Characterization of a fungicidal substance produced by *Eupenicillium brefeldianum* isolated from soil for plant disease control and its significance in nature

Yen-Ting Chen and Wen-Hsiung Ko *

**Abstract**

**Background:** A fungus identified as *Eupenicillium brefeldianum* was isolated from soil amended with vegetable tissues.

**Result:** When grown in liquid medium prepared from the same vegetable tissues, *E. brefeldianum* produced a substance capable of preventing disease development of leaf spots of mustard cabbage caused by *Alternaria brassicicola* and inhibiting the germination of *A. brassicicola* conidia. The inhibitory substance was fungicidal and was very stable under high temperature and extreme pH. It was soluble in polar solvents but not soluble in non-polar solvents, and did not have charges on its molecule. This is the first discovery of the production of a fungicidal substance by this fungus.

**Conclusion:** Results from this study suggest the possession of a strong competitive saprophytic ability by *E. brefeldianum*, which in turn may explain the widespread occurrence of this fungus in soils.

**Keywords:** *Alternaria brassicicola*; Competitive saprophytic ability; *Eupenicillium brefeldianum*; Fungicidal substance

**Background**

Leaf spot of crucifers caused by *Alternaria brassicicola* (Schwein) Wiltshire, *A. brassicae* (Berk.) Sacc., and *A. raphani* Groves & Skolko are of worldwide importance (Farr et al. 1989). These pathogens have caused considerable losses in cruciferous oil production and vegetable crop yield (Humpherson-Jones and Maudo 1982; Kolte et al. 1987). In Taiwan, *A. brassicicola* is the causal organism of leaf spots on crucifers. The disease causes reduction in quality and yield of cruciferous vegetables all year round (Ho et al. 2007).

Control of foliar plant diseases relies heavily on the use of chemical fungicides (Agrios 2005; Hewitt 1998). However, development of alternative methods for the control of these diseases is needed because of the high concerns about the adverse effects of pesticides on human health and the environment (World Health Organization 1990). A project was, therefore, initiated to screen liquid cultures of soil microorganisms directly on leaves of mustard cabbage for ability to control leaf spots caused by *A. brassicicola* (Chen et al. 2011). Liquid culture of a fungus identified as *Eupenicillium brefeldianum* was found to be very effective in controlling the disease in the greenhouse. The fungus is famous for its ability to produce brefeldin A which has multiple functional and biochemical effects on experimental animals (Kamata et al. 1983; Klausner et al. 1992; Nebenfuhr et al. 2002). This fungus is found throughout the world in soil including agricultural, forest, orchard and pasture soils (Keller and Bidochka 1998; Souza-Motta et al. 2003). Little is known about the mechanism of its widespread occurrence in soil. The objectives of the study are to characterize the inhibitory substance produced by *Eupenicillium brefeldianum* and to explain why this fungus is so widespread in nature.

**Methods**

**Isolation of soil microorganisms**

Soil samples collected from central Taiwan were taken from a depth of 0–10 cm, sifted and moistened to about 65% water-holding capacity (Tsai et al. 2012). Selective media for isolation of fungi, actinomycetes and bacteria...
were prepared as described previously (Ko et al. 2010b). Approximately 1.3 g soil was mixed with 100 ml sterile distilled water in an Omni mixer chamber at 5000 rpm for 30 s. The suspension was diluted to $10^{-4}$, $10^{-5}$ and $10^{-6}$ for fungi, $10^{-5}$, $10^{-6}$ and $10^{-7}$ for actinomycetes and bacteria to determine the dilution needed to obtain soil suspension essentially free of each group of microorganisms (Ko et al. 2011). A 1-ml aliquot of diluted soil suspension was mixed with 20 ml of a molten selective medium at 45°C in a Petri plate. Five plates were used for each treatment.

Vegetables including fruit of tomato (Lycopersicon esculentum), tubers of sweet potato (Ipomoea batatas), and leaves and stems of spinach (Spinacia oleracea), ong choy (Ipomoea aquatica) and common purslane (Portulaca oleracea) were purchased from local markets. For soil amendment, about 500 g soil was mixed with 4% each of chopped vegetables in a 1000-ml bottle and incubated at 24°C for at least two weeks before use. To isolate microorganisms with ability to utilize amended nutrients for multiplication, suspension of amended soil was diluted to the concentration pre-determined for each group of microorganisms and plated on the selective medium as described above. After incubation at 24°C for seven days, colonies appeared were individually transferred to 10% V-8 agar (10% V-8 juice, 0.02% CaCO$_3$ and 2% agar) plates (Ko et al. 2010a).

**Cultivation of isolated microorganisms in liquid medium**

Liquid medium was prepared by grinding 4 g each of the five chopped vegetables in 100 ml water in an Omni mixer at 4000 rpm for 3 min and dispensing 50 ml broth in a 250-ml flask. After autoclaving, each flask was inoculated with two loopfuls of bacterium, or a piece (ca. 4 × 5 × 3 mm) of actinomycete or fungus agar culture. Inoculated flasks were incubated on a shaker for two weeks. After incubation, cultures were separately ground in an Omni mixer at 60, 80 or 100°C for 30 min or autoclaved for 15 min. To study the effect of high temperature on the activity of the fungicidal substance under extreme pH, the pH of the extract was adjusted to 2 or 12 for 24 h and then readjusted back to 4.

**Characterization of the fungicidal substance**

To study the effect of pH on the activity of the fungicidal substance, the pH of the culture extract of *E. brefeldianum* was adjusted from the original 4 to 3 with 1 N HCl, or 5 to 10 with 1 N NaOH (Ko et al. 2010b). To study the stability of the fungicidal substance under extreme pH, the pH of the extract was adjusted to 2 or 12 for 24 h and then readjusted back to 4.

**Extraction of the fungicidal substance**

Culture fluid of *E. brefeldianum* isolate V3F-3 was freeze dried. One gram dry powder, obtained from approximately 50 ml of culture fluid was extracted with 25 ml of water, ethanol, methanol, acetone, ethyl acetate, or chloroform in a 250-ml flask by shaking on a shaker for 24 h (Ko et al. 2010b). The mixture was centrifuged at 1500 × g for 5 min to obtain clear extract. For bioassay and characterization of the fungicidal substance, 10 ml extract was evaporated to 2 ml followed by addition of 2 ml water and evaporation to 2 ml again. To test the ability of different solvents to extract the inhibitory substances, 10-fold concentration of the extracts was used.

**Germination tests**

To test the effect of the culture extract on spore germination, 10 μl of conidial suspension ($2 \times 10^4$ spores/ml) of *A. brassicicola* was mixed with 10 μl of extract in a cavity of a sterile eight-cavity slide. Slides with spores were kept moist by placing each on a L-shaped glass rod in a 9-cm Petri plate containing 10 ml sterile distilled water. Germination was recorded after incubation at 24°C for 4 h, and 100 spores were counted in each of the three replicates. All experiments were done twice.
For the study of the effect of the treatment of culture extract with cation exchange resins, anion exchange resins or activated charcoal on its activity against germination of *A. brassicicola*, 5 g of Diaion SK1B cation exchange resins (equivalent to Amberlite 1R-120), Diaion SA 12A anion exchange resins (equivalent to Amberlite 1RA-420; Tai-Young Chemical Co., Kaoshiung, Taiwan) or activated charcoal (Sigma-Aldrich) was washed with 50 ml of distilled water three times by shaking over a six-hour period to remove possible inhibitory substances. Ten milliliter extract was shaken with 1 g cation exchange resins, anion exchange resins or activated charcoal in a 150-ml flask at 100 strokes/min for 24 h and filtered through a Whatman no. 1 filter paper. The filtrate was then used for germination tests (Ko et al. 2010b). In each of the three replicates, 100 spores were counted and all experiments were done at least twice.

### Results

#### Effect of culture on leaf spots

Among the 241 microorganisms isolated from soils amended with vegetable tissues tested, liquid cultures from one bacterium and five fungi were able to reduce the disease incidence of leaf spots of mustard cabbage caused by *A. brassicicola* from 90% in the control to 0-30%. Liquid culture of the fungus identified as *Eupenicillium brefeldianum* was able to completely suppress the disease development and inhibit spore germination of the pathogen (Table 1). Moreover the fungus is famous for its production of brefeldin A (Nebenfuhr et al. 2002) and widespread occurrence in nature (Keller and Bidochka 1998; Souza-Motta et al. 2003). It was, therefore, selected for further study.

#### Isolation of inhibitor and nature of its inhibition

Cultures of *Eupenicillium brefeldianum* were each extracted with a different solvent. The extracts were tested for their ability to inhibit spore germination of *A. brassicicola* after solvent evaporation with the exception of water extract which was tested directly without evaporation treatment.

### Table 1 Effect of liquid culture of *Eupenicillium brefeldianum* on symptom development on mustard cabbage leaves inoculated with conidia of *Alternaria brassicicola* and on conidial germination of the pathogen

| Treatment       | Symptom development (%) | Conidial germination (%) |
|-----------------|-------------------------|--------------------------|
|                 | Disease incidence (%)   | Lesion size (mm)         | Conidial germination (%) |
| Culture         | 0                       | 0                        | 0                        |
| Water (control) | 90                      | 3.7                      | 97                       |

*Two leaves were sprayed with the liquid culture once a day three times before inoculation on the 4th day. Each leaf was inoculated at five sites, and the disease incidence and lesion diameter were recorded after 3 days.*

*Spore suspension mixed with the same amount of extract was incubated at 28°C for 4 h. Data were means of three replicates.*

#### Table 2 Effectiveness of different solvents in extracting inhibitory substances from freeze-dried powder of liquid culture of *Eupenicillium brefeldianum* against germination of *Alternaria brassicicola* conidia

| Solvent       | Germination (%) | Control |
|---------------|-----------------|---------|
| Extract       | 4 h             | 24 h    | 4 h    |
| Water         | 2               | 67      | 100    |
| Ethanol       | 0               | 79      | 99     |
| Methanol      | 0               | 20      | 100    |
| Acetone       | 89              | 99      | 99     |
| Ether         | 89              | 99      | 99     |
| Chloroform    | 97              | 96      | 95     |
| Water (control)| 97              |         |        |

*Spore suspension mixed with the same amount of extract was incubated at 28°C. Data were means of three replicates.*

Germination rates were recorded after incubation for 4 h. The result showed that spore germination was inhibited nearly completely in water extract and completely in ethanol and methanol extracts. Extracts of acetone, ethyl acetate, ether and chloroform were essentially non-inhibitory, supporting 89 to 97% germination in comparison with 97% in water control (Table 2). When germination rates were recorded after incubation for 24 h, *A. brassicicola* germinated 67, 79 and 20% in extracts of water, ethanol and methanol, respectively. Methanol was, therefore, selected for extraction of the inhibitor from the freeze dried powder of *E. brefeldianum* for subsequent studies.

To determine if the antifungal effect of the inhibitor is fungicidal or fungistatic, conidial suspension of *A. brassicicola* was mixed with culture extract of *E. brefeldianum*. The mixture was incubated at 28°C for 24 or 48 h before centrifugation at 1500 × g for 5 min to remove the inhibitor. The conidial suspension was then spread on water agar and incubated at 28°C for 4 h to test their viability. The result showed that after exposure of conidia to the inhibitor for 24 and 48 h, essentially all of them became non-viable (Table 3), indicating that the antifungal effect of the inhibitor was fungicidal.

### Table 3 Germination of conidia of *Alternaria brassicicola* on water agar after exposure to culture extract of *Eupenicillium brefeldianum* for 24 or 48 h

| Exposure time (h) | Germination (%) |
|-------------------|-----------------|
| Extract           | Water agar      | Control |
| 24                | 0               | 7       | 99     |
| 48                | 0               | 4       | 100    |

*Spore suspension mixed with the same amount of extract was incubated at 28°C for 4 h. Data were means of three replicates.*
Effect of pH on the inhibitory activity
The inhibitory effect of the culture extract of *E. brefeldianum* against germination of *A. brassicicola* conidia was not affected when its pH was adjusted from the original 4.0 to 3.0. At pH 3.0, the germination rate was decreased from 91% in the control to 0% (Table 4). When the pH was increased from 4.0 to 5.0, the germination rate was increased from 0 to 59%. At pH ranging from 6.0 to 10.0, the extract was no longer inhibitory.

Effect of the extreme pH and high temperature on the stability of the fungicidal substance
To determine the effect of extreme pH on the stability of the fungicidal substance produced by *E. brefeldianum*, its culture extract was adjusted to pH 2.0 or 12.0 for 24 h before being adjusted back to the original pH of 4.0. The result showed that the fungicidal substance was very stable at pH 2.0 and 12.0. After 24 h exposure, the extract still inhibited conidial germination of *A. brassicicola* completely in comparison with the 98% germination in the water control (Table 5).

To study the effect of high temperature on the stability of the fungicidal substance, the extract was treated at 60, 80 or 100°C for 30 min or autoclaved for 15 min. The result showed that the fungicidal substance was very stable at temperature ranging from 60 to 100°C (Table 6). After 30 min exposure, the extract still caused complete inhibition of conidial germination of *A. brassicicola* in comparison with the 97% germination in the water control. Even after autoclaving for 15 min, the inhibitory effect of the fungicidal substance still remained unaffected.

Effect of the adsorptive material treatment on inhibitory effect of the fungicidal substance
When the culture extract of *E. brefeldianum* was shaken with cation exchange resins or anion exchange resins, the fungicidal substance still inhibited the germination of *A. brassicicola* completely in comparison with 98% germination in water control (Table 7). However, after shaking with activated charcoal, the fungicidal substance was no longer inhibitory to spore germination. The germination rate increased from 0 to 99%.

Discussion
Since soil contains abundant and diverse microorganisms (Alexander 1977), ability to colonize plant tissues under such environment is a good indication that *E. brefeldianum* possesses a strong competitive saprophytic ability

### Table 4
Conidial germination of *Alternaria brassicicola* in culture extract of *Eupenicillium brefeldianum* adjusted to various pH values

| pH value after adjustment | Germination (%)<sup>a</sup> | Extract | Water (control) |
|---------------------------|-----------------------------|---------|-----------------|
| 3.0                       | 0                           | 91      |
| 4.0 (original)            | 0                           | 89      |
| 5.0                       | 59                          | 91      |
| 6.0                       | 98                          | 94      |
| 7.0                       | 98                          | 93      |
| 8.0                       | 93                          | 93      |
| 9.0                       | 95                          | 87      |
| 10.0                      | 91                          | 93      |

<sup>a</sup>Spore suspension mixed with the same amount of extract was incubated at 28°C. Data were means of three replicates.

### Table 5
Conidial germination of *Alternaria brassicicola* in culture extract of *Eupenicillium brefeldianum* adjusted to pH value of 2.0 or 12.0 for 24 h before being adjusted back to the original pH of 4.0

| pH            | Germination (%)<sup>a</sup> |
|---------------|-------------------------------|
| 2.0           | 0                             |
| 12.0          | 0                             |
| 4.0 Extract   | 0                             |
| 4.0 Water     | 98                            |

<sup>a</sup>Spore suspension mixed with the same amount of extract was incubated at 28°C for 4 h. Data were means of three replicates.

### Table 6
Effect of high temperature treatment of culture extract of *Eupenicillium brefeldianum* on its inhibitory activity against germination of *Alternaria brassicicola* conidia

| Treatment<sup>a</sup> | Germination (%)<sup>b</sup> |
|------------------------|-----------------------------|
| 60                     | 0                           |
| 80                     | 0                           |
| 100                    | 0                           |
| Autoclave              | 0                           |
| Non-treated extract    | 0                           |
| Water (control)        | 97                          |

<sup>a</sup>Extract was treated at 60, 80 or 100°C for 30 min or autoclaved for 15 min. <sup>b</sup>Spore suspension mixed with the same amount of extract was incubated at 28°C for 4 h. Data were means of three replicates.

### Table 7
Effect of treatment of culture extract of *Eupenicillium brefeldianum* with cation exchange resins, anion exchange resins or activated charcoal on its inhibitory activity against germination of *Alternaria brassicicola* conidia

| Treatment<sup>a</sup> | Germination (%)<sup>b</sup> |
|------------------------|-----------------------------|
| Extract without treatment | 0                         |
| Cation exchange resins  | 0                           |
| Anion exchange resins   | 0                           |
| Activated charcoal      | 99                          |
| Water (control)         | 98                          |

<sup>a</sup>The pH value of the treated extract was adjusted to the original pH of 4 before spore germination test. <sup>b</sup>Spore suspension mixed with the same amount of extract was incubated at 28°C for 4 h. Data were means of three replicates.
(Garrett 1970). This may explain its widespread occurrence in soil (Keller and Bidoehka 1998; Souza-Motta et al. 2003). The fungus produced a stable fungidal substance when grown in broth prepared from the same vegetable tissues used in the soil amendment. This indicates the possibility of production of this fungidal substance by *E. brefeldianum* during its colonization of plant tissues in soil. It is conceivable that suppression of substrate competitors by this fungidal substance may account at least in part for the strong competitive saprophytic ability of this fungus in soil. Such possibility deserves further investigation.

The fungidal substance produced by *E. brefeldianum* was very stable under high temperature and high or low pH, indicating that the compound may last very long in nature after being applied to plant leaves. This may explain why the fungidal compound was very effective in controlling leaf spots of mustard cabbage caused by *A. brassicicola*. Although the compound was effective in controlling leaf spots of mustard cabbage in the greenhouse, its possibility of being developed into a commercial product is far from certain. Its disease control ability in the field and its toxicity to the non-target organisms in nature remain to be investigated.

The fungidal substance produced by *E. brefeldianum* was very effective at pH 3.0 to 4.0, partially effective at pH 5.0, but not at pH ranging from 6.0 to 10.0. Several microbiotics have been shown to be more active against microorganisms in acidic medium than in the more alkaline medium (Poala et al. 1970).

Results from this study showed that the inhibitory substance produced by *E. brefeldianum* was fungidal. The fungidal substance in the freeze dried powder of the culture was extractable with water, ethanol and methanol, but not acetate, ethyl acetate, ether and chloroform, indicating that it is soluble in polar solvents but not soluble in non-polar solvents. The ion exchange resin test suggested that the compound did not have charges on its molecules.

### Conclusion

Results from this study suggest that *E. brefeldianum* possesses a strong competitive saprophytic ability because of its ability to produce a fungidal substance during its colonization of plant tissues in soil. This may explain the widespread occurrence of this fungus in nature.

### Competing interests

The authors declare that they have no competing interests.

### Authors’ contributions

WHK conceived and designed the experiments, and wrote the manuscript. YTC performed the experiments. Both authors read and approved the final manuscript.

### Acknowledgments

We thank Dr. Sheng-Shong Tzean for identification of V3F-3, and Miss Hsu-Fang Cheng for manuscript typing. This study was supported by grants from Council of Agriculture and National Science Council of Taiwan (NSC 101-2321-B-005-004).

### Received

24 October 2013 Accepted: 22 January 2014

Published: 22 July 2014

### References

Agrios GN (2005) Plant Pathology, 5th edn. Elsevier Academic Press, San Diego, California

Alexander M (1977) Introduction to Soil Microbiology, 2nd edn. Wiley, New York

Ann PJ, Tsai JN, Wang TC, Chen CH, Lin MJ, Ko WH (2010) Reevaluation of the report of the A2 mating, type of *Phytophthora infestans* on Tomato in Taiwan. Bot Stud 51:203–207

Chen YT, Lin MJ, Yang CH, Ko WH (2011) Characterization of a fungistatic substance produced by Aspergillus flavus isolated from soil and its significance in nature. New Biotechnol 286:99–103

Farr DF, Bills GF, Chanurs GR, Rosenblatt (1989) Fungi on Plants and Plant Products in the United States. APS Press, St. Paul, Minnesota

Garrett SD (1970) Pathogenic Root-Infecting Fungi. Cambridge University Press, London

Hewitt HG (1998) Fungicides in Crop Protection. CAB International, Wallingford, Oxford, UK

Ho WC, Wu TY, Su HJ, Ko WH (2007) Effect of oriental medicinal plant extracts on spore germination of *Alternaria brassicicola* and nature of inhibitory substances from speedweed. Plant Dis 91:1621–1624

Humpherson-Jones FM, Maudo RB (1982) Control of dark leaf spot (Alternaria brassicicola) of Brassica oleracea seed production crops with foliar spray of iprodione. Ann Appl Biol 100:99–104

Kamata K, Okubo M, Ishigamoi E, Masaki Y, Uchida H, Watanabe K, Kashiwagi N (1983) Immunosuppressive effect of bredinin on cell-mediated and humoral immune reactions in experimental animals. Transplantation 35:144–149

Keller L, Bidoehka MJ (1998) Habitat and temporal differences among soil microfungal assemblages in Ontario. Can J Bot 76:1798–1803

Klausner RD, Donaldson JG, Lippincott-Schwartz J (1992) Brefeldin A: Insights into the control of membrane traffic and organelle structure. J Cell Biol 116:1071–1080

Ko WH, Lin MJ, Hu CY, Ann PJ (2010a) Aquaperonospora taiwanensis gen. et sp. nov. in Peronosporaceae of Peronosporales. Bot Stud 51:343–350

Ko WH, Tsou YJ, Lin MJ, Chem LL (2010b) Activity and characterization of secondary metabolites produced by a new microorganism for control of plant diseases. New Biotechnol 27:397–402

Ko WH, Yang CH, Lin MJ, Chen CY, Tsou YJ (2011) *Humicola philosophorides* sp. nov. from soil with potential for biological control of plant diseases. Bot Stud 52:197–202

Kolte SJ, Awasthi RP, Vishwanath M (1987) Assessment of yield losses due to *Alternaria* blight in rape and mustard. Ind Phytopathol 40:209–211

Nebenfuhr A, Ritzenhalter C, Robinson DG (2002) Brefeldin A: deciphering an enigmatic inhibitor of secretion. Plant Physiol 130:1071–104

Poala P, Wilcox C, Finland M (1970) Effect of pH of medium and size of inoculum on activity of antibiotics against group D Streptococcus (*Enterococcus*). Appl Microbiol 19:623–637

Souza-Motta CM, Cavalcanti MAQ, Fernandes MJ, Lima DMM, Nascimento JP, Laranjeira D (2003) Identification and characterization of filamentous fungi isolated from the sunflower (*Helianthus annuus* L.) rhizosphere according to their capacity to hydrolyse inulin. Braz J Microbiol 34:273–280

Tsai YN, Lin MJ, Ko WH (2012) A simple method for production of uniform inoculum of *Rhizoctonia solani* with strong pathogenicity. Biocat and Agric Biotechnol 1:85–88

Wang PH, Chen YS, Lin MJ, Tsou YJ, Ko WH (2010) Severe decline of wax apple trees caused by *Fusarium solani* in northern Taiwan. Bot Stud 51:343–350

World Health Organization (1990) Public Health Impact of Pesticides Used in Agriculture. World Health Organization, Geneva, Switzerland

---

doi:10.1186/1999-3110-55-39

Cite this article as: Chen and Ko: Characterization of a fungidal substance produced by *Eupenicillium brefeldianum* isolated from soil for plant disease control and its significance in nature. *Botanical Studies* 2014 55:39.