD are well characterized caspase-3 substrates (generating signature fragments of 89 and 40 kD, respectively (23)), their ability to be cleaved by caspase-3 in IAA-treated lysates was used in the following defined test system: (i) HeLa lysates containing 0.8 mM caspase-3 and 5 mM DTT were incubated for 1 h at 37°C; the extent of PARP cleavage (66%) and U1-70kD (25%) cleavage obtained under these conditions represented maximum possible caspase-3 activity. (ii) 1 mM IAA was added to HeLa lysates prepared in the absence of DTT. After incubating for 15 min at 4°C, 0.8 nM caspase-3 was added and the lysates were further incubated (1 h at 37°C); PARP and U1-70kD were not cleaved under these conditions. (iii) HeLa lysates were incubated as in (ii), except that 5 mM DTT was added simultaneously with caspase-3 after the IAA poisoning step. Both PARP and U1-70kD were cleaved (the amounts of cleavage were 80 and 70%, respectively, of that noted in approach [i]), confirming that exogenous caspases are active in these lysates in which endogenous caspases have previously been inactivated if subsequently added with DTT. Abolition of the contribution of endogenous caspases under these circumstances accounts for the decreased total substrate cleavage observed.

In Vitro Cleavage of [35S]Methionine-Labeled A Uantoigens Substrates Generated by In Vitro Transcription/Translation. [35S]Methionine-labeled CENP-B, fibrillarin, PM/S1, and PM/S2 were generated by coupled in vitro transcription/translation (23). Cleavage reactions were performed in buffer A consisting of 10 mM Hepes, pH 7.4, 2 mM EDTA, and 1% NP-40. (5 mM DTT was added to the reactions containing caspases). Reactions contained the amounts of caspase-3 and granzyme B indicated in the figure legends and were incubated at 37°C for either 15 or 60 min, as specified. After terminating the reactions by adding gel buffer and boiling, samples were electrophoresed on 10% SDS-PAGE (CENP-B, PM/S1, and PM/S2) or 12% SDS-PAGE (fibrillarin), and their fragments were visualized by fluorography.

In Vitro Cleavage of [35S]Methionine-Sulfone-Labeled Endogenous A Uantoigens Substrates After Immunoprecipitation With Patient Sera. HeLa cells were labeled for 2 h with [35S]methionine/cysteine, and the lysates were immunoprecipitated with patient sera recognizing histidyl tRNA synthetase, RNA polymerase II large subunit, PM/S1, or alanyl tRNA synthetase, followed by protein A–agarose. The beads, containing washed, radiolabeled endogenous proteins were resuspended in buffer A in the presence or
absence of granzyme B and incubated for 15 min at 37°C. The following amounts of granzyme B were used: 46 nM histidyl and alanyl tRNA synthetases and 23 nM RNA polymerase II and PM Sci. Samples were electrophoresed on 10% SDS-PAGE, and radiolabeled proteins and their fragments were visualized by fluorography.

In vitro cleavage of endogenous and purified Nontargeted substrates. HeLa lysates were incubated with granzyme B for 1 h at 37°C and then electrophoresed on 8 or 10% SDS-PAGE and immunoblotted with a rabbit polyclonal antibody to Cdc2p34 (Santa Cruz Biotechnology) or mAbs to vinculin or β-tubulin (Sigma Chemical Co.). Cleavage of purified human lactoferrin, apotransferrin, and thrombin (all from Sigma Chemical Co.) was performed by incubating 20 μg of each substrate in the absence or presence of 30 nM granzyme B for 60 min at 37°C. The reactions were terminated by adding gel application buffer, and the samples were electrophoresed on 10% SDS-PAGE and visualized by Coomassie blue staining. Similar amounts of purified La were well cleaved under identical conditions (data not shown).

Calculation of catalytic constant (k_cat/K_m) values. k_cat/K_m values were determined as described (21, 23) using endogenous substrates in cell lysates, radiolabeled endogenous immunoprecipitated substrates, or radiolabeled substrates generated by coupled in vitro transcription/translation. The percent cleavage of each substrate was determined by densitometry; these values were fit to the first order rate equation,

\[
\frac{\% \text{ substrate cleavage}}{100} = 1 - e^{-k_{cat} \left( E/K_m \right) t}
\]

to calculate k_cat/K_m. For several of the autoantigens listed in Table I, k_cat/K_m values were obtained using two or all three of these methods, and equivalent results were obtained (reference 21; data not shown).

Cleavage of endogenous substrates after in vitro incubation of Fas-negative K562 CEs with YTC cell GC or Lymphokine-activated Killer CEs. K562 cells were incubated with YT cell GC as described (21), except that the incubations were done in HBSS containing 10 mM Hepes, pH 7.4. Cells were incubated for 2 h at 37°C in the absence or presence of added GC, before terminating reactions by adding 2× SDS-gel buffer directly to cell suspensions. Similar experiments were performed using cultured HeLa cells, human umbilical vein endothelial cells, human myoblasts, and Jurkat cells with identical results (data not shown). LAK cells were prepared and incubated with K562 target cells at a 3:1 ratio in the absence or presence of 100 μM Ac-DEVD-CHO or 50 μM Z-IETD-fluoromethylketone (FMK; Calbiochem Corp.) as described (21). Incubation of either LAK or K562 cells individually with these inhibitors did not affect the autoantigens analyzed (data not shown).

Confirmation of granzyme B cleavage sites by mutagenesis of P1 Asp residues. CDNA clones encoding fibrillarlin, La, U1-70kD, M1-2, PM S1, PM S2, and topoisomerase I were used as templates for mutagenesis by overlap-extension PCR to generate clones containing putative granzyme B site P1 Asp→Ala substitutions (21). [35S]Methionine-labeled polypeptides were generated by coupled in vitro transcription/translation and used as substrates for granzyme B cleavage as described above.

### Results

Granzyme B Specifically and Efficiently Cleave Numerous Autoantigens Targeted in Systemic Autoimmune Diseases. We initially determined whether and how efficiently a variety of well-defined autoantigens across the spectrum of systemic autoimmune diseases were cleaved by purified granzyme B (Table I). We used autoantibodies of known specificity to immunoblot lysates of HeLa cells that had been incubated in vitro with or without granzyme B. Lysates were pre-treated with IAA (which covalently modifies the active site cysteine of caspases) to prevent endogenous caspase activity. Interestingly, several autoantigens that have previously been shown to be cleaved by caspases during apoptosis were also efficiently cleaved by granzyme B, generating unique fragments in every case. These substrates included U1-70kD, topoisomerase I, SR P-72, PAR P (Fig. 1), and N O R-90 (data not shown). We also identified M1-2, K1-67, PM S1, PM S2, and La as additional autoantigens that are cleaved by both caspases and granzyme B; again, distinct

| No. | Autoantigen | Cleavage site | k_cat/K_m |
|-----|-------------|---------------|-----------|
| 1   | DNA-PKcs    | VGPD2698-F    | 2.5 × 10^6 |
| 2   | Topoisomerase I | IED15-F | 1.6 × 10^6 |
| 3   | Nu MA       | VATD1705-A    | 5.4 × 10^5 |
| 4   | Mi-2        | VDPD1312-Y    | 8.5 × 10^4 |
| 5   | La          | LEED220-A     | 6.1 × 10^4 |
| 6   | PM S1       | ISAD496-E     | 6.9 × 10^4 |
| 7   | Fibrillarlin | VGPD188-G     | 3.3 × 10^4 |
| 8   | PAR P       | VDPD538-S     | 2.3 × 10^4 |
| 9   | U1-70kD     | LGND409-S     | 1.3 × 10^4 |
| 10  | PM S2       | VEKD493-S     | 1.4 × 10^4 |
| 11  | Isoleucyl tRNA synthetase | VTPD563-Q | 7.8 × 10^4 |
| 12  | Histidyl tRNA synthetase | LGPD48-E | 2.3 × 10^4 |
| 13  | Alanyl tRNA synthetase | VAPD632-R | 1.8 × 10^4 |
| 14  | RNA polymerase I | ICPD448-M | 1.3 × 10^4 |
| 15  | Ki-67       | VCTD1481-K    | 8.1 × 10^3 |
| 16  | PM ScI      | VEQD253-M     | 7.5 × 10^3 |
| 17  | CENP-B      | VSDD536-E     | 5.9 × 10^3 |
| 18  | RNA polymerase II | ITPD470-P | ND |
| 19  | SR P-72     | VTPD573-P     | ND |
| 20  | Ku-70       | ISSD79-R      | ND |
| 21  | N O R-90    | VR PD220-A    | ND |

Catalytic constant (k_cat/K_m) values were determined as described in Materials and Methods using defined amounts of purified granzyme B to cleave either radiolabeled substrates expressed by in vitro transcription/translation, endogenous substrates in cell lysates, or radiolabeled endogenous immunoprecipitated substrates. Likely granzyme B cleavage sites were predicted using fragment sizes and the known cleavage specificity of granzyme B (29, 30). For autoantigens 1-10, the predicted cleavage sites were confirmed by showing that molecules having P1Asp→Ala substitutions were no longer cleaved by granzyme B (see Fig. 8).
fragments were generated by the two proteases (Figs. 1, 2, and 8). Whenever possible, efficient cleavage by granzyme B and generation of novel fragments was confirmed using both endogenous substrates in lysates (detection by immunoblotting) and in vitro–translated substrates (detection by fluoro- 
graphy; this was done for U1-70kD, PARP, NuMA, topoisomerase I, La, PM S1, and Mi-2). The $k_{\text{cat}}/K_m$ values varied between $1.3 \times 10^4$ M$^{-1}$s$^{-1}$ (U1-70kD) and $1.6 \times 10^8$ M$^{-1}$s$^{-1}$ (topoisomerase I) (Table I). Several substrates were cleaved by granzyme B at multiple sites (e.g., Mi-2, K i-67, PM S1, and PM S2; Figs. 1 and 2), generating a complex profile of cleavage fragments.

Previous studies have identified several autoantigens that are not susceptible to cleavage by caspases during apoptosis (7, 9). Using the immunoblotting system described above and/or cleavage assays using in vitro–translated substrates, we demonstrated that many of these autoantigens are efficiently cleaved by granzyme B. These molecules included fibrillarin, Ku-70, and RNA polymerase II large subunit. (Figs. 2 and 3; see Fig. 5). As some well-defined autoantibodies do not recognize their antigens by immunoblotting, we also addressed the susceptibility of radiolabeled endoge- 
nous substrates to cleavage by granzyme B in vitro. To perform these studies, HeLa cells were radiolabeled with $[35S]$methionine/cysteine, and proteins were immunoprecipi-
tated using human autoantibodies. Protein A–agarose beads containing washed, precipitated proteins were resus-
pended in buffer supporting the activity of granzyme B and incubated with or absence of added purified granzyme B. To confirm the validity of this approach, we tested several different autoantigens known to be cleaved by granzyme B, as well as several autoantigens and nonau-
toantigens that are not cleaved by granzyme B. The cleavage profiles obtained after incubating granzyme B with (a) lysates (followed by detection with immunoblotting) or (b) immunoprecipitated radiolabeled antigens (followed by detection with fluoro- 
graphy) were compared. Identical results were obtained using these two methods for cleaved autoantigens (topoisomerase I, Mi-2, RNA polymerase II large subunit, Ku-70, PARP, La, and NOR-90), uncleaved autoantigens (Ku-80, R o-60kD), and control sub-
strates (B-tubulin, vinculin). Using this immunoprecipita-
tion approach, we demonstrated that several additional autoantigens (PM Scl, RNA polymerase I and II large subunits, histidyl tRNA synthetase, isoleucyl tRNA synthetase, and alanyl tRNA synthetase) were indeed cleaved by granzyme B, generating unique fragments (Table I; Fig. 3 contains representative examples). Using identical immuno-
precipitation conditions, these substrates were not cleaved by caspase-3, whereas Mi-2 cleavage by caspase-3 occurred normally (data not shown). Several autoantigens were not susceptible to cleavage by either caspases or granzyme B (Fig. 4 and references 7, 9, 24). These included R o-52kD and -60kD, ribosomal protein P, histones, Sm proteins, threonyl tRNA synthetase, glycy1 tRNA synthetase, and Ku-80.

Susceptibility to cleavage by granzyme B was a highly specific feature of autoantigens; nonautoantigens were ei-
Figure 2. [35S]methionine-labeled autoantigen substrates generated by in vitro transcription/translation are cleaved by purified granzyme B. [35S]methionine-labeled CENP-B, fibrillarin, PSMA, and PSM2 were incubated for 60 min at 37°C with the following amounts of caspase-3: 1 nM (PSM2) or 2.5 nM (CENP-B, fibrillarin, and PSMA). Separate incubations were performed as follows with granzyme B: 62.5 nM, 15 min (fibrillarin, PSMA); 42 nM, 60 min (CENP-B); and 28 nM, 60 min (PSM2). The samples were electrophoresed and visualized as described in Materials and Methods. Open arrowheads denote intact antigens (CENP-B, fibrillarin, PMS1); 42 nM, 60 min (CENP-B); and 28 nM, 60 min (PSM2). Solid arrowheads mark the granzyme B–specific fragments (CENP-B, 58 and 40 Kd; fibrillarin, 37 Kd; PMS1, 110 Kd; PMS2, 105 Kd). Solid arrowheads mark the granzyme B–specific fragments (CENP-B, 58 and 40 Kd; fibrillarin, 17 Kd; PSM1, 50 and 60 Kd; PSM2, 60, 50, and 36 Kd).

Figure 3. Immunoprecipitated endogenous autoantigens are cleaved by purified granzyme B. [35S]methionine/cysteine-labeled endogenous histidyl tRNA synthetase, RNAP1 (large subunit), RNAP2 (small subunit), and PMScl were immunoprecipitated and cleaved by granzyme B as described in Materials and Methods. Solid arrowheads denote granzyme B–specific fragments (histidyl tRNA synthetase, 40 Kd; RNAP1 (large subunit, 190, 110, and 92 Kd; RNAP2 (small subunit, 85 and 74 Kd; alanyl tRNA synthetase, 58 Kd).
fold more efficient than cleavage by caspase-3, and granzyme B–specific fragments are the most prominent (Fig. 6, lane 3; note that in lanes 3 and 4, the majority of topoisomerase is in the 98-kD granzyme B–cleaved form). Where the efficiency of substrate cleavage by granzyme B and caspase-3 are similar (e.g., DNA-PK, and M-i-2), both caspase- and granzyme B–specific fragments are generated. Inhibition of caspases abolishes the caspase-specific fragments only. In contrast, where substrates are cleaved ~200-fold more efficiently by caspase-3 than by granzyme B (PARP, U1-70kD), no granzyme B–specific fragments were observed in the intact cell killing assay unless caspases were inhibited by adding Ac-DEVD-CHO (Fig. 6; compare lanes 3 and 4).

Production of unique autoantigen fragments in the LAK cell assay was decreased by inhibitors of granzyme B (28). Thus, 50 μM of Z-IETD-FMK (Fig. 7 A) greatly diminished cleavage of NuMA, M-i-2, and topoisomerase I, as evidenced by the increased amount of intact antigen detected by blotting (Fig. 7 A; compare lanes 3 and 4). This was accompanied by an abolition of the unique cleavage fragments of NuMA (175 kD) and topoisomerase I (98, 75, and 72 kD) and markedly inhibited the production of granzyme B–specific fragments of M-i-2 (48 kD). This pattern of inhibition correlated well with the efficiency of cleavage of these substrates by granzyme B (Table I). As noted above, production of granzyme B–specific fragments of autoantigens was either unchanged (M-i-2) or enhanced (PARP, U1-70kD, SRP-72) by treatment with the caspase–specific inhibitor, Ac-DEVD-CHO (Fig. 6, lane 4). Similar results were also obtained when 143 nM CrmA was used to inhibit granzyme B activity in cell lysates in which granzyme B–specific fragments of U1-70kD and PARP were abolished (Fig. 7 B), with concomitant recovery of intact antigen. Although both Z-IETD-FMK and CrmA also inhibit group III caspases, unique fragments of these autoantigens are not generated by these proteases (see Fig. 9).

Granzyme B Cleaves Autoantigens at Highly Conserved Sites. The specificity of granzyme B has recently been defined using a positional scanning combinatorial tetrapeptide library (29, 30). The protease has a preference for I, V, or L in P2, E, G, or G in P3, and P, S, N, A, Q, H, T, V, E, or D in P3, with a near absolute preference for D in P1. This specificity and the sizes of the fragments generated by granzyme B cleavage were used to predict cleavage sites. Using site-directed mutagenesis to make a series of P1 Asp→Ala substitutions in several of the granzyme B substrates, we addressed the effects of mutation on the efficiency of cleavage by the protease and thus defined the granzyme B cleavage sites in fibrillarin, M-i-2, topoisomerase I, PM S1, PM S2, La, and U1-70kD (Table I; Fig. 8 contains representative examples). The granzyme B cleavage sites in PARP and DNA-PKcs were defined previously (21, 31). In every case, the cleavage site sequence is in accord with the specificity determined by the combinatorial library. The P2 and P3 residues in these cleavage sites were quite restricted, with prominent representation of P, A, and S in P2, and G, E, T, D, and S in P3. Interestingly, many of these residues are preferred by granzyme B but not by group III caspases (29). Using fragment sizes to predict likely granzyme B cleavage sites in other autoantigens, we readily identified probable cleavage sites in these proteins (Table I). Other than the consensus tetrapeptide sequence preceding the cleavage site in these autoantigens, there were no obvious similarities in primary sequence either upstream or downstream of this site. Interestingly, several granzyme B substrates also have consensus tetrapeptide sequences that are not cleaved, indicating that additional conformational information influences susceptibility to cleavage at these sites.
Autoantigens That Are Cleaved by Granzyme B Are Not Substrates for Caspase-8.

In addition to the granule exocytosis pathway, CTLs can also induce target cell proteolysis and apoptosis through ligation of target cell Fas by CTL Fas ligand (32). As caspase-8 is prominently activated when CTLs induce target cell death through the Fas pathway (33) and because group III caspases have a very similar substrate specificity to granzyme B (29), we determined if caspase-8 could generate the same proteolytic fragments of endogenous autoantigens in cell lysates that are generated by granzyme B. Caspase-8 efficiently activates precursor effector caspases in cell lysates (which in turn efficiently cleave downstream substrates), we first irreversibly inactivated these caspase precursors with IAA. Exogenous caspase-8 was then added in the presence of excess DTT, and substrate cleavage was assayed by immunoblotting. We initially confirmed that caspase-8 was active in the lysate system by using a coupled proteolysis system to demonstrate cleavage of downstream caspase-3 substrates (see Table II).

Table II. Relative Cleavage Efficiencies Determine Ratio of Substrate Fragments Generated In Vivo

| Autoantigen       | $k_{cat}/K_m$ (M$^{-1}$s$^{-1}$) for cleavage | Prominent fragment type generated in: |
|-------------------|---------------------------------------------|---------------------------------------|
|                   | Caspase-3                                  | Granzyme B                            |
| Topoisomerase-I   | $3.4 \times 10^4$                          | $1.6 \times 10^6$                     |
| DNA-PKcs          | $7.5 \times 10^6$                          | $2.5 \times 10^6$                     |
| Mi-2              | $2.0 \times 10^6$                          | $8.5 \times 10^4$                     |
| U1-70kD           | $2.5 \times 10^6$                          | $1.3 \times 10^4$                     |
| PARP              | $5.0 \times 10^6$                          | $2.3 \times 10^4$                     |

Autoantigens that are cleaved by granzyme B with similar or greater efficiency than by caspase-3 give rise to prominent granzyme B fragments in the LAK/K562 system. In this system, granzyme B fragments are not observed when substrates are cleaved less efficiently by granzyme B than by caspase-3, unless caspases are inhibited (see Fig. 6). $k_{cat}/K_m$ values for caspase-3 cleavage of DNA-PKcs, PARP, and U1-70kD are from reference 23, and values for granzyme B cleavage of DNA-PKcs and PARP are from reference 21. The remaining $k_{cat}/K_m$ values were calculated as described in Materials and Methods.

Figure 7. Generation of unique autoantigen fragments is abolished by Z-IETD-FMK or CrmA. Indicator lines denote the positions of intact antigens; solid and open arrowheads denote the granzyme B-specific and caspase-specific fragments, respectively. (A) LAK and K562 cells were preincubated in the absence or presence of 50 μM Z-IETD-FMK and then coincubated (E/T ratio of 3:1) for an additional 4 h (21). The samples were electrophoresed and autoantigens were blotted as described in the Fig. 6 legend. (B) GC were preincubated in the absence or presence of an equimolar amount of CrmA (amounts based on the granzyme B content of GC) for 5 min at 37°C. HeLa lysates were then incubated with these GC for a further 60 min at 37°C (0.5 μl GC per 75 μg lysate) before electrophoresis and immunoblotting.

Figure 8. Granzyme B cleavage of fibrillarin and La is abolished by P1DfiA mutation. Site-directed mutation of aspartic acid residues in the P1 positions of fibrillarin (D$^{184}$→A) and La (D$^{220}$→A) was performed. Wild-type (WT) and mutated products were generated by in vitro transcription/translation before cleavage with granzyme B as described in Materials and Methods. Open and solid arrowheads mark the positions of intact molecules and granzyme B-specific fragments, respectively. Similar results were obtained when P1 aspartic acid residues in the granzyme B cleavage sites of U1-70kD, Mi-2, PM51, PM52, and topoisomerase I were mutated (data not shown).
Susceptibility to Granzyme B Cleavage Predicts Autoantigen Status

**Discussion**

Autoantibodies found in human autoimmune diseases recognize a group of substrates efficiently cleaved by granzyme B during CTL-mediated apoptosis. We have demonstrated that components of 21 out of 29 well-defined autoantigens are susceptible to efficient cleavage by granzyme B, generating unique fragments. Although the cleaved molecules differ markedly in subcellular location, function, and extended primary sequence, they share two features: (a) all are autoantigens targeted by a high titer autoantibody response in human autoimmune diseases, including systemic lupus erythematosus, Sjogren’s syndrome, diffuse and limited scleroderma, and autoimmune myositis, and (b) molecules are unified by containing a granzyme B cleavage site not susceptible to cleavage by caspase-8. The status of a molecule as an autoantigen and its unique susceptibility to cleavage by granzyme B but not by caspase-8 are therefore highly related ($P < 0.0001$, chi-square analysis). The positive predictive value of susceptibility to unique cleavage by granzyme B and status as an autoantigen is 100% for the 48 molecules studied, whereas the negative predictive value is 73%, indicating that additional mechanisms play a role in selection of some molecules as autoantigens. It is noteworthy that most of the uncleaved molecules are nucleoprotein complexes (e.g., components of nucleosomes and small nuclear ribonucleoproteins); we are in the process of defining whether these molecules undergo distinct structural modifications during unique forms of apoptotic cell death. Indeed, there are several other components of CTL granules that might play a role in this regard.

Interestingly, the autoantigens susceptible to cleavage by granzyme B include antigens that are targeted across the spectrum of autoimmune rheumatic diseases. The striking correlation of specific autoantibody response with unique biologic phenotype (e.g., Mi-2 in dermatomyositis and topoisomerase I in diffuse scleroderma) raises the question of how this disease specificity arises if a common mechanism (CTL granule-induced death) is responsible for targeting of this group of molecules. Although this remains unclear, the immunizing tissue and initiating stimulus may play important roles in focusing subsequent, self-sustaining injury.

Several of the cleaved molecules (PARP, U1-70kD, Ku-70, SRP-72, topoisomerase I, DNA-PKcs, Ku-70, SRP-72, topoisomerase I, [Fig. 1 A], DNA-PKcs, NuMA [reference 21], Mi-2, La, PM51, PM52, and NOR-90 [data not shown]) are also cleaved by effector caspases during apoptosis but at different sites in each case. Frequently, the granzyme B and caspase-3 cleavage sites are close to each other in the primary sequence. The striking linkage of susceptibility to cleavage by both caspases and granzyme B in these substrates (but at different sites) suggests that the two different families of apoptotic proteases recognize two distinct structural features of a common motif that have been targeted during independent evolution of the apoptotic cysteine and serine protease families. The likelihood that a functional motif has been targeted by the apoptosis-specific proteases is further underscored by the observation that the granzyme B cleavage sites in several molecules (e.g., fibrillarin, PARP, and PM51) are highly conserved, even in drosophila and yeast. This striking conservation of sequence at granzyme B cleavage sites in organisms in which CTLs have not yet evolved implies that an important, as yet undefined function is served by these regions that is altered by proteolysis. This new, extended family of granzyme B substrates therefore provides a powerful tool with which to explore the evolution and biological functions of the aspartic acid-specific apoptotic proteases and the specific mechanisms underlying CTL granule-induced cell death.

**Figure 9.** Caspase-8 does not cleave endogenous autoantigens that are granzyme B substrates. HeLa lysate (in which endogenous caspases had been inactivated by 1 mM IAA) was incubated with 5 mM DTT in the absence (lanes 1, 4, 7, 9, 11, and 13) or presence (lanes 2, 5, 8, 10, 12, and 14) of 50 nM caspase-8 for 1 h at 37°C. In the lysates shown in lanes 3 and 6, omission of IAA permitted caspase-8 to activate endogenous effector caspases. After terminating the reactions, 75-μg amounts of sample were electrophoresed in each gel lane. Autoantigens were detected by blotting with monospecific patient sera. In lanes 1–6, open and solid arrowheads denote the SDS-PAGE migration positions of the intact antigens and the caspase-8–induced fragments, respectively. Radiolabeled PM51 and PM52 (generated by in vitro transcription/translation) were also not cleaved by purified caspase-8 (data not shown).
demonstrated that DNA-PKcs, NuMA, and PARP are favored when Caspases are inhibited. We have previously observed that members of the group III caspases, which include caspase-8, caspase-10, caspase-11, and caspase-12, are preferentially present in lymphoid and hematopoietic tissues and are involved in the regulation of programmed cell death (apo- tosis) and cell proliferation (21). The recent observation of tumor-specific CTLs in paraneoplastic syndromes (50) suggests that such a mechanism may be more broadly applicable to other autoimmune syndromes.

Caspases are responsible for the proteolytic cleavage of several autoantigens away from P2 and P3 residues that are preferentially captured and presented by dendritic cells (46–49). The resulting immune response is directed against products of CTL granule–induced death, generating an autoamplifying loop that is self-sustaining and leads to the development of autoimmune diseases. The recent observation of tumor-specific CTLs in paraneoplastic syndromes suggests that such a mechanism may be more broadly applicable to other autoimmune syndromes (50).

Generation of Granzyme B–specific Autoantigen Fragments Is Favorable when Caspases Are Inhibited. We have previously demonstrated that DNA-PKcs, NuMA, and PARP are cleaved by both caspase-8 and granzyme B at the same sites, generating identical fragments (33). This marked skewing of granzyme B cleavage sites in autoantigens away from P2 and P3 residues that are preferentially cleaved by both caspase-8 and granzyme B strongly suggests that unique cleavage by granzyme B plays a role in selection of targets in this spectrum of autoimmune disease.

In contrast, the group III caspases prefer tetrapeptide substrates with I, V, or L in P4, E, D, or Q in P3, and H, I, T, W, or V in the P2 position (29). Whereas granzyme B has a similar specificity to the group III caspases, it has a broader substrate specificity in the P1 and P2 positions. Thus, granzyme B will robustly cleave substrates containing G or S in P3 (not tolerated by group III caspases) and prefers P, A, N, and Q in P2 (none of which are tolerated by group III caspases). Interestingly, 10 of 11 proven granzyme B cleavage sites in autoantigens contain residues that are preferred by granzyme B but are poorly tolerated by group III caspases (molecules contain P [7], A [2], or S [1] in the P2 position; four cleavage sites also contain G or S in P3; Table I). Consistent with this observation is the demonstration that none of these substrates could be cleaved by caspase-8 (Fig. 9). In contrast, procaspase-3 and -7, which are not autoantigens, are efficiently cleaved by both caspase-8 and granzyme B at the same sites, generating identical fragments (33). This marked skewing of granzyme B cleavage sites in autoantigens away from P2 and P3 residues that are preferentially cleaved by both caspase-8 and granzyme B strongly suggests that unique cleavage by granzyme B plays a role in selection of targets in this spectrum of autoimmune disease.

Generation of Granzyme B–specific Autoantigen Fragments Is Favorable when Caspases Are Inhibited. We have previously demonstrated that DNA-PKcs, NuMA, and PARP are cleaved by both caspase-8 and granzyme B and that the most prominent fragments generated in lysates and intact cells reflect the relative cleavage efficiencies of the two proteases (21). The studies reported here extend those observations to numerous additional autoantigens (Mi-2, Ki-67, U1-70kD, topoisomerase I, and SR P-72; Fig. 1 A and Fig. 6). Where cleavage of a substrate by granzyme B is equal to or more efficient than that by caspase-8, granzyme B–specific fragments are generated in the LAK/K562 system (e.g., DNA-PKcs Mi-2, and topoisomerase I; Table I). In contrast, where cleavage by granzyme B is less efficient than cleavage by caspases (e.g., PARP, U1-70kD, SR P-72, and topoisomerase I [72- and 74-kD fragments]), effective generation of the granzyme B–specific fragments is only seen in intact cells when caspase activity is inhibited. This observation focuses attention on defining potential immunizing microenvironments and proimmune insults in which such circumstances may arise in vivo. Relevant possibilities include conditions where viral or endogenous caspase inhibitors are expressed (35–40), as well as in long-lived cells or tumor cells that express low levels of specific effector caspases (41, 42).

CTL Granule–induced Death and Autoimmunity. Human systemic autoimmune diseases represent a highly complex disease spectrum, with numerous variables affecting individual susceptibility, initiation, and tissue targets. By demonstrating that the autoantigens targeted across the spectrum of these diseases are unified by their susceptibility to efficient cleavage by granzyme B, with the generation of unique fragments not produced during any other form of cell death, these studies focus attention on the role of the CTL granule pathway in initiation of autoimmunity. Where substrates are cleaved by both caspases and granzyme B, effective generation of unique granzyme B fragments is dependent upon relative inhibition of the caspases (Fig. 6; reference 21). We therefore propose that during proimmune (43) intracellular infections occurring in a microenvironment in which caspase activity is under relative inhibition, production of unique granzyme B fragments is favored. In susceptible individuals in whom clearance of apoptotic material might be impaired (44, 45), suprathermal amounts of these fragments accumulate and are effectively captured and presented by dendritic cells (46–49). The resulting immune response is directed against products of CTL granule–induced death, generating an autoamplifying injury characteristic of these self-sustaining diseases. The recent observation of tumor-specific CTLs in paraneoplastic cerebellar degeneration suggests that such a mechanism may be more broadly applicable to other autoimmune syndromes (50).

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Address correspondence to Antony Rosen, Johns Hopkins University School of Medicine, 720 Rutland Ave., Ross 1059, Baltimore, MD 21205. Phone: 410-955-0139; Fax: 410-955-0964; E-mail: arasen@jhmi.edu

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