A pharmacokinetic comparison of intravenous versus intra-arterial folinic acid

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Summary Recent clinical trials have suggested that a combination of folinic acid and 5-fluourouracil (5-FU) may improve response rates and survival in patients with advanced colorectal cancer. However, this regimen has been complicated by potentially life threatening toxicity. Regional delivery of folinic acid via a hepatic artery catheter might be expected to reduce systemic exposure and subsequent adverse effects. The present study compared the pharmacokinetic profiles of intravenous and intra-hepatic arterial infusions of folinic acid in patients with colorectal liver metastases (n = 6) who were being treated with weekly regional infusions of 5-FU. The mean area under the plasma concentration–time curve, the peak plasma concentration and the steady state volume of distribution were 163 μg ml⁻¹ h⁻¹ (SD 41), 18.5 μg ml⁻¹ (SD 1.2) and 7.4 ml⁻¹ (SD 0.44) respectively following intravenous administration of folinic acid compared with 142 μg ml⁻¹ h⁻¹ (SD 45), 14.8 μg ml⁻¹ (SD 2.4) and 11.2 ml⁻¹ (SD 1.22) following intra-hepatic arterial administration (P < 0.05). Regional reduction in systemic exposure compared with the intravenous route.

The outlook for patients with colorectal liver metastases remains depressing; the mean survival for patients in the West of Scotland is approximately 3 months (Wood et al., 1976). The results of systemic chemotherapy have been disappointing. Average response rates of only 10–15% have been reported following treatment with 5-FU and response has not been accompanied by increased survival (Kemeny, 1983). These reports have lead to a third of surgeons in England and Wales opting not to actively treat patients with non-resectable colorectal liver metastases (Karanjia et al., 1990) and trials in the UK continue to include “no active treatment” control arms (Hunt et al., 1990).

Recent studies, however, have suggested that the addition of folinic acid may significantly improve survival amongst advanced colorectal cancer patients receiving 5-FU (Erlrichman, 1988, Poon et al., 1989; Kerr, 1989). Briefly, folinic acid enhances 5-FU activity by stabilising the binding of the 5-FU metabolite, fluorodeoxyuridine monophosphate, to the enzyme thymidylate synthase. Unfortunately, therapy has been complicated by systemic toxicity which can be occasionally life-threatening.

The rationale for regional chemotherapy is the delivery of high drug concentrations to the compartment harbouring the tumour with relatively less drug escaping into the systemic vascular compartment. There is a large literature on intra-hepatic arterial administration of 5-FU and FUDR to patients with hepatic metastases from colorectal primary cancers. In summary, the data are suggestive that tumour response rates are higher comparing regional with systemic administration, but there is no convincing evidence that intra-arterial chemotherapy significantly prolongs survival.

The aim of the present study was to compare the pharmacokinetic profiles of intravenous and intra-hepatic arterial folinic acid in patients who were receiving a 24 h infusion of 5-FU via an indwelling hepatic artery catheter.

Patients and methods

Patients

Patients with biopsy-proven, metastatic, colorectal adenocarcinoma, confined to the liver were recruited. All subjects had previously undergone a resection of an adenocarcinoma of the colon or rectum and surgical placement of a hepatic artery catheter (Infusaport 38940). Shiley Infusaid Inc., Norwood, MA, USA. Entry criteria were; WHO performance status < 2, life expectancy > 2 months, white cell count > 4 × 10⁹ l⁻¹ platelets > 150 × 10⁹ l⁻¹ and bilirubin < 30 μmol l⁻¹. Pre-treatment staging consisted of physical examination, abdominal CT scan and chest radiograph. All patients gave informed consent prior to entering the study.

Treatment

Patients received a 24 h arterial infusion of 5-FU, 600 mg m⁻² per week, for 6 consecutive weeks. On the fourth and sixth weeks of this regimen, folinic acid (100 mg m⁻²) was administered as a 2 h infusion. On one occasion the folinic acid was given intravenously and on the other occasion it was administered via the hepatic artery. 5-FU and folinic acid infusions were commenced simultaneously. The order of the route of folinic acid delivery was random.

Pharmacokinetic assessments

Five ml of peripheral venous blood was sampled before each folinic acid infusion and at 1, 2, 2.5, 2.5, 3, 4, 4.5, 5, 6, 8, 14 and 26 h after commencement of the infusion. Blood was placed in tubes containing lithium heparin which were stored in ice. With minimal delay (less than 5 min) the blood was centrifuged at 5,000 r.p.m. for 5 min then the plasma was removed and stored at −20°C in light protected vials.

As folinic acid has the propensity to be oxidised, control experiments were performed, spiking blood samples with known quantities of folinic acid, to ensure that plasma plates were stable during collection and separation. Folinic acid degradation was always less than 10% under these conditions.

Serum levels of folinic acid were assayed using a sensitive and specific HPLC assay system (inter and intra-assay coefficients of variation < 5%) with a limit of detection of 50 ng ml⁻¹. A C18 microbondapack (10 microns) column is used. The mobile phase consists of two components. Mobile phase A; 0.25 M phosphate buffer (pH 5); Mobile phase B, 50% 0.25 M phosphate buffer (pH 5), 50% methanol. The mobile phase consists of phase A throughout with a 50% contribution from phase B between 10–15 min run time. The extraction method depends on the addition of cold methanol to plasma (1.5:1 vol/vol) and subsequent vortex evaporation of the supernate prior to redissolution and injection onto the column in H₂O. Detection was performed by UV analysis at 310 nm.

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Plasma levels of folinic acid were plotted against time and the data were fitted to an infusional compartment open model, by the method of least squares, using an ‘in house’ computer programme based on the Marquhardt algorithm. The area under the plasma concentration-time curve was extrapolated to infinity. Peak concentrations were actual end of infusion concentrations and the volume of distribution was computed from the pharmacokinetic model parameters (A, α, B, β) corrected for duration of infusion.

Statistical methods

The data were analysed using the paired student’s t-test.

Results

Patients

Six patients entered the study (four males, two females). Mean age was 61 years (range 39–75). Three patients had previous treatment for their metastases (two with mitomycin C and one with regional $^{90}$Y-glass microspheres) but their disease had progressed despite these therapies.

Pharmacokinetics

The pharmacokinetic parameters for folinic acid are summarised in Table I. The plasma profiles for patient 1 are shown in Figure 1. The mean area under the time/concentration curve was 163 µg ml$^{-1}$ h (SD 41) following intravenous administration of folinic acid and 142 µg ml$^{-1}$ h (SD 45) following the arterial route ($P < 0.02$). The mean peak plasma concentration was 18.5 µg ml$^{-1}$ (SD 1.2) following intravenous administration and 14.8 µg ml$^{-1}$ (SD 2.4) with the arterial route ($P = 0.02$). The mean volume of distribution at steady state was 7.421 m$^{-3}$ (SD 0.44) following intravenous administration and 11.21 m$^{-3}$ (SD 1.22) with the arterial route ($P = 0.028$). There was no significant difference for clearance, $t_{1/2}$ or $t_{1/2}$ of folinic acid comparing intravenous and regional delivery.

Toxicity

No patients exhibited any evidence of treatment related toxicity. In particular, there were no signs of myelosuppression, liver failure, diarrhoea, stomatitis or vomiting. Subsequent to completion of the pharmacokinetic study, three patients, who received further intra-arterial infusions of folinic acid, experienced occlusion of their hepatic artery catheter.

Discussion

Regional infusion of 5-FU reduces systemic exposure to the drug and therefore diminishes systemic toxicity (Goldberg et al., 1990). Our attempts, in the present study, to apply these principles to folinic acid administration have revealed a statistically significant regional advantage for the arterial route. The pharmacokinetic parameters following intravenous administration of folinic acid are similar to those described in previous studies (Straw et al., 1984; McGuire et al., 1988; Trave et al., 1988; Rustum, 1989).

Intra-arterial infusion of folinic acid reduces the plasma AUC and end of infusion plasma concentration and increases its volume of distribution relative to intravenous administration. We are not sure why the volume of distribution is greater following loco-regional infusion but it is possible that this could be due to increased first pass folinic acid extraction by the liver. We plan to clarify this point by measuring drug concentrations in tumours and normal liver biopsies taken during peroperative folinic acid infusions.

There was a trend towards prolongation of drug half-life following arterial infusion, but this did not reach significance. Folinic acid is cleared predominantly by the liver by metabolism to 5 methyl tetrahydrofolate, however it was not

| Table 1 Comparison of pharmacokinetic parameters for folinic acid following intravenous or intra-hepatic arterial administration |
|---------------------------------------------------------------|
| **Parameter** | **Patient** | 1 | 2 | 3 | 4 | 5 | 6 | Mean | SE |
|----------------|------------|---|---|---|---|---|---|------|----|
| Area under curve (µg ml$^{-1}$ h$^{-1}$) | IV | 100 | 132 | 210 | 166 | 179 | 191 | 163 | 16.6 |
| | IA | 69 | 115 | 203 | 156 | 161 | 148 | 142 | 18.5 |
| Peak concentration (µg ml$^{-1}$) | IV | 18.7 | 17.5 | 19.2 | 17.3 | 18.1 | 20.4 | 18.5 | 0.5 |
| | IA | 11.7 | 12.5 | 16.8 | 16.9 | 13.9 | 13.9 | 14.8 | 0.9 |
| Clearance (1 h$^{-1}$ m$^{-3}$) | IV | 1.0 | 0.76 | 0.48 | 0.6 | 0.56 | 0.52 | 0.65 | 0.08 |
| | IA | 1.44 | 0.87 | 0.49 | 0.64 | 0.62 | 0.68 | 0.79 | 0.13 |
| Steady state volume of distribution (1 m$^{-3}$) | IV | 6.1 | 7.6 | 6.7 | 9.2 | 7.0 | 7.9 | 7.4 | 0.44 |
| | IA | 12.3 | 16.2 | 10.8 | 9.2 | 7.4 | 11.1 | 11.2 | 1.22 |
| $t_{1/2}$ (h) | IV | 0.3 | 0.1 | 0.2 | 0.6 | 1.5 | 0.5 | 0.53 | 0.21 |
| | IA | 0.7 | 2.9 | 5.3 | 1.2 | 0.5 | 2.3 | 2.15 | 0.74 |
| $t_{1/2}$ (h) | IV | 4.8 | 7.9 | 14.9 | 17.3 | 11.0 | 16.4 | 12.1 | 2.05 |
| | IA | 7.5 | 32.0 | 36.7 | 16.5 | 10.7 | 20.1 | 20.7 | 5.81 |
possible to measure this metabolite in the present study. The regional advantage conferred by hepatic arterial of folic acid can be assessed by the ratio of plasma AUC's or end of infusion peak plasma concentrations; the larger the ratio, the greater the regional advantage (AUCreg:AUCsys = 1.15, Cmaxreg:Cmaxsys = 1.25). The regional advantage conferred by hepatic arterial infusion of folic acid is relatively small compared to drugs such as 5-fluorouracil (Goldberg et al., 1990).

Although there is an apparent pharmacokinetic advantage, this is offset by the potential for catheter thrombosis. In our experience, which includes placement of hepatic artery catheters in more than 70 patients, 5-FU can be infused regionally without complications. However, the addition of regional folic acid to this treatment regimen has been associated with catheter occlusion in three out of six patients so treated. This information is anecdotal and has not been the subject of a formal study but we believe that it is important to supply these data. There was no other apparent treatment related toxicity.

Clinically, there are two strategies to try to overcome the problem of systemic relapse following loco-regional therapy: combination of regional with systemic therapy or dose escalating the regional therapy and quality of life in patients with advanced colorectal carcinoma. We have decided to adopt the latter approach and we are currently undertaking a phase I trial of intravenous folic acid in combination with a 24 h intrahepatic arterial infusion of 5-FU. The folic acid schedule has been fixed and the dose of 5-FU will escalated until plasma concentrations are similar to those achieved with intravenous infusions. This should allow the generation of high levels within the liver, the site of predominant bulk disease, but maintain adequate systemic levels.

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