Modifying ICCA with Trp-Phe-Phe to Enhance in vivo Activity and Form Nano-Medicine

Background: 1-(4-isopropylphenyl)-β-carboline-3-carboxylic acid (ICCA) was modified by Trp-Phe-Phe to form 1-(4-isopropylphenyl)-β-carboline-3-carbonyl-Trp-Phe-Phe (ICCA-WFF).

Purpose: The object of preparing ICCA-WFF was to enhance the in vivo efficacy of ICCA, to explore the possible targeting action, and to visualize the nano-feature.

Methods: The advantages of ICCA-WFF over ICCA were demonstrated by a series of in vivo assays, such as anti-tumor assay, anti-arterial thrombosis assay, anti-venous thrombosis assay, P-selectin expression assay, and GPIIb/IIIa expression assay. The nano-features of ICCA-WFF were visualized by TEM, SEM and AFM images. The thrombus targeting and tumor-targeting actions were evidenced by FT-MS spectrum analysis.

Results: The minimal effective dose of ICCA-WFF slowing tumor growth and inhibiting thrombosis was 10-fold lower than that of ICCA. ICCA-WFF, but not ICCA, formed nano-particles capable of safe delivery in blood circulation. In vivo ICCA-WFF, but not ICCA, can target thrombus and tumor. In thrombus and tumor, ICCA-WFF released Trp-Phe-Phe and/or ICCA.

Conclusion: Modifying ICCA with Trp-Phe-Phe successfully enhanced the anti-tumor activity, improved the anti-thrombotic action, formed nano-particles, targeted tumor tissue and thrombus, and provided an oligopeptide modification strategy for heterocyclic compounds.

Keywords: ICCA, modification, Trp-Phe-Phe, anti-tumor, thrombus targeting, release, toxicity, nano-species

Introduction

The malignant cancer is a heterogeneous disease and characterized by the different phenotypes.1–3 Through early recognition, surgical extirpation and clinical therapy did reduce the death rate,4 the diseases, such as deep venous and arterial embolisms can induce bad outcomes of cancer patients, thereby the diseases made malignant cancer to be a response for the elevated mortality of human being of the world.5–9 In fact, the correlation between malignant cancer and thromboembolism was established for more than 150 years. Later, it was reported that the risk of hospitalized patients with cancer to develop venous thromboembolism was 4.1 times higher than that of ICCA-WFF released Trp-Phe-Phe and/or ICCA.

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simultaneously inhibiting GPIIb/IIa, P-selectin, arterial thrombosis, venous thrombosis and tumor growth. However, its oral dose (5 μmol/kg) was a bit high. To lower the dose, some oligopeptides were previously designed, prepared and evaluated for modifying ICCA. Among them, Trp-Phe-Phe was found having anti-tumor and anti-thrombotic activities, as well as depressing serum P-selectin and GPIIIa in particular. This led to a hypothesis that coupling ICCA with Trp-Phe-Phe should form the derivative capable of binding GPIIb/IIa and P-selectin. In this case, 1-(4-isopropylphenyl)-β-carboline-3-carbonyl-Tyr-Phe-Phe (ICCA-WFF) was designed and docked into the active sites of P-selectin and GPIIIa.

Materials and Methods

General
Trp and Phe purchased from Sigma were L-configuration. All reactions were done in nitrogen (1 bar). On Bruker AMX-300 and AMX-800 spectrometer $^1$H (300 and 800 MHz) and $^{13}$C (75 and 200 MHz) NMR spectra were measured for the intermediates or ICCA-WFF, for which dimethylsulfoxide (DMSO-d$_6$) was the solvent and tetramethylsilane was the internal standard. Perkin-Elmer 13001 instrument was used to record IR spectra. ESI(±)-ICR-FT-MS spectrum was recorded on a 9.4 T solariX Fourier transform ion cyclotron resonance mass spectrometer (Bruker Corp, Billerica, MA, USA) to get the exact mass of ICCA-WFF. Chromatography was performed by using Sephadex-LH20 or Qingdao silica gel GF254 or Qingdao silica gel H$_{60}$. All solvents were distilled and dried by the literature procedures for use. HPLC purity (Waters, C$_{18}$ column 4.6×150 mm) of the intermediates and ICCA-WFF were more than 96% and 99.63%, respectively. HPLC was performed with Agilent Technologies 1200 Series HPLC system (Agilent Technologies, Santa Clara, CA, USA) on Eclipse XDB C18 column (5 μm, 4.6 mm × 150 mm). The column temperature was 40°C, the mobile phase consisted of water and methanol, and the gradient consisted of 60% methanol (0–5 min), 70% methanol (5–10 min), 80% methanol (10–20 min) and 90% methanol (20–30 min), while the flow rate was 0.8 mL/min. Ultraviolet (UV) absorption spectrum was recorded online. The UV detector was set to a scanning range of 200–400 nm, and the chromatogram of ICCA-WFF was monitored at a wavelength of 280 nm.

Animals and Cell Lines
Human myeloid leukemia cells (K562), human non-small cell lung cancer cells (A549), non-small cell lung cancer cells (95D) and ascites tumor cells (S180) were purchased from keyGEN BioTECH (Nanjing China).

From the Laboratory Animal Center of Capital Medical University male ICR mice (22–24 g) and male Sprague Dawley rats (240–270 g) were purchased. Ethics Committee of Capital Medical University reviewed and approved the protocol of the evaluations. In particular, the animal welfare fulfilled the requirements of the Animal Welfare Act and NIH Guide for Care and Use of Laboratory Animals. Biological data were statistically analyzed with ANOVA. LSD for multiple group comparison. SPSS 19.0-program was used to perform the analysis and a P-value less than 0.05 was considered statistically significant.

Synthesis
The preparations and the physical-chemical data of (1R,3S)-1-(4-isopropylphenyl)-2,3,4,9-tetrahydro-β-carboline-3-carboxylic acid benzyl esters(I), benzyl 1-(4-isopropylphenyl)-β-carboline-3-carboxylate (2) and ICCA were the same as those of the literature.

Preparation of Boc-Phe-Phe-OBzl
At 0°C to a solution of 13.00 g (49.02 mmol) of Boc-Phe in 200 mL of anhydrous THF 6.02 g (44.59 mmol) of HOBT and 11.00 g (53.39 mmol) of DCC were added. This solution was at 0°C stirred for 30 min and as solution A for the coming use. A solution of 13.00 g (44.56 mmol) of HCl-Phe-OBzl in 100 mL of anhydrous THF was adjusted to pH 9 with NMM. This solution was firstly combined with solution A to form a reaction mixture and then stirred at room temperature, of which pH 9 was maintained by adding NMM. Six hours after the stirring TLC (petroleum/acetone,2/1) indicated the complete disappearance of HCl-Phe-OBzl. The reaction mixture was filtered, the filtrate was evaporated in vacuum and the residue was dissolved in 150 mL of ethyl acetate. This solution was successively washed with saturated aqueous NaHCO$_3$ (50 mL×3), 5% aqueous KH$_2$PO$_4$ (50 mL×3), saturated aqueous NaCl (50 mL×3), saturated aqueous NaHCO$_3$ (50 mL×3) and saturated aqueous NaCl (50 mL×3). Ethyl acetate phase was isolated, dried with anhydrous Na$_2$SO$_4$ for 12 hrs, filtered, the filtrate was evaporated in vacuum and the residue was purified on silica gel column (petroleum/acetone,5/1) to give 20.2 g (90%) of title compound as colorless powders. $^1$H NMR (300 MHz, DMSO-d$_6$): δ/ppm = 8.45 (d, J = 7.2 Hz, 1H), 7.24 (m, 15H), 6.88 (d, J = 8.7 Hz, 1H), 5.07 (m, 2H), 4.56 (m, 1H), 4.20 (m, 1H), 3.04 (m, 1H).
2.87 (m, 1H), 2.60 (m, 1H), 1.30 (s, 9H). ESI(+)-MS (m/z): 503 [M+H]+.

Preparation of Phe-Phe-OBzl
At 0°C to a solution of 10.0g (19.9 mmol) of Boc-Phe-Phe-OBzl in 50 mL of anhydrous ethyl acetate, a solution of hydrogen chloride in anhydrous ethyl acetate (50 mL, 4M) was added. Four hours after stirring TLC (petroleum ether/acetone, 2/1) indicated the complete disappearance of Boc-Phe-OBzl. This reaction mixture was evaporated in vacuum and the residue was dissolved in 50 mL of anhydrous ethyl acetate for evaporation. This procedure was repeated for at least 3 times to remove the access hydrogen chloride thoroughly, which provided 7.72 g(96.5%) of the title compound as colorless powders. ESI(+)-MS (m/z): 403 [M+H]+.

Preparation of Boc-Trp-Phe-Phe-OBzl
By using the procedure of item 2 from 12.39 g (40.7 mmol) of Boc-Trp and 14.95 g (37.02 mmol) of Phe-Phe-OBzl. This reaction mixture was evaporated in thorough, which provided 7.72 g(96.5%) of the title compound as colorless powders. ESI(+)-MS (m/z): 689 [M+H]+ (calculated value: 497.2188).

Preparation of 1-(4-isopropylphenyl)-β-carboline-3-carbonyl-Tyr-Phe-Phe-OBzl (3)
By using the procedure of item 2 from 2.75 g (8.3 mmol) of ICCA and 5.89 g (10.0 mmol) of Trp-Phe-Phe-OBzl 5.1 g (68%) of title compound was obtained as colorless powders. IR (KBr): 3396, 3295, 3064, 3029, 2960, 2921, 2862, 1725, 1651, 1523, 1495, 1454, 1392, 1356, 1212, 1173, 1100, 739, 698 cm⁻¹. 1H NMR (300 MHz, DMSO-d6): δ/ ppm = 10.76 (s, 1H), 8.62 (d, J = 7.5 Hz, 1H), 7.90 (d, J = 8.4 Hz, 1H), 7.52 (d, J = 7.8 Hz, 1H), 7.27 (m, 16H), 7.02 (m, 3H), 6.75 (d, J = 8.4 Hz, 1H), 5.07 (m, 2H), 4.64 (m, 2H), 4.57 (q, J = 7.2 Hz, 1H), 4.14 (m, 1H), 2.90 (m, 6H), 1.27 (s, 9H). ESI(+)-MS (m/z): 589 [M+H]+.

Preparation of Trp-Phe-Phe-OBzl
Into a suspension of 1.60 g (2.7 mmol) of Trp-Phe-Phe-OBzl and 160 mg of Pd/C in 30 mL of CH₂Cl₂ hydrogen was introduced for 10 hrs, and TLC (CH₂Cl₂/CH₃OH, 10/1) indicated the complete disappearance of Trp-Phe-Phe-OBzl. The reaction mixture was filtered, the filtrate was evaporated under vacuum and the residue was grindned in anhydrous ether to give 720 mg (53%) of the title compound as the colorless powders. Mp 187–189°C; [α]D²⁵ = −72.4 (c=0.106, CH₃OH). IR (KBr): 3290, 3060, 3029, 2926, 2853, 1656, 1639, 1519, 1495, 1454, 1435, 739, 697 cm⁻¹. 1H NMR (300 MHz, DMSO-d6): δ/ppm = 10.91 (s, 1H), 8.33 (d, J = 7.5 Hz, 1H), 8.25 (d, J = 7.2 Hz, 1H), 7.59 (d, J = 7.5 Hz, 1H), 7.34 (d, J = 7.8 Hz, 1H), 7.16 (m, 14H), 6.97 (t, J = 7.5 Hz, 1H), 4.57 (m, 1H), 4.37 (m, 1H), 3.57 (m, 1H), 3.17 (s, 1H), 3.1 (m, 1H), 2.99 (m, 3H), 2.76 (m, 2H), 13C NMR (75 MHz, DMSO-d6): δ/ppm = 173.24, 172.78, 170.88, 138.38, 137.83, 136.78, 129.84, 129.71, 128.51, 128.39, 127.79, 126.67, 124.68, 121.42, 118.94, 118.75, 111.84, 109.85, 54.84, 54.47, 53.73, 38.18, 37.38, 30.04; HPLC purity: 99.17%; ESI(-)-ICR-FT-MS: 497.2179 [M-H]⁻.
(q, J = 7.2 Hz, 1H), 4.64 (m, 1H), 4.47 (q, J = 6.4 Hz, 1H), 3.38 (dd, J = 14.4 Hz, 7.2 Hz, 1H), 3.18 (m, 2H), 3.05 (m, 3H), 2.81 (m, 1H), 1.33 (s, 6H).\(^{13}\)C NMR (200 MHz, DMSO-\( d_6 \)) : \( \delta /ppm = 173.14, 171.45, 171.41, 164.72, 149.81, 142.01, 141.13, 139.49, 138.01, 136.58, 135.48, 134.64, 130.13, 129.71, 129.57, 129.02, 128.93, 128.68, 128.35, 128.01, 127.33, 126.90, 126.58, 124.31, 122.41, 121.65, 121.30, 120.66, 118.98, 118.71, 113.20, 111.72, 110.06, 54.04, 53.96, 53.74, 38.00, 37.26, 33.91, 28.38, 24.37, 24.31. HPLC purity: 99.63%; ESI(-)-ICR-FT-MS (m/z): 809.3448 [M-H] \(^{-}\) (calculated value, 809.3451).

**FT-ICR-MS and qCID Spectra**
To reveal ICCA-WFF’s molecule assembly its FT-ICR-MS and qCID spectra were measured on Bruker 9.4 T solariX FT-ICR mass spectrometer. The measurement was performed by following the procedure of the equipment.

**NOESY 2D \(^{1}\)H NMR Spectrum**
To reveal the manner of ICCA-WFF’s molecule assembly its NOESY 2D NMR spectrum was measured on Bruker 800 MHz spectrometer. The measurement was performed by following the procedure of the equipment.

**SEM, TEM and AFM Feature**
To visualize the nano-feature of ICCA-WFF in lyophilized powders, in water and in rat plasma its scanning electron microscopy (SEM) image, transmission electron microscopy (TEM) image and atomic force microscopy (AFM) image were recorded by following the procedure. In addition, the nano-property of aqueous ICCA-WFF was identified by Faraday-Tyndall effect.\(^{22}\)

**Mesoscale Simulation of Nano-Particle**
The structure and conformation of ICCA-WFF were generated with ChemDraw Ultra 10.0 (Cambridge Soft) and Materials Studio 3.2 (Accelrys, Inc. USA), and then the conformation optimization, the molecular dynamic simulation, the equilibration of the structure, the building of a rigid coarse grain model of 4 connected spherical beads, and the simulation of 15,000 ps was performed by following the method.\(^{21}\)

**Bioassays**

**In vitro Anti-Proliferation Assay**
The in vitro viability assays of human myeloid leukemia cells (K562), ascites tumor cells (S180), human non-small cell lung cancer cells (A549) and non-small cell lung cancer cells (95D) were performed by using the method of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT).\(^{23}\)

**In vivo Anti-Tumor Assay**
On S180 mouse model this assay was carried out, and the mice were divided into 7 groups (each 12).\(^{24}\) The mice in positive control group were intraperitoneally injected doxorubicin (Dox, 2 µmol/kg/day) for 9 consecutive days, the mice in negative control group were orally given CMCNa (carboxymethyl cellulose sodium, 0.5%) for 9 consecutive days, the mice in parent compound control group were orally given ICCA (5 µmol/kg/day) for 9 consecutive days, the mice in object compound control group were orally given Trp-Phe-Phe (5 µmol/kg/day) for 9 consecutive days, the mice in object compound group were orally given ICCA-WFF (5, 0.5 and 0.05 µmol/kg/day) for 9 consecutive days.

**Testing Serum ALT, AST and Cr**
The serum was prepared by centrifuging (1000 g, 10 min) blood (0.5 mL) of S180 mice treated by CMCNa or ICCA-WFF (5 µmol/kg/day). Serum glutamic pyruvic transaminase (ALT), glutamic oxaloacetic transaminase (AST) and creatinine (Cr) were tested by using the procedure of literature and ALT/GPT Assay Kit (Nanjing Jiancheng Bioengineering Institute, PR China).\(^{25}\)

**Testing Serum BUN**
The serum was prepared by centrifuging (3000 g, 10 min) the blood (0.5 mL) of S180 mice treated by CMCNa or ICCA-WFF (5 µmol/kg/day). Blood urea nitrogen (BUN) was tested by using the procedure of literature and the procedure of the kit (Shanghai Lianshuo Biological Technology Co., Ltd, PR China).\(^{25}\)

**Arterial Thrombus Weight Assays**
Arterial thrombus weight assay was performed on male ICR mouse (22–24 g, each 12) model or on male Sprague Dawley rat (240–270 g, each 12) model by following the procedure of the literature.\(^{25,26}\) In mouse arterial thrombus weight assay 0.5% CMCNa (10 mL/kg, negative control) or a suspension of aspirin in 0.5% CMCNa (240 µmol/kg, positive control) or a suspension of ICCA in 0.5% CMCNa (5 µmol/kg) or a suspension of WFF in 0.5% CMCNa (5 µmol/kg) or a suspension of ICCA-WFF in 0.5% CMCNa (5, 0.5 and 0.05 µmol/kg) were orally given. In rat arterial thrombus weight assay 0.5% CMCNa (3 mL/kg) or suspension of aspirin in 0.5% CMCNa (167 µmol/kg) or a suspension of
ICCA in 0.5% CMCNa (5 µmol/kg) or a suspension of WFF in 0.5% CMCNa (5 µmol/kg) or a suspension of ICCA-WFF in 0.5% CMCNa (5, 0.5 and 0.05 µmol/kg) were orally given.

**P-selectin and GPIIb/IIIa Expression Assays**

The level of P-selectin expression was identified with an enzyme-linked immune-sorbent assay (ELISA). The procedure of the literature and the kit (rat P-selectin ELISA kit; Wuhan Huamei Biotech Co., Ltd., Wuhan, Hubei Province, PR China) was used. GPIIb/IIIa expression was similarly identified with ELISA. The procedure of the literature and the kit (rat GPIIb/IIIa ELISA kit; Wuhan Huamei Biotech Co., Ltd., Wuhan, Hubei Province, PR China) was used.

In ELISA the serum was prepared from the blood of the rats treated by ICCA (5 µmol/kg) or by aspirin (167 µmol/kg, positive control) or by ICCA-WFF (5, 0.5 and 0.05 µmol/kg) or by 0.5% CMCNa (3 mL/kg, negative control).

**Rat Tail Bleeding Time Assay**

This assay was performed by following the procedure of the literature. In assay the male Sprague Dawley rats (240–270 g, each 12) were orally treated with 0.5% CMCNa (3 mL/kg, negative control) or a suspension of aspirin in 0.5% CMCNa (167 µmol/kg, positive control) or a suspension of ICCA-WFF in 0.5% CMCNa (5 µmol/kg) or a suspension of ICCA-WFF in 0.5% CMCNa (5, 0.5 and 0.05 µmol/kg).

**Venous Thrombus Weight Assay**

This assay was performed by following the procedure of the literature. In assay male Sprague Dawley rats (240–270 g, each 12) were orally treated with 0.5% CMCNa (3 mL/kg, negative control) or warfarin in 0.5% CMCNa (4.87 µmol/kg, positive control) or ICCA in 0.5% CMCNa (5 µmol/kg) or ICCA-WFF in 0.5% CMCNa (5, 0.5 and 0.05 µmol/kg).

**Testing PT, TT, APTT and FIB**

To estimate the effect of ICCA-WFF on the prothrombin time (PT), thrombin time (TT), activated partial thromboplastin time (APTT) and plasma fibrinogen (FIB) the blood of 0.5% CMCNa (3 mL/kg, negative control) or warfarin in 0.5% CMCNa (4.87 µmol/kg, positive control) or ICCA in 0.5% CMCNa (5 µmol/kg) or ICCA-WFF in 0.5% CMCNa (5, 0.5 and 0.05 µmol/kg).

Docking Toward the Active Sites of P-selectin and GPIIb/IIIa

To show the superiority of ICCA-WFF to ICCA in fitting the active site of both P-selectin and GPIIb/IIIa the CDOCKER interaction energies were calculated. For this purpose, the conformations of ICCA-WFF and ICCA were generated, and 10 energy optimized conformations of ICCA-WFF and ICCA were docked into the active sites of P-selectin and GPIIb/IIIa with CDOCKER protocol and using the procedure of the literature.

Molecular Assembly and Thrombus Targeting Action

**FT-ICR-MS and Corresponding Characterization**

Molecular assembly of ICCA-WFF was identified with FT-ICR-MS spectra, while the manner of the molecular assembly was identified with the nuclear overhauser effect spectroscopy (NOESY) and energy optimization conformation. These methods were in accordance with the literature.

**FT-ICR-MS Analysis of Homogenate Extracts**

To explore the targeting action the blood, brain, heart, kidney, liver, spleen and thrombus of arterial thrombosis rats treated with 0.5% CMCNa (3 mL/kg, negative control, 12 rats) or ICCA-WFF in 0.5% CMCNa (5 µmol/kg, 12 rats) were homogenized, the homogenates were extracted with methanol and the extracts received FT-ICR-MS tests. Besides, the tumor tissue, blood, brain, heart, kidney, liver and spleen of S180 mice treated with 0.5% CMCNa (3 mL/kg, negative control, 12 rats) or a suspension of ICCA-WFF in 0.5% CMCNa (5 µmol/kg, 12 rats) were homogenized, the homogenates were extracted with methanol and the extracts received FT-ICR-MS analysis. These methods were in accordance with literature.

**Confocal Microscope Imaging**

To show the cellular uptake of ICCA-WFF the confocal microscope image was visualized. At 37°C 95D cells (1×10^5) cells/mL) were incubated in complete RPMI 1640 medium containing 10% fetal calf serum, penicillin (60 µg/mL), and streptomycin (100 µg/mL), as well as CO₂ (a humidified atmosphere, 5%) for 24 hrs. After removing the original medium, the complete RPMI 1640 medium or a solution of ICCA-WFF in the complete RPMI 1640 medium (final concentration: 55 µM, 27.5 µM and 13.5 µM) was added (1 mL/well), and the cells were incubated for another 12 h. After removing the medium, the cells were washed with fresh medium.
(1 mL×3) and resuspended in fresh medium (1 mL/well). The cells treated by the complete RPMI 1640 medium or a solution of ICCA-WFF in the complete RPMI 1640 medium were analyzed by the use of TCS SP8 STED confocal microscopy (Leica, Leica Microsystems, Wetzlar, Germany). The excitation wavelength and the emission wavelength were 405 nm and 575 nm, respectively.

**Results**

**Trp-Phe-Phe Was Found to Be Suitable for Modifying ICCA**

To lower the dose of ICCA some oligopeptides were previously designed, prepared and evaluated. Among them Trp-Phe-Phe was found to be capable of slowing tumor growth, inhibiting thrombosis, and depressing serum P-selectin and GPIIb/IIIa in particular (Figure 1).

Thus, a derivative of ICCA coupling Trp-Phe-Phe could bind GPIIb/IIIa and P-selectin was hypothesized, and a docking investigation was performed. Figure 2 shows that the CDOCKER interaction energies of ICCA and ICCA-WFF with P-selectin are −20.5 kcal/mol and −40.3 kcal/mol, respectively. Figure 2 also shows that the CDOCKER interaction energies of ICCA and ICCA-WFF with GPIIb/IIIa are −23.9 kcal/mol and −41.6 kcal/mol, respectively. For P-selectin and GPIIb/IIIa, the CDOCKER interaction energies of ICCA-WFF are significantly lower than those of ICCA.

The multiple activities of Trp-Phe-Phe in vivo with the CDOCKER interaction energy of ICCA-WFF to be lower than that of ICCA together suggested that Trp-Phe-Phe was suitable for modifying ICCA.

**The Route Is Suitable to the Preparation of ICCA-WFF**

To process the forward investigations, ICCA-WFF was synthesized by following the route of Figure 3. This route consists of the synthesis of ICCA, the preparation of Trp-Phe-Phe-OBzl and the coupling of ICCA with Trp-Phe-Phe-OBzl. It was found that the overall yields of the synthesis of ICCA, the preparation of Trp-Phe-Phe-OBzl and the coupling of ICCA with Trp-Phe-Phe-OBzl were 73%, 83% and 61%, respectively, while the HPLC purity of ICCA-WFF was 99.63%. These data emphasize that this route is suitable for preparing ICCA-WFF.

**ICCA-WFF Forming Trimer and Having Z-Like Conformation**

FT-ICR-MS spectrum shows that the molecular assembly of ICCA-WFF leads to the formation of a trimer. Figure 4A shows that the FT-ICR-MS spectrum of ICCA-WFF gives an anion peak at 809.34135 (the mass of monomer minus H, calculated value: 809.34459), an anion peak at 1619.69296 (the mass of dimer minus H, calculated value: 1619.69296) and an anion peak at 1214.52111 (the mass of trimer minus 2H, calculated value: 1214.52107). Figure 4B shows that the qCID of the trimer minus 2H at 1214.52895 gives an anion peak of monomer minus H at 809.34930 and an anion peak of dimer minus H at 1619.71182. Therefore, the anion peaks of the monomer minus H and the dimer minus H occur in Figure 4A are the fragments of the trimer, and the trimer is the sole form of ICCA-WFF to exist.

NOESY 2D 1H NMR spectrum and energy optimization conformation define the manner of molecular assembly of ICCA-WFF to form the trimer. Figure 4C shows that there is only one interesting cross-peak marked by a red circle. According to the concept of NOESY, this cross-peak reflects an interaction of the methine H of the isopropyl moiety of one molecule and the amino H of the carboxyl terminal Phe of Trp-Phe-Phe moiety of another molecule. This cross-peak also means that the distance between these H is less than 4 Å. Based on the distance between two H atoms, three energy minimized monomers of ICCA-WFF were manually

![Figure 1](https://example.com/figure1.png)

**Figure 1** In vivo activities of Trp-Phe-Phe (WFF), n=12: (A) anti-tumor activity of WFF against S180 mice; (B) anti-thrombotic activity of WFF on rat model; (C) inhibition of WFF to serum P-selectin of arterial thrombosis rats; (D) inhibition of WFF to serum GPIIb/IIIa of arterial thrombosis rats.

**Abbreviation:** WFF, Trp-Phe-Phe.
accessed to form a trimer. This manual operation gives the trimer of ICCA-WFF a Z-like conformation (Figure 4D).

**Aqueous ICCA-WFF Is a Nano-Solution**

To show the nano-property of aqueous ICCA-WFF the Faraday-Tyndall effect was tested. Figure 5A, E, I and M indicate that the ultrapure water of various pH occurs no Faraday-Tyndall effect. Figure 5B, F, J and N indicate that the radiation of 650 nm laser does not induce the ultrapure water of various pH to occur Faraday-Tyndall effect. Figure 5C, G, K and O indicate that 3.7 μM solutions of ICCA-WFF in ultrapure water of various pH occur no Faraday-Tyndall effect. Figure 5D, H, L and P indicate that the radiation of 650 nm laser induces 3.7 μM solutions of ICCA-WFF in ultrapure water of various pH to occur.
Faraday-Tyndall effect. These observations mean that the aqueous ICCA-WFF is nano-solution.

In Aqueous Solution ICCA-WFF Exists as Nano-Particles
To visualize the nano-feature of ICCA-WFF in the ultra-pure water of pH 6.8 the TEM images were provided. Figure 6 shows that in 1.23 nM, 123 nM and 12.3 μM solutions of ICCA-WFF form nano-particles of 61.5–230.7 nm in diameter. Besides, the size of the nano-particles is independent of the concentration of the solution.

In Solid State ICCA-WFF Exists as Nano-Particles
To visualize the nano-feature of ICCA-WFF in solid state the lyophilized powders were prepared from solution with various concentration and pH values, and the SEM images of the lyophilized powders tested. Figure 7 shows that the lyophilized powders of ICCA-WFF consist of nanoparticles, the diameter of the nano-particles is less than 70 nm, and the diameter of the nano-particles is independent of the concentration and the pH value of the solution used for preparing the lyophilized powders.

In Rat Serum ICCA-WFF Exists as Nano-Particles
To visualize the nano-feature of ICCA-WFF in rat plasma the AFM images were recorded. Figure 8A shows that the rat plasma alone has no interesting particles. Figure 8B–D shows that in the rat plasma ICCA-WFF exists as nanoparticles of 12.3–59.2 nm in height, and the height is independent of the concentration.
ICCA-WFF Has Similar Cytotoxicity to ICCA in vitro

To know if the modification of ICCA by Trp-Phe-Phe greatly alters the cytotoxicity of ICCA, the viabilities of A549, K562, S180 and 95D cells treated by ICCA-WFF were measured by using MTT method. Figure 9 shows that the IC$_{50}$ values of ICCA-WFF against A549, K562, S180 and 95D cells are ~50–100 μM. The IC$_{50}$ of ICCA-WFF against 95D cells has the lowest value. In was known that the IC$_{50}$ values of ICCA against A549, K562, S180 and 95D cells are also ~50–100 μM. Thus, for these cells,
ICCA-WFF and ICCA have similar IC_{50}s, and Trp-Phe-Phe modification does not greatly alter the cytotoxicity of ICCA.

Trp-Phe-Phe Modification Increases Anti-Tumor Activity of ICCA in vivo
To evidence the enhanced antitumor activity of Trp-Phe-Phe modification of ICCA in vivo the tumor growth assay was performed on S180 mouse model. Figure 10A shows that the tumor weight of S180 mice orally treated with 5 μmol/kg/day of ICCA for 9 consecutive days is significantly lower than that of S180 mice orally treated with CMCNa. Figure 10A also shows that ICCA-WFF dose-dependently slows tumor growth, and the tumor weight of S180 mice orally treated with 0.5 μmol/kg/day
of ICCA-WFF for 9 consecutive days is equal to that of S180 mice orally treated with 5 μmol/kg/day of ICCA. Therefore, Trp-Phe-Phe modification leads the anti-tumor activity of ICCA to 10-fold increase.

**Trp-Phe-Phe Modification Increases the Inhibition of ICCA to Arterial Thrombosis in vivo**

To evidence the enhanced anti-thrombotic activity of Trp-Phe-Phe modification of ICCA in vivo the arterial thrombotic assays were performed on mouse and rat models. Figure 10B shows that ICCA-WFF dose-dependently decreases the arterial thrombus weight of the mice, and the arterial thrombus weight of the mice orally treated by 0.5 μmol/kg of ICCA-WFF is equal to that of the mice orally treated by 5 μmol/kg of ICCA. Therefore, Trp-Phe-Phe modification leads the anti-tumor activity of ICCA to 10-fold increase.

**ICCA-WFF Down regulates P-selectin and GPIIb/IIIa Expression in vivo**

To correlate P-selectin and GPIIb/IIIa with the inhibition of ICCA-WFF to arterial thrombosis in vivo the expression levels of P-selectin and GPIIb/IIIa were examined by using the reported methods. Figure 11A shows that P-selectin level in the serum of the rats treated with 5 μmol/kg ICCA is significantly lower than that of the rats treated with CMCNa, and is significantly higher than that of the rats treated with 5 μmol/kg ICCA-WFF. Thus, by downregulating P-selectin expression ICCA-WFF inhibits arterial thrombosis.

**Trp-Phe-Phe Modification Does Not increase the Inhibition of ICCA to Venous Thrombosis in vivo**

To show the effect of Trp-Phe-Phe modification on the anti-venous thrombotic activity of ICCA the venous thrombotic assays were performed on rat models. Figure 10D shows that ICCA-WFF dose-dependently decreases the venous thrombus weight of the treated rats, and the minimal effective dose is 0.5 μmol/kg. Besides, at 5 μmol/kg of dose ICCA-WFF, ICCA and warfarin have equal activity. This means that Trp-Phe-Phe modification does not enhance the inhibition of ICCA to venous thrombosis in vivo.
effectively inhibits arterial thrombosis. Besides, the efficacy of ICCA-WFF downregulating the expression of P-selectin and GPIIb/IIIa is 33 folds higher than that of aspirin.

ICCA-WFF Has No Bleeding Risk

To show the bleeding risk of ICCA-WFF the rats of arterial thrombosis assay to be given tail bleeding time assay by using the reported method. Figure 11C shows
that at 5 μmol/kg of oral dose both ICCA and ICCA-WFF do not prolong tail bleeding time of the treated rats. Thus, Trp-Phe-Phe modification does not increase the bleeding risk of ICCA. Besides, the tail bleeding time of the rats treated by 167 μmol/kg of aspirin is significantly longer than that of the rats treated by 5 μmol/kg of ICCA-WFF.

By Adhering Rat Platelets ICCA-WFF Inhibits P-selectin and GPIIb/IIIa Expression

The in vitro action of ICCA-WFF on the resting rat platelets and arachidonic acid (AA) activated rat platelets was visualized by AFM. Figure 12A shows that the resting rat platelets have a smooth surface and have no pseudopodia. Figure 12B–D shows that in the presence of ICCA-WFF (1.2×10^{-2} nM, 1.2×10^{-3} nM and 1.2×10^{-4} nM) the resting rat platelets neither have smooth surface nor have pseudopodium. Figure 12E shows that the activation of AA induces rat platelets to extend pseudopodia and to form aggregation. Figure 12F–H show that ICCA-WFF concentration (1.2×10^{-2} nM, 1.2×10^{-3} nM and 1.2×10^{-4} nM) dependently inhibits AA activated rat platelets to extend pseudopodia and to form aggregation. These phenomena suggest that by adhering onto the surfaces of the rat platelets, ICCA-WFF downregulates the expression of P-selectin and GPIIb/IIIa.

Mesoscale Simulation of ICCA-WFF Forming Nano-Particle

To dissect the process of the nano-particle formation and to predict the molecular number of ICCA-WFF in a nano-particle of definite diameter a mesoscale simulation was performed. Figure 13 shows that the formation of the nano-particle consists of 3 steps, and a nano-particle of 99.96 nm in diameter contains 1187 molecules of ICCA-WFF. This demonstrates that mesoscale simulation not only reveals the course of nano-particle formation but also estimates how much molecules will be involved in a nano-particle of the exact size.

ICCA-WFF Has No Liver and Renal Toxicity

Serum ALT, AST, Cr and BUN are usually for estimating the toxicity of the drug to kidney and liver. In this profile, the ALT, AST, Cr and BUN of S180 mice treated with 5 μmol/kg/day of ICCA-WFF or CMCNa for 9 consecutive days were measured and are shown in Figure 14A. As seen, the

Figure 12 Imaging the interaction of ICCA-WFF and rat platelets by AFM: (A) AFM image of resting rat platelets; (B) AFM image of resting rat platelets with ICCA-WFF (1.2×10^{-2} nM); (C) AFM image of resting rat platelets with ICCA-WFF (1.2×10^{-3} nM); (D) AFM image of resting rat platelets with ICCA-WFF (1.2×10^{-4} nM); (E) AFM image of AA activated rat platelets; (F) AFM image of AA activated rat platelets with ICCA-WFF (1.2×10^{-3} nM); (G) AFM image of AA activated rat platelets with ICCA-WFF (1.2×10^{-4} nM); (H) AFM image of AA activated rat platelets with ICCA-WFF (1.2×10^{-5} nM). (Abbreviations: AFM, atomic force microscopy; ICCA-WFF, 1-(4-isopropylphenyl)-β-carboline-3-carbonyl-Tyr-Phe-Phe; AA, arachidonic acid.)
ALT, AST, Cr and BUN of S180 mice treated by ICCA-WFF are equal to those of S180 mice treated by CMCNa. These data mean that, similarly to CMCNa, 5 μmol/kg of ICCA-WFF does not injure the kidney and the liver of S180 mice.

ICCA-WFF Affects No Coagulation Function

To estimate the effect of ICCA-WFF on coagulation function the PT, TT, APTT and FIB of the rats in the venous thrombosis assay were measured. Figure 14B shows that the PT, TT, APTT and FIB of the rats treated by ICCA-WFF (5 μmol/kg, 12 rats) are equal to those of the rats treated by CMCNa (3 mL/kg, negative control, 12 rats). The PT, TT, APTT and FIB suggest that, similarly to CMCNa, 5 μmol/kg of ICCA-WFF does not injure the coagulation system of the thrombosis rats.

ICCA-WFF Targets Arterial Thrombus and Tumor Tissue

To ensure the targeting action and release the extracts of the homogenates of the blood, brain, heart, kidney, liver, spleen, arterial thrombus or venous thrombus of the rats treated with CMCNa or ICCA-WFF (5 μmol/kg), as well as the extracts of the homogenates of the blood, brain, heart, kidney, liver, spleen and the tumor tissue of S180 mice treated with CMCNa or ICCA-WFF (5 μmol/kg/day) received FT-ICR-MS spectrum analysis. It was found that the FT-ICR-MS spectra of the extract of the homogenate of the arterial thrombus, but not the blood and the organs, of the arterial thrombosis rats treated by ICCA-WFF gave the ion peaks related to ICCA-WFF. It was also found that the FT-ICR-MS spectra of the extract of the homogenate of the venous thrombus, but not the blood and the organs, of the venous thrombosis rats treated by ICCA-WFF gave the ion peaks related to ICCA-WFF. It was again found that the FT-ICR-MS spectra of the extract of the homogenate of the tumor tissue, but not the blood and the organs, of the S180 mice treated by ICCA-WFF gave the ion peaks related to ICCA-WFF. These observations are shown in Figure 15.

Figure 15A and B are the FT-ICR-MS spectra of the extracts of the homogenates of the arterial thrombus and the venous thrombus, respectively. In both of the spectra,
there are the ion peaks of ICCA and Trp-Phe-Phe. These data evidence that ICCA-WFF targets thrombus and releases anti-thrombus pharmacophores ICCA and Trp-Phe-Phe. Figure 15C is FT-ICR-MS spectrum of the extract of the homogenate of the tumor tissue. In the spectrum, there is the ion peak of ICCA. This data evidences that ICCA-WFF targets tumor tissue and releases anti-tumor pharmacophore ICCA.

ICCA-WFF Enters Cytoplasm of 95D Cells

To show the cellular uptake of ICCA-WFF the confocal microscope images of 95D cells treated by complete RPMI 1640 medium and ICCA-WFF (55 μM, 27.5 μM and 13.5 μM) were recorded, and the images are visualized. Figure 16A shows that at 405nm of excitation wavelength and 575 nm of emission wavelength the nucleus of the cells treated by the complete RPMI 1640 medium exhibits weak red. Figure 16B–D show that at 405nm of excitation wavelength and 575 nm of emission wavelength the cytoplasm, but not nucleus of the cells, treated by ICCA-WFF exhibits strong red. This strong red is from the UV absorption of ICCA-WFF at 254 nm, and the absorption strength depends on the concentration of ICCA-WFF. Thus, confocal microscope images reflect the uptake of ICCA-WF for 95D cells in a concentration-dependent manner.

Discussion

The synthetic route of Figure 2 provides ICCA-WFF in proper yield and high HPLC purity. These results demonstrate that the route is suitable for preparing ICCA-WFF. FT-
ICR-MS spectrum approves that due to molecular assembly the trimer is the sole existence form of ICCA-WFF. To explore whether the trimer can further assemble the aqueous ICCA-WFF is radiated with 650 nm laser to test Faraday-Tyndall effect. The appearance of Faraday-Tyndall effect suggests that aqueous ICCA-WFF has nano-property and this nano-property should contribute to the assembly of ICCA-WFF’s trimer. The assembly of the trimer is further visualized with TEM, ESM and AFM images. The images show that even though in ultrapure water of pH 6.8 the diameter of the nano-particles is 61.5–230.7 nm, in the lyophilized powders, the diameter of the nano-particles is less than 70 nm and in rat plasma the height of the nanoparticles is less than 60 nm in particular. The size of the nanoparticles suggests that in the blood circulation ICCA-WFF is able to avoid macrophage phagocytosis and is capable of safe delivery. AFM images also visualize that the size of the nano-particles benefits ICCA-WFF adhering onto the surfaces of AA activated rat platelets. Logically, this kind of adhering property benefits ICCA-WFF depressing the expression of P-selectin and GPIIb/IIIa.

The minimal effective dose of ICCA-WFF to slow tumor growth is 0.5 μmol/kg/day, and the minimal effective dose of ICCA-WFF to inhibit the arterial thrombosis and the venous thrombosis is 0.5 μmol/kg. On the other hand, however, Trp-Phe-Phe modification increases the inhibition of ICCA to tumor growth and arterial thrombosis, but does not increase the inhibition of ICCA to venous thrombosis. In addition, the in vivo activities of ICCA-WFF inhibiting the expression of P-selectin and GPIIb/IIIa are significantly higher than those of ICCA. FT-ICR-MS spectrum analysis ensures that after oral administration ICCA-WFF selectively distributes to tumor tissue and thrombus as well as releasing anti-tumor and anti-thrombotic pharmacophores wherein. ICCA-WFF has no bleeding risk, no effect on the coagulation function and no liver or kidney toxicity. ICCA-WFF is a promising nano-medicine.

Conclusions

Coupling 1-(4-isopropylphenyl)-β-carboline-3-carboxylic acid (ICCA) and Trp-Phe-Phe smoothly provides 1-(4-isopropylphenyl)-β-carboline-3-carbonyl-Trp-Phe-Phe (ICCA-WFF). The molecular assembly leads ICCA-WFF been able to form nano-particles capable of safe delivery in the blood circulation, adhering on the surfaces of platelets, targeting tumor tissue and thrombus as well as releasing anti-tumor and anti-thrombotic pharmacophores wherein. ICCA-WFF has no bleeding risk, no effect on the coagulation function and no liver or kidney toxicity. ICCA-WFF is a promising nano-medicine.

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Disclosure

The authors report no conflicts of interest in this work.

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