Lentinula edodes (shiitake mushroom): An assessment of in vitro anti-atherosclerotic bio-functionality

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Abstract Mushrooms have been highly regarded as possessing enormous nutritive and medicinal values. In the present study, we evaluated the anti-oxidative and anti-atherosclerotic potential of shiitake mushroom (Lentinula edodes) using its solvent–solvent partitioned fractions that consisted of methanol:dichloromethane (M:DCM), hexane (HEX), dichloromethane (DCM), ethyl acetate (EA) and aqueous residue (AQ). The hexane fraction (1 mg/mL) mostly scavenged (67.38%, IC50 0.55 mg/mL) the 2,2-diphenyl-1-picryl hydrazyl (DPPH) free radical, contained the highest reducing capacity (60.16 mg gallic acid equivalents/g fraction), and most potently inhibited lipid peroxidation (67.07%), low density lipoprotein oxidation and the activity of 3-hydroxy 3-methyl glutaryl co-enzyme A reductase (HMGR). GC–MS analyses of the hexane fraction identified a-tocopherol (vitamin E), oleic acid, linoleic acid, ergosterol and butyric acid as the bio-functional components present in L. edodes. Our findings suggest that L. edodes possesses anti-atherosclerotic bio-functionality that can be applied as functional food-based therapeutics against cardiovascular diseases.

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1. Introduction

Food and food bio-components having anti-oxidative and anti-hypercholesterolemic bio-functionalities have been highly advocated as remedies for oxidative stress-induced pathophysiological complications (Orekhov et al., 2013). Atherosclerosis, a cardiovascular disease (CVD) hallmark, has been intricately linked with an oversupply of food-based cholesterol and excessive production of oxidised low density lipoproteins (ox-LDL). The International Atherosclerotic Society (IAS) formulated...
global recommendations for the management of hyperlipidemia, emphasising control of the level of LDL in order to reduce the risk of atherosclerosis (Grundy et al., 2013). As atherosclerosis is a long-term cause and effect process, antiatherosclerosis therapy should parallel its course. In this endeavour, inhibitors of the key enzyme in cholesterol biosynthesis, 3-hydroxy 3-methyl glutaryl co-enzyme A reductase (HMGR), have been screened from micro-organisms followed by their synthetic development designated as “statins”. But the long-term use of statins and other cholesterol lowering drugs seem inept when considering tachyphylaxis, drug toxicity and cost management (Martin et al., 2012). In addition, not all people can tolerate statins and their application comes with some downside along with muscle aches upon discontinuation (Karanth et al., 2013). In order to mitigate oxidative stress towards LDL, boosting the anti-oxidative defence arsenal has been suggested (Cherubini et al., 2005). In this connection, food and food bio-component based approaches have received due attention (Sirtori et al., 2009). The European Food Safety Association (EFSA) recommended two types of functional foods: phytosterol-based and β-glucan-based, for their potency in reducing the risk of atherosclerosis en route to CVD (EFSA Panel on Dietetic Products, Nutrition and Allergies, 2010). These bio-components work by lowering the absorption of cholesterol and/or curbing its biosynthesis. The anti-oxidative mode of action of these bio-functional food components can be divided into two broad spectra: electron transfer (ET) and hydrogen atom transfer (HAT) through which the anti-oxidant itself becomes a “less harmful oxidant for a while” followed by its return to the main structural form with the help of other “network anti-oxidants”, ultimately mitigating oxidative stress.

The macro fungi, mushrooms have been reported to possess immense nutritional and medicinal bio-components and spur excellence in maintaining global public health. Mushrooms abound with phytosterol derivatives, including ergosterol and fucosterol with the aqueous fraction with ethyl acetate (EA, 3

2.1. Preparation of gradually partitioned solvent fractions of L. edodes

Proper separation and purification of the functionally active bio-components is an important criterion in demonstrating their bio-functionality. Based on the polarity and density gradient differences among five solvents (methanol, dichloromethane, ethyl acetate, hexane and water), we gradually fractionated the mushroom bio-components following a method described elsewhere (Rahman et al., 2014). Briefly, L. edodes fruiting bodies were sliced, sun-dried and ground to powder. Two hundred grams of powder was used for extraction with 4 L of methanol:dichloromethane (2:1) in conical flasks at room temperature with occasional stirring and shaking for 3 days followed by filtration through Whatman No. 1 filter paper. The extraction was repeated twice and the total organic solution, collected from each step of extraction, was evaporated using a rotary evaporator (Büchi Rotavapor R-114, Switzerland) that yielded the crude (M:DCM) extract. The dried, crude extract was dissolved in 90% aqueous methanol and partitioned with hexane (3 × 100 mL). The upper hexane layer was separated using a separatory funnel and later rota-evaporated. The bottom aqueous methanolic layer left was rota-evaporated that yielded a semisolid fraction. Re-dissolving of the semisolid fraction in distilled water (100 mL) was followed by successive partitioning with dichloromethane (DCM, 3 × 100 mL). We collected the partitioned DCM fraction and rota-evaporated. Then, we re-partitioned the aqueous fraction with ethyl acetate (EA, 3 × 100 mL) followed by collection of the resultant upper EA layer and rota-evaporation. Finally, we freeze-dried the lower aqueous part and obtained the aqueous fraction.

2.2. Evaluation of the anti-oxidant properties of L. edodes fractions

We evaluated the anti-oxidative effects of the L. edodes fractions compared with the positive control quercetin while carrying out the standard tests described below.

2.2.1. Scavenging effect on 2,2-diphenyl-1-picrylhydrrazil (DPPH) radical

We followed a previously developed method in our lab for determining the DPPH free radical scavenging effects of the L. edodes fractions (Abdullah et al., 2012). In brief, 3.9 mM of 0.06 mM DPPH dissolved in methanol was mixed with 0.1 mL of each solvent fraction (1.0 mg/mL conc.). After shaking the mixture in darkness, we measured the absorbance at 515 nm. Using methanol as the blank, we calculated the percentage of DPPH free radical scavenging using the following equation:
Radical scavenging activity (\%) = \left( \frac{A_0 - A_s}{A_0} \right) \times 100

where \( A_0 \) is the absorbance of the 0.06 mM methanolic DPPH alone, and \( A_s \) is the absorbance of the reaction mixture. The IC\(_{50}\) value (concentration of the fraction necessary to produce half maximal inhibition/scavenging) of the most potent solvent fraction was calculated from the graph of the radical scavenging activity against fraction concentration.

2.2.2. Folin–Ciocalteu assay using the \( L. \) edodes fractions

Folin–Ciocalteu reagent (10%, 250 mL) was added to an equal volume of the solvent fraction and kept at darkness for 3 min while shaking. To this mixture, 500 mL of 10% sodium carbonate was then added, followed by incubation in the dark for 1 h, followed by measurement of the absorbance at 750 nm. The calibration curve of gallic acid (2–10 mg/mL) was used to express the activity of the Folin–Ciocalteu assay as milligrams of gallic acid equivalents (mg GAE) per gram of fraction (Berger et al., 2004).

2.2.3. Inhibitory effects of the \( L. \) edodes fractions upon lipid peroxidation

We applied our previously established method to determine the inhibitory effect of each of the fractions on lipid peroxidation of buffered egg yolk (Rahman et al., 2014). Briefly, Fowl egg yolk was emulsified with 0.1 M phosphate buffer (pH 7.4) to prepare a solution of 25 g/L. Lipid peroxidation was induced by ferrous sulphate (1 M, 100 mL). The \( L. \) edodes fractions (100 mL) were then added, shaken and incubated at room temperature for 1 h. Then, 15% (v/v) of trichloroacetic acid (TCA, 500 mL) and thiobarbituric acid (TBA, 1 mL), both freshly prepared, were added. Following incubation in a boiling water bath for 10 min, the reaction mixtures were cooled and centrifuged at 3500 g for 10 min to precipitate the proteins. The absorbance of an aliquot of the supernatant (100 mL) was measured at 532 nm. Buffered egg yolk with Fe\(^{2+}\) alone was used as the control. Calculation of the percentage inhibition of lipid peroxidation was performed according to the following equation:

\[
\text{Inhibition (\%)} = \left( \frac{A_0 - A_s}{A_0} \right) \times 100
\]

where \( A_0 \) is the absorbance of the control and \( A_s \) is the absorbance of the reaction mixture containing the fraction. The IC\(_{50}\) value of the most potent solvent fraction was calculated from the graph of the inhibition of lipid peroxidation against fraction concentration.

2.2.4. FeSO\(_4\)-induced LDL oxidation and \( L. \) edodes fraction-mediated inhibition

Following our previously established method, we determined the \( L. \) edodes fraction-mediated inhibition of human LDL (Sigma–Aldrich, St. Louis, USA) oxidation in vitro (Rahman et al., 2014). The observed aspects included:

2.2.4.1. Effect of the \( L. \) edodes fractions upon the lag time of conjugated diene (CD) formation. We induced Fe\(^{2+}\) (FeSO\(_4\), 50 μg/mL)-mediated oxidative stress towards human LDL (150 μg/mL) at room temperature and pH 7.4. Subsequently, we observed the kinetics of both Fe\(^{2+}\)-induced LDL oxidation and the \( L. \) edodes fraction-mediated (1 μg/mL) corresponding inhibition at 234 nm at 20 min intervals for a period of 180 min. FeSO\(_4\) in ultrapure water (pH 7.4) was used as the blank. Along the time course of oxidation, the \( L. \) edodes fraction-mediated protection phase denoted as the “lengthened lag time of CD formation” was identified and measured.

2.2.4.2. Effect of \( L. \) edodes fractions upon malondialdehyde (MDA) production. Oxidative modification of human LDL was performed using 10 mM ferrous sulphate followed by the addition of \( L. \) edodes fractions at a 1 mg/mL concentration. Trichloroacetic acid (TCA, 500 μL, 15% v/v) and thiobarbituric acid (TBA, 1 mL, 1% v/v) were added and incubated at 100 °C for 10 min. After cooling, an aliquot (300 μL) was taken to the ELISA reader and the absorbance read at 532 nm. For the blank, FeSO\(_4\) in water, pH 7.4, was used. We used a standard assay kit (Cayman Chemicals’ TBARS assay kit, item No. 1000955) and followed the manufacturer’s recommendations to calculate the production of MDA.

2.3. Determination of HMG Co-A reductase (HMGR) inhibitory effect

The in vitro HMG Co-A reductase (HMGR) inhibitory effect of the various fractions of \( L. \) edodes was tested using the HMGR assay kit (Sigma-Aldrich, catalogue No. CS1090) (Gholamhoseinian et al., 2010). The human catalytic domain of the HMGR enzyme (concentration 0.6 mg protein/mL) and pravastatin as a positive control were used according to the recommended conditions. In order to attain a final mixture of volume 200 μL and concentration 400 μM, 4 μL of NADPH and 12 μL of HMG-CoA substrate were added with 2 μL of the catalytic domain of human recombinant HMGR together with an appropriate amount of 100 mM potassium phosphate buffer (containing 120 mM KCl, 1 mM EDTA, and 5 mM dithiothreitol, pH 7.4). Aliquots (1 μL) of pravastatin and \( L. \) edodes fractions (1 mg/mL) were added and the rate of NADPH oxidation by HMGR was monitored every 20 s at 340 nm for a period of 10 min using a Bio Tek H1 synergy hybrid multi-mode plate reader equipped with Gen 5 data analysis software (Bio Tek, VT, USA).

2.4. Identification of bio-functional components by GC–MS

As the hexane fraction of \( L. \) edodes showed the best performance in terms of anti-oxidative and HMG Co-A reductase inhibitory activities, it was subjected to GC–MS analysis to identify the bio-functional and anti-atherosclerotic components. We utilised gas chromatography directly coupled to a mass spectrometer system (Agilent 7890 C triple quadruple GC/MS system, USA). Utilising a HP-5 ms silica capillary column (30 m × 250 μm, 0.25 μm film) the instrumental conditions maintained were: oven temperature 70 °C–300 °C (finally maintained for 29 min); inert helium gas as the carrier, flow rate 1 mL/min; injection volume 1.5 μL; injection technique split less; injector temperature 250 °C; ionisation energy 70 eV; modode electronic ionisation (EI); temperature of ion source 200 °C; range of masses scanned 50–550 m/z; and interface line temperature 300 °C. Mass spectra of the National Institute of Standards and Technology (NIST 08 and NIST08 s) library data were used as a reference to identify the peaks.
2.5. Statistical analyses

We conducted all the experiments in triplicate and the data are presented as mean ± SD. Using statistical package SPSS version 16 we performed one-way analysis of variance (ANOVA). The differences among means were further analysed by the least significance difference (LSD) test at a 95% confidence level (p < 0.05).

3. Results and discussion

3.1. DPPH-free radical scavenging activity

The DPPH-free radical scavenging performance of the *L. edodes* fractions was evaluated at a concentration of 1 mg/mL and the results are presented in Fig. 1. All the fractions were capable of scavenging free radicals but varied in their potency. The hexane fraction had the highest scavenging activity (67.38%, IC₅₀ 0.55 mg/mL), followed by that of DCM (50.85%, IC₅₀ 0.68 mg/mL). The other three fractions did not differ significantly in terms of DPPH free radical scavenging.

The anti-oxidative test evaluating the scavenging effect of the *L. edodes* fractions upon the DPPH free radical is an ET based assay. All the fractions possessed free radical scavenging potential but the scavenging effect of the hexane fraction surpassed that of all the others (Fig. 1). It measured the cumulative antioxidant potential of the mushroom fractions whereby the scavenging antioxidants present in the respective fractions reduced the stable, purple radical DPPH and converted it into the yellow coloured, non-radical form (DPPH-H) (Fig. 2).

3.2. Folin–Ciocalteu assay

The reducing capacity of the bio-components is a special feature of their anti-oxidative potency. In connection with this, we performed the Folin–Ciocalteu assay compared to quercetin. Reducing prowess was expressed in terms of milligrams of gallic acid equivalents per gram of fraction (mg GAE/g fraction). The results are presented in Fig. 2, indicating the hexane fraction possessed the highest reducing capacity (60.16 GAE/g fraction), followed very closely by those of DCM (58.86 mg GAE/g fraction) and EA (55.32 mg GAE/g fraction).

The Folin–Ciocalteu assay performed in the present experiment (Fig. 3) is another ET-based assay through which the total reducing capacities of the *L. edodes* bio-components were measured. The data presented in Fig. 2 are in line with a previous study undertaken by Abdullah et al. (2012). As compared to the natural reducing agent gallic acid, *L. edodes* showed promising results and thus we further studied its effects upon lipid peroxidation and LDL oxidation.

3.3. Lipid peroxidation inhibitory effect

The inhibitory effect of the *L. edodes* fractions upon lipid peroxidation is shown in Fig. 3. We induced oxidative modification of lipid structures with a transition metal, Fe⁺², and assessed whether the *L. edodes* fractions could lower the inhibition or result in a reduced rate of oxidised lipid structural byproduct generation. We found that the hexane fraction had the most potent inhibitory effect on lipid peroxidation (67.07%), followed by DCM (64.23%) (Fig. 4). This finding was in line with the free radical scavenging and reducing capacities of the fractions and suggested these effects might be attributed to the lipophilic bio-components present in the hexane fraction of *L. edodes*.

3.4. Inhibition of low density lipoprotein (LDL) oxidation by *L. edodes* fractions

The kinetics of LDL oxidation incorporates an initial lag period of slow oxidation followed by a log period of enhanced oxidation. During the lag period conjugated dienes (CD) are formed, and during the log period malondialdehydes (MDA) are produced.
found that all MDA corresponds to the severity of LDL oxidation. We added the lipid hydro-peroxide double bonds. Thus, the level of oxidized lipids are generated (Yoshida and Kisugi, 2010). The anti-atherosclerotic performance of bio-components involves their mediating effect upon increasing the lag time and decreasing MDA production (Li and Mehta, 2005). The hexane fraction of L. edodes also performed the best in this regard.

3.4.1. Effect of L. edodes fractions upon the lag time of CD formation

Oxidative stress exerted upon LDL molecules by Fe^{2+} affects the polyunsaturated fatty acids (PUFAs) present in the LDL molecule and produces conjugated dienes (CD). Fig. 5 represents the kinetics of CD production and the effect of L. edodes fractions upon this kinetic pattern. The hexane fraction lengthened the lag time (125 min) of CD formation the most, indicating it possessed the highest inhibitory activity against LDL oxidation.

3.4.2. Effect of L. edodes fractions upon MDA formation

In the LDL oxidation cascade, CD formation is rapidly followed by the production of MDA through the breakage of the lipid hydro-peroxide double bonds. Thus, the level of MDA corresponds to the severity of LDL oxidation. We found that all L. edodes fraction-treated samples had a lower MDA production (data not shown for the sake of brevity). Among all the solvent fractions of L. edodes, the hexane fraction rendered the smallest amount of MDA (21.57 μM/mg LDL), indicating it had the highest content of bio-components capable of withstanding LDL oxidation.

3.5. Inhibitory effect of L. edodes fractions upon HMG Co-A reductase (HMGR) activity

We assessed whether the L. edodes fractions could inhibit the activity of the cholesterol biosynthetic regulatory enzyme HMG Co-A reductase (HMGR) in vitro. All the fractions could inhibit the activity of the catalytic domain of human HMGR at 1 mg/mL concentration. As depicted in Fig. 6, here also, the hexane fraction-mediated inhibitory effect was the most potent (44.26%).

An increased level of total cholesterol is an independent risk factor for atherosclerosis. Interruption of cholesterol biosynthesis through direct inhibition of its regulatory enzyme HMG Co-A reductase has been a target for the last few decades. We also evaluated whether L. edodes could inhibit HMG Co-A reductase activity in vitro. Consistent with our other findings, the hexane fraction of L. edodes inhibited the activity of HMG Co-A reductase the most (44.26%) as compared to the synthetic statin drug pravastatin (87.64%). The anti-oxidative effect of the statin drugs has been regarded as conferring their anti-atherosclerotic effect (Puttananjaiah et al., 2011). L. edodes possesses the top-ranking position among 26 different edible and medicinal mushrooms in terms of exerting inhibitory effects towards HMG Co-A reductase activity (Gil-Ramirez et al., 2013a,b). However, different parts of the mushroom body differ from one another in terms of potency, with the L. edodes cap possessing the best HMG Co-A reductase inhibitory activity.

3.6. Identification of the anti-atherosclerotic bio-functional components by GC–MS

The bio-functional components in the hexane fraction of L. edodes that exhibited anti-atherosclerotic effects were...
identified through GC–MS analyses. The chromatograph presented in Fig. 7 demonstrates the peaks of the identified bio-components. Among them, the most abundant were \( \alpha \)-tocopherol (vitamin E), oleic acid, linoleic acid, ergosterol and butyric acid (Table 1).

GC–MS analysis identified the presence of the chain breaking anti-oxidant \( \alpha \)-tocopherol (vitamin E) in the hexane fraction. The inhibitory effect of \( L. \) edodes towards LDL oxidation observed in the present study might be attributed to the \( \alpha \)-tocopherol content of this mushroom. Vitamin E scavenges free radicals by following hydrogen atom transfer (HAT) mechanism. It reacts with a free radical such as a lipid peroxyl molecule and interrupts further peroxidation by producing a hydroperoxide and a radical form of \( \alpha \)-tocopherol. Both of these intermediates are less harmful as they are almost unreactive. The radical form of \( \alpha \)-tocopherol can either react with another peroxyl molecule to turn into a functionally unreactive product or revert back to its original functional form of vitamin E (Fig. 8).

Despite some exceptions, studies involving animal models and clinical trials as well as human epidemiological data suggest the protective role of \( \alpha \)-tocopherol against atherosclerosis (O’Byrne et al., 2002; Munteanu et al., 2004). Vitamin E was able to reduce atherosclerotic lesions up to 60% in experimentally induced atherosclerosis-prone mice (Peluzio et al., 2001; Schwenke et al., 2002). Its anti-atherosclerotic role in hypercholesterolemic rabbits has also been documented (Schwenke et al., 2002). At physiological concentrations (10–30 \( \mu \)M), \( \alpha \)-tocopherol could lengthen the lag time of CD formation from 59.6 to 98.9 min (Baldi et al., 2003).

Ergosterol, another bio-component identified in the hexane fraction of \( L. \) edodes, has been suggested to act as a membrane antioxidant upon ingestion. Ohnuma et al. (2000) isolated and elucidated the structural features of ergosterol from \( L. \) edodes. The inhibitory effect of ergosterol peroxide isolated from the mushroom \( Armillariella mellea \) on lipid peroxidation was greater than those of \( \alpha \)-tocopherol and thiourea (Wiseman et al., 1993). Supplementation of ergosterol and oleic acid in medium containing \( Saccharomyces cerevisiae \) has been reported to lower the production of reactive oxygen species, oxidative stress and the resultant damage at the cellular and biomolecular levels (Ohnuma et al., 2000). Ergosterol peroxide

![Figure 7](image-url)
and ergosterol have been reported to suppress LPS-induced inflammatory responses through inhibiting NF-κB and C/EBPα transcriptional activity, and phosphorylation of MAPKs (Kim et al., 1999). Mushrooms’ antioxidant effects relating to DPPH free radical scavenging and Folin–Ciocalteu assay (reducing capacity test) has been attributed towards its content of different bio-components (Landolfo et al., 2010). However, anti-oxidative performance varies depending on mushroom species and also based on the extracting solvent utilised. In case of polar solvent-based mushroom bioseparation, phenolic and polyphenolic group of reducing substances have been regarded for providing with DPPH free-radical scavenging and Folin–Ciocalteu assay performance while triterpenoids have been linked with metal chelating activities (Landolfo et al., 2010; Kobori et al., 2007). For non-polar solvent such as hexane, ergosterol has been identified as the principal anti-oxidative mushroom bio-component (Kalogeropoulos et al., 2013). Thus, 5,6-dihydroxy ergosterol and structurally relevant components such as 7,22-ergostadienol, ergosta 7,22-dien-3-ol present in the hexane fraction might have been involved in chelating transition metal ion, Fe^{2+} and thus mediating reduced oxidative stress and consequent lowered LDL oxidation. In case of Grifola frondosa, the anti-oxidative pattern of ergosterol has been documented to be second only to fatty acids (Nieto and Chegwin, 2008).

Fatty acids are important functional bio-components of both edible and medicinal fungi. In the hexane fraction of L. edodes we identified the presence of oleic acid, linoleic acid and butyric acid. Oleic acid is a monounsaturated fatty acid that has been reported to provide anti-atherosclerotic effects in different diets, especially the Mediterranean diet (Shao et al., 2010). Its anti-atherosclerotic mode of action involves dampening of endothelial activity by reducing the expression of vascular cell adhesion molecule – 1 (VCAM-1) and the activity of NF-κB (Zhang et al., 2002). As a result, the adherence of monocytes to endothelial walls is reduced and the associated pro-inflammatory and pro-atherogenic processes slowed down. Butyric acid, another short chain fatty acid found in the L. edodes fraction, has also been reported to confer an anti-atherosclerotic effect through modulating VCAM-1 and NF-κB activity (Massaro et al., 1999).

Linoleic acid (LA, cis-9, cis-12 octadecadienoic acid) has been among the most abundant fatty acids found in some other species of edible mushrooms (Kalogeropoulos et al., 2013). Linoleic acid and its constitutional and stereoisomers

| Table 1 | Identification of the bio-active components of the hexane fraction of L. edodes by GC–MS. |
|---------|------------------------------------------------------------------------------------------|
| No      | Bio-component | Structure | Molecular formula | Molecular weight (g) |
| 1       | α-Tocopherol | ![Structure](image1) | C_{29}H_{50}O_{2} | 430.71 |
| 2       | Oleic acid  | ![Structure](image2) | C_{18}H_{34}O_{2} | 282.46 |
| 3       | Linoleic acid | ![Structure](image3) | C_{18}H_{32}O_{2} | 280.45 |
| 4       | Ergosterol | ![Structure](image4) | C_{28}H_{44}O | 396.65 |
| 5       | Butyric acid | ![Structure](image5) | C_{4}H_{8}O_{2} | 88.11 |

Figure 8  Basic mechanisms demonstrating the lipid peroxidation inhibitory effect of α-tocopherol.

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termed “conjugated linoleic acid (CLA)” have become an attractive subject of anti-atherosclerotic studies. Research involving the effects of linoleic acid upon animal models has demonstrated its considerable inhibitory effect on the establishment and progression of atherosclerosis. Dietary supplementation of conjugated linoleic acid (CLA) could lower blood levels of LDL and triacylglycerol as well as slow down the progression of atherosclerotic lesion formation (Carluccio et al., 1999). The proposed mechanism involved CLA-induced enhanced expression of the nuclear transcription factor peroxisome proliferator activated receptor γ (PPARγ), especially in vascular smooth muscle cells. Although some in vitro studies indicate a pro-oxidative and pro-atherogenic role of linoleic acid, compelling in vivo evidence has yet to be established. In human subjects, linoleic acid rich diets have been credited with reducing the risk of coronary heart diseases (Zapolska-Downar et al., 2004).

Inhibitory effect towards the activity of HMG Co-A reductase might also be attributed to ergosterol present in the hexane fraction of L. edodes. Our proposition is backed by the findings of both in vitro, ex vivo and in vivo studies (Rudel, 1999; Kritchevsky et al., 2000; Gil-Ramirez et al., 2013a,b). Ergosterol peroxide from the edible mushroom Sarcoodon aspratus, had been found to down-regulate the expression of low-density lipoprotein receptor (LDLR) and HMG-CoA reductase (HMGCR) genes in RAW264.7 cells (Kim et al., 1999). Supplementation of ergosterol and oleic acid in the medium containing Saccharomyces cerevisiae had been reported to lower the production of reactive oxygen species, oxidative stress and the resultant damage towards cellular and biomolecular levels (Ohnuma et al., 2000). The HMG Co-A reductase inhibitory effect is mediated by the statins. Contrary to the established notion of the statins for hypcholesterolemic performance, the present study reports a different mechanistic approach of the mushroom bio-components. As the present study had not searched for the presence of statins in the L. edodes fractions, this remains as an important scope of our future studies.

4. Conclusion

L. edodes (shiitake mushroom) has been found possessing bioactive food components capable of conferring anti-oxidative defence and curtailting LDL oxidation as well as being potent in inhibiting the activity of the rate limiting enzyme in cholesterol biosynthesis, thus supporting its use as an anti-atherosclerotic agent. Food bio-components including α-tocopherol (vitamin E), oleic acid, linoleic acid, ergosterol and butyric acid present in L. edodes enabled it to act as an anti-atherosclerotic agent. The current findings can be further explored in in vivo models to substantiate the therapeutic use of L. edodes with a view to aiding atherosclerosis-affected people around the world.

Author contribution

Noorlidah Abdullah and Norhaniza Aminudin planned and supervised the research, edited the manuscript. Mohammad Azizur Rahman conducted the research, statistical analyses and prepared the manuscript.

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