Functional Heterogeneity of Reelin in the Oral Squamous Cell Carcinoma Microenvironment

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Background: Reelin, an extracellular glycoprotein, is expressed on neuronal cells and participates in neuronal migration during brain development. Recently, Reelin also has a vital role in carcinogenesis. However, its role in oral squamous cell carcinoma (OSCC) remains to be explored. The purpose of this study was to explore the roles of Reelin in OSCC.

Methods: The expression of Reelin in cancer-associated fibroblasts (ReelinCAF) and tumor cells (ReelinTC) was analyzed by the Gene Expression Omnibus (GEO) database. Immunohistochemistry (IHC) was used to detect the spatial pattern of Reelin in 75 OSCCs. The diagnostic and prognostic values of Reelin were evaluated and also verified by The Cancer Genome Atlas (TCGA) database. Primary CAFs from 13 OSCC patients were isolated to confirm Reelin expression. Thirty-nine OSCC peripheral blood samples were used to analyze the change of immunocytes based on Reelin levels by flow cytometry. The relationship between Reelin and tumor immune microenvironment in head and neck squamous cell carcinoma (HNSCC) tissues was determined by TISIDB and the Tumor Immune Estimation Resource (TIMER) database.

Results: In breast cancer, pancreatic cancer and rectal cancer, Reelin in CAFs was significantly upregulated compared with Reelin in TCs. The IHC results in OSCC also showed that Reelin levels were higher in CAFs. Upregulated ReelinTC was related to a decreased pN stage and distant metastasis. Strikingly, patients with enhanced ReelinCAF had a high risk of lymph node metastasis, poor worst pattern of invasion (WPOI), and distant metastasis, but showed comparable Ki-67 level in all OSCC patients, resulting in shorter overall survival (OS) and disease-specific survival (DSS). Unexpectedly, Reelin in tumor-infiltrating lymphocytes (ReelinTIL) was correlated with postoperative relapse. Patients with high ReelinTIL, but not ReelinTC and ReelinCAF, had poor cytotoxicity of CD8+ T cells and higher ratio of CD4/CD8 in peripheral blood. However, Reelin was positively associated with tissue-resident B cells and NK cells in the tumor microenvironment.
INTRODUCTION

Head and neck squamous cell carcinoma (HNSCC) was the sixth most common cancer worldwide in 2018. Oral squamous cell carcinoma (OSCC) is the most common malignant tumor of the head and neck with a high mortality rate (1–3). Therefore, the potential predictors of OSCC are awaited to identify for guiding diagnosis and treatment. Reelin is a secreted extracellular matrix glycoprotein that is formed with 3,461 amino acids. It is secreted by Cajal-Retzius cells in the marginal zone and encoded by the RELN gene, which is located on chromosome 7q22. The molecular weight of Reelin is about 400 kDa (4). To date, a majority of Reelin-related studies have focused on neurological diseases, such as schizophrenia, autism, and Alzheimer’s disease because Reelin helps modulate the processes of neuronal migration and positioning during brain development (7–10). Importantly, Reelin is expressed in many extraneuronal tissues, including plasma, blood cells, liver, and intestine. However, the roles of peripheral Reelin remain unknown, especially in oral tissues with a large number of nerves (4).

Notably, alterations in Reelin expression are observed in cancers of nonneuronal origin (11). Specifically, the expression of Reelin is reduced in hepatocellular carcinoma, gastric carcinoma, colorectal cancer, glioblastoma, breast cancer, and pancreatic cancers (12–17). In contrast, upregulation of Reelin is observed in retinoblastoma, esophageal carcinoma, high-grade prostate cancer, multiple myeloma, and non-Hodgkin lymphoma (18–22). The alterations in Reelin expression are related to the tumor metastasis and invasion (23, 24). For example, in a human esophageal carcinoma cell line, RELN was related to TGF-β, which was related to metastasis, and knockdown of RELN increased the rate of cell migration (25). However, the role of Reelin in OSCC is far from known.

Although the studies related to Reelin mainly focuses on brain development and tumor progression, some studies have proved that Reelin is also linked to immune function. For example, some studies pointed out that interleukin (IL)-1β mRNA was highly expressed in mice with RELN mutation by stimulating with lipopolysaccharide (26, 27). Decreased Reelin was associated with the alterations in serotonin transporter (SERT) clustering in blood lymphocytes, which may be essential in some cardiovascular or immune system alterations (28). Thus, correlation between Reelin and immune balance in OSCC remains elucidated.

Therefore, in the present study, we focused on the expression pattern of Reelin in OSCC, including tumor cells (ReelinTC), cancer-associated fibroblasts (ReelinCAF), and tumor-infiltrating lymphocytes (ReelinTI). Furthermore, we evaluated the correlation between Reelin and clinicopathological characteristics, its prognostic significance, and immune cell infiltrations.

Conclusion: Reelin has a versatile function in distinct cell types during the development of OSCC via governing tumor cell and stroma microenvironment.

Keywords: Reelin, oral squamous cell carcinoma, cancer-associated fibroblasts, prognosis, lymphocyte subsets

MATERIALS AND METHODS

Patients and Samples

All methods used for this study were approved by the Ethics Committee of Nanjing Stomatoly Hospital [2019NL-009(KS)]. The study was carried out in accordance with the Declaration of Helsinki. Written informed consent was obtained from all the patients. Seventy-five OSCC patients were recruited in this study. All the 75 cases included 14 cases of cheek cancer, 30 cases of tongue cancer, 16 cases of gingival cancer, and 15 cases of other types of oral cancer. The inclusion and exclusion criteria of patients were the same as those of our previous studies (29). Specifically, the patients must have primary OSCC without anti-cancer therapies including chemotherapy and/or radiotherapy before surgery, and no distant metastasis prior to surgery, and these patients underwent curative resection between 2013 and 2016 at Nanjing Stomatoly Hospital. Lactating individuals, pregnant women, and patients who were diagnosed with other malignant diseases or certain autoimmune diseases were excluded. These patients with primary tumor were diagnosed by hematoxylin and eosin staining by two experienced pathologists. All 75 patients were followed-up until April 2020. Paraaffin-embedded OSCC tissue slices were obtained from the pathology department and used for IHC study. Thirty-nine blood samples from OSCC patients were obtained for flow cytometry assay before any related treatments.

Immunohistochemistry and Quantification

The protocol of IHC of formalin-fixed paraffin-embedded sections and scoring details of IHC was performed as previously described (29, 30). Specifically, IHC was performed on 3-µm formalin-fixed paraffin-embedded sections using anti-Reelin (monoclonal, 1:100 dilution; MAB5366; Abcam) and anti-Ki-67 (monoclonal, 1:200 dilution; ab16667; Abcam) and anti-Ki-67 (monoclonal, 1:200 dilution; ab16667; Abcam). Slides were deparaffinized with xylene and rehydrated in an ethanol series. Antigen retrieval was performed with 10 mmol/L citric acid (pH 6.0) in a pressure cooker. Then, endogenous peroxidase activity was blocked with a 3% hydrogen peroxide solution. After washing in phosphate-buffered saline (PBS; pH 7.4) three times, slides were incubated with primary antibodies against Reelin (MAB5366; Abcam) at 4°C overnight. Polink-2 plus HRP Detection Kit was used as the secondary antibody at 37°C for 40 min. Finally, slides were developed in diaminobenzidine (DAB). The primary antibody was replaced by PBS as negative control. The IHC staining results of Reelin and Ki-67 were independently and double blindly evaluated by two senior pathologists who did not know the patients’ data, and the average values were calculated for further analysis. IHC staining was scored according to the percentage of positive cells and staining intensity. The percentage of stained cells was defined as 0 = 0–5%; 1 = 6%–25%; 2 = 26%–50%;
3 = 51%–75%; and 4 = 75–100%. Staining intensity was defined as follows: 0 = negative staining; 1 = weak staining; 2 = moderate staining; and 3 = strong staining (29, 30). The IHC score was calculated by multiplying the grade of the staining intensity by that of the staining percentage. High and low expressions of Reelin were defined by the median of IHC scores.

**Primary CAFs Cultivation**

CAFs used in this study were isolated from fresh 13 OSCC tumor tissues immediately after surgical resection. Details of this protocol were the same as previously described (29).

**Immunofluorescence**

Immunofluorescence performed on primary CAFs was carried out as previously described (29): primary antibodies, α-SMA (1:200, Abcam, England), FSP-1 (1:200, Abcam, England), and PDGFR-β (1:200, Abcam, England); secondary antibodies, Alexa Fluor 647-conjugated (1:100, Abcam, England), Alexa Fluor 555-conjugated (1:200, Abcam, England), and Alexa Fluor 488-conjugated (1:400, Abcam, England).

**mRNA Isolation and Quantitative PCR**

Thirteen cultured primary CAFs were classified as WPOI = 1–3 or WPOI = 4–5. The details of extracting total mRNA and conducting quantitative PCR (q-PCR) were performed as previously described (29). Comparative 2−ΔΔCt or 2−ΔCT method was used to quantify the relative mRNA expression as indicated.

**Flow Cytometry Assay**

For the cell subtypes of peripheral blood mononuclear cell (PBMC) analysis, details of flow cytometry were the same as previously described (31).

**Analysis of GEO, TCGA, TISIDB, and TIMER Databases**

The differential expression of Reelin between CAFs and TCs was performed by Gene Expression Omnibus (GEO, https://www.ncbi.nlm.nih.gov/geo/) database analysis in specific tumors. The Cancer Genome Atlas (TCGA, https://cancergenome.nih.gov/abouttcga/overview) database was used to further validate

**FIGURE 1**

Reelin was upregulated in CAFs compared with TCs derived from OSCC and other several cancers. Enhanced level of RELNCAF compared with RELNTC in breast cancer (A), pancreatic cancer (B), and rectal carcinoma (C) using the GEO database. Representative immunohistochemistry staining of Reelin in OSCC, according to TCs, FLCs and TILs (D). The IHC score of ReelinTC and ReelinFLC in OSCC (E). **** denoted that differences were considered statistically significant with p < 0.0001.
survival implication of Reelin. Tumor Immune Estimation Resource (TIMER, https://cistrome.shinyapps.io/timer/) was used to learn the association between Reelin mRNA expression and the expression of particular immune infiltrating cell subset markers according to the TIMER’s online instructions. An integrated repository portal for tumor–immune system interactions (TISIDB, http://cis.hku.hk/TISIDB/index.php) was utilized to examine Reelin mRNA and immune system interactions in 28 types of TILs across human cancers.

Statistical Analysis

SPSS 25.0 software and GraphPad Prism 8.0 were used for statistical analyses and figure process. The results of the experiments were presented as mean ± SEM. Chi-square test, Pearson’s chi-square test, or Fisher’s exact test was used to compare clinicopathological parameters of the patients. Kolmogorov-Smirnov tests were used to learn the relationship between Reelin and Ki-67. The Mann–Whitney U test was used to compare the two groups (e.g., Reelin in CAFs with relapse, with relapse versus without relapse). Survival analysis including overall survival (OS), disease-specific survival (DSS), and progression-free survival (PFS) were evaluated by Kaplan–Meier survival test. The Cox regression model was used to evaluate the correlations between different prognostic factors in a multivariate analysis. The immune infiltration level was compared with the t test. All the analyses were two-sided test and differences were considered statistically significant with p < 0.05.

RESULTS

The Reelin Levels Were Higher in CAFs Than That in TCs

Based on the GEO database, we found that Reelin was significantly upregulated in CAFs compared with TCs (p = 0.0063, p = 0.0021, and p = 0.0033; Figures 1A–C). Thus, to validate Reelin expression in OSCC, IHC staining was performed to assess the expression of Reelin in different cell types including TCs, FLCs, and TILs. Consistently with the database, the IHC score of ReelinFLC was significantly higher than that of ReelinTC in OSCC (p < 0.0001 Figure 1E). In addition, representative IHC staining is shown in Figure 1D.

ReelinCAF Positively Correlated With pN stage and WPOI, but ReelinTC Negatively Correlated With pN Stage

After confirming the contrary expression pattern of ReelinTC and ReelinFLC in OSCC, we further evaluated the correlation between Reelin and various clinicopathological variables according to different cell types in Table 1. The results demonstrated that patients with low-expressed ReelinTC (p = 0.041, Figure 2A) but high-expressed ReelinFLC (p = 0.007, Figure 2B) had a high risk of lymph node metastasis. Also, more ReelinFLC was closely linked to poor WPOI (p = 0.034, Figure 2C). In addition, we continued to study the role of Reelin in primary CAFs from OSCC patients. CAFs were successfully isolated and cultured
from fresh OSCC tissues with reliable phenotypic identification using immunofluorescence (Figure 2D). We divided CAFs into WPOI (1–3) and WPOI (4–5) groups. Consistently with the IHC results, high-expressed Reelin in primary CAFs had a poor WPOI using qPCR analysis (p = 0.0016, Figure 2E). However, there were no differences between Reelin and sex, age, smoking habit, T stage, or differentiation (all p > 0.05). Ki-67 IHC staining also showed that there was no difference between low Reelin levels and high Reelin levels in TCs (p = 0.1232, Figure 3A), CAFs (p = 0.6257 Figure 3B), and TILs (p = 0.4325, Figure 3C).

Upregulated ReelinCAF, but Downregulated ReelinTC Correlated With High Risk of Postoperative Relapse, Metastasis, and Poor Clinical Outcome

Considering that decreased ReelinTC, but increased ReelinCAF was correlated with poor clinicopathological characteristics, we further analyzed the association between Reelin and postoperative relapse as well as distant metastasis according to distinct cell types. The results indicated that patients with postoperative relapse had a low expression of ReelinTC (p = 0.0465, Figure 3D), but a high expression of ReelinCAF (p = 0.0015, Figure 3E). Moreover, patients with low-expressed ReelinTC (p = 0.0221, Figure 3G) but high-expressed ReelinCAF (p = 0.0165 Figure 3H) were prone to postoperative metastasis. Strikingly, patients with high ReelinTIL also had a high risk of postoperative relapse (p = 0.0007, Figure 3F). Reelin in TILs didn’t have relationship with metastasis (P = 0.0565, Figure 3F).

Considering that the role of Reelin in TCs was contrary to that in CAFs in cancer metastasis and relapse, we further evaluated its prognostic value in OS and DSS. Kaplan–Meier curve analysis showed that patients with lower-expressed ReelinTC but higher-expressed ReelinCAF had a shorter OS (p = 0.0391, Figure 4A; p < 0.0001, Figure 4B) and DSS (p = 0.0005, Figure 4D; p = 0.0005, Figure 4E). Reelin in TILs wasn’t linked with OS and DSS (P = 0.7794, Figure 4C; P = 0.5798, Figure 4F). Consistent with our results, the TCGA database (TCGA.HNSC.sampleMap/HiSeqV2) also showed that patients with high-expressed ReelinTC had a longer DSS (p = 0.0128, Figure 4G) and PFS (p = 0.0173, Figure 4H).

ReelinCAF Was an Independent Prognostic Factor of OS and DSS

Univariate and multivariate analyses suggested that ReelinCAF was a valuable prognostic factor among the clinicopathologic variables examined, including gender, age, smoking, T stage, N stage, differentiation, and WPOI. Note that patients with high-expressed ReelinCAF had a worse OS (p = 0.004, Table 2) and.
DSS ($p = 0.009$, Table 2). These data clearly showed that Reelin$^{CAF}$ level may further refine the prognostic criteria so as to find OSCC patients who may need extra therapy.

**Reelin$^{TIL}$ Disrupted the Balance of CD4$^+$/CD8$^+$ T Cell in Peripheral Blood**

Several previous studies have pointed out that Reelin also played a vital role in immune function. Our data showed that patients with high-expressed Reelin$^{TIL}$ had a high risk of postoperative relapse. We further analyzed the correlation between Reelin and lymphocyte subsets from PBMCs of OSCC using flow cytometry. Lymphocytes were gated as shown in Figure 5A. The results demonstrated that patients with high-expressed Reelin$^{TIL}$, not Reelin$^{TC}$ or Reelin$^{CAF}$, had a low ratio of CD3$^+$CD8$^+$ T cells ($p = 0.0085$, Figure 5B) and a high ratio of CD3$^+$CD4$^+$ T cells ($p = 0.0118$, Figure 5B). A study reported that elevated CD4/CD8 ratio was related to poor survival in head and neck squamous cancer (32). Therefore, we evaluated the CD4/CD8 ratio between

**FIGURE 3** | Correlation between Ki-67 in TCs and Reelin in TCs, CAFs, and TILs (A–C). Correlation between Reelin and postoperative recurrence as well as distant metastasis, according to TCs (D, G), CAFs (E, H), and TILs (F, I). *, ** and *** denote that differences were considered statistically significant with $p < 0.05$, $p < 0.01$ and $p < 0.001$ respectively. ns represented no statistical differences.
low and high expression of ReelinTIL. The result showed that more ReelinTIL had a higher ratio of CD4/CD8 (p = 0.0018, Figure 5C). Besides, our data showed no relationship between Reelin and B cells as well as NK cells from peripheral blood.

**ReelinTIL Promoted Multitype Immune Cells Infiltration in the Tissue Microenvironment**

We furthermore evaluated the relationship between RELN and tumor immune microenvironment. We found correlations of RELN with 28 types of TILs across human heterogeneous cancers according to the TISIDB database (Figure 5D). In HNSCC, RELN positively correlated with abundance of central memory CD8 T cells (Tcm_CD8 T cells; rho = 0.288, p < 0.001, Figure 5E), central memory CD4 T cells (Tcm_CD4 T cells; rho = 0.236, p < 0.001, Figure 5E), natural killer cells (NK cells; rho = 0.357, p < 0.001, Figure 5E), and activated B cells (B cells; rho = 0.313, p < 0.001, Figure 5E). Meanwhile, the Spearman’s correlation test was further used to examine the correlation among RELN and immune cell infiltration of HNSCC according to the TIMER database (Table 3). Elevated RELN was significantly associated with T-cell, B-cell, monocyte, macrophage, and dendritic cell infiltration (all p < 0.05), leading to a multitype immune cell infiltration in the tissue microenvironment.

**DISCUSSION**

Reelin has a great impact on carcinogenesis and tumor progression. However, the expression of Reelin in different cancers was different. For example, Reelin is decreased in hepatocellular carcinoma, but increased in esophageal carcinoma (13, 22). Based on the GEO database, we found that Reelin was significantly upregulated in CAFs compared with TCs in breast cancer, pancreatic cancer, and rectal carcinoma. However, the expression of Reelin in OSCC had not yet been investigated. In this study, we examined the expression pattern of Reelin in OSCC for the first time. The result also demonstrated that Reelin levels were higher in CAFs compared with TCs in OSCC. Besides, TIL-derived Reelin was expressed distinctively according to different OSCC patients. Therefore, we concluded that Reelin might have a versatile function in different cell types during the development of OSCC via governing tumor cell and stroma microenvironment.

According to the significant role of Reelin in tumor progression, some studies showed that upregulated Reelin was found to be correlated with proliferation of cancer cells in multiple myeloma (33). However, stimulating Reelin signaling showed that the proliferation of tumor cells in glioblastoma decreased significantly (15). Moreover, in breast cancer, a previous study showed that enhanced Reelin in MDA-MB231 cells suppressed invasiveness of cancer cells in vitro (17).
In OSCC, to investigate the association between Reelin and tumor proliferation, we analyzed Ki-67 among high- and low-expressed groups of Reelin. The result showed that there was no statistical relationship between low and high Reelin levels. However, patients with high-expressed ReelinCAF were predicted to have a poor WPOI. Notably, low-expressed ReelinTC, but high-expressed ReelinCAF was related to the lymph node metastasis, which indicated that Reelin has an opposite function in CAFs and TCs during the development of OSCC.

Besides being related to clinicopathological parameters, some previous studies also determined that Reelin is also related to the prognosis of patients. For example, in hepatocellular carcinoma (HCC), relapse was more likely to happen when the expression of Reelin in tumor cells was high (13). In addition, comparing metastatic colorectal diseases with primary tumors, the expression of Reelin increased significantly in metastatic tumors (23). These data indicated that Reelin might play a vital role in local relapse and distant metastasis. In our survival analysis, the result showed that patients with higher-expressed ReelinCAF had a greater risk of relapse and distant metastasis. Instead, patients with higher-expressed ReelinTC had a lower risk of relapse and distant metastasis. Strikingly, patients with high-expressed ReelinTIL also had a high risk of relapse. This finding further suggested that ReelinCAF and ReelinTC play a diverse role in the prognosis of OSCC patients. Also, ReelinTIL might be a potential indicator with an increased risk of OSCC relapse.

### TABLE 2 | Cox-regression analyses of overall survival (OS) and disease-specific survival (DSS) in OSCC patients.

| Variables | Univariate analysis | Multivariate analysis |
|-----------|---------------------|-----------------------|
|           | Hazard ratio | 95% CI | p | Hazard ratio | 95% CI | p |
| **OS**    |            |       |       |            |       |       |
| Gender    | Male vs. Female | 1.46 | 0.625–3.413 | 0.382 | 7.618 | 2.242–25.887 | 0.001* |
| Age       | <60 vs. ≥60 | 0.992 | 0.446–2.0210 | 0.985 | 1.374 | 0.513–3.684 | 0.527 |
| Smoking   | No vs. Yes | 1.073 | 0.408–2.0820 | 0.886 | 9.828 | 2.919–33.088 | <0.001* |
| T Stage   | 1–2 vs. 3–4 | 0.374 | 0.153–0.912 | 0.027 | 2.164 | 0.645–7.260 | 0.211 |
| N Stage   | No vs. Yes | 2.589 | 0.966–6.941 | 0.059 | 1.014 | 0.351–3.053 | 0.972 |
| Differentiation | Well vs. Moderate to poor | 2.954 | 1.102–7.921 | 0.031* | 2.767 | 1.025–7.471 | 0.045* |
| WPOI      | 1–3 vs. 4–5 | 0.148 | 0.051–0.435 | 0.001* | 0.202 | 0.068–0.599 | 0.004* |
| ReelinTC  | Low vs. High | 2.589 | 0.966–6.941 | 0.059 | 0.922 | 0.413–2.061 | 0.844 |
| ReelinCAF | Low vs. High | 1.374 | 0.513–3.684 | 0.527 | 9.828 | 2.919–33.088 | <0.001* |
| ReelinTIL | Low vs. High | 8.504 | 2.492–29.016 | 0.001* | 4.776 | 1.345–16.962 | 0.016* |
| **DSS**   |            |       |       |            |       |       |
| Gender    | Male vs. Female | 1.461 | 0.590–3.622 | 0.413 | 2.164 | 0.645–7.260 | 0.211 |
| Age       | <60 vs. ≥60 | 0.992 | 0.383–2.125 | 0.814 | 1.852 | 0.545–6.293 | 0.323 |
| Smoking   | No vs. Yes | 1.024 | 0.364–2.878 | 0.964 | 2.499 | 0.914–6.831 | 0.074 |
| T Stage   | 1–2 vs. 3–4 | 1.235 | 0.415–3.673 | 0.705 | 6.485 | 1.509–27.866 | 0.012* |
| N Stage   | No vs. Yes | 0.173 | 0.058–0.516 | 0.002* | 0.811 | 0.341–1.928 | 0.636 |
| Differentiation | Well vs. Moderate to poor | 0.922 | 0.413–2.061 | 0.844 | 8.504 | 2.492–29.016 | 0.001* |
| WPOI      | 1–3 vs. 4–5 | 0.922 | 0.413–2.061 | 0.844 | 1.014 | 0.351–3.053 | 0.972 |
| ReelinTC  | Low vs. High | 6.485 | 1.509–27.866 | 0.012* | 4.122 | 0.941–18.053 | 0.06 |
| ReelinCAF | Low vs. High | 0.173 | 0.058–0.516 | 0.002* | 0.226 | 0.074–0.691 | 0.009* |
| ReelinTIL | Low vs. High | 0.811 | 0.341–1.928 | 0.636 | 2.954 | 1.102–7.921 | 0.031* |

CI, confidence interval; ReelinCAF, Reelin in cancer-associated fibroblasts; ReelinTC, Reelin in tumor cells; ReelinTIL, Reelin in tumor-infiltrating lymphocytes; WPOI, worst pattern of invasion. *Denotes that differences were considered statistically significant with p < 0.05.
FIGURE 5 | Flow cytometry analysis of lymphocytes, gated based on characteristic light-scatter properties, and single lymphocytes were gated based on forward scatter height vs. forward scatter area (FSC-A). The numbers in the quadrants or over line indicate the percentage of cells (A). Correlation of Reelin expression with immune infiltration level in OSCC before surgery, according to TCs, CAFs, and TILs (B). Correlation of CD4/CD8 ratio with Reelin in TILs (C). Relations between expression of RELN and 28 types of TILs across human heterogeneous cancers (D). RELN significantly correlated with CD8⁺ T cells, CD4⁺ T cells, NK cells, and B cells in HNSCC (E). * and ** denote that differences were considered statistically significant with \( p < 0.05 \) and \( p < 0.01 \), respectively. ns represented no statistical differences.
### TABLE 3 | Correlation analysis between RELN and immune cell infiltrations in HNSCC samples using TIMER.

| Description          | Gene markers | RELN               | None | p  | RELN               | Purity | p  |
|----------------------|--------------|--------------------|------|----|--------------------|--------|----|
|                      |              | Cor                |      |    | Cor                |        |    |
|                      |              |                    |      |    | None               |        |    |
| CD8+ T cell          | CD8A         | 0.185              | ***  |    | 0.134              | **     |    |
|                      | CD8B         | 0.162              | ***  |    | 0.114              | *      |    |
| T cell (general)     | CD3D         | 0.182              | **** |    | 0.129              | *      |    |
|                      | CD3E         | 0.264              | **** |    | 0.217              | ****   |    |
|                      | CD2          | 0.258              | **** |    | 0.211              | ****   |    |
| B cell               | CD19         | 0.23               |      |    | 0.186              | ****   |    |
|                      | CD79A        | 0.236              |      |    | 0.196              | ****   |    |
| Monocyte             | CD86         | 0.345              |      |    | 0.307              | ****   |    |
|                      | CSF1R        | 0.397              |      |    | 0.358              | ****   |    |
| TAM                  | CCL2         | 0.413              | **** |    | 0.383              | ****   |    |
|                      | CD68         | 0.279              | **** |    | 0.245              | ****   |    |
|                      | IL10         | 0.424              | **** |    | 0.395              | ****   |    |
| M1 macrophage        | NOS2         | 0.166              |      |    | 0.185              | ****   |    |
|                      | IRF5         | 0.028              | 0.525|    | 0.025              | 0.578  |    |
| M2 macrophage        | CD163        | 0.348              |      |    | 0.308              | ****   |    |
|                      | VSG4         | 0.261              |      |    | 0.219              | ****   |    |
|                      | MS4A4A       | 0.3                |      |    | 0.252              | ****   |    |
| Neutrophils          | CEACAM9      | −0.017             | 0.703|    | −0.026             | 0.564  |    |
|                      | ITGAM        | 0.27               |      |    | 0.24               | ****   |    |
|                      | CCR7         | 0.383              | **** |    | 0.342              | ****   |    |
| Natural killer cell  | KIR2DL1      | 0.067              | 0.127|    | 0.061              | 0.18   |    |
|                      | KIR2DL2      | 0.055              | 0.209|    | 0.038              | 0.406  |    |
|                      | KIR2DL4      | 0.058              | 0.186|    | 0.044              | 0.334  |    |
|                      | KIR3DL1      | 0.164              |      |    | 0.144              | **     |    |
|                      | KIR3DL2      | 0.167              |      |    | 0.137              | **     |    |
|                      | KIR3DL3      | −0.027             | 0.544|    | −0.04              | 0.373  |    |
|                      | KIR2DS4      | 0.035              | 0.423|    | 0.007              | 0.878  |    |
| Dendritic cell       | HLA-DPB1     | 0.28               |      |    | 0.232              | ****   |    |
|                      | HLA-DQB1     | 0.217              |      |    | 0.182              | ****   |    |
|                      | HLA-DRA      | 0.285              |      |    | 0.236              | ****   |    |
|                      | HLA-DPA1     | 0.299              |      |    | 0.252              | ****   |    |
|                      | CD1C         | 0.382              |      |    | 0.35               | ****   |    |
|                      | NRP1         | 0.458              |      |    | 0.429              | ****   |    |
|                      | ITGAX        | 0.272              |      |    | 0.232              | ****   |    |
| Th1                  | TBX21        | 0.191              |      |    | 0.15               | ***    |    |
|                      | STAT4        | 0.287              |      |    | 0.25               | ****   |    |
|                      | STAT1        | 0.208              |      |    | 0.176              | ****   |    |
|                      | IFNG         | 0.034              | 0.436|    | −0.01              | 0.822  |    |
|                      | TNF          | 0.2                |      |    | 0.193              | ****   |    |
| Th2                  | GATA3        | 0.307              |      |    | 0.273              | ****   |    |
|                      | STAT6        | 0.249              |      |    | 0.27               | ****   |    |
|                      | STAT5A       | 0.258              |      |    | 0.24               | ****   |    |
|                      | IL13         | 0.204              |      |    | 0.186              | ****   |    |
| Tfh                  | BCL6         | 0.097              | 0.0271|    | 0.126              | **     |    |
| Th17                 | STAT3        | 0.312              |      |    | 0.317              | ****   |    |
|                      | IL17A        | 0.195              |      |    | 0.162              | **     |    |
| Treg                 | FOXP3        | 0.391              |      |    | 0.356              | ****   |    |
|                      | CCR8         | 0.49               |      |    | 0.468              | ****   |    |
|                      | STAT5B       | 0.395              |      |    | 0.387              | ****   |    |
|                      | TGFB1        | 0.251              |      |    | 0.246              | ****   |    |
| T cell exhaustion    | PDCD1        | 0.165              |      |    | 0.114              | *      |    |
|                      | CTLA4        | 0.247              |      |    | 0.203              | ****   |    |
|                      | LAG3         | 0.107              |      |    | 0.068              | 0.133  |    |
|                      | HAVCR2       | 0.279              |      |    | 0.238              | ****   |    |
|                      | GZMB         | 0.087              |      |    | 0.045              | 0.315  |    |

**TIMER**, Tumor IMmune Estimation Resource; TAM, tumor-associated macrophage; Th, T helper cell; Tfh, Follicular helper T cell; Treg, regulatory T cell; Cor, R value of Spearman’s correlation; None, correlation without adjustment; Purity, correlation adjusted by purity.

*, **, *** and **** denote that differences were considered statistically significant with $p < 0.05$, $p < 0.01$, $p < 0.001$, and $p < 0.0001$, respectively.
Moreover, as a previous study showed, high-expressed Reelin was negatively associated with PFS and OS in multiple myeloma (34). However, low-expressed Reelin significantly correlated with poor clinical outcome in breast cancer (17). Also, in glioblastoma, the expression of Reelin was positively related to survival (15). Just as we predicted, the result showed that higher-expressed ReelinCAF, but lower-expressed ReelinTC is correlated with shorter OS and DSS in this study. Meanwhile, ReelinCAF was an independent prognostic factor of OS and DSS for OSCC patients.

Furthermore, Reelin-related studies had focused on not only the tumor progression but also the immune function. For example, alterations in serotonin transporter (SERT) clustering in blood lymphocytes associated with a decrease in Reelin expression may be operative in some cardiovascular or immune system alterations (28). Our data showed that patients with high-expressed ReelinTIL had a high risk of postoperative relapse. We identified the relationship between the expression of Reelin and lymphocyte subsets from PBMCs of OSCC. The result showed that patients with high ReelinTIL, but not ReelinTC and ReelinCAF had poor cytotoxicity of CD8+ T cells and higher ratio of CD4/CD8 in peripheral blood, which might result in relapse of OSCC patients. However, the database showed that Reelin was positively associated with tissue-resident B cells and NK cells in the tumor microenvironment, suggesting that Reelin regulates immune response depending on the tissue or blood microenvironment.

In conclusion, we identified that increased ReelinCAF had a higher risk for metastasis and relapse. Therefore, ReelinCAF can be used as a potential biomarker in diagnosis and treatment of OSCC patients. Moreover, Reelin could regulate the balance of CD4+ /CD8+ T cells in peripheral blood, which indicated that Reelin might affect the prognosis of OSCC patients. However, it is positively correlated with tissue-resident B cells and NK cells, suggesting that Reelin regulates immune response depending on the tissue or blood microenvironment. Altogether, the study mainly explored the prognostic value of Reelin and the role in immune imbalance in OSCC. However, the molecular function and regulation pathway of Reelin in tumorigenesis of OSCC remained unexplored. In addition, the molecular mechanism of Reelin on immunomodulatory effects was still quite unclear. Future research is needed to unravel the molecular function and regulation pathway of Reelin in tumorigenesis of OSCC. Moreover, the molecular mechanism of Reelin on immunomodulatory effects need to be further explored.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/supplementary material. Further inquiries can be directed to the corresponding authors.

ETHICS STATEMENT

All the patients have signed the written informed consent before the study. And the present study was approved by the Research Ethics Committee of Nanjing Stomatological Hospital.

AUTHOR CONTRIBUTIONS

All authors contributed to the article and approved the submitted version.

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