Green Synthesis of Silver Nanoparticles with Antibacterial Activities Using Aqueous *Eriobotrya Japonica* Seed Extract

Bo RAO\(^a\) and Ren-Cheng TANG\(^b\)\(^*\)

National Engineering Laboratory for Modern Silk, College of Textile and Clothing Engineering, Soochow University, 199 Renai Road, Suzhou 215123, China
\(^a\) 535184812@qq.com.cn, \(^b\) tangrencheng@suda.edu.cn

**Keywords:** Antibacterial activity, *Eriobotrya japonica*, Green synthesis, Silver nanoparticles.

**Abstract.** An eco-friendly approach for the preparation of silver nanoparticles (AgNPs) from silver nitrate solution using aqueous *Eriobotrya japonica* seed extract was investigated. The reduction of silver ions in solution was monitored using UV-vis spectroscopy, and the surface plasmon resonance of AgNPs at 428 nm was observed. The biosynthesized nanoparticles were characterized using transmission electron microscopy (TEM), scanning electron microscopy, energy dispersive X-ray spectroscopy, dynamic light scattering and X-ray diffraction. The prepared AgNPs were polydispersed and spherical in shape, and their average particle size determined by TEM was 25 nm. Furthermore the biosynthesized AgNPs were found to exhibit effective antibacterial activities against *Escherichia coli* and *Staphylococcus aureus*.

**Introduction**

In the past ten years, silver nanoparticles (AgNPs) have attracted a great deal of attention due to their unique physical, chemical, optical and biological properties, and found tremendous applications in various industries such as biomedicine, drug delivery, electronics, optics, catalysis, food industry, textile industry, etc. \[1\]. A variety of chemical and physical approaches have been developed to prepare AgNPs. Among them, the chemical reduction is most widely used. However, these approaches are inevitably associated with the use of hazardous chemicals such as reductants, stabilizers and organic solvents, or have special requirements for the employed techniques such as high energy radiation, microwave irradiation, etc. \[2, 3\]. In recent years, the biological approach using plant extracts has become a valuable alternative to physical and chemical synthesis. Many researches have been reported on the biosynthesis of AgNPs plant extracts \[4\].

*Eriobotrya japonica* (loquat) is a large evergreen shrub or small tree and cultivated worldwide in all warm and subtropical regions for its fruit or as an ornamental plant. But up to now the seeds of *E. japonica* have usually been disposed of as waste after consuming its fruits. In terms of the comprehensive utilization of *E. japonica* seeds as well as the recent developments of plant-mediated AgNPs synthesis, it is significant to carry out the research on the biosynthesis of AgNPs using the extracts of *E. japonica* seeds. The aqueous *E. japonica* seed extract is rich in nutritional and bioactive components such as amygdalins, polyphenols, proteins, etc. \[5\], which could be used as reductants for the synthesis of AgNPs from silver nitrate solution and also stabilizers of generated AgNPs. In this study, a green approach for the synthesis of AgNPs using aqueous *E. japonica* seed extract was described. And the AgNPs obtained were afterwards subjected to a sequence of characterizations. In addition, the antibacterial activities of AgNPs against *Escherichia coli* and *Staphylococcus aureus* were examined.

**Experimental**

**Materials.** Silver nitrate was purchased from Sinopharm Chemical Reagent Co. Ltd., China. *Eriobotrya japonica* seed was collected from the orchard in the suburb of Suzhou, China.
Preparation of *E. japonia* Seed Extract. To prepare the aqueous *E. japonia* seed extract, the mixture containing 5 g of the seed powder and 100 mL of deionized water in the 250 mL Erlenmeyer flask was heated in the oscillator at 90 °C for 60 min. Subsequently, the extract was centrifuged at 4000 rpm for 20 min to remove impurities. The supernatant was collected, diluted by deionized water to 100 mL, and finally stored at -4 °C for further experiments.

**Biosynthesis of AgNPs.** For the typical reduction of silver ions to AgNPs, 5 mL of the freshly prepared extract was added to 50 mL of the aqueous silver nitrate solution (1 mM), and then the pH value of the mixture was immediately adjusted to 8 using 0.1 M sodium hydroxide before incubation at 60 °C. As the reaction proceeded, the color change of the solution from colorless to yellowish brown was observed and recognized for the formation of AgNPs.

**Measurements.** The UV-vis spectra of the mixture before and after reaction were measured by UV-1800 UV-vis spectrophotometer from 200 to 800 nm. The particle size distribution of the prepared AgNPs colloid was determined by Malvern Zetasizer Nano ZS 90 at a scattering angle of 90°. The detailed size and morphology of biosynthesized AgNPs was investigated by using S-4800 scanning electron microscope with energy dispersive spectrometer and HT700 transmission electron microscope. The X-ray diffraction measurement of AgNPs was performed on the X’Pert-Pro MPD X-ray diffractometer equipped using the Cu-Kα radiation of wavelength 0.15418 nm. The antibacterial activity of biosynthesized AgNPs against *E. coli* and *S. aureus* was determined using the well diffusion method.

**Results and Discussion**

**UV-vis Spectral Studies.** Fig. 1a displays the color change of the mixture before and after reaction. The formation of AgNPs in the solution was evidenced by the color change from colorless to yellowish brown owing to the excitation of the surface plasmon resonance of AgNPs [6]. Fig. 1b shows the UV-vis absorption spectra of the mixture of *E. japonia* seed extract and silver nitrate as a function of time. The bioreduction of silver ions gave rise to the appearance of the characteristic absorption peak at around 428 nm. The peak intensity increased with an increase in reaction time, indicating the increasing number of AgNPs formed in the mixture. The UV-vis spectrum recorded after 90 min showed no obvious increase, suggesting the reaction tended towards equilibrium.

**Characterizations of Biosynthesized AgNPs.** The SEM, EDX, TEM and DLS techniques were applied to determine the size, shape and morphologies of AgNPs in order to give further insights into the detailed features of AgNPs. The SEM image obtained at a high magnification is shown in Fig. 2a, and the related EDX spectrum is given in Fig. 2b. The AgNPs obtained were polydispersed and spherical in shape with a size distribution in the range from 14 to 76 nm. A small number of large
particles viewed probably resulted from the agglomeration of small ones. The EDX profile showed strong signal peak at 3 keV, which is typical of the absorption of silver nanocrystallites [7], demonstrating the presence of AgNPs. Interestingly, some adjoint elements such as C, O and Cl were also detected. This is very likely associated with the organic moieties from the extract adsorbed on the surface of AgNPs, which play a crucial role in the reduction and stabilization of AgNPs.

![Fig. 2. SEM image of biosynthesized AgNPs using *E. japonia* seed extract (a) and EDX spectrum of biosynthesized AgNPs (b).](image)

The morphology and size dimension of the biosynthesized AgNPs can be intuitively obtained from the TEM image (Fig. 3a) and the size distribution in the colloid can be given by the DLS analysis (Fig. 3c). The AgNPs were spherical in shape with different size distribution. It should be noted that the average particle size (around 25 nm) shown by Fig. 3b obtained from TEM analysis was significantly smaller than that (about 100 nm) measured by the DLS method. This discrepancy could be possibly due to the adsorption of organic stabilizers from the extract on the surface of AgNPs, the aggregation of some small particles and the adsorption of water on the stabilized AgNPs [8, 9], all of which could have a negative effect on the average particle size obtained by the DLS method.

![Fig. 3. TEM image of biosynthesized AgNPs using *E. japonica* seed extract (a), histogram of the AgNPs size distribution (b) and DLS histogram of biosynthesized AgNPs in colloidal solution (c).](image)

Fig. 4 illustrates the XRD pattern of the biosynthesized AgNPs using *E. japonica* seed extract, which can confirm the crystalline nature of the prepared AgNPs. The diffraction peaks at the 2θ values of 32.22°, 38.17°, 46.26°, 54.91°, 57.79° and 77.14° corresponded to the 122, 123, 231, 142, 241 and 311 crystallographic planes, respectively. This suggests that the prepared AgNPs are biphasic in nature. And the similar crystalline structure obtained by XRD was also found in the biosynthesized AgNPs using *Ocimum canum* leaf extract [10].

**Antibacterial Activity.** As shown by Fig. 5, the zone of inhibition was clearly observed in the vicinity of wells filled with silver nitrate (9.3 mm for *E. coli*, and 7.5 mm for *S. aureus*), and AgNPs (8.5 mm for *E. coli*, and 7.3 mm for *S. aureus*). The *E. japonica* seed extract did not display the obvious zone of inhibition. The tests prove that both silver nitrate and biosynthesized AgNPs have good antibacterial activities against *E. coli* and *S. aureus.*
Fig. 4. XRD pattern of biosynthesized AgNPs using *E. japonica* seed extract.

Fig. 5. Antibacterial activities of biosynthesized AgNPs using *E. japonica* seed extract against *E. coli* (a) and *S. aureus* (b).

**Conclusions**

An eco-friendly and cost-effective protocol for the synthesis of AgNPs by utilizing a renewable natural resource *E. japonica* seed was proposed. The biosynthesized AgNPs were characterized by UV-vis, SEM, EDX, TEM, DLS and XRD. The AgNPs were polydispersed in solution and spherical in shape, and their average particle sizes determined by TEM and DLS were about 25 and 100 nm, respectively. The biogenic AgNPs exhibited good antibacterial activities against *E. coli* and *S. aureus*. Further research on the AgNPs biosynthesized using *E. japonica* seed extract could bring a promising application in the fields of medicine and hygiene.

**Acknowledgement**

This study was funded by Jiangsu Provincial Natural Science Foundation of China (BK20131178), and Jiangsu Provincial Key Research and Development Program of China (BE2015066).

**References**

[1] V.K. Sharma, R.A. Yngard, Y. Lin, Adv. Colloid Interfac. Sci. 145 (2008) 83-86.
[2] H.S. Shin, H.J. Yang, S.B. Kim, M.S. Lee, J. Colloid Interf. Sci. 274 (2004) 89-94.
[3] M. Noroozi, A. Zakaria, M.M. Moksin, Z.A. Wahab, A. Abedini, Int. J. Mol. Sci. 13 (2012) 8086-8096.
[4] M.S. Akhtar, J. Panwar, Y.-S. Yun, ACS Sustain. Chem. Eng. 1 (2013) 591-602.
[5] E.-N. Li, J.-G. Luo, L.-Y. Kong, Phytochem. Anal. 20 (2009) 338-343.
[6] A. Henglein, J. Phys. Chem. 97 (1993) 5457-5471.
[7] Y. Liu, M. Hussain, H. Memon, S. Yasin, Dig. J. Nanomater. Bios. 10 (2015) 1019-1024.
[8] A. Bootz, V. Vogel, D. Schubert, J. Kreuter, Eur. J. Pharm. Biopharm. 57 (2004) 369-375.
[9] S. Das, P. Roy, S. Mondal, T. Bera, A. Mukherjee, Colloid. Surface. B 107 (2013) 27-34.
[10] C. Jayaseelan, A.A. Rahuman, Parasitol Res. 111 (2012) 1369-1378.