Role and mechanism of pancreatic \(\beta\)-cell death in diabetes: The emerging role of autophagy

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ABSTRACT

Pancreatic \(\beta\)-cell failure resulting from decreased \(\beta\)-cell mass or dysfunction is the ultimate step towards most types of diabetes. Even if insulin resistance exists, diabetes does not develop unless pancreatic \(\beta\)-cell function or its adaptation is compromised. Classically, two types of cell death (apoptosis and necrosis) have been studied in the diabetes field. Recently, a third type of cell death (autophagy, sometimes called type 2 programmed cell death in comparison with apoptosis, type 1 programmed cell death) and its pathophysiological role have been recognized and are being investigated. In the present review, we will discuss the role of various types of cell death in the development of type 1 and type 2 diabetes. Specifically, we will briefly cover recent progress regarding the role of autophagy in diabetes, which is becoming a hot topic in diabetes and metabolism. (J Diabetes Invest, doi: 10.1111/j.2040-1124.2010.00054.x, 2010)

KEY WORDS: Diabetes, \(\beta\)-cell death, Autophagy

INTRODUCTION

Pancreatic \(\beta\)-cell failure emanating from \(\beta\)-cell death or dysfunction is the critical precipitating event in the development of diabetes. In type 1 diabetes, \(\beta\)-cell death is the final and critical step in the development of the disease, whereas both \(\beta\)-cell death and dysfunction contribute to \(\beta\)-cell failure in type 2 diabetes. Among diverse types of cell death, \(\beta\)-cell apoptosis (type 1 programmed cell death) plays a major role in the reduction of \(\beta\)-cell mass; however, macroautophagy (here referred to as autophagy), which is also called type 2 programmed cell death, might also play a role in the regulation or execution of \(\beta\)-cell death. Here, we describe the role of apoptosis and necrosis in the \(\beta\)-cell loss of diabetes. Additionally, we review the role of autophagy in pancreatic \(\beta\)-cell physiology and in the development of diabetes shown in recent publications.

\(\beta\)-CELL APOPTOSIS IN TYPE 1 DIABETES

Diverse types of cell death modes contribute to \(\beta\)-cell injury in diabetes. Among them, apoptosis is the most prominent type of cell death in diabetes. In type 1 diabetes, it is generally agreed that apoptosis of pancreatic \(\beta\)-cells is the most important and final step in the progression of the disease. However, it has not been elucidated which molecule(s) are the real culprit(s) in pancreatic \(\beta\)-cell apoptosis. Perforin, Fas ligand (Fasl), tumor necrosis factor (TNF)-\(\alpha\), interleukin (IL)-1, interferon (IFN)-\(\gamma\) and nitric oxide (NO) have been claimed as the effector molecules; however, they, as a single agent, might explain only part of \(\beta\)-cell death in type 1 diabetes. Whereas FasL was initially considered as a strong candidate for the most important death effector, subsequent experiments by others, including the present authors, showed discordant results. Combinations or synergism between IFN-\(\gamma\) and TNF-\(\alpha\) or IL-1 are being revisited as possible death effectors and the molecular mechanism explaining such a synergism has been addressed in several recent papers. However, it is still controversial as to which combination between IFN-\(\gamma\)/TNF-\(\alpha\) vs IFN-\(\gamma\)/IL-1 is dominant in the development of type 1 diabetes in vivo. Signal transducer and activator of transcription-1 (STAT1) is phosphorylated by IFN-\(\gamma\) and modulates signal transduction downstream of both IFN-\(\gamma\)/TNF-\(\alpha\) and IFN-\(\gamma\)/IL-1 combinations. The present authors and others have reported that mice with targeted disruption of STAT1 are resistant to the development of natural type 1 diabetes or that after multiple streptozotocin (STZ) treatment (Figure 1). However, because STAT1 is downstream of both IFN-\(\gamma\)/TNF-\(\alpha\) and IFN-\(\gamma\)/IL-1, the abrogation of type 1 diabetes in STAT1-knockout mice cannot tell which combination is dominant. Although nuclear factor-\(\kappa\)B (NF-\(\kappa\)B) can be activated by both TNF-\(\alpha\) and IL-1, NF-\(\kappa\)B activation might have a different role in \(\beta\)-cell death according to the death signals. NF-\(\kappa\)B activation by TNF-\(\alpha\) has been reported to play an anti-apoptotic role. However, NF-\(\kappa\)B activation could be detrimental to the cells after its activation by other (death) effectors, such as IL-1 (Figure 1). Because of such differences, gene targeting of the NF-\(\kappa\)B-related pathway might be able to tell which cytokine combination is more important in the development of type 1 diabetes. In fact, our data showed that transgenic mice expressing a NF-\(\kappa\)B inhibitor, IkB\(\alpha\)-superrepressor (IkB\(\alpha\)-SR),
specifically in pancreatic β-cells driven by rat insulin promoter, developed accelerated diabetes, suggesting that the net effect of NF-κB in β-cells in vivo is anti-apoptotic. These genetic approaches might be able to provide important information regarding the development of preventive or therapeutic agents in type 1 diabetes. For instance, inhibitors of STAT1 or its upstream Janus kinase 2 (JAK2) signaling could be used to inhibit the development of type 1 diabetes or recurrence of type 1 diabetes after islet transplantation. Downstream molecules of NF-κB activation, such as X-linked inhibitor of apoptosis protein, that have a strong anti-apoptotic activity can be used to inhibit β-cell death in vivo and to prevent the development or recurrence of type 1 diabetes.

β-CELL APOPTOSIS IN TYPE 2 DIABETES

The role of pancreatic β-cell death in type 2 diabetes is less clear. In the preclinical period of type 2 diabetes characterized by insulin resistance, hyperinsulinemia and β-cell hyperplasia develop to compensate for the relative lack of insulin action, which is clearly shown in animal models of type 2 diabetes. For the development of overt type 2 diabetes, relative insulin deficiency is critical in addition to insulin resistance. Insulin deficiency of type 2 diabetes could be a result of decreased insulin release from β-cells and/or decreased β-cell mass. Decreased β-cell mass in full-blown or clinically overt type 2 diabetes has been reported in type 2 diabetes patients. The mechanism of decreased β-cell mass in type 2 diabetes is likely to be a result of pancreatic β-cell apoptosis or death. However, only a few papers reported apoptosis of pancreatic β-cells in type 2 diabetes, which is due to very low probability of detecting ongoing β-cell death in given pancreatic sections of slowly progressive type 2 diabetes.

Even more obscure is the effector molecule(s) in pancreatic β-cell apoptosis of type 2 diabetes. Besides insulin, the concentration of several molecules such as TNF-α or advanced glycation end-products (AGE) is elevated, which could be potentially harmful to β-cell function or β-cell viability in the long term. Amelioration secreted together with insulin, endoplasmic reticulum (ER) stress resulting from prolonged high insulin production or lipid molecules, such as free fatty acids (FFA), might also contribute to the β-cell apoptosis in type 2 diabetes.

FFA also has been reported as a possible effector of pancreatic β-cell dysfunction or death (lipotoxicity). FFA released from the visceral fat of obese subjects is one of the strong culprits in the pathogenesis of insulin resistance that is a prerequisite for the development of type 2 diabetes. The role of FFA in insulin resistance has been well established. Recently, papers suggesting the potential role of FFA in relative insulin deficiency, as well as in insulin resistance, were published. According to such papers, FFA plays a role as an effector of pancreatic β-cell dysfunction or apoptosis (lipapoptosis). However, the detailed molecular and cellular mechanism of lipapoptosis is not elucidated. Previous papers have reported that ceramide produced from FFA plays an important role in lipapoptosis of pancreatic β-cells in type 2 diabetes. In such events, c-Jun N-terminal kinases (JNK) activation by lipid intermediates produced from FFA might contribute to the lipapoptosis of pancreatic β-cells in obesity-induced diabetes. JNK might also be involved in FFA-induced pancreatic β-cell dysfunction or decreased insulin production associated with obesity. Thus, in addition to the well-established role of JNK activation in insulin resistance, JNK might contribute to the β-cell death and β-cell dysfunction in lipid injury or those associated with obesity. Although ceramide has been implicated as a death effector in lipapoptosis of pancreatic β-cells, the role of lipid intermediates produced from FFA other than ceramide cannot be eliminated. For instance, diacylglycerol (DAG), triglyceride (TG) or other metabolites resulting from incomplete β-oxidation of fatty acids have been implicated as the final effector molecules in FFA-induced lipapoptosis of hepatocytes. However, the role for LPC in lipapoptosis of pancreatic β-cells remains to be tested.

β-CELL NECROSIS IN DIABETES

Necrosis of pancreatic β-cells might also contribute to the development of diabetes. In the case of diabetes induced by exogenous agents such as STZ or virus, necrosis might play a major role in the development of disease. In the case of natural
type 1 diabetes, β-cells undergoing secondary necrosis after physiological apoptosis can contribute to the induction of insulitis. During the neonatal period in mice, the β-cell population increases rapidly, and this postnatal expansion of β-cell mass is followed by a transient wave of physiological β-cell apoptosis. Normally, these apoptotic β-cells are engulfed by phagocytes, such as macrophages, thereby preventing the release of potentially cytotoxic or inflammatory cellular contents. However, in non-obese diabetic (NOD) mice with classic type 1 diabetes, the remaining apoptotic β-cells that are not properly cleared due to defective phagocytosis of apoptotic cells might undergo secondary necrosis. Endogenous ligands from the cells undergoing secondary necrosis might be able to stimulate Toll-like receptors 2 (TLR2) on antigen-presenting cells (APC) in the pancreatic lymph nodes (PLN), which might ultimately lead to macrophage activation, dendritic cell (DC) maturation, and priming of naïve diabetogenic T cells. This model is substantiated by the significantly decreased incidence of diabetes and attenuated severity of insulitis in TLR2-knockout NOD mice compared with wild-type NOD mice. In contrast, the development of type 1 diabetes was not inhibited in TLR4-knockout NOD mice. These findings suggest that β-cell necrosis and its sensing through TLR2 might be one of the initial events for the stimulation of APC and the development of type 1 diabetes.

OVERVIEW OF AUTOPHAGY

Autophagy is an intracellular bulk degradation/recycling process that promotes cell survival under nutrient deprivation, but can also execute the non-apoptotic form of programmed cell death under certain conditions. Autophagy is derived from Greek roots: auto, meaning ‘self’, and phag, ‘to eat’. Although the term literally translates to ‘self-eating’, this non-specific degradation process that involves recycling of cellular components plays a crucial role in cellular homeostasis. The ubiquitin-proteasome system (UPS) and the autophagy-lysosome pathway are the two main routes for eukaryotic intracellular protein clearance. Proteasomes predominantly degrade short-lived, ubiquitinated, soluble proteins. In contrast, autophagy is critical in the sequestration of cytoplasm by forming double-membrane vesicles called autophagosomes. The outer membrane of the autophagic vacuoles fuses with lysosomes to form autophagolysosomes, where their contents are degraded by lysosomal enzymes. Because intracellular protein degradation affects many cellular processes, dysregulated or impaired autophagy is likely to be involved in a wide variety of disease states, such as neurodegeneration or cancer.

By degrading intracellular proteins, the autophagy pathway functions in adaptation to cellular starvation. After nutrient stress, autophagy is activated to produce amino acids that can serve as an alternative energy source. Therefore, autophagy can act as a pro-survival mechanism during nutrient starvation. In addition to the aforementioned stimuli, inhibition of the proteosome function has been shown to activate autophagy, suggesting that the two systems can act in a compensatory fashion to one another, although inconsistent results have been reported. Even in the presence of sufficient nutrients or under basal conditions, autophagy is necessary for the sequestration of denatured long-lived proteins and damaged cell organelles as a measure of organelle recycling constitutively or in response to the environmental insults.

Because autophagy is important for the supply of energy sources in cellular starvation, inhibition of autophagy might lead to a shortage of bioenergetic sources, such as NADPH₂ and ATP, which might trigger apoptosis. Thus, the constitutive turnover of proteins and organelles is important for the supply of basic nutrients and the maintenance of organelle function or cell viability. Diverse forms of autophagosome formation have been reported depending on the substrates or organelle being degraded, suggesting that autophagy is a heterogeneous process.

AUTOPHAGY OF PANCREATIC β-CELLS

The main function of pancreatic β-cells is the regulation of glucose homeostasis. In response to elevated glucose, exocytosis of insulin-containing secretory granules occurs in β-cells. To replenish the lost insulin, insulin translation should be stimulated. Even under non-stimulatory glucose concentrations, insulin synthesis continues, showing that constant protein synthesis and degradation occur in pancreatic β-cells. To maintain secretory granules of β-cells in an optimal state, aged β-cell granules should undergo degradation through autophagy or a related phenomenon, crinophagy. Hence, it is possible that defects in the regulation of autophagy might play a role in the pathogenesis of diabetes.

In addition to rapid turnover of insulin granules, pancreatic β-cells have to deal with an overwhelming accumulation of misfolded or aggregated proteins provoked by chronic ER or oxidative stress, such as hyperglycemia, cytokines or FFA. For example, chronic hyperglycemia itself induces the formation of ubiquitinated-protein aggregates in a pancreatic β-cell line (INS-1 832/13), although the nature of those proteins needs to be clarified. These ubiquitinated proteins are cleared by a ubiquitin-proteasome system or autophagy, showing that autophagy can act as a defense mechanism against diabetes-induced cellular damage. Furthermore, autophagy is important in the turnover of dysfunctional mitochondria in β-cells in a process called ‘mitophagy’, and dysregulation of this process might predispose to diabetes.

In addition to its role in the turnover of insulin granules, protein aggregates or dysfunctional organelles, several studies using cell lines and rodent models have shown the importance of autophagy in regulating β-cell survival and function. In animal models, low-level constitutive autophagy in β-cells has been reported in C57BL/6 mice fed a standard diet; however, autophagy was upregulated in mice fed a high-fat diet (HFD), suggesting compensatory autophagy activation to relieve metabolic stress of β-cells. Similarly, β-cells of diabetic db/db mice contained large numbers of autophagosomes, compared with non-diabetic control mice. To prove the role of autophagy in β-cells, two groups
reported mice with β-cell specific deletion of the Atg7 (autophagy-related 7)38,39. β-cell specific Atg7-knockout (Atg7Δβ-cell) mice showed hypoinsulinaemia and hyperglycaemia. Atg7Δβ-cell mice also showed increased apoptosis and decreased proliferation of β-cells leading to decreased β-cell mass (Figure 2a–d). Consistent with the decreased β-cell mass, insulin content in the pancreas was significantly lower in Atg7Δβ-cell mice compared with control mice (Figure 2e). Basal and high glucose-stimulated insulin secretion from viable primary islets of Atg7Δβ-cell mice was reduced compared with control mice (Figure 2f). Furthermore, glucose-induced cytosolic Ca²⁺ transients were defective in primary islet cells from Atg7Δβ-cell mice (Figure 2g), suggesting that β-cell function is impaired, even in apparently healthy β-cells of Atg7Δβ-cell mice, and that autophagy is necessary for the physiological function of β-cells. Electron microscopy (EM) showed more ultrastructural abnormalities, such as swelling of mitochondria and cisternal distension of rough ER and Golgi complex, even in apparently normal-looking β-cells of Atg7Δβ-cell mice38,39. An accumulation of ubiquitinated proteins was also noted. These results suggest that autophagy is necessary to maintain structure, mass and function of pancreatic β-cells. If the adaptive autophagic machinery is impaired, it might predispose to the development of type 2 diabetes. Autophagy might also function as a protective mechanism against insulin resistant states, such as obesity, and alteration of the autophagic process could be a predisposing factor to type 2 diabetes.

Figure 2 | Morphological and functional changes in β-cells of Atg7Δβ-cell mice. (a) Hematoxylin–eosin staining showing variable-sized vacuolated cells in pancreatic islets of Atg7Δβ-cell mice (arrows). Some vacuolated cells stained positive for insulin. (b) Paraformaldehyde–embedded pancreatic sections were subjected to insulin immunohistochemistry, and relative β-cell area was determined by point counting (*P < 0.05 vs Atg7FF and Atg7F/W, Cre+ mice; n = 9 each). (c) Combined insulinoimmunohistochemistry and TUNEL staining was carried out to count apoptotic β-cell number/islet (*P < 0.05 vs Atg7FF and Atg7F/W, Cre+ mice; n = 9 each). Representative apoptotic β-cells are shown on the right (arrow heads). (d) Double immunohistochemistry for insulin and BrdU after intraperitoneal injection of BrdU was carried out to evaluate β-cell proliferation (P < 0.05 by ANOVA, *P < 0.05 vs Atg7FF mice by post-hoc analysis; n = 9 each). (e) Pancreatic insulin was extracted by acid-ethanol, and insulin content was measured by radioimmunoassay (*P < 0.001 vs Atg7FF, Cre+ mice; n = 8 each). (f) Insulin secretion ex vivo. Pancreatic islets from 20-week-old mice (n = 3 each) were incubated in KRB8 containing 3.0 mmol/L glucose for 1 h and then in KRB8 containing 16.7 mmol/L glucose for 1 h. Buffer was collected for insulin radioimmunoassay. Results are representative of two independent experiments carried out in triplicate (*P < 0.05 vs Atg7FF and Atg7F/W, Cre+ mice; *P < 0.05 vs Atg7FF mice). (g) Glucose-induced Ca²⁺ transients in isolated islets (n = 5 each). [Ca²⁺]c was determined in Tyrode solution containing 3.0 or 16.7 mmol/L glucose (36°C). In panels (b)–(f), error bars represent SEM. Reproduced from the original paper in Cell Metabolism 2008; 8: 318–325.
In addition to the mouse model, β-cells in human type 2 diabetes patients also show an accumulation of autophagic vacuoles and autophagosomes, which might represent the adaptive process rather than cause of β-cell loss. However, further studies are warranted to substantiate this point, because a blockade in the autophagic process after the completion of autophagosome formation can lead to the increased number of autophagosomes and increased autophagosome number does not necessarily mean increased autophagic activity. The protective role of autophagy against β-cell lipaoapoptosis in vitro that otherwise could be potentially involved in the decrease of β-cell mass and β-cell failure of type 2 diabetes has also been suggested. However, this interpretation should be based on the cellular or environmental context. For example, a recent report suggested that inhibition of autophagy prolonged pancreatic β-cell survival and delayed cell death in Pdx1-deficient mice.

**AUTOPHAGY AND CELL DEATH**

Autophagy can promote cell survival, but under certain circumstances it might contribute to the execution of cell death. However, the molecular basis underlying its dual role remains unclear. It is also not certain whether or not autophagy is a real effector mechanism of cell death. The visualization of a large-scale accumulation of autophagosomes in areas undergoing cell death has led to the belief that autophagy is a non-apoptotic form of programmed cell death. In many cases, however, it is agreed that this autophagic activity in dying cells does not cause cell death, but it simply occurs as a death process. In fact, activation of autophagy machinery might actually be a compensatory survival mechanism of the cells to provide energy substrates. Furthermore, autophagy enhances the quality control of mitochondria, preventing reactive oxygen species (ROS) production and ROS-mediated DNA damage. By preventing ROS accumulation, autophagy might have a role in the inhibition of the progression of cell death mode to necrosis. Most evidence shows that, at least in cells with intact apoptotic machinery, autophagy is primarily a pro-survival rather than a pro-death mechanism.

However, it can also mediate cellular demise, depending on cell context and specific circumstances. Massive autophagy might result in direct self-destruction of cells, initially suggested as type 2 cell death. Alternatively, autophagy might somehow trigger cell death by switching pro-survival signals to pro-apoptotic signals. Further research will be necessary to fully define the complex interplay between autophagy and apoptosis pathways or even necrosis, because there are increasing research interests to develop autophagy-enhancing drugs in various medical conditions, such as neurodegenerative diseases, diabetes and cancer.

**AUTOPHAGY AND AGE-ASSOCIATED DISEASES**

Because autophagy degrades aggregated proteins and recycles damaged organelles, the decline of autophagic activity might be one of the mechanisms leading to aging and age-associated diseases. In fact, autophagic activity has been reported to diminish with age, and research focusing on inducing autophagy to ameliorate age-induced disease is now emerging. One of the mechanisms of anti-aging effects of autophagy could be the elimination of damaged mitochondria. Increased mitochondrial production of ROS subsequent to the loss of mitochondrial function has been implicated in the aging process, which constitutes the ‘mitochondrial theory of aging.’ Because impaired glucose tolerance and diabetes are frequently associated with aging, dysregulated autophagy might play a role in the development of diabetes in aged subjects or other age-associated diseases, such as neurodegeneration.

**AUTOPHAGY AND INSULIN RESISTANCE**

In addition to its role in pancreatic β-cells, autophagy might have a function in insulin sensitive tissues. A recent paper reported that autophagy regulates lipid content in the liver. Singh et al. reported that starvation induced the recruitment of several autophagy-related proteins to lipid droplets to form autolipophagosomes and induced triglyceride (TG) breakdown. However, the efficiency of ‘lipophagy’ was impaired when intracellular lipid content was chronically increased, such as HFD. In such a case, autophagic clearance was impaired, as shown by reduced association of autophagic vacuoles with lipid droplets in response to starvation in HFD-fed mice. Subsequently, autophagy-mediated breakdown of lipid stores was further diminished in the liver, inducing a vicious circle in which increased fat ingestion was associated with decreased fat removal and excessive lipid deposition in the liver. However, seemingly inconsistent findings have also been published. Although autophagy is likely to be involved in lipid metabolism and lipid injury, further studies will be necessary to understand the role of autophagy in lipid metabolism in more detail.

The role of autophagy in adipose tissue, another insulin target tissue, was also reported. The mutant mice with a targeted deletion of Atg7 in adipose tissue were slim and contained less white adipose tissue mass. Autophagy-deficient white adipocytes were smaller and contained more mitochondria, showing increased rates of β-oxidation and reduced rates of hormone-induced lipolysis. Consistently, the mutant mice had lower fatty acids and their levels decreased at faster rates on insulin stimuli. Those mutant mice were resistant to HFD-induced obesity, suggesting that autophagy is important in adipogenesis.

Another paper reported that autophagy participates in the downregulation of insulin receptors and ER stress-mediated insulin resistance as an adaptive process to ER stress. Conversely, insulin resistance has been reported to suppress autophagy in the liver. A recent paper showed the role of hepatic autophagy in insulin resistance in vivo. In both genetic and dietary models of obesity, suppression of Atg7 was observed in the liver, resulting in defective insulin signaling and elevated ER stress. These metabolic abnormalities were restored by Atg7 expression, suggesting that loss of autophagy is a critical component of defective insulin action seen in obesity. However, it is not clear whether or not decreased expression of...
autophagy-related genes in obesity is a general phenomenon or if autophagic activity could be restored by overexpression of such genes.

CONCLUSION

β-cell death of various types play diverse and important roles in the development of type 1 and type 2 diabetes. Because β-cell death is the ultimate final step in the development of diabetes, current investigation of anti-apoptotic therapies targeting β-cells is an emerging area in diabetes research. These results also show that autophagy is important in the maintenance of β-cell mass, structure and function. Dysregulation of autophagy in insulin producing β-cells or insulin target tissues might predispose to metabolic disorders, such as type 2 diabetes. Exploring measures to modulate autophagic activity to protect against β-cell failure in type 2 diabetes or aging could be a promising area of research. Such approaches might also be applicable to insulin-resistant states, such as obesity and metabolic syndrome.

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