ABSTRACT

Aim: The study evaluates the phytochemical screening, atomic absorption spectroscopy (AAS), Gas chromatography–mass spectrometry (GC-MS) and antibacterial activities of aqueous and methanolic extracts of turmeric (Curcuma longa) rhizome against Escherichia coli (E. coli) and Staphylococcus aureus (S. aureus).

Place and Duration of Study: The study was carried out for six months in 2020 in Biochemistry Laboratory, Department of Chemical Sciences, College of Basic Sciences, Lagos State University of Science and Technology (LASUSTECH), Ikorodu, Lagos State, Nigeria.

Methodology: The phytochemical screening, GC-MS and AAS were determined using standard methods. Antibacterial activities were evaluated by disc diffusion and agar well diffusion methods. The minimum inhibitory concentrations (MIC) and minimum bactericidal concentration (MBC) were determined using standard procedures.

Results: The aqueous and methanolic extracts of turmeric (Curcuma longa) rhizome showed the presence of phytochemicals like tannins, flavonoids, alkaloids, reducing sugar and saponin. Mineral
composition analysis shows that the plant contains Na, Ca, Mg, K and Fe. Nineteen compounds were identified using GC-MS analysis of turmeric with a R-Turmerone being the most abundant with peak area of 50.05%. The results revealed that at 250 and 500 mg/mL for both aqueous and methanolic root extract of C. longa were sensitive to both organism, with zone of inhibition of 22.29±2.35, 29.56±2.23, 21.79±1.04 and 29.95±1.83 against E. coli and 22.31±1.59, 28.67±1.42, 22.96±0.96 and 30.13±1.94mm against S. aureus respectively. Azithromycin has zone of inhibition values that ranges from 19.35±1.02 to 32.03±1.23 mm for both organisms tested at 12.50 and 25.00 mg/mL respectively. E. coli and S. aureus were susceptible to erythromycin, ciprofloxacin, roceplin, and streptomycin and resistant to chloramphenicol and septrin for only S. aureus. The MIC of the aqueous and methanolic root extract of turmeric on E. coli and S. aureus were 62.50, 31.25, 31.250 and 15.625 mg/mL while their MBC values were 250.00, 62.500, 62.500 and 31.2500 mg/ml respectively. MBC/MIC values show that both extracts had bactericidal effects.

Conclusions: Curcuma longa has essential minerals, phytochemicals, antibacterial activity and may prevent pathogenic diseases caused by Escherichia coli and Staphylococcus aureus.

Keywords: Antibacterial activity; turmeric (curcuma longa); AAS and GC-MS analyses.

1. INTRODUCTION

Curcuma longa L. is commonly called turmeric and is a member of the ginger family. Turmeric is a golden spice derived from the rhizome of the Curcuma longa plant, which belongs to the Zingiberaceae family [1]. Curcuma longa has been used as the principal ingredient of dishes used from Nigeria, India and Bangladesh for its color, flavor, and taste. In West Africa it’s mainly used as a dye to color products, such as cotton cloth, tanned leather, palm fibers and thread to a golden yellow. The use of the yellow color of turmeric rhizome and other plant derivatives as dyes is on the increase toward replacing synthetic additives with natural compounds [2].

The yellow color of turmeric is due to the presence of three main curcuminoïds in the rhizome namely: curcumin, demethoxycurcumin, and bis-demethoxycurcumin. Dry turmeric contains: 5.1% oils, 6.3% proteins, 69.43% carbohydrates, 3.5% minerals, and other elements [3]. The bioactive chemical constituents in turmeric have been investigated. Approximately 235 compounds, primarily terpenoids and phenolics, have been identified from various species of turmeric, including 22 diarylheptanoids and diarylpentanoids, 8 phenylpropenes as well as other phenolics, 109 sesquiterpenes, 68 monoterpenes, 5 diterpenes, 4 sterols, 3 triterpenoids, 2 alkaloids, and 14 other compounds [4]. Curcuminoïds (mostly curcumin) and essential oils (primarily monoterpenes) are the major bioactive constituents showing different bioactivities. Calebin-A, vanillic acid, vanillin, quercetin, and other phenolic compounds have also previously been identified from turmeric [1, 5]. Studies have shown that aqueous extract of turmeric rhizomes exhibited antibacterial activity against S aureus ATCC 25923, Staphylococcus epidermis ATCC 12228, Escherichia coli ATCC 25922 and Klebsiella pneumoniae ATCC 10031 [6, 7].

Escherichia coli is a Gram negative, rod-shaped bacterium that is commonly found in the lower intestine of warm-blooded animals. E. coli causes severe infectious diseases associated with high rates of mortality and morbidity [8]. Staphylococcus aureus are Gram positive bacteria and they cause wide range of infections in human and animals. They are found on human skin and mucous membranes. However, it can also be found in other areas of human contact including soil, water, and food products [9]. They causes serious infections like bacteremia, septicemia, osteomyelitis, pneumonia, septic arthritis, wound sepsis, endocarditis, bone and joint infections, toxic shock syndrome and food poisoning [10]. The study evaluates the phytochemical screening, Gas chromatography–mass spectrometry (GC-MS), atomic absorption spectroscopy (AAS) and antibacterial activities of aqueous and methanolic extracts of turmeric (Curcuma longa) rhizome against Escherichia coli and Staphylococcus aureus.

2. MATERIALS AND METHODS

2.1 Collection and Identification of Plant Extract

The turmeric was purchased from Ikorodu market and was authenticated by Momoh Johnson from Department of Chemical Sciences (Biochemistry
2.2 Mineral Analysis of Turmeric

Two grams of turmeric was digested with 10 mL of aqua regia (Trioxonitrate (v) acid and hydrochloric acid in the ratio 1:3) and the total mixture of the plant and the acids were heated in a crucible for some minutes until brown fumes produced in the process disappeared leaving white fumes. It was then later filtered with filter paper into universal bottle. The micro and macro elements present in the turmeric sample were determined using AGILENT 720 ICP-OES Atomic Absorption Spectrophotometer (AAS). The minerals that were analyzed for were: Ca, Fe, K, Na, Mg, Cu, Zn and Pb.

2.3 Gas Chromatography-Mass Spectrometry (GC-MS) analysis of Turmeric

GC-MS analysis of the Curcuma longa rhizome was carried out on an Agilent technology 7890 GC system equipped with a mass spectrometric detector (MSD) as described by Momoh et al. [11].

2.3.1 Detection of components

Analysis of mass spectrum GC-MS was conducted by the database of the National Institute Standard and Technique (NIST) which contained more than 62,000 patterns. The spectrum of the unidentified compound was compared with the spectrum of the identified compounds stored in the National Institute Standard and Technique library. The names, molecular weight, structure of the compounds in the test material were ascertained.

2.3.2 Preliminary phytochemical analysis

The presence of saponin, tannins, alkaloids, flavonoids, anthraquinones, glycosides and reducing sugars were determined by qualitative methods [12-14]. The simple qualitative analyses of the extract were based on the intensity of the colour change.

2.3.3 Test organisms

To study the antimicrobial activity of aqueous and methanolic root extracts of turmeric (Curcuma longa) extract against two bacterial strains (Escherichia coli (Gram negative ATCC # 25922) and Staphylococcus aureus (Gram positive, clinical isolates ATCC #6538) were used for the study. The two microorganisms were maintained at 4°C on Nutrient Agar slant in the Department of Chemical Sciences and fresh subcultures were made before use.

2.3.4 Inoculum preparation

A loopful of isolated colonies of the two organisms were inoculated separately into 4 mL of peptone water, incubated at 37°C for 4 hours. These actively growing bacterial suspensions were then adjusted with peptone water to obtain turbidity visually comparable to that of 0.5 McFarland standards using standard procedure [8]. The 0.5 McFarland standard was prepared by mixing 0.5mL of 1.75% (w/v) barium chloride dehydrate (BaCl₂. 2H₂O) with 99.5 mL of 1% (v/v) H₂SO₄. This turbidity was equivalent to approximately 1 x 10⁸ colony forming units per mL (CFU/mL) [8].

2.3.5 Antibiotic susceptibility testing

The susceptibility of the organisms to different antibiotics were tested using the disk diffusion method as described [15, 16]. On freshly prepared Mueller Hinton agar and standardized by the method of Famuyide et al. [17] and National Committee for Clinical Laboratory Standard (NCCLS), 2000 [18] using some selected antibiotics namely: Roceplin (25μg/ disk), chloramphenicol (30μg/ disk), streptomycin (30μg/ disk), erythromycin (10μg/ disk), ciprofloxacin (10μg/ disk), and septrin (30μg /disk). For each combination of the antibiotics and the bacterial strains, the experiment was performed in triplicate.

2.3.6 Determination of diameter of zone of inhibition using agar well diffusion method

Agar well-diffusion method was employed to determine the antimicrobial activity of aqueous and methanolic root extracts of turmeric (Curcuma longa) extract. Eighteen hours of broth culture of the two microorganisms were suspended into the sterile nutrient broth. It was standardized by gradually adding 9% normal saline to compare its turbidity to McFarland standard of 0.5 which is approximately 1 x 10⁸ colony forming units per mL. Petri dishes were prepared by loading about 25 mL of an autoclaved nutrient agar on sterile plates and left to solidify. Then, the surface of each plate was
drilled using a sterile cork borer (6 mm) and 3 wells were punched out on each plate. A total of 100 μL of a standardized culture (adjusted to 0.5 McFarland) of the two organisms were added into the different agar plates followed by loading of 100 μL of the aqueous and methanolic root extracts of turmeric extract in the wells and allowed to diffuse at room temperature for 2 hours. The plates were incubated at 37ºC for 18-24 hours for bacterial pathogens. The diameters of the inhibition zone (mm) were measured. The susceptibility of the two different organisms (Staphylococcus aureus and Escherichia coli) to aqueous and methanolic extracts of turmeric were assayed using standard method [8]. The experiment was repeated thrice, for each replicate, the readings were taken in three different fixed directions and the average values were recorded [8]. The inhibitory responses were classified as potent response, +++++, zone diameter >30 mm; strong response, +++, zone diameter between 21-30 mm; moderate response, ++, zone diameter between 16-20 mm; weak response, +, zone diameter between 10-15 mm; and little or no response, zone diameter <10 mm [19].

2.3.7 Minimum inhibitory concentration (MIC) of aqueous and methanolic root extracts of turmeric (Curcuma longa)

Minimum inhibition concentration is the lowest extract concentration that inhibited the growth of the test organisms as indicated by the absence of visible turbidity in the tube compared with the control tubes. The MIC of the aqueous and methanolic root extracts of turmeric rhizome extracts were determined according to standard method [8]. The MIC of the aqueous and methanolic root extracts of turmeric extract were assayed using serial dilution method. Briefly, a total of 1 mL of Mueller-Hinton broth was poured to a set of different test tubes and autoclaved. Subsequently, 1 ml of 100% aqueous and methanolic root extracts of turmeric (2g/mL) were poured to the first separate test tubes to make a concentration of 50%, and two-fold serial dilutions were made by transferring 1 mL from one tube to another to get the following series: 50%, 25%, 12.5%, 6.25%, 3.12%, 1.56%, 0.78%, 0.39% etc. Then, an overnight broth culture of the different test organisms were adjusted to McFarland turbidity standard and 100 μL of the different cell suspensions were added to each of the separate tubes. The tubes were incubated aerobically at 37°C for 18 hours. Negative control tube was made by pouring 1mL of normal saline instead of the aqueous and methanolic root extracts of turmeric extract. The lowest concentration of the dilution without bacterial growth was considered as the minimum inhibition concentration.

2.3.8 Minimum Bactericidal Concentration (MBC) of the aqueous and methanolic root extracts of turmeric extract

The MBC of the aqueous and methanolic root extracts of Turmeric extract were carried out by standard method [8]. In the procedure, 0.1 mL aliquots of test samples taken from the non-turbid tubes of the minimum inhibition concentration assay test tubes were sub-cultured onto nutrient agar plates. The resulting plates were then incubated aerobically at 37°C for 24 hours. The lowest

3. RESULTS

| Elements | Conc. in mg/L | %RSD |
|----------|--------------|------|
| Na       | 1.3638 ±0.002 | NIA  |
| Mg       | 0.8025±0.001  | NIA  |
| Ca       | 0.7973 ±0.001 | 1.14 |
| K        | 0.0018 ±0.000 | 0.53 |
| Fe       | 1.0109 ±0.002 | 0.4  |
| Zn       | 0.0484 ± 0.000 | 30.3 |
| Ag       | 0.0019 ± 0.000 | 23.6 |
| As       | 0.0097 ± 0.000 | 50.0 |
| Cd       | 0.0059 ±0.001 | 80.0 |
| Co       | 0.0086 ± 0.000 | 140.5|
| Cu       | 0.0061 ± 0.000 | 307.1|
| Ni       | 0.0016 ±0.000  | 12.1 |
| Pb       | -0.0028 ± 0.000 | 363.8|

NIA indicate not available. Values are mean ± standard deviation for triplicate determinates
Table 2. Compounds found in the turmeric analyzed using Gas Chromatography–Mass Spectrometry

| Pk# | RT | Name of the compound | Molecular Formulae | Molecular Weight (g/mol) | Peak Area (%) | Ref# | CAS# |
|-----|----|----------------------|--------------------|--------------------------|--------------|------|------|
| 1   | 8.358 | Benzene, 1-(1,5-dimethyl-4-hexenyl)-4-methyl- | C₁₅H₂₂ | 202.3352 | 1.67 | 66865 | 000644-30-4 |
| 2   | 8.877 | Cyclohexene, 3-(1,5-dimethyl-4-hexenyl)-6-methylene-, [S-(R*,S*)]- | C₁₅H₂₄ | 204.3511 | 1.88 | 68734 | 020307-83-9 |
| 3   | 9.611 | Benzene, 1-ethyl-3,5-dimethyl- | C₁₀H₁₄ | 134.2182 | 1.85 | 15214 | 000934-74-7 |
| 4   | 9.987 | Benzene, 1-(1,5-dimethylhexyl)-4-methyl- | C₁₅H₂₄ | 204.3511 | 5.83 | 68654 | 001461-02-5 |
| 5   | 10.811 | aR-Turmerone | C₁₅H₂₀O | 216.3187 | 50.05 | 79922 | 000532-65-0 |
| 6   | 11.244 | 2-Methyl-6-(4-methylene cyclohex-2-en-1-yl) hept-2- en-4-one | C₁₅H₂₂O | 218.335 | 20.03 | 81679 | 082508-14-3 |
| 7   | 11.592 | 3-Methyl-6-(6-methylhept-5-en-2-yl)-cyclohex-2- enone | C₁₅H₂₄O | 220.3505 | 1.42 | 83600 | 066964-98-5 |
| 8   | 11.696 | Gamma-Terpine | C₁₀H₁₆ | 136.2340 | 1.12 | 16078 | 000099-85-4 |
| 9   | 11.787 | Binaparicyl | C₁₅H₁₈N₂O₆ | 322.317 | 1.26 | 180130 | 000485-31-4 |
| 10  | 11.892 | Benzonitrile, 3-hydroxy | C₂H₇NO | 119.1207 | 1.30 | 9294 | 000873-62-1 43 |
| 11  | 11.925 | (E)-Atlantone | C₁₅H₂₂O | 218.3346 | 2.14 | 81630 | 108645-54-1 |
| 12  | 12.096 | Cumene, angelate, o | C₁₄H₁₈O₈ | 218.29 | 0.98 | 81511 | 1000383-67-2 38 |
| 13  | 12.187 | 3,5-Dimethylanisole | C₉H₁₂O₂ | 136.1910 | 1.29 | 16778 | 000874-63-5 |
| 14  | 12.296 | Prop-2-ynyl (E)-2-methylbut-2-enoate | C₅H₉O₂ | 138.16 | 1.99 | 17804 | 1000373-72-5 22 |
| 15  | 13.106 | Diglycolic acid, nonyl 3-phenylpropyl ester | C₂₂H₃₆O₅ | 378.5 | 3.35 | 241428 | 1000382-18-0 35 |
| 16  | 13.287 | (S)-(S)-3-Methyl-6-((S)-6-methyl-4-oxohept-5-en-2-yl)cyclohex-2-enone | C₁₅H₂₀O₂ | 234.3398 | 0.88 | 96682 | 949081-10-1 |
| 17  | 13.696 | But-2-enoamide, N-ethyl-N-(3-methylphenyl)-3-methyl- | C₁₄H₁₉NO | 217.31 | 1.30 | 80637 | 1000308-23-6 38 |
| 18  | 14.196 | Cyclohexanecarboxylic acid, 4-nitrophenyl ester | C₁₃H₁₅NO₄ | 249.2625 | 1.26 | 110342 | 1000307-70-8 |
| 19  | 15.601 | 9,12-Octadecadienoic acid (Z,Z)- | C₁₈H₃₂O₂ | 280.4455 | 0.40 | 140138 | 000060-33-3 |
Fig. 1. Gas-Chromatography–Mass Spectrometry chromatogram of turmeric (*Curcuma longa*)

Fig. 2a. Mass spectrum of Benzene, 1-(1,5-dimethyl-4-hexenyl)-4-methyl- structure.

Fig. 2b. Mass spectrum of Cyclohexene, 3-(1,5-dimethyl-4-hexenyl)-6-methylene-, [S-(R*,S*)]- structure

Fig. 2c. Mass spectrum of Benzene, 1-ethyl-3,5-dimethyl- structure

Fig. 2d. Mass spectrum of Benzene, 1-(1,5-dimethylhexyl)-4-methyl- structure
Fig. 2e. Mass spectrum of a R-Turmerone structure

Fig. 2f. Mass spectrum of 2-Methyl-6-(4-methylene)cyclohex-2-en-1-yl)hept-2-en-4-one structure

Fig. 2g. Mass spectrum of 3-Methyl-6-(6-methylhept-5-en-2-yl)cyclohex-2-enone structure.

Fig. 2h. Mass spectrum of gamma-Terpinene structure

Fig. 2i. Mass spectrum of Binapacryl structure

Fig. 2j. Mass spectrum of (E)-Atlantone structure

Fig. 2k. Mass spectrum of (E)-Atlantone structure

Fig. 2l. Structure of Cumenyl angelate, o-
Fig. 2m. Mass spectrum of 3,5-Dimethylanisole
Structure

Fig. 2n. Structure of Prop-2-ynyl (E)-2-methylbut-2-enoate

Fig. 2o. Structure of Diglycolic acid, nonyl 3-phenylpropyl ester

Fig. 2p. Mass spectrum of (S)-3-Methyl-6-((S)-6-methyl-4-oxohept-5-en-2-yl)cyclohex-2-enone structure

Fig. 2q. Structure of But-2-enamide, N-ethyl-N-(3-methyl phenyl)-3-methyl-

Fig. 2r. Mass spectrum of Cyclohexanecarboxylic acid, 4-nitrophenyl ester structure
Fig. 2. Mass spectrum and structure of 19 different compounds obtained during GC-MS analysis of *Curcuma longa*

Table 3. Phytochemistry of aqueous and methanolic extracts of turmeric (*Curcuma longa*)

| Phytochemical constituent | Test performed                  | Water | Methanol |
|---------------------------|----------------------------------|-------|----------|
| Alkaloids                 | Mayer’s test                     | +     | +        |
| Tannins                   | Ferric chloride test             | +     | +        |
| Saponins                  | Froth test                       | +     | +        |
| Flavonoids                | Lead acetate test                | +     | +        |
| Simple phenolics          | Ferric chloride test             | +     | +        |
| Steroid                   | Liebermann-Burchard’s test       | -     | +        |
| Protein                   | Biuret test                      | +     | +        |
| Reducing sugar            | Fehling’s solution test          | +     | +        |
| Carbohydrate              | Molisch’s test                   | +     | +        |

Notes: (+) indicates present, (-) indicates absent

Fig. 3a. Zone of inhibition at 250 mg/ml of the aqueous extract of *Curcuma longa* against *Escherichia coli*

Fig. 3b. Zone of inhibition at 250 mg/ml of the methanolic extract of *Curcuma longa* against *Escherichia coli*
Fig. 3c. Zone of inhibition at 25 mg/ml for azithromycin solution against *Staphylococcus aureus*

Fig. 3d. Zone of inhibition at 25 mg/ml for azithromycin solution against *Escherichia coli*

Fig. 3. Zone of inhibition of azithromycin solution, aqueous and methanolic extracts of *Curcuma longa* rhizome against *Staphylococcus aureus* and *Escherichia coli* at 25 and 250 mg/mL

Table 4. Antimicrobial susceptibility pattern of standard antibiotics agent against *Escherichia coli*

| Antibiotic sensitive disc | Concentration (µg) | Diameter of zone of inhibition (mm) | Interpretation |
|--------------------------|-------------------|---------------------------------------|----------------|
| Chloramphenicol (CH)     | 30                | 14.80 ± 0.37                          | +              |
| Ciprofloxacin (CPX)      | 10                | 19.16 ± 1.250                         | ++             |
| Streptomycin (S)         | 30                | 21.37 ± 1.32                          | +++            |
| Roceplin (R)             | 25                | 23.00 ± 1.95                          | +++            |
| Septrin (SXT)            | 30                | 18.07 ± 0.64                          | ++             |
| Erythromycin (E)         | 10                | 20.00 ± 1.66                          | ++             |

Table 5. Antimicrobial susceptibility pattern of standard antibiotics agent against *Staphylococcus aureus*

| Antibiotic sensitive disc | Concentration (µg) | Diameter of zone of inhibition (mm) | Interpretation |
|--------------------------|-------------------|---------------------------------------|----------------|
| Chloramphenicol (CH)     | 30                | 15.13 ± 0.60                          | +              |
| Ciprofloxacin (CPX)      | 10                | 18.55 ± 0.27                          | ++             |
| Streptomycin (S)         | 30                | 18.21 ± 0.17                          | ++             |
| Roceplin (R)             | 25                | 18.00 ± 0.58                          | ++             |
| Septrin (SXT)            | 30                | 16.83 ± 0.24                          | ++             |
| Erythromycin (E)         | 10                | 16.64 ± 0.39                          | ++             |

Concentration of the aqueous and methanolic root extracts at which no colonies of *Escherichia coli* and *Staphylococcus aureus* were taken as the minimum bactericidal concentration. The results were compared with that of control tube using sterilized distilled water. The experiment was performed in triplicate. The MBC was taken as the concentration of the aqueous and methanolic root extracts of turmeric that did not show any growth on a new set of agar plates. The lowest MIC value that revealed no visible growth was regarded as the minimum bactericidal concentration. The MBC/MIC value was also calculated as either bactericidal or bacteriostatic.

3.1 Statistical Analysis

All analyses were carried out in triplicate determination and results were expressed as mean ± SD. Students t-test was used for comparison. The data analysis was done using one way analysis of variance (ANOVA) Post Hoc Turkey Graph Pad prism computer software version 5.01. *P*-value < 0.05 was considered significant.
Table 6. Zone of inhibition of *Curcuma longa* aqueous and methanolic extract against *Escherichia coli* and *Staphylococcus aureus*

| Test organisms                  | Aqueous extract of Tumeric concentration (mg/ml) | Zone of inhibition for aqueous extract of Tumeric (mm) | Methanolic extract of Tumeric concentration (mg/mL) | Zone of inhibition of methanolic extract of Tumeric (mm) | Concentration of azithromycin solution used (mg/mL) | Zone of inhibition of azithromycin solution (mm) |
|---------------------------------|-------------------------------------------------|-----------------------------------------------------|---------------------------------------------------|-----------------------------------------------------|---------------------------------------------------|--------------------------------------------------|
| *Escherichia coli*              | 250                                             | 22.29±2.35<sup>b</sup>                              | 250                                               | 21.79±1.04<sup>b</sup>                              | 12.50                                             | 21.67±1.04<sup>c</sup>                            |
| *Staphylococcus aureus*         | 250                                             | 22.31±1.59<sup>b</sup>                              | 250                                               | 22.96±0.96<sup>b</sup>                              | 12.50                                             | 19.35±1.0<sup>c</sup>                             |
| *Escherichia coli*              | 500                                             | 29.56±2.231<sup>a</sup>                             | 500                                               | 29.95±1.83<sup>a</sup>                              | 25                                                | 32.03±1.23<sup>a</sup>                            |
| *Staphylococcus aureus*         | 500                                             | 26.75±1.42<sup>a</sup>                              | 500                                               | 30.13±1.94<sup>a</sup>                              | 25                                                | 28.76±0.92<sup>a</sup>                            |

Comparisons across the column was done using One way ANOVA Post Hoc Turkey test. The superscript a has the highest value followed by b and c has the lowest value. *A. P<0.05 was considered statistically significant*.

Table 7. Minimum inhibitory concentration (MIC) and Minimum bactericidal concentration (MBC) of *Curcuma longa* extracts against *Escherichia coli* and *Staphylococcus aureus*

| ORGANISMS                      | *Escherichia coli* | *Staphylococcus aureus* |
|--------------------------------|--------------------|-------------------------|
| MIC for aqueous extract of Tumeric (mg/mL) | 62.500             | 31.250                  |
| MIC for methanolic extract of Tumeric (mg/mL) | 31.250             | 15.6250                 |
| MBC for aqueous extract of Tumeric (mg/mL)   | 250.00             | 62.500                  |
| MBC for methanolic extract of Tumeric (mg/mL) | 62.500             | 31.250                  |
| MBC/MIC for aqueous extract of tumeric       | 4.00               | 2.00                    |
| MBC/MIC for aqueous extract of tumeric       | 2.00               | 2.00                    |
4. DISCUSSION

The result of this study shows that sodium (1.3638 ± 0.002) was the most abundant element present in the turmeric followed by iron (1.0109 ± 0.002), magnesium (0.8025±0.001) and calcium (0.7973 ±0.001). Other elements like: K, Zn, Ag, As, Cd, Co, Cu, Ni and Pb were found to be present in very small quantities that are not significant (Table 1). Enemor et al. [20] study shows that *Curcuma longa* rhizomes had higher contents of calcium, magnesium, potassium and sodium in parts per million (ppm) at 38.68 ± 0.114, 19.75 ± 0.001, 9.20 ± 0.002 and 7.06 ± 0.014 respectively. Their result is similar to the result obtained in our study. Ogidi et al. [21] study indicates that sodium element helps in the treatment of heart diseases. In a research work carried out by Hartwig [22], magnesium plays fundamental roles in genomic stability and DNA repair processes. Other studies show that magnesium activates over 300 different enzymes and thus participates in many metabolic processes, which makes it an important micronutrient, and also helps in electrolyte transport across cell membranes [23, 24]. Okwu, [25] study shows that Magnesium and calcium are used for the formation of strong bone and teeth. The presence of Calcium ions help to convert prothrombin to thrombin during blood coagulation and are also used in milk clotting. Calcium ions help in the activation of numerous enzymes activities in the body. Iron is an important element that is used in the formation of red blood cells.

Fig. 1 shows the Gas-Chromatography–Mass Spectrometry chromatogram of *Curcuma longa* (turmeric). A total of 19 compounds were identified consisting of 2 prominent compounds and 17 minor compounds (Table 2). The two major compounds and their percentage abundance are: ar-Turmerone (RT=10.811 and peak area = 50.05%) and 2-Methyl-6-(4-methylene-cyclohex-2-en-1-yl)-hept-2-en-4-one (RT=11.244 and peak area = 20.03%). ar-Turmerone (peak area = 50.05%), is the most abundant compound followed by 2-Methyl-6-(4-methylene-cyclohex-2-en-1-yl)-hept-2-en-4-one (peak area = 20.03%). Ar-turmerone, the major volatile component in the rhizome, showed potent α-amyrase (IC₅₀ of 24.5 μg) and α-glucosidase (IC₅₀ of 0.28 μg) inhibition [26]. Hoi-Seon, [27] study shows that at 2 and 1 mg/disk, ar-turmerone strongly inhibited the growth of *C. perfringens* and moderately inhibited the growth of *E. coli* without any adverse effects on the growth of four lactic acid bacteria (*B. adolescentis, B. bifidum, B. longum, and L. casei*) at 2 mg/disk. Marilyana et al. [28] study shows that ar-turmerone was not active against *Escherichia coli* ATCC 25922, *Staphylococcus aureus* ATCC 25923, *Pseudomonas aeruginosa* ATCC 27853 and *Klebsiella pneumoniae* ATCC 13883 using disc diffusion method. Hucklenbroich et al [29] experimental findings show that in-vitro and in-vivo study of aromatic (ar-) turmerone induces neural stem cells (NSC) proliferation. Ar-turmerone support regeneration in neurologic disease. Studies have shown that antitumor properties, exerted via the induction of apoptosis [30] and inhibition of tumor cell invasion have been attributed to ar-turmerone [31]. Park et al. [31] study shows that ar-turmerone also possesses anti-inflammatory properties resulting from the blockade of key signaling pathways in microglia. Microglia activation is a hallmark of neuroinflammation and is associated with various neurologic disorders, including neurodegenerative diseases [32, 33] and stroke [34, 35].

The preliminary qualitative analysis of the different secondary metabolites present in both extracts of *Curcuma longa* was investigated. The aqueous and methanolic root extract of turmeric showed that they contain alkaloids, flavonoids, tannins, saponins, simple phenolics, steroids (steroids was absent in the aqueous root extract), protein, reducing sugar and carbohydrate. Flavonoids are generally more soluble in water or polar solvents because they bonds with hydroxyl groups. Glycosides are compounds that contain sugar and non-sugar components. Saponins are generally in the form of glycosides so they tend to be polar. Saponins are surface active compounds that produce foam if shaken in water. This happens because saponins have polar and non-polar groups that will form micelles. When the micelle is formed the polar group will face out while the non-polar groups face inside so it looks like foams. Tannins which are phenolic compounds tend to dissolve in water and tend to be polar. Terpenoids are fat soluble. One of the terpenoids which has the potential as an antimicrobial is triterpenoid, while steroids are fat groups and are part of the triterpenoid.

*Staphylococcus aureus* and *Escherichia coli* were selected for the study and tested against some selected antibiotics, aqueous and methanolic extracts of turmeric. Rocreplin and streptomycin antibiotics showed strong response
with zone diameter between 21-30 mm against *Escherichia coli*, ciprofloxacin, septrin and erythromycin showed moderate response with zone diameter between 16-20 mm while chloramphenicol showed weak response with zone diameter less than 16 mm (Table 4). Ciprofloxacin, streptomycin, roceplin, septrin and erythromycin antibiotics showed moderate response to *Staphylococcus aureus* with zone diameter between 16-20 mm, while chloramphenicol showed weak response with zone diameter less than 16 mm (Table 5). In the present study, the aqueous and methanolic extracts of turmeric extract exhibited strong response antimicrobial activity against *Escherichia coli* and *Staphylococcus aureus* with zone of inhibition ranging from 21.79±1.04 to 30.13±1.94 at concentration of 250 and 500 mg/dl respectively. The study shows that 500 mg/ml aqueous and methanolic rhizome extracts of turmeric were sensitive to the tested organisms and were significantly (P< 0.004) different from the 250 mg/ml of the different extracts used in the study. *Staphylococcus aureus* was more sensitive to the two different extracts used in the study (Table 6).

Azithromycin solution at 25 mg/ml showed potent response against *Escherichia coli* with zone diameter greater than 30 mm and strong response against *Staphylococcus aureus* with zone diameter less than 30 mm. At 12.50 mg/ml, azithromycin exhibited strong response against *Escherichia coli* and moderate response against *Staphylococcus aureus* (Table 6).

Kim et al. [36] and Chandrana et al. [37] studies reported that turmeric extract was effective against *Bacillus subtilis*, *Escherichia coli*, and *Staphylococcus aureus* which may be due to the presence of curcuminoid which is a phenolic compound. Negi et al. [38] research work reported that curcune and turmerone components of turmeric possessed better antibacterial activity against a wide range of microbes including: *Bacillus subtilis, Bacillus cereus, Staphylococcus aureus, Bacillus coagulans, Psuedomonas aeruginosa* and *Eschericha coli*. The antimicrobial activity of turmeric is reported to be due to the presence of curcumindins, turmerol, curcumin, veleric acid, essential oil and turmeric oil [39-41].

The antibacterial activity of aqueous and methanolic extracts of turmeric against *S. aureus* and *E. coli* pathogens were investigated for their MIC and MBC values. MIC or MBC values are the lowest concentration of an antimicrobial agent necessary to inhibit bacterial growth or kill bacteria respectively [8]. Aderere et al. [8] study shows that MIC test is important in the laboratory to confirm the resistance of microorganisms to an antimicrobial agent and also used it to monitor the activity of new antimicrobial agents. The aqueous and methanolic rhizome extracts of turmeric have MIC values of 62.500 and 31.250 mg/ml for *E. coli*, 31.250 and 15.6250 mg/ml for *S. aureus* respectively. The two extracts also have MBC values of 250.00 and 62.500 for *E. coli*, 62.500 and 31.250 mg/ml for *S. aureus* respectively (Table 7). The result of this study showed that the gram-negative bacterium (*Escherichia coli*) was less susceptible to the two rhizome extracts when compared to the gram-positive bacterium (*Staphylococcus aureus*).

Study has shown that curcumin, the active constituent of turmeric exhibited inhibitory activity on methicillin-resistant *S. aureus* strains (MRSA) with MIC values ranging from 125–250 μg/mL [38]. This compound also displayed good antibacterial activity with MIC values ranging from 5 to 50 μg/mL against 65 clinical isolates of *Helicobacter pylori* [42].

Different studies show that aqueous extract of turmeric rhizomes exhibited antibacterial effects against *S. aureus* ATCC 25923, *Staphylococcus epidermis* ATCC 12228, *Escherichia coli* ATCC 25922 and *Klebsiella pneumoniae* ATCC 10031 with MIC values ranging from 4–16 μg/mL [6, 7]. The methanolic extract of *C. longa* has inhibitory effects against *S. aureus* (MIC value 128 μg/mL) and *Bacillus subtilis* (MIC value of 16 μg/mL) [43]. In another study carried out by Lawhavini et al. [44], the methanol and hexane extracts of turmeric also showed antibacterial effect against an array of bacteria including, *Vibrio vulnificus, Vibrio harveyi, Vibrio parahaemolyticus, Vibrio alginolyticus, Vibrio cholerae, Bacillus cereus, B. subtilis, Aeromonas hydrophila, S. aureus, Streptococcus agalactiae, Staphylococcus epidermidis, Edwardsiella tarda* and *Staphylococcus intermedius* with MIC values ranging from 125–1000 μg/mL [44]. All the above studies support the result obtained in our research concerning the antibacterial activity of turmeric against *Staphylococcus aureus* and *Escherichia coli*. Aderere et al., [8] 2020 study has shown that calculated MBC/MIC ratio is bactericidal if the values of MBC/MIC ratio are less than or equal to 4 and bacteriostatic if the MBC/MIC ratio is > 4. The aqueous and methanolic extracts of turmeric rhizome have...
bactericidal effects on *Escherichia coli* and *Staphylococcus aureus* respectively.

5. CONCLUSION

*Curcuma longa* has essential minerals, phytochemicals and other natural therapeutic agents that possess antibacterial activity against *Escherichia coli* and *Staphylococcus aureus* and may prevent pathogenic diseases caused by these organisms.

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COMPETING INTERESTS

Authors have declared that no competing interests exist.

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