Identification of hippocampal cortical microinfarcts on postmortem 3-T magnetic resonance imaging

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Received: 20 February 2021 / Accepted: 12 April 2021 / Published online: 28 April 2021
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Abstract
Cortical microinfarcts (CMI) are increasingly recognized in the neurological community as a biomarker related to cognitive impairment and dementia. If their radiological depiction has been largely described in experimental settings using ultra-high-field magnetic resonance imaging (MRI), less is known about their visibility on routinely used 3-T MRI. In this radiologic-pathologic correlation study, using 3-T post-mortem MRI, we searched for hippocampal CMI, in a double-blinded fashion, and found that only 4/36, or 11%, were clearly demonstrated on both radiological and histopathological exams.

Keywords Cerebral microinfarcts · Hippocampus · Dementia · 3-T post-mortem MRI · Histopathology

Background
Cortical microinfarcts (CMI) correspond to small ischemic events of 0.05–4 mm in size, mostly measuring ≤ 1 mm [1]. They are associated with ß-amyloid deposits (57.1% of cases in one study [2]) and, depending on the affected brain region, atherosclerotic microangiopathy [2]. They are increasingly recognized in the neurological community as a radiological biomarker of vascular cognitive impairment [3, 4]. One meta-analysis of 32 neuropathological studies [5] found CMI in 62% of patients with vascular dementia (VaD), in 43% of patients with Alzheimer’s disease (AD), and in 33% of patients with mixed AD and VaD, compared to 24% of non-demented older individuals. Two in vivo magnetic resonance imaging (MRI) studies in the context of VaD found CMI in 20% and 32% of patients respectively [6, 7].

CMI of the hippocampal region were detected in an autopsy study in 44% of a total of 57 subjects with AD and cerebral infarcts [8]. Equally, over 50% of patients with vascular dementia showed hippocampal lesions in a neuropathological study of 20 cases [9].

The hippocampi are known to lie in a watershed territory between the internal carotid (anterior choroidal artery) and posterior cerebral artery perfusion territories, and the CA1 (CA, cornu ammonis) subfield is of particular vulnerability to ischemic events. The question was thus raised if the...
occurrence of CMI could be an indicator of hypoperfusion in the hippocampi [10].

Radiologically, CMI depiction was described in experimental settings by Van Veluw et al. [2] using post-mortem ultra-high-field MRI, but also more recently using 3-T MRI, despite their little size, using extended acquisition times of for example 1 h and 52 min for a T2-weighted sequence [11].

In our radiologic-pathologic correlation study using 3-T post-mortem MRI, we aimed to determine the number of hippocampal CMI that could be seen on both radiological and histopathological exams, using a double-blinded radiologic-pathologic approach.

Material and methods

Post-mortem whole-brain 3-T MRI (Magnetom Skyra, Siemens, Erlangen, Germany) using a 64-channel head coil was performed on autopsy specimens of 21 patients. All brain specimens were prepared and scanned following a technique described earlier [12], including a T2-weighted 3D sequence that was used for the radiological analysis in this study by one neuroradiologist (M.S.), in line with previous reports on the MRI detection of CMI [13]. Acquisition parameters were as follows: TR, 1400 ms; TE, 158 ms; bandwidth, 289 Hz/pixel; flip angle, 120°; matrix size, 384×384 pixels; voxel size, 0.1 mm³; acquisition time, 20 min 25 s. CMI were defined as focal and well-delineated spots of elevated signal intensity in any of the three portions of the right hippocampus, first identified on a reconstructed coronal oblique plane and then confirmed on an orthogonal axial plane.

Secondly, the entire hippocampi of the right hemisphere were embedded in paraffin and cut into 50-µm-thick coronal-oblique slices, perpendicular to the long axis of the hippocampal corpus. Each 20th slice was stained with cresyl violet (Nissl stain). The slides were examined by a neuropathologist (E.V.K.) to identify CMI. The neuropathologist’s results, considered the gold standard, were blinded as referred to the radiologist’s findings. Consensus reading was performed thereafter, for comparison of images positivized by both readers.

Results

The results are presented in Tables 1, 2, and 3. In total, 36 CMI of the right hippocampus were found on the Nissl-stained slides of 21 autopsy exams, only 4 of which had a correlate on 3-T MRI, corresponding to a radiological true positive finding (Table 1).
positive rate of 11%. No larger infarcts were present in the resected right-sided hippocampi. Ten right hippocampal signal hyperintensities on 3-T MRI remained without a histopathological correlate and were considered radiological false positives. Radiological sensitivity, specificity, and accuracy were 11.1%, 23.1%, and 2%, respectively. Illustrative examples of true positive, false negative, and false positive CMI after radiologic-pathologic consensus readings are provided in Fig. 1.

Discussion

In the neurological community, an increasing interest is paid to CMI in patients suffering from cognitive decline in a known context of VaD, AD, or mixed dementia (AD plus VaD). With the hippocampi constituting a region of particular interest for cognitive decline and dementia, we decided to focus our study on lesions of this organ. Our work on 21 patients showed CMI to be of limited visibility on post-mortem 3-T MRI with imaging parameters resembling clinical routine imaging, with a radiological true positive rate of only 11% after radiologic-pathologic correlation. This might be attributed to small CMI size in our series, far below an accepted upper limit of 4 mm, and approaching the limits of spatial resolution on 3-T MRI. On the other hand, 7-T MRI studies showed promising results in detecting small CMI [11], but analyses were not focused so far on the hippocampal region. The important question of the correlation between CMI size and clinical repercussion is beyond the scope of this report but needs to be further clarified.

In addition to the limited number of true positives on 3-T MRI, we found a significant number of radiological false positives (10/21), thought to be due to superficial preparation-related lesions in the brain specimens and small hippocampal remnant cysts (between the dentate gyrus and the cornu ammonis). As a consequence, overall performance of the method was low, with an accuracy of only 2%.

Contrarily to earlier studies, we used a double-blinded approach between the radiologist and the pathologist, instead of using MRI-detected lesions as targets for histological analysis. We also opted for a shorter imaging time as compared to previous reports, closer to routine scanning of alive patients, as our main interest was clinical applicability in the case of positive results.

Several limitations of the study should be noted. If the detectability of microinfarcts is already low for 1.5-T and 3-T MRI in the hippocampal region, small physiologic cysts (of the abovementioned type) are frequent lesions that may cause further false positive findings. Another limitation lies in the lack of correlation between in vivo and post-mortem MRI, where future studies will have to further investigate if the process of formalin fixation changes the volume of small cystic lesions within the brain parenchyma. Finally, it has to be kept in mind that even the histopathological exam, considered the gold standard, remains incomplete. In our study only each 20th slice of the right hippocampus underwent analysis, and it cannot be excluded that small CMI lesions remained unseen in the 1-mm interslice gaps. In a future study with a similar design to ours, a systematic “second look” approach might be useful, where all lesions positivized by MRI are subject to novel analysis of additional “between gaps” histopathological slices.

In conclusion, CMI are increasingly recognized to in dementia research, possibly contributing independently to cognitive decline. Post-mortem 3-T MRI does not seem to be an adequate technique for the detection of these lesions when located in the hippocampi. Moreover, false positive findings might be secondary to small superficial defects occurring during specimen preparation, or small physiologic cysts frequently found in this region.

Table 2 A 2×2 table showing relation of radiological and histological findings

| MRI CMI negative | MRI CMI positive |
|------------------|-----------------|
| Histological CMI negative | 3 | 10 |
| Histological CMI positive | 32 | 4 |

CMI, cortical microinfarct; MRI, magnetic resonance imaging

Table 3 A 3×3 table summarizing findings of Tables 1 and 2 to show sensitivity, specificity, and accuracy of 3-T MRI to detect hippocampal CMI, with histological analysis presenting the reference standard

| Reference standard | Reference standard |
|--------------------|--------------------|
| Histological CMI positive | Histological CMI negative |
| Index test MRI CMI positive | 4 (TP) | 10 (FP) | PPV 28.6% |
| Index test MRI CMI negative | 32 (FN) | 3 (TN) | NPV 8.6% |
| Sensitivity 11.1% | Specificity 23.1% | Accuracy 2.0% |

CMI, cortical microinfarct; MRI, magnetic resonance imaging; TP, true positive; FP, false positive; FN, false negative; TN, true negative; PPV, positive predictive value; NPV, negative predictive value
Fig. 1 Post-mortem 3-T magnetic resonance imaging (MRI) T2-weighted image in coronal reconstruction shows millimetric signal hyperintensity in right hippocampal head (a, arrow), due to a cortical microinfarct (CMI) on the corresponding Nissl-stained histopathological image (b, asterisk on the main image, arrow on the inset). False negative MRI image (e) shows homogenous right hippocampal tail, without depiction of histopathologically evident CMI (d, asterisk on the main image, arrow on the inset). 3-T MRI T2-weighted image in coronal reconstruction shows focal signal hyperintensity in right hippocampal body (e, arrowhead), without a correlate on the subsequent histopathological exam (f). The image was considered a false positive finding.

Author contribution MS performed the main writing of the article. MS and EVK prepared the article’s figure. MS and SH prepared the article’s tables. EM and RS gave major contributions to improve the presented MR acquisition sequence and helped optimizing the workflow, including preparation and handling of the brain specimens for the MRI studies, together with EVK. Critiques and corrections were provided by EM, RS, MLM, and EVK. SH designed the work and oversaw its progress and final presentation giving continuous feedback. All authors approved the manuscript.

Funding Open Access funding provided by Université de Genève. This study was funded by a Swiss National Science Foundation grant (grant decision no. 320030_159990/1).

Declarations

Conflict of interest The authors declare that they have no conflict of interest.

Ethical approval The retrospective study involving human participants was approved by the Research Ethics Committee of Geneva (project no. 14–213). All procedures performed in the analysis were in accordance with the 1964 Helsinki Declaration and its later amendments.

Informed consent Informed consent was obtained from all individual participants included in the study or from their next of kin.

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References

1. Damasceno BP (2012) Relationship between cortical microinfarcts and cognitive impairment in Alzheimer’s disease. Dement Neuropsychol 6(3):131–136
2. Van den Brink H, Zwiers A, Switzer AR, Charlton A, McCreary CR, Goodyear BG, Frayne R, Biessels GJ, Smith EE (2018) Cortical microinfarcts on 3T magnetic resonance imaging in cerebral amyloid angiopathy. Stroke 49(8):1899–1905
3. Kövari E, Gold G, Herrmann FR, Canuto A, Hof PR, Michel JP, Bouras C, Giannakopoulos P (2004) Cortical microinfarcts and demyelination significantly affect cognition in brain aging. Stroke 35(2):410–414
4. Gold G, Kövari E, Herrmann FR, Canuto A, Hof PR, Michel JP, Bouras C, Giannakopoulos P (2005) Cognitive consequences of thalamic, basal ganglia, and deep white matter lacunes in brain aging and dementia. Stroke 36(6):1184–1188
5. Brundel M, de Bresser J, van Dillen JJ, Kappelle LJ, Biessels GJ (2012) Cerebral microinfarcts: a systematic review of neuropathological studies. J Cereb Blood Flow Metab 32(3):425–436
6. Ferro DA, van Veluw SJ, Koek HL, Exalto LG, Biessels GJ (2017) Cortical cerebral microinfarcts on 3 Tesla MRI in patients with vascular cognitive impairment. J Alzheimers Dis 60(4):1443–1450
7. Van Veluw SJ, Hilal S, Kuijf HJ, Ikram MK, Xin X, Yeow TB, Venkatesubramanian N, Biessels GJ, Chen C (2015) Cortical microinfarcts on 3T MRI: clinical correlates in memory-clinic patients. Alzheimers Dement 11(12):1500–1509
8. Del Ser T, Hachinski V, Merskey H, Munoz DG (2005) Alzheimer’s disease with and without cerebral infarcts. J Neurol Sci 231(1–2):3–11
9. Vinters HV, Ellis WG, Zarow C, Zaias BW, Jagust WJ, Mack WJ, Chui HC (2000) Neuropathologic substrates of ischemic vascular dementia. J Neuropathol Exp Neurol 59(11):931–945
10. Suter OC, Sunthorn T, Kraftsik R, Straubel J, Darekar P, Khalili K, Miklossy J (2002) Cerebral hypoperfusion generates cortical watershed microinfarcts in Alzheimer disease. Stroke 33(8):1986–1992
11. Van Veluw SJ, Zwanenburg JJ, Engelen-Lee J, Spliet WG, Hendriks J, Luijten PR, Biessels GJ (2013) In vivo detection of cerebral cortical microinfarcts with high-resolution 7T MRI. J Cereb Blood Flow Metab 33(3):322–329
12. Scheffler M, Maturana E, Salomir R, Haller S, Kovari E (2018) Air bubble artifact reduction in post-mortem whole-brain MRI: the influence of receiver bandwidth. Neuroradiology 60(10):1089–1092
13. Miyata M, Kakeda S, Yoneda T, Ide S, Watanabe K, Moriya J, Korogi Y (2018) Signal change of acute cortical and juxtacortical microinfarction on follow-up MRI. AJNR Am J Neuroradiol 39(5):834–840

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