ValORIZATION OF BY-PRODUCTS FROM COMMERCIAL TUNA CANNERY FOR NEW FORMULATION OF PET’S FOOD PRODUCT: PHYSICOCHEMICAL CHARACTERIZATION AND DLC DETERMINATION

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ABSTRACT

Tunisia has natural resources favorable to fishing with a coastline that extends for a total length of 1,300 kilometers. The large quantities processed show that tuna canneries generate an important amount of discarded high-quality species. These wastes are, of course, problems, but they also offer excellent opportunities for biotechnological exploitation. The disposal of these wastes has always been expensive and has often a harmful impact on the environment, but thanks to the evolution of techniques and the development of markets, it is now possible to transform this waste stream into useful and marketable products. Thanks to their high protein content, tuna discarded products represent an important source for the development of a new animal feed product, including the young dog. In the present work, we focused primarily on the characterization of the red muscles of a tuna cannery for inclusion in the formulation of young dog’s wet food. The composition of these co-products indicates a high protein content (80%), and a moderate fat and mineral content (11% and 8%, respectively). Nevertheless, the carbohydrate content in the red muscles of this fish remains low (2%). To increase the carbohydrate content which constitute the principal energy source and fiber proportion to improve product digestibility by the animals, cereals (maize flour and rice flour) were used. Four formulae were tested, using the same manufacturing process. A physicochemical study of the finished products was carried out and the Limit Date of Consumption was determined and estimated to 3 years and 3 months.

INTRODUCTION

Over the past five decades, world fish production has progressively increased and the amount of fish for consumption has increased with an average annual growth rate of 3.2%, a higher rate than the global population, which stood at 1.6% [1]. In 2011, total global fishing accounted for 154 million tons of fish (91.6 million tons of fish and marine animals from capture and 62.4 million tons from aquaculture) [2]. The differences in the arrangement of sea food products concern all fresh products which are damaged when caught and fished, transported, handled or removed after sorting in packing plants, freezing or canning of fish. In general, about half of the world’s catches are consumed directly by humans. The fish processing industry is responsible for a huge amount of wastes, including skins, bones, fins, scales, and swim bladders, representing nearly 36% (and may represent up to 60%) of the raw material mass [3]. Given that ecological laws restricting the removal of fish discards are increasing, there is a strong demand towards the complete exploitation of marine resources. These releases contain high-nutritional protein, unsaturated fatty acids, vitamins, antioxidants, minerals, and essential amino acids and peptides that are beneficial to the body. It is therefore essential to find ways of...
making the most of these co-products by integrating the concept of sustainable development: the techniques used must be non-aggressive to the environment and must not involve excessive energy expenditure or investment [4]. Today, the fisheries resource development industry is a future component of the fishing industry.

The added value of these products is its contribution to reducing the loss of valuable protein. It may also be the source of new products [5]. Potential field of applications are very broad [6,7]. The most prevalent field is the use for human or pet food. The second is dietary and nutraceutical ingredients. The third is cosmetic and pharmaceutical ingredients and the latter covers various fields of application (agriculture, energy, leather, industry, etc.). The present study concerns the valorization of fishery products and aims at the development of food for young dogs and cats from red muscle co-products, resulting from the processing of tuna. The marine co-product advancement industry is part of the fish industry branch of the IAA sector. It is also included in the list of primary agricultural processing activities JORT [8]. In fact, valorization is of big interest to both the canned and semi-preserved fish industry, the frozen seafood industry, and the fish-based food industry. The valorization of fish muscle proteins from high quality discarded species has been studied, and these proteins are considered suitable for the preparation of different seafood products with enhanced added value in comparison to the traditional preparation of fish meals or fish oils [9-11]. However, we investigate the physicochemical characterization of the red muscles of tuna revealed the richness of these co-products in protein, minerals and fat. The use of these co-products in Tunisia is extremely rare and is limited to the manufacture of fish meal. The incorporation of these red muscles with cereals and sunflower oil in wet foods for young dogs has made it possible to have balanced and stable foods. A physicochemical study of the finished products was carried out and the Limit Date of Consumption was determined and estimated to 3 years and 3 months.

Materials and Methods

Raw Material

The selected fish species "Atlantic red thon" (Thunnus Thynnus) "yellow thon" (Thunnus albacares) and "pink thon" named also Skip Jack Tuna (Katsuwonus pelamis) were collected from local tuna cannery. Twenty kg of each selected species was separated from commercial ones and distributed in 12 kg iced boxes until further processing. Rice flour, corn flour and sunflower oil were purchased from a local provider.

Physicochemical Characterization

Determination of the Ashes

The percentage of residue is determined by calcinating 1g of dried sample in triplicate for 4 hours at 550°C to constant mass [12].

Minerals Determination

The resulting residues were treated with 5 ml of nitric acid and 5 ml of concentrated chloric acid, the solution is heated until white smoke is released and the acids evaporate. 15 ml of distilled water is added, while continuing the heating for 10 min. The solution was filtered in a 50 ml flask and the volume completed by the distilled water. Mineral contents (K, Mg, Ca, Na, Zn and Fe) were determined by atomic absorption according to the Official methods of analysis [13].

Determination of Proteins by the Kjeldahl Method:

The percentage of total nitrogen is expressed using the standard Kjeldahl method. The protein assay is based on a mineralization of organic matter then a distillation and titration with 0,1N sulphuric acid [14]. Protein (%)=(N*W*14*100)/m) 6.25V: Volume of sulphuric acid 0,1N (ml); N: TITLE of sulphuric acid solution (0,1N); m=mass of sample analyzed in mg; 6,25: Coefficient of conversion of organic nitrogen into proteins. This factor is since proteins contain an average of 16% nitrogen.

Lipids

Extraction of the fat in the sample was performed using the Soxhlet technique, using petroleum ether as an organic solvent [14].

Assays

Fatty acids were methylated by boron trifluoride in methanol (Sigma) according to the Wetzel and Schmitz [15] method. The Rapid Preparation of Fatty Acid Esters for Gas Chromatographic Analysis [16] and fatty acid methyl esters (FAMES) were analyzed by gas chromatography (GC) on a Shimadzu (GC-2014) equipped with non-polar DANI-SPA column (50 m length, 0.32 mm i.d.) and a flame ionization detector. Nitrogen was used as a carrier gas. The temperatures of the column oven, the injection ports, and the detector were maintained at 165, 250, and 250°C, respectively. Two microliters of the sample were injected into the column. Chromatographic peaks were identified by comparing the retention times with those of known standards, maintained at 165, 250, and 250°C, respectively. Two microliters of the sample were injected into the column. Chromatographic peaks were identified by comparing the retention times with those of known standards.

Determination of total Sugar Content

Total sugars were determined using the method described by Dubois et al. [17]. Optical density (DO) was measured at 490nm.

DLC Determination Test

The protocols for performing the DLC analyses or aging tests are described in the French standard NF V 01-003 [18] of February 2004. The principle is to keep a number of product units, by varying the temperature to create a "cold chain break" that will result in an increase in the different reactions leading to aging of the product and periodically we count the total germs. The resolution of
Arrhenius’ law, allows calculating the microbiological DLC of the products tested.

\[ r = \frac{d[A]}{dt} = K[A]^n \]

r: degradation rate; A: Parameter to be studied; K: Speed constant of the reaction; n: Order of reaction

Reaction of order 0: A-A0=kt, characterized by a linear relationship between the deterioration and reaction time. Reaction of order 1: ln (A0/A)=kt, characterized by a logarithmic change in the A concentration as a function of time. Order reaction n: 1/A n-1 -1/A0 n-1=kt, with: A0 the initial value of factor A on the day of its manufacture.

**Results and Discussion**

1-Physicochemical Characterization of Tuna Red Muscles:

**Table 1:** Physiochemical composition of red tuna muscles (%).

| Moisture (%) | Ashes (%) | Proteins (%) | Lipids (%) | Minerals (%) | Sugars (%) |
|--------------|-----------|--------------|------------|-------------|-----------|
| Muscle rouge | 75.35±    | 38.48±       | 80.09±     | 11.3±       | 7.67±     |
| (% MS)       | 0.33      | 0.56         | 0.65       | 0.03        | 0.09      |

**Table 2a:** CPG analyses of the lipid fraction of tuna’s red muscles.

| Fatty acids | (%) in the lipid fraction of the red muscles |
|-------------|-------------------------------------------|
| ΣA.G.S      | 25.83                                     |
| C12:0       | 5.64                                      |
| C14:0       | 1.41                                      |
| C16:0       | 4.92                                      |
| C18:0       | 13.86                                     |
| ΣA.G.MI     | 74.17                                     |
| C16:1       | 47.63                                     |
| C18:1       | 26.54                                     |

**Table 2b:** Red muscle minerals and trace elements (mg/100g).

| Minéraux  | Teneurs |
|-----------|---------|
| Calcium   | 0.36    |
| Potassium | 170.15  |
| Sodium    | 240.6   |
| Magnésium | 100.2   |
| Fer       | 0.48    |
| Cuivre    | <0.001  |
| Zinc      | 0.28    |

Characterization of the raw material is essential before any investigation into the formulation of a food product. In fact, the composition of primary material is critical to choose the suitable product to develop. The physicochemical characterization of tuna red muscles includes a dry matter, fat, protein and minerals is presented in Table 1. The results show that the red muscles have a high moisture content reaching around 75%. This could be explained by the high-water retention capacity of these co-products during the storage and cooking stages. Furthermore, the composition of these co-products indicates a high protein content (80% MS), and moderate fat (11% MS) and mineral content (8% MS). Nevertheless, the carbohydrate content in the red muscles of the tuna remains low (2%). These data point us towards an outlet for animal feed recovery. To emphasize the richness and benefits
of these red muscles in unsaturated fat, the fatty acid composition of the fat fraction of the co-products was achieved and presented in the Table 2a. Red muscle contains a diversity of fatty acids in the lipid fraction. It contains a moderate content of saturated fatty acids (26%) compared to beef (46%) and lamb (51%) [20], whose excess of this type of fat increases cardiovascular risk. In addition, it is an appreciable source of monounsaturated fat (74%). For food application, further characterization of the mineral composition is required. The results are summarized in Table 2b. Tuna red muscles contain high amounts of sodium (240mg/100g MS) and potassium (170mg/100g MS). The magnesium concentration is in the order of 100 mg/100 g, which is significantly higher (4 times higher) than those described for beef (24mg/100g) [19] or shrimp (61mg/100g) [20].

**Table 3:** Formulation of wet food product for young dogs: experimental values.

| Formulations | Muscles red tuna | Corn flour | Rice flour | Sunflower oil | Water |
|--------------|-----------------|-----------|-----------|--------------|-------|
| F1           | 65              | 5         | 10        | 5            | 15    |
| F2           | 67              | 5         | 10        | 5            | 13    |
| F3           | 70              | 5         | 5         | 5            | 15    |
| F4           | 70              | 5         | 10        | 5            | 10    |

**Physicochemical Characterization of Prepared Formulae**

The experimental values (Table 5a) confirm that the formula 2, 3 and 4 meet the standards required by the AAFCO [21] and the FEDIAF [22] in terms of amount of proteins and fat composition (Table 4). The third formulation present the highest moisture percentage. Meanwhile, formulation number 2 and 4 are richer in terms of dry matter, proteins and minerals matter, and shows the highest energy values. This allows us to validate the choice of these two formulations for further microbiological and physicochemical analysis. Their energy inputs represent respectively about 16% and 17% of the need daily energy (1297.16 kcal). Their mineral contents are represented in the Table 5a. In fact, some minerals and trace elements remain below the AAFCO requirements, such as calcium, copper and zinc. These deficiencies can be corrected by using mineral-vitamin complexes in food formulation. To highlight the nutritional benefits of these foods and their unsaturated fats, the fatty acid composition of the fat fraction has been achieved and presented in Table 5b. These two formulas contain a diversity of fatty acids in the fat fraction. They are rich in unsaturated fatty acids, especially oleic acid which accounts for 27% of formula 2 and about 39% of formula 4. Formula 4 is a source of linoleic acid (Ω6) (55.43%) compared to 50.28% of formula 2.

**Table 4:** Composition of prepared dogs food product (%).

| Formulation | Moisture (%) | Ashes (%) | Proteines | Lipids | Minerals | Sugar | VE |
|-------------|--------------|-----------|-----------|--------|----------|-------|----|
| 1           | 57.29±2.00   | 40.55±1.055 | 18.29±2.2 | 6.72±0.31 | 1.95±0.28 | 15.66±1.83 | 196.28 |
| 2           | 58.92±1.03   | 42.41±1.524 | 23.62±0.8 | 7.19±0.49 | 2.21±0.5 | 13.5±0.85 | 213.19 |
| 3           | 63.74±2.21   | 38.52±0.85 | 23.51±0.1 | 6.23±0.51 | 1.93±0.23 | 12.13±0.98 | 198.63 |
| 4           | 57.33±1.88   | 42.14±1.54 | 27.12±2.5 | 7.50±0.86 | 2.1±0.35 | 12.35±2.01 | 225.38 |
| AAFCO (2014)| 60           | 30        | 22.5      | 0.58    | 3.6      | ND    |

a= % of MS; VE: energy value in kcal/100 g of the product.

**Table 5a:** Mineral and trace element content in formulation 2 and 4.

| Minéraux | 2  | 4  | AAFCO (%) |
|----------|----|----|-----------|
| Calcium  | 0.83 | 0.76 | 1         |
| Potassium| 0.7 | 0.49 | 0.6       |
| Sodium   | 1.55 | 2.25 | 0.3       |
| Magnesium| 0.14 | 0.18 | 0.06      |
|     | Fer | Cuivre | Zinc |
|-----|-----|--------|------|
| 184 | 79.4 | 88     |
| 4.9 | 5.1  | 12.4   |
| 69.8| 97.4 | 100    |

Table 5b.

| Fatty acids | (%) in lipids fraction of       |
|-------------|--------------------------------|
|             | 2                              |
|             | 4                              |
| ΣA.G.S      |                                |
| C_{12:0}    | 0.86                           |
| C_{14:0}    | 0.17                           |
| C_{16:0}    | 11.78                          |
| C_{18:0}    | 3.74                           |
| ΣA.G.I      |                                |
| C_{16:1}    | 0.66                           |
| C_{18:1}    | 26.67                          |
| C_{18:2}    | 50.28                          |
| C_{18:3}    | 0.83                           |
| C_{20:1}    | 0.25                           |
| C_{20:2}    | 0.36                           |
| C_{20:4}    | 0.32                           |
| C_{20:5}    | 0.87                           |
| C_{22:6}    | 3.14                           |

Microbiological Stability Tests and DLC Determination

The microbiological analysis includes total aerobic mesophyll flora, staphylococcus aureus, fecal and total coliforms, ad anaerobic sulphur to reducer flora. Results confirm that the products tested are of good microbiological quality and meet the requirements of the fish canning standard. Despite the relatively high value of water activity (0.941 and 0.916 for the F2 and F4, respectively), the reference germs are absent, this can be explained by the efficiency of the heat treatment at 116°C for one hour after the sterilization procedure [23,24].

Aging Test

Figure 1a: Evolution of total germs over storage time at 37 and 45°C.
Measurement of total germs as a function of storage time and temperature (37 and 45°C) are reported in Figure 1. During storage, the total germ concentration increases linearly over time and temperatures rise, showing the positive effect of increasing temperatures (37 to 45°C) on the development of microorganisms. The linear regression lines representing number of germs as f(t) shown in Figure 1a, gives regression coefficients R² nearly of 1. Degradation equations are of zero order of type: \([\text{GT}]= [\text{GT}]_0 + kt\). This equation makes it possible to deduce the reaction constants at different temperatures: \(K = 83.4\) and \(185.3\) at 37°C and 45°C respectively.

**Determinations of Activation Energy and DLC:**

Determination of the DLC requires the measurement of the activation energy and the constant \(K_0\). For this purpose, the \(\ln K = f(1/T)\) curve was plotted where \((-E_a/R)\) is the slope of the linear regression line, and \(\ln K_0\) was originally ordered (Figure 1b). On the basis of the results of the total germ analyses, the determination of the DLC of the product is obtained by equation (2). The values obtained show the activation energy is of about 15862.22 (Cal/g/mol) for \(K_0\) of 10.37*1012 which correspond to a DLC of 1198 days at 37°C. The results obtained enable the DLC of the product to be calculated by considering the evolution of the fungal flora. And thus, counting the total germs in the food produced results in a consumption limit of three years and three months, without the addition of preservatives or other additives.

**Conclusion**

Current study is part of the valorization of the red muscles discarded during the processing of tuna in canneries of South Tunisian, in order to obtain a wet food for young dogs. The physicochemical characterization of these by-products has shown their richness in proteins, fats and minerals, hence the idea of integrating them into animal feed. The low carbohydrate and fibre content leads us to use foods rich in these elements, thus the use of both types of cereals (corn flour and rice flour) as a source of energy and fibers to have a finished product digestible by the animal. Four formulae were tested using the same manufacturing process. Subsequently, a physicochemical study and a determination of the DLC were carried out. The DLC determined for one of the prepared foods is three years and three months. The physicochemical characterization showed the product’s rich protein, carbohydrate and fat content, and consequently it was considered with an important energy value. The composition is also characterized by high concentrations of minerals (Mg, Na), confirming the nutritional richness of the product. Microbiological analyses were carried out to ensure the safety of the finished product and the results showed the absence of pathogens and the compliance of the product with current standards.

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