Abstract

Background: Endothelin-I and angiotensin II are strong vasoconstrictors. Patients with ischemic heart disease have elevated plasma levels of endothelin-I and angiotensin II and show increased vascular tone. The aim of the present study was to examine the endothelin and angiotensin II receptor expression in subcutaneous arteries from patients with different degrees of ischemic heart disease.

Methods: Subcutaneous arteries were obtained, by biopsy from the abdomen, from patients undergoing coronary artery bypass graft (CABG) surgery because of ischemic heart disease (n = 15), patients with angina pectoris without established myocardial infarction (n = 15) and matched cardiovascular healthy controls (n = 15). Endothelin type A (ET\textsubscript{A}) and type B (ET\textsubscript{B}), and angiotensin type 1 (AT\textsubscript{1}) and type 2 (AT\textsubscript{2}) receptors expression and function were examined using immunohistochemistry, Western blot and in vitro pharmacology.

Results: ET\textsubscript{A} and, to a lesser extent, ET\textsubscript{B} receptor staining was observed in the healthy vascular smooth muscle cells. The level of ET\textsubscript{B} receptor expression was higher in patients undergoing CABG surgery (250\% ± 23\%; P < 0.05) and in the patients with angina pectoris (199\% ± 6\%; P < 0.05), than in the healthy controls (100\% ± 28\%). The data was confirmed by Western blotting. Arteries from CABG patients showed increased vasoconstriction upon administration of the selective ET\textsubscript{B} receptor agonist sarafotoxin S6c, compared to healthy controls (P < 0.05). No such difference was found for the ET\textsubscript{A} receptors. AT\textsubscript{1} and, to a lesser extent, AT\textsubscript{2} receptor immunostaining was seen in the vascular smooth muscle cells. The level of AT\textsubscript{1} receptor expression was higher in both the angina pectoris (128\% ± 25\%; P < 0.05) and in the CABG patients (203\% ± 41\%; P < 0.05), as compared to the healthy controls (100\% ± 25\%). The increased AT\textsubscript{1} receptor expression was confirmed by Western blotting. Myograph experiment did however not show any change in vasoconstriction to angiotensin II in CABG patients compared to healthy controls (P = n.s).

Conclusion: The results demonstrate, for the first time, upregulation of ET\textsubscript{B} and AT\textsubscript{1} receptors in vascular smooth muscle cells in ischemic heart disease. These receptors may play a role in the pathophysiology of ischemic heart disease and could provide important targets for pharmaceutical interventions.
Background
The renin-angiotensin and the endothelin systems are essential in vascular homeostasis and may become maladaptive in cardiovascular diseases [1]. Angiotensin II and endothelin-1 are formed in the endothelium and induce potent vasoconstriction and proliferation of vascular smooth muscle cells [2,3]. The continuous production of endothelin-1 and angiotensin II in the endothelium is important for the control of vessel tone and changes in the endothelin- and renin-angiotensin-systems can give rise to dysfunctional vessels such as those seen in patients with cardiovascular risk factors [4]. Endothelin-1 and angiotensin II have therefore been suggested to play a role in the development of cardiovascular diseases, including hypertension [5], chronic heart failure [6] and atherosclerosis [7].

Endothelin-1 mediates its effects through two distinct G-protein coupled receptors; the endothelin type A (ET\(_A\)) and type B (ET\(_B\)) receptors. During physiologic conditions, the ET\(_A\) receptor is the dominant receptor subtype expressed in vascular smooth muscle cells and mediates contraction, while the ET\(_B\) receptor is primarily located on endothelial cells and mediates vasodilatation via the release of nitric oxide and prostaglandins [8]. ET\(_B\) receptors on vascular smooth muscle cells have however been observed to be upregulated during pathological conditions such as atherosclerosis [9] and congestive heart failure [10]. Endothelin receptors on vascular smooth muscle cells are both mitogenic, leading to atherosclerosis and can induce strong vasoconstriction, resulting in elevated vascular tone that contributes to the development of ischemic cardiovascular disease.

Two angiotensin II receptors have been identified in man, AT\(_1\) and AT\(_2\) receptors, which are members of the G-protein coupled seven-transmembrane domain receptor family. The vascular effects of angiotensin II are primarily mediated by AT\(_1\) receptors located on smooth muscle cells which induce vasoconstriction and mitogenesis. Conversely, AT\(_2\) receptors are located on endothelial cells and are known to induce vasodilatation, inhibit cell growth and stimulate apoptosis [11]. AT\(_2\) receptors have been shown, although to a lesser extent, in vascular smooth muscle cells. Angiotensin II acts, apart from being a potent vasoconstrictor also as a growth factor that regulates cell growth, differentiation and fibrosis, as well as being implicated in the pathology of heart failure, hypertension and atherosclerosis [11].

In vivo studies on the effects of endothelin-1 and angiotensin II in the human peripheral vasculature have previously mainly been performed using a forearm blood flow model. To the best of our knowledge, this is the first in vitro study using peripheral vascular tissue samples isolated from patients with different degrees of cardiovascular disease. The peripheral vasculature is contributing significantly to total peripheral resistance leading to our use of small peripheral arteries and arterioles, obtained from the subcutaneous tissue in patients. Patients with angina pectoris without established myocardial infarction, patients undergoing coronary artery bypass graft (CABG) surgery because of ischemic heart disease and cardiovascular healthy controls, were included in the study. The expressions of endothelin ET\(_A\) and ET\(_B\), and angiotensin AT\(_1\), and AT\(_2\) receptors in vascular smooth muscle cells were studied by using immunohistochemistry, Western blot and in vitro pharmacology.

Methods

Ethics
The project was approved by the Ethics Committee of Lund University in Sweden (LU Dnr: 308/2004) and conforms to the principles outlined in the Declaration of Helsinki. Each individual provided written consent to the procedure.

Study groups
Three different groups of patients were included in the immunohistochemistry part of the study.

1. Cardiovascular healthy controls (controls). This group consisted of ten healthy volunteers without previous history of chest pain, cardiovascular disease or prior cardiac medications. Blood samples for laboratory analysis were taken prior to the biopsy.

2. Fifteen patients with angina pectoris without established myocardial infarction (patients with angina pectoris). These patients were admitted to the medical emergency unit with angina pectoris without signs of prior or ongoing myocardial infarction as measured by electrocardiogram and biomarkers for myocardial injury. The patients had no history of heart failure. Blood samples for laboratory analysis were taken at the telemetry unit after the chest pain had resolved.

3. Patients undergoing coronary artery bypass graft surgery (CABG) (n = 10). These patients underwent elective CABG because of stable ischemic heart disease confirmed by angiography revealing coronary disease in three arteries. The patients had no prior known myocardial infarction but showed different degrees of heart failure secondary to their heart vessel disorder. Blood samples for laboratory analysis were taken the day before the surgical intervention.

The patient background characteristics are defined in Table 1.
cold isopentane and stored at -80°C. The tissue was frozen in ice-water. The biopsies were stored at -80°C prior to sectioning into cutaneous tissue (1 cm³), containing small arteries, was undergoing CABG surgery the biopsy was taken under general anaesthesia during the initiation of surgery. A subcutaneous tissue of the abdomen. For the control group and the patient with angina pectoris, this was done during local anaesthesia (2 ml, 1% Xylocain; AstraZeneca, Sweden). For the patients undergoing CABG surgery the biopsy was taken under general anaesthesia during the initiation of surgery. A subcutaneous tissue (1 cm³), containing small arteries, was removed from the abdomen. The tissue was frozen in ice-cold isopentane and stored at -80°C.

**Tissue collection**

A biopsy was taken from the subcutaneous tissue of the abdomen. For the control group and the patient with angina pectoris, this was done during local anaesthesia (2 ml, 1% Xylocain; AstraZeneca, Sweden). For the patients undergoing CABG surgery the biopsy was taken under general anaesthesia during the initiation of surgery. A subcutaneous tissue (1 cm³), containing small arteries, was removed from the abdomen. The tissue was frozen in ice-cold isopentane and stored at -80°C.

**Immunohistochemistry**

The biopsies were stored at -80°C prior to sectioning into 8-μm-thick slices in a calibrated Microm HM500M cryostat (Microm). The sections were fixed and rehydrated in phosphate-buffered saline (PBS). The sections were incubated overnight with rabbit anti-human ETB receptor agonist, sarafotoxin 6c at increasing concentrations (10⁻¹¹-10⁻⁶ M) and returned to baseline and endothelin-1 was then added at increasing concentrations (10⁻¹¹-10⁻⁶ M) when endothelin ETB receptors were desensitized [13] facilitating endothelin-1 to act solely on the endothelin ETB receptors. For details see Nilsson et al. [14].

**In vitro pharmacology**

The resistance arteries were removed from the abdomen from cardiovascular healthy patients undergoing abdominal reduction surgery (n = 5) or during CABG surgery (n = 5). The subjects had the same clinical characteristics as those for immunohistochemistry (Table 1). Vessels were placed in saline buffer on ice, transported to the laboratory and subsequently analyzed for their contractile properties in a myograph recording their isometric tension. The vessels were cut into 1-mm-long cylindrical segments and mounted on two L-shaped metal prongs, unilaterally a force displacement transducer continuously recorded the isometric tension [12]. Mounted vessel segments were immersed in vessel baths at 37°C containing a bicarbonate based buffer solution; NaCl (119 mM), NaHCO₃ (15 mM), KCl (4.6 mM), CaCl₂ (1.5 mM), MgCl₂ (1.2 mM), NaH₂PO₄ (1.2 mM), and glucose (5.5 mM). The buffer was continuously aerated with oxygen enriched with 5% CO₂ resulting in pH 7.4. Initially vessels were stretched to a resting tone of 2 mN and was then allowed to stabilize at this tension for 1 h. Contractile capacity of each arterial vessel segment was controlled by exposure to a potassium rich (63.5 mM) buffer solution. Concentration-response curves were obtained by cumulative application of endothelin ET₄ receptor agonist, sarafotoxin 6c at increasing concentrations (10⁻¹¹-10⁻⁶ M), ET-1 (AnaSpec, San Jose, CA) in the concentration range 10⁻¹⁴ to 10⁻⁷ M and angiotensin II in the concentration range 10⁻¹² to 10⁻⁶ M. Before the application of angiotensin II, the arteries were pretreated with the AT₁ receptor antagonist PD-123319 (10⁻⁵.5 M) for 30 min. After washout, the vessels returned to baseline and endothelin-1 was then added at increasing concentrations (10⁻¹¹-10⁻⁶ M) when endothelin ETB receptors were desensitized [13] facilitating endothelin-1 to act solely on the endothelin ETB receptors. For details see Nilsson et al. [14].

**Table 1: Physical and Biochemical Characteristics in the Study Subjects**

| Variable                  | Control n = 10 | Angina n = 15 | CABG n = 10 |
|---------------------------|----------------|--------------|-------------|
| Age (years)               | 62 ± 13        | 64 ± 13      | 70 ± 5      |
| Smokers                   | 2              | 4            | 1           |
| SBP (mmHg)                | 125 ± 9        | 142 ± 15*    | 140 ± 14*   |
| DBP (mmHg)                | 72 ± 8         | 76 ± 9       | 78 ± 12     |
| Total cholesterol (mg/dl) | 4.9 ± 1.6      | 5.3 ± 1.2    | 4.4 ± 2.4   |
| LDL (mg/dl)               | 3.5 ± 0.7      | 3.5 ± 1.1    | 2.5 ± 1.6   |
| HDL (mg/dl)               | 1.5 ± 0.5      | 1.5 ± 0.4    | 1.3 ± 0.5   |
| Triglycerides (mg/dl)     | 1.5 ± 0.9      | 1.7 ± 1.0    | 1.9 ± 0.9   |
| Apo lipoprotein A (mg/dl)| 1.5 ± 0.3      | 1.6 ± 0.2    | 1.3 ± 0.5   |
| Apo lipoprotein B (mg/dl)| 0.9 ± 0.2      | 1.0 ± 0.3    | 0.9 ± 0.3   |
| NYHA class                |                |              |             |
| I                         | 10             | 10           | 4           |
| II                        | 0              | 0            | 4           |
| III                       | 0              | 0            | 2           |
| IV                        | 0              | 0            | 0           |
| NT pro-BNP (ng/L)         | 106 ± 112      | 218 ± 243    | 1626 ± 2157*|
| C reactive protein (mg/dl)| 2.1 ± 1.9      | 6.2 ± 15.3*  | 6.9 ± 6.1*  |
| Beta blockers (n)         | 0              | 6            | 7           |
| Statins (n)               | 0              | 5            | 8           |
| ARB or ACEI (n)           | 0              | 3            | 6           |
| HbA1c (%)                 | 4.5 ± 0.5      | 4.7 ± 0.4    | 4.7 ± 0.8   |
| BMI (kg/m²)               | 28 ± 2         | 29 ± 5       | 27 ± 3      |

Values are expressed as mean ± SD. Statistical analysis was performed using ANOVA with Dunnet’s post-test for multiple comparisons. Significance was defined as P < 0.05 (*). Abbreviations are: CABG, coronary artery bypass graft surgery; SBP, systolic blood pressure; DBP, diastolic blood pressure; LDL, low-density lipoprotein cholesterol; HDL, high-density lipoprotein cholesterol; NYHA, New York Heart Association Functional Classification; NT-proBNP, N-terminal pro B-type natriuretic peptide; ARB, angiotensin receptor blockers; ACEI, angiotensin converting enzyme inhibitors and BMI, body mass index.
The drugs used, endothelin-1 (ET-1), sarafotoxin 6c (S6c), angiotensin II and PD-123319 were purchased from Sigma Chemical Co (St. Louis, MO). They were dissolved in 0.9% NaCl with 10% albumin.

**Western Blot Analysis**

Human vessels (4 from healthy donators and 5 from CABG patients) were frozen in liquid nitrogen and homogenized in cell extract denaturing buffer (BioSource, USA) with addition of a protease inhibitor cocktail (Sigma, USA). Whole cell lysates were sonicated for 2 min on ice, centrifuged at 15,000 × g at 4°C for 30 min, and the supernatants were collected as protein samples. The protein concentrations were determined using the protein assay reagents (Bio-Rad, Hercules, CA, USA) and stored at -80°C until immunoblotting assay. The protein homogenates were diluted 7:2:1(v/v) with 5× LDS Sample Buffer and 10× DTT Reducer (PAGEgel, Inc San Diego, CA, USA). A total of 4050 μg of protein was boiled for 5 min and separated by 412% SDS Ready Gels (PAGEgel, Inc., USA) for 120 min at 100 v, and transferred electrophoretically to nitrocellulose membranes (Bio-Rad) at 100 v for 60 min. The membrane was then blocked for 1 h at room temperature with phosphate buffered saline (PBS) containing 0.1% Tween-20 (Sigma) and 5% non-fat dried milk, and incubated with sheep anti-ETB (Enzo Life Sciences Inc., Farmingdale, NY, USA), rabbit anti-AT1 (Abcam plc., Cambridge, UK), or rabbit anti-β-actin (Cell Signaling Technology, Inc., Boston, MA, USA) polyclonal primary antibodies diluted 1:200-500 overnight at 4°C, followed by incubation with anti-sheep (Abcam plc., UK) or rabbit anti-rabbit (Amersham Biosciences, Piscataway, NJ, USA) IgG, horseradish peroxidase (HRP)-conjugated secondary antibodies diluted 1: 5000 for 1 h at room temperature. The probed proteins were developed by Lumisensor Chemiluminescent HRP Substrate kit (GenScript Corp., Piscataway, NJ, USA). To detect multiple signals using a single membrane, the membrane was incubated for 515 min at room temperature with restored plus Western blot stripping buffer (Pierce Biotechnology, Inc., Rockford, IL, USA). The membranes were visualized using a Fujifilm LAS-1000 Luminiscent Image Analyzer (Stamford, CT, USA.). The quantification of band intensity was analyzed with Image Gauge Ver. 4.0 (Fuji Photo Film Co., LTD., Japan). The expression of target proteins were presented as a relative extent to the level of β-actin and normalized to the percentage of control.

**Calculations and statistics**

**Immunohistochemistry**

The experiments were performed using arteries from 15 patients with angina pectoris, 10 patients undergoing CABG surgery and 10 healthy controls. The samples were examined in a fluorescence microscope (Olympus optical Co, LTD, Bx60F5) and the absolute fluorescence intensity was measured with ImageJ http://rsb.info.nih.gov/ij/. Mean fluorescence values in the area selected were obtained in three sections and at four different regions, and the mean values were calculated. Statistical analysis was performed using ANOVA with Bonferroni’s or Dunnett’s post-test for multiple comparisons. Significance was defined as P < 0.05 (*). The data are given in the text as percentage difference relative to the mean fluorescence seen in the control. The results are presented as mean values ± standard deviation (SD) and N equals number of patients. Values are presented as percent fluorescence in the angina and CABG groups compared with the control patients group, where the control group is set to 100%.

**In vitro pharmacology**

The contractile reaction in each segment is expressed as percentage of the 63.5 mM potassium induced contraction. E\text{max} value represents the maximum contractile response elicited by an agonist, and the pEC50, the negative logarithm of the drug concentration that elicited half the maximum response. Linear regression analysis was applied using values obtained from above and below half-maximum response for calculating the negative logarithm of the concentrations that produced 50% contraction (pEC50). Data are presented as mean ± standard error of the mean (S.E.M). Statistical analyses was performed using Student’s t-test.

**Western Blot**

Comparisons between two groups were performed using two-tailed unpaired Student’s t-test. A p-value less than 0.05 were considered to be significant. Results were presented as mean ± S.E.M.

**Results**

**Endothelin receptor expression**

In healthy controls, the vascular smooth muscle cells stained for ET\text{A} and, to a lesser extent, ET\text{B} receptors. The immunostaining intensities for ET\text{B} receptors were higher in arteries from the patients undergoing CABG surgery (250% ± 23%; P < 0.05) and from the patients with angina pectoris (199% ± 6%; P < 0.05), than from the healthy controls (Figure 1 and 2). Furthermore, the levels of ET\text{B} receptor expression were higher in the smooth muscle cell layer in the arteries from the patients undergoing CABG surgery than in the arteries from the patient with angina pectoris (Figure 1B). The levels of ET\text{B} receptor expression, in the smooth muscle cells, were similar in all three groups (P = n.s.). In endothelial cells ET\text{B} receptor immunostaining was primarily observed, however, it was not different among the groups.

Western blot for ET\text{B} receptors detected a single band with the approximate molecular weight of 53 kDa in the artery homogenates from all samples including healthy controls and patients undergoing CABG surgery (Figure 3). Densitometric analysis showed that patients undergoing CABG
surgery had a mean increase of ETB receptors expression (133 ± 8%; P < 0.05), which was significantly more than in healthy controls (Figure 3).

**Angiotensin II receptor expression**
The vascular smooth muscle cells stained for AT1 and, to a lesser extent, for AT2 receptors. The levels of AT1 receptor expression were higher in both the angina pectoris (128% ± 25%; P < 0.05) and in the CABG patients (203% ± 41%; P < 0.05), than from the healthy controls (Figure 2 and 4). In addition, the levels of AT1 receptor expression were higher in the smooth muscle cell layer in arteries from patients undergoing CABG surgery than in arteries from patient with angina pectoris (P < 0.05). AT2 receptor expression was similar in all three groups. In endothelial cells, only AT2 receptor immunostaining was observed. Since the numbers of cells in the endothelial cell layer are limited, no comparisons were made between the groups. Western blot for AT1 receptors detected a single band with the approximate molecular weight of 41 kDa in the artery homogenates from all samples including healthy controls and patients undergoing CABG surgery (Figure 3). Densitometric analysis showed that patients undergoing CABG surgery had a mean increase of AT1 receptors expression (137 ± 9%; P < 0.05) which was significantly more than in healthy controls (Figure 3).

**Actin and CD-31 immunostaining**
Double immunostaining showed co-localization between on one hand the AT1 and ETB receptors and on the other hand smooth muscle cell alpha-actin in resistance arteries. As shown in the illustration (Figure 5) there was clear co-localization between the two receptor subtypes and alpha-actin both in control and in CABG subjects. CD-31 was used to stain endothelial cells; we observed weak co-localization between both receptor and CD31 (Figure 5).

**In vitro Pharmacology**
The selective ETB receptor agonist S6c was used to study ETB mediated contraction. Afterwards ET-1 induced vasoconstriction was studied when endothelin B receptors were desensitized by the previous application of S6c. The selective ETB receptor agonist S6c was used to study ETB mediated contraction. Afterwards ET-1 induced vasoconstriction was studied when endothelin B receptors were desensitized by the previous application of S6c.
Sarafotoxin 6c induced no vasoconstriction in healthy control vessels. On the other hand patients undergoing CABG showed a fairly strong and significant contractile response to the cumulative application of S6c (P < 0.05 n = 5) suggesting upregulation of ETB receptors at the functional level (Figure 6A). Endothelin-1 induced a potent vasoconstriction in both healthy controls and in patients undergoing CABG surgery (P = ns) indicating that there were no obvious changes in the ETA receptor responses between the two groups (Figure 6B). Angiotensin II induced concentration-dependent contractions of the arteries were strong but there were no differences between the groups (Figure 6C). The potassium rich buffer (124 mM) resulted in strong persistent contractions of the resistance arteries with no differences between the groups (data not shown).

**Demographics**

The patients’ background characteristics are described in Table 1. Taken together, the systolic blood pressure and C reactive protein were higher in the patient with ischemic heart disease (both CABG and angina pectoris). The plasma levels of N-terminal pro B-type natriuretic peptide (NT-proBNP) were higher in the CABG and the angina pectoris groups than in the cardiovascular healthy controls.

**Discussion**

**Endothelin receptor expression and function**

Our results clearly show that the level of ETB receptor expression is higher in the vascular smooth muscle cell layer of arteries from patients with angina pectoris than from healthy controls. The levels of ETB receptor expression are even higher in arteries from patients undergoing CABG surgery. We hypothesize that there is a progressive upregulation of this receptor subtype that may correlate with the degree of cardiovascular disease.

Upregulated ETB receptors on vascular smooth muscle cells have previously been shown in diabetes, hypertension and in human atherosclerotic coronary arteries and in atherosclerotic plaques [9,15,16]. Plasma levels of ET-1 are elevated in patients with ischemic heart disease and in
heart failure; it has been suggested as a prognostic marker [17]. The circulating ET-1 levels are further increased in patients undergoing CABG surgery [18]. This increased activity in the endothelin system may contribute to smooth muscle cell proliferation, vasoconstriction and decreased perfusion in atherosclerotic disease [19-21]. Furthermore, CABG patients had significantly increased pro-BNP levels indicating some degree of heart failure, a condition associated with increased ETB receptor mediated systemic vasoconstriction [22]. The elevated endothelin ETB contraction in the present study, also verified by Western blot, indicates that the increased levels of contractile endothelin ETB receptors on the vascular smooth muscle cells could play an important role in ischemic heart disease since ETB receptor activation in healthy controls did not induce any contractions at all.

Mechanisms underlying the upregulation of ETB receptors
The mechanism underlying the increased ETB receptor expression is not known, but could depend on increased transcription of ETB receptor mRNA triggered by some of the many humoral factors that are increased in ischemic heart disease. Upregulation of ETB receptors is known to rely on increased transcription and subsequent translation of receptor mRNA [23]. In the human genome, the 5’-flanking region of the genes encoding the endothelin receptors contain several regulatory elements, like GATA-motifs and E-boxes [24,25]. This indicates that the genes might be activated by for example inflammatory components. Indeed, the presence of interleukin-1β and TNF-α may enhance the upregulation of vascular ETB receptors [26]. In agreement, the patients with ischemic heart disease had higher plasma levels of C reactive protein. Atherosclerosis is known to induce a proinflammatory response [27] and this may in part regulate endothelin receptor expression.

Angiotensin II receptor expression
In the present study, the levels of AT1 receptor expression were higher in subcutaneous arteries from patients with ischemic heart disease than in the healthy controls as verified by immunohistochemistry and Western blot. The levels of expression were even higher in arteries from patients undergoing CABG surgery. The plasma level of angiotensin II is known to be elevated in different heart conditions, such as heart failure [28], hypertension [29], hypoxia [30], hypercholesterolemia [31] and hyperglycemia [32]. We have found, herein, that this may be associated with enhanced expression of vascular angiotensin II receptors. Angiotensin II is a potent vasoconstrictor and mitogen of coronary artery smooth muscle cells. The stimulation of AT1 receptors results in progression of atherosclerotic lesions, inflammation and plaque rupture. Increased expression of AT1 receptors might make the vasculature prone to develop spasm and atherosclerotic plaques, and thus further increase peripheral vascular resistance and reduce lumen. However, we did not observe any enhanced contraction of angiotensin II mediated via AT1 receptors as a result of an increased expression in the myograph experiments. One explanation to this could be the increased use of AT1 receptor antagonists in patients undergoing CABG, thus blocking the AT1 receptors. It is important, however, not to forget that AT1 receptors apart from their contractile properties also possess other abilities such as regulating cell growth, differentiation and fibrosis which are important in the pathology of heart failure, hypertension and atherosclerosis.
Figure 5
Co localization of the AT₁ and ET₄ receptors to vascular smooth muscle cells and endothelial cells. Immunostaining and co localization of the AT₁ and ET₄ receptors to alpha-SMC and endothelin in resistance arteries of control patients. Antibodies to AT₁ (green, A1, arrow) and smooth muscle actin (red, A2 arrow), A3 (merged A1 and A2). Antibodies to ET₄ (green, B1, arrow) and smooth muscle actin (red, B2, arrow), B3 (merged B1 and B2). Antibodies to AT₁ (green, C1, arrow) and CD-31 (red, C2, arrow), C3 (merged C1+C2). Antibodies to ET₄ (green, D1, arrow) and CD-31 (red, D2, arrow), D3 (merged D1 and D2). Magnification x 400 for all photomicrographs.
Mechanisms underlying the upregulation of AT_1 receptors
The mechanisms underlying the upregulation of AT_1 receptor expression in patients with ischemic heart disease cannot be deduced from the present study. Inflammatory mediators, such as IL-1 and IL-6, may upregulate AT_1 receptors and enhance angiotensin II stimulated vessel hypertrophy [33]. In the present study, the observed plasma levels of C reactive protein was elevated in patients with ischemic heart disease. It has previously been shown that high levels of C reactive protein correlate with high levels of AT_1 receptor expression [34]. Increased AT_1 receptor expression has therefore been associated with vascular inflammation. C reactive protein is known to independently predict risk for myocardial infarction, stroke, peripheral artery disease and sudden cardiac death even among apparently healthy individuals [35].

Clinical perspective
Both ET-1 and angiotensin II have strong impact on cardiovascular diseases as discussed above; herein we will discuss their involvements in other situations: CABG surgery is hampered by deleterious vasospasm in arterial grafts. It has been suggested that impaired vasomotor function may contribute to this vasospasm [36]. Plasma levels of ET-1 and angiotensin II are elevated during CABG surgery [18]. Both endothelin and angiotensin II receptors have been shown to mediate strong vasoconstrictor effects in bypass grafts [28]. Many measures have been undertaken to control the vessel tone during surgery, including dilating the vessels with potent vasodilators such as papaverine, sodium nitroprusside and nifedipine, or mechanically distending the vessels with saline. It might be important to consider endothelin and angiotensin II receptor antagonists for this indication especially since the present results show increased levels of endothelin and angiotensin II receptor expression in patients undergoing CABG surgery.

Angiotensin converting enzyme inhibitors (ACEI) and angiotensin AT_1 receptor antagonists are frequently being used to prevent the development of target organ damage in atherosclerotic disease and in hypertension [37]. Whether endothelin receptor antagonists will become part of the therapeutic armamentarium in ischemic heart disease remains unclear, and none of these agents is currently being developed for this indication. However, the mixed endothelin blocker Bosentan is used in treatment of pulmonary hypertension [38] and other non-selective ET receptor antagonists have been shown to inhibit the development of atherosclerosis in experimental models [39]. New endothelin antagonists devoid of side effects and perhaps specific for either of the receptor subtypes are discussed in treatment of vasospasm after subarachnoid haemorrhage [40], or alternative inhibitors of the endothelin converting enzymes that generate endothelin-1 may in the future become available to block the endothelin system [5].

Limitations
The present study has demonstrated increased expression of ET_0 and AT_1 receptors in patients with ischemic heart disease. However, the patients have confounding factors. The systolic blood pressure was higher in patients with ischemic heart disease compared to the healthy controls. Furthermore, the prevalence of medication with statins, beta blockers, ACEI, and AT_1 receptor antagonists was
common in patients with ischemic heart disease while they were non-existing in our healthy controls. The effect of these confounding factors on the endothelin and angiotensin II receptor expression cannot be concluded from the present study.

Conclusion
The present study clearly shows that patients with ischemic heart disease have upregulated ET$_{B}$ and AT$_{1}$ receptors in the smooth muscle cells of peripheral resistance arteries. The level of ET$_{B}$ and AT$_{1}$ expression correlates with the degree of ischemic heart disease, being highest in the patients undergoing CABG surgery. There was no difference in the expression of ET$_{A}$ or AT$_{2}$ receptors between the study groups. The changes in ET$_{B}$ and AT$_{1}$ receptor expression may be signs of remodeling of the vasculature, which is characteristic of ongoing ischemic heart disease.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
ID planned and executed the immunofluorescence experiments, analyzed the data, preformed the surgical procedure and wrote the manuscript. MLE acquired the patient data, the biopsies and reviewed the manuscript. QC preformed the Western blot experiments and wrote the associated texts. MM participated in the writing of the manuscript. POK performed the thoracic surgical procedure, removed the vessels and reviewed the manuscript. LE conceived and planned the study, and carefully reviewed the data and assisted in writing of the manuscript.

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