Comparison of Placenta PCR and Maternal Serology of Aborted Women for Detection of *Toxoplasma gondii* in Ardabil, Iran

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Abstract: Primary maternal infection with toxoplasmosis during pregnancy is frequently associated with transplacental transmission of the parasite to the fetus. This study was conducted to test the utility of PCR assay to detect recent infections with *Toxoplasma* in aborted women at various gestational ages who referred to Obstetrics and Gynecology Department of Alavi Hospital in Ardabil during 2014 and 2016. Two hundred women with a history of single or repeated abortion were investigated in this study. Blood samples were tested for specific anti-*Toxoplasma* IgM and IgG antibodies by ELISA. According to the results, 53.5% of the women under study were positive for anti-*Toxoplasma* antibodies: 4.0% of them had IgM, 43.0% had IgG, and 6.5% had both IgM and IgG. Subsequently, Nested-PCR analysis was used to detect *T. gondii* DNA in the placenta of subjects. In 10.5% of the women, the results were positive for 529 bp element of *T. gondii*. Among them, 5 (23.8%) cases were IgM positive, 1 (4.8%) case was IgG positive, and 11 (52.4%) were both IgM and IgG positive. In 4 (19.0%) patients, none of the antibodies were found to be positive. In total, 16 patients had positive results in both ELISA and PCR methods, and 174 cases had negative results for new infection. The findings of this study revealed that *T. gondii* might be one of the significant factors leading to abortion, and that the analysis of placenta can be important in order to achieve increased detection sensitivity.

Key words: *Toxoplasma gondii*, abortion, blood, nested-PCR, ELISA, Iran

INTRODUCTION

Toxoplasmosis is a parasitic infection which is induced by *Toxoplasma gondii* and has various clinical symptoms [1]. Infection with this parasite occurs as the result of eating uncooked foods as well as water and vegetables contaminated with cat feces [2]. When a woman eats *Toxoplasma* oocysts or cysts for the first time during pregnancy, tachyzoites spread all over the body through the blood [3]. The fetus becomes infected with the *Toxoplasma* parasite which enters its blood flow via the placenta. The infection of mother before pregnancy rarely puts the fetus in danger unless in patients with immune system deficiencies [4].

Congenital toxoplasmosis generally occurs when the mother gets infected with *Toxoplasma* newly during pregnancy which can lead to various infections in the fetus and infant such as abortion, stillbirth, and live birth of a child with classic symptoms of toxoplasmosis like hydrocephaly, microcephaly, cerebral calcifications, and retinochoroiditis [5,6]. The risk of transfer after primary infection varies from 25% (in the first trimester) to 65% (in the last trimester); however, the younger fetuses are more susceptible to this infection [7].

In Iran, the rate of abortion as the result of toxoplasmosis is unknown. Most of the studies to date about the abortions suspected to be the result of *Toxoplasma* have been conducted based on the serological findings about mothers. In this study, we investigated the use of nested PCR (nPCR) method for the isolation of 529 bp element in the placenta and ELISA method for the detection of anti-*Toxoplasma* antibodies in mothers who had experienced abortion in Ardabil, north-west of Iran.
MATERIALS AND METHODS

In this study, we investigated 200 women who had experienced abortion in different gestational ages and referred to Obstetrics and Gynecology Department of Alavi Hospital in Ardabil during 2014 and 2016. The study protocol was approved by the scientific committee of Ardabil University of Medical Sciences with the approval code of 9206.

Sample collection

Three ml of venous blood was drawn from each patient and their sera were isolated. About 20 g of the placenta samples of the same patients were cut in sterilized conditions and stored together with the serum samples at the temperature of -20˚C until conducting the tests. The sera of all cases were tested for the presence of specific IgM and IgG anti-Toxoplasma antibodies via ELISA kits (Biokit, Barcelona, Spain) according to the manufacturer’s instructions. For each patient, a questionnaire including the mother’s age, gestational age, and the history of prior abortion was completed.

DNA extraction and PCR detection:

*T. gondii* DNA was extracted from the placenta of the aborted women using the QIAamp DNA mini kit (Qiagen, Courtaboeuf, France) according to the manufacturer’s instructions.

Detection procedures sets were used for amplifying fragments of 529 bp element as described by Su et al. [8]. The external primers were Tox-8: GACGTCTGTGTCACGTAGAAAG and Tox-5: CGTCGACACGCTGCACTC CG AT producing an amplified product of 450 bp. Internal primers were Tox-9: AGGAGAGATCATCGCTGACGT AC and Tox-II: GCGTCGTCTCAGTATCG producing an amplified product of 162 bp. The reactions mixture contained 40 µl mixture of 5 µl 10× PCR buffer, 4 µl 25 mM MgCl₂, 4 µl dNTPs (2.5 mM each), 0.2 µl Fast Start Taq (5 U/µl), 0.30 µl external forward primer (50 µM), and 0.30 µl external reverse primer (50 µM) added with 10 µl template DNA. The following conditions were used to amplify the target DNA: one cycle of 5 min initial denaturation at 95˚C followed by 30 cycles at 94˚C for 30 sec, 55˚C for 1 min, and 72˚C for 2 min. Amplification was performed using thermal cycler. All PCR products, regardless of the presence or absence of a visible band were subjected to a second round of PCR.

The nPCR reaction was performed using 5 µl of the first PCR reaction in a mixture containing the inner primers at final concentration of 45 µl mixture of 5 µl 10× PCR buffer, 4 µl 25 mM MgCl₂, 4 µl dNTPs (2.5 mM each), 0.2 µl Fast Start Taq (5 U/µl), 0.30 µl nested forward primer (50 µM), and 0.30 µl nested reverse primer (50 µM). Amplification was carried out at 95˚C for 5 min (one cycle), then followed by 30 cycles at 94˚C for 30 sec, 55˚C for 1 min, and 72˚C for 1 min. The amplification products were detected by gel electrophoresis using 2% agarose gel in 1× Tris–borate–EDTA buffer. DNA bands were visualized in the presence of ultraviolet light following the staining with 0.5% ethidium bromide.

Analysis of data

The minimum and maximum range of *Toxoplasma*-induced abortion prevalence and the level of agreement (kappa statistics, SPSS 16, Chicago, Illinois, USA) were measured to analyze any association between congenital infection by *T. gondii* and the 2 different diagnostic techniques employed [11]. The chi-square test was used to analyze the association between congenital infection and the demographic characteristics of the mothers who had abortion.

RESULTS

Demographic characteristics of mothers who had abortion

The age range of the patients included in the study was 16-41 years and their mean age was 30.5 years. Forty-three subjects were under 20 years old, 88 of them were between 20 and 30 years old, and 69 of them were over 30 years old. According to the data collected, most of the patients (53.0%) were in the first trimester of pregnancy. The patients in the second and third trimester of pregnancy were 31.0% and 16.0%, respectively (Table 1). Moreover, 71.0% of them did not have prior abortion experience. Hypothyroidism, diabetes, and allergies were found respectively in 8 (4.0%), 9 (4.5%), and 5 (2.5%) patients while 178 cases (84.0%) had no history of diseases except miscarriage.

Serology

In this study, samples from the women who had abortion were investigated for the presence of anti-*Toxoplasma* IgG and IgM antibodies using ELISA method. The analysis indicated positive results for anti-*Toxoplasma* antibodies in 53.5% of the patients: 43.0% tested positive for only anti-*Toxoplasma* IgG antibodies, 6.5% tested positive for both anti-*Toxoplasma* IgG and IgM antibodies, and 4.0% indicated positive results for
only anti-Toxoplasma IgM antibodies (Tables 1, 2).

**PCR and nPCR**

In this study, in addition to serological diagnosis method, PCR method was also used to investigate the presence of Toxoplasma DNA in the placenta of the patients under study. All PCR products in the first round were included in the second round of PCR. Using this method, from among the 200 patients under study, Toxoplasma infection was diagnosed in 21 (10.5%) samples based on nPCR (Fig. 1). In 17 of the samples with positive results, both the 450 bp and the 162 bp bands were detectable, but in 4 of them, just the 162 bp band was detected in nPCR. Among the 21 patients for whom nPCR was reported to be positive, 5 (23.8%) cases were IgM positive, 1 (4.8%) was IgG positive, and 11 (52.4%) were both IgM and IgG positive. In 4 (19.0%) patients, none of the antibodies were found to be positive.

**Correlation between demographic characteristics of mothers and infection**

ELISA and nPCR results and data from questionnaires were analyzed employing chi-square statistical test with 95% confidence interval using SPSS software version 16. No significant association was found between the presence of Toxoplasma DNA and mother’s age ($P=0.23$), history of abortion ($P=0.08$), and gestational age ($P=0.16$). There was also no significant association between positive serological samples (IgM positive and both IgM and IgG positive) and mother’s age ($P=0.18$), history of abortion ($P=0.06$), and gestational age ($P=0.15$).

**Agreement between ELISA and nPCR methods**

Among all samples, maternal seroprevalence (anti-Toxoplasma IgM antibodies) was 5.8% to 14.2% (95% confidence interval) and the estimated abortion prevalence associated with T. gondii based on PCR was calculated to be from 6.3% to 14.7% (95% confidence interval). Results show moderate logarithmic agreement between the 2 different tests ($\kappa=0.44$). (A value of kappa equal to +1 implies perfect agreement between the 2 raters, while that of -1 implies perfect disagreement) [9].

**DISCUSSION**

The detection of infective agents in pregnancy is very important since they not only threaten the mother’s health but also cause fetal abnormalities [10]. According to the findings of the

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**Table 1. Demographic characteristics of mothers who had abortion and results of different diagnostic techniques**

|                      | PCR       | ELISA     | Both PCR and ELISA |
|----------------------|-----------|-----------|---------------------|
|                      | Positive  | Negative  | IgM+                | IgG+ | IgG&IgM+ | Ig- Positivea | Negativeb |
| Age groups (year)    |           |           |                     |      |          |              |            |
| ≤ 20                 | 43        | 3         | 2                   | 13   | 3        | 25            | 3           | 23         |
| 20-30                | 89        | 10        | 4                   | 31   | 5        | 48            | 8           | 47         |
| ≥ 30                 | 69        | 7         | 2                   | 42   | 5        | 20            | 6           | 19         |
| Gestational age      |           |           |                     |      |          |              |            |
| 1st trimester        | 106       | 11        | 4                   | 27   | 6        | 69            | 8           | 67         |
| 2th trimester        | 62        | 8         | 4                   | 28   | 6        | 24            | 8           | 22         |
| 3th trimester        | 32        | 2         | 0                   | 31   | 1        | 0             | 1           | 0          |
| History of abortion  |           |           |                     |      |          |              |            |
| Yes                  | 58        | 3         | 0                   | 31   | 2        | 25            | 1           | 23         |
| No                   | 142       | 18        | 8                   | 55   | 11       | 68            | 16          | 66         |

*aPCR positive and ELISA positive (IgG positive+IgM positive+IgG and IgM positive).

*bPCR negative and ELISA negative (Ig negative).*

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**Table 2. ELISA and nested PCR results in samples**

| Methods | nPCR | ELISA     | No. |
|---------|------|-----------|-----|
|         | Positive | Negative |     |
| IgG Positive | 1     | 85       | 86  |
| IgM Positive | 5     | 3        | 8   |
| IgG & IgM Positive | 11    | 2        | 13  |
| Negative  | 4     | 89       | 93  |
| No.      | 21    | 179      | 200 |

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**Fig. 1. Agarose gel electrophoresis of nested PCR products of tissue samples of aborted placenta with the internal 529 bp element primers. Lane M, molecular marker; lane 1, T. gondii positive control (type 1; RH strain); lane 18 negative control; lane 2-17 samples.**
previous studies, mother’s infection in the first and second trimester might lead to the death of fetus in 5% and 2% of the cases, respectively [1-11]. The incidence rate of congenital toxoplasmosis is approximately 1.5 cases per 1,000 live births, so the estimated global incidence of congenital toxoplasmosis is 190,100 cases per year [12].

A series of techniques are required to make a definitive diagnosis in the abortions occurred as the result of toxoplasmosis [13]. Suzuki et al. [14] conducted the evaluation of IgM via isolating the SAG 1 gene of *Toxoplasma* in placenta and concluded that the evaluation of placenta in order to detect *Toxoplasma* genome can be very helpful for the rapid diagnosis of congenital toxoplasmosis.

In 2000, the 200- to 300-fold repetitive 529 bp DNA fragment was identified in *T. gondii* genome and researchers reported high sensitivity and great feature of PCR method in detecting this parasite [15]. In 2003, 529 bp DNA fragment was introduced as a new alternative for 18S ribosomal DNA and B1 targets in detecting *Toxoplasma* DNA in congenital toxoplasmosis [16]. The rate of *Toxoplasma*-induced abortion has been different in studies (18.3% in Egypt [17], 5.3% in Brazil [18], 15.5% in Isfahan, Iran [19], 14.4% in Shiraz, Iran [20], and 11.0% in France [21]. Sarkari et al. [22] analyzed the aborted fetuses for which the PCR test was positive using immunohistochemistry (IHC) method and did not observe any cyst or tachyzoite in them. The reason for this might be the low number of parasites in the aborted tissues.

In our study, a moderate agreement between the 2 different tests was found with 95% confidence interval (κ = 0.44). This finding indicates the need for more research in order to shed light on the methods of infection transfer and also the level of transfer after getting infected.

Although getting serological tests is one of the main diagnostic methods used for the detection of toxoplasmosis, it has many limitations. This method might not be able to detect special anti-*Toxoplasma* antibodies during the active phase of infection since these antibodies might not be produced until several weeks after parasitaemia. Moreover, this method might be unsuccessful in detecting *T. gondii* infection in individuals with immune system deficiencies due to the lack of appropriate antibodies production. Therefore, using PCR method in detecting infection with *T. gondii* obviates the problems encountered when using serological methods, and also when dealing with patients having complicated problems. Thus, the results obtained simultaneously from PCR and ELISA methods can definitively confirm the infection [23].

The presence of *Toxoplasma* DNA in 4.3% of the women for whom the results of serological test were negative might suggest the existence of a new infection at the time of getting samples which shows that the immune system either has not been able to produce enough immunoglobulins to detect it in serology method or suffers from degrees of immunodeficiency. A positive result of a serological test only shows the presence of an infection whereas the direct detection of *T. gondii* in clinical samples can confirm the presence of parasites which themselves can be primary, reactivated, or chronic infections [24]. Our findings indicate that toxoplasmosis is a big danger and a main risk factor in increasing the probability of abortion in pregnant women in this part of Iran. Therefore, we recommend that in order to increase detection sensitivity, molecular test of placenta be conducted for each abortion even if the results of serological tests are negative.

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**CONFLICT OF INTEREST**

We have no conflict of interest related to this study.

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