Shaping the Latitudinal Diversity Gradient: New Perspectives from a Synthesis of Paleobiology and Biogeography

David Jablonski,1,* Shan Huang,2 Kaustuv Roy,3 and James W. Valentine4

1. Department of Geophysical Sciences, University of Chicago, Chicago, Illinois 60637; 2. Senckenberg Biodiversity and Climate Research Center, Senckenberganlage 25, D-60325 Frankfurt (Main), Germany; 3. Section of Ecology, Behavior, and Evolution, Division of Biological Sciences, University of California, San Diego, La Jolla, California 92093; 4. Department of Integrative Biology, University of California, Berkeley, California 94720

Submitted July 13, 2016; Accepted September 16, 2016; Electronically published December 2, 2016

Online enhancements: supplemental tables. Dryad data: http://dx.doi.org/10.5061/dryad.qd53c.

ABSTRACT: An impediment to understanding the origin and dynamics of the latitudinal diversity gradient (LDG)—the most pervasive large-scale biotic pattern on Earth—has been the tendency to focus narrowly on a single causal factor when a more synthetic, integrative approach is needed. Using marine bivalves as a model system and drawing on other systems where possible, we review paleobiologic and biogeographic support for two supposedly opposing views, that the LDG is shaped primarily by (a) local environmental factors that determine the number of species and higher taxa at a given latitude (in situ hypotheses) or (b) the entry of lineages arising elsewhere into a focal region (spatial dynamics hypotheses). Support for in situ hypotheses includes the fit of present-day diversity trends in many clades to such environmental factors as temperature and the correlation of extinction intensities in Pliocene bivalve faunas with net regional temperature changes. Support for spatial dynamics hypotheses includes the age-frequency distribution of bivalve genera across latitudes, which is consistent with an out-of-the-tropics dynamic, as are the higher species diversities in temperate southeastern Australia and southeastern Japan than in the tropical Caribbean. Thus, both in situ and spatial dynamics processes must shape the bivalve LDG and are likely to operate in other groups as well. The relative strengths of the two processes may differ among groups showing similar LDGs, but dissecting their effects will require improved methods of integrating fossil data with molecular phylogenies. We highlight several potential research directions and argue that many of the most dramatic biotic patterns, past and present, are likely to have been generated by diverse, mutually reinforcing drivers.

Keywords: extinction, speciation, geographic range dynamics, phylogeny, historical biogeography, paleobiology.

INTRODUCTION

The latitudinal diversity gradient (LDG), the most pervasive large-scale biological pattern on Earth, has been documented and debated for more than two centuries. Multiple reviews have attested to the increase in diversity of species and higher taxa from poles to tropics and cataloged the many hypotheses on underlying mechanisms (e.g., Pianka 1966; Rohde 1992; Gaston 2000; Hillebrand 2004; Mittelbach et al. 2007; Krug et al. 2009; Donoghue and Edwards 2014; Fine 2015; Schluter 2016), leading Brown (2014, p. 9) to state that “even as the patterns have become clearer . . . the explanations have remained elusive and controversial.” However, others have further to argue that a number of supposedly opposing hypotheses about the LDG actually represent different facets that should be integrated to gain a fuller understanding of the origin and maintenance of this biodiversity pattern.

One such artificial dichotomy, the chief focus of this essay, involves hypotheses that the equator-to-pole profile of the LDG is essentially shaped by local environmental factors that determine the number of species and higher taxa at a given latitude (for brevity, we will call these in situ hypotheses), as opposed to hypotheses that center on the long-term spatial dynamics of clades across latitudes. The crux of this dichotomy is the explanatory power of past and present conditions within the region where diversity is being measured, as opposed to processes that closely link diversity in the focal region to events outside it. We argue that, in fact, in situ processes and spatial dynamics have likely shaped the modern LDG in concert, as illustrated by the simple conceptual model in figure 1. Consider two adjacent regions, α and β, which can represent two adjacent latitudinal bands or biogeographic provinces anywhere along the LDG. (Note that the focus of this essay is on the mechanisms underlying the shape of the full LDG rather than
those responsible for a binary, tropical-vs.-extratropical contrast in richness; the latter would mostly fall under an in situ heading, aside from those hypotheses holding that tropical diversity is promoted by the influx of taxa from higher latitudes.) The richness of each region at a given time is partly a function of (i) the environmental and habitat characteristics of each region (\(T_a\) and \(T_b\)), (ii) the speciation rates in each region (\(s_a\) and \(s_b\)), and (iii) their respective extinction rates (\(x_a\) and \(x_b\)), which together constitute the in situ component because they operate within each region. In addition, the richness of each region is also a function of dispersal into each area (\(d_a\) and \(d_b\)), that is, the larger-scale spatial dynamic (for further details, see Goldberg et al. 2005; Jablonski et al. 2006; Roy and Goldberg 2007).

We will support this integrative view by a combination of present-day and fossil data that provide evidence for each type of dynamic and, therefore, for their simultaneous operation. We frame our discussion using the marine Bivalvia, which have become a model system for large-scale spatial and temporal analyses for at least three reasons. First, their well-documented biogeography shows a strong LDG (fig. 2) that mirrors that of other groups on land and sea (Hillebrand 2004; Tomasovych et al. 2016). Second, they exhibit relatively high taxonomic, phenotypic, and functional diversity, including a wide range of trophic groups ranging from suspension feeding through chemo- and photosymbiosis to predation and parasitism. And third, their rich fossil record allows robust estimates of origination, regional and global extinction, and geographic range dynamics through time (Crame 2000a, 2000b, 2002; Jablonski et al. 2006, 2013; Bieler et al. 2013; Berke et al. 2014; Huang et al. 2015; Mondal and Harries 2016). The ideas presented here should apply broadly across different groups of organisms, and where possible we draw on examples from other groups as well. We will also briefly touch on a related (false) dichotomy, that is, whether the tropics are a cradle or museum of biodiversity.

Evidence for In Situ Controls

Many analyses, both marine and terrestrial, have found that present-day temperature (annual mean and, to some extent, variance) along with a handful of other physical factors, such as precipitation on land and productivity in the sea, are powerful first-order predictors of the profile of the LDG on global and continental scales (Fine 2015 and references therein) and of the boundaries between biogeographic units, such as provinces (Olson et al. 2001; Lomolino et al. 2010; Belanger et al. 2012; Briggs and Bowen 2012). The consensus, reasonably, has been that such correlations provide strong evidence for in situ controls and that species and higher-taxon richness within latitudinal bands or provinces track climate almost instantaneously, in that diversity often fits well with present-day factors despite the enormous changes in climate at most locations over the past 18,000 years. However, the species-rich bathyal (deep-sea) benthos suggests that tropical temperatures are not necessary to generate and maintain high levels of diversity (Rex and Etter 2010; Valentine and Jablonski 2015). One hypothesis is that high bathyal diversity indicates a role for damped seasonality in, among other potential factors, temperature and organic carbon flux (and thus trophic stability), in contrast to the cold but more
seasonal and depauperate polar sea (Valentine and Jablonski 2015). More generally, while a strong correlation between latitudinal trends in mean annual temperature and regional species richness is frequently emphasized (including for bivalves; Roy et al. 1998), many other environmental variables covary along this gradient, as do such historical effects as changes in past climate and habitats (e.g., those driven by Pleistocene glaciations). This multitude of potential drivers makes it difficult to evaluate the role played by different present-day variables and to separate present-day and historical factors (e.g., Jetz et al. 2004; Ricklefs 2004; Erwin 2009; Field et al. 2009; Svenning et al. 2015; Valentine and Jablonski 2015).

Large-scale natural experiments can shed light on the role played by in situ climate changes in shaping the LDG, and inferences about the variability of past climatic and other environmental conditions have been used to explain regional diversity patterns. The bivalve fossil record shows that regional extinction is strongly correlated with net temperature change since the mid-Pliocene and regional extinction in Pliocene bivalve faunas (fig. 3). In particular, the striking correspondence between modest net temperature changes and regional losses in the warm-temperate Pliocene faunas is consistent with the view that such historical factors as past climatic changes and extinctions have played a major role in shaping the modern LDG (e.g., Mannion et al. 2014). The temperature differences between modern and full-glacial Pleistocene conditions were greater than those in the Pliocene, albeit with roughly parallel interregional patterns (see Annan and Hargreaves 2013; Precht and Aronson 2016), and so could have contributed to, or reinforced, the observed trend. In any case, bivalves show differential regional extinctions with latitude that are positively related to net regional cooling over the past 3 million years and, probably, to the distribution of temperature minima in the Pleistocene. Most of these extinctions were regional, with individual clades contracting equatorward in the face of climate cooling (Valentine et al. 2013), progressively sharpening the LDG as the poles shifted to a refrigerated state (albeit with smaller, repeated distributional shifts during glacial/interglacial cycles). Detailed interregional analyses comparing the effects of time-integrated temperature change or maximum regional excursions against net biotic change would be valuable.

The pattern in figure 3 also supports the more specific view that regions that experienced lesser climate change have harbored, and perhaps have generated, more diversity than regions that experienced greater changes (e.g., Jansson and Dynesius 2002; Pyron and Wiens 2013; Mannion et al. 2014; Fine 2015; Claramunt and Cracraft 2015; Pulido-Santacruz and Weir 2016). The effects of such variation can be manifest at a variety of scales and locations. For example, within the tropics, present-day local diversity in marine fishes is inversely related to distance from historically stable reef areas (Pellissier et al. 2014). In the temperate zones, the high plant diversity in the South African Cape Floristic Region has been attributed to the apparently damped climate fluctuations for its latitude (Dynesius and Jansson 2000; Schnitzler et al. 2011; for additional examples, see also Jansson 2003; Svenning et al. 2015), and climate velocity—defined as the rate of displacement of climatic conditions across a region—has been shown to influence geographic patterns of endemism in multiple vertebrate groups (Sandel et al. 2011), which in turn affects their LDGs. In the deep-sea benthos, more direct analyses using estimates of paleotemperature and paleoproductivity show that regional species richness of deep-sea invertebrates responded predictably to past changes in climate (Hunt et al. 2005; Yasuhara et al. 2012).

Taken together, figure 3, the extensive literature showing a close relation today between both marine and terrestrial diversity and regional climate, and the smaller literature relating fossil diversity to climate shifts (Erwin 2009; Mannion et al. 2014) strongly suggest that regional diversity can change substantially when the environmental pa-

Figure 3: Regional extinction in Pliocene faunas versus net temperature change from the Pliocene to the present day. Regional temperature changes are estimated using a spatially explicit model of Pliocene paleotemperatures based on stable isotopes in microfossils (Dowsett et al. 2010, 2013) and thus are independent of molluscan distribu
tional data; bivalve data are from Valentine et al. (2013) and Jablonski et al. (2013). CA = California; Indo = Indonesia; Ice = Iceland; Med = Mediterranean; Mid-Atl = Mid-Atlantic states, United States; N Sea = North Sea region; NZ = New Zealand; Venez = Venezuela.
rameters shift strongly. For bivalves, such climate-driven steepening of the LDG does not appear to be phylogenetically patterned, at least not at the family level, where comprehensive phylogenetic data are available (Bieler et al. 2014). Comparing diversity changes along each of the temperate coasts of North America from the Pliocene to the present day, a given family may show different extinction rates and/or dispersal dynamics on different coasts (Huang et al. 2015). A better predictor of clade behavior in the face of cooling climates is the geographic range size of species at the Pliocene starting point, with the narrowest-ranging species most prone to disappear from the focal region (Huang et al. 2015; see also Saupe et al. [2015], who found that a measure of niche breadth lacks the predictive power shown by geographic range per se). This result is not surprising in that those species most able to tolerate and disperse among a range of latitudes and environments might be expected to survive longest, but it usefully demonstrates that this tolerance effect can override family-level clade membership. Studies of present-day terrestrial birds and mammals, using more indirect methods, have also concluded that climate changes “can force multiple groups into similar diversity patterns even when evolutionary trajectories differ” (Hawkins et al. 2012, p. 825). These findings drive home the message that a given clade can attain similar diversities in two regions in the absence of similar diversification dynamics and that different diversities for a given clade in two regions today need not signal a long divergent history.

Evidence for Spatial Dynamics

Changing climates, and thus tracking of in situ environmental change with or without lags, are not the only impetus for large-scale biogeographic shifts. Our initial work, and that of many others, on the dynamics underlying the LDG was motivated by Stebbins’s (1974) famous question: are the tropics a cradle or a museum of diversity? Stebbins used the cradle metaphor for regions that are diversity generators, presumably with high origination rates, while museums are areas that accumulate diversity, presumably with low extinction rates. Although such a binary characterization of the evolutionary drivers of the LDG has led to considerable empirical work, this dichotomy is increasingly proving to be a false one. The fossil record shows that, over the past 12 million years, bivalve genera preferentially originated in the tropics and expanded their ranges poleward over time—even in the Pleistocene, during times of glaciated poles and steep thermal gradients—while maintaining their tropical presence (Jablonski et al. 2006, 2013 [the latter with substantially improved data based on further integration of literature and museum records]). The pattern is particularly striking given that the sampling is strongly biased to the temperate zones (Valentine et al. 2013), so that some of the clades that originated in the tropics would not have entered into the fossil record until they entered the midlatitudes, thereby underestimating the strength of the dynamic.

Figure 4: Geologic ages of bivalve genera at high and low latitudes, showing a deficiency of young taxa near both poles and a long tail of older taxa in all three regions. Frequency distributions for the Arctic and Antarctic differ significantly from that of the tropics (Komogorov-Smirnov test; tropics vs. Arctic, \( P < .001 \); tropics vs. Antarctic, \( P = .004 \)) but not from each other (\( P = .78 \)). Genus ages are given in table S1 in the Dryad Digital Repository: http://dx.doi.org/10.5061/dryad.qd53c (Jablonski et al. 2017).
Because nearly all bivalve clades that expand out of the tropics (OTT) maintain their presence in the tropics even as they push poleward and because the highest latitudes evidently have low origination rates, the polar regions harbor very few young genera while the tropics have many, with a median age roughly half that of the polar fauna (e.g., Goldberg et al. 2005). However, tropical genera also have a long tail to their age distribution, which includes ~90% of the taxa that constitute the polar tails (fig. 4). Thus, the tropics are both a cradle and a museum of diversity in that they harbor both the oldest and the youngest genera. The poles are mainly a museum of diversity, as they harbor mostly older genera. Of course, present-day age distributions in a given region are uninformative about when taxa actually arrived, but the fossil data can determine the polarity of such movement.

These latitudinal dynamics could still fit into an in situ paradigm if the OTT genera were simply filling gaps left by regional extinctions to keep the global biota in equilibrium with key environmental variables. Tests on, for example, ecological equivalency between extinctions and successful OTT range expansions would therefore be valuable. However, comparison of West Pacific and West Atlantic diversity patterns falsify any simple in situ hypothesis. Southeastern Japan and southeastern Australia, each firmly lodged today in the temperate zone, contain 30% and 14% more species, respectively, than the Caribbean (table 1; see also Tittensor et al. 2010, in which diversity maps incorporating a variety of marine groups show similar or greater coastal diversity in one or both of our two temperate regions relative to the tropical Caribbean, aside from two strongly tropical clades [corals and mangroves]). In situ models would predict the opposite: the Caribbean has an order of magnitude more shelf area than either of the temperate provinces; its mean annual sea surface temperature is 5°–6°C warmer and significantly less seasonal; it is topologically and ecologically more complex, with coral reefs and a scatter of islands of various sizes in contrast to the essentially linear temperate provinces; and it almost certainly hosts a greater variety and intensity of biotic interactions, another factor posited to promote taxonomic diversity (e.g., Mittelbach et al. 2007; Schemske et al. 2009). Diversity is evidently spilling over from the massively diverse west Pacific tropical region into the two adjacent temperate provinces, to an extent that overwhelms the simple environment-diversity relationships that are often seen when examining a single coastline or cumulative diversity in global latitudinal bands.

The very low diversity near the poles relative to diversity maxima in all tropical regions also has analytical consequences: a global model focusing exclusively on the equator-pole gradient will tend to downplay longitudinal interocean differences, which, though little studied, have the potential to be highly informative about the interaction of processes that set large-scale diversity patterns. For example, linear models linking diversity to in situ environmental factors will tend to derive strong support from extreme polar and tropical values, which then drive the conclusions. Yet the deviations from model predictions at midlatitudes and elsewhere, often dismissed as noise, actually represent biologically informative variance. To be sure, some interocean differences arise from historical events, the consequences of which are chiefly restricted to a single ocean basin. The uplift of the Panamanian Isthmus and the oceanographic response was such an event, potentially accounting in part for the surprisingly low modern Caribbean diversity. Although spatially heterogeneous within the Caribbean, this turnover event involved a 14% extinction of bivalve genera and subgenera, by one estimate, but is followed by a 20% diversity rebound on a per-taxon basis (Todd et al. 2002) and so is unlikely to account by itself for the observed discordance between environment and diversity seen in these tropical-temperate contrasts.

### Synthesis and Future Directions

Large-scale diversity patterns are set by the spatial and temporal dynamics of origination, extinction, and geographic range shifts. These dynamics must operate within an environmental framework so that constraints on diversity imposed by the physical and biotic environment are also important. The role played by environmental controls has been corroborated in both paleontological and neontological data sets via the fit of regional diversity variations—

| Region    | No. species | Shelf area (km²) | Lateral extent (°) | Mean annual sea-surface temperature (°C) |
|-----------|-------------|------------------|--------------------|----------------------------------------|
| SE Japan  | 769         | 71,200           | 7                  | 21.4                                   |
| Caribbean | 537         | 912,100          | 27                 | 27.0                                   |
| SE Australia | 612    | 61,750           | 14                 | 22.3                                   |

Note: The Caribbean Province is taken to extend from southern Florida and the Yucatan Peninsula to the mouth of the Amazon, the southeastern Japan Province is taken to extend from the southern tip of Kyushu to the Boso Peninsula, and the southeastern Australian (Peronian) Province is taken to encompass southern Queensland and New South Wales (for coastal marine biogeographic compartments, see Spalding et al. 2007; Belanger et al. 2012; Briggs and Bowen 2012). Caribbean data are available in data file S1 of Jablonski et al. (2013). Northeastern Australia and southeastern Japan species lists are provided in table S2 in the Dryad Digital Repository: http://dx.doi.org/10.5061/dryad.qd53c (Jablonski et al. 2017). Shelf area and latitudinal extent were estimated using ArcGIS. Mean annual sea-surface temperature was calculated from MODIS (http://modis.gsfc.nasa.gov/).
The intuitively appealing link between the steepening of the latitudinal thermal gradient and the steepening of the LDG, as in our bivalve analyses, is supported by other lines of post-Paleozoic fossil evidence on land and sea, which show shallow LDGs during warm intervals and steepening ones with cooling (Crame 2002; Rose et al. 2011; Fenton et al. 2016; Marcot et al. 2016). However, Powell (2007) has argued that a major late Paleozoic phase of global cooling preferentially removed tropical endemic brachiopods and therefore weakened the LDG, in contrast to most inferences for Cenozoic cooling, which as cited above evidently diminished extratropical diversity but left tropical diversity unscathed or enhanced and thereby strengthened the LDG (fig. 3). Yet to be determined is whether, for example, late Paleozoic glacial episodes differed fundamentally from late Cenozoic ones, late Paleozoic brachiopods differed fundamentally from post-Paleozoic bivalves and other groups found to have steepened their LDGs with glaciations, or late Paleozoic biotas present taxonomic or sampling issues that undermine the comparability of the patterns.

The biogeographic dynamics in greenhouse worlds, such as the early Cenozoic, have been studied less than the biotic responses to refrigerating poles, as the latter led to the present-day configuration. This gap should be remedied, as the response of the LDG to global warming in the geologic past or the anthropogenically warmed future is of much theoretical and practical interest. Assuming that shallowing LDGs under global warming is the norm, as the bulk of the available evidence indicates, the in situ hypothesis would have the shallowing occur by local origination at higher latitudes, whereas spatial dynamics would drive shallowing by range expansion from lower latitudes. Both mechanisms evidently operated during the mid-Pliocene warm interval (Huang et al. 2015), but more rigorous phylogenetic and biogeographic analyses of the Pliocene biota and analyses of other warming intervals are needed to determine the relative importance of these processes on land and sea and among clades.

Even with today’s steep thermal gradients, some clades lack a strong LDG globally, and some even show an inverse gradient, with diversity minima in the tropics. Such contrarian trends are known in a few terrestrial, marine benthic, and marine pelagic groups, usually embedded within larger clades showing more conventional LDGs (Roy et al. 2000; Krug et al. 2007; Kindlmann et al. 2007; Lucifora et al. 2011; but see Quicke 2012 and Lees et al. 2014 on sampling issues in ichneumonid wasps and gracillariid moths, respectively). Contrarians are neglected vehicles for testing hypotheses on factors promoting “normal” LDGs. For example, contrarian marine bivalve families tend to exhibit life-history strategies involving relatively low fecundities, high per-offspring maternal investment, and low dispersal abilities, which appear to be increasingly advantageous with increasing latitude, likely owing to slowing of development.
at low temperatures and high seasonality (Roy et al. 2000; Krug et al. 2007). Diversity maxima for these bivalve clades tend to be centered around the younger genera, implying higher origination rates in midlatitudes for the contrarian groups (Krug et al. 2007; but also see Pyron and Burbank [2009], who argue for early colonization of the temperate zone and strong niche conservatism in a contrarian snake tribe).

Life histories may contribute to patterns within major marine groups, but we do not know how differences in dispersal relate to marine-terrestrial comparisons. Most terrestrial groups arguably have lower dispersal abilities than marine mollusks bearing pelagic larvae, but mammals, herps, insects, and plants demonstrably tracked high-frequency, large-amplitude Pleistocene climate changes across hundreds of kilometers (Elias 1984; Holman 1995, 1998; Johnson and Blois 2015; Maguire et al. 2015) and thus are quite capable of moving rapidly over wide distances on the relevant timescales. The critical issue does not appear to be dispersal ability per se—even wide-ranging mollusks tend to track isotherms rather than cross them (Jablonski et al. 2013)—but the ability to establish viable populations in novel climate zones. This ability may still hinge on life history (e.g., per capita fecundity of colonists, as in Roy et al. 2001, 2002), but serious comparative analyses from this perspective have not begun.

Present-day distributions are powerful vehicles for evaluating links between diversity and in situ factors, and the integration of phylogenetic models and modern biogeographic data has the potential of becoming increasingly robust, particularly as paleontological data are brought into the mix. Spatial and temporal dynamics are difficult to derive with confidence from information on extant diversity alone. For example, high extinction rates in temperate zones and unexceptional origination rates in the tropics have been inferred for squamates (Pyron 2014) and birds (Pulido-Santacruez and Weir 2016), and carnivorans are inferred to have temperate extinction and dispersal into the tropics (Rolland et al. 2015), using phylogenies that include extant tips. However, some results are based on model comparisons using GeoSSE and related methods, which can be unreliable under certain circumstances (Maddison and FitzJohn 2015; Rabosky and Goldberg 2015). More importantly, analyses relying on extant taxa may not produce reliable estimates of past extinction or spatial shifts (Quental and Marshall 2010; Rabosky 2010), especially given an overarching assumption of a stochastically constant extinction rate. As discussed above, the regional extinctions in the Pli-Pleistocene provide one example of the violation of this assumption, especially since many areas have not seen later evolutionary recoveries. The spatial dynamics described above also indicate that the age of a lineage, whether inferred paleontologically or by estimates from molecular phylogenies, is a poor estimator of the arrival time of that lineage in a region.

Despite our advocacy of integrated studies, we are not arguing that paleontological data can be applied to macroevolutionary and macroecological investigations any less critically than other types of data. Paleontological data can provide a direct window to the spatial and diversification dynamics underpinning modern biodiversity, but spatially structured sampling biases in fossil preservation, discovery, and sampling as well as in publication can generate artificial signal and not simply noise (e.g., Valentine et al. 2013; Hannisdal et al. 2016). For example, the apparent inverse LDGs in Mesozoic dinosaurs, with diversity maxima at midlatitudes (e.g., Mannion et al. 2014), are difficult to interpret, given that nearly all dinosaur species are known only from the locality where they were initially discovered (Dodson 1990) and that sampling is massively biased in favor of midlatitudes (Taylor 2008; and the map in Hunt 2010). The above-cited flattened LDG in early Cenozoic mammals across North America appears to be more robust than the dinosaurian trend, as it is confined to a more evenly sampled region (Maroc et al. 2016) and is centered over a midlatitude portion of the LDG likely to be nearly flat in a greenhouse-climate world (see, e.g., Archibald et al. 2010, 2013 on low seasonality at high Eocene latitudes).

Comparative analyses among clades will require improved methods of integrating fossil data with molecular phylogenies and of developing models for clades having sparser fossil records than marine bivalves—both of these active areas of research. This integration of neontological and paleontological data, promoted by high-throughput genetic methods and the next generation of evolutionary models, will open the door to an enormous range of basic questions; we can mention only a few here. Comparative historical analysis among regions, particularly along longitudinal transects, will help determine when the significant diversity differences among tropical regions were established—an exercise that requires some knowledge of the extinct components of regional biotas. For example, the timing of the massive diversity differences between the marine biotas of the tropical West Pacific and the Caribbean (see fig. 2) is poorly known, owing to the aforementioned fossil-sampling biases (also Johnson et al. 2015). For a given region, the phylogenetic structure of the OTT dynamic may vary among clades or at different times (e.g., according to the steepness of the climate gradient and thus the sharpness of the tropical-temperate boundary). Strict monophyly of the OTT component of clades appears unlikely in clades having large numbers of bridge species (i.e., species straddling the tropical-temperate boundary; Jablonski et al. 2013) but should be stronger as the frequency of bridge species declines, as appears to occur in at least some terrestrial
lineages (e.g., Fine 2015). Accordingly, certain plant clades are highly polyphyletic in their OTT patterns, although a strong phylogenetic signal prevails at higher levels (Donoghue and Edwards 2014).

Still uncertain is whether bivalves or other groups vary strongly in their per-taxon origination with latitude, a question related to diversity-dependent dynamics. Such latitudinal variation appears likely for bivalves given the paucity of young taxa at high latitudes, whether arising in situ or arriving via range expansion, but definitive tests require robust estimates of standing diversity along the LDG through multiple climate cycles. Such estimates, at least in relative terms, can be obtained with some confidence in the well-sampled temperate zones but are more difficult for the crucial but undersampled tropics, particularly in the global diversity maximum in the tropical West Pacific (see Valentine et al. 2013; Vilhena and Smith 2013). On coarser, Phanerozoic timescales, for all marine invertebrates, Kiersling et al. (2010) estimated tropical diversities and found greater per-taxon genus origination in the tropics relative to higher latitudes, and Jablonski (1993, 2005; see also Martin et al. 2007) found excess origination of higher taxa (orders) in the tropics relative to sampling at the species level. These results are consistent with the hypothesis that the tropics have higher per-taxon origination rates in the ocean at least, but more detailed analyses—almost certainly requiring fossil data, given the need to include extinct taxa in the denominator of the equation—are needed to resolve this question.

Finally, we also know little about why some taxa exhibit such strong OTT dynamics while many others do not. In marked contrast to plants, bivalves lack strong phylogenetic patterns in clade success or failure in permanently expanding OTT. Key factors promoting the OTT dynamic in plants, such as frost tolerance, evidently fall more strongly along phylogenetic lines than the analogous traits in bivalves (see Jablonski et al. 2013; see also Fine 2015). One pattern that emerges in bivalves is the tendency for genera with higher diversification rates to contribute disproportionately to the OTT dynamic, suggesting that “speciation pressure” increases the probability that a taxon successfully crosses the tropical-temperate boundary (Jablonski et al. 2013; Tomasovich et al. 2016). In fact, speciation pressure may promote the crossing of major climate boundaries in either direction. For example, the bird clades that evidently invaded the New World tropics from the Northern Hemisphere (i.e., inverting the OTT dynamic; see Kennedy et al. 2014; Claramunt and Cracraft 2015) also show higher diversification rates than clades restricted to either region (Tomasovich et al. 2016). Thus, in groups as disparate as birds and bivalves, genera that cross the tropical-temperate divide have more species in the tropics than do tropical endemics and have more extratropical species than do genera restricted to extratropical regions. New generalizations will continue to emerge from comparative analyses at these large spatial and phylogenetic scales.

Conclusion

We have argued here that a model system that provides spatially explicit historical data yields compelling evidence for both in situ environmental control and interregional dynamics in large-scale diversity patterns, and it would be fascinating to determine whether the two sides are weighted equally across a wide variety of clades and settings. Extending a new set of phylogenetic models for diversification to include in situ climate and habitat components and the spatial dynamics described here remains a challenge but is essential for breaking down the dichotomy that is the focus of this article. The arguments presented here suggest a new generation of large-scale questions about biodiversity, which can best be answered by the integration of paleo- and neontological approaches. Given a synthetic view of today’s LDG shape as the product of origination, extinction, and past geographic shifts of taxa, all of those variables might differ among clades, functional groups, and land masses or ocean basins, even when the first-order gradient is similar among those groups and biomes. Furthermore, as implied by figure 1, these variables are not necessarily independent. For example, the evolution of new taxa in one area can fuel dispersal into an adjacent region through post-speciation range expansion (Jablonski et al. 2006). Similarly, changing climate or habitat characteristics in one area (an in situ dynamic) could promote range expansion into the area of species from adjacent regions. Owing to such interdependence, failure to take into account the dispersal parameter can, in some cases, even affect the reliability of diversification rate estimates (Roy and Goldberg 2007). Neither the data most readily available for extant species nor the fossil record fully capture the interplay of these factors on their own. But integrating spatially explicit fossil data with modern biogeography and phylogeny can provide a powerful vehicle for testing hypotheses and generating new ones. This combined approach will be the most powerful not just for a deeper understanding of past and present biodiversity but for anticipating biotic changes in the face of accelerating global change.

Synthesis of interactions among mechanisms may be fundamental for understanding many of the most dramatic biodiversity patterns. The search for unitary mechanisms is a valuable means of sharpening hypotheses and discarding the weaker ones, but the most extreme patterns may be generated by diverse, mutually reinforcing drivers (as also noted by, e.g., Gaston [2000]). In this light, the West Pacific Coral Triangle is probably the global marine hotspot because it is a “perfect storm” for biodiversity, combining an
array of in situ factors, including warm temperatures, low seasonality, a vast area, a wide range of habitat types (reefs and mangroves to lagoons and open shelves), a complex island-mainland structure, and intense biotic interactions, with a strong spatial dynamic involving the influx of taxa from other tropical regions (e.g., Leprieur et al. 2016). The end-Permian mass extinction was by far the most severe in the Phanerozoic because many mutually reinforcing factors conspired to draw down diversity, most likely with a volcanic trigger (Payne and Clapham 2012). The Cambrian explosion was the most dramatic phenotypic diversification of multicellular organisms because development, ecology, and such external cues as oxygen levels successively fell into place in a mutually reinforcing fashion (Erwin and Valentine 2013). Insects are the most diverse metazoan clade because they have many attributes promoting high speciation and low extinction rates (Mayhew 2007), and several major plant radiations, including Bromeliaceae and Orchidaceae, evidently involved the confluence of multiple innovations and environmental shifts (Donoghue and Sanderson 2015). Such multifactorial explanations do not mean that we cannot rank drivers in particular cases, but they indicate that comparative analysis and synthesis can yield new insights—why the end-Cretaceous or late Devonian extinctions were less severe than the end-Permian extinction, why the recovery from the end-Permian extinction was less dramatic than the Cambrian explosion, why the diversification of squamates or birds was less dramatic than that of insects—and new approaches to integrating fossil and present-day data are likely to be key to our understanding.

Acknowledgments

We thank G. G. Mittlebach and D. W. Schemske for the invitation to participate in the Asilomar symposium that stimulated this overview. We thank the many bivalve specialists who have been generous with taxonomic advice and access to the collections in their care; the combined D. Jablonski–T. Price laboratory for many stimulating discussions; S. M. Kidwell for her usual incisive review; J. L. Bronstein, E. M. Harper, and an anonymous reviewer for valuable comments on the manuscript; S. Edie for assistance with table 1; and the National Science Foundation, the National Aeronautics and Space Administration, and the John Simon Guggenheim Foundation for support. S.H. is supported by the Alexander von Humboldt Foundation through a postdoctoral fellowship.

Literature Cited

Annan, J. D., and J. C. Hargreaves. 2013. A new global reconstruction of temperature changes at the last glacial maximum. Climate of the Past 9:367–376.

Archibald, S. B., W. H. Bossert, D. R. Greenwood, and B. D. Farrell. 2010. Seasonality, the latitudinal gradient of diversity, and Eocene insects. Paleobiology 36:374–398.

Archibald, S. B., D. R. Greenwood, and R. W. Mathewes. 2013. Seasonality, montane beta diversity, and Eocene insects: testing Janzen’s dispersal hypothesis in an equable world. Palaeogeography, Palaeoclimatology, Palaeoecology 371:1–8.

Belanger, C. L., D. Jablonski, K. Roy, S. K Berke, A. Z Krug, and J. W. Valentine. 2012. Global environmental predictors of benthic marine biogeographic structure. Proceedings of the National Academy of Sciences of the USA 109:14046–14051.

Berke, S. K., D. Jablonski, A. Z. Krug, and J. W. Valentine. 2014. Origination and migration drive latitudinal gradients in marine functional diversity. PLoS One 9:e101494.

Bieler, R., P. M. Mikkelsen, T. M. Collins, E. A. Glover, V. L. González, D. L. Graf, E. M. Harper, et al. 2014. Investigating the bivalve tree of life—an exemplar-based approach combining molecular and novel morphological characters. Invertebrate Systematics 28:32–115.

Bieler, R., P. M. Mikkelsen, and G. Giribet. 2013. Bivalvia—a discussion of known unknowns. American Malacological Bulletin 31: 123–133.

Briggs, J. C, and B. W. Bowen. 2012. A realignment of marine biogeographic provinces with particular reference to fish distributions. Journal of Biogeography 39:12–30.

Brown, J. H. 2014. Why are there so many species in the tropics? Journal of Biogeography 41:8–22.

Claramunt, S., and J. Cracraft. 2015. A new time tree reveals Earth’s history’s imprint on the evolution of modern birds. Science Advances 2015:e1501005.

Cornell, H. V. 2013. Is regional species diversity bounded or unbounded? Biological Reviews 88:140–165.

Crame, J. A. 2000a. Evolution of taxonomic diversity gradients in the marine realm: evidence from the composition of Recent bivalve faunas. Paleobiology 26:188–214.

———. 2000b. Intrinsic and extrinsic controls on diversification in the Bivalvia. Pages 135–148 in S. J. Culver and P. F. Rawson, eds. Biotic response to global change: the last 145 million years. Cambridge University Press, Cambridge.

———. 2002. Evolution of taxonomic diversity gradients in the marine realm: a comparison of Late Jurassic and Recent bivalve faunas. Paleobiology 28:184–207.

Dodson, P. 1990. Counting dinosaurs: how many kinds were there? Proceedings of the National Academy of Sciences of the USA 87: 7608–7612.

Donoghue, M. J., and E. J. Edwards. 2014. Biome shifts and niche evolution in plants. Annual Review of Ecology, Evolution, and Systematics 45:547–572.

Donoghue, M. J., and M. J. Sanderson. 2015. Confluence, synonymity, and depauperization in plant diversification. New Phytologist 207:260–274.

Dowsett, H., K. M. Foley, D. K. Stoll, M. A. Chandler, L. E. Sohl, M. Bentzen, B. L. Otto-Bienser, et al. 2013. Sea surface temperature of the mid-Piacenzian ocean: a data-model comparison. Scientific Reports 3. doi:10.1038/srep02613.

Dowsett, H., M. Robinson, A. Haywood, U. Salzmann, D. Hill, L. Sohl, M. Chandler, M. Williams, K. Foley, and D. Stoll. 2010. The PRISM3D paleoenvironmental reconstruction. Stratigraphy 7:123–139.

Dynesius, M., and R. Jansson. 2000. Evolutionary consequences of changes in species’ geographical distributions driven by Milanko-
witch climate oscillations. Proceedings of the National Academy of Sciences of the USA 97:9115–9120.

Elías, S. E. 1984. Quaternary insects and their environments. Smithsonian Institution Press, Washington, DC.

Erwin, D. H. 2009. Climate as a driver of evolutionary change. Current Biology 19:R575–R583.

Erwin, D. H., and J. W. Valentine. 2013. The Cambrian explosion. Roberts, Greenwood Village, CO.

Fenton, I. S., P. N. Pearson, T. Dunkley Jones, A. Farnsworth, D. J. Lunt, P. Markwick, and A. Purvis. 2016. The impact of Cenozoic cooling on assemblage diversity in planktonic foraminifera. Philosophical Transactions of the Royal Society B 371:20150224.

Field, R., B. A. Hawkins, H. V. Cornell, D. J. Currie, J. A. F. Diniz-Filho, J.-F. Guégan, D. M. Kaufman, et al. 2009. Spatial species richness gradients across scales: a meta-analysis. Journal of Biogeography 36:132–147.

Fine, P. V. A. 2015. Ecological and evolutionary drivers of geographic variation in species diversity. Annual Review of Ecology, Evolution, and Systematics 46:369–392.

Foote, M. 2010. The geologic history of biodiversity. Pages 479–510 in M. A. Bell, D. J. Futuyma, W. F. Eanes, and J. S. Leviton, eds. Evolution since Darwin: the first 150 years. Sinauer, Sunderland, MA.

Gaston, K. G. 2000. Global patterns in biodiversity. Nature 405:220–227.

Goldberg, E. E., K. Roy, R. Lande, and D. Jablonski. 2005. Diversity, endemism, and age distributions in macroevolutionary sources and sinks. American Naturalist 165:623–633.

Hannisdal, B., K. A. Haaga, T. Reitan, D. Diego, and L. H. Liow. 2016. Common species link global ecosystems to climate change. bioRxiv. http://dx.doi.org/10.1101/043729.

Harmon, L. J., and S. Harrison. 2015. Species diversity is dynamic and unbounded at local and continental scales. American Naturalist 185:584–593.

Hawkins, B. A., C. M. McCain, T. J. Davies, L. B. Buckley, B. Anacker, H. V. Cornell, E. I. Damschen, et al. 2012. Different evolutionary histories underlie congruent species richness gradients of birds and mammals. Journal of Biogeography 39:825–841.

Hillebrand, H. 2004. On the generality of the latitudinal diversity gradient. American Naturalist 163:192–211.

Holman, J. A. 1995. Pleistocene amphibians and reptiles in North America. Oxford University Press, New York.

——. 1998. Pleistocene amphibians and reptiles in Britain and North Europe. Oxford University Press, New York.

Huang, S., K. Roy, J. W. Valentine, and D. Jablonski. 2015. Convergence, divergence, and parallelism in marine biodiversity trends: integrating present-day and fossil data. Proceedings of the National Academy of Sciences of the USA 112:4903–4908.

Hunt, G. 2010. Evolution in fossils lineages: paleontology and The Origin of Species. American Naturalist 176(suppl.):S61–S76.

Hunt, G., T. M. Cronin, and K. Roy. 2005. Species-energy relationship in the deep sea: a test using the Quaternary fossil record. Ecology Letters 8:739–747.

Jablonski, D. 1993. The tropics as a source of evolutionary novelty: the post-Palaeozoic fossil record of marine invertebrates. Nature 364:142–144.

——. 2005. Evolutionary innovations in the fossil record: the intersection of ecology, development and macroevolution. Journal of Experimental Zoology 304B:504–519.

——. 2008. Biotic interactions and macroevolution: extensions and mismatches across scales and levels. Evolution 62:715–739.

Jablonski, D., C. L. Belanger, S. K. Berke, S. Huang, A. Z. Krug, K. Roy, A. Tomosovych, and J. W. Valentine. 2013. Out of the tropics, but how? fossils, bridge species, and thermal ranges in the dynamics of the marine latitudinal diversity gradient. Proceedings of the National Academy of Sciences of the USA 110:10487–10494.

Jablonski, D., S. Huang, A. Z. Krug, K. Roy, and J. W. Valentine. 2017. Data from: Shaping the latitudinal diversity gradient: new perspectives from a synthesis of paleobiology and biogeography. American Naturalist, Dryad Digital Repository, http://dx.doi.org /10.5061/dryad.qd53c.

Jablonski, D., K. Roy, and J. W. Valentine. 2006. Out of the tropics: evolutionary dynamics of the latitudinal diversity gradient. Science 314:102–106.

Jackson, S. T., and J. L. Blois. 2015. Community ecology in a changing environment: perspectives from the Quaternary. Proceedings of the National Academy of Sciences of the USA 112:4915–4921.

Janssen, R. 2003. Global patterns in endemism explained by past climatic change. Philosophical Transactions of the Royal Society B 367:583–590.

Janssen, R., and M. Dynesius. 2002. The fate of clades in a world of recurrent climatic change: Milankovitch oscillations and evolution. Annual Review of Ecology and Systematics 33:741–777.

Jetz, W., C. Rahbek, and R. K. Colwell. 2004. The coincidence of rarity and richness and the potential signature of history in centres of endemism. Ecology Letters 7:1180–1191.

Johnson, K. G., W. Renema, B. R. Rosen, and N. Santodomingo. 2015. Old data for old questions: what can the historical collections really tell us about the Neogene origins of reef-coral diversity in the Coral Triangle? Palaeontologia Electronica 18:164–168.

Kennedy, J. D., Z. Wang, J. T. Weir, C. Rahbek, J. Fjeldså, and T. D. Price. 2014. Into and out of the tropics: the generation of the latitudinal gradient among New World passerine birds. Journal of Biogeography 41:1746–1757.

Kissling, W. C., Simpson, and M. Foote. 2010. Reefs as cradles of evolution and sources of biodiversity in the Phanerozoic. Science 327:196–198.

Kindlmann, P., I. Schoedlbauerova, and A. F. G. Dixon. 2007. Inverse latitudinal trends in species diversity. Pages 246–257 in D. Storch, P. A. Marquet, and J. H. Brown, eds. Scaling biodiversity. Cambridge University Press, Cambridge.

Krug, A. Z., and D. Jablonski. 2012. Long-term origination rates are reset only at mass extinctions. Geology 40:731–734.

Krug, A. Z., D. Jablonski, and J. W. Valentine. 2007. Contrarian clade confirms the ubiquity of spatial origination patterns in the production of latitudinal diversity gradients. Proceedings of the National Academy of Sciences of the USA 104:18129–18134.

——. 2009a. Signature of the end-Cretaceous mass extinction in the modern biota. Science 323:517–521.

Krug, A. Z., D. Jablonski, J. W. Valentine, and K. Roy. 2009b. Generation of Earth’s first-order biodiversity pattern. Astrobiology 9: 113–124.

Lees, D. C., A. Y. Kawahara, R. Rougerie, I. Ohshima, A. Kawakita, O. Bouteleux, J. De Prins, and C. Lopez-Vaamonde. 2014. DNA barcoding reveals a largely unknown fauna of Garcillariidae leaf-mining moths in the Neotropics. Molecular Ecology Resources 14:286–296.

Leprieur, F., P. Descombes, T. Gaboriau, P. F. Cowman, V. Paravicini, M. Kulbicki, C. J. Melian, et al. 2016. Plate tectonics drive tropical reef biodiversity dynamics. Nature Communications 7:11461.
Maddin, W. P., and R. G. FitzJohn. 2015. The unsolved challenge to phylogenetic correlation tests for categorical characters. *Systematic Biology* 64:127–136.

Maguire, K. C., D. Nieto-Lugilde, M. C. Fitzpatrick, J. W. Williams, and J. L. Blois. 2015. Modeling species and community variation to past, present, and future episodes of climatic and ecological change. *Annual Review of Ecology, Evolution, and Systematics* 46:343–368.

Mannion, P. D., P. Upchurch, R. B. J. Benson, and A. Goswami. 2016. Late Cenozoic on-figuration and the need to incorporate time-varying carrying capacities. *Philosophical Transactions of the Royal Society B* 371:20150217.

Martin, P. R., F. Bonier, and J. J. Tewksbury. 2007. Revisiting Jablonski (1993): cladogenesis and range expansion explain latitudinal variation in taxonomic richness. *Journal of Evolutionary Biology* 20:930–936.

Mayhew, P. J. 2007. Why are there so many insect species? perspectives from fossils and phylogenies. *Biological Reviews* 82:425–454.

Marcot, J. D., D. L. Fox, and S. R. Niebuhr. 2016. Late Cenozoic onset of the latitudinal diversity gradient of North American mammals. *Proceedings of the National Academy of Sciences of the USA* 113:7189–7194.

Marshall, C. R., and T. B. Quental. 2016. The uncertain role of diversity dependence in species diversification and the need to incorporate time-varying carrying capacities. *Philosophical Transactions of the Royal Society B* 371:20150217.

Martin, P. R., F. Bonier, and J. J. Tewksbury. 2007. Revisiting Jablonski (1993): cladogenesis and range expansion explain latitudinal variation in taxonomic richness. *Journal of Evolutionary Biology* 20:930–936.

Mayhew, P. J. 2007. Why are there so many insect species? perspectives from fossils and phylogenies. *Biological Reviews* 82:425–454.

Miller, A. I., and J. J. Sepkoski Jr. 1988. Modeling bivalve diversification: the effect of interaction on a macroevolutionary system. *Paleobiology* 14:364–369.

Mittelbach, G. G., D. Schemske, H. V. Cornell, A. P. Allen, J. M. Brown, M. Bush, S. P. Harrison, et al. 2007. Evolution and the latitudinal diversity gradient: speciation, extinction, and biogeography. *Ecology Letters* 10:315–331.

Mondal, S., and P. J. Harries. 2016. Phanerozoic trends in ecospace utilization: the bivalve perspective. *Earth-Science Reviews* 152:105–126.

Olson, D. M., E. Dinerstein, E. D. Wikramanayake, N. D. Burgess, G. V. N. Powell, E. C. Underwood, J. A. D’Amico, et al. 2001. Terrestrial ecoregions of the world: a new map of life on Earth. *Science* 287:583–592.

Payne, J. L., and M. E. Clapham. 2012. End-Permian mass extinction in the oceans: an ancient analog for the twenty-first century? *Annual Review of Earth and Planetary Sciences* 40:89–111.

Pellissier, L., F. Leprieur, V. Parravicini, P. F. Cowman, M. Kulbicki, G. Litsios, S. M. Olsen, M. S. Wisz, D. R. Bellwood, and D. Moullot. 2014. Quaternary coral reef refugia preserved fish diversity. *Science* 344:1016–1019.

Pianka, E. R. 1966. Latitudinal gradients in species diversity: a review of concepts. *American Naturalist* 100:33–46.

Pires, M. M., D. Silvestro, and T. B. Quental. 2015. Continental faunal exchange and the asymmetrical radiation of carnivores. *Proceedings of the Royal Society B* 282:20151952.

Powell, M. G. 2007. Latitudinal diversity gradients for brachiopod genera during late Paleozoic time: links between climate, biogeography and evolutionary rates. *Global Ecology and Biogeography* 16:519–528.

Precht, W. F., and R. B. Aronson. 2016. Faunal tracking and the stability of reef-coral assemblages in the Quaternary. Pages 257–289 in D. K. Hubbard, C. S. Rogers, J. H. Lipp, and G. D. Stanley Jr., eds. Coral reefs at the crossroads. Springer, New York.

Pulido-Santacruz, P., and J. Weir. 2016. Extinction as a driver of avian latitudinal diversity gradients. *Evolution* 70:860–872.

Pyron, R. A. 2014. Temperate extinction in squamate reptiles and the roots of latitudinal diversity gradients. *Global Ecology and Biogeography* 23:1126–1134.

Pyron, R. A., and F. T. Burbank. 2009. Can the tropical conservatism hypothesis explain temperate species richness patterns? an inverse latitudinal biodiversity gradient in the New World snake tribe Lampropeltini. *Global Ecology and Biogeography* 18:406–415.

Pyron, R. A., and J. J. Wiens. 2013. Large-scale phylogenetic analyses reveal the causes of high tropical amphibian diversity. *Proceedings of the Royal Society B* 280:20131622.

Querci, L., F. Leprieur, V. Parravicini, P. F. Cowman, M. Kulbicki, G. Litsios, S. M. Olsen, M. S. Wisz, D. R. Bellwood, and D. Moullo. 2014. Quaternary coral reef refugia preserved fish diversity. *Science* 344:1016–1019.

Rabosky, D. L. 2010. Extinction rates should not be estimated from molecular phylogenies. *Evolution* 64:1816–1824.

Rabosky, D. L., and E. E. Goldberg. 2015. Model inadequacy and mistaken inferences of trait-dependent speciation. *Systematic Biology* 64:340–355.

Rabosky, D. L., and A. H. Hurlbert. 2015. Species richness at continental scales is dominated by ecological limits. *American Naturalist* 185:572–583.

Rex, M. A., and R. J. Etter. 2010. Deep-sea biodiversity. *Harvard University Press*, Cambridge, MA.

Ricklefs, R. E. 2004. A comprehensive framework for global patterns in biodiversity. *Ecology Letters* 7:1–15.

———. 2007. History and diversity: explorations at the intersection of ecology and evolution. *American Naturalist* 170(suppl.):S56–S70.

Rohde, K. 1992. Latitudinal gradients in species diversity: the search for the primary cause. *Oikos* 65:514–527.

Rolland, J., F. L. Condamine, C. R. Beeravolu, F. Liguet, and H. Morlon. 2015. Dispersal is a major driver of the latitudinal diversity gradient of Carnivora. *Global Ecology and Biogeography* 24:1059–1071.

Rose, P. J., D. L. Fox, J. Marcot, and C. Badgley. 2011. Flat latitudinal gradient in Paleocene mammal richness suggests decoupling of climate and biodiversity. *Geology* 39:163–166.

Rosenzweig, M. L. 2001. The four questions: what does the introduction of exotic species do to diversity? *Evolutionary Ecology Research* 3:361–367.

Roy, K., and E. E. Goldberg. 2007. Origination, extinction and dispersal: integrative models for understanding present-day diversity gradients. *American Naturalist* 170(suppl.):S71–S85.

Roy, K., D. Jablonski, and J. W. Valentine. 2000. Dissecting latitudinal diversity gradients: functional groups and clades of marine bivalves. *Proceedings of the Royal Society B* 267:293–299.

———. 2001. Climate change, species range limits and body size in marine bivalves. *Ecology Letters* 4:366–370.

———. 2002. Body size and invasion success in marine bivalves. *Ecology Letters* 5:163–167.

Rohde, K. 1992. Latitudinal gradients in species diversity: the search for the primary cause. *Oikos* 65:514–527.

Rolland, J., F. L. Condamine, C. R. Beeravolu, F. Liguet, and H. Morlon. 2015. Dispersal is a major driver of the latitudinal diversity gradient of Carnivora. *Global Ecology and Biogeography* 24:1059–1071.

Rose, P. J., D. L. Fox, J. Marcot, and C. Badgley. 2011. Flat latitudinal gradient in Paleocene mammal richness suggests decoupling of climate and biodiversity. *Geology* 39:163–166.

Rosenzweig, M. L. 2001. The four questions: what does the introduction of exotic species do to diversity? *Evolutionary Ecology Research* 3:361–367.

Roy, K., and E. E. Goldberg. 2007. Origination, extinction and dispersal: integrative models for understanding present-day diversity gradients. *American Naturalist* 170(suppl.):S71–S85.

Roy, K., D. Jablonski, and J. W. Valentine. 2000. Dissecting latitudinal diversity gradients: functional groups and clades of marine bivalves. *Proceedings of the Royal Society B* 267:293–299.

———. 2001. Climate change, species range limits and body size in marine bivalves. *Ecology Letters* 4:366–370.

———. 2002. Body size and invasion success in marine bivalves. *Ecology Letters* 5:163–167.
Roy, K., D. Jablonski, J. W. Valentine, and G. Rosenberg. 1998. Marine latitudinal diversity gradients: tests of causal hypotheses. *Proceedings of the National Academy of Sciences of the USA* 95:3699–3702.

Sandel, B., L. Arge, B. Dalsgaard, R. G. Davies, K. J. Gaston, W. J. Sutherland, and J.-C. Svenning. 2011. The influence of Late Quaternary climate-change velocity on species endemism. *Science* 334:660–664.

Saupe, E. E., H. Qiao, J. R. Hendricks, R. W. Portell, S. J. Hunter, J. Soberón, and B. S. Lieberman. 2015. Niche breadth and geographic range size as determinants of species survival on geological time scales. *Global Ecology and Biogeography* 24:1159–1169.

Sax, D. F., and S. D. Gaines. 2008. Species invasions and extinction: the future of native biodiversity on islands. *Proceedings of the National Academy of Sciences of the USA* 105:11490–11497.

Schemske, D. W., G. G. Mittelbach, H. V. Cornell, J. M. Sobel, and K. Roy. 2009. Is there a latitudinal gradient in the importance of biotic interactions? *Annual Review of Ecology, Evolution, and Systematics* 40:245–269.

Schulte, D. 2016. Speciation, ecological opportunity, and latitude. *American Naturalist* 187:1–18.

Schnitzler, J., T. G. Barraclough, J. S. Boatwright, P. Goldblatt, J. C. Manning, M. P. Powell, T. Rebelo, et al. 2011. Causes of plant diversification in the Cape biodiversity hotspot of South Africa. *Systematic Biology* 60:343–357.

Spalding, M. D., H. E. Fox, G. R. Allen, N. Davidson, Z. A. Ferdana, M. Finlayson, B. S. Halpern, et al. 2007. Marine ecoregions of the world: a bioregionalization of coastal and shelf areas. *BioScience* 57:573–583.

Stebbins, G. L. 1974. Flowering plants: evolution above the species level. Belknap Press of Harvard University Press, Cambridge, MA.

Svenning, J.-C., W. L. Eisernhard, S. Normand, A. Ordóñez, and B. Sandel. 2015. The influence of paleoclimate on present-day patterns in biodiversity and ecosystems. *Annual Review of Ecology, Evolution, and Systematics* 46:551–572.

Taylor, M. P. 2008. Dinosaur diversity analysed by clade, age, place and year of description. Pages 134–138 in P. M. Barrett and S. E. Evans, eds. Ninth international symposium on Mesozoic terrestrial ecosystems and biota. Natural History Museum, London.

Tittensor, D. P., C. Mora, W. Jetz, H. K. Lotze, D. Ricard, E. Vanden Berghe, and B. Worm. 2010. Global patterns and predictors of marine biodiversity across taxa. *Nature* 466:1098–1101.

Todd, J. A., J. B. C. Jackson, K. G. Johnson, H. M. Fortunato, A. Heitz, M. Alvarez, and P. Jung. 2002. The ecology of extinction: Molluscan feeding and faunal turnover in the Caribbean Neogene. *Proceedings of the Royal Society B* 269:571–577.

Tomasovych, A., J. D. Kennedy, T. J. Betznr, N. Bitler Kuehne, S. Edie, S. Kim, K. Supriya, et al. 2016. Unifying latitudinal gradients in range size and richness across marine and terrestrial systems. *Proceedings of the Royal Society B* 283:20153027.

Valentine, J. W., and D. Jablonski. 2015. A twofold role for global energy gradients in marine biodiversity trends. *Journal of Biogeography* 42:997–1005.

Valentine, J. W., D. Jablonski, A. Z. Krug, and S. K. Berke. 2013. The sampling and estimation of marine paleodiversity patterns: implications of a Pliocene model. *Paleobiology* 39:1–20.

Valentine, J. W., D. Jablonski, A. Z. Krug, and K. Roy. 2008. Incumbency, diversity, and latitudinal gradients. *Paleobiology* 34:169–178.

Vermeij, G. J. 2005a. Invasion as expectation: a historical fact of life. Pages 315–339 in D. F. Sax, J. S. Stachowicz, and S. D. Gaines, eds. Species invasion: insights into ecology, evolution and biogeography. Sinauer, Sunderland.

———. 2005b. One-way traffic in the western Atlantic: causes and consequences of Miocene to early Pleistocene molluscan invasions in Florida and the Caribbean. *Paleobiology* 31:624–642.

Vilhena, D. A., and A. B. Smith. 2013. Spatial bias in the marine fossil record. *PLoS One* 8:e74470.

Walker, T. D., and J. W. Valentine. 1984. Equilibrium models of evolutionary species diversity and the number of empty niches. *American Naturalist* 124:887–899.

Wiens, J. J. 2011. The causes of species richness patterns across space, time, and clades and the role of "ecological limits." *Quarterly Review of Biology* 86:75–96.

Yasuhara, M., G. Hunt, T. M. Cronin, N. Hokanishi, H. Kawahata, A. Tsujimoto, and M. Ishitake. 2012. Climatic forcing of Quaternary deep-sea benthic communities in the North Pacific Ocean. *Paleobiology* 38:162–179.

Editor: Judith L. Bronstein

The common mussel (Mytilus edulis). From “Life Histories of the Mollusca” by A. S. Packard Jr. (The American Naturalist, 1875, 9:282–307).