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Mineral and Organic Matter Characterization of Density Fractions of Basalt- and Granite-Derived Soils in Montane California

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1. Introduction

There is ample evidence that soil mineralogy affects carbon cycling rates (e.g. Feller and Beare, 1997; Masiello et al., 2004; Torn et al., 1997). Humus may be physically protected from biological attack by (1) direct adhesion to clay surfaces via electrostatic interactions, hydrogen bonds, and cation bridges; (2) complexation with Fe and Al cations, amorphous oxides, and terminal atoms within the mineral structure; and (3) occlusion within mineral aggregates (Baldock and Skjemstad, 2000; Krull et al., 2003). In addition, the mineral matrix affects the stability of soil organic matter (OM) via the distribution of soil pores, water retention, O₂ diffusion, and the pH of soil water. But while methods for measuring the effects of texture, aggregation, and structure are relatively well-established (see Christensen, 1992 for a review), there is no generally accepted way of isolating discrete organo-mineral complexes, so that the effects of distinct minerals can be compared and ultimately extrapolated over a wide range of mineral and environmental conditions.

Density has routinely been used to separate soil OM fractions based on their degree of association with mineral particles (Baisden et al., 2002; Christensen, 1992; Golchin et al., 1995; Monnier et al., 1962). In addition, due to variations in the specific gravity of different minerals, it has also served as a proxy for mineral species in clay (Jaynes and Bigham, 1986; Sypcher and Young, 1979) and silt (Shang and Tiessen, 1998). In this study we evaluate density as a means of separating organo-mineral complexes, and use this method to explore the role of mineralogy on OM storage and cycling.

In an early study along the western flank of California’s Sierra Nevada, mafic soils were found to have higher levels of clay, carbon (C), and nitrogen (N), but lower levels of OM per unit of clay (Harradine and Jenny, 1958; Harradine, 1954). To learn more about the reasons
for these biogeochemical differences, and evaluate the direct influence of mineralogy on OM stability, we separated granitic and basaltic soils into density classes designed to isolate distinct mineral species and associated organic matter (Figure 1). In the resulting fractions, we used powder X-ray diffraction to identify the dominant mineral species, C/N and stable isotopes of C and N as indices of the degree of decomposition of the OM (Baisden et al., 2002; Ehleringer et al., 2000; Nadelhoffer and Fry, 1988), and 14C measurements to infer C turnover times (Trumbore, 1993; Trumbore and Zheng, 1996). Following this detailed analysis we conducted a profile analysis of the 0-2, 2-3, and >3 g cm⁻³ fractions of the basaltic soil.

![Fig. 1. Expected minerals and associated density classes for the granite and basalt soils.](image)

### 2. Materials and methods

#### 2.1 Study sites and sample selection

This study is based on three well-drained, unglaciated, forest soils: The Jimmerson, Musick, and Shaver series (Table 1). The dominant plant species are (1) ponderosa pine (*Pinus ponderosa*) and incense cedar (*Libocedrus decurrens*) at the Jimmerson site, (2) ponderosa pine, incense cedar, and manzanita (*Arctostaphylos manzanita*) at the Musick site, and (3) white fir (*Abies concolor*), sugar pine (*Pinus lambertiana*), and incense cedar at the Shaver site. The parent material of the Jimmerson soil is mapped as olivine basalt and that of the Musick and Shaver soils as granodiorite. The Jimmerson and Musick sites, with almost identical climates, are located just below the permanent winter snowline in a zone of rapid soil development, whereas the higher Shaver site is slightly cooler and wetter, and subject to a thick snowpack (Dahlgren et al., 1997). These soils represent a subset of two climate transects on which we are conducting a longer-term OM cycling study and were selected to maximise the variability in mineralogy across those transects and enable limited parent material and climate comparisons; i.e. basaltic Jimmerson versus granitic Musick and warmer Musick versus cooler Shaver.

![Table 1](table)

| Density (g cm⁻³) | Expected Minerals          | Density (g cm⁻³) | Expected Minerals          |
|------------------|----------------------------|------------------|----------------------------|
| 0                | (None)                     | 0                | (None)                     |
| 1                |                             | 1                | Noncrystalline minerals    |
| 2                | Halloysite                 | 2.0              | Illite, Halloysite         |
| 2.2              | Gibbsite, Vermiculite      | 2.2              | Gibbsite                   |
| 2.5              | Feldspars, Kaolinite       | 2.5              | Feldspars, Kaolinite, Montmorillonite |
| 2.6              | Quartz                     | 2.6              | Quartz                     |
| 2.7              | Muscovite, Hornblende      | 2.7              | Anorthite                  |
| 3.2              | Goethite, Hematite         | 3.2              | Augite, Hornblende         |

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Mineral and Organic Matter Characterization of Soil Density Fractions

Table 1. Characteristics of the study sites (Allardice et al. 1983; Begg et al. 1985).

| Soil series | Jimmerson | Musick | Shaver |
|-------------|-----------|--------|--------|
| Parent material | Pleistocene basaltic andesite | Jurassic granodiorite | Jurassic granodiorite |
| Mean annual precipitation \(cm\) | 91 | 94 | 102 |
| Mean annual temperature \(^\circ C\) | 13.9 | 11.7 | 8.9 |
| Elevation \(m\) | 774 | 1,240 | 1,780 |
| Classification | fine oxidic mesic Ultic Palexeralf | fine-loamy Ultic Haploxeralf | coarse-loamy Pachic Xerumbrept |
| County | Shasta | Fresno | Fresno |
| Township & Range | NE/4 S26, T31N, R1W, MMD | S.29, T10S, R25E, MMD | S.25, T10S, R25E, MMD |
| Latitude & Longitude | 40.52 ° N, 121.94 ° W | 37.02 ° N, 119.27 ° W | 37.03 ° N, 119.18 ° W |

These soils were originally sampled, analyzed, and archived during the California Cooperative Soil-Vegetation Survey – a 47-year long reconnaissance of California’s upland forests (Allardice et al., 1983; Begg et al., 1985). For the Survey, C was determined based on the mass increase of an ascarite CO\(_2\) trap after complete combustion under a constant stream of O\(_2\) at 900 ° C; N was determined by Kjeldahl digestion; and particle size distribution was determined by the pipette or hydrometer method.

2.2 Separation of density fractions

Density separations were performed on modern (1992) samples of the granitic soils and on archived (1961) samples of the basaltic soil. Detailed density separations were performed on the granitic Musick A1 and Shaver A2 horizons and on the A2 and Bt1 horizons of the basaltic Jimmerson soil (Table 2). Based on the specific gravity of the minerals commonly encountered in soils derived from each parent material, we isolated eight and twelve density fractions from the granite and the basalt samples, respectively (Figure 1). The <2 g cm\(^{-3}\) material of the granitic samples was separated into two fractions, but in anticipation of the possible presence of low-density allophanic minerals in the basaltic soil, the <2 g cm\(^{-3}\) material of the basaltic samples was separated into five fractions: <1, 1-1.4, 1.4-1.6, 1.6-1.8, and 1.8-2 g cm\(^{-3}\). Following on the results of the detailed density separation, the A1 and A3 Jimmerson soil horizons were subsequently separated into just three fractions: 0-2, 2-3, and >3 g cm\(^{-3}\).

Soil horizon samples were processed as outlined in Table 3. Successively higher density separates were obtained using a modification of the Golchin et al. (1994) method. Air-dry soils were sieved to < 2 mm, split into four ~6 g replicates, freeze-dried, and weighed into 50 ml polypropylene tubes to which ~30 ml of deionized water was added. Capped tubes were gently mixed by inverting six times and allowed to sit for at least one hour to fully wet the sample. After this period the tubes were mixed using a Vortex® mixer for ten seconds, immersed in an ice bath ice, and sonicated at 200 W for three minutes using a 350 W Branson™ sonicator with a 12.5 mm probe immersed to ~3 cm depth. Tubes were then
shaken at 180 RPM for ten minutes and centrifuged at 20,000 g to settle 1.05 g cm\(^{-3}\) and 0.2 \(\mu\)m diameter spherical particles. The floating material, corresponding to the 0-1 g cm\(^{-3}\) fraction, was isolated by decanting.

| Series  | Sampling date | Horizons | Depth cm | Texture | Clay % | Hue \(^a\) | Structure \(^b\) |
|---------|---------------|----------|----------|---------|--------|-----------|-----------------|
| Jimmerson | 1961          | A1       | 0-3      | loam    | 25     | 3:1       | weak, medium, subangular blocky |
|         |               | A2*      | 3-25     | clay loam | 33     | 3:1       | moderate, medium, angular blocky |
|         |               | A3       | 25-61    | clay loam | 36     | 3:1       | moderate, medium, angular blocky |
|         |               | B,1*     | 61-122   | clay     | 51     | 3:1       | subangular blocky |
| Musick  | 1992          | A1       | 0-7      | loam    | 20     | 1:1       | moderate, medium, granular |
| Shaver  | 1992          | A2       | 5-10     | sandy loam | 7      | 1:1       | weak, fine, granular |

\(^a\) The relative proportions of red and yellow.
\(^b\) The strength, size, and type, respectively, of structural aggregates.

Table 2. The properties of the soil horizons examined in this study.

| Step | Description |
|------|-------------|
| 1. Whole air-dry soil: | Coarse split <br>Gently crush with mortar and pestle |
| 2. Sieved soil: | Riffle split to obtain two replicates <br>Add replicates to centrifuge tubes <br>Freeze dry <br>Weigh |
| 3. Sequential density fractionation: | Adjust tube contents to target density <br>Vortex <br>Shake <br>Sonicate <br>Centrifuge <br>Isolate the floating fraction and rinse it <br>Repeat previous steps at successively higher densities |
| 4. Density fractions: | Freeze dry <br>Weigh <br>Photograph <br>Grind |

Table 3. The soil processing and density separation steps.
content of the Jimmerson Bt1 horizon made it extremely difficult to disengage and disperse the pellet from the bottom of the tube; for these samples, shaking and sonication times were increased as necessary). Following sonication, tubes were centrifuged to settle 0.2 µm diameter spherical particles with a density 0.05 g cm⁻³ higher than the solution. Following centrifugation the floating fractions were isolated: Floating fractions with densities less than 2 g cm⁻³ were decanted onto precombusted and tared quartz fiber filters and rinsed with 1 L deionized water. Higher density floating fractions were decanted into clean 50 ml tubes, diluted with enough water to allow them to settle, centrifuged, and rinsed 3-4 times with deionized water. All fractions were freeze-dried and weighed.

2.3 Characterization of density fractions

Morphology. Density fraction were observed and photographed through a Leica Stereo Zoom visual microscope using a Sony Cybershot digital camera.

Mineralogy. Single laboratory replicates were ground to <100 µm and placed in a Rigaku Geigerflex (Cu Kα) X-ray diffractometer. The diffraction intensity was recorded every 0.05° for 2.5 seconds by Theta software and the mineral species were identified based on the resulting powder X-ray diffraction (XRD) spectra (Barnhisel and Bertsch, 1989; Brindley and Brown, 1984).

Carbon, nitrogen, and stable isotopes. Two laboratory replicates were ground to <200 µm and analyzed on a Europa 20/20 continuous flow stable isotope ratio mass spectrometer at the Center for Stable Isotope Biogeochemistry, University of California, Berkeley. The C and N isotope ratios are reported as δ¹³C and δ¹⁵N values, where the standard is Pee Dee Belemnite carbonate for C (Kendall and Caldwell, 1998) and atmospheric N₂ for N. If replicate size was insufficient for a stable isotope measurement, replicates were either pooled or analyzed on a Carlo Erba CN Analyzer.

Radiocarbon. Single samples were weighed and sealed in evacuated Vycor tubes with 0.5 g Cu, 1 g CuO, and a strip of Ag foil (Boutton, 1991), combusted for three hours at 875 °C, then cooled (Minagawa et al., 1984). The evolved CO₂ was cryogenically purified under vacuum and measured manometrically. At the Center for Accelerator Mass Spectrometry, Lawrence Livermore National Labs, the CO₂ gas was reduced to graphite on which ¹⁴C was measured and reported as Δ¹⁴C (‰):

\[ \Delta^{14}C = (F-1) \times 1000 \]  

(1)

where

\[ F = \frac{^{14}C}{^{12}C}_{\text{sample}} \times e^{(1950-y)} \]  

(2)

F is the absolute fraction modern, the ratio between the ¹⁴C/¹²C of the samples, (normalized to δ¹³C=-25 ‰) and that of the international standard (95% of the activity of the NBS oxalic
acid standard in AD 1950 normalized to $\delta^{13}C=-19$‰). This value is corrected for the radioactive decay of the standard between 1950 and $y$, the year of the measurement (Stuiver and Polach, 1977). The radioactive decay constant, $\lambda$, is 1.21E-4 year$^{-1}$. The F values of pre-bomb atmospheric CO$_2$ correspond to ~1, values <1 indicate that radioactive decay has taken place, and values >1 indicate that “bomb carbon” has been incorporated into the sample.

### 2.4 The carbon turnover models

A mass balance model of soil organic C states that:

$$\frac{dC_i}{dt} = I_i - k_iC_i$$

where $C_i$ is the carbon inventory in pool i, $I$ is annual carbon inputs (mass year$^{-1}$), and $k_i$ is the first-order decomposition constant (year$^{-1}$). Similarly, the balance of $^{14}$C atoms in reservoir i, $F_iC_i$, can be described by:

$$\frac{d(F_iC_i)}{dt} = F_{atm}I - F_iC_i(k_i+\lambda)$$

where, $F_i$ is the $^{14}$C value of pool i, $F_{atm}$ is the $^{14}$C value of the atmosphere. Starting with the common assumption that the system is in steady state with respect to $^{12}$C, and hence, $I_i = k_iC_i$ (from Equation 3), we used two distinct approaches to translate the $^{14}$C values of density fractions into their turnover times ($T_i$), defined as $1/k_i$:

1. For the 1961 basalt soil fractions, which lack bomb-derived carbon, it is assumed that $F_{atm} = 1$ (pre-bomb conditions). Thus, from Equation 4, $F_{atm} = F_iC_i(k_i+\lambda)$, and

$$F_i = \frac{k_i}{k_i+\lambda}$$

2. For the 1992 granite soil fractions, which contain bomb carbon, the $^{14}$C value of the density fractions was translated into turnover times using a time-dependent box model. The time series was initialized in 1890, using Equation 5. In each subsequent year ($t$):

$$C_tF_t = I F_{atm}(t-lag) + C_{t-1}F_{t-1}(1-k-\lambda)$$

where $C_tF_t$ is the $^{14}$C inventory of a soil fraction in year $t$; $F_{atm}$ is the $^{14}$C value of the atmosphere (Levin and Hesshaimer, 2000), and lag is the average number of years that atmospheric carbon is retained in plant tissue before becoming part of the soil OM pool. The remaining terms are defined as above. Given that at steady state, $C_t = C_{t-1}$ (and $I_i = k_iC_i$ as above), we divide equation 6 by $C_t$ and obtain

$$F_t = F_{atm}(t-lag) k + F_{t-1}(1-k-\lambda)$$

By matching the modelled and measured F values for the year in which the soil was sampled, the decomposition constant, $k$ (and corresponding turnover time) can be extracted. This model assumes the fraction being modelled is homogeneous; i.e. that the decomposition rate is the same for every C atom of the population. While this assumption may be erroneous, the average turnover times derived using this approach allow for comparisons among soils and fractions.
3. Results

3.1 Morphology and mineralogy

- Whole soil

The whole soil morphological attributes (Table 2) indicate that the degree of development of the three soils increases from the Shaver, to the Musick, to the Jimmerson series. The Shaver soil had a low clay content and poorly developed structure (fragile granules 1-2 mm in diameter), whereas the Musick soil had higher clay and stronger structure (spherical granules 2-5 mm in diameter). The Jimmerson soil had the highest clay content, a distinctly redder hue (reflecting abundant iron oxides), and very strong structure (3-dimensional blocks 10-20 mm in diameter). As will become evident, these morphological differences, especially between the granite and the basalt soils, influenced the density separation results.

- Density fractions

Granitic soils. We found striking differences in the morphology and powder X-ray diffraction (XRD) patterns across the density fractions of the granite soils (Figure 2). We found a mixture of organic matter, fine roots, root bark, mineral grains, and charcoal in the 1-2 g cm$^{-3}$ fractions; decomposed organic matter, fine roots, and kaolin clays (kaolinite and halloysite) in the 2.0-2.2 g cm$^{-3}$ fractions; kaolins and very few roots in the 2.2-2.5 g cm$^{-3}$ fractions; dickite (a kaolin) and feldspars (principally anorthite, but also microcline and sandine) in the 2.5-2.6 g cm$^{-3}$ fractions; quartz (large peak at $2\theta = 26.6^\circ$, smaller peaks at 20.8, 50, and 59.9$^\circ$) and anorthite in the 2.6-2.7 g cm$^{-3}$ fractions; phlogopite (mica) and some anorthite or albite in the 2.7-3.2 g cm$^{-3}$ fraction; and magnesiohornblende grains in the >3.2 g cm$^{-3}$ fractions.

The main difference in the mineralogy of the Shaver and Musick was that, in addition to kaolins, we found gibbsite (peak at $2\theta = 18.3^\circ$) and hydroxyl-interlayered vermiculite in the 2.2-2.5 g cm$^{-3}$ Shaver fraction.

Basaltic soil. From 0 to 2 g cm$^{-3}$ in the A2 and Bt1 horizons of the Jimmerson soil, OM content and sample heterogeneity decreased steadily and the OM changed from recognizable plant parts to more disintegrated and decomposed material (Figure 3a). The diffraction patterns of the mineral density fractions, which were similar for A and B horizons, changed gradually with density (Figure 3b). Halloysite dominated the spectrum between 2.4 and 2.6 g cm$^{-3}$, and remained an important phase up to 2.9 g cm$^{-3}$. We found cristobalite in the 2.2-2.4 g cm$^{-3}$ fraction; feldspar and quartz grains between 2.4 and 2.9 g cm$^{-3}$ (A horizon) or between 2.6 and 2.9 g cm$^{-3}$ (B horizon); quartz in the 2.6-2.7 g cm$^{-3}$ fractions (its peak dwarfed all others and only the base is shown); and anorthite and/or albite in the 2.7-2.9 g cm$^{-3}$ fraction. The orange skins we observed in the 2.2-2.7 g cm$^{-3}$ fractions in the B horizon, which did not produce diagnostic XRD patterns, were presumably amorphous iron oxides. From 2.7 to 3.2 g cm$^{-3}$ a transition occurred from fine halloysite particles to large reddish and metallic silver particles (A horizon) or to red/yellow particles (B horizon). Above 3.2 g cm$^{-3}$ goethite and hematite phases dominate. The peak ratio at 35.6$^\circ$ versus 33.2$^\circ$ signifies relatively more goethite in the B horizon, which agrees with the difference in hue: More red in the A horizon and more yellow in the B horizon.
Density fractions are labeled in g cm\(^{-3}\). Powder X-ray diffraction spectra are shown for the Musick soil only - with the exception of the 2.2-2.5 g cm\(^{-3}\) fraction, where the Shaver soil spectrum is in gray. Diagnostic peaks for Halloysite (Y), Kaolinite (K), Vermiculite (V), Quartz (Q), Anorthite (A), Sillimanite (S), Dickite (D), Phlogopite (P), and Magnesiohornblende (MgH) are labeled. See text in Methods and in Results for more details.

Fig. 2. Morphology and mineralogy of the density fractions obtained from the two granite soils, Musick (left hand side) and Shaver (right hand side)
Fig. 3a. Morphology and mineralogy of the low-density mineral-free fractions obtained from the A2 and Bt1 horizons of the Jimmerson (basalt) soil. Density fractions are labelled in g cm$^{-3}$. Powder X-ray diffraction spectra are shown for the A2 horizon in black and for the Bt1 horizon in gray.
Fig. 3b. Morphology and mineralogy of the high-density mineral-associated fractions obtained from the A2 and Bt1 horizons of the Jimmerson (basalt) soil. Density fractions are labelled in g cm\(^{-3}\). Powder X-Ray diffraction spectra are shown for the A2 horizon in black and for the Bt1 horizon in gray. Peaks for Halloysite (Y), Cristobalite (C), Quartz (Q), Anorthite (A), Goethite (G), and Hematite (H) are labeled. See text in Methods and in Results for more details.
3.2 Chemistry

- Whole soil

The % C profiles for the three soils were very similar, but the C/N and % clay profiles were quite distinct (Figure 4). There was no clear association between % C and clay values. The % clay and C/N ratios were inversely correlated, however, indicating that clay has a positive effect on the overall state of OM decay. The overall linear R-square=0.88, p<0.0001, n=10. For the Musick and Jimmerson soils (n=4 sampling depths each) the R² and p-values for the linear regression of C/N on % Clay are 0.99 and <0.004 for both cases. For the Shaver soil, with only two sampling depths, a regression analysis was not warranted. The whole soil δ¹³C and δ¹⁵N values increased with depth, a trend that has been observed in a number of soils (e.g. Nadlehofer and Fry 1988).

![Fig. 4. Depth profiles of whole soil a. % C, b. C/N, and c. % clay for the Jimmerson, Musick, and Shaver soils (Allardice et al. 1983; Begg et al. 1985). The basalt soil is represented by closed symbols and the granite soils are represented by open symbols.](image)

- Density fractions

The complete C, N, and isotope results of the detailed density separations are tabulated in Tables 4 and 5. Table 6 shows the results of the ANOVAs and subsequent multiple comparison tests among density fractions for each soil. To compare trends in the density fractions across sites, some of the data from tables 4 and 5 are shown in figure 5, where stable isotope values were plotted as Δ, the difference between the soil fraction δ values and the root δ values; this correction accounts for site differences in the isotopic composition of the plant inputs.

The 0-1 g cm⁻³ density class was distinguished by high % C, low C:N ratios and high Δ¹⁵N values. In the different fractions of the 1-2 g cm⁻³ continuum of the basaltic Jimmerson soil, % C and C/N decreased while stable isotope values generally increased (Table 6). The presence of charcoal in the 1-1.6 g cm⁻³ density classes was reflected in C/N ratios > 40 (Table 5, Figure 5b, c). In all three soils the 2.0-2.4 g cm⁻³ density fractions corresponded to a transition between less decomposed mineral-free and more decomposed mineral-bound organic matter where % C and C/N decreased- and δ¹³C and δ¹⁵N increased. Higher stable isotope values and lower C/N ratios occurred at densities greater than 2.2 g cm⁻³, a class dominated by minerals with little associated OM. The primary mineral-dominated fractions, with densities > 2.5 g cm⁻³, were relatively unimportant with respect to C and N storage.
Table 4. Granite soil density fraction characteristics. Proportion of total mass (Mt), proportion of total nitrogen (Nt), % C, % N, C/N, \( \delta^{13}C \), \( \delta^{15}N \), and \( 14C \) (F) for the bulk soil, roots, and mineral/density fraction.
| Density fraction /g cm$^{-3}$ (mineral in fraction) | Mean        | S.E.   | Mean        | S.E.   | Mean        | S.E.   | Mean        | S.E.   | Mean        | S.E.   | Mean        | S.E.   |
|-----------------------------------------------|-------------|--------|-------------|--------|-------------|--------|-------------|--------|-------------|--------|-------------|--------|
| A2 horizon                                    |             |        |             |        |             |        |             |        |             |        |             |        |
| Bulk                                          | 2.0         | 0.0    | 0.11        | 0.00   | 18.2        | 0.4    | -25.0       | 0.1    |             |        |             |        |
| Roots                                         | 38.0        | 1.6    | 1.08        | 0.03   | 35.1        | 9.5    | 27.0        | 0.0    |             |        |             |        |
| 0-1 (free OM)                                 | 0.000       | 0.00   | 0.01        | 0.00   | 43.9        | 1.5    | 11.5        | 0.07   | 38.4        | 2.7    | -27.8       | 0.1    |
| 1-1.4 (free OM)                               | 0.003       | 0.00   | 0.07        | 0.04   | 48.8        | 0.7    | 0.64        | 0.03   | 76.9        | 3.3    | -27.7       | 0.1    |
| 1.4-1.6 (free OM)                             | 0.008       | 0.00   | 0.15        | 0.00   | 45.0        | 0.0    | 0.93        | 0.06   | 48.6        | 3.0    | -26.0       | 0.0    |
| 1.6-1.8 (free OM)                             | 0.006       | 0.00   | 0.10        | 0.02   | 35.2        | 0.4    | 0.89        | 0.03   | 39.5        | 0.5    | -25.7       | 0.1    |
| 1.8-2.0 (free OM)                             | 0.007       | 0.00   | 0.08        | 0.01   | 23.6        | 1.1    | 0.86        | 0.02   | 27.6        | 1.4    | -25.8       | 0.0    |
| 2.0-2.2 (Halloysite)                          | 0.006       | 0.00   | 0.03        | 0.00   | 9.5         | 0.7    | 0.45        | 0.01   | 21.0        | 1.7    | -25.4       | 0.1    |
| 2.2-2.4 (Halloysite)                          | 0.064       | 0.01   | 0.07        | 0.00   | 2.2         | 0.6    | 0.18        | 0.01   | 12.9        | 3.3    | -24.6       | 0.2    |
| 2.4-2.6 (Halloysite)                          | 0.601       | 0.02   | 0.28        | 0.01   | 0.9         | 0.0    | 0.08        | 0.00   | 11.0        | 0.4    | -23.7       | 0.1    |
| 2.6-2.7 (H+Quartz)                            | 0.117       | 0.01   | 0.02        | 0.00   | 0.4         | 0.0    | 0.04        | 0.00   | 10.6        | 0.4    | -23.8       | 0.1    |
| 2.7-2.9 (H+Anorthite)                         | 0.082       | 0.00   | 0.02        | 0.00   | 0.5         | 0.0    | 0.04        | 0.00   | 12.8        | 1.1    | -23.9       | 0.0    |
| 2.9-3.2 (H+Anorthite)                         | 0.031       | 0.00   | 0.01        | 0.00   | 0.7         | 0.0    | 0.06        | 0.00   | 12.2        | 0.4    | -23.9       | 0.1    |
| 3.2+ (Iron oxides)                            | 0.097       | 0.00   | 0.03        | 0.00   | 0.6         | 0.0    | 0.03        | 0.00   | 20.4        | 1.7    | -23.9       | 0.1    |
| Bulk recovered                                | 1.023       | 0.01   | 0.87        | 0.05   | 0.87        | 0.03   | 1.8         | 0.1    | 15.9        | 1.0    | -21.7       | 1.2    |
| Bt1 Horizon                                   |             |        |             |        |             |        |             |        |             |        |             |        |
| Bulk                                          | 0.4         | 0.0    | 0.03        | 0.00   | 14.0        | 0.4    | -23.4       | 0.1    |             |        |             |        |
| 0-1 (free OM)                                 | 0.000       | 0.00   | -0.02       | 0.02   | 49.9        | 1.1    | 0.76        | 0.16   | 68.8        | 14.7   |             |        |
| 1-1.4 (free OM)                               | 0.001       | 0.00   | 0.05        | 0.05   | 51.3        |        |             |        |             |        |             |        |
| 1.4-1.6 (free OM)                             | 0.001       | 0.00   | 0.06        | 0.02   | 41.6        | 1.5    | 0.45        |        | 95.8        |        | -25.7       |        |
| 1.6-1.8 (free OM)                             | 0.000       | 0.00   | -0.03       | 0.01   | 36.2        | 0.6    | 0.40        | 0.04   | 90.8        | 10.0   |             |        |
| 1.8-2.0 (free OM)                             | 0.001       | 0.00   | 0.05        | 0.00   | 34.0        | 0.64   |             |        | 53.3        |        |             |        |
| 2.0-2.2 (Halloysite)                          | 0.000       | 0.00   | 0.00        | 0.00   | 11.8        | 0.40   |             |        | 29.7        |        |             |        |
| 2.2-2.4 (Halloysite)                          | 0.010       | 0.00   | 0.02        | 0.00   | 0.8         | 0.2    | 0.06        | 0.01   | 12.5        | 3.4    | -25.7       |        |
| 2.4-2.6 (Halloysite)                          | 0.083       | 0.00   | 0.52        | 0.02   | 0.69        | 0.10   | 0.3         | 0.03   | 9.4         | 0.4    | -22.9       | 0.0    |
| 2.6-2.7 (H+Quartz)                            | 0.053       | 0.004  | 0.04        | 0.00   | 0.3         | 0.05   | 0.03        | 0.00   | 11.0        | 0.4    | -24.0       | 0.0    |
| 2.7-2.9 (H+Anorthite)                         | 0.035       | 0.001  | 0.03        | 0.01   | 0.3         | 0.1    | 0.02        | 0.00   | 13.4        | 3.0    | -24.6       | 0.1    |
| 2.9-3.2 (H+Anorthite)                         | 0.018       | 0.001  | 0.02        | 0.00   | 0.1         | 0.05   | 0.03        | 0.00   | 17.8        |        |             |        |
| 3.2+ (Iron oxides)                            | 0.012       | 0.001  | 0.12        | 0.00   | 0.05        | 0.00   | 0.4         | 0.00   | 36.7        | 1.1    | -23.3       | 0.1    |
| Bulk recovered                                | 0.985       | 0.004  | 0.85        | 0.00   | 0.85        | 0.00   | 0.4         | 0.00   | 9.4         | 1.0    | -19.9       | 1.2    |

Table 5. Jimmerson (basalt) A2 and Bt1 horizon density fraction characteristics. Proportion of total C (Ct), proportion total N (Nt), % C, % N, C/N, $\delta^{13}C$, $\delta^{15}N$, and $\delta^{14}C$ (F) for the bulk soil, roots, and.
Fig. 5 a-d. The carbon and nitrogen chemistry of the A horizons plotted against the midpoint of each density fraction. The basalt soil is represented by closed symbols and the granite soils are represented by open symbols.

The proportion of total C (Ct) in each density fraction, i, is Ct(i) = %C(i) × Ml(i) ÷ %C_{bulk(avg)}, where, x is the laboratory replicate for i. Reported Ct values are the mean of two laboratory replicates. Nt is calculated analogously. Where reported, the standard error (s.e.) for Mt, C, N, δ¹³C, and δ¹⁵N is the absolute standard error of the mean of laboratory replicates (n=4 for mass fractions and n=2 for others). Standard errors for C/N, Ct, Nt, and recovered bulk are calculated using gaussian error propagation. The error for δ¹⁴C is the analytical error reported by CAMS after rerunning one sample several times. The 0-1 and 1-2 g cm⁻³ fractions as well as the >2.5 g cm⁻³ were composited for δ¹⁴C analysis.

The granitic soils yielded density fractions that clearly differed from one another with respect to mineral and OM composition. Radiocarbon values decreased as a linear function of density, albeit with different slopes for Musick and Shaver (Table 7). The 2 g cm⁻³ boundary separating C cycling times of less than and greater than 100 years but differences across higher density fractions were relatively small. The only outstanding C turnover pool corresponded to the mica-dominated fraction of the Musick soil, with a ~600 year mean residence time. The 1992 granitic soil samples reflect the incorporation of δ¹⁴C from nuclear weapons, whereas the 1961 basalt soil samples do not. As a result we cannot directly compare these two sets of δ¹⁴C values and derived turnover times.
| Soil series | Horizon | Density / gcm$^{-3}$ | C / % | N / % | CN | $\delta^{13}$C / ‰ | $\delta^{15}$N / ‰ |
|-------------|---------|----------------------|-------|-------|----|----------------|----------------|
| Jimmerson A2 | 0-1     | a                    | b     | a     | b  | a               | bc             |
|             | 1.4     | a                    | c     | a     | a  | a               | -              |
|             | 1.4-1.6 | b                    | b     | b     | b  | a               | -              |
|             | 1.6-1.8 | c                    | b     | b     | bc | a               | -              |
|             | 1.8-2   | d                    | b     | c     | bc | a               | abc            |
|             | 2.2-2   | e                    | d     | cd    | c  | bc              |
|             | 2.2-2.4 | f                    | e     | d     | d  | cd              |
|             | 2.4-2.6 | f                    | e     | d     | e  | d               |
|             | 2.6-2.7 | f                    | e     | d     | e  | cd              |
|             | 2.7-2.9 | f                    | e     | d     | e  | cd              |
|             | 2.9-3.2 | f                    | e     | d     | e  | cd              |
|             | 3.2+    | f                    | e     | cd    | e  | cd              |

**ANOVA P values:**

-0.0001  0.0001  0.0001  0.0001  0.0001

| Jimmerson Bt1 | 0-1     | a                    | a     | a     | a  | -               | -              |
|              | 1-1.4   | a                    | -     | a     | a  | -               |
|              | 1.4-1.6 | b                    | ab    | a     | b  | a               |
|              | 1.6-1.8 | c                    | ab    | ab    | -  | -              |
|              | 1.8-2   | c                    | ab    | ac    | -  | -              |
|              | 2.2-2   | d                    | ab    | ab    | -  | -              |
|              | 2.2-2.4 | e                    | b     | b     | b  | b               |
|              | 2.4-2.6 | e                    | b     | b     | e  | c               |
|              | 2.6-2.7 | e                    | b     | bc    | d  | c               |
|              | 2.7-2.9 | e                    | b     | bc    | c  | b               |
|              | 2.9-3.2 | e                    | b     | bc    | -  | -              |
|              | 3.2+    | e                    | b     | bc    | e  | c               |

**ANOVA P values:**

-0.0001 | 0.0002 | 0.0002 | 0.0001 | 0.0001

| Musick A1 | 0-1     | b                    | a     | b     | a  | ab              |
|           | 1-2     | a                    | b     | a     | a  | a               |
|           | 2-2.2   | c                    | c     | b     | b  | bc              |
|           | 2.2-2.5 | d                    | d     | c     | c  | c               |
|           | 2.5-2.6 | e                    | e     | c     | c  | c               |
|           | 2.6-2.7 | e                    | e     | cd    | b  | bc              |
|           | 2.7-3.2 | e                    | e     | d     | b  | b               |
|           | 3.2+    | e                    | e     | cd    | b  | d               |

**ANOVA P values:**

-0.0001 | 0.0001 | 0.0001 | 0.0001 | 0.0001

| Shaver A2 | 0-1     | a                    | a     | a     | ab | bc              |
|           | 1-2     | a                    | b     | a     | ab | a               |
|           | 2-2.2   | c                    | b     | b     | ab | cde             |
|           | 2.2-2.5 | c                    | c     | b     | a  | de              |
|           | 2.5-2.6 | c                    | c     | b     | ab | cde             |
|           | 2.6-2.7 | c                    | c     | b     | ab | b               |
|           | 2.7-3.2 | c                    | c     | b     | a  | bce             |
|           | 3.2+    | c                    | c     | b     | b  | e               |

**ANOVA P values:**

-0.0001 | 0.0001 | 0.0153 | <0.0001

Table 6. Results of the ANOVA and Tukey-Kramer Honestly Significant Differences test conducted for each soil horizon and chemical analysis. P-values are for the overall ANOVA and within each of these groupings. Different letters denote significant differences among all density fractions for alpha=0.050.
In both the A2 and Bt1 horizons of the basaltic soil, high levels of clay, iron oxides, and resulting aggregation hindered the segregation of its constituent minerals. Morphology and mineralogy changed very gradually with density, such that the seven mineral density fractions yielded only two discrete organo-mineral fractions dominated by halloysite versus iron oxides. In our attempts to disperse the soil minerals, we used relatively high levels of ultrasonic energy on all samples, and this contributed to substantial losses of dissolved C (which, by difference, is $^{13}$C-depleted) into the polytungstate density solution (Tables 4, 5). Most of the recovered C and N corresponds to the mineral-free (<2 g cm$^{-3}$) and kaolin-bearing (2-2.6 g cm$^{-3}$) fractions.

| Density fraction / g cm$^{-3}$ | Radiocarbon values / FM | Musick | Shaver |
|-------------------------------|-------------------------|--------|--------|
| 0-2                           | 1.137                   | 1.084  |        |
| 2-2.2                         | 1.064                   | 1.039  |        |
| 2.2-2.5                       | 1.068                   | 1.049  |        |
| 2.5-2.6                       | 1.038                   | 1.045  |        |
| 2.7-3.2                       | 0.952                   | 1.044  |        |

Table 7. Radiocarbon values of the granitic soil density fractions. For Musick, $^{14}$C (FM) = 1.235 - 0.084 density midpoint (g cm$^{-3}$), with $R^2=0.85$, $p=0.026$, $n=5$ and for Shaver, $^{14}$C (FM) = 1.099 - 0.021 density midpoint (g cm$^{-3}$), with $R^2=0.75$, $p=0.059$, $n=5$.

3.3 Jimmerson soil profile analysis

In both A2 and Bt1 horizons the large 2.4-2.6 g cm$^{-3}$ halloysite fraction captured over 25 % of the total C; with the largest pool of remaining C in either the 1.4-1.6 g cm$^{-3}$ mineral-free fraction (A2 horizon) or the >3 g cm$^{-3}$ goethite/hematite fraction (Bt1 horizon). To avoid costly AMS $^{14}$C measurements on fractions with minor quantities of C we limited the $^{14}$C measurements to these three fractions and the bulk soil. The weighted $^{14}$C value of these three fractions (free, halloysite, and iron-oxide) is not very different from that of the bulk soil, and this provides some assurance that we did not miss a substantial, distinct, C pool.

To complete the Jimmerson soil profile analysis (Figure 6) we separated the remaining A1 and A3 horizons into just three fractions, 0-2, 2-3, and >3 g cm$^{-3}$, corresponding to free, halloysite, and iron oxide-bound OM, respectively. The high C/N values in the mineral-free fraction of the A2, A3, and Bt1 horizons indicate the presence of charcoal. Before deriving turnover times we made a conservative adjustment of the $^{14}$C values of the free fractions with C/N > 40 by assuming that the charcoal has 85 % C, 0 % N, and a $^{14}$C value equal to that of the slowest cycling pool of each horizon. In the halloysite fraction, the proportion of total C, as well as its $^{14}$C-derived turnover time and stable isotope value increased steadily and predictably with depth (i.e. deeper soil → slower turnover time → higher $\delta^{15}$N and $\delta^{13}$C). In contrast, the isotope patterns associated with the iron-rich fractions do not vary as regularly. From the A1 to the A3 horizons the $\delta^{15}$N values were negatively correlated with turnover time (i.e. slower turnover time → less $^{15}$N) and only in the B horizon did both values increase.
Mineral and Organic Matter Characterization of Soil Density Fractions

4. Discussion

The objectives of this study were to use density to separate discrete organo-mineral complexes from both granite and basalt-derived soils so as to evaluate the direct effect of parent material, and resulting soil mineralogy, on OM dynamics. Our results allow us to compare (1) the chemistry and turnover time of different density fractions within a given soil sample and (2) the chemistry of A horizons from soil with differing parent material and/or climate. In addition, (3) we examined depth related trends in chemistry and turnover of the basalt soil profile.

4.1 Evaluation of the effectiveness of the density separation method

The absence of broad bands around \( \theta = 5.9, 26.2, \text{ and } 35.9 \) confirmed that the basaltic soil did not contain allophane or imogolite phases; its low density fractions were all...
mineral-free. The frequently high $\Delta^{15}N$ and low C/N values of the 0-1 g cm$^{-3}$ are probably associated with the increased presence of ectomycorrhizal fungal products (Hogberg, 1997), such as spores and hyphae corresponding to the black spheres and spongy material visible in figures 2 and 3a. The entire 1-2.4 g cm$^{-3}$ interval of both soil types represented a continuum of alteration from fresh litter, with high % C, N, and C/N, and low $\Delta^{13}C$ and $^{15}N$, to highly humified kaolin-associated OM at the other end of the spectrum (Ehleringer et al., 2000; Nadelhoffer and Fry, 1988). This result, which was true for both the basalt and the granite samples, is consistent with both Golchin et al. (1995) and Baisden et al. (2002), and suggests that the quantity of OM associated with the mineral surface is inversely related to its degree of decomposition. The corresponding $^{14}C$ analyses indicate, however, that neither of these two parameters is useful for predicting the mean residence time of the organic C.

Previous radiocarbon work on the granite soils has shown that acid/base hydrolysis of the >2 g cm$^{-3}$ mineral fraction can leave a substantial $^{14}C$-depleted residue (Trumbore and Zheng 1996). The only density separates obtained in this study that are large enough to account for this pool of distinctly older C are the secondary clay-dominated fractions. If some of the C in these fractions is much older, the average residence times we report actually represent a heterogeneous mixture of faster and more slowly cycling C components. This condition violates the assumption of homogeneity inherent in our C cycling model (Equation 7), and the turnover times of the kaolin-rich, and possibly other, fractions should therefore be cautiously interpreted as average $^{14}C$ ages of mixed C pools. We conclude that acid/base hydrolysis may be the best way to separate very long-lived C from these clay fractions. But we also note that the sodium pyrophosphate used in the base hydrolysis dissolves metastable iron chelates that can bind long-lived C (McKeague et al., 1971; Trumbore and Zheng, 1996). Thus, in iron-rich samples such as the Jimmerson B horizon, the very heavy fraction (> 3 g cm$^{-3}$) may be appropriate for separating this potentially important pool.

4.2 Organic matter dynamics as a function of parent material and climate

Harradine and Jenny (1958) observed that basaltic soils in California have more OM than granitic soils. In this study we saw no difference in the total C % profiles of the basaltic Jimmerson and the granitic Musick soils (Figure 4) and when plotted as a function of depth, the mineral-bound C per unit clay or the stable isotope enrichment values in the clay-dominated density fractions were indistinguishable (Figure 7, 8). The observations by Harradine and Jenny (1958) may have been due to the presence of allophanes in younger basaltic soils that likely were part of their data set, since allophane minerals can store large amounts of C. In this study, the Jimmerson soil we examined is on an old relatively stable landscape, and any allophanes initially formed have weathered to more crystalline secondary mineral phases, ultimately making the difference between the granitic and basaltic parent materials less striking.

The big mineralogical difference between basaltic and granitic soils in our study is in the amount of iron oxides, minerals which are known to coat kaolin clays and increase their specific surface area and OM stabilizing potential (e.g. Baldock and Skjemstad, 2000).
Fig. 7. Mineral-bound C normalized for clay by sampling depth. The basalt soil is represented by closed symbols and the granite soils are represented by open symbols. Standard error values are based solely on the error associated with the mineral-associated C (n=2), the error associated with % clay is unknown (n=1).

Fig. 8. Enrichment of the kaolin-dominated (2-2.6 g cm\(^{-3}\)) \(^{13}\)C and \(^{15}\)N values (\(\Delta^{13}\)C=\(\delta^{13}\)C\(_{\text{soil}}\)-\(\delta^{13}\)C\(_{\text{roots}}\) and \(\Delta^{15}\)N=\(\delta^{15}\)N\(_{\text{soil}}\)-\(\delta^{15}\)N\(_{\text{roots}}\)) as a function of depth. The basalt soil is represented by closed symbols and the granite soils are represented by open symbols. Standard error bars reflect the simple rules for propagation of error of sums and differences.

But neither the iron-coated kaolin fractions nor the iron oxide fractions had particularly high concentrations of OM. We conclude that the effect of iron oxides in C stabilization occurs indirectly through the formation of highly stable aggregates and is also relatively more pronounced in B horizons, where iron is most abundant.

The Musick and Jimmerson sites lie just below, and the Shaver soil just above, the permanent winter snow line. The Shaver soil was the least \(^{15}\)N-enriched at all depths,
possibly due to more effective retention of mineral forms of N or a preferential loss of organic N (Amundson et al., 2003) during snowmelt at that site. Intense leaching at snowmelt retards the accumulation of kaolinite and promotes the formation of vermiculite (Dahlgren et al., 1997), with about fifty times more specific surface area. This mineral difference led to higher levels of mineral-bound C per unit clay in the Shaver than in the Musick soil (Figure 8). This effect is not an artefact of sampling depth, which would have had the opposite effect, but may be enhanced by the direct effect of temperature on biodegradation.

Mineralogy and/or climate may also influence C turnover, which was slightly slower in the Shaver than the Musick clay fractions (142-167 versus 110-115 years, Table 7). Wattel-Koekkoek and Buurman (2004) report similarly modest differences in the cycling rates of OM on high activity smectite versus low activity kaolinite species, and we suspect that much larger differences in clay activity, such as between noncrystalline and crystalline species, are required to highlight the direct effect of mineralogy on C cycling rates (Torn et al. 1997).

4.3 The Jimmerson soil mineralogy and profile development

The mineral analysis of the Jimmerson soil revealed an absence of allophanes and the presence of quartz. The occurrence of quartz and cristobalite on this broad flat lava flow suggest that either (1) quartz grains arrived by aeolian transport from a nearby sandstone formation; or (2) the parent material, which is mapped as a broad Pleistocene vesicular olivine basalt lava flow (MacDonald and Lydon, 1972), may in fact be a basalt-andesite intergrade.

It has been observed (Allen and Hajek, 1989) that halloysite-dominated soils are rare, however the XRD data for the Jimmerson soil clearly indicate that halloysite is the main secondary mineral in both the A and B horizons. Volcanic glass and allophanes are precursors to halloysite, which can also precipitate from the desilication of smectites (2:1 layer silicates) and may subsequently recrystallize to produce kaolinite (Allen and Hajek, 1989; Hendricks and Whittig, 1968; Southard and Southard, 1989). The absence of allophanes is related to the relative age of the soil. This soil, which is not glaciated and has been undergoing weathering for thousands of years, is presumably at an advanced stage of development that lies between the allophanic (or smectitic) and kaolinitic endmembers. As noted above, this mineralogical stage has muted parent material effects on soil C storage that might be expected in less developed soils.

The depth profile of the >3 g cm\(^{-3}\) iron oxide fraction in the Jimmerson soil exhibited an intriguing pattern for which we present two interpretations: (1) The suite of organic compounds associated with the goethite and hematite may change with depth – a hypothesis that could be tested using \(^{13}\)C NMR spectroscopy (e.g. Golchin et al.1994); and/or (2) young humus with relatively high stable isotope values may be preferentially ‘cheluviated’ from the A1 to the A2 of horizon. Because the crystalline iron oxides we identified in the XRD patterns are almost certainly immobile, a corollary to this hypothesis is that the > 3 g cm\(^{-3}\) fractions also captured non-crystalline iron phases, an assumption that could be tested by directly examining the \(^{13}\)C and \(^{15}\)N contents of OM in various horizons.
mineralogical phases separated by sodium pyrophosphate and ammonium oxalate extractions (e.g. Masiello et al., 2004).

5. Conclusions

In this study we examined three northern California forest soils: the Jimmerson (warm, basaltic), the Musick (warm, granitic), and the Shaver (cool, granitic). As expected, we found levels of clay, iron, and aggregation to be highest in the warmer basaltic soil and lowest in the cooler granitic soil. But although clay content was correlated with degree of humification as indicated by C/N ratios, there was no difference in % C depth profiles and total C storage across these three soils. In the A horizons, where most OM resides, the iron oxide fraction of the basaltic soil was associated with a very minor proportion of the OM. Overall, it was climate differences between the two granitic soils, rather than differences in the mineral composition of the parent materials, that resulted in the most obvious mineral effect on C. In the cooler granitic soil, more intense leaching has promoted the formation of more reactive clay and hence more mineral-bound C per unit clay.

The impetus of this study was to evaluate the mineral density separation method in the context of understanding the role of parent material and mineralogy in organic matter storage and turnover. Density is clearly appropriate for separating minerals with different specific gravities – such as allophanes, crystalline silicate clays, and iron oxides – and it is more effective when aggregation is weak – such as in ashy or sandy soils. In part because they span a relatively wide range of densities, primary minerals are easy to separate. They are also easy to subsequently identify. They do not, however, harbour much OM, so their utility in OM studies may be limited to special cases, such as ammonium fixation by altered mica.

Conversely, distinct silicate clay species, which are associated with high OM content, have a relatively high degree of overlap in their specific densities, especially in light of their OM as well as iron oxide coatings. For example, this study corroborates the notion that the density of organo-clay complexes clearly reflects the OM:clay ratio (as well the degree of alteration of the clay-associated OM). But, although the slowest cycling C is indeed associated with clays, as most studies indicate, so is fast cycling C, and separating these two pools requires additional chemical treatment. In conclusion, it will often be necessary to follow preliminary density separations with complementary techniques.

If certain groups of mineral species occur and function together, as is the case with kaolins and their likely ferric coating, there may be little value in attempting to separate them. In fact, our density separation may have lifted some colloidal ferric coatings from the kaolin surfaces and combined them with crystalline iron oxides, thereby obscuring our ability to distinguish the effect of these ecologically distinct species on soil OM patterns. We note, however, that the density separation technique can be used to exploit differences in particle size as well as density. Iron species can range from sand sized macro crystals to colloidal coatings and can be separated on the basis of their size – another determinant of settling velocity.

Finally, although sonication is required in order to disaggregate the constituent minerals of most soils, when the aggregation is strong it may be impractical to deliver enough energy to accomplish this. For example, the sonication of the basalt soil was insufficient to fully lift the
iron coatings off the kaolin, but excessive insofar as it resulted in a significant amount of OM becoming dissolved in the sodium polytungstate solution. This kind of trade-off needs to be evaluated on a case by case basis.

This study highlights advantages and difficulties associated with the density fractionation approach. The success of the density separation is highly dependent on the individual soil characteristics - such as aggregation and the degree of overlap in the densities of the constituent clay minerals - and on the purpose of the separation, that is how the OM attributes of interest are related to the mineral density differences. We conclude that the principle of separating intact soil into discrete organo-mineral complexes on the basis of gravity is reproducible and effective, but the organic matter pools associate with these fractions are still heterogeneous with respect to composition and turnover time.

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