NUCLEAR AND CYTOPLASMIC OESTROGEN RECEPTORS IN SQUAMOUS CARCINOMA OF THE CERVIX

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Summary.—Nuclear and cytoplasmic oestrogen receptors (RE_N and RE_C) were sought in 5 normal cervices and in 43 specimens of squamous carcinoma of the cervix. All 3 tissue components of the 5 normal cervices contained both RE_N and RE_C. Thirty-five (81%) of the tumours contained receptors, but in only 9 (21%) were they found in both subcellular compartments. Twenty-four tumours (56%) had only RE_C and 2 had only RE_N. The potential therapeutic significance of these findings is not yet known, but it seems possible that tumours with an intact receptor mechanism might benefit from oestrogen therapy and have a more favourable prognosis.

All three tissue components of the normal cervix respond to oestrogens. Thus the growth of cervical stroma at puberty, during pregnancy and during oral contraceptive usage, the mid-cycle secretion of mucus by the columnar epithelium of the endocervix, and the growth of the squamous epithelium itself that may be seen on the atrophic postmenopausal cervix treated with hormone replacement are all due to oestrogens. Invasive squamous carcinoma of the cervix develops from abnormal squamous metaplasia in the transformation zone previously created by oestrogen-promoted stromal hypertrophy and consequent eversion of the endocervix. This study of oestrogen receptors was begun in order to determine whether oestrogens play any part in the transition from normal to neoplastic tissue in the human cervix. Nuclear and cytoplasmic receptors (RE_N and RE_C, respectively) were measured because of the extensive evidence in animals (Anderson et al., 1974, 1975; O’Malley & Means, 1974) and in humans (McGuire et al., 1978; Barnes et al., 1979) that it is the nuclear-bound oestrogen receptor that is responsible for effecting the expression of oestrogenic stimulation. An intact RE mechanism in breast cancer implies an improved prognosis (Leake et al., 1981a) and a high probability of response to endocrine therapy, if required (Leake et al., 1981b). If REs are present in squamous cervical cancer, and have the same significance, the clinical value of these observations would be considerable.

MATERIALS AND METHODS

The uteri from 5 women undergoing hysterectomy for fibroids were obtained immediately after removal. Endometrium was scraped out of the uterine cavity and samples of endocervical, ectocervical and stromal tissue were dissected from the cervix, placed in buffered saline (BS: 0.15M NaCl, 20mM Hepes, pH 7.4) on ice and transported to the laboratory for immediate assay. The uterus was sent for routine histological examination to confirm the normality of the tissues obtained.

Biopsy specimens were obtained from the tumours of 43 women with squamous carcinoma of the cervix. One portion was placed
in formol saline for histological confirmation of the diagnosis, and the remainder was transported immediately to the laboratory in BS on ice. From those biopsy specimens that were large enough, another portion of the tumour, adjacent to the part that would be assayed, was placed in formol saline for a histological assessment of the proportion of tumour cells in the sample. The remaining tissue was either assayed immediately or stored in liquid N₂ for not more than 4 weeks.

[2,4,6,7-³H]-Oestradiol-17β (85–110 Ci/mmol) was obtained from the Radiochemical Centre, Amersham, and its purity confirmed by ascending paper chromatography (Bush, 1952). Unlabelled steroids were obtained from Sigma Co. (St Louis, Missouri, U.S.A.) and, unless otherwise stated, all other chemicals were obtained from E. Merck (Darmstadt, Germany). The assay has been described in detail previously (Soutter et al., 1979; Peggaro et al., 1980) and therefore will be outlined only briefly here, indicating modifications in technique. Normal stroma and squamous epithelium were more readily homogenized, using a microdismembrator (Braun, Melsunge, West Germany) but less fibrous tissues from the endometrium, the endocervix and most tumour samples were disrupted using a Teflon-glass homogenizer. The homogenizing buffer (HED) used was: 20mM Hepes; 1-5mM EDTA; 0-25mM dithiothreitol; pH 7-4. Only 150 mg of tissue (50 mg/ml HED) was required for a full assay. A crude nuclear pellet was prepared by centrifugation at 700 g for 10 min, the supernatant was decanted and a portion stored at --20°C for protein estimation by Lowry's method. The crude nuclear pellet was washed once in BS and then resuspended in 0-15M NaCl, 20mM Hepes, pH 6-2, and an aliquot kept at --20°C for DNA estimation (Katzenellenbogen & Leake, 1974). The binding of [³H]-oestradiol-17β in both the 700g supernatant (cytoplasmic binding) and in the resuspended nuclear pellet was measured in 8 150μl portions after incubation for 18 h at 4°C with [³H]-oestradiol-17β at final concentrations from 0-1 to 0-8nM. An additional portion from both fractions was incubated with 0-8nM [³H]-oestradiol-17β plus a 100-fold excess of non-radioactive diethylstilboestrol to measure nonspecific binding. Appropriate blanks were included in the cytoplasmic assay. Free and loosely bound ligand was removed from the cytoplasmic preparation by dextran-coated charcoal, and from the nuclear suspension by washing with 20 ml 0-15M NaCl on Whatman GF/C filters using a Millipore filter unit. Total and bound radioactivity were measured in a Packard Tricarb 3390, using Instagel (Packard) as scintillant, at average efficiencies of 26% for the cytoplasmic assay and 42% for the nuclear assay. The data was plotted according to Scatchard (1949) and a straight line was drawn by the method of least squares, using the linear part of the plot. The correlation coefficient of this line was required to be significant to the 5% level. The number of binding sites was calculated from the intercept of this line and the abscissa, and the dissociation constant, which is inversely proportional to the strength of the binding measured, was calculated from the reciprocal of the slope. Receptors were deemed to be present if the dissociation constant was less than 8 × 10⁻¹⁰M. Thus it was possible in every case to measure the number of receptors present, and the strength of the binding.

| Table I. — Mean (s.e.) oestrogen-receptor concentrations and dissociation constants in normal uterine tissue |
|---------------------------------|----------------|-------------|----------------|-------------|
| RE  | (fmol/mg protein) | Kₓ | RE  | (fmol/mg DNA) | Kₓ |
| No. |                 |    | No. |                |    |
| Endometrium 5  | 156 (46) | 0-58 (0-21) | 5  | 595 (374) | 1-99 (0-74) |
| Endocervix  5  | 89 (37)  | 2-17 (0-55) | 4* | 414 (79)  | 3-40 (0-85) |
| Ectocervix  5  | 86 (36)  | 2-25 (0-67) | 5  | 664 (88)  | 3-04 (1-01) |
| Stroma      5  | 51 (18)  | 3-44 (1-12) | 4* | 1003 (234) | 3-49 (0-79) |

Kₓ = Dissociation constant × 10⁻¹⁰M.

* One nuclear assay from this group was technically unsatisfactory.
RESULTS

All 3 components of the 5 normal cervices contained both RE\textsubscript{N} and RE\textsubscript{C} (Table I). Although there seemed to be lower concentrations of RE\textsubscript{C} in the cervical tissues than in the endometrium, no such difference was apparent for RE\textsubscript{N}.

In contrast, samples of squamous carcinoma contained both REs in only 21% of cases, RE\textsubscript{C} alone in 56% of cases and RE\textsubscript{N} alone in 2 specimens (4.7%) (Table II). The levels of RE concentrations found were lower than in normal cervical tissue (Table III). Twelve samples were large enough to be assayed more than once (Table IV). Ten contained RE\textsubscript{C} on each occasion they were assayed, and 2 contained RE\textsubscript{N} on each occasion. One tumour was assayed 4 times, and evidence of nuclear binding was found every time. However, the line drawn on the Scatchard plot was not quite significant at the 5% level on two of these occasions, so those assays were reported as negative. In one of the 4 replicate cytoplasmic assays, a low concentration of binding was found. These were the only examples of non-concordance of results. There was no evidence of degra-

**TABLE II.**—Oestrogen receptors in squamous carcinoma of the cervix

| REC/REN | No. | %  |
|---------|-----|----|
| +/-     | 9   | 20.9|
| +/−     | 24  | 55.8|
| −/+     | 2   | 4.7 |
| −/−     | 8   | 18.6|
| Total   | 43  | 100 |

**TABLE III.**—Oestrogen-receptor concentrations and dissociation constants in squamous carcinoma of the cervix

| REC   | KD | RE\textsubscript{N} | KD |
|-------|----|---------------------|----|
| Mean  | 26.8 | 2.19 | 181 | 2.76 |
| S.e.  | 3.3  | 0.24 | 29.6 | 0.26 |
| Range | 5-96 | 0.30-6.28 | 61-397 | 1.53-4.47 |
| n     | 33   | 33    | 11  | 11  |

**TABLE IV.**—The results of replicate assays of squamous carcinoma

| Patient | REC   | KD  | RE\textsubscript{N} | KD  |
|---------|-------|-----|---------------------|-----|
| J.N.    | 63    | 0.88| 144                 | 2.64|
| C.S.    | 51    | 4.38| 260                 | 4.80|
| M.T.    | 25    | 2.80| 0                   | —   |
| B.D.    | 11    | 2.98| 0                   | —   |
| I.H.    | 27    | 2.11| 0                   | —   |
| P.D.    | 23    | 3.21| 159                 | 2.72|
| L.S.    | 59    | 0.30| 0                   | —   |
| S.M.    | 39    | 5.87| 0                   | —   |
| S.M.    | 62    | 3.61| 0                   | —   |
| P.M.    | 30    | 2.87| 0                   | —   |
| R.Z.    | 45    | 0.58| unsatisfactory assay | —   |
| 38     | 0.22 | 397  | 3.18  |
| R.Z.    | 13    | 5.66| 146                 | 2.78|
| 0            | 0   | 0    | 0              |
| 0            | 0   | 0    | 0              |
| 0            | 0   | 0    | 0              |
| 0            | 0   | 0    | 0              |
| 0            | 0   | 0    | 0              |
| 0            | 0   | 0    | 0              |
| 0            | 0   | 0    | 0              |
| 0            | 0   | 0    | 0              |
| 0            | 0   | 0    | 0              |
| 0            | 0   | 0    | 0              |
| 0            | 0   | 0    | 0              |
| 0            | 0   | 0    | 0              |
| 0            | 0   | 0    | 0              |
| 0            | 0   | 0    | 0              |

**TABLE V.**—Oestrogen receptors in squamous carcinoma of the cervix compared with the histological assessment of the percentage of tumour cells in the sample

| % Tumour cells | REC | RE\textsubscript{N} |
|----------------|-----|---------------------|
| 45             | 15  | 0                   |
| 50             | 31  | 203                 |
| 50             | 11  | 0                   |
| 50             | 23  | 0                   |
| 55             | 5   | 160                 |
| 60             | 27  | 196                 |
| 60             | 18  | 0                   |
| 65             | 24  | 0                   |
| 70             | 0   | 0                   |
| 75             | 22  | 0                   |
| 75             | 27  | 0                   |
| 80             | 0   | 0                   |
| 80             | 0   | 0                   |
| 80             | 45  | 0                   |
| 80             | 62  | 0                   |
| 80             | 38  | 397                 |
| 85             | 96  | 132                 |
| 90             | 0   | 182                 |
| 90             | 23  | 159                 |
| 90             | 37  | 144                 |
| 90             | 36  | 0                   |
| 90             | 0   | 0                   |
| 90             | 0   | 320                 |
| 100            | 16  | 0                   |
| 100            | 0   | 0                   |
| 100            | 10  | 0                   |
| 100            | 16  | 0                   |
| 100            | 19  | 0                   |
| 100            | 30  | 0                   |
| 100            | 53  | 0                   |
dation of receptors during storage in liquid N₂. Examples of Scatchard plots of the data are shown in Figs 1–3.

In 33 samples there was enough tissue in the biopsy portion for a histological assessment of the proportion of malignant cells. In 16 of these, 90% or more of the sample consisted of tumour; in 12, 60–85% and in 5, 45–55% of the cells seen were neoplastic. The proportion of tumour cells present, within these limits, seemed not to affect the results of receptor measurement (Table V).

DISCUSSION

The limited study of normal cervical tissues reported here was intended only to demonstrate that oestrogen receptors are normally present in the nucleus of the cell as well as in the cytoplasm. The cytoplasmic levels found are similar to those demonstrated by Sanborn et al. (1978).

In earlier studies of carcinoma of the cervix, a smaller proportion of tumours showed evidence of oestrogen binding. Whole-tissue uptake studies showed low levels of binding in 10 out of 26 samples (Terenius et al., 1971). REC measurement gave positive results at very low levels of binding in 17% of 42 samples of squamous carcinoma but in 3 of 4 samples of adenocarcinoma of the cervix (Hähnel et al., 1979). However, in a study of several tissues and tumours, the 3 samples of carcinoma of cervix which were assayed were all found to contain REC (Syrjälä et al., 1978).

Since the assay used in this study was able to detect receptors in normal tissue, it might have been possible that the low levels of receptors found in the tumours were due to an admixture of normal cells
in the biopsy specimen. The histological assessment of this "contamination" showed that in most cases the amount of normal tissue was too small to account for the concentrations of receptor detected; nor was there any correlation between the percentage of tumour cells in the biopsy specimen and the receptor concentration. Thus it seems unlikely that the receptors detected in biopsy specimens of tumour tissue were present only in normal cells contained in the sample.

The use of strict criteria for a positive result may have led to some assays being declared negative when receptors were in fact present. The opposite error was less likely. This is borne out by the good replication found in samples assayed more than once.

Tumours containing both REs may be susceptible to oestrogen regulation. Seventy-one per cent of breast cancers with this receptor configuration respond to endocrine therapy (Leake et al., 1981b).

Tumours with cytoplasmic but no nuclear binding may have abnormal receptors that are incapable of translocation to the nucleus. This type of tumour is much less common in breast cancer, where it accounts for only 12–17% of cases (Leake et al., 1981b; Pegoraro et al., 1980) of which a surprisingly high 24% respond to endocrine therapy (Leake et al., 1981b). The interesting tumours with receptor found only in the nucleus remain an enigma in breast (Leake et al., 1981b) and in cervical carcinoma, and challenge current concepts of oestrogen action. Few breast tumours of this type respond to endocrine therapy (Leake et al., 1981b). Similarly, by extrapolation from breast-cancer experience, few tumours with no evidence of RE in either compartment would be expected to respond to endocrine therapy and might be expected to carry a poorer prognosis (Leake et al., 1981a).

It would be quite wrong to draw conclusions from experience with breast cancer about the outcome of endocrine therapy or the prognosis in carcinoma of the cervix, simply because some of both types of cancer contain REs. The concentrations of both RE_C and RE_N are lower in carcinoma of cervix than in carcinoma of the breast (Leake et al., 1981b; Pegoraro et al., 1980) and other, more subtle, differences may exist. However, it is most encouraging to note that two controlled studies of oestrogen therapy combined with radiotherapy, as the primary treatment of carcinoma of the cervix, have shown a clear and substantial improvement in the 5-year survival of patients given oestrogens (Runge, 1959; Sugimori et al., 1976).

If the mode of action of oestrogens in carcinoma of the cervix is similar to that in carcinoma of breast, anti-oestrogens may be equally effective (Ingle et al., 1981) and the measurement of progesterone receptors may also be valuable (Barnes et al., 1980).

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