Differential contribution of Bud6p and Kar9p to microtubule capture and spindle orientation in S. cerevisiae

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In Saccharomyces cerevisiae, spindle orientation is controlled by a temporal and spatial program of microtubule (MT)–cortex interactions. This program requires Bud6p/Aip3p to direct the old pole to the bud and confine the new pole to the mother cell. Bud6p function has been linked to Kar9p, a protein guiding MTs along actin cables. Here, we show that Kar9p does not mediate Bud6p functions in spindle orientation. Based on live microscopy analysis, kar9Δ cells maintained Bud6p-dependent MT capture. Conversely, bud6Δ cells supported Kar9p-associated MT delivery to the bud. Moreover, additive phenotypes in bud6Δ kar9Δ or bud6Δ dyn1Δ mutants underscored the separate contributions of Bud6p, Kar9p, and dynein to spindle positioning. Finally, tub2C354S, a mutation decreasing MT dynamics, suppressed a kar9Δ mutation in a BUD6-dependent manner. Thus, Kar9p-independent capture at Bud6p sites can affect spindle orientation provided MT turnover is reduced. Together, these results demonstrate Bud6p function in MT capture at the cell cortex, independent of Kar9p-mediated MT delivery along actin cables.

Introduction

The fidelity of cell division rests on a precise spatial and temporal relationship between chromosomal segregation and the division plane. The axis for chromosomal segregation is defined by the position of the spindle poles, which generate both the mitotic spindle and astral microtubules (MTs) that interact with the cell cortex. Cell division occurs perpendicular to this axis, once chromosomal segregation is complete (Rappaport, 1996; Glotzer, 2001).

The budding yeast Saccharomyces cerevisiae is a powerful model for understanding the multiple layers of control operating to orient the mitotic spindle in a cell dividing asymmetrically. Yeast cells position the preanaphase spindle along the mother-bud axis near the bud neck. As the spindle elongates during anaphase, one pole translocates into the bud and chromosomal segregation between the mother and daughter cell is accomplished (Segal and Bloom, 2001).

Yeast cells control spindle position through the functional counterpart of the centrosome, the spindle pole body (SPB), which interacts with the cell cortex via astral MTs (Carminati and Stearns, 1997; Shaw et al., 1997; Segal et al., 2000a). These interactions are confined to specific areas of the cell cortex according to a program that appears to follow the cortical distribution of the actin-interactor Bud6p/Aip3p throughout the cell cycle (Amberg et al., 1997; Segal et al., 2002).

Current models assign a key role to Kar9p in enforcing daughter-bound fate of a single SPB after spindle assembly (Kusch et al., 2003). Kar9p interacts with the MT-binding protein Bim1p, the yeast EB1 homologue (Tirnauer et al., 1999; Korinek et al., 2000; Lee et al., 2000; Miller et al., 2000), and is preferentially recruited at the daughter-bound SPB (SPBd) to travel toward the MT plus end (Liakopoulos et al., 2003). In turn, Kar9p-bound MTs interact with the cargo domain of Myo2p to be guided along actin cables to the bud (Beach et al., 2000; Yin et al., 2000; Hwang et al., 2003). This model explains the dependency of spindle orientation on the actin cytoskeleton (Palmer et al., 1992; Theesfeld et al., 1999). However, it does not address the issue of how SPB asymmetry is established or the nature of the relationship between Kar9p and Bud6p in MT capture. A more integral model must also incorporate the underlying asymmetric behavior between the old and new SPB with regard to astral MT organization (Shaw et...
This intrinsic asymmetry requires Cdk activity (Segal et al., 2000a) but is independent of cortical determinants or Kar9p (Yeh et al., 2000). In light of refined proposals suggesting that Kar9p provides a bridge between MTs and actin transport to be delivered to the bud while bound to MTs exclusively (Hwang et al., 2003; Kusch et al., 2003; Liakopoulos et al., 2003), the role of Bud6p in capture at the cell cortex must be revisited and incorporated into these models. Alternatively, Bud6p has been implicated in organization of actin cables (Amberg et al., 1997; Moseley et al., 2004). According to this view, Bud6p role in MT capture is limited to supporting Kar9p function (Heil-Chapdelaine et al., 1999).

Here, we have explored the relationship between Kar9p and Bud6p. We used time-lapse analysis to examine Bud6p–MT interactions in kar9Δ cells and, conversely, the impact of a bud6Δ mutation on Kar9p-mediated MT delivery to the bud. We also analyzed kar9Δ bud6Δ cells to test for synergism between the two mutations. Finally, we showed the ability of a tub2 allele that decreases MT turnover (Gupta et al., 2002), to suppress a kar9Δ mutation in a Bud6p-dependent manner. Our data suggest that Bud6p role in MT capture at the cell cortex is separable from Kar9p-mediated delivery of MTs to the bud along actin cables.

### Results

#### MT-Bud6p dynamic interactions in kar9Δ cells

We have previously studied the dynamic behavior of astral MT–cortex interactions in wild-type cells expressing GFP-Bud6 and GFP-Tub1. This analysis showed a high incidence of MT interactions at GFP-Bud6 cortical sites throughout the cell cycle. Moreover, shrinkage of MTs at the cell cortex as an indicator of a Bud6p-dependent mode of interaction (Segal et al., 2002). To evaluate the involvement of Kar9p in Bud6p-associated MT capture, we undertook a comparative analysis of wild-type versus kar9Δ cells coexpressing GFP-Bud6 and GFP–α tubulin (Tub1p) fusions. Interactions were studied along the cell cycle divided arbitrarily into three stages based on spindle pathway landmarks and the program of Bud6p localization (Fig. 1 A).

The kar9Δ mutation did not affect MT–Bud6p contacts from mitotic exit to generation of a new budding site (ME to...
BE: 54.7%, n = 1105 in kar9Δ vs. 53.8%, n = 557 in wild type, Fig. 1 B). Once the Bud6p ring at the previous division site disassembled and accumulation began at the prebud site, MTs reoriented to this new area of capture (Fig. 2 A). In addition, interactions at Bud6p sites occurred at wild-type frequencies from onset of anaphase to mitotic exit in kar9Δ cells (Fig. 1 B, SE to ME: 76.1%, n = 822 in kar9Δ vs. 78.1%, n = 420 in wild type) and MT shrinkage at Bud6p sites was unperturbed (Fig. 1 B, open boxes within black bars).

In contrast, kar9Δ cells exhibited a marked decrease in interactions with Bud6p decorated areas after bud emergence (Fig. 1 B, BE to SE). Cells showed repeated MT interactions with the bud tip in small-budded cells (100% cells within 10 min from bud emergence, n = 32). Yet, as the bud continued to grow, cells failed to maintain MTs oriented in the bud (Fig. 2 B, arrowheads). As a result, MTs spent significant time probing the mother cell cortex away from Bud6p marked regions. However, once GFP-Bud6 decorated the bud neck, MTs resumed interactions with this discrete area (Fig. 2, C and D, arrows). In spite of the reduction in Bud6p–MT contacts in S phase, kar9Δ cells still showed MT shrinkage at Bud6p sites (Fig. 1 B).

Bud6p–astral MT interactions during anaphase in kar9Δ cells occurred at wild-type levels. These interactions contributed to spindle positioning even if anaphase began within the mother cell (Fig. 2 E).

The correlation between MT–Bud6p interactions and the duration of cortical contacts previously reported in wild-type cells (Segal et al., 2002) was still apparent in kar9Δ cells. On average, interactions at Bud6p sites lasted 2.2 ± 1.2 min (n = 75) from mitotic exit to bud emergence, 2.0 ± 0.3 min (n = 42) from bud emergence to spindle assembly and increased to 3.0 ± 1.8 min (n = 92) from spindle elongation to mitotic exit. In contrast, interactions away from Bud6p sites lasted 0.6 ± 0.2 min (n = 112). These values were in agreement with those in wild-type cells (for review see Segal et al., 2002).

Together, these results confirmed that a kar9Δ mutation did not alter the dynamic characteristics of cortical Bud6p–MT interactions, instead, it perturbed the maintenance of astral MT interactions during spindle assembly.
orientation toward the bud from late bud emergence through spindle assembly and thus, indirectly, decreased the incidence of capture at Bud6p sites.

Analysis of Kar9p-driven MT delivery to the bud in wild-type or bud6Δ cells

We then examined wild-type or bud6Δ cells expressing CFP-Tub1 and Kar9-GFP to determine the dynamic behavior of Kar9p bound to MTs, the effect on astral MT orientation to cell cortex areas and the relationship to MT-driven SPB movement toward the bud.

Decoration of MTs by Kar9-GFP was initiated by recruitment at the SPB in most cases, both in wild-type or bud6Δ cells (93.5%, n = 113 MTs and 96.2%, n = 104, respectively). Kar9p was detected at both SPBs at onset of spindle assembly (14 of 16 time-lapse series spanning spindle assembly) but was clearly asymmetric in spindles longer than 1.2 μm (Fig. 3). Kar9p traveled along MTs toward the plus or minus end. In addition, Kar9p moved while fixed at the plus end of a growing or shrinking MT (Fig. 4 A). These modes of dynamic behavior occurred significantly in bud6Δ cells. However, the bud6Δ mutation slightly reduced Kar9p translocation along persistent MTs (Fig. 4 A, black and gray bars in wild type vs. bud6Δ).

The distribution of cortical interactions involving Kar9p-bound MTs by cell compartment was altered in bud6Δ cells. In general, MTs decorated by Kar9p were already directed toward the bud or became oriented when Kar9p occupied the plus end (Fig. 4, B–D). A bud6Δ mutation markedly decreased interactions confined to the bud neck (Fig. 4 A, open portion of bars for each mode of dynamic behavior). Thus, Kar9p-directed MTs to the bud, yet, Bud6p appeared to dictate bud neck capture.

Of all Kar9p-bound MTs in wild-type cells, 31% (n = 113 MTs) were associated with SPB movement toward the bud (24% in bud6Δ cells, n = 104 MTs). To correlate the direction of movement with Kar9p dynamic behavior, all events involving Kar9p return to the spindle pole were scored for coupled SPB movement toward the cortex. As shown in Fig. 5 A, there was no correlation between the direction of SPB movement and shrinkage of Kar9p-bound MTs in wild-type or bud6Δ cells (< 7%, n > 120). In fact, the SPB remained stationary relative to the cortex in the majority of Kar9p returns to the pole.

In wild-type cells, Kar9p dynamic behavior in the absence of SPB movement was confined to the vicinity of the bud neck within the mother (Fig. 5 B). This was associated with angular movement of MTs of constant length similar to previously reported Kar9p or Myo2p-dependent transports (Hwang et al., 2003; Liakopoulos et al., 2003). Alternatively, Kar9p-bound MTs shrank away from the cortex without causing SPB movement (Fig. 5 C, arrow). In conclusion, Kar9p did not mediate SPB movement via changes in MT length. This was in contrast with, spindle orientation associated with cortical Bud6p relying primarily on MT shrinkage.
In \textit{bud6Δ} cells SPBs were initially present away from the bud and became quickly repositioned as a Kar9p-bound MT moved toward the bud without observable shrinkage, presumably, along an actin cable (Fig. 6, arrows). This type of proessive MT movements toward the bud were absent in \textit{bud6Δ kar9Δ} cells ($n > 150$ MTs). Thus, a \textit{bud6Δ} mutant supported Kar9p-dependent MT orientation and relied on long-range Kar9p-bound transport to compensate for lack of early MT–cortex interactions with the new bud and the inability to mobilize SPBs by MT shrinkage (Segal et al., 2002).

**Spindle orientation in \textit{bud6Δ kar9Δ} or \textit{bud6Δ dyn1Δ} mutants**

Genetic analysis of spindle orientation has assigned motor activities and cortical determinants to putative early and late pathways required for spindle position (Heil-Chapdelaine et al., 1999). Kar9p and dynein are regarded as key components of the “early” and “late” pathways, respectively. To further assess the contribution of Bud6p in these pathways, the phenotypes resulting from double mutant combinations \textit{bud6Δ kar9Δ} or \textit{bud6Δ dyn1Δ} (DYN1/DHC1, dynein heavy chain; Eshel et al., 1993; Li et al., 1993) were characterized.
Astral MT behavior and spindle orientation defects observed in bud6Δ kar9Δ cells demonstrated the additive impact of deleting BUD6 over a single kar9Δ mutation. Early orientation of astral MTs toward the emerging bud was abolished as in single bud6Δ mutants (not depicted; Segal et al., 2002), whereas spindle positioning in the double mutant was markedly impaired relative to either single mutant (Table I and Fig. 7). A range of phenotypes in bud6Δ kar9Δ cells highlighted the contribution of Bud6p to cortical capture in the absence of Kar9p.

First, retention of the spindle at the bud neck was compromised causing transient positioning within the bud (Fig. 7 A, 17.0–18.5 min). This phenotype was as prevalent (26%, n = 39 cells recorded) as in bud6Δ cells and never observed in kar9Δ cells (Table I).

Second, onset of spindle elongation along the mother-bud axis was markedly reduced (33%, n = 39), possibly due to the additional absence of MT interactions with the bud neck (Fig. 7 B and see last section of Results describing Fig. 9 A). This contrasted with kar9Δ cells (Table I), in which MT interactions with the bud neck still contribute toward orientation (Segal et al., 2000b; and see last section of Results describing Fig. 9 A). Initial spindle elongation within the mother cell in kar9Δ bud6Δ cells was followed by dynein-driven positioning of the spindle part way through anaphase (Fig. 7 B) as in kar9Δ cells (Segal et al., 2000b; Yeh et al., 2000).

Third, aberrant loss of spindle orientation in mid-anaphase of bud6Δ kar9Δ mutants (Fig. 7 C, 7.5–9.5 min), as observed in 13% (n = 39) of cells recorded (Table I), underscored Bud6p-dependent capture in anaphase. This indicated that dynein-driven events might be impaired yet sufficient to sustain viability of bud6Δ kar9Δ cells.

We also determined the effects of deleting BUD6 on spindle phenotypes of dyn1Δ cells, with particular focus on anaphase. In wild-type cells, the “fast phase” of spindle elongation (Fig. 8 A, 0–2.5 min and 6.0–10 min) is typically coupled to translocation of the SPB into the bud (Yeh et al., 1995). SPB translocation is delayed in dyn1Δ mutants, although spindle elongation still begins along the mother-bud axis (Table II).

### Table I. Time-lapse analysis of spindle behavior at anaphase onset in bud6, kar9, and bud6kar9 mutants

| Strain          | % lack of retention at the bud neck | % correct alignmenta | % mid-anaphase loss of positioning | n† |
|-----------------|------------------------------------|----------------------|-----------------------------------|----|
| wild type       | 0                                  | 100                  | 0                                 | 28 |
| bud6Δ           | 27                                 | 87                   | 13                                | 15 |
| kar9Δ           | 0                                  | 60                   | 0                                 | 25 |
| kar9Δ bud6Δ     | 26                                 | 33                   | 13                                | 39 |

Results are expressed as percentage relative to the total number of cells recorded.

aAlignment of the spindle at anaphase onset was considered correct if an imaginary line drawn through the long axis of the spindle transversed the mother-bud neck (Theesfeld et al., 1999) as separation of the poles began at the “fast phase” of anaphase B (Yeh et al., 1995).

†n = total number of anaphase cells recorded.

### Table II. Spindle behavior in bud6, dyn1, and bud6dyn1 mutants

| Strain          | % correctly aligned spindles at onset of anaphasea | % late anaphase spindles exhibiting correct polarityb |
|-----------------|---------------------------------------------------|---------------------------------------------------|
| wild type       | 100 (n = 28)                                      | 100 (n = 200)                                      |
| bud6Δ           | 87 (n = 15)                                       | 92 (n = 300)                                       |
| dyn1Δ           | 94% (n = 35)                                      | 90 (n = 210)                                       |
| bud6Δ dyn1Δ     | 72% (n = 18)                                      | 60 (n = 302)                                       |

aAlignment of the spindle at anaphase onset was scored as described in Table I. Results are expressed as percentage of total anaphase cells recorded (n).
bPolarity in late anaphase spindles was considered correct if a single pole was still connected to the bud cortex, irrespective of spindle alignment. Results are expressed as percentage of total anaphase cells counted (n) in asynchronous populations of the indicated strains.

In 9% of anaphase cells recorded, the spindle became overtly misaligned during anaphase, after initial elongation along the mother-bud axis (Fig. 8 C).
or

lations (onstrated by the more complex spindle defects of dynamics and progression (Segal et al., 2000b) was further dem-
dependent MT capture at the bud neck for correct spindle dy-
Bud6p to spindle orientation. In addition, the impact of Bud6p-
umsery SPBs (Fig. 8, E and F). Such cells were not present in
copy (Fig. 8 F, 6.0–10.0 min). Accordingly, asynchronous
was sufficiently frequent to be recorded by real-time micros-
Progression of the spindle pathway past spindle disassembly
phase spindles within the mother failed to restrain mitotic exit.

In contrast, bud6Δ dyn1Δ mutants already showed a mild
preanaphase spindle orientation defect and concomitant reduction
of spindle elongation along the mother-bud axis (Table II).
Spindle alignment could be rectified during early anaphase,
presumably, through Kar9p-directed astral MT delivery to the
bud (Fig. 8 D).

Impaired MT capture at the bud neck in dyn1Δ bud6Δ
cells led to loss of polarity of mid-anaphase spindles held
within the mother cell, as astral MTs emerging from both poles
entered the bud (Fig. 8 E; for review see Yeh et al., 2000). In
asynchronous populations, only 60% (n = 302) of late ana-
phase spindles in dyn1Δ bud6Δ cells retained apparent polarity
(only one SPB connected to the bud) in contrast to 90% (n = 210) in dyn1Δ cells (Table II).

Finally, bud6Δ dyn1Δ cells containing misaligned ana-
phase spindles within the mother failed to restrain mitotic exit.
Progression of the spindle pathway past spindle disassembly
was sufficiently frequent to be recorded by real-time micros-
Fig. 8 F, 6.0–10.0 min). Accordingly, asynchronous
bud6Δ dyn1Δ cultures contained 10% of cells exhibiting super-
numeryary SPBs (Fig. 8, E and F). Such cells were not present in
bud6Δ kar9Δ or single bud6Δ, kar9Δ, or dyn1Δ mutant popu-
lations (n = 1,000 cells).

The observed interactions between bud6Δ and kar9Δ or
dyn1Δ mutations stress the separate contributions of Kar9p and
Bud6p to spindle orientation. In addition, the impact of Bud6p-
dependent MT capture at the bud neck for correct spindle dy-
namics and progression (Segal et al., 2000b) was further dem-
strated by the more complex spindle defects of bud6Δ kar9Δ
or bud6Δ dyn1Δ mutants during anaphase.

Effect of reduced MT turnover on spatial distribution of MT-cortex interactions in kar9Δ cells

The observed dynamic behavior of Kar9p was difficult to re-
ocnicle with the proposed role for Kar9p in cortical capture of
MTs. Kar9p kept a fixed distance from the SPB during MT
tracking along actin cables, and frequently decorated MTs al-
ready reaching within the bud. Moreover, the phenotype of
kar9Δ cells after bud emergence (Fig. 2 B), suggested that
Kar9p might additionally control MT dynamic behavior to pro-
mote persistence of MTs in the bud. This raised the possibility
that the requirement for Kar9p could be bypassed by a decrease
in MT instability. Under these conditions, MT capture coinci-
dent with cortical Bud6p in the bud might prevail and suppress
the spindle orientation defect of kar9Δ mutants.

To address this question, time-lapse analysis was per-
formed to compare wild-type, kar9Δ, and kar9Δ tub2C354S cells
for the distribution of MT–cortex interactions by cell compart-
mament from bud emergence to assembly of ~2-μm-long spindles
(Fig. 9 A). The tub2C354S allele was introduced to reduce MT
dynamics (Gupta et al., 2002).

Wild-type cells exhibited a characteristic distribution of
MT–cortex interactions in the mother, the bud and at the bud
neck. These interactions (Fig. 9 A) led to correctly oriented pre-
anaphase spindles, which initiated elongation along the mother-
bud axis (Table III). In kar9Δ cells, interactions within the bud
were selectively decreased (Fig. 9 A). As a result, correct align-
ment at anaphase onset was reduced to 60% (n = 25; Table III).

Introduction of the tub2C354S allele in the kar9Δ mutant
restored astral MT–cortex interactions with the bud (Fig. 9, A
and B) and significantly improved preanaphase spindle ori-
tentation (Fig. 9, C–E; Table III). Astral MTs became oriented
ward the bud (Fig. 9 B) and continued to interact with the bud
cortex until anaphase (Fig. 9 E). Elongation of the spindle
and B) and significantly improved preanaphase spindle orienta-
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hward the bud (Fig. 9 B) and continued to interact with the bud
cortex until anaphase (Fig. 9 E). Elongation of the spindle

Figure 6. Kar9p-mediated MT transport in a bud6Δ cell. Selected frames from a time-lapse series showing Kar9p-mediated orientation of the SPB in a
bud6Δ cell expressing CFP-Tub1 and Kar9-GFP. Overlays of CFP-Tub1 (red) and Kar9-GFP (green) images are shown. The DIC images correspond to 0
and 9 min [arrowhead indicates the bud]. Initially, the SPB was positioned far from the prebud site (0–1.5 min). As Kar9-GFP occupied the plus end of an
MT (1.5–3.0 min, arrows), the SPB became rapidly oriented toward the budding site (5 min). Kar9-GFP then continued to direct MTs into the growing bud
(5–9 min). Numbers indicate time elapsed in minutes. Bar, 2 μm.
Thus, decreased MT turnover suppressed spindle orientation defects of kar9Δ/H9004 cells, in a Bud6p-dependent manner. This result confirmed the participation of Bud6p in MT capture at the cell cortex and pointed to an additional role for Kar9p in control of MT dynamic behavior. Indeed, Kar9p-bound MTs entering the bud underwent repeated cycles of recovery (unpublished data). Such cycles were confirmed by time-lapse analysis of wild-type cells expressing GFP-Tub1 (at least 2 cycles in 60% of MTs entering the bud, n/H11005 135) but were markedly decreased in a kar9Δ/H9004 mutant in which 97% of MTs entering the bud (n/H11005 105) underwent a single cycle of growth and shrinkage past the bud neck (Fig. S1, available at http://www.jcb.org/cgi/content/full/jcb.200407167/DC1).

Discussion

Kar9p is not directly involved in cortical capture at Bud6p sites

The presumed involvement of Bud6p in actin organization has prompted models in which the role of Bud6p in spindle orientation is based on supporting Kar9p function (Heil-Chapdelaine, et al., 1999; Miller et al., 1999). These models assumed that either Kar9p localization at the bud cortex requires actin or, alternatively, Kar9p bound to MT plus ends interfaces with Myo2p-dependent transport. Accordingly, by perturbing the actin cytoskeleton, a bud6Δ mutation disrupts Kar9p-mediated spindle orientation.

Here, we show that the dynamic properties of Bud6p–MT interactions are unchanged by a kar9Δ mutation. MTs kinetically followed Bud6p from onset of anaphase until bud emergence in the next cell cycle as in wild-type cells (Figs. 1 and 2). In addition, Bud6p-dependent MT shrinkage coupled to SPB movement, was unperturbed. A kar9Δ mutation decreased the orientation and maintenance of MTs toward the bud in small-
budded cells (Fig. 1 B and Fig. 2 B) thus, reducing Bud6p–MT interactions during S phase. Yet, kar9Δ cells were proficient in MT–Bud6p interactions at the bud neck after SPB separation (Fig. 2, C and D).

Together, these results indicated that MT capture at Bud6p sites is independent of Kar9p, yet, the efficiency by which MTs entered and were maintained in the bud depended on Kar9p. The role of Kar9p became critical during S phase un-
Figure 9. Effect of decreased MT turnover on orientation of MT-cortex interactions in kar9A cells. (A) Astral MT-cortex interactions in the indicated strains expressing GFP-Tub1 (number of cells recorded: 81 wild type, 77 kar9A, 64 kar9A tub2C354S, 52 kar9A tub2C354S bud6Δ, 65 kar9A bud6Δ, 51 bud6Δ tub2C354S, and 45 bud6Δ) were scored by cell compartment (mother, bud neck, or bud) from bud emergence to preanaphase spindle assembly. Error bars indicate 95% confidence limits. (B–E) Selected frames from representative time-lapse series showing orientation of MT-cortex interactions in kar9A tub2C354S GFP-TUB1 cells. (B) Early orientation of astral MT–cortex interactions toward the prebud site occurred after mitotic exit (9.0 min). Orientation was maintained throughout bud emergence (16.0–34.5 min). DIC images corresponding to 0, 9.0, 16.5, and 34.5 min are shown. (C) A small budded cell showing an astral MT oriented toward the bud (0 min) maintained its orientation throughout spindle assembly and alignment (18.0–23.0 min) of the preanaphase spindle along the mother-bud axis. DIC images correspond to the first and last frame of the series. (D) Selected frames from a time-lapse series showing preanaphase spindle orientation. A cell completed spindle assembly (0–7.5 min), maintaining its orientation throughout spindle assembly and alignment (18.0–23.0 min) of the preanaphase spindle along the mother-bud axis. DIC images correspond to the first and last frame of the series. (E) Selected frames from a time-lapse series showing failure to orient MTs toward the emerging bud in a kar9A bud6Δ tub2C354S GFP-TUB1 cell. (F and G) Time-lapse series showing failure to orient MTs toward the emerging bud in a kar9A bud6Δ tub2C354S GFP-TUB1 cell. (F) Astral MTs interacted with the cell cortex away from the bud. DIC images correspond to the first and last frame. (G) Defective preanaphase spindle orientation (0–15.0 min) and misaligned spindle elongation (20.0–29.0 min) followed by spindle orientation through dynein-dependent interactions (44.5–53.0 min) in mid-anaphase. Numbers indicate time elapsed in minutes. Bars, 2 μm.
til preanaphase orientation was accomplished, demarking the end of the actin-sensitive period of spindle orientation (Theesfeld et al., 1999). Bud6p functions in cortical capture beyond this period further support a role that is separable from its links with the actin cytoskeleton (Moseley et al., 2004).

Kar9p-mediated delivery of astral MTs to the bud occurs in bud6Δ cells

To correlate the behavior of Kar9p with parameters of astral MT function, we examined wild-type cells coexpressing CFP-Tub1 and Kar9-GFP. We also asked whether a bud6Δ mutation perturbed the behavior of Kar9p-bound MTs in support of a functional link between Bud6p and Kar9p.

Kar9p was present at both SPBs during spindle assembly but became asymmetric once the identity of the poles was defined (Fig. 3). This dependency on correctly polarized MT–cortex interactions for asymmetric recruitment to the SPBx resembled that reported for mitotic exit regulators (Pereira et al., 2001; Smeets and Segal, 2002). Accordingly, Maekawa and Schiebel (2004) demonstrated that Cdc28p-Clb4p is recruited, in fact, to the SPBx via asymmetrically localized Kar9p. This precludes the converse view that Cdc28p-Clb4p prevents recruitment of Kar9p at the mother-bound SPB (SPBm) to generate polarity.

Kar9p traveled along MTs or moved when fixed at the MT plus end during periods of MT shortening or growth. Yet, association of Kar9p with an MT plus end did not always coincide with its interaction with the cell cortex (Fig. 4, B–D) or necessarily precede orientation of MTs toward the bud. These findings are not compatible with Kar9p serving solely as a cortical anchor or a bridge with the actin cytoskeleton.

Kar9p-bound MTs were directed to the bud in bud6Δ cells (Figs. 4 and 6), whereas events ending at the bud neck were markedly reduced (Fig. 4 A). Thus, a bud6Δ mutation did not perturb Kar9p-based MT delivery but impaired interactions with the bud neck or bud cortex.

The role of septins in MT capture at the bud neck has been previously addressed but was inferred to reflect Kar9p-dependent capture (Kusch et al., 2002). Yet, a bud6Δ mutation was not evaluated in that work. Because MT capture at the bud neck is proficient in kar9Δ cells (Fig. 2), whereas defective in a bud6Δ mutant (Fig. 4; Segal et al., 2002), it is clear that Bud6p, rather than Kar9p, creates an area of specialized capture at the bud neck. Indeed, disruption of the septin ring after a tempera-

![Figure 10. The role of Bud6p and Kar9p in cortical capture. A model summarizing astral MT dynamic behavior associated with Bud6p or Kar9p during MT capture and establishment of spindle polarity.](image-url)
ture shift in cdc3Δ, cdc10Δ, or cdc11Δ strains impaired Bud6p localization to the bud neck in S phase (Fig. S2, available at http://www.jcb.org/cgi/content/full/jcb.200407167/DC1), explaining the disruption of astral MT–bud neck interactions in these mutants.

Finally, Kar9p-bound MTs did not support shrinkage coupled to SPB movement toward the cortex (Fig. 5 A). Interestingly, although the SPBd is associated with, on average, three MTs in wild-type cells only one is usually decorated by Kar9p at any given time. Thus, MT shrinkage coincident with Bud6p or Kar9p-associated transport may take place on different MT subpopulations (Fig. 10). Indeed, contrary to the high frequency of MT interactions with cortical Bud6p, Kar9-GFP rarely colocalized with CFP-Bud6 (unpublished data).

The analysis presented here showed separable contributions of Bud6p and Kar9p toward spindle orientation summarized in Fig. 10. Earlier events would be dictated by the Bud6p cortical program of MT interactions and enforced by progressive asymmetric recruitment of Kar9p to the SPBd. Bud6p would couple SPB movement with cortical capture, whereas Kar9p would secure delivery of MTs from the SPBd to the bud. In addition, Kar9p transits along MTs within the bud may preserve these MTs from shrinking past the bud neck and instead redirect them to interact with actin cables via Myo2p.

**Genetic analysis places Bud6p in the “early” and “late” pathways of spindle orientation**

Genetic analysis has assigned motors, MT binding proteins and putative cortical anchors to “early” and “late” pathways participating in spindle orientation (Heil-Chapdelaine et al., 1999; Pearson and Bloom, 2004). Bud6p cortical program underlies the orientation of MT–cortex interactions throughout the spindle pathway (Segal et al., 2002). Yet, Bud6p relationship with actin organization (Moseley et al., 2004) pointed to links with the early Kar9p-dependent pathway.

Here, we show that kar9Δ bud6Δ cells exhibited phenotypes suggestive of synergism between the two mutations (Fig. 7 and Table I). These cells still relied on dynein-driven MT sliding to orient anaphase spindles. These interactions, however, were not completely proficient as evidenced by transient loss of orientation in the late portion of anaphase (Fig. 7 C).

Additive effects were also observed in bud6Δ dyn1Δ cells. Preanaphase spindle orientation was comparable to that of bud6Δ single mutants, yet, defective cortical capture at the bud neck also resulted in disruption of spindle polarity in anaphase cells (Fig. 8 and Table II).

Failure to translocate a spindle pole into the bud during anaphase blocks mitotic exit and cell division, under the control of the mitotic exit network (Bardin and Amon, 2001; McCollum and Gould, 2001). This surveillance mechanism is based on the separation between the upstream activator of the network Lte1p confined to the bud cortex, and its effector, the small GTPase Tem1p, localized to the SPBd. Once the SPBd translocates into the bud, these two regulators may interact to trigger mitotic exit. Adames et al. (2001) have proposed that cortical capture at the bud neck is also critical for this control.

In agreement with this proposal, dyn1Δ bud6Δ cells failed to restrain spindle disassembly within the mother cell. This underscores the importance of Bud6p in cortical capture at the bud neck as budd1Δ cells still compartmentalize Lte1p to the bud (Jensen et al., 2002). Conversely, budd1Δ cells experience a delay in mitotic exit when astral MTs from the SPBd, already positioned in the bud, grow aberrantly past the bud neck and interact with the mother cell cortex (Segal et al., 2002).

To orient the spindle, MT-based motors must generate force relative to an attachment site to result in net displacement of the spindle pole (Pearson and Bloom, 2004). Num1p provides an anchor for dynein-driven sliding movements along the cortex as the spindle translocates into the bud (Heil-Chapdelaine et al., 2000). In contrast, the cortical anchor promoting dynein-dependent MT shrinkage (Carminati and Stearns, 1997) has not been assigned. Our studies point to the involvement of Bud6p in this mode of interaction. Indeed, Bud6p-dependent SPB repositioning after cytokinesis, which relies on MT shrinkage in association to the Bud6p ring (Segal et al., 2002), is abolished by a dyn1Δ mutation (unpublished data). Moreover, MT–Bud6p interactions during anaphase are preserved in the absence of Num1p (Segal et al., 2002). Thus, different modes of MT–cortex interactions might be modulated by association of MT-bound dynein–dynactin complex with alternative cortical anchors. This would add an additional layer of complexity to the control of competing modes of MT dynamic behavior accompanying the transitions from Kar9p to dynein-driven spindle positioning (Segal and Bloom, 2001).

**Is MT plus end delivery to the bud the sole function of Kar9p?**

Oriented MT capture at Bud6p sites in the bud still occurred in kar9Δ cells. Yet, orientation was not maintained as the bud continued to grow (Fig. 2). Therefore, we tested whether reduced MT instability could suppress kar9Δ phenotypes. Indeed, reduced MT turnover restored spindle orientation in kar9Δ cells dependent on Bud6p (Fig. 9), pointing to a role for Kar9p in maintaining MTs within the bud once Bud6p–dependent capture had targeted these MTs to the bud cortex. Consistent with this notion, Kar9p appeared to prevent MTs from undergoing shrinkage past the bud neck once inside the bud (Fig. S1).

We have previously proposed that Kar9p may affect MT dynamics in addition to its role as a bridge to the actin cytoskeleton (Segal and Bloom, 2001). Interestingly, the human adenomatous polyposis coli, the proposed counterpart of Kar9p, is critical for EB1–dependent MT polymerization. Adenomatous polyposis coli is subject to phosphorylation by Cdk controlling its binding to EB1 (Nakamura et al., 2001). Similarly, phosphorylation by Cdk may control Kar9p binding to Bim1p although the significance of this control is unclear (Liakopoulos et al., 2003). Finally, another work implicates Kar9p in modulation of MT dynamic behavior (Maekawa and Schiebel, 2004). According to this report, Kar9p targets Cdk to plus ends of MTs generated by the SPBd to control differential MT–cortex interactions at the bud tip or bud neck. The substrates mediating this control remain unknown.
Mechanisms of cortical capture are conserved between yeast and higher eukaryotes. Thus, the continued effort to dissect the regulation of dynamic aspects of MT–cortex interactions in yeast will yield new clues to understand the spatial control of MT capture, and its impact on developmental programs relying on regulated spindle orientation for the generation of cell division in metazoans.

Materials and methods

Yeast strains, plasmids, and genetic procedures
Yeast strains were isogenic to 15Dau (Segal et al., 2000a). The deletion alleles budΔ, kar9Δ, and dyn1Δ were generated using KAN4 cassettes amplified by PCR (Segal et al., 2000b). Strains expressing a mutant β-tubulin conferring reduced MT dynamics were created by transforming yeast cells with a linear SacI–SphI fragment from pCS3-C354S (provided by R. Himes, University of Kansas, Lawrence, KS) encoding tub2ΔC354S (Gupta et al., 2002). A cdc3Δ strain and isogenic wild-type strains were a gift from M. Longtime [Oklahoma State University, Stillwater, OK]. Strains expressing a GFP-Tub1 and GFP-Bud6 fusion were obtained by transformation with pAFS2Z [Straight et al., 1997] and pR82190 (Amberg et al., 1997), respectively. pCFP-TUB1 was used to express a CFP-tubulin fusion (Jensen et al., 2001). pRS404bKARP-GFP contained a 495bp SacI–NotI fragment generated by PCR for 3’in-frame fusion to GFP. Digestion with NotI and SacI targeted the construct for integration at the endogenous KARP. Standard yeast genetic procedures were used (Sherman et al., 1986). Cells were grown at 25°C unless indicated.

Microscopy methods
Cells were grown to ~5 × 10^6 cells/ml in selective dextrose medium and then mounted in the same medium containing 25% gelatin to perform time-lapse recordings at RT (Maddox et al., 1999) using a Nikon Eclipse E800 microscope equipped with a CFI Plan Apochromat 100X, N.A. 1.4 objective, Chroma Technology Corp. Filter sets and a Cool SNAP HQ CCD camera (Roper Scientific). Images were acquired using 2 × 2 binning. For cells expressing GFP fusions, five fluorescence images were acquired at a z-distance of 0.8 μm between planes. A single differential interference contrast (DIC) image was taken in the middle focal plane. This acquisition regime was repeated at 10-, 15-, or 30-s intervals. Images were processed using Metamorph software (Universal Imaging Corp., Middletown, PA). The validity and considerations on the resolution of the analysis to assess dynamic interactions between cortical Bud6p and MTs have been addressed previously (Segal et al., 2002).

Interactions at or away from Bud6p were categorized as described previously (Segal et al., 2002). For simplicity, MT shrinkage at the cortex was the only category scored individually. This mode of interaction occurs at Bud6p, cortical sites and is abolished in budΔ cells (Segal et al., 2002). For this analysis, the cell cycle was arbitrarily divided into three stages (Fig. 1): (1) from mitotic exit (ME) to generation of the new prebud site (from spindle disassembly and formation of a double ring of Bud6p at the division site till appearance of the Bud6p ring and relocation to the new bud site); (2) from bud emergence (BE) to spindle assembly and orientation; and (3) from spindle elongation (SE) to ME. For the third stage, only interactions involving astral MTs from the SPB were considered.

Duration of cortical interactions was determined by following the history of 50 MTs. Mean values correspond to the total time MTs contacted the cortex divided by the number of contact events (n). Results were expressed as mean ± SD.

Cells expressing CFP-Tub1 and Kar9-GFP fusions were recorded using a modification of the protocol that discriminates between CFP and GFP using a CFP/YFP filter set (Pearson et al., 2001). Three fluorescence images were acquired at a z-distance of 0.8 μm. Z-stacks were acquired at 12-, 15-, 30-, or 60-s intervals. Kar9-GFP dynamic behavior was categorized as follows: (1) Kar9p movement along an MT toward the plus end; (2) Kar9p movement toward the SPB along a persistent MT; (3) Kar9p movement at the plus end of a growing MT; (4) Kar9p movement at the plus end of a shrinking MT; and (5) Kar9p association to a plus end without recruitment via the SPB. Categories 1 and 2 included all events involving Kar9p presence along an MT, whereas categories 3 and 4 corresponded to all events in which Kar9p at the MT plus end moved as a consequence of MT growth and shrinkage.

To correlate Kar9p-bound MT shortening and SPB movement, Kar9p returns toward the spindle pole were scored for associated movement of the SPB as follows: (a) no movement (the SPB remained stationary); (b) movement away from the cortex (unrelated to the direction of the shortening MT); and (c) movement coupled to shortening (toward the cortex).

Quantitations described in Fig. 2 included data derived from 54 wild-type cells (total of 65.5 h recorded) and 172 kar9Δ cells (140 h). Analysis of cells coexpressing GFP-Tub1 and Kar9-GFP included recorded data of 107 wild-type cells [50 h] and 99 budΔ cells [31 h]. Quantitations described in Fig. 9 represent recorded data from the following strains expressing GFP-Tub1: 1) wild-type cells [89 h]; 77 kar9Δ cells [76 h]; 64 kar9Δ tub2ΔC354S cells [73 h]; 51 budΔa tub2ΔC354S cells [50.5 h]; 52 budΔa kar9Δ tub2ΔC354S [56.8 h]; 65 kar9Δ budΔa cells [83 h]; and 45 budΔa cells [70 h].

Comparative analysis of budΔa, kar9Δ, dyn1Δ, budsΔ kar9Δ and dyn1Δ budsΔ cells expressing GFP-Tub1 was performed using the same set of recordings described above. In addition, 31 dyn1Δ budsΔ cells [26.2 h] and 36 dyn1Δ cells [50 h] were recorded.

Online supplemental material
Fig. S1 shows representative time-lapse series of wild-type or kar9Δ cells expressing GFP-Tub1 showing dynamic behavior of MTs entering the bud. Fig. S2 shows GFP-Bud6 localization in cdc3Δ cells. Online material is available at http://www.jcb.org/cgi/content/full/jcb.200407167/DC1/C.

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