Metabolic reprogramming: a new relevant pathway in adult adrenocortical tumors

Céline Pinheiro¹,²,³,⁴,*, Sara Granja¹,²,*, Adhemar Longatto-Filho¹,²,⁴,⁵, André M. Faria⁶, Maria C. B. V. Fragoso⁶,⁷, Silvana M. Lovisolo⁸, Antonio M. Lerário⁷, Madson Q. Almeida⁶,⁷, Fátima Baltazar¹,²,*, Maria C. N. Zerbini⁹,*

¹Life and Health Sciences Research Institute (ICVS), School of Health Sciences, University of Minho, Braga, Portugal
²ICVS/3B’s-PT Government Associate Laboratory, Braga/Guimarães, Portugal
³Barretos School of Health Sciences Dr. Paulo Prata – FACISB, Barretos, Sao Paulo, Brazil
⁴Molecular Oncology Research Center, Barretos Cancer Hospital, Barretos, Sao Paulo, Brazil
⁵Laboratory of Medical Investigation (LIM-14), Faculdade de Medicina da Universidade de São Paulo, São Paulo, Brazil
⁶Unidade de Suprarrenal, Disciplina de Endocrinologia e Metabologia, Laboratório de Hormônios e Genética Molecular LIM42, Hospital das Clínicas, Faculdade de Medicina da Universidade de São Paulo, São Paulo, Brazil
⁷Instituto do Câncer do Estado de São Paulo - ICESP, Hospital das Clínicas, Faculdade de Medicina da Universidade de São Paulo, São Paulo, Brazil
⁸Hospital Universitário, Faculdade de Medicina da Universidade de São Paulo, São Paulo, Brazil
⁹Departamento de Patologia, Faculdade de Medicina da Universidade de São Paulo, São Paulo, Brazil

*These authors have contributed equally to this work

Correspondence to:
Céline Pinheiro, e-mail: celinepinheiro@gmail.com

Keywords: adrenocortical tumors, metabolic reprogramming, monocarboxylate transporter, Warburg effect

Received: May 09, 2015  Accepted: November 06, 2015  Published: November 16, 2015

ABSTRACT

Adrenocortical carcinomas (ACCs) are complex neoplasias that may present unexpected clinical behavior, being imperative to identify new biological markers that can predict patient prognosis and provide new therapeutic options. The main aim of the present study was to evaluate the prognostic value of metabolism-related key proteins in adrenocortical carcinoma. The immunohistochemical expression of MCT1, MCT2, MCT4, CD147, CD44, GLUT1 and CAIX was evaluated in a series of 154 adult patients with adrenocortical neoplasia and associated with patients’ clinicopathological parameters. A significant increase was found for membranous expression of MCT4, GLUT1 and CAIX in carcinomas, when compared to adenomas. Importantly MCT1, GLUT1 and CAIX expressions were significantly associated with poor prognostic variables, including high nuclear grade, high mitotic index, advanced tumor staging, presence of metastasis, as well as shorter overall and disease free survival. In opposition, MCT2 membranous expression was associated with favorable prognostic parameters. Importantly, cytoplasmic expression of CD147 was identified as an independent predictor of longer overall survival and cytoplasmic expression of CAIX as an independent predictor of longer disease-free survival. We provide evidence for a metabolic reprogramming in adrenocortical malignant tumors towards the hyperglycolytic and acid-resistant phenotype, which was associated with poor prognosis.

INTRODUCTION

Adrenocortical carcinoma (ACC) is a rare and highly aggressive malignancy with an annual incidence of 0.7–2.0 cases per million [1–3]. Surgery is the standard of care for localized adrenocortical carcinomas. To date, systemic treatment of advanced ACC has shown unsatisfactory overall response [4, 5]. Over the past two decades, considerable advances have been made into the understanding of molecular mechanisms, but is still not yet satisfactory [1, 2, 6].

Adrenocortical neoplasms have been a great challenge since they may present an unexpected biological...
behavior when using classic features commonly used for histopathological classification of neoplastic lesions, particularly in the borderline Weiss (scores 3 and 4) group. Therefore, it is imperative to find alternative information that can help determine with greater accuracy the possible biological behavior of patients with this disease, and, consequently, establish a correct therapeutic strategy [7, 8].

During carcinogenesis and tumor progression, neoplastic cells may reprogram their metabolism, showing a preference for glycolysis pathway for energy production, even in the presence of oxygen, a phenomenon known as the "Warburg effect". This aerobic glycolysis implies the conversion of pyruvate to lactic acid, leading to a reduction in intracellular pH [9]. To prevent acid-induced apoptosis as well as glycolysis inhibition by accumulation of the end product, cancer cells upregulate proteins related to pH regulation and lactate transport, including the glucose transporter GLUT1, the pH regulator carbonic anhydrase IX (CAIX) [10, 11] and monocarboxylate transporters (MCTs) [11]. Therefore, many malignancies show a significant increase in the expression of these plasma membrane transporters, with associations with poor patient’s prognosis [12–15]. The MCT family has 14 members, being isoforms 1, 2 and 4 the most well studied isoforms responsible for the transport of monocarboxylates, including lactate, coupled with a proton across the plasma membrane. As consequence from their substrate affinities, MCT1 and MCT4 mediate monocarboxylate efflux from cells, while MCT2 is involved in monocarboxylate uptake [16]. Importantly, these transporters require co-expression with chaperones for proper plasma membrane localization and activity. The main chaperone of MCT1 and MCT4 is CD147 [17, 18], while MCT2 is mainly associated with gp70 [18]. CD44 has also been recently described as a MCT chaperone [19]; however CD147 and CD44 expressions do not account for all MCT1/4 positive cases, suggesting that an additional MCT chaperone still remains to be identified [20].

To the best of our knowledge, the metabolic profile of adrenocortical tumors is very little explored [21]. However, 18F-fluorodeoxyglucose positron emission tomography (FDG-PET) data suggest that adrenocortical carcinomas show high levels of glucose consumption [22–25], indicating a possible clinical relevance of the glycolytic metabolism for the management of this neoplasia. Given the importance of these neoplasms, particularly in the Brazilian environment [26, 27], and the potential use of molecular players of aerobic glycolysis as prognostic tools and therapeutic targets [10, 12, 28], the aim of the present work is to study the pattern of expression of the metabolism-related proteins MCT1, MCT2, MCT4, their chaperones CD147 and CD44, as well as GLUT1 and CAIX in adult ACC, and to determine whether these proteins have some biological and/or prognostic predictive value.

RESULTS

Expression of MCTs, CD147, CD44, GLUT1 and CAIX in adrenocortical adenomas and carcinomas

The immunohistochemical evaluation of these metabolism-related proteins in adrenocortical adenomas and carcinomas shows that all proteins can exhibit cytoplasmic, plasma membrane or simultaneous expression in both localizations, with a predominance of plasma membrane expression (exception for CAIX, Figures 1 and 2). As can be seen in Figure 2, all 3 MCT isoforms were expressed in the plasma membrane of most adrenocortical carcinoma samples (around 60–65%) and, MCT4 expression was significantly higher in carcinomas than in adenoma cells (66.7% versus 20.0%, respectively, p < 0.001). Cytoplasmic expression frequencies were more heterogeneous among MCT isoforms and did not differ between adenomas and carcinomas. CD147 was the protein more frequently expressed in the plasma membrane, but, similarly to CD44, no significant difference was observed between adenomas and carcinomas (Figure 2). In the carcinoma group, an increased expression was observed in the cytoplasm and plasma membrane for GLUT1 (p < 0.001 and p < 0.001, respectively, Figure 2), as well as in the plasma membrane for CAIX (p < 0.001, Figure 2). When evaluating the co-expression of MCTs with the other proteins at the plasma membrane (Table 1), we found co-expression of MCT1 and either CD147 (p = 0.001) or GLUT1 (p = 0.007), MCT2 co-expressed positively with CD147 (p < 0.001) and inversely with CAIX (p = 0.023), and MCT4 co-expressed with CD147 (p = 0.005), CD44 (p = 0.031), GLUT1 (p = 0.001) and CAIX (p < 0.001).

Clinicopathological significance of the metabolism-related proteins

The analysis of MCT plasma membrane expression association with the clinicopathological parameters is shown in Table 2. MCT1 was significantly associated with stage III+IV (p = 0.029), MCT2 was significantly associated with smaller tumor size (p = 0.004), lower mitotic index (p = 0.008), absence of sinus invasion (p = 0.024) and absence of metastasis (p = 0.022), while MCT4 showed no significant associations with the clinicopathological data. Table 3 shows the associations of CD147, CD44, GLUT1 and CAIX plasma membrane expression with the clinicopathological parameters. CD147 was significantly associated with smaller tumor size (p = 0.009), GLUT1 was significantly associated with higher mitotic index (p = 0.011) and presence of metastasis (p = 0.004), CAIX was significantly associated with higher nuclear grade (p = 0.048) and presence of metastasis (p = 0.021), while CD44 showed no significant associations with the clinicopathological data. When
evaluating the cytoplasmic expression of the proteins under study, MCT4 and CD147 were significantly associated with absence of metastasis and CAIX with absence of necrosis (data not shown).

Survival analysis

Overall survival analysis (Figure 3) shows MCT1 and GLUT1 plasma membrane expression significantly associated with shorter overall survival ($p = 0.033$ and $p = 0.005$, respectively), MCT2 plasma membrane expression significantly associated with longer overall survival ($p = 0.009$), a tendency for CD147 cytoplasmic expression to be associated with longer overall survival ($p = 0.058$, data not shown) and CAIX plasma membrane expression associated with shorter overall survival ($p = 0.051$). Disease-free survival analysis (Figure 4) shows MCT4, CD147 and CAIX cytoplasmic expression to be significantly associated with longer disease-free survival ($p = 0.045$, $p = 0.019$ and $p = 0.045$, respectively), GLUT1 plasma membrane expression associated with shorter disease-free survival ($p = 0.007$) and a tendency for CAIX plasma membrane expression to be associated with shorter disease-free survival ($p = 0.064$, data not shown). The predictive prognostic values of the proteins were analyzed by means of Cox regression analysis.

Figure 1: Immunohistochemical expression of MCT1 A, MCT2 B, MCT4 C, CD147 D, CD44 E, GLUT1 F, and CAIX G in adrenocortical carcinomas. All the proteins were more importantly found in the plasma membrane of cells.

Figure 2: Frequency of staining of the different proteins analyzed in adrenocortical adenomas and carcinomas. Pearson’s chi-square ($\chi^2$) test was used to assess differences of expression frequency between adenomas and carcinomas. *$p < 0.001$
proportional hazards regression models (Table 4). Univariate analysis showed similar results to the ones obtained with Kaplan-Meier analysis. Multivariate analysis showed that CD147 cytoplasmic expression was the only factor with predictive value for overall survival, with a hazard ratio of 0.32 ($p = 0.008$), and that CAIX cytoplasmic expression was the only factor with predictive value for disease-free survival, with a hazard ratio of 0.31 ($p = 0.036$). Tumor stage according to ENSAT system confirmed to be a strong prognostic factor for both overall survival and disease-free survival in the univariate and multivariate analysis.

**Glycolytic versus oxidative phenotype**

The major findings described so far allowed us to establish 2 predominant expression profiles, the glycolytic and the oxidative phenotypes (summarized in Figure 5), which were associated with the clinicopathological parameters. The staining patterns of the metabolism-related proteins in cases representative of the glycolytic and oxidative phenotypes are shown in Figure 6 and Figure 7, respectively. The glycolytic phenotype was significantly associated with the mitotic index of the cases [8.3% (2/24) of cases with low mitotic index show a glycolytic phenotype versus 35.3% (6/17) of cases with high mitotic index; $p = 0.049$] and tumor p53 status [10.5% (2/19) of cases with normal p53 show a glycolytic phenotype versus 80.0% (4/5) of cases with mutated p53; $p = 0.006$].

**DISCUSSION**

The increased frequency of MCT4, GLUT1 and CAIX plasma membrane expression of in adrenal cortical carcinomas suggests a metabolic remodeling of malignant cells towards a hyperglycolytic and acid-resistant phenotype, which is compatible with data from FDG-PET [22–25]. MCT2, mainly responsible for the uptake of monocarboxylates into cells [16], as expected, showed a slight but not significant decrease in the frequency of expression from adenomas to carcinomas, which is compatible with the metabolic reprogramming towards aerobic glycolysis in carcinomas. Although CD147 and CD44 showed no increase in expression in carcinomas, they were more frequently expressed in MCT positive tumors, which is in accordance with their role as MCT chaperones [17–19]. Notably, CD147 and CD44 did not account for all MCT1 or MCT4 positive tumors, suggesting the existence of an additional MCT chaperone, as already advocated by other study [20].

As vastly described, the metabolic reprogramming of cancer cells is also involved in cancer cell aggressiveness and therapeutic resistance, rendering patients a poor prognosis [10]. The prognostic value of the metabolism-related proteins evaluated in the present study has been studied in a variety of tumor types; however, only GLUT1 has been evaluated in adrenocortical carcinoma [21]. In Fenske and colleagues study, GLUT1 was expressed in 33% of adrenocortical carcinoma samples, which is

| CD147 | CD44 | GLUT1 | CAIX |
|-------|------|-------|------|
| $n$   | $n$  | $n$   | $n$  |
| Positive (%) | Positive (%) | Positive (%) | Positive (%) |
| p     | p     | p     | p     |
|       |       |       |       |
| MCT1  |       |       |       |
| Negative | 66   | 65    | 66    | 66 |
|         | 39 (59.1) | 11 (16.9) | 8 (12.1) | 14 (21.2) |
| Positive | 88   | 87    | 88    | 88 |
|         | 74 (84.1) | 24 (27.6) | 27 (30.7) | 29 (33.0) |
| MCT2  |       |       |       |       |
| Negative | 50   | 50    | 50    | 50 |
|         | 26 (52.0) | 8 (16.0) | 14 (28.0) | 20 (40.0) |
| Positive | 103  | 102   | 103   | 103 |
|         | 87 (84.5) | 27 (26.5) | 21 (20.4) | 23 (22.3) |
| MCT4  |       |       |       |       |
| Negative | 86   | 85    | 86    | 86 |
|         | 56 (65.1) | 14 (16.5) | 11 (12.8) | 13 (15.1) |
| Positive | 67   | 67    | 67    | 67 |
|         | 57 (85.1) | 21 (31.3) | 24 (35.8) | 30 (44.8) |
Table 2: Association of plasma membrane expression of MCTs with the clinicopathological parameters in adult adrenocortical carcinomas

| Parameter          | MCT1 | p     | MCT2 | p     | MCT4 | p     |
|--------------------|------|-------|------|-------|------|-------|
|                    | Positive (%) |   | Positive (%) |   | Positive (%) |   |
| Tumour size        |                  | 0.111 |                  | 0.003 |                  | 0.536 |
| <8 cm              | 18 | 14 (77.8) |   | 17 (94.4) |   | 11 (61.1) |   |
| ≥8 cm              | 58 | 33 (56.9) |   | 33 (56.9) |   | 40 (69.0) |   |
| Tumour weight      |                  | 0.417 |                  | 0.284 |                  | 0.781 |
| <467.2 mg          | 30 | 18 (60.0) |   | 21 (70.0) |   | 21 (70.0) |   |
| ≥467.2 mg          | 30 | 21 (70.0) |   | 17 (56.7) |   | 20 (66.7) |   |
| Weiss score        |                  | 0.382 |                  | 0.400 |                  | 0.881 |
| <6                 | 33 | 19 (57.6) |   | 20 (60.6) |   | 22 (66.7) |   |
| ≥6                 | 40 | 27 (67.5) |   | 28 (70.0) |   | 26 (65.0) |   |
| Nuclear grade      |                  | 0.304 |                  | 1.000 |                  | 0.620 |
| Low                | 5  | 2 (40.0)   |   | 4 (80.0)   |   | 3 (60.0)   |   |
| High               | 36 | 26 (72.2)  |   | 27 (75.0)  |   | 26 (72.2)  |   |
| Mitotic index      |                  | 0.790 |                  | 0.008 |                  | 0.296 |
| Low                | 24 | 16 (66.7)  |   | 22 (91.7)  |   | 15 (62.5)  |   |
| High               | 17 | 12 (70.6)  |   | 9 (52.9)   |   | 14 (82.4)  |   |
| Atypical mitosis   |                  | 0.164 |                  | 0.241 |                  | 0.276 |
| Absent             | 28 | 17 (60.7)  |   | 23 (82.1)  |   | 18 (64.3)  |   |
| Present            | 13 | 11 (84.6)  |   | 8 (61.5)   |   | 11 (84.6)  |   |
| Tissue p53 status  |                  | 0.118 |                  | 1.000 |                  | 0.272 |
| Normal             | 19 | 10 (52.6)  |   | 10 (52.6)  |   | 12 (63.2)  |   |
| Mutated            | 5  | 5 (100.0)  |   | 3 (60.0)   |   | 5 (100.0)  |   |
| Necrosis           |                  | 1.000 |                  | 0.700 |                  | 0.701 |
| Absent             | 11 | 8 (72.7)   |   | 9 (81.8)   |   | 7 (63.6)   |   |
| Present            | 30 | 20 (66.7)  |   | 22 (73.3)  |   | 22 (73.3)  |   |
| Venous invasion    |                  | 0.481 |                  | 0.064 |                  | 0.165 |
| Absent             | 27 | 17 (63.0)  |   | 23 (85.2)  |   | 17 (63.0)  |   |
| Present            | 14 | 11 (78.6)  |   | 8 (57.1)   |   | 12 (85.7)  |   |
| Sinus invasion     |                  | 0.645 |                  | 0.024 |                  | 1.000 |
| Absent             | 35 | 23 (65.7)  |   | 29 (82.9)  |   | 25 (71.4)  |   |
| Present            | 6  | 5 (83.3)   |   | 2 (33.3)   |   | 4 (66.7)   |   |
| Capsular invasion  |                  | 1.000 |                  | 0.700 |                  | 1.000 |
| Absent             | 27 | 19 (70.4)  |   | 21 (77.8)  |   | 19 (70.4)  |   |
| Present            | 13 | 9 (69.2)   |   | 9 (69.2)   |   | 9 (69.2)   |   |
| Staging            |                  | 0.029 |                  | 0.186 |                  | 0.674 |

(Continued)
|                | MCT1          |          | MCT2          |          | MCT4          |          |
|----------------|---------------|----------|---------------|----------|---------------|----------|
|                | Positive (%)  | p        | Positive (%)  | p        | Positive (%)  | p        |
| I+II           | 43            |          | 22 (51.2)     |          | 31 (72.1)     |          |
|                |               |          | 19 (57.6)     |          | 23 (69.7)     |          |
| III+IV         | 33            |          | 25 (75.8)     |          | 19 (57.6)     |          |
|                |               |          | 28 (65.1)     |          | 23 (69.7)     |          |
| Metastasis     |               | 0.079    | 0.022         | 0.200    |               |          |
| Absent         | 37            |          | 19 (51.4)     |          | 29 (78.4)     |          |
|                |               |          | 22 (53.7)     |          | 30 (73.2)     |          |
| Present        | 41            |          | 29 (70.7)     |          | 22 (53.7)     |          |

Table 3: Association of plasma membrane expression of CD147, CD44, GLUT1 and CAIX with the clinicopathological parameters in adult adrenocortical carcinomas

|                | CD147         |          | CD44          |          | GLUT1         |          | CAIX         |          |
|----------------|---------------|----------|---------------|----------|---------------|----------|--------------|----------|
|                | n             | Positive (%) | p          | n             | Positive (%) | p        | n             | Positive (%) | p        |
| Tumour size    |               | 0.009    | 0.238         | 0.299    | 0.075         |
| <8 cm          | 18            | 18 (100.0) |          | 18            | 7 (38.9)     |          | 18            | 5 (27.8)     |          |
|                |               |          | 5 (27.8)     |          | 5 (27.8)     |          |
| ≥8 cm          | 58            | 42 (72.4) |          | 57            | 14 (24.6)    |          | 58            | 24 (41.4)    |          |
|                |               | 30 (51.7) |          | 30 (46.7)     |            |          |
| Tumour weight  | 0.347         | 0.937    | 0.592         | 1.000    |               |
| <467.2 mg      | 30            | 25 (83.3) |          | 29            | 8 (27.6)     |          | 30            | 10 (33.3)    |          |
|                |               | 14 (46.7) |          | 14 (46.7)     |            |          |
| ≥467.2 mg      | 30            | 22 (73.3) |          | 30            | 8 (26.7)     |          | 30            | 12 (40.0)    |          |
|                |               | 14 (46.7) |          | 14 (46.7)     |            |          |
| Weiss score    | 0.895         | 0.752    | 0.958         | 0.969    |               |
| <6             | 33            | 26 (78.8) |          | 33            | 8 (24.2)     |          | 33            | 13 (39.4)    |          |
|                |               | 15 (45.5) |          | 15 (45.5)     |            |          |
| ≥6             | 40            | 31 (77.5) |          | 40            | 11 (27.5)    |          | 40            | 16 (40.0)    |          |
|                |               | 18 (45.0) |          | 18 (45.0)     |            |          |
| Nuclear grade  | 1.000         | 0.645    | 0.382         | 0.048    |               |
| Low            | 5             | 5 (100.0) |          | 5             | 1 (20.0)     |          | 5             | 1 (20.0)     |          |
|                |               | 0 (0.0)  |          | 0 (0.0)       |            |          |
| High           | 36            | 35 (97.2) |          | 36            | 13 (36.1)    |          | 36            | 16 (44.4)    |          |
|                |               | 20 (55.6) |          | 20 (55.6)     |            |          |
| Mitotic index  | 1.000         | 0.591    | 0.011         | 0.279    |               |
| Low            | 24            | 23 (95.8) |          | 24            | 9 (37.5)     |          | 24            | 6 (25.0)     |          |
|                |               | 10 (41.7) |          | 10 (41.7)     |            |          |
| High           | 17            | 17 (100.0)|          | 17            | 5 (29.4)     |          | 17            | 11 (64.7)    |          |
|                |               | 10 (58.8) |          | 10 (58.8)     |            |          |
| Atypical mitosis| 1.000       | 0.734    | 0.075         | 0.265    |               |
| Absent         | 28            | 27 (96.4) |          | 28            | 9 (32.1)     |          | 28            | 9 (32.1)     |          |
|                |               | 12 (42.9) |          | 12 (42.9)     |            |          |
| Present        | 13            | 13 (100.0)|          | 13            | 5 (38.5)     |          | 13            | 8 (61.5)     |          |
|                |               | 8 (61.5)  |          | 8 (61.5)      |            |          |
| Tissue p53 status| 0.272       | 1.000    | 0.122         | 0.317    |               |
| Normal         | 19            | 12 (63.2) |          | 19            | 3 (15.8)     |          | 19            | 6 (31.6)     |          |
|                |               | 8 (42.1)  |          | 8 (42.1)      |            |          |
| Mutated        | 5             | 5 (100.0) |          | 5             | 1 (20.0)     |          | 5             | 4 (80.0)     |          |
|                |               | 4 (80.0)  |          | 4 (80.0)      |            |          |
| Necrosis       | 0.268         | 0.140    | 0.736         | 0.335    |               |

(Continued)
similar to the expression frequency found in the present study [21]. Herein, we show associations between GLUT1 and high mitotic index, presence of metastasis, and both longer overall and disease-free survival. This evidence is in agreement with the crucial role of GLUT1 in the metabolic reprogramming, as cancer cells are only able to depend on glycolytic pathway for energy production if the glycolytic flow is highly increased. This demands an increased uptake of glucose, mainly provided by GLUT1 in cancer cells [29].

MCT1 expression at the plasma membrane suggests a high lactate efflux from cancer cells, which, besides allowing glycolytic flux for energy production, also detains an important role in the microenvironment, by decreasing the immune response against tumor cells [30], increasing tumor cell motility [31], inducing angiogenesis [32], as well as stimulating hyaluronan and its receptor CD44, molecules involved in the process of cancer invasion and metastasis [33]. These roles of lactate in the extracellular milieu are in accordance with the association of MCT1 expression with high stage and shorter overall survival. In opposition, plasma membrane expression of MCT4, which was increased from adenomas to carcinomas, showed no association with the clinicopathological parameters. However, cytoplasmic expression of MCT4 and CD44, as well as CD147, was associated with absence of metastasis. In accordance, cytoplasmic expression of MCT4 and CD147 was also associated with longer disease-free survival, while cytoplasmic expression of CD147 also showed a tendency to be associated with longer overall survival. Further studies are warranted to elucidate the role of MCT4 in the cytoplasm. Importantly, MCT2 plasma membrane expression was associated with good
prognostic variables, including small tumor size, low mitotic index, absence of sinus invasion, absence of metastasis and longer overall survival. MCT2 expression at the plasma membrane suggests monocarboxylate consumption (likely lactate) from the extracellular environment. Since MCT2 plasma membrane expression is independent from MCT1 or MCT4 (data not shown), MCT2 positive tumors may present a metabolic symbiosis between cancer cells, in which the peripheral and oxygenated oxidative cells consume, through MCT2, the lactate released, through MCT4, by the central and less oxygenated glycolytic cells [34]. As a result, lactate will no longer act in the microenvironment to stimulate tumor aggressiveness. This rationale is in line with the evidence that MCT2 expressing tumors show a less aggressive phenotype than MCT2 negative tumors. Finally, we found CAIX to be more frequently expressed at the plasma membrane of tumors showing high nuclear grade and presence of metastasis, which is in accordance with the contribution of CAIX to the acid-resistant phenotype, rendering cancer cells a survival advantage that will contribute to cancer progression. Also, since CAIX contributes to the extracellular acidification, this pH regulator has a role in the acid-mediated cancer cell invasive behavior [35].

In line with the role of the different proteins in the metabolic remodeling of cancer cells and the evidence from the associations with the clinicopathological data, plasma membrane expression of MCT1 or GLUT1 was identified as a poor prognostic factor and plasma membrane expression of MCT2 or GLUT1 was identified as a poor prognostic factor.

Figure 3: Overall survival curves of adrenocortical carcinomas' patients. The results are stratified according to protein immunohistochemical expression. Only significant (or borderline) results are shown. Continuous line refers to positive expression while interrupted line refers to negative expression. A. Plasma membrane expression of MCT1; B. Plasma membrane expression of MCT2; C. Plasma membrane expression of GLUT1; D. Plasma membrane expression of CAIX.
membrane expression of MCT2 was identified as a good prognostic factor for overall survival in univariate analysis. Multivariate analysis showed that none of these markers has a stage-independent prognostic value, differently from results previously described for GLUT1 [21]. Importantly, CD147 cytoplasmic expression was identified as a stage-independent good prognostic factor for overall survival. In disease-free survival, while plasma membrane of GLUT1 was identified as a poor prognostic factor and plasma membrane expression of MCT2 and cytoplasmic expression of CD147 were identified as good prognostic value in univariate analysis, multivariate analysis showed that none of these markers has a stage-independent prognostic value. Importantly, CAIX cytoplasmic expression, while showing borderline significance in the univariate analysis, was identified as a stage-independent good prognostic factor for disease-free survival.

In the present study, we provided evidence for a metabolic reprogramming of adrenocortical malignant tumors towards the hyperglycolytic and acid-resistant phenotype characteristic of the Warburg effect, which associates with a worse prognosis. Since only few studies are available on the metabolic reprogramming of adult adrenocortical carcinoma, we believe that the results from the present study may bring new and relevant knowledge about the biology of this rare type of cancer, with possible developments in adrenocortical carcinoma management and the search for new target-directed therapeutic resources.

Figure 4: Disease-free survival curves of adrenocortical carcinomas’ patients. The results are stratified according to protein immunohistochemical expression. Only significant results are shown. Continuous line refers to positive expression while interrupted line refers to negative expression. A. Cytoplasmic expression of MCT4; B. Cytoplasmic expression of CD147; C. Plasma membrane expression of GLUT1; D. Cytoplasmic expression of CAIX.
Table 4: Prognostic factors for overall survival and disease-free survival in adult adrenocortical carcinomas

|                           | Univariate analysis | Multivariate analysis |
|---------------------------|---------------------|-----------------------|
|                           | Overall survival    |                       |
|                           | HR                  | 95% CI                | p         | HR                  | 95% CI                | p         |
| Male sex                  | 0.73                | 0.28–1.87             | 0.51      | 3.6                 | 0.7–17.5              | 0.13      |
| Age (yrs)                 | 0.99                | 0.98–1.02             | 0.76      |                     |                       |           |
| Tumor size ≥ 8 cm         | 3.86                | 1.37–10.9             | 0.01      | 2.3                 | 1.5–8.8               | 0.005     |
| Weiss score ≥ 6           | 0.86                | 0.44–1.67             | 0.64      |                     |                       |           |
| Tumor stage (ENSAT 3)     | 5.1                 | 2.4–10.8              | 0.0001    | 2.3                 | 0.91–5.7              | 0.077     |
| MCT1 cytoplasm            | 1.05                | 0.47–2.3              | 0.91      |                     |                       |           |
| MCT1 plasma membrane      | 2.27                | 1.04–4.91             | 0.038     | 2.3                 | 0.9–9.7               | 0.077     |
| MCT2 cytoplasm            | 0.83                | 0.41–1.67             | 0.6       |                     |                       |           |
| MCT2 plasma membrane      | 0.409               | 0.20–0.82             | 0.01      | 0.78                | 0.34–0.74             | 0.548     |
| MCT4 cytoplasm            | 0.53                | 0.24–1.14             | 0.1       |                     |                       |           |
| MCT4 plasma membrane      | 1.59                | 0.71–3.55             | 0.26      |                     |                       |           |
| CD147 cytoplasm           | 0.49                | 0.23–1.04             | 0.063     | 0.32                | 0.14–0.74             | 0.008     |
| CD147 plasma membrane     | 0.94                | 0.44–2.05             | 0.89      |                     |                       |           |
| CD44 cytoplasm            | 0.63                | 0.2–1.8               | 0.39      |                     |                       |           |
| CD44 plasma membrane      | 0.9                 | 0.4–2.0               | 0.79      |                     |                       |           |
| GLUT1 cytoplasm           | 1.3                 | 0.6–2.81              | 0.5       |                     |                       |           |
| GLUT1 plasma membrane     | 2.63                | 1.3–5.3               | 0.007     | 1.97                | 0.9–4.2               | 0.088     |
| CAIX cytoplasm            | 0.545               | 0.27–1.1              | 0.092     | 0.72                | 0.32–1.62             | 0.43      |
| CAIX plasma membrane      | 1.98                | 0.98–3.99             | 0.056     | 0.96                | 0.42–2.18             | 0.93      |
|                           |                     |                       |           |                     |                       |           |
| Disease-free survival     |                     |                       |           |                     |                       |           |
|                           | HR                  | 95% CI                | p         | HR                  | 95% CI                | p         |
| Male sex                  | 1.1                 | 0.45–2.8              | 0.81      |                     |                       |           |
| Age (yrs)                 | 1.06                | 0.98–1.03             | 0.63      |                     |                       |           |
| Tumor size ≥ 8 cm         | 3.49                | 1.2–10.4              | 0.022     | 1.76                | 0.45–6.8              | 0.41      |
| Weiss score ≥ 6           | 0.695               | 0.33–1.46             | 0.33      |                     |                       |           |
| Tumor stage (ENSAT 3)     | 4.46                | 2.1–9.5               | 0.0001    | 3.96                | 1.4–10.9              | 0.008     |
| MCT1 cytoplasm            | 1.44                | 0.58–3.53             | 0.43      |                     |                       |           |
| MCT1 plasma membrane      | 2.1                 | 0.84–5.1              | 0.11      |                     |                       |           |
| MCT2 cytoplasm            | 1.32                | 0.56–3.1              | 0.52      |                     |                       |           |
| MCT2 plasma membrane      | 0.42                | 0.18–0.97             | 0.04      | 0.74                | 0.28–1.9              | 0.527     |
| MCT4 cytoplasm            | 0.42                | 0.16–1.08             | 0.071     | 1.12                | 0.36–3.5              | 0.843     |
| MCT4 plasma membrane      | 1.22                | 0.497–2.99            | 0.66      |                     |                       |           |
| CD147 cytoplasm           | 0.36                | 0.14–0.92             | 0.033     | 0.43                | 0.16–1.18             | 0.1       |

(Continued)
Univariate analysis | Multivariate analysis
--- | ---
| Overall survival | Overall survival |
| | HR | 95% CI | p | HR | 95% CI | p |
| CD147 plasma membrane | 1.02 | 0.37–2.76 | 0.97 | | | |
| CD44 cytoplasm | 0.23 | 0.03–1.85 | 0.17 | | | |
| CD44 plasma membrane | 0.99 | 0.35–2.77 | 0.98 | | | |
| GLUT1 cytoplasm | 0.55 | 0.18–1.61 | 0.27 | | | |
| GLUT1 plasma membrane | 3.84 | 1.52–9.68 | 0.004 | 2.4 | 0.86–6.74 | 0.097 |
| CAIX cytoplasm | 0.47 | 0.2–1.1 | 0.079 | 0.31 | 0.1–0.92 | 0.036 |
| CAIX plasma membrane | 1.95 | 0.84–4.53 | 0.12 | | | |

Figure 5: Schematic representation summarizing the major findings of the present work. The schematic representation distinguishes the major findings for plasma membrane and cytoplasmic expression of the different proteins studied. Red arrow: significant increase in expression frequency when comparing adenomas to carcinomas. * Independent predictors of survival as determined by multivariate analysis.
MATERIALS AND METHODS

Human adrenocortical tumor samples

The series analyzed included 154 formalin-fixed paraffin-embedded adrenocortical neoplasias (76 adenomas and 78 carcinomas, according to Weiss score), retrieved from the files of the Pathology Department of the Clinical Hospital, School of Medicine, University of Sao Paulo, Brazil. No case included in this series fulfills the criteria for oncocytic tumor. Samples were organized into tissue microarrays (TMA) containing 208 cores each (1.0 mm diameter each core). Each case was represented in TMAs by three cores and control samples (kidney) were also included for TMA orientation. Clinico pathological data for the adrenocortical carcinomas included age at diagnosis (normal distribution, mean 40.6 years, range: 15 to 81 years), gender, tumor size (normal distribution, mean 11.8 cm, range: 2.2 to 23.0 cm; categorized using 8 cm as cut-off [36]) and weight (non-normal distribution, median 467.2 mg, range: 10 to 2600 mg), Weiss score and each of its individual histological parameters [37], tumor p53 status, staging (according to ENSAT system [38]), metastasis, disease-free survival and overall survival (median 13.5 and 30.5 months, respectively). Cases that could not be evaluated by Weiss score were classified as carcinomas if metastatic disease was detected (clinically malignant). Detailed information of the clinico pathological data for the adrenocortical adenomas as well as the adrenocortical carcinomas is presented in Tables 5 and 6, respectively. The present study was approved by the Local Ethic Committee (number 11090).

Immunohistochemistry

MCT1 immunohistochemistry was performed according to the avidin-biotin-peroxidase complex method (R.T.U. VECTASTAIN Elite ABC Kit (Universal), Vector Laboratories, Burlingame, CA), as previously described [39]. Immunohistochemistry for MCT2, MCT4, GLUT1, CD44 and CAIX was performed according to the streptavidin-biotin-peroxidase complex principle (Ultra vision Detection System Anti-polyvalent, HRP, Lab Vision Corporation, Fremont, CA), as previously described [20, 40, 41]. CD147 immunostaining was performed using a polymer system (UltraVision ONE Detection System: HRP Polymer Lab Vision Corporation, Fremont, CA) as already described [42]. Negative controls were performed by the use of appropriate serum controls for the primary antibodies (N1698 and N1699, Dako, Carpinteria, CA). Colon carcinoma tissue was used as positive control for MCT1, MCT4, CD147 and CD44, head and neck squamous cell carcinoma was used for GLUT1, and normal stomach was used for CAIX. Tissue sections were counterstained with hematoxylin and permanently mounted. Please refer to Table 7 for detailed aspects for each antibody used.

Figure 6: Immunohistochemical staining patterns of metabolism-related proteins in a case representative of the glycolytic phenotype. In this case, MCT1 A, MCT4 C, CD147 D, GLUT1 E, and CAIX F, were found in the plasma membrane while MCT2 B, was observed in the cytoplasm.
**Immunohistochemical evaluation**

Sections were scored semi-quantitatively for expression in cancer cells as follows: 0: no immunoreactive cells; 1: < 5% of immunoreactive cells; 2: 5–50% of immunoreactive cells; and 3: > 50% of immunoreactive cells. Also, intensity of staining was scored semi-qualitatively as follows: 0: negative; 1: weak; 2: intermediate; and 3: strong. The final score was defined as the sum of both parameters (extension and intensity), and, for the metabolism-related proteins, grouped as negative (score 0 and 2) and positive (score 3–6), as previously described [39]. Protein expression in the different cellular localizations (cytoplasm and plasma membrane) was evaluated separately.

**Statistical analysis**

Data were stored and analyzed using the IBM SPSS Statistics software (version 20, IBM Company, Armonk, NY). All comparisons were examined for statistical significance using Pearson’s chi-square ($\chi^2$) test and Fisher’s exact test (when $n < 5$). Overall survival was defined as the time from the date of primary diagnosis to death related to adrenocortical cancer or last follow-up. Disease-free survival was defined as the time from the date of complete tumor resection to the first radiological evidence of disease relapse or death. Overall and disease-free survival curves were estimated by the method of Kaplan-Meier and data compared using the log-rank test. Predictive factors of prognosis were identified by means of Cox proportional hazards regression models, which were used to estimate hazard ratios (HR) and their 95% confidence intervals in univariate and multivariate analysis. All variables that reached a $p$ value < 0.1 at Kaplan-Meier estimates were included in the Cox survival analysis. For multivariate analysis, variables that reached a $p$ value < 0.1 at univariate analysis were included. The threshold for significant $p$ values was established as $p < 0.05$. 

Figure 7: Immunohistochemical staining patterns of metabolism-related proteins in a case representative of the oxidative phenotype. In this case, MCT2 A. was found in the plasma membrane, while MCT4 B. CD147 C. and CAIX D. were found in the cytoplasm.
Table 5: Clinicopathological data of the adrenocortical adenoma patients

| Variable                          | n  | %   |
|----------------------------------|----|-----|
| **Age** ($n = 76$)               |    |     |
| ≥ 44.2 years                     | 35 | 46.1|
| < 44.2 years                     | 41 | 53.9|
| **Gender** ($n = 76$)            |    |     |
| Female                           | 66 | 86.8|
| Male                             | 10 | 13.2|
| **Tumor size** ($n = 73$)        |    |     |
| < 8 cm                           | 71 | 97.3|
| ≥ 8 cm                           | 2  | 2.7 |
| **Tumor weight** ($n = 64$)      |    |     |
| < 20.0 mg                        | 29 | 45.3|
| ≥ 20.0 mg                        | 35 | 54.7|
| **Weiss score** ($n = 76$)       |    |     |
| 0                                | 35 | 46.1|
| 1                                | 22 | 28.9|
| 2                                | 19 | 25.0|
| **Nuclear grade** ($n = 42$)     |    |     |
| Low                              | 37 | 88.1|
| High                             | 5  | 11.9|
| **Mitotic index** ($n = 42$)     |    |     |
| Low                              | 41 | 97.6|
| High                             | 1  | 2.4 |
| **Atypical mitosis** ($n = 42$)  |    |     |
| Absent                           | 42 | 100.0|
| Present                          | 0  | 0.0 |
| **Necrosis** ($n = 42$)          |    |     |
| Absent                           | 38 | 90.5|
| Present                          | 4  | 9.5 |
| **Venous invasion** ($n = 42$)   |    |     |
| Absent                           | 42 | 100.0|
| Present                          | 0  | 0.0 |
| **Sinus invasion** ($n = 42$)    |    |     |
| Absent                           | 41 | 97.6|
| Present                          | 1  | 2.4 |
| **Capsular invasion** ($n = 41$) |    |     |
| Absent                           | 38 | 92.7|
| Present                          | 3  | 7.3 |

(Continued)
Variable  |  n  |  %  
--- | --- | ---  
**Staging (n = 56)**  
I  | 49  | 87.5  
II  | 7  | 12.5  

*The mean value was used for age cut-off, while the median value was used for weight cut-off as these variables followed a normal and non-normal distribution, respectively.  
*Nuclear grade and mitotic index were defined according to Weiss score definitions (30).  

**Table 6: Clinicopathological data of the adrenocortical carcinoma patients**  

| Variable  | n  | %  
--- | --- | ---  
**Age**  
≥ 40.6 years  | 34  | 43.6  
< 40.6 years  | 44  | 56.4  
**Gender (n = 78)**  
Female  | 61  | 78.2  
Male  | 17  | 21.8  
**Tumor size (n = 76)**  
< 8 cm  | 18  | 23.7  
≥ 8 cm  | 58  | 76.3  
**Tumor weight**  
< 467.2 mg  | 30  | 50.0  
≥ 467.2 mg  | 30  | 50.0  
**Weiss score (n = 78)**  
≥ 3  | 5  | 6.4  
3  | 13  | 16.7  
4  | 13  | 16.7  
5  | 7  | 9.0  
6  | 11  | 14.1  
7  | 11  | 14.1  
8  | 14  | 17.9  
9  | 4  | 5.1  
**Nuclear grade**  
Low  | 5  | 12.2  
High  | 36  | 87.8  
**Mitotic index**  
Low  | 24  | 58.5  
High  | 17  | 41.5  
**Atypical mitosis (n = 41)**  
Absent  | 28  | 68.3  
Present  | 13  | 31.7  

(Continued)
| Variable                      | $n$ | %  |
|-------------------------------|-----|-----|
| **Necrosis ($n = 41$)**       |     |     |
| Absent                        | 11  | 26.8|
| Present                       | 30  | 73.2|
| **Venous invasion ($n = 41$)**|     |     |
| Absent                        | 27  | 65.9|
| Present                       | 14  | 34.1|
| **Sinus invasion ($n = 41$)** |     |     |
| Absent                        | 35  | 85.4|
| Present                       | 6   | 14.6|
| **Capsular invasion ($n = 40$)**|     |     |
| Absent                        | 27  | 67.5|
| Present                       | 13  | 32.5|
| **Staging ($n = 76$)**        |     |     |
| I                             | 7   | 9.2 |
| II                            | 36  | 47.4|
| III                           | 15  | 19.7|
| IV                            | 18  | 23.7|
| **Metastasis ($n = 78$)**     |     |     |
| Absent                        | 37  | 47.4|
| Present                       | 41  | 52.6|
| **Evolution ($n = 78$)**      |     |     |
| Alive                         | 46  | 59.0|
| Dead                          | 32  | 41.0|

*The mean value was used for age cut-off, while the median value was used for weight cut-off as these variables followed a normal and non-normal distribution, respectively.

*Cases classified as carcinoma (Weiss ≥ 3) based on the presence of metastatic disease.

*Nuclear grade and mitotic index were defined according to Weiss score definitions (30).

Table 7: Detailed aspects for each antibody used in immunohistochemistry

| Protein | Antigen retrieval | Antibody | Antibody dilution and incubation time |
|---------|-------------------|----------|---------------------------------------|
| MCT1    | Citrate buffer (0.01 M, pH = 6), 98°C, 20’ | AB3538P Chemicon International | 1:200, overnight |
| MCT2    | Citrate buffer (0.01 M, pH = 6), 98°C, 20’ | sc-50322 Santa Cruz Biotechnology | 1:200, 2 hours |
| MCT4    | Citrate buffer (0.01 M, pH = 6), 98°C, 20’ | sc-50329 Santa Cruz Biotechnology | 1:500, 2 hours |
| CD147   | EDTA (1 mM, pH = 8), 98°C, 20’ | sc-71038 Santa Cruz Biotechnology | 1:400, overnight |

(Continued)
This work was supported by FAPESP to Maria Claudia Nogueira Zerbini (2013-26344-8). SG received the post-doctoral fellowship UMINHO/BPD/18/2014.

The authors declare no conflicts of interest.

1. Fassnacht M, Libe R, Kroiss M, Alloio B. Adrenocortical carcinoma: a clinician’s update. Nat Rev Endocrinol. 2011; 7:323–335.
2. Fassnacht M, Kroiss M, Alloio B. Update in adrenocortical carcinoma. J Clin Endocrinol Metab. 2013; 98:4551–4564.
3. Else T, Kim AC, Sabolch A, Raymond VM, Kandathil A, Caoili EM, Jolly S, Miller BS, Giordano TJ, Hammer GD. Adrenocortical carcinoma. Endocr Rev. 2014; 35:282–326.
4. Fassnacht M, Terzolo M, Alloio B, Baudin E, Haak H, Berutti A, Welin S, Schade-Brittinger C, Lacroix A, Jarzab B, Sorbye H, Torpy DJ, Stepan V, Kroiss M, et al. Combination chemotherapy in advanced adrenocortical carcinoma. N Engl J Med. 2012; 366:2189–2197.
5. Berutti A, Baudin E, Gelderblom H, Haak HR, Porpiglia F, Fassnacht M, Pentheroudakis G, Group EGW. Adrenal cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. Ann Oncol. 2012; 23: vii131–138.
6. Faria AM, Almeida MQ. Differences in the molecular mechanisms of adrenocortical tumorigenesis between children and adults. Mol Cell Endocrinol. 2012; 351:52–57.
7. Lucon AM, Pereira MA, Mendonca BB, Zerbini MC, Saldanha LB, Arap S. Adrenocortical tumors: results of treatment and study of Weiss’s score as a prognostic factor. Rev Hosp Clin Fac Med Sao Paulo. 2002; 57:251–256.
8. Duregon E, Volante M, Giorcelli J, Terzolo M, Lalli E, Papotti M. Diagnostic and prognostic role of steroidogenic factor 1 in adrenocortical carcinoma: a validation study focusing on clinical and pathologic correlates. Hum Pathol. 2013; 44:822–828.
9. Gatenby RA, Gillies RJ. Why do cancers have high aerobic glycolysis? Nat Rev Cancer. 2004; 4:891–899.
10. Tennant DA, Duran RV, Gottlieb E. Targeting metabolic transformation for cancer therapy. Nat Rev Cancer. 2010; 10:267–277.
11. Chiche J, Brahim-Horn MC, Pouyssegur J. Tumour hypoxia induces a metabolic shift causing acidosis: a common feature in cancer. J Cell Mol Med. 2010; 14:771–794.
12. Pinheiro C, Longatto-Filho A, Azvedo-Silva J, Casal M, Schmitt FC, Baltazar F. Role of monocarboxylate transporters in human cancers: state of the art. J Bioenerg Biocembr. 2012; 44:127–139.
13. de Oliveira AT, Pinheiro C, Longatto-Filho A, Brito MJ, Martinho O, Matos D, Carvalho AL, Vazquez VL, Silva TB, Scapulatempo C, Saad SS, Reis RM, Baltazar F. Co-expression of monocarboxylate transporter 1 (MCT1) and its chaperone (CD147) is associated with low survival in patients with gastrointestinal stromal tumors (GISTs). J Bioenerg Biocembr. 2012; 44:171–178.
14. Miranda-Goncalves V, Honavar M, Pinheiro C, Martinho O, Pires MM, Pinheiro C, Cordeiro M, Bebiano G, Costa P, Palmeirim I, Reis RM, Baltazar F. Monocarboxylate transporters (MCTs) in gliomas: expression and exploitation as therapeutic targets. Neuro Oncol. 2013; 15:172–188.
15. Pinheiro C, Penna V, Morais-Santos F, Abrahao-Machado LF, Ribeiro G, Curelli EC, Olivieri MV, Morini S, Valencas I, Ribeiro D, Schmitt FC, Reis RM, Baltazar F. Characterization of monocarboxylate transporters (MCTs) expression in soft tissue sarcomas: distinct prognostic impact of MCT1 sub-cellular localization. J Transl Med. 2014; 12:118.
16. Halestrap AP. The monocarboxylate transporter family—Structure and functional characterization. IUBMB Life. 2012; 64:1–9.
17. Kirk P, Wilson MC, Heddle C, Brown MH, Barclay AN, Halestrap AP. CD147 is tightly associated with lactate transporters MCT1 and MCT4 and facilitates their cell surface expression. EMBO J. 2000; 19:3896–3904.
18. Wilson MC, Meredith D, Fox JE, Manoharan C, Davies AJ, Halestrap AP. Basigin (CD147) is the target for organomercurial inhibition of monocarboxylate transporter isoforms 1 and 4: the ancillary protein for the insensitive MCT2 is EMBIGIN (gp70). J Biol Chem. 2005; 280:27213–27221.

19. Slomiany MG, Grass GD, Robertson AD, Yang XY, Maria BL, Beeson C, Toole BP. Hyaluronan, CD44, and emmprin regulate lactate efflux and membrane localization of monocarboxylate transporters in human breast carcinoma cells. Cancer Res. 2009; 69:1293–1301.

20. Pinheiro C, Reis RM, Ricardo S, Longatto-Filho A, Schmitt F, Baltazar F. Expression of monocarboxylate transporters 1, 2, and 4 in human tumours and their association with CD147 and CD44. J Biomed Biotechnol. 2010; 2010:427694.

21. Fenske W, Volker HU, Adam P, Hähner S, Johanssen S, Wortmann S, Schmidt M, Marcos M, Müller-Hermelink HK, Allolio B, Fassnacht M. Glucose transporter GLUT1 expression is a clinical-independent predictor of clinical outcome in adrenocortical carcinoma. Endocr Relat Cancer. 2009; 16:919–928.

22. Nunes ML, Rault A, Teynie J, Valli N, Goyut M, Gaye D, Belleeannee G, Tabarin A. 18F-FDG PET for the identification of adrenocortical carcinomas among indeterminate adrenal tumors at computed tomography scanning. World J Surg. 2010; 34:1506–1510.

23. Deandreis D, Leboulleux S, Caramella C, Schlumberger M, Baudin E. FDG PET in the management of patients with adrenal masses and adrenocortical carcinoma. Horm Cancer. 2011; 2:354–362.

24. Takeuchi S, Balachandran A, Habra MA, Phan AT, Bassett RL, Jr., Macapinlac HA, Chuang HH. Impact of (1)8 F-FDG PET/CT on the management of adrenocortical carcinoma: analysis of 106 patients. Eur J Nucl Med Mol Imaging. 2014; 41:2066–2073.

25. Bourdeau I, MacKenzie-Feder J, Lacroix A. Recent advances in adrenocortical carcinoma in adults. Curr Opin Endocrinol Diabetes Obes. 2013; 20:192–197.

26. Ribeiro RC, Sandrini F, Figueiredo B, Zambetti GP, Michalkiewicz E, Lafferty AR, DeLacerda L, Rabin M, Cadwell C, Sampaio G, Cat I, Stratakis CA, Sandrini F, Figueiredo B, Zambetti GP, Beeson C, Toole BP. Hyaluronan, CD44, and emmprin regulate lactate efflux and membrane localization of monocarboxylate transporters in human breast carcinoma cells. Cancer Res. 2009; 69:1293–1301.

27. Fenske W, Volker HU, Adam P, Hähner S, Johanssen S, Wortmann S, Schmidt M, Marcos M, Müller-Hermelink HK, Allolio B, Fassnacht M. Glucose transporter GLUT1 expression is an stage-independent predictor of clinical outcome in adrenocortical carcinoma. Endocr Relat Cancer. 2009; 16:919–928.

28. Park SK, Chiche J, Pouysssegur J. Disrupting proton dynamics and energy metabolism for cancer therapy. Nat Rev Cancer. 2013; 13:611–623.

29. Ganapathy V, Thangaraju M, Prasad PD. Nutrient transporters in cancer: relevance to Warburg hypothesis and beyond. Pharmacol Ther. 2009; 121:29–40.

30. Fischer K, Hoffmann P, Voelkl S, Meidenbauer N, Ammer J, Edinger M, Gottfried E, Schwarz S, Rothe G, Hoves S, Renner K, Timischl B, Mackensen A, Kunz-Schughart L, Andreessen R, Krause SW, et al. Inhibitory effect of tumor cell-derived lactic acid on human T cells. Blood. 2007; 109:3812–3819.

31. Walenta S, Schroeder T, Mueller-Klieser W. Metabolic mapping with bioluminescence: basic and clinical relevance. Biomol Eng. 2002; 18:249–262.

32. Hunt TK, Aslam RS, Beckert S, Wagner S, Ghani QP, Hussain MZ, Roy S, Sen CK. Aerobically derived lactate stimulates revascularization and tissue repair via redox mechanisms. Antioxid Redox Signal. 2007; 9:1115–1124.

33. Stern R, Huster S, Neudecker BA, Formby B. Lactate stimulates fibroblast expression of hyaluronan and CD44: the Warburg effect revisited. Exp Cell Res. 2002; 276:24–31.

34. Sonveaux P, Veyrag F, Schroeder T, Wergin MC, Verraz B, Rabbani ZN, De Saedeleer CJ, Kennedy KM, Diepart C, Jordan BF, Kelley MJ, Gallez B, Wahl ML, Feron O, Dewhirst MW. Targeting lactate-fueled respiration selectively kills hypoxic tumor cells in mice. J Clin Invest. 2008; 118:3930–3942.

35. Smallbone K, Gavaghan DJ, Gateby RA, Maini PK. The role of acidity in solid tumour growth and invasion. J Theor Biol. 2005; 235:476–484.

36. Faria AM, Sbiera S, Ribeiro TC, Soares IC, Mariani BM, Freire DS, de Sousa GR, Lerario AM, Ronchi CL, Deutschbein T, Wakamatsu A, Alves VA, Zerbini MC, Mendonca BB, Fragoso MC, Latronico AC, et al. Expression of LIN28 and its regulatory microRNAs in adult adrenocortical carcinomas: proposal for a Revised TNM Classification. Clin Endocrinol (Oxf). 2015; 82:481–488.

37. Weiss LM. Comparative histologic study of 43 metastasizing and nonmetastasizing adrenocortical tumors. Am J Surg Pathol. 1984; 8:163–169.

38. Fassnacht M, Johanssen S, Quinkler M, Buesky P, Willenberg HS, Beuschlein F, Terzolo M, Mueller HH, Hahner S, Allolio B, German Adrenocortical Carcinoma Registry G, European Network for the Study of Adrenal T. Limited prognostic value of the 2004 International Union Against Cancer staging classification for adrenocortical carcinoma: proposal for a Revised TNM Classification. Cancer. 2009; 115:243–250.

39. Pinheiro C, Longatto-Filho A, Scapulatempo C, Ferreira L, Ronchi CL, Ferreira L, Ronchi CL, Pinheiro C, Sousa B, Albergaria A, Paredes J, Dufloth R, Mota J, Malinich E, Ferschl M, Hahner S, Johanssen S, Pratschke J, Eberhardt W, et al. Inhibitory effect of tumor cell-derived lactic acid on human T cells. Blood. 2007; 109:3812–3819.

40. Pinheiro C, Sousa B, Albergaria A, Paredes J, Dufloth R, Vieira D, Schmitt F, Baltazar F. GLUT1 and CAIX expression profiles in breast cancer correlate with adverse prognostic factors and MCT1 overexpression. Histol Histopathol. 2011; 26:1279–1286.
41. Queiros O, Preto A, Pacheco A, Pinheiro C, Azevedo-Silva J, Moreira R, Pedro M, Ko YH, Pedersen PL, Baltazar F, Casal M. Butyrate activates the monocarboxylate transporter MCT4 expression in breast cancer cells and enhances the antitumor activity of 3-bromopyruvate. J Bioenerg Biomembr. 2012; 44:141–153.

42. Granja S, Morais-Santos F, Miranda-Goncalves V, Viana-Ferreira M, Nogueira R, Nogueira-Silva C, Correia-Pinto J, Baltazar F. The monocarboxylate transporter inhibitor alpha-cyano-4-hydroxycinnamic acid disrupts rat lung branching. Cell Physiol Biochem. 2013; 32:1845–1856.