Increased Nuclear Localization of $\beta$ - catenin by rhBMP - 2 in the Induced Oral Squamous Cell Carcinoma in the Syrian Hamster Cheek Pouch

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Abstract

Background: Recombinant human bone morphogenetic protein 2 (rhBMP - 2) regulates several functions within human cells, including invasion, adhesion, differentiation, proliferation, and programmed death. The intercellular role of this protein in cancer progression has been extensively investigated in recent studies. Alterations in $\beta$ - catenin could also promote cancer progression through the Wnt/$\beta$ - catenin signaling pathway. The present study aimed to evaluate the effect of rhBMP - 2 on the expression and localization of $\beta$ - catenin in the induced oral squamous cell carcinoma (OSCC) in the buccal pouches of golden Syrian hamsters.

Methods: Buccal cavities of 57 hamsters were scrubbed by 5% 7,12 - dimethylbenz[a]anthracene (DMBA) three times per week for 3.5 months. In addition, the animals in the test group (n = 29) were injected with 0.25 $\mu$g of rhBMP-on days one, seven, and 49 of the study, and biopsies were extracted afterwards. The expression of $\beta$ - catenin was immunohistochemically studied in the affected tissues.

Results: Membranous $\beta$ - catenin expression was observed in 71.1% of the cells in the animals in the test group and 64.8% of the control subjects. However, no significant difference was observed between the study groups (P > 0.05). Moreover, nuclear $\beta$ - catenin expression was detected in 39.1% and 23.6% of the cells in the test and control groups, respectively with a significant difference in this regard (P < 0.05).

Conclusions: According to the results, rhBMP - 2 increased the nuclear localization of $\beta$ - catenin in the OSCC and activated the Wnt/$\beta$ - catenin signaling pathway. However, evidence is scarce on the increased invasiveness due to the loss of $\beta$ - catenin membranous expression.

Keywords: rhBMP - 2, $\beta$ - catenin, Oral Squamous Cell Carcinoma

1. Background

Bone morphogenetic proteins (BMPs) are the largest subfamily of the TGF/$\beta$ protein superfamily, which participate in several developmental processes and signaling pathways within living cells; as such, researchers have recommended changing the name to body morphogenetic proteins (1). This family of proteins control many stages of cellular differentiation in various embryonic cell types in order to generate distinct constituents (2). During embryonic maturation, BMPs play a key role in the induction of apoptosis, which is essential to the elimination of ineffective cells and tissues (3).

Recent studies have revealed that BMPs are also involved in tumor development, exerting antitumorigenic effects on the primary stages and protumorigenic effects on progressive carcinogenesis (4). Although BMPs are considered to be multifunctional proteins, the implantation of osteoinductive proteins, such as BMP - 2, in the bones or other tissues results in bone formation. BMP - 2 particles are dissolvable and act locally to bind to the definite receptors on the cell membrane. These receptors transfer signals through specific molecules known as small mothers against decapentaplegic (SMADs), which are able to stimulate specific genes (5).

BMP receptors (BMPRs) are located on the cell membranes containing serine/threonine kinase receptors,
which are comprised of an extracellular short division consisting of 10 - 12 cysteine tails, a transmembrane division, and a serine - threonine kinase division within the cell.

Researchers have defined five categories of type I BMP-PRs and three categories of type II BMP-PRs (6). Accordingly, several studies have indicated the sequence of BMP-2 signaling, including the SMAD signals from the clathrin-coated pits and SMAD pathways starting within detergent-resistant membranes, which contain caveolae and are responsible for the regulation of SMAD signaling (7). However, the process remains unclear due to the contradictory reports and lack of studies in this regard.

The current pattern of BMP-2 signals mainly involves the controlling of SMADs (1, 4, 5), which are combined with LEF1 - TCF in order to induce several critical processes in cells and tissues, such as reproduction, mobility, and metaplasia (8-10).

Translocation of β-catenin to the nucleus seems to be the final stage of the Wnt cascade, which promotes β-catenin gene transcription. As the transcriptional mediator in the nucleus, β-catenin binds to the TCF and LEF factors, stimulating the essential β-catenin collaboration for genes transcription (11).

Two systems are involved in the process mentioned in Figure 1, including the binding of 1-TCF to the Groucho receptors to enhance the deacetylation of the histones and chromatin compression and suppress the expression of the targeted genes, and Wnt-induced β-catenin stability in the cell plasma to enable the transfer of β-catenin to the cell nuclei for TCF binding. With the assistance of other mediators, this process loosens the chromatin and enables genomic expression (12).

2. Methods

2.1. Studied Animals

The study was conducted on 60 male golden Syrian hamsters aged 8 - 10 weeks with the weight of 80 - 120 grams. The animals were equally divided into two groups of test and control in the animal houses of the Faculty of Pharmacy at Damascus University, Syria. The subjects were kept in polypropylene enclosed cages and subjected to a standard nutritional system comprised of lipids (6%), protein (20%), ash (12%), calcium (0.9%), glucose (3.5%), vitamins (2.2%), nitrogen - free nutrients (56%), and water. In addition, stable temperature (29 ± 1°C) and humidity (48 ± 9%).

One hamster in the test group (BMP - 2) and two animals in the control group did not survive to the end of the experiment. In this study, 5% 7,12 - dimethylbenz[a]anthracene (DMBA) dissolvable in paraffin wax (Sigma Aldrich Chemical Pvt. Ltd) and rhBMP - 2 (0.25 µg/ml) soluble in physiological serum were purchased from Cowellmedi Co. Ltd, Korea.

2.2. Tumor Induction and Experimental Procedures

Carcinogenesis was stimulated in the buccal pouches of 75 golden Syrian hamsters by the applying 0.5% DMBA in the liquid wax of topical paraffin and repeated three times per week for 3.5 months. The animals in the test group (n = 29) were administered with rhBMP - 2 (0.25 µg/ml) through injection into the submucosal tissue of the buccal pouch on days one, seven, and 49. Afterwards, they were sacrificed, and tissue samples were extracted and preserved in 10% buffered formalin.

2.3. Histopathological Studies

The mucosal tissues of buccal pouches were examined histopathologically in the test and control groups. Following that, the biopsies were fixed in 10% buffered formaldehyde, processed, and embedded in paraffin wax. Two serial cross-sections (thickness: 4 µm) were placed on silanized slides for deparaffinization and rehydration with dimethylbenzene and three descending grades of alcohol.

Antigen retrieval was performed under pressure in an oven for five minutes. The samples were incubated after covering with 3% hydrogen peroxide for 15 minutes in order to prevent endogenous peroxidase activity. Additionally, the primary anti-β-catenin monoclonal antibody (Dako Cytomation, USA, clone β-catenin-1) was added to the slides (n = 57), which were covered again for four hours at the temperature of 27°C using a dilution (6 µg/ml) in accordance with the instructions of the manufacturer.

After additional incubation with the secondary antibody (45 minutes) and streptavidin peroxidase (30 minutes), visualization was carried out for 10 minutes using a freshly arranged diaminobenzidine (DAB) chromogen. Finally, the specimens were stained in Harris hematoxylin (Sigma Aldrich Chemical Pvt. Ltd) and rhBMP-2 (0.25 µg/ml) soluble in physiological serum were purchased from Cowellmedi Co. Ltd, Korea.
Figure 1. Link Between BMP-2 and β-catenin Through SMADS (8)

Figure 2. Buccal Pouch of Animals After 14 Days of DMBA Application

Figure 3. Expression of β-catenin in BMP-2 Group

These fields were determined and investigated for the localization of β-catenin. The epithelial cell showing normal, intact membranous staining and the cells with cytoplasmic, nuclear or incomplete membranous expression were considered as irregular cells for β-catenin.

3. Results

Most of the tissue samples exhibited positive expression patterns for β-catenin. Membranous expression of β-catenin was observed in 71.1% of the cells in the test group and 64.8% of the cells in the control group; however, no significant difference was observed in this regard (P > 0.05) (Table 1, Figures 3 and 4).

Nuclear β-catenin expression was detected in 39.1% of the cells in the test group and 23.6% of the cells in the control group, showing a significant difference in this regard (P < 0.05) (Table 2 and Figure 5).
Table 1. Expression of \( \beta \)-catenin in Study Groups

| Group   | N | Median | Maximum Value | Minimum Value | Standard Deviation |
|---------|---|--------|---------------|---------------|-------------------|
| Control | 28| 64.8%  | 82.2%         | 50.6%         | 10.1%             |
| BMP - 2 | 29| 71.1%  | 81.3%         | 46.2%         | 13.7%             |

Table 2. Nuclear Expression of \( \beta \)-catenin in Study Groups

| Group   | N  | Median | Maximum Value | Minimum Value | Standard Deviation |
|---------|----|--------|---------------|---------------|-------------------|
| Control | 28 | 23.6%  | 33.9%         | 10.7%         | 8.1%              |
| BMP - 2 | 29 | 39.1%  | 47.6%         | 17.2%         | 11.7%             |

4. Discussion

BMPs play a key role in tissue growth, organizing multiple processes during the formation of the skeleton, hematopoiesis, formation of the nerve tissues, and determining the differentiation plan in embryonic cells. These proteins were discovered in the 1960’s and have been extensively studied over the past 60 years. Nevertheless, further investigation is required to reveal the other aspects of their functions (13).

BMP signals initiate with their binding to specific type I and II receptors. The genetic control is mediated by various pathways, which may involve SMAD - independent and SMAD - dependent signals. The current study has focused on the association of BMP - 2 and one of the most critical agents in the Wnt signaling pathway, known as the \( \beta \)-catenin protein.

\( \beta \)-catenin is responsible for organizing the harmony of cell - cell adhesion and gene transcription; therefore, so it has a dual function within the cells. In humans, \( \beta \)-catenin is coded by the CTNNB1 gene. The shift between these functions is organized by several factors, including molecular structure and stabilization and the presence of epithelial - cadherin-mediated cell - cell bonding (14).

In the present study, no statistical difference was observed in the expression of \( \beta \)-catenin after the injection of rhBMP - 2 (0.25 \( \mu \)g/ml) into the submucosal tissue of the buccal pouch in golden Syrian hamsters with induced squamous cell carcinoma. However, nuclear localization had a significant difference between the study groups, and \( t \) was more significant in the BMP - 2 group compared to the control group. This finding indicated the interaction between BMP - 2 and Wnt signaling pathway, where \( \beta \)-catenin may differ based on its cellular localization.

As \( \beta \)-catenin translocates into the cell nuclei, it produces specific compounds with the TCF/LEF transcriptional factor. The connection enhances the expression of many targeted genes and activates the Wnt signaling pathway (15). As a result, rhBMP - 2 leads to cell multiplication, dislocation, invasion, and epithelial - mesenchymal transition through the nuclear translocation of \( \beta \)-catenin (8). Interestingly, \( \beta \)-catenin expression in the nuclei is often associated with the LEAST favorable results in the patients undergoing postoperative chemotherapy or radiotherapy.

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In the current research, the results of multivariate analysis indicated that β-catenin expression in the nuclei is an independent prognostic indicator in cervical squamous cell carcinoma (16). Furthermore, several studies have confirmed the association between the nuclear localization of β-catenin and the high-grade serous carcinomas of the ovaries (17). It has also been reported that the abnormal expression of β-catenin is an essential predictor of histological differentiation in the patients with oral squamous cell carcinoma (OSCC) and is associated with poorly differentiated OSCC (14). Therefore, disorganization of β-catenin may contribute to the pathogenesis of many types of carcinogenesis.

In conclusion, the results of the present study indicated that rhBMP-2 increases the nuclear localization of β-catenin in OSCC and activates Wnt/β-catenin signaling pathway. However, evidence is scarce regarding the role of rhBMP-2 in increasing invasiveness due to the loss of β-catenin membranous expression. Our findings could be beneficial in recognizing the signaling and mechanism of action of BMPs, allowing clinical applications in the future. It is hoped that all the influential factors and molecules and signaling pathways of these proteins will be established clearly in further investigations.

References
1. Wagner DO, Sieber C, Bhushan R, Borgermann JH, Graf D, Knaus P. BMPs: from bone to body morphogenetic proteins. Sci Signal. 2010;3(107):mr1. doi: 10.1126/scisignal.3107mr1. [PubMed: 20124549].
2. Varga AC, Wrana JL. The disparate role of BMP in stem cell biology. Oncogene. 2005;24(37):5713-21. doi: 10.1038/sj.onc.1208919. [PubMed: 1621804].
3. Xiao YT, Xiang LX, Shao JZ. Bone morphogenetic protein. Biochem Biophys Res Commun. 2007;362(3):550-3. doi: 10.1016/j.bbrc.2007.08.045. [PubMed: 177956].
4. Blanco Calvo M, Bolos Fernandez V, Medina Villamil V, Aparicio Gallego G, Diaz Prado S, Grande Pulido E. Biology of BMP signalling and cancer. Clin Transl Oncol. 2009;11(3):326-37. [PubMed: 19293049].
5. Ebara S, Nakayama K. Mechanism for the action of bone morphogenetic proteins and regulation of their activity. Spine (Phila Pa 1976). 2002;27(16 Suppl):S50-5. [PubMed: 12205418].
6. Nohe A, Keating E, Knaus P, Petersen NO. Signal transduction of bone morphogenetic protein receptors. Cell Signal. 2004;16(3):298-9. [PubMed: 14687659].
7. Saldanha S, Bragdon B, Moseychuk O, Bonor J, Dhurjati P, Nohe A. Caveolae regulate Smad signaling as verified by novel imaging and system biology approaches. J Cell Physiol. 2011;228(5):1060-9. doi: 10.1002/jcp.22453. [PubMed: 23049790].
8. Zaid KW, Chantiri M, Bassit G. Recombinant human bone morphogenetic Protein-2 in development and progression of oral squamous cell carcinoma. Asian Pac J Cancer Prev. 2016;17(3):2927-32. [PubMed: 27039814].
9. Sieber C, Kopf J, Hiepen C, Knaus P. Recent advances in BMP receptor signaling. Cytokine Growth Factor Rev. 2009;20(5-6):343-55. doi: 10.1016/j.cytogfr.2009.10.007. [PubMed: 19897402].
10. Hartung A, Bitton-Worms K, Rechtman MM, Wenzel V, Borgermann JH, Hassel S, et al. Different routes of bone morphogenetic protein (BMP) receptor endocytosis influence BMP signaling. Mol Cell Biol. 2006;26(20):7791-805. doi: 10.1128/MCB.00022-06. [PubMed: 16929661].
11. Arce I, Yokoyama NN, Waterman ML. Diversity of LEF/TCF action in development and disease. Oncogene. 2006;25(57):7492-504. doi: 10.1038/sj.onc.1208056. [PubMed: 1743291].
12. Gonzalez-Moles MA, Ruiz-Avila I, Gil-Montoya JA, Plaza-Campillo J, Scully C. beta-catenin in oral cancer: an update on current knowledge. Oral Oncol. 2014;50(9):818-24. doi: 10.1016/j.oraloncology.2014.06.005. [PubMed: 2499889].
13. Bragdon B, Moseychuk O, Saldanha S, King D, Julian J, Nohe A. Bone morphogenetic proteins: a critical review. Cell Signal. 2011;23(4):609-20. doi: 10.1016/j.cellsig.2010.10.003. [PubMed: 2095940].
14. Zaid KW. Immunohistochemical assessment of E-cadherin and beta-catenin in the histological differentiations of oral squamous cell carcinoma. Asian Pac J Cancer Prev. 2014;15(20):8847-53. [PubMed: 25374218].
15. Ebert MP, Yu J, Hoffmann J, Rocco A, Rocken C, Kahmann S, et al. Loss of beta-catenin expression in metastatic gastric cancer. J Clin Oncol. 2003;21(9):1708-14. doi: 10.1200/JCO.2003.10.077. [PubMed: 12271245].
16. Zhang Y, Liu B, Zhao Q, Hou T, Huang X. Nuclear localization of beta-catenin is associated with poor survival and chemotherapeutic resistance in human cervical squamous cell cancer. Int J Cell Biol. 2014;2014:7(7):2908-17. [PubMed: 252076].
17. Lee CM, Shvartsman H, Deavers MT, Wang SC, Xia W, Schmandt R, et al. beta-catenin nuclear localization is associated with grade in ovarian serous carcinoma. Gynecol Oncol. 2004;14(3):363-8. [PubMed: 12648588].