Can Lung Imaging Scores and Clinical Variables Predict Severe Course and Fatal Outcome in COVID-19 Pneumonia Patients? A Single-Center Observational Study

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Abstract: COVID-19 prediction models mostly consist of combined clinical features, laboratory parameters, and, less often, chest X-ray (CXR) findings. Our main goal was to propose a prediction model involving imaging methods, specifically ultrasound. This was a single-center, retrospective cohort observational study of patients admitted to the University Hospital Split from November 2020 to May 2021. Imaging protocols were based on the assessment of 14 lung zones for both lung ultrasound (LUS) and computed tomography (CT), correlated to a CXR score assessing 6 lung zones. Prediction models for the necessity of mechanical ventilation (MV) or a lethal outcome were developed by combining imaging, biometric, and biochemical parameters. A total of 255 patients with COVID-19 pneumonia were included in the study. Four independent predictors were added to the regression model for the necessity of MV: LUS score, day of the illness, leukocyte count, and cardiovascular disease ($\chi^2 = 29.16$, $p < 0.001$). The model accurately classified 89.9% of cases. For the lethal outcome, only two independent predictors contributed to the regression model: LUS score and patient’s age ($\chi^2 = 48.56$, $p < 0.001$, 93.2% correctly classified). The predictive model identified four key parameters at patient admission which could predict an adverse outcome.

Keywords: lung ultrasound; COVID-19; prognostic; pneumonia; CT; chest X-ray

1. Introduction

Since the end of 2019, the world has battled the growing coronavirus disease 2019 (COVID-19) epidemic caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), with more than 290 million confirmed cases worldwide [1]. The virus is designated as severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), and the disease is coronavirus disease 2019 (COVID-19). Although the understanding of COVID-19 is evolving, there are still major issues concerning the immune response to SARS-CoV-2, COVID-19 diagnosis, management, prevention, and emerging new variants which make it obvious that SARS-CoV-2 is here to stay and potentially becoming endemic [2,3].

Thoracic imaging, including chest X-ray (CXR) or computed tomography (CT), is essential in the diagnosis of COVID-19 pneumonia. However, previous studies showed that CXR was less sensitive in the detection of COVID-19 lung disease compared to CT, with a reported baseline CXR sensitivity of 69% [4]. The sensitivity of CT is 83–100%, considering the results of reverse transcription-polymerase chain reaction (RT-PCR) tests as
the gold standard for diagnosis of COVID-19 [5]. High-resolution CT (HRCT) is the “gold” standard imaging method to evaluate the severity of lung involvement in COVID-19 patients. Despite the above, CXR is still extensively being used in the diagnosis of COVID-19 pneumonia due to its wide availability and relative inexpensiveness. COVID-19 pneumonia changes on CXR are typically ill-defined bilateral alveolar opacities of peripheral distribution. Similarly, the most commonly reported HRCT findings of COVID-19 pneumonia include airspace opacities (ground-glass and/or consolidation), typically subpleural and multilobar involvement, sometimes associated with septal thickening [6].

In addition to the aforementioned imaging methods that use ionizing radiation, bedside lung ultrasonography (LUS) is a rapid, non-ionizing, repeatable, and reliable examination technique that is increasingly used by clinicians in the COVID-19 pandemic, especially for hospitalized and critically ill patients reducing the need for transportation. Growing evidence shows that LUS sensitivity is close to that of chest CT and is much higher than that of CXR; its usefulness for the management of patients with COVID-19 pneumonia, from diagnosis to monitoring, follow-up, and even outcome prediction, is also demonstrated [7–12].

The vast majority of patients with more severe symptoms of the disease have one or more comorbidities, such as obesity and cardiovascular disease, with high mortality among elderly patients [13]. Biochemical and hematological laboratory factors such as lymphopenia, elevated serum ferritin, d-dimer, troponin, C-reactive protein, lactate dehydrogenase, and IL-6 are associated with severe disease, poor prognosis, and increased mortality [14–16].

Identifying risk factors at hospital admission that can predict the clinical course of the disease would help physicians to provide appropriate and timely therapeutic interventions. The COVID-19 prediction models developed so far have mostly combined clinical features, laboratory parameters, and, less often, CXR findings [17–20]. To the best of our knowledge, none of the proposed combined prediction models involved lung ultrasound, and that was the main goal of our work.

2. Materials and Methods
2.1. Study Design

This was a single-center, retrospective cohort observational study.

2.2. Inclusion and Exclusion Criteria

The study included a consecutive cohort of patients admitted with the diagnosis of COVID-19 pneumonia in the University Hospital of Split, Croatia, from November 2020 to May 2021, before the COVID-19 vaccine was widely available. Inclusion criteria were WHO diagnostic criteria for pneumonia COVID-19, SARS-CoV-2 infection confirmed by PCR [1], and the existing LUS exam after admission. Exclusion criteria were pulmonary edema associated with heart failure, severe lung emphysema, chronic interstitial lung disease, severe hemodynamic instability and inability to change body position, severe chest deformity, extensive subcutaneous emphysema, any other pulmonary diseases impeding ultrasound image acquisition (i.e., significant pleural effusion, previous pneumonectomy), and an inability to undergo LUS examination (Figure 1).

2.3. Outcomes

The study’s primary outcome was the definition of biometric (e.g., age), biochemical (e.g., spO2, LDH), and radiological predictive factors (LUS, CXR, and CT scores) for the necessity of mechanical ventilation (MV) in the treatment of pneumonia or a lethal outcome for the patients.

The secondary outcomes were the comparisons of the reliability and applicability of the three radiological scores assessed.
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2.4. Data Extraction

Patient demographics, comorbidities, symptoms, laboratory tests, imaging findings, treatment modalities, disease severity, and mortality data were extracted from electronic medical records.

2.5. Acquisition Protocol

Lung ultrasound examinations were performed by a trained sonographer (IS) on admission to the hospital using ultrasound equipment (Toshiba Nemio XG Istyle, Tokyo, Japan) with a 1–6 MHz convex transducer. The extent and severity of pulmonary infiltrations were described by a numerically repeatable LUS (Lung Ultrasound Score) coefficient proposed for COVID-19 pneumonia by Soldati et al. [21]. Fourteen areas (three posterior, two lateral, and two anterior for each lung) were examined completely intercostally to cover the widest possible area with a single scan. Changes were scored from 0 to 3. Zero (0) is a regular finding, the existence of a regular and not thickened pleural line, with a sliding sign and the presence of A-lines; 1: denotes an irregular pleural line with some B

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Figure 1. Flowchart of the records included or excluded from the analysis. Legend: CT score—Computerized tomography score; CXR score—chest X-ray score; LUS score—Lung ultrasound score.
lines suggesting some loss of aeration; 2: suggests a severe loss of aeration by a broken pleural line and small-to-large consolidated areas with associated areas of white below the consolidated area; 3: is attributed if the scanned area shows large dense consolidations which signify a complete loss of aeration called “white lung” (Table 1). For each patient, the stated scores in all 14 zones were added up (ranging from 0 to 42) to obtain the total LUS score [21]. According to the part of the lung they are positioned in, the 14 areas were grouped in apical, middle, and basal for further statistical analysis.

Table 1. Grading of LUS score, CT score, and CXR score.

| Score/Grade | LUS Score                                                                 | CT Score                        | CXR Score                        |
|-------------|---------------------------------------------------------------------------|--------------------------------|---------------------------------|
| 0           | regular finding: the existence of a regular and not thickened pleural line, with the sliding sign, and the presence of A-lines | no abnormalities               | no abnormalities                |
| 1           | some loss of aeration: irregular pleural line with some B lines severe loss of aeration: broken pleural line | prevalent ground-glass opacities (GGOs) | interstitial infiltrates         |
| 2           | small-to-large, consolidated areas with associated areas of white below the consolidated area complete loss of aeration: scanned area shows large dense consolidations; “white lung” | GGOs mixed with consolidations | interstitial and alveolar infiltrates (interstitial predominance) |
| 3           |                                                                           | prevalent consolidations        | interstitial and alveolar infiltrates (alveolar predominance) |

Acronyms: CT—computerized tomography; CXR—Chest X-ray; LUS—lung ultrasound.

CT scans were acquired during hospitalization, as indicated by attending physicians, thus for a limited number of patients. Scanning was acquired on 128 slice multi-slice CT (Philips, Ingenuity Elite). Fourteen CT areas (three posterior, two lateral, and two anterior for each lung) on non-contrast native scans corresponded to fourteen LUS areas. CT findings in each area were classified as follows: 0—no abnormalities, 1—prevalent ground-glass opacities (GGOs), 2—GGOs mixed with consolidations, and 3—prevalent consolidations (Table 1) [22]. For each patient, these scores in all 14 areas were added up (ranging from 0 to 42) to obtain a total CT score.

Chest X-rays were also acquired upon hospital admission, but the CXR score was assessed retrospectively. For CXR scoring, we used the Brixia score [23]. Chest X rays were divided into 3 zones per lung (upper, middle, and lower) in a total of 6 zones: the upper zone extends above the inferior wall of the aortic arch; the mid-zone is the space below the inferior wall of the aortic arch and above the inferior wall of the right inferior pulmonary vein; the lower zone extends below the inferior wall of the right inferior pulmonary vein [23]. Given that, we agreed that anatomically upper CXR zones (zone 1—left, 2—right) approximately corresponded to areas 6 (left) and 3 (right) on the LUS and CT; middle CXR zones (3—left, 4—right) corresponded to areas 2, 10, 14 (left) and 5, 8, 12 (right) on the LUS and CT, and lower CXR zones (5—left and 6—right) corresponded to areas 4, 9, 13 (left) and 1, 7, 11 (right) on the LUS and CT. CXR changes within each zone were scored: 0—no abnormalities, 1—interstitial infiltrates, 2—interstitial and alveolar infiltrates (interstitial predominance), and 3—interstitial and alveolar infiltrates (alveolar predominance) see Table 1. For each patient, the stated scores in all 6 zones were added up (ranging from 0 to 18) to obtain the total CXR score.

2.6. Bias

All patient records with an LUS exam were consecutively analyzed without exclusions to minimize selection bias. The only reduction in sample numbers was due to the availability of the other radiological scores, duplicate entries, or based on statistical tests (far out values detection).

The radiological exams were performed by a single clinician (LUS score—IS; CXR score—DBM; CT score—DBM), thus not as a two-person consensus. However, both authors (IS, DBM) are highly experienced in their respective clinical fields. HRCT and LUS exams were blinded and performed without knowledge of laboratory parameters, current
treatment, and further involvement in the treatment of the patient. However, the decision to mechanically ventilate the patient was up to the third party without knowledge (blinded) of the LUS score. The CXR and CT scores were assessed upon writing this manuscript with a clinician blinded to any of the observed predictors or clinical outcomes.

2.7. Study Size

This was a sample of consecutive patients collected during the 6-month pandemic peak in Croatia, and the expected enrollment was over 240 cases. We expected the area under the receiver operating curve to be above 0.7 with a ratio of positive outcome cases around 1:10. To accommodate statistical power of 80%, this required a minimum sample of 190 patients.

2.8. Statistical Analysis

Descriptive statistics were performed: categorical data were presented by absolute and relative frequencies; continuous data with normal distribution were presented as mean and standard deviation (SE) when highly variable by the median and interquartile range (IQR). The outliers and far-out values were detected by the Tukey method [24]. The normality of the distribution of continuous variables was tested by the Shapiro–Wilk test. The t-test was used to compare the means and the Mann–Whitney U test to compare the medians between two groups, while the one-way Chi-square test was used to compare proportions for dichotomous variables. Logistic regression analysis (univariate, multivariate—stepwise method) was used to analyze independent factors associated with the necessity for mechanical ventilation or lethal outcome. The continuous variables included in the models were added to the combined model with their respective slope coefficients as follows: combined score = \( \beta_1 \times \text{var}1 + \beta_2 \times \text{var}2 + \beta_3 \times \text{var}3 + \ldots + \beta_n \times \text{var}n \). The receiver operating curve (ROC) was used to determine the optimal threshold, the area under the curve (AUC), specificity, and sensitivity of the tested parameters. Regression analysis was used to describe the relationship between radiological scores. The type I error (alpha) was set to 0.05, and the type II error (beta, statistical power) was set to 80%. The statistical software used for analysis was MedCalc® Statistical Software version 19.6 (MedCalc Software Ltd., Ostend, Belgium; https://www.medcalc.org, accessed on 15 July 2021).

2.9. Reporting

We reported the study in line with the STROBE reporting guideline for cohort studies; the STROBE checklist is available in Supplementary File S1.

3. Results

3.1. Patients and Characteristics

We examined 299 patient records hospitalized in the University Hospital of Split due to COVID-19 from November 2020 to May 2021. Eleven duplicate records or control LUS were excluded, and the rest of the 288 eligible records of patients who underwent LUS were analyzed (Figure 1). Out of 265 records of patients that had a chest X-ray, 10 were excluded due to the dates between LUS and CXR exams being too far apart. Two records out of 42 patients who underwent CT scans were excluded for the same reason. Finally, 255 cases with CXR and LUS and 40 with CT and LUS scores were analyzed.

Patient characteristics between groups not requiring and requiring MV differed only when cardiovascular disease, hemiplegia, or leukemia were present as a negative prognostic factor. Almost the same was noticed in the case of a death outcome (Table 2). Similarly, factors for both the necessity for MV and fatal outcome were the age of the patients and the day of the illness on admission (Table 2). Furthermore, a difference was found in biochemical parameters such as elevated LDH in the group requiring MV, lower spO2, and higher troponin among patients with a fatal outcome. Leukocyte counts were lower in both mechanically ventilated and deceased patients, and respective LUS scores were significantly higher (Table 2).
Table 2. Patient characteristics according to need for mechanical ventilation or death outcome.

| Comorbidity/habit                        | No MV \(\text{n/N (\%)}\), Median (IQR) | MV Required \(\text{n/N (\%)}\), Median (IQR) | \(p^*\) | No Death \(\text{n/N (\%)}\), Median (IQR) | Death \(\text{n/N (\%)}\), Median (IQR) | \(p^*\) |
|-----------------------------------------|-----------------------------------------|-----------------------------------------------|--------|-------------------------------------------|----------------------------------------|--------|
| Arterial hypertension                   | 132/253 (52.2%) 14/28 (50.0%) 0.827    | 126/253 (49.8%) 20/28 (71.4%)                 | 0.030  |
| Cardiovascular disease                  | 34/253 (13.4%) 8/28 (28.6%) 0.033      | 31/253 (12.3%) 11/28 (39.3%)                 | 0.000  |
| COPD                                    | 14/253 (5.5%) 1/28 (3.6%)              | 14/253 (5.5%) 1/28 (3.6%)                   | 0.662  |
| CVI or TIA                              | 3/253 (1.2%) 0/28 (0.0%)               | 3/253 (1.2%) 0/28 (0.0%)                    | 0.563  |
| Dementia                                | 3/253 (1.2%) 0/28 (0.0%)               | 1/253 (0.4%) 2/28 (7.1%)                    | 0.001  |
| Diabetes                                | 50/253 (19.8%) 6/28 (21.4%)            | 50/253 (19.8%) 6/28 (21.4%)                 | 0.835  |
| Hemiplegia                              | 0/253 (0.0%) 1/28 (3.6%)               | 0/253 (0.0%) 1/28 (3.6%)                    | 0.662  |
| Kidney failure                          | 12/253 (4.7%) 1/28 (3.6%)              | 10/253 (4.0%) 3/28 (10.7%)                  | 0.107  |
| Leukemia                                | 3/253 (1.2%) 2/28 (7.1%)               | 4/253 (1.6%) 1/28 (3.6%)                    | 0.451  |
| Liver failure                           | 7/253 (2.8%) 0/28 (0.0%)               | 6/253 (2.4%) 1/28 (3.6%)                    | 0.700  |
| Myocardial infarct                      | 10/253 (4.0%) 3/28 (10.7%)             | 10/253 (4.0%) 3/28 (10.7%)                  | 0.107  |
| Perforated ulcer                        | 2/253 (0.8%) 1/28 (3.6%)               | 2/253 (0.8%) 1/28 (3.6%)                    | 0.175  |
| Rheumatological disease                | 8/253 (3.2%) 0/28 (0.0%)               | 8/253 (3.2%) 0/28 (0.0%)                    | 0.341  |
| Smoker                                  | 33/243 (13.6%) 4/27 (14.8%)            | 34/246 (13.8%) 3/24 (12.5%)                 | 0.858  |
| Biometrics                              |                                         |                                               |        |
| Female gender                           | 68/252 (27.0%) 10/28 (35.7%)           | 66/252 (26.2%) 12/28 (42.9%)                | 0.035  |
| Age (years)                             | 62 (55–70) 69 (62–75)                  | 62 (54–68) 75 (69–82)                       | 0.000  |
| Weight (kg)                             | 91 (84–103) 94 (82–103)                | 91 (84–103) 92 (80–97)                      | 0.388  |
| Height (cm)                             | 180 (173–186) 175 (170–186)            | 179 (173–187) 176 (168–180)                 | 0.181  |
| BMI (kg/m\(^2\))                       | 28.3 (26.1–30.9) 29.4 (27.8–33.1)      | 28.4 (26.1–31.1) 28.7 (26.9–32.9)          | 0.803  |
| Day of the illness                      | 9 (6–12) 6 (5–8)                      | 9 (6–12) 7 (4–10)                           | 0.035  |
| Biochemical parameters                  |                                         |                                               |        |
| CRP (mg/L)                              | 76.3 (47.8–136.8) 98 (67.8–146.6)      | 79.9 (49.3–136.8) 95.8 (61.4–146.6)         | 0.416  |
| D–dimer (ng/L)                          | 0.92 (0.61–1.49) 1.05 (0.61–2.20)      | 0.88 (0.60–1.48) 1.48 (0.72–2.66)           | 0.051  |
| LDH (U/L)                               | 351 (282–424) 434 (306–456)            | 335 (285–429) 366 (285–448)                 | 0.545  |
| Lymphocytes count (10\(^9\)/L)          | 7.9 (5.7–10.8) 5.7 (4.5–8.7)           | 7.9 (5.7–10.6) 5.4 (4.4–10.5)               | 0.042  |
| Neutrophils (%)                         | 13.0 (8.4–18.4) 16.3 (10.8–23.9)       | 13.0 (8.5–18.4) 16.3 (9.2–23.8)             | 0.272  |
| Neutrophils (%)                         | 81.2 (74.6–86.9) 75.8 (70.9–83.1)      | 81.2 (74.8–86.7) 75.8 (69.4–83.0)           | 0.123  |
| pO2 (kPa)                               | 7.30 (6.5–7.97) 7.27 (6.18–7.78)       | 7.30 (6.50–7.95) 7.20 (6.06–7.97)           | 0.506  |
| spO2 (%)                                | 91 (88–93) 89 (83–94)                  | 91 (88–93) 88 (82–93)                      | 0.022  |
| hs-Troponin (ng/L)                      | 9.45 (6.50–15.30) 10.9 (8.0–20.9)      | 9.25 (6.55–14.65) 14.6 (8.60–44.10)         | 0.012  |

* for dichotomous \(\text{n/N}\) one-way classification Chi-squared test was used; for continuous variables without normal distribution, Mann–Whitney U-test was used, and respective \(p\)-values stated. Acronyms: BMI—body mass index; COPD—chronic obstructive pulmonary disease; CXR—chest X-ray score; CT—chest computerized tomography score; IQR—interquartile range; LUS—lung ultrasound score; MV = 1—mechanical ventilation required, MV = 2—MV not required, death = 0—no death, death = 1—death outcome; SD—standard deviation; TIA—transitory ischemic attack.

3.2. Predictors of Necessity for MV or a Lethal Outcome

When combined in stepwise multivariate analysis, both CXR and CT scores were excluded from models for either MV or lethal outcome. Only the LUS score was retained in both models as follows (univariate analysis details available in Supplementary File S2).

Four independent predictors gave a unique statistically significant contribution to the regression model for the necessity of MV, and these are LUS score, day of the illness at admission, leukocyte count, and presence of cardiovascular disease (\(\chi^2 = 29.16, p < 0.001\)). The model accurately classified 89.9% of cases (Table 3, Figure 2A).
Table 3. Stepwise multivariate logistic regression according to the necessity of MV or lethal outcome.

| Variable                        | Coefficient | \( p \)  | Odds Ratio | 95% CI         | Cutoff | Sensitivity | Specificity |
|---------------------------------|-------------|----------|------------|----------------|--------|-------------|-------------|
| Mechanical ventilation          |             |          |            |                |        |             |             |
| LUS score                       | 0.101       | <0.001   | 1.11       | 1.04–1.17      | >27    | 72.4%       | 60.8%       |
| Day of illness                  | -0.131      | 0.022    | 0.88       | 0.78–0.98      | ≤7     | 72.4%       | 64.1%       |
| Leukocyte count                 | -0.160      | 0.021    | 0.85       | 0.74–0.98      | ≤6.3   | 64.3%       | 66.7%       |
| Cardiovascular disease present  | 1.019       | 0.041    | 2.77       | 1.04–7.35      | positive | 28.6%       | 86.6%       |

| Combined score                  | Correctly Classified | AUC | 95% CI | Cutoff | Sensitivity | Specificity |
|---------------------------------|----------------------|-----|--------|--------|-------------|-------------|
| Mechanical ventilation          | 88.9%                | 0.807 | 0.735–0.851 | >0.51 | 85.7%       | 66.4%       |
| LUS score                       | 93.2%                | 0.859 | 0.841–0.901 | >9.6  | 96.5%       | 57.9%       |

For the lethal outcome, only two independent predictors gave a unique statistically significant contribution to the regression model: LUS score and age of the patient (\( \chi^2 = 48.56, p < 0.001 \)). The model accurately classified 93.2% of cases (Table 3, Figure 2B).

3.3. Relationship between LUS and CXR Scores

The regression model between the CXR score and LUS score demonstrated a strong trend (slope 0.160, 95%CI 0.109 to 0.212, \( p < 0.001 \)); however, there was significant variability around the regression line (\( R^2 = 0.128 \); Supplementary Files S2 and S3).

Prediction models for MV based on LUS score (AUC = 0.693 ± 0.058) and CXR score (AUC = 0.586 ± 0.054) showed no significant difference of 0.106, \( p = 0.136 \) (Figure 3A).
Figure 3. Receiver operating curves. (A) LUS score vs. CXR score—prediction of mechanical ventilation. (B) LUS score vs. CXR score—prediction of lethal outcome. (C) LUS score vs. CT score—prediction of mechanical ventilation. (D) LUS score vs. CT score—prediction of lethal outcome. Legend: AUC—Area under hierarchical receiving operator curve; CT score—Computerized tomography score; CXR score—chest X-ray score; LUS score—Lung ultrasound score.

Additionally, models for death based on LUS score (AUC = 0.697 ± 0.064) and CXR score (AUC = 0.645 ± 0.059) showed no significant difference of 0.052, p = 0.449 (Figure 3B).

3.4. Relationship between LUS and CT Scores

The regression model between the CT score and LUS score demonstrated a strong trend (slope 0.502, 95%CI 0.292 to 0.711, p < 0.0001); however, there was significant variability around the regression line (R² = 0.396; Supplementary File S2).
Prediction models for MV based on LUS score (AUC = 0.871 ± 0.090) and CT score (AUC = 0.843 ± 0.077) showed no significant difference of 0.023, \( p = 0.819 \) (Figure 3C).

Additionally, models for death based on LUS score (AUC = 0.885 ± 0.057) and CT score (AUC = 0.836 ± 0.080) showed no significant difference of 0.049, \( p = 0.582 \) (Figure 3D).

4. Discussion

The results of our study draw us to conclusions defining four independent predictors regarding the regression model for the necessity of MV: LUS score, day of the illness at admission, leukocyte count, and presence of cardiovascular disease. For the lethal outcome, the two independent predictors were the LUS score and the age of the patient. Although LUS' predictive value was shown in several previous studies, as far as we know, it was not combined with demographics, clinical data, and laboratory parameters, creating prediction models for disease severity.

The average CXR score was not previously shown to be different between the group of patients who needed MV and the group of patients who did not. Although it is a possible prognostic factor for death in the univariate model, the CXR score was dropped from the multivariate model as insignificant in favor of, for example, the patient’s age. Therefore, we can conclude that the CXR score is inferior to the LUS score in the prediction of the need for MV or death.

For both the CT score and the CXR score, we showed a significant correlation with the LUS score by linear regression, although in both cases, there was significant variability. We attributed this to subjectivity in the scoring system based on the decision of the clinician interpreting the finding, the lower sensitivity and specificity of the CXR, and the time interval between CT, LUS, and CXR acquisition, in comparison with other studies that used shorter time delays among them [25]. On the other hand, the narrower CXR score scale (0–18) allowed for a smaller distribution of the total score. It was expected that the relative cutoff value (out of maximal score) would be higher in CXR than in the LUS score, which did not prove to be exact. The cutoff CXR score for the prediction of MV was already at 28% of the maximum score, while for the fatal outcome, it was at 38% and was therefore clinically completely irrelevant for these critical outcomes for the remaining upper ~60% of the score scale. We believe that based on plain CXR, very little can be described, such as the gentle difference of changes in the lung parenchyma essential for the outcomes. The non-specificity of CXR is a possible reason for the relatively low cutoff values obtained.

On the other hand, a much better correlation between LUS score and CT score, with significantly less variability and equal width of the scoring scale, speaks in favor of LUS as an excellent method of choice in monitoring patients with pneumonia; this was confirmed in other studies as well [25,26]. Despite this, previously published studies of predictive models involving radiological imaging have exceptionally included ultrasound more commonly than CT or chest CXR. Huang J. et al. suggested a diagnostic model obtained to distinguish between moderate and severe/critical COVID-19 using CT as a part of the diagnostic procedure. Their model included CT imaging in all patients and even those with mild symptoms [27]. Although CT is the gold standard, of course, the widespread use of such a model, especially in times of intense epidemics, is not possible. The Dutch COVID-19 risk model presented by Schalekamp et al. used similar demographic and laboratory parameters combined with the chest CXR score, which, unlike ours, used four zones in determining the CXR score. As in our study, patients who developed critical illness had leukopenia and higher lactate dehydrogenase levels more often. Contrary to our results, no significant difference was found for the duration of symptoms, although the proportion of patients with a symptom duration of more than 7 days was slightly smaller among those with critical disease [27]. As previously reported, age was a strong predictor of a lethal outcome in our population as well [28].

Multiple studies reported a significant association between elevated leukocyte counts and decreased lymphocyte counts among patients with severe cases of COVID-19 compared
with those with mild cases [15,29]. However, in our study population, a decreased leukocyte count was predictive of more severe disease.

Various biochemical markers were researched for their predictive usefulness, for example, LDH, CRP, and D-dimer [30–32]. In our study, patients who needed MV had significantly higher LDH values, although LDH did not appear to have predictive power in regression analysis. CRP and D-dimer were not significantly higher in the MV or deceased groups.

In our study, a lower day of illness at hospital admission showed to be a predictive factor for disease severity (MV), as it might suggest a more rapidly evolving and progressive disease in these patients. Similar results concerning onset to hospitalization time were found as a part of the risk nomogram established to predict the incidence of severe or critical COVID-19 in elderly patients in the study by Zeng et al. [20].

The percentage of smokers in the hospitalized population was significantly lower than in the overall population of Croatia, regarding which there is still controversy in the available literature, but most studies still link smoking to a higher risk of severe COVID-19 [32–35].

The proportion of patients who ended up on MV or died was similar to our previous study [11]. Only the therapeutic options in the patients changed, so we believe that the lower predictability of the LUS score in this study could be related to a modified corticosteroid therapy that the patients received (methylprednisolone 1 mg/kg vs. dexamethasone 6 mg used previously), which might have affected the clinical course and mortality of this disease.

4.1. Strengths

Our study was conducted on a large sample of hospitalized patients, and data were analyzed without exclusion by clinical characteristics using only statistically relevant and transparent figures.

4.2. Limitations

As mentioned in the bias section of the methods, we did not do a double, independent check of the radiological scores, and this could be a limiting factor. Although we demonstrated a strong association between the LUS score and the CT score, the sample for the CT score was relatively small and did not reach the test strength of 80%.

5. Conclusions

The predictive model obtained in our study identified four key parameters at the patient admission to the hospital, LUS score, day of the illness, leukocyte count, and presence of cardiovascular disease, that can predict an adverse COVID outcome. Hopefully, it will help physicians provide appropriate and timely therapeutic interventions.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/xxx/s1, Supplementary File S1—STROBE checklist, Supplementary File S2—Univariate regression of predictors for mechanical ventilation or death, Supplementary File S3—Scatter diagram and regression line between LUS score and CXR score.

Author Contributions: Conceptualization, I.S., M.P.I., K.D. and D.B.M.; methodology, I.S., I.J., J.D. and D.B.M.; software, O.B.; validation, I.S. and O.B.; formal analysis, I.S., I.J., J.D. and D.B.M.; investigation, I.S., I.J., J.D. and D.B.M.; resources, I.S., I.J., J.D. and D.B.M.; data curation, I.S., I.J., J.D., D.B.M. and O.B.; writing—original draft preparation, I.S., M.P.I., O.B. and D.B.M.; writing—review and editing, I.S., I.J., J.D., M.P.I., D.B.M., O.B. and K.D.; visualization I.S., I.J., J.D. and D.B.M.; supervision, K.D. and M.P.I.; project administration, I.S., M.P.I. and D.B.M.; funding acquisition, K.D. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.
Institutional Review Board Statement: The study was approved by the Institutional Ethics Board of University Hospital Centre Split, Croatia (Klasa: 500-03/21-01/01, Ubr.br. 2181-147-01/06/M.S.-20-02, 29 January 2021).

Informed Consent Statement: Written informed consent was not required for participants with emerging infectious diseases, LUS is a standard of care in our center, and this study had no interventions.

Data Availability Statement: The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Conflicts of Interest: The authors declare no conflict of interest.

References
1. WHO. Diagnostic Testing for SARS-CoV-2. Available online: https://www.who.int/publications/i/item/diagnostic-testing-for-sars-cov-2 (accessed on 18 April 2021).
2. Cascella, M.; Rajnik, M.; Cuomo, A.; Dulebohn, S.C.; Di Napoli, R.S. Features, Evaluation, and Treatment of Coronavirus; StatPearls: Treasure Island, FL, USA, 2020.
3. Pascarella, G.; Strumia, A.; Piliego, C.; Bruno, F.; del Buono, R.; Costa, F.; Scarlata, S.; Agrò, F.E. COVID-19 diagnosis and management: A comprehensive review. J. Intern. Med. 2020, 288, 192–206. [CrossRef] [PubMed]
4. Borakati, A.; Perera, A.; Johnson, J.; Sood, T. Diagnostic accuracy of X-ray versus CT in COVID-19: A propensity-matched database study. BMJ Open 2020, 10, e042946. [CrossRef] [PubMed]
5. Kovacs, A.; Palásti, P.; Vereb, D.; Bozsik, B.; Palkó, A.; Kincses, Z.T. The sensitivity and specificity of chest CT in the diagnosis of COVID-19. Eur. Radiol. 2021, 31, 2819–2824. [CrossRef] [PubMed]
6. Kaufman, A.E.; Naidu, S.; Ramachandran, S.; Kaufman, D.S.; Fayad, Z.A.; Mani, V. Review of radiographic findings in COVID-19. World J. Radiol. 2020, 12, 142–155. [CrossRef]
7. Jackson, K.; Butler, R.; Aujayeb, A. Lung ultrasound in the COVID-19 pandemic. Postgrad. Med. J. 2021, 97, 34–39. [CrossRef]
8. Lieveld, A.W.; Kok, B.; Schuit, F.H.; Azijli, K.; Heijmans, J.; van Laarhoven, A.; Assman, N.L.; Kootse, R.S.; Olgers, T.J.; Nanayakkara, P.W.; et al. Diagnosing COVID-19 pneumonia in a pandemic setting: Lung Ultrasound versus CT (LUVCT)—A multicentre, prospective, observational study. ERJ Open Res. 2020, 6, 00539-2020. [CrossRef]
9. Ottaviani, S.; Franc, M.; Ebstein, D.; Demaria, L.; Lheure, C.; Debray, M.; Khalil, A.; Crestani, B.; Borie, R.; Dieudé, P. Lung ultrasonography in patients with COVID-19: Comparison with CT. Clin. Radiol. 2020, 75, 877.e1–877.e6. [CrossRef]
10. Nouvenne, A.; Zani, M.D.; Milanese, G.; Parise, A.; Baciarello, M.; Bignami, E.G.; Odone, A.; Sverzellati, N.; Meschi, T.; Ticinesi, A. Lung Ultrasound in COVID-19 Pneumonia: Correlations with Chest CT on Hospital admission. Respir. Int. Rev. Thorac. Dis. 2020, 99, 617–624. [CrossRef]
11. Skopljanac, I.; Ivelja, M.; Barcot, O.; Brdar, I.; Dolic, K.; Polasek, O.; Radic, M. Role of Lung Ultrasound in Predicting Clinical Severity and Fatality in COVID-19 Pneumonia. J. Pers. Med. 2021, 11, 757. [CrossRef]
12. Gutsche, H.; Lesser, T.G.; Wolfram, F.; Doenst, T. Significance of Lung Ultrasound in Patients with Suspected COVID-19 Infection at Hospital Admission. Diagnostics 2021, 11, 921. [CrossRef]
13. Mohammad, S.; Aziz, R.; Al Mahri, S.; Malik, S.S.; Haji, E.; Khan, A.H.; Khatlani, T.S.; Bouchama, A. Obesity and COVID-19: What makes obese host so vulnerable? Immun. Ageing 2021, 18, 1. [CrossRef] [PubMed]
14. Ji, D.; Zhang, D.; Xu, J.; Chen, Z.; Yang, T.; Zhao, P.; Chen, G.; Cheng, G.; Wang, Y.; Bi, J.; et al. Prediction for Progression Risk in Patients With COVID-19 Pneumonia: The CALL Score. Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am. 2021, 71, 1393–1399. [CrossRef] [PubMed]
15. Huang, G.; Kovalic, A.J.; Graber, C.J. Prognostic Value of Leukocytosis and Lymphopenia for Coronavirus Disease Severity. Emerg. Infect. Dis. 2020, 26, 1839–1841. [CrossRef]
16. Luo, X.; Zhou, W.; Yan, X.; Guo, T.; Wang, B.; Xia, H.; Ye, L.; Xiong, J.; Jiang, Z.; Liu, Y.; et al. Prognostic Value of C-Reactive Protein in Patients with Coronavirus 2019. Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am. 2020, 71, 2174–2179. [CrossRef]
17. Fink, D.L.; Khan, P.Y.; Goldman, N.; Cai, J.; Hone, L.; Mooney, C.; El-Shakankery, K.H.; Sisemore, G.; Whitford, V.; Marks, M.; et al. Development and internal validation of a diagnostic prediction model for COVID-19 at time of admission to hospital. QJM Mon. J. Assoc. Physicians 2021, 114, 699–705. [CrossRef]
18. Guan, X.; Zhang, B.; Fu, M.; Li, M.; Yuan, X.; Zou, Y.; Peng, J.; Guo, H.; Lu, Y. Clinical and inflammatory features based machine learning model for fatal risk prediction of hospitalized COVID-19 patients: Results from a retrospective cohort study. Ann. Med. 2021, 53, 257–266. [CrossRef] [PubMed]
19. Qi, J.; He, D.; Yang, D.; Wang, M.; Ma, W.; Cui, H.; Ye, F.; Wang, F.; Xu, J.; Li, Z.; et al. Severity-associated markers and assessment model for predicting the severity of COVID-19: A retrospective study in Hangzhou, China. BMC Infect. Dis. 2021, 21, 774. [CrossRef] [PubMed]
20. Zeng, F.; Deng, G.; Cui, Y.; Zhang, Y.; Dai, M.; Chen, L.; Han, D.; Li, W.; Guo, K.; Chen, X.; et al. A predictive model for the severity of COVID-19 in elderly patients. Aging 2020, 12, 20982–20996. [CrossRef]
21. Soldati, G.; Smargiassi, A.; Inchingolo, R.; Buonsenso, D.; Perrone, T.; Briganti, D.F.; Perlini, S.; Torri, E.; Mariani, A.; Mossolani, E.E.; et al. Proposal for International Standardization of the Use of Lung Ultrasound for Patients With COVID-19: A Simple, Quantitative, Reproducible Method. J. Ultrasound Med. Off. J. Am. Inst. Ultrasound Med. 2020, 39, 1413–1419. [CrossRef]

22. Smargiassi, A.; Soldati, G.; Torri, E.; Mento, F.; Milardi, D.; del Giacomo, P.; de Matteis, G.; Burzo, M.L.; Larici, A.R.; Fompili, M.; et al. Lung Ultrasound for COVID-19 Patchy Pneumonia: Extended or Limited Evaluations. J. Ultrasound Med. Off. J. Am. Inst. Ultrasound Med. 2021, 40, 521–528. [CrossRef]

23. Borghesi, A.; Maroldi, R. COVID-19 outbreak in Italy: Experimental chest X-ray scoring system for quantifying and monitoring disease progression. La Radiol. Med. 2020, 125, 509–513. [CrossRef] [PubMed]

24. Tukey, J.W. Exploratory Data Analysis; THETA: Reading, MA, USA, 1977; Volume 2.

25. Gibbons, R.C.; Magee, M.; Goett, H.; Murrett, J.; Genninger, J.; Mendez, K.; Tripod, M.; Tyner, N.; Costantino, T.G. Lung Ultrasound vs. Chest X-Ray Study for the Radiographic Diagnosis of COVID-19 Pneumonia in a High-Prevalence Population. J. Emerg. Med. 2021, 60, 615–625. [CrossRef] [PubMed]

26. Rizzetto, F.; Perillo, N.; Artioli, D.; Travaglini, F.; Cuccia, A.; Zannoni, S.; Tombini, V.; di Domenico, S.L.; Albertini, V.; Bergamaschi, M.; et al. Correlation between lung ultrasound and chest CT patterns with estimation of pulmonary burden in COVID-19 patients. Eur. J. Radiol. 2021, 138, 109650. [CrossRef] [PubMed]

27. Schalekamp, S.; Huisman, M.; van Dijk, R.A.; Boomsma, M.F.; Jorge, P.J.F.; de Boer, W.S.; Herder, G.J.M.; Bonarius, M.; Groot, O.A.; Jong, E.; et al. Model-based Prediction of Critical Illness in Hospitalized Patients with COVID-19. Radiology 2021, 298, E46–E54. [CrossRef] [PubMed]

28. Mi, J.; Zhong, W.; Huang, C.; Zhang, W.; Tan, L.; Ding, L. Gender, age and comorbidities as the main prognostic factors in patients with COVID-19 pneumonia. Am. J. Transl. Res. 2020, 12, 6537–6548. [PubMed]

29. Tong, X.; Cheng, A.; Yuan, X.; Zhong, X.; Wang, H.; Zhou, W.; Xu, X.; Li, Y. Characteristics of peripheral white blood cells in COVID-19 patients revealed by a retrospective cohort study. BMC Infect. Dis. 2021, 21, 1236. [CrossRef]

30. Chen, R.; Liang, W.; Jiang, M.; Guan, W.; Zhan, C.; Wang, T.; Tang, C.; Sang, L.; Liu, J.; Ni, Z.; et al. Risk Factors of Fatal Outcome in Hospitalized Subjects with Coronavirus Disease 2019 From a Nationwide Analysis in China. Chest 2020, 158, 97–105. [CrossRef]

31. Liu, F.; Li, L.; Xu, M.; Wu, J.; Luo, D.; Zhou, Y.; Li, B.; Song, X.; Zhou, X. Prognostic value of interleukin-6, C-reactive protein, and procalcitonin in patients with COVID-19. J. Clin. Virol. Off. Publ. Pan Am. Soc. Clin. Virol. 2020, 127, 104370. [CrossRef] [PubMed]

32. Martha, J.W.; Wibowo, A.; Pranata, R. Prognostic value of elevated lactate dehydrogenase in patients with COVID-19: A systematic review and meta-analysis. Postgrad. Med. J. 2021. [CrossRef]

33. Ismail, N.; Hassan, N.; Abd Hamid, M.H.N.; Yusoff, U.N.; Khamal, N.R.; Omar, M.A.; Wong, X.C.; Pathmanathan, M.D.; Mohd Zin, S.; Muhammad Zin, F.; et al. Association of smoking and severity of COVID-19 infection among 5889 patients in malaysia: A multi-center observational study. Int. J. Infect. Dis. 2022, 116, 189–196. [CrossRef]

34. Zhao, Q.; Meng, M.; Kumar, R.; Wu, Y.; Huang, J.; Lian, N.; Deng, Y.; Lin, S. The impact of COPD and smoking history on the severity of COVID-19: A systematic review and meta-analysis. J. Med. Virol. 2020, 92, 1915–1921. [CrossRef] [PubMed]

35. European Commission; Directorate-General for Communication; Directorate-General for Health and Food Safety. Attitudes of Europeans towards tobacco and electronic cigarettes: Report, European Commission. 2021. Available online: https://op.europa.eu/en/publication-detail/-/publication/c070c04c-6788-11eb-aeb5-01aa75ed71a1 (accessed on 19 April 2022). [CrossRef]