Review Article
The Molecular Pathogenesis and Clinical Implications of Hepatocellular Carcinoma

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1. Introduction
Hepatocellular carcinoma (HCC) is the sixth most common malignant disease worldwide and the third greatest cause of cancer-related death [1]. The etiology of HCC has been reported to be related to a variety of diseases such as viral hepatitis [2, 3], alcoholic hepatitis [4], nonalcoholic fatty liver disease (NAFLD) [5, 6], and metabolic syndrome including diabetes mellitus [7, 8]. The sequences from chronic hepatitis and liver cirrhosis cause de novo HCC [9] (Figure 1). HCC is considered to have increased invasiveness with malignant transformation and metastatic potential [10, 11]. Therefore, it is difficult to select the proper management of the disease.

The clinical therapy for HCC includes various modalities such as liver resection [12], liver transplantation [13], transarterial chemoembolization (TACE) [14], percutaneous ethanol injection therapy (PEIT) [15], radiofrequency ablation (RFA) [16], and chemotherapy including molecular-targeted therapy [17]. However, the high recurrence rate is a major concern after any treatment [18]. The reason for the high recurrence rate of HCC could be proliferation of epithelial cells and increased arterial vascularity [19, 20]. Furthermore, HCC cells themselves express various growth factors such as vascular endothelial growth factor (VEGF) [19], platelet-derived growth factor (PDGF) [20], epidermal growth factor (EGF) [21], fibroblast growth factor (FGF) [22], and insulin-like growth factor (IGF) [23], which induce cell proliferation in an autocrine fashion [24]. The receptors of these growth factors activate intracellular signals such as the RAF/MEK/ERK pathway [25] and the PI3K/AKT/mTOR pathway [26], which induce proliferation of both cancer and endothelial cells (Figure 2). These growth factors, including their intracellular molecules, are considered to be a specific target for HCC treatment.

In clinical trials, sorafenib, which is an inhibitor of the VEGF receptor (VEGFR) and PDGF receptor (PDGFR), has been proven to have a survival benefit for nonresectable HCC compared a placebo in the best supportive care (BSC) setting [27, 28]. Phase III trials are ongoing to determine the survival benefit in patients who receive surgery or ablation [29]. On the other hand, the survival benefit of sorafenib has been limited to a few of months and other pathways need to be blocked to achieve longer survival. Side effects of...
The etiology of HCC has been reported to be related to a variety of diseases such as viral hepatitis, alcoholic hepatitis, nonalcoholic fatty liver disease (NAFLD), and metabolic syndrome including diabetes mellitus. Regeneration of damaged hepatocytes reveals the activation of stem cells. The abilities of self-renewal and infinite proliferation are closely related to the development of hepatocellular carcinoma (HCC). Stem cells in the liver are divided into several types, including oval cells, small hepatocytes, and progenitor cells. HCC cells and liver cancer stem cells could derive from mutation of these stem cells. The origin of the stem cells could be from either mature hepatocytes or bone marrow cells.

Sorafenib therapy are obstacles to continuation of the therapy because normal cells also express VEGFR and PDGFR, and sorafenib severely damages both normal and cancer cells. The development of anticancer drugs must focus on a specific target that is restricted to the cancer cells.

Cancer stem cells have been shown to be a target for cancer-specific therapy recently. The abilities of self-renewal and infinite proliferation are closely related to the nature of HCC development [30, 31]. Stem cells in the liver are divided into several cell types, including oval cells [32], small hepatocytes [33], and progenitor cells [34] (Figure 1). Liver cancer stem cells and HCC cells could derive from mutation of these stem cells. The origin of the stem cells could be either from mature hepatocytes or from bone marrow cells [35] (Figure 1). Thus, specific stem cell-based therapy could be another strategy to overcome the high recurrence rate of HCC [36]. We describe the molecular pathogenesis, molecular therapy, and stem cell-targeted therapy for HCC treatment in this review.

2. Role of Growth Factors and Angiogenesis in HCC

A specific pathological feature of HCC is high vascularity of the tumor. It is necessary to increase vascularity for cancer cell proliferation. VEGF, PDGF, EGF, FGF, and IGF, growth factors that facilitate high vascularity and cancer cell proliferation, are expressed not only in cancer cells but also in other surrounding cells. The high expression of the growth factors is also associated with tumor invasion and portal thrombosis [19, 20, 22].

Among the growth factors, high expression of EGF is related to differentiation and invasion by the cancer cells [21]. On the other hand, PDGF is related to metastatic behavior of HCC cells [20]. The proliferation of endothelial cells is important for metastasis and invasion by cancer cells. Therefore, growth factors play an important role in proliferation of cancer cells not only in an autocrine fashion but also in a paracrine fashion through surrounding cells [24]. In addition, antivascular factors decrease in the serum and tissue of HCC patients [37]. These reports indicated that specific growth factors can be targets for HCC treatment.

Based on the high vascularity of HCC, endothelial cells could be a target for HCC treatment. This approach could be promising because endothelial cells have normal cell physiology with stable genetic regulation, which can be easily manipulated by molecular target therapy.

3. Role of RAF/MEK/ERK Signaling Pathway in Developing HCC

Tyrosine kinase type receptors, such as VEGFR, PDGFR, EGFR, FGFR, and IGFR, activate intracellular RAS in the RAF/MEK/ERK pathway [19–23]. Subsequently, AP-1 family members such as c-JUN and c-FOS activate expression of various genes that induce cell proliferation and vasculogenesis [38] (Figure 2). The activation of the RAF/MEK/ERK pathway is related to the disease progression of HCC [39] and
Intracellular signaling pathways

Cell surface tyrosine kinase receptors

- RAF/MEK/ERK pathway
- PI3K/ALT/mTOR pathway

RAS $\rightarrow$ RAF $\rightarrow$ MEK $\rightarrow$ ERK

PI3K $\rightarrow$ AKT $\rightarrow$ mTOR

PTEN

BAD $\rightarrow$ Apoptosis

Cancer cell proliferation

Angiogenesis of endothelial cells

**Figure 2:** The RAF/MEK/ERK and the PI3K/AKT/mTOR signaling pathways are shown. Proangiogenic and proliferative growth factors activate the RAF/MEK/ERK pathway. The small GTPase RAS and the serine/threonine kinase RAF are the key molecular signal regulators. Intermediate signaling is regulated by MEK, which is responsible for phosphorylating and activating the final downstream signaling ERK molecules. ERK regulates cellular activity, indirect inducers of gene expression, and transcription factors in the AP-1 family such as c-JUN and c-FOS and cell cycle-related kinases. Binding of these growth factors to their receptors also activates PI3K, which subsequently produces the lipid second messenger, and in turn activates serine/threonine kinase AKT. Activated AKT also phosphorylates several cytoplasmic proteins, most notably mTOR. The activation of mTOR increases cellular proliferation, and inactivation of BAD decreases apoptosis and increases cell survival. This pathway is negatively regulated by the phosphatase and tensin homolog deleted on chromosome 10 (PTEN), which targets the lipid products of PI3K for dephosphorylation.

HBV-related HCC development [40]. Furthermore, HCV core protein activates RAF and is considered to play a role in the development of HCC [41].

RAS and RAF play important roles in which intracellular signals activate expression of various genes [42] (Figure 2). RAS activates RAF, which induces activation of MEK [43]. MEK activates ERK and its phosphorylation [43]. ERK regulates more than one hundred intracellular substrates directly and gene expression indirectly as cell kinase to activate transcription factors and cell cycle regulators [44, 45]. Activation of ERK is closely related to cancer cell proliferation and, thus, inhibition of ERK could have an anticancer effect [46].

### 4. Role of PI3K/AKT/mTOR Signaling Pathway in Developing HCC

The phosphatidylinositol-3 kinase (PI3K) pathway plays an important role in the proliferation and survival of cancer cells in various solid tumors, including HCC [26] (Figure 2). PI3K activates AKT, which is a lipid second messenger [42]. Subsequently, AKT phosphorylates various intracellular proteins, including mTOR [42]. The activation of mTOR induces cell proliferation and inactivates BAD [47]. Inactivation of BAD is important for cancer cells to survive by regulating apoptosis [47]. Inactivation of AKT has been shown to improve the antitumor effect of sorafenib in an animal model and thus it could have potential use for HCC treatment [48]. The PI3K pathway is regulated by phosphatase and tensin homolog deleted on chromosome 10 (PTEN) negatively and the expression of PTEN is suppressed in half of HCC cells clinically [49] (Figure 2). In fact, PTEN expression is suppressed by HBx protein in HBV hepatitis patients [23], and downregulation of PTEN is associated with tumor grade progression, tumor stage, and poor overall prognosis [49].

### 5. Molecular-Targeting Clinical Therapies and Trials for HCC

Sorafenib is an inhibitor of RAF that is activated by VEGF and PDGF [50–52]. It has been tested in phase III clinical
| Agent                          | Target                  | Year | Phase | Patients | CR (%) | PR (%) | SD (%) | RR (%) | PFS (months) | TTP (months) | OS (months) | Authors            | Reference number |
|-------------------------------|-------------------------|------|-------|----------|--------|--------|--------|--------|--------------|--------------|-------------|--------------------|------------------|
| Sorafenib SHARP study versus placebo | RAF, VEGFR, PDGFR       | 2008 | III   | 299      | 0      | 2.3    | 71.0   | 2.3    | 4.1          | 5.5          | 10.7        | Llovet et al.      | [27]             |
|                               |                         | 2008 | III   | 303      | 0      | 0.7    | 67.0   | 0.7    | 4.9          | 2.8          | 7.9         |                    |                  |
| Sorafenib Asia-Pacific study versus placebo |                         | 2009 | III   | 150      | 0      | 3.3    | 54.0   | 3.3    | 3.5          | 2.8          | 6.5         | Cheng et al.       | [28]             |
|                               |                         | 2009 | III   | 76       | 0      | 1.3    | 27.6   | 1.3    | 3.4          | 1.4          | 4.2         |                    |                  |
| Sunitinib                     | VEGFR, PDGFR            | 2009 | II    | 37       | 0      | 2.7    | 35.1   | 2.7    | 3.7          | 5.3          | 8.0         | Faivre et al.      | [53]             |
|                               |                         | 2009 | II    | 34       | 0      | 2.9    | 50.0   | 2.9    | 3.9          | 4.1          | 9.8         | Zhu et al.         | [54]             |
| Brivanib                      | VEGFR, FGFR             | 2009 | II    | 55       | NR     | NR     | NR     | 5.0    | NR           | 2.8          | 10.0        | Raoul et al.       | [55]             |
| Erlotinib                     | EGF                     | 2005 | II    | 38       | 0      | 9.0    | 50.0   | 9.0    | NR           | 3.2          | 13.0        | Philip et al.      | [56]             |
| Erlotinib + bevacizumab       |                         | 2009 | II    | 40       | 0      | 25.0   | 38.0   | 25.0   | 9.0          | NR           | 15.7        | Thomas et al.      | [60]             |
| TSU-68                        | VEGFR, PDGFR, FGFR      | 2008 | I/II  | 35       | 2.9    | 5.7    | 42.9   | 8.6    | NR           | NR           | NR          | Kanae et al.       | [57]             |
| Bevacizumab                   | VEGF                    | 2009 | II    | 46       | 2      | 11.0   | 13.0   | 6.9    | NR           | 12.4         | 9.6         | Siegel et al.      | [58]             |
| Cetuximab                     | EGF                     | 2007 | II    | 30       | 0      | 0.0    | NR     | 0.0    | 1.4          | NR           | 9.6         | Zhu et al.         | [59]             |

CR: complete response; PR: partial response; SD: stable disease; RR: response rate; PFS: progression-free survival; TTP: time to progression; OS: overall survival; NR: not reported.
Table 2: Ongoing clinical trials using molecular-targeted agents for hepatocellular carcinoma.

| Acronym | Phase | Active arm | Control arm | Design of the clinical trials |
|---------|-------|------------|-------------|-----------------------------|
| STORM   | III   | Sorafenib  | Placebo     | Adjuvant therapy after resection or ablation |
| SILLIUS | III   | Sorafenib + TACE | Sorafenib | Combination therapy with hepatic arterial infusion chemotherapy (TACI) |
| SPACE   | II    | Sorafenib + TACE | Placebo + TACE | Combination therapy with transarterial chemoembolization (TACE) |
| TACTICS | II    | Sorafenib + TACE | Placebo + TACE | Combination therapy with transarterial chemoembolization (TACE) |
| BRISK-PS| III   | Brivanib    | Placebo     | Second-line therapy in sorafenib-resistant HCC |
| BRISK-TA| III   | Brivanib    | Placebo     | Combination therapy with transarterial chemoembolization (TACE) |
| BRISK-FL| III   | Brivanib    | Sorafenib   | First-line clinical trial for brivanib versus sorafenib |

Table 3: Markers for cancer stem cell of HCC in recent reports.

| Markers          | References |
|------------------|------------|
| CD133            | [61]       |
| CD90             | [62]       |
| CD44             | [62]       |
| EPCAM            | [63, 64]   |
| ABC transporters | [72]       |
| CD13             | [79]       |

trials, such as the SHARP trial [27] and Asia-Pacific trial [28] (Table 1). It improves overall survival in patients with advanced HCC compared to patients administered a placebo in the BSC setting. Other tyrosine kinase inhibitors were also tested in clinical trials. Sunitinib is an inhibitor of VEGFR and PDGFR [53] (Table 1). The clinical phase II trial of sunitinib for HCC treatment showed severe grade 3 to 4 side effects [53]. Therefore, the comparative study between sorafenib and sunitinib has ceased in April 2010.

Brivanib, erlotinib, and TSU-68, which are inhibitors of growth factor receptors, have been clinically tested for advanced HCC patients as well. The response rates to single doses of sorafenib [27, 28], sunitinib [53, 54], brivanib [55], erlotinib [56], and TSU-68 [57] were 2.3–3.3%, 2.7–2.9%, 5.0%, 9.0%, and 8.6% respectively (Table 1). Phase II clinical trials using bevacizumab [58], a VEGFR inhibitor, and cetuximab [59], an EGFR inhibitor, had 13% and 0% complete responses while cetuximab and TSU-68 achieved 2–3% complete responses [60] (Table 1). Therefore, combination of these agents with appropriate management of the side effects could improve survival of patients with advanced HCC in the future.

Ongoing clinical trials using molecular-targeted agents for HCC are shown in Table 2. The STORM trial is a phase III clinical trial using a single dose of sorafenib alone for adjuvant therapy after liver resection and ablation [29]. The SILLIUS trial is a phase III clinical trial using combination therapy with transarterial chemoembolization (TACE) and sorafenib for advanced HCC patients. The SPACE trial and TACTICS trial are a phase II clinical trial using TACE and sorafenib for advanced HCC patients. The BRISK-PS trial is designed for second therapy using brivanib for advanced HCC patients resistant to sorafenib. The BRISK-TA employs adjuvant therapy using brivanib after TACE, and the BRISK-FL trial is a comparative clinical trial using sorafenib alone and brivanib alone. These BRISK trials are a phase III clinical trial. The clinical results of these molecular-targeted therapies have not all been published yet and we will need to interpret the results carefully in the future.

6. Molecular Markers of Cancer Stem Cells in HCC

Molecular markers of cancer stem cells are shared with either normal stem cells or progenitor cells. CD133 [61], CD90 [62], CD44 [62], and EPCAM [63, 64] have been shown to be markers for cancer stem cells in HCC patients (Table 3). These biomarkers can be useful to estimate the prognosis of HCC and they could be useful for specific targeted therapy for cancer cells.

CD133 has been shown to be related to prognosis and metastasis in HCC patients [65]. Tumor proliferation was suppressed by anti-CD133 antibodies in a mouse model [66]. NSC74859 is a specific inhibitor of signal transducer and activator of transcription 3 (STAT3) activation and it decreases CD133-positive cells with suppression of cancer development [67]. In addition, CD133 cells increase in PTEN deleted mice [68], which indicates that PTEN can play an important role to regulate CD133-positive cancer stem cells. These basic studies suggest that CD133 can be a molecular target for HCC treatment.

The ATP-binding cassette (ABC) transporters are a family of membrane transporters such as MDR1 [69], ABCG2 [70], and ABCB2 [71]. ABC transporters protect cells from cytotoxic agents to reduce the drug sensitivity. A combination of chemotherapy and inhibitors of ABC transporters could decrease not only the number of HCC cells but also that of cancer stem cells [72] (Table 3).

CD90 is expressed in oval cells and progenitor cells and its expression is related to tumor development [73]. CD44 is a receptor of hyaluronate expressed on the cell surface and is often coexpressed with CD90 [62, 74]. Anti-CD44 treatment induces apoptosis in CD90-positive cells and thus CD44 plays an important role in the survival of cancer cells [75, 76]. EPCAM is another cell marker for progenitor cells and the direct target of the Wnt/beta catenin pathway.
The knockdown of the Wnt/beta catenin by small interfering RNA (siRNA) of Wnt/beta catenin decreases the number of EPCAM-positive cells with suppression of tumor development and induces apoptosis [78]. CD13-positive cells are also potential cancer stem cells [79] (Table 3). These cells can be found in the peripheral areas of HCC after TACE treatment, which is considered to be related to tumor recurrence [80]. Furthermore, inhibitors of CD13 such as 24F can suppress the invasion and angiogenesis of HCC [81].

7. Summary

The molecular pathogenesis of HCC is important to understand the mechanism of tumor development as well as the high-recurrence behavior of HCC. Furthermore, each step of the molecular signals could be a target to control tumor progression. Further clinical studies using single agents and combination therapies need to be conducted for HCC treatment. The clinical benefits of cell-targeted therapy for cancer stem cells are eagerly awaited.

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