Sensitivity and Specificity of Gene Xpert in the Diagnosis of Spinal Tuberculosis: A Prospective Controlled Clinical Study

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Abstract

Study Design: Prospective matched cohort study

Objective: To evaluate the sensitivity and specificity of Gene Xpert in the diagnosis of spinal tuberculosis.

Methods: From January 2016 to August 2018, Gene Xpert results were prospectively studied in 68 patients of clinicoradiologically suspected spinal tuberculosis (STB) and a control group (CG) of 92 patients, all of whom underwent computed tomography–guided/C-arm-guided/open surgical biopsy. Sensitivity, specificity, positive predictive value, and negative predictive value are obtained using standard equations.

Results: Out of 68 cases of STB, Gene Xpert was positive in 62 (true positive: 62/68) and negative in 6 (false negative: 6/68). Gene Xpert was negative for all 92 cases of CG (true negative: 92/92, false positive 0/92). Thus, in our series, sensitivity of Gene Xpert is 91.18%, specificity is 100%, positive predictive value is 100%, and negative predictive value is 93.88%. Out of all cases of STB, 62/68 (91.18%) were Gene Xpert positive, but only 35/64 (54.69%) was acid-fast bacilli (AFB) culture positive and 53/60 (88.33%) was histopathologically conclusive of TB. Also, Gene Xpert was positive in 7/7 (100%) cases of STB in which histopathology were inconclusive and 25/29 (86.21%) cases of STB in which AFB cultivation was negative.

Conclusion: In STB, Gene Xpert clearly outperforms AFB culture and histopathology due to its high sensitivity and specificity apart from being rapid in diagnosis. Hence it is justified to diagnose spinal tuberculosis by Gene Xpert though histopathology is confirmative and AFB culture remains the gold standard.

Keywords
spinal, tuberculosis, Gene Xpert, sensitivity, specificity

Introduction

*Mycobacterium tuberculosis* is the most common microbiological agent that causes granulomatous, chronic infection in India and many developing countries. The spine is the most common site of skeletal tuberculosis and accounts for 50% of cases.1 Early diagnosis and management of spinal tuberculosis (STB) have special importance in preventing serious complications including the neurological deficit, irreversible disability, and spinal deformity.

Confirmation of *M tuberculosis* by positive culture takes 6 to 8 weeks, which limits its usefulness.2 Histopathology is an important diagnostic tool but is technically demanding and time consuming.3 Xpert MTB/RIF assay (Gene Xpert) is a rapid automated molecular test with high accuracy for pulmonary4 and various extrapulmonary samples of TB such as cerebrospinal fluid, urine, lymph node and other tissues.5,6

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However, adequate data regarding sensitivity and specificity in samples collected from spinal tuberculosis is not available and comparable control group is not available in studies in published literature. We tried to evaluate sensitivity and specificity of Gene Xpert in relation to patients with spinal tuberculosis diagnosed on clinicoradiological parameters, HP (histopathology) and AFB (acid-fast bacilli) culture and compared it to a control group of patients who were diagnosed to be nontubercular lesions based on histopathological and/or microbiological diagnosis.

**Methods**

After approval from institutional review board, we prospectively studied 68 patients with clinicoradiologically suspected STB who underwent biopsy procedures by computed tomography (CT)–guided, percutaneous C-arm-guided, or open surgical procedures from January 2016 to August 2018. All samples were tested for AFB stain, standard AFB culture, histopathology, and Gene Xpert. Similarly, a control group (CG) of 92 patients was studied, who were clinicoradiologically and histopathologically non-TB lesions who had similar investigations from a biopsy procedure. The results of Gene Xpert were analyzed in relation to histopathology and AFB culture report at 42 days.

Patients’ clinical details, procedure details, and follow-up data were recorded in electronic medical record system along with case sheet. Radiological studies, including X-ray, magnetic resonance imaging (MRI) and CT scan were studied using PACS (picture archiving and communication system). CT-guided biopsy was performed under local anesthesia and all percutaneous C-arm-guided biopsy were done under general anesthesia using standard protocols. The rest of the samples were obtained by open surgical procedures.

All samples underwent following laboratory examinations,

1. Microscopic smear examination for AFB by Ziehl–Neelsen technique and examined under 100× oil immersion lens.
2. TB culture was done using incubating culture for 42 days using BaCT/ALERT 3D mycobacterial detection system (Culture bottles—BaCT/ALERT MP medium-enriched middle brook 7Hg broth and LJ media [Lowenstein-Jensen medium] containing p-nitrobenzoic acid [PNB]).
3. Gene Xpert (Xpert MTB/RIF assay) (Cepheid Inc, Sunnyvale, CA, USA) was done with adherence to the protocol in a real-time format using 6 fluorescent probes called molecular beacons (real-time PCR-CBNAAT—cartridge-based nucleic acid amplification test—completely closed system—Sensitivity of the test 131 test CFU/mL).

Tissue samples were grounded first followed by treatment by N-acetyl-L-cysteine (NALC)-NaOH solution (digestion, decontamination, and concentration procedure). Four percent NaOH was mixed with an equal quantity of 2.9% sodium citrate solution. NALC powder was then added to this solution of NaOH–Na citrate to achieve a final concentration of 0.5%, that is, for 100 mL NaOH-NALC solution; 0.5 g NALC powder was added. This final solution was added in a volume equal to that of the specimen. Whole material was vortexed and left for 15 to 20 minutes taking care not to exceed treatment time for more than 20 minutes.

Post decontamination another treatment was done with the buffer provided in the kit. The sample was then inoculated in cartridge taking care that no particulate matter entering the cartridge.

Pus sample did not need prior treatment and was directly treated with the buffer provided in the kit.

4. Histopathology study: Tissue was stained with hematoxylin and eosin stain and was seen under a microscope for AFB and epithelioid cell granuloma, Langerhans cell with or without caseation (except for abscess/liquid samples—where histopathology was not possible for practical purposes).

All laboratory investigations—AFB stain, standard AFB culture, histopathology, and Gene Xpert—were done in the same LABORATORY. All reports were confirmed by the concerned senior pathologist.

Diagnostic accuracy was first assessed by simple comparison of results of Gene Xpert in relation to confirmation by histopathological evidence of granulomatous inflammation and/or isolation of *M tuberculosis* bacilli by culture. Additionally, clinicoradiological presentation and response to antitubercular therapy at follow-up were added for further confirmation of the diagnosis of tuberculosis. In addition, the results of Gene Xpert in the STB group were further analyzed in relation to histopathology and AFB culture.

**Results**

This study included 160 specimens, all of which were sent to the same laboratory between January 2016 and August 2018. Among the 160 tissue samples, CT-guided fine-needle aspiration cytology (FNAC) was done in 29 cases, percutaneous wide bore needle biopsy in 65 cases, and open surgical biopsy was done in 66 cases (Table 1).

Out of 68 cases of STB confirmed by either histopathology or AFB culture or both, in 62 cases Gene Xpert detected *M tuberculosis* (true positive: 62/66) and in 6 cases *M tuberculosis* was not detected by Gene Xpert (false negative: 6/68). Gene Xpert was negative for all 92 cases that were clinicoradiologically and histopathologically non-TB lesions (true negative: 92/92, false positive: 0/92). Thus, the sensitivity of Gene Xpert for spinal tuberculosis in our series is 91.18%, specificity is 100%, a positive predictive value is 100%, and negative predictive value is 93.88% (Table 2). CT-guided FNAC was done in 16 cases of STB and 13 cases of CG. Because of such limited sample size in these subgroups, we could not analyze sensitivity and specificity of each biopsy method.
Table 1. Types of Biopsy Procedures.

| Disease | CT-Guided FNAC | Percutaneous C-Arm-Guided Biopsy | Open Surgical Biopsy |
|---------|----------------|----------------------------------|----------------------|
| STB     | 16             | 20                               | 32                   |
| CG      | 13             | 45                               | 34                   |
| Total   | 29             | 65                               | 66                   |

Abbreviations: CT, computed tomography; FNAC, fine-needle aspiration cytology; STB, spinal tuberculosis; CG, control group.

Table 2. Results of Gene Xpert in STB and CG.

| Disease | Positive | Negative | Total |
|---------|----------|----------|-------|
| Test    |          |          |       |
| Positive| FP = 0   | TP = 62  | 62    |
| Negative| FN = 6   | TN = 92  | 98    |
| Total   | 68       | 92       | 160   |

Abbreviations: STB, spinal tuberculosis; CG, control group; TP, true positive; FP, false positive; FN, false negative; TN, true negative.

Out of 68 cases of spinal tuberculosis, 8 were pus samples and hence histopathology was not feasible—in the remaining 60 cases, the sample was obtained for histopathology, which was positive in 53 cases (53/60 = 88.33%) and was inconclusive in 7 cases (7/60 = 11.67%). Similarly, out of 68 cases of spinal tuberculosis, in 4 cases AFB culture could not be done—out of the remaining 64 cases, AFB culture grew *M. tuberculosis* in only 35 cases (35/64 = 56.69%) (Table 3). Thus, the sensitivity of Gene Xpert, histopathology, and AFB culture with regard to comprehensive final diagnosis is 91.18%, 88.33%, and 56.69%, respectively.

On further subgroup analysis, among 53 patients in whom histopathology was positive for tuberculosis, Gene Xpert was positive in 47 cases (47/53 = 88.68%) and Gene Xpert was negative in 6 cases (6/53 = 11.32%). Similarly, among 29 cases in which AFB culture grew *M. tuberculosis*, Gene Xpert was positive in 23 cases (23/29 = 82.76%) and negative in 6 cases (6/29 = 20.69%). Also, Gene Xpert was positive in all 7 cases of STB in which histopathology was inconclusive (7/7 = 100%). Similarly, out of 29 cases of STB in which AFB culture was negative, Gene Xpert was positive in 25 cases (25/29 = 86.21%) (Table 4).

Out of all the samples in which Gene Xpert was positive to *M. tuberculosis*, rifampicin resistance was detected in only 1 case and was replicated in the drug sensitivity report. The patient was put on appropriate second-line antitubercular drugs (ATD) and responded well to therapy. The final diagnosis of patients in the CG is presented in Table 5.

**Discussion**

Tuberculosis of the spine requires prompt diagnosis and early initiation of treatment, otherwise patient may develop kyphosis and/or neurological complication (paraplegia) and other adverse consequences. For diagnosis of osteoarticular tuberculosis, no single modality like AFB culture, AFB staining, histopathology is definite of ascertaining it.

The sensitivity of AFB staining in various published studies was reported in the range of 20% to 60%, which can be increased to a small extent using special physical and/or chemical methods of sample processing and special equipment like fluorescent microscopy/LED fluorescent microscopy. Lakhanpal et al. reported 49.53% positivity by AFB culture and a range of 48.6% to 80% has been reported in the literature. Various factors accounted for low AFB culture sensitivity such as paucibacillary as nature of disease, type of species, stain used, experience of the technical person, and so on. In addition to these, it has other limitations like the requirement of live organisms, the long incubation period of 2 to 8 weeks, and biosafety issues. These causes delay in diagnosis and result in poor disease control and increased health care cost.

CT/fluoroscopic-guided FNAC biopsy technique has been described as a potential “game changer” for TB control. This is a new approach to DNA sequence analysis that uses fluorogenic reporter molecules—nucleic acid hybridization probes—called molecular beacons. The Gene Xpert detects *M. tuberculosis* and rifampicin resistance by PCR amplification of the rifampin resistance determining region (RRDR) of the *M. tuberculosis* *rpoB* gene and subsequent probing of this region for mutations that are associated with rifampicin resistance.

This multifunctional diagnostic platform has many advantages over other diagnostic tools. It is an automated, closed system that requires minimal technical expertise, providing a diagnosis of TB and at the same time assessment of rifampicin resistance within 2 hours as opposed to 1 day for smear microscopy, 16 days using liquid culture and 20 days using solid culture. The test reduces the time to start definite treatment from 56 days (interquartile range [IQR] 39-81) to 5 days (IQR, 2-8). It has an analytic sensitivity of 5 genome copies of purified DNA and 131 CFU/mL of *M. tuberculosis* spiked into sample. Also it has less cross-contamination, requires much less biosafety infrastructure and minimal training.

Molecular techniques have been a great revolution in the diagnosis of tuberculosis providing rapid results while being highly sensitive in pulmonary tuberculosis detection and drug-resistant cases. Also role of Gene Xpert has been established in many types of extrapulmonary samples, such as for lymphomas.
node samples, sensitivity ranged from 50% to 100% whereas for cerebrospinal fluid (TB meningitis), the pooled sensitivity was 62.8% (95% CI 47.7%-75.8%) and pooled specificity was 98.8% (95% CI 95.7%-100%). For diagnosis of joint tuberculosis, sensitivity, specificity, accuracy, positive and negative predictive values of PCR were 82.65% (81/98), 91.00% (91/100), 86.87% (172/198), 90.00% (81/90), and 84.26% (91/108), respectively as demonstrated by Sun et al in their study.

In this study, we have tried to find sensitivity and specificity of Gene Xpert in spinal tuberculosis cases, for which adequate literature data is not available yet. This is a prospective study and observed sensitivity of Gene Xpert is 91.18% and specificity is 100%, which correlates well with similar study by Held at al showing sensitivity of 95.6% and specificity of 96.2% for STB and study done by Arockiaraj et al demonstrating sensitivity of 71.2% and specificity of 100% compared with composite reference standard. Positive predictive value is 100% and negative predictive value is 93.88% in our study, that is, negative result reliably excludes TB. This is the only study that compares Gene Xpert results in TB spine with controlled group of established non-TB cases.

However, these tests are not suitable for patient monitoring as these tests detect DNA from both viable and nonviable bacilli. In our series, out of 6 patients of STB group in which Gene Xpert was negative (false negative), all 5 patients were on ATD (2 weeks to 1.5 years) before biopsy procedure, which might have affected the result of Gene Xpert. But, in the same STB group there were another 12 patients who were on ATD for a different duration (1 week to 2 years) before biopsy procedure but their Gene Xpert was positive. So, starting of ATD before sample collection cannot be solely attributed to negative results of Gene Xpert, nor can it be used as an ideal tool for monitoring the treatment as per this study. But, role of Gene Xpert after treatment with ATD in diagnosis and monitoring cases of STB will be of interest to learn in future studies.

Thus, from a purely technical perspective, no test for TB is perfect. Microscopy, conventional culture, and drug sensitivity test (both phenotypic and genotypic) all have shortcomings and limitations related to accuracy and effectiveness, operator dependency, training and resource requirements, and biosafety. Gene Xpert clearly outperforms microscopy and should be used as the initial diagnostic test but clinicoradiological presentation; culture and histopathology should be rechecked before excluding the diagnosis of STB. Patients identified positively by Gene Xpert without rifampicin resistance should receive appropriate first-line ATD immediately. Rapid drug sensitivity testing for rifampicin is recommended by the World Health Organization.

Patients at risk of drug resistance in whom rifampicin resistance is detected by Gene Xpert should be placed on an appropriate multidrug-resistant TB regimen immediately and isoniazid continued until the drug sensitivity test result for isoniazid is available. Additional specimen should be provided for conventional culture and DST against other first- and second-line drugs according to World Health Organization recommendations and their treatment adjusted accordingly.

### Table 3. Gene Xpert Results Compared With Histopathology and Acid-Fast Bacilli Culture.

|                      | Histopathology |                      | Acid-Fast Bacilli |
|----------------------|----------------|----------------------|-------------------|
|                      | Positive | Inconclusive | Not Done | Total | Positive | Negative | Not Done | Total |
| Gene Xpert           | Positive  | 47          | 7        | 8      | 62       | 33       | 25       | 4      | 62    |
|                      | Negative  | 6          | 0        | 0      | 6        | 2        | 4        | 0      | 6     |
| Total                | 53       | 7          | 8        | 68     | 35       | 29       | 4        | 68     |

### Table 4. Results of Gene Xpert, Histopathology, and AFB Culture in the Study.

|                  | Gene Xpert | Histopathology | AFB Culture |
|------------------|------------|----------------|-------------|
| STB (68)         | Positive (62) | Positive (47) | Positive (25) |
|                  | Negative (0)  | Negative (19)  | Not done (3) |
|                  | Inconclusive (7) | Positive (3) | Negative (3) |
|                  | Not done (8)  | Not done (0)   | Not done (0) |
| Negative (6)     | Positive (6)  | Positive (2)   | Negative (4) |
|                  | Negative (0)  | Negative (0)   | Not done (0) |
|                  | Inconclusive (0) | Positive (0) | Not done (0) |
| CG (92)          | Negative (92) | Negative (84) | Positive (0) |
|                  | Not done (8)  | Negative (82)  | Not done (2) |
|                  |              | Positive (0)   | Negative (0) |
|                  |              | Negative (7)   | Not done (1) |

### Table 5. Final Diagnosis of Patients in the Control Group.

| Final Diagnosis                              | No. of Cases |
|----------------------------------------------|--------------|
| Pyogenic infection/discitis                   | 39           |
| Neoplastic lesions                           | 23           |
| Pathological fractures (osteoporosis)        | 30           |
| Total                                        | 92           |

Abbreviations: AFB, acid-fast bacilli; STB, spinal tuberculosis; CG, control group.
Conclusion

In STB, Gene Xpert outperforms AFB culture and histopathology due to its high sensitivity and specificity apart from being rapid in diagnosis. Hence it is justified to diagnose spinal tuberculosis by Gene Xpert though histopathology is confirmative and AFB culture remains the gold standard. It might be useful to have confirmation by clinicoradiological features, histopathology, and AFB culture before excluding the diagnosis of spinal tuberculosis.

Acknowledgments

The authors thank Dr. Roy & Tribedi Diagnostic Laboratory.

Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The author(s) received no financial support for the research, authorship, and/or publication of this article.

References

1. Rajasekaran S. Spinal Infections and Trauma. 1st ed. New Delhi, India: Jaypee Brothers; 2011.
2. Siddiqui K, Lambert M, Walley J. Clinical diagnosis of smear-negative pulmonary tuberculosis in low-income countries: the current evidence. Lancet Infect Dis. 2003;3:288-296.
3. Maynard-Smith L, Larke N, Peters JA, Lawn SD. Diagnostic accuracy of the Xpert MTB/RIF assay for extrapulmonary and pulmonary tuberculosis when testing non-respiratory samples: a systematic review. BMC Infect Dis. 2014;14:709. doi:10.1186/s12879-014-0709-7.
4. Boehme CC, Nabeta P, Hillemann D, et al. Rapid molecular detection of tuberculosis and rifampin resistance. N Engl J Med. 2010;363:1005-1015. doi:10.1056/NEJMoA0907847.
5. Denkinger CM, Schumacher SG, Boehme CC, Dendukuri N, Pai M, Steingart KR. Xpert MTB/RIF assay for the diagnosis of extrapulmonary tuberculosis: a systematic review and meta-analysis. Eur Respir J. 2014;44:435-446. doi:10.1183/09031936.0007814.
6. Lawn SD, Nicol MP. Xpert® MTB/RIF assay: development, evaluation and implementation of a new rapid molecular diagnostic test for tuberculosis and rifampicin resistance. Future Microbiol. 2011;6:1067-1082. doi:10.2217/fmb.11.84.
7. World Health Organization. Fluorescent Light-Emitting Diode (LED) Microscopy for Diagnosis of Tuberculosis Policy Statement. Geneva, Switzerland: World Health Organization; 2011. http://apps.who.int/iris/bitstream/10665/44620/1/9789241598811_eng.pdf. Accessed June 4, 2019.
8. Lakhanpal VP, Tuli SM, Singh H, Sen PC. The value of histology, culture and guinea pig inoculation examination in osteo-articular tuberculosis. Acta Orthop Scand. 1974;45:36-42. doi:10.3109/1745367408989119.
9. Chakravorty S, Sen MK, Tyagi JS. Diagnosis of extrapulmonary tuberculosis by smear, culture, and PCR using universal sample processing technology. J Clin Microbiol. 2005;43:4357-4362. doi:10.1128/JCM.43.9.4357.
10. Chang K, Lu W, Wang J, et al. Rapid and effective diagnosis of tuberculosis and rifampicin resistance with Xpert MTB/RIF assay: a meta-analysis. J Infect. 2012;64:580-588. doi:10.1016/j.jinf.2012.02.012.
11. New Diagnostics Working Group of the Stop TB Partnership. Pathways to Better Diagnostics for Tuberculosis. Geneva, Switzerland: World Health Organization; 2009. http://apps.who.int/iris/bitstream/10665/44230/1/9789241598811_eng.pdf. Accessed June 4, 2019.
12. Jain AK, Jena SK, Singh MP, Dhammi IK, Ramachadran VG, Dev G. Evaluation of clinico-radiological, bacteriological, serological, molecular and histological diagnosis of osteoarticular tuberculosis. Indian J Orthop. 2008;42:173-177. doi:10.4103/0019-5413.40253.
13. Kang M, Gupta S, Khandelwal N, Shankar S, Gulati M, Suri S. CT-guided fine-needle aspiration biopsy of spinal lesions. Acta Radiol. 1999;40:474-478. doi:10.3109/02841859909175570.
14. Mondal A. Cytological diagnosis of vertebral tuberculosis with fine-needle aspiration biopsy. J Bone Joint Surg Am. 1994;76:181-184.
15. Small PM, Pai M. Tuberculosis diagnosis—time for a game change. N Engl J Med. 2010;363:1070-1071. doi:10.1056/NEJMe1008496.
16. Piatak AS, Telenti A, Murray MR, et al. Genotypic analysis of Mycobacterium tuberculosis in two distinct populations using molecular beacons: implications for rapid susceptibility testing. Antimicrob Agents Chemother. 2000;44:103-110. doi:10.1128/AAC.44.1.103-110.2000.
17. El-Hajj HH, Marras SA, Tyagi S, Kramer FR, Alland D. Detection of rifampin resistance in Mycobacterium tuberculosis in a single tube with molecular beacons. J Clin Microbiol. 2001;39:4131-4137. doi:10.1128/JCM.39.9.4131.
18. Blakemore R, Story E, Helb D, et al. Evaluation of the analytical performance of the Xpert MTB/RIF test for diagnosis of tuberculosis and rifampicin resistance with Xpert MTB/RIF in a clinic in a single tube with molecular beacons. J Clin Microbiol. 2011;49:2495-2501. doi:10.1128/JCM.00128-10.
19. Boehme CC, Nicol MP, Nabeta P, et al. Feasibility, diagnostic accuracy, and effectiveness of decentralised use of the Xpert MTB/RIF test for diagnosis of tuberculosis and multidrug resistance: a multicentre implementation study. Lancet. 2011;377:1495-1505. doi:10.1016/S0140-6736(11)60438-8.
20. Weyer K, Mirzayev F, Migliori GB, et al. Rapid molecular TB diagnosis: evidence, policy making and global implementation of Xpert MTB/RIF. Eur Respir J. 2013;42:252-271. doi:10.1183/09031936.00157212.
21. Dheda K, Ruhwald M, Theron G, Peter J, Yam WC. Point-of-care diagnosis of tuberculosis: past, present and future. Respiratory. 2013;18:217-232. doi:10.1111/resp.12022.
22. Ghariani A, Jaouadi T, Smaoui S, et al. Diagnosis of lymph node tuberculosis using the GeneXpert MTB/RIF in Tunisia. Int J Mycobacteriol. 2015;4:270-275. doi:10.1016/j.ijmyco.2015.05.011.
23. Nhu NT, Heemskerk D, Thu do DA, et al. Evaluation of GeneXpert MTB/RIF for diagnosis of tuberculous. *J Clin Microbiol*. 2014;52:226-233. doi:10.1128/JCM.01834-13.

24. Sun YS, Lou SQ, Wen JM, et al. Clinical value of polymerase chain reaction in the diagnosis of joint tuberculosis by detecting the DNA of *Mycobacterium tuberculosis*. *Orthop Surg*. 2011;3:64-71. doi:10.1111/j.1757-7861.2010.00115.x.

25. Held M, Laubscher M, Zar HJ, Dunn RN. GeneXpert polymerase chain reaction for spinal tuberculosis: an accurate and rapid diagnostic test. *Bone Joint J*. 2014;96-B:1366-1369. doi:10.1302/0301-620X.96B10.34048.

26. Arockiaraj J, Michael JS, Amritanand R, David KS, Krishnan V. The role of Xpert MTB/RIF assay in the diagnosis of tubercular spondylodiscitis. *Eur Spine J*. 2017;26:3162-3169. doi:10.1007/s00586-017-5076-9.

27. TB CARE I. *International Standards for Tuberculosis Care*. 3rd ed. Hague, Netherlands: TB CARE I; 2014.

28. World Health Organization. *Implementing Tuberculosis Diagnostics: A Policy framework*. Geneva, Switzerland: World Health Organization; 2015.

29. World Health Organization. *Guidelines for the Programmatic Management of Multidrug-Resistant Tuberculosis*. Geneva, Switzerland: World Health Organization; 2011.

30. World Health Organization. *WHO Treatment Guidelines for Drug-Resistant Tuberculosis*. Geneva, Switzerland: World Health Organization; 2016.