Article

Glycosyl Formates: Glycosylations with Neighboring-Group Participation

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Abstract: Protected 2-O-benzylated glycosyl formates were synthesized in one-step from the corresponding orthoester using formic acid as the sole reagent. Glucopyranosyl, mannopyranosyl and galactopyranosyl donors were synthesized and their glycosylation properties studied using model glycosyl acceptors of varied steric bulk and reactivity. Bismuth triflate was the preferred catalyst and KPF6 was used as an additive. The 1,2-trans-selectivities resulting from neighboring-group participation were excellent and the glycosylations were generally high-yielding.

Keywords: glycosylation; catalysis; selectivity; formates; neighboring-group participation; atom economy

1. Introduction

Atom-economic glycosylation methods are receiving increasing interest as a consequence of the focus on green and sustainable chemistry. Early examples of simple catalytic glycosylations [1] in oligosaccharide synthesis came from the Kochetkov group, who introduced glycosylations using orthoesters [2]. Glycosyl fluorides [3] and trichloroacetimidates (TCAs) [4] appeared as glycosylation reagents in the early nineteen eighties and have since then been among the most popular glycosyl donors for catalytic glycosylations. 1 N-Phenyl trifluoroacetimidates have also received increasing attention as a glycosyl donor in recent years [5]. Although the abovementioned glycosyl donors are all effectively activated by simple Brønsted or Lewis acid catalysis, there has been a recent trend towards developing increasingly complex glycosylation methods involving expensive metal catalysts, and, often, complex leaving groups. The gold-catalyzed activation of alkyne derivatives has been a “hot topic” and has been used successfully in natural product synthesis [6,7]. Many of the newly developed methods are both elegant and useful [8], but most are restricted to small-scale synthesis due to the high cost of the specialized substrates and catalysts [9]. The use of highly specialized and sensitive reagents is to be avoided and heavy-metal promoters are environmentally concerning upon upscaling. Furthermore, large leaving groups generate more waste and, hence, a better atom economy is desirable. None of the methods mentioned above fulfill the demands for a scalable, cheap and environmentally friendly glycosylation method. Glycosyl esters are an interesting group of glycosyl donors, which are commonly used as anomeric protective groups. When used as an anomeric leaving group, relative harsh conditions are needed, which limits their use in oligosaccharide synthesis [1,10]. Methods using a less trivial remote activation have been developed to increase leaving group ability, but these methods require expensive reagents, such as Tf2O, in stoichiometric amounts [11]. Recently, several groups have reported glycosylation methods using ester leaving groups, catalyzed by metal triflates, but the scope is still limited [1,12–15]. Recently, we introduced glycosyl formates as glycosyl donors [16]. In addition to their simple synthesis using formic acid, this new donor type could be catalytically activated, and we have, furthermore, demonstrated that, under glycosylation conditions, the formic acid produced could be transformed into CO2 and hydrogen gas making the glycosylation “traceless”. In
this preliminary study, it was found that Bi(OTf)$_3$ was an excellent catalyst for activating the formyl group. Bi(OTf)$_3$ has also been demonstrated to activate glycosyl halides [13,17] and the Ferrier rearrangement [18]. Working with the synthesis of glycosyl formates, it was surprising to find that this type of functional group had been only poorly investigated in glycosides. Hough and Lewis isolated 1-O-formoyl-β-D-glucopyranose tetraacetate as an undesired by-product, which represents the first description of a glycosyl formate [19]. Later, Vodcadlo and co-workers used formates as a precursor for the enol ether, which can be obtained via a Wittig-type reaction [20]. Frauenrath and coworkers prepared 1-O-vinyl glycosides from the corresponding formates using Tebbe’s reagent [21]. In this paper, our progress in using glycosyl formates as simple glycosyl donors is reported. The focus is on the synthesis of this new class of glycosyl donors and on their glycosylation properties, where neighboring-group participation is used for anomeric stereo control.

2. Results

With the goal of simplifying the glycosylation reaction and making it more atom-economic, we previously developed a new efficient synthesis of glycosyl formates, which avoids the use of reagents and catalysts. The hemiacetal of the donor was simply treated with formic acid giving the new donor and recovered starting material (Scheme 1) [16]. The simplicity of the method contrasts with the previous syntheses of glycosyl formates, where glycosyl halides [19–21] thioglycosides [21] or coupling reactions (EDC) [21] were required. All these procedures produce at least a stoichiometric amount of waste.

![Scheme 1](image1.png)

**Scheme 1.** Synthesis of glycosyl formates using HCOOH as the sole reagent and solvent.

It was observed that the glycosyl formate synthesis mainly achieved the α-product ($\alpha:\beta = 9:1$), which could suggest thermodynamic control. Additionally, full conversion of the hemiacetal was not achievable, which is consistent with the establishment of an equilibrium between the products and the substrate.

When synthesizing 2-O-benzoyl-3,4,6-tri-O-benzyl-β-D-glucopyranosyl formate D1 from the 1,2-orthoester 1, it was found that the conditions using neat formic acid gave rise to partly debenzylation and, therefore, it was decided to use diluted solutions of formic acid in CH$_2$Cl$_2$, which resulted in a very satisfying 89% isolated yield (Scheme 2) [16]. The problems with incomplete formylations appear to be related to using the hemiacetal as the substrate, which liberated equivalent amounts of water upon esterification. Obviously, the use of orthoesters as substrates is desirable as these are readily available and the formed 2-benzoyl-group would be expected to impose neighboring-group participation, and even anchimeric assistance, in the otherwise relatively slow glycosylation reaction using formates [22,23]. Perbenzylated 1,2-orthoesters with galacto- and manno- stereochemistry (Supplementary Materials: S9 and S13) were therefore synthesized using literature protocols (see Supplementary Materials for details). Treating these orthoesters with diluted formic acid resulted in a clean transformation into the corresponding glycosyl formates. Isolated yields of almost 90% with high anomeric selectivity clearly demonstrated the feasibility of this protocol (Figure 1; see Supplementary Materials for details).

![Scheme 2](image2.png)

**Scheme 2.** Synthesis of donor D1 from the orthoester.
A common side-reaction, when using ester leaving groups in glycosylations, is hemiacetal formation from the nucleophilic attack on the ester carbonyl carbon, i.e., a transesterification. This side-reaction, quantified by the “GT” parameter, has previously been shown to be dependent on additives, catalyst loading, and the nucleophile used [16]. In this investigation, our hypothesis was that having neighboring-group participation would reduce this reaction pathway via the formation of a dioxolenium ion intermediate faster than the transesterification reaction takes place.

With access to 2-O-benzoylated glycosyl formates (D1 to D3) with different stereochemistry, evaluation of their glycosylation properties could begin. The glucosyl donor D1 has previously been used [16] and, hence, served as a reference for the new donors. The previous results are included in the table for completeness.

Due to the relative long reaction times with armed glycosyl donors [24], disarmed glycosyl donors carrying more acyl groups were not investigated in depth and an initial screening confirmed the low reactivity.

Four model glycosyl acceptors, with different steric bulk and nucleophilicities, were chosen for this study (Figure 1). Cyclohexanol A1 was used as a secondary achiral alcohol with medium steric bulk. 1-Adamantanol A2 is a sterically more demanding glycosyl acceptor although still nucleophilic despite being a tertiary alcohol [25]. A 6-OH and a 4-OH glucosyl acceptor (A3 and A4) were also included as more relevant and challenging nucleophiles, with different reactivities. Glycosylation using D1 was carried out under standard conditions, i.e., THF, 25 mol%, Bi(OTf)3 and KPF6 as an additive. To our surprise, the least reactive nucleophile A4 was the best performing, giving an 81% isolated yield of the β-anomer G4. Both A2 and A3 also gave good, isolated yields around 70% of the β-anomers (G2 and G3, respectively). Glycosylations with donor A1 improved the β-selectivity and the yield remained similar to the reactions with a 2-OBn group (Table 1, entry 1–4, and previous results with the perbenzylated glucosyl formate [16]). Turning to the galactosyl donor D2, acceptors A1–A3 all gave similar yields of 70% and, exclusively, the β-galactosides G5–G7, consistent with the results from the glucosylations (Table 1, entry 5–7). However, when using the more challenging 4-OH acceptor A4, the yield was reduced and only 38% of the β-galactoside G8 could be isolated (Table 1, entry 8). Attempts
to increase this yield significantly were not successful and, hence, the current methods have
some limitations when combining the galactosyl donor with bulky, less-reactive glycosyl
acceptors. The main side-product was found to be the hemiacetal, suggesting reaction at
the formate carbonyl by the nucleophile and, hence, a more dominant transesterification
pathway. The axial 4-O-Bn in the galactosyl formate is assumed to hinder the access to
the β-side and, hence, to disfavor galactoside formation, when a bulky acceptor is used.
This pushes the reaction towards transesterification, as illustrated with the blue arrows
(Scheme 3). Mannosylations using D3 (entry 9–12) followed the trend observed with the
glucosylation and gave good isolated yields of G9–12, all with excellent 1,2-trans selectivity.
The more hindered nucleophiles again performed slightly better, as was observed with
the glucosyl donor D1. This can be explained by a slower reaction, which predominantly
passes through a dioxolenium ion intermediate, and, hence, diminishes the reaction of the
formate carbonyl.

Table 1. General procedure for the glycosylations: Glycosyl donor (1 equiv.) was dissolved in THF
(67 mM) and the glycosyl acceptor (1.15 equiv.) was added together with the catalyst (Bi(OTf)3
25 mol%) and KPF6 (1.15 equiv.) as an additive. The reaction was stirred at rt under an atmosphere
of nitrogen.

| Entry | Glycosyl donor | Glycosyl acceptor | Glycoside | Yield a |
|-------|----------------|-------------------|-----------|---------|
| 1     | D1             | A1                | G1        | 65      |
| 2     | D1             | A2                | G2        | 67      |
| 3     | D1             | A3                | G3        | 72      |
| 4     | D1             | A4                | G4        | 81      |
| 5     | D2             | A1                | G5        | 71      |
| 6     | D2             | A2                | G6        | 72      |
| 7     | D2             | A3                | G7        | 73      |
| 8     | D2             | A4                | G8        | 38      |
| 9     | D3             | A1                | G9        | 62      |
| 10    | D3             | A2                | G10       | 68      |
| 11    | D3             | A3                | G11       | 71      |
| 12    | D3             | A4                | G12       | 76      |
| 13    | D4             | A1                |           |         |

* Isolated yields.

Scheme 3. Proposed mechanism explaining the lower yield when galactosylating the hindered 4-OH
acceptor A4.

Motivated by the generally good, isolated yields and excellent stereoselectivity, the
study was expanded to include another common class of protective groups known to take
part in neighboring-group participation, i.e., carbamates. The N-Alloc-protected glucosyl donor D4 was, therefore, prepared and studied using the standard conditions. As this donor cannot be synthesized directly from an orthoester, a new approach had to be developed. As described in our recent paper, hemiacetals can be used as substrates for the introduction of the formyl group. The corresponding hemiacetal was therefore prepared, inspired by literature procedures [26,27]. Formylation using formic acid could, however, not be used on this substrate, and mixed anhydrides were, therefore, investigated. Acetic formic anhydride was found to be the best choice, giving the donor D4 in 87% (brsm) yield. In contrast to the 2-O-benzoyl-protected glycosyl donors, no glycosylation products were observed, and the main product obtained was, disappointingly, the hemiacetal. This suggests that the alloc group does not effectively participate in the reaction by neighboring-group participation and that the nucleophile, therefore, primarily reacts at the formyl group. Together with similar results using per benzoylated glycosyl formates (not shown), this suggests that, when the glycosyl donor is becoming too disarmed, i.e., less reactive, the reaction pathway leading to the hemiacetals becomes dominant. Changing the acceptor did not change this outcome of the glycosylations and, hence, this method has a limitation when it comes to the functionality on position 2.

3. Conclusions

In conclusion, we have developed a general method for synthesizing various glycosyl formates with a 2-O-benzoyl group from the corresponding orthoester. The reaction proceeds under mild conditions with formic acid as the sole reagent. The 2-O-benzoyl group installed secures neighboring-group participation and, hence, 1,2-trans-selective glycosylations. The new donors were found to be storable and easy to handle. Glycosylations were performed using Bi(OTf)3 in catalytic amounts and with KPF6 as an additive. The glycosides were generally isolated in high yields and with high 1,2-trans-selectivity.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/molecules27196244/s1: Full experimental details and NMR spectra (Citation [16,22,26–39]).

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