The mechanism underlying the pubertal increase in pulsatile GnRH release in primates

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Abstract
In primates, the gonadotropin-releasing hormone (GnRH) neurosecretory system, consisting of GnRH, kisspeptin, and neurokinin B neurons, is active during the neonatal/early infantile period. During the late infantile period, however, activity of the GnRH neurosecretory system becomes minimal as a result of gonadal steroid independent central inhibition, and this suppressed GnRH neurosecretory state continues throughout the prepubertal period. At the initiation of puberty, the GnRH neurosecretory system becomes active again because of the decrease in central inhibition. During the progress of puberty, kisspeptin and neurokinin B signaling to GnRH neurons further increases, resulting in the release of gonadotropins and subsequent gonadal maturation, and hence puberty. This review further discusses potential substrates of central inhibition and subsequent pubertal modification of the GnRH neurosecretory system by the pubertal increase in steroid hormones, which ensures the regulation of adult reproductive function.

KEYWORDS
GnRH, human induced pluripotent stem cells, kisspeptin, NKB

1 | INTRODUCTION

Unlike the neurohypophysis, the adenohypophysis does not receive direct innervation from the hypothalamus. Yet, the hypothalamus influences the secretory activity of adenohypophyseal hormones luteinising hormone (LH) and follicle-stimulating hormone (FSH), which are indispensable for puberty onset and maintenance of reproductive function. As early as early last century, the concept that sexual maturation is controlled by the central nervous system has been proposed, as a result of a clinical observation showing that delayed puberty in patients with Frohlich’s syndrome is associated with damage of the ventral region of the brain.1 Three decades later, Hohlweg and Junkmann2 advanced the concept that a neural sex center is involved in controlling puberty and reproductive cyclicity. Meanwhile, during the 1940s to 1950s, Everett3 and Sawyer4 showed the influence of the hypothalamus on cyclic ovulation in rats with neuropharmacological tools in several publications. It was not, however, until 1955 that Harris5 showed clear evidence implicating the hypothalamus as central for the initiation of puberty. Harris and Jacobson6 demonstrated that either prepubertal pituitary glands transplanted into adult hypophysectomized rats or immature ovaries grafted into adult ovariectomized rats were able to sustain estrous cyclicity, whereas grafting adult pituitary gland or ovaries into sexually immature hypophysectomized or ovariectomized rats failed to maintain the estrous cyclicity. Finally, in 1971, two competitive research groups, comprising Amoss et al.7 and Matsuo et al.8 isolated and sequenced the mammalian form of the gonadotropin-releasing hormone (GnRH) peptide.

Puberty is one of the most prominent developmental events in life. During puberty, major hormonal, morphological, physiological, and behavioral changes occur such that after puberty the full capacity of reproductive function is established. Because puberty is a transitional period between childhood and adulthood, in primates it takes a
During the past 50 years, we have learned that the basic distribution pattern of GnRH neurons and pituitary-gonadal axis during the embryonic stage in rhesus monkeys and humans is already established at ED55–60. In infantile male monkeys, GnRH neurons are active because gonadotrophs in the pituitary are functional. This review summarizes what we know about the mechanism of puberty onset and demonstrates how activity of the GnRH neuron increases during the transitional period along with changes in upstream regulatory neuronal systems. This review summarizes what we know about the mechanism of the pubertal increase in GnRH release in primates.

## 2 | Ontogeny of GnRH Neurons and Changes in GnRH Release Pattern During Embryonic Development

Unlike most neurons in the brain, mammalian GnRH neurons originate from the nasal epithelium during the early gestational period and migrate into the brain, namely the preoptic area (POA) and medial basal hypothalamus (MBH). In the rhesus macaque, GnRH neurons are found in the nasal placode as early as embryonic day ED32 (E32) and commonly at ED34–36. GnRH neurons migrate along the nasal septum and then terminal nerve, enter the forebrain through the cribiform sieve at ED38 and subsequently migrate into the MBH by ED47. The basic distribution pattern of GnRH neurons in the brain is already established at ED55 (E55) and ED58 (E58) (Figure 1), although most of them originate in the olfactory pit at ED36. GnRH neurons in vitro derived from E36 rhesus macaque embryos dramatically increases after 3 weeks in culture, which is equivalent to approximately EDS6–57 in vivo. The negative feedback loop in the male hypothalamic-pituitary-gonadal axis is operative during the second trimester because gonadectomy in male monkeys at ED98–104, but not in female macaques, results in the elevation of LH and FSH.

A similar ontogenic profile of the reproductive neuroendocrine system has been also described in humans. In human fetuses, GnRH neurons are found in the nasal placode as early as embryonic week (EW) 5.5, although most of them originate in the olfactory pit at EW6.0–6.5. GnRH neurons enter the forebrain through the terminal nerve and then cribiform sieve by EW6.5, and they migrate into the hypothalamus by EW9.0. This is illustrated in Figure 1. FSH and LH are detectable in the pituitary by EW10, the pituitary starts to respond to GnRH and release gonadotropins into the general circulation by EW11–12, and their content increases until EW25–29. Circulating gonadotropins reach peak levels at mid-gestation, and both LH and FSH levels subsequently decline during late gestation.

### 3.1 Neonatal/infantile period

The direct measurement of GnRH release during the neonatal and early juvenile/prepubertal period has not been conducted. As such, GnRH neural activity in primates during this period is assessed by changes in circulating gonadotropins. In infantile male monkeys, GnRH neurons are active: (1) Circulating LH and testosterone are elevated during the first 2–3 postnatal months and (2) diurnal variations of testosterone are observed. Importantly, the diurnal variation in infantile males is quantitatively similar to that seen in sexually mature males. Plant shows that the hypothalamic-pituitary axis in males is already fully mature at the neonatal stage because bilateral orchidectomy at 1 week of age results in an increase in LH and FSH secretion, with a pulse pattern very similar to that in castrated adult males. By contrast, in female monkeys, circulating LH is only slightly elevated during the first 3 months. A moderate elevation of estrogen levels is observed during late gestation through the neonatal period in females, and ovariectomy in females at 1 week of age induces an attenuated and abbreviated elevation of LH release with slower pulse frequency compared to those in orchidectomized male infants. As such, in females, the negative feedback mechanism appears to be only partially operative in the late gestational period through the neonatal period. Presently, the precise underlying mechanisms of the pubertal increase in GnRH release in primates.
mechanism of the sex difference in activity of GnRH neurons during the neonatal period remains unclear. Both LH and FSH levels in male monkeys decrease after 3–4 months of age.  

Similar to the rhesus macaque, GnRH neurons in human male neonates are already active. LH levels abruptly increase within the first few minutes after birth, followed by an increase in serum concentrations of testosterone during the first 3–21 h. High levels of LH in the human male infant decline within 6 months and remain low until the time of puberty. FSH levels in human males are slightly elevated for the first three postnatal months, after which they become low. Circulating levels of testosterone are also elevated for 2–4 months postnatally. By contrast, in female neonates LH levels are only slightly elevated during the first few months of postnatal life, but FSH levels are high for the first 5 months. After the first 6 months of life, circulating levels of FSH, LH and gonadal steroids become low, and the hypothalamic-pituitary-gonadal system enters a quiescent stage until the time of puberty. Again, similar to that seen in macaques, the hypothalamic-pituitary suppression is attributable to non-gonadal origin because elevated concentrations of LH and FSH in infants with gonadal dysgenesis declines when they reach the juvenile period, as seen in eugonadal children. The transient increase in circulating gonadotropin and gonadal steroids during the neonatal period is also called “mini-puberty”. Physiological significance of mini-puberty in males is two fold: (1) transient elevation of testosterone would be important for development of male genitalia, such as penile and testicular growth as well as the proliferation of gonadic cells in humans and (2) neurobehavioral development in later human life.

### 3.2 | Juvenile/prepubertal period

Activity of the hypothalamic-pituitary-gonadal system is minimal during the prepubertal period. This state of quiescence, unique in primates, represents a period of non-gonadal inhibition upon GnRH release. Indeed, direct measurements of GnRH release in the hypothalamus indicate that during the entire prepubertal period both the pulse frequency and pulse amplitude of GnRH release are low until the time of puberty in both males and females and regardless of the
Importantly, the GnRH neurosecretory system in juvenile monkeys is gonadal steroid independent because developmental reduction in GnRH/LH release is observed in both male and female gonadectomized monkeys and administration of gonadal steroids do not alter either GnRH or gonadotropin levels. Furthermore, although the GnRH neurosecretory system does not respond to estradiol or androgens, it responds to various neurostimuli, such as electrical stimulation and NMDA challenge. Electrical stimulation of the basal hypothalamus in prepubertal female monkeys results in GnRH release as high as that seen in pubertal female monkeys and infusion of NMDA into the median eminence (ME) of prepubertal female monkey results in GnRH release. Importantly, long-term administration of NMDA at 3 h-intervals in prepubertal male monkeys results in precocious LH elevation followed by precocious puberty. Therefore, GnRH neurons are under the central inhibition during the prepubertal period. Presently, the precise mechanism of the central inhibition remains to be investigated. Current knowledge on this topic is discussed in a later section.

In prepubertal humans, both LH and FSH levels are very low. However, results of sensitive assays indicate that during the prepubertal period a minimum GnRH neuronal activity remains because release of LH and FSH is pulsatile and the nocturnal LH and FSH rhythm (i.e., evening levels are higher than those during daytime) is detected. In non-human primates, although nocturnal increases in GnRH release are seldom seen during the prepubertal period, it is readily observed during the early pubertal period.

### 3.3 Pubertal period

The first hormonal sign of puberty precedes the first physical sign of puberty. At an early phase of puberty (i.e., early pubertal stage), the basal level, pulse frequency and pulse amplitude of LH/FSH release all increase before nipple growth and menarche in girls. In rhesus females as well, the basal level, frequency, and amplitude of GnRH, LH, and FSH start to increase several months before menarche. Furthermore, it has been well documented that at an early stage of puberty, nocturnal increase in the LH pulse amplitude becomes prominent in both males and females in humans and rhesus monkeys. The pubertal increase in gonadotropin release leads to secretion of estradiol from the ovary and testosterone from the testis, resulting in secondary sex characteristics, such as nipple growth, sex-sk in development, and subsequently, menarche in females. In males, however, external signs of puberty (i.e., an increase in testicular size) are difficult to detect until the mid-pubertal stage.

During the mid-pubertal stage, increases in GnRH, LH and estradiol/testosterone accelerate. Although GnRH pulse frequency remains stable after the onset of puberty, the basal level and pulse amplitude of GnRH and LH continue to increase throughout puberty, until first ovulation in females and full spermatogenesis in males occur. During the mid-pubertal period the nocturnal increases in GnRH release become increasingly prominent. Importantly, a pubertal increase in GnRH release is independent from the presence or absence of gonadal steroids in circulation in both sexes because release of GnRH and LH in neonatally or prepubertally gonadectomized male and female monkeys also starts to increase at a similar age as gonadally intact counter parts.

Collectively, active GnRH neurons at the neonatal period are suppressed by central inhibition throughout the prepubertal period until the time of puberty, when a sustained increase in GnRH release, followed by elevated gonadotropin release occurs. The importance of the increased GnRH release at puberty onset is also experimentally shown. Then, what is the mechanism to initiate puberty?

### 4 NEUROENDOCRINE SIGNALING INITIATING THE PUBERTAL INCREASE IN GnRH RELEASE

Clinical studies in human genetics indicate that kisspeptin and neurokinin B (NKB) signaling play critical roles in puberty, as patients with mutations in the genes encoding KISS1 or NK3 and their receptors, KISS1R or NK3R, respectively, exhibit abnormal timing of puberty or no puberty. Subsequent studies in animal experiments suggest that (1) kisspeptin neurons express estrogen receptor alpha (ERα); (2) kisspeptin neurons in the anterior ventral nucleus (AVPV) innervate cell bodies of GnRH neurons, whereas kisspeptin neurons in the arcuate nucleus (ARC) innervate GnRH neuroprocesses; and (3) kisspeptin is an upstream regulator of basal as well as preovulatory GnRH release. Herde et al. renamed GnRH neuroprocesses in the ME as “dendrones” because these neuroprocesses have properties of both dendrites and axons. Moreover, because a subset of kisspeptin neurons in the ARC of the mouse and sheep co-expresses NKB and dynorphin, Goodman and colleagues have named them “KNΔy neurons”. In primates, however, only a small subset of kisspeptin neurons in the ARC co-express both NKB and dynorphin and co-expression of kisspeptin, NKB, and dynorphin within a single ARC neuron may not be critical for pulse-generation because interaction between the three peptides could occur at their neuroterminals. As such, in this review, we discuss these three types of neurons, as the independent units forming the regulatory network for GnRH release (Figure 3).

Our recent studies suggest that during the prepubertal period the release of NKB and kisspeptin neurons is also very low and release of both peptides increases along with the pubertal increase in GnRH release (Figure 2; also unpublished observations by JP Garcia and E Terasawa). Therefore, central inhibition governs the entire GnRH neurosecretory system and the removal of central inhibition is essential for the onset of puberty.

### 4.1 Central inhibition

Underlying mechanisms and neuronal substrates composing central inhibition are not well understood. Many years ago, we proposed the hypothesis that the GABA neuron is a part of the neural substrate of central inhibition. This hypothesis is based on the observations that...
GABA release in the ME in prepubertal monkeys was higher than that in early- and mid-pubertal monkeys; (2) infusion of the GABA<sub>α</sub> receptor blocker, bicuculline, into the ME stimulates GnRH release in prepubertal, but not pubertal monkeys; (3) infusion of antisense oligonucleotides to the rate limiting enzyme for GABA synthesis, GAD-67 and GAD-65 mRNAs, into the ME of prepubertal monkeys resulted in a large increase in GnRH release, whereas the same procedure in pubertal monkeys stimulated only a small increase in GnRH release; and finally (4) pulsatile infusion of bicuculline into the base of the third ventricle results in precocious menarche and first ovulation. More recent studies further indicate that infusion of bicuculline into the ME stimulates kisspeptin release and NKB release in prepubertal, but not pubertal monkeys. Importantly, the results from a follow-up study of the antisense mRNA infusion study that a decrease in GABA synthesis by interference with GAD67 synthesis followed by the reduction in GABA release triggers an increase in GnRH release accompanied with increase in glutamate release, suggesting the contribution of glutamate for the pubertal increase in GnRH release.

**FIGURE 3** Schematic illustration summarizing developmental changes in the neuroendocrine pathways involved in the mechanism of puberty onset. Possible interactions between kisspeptin (red), neurokinin B (NKB) (blue), opioid (gray), and gonatotropin-releasing hormone (GnRH) (black) neurons in the hypothalamus in prepubertal and pubertal male (A) and female (B) monkeys are shown. Although, in this schema kisspeptin, NKB, opioid neurons are all clustered in the arcuate nucleus (ARC), interaction between them takes place at the GnRH cell body levels as well as GnRH neuroterminals in the median eminence (ME). The number of colored dots at the neuroterminals reflects the estimated amount of neuropeptide release, based on the results from a series of experiments in this lab. A black X indicates signaling pathways are not operative. Note that major remodeling of kisspeptin and NKB signaling pathways takes place during puberty, such that their regulation of the GnRH neurosecretory system is most effective in adult reproductive function. Specifically, in prepubertal males, there is a reciprocal interaction between kisspeptin and NKB signaling (kisspeptin neurons mediate NKB signaling and NKB neurons mediate kisspeptin signaling) and both kisspeptin and NKB signaling modify the activity of GnRH neurons directly and independently. Perhaps the high activity of GnRH neurons during the neonatal period in males reflects the presence of reciprocal pathways in prepubertal males, although subsequent central inhibition during the prepubertal/juvenile keeps GnRH release low (top left). When males reach puberty, direct NKB signaling to GnRH neurons is lost, kisspeptin signaling through NKB neurons is no longer available, and NKB signaling to GnRH neurons needs to be mediated through kisspeptin neurons (top right). It is speculated that a simple NKB upstream signaling mechanism mediated through kisspeptin neurons is sufficient in the regulation of GnRH release in adult male reproductive function. By contrast, in prepubertal females, although both kisspeptin and NKB signaling can directly and independently influence activity of GnRH neurons, there is no reciprocal interaction between kisspeptin and NKB signaling (bottom left). In pubertal females, however, kisspeptin and NKB signaling not only influence GnRH neurons directly and independently, but also new reciprocal interactions between kisspeptin and NKB signaling are established (bottom right). These reciprocal and cooperative kisspeptin and NKB signaling pathways would provide more power and flexibility to regulate GnRH neurons in adult females, such that cyclic ovulations and pregnancy can be achieved. The role of opioid neurons is rather hypothetical at this point. Adapted and reproduced from Terasawa.
A similar prepubertal GABA dominant inhibitory mechanism appears to exist in male macaques because a recent study indicates that the tonic GABA inhibition over release of kisspeptin and NKB in the ME was observed in prepubertal, not pubertal, male rhesus monkeys. Collectively, it appears that GABA and glutamate neurons located up-stream of the kisspeptin and NKB neurons are a part of neuronal circuits involved in central inhibition.

Because the direct action of GABA is stimulatory for the GnRH neuron, clarification may be needed regarding the consistent findings on the inhibitory role of GABA in pubertal GnRH neurons. Although the stimulatory action of GABA on GnRH neurons is shown by recording from single cell studies with anatomically identified GnRH neurons, inhibitory action of GABA is derived from the direct measurement of GnRH release in the hypothalamus under the presence of GABA or GABA receptor agonists and antagonists in prepubertal macaques. As such, GABA action on GnRH release that we described is involved in multiple neural pathways. Importantly, the inhibitory role of GABA is seen in prepubertal but not pubertal monkeys, whereas excitatory action of GABA on mouse GnRH neurons occurs throughout the developmental stages, clearly suggesting that the neural pathway can be changed across puberty, but direct action of GABA is consistently stimulatory. Therefore, the difference between “inhibitory” versus “excitatory” role of GABA in GnRH neurons in the context stems from differences in experimental approaches.

In 2013, a striking report indicative of the mechanism of central inhibition was published. In their paper, Abreu et al. reported that human patients with frame shift mutations of MKRN3, the gene encoding makorin RING-finger protein 3, exhibit central precocious puberty and that MKRN3 mRNA expression in the ARC of male and female mice was higher on postnatal day (P)10 (P10) than P20. Importantly, the mutation in MKRN3 occurs at the zinc-finger domain, consistent with reports of gene-wide genome association studies regarding gene associated with the age of puberty.

Involvement of zinc-finger protein-mediated transcriptional repression of GnRH neurons as the central inhibition has also been reported in prepubertal monkeys. A more recent study by Abreu et al. further indicates that MKRN3 mRNA expression in the female rhesus monkey hypothalamus at less than 6 months of age were highest and gradually decreased at 6–12 months reaching the lowest at 12–30 months of age, and MKRN3 mRNA expression in the hypothalamus of both male and female mice was gonadal steroid independent because the hpg mouse that lacks active gonads exhibits a similar developmental decrease in MKRN3 mRNA expression in the whole hypothalamus. It was further shown that the developmental decrease in MKRN3 mRNA also occurs in the ARC and AVPV of mice in both sexes and that MKRN3 mRNA is colocalized in mice kisspeptin neurons in the ARC and AVPV. Finally, it was shown that MKRN3 inhibited Kiss1 and Tac3 promoter activity by repressing KISS1 and TAC3 transcription. Together, it was proposed the hypothesis that MKRN3 represses activity of kisspeptin and NKB neurons through an MKRN3-directed ubiquitination-mediated mechanism and developmental decrease in MKRN3 activity results in disinhibition of GnRH neurons, resulting in the pubertal increase in the GnRH peptide.

The study by Abreu et al. is one of the most impressive works in recent years. However, because the assessment of changes in MKRN3 mRNA in the monkey study was conducted with ill-defined developmental stages in females and Abreu et al. did not confirm changes in MKRN3 mRNA with developmental changes in hormones (LH, FSH, and gonadal steroids), it is difficult to apply the concept to monkey puberty at this time. For example, grouping monkeys at 0–6 months of age when the highest MKRN3 mRNA level seen means that this group includes the brains from monkeys at “neonatal mini-puberty” and the beginning of the prepubertal period. Similarly, grouping female monkeys at 12–30 months of age means females at prepubertal and early pubertal period are grouped together. An additional study with more precise age groups in both sexes independently with monitoring gonadotropin levels is clearly needed. This fundamental flaw aside, however, many questions arise. First, it is unclear whether the mechanism observed in mice is applicable to primate puberty, including humans: (1) the time course between birth and puberty onset in primates is much longer than in mice and (2) convincing evidence for central inhibition in the mouse has not been reported, and in the mouse, gonadal steroid sensitive GnRH suppression dominates prior to puberty onset; there is also a species difference issue. Second, as discussed earlier, at the neonatal period GnRH release and gonadotropin secretion in primates are elevated, resulting in “mini-puberty”. If MKRN3 mRNA levels are maximal elevated in animals at the neonatal period, how can high activity of GnRH neurons be achieved? Third, assuming a similar mechanism to that seen in mice can take place in primates, it is still unclear what triggers a decrease in MKRN3 mRNA in neurons of the ARC and AVPV? Fourth, does decrease in MKRN3 mRNA in the ARC and AVPV play a similar role in both sexes? Unlike that seen in rodents, there is little sex difference in the function of AVPV kisspeptin neurons in primates, although it is known that male puberty occurs much later than in female puberty in both species. Fifth, as discussed above, a series of studies consistently indicate that the GABA neuron is a part of neuronal circuits for central inhibition in non-human primates. Then, how do changes in GABA neurons at the onset of puberty relate to the developmental changes in the ubiquitin ligase MKRN3 in kisspeptin and NKB neurons? Presently, we have more questions than answers.

Note that there are consistent reports showing that neuropeptide Y (NPY) also plays a role in central inhibition in male monkeys. Because GABA and NPY are colocalized in ARC neurons, there is a possible common mechanism between GABA and NPY in central inhibition. This hypothesis needs to be tested in a series of future studies.

4.2 Activity of kisspeptin and NKB neurons increases at puberty onset

After the reduction of central inhibition, activity of kisspeptin and NKB signaling over GnRH neurons increases. We have systematically
examined the manner in which kisspeptin and NKB signaling stimulates the pubertal increase in GnRH release in non-human primates. As described below, kisspeptin and NKB signaling accelerates the pubertal increase in GnRH release through two mechanisms: (1) kisspeptin and NKB neurons release a higher amount of respective peptides and (2) an increased sensitivity of KISS1R on GnRH neurons.

4.2.1 | Kisspeptin release

Kisspeptin release in the ME is pulsatile and the pulse frequency is similar to that in GnRH release. Indeed, kisspeptin pulses are highly correlated to GnRH pulses.\textsuperscript{101} There is a clear developmental change in kisspeptin release: Although, in prepubertal female monkeys, the mean release, pulse frequency, and pulse amplitude of kisspeptin release are all low, in pubertal females, all components of kisspeptin release significantly increase\textsuperscript{53} (Figure 2). Importantly, in an early phase after puberty onset, the pubertal increase in kisspeptin release is gonadal steroid independent because an increase in kisspeptin release also occurs in gonadectomized animals,\textsuperscript{53} but the later phase of the kisspeptin increase is partly a result of the pubertal increase in circulating gonadal steroid hormones.\textsuperscript{53,102} In males, kisspeptin release also gradually increases after puberty onset.\textsuperscript{103}

4.2.2 | NKB release

NKB release in the ME of prepubertal males is low and it increases at puberty onset. Again, the pubertal increase in NKB release in males is gonadal steroid independent (JP Garcia and E Terasawa, unpublished observations). Whether the pubertal increase in NKB release occurs in females has not been examined. As seen in kisspeptin release, we assume that NKB release is pulsatile, although we have not confirmed this systematically yet.

4.3 | Changes in the sensitivity of GnRH neurons to kisspeptin and NKB signaling

In female monkeys GnRH neurons respond to both kisspeptin (kisspeptin10) and NKB (senktide) signaling in a dose-responsive manner.\textsuperscript{53,104} Because the GnRH response to the same dose of kisspeptin10 in pubertal females is larger than in prepubertal females, the sensitivity of GnRH neurons to kisspeptin signaling clearly increases after puberty onset. Importantly, the pubertal increase in responsiveness of GnRH neurons to kisspeptin signaling is a result of the pubertal increase in circulating gonadal steroids.\textsuperscript{104} By contrast, in males, although both kisspeptin and NKB are stimulatory to GnRH release and the response of GnRH neurons to kisspeptin (kisspeptin10) signaling is dose-dependent, the GnRH response to NKB signaling is not dose-dependent.\textsuperscript{54,102,103} Moreover, in male monkeys, developmental amplification of the GnRH response to kisspeptin10 is seen only at a higher dose.\textsuperscript{103} Interestingly, GnRH neurons in females are more sensitive to kisspeptin signaling than in males because a 1/100-fold smaller dose of kisspeptin induces a larger response in females than in males at both the prepubertal and pubertal stages.\textsuperscript{102,103} The underlying mechanism of the sex difference in GnRH response to kisspeptin signaling is currently unknown.

4.4 | Kisspeptin and NKB signaling pathways to GnRH neurons undergo pubertal change

Although it is well documented that GnRH neurons express kisspeptin receptors\textsuperscript{105,106} and kisspeptin directly modulates GnRH release, the direct action of NKB on GnRH release remains controversial. NKB neurons, however, can modulate GnRH neurons directly through dendrites because GnRH neuroterminal fibers in the ME express NK3R,\textsuperscript{107} and a study using fast scan cyclic voltammetry with a carbon-fiber microelectrode indicates that the direct action of the NK3R agonist senktide on GnRH neurons occurs in the neuroterminal region, but not in the POA.\textsuperscript{108} Therefore, microdialysis experiments examining interactions between kisspeptin and NKB signaling to GnRH release conducted in the ME in our laboratory are physiologically relevant.

To clarify the hierarchy between kisspeptin and NKB signaling that influences activity of GnRH neurons, we have conducted a series of experiments using agonists and antagonists for KISS1R and NK3R. Through such experiments, we have observed quite surprising results: Kisspeptin and NKB signaling pathways to GnRH neurons undergo pubertal change in both males and females, and the direction of the pathway changes in the two sexes is almost opposite. Because detailed experimental designs and results have been published previously,\textsuperscript{103,104,109} in this review, we just summarize the results and interpretations. In prepubertal males, there is a reciprocal interaction between kisspeptin and NKB signaling (kisspeptin neurons mediate NKB signaling to GnRH neurons needs to be mediated through kisspeptin signaling) and both kisspeptin and NKB signaling modify the activity of GnRH neurons directly and independently. We speculate that the powerful reciprocal pathways in prepubertal males might reflect the high activity of GnRH neurons during the neonatal period in males but subsequent Central Inhibition during the prepubertal/early stages keeps GnRH release low (Figure 3, top left). When males reach puberty (i.e., in pubertal males), direct NKB signaling to GnRH neurons is lost, kisspeptin signaling through NKB neurons is no longer available, and NKB signaling to GnRH neurons needs to be mediated through kisspeptin neurons (Figure 3, top right). The upstream regulation of NKB signaling through kisspeptin neurons has also been reported in male monkeys.\textsuperscript{110} Perhaps a simple mechanism underlying NKB signaling through kisspeptin neurons is sufficient in the regulation of GnRH release, and hence male reproductive function in adulthood. By contrast, in prepubertal females, although both kisspeptin and NKB signaling can directly and independently influence activity of GnRH neurons, there is no reciprocal interaction between kisspeptin and NKB signaling (Figure 3, bottom left). In pubertal females, however, kisspeptin and NKB signaling not only influences
GnRH neurons directly and independently, but also new reciprocal interactions between kisspeptin and NKB signaling are established (Figure 3, bottom right). These reciprocal and cooperative kisspeptin and NKB signaling pathways would provide more power and flexibility to regulate GnRH neurons in adult females, such that cyclic ovulations and pregnancy can be achieved.

In summary, we now know that during the postnatal development, a sustained elevation of pulsatile GnRH release results in puberty in both males and female primates. Indeed, transient elevation of the GnRH release does not lead to puberty. As described earlier, the GnRH neurosecretory system in males shortly after birth is fully active, similar to that observed in adulthood, but, at the late neonatal period, central inhibition comes in, suppressing the elevated GnRH neurosecretory activity. In females as well, a similar, but smaller degree of the transient GnRH elevation is seen during the early neonatal period, and central inhibition comes in, suppressing the GnRH neurosecretory system. Although the neural substrates responsible for central inhibition remain unclear, the suppression of GnRH release continues throughout the prepubertal period. It is considered that central inhibition for reproductive function provides the time for maturation of higher brain function, including intellectual, judgement, memory function, and sensory-motor function.

Recent studies investigating the role of the KNDy neurons forming the GnRH neurosecretory system further suggest that the reduction in central inhibition comes first. During the early stage of the pubertal period, active interaction is observed between kisspeptin and NKB signaling to GnRH neurons, which is reminiscent of the very active GnRH neurosecretory system during the neonatal period in males, whereas, during the late pubertal period, a simple NKB dominant pathway to GnRH neurons (i.e., NKB signaling → kisspeptin signaling → GnRH neurons) is established, such that male adult reproductive function does not require complex neuroendocrine mechanisms. By contrast, whereas, during the early stage of pubertal period in females, there is no cooperative interaction between kisspeptin and NKB signaling to GnRH neurons, during the late pubertal period to adulthood, a flexible and cooperative relationship between kisspeptin and NKB signaling is established to be suitable for adult female reproductive function. Therefore, the pubertal period can be redefined as the time when the GnRH neurosecretory system is reorganized/remodeled to tailor the need for adult reproductive function.

Finally, brief comments on the role of opioid neurons including dynorphin neurons in the pubertal increase in pulsatile GnRH. We already know that opioid neurons are not involved in central inhibition, whereas the preliminary data indicate that release of β-endorphin increases after puberty onset. Therefore, we speculate that opioid peptides play an important role for the pubertal increase in pulsatile GnRH release, as a part of the break. Indeed, a recent report showed that tonic infusion of the opioid antagonist naloxone increased the LH pulse frequency in patients with deficiency of NKB signaling genes, indicating the role of opioid neurons in GnRH pulse generation. Future studies warrant clarification of the role of opioid neurons in the pubertal increase in GnRH release.

5 | CONCLUSIONS AND FUTURE PERSPECTIVES

Since the discovery of the GnRH molecule in 1971, great advancements have been made in neuroendocrine research. Specifically, the isolation and molecular identification of the GnRH molecule from the hypothalamus, followed by the discovery of many other small peptides including kisspeptin, NKB, and several opioids in the brain, have played a great role. Although, in this article the author did not discuss non-peptidergic neurotransmitters, such as acetylcholine, serotonin, dopamine, epinephrine, norepinephrine, and nitric oxide, they are also known to modulate GnRH neuronal activity directly or indirectly. Nevertheless, the concept that an increase in pulsatile GnRH release initiates puberty, and the pubertal increase in GnRH release is accompanied by the increased activity of kisspeptin and NKB neurons, is firmly established. We now know that the presence and proper connections between GnRH, kisspeptin and NKB neurons in the hypothalamus are all necessary for puberty onset and subsequent maintenance of normal reproduction functions. Neural substrates and mechanisms of central inhibition remain to be investigated, but the availability of various in vivo and in vitro models including GnRH neurons derived from human stem cells would provide a bright future for research in GnRH neurobiology.

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CONFLICTS OF INTEREST

The author declares that she has no conflicts of interest.

AUTHOR CONTRIBUTIONS

The manuscript has been designed and written by Ei Terasawa.

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DATA AVAILABILITY STATEMENT

Data sharing is not applicable to this article as no new data were created or analyzed in this study.

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REFERENCES

1. Erdheim J. Über hypophysengangs geschwulste und hirncholesteatome. Sitzungsb Kais Akad Wissen Math Naturw Klin. 1904;113:537-726.
2. Hohlweg W, Junkmann K. Die hormonal-nervose regulierung der funktion des hypophysenvorderlappens. Klin Wochenschr. 1932;11:321-323.
3. Everett JW. Neurobiology of reproduction in the female rat. A fifty-year perspective. Monogr Endocrinol. 1989;32:1-133.
4. Sawyer CH. First Geoffrey Harris Memorial lecture. Some recent developments in brain-pituitary-ovarian physiology. Neuroendocrinology. 1975;17:97-124.
5. Harris GW. Neural Control of the Pituitary Gland. London, UK: Edward Arnold; 1955.
6. Harris GW, Jacobson D. Functional grafts of the anterior pituitary gland. Proc R Soc London B Biol Sci. 1952;139:263-276.
7. Amoss M, Burgess R, Blackwell P, Vale W, Fellows R, Guillemin R. Purification amino acid composition and N-terminus of the hypothalamic luteinizing hormone releasing factor (LRF) of ovine origin. Biochem Biophys Res Commun. 1971;44:205-210.
8. Matsuo H, Baba Y, Nair RMG, Arimura A, Schally AV. Structure and function of the hypothalamus. Proc R Soc London B: Biol Sci. 1989;233:119-133.
9. Hamilton AS, Mack TM. Puberty and genetic susceptibility to breast cancer in a case-control study in twins. N Engl J Med. 2003;348:2313-2322.
10. Wardle J, Brodersen NH, Cole TJ, Jarvis MJ, Boniface DR. Development of adiposity in adolescence: five year longitudinal study of an ethnically and socioeconomically diverse sample of young people in Britain. BMJ. 2006;332:1130-1135.
11. Jasik CB, Lustig RH. Adolescent obesity and puberty: the “perfect storm”. Ann N Y Acad Sci. 2008;1135:265-279.
12. Dwyer AA. Psychosexual effects resulting from delayed, incomplete, or absent puberty. Curr Opin Endocrinol. 2020;14:15-21.
13. Schwanzel-Fukuda M, Pfaff DW. Origin of luteinizing hormone-releasing hormone neurons. Nature. 1989;338:161-164.
14. Wray S, Grant P, Gainer H. Evidence that cells expressing luteinizing hormone-releasing hormone mRNA in the mouse are derived from progenitor cells in the olfactory placode. Proc Natl Acad Sci U S A. 1989;86:8132-8136.
15. Ronneklev OK, Resko JA. Ontogeny of gonadotropin releasing hormone-containing neurons in early fetal development of rhesus macaques. Endocrinology. 1990;126:498-511.
16. Quanbeck C, Sherwood NM, Millar RP, Terasawa E. Two populations of luteinizing hormone-releasing hormone neurons in the forebrain of the rhesus macaque during embryonic development. J Comp Neurol. 1997;380:293-309.
17. Terasawa E, Quanbeck CD, Schulz CA, Burich AJ, Luchansky LL, Claude P. A primary cell culture system of luteinizing hormone releasing hormone neurons derived from embryonic olfactory placode in the rhesus monkey. Endocrinology. 1993;133:2379-2390.
18. Terasawa E, Keen KL, Mogi K, Claude P. Pulsatile release of luteinizing hormone-releasing hormone (LHRH) in cultured LHRH neurons derived from the embryonic olfactory placode of the rhesus monkey. Endocrinology. 1999;140:1432-1441.
19. Terasawa E, Busser BW, Luchansky LL, et al. Presence of luteinizing hormone-releasing hormone fragments in the rhesus monkey forebrain. J Comp Neurol. 2001;439:491-504.
20. Resko JA, Ellinwood WE, Pasztor LM, Buhl AE. Sex steroids in the umbilical circulation of fetal rhesus monkeys from the time of gonadal differentiation. J Clin Endocrinol Metab. 1980;50:900-905.
21. Kurian JR, Keen KL, Terasawa E. Epigenetic changes coincide with in vitro primate GnRH neuronal maturation. Endocrinology. 2010;151:5359-5368.
22. Resko JA, Ellinwood EE. Negative feedback regulation of gonadotropin secretion by androgens in fetal rhesus macaques. Biol Reprod. 1985;3:346-352.
23. Grumbach MM, Kaplan SL. The neuroendocrinology of human puberty, an ontogenetic perspective. In: Grumbach MM, Sizonenko PC, Aubert ML, eds. Control of the Onset of Puberty. Philadelphia, PA: Williams & Wilkins; 1990:1-62.
24. Schwanzel-Fukuda M, Crossin KL, Pfaff DW, Bouloux PM, Hardelin JP, Petit C. Migration of luteinizing hormone-releasing hormone (LHRH) neurons in early human embryos. J Comp Neurol. 1996;366:547-557.
25. Verney C, al Amraoui A, Zecевич N. Conimigration of tyrosine hydroxylase- and gonadotropin-releasing hormone-immunoreactive neurons in the nasal area of human embryos. Brain Res Dev Brain Res. 1996;97:251-259.
26. Grumbach MM, Roth JC, Kaplan SL, Kelch RP. Hypothalamic-pituitary regulation of puberty in man: evidence and concepts derived from clinical research. In: Grumbach MM, Grave GD, Mayer FE, eds. Control of the Onset of Puberty. New York, NY: John Wiley & Sons; 1974:115-166.
27. Kaplan SL, Grumbach MM, Aubert ML. The ontogenesis of pituitary hormones and hypothalamic factors in the human fetus: maturation of central nervous system regulation of anterior pituitary function. Recent Prog Horm Res. 1976;32:161-243.
28. Clements JA, Reyes F, Winter JS, Faiman C. Studies on human sexual development. Ill. Fetal pituitary and serum, and amniotic fluid concentrations of LH, CG, and FSH. J Clin Endocrinol Metab. 1976;42:9-19.
29. Winter JSD, Faiman C, Reyes F. Sex steroid production by the human fetus: its role in morphogenesis and control by gonadotropins. Birth Defects Orig Artic Ser. 1977;13:41-58.
30. Castillo RH, Matteri RL, Dumesic DA. Luteinizing hormone synthesis in cultured fetal human pituitary cells exposed to gonadotropin-releasing hormone. J Clin Endocrinol Metab. 1992;75:318-322.
31. Styne DM, Grumbach MM. Puberty, ontology, neuroendocrinology, physiology, and disorders. In: Kroenenberg H, Melmed S, Polonsky KS, Larsen PR, eds. Williams Textbook of Endocrinology. 11th ed. Philadelphia, PA: Saunders Elsevier. 2008:970-1166.
32. Frawley LS, Neill JD. Age related changes in serum levels of gonadotropins and testosterone in infantile male rhesus monkeys. Biol Reprod. 1979;20:1147-1151.
33. Robinson JA, Bridson WE. Neonatal hormone patterns in the macaque. I. Steroids. Biol Reprod. 1978;19:773-778.
34. Plant TM. Neuroendocrine basis of puberty in the rhesus monkey (Macaca mulatta). In: Martini L, Ganong WF, eds. Frontiers in Neuroendocrinology. Raven Press; 1988:215-238.
35. Plant TM. Pulsatile luteinizing hormone secretion in the neonatal male rhesus monkey (Macaca mulatta). J Endocrinol. 1982;93:71-74.
36. Plant TM. A striking diurnal variation in plasma testosterone concentrations in infantile male rhesus monkeys (Macaca mulatta). Neuroendocrinology. 1982;35:370-373.
37. Pohl CR, de Ridder CM, Plant TM. Gonadal and nongonadal mechanisms contribute to the prepubertal hiatus in gonadotropin secretion in the female rhesus monkey (Macaca mulatta). J Clin Endocrinol Metab. 1995;80:2094-2101.
38. Plant TM. A striking sex difference in the gonadotropin response to gonadectomy during infantile development in the rhesus monkey (Macaca mulatta). Endocrinology. 1986;119:539-545.
39. Steiner RA, Bremner WJ. Endocrine correlates of sexual development in the male monkey, Macaca fascicularis. Endocrinology. 1981;90:191-919.
40. Plant TM. A study of the role of the postnatal testes in determining the ontogeny of gonadotropin secretion in the male rhesus monkey (Macaca mulatta). Endocrinology. 1985;116:1341-1350.
41. Corbier P, Dehennin M, Castanier A, Mebazaa A, Edwards DA, Roffi J. Sex differences in serum luteinizing hormone and testosterone in the human neonate during the first few hours after birth. J Clin Endocrinol Metab. 1990;71:1344-1348.

42. Faiman C, Winter JS. Gonadotropins and sex hormone patterns in puberty, clinical data. In: Grumbach MM, Grave GD, Mayer FE, eds. Control of the Onset of Puberty. New York, NY: Wiley; 1974:32-55.

43. Winter JS, Faiman C. Serum gonadotropin concentrations in agonal children and adults. J Clin Endocrinol Metab. 1972:35:561-564.

44. Andersson AM, Toppari J, Haavisto AM, et al. Longitudinal reproductive hormone profiles in infants: peak of inhibin B levels in infant boys exceeds levels in adult men. J Clin Endocrinol Metab. 1998;83:675-681.

45. Winter JS, Faiman C, Hobson WC, Prasad AV, Reyes FI. Pituitary-gonadal relations in infancy. I. Patterns of serum gonadotropin concentrations from birth to four years of age in man and chimpanzee. J Clin Endocrinol Metab. 1975:40:545-551.

46. Conte FA, Grumbach MM, Kaplan SL. A diphasic pattern of gonadotropin secretion in prepubertal, but not prepubertal, ovariectomized female rhesus monkeys. J Clin Endocrinol Metab. 1998;83:57:288-893.

47. Kuiri-Hanninen T, Sankilampi U, Dunkel U. Activation of hypothalamic-pituitary-gonadal axis in infancy. Horm Res Paediatr. 1999;51:156-164.

48. Wiltz L, Marshall G, Knoobl E. Experimental induction of puberty in the infantile female rhesus monkey. Science. 1980;207:1373-1375.

49. Seminara SB, Messager S, Chatzidaki EE, et al. The GPR54 gene as a key. Endocrinology. 2003;148:5752-5760.

50. Oakley AE, Clifton DK, Steiner RA. Kisspeptin signaling in the brain. Endocr Rev. 2009;30:713-743.

51. Garcia JP, Guerriero KA, Keen KL, Kenealy BP, Seminara SB, Terasawa E. Kisspeptin and neurokinin B signaling network underlies the pubertal increase in GnRH release in female rhesus monkeys. Endocrinology. 2017;158:3269-3280.

52. Chongthammakun S, Claypool LE, Terasawa E. Ovarioctomy increases in vivo luteinizing hormone-releasing hormone release in pubertal, but not prepubertal, female rhesus monkeys. J Neuroendocrinol. 1995;5:41-50.

53. Guerriero KA, Keen KL, Terasawa E. Experimental increase in kisspeptin-54 release in vivo is independent of the pubertal increase in estradiol in female rhesus monkeys (Macaca mulatta). Endocrinology. 2012;153:1887-1897.

54. Garcia JP, Guerriero KA, Keen KL, Kenealy BP, Seminara SB, Terasawa E. Kisspeptin and neurokinin B signaling network underlies the pubertal increase in GnRH release in female rhesus monkeys. Endocrinology. 2017;158:3269-3280.

55. Chongthammakun S, Terasawa E. Negative feedback effects of estrogen on luteinizing hormone-releasing hormone release occur in pubertal, but not prepubertal, ovariectomized female rhesus monkeys. Endocrinology. 1993;132:735-743.

56. Claypool LE, Watanabe G, Terasawa E. Effects of electrical stimulation of the medial basal hypothalamus on the in vivo release of luteinizing hormone-releasing hormone in the pubertal and peripubertal female rhesus monkey. Endocrinology. 1990;127:3014-3022.

57. Claypool LE, Kasuya E, Saitoh Y, Marzban F, Terasawa E. N-methyl D,L-aspartate induces the release of luteinizing hormone-releasing hormone in the pubertal and pubertal female rhesus monkey as measured by in vivo push-pull perfusion in the stalk-medial eminence. Endocrinology. 2000;141:219-228.

58. Plant TM, Gay VL, Marshall GR, Arslan M. Puberty in monkeys is triggered by chemical stimulation of the hypothalamus. Proc Natl Acad Sci U S A. 1989;86:2506-2510.

59. Wrennik JM, Delemarre-van de Waal HA, Schoemaker R, Schoemaker H, Schoemaker J. Luteinizing hormone and follicle stimulating hormone secretion patterns in girls throughout puberty measured using highly sensitive immunoradiometric assays. Clin Endocrinol (Oxf). 1990;33:333-344.
77. Lehman MN, He W, Coolen LM, Levine JE, Goodman RL. Does the KNDy Model for the control of gonadotropin-releasing hormone pulses apply to monkeys and humans? Semin Reprod Med. 2019;37: 71-83.

78. Hrabovszky E, Siptos MT, Molnár CS, et al. Low degree of overlap between kisspeptin, neuropeptide Y, and dynorphin immunoreactivities in the infundibular nucleus of young male human subjects challenges the KNDy neuron concept. Endocrinology. 2012;153:4978-4989.

79. Skrapits K, Borsay A, Herczeg L, Ciofi P, Liposits Z, Hrabovszky E. Neuropeptide co-expression in hypothalamic kisspeptin neurons of laboratory animals and the human. Front Neurosci. 2015;9:29.

80. Mitsushima D, Hei DL, Terasawa E. Gamma-Aminobutyric acid is an inhibitory neurotransmitter restricting the release of luteinizing hormone-releasing hormone before the onset of puberty. Proc Natl Acad Sci U S A. 1994;91:395-399.

81. Mitsushima D, Marzban F, Luchansky LL, et al. Role of glutamic acid decarboxylase in the prepubertal inhibition of the luteinizing hormone-releasing hormone release in female rhesus monkeys. J Neurosci. 1996;16:2563-2573.

82. Kasuya E, Nyberg CL, Mogi K, Terasawa E. A role of gamma-amino butyric acid (GABA) and glutamate in control of puberty in female rhesus monkeys: effect of an antisense oligodeoxynucleotide for GAD67 messenger ribonucleic acid and MK801 on luteinizing hormone-releasing hormone release. Endocrinology. 1999;140:705-712.

83. Keen KL, Burich AJ, Mitsushima D, Kasuya E, Terasawa E. Effects of pulsatile infusion of the GABA(A) receptor blocker bicuculline on the onset of puberty in female rhesus monkeys. Endocrinology. 1999;140:5257-5266.

84. Kurián JR, Keen KL, Guerriero KA, Terasawa E. Tonic control of kisspeptin release in prepubertal monkeys: implications to the mechanism of puberty onset. Endocrinology. 2012;153:3331-3336.

85. García JP, Keen KL, Anderson RM, Hirshfeld BC, Seminara SB, Terasawa E. Prepubertal tonic gamma-amino butyric acid (GABA) inhibition is upstream of neurokinin B (NKB), kisspeptin, and gonadotropin-releasing hormone (GnRH) neuronal network in male rhesus monkeys. Abstracts of the 101st Annual Meeting of the Endocrine Society, held on March 23-26, 2019, at New Orleans, LA (No. P1-411).

86. Terasawa E, Luchansky LL, Kasuya E, Nyberg CL. An increase in glutamate release follows a decrease in gamma aminobutyric acid and the pubertal increase in luteinizing hormone-releasing hormone release in the female rhesus monkeys. J Neuroendocrinol. 1999;11:275-282.

87. Herbison AE, Moenter SM. Depolarising and hyperpolarising actions of GABA(A) receptor activation on gonadotrophin-releasing hormone neurons: towards an emerging consensus. J Neuroendocrinol. 2011;23:557-569.

88. Berg T, Silveira MA, Moenter SM. Prepubertal development of GABAergic transmission to gonadotropin-releasing hormone (GnRH) neurons and postsynaptic response are altered by prenatal androgenization. J Neurosci. 2018;38:2283-2293.

89. Abreu AP, Dauber A, Macedo DB, et al. Central precocious puberty caused by mutations in the imprinted gene MKRN3. N Engl J Med. 2013;368:2467-2475.

90. Perry JR, Stolk L, Franceschini N, et al. Meta-analysis of genomewide association data identifies two loci influencing age at menarche. Nat Genet. 2009;41:648-650.

91. Perry JR, Elks CE, Sulem P, et al. Parent-of-origin-specific allelic associations among 106 genomic loci for age at menarche. Nature. 2014;514:92-96.

92. Lomniczi A, Wright H, Castellano JM, et al. Epigenetic regulation of puberty via zinc finger protein-mediated transcriptional repression. Nat Commun. 2015;6:10195.
action of neurokinin B to trigger gonadotropin-releasing hormone release is upstream from the kisspeptin receptor. Neuroendocrinology. 2011;94:237-245.

111. Terasawa E, Fernandez DL. Neurobiological mechanisms of the onset of puberty in primates. Endocr Rev. 2001;22:111-151.

112. Terasawa E. Chongthammakun S. Developmental Changes in In Vivo Release of β-Endorphin (β-END) from the Stalk-Median Eminence (S-ME) in Female Rhesus Monkeys. Abstracts of the 21st annual meeting of the neuroscience society, No. 361.1, New Orleans, LA. 1991

113. Lippincott MF, León S, Chan YM, et al. Hypothalamic reproductive endocrine pulse generator activity independent of neurokinin B and dynorphin signaling. J Clin Endocrinol Metab. 2019;104:4304-4318.

114. Keen KL, Petersen AJ, Figueroa AG, et al. Physiological characterization and transcriptomic properties of GnRH neurons derived from human stem cells. Endocrinology. 2021;162(9):bqab120.

115. Kenealy BP, Keen KL, Kapoor A, Terasawa E. Neuroestradiol in the stalk median eminence of female rhesus macaques decreases in association with puberty onset. Endocrinology. 2016;157:70-76.

116. Terasawa E. International neuroendocrine federation masterclass series: the GnRH neuron and its control, first edition. In: Herbison AE, Plant TM, eds. Postnatal Development of GnRH Neuronal Function. Hoboken, NJ: John Wiley & Sons; 2018:61-91.

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