A randomized study comparing chemotherapy followed by G-CSF alone or in combination with GM-CSF for mobilization of peripheral blood stem cells in patients with non-Hodgkin’s lymphomas

Objective: Granulocyte colony-stimulating factor (G-CSF) and granulocyte-macrophage colony stimulating factor (GM-CSF) are the two most commonly used cytokines for mobilization of stem cells in patients undergoing high dose chemotherapy with stem cell support. Both cytokines increase the peripheral blood white blood cell count and the stem cell count but there are other differences in the stem cell products mobilized by G-CSF versus those mobilized with GM-CSF. Generally higher numbers of dendritic cells are mobilized with GM-CSF than by G-CSF. The primary objective of this randomized study was to evaluate the safety and efficacy of chemotherapy plus G-CSF versus chemotherapy plus G-CSF and GM-CSF in patients with B-cell non-Hodgkin’s lymphoma (NHL) who were undergoing chemo-mobilization. Secondary objectives were to determine the expression of various dendritic cell subsets in the two groups and to determine the incidence of disease progression or relapse at 12 months.

Methods: We prospectively evaluated 84 patients with relapsed NHL who were candidates for high dose therapy (HDT). All patients underwent chemo-mobilization using ifosfamide, etoposide, and rituximab. All patients were randomized in an adaptive manner to receive either G-CSF or G-CSF plus GM-CSF (G+GM) starting 24 hours after completion of chemotherapy and continuing until completion of apheresis. The stem cell yield/kg, the number of apheresis procedures needed in the two groups, and the toxicity were recorded. We also enumerated dendritic cell subsets, myeloid DCs (mDC) and plasmacytoid DCs (pDC), in apheresis products and in peripheral blood (PB) samples collected pre-chemotherapy. The data were expressed as a percentage of peripheral blood mononuclear cells.

Results: A total of 84 patients were treated. Forty-three patients received G-CSF and 41 received G+GM. Both regimens were well tolerated. The median CD34+ cell dose collected was similar in the two groups. A total of 54 (G-CSF N = 25 and G+GM N = 29) paired samples from baseline and post-apheresis were available for analysis of dendritic cell subsets. There was no significant difference in the percentages of mDC subsets between baseline and post-apheresis collected with G-CSF or G+GM mobilization. However, there was a significant increase in the percentage of pDC subsets in the G-CSF alone when compared to the G+GM arm (P = 0.002). Furthermore, the ratio of mDC and pDC was significantly lower after mobilization with G-CSF versus G+GM (P = 0.029).

Conclusion: Addition of GM-CSF to G-CSF to the mobilization regimen resulted in lower percentages of pDC in the apheresis products when compared to those with G-CSF alone. This shifts the mDC/pDC ratio in the apheresis grafts in favor of mDC in the combination arm. However, these differences did not seem to impact the clinical outcomes in the two groups. (ClinicalTrials.gov Identifier: NCT00499343).

Keywords: lymphoma, filgrastim, sargramostim, stem cell mobilization
**Introduction**

Peripheral blood stem cells (PBSC) are the preferred source of stem cells for autologous transplantation because of the technical advantage and the shorter time to engraftment. Mobilization of CD34+ cells into the peripheral blood can be achieved by the administration of filgrastim (G-CSF or Neupogen®), sargramostim (GM-CSF or Leukine®), or both, either alone or in combination with chemotherapy. Both cytokines differ somewhat in the number and composition of PBSCs and effector cells mobilized to the peripheral blood. Previous studies have shown a correlation between clinical outcome and the graft composition. As a single agent, G-CSF mobilizes more CD34+ cells than does GM-CSF. Studies using concurrent or sequential mobilization with G+GM suggest that the combination of the two growth factors is superior to either GM-CSF or G-CSF alone in mobilizing CD34+ cells. There are other differences in the stem cell product mobilized by G-CSF versus those mobilized with GM-CSF. Generally higher numbers of dendritic cells are mobilized with GM-CSF than by G-CSF. Dendritic cells (DCs) are important antigen presenting cells that are necessary for priming naïve T cells. They play an important role in the development of anti-tumor responses. In humans two types of DCs have been described: mDC and pDC.

It is unclear if mobilization of higher numbers of dendritic cells has any clinical advantage for patients undergoing high-dose chemotherapy and autologous PBSC transplantation.

The purpose of this randomized study was to evaluate the safety and efficacy of chemotherapy plus G-CSF versus chemotherapy plus G+GM in patients with B-cell NHL who were undergoing chemo-mobilization. Secondary objectives were to determine the degree of expression of various dendritic cell subsets in the two groups and determine the incidence of disease progression or relapse at 12 months in the two groups.

**Patients and methods**

Patients up to 70 years of age with relapsed or primary refractory CD20-positive NHL who were to undergo autologous stem cell collection for SCT at The University of Texas MD Anderson Cancer Center were eligible for this prospective study. All patients were required to have adequate hematological, renal, and hepatic function and a Zubrod performance status of <3. Patients with active infections or lymphoma in the central nervous system, prior pelvic radiation, more than three prior chemotherapy regimens, and more than six cycles of fludarabine-based chemotherapy were excluded. Patients had to be at least three weeks from their last chemotherapy. All patients provided written informed consent. The protocol was reviewed and approved by the Institutional Review Board of The University of Texas MD Anderson Cancer Center, and the study was reviewed annually by The University of Texas MD Anderson Data Safety Monitoring Board (DSMB). The data for this analysis were collected from the institution’s Department of Stem Cell Transplantation database and from patients’ medical records.

**Chemotherapy**

Patients received ifosfamide, 10 g/m², given by continuous intravenous infusion over 72 hours on days 1–3, etoposide, 150 mg/m², given intravenously every 12 hours for a total of six doses on days 1–3, and rituximab intravenously at 375 mg/m² on day 1 and at 1000 mg/m² on day 8. Patients randomized to the G-CSF arm started G-CSF on day +6 at 6 µg/kg (rounded off to the nearest vial) every 12 hours until completion of apheresis. Patients randomized to the G+GM arm started G-CSF 6 µg/kg (rounded off to the nearest vial) every 12 hours on day +6 along with GM-CSF at 250 µg/m² daily (rounded off to the nearest vial) till the completion of apheresis.

**Peripheral blood stem cell collection**

Upon recovery of counts PBSCs were collected using standard apheresis procedures. Apheresis was started when the peripheral blood CD34 counts reached $15 \times 10^6/mL$ and continued till the target CD34+ cell dose of $\geq 4 \times 10^6/kg$ was reached. All patients underwent leukapheresis using the COBE Spectra cell separator (COBE BCT, Inc., Lakewood, CO). Three times the estimated blood volume was processed during each collection. Anticoagulant citrate dextrose solution was used as an anticoagulant. Calcium was given by continuous infusion through the return line. The total nucleated cell count and the CD34+ cell concentration was measured immediately after completion of apheresis. Patients who failed to reach a peripheral blood CD34 count of $\geq 15 \times 10^6/mL$ or failed to collect a CD34+ cell dose of $\geq 2 \times 10^6/kg$ after four apheresis procedures were classified as mobilization failures.

**Graft composition**

Dendritic cell subsets, mDC and pDC, were enumerated by immunophenotyping. DCs were identified by their lack of
leukocyte lineage-specific markers, and expression of HLA DR, CD11c (mDC) and CD123 (pDC). The expression of these molecules in the apheresis product was compared with that of PB samples collected pre-chemotherapy as baseline. Additionally, the percentage of leukocytes expressing these differentiation antigens was compared for patients in the G-CSF arm with those in the GM-CSF arm by the Mann–Whitney U test; differences were considered significant for \( P \leq 0.05 \).

**Statistical methods**

This was a randomized trial to compare chemotherapy followed by G-CSF or G+GM for stem cell mobilization. The primary objective of the study was to determine the efficacy of *in vivo* purging achieved by rituximab in the two groups and to determine the number of apheresis procedures, total stem cell yield/kg patient body weight, and the toxicity profile in the two groups. Secondary objectives were to determine the expression of various dendritic cell subsets in the two groups and determine the incidence of disease progression or relapse at 12 months in the two groups.

Treatment success was defined as successful purging of stem cell product of monoclonal B-cells and the ability to collect \( \geq 4 \times 10^6 \) CD34+ cells/kg in four, or less, apheresis procedures. As part of the pretreatment evaluation patients provided bone marrow (BM)/PB for the measurement of monoclonal B-cells by flow cytometry. We assumed that if the BM/PB was not involved with monoclonal B-cells then the collected product did not have to be re-tested. Initially, patients were randomized fairly to the two treatment arms. After 40 patients had been randomized, adaptive randomization was employed to unbalance the randomization in favor of the treatment arm having the higher observed success rate. We assumed a beta (1.8, 0.2) prior distribution for the probability that the treatment arm having the higher observed success rate than the other with probability more than 0.90 was either treatment arm would be found to have a higher success rate than the other with probability more than 0.90 was less than 0.02.

Patient demographics are summarized in Table 1. There were no statistically significant differences between the two treatment groups with regard to demographic or clinical characteristics.

**Apheresis data**

All the apheresis products were negative for monoclonal B cells by flow cytometric analysis. Thirty-nine of 43 patients (90.7% with 90% CI: 82.7–96.6%) in the G-CSF arm and 35 of 41 patients (85.4% with 90% CI: 76.0–93.3%) in the G+GM arm successfully collected \( \geq 4 \times 10^6 \) CD34+ cells/kg body weight. The probability that the G-CSF arm

| Table 1 Baseline patient characteristics |
|----------------------------------------|
| Characteristic                         | G-CSF | G-CSF + GM-CSF |
| Total treated                         | 43    | 41            |
| Age (years)                           |       |               |
| 20 to 39                              | 4     | 3             |
| 40 to 59                              | 29    | 26            |
| 60 to 79                              | 10    | 12            |
| Gender                                |       |               |
| Male/Female                           | 29/14 | 24/17         |
| Histology                             |       |               |
| Low grade                             | 4     | 7             |
| Intermediate grade                    | 39    | 34            |
| Ann Arbor stage                       |       |               |
| 0–1                                   | 25    | 23            |
| >1                                    | 18    | 18            |
| LDH*                                  |       |               |
| Normal                                | 28    | 29            |
| >Normal                               | 15    | 11            |
| Number of prior chemotherapies        |       |               |
| Median (range)                        | 2 (1–4)| 2 (1–3)       |

*Notes:* One patient on the G-CSF + GM-CSF treatment arm has no data on LDH.

**Abbreviations:** G-CSF, granulocyte colony-stimulating factor; GM-CSF, granulocyte-macrophage colony-stimulating factor; LDH, lactate dehydrogenase.
has a higher success rate than the G+GM arm is 0.778. The median CD34+ cell dose collected was $10.34 \times 10^6$/kg (range, 0.14–59) in G-CSF arm and $7.5 \times 10^6$/kg (range, 0.79–73.7) in the G+GM arm ($P = 0.65$). Median numbers of apheresis procedures required were two in both arms ($P = 0.64$). Collection started a median of 17 days (range, 12–26) from the start of chemotherapy in the G-CSF arm and 14 days (range, 13–26) in the G+GM arm ($P = 0.06$). These results are summarized in Table 2.

**Count recovery**
The median time to absolute neutrophil count (ANC) of $0.5 \times 10^9$/L was 14 days (range, 0–20) in the G-CSF arm and 13 days (range, 0–21) in the G+GM arm ($P = 0.05$) as shown in Table 2.

**Toxicity data**
No unexpected toxicities were observed. Commonly experienced toxicities included bone pain and myalgias secondary to cytokine administration, cytopenias related to chemotherapy, and febrile neutropenia. No one experienced grade 4 or 5 toxicity. The most common grade 3 toxicities that were observed are summarized in Table 3a (CTCAE v 3.0). The number of patients who experienced a grade 3 adverse event was similar in the G-CSF and G+GM group (11 versus 13 patients, respectively; $P = 0.3$) (Table 3b).

**Overall and disease-free survival**
After a median follow up time of 14.5 months (range, 0.6–38.5) in the G-CSF arm and 14.0 months (range, 1.1–39.9) in the G+GM arm, the 3-year overall survival (OS) was 85% (95% CI: 69.5%–100%) and 86.6% (95% CI: 76.2%–98.3%), respectively ($P = 0.489$). The 3-year disease-free survival (DFS) was 75.8% (95% CI: 57.9%–99.4%) in the G-CSF arm and 77.1% (95% CI: 65.0%–91.5%) in the G+GM arm as shown in Figure 1.

The OS for patients with intermediate grade NHL was 83.7% (95% CI: 72.2%–97.0%) at 3 years. None of the patients with low grade NHL died. The DFS for patients with intermediate grade NHL was 72.8% (95% CI: 60%–88.4%) at 3 years. None of the patients with low grade NHL had recurrent disease.

**Graft composition**
A total of 54 patients (25 G-CSF and 29 G+GM) provided paired baseline and post-apheresis samples DC subset

| Table 2 Apheresis data | G-CSF | G-CSF + GM-CSF | $P$ value |
|------------------------|-------|----------------|-----------|
| Days from start of chemotherapy to apheresis Median (range) | 17 (12–26) | 14 (13–26) | 0.05 |
| Failure to mobilize | 2 | 3 | 0.67 |
| Failure to collect $4 \times 10^6$ CD34+ cells/kg | 2* | 2 | 0.99 |
| No. of apheresis needed Median (range) | 2 (1–6) | 2 (1–7) | 0.64 |
| CD34+ cells/kg Median (range) | $10.34 \times 10^6$ (0.14–59) | $7.5 \times 10^6$ (0.8–74) | 0.65 |
| ANC $\geq 0.5 \times 10^9$/L after mobilization (days) Median (range) | 14 (0–20) | 13 (0–21) | 0.06 |

**Notes:** *One patient was noncompliant with therapy.
**Abbreviations:** ANC, absolute neutrophil count; G-CSF, granulocyte colony-stimulating factor; GM-CSF, granulocyte-macrophage colony-stimulating factor.

| Table 3a Toxicity data: Grade 3 adverse events (CTCAE v 3.0) | G-CSF | G-CSF/GM-CSF | Total |
|-----------------------------------------------------------|-------|--------------|-------|
| Bone pain | 1 | 0 | 1 |
| Cytokine release syndrome | 0 | 2 | 2 |
| Drug fever | 0 | 1 | 1 |
| Esophagitis | 1 | 0 | 1 |
| Febrile neutropenia | 6 | 7 | 13 |
| Fever/infection without neutropenia | 1 | 1 | 2 |
| Muscle weakness (generalized) | 0 | 1 | 1 |
| Nausea/vomiting | 2 | 0 | 2 |
| Thrombocytopenia | 1 | 0 | 1 |
| Anemia | 2 | 1 | 3 |
| Syncope | 0 | 1 | 1 |
| **Total** | 14 | 14 | 28 |

**Abbreviations:** G-CSF, granulocyte colony-stimulating factor; GM-CSF, granulocyte-macrophage colony-stimulating factor.

| Table 3b Number of patients by maximum grade of adverse event | G-CSF | G+GM | Total |
|-------------------------------------------------------------|-------|------|-------|
| Maximum grade | 3 | 11 | 13 | 24 |
| 2 | 10 | 15 | 25 |
| 1 | 19 | 12 | 31 |
| **Total** | 40 | 40 | 80 |

**Notes:** There appear to be no statistically significant differences between treatment groups with respect to the number of patients with maximum grade adverse event. Fisher’s exact test $P$-value $= 0.2704$. 


analysis. There was no significant difference between the percentages of mDC subsets in the apheresis products in the two arms. However, there was a significant increase in the percentage of pDC subsets in the apheresis product in the G-CSF arm when compared to G+GM ($P = 0.002$); consequently, there was a significant decrease in the ratio of mDC and pDC (mDC/pDC) with G-CSF when compared to G+GM ($P = 0.029$) (Table 4 and Figure 2).

**Discussion**

Although G-CSF and GM-CSF are widely used for stem cell mobilization in normal donors and in patients with hematological malignancies who are to undergo an autologous stem cell transplant, there is still controversy regarding the role of the two cytokines in mobilizing DC cells and their subsets. The clinical impact, of either of the two mobilization strategies in patients undergoing autologous PBSC transplantation is also unclear. Our study shows that both regimens are well tolerated. The most common grade 3 toxicity was febrile neutropenia. The stem cell yield, number of apheresis procedures required to reach a target cell dose of $4 \times 10^6$/kg and the overall and disease free survival were similar in the two groups. Previous studies have shown that GM-CSF when used alone or in combination with G-CSF mobilizes more mature (CD14-/CD80+ or CD86-) cells and precursor dendritic cells when compared to G-CSF alone.5,6,8–12 What is unclear, however, is whether there is polarization of these dendritic cells to myeloid dendritic cells or plasmacytoid dendritic cells and its clinical significance. Several groups have reported polarization to pDC cells in normal donors who received G-CSF for stem cell mobilization.13–15 In contrast Shaughnessy et al did not observe any polarization of DCs by either G-CSF or GM-CSF in normal donors, however donors receiving G-CSF mobilized more mDC and pDC cells.16 Gazitt et al did not find a major difference in the ratio of mDC/pDC cells in the PB of patients with NHL mobilized with cyclophosphamide plus G-CSF compared with patients mobilized with either cyclophosphamide plus GM-CSF or cyclophosphamide plus GM-CSF followed by G-CSF.12 Others have reported polarization to pDC with G-CSF17–18 or no polarization of DC subsets.19 Sanchez et al11 found that in patients with acute myelogenous leukemia who were mobilized with G-CSF alone, there was polarization towards the mDC subset.

Our data show that there was no difference in the total numbers of DC cells mobilized in the two groups; however, we found a polarization towards the pDC subset in

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**Table 4** Changes in the percentages of mDC, pDC and the mDC/pDC pre and post mobilization with G-CSF or GM-CSF

| Median (mean ± SEM) | G-CSF N = 25 | G+GM N = 29 | $P$ value |
|---------------------|--------------|--------------|-----------|
| %mDC                |              |              |           |
| Pre                 | 55.6 (49.9 ± 3.53) | 56.9 (57.2 ± 2.78) | 0.125     |
| Post                | 17.6 (22.5 ± 2.76) | 17.8 (19.3 ± 2.11) | 0.633     |
| %pDC                |              |              |           |
| Pre                 | 18.9 (21.6 ± 2.83) | 22.5 (22.1 ± 1.84) | 0.549     |
| Post                | 7.6 (9.25 ± 1.32) | 4.8 (4.82 ± 0.57) | 0.002     |
| mDC/pDCmDC          |              |              |           |
| Pre                 | 2.67 (4.80 ± 1.47) | 2.68 (3.53 ± 0.52) | 0.924     |
| Post                | 2.52 (3.72 ± 0.80) | 4.02 (7.21 ± 2.30) | 0.029     |

**Abbreviations:** DC, dendritic cells; G-CSF, granulocyte colony-stimulating factor; GM, granulocyte-macrophage; SEM, standard error of mean.
patients mobilized with G-CSF alone. We did not observe a survival difference in the two arms of the study at least with the short follow-up time of 14 months. On the contrary Gazitt et al\textsuperscript{12} reported that there was a survival advantage for patients mobilized with GM-CSF containing regimens. Similarly Dean et al also found that higher numbers of mDC cells and total DCs were significantly associated with improved survival.\textsuperscript{1} These differences may be explained by the different patient populations studied and whether the evaluations were done on the PB or in apheresis products.

**Conclusion**

Mobilization of peripheral blood progenitor cells with a combination of G-CSF plus GM-CSF results in the mobilization of higher numbers of pDC cells. However, this did not seem to have any clinical advantage.

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