Case report

Transient atelectasis due to hilar lymph node swelling affected by lenalidomide-induced tumor flare reaction

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Tumor flare reaction (TFR) is a unique immune-mediated tumor recognition phenomenon presenting as rapid enlargement of the tumor, which mimics disease progression, developing in the early stage of treatment using immunomodulatory drugs or immune checkpoint inhibitors. A 59-year-old man with follicular lymphoma had residual tumor burden in the left hilar lymph nodes after R-CHOP therapy, and received lenalidomide and rituximab (R2) therapy. He developed respiratory distress on day 11 of R2 therapy. Chest X-ray and CT demonstrated left lung atelectasis due to left hilar lymph node swelling. We performed transbronchial lung biopsy on day 20 of R2 therapy. The biopsied left bronchus tissue exhibited extensive necrosis, which had a B-cell phenotype consistent with that of follicular lymphoma. Neither NK cells nor cytotoxic T cells were detected. It was unclear whether the immune effector cells disappeared at the time of transbronchial lung biopsy. Atelectasis in our patient improved by continuing R2 therapy beyond TFR.

Keywords: Tumor flare reaction, Pseudoprogression, Immunomodulatory drug

INTRODUCTION

Lenalidomide, an immunomodulatory drug, was reported to reactivate dysfunctional T and natural killer (NK) cells ex vivo by increasing their proliferative capacity and T-helper cell type 1 (Th1) cytokine release.1 Tumor flare reaction (TFR) is a unique immune-mediated tumor recognition phenomenon presenting as an increase in tumor burden, low-grade fever and rash. Lenalidomide-induced TFR involves the activation of NK cells and T cells, and their infiltration into the tumor sites.2 TFR was originally described in patients with chronic lymphocytic leukemia (CLL) treated using immunomodulatory drugs (IMiDs) (thalidomide and lenalidomide)3,4 TFR was also observed in mantle cell lymphoma, indolent non-Hodgkin lymphoma (NHL), aggressive NHL and Hodgkin lymphoma treated by lenalidomide.5

TFR mimics disease progression on imaging before an effective anti-tumor response occurs. A similar phenomenon, ‘pseudoprogression’, was also reported in multiple solid tumor types treated using immune checkpoint inhibitors (ICIs) resulting from T cells infiltrating the tumor site.6

We report a patient with refractory follicular lymphoma who exhibited transient atelectasis due to hilar lymph node swelling affected by lenalidomide-induced TFR.

CASE REPORT

A 58-year-old man visited a hospital in October 2019 for lymph node swelling and the left inguinal lymph node was biopsied. He was diagnosed with follicular lymphoma grade 3a (Figure 1 A–B) and referred to our hospital. We diagnosed his lymphoma as follicular lymphoma grade 3a, stage IV and FLIPI: high. He administered bendamustine, but it was ineffective. He then received R-CHOP therapy and had a partial response. After 5 courses of R-CHOP therapy, left hilar lymph node swelling remained on PET/CT with a SUV of 20.5 as the main lesion (Figure 2).

We started R2 (lenalidomide at 20 mg/day, days 1-21 and rituximab at 375 mg/m2, day 1) therapy in June 2020 (day 1). At that time, the WBC count was 3,870/μl (3,500-8,500), containing 2,593/μl of neutrophils and 368/μl of monocytes. Hb was 12.4 g/dl (11.5-17.0), platelet count was 25.4 × 104/μl (15.0-35.0), LDH was 387 U/L (120-200) and CRP was 0.57 mg/dl (≤ 0.30). He was in a good condition on day 4 of R2 therapy. Eleven days after starting R2 therapy, he visited our hospital for respiratory discomfort. His body temperature was 36.5˚C, blood pressure was 129/62 mmHg and SpO2 (room air) was 89-90%. He exhibited grade 1 rash, but had no pain. The WBC count was 2,120/μl, containing 922/μl of neutrophils and 424/μl of monocytes. Atelectasis in our patient improved by continuing R2 therapy beyond TFR.

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Fig. 1. Pathological findings: The inguinal lymph node at onset (A x 40, B x 400); The neoplastic follicles show a vaguely nodular pattern (A). Both centrocytes and centroblasts were present (B). The biopsied left bronchus tissue (C–J x 200); On hematoxylin and eosin staining, dense infiltration of lymphocytes, which were almost all necrotic, was observed under the bronchial epithelium (C). Although they had necrotic change, they were CD10-positive (D), CD20-positive (E), PAX-5-positive (not shown) and bcl-2-positive cells (F), which is consistent with the phenotype of follicular lymphoma. There were few CD3-positive cells (G), which were negative for granzyme B (H), i.e., not cytotoxic T cells. CD56-positive cells, i.e. NK cells, were not detected (I). CD68-positive cells considered to be macrophages were well noted (J).
was relatively high at 20% of the WBC. Hb was 13.6 g/dl, platelet count was 29.3 × 10^4/μl, LDH was 264 U/L and CRP was 0.99 mg/dl. Chest X-ray revealed a left lung severe shadow (Figure 3). CT demonstrated left lung atelectasis due to left hilar obstruction by lymph node swelling (Figure 2).

After emergency hospitalization on day 11 of R2 treatment, he was stable on oxygen inhalation of 0.5 L/min at rest and 2 L/min when walking. Antihistamine was prescribed for rash. We continued the combination lenalidomide-rituximab (R2) immunotherapy. Although rituximab was administered 6 times through 5 courses of R-CHOP therapy and on day 1 of this course, it was added on day 13. The left hilar lymph node swelling and atelectasis did not improve on chest X-ray on day 18 (Figure 3). If the obstruction of the left bronchus was not due to TFR but to true progression, radiation therapy was considered necessary. We thus carried out transbronchial lung biopsy on day 20 of R2 treatment. The biopsied left bronchus tissue exhibited dense infiltration of lymphocytes, which were almost all necrotic. Although they had necrotic change, their phenotype was consistent with that of follicular lymphoma. Neither NK cells nor cytotoxic T cells were detected. Macrophages were well noted. (Figure 1 C–J).

Rituximab was added just after transbronchial lung biopsy on day 20. From around this time, his respiratory state improved. Chest X-ray on day 25 revealed a decrease in left hilar lymph node swelling and improvement of atelectasis. He no longer needed oxygen inhalation and was discharged. As chest X-ray on day 28 (Figure 3) revealed left lung expansion, he received a second cycle of R2 therapy from day 29. The left lung atelectasis expanded through the second cycle of R2 therapy (Figure 3). Central nervous system involvement developed thereafter and we changed R2 therapy to R-CHASE therapy, which contains high-dose cytarabine, and intrathecal administration of methotrexate, cytarabine and prednisolone.

**DISCUSSION**

TFR is acutely dependent on NK cell function, and is then maintained by the rapid recruitment and proliferation of T cells. Andritsos et al. reported increased CD3-positive, CD4-positive, CD8-positive and granzyme B-positive T-cells in an excised swollen tonsil after lenalidomide treatment. In the present case, the biopsied left bronchus tissue on day 20 of R2 therapy did not contain NK cells or cytotoxic T cells. Macrophages were relatively conspicuous. It is unclear whether NK cells and cytotoxic T cells disappeared at the time of transbronchial lung biopsy when the respiratory state began improving. Although it is possible that the left hilar lymph nodes swelled due to tumor progression without TFR, rapid enlargement just after the start of R2 therapy may have been related to TFR.

R2 therapy was reported to have favorable activity in patients with relapsed/refractory follicular lymphoma.
Wang et al. found that lenalidomide with rituximab is effective even for transformed large cell lymphoma originating from follicular lymphoma. In a randomized study for relapsed/refractory diffuse large B-cell lymphoma, patients treated using lenalidomide had a longer progression-free survival than those treated at the investigator’s discretion (gemcitabine, rituximab, etoposide or oxaliplatin). Based on the resistance to chemotherapy in the present case, we expected complete remission via a unique mechanism of action from the combination of lenalidomide and rituximab rather than the intensity of salvage chemotherapy. Therefore, we selected R2 therapy even if transformation was possible.

Immunotherapy, such as IMiDs and ICIs, works differently from chemotherapy and takes more time to exhibit effects than cytotoxic drugs. Chanan-Khan et al. reported that TFR induced by immunomodulatory drugs, such as lenalidomide, develops in >90% of patients during the first treatment cycle and the median time to onset is 6 days. Goy et al. also found that TFR generally developed during the first cycle of lenalidomide, with few events during later cycles. Our patient exhibited a typical clinical course regarding the time of TFR. Steroids are used for the management of severe cases of TFR and prophylaxis. Chong et al. reported that patients who received once weekly low-dose (10 mg) dexamethasone had fewer dose interruptions for TFR during the first 8 weeks of lenalidomide. Our patient did not receive dexamethasone, but he was administered 100 mg of hydrocortisone once just before the start of rituximab on day 1, day 13 and day 20. Hydrocortisone acts for a short time, unlike dexamethasone. The present patient did not receive any steroids after hydrocortisone on day 1, until he received hydrocortisone on day 13. If dexamethasone had been administered weekly in the first course, TFR may have been less severe.

It is difficult to differentiate pseudo-progression due to TFR from true progression only by imaging, which largely relies on the tumor size. The identification of TFR is important to avoid the premature discontinuation of effective therapy because the intensity of TFR is correlated with the probability of achieving a complete response. TFR should be paid attention to in the early stage of treatment using immunomodulatory drugs.

CONFLICT OF INTEREST

The authors declare that they have no conflicts of interest.

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