INTRODUCTION

Hen eggs are an affordable foodstuff and are nutrient-dense food component that can contribute to healthy diet. Eggs contain good-quality protein and lipids as well as many micronutrients, including essential and non-essential vitamins and minerals (ENC, 2014). During the last three decades, the world’s egg production has increased dramatically, and Asia is the major consumer (FAO, 2018). Egg nutrient composition and quality are affected by diet, heredity, environment and the hen’s age (Koelkebeck, 2014). Variation in nutrients composition of laying hens’ feed can greatly impact the egg nutrition components such as vitamins, minerals, omega-3 polyunsaturated fatty acids (PUFA) and lutein that is favourable to human health (Walker et al., 2012; Yao et al., 2013). Improved performance and egg quality traits and composition have been observed in laying hens conventional diets containing ingredients such as maize, wheat, barley, oil cake and fishmeal (Swain et al., 2014). However, in the past decade, a dramatic global increase in the price of conventional feedstuffs has occurred due to shortages resulting from the competition between humans and livestock for conventional foodstuffs (FAO, 2018). To sustain
the optimal laying hen’s performance and egg productivity at the lowest cost, incorporation of cheaper and more readily available alternative feedstuffs into livestock diets is vital.

The oil palm (*Elaeis guineensis*), belonging to the family Palmae, is a valuable agricultural crop that is used to produce palm oil and other oil palm products. Palm oil is extracted from fresh fruit bunches (FFB) by a mechanical process. *Figure 1* presents a typical palm oil processing flow diagram. Solid waste materials and co-products are generated from the palm oil milling process. Inclusion of oil palm industry products and co-products as a non-conventional alternative food source for poultry is of primordial prominence in tropical countries due to its availability and cost-effectiveness compared to conventional feedstuffs. A schema showing how oil palm primary products and co-products that are available to the feed industry are produced during the palm oil milling process is shown in *Figure 2*. The palm oil mill and palm kernel crushing plant produce crude palm oil (CPO) and crude palm kernels oil (CPKO), respectively, as primary products, and biomass such as palm kernel cake (PKC) and palm oil mill effluent (POME) or palm oil sludge as secondary/co-products from fresh fruit bunches (Foong *et al.*, 2019). CPO, PKC and POME having considerable potential nutritive value have been used as feed ingredients for laying hens, for decades. The nutritive properties of these products, particularly their high content of metabolisable energy (ME), crude protein (CP), antioxidant compounds (especially vitamin E and carotenes) and minerals, could provide significant benefits for laying hen’s performance, egg production and egg composition, and the prospect for increasing their usage in the poultry diet is huge (Tafsin *et al.*, 2017). There are extensive information on the nutritive value, biochemical properties, development methods and feeding response of laying hens fed oil palm products and co-product-based diets. This article reviews the potential use of CPO and the co-products from palm oil milling (PKC and POME) to improve laying hens’ performance and egg production and quality traits.

*Figure 1. Palm oil processing flow chart.*

Source: FAO (2002).
EFFECT OF CRUDE PALM OIL ON LAYING HEN’S PERFORMANCE AND EGG QUALITY

Digestibility of CPO in poultry is influenced profoundly by saturation, the content of unesterified fatty acids, and by age of the birds (Palmquist, 2004). Tomkins and Drackley (2010) suggested the digestibility of vegetable oil is lower in young birds than in mature birds due to undeveloped bile salt system in the young animal and the emulsifiers improve the digestion of dietary lipids in poultry. Higher saturation and unesterified fatty acids greatly decreased digestibility in birds at 1.5 weeks of age compared with 7.5 weeks (Palmquist, 2004). CPO has been traditionally used as an ingredient (approximately 3%-5% of the diet) in feed for laying hens due to its tremendous value as a source of lipids, an energy source and a source of vitamins A and D as well as to reduce the dustiness of the diet (Tomkins and Drackley, 2010; Akter et al., 2014). CPO is particularly rich in natural vitamin E (specifically tocotrienols and tocopherols) and may confer specific health benefits such as hypocholesterolaemic, neuroprotective and anti-carcinogenic properties. Eggs can be enriched with functional compounds by biontransfer of these substances from feed to eggs.

Egg yolks enriched in vitamin E can be obtained by incorporating CPO in the laying hens’ diet (Kang et al., 1998). Due to vitamin E is not usually among the ingredients naturally present in feeds, their enhancement in feeds via the incorporation of CPO results in a stable improvement in eggs that is dose-dependent. Vitamin E from CPO, which is known to be efficient lipid-soluble chain-breaking antioxidants, prevent egg cell membranes from being attacked by lipid molecular oxygen (peroxyl) radicals in eggs (Atabo et al., 2018). CPO is also rich in carotenoids (mainly α-carotene and β-carotene). Incorporation of CPO could result in significant integration of retinol in the eggs, where β-carotene is the precursor (inactive form) for the formation of retinol from enzymatic oxidative cleavage of β-carotenoid by enzyme β-carotenoid 15, 15'-dioxygenase (Wu et al., 2016).

A recent study showed that dietary inclusion of CPO (2%-4% of the diet) significantly increased egg production, egg yolk colour, and egg and yolk weight but not shell weight, albumen weight or specific gravity compared to the control group (Areerob et al., 2018). Importantly, Areerob et al. (2018) observed that the diet supplemented with the highest level of CPO (4%) reduced egg yolk cholesterol levels and enhanced the total vitamin E and carotenoid levels in eggs. Earlier, Hodzic et al. (2008) also found that incorporation of CPO into layer rations could decrease egg and yolk cholesterol levels. In another study, Punita and Chaturvedi (2000) observed a maximum reduction in the total lipid content of hens’ eggs with the dietary inclusion of two hypocholesterolemic agents, CPO and grain.

Figure 2. Schema for palm oil manufacturing and co-products for poultry feed.
amaranth, either individually or in combination. The hypocholesterolemic effect of the CPO may be linked to its high tocotrienol content, which has been reported to decrease hepatic 3-hydroxy-3-methylglutaryl-coenzyme A (HMG-CoA) reductase activity and to act as an oxidised sterol, thereby reducing cholesterol levels (Qureshi et al., 1991).

In contrast, Akter et al. (2014) found that supplementation of laying hens’ diets with CPO at levels of 1.5%, 3.0% and 5.0% had no significant effect on egg weight, Haugh unit (HU), shape index, egg shell weight or albumen or yolk weight as a percentage of egg weight. They also found that CPO, which is known to be rich in saturated fatty acids, did not result in a significant increase in egg shell thickness at any of the inclusion levels tested. The reason could be that several dietary oils supplementation might impair calcium (Ca) and phosphorus (P) absorption due to increased saponification with feed minerals and decreased absorption of Ca and P by the gut system of hens and thus affect the egg shell thickness and reduce egg specific gravity (Abdulla et al., 2017). A number of studies has shown that dietary fat sources such as CPO had no significant effect on albumen weight, albumen height or HU because all the diets used had similar energy and protein (isonitrogenous and isocaloric) contents according to the hens’ nutrient requirements. On the other hand, the rate of hepatic synthesis of fats might be sufficient to supply the amount of lipids needed to achieve optimal egg albumen and yolk weights, and exogenous fat may not be necessary to meet these requirements.

The inclusion of increasing amounts of CPO in poultry feed is positively correlated with the yolk colour score, as explained by Ping and Gwendolin (2006) and Tranbarger et al. (2011). As hens are unable to synthesise egg colouration pigments, the colouration of the egg yolk depends on the absorption and deposition of pigmenting agents from the diet. Dietary carotenoids are commonly used to produce yellow-coloured yolks, whereas capsanthin, capsorubin and canthaxanthin are often used to produce red-coloured yolks (Sandeski et al., 2014). Kang et al. (1998) explained that CPO contains adequate amounts of β-carotenoids, which escapes from absorption in gut and impairs transportation to ovarian follicles and thus have a major impact on yolk colour. Moreover, CPO naturally consists about 90% carotene (α- and β-carotene) and 10% xanthophylls of the total carotenoids, which are essential for production of eggs with yellow-coloured yolks (Goh et al., 1985). Akter et al. (2014) found that yolk colour was significantly enhanced when the amount of CPO in the diet was increased; eggs from hens in the group that received 5.0% of CPO showed the highest yellow-coloured yolk score (5.69), followed by eggs from hens that received 3.0% of CPO (4.80), 1.5% of CPO (3.69) and basal diets (2.19), respectively. The improvement in egg yolk colour score from hens fed different diets can be attributed to the increasing dietary level of CPO, which is enriched in the yellow pigment xanthophyll, a lipid-like compound derived from the carotenoid group (Ping and Gwendolin, 2006).

CPO-supplemented laying hens may have lower levels of thiobarbituric acid-reactive substances (TBARS) and thus produce eggs with relatively lower levels of TBARS, resulting in an increase in the vitamin E (tocotrienols and tocopherols) and retinol content of the eggs (Kang et al., 1998; 2001). In agreement with previous findings, Akter et al. (2014) revealed that among the CPO treatments, eggs from hens that received diets higher in CPO had lower TBARS, indicating that absorption of more natural antioxidants from CPO resulted in increased lipid stability in eggs without affecting the egg quality characteristics. The synergistic effect of vitamin E (tocotrienol and tocopherol) and carotenoids present in the CPO might improve the oxidative stability of egg yolk, egg quality, and reduce the development of undesirable flavours while increasing egg production (Panda and Cherian, 2014).

**EFFECTS OF PKC INCLUSION ON LAYING HEN PERFORMANCE AND EGG QUALITY**

PKC is an important residue produced by the oil palm industry. It is also known as palm kernel meal (PKM) or palm kernel expeller (PKE). In an ideal milling process, oil can be extracted commercially from palm kernels via two key methods, *i.e.*, mechanical expression and solvent extraction. PKM or PKC, and PKE are the residue remaining after the solvent and expeller extraction of oil from palm kernels, respectively. Due to the general lack of discrimination between these two products, the term ‘PKC’ will be used throughout this review to refer to both products. Most of the PKC are used in animal feeding for fattening, as well as providing protein, energy, vitamins and minerals that required by various livestock for optimal growth performance (Abdeltawab and Khattab, 2018).

**Growth Performance**

PKC has been widely studied for use as a source of protein and energy in both laying hens and broiler chickens. It has been suggested that the incorporation of PKC in broiler or laying hen rations at the optimal level of 20%-30% as a partial replacement for corn and soyabees results in a reduction in the cost of poultry feed (Alimon and Wan Zahari, 2012; Yusrizal et al., 2013; Chen et al., 2015). Mohamed et al. (2012) suggested that utilisation of PKC as poultry feed ingredient at higher inclusion levels become limited due to its
high fibre and shell content. However, to date, reports on the optimal feeding level of PKC in layers’ diets and the effects of these diets on growth performance have been inconsistent. Sundu et al. (2006) speculated that these dissimilarities are due to differences in the total fibre content, the degree of grittiness and the amino acid content of PKC. Several studies have reported that inclusion of 30%-40% of PKC in the layer’s diet did not significantly affect body weight gain (BWG) compared to a diet containing maize-groundnut cake (Onwudike, 1988; Radim et al., 2000; Odunsi et al., 2002). Shakila et al. (2012) found that BWG of breeder-type layers fed 7.5 and 15% PKC were comparable with those fed with basal diet containing de-oiled rice bran (DORB) at 41 and 60 weeks of age. In contrast, Zanu et al. (2012) found that the inclusion of PKC in the diet at increasing levels (5%, 10% and 15%) significantly reduced the BWG of Lohmann Brown laying hens compared to a basal diet (0% PKC). A negative correlation between the level of PKC included in the ration and the BWG of hens may be attributed to the higher fibre content and lower nutrient digestibility of diets containing PKC. PKC might also undergo the Maillard reaction (a reaction between amino groups and the mannanose in PKC that forms a brown complex matter) during oil extraction under heat, and this may adversely affect its digestibility (Sundu and Dingle, 2003). Furthermore, high shell content of PKC might affect the feeding value for animal in terms of low protein content and energy availability. Recently, a premium grade PKC which is less shell (< 4%) content has been commercially developed in concordance of its better usage as a feed ingredient for broiler chicken that able to enhance the growth performance comparable to that of commercial chicken (Halim et al., 2017).

The feed intake (FI) of layer hens’ diets containing PKC is typically higher than that of hens fed a maize-based diet due to the more rapid transit rate of digesta derived from the former diets in the gastrointestinal tract (GIT) and their lower water-holding capacity and higher bulk density, all of which are believed to increase FI in birds (Akpodiete, 2008). In addition, PKC is a fibrous feed that releases less of its nutrient content because the nutrients are diluted by the fibre. Thus, layer hens are able to regulate their FI to obtain the necessary daily dietary energy level for optimal production performance. An increase in the FI of hens fed PKC-based layer rations could also be due to an increase in the palatability of feed as a result of presence of residual palm oil in PKC. Consistent with this, Odunsei et al. (2002) and Adrizal et al. (2011) found that cumulative FI was increased in native layers fed an isocaloric diet containing 30% of PKC compared to layers fed a diet containing maize-groundnut cake. Chong et al. (2008) suggested that when the poultry diet is isocaloric, birds might consume more feed to meet their amino acid requirements. An increase in FI may be due to the fibrouness of the diet (4.45% to 7.78% crude fibre (CF)) of the ration because PKC is defined as high CF (16.2%-25.3%) considering that fibre accounts for 50% of the non-starch polysaccharides (NSP) of the dry matter (DM).

However, studies by Perez et al. (2000) and Iyayi and Aderolu (2004) on the effect of the inclusion of PKC in laying hen diets reported that addition of PKC at various levels (7.5%-50%) did not influence FI or feed conversion ratio (FCR) compared to a maize-groundnut cake-based control diet. These findings suggest that PKC has relative benefit as a potential feed ingredient for laying hens in terms of digestion and nutrient availability. In contrast, Onwudike (1988) observed that layers’ diets containing 50%-70% of PKC impaired FI in layer hens (31 weeks old) over a six-month experimental period. A study conducted by Afolabi et al. (2012) showed that the daily FI of local hens (63.97 to 73.44 g/bird per 12 weeks) was significantly lower when PKC was included at levels of 10%, 20% and 40% than birds fed a basal diet (64.02 g/bird per day) but not when they were fed a diet containing 50% PKC. In the same study, lower FCR was observed in hens fed diets containing 20% to 40% of PKC, indicating more efficient and more favourable FCR than that found in hens fed diets containing 50% PKC. The reasons for the inconsistency of these results are unclear, but it is hypothesised that the differences could be associated with the biochemical properties of the PKC, including its amino acid composition, its high CF and energy content, its mineral nutrient content and its unpalatability (Sharmila et al., 2014).

The nutritional value of PKC for animal feed can be improved by the use of solid-state fermentation (SSF) and by the addition of exogenous enzymes. Mannanase can be used for the fermentation of NSP (mannans), resulting in an improved bioavailability of simple sugars as an energy source (Düsterhöft et al., 1992; Daud and Jarvis, 1992). Enzymatic depolymerisation of mannans may release mono- or di-saccharides that will be easily absorbed and metabolised by poultry (Saenphoom et al., 2013). Chemical treatment of PKC with acidic (acetic and formic acids) or alkaline (ammonium hydroxide) solutions has also been proposed to improve the PKC nutritive values by increasing the CP content and decreasing the fibre content (Adilah and Alimon, 2011). Fermentation of PKC using microbial biotechnology increased the CP content (15%-35%) as well as the true metabolisable energy (TME) value (5.5-8.1 MJ kg⁻¹) of PKC. Dairo and Fasuyi (2008) found that fermented PKC could increase the CP content from 20.04% to 23.42% and decreased the CF from 15.47% to 12.44%. This implies that fermented PKC can be used to substitute for the expensive soyabean meal protein in laying hens’ diets. Several studies have reported improvements in nutrient
digestibility and nutritional values of PKC via SSF using potential cellulolytic and hemicellulolytic bacteria, proteolitc bacteria and fungi for poultry feeding (Rahim et al., 2007; Alshelmani et al., 2013; 2014; 2016; 2017; Muangkeow and Chinajariyawong, 2009; 2013). These microbial cultures can change the chemical or physico-chemical properties of the PKC substrate as well as having the ability to produce multiple enzymes to breakdown the β-glycosidic bonds that link the cellulose, xylan and mannan in the PKC (Table 1). Therefore, releasing more sugars improve the nutritive value of PKC. In addition, the nutritive value of PKC fermented by microbes was improved, and it could be beneficial in terms of nutrient digestibility, intestinal microflora, ileal microbial fermentation and poultry performance (Marini et al., 2005).

Dairo and Fasuyi (2008) reported that overall BWG was significantly improved in 37-week old layers fed fermented PKC at 75% as a replacement for soyabean meal protein for 12 weeks. Chong et al. (2008) found that FI was significantly lower in layers fed diets containing PKC at 25% with 0.1% enzyme supplementation. However, laying hens fed diets containing < 25% of PKC with the inclusion of NSP enzymes showed better FCR than hens fed PKC without enzyme supplementation (Chong et al., 2008). Apart from laying hens, Soltan (2009) found that the negative effect of feeding a 20% PKC ration on broiler growth performance could be improved by the addition of a mixture of enzymes that included cellulose, xylanase, amylase, protease, β-glucosan and lipase. Chong et al. (2008) reported that inclusion of PKC at 12.5% and 25% with addition of an NSP enzyme in the diets of laying hens did not adversely affect the animals’ growth performance. The enhanced growth performance observed in the birds fed fermented PKC could be attributed to the reduction in the amounts of NSP and complex carbohydrates in PKC when the feed was supplemented with enzymes.

**Egg Production and Egg Quality Traits**

It is acceptable to include up to 25% of PKC in laying hens’ feed to achieve optimal growth performance without any detrimental impact on egg weight on external and internal quality (Yeong et al., 1981; Yeong, 1987; Radim et al., 2000). Consistent with this, Chong et al. (2008) found that dietary inclusion of 25% of PKC containing NSP enzyme in the layer diet did not show detrimental effects on egg quality traits. While, several studies have found no negative effects on egg production or quality traits in laying hens fed rations containing up to 40% of PKC (Onwudike, 1988; Perez et al., 2000; Akpodiete, 2008). Notwithstanding that, a study by Longe (1984) found lower tolerable levels of PKC (< 20%) in laying hens.

Afolabi et al. (2012) evaluated the effects of different dietary levels of PKC with added palm oil on the egg quality traits of Nigerian hens. The results showed that dietary PKC levels of 20%-40% resulted in significantly higher hen-day production (HDP) compared with hens fed a basal diet without PKC or a diet consisting of 50% PKC. However, the positive effects of PKC on egg quality traits were not observed in laying hens’ fed diets containing any of the PKC ration levels tested (10%-50%). A significant positive impact on HDP associated with the inclusion of varying levels (0%-50%) of PKC in the diet was also reported by Longe and Adekoya (1988) and Perez et al. (2000). Onwudike (1992) obtained FCR values in the range of about 3.09-3.12

| Microorganism                        | Enzymes involved during fermentation | Bioreactor (duration and temperature) | References                    |
|-------------------------------------|--------------------------------------|---------------------------------------|-------------------------------|
| *Rhizopus* sp.                      | Protease, lipolise and amylase        | -                                     | Rahim et al. (2007)           |
| Fibre degrading bacteria            | Cellulase, mannanase, xylanase        | Flask (4 to 7 d at 30°C)              | Alshelmani et al. (2013)      |
| *Cellulolytic and hemicellulolytic*| Cellulase, mannanase, xylanase        | Flask (12 d at 30°C)                  | Alshelmani et al. (2016)      |
| bacteria                            | Cellulase, mannanase, xylanase        | Flask (9 d at 30°C)                   | Alshelmani et al. (2017)      |
| *Paenibacillus polymyxa*            | Cellulase, mannanase, xylanase        | Flask (4 to 7 d at 30°C)              | Muangkeow and Chinajariyawong (2009) |
| *Cellulolytic bacteria (Paenibacillus polymyxa and Paenibacillus curdlanolyticus)* | Cellulase, mannanase, xylanase | Flask (9 d at 30°C)                   | Muangkeow and Chinajariyawong (2013) |
| *Aspergillus wentii*                | Amylase, lipase, dextranase, cellulase, pectinase, β-glucosidase, mannanase, xylanase | -                                     | Muangkeow and Chinajariyawong (2009) |
| *Aspergillus wentii*                | Amylase, lipase, dextranase, cellulase, pectinase, β-glucosidase, mannanase, xylanase | -                                     | Muangkeow and Chinajariyawong (2013) |

Note: d - days.
and a better HDP when feeding the Harco strain of laying hens with diets 20%-40% PKC-based. In the majority of cases, the incorporation of PKC at various levels (0%-50%) has no negative impact on egg quality traits, indicating that the PKC-based feed was nutritious and useful despite its fibrousness and imbalanced amino acid profile. The optimal level of inclusion of PKC could reduce the cost of egg production and improved the efficiency of production by substituting costly conventional feed ingredients with PKC ration.

Adrizal et al. (2011) indicated that native laying hens diets containing 15% or 30% of PKC with or without enzyme supplementation had similar trend in enhancement on egg production and egg internal and external quality traits (albumen height, HU and egg shell surface and thickness) compared to a corn-soyabean diet, but the yolk colour score (5-7) was significantly lower (paler) in the PKC-fed hens. A significantly lower yolk colour has also been reported for laying hens fed diets containing 25% of PKC (Panigrahi, 1998; Chong et al., 2008). Oil extraction of PKC at processing plant could lead to the removal of 80% of the oil, which consequently might eliminate most of the fat-soluble vitamins from PKC, including carotenes as well. This could be the main reason of declining effects of layer feeding dietary PKC on egg yolk colour. This is in line with study by Adrizal (2010) which observed beneficial effects of PKC in combination with cassava meal supplementation as a β-carotene source in laying hen’s diet has improved the egg yolk colour compared to the PKC diet alone. However, Zanu et al. (2012) revealed that egg yolk colour was significantly improved with a corresponding increase in the levels of PKC in layer ration. However, Longe (1984) discovered that layer hens fed 20% of PKC-based ration had lower egg production than hens fed a corn-soyabean-based diet, although the daily energy consumption values of the two groups of hens were comparable. In addition, Zanu et al. (2012) indicated that diets containing 15% PKC adversely affected egg production but did not affect egg weight (which ranged from 62.48 g to 68.53 g), egg shell thickness or HU score compared to the control diet. Whereas, Adrizal et al. (2011) indicated that feeding laying hens with diet containing 15% PKC had better effects on egg production and egg quality. Differences in PKC effects on egg production and quality at similar inclusion level indicated that, PKC might exert beneficial or detrimental effect depending on their substance chemical properties such oil content, CF level and shell content of the PKC obtained from different milling industry.

Nevertheless, more studies revealed that egg production and egg quality decline with the inclusion of >50% of PKC in laying hens’ diet (Pérez et al., 2000; Afolabi et al., 2012; Sharmila et al., 2014). Laying hen fed with high volume of PKC might show poor efficiency of food conversion into eggs and lower egg production due to the high CF content of PKC diets, which can lead to impaired nutrient digestibility and absorption, particularly in the case of amino acids, and also to nutrient imbalance rather than their energy requirements. Low availability of amino acids at the increasing levels of PKC may not meet the requirements for optimum egg production, hence the reduced rate of lay.

**EFFECTS OF POME ON LAYING HENS’ EGG PRODUCTION AND EGG QUALITY TRAITS**

POME, a by-product of CPO production that contains mainly fruit cell debris and oil from the palm mesocarp, is obtained by decanting the liquid waste. Commonly, dehydrated POME contains 12.5% CP and 20.1% CF and can be used as an alternative feed ingredient for poultry diet (Yeong, 1980). However, based on animal feeding trials, it was recommended that dehydrated POME should be used in poultry feed in limited amounts, i.e., 5% and 10%-15% in the diets of broilers and laying hens, respectively, owing to its high CF (11.5%-32.7%) and ash content (9%-25%) and its low digestible amino acid content (Sinurat, 2003). Appropriate amount of dehydrated POME in layer ration could result in optimal egg production of laying hens.

There are several commercial feeds derived from POME with enhanced nutritive value. The POME have been converted to potential feedstuff by using a fermentation process (anaerobic, acidophilic and thermophilic), followed by concentration (decantation down to 15%-20% DM) (Pérez et al., 1999; Chavalparit et al., 2006). The sludge-fermented product (SFP) has been known as treated POME had increased protein, metabolisable energy and in vitro soluble phosphorus, and reduced the CF content that can be used as feed ingredients for poultry (Mirmawati et al., 2019). The treated POME can be incorporated into broiler or layer diets as a protein source to replace 30% of soyabean meal. However, the recommended optimal level of the treated POME or SFP inclusion rate in laying hens diet was 10% (Yeong, 1980).

A study by Yeong (1987) had found average percent HDP, total egg mass and feed conversion (F:gain in egg mass) were 76.4%, 8.9 kg and 2.77:1, respectively, was obtained in hens fed the diet containing 10% treated POME compared with 77.9%, 9.2 kg and 2.52:1, respectively, for the corn-soyabean meal reference diet. Inferior results were apparent in those layers fed diets with more than 10% treated POME. This result implies that diets containing higher levels of treated POME (>10%) may be lacking in some of the nutrients required
to support optimal egg production, particularly because neither diet contains animal protein. However, egg quality (egg specific gravity, percent yolk, albumen and shell weight, yolk index and HU) was not affected by the inclusion of different levels of treated POME in laying hens’ diets. Yeong (1987) also reported that dehydrated POME with 13.0% CP could be used at levels of up to 20% in laying hens’ diets without detrimental effects on egg production. In general, dried portion of POME, which was in fermented form or dehydrated mechanically in the raw form is likely incorporated in layers or broilers rations.

The nutrient levels and quality of dry POME could also be increased by fermentation using lignocellulolytic fungi (Nuraini and Trisna, 2017). Fermentation with lignocellulolytic fungi, produce a variety of lignocellulolytic enzymes which are responsible for the biodegradation of lignocellulosic agro-wastes in nature (Govumoni et al., 2015). Lignocellulolytic enzymes such as cellulase, ligninase, xylanase and mannanase can reduce the CF levels and, improve the apparent metabolisable energy (AMEn) and availability of nutrients in dehydrated POME. Since PKC and POME are both obtained from oil palm fruit, a similar pattern of NSP consisting of mannan-oligosaccharide as a prebiotic source may occur in both materials. Sinurat et al. (2008) reported that substitution of 25% corn with dry POME or with the multienzyme treated POME slightly improved egg production, FI and FCR in laying hens. However, substitution of 50% corn with the multienzyme treated POME slightly reduced the FI and impaired the egg production and the FCR.

There has recently been much interest in using POME, compared to PKC by researchers as an alternative feedstuff for laying hens with regard to its nutritional values, abundant availability and low cost, also the advantage of its high-fat content and high energy ingredient source to be used in layer ration formulations. Moreover, practically there are fewer studies reported on using various biotechnological treatment methods of POME, which might be contributing to increased optimum inclusion level of POME in laying hens’ feeding and improved laying hen’s performance and egg production. Increasing the consumption of POME by livestock, particularly in poultry has been proposed as a means of an efficient approach to minimise environmental problems due to its highly polluting properties.

### TABLE 2. ADVANTAGES AND DISADVANTAGES OF CRUDE PALM OIL AND CO-PRODUCTS OF PALM OIL MILLING ON LAYING HENS’ FEEDING AND EGG QUALITY TRAITS

| Oil palm products and co-products | Advantages | Disadvantages |
|----------------------------------|------------|---------------|
| Crude palm oil (CPO)             | Improve the performance of pullets when fed up to 5% of diet. | Lower digestibility of CPO in young birds. |
|                                  | Economically feasible to substitute part of the maize in feed. | Higher saturation and unesterified fatty acids content decrease digestibility in bird. |
|                                  | Stimulates feed and metabolisable energy (ME) consumption. | Saponification with feed minerals and decreased absorption of calcium (Ca) and phosphorus (P). |
|                                  | Increase vitamin E (tocopherol and tocotrienol) and carotenoids contents in eggs. | |
|                                  | Increase egg weight and size. | |
|                                  | Reduce egg cholesterol. | |
|                                  | Inhibition of lipid oxidation in egg. | |
|                                  | Improve yolk colour. | |
| Palm kernel cake (PKC)           | Produce in large amounts and low cost. | Lower crude protein (CP) level. |
|                                  | Satisfactory dietary substitute for wheat middlings in layer diet. | High in fibre content. |
|                                  | Optimum inclusion up to 20%-30% in diet with no adverse effects on growth performance or egg quality traits. | Low starch polysaccharides. |
|                                  | Incorporation of 30% premium grade PKC as substitute for 25% of corn and 5% of soyabean in diet improve growth performance. | Lower protein digestibility of PKC diet. |
| Palm oil mill effluent (POME)/palm oil sludge (POS) | Economical replacement for maize (regular diet constituent) and soyabean. | More than 40% of PKC show deleterious effects on egg production and quality. |
|                                  | Dehydrated POME used to replace part of the protein and energy sources in layer diets. | Deficient in lysine, methionine and threonine contents. |
|                                  | Rich content of organic matter. | Reduced feed intake. |
|                                  | Optimum inclusion level is 5%-10% depending on the process (fresh, dried, and fermented). | |
|                                  | Dry POS is low in fibre (16.8%) content and high in protein content (13.0%) improve egg production. | Low in protein content and digestible amino acids. |
|                                  | Fat-rich substrate help improve egg quality. | High fibre content. |

|                                  | |
|                                  | High moisture content makes it difficult to incorporate in layer diet. |
|                                  | More than 10% of POME depress bird feed intake. |
The advantages and disadvantages of CPO and co-products of palm oil milling (PKC and POME) as feed ingredient on growth performance of laying hens, egg production and egg quality traits are summarised in Table 2.

CONCLUSION

Substitution of conventional feed ingredients in the rations fed to laying hens with primary products or co-products from palm oil milling was found to be economically beneficial in improving layer performance and egg quality traits. CPO could effectively be utilised as energy sources in layer rations for the best laying performance and egg quality when added at assigned ME values. Inclusion of PKC and POME at levels of up to 30% and 10%, respectively, had no detrimental impact on laying hen’s performance or on egg production. These co-products have the potential for use as ingredients in commercial layer rations in oil palm producing countries due to their contribution to developing a cost-effective feeding system and possibly to reducing the adverse environmental impact of liquid waste (POME) from milling. Use of the primary and co-products from the palm oil milling industry should be intensified to further expand their use in improving the egg production and quality as well as improving the growth performance in laying hens.

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