Research Article

Predictive and Prognostic Role of Lipocalin-2 Expression in Prostate Cancer and Its Association with Gleason Score

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Received 19 September 2020; Revised 7 January 2021; Accepted 9 January 2021; Published 20 January 2021

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Lipocalin-2 has an important role in tumor progression, invasion, and metastasis. However, its role in prostate cancer remains unclear. The objective of this study is to determine the expression level of lipocalin-2 in human prostate cancer tissues and to evaluate the relationship between its expression level and clinicopathologic parameters including response to docetaxel treatment, Gleason score, progression-free survival (PFS), and overall survival (OS). We retrospectively analyzed paraffin-embedded tissue sections from 33 metastatic castrate-resistant prostate cancer (mCRPC) patients whose clinical outcomes had been tracked after docetaxel treatment. The expression status of lipocalin-2 was defined by immunohistochemistry (IHC) using the anti-lipocalin-2 antibody. Lipocalin-2 was highly expressed in 36% of the examined specimens. There was no significant correlation between high lipocalin-2 expression and docetaxel response (p = 0.09). High lipocalin-2 expression was significantly associated with a higher Gleason score (p = 0.027). Kaplan–Meier survival analysis failed to show a significant correlation between expression levels of lipocalin-2 and both OS and PFS although patients with high lipocalin-2 levels had a numerically shorter PFS and OS time compared to patients with low levels. Consequently, it is clear that further studies are needed to evaluate the predictive and prognostic role of lipocalin-2 in prostate cancer patients.

1. Introduction

Prostate cancer is the leading reason of death from cancer in men [1]. Despite the advances in treatment, many patients develop recurrence after radical prostatectomy or definitive radiotherapy. All recurrent and metastatic patients who initially respond to antiandrogen therapy inevitably progress to the castration-resistant status. In this clinical situation, docetaxel and new hormonal treatments (abiraterone and enzalutamide) are the main treatment options [2]. However, these treatment options are not effective for all metastatic castrate-resistant prostate cancers (mCRPC). About 50–60% of these patients do not benefit from docetaxel [3]. Therefore, it is necessary to develop a marker predicting sensitivity to docetaxel in order to define the patient group who will gain benefit with docetaxel.

Lipocalin-2 (also known as oncogene 24p3, neutrophil gelatinase-associated lipocalin, and uterocalin) is a glyco-sylated protein originally purified from neutrophil granules [4, 5]. Lipocalin-2 functions as a transporter of some lipophilic molecules, such as retinoids, fatty acids, and cholesterol. Moreover, it is involved in cell differentiation, iron delivery, cell migration, and apoptosis. The best-characterized function of lipocalin-2 is to deprive bacteria of iron essential to their growth. Thus, lipocalin-2 plays an important role in the innate immune response against bacterial infection [6, 7]. Surprisingly, lipocalin-2 expression has been shown to be increased in many types of cancer in recent years, including esophageal [8, 9], pancreas [10], ovarian [11], breast [12], lung [13], and gastric [14] cancer. It increases the activity of matrix metalloproteinase 9 (MMP9),...
which breaks down the extracellular matrix and basement membranes, by preventing its autodegradation, and results in tumor progression, invasion, and metastasis [15].

Previous in vitro studies have shown that lipocalin-2 overexpression facilitates the progress of CRPC by increasing androgen receptor transcriptional activity [16]. In addition, in the xenograft prostate cancer model, lipocalin-2 overexpression has significantly promoted tumor growth. In another study, it has been reported that high lipocalin-2 expression correlated significantly with tumor differentiation (p < 0.017) and Gleason’s grade [17]. In addition, previous studies have reported that expression level of lipocalin-2 in cancer patients may be associated with survival. In our study, we retrospectively evaluated the lipocalin-2 expression immunohistochemically in sections obtained from paraffin-embedded human prostate adenocarcinoma tissue samples, investigated the role of lipocalin-2 expression in predicting treatment response to docetaxel, and evaluated whether there is a relationship between its expression levels and Gleason’s grade in the patients with metastatic prostate adenocarcinoma. Survival was also analyzed to determine the prognostic value of lipocalin-2 in these patients.

2. Methods

2.1. Patient and Tissue Samples. Cases were selected retrospectively from records of Canakkale Onsekiz Mart University Health Research and Practice Hospital and Canakkale Government Hospital between the years 2015–2019. The study has been approved by the Çanakkale Onsekiz Mart University Ethics Committee (approval number: 2019-17). Patients who were diagnosed for adenocarcinoma of prostate and administered at least three cycles of docetaxel (75 mg of docetaxel per square meter as a 1 h intravenous infusion on day 1 every 21 days together with 10 mg/day prednisone continue) for mCRPC to exclude prostate specific antigen (PSA) flare as a bias source in the selection of patients were included into the study. All patients had regular measurements of PSA (every 3 weeks during active treatment). A total of 33 patients who met eligibility criteria were stratified according to treatment responses to docetaxel into two groups as responders and nonresponders. In the responder group (patients with response, partial response, stable disease, and flare phenomena) PSA response was defined according to the prostate-specific antigen working group consensus criteria [18]:

(i) Response: PSA decline ≥ 50% from baseline.
(ii) Partial response and stable disease: PSA decline <50% but >0.
(iii) Flare phenomena: initial PSA elevation followed by a decline below the PSA baseline (including patients with PSA decline ≥ 50% from baseline).

Disease progression under docetaxel treatment (nonresponder group) was defined according to the Prostate Cancer Working Group (PCWG3 2015) [19] criteria as continuous, irreversible rise of PSA (two consecutive PSA increase of 25% and 2 µg/l above the nadir) despite therapy and/or progression of measurable disease according to the Response Evaluation Criteria in Solid Tumor (RECIST) and/or appearance of 2 or more new bone lesion. In addition, patients were stratified into two groups according to Gleason score as grade group 1–4 (Gleason score ≤ 8) and grade group 5 (Gleason score 9-10).

2.2. Immunohistochemical Evaluation. Formalin-fixed and paraffin-embedded tissue specimens of all patients were obtained from pathology department archive of Canakkale Onsekiz Mart University Health Research and Practice Hospital and Canakkale Government Hospital, and these paraffinized tissues were used for IHC staining. These specimens were cut (4 µm) and stained with hematoxylin and eosin. A representative slide of each case was stained with lipocalin-2 antibody (Sigma Aldrich clon PA348-26.3.5.1/1000 dilution) in the Leica Bond Max fully automated IHC device (Leica Biosystems). IHC staining was carried out according to the manufacturer’s instructions for both the antibody and device. Stomach tissue was used as a positive control, and a section without primary antibody was used as a negative control. Lipocalin-2 expression was generally cytoplasmic. As normal prostate epithelium showed weak staining with lipocalin-2 antibody, lipocalin-2 expression on tumor cells was evaluated according to a score corresponding to the sum of both: (a) staining intensity (0, negative (Figure 1(a)); 1, weak (Figure 1(b)); 2, intermediate (Figure 1(c)); and 3, strong (Figure 1(d)) and (b) percentage of positive cells (0, 0% positive cells; 1, <25% positive cells; 2, 26–50% positive cells; and 3, >50% positive cells), as described elsewhere [20]. The sum of a + b reached a maximum score of 6. Slides were scored in the absence of any clinical data, and a score 3 and greater than 3 was evaluated as high expression.

2.3. Statistics. Each clinicopathological variable was compared between the lipocalin-2-positive and -negative expression groups and evaluated with X² test or Fisher’s exact test. OS time was calculated using the Kaplan–Meier method as the time from the date of diagnosis to the date of death or last follow-up. Differences between low and high expression groups were compared using the log-rank test. p < 0.05 (two-tailed) was considered statistically significant. All statistical analyses were performed using SPSS, version 20.

3. Results

3.1. Lipocalin-2 Expression in Prostate Cancer and Its Relationship to the Clinical Effectiveness of Docetaxel. Twelve tumors (36%) showed positive expression for lipocalin-2. Of all patients, 60% (20/33) were responsive to docetaxel. Of patients with lipocalin-2-positive tumors, 41% (5/12) had docetaxel-responsive disease, compared with 71% (15/25) of lipocalin-2-negative tumors (p = 0.095). There was no significant association between the lipocalin-2-positive and lipocalin-2-negative groups with respect to the clinical effectiveness of docetaxel. The association between the
expressions level of lipocalin-2 and clinicopathological parameters is summarized in Table 1.

### 3.2. Association of Lipocalin-2 Expression with Gleason Score

Of lipocalin-2-positive tumors, 83% (10/12) had a Gleason score of 9 or 10 (grade group 5), compared with 42% (9/21) of lipocalin-2-negative tumors \((p = 0.027)\). There was significant association between the expression levels of lipocalin-2 protein and Gleason score (Table 1).

### 3.3. Association of Lipocalin-2 Expression with Survival

PFS was defined as the time from treatment initiation with docetaxel to disease progression or last visit. All but seven patients in the study group showed progression at the time of analysis. The longest PFS was 37 months. Median PFS was 13, 1 month in patients with lipocalin-2-negative tumors compared to 7, and 4 months for those with lipocalin-2-positive tumors \((p = 0.14)\). There was no significant difference between two groups with respect to PFS. Sixteen of 33 patients (48%) died from the date of diagnosis to the last follow-up. The longest OS was 119 months. The median survival time in patients with positive and negative lipocalin-2 expression was 40.5 and 65.7 months, respectively (log-rank test, \(p = 0.16\), Figure 2).

### 4. Discussion

Docetaxel which is a microtubule-stabilizing agent leads to defective spindle formation by increasing polymerization and inhibiting depolymerization of microtubules. Defective spindle formation activating the mitotic checkpoint results in cell cycle arrest and apoptosis [21]. Various molecules such as microtubule-associated proteins, tau, Aurora A, spindle assembly checkpoint proteins (MAD1-3, BUB1-3, and BubR1) and class III \(\beta\)-tubulin have been reported as predictive markers for microtubulising agents [22].

A previous in vitro study showed that lipocalin-2 gene was one of the highly amplified genes in paclitaxel-resistant PC-3-TxR cell obtained from parent prostate cancer cell line (PC-3) [23]. However, there is only one clinical study investigating the predictive role of lipocalin-2 in the current literature. In this study, it has been reported that lipocalin-2 was a predictive marker for pathological complete response after neoadjuvant chemotherapy [24] in the low-risk subgroup of breast cancer. In this clinicopathological study that we conducted to confirm the in vitro result and determine the prognostic role of lipocalin-2 in prostate cancer patients, statistical analysis failed to show significant association between the levels of lipocalin-2 expression and the effectiveness of docetaxel although patients with high lipocalin-2 expression were more resistant to docetaxel \((p = 0.09)\).

In some studies, contradictory results have been reported for the association between some clinicopathologic parameters such as tumor differentiation, tumor stage, and lymph node metastasis and lipocalin-2 expression. Pancreatic cancer study [10] reported that high expression of lipocalin-2 was related to negative lymph node metastasis and earlier TNM stage. Conversely, cervical [25], colorectal [26], ovarian [27], and breast cancer studies reported that the high level of lipocalin-2 expression was associated with poor differentiated tumors, higher lymph node metastasis, and...
higher tumor stage. In addition, two in vitro studies conducted in oral squamous cell carcinoma (OSCC) and colon carcinoma cell line have been shown that lipocalin-2 expression reduces metastatic potential of these cells [28, 29]. There are only two studies investigating the relationship between prostate cancer and lipocalin-2 expression in the current literature. Tung MC et al. showed that high lipocalin-2 expression was significantly correlated with tumor differentiation ($p < 0.017$) and Gleason’s grade, and knockdown of lipocalin-2 suppresses the growth and invasion of prostate cancer cells [17]. In their study, lipocalin-2 was highly expressed in 37% of tissues of prostate cancer according to normal prostate tissues. Similarly, in our study, we found that lipocalin-2 was highly expressed in 36% of patients, and high lipocalin-2 expression was significantly associated with higher Gleason score ($p = 0.027$).

Different results have also been reported for the relationship between lipocalin-2 expression and survival. OSCC [28] and pancreatic cancer [10] study reported that high lipocalin-2 expression was related to a longer survival time. Conversely, cervical, lung, colon, ovarian, and breast cancer studies reported that high lipocalin-2 expression significantly correlated with a shorter survival time [13, 25–27, 30]. There is no clinical study investigating the relationship between the lipocalin-2 expression in patients with prostate cancer and survival in the current literature, and unfortunately, we failed to show a correlation between the lipocalin-2 expression and both PFS and OS in our study. However, patients with high lipocalin-2 expression had a numerically shorter PFS and OS time compared to patients with low levels.

To the best our knowledge, this is the first study investigating the effect of lipocalin-2 expression upon survival and clinical effectiveness of docetaxel treatment. However, some limitations such as the presence of censored data, the small size of the sample, and difficulties in the interpretation of IHC evaluation can be listed among the weaknesses of the study.

### 5. Conclusion

In summary, lipocalin-2 was highly expressed in about 36% of patients with prostate adenocarcinoma. High lipocalin-2 expression was significantly associated with a higher Gleason score. Although no statistical significance was observed, patients with high lipocalin-2 expression tended to have lower effectiveness of docetaxel and shorter survival time. Despite significant correlation with Gleason score, our results did not support the predictive and prognostic role of lipocalin-2. It is clear that further studies are needed to

### Table 1: Associations between the clinicopathological variables and lipocalin-2 expression.

| Variable            | $n$ | Lipocalin-2 expression | $p$ value |
|---------------------|-----|------------------------|-----------|
|                     |     | Low  | High  |                     |
| Responders          | 20  | 15   | 5     | 0.09                 |
| Nonresponders       | 13  | 6    | 7     |                      |
| Grade group 1–4     | 14  | 12   | 2     | 0.02                 |
| Grade group 5       | 19  | 9    | 10    |                      |

![Figure 2: Progression-free survival (a) and overall survival (b) in lipocalin-2-positive and lipocalin-2-negative patients.](image-url)
evaluate the predictive and prognostic role of lipocalin-2 in prostate cancer patients.

Data Availability

The clinocopathologic data used to support the findings of this study are restricted by the Ethics Board of the Çanakkale Onsekiz Mart University. Data are available from corresponding author for researchers who meet the criteria for access to confidential data.

Ethical Approval

The study has been approved by the Çanakkale Onsekiz Mart University Ethics Committee (approval number: 2019-17).

Conflicts of Interest

The authors declare that there are no conflicts of interest.

References

[1] F. Bray, J. Ferlay, I. Soerjomataram, R. L. Siegel, L. A. Torre, and A. Jemal, "Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries," CA: A Cancer Journal for Clinicians, vol. 68, no. 6, pp. 394–424, 2018.

[2] A. Delli, F. Zagouri, D. Mitropoulos, A. Bamias, and A. G. Papatsoris, "Management of advanced prostate cancer: a systematic review of existing guidelines and recommendations," Cancer Treatment Reviews, vol. 73, pp. 54–61, 2019.

[3] I. F. Tannock, R. De Wit, W. R. Berry et al., "Docetaxel plus prednisone or mitoxantrone plus prednisone for advanced prostate cancer," New England Journal of Medicine, vol. 351, no. 15, pp. 1502–1512, 2004.

[4] L. Kjeldsen, A. H. Johnsen, H. Sengeløv, and N. Borregaard, "Isolation and primary structure of NGAL, a novel protein associated with human neutrophil gelatinase," Journal of Biological Chemistry, vol. 268, no. 14, pp. 10425–10432, 1993.

[5] S. Triebel, J. Bläser, H. Reinkes, and H. Tschesche, "A 25 kDa α2-microglobulin-related protein is a component of the 125 kDa form of human gelatinase," FEBS Letters, vol. 314, no. 3, pp. 386–388, 1992.

[6] D. H. Goetz, M. A. Holmes, N. Borregaard, M. E. Bluhm, K. N. Raymond, and R. K. Strong, "The neutrophil lipocalin NGAL is a bacteriostatic agent that interferes with siderophore-mediated iron acquisition," Molecular Cell, vol. 10, no. 5, pp. 1033–1043, 2002.

[7] U. E. Schaible and S. H. E. Kaufmann, "Iron and microbial infection," Nature Reviews Microbiology, vol. 2, no. 12, pp. 946–953, 2004.

[8] H. Zhang, L. Xu, D. Xiao et al., "Upregulation of neutrophil gelatinase-associated lipocalin in oesophageal squamous cell carcinoma: significant correlation with cell differentiation and tumour invasion," Journal of Clinical Pathology, vol. 60, no. 5, pp. 555–561, 2007.

[9] Z.-P. Du, B.-L. Wu, S.-H. Wang et al., "Shortest path analyses in the protein-protein interaction network of NGAL (neutrophil gelatinase-associated lipocalin) overexpression in esophageal squamous cell carcinoma," Asian Pacific Journal of Cancer Prevention, vol. 15, no. 16, pp. 6899–6904, 2014.

[10] B. Xu, D.-Y. Jin, W.-H. Lou, and D.-S. Wang, "Lipocalin-2 is associated with a good prognosis and reversing epithelial-to-mesenchymal transition in pancreatic cancer," World Journal of Surgery, vol. 37, no. 8, pp. 1892–1900, 2013.

[11] R. Lim, N. Ahmed, N. Borregaard et al., "Neutrophil gelatinase-associated lipocalin (NGAL) an early-screening biomarker for ovarian cancer. NGAL is associated with epidermal growth factor-induced epithelio-mesenchymal transition," International Journal of Cancer, vol. 120, no. 11, pp. 2426–2434, 2007.

[12] J. Yang, B. McNeish, C. Butterfield, and M. A. Moses, "Lipocalin 2 is a novel regulator of angiogenesis in human breast cancer," The FASEB Journal, vol. 27, no. 1, pp. 45–50, 2013.

[13] T. Shimura, A. Dagher, M. Sachdev et al., "Urinary ADAM12 and MMP-9/NGAL complex detect the presence of gastric cancer," Cancer Prevention Research, vol. 8, no. 3, pp. 240–248, 2015.

[14] J. M. Ruiz-Morales, R. Dorantes-Heredia, O. Arrieta, N. C. Chávez-Tapia, and D. Motola-Kuba, "Neutrophil gelatinase-associated lipocalin (NGAL) and matrix metalloproteinase-9 (MMP-9) prognostic value in lung adenocarcinoma," Tumor Biology, vol. 36, no. 5, pp. 3601–3610, 2015.

[15] L. Yan, N. Borregaard, L. Kjeldsen, and M. A. Moses, "The high molecular weight urinary matrix metalloproteinase (MMP) activity is a complex of gelatinase B/MMP-9 and neutrophil gelatinase-associated lipocalin (NGAL)," Journal of Biological Chemistry, vol. 276, no. 40, pp. 37258–37265, 2001.

[16] G. Ding, J. Wang, C. Feng, H. Jiang, J. Xu, and Q. Ding, "Lipocalin 2 over-expression facilitates progress of castration-resistant prostate cancer via improving androgen receptor transcriptional activity," Oncotarget, vol. 7, no. 39, pp. 64309–64317, 2016.

[17] M.-C. Tung, S.-C. Hsieh, S.-F. Yang et al., "Knockdown of lipocalin-2 suppresses the growth and invasion of prostate cancer cells," The Prostate, vol. 73, no. 12, pp. 1281–1290, 2013.

[18] G. J. Bubley, M. Carducci, W. Dahut et al., "Eligibility and response guidelines for phase II clinical trials in androgen-independent prostate cancer: recommendations from the prostate-specific antigen working group," Journal of Clinical Oncology, vol. 17, no. 11, pp. 3461–3467, 1999.

[19] H. I. Scher, M. J. Morris, W. M. Stadler et al., "Trial design and objectives for castration-resistant prostate cancer: updated recommendations from the prostate cancer clinical trials working group 3," Journal of Clinical Oncology, vol. 34, no. 12, pp. 1402–1418, 2016.

[20] T. Shibusa, N. Shijubo, and S. Abe, "Tumor angiogenesis and vascular endothelial growth factor expression in stage I lung adenocarcinoma," Clinical Cancer Research: An Official Journal of the American Association for Cancer Research, vol. 4, no. 6, pp. 1483–1487, 1998.

[21] B. T. McGrogan, B. Gilmartin, D. N. Carney, and A. McCann, "Taxanes, microtubules and chemoresistant breast cancer," Biochimica et Biophysica Acta (BBA) - Reviews on Cancer, vol. 1785, no. 2, pp. 96–132, 2008.

[22] S. Noguchi, "Predictive factors for response to docetaxel in human breast cancers," Cancer Science, vol. 97, no. 9, pp. 813–820, 2006.

[23] M. Takeda, A. Mizokami, K. Mamiya et al., "The establishment of two paclitaxel-resistant prostate cancer cell lines and the mechanisms of paclitaxel resistance with two cell lines," The Prostate, vol. 67, no. 9, pp. 955–967, 2007.
A. S. Wenners, K. Mehta, S. Loibl et al., “Neutrophil gelatinase-associated lipocalin (NGAL) predicts response to neoadjuvant chemotherapy and clinical outcome in primary human breast cancer,” *PLoS One*, vol. 7, no. 10, 2012.

I.-H. Chung, T.-I. Wu, C.-J. Liao et al., “Overexpression of lipocalin 2 in human cervical cancer enhances tumor invasion,” *Oncotarget*, vol. 7, no. 10, pp. 11113–11126, 2016.

H. T. Maier, F. Aigner, B. Trenkwalder et al., “Up-regulation of neutrophil gelatinase-associated lipocalin in colorectal cancer predicts poor patient survival,” *World Journal of Surgery*, vol. 38, no. 8, pp. 2160–2167, 2014.

H. Cho and J.-H. Kim, “Lipocalin 2 expressions correlate significantly with tumor differentiation in epithelial ovarian cancer,” *Journal of Histochemistry & Cytochemistry*, vol. 57, no. 5, pp. 513–521, 2009.

C.-W. Lin, W.-E. Yang, W.-J. Lee et al., “Lipocalin 2 prevents oral cancer metastasis through carbonic anhydrase IX inhibition and is associated with favourable prognosis,” *Carcinogenesis*, vol. 37, no. 7, pp. 712–722, 2016.

H.-J. Lee, E.-K. Lee, K.-J. Lee, S.-W. Hong, Y. Yoon, and J.-S. Kim, “Ectopic expression of neutrophil gelatinase-associated lipocalin suppresses the invasion and liver metastasis of colon cancer cells,” *International Journal of Cancer*, vol. 118, no. 10, pp. 2490–2497, 2006.

M. Bauer, J. C. Eickhoff, M. N. Gould, C. Mundhenke, N. Maass, and A. Friedl, “Neutrophil gelatinase-associated lipocalin (NGAL) is a predictor of poor prognosis in human primary breast cancer,” *Breast Cancer Research and Treatment*, vol. 108, no. 3, pp. 389–397, 2008.