Non-Contact Measurements of Electrocardiogram and Cough-Associated Electromyogram from the Neck Using In-Pillow Common Cloth Electrodes: A Proof-of-Concept Study

Akira Takano, Hiroshi Ishigami and Akinori Ueno

Abstract: Asthma and chronic obstructive pulmonary disease are associated with nocturnal cough and changes in heart rate. In this work, the authors propose a proof-of-concept non-contact system for performing capacitive electrocardiogram (cECG) and cough-associated capacitive electromyogram (cEMG) measurements using cloth electrodes under a pillowcase. Two electrodes were located along with the approximate vector of lead II ECG and were used for both cECG and cEMG measurements. A signature voltage follower was introduced after each electrode to detect biopotentials with amplitudes of approximately 100 µV. A bootstrapping technique and nonlinear electrical component were combined and implemented in the voltage follower to attain a high input impedance and rapid static discharge. The measurement system was evaluated in a laboratory experiment for seven adult males and one female (average age: 22.5 ± 1.3 yr). The accuracy of R-wave detection for 2-min resting periods was 100% in six subjects, with an overall average of 87.5% ± 30.0%. Clearly visible cEMGs were obtained for each cough motion for all subjects, synchronized with reference EMGs from submental muscle. Although there remains room for improvement in practical use, the proposed system is promising for unobtrusive detection of heart rate and cough over a prolonged period of time.

Keywords: cough-associated EMG; textile electrodes; capacitive ECG; non-contact capacitive measurement; bootstrapping technique; heart rate from the neck

1. Introduction

Cough is the most common symptom for which patients seek medical advice [1–3]. Nocturnal cough can occur in asthma [4,5], chronic obstructive pulmonary disease (COPD) [6–8], obstructive sleep apnea [9–11], and gastroesophageal reflux disease (GERD) [12,13], among others. In addition, nocturnal morbidity and mortality are reported for asthma and COPD [14–16]. Nocturnal GERD is considered to be more harmful than daytime GERD [13,17]. As subjective assessments of nocturnal cough frequency are unreliable [18,19], cough detection devices are required for reliable objective assessments and diagnoses. Meanwhile, recordings must be obtained over a prolonged time period for cough detection due to the paroxysmal nature of coughs [20]. Therefore, a non-intrusive approach is desired for cough detection.

The diseases mentioned above are associated not only with cough but also with changes in heart rate (HR) or HR variability [21–25]. Pneumonia presents with both cough and increased HR [26,27]. Hence, a non-obtrusive method for measuring nocturnal cough and HR may be useful for diagnosing and assessing these diseases.

HR is generally calculated from the interval of R-waves (RRI) in electrocardiogram (ECG) measurements obtained with adhesive electrodes. In addition, some methods have been developed for cough detection. Previously reported methods include the use of
microphones [20,28–36], video recordings [37], electromyography (EMG) [38,39], and accelerometers [40]. In routine cough measurements over a prolonged time period, however, audio sensing using a microphone or video recording may undermine the patient’s privacy [40,41]. Prolonged attachment of the adhesive electrodes used in existing ECG and EMG measurement devices can cause skin reactions or inflammation as the measurement period increases. Therefore, new methods for long-duration measurements of HR and cough are desirable to ensure broad acceptance among users.

One possible approach is the use of dry cloth electrodes via clothing for ECG and/or EMG measurements. Some researchers have successfully performed ECG [42,43] or EMG [44,45] measurements in such a way without direct skin contact. This approach can simultaneously reduce the risk of inflammation and privacy invasion. For cough detection, the laryngeal muscles [46] and surrounding muscles can be a target for EMG measurement. For HR detection, the R-wave in ECG has been shown to arise in the EMG signal obtained from hyoid muscles during polysomnography (PSG) [47]. Therefore, both EMG and ECG can potentially be measured from the neck region, neighboring the above described muscles.

Pillows are widely used during sleep almost daily, and the pillowcase is in contact with the posterior neck region over a long time period. Thus, dry cloth electrodes under the pillowcase may detect both cough-associated EMG and ECG in a contact-free manner over a long period of time without invading privacy. Based on these considerations, we developed and tested a prototype non-contact system for measuring cough-associated EMG and ECG from the neck using in-pillow cloth electrodes as a proof-of-concept study.

2. Materials and Methods

2.1. In-Pillow Cloth Electrodes

The cloth electrodes shown in Figure 1 were designed for passively measuring both the electrocardiographic and electromyographic signals from the neck. The measurements are mediated by capacitive couplings of the neck skin (conductor), pillowcase (insulator), and electrode (conductor). Accordingly, the alternating (i.e., time-varying) ionic displacement in the body (ECG and EMG signals) is converted to alternating electronic displacements without direct skin–electrode contact [42–45]. Each electrode plays a dual role of electrode and coupling capacitor at the front-end of the measurement circuit. Measurements by this approach are realized when the coupling capacitances are sufficiently large (meaning that the capacitive reactance of the coupling is sufficiently small), or when the input impedance of a front-end circuit connected to each electrode is sufficiently high to suppress the voltage loss of each coupling. Hereafter, the ECG and EMG signals measured via capacitive couplings are referred to as cECG and cEMG, respectively.

Sensing electrodes, depicted as (−elec.) and (+elec.) in Figure 1a, were arranged diagonally along with an approximate vector of lead II ECG according to the results of the following preliminary experiment, which was conducted on three subjects. In the preliminary experiment, four contact-type electrodes (F-150S, Nihon Kohden) were attached at the corner of the quadrangular posterior surface of the neck of the subject. Two sets of potential difference between the diagonal electrodes were simultaneously measured using a commercially available electrocardiograph (ECG100C, BIOPAC Systems). One set (Figure 1a) was arranged as described by Walter et al. [48] so that the detected electrical vector roughly followed the lead II ECG vector. The other set used the remaining two electrodes for measuring the flipped electrical vector. The amplitudes of ten R-waves of the obtained ECG were measured for each vector and compared among the vectors of each subject. The electrode layout of the prime experiment was that in Figure 1a, because in this layout, the amplitude of the R-waves was significantly larger than that obtained using the flipped layout (as confirmed by significance tests in all subjects). The common use of sensing electrodes for both cECG and cEMG reduced the required number of electrodes and increased the distance and size of the incorporated electrodes.
Figure 1. In-pillow capacitive cloth electrodes. (a) Top-view configuration of the electrodes. (b) Cross-sectional configuration of (a) along the dotted line (1)–(2). The electrodes have a five-layered structure. The top layers consist of a negative electrode (−elec.) and a positive electrode (+elec.).

As shown in Figure 1b, a five-layered structure with a sensing layer, a shield layer and a ground (GND) layer was adopted [49,50]. Each sensing electrode was connected to a front-end voltage follower (FVF) as shown in Figure 2a. The output of the FVF (i.e., sensed voltage) was fed back to the driven shield layer via a 150-Ω resistor. The driven shield layer performs not only as a shield but also as a protector against current leakage from the sensing layer to the GND layer, decreasing the stray capacitance between the sensing and GND layers and increasing the common-mode rejection ratio (CMRR) [51]. Furthermore, the driven shield layer is known to protect the sensing layer from undesirable current caused by moving artifacts and electrostatic disturbances, thereby improving the artifact tolerance [49,50].

To prevent sleep disruption, the electrode is constructed from a thin, soft conductive fabric (CSTK, Kitagawa Industries) and insulating textiles (100% polyester), with a total thickness of 0.45 µm. The electrodes were fixed on a pillow (stuffed with 100% latex cushioning) to be placed at the neck and were covered with a cotton pillowcase (308 µm thick). When the patient lays their head and neck on the pillow, the electrodes form capacitive couplings with the neck via the pillowcase.

2.2. Measurement System

Figure 2a shows a block diagram of the constructed measurement system and Figure 2b illustrates the function of FVF in capacitive biopotential measurements. The source biopotential $e_s$ is divided by the impedance of the coupling capacitance $Z_{cap}$, and the input impedance of FVF $Z_{in,FVF}$. The divided voltage becomes an input (output) of the FVF $v_{in,FVF}$ ($v_{out,FVF}$) as follows:

$$\frac{\dot{Z}_{in,FVF}}{Z_{cap} + \dot{Z}_{in,FVF}} e_s = \frac{1}{\dot{Z}_{cap} / \dot{Z}_{in,FVF} + 1} e_s = v_{in,FVF} (= v_{out,FVF}).$$

(1)

When $\dot{Z}_{in,FVF}$ is sufficiently larger than $\dot{Z}_{cap}$ (i.e., $\dot{Z}_{in,FVF} \gg \dot{Z}_{cap}$), a voltage close to the source voltage is input to the FVF.
The FVF usually employs a simple voltage follower with high input impedance. However, there is a prolonged period during which the voltage input is out-of-range (for example, by static electricity with amplitudes exceeding 100 V). During this time, measurements are not possible. To achieve compatibility between the high input impedance of signal inputs with amplitudes of approximately 100 µV and the low input impedance of out-of-range voltage inputs, a bootstrapped varistor was incorporated in the FVF (see Figure 2c). The bootstrapping component supports a high input impedance when the input voltage is lower than the varistor threshold. When the input voltage exceeds the threshold, the input impedance of the varistor drops and the discharge time is reduced [52,53].

Subsequent circuits for measuring the cECG and cEMG signals consisted of an instrumentation amplifier, filters, and an inverting amplifier. The passband and gain were set to 0.5–100 Hz and 60.0 dB, respectively, for the cECG measurement and 40–1000 Hz and 72.0 dB, respectively, for the cEMG measurement. The cEMG component below 40 Hz was attenuated to suppress R-waves in the cECG.

2.3. Experimental Methods

All experimental procedures were approved by the Human Life Ethics Committee of Tokyo Denki University. All subjects provided informed consent prior to participation in our experiments. Eight healthy subjects (seven males, one female) participated in the experiment, as listed in Table 1. Subjects with long hair were asked to expose the skin on their neck. All subjects were instructed to lay their head on the pillow so that the electrodes were positioned beneath bare neck skin, as shown in Figure 2a. Prior to the experiment, subjects practiced the action of “coughing twice” with an interval such that the corresponding submental EMG (EMG_ref) trains could be distinguished into two groups.
Then, the subjects remained in a resting state in the supine position until the noise in the cEMG signal was attenuated.

Table 1. Subject information and experimental conditions.

| Subject | Weight (kg) | Height (m) | BMI (kg/m²) | Age (yr) | Temp (°C) | RH (%) | VH (g/m³) |
|---------|-------------|------------|-------------|----------|-----------|--------|-----------|
| A       | 74          | 1.61       | 28.5        | 23       | 23.5      | 36.0   | 7.62      |
| B       | 60          | 1.75       | 19.6        | 23       | 24.0      | 28.0   | 6.10      |
| C       | 70          | 1.63       | 26.3        | 23       | 23.8      | 33.0   | 7.11      |
| D       | 48          | 1.53       | 20.5        | 20       | 23.8      | 43.8   | 9.44      |
| E       | 57          | 1.66       | 20.7        | 24       | 21.8      | 28.0   | 5.38      |
| F       | 90          | 1.65       | 33.1        | 23       | 23.6      | 35.0   | 9.34      |
| G       | 64          | 1.61       | 24.7        | 23       | 25.2      | 22.0   | 5.13      |
| H       | 77          | 1.80       | 23.8        | 21       | 23.5      | 22.0   | 4.66      |

Mean ± SD | 67.5 ± 13.1 | 1.7 ± 0.1 | 24.6 ± 4.6 | 22.5 ± 1.3 | 23.7 ± 0.9 | 31.0 ± 7.5 | 6.85 ± 1.85 |

BMI: body mass index, Temp: temperature, RH: relative humidity, VH: volumetric humidity.

The experimental protocol is shown in Figure 3. After the first rest period, the subjects were instructed to “cough twice” seven times as they trained. The time stamp for each “two-cough” cycle was recorded for reference. When a natural cough occurred after the last (i.e., seventh) two-cough cycle, the initiation of the second rest period was postponed to the end of the natural cough. cECG and cEMG signals were recorded with the proposed system throughout the entire experiment. As a reference ECG signal (ECG\text{ref}), the lead II ECG was simultaneously measured from the chest with a commercially available telemetry system (BN-RSPEC, BIOPAC Systems) and contact-type electrodes (F-150S, Nihon Kohden). The EMG\text{ref} signal was simultaneously measured from the submental by reference to PSG [47] with a commercially available EMG amplifier (BA1104M, TEAC Instruments), telemetry system (TU-4, TEAC Instruments), and the same type of electrodes. Measured cECG and cEMG signals, as well as reference ECG\text{ref} and EMG\text{ref} signals, were digitized at 10 kHz with 16-bit resolution between −10 and +10 V by a commercially available A/D converter (BIOPAC Systems, MP150) and recorded on a personal computer.

Figure 3. Protocol of experiment in this study.

2.4. Analytical Methods

2.4.1. Analysis of ECG Signals

The following analysis was applied to a 2-min segment before the first two-cough cycle. Moving averaging (20-ms time window), bandpass filtering (IIR, 10–20 Hz), and differential processing (10-ms interval) were applied to both ECG\text{ref} and cECG segments as preprocessing. Five assumed R-waves were selected in a random manner from each preprocessed signal, and their mean amplitude was calculated. For R-wave detection, 80% of each mean amplitude was used as the threshold through the segment. The RRI was calculated from two consecutively detected R-waves. If the RRI of a cECG signal was within ±10 ms of the RRI of ECG\text{ref}, the latter R-wave of the cECG was deemed as correctly detected. The sensitivity (P_{SNS}), accuracy (P_{ACC}), and positive predictive value (P_{PPV}) of the detected R-waves in cECG measurements were calculated by (2), (3), and (4), respectively:
\[ P_{SNS} = \frac{N_{TP}}{N_{TP} + N_{FN}} \times 100, \quad (2) \]
\[ P_{ACC} = \frac{N_{TP} + N_{TN}}{N_{TP} + N_{TN} + N_{FP} + N_{FN}} \times 100, \quad (3) \]

and
\[ P_{PPV} = \frac{N_{TP}}{N_{TP} + N_{FP}} \times 100, \quad (4) \]

where \( N_{TP} \), \( N_{FP} \), and \( N_{FN} \) are the numbers of R-waves correctly detected, falsely detected, and undetected, respectively. As there are no countable valid events corresponding to \( N_{TN} \), \( N_{TN} \) was set at zero when calculating the accuracy [54].

2.4.2. Analysis of EMG Signals

To verify that cough-associated cEMG measured from the posterior neck was synchronized with the EMG\(_{\text{ref}}\) obtained from submental muscle, the following analysis was performed. First, bandpass filtering (IIR, 40–500 Hz) was applied to both EMG\(_{\text{ref}}\) and cEMG signals as preprocessing. The analysis segment of each two-cough cycle in EMG\(_{\text{ref}}\) and cEMG was determined as shown in Figure 4. For each two-cough cycle in the EMG\(_{\text{ref}}\) signal, the segment (i.e., beginning and ending times) and its middle point were identified with reference to the recorded time stamp of the two-cough cycle, and the duration of the two-cough segment was measured (step 1). The first and last (seventh) two-cough segments were excluded for stability purposes, and the mean duration (MD) of the remaining segments was calculated for each subject (step 2). Extended segments with a duration 1.2-fold longer than the MD were selected for discrete-time Fourier transform (DFT) analysis (step 3). Hereafter, the extended segment with a common duration is called the coughing segment. Coughing segments in the cEMG signal corresponding to those in the EMG\(_{\text{ref}}\) signal were also selected for each subject. A DFT was applied to each coughing segment in the EMG\(_{\text{ref}}\) and cEMG data. Five stable resting segments with the same duration as the coughing segment were selected from the 2-min first rest period of the EMG\(_{\text{ref}}\) and cEMG signals, and were also analyzed by DFT. In the selection of the resting segment, any segments with visible EMG firings due to swallowing or inhalation were excluded.

![Figure 4. Flowchart of selecting the coughing segment in the EMG\(_{\text{ref}}\) and cEMG data for the following discrete-time Fourier transform (DFT) analysis.](image-url)
The total spectral amplitude (TSA) from 40 to 500 Hz was computed in each DFT spectrum obtained from a segment. The TSAs were averaged over five segments of resting and five segments of coughing in the EMG_{ref} and cEMG signals of each subject. The mean TSAs between resting and coughing in the EMG_{ref} and cEMG signals were then compared for each subject, and the comparisons were quantified in a significance test. Based on the normality results, the EMG_{ref} and cEMG signals were evaluated by a paired t-test and a Wilcoxon signed-rank test, respectively.

3. Results
3.1. Simultaneous Measurements of cECG and Cough-Associated cEMG

Figure 5a shows example recordings of ECG_{ref}, cECG, EMG_{ref}, and cEMG, including the fifth and sixth two-cough segments obtained with the reference or proposed systems from subject B. Additionally, filtered and differentiated cECG signals are also depicted. Clearly visible periodic R-waves were observed in the cECG signal during the stable period, which were synchronized with those of the ECG_{ref} signal, as shown in Figure 5b. Although R-waves were unclear during and after the two-cough cycle due to fluctuations in the signal baseline, the timings of R-waves could be detected by applying a bandpass filter and differentiation to the cECG signal (see the third stage in Figure 5a). Similarly, most of the R-wave timings after the two-cough cycle were detected in all subjects except subject E. Synchronized R-waves similar to those in this figure were confirmed in all subjects except subject E. For cEMG signals, EMG firings during the two-cough period were synchronized with those in the simultaneously measured EMG_{ref} signal, as shown in Figure 5c. These synchronized firings were observed for all two-cough cycles in all subjects. Firings considered to be associated with inhalation and swallowing were also confirmed before and/or after the two-cough cycles in all subjects except subjects D and H.

Figure 5. Simultaneous measurements of subject B with the proposed or commercial system obtained during a two-cough experiment. (a) Recordings of ECG_{ref}, cECG, differentiated cECG, EMG_{ref}, and cEMG. (b) Enlarged ECG_{ref} and cECG waveforms and (c) enlarged EMG_{ref} and cEMG signals corresponding to the dotted rectangular sections in (a).
3.2. Evaluation of cECG and cEMG Signals Measured in the Proposed System

3.2.1. Evaluation of cECG Signals

The $P_{SNS}$, $P_{ACC}$, and $P_{PPV}$ values of R-waves in the cECG signals for each subject are given in Table 2, with averages of 90.7%, 87.5%, and 89.0%, respectively. Aside from those for subjects D and E, the $P_{SNS}$, $P_{ACC}$, and $P_{PPV}$ values were all 100%. In subject E, the R-wave amplitudes were considerably smaller than those of the other subjects, as shown in Figure 6. Consequently, approximately 85% of the waves were not detected by the current detection algorithm, resulting in a low $P_{SNS}$ (14.3%). Moreover, the number of falsely detected ($N_{FP} = 154$) and undetected ($N_{FN} = 79$) waves was substantially higher than that of correctly detected waves ($N_{TP} = 39$), because many T-waves had a larger amplitude than the R-waves and were falsely detected as R-waves (see Figure 6).

Table 2. Sensitivity ($P_{SNS}$) (%), accuracy ($P_{ACC}$) (%), and positive predictive value ($P_{PPV}$) (%) of R waves.

| Subject | $P_{SNS}$ (%) | $P_{ACC}$ (%) | $P_{PPV}$ (%) | $N_{TP} + N_{FN}$ |
|---------|--------------|--------------|--------------|------------------|
| A       | 100          | 100          | 100          | 163              |
| B       | 100          | 100          | 100          | 174              |
| C       | 100          | 100          | 100          | 172              |
| D       | 92.5         | 85.4         | 91.8         | 133              |
| E       | 33.1         | 14.3         | 20.2         | 118              |
| F       | 100          | 100          | 100          | 141              |
| G       | 100          | 100          | 100          | 122              |
| H       | 100          | 100          | 100          | 135              |

Mean ± SD 90.7 ± 23.4 87.5 ± 30.0 89.0 ± 27.9 145 ± 22

Figure 6. Simultaneous recordings of ECG$_{ref}$, cECG and filtered and differentiated cECG of subject E obtained with the commercial system (first stage) or the proposed system (second and third stages) during the 2-min segment before the first two-cough experiment. Inverted triangles marks in the third stage show the detected bottoms of the R-waves.

3.2.2. Evaluation of cEMG Signals

Figure 7 shows DFT spectra of the analyzed segments in the EMG$_{ref}$ and cEMG signals depicted in Figure 5c. When comparing the target frequency components (40–500 Hz) of EMG$_{ref}$ and cEMG, the spectral amplitudes for the coughing segment of both signals were higher than those of the resting segment. This increase in spectral amplitude for the coughing segment was observed in the cEMG signal for all subjects.
Figure 7. Comparison of frequency–amplitude spectra between coughing and resting segments for (a) EMG \(_{\text{ref}}\) and (b) cEMG data shown in Figure 5c. The red line corresponds to the spectra for coughing segments, while the blue line corresponds to that for resting segments. (Subject #B).

Figure 8a compares the TSAs of the resting and coughing segments, respectively, in the EMG\(_{\text{ref}}\) and cEMG signals of each subject. Consistent with the spectral amplitudes of each subject, the mean TSA values were higher during coughing than during resting in both the cEMG and EMG\(_{\text{ref}}\) signals of each subject. Figure 8b compares the group average of the mean TSAs over all subjects during resting and coughing in the EMG\(_{\text{ref}}\) and cEMG signals. As evidenced in the figure, the mean TSA was significantly higher in the coughing segments than in the resting segments, in both the EMG\(_{\text{ref}}\) signals (\(p < 0.01\)) and cEMG signals (\(p < 0.01\)).

Figure 8. Comparison of mean total spectral amplitudes (TSAs) between resting and coughing in the EMG\(_{\text{ref}}\) and cEMG signals of (a) each subject, and (b) the group of all subjects.

As the mean TSAs were higher in the coughing segments of EMG\(_{\text{ref}}\) in all subjects (Figure 8a) and the difference was significant (Figure 8b), we confirmed that the segments segregated the periods including cough-associated muscle activities. Moreover, the mean TSAs were significantly increased in the coughing segments during the same periods of cEMG (Figure 8a,b), indicating that the proposed system detected cough-associated muscle activities via the pillowcase holding the posterior neck.

4. Discussions

4.1. Advantages of the Adopted Electrode Configuration and Novel FVF

As shown in Figure 5, the proposed system succeeded in detecting both cECG and cough-associated cEMG signals with amplitudes of approximately 100 \(\mu\)V for subject B from the neck via a pillowcase in an environment with a low humidity of 6.10 g/m\(^3\). Similar results were obtained from seven of eight subjects, and the average R-wave de-
tection accuracy exceeded 85%, as shown in Table 2. In the cEMG signal, an increase in spectral amplitude from 40 to 500 Hz was observed for the recording segment with cough action in all eight subjects. The mean volumetric humidity during the experiment was 6.85 ± 1.85 g/m², and the respective values of relative humidity were lower than 40% for seven of eight subjects. The successful detection of both ECG and EMG signals from the neck is attributable to the following two points: (I) the total number of sensing electrodes was reduced to two by combining the electrode and the FVF for detecting both signals, and (II) the residual space allowed us to arrange the electrodes diagonally along with the approximate vector of the lead II ECG measurement. As the typical electrical axis of the heart for R-waves varies between 30° and 60° with respect to the horizontal axis when frontally measured on the chest [48], the typical axis is a good fit for our diagonal arrangement of posterior neck measurements.

The successful detection of signals with amplitudes of approximately 100 µV via cloth with a thickness of 308 µm is attributed to the following three points: (a) an FVF with ultra-high input impedance was introduced to suppress the voltage loss at the electrode–skin coupling, (b) no ground electrode was used to secure the area for the two sensing electrodes in the limited neck space, and (c) a pillow with an indentation in the neck region was adopted to maintain coupling between the electrodes and the neck. Regarding point (a), the input impedance $Z_{in, bt}$ of the proposed FVF shown in Figure 2c, is expressed as follows [53]:

$$Z_{in, bt} = R_1 + R_{by} + \frac{R_1 R_{by}}{R_3}.$$  

(5)

Assuming that the varistor used in this study (TDK, AVR-M1608C080MTAAB) has a resistance $R_{by}$ of 250 MΩ when the voltage applied to the varistor $v_{vrst}$ is less than the threshold voltage (8 V), $Z_{in, bt}$ is approximately 2.5 TΩ according to Equation (4) for the resistances $R_1$ (99 MΩ) and $R_3$ (10 kΩ) shown in Figure 2c. Furthermore, because the input impedance of the OP amplifier used in this study (Texas Instruments, OPA129) is 10 TΩ according to the data sheet, the input impedance $Z_{in, syn}$ of the analog front-end circuit synthesized in parallel with the bootstrapping component is calculated as approximately 2.0 TΩ. Meanwhile, the coupling capacitance is calculated as 57.5 pF based on $C = \varepsilon S/d$ with a dielectric constant of $\varepsilon_r = 8.85 \times 10^{-12}$ F/m, a coupling area of $S = 2.00 \times 10^{-3}$ m², and a coupling distance of $d = 3.08 \times 10^{-4}$ m. Assuming that the dominant frequency band of the ECG R-wave is 10–20 Hz, the capacitive reactance of the coupling $X_C = d/(2\pi f \varepsilon S)$ is 277–138 MΩ. Therefore, a sufficiently large $Z_{in, syn} >> X_C$ is achieved by the FVF used in this study. Moreover, both points (b) and (c) contributed to the decreased coupling reactance $X_C$ associated with an increased coupling area $S$ and decreased coupling distance $d$. As a result, the voltage loss at the coupling was considered to be suppressed.

The successful detection of both ECG and EMG signals in a low-humidity environment is attributable to the following three points: (i) the adoption of an FVF with a voltage-dependent input impedance, (ii) the adoption of five-layered electrodes with rapid static discharge characteristics [50], and (iii) the adoption of a pillow with an indentation in the neck. Regarding point (i), the varistor has a nonlinear resistance that can be approximated by the following equation:

$$R_{by} \approx \begin{cases} 
R_{by} f - V_{thr} < v_{vrst} < +V_{thr}, \\
0, \text{else}
\end{cases}$$  

(6)

where $v_{vrst}$ is the voltage applied to the varistor and $V_{thr}$ is the threshold voltage. It has been reported that the bootstrapping component, including the varistor shown in Figure 2c, has a voltage-dependent impedance, as given by Equation (7) [53]:

$$Z_{in, bt} = R_1 + R_{by} + \frac{R_1 R_{by}}{R_3} \approx \begin{cases} 
\frac{R_1}{R_3} R_{by}, & \text{if } -V_{thr} < v_{vrst} < +V_{thr} \\
R_1, & \text{else}
\end{cases}$$  

(7)
Because \( v_{\text{vrst}} = \frac{R_h}{R_h + R_3} v_{\text{in}} \) \( Z_{\text{in, bt}} \) has a low impedance \( R_1 \) when \( v_{\text{in}} \) exceeds a threshold \( \pm V_{\text{thr}} (1 + \frac{R_3}{R_h}) \). As the time constant of the measurement system is the product of the input impedance and coupling capacitance of the measuring site, a smaller decrease in the input impedance of the FVF corresponds to a shorter time constant. Consequently, the discharge duration for static electricity on the body or clothing, which commonly develops under low-humidity conditions, becomes shorter, leading to a stable measurement.

As for point (ii), it has been shown that five-layered (i.e., doubly shielded) electrodes have superior discharge characteristics over single- and three-layered electrodes when a 5000-V electrostatic discharge is applied to the electrodes in an electrostatic discharge immunity test (IEC61000-4-2) [50]. Five-layered electrodes are reported to mitigate the motion artifacts caused by an impact hammer [49]. Therefore, it was considered that the electrode configuration mitigated the artifacts induced by cough-associated body movements in the present study, and R-waves were detected after the two-cough cycle (see Figure 5a). Concerning point (iii), the rapid coupling between the electrodes and the neck caused by the indentation may reduce the occurrence of friction between the skin and the pillowcase.

4.2. Comparison of Heart Rate Variability between the Proposed and Commercial Systems

Heart rate variability (HRV) analysis of the ECG\textsubscript{ref} and cECG data was performed on six subjects with a \( P_{\text{ACC}} \) of 100\% (Table 2). Timing data of the R-waves detected in the ECG analysis of 2.4.1 were analyzed by the “HRV analysis” function in commercially available software (AcqKnowledge4.1, BIOPAC Systems). Spectral power densities (PSDs) of very low frequency (VLF, DC–0.04 Hz), low frequency (LF, 0.04–0.15 Hz), high frequency (HF, 0.15–0.40 Hz), and very high frequency (VHF, 0.40–3.00 Hz) were respectively calculated, and are summarized in Table 3.

| Subject | VLF \( \times 10^{-4} \) (s\(^2\)/Hz) | Error Rate (%) | LF \( \times 10^{-4} \) (s\(^2\)/Hz) | Error Rate (%) | HF \( \times 10^{-4} \) (s\(^2\)/Hz) | Error Rate (%) | VHF \( \times 10^{-4} \) (s\(^2\)/Hz) | Error Rate (%) |
|---------|--------------------------------|----------------|--------------------------------|----------------|--------------------------------|----------------|--------------------------------|----------------|
| ECG\textsubscript{ref} | cECG | ECG\textsubscript{ref} | cECG | ECG\textsubscript{ref} | cECG | ECG\textsubscript{ref} | cECG | ECG\textsubscript{ref} | cECG |
| A | 8.9283 | 8.9286 | 0.004 | 62.607 | 62.615 | 0.014 | 16.102 | 16.098 | 0.02 | 8.539 | 8.531 | 0.09 |
| B | 8.0537 | 8.0543 | 0.008 | 54.646 | 54.645 | 0.002 | 14.659 | 14.657 | 0.02 | 7.804 | 7.811 | 0.10 |
| C | 8.1603 | 8.1624 | 0.026 | 56.258 | 56.273 | 0.027 | 15.446 | 15.435 | 0.08 | 7.714 | 7.747 | 0.43 |
| F | 11.9863 | 11.986 | 0.002 | 82.874 | 82.879 | 0.006 | 23.857 | 23.815 | 0.17 | 11.395 | 11.415 | 0.17 |
| G | 15.3784 | 15.378 | 0.000 | 105.311 | 105.247 | 0.061 | 29.195 | 29.141 | 0.19 | 15.340 | 15.299 | 0.27 |
| H | 13.2398 | 13.244 | 0.028 | 88.198 | 88.259 | 0.070 | 23.870 | 23.837 | 0.14 | 12.350 | 12.333 | 0.14 |
| Mean | 10.9578 | 10.9589 | 0.011 | 74.982 | 74.986 | 0.030 | 20.521 | 20.497 | 0.10 | 10.524 | 10.523 | 0.20 |

As evidenced in Table 3, the maximum PSD error rate between cECG and ECG\textsubscript{ref} was 0.43\%. The mean error rate in each band was less or equal to 0.2\%. Therefore, the HRV values were measured accurately in all six subjects during the resting period. The next step is validation of the system in long-duration measurements.

4.3. Challenges to Be Tackled

As this study was pursued as a proof-of-concept, the method has limited applicability to real scenarios. We note that the accuracy of R-wave detection in subject E was below 15\% (see Table 2), and the cECG recording in Figure 6 revealed a more distorted waveform, with the R-wave having a smaller amplitude than the T-wave. Because the frequency components of R-waves are generally larger than those of T-waves, the observed distortion (i.e., large attenuation at higher frequencies) is attributable to stray capacitance between the lead wire of each sensing electrode and the circuit ground, as reported in [43]. Nakamura et al. recently reported on a novel analog front-end circuit that exhibited characteristics of stray capacitance reduction. Therefore, the substitution of our FVF for this analog front-end
A signature analog front-end circuit would mitigate the distorted attenuation observed in the cECG waveform. The recording in Figure 6 also implies that the small amplitude of the original R-wave in subject E caused the low detection ratio and that noise reduction can improve the R-wave detection accuracy. So-called driven shield ground (DSG) is known to suppress common-mode noise such as power line noise [55] in capacitive biopotential measurements. Thus, the introduction of a DSG may enhance the detection accuracy for subjects with small R-wave amplitudes. A DSG also suppresses common-mode components in motion artifacts [55]. Therefore, introducing a DSG might further improve the detection of R-waves during or after the cough-associated body movements.

In addition to the above described hardware approaches, the application of a more sophisticated algorithm (e.g., publicly available toolboxes [56]) instead of the current threshold approach may enhance the detectability of R-waves. For patients with a persistent cough, the total number of coughs at night can exceed 100 [28]. To apply the proposed system to these patients, an algorithm is needed to count the coughs in a recorded signal. However, the proposed system detected cEMG signals associated not only with coughs but also with inhalation and/or swallowing in the experiment, as shown in Figure 9. Therefore, an algorithm for distinguishing among coughs, inhalation, and swallowing would be important for clinical applications. Aside from electromyographic interference, mechanical vibrations from speech or sound did not affect the outputs from the electrodes, as evidenced by the time series waveforms and their DFT spectra (data not shown).

Figure 9. Recordings of EMG_ref and cEMG including a two-cough cycle for subjects E and C. In the cEMG signals, firings considered to be associated with inhalation (a) and swallowing (b) were also confirmed by the two-cough procedure.

The configuration and dimensions of the current electrode narrow the applicable subjects and situations. The subjects include infants and bedridden elderly with thin hair who do not roll over. To widen the acceptance of the device, the measurements of cECG and cEMG in lateral postures through thick hair should be attained in a wide age range of the subjects.

5. Conclusions

In this study, the authors developed a proof-of-concept non-contact system for performing cECG and cough-associated cEMG measurements using in-pillow electrodes. The sensing electrodes were arranged under a pillow cover so that the vector between the electrodes was similar to that of lead II ECG. The same sensing electrodes were used for the cEMG measurement. To detect biopotentials with amplitudes of approximately 100 µV, a signature analog front-end circuit was introduced in the measurement system. A
The bootstrapping technique and a nonlinear electrical component (varistor) were combined to attain high input impedance and rapid static discharge characteristics. cECG and cEMG signals were measured with a 308-µm-thick pillow cover on the fabricated electrodes for seven adult males and one female (average age: 22.5 ± 1.3 yr) during resting or coughing. The accuracy of R-wave detection for a 2-min resting period was 100% in six subjects, with an average of 87.5% ± 30.0% for all subjects. Clearly visible cEMGs were measured for all 14 cough motions in each subject, corresponding to the reference EMGs obtained from submental muscle. The power spectra of non-contact cEMGs during coughing showed an increase in 40–500 Hz compared with that obtained during resting. Although its practical applicability could be improved, the proposed method is a potentially useful method for routine measurement for ECG and cough-associated EMG through cervical pillows.

Future work includes the introduction of a negative capacitance circuit and a bandpass amplifier at the analog front-end to improve the accuracy of R-wave detection for low signal-to-noise ratios, such as that observed in subject E. In addition, a more sophisticated algorithm is to be introduced for the improvement of the accuracy of R-wave detection.

**Author Contributions:** Conceptualization, A.T. and A.U.; methodology, A.T. and A.U.; software, A.T.; validation, A.T. and A.U.; formal analysis, A.T.; investigation, A.T. and A.U.; resources, A.T., H.I., and A.U.; data curation, A.T. and A.U.; writing—original draft preparation, A.T.; writing—review and editing, A.U.; visualization, A.T. and A.U.; supervision, A.U.; project administration, A.U.; funding acquisition, A.U. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was supported in part by SECOM Science and Technology Foundation and in part by Private University Research Branding Project supported by MEXT, Japan.

**Institutional Review Board Statement:** The study was conducted according to the guidelines of the Declaration of Helsinki, and approved by the Human Life Ethics Committee of Tokyo Denki University.

**Informed Consent Statement:** Informed consent was obtained from all subjects involved in the study.

**Acknowledgments:** We would like to thank reviewers for their helpful comments.

**Conflicts of Interest:** The authors declare no conflict of interest.

**References**

1. Schappert, S.M. National ambulatory medical care survey: 1991 summary. Vital Health Stat13 1993, 116, 1–20.
2. Irwin, R.S.; Boulet, L.P.; Cloutier, M.M.; Fuller, R.; Gold, P.M.; Hoffstein, V.; Ing, A.J.; McCool, F.D.; O’Byrne, P.; Poe, R.H.; et al. Managing cough as a defense mechanism and as a symptom. A consensus panel report of the American College of Chest Physicians. Chest 1998, 114, 1335–1815. [CrossRef] [PubMed]
3. Kono, S. Prolusion. In Clinical Practice Guidelines: Cough, 2nd ed.; The Japanese Respiratory Society: Tokyo, Japan, 2012; pp. 2–3, (In Japanese). Available online: https://www.jrs.or.jp/uploads/uploads/files/photos/1048.pdf (accessed on 4 November 2020).
4. Brooke, A.M.; Lambert, P.C.; Burton, P.R.; Clarke, C.; Luyt, D.K.; Simpson, H. Night cough in a population-based sample of children: Characteristics, relations and associations with measures of asthma severity. Eur. Respir. J. 1996, 9, 65–71. [CrossRef] [PubMed]
5. Bentur, L.; Beck, R.; Shinawi, M.; Naveh, T.; Gavriely, N. Wheeze monitoring in children for assessment of nocturnal asthma and response to therapy. Eur. Respir. J. 2003, 21, 621–626. [CrossRef]
6. Omachi, T.A.; Blanc, P.D.; Claman, D.M.; Chen, H.; Yelin, E.H.; Julian, L.; Katz, P.P. Disturbed sleep among COPD patients is longitudinally associated with mortality and adverse COPD outcomes. Sleep Med. 2012, 13, 476–483. [CrossRef]
7. Stephenson, J.J.; Cai, Q.; Mocarski, M.; Tan, H.; Doshi, J.A.; Sullivan, S.D. Impact and factors associated with nighttime and early morning symptoms among patients with chronic obstructive pulmonary disease. Int. J. Chron. Obstruct. Pulmon. Dis. 2015, 10, 577–586. [CrossRef]
8. Ding, B.; Small, M.; Bergström, G.; Holmgren, U. A cross-sectional survey of night-time symptoms and impact of sleep disturbance on symptoms and health status in patients with COPD. Int. J. Chron. Obstruct. Pulmon. Dis. 2017, 12, 589–599. [CrossRef]
9. Birring, S.S.; Ing, A.J.; Chan, K.; Cossa, G.; Matos, S.; Morgan, M.D.L.; Padvor, I.D. Obstructive sleep apnoea: A cause of chronic cough. Cough 2007, 3, 7. [CrossRef]
10. Chan, K.K.Y.; Ing, A.J.; Laks, L.; Cossa, G.; Rogers, P.; Birring, S.S. Chronic cough in patients with sleep-disordered breathing. Eur. Respir. J. 2010, 35, 368–372. [CrossRef]
11. Chan, K.; Ing, A.; Birring, S.S. Cough in obstructive sleep apnoea. *Pulm. Pharmacol. Ther.* 2015, 35, 129–131. [CrossRef]

12. Gaudie, G.S. Pulmonary manifestations of gastrooesophageal reflux disease. *Ann. Thorac. Med.* 2009, 4, 115–123. [CrossRef] [PubMed]

13. Emilsson, Ö.I.; Bengtsson, A.; Franklin, K.A.; Torén, K.; Benadiksdóttir, B.; Farkhoooy, A.; Weyler, J.; Dom, S.; Backer, W.D.; Gislason, T.; et al. Nocturnal gastro-oesophageal reflux, asthma and symptoms of OSA: A longitudinal, general population study. *Eur. Respir. J.* 2013, 41, 1347–1354. [CrossRef] [PubMed]

14. Cochrane, G.M.; Clark, J.H. A survey of asthma mortality in patients between ages 35 and 64 in the Greater London hospitals in 1971. *Thorax* 1975, 30, 300–305. [CrossRef] [PubMed]

15. McNicholas, W.T.; Fitzgerald, M.X. Nocturnal deaths among patients with chronic bronchitis and emphysema. *Br. Med. J. (Clin. Res. Ed.)* 1984, 289, 878. [CrossRef] [PubMed]

16. Owens, R.L.; Maceira, M.M.; Teodorescu, M. The overlaps of asthma or COPD with OSA: A focused review. *Respirology* 2017, 22, 1073–1083. [CrossRef]

17. Orr, W.C. Review article: Sleep-related gastro-oesophageal reflux as a distinct clinical entity. *Aliment. Pharmacol. Ther.* 2010, 31, 47–56. [CrossRef] [PubMed]

18. Archer, L.N.J.; Simpson, H. Night cough counts and diary card scores in asthma. *Arch. Dis. Child.* 1985, 60, 473–474. [CrossRef]

19. Falconer, A.; Oldman, C.; Helms, P. Poor agreement between reported and recorded nocturnal cough in asthma. *Pediatr. Pulmonol.* 1993, 15, 209–211. [CrossRef]

20. Barry, S.J.; Dane, A.D.; Morice, A.H.; Walmsley, A.D. The automatic recognition and counting of cough. *Cough* 2006, 2, 8. [CrossRef]

21. Garrett, C.S.; Seidler, A.; McKibben, A.; McAlpine, L.E.; Gordon, D. Spectral analysis of heart rate variability in bronchial asthma. *Clin. Auton. Res.* 1992, 2, 105–111. [CrossRef]

22. Jensen, M.T.; Marott, J.L.; Lange, P.; Vestbo, J.; Schnohr, P.; Nielsen, O.W.; Jensen, J.S.; Jensen, G.B. Resting heart rate is a predictor of mortality in COPD. *Eur. Respir. J.* 2013, 42, 341–349. [CrossRef] [PubMed]

23. Gula, L.J.; Krahn, A.D.; Skanes, A.; Ferguson, K.A.; George, C.; Yee, R.; Klein, G.J. Heart rate variability in obstructive sleep apnea: A prospective study and frequency domain analysis. *Ann. Noninvasive Electrocardiol.* 2003, 8, 144–149. [CrossRef] [PubMed]

24. Lado, M.J.; Vila, X.A.; Rodriguez-Liñares, L.; Méndez, A.J.; Olivieri, D.N.; Félix, P. Detecting sleep apnea by heart rate variability analysis: Assessing the validity of databases and algorithms. *J. Med. Syst.* 2011, 35, 473–481. [CrossRef] [PubMed]

25. Lee, Y.C.; Wang, H.P.; Lin, L.Y.; Lee, B.C.; Chiu, H.M.; Wu, M.S.; Chen, M.F.; Lin, J.T. Heart rate variability in patients with different manifestations of gastroesophageal reflux disease. *Auton. Neurosci.* 2004, 116, 39–45. [CrossRef]

26. Halm, E.A.; Teirstein, A.S. Management of community-acquired pneumonia. *N. Engl. J. Med.* 2002, 347, 2039–2045. [CrossRef]

27. Ewig, S.; Bauer, T.; Hasper, E.; Pizzulli, L.; Kubini, R.; Lüderitz, B. Prognostic analysis and predictive rule for outcome of hospital-treated community-acquired pneumonia. *Eur. Respir. J.* 1995, 8, 392–397. [CrossRef]

28. Hsu, J.Y.; Stone, R.A.; Sinclair, R.B.L.; Worsdell, M.; Bussel, C.M.; Chung, K.F. Coughing frequency in patients with persistent cough: Assessment using a 24 h ambulatory recorder. *Eur. Respir. J.* 1994, 7, 1246–1253. [CrossRef]

29. Coyle, M.A.; Keenan, D.B.; Henderson, L.S.; Watkins, M.L.; Haumann, B.K.; Mayleben, D.W.; Wilson, M.G. Evaluation of an ambulatory system for the quantification of cough frequency in patients with chronic obstructive pulmonary disease. *Cough* 2005, 1, 3. [CrossRef]

30. Birring, S.S.; Fleming, T.; Matos, S.; Raj, A.A.; Evans, D.H.; Pavord, I.D. The Leicester Cough Monitor: Preliminary validation of an automated cough detection system in chronic cough. *Eur. Respir. J.* 2008, 31, 1013–1018. [CrossRef]

31. Vizel, E.; Yigla, M.; Goryachev, Y.; Dekel, E.; Felis, V.; Levi, H.; Kroin, I.; Godfrey, S.; Gavirol, N. Validation of an ambulatory cough detection and counting application using voluntary cough under different conditions. *Cough* 2010, 6, 3. [CrossRef]

32. McGuinness, K.; Holt, K.; Dockry, R.; Smith, J. Validation of the vitaioAK™ 24 h ambulatory cough monitor. *Thorax* 2012, 67, A131. [CrossRef]

33. Fischer, P.; Sohrabi, K.; Ashraf, O.; Schneider, H.; Weissflog, A.; Gross, V. Home monitoring of nocturnal cough in COPD patients. *Curr. Dir. Biomed. Eng.* 2017, 3, 269–271. [CrossRef]

34. Umayahara, Y.; Soh, Z.; Sekikawa, K.; Kawae, T.; Otsuka, A.; Tsuji, T. Clinical significance of cough peak flow and its non-contact measurement viac sounds: A narrative review. *Appl. Sci.* 2020, 10, 2782. [CrossRef]

35. Umayahara, Y.; Soh, Z.; Sekikawa, K.; Kawae, T.; Otsuka, A.; Tsuji, T. Estimation of Cough Peak Flow Using Cough Sounds. *Sensors* 2018, 18, 2381. [CrossRef]

36. Umayahara, Y.; Soh, Z.; Sekikawa, K.; Kawae, T.; Otsuka, A.; Tsuji, T. A Mobile Cough Strength Evaluation Device Using Cough Sounds. *Sensors* 2018, 18, 3810. [CrossRef]

37. Corrigan, D.L.; Paton, J.Y. Pilot study of objective cough monitoring in infants. *Pediatr. Pulmonol.* 2003, 35, 350–357. [CrossRef]

38. Chang, A.B.; Newman, R.G.; Phelan, P.D.; Robertson, C.F. A new use for an old Holter monitor: An ambulatory cough meter. *Eur. Respir. J.* 1997, 10, 1637–1639. [CrossRef]

39. Ranjani, S.; Santhiya, V.; Jayapreetha, A. A real time cough monitor for classification of various pulmonary diseases. In Proceedings of the Third International Conference on Emerging Applications of Information Technology, Kolkata, India, 30 November–1 December 2012; pp. 102–105.

40. Paul, I.M.; Wai, K.; Jewell, S.J.; Shaffer, M.L.; Varadan, V.V. Evaluation of a new self-contained, ambulatory, objective cough monitor. *Cough* 2006, 2, 7. [CrossRef]
41. Liaqat, D.; Nemati, E.; Rahman, M.; Kuang, J. A method for preserving privacy during audio recordings by filtering speech. In Proceedings of the IEEE Life Sciences Conference (LSC), Sydney, Australia, 13–15 December 2017; pp. 79–82.
42. Kato, T.; Ueno, A.; Kataoka, S.; Hoshino, H.; Ishiyama, Y. An application of capacitive electrode for Detecting Electrocardiogram of Neonates and Infants. In Proceedings of the 28th Annual International conference of IEEE EMBS, New York, NY, USA, 31 August–3 September 2006; pp. 916–919.
43. Ueno, A.; Akabane, Y.; Kato, T.; Hoshino, H.; Kataoka, S.; Ishiyama, Y. Capacitive sensing of electrocardiographic potential through cloth from the dorsal surface of the body in a supine position: A preliminary study. IEEE Trans. Biomed. Eng. 2007, 54, 759–766. [CrossRef]
44. Ueno, A.; Yamaguchi, T.; Iida, T.; Fukuoka, Y.; Uchikawa, Y.; Noshiro, M. Feasibility of capacitive sensing of surface electromyographic potential through cloth. Sens. Mater. 2012, 24, 335–346.
45. Borelli, G.; Bonnet, J.J.; Hernandez, Y.R.; Matsuda, K.; Damerau, J. Spectral-distance-based detection of EMG activity from capacitive measurements. IEEE Sens. J. 2018, 18, 8502–8509. [CrossRef]
46. Fontana, G.A.; Lavorini, F. Cough motor mechanisms. Respir. Physiol. Neurobiol. 2006, 152, 266–281. [CrossRef] [PubMed]
47. Yoshizumi, N.; Hori, T. Methods for sleep study. In Handbook of Sleep Science and Sleep Medicine; Japanese Society of Sleep Research, Ed.; Asakura Publishing Co.: Tokyo, Japan, 1994; pp. 452–453. (In Japanese)
48. Walter, M.; Eilebrecht, B.; Wartzek, T.; Leonhardt, S. The smart car seat: Personalized monitoring of vital signs in automotive applications. Pers. Ubiquit. Comput. 2011, 15, 707–715. [CrossRef]
49. Fukuyama, Y.; Suzuki, R.; Takayama, S.; Ueno, A. Multi-layered fabric electrode for movement artifact reduction in capacitive ECG Measurement. In Proceedings of the 35th Annual International Conference of IEEE (EMBS), Osaka, Japan, 3–7 July 2013; pp. 555–558.
50. Kato, Y.; Motohara, S.; Omuta, T.; Azran, A.; Ueno, A. Artifact tolerance test for capacitive wearable chest-belt electrocardiograph-effect of electrode configuration. IEEJ Trans. Electron. Inf. Syst. 2017, 137, 607–615. (In Japanese)
51. Bednar, T.; Babusiak, B.; Smondrk, M.; Cap, I.; Borik, S. The impact of active electrode guard layer in capacitive measurements of bio-signals. Measurement 2020, 171, 108740. [CrossRef]
52. Ishigami, H.; Tanaka, S.; Ueno, A. Non-contact measurements of blink-associated electrooculogram using in-pillow cloth electrodes: Potential application as a communication aid. In Proceedings of the 42nd Annual International Conference of the IEEE Engineering in Medicine & Biology Society (EMBC), Montreal, QC, Canada, 20–24 July 2020; pp. 4147–4150.
53. Nakamura, H.; Sakajiri, Y.; Ishigami, H.; Ueno, A. A novel analog front end with voltage-dependent input impedance and bandpass amplification for capacitive biopotential measurements. Sensors 2020, 20, 2476. [CrossRef]
54. Lee, J.S.; Heo, J.; Lee, W.K.; Lim, Y.G.; Kim, Y.H.; Park, K.S. Flexible capacitive electrodes for minimizing motion artifacts in ambulatory electrocardiograms. Sensors 2014, 14, 14732–14743. [CrossRef]
55. Kim, K.K.; Lim, Y.K.; Park, K.S. Common mode noise cancellation for electrically non-contact ECG measurement system on a chair. In Proceedings of the IEEE Engineering in Medicine and Biology Society 27th Annual Conference, Shanghai, China, 1–4 September 2005; pp. 5881–5883.
56. The BioSig Project. Available online: http://biosig.sourceforge.net/ (accessed on 5 January 2021).