A Screen of *Coxiella burnetii* Mutants Reveals Important Roles for Dot/Icm Effectors and Host Autophagy in Vacuole Biogenesis

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Abstract

*Coxiella burnetii* is an intracellular pathogen that replicates in a lysosome-derived vacuole. The molecular mechanisms used by this bacterium to create a pathogen-occupied vacuole remain largely unknown. Here, we conducted a visual screen on an arrayed library of *C. burnetii* NMII transposon insertion mutants to identify genes required for biogenesis of a mature *Coxiella*-containing vacuole (CCV). Mutants defective in Dot/Icm secretion system function or the PmrAB regulatory system were incapable of intracellular replication. Several mutants with intracellular growth defects were found to have insertions in genes encoding effector proteins translocated into host cells by the Dot/Icm system. These included mutants deficient in the effector proteins Cig2, CoxCC8 and Cbu1754. Mutants that had transposon insertions in genes important in central metabolism or encoding tRNA modification enzymes were identified based on the appearance filamentous bacteria intracellularly. Lastly, mutants that displayed a multi-vacuolar phenotype were identified. All of these mutants had a transposon insertion in the gene encoding the effector protein Cig2. Whereas vacuoles containing wild type *C. burnetii* displayed robust accumulation of the autophagosome protein LC3, the vacuoles formed by the cig2 mutant did not contain detectable amounts of LC3. Furthermore, interfering with host autophagy during infection by wild type *C. burnetii* resulted in a multi-vacuolar phenotype similar to that displayed by the cig2 mutant. Thus, a functional Cig2 protein is important for interactions between the CCV and host autophagosomes and this drives a process that enhances the fusogenic properties of this pathogen-occupied organelle.

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Introduction

*Coxiella burnetii* is a highly infectious human pathogen responsible for a global zoonotic disease called Q fever. Inhalation of contaminated aerosols by humans can lead to an acute systemic illness or a more serious chronic infection that commonly presents as endocarditis [1,2]. The animal reservoirs for *C. burnetii* include domesticated livestock, and transmission to humans from these animals can lead to outbreaks of disease, such as the Q-fever epidemic that was linked to dairy goat farms in the Netherlands [2].

Phase I strains of *C. burnetii* produce a lipopolysaccharide molecule that has a complex O-antigen polysaccharide chain that protects the bacteria from being killed by host serum [3]. Phase II variants of *C. burnetii* produce a truncated O-antigen polysaccharide and can be isolated from both infected animals and bacteria cultured *in vivo* [3,4]. Although most strains of *C. burnetii* exhibit phase variation and switch between phase I and phase II spontaneously, a phase II variant of the exhibit phase variation and switch between phase I and phase II, giving rise to the phase I and phase II strains. The phase I strain is host-directed transport of the CCV through the endocytic pathway to eliminate the bacteria from the low pH environment of a lysosome [7,8]. Intracellular replication of *C. burnetii* requires formation of a specialized membrane-bound compartment termed the *Coxiella*-containing vacuole (CCV). After cells internalize *C. burnetii* there is host-directed transport of the CCV through the endocytic pathway, which delivers the bacteria to the low pH environment of a lysosome [7,8]. Intracellular *C. burnetii* resist the hydrolytic and bactericidal activities inside the lysosome and the acidic pH of this organelle is required to stimulate *C. burnetii* metabolism, which enables the bacteria to survive and replicate intracellularly [9,10]. Although the molecular mechanisms used by *C. burnetii* to
Author Summary

Coxiella burnetii is the causative agent of the human disease Q fever. This bacterium uses the Dot/Icm type IV secretion system to deliver effectors into the cytosol of host cells. The Dot/Icm system is required for intracellular replication of C. burnetii. To determine the contribution of individual proteins to the establishment of a vacuole that supports C. burnetii replication, we conducted a visual screen on a library of C. burnetii transposon insertion mutants and identified genes required for distinct stages of intracellular replication. This approach was validated through the identification of intracellular replication mutants that included insertions in most of the dot and icm genes, and through the identification of individual effector proteins delivered into host cell by the Dot/Icm system that participate in creating a vacuole that supports intracellular replication of C. burnetii. Complementation studies showed convincingly that the effector Cig57 was critical for intracellular replication. The effector protein Cig2 was found to play a unique role in promoting homotypic fusion of C. burnetii vacuoles. Disrupting host autophagy phenocopied the defect displayed by the cig2 mutant. Thus, our visual screen has successfully identified effectors required for intracellular replication of C. burnetii and indicates that Dot/Icm-dependent subversion of host autophagy promotes homotypic fusion of CCVs.

definition of axenic culture conditions. Formerly classified as an obligate intracellular bacterium, it has been shown that C. burnetii replicates axenically in Acidified Cysteine Citrate Media (ACCM) with 5% CO₂ and low oxygen conditions [23,24]. Subsequently, genetic approaches were developed to isolate transposon-insertion mutants and mutants with targeted gene deletions [25–27], which were used to demonstrate that the Dot/Icm type IVB secretion system encoded by C. burnetii is essential for intracellular replication [27–29]. This secretion system is genetically and functionally related to the Dot/Icm system of the human pathogen Legionella pneumophila [30–32]. In L. pneumophila, the Dot/Icm system facilitates intracellular replication by translocating into the host cytosol approximately 300 different effector proteins [33]. These effectors rapidly modulate the host cell biology to direct the remodeling of the Legionella-containing vacuole (LCV), which prevents fusion with lysosomes and promotes fusion of secretory vesicles to create a vacuole that supports intracellular replication [33]. The biochemical function of a small proportion of these effectors has been elucidated but correlating these functions to pathogenesis is hampered by a large degree of functional redundancy both in terms of multiple paralogs with mirroring functions [34] and dissimilar effectors targeting the same host cell pathways [35,36]. With few exceptions, deletion of a gene encoding a L. pneumophila effector does not typically have a measurable impact on the ability of the bacterium to replicate intracellularly. It is thought that the diversity of the natural protozoan hosts L. pneumophila encounters in nature has resulted in the selection of functionally-redundant effectors that mediate survival in specific protozoan hosts [36].

The L. pneumophila Dot/Icm system initiates effector translocation immediately upon contact with a host to prevent the LCV from engaging the host endocytic pathway [37,38]. By contrast, the C. burnetii Dot/Icm system does not translocate effectors until the bacteria are delivered to lysosomes and become metabolically active in an acidified vacuole [39]. Given their divergent intracellular infection strategies it is predicted that there will be minimal overlap in the function of the effectors encoded by L. pneumophila when compared to C. burnetii, which is supported by the observation that few bone fide homologs of L. pneumophila effectors are encoded in the C. burnetii genome. To date, over 100 C. burnetii Dot/Icm effectors have been identified using a range of methods [28,40–46]. The translocation of the majority of these effectors was observed using L. pneumophila as a surrogate effector delivery platform. Several C. burnetii effectors have been implicated in preventing apoptosis [43,47], including the ankyrin repeat-containing protein AnkG that infers an anti-apoptotic role [48], and to manipulate other aspects of the host biology important for intracellular replication. Because C. burnetii replicates exclusively inside mammalian hosts it is predicted that there will be less functional redundancy in the cohort of C. burnetii effector proteins compared to what is observed for L. pneumophila effector proteins, and that loss of single effectors may impact CCV formation. This means that it should be possible to identify effectors important for intracellular replication, and that determining the function of these effectors will increase our understanding of CCV development.

Here, we conducted a visual screen on an arrayed library of random transposon insertion mutants of C. burnetii NMII to identify genes important for formation of the mature CCV. This approach was successful and resulted in the identification of genes that are important for the intracellular lifestyle of C. burnetii. The

transform a lysosome into a replication-permissive compartment remain unclear, there is evidence that this compartment interacts with vesicles derived from the host autophagic and secretory pathways [11–13]. This results in a compartment containing vesicles derived from the host autophagic and secretory pathways remaining unclear, there is evidence that this compartment interacts with vesicles derived from the host autophagic and secretory pathways [11–13]. This results in a compartment containing vesicles derived from the host autophagic and secretory pathways. Homotypic fusion of lysosome-derived compartments will lead to the formation of a large lysosome-derived compartment in the infected cell, which consumes cellular lysosomes and results in the transformation of axenic culture conditions. Formerly classified as an obligate intracellular bacterium, it has been shown that C. burnetii replicates axenically in Acidified Cysteine Citrate Media (ACCM) with 5% CO₂ and low oxygen conditions [23,24]. Subsequently, genetic approaches were developed to isolate transposon-insertion mutants and mutants with targeted gene deletions [25–27], which were used to demonstrate that the Dot/Icm type IV secretion system encoded by C. burnetii is essential for intracellular replication [27–29]. This secretion system is genetically and functionally related to the Dot/Icm system of the human pathogen Legionella pneumophila [30–32]. In L. pneumophila, the Dot/Icm system facilitates intracellular replication by translocating into the host cytosol approximately 300 different effector proteins [33]. These effectors rapidly modulate the host cell biology to direct the remodeling of the Legionella-containing vacuole (LCV), which prevents fusion with lysosomes and promotes fusion of secretory vesicles to create a vacuole that supports intracellular replication [33]. The biochemical function of a small proportion of these effectors has been elucidated but correlating these functions to pathogenesis is hampered by a large degree of functional redundancy both in terms of multiple paralogs with mirroring functions [34] and dissimilar effectors targeting the same host cell pathways [35,36]. With few exceptions, deletion of a gene encoding a L. pneumophila effector does not typically have a measurable impact on the ability of the bacterium to replicate intracellularly. It is thought that the diversity of the natural protozoan hosts L. pneumophila encounters in nature has resulted in the selection of functionally-redundant effectors that mediate survival in specific protozoan hosts [36].

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requirement for a functional Dot/Icm system in biogenesis of the CCV was evident. Insertions that inactivated genes encoding structural components of this secretion apparatus were identified in addition to insertions in genes encoding regulatory factors that govern expression of the Dot/Icm system. Importantly, these studies have identified several effector proteins that play important and distinct roles during intracellular replication. Specifically, using this approach we reveal a genetic interaction between the effector Cig2 and the host autophagy pathway. These data indicate that Cig2 function is required for robust interactions between the CCV and host autophagosomes, and that this maintains the CCV in an autolysosomal stage of maturation.

**Results**

**Construction of an arrayed library of *C. burnetii* transposon mutants**

The plasmid pKM225 encoding a Himar1 Tn7 transposase was used to introduce a transposon encoding chloramphenicol resistance and a mChERRY fluorescent protein randomly onto the genome of the *C. burnetii* NMII strain RSA493. The mutagenesis procedure was optimized to reduce isolation of siblings containing identical transposon insertions and to reduce the number of spontaneous mutants defective for Dot/Icm function (described in Materials and Methods). After optimization, 3,840 transposon insertion mutants were isolated as single clones from 40-independent pools. These clones were then arrayed and expanded in 96-well plates containing ACCM-2. We were successful in expanding 84.6% of the clones (3,237 mutants) in liquid ACCM-2 under antibiotic selection. To determine the degree to which these clones represented independent mutants with different sites of transposon insertion, we determined the site of insertion for a total of 459 mutants using a two-stage semi-degenerate PCR amplification and sequencing protocol. This analysis confirmed that isolated clones had single transposon insertions distributed randomly across the *C. burnetii* genome. Additionally, this analysis revealed several mutants that had transposon insertions in genes encoding known effectors of the Dot/Icm system (Tables S1, S2, S3, S4, S5).

**A visual screen identifies *C. burnetii* mutants with vacuole biogenesis defects**

The arrayed library of *C. burnetii* NMII transposon mutants was analyzed using a visual assay that assessed the ability of individual mutants to form vacuoles that support intracellular replication. Specifically, HeLa 224 cells distributed in 96-well glass-bottom plates were infected with individual mutants at an MOI of approximately 500 and then the cells were incubated for 96 h. Cells were fixed and stained with antibodies specific for *Coxiella* (red) and LAMP1 (green), and the DNA was labeled with Hoechst dye (blue). The ability of each mutant to form a vacuole that supported intracellular replication was assessed by direct examination using fluorescence microscopy. Importantly, the parental NMII control strain and most of the *C. burnetii* transposon insertion mutants formed a single spacious vacuole filled with replicating bacteria (Figure 1A). Additionally, of the 459 mutants for which the transposon insertion site was determined, we found that 324 mutants (71%) did not display a discernable vacuole formation defect in this visual assay, which included several mutants having insertions in genes encoding known effectors of the Dot/Icm system (Table S5). Lastly, as described in detail below, many of the mutants that displayed vacuole formation defects had insertions in genes that would be predicted to affect intracellular replication. Thus, confidence was high that this screen would identify a unique cohort of genes important for *C. burnetii* replication in mammalian cells.

There were four distinct mutant phenotypes revealed in this visual screen (Figure 1). A severe defect characterized as no detectible intracellular replication in the visual screen was observed for 74 mutants having transposon insertions that were distributed among 21 different protein-coding regions and six different intergenic regions (Table S1). At 96 h post-infection these mutants were observed as single bacteria inside of LAMP1-positive vacuoles (Figure 1B). Forty-two transposon mutants displayed a reduced ability to replicate intracellularly as determined by their presence in small vacuoles containing fewer bacteria compared to vacuoles containing the control strain (Figure 1C, Table S2). Nine transposon mutants displayed filamentous growth inside of host cells (Figure 1E, Table S3), suggesting that these bacteria were under stress or defective for cell division. Lastly, there were 10 transposon mutants isolated independently that displayed a multi-vacuolar phenotype, which was characterized by the appearance of infected host cells that contained multiple vacuoles each supporting replication of *C. burnetii* (Figure 1D, Table S4). Importantly, every mutant we identified that displayed this multi-vacuolar phenotype had a transposon insertion in the 2,430 bp region encoding the protein Cig2 (Cbu0021).

**Genes essential for Dot/Icm function are highly represented among the mutants defective for intracellular replication**

Mutants with transposon insertions in the genes *icmL*·2 or *icmD* and mutants with targeted deletions of the genes *dotA* or *dotB* were shown previously to be defective for intracellular replication [27–29], which established the essential role the Dot/Icm system has in *C. burnetii* pathogenesis. Here, we identified 66 different intracellular growth mutants harboring a transposon insertion in *dot* and *icm* loci and three mutants that were severely attenuated for intracellular replication with insertions in this region (Figure 2A). The observation that many of the intracellular growth mutants have insertions in the region encoding the Dot/Icm system, as well as the extensive coverage of this region that was obtained, provides addition evidence that the arrayed mutant library contains a random distribution of transposon insertions. Additionally, this analysis indicates that spontaneous unlinked mutations that affect Dot/Icm function did not occur at a high frequency. This was a concern because if spontaneous *dot* and *icm* mutants were encountered at a high frequency then the mutant library would not be effective at identifying effectors essential for intracellular replication, and most intracellular growth phenotypes would not be complemented in trans upon introducing plasmids encoding the wild type allele of the disrupted gene. In the pool of 450 transposon mutants that were sequenced, we identified multiple insertion mutations in the coding region located between the *icmQ* and *icmT* genes (Figure 2A). No defects in CCV biogenesis were observed for these mutants, indicating that the genes in this region are not essential for Dot/Icm function. Also, we determined that the hypothetical protein Cbu1651 encoded in this region was not essential for Dot/Icm function because the mutant 16-E10 had an insertion in the *cbu1651* coding region but did not have a vacuole biogenesis defect. Two mutants with the transposon inserted at the 3′ end of the *cbu1651* gene displayed a vacuole biogenesis defect, however, these insertions are predicted to negatively affect expression of *icmX*, which is a gene essential for Dot/Icm function [49].

Previously, it was demonstrated that the *C. burnetii* *icmD* gene was required for intracellular replication [29]. Additionally, studies on *L. pneumophila* predict that *icmC*, *icmN*, *icmT* and *dotD* should
also be important for function of the C. burnetii Dot/Icm system [50–53], and a recent independent study has shown that C. burnetii mutants deficient in dotD, icmC and icmN display intracellular replication defects [54]. Although complementation studies and in-frame deletion analysis was not used to more precisely determine the specific dot and icm genes that were essential for intracellular replication, the region itself was highly represented among mutants with severe intracellular growth.

Figure 1. Transposon mutants of C. burnetii display different intracellular phenotypes. Transposon mutants were subject to a vacuole formation assay in which 96-well plates of HeLa 229 cells were infected, at an MOI of approximately 500, with individual transposon mutants. Following a 96 h infection period, the infection was fixed and stained with anti-Coxiella (red), anti-LAMP1 (green) and Hoechst dye (blue) and observed with low magnification fluorescence microscopy. (A) The parental strain C. burnetii NMII displayed a large CCV in the majority of HeLa cells. (B) A cohort of mutants showed no intracellular replication as demonstrated by dotA::Tn, (C) another category produced smaller replicative vacuoles such as that seen with cig57::Tn, (D) transposon insertions that disrupted cig2 resulted in the appearance of multiple vacuoles in a single cell, and (E) a small proportion of mutants, such as gidA::Tn, displayed CCVs with an abnormal shape due to filamentous replication of the C. burnetii.

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defects, which suggested that it should be possible to identify other important genes required for intracellular replication using this library of transposon mutants.

The two-component regulatory system PmrAB is required for Dot/Icm effector translocation

The response regulator PmrA of *L. pneumophila* is important for intracellular growth of this pathogen as it controls the expression of genes encoding components of the Dot/Icm system and many effectors [55]. The *C. burnetii cbu1227* gene encodes a PmrA homologue [55], which was initially annotated as QseB [30]. The prediction is that PmrA activity is controlled by the sensor kinase PmrB encoded by an adjacent gene in the operon. At least 68 promoter regions in *C. burnetii* contain a consensus PmrA binding site, which included five promoter regions upstream of operons that encode most of the *dot* and *icm* genes [55]. Thus, the prediction is that expression of most *C. burnetii dot* and *icm* genes will require a functional PmrAB system. Consistent with this hypothesis, among the *C. burnetii* mutants identified that were defective for vacuole biogenesis, we isolated three mutants with a transposon insertion in *pmrB*, one mutant with a transposon insertion in *pmrA*, and one mutant with a transposon insertion in the regulatory region upstream of *pmrA* (Figure 2B). To determine if the Dot/Icm system was operational in mutants defective for PmrAB function we introduced a plasmid encoding a β-lactamase reporter (BlaM) fused to the effector protein Cbu0077 that produces the hybrid protein BlaM-77. The BlaM-77 protein was produced in the *pmrA::Tn* mutant strain and effector protein translocation was assayed during host cell infection (Figure 2C). To determine if the Dot/Icm system was operational in mutants defective for PmrAB function we introduced a plasmid encoding a β-lactamase reporter (BlaM) fused to the effector protein Cbu0077 that produces the hybrid protein BlaM-77. The BlaM-77 protein was produced in the *pmrA::Tn* mutant strain and effector protein translocation was assayed during host cell infection (Figure 2C). To determine if the Dot/Icm system was operational in mutants defective for PmrAB function we introduced a plasmid encoding a β-lactamase reporter (BlaM) fused to the effector protein Cbu0077 that produces the hybrid protein BlaM-77. The BlaM-77 protein was produced in the *pmrA::Tn* mutant strain and effector protein translocation was assayed during host cell infection (Figure 2C).

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**Figure 2.** The Dot/Icm system and the PmrAB system are essential for intracellular replication and delivery of *C. burnetii* effector proteins. (A) Indicated are the locations of transposon insertions in the chromosomal regions encoding the Dot/Icm system from 1559100 bp to 1591800 bp, and (B) the PmrAB two-component regulatory system from 1176500 to 1179500 bp. The site of each transposon insertion is represented by an arrow. The mutants that could not replicate intracellularly were assigned red arrows, mutants that produced normal CCVs were assigned black arrows, and mutants that formed small vacuoles were assigned grey arrows. (C) The plasmids pBlaM and pBlaM-77 were introduced into *C. burnetii* NMII (black bars) and *pmrA::Tn* (grey bars) to determine whether the PmrAB system was essential for effector translocation. Cleavage of the fluorescent β-lactamase substrate CCF4-AM was determined by calculating the ratio of fluorescence at 460 nm to 535 nm relative to uninfected cells. The graph shows the mean ± SD calculated for three independent samples. doi:10.1371/journal.ppat.1004286.g002

The effector protein Cig57 is important for intracellular replication

Many *C. burnetii* NMII transposon mutants had an intracellular replication defect that resulted in a significant reduction in the size of vacuoles and the number of bacteria in each vacuole. Included in this category were three transposon insertions that were predicted to result in partial loss-of-function in the Dot/Icm system. These mutants included transposon insertions in the *icmS* gene encoding a chaperone protein that assists in effector translocation [36], a mutant with an insertion located in the 3′ region of *icmX* that would result in the production of an IcmX protein with a small C-terminal deletion, and a mutant with an insertion in the *cbu1651* gene that likely affects the expression of adjacent *dot* and *icm* genes.

Reduced intracellular replication was also observed in mutants having insertions in genes encoding three different effector proteins, which were Cig57, CoxCC8 and Cbu1754. We isolated 10 intracellular growth mutants having a transposon insertion in *cig57* and the *cig57::Tn* mutant called 3-H3 was analyzed in detail. When intracellular replication was measured over a seven-day period, the 3-H3 strain displayed only a 5-fold increase in genome equivalents (GEs, Figure 3). Introduction of a plasmid-encoded triple FLAG-tagged version of 3×FL-Cig57 (pFLAG-Cig57) into the 3-H3 *cig57::Tn* mutant restored intracellular replication.
replication to levels that were equivalent to wild type *C. burnetii*, as determined by 257-fold increase in GE after a 7-day infection period and the appearance by immunofluorescence microscopy of large spacious vacuoles containing replicating 3-H3 (pFLAG-Cig2) bacteria (Figure 3B & 3C). Importantly, there were no obvious defects observed in the maturation of vacuoles containing 3-H3 as indicated by the presence of both LAMP1 (Figure 3) and cathepsin D (Figure S1) on the vacuoles formed by this mutant. Thus, the effector protein Cig57 likely has a role in modulating host processes important for replication that occur after *C. burnetii* is delivered to a lysosome-derived organelle.

**Screening for intracellular growth mutants identifies a new Dot/Icm effector protein**

We identified a strain of *C. burnetii* having a transposon insertion in the gene *cbu1780* and a strain having an insertion in the gene *cbu2072* that had both displayed a severe intracellular growth defect. Both of these genes encode hypothetical proteins, which raised the possibility they might encode novel effectors. To determine if these proteins might encode effectors both Cbu1780 and Cbu2072 were tested for Dot/Icm-dependent translocation using fusion proteins having an amino-terminal BlaM reporter. The resulting BlaM-1780 and BlaM-2072 fusion proteins were produced in *C. burnetii* NM II and the isogenic icmL::Tn strain. The BlaM-1780 fusion protein was translocated into the host cytosol when produced in *C. burnetii* with a functional Dot/Icm system, as determined by a significant increase in the 460:535 nm fluorescence ratio to 24.62±2.70. By contrast, no translocation was detected by BlaM-2072, 1.07±0.06. Similarly, no translocation was detected for the controls, which included BlaM-1780 and BlaM-2072 produced in the Dot/Icm-deficient mutant and BlaM alone produced in the parental *C. burnetii* NMII strain (Figure S2). Thus, Cbu1780 is an effector protein that has an important role during infection.

**The effector Cig2 is required for homotypic fusion of CCVs**

A striking phenotype that resulted in the appearance of multiple CCVs in HeLa cells that were infected by *C. burnetii* was observed for 10 independent transposon insertion mutants (Figure 1D). Because individual vacuoles containing replication-competent *C. burnetii* will undergo homotypic fusion inside of an infected cell this phenotype suggests that these mutants were defective in promoting homotypic fusion of the CCV. All of the mutants identified that displayed this multi-vacuolar phenotype had a transposon insertion in the gene encoding the hypothetical protein Cig2 (Chut0021), which was recently postulated to be an effector because it could be translocated by the *L. pneumophila* Dot/Icm system [45]. The mutants 3-C3 and 1-D12 producing normal cig2::Tn expression of downstream genes. When a plasmid encoding the cig2::Tn mutant strain 2-E1 the resulting 2-E1 (pFLAG:Cig2) was translocated during host cell infection and that translocation of BlaM-Cig2 by *C. burnetii* required the Dot/Icm system (Figure 4D). Thus, these data indicate that the Cig2 protein is a translocated effector required for homotypic fusion of the CCV.

**Host autophagy is required for Cig2-dependent homotypic fusion of CCVs**

Previous studies have revealed a multi-vacuolar phenotype when the host gene encoding Syntaxin-17 (STX17) was silenced in HeLa cells and the STX17-silenced cells were then infected with NMII [8]. The similarity between the phenotype in STX17-silenced cells and the multi-vacuole phenotype observed for the cig2::Tn mutant suggested a genetic interaction between Cig2 and STX17. Recent data has shown that STX17 has an essential role in the host process autophagy [57,58], which would suggest autophagy might be required for homotypic fusion of CCVs. LC3 is a protein that is attached to autophagosomal membranes [59], and is important for autophagosome biogenesis and the selection of intracellular cargo that will be enveloped by autophagy. Consistent with the hypothesis that autophagy may be subverted during *C. burnetii* infection, it has been shown that LC3 is present on the CCV [12]. To determine if autophagy is required for CCV homotypic fusion, we used siRNA to silence the genes encoding the essential autophagy factors ATG5 and ATG12, and vacuole biogenesis was assayed by immunofluorescence microscopy. Compared to mock-transfected cells or cells where the control protein syntaxin-18 (STX18) was silenced, there was a significant increase in the percentage of *C. burnetii*-infected cells having two or more vacuoles per cell when the genes encoding the autophagy factors ATG5, ATG12, or STX17 were silenced (Fig. 5A & B). Thus, a functional host autophagy system is required for Cig2-dependent homotypic fusion of the CCV.

The *L. pneumophila* Dot/Icm effector RavZ is translocated into the host cell during infection and inhibits autophagy by directly uncoupling ATG8 proteins attached to autophagosomal membranes, which includes LC3 [60]. We generated a *C. burnetii* strain that produces 3×FL-RavZ to determine if autophagy is important during the initial stage of infection when the Dot/Icm system is silent or during a later stage of infection when effectors are delivered into host cells. HeLa cells infected with *C. burnetii* producing 3×FL-RavZ had an autophagy defect as determined by the reduction in lipidated LC3-II protein when compared to uninfected cells or cells infected with *C. burnetii* producing the catalytically inactive 3×FL-RavZC258A protein (Fig. 5D). Importantly, most of the cells infected with *C. burnetii* producing functional 3×FL-RavZ displayed the multi-vacuolar phenotype defined by the presence of two or more vacuoles containing *C. burnetii*, whereas cells producing the catalytically inactive 3×FL-RavZC258A protein did not (Fig. 5C & E). Thus, Dot/Icm-mediated delivery of 3×FL-RavZ interfered with homotypic fusion of the CCV by blocking autophagy after bacteria had been transported to a lysosome-derived compartment in the cell, which indicates that Cig2-mediated homotypic fusion of the CCV requires membranes that display lipidated ATG8 proteins.

**The effector Cig2 is important for CCV fusion with autophagosomes**

The finding that defects in host autophagy or loss-of-function mutations in cig2 both result in a multi-vacuolar phenotype suggested that *C. burnetii* might subvert host autophagy by a Cig2-
**Figure 3.** The Dot/Icm effector Cig57 is required for efficient intracellular replication of *C. burnetii*. (A) Intracellular replication of *C. burnetii* NMII (black squares), cig57::Tn (black triangles) and cig57::Tn pFLAG-Cig57 (open circles). The fold-increase in GE relative to the inoculum was determined by dotA-specific qPCR and is represented here as the mean ± SD of 3 independent infections at days 1, 3, 5 and 7 post-infection. (B,C) Representative micrographs of HeLa cells infected for either 3 days (B) or 5 days (C) with *C. burnetii* NMII, cig57::Tn or cig57::Tn pFLAG-Cig57. Cells were stained with anti-Coxiella antibody (red), anti-LAMP1 antibody (green) and Hoechst dye (blue). Note that the vacuoles containing cig57::Tn were smaller and contained few bacteria at each time point compared to the control NMII strain or the complemented cig57::Tn pFLAG-Cig57 strain. Scale bars represent 10 μm.

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**Figure 4.** The Dot/Icm effector Cig2 is necessary for homotypic fusion of CCVs. (A) HeLa cells were infected with *C. burnetii* NMII strains at an MOI of 500. Five days post-infection the infections were fixed and stained with anti-Coxiella (red), anti-LAMP1 (green) and Hoechst dye (blue). Representative micrographs demonstrate the multiple vacuole phenotype observed for the cig2::Tn mutant strain. * indicates the location of individual CCVs within a chosen cell. These CCVs were identified both by LAMP1 staining and phase contrast microscopy. Scale bars represent 10 μm. (B) The average number of vacuoles containing *C. burnetii* NMII (black bars), cig2::Tn (white bars) and cig2::Tn pFLAG:Cig2 (grey bars) was determined at day 3 post-infection for at least 100 infected cells on duplicate coverslips. Data are the mean ± SD from 3 independent experiments. (C) Replication of cig2::Tn (open triangles) and the parental *C. burnetii* NMII strain (black squares) was determined by measuring genome equivalents over a 7 day infection period. (D) Micrographs from translocation assays using *C. burnetii* NMII and the icmL::Tn strains producing either BlaM alone (pJB-Cm:BlaM) or BlaM-Cig2 (pJB-Cm:BlaM-Cig2). Fluorescence intensity at 535 nm of uncleaved CCF4-AM is shown in green. Fluorescence intensity of cleaved CCF4-AM at 460 nm generated by the enzymatic activity of BlaM fusion proteins delivered into the host cell cytosol is shown in blue. These images are representative of 3 independent experiments.

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Figure 5. Autophagy is required for homotypic fusion of CCVs. (A) HeLa cells in which the indicated genes were silenced using siRNA were infected with C. burnetii and fixed 3 days post-infection before immunostaining with anti-C. burnetii (red) and anti-LAMP1 (green) antibodies. DNA was stained with Hoechst (blue). Shown are representative micrographs for mock-transfected cells and cells transfected with siRNA specific to human ATG5, ATG12, STX17, and STX18. (B) Single-cell quantification of C. burnetii vacuole counts in ATG5, ATG12, and STX17 siRNA-transfected HeLa cells compared to STX18-transfected cells and mock-transfected cells at 3 days post-infection. The percentages of infected cells with two or more C. burnetii vacuoles were counted. Shown is the mean ± SD from 3 independent experiments. (C) HeLa cells infected with C. burnetii expressing the L.
Autophagy Subversion by Coxiella burnetii

deeply dependent mechanism. Consistent with this hypothesis we found that the host autophagy protein LC3 was abundant on large vacuoles containing the parental NMII strain, whereas vacuoles containing the isogenic cig2::Tn mutant had a severe defect in LC3 accumulation (Figure 6A & B). LC3 accumulation at the CCV was restored when a plasmid encoding 3×FLAG-Cig2 was introduced into the cig2::Tn mutant. To determine if Cig2 may increase autophagy flux in infected cells the autophagy rates were assessed after infection by measuring the accumulation of lipophilic LC3-II in cells infected with the cig2::Tn mutant strain were compared with cells infected with the parental NMII strain of C. burnetii (Figure S2). These data indicate that infection by C. burnetii does not elevate the basal rate of autophagy and that Cig2 function does not affect autophagy flux during infection.

Lastly, we asked whether the CCV created by the cig2::Tn mutant was still accessible to fluid-phase endocytic transport and whether the lumen of the vacuole remained hydrolytic. This question was addressed by pulsing infected macrophages with soluble DQ Green BSA added to the extracellular medium. Endocytic transport and cleavage of DQ Green BSA by lysosomal proteases generates fluorescent peptides that permit visualization of hydrolytic organelles by fluorescence microscopy. Similar to the organelles formed by the parental NMII strain and the cig2::Tn mutant complemented with the plasmid producing 3×FLAG-Cig2, the vacuoles containing the Cig2-deficient mutants retained the ability to cleave DQ Green BSA as indicated by the green fluorescence localized to the CCV (Figure 6D), which is consistent with the finding that these vacuoles contain the lysosomal protease Cathepsin D (Figure S1). These data indicate that compared to vacuoles formed by the parental NMII strain, the lumen of the vacuole containing the cig2::Tn mutant has a similar capacity to receive endocytic cargo and hydrolyze proteins. This, Cig2 function is required to promote fusion of autophagosomes with the initial acidified lysosome-derived vacuole in which C. burnetii resides.

Discussion

Here we employed large-scale transposon mutagenesis to create an arrayed library of 3,237 C. burnetii transposon insertion mutants. The C. burnetii NMII RSA 493 genome is comprised of a chromosome that is 1,995,275 bp and a 37,393 bp plasmid called QpH1 [30]. The total number of mutants obtained would correlate with at least one insertion for every 628 bp of DNA assuming the transposon we used inserted randomly in the genome. Given that the average size of a C. burnetii open reading frame is 849 bp most non-essential genes should be present in the library. Consistent with these calculations, our screen identified insertions in most of the dot and icm genes predicted to be non-essential for axenic growth of C. burnetii. There were, however, also 10 independent insertions isolated in the 2,430 bp gene cig2, which is higher than would be predicted given random distribution of the transposon throughout the genome. Thus, we are confident that insertions in most of the genes required for intracellular infection by C. burnetii but not for replication in axenic medium were represented in this arrayed library of mutants; however, we acknowledge that there are difficulties in reaching saturation of the genome by transposon mutagenesis that could result in a several genes required for intracellular replication not being present in this library.

We found that loss-of-function mutations in the PmrAB two-component regulatory system abolished intracellular replication of C. burnetii, which is consistent with independent data reported in two recent studies [54,61]. Here we employed large-scale transposon mutagenesis to create an arrayed library of mutants; however, we acknowledge that there are difficulties in reaching saturation of the genome by transposon mutagenesis that could result in several genes required for intracellular replication not being present in this library.
Figure 6. The effector Cig2 is important for CCV fusion with autophagosomes. (A) HeLa cells were infected with C. burnetii N41, cig2::Tn or cig2::Tn pFLAG:Cig2 for 5 days and stained with anti-LC3 (green) and anti-Coxella (red) antibodies. (B) The association of LC3 with the CCV was quantified by observing 100 infected cells per sample in 3 independent experiments. (C) LC3 and actin immunoblots from HeLa cells infected with parental C. burnetii N41, the cig2::Tn mutant, the parental C. burnetii N41 producing either RvA2 or RvA2C258A, or uninfected cells. Cells were untreated (top image), or treated (bottom image) with bafilomycin A1 and rapamycin for 2 h prior to lysis. (D) J774A.1 cells were infected with C. burnetii N41, cig2::Tn, or cig2::Tn pFLAG:Cig2 for 36 h before loading with 50 μg/ml DQ Green BSA for 16 h. Micrographs show phase contrast images to reveal vacuoles and the green fluorescence that results from DQ Green BSA degradation by proteases in lysosomal compartments. doi:10.1371/journal.ppat.1004286.g006

given the high oxidative stress caused by residing in a lyosome-like organelle.

Several C. burnetii mutants were identified in the visual screen because they displayed a filamentous growth phenotype. Disruption of the two-component regulatory system encoded by cbu0505 and cbu0606, mnaA, pspP and gidA resulted in filamentous replication intracellularly. The protein Cbu0745 is predicted to be the C. burnetii homolog of ribosome-associated factor Y, and the proteins MnmA and GidA are enzymes involved in tRNA modification. Three independent mutants that displayed a filamentous growth phenotype were found to have insertions in the gidA gene, and previous studies indicate that disruption of gidA in Salmonella also results in bacteria that have defect in cell division resulting in filamentation [66,67]. This gidA mutant phenotype has been attributed to an altered expression of genes responsible for cell division and chromosome segregation [66]. Thus, it is likely that many of the C. burnetii mutants that demonstrate filamentation have defects in fundamental cellular processes including translation and chromosome segregation that affect cell division.

Specific Dot/Icm effector proteins critical for CCV biogenesis and intracellular replication of C. burnetii were identified in this visual screen. Three other recent studies have reported C. burnetii intracellular replication defects resulting from mutations in specific Dot/Icm effectors [46,48,54]. By contrast, genetic screens to isolate intracellular replication mutants in L. pneumophila identified the Dot/Icm secretion system as being critical for intracellular replication, but did not reveal effector proteins that are essential for intracellular replication. To illustrate this point, it was shown that a Legionella strain having five large chromosomal deletions that eliminated the production of 71 different effector proteins could still replicate inside macrophages [68], which provides further evidence that there is extensive functional redundancy built into the Legionella effector repertoire and this makes it difficult to identify effectors required for virulence by screening mutants for intracellular replication defects. Thus, the identification of effector mutants with strong intracellular growth phenotypes suggests that there is slightly less functional redundancy in the C. burnetii effector repertoire compared to Legionella. However, we identified mutants having transposon insertions in genes encoding 16 different effector proteins and were unable to detect any defects in intracellular replication or vacuole morphology for these effector mutants. Thus, it remains likely that there are functionally redundant effectors that modulate some of the host functions required for intracellular replication of C. burnetii. Additionally, it is likely that some of the effectors that are encoded by C. burnetii play important roles during infection of animals even though these effectors are not required for C. burnetii replication in host cells cultured ex vivo. Hypothetically, there could be effectors that modulate inflammation by preventing detection of C. burnetii by either innate or adaptive immune surveillance that would be predicted to fall into this category.

In our initial attempts at using transposon insertion mutagenesis to identify genes important for intracellular replication we were befuddled by loss-of-function mutations presumably arising spontaneously at a high frequency in dot and icm genes, which resulted in intracellular growth defects that were not linked to the site of transposon insertion. We optimized the mutagenesis protocol to reduce the probability of phenotypic differences being the result of spontaneous unlinked mutations. By either isolating multiple independent insertions in a gene where all mutants display the same phenotype or by complementing a phenotype by introducing a wild type allele on a plasmid, we demonstrate here that there are distinct phenotypes that are linked to transposon-mediated inactivation of a specific gene. However, it remains possible that some of the mutant phenotypes reported for insertion mutants described in the Supplemental Tables could be due to unlinked mutations and further studies are needed to support this initial analysis. Our data also suggest that unlinked mutations may have complicated results in a recent study where transposon insertions in genes encoding effector proteins were found to affect intracellular replication [46]. Complementation studies were not included in this study, which made it difficult to rule out the possibility that some of the transposon insertion mutants with intracellular growth defects had unlinked mutations that affect Dot/Icm function or the function of some other gene important for infection. For example, it was reported that a cbu2052 transposon insertion mutant had an intracellular replication defect, however, we obtained two independent mutants with insertions in the cbu2052 gene and immediately upstream of cbu2052 (Table S5) and both of these mutants formed CCVs that were indistinguishable from the vacuoles formed by the parental strain of C. burnetii. Thus, it is important that transposon insertion phenotypes in C. burnetii are validated using either complementation or allelic replacement approaches before important functions are assigned to effector proteins.

Ten intracellular replication mutants isolated in the screen were found to have independent insertions in the cig57 gene and the intracellular replication defect displayed by a cig57::Tn mutant was complemented using a plasmid encoded cig57 allele. Cig57 is highly conserved among sequenced C. burnetii strains, however, database searches did not identify other proteins with homology to Cig57. Thus, we can conclude with high confidence that Cig57 represents a unique effector protein that has an activity that is important for C. burnetii intracellular replication.

In addition to identifying mutants defective for intracellular replication, the visual screen revealed that C. burnetii cig2 mutants display a multi-vacuole phenotype. Whereas infection of a single cell by multiple C. burnetii usually leads to formation of a single vacuole due to homotypic fusion of the individual CCVs, the vacuoles containing cig2 mutants do not display the same propensity to fuse with each other inside the host cell, which results in a single host cell having multiple CCVs that each display LAMP1 and cathepsin D localization at the limiting membrane of the proteolytic lysosome-derived organelle. The cig2 gene encodes a protein with a predicted molecular weight of 92.9 kDa. The Cig2 protein is encoded in the genomes of all sequenced strains of C. burnetii, however, the protein does not have homologs in other organisms and there are no conserved domains that might aid in predicting the biochemical functions of this protein. Our data demonstrate that Cig2 is translocated into host cells during infection by C. burnetii using a mechanism that requires the Dot/
Icm system. Additionally, it has been shown that Cig2 produced in *Legionella* can be translocated into host cells by the Dot/Icm system [45]. Thus, Cig2 represents a functional Dot/Icm effector protein that modulates vacuole biogenesis.

The multi-vacuolar phenotype displayed by cig2::Tn mutants was similar to the multi-vacuolar phenotype displayed after STX17 was silenced and HeLa cells were infected with the parental NMII strain [8]. Why silencing of STX17 would result in a multi-vacuole phenotype was unclear initially, however, recent studies have shown that STX17 function is critical for autophagy in mammalian cells [57,58]. This suggested that host autophagy was required for homotypic fusion of CCVs. Indeed, our data show that silencing host genes encoding essential component of the autophagy machinery resulted in the multi-vacuole phenotype in *C. burnetii*-infected cells. Additionally, when the LC3-deconjugating effector RavZ was introduced into *C. burnetii*, the RavZ-producing *C. burnetii* were able to disrupt host autophagy and this resulted in a multi-vacuole phenotype. These data provide a clear phenotypic link between the host autophagy system and Cig2 function.

Similar to the unregulated fusion that occurs between pre-existing phagolysosomes and the CCV in infected cells, upregulation of autophagy in mammalian cells generates large autolysosomal organelles as autophagosomes consume lysosomes through rapid fusion [69]. Importantly, independent studies have shown that LC3 associates with the CCV during vacuole biogenesis by an active process mediated by viable *C. burnetii* [12,13]. Additionally, it has been shown that the presence of LC3 on phagosome membranes will promote rapid fusion with lysosomes by a process known as LC3-associated phagocytosis [70]. Thus, we hypothesized that the reason a *C. burnetii* cig2 mutant displays a multi-vacuolar phenotype is because this effector is important for autophagy subversion by *C. burnetii*. Finding that there is defect in the localization of LC3 to vacuoles formed by Cig2-deficient *C. burnetii* supports this hypothesis. Finding that the rates of autophagy were similar following infection of cells with NMII or the isogenic cig2::Tn mutant indicates that Cig2 does not stimulate a general upregulation of autophagy flux in the infected cells. This suggests that Cig2 function is required to promote fusion of autophagosomes that are generated at a basal level in the infected cells with the CCV.

Based on these data, we propose a model whereby autophagy subversion by Cig2 is required to constitutively promote the fusion of autophagosomes with the CCV during infection. This would enable Atg8 proteins such as LC3 to be maintained on the CCV membrane and keep the CCV in autolysosomal stage of maturation. We postulate that by locking the CCV in an autolysosomal stage of maturation, we postulate that by locking the CCV in an autolysosomal stage of maturation this vacuole would remain highly fusogenic, and this would promote homotypic fusion and fusion of the CCV with other lysosome-derived organelles in the cell. The result would be formation of a spacious CCV and the fusion of lysosome-derived organelles containing other bacteria or inert particles with the CCV. Determining whether this model is correct will require elucidating the biochemical function of Cig2 and a better understanding of the role autophagy subversion plays in generating the vacuole that *C. burnetii* occupies.

**Materials and Methods**

**Bacterial strains and host cell lines**

Plaque purified *C. burnetii* Nine Mile phase II (NMII), strain RSA93 clone 4, was aerobically grown in liquid ACCM-2 or ACCMagarose at 37°C in 5% CO2 and 2.5% O2 as previously described [24,71]. When appropriate, kanamycin and chloramphenicol were added to ACCM-2 at 300 µg/ml and 3 µg/ml respectively. HeLa 229 cells (CCL-2; ATCC, Manassas, VA) and J774.1 cells were maintained in Dulbecco’s Modified Eagle Media (DMEM) supplemented with 10% heat inactivated fetal bovine serum (FBS) at 37 °C in 5% CO2.

**Transposon mutagenesis of *C. burnetii* NMII**

pKM225 was introduced into the stationary phase *C. burnetii* NMII via electrotransformation at 18 kV, 500 Ω and 25 µF as previously described [28,71]. Following electroporation, the bacteria were recovered in 20 ml of ACCM-2 for 24 h before being plated on ACCMagarose plates containing chloramphenicol. After 6 days incubation, single colonies were isolated and resuspended in 1 ml aliquots of ACCM-2 with chloramphenicol in 24 well plates. Following a further 6 days, each 1 ml *C. burnetii* culture was passaged 1:1000 to provide bacteria for the vacuole formation assay and determination of the transposon insertion site. The remaining culture was pelleted via centrifugation and resuspended in 100 µl of DMEM containing 10% FBS and 10% Dimethyl sulfoxide for storage in 96 well plates.

**Determination of transposon insertion site**

The genomic location of the transposon insertion sites was determined for transposon mutants with distinct intracellular phenotypes and a wider random selection of recovered mutants. Nested primers within the transposon, facing the transposon-genome junction site, were used to amplify the insertion site either from *C. burnetii* cell lysate or purified genomic DNA. The first round of amplification used primer 1: GGGGGGAACGCCCTGG-TATC and a pool of random oligonucleotides with a common arm. The product of this PCR was used as a template for the second round of amplification with primer 2: GTGGGGTTTCGCACCTC and primer ARB2: GGCAGGCGCCTGACAGATG. The second PCR product was sequenced using primer 3: TCGATTTTTGTGATGCTCGTC. Sequencing results were analyzed using 4Peaks and BLAST programs.

**Vacuole formation assay and immunofluorescence microscopy**

1.5 x 10⁵ HeLa 229 cells were added into 96 well tissue culture plates. The next day monolayers were infected with stationary phase *C. burnetii* transposon mutants at a multiplicity of infection (MOI) of approximately 500 in DMEM with 5% FBS. The infection was allowed to progress for approximately 96 h, with the media changed 24 h after infection. After 96 h, cells were fixed with 4% paraformaldehyde and then blocked and permeabilized in blocking buffer (PBS containing 2% BSA and 0.05% saponin). The cells were stained with anti-LAMP1 monoclonal H4A3 (Developmental Studies Hybridoma Bank) and rabbit anti-*C. burnetii* polyclonal antibody in blocking buffer at 1:1000 and 1:10000 respectively. Secondary antibodies, Alexa Fluor 488 and 594 (Invitrogen) were used at 1:3000 also in blocking buffer. During final PBS washes bacterial and host DNA was stained with Hoechst 33342 (Invitrogen). Stained infections were visually analyzed using 4Peaks and BLAST programs.

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**Vacuole formation assay and immunofluorescence microscopy**

1.5 x 10⁵ HeLa 229 cells were added into 96 well tissue culture plates. The next day monolayers were infected with stationary phase *C. burnetii* transposon mutants at a multiplicity of infection (MOI) of approximately 500 in DMEM with 5% FBS. The infection was allowed to progress for approximately 96 h, with the media changed 24 h after infection. After 96 h, cells were fixed with 4% paraformaldehyde and then blocked and permeabilized in blocking buffer (PBS containing 2% BSA and 0.05% saponin). The cells were stained with anti-LAMP1 monoclonal H4A3 (Developmental Studies Hybridoma Bank) and rabbit anti-*C. burnetii* polyclonal antibody in blocking buffer at 1:1000 and 1:10000 respectively. Secondary antibodies, Alexa Fluor 488 and 594 (Invitrogen) were used at 1:3000 also in blocking buffer. During final PBS washes bacterial and host DNA was stained with Hoechst 33342 (Invitrogen). Stained infections were visually inspected for formation of large CCVs and the transposon mutants the exhibited abnormal CCV formation were investigated further. For additional immunofluorescence microscopy HeLa cells were added in 24-well dishes containing 10 mm glass coverslips. At the indicated times post-infection the cells were fixed and stained as above before being mounted on slides using ProLong Gold (Invitrogen). For cathepsin D staining of infected cells, anti-cathepsin D (Novus Biologicals) was used at 1:50 following fixation and permeabilized with ice-cold MeOH and blocking in PBS with 2% BSA. For endogenous LC3 staining
HeLa cells were infected for five days before fixing in cold methanol on ice for five minutes. Coverslips were blocked in 2% BSA, and stained with mouse anti-LC3 (NanoTools clone 2G6) and rabbit anti-Coxiella antibodies at 1:200 and 1:1,000, respectively, in blocking solution. Cells were washed three times in PBS and stained with anti-mouse 488 and anti-rabbit 546 at 1:2,000. Coverslips were washed three times with PBS and mounted on slides using ProLong Gold Antifade reagent (Invitrogen). Images for endogenous LC3 were acquired using an LSM510 confocal microscope equipped with a 100×/1.4 numerical aperture objective lens. Images were analyzed in Image J and Photoshop. For DQ Green BSA experiments, J774A.1 cells were infected with C. burnetii NMII, cig2::Tn, or cig2::Tn pFLAG-Cig2 in 35 mm glass bottom dishes. Cells were incubated in medium containing 50 μg/ml DQ Green BSA at 36 h post-infection and allowed to incubate for a further 16 h. Cells were washed three times with PBS and placed in fresh 5% FBS/DMEM (no phenol red) as described previously [5]. Live images were acquired after one additional hour of incubation with the fresh media. Digital images were acquired with a Nikon Eclipse TE2000-S inverted fluorescence microscope using a 60×/1.4 or 100×/1.4 numerical aperture objective lens and a Photometrics CoolSNAP EZ camera controlled by SlideBook v.5.5 imaging software.

C. burnetii intracellular growth curves

The day before infection, HeLa 229 were plated at a density of 5 × 10^5 into 24 well plates with or without 10 mm glass coverslips. Axenically grown stationary phase C. burnetii strains were quantified by qPCR using dotA specific primers [10] and diluted in DMEM with 5% FBS to an MOI of 50. Following a 4 h infection period, cells were washed once with PBS and incubated with fresh DMEM with 5% FBS. This point was considered Day 0 and a sample was collected to provide the inoculum amount of C. burnetii. Infection lysate was collected at the time of infection, 24 (Day 1), 72 (Day 3), 120 (Day 5) and 168 (Day 7) h after this initial time point. Genomic DNA was extracted from these samples using the Illustra Bacteria GenomicPrep Mini Prep Kit (GE Healthcare, Piscataway, NJ) and was used to quantify genomic equivalents by specific qPCR. In addition, replicate wells were fixed with 4% paraformaldehyde at Day 3 and Day 5 for subsequent immunofluorescence staining with anti-LAMP1 and anti-C. burnetii as described above.

BlA translocation assays

Translocation assays were performed as described previously [28,39]. Genes of interest were cloned into the SalI site of pJB-CAT-BlA and these constructs were introduced into C. burnetii NMII via electroporation. BlA fusion protein expression of isolated clones was confirmed by western blot with anti-BlA M (1:1000), (QED Bioscience Inc, San Diego, CA). 2 × 10^6 HeLa cells were plated in black clear bottom 96 well trays and, the following day, were infected with stationary phase C. burnetii NMII strains at an MOI of 100. The infection was allowed to proceed for 24 h before cells were loaded with the fluorescent substrate CCF4/AM according to the instructions for the LiveBLAzer-FRET B/G Loading Kit (Invitrogen, Carlsbad, CA). Fluorescence, with an excitation of 415 nm and emission at 460 nm and 355 nm, was quantified using a Tecan M1000 plate reader. The ratio of signal at 460 nm to 355 nm (blue:green) was calculated relative to uninfected cells. In addition, cells were visualized by fluorescence microscopy using an inverted Nikon Eclipse TE-2000 S microscope and a 20× objective.

RNA interference

In 24-well plates, HeLa 229 cells were reverse-transfected with small-interfering RNA (siRNA) SMARTpools specific for human ATG5 (NM_004849, ATG12 (NM_004707), STX17 (NM_017919), or STX18 (NM_016930) using Dharmafect-1 (Thermo Scientific) at a final concentration of 50 nM total siRNA in DMEM with 5% FBS. Transfected cells were incubated overnight, washed, and the adherent cells were subjected to a second round of siRNA transfection at the same concentration. After a two-day incubation, the cells were infected with C. burnetii NMII at a MOI of 50. At one day post-infection, the cells were lifted and replated at a lower cell density into a 24-well plate containing 12-mm-diameter glass coverslips and incubated for an additional two days. Cells were processed for immunofluorescence as described above.

Construction of plasmids encoding RavZ

Primers were designed to amplify ravZ or ravZC258A from plasmids pGFP-RavZ, or pGFP-RavZC258A [50] by PCR and to contain extended overhangs specific for sequence- and ligasation-independent cloning (SLIC) into the pJB-CAT-3×FLAG destination vector [72]. The following primers show the destination vector sequence underlined and the ravZ-specific sequences italicized: RavZ forw ard, 5′-ATATCGATTTAAGGATGCAGTAACAAGGTGCACGTGAATGCAAGATTACGGAGG-3′ and RavZ reverse, 5′-GGGGCCGGGTCTAAGAGGTGCGTGGGCTTACGTCGGACCTTTCTCATCTAAAGGACACACACCC-3′. The resulting vectors pJB-CAT3×FLAG-RavZ and pJB-CAT-3×FLAG-RavZC258A encode RavZ proteins that have three tandem copies of the FLAG epitope tag (3×FL) fused to the amino terminus of the protein.

Infection with C. burnetii expressing RavZ

Plasmid DNA (pJB-CAT-3×FLAG-RavZ or pJB-CAT-3×FLAG-RavZC258A) was electroporated into C. burnetii NM II, and chloramphenicol-resistant C. burnetii were clonally isolated as described previously [28]. Immunoblots of C. burnetii lysates using anti-Flag M2 antibody (Sigma) confirmed that 3×FL-RavZ protein was expressed. C. burnetii transformed with pJB-CAT-3×FLAG, pJB-CAT-3×FLAG-RavZ or pJB-CAT-3×FLAG-RavZC258A were used to infect HeLa cells at a MOI of 50. After 10 h incubation, cells were washed and incubated for three days before either fixation with 4% PFA for immunofluorescence, or lysis for immunoblot analysis.

LC3 immunoblot analysis

Uninfected or three-day post-infection C. burnetii-infected HeLa cells were lysed as described previously for Figure 5 [60]. Lysates were centrifuged and the supernatant separated by SDS-PAGE for immunoblot analysis using an anti-LC3 antibody (Novus) at 1:3000 and an anti-actin antibody (Sigma) at 1:5000. For Figure 6, uninfected or five-day post infection C. burnetii infected HeLa cells were maintained in 6-well dishes before harvesting with a cell scraper and lysing in buffer containing 2% Triton-X as described in Tanida et al., 2008 [73]. Cells were either left untrreated prior to lysis, or were incubated in media containing 200 nM rapamycin and 100 ng/ml bafilomycin A1 2 h prior to lysis.

Supporting Information

Figure S1 The CCVs formed by cig57::Tn and cig2::Tn contain the lysosomal protease cathespin D. HeLa 229 cells were infected with C. burnetii NMII, cig2::Tn or cig57::Tn at a multiplicity of 50 bacteria to 1 host cell. At 72 h post-infection the samples were fixed and stained with anti-cathepsin D (green),
anti-Coxiella (red) and Hoechst dye (blue). Representative images demonstrate that cathepsin D is localized to the CCVs formed by all three strains. Scale bars represent 10 μm. (PDF)

Figure S2 Cbu1780 is a novel Dot/Icm effector. The hypothetical proteins Cbu1780 and Cbu2072 were tested for Dot/Icm-dependent translocation using the β-lactamase translocation assay. C. burnetii NMIII and the icmL::Tn mutant expressing BlaM-1780 or BlaM-2072 from a plasmid were used to infect HeLa cells. At 24 h post-infection, CCF4-AM was incorporated into the cells and translocation was assayed through the quantitative measure of fluorescence at 460 nm and 535 nm (A) and visually (B). Translocation positive cells (blue) were only observed for C. burnetii NMIII expressing BlaM-1780. Results represent the mean 460::535 nm ratio ± standard deviation from three independent experiments and representative images. (PDF)

Table S1 Coxiella burnetii transposon mutants that do not replicate intracellularly. (DOCX)

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