Brain metastases remain a major problem in patients with advanced non-small cell lung cancer (NSCLC). The permeability of the blood–brain barrier (BBB) is highly increased during lung cancer brain metastasis; however, the underlying mechanism remains largely unknown. We previously found that Inc-MMP2-2 is highly enriched in tumor growth factor (TGF)-β1-mediated exosomes and regulates the migration of lung cancer cells. This study aimed to explore the role of exosomal Inc-MMP2-2 in the regulation of BBB and NSCLC brain metastasis. Here, using endothelial monolayers and mouse models, we found that TGF-β1-mediated NSCLC-derived exosomes efficiently destroyed tight junctions and the integrity of these natural barriers. Overexpression of Inc-MMP2-2 in human brain microvascular endothelial cells increased vascular permeability in endothelial monolayers, whereas inhibition of Inc-MMP2-2 alleviated these effects. Furthermore, Inc-MMP2-2 knockdown markedly reduced NSCLC brain metastasis in vivo. Mechanistically, through luciferase reporter assays, RNA pull-down assay, and Ago2 RNA immunoprecipitation assay, we showed that Inc-MMP2-2 served as a microRNA sponge or a competing endogenous RNA for miR-1207-5p and consequently modulated the derepression of EPB41L5. In conclusion, TGF-β1-mediated exosomal Inc-MMP2-2 increases BBB permeability to promote NSCLC brain metastasis. Thus, exosomal Inc-MMP2-2 may be a potential biomarker and therapeutic target against lung cancer brain metastasis.

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INTRODUCTION
Lung cancer is the most common cancer in men and is associated with a high mortality rate worldwide [1]. Distant metastasis remains the primary cause of poor prognosis in lung cancer [2]. Brain metastases are a frequent complication in patients with advanced lung cancer and a major cause of mortality [3]. The blood–brain barrier (BBB) consists of endothelial cells (ECs), lined by pericytes, basement membrane, and astrocytes. This forms a tight barrier around blood vessels to restrict the flow of essential components in and out of the central nervous system (CNS) [4]. The integrity of the BBB is essential in preventing the invasion of tumor cells into the brain. In lung cancer, the BBB permeability is increased in brain metastases [5]. However, the mechanisms associated with the alterations in BBB permeability during lung cancer brain metastases are still unclear.

Exosomes have been recently a topic of interest owing to their role in multiple diseases [6–8]. A previous study has reported that exosomes are early contributors to the pathogenesis of different diseases [9, 10]. Tumor-derived exosomes, which are released from the primary tumor into the circulation, can travel to distant organs and modulate the microenvironment in premetastatic organs to facilitate future metastasis [11]. Tumor growth factor (TGF)-β1 is a multifunctional cytokine, having many biological activities, including regulating cell proliferation, differentiation, and apoptosis. We have previously reported that TGF-β1-mediated non-small cell lung cancer (NSCLC) cell exosomes might increase the permeability of lung vascular ECs and downregulate its tight junctions via lnc-MMP2-2 [12]. However, there is limited data on the significance of exosomal lnc-MMP2-2 in BBB and brain metastasis.

Previous studies have demonstrated that lncRNAs exhibit highly cell type-specific expression and function and have roles in carcinogenesis, metastasis, and drug resistance [13–15]. However, the role of lncRNAs in the regulation of BBB is yet to be fully explored. In this study we explored the role of exosomal lnc-MMP2-2 in the regulation of BBB and NSCLC brain metastasis. Through immunofluorescence (IF) and endothelial permeability assay we determined that Inc-MMP2-2 increases the permeability and destroys the integrity of BBB in vitro and in vivo. To identify the mechanisms by which Inc-MMP2-2 functioned in regulation of BBB permeability, luciferase reporter assays, RNA pull-down, and AGO2-RIP assay were performed and we found that Inc-MMP2-2 served as a microRNA sponge or a competing endogenous RNA (ceRNA) for miR-1207-5p. Furthermore, miR-1207-5p consequently modulated the derepression of EPB41L5. Additionally, we found that Inc-MMP2-2 knockdown drastically reduced NSCLC brain metastasis in mice. We aimed to elucidate the regulatory mechanism of
exosomal Inc-MMP2-2 in lung cancer brain pro-metastases and examine whether it could be a promising therapeutic target for patients with NSCLC.

MATERIALS AND METHODS

Reagents

Antibodies against N-cadherin (22018-1-AP), Zo-1 (21773-1-AP), occludin (13409-1-AP), CD63 (25682-1-AP), CD81 (27855-1-AP), Alix (12422-1-AP), GAPDH (iFluoro-Goat IgG), and anti-Ago2 (10686-1-AP) were purchased from Proteintech (Wuhan, China). Antibodies to claudin-5 (343214) and EPB41L5 (614203) were purchased from Zenbio (Guangzhou, China). Anti-VE-cadherin antibodies (2500) were obtained from CST (Beverly, MA, USA). Rhodamine B isoioxyanate-dextran was purchased from Sigma-Aldrich (St. Louis, MO, USA). Exo-Fect Exosome Transfection Kit was purchased from System Biosciences (Beijing, China). Recombinant human transforming growth factor β (TGF-β1) was purchased from PeproTech (Rocky Hill, USA).

Cell cultures and transfections

The NSCLC cell line A549 was obtained from Cell Bank of the Chinese Academy of Sciences (Shanghai, China). Non-fetal-derived human brain microvascular endothelial cells (HBMECs) were purchased from Bioloo Biotechnology (Shanghai, China). A549 and HBMECs were grown in Roswell Park Memorial Institute-1640 medium (Invitrogen, Carsbad, CA, USA) containing 10% fetal bovine serum at 37 °C with 5% CO2. The Inc-MMP2-2 overexpression and silencing lentivirus and their control lentiviruses were packaged by Genomeditech (Shanghai, China). EPB41L5-targeting siRNA and scramble control siRNA were obtained from Ribobio (Guangzhou, China). Two reporter plasmids or vector control and control agomir or mir-1207-5p agomir were co-transfected into HBMECs. The luciferase activity was measured as per our method previously described [12].

RNA fluorescence in situ hybridization and luciferase reporter assay

Fluorescence in situ hybridization (FISH) assay was performed according to a previous method [12]. The probe used for Inc-MMP2-2 was 5′-FAM-acagtacctgcct gccgcttgg-3′; shRNA#1, 5′-GCTGCAAGAACATCTTCCG-3′; shRNA#2, 5′-GGAATGACT CTGGTCCATC-3′; shRNA#3: 5′-GCCCTCTGCC AACATTCCCAAT-3′. Cell transduction was performed following the manufacturer’s instructions. Stable cells were selected using medium containing 0.5 μg/ml puromycin.

Detection analysis for BBB integrity in vivo

The BBB integrity was evaluated by using Evans blue (EB) and rhodamine B isothiocyanate-Dextran injection. EB (2% in PBS, 4 ml/kg) or rhodamine B (100 mg/kg) was injected into the mouse tail vein pretreated with exo or Texo for 24 h. After 3 h of circulation, the mice were anesthetized, and the heart was perfused. The mouse brains injected with rhodamine B were embedded into frozen blocks and sectioned into 15 μm by cryostat sectioning. The distribution of rhodamine B in the cerebra was observed through fluorescence microscopy. The EB fluorescence detection assay was performed according to a previous method [17].

Assembly of Inc-MMP2-2 lentivirus

The Inc-MMP2-2 overexpression, silencing lentivirus, and the empty vector were packaged by Genomeditech (Shanghai, China). The Inc-MMP2-2-silencing targeting sequences were as follows: shControl (NC), 5′-TT CTCCGAAAGTCTGCAGCAGT-3′; shRNA#1, 5′-GCTGCAAGAACATCTTCCG-3′; shRNA#2, 5′-GGAATGACT CTGGTCCATC-3′; shRNA#3: 5′-GCCCTCTGCC AACATTCCCAAT-3′. Cell transduction was performed following the manufacturer's instructions. Stable cells were selected using medium containing 0.5 μg/ml puromycin.

RNA immunoprecipitation

RNA immunoprecipitation (RIP) was performed using the EZMagna RIP kit (Millipore, Billerica, MA, USA) following the manufacturer’s protocol. Briefly, HBMECs were lysed in complete RIP lysis buffer, after which 100 μl of whole cell extract was incubated with RIP buffer containing magnetic beads conjugated with human anti-Ago2 antibody or control IgG. Samples were incubated with Proteinase K (1X), and then immunoprecipitated RNA was isolated. The RNA concentration was measured using a NanoDrop (Thermo Scientific). Moreover, purified RNA was subjected to qRT-PCR analysis to demonstrate the presence of the binding targets using respective primers.

Biotin pull-down assay

HBMECs were transfected with biotinylated wild-type (WT) mir-1207-5p, mutant mir-1207-5p (Genomeditech, Shanghai, China). First, cell lysates were harvested 48 h after transfection and incubated with Dynabeads M-280 Streptavidin (Invitrogen, Carlsbad, CA, USA) for 3 h at 4 °C according to the manufacturer’s protocol. Then, the beads were washed three times with ice-cold lysis buffer and once with high-salt buffer according to a previous method [18]. The bound RNAs were purified using a RNA extraction kit (Solarbio, Beijing, China) for the qRT-PCR analysis.

Mouse brain metastasis models

All experimental protocols were approved by the Laboratory Animal Ethical Committee at Chengdu Medical College. Female nude mice (5 weeks old, purchased from Chengdu Dossy Experimental Animals Co, Ltd, Sichuan, China) were injected intracardially with control A549, Inc-MMP2-2-knocked down A549, TGF-β1 pretreated (10 ng/ml for 24 h) control A549, or TGF-β1-pretreated Inc-MMP2-2-knocked down A549 cells (1 × 106 cells in 100 μL PBS) and were monitored for brain metastasis using an IVIS Imaging System. Approximately 3 weeks post-injection, all the mice were euthanized, and whole brains were collected. The presence of brain metastases was confirmed by fluorescence photography and hematoxylin-eosin staining.

Statistical analysis

Each in vitro experiment was performed independently at least three times. The results are presented as means ± SD. Student’s t tests was used for the comparison of two groups. One-way analysis of variance was conducted for comparison of multiple groups. All statistical analyses were performed with GraphPad Prism 5 (GraphPad Software, San Diego, CA, USA).
USA). Statistical significance was assigned at $p < 0.05$ (*), $p < 0.01$ (**), or $p < 0.001$ (***)..

**RESULTS**

**TGF-β1-mediated A549-derived exosomes promote EndoMT, downregulate the expression of tight junction proteins, and increase HBMECs monolayer permeability.**

Our previous studies have shown that TGF-β1-mediated A549 cell exosomes may increase lung vascular EC permeability and downregulate its tight junctions [12]. Figure 1A shows TEM images of exosomes secreted by A549 cells (exo) and of TGF-β1-pretreated (10 ng/ml, 24 h) A549 cells (Texo). The expression of exosomes characteristic proteins Alix, CD81, and CD63 are detected using western blotting. **D** Expression of EndoMT markers (VE-cadherin, N-cadherin) and tight junction proteins (ZO-1, occludin, and claudin-5) in exo- and Texo-treated HBMECs as measured using IF (scale bar, 50 μm) and western blotting assays. **F** Schematic representation of the Transwell chamber used for assaying transport across an endothelial monolayer. **G** The permeability of HBMECs (exo or Texo pre-cocultured) monolayer grown on 0.4-μm filters as measured according to the appearance of rhodamine B isothiocyanate-dextran, which was added in the upper well at the beginning of the experiment and in the bottom well after 2-h incubation. ****$p < 0.01$.  

Exosomes showed that no obvious difference exists between the two exosomes in appearance and size. Additionally, the exosomes characteristic proteins Alix, CD9, and CD63 detected using western blotting are shown in Fig. 1C. This analysis revealed that the size, concentration, and characteristic protein expression between exo and Texo were similar. IF staining and western blot analysis revealed that Texo, but not exo, downregulated expression of the endothelial marker VE-cadherin and upregulated expression of the mesenchymal marker N-cadherin. A decreased expression of tight junction proteins (ZO-1, occludin, and claudin-5) was also observed in HBMECs (Fig. 1D, E). We then performed an in vitro permeability assay by measuring the traversing of rhodamine-labeled dextran through HBMECs monolayers growing on 0.4-μm filters. The results showed that vascular EC permeability was markedly higher in Texo than that in exo (Fig. 1F, G).
TGF-β1-mediated A549-derived exosomes alter the integrity of BBB in vivo

Next, to further demonstrate the effect of Texo in vivo BBB, we injected Texo or exo into the tail vein of mice and examined the integrity of BBB after exosomes treatment (Fig. 2A). The in vivo integrity of BBB was determined according to the appearance of intravenously injected rhodamine-dextran and EB and the expression of tight junction proteins. Notably, Texo significantly enhanced the brain vascular permeability (Fig. 2B, C) and diminished ZO-1 and VE-cadherin expression in CD31+ ECs (Fig. 2D).

lnc-MMP2-2 inhibition in Texo reverses Texo-induced downregulation of tight junction proteins and HBMECs monolayer permeability

A previous study showed that exosomes play important roles as carriers of intercellular signals during vascular remodeling and cancer invasion [19]. Given the role of exosomal lnc-MMP2-2 in regulating HMVEC-L monolayer permeability [12], we explored its role in HBMECs. Exosomes in Texo were isolated, and lnc-MMP2-2 Smart Silencer were loaded in exosomes using Exo-Fect Exosome Transfection Kit. As expected, Texo, which was transfected with lnc-MMP2-2 Silencer, upregulated the expression of tight junction proteins in vitro and in vivo and attenuated the permeability of HBMECs monolayer (Fig. S1).

Inc-MMP2-2 promotes EndoMT, destroys tight junctions, and induces HBMECs monolayer permeability in vitro

To directly elucidate the biological roles of Inc-MMP2-2 in BBB, we first examined the distribution of Inc-MMP2-2 in HBMECs. The FISH assay showed that Inc-MMP2-2 was mainly expressed in the cytoplasm (Fig. 3A). Next, HBMECs were infected with Inc-MMP2-2 overexpression and silencing lentivirus. Stably infected cells were selected using puromycin, and Inc-MMP2-2 expression was confirmed using qRT-PCR (Fig. 3B, C). Based on these data, shlnc-MMP2-2 #3 was selected for all subsequent experiments. IF staining and western blotting showed that Inc-MMP2-2 overexpression markedly downregulated the levels of the endothelial marker VE-cadherin but upregulated those of mesenchymal markers (N-cadherin and the tight junction proteins ZO-1, claudin-5, and occludin; Fig. 3D, E). Moreover, permeability assays revealed that Inc-MMP2-2 overexpression significantly increased HBMECs monolayer permeability (Fig. 3F). Interestingly, Inc-MMP2-2 silencing showed the opposite effects (Fig. 3D–F).

Inc-MMP2-2 functions as a molecular sponge for miR-1207-5p in HBMECs

To identify the potential mechanisms by which Inc-MMP2-2 functioned in HBMECs, bioinformatic tools (microRNA.org and miRBase) were employed to analyze the potential targets of
The results revealed that miR-1207-5p had putative lnc-MMP2-2 binding sites (Fig. 4A). The qPCR assay showed that knockdown of lnc-MMP2-2 significantly increased miR-1207-5p expression, whereas ectopic lnc-MMP2-2 markedly inhibited miR-1207-5p expression (Fig. 4B, C). Interestingly, knockdown or overexpression of miR-1207-5p also affected lnc-MMP2-2 expression (Fig. 4D, E). Moreover, luciferase reporter assay revealed that miR-1207-5p overexpression lowered the luciferase activity of pGL3-lnc-MMP2-2-Wt but not that of pGL3-lnc-MMP2-2-mut (Fig. 4F).

In addition, the anti-Ago2 RIP assay further confirmed that miR-1207-5p was a target of lnc-MMP2-2 in HBMECs (Fig. 4G). Subsequently, biotin-labeled pull-down assays showed markedly higher lnc-MMP2-2 expression in HBMECs transfected with biotin-labeled miR-1207-5p than that in those labeled with the mutagenesis of the binding sites for lnc-MMP2-2 in miR-1207-5p (Fig. 4H). Collectively, these data demonstrated that lnc-MMP2-2 could directly bind to miR-1207-5p in HBMECs and showed a reciprocal repression of miR-1207-5p and lnc-MMP2-2.

MiR-1207-5p upregulates the expression of tight junction proteins and attenuates HBMECs monolayer permeability

Next, we confirmed the roles of miR-1207-5p in regulating HBMECs monolayer permeability. miR-1207-5p silencing and knockdown following transfection was confirmed via qRT-PCR (Fig. S2A). IF staining and western blotting showed that miR-1207-5p inhibition markedly lowered the levels of the endothelial marker VE-cadherin but upregulated those of the mesenchymal marker N-cadherin. It also downregulated the expression of the tight junction proteins ZO-1, claudin-5, and occludin (Fig. S2B, C).

Moreover, permeability assays revealed that miR-1207-5p inhibition significantly increased HBMECs monolayer permeability (Fig. S2D). As expected, miR-1207-5p overexpression showed the opposite effect (Fig. S2B–D).

EPB41L5 is a direct target of miR-1207-5p in HBMECs

To elucidate the underlying molecular mechanism by which miR-1207-5p exerts its effects on HBMECs, bioinformatic tools (microRNA.org and miRBase) were used for searching the candidate targets of miR-1207-5p. As shown in Fig. 5A, binding sequences of miR-1207-5p were identified in the 3′ untranslated region of EPB41L5 mRNA. Subsequent luciferase reporter assays, qRT-PCR, and western blot analysis revealed that miR-1207-5p could directly target EPB41L5 and negatively modulate the expression of EPB41L5 in HBMECs (Fig. 5B–D). In addition, we explored whether lnc-MMP2-2 could regulate the expression of EPB41L5. As shown in Fig. 5E, F, EPB41L5 expression could be positively regulated by lnc-MMP2-2 in HBMECs. This suggests that EPB41L5 is a direct target of miR-1207-5p and is positively modulated by lnc-MMP2-2 in HBMECs.

EPB41L5 promotes EndoMT, destroys tight junctions, and induces HBMECs monolayer permeability

EPB41L5 has been confirmed to play an oncogenic role in glioblastoma [20], gastric cancer [21], and breast cancers [22]. To explore whether EPB41L5 is also critical for BBB permeability, EPB41L5 overexpression or knockdown assays were performed in HBMECs. First, we identified an effective RNAi oligonucleotide to silence EPB41L5 expression in HBMECs (Fig. 6A, B).
Fig. 4  Inc-MMP2-2 functions as molecular sponge for miR-1207-5p in HBMECs. A The potential binding sites between miR-1207-5p and Inc-MMP2-2. B, C Stable Inc-MMP2-2-knocked down or -overexpressed HBMEC are subjected to qRT-PCR for analysis of miR-1207-5p expression. D, E Inc-MMP2-2 expression is decreased after transfection with miR-1207-5p inhibitor or agomir. F Complementary sequence between miR-1207-5p and wild-type (Wt) Inc-MMP2-2. The putative binding sites of miR-1207-5p is mutated in Inc-MMP2-2 vector. G Anti-Ago2 RIP is performed in HBMECs transiently overexpressing miR-1207-5p. H The sequences for Wt and Mt forms of miR-1207-5p are shown. lnc-MMP2-2 is highly enriched in the sample pulled down by biotinylated Wt miR-1207-5p but not in Mt miR-1207-5p. **p < 0.01, ***p < 0.001.

Fig. 5  EPB41L5 is a direct target of miR-1207-5p in HBMECs. A Bioinformatics analysis showed that miR-1207-5p could directly target the 3′-UTR of EPB41L5 wild-type (Wt). EPB41L5-mutant (Mt) means mutation of binding sites in the 3′-UTR of EPB41L5. B miR-1207-5p negatively regulates the luciferase activity of Wt EPB41L5, rather than that of Mt EPB41L5 in HBMECs. C, D miR-1207-5p inversely regulates the level of EPB41L5 mRNA and protein in HBMECs. E, F Inc-MMP2-2 positively regulates the abundance of EPB41L5 mRNA and protein in HBMECs. **p < 0.01, ***p < 0.001.
knockdown and overexpression were confirmed via qRT-PCR (Fig. 6C). Subsequent IF staining, western blotting, and monolayer permeability analysis revealed that EPB41L5 overexpression significantly promotes EndoMT, destroys tight junctions, and induces HBMECs monolayer permeability (Fig. 6D–F).

**Inc-MMP2-2 knockdown suppresses NSCLC brain metastasis in vivo**

To further elucidate the effect of Inc-MMP2-2 in NSCLC brain metastasis in vivo, a brain metastasis mouse model was established by implanting stably transduced A549 cells with control or shInc-MMP2-2 lentivirus treated with or without TGF-β1. Inc-MMP2-2 knockdown significantly inhibited the occurrence of brain metastasis (Fig. 7A–E).

**DISCUSSION**

BBB permeability is highly increased during lung cancer brain metastasis, but the mechanism still remains largely unknown. The current study found that TGF-β1-mediated NSCLC-derived exosomes promote EndoMT, downregulate the expression of tight junction proteins, increase HBMECs monolayer permeability, and alter the integrity of BBB in vivo. Inhibition of exosomal Inc-MMP2-2 reversed this effect. Inc-MMP2-2 overexpression and knockdown assays showed that Inc-MMP2-2 remarkably promotes EndoMT, destroys tight junctions, and induces HBMECs monolayer permeability.

Despite recent advances in treatment modalities, lung cancer remains the leading cause of cancer-related deaths worldwide, and metastasis remains the major determinant of poor outcomes [23]. A previous study showed that brain metastasis eventually occurs in ~40% of NSCLC patients, and 10% of NSCLC patients have brain metastasis at the time of diagnosis. The BBB is a barrier between blood circulation and brain tissue that provides anatomical and physiological protection for the CNS, supplies brain tissue with nutrients, and restricts the flow of essential components in and out of the CNS [24]. The BBB permeability is highly increased in lung cancer brain metastases [5], enabling the penetration of circulating tumor cells into the brain and promoting brain metastases. However, the mechanisms by which the BBB permeability is altered remain incompletely understood.

Human brain microvascular ECs is the main component of BBB. Thus, the cell morphology and tight junctions between HBMECs is essential for BBB permeability. EndoMT is a specific form of epithelial-to-mesenchymal transition characterized by a loss of endothelial features and the acquisition of mesenchymal features. Studies have shown that TGF-β1 is a potent inducer...
of EndoMT [25, 26]. TGF-β1-mediated IncRNAs have been shown to play important regulatory roles in cancer progression [27–29]. Interestingly, our previous study showed that TGF-β1-mediated exosomal Inc-MMP2-2 promotes lung cancer cell invasion into the vasculature by regulating the permeability of lung vascular ECs [12]. Thus, we hypothesized that TGF-β1-mediated exosomal Inc-MMP2-2 might be a regulator in HBMECs and eventually induce BBB dysfunction.

Fig. 7 Inc-MMP2-2 knockdown inhibits the NSCLC brain metastasis in vivo. A549 cells stably transfected with control and shInc-MMP2-2 lentivirus and treated with or without 10 ng/ml TGF-β1 for 24 h. All the cells are then injected into the heart of nude mice (N = 6). Three weeks after the injection, mice are photographed and sacrificed. A Representative IVIS imaging in brain metastasis mice. B, C Representative bioluminescent images of brain metastases and paired HE images (scale bar, 200 μm). D Brain tissues are subjected to double-label IF for analysis of ZO-1 (red) expression in CD31+ (green) endothelial cells (scale bar, 50 μm). E The average number of metastatic nodules in the brain metastasis mice. **p < 0.01. F Schematic of the proposed mechanism of action of the Inc-MMP2-2/miRNA-1207-5p/EPB41L5 axis. A detailed description is shown in the main text. sC: shControl, sM: shInc-MMP2-2, sC + T: shControl + TGF-β1, sM + T: shInc-MMP2-2 + TGF-β1.
One of the most well-known function of IncRNAs is that they function as competing endogenous RNAs (ceRNAs) to sponge microRNAs [30, 31]. The location of IncRNA is closely related to its function. For example, nuclear localization is related to its transcriptional modulator effect, and cytoplasmic localization can play the role of ceRNA [31]. We particularly detected Inc-MMP-2-2 location in HBMECs and found that IncMMP-2-2 is mainly expressed in the cytoplasm; this result corresponds to its ceRNA mechanism. Further, our bioinformatic analyses revealed that miR-1207-5p has putative Inc-MMP-2-2 binding sites. Inc-MMP-2-2 knockdown markedly upregulated miR-1207-5p expression, whereas Inc-MMP-2-2 overexpression downregulated miR-1207-5p expression. Moreover, luciferase reporter assay, biotin pull-down assay, and RIP revealed that miR-1207-5p was a target of Inc-MMP-2-2 in HBMECs. Thus, we considered that Inc-MMP-2-2 increases the permeability of vascular ECs in the BBB by downregulating miR-1207-5p expression.

EPB41L5 belongs to the NBL4 subgroup of the band 4.1 superfamily, which has a conserved 4.1-ezrin-radixin-moesin (FERM) domain at the N-terminus and a nonhomologous sequence at the C-terminus [20]. Recent studies confirmed that EPB41L5 is involved in the occurrence and development of squamous cell carcinoma [32], gastric cancer [21], and breast cancer [22]. In this study, EPB41L5 was identified as a direct target of miR-1207-5p in HBMECs. Notably, Inc-MMP-2-2 positively regulated EPB41L5 expression in HBMECs, while miR-1207-5p showed an opposite regulatory effect. Importantly, both ectopic EPB41L5 or MiR-1207-5p silencing in HBMECs directly promoted EndoMT, destroyed tight junctions, and induced HBMECs monolayer permeability.

Furthermore, this study has some limitations. Endothelial glyocalyx, a carbohydrate-rich layer that lines the luminal side of the vascular endothelium, is essential in vascular permeability and is regarded as the first BBB barrier. [33]. Numerous experiments have proved that the glyocalyx abscission is related to the pathophysiology of inflammatory diseases, atherosclerosis, and cancer metastases [34–36]. However, few studies have shown the effect of long non-coding RNA in regulating endothelial glyocalyx. Therefore, we only focused on the vascular EC without involving endothelial glyocalyx in this research. Hence, the role of Inc-MMP-2-2 in regulating endothelial glyocalyx remains unclear, which is our future study plan.

In conclusion, we identified a novel ceRNA regulatory pathway in which Inc-MMP-2-2 upregulates EPB41L5 expression by sponging miR-1207-5p. EPB41L5 directly promotes EndoMT, destroys tight junctions, increases BBB permeability, ultimately promoting brain metastasis in NSCLC (Fig. 7F). These findings support the potential of exosomal Inc-MMP-2-2 as a novel biomarker and therapeutic target against NSCLC brain metastasis.

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AUTHOR CONTRIBUTIONS
Conducted the study design: YX, DW, and SD; Carried out experiments and data analysis: DW and SD; Provided technical support and material: LL, TL, TZ, JL, and YY; Wrote the manuscript: DW and SD; Read and approved the final manuscript: all authors.

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COMPETING INTERESTS
The authors declare no competing interests.

ETHICS APPROVAL
The study was approved by the Ethics Committee of Chengdu Medical College.

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