Biopreservation of Apricot pulp and Apricot Ready to Serve by using Bacteriocin of *Lactobacillus brevis* SM6 and their Comparison with Chemical Preservative

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**Abstract**

Food safety is a growing public health concern of both in the developing, and developed nations. The consumption of chemically preserved foods raises the concern among the consumer for naturally produced antimicrobial agents. Therefore, in the present study an attempt has been made for the preparation of apricot RTS and apricot plum with bacteriocin *Lactobacillus brevis* SM6 (crude and purified) and was compared with standard chemical preservative (sodium benzoate) to evaluate the effect of biopreservation in the food. Bacteriocin was added in the food products following direct and indirect approach. The effect of preservatives in the prepared food products was studied and found that bacteriocin of *Lactobacillus brevis* SM6 was at par with chemical preservative in preserving apricot RTS and apricot pulp efficiently by preventing the growth of spoilage causing pathogens. The effect of bacteriocin was also compared statistically with chemical preservative to prove its efficacy for biopreservation.

**Introduction**

Nature has given many types of foods to the mankind. Everybody expects that the food they eat is wholesome and safe for consumption. The greatest threat to quality and safety of our food comes from the microbial spoilage (Pal, 2013). Preservatives are a type of food additive which are added to food to prolong shelf life, and keep the products from being broken down by microorganisms (Pal, 2014), though chemical preservatives for the preservation of food are successful to some extent, their quality is not as satisfying as fresh food. Therefore there is an increase demand in the market for the use of natural source as biopreservative to combat food spoilage. Among bio-preservatives, bacteriocin has caught the attention of food scientists to be used as a natural food biopreservative due to its antimicrobial activity against food spoilage and pathogenic bacteria.

Bacteriocins are ribosomally-synthesized antimicrobial peptides or proteinaceous compounds produced by bacterial strains. They are generally effective in inhibiting the growth of similar or closely related bacterial
strains. Among biopreservatives, bacteriocin has caught attention of food scientists to be used as a natural food biopreservative due to its antimicrobial activity against serious pathogenic and food spoilage bacteria (Gupta et al., 2015). A high diversity of various bacteriocins is produced by many lactic acid bacteria (LAB) and is found in numerous fermented and non-fermented foods. LAB is used as natural or selected starters in food fermentations in which they perform acidification due to the production of lactic and acetic acids (Gupta and Sharma, 2017). Several bacteriocins from LAB extend potential applications in food preservation, thus help foods to be naturally preserved and richer in organoleptic and nutritional properties. Among biopreservatives, nisin has already been used successfully at commercial scale in dairy products (Sharma and Gautam, 2007). Biopreservation, defined as the extension of shelf life and enhanced safety of foods by the use of natural or controlled microbiota and/or antimicrobial compounds. Two approaches of food biopreservation are generally followed:

Direct approach: Addition of bacteriocin to the food product

Indirect approach: Addition of bacteriocinogenic lactic acid bacteria to the food product.

These methodologies of food biopreservation are selected on the basis of many intrinsic and extrinsic factors such as nature of the food product. As in non fermented food products or beverages bacteriocin mediated biopreservation is more productive than the use of cell cultures of probiotics. Since, bacteriocins are bioproducts they do not tend to change the properties of food product such as, its flavor, colour, texture physicochemical properties etc. but their antimicrobial properties tend to eliminate or restrict the pathogenic bacteria thus, enhancing the stability and shelf life of food products. Live bacteriocinogenic probiotics can be directly added to food products which can be preserved by fermentation as the probiotics contribute to preservation through production of various antimicrobial compounds. In the present investigation both these methods of biopreservation were applied and their effects were studied. The majority of bacteriocin producers are natural food isolates and hence they are suited for food applications (Raja et al., 2010). Application of bacteriocin may help to reduce the use of chemical preservatives and the intensity of heat and other physical treatments, satisfying the demand of consumers for foods that are fresh in taste, ready to eat and lightly preserved. Moreover, bacteriocins being proteinaceous in nature, once administered in gastrointestinal tract, do not have any residual effect, contrary to chemical preservatives which are highly health hazardous in nature. In the present study, therefore an attempt has been made to study the biopreservative effective of bacteriocin on apricot RTS and apricot pulp prepared and its comparison with the generally used chemical biopreservative (sodium benzoate).

Materials and Methods

Food biopreservation

Apricot (Prunus armeniaca L.) RTS beverage and pulp were taken in this study to determine the effect of bio-preservation in them during storage. Each product was treated separately with purified and partially purified bacteriocin of Lactobacillus brevis SM6, chemical preservative sodium benzoate and the bacteriocin producer LAB i.e. Lactobacillus brevis SM6. Purified and partially purified bacteriocin were added @ 5000 AU/ml each and the chemical preservative sodium benzoate was added in concentration of 2000
Controls were also run in parallel i.e. food products without any preservative. The products were kept for storage under refrigerated conditions for 30 days. The quality evaluation of the products was done periodically on 0, 7, 14, 21 and 30 days.

In total 6 sets were made with each product as given below.

**Bacteriocin mediated preservation**

**SET-A** - Control (pasteurized sample) + (pathogen) *Listeria monocytogenes* (10^8 CFU/ml)

**SET-B** - Product + (pathogen) *Listeria monocytogenes* (10^8 CFU/ml) + Crude bacteriocin (5000 AU/ml)

**SET-C** - Product + (pathogen) *Listeria monocytogenes* (10^8 CFU/ml) + purified bacteriocin (5000 AU/ml)

**SET-D** - Product + (pathogen) *Listeria monocytogenes* (10^8 CFU/ml) + chemical preservative (2000 ppm)

**Potential bacteriocinogenic lactic acid bacteria mediated preservation**

**SET-A** - Control (pasteurized sample) + (pathogen) *Listeria monocytogenes* (10^8 CFU/ml)

**SET-E** - Product + (pathogen) *Listeria monocytogenes* (10^8 CFU/ml) + *Lactobacillus brevis* SM6 (10^8 CFU/ml)

**SET-F** - Product + *L. monocytogenes* (10^8 CFU/ml)

**Pulp preparation**

**Ingredients**

i. Ripened apricot: 600 g

ii. Water: 200 ml

**Recipe**

1. Ripened fruits were taken
2. Fruits were peeled
3. Peeled fruits were boiled in minimum amount of water for 10 minutes
4. The fruits were then mashed
5. 6 sets were made accordingly

**SET A** **SET B** **SET C** **SET D** **SET E** **SET F**

Finished apricot pulp

Refrigeration at 4°C

Evaluation of quality attributes.
Sensorial evaluation

Sensorial evaluation of each set was done in terms of appearance, texture, flavor, odor and overall acceptability. Nine point hedonic scale method as given by Amerine et al., (1965) was followed for conducting the sensory evaluation of the food product.

The panel of 10 judges was selected to evaluate the products for sensory parameters such as appearance, flavor, texture, taste, odor and overall acceptability depending upon the type of product.

Efforts were made to keep the same panel for sensory evaluation throughout the course of study.

Physicochemical changes during storage

pH

pH of each set was measured using pH meter at an interval of 0, 7, 14, 21 and 30 days.

Total soluble solids (TSS)

TSS was measured by placing 1-2 drops of sample on the prism of a hand refractometer. For 0, 7, 14, 21 and 30 days and the results were expressed as °B (Ranganna, 2009).

Acidity in terms of lactic acid

An aliquot of the sample prepared was diluted with recently boiled distilled water. 2-3 drops of 1% phenolphthalein solution was used as an indicator and titration was done with 0.1N NaOH. Titre value was noted and calculations were done as percent anhydrous lactic acid.

Ascorbic acid

Ascorbic acid was determined as per AOAC (1995) method for 0, 7, 14, 21 and 30 days.

Total carbohydrates

Total carbohydrates were determined as per Sadasivam and Manickam (1992).

Total proteins

Total proteins were determined by following the method given by Ranganna (2009).

Crude fibers

Crude fibers were determined as per the method described in AOAC (1995).

Free radical scavenging activity (FRSA) (Brand et al., 1995)

Free radical scavenging activity was measured as per the method of Brand et al., (1995). DPPH (2, 2-diphenyl-1-picrylhydrazyl) was used as a source of free radical.

Microbial study during storage

The colony count was observed during storage period of 0, 7, 14, 21 and 30 days by standard spread plate count method. MRS agar was used to enumerate lactic acid bacteria while nutrient agar was used to enumerate Listeria monocytogenes.

Statistical analysis

Data pertaining to the physicochemical attributes and data on sensorial evaluation of prepared food product (apricot RTS) was analyzed by completely randomized design (CRD) factorial as described by O’Mahony and Decker (1985).

RTS preparation

Ingredients

Apricot pulp: 60 g
Sugar: 52.8 g
Water: 545.16 ml
Acid: 1.56 g
Recipe

Apricot (pulp/juice)

Mixing with strained syrup solutions (sugar+water+acid in quantities as mentioned above)

Homogenization

Bottling (100 ml in each bottle)

Corking

Pasteurization (at 90°C for 25 min)

Cooling (to room temperature)

Treatments (6 Sets were made accordingly)

SET A SETB SET C SET D SET E SET F

Refrigeration at 4°C

Evaluation of quality attributes

Sensorial evaluation

Sensorial evaluation of each set was done in terms of appearance, texture, flavor, flavor, odor and overall acceptability as given by Amerine et al., (1965)

Physicochemical changes during storage

The physiological changes i.e. pH, total proteins, total carbohydrates and crude fibers were performed as mentioned in section 2.2.4.

Microbial study during storage

The microbial study was done same as mentioned in section 2.2.5

Statistical analysis: was done according to the method described by O’Mahony and Decker (1985).
**Results and Discussion**

In the present study, apricot RTS and apricot plum preparations with bacteriocin (crude and purified) was compared with standard chemical preservative (sodium benzoate) to evaluate the effect of bio-preservation in the food and a negative control was run along with these to detect the efficacy of the preservative. The food products selected for experiment were kept for 30 days and the effect was noted at 0, 7th, 14th, 21st and 30th day. The chemical preservative was added at permissible limits i.e. 2000 ppm. The bacteriocin activity which was found to be 8000 AU/ml and 6000 AU/ml in case of purified and partially purified bacteriocin was kept at 5000 AU/ml (Ray *et al.*, 2001).

**Prunus armeniaca L. (Apricot)**

Apricot (*Prunus armeniaca* L.) is a very important stone fruit of temperate as well as mid hills of Himachal Pradesh. The fruit has a nutritional value of interest having a good source of fiber, minerals (especially potassium but also calcium, iron, magnesium, zinc, phosphorus and selenium) and vitamins such as vitamin A, vitamin C, thiamin, riboflavin, niacin and pantothenic acid. The total mineral content of the pulp, as represented by its ash, is 2.452%. The percentage content of different mineral elements in the pulp is phosphorus, 0.083; potassium, 0.996; calcium, 0.042; magnesium, 0.033; and iron, 0.010. The protein is only 0.67 per cent. Dry matter in apricot is about 10-20%; main constituents are sugar, polysaccharides and organic acids (*Belitz et al.*, 2009). Fruits contain 0.1-1.5% nitrogen compounds; 35-75% of these nitrogen compounds are proteins. Free amino acids are also abundant. Moreover, apricots contain a number of main secondary metabolites such as polyphenols, carotenoids, fatty acids, volatiles and polysaccharides. In particular, phenolic compounds are one of the main sources of antioxidant activity which are able to prevent oxidative stress scavenging free radicals and nitrogen species (*Erdogan and Kartal*, 2011).

**Apricot pulp**

Apricot pulp was prepared as per the method given by Rangana (2009). The ingredients required were- ripened apricot (600 g) and water (150 ml). First of all the ripened fruits were taken and peeled. Peeled fruits were then boiled in minimum amount of water for 10 min and were then mashed. Different treatments were then applied accordingly as described above. All the sets were kept for 1 month for further evaluation. Nutritional facts of fresh apricot pulp without any fortification has been presented in Table 1.

Sets: In total 6 sets were made

**Direct approach:** Bacteriocin mediated

- **SET-A** - Product + *L. monocytogenes* (10⁸ CFU/ml)
- **SET-B** - Product + *L. monocytogenes* (10⁸ CFU/ml) + Crude bacteriocin (5000 AU/ml)
- **SET-C** - Product + *L. monocytogenes* (10⁸ CFU/ml) + purified bacteriocin (5000 AU/ml)
- **SET-D** - Product + *L. monocytogenes* (10⁸ CFU/ml) + chemical preservative (2000 ppm)

**Indirect approach:** Bacteriocinogenic bacteria mediated

- **SET-A** - Product + *L. monocytogenes* (10⁸ CFU/ml)
- **SET-E** - Product + *L. monocytogenes* (10⁸ CFU/ml) + *L. brevis* SM6 (10⁸ CFU/ml)
- **SET-F** - Product + *L. brevis* SM6 (10⁸ CFU/ml)

To study the effects at experimental level 25 ml of product was taken in separate tubes...
(Fig. 1 a & b) to which the preservatives were added accordingly. Each set contained 25 ml pulp and 1 ml (10^8 CFU/ml) L. monocytogenes was added to each set to desirably cause spoilage and to study the effects of various treatments. Crude bacteriocin (20.75 ml) was added to set B and purified bacteriocin (15.63 ml) was added to set C. Further evaluation of all the sets was done periodically for one month.

**Sensorial evaluation**

Freshly prepared apricot pulp samples were assessed by 10 panelists using a 9 point sensory hedonic scale for some sensory parameters (viz. appearance, flavor, texture, taste and overall acceptability), as described by Amerine et al., (1965). In a sensory evaluation set A was least accepted (with overall acceptability 4.94) whereas set C had a maximum acceptability (8.42) (Fig. 2). Statistically sensorial evaluation was carried out by Completely Randomized Design (CRD). The results showed a significant effect of different treatments on sensory attributes of apricot pulp and also indicate that the type of bacterial strain contributes a significant influence on the overall acceptability of the product. Castro et al., (2017) did sensory evaluation of banana pulp in terms of colour, texture, flavor, taste, odor and overall acceptability.

**Microbiological study of apricot pulp**

During biopreservation studies it was noticed that control (i.e. without addition of bacteriocin) has shown rapid growth of L. monocytogenes leading to spoilage within three days of product formulation. On the other hand crude bacteriocin (Set B) has created the preservative effect till 7th day while purified bacteriocin (Set C) and chemical preservatives (Set D) were found the most efficient to check the growth of L. monocytogenes and keeping the product preserved till 30th day with the meager increase in L. monocytogenes log CFU/ml i.e. 4.50 and 5.99 respectively. This experiment proves that purified bacteriocin is capable of preserving apricot pulp efficiently.

In case of direct addition of bacteriocinogenic LAB it was found that LAB number started decreasing gradually from 7th day (log CFU/ml 4.85) onwards upto 30 days (2.80 log CFU/ml) in the product probably due to unfavourable environment. The preservation effect of bacteriocin produced by L. brevis OG1 was evaluated by Ogunbanwo and Okanlawon (2006), for chicken immobilized inedible films. L brevis OG1 produce bacteriocin that has a broad spectrum of inhibition against pathogens and food spoilage organisms created during storage as evident from physicochemical parameters (pH, TSS, TA, ascorbic acid). As far as growth of L. monocytogenes is concerned in these products biopreservative effect was observed with minimal increase in their log CFU/ml i.e. 4.90 (0 day) to 5.15 (30th day) (Fig. 3 & 4). The biopreservative effect in this case is due to different antimicrobial compounds secreted by L. brevis SM6 in this culture medium (lactic acid, H2O2, acetic acid alongwith bacteriocin). Upon comparison between direct and indirect approach, it is clear that addition of purified bacteriocin is the best option for food biopreservation probably because of its powerful antagonistic potential expressed i.e. 8000 AU/ml present in it. Addition of bacteriocin/bacteriocinogenic LAB for food preservation is preferred according to the nature of the food products as many intrinsic and extrinsic parameters affect food processing and preservation, e.g. in non-fermented refrigerated products only purified or semi-purified bacteriocin preparations are applied for preservation rather than applying cell culture. The direct addition of purified
bacteriocin provides a more controlled preservative tool in such products. Furthermore, bacteriocin producing cells of lactic acid bacteria having probiotic potential are also added in food products which can be preserved by means of fermentation. These lactic acid bacteria contribute in preservation, as during preservation these will produce different antimicrobial compounds such as lactic acid, acetic acid, hydrogen peroxide, carbon dioxide besides bacteriocins which altogether inhibit pathogenic and spoilage causing microorganisms, extending their shelf life and enhancing the safety of food products.

Ogunbanwo and Okanlawon (2006), observed that immobilization of bacteriocin produced by L. brevis OG1 in calcium alginate gel and application to the surfaces of lean tissue (cutaneous) of broiler chicken was effective for reducing microbial load up to 21 days of refrigerated storage.

**Changes in quality attributes of apricot pulp during storage**

Prepared apricot pulp samples were kept for storage for 30 days and the quality attributes (pH, total soluble solid, titratable acidity and ascorbic acid) of each set were noted on 0, 7th, 14th, 21st and 30th days of storage as described in Table 2. pH followed the same trends as apricot RTS, it was observed that the pH values slightly decreased during the storage. On 30th day maximum decrease was noticed in set A (2.56) and minimum in set C (3.33). Total soluble solid of apricot RTS remained almost same for sets A to E but there was decrease in TSS in set F (11.12%).

Initially TSS values were same for all sets (12%). After 30 days TSS was maximum for set A and minimum for set H. Titratable acidity was measured in terms of lactic acid. Titratable acidity is negatively correlated with pH that means while pH decreased, titratable acidity increased. The initial lactic acid was 0.45% for every RTS set. On 30th day of storage lactic acid % had increased most in set F to 0.96% and was at par in case of set C and D (0.91). Ascorbic acid content of apricot pulp also showed variation in its values during storage period. Initially ascorbic acid content ranges in between 9.85 to 9.88 mg/ml in all sets which rose in every treatment. Statistically changes in quality attributes of apricot during storage period were analyzed by CRD factorial which revealed that there are non-significant results of change in quality attributes of apricot RTS during storage conditions.

**Apricot Ready to Serve (RTS) beverage**

80 g apricot pulp was taken which was mixed in strained sugar syrup (containing 70.4 g sugar and 647.52 ml water) and was properly homogenized. Bottling of the product (100 ml in each bottle) was done, and the bottles were properly corked which were then pasteurized at 90°C for 25 min. They were left for cooling at room temperature and the treatments were applied accordingly. Nutritional facts of fresh apricot RTS without any fortification has been presented in Table 3.

**Sets:** In total 6 sets were made

Direct approach: Bacteriocin mediated

- **SET-A** - Product + L. monocytogenes (10⁸ CFU/ml)
- **SET-B** - Product + L. monocytogenes (10⁸ CFU/ml) + crude bacteriocin (5000 AU/ml)
- **SET-C** - Product + L. monocytogenes (10⁸ CFU/ml) + purified bacteriocin (5000 AU/ml)
- **SET-D** - Product + L. monocytogenes (10⁸ CFU/ml) + chemical preservative (2000 ppm)

Indirect approach: Bacteriocinogenic bacteria mediated

- **SET-A** - Product + L. monocytogenes (10⁸...
CFU/ml) 
SET-E - Product + *L. monocytogenes* (10⁸ CFU/ml) + *L. brevis* SM6 (10⁸ CFU/ml) 
SET-F - Product + *L. brevis* SM6 (10⁸ CFU/ml) 

To study the effects at experimental level 25 ml of product was taken in separate tubes (Plates) to which the preservatives were added accordingly (Fig. 5 a&b). Each set contained 25 ml RTS and 1 ml of *L. monocytogenes* (10⁸ CFU/ml) was added to each set to desirably cause spoilage and to study the effects of various treatments. Crude bacteriocin (20.75 ml) was added to set B and purified bacteriocin (15.63 ml) was added to set C. Further evaluation of all the sets was done periodically for one month.

**Sensorial evaluation**

Freshly prepared apricot RTS samples were assessed by 10 panelists using a 9 point sensory hedonic scale for some sensory parameters (viz. appearance, texture, taste, odor and overall acceptability), as described by Amerine *et al.*, (1965). In sensory evaluation set A was least accepted (with overall acceptability of 5.05), set C had a maximum acceptability (8.46 out of 10) and set B and set D were almost equally acceptable (8.08 and 7.92 respectively) (Fig. 6). The results showed a significant effect of different treatments on sensory attributes of apricot RTS and also indicate that the type of bacterial strain contributes a significant influence on the overall acceptability of the product. Dambalkar *et al.*, (2015) performed sensory evaluation of Beetroot orange RTS drink which was accessed in terms of colour, flavor, texture, taste and overall acceptability. A sensory evaluation of carrot juice supplement with the *L. acidophilus* DSM 20079 was carried out by Goderska *et al.*, (2007) and they found that bacterial strain exerts a significant influence on the sensory evaluation of the examined product.

**Microbiological study of apricot RTS**

During biopreservation studies it was noticed that control (i.e. without addition of bacteriocin) has shown rapid growth of *L. monocytogenes* leading to spoilage within three days of product formulation. On the other hand crude bacteriocin (Set B) has created the preservative effect till 7th day while purified bacteriocin (Set C) and chemical preservatives (Set D) were found the most efficient to check the growth of *L. monocytogenes* and keeping the product preserved till 30th day with the meager increase in *L. monocytogenes* log CFU/ml i.e. 4.57 and 5.78 respectively. This experiment proves that purified bacteriocin is capable of preserving apricot RTS efficiently.

In case of direct addition of bacteriocinogenic LAB it was found that LAB number started decreasing gradually from 7th day (log CFU/ml 4.55) onwards upto 30 days (2.32 log CFU/ml) in the products probably due to unfavourable environment created during storage as evident from physicochemical parameters (pH, TSS, TA, ascorbic acid). As far as growth of *L. monocytogenes* is concerned in these products biopreservative effect was observed with minimal increase in their log CFU/ml i.e. 4.85 (0 day) to 5.75 (30th day) (Fig. 7 & 8). The biopreservative effect in this case is due to different antimicrobial compounds secreted by *L. brevis* SM6 in this culture medium (lactic acid, H₂O₂, acetic acid along with bacteriocin). Upon comparison between direct and indirect approach, it is clear that addition of purified bacteriocin is the best option for food biopreservation probably because of its powerful antagonistic potential expressed i.e. 8000 AU/ml present in it. Addition of bacteriocin/bacteriocinogenic LAB for food preservation is preferred according to the nature of the food products as many intrinsic and extrinsic parameters affect food processing and preservation, e.g. in non-
fermented refrigerated products only purified or semi-purified bacteriocin preparations are applied for preservation rather than applying cell culture. The direct addition of purified bacteriocin provides a more controlled preservative tool in such products. Furthermore, bacteriocin producing cells of lactic acid bacteria having probiotic potential are also added in food products which can be preserved by means of fermentation. These lactic acid bacteria contribute in preservation, as during preservation these will produce different antimicrobial compounds such as lactic acid, acetic acid, hydrogen peroxide, carbon di oxide besides bacteriocins which altogether inhibit pathogenic and spoilage causing microorganisms, extending their shelf life and enhancing the safety of food products.

Changes in quality attributes of apricot RTS during storage

Prepared apricot RTS samples were kept for storage for 30 days and the quality attributes (pH, total soluble solid, titratable acidity and ascorbic acid) of each set were noted on 0, 7th, 14th, 21st and 30th days of storage as described in Table 4. It was observed that the pH values slightly decreased during the storage. On 30th day maximum decrease was noticed in set A and minimum in set E. Total soluble solid of apricot RTS remained almost same for sets A to E but there was significant decrease in TSS in Set A (9.19% ß). Initially all sets had a TSS of 10%β during storage for 30 days. Titratable acidity was measured in terms of lactic acid. Titratable acidity is negatively correlated with pH that means while pH decreased, titratable acidity increased. The initial lactic acid was measured to be 0.15% (Rangana, 2009) for every RTS set. On 30th day of storage lactic acid % had increased most in set A and was minimum in case of set C (0.66%) where it remained same. Ascorbic acid content of apricot RTS also showed variation in its values during storage period. Initially ascorbic acid content was found to be 9.5 mg/ml (Rangana, 2009) in all sets which increased in every treatment. Statistically changes in quality attributes of apricot during storage period were analysed by CRD factorial which revealed that there is non-significant change in results of quality attributes of apricot RTS during storage conditions. The use of L. brevis SM6 for the biopreservation of apricot RTS and apricot pulp proved to be more effective against the spoilage causing pathogens as compared to the chemical preservative (Sodium benzoate) used. Thus, successfully fulfilling the main objective of this study.

Table 1. Nutritional chart of apricot pulp

| Nutritional facts per 100 ml | Samples |
|-----------------------------|---------|
|                            | SET A   | SET B   | SET C   | SET D   | SET E   | SET F   |
| *Antioxidant activity (%)   | 45.50   | 45.51   | 45.50   | 45.51   | 45.51   | 45.50   |
| *Protein (g/ 100 g)        | 1.4     | 1.41    | 1.42    | 1.41    | 1.41    | 1.40    |
| *Fiber (%)                 | 3.30    | 3.31    | 3.30    | 3.30    | 3.31    | 3.30    |
| *Carbohydrate (g/ 100 g)   | 12      | 11.9    | 12      | 12      | 11.9    | 12.1    |
| *Total sugars (%)           | 9.47    | 9.48    | 9.48    | 9.47    | 9.47    | 9.46    |

*values evaluated in lab
Table 2: Physicochemical characteristics of apricot pulp

| Treatments (T) | pH  | TSS (°Brix) | Titratable acidity (%) | Ascorbic acid (mg/100g) |
|----------------|-----|-------------|------------------------|-------------------------|
|                | 0   | 7 | 14 | 21 | 30 | Mean | 0 | 7 | 14 | 21 | 30 | Mean | 0 | 7 | 14 | 21 | 30 | Mean | 0 | 7 | 14 | 21 | 30 | Mean |
| Set A          | 4.40 | 4.30 | 3.12 | 3.01 | 2.56 | 3.47 | 12.0 | 12.0 | 11.51 | 11.34 | 11.22 | **11.61** | 0.45 | 0.69 | 0.75 | 0.85 | 0.95 | **0.73** | 9.86 | 9.82 | 9.73 | 9.60 | 9.47 | **9.69** |
| Set B          | 4.41 | 4.35 | 4.27 | 4.19 | 3.16 | **4.07** | 12.0 | 12.0 | 11.77 | 11.66 | 11.37 | **11.76** | 0.45 | 0.57 | 0.60 | 0.67 | 0.92 | **0.78** | 9.87 | 9.74 | 9.62 | 9.55 | 9.51 | **9.65** |
| Set C          | 4.41 | 4.31 | 4.24 | 4.14 | 3.33 | **4.06** | 12.0 | 12.0 | 11.84 | 11.72 | 11.57 | **11.82** | 0.45 | 0.57 | 0.69 | 0.75 | 0.91 | **0.81** | 9.88 | 9.81 | 9.75 | 9.68 | 9.45 | **9.71** |
| Set D          | 4.41 | 4.34 | 4.22 | 4.13 | 3.10 | **4.10** | 12.0 | 12.0 | 11.74 | 11.53 | 11.46 | **11.74** | 0.45 | 0.58 | 0.66 | 0.76 | 0.91 | **0.81** | 9.85 | 9.76 | 9.67 | 9.60 | 9.57 | **9.69** |
| Set E          | 4.40 | 4.03 | 3.77 | 3.45 | 3.24 | **3.79** | 12.0 | 12.0 | 11.66 | 11.56 | 11.41 | **11.72** | 0.45 | 0.76 | 0.80 | 0.85 | 0.93 | **0.75** | 9.86 | 9.74 | 9.67 | 9.53 | 9.45 | **9.65** |
| Set F          | 4.41 | 4.12 | 3.84 | 3.40 | 3.21 | **3.79** | 12.0 | 12.0 | 11.64 | 11.21 | 11.12 | **11.59** | 0.45 | 0.54 | 0.83 | 0.89 | 0.96 | **0.73** | 9.88 | 9.76 | 9.64 | 9.50 | 9.42 | **9.64** |
| Mean           | 4.40 | 4.24 | 3.91 | 3.72 | 3.10 | 12 | 12 | **11.69** | 11.50 | 11.35 | **11.35** | 0.45 | 0.61 | 0.72 | 0.79 | 0.93 | **0.93** | 9.86 | 9.77 | 9.68 | 9.57 | 9.47 |

CD_{0.05} Treatments (T) 0.30
Storage interval (S) 0.31
T x S 0.59

Table 3: Nutritional chart of apricot RTS

| Nutritional facts per 100 ml | SET A | SET B | SET C | SET D | SET E | SET F |
|-----------------------------|-------|-------|-------|-------|-------|-------|
| *Antioxidant activity (%)   | 4.10  | 4.11  | 4.10  | 4.10  | 4.11  | 4.12  |
| *Protein (g/100 ml)         | 0.10  | 0.10  | 0.11  | 0.10  | 0.10  | 0.11  |
| *Fiber (%)                  | 0.30  | 0.31  | 0.31  | 0.30  | 0.29  | 0.30  |
| *Carbohydrate (g/100g)      | 11    | 10.9  | 11    | 10.9  | 10.8  | 10.9  |
| *Total sugars (%)           | 3.03  | 3.04  | 3.03  | 3.04  | 3.03  | 3.03  |

*values evaluated in lab
### Table 4 Physicochemical characteristics of apricot RTS

| Treatments (T) | pH     | TSS (°B) | Titrable acidity (%) | Ascorbic acid (mg/100g) |
|---------------|--------|----------|-----------------------|--------------------------|
|               | 0      | 7        | 14       | 21      | 30      | Mean | 0    | 7    | 14    | 21    | 30      | Mean | 0 | 7    | 14    | 21    | 30      | Mean |
| Set A         | 4.62   | 4.50     | 3.32     | 3.06    | 2.42    | **3.58** | 10.00| 9.80 | 9.65  | 9.34  | 9.19    | **9.59** | 0.40| 0.48| 0.59  | 0.67  | 0.79    | **0.58** | 9.50| 9.42 | 9.53  | 9.50  | 9.37    | **9.46** |
| Set B         | 4.64   | 4.42     | 4.33     | 4.18    | 3.07    | **4.12** | 10.00| 10.00| 9.72  | 9.57  | 9.49    | **9.69** | 0.40| 0.45| 0.51  | 0.62  | 0.71    | **0.53** | 9.51| 9.48 | 9.62  | 9.57  | 9.55    | **9.54** |
| Set C         | 4.64   | 4.39     | 4.24     | 4.13    | 3.08    | **4.09** | 10.00| 10.00| 9.81  | 9.71  | 9.61    | **9.78** | 0.40| 0.48| 0.54  | 0.59  | 0.66    | **0.53** | 9.51| 9.45 | 9.38  | 9.35  | 9.31    | **9.37** |
| Set D         | 4.64   | 4.21     | 4.17     | 4.11    | 3.15    | **4.05** | 10.00| 10.00| 9.73  | 9.53  | 9.31    | **9.64** | 0.40| 0.47| 0.51  | 0.58  | 0.69    | **0.53** | 9.49| 9.41 | 9.36  | 9.29  | 9.21    | **9.31** |
| Set E         | 4.63   | 4.20     | 3.92     | 3.62    | 3.42    | **3.95** | 10.00| 10.00| 9.70  | 9.50  | 9.39    | **9.64** | 0.40| 0.49| 0.55  | 0.63  | 0.74    | **0.56** | 9.52| 9.44 | 9.37  | 9.33  | 9.29    | **9.35** |
| Set F         | 4.64   | 4.30     | 3.15     | 3.54    | 3.38    | **4.01** | 10.00| 10.00| 9.82  | 9.43  | 9.23    | **9.56** | 0.40| 0.48| 0.53  | 0.65  | 0.76    | **0.56** | 9.54| 9.43 | 9.32  | 9.24  | 9.18    | **9.34** |
| Mean          | 4.64   | 4.30     | 3.15     | 3.54    | 3.38    | **3.38** | 10.00| 10.00| 9.82  | 9.43  | 9.23    | **9.56** | 0.40| 0.47| 0.53  | 0.62  | 0.72    | **0.56** | 9.51| 9.43 | 9.42  | 9.37  | 9.30    | **9.34** |

**CD₀.₀₅**

- Treatments (T) 0.224
- Storage interval (S) 0.245
- T x S 0.549

- Treatments (T) 0.221
- Storage interval (S) NS
- T x S NS

- Treatments (T) 0.218
- Storage interval (S) NS
- T x S NS

- Treatments (T) NS
- Storage interval (S) NS
- T x S NS
**Fig. 1 (a & b)** Biopreservation of apricot pulp by purified bacteriocin produced by *Lactobacillus brevis* SM6

**Fig. 2** Sensorial evaluation of apricot pulp
Fig. 3 Comparison of crude and purified bacteriocin of *Lactobacillus brevis* SM6 with chemical preservative against *Listeria monocytogenes* to enhance shelf life of apricot pulp

![Graph showing log CFU/ml (L. brevis SM6) and log CFU/ml (L. monocytogenes) over days for different sets.]

Fig. 4 Comparison of *Lactobacillus brevis* SM6 against *Listeria monocytogenes* to enhance shelf life of apricot pulp

![Graph showing log CFU/ml (L. monocytogenes) for different sets.]

Fig. 5 (a&b) Biopreservation of apricot RTS by purified bacteriocin produced by *Lactobacillus brevisSM6*

![Image of bottles labeled Set-A to Set-F.]

a
**Fig. 6** Sensorial evaluation of apricot RTS

**Fig. 7** Comparison of crude and purified bacteriocin of *Lactobacillus brevis* SM6 with chemical preservative against *Listeria monocytogenes* to enhance shelf life of apricot RTS
In conclusion, the bacteriocin Lactobacillus brevis SM6 has been found quite effective as compared to the chemical preservative and serve as a good candidate to extend the storage life of apricot RTS and apricot plum by preventing the growth of harmful spoilage causing pathogens.

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