Bacteriology

Isolation of Atypical Genotype Actinobacillus pleuropneumoniae Serotype 6 in Japan

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ABSTRACT. We describe here isolation of genetically atypical serotype 6 Actinobacillus pleuropneumoniae in Japan indistinguishable by the multiplex PCR that can discriminate between immunologically cross-reactive serotypes 3, 6 and 8. Nucleotide sequence analysis of capsular export and biosynthesis genes revealed that the atypical isolates have capsular polysaccharide export and synthesis gene sequences that are distinct from those of the serotype 6 reference strain. The atypical strains contain a sequence that is identical with both serotype 3- and 6-specific primers, which causes cross-reactions in multiplex PCR.

KEY WORDS: Actinobacillus pleuropneumoniae, atypical serotype 6, capsular polysaccharide export and biosynthesis genes, multiplex PCR, nucleotide sequence.

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Actinobacillus pleuropneumoniae is the etiological agent of porcine pleuropneumonia, which causes serious economic losses in the pig-rearing industry [4]. To date, 15 serotypes have been recognized mainly based on the antigenic diversity of capsular polysaccharides in the organisms [2, 18]. The prevalent serotypes differ by country, region and farm [3]. In Japan, serotype 2 is the most common; serotypes 1 or 5 follow [3, 7, 15, 16, 22]. Serotypes 3, 6, 7, 8, 9, 11, 12 and 15 are isolated in only sporadic cases [3, 7]. Serotyping is widely performed in veterinary diagnostic laboratories, since virulence differs in serotypes and vaccines for A. pleuropneumoniae are serotype specific [4, 17, 20]. However, only a few veterinary diagnostic laboratories can prepare a full set of serotype-specific antisera for serotyping. Furthermore, cross-reactions are often observed among different serotypes in a number of conventional immunological tests, for example, between serotypes 1, 9 and 11, serotypes 4 and 7 and between serotypes 3, 6 and 8, which prevent accurate and rapid typing of field strains [11–14].

To overcome such problems, genotypic methods, such as PCR amplification of serotype-specific capsular biosynthesis or export genes, have been developed to enable precise serotyping [1, 7, 8, 10, 19, 23, 24]. Among them, a specific multiplex PCR that can discriminate among the problematic cross-reactive serotype 3, 6 and 8 strains in a single tube has been developed [24]. Recently, we isolated two serotype 6 strains from different prefectures with an immunodiffusion test. Then, we tried to genetically confirm them as serotype 6 by the single-tube multiplex PCR [24]. However, the Japanese isolates could not be typed with the multiplex PCR due to cross-reaction. In this study, we report serological and genetic characterization of the atypical Japanese A. pleuropneumoniae serotype 6 isolates.

An A. pleuropneumoniae designated as strain QAS59 was isolated from an abscess of a pig’s backbone in Fukui, Japan, in 2009, and another strain designated as HYT2 was isolated from an abscess of a pig’s lung at an abattoir in Tochigi, Japan in 2010. They were grown at 37°C in 5% CO2 on chocolate agar or tryptic soy agar supplemented with 5% horse blood and 50µg/mL nicotinamide dinucleotide as previously described [7]. Serotyping was carried out by slide agglutination and immunodiffusion tests using rabbit hyperimmune sera against reference strains of the 15 serotypes of A. pleuropneumoniae [13, 14]. The results are shown in Table 1. The two Japanese isolates were serotyped as serotype 6 by the immunodiffusion test, although these strains showed cross-reactivity against more than two antisera against serotype reference strains by the slide agglutination test.

Genotyping was then carried out by a multiplex PCR that can distinguish between serotypes 3, 6 and 8 based on the cps and/or cpx gene [24], a PCR typing system based on the A. pleuropneumoniae toxin (Apx toxin) and outer membrane lipoprotein genes (apx and omlA, respectively) [5]. In addition, a monoplex PCR with either serotype 3-, 6- or 8-specific primer sets only was also performed. Results for the genotyping are shown in Table 1 and Fig. 1. In the multiplex PCR, both Japanese strains had two amplicons (Table 1, Fig. 1A and 1C). Therefore, we also tried a monoplex PCR with only serotype 3-, 6- or 8-specific primers. In these experiments, each Japanese strain had an amplicon in each capsular serotype 3 and 6 monoplex PCR (Table 1, Fig. 1B and 1C). Apx-toxin gene profiling resulted in the same apx gene combination as the serotype 4, 6, 8 and 15
reference strains in strain HYT2, while the same apx gene combination was seen in strain QAS59 as in the serotype 3 reference strain (Table 1) [5]. The omlA gene typing scheme for both Japanese strains showed an omlAIII pattern, which was the same as that found for the serotype 3, 6 and 7 reference strains (Table 1) [5].

The ultimate goal in this study was to clarify why two amplicons were observed in the Japanese serotype 6 isolates with the multiplex PCR. We speculated that the Japanese serotype 6 isolates might have possessed identical or highly similar omlA and apx genes.

### Table 1. Serotyping and genotyping of atypical Japanese A. pleuropneumoniae isolates

| Tests                          | Strains                        |
|-------------------------------|--------------------------------|
| Serotyping tests              | Serotype 3 reference strain    | Serotype 6 reference strain | Strain QAS59 | Strain HYT2 |
| Slide agglutination test      | NT a)                         | NT                          | 3, 6, 8, 15  | 6, 10, 11, 15 |
| Immunodiffusion test          | 3 b)                          | 6                           | 6            | 6            |
| Genotyping tests              |                                |                              |              |
| Multiplex PCR                 | Serotypes 3, 6 and 8           | 3                           | 6            | 3, 6         |
| Monoplex PCR                  | Serotype 3                     | +                           | +            | +            |
|                               | Serotype 6                     | –                           | +            | +            |
|                               | Serotype 8                     | –                           | –            | –            |
| Apx toxin PCR profiling       | apxICA                        | –                           | –            | –            |
|                               | apxIICA                       | +                           | +            | +            |
|                               | apxIIICA                      | +                           | +            | +            |
|                               | apxIBD                        | –                           | +            | –            |
|                               | apxIIIBD                      | +                           | +            | +            |
| omlA typing                   | omlAIII                       | omlAIII                     | omlAIII      | omlAIII      |

a) Not tested. b) The numbers indicate the positive serotypes.
similar sequences with serotype 3-specific primers (APN3F, 5'-TTT GCG CTG TAG TGC TCC AAC-3', and APN3R, 5'-AAT AAA TAA AGT TGC TCG AAA GTA) in the multiplex PCR [24], resulting in cross-reaction in the multiplex PCR. Therefore, the nucleotide sequences of the cpxD-cpsA region, in which the serotype 6-specific PCR primers for the multiplex PCR [24] are located, of the Japanese isolates were determined. The DNA regions were PCR amplified, purified and sequenced as previously described [6]. The primers used for amplification of the DNA region (5'-CAC ACC ATA AAC CTT TGC TAG ATC-3' and 5'-GTC ATC GCC TAA TGA TCT TGC TCT TTC-3') were designed in this study based on the cpx6D and cps6A sequence (accession number (no.) AY534316) [9]. The nucleotide sequences (2,410 base pairs (bp)) of the cpxD-cpsA region of QAS59 and HYT2 were identical and deposited into the DDBJ/GenBank/EMBL databases under accession no. AB809625. As we expected, QAS59 and HYT2 had sequences identical to serotype 3-specific primers APN3F and APN3R as well as serotype 6-specific primers, resulting in amplification of two amplicons from both serotypes 3- and 6-specific primer sets.

The nucleotide sequences of the entire cpsA and partial cpxD genes of the Japanese isolates were then compared with nucleotide sequences in databases. Consequently, the entire nucleotide sequences of the cps6A gene (1,137 bp) of QAS59 and HYT2 showed highest identity with that of A. pleuropneumoniae serotype 7 (97.3%, accession no. CP001091) followed by those of serotypes 8 (96.9%, accession no. AY356552), 6 (92.9%, accession no. AY354316) and 3 and 2 (91.3%, accession no. CP000687 and AY377726, respectively). A high degree of homology between cpsA genes, which is involved in capsular biosynthesis of A. pleuropneumoniae serotypes 2, 3, 6, 7 and 8, has already been reported [9], indicating a similar function for CpsA encoded by cpsA genes in the five serotypes [9]. It has also been suggested that genes other than cpsA genes determine the capsular antigenic differences among these serotypes [9, 21]. For example, cps6E [9, 21] and cpx6F [21] were present only in serotype 6. It is of interest that the cpx6A genes of strains QAS59 and HYT2 did not show the highest identity with those of the serotype 6 reference strains despite the fact that we expected that the CPS synthesis genes were conserved in the serotype. Furthermore, the partial nucleotide sequence of the cpxD gene (993 bp), which is not involved in capsular antigenicity but is involved in capsular export, of the Japanese isolates revealed the highest similarity (97.2%, accession no. AY534316) with that of the serotype 6 reference strain, followed by serotype 2 (95.5%, accession no. AY377726); serotype 7, 3 and 1 (94.9%, accession no. CP001091, CP000687 and AF518558, respectively); serotype 12 (91.5%, accession no. AY496881); serotype 4 (91.1%, accession no. GU585380); and serotype 5 (89.4%, accession no. CP000569), indicating that the capsular export gene, cpxD, is conserved more than the capsular synthesis gene, cpsA, in serotype 6 strains.

In conclusion, two Japanese serotype 6 A. pleuropneumoniae isolates are genetically distinct from those isolated from countries other than Japan [24] due to unique cpxD and cpsA genes, resulting in the inability to use multiplex PCR for serotypes 3, 6 and 8 A. pleuropneumoniae [24]. Development of an alternative typing method is needed for genetically atypical serotype 6 A. pleuropneumoniae isolates.

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