The ubiquitin–proteasome system (UPS) of protein degradation has been evaluated in different forms of neural plasticity and memory. The role of UPS in such processes is controversial. Several results support the idea that the activation of this system in memory consolidation is necessary to overcome negative constraints for plasticity. In this case, the inhibition of the UPS during consolidation impairs memory. Similar results were reported for memory reconsolidation. However, in other cases, the inhibition of UPS had no effect on memory consolidation and reconsolidation but impeded the amnesic action of protein synthesis inhibition after retrieval. The last finding suggests a specific action of the UPS inhibitor on memory labilization. However, another interpretation is possible in terms of the synthesis/degradation balance of positive and negative elements in neural plasticity, as was found in the case of long-term potentiation. To evaluate these alternative interpretations, other reconsolidation-interfering drugs than translation inhibitors should be tested. Here we analyzed initially the UPS inhibitor effect in contextual conditioning in crabs. We found that UPS inhibition during consolidation impaired long-term memory. In contrast, UPS inhibition did not affect memory reconsolidation after contextual retrieval but, in fact, impeded memory labilization, blocking the action of drugs that do not affect directly the protein synthesis. To extend these findings to vertebrates, we performed similar experiments in contextual fear memory in mice. We found that the UPS inhibitor in hippocampus affected memory consolidation and blocked memory labilization after retrieval. These findings exclude alternative interpretations to the requirement of UPS in memory labilization and give evidence of this mechanism in both vertebrates and invertebrates.
This phenomenon, called reconsolidation, is present in different types of learning and in diverse phylogenetic lines such as mollusks, crustaceans, birds, rodents, and humans (Dudai 2006). In the case of associative learning, memory is usually reactivated by the presentation of the conditioned stimulus (CS) without the unconditioned stimulus (US). In associative contextual models, the CS presentation consists in the re-exposure of the animal to the training context. Memory reactivation by the CS can trigger two apparently competing mechanisms depending on the duration of the re-exposure: reconsolidation and extinction, as initially characterized in crabs (Pedreira and Maldonado 2003). The role of the UPS in reconsolidation is controversial. In spatial memory in mice, the inhibition of proteasome-dependent protein degradation in hippocampus during reconsolidation impaired subsequent long-term memory (Artinian et al. 2008). In contrast, no effect was observed for fear conditioning upon inhibition in either hippocampus (Lee et al. 2008) or amygdala (Jarome et al. 2011), or for cocaine-induced conditioned place preference in the nucleus accumbens (Ren et al. 2013). Similar results were reported in sensitization and facilitation in Aplysia (Lee et al. 2012). Interestingly, however, inhibition of proteasome-dependent protein degradation blocked the amnesic action of protein synthesis inhibitors (Lee et al. 2008, 2012; Jarome et al. 2011; Ren et al. 2013). These findings were interpreted as a specific action of the UPS inhibitor on memory labilization, suggesting that the translation blocker was no longer amnesic because memory could not become labile after retrieval. However, another interpretation is possible in terms of the balance between synthesis and degradation of positive and negative elements in neural plasticity, as was found in the case of LTP (Fonseca et al. 2006). In this work, the pharmacological inhibition of either UPS or protein synthesis disrupted the expression of the late phase of LTP. However, when protein degradation and protein synthesis were inhibited at the same time, LTP was no affected. These findings suggest that LTP, and perhaps other forms of plasticity and memory, may require the combined action of degradation and synthesis of plasticity proteins. It is therefore conceivable that the balance of degradation and synthesis of some key proteins could be a crucial factor regulating memory re-stabilization in reconsolidation. The results obtained in fear conditioning, conditioned place preference and in Aplysia sensitization, in which the simultaneous inhibition of both UPS and translation caused no effect in reconsolidation could also be attributable to the interpretation of a balance between synthesis and degradation. In these cases, protein degradation may not be critical for reconsolidation but when protein degradation is inhibited, new protein synthesis is no longer required because the preexistent proteins are enough to sustain the plasticity process.

To evaluate these alternative interpretations other reconsolidation-interfering drugs than translation inhibitors should be tested. If UPS inhibitors are able to block memory impairment or enhancement induced by such drugs, then the role of protein degradation in memory labilization could be confirmed beyond its effect on the synthesis/degradation balance. Here we used a contextual conditioning task in crabs to analyze the effect of UPS inhibition in combination with the memory-disrupting drugs MK801 and sulfasalazine, and with the memory-enhancing drug bicuculline. To extend these findings to vertebrates, we performed similar experiments with sulfasalazine as memory-disrupting drug and the calcineurin (CaN) inhibitor FK506 as memory-enhancing drug in another example of contextual conditioning, the contextual fear memory in mice.

**Results**

Inhibition of the UPS during consolidation impairs long-term memory in crab contextual conditioning

As outlined above, the results available in the literature about the action of UPS inhibition during memory consolidation are contradictory, with most studies reporting memory impairment, whereas in two cases no effect was found (Lee et al. 2008, 2012). To further explore this issue, we initially studied the effect of UPS inhibition on long-term memory in contextual conditioning of the crab Chasmagnathus. In this task, a context (conditioned stimulus, CS) was presented with a coexisting unconditioned stimulus (US), a visual danger stimulus (Fustiñana et al. 2013). We administered the proteasome inhibitor MG132 at a 2.38 \( \mu \text{g}/\text{animal dose or dimethyl sulfoxide as vehicle (VEH) systemically before training (15-trials) to either trained animals (TR group) or control animals that did not receive the danger stimulus (CT group). The experiment thus had four groups: CT-VEH, CT-MG132, TR-VEH, and TR-MG132. Twenty-four hours later we tested these groups using a single CS–US presentation (Fig. 1A). These
data show that the TR group injected with VEH had significantly lower levels of escape responses compared with the CT-VEH group. In contrast, the TR-MG132 level of response was similar to the CT-MG132 group, indicating that the drug caused a memory impairment ($F_{(3,116)} = 3.35; P < 0.05$; CT-VEH vs. TR-VEH: $P < 0.05$ and CT-MG132 vs. TR-MG132: $P$ NS) (Fig. 1B). Together, these results suggest that proteasome inhibition disrupts consolidation of contextual memory in the crab model.

**Inhibition of the UPS during reconsolidation does not impair long-term memory**

In the following experiment we evaluated the effect of a UPS inhibitor during memory reconsolidation. In contextual conditioning of *Chasmagnathus*, memory retrieval is induced by a brief re-exposure of the animals to the CS without US presentation (Fustiñana et al. 2013). The retrieval session consisted of 27 sec of re-exposure to the CS with 5 min of total re-exposure to the experimental device. Under these conditions, memory is labilized and the reconsolidation process is induced (Fustiñana et al. 2013). We trained two pairs of TR and CT groups, and 24 h later one CT–TR pair was injected with MG132 at the same dose used in the previous experiment and the other CT–TR pair was injected with vehicle (four groups: CT-VEH, TR-VEH, CT-MG132, and TR-MG132). Twenty minutes after injection all crabs received a retrieval session and 24 h later they were tested with one CS–US presentation (Fig. 1C). We observed a significant difference between the two CT groups and their matched TR groups ($F_{(3,156)} = 4.13$; $P < 0.05$; CT-VEH vs. TR-VEH: $P < 0.05$ and CT-MG132 vs. TR-MG132: $P < 0.05$) (Fig. 1D). These results indicated that the MG132 administration, at the same dose which caused memory impairment during consolidation in the previous experiment, had no effect on memory reconsolidation.

**Inhibition of the UPS during reconsolidation blocks the amnesic effect of sulfasalazine**

Inhibition of the UPS has been reported to rescue memory impairment induced by inhibition of translation during reconsolidation (Lee et al. 2008, 2012; Jarome et al. 2011; Ren et al. 2013; see above). To test whether this action extends to reconsolidation inhibitors interfering a different molecular mechanism, we focused our attention on the transcription factor nuclear factor kappa B (NF-κB). The activation and translocation to the nucleus of NF-κB is an important regulatory mechanism of gene expression during memory formation in both vertebrates and invertebrates (for review, see Meffert and Baltimore 2005; Romano et al. 2006b; Alberini 2009). This transcription factor is also involved in memory reconsolidation (Merlo et al. 2005; Boccia et al. 2007; Lubin and Sweatt 2007; de la Fuente et al. 2011; Si et al. 2012). A key mechanism in the NF-κB pathway is the activation of IκB kinase (IKK). Upon its activation, IκK phosphorylates IκB, a regulatory protein that inhibits NF-κB. Once phosphorylated, IκB is degraded by UPS allowing NF-κB nuclear translocation (Roff et al. 1996). Sulfasalazine is a potent and specific inhibitor of IKK (Wahl et al. 1998) that interferes with the ATP-binding site of the kinase (Weber et al. 2000; Merlo et al. 2002). This drug was extensively used for memory disruption in both consolidation and reconsolidation (Merlo et al. 2002, 2005; Freudenthal et al. 2005; Boccia et al. 2007; Lubin and Sweatt 2007; Si et al. 2012). Recently, we found that sulfasalazine impaired crab contextual memory specifically when it was reactivated by presentation of the training context without reinforcement (Fustiñana et al. 2013). To test whether the proteasome inhibitor MG132 is able to block this memory impairment of sulfasalazine in reconsolidation, we trained three matched pairs of TR and CT groups. Twenty-four hours later one CT–TR pair was injected with sulfasalazine at a 20 μg/animal dose, one pair was injected with a cocktail of MG132 and sulfasalazine and the other CT–TR pair was injected with vehicle, yielding six groups: CT–VEH, TR–VEH, CT–SSZ, TR–SSZ, CT–MG132/SSZ, and TR–MG132/SSZ. Twenty minutes after injection all crabs received a retrieval session. These groups were tested 24 h after re-exposure (Fig. 2A). Consistent with previous results, the CT–SSZ versus TR–SSZ pair showed no significant differences in planned comparisons. In contrast, the other pair comparisons displayed a significantly lower level of response for TR groups versus CT groups (Fig. 2B; $F_{(5,174)} = 3.08$; $P < 0.05$; CT-VEH vs. TR-VEH: $P < 0.05$, CT-SSZ vs. TR-SSZ: $P$ NS and CT-MG132/SSZ vs. TR-MG132/SSZ: $P < 0.05$). These results are in agreement with previous findings in mouse and *Aplysia*, which indicated that the inhibition of UPS blocked the action of anamnesic drugs.

**Inhibition of the UPS does not interfere with memory retrieval**

The results of the previous experiment suggest that MG132 is able to block the effect of IKK/NF-κB pathway inhibition, suggesting that UPS inhibition prevents labilization of the memory. However,
alternatively it is conceivable that MG132 interferes with memory retrieval, a process that is required for memory labilization. To evaluate this possibility of retrieval interference, we performed an experiment with two pairs of CT–TR groups that were injected with MG132 or VEH 20 min before the memory test, 24 h after training (Fig. 2C). The results indicated significant differences in CT versus TR comparisons for both pairs, suggesting that MG132 did not interfere with memory retrieval (Fig. 2D). F(1,110) = 3.81; P < 0.05; CT–VEH vs. TR–VEH: P < 0.05 and CT–MG132 vs. TR–MG132: P < 0.05).
on memory reconsolidation by means of blocking the labilization process. To extend these finding to vertebrates, we performed similar experiments using contextual fear conditioning in mice. In this task, mice receive three footshocks in a particular context. Memory labilization and reconsolidation is induced by a re-exposure (5 min) to the training context without footshock 1 d after training (de la Fuente et al. 2011). After the re-exposure session, four groups of animals were bilaterally infused into hippocampus with either vehicle (VEH group), 0.5 μg/hemisphere of sulfasalazine (SSZ group), 0.05 μg/hemisphere of MG132 (MG132 group), or the sulfasalazine/MG132 cocktail (SSZ/MG132 group). On Day 2 all groups of animals showed high percentage of freezing without difference between them, revealing retention of memory \((F_{3,36} = 1.99, P = 0.13)\); VEH vs. SSZ: \(P > 0.05\); VEH vs. MG132: \(P > 0.05\); VEH vs. MG132/SSZ: \(P > 0.05\); SSZ vs. MG132: \(P > 0.05\); SSZ vs. MG132/SSZ: \(P > 0.05\); SSZ: \(P > 0.05\); data not shown. On Day 3, mice were tested by a second re-exposure of 5 min (Fig. 4A). During testing, the percentage of freezing was determined (Fig. 4B). As expected, the VEH group showed a high level of freezing, above 40%, and the SSZ group showed a significantly lower level of freezing \((F_{3,36} = 6.31, P < 0.05)\); VEH vs. SSZ: \(P < 0.05\); SSZ vs. MG132: \(P < 0.05\); SSZ vs. MG132/SSZ: \(P < 0.05\). These results indicate that inhibition of the IKK/NF-κB pathway during memory reconsolidation causes memory impairment. Conversely, the MG132 group showed high levels of freezing that were not significantly different to the VEH group levels, but significantly higher than the SSZ group, suggesting that administration of MG132 had no effect on memory. Similarly, the group SSZ/MG132 showed levels of freezing comparable with the VEH control group, indicating that UPS inhibition blocked the amnesic effect of sulfasalazine. These results are in agreement with previous observation (Lee et al. 2008), where administration of the UPS inhibitor clasto-lactacystin-β-lactone into hippocampus did not impair contextual fear memory in mice, but impeded the amnesic effect of the protein synthesis inhibitor anisomycin. The authors also found that the same dose of the UPS inhibitor had no effect on memory during consolidation. In the following experiment we administered MG132 after the training session to compare with those results utilizing the drug clasto-lactacystin-β-lactone. One group of mice was trained with the footshock (SH) and another group was exposed to the training chamber in the absence of footshocks (NS). After the training session, mice received a bilateral intra-hippocampal administration of either vehicle or MG132 in the same dose as the previous experiment (NS–VEH, SH–VHS, NS–MG132, and SH–MG132). On Day 2, all groups were tested (Fig. 4C). The non- shocked groups displayed significantly lower levels of freezing than the shocked groups, but no effects of the drug were found \((F_{3,36} = 10.45, P < 0.05)\); NS–VEH vs. SH–VEH: \(P < 0.05\); NS–MG132 vs. SH–MG132: \(P < 0.05\); SH–VEH vs. SH–MG132: \(P > 0.05\); SS–VEH vs. NS–MG132: \(P > 0.05\); NS–VEH vs. NS–MG132: \(P > 0.05\); data not shown). Next, we used the same experiment design but with a higher dose of MG132, 0.15 μg/hemisphere. In contrast to the previous experiment, we found a significantly lower level of freezing in the SH–MG132 group with respect to SH– VEH \((F_{3,36} = 16.15, P < 0.05)\); NS–VEH vs. SH–VEH: \(P < 0.05\); NS–MG132 vs. SH–MG132: \(P < 0.05\); SH–VEH vs. SH–MG132: \(P < 0.05\); NS–VEH vs. NS–MG132: \(P < 0.05\); NS–MG132 vs. NS–VEH: \(P > 0.05\) (Fig. 4D), indicating a memory impairment due to UPS inhibition during consolidation. The last two experiments support the interpretation that, in the mouse hippocampus, a higher dose of UPS inhibitor is required to impair memory during consolidation compared with the one needed to block memory labilization after retrieval.

Inhibition of the UPS during reconsolidation blocks the fear memory enhancing effect of the calcineurin inhibitor FK506

Taking into account the result described above for enhancement of memory during reconsolidation in crabs, we aimed at extending the results to mice. We focused our attention on Ca\(^{2+}\)/calmodulin-dependent phosphatase 2B (calcineurin, CaN), an important regulator of the level of protein phosphorylation in the nervous system because it is activated directly by Ca\(^{2+}\)/calmodulin signals in synaptic spines. Some studies support its role as a memory constraint in different memory tasks (Baumgartel et al. 2008; Havekes et al. 2008), and it was found to be involved in memory extinction (Lin et al. 2003; de la Fuente et al. 2011). More recently, we found evidence for the role of CaN as a negative constrain in contextual fear memory in mice, and that inhibition of CaN by the specific inhibitor FK506 in hippocampus-enhanced memory (V de la Fuente, N Federman, M Sustiñana, G Zalcman, and A Romano, in prep.). In the following experiment we used FK506 to induce memory enhancement, and to test whether UPS inhibition blocks this memory facilitation during reconsolidation. One group of mice underwent contextual fear conditioning with three footshock presentations, while another three groups
received a weak training with only one footshock. On Day 2, all groups were re-exposed to the training context for 5 min and the percentage of freezing was determined; 15 min before re-exposure we bilaterally infused into hippocampus either vehicle, 5 µg/hemisphere of FK506, 0.05 µg/hemisphere of MG132, or FK506/MG132 cocktail (FK506/MG132 group). The group that received three trials was injected with vehicle (3SH-VEH group) and was used as a positive control for memory retention at testing. One of the groups trained with one trial also received vehicle (1SH-VEH group), and the other two weakly trained groups were injected with either FK506 (1SH-FK506 group) or FK506/MG132 cocktail (1SH-FK506/MG132 group). On Day 2 only the group of animals that received three trials show high levels of freezing while the three groups that received only one trial on Day 1 elicited significantly low levels of freezing compared with the first group (F(3,36) = 6.64, P < 0.05; 1SH-VEH vs. 3SH-VEH: P < 0.05; 1SH-VEH vs. 1SH-FK506: P = NS; 1SH-VEH vs. 1SH-FK506/MG132: P = NS; 1SH-FK506/MG132 vs. 3SH-VEH: P < 0.05; 1SH-FK506/MG132 vs. 1SH-FK506: P = NS; 1SH-FK506 vs. 3SH-VEH: P < 0.05) (data not shown). On Day 3, all mice were tested with a second re-exposure of 5 min (Fig. 5A), and the percentage of freezing was determined (Fig. 5B). As expected, the 3SH-VEH group showed high levels of freezing and the 1SH-VEH group displayed significantly lower freezing levels. The 1SH-FK506 group displayed high freezing values similar to the 3SH-VEH group, but the group injected with the cocktail showed a significantly lower level of freezing (F(3,36) = 5.52, P < 0.05; 1SH-VEH vs. 3SH-VEH: P < 0.05; 1SH-VEH vs. 1SH-FK506: P < 0.05; 1SH-VEH vs. 1SH-FK506/MG132: P = NS; 1SH-FK506/MG132 vs. 3SH-VEH: P < 0.05; 1SH-FK506/MG132 vs. 1SH-FK506: P < 0.05; 1SH-FK506 vs. 3SH-VEH: P = NS). These results indicate that hippocampal administration of an inhibitor of calcineurin during reconsolidation enhances fear memory. In contrast, coadministration with an inhibitor of UPS blocked this enhancement.

**Discussion**

In this work we analyzed the role of the ubiquitin–proteasome system (UPS) in memory consolidation and reconsolidation of two examples of contextual-associative memories, the contextual conditioning of the crab *Chasmagnathus* and the contextual fear conditioning in mice. The administration of the UPS inhibitor MG132 impaired long-term memory when administered shortly before training, indicating that the UPS is necessary for memory consolidation. This effect, observed in the two models, is in line with the view that negative constrains need to be removed to activate key neural plasticity mechanisms such as the NF-κB pathway of gene expression regulation, as was evidenced previously (Lopez-Salon et al. 2001; Merlo and Romano 2008). The finding that the administration of an inhibitor of UPS in mouse hippocampus induced memory impairment in contextual fear conditioning differs from previous findings of Lee et al. (2008), who found that the UPS inhibitor was unable to induce memory impairment. The different results could be explained by the use of different drugs, clasto-lactacystin-β-lactone versus MG132. However, it is important to note that we initially found no effect of MG132 on memory consolidation in mice at a dose that was effective during reconsolidation. An increment of the dose was required to observe memory impairment during consolidation. Thus, the lack of effect of clasto-lactacystin-β-lactone in that report could also be explained by an insufficient dose to impair memory retention.

Conversely to the effect of the UPS inhibitor on memory consolidation, no effect was found during reconsolidation. A simple interpretation of these results is that, in contrast to consolidation, the UPS is no longer required during reconsolidation. However, when the UPS inhibitor is injected in combination with the amnesic drug sulfa-salazine, the expected memory impairment was not observed in either memory model. The UPS inhibitor MG132 does not have a direct action on memory reconsolidation but in memory labilization. These findings are in keeping with previous studies using inhibitors of protein synthesis as amnesic agents (Lee et al. 2008, 2012; Jarome et al. 2011; Ren et al. 2013), and support the interpretation that protein degradation is required for memory to become labile after retrieval induced by a brief re-exposure to the training context. However, another interpretation could be that the balance between synthesis and degradation of positive and negative elements governs neural plasticity, as was found in the case of LTP (Fonseca et al. 2006). In that study, it was proposed that protein degradation may be not critical for neural plasticity. Instead, if degradation is inhibited, de novo protein synthesis would no longer be required because preexisting proteins would be enough to sustain the neural plasticity process. Here we used sulfa-salazine, which does not interfere directly with the balance between protein synthesis and degradation. Our data extend the previous finding, and support a direct action of the UPS during memory labilization. In addition, our results show that the UPS inhibition not only blocked the amnesic effect of reconsolidation-disrupting drugs, but also blocked the memory enhancing effect of the reconsolidation-enhancing drugs bicuculine and FK506. These findings further support the direct and specific role of the UPS in memory labilization. In other words, labilization process is required for memory modulation after retrieval. If this process is blocked, drugs that enhance or disrupt memory during reconsolidation have no effect.

In the experiments displayed in Figure 4, we found that the dose of MG132 that was effective to block memory labilization

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**Figure 5.** Hippocampal proteasome inhibition impairs the enhancement effect of FK506 during reconsolidation fear conditioning. (A) Experimental protocol. On Day 1 one group of animals were trained with three pairings of CS–US and three groups received only one pairing of CS–US. On Day 2 all groups received a bilateral intra hippocampal injection and 15 min later were re-exposed to the training context during 5 min. The three trials group and one of trial received VEH (1E-VEH and 3E-VEH, respectively), another one trial group received FK506 (5 µg/hemisphere) and the other received a cocktail of FK506 and MG132 (FK/MG). On Day 3 the testing session was performed. The arrow indicates the moment of the injection. (B) Testing session. Percentage of freezing ± SEM on Day 3; (* P < 0.05, n = 10, per group.)
Protein degradation in contextual memory

did not affect consolidation. Instead, a three times higher dose was required to impair memory during consolidation. These data suggest that the requirement and potentially the duration of protein degradation may be different during consolidation than during labilization and a higher dose of UPS inhibitor is required to block such a process.

In the present work we used sulfasalazine, a drug that interferes with an important mechanism of neural plasticity, the IKK/NF-κB pathway. This pathway is considered critical for synaptogenesis (Boersma et al. 2011), and has a key role in different memory models in diverse species (Romano et al. 2006a). Sulfasalazine is a drug that specifically interferes with IKK activity and was used in different memory tasks (Merlo et al. 2002; Boccia et al. 2007; Lubin and Swatt 2007). In addition, we used here two drugs to produce memory enhancement. One of them, bicusculine, is a competitive antagonist of the GABA receptor type A and was previously used to enhance memory in different taxa and particularly in crabs (Carbo Tano et al. 2009). Alternatively, we administered in mice FK506, a specific inhibitor of CaN. The enhancing effect of FK506 is attributed to the role of CaN as a negative element in consolidation and reconsolidation (Baumgärtel et al. 2008; Havekes et al. 2008; V de la Fuente, N Federman, MS Fustiñana, G Zalcman, and A Romano, in prep.). Moreover, this enhancing effect is specific for reconsolidation, because nonre-exposed animals injected with FK506 did not show memory improvement (V de la Fuente, N Federman, MS Fustiñana, G Zalcman, and A Romano, in prep.). However, this protein phosphatase plays an important role as a positive element in the formation of fear extinction memory (de la Fuente et al. 2011).

The ubiquitin–proteasome mechanism of protein degradation is evolutionarily conserved. The use of two models of associative contextual memory in different phyla allowed us to determine the role of UPS in memory consolidation and reconsolidation. In conclusion, the UPS plays a similar role with similar kinetics although the nervous system organization is completely different in these two species. Our results, together with the previous evidence, support the notion that during consolidation the degradation of proteins which act as negative elements is important for the neural plasticity required for memory storage. During reconsolidation, the degradation of proteins is also critical to allow the process of labilization in which the stored memory becomes susceptible to modification on the basis of new experience.

Materials and Methods

Animals

Crabs
Adult male Chasmagnathus granulatus (recently renamed as Neohelice granulata), Crustacea, Grapsidae, inter-tidal crabs, 2.6–2.9 cm across the carapace, weight 17 ± 0.2 g (n = 60), were collected from water <1 m deep in the estuarine coasts of San Clemente del Tuyú, Argentina, and transported to the laboratory where they were lodged in plastic tanks (30 × 45 × 20 cm) filled to 0.5 cm depth with diluted (12‰, pH 7.4–7.6) marine water (prepared from Cristalsea Marinemix salts) to a density of 20 crabs per tank. The holding room was maintained on a 12-h light–dark cycle (light on 07:00–19:00 h). Temperature of both holding and experimental rooms was maintained within a range of 22°C–24°C. Experiments were performed between the third and the tenth day after the animals’ arrival. Each crab was used in only one experiment. Experiments were performed in accordance with the local regulations for the care and use of laboratory animals. All efforts were made to minimize animal suffering and to reduce the number of animals used. Furthermore, all the groups included the same number of animals in each experiment, 30 crabs.

Consequently, considering that each experiment included four groups, the number of crabs used was 120.

Mice
Balb C male mice, 60–70 d of age, weighing 25–30 g, were used (La Plata University Animal Facilities, La Plata, Argentina). The animals were individually caged and singly housed throughout the experimental procedures, with water and food ad libitum, under a 12:12 h light/dark cycle (lights on at 07:00 am) at an ambient temperature of 21°C–22°C. Experiments were performed in accordance with local regulations and the National Institutes of Health (NIH) Guide for the Care and Use of Laboratory Animals (NIH publication 80-23/96). All efforts were made to minimize animal suffering and to reduce the number of animals used.

Experimental devices and behavioral procedures

Crabs
The experimental device has been described in detail elsewhere (Lozada et al. 1990). Briefly, the experimental unit was a bowl-shaped opaque container surrounded by a steep concave wall 12 cm high (23-cm top diameter and 9-cm floor diameter). The container was filled with marine water to a depth of 0.5 cm. The crab was placed in the container, which was suspended from an upper wooden framework (23 × 23 × 30 cm) by three strings. A motor-operated screen (US, an opaque rectangular strip of 25.0 × 7.5 cm) was moved horizontally over the animal from left to right, and vice versa. The screen’s movements were cyclical. The screen displacements provoked the escape response of the crab and subsequent container vibrations. Each trial lasted 9 sec and consisted of two successive cycles of movement. Four microphones were attached to the center of the outside base of the container. The microphones recorded the vibrations that were produced by the animal’s response. These signals were amplified, integrated during the entire trial (9 sec), and translated into arbitrary numerical units ranging from zero to 8000. During the experiment, the crabs were illuminated using a 5-W bulb placed either above or below the container (Perez-Cuesta et al. 2007). A computer was used to program the trial sequences, trial illumination, trial duration, and inter-trial intervals and to monitor the experimental events. The experimental room contained 40 experimental devices that were separated from each other by partitions.

The training and other treatment sessions were preceded by 10 min of adaptation to the experimental device, which was illuminated from below throughout. A standard CSM training session consisted of 15 US presentations without light shift (above illumination) with an inter-trial interval (ITI) of 171 sec. A standard contextual Pavlovian conditioning (CPC) training session consisted of 15 trials. Each trial lasted 27 sec with above illumination (CS), and the US was presented during the last 9 sec. The ITI between US presentations was 171 sec, as in the standard CSM protocol, but the ITI between CS presentations was 153 sec. During the ITI, the experimental unit was illuminated from below, which provoked a virtual change in the environmental features. The untrained animals were kept in the experimental unit during the entire training session. These animals were not presented with the US, but were presented with the same pattern of light shift. Immediately after each session, the crabs were moved from the experimental unit to individual resting containers, which were plastic boxes that were filled with water to a depth of 0.5 cm. The resting containers were kept inside dimly lit drawers. One trial of the US was presented before the training to measure the responsiveness of each animal. No differences were found between groups in this pre-training trial for any of the experiments (Fustiñana et al. 2013).

Mice
The conditioning chamber (context A) was made of transparent acrylic (24.5 × 24.5 × 42 cm) in a wooden box with a clear front
Surgery and injections

Crabs

Fifty microliters of the vehicle (VEH) or drug solution was injected through the right side of the dorsal cephalothoracic-abdominal membrane by a syringe that was fitted with a sleeve to control the depth of penetration to 4 mm, thus ensuring that the injected solution was released into the pericardial sac.

Mice

Mice were implanted under deep anesthesia (ketamine and xylazine) with a 23-gauge guide cannulae 1 mm dorsally to the dorsal hippocampus, at coordinates of anterior, zine) with a 23-gauge guide cannulae 1 mm dorsally to the dorsal hippocampus, at coordinates of anterior, 1.2, in accordance with the atlas of Paxinos and Franklin (2001). Guide cannulae were fixed to the skull with dental acrylic. Experiments were performed after animal recovery and injections were administered immediately before or after context re-exposure (see Results) without anesthesia. The injection device consisted of a 30-gauge cannula connected to a 5 μL Hamilton syringe with tubing. Initially, the infusion device was filled with distilled water and a small air bubble was sucked into the injection cannula, followed by the injection solution. The air bubble allowed for visual inspection of the injection progress. The injection cannula was inserted into the guide cannula with its tip extending beyond the guide by 1 mm to reach the dorsal hippocampus. The injections were administered during 30 sec and operated by hand. The injection cannula was removed after 60 sec to avoid reflux and to allow the diffusion of drugs. The volume of each intra-hippocampal infusion was 0.5 μL/hemisphere. Different injection devices were used for drug and vehicle. After behavioral procedures, the animals were injected with black ink and were decapitated. The brains were placed in 4% paraformaldehyde for 1 d followed by 30% sucrose for an additional 24 h. To verify cannulae placement, frozen brains were sliced using a cryostat and analyzed with a magnifying glass. The deepest position of the needle was superimposed on serial coronal maps. Only data from animals with cannulae located in the intended sites were included in the analysis.

Protein degradation in contextual memory

Drugs

The following drugs were used: sulfasalazine (SSZ) [2,4-hydroxy-((4-(2-pyridinylamino)-sulphonyl)-phenyl)-azo]benzoic acid] (Sigma-Aldrich, St. Louis, MO, USA), which is an NF-κb inhibitor, was freshly dissolved at a final dose of 1.2 μg/g of animal (5 mM) in crab (Merlo et al. 2002) and 0.5 μg per hemisphere (2.5 mM) in mice (Boccia et al. 2007). The proteasome inhibitor MG132 (N-CBZ-LEU-LEU-LEU-AL) (Sigma-Aldrich) was injected at a final dose of 2.69 μg/g of animal and MK801 [(+)-5-methyl-10,11-dihydro-5H-indenzo[α,a][cyclohepten-5,10-imine] (Sigma-Aldrich) at a final dose of 0.6 μg/g of animal. In mice we also used (3S,4R,5S,8R,9E,12S,14S,15R,16S,18R,19R,20S)-6, 5, 11, 12, 13, 14, 15, 16, 17, 18, 19, 24, 25, 26α-hexadecahydro-5,19-dihydroxy-3-[((1E)-2-[(1R,3R,4R)-4-hydroxy-3-methoxyxyclohexyl]-1-methylphenyl]-14,16-dimethoxy-4,10,12,18-tetramethyl-8-(2-propan-1-yl)-15, 19-epoxy-3H-pyrido[2,1-c][1,4]oxaazacyclotricoso-1,7,20,21(4H,23H)-tetrone, monohydrate [FK-506 (FK)] (Sigma-Aldrich), an inhibitor of the calcineurin phosphatase (CaN) administered at a final dose of 5 μg/hemisphere (10 μg/μL) (V de la Fuente, N Federman, MS Fustiñana, G Zalcman, and A Romano, in prep.). All the drugs were diluted in dimethyl sulfoxide (DMSO) which was used as VEH.

Data analysis and drug effect evaluation

In the crab CPC, retention of the learning acquired during training was considered to have occurred when a significantly lower level of response in the testing session was found for the trained group compared with its control group (i.e., both groups were injected with the same solution or treated with the same behavioral manipulation). The rationale for this criterion is based on the previous experiments performed in our laboratory. In these experiments, a significant difference (t-test, α = 0.05) between the trained (TR) and untrained (UT) groups was invariably identified at testing sessions that took place 24 h or more after training. The experiments demonstrating this difference included 15 or more training trials with 171-sec inter-trial intervals. Accordingly, for the current experiment, a significant difference was predicted at testing between the UT and TR groups. Therefore, throughout the current paper, the results from the behavioral study were analyzed using a priori planned comparisons via ANOVA with p (per comparison error rate) = 0.05, according to the standard method (Howell 1987). A lack of difference between the UT and TR groups was assumed to indicate a lack of memory retention. The statistical analysis of the test data included a set of three a priori planned comparisons, namely each pair of UT–TR groups and the comparison between the two UT groups, using planned comparisons of least squares means with p (per comparison error rate) < 0.05 (Rosenthal and Rosnow 1985; Howell 1987). A comparison between the control groups that received different treatments was necessary to determine the possible drug or behavioral manipulation side effects that may have affected the response level at testing in a manner that was unrelated to training experience. In all the experiments of the current paper, no differences were found in this comparison. For each experiment, a prediction was made based on the experimental design. In general, we could predict that, in the first comparison, a difference between the UT and TR groups was expected due to the reduction in the response level that was induced by training in the latter group (e.g., this group received a strong training protocol without any other treatment). Conversely, in the second comparison, if the drug impaired retention, then no differences were expected between the UT–TR pair. Finally, as long as the behavioral manipulation or the drug did not affect the level of response at testing, then no difference was expected in the comparison between control groups. All of the values were represented as the normalized mean ± the standard error with respect to the main control group (100%, e.g., CPC, P, or VEH).
In mice, behavioral data were analyzed by two-way ANOVA, followed by Newman–Keuls multiple comparisons post hoc test, comparing the values of the percentages of freezing.

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