Evaluation of *Helicobacter pylori* Infection, Neutrophil–Lymphocyte Ratio and Platelet–Lymphocyte Ratio in Dyspeptic Patients

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Abstract: Recent studies have shown a correlation between *Helicobacter pylori* (*H. Pylori*) infection and the neutrophil/lymphocyte ratio (NLR) and platelet/lymphocyte ratio (PLR). The aim of this study was to investigate the relationship between *H. Pylori* infection and hematimetric indices in patients with dyspepsia symptoms. Overall, 448 patients who underwent gastroscopy were analyzed retrospectively. Histopathological evaluation of biopsies according to *H. pylori* presence was classified as *H. Pylori* positive and negative groups, which are analyzed in relation with hematimetric indices. NLR and PLR measurements did not show a statistically significant difference between *H. pylori* negative and positive groups (p > 0.05). NLR revealed a negative correlation between hemoglobin (HGB), iron, and ferritin measurements in the correlation analysis of the *H. Pylori* positive group (r = −0.133, p = 0.031; r = −0.270, p = 0.002; r = −0.162, p = 0.032). Again, with PLR, there was a negative correlation between HGB, mean corpuscular volume (MCV), iron, and ferritin measurements (r = −0.310, p = 0.001, r = −0.187, p = 0.002, r = −0.335, p = 0.001; r = −0.290; p = 0.001). The results of our study do not reveal an association between *H. pylori* presence and inflammatory response, which is evaluated by NLR and PLR measurements in patients with dyspepsia. However, low serum iron and ferritin values of *H. pylori*-positive patients suggest the effect of *H. pylori* on iron metabolism.

Keywords: *Helicobacter pylori*; dyspepsia; neutrophil; lymphocyte; platelet; iron

1. Introduction

*Helicobacter pylori* (*H. pylori*) is one of the most common chronic bacterial infections in humans. It affects about 50% of the world’s population. Generally, the infection is seen more frequently and acquired earlier in developing countries when compared to developed countries due to various factors, such as geographical location, ethnic features, sanitation conditions, economical structure of the country [1].

Dyspepsia is a collection of symptoms, which has a differential diagnosis and heterogeneous pathophysiology. It is seen at least in 20% of the population [2]. While 25% of patients with dyspepsia symptoms have an underlying organic cause (organic dyspepsia), 75% of them have no underlying organic cause in diagnostic evaluation (functional dyspepsia) [3,4]. *H. pylori* gastritis is one of the prominent factors in the pathophysiology of functional dyspepsia (FD). *H. pylori* infection among dyspeptic patients is estimated to be up to 70%, although it showed regional differences [5].
The number of white blood cells (WBC), neutrophil, lymphocyte, platelet count (PLT) and mean platelet volume (MPV) values and ratios between them are used as an inflammatory indicator. Neutrophil/lymphocyte ratio (NLR) and platelet/lymphocyte ratio (PLR) are the most important of these indicators and they were studied recently to determine the severity of various diseases, gastric cancers, and H. pylori infection [6,7]. Neutrophils are the representative components of the innate immune system and lymphocytes represent the adaptive immune system. Furthermore, most of the studies have demonstrated that increased NLR and PLR are associated with active disease associated with known inflammatory markers [8].

Iron deficiency is associated with increased susceptibility of infections, as well as impaired physical and cognitive functions with or without anemia. Since the first case of H. pylori infection with iron deficiency anemia reported in 1991, several explanations for the role of H. pylori on iron metabolism have been speculated [9]. Although the relationship is not fully understood, bleeding due to chronic gastritis, absorption defects due to achlorhydria or recently elevated hepcidin levels are blamed [10].

From the point of view that H. pylori had been linked to various hematologic manifestations, we aimed to determine the relation between the H. pylori infection and the hematimetric parameters, particularly NLR and PLR values in patients with dyspeptic symptoms.

2. Materials and Methods

This study is conducted retrospectively at our Internal Medicine Department among 448 outgoing patients and patients who had gastroscopy. Of these, we tested for H. pylori and simultaneously performed a complete blood count to make our selection according to inclusion and exclusion criteria. The study includes the adult patients between 17 and 83 years old who had a gastroscopy and checked for H. pylori levels in our hospital’s pathology and endoscopy records. Endoscopic-pathological diagnosis, H. pylori positive–negative cases, white blood cell count (WBC), neutrophil, lymphocyte, platelet count, iron, vitamin B12, folic acid, total iron binding capacity (TIBC), ferritin, and C-reactive protein (CRP) levels of the patients who are subjects of this study are checked. H. pylori positive and negative cases gathered together in the pathology database according to the population at hand. Patients with (1) gastric cancer, (2) gastric lymphoma, (3) pregnancy, (4) the gastroduodenal surgery, (5) patients who have used antibiotics for up to 4 weeks before the study and (6) who have already undergone H. pylori eradication treatment, (7) chronic systemic inflammatory disease were excluded from the study.

Neutrophil/lymphocyte ratio (NLR) and platelets/lymphocyte ratio (PLR) are the leukocyte indexes recommended as inflammatory indicators. In our study, the relation between H. pylori infection results and neutrophil/lymphocyte ratio (NLR) and platelet/lymphocyte ratio (PLR) is determined.

3. Endoscopic and Histopathologic Evaluation

The study includes the cases where gastroscopy had been performed (OLYMPUS GIF TYPE XP190N). Biopsies were taken from antrum and corpus, which were evaluated in terms of H. pylori. Findings of the biopsies for H. pylori by gastroscopy are examined and recorded retrospectively. All subjects gave their informed consent for inclusion before they participated in the study. This study was conducted in accordance with the Declaration of Helsinki, and the protocol was approved by the Ethics Committee of University of Health Sciences GOP Taksim Education and Research Hospital on 3 May 2017. (Protocol number: 39).

4. Statistical Analyses

Frequency, ratio, mean, minimum, maximum, and standard deviation values were used in the descriptive statistics to determine continuous variables. A Student t-test was used for comparison of independent and normally distributed two variables. A Mann–Whitney U test was performed for comparison of independent and non-normally distributed variables. The Pearson chi-square test and Fisher’s exact test were performed to determine differences between categorical variables. Multiple
linear logistic regression analysis was performed to determine the effect levels of the parameters. Spearman’s correlation tests were used for the correlation analysis. Statistical significance was assessed at \( p < 0.05 \). Statistical analysis was performed by using Number Cruncher Statistical System (NCSS, Kaysville, UT, USA, 2007).

5. Results

Clinical and biochemical characteristics according to the presence of \( H. pylori \) are summarized in Table 1. A total of 263 \( H. pylori \)-positive patients with a mean age of 47.87 ± 13.98 years, comprising 188 females and 75 males, and a total of 185 \( H. pylori \)-negative patients with a mean age of 51.30 ± 14.10 years, comprising 130 females and 55 males, were enrolled in the study. The mean age of \( H. pylori \) positive group was significantly lower than the \( H. pylori \) negative group (\( p = 0.011, p < 0.05 \)). There were no differences in hemoglobin (HGB), mean corpuscular volume (MCV), leukocyte, neutrophil, lymphocyte, platelet, NLR, and PLR measurements between \( H. pylori \)-negative patients and \( H. Pylori \)-positive patients (\( p > 0.05 \)). Patients with \( H. pylori \) had statistically significantly lower iron and ferritin compared to those without \( H. pylori \) (\( p = 0.029; p = 0.003 \)). NLR revealed a statistically significant negative correlation between HGB, iron and, ferritin measurements in the correlation analysis of \( H. pylori \) positive group (\( r = -0.133, p = 0.031; r = -0.270, p = 0.002; r = -0.162, p = 0.032 \). Again, with PLR, there was a statistically negative correlation between HGB, MCV, iron and, ferritin measurements (\( r = -0.310, p = 0.001, r = -0.187, p = 0.002, r = -0.335, p = 0.001; r = -0.290; p = 0.001 \) (Table 2). There was no statistical significance regarding both the PLR and NLR ratio distributions between the two groups (Figure 1).

Table 1. Demographic and biochemical characteristics according to the presence of \( H. pylori \).

| \( H. pylori \) | Negative (185) Mean ± sd, n (%) | Positive (263) Mean ± sd, n (%) | \( p \) |
|----------------|---------------------------------|---------------------------------|------|
| Age (years)    | 51.30 ± 14.10                  | 47.87 ± 13.98                  | 0.011* |
| Gender Male    | 55 (29.7)                      | 75 (28.5)                      | 0.781 |
| Female         | 130 (70.3)                     | 188 (71.5)                     |      |
| HGB (g/dL)     | 12.66 ± 1.56                   | 12.59 ± 1.67                   | 0.501 |
| MCV (fL)       | 87.57 ± 6.85                   | 86.55 ± 7.57                   | 0.185 |
| Leukocyte (µL) | 682.70 ± 1607.57               | 694.12 ± 1973.45               | 0.647 |
| Neutrophil (µL)| 3781.03 ± 1273.28              | 3995.82 ± 1559.09             | 0.278 |
| Lymphocyte (µL)| 2278.65 ± 580.43              | 2380.87 ± 1013.54             | 0.373 |
| NLR (%)        | 1.74 ± 0.65                    | 1.92 ± 1.00                    | 0.338 |
| PLR (%)        | 118.91 ± 38.66                 | 125.59 ± 47.42                 | 0.237 |
| Platelet (µL)  | 258,718.92 ± 66,735.34         | 264,551.33 ± 75,162.24         | 0.502 |
| Iron (µg/dL)   | 67.28 ± 39.82                  | 56.45 ± 33.64                  | 0.029* |
| TIBC (µg/dL)   | 356.01 ± 60.33                 | 363.94 ± 60.20                 | 0.268 |
| Ferritin (ng/mL)| 36.04 ± 42.36                 | 30.11 ± 52.80                  | 0.003** |
| B12 (pg/mL)    | 268.77 ± 158.22                | 278.53 ± 186.21                | 0.859 |
| Folic acid (ng/mL)| 8.72 ± 4.15               | 7.27 ± 2.77                    | 0.081 |
| CRP (mg/L)     | 6.04 ± 6.22                    | 5.28 ± 4.48                    | 0.530 |
| Intestinal metaplasia | 26 (14.1)           | 48 (18.3)                      | 0.239 |
| Atrophy        | 6 (3.2)                        | 18 (6.8)                       | 0.096 |
| Gastritis      | 156 (84.3)                     | 219 (83.3)                     | 0.766 |
| Gastric ulcer  | 31 (16.8)                      | 45 (17.1)                      | 0.933 |
| Duodenal ulcer | 14 (7.6)                       | 27 (10.3)                      | 0.329 |
| Esophagitis    | 49 (26.5)                      | 77 (29.3)                      | 0.518 |

Abbreviations: HGB, hemoglobin; MCV, mean corpuscular volume; TIBC, total iron binding capacity; NLR, neutrophil/lymphocyte ratio; PLR, platelet/lymphocyte ratio; CRP, c-reactive protein. * Student t-test, ** Mann–Whitney U test, \( \chi^2 \) Pearson’s chi-squared test, \( * p < 0.05, ** p < 0.01 \).
While prevalence is 40–50% in developed countries, it is 70–90% in underdeveloped or developing countries and even for some age groups, it reaches 100% [11]. One of the attention-grabbing findings of our study is the fact that H. pylori-positive patients are younger when compared to H. pylori-negative patients. This shows that H. pylori infection is acquired in early ages. In a similar manner, study from Eastern Europe revealed 72.1% positivity for H. pylori among 459 adults who were tested by urea breathe test [12]. Another study from Norway with 1624 participants from all ages demonstrated that the prevalence increases gradually with age, except for in the age group above 70 years. They hypothesized that age-related positivity might be the result of lifestyle factors and social activity which broadens common use areas [13].

It is shown that H. pylori has a large spectrum of both metabolic and non-metabolic extra-gastric manifestations. A striking study by Isave et al. demonstrated the colonization of H. pylori in the hepatobiliary system of cirrhosis patients with a frequency of 50% [14]. Their findings support the

### Table 2. Comparison of NLR and PLR with other parameters in H. pylori-positive patients.

| Parameter                      | N   | NLR | PLR |
|-------------------------------|-----|-----|-----|
| Age (years)                   | 263 | 0.020 | 0.743 |
| HGB                           | 263 | -0.133 | 0.031* |
| MCV                           | 263 | -0.084 | 0.173 |
| Leukocyte (µL)                | 263 | 0.302 | 0.001** |
| Neutrophil (µL)               | 263 | 0.644 | 0.001** |
| Lymphocyte (µL)               | 263 | -0.596 | 0.001** |
| Platelet (µL)                 | 263 | 0.032 | 0.606 |
| Iron (µg/dL)                  | 131 | -0.270 | 0.002** |
| TIBC (µg/dL)                  | 129 | 0.016 | 0.853 |
| Ferritin (ng/mL)              | 175 | -0.162 | 0.032* |
| B12 (pg/mL)                   | 177 | -0.035 | 0.644 |
| Folic acid (ng/mL)            | 81  | -0.078 | 0.490 |
| CRP (mg/L)                    | 52  | 0.006 | 0.967 |

Abbreviations: HGB, hemoglobin; MCV, mean corpuscular volume; TIBC, total iron binding capacity; NLR, neutrophil/lymphocyte ratio; PLR, platelet/lymphocyte ratio; CRP, c-reactive protein. r: Spearman’s correlation coefficient, *p < 0.05, **p < 0.01.

![Figure 1. PRL and NLR distribution according to H. pylori presence.](image-url)
studies that were pointed out toward the \textit{H. pylori} infection and hepatocellular carcinoma \cite{15}. On the other hand, the relationship between \textit{H. pylori} infection and cardiovascular diseases, chronic metabolic diseases, such as diabetes mellitus, and hyperlipidemia has been argued \cite{16,17}. \textit{H. pylori} on a human host causes neutrophils and monocyte stimulation and many pro-inflammatory cytokine releases that cause gastric mucosal injury. Although \textit{H. pylori} is a noninvasive organism, because of the antigenic substances it produces, such as heat shock protein, urease, and lipopolysaccharide, it activates T cells. With an improved antigen presentation, IL-1, IL-6, IL-8, inflammatory cytokines, such as tumor necrosis factor-alpha (TNF-alfa), are released. In addition to that, B-cell response is produced both locally and systemically \cite{18}. Activation of leukocytes occurs as a result of chronic inflammation; in other words, leukocytes have a role in chronic diseases, such as diabetes, hypertension, atherogenesis, thrombosis formation and other inflammatory disorders \cite{19}.

In our study, we investigated the \textit{H. pylori} infection activity in patients with dyspepsia symptoms and the relationship between inflammatory markers, neutrophil/lymphocyte ratio and platelets/lymphocyte ratio. In this study we conducted, we could not observe a significant difference in leukocyte, lymphocyte, platelet, NLR and PLR levels between positive/negative \textit{H. pylori} patients. Guclu et al. have evaluated patients with antral gastritis with endoscopy in the study they carried out in 2017. These patients were classified in four groups as \textit{H. pylori} negative, mild, moderate and severe and in the study conducted on 199 patients, it was observed that patients that were \textit{H. pylori} positive had a lower NLR compared to patients with negative \textit{H. pylori}. However, peripheral blood lymphocyte values and platelet counts in patients that were severely \textit{H. pylori} positive had been detected to be higher compared to negative patients \cite{20}. In another study, patients that were \textit{H. pylori} positive had been observed to have higher leucocyte, neutrophil and NLR levels compared to \textit{H. pylori}-negative patients, independent from the bacterial Cytotoxin-linked genes, without taking the Cag A conditions and peptic ulcer into consideration \cite{7}. Nalbant et al. conducted a study on 91 patients who had gastrointestinal endoscopy upon dyspeptic complaints. They found out that patients who were \textit{H. pylori} positive had an apparent reduction in the number of neutrophils, lymphocytes and NLR in their blood, where only the neutrophils decrease was determined to be statistically significant \cite{21}. We think that the lower number of NLR in the \textit{H. pylori} positive group might be related to the small sample size. Nalbant et al. also determined that within the same study, the platelet values of the \textit{H. pylori} positive group were lower, compared to the \textit{H. pylori} negative group. In a study conducted by Asil et al. in 2016 with 286 patients who had chronic \textit{H. pylori} infection and 130 \textit{H. pylori} negative control group, it was found that patients with chronic \textit{H. pylori} infection had a higher NLR level compared to the \textit{H. pylori} negative group and they observed a decline in NLR with \textit{H. pylori} eradication. In this study, Asil et al. report a remarkable finding that \textit{H. pylori} eradication leads to a decline in NLR rates \cite{22}.

Reviewing the literature, it is suggested that this might be an important marker to foresee the PLR in prognosis of hepatocellular carcinoma, breast cancer and lung cancer \cite{23–25}. In addition, it may be effective in demonstrating the seriousness of coronary artery disease and cardiac damage due to atherosclerotic plaque damage \cite{26}. When all these studies are evaluated together, PLR is expected to be affected by \textit{H. pylori} infection, which causes a chronic inflammation. Farah et al. have conducted studies on this topic in 2014 and 2017, suggesting that NLR and PLR of \textit{H. Pylori}-positive patients are higher than \textit{H. Pylori}-negative patients \cite{27,28}. Shimoyama et al. have studied the relationship between NLR–PLR and mortality in the presence of a gastrointestinal perforation, which is a heavy inflammatory case and scales that reflect other inflammatory cases. Their study has retrospectively evaluated 32 patients with gastrointestinal perforation; it has found NLR and PLR to demonstrate mortality with patients who have gastrointestinal perforation much better compared to other prognostic indices that show inflammation \cite{29}. We could not, however, find a correlation between the presence of \textit{H. pylori} infection and NLR and PLR, in our study. Umit et al. have observed that patients with \textit{H. Pylori} have lower platelet counts compared to patients who are \textit{H. Pylori} negative, in their retrospective search conducted on 4823 patients in 2015. Even though these patients had platelet counts within the normal range, since they had a lower platelet count compared to the \textit{H. pylori} negatives, it was suggested
that *H. pylori* reduces the platelet values before immune cytopenia effects the platelets [30]. When we evaluate our work, *H. pylori*-positive patients and *H. pylori*-negative patients have not demonstrated a statistically significant difference. This finding can be explained by the fact that our study had fewer patients contributing compared to the study of Umit et al.

In our study, *H. pylori*-positive patients had lower folic acid, iron and ferritin levels than the negative ones (Table 1). Several pathways are blamed for decreased iron stores in the *H. pylori*-positive patients, including chronic gastritis with achlorhydria, reduced ascorbic acid secretion in the gastric mucosa, iron uptake by the bacterium, and bleeding ulcers related to *H. pylori* [31]. However, the studies that assess the effect of *H. pylori* eradication therapy on the improvement of iron deficiency are controversial [32]. Some of the studies hypothesize that eradication therapy for *H. pylori* improves the response of iron replacement therapy, which was previously refractory [33,34]. Therefore, additional studies are needed to clarify whether there is a casual relationship between *H. pylori* and iron deficiency. Furthermore, we determined that the NLP and PLR are inversely correlated with serum iron and ferritin levels (Table 2). The importance of iron for immunity appears in immune cell proliferation, especially lymphocytes [35]. This may be a possible explanation for the correlation between iron related parameters and NLP and PLR.

There are some limitations of this study. First the retrospective design of the study makes it difficult to establish the cause and effect relationship. Second, we practiced on hematimetric indices to assess the inflammation of *H. pylori*-positive patients. However, it would certainly be better to add the measurement of other known inflammatory markers such as TNF-alfa, IL-1, IL-6 and IL-8. Finally, some of the patients had a limited number of biopsies, which can reduce the accuracy regarding *H. pylori* infection.

7. Conclusions

No correlation between *H. pylori* infection, which is a case of chronic inflammation, and NLR–PLR, which are the strongest markers of inflammatory condition, was observed. On the other hand, *H. pylori* infection is associated with the presence of iron deficiency. Considering what *H. pylori* infection does in relation to the chronic inflammatory response and the markers of *H. pylori* infection, we believe more extensive and prospective studies are required in this field.

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