Transmission and pathology of *Streptococcus iniae* in monosex Nile tilapia (*Oreochromis niloticus*) in aquaculture of Bangladesh

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**Abstract**

*Streptococcus iniae* is a major fish pathogen, recently emergent outbreaks were recorded in commercially cultured monosex Nile tilapia (*Oreochromis niloticus*) result in significant losses termed “streptococcosis”- causes unusual appearances with multi-focal pin-point haemorrhages, abscesses, necrosis and asites in skin, fin, muscle, liver, spleen, kidney, blood, interstitial fluid specially in central nervous system and brain. This disease was more prevalent (>26%) at summer when the water temperature was approximately >25°C, percentage of mortality was higher >41% during the overcrowding and improper water chemistry. Raised levels of glucose and ammonium in blood serum causes reduced number of free blood cells released into the haemolymph to stomach and gut, result in refrain from eating in diseased tilapia. Stocking density (200 fish/decimal; class IV) had significant effect (*P*<0.01) on the total production (5,000 to 5,500 kg/ha). *S. iniae* in the circulating blood cells, extra-tubular haemal spaces containing blood vessels, fixed phagocytes in the hepatopancreas (gastrointestinal tract), bacteria-like particles in the brain tissue, vacuum and necrosis in hepatocytes revealed with histopathology. In vitro study revealed that cohabitation of dead or infected fish with healthy fish resulted infection (horizontal transmission mechanism) to the healthy fish.

**Keywords:** Streptococcosis, Monosex tilapia, histopathology, transmission

**INTRODUCTION**

There is increasing interest in the likely effect of global climate change on diseases in both wild and cultured organisms (Lafferty *et al.* 2004). In particular, aquatic animals are highly sensitive to temperature change; overcrowding and improper water chemistry can reduced oxygen tension in water, higher microbial growth and immunosuppression to fish, resulted higher prevalence of disease in monosex Nile tilapia (*Oreochromis niloticus*) (Le Moullac and Haffner 2000) caused by *Streptococcus iniae* (Shoemaker and Klesius 1997).

*S. iniae* is the main etiological agent of streptococcosis also caused by *Streptococcus* and *Lactococcus* sp., in a variety of fresh and saltwater fish species in worldwide and results in estimated losses US$150 million in annually (Shoemaker and Klesius 1997). Temperature, dissolved oxygen, nitrite level (Bunch and Bejerano 1997), various stressor factors (Shoemaker and Klesius 1997), high fish density and infectious dose in intensive culture system (Stoffregen *et al.* 1996, Eldar *et al.* 1997) caused high mortality in tilapia with *S. iniae* (Shoemaker *et al.* 2001).

*S. iniae* was first isolated from a skin lesion of a captive Amazon River fish, *Inia geoffrensis* (Pier and Madin 1976). Since then, the susceptible bacterium has been reported
in many species of fresh, estuarine and marine fish species from 15 countries in 6 continents, including Africa, Asia, Australia, Europe, as well as North and South Africa with ayu (Kitao et al. 1981), barramundi (Bromage et al. 1999), salmon (Eldar et al. 1995), European seabass (Zlotkin et al. 1998), grey mullet (Eldar et al. 1994), grouper (Kvitt and Colorni 2004), rainbow trout (Eldar et al. 1997), red drum (Shen et al. 2005), snapper (Ferguson et al. 2000), silver bream (Bromage and Owens 2002), channel catfish (Shoemaker et al. 2001), tilapia (Abutbul et al. 2004), and yellowtail (Kaige et al. 1984). Among other, *S. iniae* has been isolated from humans with bacteraemia, cellulitis, meningitis, and osteomyelitis (Facklam et al. 2005). The source of human infections has been associated with the preparation of *S. iniae* infected tilapias for cooking (Lehane and Rawlin 2000).

The current study investigates the nature of the causative agent including epidemiology, mode of transmission, seasonal effects on disease prevalence, interaction between the causative agent with host cells or tissues or organs by histopathological analysis and gross pathological observations. The present study also aims to evaluate the growth performance of the all-male Nile tilapia population at various stocking densities and determine an ideal stocking density as well as appropriate water quality under the climatic and ecological conditions prevailing in the cultured systems in southern part of Bangladesh.

**METHODOLOGY**

**Experimental fish and design:** Adult tilapia (*Oreochromis niloticus*) affected by satreptococcal disease were collected from Naolli, Bisali, Kashiupur, Chachuri, Kalia, Shingashul in Jessore and Narail districts (arrow indicates the working area in map; Figure 1) from a pilot farm of South-West Area Integrated Water Resources Planning and Management Project (SAIWRMP) funded by Asian Development Bank (ADB), Ministry of Water Resource and Bangladesh Water Development Board (BWDB) during the period of 2012-2013. Independent experiments were designed in *vitro* at the Department of Fisheries and Marine Bioscience laboratory, Jessore University of Science and Technology and Faculty of Fisheries, Bangladesh Agricultural University, to evaluate the health, dietary needs, density, dose of tilapia (diseased or disease-free) at different replicable stocked such as: 80, 120, 160, 200, 240 and 280 fry/decimal corresponding to density classes (I, II, III, IV, V and VI), bacterial strain, infection trails and histopathology.

**Health status assays:** Blood and interstitial fluid (haemolymph) was withdrawn from tilapia, *O. niloticus* by puncturing the unsclerotized membrane with a 21 gauge needle and syringe. For total haemocyte (blood cell) counts (THC), the blood was mixed with an equal volume of ice-cold fixative (5% formaldehyde in 3% sodium chloride solution) and cells counted with a Neubauer haemocytometer. For blood chemistry (total protein, ammonium and glucose) blood was allowed to clot for ~60 min on ice, centrifuged (5000 g; 10 min) and the resulting serum stored at -80°C. Total protein was determined using a biocinchromic acid assay kit (Pierce and Warriner) using BSA as a standard, while glucose and ammonium concentrations were determined using methods (Bergmeyer 1984, Bolz and Howel 1978) modifications. Glycogen levels of the hepatopancreas (digestive gland) were determined using the anthrone method (Van Handel 1965) with type II glycogen (Sigma-Aldrich) as the standard. All assays were performed in 96-well microtitre plates and absorbance’s read on an Anthos II plate reader.

**Culture pond:** Fifty newly excavated 0.02 to 0.05 ha freshwater earthen ponds were completely drained, tilled, dried, leveled and limed at the rate of 247 kg/ha during the sunny days after plowing. Ponds limed and one week later water level was increased up to 1.37 m to support natural productivity. Organic fertilizers (cattle manure 1235 kg/ha and compost 1235 kg/ha) and inorganic fertilizers (urea and TSP; 24.7 kg/ha) were applied in time intervals (15 days) during the culture period.

**Monosex Nile tilapia production:** Three days old mixed sex juveniles of Nile tilapia (mean weight 0.0021±0.005 g; mean length 1.20±0.010 cm) were collected from the hatchery. Fish stocked in each of the culture systems at a density of 50 individuals/m² and hormone treated diets were prepared by the 99% ethyl alcohol evaporation technique (Shelton et al. 1978).

Fish (mean weight 18±2 g) acclimatized and released in the working area in map; Figure 1) from a pilot farm of Shingashul in Jessore and Narail districts (arrow indicates collected from Naolli, Bisali, Kashipur, Chachuri, Kalia, *O. niloticus* affected by satreptococcal disease were collected from Naolli, Bisali, Kashiupur, Chachuri, Kalia, Shingashul in Jessore and Narail districts (arrow indicates the working area in map; Figure 1) from a pilot farm of South-West Area Integrated Water Resources Planning and Management Project (SAIWRMP) funded by Asian Development Bank (ADB), Ministry of Water Resource and Bangladesh Water Development Board (BWDB) during the period of 2012-2013. Independent experiments were designed in *vitro* at the Department of Fisheries and Marine Bioscience laboratory, Jessore University of Science and Technology and Faculty of Fisheries, Bangladesh Agricultural University, to evaluate the health, dietary needs, density, dose of tilapia (diseased or disease-free) at different replicable stocked such as: 80, 120, 160, 200, 240 and 280 fry/decimal corresponding to density classes (I, II, III, IV, V and VI), bacterial strain, infection trails and histopathology.

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The fish were fed with 17α-methyltestosterone (17αMT) hormone treated diet (35% protein) with a dose of ≤ 60 mg MT/kg feed at a rate of 50%, 30% and 20% body weight/day for 21 days (Haylor and Pascual 1991, Popma and Green 1990) and other report showed 10 mg MT/kg feed at a rate of 20% body weight/day for 30 days (Chakraborty et al. 2007, Chakraborty and Banerjee 2009) and hormone treated diets were prepared by the 99% ethyl alcohol evaporation technique (Shelton et al. 1978). Fish (mean weight 18±2 g) acclimatized and released in
the 50 pilot ponds at six different stocking densities of 19760, 29640, 39520, 49400, 59280 and 69160 fry/ha corresponding to density classes (I, II, III, IV, V and VI). After first month, the fish were fed twice to four times daily with hormone untreated control diet (crude protein content 25 to 30%) and total digestibility 2500±300 kcal/kg feed) at a constant rate of 10, 8, 7, 6, 5, 4, 3, 2 and 1% of body weight/day for the next 120 days respectively.

The entire experiment was conducted in three replicating units for statistical validation. Throughout the entire culture period different water quality parameters like temperature, DO, free CO$_2$, pH, total alkalinity and transparency were monitored daily using standard procedures (Apha 1998). Besides, growth parameters like specific growth rate (SGR), relative growth rate (RGR), daily weight gain (DWG), food conversion ratio (FCR) and protein efficiency ratio (PER) were measured according to production was also calculated.

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**Results for Water Quality Parameters**

| Parameter       | Density Class (fish/ha) |
|-----------------|-------------------------|
| Temperature     | 200 (25.2°C to 33.7°C)   |
| pH              | 7.2 to 8.0 (max. 8.0±0.1) |
| Alkalinity      | (122.5 mg/l to 145.6 mg/l) |
| Transparency    | (25.2 cm to 39.5 cm)     |
| Free CO$_2$     | (8.2±0.4 mg/l; lowest (3.1±0.4 mg/l), and average survival rate 73.92%. Stocking density had significant effect (P<0.01) on the survival percentage, maximum (84.5±1.55%) and minimum (62±1.08%) survival were found in the density classes I and VI, respectively (Figure 2). Stocking density (200 fish/decimal; class IV) had significant effect (P<0.01) on the total production (5,000 to 5,500 kg/ha) and average weight 300 to 320 g and also showed the highest weight,
length, RGR, SGR, PER and protein content (Table 2). Among the six different density categories, fishes of class IV showed maximum yield while the minimum yield was observed in density class I (Figure 2). FCR was maximum for the density class I where stocking density was 19760 fish/ha and minimum for density class VI corresponding to 59280 fish/ha (Table 2). The overall average production of all the density groups was 3195.6±287.8 kg/ha.

**Table 1.** Weekly values of physicochemical parameters of different density class ponds during monosex tilapia culture

| Density classes | Temperature (°C) | DO (mg/l) | Free CO₂ (mg/l) | pH | Transparency (cm) | Alkalinity (mg/l) |
|----------------|-----------------|-----------|-----------------|----|------------------|------------------|
| I              | 25.2±0.8        | 6.1±0.08  | 5.1±0.6        | 7.2±0.08 | 39.5±0.3 | 123.4±3.3        |
| II             | 28.6±0.8        | 7.5±0.08  | 6.5±0.4        | 7.4±0.1  | 35.0±2.2 | 139.7±2.0        |
| III            | 31.0±0.7        | 7.1±0.1   | 3.1±0.4        | 8.0±0.1  | 35.3±0.3 | 145.6±1.8        |
| IV             | 30.7±0.7        | 7.1±0.09  | 5.4±1.0        | 7.4±0.08 | 30.2±1.0 | 124.9±4.6        |
| V               | 31.2±0.8       | 7.4±0.1   | 6.5±0.4        | 7.8±0.09 | 25.2±0.8 | 122.5±2.5        |
| VI              | 33.7±1.0        | 4.1±0.1   | 8.2±0.4        | 7.2±0.09 | 27.8±1.0 | 132.1±4.3        |

**Table 2: Growth performances of monosex Nile tilapia at various stocking densities during pond culture**

| Density classes | Weight (g) | Length (cm) | DWG (g/day) | RGR (%) | SGR (%) | FCR | PER | Protein (% wet weight) |
|----------------|------------|-------------|-------------|---------|---------|-----|-----|------------------------|
| I              | 192.7f     | 21.2f       | 1.74f       | 448.4f  | 1.9d    | 2.5a | 1.3d | 13.8e                  |
| ±2.3           | ±0.2       | ±0.02       | ±0.6        | ±0.04   | ±0.04   | ±0.03| ±0.04|                        |
| II             | 231.0d     | 22.6d       | 2.19d       | 557.48d | 2.1c    | 2.3b | 1.4c | 14.18d                 |
| ±2.3           | ±0.2       | ±0.08       | ±0.6        | ±0.06   | ±0.02   | ±0.02| ±0.02|                        |
| III            | 291.6b     | 24.7b       | 2.87b       | 729.95b | 2.3a    | 1.9d | 1.5b | 16.03b                |
| ±2.4           | ±0.1       | ±0.02       | ±0.3        | ±0.05   | ±0.03   | ±0.01| ±0.01|                        |
| IV             | 309.6a     | 25.4a       | 3.13a       | 780.22a | 2.4a    | 2.1c | 1.6a | 16.58a               |
| ±2.6           | ±0.2       | ±0.03       | ±0.5        | ±0.03   | ±0.03   | ±0.01| ±0.01|                        |
| V               | 261.5c     | 23.5c       | 2.49c       | 644.15c | 2.2b    | 1.8e | 1.5b | 15.73c               |
| ±1.6           | ±0.1       | ±0.02       | ±0.3        | ±0.02   | ±0.02   | ±0.02| ±0.02|                        |
| VI              | 208.9e     | 21.8e       | 1.93e       | 494.23e | 1.9d    | 2.4a | 1.4c | 14.15e               |
| ±2.2           | ±0.1       | ±0.02       | ±0.4        | ±0.02   | ±0.03   | ±0.02| ±0.06|                        |

**Culture of the putative streptococcal disease agent:** Attempts to isolate and grow the potential causative agent on a wide range of agar-based culture media failed despite the use of long incubation periods and a range of dilutions of the infected haemolymph. These procedures were attempted using haemolymph from different streptococcal diseased affected fish. In all cases, the plates were devoid of colonies apart from the occasional contaminant (<0.001% of the total number of bacteria seen in blood by direct microscopy). Such colonies were not found on a routine basis from streptococcal disease affected fish.

**Infection:** Mortalities in the low, medium and high-density treatments began occurring 2 to 4 days post infection. Typical signs of streptococcal disease i.e., erratic swimming, darkening of the fish, exophthalmia and cloudy eyes were observed (Plumb 1997) in the dead and dying tilapia following infection. *S. iniae* was isolated from both brain and anterior kidney of fresh dead tilapia on blood agar. Density was shown to significantly affect mortality of tilapia infected by immersion exposure to *S. iniae* (Table 3).

A significant difference (*P*<0.05) was seen in the mortality when compared low (6.4%) and medium (30.6%) and low and high (26.4%) density treatments. No significant difference was observed when compared medium and high-density treatments. Two-way analysis of variance demonstrated density had a significant effect on *S. iniae* mortality (*P*=0.0001). Dose utilized had little effect on mortality. High density by dose however, showed a significant interaction (*P*=0.001), which is evident when looked at the mortality of each dose in the high-density trial (Table 3). Mortalities in the cohabitation treatment began to occur between day 5 and day 7 after addition of the dead moribund fish.

Dead fish were seen earlier (day 2) in the immersion treatments. Signs of streptococcal disease were observed in both cohabitation and immersion treatments and *S. iniae* was isolated from fresh dead fish on blood agar. No mortality occurred in the noninfected tilapia for the 28 day trial. No significant differences (*P*<0.05) in mortality was observed between replicates 1 and 2 of the immersion exposed treatments (41.3% and 37.3%, respectively) and of the cohabitation treatment (26.6%, Table 4). All the infected treatments exhibited significantly higher (*P*=0.0008) mortality when compared to the control treatment where no fish died.
**Gross pathology and disease prevalence:** Infected tilapia initially refrained from eating, the stomach and gut becomes empty or nearly empty, in most cases fibrous material seen in the peritoneal cavity; seriously weakened tilapia ultimately died (within 7 days of initial sampling). Diseased fish had significantly higher levels of ammonium in the blood than non-diseased individuals sampled at the same times (18.3±1.8 µg ml-1 blood in diseased fish compared with 3.4±0.4 µg ml-1 blood in non-diseased fish; mean values±SEM, n=20–81, P<0.001, Student’s t-test).

During routine sampling employed as a method of identifying affected individuals showed eye lesions, haemorrhages (endophthalmia or exophthalmia) and also lead to unilateral (one eye) or bilateral (both eyes) opacification of the eye (Figure 3A and 3B). Streptococcosis developed abscesses on the inferior jaw lead to burst and turn into haemorrhagic ulcers (Figure 3B and 3C). This disease affected the central nervous system of tilapia and infected fish engaged in various forms of abnormal behavior, such as lethargy, disorientation, swirling, bending of the body and ascites was accompanied by a protruding anus (Figure 3A-3D). Skin was found to exhibit the characteristic multi-focal pin-point haemorrhage, normally located near the mouth, anus, genital papilla and the base of the fins (Figure 3C and 3D).

**Effects of disease on blood cells, serum protein, glucose and ammonium, and hepatopancreatic glycogen:** Blood from the streptococcal disease affected tilapia showed THC of apparently healthy fish decreased significantly upon development of streptococcal disease (Figure 6A). Similarly, the amount of total protein in serum also significantly declined before the initial diagnosis of disease (Figure 6A), while in the case of glucose and ammonium (Figure 6B), a significant increase in these factors was found at the time of diagnosis. Levels of glycogen in the hepatopancreas (digestive gland) of affected and non-affected fish were not found to differ significantly.

**Histopathology:** Figure 7A shows variety of epithelial cells, extra-tubular haemal spaces containing blood vessels, fixed phagocytes, and freely circulating

![Figure 3: Gross signs of disease in an infected tilapia with *Streptococcus iniae*](image1)

![Figure 4: Effect of season and water temperature on the prevalence of streptococcal disease in a pilot farm](image2)

![Figure 5: (A) Appearance of haemolymph from streptococcal disease affected tilapia (1, 2) and non-diseased tilapia (3, 4). (B) Phase-contrast micrograph of blood from a streptococcal disease affected tilapia](image3)
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haemocytes in gastrointestinal tract. A gram stained, impression print of *S. iniae* infected brain tissue illustrating the gram-positive chains of cocci (Figure 7B). Occasionally, haemocyte accumulations, termed nodules, were found in the haemal space; some of these were seen to contain debris and bacteria-like particles in brain tissue of diseased tilapia (Figure 7C). All other tissues examined (gonad, alimentary canal and muscle) were structurally normal (not shown). The degeneration in hepatocytes, vaculation, congestion and necrosis in hepatocytes *O. niloticus* injected with *S. iniae* (Figure 7D).

Transmission of disease and temperature stress experiments: Asymptomatic carriers have been found and horizontal transmission has been reported during outbreaks of streptococcal disease. This disease transmitted horizontally and major routes of transmission apparently via infected fish to disease-free tilapia with fed skin, organs and trash fish or via cannibalism, injuries, etc., and also from the contaminated water to the fish after 6 to 8 weeks post-challenge (Table 3 and 4). Although the disease caused mortality in fish as small as 10 g, however, bigger fish (from 100 g to market size) were more susceptible to the disease. The greatest losses occurred in large fish close to the market size (300 to 400 g). Outbreaks occurred when fish exposed to stress with suboptimal oxygen levels (<2ppm) in the water and overcrowding (50 fish/m³) for a long period of time (Table 3 and 4). Fish also showed acute symptoms with peaks of mortality lasting 2-3 weeks during the high water temperature (>25°C), it was chronic, when water temperature was low (<10°C) and showed persistent level of mortality (Table 3 and 4). Furthermore, during this time, the THC of these individuals was differing from that affected fish to non disease affected animals (data not shown). Intrahepatic injection of apparently healthy tilapia with blood (100-300 ml containing ~108 bacteria) from disease affected individuals resulted in large mortalities (100%) within 24 hours of injection. Similarly, injection of fish with the same volume of cell-free haemolymph (i.e. with neither haemocytes nor bacteria) from disease affected fish resulted in 100% mortality at 24 hours post-challenge. In the case of fish injected with the same volume of whole blood from uninfected fish, no mortalities were observed 24-48 hours post-challenge.

**Figure 6:** (A) Changes in total haemocyte counts and total serum protein, and (B) serum glucose and serum ammonium levels prior to the initial identification of milky disease in crabs. Values shown are means±SD, n=12–17. *P<0.05, **P<0.01 and ***P<0.001 compared with time 0

**DISCUSSION**

The current study has revealed the epidemiology and pathology of streptococcal disease in tilapia cultured farms during summer months when high water temperatures prevail. It is well established that tilapia, like all aquatic animals, are highly sensitive to environmental degradation such as elevated or depressed temperature, reduced oxygen tension, overcrowding and changes in the concentrations of xenobiotics or waste products in the water (Le Moullac and Haffner 2000). Temperature affected on the immune systems of tilapia...
resulting in immune suppression and also the antibacterial activity of the haemocytes of tilapia which enhanced disease prevalence and severity (Chang and Plumb 1996) which was similarly observed in the farm and in vitro conditions during the summer months in the present study. In the present study THC was found rapid decline in early stages of streptococcal disease and significant elevation in the amounts of glucose and ammonia in the blood of affected tilapia due to environmental stress (Yoganandhan et al. 2003). Intrahepatic injection of healthy tilapia with blood from streptococcal disease affected tilapia resulted in their rapid (>24 hours) mortality but without any of the key symptoms (septicaemia-like infection) of the disease as seen in natural infections in the farm and in vitro conditions.

Table 3. Density and dose effect on mortality following S. iniae immersion challenge in tilapia held for 28 days

| Dose CFU/ml plate count | No. fish per tank five tanks for each dose | No. dead/No. total* | % Mortality (SEM)** |
|-------------------------|------------------------------------------|---------------------|---------------------|
| 3.2x10^7                | 25                                       | 8/125               | 6.4 (±1.5)**        |
| 6.1x10^7                | 25                                       | 6/125               | 4.8 (±0.8)**        |
| 2.5x10^8                | 25                                       | 10/125              | 8.0 (±1.6)**        |
| **Low density**         | Mean % mortality (SEM) 6.4 (±0.8)**       |
| 2.5x10^7                | 50                                       | 93/250              | 37.2 (±2.1)**       |
| 3.8x10^7                | 50                                       | 64/250              | 25.6 (±2.6)**       |
| 7.1x10^7                | 50                                       | 73/250              | 29.2 (±3.7)**       |
| **Medium density**      | Mean % mortality (SEM) 30.6 (±2.0)**      |
| 2.7x10^7                | 100                                      | 83/500              | 16.6 (±1.9)**       |
| 4.9x10^7                | 100                                      | 137/500             | 27.4 (±5.1)**       |
| 1.9x10^8                | 100                                      | 176/500             | 35.2 (±4.6)**       |
| **High density**        | Mean % mortality (SEM) 26.4 (±3.0)**      |

* Fresh dead fish were cultured on blood agar and S. iniae was isolated from the brain and kidney.

** Lower case superscripts denote differences in the means on each group of mortality data in low, medium and high treatments (i.e., dose) by Duncan’s Multiple Range Test.

*** Low density = 5.6 g/l

† Upper case superscripts denote differences in the mean percent mortality as compared by two-way analysis of variance using Duncan’s multiple range test to compare means. Significance level was equal to P<0.05.

§ Medium density = 11.2 g/l

‖ High density = 22.4 g/l

Lethargy, loss of appetite, sluggish movement, dark discoloration of the skin and respiratory manifestations were found as common clinical sings and histopathological observations revealed lesions, necrosis, haemorrhage and ulcer in all vital organs of necropsied fish with accumulation of fluids in abdomen, congested gills and dark liver in this study. Similar clinical sings and histopathological hepatic lesions, severe congestion, vascular degeneration of hepatocytes, focal coagulative necrosis, focal replacements of the hepatic parenchyma with extravagated blood, hemorrhage, activation of melanomacrophages were reported of aflatoxicated Nile tilapia (Mehrim et al. 2006).

Table 4: Density effect on route of infection with Streptococcus iniae (100 fish/tank; 3 tanks/treatment)

| Treatment | No. dead/no. total* | % Mortality (SEM)** |
|-----------|---------------------|---------------------|
| Non infected*** | 0/300               | 0.0 (±0.0)**        |
| 9.6x10^7 (re 1)* | 124/300             | 41.3 (±7.3)**       |
| 9.6x10^7 (rep 2) | 112/300             | 37.3 (±2.8)**       |
| Cohabitation*    | 80/300              | 26.6 (±2.5)**       |

* Fresh dead fish were cultured on blood agar and S. iniae was isolated from the brain.

** Significant differences in the means are noted by different letters (P=0.0008). The standard error of the mean is presented in parentheses.

*** Non infected tilapia (controls) which were immersed in tryptic soy broth only.

† Tilapia infected with S. iniae (CFU/ml) as determined from triplicate plate counts of immersion water on blood agar plates.

* Cohabitation infection was accomplished by adding five moribund or fresh dead tilapia to each tank for 48 h before removal.

For tilapia culture generally considered the optimum temperature 20-30°C or above; DO: 4-5 mg/l; free CO2: 20 mg/l; pH: 6.5 to 11.0; transparency: 25-35 cm and stocking density: 200-250 fish/decimal (Tran-Duy et al. 2008), similar results were observed more or less stable during the experimental period. The cannibalism had found a main cause of tilapia fry mortality at high stocking densities (El-Sayed 2002) this study also indicates an inverse relationship between the stocking density and growth potential of the fish when it is stocked at a very high density. But interestingly, the maximum growth and well survival percentage of the fish is obtained at an intermediate stocking level. Monosex Nile tilapia stocked in ponds at a low density showed better growth than at a higher density, due to higher density showed intense antagonistic behavioral interaction, competition for food and living space, environmental stress, social dominance, lower individual growth rates, deterioration in water quality, rise plasma cortisol concentrations (Diana et al. 2004). The present study established an optimum stocking density level of tilapia for maximum utilization of food and space with minimum stress and energy expenditure resulting in higher growth potential of the fish, on other hand at the low stocking densities showed decreased feed utilization efficiency, stunted growth, feed wastage and increase production cost.

In our study, fish were housed in aquaria that allowed for one-half water exchanges per hour. But in closed culture systems, tilapia held at high densities, little or no water is exchanged, which leads the significant increases in mortality 70% or more (Craig et al. 2000). Dose of S. iniae
applied to infect the tilapia via immersion exposure with highest density treatment, showed significantly higher mortality with the greater number of \textit{S. iniae} and low-density treatments resulted in little mortality. Increased density enhances direct contact with a greater concentration of \textit{S. iniae} in water from dead diseased fish that favors transmission of \textit{S. iniae} and subsequent inoculation via abrasion of the infected fish skin, resulted in large mortality (Perera et al. 1997). We were successful in inducing mortality in tilapia following cohabitation by placing previously infected tilapia in the same tank for 48 hours exposure before removal (Robinson and Meyer 1966). While the dead or moribund fish were in the tanks, we observed cannibalism of eyes and viscera. This suggests an oral and/or olfactory mode of infection; able to infect tilapia using direct delivery of \textit{S. iniae} to the gut which was responsible for infection by waterborne \textit{S. iniae}.

Many studies have been carried out to reveal the transmissions of \textit{Streptococcus} sp. from the newly introduced fish into the farm (Nguyen et al. 2002), bacteria on excreted feces of infected fishes, survive in the water, be infectious to other healthy fish (Nguyen et al. 2002), infected thrashed fish as fish feed also responsible for the outbreaks of streptococcosis in farm (Kim et al. 2007). The present study revealed the horizontal transmission of the pathogens between fish in cohabitation experiment, similarly this mechanism usually involved in fish that were cultured in high densities, same as different species of wild and cultured fish (Evans et al. 2002), infection have been occurred through wounds and abrasions of the skin (Xu et al. 2007). The fish cohabiting barramundi pens had the same \textit{S. iniae} strains as the barramundi, in addition, the transmission among the species of reef fish to fish via contact and abrasion were also observed (Bromage and Owens 2002).

CONCLUSION

The number of \textit{S. iniae} or dose used in the cohabitation experiment (cannibalism) and seasonal effect was properly quantified for streptococcal outbreaks. Stocking density (49400 fish/ha; class IV) had significant effect (P<0.01) on the total production (5,000 to 5,500 kg/ha) and average weight 300 to 320 g and also showed the highest weight, length, RGR, SGR, PER and protein content. High density (100 fish/4 l tank) effect on route of infection i.e., percentage of mortality was 41.3±7.3, low density (25 fish/4 l tank) showed the percentage of mortality was 4.8±0.8. Reducing fish density may be another management strategy to limit transmission of \textit{S. iniae} in intensively farmed tilapia. This disease was more prevalent (>26%) at summer when the water temperature was approximately >25°C, percentage of mortality was higher >41% during the overcrowding and improper water chemistry. Raised levels of glucose and ammonium in blood serum causes reduced number of free blood cells released into the haemolymph to stomach and gut, result in refrain from eating in diseased tilapia. The transmissions routes of infection, reduced free blood cell numbers and total serum protein, raised levels of glucose and ammonium in blood and histopathological observation suggested adequate control measures and a best management strategy should be employed to quickly remove dead and moribund fish from intensive culture operations. Reducing fish density, lowering water temperature, stopping overfeeding and water exchange also suggested a new approach for management strategy to limit transmission of \textit{S. iniae} in intensively farmed tilapia.

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