Review Article

Oxidative Stress-Related Mechanisms in SARS-CoV-2 Infections

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The COVID-19 pandemic caused relatively high mortality in patients, especially in those with concomitant diseases (i.e., diabetes, hypertension, and chronic obstructive pulmonary disease (COPD)). In most of aforementioned comorbidities, the oxidative stress appears to be an important player in their pathogenesis. The direct cause of death in critically ill patients with COVID-19 is still far from being elucidated. Although some preliminary data suggests that the lung vasculature injury and the loss of the functioning part of pulmonary alveolar population are crucial, the precise mechanism is still unclear. On the other hand, at least two classes of medications used with some clinical benefits in COVID-19 treatment seem to have a major influence on ROS (reactive oxygen species) and RNS (reactive nitrogen species) production. However, oxidative stress is one of the important mechanisms in the antiviral immune response and innate immunity. Therefore, it would be of interest to summarize the data regarding the oxidative stress in severe COVID-19. In this review, we discuss the role of oxidative and antioxidant mechanisms in severe COVID-19 based on available studies. We also present the role of ROS and RNS in other viral infections in humans and in animal models. Although reactive oxygen and nitrogen species play an important role in the innate antiviral immune response, in some situations, they might have a deleterious effect, e.g., in some coronaviral infections. The understanding of the redox mechanisms in severe COVID-19 disease may have an impact on its treatment.

1. Introduction

Patients with pneumonia of unknown etiology had been diagnosed in mid-December 2019 in Wuhan (Hubei province, China). Later, the SARS-CoV-2 (severe acute respiratory syndrome) coronavirus started to spread all over the world, without any exemptions. As for today, more than 182 million of patients have been infected, and more than 3.9 million died due to COVID-19 [1], providing an estimate of the mortality rate at 3.3%. When compared to the seasonal flu, COVID-19 related mortality is at least 60 times higher. Seasonal flu outbreak annually causes the infection of 3 to 5 million people, both asymptomatic and symptomatic, with mortality rate not exceeding 0.05% [2]. Clinical course of COVID-19 may, in most cases, consist of three periods [3]. After a short incubation period lasting from 2 to 5 days, patients become symptomatic, with the loss of sense of taste and olfactory dysfunction, dry cough, fever exceeding 38°C, and dyspnoea. Other symptoms, including headache, fatigue, diarrhoea, and conjunctivitis, are less frequent. Additionally, most patients develop a bilateral interstitial pneumonia [4]. After 7-10 days, dyspnoea decreases in majority of patients, inflammatory changes in the lungs resolving to some extent, and the patients are free from the virus in most cases. In severe COVID-19, the pneumonia causes a rapid drop in arterial pO₂, levels with the transcutaneous saturation measurement, usually below 60% when breathing ambient air. The progressive respiratory failure due to the loss of lung active surface of gas exchange and vascular abnormalities leads to the need of noninvasive ventilation support. In most severe cases, patients suffer from disseminated intravascular coagulation (DIC) or a septic shock and have to be sedated and undergo ventilation support [5].

Some experimental data available so far has suggested that the severe COVID-19 course might be related to the viral load during the SARS-CoV-2 exposure [6]. A recent study performed in 1145 patients suggested a significant independent association between viral load and mortality (with the hazard ratio of 1.07 [95% CI 1.03–1.11], p = 0.0014) implying that the 7% increase in mortality risk was...
present for each log transformed copy of viral RNA per mL of nasopharyngeal swab sample [7]. Another important factor, probably protecting from the development of severe COVID-19, is a normal to high level of serum vitamin D [8, 9]. Smoking cigarettes, however, may increase the risk of severe course of the disease, even in the absence of smoking-related disease [10–12]. The well-known and widely accepted hypothesis is that the male sex, hypertension, COPD, diabetes, or cancer may deeply influence the severity of the disease [13–16].

Today, it is not clear whether bronchial asthma may have any effect on the infection rate or the severity of COVID-19. Moreover, the question of how and why the viral pneumonia leads to DIC and septic shock with cytokine and bradykinin storms remains to be elucidated. ROS and RNS play an important role in the innate immune response, which is also directed against viruses [17]. In this review, we focus on the possible role of ROS and RNS in severe COVID-19 pathogenesis.

2. Antiviral Immune Response Mechanisms

The immune system has the potential to effectively control viral infections, and thus, it can limit their effect on the host organism. The processes of virus entry into the host cell, its replication, stimulation, and regulation of the antiviral immune response trigger a complex series of interactions between the virus and the host [18] (Figure 1(a)). There are two defense mechanisms: specific, acquired immunity and nonspecific, innate immunity. Nonspecific immunity is the first line of defense against infection and does not depend on prior contact with the pathogen. Mast cells, NK (natural killers), NKT (natural killer T cells), NHC (natural helper cells), natural lymphoid cells, granulocytes, macrophages, and monocytes are responsible for innate immunity. The pathophysiology of the extremely high pathogenicity of coronaviruses is not fully understood [19, 20]. It is worth noting that the immune system must develop a specific cytotoxic T cell (CTL) response. CTLs have the ability to recognize the viral-derived peptide on the surface of the infected cell, specifically in the MHC (the major histocompatibility complex) class I binding groove. Then, lymphocytes recognize the infected cell and destroy it by secreting cytolytic granules or activating programmed death in the cell through receptors such as FAS. In parallel with the development of the cellular response, a humoral immune response develops—associated with the activation of B cells and the subsequent release of specific antibodies. The helper T cells are at the center of the activation of adaptive immunity.

The lung epithelium is the largest surface that comes into contact with the environment. In the airways, viruses are detected by airway epithelial cells, mast cells, and cells of the mononuclear phagocyte system. The sensor cells are equipped with pattern recognition receptors such as Toll-like receptors (TLR), PAMPs (pathogen associated molecular patterns), derived from viruses, trigger a specific combination of PRRs (pattern recognition receptors) and adapter molecules, leading to the immune response adapted to the pathogen [21]. Coronavirus replication leads to, e.g., disruption of lysosomes, damage of mitochondria, and imbalanced ion concentrations [22, 23]. As a consequence, pyroptosis occurs, which initiates the secretion of proinflammatory molecules of the interleukin-1 family [24, 25] (Figure 1(b)). Coronavirus SARS-CoV-2-induced cell death releases histones and a high-mobility group box 1, which are normally hidden from recognition by PRRs. Then, additional proinflammatory cytokines and chemokines are produced, e.g., IL-6, IP-10, MIP1αβ (macrophage inflammatory proteins-1αβ), and MCP1 (monocyte chemotractant protein-1). Only in theory, detection of CoVs by pattern recognition receptors triggers an innate immune response that would be effective to limit viral replication. Interferons (IFN)-αβ, and type III are released to help control/eliminate viral infection. Their function is to remove the virus from infected cells by activating ISGs (IFN-stimulated genes) which exert direct antiviral effects, i.e., recruit antiviral immune effector cells. It has been observed that during zoonosis, the antiviral immune response can be detrimental to the body if the timing and target tissue of the immune response are inadequate [26].

The mechanism of innate immunity leads to inflammation, release of IFN-αβ, and activation of NK cells, which allows the suppression of local infection. Unfortunately, coronaviruses have developed strategies to protect themselves or their by-products from being recognized by the host [27]. In addition, viruses inhibit interferon induction and block IFN signaling. For example, SARS-CoV-1 (the coronavirus emerged in 2003, causing severe acute respiratory syndrome coronavirus) can effectively suppress interferon expression by nonstructural and structural proteins [28]. Coronaviruses circumvent the early phase of the innate immune response. Generalizing, the virus is recognized due to the stimulation of Toll-like receptors located on the epithelium and on dendritic cells, which are designed to inform B and T lymphocytes about the invasion of the pathogen. In the case of coronaviruses, these are Toll-like receptors 7 and TLR8 receptors that recognize viral RNA. Viral proteins are recognized by TLR2 and TLR4 receptors. During SARS-CoV-2 infection, the level of these receptors decreases, and their expression is lower in the elderly. SARS-CoV-2 infection is dangerous when a patient lacks specific antibodies and specific CTLs, because it can progress to severe pneumonia and ARDS [29].

3. The Role of ROS and RNS in Antiviral Response

The generation of ROS is one of the major mechanisms leading to infected cell death through apoptosis or necrosis, specifically during the very early stages of the immune response [30]. Both ROS and RNS also play an important role in signal transduction. Viral proteins or nucleic acids triggering the pattern recognition receptors lead to activating the interferon response through TLR-domain-containing adapter-inducing interferon (TRIF) and interferon regulatory factors (IRFs) as well as increasing in the inducible nitric oxide synthase (iNOS) expression and activity through the myeloid differentiation primary response-88 (MyD-88) adapter protein [31]. These
processes lead to an increase in the RNS production. The RNS might inhibit viral proliferation in infected cells [32, 33]. Similarly, both PRRs and interferon type I pathways lead to an increase in ROS production from the xanthine oxidase, nitric oxide synthase, or the mitochondrial respiratory reactions. These processes have been crucial in the innate immune response to various viruses including human respiratory viruses (influenza viruses, HRSV (human respiratory syncytial virus), and rhinoviruses). ROS are signaling molecules regulating a wide variety of physiological functions. ROS are a part of the mechanisms leading to the elimination of virus-infected cells and patient recovery. In some rare cases, specifically in the case of influenza infection, a severe course of the disease develops, leading to a severe adult respiratory distress syndrome (ARDS) with significant mortality [34]. Why ARDS is more frequent in some coronavirus infections (SARS, MERS (Middle East respiratory syndrome coronavirus), and SARS-CoV-2)
remains unknown. Therefore, ROS and RNS might be at least one of the important diseases modifying pathways in severe COVID-19.

The effects caused by the reactive forms of oxygen and nitrogen might depend on the source of their origin. For instance, many RNA viruses activate endosomal NADPH (nicotinamide adenine dinucleotide phosphate hydrogen) oxidase via Toll-like receptor 7 mechanism, activated in turn by binding to single-stranded RNA [35, 36]. This is likely because these viruses, when attached to the cell, are built into the endosomes and their RNA can interact with TLR7; SARS-CoV-2 might activate Nox2 (NADPH oxidase 2) through TLR7 and that might have a negative impact on the defense mechanism against viruses. This is due to the fact that Nox2 activation is used by viruses in order to restrain immune reactions and develop the infection [36, 37].

Overproduction of toxic ROS and excessive inflammation are harmful for tissues and may cause their damage [38–41]. As a result of an uncontrolled inflammatory response, oxidative stress (an imbalance between oxidants and antioxidants) arises, which in turn stimulates inflammatory cells to further produce cytokines and a “vicious circle” occurs (Figure 2).

The characteristic features of the severe form of COVID-19 include, but are not limited to, severe lymphopenia, lung tissue damage, and a “cytokine storm” leading to acute respiratory distress and multiorgan failure. Despite a central role of mitochondria in ROS generation, many questions remain unanswered about their role during the “cytokine storm” and pathogenesis of infections with coronaviruses. Lymphopenia causes, among others, a defect in the regulation of anti-viral immunity. The cytokine storm begins with the intense activation of cytokine-secreting cells with innate and acquired immune mechanisms [42] (Figure 3). It should be pointed out that in the case of a “cytokine storm”, neutrophil apoptosis does not occur. Patients have a huge amount of neutrophils that have undergone NETosis (NET-neutrophil extracellular traps). During NETosis, neutrophil extracellular trap is formed, and along with the “spilling out” of neutrophil DNA outside the cell, toxic enzymes are released, such as elastase, which damages lung tissue [43]. Moreover, microclots in the pulmonary circulation are formed. In the blood of COVID-19 patients, immune changes characteristic of viral infections were observed, i.e., increased levels of ASC-producing cells, activated CD4+ T cells and CD8+ T cells and IgM and IgG antibodies [44, 45]. Importantly, “cytokine storm” may occur, responsible for lung tissue damage during viral respiratory infections [46, 47]. Such sustained ROS production leads to the vicious circle that results in inflammatory damage but also hinders treatment of damage [48].

4. ROS and RNS Generation in SARS, MERS, and COVID-19

The high mortality rates of SARS-CoV-1, SARS-CoV-2, and MERS motivate scientists to study these infections in a variety of ways to find any effective therapeutic options. While numerous studies confirm a strong association between oxidative and nitrosative stress and severity of various viral infections (HCV (hepatitis C virus) [49], HBV (Hepatitis B virus) [50], and HRSV [51]), there is still limited clinical data showing such dependence in case of the SARS-CoV, SARS-CoV-2, and MERS infection—their severity or progression [52]. Previous research demonstrated that in SARS-CoV-infected human lung samples, explicit production of oxidized phospholipids followed by ROS generations was observed in the injured air spaces, pneumocytes, and alveolar macrophages [53]. Moreover, in macrophages, the oxidized phospholipids have been shown to modulate lung injury severity by TLR4-TRIF-TRAF6 expression and trigger cytokine production [22]. Lin et al. published a study showing that the ROS-activated NF-κB (nuclear factor kappa-light-chain-enhancer of activated B cells) signal transduction pathway is induced by SARS-CoV-1 protease-3CLpro and therefore might be involved in the SARS-CoV infection development [54].

Angiotensin converting enzyme-2 (ACE2), known as the cell entry receptor of the SARS-CoV-2, is a multifunctional transmembrane protein. ACE2 plays a double-edged role in SARS-CoV-2 infection, and apart from being the cellular receptor for SARS-CoV-2 spike proteins, it is the critical molecule in combating inflammatory and oxidative damage of tissues by COVID-19. This enzyme decreases angiotensin II which is stimulant of NADPH oxidase. In addition, the product of ACE2 enzymatic activity, angiotensin 1-7, has a strong antioxidant effect [55, 56].

The virus binding to ACE2 receptor initiates its entry to the cell, and after attachment and virion-membrane fusion, ACE2 expression is downregulated [57, 58]. The viral
Consequently, even death of the patient [24].

Angiotensin II availability is increased through the high hippuric acid and other viral infections [59]. In the course of SARS-CoV-2 infection, during other viral infections but also releasing in oxidative stress mechanisms (in contrast to what happens to the redox balance, i.e., accumulated oxidative damage and a deteriorated antioxidative defense system followed by increased reactive oxygen species [66]. As a consequence, induction of proinflammatory cytokine expression occurs (such as TNF-α, interleukin (IL) 6, IL-8, and IL-1β), via redox-sensitive transcription factors, e.g., NF-κB [67, 68].

Previous genomic analyses of SARS-CoV-1 on aged macaques demonstrated that old subjects presented stronger host response to virus and more severe infection pathology than young ones; this was associated with a reduced expression of type I interferon and an increase in the differential expression of inflammatory genes related to NF-κB [66].

Recent study demonstrated that patients suffering from severe COVID-19 disease, requiring intensive care unit treatment, presented higher levels of Nox2 activation, and thus, Nox2 seems to be a pivotal agent in COVID-19 aggravation [37]. However, Li et al. published data suggesting that the SARS-CoV nonstructural protein nsp10 might impair the redox system in the mitochondria, another ROS source, by a loss in the cellular inner mitochondrial membrane potential. This effect probably enhanced the cytopathic effect of SARS-CoV-1 [69]. Interestingly, it has been shown recently that coronaviruses, thanks to the protein nsp10 in combination with nsp16, can methylate the 5′ ends of their mRNAs, thus resembling the host mRNA and protecting them from the innate immune response [70].

Moreover, inflammatory cytokines-TNF-α and IL-6, which may initiate mitochondrial ROS production and are associated with ATP production, were found in COVID-19 serum (Figure 4) [71, 72]. In fact, Saleh et al. proposed recently a hypothesis that, apart from the intracellular mitochondria failure that plays a key role in COVID-19 disease, the extracellular mitochondria are important mediators [73–76]. They provoke the immune response, regulate cell-to-cell communication, and danger sensing [77]. According to the authors, this complex interplay between platelet mitochondrial dysfunction, oxidative stress, and mitophagy would provide useful therapeutic strategies [73].

The severity of coronavirus infections is generally age related [65], which might be attributed to a disruption in the redox balance, i.e., accumulated oxidative damage and a deteriorated antioxidative defense system followed by increased reactive oxygen species [66]. As a consequence, induction of proinflammatory cytokine expression occurs (such as TNF-α, interleukin (IL) 6, IL-8, and IL-1β), via redox-sensitive transcription factors, e.g., NF-κB [67, 68].

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The above mentioned Nox2 is a multisubunit protein, and its activation requires translocation of the cytosolic subunits—p47phox, p67phox, and Rac to the NOX/p22phox membrane complex [79]. Superoxide produced by Nox2 is implicated in influenza-mediated lung pathology [80]. Tang et al. published studies suggesting that endosomes are the

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**Figure 3:** Immune changes characteristic of severe COVID-19. Deregulation of cytokines and influx of inflammatory cells can lead to lung infiltration and critical symptoms; a “cytokine storm” may lead to a dramatic disruption of the homeodynamics of the whole organism and, consequently, even death of the patient [24].

- **Immunosuppression**
- **Lymphopenia**
- **Inefficient T cell immunity**
- **Inefficient B cell immunity**
- **Severe COVID-19**
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regulate cell-to-cell communication and danger sensing (11). Peroxynitrite is formed by the reaction of nitrite (NO$\cdot$) and superoxide, which is a product of the NADPH oxidase complex (2). Additionally, extracellular mitochondria, which are also ROS source, are able to provoke the immune response, proinflammatory cytokine expression (9). Based on the analogy and similarities between SARS-Cov-1 and SARS-Cov-2, it may be assumed that the nonstructural protein nsp10 causes mitochondrial impairment (10). Additionally, extracellular mitochondria, which are also ROS source, are able to provoke the immune response, regulate cell-to-cell communication and danger sensing (11). Peroxynitrite is formed by the reaction of nitrite (NO$\cdot$) and hydrogen peroxide (12), and it has been proved to damage lung tissue and thus playing an important role in lung destruction in viral infections.

**Figure 4: Action of SARS-CoV-2 in proposed mechanisms in the context of reactive oxygen and nitrogen species.** SARS-CoV-2 may affect the induction of reactive oxygen species by inducing both of their sources—NADPH oxidase and mitochondria. The increase in Nox2 activity in COVID-19 patients may be related to the activation of this enzyme by TLR7 (1), as is the case with other RNA viruses. Activated NADPH oxidase is responsible for the production of ROS (2), which are related to the activation of NF-$\kappa$B (3). The activity of this transcription factor results in the expression of proinflammatory cytokines like IL-6 and-1$\beta$ (4), which in turn can induce the production of mitochondrial ROS (5). On the other hand, ROS, if produced in excess, regardless of the source, may cause cell damage, enzymatic protein activity failure, virus mutation, and nucleic acid damage (6). NF-$\kappa$B, activated by ROS, has been proved to induce the expression of iNOS (7). The enzyme, responsible for the production of nitric oxide (8), has been shown to inhibit SARS-Cov virus replication (the coronavirus causing severe acute respiratory syndrome coronavirus, emerged in 2003), (9). Based on the analogy and similarity between SARS-Cov and SARS-Cov-2, it may be assumed that the nonstructural protein nsp10 causes mitochondrial impairment (10). Additionally, extracellular mitochondria, which are also ROS source, are able to provoke the immune response, regulate cell-to-cell communication and danger sensing (11). Peroxynitrite is formed by the reaction of nitrite (NO$\cdot$) and hydrogen peroxide (12), and it has been proved to damage lung tissue and thus playing an important role in lung destruction in viral infections.

Main site of ROS production under the influenza virus infection [46]. In addition, the authors indicated that ROS generation might be triggered by influenza virus in endosome via four different ways, one of which is TLR7 activation through the single-stranded RNA and protein kinase C activation. This results in phosphorylation of p47phox and by the assembly of the Nox2 oxidase complex at the endosomal membrane. The importance of Nox2 in influenza A infection was confirmed by literature, showing that in the absence of Nox2, influenza A virus results in lower viral burden and consequently results in significantly less lung injury, suggesting that ROS generated by Nox2 promotes rather than inhibits viral infection [80–83] (Figure 5.).

As mentioned earlier, apart from Nox2, also Nox1, Nox 4, and Duox2 might play a role in the ROS formation of viral infections [84–86]. Nox1 was shown to critically inhibit the early burst of proinflammatory cytokine expression in the lung and subsequently—oxidative stress followed by influenza A virus infection [85]. Nox1 oxidase has been proved to suppress early proinflammatory cytokine expression burst. Taking into consideration that ROS contribute to dysfunction and injury of the lung during influenza virus infection, this role of Nox1 seems surprising [85]. On the contrary, the study of Hofstetter et al. demonstrated that Nox1 presents activity promoting mortality during the peak of influenza infection, through restrain of the early phase of the adaptive immune response [87].

One of the key mediators of cytokines/chemokines induction is NF-$\kappa$B. The pathway of this transcription factor is directly activated by ROS and by certain proinflammatory cytokines, such as TNF-$\alpha$ and IL-1$\beta$. A wide spectrum of cytokines and chemokines may be expressed as a consequence of NF-$\kappa$B action, including IL-1$\beta$, IL-6, and IL-8, produced by most viruses (e.g., influenza virus, HBV, HIV (human immunodeficiency virus), EBV, SARS-CoV-2, and MERS); moreover, many respiratory viruses induce NF-$\kappa$B signaling both in vitro and in vivo in a ROS-dependent fashion [88–92]. During viral infections, NF-$\kappa$B binds to distinct sites of the iNOS promoter, causing iNOS enhanced expression. NO overproduction is predominantly caused by iNOS, which might be expressed, e.g., by inflammatory phagocytic cells [93–95]. Reactive nitrogen species play an important role in viral infections, in fact, some viruses, e.g., HCV, HRSV, or HIV, might upregulate the expression of iNOS [96–98]. On the other hand, IL-10, produced by many viruses (e.g., EBCV, HBV, HIV, SARS-CoV-2, and MERS), indirectly inhibits iNOS by inducing arginase, which reduces the availability of L-arginine, the substrate of iNOS [99, 100]. A previous study shows that HRSV directly upregulated iNOS in human type 2 alveolar epithelial cells, suggesting...
that the expression increase might be associated with interferon regulatory factor 1, instead of cytokines [101]. The regulatory factor mentioned has been involved in iNOS expression activation together with NF-κB and double-stranded RNA-activated protein kinase, as shown previously in influenza virus infection [102].

5. Molecular Mechanisms of ROS and RNS Generation in Response to Viruses

Reactive oxygen and nitrogen radicals are generated, among others, in viral infections. In conditions of infection, ROS and RNS production might be activated either by viral components or by cytokines, in response to the pathogen. Apart from the influenza virus, several viruses are associated with ROS generation. Epstein-Barr virus (EBV) may induce generation of ROS by NADPH oxidase in B lymphocytes through upregulating Nox2 [103]. In turn, Nox4 is activated by core protein of HCV, but this oxidase initiates mitochondrial ROS production, showing that this virus induces other proteins to trigger ROS generation [104, 105]. Mitochondrial ROS generation pathway is activated by viruses in various ways—the rubella virus produces N protein which induces the production of ROS by increasing the activity of mitochondrial respiratory chain [106]. Yuan et al. demonstrated that hepatitis B virus increases mitochondrial ROS generation resulting in the elevated IL-6 expression [107].

Apart from mitochondrial source, ROS are produced by enzymatic activity of NADPH oxidases in a highly regulated manner and play roles in both physiology and disease [108]. Out of seven NADPH oxidase homologs, four are implicated in ROS generation under viral infections: Nox1, Nox2, Nox4, and Duox2, but the primary source of inflammatory cell ROS is the Nox2 oxidase enzyme [36, 109, 110]. Although Nox2 is a phagocytic enzyme playing a role in killing bacteria and fungi, it is also known for contributing to virus-induced ROS production during viral infections, e.g., with IV (Influenza virus) [111], HRSV [91], HRV (human rhinovirus) [112], SARS-CoV-2 [37], and SeV (sendai virus) [91, 92].

In severe COVID-19, the major cytokines generated as part of immune response are IL-1β, IL-2, IL-6, and TNF. Also, IFN-ϒ seems to play an important role in antiviral response, although the data may suggest some defective interferon synthesis and release in severe patients infected with SARS-CoV-2. IL-1β is a well-known ROS and RNS generation activator [113]. Similarly, IL-2 stimulates RNS to generate nitrogen radicals [114]. Interleukin-6 activates human neutrophils and monocytes increasing the generation of free oxygen radicals [115]. Similarly, IFN-ϒ and TNF stimulate the generation of RNS in human [114]. On
the other hand, free oxygen radicals may increase IL-6 production and free nitrogen radicals are responsible, at least in part for its synthesis [116, 117]. Moreover, high levels of IL-6 are associated with the higher mortality rate in ICU-(intensive care unit-) treated COVID-19 patients [118, 119].

The effects of the antiviral potential of nitric oxide (NO) against SARS coronavirus have been described in Vero E6 cells and revealed, that NO donor, S-nitroso-N-acetylpenicillamine inhibited the replication cycle of SARS-CoV in a dose-dependent manner [67]. In patients with SARS, NO was associated with oxygenation amelioration. Moreover, endogenous but also exogenous NO inhibited SARS-CoV viral replication [120–122]. NO reacts with superoxide radicals yielding peroxynitrite, and both peroxynitrite and NO are toxic to mitochondria.

Apart from iNOS induction in response to viruses and viral components, interferon gamma has been reported as a major cytokine to induce iNOS and NO overproduction in the pathogenesis of virus infection [123, 124]. This cytokine is associated to Th1 cell response, as it is acknowledged that antiviral adaptive response is Th1 type [125]. Nevertheless, some viruses (such as influenza virus and HSV) might inhibit Th1 response through downregulation of interferons production. This type of immune response manipulation may prominently influence the consequence of the infection [126, 127]. Moreover, produced in excess during viral infection, reactive nitrogen species, are likely to influence mutagenesis in the virus [128].

6. The Possible Therapeutic Approach Related to Oxidative Stress Tampering in COVID-19

Several strategies for treating the SARS-CoV-2 infection are currently under consideration. Scientists and doctors have developed therapies based on the use of interferons, antibodies, inhibitors of viral/host proteases, and host-directed therapies. To date, no clinically effective antiviral therapy against SARS-CoV-2 has been confirmed; therefore, patients receive mainly supportive treatment which is often supplemented with various drug combinations. Many authors have documented elevated chemokines and interleukins levels in COVID-19 patients, so future efforts should focus inter alia on drugs that can be rapidly deployed and have immunomodulatory properties [129–132]. The use of interleukin 1 receptor antagonist in nine patients with moderate to severe COVID-19 pneumonia was effective in improving clinical and biological indices [133]. IL-1 receptor blocker reduced the need for invasive mechanical ventilation in the intensive care unit as well as mortality in patients with severe COVID-19 [134]. Shakoory et al. [135] in their randomized controlled trial confirmed that the inhibition of IL-1 receptor significantly decreased mortality in sepsis patients with features of macrophage activation syndrome. Patients who received IL-6 receptor antagonists had a marked reduction in pyrexia within days after treatment and a reduction in oxygen demand [136]. In the TESEO (the tocilizumab in patients with severe COVID-19 pneumonia) study, the use of a recombinant humanized anti-human IL-6 receptor monoclonal antibody (i.v. or s.c.) was associated with a reduced risk of mechanical ventilation and death [137]. Another IL-6 receptor blocker was effective only in critically ill COVID-19 patients requiring mechanical ventilation or high-flow oxygenation or requiring intensive care treatment [138]. Recent studies have highlighted the role of optimal nutritional status in boosting the immune system, focusing on the most important ingredients that reduce inflammation and oxidative stress parameters [139]. Interestingly, hydroxychloroquine (HCQ), the antimalarial drug, used to treat COVID-19, has been recently demonstrated to inhibit Nox2 activity through the ability to alkalize endosomes and therefore impedes antiphospholipid antibody activity (aPL) [35, 140]. The aPL, as a proinflammatory factor, has been proved to act via the pathway in which NADPH oxidase takes part [141]. There are many mechanisms for neutralizing free radicals, e. g., glutathione which is capable of affecting viral replication; the glutathione peroxidase/reductase enzyme system that allows reduced glutathione to bind to free radicals to produce oxidized glutathione, which is then regenerated to GSH; peroxiredoxin system that neutralizes lipid peroxidation; superoxide dismutase neutralizing superoxide anion; catalase eliminating hydrogen peroxide; carotenoids and polyphenols with scavenging effects; vitamins E and C; and finally, zinc and selenium, which have antioxidant properties as cofactors of antioxidant enzymes [142]. Providing substances that strengthen the antioxidant system will reduce the level of oxidative stress parameters during infection. Moreover, the use of molecular techniques to target antioxidants to organs of interest is an approach that might enhance the effectiveness of the antioxidant and circumvent toxicity [143].

Resveratrol is a wide studied antioxidant agent, which plays a role in mitochondria-derived ROS [144] but also down regulates the expression and activity of the NADPH oxidase [145]. In the case of MERS-CoV, resveratrol appeared to inhibit MERS-CoV infection. Moreover, the authors of a recently published study point out that as MERS-CoV infection leads to inflammatory cytokines production, resveratrol, via hindering NF-κB pathway, may reduce the inflammation [146–148]. They also found that the expression of the nucleocapsid (N), which is essential for MERS-CoV replication, was decreased after resveratrol treatment [61]. MERS-CoV next to SARS-CoV-1 and SARS-CoV-2 has been demonstrated to depend on TMPRSS2 (transmembrane serine protease 2) which plays an important role during the virus entry to the cell. Presumably, TMPRSS2 might regulate mitochondrial function [149–151].

Recently, many others antioxidants have been tested for the highly conserved SARS-CoV-2 main protease using molecular docking. Of all the compounds that were investigated, the lowest predicted IC50 value was observed for taxifolin. Moreover, taxifolin along with other compounds such as eriodictyol did not show any toxicity against the toxicity parameters used in the experiment [152]. This flavonoid was found to be a powerful antiradical and antioxidant activities in different in vitro bioassays when compared with standard antioxidant compounds [153]. This compound inhibits NF-
κB pathway and downregulates STAT3 of the JAK/STAT pathway [154]. Thus, taxifolin could be a potential inhibitor against Mpro but further in vivo studies are needed [155]. Another analyzes also point to the natural compounds, taxifolin and rhamnetin, as potential inhibitors of Mpro [156]. Rutin (a polyphenolic flavonoid) may able a potential inhibitor as it is able to form several hydrogen bonds and σ–π stacking interactions with various amino acids of Mpro in anchoring and blocking the substrate into the active pocket of the catalytic center [157]. In vivo and in silico studies have demonstrated that silymarin and its derivative silybin (a flavonoid from the group of flavonolignans) are able to inhibit SARS-CoV-2 main protease [158]. Another authors found luteolin to be effective in blocking the S2 protein of SARS-CoV [159]. It is already known that the SARS-CoV and SARS-CoV-2 S proteins share about 76% amino acid similarity [142]. Several other herbal compounds like quercetin, naringenin, kaempferol, allicin, demethoxycurcumin, catechin, apigenin-7-glucoside, oleuropein, curcumin, zingerol or gingerol have been also investigated [57].

The approach of using antioxidants both to reduce viral replication and to reduce viral-induced oxidative damage may prove to be particularly useful for those viruses, which have thus far eluded attempts at antiviral therapies.

7. Conclusion

In conclusion, the literature demonstrates an important role of reactive oxygen and nitrogen species during SARS-CoV-2 infections, associated with a weakened antioxidant defense. Nevertheless, it must be noted that some of the understanding, background, and supporting data presented in the current review come from the experience with other human coronaviruses or viruses, such as RSV/HBV/HCV, and may not necessarily be known to be appropriate with respect to SARS-CoV-2.

The oxidative stress mechanism coupled with innate immunity activates transcription factors, such as NF-κB, which results in an exacerbated proinflammatory host response. The importance of ROS and RNS is also connected with the fact that this virus is especially dangerous for the elderly, and their deteriorated antioxidative/nitrosative defense system affected by increased reactive oxygen and nitrogen species. Moreover, only treatments diminishing the ROS and RNS production such as dexamethasone and tocilizumab deliver substantial benefits to severe COVID-19 patients. Therefore, there is a strong need to deeply investigate this issue, as it would be of interest to use the antioxidants as potential therapeutic tools.

Abbreviations

ACE2: Angiotensin-converting enzyme 2
ADCC: Antibody dependent cellular cytotoxicity
aPL: Antiphospholipid antibodies
ARDS: Acute respiratory distress syndrome
CTL: Cytotoxic T leukocytes
DIC: Disseminated intravascular coagulation
EBV: Epstein-Barr virus
HBV: Hepatitis B virus
HCV: Hepatitis C virus
HCQ: Hydroxychloroquine
HIV: Human immunodeficiency virus
HRSV: Human respiratory syncytial virus
HRV: Human rhinovirus
ICU: Intensive care unit
IFN: Interferon
IL: Interleukin
iNOS: Inducible nitric oxide synthase
IRFs: Interferon regulatory factors
ISGs: IFN-stimulated genes
IV: Influenza virus
MCP-1: Monocyte chemoattractant protein-1
MERS: Middle East respiratory syndrome coronavirus
MHC: Major histocompatibility complex
MIP1αβ: Macrophage inflammatory proteins-1αβ
MyD-88: Myeloid differentiation primary response-88
NADPH: Nicotinamide adenine dinucleotide phosphate hydrogen
NET: Neutrophil extracellular traps
NF-κB: Nuclear factor kappa-light-chain-enhancer of activated B cells
NHC: Natural helper cells
NK: Natural killers
NKT: Natural killer T cells
NOX2: NADPH oxidase 2
PAMPs: Pathogen associated molecular patterns
PRR: Pattern recognition receptors
SARS: Severe acute respiratory syndrome
SeV: Sendai virus
TLR: Toll-like receptors
TMPRSS2: Transmembrane serine protease 2
TNF-α: Tumor necrosis factor alpha
TRAF6: TNF receptor associated factor 6
TRIF: TIR-domain-containing adapter-inducing interferon-β.

Data Availability

The data supporting this review article are from previously reported studies and datasets, which have been cited. The processed data are available from the corresponding authors of upon request.

Conflicts of Interest

The authors declare no conflict of interest.

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References

[1] 2020, https://covid19.who.int/.
[2] vol. 27, p. 12, 2021, https://www.who.int/news-room/fact-sheets/detail/influenza-(seasonal).
[3] J. M. Mostaza, F. García-Iglesias, T. Gonzalez-Alegre et al., “Clinical course and prognostic factors of COVID-19 infection in an elderly hospitalized population,” Archives of Gerontology and Geriatrics, vol. 91, article 104204, 2020.

[4] K. Hushmandi, S. Bokaie, M. Hashemi et al., “A review of medications used to control and improve the signs and symptoms of COVID-19 patients,” European Journal of Pharmacology, vol. 887, article 173568, 2020.

[5] C. Ryan, A. Minc, J. Caceres et al., “Predicting severe outcomes in COVID-19 related illness using only patient demographics, comorbidities and symptoms,” The American Journal of Emergency Medicine, vol. 45, pp. 378–384, 2020.

[6] L. F. Westblade, G. Brar, L. C. Pinheiro et al., “SARS-CoV-2 viral load predicts mortality in patients with and without cancer who are hospitalized with COVID-19,” Cancer Cell, vol. 38, no. 5, article e662, pp. 661–671.e2, 2020.

[7] E. Pujadas, F. Chaudhry, R. McBride et al., “SARS-CoV-2 viral load predicts COVID-19 mortality,” The Lancet Respiratory Medicine, vol. 8, no. 9, p. e70, 2020.

[8] A. K. J. Mandal, V. Baktash, T. Hosack, K. Van den Abbee, and C. G. Missouris, “Vitamin D status may indeed be a prognostic factor for morbidity and mortality in patients with COVID-19,” Journal of Medical Virology, vol. 93, no. 3, p. 1225, 2021.

[9] C. E. Hastie, J. P. Pell, and N. Sattar, “Vitamin D and COVID-19 infection and mortality in UK biobank,” European Journal of Nutrition, vol. 60, pp. 545–548, 2021.

[10] A. L. Nyman, C. A. Spears, V. Churchill et al., “Associations between COVID-19 risk perceptions and smoking and quitting behavior among U.S. adults,” Addictive Behaviors Reports, vol. 14, article 100394, 2021.

[11] S. F. Maloney, M. Combs, R. L. Scholtes et al., “Impacts of COVID-19 on cigarette use, smoking behaviors, and tobacco purchasing behaviors,” Drug and Alcohol Dependence, vol. 229, Part B, article 109144, 2021.

[12] L. A. Wilkinson, K. A. Mengenhagen, M. T. Carter et al., “Smoking status related to Covid-19 mortality and disease severity in a veteran population,” Respiratory Medicine, vol. 190, article 106668, 2021.

[13] K. Hu, L. Lin, Y. Liang et al., “COVID-19: risk factors for severe cases of the delta variant,” Aging (Albany NY), vol. 13, no. 20, pp. 23459–23470, 2021.

[14] A. Akbar Asadi-Pooya, A. Akbari, A. Emami et al., “Risk factors associated with long COVID syndrome: a retrospective study,” Iranian Journal of Medical Sciences, vol. 46, no. 6, pp. 428–436, 2021.

[15] H. Holt, M. Talaei, M. Greenig et al., “Risk factors for developing COVID-19: a population-based longitudinal study (COVIDENCE UK),” Thorax, pp. 1–13, 2021.

[16] M. Bonato, U. Semenzato, M. Tine et al., “Risk factors for development and severity of COVID-19 in COPD patients,” Frontiers in Medicine, vol. 8, article 714570, 2021.

[17] S. Banerjee, S. Ghosh, A. Mandal, N. Ghosh, and P. C. Sil, “ROS-associated immune response and metabolism: a mechanistic approach with implication of various diseases,” Archives of Toxicology, vol. 94, pp. 2293–2317, 2020.

[18] M. Catanzaro, F. Fagiani, M. Racchi, E. Corsini, S. Govoni, and C. Lanni, “Immune response in COVID-19: addressing a pharmacological challenge by targeting pathways triggered by SARS-CoV-2,” Signal Transduction and Targeted Therapy, vol. 5, p. 84, 2020.

[19] J. A. Otter, C. Donskey, S. Yezli, S. Douthwaite, S. D. Goldenberg, and D. J. Weber, “Transmission of SARS and MERS coronaviruses and influenza virus in healthcare settings: the possible role of dry surface contamination,” The Journal of Hospital Infection, vol. 92, no. 3, pp. 235–250, 2016.

[20] Y. Yin and R. G. Wunderink, “MERS, SARS and other coronaviruses as causes of pneumonia,” Respirology, vol. 23, pp. 130–137, 2018.

[21] E. de Wit, N. van Doremalen, D. Falzarano, and V. J. Munster, “SARS and MERS: recent insights into emerging coronaviruses,” Nature Reviews. Microbiology, vol. 14, pp. 523–534, 2016.

[22] K. T. Chow, M. Gale Jr., and Y. M. Loo, “RIG-I and other RNA sensors in antiviral immunity,” Annual Review of Immunology, vol. 36, pp. 667–694, 2018.

[23] A. Iwasaki, E. F. Foxman, and R. D. Molony, “Early local immune defences in the respiratory tract,” Nature Reviews. Immunology, vol. 17, pp. 7–20, 2017.

[24] K. Tsuchiya, “Inflammasome-associated cell death: pyroptosis, apoptosis, and physiological implications,” Microbiology and Immunology, vol. 64, no. 4, pp. 252–269, 2020.

[25] C. Zhao and W. Zhao, “NLRP3 inflammasome-a key player in antiviral responses,” Frontiers in Immunology, vol. 11, p. 211, 2020.

[26] G. Schonrich, M. J. Raftery, and V. Samlid, “Devilishly radical NETwork in COVID-19: oxidative stress, neutrophil extracellular traps (NETs), and T cell suppression,” Advances in Biological Regulation, vol. 77, article 100741, 2020.

[27] P. V’Kovski, A. Krtatzel, S. Steiner, H. Stalder, and V. Thiel, “Coronavirus biology and replication: implications for SARS-CoV-2,” Nature Reviews. Microbiology, vol. 19, pp. 155–170, 2020.

[28] J. Mu, Y. Fang, Q. Yang et al., “SARS-CoV-2 N protein antagonizes type I interferon signaling by suppressing phosphorylation and nuclear translocation of STAT1 and STAT2,” Cell Discovery, vol. 6, pp. 1–4, 2020.

[29] G. Morris, C. C. Bortolasci, B. K. Puri et al., “The pathophysiology of SARS-CoV-2: a suggested model and therapeutic approach,” Life Sciences, vol. 258, article 118166, 2020.

[30] A. L. Valadao, R. S. Aguiar, and L. B. de Arruda, “Activation of toll-like receptor signaling pathways leading to nitric oxide-mediated antiviral responses,” Archives of Virology, vol. 161, no. 8, pp. 2075–2086, 2016.

[31] D. Poud, “Nitric oxide and the common cold,” Current Opinion in Allergy and Clinical Immunology, vol. 5, no. 1, pp. 37–42, 2005.

[32] R. Koetzler, R. S. Zaheer, S. Wiehler, N. S. Holden, M. A. Giembycz, and D. Proud, “Nitric oxide inhibits human rhinovirus-induced transcriptional activation of CXCL10 in airway epithelial cells,” The Journal of Allergy and Clinical Immunology, vol. 123, no. 1, pp. 201–208, 2009.

[33] A. C. Kalil and P. G. Thomas, “Influenza virus-related critical illness: pathophysiology and epidemiology,” Critical Care, vol. 23, no. 1, pp. 1–7, 2019.

[34] J. J. DiNicolantonio and M. McCarty, “Thrombotic complications of COVID-19 may reflect an upregulation of endothelial tissue factor expression that is contingent on activation of endosomal NADPH oxidase,” Open Heart, vol. 7, no. 1, article e001337, 2020.
M. Mahmudpour, J. Roozbeh, M. Keshavarz, S. Farrokhi, and F. Violi, A. Oliva, R. Cangemi et al., “Cytokine storm in COVID-19,” *Viruses*, vol. 12, no. 12, pp. 647–657, 2020.

J. Checa and J. M. Aran, “Reactive oxygen species: drivers of physiological and pathological processes,” *Journal of Inflammation Research*, vol. Volume 13, pp. 1057–1073, 2020.

J. P. Kehrer and L. O. Klotz, “Free radicals and related reactive species as mediators of tissue injury and disease: implications for health,” *Critical Reviews in Toxicology*, vol. 45, pp. 765–798, 2015.

M. Valko, D. Leibfritz, J. Moncol, M. T. Cronin, M. Mazur, and J. Telser, “Free radicals and antioxidants in normal physiological functions and human disease,” *The International Journal of Biochemistry & Cell Biology*, vol. 39, no. 1, pp. 44–84, 2007.

A. A. Alfadda and R. M. Sallam, “Reactive oxygen species in health and disease,” *Journal of Biomedicine & Biotechnology*, vol. 2012, Article ID 936486, 14 pages, 2012.

A. K. Azkur, M. Akdis, D. Azkur et al., “Immune response to SARS-CoV-2 and mechanisms of immunopathological changes in COVID-19,” *Allergy*, vol. 75, no. 7, pp. 1564–1581, 2020.

S. Gupta and M. J. Kaplan, “The role of neutrophils and NETosis in autoimmune and renal diseases,” *Nature Reviews. Nephrology*, vol. 12, no. 7, pp. 402–413, 2016.

F. Gong, Y. Dai, T. Zheng et al., “Peripheral CD4+ T cell subsets and antibody response in COVID-19 convalescent individuals,” *The Journal of Clinical Investigation*, vol. 130, no. 12, pp. 6588–6599, 2020.

C. Pallotto, L. R. Suardi, S. Esperti et al., “Increased CD4/CD8 ratio as a risk factor for critical illness in coronavirus disease 2019 (COVID-19): a retrospective multicentre study,” *Infectious Diseases*, vol. 52, no. 9, pp. 675–677, 2020.

Y. Tang, J. Liu, D. Zhang, Z. Xu, J. Ji, and C. Wen, “Cytokine storm in COVID-19: the current evidence and treatment strategies,” *Frontiers in Immunology*, vol. 11, p. 1708, 2020.

M. Mahmudpour, J. Rozobeh, M. Keshavarz, S. Farrokhi, and I. Nabipour, “COVID-19 cytokine storm: the anger of inflammation,” *Cytokine*, vol. 133, article 155151, 2020.

M. Xu, L. Wang, M. Wang et al., “Mitochondrial ROS and NLRP3 inflammasome in acute ozone-induced murine model of airway inflammation and bronchial hyperresponsiveness,” *Free Radical Research*, vol. 53, pp. 780–790, 2019.

R. Cardin, G. Saccoccio, F. Masutti, S. Bellentani, F. Farinati, and C. Tiribelli, “DNA oxidative damage in leukocytes correlates with the severity of HCV-related liver disease: validation in an open population study,” *Journal of Hepatology*, vol. 34, no. 4, pp. 587–592, 2001.

C. Bolukbas, F. F. Bolukbas, M. Horoz, M. Aslan, H. Celik, and O. Erel, “Increased oxidative stress associated with the severity of the liver disease in various forms of hepatitis B virus infection,” *BMC Infectious Diseases*, vol. 5, no. 1, pp. 1–7, 2005.

G. Moreno-Solis, M. J. Dela Torre-Aguilar, J. Torres-Borrego et al., “Oxidative stress and inflammatory plasma biomarkers in respiratory syncytial virus bronchiolitis,” *The Clinical Respiratory Journal*, vol. 11, no. 6, pp. 839–846, 2017.

L. Delgado-Roche and F. Mesta, “Oxidative stress as key player in severe acute respiratory syndrome coronavirus (SARS-CoV) infection,” *Archives of Medical Research*, vol. 51, no. 5, pp. 384–387, 2020.

Y. Imai, K. Kuba, G. G. Neely et al., “Identification of oxidative stress and Toll-like receptor 4 signaling as a key pathway of acute lung injury,” *Cell*, vol. 133, no. 2, pp. 235–249, 2008.

C. W. Lin, K. H. Lin, T. H. Hsieh, S. Y. Shiu, and J. Y. Li, “Severe acute respiratory syndrome coronavirus 3C-like protease-induced apoptosis,” *FEBS Immunology and Medical Microbiology*, vol. 46, no. 3, pp. 375–380, 2006.

Y. Liu, G. Hu, Y. Wang et al., “Functional and genetic analysis of viral receptor ACE2 orthologs reveals a broad potential host range of SARS-CoV-2,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 118, no. 12, 2021.

M. Soto, G. diZerega, and K. E. Rodgers, “Countermeasure and therapeutic: a(1–7) to treat acute respiratory distress syndrome due to COVID-19 infection,” *Journal of the Renin-Angiotensin-Aldosterone System*, vol. 21, no. 4, 2020.

M. Hoffmann, H. Kleine-Weber, S. Schroeder et al., “SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor,” *Cell*, vol. 181, no. 2, pp. 271–280, 2020.

X. Zou, K. Chen, J. Zou, P. Han, J. Hao, and Z. Han, “Single-cell RNA-seq data analysis on the receptor ACE2 expression reveals the potential risk of different human organs vulnerable to 2019–nCoV infection,” *Frontiers in Medicine*, vol. 14, no. 2, pp. 185–192, 2020.

C. Sindona, G. Schepici, V. Contestabile, P. Bramanti, and E. Mazzon, “NOX2 Activation in COVID-19: Possible Implications for Neurodegenerative Diseases,” vol. 57, no. 6, p. 604, 2021.

A. Kouountouri, E. Korakas, I. Ikonomidou et al., “Type 1 diabetes mellitus in the SARS-CoV-2 pandemic: oxidative stress as a major pathophysiological mechanism linked to adverse clinical outcomes,” *Antioxidants*, vol. 10, no. 5, p. 752, 2021.

G. Li, X. He, L. Zhang et al., “Assessing ACE2 expression pattern in lung tissues in the pathogenesis of COVID-19,” *Journal of Autoimmunity*, vol. 112, article 102463, 2020.

F. Lovren, Y. Pan, A. Quan et al., “Angiotensin converting enzyme-2 confers endothelial protection and attenuates atherosclerosis,” *American Journal of Physiology. Heart and Circulatory Physiology*, vol. 295, no. 4, pp. H1377–H1384, 2008.

F. Ferretta, M. G. Matera, M. Cazzola, and A. Bianco, “Severe respiratory SARS-CoV2 infection: does ACE2 receptor matter?,” *Respiratory Medicine*, vol. 168, article 105996, 2020.

A. R. Bourgonje, A. E. Abdulle, W. Timens et al., “Angiotensin-converting enzyme 2 (ACE2), SARS-CoV-2 and the pathophysiology of coronavirus disease 2019 (COVID-19),” *The Journal of Pathology*, vol. 251, no. 3, pp. 228–248, 2020.

L. S. Y. Wong, E. X. L. Loo, A. Y. H. Kang, H. X. Lau, P. A. Tambyah, and E. H. Tham, “Age-related differences in immunological responses to SARS-CoV-2,” *The Journal of Allergy and Clinical Immunology. In Practice*, vol. 8, no. 10, pp. 3251–3258, 2020.
[67] H. Y. Chung, B. Sung, K. J. Jung, Y. Zou, and B. P. Yu, “The molecular inflammatory process in aging,” *Antioxidants & Redox Signaling*, vol. 8, pp. 572–581, 2006.

[68] T. Lawrence, “The nuclear factor NF-kappaB pathway in inflammation,” *Cold Spring Harbor Perspectives in Biology*, vol. 1, article a001651, 2009.

[69] Q. Li, L. Wang, C. Dong et al., “The interaction of the SARS coronavirus non-structural protein 10 with the cellular oxido-reductase system causes an extensive cytotoxic effect,” *Journal of Clinical Virology*, vol. 34, pp. 133–139, 2005.

[70] T. Viswanathan, S. Arya, S. H. Chan et al., “Structural basis of RNA cap modification by SARS-CoV-2,” *Nature Communications*, vol. 11, no. 1, pp. 1–7, 2020.

[71] Z. S. Ulhaq and G. V. Soraya, “Interleukin-6 as a potential biomarker of COVID-19 progression,” *Medecine et Maladies Infectieuses*, vol. 50, no. 4, pp. 382–383, 2020.

[72] D. M. Del Valle, S. Kim-Schulze, H. H. Huang et al., “An inflammatory cytokine signature predicts COVID-19 severity and survival,” *Nature Medicine*, vol. 26, pp. 1636–1643, 2020.

[73] J. Saleh, C. Peyssonnaux, K. K. Singh, and M. Edeas, “Mitochondria and microbiota dysfunction in COVID-19 pathogenesis,” *Mitochondrion*, vol. 54, pp. 1–7, 2020.

[74] D. Giannis, L. A. Zingas, and P. Gianni, “Coagulation disorders in coronavirus infected patients: COVID-19, SARS-CoV-1, MERS-CoV and lessons from the past,” *Journal of Clinical Virology*, vol. 127, article 104362, 2020.

[75] T. J. Oxley, J. Mocco, S. Majidi et al., “Large-vessel stroke as a presenting feature of Covid-19 in the young,” *The New England Journal of Medicine*, vol. 382, p. e60, 2020.

[76] X. Jiang, Z. Zhang, C. Wang et al., “Bimodal effects of D614G mutation on the spike glycoprotein of SARS-CoV-2 enhance protein processing, membrane fusion, and viral infectivity,” *Signal Transduction and Targeted Therapy*, vol. 5, no. 1, p. 268, 2020.

[77] S. Milliotis, B. Nicolalde, M. Ortega, J. Yopez, and A. Caicedo, “Forms of extracellular mitochondria and their impact in health,” *Mitochondrion*, vol. 48, pp. 16–30, 2019.

[78] F. Zhou, T. Yu, R. Du et al., “Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study,” *Lancet*, vol. 395, no. 10229, pp. 1054–1062, 2020.

[79] J. Wiecezinska, M. Sokolowska, and R. Pawliczak, “NOX modifiers—just a step away from application in the therapy of airway inflammation?,” *Antioxidants & Redox Signaling*, vol. 23, pp. 428–445, 2015.

[80] E. E. To, B. R. Broughton, K. S. Hendricks, R. Vlahos, and S. Selemidis, “Influenza A virus and TLR7 activation potentiate NOX2 oxidase-dependent ROS production in macrophages,” *Free Radical Research*, vol. 48, pp. 940–947, 2014.

[81] R. Vlahos, J. Stambas, S. Bozinovski, B. R. Broughton, G. R. Drummond, and S. Selemidis, “Inhibition of Nox2 oxidase activity ameliorates influenza a virus-induced lung inflammation,” *PLoS Pathogens*, vol. 7, no. 2, article e1001271, 2011.

[82] R. Vlahos, J. Stambas, and S. Selemidis, “Suppressing production of reactive oxygen species (ROS) for influenza a virus therapy,” *Trends in Pharmacological Sciences*, vol. 33, no. 1, pp. 3–8, 2012.

[83] R. J. Snellgrove, L. Edwards, A. J. Rae, and T. Hussell, “An absence of reactive oxygen species improves the resolution of lung influenza infection,” *European Journal of Immunology*, vol. 36, no. 6, pp. 1364–1373, 2006.

[84] S. Carnesecchi, J. C. Pache, and C. Barazzzone-Argiroff, “NOX enzymes: potential target for the treatment of acute lung injury,” *Cellular and Molecular Life Sciences*, vol. 69, pp. 2373–2385, 2012.

[85] S. Selemidis, H. J. Seow, B. R. Broughton et al., “Nox1 oxidase suppresses influenza a virus-induced lung inflammation and oxidative stress,” *PLoS One*, vol. 8, no. 4, article e60792, 2013.

[86] H. J. Kim, C. H. Kim, M. J. Kim et al., “The induction of pattern-recognition receptor expression against influenza a virus through Duox2-derived reactive oxygen species in nasal mucosa,” *American Journal of Respiratory Cell and Molecular Biology*, vol. 53, pp. 525–535, 2015.

[87] A. R. Hofstetter, J. A. De La Cruz, W. Cao et al., “NADPH oxidase 1 is associated with altered host survival and T cell phenotypes after influenza a virus infection in mice,” *PLoS One*, vol. 11, article e0149864, 2016.

[88] E. Flory, M. Kunz, C. Scheller et al., “Influenza virus-induced NF-kappaB-dependent gene expression is mediated by over-expression of viral proteins and involves oxidative radicals and activation of Ikappab kinase,” *The Journal of Biological Chemistry*, vol. 275, pp. 8307–8314, 2000.

[89] N. W. Bartlett, L. Slater, N. Glanville et al., “Defining critical roles for NF-kB p65 and type I interferon in innate immunity to rhinovirus,” *EMBO Molecular Medicine*, vol. 4, no. 12, pp. 1244–1260, 2012.

[90] S. F. Dosch, S. D. Mahajan, and A. R. Collins, “SARS coronavirus spike protein-induced innate immune response occurs via activation of the NF-kB pathway in human monocyte macrophages in vitro,” *Viruses Research*, vol. 142, no. 1-2, pp. 19–27, 2009.

[91] K. Fink, A. Duval, A. Martel, A. Soucy-Faulkner, and N. Grandvaux, “Dual role of NOX2 in respiratory syncytial virus- and Sendai virus-induced activation of NF-kB in airway epithelial cells,” *Journal of Immunology*, vol. 180, no. 10, pp. 6911–6922, 2008.

[92] O. A. Khomich, S. N. Kochetkov, B. Bartosch, and A. V. Ivanov, “Redox biology of respiratory viral infections,” *Viruses*, vol. 10, no. 8, p. 392, 2018.

[93] T. Akaie, “Role of free radicals in viral pathogenesis and mutation,” *Reviews in Medical Virology*, vol. 11, pp. 87–101, 2001.

[94] C. Nathan and M. U. Shiloh, “Reactive oxygen and nitrogen intermediates in the relationship between mammalian hosts and microbial pathogens,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 97, no. 16, pp. 8841–8848, 2000.

[95] S. Moncada and A. Higgs, “The L-arginine-nitric oxide pathway,” *The New England Journal of Medicine*, vol. 329, pp. 2002–2012, 1993.

[96] K. Machida, K. T. Cheng, V. M. Sung, K. J. Lee, A. M. Levine, and M. M. Lai, “Hepatitis C virus infection activates the immunologic (type II) isoform of nitric oxide synthase and thereby enhances DNA damage and mutations of cellular genes,” *Journal of Virology*, vol. 78, no. 16, pp. 8835–8843, 2004.

[97] E. Polazzi, G. Levi, and L. Minghetti, “Human immunodeficiency virus type 1 Tat protein stimulates inducible nitric oxide synthase expression and nitric oxide production in..."
Oxidative Medicine and Cellular Longevity

microglial cultures,” *Journal of Neuropathology and Experimental Neurology*, vol. 58, no. 8, pp. 825–831, 1999.

[98] V. Mgbemena, J. Segovia, T. H. Chang, and S. Bose, “KLF6 and iNOS regulates apoptosis during respiratory syncytial virus infection,” *Cellular Immunology*, vol. 283, no. 1-2, pp. 1–7, 2013.

[99] C. Santiago-Olivares, E. Rivera-Toledo, and B. Gomez, “Nitríc oxide production is downregulated during respiratory syncytial virus persistence by constitutive expression of arginase 1,” *Archives of Virology*, vol. 164, pp. 2231–2241, 2019.

[100] C. G. Molteni, N. Principi, and S. Esposito, “Reactive oxygen and nitrogen species during viral infections,” *Free Radical Research*, vol. 48, pp. 1163–1169, 2014.

[101] Y. J. Kao, P. A. Piedra, G. L. Larsen, and G. N. Colasurdo, “Induction and regulation of nitric oxide synthase in airway epithelial cells by respiratory syncytial virus,” *American Journal of Respiratory and Critical Care Medicine*, vol. 163, pp. 532–539, 2001.

[102] K. Uetani, S. D. Der, M. Zamanian-Daryouh et al., “Central role of double-stranded RNA-activated protein kinase in microbial induction of nitric oxide synthase,” *Journal of Immunology*, vol. 165, pp. 988–996, 2000.

[103] S. Lassoued, R. Ben Ameur, W. Ayadi, B. Gargouri, R. Ben Mansour, and H. Attia, “Epstein-Barr virus induces an oxidative stress during the early stages of infection in B lymphocytes, epithelial, and lymphoblastoid cell lines,” *Molecular and Cellular Biochemistry*, vol. 313, pp. 179–186, 2008.

[104] Y. Li, D. F. Boehning, T. Qian, V. L. Popov, and S. A. Weiman, “Hepatitis C virus core protein increases mitochondrial ROS production by stimulation of Ca²⁺ uniporter activity,” *The FASEB Journal*, vol. 21, pp. 2474–2485, 2007.

[105] N. Dionisio, M. V. Garcia-Mediavilla, S. Sanchez-Campos et al., “Hepatitis C virus NS5A and core proteins induce oxidative stress-mediated calcium signalling alterations in hepatocytes,” *Journal of Hepatology*, vol. 50, no. 5, pp. 872–882, 2009.

[106] C. Claus, K. Schonefeld, D. Hubner, S. Chey, U. Reibetanz, and U. G. Liebert, “Activity increase in respiratory chain complexes by rubella virus with marginal induction of oxidative stress,” *Journal of Virology*, vol. 87, no. 5, pp. 8481–8492, 2013.

[107] K. Yuan, Y. Lei, H. N. Chen et al., “HBV-induced ROS accumulation promotes hepatocarcinogenesis through snail-mediated epigenetic silencing of SOCS3,” *Cell Death and Differentiation*, vol. 23, no. 4, pp. 616–627, 2016.

[108] K. Bernard, L. Hecker, T. R. Luckhardt, G. Cheng, and V. J. Thannickal, “NADPH oxidases in lung health and disease,” *Antioxidants & Redox Signaling*, vol. 20, pp. 2838–2853, 2014.

[109] R. Vlahos and S. Selemidis, “NADPH oxidases as novel pharmacologic targets against influenza a virus infection,” *Molecular Pharmacology*, vol. 86, no. 6, pp. 747–759, 2014.

[110] H. E. Boudreau, S. U. Emerson, A. Korzeniowska, M. A. Jendrysik, and T. L. Leto, “Hepatitis C virus (HCV) proteins induce NADPH oxidase 4 expression in a transforming growth factor beta-dependent manner: a new contributor to HCV-induced oxidative stress,” *Journal of Virology*, vol. 83, no. 24, pp. 12934–12946, 2009.

[111] G. Laghali and M. D. Tate, “Targeting endosomal NOX2 as a potential therapy to limit influenza a virus infection,” *Respiratory Research*, vol. 24, pp. 935-936, 2019.

[112] P. Kauf, M. C. Biagioli, I. Singh, and R. B. Turner, “Rhinovirus-induced oxidative stress and interleukin-8 elaboration involves p47-phox but is independent of attachment to intracellular adhesion molecule-1 and viral replication,” *The Journal of Infectious Diseases*, vol. 181, no. 6, pp. 1885–1890, 2000.

[113] B. Fubini and A. Hubbard, “Reactive oxygen species (ROS) and reactive nitrogen species (RNS) generation by silica in inflammation and fibrosis,” *Free Radical Biology & Medicine*, vol. 34, pp. 1507–1516, 2003.

[114] J. B. Hibbs Jr., C. Westenfelder, R. Taintor et al., “Evidence for cytokine-inducible nitric oxide synthesis from L-arginine in patients receiving interleukin-2 therapy,” *The Journal of Clinical Investigation*, vol. 89, pp. 867–877, 1992.

[115] A. Kharazmi, H. Nielsen, C. Rechnitzer, and K. Bendtzen, “Interleukin 6 primes human neutrophil and monocyte oxidative burst response,” *Immunology Letters*, vol. 21, pp. 177–184, 1989.

[116] A. C. Makris, Y. Sotzios, Z. Zhou et al., “Nitric oxide stimulates interleukin-6 production in skeletal myotubes,” *Journal of Interferon & Cytokine Research*, vol. 30, pp. 321–327, 2010.

[117] R. A. Willis, A. K. Nussler, K. M. Fries, D. A. Geller, and R. P. Phipps, “Induction of nitric oxide synthase in subsets of murine pulmonary fibroblasts: effect on fibroblast interleukin-6 production,” *Clinical Immunology and Immunopathology*, vol. 71, no. 2, pp. 231–239, 1994.

[118] D. R. J. Arachchilage and M. Laffan, “Abnormal coagulation parameters are associated with poor prognosis in patients with novel coronavirus pneumonia,” *Journal of Thrombosis and Haemostasis*, vol. 18, no. 5, pp. 1233–1234, 2020.

[119] N. Tang, D. Li, X. Wang, and Z. Sun, “Abnormal coagulation parameters are associated with poor prognosis in patients with novel coronavirus pneumonia,” *Journal of Thrombosis and Haemostasis*, vol. 18, no. 4, pp. 844–847, 2020.

[120] S. Akerstrom, M. Mousavi-Jazi, J. Klingstrom, M. Lejon, A. Lundkvist, and A. Mirazimi, “Nitríc oxide inhibits the replication cycle of severe acute respiratory syndrome coronavirus,” *Journal of Virology*, vol. 79, pp. 1966–1969, 2005.

[121] R. A. Alvarez, L. Berra, and M. T. Gladwin, “Home nitric oxide therapy for COVID-19,” *American Journal of Respiratory and Critical Care Medicine*, vol. 202, no. 1, pp. 16–20, 2020.

[122] L. Chen, P. Liu, H. Gao et al., “Inhalation of nitric oxide in the treatment of severe acute respiratory syndrome: a rescue trial in Beijing,” *Clinical Infectious Diseases*, vol. 39, no. 10, pp. 1531–1535, 2004.

[123] A. M. Chavez, M. J. Morin, N. Unno, M. P. Fink, and R. A. Hodin, “Acquired interferon gamma responsiveness during Caco-2 cell differentiation: effects on iNOS gene expression,” *Gut*, vol. 44, pp. 659–665, 1999.

[124] N. Jarasch, U. Martin, E. Kamphausen, R. Zell, P. Wutzler, and A. Henke, “Nitric oxide inhibits the replication cycle of severe acute respiratory syndrome coronavirus,” *Journal of Virology*, vol. 79, pp. 1966–1969, 2005.

[125] T. C. Lewis, E. E. Metitiri, G. B. Mentz et al., “Epstein-Barr virus induces an oxidative burst response,” *Gut*, vol. 759, 2013.

[126] M. Mibayashi, L. Martinez-Sobrido, Y. M. Loo, W. B. Cardenas, M. Gale Jr., and A. Garcia-Sastre, “Inhibition of retinoic acid-inducible gene I-mediated induction of antiviral cytokine-inducible nitric oxide synthesis from L-arginine in patients receiving interleukin-2 therapy,” *The Journal of Clinical Investigation*, vol. 89, pp. 867–877, 1992.
beta interferon by the NS1 protein of influenza A virus,” *Journal of Virology*, vol. 81, no. 2, pp. 514–524, 2007.

[127] J. Ban, N. R. Lee, N. J. Lee, J. K. Lee, F. S. Quan, and K. S. Inn, “Human respiratory syncytial virus NS1 targets TRIM25 to suppress RIG-I ubiquitination and subsequent RIG-I-mediated antiviral signaling,” *Viruses*, vol. 10, no. 12, p. 716, 2018.

[128] T. Akaike, S. Fujii, A. Kato et al., “Viral mutation accelerated by nitric oxide production during infection in vivo,” *The FASEB Journal*, vol. 14, no. 10, pp. 1447–1454, 2000.

[129] C. Huang, Y. Wang, X. Li et al., “Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China,” *The Lancet*, vol. 395, no. 10223, pp. 497–506, 2020.

[130] C. Qin, L. Zhou, Z. Hu et al., “Dysregulation of immune response in patients with coronavirus 2019 (COVID-19) in Wuhan, China,” *Clinical Infectious Diseases*, vol. 71, no. 15, pp. 762–768, 2020.

[131] C. Wu, X. Chen, Y. Cai et al., “Risk factors associated with acute respiratory distress syndrome and death in patients with coronavirus disease 2019 pneumonia in Wuhan,” *JAMA Internal Medicine*, vol. 180, pp. 934–943, 2020.

[132] A. Tufán, A. Avanoglu Guler, and M. Matucci-Cerinic, “COVID-19, immune system response, hyperinflammation and repurposing anti-rheumatic drugs,” *Turkish Journal of Medical Sciences*, vol. 50, no. SI-1, pp. 620–632, 2020.

[133] A. Aouba, A. Baldolli, L. Geffray et al., “Targeting the inflammatory cascade with anakinra in moderate to severe COVID-19 pneumonia: case series,” *Annals of the Rheumatic Diseases*, vol. 79, no. 10, pp. 1381-1382, 2020.

[134] T. Huet, H. Beussier, O. Voisin et al., “Anakinra for severe forms of COVID-19: a cohort study,” *The Lancet Rheumatology*, vol. 2, no. 7, pp. e393–e400, 2020.

[135] B. Shakoory, J. A. Carcillo, W. W. Chatham et al., “Interleukin-1 receptor blockade is associated with reduced mortality in sepsis patients with features of macrophage activation syndrome: reanalysis of a prior phase III trial,” *Critical Care Medicine*, vol. 44, pp. 275–281, 2016.

[136] X. Xu, M. Han, T. Li et al., “Effective treatment of severe COVID-19 patients with tocilizumab,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 117, pp. 10970–10975, 2020.

[137] G. Guaraldi, M. Meschiari, A. Cozzi-Lepri et al., “Tocilizumab in patients with severe COVID-19: a retrospective cohort study,” *The Lancet Rheumatology*, vol. 2, pp. e474–e484, 2020.

[138] https://www.genengnews.com/news/sanofi-regeneron-narrow-phase-iii-kevzara-trial-to-critical-covid-19-patients/

[139] M. Ildir, A. Brito, G. Dingo et al., “Strengthening the immune system and reducing inflammation and oxidative stress through diet and nutrition: considerations during the COVID-19 crisis,” *Nutrients*, vol. 12, no. 6, p. 1562, 2020.

[140] N. Muller-Calleja, D. Manukyan, A. Canisius, D. Strand, and K. J. Lackner, “Hydroxylcholroquine inhibits proinflammatory signalling pathways by targeting endosomal NADPH oxidase,” *Annals of the Rheumatic Diseases*, vol. 76, no. 5, pp. 891–897, 2017.

[141] N. Prinz, N. Clemens, A. Canisius, and K. J. Lackner, “Endosomal NADPH-oxidase is critical for induction of the tissue factor gene in monocytes and endothelial cells. Lessons from the antiphospholipid syndrome,” *Thrombosis and Haemostasis*, vol. 109, no. 3, pp. 525–531, 2013.

[142] M. P. Ntynonga-Pono, “COVID-19 infection and oxidative stress: an under-explored approach for prevention and treatment?” *The Pan African Medical Journal*, vol. 35, Suppl 2, p. 12, 2020.

[143] K. B. Schwarz, “Oxidative stress during viral infection: a review,” *Free Radical Biology & Medicine*, vol. 21, no. 5, pp. 641–649, 1996.

[144] E. D. Danz, J. Skramsted, N. Henry, J. A. Bennett, and R. S. Keller, “Resveratrol prevents doxorubicin cardiotoxicity through mitochondrial stabilization and the Sirt1 pathway,” *Free Radical Biology & Medicine*, vol. 46, no. 12, pp. 1589–1597, 2009.

[145] N. Xia, A. Daiber, U. Forstermann, and H. Li, “Antioxidant effects of resveratrol in the cardiovascular system,” *British Journal of Pharmacology*, vol. 174, no. 12, pp. 1633–1646, 2017.

[146] S. C. Lin, C. T. Ho, W. H. Chuo, S. Li, T. T. Wang, and C. C. Lin, “Effective inhibition of MERS-CoV infection by resveratrol,” *BMC Infectious Diseases*, vol. 17, p. 144, 2017.

[147] J. Zhou, H. Chu, C. Li et al., “Active replication of Middle East respiratory syndrome coronavirus and aberrant induction of inflammatory cytokines and chemokines in human macrophages: implications for pathogenesis,” *The Journal of Infectious Diseases*, vol. 209, no. 9, pp. 1331–1342, 2014.

[148] P. B. Jakus, N. Kalman, C. Antus et al., “TRAF6 is functional in inhibition of TLR4-mediated NF-κB activation by resveratrol,” *The Journal of Nutritional Biochemistry*, vol. 24, no. 5, pp. 819–823, 2013.

[149] M. K. Leow, “Correlating cell line studies with tissue distribution of DPP4/TMPRSS2 and human biological samples may better define the viral tropism of MERS-CoV,” *The Journal of Infectious Diseases*, vol. 208, no. 8, pp. 1350-1351, 2013.

[150] D. Bestle, M. R. Heindl, H. Limburg et al., “TMPRSS2 and furin are both essential for proteolytic activation of SARS-CoV-2 in human airway cells,” *Life Science Alliance*, vol. 3, no. 9, e02000786, 2020.

[151] L. M. Reinke, M. Spiegel, T. Plegge et al., “Different residues in the SARS-CoV spike protein determine cleavage and activation by the host cell protease TMPRSS2,” *PLoS One*, vol. 12, no. 6, article e0179177, 2017.

[152] N. Gogoi, P. Chowdhury, A. K. Goswami, A. Das, D. Chetia, and B. Gogoi, “Computational guided identification of a citrus flavonoid as potential inhibitor of SARS-CoV-2 main protease,” *Molecular Diversity*, vol. 25, no. 3, pp. 1745–1759, 2021.

[153] F. Topal, M. Nar, H. Gocer et al., “Antioxidant activity of taxifolin: an activity-structure relationship,” *Journal of Enzyme Inhibition and Medicinal Chemistry*, vol. 31, no. 4, pp. 674–683, 2016.

[154] C. Sunil and B. Xu, “Potential inhibitors for novel coronavirus protease identification by virtual screening of 606 million compounds,” *International Journal of Molecular Sciences*, vol. 21, no. 10, p. 3626, 2020.
[157] A. K. Bepari and H. M. Reza, "Identification of a novel inhibitor of SARS-CoV-2 3CL-Pro through virtual screening and molecular dynamics simulation," PeerJ, vol. 9, article e11261, 2021.

[158] A. M. Sardanelli, C. Isgro, and L. L. Palese, "SARS-CoV-2 main protease active site ligands in the human metabolome," Molecules, vol. 26, no. 5, p. 1409, 2021.

[159] L. Yi, Z. Li, K. Yuan et al., "Small molecules blocking the entry of severe acute respiratory syndrome coronavirus into host cells," Journal of Virology, vol. 78, no. 20, pp. 11334–11339, 2004.