Enhanced photosynthetic output via dichroic beam-sharing

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Abstract  Microbial solar biofuels offer great promise for future sustainable food, fuels and chemicals but are limited by low productivities and a requirement for large land areas to harvest sunlight. A 71 % increase in combined photosynthetic activity was achieved by illuminating both Rhodobacter sphaeroides and Arthrospira (Spirulina) platensis from a single beam of simulated sunlight, divided using a dichroic mirror. Therefore, this technique is termed ‘dichroic beam-sharing’, in which the complementary action spectra of two different useful micro-organisms, belonging to green and purple groups, is exploited and allows a single beam of sunlight to be shared efficiently between separate photobioreactors. Because the action spectra of these two organisms are typical of large groups, this novel method could increase the productivity of photosynthetic micro-organisms in the production of diverse commodities.

Keywords  Arthrospira (Spirulina) platensis · Bioenergy · Biohydrogen · Biofuel · Dichroic beam-sharing · Rhodobacter sphaeroides

Introduction

Despite the concerns over fossil fuels (Kerr 2011), the adoption of biofuels is still limited, primarily by photosynthetic efficiency (PE). For crop plants, PE is just 0.5–1 % (Archer and Barber 2004) which, if biofuels are to replace fossil fuels so atmospheric carbon is stabilised by 2100, would necessitate a doubling of agricultural land area (Gurgel et al. 2008). In contrast, photosynthetic microorganisms can produce biofuels with a significantly greater PE (up to 9 % e.g. Guo et al. 2011) and can be cultivated in areas where crops cannot, such as on steep slopes, contaminated land, buildings or water, and provide a range of valuable co-products (Fig. 1) (Blankenship et al. 1995; Metzger and Largeau 2005). However, these benefits are currently offset by the cost of photobioreactors and so further improvements in PE are required for microbial solar biofuels to become viable. Attempts to enhance PE biochemically have, so far, met with limited success (e.g. Work et al. 2012 and...
references therein). In this study we demonstrate a complementary approach: dichroic beam-sharing.

Oxygenic photosynthetic microorganisms (including cyanobacteria, algae and chloroplasts; usually green) and anoxygenic purple non-sulfur bacteria have complementary action spectra and so their co-cultivation has the potential to improve overall photobioreactor efficiency. A co-culture of these is not suitable because oxygen generated by the green microorganisms inhibits the purple, and a bi-layer approach (Miyamoto et al. 1987) is limited by light scattering and absorption in the upper layer reducing light penetration to the lower. However, a dichroic mirror (DCM) has a reflection spectrum, \( R(\lambda) \), (simply related to the transmittance spectrum, \( T(\lambda) = 1 - R(\lambda) \)) that can be engineered by control of its surface coating and so can be used to direct to each microorganism its favoured part of the solar spectrum (Fig. 1).

In this study, we demonstrate this approach for *Arthrospira* (*Spirulina*) *platensis* (a foodstuff; Raoof et al. 2006) and *Rhodobacter sphaeroides* ZX-5 (a hydrogen producer; Redwood et al. 2012a) and show that up to 100 % increase in combined PE could be achieved.

**Methods**

**Solar simulator**

Xenon lamps are commonly used as solar simulators because their emission spectrum resembles that of the sun over most of the spectrum. This match is often further improved using specially designed ‘AM1.5’ filters. The American Society for Testing and Materials (ASTM) define a solar simulator as ‘class A’ if its emission spectrum differs from the solar spectrum by no more than 25 % over all of several non-equal bands. Filtered xenon lamps are class A over the visible and near-infrared region (400–1,000 nm) as a whole but are only class B (<40 % difference) between 800 and 900 nm because in this region they emit several strong spectral lines absent from the solar spectrum. The poorer spectral match in this region is not significant for many applications but is relevant to the current study because this region corresponds to the most important part of the action spectrum for purple non-sulfur bacteria. Hence, to ensure a good spectral match in this region, a filtered xenon lamp (LOT Oriel P/N LSO104; 150 W) was combined with a quartz tungsten halide (QTH) lamp (Oriel P/N 60000; 100 W), the emission spectrum of which is much closer to that of sunlight in the 800–900 nm spectral region. The spectrum of the Xe lamp was filtered to remove wavelengths longer than 725 nm whilst that of the QTH lamp was filtered to remove wavelengths shorter than 725 nm. A water-filter further improved the spectral match in the near-infrared by mimicking the absorbance of atmospheric H\( _2 \)O. The beams from each lamp were overlaid (Fig. 2) to produce a combined field of illumination that was a close match to the solar spectrum, including...
The spectra of the xenon lamp, the combined lamps and the solar spectrum were measured and compared using a spectrometer (Ocean Optics USB4000 VIS–NIR with Spectrasuite software in relative irradiance mode) calibrated using a blackbody source traceable to the UK National Physical Laboratory. The intensity of the combined light field could be adjusted by focusing or defocusing the beams from the lamps and was set to produce an intensity of 10 W/m², as measured by a PAR sensor (thermopile; Skye, UK, 400–1,000 nm), over an area of 750 cm². This intensity was chosen so that any light saturation effects could be neglected. Light limitation was confirmed practically; activity was proportionate to light intensity in the range 2.5–25 W/m² (see Supplementary Fig. 1).

Two identical dichroic mirrors (Thorlabs P/N DMLP638L) were used to spectrally divide the combined-lamp illumination so that the transmitted light was incident onto *A. platensis* whilst the reflected light was incident onto *R. sphaeroides*.

### Strains and culture conditions

*Arthrosira* (*Spirulina*) *platensis* (strain no. 86.79) was purchased from Sammlung von Algenkulturen der Universität Göttingen and maintained in 400 ml Raoof’s low-cost medium (Raoof et al. 2006) at 30 °C with continuous mechanical mixing and illumination (9 W fluorescent lamp; 14 W/m²) in a 1 l water-jacketed glass vessel. 300 ml of the culture (75 % of total) was discarded weekly and replaced with fresh Raoof’s medium. Chlorophyll was measured spectrometrically according to Raoof et al. (2006) and found to be proportional to dry weight (DW) and the OD₆₆₀ during the first 7 days of cultivation, after which, the chlorophyll concentration declined while DW continued to increase.

*Rhodobacter sphaeroides* ZX-5 was cultured as described previously (Redwood et al. 2012b), using sodium butyrate (30 mM) as the primary carbon source. To provide consistent stocks for H₂ production assays a single batch of *R. sphaeroides* cells was divided and aliquots were preserved at −80 °C. Cells were grown for 72 h (30 °C, intermittent manual shaking) in completely filled bottles under 75 W/m² from a 300 W halogen lamp, harvested by centrifugation (4,000 × g, 15 min) and resuspended in butyrate medium with 15 % (v/v) glycerol, before freezing in liquid N₂.

Experiments used a glass waterbath (30 °C) receiving illumination at 10 W/m² from below by the solar simulator, in which reactors (12 ml total internal volume) were positioned in the beam. For *A. platensis* growth tests, inocula were taken from an actively growing maintenance culture, diluted to an OD₆₆₀ of 0.04 (25 mg DW/l) with fresh medium, transferred (8 ml) into reaction vials and incubated under irradiance from the solar simulator with continuous mixing by bubbling with moistened air through 18G needles. OD₆₆₀ was recorded after 48 h. For *R. sphaeroides* H₂ production tests, aliquots were thawed and diluted to 1,000 mg DW/l (OD₆₆₀ = 0.302) with fresh butyrate medium, then dispensed (4 ml) into reaction bottles (12 ml internal volume), sealed with anaerobic stoppers, purged with argon (30 min) and incubated under irradiance from the solar simulator. The H₂ concentration in the headspace was measured after 36 h as described previously (Orozco et al. 2012).

### Results

Figure 3a, b compare the action spectra of green micro-organisms and purple bacteria with the transmission and reflectance spectra of the dichroic mirror (manufacturer’s data) respectively, showing a good match in both cases. Manufacturer’s data were
confirmed (Supplementary Fig. 2). The spectra of the filtered xenon lamp and the combined lamps relative to the solar spectrum are given in Fig. 3c, and show an improved spectral match in the 800–900 nm region, now class A rather than B according to ASTM classification.
Application of the dichroic mirror reduced the light supplied to *A. platensis* (transmitted light) by 62% but the growth only by 25%. The difference was statistically significant (*t* test, *P* < 5%). Figure 4a shows that, in relation to the total light intensity, the cultures with transmitted light were twice as efficient as those using a direct beam. Conversely, for H₂ production by *R. sphaeroides* the dichroic mirror reduced the supplied light (reflected) by 38% causing no significant difference in photosynthetic activity with an average productivity equivalent to 95% of the control (full spectrum). Figure 4b shows that, in relation to the total light intensity, the cultures with reflected light were 1.5 times as efficient as those using a direct beam. Figure 4c shows that the combined photosynthetic activity was equivalent to 170% in comparison to either reactor individually.

**Discussion**

The dichroic mirror provided almost complete reflection of blue and NIR wavelengths corresponding to the action maxima of purple bacteria (Fig. 3) and, accordingly, the mirror caused no detectable reduction in photosynthetic activity. Similarly, the mirror provided almost complete transmission of red wavelengths corresponding to the action maximum of green organisms. However, ~50% of light was lost through reflection in the 400–550 nm band, which represents ~50% of the total action for green organisms. Therefore, the overall reduction in active radiation was ~25% which corresponds to the observed 25% reduction in growth for *A. platensis*. Hence, the observed 25% reduction in activity is attributed to the partial reflection of useful “blue” (400–550 nm) light. Further mirror development of dichroic mirrors would aim to increase transmission in the “blue” band, potentially increasing the combined photosynthetic activity from 170% towards the limit of 200%.

The present study demonstrates the biological compatibility of dichroic beam-sharing with one pair of useful organisms. However, the action spectrum of *A. platensis* is typical of green micro-organisms of diverse genera and the chloroplasts of higher plants (Fig. 3), and so this technique should be applicable to other organisms with useful products. Further work is also required to establish to what degree the additional cost of using this technique would offset the benefit of doubled overall PE, set against food security and land use issues which may ultimately prove a critical socio-economic factor. This additional cost is likely to be minimised for photobioreactor designs utilising optical fibre light-delivery (Erickson et al. 2011) which would require only small dichroic mirrors. The optical damage threshold of the mirrors is typically very high (>1,000 MW/m²; manufacturer’s specification for the mirror used in this study), three orders of magnitude...
greater than the peak solar intensity at a concentration factor of 1,000.

**Conclusion**

Dichroic beam-sharing offers a practical opportunity for increasing photosynthetic activity by up to 100%. This study demonstrated a 71% increase in the combined photosynthetic activity of *A. platensis* and *R. sphaeroides*, which have been studied in the production of sustainable food and H₂ fuel, respectively. Losses were associated mainly with *A. platensis* and attributed to the partial reflection of useful ‘blue’ (400–550 nm) light by the dichroic mirror. The demonstrated principle indicates that solar photosynthetic activity could be increased without requiring additional land or sunlight, using existing technology to enhance the sustainable production of foods, fuels and chemicals via duplexed photobioreactor systems.

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