How to characterise shared space use networks

Klara M. Wanelik¹* & Damien R. Farine²*

¹ Department of Evolution, Ecology and Behaviour, Institute of Infection, Veterinary and Ecological Sciences, University of Liverpool, Liverpool, UK.

² Department of Evolutionary Biology and Environmental Studies, University of Zurich, Zurich, Switzerland.

* Corresponding authors

Klara M. Wanelik

Current address: Department of Zoology, University of Oxford, Oxford, UK.

Email: klara.wanelik@zoo.ox.ac.uk

Damien R. Farine

Email: damien.farine@ieu.uzh.ch
Abstract

Studying the social behaviour of small or cryptic species often relies on constructing shared space use networks from sparse point-based observations of individuals (e.g. live trapping data). Such an approach assumes that individuals that have more observed space sharing events (e.g. detections in the same trapping location) will also have interacted more. However, there is very little guidance on how to construct shared space use networks, how much data are required for making such networks, or how to interpret the relationships they generate. In this study, we quantify the robustness of shared space use networks to different sampling regimes and network-generation algorithms. We first use empirical data to highlight that characteristics of how animals use space can help us to establish new ways to model the potential for individuals to co-occur. We then show that a method that explicitly models individuals’ home range and subsequent overlap in space among individuals (spatial overlap networks) requires fewer data for inferring observed networks that are correlated with the true shared space use network (relative to networks constructed from space sharing events). As a result, we show that shared space use networks based on estimating spatial overlap are also more powerful for detecting biological effects present in the true shared space use network. Finally, we discuss when it is appropriate to make inferences from shared space use about social interactions. Our study confirms the potential for using sparse trapping data from cryptic species to address a range of important questions in ecology and evolution.

Keywords

home range, point-based observations, social network, space-sharing, sparse observations, trapping data
Introduction

Social networks are central to addressing many of the key questions in ecology and evolution (Cantor et al. 2021). However, network construction remains a major challenge in many systems because large numbers of observations are needed to construct meaningful networks (Whitehead 2008a; Farine and Whitehead 2015). Recent technological improvements for collecting proximity, contact, or interaction data allow much more detailed networks to be constructed by improving the temporal resolution at which the data are collected (Douglas, Ji, & Clout, 2006; Rutz et al., 2012; Ryder, Horton, van den Tillaart, Morales, & Moore, 2012; Berkvens, Olivares, Mercelis, Kirkpatrick, & Weyn, 2019). However, for many smaller or more cryptic species, where observation remains difficult, most studies still rely on collecting data by trapping individuals and modelling social contacts as observations of different individuals in the same trap across different trapping events (e.g. Perkins, Ferrari, & Hudson, 2008; Porphyre, Stevenson, Jackson, & McKenzie, 2008; Grear, Perkins, & Hudson, 2009; Grear, Luong, & Hudson, 2013; VanderWaal, Atwill, Hooper, Buckle, & McCowan, 2013; Davis et al., 2014). To date, no study has quantified whether the sparse observations typical of such studies allow us to construct networks that represent the real underlying patterns of shared space use (an issue highlighted in relation to disease transmission; Tompkins, Dunn, Smith, & Telfer, 2011; White, Forester, & Craft, 2017) and, therefore, whether we can extract meaningful biological relationships from such shared space use networks.

Shared space use networks can be constructed from a range of different data-collection traps, most commonly live-capture traps and camera traps, but also increasingly using RFID detections (Sabol et al. 2018). These methods are characterised by typically detecting single individuals at any one time, but having the capability to observe multiple individuals in the
same location(s) over time. Individuals that are then observed (trapped) at the same location are considered to be connected, with binary edges (there or not) between them, or, if constructing a weighted network, with the number of space sharing events (e.g. trapping locations) defining the strength of their connection. However, the definition of these edges, and therefore the interpretation of the resulting shared space use network, can be unclear. For example, both VanderWaal et al. (2013) and Grear, Luong & Hudson (2013) use networks generated from such observed space sharing events to represent both shared space use and the likelihood of social interaction. The utility of these networks in representing social interactions comes from the assumption that two animals more often observed in the same locations are more likely to come into direct contact. The validity of this assumption will depend on the biology of the system (Farine 2015).

Beyond the interpretation of the meaning of network connections, there has been no direct quantification of the robustness of shared space use networks to different sampling regimes. It has been suggested that the data-intensive nature of networks may act as a barrier to the more widespread use of networks in the fields of ecology and evolution, with wildlife systems often being data limited (Craft and Caillaud 2011). Recent investigations into networks based on co-occurrence data (Farine and Strandburg-Peshkin 2015; Hart et al. 2021) and direct observation methods (Davis, Crofoot, & Farine, 2018) have highlighted how networks are data-hungry. Constructing a meaningful network requires sufficient observations to accurately estimate each of the many relationships (both present and absent) that connect all individuals in a population (specifically: \( \frac{n(n-1)}{2} \) edges in an undirected network). Thus, the sampling intensity needed to maintain a minimum number of observations per individuals and, critically, the co-observations of pairs of individuals (dyads) grows quadratically with the number of individuals represented in a given network.
Previous studies quantifying the data required to construct meaningful social networks have suggested that a good rule-of-thumb is that an average of 15 potential co-observations between all pairs of individuals are needed (i.e. of potential associations or interactions, which could be both individuals together or just one individual in the absence of the other; Farine & Strandburg-Peshkin, 2015; Davis et al., 2018). Importantly, more observations are required for accurately defining network structure when the differences in the relationship among dyads are more uniform through a population (Whitehead 2008a, b; Hart et al. 2021), such as we expect in less social species. However, these estimates of effort may not translate well to quantifying shared space use networks, because estimating shared space use could require much fewer data than estimating relationship strengths, as space use is a property that can be characterised at the level of individuals. Further, the resulting networks do not aim to characterise relationship strengths but rather a broader likelihood of encounter. Addressing the question of how much data need to be collected is also crucially important because many studies that have constructed shared space use networks do not report the mean number of observations per individual (Webber and Vander Wal 2019). Furthermore, the majority of those that do have been based on relatively few observations per individual. For example, VanderWaal (2013) had a mean of 11 trapping events per individual, meaning that they would have a mean of 22 potential observations from which to characterise dyadic edge weights in their network (observing two individuals apart 11 times each would give the denominator of an association index—e.g. the proportion of time individuals were associated—of 22, see Hoppitt & Farine, 2018). Understanding how the number of trapping events relates to the robustness of network estimates remains a major gap in knowledge.
The choice of which data to include when constructing edges in a shared space use network can also have an impact on how meaningful the resulting network is. Many studies have defined a temporal threshold within which the observation of two individuals in the same place must occur for these observations to be counted as a connection (i.e. a space sharing event). These are often inspired by biology, commonly the lifetime of a disease vector or pathogen in studies using networks to characterise disease transmission. For example, Porphyre et al. (2008) used 28 days (maximum survival of *Mycobacterium bovis* in the environment) and Perkins, Ferrari & Hudson (2008) used 14 days (the time needed for an infective L3 larval stage of *Heligmosomoides polygyrus* to develop from the eggs of an infected host). However, such definitions can be at odds with the definition and biological motivation behind applying a network approach. For studies that rarely observe individuals, a shared space use network is most powerful when used as a proxy defining the potential for individuals to co-occur (and possibly encounter one-another) anywhere within their respective home ranges, rather than whether they actually did encounter one-another in any particular location. In other words, shared space use networks model potential contacts, rather than estimating any actual contacts. This definition contrasts with studies that can regularly observe or recapture individuals (such as Smith et al., 2018 who used PIT-tag readers at the entrance of burrows) and can directly relate observed space sharing events (e.g. within a burrow on a given visit) at the same spatial and temporal scales as social behaviours or disease transmission events that form part of the study of interest (see Farine, 2018 for further discussion). Restricting the temporal scale for defining connections also reduces the data available from which the strength of connections between individuals can be estimated. Using observations spaced more widely apart in time forms connections between individuals that describe the system more generally as opposed to quantifying with precision the
connections among specific individuals (although these can correlate, e.g. Sabol, Solomon, & Dantzer, 2018).

In this study, we conduct a quantitative evaluation of the robustness of networks to different sampling regimes and network definitions to provide guidance on how to construct shared space use networks. We first use empirical data to highlight that characteristics of how animals use space can help us to establish new ways to model the potential for individuals to co-occur (and potentially encounter one-another). We then describe a new method for characterising shared space use network that more generally estimates home range overlap. We show that using this method can generate a network which generally (1) is more strongly correlated with the true shared space use network; (2) is a more accurate representation of the true space sharing network and, therefore; (3) has greater power to detect biological effects present in the true shared space network, relative to networks generated from the observed space sharing events only. Importantly, the spatial overlap method requires many fewer observations than using space sharing events to reconstruct meaningful shared space use networks, and many fewer observations than what has been suggested in the more general guidelines for social networks (e.g. 15 observations per dyad, see above). Finally, we discuss the topic of inference from shared space use networks, and how appropriate it is to link network data with biological processes.

Materials & Methods

Our study consists of three core components. First, we use a large empirical dataset to highlight characteristics of how animals use space, specifically that they have a core and a periphery to their home range. Second, we estimate the ability for data on space sharing
events by individuals to generate networks that are robust to different sampling regimes using simulated data. Third, we describe a new method, inspired by the core-peripheral nature of animal home ranges, for defining network edges. We use the same simulated data to show that this method generates observed networks which are, for a given number of captures per individual, more strongly correlated with the true shared space use network, are a more accurate representation of the true shared space use network, and are more powerful at detecting biological effects present in the true shared space use networks (i.e. detect a positive correlation present in the network from which a given simulated dataset is based) relative to networks generated from observed space sharing events only. All simulations were run in R version 3.3.1 (R Core Team 2016) using the packages vegan version 2.4-3 (Oksanen and et al 2017) and sna version 2.4 (Butts 2016).

**Modelling home ranges in an empirical dataset**

There is a large body of literature on how best to model animal space use, and it is widely accepted that many animals have a core and a periphery to their home range (Hayne 1949, 1950; Calhoun and Casby 1958; Jennrich and Turner 1969; Schoener 1981; Swihart and Slade 1989; Spencer et al. 1990; Slade and Russell 1998; Zamora and Moreno-Amich 2002; Klein and Cameron 2012). We test whether this holds true in a large-scale empirical dataset for a population of field voles (*Microtus agrestis*), and in so doing, the utility of this simple representation of home range for modelling the space-use behaviour of large numbers of animals.

We use part of a dataset from a study of *M.* *agrestis* in Kielder Forest, UK (55˚13’ N, 2˚3’ W) that involved capturing individuals using live-trapping methods. Access to the study site was provided by the Forestry Commission. Full details are given in Jackson et al. (2014). The
site was monitored across two years (2009-2010) by monthly trapping sessions between February and November, and contained a live-trapping grid (0.375 ha) of 150 (10 × 15) regularly spaced traps (3–5 m intervals) placed in optimal habitat. Animals were marked with passive radio frequency transponders (AVID plc, East Sussex, UK) and monitored over time, thus providing sequences of capture and recaptures. This dataset is comprised of 347 individuals and 678 trapping events.

*M. agrestis* is a polygynous species, with strictly territorial males. Home ranges of *M. agrestis* vary across different locations, habitats and across different times of the year. A review of nine studies, all conducted in later summer, but across a range of locations and habitats and using a range of different home range estimation methods, found that female home ranges varied in size from 30 to 900 m² while males home ranges varied in size from 200 to 1500 m² (Borowski 2003). Large males also have the largest home ranges (Borowski 2003). In our own study population, there is evidence for differences in the degree to which large males, small males and females are discouraged by distance (Davis et al., 2014). We estimate home range parameters for females and large males (mean weight ≥ 25 g) as an example, which we go on to use in our simulations (see *Simulation procedure*).

*Simulation procedure*

In brief, our study used the following procedure (see Fig. 1):

1. We simulated a set of 100 individuals with home ranges defined by a centroid and characterized by a negative sigmoidal curve that highlights the declining probability $P$ of an individual to be detected at an increasing distance ($d$) away from the centroid of its home range:
$P(d) = \frac{1}{1+e^{-a-bd}}$  \hspace{1cm} (1)

where, $a$ describes the overall size of the home range, $b$ describes the steepness of the edge of the home range, and $d$ is the logarithmic distance from the centroid. Our choice of negative sigmoidal curve was inspired by the core-peripheral nature of animal home ranges, and a large-scale empirical dataset for a population of field voles ($M. agrestis$), but in the discussion we highlight that different home range shapes can be used. We defined the true shared space use network as the amount of overlap in the home range profiles across all combinations of individuals (see detailed methods below).

(2) We randomly placed simulated individuals in a spatial area containing $T$ traps laid out in a stratified grid. We then simulated observation datasets that contained detections of individuals at traps, where the detection probability for a given individual in a given trap was determined by the position of the trap relative to the home range profile of the individual defined in Equation 1 (higher closer to the centroid, lower further away from the centroid).

(3) From the simulated observation datasets, we first constructed a network based on observed space sharing events only.

(4) Finally, we applied a novel method to construct an overlap network, based on estimating individual home ranges and estimating home range overlap among individuals.

Below we describe steps 1-4 in more detail:

1. **Simulating true networks**

We first drew $N$ sets of $x$ and $y$ coordinates from a uniform distribution, where the boundaries of the distribution correspond to the edges of our study area (in our case, from 0 to 10 in each
For each individual, we also randomly allocated a sex (male or female) and drew home range parameters \((a\) and \(b\) in Equation 1) based on the sex, giving males a larger home range than females. Home range parameters for males and females were based on the empirical data (see Modelling home ranges in an empirical dataset), with added noise drawn from a normal distribution with standard deviation equal to 0.05 times the home range parameter in question \((a\) or \(b\)) to simulate individual-level variation in home range profile.

For each simulation, we generated a true shared space use network, with edge weights representing the amount of overlap in the home range between each pair of individuals. This was done numerically by overlaying the two individuals’ 2D home range profiles and calculating the area under the two surfaces (Fig. 1a). Specifically, we predicted the probability of detecting each individual in grid overlapping both their home ranges, using equation (1), and calculated the overlap by dividing the sum of the lowest values at each point on the grid \((x, y)\) by the sum of the largest value at each point, according to the following equation:

\[
E_{1,2} = \frac{\sum_{x=-l}^{m} \sum_{y=-l}^{n} \min(P_1(\sqrt{(x_1-x)^2+(y_1-y)^2}), P_2(\sqrt{(x_2-x)^2+(y_2-y)^2}))}{\sum_{x=-l}^{m} \sum_{y=-l}^{n} \max(P_1(\sqrt{(x_1-x)^2+(y_1-y)^2}), P_2(\sqrt{(x_2-x)^2+(y_2-y)^2}))}
\]  

(2)

where \(P_n(\sqrt{(x_n-x)^2+(y_n-y)^2})\) is the probability of observing individual \(n\) at location \((x, y)\) from Equation 1, and \((m, n, l)\) are coordinates capturing an area overlapping both home ranges. In our simulations we first moved both individuals’ coordinates such that the centre of one individuals home range was at \((x = 0, y = 0)\), and defined \(m\) as \(-2\), \(n\) as \(x_2+2\), and \(l\) as \(y_2+2\).
2. Simulating observations of individuals in traps

We first calculated the probability for a given individual to be observed in a given trap. We defined this probability based on the distance of the trap to the centre of the individual’s home range using Equation 1. We repeatedly did this for all combinations of individuals and traps (‘Trapping Probability’ in Fig. 1b). We then used these probabilities to simulate observations by drawing from a binomial distribution \{0,1\} (‘Simulated Detections’ in Fig. 1b). We incremented the number of draws from this sampling process to generate more observations. Because draws resulted in variable numbers of observations, we then calculate the mean number of observations per individual, allowing us to make our results more easily interpretable.

3. Generating shared trap networks

Each simulated dataset contained the number of detections of each individual in each trap. We generated a shared trap network for each simulated dataset with the edge weight between individual 1 and individual 2 \(E_{1,2}\) defined as:

\[
E_{1,2} = \frac{|\text{traps}_1 \cap \text{traps}_2|}{|\text{traps}_1 \cup \text{traps}_2|}
\]  

(3)

where \(\text{traps}_1 \cap \text{traps}_2\) is the set of traps in which both individuals were detected, and \(\text{traps}_1 \cup \text{traps}_2\) is the set of traps in which either or both individuals were detected (Fig. 1c).

4. Generating networks based on overlapping home ranges

We then applied a novel method for generating shared space use networks based on first estimating a population’s home range profile(s) from a simulated observation dataset, and
then calculating the overlap in the observed home range profiles of each pair of individuals based on the distance between their observed centroids. Our method operates as follows. First, we calculate each individual’s observed centroid by taking the mean of the positions it was observed in weighted by the number of observations at each of these positions. Second, we calculate the distance between this centroid and all of the traps where it could have been captured. Third, we calculate the observed home range profile for individuals, which (for a representative, sparse dataset) we achieve by fitting a negative sigmoidal curve (Equation 1; fitted using a Bernoulli GLM, with 0 indicating an individual was not detected at a particular trap, and 1 indicating an individual was detected at a particular trap) for males and females separately, thereby generating a relationship representing the average home range profile for each sex (see discussion for justification for this strategy of combining individuals, as well as alternative strategies). Fourth, we use the observed profiles calculated for each individual to estimate overlap in space use between each pair of individuals using Equation 2 (Fig. 1d).

Estimating the robustness of shared space use networks to different sampling regimes

The ultimate aim of any network we construct is to be able to reliably test a hypothesis of interest. To test the performance of our novel spatial overlap network method against traditional shared trap networks, we generated 1000 true networks (Fig. 1a). For each true network, we produced simulated observation datasets that varied in sampling intensity (number of draws from a binomial distribution \( \{0,1\} \) given a probability of observing an individual in a trap; Fig. 1b). We designed this such that the sampling intensity corresponded to a mean number of observations per individual ranging between 1 and 50 (regardless of trapping grid density), thus capturing the spectrum of what has been reported in the literature. For each simulated observation dataset, we generated a shared trap network, and a network
using the overlap approach by reconstructing separate negative sigmoidal curves (or home range profiles) for large males and females (Fig. 1c).

We assessed the performance of each of these observed networks for three metrics. First, we calculated the correlation between the edge weights in the observed network and the edge weights in the true network using a Mantel test. The correlation provides a measure of relative position of each edge, such that when the correlation is 1 the position of each edge from the observed network is the same as the positions from the true network, irrespective of any changes in scale. Second, we calculated a measure of accuracy by taking the sum of the absolute differences between the observed and true network edge weights. The accuracy provides a measure of whether the estimated edges in the observed network are on the same scale as those in the true network. Third, we calculated a measure of power by finding the proportion of observed networks in which we could detect a significant biological effect—here the difference in mean degree (sum of edge weights) between large males and females (large males were given a larger home range than females, see point 1 of the simulation procedure)—that is present in the true network. We estimated significance for each simulated observation dataset by comparing the observed difference in mean degree between large males and females to the distribution of differences in 100 permuted networks. We used node permutations, which involved randomising the assignment of sex to the identities of each individual. We deemed the effect from an observed network to be significant if fewer than three of the randomised networks generated a difference that was larger than the observed one (two-tailed test at \( p = 0.05 \), see Farine, 2017).

_Variants_
We repeated the procedure described above for true networks with varying effect sizes for the difference in mean degree (sum of edge weights) between large males and females by varying the $b$ parameter, resulting in: (a) an effect size half that in our empirical dataset, (b) an effect size equal to that in our empirical dataset, (c) an effect size twice that in our empirical dataset (see Results). We also repeated the procedure using trapping grids of differing densities: (a) a $10 \times 10$ grid, and (b) a $19 \times 19$ grid within the same area.

**Results**

*Modelling home ranges in an empirical dataset*

Consistent with space-use theory, we found evidence for a declining probability of an individual field vole to use space further away from the centre of its home range. Furthermore, we characterised this empirical relationship, between probability of detection and distance from centroid, using a negative sigmoidal curve (Equation 1; fitted using a Bernoulli GLM, with 0 indicating an individual was not detected at a particular trap, and 1 indicating an individual was detected at a particular trap). We found evidence for large males having a larger home range ($a = 2.08, b = -4.82$) than females ($a = 2.83, b = -6.21$; Fig. 2), resulting in a difference in mean degree of 1.8 between these two classes. We used these class-specific curves, and the resulting difference in mean degree, to generate true shared space use networks in our simulations.

*Performance of simulated observed networks with varying number of captures per individual*

The number of individuals detected at least once at a trap increases with the number of captures per individual, starting from a mean of 31.2 individuals (out of a total population of 100 individuals) present at a mean of 1.3 captures per individual, and reaching a mean of
100.0 individuals (i.e. the whole population) present at approximately 10 captures per individual (Fig. 3c).

**Correlation**

As the mean captures per individual increases, the shared trap network becomes more strongly correlated with the true network. At a mean of 1.9 captures per individual, the Mantel correlation coefficient between the shared trap network and the true network is 0.4. The correlation coefficient plateaus from a mean of approximately 20 captures per individual, reaching a maximum of 0.8 at a mean of 28.7 captures per individual. The overlap network shows broadly the same pattern, but is, for a given number of captures per individual, typically more strongly correlated with the true network than the shared trap network. At a mean of 1.9 captures per individual, the correlation coefficient between the overlap network and the true network is 0.5. The correlation coefficient also plateaus earlier, from a mean of approximately 10 captures per individual, and reaches a higher maximum of 1.0 (Fig. 3a).

**Accuracy**

As the mean number of captures per individual increases, the shared trap network becomes more accurate. At a mean of 1.9 captures per individual, the sum of the absolute differences in edge weights between the true space use and shared trap networks is 630.8, which reaches a minimum of 386.1 at a mean of 28.7 captures per individual. The overlap network shows broadly the same pattern, but is more accurate for a given number of captures per individual. For example, at a mean of 1.9 captures per individual, the absolute difference in edge weights to the true network is 609.1. The sum of differences for the overlap network also reaches a lower minimum of 224.0 (Fig. 3b).
As the mean captures per individual increases, the ability to detect a true biological relationship (i.e. the power) of the shared trap network also increases. For example, there is nearly double the chance of detecting a true positive at a mean of 4.3 captures (5.2%) compared to 1.9 captures (3.2%). However, the power remains low for small effect sizes even after large numbers of captures. The overlap network shows broadly the same pattern, but has consistently greater power to detect an effect for a given number of captures per individual, above a mean of approximately 3 captures per individual. For example, at 4.3 captures per individual there is more than double the chance of detecting a true positive in the overlap network (11.1%) compared to the shared trap network (5.2%). The power of the shared trap network increases continuously and reaches a maximum power of 35.2% at a mean of 28.7 captures per individual. The power of the overlap network plateaus at a mean of approximately 10 captures per individual, reaching a much higher maximum of 82.0% (Fig. 4a). Below 3 captures per individual, the shared and overlap networks have similar power to detect an effect.

**Performance of observed networks with varying effect sizes**

Only the power of the observed networks changes as a result of varying effect size.

**Power**

As the effect size increases, there is a corresponding increase in the power of the shared trap network, above a mean of approximately 3 captures per individual. For example, at a mean of 4.3 captures per individual, there is a 2.3% chance of detecting a true positive if the effect size is half that found in our empirical data, 5.2% chance is the effect size is equivalent to that found in our empirical data, and a 24.5% chance if the effect size is twice that found in
our empirical data. The overlap network shows broadly the same pattern. At a mean of 4.3 captures per individuals, there is a 7.3% chance of detecting a true positive if the effect size is half that found in our empirical data, 11.1% chance if the effect size is equivalent to that found in our empirical data, and 28.8% chance if the effect size is twice that found in our empirical data. Below approximately 3 captures per individual, shared and overlap networks have similar power, regardless of effect size.

**Changing trapping grid density**

Observed networks change in all three metrics (correlation, accuracy and power) as a result of varying trapping grid density. Grid density also changes the number of individuals present in both observed networks when the number of captures per individual is low. For example at 1–2 captures per individual, on a 19 × 19 grid, a mean of 77.1 individuals (out of a total population of 100 individuals) are present in the observed networks (compared to 31.2 on a 10 × 10 grid; see above). However, all individuals in the population are present in the observed networks (i.e. mean of 100.0 individuals) from a mean of approximately 10 captures per individual, regardless of grid density (Fig. 5c).

**Correlation**

A higher density grid leads to a weaker correlation between the shared trap network and the true network, for a given number of captures per individual (Fig. 5a). For example, at a mean of approximately 2 captures per individual, the Mantel correlation coefficient between the shared trap network and the true network is 0.4 on a 10 × 10 grid, and 0.2 on a 19 × 19 grid. The shared trap network reaches the same maximum correlation coefficient of 0.8 on a 19 × 19 grid and on a 10 × 10 grid (see above). The correlation between the overlap network and true network differs very little between the 10 × 10 and 19 × 19 grid (Fig. 5a). At a mean of
approximately 2 captures per individual, the correlation coefficient between the overlap network and the true network is 0.5 on a $10 \times 10$ grid, and 0.6 on a $19 \times 19$ grid. The overlap network also reaches the same maximum correlation coefficient of 1.0 on a $19 \times 19$ grid and on a $10 \times 10$ grid (see above).

**Accuracy**

A higher density grid leads to a slightly less accurate shared trap network for a given number of captures per individual (Fig. 5b). For example, at a mean of approximately 2 captures per individual, the absolute difference in edge weights to the true network is 630.8 on a $10 \times 10$ grid, and 644.1 on a $19 \times 19$ grid. The minimum absolute difference in edge weights for the shared trap network is also higher on a $19 \times 19$ grid (434.8) compared to a $10 \times 10$ grid (386.1; see above). The overlap network shows the opposite pattern i.e. a higher density grid leads to a slightly more accurate overlap network for a given number of captures per individual (Fig. 5b). For example, at a mean of approximately 2 captures per individual, the absolute difference in edge weights of the overlap network to the true network is 609.1 on a $10 \times 10$ grid, and 569.9 on a $19 \times 19$ grid. The minimum absolute difference in edge weights for the overlap network is also lower on a $19 \times 19$ grid (211.8) compared to a $10 \times 10$ grid (224.0; see above).

**Power**

A higher density grid leads to a slightly less powerful shared trap network for a given number of captures per individual (Fig. 5d). For example, at a mean of approximately 2 captures per individual, the chance of detecting a true positive is 3.2% on a $10 \times 10$ grid, and 2.5% on a $19 \times 19$ grid. The power of the overlap network does not change with grid density (Fig. 5d). At a
mean of approximately 2 captures per individual, the chance of detecting a true positive is 3.1% on a 10 × 10 grid, and on a 19 × 19 grid.

**Discussion**

In this study, we quantify the robustness of shared space use networks to different sampling regimes. In doing so, we provide much needed guidance for informing the choice of sampling regime when designing studies to accurately quantify space sharing among individual animals. Using a large-scale empirical dataset for a population of field voles (*M. agrestis*), we also demonstrate the utility of modelling space-use behaviour on the basis that individuals have a core and a periphery to their home range. Finally, we use these insights to develop a new method for generating shared space use networks based on estimating overlapping home ranges. We show that networks generated using the overlap method are generally more strongly correlated with the true shared space use network, are a more accurate representation of the true shared space use network, and are more powerful to detect effects present in the true shared space use network relative to networks generated from observed space sharing events only.

Our overlap method works particularly well when the mean number of captures per individual is low and provides the potential to generate meaningful networks even from sparse point-based observations of individuals. While standard methods are restrictive, by relying only on joint observations at a trap, and sometimes imposing a temporal threshold within which the observation of two individuals in the same place must occur, our method pools data among individuals to arrive at a more general estimate of home range profile. In doing so, our method accounts for imperfect and heterogeneous observations (as in e.g.
Gimenez et al., 2019). Using these general profiles, we then calculate the extent of two individuals’ home range overlap, as a function of their observed centroids, to estimate their overlap in space. Our simulation results confirm that this approach results in more accurate and more representative networks than existing methods. We also model differences in home range profile between large males and females, based on our empirical data. However, classes of individuals which differ in their space use will vary between systems, and prior knowledge will be necessary to identify these classes e.g. males and females, larger and smaller individuals or younger and older individuals (Wolton and Flowerdew 1985; Mikesic and Drickamer 1992; Dahle and Swenson 2003; Godsall et al. 2014). Our results further show that an increase in correlation and accuracy of the overlap method translates to greater power at extracting biological effects present in the true shared space use network. In our case, we modelled differences in mean degree (sum of edge weights) between large males and females, but these outcomes should be generalisable to other hypotheses.

An important, and perhaps unexpected, finding is that denser grids of data-collection traps can make networks based on space sharing events (at least when the mean number of captures per individuals is below 30) less strongly correlated with the true network, less accurate and less powerful. This result makes sense when considering sampling stochasticity. If there are more available traps, then it is less likely that two individuals, which overlap in space, will be trapped in exactly the same trap (unless the number of captures is very high and in no way limiting). Subsequently, constructing networks from occurrences at the same trap reduces the numerator of the edge weight calculation in Equation 3 (the set of traps in which both individuals were detected) and increases the denominator (the set of traps in which at least one individual was detected). By contrast, we show that, in terms of correlation, accuracy and power, our overlap method performs equally well, or better, on a
denser trapping grid. This is because finer-scaled grids provide better estimations of individuals’ space use.

It is important to note that networks generated using the overlap method do not always perform better than those based on observed space sharing events only. We show here that a shared trap network is more accurate and more powerful at detecting biological effects present in the true network when the effect size is low and the number of captures per individual is high. In other words, if many space sharing events are observed, then a network based on these alone deviates less from the true network and is more likely to detect a subtle biological effect present in the true network, relative to the process of pooling data and generating population-wide home range profiles. This pattern is likely to be driven by noise in the process of estimating the centroids of individuals, which can be susceptible to outlying observations. Thus, studies that use methods, such as RFID detections, that produce substantially larger datasets than trapping does (e.g. Sabol et al., 2018) should model the sampling process to determine the most powerful approach for a given effect strength.

We further note that the performance of shared trap networks may also depend on the biological system, or the ecological conditions it experiences (as in Perkins, Cagnacci, Stradiotto, Arnoldi, & Hudson, 2009). Our study is based on empirical data on *M. agrestis* sampled during the breeding season. During this time field voles maintain relatively fixed home ranges which can be estimated with some certainty (Myllymaki 1977; Niethammer and Krapp 1982). However, this is likely to be problematic if individuals are highly mobile, resulting in constantly shifting home ranges. We would expect a shared trap network to be better suited to such a dynamic system. Our study population also inhabits a relatively homogenous landscape, in the form of grassy clear-cuts within a coniferous forest. As a
result, individuals are not expected to vary a great deal in the size of their home range. However, landscape features, such as hills, in a more heterogeneous landscape could result in more variability in home range size among individuals, making it difficult to quantify an ‘average’ home range. A shared trap network could again be better suited to such a system. However, if observations of individuals are sufficiently numerous (Noonan et al. 2019), individual differences in home range profile could be accounted for when using the overlap method. This could be done, for example, by fitting a random effect for individual within class-specific regressions. Ultimately, the strongest approach (and the aim of generating shared space use networks) is to estimate a precise home range for each individual in the population. These individual home range profiles could vary in size (by changing the $a$ and $b$ parameters of the negative sigmoidal curve) and/or in shape (by replacing the negative sigmoidal curve with a different function or an explicit home range model, e.g. Fleming & Calabrese, 2017). Finally, given sufficient data, useful tools exist for estimating home range overlap among individuals (Winner et al. 2018).

Shared space use networks are, and will continue to be, widely used to shed light on various biological processes. For example, individuals who share more space may be more likely to compete for resources. Many parasites and pathogens are also transmitted through the environment, and so knowing who shares space with whom, can tell us something about who is likely to transmit infection to whom (VanderWaal et al. 2013). It is also true that shared space use, or proximity, is a prerequisite for interaction (Farine 2015), but whether or not individuals that share space do indeed associate, or interact, and thus how far point-based observations can be used to draw meaningful inferences, will depend on the biology of the system. Behaviour in particular is important to consider, with some animals actively avoiding each other (Davis et al., 2014) and others actively seeking each other out (Raulo et al. 2021).
It is therefore important to take care when making biological inferences from any network data.

One point we highlight in our study is that the process of network generation makes explicit assumptions about the biological processes being modelled. Calculating space sharing events, for example the presence of two individuals in the same location within a given pathogen transmission period (defined by the lifetime of the pathogen in the environment, or the time taken for a pathogen to develop into an infective stage) produces networks aimed at estimating actual transmission events. When observation data are sparse, these observed events are likely to represent only a small proportion of all events that took place, and thus the power of the network to detect biological effects is low. This could explain why observed transmission networks are not always robust estimates of transmission processes (Wohlfiel et al. 2013). By contrast, modelling the overlapping space use among individual captures the relative probability of transmission events taking place among all the dyads in a population, which will include both observed and unobserved events. Our simulations confirm that defining networks in this way can produce networks that are more powerful at detecting biological effects, especially when observations are sparse. We use pathogen transmission as an example to illustrate our point, but this should be generalisable to other questions.

Our method provides a novel opportunity to generate meaningful shared space use networks, and if appropriate, to make inferences from shared space use about social interactions, even from sparse point-based observations of individuals. It therefore unlocks the potential of these data, still the most common form of data available for many smaller or more cryptic species, to address a range of key questions in ecology and evolution.
Figures

Fig. 1 Schematic showing the simulation process: (a) Simulation of the true shared space use network with edge weights between two individuals (e.g. Individual 1 and Individual 2) equal to the overlap between their respective home ranges (here depicted along a one-dimensional slice). Each home range modelled using a negative sigmoidal curve with class-specific parameters \((a\) and \(b\); Equation 1) that captures the decreasing probability of observing individuals as the distance away from the centroid increases, and the overlap between the two-dimensional surfaces produced by the negative sigmoidal curves being calculated using Equation 2. (b) Generating a simulated observation dataset by calculating the probability for a given individual to be observed in a given trap based on its home range profile (crosses represent centroids; circles represent trapping probabilities; the bigger the circle, the greater the probability of detecting an individual at a trap), then simulating observations by drawing from a binomial distribution \(\{0,1\}\) with the probability of getting a 1 for a given individual in a given trap defined by this trapping probability. (c) Generating a shared trap network, where nodes represent individuals, and where edge weights are calculated using Equation 3. (d) Generating an overlap network, where nodes represent individuals, and where edge weights represent the overlap between two individuals’ observed home range profiles. First, the observed centroid was calculated for each individual. Then we modelled class-specific home ranges using a negative sigmoidal curve (using a GLM regression). Third, we used Equation 2 to calculate home range overlaps as an estimate of shared space use (i.e. the edge weights), as in (a) but with the observed home range profiles and centroid values.
Fig. 2 Class-specific negative sigmoidal curves for *M. agrestis* describing the change in probability of detection with increasing distance from the centre of an individual’s home range. Line shows the fitted home range profile for large males and females. Points show the raw data (whether, 1, or not, 0, an individual was detected at a location). Distances are measured in trapping grid cells (1 grid cell = 3–5 m)
**Fig. 3** Performance of observed networks with varying number of captures per individual on a 10 × 10 trapping grid, as measured by (a) Correlation: Mantel correlation between edge weights in observed and true networks, (b) Accuracy: Absolute difference in edge weights between observed and true networks (lower values = more accurate networks), and (c) Number of individuals in observed networks. LOESS smoother added to aid visual interpretation. Panel (c) refers to the data in the simulated observation dataset, which is identical for both methods.
**Fig. 4** Performance of observed networks with varying number of captures per individual on a $10 \times 10$ trapping grid, as measured by the power of observed networks to detect a biological effect present in the true network. Proportion of true positives shown on y-axis, and mean of mean captures per individual shown on x-axis. Repeated for true networks with varying effect sizes (a) an effect size half that in our empirical dataset, (b) an effect size equal to that in our empirical dataset, (c) an effect size twice that in our empirical dataset.
Fig. 5 Performance of observed networks with varying numbers of captures per individual on a 19 × 19 trapping grid, as measured by (a) Correlation: Mantel correlation between edge weights in observed and true networks, (b) Accuracy: Absolute difference in edge weights between observed and true networks (lower values = more accurate networks), (c) Number of individuals in observed networks, and (d) Power: Proportion of true positives. LOESS smoother added to (a–c) to aid visual interpretation. Panel (c) refers to the data in the simulated observation dataset, which is identical for both methods.
Glossary

**True shared space use network:** A network which is a true representation of space-sharing amongst individuals in a population, against which observed shared space use networks are compared. Edge weights represent the amount of overlap in the home range between each pair of individuals.

**Sampling intensity:** The number of draws from a binomial distribution \{0,1\} given a probability of observing an individual in a trap. Combined with trapping grid density, results in a mean number of observations per individual.

**Sampling regime:** A combination of (i) the trapping grid density used in a study, and (ii) the sampling intensity in this grid, resulting in some mean number of observations per individual for a study.

**Individual centroid:** The centre of an individual’s home range. Calculated by taking the mean of the positions it was observed in weighted by the number of times it was observed at each of these positions.

**Home range profile:** The change in probability of detection with increasing distance from the centroid of an individual (here described using a negative sigmoidal curve, but see Discussion). Assumes an individual has a core and periphery to their home range, with the probability of detection higher closer to the core, and lower further away from the core.
Simulated observation dataset: A dataset that contains detections of individuals at traps, where the detection probability for a given individual in a given trap is determined by the position of the trap relative to the individual’s true home range profile.

Space sharing event: The observation of two individuals in the same spatial locations, within a given defined temporal boundary (if applicable).

Shared trap network: A network based on observed trap-sharing events only. Edge weights represent the number of traps in which both individuals were detected divided by the number of traps in which either or both individuals were detected.

Spatial overlap network: A network based on the amount of spatial overlap in the home range among individuals. Edge weights represent the overlap between two individuals’ home ranges. In our case, we calculated home range overlap numerically by overlaying the two individuals’ home range profiles and calculating the area under the two curves using Equation 2.

Observed network: A network in which the estimation of the relationships (here space-sharing) among individuals in a population is based on an observed dataset. In our case, we simulated observed datasets and created observation networks using both space sharing (shared traps) and spatial overlap approaches.

Power of observed networks: The proportion of observed networks in which one can detect a biological effect known to be present in the true network.
**Correlation of observed network:** The correlation between the edge weights in an observed network and the edge weights in a true network using a Mantel test.

**Accuracy of observed network:** The sum of the absolute differences between the edge weights in an observed network and the edge weights in a true network.

**Acknowledgements**

KMW was supported by a Natural Environment Research Council (NERC) research grant NE/L013452/1 awarded to Mike Begon, Steve Paterson (University of Liverpool), Janette Bradley (Universiy of Nottingham) and Joseph Jackson (University of Salford), and a Johnston Postdoctoral Development Award from the University of Liverpool. DRF received funding from the Max Planck Society, an Eccellenza Professorship Grant of the Swiss National Science Foundation (Grant Number PCEFP3_187058), the European Research Council (ERC) under the European Union’s Horizon 2020 research and innovation programme (grant agreement No. 850859). The empirical data used in this study were collected as part of the NERC research grant NE/E015131/1 awarded to Mike Begon, Steve Parterson, Janette Bradley and Richard Birtles (University of Salford). We would like to thank the many individuals involved in generating this dataset, particularly Joseph Jackson, and the Forestry Commission for access to the study sites.
Declarations

Data availability

Data available from the Dryad Digital Repository:

https://datadryad.org/stash/dataset/doi:10.5061/dryad.bk537. R code demonstrating how to apply our method and to replicate the simulations can be found at:

https://github.com/kwanelik/Shared-space-use-networks.

Competing interests

The authors declare no competing interests.

Author contribution statement

Both authors designed the study, analysed the data and wrote the manuscript.
References

Berkvens R, Olivares IH, Mercelis S, et al (2019) Contact detection for social networking of small animals. In: Advances on P2P, Parallel, Grid, Cloud and Internet Computing. Taichung, Taiwan

Borowski Z (2003) Habitat selection and home range size of field voles Microtus agrestis in Słowiński National Park, Poland. Acta Theriol (Warsz) 48:325–333.
https://doi.org/10.1007/BF03194172

Butts C (2016) sna: Tools for Social Network Analysis. R package version 2.4.
https://CRAN.R-project.org/package=sna

Calhoun JB, Casby JU (1958) Calculation of home range and a density of small mammals. United States Public Heal Monogr 55:1–24

Cantor M, Maldonado-Chaparro AA, Beck KB, et al (2021) The importance of individual-to-society feedbacks in animal ecology and evolution. J Anim Ecol 90:27–44.
https://doi.org/10.1111/1365-2656.13336

Craft ME, Caillaud D (2011) Network Models: An Underutilized Tool in Wildlife Epidemiology? Interdiscip Perspect Infect Dis 2011:.
https://doi.org/10.1155/2011/676949

Dahle B, Swenson JE (2003) Home ranges in adult Scandinavian brown bears (Ursus arctos): Effect of mass, sex, reproductive category, population density and habitat type. J Zool 260:329–335. https://doi.org/10.1017/S0952836903003753

Davis GH, Crofoot MC, Farine DR (2018) Estimating the robustness and uncertainty of animal social networks using different observational methods. Anim Behav 141:29–44. https://doi.org/10.1016/j.anbehav.2018.04.012

Davis S, Abbasi B, Shah S, et al (2014) Spatial analyses of wildlife contact networks. J R Soc Interface 12:20141004. https://doi.org/10.1098/rsif.2014.1004
Douglas M, Ji W, Clout MN (2006) MateID: Design and Testing of a Novel Device For Recording Contacts Between Free-Ranging Animals. Wildl Soc Bull 34:203–207. https://doi.org/10.2193/0091-7648(2006)34[203:mdatoa]2.0.co;2

Farine DR (2015) Proximity as a proxy for interactions: Issues of scale in social network analysis. Anim Behav 104:e1–e5. https://doi.org/10.1016/j.anbehav.2014.11.019

Farine DR (2018) When to choose dynamic vs. static social network analysis. J Anim Ecol 87:128–138. https://doi.org/10.1111/1365-2656.12764

Farine DR (2017) A guide to null models for animal social network analysis. Methods Ecol Evol 8:1309–1320. https://doi.org/10.1111/2041-210X.12772

Farine DR, Strandburg-Peshkin A (2015) Estimating uncertainty and reliability of social network data using Bayesian inference. R Soc Open Sci 2:150367. https://doi.org/10.1098/rsos.150367

Farine DR, Whitehead H (2015) Constructing, conducting, and interpreting animal social network analysis. J Anim Ecol 84:1144–1163. https://doi.org/10.1111/1365-2656.12418

Fleming CH, Calabrese JM (2017) A new kernel density estimator for accurate home-range and species-range area estimation. Methods Ecol Evol 8:571–579. https://doi.org/10.1111/2041-210X.12673

Gimenez O, Mansilla L, Klaich MJ, et al (2019) Inferring animal social networks with imperfect detection. Ecol Modell 401:69–74. https://doi.org/10.1016/j.ecolmodel.2019.04.001

Godsall B, Coulson T, Malo AF (2014) From physiology to space use: Energy reserves and androgenization explain home-range size variation in a woodland rodent. J Anim Ecol 83:126–135. https://doi.org/10.1111/1365-2656.12116

Grear DA, Luong LT, Hudson PJ (2013) Network transmission inference: Host behavior and parasite life cycle make social networks meaningful in disease ecology. Ecol Appl
Grear DA, Perkins SE, Hudson PJ (2009) Does elevated testosterone result in increased exposure and transmission of parasites? Ecol Lett 12:528–537. https://doi.org/10.1111/j.1461-0248.2009.01306.x

Hart JDA, Franks DW, Brent LJN, Weiss MN (2021) Accuracy and power analysis of social networks built from count data. Methods Ecol Evol 00:1–10. https://doi.org/10.1111/2041-210X.13739

Hayne D (1949) Calculation of size of home range. J Mammal 30:1–18

Hayne D (1950) Apparent Home Range of Microtus in Relation to Distance between Traps. Am Soc Mammalogists 31:26–39

Hoppitt WJE, Farine DR (2018) Association indices for quantifying social relationships: how to deal with missing observations of individuals or groups. Anim Behav 136:227–238. https://doi.org/10.1016/j.anbehav.2017.08.029

Jackson JA, Hall AJ, Friberg IM, et al (2014) An immunological marker of tolerance to infection in wild rodents. PLoS Biol 12:e1001901. https://doi.org/10.1371/journal.pbio.1001901

Jennrich RI, Turner FB (1969) Measurement of non-circular home range. J Theor Biol 22:227–237. https://doi.org/10.1016/0022-5193(69)90002-2

Klein GP, Cameron GN (2012) Effect of habitat gradients on space use by white-footed mice (Peromyscus leucopus). J Mammal 93:706–715. https://doi.org/10.1644/11-MAMM-A-258.1

Mikesic D, Drickamer L (1992) Factors Affecting Home-Range Size in House Mice (Mus musculus domesticus) Living in Outdoor Enclosures. Am Midl Nat 127:31–40

Myllymaki A (1977) Intraspecific competition and home range dynamics in the field vole Microtus agrestis. Oikos 29:553–569
Niethammer J, Krapp F (1982) Microtus agrestis (Linnaeus, 1761). In: Niethammer J, Krapp F (eds) Handbuch der saugetierkunde Europas, Volume 2/I, Nagetiere II. Akademische Verlag, Wiesbaden, Germany, pp 350–373

Noonan MJ, Tucker MA, Fleming CH, et al (2019) A comprehensive analysis of autocorrelation and bias in home range estimation. Ecol Monogr 89:e01344. https://doi.org/10.1002/ecm.1344

Oksanen J, et al (2017) vegan: Community Ecology Package. R package version 2.4-3. https://CRAN.R-project.org/package=vegan

Perkins S, Ferrari M, Hudson P (2008) The effects of social structure and sex-biased transmission on macroparasite infection. Parasitology 135:1561–1569. https://doi.org/10.1017/S0031182008000449

Perkins SE, Cagnacci F, Stradiotto A, et al (2009) Comparison of social networks derived from ecological data: Implications for inferring infectious disease dynamics. J Anim Ecol 78:1015–1022. https://doi.org/10.1111/j.1365-2656.2009.01557.x

Porphyre T, Stevenson M, Jackson R, McKenzie J (2008) Influence of contact heterogeneity on TB reproduction ratio R0 in a free-living brushtail possum Trichosurus vulpecula population. Vet Res 39:31. https://doi.org/10.1051/vetres:2008007

R Core Team (2018) R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL https://www.R-project.org/.

Raulo A, Allen BE, Troitsky T, et al (2021) Social networks strongly predict the gut microbiota of wild mice. ISME J 15:2601–2613. https://doi.org/10.1038/s41396-021-00949-3

Rutz C, Burns ZT, James R, et al (2012) Automated mapping of social networks in wild birds. Curr Biol 22:R669–R671. https://doi.org/10.1016/j.cub.2012.06.037

Ryder TB, Horton BM, van den Tillaart M, et al (2012) Proximity data-loggers increase the
quantity and quality of social network data. Biol Lett 8:917–920.
https://doi.org/10.1098/rsbl.2012.0536

Sabol AC, Solomon NG, Dantzer B (2018) How to Study Socially Monogamous Behavior in Secretive Animals? Using Social Network Analyses and Automated Tracking Systems to Study the Social Behavior of Prairie Voles. Front Ecol Evol 6:178.
https://doi.org/10.3389/fevo.2018.00178

Schoener TW (1981) An empirically based estimate of home range. Theor Popul Biol 20:281–325. https://doi.org/10.1016/0040-5809(81)90049-6

Slade NA, Russell LA (1998) Distances as Indices to Movements and Home-Range Size From Trapping Records of Small Mammals. J Mammal 79:346–351.
https://doi.org/10.2307/1382871

Smith JE, Gamboa DA, Spencer JM, et al (2018) Split between two worlds: Automated sensing reveals links between above- and belowground social networks in a free-living mammal. Philos Trans R Soc B Biol Sci 373:20170249.
https://doi.org/10.1098/rstb.2017.0249

Spencer SR, Cameron GN, Swihart RK (1990) Operationally Defining Home Range: Temporal Dependence Exhibited by Hispid Cotton Rats. Ecology 71:1817–1822

Swihart RK, Slade NA (1989) Differences in Home-Range Size between Sexes of Microtus ochrogaster. J Mammal 70:816–820. https://doi.org/10.2307/1381718

Tompkins DM, Dunn AM, Smith MJ, Telfer S (2011a) Wildlife diseases: From individuals to ecosystems. J Anim Ecol 80:19–38. https://doi.org/10.1111/j.1365-2656.2010.01742.x

Tompkins DM, Dunn AM, Smith MJ, Telfer S (2011b) Wildlife diseases: From individuals to ecosystems. J Anim Ecol 80:19–38. https://doi.org/10.1111/j.1365-2656.2010.01742.x

VanderWaal KL, Atwill ER, Hooper S, et al (2013) Network structure and prevalence of
Cryptosporidium in Belding’s ground squirrels. Behav Ecol Sociobiol 67:1951–1959. https://doi.org/10.1007/s00265-013-1602-x

Webber QMR, Vander Wal E (2019) Trends and perspectives on the use of animal social network analysis in behavioural ecology: a bibliometric approach. Anim Behav 149:77–87. https://doi.org/10.1016/j.anbehav.2019.01.010

White LA, Forester JD, Craft ME (2017) Using contact networks to explore mechanisms of parasite transmission in wildlife. Biol Rev 92:389–409. https://doi.org/10.1111/brv.12236

Whitehead H (2008a) Analyzing Animal Societies. University of Chicago Press, Chicago, IL, USA

Whitehead H (2008b) Precision and power in the analysis of social structure using associations. Anim Behav 75:1093–1099. https://doi.org/10.1016/j.anbehav.2007.08.022

Winner K, Noonan MJ, Fleming CH, et al (2018) Statistical inference for home range overlap. Methods Ecol Evol 9:1679–1691. https://doi.org/10.1111/2041-210X.13027

Wohlfiel CK, Leu ST, Godfrey SS, Bull CM (2013) Testing the robustness of transmission network models to predict ectoparasite loads. One lizard, two ticks and four years. Int J Parasitol Parasites Wildl 2:271–277. https://doi.org/10.1016/j.ijppaw.2013.09.005

Wolton RJ, Flowerdew JR (1985) Spatial distribution and movements of wood mice, yellow necked mice and bank voles. Symp Zool Soc London 55:249–275

Zamora L, Moreno-Amich R (2002) Quantifying the activity and movement of perch in a temperate lake by integrating acoustic telemetry and a geographic information system. Hydrobiologia 483:209–218. https://doi.org/10.1023/A:1021396016424