Dexamethasone inhibits NF-κBp65 and HMGB1 expression in the pancreas of rats with severe acute pancreatitis

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Abstract. Severe acute pancreatitis (SAP) starts as a local inflammation of pancreatic tissue that induces the development of multiple extra-pancreatic organ dysfunction; however, the underlying mechanisms remain unclear. The present study was designed to evaluate the effect of dexamethasone (DXM) on pancreatic damage and to investigate the role of high-mobility group box-1 (HMGB1) and nuclear factor-κB (NF-κBp65) in the development of SAP in animal and cell models. For the in vivo experiment, 35 Sprague-Dawley rats were randomly assigned to three groups: The sham-operation control group, the SAP group and the DXM treatment group. Histological analysis revealed that, when DXM was infused into SAP rats, edema formation and structural alterations with necrosis were reduced, and the number of apoptotic cells was markedly reduced. In addition, compared with the SAP group, the expression level of HMGB1 was significantly decreased in the nucleus and the expression level of NF-κBp65 was significantly decreased in the cytoplasm from rats treated with DXM. In vitro, DXM was able to suppress the apoptosis and cell death induced by caerulein (CAE), and DXM could suppress the expression of NF-κBp65 and HMGB1 induced by CAE, as demonstrated by western blotting and immunofluorescence analysis. Therefore, these results provide an experimental basis for investigating the underlying therapeutic mechanisms of DXM treatment for SAP.

Introduction

Acute pancreatitis is a relatively common disease and severe acute pancreatitis (SAP) is associated with a high mortality rate, ranging from 15-40% (1). SAP starts as a local inflammation of pancreatic tissue and is characterized by the development of systemic inflammatory response syndrome (SIRS) and multiple organ dysfunction (2,3). Unfortunately, despite several years of experimental and clinical research, the precise pathophysiology of SAP, particularly in the clinical context remains unclear. This has precluded the development of definitive treatment modalities for this potentially life-threatening illness.

Experimental and clinical studies over the past decade have reported the profiles of intrapancreatic and circulating cytokines, chemokines, adhesion molecules, transcription factors including nuclear factor (NF)-κB and high-mobility group box 1 (HMGB1), the nucleotide-binding oligomerization domain receptor, Toll-like receptor (TLR)9 (6) and protective pathways, including the heme oxygenase-1 pathway (7). The HMGB1 protein, as a late mediator of endotoxin lethality, was demonstrated to exhibit a delayed release by cultured macrophages of more than 8 h following stimulation with endotoxin, tumor necrosis factor (TNF), or interleukin (IL)-15 (11,12). Furthermore, previous studies have demonstrated that HMGB1 may serve an important role in SAP (9,11) and it may act as a key inflammatory mediator that participates in the development of SIRS and multiple organ damage in SAP (10). HMGB1, TLR4 and NFκB are possible therapeutic targets for SAP treatment; therefore, HMGB1 antagonists, TLR4 antagonists and NFκB inhibitors should be considered. TLR4 antagonists are the closest to being used in a clinical setting for SAP treatment. Two TLR4 antagonists, namely VGX-1027 and eritoran are already in clinical development (13-16).

The inhibition of HMGB1 by sodium butyrate has been reported to have beneficial effects on SAP development;
however, sodium butyrate is not a specific HMGB1 inhibitor (17).

Dexamethasone (DXM), a type of steroid medication, can improve microcirculation and inhibit enzymes and inflammatory mediators (18). It has been used to treat SAP, but its protective effects and associated mechanisms on pancreatic injury remain unclear. In the present study, it was hypothesized that HMGB1 and NF-κBp65 are involved in the therapeutic mechanism through which DXM acts on SAP. Experiments were performed to investigate the influence of DXM on the expression levels of NF-κBp65 and HMGB1, as well as on apoptosis, in pancreatic cells of SAP rats, to observe the therapeutic efficacy and investigate the underlying therapeutic mechanisms of DXM treatment.

Materials and methods

Materials and reagents. Sodium taurocholate solution (TCA) was obtained from Beijing Solarbio Science & Technology Co., Ltd. (Beijing, China) and DXM injections were obtained from Hubei Tianyao Pharmaceutical Co., Ltd. (Fancheng, China). Chloral hydrate was purchased from Sangon Biotech Co., Ltd. (Shanghai, China). The HMGB1 antibody was from Abcam (Cambridge, UK) and the NF-κB p65 antibody was from Cell Signaling Technology, Inc. (Danvers, MA, USA).

Animals. In total, 35 male Sprague Dawley rats, 6-8 weeks old and weighing between 200-250 g, were purchased from Hunan Slake Jingda Experimental Animal Co. Ltd. (Changsa, China). The study was approved by the Ethics Committee of Central South University (Changsa, China).

Experimental model and groups. Rats were raised in rooms with a 12-h light/dark cycle at 25°C for 1 week prior to the experiment. Rats were fasted for 12 h and given food and fresh tap water ad libitum up to the experiment. Anesthesia was administered by intraperitoneal injection of 10% chloral hydras (0.3 ml/100 g). The SAP model was induced by the standard retrograde infusion of a freshly prepared 5% TCA into the biliopancreatic duct following laparotomy. The 35 rats were randomly divided into three groups: (i) Sham-operation control group (sham group, n=5), where rats received an equivalent volume of normal saline following a successful sham operation; (ii) SAP model group (SAP group, n=15), where rats received an equivalent volume of 5% TCA; and (iii) DXM treatment group (DXM group, n=15), where rats were administered one dose of DXM (0.5 mg/100 g) intravenously in the tail following successful SAP induction. The rats were sacrificed by dislocation of the neck in the state of anesthesia at designated time-points following the induction of pancreatitis.

Histological examination. For histological analysis, pancreas tissue specimens were fixed in 4% formaldehyde at room temperature for 3 days, embedded in paraffin, sectioned at 4 μm and treated with hematoxylin-eosin (HE) staining for 10 min at room temperature. The terminal deoxynucleotidyl transferase (TdT) dUTP nick-end labeling (TUNEL) assay was used to examine the apoptotic cells in the pancreas. Following staining with 0.5% hematoxylin for 15 min at room temperature, the nuclei of healthy cells were stained blue, while those in apoptotic cells presented brown/yellow staining. Integrated optical density (IOD) analysis was used to indirect reaction the apoptosis. The expression levels of HMGB1 and NF-κBp65 in the pancreas were examined by immunohistochemistry (IHC). Specimens were mounted in Permount and examined using routine light microscopy.

Cell culture. AR42J pancreatoma cells were obtained from American Type Culture Collection (Manassas, VA, USA) and cultured in a humidified atmosphere with 5% CO₂ at 37°C. The cells were treated with 10⁻⁸ mol/L caerulein (CAE) and/or DXM. Experiments were assigned to three groups: CK group, CAE group and DXM group. In the DXM group, cells were co-treated with CAE and 10⁻⁶, 10⁻⁷ or 10⁻⁸ mol/L DXM for 24 h. In the CK group, which acted as the control group, the AR42J cells were only treated with PBS. Cells in the CAE group were treated with CAE alone.

Detection of cellular apoptosis assay. For this experiment, 2x10⁶ cells were plated into 60-mm dishes and then treated with or without CAE/DXM for 24 h. Cellular apoptosis was detected using the AnnexinV-fluorescein isothiocyanate Apoptosis kit (BD Biosciences, Franklin Lakes, NJ, USA) and a FACScanto II flow cytometer (BD Biosciences), following the manufacturer’s protocol.

Cell survival assay. AR42J cells (5x10⁵ cells/well) were seeded into a 96-well plate and a MTT-based assay was performed after 24 h. DMSO (150 μl) was used to dissolve the purple formazan and the absorbance at 570 nm was measured to determine the cell survival rates.

Immunofluorescence assay. Immunofluorescence was performed on AR42J cells following blocking with 1X PBS/5% Normal Goat Serum (005-000-121; Jackson ImmunoResearch Laboratories, Inc., West Grove, PA, USA)/0.3% Triton X-100 at room temperature for 60 min using anti-HMGB1 (1:500; cat. no. ab79823; Abcam), and anti-NF-κBp65 (1:500; cat. no. 8242; Cell Signaling Technology, Inc.) primary antibodies and cy3-conjugated secondary antibodies (1:500; Abcam). Nuclei were counterstained with DAPI (1:1,000; Sigma Aldrich; Merek KGaA, Darmstadt, Germany) for 1 h at room temperature and cover slips were mounted with Fluorescence Mounting Medium (Dako; Agilent Technologies, Inc., Santa Clara, CA, USA) and examined using a fluorescence microscope.

Western blotting. AR42J cells were harvested using RIPA buffer (002A; Auragene; Hunan Aijia Biotechnology Co., Ltd., Hunan, China) and a BCA Protein Quantitation kit used for protein determination. Proteins were subjected to SDS-PAGE and immunoblotting, as previously described (19). A total of 30 μg protein per lane was separated by 10% SDS-PAGE, transferred onto polyvinylidene fluoride membranes (EMD Millipore, Billerica, MA, USA), blocked with 5% skimmed milk for 1 h at room temperature and incubated with primary antibodies at 4°C overnight: Anti-HMGB1 (1:1,0000; cat. no. ab79823; Abcam), anti-NF-κBp65 (1:1,1000; cat. no. 8242; Cell Signaling Technology, Inc.) and anti-β-actin (1:1,1000; cat. no. 3700; Cell Signaling Technology, Inc.). Then the blots were incubated with the corresponding secondary antibodies.
Compared with the CK group, cell survival was significantly improved by DXM treatment (P<0.05). Additionally, a positive association was demonstrated between cell viability and DXM concentration (Fig. 3C).

Effect of DXM on HMGB1 and NF-κBp65 expression in AR42J cells. Immunofluorescence (Fig. 4A) and western blotting (Fig. 4B) analyses demonstrated that HMGB1 and NF-κBp65 were expressed at low levels in AR42J cells treated with PBS (CK group), were expressed at a high level in the CAE group, and were expressed at an intermediate level in the DXM and CAE co-treated group. The expression of HMGB1 and NF-κBp65 decreased in a dose-dependent manner with DXM treatment and reached the lowest levels when treated with 10⁻⁶ M DXM, compared with the CAE group (Fig. 4).

Discussion

SAP is an acute, critical illness with rapid onset, long duration and rapid progress that results in massive necrosis of pancreatic tissue, extrapancreatic multiple organ failure, and a high mortality rate. Despite advances in treatment techniques, the mortality of SAP has improved slowly over the past several decades and the pathogenesis of SAP has not been completely clarified (20,21). However, experts have focused on inflammatory mediators and their corresponding antagonists as potential therapeutic targets, due to their contribution to the injury of the pancreas and other organs, frequently resulting in patients succumbing to the condition.

DXM is a long-acting corticosteroid. Its therapeutic effects on SAP are primarily associated with suppressing the production and/or actions of inflammatory mediators, enhancing the anti-stress capacity of the body, reducing endotoxemia, and scavenging oxygen free radicals (22,23). In the present study, to investigate the protective effects of DXM on the pancreas of SAP rats the effect of DXM treatment on the expression levels of NF-κBp65 and HMGB1, as well as on the histopathology of the pancreas in rats with SAP, was investigated.

NF-κBp65 is a transcription factor that regulates various genes involved in inflammatory and immune responses, including cytokine and adhesion molecules (24). TNF-α expression is directly regulated by NF-κBp65, as there are NF-κBp65 binding sites on the TNF-α promoter. Research has demonstrated that the inhibition of NF-κBp65 can result in decreased expression levels of cytokines, including TNF-α, reducing the inflammatory response in organisms. The role of NF-κBp65 activation in the pathogenesis of SAP was previously reviewed (25) and in a later investigation, this laboratory demonstrated evidence that the upregulation of NF-κBp65 could aggravate SAP-induced pancreatic injury (26).

NF-κBp65 inhibitors in in vitro and in vivo models of SAP have already been studied (27,28). In the present study, DXM was also demonstrated as an inhibitor of NF-κBp65 and that DXM can downregulate the expression of NF-κBp65 protein in the pancreatic tissue of SAP rats and in AR42J cells treated with CAE. Previous studies have demonstrated that HMGB1 acts through multiple mechanisms, including through the NF-κBp65 pathway (29). However, it is not certain whether there are binding sites for NF-κBp65 on the HMGB1 promoter.
Figure 1. Pathological changes in the pancreas. Pancreases from the sham, SAP and DXM groups were harvested 3, 6 or 12 h following operations and were stained. (A) Mass hemorrhages, edema and structural alterations with necrosis were observed in the pancreas by hematoxylin and eosin staining (magnification, x400). Scale bar represents 250 µm. (B) The apoptotic acinar cells in the pancreas were observed by TUNEL staining. (C) IOD analysis of TUNEL. Data are presented as the mean ± standard error of the mean (n=3). **P<0.01, ***P<0.001 vs. SAP group; ***P<0.001 vs. Sham group. SAP, severe acute pancreatitis; DXM, dexamethasone-treated; TUNEL, terminal deoxynucleotidyl transferase dUTP nick-end labeling; IOD, integrated optical density.
Therefore, it remains unclear whether HMGB1 expression is directly regulated by NF-κBp65.

HMGB1, a DNA-binding intranuclear protein, is a late activator of the inflammatory cascade when released into the extracellular space. HMGB1 is released from necrotic cells or secreted by activated monocytes or macrophages. HMGB1 induces pro-inflammatory cytokines in human monocytes via TLR4 and NF-κB activation (30). HMGB1 can mediate cell-to-cell signaling by binding to the receptor for advanced glycation end products and toll-like receptors (TLR), especially TLR-2 and TLR-4, to enhance the inflammatory response (29,31). As a late-phase mediator, HMGB1 was previously discovered to be upregulated in a number of acute and chronic inflammatory conditions, including sepsis, acute lung injury and rheumatoid arthritis (32-34). In contrast to other known pro-inflammatory cytokines, the delayed kinetics of HMGB1 provide a wide

Figure 2. Immunohistochemical analysis. To detect the effects of DXM on HMGB1 and NF-κB protein expression levels in pancreatic tissue from sodium taurocholate solution-treated rats, HMGB1 and NF-κBp65 proteins were detected in pancreatic tissue from animals sacrificed 3, 6, or 12 h following operation in the sham, SAP and DXM groups. (A) The expression of HMGB1. (B) The expression of NF-κBp65 (magnification, x200). Scale bar represents 500 µm. HMGB1, high-mobility group box 1; NF-κB, nuclear factor-κB; DXM, dexamethasone; SAP, severe acute pancreatitis.
5350

ZHOU et al: REGULATION OF SEVERE ACUTE PANCREATITIS BY DEXAMETHASONE

window of opportunity for therapeutic approaches (35). The present results also highlight the possible beneficial role of anti-inflammatory cytokines, including IL-10 and IL-13, in the pathogenesis of SAP. These cytokines may counteract the type-1 pro-inflammatory cytokines secreted in response to the activation of the TLR4-NF-κB pathway induced by HMGB1.
As a result, HMGB1 offers the hope of developing an anti-inflammatory therapy that is practical and effective. In a previous study serum HMGB1 levels, were demonstrated to be significantly elevated in patients with SAP and were correlated with disease severity (36). Therefore, it has been speculated that HMGB1 may be a target for anti-inflammatory treatment in SAP. In the present study, it was demonstrated that HMGB1 expression levels increased in the pancreas with SAP and decreased following treatment with DXM, suggesting that HMGB1 may be pivotal for the inflammatory response and organ injury observed in SAP and may be a therapeutic target for SAP. It was hypothesized that elevated HMGB1 levels may represent an aggravating factor in the SAP and the associated multiple organ damage. However, HMGB1 affects a number of angiogenesis-associated conditions, including cancer, proliferative diabetic retinopathy and wound healing through the p53 pathway, and it is a promising therapeutic target in a number of tumors, including epidermal tumors, prostate cancer and colon cancer (37-39). Therefore, the exact function of HMGB1 and its mechanism still need to be elucidated. Notably, while investigating the relevance of HMGB1-TLR4-NF-κB-induced pro-inflammatory cytokines in SAP, it was demonstrated that the antibiotic and immunomodulatory agent fusidic acid may prevent pancreatitis through the reduction of TNF-α and IL-8. This is of interest because sodium fusidate is clinically available and could be immediately considered for the treatment of SAP (40,41).

In conclusion, DXM can reduce the levels of NF-κBp65 and HMGB1 and mitigate the pathological alterations in the pancreas of rats with SAP and in AR42J cells treated with CAE. In in vivo and vitro experiments, DXM was identified to have a therapeutic effect on SAP. Therefore, NF-κBp65 and HMGB1 may serve auxiliary roles in the treatment of SAP. Overall, each of these inflammatory mediators has benefits and should be used appropriately in future clinical practice.

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Availability of data and materials

The datasets used during the present study are available from the corresponding author upon reasonable request.

Authors’ contributions

SZ, LM and KX conceived and designed the study. SZ, TL, JZ and JY performed the experiments. SZ and JY wrote the manuscript. KX reviewed and edited the manuscript. All authors read and approved the manuscript and agree to be accountable for all aspects of the research in ensuring that the accuracy or integrity of any part of the work were appropriately investigated and resolved.

Ethics approval and consent to participate

The present study was approved by the Ethics Committee of Central South University (Changsa, China).

Patient consent for publication

Not applicable.

Competing interests

The authors declare they have no competing interests.

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5352

ZHAO et al: REGULATION OF SEVERE ACUTE PANCREATITIS BY DEXAMETHASONE

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