Supplemental Figure 1

A. Depletion of rRNA in PolyA+ RNA samples

Normox Hypox

Fold Depletion

-120 -100 -80 -60 -40 -20 0 28s

18s

B. Normoxia

h18s h28s 40S 60S 5.8s 5s

Fraction Numbers

Percent rRNA/Fraction

0% 10% 20% 30% 40% 50%

1 2 3 4 5 6 7 8 9 10 11 12

Hypoxia

h18s h28s 40S 60S 5.8s 5s

Fraction Numbers

Percent rRNA/Fraction

0% 10% 20% 30% 40% 50%

1 2 3 4 5 6 7 8 9 10 11 12

Supplemental Figure 1:

(A) qRT-PCR verification of rRNA depletion in PolyA+ RNA. Both 18 and 28s rRNA from cells cultured in Normoxic or 24 hours Hypoxic conditions were depleted over 70-fold indicating greater than 98% depletion of rRNA. (N of 2)

(B) 40 and 60S ribosomal subunits from HEK-293T cells grown in normoxic or hypoxic conditions for 24 hours were separated by differential centrifugation followed by sucrose gradient fractionation. qRT-PCR of individual fractions reveals purity of respective rRNAs in each fraction. Fractions containing 40S and 60S peaks are marked. Fractions with highest purity were used in immunoblot in Figure 1D. (Graph represents 1 of 3 experiments).
Supplemental Figure 2: UPLC-MS/MS of Poly A+ RNA from HEK-293T cells cultured in Normoxic or 24 hours Hypoxic conditions. Values represent the amount of the modification divided by total parent levels (N of 2). *P ≤ 0.05 by Paired Student’s t-test. Error bars represent Standard error of the mean (SEM).
Supplemental Figure 3: UPLC-MS/MS of Total RNA from HEK-293T cells. Values represent the amount of the modification divided by total parent levels (N of 2). *P ≤ 0.05 by Paired Student’s t-test. Error bars represent Standard error of the mean (SEM).
Supplemental Figure 4: UPLC-MS/MS of Ribosomal RNA from HEK-293T cells. Values represent the amount of the modification divided by total parent levels (N of 3). *P ≤ 0.05 by Paired Student’s t-test. Error bars represent Standard error of the mean (SEM).
Supplemental Figure 5:

**Transfection with METTL3 and METTL14 siRNAs decrease M6A levels.**

HEK-293Ts transfected with METTL3 and 14 siRNAs (KD) or negative control siRNA (Neg) for 72 hours and 24 hours of normoxic or hypoxic exposure followed by MeRIP and qRT-PCR quantitation shows decreased M6A levels in mRNA targets after Mettl3/14 knockdown.

(N of 1)
Supplemental Figure 6

A. 

B. 

C. 

D. 

E. 

F.
Supplemental Figure 6: Polysome Profile traces.

(A-D) Traces of Polysome Profiles from Figure 4. (A), Normoxia, (B), Normoxia M3/14 siRNA, (C), Hypoxia, (D) Hypoxia M3/14 siRNA. (E-F) Traces of Polysome Profiles from Supplemental Figure 6. (E) Normoxia, (F), Hypoxia. (Traces represent 1 of 3 experiments).
Supplemental Figure 7: Reoxygenation Polysome Profile traces.

(A-F) Traces of Polysome Profiles from Figure 5. (A), 30min reoxygenation negative siRNA, (B), 30min reoxygenation m3/14 siRNA, (C), 1hr reoxygenation negative siRNA, (D), 1hr reoxygenation m3/14 siRNA, (E), 4hr reoxygenation negative siRNA, (F), 4hr reoxygenation m3/14 siRNA. (Traces represent 1 of 3 experiments).
Supplemental Figure 8. Mettl3/14 Knockdown decreased the ability of messages to recover from hypoxic stress after 4 hours re-oxygenation.

HEK-293T cells harvested after 72 hours transfection with Mettl3/14 (M3/14 KD) or negative control (Neg) siRNA and 24 hours of hypoxic conditions and either 30 minutes, 1 hour or 4 hours of room level re-oxygenation recovery. Polysome Profiling of extracts separated by differential centrifugation followed by sucrose gradients. qRT-PCR analysis followed by binning into estimated non-polysome (1,2,3), light polysome (4,5,6), moderate polysome (7,8,9), and heavy polysome (10,11,12) bound fractions show percentage of individual mRNA in each bin. Error bars represent SEM of 3 experiments in the no re-oxygenation, 1 and 4 hour re-oxygenation experiments and SEM of 2 experiments in the 30 minute re-oxygenation experiment. N of 1 Myc hypoxia experiment is shown. *P ≤ 0.05 by Paired Student’s t-test compared to binned negative control siRNA.
Supplemental Figure 9: EDTA-resistant RNP particles are observed for several hypoxia-modified mRNAs.

Polysome Profiling following addition of 25mM EDTA added to the lysis buffer. qRT-PCR analysis of the fractions shows percentage of the specific mRNA in each fraction. Fraction containing 80S peak is marked.

Compare to Figure 3.
Supplemental Figure 10: Mettl3/14 Knockdown (m3/14 KD) had no significant effect on cell proliferation in HEK-293T cells grown in Normoxic (Normox) or 24 hour Hypoxic (Hypox) conditions when compared to negative control siRNA (Neg). (N of 3)
### Supplemental Table 1

#### Primer List

| Gene     | Forward (5'-3') | Reverse (5'-3') |
|----------|----------------|-----------------|
| 18s rRNA | CTGAGAAACGGCTACCACATC | GCCTCGAAAGAGTCCTGTATTG |
| 28s rRNA | GGGTGGTAAACTCCTTAAGG | GCCCTCTTTGAACCTCTCTCTTC |
| 5.8s rRNA | CTCGTGCCTCGATGAAGAA | TCGAAGTGTCGATGATCAATGT |
| 5s rRNA  | CGTCTGATCTCGGAAGCTAAG | CCTACAGCAACCGGTATTC |
| β2M      | AGATGTCTCGCTTCGTGGCTTA | TGTCGGATGGATGAAACCCAGACA |
| Dusp1    | CAACCACAAGGCAGACATCA | CAGTGGGCAAAACACCCCTTCC |
| eEF1A1   | CGGTCTCAGAACTGGTGTGTTTC | AAACCAAGTGTTGCCACAA |
| GapDH    | AAGGTCAAAGATCCGGATTGCTGTT | AGCCTTGACCGTGCCATGGAATTT |
| Glut1    | TATCGTCACACGGGCTTCCTGCT | CACAAAGCCCAAGATGGCCACGAT |
| Hes1     | GAAGGCGGACATTCTGGAAAT | GTCACTCCGTTCTGCACTC |
| HuR      | CCTGTTCAGCAGCATTGGTGAAGT | TCCAGCGTGTTGATCGCTCTCTCT |
| Jun      | TTCTATGACGATGCCCTCAAC | TCAGGGTCACTGCTCTGTTTC |
| METTL3   | AGCCTTTGCAACCAACGTCC | CCGACCTCAGACAGCAAA |
| Myc      | TCCTCGGATTCTCTGCTCTCTCT | AGAAGGGTACAGACTCTGACCT |
| VHL      | TCTCTCAATTTGACCGACAGCCT | GGTCTTTTCTGCACATTGGGTG |

### Supplemental Table 2

**siRNA’s**

| siRNA          | Target Sequence                      |
|----------------|--------------------------------------|
| siMETTL3       | CTGCAAGTATGTTCACTATGA                |
| siMETTL14      | AAGGATGAGTTAATAGCTAAA                |
| Negative Control #1 siRNA | Ambion cat. # AM4635               |
## Supplemental Table 3

### Antibodies

| Antibody                  | Catalogue # | Vendor                  | WB Dilution | IP Amount |
|---------------------------|-------------|-------------------------|-------------|-----------|
| c-Jun                     | MA5-15119   | Thermo Fisher           | 1/100       |           |
| c-Myc                     | NBP1-19671  | Novus Biologicals      | 1/1000      |           |
| GAPDH (G-9)               | sc-365062   | Santa Cruz Biotechnology| 1/100       |           |
| Glut1                     | PA5-16793   | Thermo Fisher           | 1/250       |           |
| METTL14                   | HPA038002   | Sigma Life Science      | 1/1000      |           |
| METTL3/MT-A70             | A301-567A   | Bethyl Laboratories     | 1/5000      |           |
| MKP-1 (c-19) (DUSP1)      | sc-370      | Santa Cruz Biotechnology| 1/100       |           |
| N6-methyladenosine (m6a) | MABE1006    | EMD Millipore           | 5 µg        |           |