Vitamin D receptor polymorphism rs2228570 is significantly associated with risk of dyslipidemia and serum LDL levels in Chinese Han population

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Abstract

Background: The goal of this study was to determine if vitamin D receptor (VDR) gene polymorphisms underlie susceptibility to dyslipidemia in a Chinese Han population.

Methods: Three tag single nucleotide polymorphisms (SNPs) (rs11574129, rs2228570, and rs739837) were genotyped using TaqMan assays to determine VDR SNP associations with dyslipidemia. We genotyped 877 cases of dyslipidemia from a normotensive, non-diabetes mellitus population and 1822 non-dyslipidemia subjects in a stage I study. In a follow-up stage II study, we included a larger sample of 3124 controls and 1679 cases with dyslipidemia. Finally, we explored the potential molecular mechanism for the SNP associations using molecular modeling analysis.

Results: We found a significant association between SNP rs2228570 and dyslipidemia in the additive (adjusted odds ratio (OR) = 1.255, 95% Confidence Interval (CI) = (1.118–1.409), \( P < 0.001 \)), dominant (OR = 1.384, 95% CI = 1.384 (1.136–1.6), \( P = 0.001 \)) and recessive models (OR = 1.356, 95%CI = 1.1–1.671, \( P = 0.004 \)) in stage I. We further established that the rs2228570 variant was significantly associated with dyslipidemia in the additive (adjusted OR = 1.146, 95% CI = 1.053–1.247, \( P = 0.002 \)), dominant (OR = 1.184, 95%CI = 1.018–1.376, \( P = 0.028 \)) and recessive models (OR = 1.209, 95%CI = 1.064–1.374, \( P = 0.004 \)) in stage II. The TT genotype was significantly higher (4.93 ± 0.75 mmol/L) compared to the TC (4.67 ± 0.47 mmol/L) or CC (4.66 ± 0.44 mmol/L) genotypes (\( P = 0.01 \)) in cases with elevated low-density lipoprotein cholesterol (LDL-C) levels. In contrast, the cases with the TT genotype had significantly lower serum 25(OH)D levels (18.43 ± 5.04 ng/ mL) compared to the TC (26.24 ± 4.16 ng/mL) and CC (36.76 ± 8.10 ng/ mL) genotypes (\( P < 0.001 \)). Multivariable linear regression analysis indicated that the rs2228750 genotype significantly correlated with serum low-density lipoprotein-C (LDL-C) levels in cases with dyslipidemia. Using molecular modeling analysis, we further found that the rs2228570 variant changed the structure and the stability of VDR and altered the binding energy of its ligand.

Conclusions: The VDR rs2228570 variant may increase susceptibility to dyslipidemia in the Chinese Han population.

Keywords: Vitamin D receptor, Polymorphism, Dyslipidemia, Chinese
Background
The prevalence of lipid abnormalities is increasing in the Chinese Han population, and is considered as a major risk factor for cardiovascular disease. Low vitamin D status has been reported to be correlated with an increased risk of hyperlipidemia [1, 2]. Furthermore, a correlation was found between vitamin D deficiency and increased serum total cholesterol (TC) and low-density lipoprotein-cholesterol (LDL-C) levels, and decreased serum high-density lipoprotein-C (HDL-C) levels [3–5]. Nonetheless, the mechanism of the lipid-lowering effect of vitamin D remains unknown. Vitamin D likely increases serum calcium by enhancing intestinal calcium absorption. Subsequently, this elevated calcium level reduces serum triglycerides by suppression of hepatic triglyceride formation and secretion [6]. The second possible mechanism may be through the inhibitory effect of vitamin D on serum PTH concentrations. The lowered level of PTH may reduce serum triglycerides by increasing their peripheral uptake. Another potential mechanism to explain the association between 25(OH) D and triglycerides would be through insulin resistance. In cases of vitamin D deficiency, the risk of insulin resistance increases, which is associated with elevated levels of very-low-density lipoprotein (VLDL) cholesterol and triglycerides [7].

Dyslipidemia is a multifactorial disease, which can be influenced by both genetic and environmental factors. Vitamin D is involved in lipid metabolism. The actions of vitamin D are mediated by the Vitamin D Receptor (VDR), a nuclear receptor [8]. The VDR gene, which codes for VDR, is located at q11–q13 on chromosome 12. A recent study showed that the CC genotype of VDR, single nucleotide polymorphism (SNP) rs2228570, is a risk factor for diabetic dyslipidemia in elderly males in North China [9]. Another investigation, conducted in Lebanon, suggested that VDR polymorphisms were associated with triglyceride and HDL levels in a healthy young population [10]. VDR FokI polymorphisms appear to be associated with coronary heart disease in Han Chinese. Furthermore, the GG genotype predicts higher HDL-cholesterol levels in adults with coronary heart disease [11]. Considering the lack of studies evaluating the link between VDR and dyslipidemia, we investigated and determined the association between VDR variants and dyslipidemia in a large Chinese Han sample.

Methods
Ethics Statement
The study was approved by the Ethics Committees of the First Affiliated Hospital with Nanjing Medical University, Nanjing, Jiangsu, China (NO2015-SR-032). All participants signed written informed consent.

Subjects
A two-stage designed study was used to evaluate whether genetic variations in VDR are associated with dyslipidemia in the Chinese Han population. In stage I, we genotyped three SNPs (rs11574129, rs2228570, and rs739837) in 877 cases with dyslipidemia from normotensive and non-diabetes mellitus population and 1822 non-dyslipidemia subjects from a community-based epidemiological survey in Jiangsu Province, China. We followed-up the stage I study to confirm the association between rs2228750 and dyslipidemia using a larger sample, including 1679 cases with dyslipidemia and 3124 age-, sex-, and ethnic origin-matched controls, in a stage II study.

Dyslipidemia was identified according to the Guidelines on Prevention and Treatment of Dyslipidemia in Chinese Adults (2007) [12]: triglyceride (TG) ≥ 2.26 mmol/L as high, total cholesterol (TC) ≥ 6.22 mmol/L as high, low-density lipoprotein cholesterol (LDL-C) ≥ 4.14 mmol/L as high, and high-density lipoprotein cholesterol (HDL-C) < 1.04 mmol/L as low. In our study, high TG, high TC, high LDL-C, or low HDL-C was regarded as dyslipidemia. The exclusion criteria included a history of cancer, stroke, thyroid disorder, kidney, or liver disease. Individuals who had family history of obesity, dyslipidemia, and other metabolic diseases were also excluded. All individuals answered standardized questionnaires inquiring about age, sex, ethnicity, history of smoking, and family medical history. Body-mass index (BMI) was calculated as weight divided by height squared in kg/m^2. Blood pressure was measured using a mercury sphygmomanometer. LDL-C, HDL-C, TC, TG, and fasting blood glucose (GLU) serum levels were measured enzymatically using an automatic clinical analyzer (Hitachi Inc., Tokyo, Japan). Serum 25(OH) D concentration was determined using an enzyme-linked immunosorbent assay (ELISA) according to the product specifications (Eagle Biosciences Inc., Nashua, New Hampshire, USA).

Genomic DNA extraction and genotyping
Genomic deoxyribonucleic acid (DNA) was extracted from whole-blood samples using the TIANGEN RelaxGene Blood DNA System (TianGen Biotech Co, Ltd, Beijing, China; NO. DP319). Three tagSNPs (rs11574129, rs2228570, and rs739837) were selected using the linkage disequilibrium (LD) method and SNP function prediction software (https://manticore.nih.gov/snpfunc/snpfunc.html). The rs11574129 and rs739837 variants regulating the microRNA binding were positioned in the 3’untranslated region. The rs2228570 was predicted as a splicing enhancer in the exon-coding region. Genotyping was performed using the TaqMan SNP Genotyping Assay (Applied Biosystems, Foster City, CA, USA). For quality control, 5% of samples were re-genotyped in a blinded fashion.
Molecular Modeling Analysis
The VDR molecular modeling studies using Discovery Studio 3.0 (DS 3.0, Accelrys Inc., Boston, MA, USA) provided insight into two VDR isoforms (wild type and RS2228570 variant). These VDR isoforms were produced by I-TASSER (http://zhanglab.ccmb.med.umich.edu/I-TASSER/). We selected the most appropriate structure from five models to model VDR by FG-MD (http://zhanglab.ccmb.med.umich.edu /FG-MD/). Then, we used DS 3.0 to set minimization (steepest descent, conjugate gradient) and equilibration steps, followed by CHARMM to understand the molecular dynamics.

Statistical analysis
Data analyses were performed using SPSS 19.0 (SPSS, Inc., Chicago, IL, USA). Quantitative variables were expressed as mean ± standard deviation (SD). Comparisons of the means between the two groups were determined by t-test, and the multiple-group means were compared using one-way ANOVA, with adjustments for age and sex. The Chi-square ($\chi^2$) test or Fisher’s exact test was employed for categorical variables. Multiple logistic regression analysis was applied to adjust covariates including age and sex. In addition, multivariable linear regression was utilized to evaluate the association of genotypes with lipid levels. Hardy–Weinberg equilibrium (HWE) was assessed by Fisher’s exact $\chi^2$ test in the control. Statistical significance was assumed at $P < 0.05$.

Results
Study subject characteristics
A total of 436 males and 441 females were included in the case group and 926 males and 956 females were included in the control group in the stage I study. The clinical characteristics of the cases in stage II study are presented in Additional file 1: Table S1. We found that the SNP rs2228570 statistically correlated with an increased risk of dyslipidemia; after adjusting for confounding factors, the odds ratios (ORs) with 95% confidence interval (CI) of the additive, dominant, and recessive models were 1.255 (1.118–1.409) ($P < 0.001$), 1.384 (1.136–1.6) ($P = 0.001$) and 1.356 (1.1–1.671) ($P = 0.004$) (Table 1).

| SNP       | Group | Genotype OR (95% CI)* | Allele     | OR (95% CI) | P-value |
|-----------|-------|-----------------------|------------|-------------|---------|
| rs11574129|       |                       |            |             |         |
|           |       | WT Ht+ MT Additive Dominant Recessive | Major/Minor | C/T         |         |
| Control   |       | 1265 560 + 55 1.069 (0.923–1.238) 1.055 (0.89–1.25) 1.26 (0.809–1.964) | 3090/670 | 1.07 (0.92–1.24) | 0.46c |
| case      |       | 578 265 + 32 P = 0.376 P = 0.536 P = 0.306 | 1421/329 | 0.38b        |         |
| rs2228570 |       |                       |            |             |         |
|           |       | CC TC + TT            |            |             |         |
| Control   |       | 539 939 + 401 1.255 (1.118–1.409) 1.384 (1.136–1.6) 1.356 (1.1–1.671) | 2017/1741 | 1.25 (1.12-1.41) | 0.832c |
| case      |       | 308 422 + 146 P < 0.001 P = 0.001 P = 0.004 | 1038/714 | 0.001b       |         |
| rs739837  |       |                       |            |             |         |
|           |       | CC TC + TT            |            |             |         |
| Control   |       | 991 749 + 139 1.094 (0.965–1.241) 1.108 (0.944–1.301) 1.157 (0.862–1.553) | 2731/1027 | 1.09 (0.96–1.24) | 0.877c |
| case      |       | 439 362 + 74 P =0.161 P = 0.209 P = 0.33 | 1240/510 | 0.17b        |         |

* Adjusted for sex and age
b P-value of Fisher’s exact $\chi^2$ test for HWE
C Odds ratio
c CI Confidence interval
that SNP rs2228570 significantly correlated with an increased risk of dyslipidemia; after adjusting for confounding factors, the ORs with 95% CI of the additive, dominant and recessive models were 1.146 (1.053–1.247) \( (P = 0.002) \), 1.184 (1.018–1.376) \( (P = 0.028) \) and 1.209 (1.064–1.374) \( (P = 0.004) \) (Table 2). To reduce potential bias in the stage II study, we adjusted for confounding factors, such as blood pressure, smoking, BMI, and GLU. In the GLU-adjusted model, we found a significant association between the rs2228570 variant and increased risk of dyslipidemia. After adjusting for confounding factors, the ORs with 95% CI of the additive, dominant, and recessive models were 1.145 (1.052–1.246) \( (P = 0.002) \), 1.177 (1.013–1.369) \( (P = 0.034) \), and 1.211 (1.065–1.377) \( (P = 0.003) \). A similar trend was observed in the blood pressure-, smoking-, and BMI-adjusted models (Table 3).

Serum lipid levels in the cases with different genotypes are shown in Fig. 1, in which it can be seen that the rs2228750 genotype was significantly associated with serum LDL levels in the dyslipidemia cases. The cases with the TT genotype had significantly higher LDL levels (4.93 ± 0.75 mmol/L) than those with the TC (4.67 ± 0.47 mmol/L) or CC (4.66 ± 0.44 mmol/L) genotype \( (P = 0.01) \). There were no significant differences among the three genotypes in the levels of serum TC, TG, or HDL \( (P > 0.05) \) (Additional file 1: Table S3). The levels of serum 25(OH)D among the different genotype cases with high LDL levels \( (\geq 4.41 \text{ mmol/L}) \) is illustrated in Fig. 2. In the cases with high LDL, the cases with the TT genotype had significantly lower serum 25(OH)D levels (18.43 ± 5.04 ng/mL) than those with the TC (26.24 ± 4.16 ng/mL) or CC (36.76 ± 8.10 ng/mL) genotype \( (P < 0.001) \) (Additional file 1: Table S4).

Multiple linear regression analysis revealed that the rs2228570 variant \( (P = 0.041) \), age \( (P = 0.01) \), and sex \( (P = 0.021) \) were independently related to the level of serum LDL (Table 4).

Molecular modeling study
The molecular modeling analysis showed that VDR protein stability was reduced in the rs2228570 variant. The binding energy of VDR to its ligand was also decreased in the rs2228570 variant (Additional file 1: Tables S5, S6). As can be observed in Fig. 3, the conformation and interaction changes are in the polymorphic region (rs2228570), where the replacement of threonine with methionine at amino acid position 1 changed the structure of VDR. This modification resulted in a shift of the mode in which the amino acid residue Tyr236 in the protein VDR was bound to hydrogen in its ligand. The change also provided interactions with neighboring residues.

### Table 2

| SNP | Group | Genotype (95% CI) \( ^{a} \) | Additive | Dominant | Recessive | Allele |
|-----|-------|-----------------------------|---------|----------|----------|-------|
| rs2228570 | WT | 1.146 (1.053–1.247) | 1.194 (1.018–1.376) | 1.209 (1.064–1.374) | 1.146 (1.053–1.247) | 1.146 (1.053–1.247) |
|       | Ht + MT | 1.146 (1.053–1.247) | 1.184 (1.018–1.376) | 1.209 (1.064–1.374) | 1.184 (1.018–1.376) | 1.184 (1.018–1.376) |
|       | Additive | OR (95% CI) \( ^{b} \) | 1.184 (1.018–1.376) | 1.209 (1.064–1.374) | 1.209 (1.064–1.374) | 1.209 (1.064–1.374) |
|       | Dominant | OR (95% CI) \( ^{c} \) | 1.184 (1.018–1.376) | 1.209 (1.064–1.374) | 1.209 (1.064–1.374) | 1.209 (1.064–1.374) |
|       | Recessive | OR (95% CI) \( ^{d} \) | 1.184 (1.018–1.376) | 1.209 (1.064–1.374) | 1.209 (1.064–1.374) | 1.209 (1.064–1.374) |
|       | Major/Minor | OR (95% CI) \( ^{e} \) | 3.362 (2.880) | 1.15 (1.05–1.25) | 0.806 \( ^{f} \) | 0.002 \( ^{g} \) |

\( ^{a} \) Adjusted for sex and age
\( ^{b} \) P-value of \( \chi^{2} \) test for comparison of allele between case and control group
\( ^{c} \) P-value of Fisher’s exact \( \chi^{2} \) test for HWE
\( ^{d} \) WT wild type, Ht heterozygote, MT mutant type
\( ^{e} \) OR Odds ratio
CI Confidence interval

### Table 3

| Genotype | Genotype OR (95% CI) \( ^{a} \) | OR (95% CI) \( ^{b} \) | OR (95% CI) \( ^{c} \) | OR (95% CI) \( ^{d} \) |
|----------|-------------------------------|-------------------|-------------------|-------------------|
| Additive | 1.145 (1.052–1.246) | 1.145 (1.052–1.247) | 1.144 (1.051–1.246) | 1.142 (1.047–1.244) |
|         | \( P = 0.002 \) | \( P = 0.002 \) | \( P = 0.002 \) | \( P = 0.003 \) |
| Dominant | 1.177 (1.013–1.369) | 1.188 (1.021–1.381) | 1.181 (1.016–1.373) | 1.18 (1.013–1.375) |
|         | \( P = 0.034 \) | \( P = 0.026 \) | \( P = 0.03 \) | \( P = 0.034 \) |
| Recessive | 1.211 (1.065–1.377) | 1.204 (1.059–1.369) | 1.207 (1.062–1.371) | 1.201 (1.054–1.368) |
|         | \( P = 0.003 \) | \( P = 0.005 \) | \( P = 0.004 \) | \( P = 0.006 \) |

\( ^{a} \) GLU adjusted model
\( ^{b} \) SBP and DBP adjusted model
\( ^{c} \) Smoking adjusted model
\( ^{d} \) BMI adjusted model
OR Odds ratio
CI Confidence Interval
Discussion

The active form of vitamin D is 1, 25-dihydroxyvitamin D. VDR tightly regulates vitamin D levels in cells and in the circulation [13]. A previous study [14] revealed that VDR activation repressed the increased levels of mouse and human CYP7A1 and reduced the cholesterol concentrations exerted by hepatic small heterodimer partner (SHP). Recent research found that the deletion of macrophage VDR promoted insulin resistance and monocyte cholesterol transport, which accelerated atherosclerosis in mice [15]. Based on the findings of the abovementioned studies, we hypothesized that VDR expression is associated with serum lipid levels. Hence, further studies on VDR polymorphisms would improve the understanding of the association of VDR with dyslipidemia.

In our study, we found a significant association of rs2228570 polymorphism with dyslipidemia and serum LDL levels. Moreover, cases with the TT genotype had significantly lower serum 25(OH)D levels compared to cases with other genotypes with high LDL.

As is well known, vitamin D regulates lipid metabolism; it inhibits lipid synthesis and promotes adipopexis.

![Fig. 1](image1) Serum lipid levels in the different genotypes cases. a Levels of HDL distribution in the three genotype cases; b Levels of LDL distribution in the three genotypes cases; c Levels of TG distribution in the three genotypes cases; d Levels of TC distribution in the three genotypes cases. The values are means ± SD

![Fig. 2](image2) Serum 25(OH)D levels in the different genotypes in the cases with high LDL levels. The three bars indicate serum 25(OH)D distribution in the three genotype cases. The values represent means ± SD

| Variable   | Beta  | B (95% CI)       | P   |
|------------|-------|------------------|-----|
| rs2228570  | -0.117| -0.089 (0.173–0.004) | 0.041 |
| Age        | 0.152 | 0.007 (0.002–0.012)  | 0.01 |
| Sex        | 0.141 | 0.146 (0.022–0.27)   | 0.021 |
| GLU        | 0.09  | 0.065 (-0.019–0.149) | 0.131 |
| BMI        | 0.096 | 0.017 (-0.003–0.038) | 0.365 |
| Smoking    | 0.129 | 0.169 (0.014–0.325)  | 0.053 |

GLU, fasting blood glucose; BMI, body mass index
in adipocytes [16]. However, the underlying mechanisms remain largely unknown. It is possible that the potential underlying mechanism operates through facilitating calcium homeostasis via vitamin D. It is commonly known that vitamin D and parathyroid hormone (PTH) are the two most important hormones that maintain calcium levels [17, 18]. Therefore, the reduction in lipid concentrations observed in this study may have been caused by increased calcium levels due to a decrease in fatty acid absorption and an increase in fecal fatty acid content, probably arising from the formation of insoluble calcium-fatty soaps in the gut [19, 20]. Such reduced absorption of fat diminishes serum total and LDL cholesterol levels [21, 22]. Calcium can also bind bile acid [23, 24] and increase the conversion of cholesterol to bile acids, thus causing increased cholesterol excretion [25]. Furthermore, microsomal triglyceride transfer protein (MTP) is involved in the secretion of triglyceride-rich VLDL, which leads to hypertriglyceridemia and atherosclerosis [26]. In addition, hepatocellular calcium activates MTP. The increase in hepatocellular calcium could be restrained by increasing serum concentrations [23]. The inhibition of hypertriglyceridemia may be caused by the inhibition of MTP activity via the suppression of hepatocellular calcium. Another potential mechanism is likely to be associated with a decrease in cholesterol absorption [27]. Vitamin D increases intestinal calcium absorption by affecting the rate of fat absorption, which results from the ability of calcium to bind to fatty and bile acids, forming calcium-fatty acid soaps [27], which are excreted in the

Fig. 3 2D diagram of the changes in bound ligand conformations of the wild-type and mutant VDR. The superposition of 1,25(OH)2D3 molecules is shown in gray. Circles represent the amino acids of the receptor molecule. Blue halos around the circles denote the solvent-accessible surface of the interacting residue. The dashed lines indicate the hydrogen bond interactions between the ligands and the receptor molecule. The green dashed lines represent hydrogen bond interactions with amino acid main chains; blue-dashed lines show the hydrogen bond interactions with the main chains of the amino acids (the arrow head is directed towards the electron donor). The numbers (e.g., 2.2) represent the hydrogen bond interaction distances; a Conformation derived from the VDR receptor wild-type model structure; b Conformation derived from the VDR receptor mutant model structure (MET1THR)
feces. Moreover, vitamin D may play a role in insulin secretion and sensitivity, thereby indirectly influencing lipid metabolism [28].

Using our molecular model, we further established that rs2228570 changed the structure and stability of VDR and altered the binding energy of its ligand. Earlier investigations revealed that an association between low levels of vitamin D and rs2228570 located in VDR’s 5’end. rs2228570 has a T to C variation in the translation initiation codon (AGT) in exon 2 [29]. This change induces the synthesis of a small (49.5 kD) protein with increased biological activity [30]. The reason for the differences between the activities of the two proteins, such as their abilities to bind 1,25-dihydroxy vitamin D₃ or induce transcription, still remain to be determined.

Conclusions
In conclusion, our results suggest that the rs2228570 polymorphism correlates with an increased risk of higher LDL and lower serum 25(OH)D levels in Chinese patients. The variation in rs2228750 reduced the stability of VDR and changed the binding energy of its ligand, which may be the underlying molecular mechanism for variant associated dyslipidemia. Our findings provide new insights into the prevention and treatment of dyslipidemia. Nevertheless, further studies are required to verify these results in different ethnic groups. Finally, additional functional investigations of the VDR gene are needed to confirm the present findings.

Additional file

Additional file 1: Table S1. Clinical characteristics of cases and controls (stage I). Table S2. Clinical characteristics of cases and controls (stage II). Table S3. Analysis of rs2228570 association with lipid levels. Table S4. Analysis of the association of rs2228570 and serum 25(OH)D levels in cases with high LDL. Table S5. Changes in VDR stability. Table S6. Changes in binding energy of VDR. (DOC 241 kb)

Abbreviations

LDL-C: Low density lipoprotein cholesterol; SNPs: Single nucleotide polymorphisms; TC: Total cholesterol; VDR: Vitamin D receptor

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Availability of data and materials

All data generated or analysed during this study are included in this MS and its Additional file 1.

Authors' contributions

YYZ contributed to the conception of the study. CS and NZ contributed significantly to analysis and manuscript preparation; JJ and YYT performed the data analyses and wrote the manuscript; HXD helped perform the analysis with constructive discussions. All authors approved the final version for publication.

Ethics approval and consent to participate

The experimental protocol was approved by the Institutional Ethics Board of the First Affiliated Hospital with Nanjing Medical University (No2015-SR-032). All patients signed written informed consent.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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