**Sex Determination across Evolution: Connecting the Dots**

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Evolutionary developmental biology is motivated by the premise that the differences we see between species are caused by changes that have occurred in the genes that regulate their developmental programs. Beginning in the 1980s, general principles began to emerge about the evolution of development in animals. The identification of the *Hox* genes in *Drosophila melanogaster* and the subsequent discovery of their conservation and similar expression in different Metazoa led to the revolutionary realization that many of the mechanisms critical to basic animal development have been conserved across more than 500 million years of evolution. Many other developmental pathways, such as those specifying the heart and the central nervous system, have since been elucidated and promptly subjected to successful comparative analysis.

These celebrated discoveries illustrate ways that very different organisms are, at a fundamental level, similar to one another. But not all developmental processes are so conservative; an outstanding example is sex determination. The majority of animal species produce two sexes, and current phylogenies (e.g., [1]) suggest that sexual dimorphism was likely a feature of the last common ancestor of the coelomate bilaterians, a vast clade of animals that excludes only sponges, ctenophores, cnidarians, and acocel flatworms. However, though critical for development and reproduction, the mechanisms that specify sex determination are among the least-conserved known. Marked variation exists in both the primary sex determination signal and in the downstream genetic pathways that interpret the signal. We are thus presented with our first conundrum: sexual differentiation appears to be an ancient, and potentially homologous, feature of animal biology, yet its genetic specification suggests multiple origins.

**Bewildering Variety**

The variety of primary sex determination cues was appreciated long before the advent of molecular genetics [2]. The two broadest categories are genetic sex determination (GSD), in which the sex of offspring is set by a sex chromosome or an autosomal gene, and environmental sex determination (ESD), in which sex is determined by temperature (as with turtles), local sex ratio (as with some tropical fish), or population density (as with mermithid nematodes). Though little is known about the molecular mechanisms of ESD, within the GSD systems many different mechanisms have been uncovered. Dual sex chromosome systems, in which either the female (ZW/ZZ) or the male (XX/XY) is heterogametic, are common, as are systems set by the ratio of the number of X chromosomes to sets of autosomes (X:A). There are also systems in which heterozygosity at a single locus is required for female development (known as complementary sex determination; [3]), as well as systems involving sex determination via multiple genes with additive effects.

Molecular genetic investigations of GSD in model systems such as *Drosophila*, *Caenorhabditis*, and mice have revealed a clear lack of conservation, underscoring the diversity. For example, although the primary sex determination signal in both *D. melanogaster* and *C. elegans* is the X:A ratio, the fruit fly pathway consists of a cell-autonomous cascade of regulated mRNA splicing, while that of the nematode follows a *Hedgehog*-like intercellular signaling pathway [4]. GSD in mammals depends (with some interesting exceptions—see [5]) upon a Y-specific dominant gene (*Sry*) encoding a transcription factor. In the face of such impressive differences, perhaps we should question our assumption of homology: could it be that sex determination in different taxa has arisen independently over and over again in evolution? Until 1998, this seemed like a good bet.

The discovery of the homology of the key sex-determining genes *doublesex* in *Drosophila* and *mab-3* in *C. elegans* provided the first evidence for a common evolutionary basis of sex determination in animals [6]. Soon, related *doublesex-mab-3* (DM)-family genes with roles in male sexual development were discovered in vertebrates and even cnidarians [7,8]. Here at last was a smoking gun that could link the diverse metazoan sex determination systems (Figure 1). But as satisfying as the result was, it immediately gave birth to another mystery: if the enormous diversity of sex determination systems are all derived from a common ancestor, how could they possibly have been modified so radically? After all, sexual differentiation and reproduction are hardly unimportant developmental processes!

**Focusing on Close Relatives**

To understand how such diversity came to be, we need to look at the differences between closely related species. This approach allows the discovery and interpretation...
of small-scale sex determination changes before they are obscured by subsequent changes. The processes discovered in this way might then be reasonably extrapolated to explain the seemingly unrelated systems of more deeply diverged taxa. Work in dipterans [9] and nematodes [10] has revealed three evolutionary phenomena that characterize shorter-term sex determination evolution.

The first of these is the often astounding rate of molecular evolution at the level of nucleotide and amino-acid sequences. Although some sex-determining genes are well conserved, many show unprecedented substitution rates [11]. An extreme example is the central integrator of the X:A ratio in *Caenorhabditis*, *xol-1*. The *xol-1* orthologues of the closely related nematodes *C. elegans* and *C. briggsae* are a mere 22% identical [12], even though genes surrounding *xol-1* are much better conserved (Figure 2A). Remarkably, the 3′ neighbor of *xol-1*, the immunoglobulin dim-1, is only 5 kb away and is essentially identical between species.

A second phenomenon, best exemplified by dipteran insects, is the modification of genetic control pathways through the gain or loss of key pathway components (Figure 2B). In *Drosophila*, the first gene to respond to the X:A ratio is *Sxl*, whose transcription is regulated by both autosomal and X-linked factors very early in development [4,13]. When X:A = 1 (i.e., in female embryos), *Sxl* transcription occurs and produces *Sxl* protein. Later in development, transcription from a second promoter occurs in both sexes, but these transcripts cannot be productively spliced without the earlier burst of *Sxl* expression. As a result, only females sustain *Sxl* expression, and in turn only females can productively splice the mRNA of *tra*, its downstream target. Productive splicing of *tra* is required to produce the female-specific form of *Dsx*, a founding member of the DM family mentioned above.

In a series of groundbreaking papers, Saccone and colleagues investigated the pathway in the more distantly related heterogametic Mediterranean fruit fly *Ceratitis capitata*. The first surprise was that although a highly conserved *Sxl* homologue exists in *Ceratitis*, it does not undergo sex-specific regulation similar to that of *Drosophila*, which suggests that it does not play a key switch role (Saccone et al. 1998). Similar results have also been found for the housefly, *Musca domestica* [14], indicating that the role of *Sxl* in sex determination may be restricted to *Drosophila* and its closest relatives. In contrast, *tra* and *dsx* are key sex regulators in all dipterans examined thus far.

A further surprise came when the *Ceratitis tra* homologue was characterized [15]. In the case of this gene, clear evidence for sex-specific regulation was found, and as with *Drosophila*, only females productively splice *tra* mRNA. However, this splicing difference can be explained nicely by a positive feedback, similar to that seen in *Drosophila Sxl*, in which *Tra* protein regulates its own splicing. In 2002, Pane et al. proposed that the dominant, male-specifying M factor on the Y chromosome inhibits this autoregulation [15]. As a result, males cannot make functional *Tra* protein, and the male form of *Dsx* is produced. These experiments show not only how a pathway can evolve, but also, importantly, how X:A and heterogametic GSD systems can be interconverted by modifying the cue that regulates a conserved molecular switch gene (the splicing of *tra* mRNA). A detailed scenario for how this might occur has recently been proposed [16].

Finally, recent studies of *Caenorhabditis* nematodes have shed light on the genetic basis of the convergent evolution of sex determination related to mating system adaptations. An important factor in this area are new phylogenies of the genus *C. elegans* and *C. briggae* acquired self-fertilization independently, from distinct gonochoristic (male/female) ancestors (Figure 2C). Although this scenario is somewhat uncertain purely on parsimony grounds, recent work on the genetic control of the germline bisexual that defines hermaphroditism has tipped the balance toward parallel evolution.

Working with *C. elegans*, Clifford et al. [19] cloned *fog-2*, a gene required for spermatogenesis in hermaphrodites but not in males. Upon doing so, it became clear that *fog-2* is part of a large family of F-box genes and was produced by several recent rounds of gene duplication. The *C. briggae* genome sequence suggested that while *C. briggae* possesses a similarly large family of F-box proteins, the duplication event giving rise to *fog-2* was specific to the *C. elegans* lineage. In this issue of *PLoS Biology*, Nayak et al. [20] extend this work by rigorously demonstrating that *fog-2* is indeed absent in *C. briggae*. The authors also identify a short, C-terminal domain
that makes FOG-2 uniquely able to perform its germline sex-determining function. This domain is probably derived from a frame-shifting mutation in an ancestral gene. Working with *C. briggsae*, Stothard et al. [21], Haag et al. [22], and Hill et al. (unpublished data) have also found evidence of important species-specific regulation of germline sex determination.

RNA interference and gene knockout approaches have shown that while *C. elegans* requires the male-promoting genes *fem-2* and *fem-3* to produce sperm in hermaphrodites, *C. briggsae* requires neither. Given that both genes have conserved roles in male somatic sex determination, this suggests that *C. briggsae* evolved hermaphroditism in a way that bypasses these genes.

The long-standing mystery of sex determination and its diversity began by comparisons between distantly related species. Recent work on closer relatives has uncovered processes that through a reasonable extrapolation enable the connection of these disparate dots into a fascinating picture of developmental evolution. Though the divergence is extreme, it is likely that a better understanding of the evolution of sex determination genes and pathways holds lessons about the evolution of development in general. The next major challenge will be to integrate the comparative developmental data with the ecological and population processes that are driving the evolution of sex determination. Only then will we be able to say that the picture is complete.

**References**

1. Peterson K, Eernisse D (2001) Animal phylogeny and the ancestry of bilaterians: Inferences from morphology and 18S rDNA gene sequences. Evol Dev 3: 170–205.
2. Bull J (1983) Evolution of sex determining mechanisms. Menlo Park (California): Benjamin Cummings Publishing. 316 p.
3. Cook J (1993) Sex determination in the hymenoptera: A review of models and evidence. Heredity 71: 421–435.
4. Cline T, Meyer B (1996) Vive la difference: Males vs. females in flies vs. worms. Annu Rev Genet 30: 637–702.
5. Graves JAM (2002) Evolution of the testis-determining gene—The rise and fall of Sry. In: Chadwick D, Goode J, editors. The genetics and biology of sex determination. West Sussex (United Kingdom): J. Wiley and Sons. 276 p.
6. Raymond C, Shamu C, Shen M, Seifert K, Hirsch B, et al. (1998) Evidence for evolutionary conservation of sex-determining genes. Nature 391: 875–881.
7. Raymond C, Kettlewell J, Hirsch B, Bardwell V, Zarkower D (1999) Expression of Dmrt1 in the genital ridge of mouse and chicken embryos suggests a role in vertebrate sexual development. Dev Biol 215: 208–220.
8. Miller SW, Harvard DC, Bunch TA, Miller DJ, Ball EE, et al. (2003) A DM domain protein from a coral, *Acropora millepora*, homologous to proteins important for sex determination. Evol Dev 5: 251–258.
9. Graham P, Penn JK, Schell P (2003) Masters change, slaves remain. Bioessays 25: 1–4.
10. Stothard P, Pilgrim D (2003) Sex determination gene and pathway evolution in nematodes. Bioessays 25: 221–231.
11. Civetta A, Singh R (1998) Sex-related genes, directional sexual selection, and speciation. Mol Biol Evol 15: 901–909.
12. Luz JG, Hassig CA, Pickle C, Godzik A, Meyer BJ, et al. (2003) XOL-1, primary determinant of sexual fate in C. elegans, is a GHMP kinase family member and a structural prototype for a class of developmental regulators. Genes Dev 17: 977–990.
13. Saccone G, Peluso I, Artiaco D, Giordano E, Bopp D, et al. (1998) The Ceratitis capitata homologue of the Drosophila sex-determining gene sex-lethal is structurally conserved, but not sex-specifically regulated. Development 125: 1495–1500.
14. Meise M, Hilfer-Kleiner D, Dubendorfer A, Brunner C, Nothiger R, et al. (1998) Sex-lethal, the master sex-determining gene in Drosophila, is not sex-specifically regulated in Musca domestica. Development 125: 1487–1494.
15. Pane A, Salvemini M, Delli Bovi P, Polito C, Saccone G (2002) The transformer gene in Ceratitis capitata provides a genetic basis for selecting and remembering the sexual fate. Development 129: 5715–5725.
16. Pommankowski A, Nothiger R, Wilkins A (2004) The evolution of the Drosophila sex-determination pathway. Genetics 166: 1761–1773.
17. Kiontke K, Gavin NP, Raynes Y, Roehrig C, Piano F, et al. (2004) Caenorhabditis phylogeny predicts convergence of hermaphroditism and extensive intron loss. Proc Natl Acad Sci U S A 101: 9003–9008.
18. Cho S, Jin SW, Cohen A, Ellis RE (2004) A phylogeny of caenorhabditis reveals frequent loss of introns during nematode evolution. Genome Res 14: 1207–1220.
19. Clifford R, Lee M, Nayak S, Ohnachi M, Giorgini F, et al. (2000) FOG-2, a novel F-box-containing protein, associates with the GLD-1 RNA-binding protein and directs male sex determination in the C. elegans hermaphrodite germline. Development 127: 5265–5276.
20. Nayak S, Goree J, Schell T (2004) fog-2 and the evolution of self-fertile hermaphroditism in Caenorhabditis. PLoS Biol 3: e6.
21. Stothard P, Hansen D, Pilgrim D (2002) Evolution of the PP2C family in Caenorhabditis: Rapid divergence of the sex-determining protein FEM-2. J Mol Evol 54: 267–282.
22. Haag E, Wang S, Kimble J (2002) Rapid coevolution of the nematode sex-determining genes fem-3 and tra-2. Curr Biol 12: 2035–2041.
23. Muller H (1948) Evidence of the precision of genetic adaptation. Harvey Lect 43: 165–229.