Effects of Tianeptine on Adult Rats Following Prenatal Stress

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Objective: Exposing a pregnant female to stress during the critical period of embryonic fetal brain development increases the risk of psychiatric disorders in the offspring. The objective of this study was to investigate the effect of antidepressant tianeptine on prenatally stressed (PNS) rats.

Methods: In this study, a repeated variable stress paradigm was applied to pregnant rats during the last week of gestation. To investigate the effects of antidepressant tianeptine on PNS rats, behavioral and protein expression analyses were performed. Forced swim test, open field test, and social interaction test were performed to determine changes in PNS rats compared to non-stressed offspring. Haloperidol was used as a positive control as an antipsychotic drug based on previous studies.

Results: Behavioral changes were restored after treatment with tianeptine or haloperidol. Western blot and immunohistochemical analyses of the prefrontal cortex revealed downregulation of several neurodevelopmental proteins in PNS rats. After treatment with tianeptine or haloperidol, their expression levels were increased.

Conclusion: Downregulation of several proteins in PNS rats might have caused subsequent behavioral changes in PNS rats. After tianeptine or haloperidol treatment, behavioral changes in PNS rats were restored. Therefore, tianeptine might decrease incidence of prenatal stress-related psychiatric disorders such as depression and schizophrenia.

KEY WORDS: Prenatal stress; Laboratory animal model; Tianeptine; Haloperidol; Behavior test; Psychiatric disorder.

INTRODUCTION

Several animal studies have demonstrated that prenatal stress (PNS) such as elevated glucocorticoid level during gestation is associated with biochemical, physiological, and behavioral changes in the offspring, including reduced birth weight, cardiovascular and neuroendocrinological abnormalities, attention dysfunctions, enhanced anxiety-related behaviors, and cognitive deficits.¹⁻¹⁰ Pregnant rats are commonly exposed to stressful manipulations during the third week of pregnancy which is approximately equivalent to the second trimester of human gestation in terms of neural development.²⁻¹⁰ Early life exposure to PNS causes neural developmental deficits with decreased number of hippocampal synapses and neurons as well as changed dendritic length and spine density. These deficits are different from those of non-stressed (NS) controls.¹¹,¹² Increased corticosterone secretion prolonged by PNS stimulation is associated with decreased levels of corticosterone receptors in the brain. On the other hand, the amplitude and duration of stress-induced corticosterone secretion can be reduced by better postnatal handling such as short periodic maternal deprivation and early adoption in adult offspring.¹³,¹⁴ PNS or maternal exposure to glucocorticoids may affect the responsivity of the hypothalamic-pituitary-adrenal (HPA) axis. It can induce cognitive deficits in offspring.²,⁸,¹⁰ Cognitive deficits resulting from disrupted hippocampal anatomy and impaired function of the HPA axis are also observed in patients with schizophrenia.⁹ PNS can also...
result in various gene expression changes in rat brain, including genes associated with neural development, cell differentiation, and neurotransmitter function. We have previously reported that dihydropyrimidinase-like 2 (Dpysl2) and LIM and SH protein 1 (Lasp1) are down-regulated in the prefrontal cortex of PNS rats. We have also found that Dpysl2 gene polymorphisms in human might be associated with the development of schizophrenia.

Cyclic adenosine monophosphate (cAMP) response element-binding protein is a transcription factor with well-established role in learning-related synaptic plasticity. It has been reported that prolonged administration of serotonin and/or noradrenaline reuptake inhibitors can enhance the expression and phosphorylation of cAMP response element-binding protein. Long-term pharmacotherapy can also attenuate stress-induced dendritic reorganization and impairment in hippocampal neurogenesis. In this context, antidepressant tianeptine can improve stress-induced morphological changes in brain areas such as the hippocampus, prefrontal cortex, and amygdala. Tianeptine can also trigger a cascade of neuronal adaptations, including increased phosphorylation of N-methyl-D-aspartate (NMDA) and α-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) among different subtypes of glutamate receptors. Haloperidol is an antipsychotic drug that affects the glutamatergic system. It can change the expression level of two main subunits or neuro-receptors of metabotropic and ionotropic glutamate receptors. Glutamate is the predominant neurotransmitter controlling neuronal excitability and neuropasticity in majority of brain circuits. Glutamatergic dysfunctions have been documented in patients with depression. It has been reported that tianeptine might be able to attenuate impaired neuropasticity by modulating the glutamatergic system in key brain structures, including medial prefrontal cortex, amygdala, and hippocampus. In addition, tianeptine treatment can induce antidepressive-like effects in rats.

These findings led us to hypothesize that tianeptine treatment might affect the pathophysiology of PNS-related psychiatric disorders such as depression and schizophrenia. In the present study, offspring behavioral patterns and changes in protein levels were examined in the prefrontal cortex of PNS rats. The objective of this study was to determine whether changes caused by PNS might be affected by tianeptine treatment.

METHODS

Drugs and Animals
Tianeptine and haloperidol (Sigma Aldrich, St. Louis, MO, USA; Merck KGaA, Darmstadt, Germany) were dissolved in 0.9% NaCl. Separate groups of adolescent rats were treated with tianeptine (10 mg/kg) or haloperidol (1.5 mg/kg) once daily for 21 consecutive days (postnatal day 35 to 56). The duration of experimental manipulation used for rats (P35-P49) was selected because it roughly approximated mid-adolescence in human. Each drug treatment was performed at 10 a.m. every day.

PNS Procedures
Pregnant Sprague-Dawley rats (n=6 per group) were purchased from Central Lab Animal, Inc. (Seoul, Korea). They arrived at the animal facility on day 7 of gestation. These rats were housed under standard conditions under a 12/12-hour light/dark cycle (lights on at 06:30). They were provided free access to food and water. All animal procedures were performed in accordance with guidelines for the care and use of laboratory animals of the United States National Institutes of Health. Animal study protocol used in this study was approved by the Institutional of Animal Care and Use Committee of Soonchunhyang University, Korea (approval No.: SCH16-0058). The PNS rat model was prepared as described previously. PNS exposure was initiated on day 14 of gestation. It included the following: 1) 1-hour restraint in a well-ventilated cylindrical Plexiglas restraint, 2) 6-hour exposure to a cold environment (4°C), 3) overnight fasting, 4) 15 minutes of swimming stress in room-temperature water, 5) reversal of the light-dark cy-

| Table 1. Prenatal stress paradigm |
|----------------------------------|
| Gestation day | Time of day | \( \text{a.m.} \) | \( \text{Mid-day} \) | \( \text{p.m.} \) |
|----------------|-------------|----------|---------------|----------------|
| 14 | Restraint 60 min | Swin 15 min | Restraint 60 min |
| 15 | Cold stress 6 hr | Swin 15 min | Fast overnight |
| 16 | Swin 15 min | Restraint 60 min | Swin 15 min |
| 17 | - | Swin 15 min | Light on overnight |
| 18 | Social stress 12 hr | - | - |
| 19 | Restraint 60 min | Swin 15 min | Restraint 60 min |
| 20 | Cold stress 6 hr | - | - |
| 21 | Swin 15 min | Restraint 60 min | Swin 15 min |
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cle, and/or 6) social stress induced by overcrowded housing conditions during the dark phase of the cycle (Table 1). Pregnant dams used as controls remained in the animal room during gestational days 14 to 21. They were only exposed to normal animal room husbandry procedures. Following birth, rats and their pups were left undisturbed in their cages until weaning on postnatal day 23. At this time, male and female offspring were separated and housed in cages with one or two littermates of the same gender. They were provided free access to rat chow and water. All groups contained litters of five to 12 pups with similar number of males and females. Extremely large or small litters were eliminated. Offspring were weaned 23 days after birth and group-housed. Male offspring were selected and used for further experiments. A total of four experimental groups of adolescents were tested: 1) control group, offspring of unstressed mothers; 2) PNS group, offspring of mothers subjected to stress before parturition; 3) tianeptine treated group, tianeptine was administered to offspring of mothers subjected to PNS; and 4) haloperidol treated group, haloperidol was administered to offspring of mothers subjected to PNS. For each group, 14 male rats were used.

Behavior Test

Modified behavioral tests including social interaction test (SIT), open field test (OFT), and forced swim test (FST), were performed as described previously and summarized below. All behavioral tests were commenced at 9 a.m. Rats were allowed to adapt to test environment for at least 30 minutes before behavior test.

SIT

SIT protocol was adapted from previous studies. Test was conducted in a black Polygal chamber (77×77×25 cm) using rats at 57 days of age. The room in which the chamber was located was darkened during testing. The chamber was illuminated by a single 25 W red light bulb placed approximately 100 cm above the base of the chamber. Social interaction partners were same-sex siblings who resided in the same cage after weaning. Their body weights were approximately the same. Each session lasted 20 minutes. Total duration of social play, the number of interaction, and type of interaction were recorded. Behaviors were scored manually by analyzing video tapes. This was achieved by a time-sampling technique performed by an observer blinded to experimental groups. Based on the video, a rater blinded to treatment conditions scored the behavioral activity as aggressive (fighting [kicking, boxing, and wrestling], aggressive grooming, and biting) or non-aggressive (sniffing, following and grooming partner). Experimental and target rats were not used in this test more than once. The arena was cleaned with 70% ethanol after each trial.

OFT

To assess exploratory activity and reactivity to novel environment, OFT was performed as described in previous studies. Briefly, rats were removed from their home cage on the test day (age, 58 days) and placed individually in the 15×15×20 cm black Polygal open-field start-box linked to another black polygal OFT box (77×77×25 cm) for 5 minutes. No background noise was provided. The experimenter exited the room and behaviors of rats in the OFT box were recorded for 20 minutes. Central boxes entered, line crossings, runs, rears, grooming, cage sniff action, and immobile behavior were scored. For each score, frequency and time were recorded.

FST

FST was performed as described in previous studies. Briefly, rats (age, 59 days) were lowered individually into a cylinder filled with fresh warm tap water (25±2°C). Each rat was removed after 15 minutes of test and wiped with a clean towel to remove excess water before being returned to its home cage. Each rat was placed in the cylinder again for 5 minutes on the following day. Swimming, climbing, and immobility behaviors were recorded with a video camera and evaluated by an observer with a stopwatch. Predominant behaviors were counted every 5 seconds. Test scores for swimming activity (horizontal movement throughout the chamber and crossing quadrants), climbing activity (upward movements along the side of the chamber and jump-ups from the bottom of the chamber), and immobility (no additional activity other than keeping the head above water or tiny whip kicks) were recorded as described previously.

Corticosterone Level

Following behavior tests, adolescent male offspring from each group were moved to the laboratory. On the last behavior test day, experimental subjects were placed
in cylindrical plastic restraint tubes for 60 minutes (5 to 6 animals/group). They were sacrificed by decapitation. Trunk blood was collected immediately into plastic tubes. Blood sample was centrifuged at 13,000 rpm for 10 minutes. Serum was placed in a fresh tube. Brains were removed rapidly from the skull. Each prefrontal cortex was separated, placed in a fresh tube, and frozen in liquid nitrogen. Brain tissue and serum samples were stored at $-80^\circ$C until use. Serum corticosterone levels were determined by immunoassay using a corticosterone ELISA kit (cat no. 501320; Cayman Chemical, Ann Arbor, MI, USA). Assays were conducted according to the manufacturer’s instructions.

**Western Blot**

Prefrontal cortical tissues were lysed in RIPA buffer (EBA-1149; Elpis-Biotech, Inc., Daejeon, Korea) containing protease inhibitors and centrifuged at 17,968 g for 10 minutes at 4°C. To detect Dpysl2 and neurofilament protein, 100 μg of lysed proteins were subjected to 10% and 8% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and transferred to polyvinylidene difluoride membrane (Millipore, Milford, MA, USA). After blocking with 5% skim milk, the polyvinylidene difluoride (PVDF) membranes were probed with anti-Dpysl2 (1:1,000; #9393; Cell Signaling Technology, Danvers, MA, USA), anti-LIM and SH3 protein 1 (Lasp1, 1:2,000, MAB8991; Millipore), anti-neurofilament H (NF-H, 1:10,000; ab8135; Abcam, Cambridge, UK), or anti-β-actin (Actb, 1:1,000; sc-81178; Santa Cruz Biotechnology, Inc., Santa Cruz, CA, USA) antibodies overnight at 4°C, after which they were incubated with peroxidase-conjugated secondary antibody (1:10,000, N4142; Sigma-Aldrich) for 1 hour at room temperature. Immunoreactive bands were detected using enhanced chemiluminescence kit (Elpis-Biotech, Inc.). Quantitative measurements of Dpysl2, Lasp1, NF-H, and Actb proteins were performed using Image J software (NIH, Bethesda, MD, USA).

**Immunohistochemistry**

Rats (4 to 5 animals/group) were deeply anesthetized with ethyl ether and perfused with 4% paraformaldehyde. Rixed brains were frozen and cut into 30-μm sections. Frozen sections from the prefrontal cortex were blocked with normal horse serum (S-2000; Vector Laboratories, Inc., Burlingame, CA, USA), incubated with anti-Dpysl2 (1:1,000, HPA002381; Atlas Antibodies AB, Stockholm, Sweden), anti-NF-H (1:10,000, ab8135; Abcam), or NeuN antibodies (1:100, MAB377; Merck Millipore, Darmstadt, Germany). Sections were incubated with Cy3-conjugated anti-rabbit (1:500; 715-545-151) or anti-mouse (1:800; 111-165-003) secondary antibodies (Jackson ImmunoResearch Laboratories, Inc., West Grove, PA, USA). Fluorescent images were captured using a confocal laser-scanning microscope (FV10-ASW; Olympus, Tokyo, Japan). Images were quantified with Image J software using a protocol described previously with slight modification.

**Statistical Analyses**

All data are expressed as mean±standard error of mean. They were compared using Student’s $t$ test and/or ANOVA. If results of ANOVA indicated significant differences, post-hoc analysis was performed with Tukey’s honest significant difference (HSD) test. All statistical analyses were performed using IBM SPSS Statistics ver. 19 software (IBM Co., Armonk, NY, USA). Statistical significance was considered at $p<0.05$.

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**Table 2. Behavior of non-prenatal stress and prenatal stress and tianeptine treatment of prenatal stress in a forced swim test**

| Forced swim test | NS       | PNS     | HP       | TN       |
|------------------|----------|---------|----------|----------|
| Climbing         | 12.29±3.11 | 5.57±2.72* | 22.71±3.40† | 12.71±3.58 |
| Swimming         | 35.57±1.73 | 32.71±4.20 | 27.86±3.79 | 36.00±2.70 |
| Immobility       | 12.14±2.13 | 21.71±2.58* | 9.00±2.20† | 11.29±1.60† |

Data are presented as mean±standard deviation.

PNS, prenatal stressed offspring; NS, non-prenatal stressed offspring; HP, haloperidol treatment with PNS offspring; TN, tianeptine treatment with PNS offspring.

$p<0.05$ *between NS and PNS, †between PNS and haloperidol, and ‡between PNS and TN.
RESULTS

FST

Significant differences in FST results were detected among the four groups (Table 2). PNS offspring exhibited significantly ($p<0.05$, Tukey’s HSD test) increased immobility behaviors compared to rats in the NS group. Decreased climbing function in PNS offspring was significantly ($p<0.05$, Tukey’s HSD test) increased after haloperidol treatment (Table 2). The increased immobility in PNS offspring was significantly ($p<0.05$, Tukey’s HSD test) decreased after treatment with haloperidol or tianeptine (Table 2).

SIT

The majority of non-aggressive behaviors (sniff, follow, grooming) and aggressive behaviors (fight, aggressive grooming, biting) in SIT (except the number and duration

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Table 3. Behavior of non-prenatal stress and prenatal stress and prenatal stress with tianeptine treatment for adolescence in a social interaction test

| Social interaction test | NS           | PNS          | HP            | TN            |
|-------------------------|--------------|--------------|---------------|---------------|
| Behavior (n)            |              |              |               |               |
| Sniff                   | 66.86±3.53   | 27.86±2.11‡  | 73.00±3.48†   | 64.43±5.08‡   |
| Follow                  | 17.57±1.00   | 1.14±0.46†   | 6.00±1.65‡    | 3.14±0.91†    |
| Grooming the partner    | 8.14±0.86‡   | 8.00±1.23†   | 8.57±1.54‡    | 0.00±0.00‡    |
| Fight                   | 2.86±0.94    | 4.28±0.74‡   | 0.00±0.00‡    | 0.00±0.00‡    |
| Aggressive grooming     | 0.43±0.20‡   | 0.29±0.18‡   | 0.14±0.14‡    | 0.14±0.14‡    |
| Biting                  | 0.43±0.30‡   | 0.29±0.18‡   | 0.14±0.14‡    | 0.14±0.14‡    |
| Duration (sec)          |              |              |               |               |
| Sniff                   | 111.57±6.69  | 85.57±4.88*  | 122.29±3.96†  | 99.43±3.64‡   |
| Follow                  | 26.71±2.09   | 1.71±0.71†   | 7.57±1.78†    | 4.86±1.35†    |
| Grooming the partner    | 19.14±1.45   | 3.00±1.46†   | 31.00±2.84†   | 28.43±2.93†   |
| Fight                   | 10.24±4.61   | 12.14±4.73†  | 0.00±0.00†    | 0.00±0.00†    |
| Aggressive grooming     | 0.71±0.36‡   | 0.71±0.71†   | 0.43±0.43†    | 0.43±0.43†    |
| Biting                  | 0.43±0.30‡   | 0.29±0.18‡   | 0.14±0.14‡    | 0.14±0.14‡    |

Data are presented as mean±standard error of mean.
PNS, prenatal stressed offspring; NS, non-prenatal stressed offspring; HP, haloperidol treatment with PNS offspring; TN, tianeptine treatment with PNS offspring.

Table 4. Behavior of non-prenatal stress and prenatal stress and prenatal stress with tianeptine treatment for adolescence in an open field test

| Open field test       | NS           | PNS          | HP            | TN            |
|-----------------------|--------------|--------------|---------------|---------------|
| Behavior (n)          |              |              |               |               |
| Central entered       | 18.29±2.18   | 7.00±1.36*   | 19.86±1.82‡   | 16.14±1.84‡   |
| Line crossing         | 7.29±1.51    | 1.86±0.59†   | 9.66±0.86†    | 8.14±1.60†    |
| Run                   | 3.57±2.16‡   | 3.86±0.94    | 5.14±1.72†    | 12.57±4.71    |
| Rear                  | 84.86±7.60‡  | 48.86±7.97*  | 87.14±4.47†   | 83.71±6.57†   |
| Grooming              | 11.00±1.83   | 12.29±1.25   | 17.14±1.88†   | 15.29±3.39†   |
| Cage sniff            | 115.00±7.90  | 80.86±15.19  | 114.86±5.01†  | 140.71±6.57†  |
| Immobility            | 0.14±0.14    | 3.00±1.11*   | 0.00±0.00†    | 0.00±0.00†    |
| Duration (sec)        |              |              |               |               |
| Run                   | 3.86±2.27‡   | 5.71±1.63    | 5.57±1.97†    | 14.00±5.45†   |
| Rear                  | 170.57±19.15 | 100.71±13.93*| 174.29±5.81†  | 156.43±21.48† |
| Grooming              | 143.57±34.65 | 182.14±26.27 | 204.29±28.42  | 179.71±46.46  |
| Cage sniff            | 406.29±27.92 | 262.71±43.02*| 391.00±37.06† | 419.14±37.23† |
| Immobility            | 4.14±4.14    | 20.14±11.52  | 0.00±0.00     | 0.00±0.00     |

Data are presented as mean±standard error of mean.
PNS, prenatal stressed offspring; NS, non-prenatal stressed offspring; HP, haloperidol treatment with PNS offspring; TN, tianeptine treatment with PNS offspring.

$p<0.05$ *between NS and PNS, †between PNS and haloperidol, and ‡between PNS and TN.
of fights or the number of biting) were changed significantly \( (p<0.05, \text{Tukey’s HSD test}) \) in the PNS group. However, these changes were restored \( (p<0.05, \text{Tukey’s HSD test}) \) by treatment with haloperidol or tianeptine (Table 3).

OFT

NS and PNS offspring were subjected to OFT for 20 minutes. The PNS group showed significant changes in the number of central entries, line crossings, number and duration of rear, duration of cage sniff, and the number of immobility compared to the NS group. These changes were restored \( (p<0.05, \text{Tukey’s HSD test}) \) after treatment with haloperidol or tianeptine (Table 4). The PNS group showed significantly decreases in the number of central entries, line-crossing, number/duration of rear behaviors, and duration of cage sniff compared to the NS group. However, these scores were increased \( (p<0.05, \text{Tukey’s HSD test}) \) after treatment with haloperidol or tianeptine. On the other hand, the number of immobility behaviors was increased in the PNS group compared to that in the NS group. After treatment with haloperidol or tianeptine, it was reverted \( (p<0.05, \text{Tukey’s HSD test}) \) to control levels (Table 4).

Western Blot and Immunohistochemistry

During development, neurons of the central nervous system express Dpysl2 that which plays a key role in axon

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**Fig. 1.** Western blot analysis of Dpysl2, Lasp1, and NF-H expression in brains of prenatal stress induced rats. (A) Dpysl2, Lasp1, and NF-H protein expression by Western blotting using Actb as an internal control. PNS rats demonstrated decreased expression of Dpysl2, Lasp1, and NF-H in the frontal cortex. Decreased expression levels of Dpysl2 and Lasp1 were recovered by tianeptine treatment. (B) Scatter gram in the graph indicating standard error mean (SEM) \( (p<0.05 \text{ compared to the prenatal stress group in Lasp1 expression}) \). (C) Scatter gram in the graph indicating SEM \( (p<0.05 \text{ compared to prenatal stress group in Dpysl2 expression}) \). (D) Scatter gram in the graph indicating SEM \( (p<0.05 \text{ compared to prenatal stress group in NF-H expression}) \).

NS, non-prenatal stressed offspring; PNS, prenatal stressed offspring; HP, haloperidol treatment offspring; TN, tianeptine treatment offspring.

\*NS and PNS, \**PNS and HP, \***PNS and TN.

Data are presented in mean±SEM; \( p<0.05 \).
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Fig. 2. Immunohistochemical analysis of NF-H expression in brains of prenatal stress induced rats. (A) Confocal microscopic image showing immunofluorescent staining for NF-H (anti-NF-H, red, Cy3) with NeuN in the frontal cortex. Fluorescent staining revealed decreased NF-H in these regions. Scale bar, FC: 50 μm. (B) Scatter gram in the graph indicating standard error of mean (SEM) (p < 0.05 compared to the prenatal stress group in the frontal cortex).

FC, frontal cortex; NS, non-prenatal stressed offspring; PNS, prenatal stressed offspring; HP, haloperidol treatment offspring; TN, tianeptine treatment offspring.

*NS and PNS, **PNS and HP, ***PNS and TN.

Data are presented in mean±SEM; p < 0.05.

formation concentrated at the synaptic sites. It also affects synaptic physiology in axons. In our previous report, Dpysl2 levels have been found to be decreased in the prefrontal cortex of PNS offspring rats. To determine the effect of PNS on neurodevelopmental proteins Lasp1, Dpysl2, and NF-H, we performed Western blots (Fig. 1) and immunohistochemical analyses (Figs. 2, 3) for prefrontal cortex areas from brains of NS, PNS, haloperidol-, or tianeptine-treated offspring. In the prefrontal cortex, levels of these proteins were significantly lower in PNS than those in NS rats (all p < 0.05, Tukey’s HSD test; Fig. 1). Expression levels of the protein were altered by haloperidol or tianeptine treatment. Despite these increases, we were unable to conclude that such increases were directly induced by haloperidol or tianeptine because drug treated NS groups were not included in this study. However, it was evident that haloperidol and tianeptine affected protein expression levels, at least partially. To confirm the expression of these proteins, we also performed immunohistochemical staining using brain tissues. Results revealed significant increases of these proteins (all p < 0.05, Tukey’s HSD test; Fig. 1). Differentially expressed protein levels of Dpysl2 and NF-H were also evident in immunofluorescent-stained brain images among the four groups. Their intensities in immunohistochemical staining showed significant differences (p < 0.05, Tukey’s HSD test; Figs. 2, 3).

Corticosterone Level

Increased stress-induced corticosterone levels were evident in the PNS group compared to those in the NS group. Corticosterone levels in haloperidol or tianeptine treated group were significantly (both p < 0.05, Tukey’s HSD test) decreased to levels in the NS group (Fig. 4).

DISCUSSION

In this study, behavioral and protein expression analy-
Fig. 3. Immunohistochemical analysis of Dpysl2 expression in brains of prenatal stress induced rats. (A) Confocal microscopic image showing immunofluorescent staining for Dpysl2 (anti-Dpysl2, red, Cy3) with NeuN in the frontal cortex. Fluorescent staining revealed decreased Dpysl2 in these regions. Scale bar, FC; 50 μm. (B) Scatter gram in the graph indicating standard error mean (SEM) (p < 0.05 compared to prenatal stress group in the frontal cortex).

NS, non-prenatal stressed offspring; PNS, prenatal stressed offspring; HP, haloperidol treatment offspring; TN, tianeptine treatment offspring.
*NS and PNS, **PNS and HP, ***PNS and TN.
Data are presented in mean±SEM; p < 0.05.

Fig. 4. Percent of corticosterone levels in adolescent male rats. These were analyzed for male offspring of rats exposed to stress during gestation in the last behavior test or male rats derived from mothers without stress during gestation. Corticosterone secretion was induced by holding rats in restraint tubes for 60 minutes. Corticosterone levels were increased in the PNS group but decreased in haloperidol or tianeptine treatment groups.
NS, non-prenatal stressed offspring; PNS, prenatal stressed offspring; HP, haloperidol treatment offspring; TN, tianeptine treatment offspring.
*NS and PNS, **PNS and HP, ***PNS and TN.
Data are presented in mean±standard error mean; p < 0.05.
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and included repeated and variable stress paradigm.\textsuperscript{10} Tianeptine treatment reduced FST immobility time in our study. Furthermore, this treatment restored abnormal number of central entries, line-crossing, number/duration of rear behaviors, duration of cage sniff, and the number of immobility in OFT. Notably, tianeptine also reduced aggressive behavior and increased non-aggressive behavior during SIT in this study.

In this manuscript, we presented therapeutic effects of an antidepressant, tianeptine, and an antipsychotic, haloperidol, in PNS model. Tianeptine works by decreasing extracellular levels of serotonin in rat brain without decreasing serotonin release,\textsuperscript{24} while haloperidol works by blocking the effect of dopamine and increasing its turnover rate.\textsuperscript{48} Although the action mechanisms of the two drugs are different, Western blot and immunohistochemical analyses of the prefrontal cortex revealed that the downregulation of several neurodevelopmental proteins in PNS rats was restored after treatment with tianeptine or haloperidol. This might explain why behavioral changes in PNS rats were restored after treatment with tianeptine or haloperidol. In this study, we focused on the effect of the two drugs on the recovery of neurodevelopmental deficit induced by PNS without focusing on the exact mechanisms involved. Haloperidol was used as a positive control as an antipsychotic drug based on previous studies. Therefore, the exact reason why effects of tianeptine and haloperidol on PNS rats are similar is currently unclear. Several studies have shown that the effect of antipsychotic drug haloperidol on dendritic architecture is at the developmental stage in animal models, corresponding to fetal or fetal plus early childhood stages in human.\textsuperscript{29} Chronic haloperidol treatment during early postnatal life induces persistent alterations in motor and associative-spatial systems.\textsuperscript{49} It also affects the expression levels of central neurotransmitters and receptors in rats. Furthermore, the effect of haloperidol depends on the stage of development when it is administered. It can induce some behavioral activity and neurochemical and molecular alterations.\textsuperscript{49} Contrarily, several studies have reported that this neuroleptic treatment has side effects with higher rates of nonresponse in children and adolescents compared to adults.\textsuperscript{50,51} However, some studies have shown that haloperidol is effective in adolescents and early adults.\textsuperscript{52-54} Previous studies have shown that adolescent male rats are highly sensitive to haloperidiol-induced suppression of locomotor activity, more than adult male rats.\textsuperscript{52-54} These results highlight the importance of early intervention for psychotic symptoms and depression of schizophrenia.\textsuperscript{55,56}

The common feature of PNS models is that critical experimental manipulation can be made to interrupt early brain and central nervous system development, thus inducing significant neurodevelopmental changes, increasing maternal stress hormones, and changing responses to prenatal stressors.\textsuperscript{2,57} Impaired social interaction behavior was observed in prepubertal PNS rats at age of 56 days and in young adult PNS rats. It has been reported that social withdrawal of adolescence period is associated with the first clinical sign of human schizophrenia.\textsuperscript{29,57,58} Therefore, social withdrawal in PNS rats might be related to schizophrenia symptoms, further supporting the relevance of this model to the schizophrenia phenotype. Such diminution in social interaction behavior reflects increased anxiety in PNS rats.\textsuperscript{59} Neurofilaments form part of the axon skeleton and functionally maintain neuronal caliber might also play a role in intracellular transport to axons and dendrites.\textsuperscript{59} It has been found that these subunits are changed in schizophrenia and mood disorder based on distribution and expression studies of pyramidal neurons in prefrontal cortex.\textsuperscript{60}

In the present report, we also investigated the expression levels of two neurodevelopment-related proteins based on previous reports.\textsuperscript{17,61} Our results revealed that application of repeated and variable PNS paradigm during critical periods of fetal brain development could result in changes in the expression of neurodevelopmental proteins, such as neurofilament proteins and Dpysl2. These proteins might have enduring effects on axonal outgrowth and synaptic function in the offspring later during adulthood.\textsuperscript{40,58} Decrease in protein expression of NF-H and Dpysl2 due to PNS was affected by haloperidol or tianeptine treatment. Therefore, these proteins might be involved in behavioral abnormalities of PNS rats directly and/or indirectly.

The present results indicate an additional role of antidepressant tianeptine in the pathogenesis of psychiatric disorders like schizophrenia. Many studies have focused on the effect of antipsychotics such as haloperidol in prenatal period and its side effects in adolescents or early adults. In contrast, we investigated the effect of tianeptine in a model geared toward adolescents and early adults by
analyzing expression levels of neurodevelopment proteins and alterations in behavioral patterns. Further research studies using cellular and animal model systems are required to characterize pharmacological functions and signal transduction pathways of drugs in this schizophrenia-like animal model.

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