High-quality imaging of endolymphatic hydrops acquired in 7 minutes using sensitive hT₂W–3D–FLAIR reconstructed with magnitude and zero-filled interpolation

Jing Zou1,3 · Luguang Chen2 · Hongbin Li1 · Guoping Zhang1 · Ilmari Pyykkö3 · Jianping Lu2

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Abstract

Background It is still challenging to detect endolymphatic hydrops (EH) in patients with Meniere’s disease (MD) using MRI. The aim of the present study was to optimize a sensitive technique generating strong contrast enhancement from minimum gadolinium–diethylenetriamine pentaacetic acid (Gd–DTPA) while reliably detecting EH in the inner ear, including the apex.

Materials and methods All imaging was performed using a 3.0 T MR system 24 h after intratympanic injection of low-dose Gd–DTPA. Heavily T2-weighted 3-dimensional fluid-attenuated inversion recovery reconstructed with magnitude and zero-filled interpolation (hT₂W–FLAIR–ZFI) was optimized and validated in phantom studies and compared with medium inversion time inversion recovery imaging with magnitude reconstruction (MIIRMR). The following parameters were used in hT₂W–FLAIR–ZFI: repetition time 14,000 ms, echo time 663 ms, inversion time 2900 ms, flip angle 120°, echo train length 271, and field of view 166 × 196 mm².

Results MRI obtained using hT₂W–FLAIR–ZFI yielded high-quality images with sharper and smoother borders between the endolymph and perilymph and a higher signal intensity ratio and more homogenous perilymph enhancement than those generated with MIIRMR (p < 0.01). There were predominantly grade II EHs in the cochlea and grade III EHs in the vestibule in definite MD. EH was detected in the apex of 11/16 ipsilateral ears, 3/16 contralateral ears in unilateral definite MD and 3/6 ears in bilateral MD.

Conclusions The novel hT₂W–FLAIR–ZFI technique is sensitive and demonstrates strong and homogenous enhancement by minimum Gd–DTPA in the inner ear, including the apex, and yields high-quality images with sharp borders between the endolymph and perilymph.

Keywords Meniere’s disease · Endolymphatic hydrops · MRI · Image reconstruction · Enhancement

Introduction

MRI has been increasingly used to assess endolymphatic hydrops (EH) in clinical practice to diagnose inner ear diseases, especially Meniere’s disease (MD), since the first report of an animal study in 2000 and following optimization of the MRI protocol [1–6]. Strong contrast enhancement of the perilymph is essential for reporting reliable results of EH, which depends on sufficient distribution of the contrast agents (gadolinium chelate) in the inner ear and the MRI sequence. However, this is not guaranteed with the current technique. Although the barrier from blood to perilymph is looser than that to endolymph, entry of contrast agents into the perilymph after intravenous injection of gadolinium chelate is still hindered by the rather tight blood-perilymph barrier unless it is severely impaired [7–9]. Aiming to boost
the uptake of contrast agents in the inner ear, intratympanic administration of gadolinium chelate was developed as an alternative method for imaging EH in the clinic, but this method still failed in 13% of patients [3, 10, 11]. Uneven diffusion of contrast agents inside the inner ear resulted in weak distribution of gadolinium chelate in the cochlear apex regardless of whether the contrast agents were delivered intratympanically or intravenously and may also affect the reliability of the method in the detection of EH because hydrops is suspected to start from the apex in patients with low-tone hearing loss [12–14]. In animal experiments with a two-phase EH model, a combination of chronic endolymphatic sac dysfunction and acute stress-induced endolymph production, the most severe degree of EH was present in the cochlear apex [15]. Therefore, there is a need in the clinic to develop a more sensitive method for the detection of EH.

Naganawa et al. aimed to improve the sensitivity of MRI methods for the detection of EH and reported that heavily T2-weighted three-dimensional fluid-attenuated inversion recovery (hT2W–3D–FLAIR) and a hybrid of reversed images of positive endolymph signals and native images of perilymph signals multiplied with heavily T2-weighted MR cisternography (HYDROPS-Mi2) imaging were optimized methods [7, 16]. The group recently further optimized the parameters of 3D-real inversion recovery imaging based on phase-sensitive reconstruction by prolonging the repetition time and increasing the refocusing flip angle, which yielded a similar value in detecting EH to HYDROPS-Mi2 imaging, which is complicated and time consuming for clinical applications [14, 17]. Zou et al. further improved the contrast enhancement using magnitude reconstruction, which was named medium inversion time inversion recovery imaging with magnitude reconstruction (MIIRMR), and demonstrated a sharper border between the perilymph and the endolymph than that when using hT2W–3D–FLAIR after intratympanic injection of 20-fold-diluted gadolinium–diethylentriamine pentaacetic acid (Gd–DTPA) [18]. During a pilot study, magnitude reconstruction of MIIRMR was compared with real reconstruction of 3D-real inversion recovery imaging as recently reported using otherwise identical parameters (Table 1) [17]. MIIRMR was significantly more sensitive than 3D-real inversion recovery (Supplementary information 1). The scan time of MIIRMR was significantly shorter than that of 3D-real inversion recovery imaging (8.6 min versus 11.4 min), although a coil with fewer channels was used (20 versus 32 channels) [17, 18]. However, the enhancement in the perilymph of the cochlear apex was still weak in most cases. Zero-filled interpolation (ZFI) is a technique used to reduce partial-volume artifacts and to improve the appearance of small structures and edges while retaining genuine information of the raw images [19]. ZFI has been applied in intracranial MR angiography to improve the contrast and continuity of examined small vessels in the range of 0.2–0.6 mm but leads to insignificant improvement in the accuracy of luminal measurement in medium-sized vessels [20, 21].

We hypothesized that T2W–3D–FLAIR reconstructed with magnitude and ZFI (hT2W–FLAIR–MZFI) may further improve the demonstration of EH in gadolinium enhancement MRI, especially in the cochlear apex, which has a rather small size within the above-mentioned range and a usually weak uptake of gadolinium chelate [22, 23].

The purpose of the present work was to demonstrate a technique employing hT2W–FLAIR–MZFI that was capable of generating strong contrast enhancement from minimum Gd–DTPA to reliably detect EH even in the cochlear apex of patients and to compare the results with those derived using the MIIRMR method.

### Table 1 The parameters for hT2W–FLAIR–MZFI, MIIRMR and 3D-real inversion recovery sequences

| Sequences            | TR/TE (ms) | TI (ms) | FA   | ETL | Matrix | FOV (mm) | Rec | Time (min) |
|----------------------|------------|--------|------|-----|--------|----------|-----|------------|
|                      |            |        |      |     | scan   |          |     |            |
| hT2W–FLAIR–MZFI*     | 14,000/663 | 900    | 120° | 271 | 324 x 384 | 648 x 768 | 166 x 196 | Mag + ZFI | 7.4 |
| hT2W–FLAIR–MZFI*     | 7000–16,000/663 | 2900   | 120° | 271 | 324 x 384 | 648 x 768 | 166 x 196 | Mag + ZFI | 3.3–8.8 |
| MIIRMR               | 16,000/663 | 2700   | 120° | 271 | 324 x 384 | 324 x 384 | 166 x 196 | Mag       | 8.8 |
| 3D real IR           | 16,000/663 | 2700   | 120° | 271 | 324 x 384 | 324 x 384 | 166 x 196 | Real      | 8.8 |

*3D real IR* 3D real inversion recovery, *ETL* echo chain length, *FA* flip angle, *FOV* field of view; hT2W–FLAIR–MZFI*: heavily T2-weighted 3D-fluid attenuated inversion recovery reconstructed with magnitude and zero-filled interpolation that was applied in patient studies; hT2W–FLAIR–MZFI*: hT2W–FLAIR–MZFI used in phantom studies for comparisons of TR-dependent sensitivity with the following TR 16,000 ms, 14,000 ms, 12,000 ms, 9000 ms, and 7000 ms, *MIIRMR* medium inversion time inversion recovery imaging with magnitude reconstruction, *Rec* reconstruction, *TE* echo time, *TI* inversion time, *Time* scan time, *TR* repetition time, *Mag* magnitude reconstruction. Other parameters: fat saturation; GRAPPA: 2; Flip angle mode: constant; number of excitations: 2; slice partial fourier: 6/8; slices per slab: 30 in hT2W–FLAIR–ZFI* and 60 in other sequences; slice thickness: 1 mm; voxel size = 0.26 × 0.26 × 1.0 mm³ for hT2W–FLAIR–MZFI* and hT2W–FLAIR–ZFI*, and 0.52 × 0.52 × 1.0 mm³ for MIIRMR and 3D real IR.
Materials and methods

Experimental optimization of the imaging method

Empirical study

Five patients diagnosed with definite MD (for the inclusion criteria, see section (“Clinical evaluation of the developed imaging method”) were selected for the empirical MRI studies with respect to the reconstruction method, parameter optimization of hT2W–FLAIR–MZFI imaging, and dilution of Gd–DTPA. MRI was performed 24 h postinjection of Gd–DTPA using a 3 T MR system (MAGNETOM Skyra, Siemens Healthcare, Erlangen, Germany) equipped with a 20-channel head/neck coil. After testing with various repetition time (TR) and inversion time (TI) values, TR 14,000 ms and TI 2900 ms were chosen (Table 1). Various dilutions of Gd–DTPA (9.38 g, 20 ml, Beijing Beilu Pharmaceutical CO., LTD, Beijing, China) (20-, 25-, and 30-fold) were tested in four patients with the diseased ear fixed at a 20-fold dilution, while the contralateral ear was exposed to a lower concentration following a previously reported protocol of intratympanic injection of 0.1 ml of the above diluted Gd–DTPA that was directly delivered onto the posterior upper surface of the tympanic medial wall using a catheter passing through the perforated tympanic membrane [18, 24, 25]. Finally, a 20-fold dilution, which was also reported to induce sufficient uptake of Gd–DTPA in the inner ear with the least individual variability [25], was defined as the optimized concentration in the following clinical study.

Phantom study

Three serial dilutions of Gd–DTPA were prepared and injected into 2-ml plastic cryopreservation tubes as follows. The first sample (phantom 1) was serially diluted from 1 × 10³-fold through 1 × 10⁸-fold with an interval of tenfold using physiological saline. Since the perilymph contains proteins at approximately 2.8 mg/ml and the perilymph/cerebrospinal fluid (CSF) protein ratios are 5 in humans and 12.5 in guinea pigs [26], bovine serum albumin (BSA) samples were prepared using physiological saline at concentrations of 11.2, 5.6, and 2.8 mg/ml to test the potential contribution of perilymphatic proteins to signal production. Based on the results of the first study, the second sample (phantom 2) was serially diluted from 2 × 10³ to 5.12 × 10⁵. To assess the impact of repetition time on the sensitivity of hT₂W–FLAIR–MZFI, the third sample (phantom 3) was serially diluted from 2 × 10³ to 1.28 × 10⁵. Tubes containing physiological saline were used as blank controls, and duplicates were prepared for each dilution. Phantoms 1 and 2 were imaged using both hT₂W–FLAIR–MZFI and MIIRMR sequences (Table 1). Phantom 3 was imaged using the hT₂W–FLAIR–MZFI sequence with the following TRs: 16,000, 14,000, 12,000, 9000, and 7000 ms (Table 1).

Clinical evaluation of the developed imaging method

Patient information

Patients who visited author JZ at the Department of Otolaryngology-Head & Neck Surgery, Changhai Hospital, Second Military Medical University, from June to November 2020 were screened. The inclusion criteria were as follows: (1) diagnosis of definite or probable MD made according to the criteria of the Barany Society in 2015 [27] and (2) sudden sensorineural hearing loss accompanied by vertigo or constant balance problems meeting the disease definitions (AAO-HNS, 2012) [28]. The exclusion criteria were as follows: (1) vertigo with other origins, such as benign paroxysmal positional vertigo and diseases in the central nervous system; and (2) sensorineural hearing loss without vertigo. In total, 22 patients diagnosed with MD (aged 28–74 years, average age 47.1 years; 10 males and 12 females) and one patient diagnosed with sudden sensorineural hearing loss (79-years old, female) were recruited for the study. The participants formed a consecutive series.

Imaging

Each patient was imaged 24 h after intratympanic injection of 0.1 ml of 20-fold-diluted Gd–DTPA that was delivered onto the posterior upper part of the tympanic medial wall as previously reported [18, 24, 25]. In total, one patient was imaged with hT₂W–FLAIR–MZFI, MIIRMR and 3D-real inversion recovery, 10 patients were imaged using both hT₂W–FLAIR–MZFI and MIIRMR, and 12 patients were imaged only with hT₂W–FLAIR–MZFI and MIIRMR are shown in Table 1. T₂-weighted-sampling perfection with application-optimized contrasts using a flip angle evolution (T₂W-SPACE) sequence was applied in all cases to image potential inner ear fibrosis or vestibular schwannoma, as previously reported [18].

Image analysis

All images were sent to an advanced workstation (Leonardo, Siemens Healthcare, Germany) for further analysis by author JZ, who has more than 21 years of experience in inner ear imaging [1, 2]. The maximum cross sectional
slices of the cochlea and vestibule were chosen for the image analysis. First, the general image quality with respect to the border between the unenhanced endolymph and enhanced perilymph was subjectively evaluated. Image sensitivity, diversity of the inner ear enhancement signal, and sensitivity in detecting EH in the cochlea, including the apex, were assessed. The sensitivity was expressed as signal intensity ratios (SIRs) in various regions of interest (ROIs) by manual drawing. The diversity of signal intensity was displayed as differences in SIR between different sites of the inner ear. Finally, the sensitivity was tested by analyzing the percentage of EH in the cochlear apex of patients with MD. Signal intensity is indicated as a gray value, and SIRs in the phantom samples were calculated using physiological saline as the reference. In patients, potential tumor appearance in the cerebellopontine angle was screened on MRI obtained using the T_{2}W-SPACE sequence. The accuracy, specificity, and sensitivity for noncontrast MRI in detecting VS were reported to reach 97%–99%, 96%–98%, and 100%, respectively [29]. T_{2}W-SPACE is superior to other sequences due to the higher resolution. SIRs in the defined ROIs in the cochlear basal turn, apex and vestibule were calculated using CSF as references. In addition to identifying EH in the cochlear apex, cochlear and vestibular EH were graded according to a previous report that applied a 3-stage method for cochlear hydrops and a 4-stage scale for vestibular hydrops based on morphological changes as follows. In the cochlea, 0 = no hydrops, the interscalar septum, the scala tympani, and scala vestibuli are recognizable with the scala media normally minimally visible; I = mild hydrops, the scala media becomes a nodular black area with narrow borders between the endolymph and perilymph; II = marked hydrops, the scala vestibuli becomes completely invisible. In the vestibule, 0 = no hydrops, the smaller saccule and larger utricle are visibly separated and occupy less than half of the surface of the vestibule; I = mild hydrops, the saccule becomes equal to or larger than the utricle but is not yet merged with the utricle; II = marked hydrops, the saccule merges with the utricle, and the surrounding perilymphatic space appears as a bright peripheral rim; and III = extreme hydrops, the perilymphatic enhancement of the vestibule is no longer visible [30].

**Statistical analysis**

IBM SPSS 27 (IBM, Chicago, IL, USA) software was used for performing the statistical analysis. The mean values of SIRs of repeated serially diluted Gd–DTPA phantom samples imaged using hT_{2}W–FLAIR–MZFI and MIIRMR were used for the statistical analysis. Univariate analysis of the general linear model and Pearson’s correlation test were applied to analyze correlations between SIR and Gd–DTPA concentrations. A paired-sample t-test was used for comparisons of SIR obtained with hT_{2}W–FLAIR–MZFI and MIIRMR at various concentrations of Gd–DTPA in phantom samples. Student’s t-test was applied to compare SIR differences between the cochlear basal turn and the apex and between the vestibule and the apex. A paired-sample t-test was used for comparison of SIR obtained with hT_{2}W–FLAIR–MZFI and MIIRMR in defined ROIs in the inner ear of 10 patients imaged using both hT_{2}W–FLAIR–MZFI and MIIRMR. p < 0.05 was considered statistically significant.

**Results**

**Experimental optimization of the imaging method**

**Comparing the effects of hT_{2}W–FLAIR–MZFI and MIIRMR in a phantom study**

In a phantom study, both the hT_{2}W–FLAIR–MZFI and MIIRMR techniques demonstrated significant linear correlations between the SIR and logarithm of gadolinium concentration (p = 0.029 and 0.019, respectively). However, the curve created with hT_{2}W–FLAIR–MZFI had a larger intercept and smaller slope than that created with MIIRMR (y = 1.44 + 0.36 x and y = 1.34 + 0.57 x, respectively) (Fig. 1). Equal SIR (cross point) was obtained using both hT_{2}W–FLAIR–MZFI and MIIRMR at 2.99 nM Gd–DTPA. A comparison between the sensitivity and repetition time (TR) shows that the longer the TR was, the more sensitive the detection. The image sensitivity became nearly saturated when TR was longer than 12,000 ms (the curves almost overlapped from TR 12,000 to 16,000 ms) (Fig. 2).

**Comparing the effects of MRI with hT_{2}W–FLAIR–MZFI and MIIRMR in patients**

The inner ear MRI obtained using hT_{2}W–FLAIR–MZFI displayed high-quality images with sharper and smoother borders between the endolymph and perilymph than those obtained using MIIRMR (Fig. 3). Each turn of the cochlea showed excellent contrast, including both the hydric and normal ears without oversaturation of the enhancement signal in either the cochlear basal turn or the vestibule imaged using hT_{2}W–FLAIR–MZFI (Fig. 3A, B). The apex of the diseased cochlea was vague due to the faint contrast on the images obtained using MIIRMR (Fig. 3C–E). Quantifications demonstrated that the SIRs in the cochlear basal turn, apex and vestibule imaged using hT_{2}W–FLAIR–MZFI were significantly higher than those obtained with MIIRMR in patients administered Gd–DTPA with the present method. However, the
SIRs in the cochlear basal turn over the apex imaged using hT_{2}W–FLAIR–MZFI were significantly lower than those obtained with MIIRMR (Table 2). In other words, the hT_{2}W–FLAIR–MZFI sequence detected a broader range of Gd–DTPA signals and could display an enhancement of the inner ear with less diversity than MIIRMR.

**Clinical evaluation of the developed imaging method**

Grading the EH in both the cochlea and the vestibule was carried out based on images acquired using hT_{2}W–FLAIR–MZFI 24 h after administration of 0.1 ml of 20-fold-diluted Gd–DTPA into the middle ear cavity (Figs. 3, 4, and 5). The grades were predominantly grade II in the cochleae (11/16 ears in unilateral and 5/6 ears in bilateral) and grade III in the vestibule (8/16 ears in unilateral and 4/6 ears in bilateral) in definite MD. EH was detected in the apex of 11/16 ipsilateral ears, 3/16 contralateral ears in unilateral definite MD and 3/6 ears in bilateral MD (Table 3).

**Discussion**

The present phantom study demonstrated that hT_{2}W–FLAIR–MZFI is more sensitive in detecting the signal of extremely low-concentration Gd–DTPA and suppressing the signal of high-concentration Gd–DTPA than MIIRMR. The SIRs in the cochlear basal turn over the apex imaged using hT_{2}W–FLAIR–MZFI were significantly lower than those obtained with MIIRMR, indicating that the hT_{2}W–FLAIR–MZFI sequence yielded a broader range of Gd–DTPA signals and could display an enhancement of the inner ear with less diversity than MIIRMR. The image quality obtained with hT_{2}W–FLAIR–MZFI was significantly higher than that obtained with MIIRMR in the same inner ear, presenting borders between the endolymph and perilymph that were sharper and smoother when using hT_{2}W–FLAIR–MZFI than when using...
Fig. 3 Comparison of hT2W–FLAIR–MZFI and MIIRMR 24 h after targeted delivery of 20-fold-diluted Gd–DTPA in a patient with Meniere’s disease. The patient suffered from episodic vertigo for 20 years and fluctuating right ear hearing loss for 1 year. The image quality and enhancement in the cochlear apex were significantly higher on MRI obtained with hT2W–FLAIR–MZFI A, B than on MRI obtained with MIIRMR C–E. The images acquired using MIIRMR were obviously inhomogeneous C–E. Grade II cochlear endolymphatic hydrops (EH) was detected on the right (R) (zoom in A) using hT2W–FLAIR–MZFI but not with MIIRMR (zoom in D). Grade I vestibular EH was demonstrated in the right ear using both hT2W–FLAIR–MZFI (zoomed in B) and MIIRMR (zoomed in E). EH was not demonstrated in the left ear (L) imaged with either hT2W–FLAIR–MZFI (zoomed in A, B) or MIIRMR. Signal intensities in the regions of interest (circled areas in A, B, D, and E) were evaluated on a single slide for statistical analysis. 1st basal turn, CSF cerebrospinal fluid, Sa saccule, Ut utricle. Scale bars = 3.0 mm

Table 2 Comparison of the signal intensity ratio in inner ear MRI obtained using hT2W–FLAIR–MZFI and MIIRMR

| Sequences      | SIR (mean ± SD)/n | Pairs<sup>a</sup> | C-BT<sup>b</sup> | C-Ap<sup>b</sup> | Vest<sup>b</sup> | C-BT/Ap |
|----------------|-------------------|-------------------|-----------------|-----------------|----------------|---------|
| hT2W–FLAIR–MZFI| 9.076 ± 5.011/60  | 9.527 ± 3.519**/44| 4.238 ± 1.888/44| 11.553 ± 4.370**/44| 2.365 ± 0.703***/44|
| MIIRMR         | 1.634 ± 1.203/60  | 1.929 ± 0.938**/21| 0.538 ± 0.435/21| 2.515 ± 1.254/**21| 4.073 ± 1.966/21|

<sup>a</sup>Signal intensity ratio in C-Ap, C-BT, and Vest of 10 cases who were imaged with both sequences were counted

<sup>b</sup>Signal intensity ratio of the defined sites over the cerebrospinal fluids
MIIRMR. The voxel size of hT2W–FLAIR–MZFI was significantly smaller than that of MIIRMR, which supported the advantages of hT2W–FLAIR–MZFI over MIIRMR (0.26 × 0.26 × 1.0 mm³ vs. 0.52 × 0.52 × 1.0 mm³). The optimized parameters were also validated. Clinical MRI with the novel hT2W–FLAIR–MZFI technique displayed a good enhancement signal in the human inner ear, corresponding to a broad range of gadolinium concentrations, and was capable of reliably demonstrating visible enhancement in the cochlear apex. EH was frequently detected in the cochlear apex of patients with MD in addition to the lower turns and vestibule.
Comparison of the current technique with reports from the literature

In an early clinical MRI study using the 3D-FLAIR technique, uptake of gadolinium in the inner ear was absent in 5% of ears and insufficient to evaluate EH in 13% of ears, although a significantly higher dose of gadolinium chelates than that in the present study was injected into the middle ear cavity (eightfold dilution, 0.4–0.5 ml/ear) [10]. The same group reported that a more sensitive sequence of hT2W–3D–FLAIR was capable of detecting gadolinium signals in the perilymph of MD patients using a 32-channel coil after intravenous injection of single-dose GdC [7, 31]. The optimized parameters were TR 9000 ms and TI 2400 ms with an imaging time of 10.7 min, which is significantly longer than that for the present study, although a
head coil with many more channels than that in our study was used. However, the image quality and contrast effect were lower than those in the present study. To improve the contrast effect, Naganawa et al. developed a technique called HYDROPS (HYbriD of reversed image of positive endolymph signal and native image of positive perilymph signal) using an image calculation that was finally updated to HYDROPS-Mi2 [16]. However, the procedure is relatively complicated in clinical practice, demands special skills in the evaluation of EH and is time-consuming (~18 min). Recently, Ohashi et al. simplified the technique by prolonging the repetition time and increasing the refocusing flip angle in 3D-real inversion recovery imaging, which resulted in a higher contrast-to-noise ratio than the previous sequence without image calculation and was capable of detecting EH in 11.4 min [14]. However, the contrast effect could be improved. The current hT2W–FLAIR–MZFI technique displayed an improved contrast effect in the inner ear from the apex to the other parts with good image quality in 7.4 min using a 3 T MR system equipped with a head coil of only 20 channels instead of 32 channels.

**Methods to improve image quality and contrast effect in the cochlear apex with hT2W–FLAIR–MZFI**

The inner ear MRI obtained using hT2W–FLAIR–MZFI displayed high-quality images with sharp and smooth borders between the scalae and good contrast enhancement in the cochlear apex, where the distribution of Gd–DTPA is usually extremely sparse while avoiding overexposure artifacts in the cochlear basal turn and vestibule where the accumulation of Gd–DTPA is significantly more pronounced than the apex. The phantom study showed that the enhancement signal generated by hT2W–FLAIR–MZFI responding to Gd–DTPA followed a curve with a smaller slope than that generated by MIIRMR. A previous study reported that signal intensity positively correlates with the TI [32]. The longer TI in the hT2W–FLAIR–MZFI sequence than that in the MIIRMR sequence possibly contributes to the higher sensitivity in detecting low concentrations of Gd–DTPA. All previously reported sequences provided weak enhancement in the cochlear apex due to the partial volume effect since the apex is small relative to the nominal pixel size; moreover, there is a loss of contrast between the target and the surrounding structures [7, 17, 18, 33]. The partial volume effect reduces the contrast and continuity of the cochlear apex and the edge between the enhanced perilymph and unenhanced endolymph. In MRI using hT2W–FLAIR–MZFI, the data matrix size (reconstruction pixel grid) was increased in all orthogonal directions, and the unknown values were then zero-filled, which resulted in any given structure being located near a reconstructed pixel’s center, thus improving the overall visualization of the enhancement in the cochlear apex while remaining genuine to the information of the raw images [19]. The unenhanced CSF and endolymph in MRI with hT2W–FLAIR–MZFI were demonstrated as dark signals, possibly attributed to the longer TI than that in the previously reported MIIRMR [18], which further improves the image contrast and additionally contributes to the sharp border. A sharp border between the enhanced and unenhanced structures is beneficial for grading the EH of patients. Attributed to the greater sensitivity, enhancement of the cochlear apex was reliably detected using the hT2W–FLAIR–MZFI sequence, avoiding false-positive or -negative reports on EH in MD.

**Methods to reduce imaging time with hT2W–FLAIR–MZFI**

The scanning time using hT2W–FLAIR–MZFI is shorter than that in previous methods. In previous reports, the imaging time was reduced by deducting slices per slab [18, 24, 25]. In the present study, the imaging time was further reduced by decreasing the TR. Although the TR in the
The Gibbs phenomenon

In addition, known as truncation artifacts, the Gibbs phenomenon is a common image defect in MR images and appears as parallel lines or spurious rings around sharp edges or tissue discontinuities, especially in situations of a high contrast effect [34, 35]. When reviewing the images in the literature, the Gibbs phenomenon definitely occurred in MRI studies of EH using previously reported techniques, although it was not described [36–38]. This drawback was also observed in the present study. In T2W-SPACE images, the Gibbs phenomenon appeared as dark dots surrounding the bright fluids of the inner ear, while in hT2W-FLAIR-MZFI images, it occurred as parallel lines distributed below the basal turn, lateral to the cochlea, around the semicircular canals, and occasionally above the apex (Supplementary Information 3). However, the Gibbs phenomenon never existed between the cochlear scalar and within the vestibular area and did not disturb the judgment of EH. It was recently reported that the Gibbs phenomenon may be simply, effectively, and robustly removed with a minimal amount of image smoothing by reinterpolating the image based on local, subvoxel shifts to sample the ringing pattern at the zero crossings of the oscillating sinc function [39].

Indicated clinical significance of hT2W-FLAIR-MZFI for EH imaging

The specificity of detecting EH using MRI was impaired by reports on EH in healthy volunteers [40]. Although the criteria for judgment of EH may be improved, a blurred border between the positive perilymph and negative endolymph may also contribute to the errors. A sharp border displayed by MRI using the hT2W-FLAIR-MZFI technique may improve the accuracy in EH detection. Improved visualization of the enhancement signal in the cochlear apex provides an excellent tool for thorough evaluation of EH in MD. Theoretically, the novel method may be applied in MRI with intravenous administration of GdC, which is favored by the radiologist [40].

Study limitations

Only 23 patients were evaluated for EH using the hT2W-FLAIR-MZFI technique. Although all cases displayed visible enhancement in the cochlear apex, a large-sample study should be conducted in the future to further confirm the present results. A large-sample study should also be carried out to compare the accuracies of the hT2W-FLAIR-MZFI and MIIRMR techniques in detecting EH. The Gibbs phenomenon does exist in the present imaging protocol, although it does not disturb the judgment of EH and may be further improved with proper techniques in the future [39].

Conclusions

The present study revealed that the novel hT2W-FLAIR-MZFI technique is capable of demonstrating strong enhancement by minimum Gd-DTPA in the inner ear, including the apex, and could homogenously display enhancement of the inner ear while improving the borders between enhanced and unenhanced structures. In addition, the imaging time of the present sequence is shorter than that of previously reported methods. The excellent imaging results provided an opportunity to accurately diagnose EH in the clinic.

Supplementary Information

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Declarations

Conflict of interest The authors declare that they have no conflicts of interest.

Ethical approval The protocol was reviewed and approved by the Ethical Committee of Shanghai Changhai Hospital (CHEC2020-107). All participants provided written informed consent.

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