Multimodal synchrotron X-ray fluorescence imaging reveals elemental distribution in seeds and seedlings of the Zn–Cd–Ni hyperaccumulator Noccaea caerulescens

Antony van der Ent 1,*, Martin D. de Jonge 2, Guillaume Echevarria 3, Mark G.M. Aarts 4, Jolanta Mesjasz-Przybyłowicz 5, Wojciech J. Przybyłowicz 5,6, Dennis Brueckner 7,8,9 and Hugh H. Harris 10

1 Centre for Mined Land Rehabilitation, Sustainable Minerals Institute, The University of Queensland, St Lucia 4072, Australia, 2 Australian Synchrotron, ANSTO, Clayton 3168, Australia, 3 Laboratoire Sols et Environnement, Université de Lorraine-INRAE, Vandoeuvre-lès-Nancy, UMR 1120, France, 4 Laboratory of Genetics, Wageningen University and Research, The Netherlands, 5 Department of Botany and Zoology, Stellenbosch University, Matieland 7602, South Africa, 6 AGH University of Science and Technology, Faculty of Physics & Applied Computer Science, 30-059 Kraków, Poland, 7 Deutsches Elektronen-Synchrotron DESY, 22607 Hamburg, Germany, 8 Department of Physics, Université Paul Cézanne, Aix-Marseille University, 13453 Marseille, France, 9 Faculty of Chemistry and Biochemistry, Ruhr-Universität Bochum, 44801 Bochum, Germany and 10 Department of Chemistry, The University of Adelaide, Adelaide 5005, Australia

*Correspondence: Centre for Mined Land Rehabilitation, Sustainable Minerals Institute (SMI), Level 5, Sir James Foots Building (No. 47A), The University of Queensland, St Lucia QLD 4072, Australia. Tel: +61 7 3346 4003; E-mail: a.vanderent@uq.edu.au

Abstract

The molecular biology and genetics of the Ni–Cd–Zn hyperaccumulator Noccaea caerulescens has been extensively studied, but no information is yet available on Ni and Zn redistribution and mobilization during seed germination. Due to the different physiological functions of these elements, and their associated transporter pathways, we expected differential tissue distribution and different modes of translocation of Ni and Zn during germination. This study used synchrotron X-ray fluorescence tomography techniques as well as planar elemental X-ray imaging to elucidate elemental (re)distribution at various stages of the germination process in contrasting accessions of N. caerulescens. The results show that Ni and Zn are both located primarily in the cotyledons of the emerging seedlings and Ni is highest in the ultramafic accessions (up to 0.15 wt%), whereas Zn is highest in the calamine accession (up to 600 µg g⁻¹). The distribution of Ni and Zn in seeds was very similar, and neither element was translocated during germination. The Fe maps were especially useful to obtain spatial reference within the seeds, as it clearly marked the vasculature. This study shows how a multimodal combination of synchrotron techniques can be used to obtain powerful insights about the metal distribution in physically intact seeds and seedlings.

Keywords: hyperaccumulator, reconstructions, seed, synchrotron, X-ray fluorescence microscopy, X-ray fluorescence tomography

Graphical abstract

Tomographic reconstruction and 2D elemental mapping of Noccaea caerulescens seeds.

Introduction

Hyperaccumulator plants are the perfect model systems for advancing our comprehension of the molecular biology and physiology of the regulation of transition elements, including Ni, Mn, and Zn.1–3 The localization of transition elements in hyperaccumulator seeds in relation to the germination processes remains understudied. Seed germination is a vital phase in the lifecycle of plants as the nascent seed has to contain all of the essential elements needed for emergence of the young plant.4 Previous studies on Berkheya coddii showed that Ni is located mainly in the micropylar area, in the embryo, and in the lower epidermis of cotyledons, whereupon it is mobilized to leaf palisade parenchyma after the first leaves emerge.5 In Noccaea praecox, Cd was translocated to the shoots (and not the roots) during germination.6

The genus Noccaea is unique in that it has over 20 taxa that hyperaccumulate different transition elements (Ni, Zn, Cd, and Pb) with several species hyperaccumulating more than one

Received: December 30, 2021. Accepted: March 18, 2022

© The Author(s) 2022. Published by Oxford University Press. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (https://creativecommons.org/licenses/by/4.0/), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.
element simultaneously. The levels of efficiency and specificity can be remarkable, e.g. *Noccaea caerulescens* can accumulate 8890 μg g⁻¹ Zn from a soil with only 139 μg g⁻¹ Zn. This species is unique in that it possesses calamine, ultramafic, and non-metallicolous ecotypes, each represented by different accessions, which differ in tolerance and hyperaccumulation. All calamine, ultramafic, and non-metallicolous *N. caerulescens* accessions hyperaccumulate Zn and Ni, while not all will accumulate Cd when it is supplied in their environment. *Noccaea caerulescens* accessions display remarkable differences in their tolerance of these metals depending on the origin. Not only is *N. caerulescens* of interest because of the spectrum of metals it tolerates and hyperaccumulates, it makes for a powerful model system because of the availability of a full molecular toolkit with the nuclear genome and transcriptome of this species having been established (Severing and Aarts, unpublished work). Consequently, candidate genes involved in Ni and, mainly, Zn uptake and translocation are now known. Zinc appears to be taken up by plasma-membrane located ZIP (Zinc-regulated, iron-regulated transporter-like Protein) Zn-importing transporters and is then loaded into the xylem by plasma-membrane located HMA (Heavy Metal ATPase 4) Zn-exporting transporters for translocation to the shoot. This most likely requires additional ZIP proteins and the vacuolar loading P-type ATPase HMA3. In addition, genetically segregating accessions have been generated by crossing different accessions, which can be used to genetically characterize different metal-adaptation traits and for three metallicolous and three non-metallicolous natural accessions, whole-genome resequencing data are generated (Wang, van den Heuvel, and Aarts, unpublished work) for population genomics purposes. Next to these non-metallicolous and calamine accessions, ultramafic accessions are known, e.g. many *N. caerulescens* accessions have been cultivated in genetically homogeneous lines by recurrent inbreeding (Schat and Aarts, unpublished work). The diversity in metal tolerance/hyperaccumulation is inheritable and not dependent on each other (meaning that hyperaccumulation can be combined with hypersensitivity and consequently lethality upon metal exposure). The calamine (as well as some non-metallicolous) accessions have been studied to much greater extent than those from ultramafic soils. Most investigations have focused on the ultramafic accession from Monte Prinzer, in Italy, including proteomics studies aimed at correlating adaptation to protein expression and transcriptome studies. Nickel, as well as Zn, xylem loading in *N. caerulescens* is facilitated by histidine, although there is no evidence that this involves 1:1 HIs–Ni complexation and the related genes are not yet known. Information on the distribution of transition elements and translocation during the germination process will shed light on the seed physiology of hyperaccumulator plants. Currently, there is no information on the redistribution and the translocation of Ni and Zn during seed germination in *N. caerulescens*. Synchrotron-based X-ray fluorescence microscopy (XFM) imaging can be used for increasing our understanding of the fundamental processes involved with hyperaccumulation at different length-scales (whole plant scale down to the subcellular level) and with sufficient sensitivity for the measurement of the entire metabolome of a plant. The ability to perform element mapping of live seedlings is contingent on the ultra-fast X-ray detection of the Maia detector system (with per-pixel dwell as low as 0.1 ms), which avoids radiation damage and associated sample degradation to the specimens. Most synchrotron experiments acquire ‘2D’ elemental maps of leaves or tissue cross-sections, but X-ray fluorescence micro-computed tomography (XFM-CT) enables reconstruction of ‘virtual cross-sections’ or 3D models of elemental data from a series of projection images. It enables ‘virtual sectioning’ of specimens, thereby entirely avoiding artefacts arising from destructive sample preparation. This method is primarily limited by self-absorption of escaping fluorescent X-rays (which is highly dependent on the X-ray energy, and hence, the element of interest). Therefore, it lends itself best to specimens that are relatively thin along the two axes perpendicular to the axis of rotation, which means that plant seeds are ideal test subjects for this approach. A pioneering study by Kim et al. applied XFM-CT to reveal Fe localization in the provascular system of intact Arabidopsis seeds.

Due to the different physiological functions of Ni and Zn, both essential nutrients that are toxic at high concentrations, but with the first required in much lower quantities than the second and with only partly overlapping associated transporter pathways, we expect differential distribution of Ni and Zn in the seeds of *N. caerulescens* and different modes of translocation during germination. This research aims to obtain information on the spatially resolved elemental distribution in the seeds and seedlings in different accessions of *N. caerulescens* contrasting in their Ni and Zn accumulation characteristics, ranging from high Ni with low Zn to low Ni with high Zn. We use synchrotron X-ray fluorescence tomography for the intact seeds and planar elemental imaging for the seedlings to elucidate elemental distribution at various stages of the germination process.

**Materials and methods**

**Seed collection and culture conditions**

The seeds were sourced from *N. caerulescens* growing in the native habitat in France and Spain from different accessions: two ultramafic accessions (Vosges in France and Cira in Spain) and a non-ultramafic (calamine) accession (‘Ganges’ near St Laurent le Minier in France). The ecology and soil chemistry of these accessions has been described in detail elsewhere. The seeds were germinated on moist [deionized water (DI)] Wettex (Vileda) at 20°C under fluorescent lights for 5 days until the cotyledons emerged and the root and hypocotyl were fully formed.

**Bulk chemical analysis of seeds**

The elemental concentrations of the seeds were determined after microwave acid digestion of ~100 mg of air-dried seeds using 4 ml HNO₃ (70%) and subsequently analysed using inductively coupled plasma atomic emission spectroscopy (ICP–AES, Thermo Scientific iCAP 7400), as described earlier.

**Scanning electron microscopy**

The seeds were carbon coated and mounted on stubs and imaged with scanning electron microscopy (SEM) using a JEOL JSM-6610 instrument. Images were acquired at 100–200x magnification at 5 kV accelerator energy (Fig. 1A–C), as described earlier.

**XFM experiments**

The XFM beamline of the Australian Synchrotron produces monochromatic radiation in the range of 4.1–20 keV with a focus down to ~1 μm and a Maia detector in backscatter geometry. The 2D elemental maps of the live seedlings were acquired using an incident energy of 15.8 keV, while the 3D tomograms of the seeds were acquired by collecting 2D XFM maps viewed between 152 and 251 angles spaced over 360°. The horizontal pixel size was either 5 microns with a dwell time of 1 ms, or 8 microns with a dwell time of 1.6 ms, while in the vertical direction the pixel size was twice that of the horizontal. In total, six seeds (two
replicates from each of the three accessions Ganges, Vosges, and Cira) were imaged (Fig. 1D). The intact seeds were mounted, using a tiny amount of cyanoacrylate glue, on a needle attached to the tomography stage. The whole intact live seedlings were held between two sheets of polyethylene (Ultralene) thin film (4 μm) held over a Perspex frame in front of the Maia Detector. The two sheets of Ultralene thin film prevent dehydration during the measurement, which on average took just 30 min per sample. To avoid potential artefacts capable of confounding the collected data due to measurement conditions, we undertook fast scanning (to minimize the effective radiation dose) and applied a cold shock (by contacting the seedlings with ice cold water for 30 min immediately prior to the analysis) to slow down metabolic processes during the analysis and to reduce beam damage.19

Data analysis

The X-ray Fluorescence Spectroscopy (XRF) event stream was analysed using the dynamic analysis method13,44 using the GeoPIXE package.45,46 The analysis of the tomographic data consisted of alignment and reconstruction steps. In case of the 3D tomograms the alignment steps were cross-correlated in the vertical direction and a combined consistency and cross-correlation method was applied in the horizontal direction. Considering that the single-slice tomograms lack vertical information, they were only aligned in horizontal direction. The aligned data were then reconstructed using a maximum-likelihood expectation-maximization algorithm based on the functions of the scikit-image Python library. Finally, the 3D renderings of the seeds were created using the open-source scientific visualization software Drishti.47

Table 1. Elemental concentrations in dry seeds of Noccaea caerulescens originating from Cira (Spain) and Vosges and Ganges (France)4

| Accession | Mg  | P    | K    | Ca  | Mn | Fe | Co | Ni | Cu  | Zn | Cd | Pb  |
|-----------|-----|------|------|-----|----|----|----|----|-----|----|----|-----|
| Cira      | 1190| 5470 | 4680 | 3680| 81 | 53 | 1.44| 1600| 2.99| 102| 4.27| 1.65|
| Vosges    | 2950| 6760 | 6000 | 3980| 143| 141| 9.91| 1000| 2.89| 191| 3.12| 1.37|
| Ganges    | 2250| 6160 | 5500 | 7650| 103| 68 | 1.05| 65  | 17  | 700| 704 | 4.08|

*All concentrations in micrograms per gram (μg g⁻¹) dry weight.

Results

Elemental concentrations in seeds

The seeds originating from the French accessions (Ganges, Vosges) and Spain (Cira) differed in their concentrations of Ni and Zn (Table 1). Seeds from ultramafic locations (Vosges, Cira) were characterized by very high concentrations of Ni (1000 and 1600 μg
g$^{-1}$ dry weight, respectively) whereas the seeds from the non-ultramafic (Ganges, Zn–Pb enriched ‘calamine’) location had only 65 μg g$^{-1}$ Ni. The Zn concentration was highest in seeds from the Ganges accession (700 μg g$^{-1}$ dry weight) while in the other two accessions it was only between 100 and 190 μg g$^{-1}$ dry weight. Furthermore, an exceptionally high concentration of Cd was recorded in seeds from the Ganges location (704 μg g$^{-1}$ dry weight) while in the other accession, Cd reached only a few micrograms per gram (μg g$^{-1}$) dry weight. Other microelements (Mn, Fe, Co, Cu) were within the normal physiological range for plants.

**Tomographic reconstructions of intact non-germinated seeds**

The 3D tomography was undertaken in seeds from the Ganges, Vosges, and Cira accessions in duplicate (Fig. 2). For each seed ~250 slices were obtained. Based on the slices, 3D tomographic reconstructions of dry N. caerulescens seeds were made (Fig. 3, Supplementary Fig. S1). The Fe maps are very similar between the accessions, with the characteristic provascular strand network of Fe enrichment in the radicle and vasculature of the cotyledons. Iron is also high in the endosperm and hilum area. Only the Vosges and Cira accessions (which are from ultramafic soils) have detectable Ni. Nickel concentrations are much higher in Vosges than in Cira seeds (although the concentrations from tomography are not quantitative, but can be gauged from raw intensity values), with Ni present in the epidermal cells of the cotyledons and radicle, and minor enrichment in the cotyledon blades (but notably not in the vascular bundles) (Fig. 2). This pattern is similar in the seeds of the Cira accessions, albeit less distinct due to the lower prevailing concentrations. The Zn concentrations are highest in Ganges (which is non-ultramafic, but calamine), lower in the Cira accession, and lowest in the Vosges accession; however, the Zn distribution patterns appear to be similar between all three accessions. Zinc concentrations are highest in the cortex of the radicle, and in the outer part of the cotyledon blades. Vascular bundles, including the central nerve, are depleted in Zn.

**Fig. 2** Single-slice X-ray fluorescence microscopy (XFM) tomograms of dry Noccaea caerulescens seeds showing Fe, Ni, and Zn signals within the physically intact seeds. The seed accessions shown here originate from Ganges (non-ultramafic, calamine) and Vosges (ultramafic) in France and Cira (ultramafic) in Spain.

**Fig. 3** Tomographic reconstruction of dry Noccaea caerulescens seeds showing Fe and Ni or Zn signals within the physically intact seeds. The seed accessions shown here originate from Ganges (non-ultramafic, calamine) and Vosges (ultramafic) in France and Cira (ultramafic) in Spain.
Elemental distribution in live seedlings at different stages of development

In all accessions, Fe is particularly enriched in the cotyledon provascular bundles as well as in the hypocotyl (Figs 4 and 5, Supplementary Figs S3 and S6). Iron concentrations locally reach up to 500 μg g⁻¹ and are highest in hotspots in the hilum of the micropylar area (Fig. 4). Iron enrichment in the provascular strands disappears when the seedling develops further. Manganese is distributed throughout the seeds (mainly cotyledons), and lower in the root (Fig. 4). The distribution of Ni and Zn is very similar in most accessions and strongly enriched in the cotyledons. Whereas Ni is especially high (up to 0.25 wt%) in the central part of the cotyledon (Fig. 4, Supplementary Fig. S3), Zn is enriched toward the apex of the cotyledon blade (up to 400 μg g⁻¹). In the calamine accession, Zn reaches up to 400 μg g⁻¹ (Fig. 6), and the distribution pattern is similar in the ultramafic accessions (Vosges, France, and Circa, Spain, which have even higher Zn enrichment, up to 600 μg g⁻¹; Supplementary Fig. S7). In the young seedlings from ultramafic accessions, Zn is evenly distributed through the cotyledon blade, with an increase toward the apex (Supplementary Fig. S7). In the calamine accession Zn is highest in the blade margins (Supplementary Figs S4 and S6).

During the first stage of germination (e.g. emergence of the cotyledons and radicle), K is strongly enriched in the radicle (especially in the root apex) and hypocotyl (Fig. 6) and this pattern remains during further development in seedlings from both ultramafic and non-ultramafic accessions (Figs 4 and 5, Supplementary Figs S2–7). In later stages of development (e.g. full extension of the hypocotyl and root), K is enriched in the radicle and can reach up to 2 wt% (Supplementary Fig. S4), before it is translocated into the hypocotyl. Comparing seedlings in different stages of development (compare K maps of seedlings with (Fig. 2) and without dehisced seed coat (Figs 3 and 5), it appears that K is translocated to the radicle from the cotyledons during seedlings’ emergence. Calcium is relatively low in the seedlings (≤0.1 wt%), but strongly enriched in the dehisced seed coat with up to 0.5 wt% Ca (Fig. 6; Supplementary Figs S2 and S7). In more developed seedlings from both ultramafic and non-ultramafic accessions (Fig. 5), Ca is enriched in the epicotyl and hypocotyl (especially in the node between epicotyl and hypocotyl).

Discussion

Synchrotron XFM-CT enables ‘virtual sectioning’ of a specimen thereby entirely avoiding artefacts arising from destructive sample preparation, beyond removal from the native environment. As such, it has broad utility across the biological and medical sciences, but this is mostly yet to be realized. Herein, topographical reconstructions of intact seeds from metal hyperaccumulating plants revealed distinct localization of Ni and Zn in the anatomical structures of the seeds, showcasing the usefulness of this analytical approach.

This study sought to investigate the distribution of Ni and Zn in intact seeds of the Ni-Cd-Zn hyperaccumulator N. caerulescens using XFM techniques, and the distribution of these and other physiologically relevant elements during seed germination. We used seeds from different accessions contrasting in their Ni and Zn hyperaccumulation characteristics (ultramafic versus calamine accessions). All accessions of N. caerulescens hyperaccumulate Zn, regardless of the nature of the soil. Nickel hyperaccumulation,
however, is limited to ultramafic soils in the field, though different accessions originating from calamine or non-ultramafic soil will be able to take up Ni at high levels above the hyperaccumulation threshold as well. While we expected differential tissue distribution and different modes of translocation of Ni and Zn during germination, this is not the case. Nickel and Zn are both located primarily in the cotyledons of the emerging seedlings and Ni is highest in the ultramafic accessions (0.16 wt% on the basis of ICP–AES data), whereas Zn is highest in the calamine accession (700 μg g⁻¹ on the basis of ICP–AES data). However, the Zn concentrations in the seeds are surprisingly low and on parity with Cd concentrations in some of the seeds (with 700 μg g⁻¹ on the basis of ICP–AES data of the Ganges accession) or compared to Ni concentrations in the seeds from ultramafic accessions. This aligns with a study on *N. praecox* in which it was found that Cd was accumulated at higher concentrations in the seeds than Zn, while in the leaves Zn reaches 10 times the concentrations of Cd.⁶ The distribution of Ni and Zn in seeds was very similar, and neither element was translocated during germination. Zinc was highest in the interveinal areas of the cotyledons. The Fe maps were especially useful to obtain spatial reference within the seeds, as it clearly marked the provascular strands. Hence, the Fe distribution pattern, visible as a network of provascular strands in the hypocotyl, radicle, and cotyledons, as first reported in *Silene vulgaris*⁴⁸ and later in other species, including *Arabidopsis thaliana*⁴⁹ *Arabidopsis halleri*⁵⁰ and *Biscutella laevigata*, is also present in *N. caeruleus*. Nickel and Zn were enriched in the epidermal cells of the cotyledons increasing toward the cotyledon margins, and not in the mesophyll cells that surround the provascular strands. This strongly suggests that Ni and Zn are co-located in the epidermal cells (likely the vacuoles), as in mature plants.

In the seeds of *Noccaea pindic*, Ni is strongly enriched in the micropylar area near the radicle as well as in the cotyledon epidermis.⁵² In other hyperaccumulator plants, the cotyledons (in *Hybanthus floribundus* subsp. *adpressus*—Violaceae), the embryonic axis (in *Pimelea leptospermoidea*—Thymelaeaceae), and the pericarp (in *Stackhousia tryonii*—Celastraceae) were the main enrichment areas for Ni.⁵³,⁵⁴ In *Berkheya codii* (Asteraceae), Ni was in the micropylar area, at the bottom of the embryo, in the lower epidermis and the margins of the cotyledons. Nickel was localized largely in the cotyledons and translocated to the leaves after emergence, where Ni was accumulated in the leaf margins and in the leaf midrib.⁵ In *Odontarrhena corsica* (synonym *Alyssum corsicum*) and *Odontarrhena chalcidica* (synonym *Alyssum murale*) (both Brassicaceae), Ni predominately accumulated in the cotyledons as well as the hypocotyl, but it was comparatively low in the seed coat.⁵⁵ Thus, it seems that the results we report here for *N. caeruleus* align very well with what has been found for other,

---

Fig. 5 Elemental X-ray fluorescence microscopy (XFM) maps of living *Noccaea caeruleus* seedlings (Cira accession, ultramafic). The scan size is 21.5 × 89 mm (1448 × 1500 pixels). The elemental image was acquired in 5 μm step size with 1.7 ms dwell per pixel, 15.8 keV, incident beam, showing K, Ca, Fe, and Ni maps.
related, and non-related, hyperaccumulator species, with respect to localization of Ni and Zn in the epidermal cells of the cotyledons.

Conclusions

Most investigations employ X-ray microanalysis (SEM–energy-dispersive X-ray spectroscopy (EDS), micro-PIXE, XFM) on hyperaccumulator plants to obtain elemental distribution data to infer fundamental molecular mechanisms a posteriori (including the present study). Such approaches have yielded a wealth of knowledge about the ecophysiology of hyperaccumulator plants. X-ray microanalysis can also be used to elicit far more insightful information when employed on plant models a priori using mutants, including those in which genes encoding for metal transport proteins have been knocked out.\textsuperscript{38,36,57} The model species N. caerulescens is perfect for these lines of inquiry as it is a polynertallic (Cd–Ni–Zn–Pb) hyperaccumulator for which there is a molecular toolkit available to study and manipulate gene expression.\textsuperscript{32,16}

Author contributions

A.v.d.E. and G.E. collected the seeds in the field. A.v.d.E. prepared the samples for the synchrotron analysis. A.v.d.E., M.D.J., J.M.P., W.J.P., and H.H.H. conducted the synchrotron XFM experiments at the Australian Synchrotron. A.v.d.E. performed the SEM and light microscopy experiments. D.B. reconstructed the tomograms and created the 3D renderings. All authors contributed to writing the manuscript.

Supplementary material

Supplementary data are available at Metallomics online.

Conflicts of interest

The authors declare no conflicts of interest.

Acknowledgements

This research was undertaken at the X-Ray Fluorescence Microscopy beamline of the Australian Synchrotron (part of the Australian Nuclear Science and Technology Organisation (ANSTO)), Victoria, Australia. We thank Rachel Mak (University of Sydney) for support during and after the experiment at the XFM beamline. The Drishti visualization software developed by the National Computational Infrastructure’s VizLab (nci.org.au/drishti) was used for 3D renderings. This work was supported by the Multimodal Australian Sciences Imaging and Visualisation Environment (MASSIVE). We thank the Centre for Microscopy and Microanalysis, The University of Queensland, for support for the SEM analysis.

Funding

Antony van der Ent was the recipient of a Discovery Early Career Researcher Award (DE160100429) from the Australian Research Council. Hugh H. Harris acknowledges the Australian Research Council for financial support (DP140100176). Wojciech J. Przybyłowicz and Jolanta Mesjasz-Przybyłowicz are recipients of the South African National Foundation incentive grants nos. 114693 and 114694, respectively.

Data availability statement

The data underlying this article will be shared on reasonable request to the corresponding author.

Fig. 6 Elemental X-ray fluorescence microscopy (XFM) maps of living Noccaea caerulescens seedlings (Ganges accession, non-ultramafic, calamine). The scan size is 21.8 x 31 mm (2366 x 1562 pixels). The elemental image was acquired in 5 μm step size with 2.5 ms dwell per pixel, 15.8 keV, incident beam, showing K, Ca, Ni, and Zn maps.
References

1. A. J. Pollard, K. D. Powell, F. A. Harper and J. A. C. Smith, The Genetic Basis of Metal Hyperaccumulation in Plants, Crit. Rev. Plant Sci., 2002, 21, 539–566.

2. U. Krämer, Metal Hyperaccumulation in Plants, Annu. Rev. Plant Biol., 2010, 61, 517–534.

3. A. Van Der Ent, A. J. M. Baker, R. D. Reeves, A. J. Pollard and H. Schat, Hyperaccumulators of Metal and Metalloid Trace Elements: Facts and Fiction, Plant Soil, 2013, 362, 319–334.

4. M. Schnell Ramos, H. Khodja, V. Mary and S. Thomine, Using uPIXE for Quantitative Mapping of Metal Concentration in Arabidopsis thaliana Seeds, Front. Plant Sci., 2013, 4. doi: 10.3389/fpls.2013.00168.

5. S. Groeber, W. Przybyłowicz, G. Echevarria, E. Montarges-Pelletier, A. Barnabas and J. Mesjasz-Przybyłowicz, Fate of Nickel and Calcium in Seedlings of the Hyperaccumulator Berckheyaoddii during Germination, Biologia Plantarum, 2015, 59, 560–569.

6. K. Vogel-Mikuš, P. Pongrac, P. Kump, M. Nečernjer, J. Simčič, P. Pelicon, M. Budnar, B. Povh and M. Regvar, Localisation and Quantification of Elements within Seeds of Cd/Zn Hyperaccumulator Thlaspi praecox by Micro-PIXE, Environ. Pollut., 2007, 147, 50–59.

7. R. D. Reeves and R. R. Brooks, Hyperaccumulation of Lead and Zinc by Two Metallophytes from Mining Areas of Central-Europe, Environmental Pollution Series A: Ecological and Biological, 1983, 31, 277–285.

8. E. Lombi, F. J. Zhao, S. J. Dunham and S. P. Mcgrath, Cadmium Accumulation in Populations of Thlaspi caerulescens and Thlaspi goeingense, New Phytol., 2000, 145, 11–20.

9. K. Vogel-Mikuš, D. Drobne and M. Regvar, Zn, Cd and Pb Accumulation and Arbuscular Mycorrhizal Colonisation of Penny-Cress Thlaspi praecox Wulf. (Brassicaceae) from the Vicinity of a Lead Mine and Smelter in Slovenia, Environ. Pollut., 2005, 133, 233–242.

10. R. D. Reeves, C. Schwartz, J. L. Morel and J. Edmondson, Distribution and Metal-Accumulating Behavior of Thlaspi caerulescens and Associated Metallophytes in France, Int. J. Phytorem., 2001, 3, 145–172.

11. A. G. L. Assunção, W. M. Bookum, H. J. M. Nelissen, R. Vooijs, H. Schat and W. H. O. Ernst, Differential Metal-Specific Tolerance and Accumulation Patterns among Thlaspi caerulescens Populations Originating from Different Soil Types, New Phytol., 2003, 159, 411–419.

12. A. G. L. Assunção, H. Schat and M. G. M. Aarts, Thlaspi caerulescens, an Attractive Model Species to Study Heavy Metal Hyperaccumulation in Plants, New Phytol., 2003, 159, 351–360.

13. T. -H. -B. Deng, C. Cloquet, Y. -T. Tang, T. Sterckeman, G. Echevarria, N. Estrade, J.-L. Morel and R.-L. Qiu, Nickel and zinc Isotope Fractionation in Hyperaccumulating and Nonaccumulating Plants, Environ. Sci. Technol., 2014, 48, 11926–11933.

14. C. Gonneau, N. Genevois, H. Frérot, C. Sirguey and T. Sterckeman, Variation of Trace Metal Accumulation, Major Nutrient Uptake and Growth Parameters and Their Correlations in 22 Populations of Noccaea caerulescens, Plant Soil, 2014, 384, 271–287.

15. C. Gonneau, N. Noret, C. Godé, H. Frérot, C. Sirguey, T. Sterckeman and M. Pauwels, Demographic History of the Trace Metal Hyperaccumulator Noccaea caerulescens (Brassica nigra L.) in Western Europe, Mol. Ecol., 2017, 26 (3), 904–922.

16. C. F. Lin, E. I. Severing, B. Te Lintel Hekkert, E. Schijlen and M. G. M. Aarts., A Comprehensive Set of Transcript Sequences of the Heavy Metal Hyperaccumulator Noccaea caerulescens, Front. Plant Sci., 2014, 5, 261.

17. A. G. L. Assunção, P. D. C. Martins and S. De Folter et al., Elevated Expression of Metal Transporter Genes in Three Accessions of the Metal Hyperaccumulator Thlaspi caerulescens, Plant, Cell & Environment, 2001, 24, 217–226.

18. R. Oomen, J. Wu, F. Lelièvre, S. Blanchet, P. Richaud, H. Barbier-Brygoo, M. G. M. Aarts and S. Thomine, Functional Characterization of NRAMP3 and NRAMP4 from the Metal Hyperaccumulator Thlaspi caerulescens, New Phytol., 2009, 181, 637–650.

19. J. Wu, F.-J., Zhao, A. Ghandilyan, B. Logoteta, M. O. Guzman, H. Schat, X. Wang and M. G. M. Aarts, Identification and Functional Analysis of Two ZIP Metal Transporters of the Hyperaccumulator Thlaspi caerulescens, Plant Soil, 2009, 325, 79–95.

20. S. O. Lochlainn, H. C. Bowen, R. G. Fray, J. P. Hammond, G. J. King, P. J. White, N. S. Graham and M. R. Bradley, Tandem Duplication of HMA4 in the Zinc (Zn) and Cadmium (Cd) Hyperaccumulator Noccaea caerulescens (I Baxter, Ed.), PLoS One, 2011, 6, e17814–e17819.

21. D. Ueno, M. J. Milner, N. Yamaji, K. Yosoka, E. Koyama, M. Clémencia Zambra, M. Kaskie, S. Ebbs, L. V. Kochian and J. F. Ma, Elevated Expression of TCHMA3 Plays a Key Role in the Extreme Cd Tolerance in a Cd-Hyperaccumulating Ecotype of Thlaspi caerulescens, Plant J., 2011, 66, 852–862.

22. P. Halimaa and Y.-F. Lin and V. H. Ahonen et al., Gene Expression Differences between Noccaea caerulescens Ecotypes Help to Identify Candidate Genes for Metal Phytoremediation, Environ. Sci. Technol., 2014, 48, 3344–3353.

23. Ya-F Lin, Z. Hassan, S. Talukdar, H. Schat and M. G. M. Aarts, Expression of the ZNT1 Zinc Transporter from the Metal Hyperaccumulator Noccaea caerulescens Confers Enhanced Zinc and Cadmium Tolerance and Accumulation to Arabidopsis thaliana (I Baxter, Ed.), PLoS One, 2016, 11, e0149750–30.

24. D. Blande, P. Halimaa, A. I. Tervahauta, M. G. M. Aarts and S. O. Kärenlampi, De Novo Transcriptome Assemblies of Four Accessions of the Metal Hyperaccumulator Plant Noccaea caerulescens, Scientific Data, 2017, 4, 160131.

25. M. Hanikenne, I. N. Talke, M. J. Haydon, C. Lanz, A. Nolte, P. Motte, J. Kroymann, D. Weigel and U. Krämer, Evolution of Metal Hyperaccumulation Required cis-Regulatory Changes and Triplication of HMA4, Nature, 2008, 453, 391–395.

26. A. G. L. Assuncao, B. Pieper, J. Vromans, P. Lindhout, M. G. M. Aarts and H. Schat, Construction of a Genetic Linkage Map of Thlaspi caerulescens and Quantitative Trait Loci Analysis of Zinc Accumulation, New Phytol., 2006, 170, 21–32.

27. A. X. Deniau, B. Pieper, W. M. Ten Bookum, P. Lindhout, M. G. M. Aarts and H. Schat, QTL Analysis of Cadmium and Zinc Accumulation in the Heavy Metal Hyperaccumulator Thlaspi caerulescens, Theor. Appl. Genet., 2006, 113, 907–920.

28. K. H. Richau and H. Schat, Intraspecific Variation of Nickel and Zinc Accumulation and Tolerance in the Hyperaccumulator Thlaspi caerulescens, Plant Soil, 2009, 314, 253–262.

29. T. L. Turner, E. C. Bourne, E. J. Von Wettberg, T. T. Hu and S. V. Nuzhdin, Population Resequencing Reveals Local Adaptation of Arabidopsis lyrata to Serpentine Soils, Nat. Genet., 2010, 42, 260–263.

30. J. P. Xing, R. F. Jiang, D. Ueno, J. F. Ma, H. Schat, S. P. Mcgrath and F. J. Zhao, Variation in Root-to-Shoot Translocation of Cadmium and Zinc among Different Accessions of the Hyperaccumulators Thlaspi caerulescens and Thlaspi praecox, New Phytol., 2008, 178, 315–325.

31. G. Visioli, S. Vincenzi, M. Marmiroli and N. Marmiroli, Correlation between Phenotype and Proteome in the Ni Hyperaccumulator
Noccaea caerulescens Subsp. caerulescens, Environ. Exp. Bot., 2012, 77, 156–164.

32. A. D. Kozhevnikova, I. V. Seregin, N. T. Erlikh, T. A. Shevrevra, I. M. Andreev, R. Verweij and H. Schat, Histidine-Mediated Xylem Loading of Zinc is a Species-Wide Character in Noccaea caerulescens, New Phytol., 2014, 203, 508–519.

33. A. van der Ent, W. J. Przybylowicz and M. D. de Jonge et al., X-ray Elemental Mapping Techniques for Elucidating the Ecophysiology of Hyperaccumulator Plants, New Phytol., 2018, 218, 432–452.

34. P. M. Kopittke, T. Punshon, D. J. Paterson, R. V. Tapper, P. Wang, F. P. C. Blamey, A. Van Der Ent and E. Lombi, Synchrotron-Based X-ray Fluorescence Microscopy as a Technique for Imaging of Elements in Plants, Plant Physiol., 2018, 178, 507–523.

35. F. P. C. Blamey, B. A. McKenna and C. Li et al., Manganese Distribution and Speciation Help to Explain the Effects of Sulfate and Phosphate on Manganese Toxicity in Four Crop Species, New Phytol., 2017, 96, 37–15.

36. F. P. C. Blamey, D. J. Paterson, A. Walsh, N. Afshar, B. A. Mckenna, M. Cheng, C. Tang, W. J. Horst, N. W. Menzies and P. M. Kopittke, Time-Resolved X-ray Fluorescence Analysis of Element Distribution and Concentration in Living Plants: an Example Using Manganese Toxicity in Cowpea Leaves, Environ. Exp. Bot., 2018, 156, 151–160.

37. M. D. De Jonge and S. Vogt, Hard X-Ray Fluorescence Tomography—an Emerging Tool For Structural Visualization, Curr. Opin. Struct. Biol., 2010, 20, 606–614.

38. S. A. Kim, T. Punshon, A. Lanzirotti, L. Li, JoseM Alonso, J. R. Ecker, J. Kaplan and M. L. Guerrero, Localization of Iron in Arabidopsis Seed Requires the Vacular Membrane Transporter VIT1, Science, 2006, 314, 1295–1298.

39. A. L. D. Paul, H. H. Harris, P. D. Erskine, W Przybylowicz, J Mesjasz-Przybylowicz, G. Eccevarria and A. Van Der Ent, Synchrotron µXRF Imaging of Live Seedlings of Berkeyia codii and Odontarrhena muralis during Germination and Seedling Growth, Plant Soil, 2020, 453 (1), 487–501.

40. D. J. Paterson, M. D. De Jonge and D. L. Howard et al., The X-Ray Fluorescence Microscopy Beamline at the Australian Synchrotron, AIP Conf. Proc., 2011, 1365, 219.

41. R. Kirkham, P. A. Dunn and A. J. Kuczewski, The Maia Spectroscopy Detector System: Engineering for Integrated Pulse Capture, Low-Latency Scanning and Real-Time Processing, AIP Conference Proceedings, 2010, 1234, 204–243. doi: 10.1063/1.3463181.

42. D. P. Siddons, R. Kirkham and C. G. Ryan et al., 2014, Maia X-ray Microprobe Detector Array System, J. Phys. Conf. Ser., 2010, 499, 012001–012010.

43. C. G. Ryan and D. N. Jamieson, Dynamic Analysis: On-Line Quantitative PIXE Microanalysis and Its Use in Overlap-Resolved Elemental Mapping, Nucl. Instrum. Methods Phys. Res., Sect. B, 1993, 77, 203–214.

44. C. G. Ryan, Quantitative Trace Element Imaging Using PIXE and the Nuclear Microprobe, Int. J. Imaging Syst. Technol., 2000, 11 (4), 219–230.

45. C. G. Ryan, D. R. Cousins, S. H. Sie and W. L. Griffin, Quantitative Analysis of PIXE Spectra in Geoscience Applications, Nucl. Instrum. Methods Phys. Res., Sect. B, 1990, 49, 271–276.

46. C. G. Ryan, B. E. Etschmann, S. Vogt, J. Maser, C. L. Harland, E. Van Achterbergh and D. Legnini, Nuclear Microprobe-Synchrotron Synergy: Towards Integrated Quantitative Real-Time Elemental Imaging Using PIXE and SXRF, Nucl. Instrum. Methods Phys. Res., Sect. B, 2005, 231, 183–188.

47. A. Limaye, Drishti: a Volume Exploration and Presentation Toolset, SPIE 8506, Developments in X-Ray Tomography VIII, 85060X.

48. J. Mesjasz-Przybylowicz, K. Grodzińska, W. J. Przybylowicz, B. Godzik and S. Szarek-Lukaszewska 1999, Micro-PIXE Studies of Elemental Distribution in Seeds of Silene vulgaris from a Zinc Dump in Olkus, Southern Poland, Nucl. Instrum. Methods Phys. Res., Sect. B, 2012, 158, 306–311.

49. S. Eroglu, R. F. H. Giehl, B. Meier, M. Takahashi, Y. Terada, K. Ignatyev, E. Andersen, H. Küpper, E. Peiter and N. Von Wrixn, Metal Tolerance Protein 8 Mediates Manganese Homeostasis and Iron Reallocation During Seed Development and Germination, Plant Physiol., 2017, 174, 1633–1647.

50. A. Babst-Kostecka, W. J. Przybylowicz, B. Seget and J. Mesjasz-Przybylowicz, Zinc Allocation to and within Arabidopsis halleri Seeds: Different Strategies of Metal Homeostasis in Accessions under Divergent Selection Pressure, Plant-Environ. Interact., 2020, 1, 207–220.

51. A. Babst-Kostecka, W. J. Przybylowicz, A. Van Der Ent, C. Ryan, C. C. Dietrich and J. Mesjasz-Przybylowicz, Endosperm Prevents Toxic Amounts of Zn from Accumulating in the Seed Embryo—an Adaptation to Metaliferous Sites in Metal-Tolerant Biscutella laevigata, Metalomics, 2020, 12 (1), 42–53.

52. G. Psaras, Nickel Localization in Seeds of the Metal Hyperaccumulator Thlaspi pindicum Hausskn, Ann. Bot., 2001, 88, 513–516.

53. N. P. Bhatia, I. Orlic, R. Siegele, N. Ashwath, A. J. M. Baker and K. B. Walsh, Elemental Mapping Using PIXE Shows the Main Pathway of Nickel Movement Is Principally Symplastic within the Fruit of the Hyperaccumulator Stockhausia trionyi, Nucl. Physiol., 2003, 160, 479–488.

54. A. G. Kachenko, N. P. Bhatia and R. Siegele et al., Nickel, Zn and Cd Localisation in Seeds of Metal Hyperaccumulators Using µ-PIXE Spectroscopy, Nucl. Instrum. Methods Phys. Res., Sect. B, 2009, 267, 12–13.

55. T. Centofanti, R. V. Tapper, A. P. Davis and R. L. Chaney, Chelator-Buffered Nutrient Solution is Ineffective in Extracting Ni from Seeds of Alyssum, J. Plant Physiol., 2011, 13 (5), 434–440.

56. T. Punshon, M. L. Guerinot and A. Lanzirotti, Using Synchrotron X-Ray Fluorescence Microprobes in the Study of Metal Homeostasis in Plants, Ann. Bot., 2008, 103, 665–672.

57. T. Punshon, F. K. Ricachenewsky, M. N. Hindt, A. L. Socha and H. Zuber, Methodological Approaches for Using Synchrotron X-Ray Fluorescence (SXRF) Imaging as a Tool in Ionomics: Examples from Arabidopsis thaliana, Metalomics, 2013, 5, 1133–1113.