Therapeutic effect of modified zengye decoction on primary Sjogren’s syndrome and its effect on plasma exosomal proteins

Yimin Zeng†, Xin Peng†, Yun Wang, Lei Hou, Wukai Ma and Peng Yang*

The Second Affiliated Hospital, Guizhou University of Traditional Chinese Medicine, Guiyang, China

Background: Modified Zengye Decoction (MZD), a traditional Chinese medicine, is an effective treatment for patients with primary Sjögren’s syndrome (pSS).

Purpose: To evaluate the efficacy of MZD and investigate its effect on plasma exosomal proteins.

Methods: Eighteen pSS patients were treated with MZD for 2 weeks. The therapeutic effect was evaluated by observing the changes in clinical symptoms, laboratory parameters, and plasma cytokines before and after treatment. Then, the differentially expressed proteins (DEPs) in the plasma exosomes before and after treatment were identified via label-free proteomics, while Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) enrichment were used to analyze the possible biological functions and signaling pathways involved in the exosomal DEPs.

Results: MZD can effectively relieve the clinical symptoms of pSS patients, downregulate the plasma IgG and IgM levels, and inhibit plasma cytokine production. Thirteen DEPs were identified via label-free proteomics in the plasma exosomes before and after MZD treatment, of which 12 were downregulated proteins. GO analysis showed that these downregulated proteins were mainly related to the insulin response involved in dryness symptoms and the Gram-negative bacterial defense response and proteoglycan binding involved in infection. KEGG enrichment analysis showed that these downregulated proteins were primarily associated with the porphyrin metabolism involved in osteoarthrosis and the NF-κB and TLR4 pathways involved in infection.

Abbreviations: American College of Rheumatology (ACR); ammonium bicarbonate (ABC); automatic gain control (AGC); biological process (BP); cartilage oligomeric matrix protein (COMP); cellular compartment (CC); ceruloplasmin (CP); differentially expressed proteins (DEPs); European League Against Rheumatism (EULAR); factor-binding protein 2 (IGFBP2); ficolin-2 (FCN2); false discovery rate (FDR); fold change (FC); Gene Ontology (GO); higher-energy collisional dissociation (HCD); Kyoto Encyclopedia of Genes and Genomes (KEGG); label-free quantification (LFQ); lipopolysaccharide–binding protein (LBP); Modified Zengye Decoction (MZD); molecular function (MF); peptide-spectrum match (PSM); primary Sjögren’s syndrome (pSS); Traditional Chinese medicine (TCM); ultracentrifugation (UC); dithiothreitol (DTT).
**1 Introduction**

Primary Sjögren’s syndrome (pSS) is an autoimmune disease caused by damage to the exocrine gland tissue, characterized by dry eyes and a dry mouth. Viral infections and hormonal factors cause abnormal epithelial cell immune function in genetically susceptible hosts, recruiting T cells, B cells, and dendritic cells that target the diseased tissue (Arvaniti et al., 2019; Maslińska, 2019). This induces the immune system to produce an immune response against its own exocrine gland epithelial cells, generating many autoantibodies and cytokines, resulting in the destruction and dysfunction of local exocrine gland tissue, while multiple systems and tissue lesions may be involved in severe cases (Riviere et al., 2020). The current treatment for the disease remains largely palliative, focusing on stimulating the secretion of the remaining exocrine glands or replacing the exocrine fluid of saliva and tears (Chowdhury et al., 2021; Fox et al., 2021). However, the existing treatments are primarily unsuccessful in improving symptoms and disease progression.

Traditional Chinese medicine (TCM) typically consists of various herbs to treat multiple conditions (Ma et al., 2016; Lu et al., 2021). It is beneficial for treating multi-system and multi-tissue diseases (Yuan et al., 2016; Shi et al., 2020). Zengye Decoction is a classic TCM prescription containing Scrophularianingpoesisis Henel (Xuanshen), Ophiopogon japonicus (Maidong) and Rheumannagiunina (Shengdihuang), which increase fluid and alleviates dryness. Zengye Decoction was first mentioned in volume II of the classic medical “Wenbing Tiaoxi,” and is mainly used to treat dryness, including constipation, thirst, and a dry tongue. Some reports showed that Zengye decoction or modified Zengye decoction could effectively alleviate SS symptoms (Wu et al., 2014; Jia et al., 2021; Wei et al., 2021). According to traditional Chinese medicinal theory, the patients in this study displayed a “toxic accumulation and blood stasis” during the early SS stage. Therefore, this study added detoxifying Artemisia carvifolia Buch (Qinghao), Oldenlandiadifissa (Baihuashecao), Andrographispaniculata (Chuanxinlian), and Gnaphalium affine (Shuqacao) during the clinical application of Zengye Decoction for SS treatment to obtain Modified Zengye Decoction (MZD). MZD displays detoxification properties by increasing moisture and alleviating dryness, increasing the therapeutic role of traditional Chinese medicine in treating SS patients. However, the specific therapeutic effect and mechanism of traditional Chinese medicine on SS remain unclear.

Exosomes are bilayer lipid membrane vesicles secreted by cells in the form of budding. Exosomes have specific markers such as CD9, CD63, and CD81 on their surfaces and contain a large number of protein, nucleic acid, and lipid molecules, which extensively participate in intercellular information transmission in a non-cellular contact manner (Gurunathan et al., 2019; Kalluri and Leliu, 2020). Studies have shown that protein information molecules in exosomes regulate T and B lymphocyte activation (Anel et al., 2019) and are associated with the immune pathogenesis of autoimmune diseases (Tan et al., 2016), including rheumatoid arthritis (RA), Sjögren’s syndrome and systemic lupus erythematosus (SLE). Since many TCM compounds play a therapeutic role based on immune regulation (Ma et al., 2013; Wang et al., 2020), and it is reported that Ophiopogon japonicus, one of the main drugs constituting Zengye decoction, can effectively regulate the differentiation of T lymphocyte subsets and inhibit the production of auto-antibodies in SS patients (Wang et al., 2007), we speculate that the therapeutic effect of Zengye Decoction on pSS may be related to the changes of content in blood exosome. This study evaluates the therapeutic impact of MZD by observing the changes in clinical symptoms, laboratory indicators, and cytokines before and after SS treatment and aims to clarify the possible action mechanism of MZD via plasma exosomal proteins using label-free proteomics and bioinformatics.

**2 Methods**

2.1 Study design

The participating pSS patients were treated with MZD for 2 weeks, and the changes in their clinical symptoms, laboratory indicators, and plasma cytokines were compared before and after treatment. Furthermore, the differentially expressed proteins (DEPs) in the plasma exosomes before and after MZD treatment were identified via label-free proteomics, while GO and KEGG enrichment were used to analyze the biological characteristics of the downregulated DEPs.

2.2 Patients

Sixty patients were recruited from the Second Affiliated Hospital of Guizhou University of Traditional Chinese
Medicine to participate in this trial. Twenty-six patients with hypertension, hyperglycemia, hyperlipidemia, gout, rheumatoid arthritis, and hepatitis B viral infection were excluded, while 16 used hydroxychloroquine hydrochloride and other Western drugs during the treatment. The remaining 18 pSS patients met the requirements of this trial and the standard classification criteria of pSS jointly presented by the American College of Rheumatology (ACR) and the European League Against Rheumatism (EULAR) (Shiboski et al., 2017). This study required a patient score exceeding or equal to 4, without the use of Western or traditional Chinese medicine in the previous 2 weeks. All subjects signed an informed consent form after reviewing the details of the trial.

2.3 Modified zengye decoction formula and usage

The granule pharmacy of the Second Affiliated Hospital of Guizhou University of Traditional Chinese Medicine provided 40 g Qinghao, 20 g Shengdihuang, 20 g Xuansheng, 20 g Maidong, 20 g Baihuashecao, 20 g Chuanxinlian, and 10 g Shuqucao (4:2:2:2:2:1) in granulated form. Here, 100 ml of boiling water was added to 7.5 g of each sample and stirred to mix thoroughly. The patients took these mixtures 30 min after meals, twice a day for 2 weeks.

2.4 Plasma preparation

Venous blood treated with EDTA was used to prepare the plasma samples of the participants. These samples were centrifuged at 3,000 g for 30 min (4°C) to remove the cells and cell debris, after which the supernatant was stored at −80°C for downstream analysis.

2.5 Exosome isolation

The plasma exosomes were isolated using ultracentrifugation (UC) with some modifications (Diaz et al., 2018). Briefly, the plasma samples were thawed at 37°C and centrifuged at 500 g for 5 min and 2,000 g for 20 min at 4°C to eliminate the cells and cell debris. The samples were then subjected to UC at 10,000 g at 4°C for 45 min to remove larger vesicles. Next, the supernatant was passed through a 0.45-μm pore filter, followed by UC at 100,000 g at 4°C for 75 min. The supernatant was discarded, after which the pelleted exosomes were resuspended in 200 μl PBS, of which 20 μl was used for electron microscopy and 10 μl for particle size measurement, while the remaining exosomes were stored at −80°C.

2.6 Label-free quantitative proteomic analysis

2.6.1 Preparation of the plasma exosome samples

The plasma exosomes of nine patients were divided into three random groups. Each group was pre-treated with a plasma exosome mixture from three patients, labeled pre-SS1, pre-SS2, and pre-SS3. Similarly, each group received post-treatment containing a mixture of the plasma exosomes from patients treated with MZD, labeled post-SS1, post-SS2, and post-SS3.

2.6.2 Exosomal protein preparation

Briefly, the plasma samples of three pSS patients were centrifuged at 1,000 g for 10 min. The subsequent supernatant was centrifuged at 17,000 g for 15 min at 4°C, followed by UC at 4°C and 200,000 g for 1 h. The supernatant was then discarded. For the 1 × PBS resuspended precipitation, urea particles were added until reaching a concentration of 8 M and shaken until fully dissolved. Ultrasonic crushing on ice, 30% energy, ultrasonic for 1 s, stop for 1 s, cumulative 2 min. The sample was centrifuged at 14,000 g for 20 min, and the supernatant was collected, 10 μl of which was used for protein quantification using a BCA protein assay kit, while the remainder was frozen at −80°C.

2.6.3 Protein digestion and desalination

A 100-μg aliquot of extracted proteins from each sample was subjected to a reduction process. A200 mM dithiothreitol (DTT) solution was added and incubated at 37°C for 1 h. The samples were diluted four times by adding 25 mM ammonium bicarbonate (ABC) buffer. Next, trypsin (trypsin: protein = 1: 50) was added and incubated overnight at 37°C, followed by the addition of 50 μl 0.1% FA to terminate the digestion process. Then, 100 μl of 100% CAN was used to wash the C18 column, followed by centrifugation at 1,200 rpm for 3 min. The column was washed once with 100 μl of 0.1% FA and centrifuged at 1,200 rpm for 3 min. After the EP tube was replaced, the sample was added and centrifuged at 1,200 rpm for 3 min. The column was washed twice with 100 μl of 0.1% FA and centrifuged at 1,200 rpm for 3 min. Then, the column was washed twice with 100 μl of 0.1% FA, followed by centrifugation at 1,200 rpm for 3 min, and then washed with 100 μl water. The EP tube was replaced, followed by elution with 70% ACN. The eluents of each sample were combined, lyophilized, and stored at −80°C until loading.

2.6.4 LC-MS/MS analysis

The mobile phase A (99.9% water and 0.1% formic acid) and B solutions (100% acetonitrile and 0.1% formic acid) were prepared. Lyophilized powder was dissolved in 10 μl of solution A and centrifuged at 14,000 g for 20 min at 4°C, after which 1 μl of the supernatant was injected into a C18 column. The peptides were separated via linear gradient elution and analyzed using a Q Exactive HF-X mass spectrometer.
(Thermo, Waltham, MA, United States) equipped with a Nanospray Flex™ (ESI) ion source at a spray voltage of 2.4 kV and an anion transport capillary temperature of 275°C. Full-scan mass spectrometry was performed in a range of 350 m/z to 1500 m/z at a resolution of 120,000 (at 200 m/z), an automatic gain control (AGC) target value of \(3 \times 10^6\), and a maximum ion injection time of 80 ms. The top 40 precursors of the highest abundance during the full scan were selected and fragmented via higher-energy collisional dissociation (HCD). The samples were analyzed via MS/MS at a resolution of 15,000 (at200 m/z), an AGC target value of \(5 \times 10^6\), a maximum ion injection time of 45 ms, and normalized collision energy of 27% (Cox et al., 2014).

2.6.5 Data analysis
The raw MS data were analyzed using Proteome Discoverer2.4 software and searched against the Homo sapiens database in the Universal Protein Resource Knowledge Base (UniProtKB). The precursor mass window was 15 ppm for the initial search and followed the enzymatic cleavage rule for trypsin. A maximum of two missed cleavage sites and a 20-ppm mass tolerance for the fragment ions were allowed. The cutoff for the global false discovery rate (FDR) in the peptide-spectrum match (PSM) and protein identification was \(p < 0.01\). The complete peptide and protein information from the triplicate analyses of the three samples is available in the ProteomeXchange Consortium via the PRIDE partner repository with the dataset identifier, PXD010556. Only proteins identified by at least two different peptides and in at least two sample replicates were regarded as present, reliably identified, and subjected to further analysis (Yan et al., 2011).

2.6.6 Protein quantification and data processing
The label-free quantification (LFQ) of each identified protein was performed using the peptide signal intensities. The MaxLFQ algorithm in MaxQuant was activated to quantify the protein abundance of the identified peptides. Match-between-runs were performed during the experimental replicates to extract the comprehensive quantification information. Perseus was used to complement a value from the normal distribution with default parameters for low-abundance proteins with missing values. Protein quantification and statistical significance analysis were performed using a two-way Student’s t-test. Proteins that were quantified with a fold change (FC) of \(>1.5\) and a significance value of \(p < 0.05\) after comparing the two groups were designated as DEPs, while \(\log_2 (\text{FC}) > 1.5\) were used for further analysis.

2.7 Enzyme-linked immunosorbent assay
The plasma B cell activating factors (BAFF), IL-4, IL-13, IL-17A, IL-22, and INF-γ concentrations were determined using ELISA kits supplied by eBioscience (San Diego, CA, United States) according to the instructions of the manufacturer. The exosomal protein ceruloplasmin (CP), cartilage oligomeric matrix protein (COMP), insulin-like growth factor-binding protein 2 (IGFBP2), ficolin-2 (FCN2), and lipopolysaccharide-binding protein (LBP) concentrations were determined using ELISA kits supplied by EIAab Science Inc. (Wuhan, China) according to the instructions of the manufacturer.

2.8 Clinical parameter analysis
All clinical parameters were obtained from the clinical laboratory according to standard operating procedures. The tear flow rate was evaluated using Schirmer’s test, with a positive result indicating that the length of the moisture on the filter paper was less than 5 mm per min (\(\leq 5\, \text{mm/5 min}\)). The unstimulated salivary flow was determined according to methods described in previous studies, with a positive result indicating that the amount of salivary per 15 min was less than 1.5 ml (\(\leq 1.5\, \text{ml/15 min}\)).

2.9 Bioinformatic analysis
The information of identified proteins was obtained from the UniProt-GOA database (http://www.ebi.ac.uk/GOA/). Both Gene Ontology (GO) analysis and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analysis were conducted by DAVID online database (https://david.ncifcrf.gov/). Main steps are as followed: Firstly, open website and click "functional annotation", paste target gene in "Enter Gene list" item, and select "Official_Gene_Symbol" in "Select Identifier" item, select "Gene list" in "list type" item, and click "use" in select "Homo sapiens" item and the system will automatically perform enrichment analysis. Secondly, click the "Gene_Ontology" drop-down option, select "GO term_BP_direct," "GO term_CC_direct" and "GO term_MF_direct" respectively, and click "Function Annotation" to get the final GO enrichment result, and to save in "up_GO.txt" file for visual analysis. Thirdly, click the "Pathway" drop-down option, select "KEGG pathway" to get the final KEGG enrichment result, and to save in "KEGG_pathway.txt" file for visual analysis. Finally, the top 20 items are used for go visual analysis and the top 15 items are used for KEGG visual analysis.

2.10 Statistical analysis
The data were shown as the median with the interquartile range or standard deviation. The differences in the clinical parameters and plasma cytokines before and after treatment
TABLE 1 MZD improves the main clinical symptoms of SS patients.

| Clinical symptoms                | Number of cases(n) | Number of improvement cases(n) | Efficiency (%) |
|----------------------------------|--------------------|---------------------------------|----------------|
| Dry mouth and tongue             | 18                 | 18                              | 100.00         |
| Dry eyes                         | 18                 | 18                              | 100.00         |
| Dry skin                         | 16                 | 16                              | 100.00         |
| Dry stool                        | 15                 | 15                              | 100.00         |
| Feverish palms and soles         | 11                 | 9                               | 81.82          |
| Enlargement of parotid gland     | 13                 | 11                              | 84.62          |
| Rash                             | 8                  | 5                               | 62.50          |
| Arthralgia                       | 16                 | 11                              | 68.75          |
| Low-grade fever                  | 8                  | 6                               | 75.00          |

FIGURE 1

The influence of MZD on SS laboratory test subjects. After the SS patients were exposed to continuous MZD intervention for 2 weeks, the differences before and after treatment were compared via a paired t-test. (A) The ESR levels. The (B) IgG, (C) IgM, and (D) IgA concentrations. (E) The salivary flow rate. (F) The tear flow rate.
were evaluated via a paired t-test. The exosomal proteins before and after treatment that needed verification via ELISA were evaluated using an unpaired t-test. A value of \( p < 0.05 \) was considered statistically significant. All statistical analyses were performed using GraphPad Prism V9 software (GraphPad Software, San Diego, CA, United States).

3 Results

3.1 The effect of modified zengye decoction on the clinical symptoms and laboratory examination parameters of Sjögren’s syndrome patients

MZD can effectively improve the clinical symptoms of SS. Effective treatment rates of 99.9% were evident for a dry tongue, dry eyes, dry stool, and dry skin. The effective treatment rates for parotid gland swelling, hand and foot heat, low fever, joint pain, and rash were 84.62%, 81.82%, 75.00%, 68.75%, and 62.50%, respectively (Table 1). In addition, MZD also significantly affected the laboratory examination parameters. Compared with pre-treatment, the erythrocyte sedimentation rate (ESR) of the patients after treatment was reduced considerably, \( p = 0.0022 \) (Figure 1A), while the plasma immune IgG and IgM content decreased substantially, with \( p \)-values of 0.0029 (Figure 1B) and 0.0253 (Figure 1C), respectively. However, no significant changes were evident in IgA (Figure 1D). A considerable increase was apparent in both the salivary flow rate, \( p = 0.01 \) (Figure 1E), and tear flow rate, \( p = 0.0008 \) (Figure 1F).

3.2 The effect of modified zengye decoction on the plasma cytokines in Sjögren’s syndrome

The imbalance between T helper cells (Th cells) subset differentiation and its related cytokines plays an important role in the pathological process of SS. To clarify the effect of MZD on the cell subset differentiation and cytokines, the changes in the plasma Treg cytokines (TGF-\( \beta \)), Th2 cytokines (IL-4 and IL-13), Th17 cytokines (IL-17A and IL-22), and BAFF were examined before and after MZD treatment. The results showed that the BAFF (Figure 2A), IL-4 (Figure 2B), IL-13 (Figure 2C), IL-17A (Figure 2D), and IL-22 (Figure 2E) plasma content decreased significantly after MZD treatment, with \( p \)-values of 0.0128, 0.0016, 0.0104, 0.0012, and 0.0191, respectively. The TGF-\( \beta \) content displayed no significant changes after treatment (Figure 2F). This suggests that although MZD can substantially downregulate the Th2 and Th17 cytokine levels, it does not significantly affect the TGF-\( \beta \) content.

3.3 The differentially expressed protein levels before and after modified zengye decoction treatment of the Sjögren’s syndrome patients

The plasma exosomes were isolated using UC, while the exosomal DEPs before and after MZD treatment were identified via label-free proteomics. Transmission electron microscopy (TEM) showed typical cup-shaped vesicles (Figure 3A), while Nanoparticle tracking analysis (NTA) indicated that these vesicles were approximately 100 nm in diameter (Figure 3B). A quantitative ratio exceeding 1.5 was considered upregulation, while a ratio below 0.667 was considered downregulation. A comparison between the results before and after MZD treatment indicated that 13 DEPs were identified in the plasma exosomes via label-free proteomic analysis, one of which was an upregulated protein, while twelve were downregulated proteins, as shown in the DEP volcano map (Figure 3C) and downregulated protein clustering heat map (Figure 3D). The detailed data of the upregulated and downregulated proteins are shown in Table2.

3.4 Analysis of the biological information of the exosomal downregulated differentially expressed proteins

To confirm that the therapeutic effect of MZD on SS is related to plasma exosomal protein reduction, this study used bioinformatics to analyze the biological functions of the downregulated proteins after MZD treatment. As shown in Figure 4A and Supplementary Table S1, the results of the GO analysis (BP) indicated that the downregulated proteins were significantly involved in the response to insulin (\( p = 0.0011 \)) and the defense response to Gram-negative bacteria (\( p = 0.0217 \)). The insulin response may be related to the dryness symptoms caused by SS, while the defense response to Gram-negative bacteria may be associated with SS infection. As shown in Figure 4B and Supplementary Table S2, the GO analysis (MF) results indicated that the downregulated proteins were significantly enriched in proteoglycan binding (\( p = 0.0079 \)), which may also be associated with SS infection. As shown in Figure 4C and Supplementary Table S3, the KEGG pathway analysis showed that the downregulated proteins were completely enriched in porphyrin metabolism (\( p = 0.0280 \)), folate biosynthesis (\( p = 0.0280 \)), antifolate resistance (\( p = 0.0280 \)), the NF-\( \kappa \)B signaling pathway (\( p = 0.0454 \)), the toll-like receptor signaling pathway (\( p = 0.0454 \)), and ferroptosis (\( p = 0.0454 \)). Porphyrin metabolism and folate biosynthesis may be related to the bone and joint injury caused by SS, while the NF-\( \kappa \)B and Toll-like receptor signaling pathways may be associated with SS infection. These results suggest that the exosomal proteins downregulated by MZD are closely related to the SS pathological mechanism.
3.5 The validation of the downregulated exosomal differentially expressed proteins (related to modified zengye decoction efficacy)

After examining the GO and KEGG analyses results, this study speculates that the exosomal proteins downregulated by MZD may include COMP, CP, FCN2, GGH, IGFBP2, and LBP. Detailed information about the BPs, MFs, and signaling pathways involved in these proteins are provided in Supplementary Tables S1–S3. The ELISA results of these exosomal proteins were consistent with the label-free proteomics findings. After treatment, the COMP, CP, FCN2, GGH, IGFBP2, and LBP expression levels were significantly decreased. The detailed detection results are shown in Figures 6A–F, respectively.

4 Discussion

Although genetic, environmental, and viral factors play an important role in the onset of SS, the exact mechanism driving this disease remains unclear (Roszkowska et al., 2021). From a clinical and pathological perspective, the primary hallmark of this disease is exocrine dysfunction, including salivary and lacrimal gland malfunction (Negrini et al., 2022). Therefore, the primary clinical manifestations of SS are xerostomia and xerophthalmia, severely impairing the quality of life of patients (Vehof et al., 2020). Treatment for the disease remains largely palliative, focusing on stimulating the remaining secretion of exocrine glands or replacing exocrine fluid of saliva and tears (Chowdhury et al., 2021). However, the existing treatments are primarily unsuccessful in improving symptoms and disease progression (Fox et al., 2021). Therefore, it is...
necessary to find new methods to improve the therapeutic effect of SS.

Since traditional Chinese medicine compounds can effectively alleviate SS symptoms (Chen et al., 2021; Hu et al., 2014; Shao et al., 2021), many patients in China opt for this treatment (Hou and Xue, 2020). This study showed that MZD could effectively improve the clinical symptoms of SS patients receiving continuous treatment for 2 weeks, including a dry tongue, dry eyes, dry skin, and dry stool. It also exhibited an excellent therapeutic effect on joint pain, parotid gland swelling, and rash. Similarly, the laboratory test results supported the therapeutic effect of traditional Chinese medicine on SS, which was mainly reflected in the significant increase in the salivary flow rate, the substantially lower ESR, and the considerable decrease in the content of the plasma immune indexes, IgG and IgM, after MZD treatment. These results suggest that MZD or other traditional Chinese medicine may provide a new strategy for SS treatment.

The main pathological feature of SS is the infiltration of the exocrine glands by a large number of lymphocytes. Some studies have shown that the abnormal differentiation of Th2 cells and Th17 cells and their cytokines play an important role in the occurrence of SS (Qi et al., 2018; Huang et al., 2020). The plasma content of Th2 cytokines, IL-4 and IL-13, and Th17 cytokines, IL-17A and IL-22, decreased significantly after MZD treatment, suggesting
that MZD inhibited Th2 and Th17 activation and the mediated inflammatory response. These results were consistent with previously published studies (Wang et al., 2007; Lee et al., 2021), which showed that MZD could effectively inhibit the activation of Th2 and Th17 cells and the production of corresponding cytokines. Furthermore, traditional Chinese medicine also inhibited BAFF production. Since BAFF activates B cells, its inhibition may be essential for the downregulation of the IgG and IgM antibody levels in traditional Chinese medicine. In addition, MZD also affected the TGF-β. Tregs are inhibitory T cells that produce TGF-β that can inhibit the inflammatory effect of Th17 cells. The results showed that Chinese herbal medicine had no significant impact on the TGF-β plasma content. Although the TGF-β level decreased after treatment, no significant differences were evident before and after treatment. This study indicated that the therapeutic effect of traditional Chinese medicine on SS may be related to

### Table 2: The plasma exosomal DEPs before and after MZD treatment.

| Accession | Gene symbol | Protein name                                      | FC    | P         |
|-----------|-------------|--------------------------------------------------|-------|----------|
| P18065    | IGFBP2      | Insulin-like growth factor-binding protein 2     | 0.4061| 0.0122   |
| Q6033     | ITIH3       | Inter-alpha-trypsin inhibitor heavy chain H3     | 0.6543| 0.0043   |
| O43147    | SGSM2       | Small G protein signaling modulator 2            | 0.7490| 0.0444   |
| P18450    | CP          | Ceruloplasmin                                    | 0.7950| 0.0122   |
| Q9UGM5    | FETUB       | Fetuin-B                                         | 0.7235| 0.0167   |
| A0A0B4J1V2| IGHV2-26    | Immunoglobulin heavy variable 2-26              | 0.7036| 0.0485   |
| P24593    | IGFBP5      | Insulin-like growth factor-binding protein 5     | 2.7084| 0.0067   |
| A0A075B6K5| IGLV3-9     | Immunoglobulin lambda variable 3-9              | 0.5856| 0.0121   |
| P26927    | MST1        | Hepatocyte growth factor-like protein            | 0.8119| 0.0350   |
| Q15485    | FCN2        | Ficolin-2                                        | 0.5233| 0.0394   |
| P18428    | LBP         | Lipopolysaccharide-binding protein                | 0.6697| 0.0180   |
| P49747    | COMP        | Cartilage oligomeric matrix protein              | 0.4523| 0.0388   |
| Q92820    | GGH         | Gamma-glutamyl hydrolase                         | 0.6058| 0.0349   |

**Figure 4**

The GO analysis of the downregulated DEPs in the plasma exosomes. (A) The MF enrichment analysis. (B) The BP enrichment analysis. The circle size represents the number of genes enriched in the GO entries. A larger circle indicates more gene enrichment in this GO entry. The color of the circle represents the significance of the enrichment. The redder the circle, the more significant the DEP enrichment on this GO entry.
the inhibition of Th2 and Th17 activation and the corresponding cytokine reduction without damaging the inhibitory function of Treg cells.

Exosomes carry several proteins, nucleic acids, and lipids that extensively participate in transmitting information between cells. Various studies have shown that exosomes play a crucial role in the occurrence of autoimmune diseases, tumors, and cardiovascular diseases (Tan et al., 2016). Exosomes can reportedly regulate the differentiation of Th2 cells and Th17 cells. Therefore, it is speculated that the therapeutic effect of MZD on SS may be related to decreased plasma exosome protein. Label-free proteomic analysis showed the presence of 13 DEPs in the plasma exosomes before and after MZD intervention, of which 12 were downregulated. The downregulated proteins were involved in several BFs, including the insulin response, the Gram-negative bacterial defense response, and proteoglycan binding. The insulin response is closely related to a dry mouth in diabetic patients (Sundaram et al., 2020). This study also speculates that the dry mouth symptoms in SS patients are also related to the insulin response. The alieving effect of traditional Chinese medicine on dryness may also be related to the downregulation of the insulin response. The defense response to Gram-negative bacteria and proteoglycan binding plays a vital role in the pathological processes of infection, consistent with the cause of SS infection. This suggests that the therapeutic effect of Chinese medicine on SS may be related to the inhibition of SS infection. The KEGG signaling pathway enrichment also showed similar results. The downregulated exosome proteins were mainly involved in the NF-κB and TLR4 pathways and porphyrin metabolism. The NF-κB and TLR4 signaling pathways are crucial in activating inflammation, further suggesting that the therapeutic effect of traditional Chinese medicine is related to the inhibition of SS infection. Porphyrin metabolism and folic acid synthesis are involved in almost all autoimmune diseases, and the influence of traditional Chinese medicine on the metabolic pathway may also highlight its efficacy. To ensure the reliability of the proteomic results, this study verified the downregulated exosomal proteins, namely COMP, CP, FCN2, GGH, IGFBP2, and LBP, involved in the BPs. The changes in these proteins before and after treatment were consistent with the proteomic results. This study suggests that the therapeutic MZD mechanism in SS may be related to the reduced expression of the exosomal proteins involved in the insulin response, porphyrin metabolism, and the TLR4/NF-κB pathway.

Therefore, this research confirms that MZD can effectively relieve the clinical symptoms of SS and verifies that its action mechanism may be related to exosomal protein reduction.
Furthermore, the MZD efficacy verification results provided evidence for the role of Th cell subset differentiation, corresponding cytokines, and exosomal proteins in SS. However, this research presents some limitations. First, this study does not sufficiently highlight the multi-target therapeutic effect of the Chinese herbal formula and lacks a single chemical component analysis of MZD efficacy. Second, the therapeutic mechanism of MZD in SS is only elucidated from the perspective of exosomal proteins, while other exosomal content that may be involved, such as microRNA and IncRNA, is not analyzed. Third, the sample size is small. Currently, SS treatment mainly combines Western and traditional Chinese medicine. Few patients are willing to accept traditional Chinese medicine alone, preventing the use of large samples for clinical intervention studies.

Data availability statement

The data presented in the study are deposited in the ProteomeXchange Consortium via the PRIDE partner repository with the dataset identifier PXD033627.

Ethics statement

The studies involving human participants were reviewed and approved by The Second Affiliated Hospital, Guizhou University of Traditional Chinese Medicine. The patients/participants provided their written informed consent to participate in this study.

Author contributions

YM and XP planned and performed the experiments; YW and LH analyzed the data; WM and PY initiated project and designed the study. PY, YM, and XP drafted and wrote the manuscript.

Funding

We gratefully acknowledge the support by Science and Technology Foundation of Guizhou (5766-07), National Natural Science Foundation of China Sponsored Project (81960819), National Natural Science Foundation of China Sponsored Project (81973835).
Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher’s note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fphar.2022.930638/full#supplementary-material

References

Anel, A., Gallego-Lleyda, A., de Miguel, D., Naval, J., and Martínez-Lostao, L. (2019). Role of exosomes in the regulation of T-cell mediated immune responses and in autoimmune disease. Cells 8 (2), 154. doi:10.3390/cells8020154

Arvaniti, P., Zachou, K., Lyberopoulos, A., Gatseli, N. K., Brooks, W. H., Dalekos, G. N., et al. (2019). Epigenetic modifications in generalized autoimmune epithelitis: Sjögren’s syndrome and primary biliary cholangitis. Epigenomes 3 (3), 15. doi:10.3390/epigenomes3030015

Chen, H. H., Lu, J. N., Yu, M. C., Chen, C. Y., Hsieh, Y. T., Hsu, Y. F., et al. (2021). Traditional Chinese medicine in patients with primary Sjögren’s syndrome: a randomized, double-blind, placebo-controlled clinical trial. Front. Med. 8, 744194. doi:10.3389/fmed.2021.744194

Chowdhury, F., Tappuni, A., and Bombardieri, M. (2021). Biological therapy in primary Sjögren’s syndrome: Effect on salivary gland function and inflammation. Front. Med. 8, 707104. doi:10.3389/fmed.2021.707104

Cox, J., Hein, M. Y., Luber, C. A., Paron, I., Nagar, N., and Mann, M. (2014). Accurate proteome-wide label-free quantification by delayed normalization and maximal peptide ratio extraction, termed MaxLFQ. Mol. Cell. Proteomics 13 (9), 2513–2526. doi:10.1074/mcp.M113.031591

Diaz, G., Bridges, C., Lucas, M., Cheng, Y., Schorey, J. S., Dobos, K. M., et al. (2018). Protein digestion, ultrafiltration, and size exclusion chromatography to optimize the isolation of exosomes from human blood plasma and serum. J. Vis. Exp. 134, 57467. doi:10.3791/57467

Fox, R. I., Fen, C. M., Gettner, J. E., and Dörner, T. (2021). Treatment of Sjögren’s syndrome: current therapy and future directions. Rheumatol. Oxf 50 (5), 2066–2074. doi:10.1093/rheumatology/kez142

Gurunathan, S., Kang, M. H., Jeyaraj, M., Qasim, M., and Kim, J. H. (2019). Correction: Gurunathan, S. et al. Review of the isolation, characterization, biological function, and multifaceted therapeutic approaches of exosomes. Cells 810 (2), 307462. doi:10.3390/cells10020146

Hou, J. Q., and Xue, L. (2020). Challenges in treatment of primary Sjögren’s syndrome and opportunities for Chinese medicine. Chin. J. Integr. Med. 26 (7), 483–485. doi:10.1007/s11684-016-0434-2

Hu, W., Qian, X., Guo, F., Zhang, M., Lyu, C., Tao, J., et al. (2014). Traditional Chinese medicine compound ShengJinRanZaoYangXue granules for treatment of primary Sjögren’s syndrome: a randomized, double-blind, placebo-controlled clinical trial. Chin. Med. J. 127 (15), 2721–2726.

Huang, Y., Yang, G., Fei, J., Wu, Y., and Yan, J. (2020). Bufotalin ameliorates experimental Sjögren’s syndrome development by inhibiting Th17 generation. Naunyn Schmiedeber. Arch. Pharmacol. 393 (10), 1977–1985. doi:10.1007/s00210-020-01817-1

Jiang, Y., Zhao, X., Yu, J., Wang, Q., Wen, C., and Huang, L. (2021). Deciphering potential pharmacological mechanism of Sha-Shun-Mai-Dong decoction on primary Sjögren’s syndrome. BMC Complement. Med. Ther. 21 (1), 79. doi:10.1186/s12906-021-03257-7

Kalluri, R., and LeBoeuf, V. S. (2020). The biology, function, and biomedical applications of exosomes. Science 367 (6478), eaau977. doi:10.1126/science.aau977

Lee, G. A., Chang, C. M., Wu, Y. C., Ma, R. Y., Chen, C. Y., Hsu, Y. T., et al. (2021). Chinese herbal medicine SS-1 inhibits T cell activation and abrogates TH responses in Sjögren’s syndrome. J. Formos. Med. Assoc. 120, 651–659. doi:10.1016/j.jfma.2020.07.024

Lu, F., Wang, D., Li, R. L., He, L. Y., Ai, L., and Wu, C. J. (2021). Current strategies and technologies for finding drug targets of active components from traditional Chinese medicine. Front. Biosci. 26 (9), 572–589. doi:10.52566/4968

Ma, H. D., Deng, Y. R., Tian, Z., and Lian, Z. X. (2013). Traditional Chinese medicine and immune regulation. Clin. Rev. Allergy Immunol. 44 (3), 229–241. doi:10.1007/s00210-012-8332-0

Ma, Y., Zhou, K., Fan, J., and Sun, S. (2016). Traditional Chinese medicine: Potential approaches from modern dynamical complexity theories. Front. Med. 10 (1), 28–32. doi:10.1007/s11684-016-0434-2

Matsilinka, M. (2019). The role of Epstein-Barr virus infection in primary Sjögren’s syndrome. Curr. Opin. Rheumatol. 31 (5), 475–483. doi:10.1097/BOR.0000000000000622

Negrini, S., Emmi, G., Greco, M., Borro, M., Sardanelli, F., Murdaca, G., et al. (2022). Sjögren’s syndrome: a systemic autoimmune disease. Exp. Clin. Med. 22 (1), 9–25. doi:10.14312/acrmed.2021-0010616

Qi, J., Li, D., Shi, G., Zhang, X., Pan, Y., Dou, H., et al. (2018). Myeloid-derived suppressor cells exacerbate Sjögren’s syndrome by inhibiting Th2 immune responses. Mol. Immunol. 101, 251–258. doi:10.1016/j.molimm.2018.07.016

Rivière, E., Pascaud, J., Tchitchek, N., Boudaoud, S., Pasletti, A., Ly, B., et al. (2020). Salivary gland epithelial cells from patients with Sjögren’s syndrome induce B lymphocyte survival and activation. Ann. Rheum. Dis. 79 (11), 1468–1477. doi:10.1136/annrheumdis-2019-216588

Roszkowska, A. M., Olivo, G. W., Aragona, E., Inferrera, L., Severo, A. A., Alessandrello, F., et al. (2021). Ophthalmologic manifestations of primary Sjögren’s syndrome. Genes 12 (3), 365. doi:10.3390/genes12030365

Shao, Q., Jin, L., Li, C., Wang, J., Wang, M., Wang, Q., et al. (2021). Comparative analysis of the efficacy and safety of herbal decoction CheReCuJin alone and combined with hydroxychloroquine for treating primary Sjögren’s syndrome: A randomized controlled trial. Explore (N Y) 15(5), 8307, 416–422. doi:10.1097/EPL.0000000000000248

Shiboski, C. H., Shiboski, S. C., Seror, R., Criswell, L. A., Labelouelle, M., Liemant, T. M., et al. (2017). 2016 American College of Rheumatology/European League against rheumatism classification criteria for primary Sjögren’s syndrome: a consensus and data-driven methodology involving three international patient cohorts. Arthritis Rheumatol. 69 (1), 35–45. doi:10.1002/art.39859

Sundaram, M., Manikandan, S., Sathesh, B., Srinivasan, D., Jayapal, D., and Kumar, D. (2020). Comparative evaluation of ketorolac among diabetic and nondiabetic subjects wearing complete denture. J. Pharm. Bioallied Sci. 12, S422–S422. doi:10.4103/jpbs.JPBS_124_20

Tan, L., Wu, H., Liu, Y., Zhao, H., Lu, C., Lu, A., et al. (2020). Potential advantages of bioactive compounds extracted from traditional Chinese medicine to inhibit bone destructions in rheumatoid arthritis. Front. Pharmacol. 11, 561962. doi:10.3389/fphar.2020.561962

Vehof, J., Uteijn, T. P., Bootma, H., and Hammond, C. J. (2020). Advances, limitations and future perspectives in the diagnosis and management of dry eye in Sjögren’s syndrome. Clin. Exp. Rheumatol. 38, 301.
Wang, Y., Yan, T., Shen, J., Guo, H., and Xiang, X. (2007). Preventive effect of Ophiopogon japonicus polysaccharides on an autoallergic mouse model for Sjögren’s syndrome by regulating the Th1/Th2 cytokine imbalance. *J. Ethnopharmacol.* 114 (2), 246–253. doi:10.1016/j.jep.2007.08.014

Wang, Y., Zhang, Q., Chen, Y., Liang, C. L., Liu, H., Qiu, F., et al. (2020). Antitumor effects of immunity-enhancing traditional Chinese medicine. *Biomed. Pharmacother.* 121, 109570. doi:10.1016/j.biopha.2019.109570

Wei, S. J., He, Q. M., Zhang, Q., Fu, K. H., Li, R. L., Peng, W., et al. (2021). Traditional Chinese medicine is a useful and promising alternative strategy for treatment of Sjögren’s syndrome: A review. *J. Integr. Med.* 19 (3), 191–202. doi:10.1016/j.jim.2021.01.008

Wu, X., Ren, C., Zhou, H., Zhang, L., Juan, C., and Yang, Y. (2014). Therapeutic effect of Zeng Ye decoction on primary Sjögren’s syndrome via upregulation of aquaporin-1 and aquaporin-5 expression levels. *Mol. Med. Rep.* 10 (1), 429–434. doi:10.3892/mmr.2014.2208

Yan, W., Luo, J., Robinson, M., Eng, J., Aebersold, R., and Ranish, J. (2011). Indexion triggered MS2 ion quantification: a novel proteomics approach for reproducible detection and quantification of targeted proteins in complex mixtures. *Mol. Cell. Proteomics* 10 (3), M110.005611. doi:10.1074/mcp.M110.005611

Yuan, H., Ma, Q., Ye, L., and Piao, G. (2016). The traditional medicine and modern medicine from natural products. *Molecules* 21 (5), 559. doi:10.3390/molecules21050559