Trophic Ecology of the Cuban Boa, *Chilabothrus angulifer* (Boidae)

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**Abstract.**—The Cuban Boa (*Chilabothrus angulifer*) is a top terrestrial predator in Cuba. References to prey species consumed by this boa date to when the first Europeans arrived in the region more than 500 years ago. However, long-term studies on its trophic ecology do not exist. The scarce and scattered records on its feeding habits indicate that this boa preys on a variety of native and domestic animals. Based on dietary information collected in the field and from the literature, we characterized the diet of this snake and tested four different hypotheses: (1) The Cuban Boa is a generalist predator; (2) the diets of boas in natural and anthropogenic habitats differ; (3) an ontogenetic shift in diet occurs; and (4) foraging strategies used in natural and anthropogenic habitats differ. We identified 49 prey species from 351 prey items obtained from 218 snakes, including 71 items (31 snakes) from the literature. Mammals represented 55% of total prey items consumed, followed by birds (41%) and ectotherms (4%). *Chilabothrus angulifer* exhibited a narrow niche breadth. However, rather than a trophic specialist, we consider this boa an opportunistic generalist predator, capable of adjusting its diet and foraging behavior according to prey availability and abundance. The diet of *Ch. angulifer* changed dramatically from mostly native mammals and birds in natural habitats to mostly livestock, pets, and human commensals in human-altered habitats. Also, mammals were consumed more frequently in natural habitats, whereas birds dominated the diet of boas associated with anthropogenic habitats. Few ectotherms were consumed in either type of habitat. We observed an ontogenetic shift in diet, but this primarily reflected a trend of consuming larger prey rather than a shift from ectotherms to endotherms as reported for some other boids. In natural habitats, *Ch. angulifer* used both ambush and active-foraging modes by day and night, whereas in anthropogenic situations, most boas used an active-foraging strategy at night. The frequent consumption of domestic animals by Cuban Boas might be the principal reason for the historical human-wildlife conflict involving this species in rural areas of Cuba.

Human-wildlife conflicts pose one of the greatest challenges in biodiversity conservation (Conover 2001; Decker et al. 2002; Anthony et al. 2010; Manral et al. 2016). Large constrictors (Boidae, Pythonidae) are among the top predators in many ecosystems. Their large size and muscular strength allow them to exploit a wide variety of prey, ranging from small ectotherms and endotherms to relatively large prey such as alligators, capybaras, monkeys, artiodactyls, and even hyenas, bears, and leopards (Murphy and Henderson 1997; Rivas 2000; Fredriksson 2005; Reed and Rodda 2009; Monroy-Vilchis et al. 2011; Greshko 2017). With the continuous development of human settlements, many large snakes have been forced to coexist with humans in a different structural habitat with a different associated fauna (Shine and Fitzgerald 1996; Fearn et al. 2001; Luisselli et al. 2001; Pearson et al. 2005; Tolson and Henderson 2006). The occurrence of these large predators in urban areas increases the probability of predation on domestic animals that are important to human wellbeing (Fearn et al. 2001; Tolson and Henderson 2006; Reed and Rodda 2009; Rodríguez-Cabrera et al. 2016a). Such interactions often are responsible for human-wildlife conflicts that are pushing many large constrictors to the edge of extinction (IUCN 2020).

With a snout-vent length (SVL) that can exceed 5.0 m and a mass of more than 30 kg (Gundlach 1880; Tolson and Henderson 1993; Petersen et al. 2015; Rodríguez-Cabrera et al. 2016a; T.M. Rodríguez-Cabrera, unpubl. data), the Cuban Boa (*Chilabothrus angulifer*) (Fig. 1) is the largest snake in the West Indies. This species is endemic to the Cuban Archipelago, where it is widely distributed on the main island, Isla de la Juventud, and some adjacent cays, where it inhabits a variety of environments at elevations from
Chilabothrus angulifer may be locally common in a few remaining natural habitats (Berovides and Carbonell 1998; Linares et al. 2011; Rodríguez-Cabrera et al. 2015). Aggregations of nearly 30 boas have been repeatedly reported in natural sites with large concentrations of food resources, especially bat caves (Berovides and Carbonell 1998). However, when analyzing the relative abundance of Ch. angulifer on a wider geographical scale, the species is uncommon. Populations of Ch. angulifer in natural habitats other than bat caves (e.g., karstic woodlands, grasslands, wetlands) appear to be much smaller and dispersed (Tolson and Henderson 2006; P.J. Tolson, in litt. 2008; T.M. Rodríguez-Cabrera, pers. obs.). However, studies on the ecology of this boa are few and almost exclusively focused on cave-associated populations (Alfonso et al. 1998; Berovides and Carbonell 1998; Morell et al. 1998; Linares et al. 2011; Rodríguez-Cabrera et al. 2015; Dinets 2017). The exception is a still-unpublished long-term field research project led by Peter J. Tolson on the grounds of the U.S. Naval Base at Guantánamo Bay (Tolson and Henderson 2006; Petersen et al. 2007, 2015; P.J. Tolson in Henderson and Powell 2009; Meeks 2018).

The available data suggest that Ch. angulifer preys on a wide variety of animals that includes reptiles, native birds, bats, hutias, introduced murid rodents, and domestic animals (Appendix I). Herein, we test the hypothesis that Ch. angulifer is a generalist predator and predict that this boa has a wide dietary niche breadth.

Even if a given species is a trophic generalist or specialist, its diet may be constrained by the abundance and composition of available dietary resources. Specifically, species assemblages and their relative abundances differ considerably between natural and anthropogenic habitats (Estrada et al. 1997; Tews et al. 2004; Gamage et al. 2011). Therefore, the expectation that prey in natural habitats will differ from that in human-altered habitats is reasonable. We also test the hypothesis that the diet of Ch. angulifer in natural habitats differs from that of boas in anthropogenic habitats and predict that both prey composition and relative abundance in the diet of snakes occurring in natural habitats are different than those in the diet of snakes associated with human-altered habitats.

Another factor that can constrain the types of prey consumed is the size of the snake. Snakes are gape-limited predators because they swallow their prey whole; hence, gape size sets an upper limit to prey size (Shine 1991; Forsman 1996; Rodríguez-Robles et al. 1999; Vincent et al. 2004, 2005; Hampton 2014). Medium-sized to large adult boas and pythons may be more than an order of magnitude larger than neonates of the same species (Henderson et al. 1987; Tolson and Henderson 1993; Rivas 2000; Pizzatto and Marques 2007; Reed and Rodda 2009). Therefore, these snakes typically exhibit an ontogenetic shift in diet. An initial shift usually involves a transition from ecto- to endothermic prey and then a second more gradual shift involves a transition from smaller to larger endothermic prey (Henderson et al. 1987; Harlow and Shine 1992; Henderson 1993; Fearn et al. 2001; Pizzatto et al. 2009; Henderson and Pauers 2012). Due to the great difference in size between neonatal and adult Ch. angulifer, we test the hypothesis that an ontogenetic shift in diet occurs in this species and predict that prey type and size changes as snakes grow.

Snakes may use sit-and-wait and/or active foraging strategies in response to a number of biotic and abiotic factors (Huey and Pianka 1981; Mushinsky 1987). Habitat structure and the characteristics and accessibility of available prey can influence the strategy employed (Mushinsky 1987; Lind and Welsh 1994; Secor 1995; Mullin and Cooper 2000; Beaupre and Montgomery 2007; Emmons et al. 2016). We test the hypothesis that the foraging strategy used by Ch. angulifer differs in natural versus anthropogenic habitats and predict that the frequency of foraging modes will be different.

Fig. 1. The Cuban Boa (Chilabothrus angulifer) is the largest and stoutest snake in the West Indies. Photograph © Raimundo López-Silvero.
Materials and Methods

Data collection.—We collected data from four different sources: direct field observations over a span of more than 30 years (1987–2020), the literature, unpublished data provided by colleagues, and testimonies from trustworthy farmers and/or local land owners. To the best of our knowledge, we screened all of the available scientific literature related to the natural history and diet of *Ch. angulifer* since the first mention of the species in the 16th century. We chose specific descriptions of predation over accounts that were anecdotal, repetitive, or speculative. Compelling data were in about 20 different publications, most of which described isolated predation events (Appendix I). Most of the data originated from about 30 montane and lowland localities in central Cuba (Cienfuegos, Villa Clara, and Sancti Spíritus Provinces) (Fig. 2). However, we also included relevant information from more than 25 additional localities in other provinces across the country. Most localities are represented by single events (i.e., one boa), but multiple cases came from a few localities (e.g., Cariblanca, Sancti Spíritus Province, and San Blas village and vicinity in the Guanahaya Massif, Cienfuegos Province).

The data gleaned from the literature were in some cases insufficient for the purposes of this work. Therefore, we verified and enhanced them with additional information by establishing direct communication with the respective authors when possible (Appendix I). In the case of the White-crowned Pigeon (*Patagioenas leucocephala*), we were unable to obtain any additional quantitative data. Therefore, this species was included in the total number of species consumed by *Ch. angulifer*, but not in the statistical analyses.

Many snakes encountered in the field had a visible bulge in the stomach. To avoid killing the snake, we used forced regurgitation by palpation of the abdomen in order to obtain the dietary information (Luiselli and Amori 2016). We quantified and identified the prey items to the lowest taxonomic resolution possible, and assigned each to one of three categories (i.e., egg, juvenile or adult). Zoological nomenclature followed the most recent compilations for each group: amphibians (AmphibiaWeb 2020; Hedges et al. 2019), reptiles (Uetz et al. 2020; Hedges et al. 2019), birds (Navarro 2020), and mammals (Mammal Diversity Database 2020). Many feeding events witnessed were incidental to other work, which sometimes precluded determination of an accurate weight for the prey item. In such situations or when prey items were in advanced states of digestion, we extrapolated the average prey mass from fresh conspecific individuals of equivalent size or assumed a mean mass for the species based on the literature (Silva 1979; Sampedro and Montañez 1989; Silva et al. 2007; Jiménez et al. 2014; Sibley 2017). When possible, we measured the SVL of snakes to the nearest centimeter using the string method (Rivas et al. 2008) but, in some cases, we visually estimated total length. Cuban Boas have relatively short tails averaging only about 10% of total length (T.M. Rodríguez-Cabrera, unpubl. data). Thus, we assumed that our visual estimates of total length approximated SVL.

Hypothesis testing.—To test the hypothesis that *Ch. angulifer* is a generalist predator, we calculated its trophic niche breadth (Krebs 1998) using the standardized version of Levins’ (1968) niche breadth (Hurlbert 1978):

\[
B' = \frac{B - 1}{n - 1}
\]

where \(B'\) is the standardized version of Levins’ niche breadth, \(B\) is Levins’ measure of niche breadth, \(n\) is the number of possible resource states, and \(p_j\) is the proportion of individuals found using resource state \(j\). Levins’ standardized niche breadth ranges from 0 to 1. This index reaches a maximum...
when each resource state is used by a similar number of individuals in proportion to its abundance, which means that the species does not discriminate between one resource state or another. It reaches a minimum when only a single resource state is used by all individuals, reflecting the narrowest possible niche and hence maximum specialization. We grouped prey items into 16 resource states according to similarities in body shape, size, and/or ecological features (i.e., anurans, lizards, snakes, turtles, aquatic birds [Gruiformes], free-ranging raptors [Accipitriformes, Cathartiformes], forest birds [Columbiformes, Cuculiformes, Passeriformes], caged birds [Columbiformes, Galliformes, Falconiformes, Passeriformes, Psittaciformes], free-ranging poultry [Anseriformes, Galliformes], bats, bovids, pigs, carnivores, rabbits [caged], hutias, and rats). Some taxonomic groups were repeated in the cases of free-ranging native and domestic birds and caged birds. However, because the way a bird is captured in the wild differs considerably from the way a caged bird is captured (i.e., little chance to evade the snake), we split them into different categories. We assumed that each of the 16 resource states involved different foraging modes, energy costs, and energy gains. We analyzed the niche breadth at two levels, for the species as a whole and separately for boas found in natural habitats and for those in anthropogenic habitats.

To test the hypothesis that the diets of boas in natural and anthropogenic habitats differ, we grouped the prey items into four classes (i.e., amphibians, reptiles, birds, and mammals) and then grouped all predation events into the two habitat categories based on qualitative criteria. We designated as natural habitats all localities without permanent evidence of human activities (Fig. 3). These are mostly karstic areas with caves and primary vegetation in the form of dense forests. Some areas with semi-natural or secondary vegetation far from permanent human settlements also were considered natural. We designated as anthropogenic habitats those localities with an ongoing incidence of human activities (Fig. 3). These are matrices of heavily disturbed habitats usually set aside for agriculture and/or stockbreeding, with isolated patches of secondary vegetation and usually located in rural, suburban, or urban areas (e.g., small villages and surroundings, farms, pastures, sugarcane fields, fruit plantations).

To test the hypothesis that an ontogenetic shift in diet occurs in *Ch. angulifer*, we grouped boas into four size classes: (1) Juveniles (<1.0 m SVL), (2) subadults and small adults (1.0–2.0 m SVL); sexual maturation occurs within this size range: ~1.15 m SVL in males, ~1.3 m SVL in females; (3) medium-sized adults (>2.0–3.0 m SVL), and (4) large adults (>3.0 m SVL). Hardy (1957) did not provide the exact sizes of five snakes preying on bats; we placed them in the subadult/small adult category based on an estimated mean value derived from the range in sizes of the snakes he studied (see Appendix I). Because of the wide range of sizes and body shapes among birds and mammals, we split those prey items into several categories, resulting in 11 total categories of prey (i.e., amphibians [50–300 g], reptiles [<100 g], small birds [<100 g], medium-sized birds [100–500 g], large birds [>500 g], bats [<50 g], rats [pups and adults, 20–120 g], hutias [young and adults, 1,200–5,000 g], rabbits [kits, 50–500 g], carnivores [domestic/feral dogs and cats, 500–4,000 g], and artiodactyls [suckling pigs, goats, and sheep, 1,000–6,000 g]). We compared prey types and their frequencies in the diets of boas in the four size classes, within the same habitat type and between different habitat types. Because of the unequal number of boas in the four size classes, we limited statistical analyses to those size classes (subadult/small adult and medium-sized adult) with sufficiently large
sample sizes. We used qualitative criteria to compare the other size classes. Using the R package “ggplot2” (Wickham 2016; R Core Team 2018), we generated a scatter plot on which we plotted the SVL of all snakes for which a measurement was available (all size classes) against prey mass. Also, to facilitate visualization and interpretation of the data, we plotted the data transformed to their cube roots (Cox 2008).

To test the hypothesis that foraging strategies of boas differed between natural and anthropogenic habitats, we used only actual predation events witnessed (i.e., snakes found while constricting or swallowing prey) to determine foraging mode and time. Although recognizing that foraging modes represent a continuum from sit-and-wait to active-foraging strategies, we classified predation events as resulting from either as sit-and-wait (ambush) or active-foraging (searching) mode (Schoener 1971). According to the time at which a predation event occurred, we classified it as diurnal or nocturnal. We also used a substantial number of stomach-content data to characterize foraging strategy. For example, we could reasonably assume which foraging strategy was used by a boa when the prey item was a domestic animal the owner was able to recognize and could provide information on its usual sleeping routine. Domestic fowl regularly use the same roosting/sleeping sites. Many of the snakes with stomach contents in anthropogenic habitats were found close to roosting sites on the next day or a few days after the prey went missing (in those cases, the assumption that an active-foraging mode was used seemed reasonable). We compared the proportion of boas using each foraging mode in natural habitats and anthropogenic habitats.

Statistical analysis.—We conducted permutational multivariate analyses of variance (PERMANOVA) to test the null hypotheses of no differences in prey composition and frequency of occurrence in the diet of Ch. angulifer between habitats and among size classes. First, we conducted a multivariate one-factor design: Factor Habitat (two levels fixed: natural and anthropogenic) to test for global differences in diet composition (amphibians, reptiles, birds, and mammals) and frequency between habitats. Second, we conducted a multivariate (11 categories) two-factor design: Factor Habitat (two levels fixed: natural and anthropogenic) and Factor Size Class (two levels fixed: subadults/small adults and medium-sized adults). We calculated Bray-Curtis similarity matrices from fourth root-transformed data and the permutation tests used 9999 unrestricted permutations of raw data for the first analysis. We also ran permutations of residuals under a reduced model for the second analysis. We used the PRIMER-E (v6.1.16) and PERMANOVA+ (v1.0.6) statistical packages (Clarke and Warwick 2001, Anderson et al. 2008) to conduct analyses. Using R software (R Core Team 2018), we conducted Pearson’s chi-squared tests with Yates’s continuity corrections ($\chi^2$) to test the null hypotheses of: (1) independence of habitat type and foraging strategy and (2) independence of habitat type and foraging time.

Results

General diet composition and trophic niche breadth.—We recorded 351 prey items obtained from 218 snakes: 71 items from the literature (n = 31 snakes) and 280 items from this work (n = 187 snakes; Table 1). We confirmed 49 different taxa in the diet of Ch. angulifer. Most prey items were endotherms (n = 337 items [96%]; n = 204 snakes), whereas ecto-therms represented only a small portion (n = 14 items [4%]; n = 14 snakes). Mammals represented 54.7% of total prey items consumed (n = 192 items; n = 105 snakes), followed by birds (n = 145 items [41.3%]; n = 99 snakes), whereas amphibians (n = 7 items [2%]; n = 7 snakes) and reptiles (n = 7 items [2%]; n = 7 snakes) comprised only a small proportion of the diet (Table 1; Fig. 4). The prey species most frequently consumed were domestic fowl (Gallus gallus) (24.8%), Desmarest’s Hutias (Capromys pilorides) (14.5%), two bat species (Jamaican Fruit-eating Bat [Artibeus jamaicensis]: 8.8%; Cuban Flower Bat [Phyllonycteris poeyi]: 6.0%), and House Rats (Rattus rattus) (7.7%); the remaining prey species were taken only sporadically.

Our data show that Ch. angulifer has a narrow trophic niche (B’ = 0.319). Most snakes (75.4%) consumed only four prey types (free-ranging poultry [32.1%], hutias [19.9%], bats [11.9%], and rats [11.5%]). When we analyzed the trophic niche breadths of boas in the two habitat types, the species had a slightly wider niche in natural habitats (B’ = 0.321) than in anthropogenic habitats (B’ = 0.135). In natural habitats, most boas (83.3%) consumed only four prey types (hutias [33.3%], bats [22.8%], rats [15.8%], and forest birds [11.4%]). In anthropogenic habitats, most boas (77.9%) consumed only two prey types (free-ranging poultry [65.4%] and caged birds [12.5%]).

Most of the snakes with stomach contents (n = 116 snakes) contained a single prey item (n = 84 snakes, 72.4%),

![Fig. 4. Frequency of occurrence of four prey types (amphibians, reptiles, birds, and mammals) in the diet of the Cuban Boa (Chilabothrus angulifer). Columns indicate the number of prey items and the number of snakes found consuming each resource.](image-url)
Table 1. Prey taxa confirmed for free-ranging Cuban Boas (*Chilabothrus angulifer*) in natural (NAT) and anthropogenic (ANT) habitats, including data from the literature and this work. The percentage of total prey items represented by each species is listed in parentheses after the name of the species. The number of prey items is followed (in parenthesis) by the number of snakes involved; question marks (?) represent unknown data. Prey growth states (GS) were defined as adult (A), juvenile/nestling/hatchling (J), and egg (E). Snake size classes are juveniles (JU: < 1.0 m TL), subadults/small adults (SA: 1.0–2.0 m TL), medium-sized adults (MA: > 2.0–3.0 m TL), and large adults (LA: > 3.0 m TL). When a prey species is reported by one or more authors, a superscript indicates the number of items referred. Sources: 1. This paper; 2. Tolson (2012); 3. Holanova and Hribal (2004), V. Holanova, in litt. 6.iv.2020; 4. Viña and Armas (1988); 5. Sampedro and Montañez (1989); 6. Sampedro (1998, in litt. 6.xii.2015; 7. Vázquez and Nieves (1980); 8. Godínez et al. (1987); 9. Segovia et al. (2013); 10. Mancina and Llanes (1997), C.A. Mancina, pers. comm. 19.v.2020; 11. Tolson and Petersen (2008), P.J. Tolson, in litt. 28.iii.2020; 12. Sheplan and Schwartz (1974); 13. Mancina (2011), pers. comm. 19.v.2020; 14. Borroto-Páez (2011a); 15. Tolson and Henderson (1993), P.J. Tolson, in litt. 8.v.2020.

*Recorded as *Anolis bartschi* (which certainly represents a different species than *A. smallwoodi*; see Appendix I). *Recorded as *Pseudemys decussata*. *Recorded as *Columba leucocephala*. *Recorded as Hirundo fulva.*

The six bats (i.e., “two Mormoops blainvillei, two Phyllonycteris poeyi, one Brachyphylla nana, and one small unidentifiable bat”) reported by Sheplan and Schwartz (1974), were in a single snake.

| Prey | Prey items (Snakes) | NAT | ANT | Prey GS | Snake Size | Class | Source |
|------|---------------------|-----|-----|---------|------------|-------|--------|
| **AMPHIBIA** | | | | | | | |
| *Anura: Hylidae* | | | | | | | |
| Cuban Treefrog, *Osteopilus septentrionalis* (1.4%) | 5 (5) | — | A | JU, SA | 1 |
| **Anura: Ranidae** | | | | | | | |
| American Bullfrog, *Lithobates catesbeianus* (0.3%) | — | 1 (1) | A | SA | 1 |
| **Anura: Bufonidae** | | | | | | | |
| Western Cuba Giant Toad, *Peltophryne fustiger* (0.3%) | 1 (1) | — | A | SA | 1 |
| **REPTILIA** | | | | | | | |
| *Squamata: Dactyloidae* | | | | | | | |
| Green-blotched Giant Anole, *Anolis smallwoodi* (0.3%) | 1 (1) | — | A | JU | 2 |
| Unidentified anole, *Anolis* sp.*a* (0.3%) | 1 (1) | — | ? | JU | 3 |
| **Squamata: Leiocephalidae** | | | | | | | |
| Saw-scaled Curlytail, *Leiocephalus carinatus* (0.6%) | 2 (2) | — | A | JU, SA | 1 |
| **Squamata: Iguanidae** | | | | | | | |
| Cuban Iguana, *Cyclura nubila* (0.3%) | — | 1 (1) | J | SA | 1 |
| **Squamata: Tropidophiidae** | | | | | | | |
| Giant Trope, *Tropidophis melanurus* (0.3%) | 1 (1) | — | J | SA | 4 |
| **Testudines: Emydidae** | | | | | | | |
| Cuban Slider, *Trachemys decussata*b (0.3%) | 1 (1) | — | J | SA | 5, 6 |
| **AVES** | | | | | | | |
| Accipitriformes: Accipitridae | | | | | | | |
| Red-tailed Hawk, *Buteo jamaicensis* (0.3%) | 1 (1) | — | A | MA | 1 |
| **Cathartiformes: Cathartidae** | | | | | | | |
| Turkey Vulture, *Cathartes aura* (0.6%) | 2 (1) | — | J | MA | 1 |
| **Anseriformes: Anatidae** | | | | | | | |
| Muscovy Duck, *Cairina moschata* (0.9%) | — | 3 (1) | J | SA | 1 |
| **Columbiformes: Columbidae** | | | | | | | |
| Rock Pigeon, *Columba livia* (1.1%) (caged) | — | 4 (3) | 2A, 2J | SA | 1 |
| Ruddy Quail-Dove, *Geotrygon montana* (0.3%) (caged) | — | 1 (1) | A | MA | 1 |
| White-crowned Pigeon, *Patagioenas leucocephala*b (%) | ? | — | E, J, A | ? | 7, 8, 1 |
| Zenaida Dove, *Zenaida aurita* (0.3%) | 1 (1) | A | SA | 1 |
| Mourning Dove, *Zenaida macroura* (1.7%) | 6 (5) | — | A | SA | 1 |
| **Cuculiformes: Cuculidae** | | | | | | | |
| Great Lizard-Cuckoo, *Coccyzus merlini* (0.3%) | 1 (1) | — | A | MA | 1 |
| **Falconiformes: Falconidae** | | | | | | | |
| Northern Crested Caracara, *Caracara cheriway* (0.3%) (caged) | — | 1 (1) | A | MA | 1 |
| **Galliformes: Phasianidae** | | | | | | | |
| Japanese Quail, *Coturnix japonica* (2.6%) (caged) | — | 9 (1) | A | MA | 1 |
| Domestic Chicken, *Gallus gallus* (24.8%) (10 caged) | 2 (2) | 85 (61) | 53J, 34A | SA, MA, LA | 1 |
| Wild Turkey, *Meleagris gallopavo* (1.1%) | — | 4 (2) | J | SA | 1 |

(continued)
| Prey                                      | Prey items (Snakes) | Prey Size | Snake Size | Source |
|-------------------------------------------|---------------------|-----------|------------|--------|
| Galliformes: Numididae                    |                     |           |            |        |
| Helmeted Guineafowl, *Numida meleagris*  | —                   | 6J, 1A    | SA         | 1      |
| Purple Gallinule, *Porphyrio martinicus* | 1 (1)               | —         | A          | SA     | 1      |
| Gruiformes: Rallidae                      |                     |           |            |        |
| Greater Antillean Grackle, *Quiscalus niger* | 1 (1)               | —         | A          | JU     | 1      |
| Unidentified blackbird (0.3%)             | 1 (1)               | —         | A          | MA     | 1      |
| Passeriformes: Icteridae                  |                     |           |            |        |
| Gray Kingbird, *Tyrannus dominicensis* (0.3%) | 1 (1)               | —         | J          | SA     | 9      |
| Passeriformes: Estrildidae                |                     |           |            |        |
| Tricolored Munia, *Lonchura malacca* (1.7%) (caged) | —                   | 6 (1)     | A          | JU     | 1      |
| Passeriformes: Tyrannidae                 |                     |           |            |        |
| Cave Swallow, *Petrochelidon fulva* (0.6%) | 2 (1)               | —         | E          | SA     | 10     |
| Passeriformes: Thraupidae                 |                     |           |            |        |
| Western Spindalis, *Spindalis zena* (0.3%) (caged) | —                   | 1 (1)     | A          | JU     | 1      |
| Passeriformes: Turdidae                   |                     |           |            |        |
| Red-legged Thrush, *Turdus plumbeus* (0.6%) | 2 (2)               | —         | A          | SA     | 1      |
| Psittaciformes: Psittacidae               |                     |           |            |        |
| Rosy-faced Lovebird, *Agapornis roseicollis* (0.3%) (caged) | —                   | 1 (1)     | A          | JU     | 1      |
| Budgerigar, *Melopsittacus undulatus* (0.3%) (caged) | —                   | 1 (1)     | A          | JU     | 1      |
| Cuban Parakeet, *Psittacula eupatria* (0.3%) (caged) | —                   | 1 (1)     | A          | JU     | 1      |
| Mammalia                                  |                     |           |            |        |
| Artiodactyla: Bovidae                     |                     |           |            |        |
| Domestic Goat, *Capra hircus* (0.6%)      | —                   | 2 (2)     | J          | LA     | 11, 1 |
| Domestic Sheep, *Ovis aries* (0.3%)       | —                   | 1 (1)     | J          | LA     | 1      |
| Artiodactyla: Suidae                      |                     |           |            |        |
| Domestic Pig, *Sus scrofa* (3.7%)         | 5 (1)               | 8 (3)     | J          | MA, LA | 1      |
| Carnivora: Canidae                        |                     |           |            |        |
| Domestic Dog, *Canis lupus familiaris* (1.1%) | —                   | 4 (2)     | J          | SA     | 1      |
| Carnivora: Felidae                        |                     |           |            |        |
| Domestic Cat, *Felis catus* (1.1%)        | 1 (1)               | 3 (3)     | 2J, 2A     | SA, MA | 1      |
| Chiroptera: Phyllostomidae                |                     |           |            |        |
| Jamaican Fruit-eating Bat, *Artibeus jamaicensis* (8.8%) | 31 (9)              | —         | A          | SA     | 12     |
| Cuban Fruit-eating Bat, *Brachyphylla nana* (0.6%) | 2 (2)               | —         | A          | SA     | 13, 15 |
| Buffy Flower Bat, *Erophylla sezekorni* (1.7%) | 6 (3)               | —         | 3J, 3A     | JU, SA | 15, 13 |
| Cuban Flower Bat, *Phyllostomus perezi* (6.0%) | 21 (10)             | —         | A          | JU, SA | 1, 14, 15 |
| Chiroptera: Mormoopidae                   |                     |           |            |        |
| Antillean Ghost-faced Bat, *Mormoops blainvilliei* (1.1%) | 4 (2)               | —         | A          | SA     | 14, 1 |
| Unidentified bats* (1.7%)                 | 6 (3)               | —         | A          | SA     | 14, 1 |
| Lagomorpha: Leporidae                      |                     |           |            |        |
| European Rabbit, *Oryctolagus cuniculus* (4.3%) (caged) | —                   | 15 (2)    | J          | SA, MA | 1      |
| Rodentia: Capromyidae                     |                     |           |            |        |
| Desmarest’s Hutia, *Capromys pilorides* (14.5%) | 51 (37)             | —         | 8J, 43A    | SA, MA, LA | 17, 18, 1, 14 |
| Prehensile-tailed Hutia, *Mysateles prehensilis* (0.3%) | —                   | 1 (1)     | A          | MA     | 1      |
| Black-tailed Hutia, *Mesocapromys melanurus* (0.3%) | 1 (1)               | —         | A          | MA     | 1      |
| Rodentia: Muridae                          |                     |           |            |        |
| House Rat, *Rattus rattus* (7.7%)         | 22 (17)             | 5 (5)     | 5J, 22A    | SA, MA, LA | 19, 20, 1, 15 |
| Brown Rat, *Rattus norvegicus* (0.3%)     | —                   | 1 (1)     | A          | MA     | 1      |
| Unidentified rats, *Rattus sp.* (0.6%)    | 1 (1)               | 1 (1)     | A          | MA     | 1      |
| Total prey items (351)                    | 184                 | 167       |            |        |
| Total snakes (218)                        | 114                 | 104       |            |        |
| Total prey species (49)                   | 29                  | 24        |            |        |
but 32 snakes (27.6%) contained two or more items (Table 2). Thus, for several prey species, the number of items recorded was higher than the number of snakes containing them (Fig. 4; Table 1). The maximum number of prey items in a single boa was nine (n = 3 snakes; Table 2). In a single instance, a snake contained more than one prey species in its stomach (all bats; n = 6 items) (Tables 1 and 2, Appendix I). One snake (1.47 m SVL) was observed swallowing an adult Buffy Flower Bat (Erophylla sezekorni) while constricting another bat of the same species. Another snake (<2.0 m SVL) was observed coiled in the nest of a Cave Swallow (Petrorhynchus fulva) a short time after the nest had been checked and had contained two eggs (Appendix I). These two cases represent the only snakes found in the act of predation involving multiple prey items in natural habitats. Seven snakes in anthropogenic habitats contained prey items in their stomach while constricting or swallowing other individuals of the same species (n = 37 items).

Ninety-five snakes (43.6%) were observed while taking prey (Table 3). Half (n = 47 snakes, 49.5%) were preying on poultry, 16 (16.8%) on bats, and ten (10.5%) on ecotermals. Fewer snakes were observed preying on huitas (n = 8, 8.4%), rats (n = 6, 6.3%), forest birds (n = 6, 6.3%), and other prey types.

**Diets in natural and anthropogenic habitats.**—We identified 29 taxa in natural habitats (n = 184 items; n = 114 snakes) and 24 taxa in anthropogenic habitats (n = 167 items; n = 104 snakes) (Table 1). We found significant differences in the dietary composition of boas in natural and anthropogenic habitats (pseudo-$F_{(1)} = 90.477$; $P = 0.0001$; Table 4A).

Only four prey taxa (rats [Rattus rattus], domestic fowl [Gallus gallus], cats [Felis catus], and pigs [Sus scrofa]) were consumed by snakes in both habitat types, albeit in different proportions (Table 1). Mammals were consumed more frequently in natural habitats (82.1%, n = 151 items; n = 84 snakes) than in anthropogenic habitats (24.6%, n = 41 items; n = 21 snakes) (Fig. 5). The most frequently taken prey items in natural habitats were bats (n = 70 items [38%]; n = 26 snakes), followed by huitias (Capromys and Mesopropus; n = 52 items [28.3%]; n = 38 snakes), and rats (Rattus; n = 23 items [12.5%]; n = 18 snakes) (Fig. 6). Domestic animals consumed in natural habitats were feral (i.e., cats) or semi-feral (i.e., chickens and pigs) and represented only a small portion of the total number of prey items consumed (n = 8 items [4.4%]; n = 4 snakes). Birds were more frequently consumed by boas in anthropogenic habitats (n = 124 items [74.3%]; n = 81 snakes) than by boas in natural habitats (n = 21 items [11.4%]; n = 18 snakes). The most frequently taken prey items in anthropogenic habitats were domestic chickens (n = 85 items; n = 61 snakes), which comprised 50.9% of all prey items and 68.6% of all birds consumed in this habitat type. All native or introduced feral birds (i.e., Caracaras [Caracara cheriway], Quail Doves [Geotrygon montana], Munias [Lonchura malaccensis], Parakeets [Psittacula eupara], and Spindalis [Spindalis zedalis]) consumed in anthropogenic habitats (n = 10 items, n = 5 snakes) were caged. Rats accounted for only 4.2% of the total number of prey items taken in anthropogenic habitats (n = 7 items; n = 7 snakes; Fig. 7). Artiodactyls (n = 16 items) were consumed only by snakes larger than 2.5 m SVL (n = 3 medium-sized adults; n = 4 large adults; Fig. 6). Except for one snake (>3.0 m SVL) in a natural habitat that contained five suckling semi-feral pigs, all artiodactyl prey were taken by boas in anthropogenic habitats. The largest measured snake included in this work was an adult female 5.65 m in total length (ca. 5.0 m SVL) and probably exceeding 40 kg that had been killed on a farm near the village of Las Vegas, Guamuhaya Massif, Cuanayagua.

**Table 2.** Individual Cuban Boas (Chilabothrus angulifer) with multiple prey items determined either by examination of stomach contents and/or observations of predation events. References (superscripts): 1. Mancia and Llanes (1997); C.A. Mancia, pers. comm. 19.v.2020; 2. Rodríguez-Cabrera et al. (2015); 3. Hardy (1957); 4. Shepler and Schwartz (1974).

| Items | Snakes | Prey species (number of prey items and growth states) |
|-------|--------|------------------------------------------------------|
| 2     | 17     | Turkey Vultures, Cathartes aura (2 chicks), Rock Pigeons, Columba livia (2 adults), Wild Turkeys, Meleagris gallopavo (2 chicks), Helmed Guineafowl, Numida meleagris (2 chicks), Cave Swallows, Petrochelidon fulva (2 eggs)¹, Mourning Doves, Zenaida macroura (2 adults), Buffy Flower Bats, Erophylla sezekorni (2 adults), Desmarest’s Huitas, Capromys pilorides (2 young and 14 adults), Antillean Ghost-faced Bats, Mormoops blainvillii (2 adults), Domestic Pigs, Sus scrofa (2 sucklings) |
| 3     | 9      | Muscovy Ducks, Cairina moschata (chicks), Domestic Dogs, Canis lupus familiaris (3 sucklings), Buffy Flower Bats, Erophylla sezekorni (3 sucklings)², Desmarest’s Huitas, Capromys pilorides (9 adults), Cuban Flower Bats, Phyllostomus hastatus (1 unidentified bat)³, Domestic Pigs, Sus scrofa (6 sucklings) |
| 4     | 1      | Unidentified bats |
| 5     | 1      | Domestic Pigs, Sus scrofa (sucklings) |
| 6     | 3      | Bats spp. (1 Cuban Fruit-eating Bat, Brachypodia nana; 2 Antillean Ghost-faced Bats, Mormoops blainvillii; 2 Cuban Flower Bats, Phyllostomus hastatus; 1 unidentified bat)⁴; European Rabbit, Oryctolagus cuniculus (6 sucklings); House Rats, Rattus rattus (5 sucklings and 1 adult female) |
| 9     | 3      | Japanese Quail, Coturnix japonica (9 adults); Domestic Chickens, Gallus gallus (8 chicks and 1 hen); Cuban Flower Bats, Phyllostomus hastatus (9 adults)³ |
Cienfuegos Province in 1987; it contained a young domestic goat (*Capra hircus*) ca. 6.0 kg (J.D. León, pers. comm. 2012).

Desmarest's Hutia was the most frequent prey species consumed in natural habitats (n = 51 items [27.7%]; n = 37 snakes; Fig. 8). Only one boa (>2.0 m SVL) in a natural habitat near Farallones, Moa, Holguín Province, contained an adult Black-tailed Hutia (*Mesocapromys melanurus*) (G. Begué, pers. comm. 2013; Fig. 9). Another boa (ca. 3.0 m SVL) found in a secondary grove associated with an anthropogenic habitat at Cariblanca in Sancti Spíritus Province contained an adult Prehensile-tailed Hutia (*Mysateles prehensilis*), the only case of predation on hutias in this type of habitat. Snakes (n = 31) containing hutias (*Capromys*, *Mesocapromys*, and *Mysateles*) in their stomachs belonged to all size classes except juveniles, and the number of items per stomach varied: one item (n =

Table 3. Observations of predation events by Cuban Boas (*Chilabothrus angulifer*) in natural (NAT) and anthropogenic (ANT) habitats. Asterisks (*) indicate that more prey items than those observed being taken were revealed later as stomach contents (e.g., four snakes preying on Domestic Chickens [*Gallus gallus*]). When a prey species has been reported by one or more authors, a superscript indicates the number of snakes involved. References: 1. This paper; 2. Holanova and Hribal (2004); 3. Viña and Armas (1988); 4. Sampedro and Montañez (1989); 5. Sampedro (1998), in litt. 19.v.2015; 6. Segovia et al. (2013); 7. Mancina and Llanes (1997), C.A. Mancina, pers. comm. 19.v.2020; 8. Dinets (2017); 9. Mancina (2011), pers. comm. 19.v.2020; 10. Rodríguez-Cabrera et al. (2015); 11. Hardy (1957); 12. Tolson and Petersen (2008), P.J. Tolson, in litt. 28.iii.2020; 13. Borroto-Páez (2011a); 14. Tolson and Henderson (1993), P.J. Tolson, in litt. 8.v.2020.

| Prey Items | Snakes | Time | Habitat | Source |
|------------|--------|------|---------|--------|
| Cuban Treefrog, *Osteopilus septentrionalis* | 5      | 5    | day     | NAT    |
| Western Cuba Giant Toad, *Peltophryne fustiger* | 1      | 1    | night   | NAT    |
| Unidentified anole, *Anolis* sp. | 1      | 1    | night   | NAT    |
| Cuban Iguana, *Cyclura nubila* | 1      | 1    | day     | ANT    |
| Giant Trope, *Tropidophis melanus* | 1      | 1    | day     | NAT    |
| Cuban Slider, *Trachemys decussata* | 1      | 1    | night   | NAT    |
| Rock Pigeon, *Columba livia* (caged) | 1      | 1    | night   | ANT    |
| Zenaida Dove, *Zenaida aurita* | 1      | 1    | day     | NAT    |
| Domestic Chicken, *Gallus gallus* (2 snakes on caged birds) | 45*    | 45   | day, night | ANT |
| Wild Turkey, *Meleagris gallopavo* | 1*     | 1    | night   | ANT    |
| Helmeted Guineafowl, *Numida meleagris* | 5      | 5    | night   | ANT    |
| Greater Antillean Grackle, *Quiscalus niger* | 1      | 1    | day     | NAT    |
| Gray Kingbird, *Tyrannus dominicensis* | 1      | 1    | night   | NAT    |
| Tricolored Munia, *Lonchura malaca* (caged) | 1*     | 1    | night   | ANT    |
| Cave Swallow, *Petrochelidon fulva* (eggs) | 2      | 1    | day     | NAT    |
| Red-legged Thrush, *Turdus plumbeus* | 1      | 1    | day     | NAT    |
| Great Lizard-Cuckoo, *Coccyzus merlini* | 1      | 1    | day     | NAT    |
| Rosy-faced Lovebird, *Agapornis roecollis* (caged) | 1      | 1    | night   | ANT    |
| Domestic Cat, *Felis catus* | 1      | 1    | day     | NAT    |
| Jamaican Fruit-eating Bat, *Artibeus jamaicensis* | 31     | 9    | night   | NAT    |
| Cuban Fruit-eating Bat, *Brachyphylla nana* | 1      | 1    | day     | NAT    |
| Buffy Flower Bat, *Erophylla sezekorni* | 2      | 1    | night   | NAT    |
| Cuban Flower Bat, *Phyllonycteris poeyi* | 4      | 4    | night   | NAT    |
| Unidentified bat | 1      | 1    | day     | NAT    |
| European Rabbit, *Oryctolagus cuniculus* (caged) | 1*     | 1    | night   | ANT    |
| Desmarest's Hutia, *Capromys pilorides* | 8      | 8    | day     | NAT    |
| House Rat, *Rattus rattus* | 6      | 6    | day     | NAT, ANT |

Table 4. Permutational multivariate analyses of variance (PERMANOVA) on the dietary composition of Cuban Boas (*Chilabothrus angulifer*) in natural and anthropogenic habitats (A) and among size classes (B): subadults/small adults in natural and anthropogenic habitats, and medium-sized adults in natural and anthropogenic habitats. All values are significant at α = 0.05. df = degrees of freedom, MS = mean squares.

| Source of Variation | df  | MS     | Pseudo-F | P (perm)  |
|---------------------|-----|--------|----------|----------|
| A Habitat           | 1   | 48581  | 90.477   | 0.0001   |
| Residual            | 225 | 536.95 |          |          |
| B Habitat           | 1   | 17998  | 19.778   | 0.0001   |
| Size class          | 1   | 14573  | 16.014   | 0.0001   |
| Habitat x Size Class| 1   | 8114.1 | 8.9164   | 0.0001   |
| Residual            | 171 | 910.01 |          |          |
Most of the snakes that preyed on *Ca. pilorides* (*n* = 37 snakes: 29 stomach contents and 8 found preying) exceeded 2.0 m SVL, with the exception of only four subadults/small adults (2 encountered during a predation event and 2 stomach contents) ranging in size from 1.7–1.8 m in total length (ca. 1.5–1.6 m SVL), each of which had taken a juvenile hutia (ca. 2.0 kg each). Mourning Doves (*Zenaida macroura*) were repeatedly reported as prey of subadult/small adult snakes in natural habitats (*n* = 6 items; *n* = 5 snakes; Fig. 10). Two snakes (>2.5 m SVL) in natural habitats contained a partially digested adult Red-tailed Hawk (*Buteo jamaicensis*) and two large nestling Turkey Vultures (*Cathartes aura*), respectively (Table 1). Ectotherms, including anurans (Cuban Treefrogs, *Osteopilus septentrionalis*; American Bullfrogs, *Lithobates catesbeianus*; and Western Cuba Giant Toads, *Peltophryne fustiger*; *n* = 7 items), lizards (anoles, *Anolis* spp.; Cuban Iguana, *Cyclura nubila*; Saw-scaled Curlytail, *Leiocephalus carinatus*; *n* = 5 items), a Giant Trope (*Tropidophis melanurus*; *n* = 1 item), and a hatchling Cuban Slider (*Trachemys decussata*; *n* = 2 subadults/small adults; *n* = 10 medium-sized adults; *n* = 8 large adults), two items (*n* = 2 medium-sized adults; *n* = 6 large adults), and three items (*n* = 3 large adults) (Table 2).
Fig. 7. A small adult Cuban Boa (*Chilabothrus angulifer*) (1.5 m SVL) containing an adult House Rat (*Rattus rattus*) (ca. 120 g) in an anthropogenic habitat in the Cienfuegos Botanical Garden, Cienfuegos Province. Photographs © T.M. Rodríguez-Cabrera.

Fig. 8. A medium-sized Cuban Boa (*Chilabothrus angulifer*) (ca. 3.0 m SVL) containing a Desmarest’s Hutia (*Capromys pilorides*) in a natural habitat in the Zapata Swamp, Matanzas Province (left). This species of hutia (1.3–6.9 kg) (right) is the most frequently taken prey item of the Cuban Boa in natural habitats, particularly by individuals >2.0 m SVL. Photographs © Rafael A. Pérez (left) and Aslam I. Castellón (right).

Fig. 9. A medium-sized Cuban Boa (*Chilabothrus angulifer*) (>2.0 m SVL) containing an adult Black-tailed Hutia (*Mesocapromys melanurus*) in natural habitat near Farallones, Moa, Holguín Province (left). This species of hutia (1.0–1.6 kg) (right) is restricted to eastern Cuban, is highly arboreal, and lives in family groups of as many as 10 individuals. Photographs © Carlos A. Pérez (left) and Samuel Reina (right).
1 item), were consumed only by juveniles (n = 6 snakes) and subadults/small adults (n = 6 snakes) (Fig. 6). Bats (n = 70 items; n = 26 snakes) also were consumed only by juveniles (n = 5 snakes) and subadults/small adults (n = 21 snakes) (Figs. 6 and 11). Indeed, the smallest measured snake included in this work was a neonate (505 mm SVL) found swallowing an adult Cuban Flower Bat (Phyllonycteris poeyi) captured on the wing at a cave entrance in central Cuba (Appendix I).

**Ontogenetic shift in diet.**—The frequency of prey types consumed by Ch. angulifer differed significantly between subadults/small adults and medium-sized adults (pseudo-F$_{(1)}$ = 16.014; P = 0.0001; Table 4B; Fig. 6) and also when the habitat type was considered (pseudo-F$_{(1)}$ = 19.778; P = 0.0001; Table 4B; Fig. 6). The interaction of habitat x size class also was significant (pseudo-F$_{(1)}$ = 8.9164; P = 0.0001; Table 4B). After merging and analyzing the data from 152...
predation events in both habitat types and for which the sizes of the snakes were known, three trends were evident: (1) Prey size increased with snake size, (2) most prey types were related to snake size, and (3) smaller prey items were omitted from the diets of larger snakes (Fig. 12).

In natural habitats, subadult/small adult snakes (n = 54) consumed 98 prey items and medium-sized snakes (n = 29) consumed 36 prey items; in anthropogenic habitats, subadult/small adult snakes (n = 63) consumed 84 prey items and medium-sized snakes (n = 30) consumed 67 prey items. The prey of subadults/small adults in natural habitats consisted of bats, non-volant mammals, forest birds, and ectotherms. Bats in particular, represented 64.3% of all prey items consumed by subadults/small adults in this type of habitat (n = 63 items, 21 snakes), followed by small and medium-sized native birds (n = 15 items [15.3%]; n = 13 snakes) (Fig. 6). The diet of medium-sized snakes in natural habitats was comprised largely of hutias, but these snakes also readily took rats and semi-feral domestic fowl. Hutias (Capromys and Mesopromys) represented 63.9% of all prey items consumed by medium-sized snakes in natural habitats (n = 23 items; n = 17 snakes), but for subadult/small adult snakes, hutias (n = 4 items [4.1%]; n = 4 snakes) made up only a small proportion of prey items taken, and those instances always involved young hutias.

The diet of subadult/small adult snakes in anthropogenic habitats included mostly small, medium-sized, and large birds, but also ectotherms, rats, and domestic mammals (Fig. 6). Medium-sized domestic birds (n = 44 items [52.4%]; 38 snakes), including adult and nestling domestic pigeons (Columba livia) and small adult poultry (Cairina, Gallus, Meleagris, and Numida), were taken most frequently by subadults/small adults. The diets of medium-sized snakes in anthropogenic habitats included most of the prey types consumed by subadult/small adult snakes, but the proportions were different. The prey types most frequently consumed by medium-sized snakes in this type of habitat were small, medium-sized, and large birds in similar proportions, altogether comprising 71.6% of the total number of prey items consumed (n = 48 items; n = 24 snakes). Medium-sized snakes also took a small number of suckling pigs. Domestic chickens represented 53.6% (n = 45 items; n = 37 snakes) and 55.2% (n = 37 items; n = 21 snakes) of the total number of prey items consumed by medium-sized adult and medium-sized snakes, respectively, in anthropogenic habitats.

Rats were consumed in similar proportions by subadult/small adult snakes in both natural (n = 9 items [9.2%]; n = 9 snakes) and anthropogenic habitats (n = 7 items [8.3%]; n = 7 snakes). Eight medium-sized snakes in natural habitats consumed eight rats (22.2% of the total number of prey items), but we found no medium-sized boas in anthropogenic habitats that had consumed rats.

We did not include juvenile snakes in the statistical analyses to test the ontogenetic shift in diet hypothesis because of small sample sizes (natural habitats: n = 14 items, n = 12 snakes; anthropogenic habitats: n = 10 items, n = 5 snakes). However, our few observations are worth mentioning. In natural habitats, juvenile snakes consumed ectotherms and endotherms in comparable numbers, whereas in anthropogenic habitats, juveniles consumed exclusively small caged birds (i.e., Agapornis, Lonchura, Melopsittacus, Psittacara, and Spindalis) (Table 1; Fig. 6). Similarly, we had a small sample size of large adult snakes in anthropogenic habitats (n = 6 items; n = 6 snakes). The few individuals belonging to this size class in this type of habitat primarily consumed large domestic fowl and suckling artiodactyls, such as pigs, goats and sheep, whereas in natural habitats, large adults (n = 36 items; n = 19 snakes) consumed hutias, rats, and semi-feral suckling pigs (Fig. 6). Hutias were consumed in a similar proportion by medium-sized and large adult snakes (n = 23 items [63.9%]; n = 15 snakes) in natural habitats (Fig. 6). Snakes in these size classes did not consume ectotherms or bats in any type of habitat.

Foraging strategy.—We documented 45 snakes in natural habitats and 50 in anthropogenic habitats engaged in preda-
tion events (Table 3). In 32 of the events in natural habitats and in 48 of the events in anthropogenic habitats, we determined the foraging mode used. We found that foraging strategy was not independent of habitat type ($\chi^2 = 33.148$, df = 1, $P = 8.542e-09$). Twenty snakes (62.5%) employed an ambush strategy and 12 snakes (38.7%) used an active-foraging mode in natural habitats. At least in one additional case involving stomach contents (juvenile bats) in a natural habitat, the snake must have used an active foraging strategy (see below). In all but one predation event observed in anthropogenic habitats and, in most if not all of the cases of stomach contents in this type of habitat, the snakes must have used an active foraging strategy (see below). Foraging time was not independent of habitat type ($\chi^2 = 29.513$, df = 1, $P = 5.553e-08$). More than half of the snakes engaged in predation events (n = 27 snakes, 60.0%) in natural habitats did so by day, whereas only 18 (40.0%) events were nocturnal (Table 3). All but three snakes engaged in predation events in anthropogenic habitats were found at night (96%; Table 3), and one of the cases found by day (constricting a rat) was in a sewer in complete darkness (Appendix I).

The snakes that were found consuming ectotherms in natural habitats apparently used both sit-and-wait and active-foraging strategies, in particular five juveniles preying on Cuban Treefrogs by day and another juvenile eating an anole at night (Table 3). Since these frogs are nocturnal and anoles are diurnal, in all instances the snakes must have actively searched for inactive prey. A small adult captured a hatchling Cuban Slider in its nest just after hatching and emergence (Tables 1 and 3). The foraging strategy used in the remaining cases involving ectotherms is uncertain.

Four of the snakes found in natural habitats were consuming forest birds by day and possibly used a sit-and-wait foraging mode. The prey items included a Greater Antillean Grackle (Quiscalus niger), a Great Lizard Cuckoo (Coccyzus merlini), a Red-legged Thrush (Turdus plumbeus), and a Zenaida Dove (Zenaida aurita) (Table 3). The boas, which apparently had been laying in ambush on the forest floor, were observed at the precise moment they captured the birds. Two other snakes probably used an active foraging mode to find two Cave Swallow ( Petrochelidon fulva) eggs and a nestling Gray Kingbird ( Tyrannus dominicensis) (Appendix I).

Four subadult/small adult snakes and one medium-sized snake were observed consuming rats in natural habitats by day (Table 3). Because rats are primarily nocturnal, most of the snakes in those cases probably used an active-foraging mode to find sleeping rats. At least in one case, a boa was observed stalking and eventually capturing a rat in a hollow tree on the Guanahacabibes Peninsula (R. Varela, pers. comm. 5.v.2020).

A boa approaching 2.0 m SVL was observed swallowing an adult feral cat (ca. 4.0 kg) early in the morning at the entrance of a hot cave near Galalón, Pinar del Río Province (R. Martínez, pers. comm. 31.i.2017). Prior to that observation, bat remains (wings) were frequently observed at the cave entrance, a typical sign of bat predation by cats. Only one cat might have been feeding on bats in that cave, as no more bat remains were observed after that cat was eaten by the boa.

All observed predation events involving Desmarest’s Hutias in natural habitats took place by day. Five snakes were observed constricking or swallowing hutias on the forest floor or on limestone rocks usually associated with karstic environments, two snakes captured the hutias in trees and fell to the ground while constricking their prey, and one snake was swallowing a hutia in a grassland by a beach (Table 3; Appendix I). The foraging strategy employed in these cases is unknown since Ca. pilorides is active by both day and night.

Most snakes observed while engaged in predations events in natural habitats (n = 16 snakes, 36.4%) were consuming bats associated with caves harboring large bat colonies. These snakes used a sit-and-wait foraging strategy. Bat-hunting always occurred in complete darkness, starting approximately 30–60 min after sunset. In some caves, boas employed two hunting sessions per night, one after sunset and another before sunrise. Snakes may take positions either in the usually narrow cave passages leading to the exit, right at the exit, or in vegetation immediately outside the cave. Boas may hang from rock crevices or coil around rock projections (e.g., stalactites) on the cave’s ceiling, around vines, branches, or aerial roots, but always in the pathway of the bats (Fig. 13). Alternatively, snakes may position themselves on the ground or climb onto a rock shelf or into a crevice in the cave chambers and passages where bats roost; in such instances, they typically elevate the anterior third of the body (Fig. 13). This appears to be especially effective when the cave roof is low enough to force the bats to fly near the ground. This same strategy is sometimes used by day as well, when for some reason (e.g., human intrusion), the bats becomes agitated and fly in circles around the chambers without leaving the cave. Contact with the extended portion of the snake’s body is necessary to trigger its attack, which consists of a rapid swing with the mouth open toward the origin of the impact. Once a bat is captured, the boa rapidly wraps it in one or two coils and begins the swallowing process, either while hanging or after capturing the bat from the ground (Fig. 14). Apparently, some boas capture multiple bats during a single hunting session. We observed at least one boa on a cave floor constricking and swallowing two freshly captured adult Buffy Flower Bats ( Erophylla sezekorni ). In addition to the successful predation events recorded, similar scenarios have been repeatedly observed by the authors in bat caves across the island, without necessarily confirming successful predation. All of the captured bats were phytophagous phyllostomids (Jamaican Fruit-eating Bats, Artibeus jamaicensis; Cuban Fruit-eating Bats, Brachyphylla nana; Buffy Flower Bats, Erophylla sezekorni, or Cuban Flower Bats, P. poeyi) (Table 3).
Other than the 50 predation events we recorded, most snakes with stomach contents in anthropogenic habitats consumed recognizable domestic animals the night (or a few nights) before they were found. These boas apparently used an active-foraging strategy. Domestic chickens, the most frequently taken prey species in anthropogenic habitats, roost in groups on tree branches, sometimes forming mixed roosting groups with other species (Fig. 15). Hens and chicks sleep on the ground during the first weeks after hatching but, as soon as the wing feathers of the chicks emerge, the mother hen begins to entice the chicks into the trees before sunset. Four snakes were observed preying on chicks hidden under hens while sleeping on the ground; only on one occasion was the snake large enough (>2.0 m SVL) to consume both the hen and eight small chicks (A. Hernández, pers. comm. 2013). In most cases, the snakes managed to slip under the hen and take only some of the chicks. Forty snakes preyed on poultry from roosting groups in trees at night. In these cases, the snakes climbed into the trees, captured, coiled around a roosting bird, and usually dropped to the ground to constrict and swallow their prey. On only a few occasions, when the birds were relatively small, did the snakes initiate ingestion while in the trees. Most predation events involving this kind of prey occurred before midnight. Local farmers and landowners stated that domestic chickens apparently are able to detect the presence of a boa, even in complete darkness, since they usually produce an alarm call. Only one snake, which
apparently had used an ambush-foraging mode was observed preying on a young rooster in an urban area by day (Fig. 16).

Other domestic animals such as caged birds, cats, dogs, and pigs were taken from diverse places in or around buildings and farms while asleep. Although Helmeted Guineafowl (*Numida meleagris*) may be as common as chickens (*Gallus gallus*) on many farms, the incidence of predation on this species was much lower and involved mostly young birds (Tables 1 and 3). In places with a high incidence of boa predation (e.g., Cariblanca; Fig. 2), we observed adult Guineafowl (with a better flight capacity) choosing higher roosting sites (often >8 m above the ground) than those selected by domestic chickens (with a poorer flight capacity), which usually perch at heights of 1–4 m. Also, Guineafowl seemed to show a preference for trees with longer, thicker, vertical trunks without forks, whereas chickens frequently perch on trees or bushes with shorter, thinner, sloping trunks, often with associated rustic wooden ladders, that are more accessible for snakes (Fig. 15).

A common situation involving caged birds (i.e., *Caracara*, *Columba*, *Coturnix*, *Gallus*, *Lonchura*, *Melopsittacus*, *Psittacara*, *Spindalis*; n = 10 snakes) and mammals (i.e., European Rabbits, *Oryctolagus cuniculus*; n = 2 snakes) involved snakes slipping into cages between the bars. However, they subsequently were unable to escape due to the newly acquired prey bulge and were found coiled in the cages the next day.

Over 30 observations involved adult and subadult domestic chickens (>2.0 kg) freshly killed in anthropogenic habitats with the anterior parts of their bodies (head, neck, and shoulders) covered by saliva. Local farmers and landowners recognized this as signs of failed predation attempts by *Ch. angulifer*. In these cases, the boas apparently were strong enough to subdue and kill their prey but lacked sufficient gape size to swallow them. Farmers and landowners in the
countryside coined the term “chupados” (i.e., sucked) for these dead chickens.

Discussion

The usually low population densities and the secretive habits of Cuban Boas render observations of predation in nature very difficult. Nevertheless, a number of historical references address the diet of *Chilabothros angulifer* (Appendix I). These fall into three more-or-less well defined periods: (1) The first decades of conquest (early 16th century) are represented by mostly ambiguous observations of West Indian chroniclers; (2) the mid-19th to mid-20th centuries also are represented by anecdotal observations but these tend to be more accurate since they were generated by naturalists; and (3) the mid-20th century to the present, represented by more accurate and detailed information. A complete gap of information exists between the early 16th and mid-19th centuries.

Most observers noted that *Ch. angulifer* is a predator of native birds, hutias, rats, and domestic animals. Before the mid-20th century, only a handful of papers made reference to specific cases of predation by free-ranging boas. Other works merely listed *Ch. angulifer* among confirmed predators of certain species without quantitative information. These include several popular articles, research project reports, and online publications related to the U.S. Naval Base at Guantánamo Bay that identified the Cuban Boa as the principal predator of hutias and other species (Appendix I).

Extensive studies on the feeding habits of large constrictors are frequently based on data from the literature and stomach contents from museum specimens, often collected over a span of several years (Henderson et al. 1987; Slip and Shine 1988; Fearn et al. 2001; Pizzatto et al. 2009). Dietary studies conducted in specific areas during relatively short periods of time are scarce (Shine and Fitzgerald 1996; Shine et al. 1998; Luissetti et al. 2001; Quick et al. 2005). *Chilabothros angulifer* is poorly represented in Cuban museum collections mostly due to preservation constraints (Rodríguez et al. 2013; Rodríguez-Schettino et al. 2014; L.M. Díaz and A. Fong, in litt., 2020). Moreover, many snakes that became museum specimens were first kept in captivity for extended periods, so the use of museum specimens for extensive dietary studies on this boa is impractical.

**General dietary composition and niche breadth.**— *Chilabothrus angulifer* showed a narrow niche breadth ($B' = 0.319$), consistent with that of a trophic specialist (Schoener 1971; Hurlbert 1978). This resulted in unexpected for a predator that consumes as many as 49 different prey species (Table 1), even after condensing those species into 16 resource categories. When analyzed separately, the trophic niche breadth of *Ch. angulifer* in natural habitats ($B' = 0.321$) was similar to the overall niche breadth, but that of boas associated with anthropogenic habitats was narrower ($B' = 0.135$). The reality that most boas exploited only a few different types of prey and that many of those were taken only sporadically explains the narrow trophic niche of the species. Nonetheless, rather than a trophic specialist, *Ch. angulifer* appears to be an opportunistic predator, capable of adjusting its diet and foraging behavior according to prey availability, abundance, and characteristics of the habitat. Despite the relatively narrow value of niche breadth, we refrain from rejecting the hypothesis that *Ch. angulifer* is a generalist predator for two main reasons: (1) Trophic specialists do not typically consume such a variety of prey species and (2) do not change their diets when in different habitat types (see below), especially when the change of habitat involves moving to human-altered environments (moves that would be of little evolutionary significance).

A general trend of exploiting the most abundant prey in each type of habitat was evident. The few prey species that form a major portion in the diet of *Ch. angulifer* in natural habitats show some degree of gregariousness and may reach high population densities. All of the species of bats exploited by boas form colonies that range from a few hundred to hundreds of thousands of individuals (Silva 1979) (Fig. 17). Hutias may form family groups comprised of as many as 10 individuals (see Silva et al. 2007 for a review). Under favorable conditions, *Ca. pilorides* reach population densities of 35.6–73.6 hutias/ha in inland second-growth forests to 78.0–153.3 hutias/ha in mangrove forests (Comas et al. 1989, 1994; Comas and Berovides 1990; Berovides and Comas 1997a, 1997b; Borroto-Páez and Mancina 2006; see Silva et al. 2007 for a review). House Rats are ubiquitous and also may achieve high population densities. Borroto-Páez et al. (1990) and Borroto-Páez (2011a, 2013) found as many as 14 rats/ha in Cuban sugarcane plantations. Gundlach (1880) also had mentioned the benefits of Cuban Boas reducing the impact of murid rodents in sugarcane plantations.

Although we could not include White-crowned Pigeons (*Patagioenas leucocephala*) in our statistical analyses due to the lack of quantitative data, these birds are highly gregarious and may form nesting colonies of 20,000 to more than 200,000 breeding adults and as many as 300 or more active nests/ha (Vázquez and Nieves 1980; Chamizo et al. 1983; Godínez et al. 1987; Godínez and Vinola 1988; Godínez 1993). The number of chicks per nest is normally two (range: 1–3 chicks). Chicks reach the fledgling stage (>200 g) two weeks after hatching (see Godínez 1993 for a review). *Chilabothrus angulifer* is a known predator of both eggs and chicks (Vázquez and Nieves 1980; Godínez et al. 1987), but certainly preys on adults as well. Local people and hunters have killed boas containing one or more adult pigeons at nesting colonies located about 10 km east of Jagüey Grande, Matanzas Province, and at “La Javira,” about 5.5 km south of Tópes de Collantes, Sancti Spíritus Province (R. Chamizo, in litt. 8.iv.2020). Those nesting colonies were extirpated in recent years due to hunting and habitat loss.
years due to deforestation, agriculture, stockbreeding, chick predation, and construction of roads and tourist facilities (Acosta and Mugica 2019; R. Chamizo, in litt. 20.v.2020). However, other nesting colonies remain active in a few locations in western Cuba, on Isla de la Juventud, and on some cays (Acosta and Mugica 2019), providing opportunities to study predation by *Ch. angulifer*.

Several species of herons in Cuba also form nesting colonies that might contain 800 to 8,000 nests/ha (Denis 2002). Although *Ch. angulifer* has not been reported preying on nesting or adult herons, some evidence suggests that it might exploit this food source as well. On 7 June 2001, a small adult boa (1.4–1.5 m total length) with a stomach bulge was found coiled in the fork of a Black Mangrove (*Avicennia germinans*) at “Wiso Colony” near the Cauto River Delta (Fig. 2) in Granma Province (D. Denis, pers. comm. 2013; for more details about this nesting colony see Denis 2001, 2002, 2006a, 2006b; Denis et al. 2003). Based on bulge size (too small to be an adult heron or a hutia), this boa must have ingested a medium-sized Cattle Egret (*Bubulcus ibis*) chick (>100 g). Cattle Egrets are the most abundant species nesting in the area (D. Denis, pers. comm. 2013; see also Denis 2001, 2002; Denis et al. 2003). However, the identity of the prey was never corroborated. In mangroves in southern Las Tunas Province (Fig. 2), boas with visible stomach bulges occasionally may be observed resting on branches or in hollow trunks close to nesting colonies that include those of various species of herons, Anhingas (*Anhinga anhinga*), and White Ibises (*Eudocimus albus*) (M. Alonso, in litt., 16.vi.2020). Wiley (2003) observed a Puerto Rican Boa (*Ch. inornatus*; 1.67 m SVL) consuming an egg and a hatchling Cattle Egret on a cay in Bahía Montalva, Puerto Rico. Daniel (2002) suggested that Indian Pythons (*Python molurus*) exploit bird concentr-
and Jamaican Boas (*Ch. subflavus*) (Gosse 1851; Rivero 1978; Schwartz and Henderson 1991; Wiley 2003).

Cuban Boas seem to show a high fidelity to sites with abundant and stable food resources. Rodríguez-Cabrera et al. (2016b) recaptured eight marked individuals associated with a hot cave in central Cuba three (n = 2), six (n = 3), nine (n = 1), and 12 (n = 1) months after being marked, and one was recaptured twice (during the 3rd and 9th months). On 23 January 2012, 14 boas were marked in “Erophylla Cave,” northwest of Yaguajay, Sancti Spíritus Province. Four of those boas were recaptured in the same cave on 5 July 2012 (ca. 5.5 months later) and one was recaptured on 14 January 2019 (ca. 7 years later; T.M. Rodríguez-Cabrera, unpubl. data). Three more boas marked in “El Abono Cave” at Cariblanca in January and April 2013 were recaptured in the same cave in December 2017 (ca. 5 years later; T.M. Rodríguez-Cabrera, unpubl. data; Fig. 18). Puente-Rolón and Bird-Picó (2004) observed a similar pattern in cave-associated Puerto Rican Boas. After implanting transmitters in nine boas and tracking them over a 10-month period, they observed that all but one boa remained in the vicinity of the bat cave where they were first captured and visited the cave repeatedly during the study period. No comparable data exist for boas associated with anthropogenic habitats, where the survival rate must be much lower due to human persecution. However, repeated loss of domestic animals of approximately the same size at regular intervals of about 8–10 days was often followed by the detection of a boa (T.M. Rodríguez-Cabrera, pers. obs.). Although such evidence is at best circumstantial, the fact that domestic animals went missing without signs indicative of avian or mammalian predation at intervals that more-or-less coincide with the digestion time of boas certainly is suggestive of boa predation. Those observations also suggest that *Ch. angulifer* might show some degree of fidelity to food-rich anthropogenic habitats.

Consumption of bird eggs by species of *Chilabothrus* seems to occur infrequently (Koenig et al. 2007). Cuban Boas are reported to have consumed eggs of only two species, White-crowned Pigeons (Godínez et al. 1987; Godínez 1993) and Cave Swallows (*Petrochelidon fulva*) (Mancina and Llanes 1997). As noted above, Wiley (2003) reported a medium-sized Puerto Rican Boa consuming a Cattle Egret egg. Ottenwalder (1980) also reported predation on 13 Ring-necked Pheasant (*Phasianus colchicus*) eggs and on an egg (and an adult) of a Hispaniolan Parakeet (*Psittacara chloropterus*) in the previously mentioned free-flight aviary at the National Zoo in Santo Domingo, Dominican Republic. Schwartz and Henderson (1991) recorded consumption of chicks and eggs of domestic chickens by Turks & Caicos Boas. Because eggshells eventually collapse in a snake’s stomach or are expelled after fluid ingestion, the only ways to document egg predation is to encounter a snake in the act or by dissection of dead snakes. Consequently, egg predation could occur more frequently than the few reports suggest. Martins and Oliveira (1998) found two avian eggshells collapsed in the stomach of a juvenile Rainbow Boa (*Epicrates cenchria*) and Ferreto and Sifuentes (2019) observed predation by another Rainbow Boa on two eggs of the ground-nesting Gray Tinamou (*Tinamus tao*).

**Diets in natural and anthropogenic habitats.**—The feeding habits of *Ch. angulifer* changed dramatically from a diet based largely on native mammals and birds in natural habitats to a diet mostly comprised of livestock, pets, and human commensals when in human-altered environments (Table 1; Figs. 5 and 6). Those results support the hypothesis that diets in natural and anthropogenic habitats differ.
These results largely coincide with previous observations on other large constrictors (Slip and Shine 1988; Shine and Slip 1990; Harlow and Shine 1992; Shine and Fitzgerald 1996; Luiselli and Angelici 1998; Luiselli et al. 1998, 2001; Shine et al. 1998; Martins and Oliveira 1999; Fearn et al. 2001; Quick et al. 2005; Pizzatto et al. 2009; Reed and Rodda 2009). Other large species of *Chilabothrus*, especially adult snakes, also seem to follow a similar pattern to that observed in *Ch. angulifer* (Reagan 1984; Henderson et al. 1987; Wiley 2003; Puente-Rolón et al. 2016; see Henderson and Powell 2009 for a review). A comparable change in diet from mostly native mammals in natural habitats to mostly domestic birds in anthropogenic habitats has been reported in some pythons. Australian Carpet Pythons (*Morelia spilota*) are large snakes (to >4.0 m, 11 kg) that consume a considerable number of domestic and commensal birds and mammals when associated with anthropogenic habitats (Shine and Fitzgerald 1996; Fearn et al. 2001). Shine and Fitzgerald (1996) noted that 89% of prey items recorded in the diet of *M. spilota* in a rural area was comprised of commensal and domestic animals. Similarly, in a sample of 97 prey items obtained from 64 pythons associated with urban and suburban habitats, a high proportion consisted of non-native animals, predominantly birds (64%), followed by mammals (35%), with reptiles representing only 1% (Fearn et al. 2001). On the contrary, of 57 prey items identified from gut and fecal contents of Carpet Pythons from a broader geographic context, 49 (86%) were mammals, five (9%) were birds, and three (5%) were reptiles (Slip and Shine 1988). Mammals seem to be the most frequent prey of large constrictors in natural habitats, whereas birds are the most common endothermic prey exploited in human-altered environments. Luiselli et al. (2001) drew similar conclusions from a study of African Rock Pythons (*Python sebae*).

Desmarest's Huitia (Fig. 8) was the prey species most frequently consumed by *Ch. angulifer* in natural habitats. This rodent is the largest native non-volant mammal in Cuba, averaging about 45.0 cm in body length and 4.0 kg in weight (Silva et al. 2007; Borroto-Páez 2011b). The two relatively large species of Cuban huitias (see Silva et al. 2007 for a review) are known prey of Cuban Boas as well, although in considerable smaller numbers (Table 1). Predation by *Ch. angulifer* on huitias was noticed by the first Europeans arriving in the region. Fernández de Oviedo (1535, officially published in 1851) mentioned snakes as large as 7.6–9.1 m in length containing 6–7 “guabiniquinax” (= huitias; see Rodríguez-Cabrera et al. 2016a for a review). Despite a lack of details, Oviedo’s (1851) comment was the first reference to the feeding habits of Cuban Boas and, for that matter, of any identifiable West Indian snake (Rodríguez-Cabrera et al. 2016a). A number of other authors have listed huitias among the prey of this boa (Appendix 1); for example, P.J. Tolson (in Tolson and Henderson 2006 and Henderson and Powell 2009) noted that the feces of every adult boa examined on the U.S. Naval base at Guantánamo Bay contained hair of Desmarest’s Huitias. Our results corroborate previous reports of huitias as prey of *Ch. angulifer* as well as Oviedo’s (1851) description of multiple huitias consumed by a single boa.

In the case of the Cuban Boa preying on an adult Red-tailed Hawk, one of the largest living raptors in Cuba (to 1.74 kg and to 1.41 m wingspan; Crossley et al. 2013), the means by which the hawk was captured is uncertain. The boa might have approached the roosting bird at night by employing a tactic similar to that used by boas in anthropogenic habitats to prey on domestic fowl. However, an alternative explanation could have been a failed predation attempt by the hawk. The Red-tailed Hawk has been widely documented as a snake predator, including in the West Indies (Guthrie 1932; Fitch et al. 1946; Knight and Erickson 1976; Fitch and Bare 1978; Sherrod 1978; Palmer 1988; Santana and Temple 1988; Global Raptors Information Network 2020). Wenner (2012) reported a case in New Mexico (USA) in which a hawk apparently initiated a predation attempt on a Gopher Snake (*Pituophis catenifer*, ca. 1.6 m total length) and instead was captured and constricted by the snake. Similar situations involving Red-tailed Hawks and other raptors, including owls, have been repeatedly observed (Shaw 2017; https://youtu.be.com). Raptors on occasion may attack snakes that they are unable to subdue. A snake exceeding 2.5 m S.V.L, such as the Cuban Boa that consumed the Red-tailed Hawk (P.N. Otero, pers. comm. 2013) certainly does not fall within the range of prey sizes consumed by this raptor (Guthrie 1932; Fitch et al. 1946; Knight and Erickson 1976; Fitch and Bare 1978; Sherrod 1978; Palmer 1988; Santana and Temple 1988; Global Raptors Information Network 2020). However, a small portion of a well camouflaged snake such as the tail might have attracted the hawk’s attention, with fatal consequences for the bird. Another boa (ca. 2.6 m S.V.L) consumed two nestling Turkey Vultures, which it probably found by searching actively. A third boa (ca. 3.0 m S.V.L) escaped from captivity in the Santa Clara Zoo, entered an aviary containing Caracaras and ate an adult. To the best of our knowledge, these are the first records of snakes preying on raptors in the West Indies and probably among the few cases reported in the world (see Guthrie 1932 for a review). Herrera and Aparicio (2019) reported a Central American Boa Constrictor (Boa imperator, ca. 2.2 m total length; referred to as *B. constrictor*, but see Reynolds and Henderson 2018 and references therein) preying on an adult Barn Owl (*Tyto alba*, referred to as *T. furcata*; but see Chesser et al. 2019).

**Ontogenetic shift in diet.**—The frequency of prey types consumed by subadult/small adult and medium-sized adult Cuban Boas differed significantly within and between habitat types. Those results lend support to the hypothesis that an ontogenetic shift in diet occurs. Most medium-sized to large
Boas from smaller to larger endothermic prey (Figs. 6 and 12). Our results suggest that boas approaching 2.0 m SVL in

Boas, particularly caves, shift their diet to hutias, rats, and medium-sized to large birds, probably because these prey are more energetically rewarding (Table 1; Figs. 6 and 12; Rodríguez-Cabrera et al. 2015). Consequently, the presence of a very large boa in or around a cave with a large bat colony does not necessarily mean that the boa is feeding on bats (although see foraging strategy below), especially since other larger and easier-to-capture prey such as hutias, rats, and birds can be associated with caves as well (Linares et al. 2009). Using stable isotopes, Puente-Rolón et al. (2016) found no significant differences in the prey types (i.e., bats and rodents) consumed by cave-associated and forest-foraging Puerto Rican Boas. In fact, their results showed that boas in both habitats acquired most of their energy from introduced murid rodents. The situation in Cuban Boas might be similar. Additionally, large boas, especially pregnant females, might exploit bat caves as resting sites to facilitate thermoregulation, and not necessarily as feeding grounds (Rodríguez-Cabrera et al. 2016b; see also Puente-Rolón and Bird-Picó 2004 and Puente-Rolón 2012 for similar observations of Puerto Rican Boas, Ch. inornatus).

The growth rate of cave-associated boas seems to be low compared to other large constrictors (Smith 1999; Madsen and Shine 2000; Reed and Rodda 2009; Rodríguez-Cabrera et al. 2016b). Recaptured boas that were marked as juveniles (<700 mm SVL) showed an average growth rate of 0.5 mm/day. However, all boas that were marked as mature individuals or approaching sexual maturity (>1.25 m SVL), showed an average growth rate of about 0.07 mm/day (Fig. 18). The growth rates of cave-associated boas with a diet largely based on bats captured on the wing might be low due to energetic constraints (see below). However, Ch. angulifer shows relatively low growth rates even in captivity (Tolson 1992; Tolson and Henderson 1993; Morell et al. 1998; Polo and Moreno 2007). Among large boas, free-ranging Green Anacondas also show relatively low growth rates (Rivas and Corey 2008; Rivas et al. 2016).

Foraging strategy.—We found differences in the frequencies of foraging modes employed by boas in natural habitats and anthropogenic habitats. This lends support to the hypothesis that foraging modes differ between boas in different habitat types. Chilabothris angulifer relied on both sit-and-wait and active foraging modes in natural habitats, depending on the prey species involved and characteristics of the habitat. Boas in natural habitats foraged almost equally frequently by day and night, whereas predation events in anthropogenic habitats occurred almost exclusively at night, generally before midnight, and the snakes very likely found their prey by searching actively. Also, most if not all the boas found with stomach contents in this type of habitat probably captured their prey using an active foraging strategy. The most frequently taken prey species of Ch. angulifer in anthropogenic habitats are
diurnal and aggregate in roosting sites at night or were confined to cages. To access them, the boas necessarily had to search actively for the prey. Also, in anthropogenic habitats the boas are more exposed to human predation during the day. Whether the marked nocturnal activity observed in boas associated with human-altered habitats is a response to the characteristics of the available prey or a recently acquired strategy to avoid human persecution is unknown and deserves further investigation. On only one occasion did a boa capture a domestic chicken by day, and it likely used an ambush strategy (Fig. 16). Similarly, although Carpet Pythons are primarily ambush predators in natural habitats (Slip and Shine 1988; Shine and Fitzgerald 1996; Fearn et al. 2001), Fearn et al. (2001) noted that most Morelia spilota associated with suburban habitats in Australia preyed on domestic animals using an active foraging mode at night. Likewise, African Rock Pythons are primarily diurnal in natural habitats and primarily nocturnal in anthropogenic habitats, with all predation events by Python sebae in suburban habitats in which a time was determined occurring at night (Luiselli et al. 2001). Burmese Pythons (Python bivittatus) introduced in Florida are primarily ambush foragers but may use an active-foraging mode in suburban areas (Reed and Rodda 2009; Reed et al. 2012). Anthropogenic habitats, with their unique structural environments and associated species and threats seem to force large constrictors to adopt a different foraging mode than that most frequently used in natural habitats.

Foraging times of cave-associated Ch. angulifer largely coincided with the exodus of phyllostomid and some mormoopid bats (e.g., Antillean Ghost-faced Bats, Mormoopis blainvillii) that leave the caves later in the evening (see Silva 1979 for activity times of bats; H. Vela, pers. comm. 8.v.2020). This is consistent with the proportion of phyllostomid bats (86%) observed in the diet of Ch. angulifer. When we analyzed the general context of cave-associated species of Chilabothrus (i.e., Ch. angulifer, Ch. inornatus, and Ch. subflavus), we observed a pattern where certain bat groups were better represented than others in the diets of these boas. Of 101 successful predation events by boas on bats, 85% involved phytophagous phyllostomids (in decreasing order of frequency: i.e., A. jamacensis [32.7%; 27–45 g], E. sezekorni [23.8%; 13–21 g], P. poeyi [20.8%; 15–29 g], Leach’s Single Leaf Bat, Monophyllus redmani [3.0%; 8–14 g], Antillean Fruit-eating Bats, Brachyphylla cavernarum [3.0%; 35–50 g], and B. nana [2.0%; 27–41 g]); only a small proportion (5%) was composed of insectivorous bats (Mormoops blainvillii [6–11 g; Mormoopidae] and Pallas’ Mastiff Bats, Molossus molossus [7–15 g; Molossidae]); with unidentified bats accounting for 8% of those taken (Hardy 1957; Sheplan and Schwartz 1974; Rodriguez and Reagan 1984; Rodriguez-Durán 1996; Prior and Gibson 1997; Vareschi and Janetzky 1998; Koenig and Schwartz 2003; Dávalos and Erickson 2004; Miersma 2010; Mancina 2011; Rodriguez-Cabrera et al. 2015; Dinets 2017; this paper). Also, in an additional sample of 117 successful bat captures by Ch. inornatus recorded at “Culebrones Cave” in Puerto Rico, most involved one of four species (in decreasing order of frequency: E. sezekorni; M. blainvillii; Sooty Mustached Bats, Pteronotus quadridens [3–6 g]; M. redmani, and B. cavernarum) (Puente-Rolón and Bird-Picó 2004; A. Puente-Rolón, in litt. 21.v.2020). Insectivorous bats that form large colonies in caves of the region have a relatively low body mass (average ca. 9 g; 3–15 g), large tail membranes, very low wing loading, and a low average aspect ratio, all of which are associated with highly maneuverable flight (Norberg and Rayner 1987; Mancina 2004; Mancina et al. 2012). On the contrary, most West Indian phytophagous phyllostomids are relatively heavy bats (average mass of species in the region ca. 30 g; 8–50 g), with reduced tail membranes, high wing loading, and a low aspect ratio, which confer them with limited maneuverability (Norberg and Rayner 1987; Mancina 2004). This suggests that phyllostomids are more vulnerable to predation by snakes while exiting or entering caves than insectivorous bats. In the case of Ch. angulifer, a more thorough study focused on cave-associated populations might reveal a different composition in the bat species consumed, depending on the relative abundance of the different bat species inhabiting each cave, but we presume phyllostomids will still dominate the diet. In particular, P. poeyi (Fig. 17) and E. sezekorni are the most abundant phyllostomid species in caves where Ch. angulifer forages for bats (Silva 1979). Consistent with our analysis, a compilation by Eshed and Vricradic (2007) showed that more than half of nearly 20 confirmed species of bats taken by Neotropical snakes were phyllostomids. In a sample of 35 bats taken by the colubrid Western Lyre Snake (Trimerorhachis bictiuctus) at “Los Laguitos Cave” in Chiapas, Mexico, one phyllostomid species (the Lesser Long-nosed Bat, Leptonycteris yerbabuenae) represented 46% of all prey items, whereas the remaining 54% was comprised of four insectivorous species (Natalidae, Mormoopidae) (Marinez-Coronel et al. 2009).

Chilabothrus angulifer is the only species in its genus with heat-sensing labial pits at birth (Tolson 1987; Reynolds et al. 2013; Rodriguez-Cabrera et al. 2015). Assuming that they play an important role (infrared detection) when foraging for flying bats in complete darkness is reasonable. However, the foraging behavior of cave-associated Ch. angulifer very much resembles that reported for other species of Chilabothrus that lack heat-sensing labial pits (Rodríguez and Reagan 1984; Rodriguez-Duran 1996; Prior and Gibson 1997; Vareschi and Janetzky 1998; Koenig and Schwartz 2003; Dávalos and Erickson 2004; Puente-Rolón and Bird-Picó 2004). Chilabothrus inornatus and Ch. subflavus hang from two-thirds to three-fourths of their bodies (>75%) with the heads slightly elevated and sometimes may adopt a double...
S-shaped striking posture once bats begin to emerge (at least *Ch. inornatus* also has been observed securing bats in the latter manner; A. Puente-Rolón, in litt. 12.v.2020). *Chilabothrus inornatus* may even move from side to side while hanging (Rodríguez-Durán, 1996; Puente-Rolón and Bird-Picó 2004). *Chilabothrus striatus* also has been observed foraging for bats at cave openings on Hispaniola, but detailed studies of its behavior do not exist (J.A. Ottenwalder in Henderson and Powell 2009). In the case of *Ch. angulifer*, it uses only the anterior third of the body, usually straight but occasionally in a double S-shaped striking posture, either hanging vertically or obliquely down, extended horizontally, or oriented upward (see also Hardy 1957; Mancina 2011), but has never been observed making oscillatory movements with its body. This suggests that the bat-hunting behavior of Puerto Rican Boas, and probably also that of Jamaican boas, is more complex and specialized than that of Cuban Boas.

Snakes are very sensitive to mechanical stimuli (Ford and Burghardt 1993; Lillywhite 2014). The three species of *Chilabothrus* mentioned above seem to use the suspended portions of their bodies as “tactile antennae,” with contact by and collisions with bats serving as triggers for strikes by snakes. The boas we observed did not predict the approach of a flying bat since they never struck unless contacted by a bat, similar to what other authors have described for West Indian boas preying on bats (Rodríguez and Reagan 1984; Rodríguez-Durán 1996; Prior and Gibson 1997; Vareschi and Janetzky 1998; Puente-Rolón and Bird-Picó 2004; Mancina 2011; Dinets 2017). The use of infrared and chemical stimuli by species of *Chilabothrus* might be more important when actively searching for prey or when employing a sit-and-wait foraging strategy when not hunting for flying bats (e.g., roosting bats, rodents, birds; Silva-Taboada and Koopman 1964). Tactile cues seem more useful when foraging for fast-moving prey such as flying bats (see Ford and Burghardt 1993 for a review on integration of sensory information in snakes), but additional studies are required to corroborate this hypothesis.

Very few of the boas observed preying on bats exceeded 2.0 m SVL. The 41 boas collected by Hardy (1957) in a cave at Guanayara, near Trinidad, ranged in length from “four to eight feet” (i.e., 1.22–2.44 m total length; ca. 1.1–2.2 m SVL). Berovides and Carbonell (1998) reported a mean SVL of 1.56 m in 19 individuals measured in “Los Majaes Cave” near Galalón, Pinar del Río Province, with no significant differences between sexes. The estimated total length of nine individuals associated with a sinkhole cave containing a colony of *A. jamaicensis* at “Desembarco del Granma” National Park ranged from 1.1–2.1 m (ca. 1.0–1.9 m SVL; Dinets 2017). The senior author has observed nearly 150 bat-hunting boas across the Cuban Archipelago and none exceeded 2.0 m SVL (mean 1.396 ± 0.287 m SD, 0.51–1.84, n = 147). We suggest that the cost of foraging for bats becomes too high as the boas approach 2.0 m SVL, making bats unprofitable prey (see Arnold 1993 for a review). The boas must drop this kind of prey from their diets just before or at the moment the energy gain from them is zero or negative (i.e., the costs of searching, capturing, ingesting, and digesting prey is higher than the energy gain from that particular prey; Arnold 1993) (Fig. 12). As far as we know, the net energy contained in West Indian bats has not been studied, but certainly they constitute relatively small prey (2–87 g) with a considerable area of skin due to wing and tail membranes (Silva 1979). Flying bats seem to represent low-energy or marginal prey, particularly for larger boas that probably incur larger foraging costs than juveniles (Schoener 1971; Arnold 1993; Koenig and Schwartz 2003). We did not quantify successful capture rates of flying bats by *Ch. angulifer* relative to snake body size, but we have observed many failed capture attempts. Prior and Gibson (1997) recorded >200 unsuccessful and no successful strikes during a hunting session (1 h 45 min, 1.6–2.5 attempts/min) of a juvenile *Ch. subflavus* (950 mm SVL, 320 g) using a similar foraging strategy. In reference to another type of unprofitable prey (i.e., “hard-to-eat prey”), Feder and Arnold (1982) studied the energy costs involved in staged predation events on Red-cheeked Salamanders (*Plethodon jordani*) by Western Terrestrial Gartersnakes (*Thamnophis elegans*). They measured the average energy content of the prey (ca. 2,000 cal), the rate of failed capture attempts (32%), and the costs involved in the various steps of the predation process (Feder and Arnold 1982; Arnold 1993), concluding that even if only 0.1% of the capture attempts are successful, salamanders should still be retained in the snake’s diet. A similar situation might apply to bats in the diet of *Ch. angulifer*, at least until they approach 2.0 m SVL. For example, every individual in a group of nine bat-hunting Cuban Boas studied during an eight-day period in a sinkhole cave succeeded in capturing bats at least twice (maximum four times) during that period (Dinets 2017; in litt. 4.v.2020). Foraging for flying bats should be classified as a special type of sit-and-wait foraging strategy, one in which a considerable amount of time and energy is invested between the phases of encountering and capturing prey (for reviews see Schoener 1971; Arnold 1993). The energetic costs of strikes at flying bats could be equivalent to costs of pursuit in active-foraging strategies (Schoener 1971; Arnold 1993), but additional studies are required to test this assumption. Also, other physical constraints apply. For example, effects of gravity on blood circulation increase as hanging snakes grow larger (Lillywhite and Henderson 1993). The size ranges observed in other species of *Chilabothrus* that exploit bats as food coincide with our observations (Rodríguez and Reagan 1984; Rodríguez-Durán 1996; Prior and Gibson 1997; Vareschi and Janetzky 1998; Koenig and Schwartz 2003; Dávalos and Erickson 2004; Puente-Rolón and Bird-Picó 2004). Consequently, the costs of bat hunting seem to be positively correlated with an
increase in snake size, apparently limiting this resource to boas <2.0 m SVL (see also Koenig and Schwartz 2003; Puente-Rolón 2012). Exceptions, however, apparently exist. In at least one hot cave with very narrow passages and small openings located north of the Sierra del Rosario in western Cuba, Cuban Boas considerably larger than 2.0 m SVL might forage for bats, presumably at a lower energetic cost, by merely opening their mouths in the midst of a dense pack of flying bats exiting the cave (A. Hernández pers. comm. 8.vi.2020; see also Hardy 1957 for relatively large boas associated with a bat cave). Similarly, Angin (2014) observed a Dominica Boa (Boa nebulosa; ca. 3.0 m total length) using a similar strategy to capture a flying Antillean Fruit-eating Bat at a cave entrance in Dominica.

A common occurrence observed in boas in both natural and human-altered habitats was dropping to the ground, (sometimes from heights >4 m) after capturing relatively large prey (e.g., hutias, adult domestic chickens) in trees. Conversely, relatively small prey (e.g., bats, chicks) captured in trees or from a cave roof were consumed while hanging. Dropping to the ground might reduce energetic costs or facilitate constriction when handling large prey.

Conservation implications.—Estrada (1994) commented that boas surviving habitat loss often acclimatize to life in groves and small forested areas near human houses and rural settlements where they prey on domestic animals like poultry. Our analysis supports that statement. To a certain extent, a high tolerance of human disturbance by Ch. angulifer could be seen as a positive trait. However, because boas moving into human-altered habitats seem to make no distinction between wild and domestic prey, a move into anthropogenic environments in search of food and shelter increases the likelihood of conflicts between boas and humans. Once a boa arrives on a farm or in a small village, it must undergo a dramatic shift in diet from mostly native species in natural habitats to almost entirely domestic and human-commensal animals. Rural residents have contradictory attitudes toward boas since they are simultaneously beneficial because they prey on introduced murid rodents and harmful because they also prey on domestic animals. However, negative attitudes usually prevail. Interviews of local people in rural areas of Cuba revealed that the first reaction toward a boa in >90% of these people is to kill it (T.M. Rodríguez-Cabrera, unpubl. data). In most cases, they argued that the justification was to prevent boas from eating their chickens. Whether domestic animals consumed by Ch. angulifer in anthropogenic habitats are livestock (i.e., poultry, pigs, goats, sheep) or pets (i.e., dogs, cats, caged birds), the loss of these animals incurs negative consequences that are either economic, emotional, or both. Therefore, that boas become “nuisance” animals that are exterminated without hesitation by most people in rural areas is not surprising (see also Estrada 1994). For example, Barbour and Ramsden (1919) stated that “The Majá [Ch. angulifer] is very much persecuted by the country folk because of its destruction of domestic fowl, turkeys, and young pigs” (see Appendix I for additional pertinent references). Other large species of Chilabothrus that consume domestic animals also trigger human-wildlife conflicts (see Henderson and Powell 2009 for a review). A number of species of boas and pythons associated with human-altered areas are well known to consume domestic animals, including some of sentimental value such as pet dogs and cats (Shine and Fitzgerald 1996; Murphy and Henderson 1997; Shine et al. 1998; Fearn et al. 2001; Luiselli et al. 2001; Quick et al. 2005; Henderson and Powell 2009; Reed and Rodda 2009).

Ever-increasing habitat loss and ongoing persecution by humans have resulted in Ch. angulifer being listed as Near Threatened on the IUCN Red List of Threatened Species (Day and Tolson 1996). More recently, it was listed in the Red Book of Cuban Vertebrates (Polo and Rodríguez 2012) and included in CITES Appendix II (https://www.cites.org/eng/app/appendices.php). Some national regulations (Ministerio de Justicia 2011) have listed this boa to prevent poaching, but intentional killing remains one of the main threats to this species and is still far from being resolved. Radiotracked Ch. inornatus associated with habitats without concentrated food resources were more likely to move than those associated with food-rich habitats (e.g., bat caves; Puente-Rolón and Bird-Picó 2004). In the case of Ch. angulifer, the combination of habitat loss and decreased sizes of prey populations (e.g., hutias) is increasingly forcing boas into anthropogenic habitats. This species has dominated Cuba’s terrestrial ecosystems for millions of years before the first humans arrived in the region just a few thousand years ago (Rodríguez-Cabrera et al. 2016a; Napolitano et al. 2019; Nägele et al. 2020). Those large individuals that we see today preying on domestic animals are merely exploiting the only available food in the remaining accessible habitats.

Considerations for further studies.—This is the first attempt to integrate and analyze the feeding habits and foraging behavior of free-ranging Cuban Boas. The relative importance of various prey taxa in the diet of Ch. angulifer may be biased as a consequence of two main factors: (1) Very small prey items and prey items in very advanced states of digestion can go unnoticed in the stomachs of some snakes, especially large individuals, and (2) smaller snakes are harder to detect (see Reed and Rodda 2009 for a review). Also, we did not assess possible seasonal and regional differences in prey availability relative to its representation in the diets of boas due to a lack of sufficiently large sample sizes from various localities within reasonable periods of time.

Further studies are required to assess local adaptations for exploiting available trophic resources throughout the year.
(e.g., in bat caves) and any possible sex-specific niche partitioning (females may be twice as heavy as males). The use of different techniques such as scar analysis and radiotelemetry to track snakes could reveal new and important information on the ecology and feeding habits of *Ch. angulifer* (Shine and Fitzgerald 1996; Petersen et al. 2007; Puente-Rolón and Bird-Picó 2004; Wunderle and Mercado 2004; Quick et al. 2005). Also, the use of stable isotopes could provide greater insights into the trophic ecology of this boa and its role in the different ecosystems of the Cuban Archipelago (Rush et al. 2014; Puente-Rolón et al. 2016; Durso and Mullin 2017).

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Appendix I. Summary of the most significant references on the feeding habits of free-ranging Cuban Boas (*Chilabothrus angulifer*). Asterisks (*) mark those included in our analyses.

| Reference | Prey species (number of prey items and growth states) |
|-----------|------------------------------------------------------|
| Fernández de Oviedo (1851) | Stated that as many as 6–7 "guabíniquínax" (= hutias) were frequently found in the stomachs of very large "culebrás ó serper" (= Ch. angulifer). |
| Poey (1866) | Commented on the damage caused by *Ch. angulifer* to henhouses and dovecotes; the largest individuals also may prey on piglets; useful when associated with barns because it controls rats; in natural habitats, it consumes hutias and birds. |
| Gundlach (1875) | Ibid. |
| Rodríguez (1876) | Ibid. |
| Gundlach (1880) | Ibid.; also commented on the benefits of boas in sugarcane plantations because of the elimination of rodents; recounted the folk story of boas capturing hutias in midair while falling to the ground where they end up coiled around their prey. |
| Miller (1904) | Referenced field notes of W. Palmer, who stated that boas were said by the country folk to forage for bats at a cave mouth near Baracoa in eastern Cuba; he did not confirm bat predation. |
| Barbour (1914) | Ibid. |
| Barbour and Ramsden (1919) | Made reference to the strong persecution of boas by humans because they prey on chickens, turkeys, and young pigs; referenced Palmer and Riley, who commented on boas that were said by the country folk to forage for bats at the openings of bat caves near Guanayaj in western Cuba; also referred to observations of V.J. Rodriguez on a boa foraging for bats in a cave near Maisí in eastern Cuba; they did not confirm bat predation. |
| Schwartz and Ogren (1956) | Observed a boa laying in a low and narrow passage connecting with a chamber containing a large colony of Jamaican Fruit-eating Bats (*Artibeus jamaicensis*) in a cave at Guajimico, Cienfuegos Province; they did not confirm bat predation. |
| Hardy (1957) | Reported three boas constricting or swallowing Cuban Flower Bats (*Phyllonycteris poiery*) and another two that regurgitated three and nine bats, respectively (same species), from a hot cave at Guanayara, west of Trinidad in central Cuba*; the snakes he studied in that cave (n = 41) ranged from four to eight feet in total length (i.e., 1.2–2.4 m), but he did not specify the size of the five snakes he found consuming bats; he was the first to confirm bat predation by *Ch. angulifer* and by any species of *Chilabothrus*. |
| Hardy (1963) | Ibid. |
| Silva-Taboada and Koopman (1964) | “The senior author was assured by local farmers that Cuban boas (*Epicrates angulifer*) often crawled up to the foliage [of the 'Jata' palm tree, *Copernicia x vespertilionum*] to eat the bats [Little Goblin Bats, *Mormoops minuta*; Broad-eared Bats, *Nyctinomops latiacaudata*]; the foliage of this palm tree is frequently used as roosting sites by these two species of bats; the authors did not confirm bat predation. |
| Vogel (1965) | Mentioned that *Ch. angulifer* preys on Desmarest’s Hutias (*Capromys pitorides*), bats, bird nests, and domestic fowl. |
| Sheplan and Schwartz (1974) | Reported a boa containing six bats (1 Cuban Fruit-eating Bat, *Brachyphylla nana*, 2 Antillean Ghost-faced Bats, *Mormoops blainvillei*, 2 Cuban Flower Bats, *Phyllonycteris poery*; 1 unidentified bat*; mentioned that the gastrointestinal tracts of other boas examined contained rats (*Rattus* sp.) or domestic chickens (pullets and half-grown adults) but provided no quantitative data on the latter. |
| Buide (1966) | Reported a large boa (3.35 m total length) killed on the Hicacos Peninsula, north of Matanzas Province, which contained a young domestic goat, *Capra hircus* (ca. 7 lbs.*; commented on the abundance of Desmarest’s Hutias (*Capromys pitorides*) in the area as potential prey. |
| Silva (1979) | Same as Hardy (1957) and Silva-Taboada and Koopman (1964); also mentioned finding boas in the foliage of the "Jata" palm tree where two bat species (Little Goblin Bat and Broad-eared Bat) roost and presented a photograph of several boas presumably moving to assume foraging positions in a cave at Guanayara (see Hardy 1957) prior to the bat exodus. |
| Vázquez and Nieves (1980) | Mentioned *Ch. angulifer* as a predator of White-crowned Pigeon (*Patagioenas leucocephala*) chicks in nesting colonies located east of Jagüey Grande, Matanzas Province.* |
| Regalado (1981) | Listed *Ch. angulifer* among what he considered occasionally harmful native species, since it preys on domestic animals. |
### Reference

| Reference | Prey species (number of prey items and growth states) |
|-----------|------------------------------------------------------|
| Buide (1985) | Made reference to hutias, rats, birds, and bats as prey of *Ch. angulifer*. |
| Buide (1986) | Ibid. |
| Godínez et al. (1987) | Mentioned *Ch. angulifer* as a predator of White-crowned Pigeon (*Patagioenas leucocephala*) chicks and eggs in a nesting colony on the Guanahacabibes Peninsula, Pinar del Río Province.* |
| Viña and Armas (1988) | Reported a boa slightly in excess of 1.0 m total length swelling a Giant Trope (*Tropidophis melanosoma*), 0.42 m in total length, tail-first, in Santiago de Cuba Province.* |
| Rams et al. (1989) | Listed *Ch. angulifer* among the potential predators of the Cuban Solenodon (*Solenodon cubanus*). |
| Sampedro and Motañez (1989) | Reported a boa preying on a Cuban Slider (*Trachemys decussata*) nest in the Zapata Swamp, Matanzas Province. |
| Schwartz and Henderson (1991) | Stated that *Ch. angulifer* is a sit-and-wait forager as an adult; diet includes domestic fowl, a number of bat species (probably based on Hardy 1957; Sheplan and Schwartz 1974), and rodents (*Capromys, Rattus*); mentioned anoles and native birds as potential prey. |
| Cruz (1992) | Mentioned *Ch. angulifer* as a predator of hutias in hot caves and briefly described foraging behavior, stating that boas take advantage of concentrations of bats, capturing them on the wing as they emerge from or return to caves. |
| Godínez (1993) | Same as Godínez et al. (1987) (in litt. 4.iv.2020). |
| Tolson and Henderson (1993) | Reported a boa (ca. 1.5 m SVL) constricting a House Rat (*Rattus rattus*) by day in a sewer (in complete darkness) in an old fort on the U.S. Naval Base at Guantánamo Bay (P.J. Tolson in litt. 23.iii.2020); noted that neonates of most other species of *Chilabotrus* prefer lizards as food, whereas neonatal Cuban Boas readily accept small rodents. |
| Estrada (1994) | Commented that those boas that survive for any length of time in human-altered habitats feed mostly on domestic animals such as poultry, for which they are seen as nuisance animals by most country people, who kill them whenever encountered. |
| Silva (1996) | Same as Buide (1985). |
| Mancina and Llanes (1997) | Provided compelling evidence of predation by a boa (1.5 m SVL) containing a Desmarest's Hutia (*Chilabotrus angulifer*); eggs (C.A. Mancina, pers. comm. 19.v.2020). |
| Sampedro (1998) | Same as Sampedro and Motañez (1989); also noted that, when first seen, the boa (ca. 2.0 m total length) had a live hatching turtle in its mouth (in litt. 6.xii.2015). |
| Legón (1998) | Same as Tolson and Henderson (2006). |
| Alberts et al. (2001) | Mentioned *Ch. angulifer* as potential predator of birds, bats, lizards, and hutias (*Capromys pilorides*), emphasizing predation on hutias. |
| Alonso and Rodríguez (2003) | Listed *Ch. angulifer* among the potential predators of cave-dwelling frogs of the genus *Eleutherodactylus*. |
| Chamizo et al. (2003) | Same as Poey (1866) and Buide (1985). |
| Rodríguez and Rodríguez (2003) | Stated that *Ch. angulifer* is persecuted and killed because it occasionally preys on poultry. |
| Holanova and Hribal (2004) | Stated that the smaller Cuban Boas prey on sleeping anoles in a context suggesting that they might be among the predators of the Pinar del Río Cliff Anole (*Anolis bartschi*); this record by J. Hribal was based on a young boa observed at night preying on an undetermined species of anole near a cave in the area of Viñales, Pinar del Río Province (V. Holanova, in litt. 6.iv.2020). |
| Fong et al. (2005) | Same as Rodríguez and Rodríguez (2003). |
| Hernandez and Pimentel (2005) | Reported a boa (1.7 m total length) containing a Desmarest's Hutia (*Capromys pilorides*) (1.9 kg) in the region of “Mil Cumbres,” Pinar del Río Province.* |
| P.J. Tolson in Tolson and Henderson (2006) | Stated that every adult *Ch. angulifer* examined (on the US Naval Base at Guantánamo Bay) contained Desmarest's Hutia (*Capromys pilorides*) hair in its feces. |
| Petersen et al. (2007) | Mentioned *Ch. angulifer* as predator of birds, bats, lizards, and Desmarest's Hutias (*Capromys pilorides*), emphasizing predation on hutias. |
| Wittmer and Lowney (2007) | Mentioned *Ch. angulifer* as a predator of Desmarest's Hutias (*Capromys pilorides*). |
| Tolson and Petersen (2008) | Presented a photograph of a large boa constricting a Desmarest's Hutia (*Capromys pilorides*) at Curco Beach on the U.S. Naval Base at Guantánamo Bay; the predation event was observed in a grassy area early in the morning (P.J. Tolson, in litt. 28.iii.2020). |
| P.J. Tolson in Henderson and Powell (2009) | Same as P.J. Tolson in Tolson and Henderson (2006). |
| Arredondo (2011) | Mentioned *Ch. angulifer* as a potential predator of hutias, solenodons, small sloths, and primates in the past. |
| Borroto-Páez (2011a) | Presented a photograph of a boa (< 2.0 m SVL) constricting a House Rat (*R. rattus*) by day in “La Barca Cave”, Guanahacabibes Peninsula, Pinar del Río Province (in litt. 25.v.2020); also mentioned *Ch. angulifer* among predators of House Mice (*Mus musculus*). |

*(continued)*
| Reference                          | Prey species (number of prey items and growth states)                                                                                                                                                                                                 |
|-----------------------------------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Mancina (2011)                    | Presented a photograph of a boa (< 2.0 m SVL) constricting a Cuban Fruit-eating Bat (*Brachyphylla nana*) by day in “La Barca Cave”, Guanahacabibes Peninsula, Pinar del Río Province (pers. comm. 19.v.2020)*; briefly described foraging behavior in cave-associated *Ch. angulifer*; stated that boas may use two basic foraging modes, either hanging down from rock projections or crevices and taking a position on the ground close to the hot chambers. |
| Amaro (2011)                      | Summarized previous records from the literature.                                                                                                                                                                                                     |
| Polo and Rodríguez (2012)         | Summarized previous records from the literature.                                                                                                                                                                                                     |
| Tolson (2012)                     | Reported entwined skeletons of a juvenile boa and a Green-blotched Giant Anole (*Anolis smallwoodi*) suggesting that the encounter was lethal for both participants.*                                                                                                                                 |
| Segovia et al. (2013)             | Reported a boa (ca. 2.0 m total length) preying upon a nestling Gray Kingbird (*Tyrannus dominicensis*) over one hour before sunrise in the “Siboney-Juticí” Ecological Reserve, Santiago de Cuba Province.* |
| Rodríguez-Cabrera et al. (2015)  | Reported predation on six bats (three juvenile Buffy Flower Bats [*Erophylla sezekorni*] and three adult Cuban Flower Bats [*Phyllonycteris poeyi*]) by four juvenile boas*; one of these boas, the smallest ever reported, had captured an adult Cuban Flower Bat on the wing; three other small boas were observed foraging for bats in caves; four of these boas had visible umbilical scars. |
| Rodríguez-Cabrera et al. (2016a) | Reviewed on the role of *Ch. angulifer* and other large species of *Chilabothrus* as top predators in the West Indies; also presented photographs of a large boa containing a Desmarest’s Hutia (*Capromys pilorides*), which is included in this report as Fig. 8 (left). |
| Dinets (2017)                     | Reported predation on 31 Jamaican Fruit-eating Bats (*Artibeus jamaicensis*) by nine boas (1.1–2.1 m total length, visually estimated) in a sinkhole cave in “Desembarco del Granma” National Park, Granma Province; five boas captured four bats each, three boas got three bats each, and one boa got two bats (in litt. 4.v.2020)*; he observed foraging activity both after sunset and before dawn and noted that boas tended to aggregate during foraging periods and that this apparently increased capture success, which suggested coordinated hunting. |