Editors’ Pick: What a pain…, or not!

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How nice, I thought as a child, would it be to not feel physical pain. Later I realized that sense of pain may exist for a reason; it was suggested that physical pain has evolved as an alarm system to protect the body from serious damage. Much later I read about human disorders involving severe insensitivity to physical pain. Still, the idea of not feeling pain remains appealing (especially anticipating my coming years when some painful age-related health problems are likely to kick in).

Previously, it was demonstrated that a loss-of-function mutation in the sodium ion channel gene SCN9A leads to the inability to experience pain [1]. In a more recent study, Leipold et al. [2] identified another sodium ion channel gene, SCN11A, which, when harboring a particular missense mutation, causes loss of pain perception. SCN11A encodes the voltage-gated sodium ion channel Na\textsubscript{\textalpha}{1,9}, which is highly expressed in nociceptors (i.e., neurons that transmit sensory information from the body periphery to the spinal cord). The authors started out with whole-exome sequencing of healthy German parents and their child diagnosed with the congenital inability to experience pain. A heterozygote non-synonymous mutation in SCN11A was identified as the only de novo event detected in this affected child. Attempting to confirm this finding in independent samples, the authors performed Sanger sequencing of SCN11A exons in 58 additional individuals diagnosed with early-onset severe sensory loss, most of them representing sporadic cases. One Swedish individual showed the very same heterozygous SCN11A mutation, and the clinical history of both individuals was also similar. From these findings, the authors hypothesized that the identified mutation that leads to a replacement of leucine with proline (p. Leu811Pro) at the distal end of one of the transmembrane segments in domain II of Na\textsubscript{\textalpha}{1,9} is disease causing and leads to loss of pain perception.

Seeking molecular proof, the authors introduced the orthologous alteration (Leu799Pro) into the mouse Scn11a gene, but found no obvious morphological changes in sensory axons or in nerve fibers in the skin of the viable heterozygote knock-in mice. However, 11% of the knock-in mice had severe lesions that were not observed in the wild-type mice. Since the lesions also appeared in knock-in mice housed solitarily, it was assumed that they were self-inflicted. Also other experimental evidence pointed towards reduced pain sensitivity in these knock-in mice.

Because Na\textsubscript{\textalpha}{1,9} had been suggested to influence the excitability of dorsal root ganglia (DRG) neurons (i.e., their property to react to stimulation), the authors then investigated the electrical potential of DRG neurons isolated from wild-type and mutant mice. They found that a significant fraction of the mutant channels were active under resting conditions, and that the duration of action potentials was significantly reduced in the mutant neurons compared with wild-type neurons. In knock-out mice, however, only a minor effect on action potentials was observed. Additional experiments showed that the excessive activity at resting conditions caused sustained depolarization of nociceptors, impaired generation of action potentials and aberrant synaptic transmission.

To study the effect of the mutation further, the authors heterologously expressed human Na\textsubscript{\textalpha}{1,9} and Na\textsubscript{\textalpha}{1,9} Leu811Pro mutant channels in ND7/23 cells (i.e., a mouse neuroblastoma x rat neuron hybrid cell line). They found that the phenotype of human Na\textsubscript{\textalpha}{1,9} Leu811Pro mutant channels resembled that of mouse Na\textsubscript{\textalpha}{1,9} Leu799Pro mutant channels, except in terms of steady-state inactivation. From the weak influence of the mutation on inactivation in the human channel, they concluded a stronger gain-of-function effect in humans than in knock-in mice.

Besides impressively demonstrating the power of whole-exome sequencing to find rare candidate disease causing mutations, this study provides a new gain-of-function view on the molecular basis of pain loss, which was previously linked to loss-of-function mutations such as in the NA\textsubscript{\textalpha}{1,7} channel encoded by SCN9A [1]. Furthermore, this finding may open up new avenues for the development of new painkilling drugs [3].
Competing interests
The author declares that they have no competing interests.

Received: 9 June 2014 Accepted: 9 June 2014
Published: 1 July 2014

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doi:10.1186/2041-2223-5-8
Cite this article as: Kayser: Editors’ Pick: What a pain…, or not.
Investigative Genetics 2014 5:8.

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