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X-ray Analysis of the NMC-A β-Lactamase at 1.64-Å Resolution, a Class A Carbapenemase with Broad Substrate Specificity*

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The treatment of infectious diseases by penicillin and cephalosporin antibiotics is continuously challenged by the emergence and the dissemination of the numerous TEM and SHV mutant β-lactamases with extended substrate profiles. These class A β-lactamases nevertheless remain inefficient against carbapenems, the most effective antibiotics against clinically relevant pathogens. A new member of the enzyme class, NMC-A, was recently reported to hydrolyze at high rates, and hence destroy, all known β-lactam antibiotics, including carbapenems and cephapymics. The crystal structure of NMC-A was solved to 1.64-Å resolution, and reveals modifications in the topology of the substrate-binding site. While preserving the geometry of the essential catalytic residues, the active site of the enzyme presents a disulfide bridge between residues 69 and 238, and certain other structural differences compared with the other β-lactamases. These unusual features in class A β-lactamases involve amino acids that participate in enzyme-substrate interactions, which suggested that these structural factors should be related to the very broad substrate specificity of this enzyme. The comparison of the NMC-A structure with those of other class A enzymes and enzyme-ligand complexes, indicated that the position of Asn-132 in NMC-A provides critical additional space in the region of the protein where the poorer substrates for class A β-lactamases, such as cephapymics and carbapenems, need to be accommodated.

The extensive use of β-lactam antibiotics has resulted in bacteria becoming resistant to these agents. The resistance is mainly mediated by the class A β-lactamases and is spread by plasmid exchanges encoding the TEM and SHV mutant enzymes (1). In nosocomial bacterial infections highly resistant to penicillins and cephalosporins, imipenem and other carbapenem antibiotics are often considered antibiotics of last resort. These compounds are active against almost all clinically important Gram-positive and Gram-negative pathogens, including β-lactamase producers (2), and were shown to be nearly ideal drugs in pediatrics (3). Carbapenems differ from the classical β-lactam antibiotics because of the presence of a carbapenem ring fused to the 4-membered β-lactam ring and by the presence of the 6α-1R-hydroxyethyl substituent instead of the acylamido group found at 6β and 7β positions of penicillins and cephalosporins, respectively (Fig. 1). The antibacterial efficiency of carbapenems arises from several factors: (i) they are resistant to hydrolysis by nearly all class A β-lactamases, including the extended-spectrum mutant enzymes (4); (ii) carbapenemases, oxacillinases, and chromosomal cephalosporinases from different bacterial strains hydrolyze imipenem at very slow rates although the apparent binding constants are in the micromolar range (5, 6); and (iii) carbapenems display a very high affinity for the penicillin-sensitive pharmacological target enzymes (PBPs) involved in the final steps of the peptidoglycan cell wall synthesis (7).

Until recently, the only enzymes known to display high hydrolytic activity against carbapenems (carbapenemases) were the class B metallo-β-lactamases (8, 9), and the high resolution x-ray structures of two such enzymes were recently reported (10, 11). Surprisingly, three highly homologous carbapenemases, NMC-A (12) and IMI-1 (13) from Enterobacter cloacae, and Sme-1 (14) from Serratia marcescens, belong to the class A family of β-lactamases (15), which have so far been characterized by their specificity for penicillins. NMC-A, IMI-1, and Sme-1 hydrolyze at significant rates a wide range of β-lactam antibiotics, including those usually considered as resistant to the class A enzymes (13, 16, 17). A preliminary x-ray analysis of Sme-1 has been reported (18). Here, we present the x-ray structure of NMC-A, solved to 1.64-Å resolution, and its comparison with those of the typical class A β-lactamases. The differences observed in the substrate-binding site, complemented with kinetic measurements, provide structural explanations for the broadened substrate profile of this enzyme.

MATERIALS AND METHODS

Protein Expression and Purification—Oligonucleotides were purchased from Eurogentec (Belgium). To overexpress the NMC-A gene, an
800-base pair DNA fragment was amplified by PCR using the Vent DNA polymerase (New England Biolabs, Beverly, MA), pPTN1 as template (12) and the following oligonucleotides as primers: 1) a 37-base sense primer: 5′-GAGGGTACCATATGTCACTTAATGTAAAGCAAA-GTAG-3′ with Asp-718 and NdeI restriction sites at the 5′ end, and the last 23 bases encoding the N-terminal MSLNVKQS peptide of the preprotein; and 2) a 35-base antisense primer: 5′-GAGGGATCCTAG-GTTTATTTAAGGTTATCAATTGC-3′ with a BamHI restriction site at the 5′ end, and the last 21 bases complementary to the sequence encoding the C-terminal AIDNLK peptide and to the TAA stop codon.

The 800-base pair purified PCR fragment and pUC20 (Boehringer Mannheim) were digested with Asp-718 and BamHI and ligated. The resulting plasmid was used to transform the Escherichia coli Top10F9 cells (Invitrogen) and for mutagenesis purposes. The correct PCR fragment digested with NdeI and BamHI was thereafter inserted into the pET22b(+) plasmid (Novagen) after modification of its ampicillin resistance by insertion of a kanamycin resistance cartridge (Amersham Pharmacia Biotech). The resulting supercoiled overexpression plasmid was isolated from E. coli Top10F′ and used to transform E. coli BL21DE3. Cells were grown at 37 °C in an 18-liter Bio-Laffite fermentor containing 15 liters of Luria-Bertani broth added with 50 μg/ml of ampicillin. When the A550 reached 1.0, IPTG was added at a 0.5 mM final concentration, and the culture was continued for 3 h. The cells were collected by centrifugation, and the periplasmic content was liberated by lysozyme treatment. The supernatant was then dialyzed against 10 mM Tris-HCl buffer, pH 8.5, and loaded onto a Q-Sepharose Fast Flow column (4.6 × 30 cm2) equilibrated with the same buffer. The enzyme was eluted with a linear NaCl gradient (0–250 mM final concentration) over 1 liter. Active fractions were pooled, dialyzed under isocratic conditions with the same buffer. Under these conditions, the production and purification yields were 100 mg/liter and 80%, respectively. Enzyme purity was verified by SDS-polyacrylamide gel electrophoresis followed by Coomassie Blue and silver staining. The protein concentration was estimated on the basis of the A280 value, and the molar extinction coefficient was from the number of tyrosine and tryptophan residues (19).

Crystallization and Structure Determination—Crystals suitable for structure determination were obtained by the hanging drop method. The initial 6-μl drop, containing 2.0 mg/ml protein in 45 mM MES buffer at pH 5.25, 5% (w/v) PEG 1500, was equilibrated against 500 μl of 260 mM MES buffer containing 20% (w/v) PEG 1500 and 6% (v/v) n-propyl

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**FIG. 1. Chemical structures of the five classes of β-lactam antibiotics.** The letters a, b, and c indicate kinetic data from Refs. 16, 42, and 17, respectively. *, not measurable.

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1 The abbreviations used are: PCR, polymerase chain reaction; IPTG, isopropyl-1-thio-β-D-galactopyranoside; MES, 4-morpholineethanesulfonic acid; PEG, poly(ethylene glycol); SIRAS, single isomorphous replacement with anomalous scattering; r.m.s.d., root mean square deviation.
alcohol. After 4 days at 22 °C, parallelepipedic crystals and thin plates were formed. Equilibration was pursued for 15 days at 4 °C prior to crystal mounting. The crystals (60 × 100 × 850 μm³) belong to the orthorhombic system, space group P2₁2₁2₁ with cell parameters a = 78.7 Å, b = 52.9 Å, c = 67.5 Å, and contain one molecule in the asymmetric unit. Diffraction synchrotron intensities of both native and derivative crystals were measured on beam line W32 at LURE (Orsay, France) on a large MAR Research imaging plate. The wavelength of the x-ray beam was 0.975 Å. Two crystals were used to collect the native x-ray intensities: one crystal for a 2.9-Å resolution data set and the other one for data to 1.64-Å resolution. The platinum derivative was obtained by soaking the native protein crystals in 3 mM K₂PtCl₄ for 65 h, and one crystal was used for collecting an anomalous data set to 2.9-Å resolution. The SIRAS phases were improved by differences given for a range of residues concern the r.m.s. deviation on the backbone atoms of these residues. Structure superimpositions were made with ProFit (SciTech Software).

### RESULTS

#### Structure Determination

The completeness, high multiplicity, and low R<sub>merge</sub> values of the data sets proved to be valuable in the phasing procedure (Table I). The quality of the SIRAS phased electron density map computed to a resolution of 2.9 Å allowed an unambiguous main-chain tracing except for residues 100–103 in a loop region and for three residues at the N and C termini. One-third of the side chains were built at this stage. Inclusion of the high resolution data, followed by three rounds of refinement led to R and R<sub>free</sub> values of 0.230 and 0.253, respectively, at 1.64-Å resolution before inclusion of water molecules. The refined structure of NMC-A includes 265 residues and 115 water molecules, and the final R and R<sub>free</sub> values for all reflections between 31 and 1.64 Å were 0.192 and 0.214, respectively (Table I). The average temperature factor was 13.3 Å<sup>2</sup>, very close to the value estimated (13.6 Å<sup>2</sup>) from the Wilson plot (27). Coordinate errors were evaluated to be 0.18 Å from a Luzzati plot (28). The N- and C-terminal amino acids and seven solvent-exposed side chains of other amino acids in the sequence showed poorly defined electron densities. Alternative conformations were observed for Ile-30 Cα and the Asn-63 side chain.

The substrate-binding site is at the interface between two domains. The first one, hereafter denoted as the β-domain (residues 26–60 and 221–291), includes a five-stranded anti-parallel β-sheet (strands S1-S5) and helices H1, H10, and H11. The second one, the helical domain (residues 69–212), is made of eight helices (H2-H9) connected by loop regions (Fig 2). The overall fold of the protein is similar to that of other class A β-lactamases (29–32), but a number of structural differences were found, particularly in the substrate-binding site. In the following paragraphs, we address the possible functional significance of these differences with respect to the penicillins, cephalosporin, and carbapenemase activities of NMC-A.

#### The Substrate-binding site

A specific feature of the class A β-lactamases is the presence of single residues and 115 water molecules, and the final R and R<sub>free</sub> values for all reflections between 31 and 1.64 Å were 0.192 and 0.214, respectively (Table I). The average temperature factor was 13.3 Å<sup>2</sup>, very close to the value estimated (13.6 Å<sup>2</sup>) from the Wilson plot (27). Coordinate errors were evaluated to be 0.18 Å from a Luzzati plot (28). The N- and C-terminal amino acids and seven solvent-exposed side chains of other amino acids in the sequence showed poorly defined electron densities. Alternative conformations were observed for Ile-30 Cα and the Asn-63 side chain.

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### Table I

| Data set          | Resolution range | Number of observations | Unique reflections | Multiplicity | Bijvoet pairs | R<sub>merge</sub> | Completeness (I/σI) | R<sub>free</sub> | R<sub>free</sub> (26) |
|-------------------|------------------|------------------------|--------------------|--------------|---------------|------------------|---------------------|---------------|---------------------|
| Native 1          | 31.0–2.9         | 22,400                 | 6495               | 3.4          | 0.029 (0.40)  | 98.1 (82.7)      | 47.9 (22.2)         |               | 0.13 (0.04)       |
| K<sub>2</sub>PtCl<sub>4</sub> | 31.6–2.9       | 40,484                 | 6494               | 6.2          | 0.038 (0.557) | 98.4 (88.9)      | 52.8 (22.9)         | 0.13 (0.04)     | 0.027 (0.041)     |
| Native 2          | 31.0–1.64        | 111,487                | 32,628             | 3.4          | 0.036 (0.192) | 91.9 (68.0)      | 32.7 (3.2)          |               |                     |

#### Diffraction data statistics<sup>a</sup>

| Resolution range | No. of reflections | No. of protein atoms | No. of water molecules | R-factor<sup>b</sup> | R<sub>free</sub> | Average B-factors Å<sup>2</sup> |
|------------------|--------------------|----------------------|------------------------|----------------------|----------------|-------------------------------|
| 31.0–1.64        | 32,628             | 2024                 | 115                    | 0.192               | 0.214          | 13.3                          |
|                  |                    |                      |                        |                      |                | 11.4                          |
|                  |                    |                      |                        |                      |                | 14.4                          |
|                  |                    |                      |                        |                      |                | 22.8                          |

#### Geometric parameters r.m.s.d.

| Bond lengths (Å) | Bond angles (°) | Dihedral angles (°) | Improper angles (°) |
|------------------|----------------|---------------------|---------------------|
| 0.006            | 1.30           | 2.38                | 1.28                |

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<sup>a</sup> The numbers given in parentheses denote the respective values of the highest resolution shell.

<sup>b</sup> R<sub>merge</sub> = Σ[I(i) - 1/ΣΣ[I(i)].

<sup>c</sup> R<sub>free</sub> = Σ[I(i) - F<sub>calc</sub>]/Σ[I(i) + F<sub>calc</sub>].

<sup>d</sup> R<sub>free</sub> = Σ[I(i) - F<sub>calc</sub>]/Σ[I(i) + F<sub>calc</sub>].

<sup>e</sup> R = Σ[F<sub>obs</sub> - F<sub>calc</sub>]/Σ[F<sub>obs</sub>].
bridge, with a C$_\text{b}$-C$_\text{g}$-C$_\text{b}$ dihedral angle of $103.4^\circ$ and a C$_\text{a}$-C$_\text{a}$ distance of 5.0 Å. This covalent bond links the N terminal of helix H2 (containing the catalytic Ser-70 residue) to strand S3 (230–237), which defines one side of the substrate-binding site (Fig. 4). The disulfide bridge has several consequences. First, this bond and the set of interactions shown in Fig. 5, would be expected to greatly diminish structural flexibility in this region of the structure. Second, the distance between the main-chain nitrogen atoms of residues 70 and 237, which define the oxyanion hole, is 0.3 Å shorter than the average value (4.7 Å) in other class A $\beta$-lactamases (33). This would result in somewhat stronger hydrogen bonding of the water molecule typically found in that position in the absence of substrate. Finally, residue 238 adopts a conformation in which its carbonyl group is flipped by $180^\circ$ from the corresponding position seen in all other structures of class A $\beta$-lactamases. As a consequence, the S3 strand breaks from Ser-237, and the infrequent glycine residue inserted at position 239 moves away from the $\Omega$ loop region.

Several polar side chains are oriented toward the substrate-
binding site (Fig. 4). The uncommon histidine residues at positions 105 and 274 face each other at the entrance of the active site cavity. The side chain of His-274 is hydrogen bonded through one water molecule to the acidic group of Asp-276 and through a second water molecule to the main-chain nitrogen and oxygen atoms of residues 239 and 243, respectively (Fig. 5). At van der Waals distance from His-274, the guanidinium group of Arg-220 is engaged in a set of interactions that likely decrease its mobility and effective charge. It forms a salt-bridge interaction with Asp-276 and is hydrogen bonded to the main-chain oxygen atoms of residues 236 and 245, and to the hydroxyl group of Ser-237 (Fig. 5). The guanidinium group is located in an area similar to that of Arg-244 in the E. coli TEM-1 β-lactamase (32), in which it interacts with the carboxylate group of the substrate in the x-ray structures of acyl-enzyme complexes (31, 34, 35).

The disulfide bridge between cysteines 69 and 238 does not seem to affect the catalytic machinery of the protein (Table II). All atoms of the conserved catalytic residues (Ser-70, Lys-73, Ser-130, Glu-166, and Lys-234; 39 atoms) in NMC-A are found in the same positions as in typical class A enzymes. A least square fit of these atoms from the Staphylococcus aureus PC1 (29), Bacillus licheniformis 749/C (30), and E. coli TEM-1 β-lactamase (32) enzymes gives an r.m.s.d. of 0.3 Å, and the same result is obtained when NMC-A is also included in the calculation. We applied the corresponding transformation matrices to all protein atoms to compare the environment of these catalytic residues and the substrate-binding sites in the superimposed protein structures (Fig. 6, A and B). We observed that the atomic positions of Asn-132, which were not taken into account to compute the superimposition matrices, were similar (r.m.s.d. = 0.2 Å) in the three enzymes devoid of carbapenemase activity but are shifted by 1.0 Å in NMC-A, when compared with their average positions in the typical class A enzymes. This difference does not arise from a local displacement of the H5 helix (residues 132–142), but seems to result from folding variations throughout the protein structure (Fig. 2). It was also apparent that the main-chain oxygen atom of Ser-237, which was shown to interact with the nitrogen atom of the 6b-acylamido substituent of penicillin substrates during catalysis (31), is displaced by 1.0 Å compared with the PC1 and TEM1 enzymes (Fig. 6, A and B). This movement likely results from the α-conformation adopted by residue 238 but seems unrelated to the insertion at

![Fig. 4. Stereo view of the NMC-A substrate-binding site.](image)

![Fig. 5. Stereo view of the hydrogen bond pattern (dotted lines) involving Arg-220 and His-274 in the NMC-A structure.](image)

### Table II

Distances in the active-site cleft

| Identificationa | Distance (Å) |
|-----------------|--------------|
|                 | NMC-A | TEM-1 |
| Ser-70 Oγ       | 2.9   | 2.8   |
| Lys-73 Nζ       | 3.1   | 3.5   |
| Ser-130 Oγ      | 3.1   | 2.8   |
| Wat-308b        | 3.1   | 4.0   |
| Ser-130 Oγ      | 2.8   | 2.9   |
| Wat-332c        | 3.4   | 3.4   |
| Lys-234 Nζ      | 5.6   | 5.6   |
| Asn-132 Nδ2     | 3.0   | 2.8   |
| Gly-236 Ca      | 10.7  | 9.8   |
| Ser-237 N      | 9.8   | 9.0   |
| Ser-237 O      | 8.2   | 7.6   |
| Glu-166 O1      | 2.6   | 2.7   |
| Glu-166 O2      | 3.0   | 2.9   |
| Asn-170 Nδ1     | 2.6   | 2.8   |
| Water 308b      | 3.0   | 3.2   |
| Ser-70 N       | 2.8   | 2.8   |
| Ser-237 O      | 2.6   | 2.8   |
| Water 332c      | 3.0   | 3.0   |
| Ser-70 N       | 4.4   | 4.8   |

a Residue name as in NMC-A.
b Located in the oxyanion hole.
c The catalytic water molecule involved in deacylation.
Indeed, compared with the TEM-1 (32) and *B. licheniformis* (30) class A enzymes, the position of the main-chain oxygen atom of residue 237, and the conformation of residue 238 are not affected in the PC1 \( \beta \)-lactamase (29) although this enzyme contains an isoleucine inserted at position 239. The 237–240 regions in NMC-A and PC1 differ significantly (r.m.s.d. = 1.5 Å) (Fig. 6B), which suggests that the specific conformation of the C-terminal edge of the S3 strand in NMC-A may be attributed to the presence of the disulfide bridge. Cysteine 69 belongs to the polypeptide stretch connecting the \( \alpha \) and \( \beta \) domains of the protein. Formation of the disulfide bridge may influence the relative orientation of these domains and contribute to the folding variations illustrated in Fig. 2.

**DISCUSSION**

NMC-A and the class A \( \beta \)-lactamases devoid of carbapenemase activity are similar with respect to their hydrolytic machinery. The positions of the essential catalytic residues are identical within a r.m.s.d. of 0.3 Å, a value close to the estimated coordinate errors of 0.2 Å in structures refined below 2-Å resolution. This similarity, which preserves the molecular bases of the catalytic mechanism (36, 37), is in line with the high penicillinase activity of NMC-A.

On the contrary, we observed specific positional differences at residues 132 and 237–240. Several studies have emphasized the implication of the 238–240 region, at the edge of the substrate-binding site, with respect to the improved hydrolysis of third-generation cephalosporins and monobactams, by class A enzymes with extended-substrate specificity. An enhanced recognition of cephalosporins was displayed by engineered *S. aureus* PC1 enzymes (A238S and Ile-239 deleted; \( \Omega \)-loop deletion), and the x-ray structures revealed an altered disposition of the C-terminal edge of the S3 strand (38, 39). Several investigations were reported on the TEM and SHV enzymes because mutations naturally occurring in that region have been found in proteins responsible for bacterial resistance to \( \beta \)-lactam antibiotics (40). It was pointed out that the size correlation between the side chains of residues 69 and 238 in the parent enzymes breaks down in extended-spectrum \( \beta \)-lactamases, and it was suggested that the steric constraints would be accommodated by pushing the lower part of the S3 strand away from the active site, by 1–2 Å (41). In NMC-A, although the disulfide bridge reduces the Ca-Ca distance between residues 69 and 238, the conformation adopted by Cys-238 induces the predicted distortion in this part of the strand. It increases the space available between residue 170 of the \( \Omega \) loop region and the S3 strand, where the large \( \beta \) substituents of third-generation cephalosporins were reported to bind (39, 41, 42). The inserted glycine 239 (\( \Phi = 78^\circ, \Psi = 18^\circ \)), with its large available conformational space, might help to satisfy the constraints associated to the formation of the adjacent disulfide bridge. In addition, the side chain of any other residue in that position

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**Fig. 6. Stereo view of the substrate-binding sites in class A enzymes.** The superimposition of the enzyme structures is based on the least square minimization of all atoms from the catalytic residues (see text). A, NMC-A (red) and TEM-1 (blue); B, NMC-A (red) and PC1 (black).
would be oriented toward the binding site and would counteract the effect promoted by the conformation of residue 238.

With respect to the hydrolytic properties of NMC-A with carbapenem and cephamycin substrates, the direct implications of the disulfide-linked Cys-238 cannot be foreseen from the x-ray structure. The residues in the 238–240 region do not provide direct binding interactions to the functionalities on the α face of the substrate, such as the 6α-hydroxyethyl group of carbapenem antibiotics. It is noteworthy that the expanded-substrate enzymes deriving from the TEM and SHV β-lactamases, which display mutations of these residues, have not been reported to hydrolyze carbapenems and cephamycins. The position of Asn-132, away from strand S3 but still at 2.8 Å from Lys-73 Nα (Table II) (43), is unusual when compared with any other class A β-lactamase. Its new location might play a major role with respect to the carbapenemase activity because additional space is provided in a critical area for protein-substrate interactions, which would permit accommodating the 6α-hydroxyethyl substituent of carbapenems. This proposal is supported by kinetic and structural data. Mobashery and co-workers (44) showed that the attenuation of the turnover rate of imipenem with the TEM-1 enzyme only arises, for steric reasons, from the 6α-hydroxyethyl group and that this group imparts resistance to turnover by TEM-1 by 10⁴-fold. These data were in line with the observations made from the crystal structure of the acyl-enzyme complex formed between TEM-1 and its inhibitor, 6α-hydroxyethyl penicillanic acid, solved at 2.0 Å resolution (34). The three-dimensional structure showed tight interactions between the inhibitor, Asn-132, and strand S3 and indicated that the larger 6α-hydroxyethyl group of carbapenems would induce steric clashes with residue 132. The superimposition of the TEM-6α-hydroxyethyl penicillanic acid and NMC-A structures, based on the best fit of the atoms of their catalytic residues (Fig. 7), suggested that the 1.0 Å displacement of Asn-132 away from strand S3 in NMC-A (Table II), would allow the 6α-hydroxyethyl substituent to be easily accommodated and the hydroxyl group to orient differently in NMC-A. Indeed, kinetic experiments indicated a $k_{cat}/K_m$ value of $10^5$ M⁻¹ s⁻¹ for this substrate (detailed kinetic studies will be presented elsewhere).

In the TEM-1-penicillin G acyl-enzyme complex (31), the oxygen and nitrogen atoms of the 6β-acylamido group of the substrate were found at 2.6 Å from Asn-132 N82 and at 2.9 Å from the main-chain oxygen of residue 237, respectively. The altered position of this atom, and its increased distance to Asn-132 N82 in NMC-A (Table II), suggested that this hydrogen bond pattern may be altered in this enzyme with penicillin substrates. To evaluate this hypothesis, we determined the $k_{cat}/K_m$ value for 6β-aminopenicillanic acid that, from its chemical structure, may only interact with the main-chain oxygen atom of residue 237. We found that the hydrolytic efficiency for this substrate by NMC-A was similar, within experimental errors, to that for penicillin G, suggesting that in this enzyme the interaction between the oxygen atom of the 6β-acylamido group of penicillin G and Asn-132 may be weakened.

The most prominent features in the substrate-binding site of NMC-A are the altered conformations at the edge of the S3 strand and the unusual position of Asn-132. The former is reminiscent of several observations made to explain the extended-spectrum activity of typical class A mutant enzymes. The position of Asn-132 seems to be important for catalytic efficiency against β-lactam antibiotics bearing a 6α substituent (carbapenems), and both 7α and 7β substituents (cephamycins).

From the clinical point of view, the gene encoding NMC-A is derived from a member of the Enterobacteriaceae family, which is a common source of nosocomial infections for which carbapenems are often the β-lactams of last resort for use in patients in intensive care units. The broad substrate specificity of NMC-A, together with the poor effect of the inhibitors of class A enzymes currently in therapeutic use, should stimulate the design of new structure- and mechanism-based inhibitors, as was successfully done with the TEM-1 enzyme (34, 35).

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