Background: The survival rate of children and adolescents with acute lymphoblastic leukemia (ALL) has progressively improved. However, ALL survivors often have adverse effects after treatment, such as an increased risk of obesity. Obesity has been associated with reduced survival.

Objective: We investigated the relationship between obesity, adipocytokine levels, and ALL short-term outcomes.

Methods: Weight and height were measured, and body mass index (BMI) was calculated at patient diagnosis and discharge. Leptin and Adiponectin levels and Minimal Residual Disease (MRD) were measured before therapy, at days 19 of remission-induction therapy, and at the end of remission-induction therapy (days 46). The relationship between BMI, adipocytokine levels, and MRD was then determined.

Results: Compared to the normal BMI group, children with an abnormal increase in BMI had an increase in MRD at day 19 and 46 ($P = 0.04$ and $P = 0.008$), and showed a positive correlation ($P = 0.014$). In addition, we found a positive correlation between weight, hip circumference at diagnosis and at day 19, and MRD at day 46. Both BMI and fat concentric distribution affected the outcome of ALL children. A higher BMI was also associated with a significant increase in Leptin levels at diagnosis. Leptin resistance should be considered in ALL children with high BMI.

Conclusion: BMI affects the outcome of ALL patients. Early interventions such as regular weight, height monitoring, and dietary assessments should be preferably initiated during remission-induction chemotherapy.

Keywords: childhood acute lymphoblastic leukemia, remission-induction chemotherapy, obesity, body mass index, minimal residual disease (MRD), adipocytokine
INTRODUCTION

The survival rate of children and adolescents with acute lymphoblastic leukemia (ALL) has improved to more than 90% in developed countries as a result of improved risk-directed treatment and supportive care (1–4). However, ALL survivors often show adverse effects after their treatment, such as an increased risk of obesity and short stature (5). In children with acute myeloid leukemia, obesity at diagnosis has been associated with worse outcomes (6, 7). These include reduced survival and increased drug toxicity (8). In the long-term, obesity could result in substantial physical and psychosocial morbidity, such as infections, hypertension, and hyperglycemia during therapy, as well as metabolic syndrome (MS) later in life (9). Hence, MS criteria, such as BP, weight, and serum triglyceride levels, should be measured at each patient visit (10).

Obesity/overweight, one of the major complications after ALL treatment, affects treatment efficacy and survival through multiple mechanisms (11). Leptin is a protein synthesized in the fatty tissues and is effective for the control of obesity. Monitoring leptin levels in acute lymphoblastic leukemia (ALL) survivors could be helpful in controlling obesity (12). Biomarkers such as homeostasis model of assessment for insulin resistance index (HOMA-IR), leptin, and leptin/adiponectin have been associated with components of MS and have been associated with a greater risk of MS among ALL children (13).

This study investigated the adverse effects of obesity in ALL children in two centers who were treated using the St. Jude total XV protocol (14). This was done during the remission-induction phase, which is the most important phase during the chemotherapy protocol. Body mass index (BMI), waist circumference, and hip circumference at diagnosis and at the end of remission-induction were measured. Leptin, Adiponectin levels, and Minimal Residual Disease (MRD) (15) were measured at day 0, 19, and 46 of remission-induction chemotherapy using marrow examinations.

MATERIALS AND METHODS

Study Design and Patients

This study was a longitudinal observational study. Between March 2017 and March 2018, ALL patients who received treatment at the Department of Hematology and Oncology of Guangzhou Women and Children's Medical Center and Department of Pediatrics at Nanfang Hospital, Southern Medical University were enrolled to this study. Patient inclusion criteria were: (1) Between 1 and 10 years of age; (2) newly diagnosed with ALL (16, 17) and received remission-induction chemotherapy based on the St. Jude total XV protocol (14) (Table 1); (3) clear consciousness and the ability to communicate, or were accompanied by their parent(s) or guardian; and (4) had stable disease. Patient exclusion criteria were: (1) extreme malnutrition or critical illness; (2) congenital metabolic or genetic disorders, such as urea cycle disorder or Prader-Willi syndrome; (3) psychiatric disorders or organic encephalopathy; (4) did not receive a normal diet or oral nutrition or required fasting and parenteral nutrition due to the disease condition.

This study was approved by the Ethics Committee of Guangzhou Women and Children's Medical Center. All parents signed informed consent before study initiation.

During the remission-induction chemotherapy, we compared the height, weight, waist, and hip circumference at discharge vs. at diagnosis. In addition, we measured Leptin and Adiponectin levels on days 0, 19, and 46 together with bone marrow examinations. A paired t-test was performed to evaluate data between days 0, 19, and 46.

Risk Classification

Risk classification was based on the CCCG-ALL-2015 protocol which was modified from the St. Jude total XV protocol (14) after considerations specific to the Chinese population (18). Risk classifications were based on the following characteristics (age, white blood cell count, immunophenotype, and cytogenetics) and treatment response as measured by MRD levels during remission induction (on day 19) and at the end of remission induction therapy (on or around day 46).

Remission-Induction Chemotherapy

Remission-induction chemotherapy was the most important and initial portion of the entire ALL chemotherapy regiment. Intravenous injection or oral administration of dexamethasone (Dex) (3 mg/m², twice per day) was on days 1–4. Afterward, prednisone (15 mg/m², three times per day) was orally administered on days 5–28. The prednisone dose was gradually reduced by half on days 29–35 until withdrawal. This was the initial remission-induction chemotherapy and was termed VDLP therapy. The second remission-induction chemotherapy was CAT therapy and consisted of cyclophosphamide (CTX), cytarabine (Ara-C), and 6-mercaptopurine (6-MP) (based on the St. Jude total XV protocol) (14) (Table 1). Children were discharged after 35 days of remission-induction chemotherapy, and then re-hospitalized for subsequent chemotherapy after 2 weeks.

Data Collection

A medical record abstraction was performed for each patient. Height and weight were measured before chemotherapy and after 46 ± 2 days. Raw BMI was then calculated. The waist and hip circumferences were measured for each course of remission-induction chemotherapy. The waist-to-hip ratio (WHR) was then calculated. In addition, Leptin and Adiponectin levels in the bone marrow were measured at days 1–4, 19 ± 2, and 46 ± 2 together with MRD using bone marrow examinations. The leptin-to-adiponectin ratio (LAR) was subsequently calculated. MRD was the main prognostic variable in this study.
### TABLE 1 | Remission-induction chemotherapy.

| Group          | Dose       | Route | Frequency | Time (days) | Note                                                                 |
|----------------|------------|-------|-----------|-------------|----------------------------------------------------------------------|
| Window         | Dex        | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Dex        | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
|                | Window     | L/I/H | 6 mg/m²·d | IV/PO bid   | 1–4 If WBC ≥ 50 × 10⁹/L, single dose (3 mg/m²) Dex was added on d0 and immature cells in peripheral blood were measured on d5. |
For children between 1 and 3 years of age, Seca416 and Seca376 were used to measure the length and weight. For children > 3 years of age, Seca704s was used to measure height and weight. Waist and hip circumferences were measured using Seca201. One decimal was used for the results of height, waist circumference, and hip circumference, while two decimals were used for the results of weight. The measuring tools were regularly verified to ensure that they were accurate and easy to use and calibrated before use. All measurements were performed by two trained nurses according to the Standard Methods for Somatometry issued by the World Health Organization (WHO) in 20081.

The RayBio® Human Leptin enzyme-linked immunosorbent assay (ELISA) kit (RayBiotech, Inc., Georgia, America) and the RayBio® Human Acrp30 ELISA kit (RayBiotech, Inc., Georgia, America) were used to measure the levels of leptin and adiponectin and were performed according to the manufacturer's instructions. Patient samples were diluted 30,000 times before measuring Adiponectin levels.

**Standardization and Definitions of BMI**

Raw BMI values were converted to age and gender-adjusted z-scores using the Centers for Disease Control and Prevention's Year 2000 growth charts for patients 2–10 years (Epi Info 3.5)2. Length for weight z-scores was substituted for patients who had measured values prior to turning 24 months of age. Patients aged 2–10 years were defined as underweight, normal weight, overweight or obese according to whether their BMI was below the 5th percentile, in or above the 5th percentile but below the 85th percentile, in or above the 85th percentile but below the 95th percentile or in or above the 95th percentile, respectively (19). Percentage of measured/median was defined as a proportion of measured value in the median of the WHO reference.

**Primary and Secondary Outcomes Variables**

The primary outcome was MRD and the change in BMI. Secondary outcomes included the changes in waist circumference, WHR, Leptin, and Adiponectin levels. MRD was determined at diagnosis, days 19 ± 2 and at days 46 ± 2 of remission-induction chemotherapy using a multiparameter 6-color flow cytometry performed on bone marrow samples with a sensitivity of 0.01% (20).

For bone marrow examinations, routine time points for bone marrow aspiration in clinical practices (days 0, 19, and 46) were selected. Patients were placed in the lateral position, and then bone marrow aspiration was performed from the posterior superior iliac spine. After obtaining 0.5–1 ml of bone marrow aspirate, samples were centrifuged and the supernatants were stored at −80°C until required (13). All bone marrow aspirations were performed by experienced physicians.

**Statistical Analysis**

All data were analyzed using SPSS 25.0 Win64 software (IBM, NY, USA). Continuous data were examined for normal distribution using the Kolmogorov–Smirnov test. Continuous variables with normal distribution were presented as means ± standard deviation (SD) and were analyzed using an independent-sample t-test (intergroup comparisons) or paired t-test (pre and post-therapy) or using repeated measure ANOVA and SNK post-hoc test (intergroup comparisons in time). Frequencies (percentages) were used to present categorical variables, and the chi-square test was used for intergroup comparisons. P < 0.05 was considered statistically significant, i.e., BMI at diagnosis was analyzed in four (underweight, normal, overweight and obese) subgroups. BMI percentile changes from diagnosis to the end of induction were analyzed as a continuous variable. MRD on day 19 and at the end of induction were compared across the groups by the chi-square tests. Complete remission was defined as having <5% leukemic blasts with restoration of normal hematopoiesis.

**RESULTS**

**Patient Recruitment and Baseline Characteristics**

In this study, 103 children with B-cell acute lymphoblastic leukemia (B-ALL) were enrolled. Six children were excluded due to severe disease needing intensive care, and three due to history of pancreatitis. Another 12 children were withdrawn during the study, which included five who were administered parenteral nutrition after refusing oral diet or enteral nutrition due to severe oral ulcers and seven who had variable blood glucose levels and received a diabetic diet. Finally, 82 children were included in the study. Baseline characteristics including age, gender, admission age, and risk classification by BMI category are presented in Table 2, while discharge status is presented in Table 3.

Minimal residual disease by BMI category is shown in Table 4. Among the four subgroups, abnormal BMI was associated with higher percentage of an MRD level of 1% or higher on day 19 (P = 0.04) and 0.01% or more on day 46 (P = 0.008). Percentage of an MRD level of 1% or higher on day 19 and 0.01% or more on day 46 was significantly higher in the higher BMI subgroups (overweight and obese) than the normal weight subgroup (7.7% vs. 0 and 30.8 vs. 6.9%, respectively).

There was no difference in Leptin and Adiponectin levels between BMI categories. However, paired t-test showed that both Leptin and Adiponectin levels raised significantly on day 19 of induction (P = 0 and P = 0.015, respectively; Table 5; Figures 1–3).

The percentage of BMI at day 0/median was defined as a proportion of BMI at diagnosis within the median of the WHO reference and represents the difference between the measured value and the mean level for the same gender and age. The greater the difference between the BMI and reference value, the worse the MRD at day 46 (P = 0.014; Table 6).

Waist and hip circumference reflected the distribution of body fat, i.e., the concentric distribution. The results suggested that fat concentric distribution is significantly correlated with poor MRD at day 46 (Table 6).
Acute lymphoblastic leukemia is the most common pediatric cancer. Childhood survivors of acute lymphoblastic leukemia (SALL) are at risk of obesity and related cardiometabolic diseases, including type 2 diabetes, hypertension, stroke, and cardiovascular events (21).

We observed changes in BMI, waist circumference and MRD of the 82 enrolled ALL children during their remission-induction therapy (Table 1). In our study, 13 (15.6%) were overweight/obese and 11 (13.4%) were malnourished at diagnosis based on the age- and sex-adjusted BMI. Waist/hip circumference ratio in most children, i.e., 60 (73.2%) reached >1.0 on Day 19 during chemotherapy. Waist circumference increased in 22 children (27%) and hip circumference increased in 33 (40%) ALL children during remission-induction. A higher BMI was associated with significantly increased MRD at day 46 and showed a positive correlation (P = 0.014). In addition, we found positive correlations between weight, hip circumference, and MRD at day 46. This suggested that an increase in MRD was associated with concentric distribution of body fat. Our study suggested that monitoring and control of body weight during chemotherapy is important to the prognosis of the disease.

A meta-analysis suggested that a higher BMI at diagnosis was associated with poorer survival in children with pediatric ALL or acute myeloid leukemia (AML). Orgel et al. performed a meta-analysis of 4,690 cases of ALL and observed poorer event-free survival (EFS) in children with a higher BMI compared to children with lower BMI. A higher BMI was associated with significantly increased mortality and a statistically non-significant trend toward the greater risk of relapse compared to children with a lower BMI (22). At a time when obesity has become a global public concern, the use of hormone drugs such as dexamethasone administered during ALL treatment could cause obesity, especially in children from developing countries. Núñez-Enríquez et al. (9) performed a multicenter cohort study...
Body composition analysis also showed an increased incidence of skeletal muscle mass loss (24). Marriott CJC conducted body composition in 75 long-term survivors of ALL (more than 10 years after initial diagnosis). Excess fat mass and the combined morbidity of sarcopenic obesity were found to be prevalent in long-term survivors of ALL. This places ALL survivors at double jeopardy from excess body fat and inadequate skeletal muscle mass (SMM). This is associated with an adverse impact on overall survival (24). In our study, we found the correlation was not only between BMI at diagnosis and MRD day 46 but also between waist circumference and MRD at day 46. This finding suggests that body composition could also affect the outcome of ALL children.

Leptin is an effective adipocytokine to control obesity (12). We found that bone marrow leptin average levels as well as Adiponectin levels increased significantly during dexamethasone administration. Hypothalamic leptin resistance should be considered in obese patients with ALL (25). Obese leukemic survivors have been reported to have lower serum adiponectin levels compared to non-obese survivors (26), while dysregulation of the adipokine pathway has been associated with carcinogenesis and ALL (27). This study also confirmed that Leptin levels of those whose BMI was high at diagnosis were also prevalent in long-term survivors of ALL. This places ALL leukemic survivors have been reported to have lower serum adiponectin levels compared to non-obese survivors (26), while dysregulation of the adipokine pathway has been associated with carcinogenesis and ALL (27). This study also confirmed that Leptin levels of those whose BMI was high at diagnosis were also prevalent in long-term survivors of ALL. This places ALL

TABLE 4 | Patient minimal residual disease and adipocytokine levels based on body mass index.

| Patient characteristics | Total (n = 82) | BMI subgroups | P-value |
|-------------------------|--------------|---------------|---------|
| MRD on day 19, n (%)    |              |               |         |
| <0.01%                  | 53 (64.6)    | 5 (45.5)      | 4 (4.4) |
| ≥0.01 to <0.1%          | 15 (18.3)    | 2 (18.2)      | 0 (0)   |
| ≥0.1% to <1             | 7 (8.5)      | 1 (9.1)       | 0 (0)   |
| ≥1                      | 3 (3.7)      | 2 (18.2)      | 0 (0)   |
| No data                 | 4 (4.9)      | 1 (9.1)       | 0 (0)   |
| MRD on day 46, n (%)    |              |               |         |
| <0.01%                  | 65 (79.3)    | 2 (18.2)      | 0 (0)   |
| ≥0.01 to <0.1%          | 8 (9.8)      | 2 (18.2)      | 0 (0)   |
| ≥0.1% to <1             | 4 (4.9)      | 2 (18.2)      | 0 (0)   |
| ≥1                      | 0 (0)        | 0 (0)         | 0 (0)   |
| No data                 | 5 (6.0)      | 2 (18.2)      | 0 (0)   |
| Leptin levels on day 0  | 60.19 ± 51.00| 63.65 ± 58.45| 54.65 ± 47.84|
| Leptin levels on day 19 | 156.55 ± 116.09| 221.37 ± 120.58| 146.22 ± 106.90|
| Leptin levels on day 46 | 125.25 ± 111.55| 91.78 ± 47.15| 125.58 ± 109.35|
| Adiponectin levels on day 0 | 1,499.94 ± 1,304.77 | 887.99 ± 626.09 | 1,595.10 ± 1,364.26 |
| Adiponectin levels on day 19 | 1,746.82 ± 932.94 | 1,965.09 ± 1,623.12 | 1,788.11 ± 857.34 |
| Adiponectin levels on day 46 | 1,501.10 ± 1,031.11 | 491.02 ± 184.96 | 1,573.21 ± 1,079.82 |
| LAR on day 0            | 0.0660 ± 0.0901 | 0.0757 ± 0.0522 | 0.0552 ± 0.0708 |
| LAR on day 19           | 0.1150 ± 0.1368 | 0.1296 ± 0.0369 | 0.0963 ± 0.0927 |
| LAR on day 46           | 0.1192 ± 0.1655 | 0.2156 ± 0.3427 | 0.1111 ± 0.1442 |

BMI, body mass index; MRD, minimal residual disease; LAR, leptin-to-adiponectin ratio. P < 0.05 are shown in bold.

in a Mexican cohort of 1,070 children. Early mortality and early relapses were the main outcomes. Overweight/obesity at diagnosis were predictors of early mortality. In this study, the correlation between BMI at diagnosis and MRD at day 46 supports the results above. Importantly, the weight gain induced during remission-induction therapy (within 6 weeks after diagnosis) after glucocorticoid administration was further exacerbated after completion of therapy. This was regardless of the patients’ clinical characteristics, such as age, race, gender, or treatment risk (23).
MRD during remission induction and consolidation therapy is the most sensitive method to evaluate treatment response and one of the strongest predictors of outcome in childhood acute lymphoblastic leukemia. Several studies have demonstrated the prognostic significance of measuring MRD in childhood ALL, suggesting that MRD positivity at serial time points during the treatment is highly predictive of relapse, and associated with poor treatment outcome (29, 30). A recent study suggested that MRD levels at the end of remission induction therapy measured using multiparameter flow cytometry have clinical significance in childhood ALL. High levels of MRD were strongly related to poor treatment outcomes (31, 32). This study found that the MRD day 46 was correlated with higher age- and sex-adjusted BMI at diagnosis \( (P = 0.014) \). MRD day 46 seemed worse in patients with fat concentric distribution, i.e., increased waist circumference on diagnosis, day 19 \( (P = 0.010 \text{ and } P = 0.007) \). However, it did not seem to affect MRD through changes in adipocytokine levels.
There were several limitations to this study. Although we focused on the BMI of ALL children, the body composition measurements were not performed to access fat-free mass and fat distribution. In addition, longer longitudinal follow-ups were not performed on ALL children until the end of maintenance therapy and analysis for EFS and overall survival (OS) (33–36).

**CONCLUSIONS**

In the study, we found that 13 (15.6%) ALL children were overweight/obese and 11 (13.4%) were malnourished (wasting) at diagnosis based on age- and sex-adjusted BMI. A higher BMI was associated with significantly increased MRD at day 46 and showed a positive correlation ($P = 0.014$). In addition, we found a positive correlation between weight, hip circumference at diagnosis, days 19, and MRD at days 46. Both BMI and fat concentric distribution affected the outcome of ALL children. A higher BMI was also associated with significantly increased Leptin levels at diagnosis. Leptin resistance should be a risk factor in ALL children with high BMI.

To address this risk of weight gain, early interventions such as regular weight and height monitoring and dietary assessment involving a multidisciplinary team of oncologists, nurses, dietitians, physical therapists, psychologists, and endocrinologists should be consulted to evaluate ALL children, preferably starting during remission-induction chemotherapy (19, 37).

**ORIGINAL DETAILS OF RISK CLASSIFICATION**

**Total Therapy Study XV for Newly Diagnosed Patients With Acute Lymphoblastic Leukemia**

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**Risk Classification**

Patients were classified into one of three categories (low-, standard-, or high-risk) based on age at admission, leukocyte counts, presence or absence of central nervous system (CNS) status, or testicular leukemia, immunophenotype, cytogenetic and molecular diagnosis, DNA index, and early response to therapy. Hence, definitive risk assignment (for
provisional low-risk cases based on presenting features) will be made after completion of remission induction therapy. The criteria and the estimated proportion of patients for each category (based on data from Total XIII study) are provided below.

**Criteria for Low-Risk ALL (≈40% of Patients)**
- B-cell precursor ALL with DNA index $\geq 1.16$, TEL-AML1 fusion gene, or age 1–9.9 years and presenting WBC $< 50 \times 10^9$/L.
- Must not have (i) CNS 3 status ($\geq 5$ WBC/L of cerebrospinal fluid with morphologically identifiable blasts or cranial nervepalsy), (ii) overt testicular leukemia (evidenced by ultrasonogram), (iii) adverse genetic features [$t(9;22)$ or BCR-ABL fusion gene; $t(1;19)$ with E2A-PBX1 fusion gene; rearranged mixed lineage leukemia gene (MLL); or hypodiploidy ($\sim 45$ chromosomes)], or (iv) poor early response ($\geq 1\%$ lymphoblasts on day 19 or 26 of remission induction, $\geq 0.01\%$ lymphoblasts by immunologic or molecular methods on remission date).

**Criteria for Standard-Risk ALL (≈50% of Patients)**
- All cases of T-cell ALL and those with B-cell precursor ALL that do not meet the criteria for low-risk or high-risk ALL.

**Criteria for High-Risk ALL (≈10% of Patients)**
- $t(9;22)$ or BCR-ABL fusion.
- Induction failure or $>1\%$ leukemic lymphoblasts in the bone marrow on remission date.
- $>0.1\%$ leukemic lymphoblasts in the bone marrow in week 7 of continuation treatment (i.e., before re-induction 1, $\sim 14$ weeks post-remission induction).
- Re-emergence of leukemic lymphoblasts by minimal residual disease (MRD) (at any level) in patients previously MRD negative.
- Persistently detectable MRD at lower levels.

### Table 7: Pearson correlation analysis of body mass index and adipokine levels.

|                      | Lep1   | Lep2   | Lep3   | Adip1  | Adip2  | Adip3  |
|----------------------|--------|--------|--------|--------|--------|--------|
| BMI day 0            |        |        |        |        |        |        |
| Pearson correlation  | 0.535* | 0.34   | 0.007  | −0.091 | −0.119 | −0.164 |
| P-value              | 0.005  | 0.198  | 0.978  | 0.65   | 0.673  | 0.515  |
| Percentage of BMI-day 0 to medium |        |        |        |        |        |        |
| Pearson correlation  | 0.451* | −0.054 | −0.159 | −0.539 | −0.245 | −0.175 |
| P-value              | 0.031  | 0.908  | 0.682  | 0.135  | 0.597  | 0.629  |

BMI, body mass index; Lep1, Leptin levels at diagnosis; Lep2, Leptin levels at days 19 ± 2; Lep3, Leptin levels at days 46 ± 2; Adip1, Adiponectin levels at diagnosis; Adip2, Adiponectin levels at days 19 ± 2; Adip3, Adiponectin levels at days 46 ± 2. $P < 0.05$ are shown in bold. *$P < 0.05$; **$P < 0.01$.

### Data Availability Statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

### Ethics Statement

The studies involving human participants were reviewed and approved by No. 2017102415 file granted by the Ethics Committee of Guangzhou Women and Children’s Medical Center. Written informed consent to participate in this study was provided by the participants’ legal guardian/next of kin.

### Author Contributions

JS: conceptualization and writing—original draft. HH, MX, and HL: data curation. HC and JS: methodology. JC, RZ, LZ, JT, and XW: resources. XW and HH: supervision. JS and JC: writing—review and editing. All authors contributed to the article and approved the submitted version.

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### References

1. Pui CH, Yang JJ, Hunger SP, Pieters R, Schrappe M, Biondi A, et al. Childhood acute lymphoblastic leukemia: progress through collaboration. *J Clin Oncol.* (2015) 33:2938–48. doi: 10.1200/JCO.2014.59.1636
2. Hunger SP, Mullighan CG. Acute lymphoblastic leukemia in children. *N Engl J Med.* (2015) 373:1541–52. doi: 10.1056/NEJMc1400972
3. Pui CH, Yang JJ, Bhakta N, Rodriguez-Galindo C. Global efforts toward the cure of childhood acute lymphoblastic leukaemia. *Lancet Child Adolesc Health.* (2018) 2:440–54. doi: 10.1016/S2352-4642(18)30066-X
4. Inaba H, Greaves M, Mullighan CG. Acute lymphoblastic leukaemia. *Lancet.* (2013) 381:1943–55. doi: 10.1016/S0140-6736(12)62187-4
5. Browne EK, Inaba H. Obesity and height in children and adolescents with acute lymphoblastic leukemia and its future management. Oncotarget. (2019) 10:2123-4. doi: 10.18632/oncotarget.26653

6. Inaba H, Surprise HC, Pounds S, Cao X, Howard SC, Ringwald-Smith K, et al. Effect of body mass index on the outcome of children with acute myeloid leukemia. Cancer. (2012) 118:5989-96. doi: 10.1002/cncr.27640

7. Lange BJ, Gerbing RB, Feusner J, Skolnik J, Sacks N, Smith FO, et al. Mortality in overweight and underweight children with acute myeloid leukemia. JAMA. (2003) 293:203–11. doi: 10.1001/jama.293.2.203

8. Aarnivala H, Pokka T, Soininen R, Mötönen M, Harila-Saari A, Niinimäki R. Trends in age- and sex-adjusted body mass index and the prevalence of malnutrition in children with cancer over 42 months after diagnosis: a single-center cohort study. Eur J Pediatr. (2020) 179:91-8. doi: 10.1007/s00431-019-04382-w

9. Núñez-Enríquez JC, Gil-Hernández AE, Jiménez-Hernández E, Fajardo-Gutiérrez A, Medina-Sansón A, Flores-Lujano J, et al. Overweight and obesity as predictors of early mortality in Mexican children with acute lymphoblastic leukemia: a multicenter cohort study. BMC Cancer. (2019) 19:708. doi: 10.1186/s12888-019-5878-8

10. Zareifar S, Haghpanah E, Shokrallah E, Karamizadeh Z. Evaluation of metabolic syndrome and related factors in children affected by acute lymphoblastic leukemia. Indian J Med Paediatr Oncol. (2017) 38:97–102. doi: 10.4103/impo.impo_69_16

11. Orgel E, Sea JL, Mittelman SD. Mechanisms by which obesity impacts survival from acute lymphoblastic leukemia. J Natl Cancer Inst Monogr. (2019) 2019:152–6. doi: 10.1093/ncimonomographs/igg020

12. Izquierdo AG, Cruejeiras AB, Casanueva FF, Carreira MC. Leptin, obesity, and leptin resistance: where are we 25 years later? Nutrients. (2019) 11:2704. doi: 10.3390/nu11112704

13. Barbosa-Cortés L, López-Alarcón M, Mejia-Aranguré JM, Klünde-Münkler M, Del Carmen Rodriguez-Zepeda M, Rivera-Mármol H, et al. Adipokines, insulin resistance, and adiposity as predictors of metabolic syndrome in the child survivors of lymphoma and acute lymphoblastic leukemia of a developing country. BMC Cancer. (2017) 17:125. doi: 10.1186/s12885-017-3097-8

14. Pui CH, Campana D, Pei D, Bowman WP, Sandlund JT, Kaste SC, et al. Treating childhood acute lymphoblastic leukemia without cranial irradiation. N Engl J Med. (2009) 360:2730–41. doi: 10.1056/NEJMoa0900386

15. van Dongen JJ, Seriu T, Panzer-Grümayer ER, Biondi A, Ponger s-Willemse and van Dongen JJ, Seriu T, Panzer-Grümayer ER, Biondi A, Ponger s-Willemse. Changes in body mass index, height, and weight in children during and after therapy for acute lymphoblastic leukemia. Cancer. (2018) 124:2428–59. doi: 10.1002/cncr.31736

16. Marriott CJ, Beaumont LF, Farcombe TH, Cranston AN, Athale UH, Yakemchuk VN, et al. Body composition in long-term survivors of acute lymphoblastic leukemia diagnosed in childhood and adolescence: a focus on sarcopenic obesity. Cancer. (2018) 124:1225–31. doi: 10.1002/cncr.31191

17. Ogden CL, Flegal KM. Changes in terminology for childhood overweight and obesity. JAMA. (2015) 210:5195–7. doi: 10.4103/0010-0015.175824

18. Argyrou C, Hatzigaiapoulos K, Theodorakidou M, Nikolou S, Vlahopoulos S, Lambrou GI. The role of adiponectin, LEPTIN, and ghrelin in the progress and prognosis of childhood acute lymphoblastic leukemia. Leuk Lymphoma. (2011) 53:2158–69. doi: 10.1080/10428191.2011.1974290

19. Skopec S, Tomasik PJ, Fijorek K, Strojny W, Wieczorek A, Balwierz W, et al. Concentrations of adipokines in children before and after hematopoietic stem cell transplantation. Pediatr Hematol Oncol. (2016) 33:21–38. doi: 10.3109/08880018.2015.1135362

20. Czogala W, Strojny W, Schab M, Grabowska A, Mikluski K, Kowalczy W, et al. FTO and PLAG1 genes expression and FTO methylation predict changes in circulating levels of adipokines and gastrointestinal peptides in children. Nutrients. (2021) 13:3585. doi: 10.3390/nu131103585

21. Czogala W, Czogala M, Strojny W, Wator G, Walkow R, Wójcik M, et al. Methylation and expression of FTO and PLAG1 genes in childhood obesity: insight into anthropometric parameters and glucose-lipid metabolism. Nutrients. (2021) 13:1683. doi: 10.3390/nu131051683

22. Skopec S, Tomasik PJ, Bikut-Multanowski W, Surmiak M, Balwierz W, Pietrzyk J, et al. Plasma levels of leptin and soluble leptin receptor and polymorphisms of leptin gene—118 G>A and leptin receptor genes K109R and Q223R, in survivors of childhood acute lymphoblastic leukemia. J Exp Clin Cancer Res. (2011) 30:66. doi: 10.1186/1756-9966-30-66

23. Borowicz MJ, Devidas M, Hunger SP, Bowman WP, Carroll AJ, Carroll WL, et al. Clinical significance of minimal residual disease in childhood acute lymphoblastic leukemia and its relationship to other prognostic factors: a Children's Oncology Group study. Blood. (2008) 111:5477–85. doi: 10.1182/blood-2008-01-132837

24. Czogala W, Strojny W, Tomasik M, Bikut-Multanowski W, Wójcik M, Mikluski K, et al. The insight into insulin-like growth factors and insulin-like growth-factor-binding proteins and metabolic profile in pediatric obesity. Nutrients. (2021) 13:2432. doi: 10.3390/nu13072432

25. Jovanovska A, Martinezova K, Kocheva S, Trajkova-Antevska Z, Coneska-Jovanova B, Panovska-Stavrids I, et al. Clinical significance of minimal residual disease at the end of remission induction therapy in childhood acute lymphoblastic leukemia. Open Access Maced J Med Sci. (2019) 7:2818–23. doi: 10.3889/oamjms.2019.752

26. Basso G, Veltroni M, Valsecchi MG, Dowerak MN, Ratei R, Silvestri D, et al. Risk of relapse of childhood acute lymphoblastic leukaemia is predicted by flow cytometric measurement of residual disease on day 15 bone marrow. J Clin Oncol. (2009) 27:5168–74. doi: 10.1200/JCO.2008.176016

27. Campana D. Minimal residual disease monitoring in childhood acute lymphoblastic leukemia. Curr Opin Hematol. (2012) 19:313–8. doi: 10.1097/MOH.0b013e328354345c
37. Lyu A, Triplett TA, Nam SH, Hu Z, Arasappan D, Godfrey WH, et al. Tumor-associated myeloid cells provide critical support for T-ALL. Blood. (2020) 136:1837–50. doi: 10.1182/blood.2020007145

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