Hydrothermal synthesis of hydroxyapatite nanoparticles and their protein adsorption behavior

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Abstract

Protein adsorption on hydroxyapatite particles with different morphologies was investigated. The nano-sized hydroxyapatite (nano-HAp) was prepared using calcium nitrate tetrahydrate [Ca(NO3)4·4H2O] and diammonium hydrogen phosphate [(NH4)2HPO4] by using a hydrothermal method with varying synthesis temperature and pH conditions. The adsorption properties of three phosphate groups (PO43− charged site (P-site) composed of oxygen ions belonging to the hydroxyapatite [Ca10(PO4)6(OH)2; HAp] has attracted immense interest in recent years for various applications such as orthopedic and dental materials, as a packing column material for affinity chromatography in order to separate various proteins, as well as an inorganic support for immobilized metal catalysts.1)–3) The crystalline and morphological controls of synthesized HAp crystals are gaining considerable importance for their utility as general adsorbents, affinity chromatographic solid supports, drug delivery system, and protein carriers.4)–6) The crystal system of HAp is hexagonal, and therefore its crystal has two crystal faces: the a- and c-face. Several studies examining protein adsorption on HAp have reported that different types of proteins adsorb on the different crystal faces of HAp.7)–10) The a-face has a positively charged site (C-site) composed of two screw-axis calcium (Ca2+) ions. Therefore, negatively charged proteins tend to adsorb on the a-face. In contrast, the c-face has a negatively charged site (P-site) composed of oxygen ions belonging to the three phosphate groups (PO43−). Therefore, positively charged proteins tend to adsorb on the c-face.7)

HAp can be synthesized by various methods such as a solid-state reaction,11) co-precipitation,12) the sol−gel method,13),14) and sputtering.15) In particular, hydrothermal methods using elevated temperatures and pressures in aqueous solutions enable us to synthesize defect-free HAp crystals with a certain shape and high crystallinity.16),17) Therefore, by hydrothermally synthesizing HAp particles, we are able to estimate their adsorption ability for different types of proteins with greater clarity.

Many researchers have reported the relationships between HAp properties and protein adsorption behavior. Kandori et al. have previously reported on the influence of the HAp texture on the adsorption of bovine serum albumin (BSA).19) They have also reported that hydroxyapatites modified with pyrophosphoric acid improved protein adsorption, and the effects of pyrophosphoric branches were further evaluated.20),21) Kawachi et al. and Takahashi et al. have reported that rod-shaped HAp crystals synthesized hydrothermally showed a high adsorption ability for the albumin.22),23) Fujii et al. and Dasgupta et al. have reported on the selective protein adsorption properties of zinc-containing hydroxyapatite crystals.24),25) As described above, the protein adsorption capacity was investigated using various types of HAp particles, modified by organic molecules or substituted with other types of ions. However, few studies have examined the systematic research on the adsorption behavior between the crystallinity of HAp and various types of proteins.

In the present study, we report on the synthesis of HAp nanoparticles (nano-HAp) controlling both the crystalline and morphological features by changing the pH and synthesis temperature; moreover, we discuss in detail the relationship between the HAp particle characteristics and the proteins adsorbed on the HAp.

1. Introduction

Hydroxyapatite [Ca10(PO4)6(OH)2; HAp] has attracted immense interest in recent years for various applications such as orthopedic and dental materials, as a packing column material for affinity chromatography in order to separate various proteins, as well as an inorganic support for immobilized metal catalysts.1)–3)

The crystalline and morphological controls of synthesized HAp crystals are gaining considerable importance for their utility as general adsorbents, affinity chromatographic solid supports, drug delivery system, and protein carriers.4)–6) The crystal system of HAp is hexagonal, and therefore its crystal has two crystal faces: the a- and c-face. Several studies examining protein adsorption on HAp have reported that different types of proteins adsorb on the different crystal faces of HAp.7)–10) The a-face has a positively charged site (C-site) composed of two screw-axis calcium (Ca2+) ions. Therefore, negatively charged proteins tend to adsorb on the a-face. In contrast, the c-face has a negatively charged site (P-site) composed of oxygen ions belonging to the three phosphate groups (PO43−). Therefore, positively charged proteins tend to adsorb on the c-face.7)

HAp can be synthesized by various methods such as a solid-state reaction,11) co-precipitation,12) the sol−gel method,13),14) and sputtering.15) In particular, hydrothermal methods using elevated temperatures and pressures in aqueous solutions enable us to synthesize defect-free HAp crystals with a certain shape and high crystallinity.16),17) Therefore, by hydrothermally synthesizing HAp particles, we are able to estimate their adsorption ability for different types of proteins with greater clarity.

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In the present study, we report on the synthesis of HAp nanoparticles (nano-HAp) controlling both the crystalline and morphological features by changing the pH and synthesis temperature; moreover, we discuss in detail the relationship between the HAp particle characteristics and the proteins adsorbed on the HAp.

2. Experimental procedure

2.1 Materials

All materials were of analytical grade and used as received without further purification. Calcium nitrate tetrahydrate [Ca(NO3)4·4H2O], diammonium hydrogen phosphate [(NH4)2HPO4], aqueous ammonia (NH4OH), and acetone was procured from Wako Pure Chemical Industries, Japan. BSA (isoelectric point (pI) = 4.7, molecular weight (MW) = 66 000), an acidic protein), myoglobin (MGB: pI = 7.0, MW = 18 000, a neutral protein), and lysozyme (LSZ: pI = 11.1, MW = 15 000, a basic protein) were purchased from Sigma–Aldrich (St. Louis, MO).
2.2 Preparation of nano-HAp by a hydrothermal method

Nano-HAp was prepared from a mixture of a 0.05 M solution of Ca(NO$_3$)$_2$·4H$_2$O and a 0.03 M solution of (NH$_4$)$_2$HPO$_4$ by a hydrothermal method. A 200-mL 0.03 M (NH$_4$)$_2$HPO$_4$ solution was added to 200 mL of the 0.05 M Ca(NO$_3$)$_2$·4H$_2$O solution. The mixture was subsequently stirred for 5 min at 20°C and further stirred at 25, 60, 120, or 180°C, respectively. The reaction temperature was increased at the rate of 1°C/min and the mixture was stirred at each temperature for another 3 h under hydrothermal conditions. The products were recovered by filtration, washed repeatedly with distilled water, and dried at 60°C overnight; the obtained samples were abbreviated as CP-1 (25°C), CP-2 (60°C), CP-3 (120°C), and CP-4 (180°C), respectively. Under other conditions, both the calcium and phosphate solutions were adjusted to pH 10 using aqueous ammonia before mixing, and then the procedure was the same as described above. The products of the pH-adjusted conditions were abbreviated as pHCP-1, pHCP-2, pHCP-3, and pHCP-4, respectively.

2.3 Characterization of synthesized nano-HAp

All the particles prepared were characterized by nitrogen gas adsorption/desorption isotherms at 77 K using a TriStar 3000 system (Shimadzu Co., Japan). Samples were preheated at 120°C under vacuum for 2 h to remove bound water. Specific surface areas were measured using the Brunauer–Emmett–Teller (BET) method. Powder X-ray diffraction (XRD) analysis was performed using a RINT2000/PC (Rigaku Co., Japan) model using a Cu Kα source. The 2θ scanning range was set between 3 and 60°; generator settings were 40 kV and 30 mA. Field emission-scanning electron microscopy (FE-SEM, S-3000 instrument operated at 20 kV, Hitachi Co., Japan) was used to study the morphology of the particles. Before observation, the samples were coated (sputtered) with platinum using a Hitachi E-1020 ion sputter. The molar ratio of Ca/P was measured by inductively coupled plasma-atomic emission spectrometry (IRIS Advantage, NIPPON THERMO Co., Japan). The ζ-potential of particles was measured using an ELSZ-2 zeta potential analyzer (Ohtsukadensi Co., Japan).

2.4 Protein adsorption on nano-HAp

Protein adsorption was performed using BSA as an acidic protein, MGB as a neutral protein, and LSZ as a basic protein. Five mg of nano-HAp particles was mixed in 1 mL of a 0.5 mg/mL protein solution dissolved in 11 mM of a phosphate buffer solution (pH 7.4). The mixture was stirred overnight at 4°C. After being centrifuged at 12 000 r.p.m. for 10 min, the amount of unadsorbed protein in the supernatant was determined by the Bradford protein assay using the absorption band at 595 nm. The amount of proteins adsorbed on the nano-HAp was calculated from the amount of proteins in the supernatant.

3. Results and discussion

3.1 Properties of HAp particles

Figure 1 shows the XRD patterns of the products prepared with different pH values and at various temperatures. The peaks associated with dicalcium phosphate dihydrate (DCPD) were observed in the pattern for CP-1 synthesized at 25°C (non-adjusted pH—approximately pH 6) [Fig. 1(a)]. In the products synthesized at temperatures above 60°C (CP-2, CP-3, CP-4) under non-pH-adjusted conditions, all the peaks were assigned to HAp (JCPDS, 9-432). The broad XRD peaks for CP-2 suggested that the product synthesized at 60°C was an incompletely crystallized hydroxyapatite. Increasing the temperature caused the peaks to sharpen, which indicates that the crystallinity of the products (CP-3, CP-4) improved. The XRD patterns of the all products synthesized at pH 10 (pHCP-1, pHCP-2, pHCP-3, pHCP-4) showed a good agreement with HAp [Fig. 1(b)]. The broad XRD peaks for pHCP-1 and pHCP-2 showed that the product crystallinity was low. The peaks for pHCP-3 and pHCP-4 became increasingly sharper with increasing synthesis temperature, indicating an improvement in the crystalline nature of the products.

The morphologies of the products are shown by FE-SEM in Fig. 2. The image for CP-1 shows plate-like particles of 5–20 µm width, which is a characteristic morphology of DCPD [Fig. 2(a)]. The images for CP-2, CP-3, and CP-4 revealed rod-like crystals of 50 nm width and 190, 220, and 270 nm length, respectively, indicating that the crystals grew along the c-axis with increasing temperature [Fig. 2(a)]. The image for pHCP-1 shows large agglomerates formed by the aggregation of smaller particles [Fig. 2(b)]. The images for pHCP-2, pHCP-3, and pHCP-4 show granule-like particles with aspect ratios of ca. 1 and 30, 50, and 75 nm size, respectively [Fig. 2(b)].

The results from the XRD and FE-SEM indicate that in the case of non-pH-adjusted conditions, DCPD was obtained at a synthesis temperature of 25°C and HAp nanoparticles were obtained at over 60°C. Moreover, we found that HAp nanoparticles were growing along the c-axis as the synthesis temperature increased. However, at pH 10, HAp nanoparticles were obtained at all temperatures studied; moreover, the HAp nanoparticles exhibited a similar aspect ratio (ca. 1) that was independent of the synthesis temperature. These results imply that the high pH
value induced HAp nucleation before heat treatment and prevented the anisotropic growth of HAp during heat treatment. In addition, each product synthesized (CP-2, CP-3, CP-4, pHCP-1, pHCP-2, pHCP-3 and pHCP-4) can be referred to as nano-HAp.

Some of the properties of the products evaluated by BET, ζ-potential, and Ca/P molar ratio are summarized in Table 1. The surface charge is an important property in protein adsorption since it influences the electrostatic interaction between the protein and the nano-HAp surface. The ζ-potential of all the products were found to exhibit negative charges, ranging from $-19.2$ to $-34.3$ mV in phosphate buffer solution (pH 7.4). Moreover, the ζ-potential tended to decrease as the synthesis temperature increased. The specific surface area measured by BET decreased with increasing synthesis temperature for the nano-HAps synthesized over 60°C, which is consistent with the morphological observation by FE-SEM. The Ca/P molar ratios of nano-HAp synthesized under the non-adjusted pH condition (CP-2, CP-3, CP-4) ranged from 1.50 to 1.62, indicating that CP-2, CP-3 and CP-4 were calcium-deficient HAp. Increasing the temperature caused an increase of the Ca/P molar ratio in nano-HAp. However, the Ca/P molar ratios of nano-HAps synthesized at pH 10 over 60°C (pHCP-2, pHCP-3, pHCP-4) exhibited comparable ratios to the theoretical value 1.67, indicating that these nano-HAps were approximately stoichiometric, while pHCP-1 synthesized at 25°C was a calcium-deficient HAp.

3.2 Adsorption of proteins onto nano-HAps

The results of the protein adsorption onto nano-HAp are shown in Fig. 3. The amount of protein adsorbed onto nano-HAps synthesized under the non-adjusted condition (CP-2, CP-3, and CP-4) showed that LSZ adsorption slightly increased and MGB adsorption slightly decreased with an increasing synthesis temperature. However, the amount of BSA exhibited no marked tendency for the synthesis temperatures. It has been reported that various properties of HAp, such as surface charge, Ca/P molar ratio, substitution content, or morphology, affect the amount of adsorbed proteins. Because CP-2, CP-3, and CP-4 have different crystal properties individually, the improvement in LSZ adsorption might be due to the increase in electrostatic interactions between nano-HAp and LSZ, but the influence of other factors, such as the Ca/P molar ratio or morphology, cannot be ignored.

In contrast, nano-HAps synthesized at pH 10 over 60°C (pHCP-2, pHCP-3 and pHCP-4) have approximately stoichiometric values of Ca/P (1.66–1.67), and a similar aspect ratio in their morphology (length/width is ca. 1). Therefore, the adsorption behavior of the proteins can be estimated by the crystallinity and ζ-potential of the nano-HAps. The adsorbed amount of LSZ onto pHCP-2, pHCP-3, and pHCP-4 showed, remarkably, an increase with increasing synthesis temperature. This result can be explained by the following two assumptions; (i) the basic protein, LSZ, having a positive charge in the phosphate buffer solution at pH 7.4 adsorbed by electrostatic interaction on the HAp surface that had a more negative ζ-potential; (ii) the improvement in the crystalline nature of the nano-HAps promoted the adsorption of the basic protein on the P-sites, because nano-HAps with high crystallinity would expose their clear crystal faces close to the theoretical α- and c-face, which originated from the regulated

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**Table 1.** Synthesis conditions and properties of the products prepared in this study

| Synthesis temperature (°C) | pH | Specific surface area (m²/g) | Ca/P | ζ-potential (mV) | Crystal phase |
|---------------------------|----|-----------------------------|------|-----------------|--------------|
| CP-1                      | 25 | 16.9                        | 1.11 | -19.2           | DCPD         |
| CP-2                      | 60 | 84.3                        | 1.50 | -23.5           | HAp          |
| CP-3                      | 120| 64.0                        | 1.57 | -28.0           | HAp          |
| CP-4                      | 180| 39.7                        | 1.62 | -32.2           | HAp          |
| pHCP-1                    | 25 | 106.3                       | 1.54 | -26.3           | HAp          |
| pHCP-2                    | 60 | 120.8                       | 1.67 | -25.8           | HAp          |
| pHCP-3                    | 120| 59.5                        | 1.67 | -33.4           | HAp          |
| pHCP-4                    | 180| 35.1                        | 1.66 | -34.3           | HAp          |

*a prepared under non-pH-adjusted conditions. a) Specific surface area by BET method. b) Ca/P molar ratio by ICP-AES. c) Surface potential of particles. d) Crystalline phase confirmation by XRD analysis."
atomic arrangement. It is noteworthy that the adsorbed amount of acidic protein, BSA onto pHCP-2, pHCP-3, and pHCP-4 also exhibited a tendency similar to that of LSZ, increasing with an increasing synthesis temperature. Since BSA has a negative charge in phosphate buffer solution at pH 7.4, this increase of the adsorbed BSA cannot be described by the ζ-potential of nano-HAps. Kandori et al. have reported that the C-site on HAp has a high adsorption affinity for BSA; nevertheless, the net surface charge of HAp is negative. Therefore, this increase of the adsorbed BSA can be attributed to the improvement in the crystalline nature of nano-HAps, that is, the highly regulated atomic arrangement. In contrast, the adsorbed amount of neutral protein, MGB onto pHCP-2, pHCP-3, and pHCP-4 shows the adsorption tendency opposite to that of LSZ and BSA, decreasing from 0.30 mg/m² to almost zero with an increasing synthesis temperature. This result may seem certainly logical because the two different binding sites (C- and P-site) on the HAp surface have specific adsorption abilities for acidic proteins and basic proteins, respectively, so neither of them would have little affinity for the neutral protein, MGB. On the basis of the above results, we can summarize as follows: the improvement of the crystallinity of HAp resulted in the high adsorption ability for the acidic protein BSA and the basic protein LSZ. Moreover, it was found that the nearly stoichiometric crystals of HAp had little affinity for the neutral protein, MGB, which can be explained by the specific interaction between the C- and P-sites on HAp and the acidic proteins and basic proteins, respectively and these would be dominant over other factors, such as electrostatic interactions.

4. Conclusion

The effects of HAp crystallinity on the protein adsorption were examined. HAp nanoparticles (nano-HAp) were synthesized using a hydrothermal method with varying synthesis temperatures and pH conditions. Nano-HAps obtained over 60°C and pH 10 had similar aspect ratios in their morphology (length/width ca. 1) and approximately stoichiometric Ca/P molar ratios. The nano-HAps merely altered their crystallinity and ζ-potential by varying the synthesis temperatures. The adsorption amounts of proteins onto the nano-HAps revealed that the improvement in the crystallinity of HAp elevated the adsorption ability for both the acidic protein BSA and the basic protein LSZ. In contrast, we found a decreased adsorption affinity to neutral protein, MGB. This unique adsorption behavior can be explained by the specific binding of the C- or P-site on HAp towards acidic or basic proteins, respectively. These results suggest that the adsorption behavior between various types of proteins and HAp with high crystallinity is strongly affected by HAp’s specific binding sites.

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