Investigation of long non-coding RNA expression profiles in patients with post-menopausal osteoporosis by RNA sequencing

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Abstract. The present study aimed to investigate the implication of long non-coding RNA (lncRNA) expression profiles in post-menopausal osteoporosis (PMOP). A total of 10 patients with PMOP and 10 age-matched healthy post-menopausal females as controls were consecutively enrolled. Their peripheral blood mononuclear cells were obtained and lncRNA as well as mRNA expression profiles were detected by RNA sequencing, followed by bioinformatics analyses. The lncRNA expression profiles were able to distinguish patients with PMOP from controls based on principal component analysis and heatmap analysis. In total, 254 upregulated lncRNAs and 359 downregulated lncRNAs were identified in patients with PMOP vs. controls. The top 5 upregulated lncRNAs were RP11-704M14.1, RP11-754N21.1, RP11-408E5.5, ANKRD26P3 and TPTEP1. The top 5 downregulated lncRNAs were RP11-310E22.4, RP11-326K13.4, FABP5P1, SERPINB9P1 and RPL13P2. Based on the interaction of dysregulated lncRNAs and mRNAs by RNA sequencing, functional annotations were then performed. Gene Ontology enrichment analysis revealed that the dysregulated lncRNAs were enriched in terms including apoptotic process and positive regulation of NF-κB transaction, and Kyoto Encyclopedia of Genes and Genomes analysis suggested enrichment in PMOP-associated signaling pathways, including osteoclast differentiation, tumor necrosis factor signaling pathway and mitogen-activated protein kinase signaling pathway. In addition, the regulatory network and circos graph further indicated the implication of lncRNA expression profiles in PMOP via interactions with mRNAs. In conclusion, the present study suggested that aberrant lncRNA expression is deeply involved in the pathogenesis of PMOP by affecting osteoclast differentiation, inflammation and apoptotic processes.

Introduction

Osteoporosis is a common bone disorder characterized by excessive bone resorption and insufficient formation of new bone, leading to reduced bone function and increased risk of fracture (1-3). Among several subtypes of osteoporosis, post-menopausal osteoporosis (PMOP) affects 9-38% of females and accounts for a large proportion of the total osteoporotic population (4,5). PMOP is a chronic disease with high prevalence among post-menopausal females. Therefore, PMOP has been considered a major public health concern during the last decades (6,7).

Long non-coding RNAs (lncRNAs) are evolutionarily conserved molecules that have been extensively studied due to their significant role in various biological processes, including signal transduction, cell growth and apoptosis (8-10). Recently, the expression levels of several specific lncRNAs, including lncRNA DANCR and lncRNA MEG3, were reported to be dysregulated in patients with PMOP and to be associated with disease progression (11,12). Although the majority of previous studies have focused on the function of specific lncRNAs, aberrations in lncRNA expression profiles associated with the pathogenesis of PMOP have been rarely investigated. In a previous study, the lncRNA expression profiles in PMOP were assessed using RNA sequencing and a total of 51 dysregulated lncRNAs were identified in patients with PMOP (13). However, the sample size of the study was relatively small (3 PMOP and 2 control samples), suggesting that the results were sensitive to noise and had low statistical power. Therefore, the lncRNA expression profiles in PMOP should be further investigated. The present study aimed to explore the implication of aberrant lncRNAs in the pathogenesis of PMOP by analyzing expression profiles.

Materials and methods

Patients. A total of 10 patients with PMOP who were treated at the Union Hospital (Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China) between January and May 2018 were enrolled in the present study. The inclusion criteria were as follows: i) Post-menopausal females;
and ii) diagnosis of osteoporosis according to the World Health Organization (WHO) criteria (14). The diagnostic cut-off for osteoporosis, based on WHO guidelines, is defined as the bone mineral density (BMD) T-score being 2.5 standard deviations (SD) below the average value of a young adult, at the femoral neck, total hip or lumbar spine (L1-L4). The exclusion criteria were as follows: i) History of diseases affecting bone metabolism, including osteomalacia, hyperparathyroidism, hyperthyroidism, hypothyroidism, rheumatoid arthritis, hypercortisolism, chronic renal insufficiency and cancer; ii) history of diabetes mellitus; and iii) previous treatment with drugs affecting bone metabolism, including glucocorticoids, estrogens and fluorides. In addition, 10 age-matched healthy females who served as the control group were enrolled in the present study. The inclusion criteria of the control group were: i) Post-menopausal females; and ii) femoral neck, total hip or spinal L1-L4 BMD T-score ≥-1.0 SD. The exclusion criteria were as follows: i) History of diseases affecting bone metabolism, including osteomalacia, hyperparathyroidism, hyperthyroidism, hypothyroidism, rheumatoid arthritis, hypercortisolism, chronic renal insufficiency and cancer; ii) history of diabetes mellitus; and iii) previous treatment with drugs affecting bone metabolism, including glucocorticoids, estrogens and fluorides.

**Samples.** Peripheral whole blood samples were collected from all subjects and the peripheral blood mononuclear cells (PBMCs) were isolated by centrifugation according to standard methods.

**RNA sequencing.** Total RNA was extracted from PBMCs using TRIzol reagent (Invitrogen; Thermo Fisher Scientific, Inc.). Subsequently, the concentration, purity and integrity of the isolated RNA were measured and adjusted. Ribosomal RNA (rRNA) was removed from the total RNA using a Ribo-Zero™ rRNA Removal Kit (Epicentre; Illumina, Inc.) and the rRNA-depleted RNAs were used to construct the sequencing library. The first and second strand of the complementary DNA (cDNA) were synthesized and the library fragments were purified using the AMPure XP system (Beckman Coulter, Inc.). The cDNA fragments, preferentially 150-200 base pairs in length, were selected. The quality of the library was assessed using the Bioanalyzer 2100 system (Agilent Technologies, Inc.). The clustering of index-coded samples was subsequently performed using the HiSeq PE Cluster Kit v4 cBot and the quality control and adapter trimming were then performed following cluster generation. Automated quality control and adapter trimming were then performed using Trim Galore, Cutadapt and FastQC software. Finally, the trimmed reads were mapped to the human genome Hg38 by HISAT2 using the default parameters, followed by mapping of the quality control using the RSeQC software. The read counts of IncRNAs and mRNAs were subsequently calculated using feature Counts based on the annotation file (Homo_sapiens. GRCh38.83.gtf) of the Ensembl database (http://www.ensembl.org).

**Bioinformatics analysis.** IncRNAs and mRNAs that were identified in ≥50% of the samples (at least 10 samples) were subjected to bioinformatics analysis using the R software (version 3.5.3; Lucent Technologies). Subsequently, the following analyses were performed: i) Principal component analysis (PCA) of IncRNA and mRNA expression profiles was performed using the Stats package. ii) Heatmap analysis of IncRNA and mRNA expression profiles was performed using the Pheatmap package. iii) Dysregulated IncRNAs and mRNAs were analyzed using the DeSeq2 package and were visualized in Volcano plots. An adjusted P-value of <0.05 and a fold change (FC) of ≥2.0 according to log_{2}(FC)>1, were considered to indicate a statistically and biologically significant difference, respectively. iv) Heatmap analysis of the dysregulated IncRNAs and mRNAs was performed using the Pheatmap package. v) Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) enrichment analyses of the dysregulated IncRNAs and mRNAs were performed using the DAVID tool. A cut-off criterion of P<0.05 was used to indicate significant differences (significant enrichment by the differentially expressed RNAs, or the co-expressed genes of the core/hub RNAs identified) for the GO terms and KEGG pathways. The top 30 significantly enriched GO terms, including the top 10 terms in the categories biological process, cellular component and molecular function, and the KEGG pathways, were determined in the present study. If <30 significantly enriched terms were obtained, all of them were presented. vi) The gene transcription and regulation data were visualized with a Circos plot using the RCircos package. In the present study, the cis-regulatory IncRNAs were defined as IncRNAs that acted within or overlapped with up to 300 kb of the mRNA gene, while the trans-regulatory IncRNAs were defined as the IncRNA molecules that acted beyond 300 kb in the genomic distance. These definitions were based on the chromosomal association of the IncRNAs with the mRNA molecules. Correlation coefficients of >0.95 were considered to indicate a significant correlation between IncRNAs and mRNAs.

**Results**

**Characteristics.** The mean age was 63.3±5.2 and 61.5±6.6 years and the body mass index (BMI) was 21.1±2.3 and 22.0±2.5 kg/m² for the PMOP patients and controls, respectively (Table I). No significant differences were noted in the parameters age

| Parameter | PMOP (n=10) | Controls (n=10) | P-value |
|-----------|-------------|----------------|---------|
| Age (years) | 63.3±5.2 | 61.5±6.6 | 0.507 |
| BMI (kg/m²) | 21.1±2.3 | 22.0±2.5 | 0.413 |
| BMD-T score | | | |
| Femoral neck | -3.18±0.39 | 0.49±0.61 | <0.001 |
| Hip | -3.03±0.46 | 0.71±0.95 | <0.001 |
| L1-L4 | -3.26±0.42 | 0.79±0.98 | <0.001 |

Values are expressed as the mean ± standard deviation. Statistical comparisons were performed using the t-test. PMOP, post-menopausal osteoporosis; BMI, body mass index; BMD, bone mineral density.
(P=0.507) and BMI (P=0.413) between PMOP patients and control subjects. However, increased BMD-T scores were noted in the femoral neck (-3.18±0.39 vs. 0.49±0.61, P<0.001), hip (-3.03±0.46 vs. 0.71±0.95, P<0.001) and lumbar spine L1-L4 (-3.26±0.42 vs. 0.79±0.98, P<0.001) in PMOP patients compared with those in control subjects (Table I).

PCA plot and heatmap analyses of lncRNA and mRNA expression profiles. Fig. 1 presents the results of the lncRNA and mRNA expression profile analysis. PCA plots are provided in Fig. 1A and C and heatmaps are displayed in Fig. 1B and D. The data revealed differential expression profiles between PMOP patients and the control group. These results suggested that lncRNA and mRNA expression profiles may be used to discriminate PMOP patients from healthy control subjects.

Volcano plot and heatmap analyses of dysregulated lncRNAs and mRNAs. The volcano plot comprising 254 upregulated and 359 downregulated lncRNAs in patients with PMOP compared with those in the control group is provided in Fig. 2A. In the heatmap, patients with PMOP and control subjects were clustered separately based on the dysregulated expression of the lncRNAs (Fig. 2B). In addition, 522 upregulated and 1,148 downregulated mRNAs in patients with PMOP compared with those of the control group were revealed in the volcano plot (Fig. 2C), further suggesting a clustering capability of the two groups based on the expression of the dysregulated mRNAs as indicated in the heatmap (Fig. 2D).

GO and KEGG enrichment analyses for dysregulated lncRNAs and mRNAs. Enrichment analyses were performed to identify
the possible implications of the dysregulated lncRNAs and mRNAs in the relevant GO terms and signaling pathways. GO enrichment analysis indicated that the dysregulated lncRNAs were accumulated in various terms in the category biological process, including the apoptotic process and positive regulation of NF-κB signaling, as well as terms in the category cellular component, including cytosol and nucleoplasm, and terms in the category molecular function, e.g. protein binding and transcription factor activity (Fig. 3A). KEGG pathway analysis revealed that the dysregulated lncRNAs were mainly involved in PMOP-associated signaling pathways, including osteoclast differentiation and tumor necrosis factor (TNF) and mitogen-activated protein kinase (MAPK) signaling pathways (Fig. 3B). In addition, the GO enrichment analysis revealed that the dysregulated mRNAs were accumulated in terms in the category biological function, including apoptosis and protein phosphorylation, as well as terms in the category cellular component, including cytosol and membrane, and in terms in the category molecular function, e.g. metal ion and ATP binding (Fig. 3C). Finally, the KEGG pathway analysis revealed that the dysregulated mRNAs were highly associated with PMOP-associated signaling pathways in patients with diabetic complications, including osteoclast differentiation, vascular endothelial growth factor and the advanced glycation end products (AGE)/receptor for AGE (RAGE) signaling pathways (Fig. 3D).
Figure 3. Functional enrichment analysis of dysregulated lncRNAs and mRNAs. (A) GO enrichment analysis of the dysregulated lncRNAs. (B) KEGG enrichment analysis of the dysregulated lncRNAs.
Figure 3. Continued. (C) GO enrichment analysis of the dysregulated mRNAs. (D) KEGG enrichment analysis of the dysregulated mRNAs. Enrichment analyses of dysregulated lncRNAs and mRNAs were performed using the DAVID online tool. FAT: a filter applied to remove very broad terms; BP, biological process; CC, cellular component; MF, molecular function; TNF, tumor necrosis factor; RAGE, receptor for advanced glycation end products; CSF1R, colony-stimulating factor 1 receptor; TGF, transforming growth factor; VEGF, vascular endothelial growth factor; lncRNA, long non-coding RNA; GO, Gene Ontology; KEGG, Kyoto Encyclopedia of Genes and Genomes; MAPK, mitogen-activated protein kinase; hsa, Homo sapiens.
The characteristics of the top 20 upregulated and downregulated lncRNAs in patients with PMOP are presented in Table II. The top 20 upregulated lncRNAs were as follows: RP11-704M14.1, RP11-754N21.1, RP11-408E5.5, ANKRD26P3, TPTEP1, LINC01468, RP11-556E13.1, AE000661.37, RP11-384P7.7, HMGB1P41, MRPL42P6, CASC15, PWAR6, RP11-701H24.8, CTB-78021.1, RP11-214K3.19, AC104088.1, MIR99AHG, RP11-310E22.4, RP11-326K13.4, FABP5P1, SERPINB9P1, RPL13P2, EEF1DP2, RP11-529H2.2, MIR3180-2, LINC00890, LINC00672, RP11-1148L6.9, CTC-137K3.1, FLJ40579, RP11-93O17.2, LINC01481, NCF1B, CTD-2023M8.1, RP11-867G23.8, RP11-988C9.15, CTD-3126B10.4, CTB-35F21.1.

The top 20 downregulated lncRNAs were as follows: RP11-310E22.4, RP11-326K13.4, FABP5P1, SERPINB9P1, RPL13P2, EEF1DP2, RP11-529H2.2, MIR3180-2, LINC00890, LINC00672, RP11-1148L6.9, CTC-137K3.1, FLJ40579, RP11-93O17.2, LINC01481, NCF1B, CTD-2023M8.1, RP11-867G23.8, RP11-988C9.15, CTD-3126B10.4, CTB-35F21.1.

Top 20 upregulated and downregulated lncRNAs were selected based on the rank of absolute value of Log2FC. lncRNA, long non-coding RNA; PMOP, post-menopausal osteoporosis; P_adj, adjusted P-value; FC, fold change.

Table II. Top 20 upregulated and 20 downregulated lncRNAs in patients with PMOP compared to controls.

A. Upregulated lncRNAs

| Gene symbol | Gene ID       | Chromosomal location | Log2FC | P-value   | P_adj   |
|-------------|---------------|----------------------|--------|-----------|---------|
| RP11-704M14.1 | ENSG00000250696 | 4                    | 3.692493 | 2.53x10^-9 | 8.01x10^-8 |
| RP11-754N21.1 | ENSG00000258084 | 12                   | 3.018311 | 7.37x10^-4  | 3.57x10^-3  |
| RP11-408E5.5 | ENSG00000232243 | 13                    | 2.940108 | 5.41x10^-6  | 5.63x10^-5  |
| ANKRD26P3  | ENSG00000237636 | 13                    | 2.917439 | 3.72x10^-11 | 8.01x10^-8  |
| TPTEP1     | ENSG00000100181 | 22                    | 2.55805  | 2.64x10^-6  | 3.1x10^-5   |
| LINC01468  | ENSG00000251002 | 10                    | 2.494046 | 5.99x10^-20 | 3.05x10^-17 |
| RP11-556E13.1 | ENSG00000228651 | 10                   | 2.444645 | 8.08x10^-12 | 5.63x10^-10 |
| AE000661.37 | ENSG00000251002 | 14                    | 2.393043 | 1.186x10^-3 | 5.22x10^-3  |
| HMGB1P41   | ENSG00000260947 | 9                     | 2.352494 | 1.47x10^-10 | 7.17x10^-9  |
| MRPL42P6   | ENSG00000242810 | 3                     | 2.275756 | 2.70x10^-5  | 1.0281x10^-2 |

B. Downregulated lncRNAs

| Gene symbol | Gene ID       | Chromosomal location | Log2FC | P-value   | P_adj   |
|-------------|---------------|----------------------|--------|-----------|---------|
| RP11-310E22.4 | ENSG00000234206 | 10                   | -3.31587 | 5.97x10^-16 | 1.13x10^-13 |
| RP11-326K13.4 | ENSG00000263823 | 18                   | -3.24126 | 2.91x10^-14 | 4x10^-12  |
| FABP5P1     | ENSG00000236972 | 13                   | -3.02139 | 1.27x10^-7  | 2.34x10^-6  |
| SERPINB9P1 | ENSG00000230438 | 6                     | -2.72421 | 5.05x10^-9  | 1.49x10^-7  |
| RPL13P2     | ENSG00000213820 | 20                   | -2.53599 | 4.73x10^-16 | 9.27x10^-14 |
| EEF1DP2     | ENSG00000226721 | 9                     | -2.53635 | 7.67x10^-20 | 3.44x10^-17 |
| RP11-529H2.2 | ENSG00000255723 | 4                     | -2.51381 | 8.33x10^-17 | 1.91x10^-14 |
| MIR3180-2   | ENSG00000257366 | 16                   | -2.4977  | 4.34x10^-10 | 1.81x10^-8  |
| LINC00672   | ENSG00000263874 | 17                   | -2.48497 | 5.27x10^-11 | 2.95x10^-9  |
| RP11-1148L6.9 | ENSG00000204620 | X                    | -2.46017 | 7.42x10^-8  | 1.47x10^-6  |
| CTC-137K3.1 | ENSG00000270137 | 8                     | -2.4554  | 2.97x10^-10 | 1.31x10^-8  |
| FLJ40579    | ENSG00000204283 | 17                   | -2.39574 | 4.27x10^-14 | 5.47x10^-12 |
| RP11-93O17.2 | ENSG00000206356 | 5                     | -2.38807 | 2.47x10^-13 | 2.58x10^-11 |
| LINC01481   | ENSG00000257613 | 12                   | -2.36361 | 1.17x10^-19 | 5.12x10^-17 |
| NCF1B       | ENSG00000182487 | 7                     | -2.36149 | 6.22x10^-21 | 4.5x10^-18  |
| CTD-2023M8.1 | ENSG00000248693 | 5                     | -2.3384  | 3.98x10^-16 | 7.96x10^-14 |
| RP11-867G23.8 | ENSG00000255468 | 11                   | -2.30296 | 1.82x10^-22 | 1.63x10^-19 |
| RP11-988C9.15 | ENSG00000263731 | 17                   | -2.26584 | 1.19x10^-9  | 1.58x10^-5  |
| CTD-3126B10.4 | ENSG00000260176 | 16                   | -2.26079 | 6.1x10^-19  | 2.39x10^-16 |
| CTB-35F21.1 | ENSG00000249526 | 5                     | -2.24419 | 2.01x10^-11 | 1.29x10^-9  |
HMGB1P41, MRPL42P6, CASC15, PWAR6, RP11-701H24.8, CTB-78O21.1, RP11-214K3.19, AC104088.1, MIR99AHG, RP1-34H18.1 and LINC00890, whereas the top 20 down-regulated lncRNAs were as follows: RP11-310E22.4, RP11-326K13.4, FABP5P1, SERPINB9P1, RPL13P2, EEF1DP2, RP11-529H2.2, MIR3180-2, LINC00672, RP11-1148L6.9, CTC-137K3.1, FLJ45079, RP11-93O17.2, LINC01481, NCF1B, CTD-2023M8.1, RP11-867G23.8, RP11-498C9.15, CTD-3126B10.4 and CTB-35F21.1.

Regulatory network of the top 20 up- and downregulated lncRNAs. The top 20 up- and downregulated lncRNAs were recorded. According to the target gene search criteria of the cis and trans lncRNAs, all lncRNAs identified at least one target mRNA. The regulatory network based on these results for the 10 upregulated and 18 downregulated lncRNAs is presented in Fig. 4. The regulatory network was generated and examined to further increase the understanding of the association between the dysregulated lncRNAs and target mRNAs. All mRNAs that were differentially expressed and were closely correlated with dysregulated lncRNAs were selected. A total of 25 lncRNAs were demonstrated to directly regulate mRNAs gene expression levels. Furthermore, the majority of the lncRNAs exerted a positive regulatory effect on gene expression. In addition, among the predicted 25 lncRNAs, 13 were indicated to directly regulate the expression of >3 mRNAs, while the remaining 12 were demonstrated to regulate the expression of <3 mRNAs (Fig. 4).

Circos graph. The roles of the dysregulated lncRNAs and mRNAs in the pathology of PMOP were displayed in the Circos plots (Fig. 5). The outermost layer of the Circos plot indicates the chromosome number. The second and third outermost layers represent all differentially expressed mRNAs and lncRNAs, respectively. Finally, the inner lines represent the cis or trans lncRNA actions on the target mRNAs (Fig. 5).

Discussion

In the present study, the lncRNA expression profiles in patients with PMOP were investigated. A total of 254 upregulated and 359 downregulated lncRNAs were identified in patients with PMOP compared with those in the control group. GO enrichment analysis revealed that the dysregulated
IncRNAs were implicated in several GO terms in the category biological process, including apoptosis and positive regulation of NF-κB signaling. The implication of the dysregulated IncRNAs in the PMOP-associated signaling pathways was assessed using KEGG pathway analysis. The results suggested that the differentially expressed IncRNAs were accumulated in PMOP-associated signaling pathways, including osteoclast differentiation, as well as TNF and MAPK signaling pathways.

Aberrant gene expression profiles have been observed in various diseases, suggesting their potential as diagnostic biomarkers and therapeutic targets (15-17). The gene expression profiles in osteoporosis remain poorly investigated. A previous study revealed 327 upregulated and 396 downregulated genes in mesenchymal stem cells of 5 patients with osteoporosis by using microarray analysis (18). In a subsequent study, a total of 1,125 dysregulated genes (373 upregulated and 752 downregulated) were identified by microarray analysis in PBMCs derived from patients with osteoporosis (15). These results provided insight that may be used for the development of novel biomarkers and identification of treatment targets for osteoporosis.

Specific IncRNA expression profiles have been increasingly investigated in the past few years, mainly due to the implication of multiple aberrant IncRNAs in the development and progression of various diseases (19,20). Recent studies revealed that aberrant IncRNA expression profiles were associated with degenerative bone diseases, including intervertebral disc degeneration (IDD) (21). The investigation of the IncRNA expression profiles in IDD using microarray technology identified 1,570 dysregulated IncRNAs in IDD patients compared with those noted in control subjects. The IncRNAs were mainly enriched in the ‘extracellular matrix’ and cell ‘apoptosis pathways’ (21). A previous study identified 1,806 IncRNAs (1,357 upregulated and 449 downregulated) and 2,307 mRNAs (1,694 upregulated and 613 downregulated) that were differentially expressed in patients with IDD compared with those in a control group (22). In addition, the levels of IncRNAs in human degenerative and normal nucleus pulposus tissues were evaluated using IncRNA-mRNA microarray. The results demonstrated that 305 IncRNAs (135 upregulated and 170 downregulated) and 3,231 mRNAs (2,133 upregulated...
and 1,098 downregulated) were dysregulated (23). Therefore, considering the important role of IncRNA expression profiles in IDD, their implication in the development and progression of PMOP should also be investigated. However, at present, limited information is available. A previous study identified 25 upregulated and 26 downregulated IncRNAs in total blood samples from 5 patients with PMOP compared with those in 2 healthy control subjects (13). However, further studies with a larger sample size are required to confirm the role of IncRNA expression profiles in PMOP. In the present study, PBMCs from 10 patients with PMOP and 10 healthy controls were isolated in order to investigate their corresponding IncRNA expression profiles. A total of 254 upregulated and 359 downregulated IncRNAs were identified in patients with PMOP compared with those in healthy controls. In the present study, the number of the dysregulated IncRNAs was higher compared with that in the above-mentioned study. This discrepancy may be due to various aspects. First, the sample size in the present study was larger compared with that in the previous study, which increased the statistical power of the data. In addition, deep RNA sequencing was utilized to investigate the IncRNA expression profiles in patients with PMOP. Therefore, a higher number of dysregulated IncRNAs, particularly novel IncRNAs, was identified. Finally, certain differences in the clinical features of the patients of the two studies were present. For instance, in the present study, the average age was 5 years lower compared with that in the previous study. These discrepancies may explain the increased number of dysregulated IncRNAs reported in the present study.

Functional enrichment analyses are frequently performed to reveal the potential functions of newly identified dysregulated IncRNAs in various diseases (24,25). In a previous study, 25 upregulated and 26 downregulated IncRNAs were identified in patients with PMOP (13). Subsequent enrichment analyses suggested that the dysregulated IncRNAs (n=51) were significantly enriched in ‘membrane’, ‘cytosol’, ‘ribosome’, ‘cancer pathways’ and in ‘osteoclast differentiation’. The enrichment analysis of the present study revealed that the dysregulated IncRNAs were enriched in various GO terms in the category biological process, including ‘apoptosis’ and ‘positive regulation of the NF-κB signaling pathway’. In addition, significant enrichment was noted in several signaling pathways associated with diabetic complications, including ‘osteoclast differentiation’ and ‘TNF’, ‘MAPK’ and ‘chemokine’ signaling pathway, as well as the ‘AGE-RAGE signaling pathway’. These observations were consistent with previously reported data (13). The results suggested that the dysregulated IncRNAs may affect the development and progression of PMOP via the regulation of osteoclast differentiation, inflammation and apoptosis. In the present study, the regulatory network of IncRNAs and mRNAs was visualized using a Circos plot. In conclusion, the IncRNA expression profiles in patients with PMOP were investigated. In addition, novel dysregulated IncRNAs and their potential functions were identified by functional enrichment analysis. The aforementioned results may deepen the current understanding of the roles of IncRNAs in PMOP and provide an application of IncRNAs as biomarkers for PMOP and novel therapeutic targets.

The present study has certain limitations. First, the majority of the patients and healthy subjects were recruited from the Middle of China. Therefore, considering broader populations, a selection bias may have occurred. Furthermore, the expression of dysregulated IncRNAs was not verified in a larger number of patients with PMOP by in vitro detection methods, e.g. PCR. Finally, the mechanistic roles of dysregulated IncRNAs in the underlying pathogenesis of PMOP should be further investigated.

In summary, investigation of the IncRNA expression profiles in patients with PMOP and controls indicated that certain IncRNAs may be associated with the pathogenesis of PMOP via the regulation of osteoclast differentiation, inflammation and apoptosis.

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Availability of data and materials
All data generated or analyzed during the present study are included in this published article.

Author’s contributions
SW designed the experiment, performed the experiments, analyzed the data and wrote and revised the manuscript.

Ethics approval and consent to participate
Ethical approval was obtained from the Ethics Committee of the Union Hospital (Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China; approval no. 2017-S927). Written informed consent was provided by all participants prior to enrollment.

Patient consent for publication
Not applicable.

Competing interests
The author declares to have no competing interests.

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