The role of allogeneic platelet-rich plasma in patients with diabetic foot ulcer: Current perspectives and future challenges

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The frequency of chronic cutaneous wounds are sharply increasing in aging populations. Patients with age-related diseases, such as diabetes, tumors, renal failure and stroke are prone to soft tissue and skin injury, compounded by slowed healing in aging. Imbalance of wound inflammation, loss of growth factor secretion, and impairment of tissue repair abilities are all possible reasons for failed healing. Therefore, it is vital to explore novel approaches to accelerate wound healing. Platelet-rich plasma (PRP) as a cell therapy has been widely applied for tissue repair and regeneration. PRP promotes wound healing by releasing antimicrobial peptides, growth factors and micro-RNAs. Medical evidence indicates that autologous platelet-rich plasma (au-PRP) can promote wound healing effectively, safely and rapidly. However, its clinical application is usually restricted to patients with chronic cutaneous wounds, generally because of other severe complications and poor clinical comorbidities. Allogeneic platelet-rich plasma (al-PRP), with abundant sources, has demonstrated its superiority in the field of chronic wound treatment. Al-PRP could overcome the limitations of au-PRP and has promising prospects in clinical applications. The aim of this review is to summarize the current status and future challenges of al-PRP in chronic cutaneous wound management. We also summarized clinical cases to further describe the application of al-PRP for chronic wounds in clinical practice.

KEYWORDS
allogeneic platelet-rich plasma, wound healing, current perspectives, future challenges, diabetic foot ulcer
Introduction

In recent decades, the frequency of chronic cutaneous wounds has significantly increased, likely due to health issues in the aging population (Jiang et al., 2011; Graves et al., 2022). Patients with diabetes mellitus (DM), tumors and surgeries can experience soft tissue injury because of long-term bed rest and mobility problems (Armstrong et al., 2017; Xu et al., 2022). Diabetic foot ulcers (DFU), pressure ulcers, venous ulcers and arterial insufficiency ulcers greatly lower quality of life, longevity, and impact financial and social burdens (Xu and Ran, 2016; Kapp and Santamaria, 2017; Graves et al., 2022). An epidemiological study of chronic wounds revealed that its prevalence in Wales was 6% in 2012–13 (Phillips et al., 2016). Chronic cutaneous wounds have different causative etiologies (Jiang et al., 2022). DFU is one of most common chronic diabetes-related injuries and has become the leading cause of non traumatic amputation and all-cause mortality, due to the high prevalence of DM (Xie et al., 2022). Here, we mainly focused on exploring a novel approach for chronic cutaneous wound healing, especially for patients with DFU.

DFU is one of most severe complications in patients with DM. The incidence of DFU is approximately 6.3% (Zhang et al., 2017). DFU wounds are difficult to heal, and the 5-year mortality after amputation reaches 40% (Wang et al., 2014; Phillips et al., 2016; Xu and Ran, 2016). It has been reported that DFU may have a worse prognosis than some forms of cancer (Armstrong et al., 2020). Patients with DFU always have a poor systemic condition, poor skin wound-healing performance and ulcer recurrence (Jiang et al., 2021). Hence, exploring a new, prompt, effective and safe approach for the promotion of DFU closure is key to reduce morbidity and mortality.

The current treatment for diabetic foot ulcers

The main reasons for the difficulty associated with DFU healing are infection, impaired tissue repair function, and loss of growth factor secretion (Pietramaggiori et al., 2008; Meyer et al., 2011; Deng et al., 2021). There are 22 clinical practice guidelines for diabetic foot. Based on the recommendations, current standard of care for DFU care mainly involves offloading of pressure, wound care, choice of shoes and adjunctive treatment. Among these recommendations, promotion of DFU healing is extremely important for prevention of ulcer and its recurrence (Sun et al., 2022). Except for wound debridement and vascular intervention treatment, cell therapy is also a promising approach for DFU, including stem cell, platelet-rich plasma (PRP) and its derived growth factors. After activation by thrombin and calcium, PRP released transforming growth factor, platelet-derived growth factor, insulin-like growth factor, fibroblast growth factor and vascular endothelial growth factor, which play an important role in tissue repair and regeneration (Carter et al., 2003). In 1998, Marx et al. demonstrated novel data on the effectiveness of PRP in the treatment of bone defects, which attracted the attention of the wound repair and regeneration fields (Marx et al., 1998). Later, researchers found that PRP can accelerate wound healing and serve as an antibiosis agent (Bielecki et al., 2007; Li and Li, 2013; López et al., 2014; Li et al., 2019a). PRP is regarded as a reservoir of host defense factors, and these secreted factors assist the host against the bacteria (Tamagawa-Mineoka, 2015). Both functions have also been proven in animal experiments (Maciel et al., 2012). Moreover, PRP can reduce the recurrence rate, infection rate, amputation rate, mortality and medical costs in the treatment of DFU (Dougherty, 2008). PRP can be harvested from autologous or allogeneic whole blood. Autologous platelet-rich plasma (au-PRP) has already been successfully applied in the treatment of chronic refractory wounds for many years. In our previous reports, diabetic patients with acute necrotizing fasciitis and refractory diabetic foot benefited from treatment with au-PRP (Deng et al., 2016; Jiang et al., 2020). Furthermore, evidence-based medical studies also supported the notion that au-PRP could improve the healing of chronic wounds, especially for DFU (Martinez-Zapata et al., 2016). However, diabetic patients with severe acute or chronic complications or coexisting diseases, such as peripheral arterial and neuropathic disease (Zhang et al., 2022), thrombosis, anemia, thrombocytopenia, or malnutrition are usually restricted from the utilization of au-PRP. Alternatively, allogeneic platelet-rich plasma (al-PRP) has already been identified to promote the healing of chronic wounds in our and other previous studies (Villela and Santos, 2010; Senemî et al., 2018; Liao et al., 2020). PRP can be harvested from patients as autologous preparations. However, under diabetic condition, poor platelet number, reduced cell activity or impaired PRP efficacy may limit their use. Administration of allogeneic PRP donated by healthy and/or younger individuals is regarded with increasing interest to overcome such limitation (He et al., 2020). Al-PRP may overcome the shortcomings of au-PRP and provide a new strategy in adjunct treatment of chronic cutaneous wounds. The aim of this review is to summarize the current and prospective challenges of PRP treatments, as well as review key case that used al-PRP in the management of chronic cutaneous wounds.

Preparation protocol for platelet-rich plasma

Platelet-rich plasma (PRP) contains high concentrations of the growth factors (Tian et al., 2019), antibacterial agent (Li et al., 2019a; Sethi et al., 2021). Based on recently reports of systematic review and meta-analysis, current evidence suggests that autologous PRP may increase wound closure, shorten healing time in patients with DFU (Li et al., 2019b; Dai et al., 2020; Qu...
et al., 2021), but both these studies and the International Working Group of the Diabetic Foot (IWGDF) working group indicate that a good design and of a high quality are needed before widely application (Game et al., 2016).

The inconsistent clinical effect of PRP for DFU, the preparation protocol may be considered as main cause. Numerous studies have demonstrated the preparation protocol of PRP by separating whole blood via gradient centrifugal stratification. However, as mentioned in a recent systematic review study, the composition of PRP was completely reported less than 30% in enrolled forty-one studies. Two ranges could be identified for platelet concentration, the first between $0.14 \times 10^6$ and $0.80 \times 10^6$ platelets/µl and the second between $1.086 \times 10^6$ and $10 \times 10^6$ platelets/µl. That is to say, efficacy of PRP depends greatly on its preparation approach. Therefore, there is a need to standardized methodology of PRP preparation as well as its guidelines creation (Anitua et al., 2022; Mastrogiacomo et al., 2022).

Au-PRP applications and limitations in diabetic foot ulcers

Au-PRP is extracted by centrifugation and separation from peripheral whole blood, and then a gel is formulated by activation with a mixed solution of thrombin and calcium. Recently, we published a report on a severely infective diabetic patient with foot necrotising fasciitis and gaseous gangrene, who successfully recovered and underwent limb salvage after a combination of clinical interventions that included PRP (Deng et al., 2016). We recently also reported another case with refractory chronic DFU in which amputation was successfully prevented through a combination approach of negative pressure wound therapy, au-PRP and antibiotic bone-cement (Jiang et al., 2020). Furthermore, a study by Babaei et al. (2017) recruited 150 patients with DFUs treated with au-PRP, and showed that DFUs with a wound size of 2–2.5 cm² completely healed after 7.2 weeks, a wound size of 5.8–8.5 cm² healed after 7.5 weeks, and a wound size of 8.5–12.5 cm² healed after 8.8 weeks. Interestingly, there was no recurrence after an 8-month follow-up. Au-PRP could promote wound healing more quickly and effectively than traditional antibiotic dressings (Deng et al., 2016; Ahmed et al., 2017; Babaei et al., 2017; Jiang et al., 2020). Based on a literature review, PRP as a cell therapy could not only accelerate wound closure but also improve wound infection as an adjunct antibacterial agent. In addition, in our previous empirical study, PRP played an important role in anti-inflammation, anti-infection and promotion of wound healing (Li et al., 2019a).

Au-PRP is now widely used in tissue repair and regeneration in medical cosmetology (Lee et al., 2019), ophthalmology (Avila et al., 2018), orthopedics (McCarrel et al., 2014), neurology (Ye et al., 2012), and oral and maxillofacial surgery (Anitua, 1999; Piccin et al., 2016). Wounds and incisions, however, are usually in an acute state, even though most patients with chronic refractory wounds have experienced a long history of chronic disease (Yotsu et al., 2015). These chronic disease states are often accompanied by acquired infection, kidney disease, vascular disease and tumors. Some of these patients may also experience malnutrition, frailty, gastrointestinal hemorrhage, anemia and cachexia. All these factors make it difficult to prepare au-PRP and ultimately restrict its applicability. Hence, based on our experience and research reports, we suggest that au-PRP treatment should not be recommended if a patient is experiencing one of the following comorbidities: current or recent chemotherapy treatment for cancer; chronic diseases such as severe high blood pressure, acute coronary disease, severe renal dysfunction, or gastrointestinal hemorrhage; hematological diseases such as anemia, thrombocytopenia, leukemia, or lymphoma; cachexia; or immune deficiency. Therefore, al-PRP may be considered an alternative to PRP for chronic wound treatment.

The methodology of al-PRP preparation

Al-PRP is extracted from banked or fresh platelets donated from healthy subjects. The preparation methods of al-PRP mainly including: manual platelet separation with venous whole blood or platelets apheresis with automatic machine only collected platelets (Figure 1). The preparation of al-PRP by the centrifugal separation method has lower requirements for preparation devices, is cost effective, and the actual procedural steps are simple and feasible. It is the most widely used preparation method at present. An increasing number of studies have identified the efficacy and safety of al-PRP for chronic wounds. Thus, the application of al-PRP has received more attention in the field of chronic wound repair, but patients and physicians worry about its safety in clinical practice (Akbarzadeh et al., 2021). We have carefully reviewed the literature through December 2021 to discuss its efficacy and safety (Table 1).

Al-PRP application for chronic wound repair

In 2007, Smrke et al. (2007) performed a combined therapy for delayed healing of severe fracture wounds using autologous cancellous bone transplantation and al-PRP. The fracture wound healed after 6 months, the patient could walk on the healed leg, and both legs were of the same length. In 2010, Scevola et al. (2010) recruited 13 patients who suffered pressure sores because of prolonged bed rest secondary to spinal injuries. Even though there was no significant difference in the final reduced ulcer area, al-PRP gel was more effective in accelerating ulcer healing in the
FIGURE 1
A schematic diagram of allogenic PRP treatment. There are two different preparation methodology of allogenic PRP donated from healthy volunteers: manual platelet separation with venous whole blood or platelets apheresis with automatic machine only collected platelets. The allogenic platelet rich gel was used to diabetic foot ulcer after activation by with a mixed solution of thrombin and calcium.

TABLE 1 Summary of studies on efficacy and safety of al-PRP for wound healing.

| Author            | Year | Study design       | Participants/Wound                                      | Intervention                                                                                      | Outcomes/Adverse effects |
|-------------------|------|--------------------|---------------------------------------------------------|---------------------------------------------------------------------------------------------------|--------------------------|
| He et al.         | 2020 | Observational cohort study | 75 patients with DLEUs al-PRP (n = 20) au-PRP (n = 25) SWT (n = 30) | Source: PRP from banked blood Centrifuge 1: 600 g for 15 min Centrifuge 2: 1,135 g for 7 min al-PRP Concentration: mean (1,043 ± 180.3) ×10^9/L au-PRP Concentration: mean (939 ± 237.4) ×10^9/L Treatment Regime 1. Standard care 2. After wound debridement with al-PRP or au-PRP | The wound healing times of the al-PRP group (56.9 ± 29.22 days) and au-PRP group (55.6 ± 33.8 days) were significantly shorter than those of the CWT group (88.0 ± 43.4 days) (p < 0.05) No obvious adverse reactions (fever, edema, pain, skin itching, rash, or other sensory abnormalities) |
| Liao et al.      | 2020 | Randomized Controlled trial | 60 patients with refractory wounds | Source: PRP from family member donation Centrifuge 1: 400 g for 10 min Centrifuge 2: 1,200 g for 20 min al-PRP Concentration: 1,200 ×10^9/L Treatment Regime 1. Wound debridement + saline dressing 2. Dressed in PRP | Significant improved rate of healing at week 1 and week 3 time points (p < 0.05) Control: conventional skin grafting No adverse reactions |
| Semenič et al.  | 2018 | Randomized Controlled trial | 60 patients with ulcers of different aetiologies | Source: ABO matched donation Centrifuge: not reported PRP Concentration: not reported Treatment Regime: 1. Wound irrigated with antiseptic 2. PRP gel applied, covered with dressing 3. A total of treatment for 3 times | Wound size decreased to 35.01% compared to 89.95% in controls (p < 0.001) at 6 months No adverse reactions observed |
| Asadis et al.    | 2014 | Case series         | 10 patients with refractory ulcer failed to healing by traditional therapies | Source: ABO matched peripheral blood Centrifuge 1: 2,000 g for 2 min Centrifuge 2: 4,000 g for 8 min PRP Concentration: not reported Treatment Regime: 1. Wound irrigated with antiseptic 2. PRP gel applied, covered with dressing 3. A total of treatment for 3 times | 9 patients completely healed, 1 patient with the area of wound reduced significantly |
| Scevola et al.   | 2010 | Randomized Controlled trial | 13 patients with pressure sores | Source: platelet concentrate from hospital Centrifuge: not reported PRP Concentration: more than 2.0×10^9 platelets transplanted Treatment Regime: 1. Clean wound bed 2. Twice weekly for 18 weeks Control: no platelet gel applied | Improved granulation tissue proliferation in first 2 weeks No changes in ulcer bacteria contents, nor serum signs of infection |
| Jeong et al.     | 2010 | Randomized Controlled trial | 100 patients with diabetic foot ulcers | Source: blood bank platelet concentrate Centrifuge: 3,000 g for 30 min PRP Concentration: mean 1.1×10^10 ± 3.9×10^9 platelets transplanted Treatment Regime: 1. Wounds debridement 2. Topical application of 12.5 or 25 ml of concentrate plus fibrinogen, 2 doses 3–4 days apart Control: topical fibrinogen and thrombin | 79% complete healing at 12 weeks Mean healing time 7.0 ± 1.9 weeks, compared to 9.1 ± 2.2 weeks in controls No adverse effects in each of the groups |

DLEUs, diabetic lower extremity ulcers; al-PRP, allogenic platelet-rich plasma; au-PRP, autologous platelet-rich plasma; CWT, conventional wound therapy; PRP, platelet-rich plasma.
first 2 weeks than treatment with the best dressing. In 2013, Tashnizi et al. (2013) recruited six patients with life-threatening, chronic sternal wounds and multidrug-resistant septicemia. After a mixture of al-PRP and fibrin application, five cases completely healed, and one case had a reduced wound area. In 2014, ten patients with refractory ulcers that failed to heal by traditional therapies were treated with al-PRP; nine of the patients completely healed, and one patient had a significantly reduced wound area (Asadis et al., 2014). In 2015, Costanzo et al. (2015) used al-PRP in a multiple myeloma patient who suffered skin flap necrosis because of the exudative lesion on the left hand caused by the leakage of chemical medicine. The wound completely healed after 12 al-PRP treatments, and no recurrence was observed at the 19-month follow-up. There were no adverse events reported in the above clinical studies.

In recent case reports, al-PRP was proven to be safe and efficacious for chronic wounds either as an application of al-PRP alone (Oliveira et al., 2017; Zhao et al., 2019) or in combination as an adjuvant method (Hernandez et al., 2012; Ju et al., 2021). Furthermore, a prospective, randomized, single-center study was conducted recently and enrolled 60 patients with chronic wounds. The wounds in the al-PRP group were treated by standard care and al-PRP, while the control group was treated only by standard care. The combined treatment of chronic wounds by standard care and al-PRP significantly shortened healing time, suggesting that al-PRP is an effective, safe, adjuvant treatment for chronic wounds (Liao et al., 2020).

Based on our long-term research on DFUs, we also performed an observational study to compare the efficacy and safety of al-PRP and au-PRP for diabetic lower extremity ulcers, one of patient with DFU was successfully healed by al-PRP (Figure 2). Seventy-five patients with diabetic lower extremity ulcers were divided into three groups according to age and ulcer size. We found that the wound healing times in the al-PRP group and au-PRP group were significantly shorter than those in the standard care group. Although there was no significant difference in the daily healing area among all groups, the trend of the healing rate in the al-PRP group, au-PRP group, and standard care group gradually decreased. No obvious adverse reactions (fever, edema, pain, skin itching, rash, or other sensory abnormalities) were observed in either the au-PRP or the al-PRP groups. Al-PRP could effectively and safely promote wound healing in patients with diabetic lower extremity ulcers as an off-the-shelf solution when au-PRP is not indicated (He et al., 2020).

Basic research trends of al-PRP in wound treatment

The promotion of wound healing depends on the amount of platelet content and whether platelet function is intact. Studies have found that the platelet turnover rate of diabetic mice is increased (Anitua, 1999), and the content of growth factors and proteins that promote ulcer healing in the autologous plasma of diabetic animals are decreased (Tyndall et al., 2003). In addition, platelet function is also related to patient age. With age, oxidative stress increases correspondingly, and the ability of nitric oxide to inhibit platelet activation is affected (Chris, 2016). One study demonstrated that platelet aggregation, integrin αIIbβ3 and P-selectin exposure were negatively correlated with age (Gilstad et al., 2009). The content and expression of platelet microRNAs (miRNAs) were also affected by age and activation approach (Simon et al., 2014; Rui et al., 2021). Al-PRP is derived from healthy adult blood donors, is less affected by the above factors and has a more stable curative effect.

In 2015, Tae-ho et al. (2015) used al-PRP to treat beagle dogs with acute pancreatitis complicated by disseminated intravascular coagulation causing large-area skin defects. After 2 days of treatment, granulation tissue began to grow. After 21 days, the wounds were completely healed. Another
study confirmed that intra-articular injection of al-PRP combined with water activity can effectively promote the repair of tendon injury in rats (Rajabi et al., 2015). In 2017, Abouelnasr et al. (2017) confirmed through dog experiments that al-PRP can repair abdominal wall defects and reduce peritoneal adhesions and recurrence rates by promoting neovascularization and increasing tissue deposition and fusion. In 2022, Saputo et al. (2022) confirmed that the application of allogeneic freeze-dried PRP was able to increase the number of fibroblasts and neovascularization in the full-thickness wound-healing process in rabbits. The allogeneic freeze-dried platelet-rich plasma increased the number of fibroblasts and neovascularization in wound healing. Allogeneic platelet concentrates derived from healthy blood donors have beneficial effects on various aspects of wound healing in vitro and are superior to plasma or platelets alone (van der Bijl et al., 2019). Al-PRP could be a promising bioactive scaffold due to its synergistic effect in supporting cell proliferation, maintaining cell viability and favoring extracellular matrix production; thus, al-PRP can likely be used as a biological scaffold for the delivery of chondroprogenitors in cartilage healing (Vinod et al., 2019).

The immunological aspects of the application of al-PRP

Platelet-associated immunomodulatory molecules are mainly stored in platelet granules. Because of extremely high platelet numbers, amount of immune mediators are released to interact with various cells of the immune system. Platelet could express molecules and cytokines with immune functions including platelet-derived growth factor (PDGF), transforming growth factor-β (TGFβ), clusters of differentiation (CD) 40, CD154 and CXCL chemokine family to regulate different types of tissue damage (Semple et al., 2011). Furthermore, the growth factors and pro-inflammatory chemokines released by microvesicles are able to modulate the function of immune cells in type 2 diabetes (Cortez-Espinosa et al., 2017). Platelets are deeply involved in the innate and adaptive immune responses in the skin via interactions with leukocytes and the endothelium (Tamagawa-Mineoka, 2015).

At present, whether al-PRP will produce a severe immune response in patients is an important issue that has been widely studied by scholars worldwide. As Table 1 described, all reported clinical trials have demonstrated no serious allergic reactions to date. Most of clinical studies have indicated that al-PRP was topically applied to wound care with ABO matched donation. In an interesting vitro study, whether the differentiation of peripheral blood monocytes to dendritic cells via stimulation with granulocyte monocyte colony stimulating factor and interleukin-4 affected by PRP was investigated in order to define al-PRP mismatched for ABO and Rh antigens before clinical practice. The results suggested that al-PRP can promote the differentiation of monocytes to a regulatory anti-inflammatory population, possibly favouring wound healing (Papait et al., 2018). From animal experiments, it was also found that after intramuscular injection of allogeneic non-homologous PRP into rabbits, the number and proportion of CD4+ and CD8+ cells in the peripheral blood of the rabbits only increased slightly. Compared with before injection, the difference was not statistically significant, and there was no significant change in the histomorphology at the injection site (Zhang et al., 2013). Actually, the immunogenicity of al-PRP is negligible for the following reasons: First, al-PRP for wound healing is a topical product and does not enter the circulatory system. The human leukocyte antigen and human platelet antigen in the gel have limited contact with host antibodies. Second, after activation, the surface antigen structure and expression level of platelets are changed, and immunogenicity is reduced. Third, PRP can be completely degraded and absorbed within a few weeks, thus eliminating the possibility of a chronic immune response.

Al-PRP could easily obtain from healthy donor and solve the risk of poor standardization procedures in preparation of PRP. So far, homologous al-PRP has not been reported due to side effect problem. Hence, immunogenicity would not be a main obstacle to the application of al-PRP. Immunological aspect to topical application of al-PRP is little paid attention, further study is needed to identify its' immune role in patients with diabetes or elderly.

In a meta-analysis study, the results indicate that al-PRP can significantly accelerate DFU healing compared with control, and no adverse events have been observed in any al-PRP experimental results (Gong et al., 2022). However, there was not a large-scale, randomized, controlled trial in the treatment of chronic DFU with al-PRP for the meta-analysis (Du et al., 2021; Mastrogiacomo et al., 2022). Of course, the promotion of each new technology is both an opportunity and a challenge for the medical industry. At present, al-PRP has not been widely used in the clinic, and certain issues still need to be further addressed. Further research is needed to determine whether the growth factors in al-PRP will not be fully expressed due to the depletion of other unknown cofactors, as well as to determine if al-PRP quality will decrease with the prolongation of storage time. Future studies should also investigate whether the use of al-PRP to repair wounds can reduce the ulcer recurrence rate, and whether the combined use of al-PRP and surgical debridement and/or other noninvasive dressings can produce synergistic effects. In addition, exosomes derived from platelets has been successfully prepared in our previous study (Rui et al., 2021). Au- or al-PRP derived exosomes could be considered as a preferred choice for
“off-the-shelf” ideal bio-material dressing for DFU treatment. Finally, further studies are needed to identify the efficacy and safety of al-PRP in the treatment of chronic wounds with a large sample size of individuals.

Conclusion

The amputation and mortality of patients with DFU have sharply increased since the COVID-19 pandemic (Jeong et al., 2010). It is especially key to promote wound healing with a novel approach for elderly patients with chronic wounds. Al-PRP could be large-scale and with standardized production because of its easy preparation and application. Al-PRP is derived from healthy adult blood donors, the quality and content of platelets are stable, and the curative effect is reliable (Papait et al., 2018). The use of Al-PRP also avoids the health burden associated with autologous blood collection for the patients, thereby overcoming the shortcomings of au-PRP in clinical settings. In sum, Al-PRP may provide a new option in the treatment of chronic refractory wounds.

Author contributions

MH, TC, YL, PS, and WD planned and designed the study, and was the primary writer of the manuscript. MH, TC, PS, BD, and XG conducted experiments, participant recruitment and data collection. SR, JB, DA, SR, JB, DA, and YM contributed to the discussion, reviewed and drafted the manuscript. All authors read and approved the final manuscript. WD takes full responsibility for this study.

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Conflict of interest

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