Mortality reduction in pediatric patients with severe fatal human adenoviral pneumonia treated with high titer neutralizing antibodies (NAbs) plasma: a retrospective cohort study

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Abstract

Background: Severe fatal human adenoviral (HAdV) pneumonia is associated with significant mortality and no effective drug is available for clinical therapy. We evaluated the association and safety of high titer neutralizing antibodies (NAbs) plasma in pediatric patients with severe fatal HAdV pneumonia.

Methods: A retrospective cohort study was performed between January 2016 to June 2021 in pediatric intensive care unit. Pediatric patients with severe fatal HAdV pneumonia were included and divided into plasma group (conventional treatment plus high titer NAbs plasma treatment) and control group (conventional treatment alone). The primary outcome was mortality in hospital. Secondary outcomes were the duration of fever after adenovirus genotype determined, duration of invasive mechanical ventilation, length of hospital stay. T-test, Mann-Whitney U-test, chi-square test, univariable and multivariable logistic regression analysis, Kaplan-Meier method and log-rank test were adopted to compare differences between two groups.

Results: A total of 59 pediatric patients with severe fatal HAdV pneumonia were enrolled. They were divided into plasma group (n = 33) and control group (n = 26). The mortality in hospital was 28.8% (17/59). Significantly fewer patients progressed to death in plasma group than control group (18.2% vs 42.3%, p = 0.042). Sequential organ failure assessment (SOFA) score, oxygen index (OI) and high titer NAbs plasma treatment were included in multivariable logistic regression analysis for mortality risk factors. Consequentially, SOFA score (Hazard Ratio [HR] 7.686, 95% Confidence Interval [CI] 1.735–34.054, p = 0.007) and without high titer NAbs plasma treatment (HR 4.298, 95%CI 1.030–17.934, p = 0.045) were significantly associated with mortality. In addition, high titer NAbs plasma treatment were associated with faster temperature recovering in survivors (p = 0.031). No serious adverse effects occurred.

Conclusions: Administration of high titer NAbs plasma were associated with a lower hazard for mortality in pediatric patients with severe fatal HAdV pneumonia. For survivors, high titer NAbs plasma treatment shorten the duration of fever.

Keywords: Human adenovirus (HAdV), Neutralizing antibodies (NAbs), Pediatric, Pneumonia, Mortality

Background

Human adenovirus (HAdV) infection is a common respiratory tract pathogen. In severe acute respiratory infection, the prevalence of HAdV has been reported to 20.1% in Northern China and 8.2% in Eastern China [1]. 88.2%
were pediatric patients in severe acute respiratory infection patients [2]. The mortality rate of immunodeficiency patients was 136 times higher than that of patients with normal immune function [3]. Adenovirus antibody levels increase with age. At present, HAdV-3 and HAdV-7 are the most important types in pediatric patients with severe respiratory diseases. In a study of adenovirus antibody levels in Chinese children found that the seropositive rates were increased against HAdV-3 (12.07, 33.96, 64.29%) and against HAdV-7 (0, 18.87, 19.05%) in different age groups (1–2, 3–5, 6–17 years, respectively) [4]. The severity of HAdV infection in respiratory tract is various from mild upper respiratory symptoms to severe fatal HAdV pneumonia. Adenovirus pneumonia was severe in 20% of patients, with a fatality rate of 5.4% [5]. Furthermore, the survivors had long term sequelae following severe HAdV pneumonia include recurrent wheeze (5.4%), bronchiolitis obliterans (15.2%) [5] and bronchiectasis (50%) [6]. However, there is no specific and effective treatment for severe fatal HAdV pneumonia.

Based on the assumption that immune plasma can establish passive immunization to control the progression of the disease before the infected patient develops a specific immunization. For almost a century, convalescent plasma and the products of convalescent blood such as specific immunoglobulin have been developed to treat virus infection disease when no specific therapy was available [7, 8]. Especially in recent years, convalescent plasma transfusion was observed in patients infected with influenza virus, middle east respiratory syndrome coronavirus (MERS-CoV), severe acute respiratory syndrome, ebola virus, and coronavirus COVID-19 [9–12]. However, these treatments are difficult to obtain or expensive to produce, and may not be able to control the progression of infection disease in emergency situations. Therefore, we suggest that high titer neutralizing antibodies (NAbs) plasma screened from healthy blood donors may be beneficial for the control of severe fatal HAdV pneumonia. To evaluate clinical curative effect and safety of high titer NAbs plasma treatment, we conducted a retrospective cohort study in our hospital for a 5-year period in pediatric patients with severe fatal HAdV pneumonia.

**Methods**

**Study design and population**

A retrospective cohort study was performed in patients with severe fatal HAdV pneumonia at Guangzhou Women and Children's Medical Center between January 2016 to June 2021. This study was approved by the ethics committee of Guangzhou Women and Children's Medical Center (Approval number: 2016102420). Signed informed consents were obtained from pediatric parents or guardians for high titer NAbs plasma treatment. Inclusion criteria were as follows: (1) age below 18 years; (2) adenoviral infection diagnosis based on polymerase chain reaction (PCR) tests by using nasopharyngeal swab, sputum or bronchoalveolar lavage. Adenovirus PCR detection kit was provided by Guangdong Hexin Health Technology Company. We carried out routine adenovirus detection for hospitalized children with severe pneumonia and monitored them by qPCR method. However, adenovirus genotype was not performed in this first test, and a relatively conservative and universal test containing 300bp target gene fragment was used. We genotyped adenovirus-positive patients by qPCR method, and differentiated them according to the different gene fragments of different adenovirus-typing. Because adenovirus genotyping technology was not mature at that time, we only tested several common adenovirus genotypes, (genotypes 3, 4, 7, 11 and 55). However, genotype 3 and 7 were mainly concentrated in our included patients, 2 patients did not belong to any of the above genotypes, so they were classified as unknown type, included in the control group and all survived. (3) pneumonia confirmed by the presence of parenchymal infiltration on chest imaging with respiratory symptoms; (4) In order to make the pediatric patients had comparability, we selected patients with critically ill adenovirus pneumonia. We defined it as severe fatal adenovirus pneumonia. Consistently with severe fatal: acute respiratory distress syndrome [13], performed invasive mechanical ventilation and Murray score [14] was greater than two. Exclusion criteria were as follows: (1) immunoglobulin or plasma allergy; (2) patients with serious underlying diseases (such as chronic lung disease, complex congenital heart disease, severe liver or kidney disease, malignant tumors, severe malnutrition, severe immunodeficiency, genetic metabolic diseases, brain dysfunction due to various causes prior to onset.).

**Clinical and laboratory data**

The information of each patient was collected from electronic medical records. Information as follows: (1) basic information: age, weight, sex, underlying comorbidity and date of admission; (2) clinical features before adenovirus genotype determined: fever days, pathogen, laboratory indicators of infection and organ function, sequential organ failure assessment (SOFA) score, oxygen index (OI). (SOFA score were grouped at the boundary of 7 based on the median SOFA score in the included patients. The median OI and quartile at the time of enrollment were 9.74(6.44–17.99), and the dividing line of mild, moderate and severe ARDS in children were 8 and 16 according to consensus recommendations.
from the Pediatric Acute Lung Injury Consensus Conference [13]. OI were grouped at the boundary of 8 based on classification limits of mild to moderate lung injury). (3) main management throughout the course of the disease: human immunoglobulin, glucocorticoid, vasoactive drugs and high titer NAb plasma; (4) primary endpoint: mortality in hospital; (5) secondary clinical outcomes: fever days after adenovirus genotype determined, duration of invasive mechanical ventilation, hospital length of stay. Due to the high mortality rate of the control group and in consideration of data integrity, we only compared secondary clinical outcomes in survivors.

Protocol of high titer plasma treatment
High titer NAb plasma were screened from fresh frozen plasma of healthy blood donors. High titer NAb plasma were classified and stored in the blood center, and all of them were managed according to blood products regulations. The proper high titer NAb plasma in this study as follows: (1) the level of neutralizing antibodies was above 1:1000 (so far, there is no standard definition of high titer NAb plasma. We retained adenovirus NAb plasma with titer > 1:1000 through Guangzhou Blood Center screening); (2) the ABO blood type was compatible with patients’ ABO blood type; (3) the NAb against HAdV type (in this study we mainly include HAdV-3 and HAdV-7) were compatible with patients’ HAdV type. High titer NAb plasma regimen were started within 24h following adenovirus genotype determined. Firstly, we confirmed whether the blood center had the proper high titer NAb plasma. Secondly, we signed the informed consent form with patient’s parents or guardians. In the study, no patient withdrew informed consent form before receiving the intervention. Finally, the volume, frequency and infusion speed of high titer NAb plasma were adjusted according to the patient’s tolerance. On the whole, the transfusion dose of high titer NAb plasma was approximately 10~20ml/kg with 2–4h under supervision of the treating physician. No high titer NAb plasma would be given if the proper high titer NAb plasma was not available. The conventional treatments include management with glucocorticoid, human immunoglobulin, other symptom control and supportive treatment [15].

Statistical analysis
Continuous variables were described as the mean and standard deviation (SD) for normally distributed variables, the median and interquartile range (IQR) for non-normally distributed variables. Categorical variables were presented as number (percentage). The independent sample T-test, Mann-Whitney U-test, chi-square test and Fisher’s exact test were adopted to compare differences between two groups. At the same time, variables with p-value were less than 0.1 in univariable analysis were selected for multivariable logistic regression analysis for mortality risk factors. Time-to-event data were analyzed by using the Kaplan-Meier method and log-rank test were used for comparing in survivors. Tests were two-tailed and p-value<0.05 were defined as statistically significant. All statistical analyses were performed with SPSS statistical software, version 26.0 (IBM, USA).

Results
Characteristics of the study population
Of the 120 pediatric patients diagnosed with adenoviral pneumonia in PICU between January 2016 to June 2021. Of these, 61 patients were excluded from the study. Consequently, 59 patients were included in the study, 33 patients got conventional plus high titer NAb plasma treatment (plasma group) and 26 patients got conventional treatment alone (control group). (Fig. 1). Baseline characteristics of these pediatric patients were illustrated in Table 1. In the study population, there were 52 (88.1%) patients with HAdV-7, 5 (8.5%) patients with HAdV-3, and 2 (3.4%) patients with unclear adenovirus genotype. The median age was 18 months (IQR, 10 to 36) and 67.8% were men. The median fever days were 10 days (IQR, 7 to 14) and 28.8% of patients were co-infection before adenovirus genotype determined. More than half of the patients had SOFA score ≥ 7 and OI ≥ 8. The final mortality in hospital was 28.8% (17 of 59 patients).

Comparison between plasma group and control group
There were no significant differences in age, gender, body weight and adenovirus genotype between plasma group and control group (p>0.05). Before adenovirus genotype was determined, there were no statistical differences in fever duration, co-infection, SOFA score, OI, blood routine, infection indicators, blood coagulation function, and other organ function indicators (include alanine transaminase, serum creatinine, creatine kinase isoenzyme) (p > 0.05). During the entire treatment, there were also no significant differences in glucocorticoid treatment, human immunoglobulin treatment (p > 0.05). There were no statistically significant differences in secondary outcomes between the two groups: fever days after adenovirus genotype determined, duration of invasive mechanical ventilation, duration of hospitalization (p > 0.05). However, plasma group had lower mortality than control group (18.2% versus 42.3%, p = 0.042). As shown in Table 1.

Comparison between survivors and non-survivors
The clinical characteristics of survivors and non-survivors were shown in Table 2.
Survivors had more patients received high titer NAb plasma treatment than non-survivors (64.3% versus 35.3%, \(p=0.042\)). The rate of SOFA score \(\geq 7\) was higher in non-survivors (42.9% versus 88.2%, \(p=0.001\)). The rate of OI \(\geq 8\) was 52.4% and 76.5% in survivors and non-survivors\(p=0.088\). There were no other significant differences in survivors and non-survivors.

**Risk factors for mortality**

In order to study risk factors for mortality in patients with severe fatal HAdV pneumonia. Variables with \(p\)-value were less than 0.1 in univariable analysis were selected for multivariable logistic regression. Consequently, SOFA score, OI and high titer NAb plasma treatment were included. When SOFA score and OI were adjusted, high titer NAb plasma treatment was still a protective factor for patients \((HR=4.298, 95\% CI 1.030–17.934, p=0.045)\). As shown in Table 3.

**Subgroup analysis in survivors**

Subgroup analysis was performed with Kaplan-Meier method and log-rank test to compare differences between plasma group and control group in survivors. High titer NAb plasma treatment was observed in faster temperature recovering to normal, but not the duration of invasive mechanical ventilation and hospitalization days. As shown in Figs. 2, 3 and 4.

**Adverse events**

Only one patient was reported sporadic red rash throughout the body during high titer NAb plasma transfusion. However, the patient recovered completely soon after intravenous promethazine and a slower rate of plasma infusion. No other adverse reactions were observed in the pediatric patients during high titer NAb plasma transfusion.

**Discussion**

In our study, the administration of high titer NAb plasma was associated with a lower hazard for mortality in pediatric patients with severe fatal HAdV pneumonia, when SOFA score and OI were adjusted. In addition, further analysis showed that the time to normal body temperature was shortened with high titer NAb plasma therapy in survivors. However, the duration of mechanical ventilation and length of hospital stay were no significant differences in survivors.

In recent years, there have been increasing reports about NAbS from convalescent patients’ plasma in the treatment of virus-associated pneumonia. A randomized and multicenter phase 2 clinical trial in the United States supported the treatment of NAbS plasma in severe influenza, the mortality of anti-influenza plasma plus standard care and standard care alone were 2% and 10%, respectively [16]. Another non-randomized prospective study also shown that convalescent plasma treatment can reduce 30-day mortality in
DESCRIPTION

Critically ill patients with COVID-19 disease [17]. The results of these observational studies supported a probable efficacy and safety of NAbs were similar to ours. Conversely, a comparative study in Guinea found the administration of NAbs from convalescent patients could not improve survival in 84 patients with Ebola virus disease, the mortality was 31% in the convalescent plasma treatment group versus 38% in the control group. However, the level of NAbs was unclear in the study, which may affect the effectiveness of treatment [18]. A recent small sample study shown that convalescent plasma therapy contributed to reduce viral load and longer survival in patients with COVID-19. However, convalescent plasma did not reduce the mortality rate in critically ill patients with end-stage COVID-19 [19].

Different criteria of inclusion and exclusion for the study population, levels of NAbs may lead to different therapeutic effects of NAbs plasma. In the current study, the total mortality of 28.8% (17 of 59 patients)

Table 1 Characteristics of the all patients, and comparison between plasma group and control group

| Variables                               | Total  | Plasma group | Control group | p value |
|-----------------------------------------|--------|--------------|---------------|---------|
| Age (months)                            | 18.00 (10.00–36.00) | 18.00 (11.00–38.50) | 16.50 (7.75–32.50) | 0.647   |
| Male (%)                                | 40 (67.8) | 23 (69.7)    | 17 (65.5)     | 0.725   |
| Weight (kg)                             | 10.00 (9.00–13.00) | 10.00 (9.75–13.00) | 10.00 (8.28–12.50) | 0.386   |
| Fever daysa                             | 10.00 (7.00–14.00) | 11.00 (8.00–14.50) | 9.50 (7.00–14.50) | 0.540   |
| Co-infection, n (%)                     | 17 (28.8) | 12 (36.4)    | 5 (19.2)      | 0.149   |

Blood routine and infection indicators

| Leucocyte, 10/L                          | 5.60 (2.70–9.70) | 5.60 (2.70–8.85) | 5.85 (2.60–11.50) | 0.867   |
| Hemoglobin, g/L                         | 92.00 ± 17.37    | 88.82 ± 14.79    | 96.04 ± 19.75    | 0.114   |
| Neutrophil, 10/L                        | 3.74 (1.48–6.85) | 3.74 (1.56–6.84) | 1.36 (1.36–8.76) | 0.963   |
| Platelets, 10/L                         | 201.00 (117.00–319.00) | 188.00 (125.00–254.50) | 252.00 (100.00–354.75) | 0.296   |
| C reactive protein, mg/L                | 36.70 (11.88–58.20) | 42.10 (12.54–70.35) | 27.31 (9.00–54.08) | 0.445   |

Organ function indicators, n (%)

| Alanine transaminase > 50 U/L           | 16 (27.1)     | 10 (30.3)     | 6 (23.1)      | 0.535   |
| Serum creatinine > 62 umol/L           | 1 (1.7)       | 1 (3.0)       | 0 (0.0)       | 1.00    |
| Creatine kinase isoenzyme > 37 U/L     | 21 (35.6)     | 10 (30.3)     | 11 (42.3)     | 0.339   |
| Prothrombin time > 15s                 | 11 (18.6)     | 5 (15.2)      | 6 (23.1)      | 0.438   |
| Activated partial prothrombin time > 45 s | 28 (47.5) | 17 (51.5)     | 11 (42.3)     | 0.482   |
| Fibrinogen < 2 g/L                     | 15 (25.4)     | 6 (18.2)      | 9 (34.6)      | 0.150   |

Adenovirus genotype, n (%)

| ADV3                                    | 5 (8.5)       | 4 (12.1)      | 1 (3.8)       | 0.156   |
| ADV7                                    | 52 (88.1)     | 29 (87.9)     | 23 (88.5)     |         |
| Unkown                                  | 2 (3.4)       | 0 (0.0)       | 2 (7.7)       |         |

Severity of Illness, n (%)

| SOFA score ≥ 7                          | 33 (55.9)     | 15 (42.4)     | 18 (69.2)     | 0.068   |
| OI ≥ 8                                  | 35 (59.3)     | 21 (63.6)     | 14 (53.8)     | 0.447   |

Treatment, n (%)

| Human immunoglobulin                    | 52 (88.1)     | 31 (93.9)     | 21 (80.8)     | 0.120   |
| Glucocorticoid                          | 35 (59.3)     | 20 (60.6)     | 15 (57.7)     | 0.821   |
| Vasoactive drug                         | 36 (61.0)     | 20 (60.6)     | 16 (61.5)     | 0.942   |

Secondary outcomes, (days)

| Fever days after adenovirus genotype determined | 3.00 (2.00–6.00) | 3.00 (2.00–5.00) | 4.50 (2.00–7.00) | 0.232   |
| Duration of invasive mechanical ventilation   | 14.00 (8.00–22.00) | 15.00 (10.00–23.00) | 11.00 (7.00–20.00) | 0.207   |
| Hospital length of stay                      | 32.00 (21.00–40.00) | 17.00 (12.50–25.00) | 15.50 (8.50–24.25) | 0.249   |

Primary outcome

| Mortality, n (%)                          | 17 (28.8)     | 6 (18.2)      | 11 (42.3)     | 0.042   |

SOFa Sequential organ failure assessment, OI Oxygen index, ADV Adenovirus

Fever daysa: Fever days before adenovirus genotype determined
was higher than 15% (11 of 75 patients) described in the other observational study [20]. It may indicate that our study population was more serious. Furthermore, the proper high titer NAbs plasma in our study met the following conditions: plasma were screened from fresh frozen plasma with the level of NAbs above 1:1000; ABO blood type were compatible with patients’ ABO blood type; NAbs against HAdV type were compatible with patients’ HAdV type. To our knowledge few investigators had treated HAdV pneumonia with high

Table 2  Characteristics between survivors and non-survivors

| Variables                  | Survivors     | Non-survivors | p value |
|----------------------------|---------------|---------------|---------|
| Age (months)               | 17.5 (7.75–30.00) | 21.00 (10.00–52.50) | 0.344   |
| Male, n (%)                | 28 (66.7)     | 12 (70.6)     | 0.770   |
| Weight (kg)                | 10.00 (9.38–12.50) | 11.00 (9.00–17.50) | 0.400   |
| Fever daysa                | 9.50 (7.00–14.25) | 11.00 (8.00–15.50) | 0.563   |
| Co-infection, n (%)        | 13 (31.0)     | 4 (23.5)      | 0.408   |

Blood routine and infection indicators

| Variable                  | Survivors     | Non-survivors | p value |
|---------------------------|---------------|---------------|---------|
| Leukocyte, 10^9/L         | 5.55 (2.70–9.23) | 6.30 (2.60–11.50) | 0.960   |
| Hemoglobin, g/L           | 92.26 ± 16.62 | 91.35 ± 19.63 | 0.857   |
| Neutrophil, 10^9/L        | 3.91 (1.56–6.78) | 3.00 (1.11–10.15) | 0.867   |
| Platelets, 10^9/L         | 195.50 (132.75–302.25) | 242.00 (67.50–356.00) | 0.947   |
| C reactive protein, mg/L  | 37.40 (11.66–73.35) | 27.52 (9.44–55.00) | 0.808   |

Organ function indicators, n (%)

| Variable                  | Survivors     | Non-survivors | p value |
|---------------------------|---------------|---------------|---------|
| Alanine transaminase>50 U/L | 9 (21.4) | 7 (41.2) | 0.122   |
| Creatinin>62 umol/L       | 0 (0.0)      | 1 (5.9)      | 0.288   |
| Creatine kinase isoenzyme >37 U/L | 13 (31.0) | 8 (47.1) | 0.242   |
| Prothrombin time>15 s     | 6 (14.3)     | 5 (29.4)     | 0.162   |
| Activated partial prothrombin time>45 s | 18 (42.9) | 10 (58.8) | 0.266   |
| Fibrinogen<2 g/L          | 9 (21.4)     | 6 (35.3)     | 0.216   |

Severity of Illness, n (%)

| Variable                  | Survivors     | Non-survivors | p value |
|---------------------------|---------------|---------------|---------|
| SOFA score ≥ 7            | 18 (42.9)    | 15 (88.2)    | 0.001   |
| OI ≥ 8                    | 22 (52.4)    | 13 (76.5)    | 0.088   |

Treatment, n (%)

| Variable                  | Survivors | Non-survivors | p value |
|---------------------------|-----------|---------------|---------|
| Human immunoglobulin      | 38 (90.5) | 14 (82.4)     | 0.320   |
| Glucocorticoid            | 27 (64.3) | 8 (47.1)      | 0.222   |
| Vasoactive drug           | 23 (54.8) | 13 (76.5)     | 0.122   |
| High titer NAbs plasma    | 27 (64.3) | 6 (35.3)      | 0.042   |

Adenovirus genotype, n (%)

| Variable                  | Survivors | Non-survivors | p value |
|---------------------------|-----------|---------------|---------|
| ADV3                      | 4 (9.5)   | 1 (5.9)       | 0.578   |
| ADV7                      | 36 (85.7) | 16 (94.1)     |         |
| Unknown                   | 2 (4.8)   | 0 (0)         |         |

Table 3  Risk factors for mortality in patients with severe fatal human adenoviral pneumonia

| Variables                  | Univariable analysis | Multivariable analysis |
|----------------------------|----------------------|------------------------|
|                            | OR 95%CI              | OR 95%CI               |
| SOFA score (<7 versus ≥7)  | 8.400 2.076–33.982   | 7.686 1.735–34.054     |
| OI (<8 versus ≥8)          | 2.955 0.827–10.561   | 3.721 0.789–17.550     |
| High titer NAbs plasma     | 3.300 1.016–10.719   | 4.298 1.030–17.934     |

SOFA Sequential organ failure assessment, OI Oxygen index, ADV Adenovirus, NAbs Neutralizing antibodies
Fever daysa: Fever days before adenovirus genotype determined
Fig. 2  Time to temperature returns to normal between plasma group and control group in survivors

Survivors

Log-rank P=0.031

Cumulative normal temperature rate, %

Days after adenovirus genotype was determined, d

Plasma group
Control group

Fig. 3  Time to withdraw ventilator between plasma group and control group in survivors

Survivors

Log-rank P=0.462

Cumulative withdraw ventilator rate, %

Days after adenovirus genotype was determined, d

Plasma group
Control group
titer NAbs plasma from healthy blood donors. In a word, our NAbs plasma had high titers and specific to HAdV type. In a retrospective cohort study found high antibody levels of convalescent plasma administration reduced risk of death compared with low antibody levels of convalescent plasma administration [21]. Another study found viral load reduced in the 2 participants who received higher levels of NAbs plasma but not in the 2 participants who received lower levels of NAbs plasma [22]. These studies showed that higher levels of NAbs plasma were beneficial to improve clinical outcomes.

Another reason for different therapeutic effect of NAbs plasma is the application time. However, the entire mechanisms and precise therapeutic components of NAbs plasma are not fully understood. The possible mechanisms of NAbs plasma in virus pneumonia, including directly neutralizing the virus and regulating immune [23]. Therefore, the transfusion of NAbs plasma may be beneficial to control the severe inflammatory response induced by virus and reduce the viral load. In addition, animal and cellular research also had shown that plasma could improve pulmonary vascular permeability, reduce leukocyte infiltration and alveolar wall thickening [24]. Typically, viremia peaks are in the first week after viral infection, and patients commonly trigger primary immune to clear viruses within 2 weeks [25]. Two weeks later, clinical deterioration is the result of inflammatory or hyperimmune attacks rather than viruses directly damage tissue. Therefore, high titer NAbs plasma should theoretically be more effective in the early course of disease. Based on these theories, convalescent plasma transfusion was failure to reduce mortality in some studies might be attributed to the late application of convalescent plasma (on median day 21.5 after onset) [26]. In contrast, one critically ill patient finally recovered after convalescent plasma infused on day 11 after onset [26]. In our study, the median time to give high titer NAbs plasma was on day 10 after onset. Therefore, we observed a reduction in mortality with high titer NAbs plasma therapy.

It is worth mentioning that high titer NAbs plasma belongs to blood products. Transfusion-related adverse events may occur in the transfusion of blood products, including fever (58.5%), anaphylaxis (1.4%), hypotension (0.9%), haemolytic reactions (5.2%), transfusion associated acute lung injury (TRALI) (0.3%), transfusion associated circulatory overload (0.6%) and so on [27]. Among them, TRALI is the leading cause of transfusion-related death. Another study had shown that the incidence of TRALI was 5.1% (114 of 2235) [28]. Patients with severe viral infection have significant acute lung injury, plasma transfusion may aggravate lung injury. In our study, most patients tolerated high titer NAbs plasma transfusion well, only 1 patient had scattered red rash throughout

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**Fig. 4** Time to hospital discharge between plasma group and control group in survivors
the body during the process of high titer NAbs plasma infusion. However, the rash quickly subsided after the treatment of promethazine and a slower rate of plasma infusion, and no other adverse reactions were observed. As everyone knows, Human leukocyte antigen antibodies (HLA-Ab) from the donor are a primary cause of TRALI [29]. Nevertheless, infectious pathogens can trigger the formation and rise of specific HLA-Ab reactivity via multiple mechanisms [30]. Recently, one study performed universal screening of HLA-Ab for all convalescent plasma. They found that 5 (7.2%) of 69 male patients were HLA-Ab positive [31]. Unlike convalescent plasma donors who have recently recovered from viral infection high titer NAbs plasma from regular blood donors may contain little HLA-Ab. As a result, high titer NAbs plasma is relatively safe, but it is still recommended to monitor and manage the possible adverse reactions according to the guidelines related to blood transfusion.

Several limitations of our study should also be recognized. First, this study was conducted in a single center. It could also indicate that the conventional treatment was more consistent. Second, because the proper high titer NAbs plasma was limited, the study was not randomized and the sample size was small. The efficacy of the study was inadequate to estimate the association between treatment and mortality with sufficient adjustment for confounding. A randomized controlled multicenter study will be required in the future. In addition, we did not dynamically monitor the viral load in patients. Viremia peaks in patients with severe fatal HAdV pneumonia may not be consistent, the dynamic monitoring of gland viral load may help to find the best time to use high titer NAbs plasma and clarify mechanism.

Conclusion
In conclusion, our study may be the first to use high titer NAbs plasma in the treatment of pediatric patients with severe fatal HAdV pneumonia. Furthermore, we found that high titer NAbs plasma therapy may be associated with reduced mortality. However, large-scale studies are required to investigate the optimal time for high titer NAbs plasma transfusion for preventing clinical deterioration and reducing mortality.

Abbreviations
NABs: Neutralizing antibodies; HAdV: Human adenoviral; SOFA: Sequential organ failure assessment; CI: Oxygen index; HR: Hazard ratio; CI: Confidence Interval; MERS-CoV: Middle east respiratory syndrome coronavirus; PICU: Pediatric intensive care unit; PCR: Polymerase chain reaction; SD: Standard deviation; IQR: Interquartile range; TRALI: Transfusion related acute lung injury; HLA-Ab: Human leukocyte antigen antibodies.

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Authors’ contributions
Conceived and designed the study: YYY and RD. Collected and analyzed data: HYP, YYY, YLZ, BLH. Wrote the paper: HYP YYY. All authors had reviewed and approved the final manuscript.

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Availability of data and materials
The datasets used and/or analyzed are not publically available in the current study. However, the datasets can be shared from the corresponding author on reasonable request.

Declarations
Ethics approval and consent to participate
This study was approved by the ethics committee of Guangzhou Women and Children’s Medical Center (Approval number: 2016102420). Signed informed consents were obtained from pediatric parents or guardians for high titer NAbs plasma treatment. All methods were performed in accordance with the provisions of the Declaration of Helsinki.

Consent for publication
Not applicable.

Competing interests
The authors declare that no competing interests exist.

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References
1. Li Y, Zhou W, Zhao Y, Wang Y, Xie Z, Lou Y, et al. Molecular typing and epidemiology profiles of human adenovirus infection among paediatric patients with severe acute respiratory infection in China. PLoS One. 2015;10(4):e123234. https://doi.org/10.1371/journal.pone.0123234.
2. Pscheidt VM, Gregianini TS, Martins LG, Veggia A. Epidemiology of human adenovirus associated with respiratory infection in southern Brazil. Rev Med Virol. 2021;31(4):e2189. https://doi.org/10.1002/rmv.2189.
3. Spaeder MC. Severe adenoviral respiratory infection in children. Intensive Care Med. 2013;39(6):1157–8. https://doi.org/10.1007/s00134-013-2893-0.
4. Tian X, Fan Y, Wang C, Liu Z, Liu W, Xu Y, et al. Seroprevalence of neutralizing antibodies against six human adenovirus types indicates the low level of herd immunity in young children from Guangzhou. China Virol Sin. 2021;36(3):373–81. https://doi.org/10.1007/s12250-020-00307-1.
5. Li L, Woo YY, de Bruyne JA, Nathan AM, Kee SY, Chan YF, et al. Epidemiology, clinical presentation and respiratory sequelae of adenovirus pneumonia in children in Kuala Lumpur, Malaysia. PLoS One. 2018;13(10):e205795. https://doi.org/10.1371/journal.pone.0205795.
6. Alharbi S, Van Caeseele P, Consunji-Araneta R, et al. Epidemiology of severe pediatric adenovirus lower respiratory tract infections in Manitoba, Canada, 1991–2005. BMC Infect Dis. 2012;12:53. https://doi.org/10.1186/1471-2334-12-53.
7. Duan K, Liu B, Li G, Zhang H, Yu T, Qu J, et al. Effectiveness of convalescent plasma therapy in severe COVID-19 patients. Proc Natl Acad Sci U S A. 2020;117(17):9490–6. https://doi.org/10.1073/pnas.2004116117.
8. Davey RJ, Fernandez-Cruz E, Markowitz N, Pett S, Babiker AG, Wentworth D, et al. Anti-influenza hyperimmune intravenous immunoglobulin for...
adults with influenza a or b infection (flu-mig): a double-blind, randomised, placebo-controlled trial. lancet respir med. 2019;7(11):951–63. https://doi.org/10.1016/s2213-2600(19)30253-x.

arabi ym, hajeeh ah, luke t, raviprakash k, balikhy h, johni s, et al. feasibility of using convalescent plasma immunotherapy for mers-cov infection, saudi arabia. emerg infect dis. 2016;22(9):1554–61. https://doi.org/10.3201/eid2209.151164.

ye h, chiuieh ts, suu lk, lin jc, chan pk, peng my, et al. experience of using convalescent plasma for severe acute respiratory syndrome among healthcare workers in a taiwan hospital. j antimicrob chemother. 2005;56(s5):919–22. https://doi.org/10.1093/jac/dki346.

mair-jenkins j, saavedra-campos m, baillie jk, cleary p, khaw fm, lim ws, et al. the effectiveness of convalescent plasma and hyperimmune immunglobulin for the treatment of severe acute respiratory infections of virology: a systematic review and exploratory meta-analysis. j infect dis. 2015;211(1):80–90. https://doi.org/10.1093/infdis/juy396.

shenoy ag, hettinger az, fernandez sj, blumenthal j, baev z. early mortality benefit with covid-19 convalescent plasma: a matched control study. br j hematol. 2021;192(4):706–13. https://doi.org/10.1111/bjh.17272.

pediatric acute respiratory distress syndrome. consensus recommendations from the pediatric acute lung injury consensus conference. pediatr crit care med. 2015;16(5):428–39. https://doi.org/10.1097/pcc.0000000000000350.

murray jf, matthey ma, luce jm, flick mr. an expanded definition of the adult respiratory distress syndrome. am rev respir dis. 1988;138(3):720–3. https://doi.org/10.1164/ajrccm/138.3.720.

jiang zt, shen kl, shen y. zhufutang practical pediatrics (8th edition) [m]. beijing: people’s medical publishing house; 2014. p. 1260–2.

beigel jh, tebas p, elle-tuennec mc, bajwa e, bell te, cairns cb, et al. immune plasma for the treatment of severe influenza: an open-label, multicentre, phase 2 randomised study. lancet respir med. 2017;5(6):500–11. https://doi.org/10.1016/s2213-2600(17)30174-1.

alsharidah s, ayed m, ameen rm, alhuraish f, Rouheldeen na, Alshammari fr, et al. covid-19 convalescent plasma treatment of moderate and severe cases of sars-cov-2 infection: a multicenter interventional study. int j infect dis. 2021;103:439–46. https://doi.org/10.1016/j.ijid.2020.11.198.

van grensen j, edwards t, de lamballerie x, semple mg, gallian p, baize s, et al. evaluation of convalescent plasma for ebola virus disease in guinea. n engl j med. 2016;374(1):33–42. https://doi.org/10.1056/nejmoa1511812.

zeng ql, yu zj, gou jj, li gm, ma sh, zhang gf, et al. effect of convalescent plasma therapy on vital shedding and survival in patients with coronavirus disease 2019. j infect dis. 2020;222(1):38–43. https://doi.org/10.1093/infdis/jiaa228.

huang h, chen y, ma ly, et al. analysis of the clinical features and the risk factors of severe adenovirus pneumonia in children. zhonghua er ke za zhi. 2021;59(1):14–9. https://doi.org/10.3760/cma.j.cn112140-20200704-00667.

joyner mj, carter re, senefeld jw, klassen sa, millis jr, johnson pw, et al. convalescent plasma antibody levels and the risk of death from covid-19. n engl j med. 2021;384(11):1015–27. https://doi.org/10.1056/nejmoa2031893.

brown jf, dye jm, tozay s, jeh-mulbah gw, wohl da, fischer wn, et al. anti-ebola virus antibody levels in convalescent plasma and viral load after plasma infusion in patients with ebola virus disease. j infect dis. 2018;218(4):555–62. https://doi.org/10.1093/infdis/jiy199.

acosta-ampudia y, monsalve dm, rojas m, rodriguez y, gallo je, salazar-unibe jc, et al. covid-19 convalescent plasma composition and immunological effects in severe patients. j autoimmun. 2021;118:102598. https://doi.org/10.1016/j.jauto.2021.102598.

potter dr, baimukanova g, keating sm, deng x, chu ja, gibb sl, et al. fresh frozen plasma and spray-dried plasma mitigate pulmonary vascular permeability and inflammation in hemorrhagic shock. j trauma acute care surg. 2015;78(suppl 1):57–17. https://doi.org/10.1097/taa.0000000000000630.

murchu eq, byrne p, walsh ka, et al. immune response following infection with sars-cov-2 and other coronaviruses: a rapid review. rev med virol. 2021;31(2):e2162. https://doi.org/10.1002/rmv.2162.

cheng y, wong r, soo yo, wong ws, lee ck, ng mh, et al. use of convalescent plasma therapy in sars patients in hong kong. eur j clin microbiol infect dis. 2005;24(1):44–6. https://doi.org/10.1007/s10096-004-1271-9.

mafrakuraeva n, khoza s, mvere da, et al. incidence and pattern of 12 years of reported transfusion adverse events in zimbabwe: a retrospective analysis. blood transfus. 2014;12(3):362–7. https://doi.org/10.2450/2014.0156-13.

vlaar ap, binnekade jm, prins d, van steen d, Hofstra JJ, schultz mj, et al. risk factors and outcome of transfusion-related acute lung injury in the critically ill: a nested case-control study. crit care med. 2010;38(3):771–8. https://doi.org/10.1093/ccrcc/388.

fabron aj, lopes lb, bordin jo. transfusion-related acute lung injury. j bras pneumol. 2007;33(2):206–12. https://doi.org/10.1590/s1806-371207000000016.

d’orsogna l, van den heuvel h, van kooten c, heidt s, claus f. infectious pathogens may trigger specific allo-hla reactivity via multiple mechanisms. immunogenetics. 2017;69(9–10):631–41. https://doi.org/10.1007/s00251-017-0989-3.

juskevitch je, stubbs jr, Gandhi MJ. Elevated rate of HLA antibodies in male COVID-19 convalescent plasma donors: a risk factor for transfusion-related acute lung injury. Mayo Clin Proc. 2021;96(2):500–2. https://doi.org/10.1016/j.mayocp.2020.11.007.

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