**ABSTRACT**

**Objective:** To develop and evaluate Transdermal patch of Maslinic acid for Transdermal drug delivery. The current study is to develop Transdermal drug delivery system.

**Methods:** Suitable method such as Solvent Casting Technique of Film Casting Technique are used for preparation of Transdermal patch.

**Results:** Various formulations were developed by using hydrophilic and hydrophobic polymers like HPMC E5 and EC respectively in single and combinations by solvent evaporation technique with the incorporation of penetration enhancer such as dimethylsulfoxide and dibutyl phthalate as plasticizer. In vitro studies concluded that HPMC E5 patches has better release than that of EC patches, which may be attributed to high water vapour permeability of HPMC patches and hydrophobic nature of EC. An attempt was made to incorporate HPMC E5 and EC to the monolithic system for better release and prolong the duration of release. Formulation F7 containing an equal ratio of HPMC E5: EC (5:5) showed maximum and sustained release of 86.816±0.264 within 24 h. Kinetic models were used to confirm the release mechanism of the formulations. Maslinic acid release from the patches F1 to F7 followed non-Fickian diffusion rate controlled mechanism.

**Keywords:** Controlled DDS, Transdermal DDS, Maslinic acid, Transdermal patch, Solvent evaporation method

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**INTRODUCTION**

A transdermal patch is a medicated adhesive patch that is placed on the skin to deliver a specific dose of medication through the skin and into the bloodstream. Often, this promotes healing to an injured area of the body. An advantage of a transdermal drug delivery route over other types of medication delivery such as oral, topical, intravenous, intramuscular, etc. is that the patch provides a controlled release of the medication into the patient, usually through either a porous membrane covering a reservoir of medication or through body heat melting thin layers of medication embedded in the adhesive. The main disadvantage to transdermal delivery systems stems from the fact that the skin is a very effective barrier; as a result, only medications whose molecules are small enough to penetrate the skin can be delivered in this method. A wide variety of pharmaceuticals are now available in transdermal patch form [1].

Administration of drug in conventional dosage form requires-large dose, frequent administration and lacks extended duration, with chances of toxicity. While in controlled drug delivery devices, there is efficient utilization of the drug desired extended duration, with very low chances of toxicity, facilitating enhanced complication of the patient, leading to better management of therapeutics. The efficacious use of drugs influences cost factor and the economy of therapy too. It seems that controlled delivery should be the goal for all products and now a day’s drug firms have been allocating large resources on the reformulation of older, existing drugs in sustained and controlled drug delivery often resulting in special economic gains [2].

A novel drug delivery approach known as a controlled release drug delivery system evolves, which facilitates the drug release into systemic circulation at a pre-determined rate [3]. A class of novel drug delivery systems is Transdermal drug delivery systems (TDDS) which can deliver medicines via the skin portal to systemic circulation at a predetermined rate and maintain clinically effective concentrations over a prolonged period of time [4, 5].

**Maslinic acid**

Maslinic acid, a naturally occurring dihydroxy triterpenoid having a 6-6-6 fused pentacyclic structure, is extractable from the fruits of olive (Oliva europaea). Maslinic acid is a compound derived from dry olive pomace oil (an olive skin wax) which is a byproduct of olive oil extraction. It is a member of the group of triterpenes known as oleananes. Maslinic acid is a pentacyclic triterpene found in a variety of natural sources, ranging from herbal remedies used in traditional Asian medicine to edible vegetables and fruits present in the Mediterranean diet. In recent years, several studies have proved that maslinic acid exerts a wide range of biological activities, i.e., antitumor, antiangiogenic, antioxidant, cardioprotective, neuroprotective, antiparasitic and growth-stimulating. Experimental models used for the assessment of maslinic acid effects include established cell lines, which have been often used to elucidate the underlying mechanisms of action, and also animal models of different disorders, which have confirmed the effects of the triterpene in vivo. Overall, and supported by the lack of adverse effects in mice, the results provide evidence of the potential of maslinic acid as a nutraceutical, not only for health promotion but also as a therapeutic adjuvant in the treatment of several disorders [6, 7].

**Methods**

**Pre-formulation studies**

**Determination of melting point**

Melting point of the drug was determined by taking a small amount of drug in a capillary tube closed at one end. The capillary tube was placed in a melting point apparatus and the temperature at which drug melts was recorded. This was performed thrice and an average value was noted [8].

**Determination of solubility**

An excess amount of the drug was taken and dissolved in a measured volume of distilled water in a volumetric flask to get a saturated solution. The solution was kept for 24 h at room temperature for the attainment of equilibrium. These solutions were kept for sonication and then supernatant were filtered using a 0.45-μm Whatman filter paper to separate the undisolved drug particles and diluted suitably and the concentration of Maslinic acid...
in the filtrate was determined spectrophotometrically by measuring at 300 nm.

**Determination of partition coefficient**

The partition coefficient of the drug was determined by taking equal volumes of 1-octanol and aqueous solution in a separating funnel. In case of water-soluble drugs, a drug solution was prepared in distilled water, and in the case of water-insoluble drugs, a drug solution of was prepared in 1-octanol. Standard solution of the drug was prepared in this phosphate buffer pH 7.4 solution.

Octanol (10 ml) was added to an equal volume of this standard drug solution in a separating funnel and was kept for 24 h at 37± ℃ with intermittent shaking. Finally, the buffer solution was separated, clarified by centrifugation and assayed for drug content.

**Determination of drug-excipients compatibility**

**FT-IR**

In the preparation of film formulation, drug and polymer may interact as they are in close contact with each other, which could lead to the instability of drug. Preformulation studies regarding the drug-polymer interaction are therefore very critical in selecting appropriate polymers. FT-IR spectroscopy was employed to ascertain the compatibility between Maslinic acid and the selected polymers. The pure drug and drug with excipients were scanned separately.

**Procedure**

Potassium bromide was mixed with drug and/or polymer and the spectra were taken. FT-IR spectrum of Maslinic acid was compared with FT-IR spectra of Maslinic acid with polymer. Disappearance of Maslinic acid peaks or shifting of the peak in any of the spectra was studied [10].

**Procurement of standard drug**

Maslinic acid was procured from Sigma Aldrich (Merck).

**Characterization of maslinic acid**

**DSC of maslinic acid**

Purity profile of the drug was determined by using differential scanning calorimetry (DSC). The latter can be assessed by the melting behavior observed in the recorded thermogram. The main application of DSC to calorimetry (DSC). The latter can be assessed by the melting behavior of the drug. The melting temperature is a strong indication of drug purity. For carrying out DSC of the model drug, 0.5 mg of sample was placed in aluminum pan. The pan was crimped using a punching machine and weighed. The moisture loss was calculated using the formula:

\[
\text{Percentage moisture absorption} = \frac{\text{Final Weight} - \text{Initial Weight}}{\text{Initial Weight}} \times 100
\]

**Calibration curve of maslinic acid**

The standard calibration curve was constructed to obtain a regression line equation to be used for finding out the concentration of drug in samples. Two calibration curves of the drug were plotted; one by RP-HPLC method and one by UV spectrophotometry. Calibration curve by RP-HPLC method was used for assay of drug in gel matrix for entrapment efficiency studies. The other one was plotted by UV spectrophotometer using Ethanolic phosphate buffer (pH 7.4) for carrying out in vitro drug release studies [12].

**Standard solution of drug**

To make a stock solution, 10 mg of Maslinic acid was weighed and transferred to 100 ml volumetric flask. Volume was made up to the mark with mobile phase to obtain a solution of 100 µg/ml. It was further diluted to obtain a secondary stock solution of 10 µg/ml. Appropriate aliquots of secondary stock solution of 10 µg/ml were taken in 10 ml volumetric flasks and diluted up to the mark with mobile phase to obtain final concentrations of 0.5, 2.0, 3.0, 4.0 and 10.0 µg/ml of the model drug.

**Preparation of standard solution**

To make a stock solution, accurately weighed 1.0 mg of Maslinic acid was dissolved in Ethanolic phosphate buffer (7.4) in 10 ml volumetric flask and volume were made up to the mark with Ethanolic phosphate buffer (7.4) to give a clear solution of 1000 µg/ml concentration. It was further diluted to obtain a secondary stock solution of 100 µg/ml. A series of different concentrations of drug were prepared from secondary stock solution i.e. 0.5, 1.0, 1.5, 2.0, 2.5 and 5.0 ml solutions were pipetted out from secondary stock solution and were transferred in to 10 ml volumetric flasks to obtain a solution of 5, 10, 15, 20, 25, 50 µg/ml respectively on making up the solution to 10 ml with Ethanolic phosphate buffer (7.4).

**Preparation of drug-phosphatidylcholine complex (D-PC)**

The complex was prepared with drug and phospholipids in a suitable molar ratio. The required amount of drug and phospholipids were put in a 250 ml round bottom flask and 20 ml reaction solvent was added. The mixture was refluxed at a suitable temperature for the required time period. To the concentrate, n-hexane was added to precipitate D-PC and filtered and dried to remove traces of solvents. The dried D-PC obtained was weighed using electronic balance and stored at room temperature for further use [13].

**Preparation of transdermal patches**

Transdermal patches containing Maslinic acid were prepared by the solvent evaporation technique in cylindrical glass molds with both sides open. The backing membrane was cast by pouring a 2 % (w/v) polynvinyl alcohol (PVA) solution followed by drying at 60 ℃ for 6 h. The drug reservoir was prepared by dissolving D-PC in Chloroform: Methanol (1:1) mixture. Dibutylphthalate 15 % (w/w of dry polymer composition) was used as a plasticizer. The drug 0.5 mg (in 5 ml solvent mixture Chloroform: Methanol) was added into the homogeneous dispersion under slow stirring with a magnetic stirrer. The uniform dispersion was cast on a PVA backing membrane and dried at room temperature. The films were stored between sheets of wax paper in desiccators [13].

**Evaluation of transdermal patches [14-17]**

**A. Physical appearance**

All the prepared patches were visually inspected for color, clarity, flexibility and smoothness.

**B. Thickness uniformity**

The aim of the present study was to check the uniformity of thickness of the formulated films. The thickness of the film was measured at 3 different points using a digital caliper and an average thickness of three readings was calculated.

**C. Weight uniformity**

For each formulation, three randomly selected patches were used. For the weight variation test, 3 films from each batch were weighed individually and the average weight was calculated.

**D. Folding endurance**

The folding endurance was measured manually for the prepared films. A strip of film (5 x 5 cm) was cut and repeatedly folded at the same place till it broke. The number of times the film could be folded at the same place without breaking/cracking gave the value of folding endurance.

**E. Percentage moisture absorption**

The films were weighed accurately and placed in the desiccators containing 100 ml of a saturated solution of potassium chloride, which maintains 80-90% RH. After 3 d, the films were taken out and weighed. The study was performed at room temperature. The percentage moisture absorption was calculated using the formula:

\[
\text{Percentage moisture absorption} = \frac{\text{Final Weight} - \text{Initial Weight}}{\text{Initial Weight}} \times 100
\]

**F. Percentage moisture loss**

The films were weighed accurately and kept in a desiccators containing anhydrous calcium chloride. After 3 d, the films were taken out and weighed. The moisture loss was calculated using the formula:
Percentage moisture loss = \( \frac{\text{Final Weight} - \text{Initial Weight}}{\text{Initial Weight}} \times 100 \)

G. Water vapors transmission rate

Glass vials of 5 ml capacity were washed thoroughly and dried to a constant weight in an oven. About 1 gm of fused calcium chloride was taken in the vials and the polymer films of 1.44 cm\(^2\) were fixed over the brim with the help of an adhesive tape. Then the vials were weighed and stored in a humidity chamber of 80-90 % RH condition for a period of 24 h. The vials were removed and weighed at the time interval of 24 h for three consecutive days to note down the weight gain.

Water vapor transmission rate = \( \frac{\text{Final Weight} - \text{Initial Weight}}{\text{Time} \times \text{Area}} \times 100 \)

H. Tensile strength

Tensile strength of the film was determined with a Universal strength testing machine (Hounsfield, Slinfold, Horsham, U. K.). The sensitivity of the machine was 1 gram. It consisted of two load cell grips. The lower one was fixed and the upper one was movable. The test film of size (4 × 1 cm\(^2\)) was fixed between these cell grips and force was gradually applied till the film broke. The tensile strength of the film was taken directly from the dial reading in kg. Tensile strength is expressed as follows;

\[ \text{Tensile strength} = \text{Tensile load at Break} / \text{Cross-Sectional Area} \]

I. Drug content uniformity of films

The patches (1 cm\(^2\)) were cut and added to a beaker containing 100 ml of phosphate-buffered saline of pH 7.4. The medium was stirred with magnetic bead. The contents were filtered using whatmann filter paper and the filtrate was examined for the drug content against the reference solution consisting of placebo films (containing no drug) at 215 nm spectrophotometrically. The experiment was repeated to validate the result.

J. In vitro drug release studies

In vitro skin permeation studies were performed by using a modified Franz diffusion cell with a receptor compartment capacity of 20 ml. The synthetic cellophane membrane was mounted between the donor and receptor compartment of the diffusion cell. The formulated patches were cut into the size of 1 cm\(^2\) and placed over the drug release membrane and the receptor compartment of the diffusion cell was filled with phosphate buffer pH 7.4. The whole assembly was fixed on a magnetic stirrer, and the solution in the receptor compartment was constantly and continuously stirred using magnetic beads at 50 rpm; the temperature was maintained at 37±0.5 °C. The samples of 1 ml were withdrawn at the time interval of 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, and 24 h, analyzed for drug content spectrophotometrically at 215 nm against blank. The receptor phase was replenished with an equal volume of phosphate buffer at each time of sample withdrawal. The cumulative amounts of drug permeated per square centimeter of patches were plotted against time.

The diffusion kinetics of the drug Maslinic acid were analyzed by graphical method.

- Zero-order graphs were made by plotting Cumulative % drug release against Time in hours.
- First-order graphs were made by using Log cumulative % drug remaining against Time in hours.
- The diffusion pattern release of the formulation was studied by plotting Higuchi’s graph using Cumulative % drug released against Square root of time.
- The Peppas exponential equation was explained by plotting a graph of Log of cumulative % drug release against Log time.

RESULTS AND DISCUSSION

Pre-formulation studies

Drug: Maslinic acid

Physical description: Solid

Color/form: Crystalline solid

Melting point: 249-250 °C

Solubility: Solubility of 0.3 mg/ml in a 1:2 solution of Ethanol: Buffered Water (pH 7.2) at 25 °C. In ethanol: 0.5 mg/ml; in DMSO: 20 mg/ml; in dimethyl formamide: 15 mg/ml. sparingly soluble in aqueous buffers.

Density: 1.15 g/cu cm

Vapor pressure: 2.3X10^-15 mm Hg at 25 °C

Drug excipients compatibility studies

FT-IR spectrum and values

![Fig. 1: Spectrum of pure maslinic acid](image-url)
Fig. 2: IR Spectrum of pure HPMC E5

Fig. 3: IR spectrum of pure EC

| S. No. | IR spectrum of | Groups       | Peak(cm⁻¹) | Stretching/Deformation |
|-------|----------------|--------------|------------|------------------------|
| 1     | Maslinic acid  | N-tertiary   | 3436       | Stretching             |
|       |                | CH2          | 2692       | Stretching             |
|       |                | CH3          | 2345       | Stretching             |
|       |                | C=O          | 1651       | Stretching             |
|       |                | C=C          | 1477       | Stretching             |
|       |                | C-N          | 1396       | Stretching             |
|       |                | C=S          | 756        | Stretching             |
| 2     | HPMC E5        | O-H          | 3463       | Stretching             |
|       |                | C-O-C        | 1064       | Stretching             |
| 3     | EC             | CH2          | 2976       | Stretching             |
|       |                | CH3          | 2873       | Stretching             |
|       |                | C-O-C        | 1056       | Stretching             |
| 4     | Physical mixture of drug and polymer | N-tertiary | 3436       | Stretching             |
|       |                | CH3          | 2927       | Stretching             |
|       |                | C=O          | 1651       | Stretching             |
|       |                | C=C          | 1473       | Stretching             |
|       |                | O-H          | 3463       | Stretching             |
|       |                | C-O-C        | 1083       | Stretching             |
Fig. 4: IR spectrum of maslinic acid+HPMC E5+EC mixture

DSC curve of maslinic acid

Fig. 5: DSC curve of maslinic acid

Formulation of transdermal patches

Table 2: Compositions of different formulations containing maslinic acid

| Formulations          | F1  | F2  | F3  | F4  | F5  | F6  | F7  |
|-----------------------|-----|-----|-----|-----|-----|-----|-----|
| Maslinic acid, mg     | 0.5 | 0.5 | 0.5 | 0.5 | 0.5 | 0.5 | 0.5 |
| Ethylcellulose, mg    | 300 | *   | 30  | 60  | 90  | 120 | 150 |
| HPMC E5, mg           | *   | 300 | 270 | 240 | 210 | 180 | 150 |
| Dibutylphthalate (2drop), ml | 0.12 | 0.12 | 0.12 | 0.12 | 0.12 | 0.12 | 0.12 |
| DMSO, ml              | 0.06| 0.06| 0.06| 0.06| 0.06| 0.06| 0.06|
| Chloroform: Ethanol(1:1), ml | 5  | 5  | 5  | 5  | 5  | 5  | 5  |

*No ingredient used. HPMC=Hydroxypropyl Methylcellulose, DMSO=Dimethyl sulfoxide

Evaluation of transdermal patches

1) Thickness uniformity

Fig. 6: Thickness uniformity
Weight uniformity

Fig. 7: Weight uniformity

Folding endurance

Fig. 8: Folding endurance

Percentage moisture absorption

Fig. 9: Percentage moisture absorption

Percentage moisture loss

Fig. 10: Percentage moisture loss
Water vapour transition rate

![Water vapour transition rate](image)

Fig. 11: percentage water vapors transition rate

Tensile strength

![Tensile strength](image)

Fig. 12: Tensile strength

Drug content

![Drug content](image)

Fig. 13: Drug content

**Characterization of standard drug**

A single endothermic peak in case of Maslinic acid at 249.62 °C almost corresponds to its melting point as shown in fig. and table represents the different concentrations and their area under the curve for Maslinic acid by RP-HPLC method. This gives a standard curve with a linear regression equation and correlation coefficient as shown in fig. and table represents the different concentrations and their absorbance for Maslinic acid spectrophotometrically. This gives a standard curve with a linear regression equation and correlation coefficient, as shown in the figure.
In vitro drug diffusion study

Fig. 14: Comparative *in vitro* release profile of maslinic acid TDDS

Fig. 15: Comparative *in vitro* release profile of maslinic acid TDDS according to zero-order kinetics

Fig. 16: Comparative *in vitro* release profile of Maslinic acid TDDS according to first-order kinetics
Fig. 16: Comparative *in vitro* release profile of maslinic acid TDDS according to higuchi plot

Fig. 17: Comparative *in vitro* release profile of maslinic acid TDDS according to peppas plot

Fig. 18: Graph showing Regression coefficient ($R^2$) values of different kinetics models and diffusion exponent ($n$) of peppas model for maslinic acid TDDS

Fig. 19: Calibration curve of maslinic acid by HPLC
CONCLUSION

Various formulations utilizing hydrophilic and hydrophobic polymers like HPMC E5 and EC were created using solvent evaporation and penetration enhancers. The tensile strength of patches improved with HPMC content. Most batches had high folding endurance ratings (above 50), indicating that the patches will be less brittle when applied to the skin and retain their integrity when folded. A little quantity of wetness keeps the patch stable and avoids dry and brittle areas. In vitro experiments showed that HPMC E5 patches released more than EC patches, perhaps due to HPMC patches' high water vapour permeability and EC's hydrophobic nature. Maslinic acid transdermal patch produced via solvent evaporation. Preliminary in vivo studies on healthy animals can assess the pharmacokinetic characteristics. batch-to-batch consistency may be achieved via in vitro/in vivo correlation.

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CONFLICTS OF INTERESTS

There are no conflicts of interest.

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AUTHORS CONTRIBUTIONS

All the authors have contributed equally.

CONFLICT OF INTERESTS

Declares none

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