Lactobacillus plantarum NDC 75017 alleviates the learning and memory ability in aging rats by reducing mitochondrial dysfunction

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Abstract. The aim of the present study was to investigate the protective effect of Lactobacillus plantarum NDC 75017 on D-galactose (D-gal)-induced mitochondrial dysfunction in the rat cerebral cortex. Fifty rats were randomly divided into five groups (n=10 in each group). The rats in the aging model group were subcutaneously injected with 100 mg/kg D-gal and those in the protective groups were additionally orally administered L. plantarum NDC 75017 at doses of 1x10^5, 1x10^6 or 1x10^9 CFU/100 mg body weight/day, respectively. The control rats were administered an equal volume of the vehicle. Following continuous treatment for seven weeks, the learning and memory abilities and mitochondrial ultrastructure, function and adenosine triphosphate (ATP) levels were examined. The results showed that the learning and memory abilities and mitochondrial levels of ATP were significantly decreased in the D-gal-induced aging model group compared with those in the control group (P<0.01). In addition, marked changes in the mitochondrial functions and ultrastructure were observed between the groups. Seven weeks of L. plantarum NDC 75017 and D-gal coadministration significantly improved the learning and memory abilities of the rats compared with the D-gal-induced aging model group. Furthermore, the combination regime significantly improved the mitochondrial ultrastructure and functions, including the mitochondrial respiratory chain, mitochondrial membrane potential and mitochondrial permeability transition. The results revealed that the L. plantarum NDC 75017 was able to alleviate learning and memory injuries in aging rats by reducing the mitochondrial dysfunction induced by D-gal.

Introduction

Mitochondrial dysfunction plays a major role in the process of aging and in aging-related neurodegenerative disorders. This is due to the crucial role of mitochondria in producing adenosine triphosphate (ATP), the main source of cellular energy (1-4). Furthermore, mitochondria are the target organelles for reactive oxygen species (ROS), which are a major source of physiologically produced oxidative stress during aging (3). A number of studies have demonstrated that mitochondrial dysfunction is closely associated with several aging-related diseases, including Alzheimer's disease, Huntington's disease (HD) and Parkinson's disease (PD) (5-7). It has been confirmed that mitochondrial protection and the consequent reduction of oxidative stress are important targets for the prevention and treatment of the early stages of these aging-related diseases (8,9). D-galactose (D-gal) is a natural reducing sugar in the body that is normally metabolized by D-galactokinase and galactose-1-phosphate uridylytransferase in animals. An excess of D-gal results in abnormal metabolism (10). The progressive deterioration in learning and memory skills, as well as the production of ROS in the brain tissue of rodents, has been previously reported in the literature (11). It has been shown that the administration of D-gal for 6-10 weeks induces mimetic aging changes in the brain tissue of rats. This has been utilized to establish animal models in studies investigating potential therapies and prevention strategies for certain age-associated diseases (12-15).

In recent years, the most frequently used antioxidant food supplements have included certain lactic acid bacteria (LAB) and medicinal plants. It has been revealed that several dietary supplements, including spinach and citrus fruits extracts, may be beneficial in protecting against age-related neurological disorders (16,17). A number of LAB strains have the
ability to scavenge free radicals, improve the activity of anti-
oxidant enzymes and inhibit lipid oxidation. Hathout et al (18)
demonstrated that treatments with *Lactobacillus casei* or
*Lactobacillus reuteri* protected rats fed an aflatoxin-contami-
nated diet from oxidative stress. Bay et al (19) reported that a
skimmed-milk culture of LAB reduced lipid peroxidation in rat
livers and brains. The abnormal expression of \( \gamma \)-aminobutyric
acid (GABA), an important neurotransmitter in the brain, is
implicated in the pathogenesis of anxiety and depression (20).
A previous study showed that chronic administration of D-gal
markedly decreased the number of GABA-immunoreactive
neurons in the cortical layers of rats with D-gal-induced aging,
which further contributed to their behavioral deficits (21). This
is one of the suggested mechanisms by which LAB regulates brain
function (22). In a previous study, the LAB strain *Lactobacillus
plantarum* NDC 75017 produced high levels of GABA and
exhibited anti-inflammatory, cholesterol-lowering and anti-
oxidant properties (23-25). In the present study, the potential
protective effect of *L. plantarum* NDC 75017 was investigated
through the establishment of a D-gal-induced aging model
in rats. The behavioral changes were examined and the ATP
levels, mitochondrial function and mitochondrial ultrastructural
changes in the cerebral cortical neurons of the rats were exam-
ined to further investigate the potential mechanism underlying
the neuroprotective effect of *L. plantarum* NDC 75017.

**Materials and methods**

**Materials.** D-gal, rhodamine 123 (Rh123), rotenone and
4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid (HEPES)
were purchased from Sigma-Aldrich (St. Louis, MO, USA).
Bicinchoninic acid protein and ATP assay kits were purchased
from Wuhan Boster Bioengineering Co. Ltd. (Wuhan, China)
and Beyotime Institute of Biotechnology (Beijing, China),
respectively. All other chemicals used were of the highest quality
that is commercially available. The JSM25610LV transmission
electron microscope used in the study was produced by Japan
Electron Optics Laboratory Co., Ltd. (Tokyo, Japan).

*Lactobacillus strain and growth conditions.* The *L. plantarum*
NDC 75017 were isolated from a traditional Chinese fermented
yogurt (from the Tongliao range of Inner Mongolia, China).
The bacteria were anaerobically grown at 30°C overnight in de
Man-Rogosa-Sharpe broth (Difco™, Beckman-Coulter, Miami,
FL, USA). The bacteria were collected by centrifugation at
8,000 x g for 5 min, washed three times with phosphate-buff-
ered saline and adjusted to 1x10<sup>8</sup>, 1x10<sup>9</sup> and 1x10<sup>10</sup> CFU/ml
for oral administration to the rats.

*Animals and experimental design.* A total of 50 male Wistar
rats (weighing 180-200 g) were obtained from the Vital River
Laboratory Animal Technology Co., Ltd. (Beijing, China). The
rats were housed in separate cages and had free access to food
and water for 2±1 week to acclimate prior to the initiation of
treatment. The animals were housed in a limited-access animal
facility where the room temperature and relative humidity
were set to 22±2°C and 55±10%, respectively. Artificial
lighting provided a 24-h cycle of 12-h light/12-h dark (light
from 7:00 a.m. to 7:00 p.m.). All of the animal experiments
were approved by the Animal Care and Use Committee of
Heilongjiang Province, China. After the one-week acclima-
tion period, the rats were randomly divided into five groups,
with 10 rats in each group. The rats were orally administered
1 ml/100 g body weight of different concentrations (CFU/ml) of
*L. plantarum* once for 49 days (seven weeks). The rats
in the control and aging model groups were only administrated
a vehicle (0.9% saline) or D-gal, respectively. The treatments
were as follows: Group I, 0.9% normal saline (control group);
Group II, D-gal (100 mg/kg) subcutaneously (D-gal group);
Group III, low-dose *L. plantarum* [1x10<sup>9</sup> CFU/100 mg, per oral
(p.o.)] plus D-gal (100 mg/kg) (L + D-gal group); Group IV,
medium-dose *L. plantarum* (1x10<sup>10</sup> CFU/100 mg, p.o.) plus
D-gal (100 mg/kg) (M + D-gal group); Group V, high-dose
*L. plantarum* (1x10<sup>9</sup> CFU/100 mg, p.o.) plus D-gal (100 mg/kg)
(H + D-gal group).

*Water maze test.* Spatial learning was investigated after the
seven weeks of D-gal injection using the Morris water escape
task according to a previous study (26). From the 44th day, the
rats were trained for four days until the 49th day, when the time
taken to climb onto the platform (escape latency) was recorded
for each rat.

*Observation of mitochondrial ultrastructure.* After seven weeks
of treatment, the mitochondrial ultrastructure of the rat cerebral
cortices was observed using a transmission electron microscope
as previously described (27). A total of 15 rats (n=3 from each
group) were sacrificed through an intraperitoneal injection of
an overdose of sodium pentobarbital (80 mg/kg). The cerebral
cortices were isolated, fixed and perfused with 4% paraformal-
hyde. The cortices were subsequently stored overnight at 4°C.
Following post-fixation in 2% osmium tetroxide for 2 h at
4°C, the tissues were dehydrated in an ascending graded ethanol
and acetone series and immersed in an acetone/Epon 812 mixture
at ratios of 1:1, 1:2 and 1:3 for 0.5, 2 and 10 h, respectively. Ultra
thin sections (70 nm) were prepared, counterstained with uranyl
acetate and lead citrate and examined using the JSM25610LV
transmission electron microscope.

*Isolation and purification of mitochondria.* Mitochondria
were isolated from the cerebral cortices of the rats through

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**Figure 1.** Effect of *Lactobacillus plantarum* NDC 75017 on the spatial
learning of D-gal-induced aging rats. Data are presented as the mean ±
standard deviation (n=10). *P<0.01 and **P<0.01 vs. the control group. **P<0.01
vs. the D-gal group. L, low-dose *L. plantarum* NDC 75017; M, medium-dose
*L. plantarum* NDC 75017; H, high-dose *L. plantarum* NDC 75017; D-gal,
D-galactose.
homogenization and differential centrifugation according to the methods performed in a previous study (28). The protein content of the isolated mitochondria samples was determined using the Bradford protein assay. Bovine serum albumin was used to construct a standard curve.

Determination of ATP content. The levels of ATP in the mitochondria were measured using the ATP Bioluminescence Assay kit (Beyotime Institute of Biotechnology) according to the manufacturers’ instructions. Briefly, the levels of ATP were determined by mixing 50 µl mitochondrial solution with 50 µl luciferase solution, which catalyzes ATP-mediated light production from luciferin. The amount of emitted light, measured using a microplate luminometer (Promega, Madison, WI, USA), was linearly associated with the ATP concentration.

Measurement of the mitochondrial permeability transition (MPT). The MPT value was determined using an ultraviolet spectrophotometer to measure the absorbance at 540 nm (A540 nm), as previously described (29). Briefly, the isolated mitochondria were diluted to 0.5 mg/ml and incubated in the assay buffer (125 mm sucrose, 65 mm KCl, 5 mm succinate, 5 mm rotenone and 10 mm Tris-HCl; pH 7.4). MPT was initiated and monitored prior to and following the addition of 50 µM calcium chloride for 5 min. The results were expressed as the decrease in the absorbance at 540 nm.

Detection of mitochondrial membrane potential (ΔΨm). ΔΨm was detected according to the methods of a previous study (29), with modifications. Briefly, fluorescence (excitation at 503 nm and emission at 527 nm) occurring in the reaction buffer (250 mm sucrose, 2 mm HEPES, 0.5 mm KH2PO4, 4.2 mm sodium succinate at pH 7.4 and 0.3 mm Rh123) was measured using an F-4500FL spectrophotometer (Hitachi High-Technologies Co., Tokyo, Japan). The diluted mitochondria (0.5 mg/ml) were added to the buffer and incubated for 3 min. The fluorescence was measured again using the F-4500FL spectrophotometer. Finally, the change in ΔΨm was expressed by the decrease in fluorescence.

Activities of the mitochondrial respiratory chain. MTT reduction was used to assess the activities of the mitochondrial respiratory chain. The methods and procedures utilized in the present study were identical to those of a previous study (28). Briefly, 0.02 ml MTT (0.1 mg/ml) was added to the mitochondrial solution containing 60 µg protein. The reaction mixture was co-incubated at 37°C for 30 min and centrifuged at 1,000 g for 5 min at room temperature. The obtained pellet was dissolved in 1 ml of acidic isopropanol and re-centrifuged at 1,000 g for 5 min at room temperature to obtain the supernatant. The absorbance was measured at 595 nm and the results were presented as A595 nm/mg protein.

Statistical analysis. Data are presented as the mean ± standard deviation. The data were analyzed using one-way analysis of variance followed by least significant difference post hoc tests to compare the different treatment groups. P<0.05 was considered to indicate a statistically significant difference.

Results

Effect of L. plantarum NDC 75017 on the spatial learning of aging rats induced by D-gal. From the 44th day, the rats were trained for four days, following which the water maze test was carried out. The escape times of the rats in D-gal group were significantly higher (P<0.01) compared with those of the control.
group rats. However, the escapes times of rats in the L + D-gal, M + D-gal and H + D-gal groups were significantly lower (P<0.01) compared with those of rats in the D-gal group (Fig. 1).

D-gal-induced ultrastructural changes in neuronal mitochondria and the effect of L. plantarum NDC 75017. Ultrastructural changes in the mitochondria of the cerebral cortical neurons were observed and the results are shown in Fig. 2. After seven weeks of D-gal treatment, marked pathological changes were observed in the mitochondria of the cerebral cortical neurons of the D-gal-induced aging group compared with the control group, including rupture and scarcity of the cristae and vacuolization (Fig. 2B). Coadministration of L. plantarum NDC 75017 and D-gal decreased the neuronal mitochondria injury in rats with dose-dependent effects (Fig. 2C-E).

Effect of L. plantarum NDC 75017 on the D-gal-induced changes in ATP content. The levels of ATP in the mitochondria in the cerebral cortical neurons of the rats were determined. The results are shown in Fig. 3. Compared with the control group, the levels of ATP in the D-gal group were significantly lower (P<0.01). However, significant increases in the levels of ATP were observed in the M and H + D-gal groups (P<0.01) compared with the D-gal group.

Effect of L. plantarum NDC 75017 on Ca²⁺-induced changes in mitochondrial membrane potential. Calcium ions induce changes in MPT. The MPT in the mitochondria of rat cerebral cortical neurons was significantly higher (P<0.01) in the D-gal group compared with that in the control group. However, the MPT in the L, M and H + D-gal groups was significantly lower than that in the D-gal group, with decreases of 27.7, 41.1, and 48.9%, respectively, relative to the D-gal model group (all P<0.01).
Effect of L. plantarum NDC 75017 on D-gal-induced changes in \( \Delta \psi \text{m} \). \( \Delta \psi \text{m} \) was determined by monitoring the dynamic fluorescence quenching of Rh123. The initial fluorescence (714±36) was markedly reduced following the addition of mitochondria. As shown in Fig. 5, mitochondria of the rats in the control group quenched the fluorescence to 446±17, and the extent of quenching exhibited by mitochondria in the D-gal group was significantly lower (374±25) (P<0.01). In the M and H + D-gal groups, fluorescence quenching was significantly increased to 416±17 and 434±38, respectively (P<0.05 and P<0.01, respectively) compared with the D-gal group (Fig. 5).

Effect of L. plantarum NDC 75017 on the D-gal-induced changes in the activity of the mitochondrial respiratory chain.

The reduction of the water-soluble tetrazolium salt MTT to formazan is regarded as an indicator of mitochondrial respiration, particularly the activity of mitochondrial succinate dehydrogenase. Compared with the control group, mitochondrial enzymatic activity was significantly lower (P<0.01) in the D-gal group, and progressive improvements were observed in the L, M and H + D-gal groups (Fig. 6).

Discussion

Several theories have been proposed to explain age-related mitochondrial dysfunction. The oxidative stress theory is the most important theory of aging proposed in past few decades (30). The targeted accumulation of ROS damages the mitochondria, which are more sensitive and vulnerable to oxidative stress than other organelles in cells due to their structural and functional characteristics (6). The decrease in mitochondrial function and the increase in mitochondrial DNA damage suggests that the progressive accumulation of oxidative DNA damage is a contributing factor to cell apoptosis or necrosis, with the generation of more ROS during aging (10,20,28,29). Chronic administration of D-gal has been widely used to mimic the process of brain aging. The D-gal-induced model of brain aging is important in the development of suitable anti-aging drug strategies (31,32). A number of studies have revealed that low doses of D-gal (such as 50 or 100 mg/kg) decrease the learning and memory abilities of mice and rats, as demonstrated through the T-maze, Y-maze and Morris water maze tests (26,33). In the present study, rats administered 100 mg/kg D-gal for seven weeks exhibited a significant decrease in learning and memory ability, as confirmed through their performance in the Morris water maze test.

At present, mitochondrial Ca\(^{2+}\) homeostasis is the center of widespread interest in scientific studies due to its modulatory role in numerous physiological processes and its involvement in cell death (6,28,29). Ca\(^{2+}\) uptake and release from the mitochondrial membrane via a variety of mechanisms control the local regulation of intracellular Ca\(^{2+}\) concentration. The mitochondrial Ca\(^{2+}\) dysfunction can cause MPT pore opening, leading to a change in the \( \Delta \psi \text{m} \) and resulting in mitochondrial swelling and dysfunction, and even cell death (28). It has been demonstrated that injecting rodents with D-gal for 6-10 weeks induces aging, affecting mitochondrial bioenergetics. This leads to the activity of the electron transport chain complex becoming compromised and a decrease in the rate of ATP synthesis (6,28). The results of the present study indicated that chronic administration of D-gal impaired the activity of the mitochondrial enzyme complex, \( \Delta \psi \text{m} \), mitochondrial membrane permeability and ATP production ability. These changes were alleviated with the administration of L. plantarum NDC 75017, which may be associated with its antioxidative properties.

GABA supplementation may activate the upstream signal survival pathways that regulate mitochondrial function, such as the phosphoinositide 3-kinase-Akt signal survival pathway (34). GABA also plays an important role in the aging process and diseases including PD and HD (22,34,35). GABA increases the circulation of serum lipids and decreases mitochondrial injury mediated by oxidative stress (21,36). Certain LABs have been demonstrated to directly regulate GABA in the microbiome-gut-brain axis in mice (22). L. plantarum NDC 75017 has the ability to produce high levels of GABA in vitro, which may contribute to its anti-aging and D-gal-induced mitochondrial dysfunction-alleviating abilities. The precise mechanisms for this should be investigated in further studies.

In conclusion, the results of the present study revealed that L. plantarum NDC 75017 was able to alleviate learning and memory-associated injuries in aging rats by reducing mitochondrial dysfunction induced by D-gal. This may be associated with its antioxidant and GABA-producing activities.

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