The Multifaceted Roles of MicroRNAs in Cystic Fibrosis

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Abstract: Cystic fibrosis (CF) is a lifelong disorder affecting 1 in 3500 live births worldwide. It is a monogenetic autosomal recessive disease caused by loss-of-function mutations in the gene encoding the chloride channel cystic fibrosis transmembrane conductance regulator (CFTR), the impairment of which leads to ionic disequilibria in exocrine organs. This translates into a chronic multisystemic disease characterized by airway obstruction, respiratory infections, and pancreatic insufficiency as well as hepatobiliary and gastrointestinal dysfunction. Molecular characterization of the mutational heterogeneity of CFTR (affected by more than 2000 variants) improved the understanding and management of CF. However, these CFTR variants are linked to different clinical manifestations and phenotypes, and they affect response to treatments. Expanding evidence suggests that multisystemic disease affects CF pathology via impairing either CFTR or proteins regulated by CFTR. Thus, altering the expression of miRNAs in vivo could constitute an appealing strategy for developing new CF therapies. In this review, we will first describe the pathophysiology and clinical management of CF. Then, we will summarize the current knowledge on altered miRNAs in CF patients, with a focus on the miRNAs involved in the deregulation of CFTR and in the modulation of inflammation. We will highlight recent findings on the potential utility of measuring circulating miRNAs in CF as diagnostic, prognostic, and predictive biomarkers. Finally, we will provide an overview on potential miRNA-based therapeutic approaches.

Keywords: cystic fibrosis; genetic disease; microRNA; circulating miRNAs; microRNA-targeted therapies; miRNA mimics; antimiRs; antagomiRs; biomarkers

1. Introduction

The dual role of time, as a fearsome enemy or important ally in the course of human diseases, has motivated researchers to advance in the frantic (and necessary) understanding of their cellular and molecular bases, hoping to identify novel approaches for patient care. The discovery of a variety of disease-modulatory molecules, including non-coding RNAs, that are present not only at the tissue level, but also in the bloodstream, has revolutionized the world of clinical molecular biology.
microRNAs (miRNAs), a class of small non-coding RNAs, are well known to regulate a diverse array of biological processes, such as proliferation, development, and metabolism [1,2]. They act as repressors of one or several genes by binding to complementary sites within the 3’ untranslated region (UTR) of target mRNAs [1,3,4].

miRNAs positively or negatively impact on the pathogenesis and/or progression of human diseases. They are (de)regulated by different mechanisms, such as epigenetic alterations, chromosomal abnormalities, as well as changes in transcriptional control [2,5]. Moreover, the altered abundance of miRNAs, along with their high stability (due to their small length of about 18–22 nucleotides) renders them highly potential biomarker candidates in non-invasive sources (i.e., blood, serum, urine, and saliva) [6,7].

The clinical potential of miRNAs as diagnostic and prognostic, as well as predictive biomarkers, has been widely described, especially in the context of human cancer [6,8–10]. Moreover, potential therapeutics harness the deregulation of miRNAs in disease conditions by means of miRNA mimics, so-called antimiRs [11,12].

Accumulating evidence suggests the involvement of miRNAs also in genetic diseases, including cystic fibrosis (CF) [13–19]. In particular, many studies showed the direct or indirect regulatory impact of deregulated miRNAs on the expression of the cystic fibrosis transmembrane conductance regulator (CFTR) mRNA [20]. Moreover, their participation in the control of the inflammation of CF airways has been demonstrated, along with their potential role as circulating biomarkers [17]. Thus, the contribution of miRNAs to disease progression and severity opens new and interesting scenarios for the clinical management of CF patients.

In this review article, we will describe the pathogenesis of CF, followed by an overview of the wide mutational spectrum harbored by the CFTR gene that translates into the large range of CF clinical phenotypes. CF care is a race against time and against the progressive and aggressive degeneration of multiple organs. Considering the benefits that non-coding RNAs could potentially offer for the treatment of human diseases, we cover current knowledge on the impact of miRNAs on CF. Thus, we will discuss the biological regulatory role of miRNAs in the progression of the disease. Moreover, we will describe circulating miRNAs as potential non-invasive biomarkers (i.e., for monitoring lung disease progression), as well as the role of single nucleotide polymorphisms (SNPs) associated with miRNAs in modulating CF phenotypes. Finally, we will summarize the therapeutic potential of miRNAs in CF.

2. Cystic Fibrosis

2.1. Cystic Fibrosis, A Multisystemic Deadly Disease

CF, also known as mucoviscidosis, is one of the most common Mendelian autosomal recessive genetic disorders [21,22]. Its incidence varies according to ethnicity, affecting approximately 1 in about 2500 newborns in Europe [23].

CF is caused by mutations in CFTR, which belongs to the ATP binding cassette (ABC) gene family [22,24]. Environmental factors, as well as non-CFTR gene modifiers, influence the manifestations and progression of the disease [25].

The CFTR gene is located on the long arm of the chromosome 7 [25]. The CFTR protein is mostly located at the apical membranes of a variety of secretory epithelia (which produce mucus, sweat, and digestive juices) harbored in multiple organs, including sweat glands, lungs, and pancreas, as well as the gut [21,25]. CFTR mainly functions as an anion channel; it acts as a cyclic adenosine monophosphate (cAMP)-dependent chloride channel and as bicarbonate channel. Moreover, the involvement of CFTR in the modulation of other ion channels, such as the epithelial sodium channel (ENaC), has been also described, not without controversy [26]. CFTR transports chloride and bicarbonate across secretory epithelia, thus regulating both the secretion and the absorption of salt and water and maintaining epithelial surface hydration.
Mutations in CFTR lead to the impairment of the expression, as well as of the function and stability at the mRNA and CFTR protein levels [27], thus unbalancing fluid and electrolyte homeostasis. For instance, an abnormal excessive secretion of salt from the sweat glands that has not been reabsorbed by the sweat duct cells is one of the main symptoms of CF [25]. At present, the measurement of chloride excreted in sweat is the gold standard to diagnose CF.

Another deleterious effect, which is due to the reduction of the bicarbonate release, is the formation of thickened and viscous secretions in bronchi, pancreas, biliary tract, intestine, and the reproductive system. Although mucus is an alley of innate immunity due to its composition made, inter alia, of antibacterial defensins and immunoglobulins, its hyperviscous form is pathogenic in CF [28].

Concerning the respiratory tract of CF patients, defects in fluid secretion and the reduced pH of the airway surface liquid affect ciliary beating and compromise mucociliary clearance, thus increasing the viscosity of the mucus but decreasing the activity of antimicrobial molecules [29]. As a consequence, mucus accumulates and obstructs the airways (bronchiectasis), promoting chronic bacterial infections and inflammatory lung damage. The airway microbiome changes across the ages [30]. Typically, CF adults are persistently colonized by Pseudomonas aeruginosa (P. aeruginosa) [31]. Altogether, these changes trigger acute pulmonary exacerbations and respiratory failure.

The gastrointestinal tract as well as the biliary ducts are also impacted by unbalanced ion secretion [32,33]. The more acidic intestinal environment, along with the sticky and thickened mucus, which is hardly cleared, alters the activity of digestive enzymes; this impairs the assimilation of digestive products, delays the intestinal transit, and causes bowel obstructions [32]. Meconium ileus is one of the earliest clinical manifestations and the most serious acute complication of CF at the intestinal level [34]. Nonetheless, older CF patients may also present distal intestinal obstruction syndrome (DIOS) and intussusception [33]. Furthermore, intestinal inflammation and dysbiosis occur in the context of CF [32,35].

Severe inflammation, viscous secretions, and fibrosis lead to pancreatic insufficiency, leading to abdominal pain, malabsorption, and weight loss [36]. Additionally, diabetes mellitus and male infertility may become manifest [37–39]. Congenital aplasia of vas deferens (CAVD) contributes to male infertility and obstructive azoospermia [40,41]. This abnormality of the urogenital tract has different clinical presentations, according, inter alia, to its bilaterally (CAVD) or unilaterally (CUAVD) occurrence [40–42]. In women with CF, fertility is compromised due to thicker cervical secretions [37].

Although CF is a chronic multisystemic disease that is ultimately lethal, daily care and personalized (expensive) treatments extend the life expectancy of most CF patients until 40 years of age [43]. The medical care for CF requires multiple interventions including high-calorie ingestion, therapy based on the replacement of pancreatic enzymes and, most importantly, the management of the pulmonary exacerbations [21,25,44]. For instance, airway-clearance techniques and aerosolized mucolytic agents, along with administration of some specific antibiotics and anti-inflammatory drugs can avoid, reduce, or eradicate bacterial infections and improve lung functions [25].

In addition, targeted therapies for CF patients with specific CFTR pathogenic mutations have been developed [43–46]. These treatments are based on a class of compounds, the CFTR modulators, which are able to modulate (enhance or restore) the expression, function, and stability of mutant CFTR proteins [43,45]. Generally, CFTR modulators are grouped into five classes: (i) “potentiators”, which restore the channel gating and conductance of the mutant CFTR protein; (ii) “correctors”, which are drugs that rescue the protein folding, thus augmenting the traffic to the plasma surface of CFTR mutant; (iii) “stabilizers” that avoid the removal and degradation by lysosomes of CFTR through its stronger stabilization on the plasma membrane; (iv) “amplifiers” that increase the expression of CFTR at the mRNA level, and, in turn, the amount of CFTR protein; and (v) “read-through” agents that restore the functional full-length protein affected by premature termination codons. To date, only two classes of modulators, potentiators and correctors, have been approved to treat CF patients [45].

Finally, lung transplantation is reserved for end-stage CF patients, but it extends survival by only 5 years on average [47].
2.2. The Mutational Landscape of the Cystic Fibrosis

Since the discovery of the first CFTR mutation in 1989, to date, more than 2000 different genetic variants have been identified (www.CFTR2.org). However, only a few of them (about 350 mutations) are considered as pathogenic (www.CFTR2.org) [43].

According to the traditional classification system, mutations harbored by CFTR are grouped into six classes, depending on their effect on the synthesis, function, or stability of the CFTR protein (Table 1) [27,44].

Table 1. Classes of mutations in the cystic fibrosis transmembrane conductance regulator (CFTR) gene according to the traditional classification system.

| Class of Mutation | CFTR Molecular Defect | Functional Abnormal Consequence | Mutation Examples | Type of Mutations | Clinical Phenotype | Therapeutic Strategy |
|-------------------|-----------------------|---------------------------------|-------------------|-------------------|-------------------|---------------------|
| I                 | No mRNA and protein synthesis | Absent protein | G542X, R553X, W1282X | Nonsense, frameshift, canonical splicing | Severe | Read-through agents |
| II                | Reduced protein processing and traffic | Misfolded protein | F508del, N1303K, I507del | Missense, aminoacid deletion, | Severe | Correctors |
| III               | Impaired channel gating | Reduced or absent channel opening | S549N, G551D | Missense, aminoacid change | Severe | Potentiators |
| IV                | Decreased channel conductance | Defect in ion transport | R347P, R117H, D1152H | Missense, aminoacid change | Mild | Potentiators |
| V                 | Reduced protein synthesis | Decreased protein | 3849 + 10 kb C>T, A455E | Splicing defect, missense | Mild | Potentiators, correctors, ASOs |
| VI                | Less protein stability and protein turnover at cell surface | Decreased half-life of the protein | 120del23, G1412X | Missense, aminoacid change | Mild | Stabilisers |

ASOs, antisense oligonucleotides; CFTR, cystic fibrosis transmembrane conductance regulator.

Specific mutations in CFTR relatively influence the severity of the disease and, therefore, translate into different clinical manifestations or CF phenotypes (Table 1) [27]. However, the severity of the disease is also related to the heterozygosity or homozygosity of the CFTR mutation [48]. The molecular diagnosis of CFTR mutations guides clinicians with regard to the management of the disease.

Severe mutations belonging to classes I, II, and III (Table 1) [27] lead to the loss of the CFTR function. In particular, class I mutations abolish the synthesis of the protein. They include nonsense mutations (i.e., premature termination codons). Class II mutations cause the retention of a misfolded protein at the endoplasmic reticulum and its subsequent degradation by the proteasome. The most common mutation in Europe, the F508del, which consists in the deletion of a phenylalanine at position 508, belongs to this class. Class III mutations affect the regulation and gating of the CFTR channel.

By contrast, class IV to VI mutations confer a milder CF phenotype (Table 1) [27]. Class IV mutations decrease the conductance of chloride and bicarbonate ions. Class V mutations lead to a reduction of the abundance (synthesis or maturation) of the normal CFTR protein. Class VI mutations destabilize the protein at the cell surface by increasing the endocytosis and lysosomal degradation of CFTR.

A new categorization of CFTR mutations into VII classes according to the therapeutic strategies has been suggested by De Boeck and Amaral [46]. The only difference compared to the traditional classification system consists in the addition of class VII mutations, which have the same functional characteristics as class I mutations but cannot be rescued by pharmacological interventions [46].
3. microRNAs in Cystic Fibrosis

**Altered microRNAs in the Regulation of CFTR**

One of the first studies on miRNAs in CF employed transcriptomic and miRNA array-based approaches to show that altered levels of miRNAs correlated with the decreased expression of CFTR under hypoxic conditions [49]. Likewise, in silico predictions, followed by in vitro validations, recently described miR-200b as a potential negative regulator of CFTR mRNA levels in human airway epithelia during hypoxia [50].

Over the years, different groups showed that a variety of miRNAs repress directly (i.e., miR-101, miR-145, miR-223, miR-494, miR-509-3p and miR-1246) or indirectly (i.e., miR-9 and miR-138) the expression of CFTR [51–58]. Concerning this latter point, miRNAs can bind to the 3' UTR of genes that modulate CFTR. For instance, miR-138 reportedly downregulates CFTR via targeting the transcriptional repressor gene *switch-independent 3 homolog A* (*SIN3A*) [58]. Another example concerns miR-9, the upregulation of which impaired the expression and activity of the gene *anoctamin 1* (*ANO1*) in bronchial epithelial cells, thus contributing to CF lung pathology [57]. *ANO1*, also known as calcium-activated chloride channel (TMEM16A), may be involved in the activation and membrane expression of CFTR [59].

By contrast, Gillen and collaborators demonstrated that miR-145 and miR-494 directly target the 3’ UTR of *CFTR* mRNA [54]. Interestingly, miR-384, miR-494, and miR-1246 are also implicated in the regulation of chloride channel transport by regulating the gene *solute carrier family (SLC) 12 member 2* (*SLC12A2*) [51]. Other investigations confirmed the deregulation of miR-101, miR-145, and miR-494 in different specimens from CF patients [51–53]. miR-101 was also found to be upregulated in the lung of patients with chronic obstructive pulmonary disease; this condition is characterized by low CFTR protein levels [52].

Concerning the involvement of the well-known miR-145 in the dysregulation of CFTR, a variety of mechanisms have been identified. For instance, elevated levels of miR-145 were found in CF versus non-CF nasal epithelial tissues and inversely correlated with the expression of *SMAD family member 3* (*SMAD3*). Furthermore, miR-145, along with miR-494 and miR-223, was increased in bronchial brushing from CF patients bearing the F508del mutation. Functional experimentation showed that the corresponding premiRs downregulated and antimiRs upregulated CFTR gene and protein expression. Moreover, defective chloride ion conductance, inflammatory and infective insults altered the levels of miR-145, miR-223, and miR-494 [60]. Recently, miR-145-5p and miR-223-3p were found to be overexpressed in F508del homozygous CF bronchial epithelial cells and specimens from children and adults with CF. High levels of these miRNAs were also found in CF and CFTR gene-corrected induced pluripotent stem cell-derived CF lung organoids. Additional studies provided evidence that the upregulation of miR-145 in CF might be partially secondary to the increased expression of transforming growth factor-β (TGF-β) [61,62].

CF and chronic bronchitis, as well as tobacco smokers, present a similar lung inflammation [63]. In this context, it has been demonstrated that air pollution and cigarette smoke (CS) play an important role in the progression of airway diseases by promoting CFTR dysfunction through miRNA deregulation. For instance, CS increased the expression of miR-101 that consequently reduced CFTR in human bronchial epithelial cells and in vivo [52]. Moreover, altered levels of miR-145-5p were involved in the TGF-β-mediated suppression of CFTR and the chloride channel, SLC26A9 [64]. Additionally, an antagoniR specific for miR-145-5p rescued CFTR and SLC26A9 expression in vitro, while the use of a neutralizing aptamer against another target of miR-145-5p, namely TGFBR2, was able to restore the suppression of CS- and TGF-β-mediated CFTR function in vivo [64].

Defective autophagy plays a critical role in the pathology of CF [65,66]. Therefore, the role of deregulated miRNAs in impairing the autophagy machinery in CF has been explored. Indeed, high levels of the cluster mirc1/mir17-92 contributed to the negative regulation of autophagy and CFTR function in CF macrophages [67].
4. microRNAs as Regulators of Inflammation in Cystic Fibrosis

Inflammation plays a critical role in the progression of CF [68]. As described before, the overproduction of hyperviscous mucus obstructs airways and impairs mucociliary clearance. This creates a perfect nutrient-rich environment for bacterial colonization as well as for pro-inflammatory modulators that lead to the progressive structural damage of the lung (Figure 1).

![Figure 1. Airway inflammation in cystic fibrosis.](image)

miRNAs can contribute to the maintenance of a pro- (mainly) or anti-inflammatory phenotype in CF patients via different ways [20]. Thus, the modulation of the abundance of miRNAs involved in these pathways may prevent lung disease and offer clinical applications in the future.
Essential fatty acid-derived lipoxins (LX), resolvins (Rv), protectins, and maresins are lipid mediators that control several aspects of the acute inflammation and resolution via specific G protein-coupled receptors (GPCRs) [69]. LXA₄ and RvD₁ activate a specific GPCR termed lipoxin A₄ receptor (ALX)/formyl peptide receptor 2 (FPR2), which signals to inhibit NF-κB activation [69,70].

The role of the ALX/FPR2 receptor has been explored in relationship with CF. Interestingly, the ALX/FPR2-dependent pathway of inflammation resolution was altered by the overexpression of miR-181b in CF respiratory cells and macrophages [71].

It has been demonstrated that a variety of altered miRNAs are able to regulate the expression of several pro-inflammatory messengers, including interleukin (IL)-6 and IL-8. For instance, miR-146a upregulation is involved in CF inflammation. Thus, the elevation of miR-146a causes macrophages isolated from CF patients to overproduce IL-6 in response to lipopolysaccharide (LPS) stimulation [72]. IL-8 is produced by macrophages and bronchial epithelial cells in the CF lung in response to infectious (i.e., Pseudomonas aeruginosa and Staphylococcus aureus) and inflammatory (i.e., IL-1β or tumor necrosis factor-α, TNF-α) stimuli via different signaling pathways (i.e., NF-κB) [73,74]. The main function of IL-8 is to attract and activate neutrophils, which dominate the inflammatory response in the CF airway [74]. The secretion of IL-8 could be directly or indirectly impaired by deregulated miRNAs. For instance, IL-8 was identified as a direct target of miR-17 and miR-93 [75,76]. Particularly, low levels of miR-17 were found in CF cell lines, also when treated with P. aeruginosa-conditioned medium, in CF bronchial brushings, and in βENaC-overexpressing (βENaC-Tg) mice with spontaneous airway neutrophilia, as well as in mucus obstructions [75]. miR-93 was identified as significantly downregulated in bronchial epithelial cells infected with P. aeruginosa through a microarray technique [76]. For both microRNAs, luciferase assays, along with other in vitro experiments, demonstrated that their deregulation directly increased the production of IL-8.

Concerning indirect modulations, high levels of miR-155 in CF cells inhibited the translation of the gene SH-2 containing inositol 5’ polyphosphatase 1 (SHIP1), thus leading to the activation of the signaling pathway mediated by phosphatidylinositol-3 kinase/protein kinase B (PI3K/AKT) and the consequent secretion of IL-8 [77]. The same authors showed that the expression of miR-155 in CF was suppressed by tristetraprolin (TTP) through the induction of miR-1 but enhanced by KH-type splicing regulatory protein (KSRP) via promoting its maturation in vitro [78]. Moreover, miR-155 may also regulate the fibrosis of CF lungs. In fact, it can suppress the expression of regulatory associated protein of mTOR complex 1 (RPTOR), thus consequently increasing the abundance of connective tissue growth factor (CTGF) in CF lung epithelial cells [79].

Recently, miRNome and transcriptomic analyses showed that miR199a-3p was less expressed in CF bronchial explants extracted from CF patients than in non-CF individuals [80]. Moreover, wet-lab experimentations revealed that the downregulation of miR-199a-3p was associated with the hyperactivation of the NF-kB pathway and, therefore, with the increased secretion of IL-8 via targeting the inhibitor of nuclear factor kappa-B kinase subunit beta (IKKβ) [80].

miRNAs can also control other signaling pathways to trigger inflammation in CF. High levels of miR-199a-5p were detected in human and murine CF macrophages and murine CF lungs [81]. The aberrant expression of miR-199a-5p, mediated by PI3K/AKT signaling, reduced the expression of caveolin 1 (CAV1), which is a mediator of inflammation processes, and in turn increased Toll-like receptor (TLR) 4. Moreover, the downregulation of miR-199a-5p was able to reduce the inflammation in CF macrophages via restoring the expression of CAV1. Furthermore, in vitro and in vivo studies demonstrated that the administration of the non-steroidal anti-inflammatory drug celecoxib rescued the altered pathway mediated by AKT/miR-199a-5p/CAV1 in CF macrophages and reduced lung inflammation in CFTR-deficient mice, respectively [81].

An elevated expression of miR-145, found in nasal airway cells from CF patients when compared to non-CF controls, correlated with the downregulation of SMAD3 [82]. SMAD3 is a negative modulator of the NF-κB–IL-8 pathway mediated by TGF-β1; therefore, miR-145 may contribute to CF-related inflammation [82].
Differently from the aforementioned miRNAs, miR-126 has an anti-inflammatory role in CF. It has been the first miRNA identified as deregulated in CF in 2010 [83]. In particular, low levels of miR-126 were found in CF airway epithelial cells. This downregulation correlated with the upregulation of the target of Myb protein 1 (TOM1), which is a negative regulator of TLR2, TLR4, IL-1, IL-1β, TNF-α, and NF-kB. When CF cells were stimulated with LPS and IL-1β, the knockdown of TOM1 significantly increased the NF-kB-mediated IL-8 secretion [83]. In addition, another study reported that miR-1343 attenuated the pathway of fibrosis by decreasing levels of activated TGF-β effector molecules, including phosphorylated (p) SMAD3 (pSMAD3) as well as pSMAD2 and consequently disturbing the cell migration and epithelial–mesenchymal transition [84].

Some miRNAs may also participate in chronic inflammation by affecting the remodeling of the pulmonary epithelium or by impacting the expression of genes involved in mucus hypersecretion. The role of miR-146a in CF inflammation has been linked to its negative impact on the production of the mucin 5AC (MUC5AC), which is a major component of the airway mucus [85]. Moreover, the knockdown of miR-146a in human bronchial epithelial cells resulted in the activation of the NF-kB and Jun N-terminal kinase (JNK) pathways [85].

In bronchial epithelial cells, miR-145, miR-494, and particularly miR-221 regulate the transcription factor 6 (ATF6), which is implicated in the airway inflammation through endoplasmic reticulum stress [86,87].

The deregulation of miR-31 enhanced the production of the cathepsin S (CTSS) via the direct inhibition of the interferon regulatory factor 1 (IRF-1) in CF epithelial cells [88]. CTSS is an elastinolytic and collagenolytic cysteine protease. High levels of CTSS in CF patients increased pulmonary neutrophilic infiltration of the lung, as well as the inactivation of antimicrobial proteins, such as lactoferrin and members of the β-defensin family, therefore contributing to lung inflammation and infection, and determining lung damage [89].

5. Circulating microRNAs as Potential Biomarkers in Cystic Fibrosis

miRNAs can be quantified in non-invasive specimens such as body fluids, which renders them easily detectable biomarkers [13,90–92]. Indeed, miRNAs are highly stable and circulate in the bloodstream as cell-free miRNAs or packaged inside microvesicles, such as exosomes [9,93]. Advances in liquid biopsy technologies (i.e., digital PCR and microfluidic single-cells technologies) enable their accurate detection [94].

Although newborn bloodspot screening (NBS) offers the opportunity of early diagnosis, CF is a pediatric disorder that most often begins as a silent disease. Thus, along with the clinical examination, the detection and the assessment of the levels of expression of a single specific miRNA or signature of miRNAs in easily obtained biological sources, could be beneficial for the care of patients affected by CF. The potential role of miRNAs as circulating biomarkers has been explored in a few studies.

The Mirc1/Mir17–92 cluster has been identified as a potential biomarker for CF disease progression [95]. In particular, high levels of the Mirc1/Mir17–92 cluster in CF sputum correlated with pulmonary exacerbations, lung function, and age.

Recently, a profiling study comparing plasma miRNAs in CF patients versus non-CF controls identified a total of 11 deregulated miRNAs. In particular, 10 miRNAs (miR-16-5p, miR-92a-3p, miR-103a-3p, miR-103b, miR-107, miR-191-5p, miR-3613-5p, let-7a-5p let-7b-5p, and hsa-let-7d-5p) were overexpressed, while only miR-598-3p was reduced in CF [96]. Furthermore, bioinformatics analyses revealed that the target genes of these miRNAs were enriched in transduction pathways, such as the mTOR and PI3K/AKT, as well as Wnt/β-catenin signaling pathways [96].

CF presents, inter alia, a gender dichotomy [97,98]. Women are affected by poorer lung function as well as lower median survival. To study this gender gap, circulating miRNAs were profiled in male and female pediatric CF plasma samples [99]. miR-885 was increased in samples from girls under six years. Moreover, in silico analyses suggested that the severity of the disease in CF females could be
related to a Ras-related C3 botulinum toxin substrate 1 (RAC1)-mediated process. However, functional experiments need to validate these theoretical predictions [99].

The involvement of circulating miRNAs in CF-related diabetes (CFRD), as well as in CF-associated liver disease (CFLD), has also been explored [100–103].

CFRD is the major extra-pulmonary co-morbidity in CF patients [38]. It significantly accelerates lung damage, leading to early mortality. The deregulation of miR-146a has been documented in CFRD [103]. In particular, high levels of the circulating miR-146 in serum samples from CF patients correlated with the onset of the CFRD. In addition, another study reported a significant increase of miR-146 expression in peripheral blood mononuclear cells cultured with CF plasma [104].

CFLD is a complication of CF that affects up to 40% of CF patients [105–107]. CFLD has been attributed to ductal cholestasis [106]. The pathogenetic mechanism of CFLD has not been fully elucidated yet. Moreover, the diagnostic criteria, captured by invasive liver biopsy, as well as the methods to monitor the progression of the disease are still inadequate [108]. In this context, miRNAs could be useful biomarkers in the management of CFLD.

Changes in the levels of circulating miR-122, miR-21, and miR-25 were able to identify CF patients affected by liver disease [100]. In detail, miR-122 was more expressed in CFLD patients when compared to CF patients without liver symptoms, as well as to healthy individuals. Moreover, miR-21 and miR-25 were overexpressed in CF patients with liver fibrosis compared to CF patients without fibrosis or controls. Additionally, circulating miRNA levels were also able to stratify CFLD patients according to the hepatic fibrosis stage (F0 = no fibrosis; F1–F4 = any histological evidence of fibrosis; F3–F4 = severe fibrosis). In particular, the combination of six miRNAs (miR-122, miR-21, miR-25, miR-210, miR-148a, and miR-19a) identified CF patients with early liver fibrinogenesis (F0–F1) and differentiated CFLD children without (F0) from those with any liver fibrosis (F1–F4) [100].

In order to diagnose and assess CFLD severity, Calvopina and collaborators investigated the combined use of serum miRNAs and aspartate aminotransferase (AST) to platelet ratio (APRI) [102]. An miRNA sequencing approach identified the circulatory miRNA signature of a total of 124 children (CF and CFLD patients, and healthy controls). Among the miRNAs detected, CFLD patients presented high levels of miR-122-5p, miR-365-33p, and miR-34a-5p, while low levels of miR-142-3p and let-7g-5p were detected with respect to CF children without liver disease. Moreover, the combination of miR-365a-3p, miR-142-3p, and let-7g-5p with APRI was able to predict liver disease in CF patients with high specificity (83%) and sensitivity (92%). Furthermore, the abundance of miR-18a-5p in serum specimens discriminated CF patients presenting severe (F3–F4) from mild/moderate (F0–F2) fibrosis [102].

In summary, emerging evidence suggests that circulating miRNAs may contribute to the diagnosis of CF complications. However, it will be important to overcome technical limitations (i.e., in sample collection, processing, and data analysis) and to obtain cognitive insights into the disease-modulatory role of such miRNAs.

6. Single Nucleotide Polymorphisms (SNPs) in microRNAs Targeting the CFTR Gene

As described above, CF is characterized by a large clinical heterogeneity. The variability of CF phenotypes is further accentuated by SNPs in the 3’ UTR of miRNA-targeted genes, as demonstrated by Amato and colleagues [36]. In particular, the SNP rs10234329, which is characterized by an adenine to cytosine (A>C) base substitution, was identified within the 3’UTR of the CFTR gene in a patient with a CFTR-related disease. Interestingly, in silico analyses demonstrated that this SNP was located in the target site of two miRNAs, including miR-433 and miR-509-3p. Moreover, in vitro experiments showed that this SNP reduced the expression of CFTR [56]. Thus, the 3’ UTR region of CFTR should be examined in CF patients that present clinical symptoms but lack mutations in the exons coding for the CFTR protein.

Another investigation identified polymorphisms in the region of a miRNA cluster (miR-99b/let-7e/ miR-125a) by examining CF F508del patients [109]. In particular, two SNPs detected (rs376594280 and
rs41275794) modulated the maturation and, therefore, altered the expression of the miR-99b/let-7e/miR-125a cluster. These results shed light on the variability of CF severity in patients bearing the same CFTR genotype.

7. Therapeutic Modulation of microRNAs in Cystic Fibrosis

7.1. microRNA Therapeutics

The modulation of the abundance of miRNAs may constitute a therapeutic strategy [11,12,110]. This can be achieved by using miRNA mimics or antimiRs [11,12,111].

miRNA mimics are synthetic double-stranded small RNAs used to re-establish the concentration of a specific downregulated miRNA [11,12]. Inversely, antimiRs suppress the function of overexpressed miRNAs. Generally, the function of miRNAs can be inhibited by three different approaches: (i) small-molecule inhibitors, which comprise specific compounds that repress the expression of a specific target miRNA (i.e., azobenzene for miR-21); (ii) miRNA sponges (expression vectors), which are transcripts that contain multiple, tandem binding sites to a microRNA of interest [112,113], thus acting as decoys of the target miRNA; and (iii) antisense oligonucleotides (ASOs) that competitively block the binding of a miRNA to its mRNA target [11,111,114].

Oligonucleotides are vulnerable to degradation by RNases present in serum or inside cells, resulting in poor pharmacokinetics [111]. Therefore, it is necessary to enhance their stability and binding affinity through two specific complementary strategies [11,111]: (i) chemical strategies relying on sugar and RNA backbone modifications, such as the addition of 2′-O-methyl (2-O′-Me) or phosphorothioate-like groups, locked nucleic acids (LNA), morpholinos, or peptide nucleic acids (PNA); and (ii) delivery strategies (i.e., viral vectors, polymer- and lipid-based delivery systems) aiming to encapsulate miRNAs, thus favoring their nuclease resistance and endosomal escape. Overall, methylation is often used for the generation of miRNA mimics, while LNA is mostly chosen for antimiRs. However, for in vivo applications, the phosphorothioate substitution is preferable [12]. In fact, this latter is not only particularly resistant to nucleases but also enhances the binding affinity with plasma proteins, so being quickly absorbed into the bloodstream [12]. To date, oligonucleotides are only administered via intravenous infusion or subcutaneous injection [12].

In spite of the interest of pharmaceutical companies for miRNA therapeutics, a variety of challenges need to be overcome, including tissue-specific targeting, toxicities, and off-targets effects [12,115]. For instance, chemical modifications could lead to adverse toxic effects in vivo, such as immune cell activation, altered coagulation, as well as hepatotoxicity [12,111]. Moreover, accurate tissue-specific delivery strategies need to be developed to minimize the risk of side effects in normal tissues as well as to attenuate the immune response.

The optimization of the design of miRNA-based drugs is another obstacle. Design factors include the size and lipophilicity of the molecules to ensure efficient adsorption by the target tissue but to avoid fast renal clearance [12].

7.2. microRNA Therapeutics in Cystic Fibrosis

Endogenous miRNAs may affect the expression of CFTR, thus exacerbating the symptoms of CF [15,17]. Thus, miRNA mimics and antimiR agents could represent an appealing therapeutic strategy [116,117]. However, because each miRNA may simultaneously target several mRNAs, this approach could lead to unpredictable off-target effects. Moreover, the tissue-specific delivery and the stability of miRNA mimics and antimiRs are critical hurdles for this approach. An example concerns the miR-138 mimic. As previously mentioned, miR-138 indirectly impairs CFTR through the downregulation of SIN3A [58]. This latter is a transcriptional repressor of CFTR. The manipulation of the expression of miR-138 was able to restore CFTR abundance, as well as the chloride channel permeability in CF bronchial epithelial cells via upregulating SIN3A [58]. Nevertheless, this strategy led to undesired side effects on other genes regulated by SIN3A.
Polymeric nanoparticles may be used to effectively deliver miRNA mimics. The pro-inflammatory miR-126, the downregulation of which leads to the overexpression of TOM1 in CF airways, has been used as a proof-of-concept [118]. Cationic nanoparticles successfully delivered miR-126-mimics into CF cell lines, thus significantly decreasing the expression of TOM1.

PNAs are also considered a valid anti-miR strategy [111]. PNAs are DNA analogues in which the sugar–phosphate backbone of the nucleic acid has been replaced by a synthetic achiral peptide backbone [111]. PNAs may be used to target specific miRNAs that are involved in the regulation of the expression of the CFTR gene [119,120]. For instance, PNAs designed to target miR-145, miR-509-3p, and miR-101-3p restored the expression of CFTR in vitro [61,121–123]. Additionally, PNAs can also be used as miRNA target protectors. In particular, PNAs can increase the expression of CFTR by competitively inhibiting the interaction of miRNAs with their mRNA targets [124]. However, these PNA-mediated approaches may present off-target effects on other miRNAs as well as on other target genes [122,125].

Another strategy involves target site blockers (TSBs) [111]. TSBs are locked nucleic acid antisense oligonucleotides that may have comparatively few off-target effects [111]. Indeed, TSBs compete with miRNAs for the binding to the miRNA target site at an mRNA. TSBs have been used to target miR-101 and miR-145 in the CFTR 3' UTR, thus increasing CFTR expression and function in CF nasal epithelial cells as well as correcting chloride efflux mediated by ANO1 [53,57]. Thus, TSBs were successfully designed to prevent the binding of miR-9 to the 3' UTR of ANO1 mRNA [57]. Recently, De Santi and colleagues reverted the inhibition of CFTR mediated by miRNAs via CFTR-specific TSBs in CF bronchial epithelial cells [126]. In particular, TSBs targeting the binding sites of miR-223-3p and miR-145-5p in the 3' UTR were encapsulated in poly-lactic-co-glycolic acid (PLGA) nanoparticles. Their delivery via nebulization, alone or in combination with CFTR modulators (ivacaftor/lumacaftor and ivacaftor/tezacaftor), effectively enhanced the protein levels of CFTR. Due to the high specificity of TSBs for CFTR and the biocompatibility of PLGA, this strategy has low off-targets, toxicity, and immunogenicity with respect to other approaches.

Manipulation of miRNAs may also be relevant to achieve F508del CFTR correction. As mentioned above, the upregulation of TGF-β increased the expression of miR-145, which consequently caused the inhibition of CFTR [61,62]. In addition, functional experiments revealed that high levels of miR-145 nullified the effects of the CFTR modulators. Thus, an antimiR strategy directed against miR-145 was able to enhance the benefits of F508del CFTR correction in CF airway epithelia.

8. Conclusions

Every CF patient is unique with respect to her or his clinical manifestations as a result of the heterogeneity of CFTR mutations. Over the past decade, major advances have been achieved in the comprehension and treatment of CF, thus allowing the implementation of personalized medicine. Patients are stratified according to their CFTR mutations, which determine the therapeutic strategy [43]. Nonetheless, the phenotypic variability in patients with the same CFTR genotype remains a major therapeutic challenge. Moreover, many rare pathogenic mutations still lack efficient therapeutic options. Future progress in CF research requires, inter alia, appropriate cellular and animal models. In vivo models (i.e., mice, pigs, and ferrets) are particularly useful to explore the pathophysiology of CF and therapeutic strategies [127]. Nevertheless, their use in research has many limitations and disadvantages [127]. CF murine models fail to spontaneously develop lung disease, as well as bacterial infections, due to their short life span [128]. Meanwhile, pigs, which are highly attractive candidates, are substantially more complicated to manage (i.e., facility spaces, food, and costs) compared to other models [128]. The advent of organoid technology provides a unique platform to study a variety of human disorders, including CF. Organoid models created from patient cells are an affordable and simple way to explore disease mechanisms and to evaluate drug effects [129]. For example, Dekkers and colleagues demonstrated that the forskolin-induced swelling of rectal organoids isolated from small endoscopic biopsies from CF patients varied according to the specific CFTR mutation [130].
Deregulated miRNAs have a significant impact on the clinical course of CF [117]. The altered abundance of miRNAs affects multiple aspects of CF pathology. Indeed, miRNAs directly modulate the expression of CFTR (as this is the case for miR-101, -145, -223, -494, and -509) or indirectly affect CF (as this is the case for miR-138) by regulating other channels or proteins that act downstream of CFTR. Moreover, miRNA deregulation critically contributes to the inflammatory process (the case for miR-126, 199a-5p) or facilitates airway obstruction by altering the composition of mucus (the case of miR-17 and miR-146).

In human pathologies, miRNAs may serve as diagnostic, prognostic, and predictive biomarkers due to their intrinsic properties, such as stability, facile detection, and disease-specific expression in non-invasive specimens [92]. There are no reliable biomarkers to monitor CF progression. It is possible that circulating miRNAs will help with evaluating and predicting fluctuations in lung inflammations as well as treatment responses. Currently, an ongoing clinical trial is measuring miRNAs extracted from blood specimens in order to stratify CF patients affected by mild or severe lung disease (NCT02992080, clinicaltrials.gov). Thus far, only a few studies have attempted to identify miRNAs as CF biomarkers. Large validation cohorts will be necessary to confirm the preliminary results obtained in these studies.

Understanding miRNA deregulation, distribution, targets, and function could unravel the biological processes that drive CF pathogenesis and, in addition, prepare the grounds for therapeutic interventions on miRNAs. Due to their small size and low antigenicity, miRNAs constitute attractive pharmacological agents for regulating the abundance of target mRNAs/proteins and pathways of interest. Most miRNA-relevant strategies rely on the use of synthetic, sequence-specific molecules that mimic or repress the expression of miRNAs. Some studies have already identified interesting miRNA-based therapeutics (i.e., TSBs targeting the binding sites of miR-223-3p and miR-145-5p) that could be used either alone or in combination with CFTR modulators [126]. However, several challenges, which include safe and organ-specific delivery, long-term efficacy, as well as side effects of prolonged treatments, need to be overcome. Pending the resolution of these issues, it is reasonable to hope that miRNA-targeting agents will eventually be introduced into the clinics.

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