Saw-scaled viper envenoming complicated with acute myocardial infarction

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Abstract
The saw-scaled viper (Echis carinatus) is considered to be a venomous snake which is especially seen in Northern Sri Lanka. Systemic manifestations are rare and reported complications include coagulopathy and renal impairment. The cardiac toxicity following snakebites is rare and cardiac involvement following the saw-scaled viper bites is extremely rare. Here, we describe a patient with acute myocardial infarction following systemic envenoming by saw-scaled viper in Northern Sri Lanka, which was successfully managed per ward protocol following national guidelines.

Keywords
Saw-scaled viper, myocardial infarction, envenoming

Introduction
Snakebites are commonly encountered in clinical practice, especially in the dry zone of Sri Lanka, that is, the Northern part of the country. The saw-scaled viper (SSV) (Echis carinatus) is considered to be a venomous snake which is especially seen in Northern Sri Lanka.1,2 The clinical profile of the SSV is not well established due to its restricted distribution in Sri Lanka.1,3 The local envenoming and hematological manifestations are common among the SSV bites.1,2 Systemic manifestations are rare and reported complications include coagulopathy and renal impairment.4,5 The cardiac toxicity following snakebites is rarely reported, especially with the SSV bites in literature. The exact pathophysiology has not been established in literature. The biological plausibility for this association is due to the thrombotic and vasoconstricting properties of snake venom. Here, we describe a middle-aged man with acute myocardial infarction following systemic envenoming by SSV in Northern Sri Lanka.

Case history
A 50-year-old previously healthy farmer was admitted to emergency unit with a history of localised pain and swelling of left foot following the SSV bite after 1 h of the incident. The dead snake was brought by the patient and identified by medical professional and subsequently was confirmed by herpetologist as SSV (E. carinatus) (shown in Figure 1). The patient had no past history or risk factors of ischaemic heart disease and had no significant past surgical history. On examination, the patient was conscious and alert. Patient had localised swelling and fang mark at the bite site without any localised bleeding manifestations. Patient’s pulse rate was 80 beats/min and his blood pressure was 120/80 mmHg. Patient’s respiratory rate was 12/min with oxygen saturation of 98% on room air. Patient had no signs of systemic envenoming. The 20-min whole blood clotting test (WBCT) showed incoagulable blood. Patient’s full blood count showed white cell count of 8520/mm3, haemoglobin of 14 g/dL and platelets of 233,000/mm3. Liver enzymes were mildly elevated (aspartate transaminase (AST): 86 IU/L, alanine transaminase (ALT): 77 IU/L). Serum electrolytes revealed sodium of 140 mmol/L and potassium of 3.8 mmol/L. Blood urea was 44 mg/dL and serum creatinine was 1.06 mg/dL. Clotting profile, including PT/INR (prothrombin time/international normalized ratio) and APTT (activated partial thromboplastin time), was very high. His
Table 1. The biochemical profile of the patient is shown with clinical progression of disease.

| Biochemical investigations          | Admission | Day 1   | Day 2     | Day 3     | Day 4     | Day 5     | Day 6     |
|-------------------------------------|-----------|---------|-----------|-----------|-----------|-----------|-----------|
| **Full blood count**                |           |         |           |           |           |           |           |
| White cell count (4000–11,000/mm³) | 6450      | 8520    | 10,800    | 10,100    | 7470      | 6820      | 6840      |
| Neutrophils (50%–70%)              | 46        | 39      | 93        | 65        | 52        | 56        | 56        |
| Lymphocytes (20%–40%)              | 43        | 52      | 4         | 30        | 33        | 29        | 36        |
| Haemoglobin (12–16 g/dL)           | 13        | 14      | 13.6      | 13.1      | 12.8      | 12.9      | 13.1      |
| MCV (80–100 fl)                    | 87        | 92      | 93        | 91        | 93        | 94        | 92        |
| Red cell count (400,000–550,000 mm³)| 448,000   | 440,000 | 410,000   | 398,000   | 398,000   | 397,000   | 410,000   |
| Platelets (150,000–450,000 mm³)    | 235,000   | 233,000 | 201,000   | 163,000   | 182,000   | 181,000   | 210,000   |
| **Renal functions tests**           |           |         |           |           |           |           |           |
| Blood urea (18–55 mg/dL)           | 43        | 44      | 40        | 38        | 22        | 28        | 32        |
| Serum creatinine (0.7–1.5 mg/dL)   | 1.03      | 1.06    | 0.99      | 1.1       | 1.06      | 1.1       | 1.1       |
| **Serum electrolytes**              |           |         |           |           |           |           |           |
| Serum sodium (135–145 mmol/L)      | 136       | 140     | 138       | 138       | 138       | 134       | 135       |
| Serum potassium (3.5–5.0 mmol/L)   | 3.9       | 3.8     | 3.7       | 4.1       | 3.6       | 3.8       | 3.9       |
| **Liver profile**                   |           |         |           |           |           |           |           |
| Serum AST (0–45 U/L)               | 46        | 86      | 78        | 44        | 42        | 40        | 40        |
| Serum ALT (0–35 U/L)               | 57        | 77      | 70        | 38        | 38        | 32        | 38        |
| Total bilirubin (0–2.0 mg/dL)      | 1.2       | 1.0     | 1.1       | 1.0       | 1.1       | 1.0       | 1.0       |
| Indirect bilirubin (0–1.6 mg/dL)   | 0.8       | 0.8     | 0.8       | 0.7       | 0.9       | 0.8       | 0.8       |
| **Clotting profile**                |           |         |           |           |           |           |           |
| PT/INR (<1.4)                      | Very high | Very high | Very high | 1.2     | 1.0     | 1.1     | 1.2     |
| APTT (<35)                         | Very high | 32       | 34       | 32       | 35       |           |           |
| **Urine full report**              |           |         |           |           |           |           |           |
| Protein (+)                         | –         | –       | –         | –         | –         | –         | –         |
| Pus cells (/HPF)                    | –         | 1.2     | –         | –         | –         | –         | 3-5       |
| Red cells (/HPF)                    | –         | 10-15   | –         | –         | –         | –         | 5-10      |
| Active sediment (+)                 | –         | Nil     | –         | Nil       | –         | –         | Nil       |
| High-sensitive Troponin I (<19 ng/L)| –         | 3051    | 1572      | 1025      | 572       | 572       | 109.2     |

MCV: mean corpuscular volume; AST: aspartate transaminase; ALT: alanine transaminase; PT/INR: prothrombin time/international normalized ratio; APTT: activated partial thromboplastin time.
Figure 2. The 12-lead electrocardiography (ECG) showed ST elevation in leads I, aVL and ST depression in II, aVF, V₄ to V₆ pre-thrombolytic therapy and resolution of ECG changes following thrombolytic therapy in patient with saw-scaled viper (SSV) (*Echis carinatus*) envenomation.

Figure 3. The coronary angiography showed normal (a) right and (b) left coronary arteries in the patient with saw-scaled viper (SSV) (*Echis carinatus*) envenomation.
Discussion

The SSV is considered to be one of the venomous snakes in Sri Lanka which is distributed in certain parts of Northern Sri Lanka and a few areas of Southern Sri Lanka.1-3 The clinical profile of the SSV is not well established due to its restricted distribution in Sri Lanka.1,2 The first case of the SSV was described by Patrick Russell in 1796 in India in 1801.6 The SSV bites are rarely reported in Sri Lanka and SSV is limited to certain parts of dry zones of Sri Lanka, especially Northern Sri Lanka.1,2

The local envenoming and haematological manifestations are common among the SSV bites which are reported in Sri Lanka.1,2 The pain, swelling and necrosis can occur due to phospholipase A2 component of the SSV venom. Spontaneous bleeding can occur due to the activation of prothrombin by metalloproteinase seen in the SSV venom.7 Consumptive coagulopathy occurs due to the activation of clotting system by procoagulant enzymes in the SSV venom which can cause hypofibrinogenemia and disseminated intravascular coagulation, resulting in multiorgan dysfunction and death.5,8 The SSV venom contains many other toxic compounds which activate to cause derangement in haemostasis such as platelet aggregation inhibitors, carinatin, echistatin, and echicetin, protein C activator, fibrinogenolysis, calcium-dependent carinactivase and disintegrins.9 However, the venom profiles differ from other geographically distinct venoms of E. carinatus due to change in the relative composition of the toxin families.10 Snake venom metalloproteinase, snaclecs and phospholipase A2 are major venom components in all the venoms.

The cardiac toxicity following snakebites is rarely reported, especially with the SSV bites in literature. Recently, some case reports of myocardial infarction have been reported following snakebites.11 However, there have been a few published case reports of cardiac toxicity following Russell’s viper12 and a hump-nosed viper bite.13,14 The exact pathophysiology has not been established in literature. The biological plausibility for this association is due to the thrombotic and vasoconstricting properties of snake venom. Various pathophysiological mechanisms behind myocardial infarction following snakebite have been proposed which includes hypovolemic and anaphylactic shock,15 coronary thrombosis,16 direct cardiac toxic effect,17 coronary vasoconstriction, myocarditis with extensive myocardial necrosis and myocardial haemorrhage and microvascular thrombin deposition.18

Our patient developed local envenoming and haematological manifestations at the time of admission. Subsequently, patient developed acute myocardial infarction after 12 h of the SSV bite. ECG is recommended as the earliest diagnostic tool to detect myocardial infarction which may not show characteristic pattern in the hyper acute phase. Therefore, further diagnostic evaluation is necessary to diagnose myocardial infarction in snakebite patients.19 Our patient is confirmed by electrocardiographic changes, positive troponin I and echocardiographic findings. Further coronary angiography findings were normal. Patient's fasting blood sugar and lipid profile were normal and he had no risk factors such as obesity, smoking, alcoholism, sedentary life style and family history of heart disease as well. Patient had no previous psychiatric disorders and did not have any anxiety following snake bite on psychiatric assessment. Furthermore, patient had not been given any premedications to predispose to myocardial infarction except anti-snake venom along with hydrocortisone and chlorpseudamine. Therefore, we conclude that acute myocardial infarction was purely due to SSV systemic envenomation. This was further evidenced by the absence of any cardiovascular risk factors or premedications or stress related to snake bite in our patient.

Immunotherapy is the only specific therapy for snake-bite envenoming.1,2 The outcome depends on the ability of immunoglobulins to bind, extract and eliminate toxins present in the body. The treatment of myocardial infarction following snakebite is major clinical challenge because snake venom causes thrombocytopenia, coagulopathy and risk of bleeding.20 These may limit the use of thrombolytic therapy. The thrombolytic therapy along with anti-anginal drugs are recommended for myocardial infarction following snakebite according to types of myocardial infarction in previous case reports in literature.11,12,19 However, our patient had already received two cycles of ASV and had normal basal tests, and was at low risk for bleeding. The prompt ASV therapy and monitoring for coagulation functions are essential for proper management of myocardial infarction following snakebite. The bacterial infections can occur at bite sites especially if bite site was incised or tampered by traditional treatments which may require local and systemic antibiotics.21 Our patient was managed with oral flucloxacillin for 5 days.

Conclusion

This case illustrates a patient with acute myocardial infarction which is a rare manifestation of the SSV envenoming following the SSV bite which was successfully managed per ward protocol following national guidelines.

Declaration of conflicting interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Ethical approval

Our institution does not require ethical approval for reporting individual cases or case series.

Funding

The author(s) received no financial support for the research, authorship, and/or publication of this article.
Informed consent

Written informed consent was obtained from the patient for their anonymised information to be published in this article.

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