MORPHOLOGICAL AND MOLECULAR CHARACTERS OF *Mimegralla* spp. (DIPTERA: MICROPEZIDAE) ON ZINGIBERACEAE IN CENTRAL JAVA

**ABSTRACT**

Rhizome fly, *Mimegralla* sp. (Diptera: Micropezidae) is a major pest on Zingiberaceae plants. Some of those fly species have been reported attacking some of Zingiberaceae plants, but in Indonesia, only one species, *Mimegralla coeruleifrons* has been reported as being a pest. Since Indonesia has many species of Zingiberaceae plants, it may raise a prediction that more than one species of Mimegralla was found on this plants. Therefore, a taxonomic research on the species of rhizome flies to find the species other than *M. coeruleifrons* on Zingiberaceae plants is urgently required. This study was conducted by using hand-picking method on Mimegralla adult inhabiting Zingiberaceae plants (ginger, turmeric, javanese ginger, and aromatic ginger), and was then identified by using morphological characters and through molecular technique by using mtCO1 gene. The results showed that *M. albimana* and *M. coeruleifrons* found at four zingiberaceae plants were the member of Mimegralla. As a conclusion, these two species have high values of phylogenic relationship (88%) and bootstrap (92).

**Keywords:** *Mimegralla* spp., molecular, morphology

INTRODUCTION

Zingiberaceae plants, including genus Curcuma, Kaempferia, Hedychium, Amomum, Zingiber, Alpinia, Elettaria, and Costus (Joy et al., 1998), are the main materials in producing herbal medicines, which is containing volatile oils and oleoresins as tonic and stimulant for human body. Rhizome fly is noted as a major pest in ginger plants (Steyskal, 1964), turmeric (Nair, 1980), and aromatic ginger (Balfas et al., 2000) in several countries, including Indonesia.

Two species of rhizome fly are reported attacking Zingiberacea in India (Maxwell-Lefroy & Howlett, 1909), those are *Mimegralla coeruleifrons* Macq. (Ghorpade et al., 1983) and *M. albimana* Doleschall (Ghorpade et al., 1988). But only one species, *M. coeruleifrons* Macq., was reported attacking Zingiberaceae in Indonesia (Balfas, 2002). However, the diverse species of Zingiberaceae planted in Indonesia raises assumption that the number of Mimegralla attacking rhizomes might be more than one species. Therefore, a thorough study should be done to clarify this issue. This study was aimed to know the species of Mimegralla attacking Zingiberaceae in Central Java by using morphological and molecular characters. In addition, the molecular identification by using
mtCO1 characters on rhizome fly (*Mimegralla* spp.) has never been done, and even the information of base nucleotide was never found in NCBI GenBank. So identification using mtCO1 need to be done to understand the phylogenetic relationship.

**MATERIALS AND METHODS**

**Sampling and Preservation**

*Mimegralla* spp. adults were collected by hand-picking from ginger (*Zingiber officinale*), turmeric (*Curcuma domestica*), Javanese ginger (temulawak, *Curcuma xanthorrhiza*), and aromatic ginger (kencur, *Kaempferia galanga*) plants in Karanganyar District (latitude of 7°37'53.674", longitude 110°56'58.957" and altitude of 204.7 m above sea level) and Purworejo District (latitude 7°42'15.660", longitude 110°1'47.537" LE and latitude of 81.5 m above the sea level). Sampling was conducted in area, where symptomatic of rhizome fly, i.e. rich of organic material rot (Hennig, 1935) and wilt (Karmawati *et al.*, 1990) were found. Sampling were conducted in October 2016, when the rainy season started and Zingiberaceae thrives at this season. Heavy rainfall is the optimum condition of the microorganisms to grow well.

Adult of rhizome flies were used for morphological and molecular identifications, and insect collection as well. Adults used for morphology identification were pinned using insect needles no. 00 and put into 96% alcohol for molecular identification.

**Identification**

Morphological and molecular identification were conducted at Laboratory of Basic Entomology and Laboratory of Virology, Department of Plant Pest and Disease, Faculty of Agriculture, Universitas Gadjah Mada, Yogyakarta.

Morphological identification of fly was done by using Leica MZ16 and Leica KL1500 LCD Microscopes and Optitab advance on the whole part of body to describe shape, size, color, and genitalia parts as specific characters of species. Magnification of microscopes showed in Table 1.

DNA extraction was done by using Genomic Mini Kit from Geneaid, and the extraction product was then amplified by PCR using LCO1490 (GGT CAA CAA ATC ATA AAG ATA TTG G) as forward primer and HCO2198 (TAA ACT TCA GGG TGA CCA AAA AAT CA) as reverse primer (Folmer *et al.*, 1994). The kit used in PCR reaction was Go Taq Green Master Mix kit: Go Taq Green Master Mix 12.5 µL; primer forward 1 µL; reverse primer 1 µL; DNA template 2 µL; and Nuclease Free Water 8.5 µL. Stages of PCR cycles were shown in Table 2. The results of subsequent DNA amplification were visualized by agarose gel electrophoresis (SIGMA) and observed using BIO-RAD UV transluminator 2000. Sequencing was carried out by sending samples to 1st Base DNA Sequencing (Selangor, Malaysia). Finally, to analyze the degree of neighborhood joining values, Bioedit software was used, and followed by Mega 5.2. and BLAST software. The Bioedit was

| Component identification | Magnification |
|---------------------------|---------------|
| *Mimegralla coeruleifrons* | *Mimegralla albimana* |
| All body                  | 11.4 x        | 20.0 x       |
| Caput                     | 51.2 x        | 64.0 x       |
| Thorax                    | 40.0 x        | 51.2 x       |
| Abdomen                   | 25.6 x        | 40.0 x       |
| Wings                     | 25.6 x        | 32.0 x       |
| Antenna                   | 128.0 x       | 128.0 x      |
| Copulatory fork           | 100.8 x       | 100.8 x      |
| Ovipositor                | 32.0 x        | 80.0 x       |

| No. | Reaction     | Temperature | Time   | Cycle    |
|-----|--------------|-------------|--------|----------|
| 1   | Predenaturasi| 95°C        | 2 minutes | 30 cycles |
| 2   | Denaturasi   | 95°C        | 30 seconds |         |
| 3   | Annealing    | 55°C        | 30 seconds |         |
| 4   | Extention    | 72°C        | 1 minute  |         |
| 5   | Final extention | 72°C   | 5 minutes  |         |
| 6   | Delay        | 4°C         | 5 minutes  |         |
used to make base paired alignment of forward and reverse nucleotide bases, and to get the percent of clustal consensus. We used MEGA 5.2. for making phylogeny tree and used BLAST for getting homology percentage.

RESULTS AND DISCUSSION

In this study 78 samples of suspected Mimegralla was found in Zingiberaceae plants (ginger, turmeric, javanese ginger, and aromatic ginger). The result of morphological identification showed that the samples was consisted of two different Mimegralla species, i.e. *Mimegralla albimana* and *M. coeruleifrons* (Figure 1). To complete the observation on the differentiation between these two species, three individual samples of each species were selected randomly, and were examined by using molecular technique.

Morphology Identification

Stilt-legged fly (*Mimegralla* spp.) is a member of family Micropezidae (Diptera: Acalyptratae), and there are 583 species of Micropezidae reported, which are divided into 52 genera and 5 subfamilies (Pape *et al*., 1758). Most of Micropezidae species were found in tropic and subtropic areas. This fly has unique characteristics by their long and slender feet.

Aczel (1959) mentioned that a specific character of *Mimegralla* is the occurrence of ocellar plate on the posterior of upper superior orbital or very close to the fore ocellus placed in a line. Frontal stripe are more than a half of total width of frons. Upper anterior orbital stand in tomentosa frontal or form a boundary line between the frontal stripe and genovertical plate. Postvertical and genal bristle always absen.

1. Forceps in apex of copulatory fork tapered at the ends with intersect bristles (in males). Shape of ovipositor oval with membranous region at the end the shorter (Figure 2). Body color is brownish red and yellowish ...................... *albimana*

2. Forceps in apex of copulatory fork show widened with a short bristle does not intersect (in males). Shape of ovipositor conical with an elongated tip at membranous region (Figure 3). Overall body color is dark blue and black ............ *coeruleifrons*

*M. albimana* and *M. coeruleifrons* could be distinguished by their color. *M. albimana* has brownish red and yellow on its body. Facet are maroon, antenna are reddish yellow, abdomen black and brownish yellow, legs bright yellow with black stripes on coxa, more dark to the apex. Whereas *M. coeruleifrons* has blackish-dark-blue body. Facet are dark blue and metallic when exposed to the light, antennae are dark brown, abdomen is black and gray, and legs are brownish with black stripes on coxa. Hennig (1935) also reported, *M. albimana* legs are continuosly dark whereas *M. coeruleifrons* legs are dark.

However, when the body of *M. albimana* and *M. coeruleifrons* were observed closer by using higher magnification, some differences were found. These differences were observed on preabdomen, ovipositor, wings, and copulatory fork. Body of *M. albimana* was stout and looks more convex than long, length of wings were 3.5 to 3.7 times than width, and length of body was 4.3 to 4.5 times than width (Figure 1). Whereas *M. coeruleifrons* was more slender and elongated and wing length was 3.8 to 4.5 times than width. Body lenght 4.6 to 4.9 times than width.

![Figure 1. Female adult of *Mimegralla albimana* (a); adult of *Mimegralla coeruleifrons* (b): frons (f), tergite 1–3 (t), apex of wing (ap), ovipositor (o), legs (l)](image-url)
Molecular Identification

DNA amplification on adult was done by using cytochrome c oxidase subunit 1 in mitochondrial (mtCO1) part and was performed by universal primers LCO1490 and HCO2198, which are mostly used for invertebrates (Folmer et al., 1994).

DNA amplification was visualized using BIO-RAD UV transluminator 2000, and a single band at 600 bp. This product then was sent to 1st Base DNA Sequencing at Selangor, Malaysia. The result of DNA sequencing of *M. albimana* showed, that it has 681 bp (base pairs) of DNA nucleotide bases, whereas *M. coeruleifrons* has 678 bp. These results were consistent with results of PCR visualization as described above. The nucleotide bases of *M. albimana* was compared with *M. coeruleifrons* and three other species (Table 3), the member of Micropezidae Family, to determine the value of homology between the first two species and the other three. The nucleotide bases of the three other species was taken from GenBank database as shown at Figure 4.

The nucleotide bases of five species were compared to determine the level of homology in percent. The closeness of phylogenetic relationship in clustal consensus
Figure 4. Clustal consensus between *Mimegralla* spp. and three others species of Micropezidae Family from GenBank database
could be determined by counting nucleotide base without an asterisk (*). The result indicated that *M. coeruleifrons* has closest relationship with *M. albimana* (88%) than with the other three species (*Hemichaeeta scutellata*, *Taeniaptera trivittata*, and *Micropeza* sp.) on homology (Table 4) and clustal consensus (Figure 4). Sokal and Sneath (1963) stated that the closeness of phylogenic relationship between organisms are indicated by the similarity of common characters. Figure 5 showed that *M. coeruleifrons* and *M. albimana* has a bootstrap value at 92, means that these species are member of Mimegralla Genus. Bootstrap analysis is used to test the validity and reliability of data (Hedges, 1992). Thus, the value of 92 means that the data in phylogeny analysis has high validity and reliability.

## CONCLUSION

*M. coeruleifrons* and *M. albimana* found in Zingiberaceae plants (ginger, turmeric, javanese ginger, and aromatic ginger) were member of the same genus, Mimegralla, by the facts that these two species have high values of phylogenetic relationship (88%) and bootstrap (92).

## ACKNOWLEDGEMENTS

The author would like to thank Prof. Steve Marshall, Dr. Awit Suwito and Dr. Rodiah Balfas for identification assistance, Siti Haryati for DNA extraction assistance, Abdu Rohman and Rizki Rajabillah for phylogeny discussion. Funding for this project was provide by BPPSDM of Ministry of Health.

| Table 3. Origin of *Mimegralla* spp. and three other species of Micropezidae Family from GenBank database were used for phylogenetic comparison |
|-----------------|-----------------|-----------------|-----------------|
| Species         | Total of nucleotide base (bp) | Origin              | Source                          |
| *Mimegralla albimana* | 681              | Karanganyar        | This research                    |
| *Mimegralla coeruleifrons* | 678              | Karanganyar        | This research                    |
| *Hemichaeeta scutellata* | 1493             | CR: Dominical      | Jackson et al. (2014)            |
| *Taeniaptera trivittata*     | 1376             | USA: SC: Georgetown Co. | Jackson et al. (2014)   |
| *Micropeza* sp.              | 1405             | Argentina          | Jackson et al. (2014)            |

| Table 4. Percentage of homology from nucleotide base of *Mimegralla albimana* and three others species of Micropezidae Family from GenBank database |
|-----------------|-----------------|-----------------|-----------------|
| No. | Species                  | 1 | 2 | 3 | 4 | 5 |
| 1   | *Mimegralla albimana*    | ID |     | 88% | 88% | 88% | 92% |
| 2   | *Mimegralla coeruleifrons* | 88% | ID | 85% | 84% | 84% | 89% |
| 3   | *Hemichaeeta scutellata* | 87% | 84% | 84% | 86% | 89% | ID |
| 4   | *Taeniaptera trivittata* | 87% | 84% | 84% | 86% | 89% | ID |
| 5   | *Micropeza* sp.          | 87% | 84% | 84% | 86% | 89% | ID |

Figure 5. Phylogeny tree of *Mimegralla* spp. and others species of Micropezidae Family
LITERATURE CITED

Aczel, M.L. 1959. *Insects of Micronesia, Diptera: Neriidae and Micropezidae*. Bernice P. Bishop Museum, Hawaii. 90 p.

Balfas, R. 2002. Status Lalat Rimpang pada Tanaman Jahe dan Strategi Pengendaliannya (Status of Rhizome Fly at Ginger and their Control). *Jurnal Litbangtan (Journal of Research and Development on Agriculture)* 21: 32–37.

Balfas, R., Supriadi, N. Karyani, & E. Sugandi. 2000. Serangan *Mimegralla coeruleifrons* Macquart dan Peranannya dalam Membawa Patogen Bakteri Penyakit Layu (Attack of *Mimegralla coeruleifrons* Macquart and its Role in Bringing Pathogen of Wilt Disease. *Jurnal Penelitian Tanaman Industri (Research Journal of Industrial Plant)* 5: 123–127.

Folmer, O., M. Black, W. Hoeh, R. Lutz, & R. Vrijenhoce. 1994. DNA Primers for Amplification of Mitochondrial Cytochrom C Oxidase Subunit I from Diverse Metazoan Invertebrates. *Molecular Marine Biology and Biotechnology* 3: 294–299.

Ghorpade, S.A., S.S. Jadhav, & D.S. Ajri. 1983. Survey of Rhizome Fly on Turmeric and Ginger in Maharashtra. *Journal Maharashtra Agriculture University* 8: 292–293.

Ghorpade, S.A., S.S. Jadhav, & D.S. Ajri. 1988. Biology of Rhizome Fly, *Mimegralla coeruleifrons* Macquart (Micropezidae: Diptera) in India, a Pest of Turmeric and Ginger Crops. *Tropical Pest Management* 34: 48–51.

Hedges, S.B. 1992. The Number of Replications Needed for Accurate Estimation of the Bootstrap p-Value in Phylogenetic Studies. *Molecular Biology and Evolution* 9: 366–369.

Hennig, W. 1935. Revision der Tyliden (Dipt., Acalypt.) II Teil: Die außeramerikanischen Taeniiapterinae, die Trepidariinae und Tylinae. Allgemeines über die Tyliden. (Fortsetzung). *Konowia* 14: 192–216.

Joy, P.P., J. Thomas, S. Mathew, & B.P. Skaria. 1998. *Zingiberaceous Medicinal and Aromatic Plants*. Aromatic and Medicinal Plants Research Station, Odakkali, Asamannoor P.O., Kerala, India. 31p.

Karmawati, E., B. Baringbing, M. Iskandar, & T.E. Wahyono. 1990. Observasi Lalat Rimpang pada Pertanaman Jahe di K.P. Sukamulya. (Observation of Rhizome Fly on Ginger Plants at Sukamulya Village). *Media Komunikasi Penelitian dan Pengembangan Tanaman Industri (Media Communication Research and Development Industrial Crops)* 6: 84–86.

Maxwell-Lefroy, H. & F.M. Howlett. 1909. *Indian Insect Life-A Manual of the Insects of the Plains (Tropical India)*. Government Press, Calcutta. 486 p.

Nair, M.R.G.K. 1980. Pests of Ginger and Turmeric, p. 101–103. In M.K. Nair, T. Premkumar, P.N. Ravindran, & Y.R. Sarma (eds.), *Proceedings of the National Seminar on Ginger and Turmeric*. Calicut, 8–9 April 1980.

Pape, T., V. Blagoderov, & M.B. Mostovski. 1758. Order Diptera Linnaeus. In Z.Q. Zhang (ed.), *Animal Biodiversity: An Outline of Higher-Level Classification and Survey of Taxonomic Richness*. *Zootaxa* 3148: 222–229.

Sokal, R.H. & P.A. Sneath. 1963. *Principle of Numerical Taxonomy*. W.H. Freeman and Co., San Fransisco. 359 p.

Steyskal, G.C. 1964. Larvae of Micropezidae (Diptera) Including the Species that Bore Ginger Root. *Annal of the Entomological Society of America* 57: 292–296.