Early-Onset Neonatal Sepsis Caused by Vertical Transmission of Pasteurella multocida

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Abstract

Early-onset neonatal sepsis contributes substantially to neonatal morbidity and mortality. Presenting signs and symptoms vary, and most causes are due to a limited number of common microbes. However, providers must be cognizant of unusual pathogens when treating early-onset sepsis (EOS). We report a case of a term neonate who presented with respiratory distress, lethargy, and hypoglycemia 5 hours after birth. He was treated for presumed EOS with blood culture, revealing an unusual pathogen, Pasteurella multocida. Sepsis from this pathogen is a rarely reported cause of early onset neonatal sepsis. Our report is one of few that implicate vertical transmission with molecular diagnostic confirmation of P. multocida, subspecies septica. The neonate was treated with antibiotics and supportive care and recovered without ongoing complications. Providers should maintain an index of suspicion for rare causes of neonatal EOS. For these unusual cases, precise microbial identification enables understanding to provide best clinical care and anticipation of complications.

Keywords

- Pasteurella multocida
- neonate
- early-onset sepsis
- vertical transmission
- gene sequencing

Background

Neonatal sepsis is a major contributing cause of neonatal morbidity and mortality. The presentation of early-onset sepsis (EOS) in the first 7 days after birth in full-term infants is variable and may include any of the following: decreased activity level and lethargy, temperature instability, hypoglycemia, respiratory distress, poor feeding, and seizures. These findings can range from subtle to severe. EOS is most often caused by group B Streptococcus; it is followed by Escherichia coli and other streptococci.1

While other gram-negative organisms are isolated less commonly, they are also responsible for neonatal EOS and must be considered to ensure appropriate management.

While EOS is typically caused by intrapartum maternal-neonatal vertical transmission, further investigation for the source of infection may be warranted when less common organisms are isolated.

We present a case of an infant with EOS due to Pasteurella multocida, subspecies septica, describe our investigation, and discuss recommendations for isolation of the source of this vertically transmitted infection, with speciation confirmed by 16S ribosomal ribonucleic acid (rRNA) gene sequencing.

Case Presentation

The mother was a 26-year-old G1P1 who delivered at 380/7 weeks. She received routine prenatal care, and serologies
were all negative or nonreactive. She went into spontaneous labor, and spontaneous rupture of membranes occurred 8 hours prior to delivery. The intrapartum course of labor and delivery of an appropriate for gestational age (2.92 kg) male were uncomplicated. No resuscitation was required, Apgar’s scores were normal, and he was admitted for normal newborn cares.

The infant developed respiratory distress with grunting 5 hours after birth. The infant was lethargic and hypoglycemic (blood glucose: 26 mg/dL). Oxygen was initiated via nasal cannula. Given concern for bacterial sepsis, intravenous (IV) access was obtained for empiric administration of ampicillin and gentamicin. Chest X-ray revealed bibasilar infiltrates, and complete blood count revealed severe leukopenia (white blood cell count: 2.3 × 10^9/L). Neonatal intensive care unit (NICU) referral was made to transport the infant for a higher level of care. The transport team intubated the infant and started fluid resuscitation given progressive respiratory failure and poor perfusion.

Upon quaternary NICU admission, laboratory studies revealed lactic metabolic acidosis, leukopenia with severe neutropenia (absolute neutrophil count: 500 × 10^9/L), and thrombocytopenia (platelet count 111 × 10^9/L). There was no coagulopathy or evidence of other end organ dysfunction. Chest radiograph revealed continued infiltrates (Fig. 1). At 12 hours of incubation, the blood culture grew gram-negative rods. Evaluation for other infectious foci included lumbar puncture, urinary catheterization, and tracheal aspiration. Cerebrospinal fluid analysis showed normal white blood cell count (17 cells/μL; 3% neutrophils, 2% lymphocytes, and 95% other cells), elevated red blood cell count (258 cells/μL), normal protein and glucose concentrations, and negative Gram’s stain. Urinalysis was unremarkable. Tracheal aspirate showed few gram-negative rods and fewer than 25 polymorphonuclear cells.

The infant had persistent pulmonary hypertension and progressive septic shock with continued hypotension and poor perfusion. Infectious diseases consultation was obtained. Antibiotics were broadened to include meropenem to cover resistant gram-negative organisms and improve central nervous system penetration. The infant received fluid resuscitation, multiple vasoactive medications, and inhaled nitric oxide to improve oxygenation in the setting of persistent fetal circulation. Blood and tracheal cultures revealed *P. multocida*. The organism was susceptible to all agents tested: ampicillin, ceftriaxone, levofloxacin, penicillin, trimethoprim-sulfa, and azithromycin. Cerebral spinal fluid and urine cultures were both negative. After the initial positive culture, subsequent blood cultures were negative. Further history revealed multiple pets in the parents’ home including two cats and one dog. Family members denied any history of bites or scratches. To evaluate more closely, the source of transmission to the neonate, maternal vaginal culture was obtained and was also positive for *P. multocida*. The mother did not endorse any vaginal symptoms.

To confirm vertical transmission, molecular diagnostic studies were performed on the positive cultures from the patient’s blood and tracheal aspirate, as well as from the maternal vaginal specimen. Initial identification as *P. multocida* was by matrix-assisted laser desorption ionization–time of flight (MALDI-TOF) mass spectrometry. Our Infectious Diseases Diagnostic Laboratory then performed 16S rRNA gene sequencing. The 16S rRNA gene region was amplified by polymerase chain reaction (PCR) and sequenced using an Applied Biosystems 3130 Genetic Analyzer. The final sequences for the three isolates were searched within the GenBank database using the basic local alignment search tool (BLAST) algorithm with these restrictions applied to the search: exclusion of the uncultured/environmental samples sequences and limited to sequence type material only. *P. multocida*, subspecies septica, was the best sequence match at 100% for all three isolates.

Furthermore, to determine the similarity among the three isolates, their sequences were aligned to one another using the National Center for Biotechnology Information (NCBI) nucleotide BLAST function which showed the three separate sequences of the three bacterial isolates were a 100% match to each other for *P. multocida*, subspecies septica. This highly suggested neonatal EOS with pneumonia due to vertical transmission from the mother. Presumably, maternal vaginal colonization was related to pet exposure in her home. No specimens from those animals were obtained.

**Case Resolution**

The Infectious Diseases Service recommended treatment of the mother with amoxicillin-clavulanic acid for a 10-day course. They also recommended that the mother sought infectious diseases consultation with any subsequent pregnancies to consider the need for vaginal colonization testing.
The infant made steady clinical progress starting 5 days after birth. The ventilator and cardiovascular support were weaned, and he was hemodynamically stable without need for respiratory support 7 days after birth. Neutropenia quickly resolved without treatment other than antibiotics. Thrombocytopenia initially worsened ( nadir of 36 \times 10^9/L) which was treated with one transfusion before eventual normalization. C-reactive protein levels peaked at 174 mg/L at approximately 60 hours and were followed to normalization.

With clinical progress and finalized culture results, antibiotic therapy was narrowed to ceftazidime, with gentamicin used a few days for synergy. He completed a 14-day course of antibiotics from the first negative blood culture with single agent ceftazidime. The infant was discharged to home with his parents 17 days after birth. Thereafter, his infancy was without noted morbidities.

**Discussion**

Our case describes a newborn with EOS and pneumonia due to a relatively rare gram-negative organism, *P. multocida*, a facultatively anaerobic gram-negative coccobacillus mainly found in the oral flora of many animals. Human infection is most commonly a cellulitis following a wound inflicted by a cat or dog. Other manifestations are possible, and exposure to colonized cats and dogs is a risk factor for infection.

Collecting a detailed exposure history can assist in determining mode of transmission.

All reports of neonatal pasteurellosis, both EOS and later onset (8–30 days), have been due to the most common species seen in human infection, *P. multocida*. For treatment of *P. multocida*, penicillin is the preferred drug treatment but some isolates may be resistant to this antibiotic. Other possible regimens include ampicillin and sulbactam; in addition, third-generation cephalosporins, such as ceftriaxone, are effective. As in our case, infants display clinical presentations seen with other causes of neonatal sepsis. *Pasteurella* is usually isolated from the neonate’s blood and or cerebral spinal fluid and has also been implicated in pneumonia, osteomyelitis, and conjunctivitis.3

Published cases implicate multiple routes of infection for neonatal *Pasteurella* infections as follows: vertical (including chorioamnionitis),3,4 animal exposure (atraumatic more often than traumatic),3,5 horizontal,6,7 and unclear route of transmission.8,10 Some reports infer transmission route from history alone. While some reports have isolated *Pasteurella* in the infant plus an additional culture (e.g., maternal vaginal culture to infer vertical transmission), ours adds to only four previous reports to confirm the route via molecular diagnostic testing.6,11–13 as summarized in Table 1. Andersson and colleagues reported a case of fatal congenital pneumonia due to vertical transmission of *Pasteurella* confirmed by Southern blot analysis of isolates from the infant, mother, and household cat (to which the infant was unexposed).6 Ours is the first report of vertical transmission via deoxyribonucleic acid (DNA) sequencing of the 16S rRNA gene of the maternal and infants’ *P. multocida* isolates with assignment of the subspecies septica. Furthermore, our identification of the subspecies, septica is an important addition to the neonatal literature, where subspecies reports are sparse. Three subspecies of *P. multocida* are known as multocida, septica, and gallicida. Though reports are limited, only subspecies septica has been reported in neonatal infection.6,12,13 Subspecies multocida has been implicated in a case of meningitis in an older infant related to traumatic animal exposure.14

More detailed identification of the isolates and subspecies septica, in particular, will improve understanding of their virulence and pathogenesis of infection when virulence associated genes are further characterized,13 hopefully relating how this organism and subspecies

**Table 1** Molecular epidemiology reports of neonatal *Pasteurella multocida* infection by age at presentation

| Study (year) | Infant’s age | Infant’s infection | Organism | Confirmed route of transmission | Source transmission | Molecular method |
|--------------|--------------|--------------------|----------|---------------------------------|---------------------|------------------|
| Andersson et al (1994)6 | < 12 hours | Pneumonia (nasopharynx, postmortem lung cultures) | *P. multocida*, subspecies septica | Vertical | Mother’s cervical canal and cat’s tonsil | Southern blot |
| This report (2022) | < 12 hours | Septicemia, pneumonia | *P. multocida*, subspecies septica | Vertical | Mother’s vaginal swab | 16S rRNA gene sequencing |
| Siahaniidou et al (2012)11 | 17 days | Meningitis, septicemia | *P. multocida* | Horizontal | Grandmother’s nasopharynxx⁴ | PFGEb² |
| Boyanton et al (2016)12 | 17 days | Meningitis, septicemia | *P. multocida*, subspecies septica | Nontraumatic animal exposure | Cats (n = 2) gingiva | Repetitive sequence-based PCRb² |
| Boerlin et al (2000)13 | 1 month | Meningitis | *P. multocida*, subspecies septica | Nontraumatic animal exposure | Cat’s tonsil | PFGEb² |

Abbreviations: PCR, polymerase chain reaction; PFGE, pulsed-field gel electrophoresis.

xGrandmother had a pet dog, and an identical organism was isolated from dog nasopharynx.

²Infant cerebrospinal fluid isolate and household cats were compared.
are opportunistic in the neonate’s relatively immature immune system.

Addition of molecular diagnostics to the literature is important to better understand its characteristics, hosts, transmission, and pathogenesis in neonatal infection. Notably, one report of neonatal sepsis and meningitis from Pasteurella multocida may have implicated a cat’s positive mouth culture in atraumatic animal transmission but molecular testing showed that the infant and cat strains were different.16

**Conclusion**

In conclusion, we present a case of neonatal Pasteurella multocida, subspecies septica, sepsis, and pneumonia with genetically confirmed vertical transmission. Unusual infections must be considered in the care of newborns with EOS. In such cases, further investigation via molecular diagnostics enhances precise microbial identification and understanding to enable best care for pregnant women and neonates.

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None.

**Conflicts of Interest**

The authors declare that there are no conflicts of interest.

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**References**

1 Simonsen KA, Anderson-Berry AL, Delair SF, Davies HD. Early-onset neonatal sepsis. Clin Microbiol Rev 2014;27(01):21–47
2 American Academy of Pediatrics. Pasteurella infections. In: Kimberlin DW BM, Jackson MA, Long SS, eds. Red Book: 2018 Report of the Committee on Infectious Diseases. Itasca, IL: American Academy of Pediatrics; 2018:606–607
3 Nakwan N, Nakwan N, Atta T, Chokeaibulkit K. Neonatal pasteurellosis: a review of reported cases. Arch Dis Child Fetal Neonatal Ed 2009;94(05):F373–F376
4 Puwanant M, Chanvitan P. Neonatal septicemia due to Pasteurella multocida: the first case report in Thailand. J Med Assoc Thai 2006;89(08):1293–1296
5 Guet-Revillet H, Levy C, Andriantahina I, et al. Paediatric epidemiology of Pasteurella multocida meningitis in France and review of the literature. Eur J Clin Microbiol Infect Dis 2013;32(09):1111–1120
6 Andersson S, Larinkari U, Vartiola T, et al. Fatal congenital pneumonia caused by cat-derived Pasteurella multocida. Pediatr Infect Dis J 1994;13(01):74–75
7 Guillet C, Join-Lambert O, Carbonnelle E, Ferroni A, Vachée A. Pasteurella multocida sepsis and meningitis in 2-month-old twin infants after household exposure to a slaughtered sheep. Clin Infect Dis 2007;45(06):e80–e81
8 Haase R, Stiefel M, Merkel N, Knopp W, Sauer H, Lieser U. [Pasteurella multocida as a rare cause of neonatal meningitis]. Z Geburtshilfe Neonatol 2006;210(06):219–221
9 Jha P, Kalyoussef S. Fulminant septic shock with Pasteurella multocida in a young infant: no bite, scratch, or lick!. Vector Borne Zoonotic Dis 2021;21(01):59–62
10 Spadafora R, Pomero G, Delogu A, Gozzoli L, Gancia P. A rare case of neonatal sepsis/meningitis caused by Pasteurella multocida complicated with status epilepticus and focal cerebritis. Pediatri Med Chir 2011;33(04):199–202
11 Siahanidou T, Gika G, Skiathtoi AV, et al. Pasteurella multocida infection in a neonate: evidence for a human-to-human horizontal transmission. Pediatr Infect Dis J 2012;31(05):536–537
12 Boyanton BL Jr., Freij BJ, Robinson-Dunn B, Makin J, Runge JK, Luna RA. Neonatal Pasteurella multocida subs. septica meningitis traced to household cats: molecular linkage analysis using repetitive-sequence-based PCR. J Clin Microbiol 2016;54(01):230–232
13 Boerlin P, Siegrist HH, Burnens AP, et al. Molecular identification and epidemiological tracing of Pasteurella multocida meningitis in a baby. J Clin Microbiol 2000;38(03):1235–1237
14 Schuur PM, Haring AJ, van Belkum A, Draaisma JM, Buiting AG. Use of random amplification of polymorphic DNA in a case of Pasteurella multocida meningitis that occurred following a cat scratch on the head. Clin Infect Dis 1997;24(05):1004–1006
15 Wilson BA, Ho M. Pasteurella multocida: from zoonosis to cellular microbiology. Clin Microbiol Rev 2013;26(03):631–655
16 Wood AS, Foraker EE, Di Pentima C. Molecular epidemiology in neonatal pasteurellosis. Pediatr Infect Dis J 2013;32(12):1402