Facial Acne: A Randomized, Double-Blind, Placebo-Controlled Study on the Clinical Efficacy of a Symbiotic Dietary Supplement

Fabio Rinaldi · Laura Marotta · Antonio Mascolo · Angela Amoruso · Marco Pane · Giammaria Giuliani · Daniela Pinto

Received: October 25, 2021 / Accepted: December 3, 2021 / Published online: January 21, 2022 © The Author(s) 2022

ABSTRACT

Introduction: Treatments other than topical and systemic antibiotics are needed to restore the dysbiosis correlated with acne onset and evolution. In this view, probiotics and botanical extracts could represent a valid adjunctive therapeutic approach. The purpose of this study was to test the efficacy of a dietary supplement containing probiotics (Bifidobacterium breve BR03 DSM 16604, Lactobacillus casei LC03 DSM 27537, and Ligilactobacillus salivarius LS03 DSM 22776) and botanical extract (lupeol from Solanum melongena L. and Echinacea extract) in subjects with mild to moderate acne over an 8-week study period.

Methods: Monocentric, randomized, double-blind, four-arm, placebo-controlled clinical study involving 114 subjects.

Results: A significant ($p < 0.05$) effect on the number of superficial inflammatory lesions was reported over the study period in the subjects taking the study agent (group II) (−56.67%), the botanical extracts (group III) (−40.00%), and the probiotics (group IV) (−38.89%) versus placebo (−10.00%). A significant ($p < 0.05$) decrease in mean desquamation score, sebum secretion rate, and porphyrin mean count versus baseline was also reported, and the effect was most evident for group II. The analysis of log relative abundance after 4 and 8 weeks of treatment compared with baseline showed a significant ($p < 0.01$) decrease in Cutibacterium acnes and S. aureus, along with a contextually and significant ($p < 0.05$) increase in Staphylococcus epidermidis, especially in group II. No significant changes were reported for group I.

Conclusion: The results from this study suggest that the administration of the dietary supplement under study was effective, safe, and well tolerated in subjects with mild to moderate acne and could represent a promising optional complement for the treatment of inflammatory acne as well as for control of acne-prone skin.

Keywords: Acne; Dietary supplement; Probiotic; Ligilactobacillus salivarius LS03; Lupeol; Echinacea
INTRODUCTION

Acne vulgaris (acne) is chronic inflammatory dermatosis with multifactorial etiology, affecting mainly adolescents [1], although it is also reported in the adult population [2]. It is ranked among the top ten diseases worldwide [3] with almost half of all women aged 21–30 years suffering this condition [4].

Acne is an inflammatory process localized to the pilosebaceous units of the face, chest, arms, and back [5]. The alteration of keratinization within the pilosebaceous unit, resulting in the formation of comedones, increased production of sebum, proliferation of *Cutibacterium acnes*, and perifollicular inflammation has been hypothesized in the pathophysiology of acne [6].

The inflammation covers the entire life cycle of the acne lesion, from onset to resolution, including the formation of closed comedones, inflammatory lesions, post-inflammatory erythema (PIE) and hyperpigmentation, and scarring [7]. There are many common treatments for acne lesions depending on the severity of acne according to the European evidence-based (S3) guidelines for the treatment of acne [8]. Systemic therapy is the first-line choice that can be eventually combined with topical therapy [9]. More specifically, oral antibiotic therapy is recommended for the management of moderate and severe inflammatory acne resistant to topical treatments, but the use of systemic antibiotics should be limited to the shortest possible time, generally 3 months. Commonly used topical therapies include benzoyl peroxide, salicylic acid, retinoids, azelaic acid, antibiotics, and their combinations [8, 10].

Antibiotic and retinoid therapies, both topical and oral, have many limitations: cheilitis; xerosis of the hands and face, including the nasal mucosa; skin fragility; and sensitivity to UV radiation [8]. For instance, isotretinoin and lymecycline treatments have been reported to modify the skin microbiota in acne [11]. Therefore, the use of antibiotics has been linked to modification in both the gut and skin microbiota [12].

A new understanding of the pathophysiology of acne leads to a paradigm shift: *Cutibacterium acnes* is not the only microorganism involved in acne development; other bacteria, mainly *Staphylococcus aureus* and *Staphylococcus epidermidis*, are also implicated [13].

In this view, treatments other than topical and systemic antibiotics are needed to restore the dysbiosis correlated with acne onset and evolution.

The usefulness of probiotics in acne has been reported in many in vitro and in vivo studies [14]. Probiotics have a two-way mechanism in managing acne; firstly, they rebalance the microbiota by preventing the growth of opportunistic bacteria [15]. In some cases, their inhibitory activity is mediated by antibacterial proteins and bacteriocins [14]. Probiotics were
also found to inhibit cytokine interleukin-8 (IL-8) in epithelial cells and to reduce inflammation by controlling the expression of inflammatory cytokines and inhibiting pathogenic CD8 T cells [16, 17]. Among cytokines involved in acne, IL-8 has been reported as the leading proinflammatory mediator in acne.

Recently, Deidda and colleagues reported the bacteriocin-dependent activity and a targeted anti-IL-8 property of the probiotic strain L. salivarius LS03 DSM 22776 [18].

Recently, there has also been growing interest in the use of botanical extract for the management of many inflammatory skin conditions, including acne vulgaris [19, 20]. A meta-analysis of Soleymani et al. [21] investigated the mechanisms of actions of promising plant-derived secondary metabolites in the management of acne vulgaris. One plant metabolite that has been reported to target the main pathogenic features of acne is lupeol, a pentacyclic triterpene, from Solanum melongena L. [22, 23].

Another effective and safe alternative for the treatment of acne is Echinacea, a botanical extract with a long history of use in inflammatory disease due to its recognized anti-inflammatory activity [24].

The purpose of this study was to test the probiotics and botanical extracts-based supplement in a randomized, double-blinded, placebo, and active-controlled, parallel groups (four arms) study to assess the effectiveness of the natural active substances in subjects with mild to moderate acne over an 8-week study period.

**METHODS**

**Subjects**

In total, 112 adult subjects (average age 23 ± 7.15 years) of both sexes with mild to moderate acne were recruited at RS Dermatologic Clinic, Milan, Italy. The main exclusion criteria included current use of any prescription treatment (oral or topical) for acne (2 months washout allowed), pregnancy and lactation, known allergy or hypersensitivity to any of the constituents of the study product, and participation in a similar study in the last 3 months.

**Compliance with Ethics**

All patients were evaluated and enrolled in the study after signing informed consent. Parental consent was obtained in the case of children. This study was approved by the Independent Ethical Committee for Clinical not Pharmacological Investigation of Genoa, Italy and was in accordance with the ethical standards of the Declaration of Helsinki of 1964.

**Study Design**

The study was structured in the form of a randomized double-blinded, placebo, and active-controlled, parallel groups (four arms) study. The study was performed between September 2018 and January 2019. The product under study was a probiotics-based dietary supplement herein referred to as study agent (Primak Integratore, Giuliani S.p.A., Milan, Italy). The supplement was in the form of a DUOCAM sachet containing two different blends: probiotics (Bifidobacterium breve BR03 DSM 16604, Lactis-sebacillus casei LC03 DSM 27537, and Ligilactobacillus salivarius LS03 DSM 22776 and botanical extract (lupeol from Solanum melongena L. and Echinacea extract), respectively.

Specifically, the probiotic chamber included \( \geq 0.5 \times 10^9 \) live cells of B. breve BR03 (DSM 16604), \( \geq 0.5 \times 10^9 \) live cells of L. casei LC03 (DSM 27537), and \( \geq 1.0 \times 10^9 \) live cells of L. salivarius LS03 (DSM 22776) (combined dose of \( \geq 2 \times 10^9 \) live cells), with maltodextrin used as a bulking agent to yield a final weight of 1.5 g. The probiotic sachets were analyzed by Biolab Research S.r.l., Novara, Italy, via flow cytometry [ISO 19344:2015 IDF 232:2015, \( \geq 2 \times 10^9 \) active fluorescent units (AFU)] and plate count method (Biolab Research Method 014-06, \( \geq 2 \times 10^9 \) CFU) to confirm target cell count.

Product stability was monitored to ensure minimum cell counts were maintained (data not shown).
Inserire Composizione Botanicals

Subjects were enrolled after verification of inclusion and exclusion criteria (Table 1) and randomized to one of the four treatment groups: (I) placebo, (II) study agent, (III) botanical extracts, and (IV) probiotics. The ingredients in the placebo were inert.

Subjects were instructed to take the supplements once a day after breakfast for 8 weeks. Subjects were screened at baseline (T0), after 4 weeks of treatment (T1), and at the end of treatment (T2).

Efficacy Endpoints

Efficacy endpoints were evaluated at each post-baseline visit and expressed as the percent change from baseline in facial comedones, papules, pustules, and nodules measured using the global acne grading system (GAGS) score that gives a weight to each region (face and back) with a severity score (1-18, mild; 19-30, moderate; 31-38, severe) [25]. Porphyrins were also evaluated at each timepoint by the VISIA system (Canfield Scientific).

Therefore, erythema, desquamation, sebum level, and microbial dysbiosis were measured. Erythema and desquamation were measured according to a four-point qualitative scale: 0 = absent, 1 = mild 2 = moderate, and 3 = severe. Sebum level was measured by a microcamera equipped with 200× Optic system (Microcamera, APR Instruments s.r.l., Milan, Italy). An area of 1.41 × 1.0 mm was displayed. Sebum value was recorded as low, < 100 μm × cm²; medium, 100–200 μm × cm²; or high, > 200 μm × cm². Microbial dysbiosis was expressed as the relative abundance of bacterial DNA of main bacterial species on the skin: Cutibacterium acnes, Staphylococcus epidermidis, and Staphylococcus aureus [26]. eNAT kit (1 ml eNAT transport and preservation medium and FLOQSwab) (Copan, Brescia, Italy) was used for sampling a 16 cm² area. Samples were stored at 4 °C until DNA extraction with QIAamp UCP Pathogen Mini Kit (Qiagen, Milan, Italy) according to the manufacturer’s protocol and then amplified with microbial polymerase chain reaction (PCR) assay kit (Qiagen, Milan, Italy) with gene-specific primers and TaqMan MGB
probe targeting *C. acnes*, *S. epidermidis*, and *S. aureus* 16S rRNA gene.

**Statistical Analysis**

Statistical analysis was performed using GraphPad statistical software version 13.0 (La Jolla, CA, USA). Change from baseline in the clinical signs of acne, sebum level, hydration, porphyrin production, and microbial dysbiosis were compared between the groups using a t-test (95% confidence interval for the between-group difference).

The sample size was determined as a minimum of 25 evaluable subjects by treatment arm based on the hypothesis that the percentage reduction in clinical signs of acne between treatments would be at least 30% larger than in the placebo arm at the end of the treatment period. Having 25 subjects per group enables the detection of the above effect with a power of > 90% and a significance level of 5%. Therefore, according to sample size calculation, a maximum of 25% of enrolled subjects could drop out during the study period.

**RESULTS**

In total, 114 of the enrolled subjects (95%) completed the study (28 in group I, 30 in group II, 29 in group III, and 27 in group IV). The mean age was 23.5 ± 8.5 years. Six subjects were withdrawn before visit T1. None of the subjects was terminated due to an adverse event. The demographics of the samples are presented in Table 2.

Percent changes from baseline in lesion counts (% of GAGS score reduction) are shown in Fig. 1. At week 4, there was a statistically significant decrease (*p* < 0.05) in the number of superficial inflammatory lesions over the study period in the subjects taking the study agent (group II), the botanical extracts (group III), and the probiotics (group IV) versus placebo (mean values −36.67%, −33.33%, and −31.11% versus −10.00% of placebo, respectively) (Fig. 1). This effect increases from T0 to T2 (mean values −56.67%, −40.00%, and −38.89% versus −18.89% of placebo, respectively (*p* < 0.05).

In general, the % of reduction in GAGS score was most evident for group II (study agent).

At the beginning of the study, the mean erythema score in the groups was 1.82 ± 0.06, which was subsequently reduced to 0.73 ± 0.69, 1.17 ± 0.83, and 1.30 ± 0.79, respectively, for groups II, III, and IV (Fig. 2) after 4 weeks of treatment, but the effect was not significant. After 8 weeks of treatment, a further and significant (*p* < 0.05) reduction of erythema score was reported for groups II–IV (0.27 ± 0.45, 0.50 ± 0.63, and 0.73 ± 0.74, respectively) (Fig. 2). Group II showed the highest reduction (85.24% reduction versus baseline). No reduction of erythema score was reported for group I during the treatment.

A significant (*p* < 0.05) decrease in mean desquamation score versus placebo (group I) was reported for group II after 4 and especially 8 weeks of treatment (Fig. 3) (−33.30% and −55.60%, respectively). A significant

| Table 2 Demographic details at baseline |
|----------------------------------------|
| **Group I** | **Group II** | **Group III** | **Group IV** |
| Age mean ± SEM (years) | 23.70 ± 7.59 | 23.53 ± 7.39 | 23.43 ± 6.61 | 23.43 ± 7.01 |
| Sex, *n* (%) | | | | |
| Male | 57 (16) | 53 (16) | 52 (15) | 48 (13) |
| Female | 43 (12) | 47 (14) | 48 (14) | 52 (14) |
| GAGS score ± SEM | 23.57 ± 5.42 | 24.61 ± 8.41 | 22.98 ± 3.32 | 25.81 ± 6.17 |

*n* number of subjects, *SEM* standard error of the mean
A decrease in desquamation after 8 weeks of treatment was reported for groups III and IV (−43.30% and −33.30%, respectively) (Fig. 3). The highest effect was reported for group II. A slight (−10%) reduction in mean desquamation score was also reported for group I.

Efficacy of the dietary supplements for acne was also measured in terms of reduction in sebum secretion rate from baseline by using a microcamera as presented in Table 3. In subjects treated with the study agent (group II) and the single components (group III and IV), the sebum secretion rate (% of subjects) was significantly reduced at the end of the study period (Table 3).

Porphyrin analysis revealed a significant (p < 0.05) reduction in the mean counts in group II after 8 weeks of use of the study agent (−45.49%) (Fig. 4). The treatment with botanical extracts (group III) produced a significant reduction in the mean porphyrin count after 8 weeks of treatment (−34.25%) compared with baseline (14.37%).

C. acnes, S. epidermidis, and S. aureus are the three major microbial species found on the skin [27, 28].

Relative abundance of predominant bacteria on the face of enrolled subjects was analyzed by mean of quantitative reverse-transcription PCR Real-time quantitative PCR (RT-qPCR). Pan-bacteria-specific targets were used as control. Student’s t-test analysis of log relative abundance (RA) after 4 and 8 weeks of treatment compared with baseline showed a significant (p < 0.01) decrease in C. acnes (from 29.7 to 21.4 and 8.8 log RA, respectively) in group II subjects (Fig. 5A).

A significant (p < 0.01) but lower reduction was also reported in group III (from 25.6 to 22.3 and 17.4 log RA, respectively) and group IV (from 26.8 to 24.2 and 21.7 log RA, respectively).

Also, S. aureus was significantly (p < 0.05) reduced in group II (from 1.4 to 1.1 and 0.2 log RA, respectively) and group III (from 1.8 to 0.9 and 0.7 log RA, respectively) after 4 and especially 8 weeks of treatment (Fig. 5B). No significant changes were reported for S. aureus in
group IV (Fig. 5B). Contextually, a significant ($p < 0.05$) increase in *S. epidermidis* was reported in group II (from 0.6 to 1.2 log RA) and group III (from 1.1 to 1.6 log RA) after 8 weeks of treatment (Fig. 5C).

No significant changes were reported for group I (Fig. 5).

Figure 6 shows a representative photographic evaluation of study agent acne in a volunteer of group II at baseline (Fig. 6A) and after 8 weeks of treatment (Fig. 6B).

**DISCUSSION**

There is growing evidence in support of the use of probiotics, mainly *Lactobacillus* and *Bifidobacterium* strains, as treatment for acne [29]. Their effect on acne pathophysiology can be attributed to their anti-inflammatory properties [30–32] but also to their ability to maintain skin hydration [33] and barrier function [34, 35].

Moreover, probiotics are reported to be well tolerated, and an improvement in user compliance versus other therapies has been reported as well [29]. Interestingly, probiotics can synergize with antibiotics treatment [36] and counteract the side effects deriving from conventional anti-acne treatment such as isotretinoin, as reported by Fabbrocini and colleagues [37]. Another important aspect of acne treatment is preserving the microbiome, both cutaneous and intestinal [12]. Probiotics can rebalance the microbiome by boosting the levels of beneficial bacteria and controlling the growth of *C. acnes*, for example, by secreting bacteriocins [38].

Because traditional acne medications have various side effects, also the use of botanical extracts and secondary metabolites from medicinal plants is currently suggested as an alternative treatment [21].

A recent study by Kwon and colleagues [23] reported lupeol as a useful agent able to target most of the major pathogenic features of acne. Lupeol is reported to decrease the level of...
proinflammatory cytokines, modulate epidermal dyskeratosis (acting on IL-1 alpha, Toll-like receptor 2 (TLR-2), and keratin 16), decrease sebum production, and reduce the synthesis of intracellular lipids via the modulation of the insulin-like growth factor type 1 receptor (IGF-1R)/phosphatidylinositol 3 kinase (PI3K)/Akt/sterol response element-binding protein-1 (SREBP-1) signaling pathway.

Another natural alternative treatment for acne is *Echinacea*, for which safety and efficacy have been reported in vivo for other skin lesions, including in wound healing [39, 40]. Both antimicrobial and anti-inflammatory activity has been attributed to *Echinacea*. An in vitro study reported that *Echinacea* was able to inactivate *C. acnes* and inhibit the derived proinflammatory cascade. *Echinacea* was also reported to exhibit antioxidant activity [39–41], which could be useful to reduce the free radical production in acne. Thus, considering the in vitro and in vivo activities on different

![% REDUCTION OF DESQUAMATION](image)

**Fig. 3** Mean desquamation score group (I) placebo, group (II) study agent, group (III) botanical extracts, and group (IV) probiotics, during the treatment. Data are expressed according to a four-point qualitative scale: 0 = absent, 1 = mild, 2 = moderate, and 3 = severe. Asterisks indicate a significant difference to the control (*p < 0.05; **p < 0.01).

| Visit | Sebum secretion rate (%), n | Group I | Group II | Group III | Group IV |
|-------|-----------------------------|---------|----------|-----------|----------|
|       |                             | 29 (8)  | 10 (3)   | 17 (5)    | 22 (6)   |
| T0    | Low                         | 42 (12) | 47 (14)  | 45 (13)   | 48 (13)  |
|       | Medium                      | 29 (8)  | 43 (13)  | 38 (11)   | 30 (8)   |
|       | High                        | 32 (9)  | 30 (9)   | 38 (11)   | 22 (6)   |
| T1    | Low                         | 50 (14) | 37 (11)  | 41 (12)   | 48 (13)  |
|       | Medium                      | 18 (5)  | 33 (10)  | 21 (6)    | 30 (8)   |
|       | High                        | 32 (9)  | 60 (18)  | 55 (16)   | 63 (17)  |
| T2    | Low                         | 54 (15) | 27 (8)   | 34 (10)   | 33 (9)   |
|       | Medium                      | 14 (4)  | 33 (4)   | 11 (3)    | 4 (1)    |

n = number of subjects

Table 3 Sebum secretion rate (%)
mechanisms involved in acne development reported for probiotics, lupeol, and *Echinacea*, a unique formulation composed of a combination of these was developed. This study compared the efficacy of a dietary supplement containing *B. breve* BR03 DSM 16604, *L. casei* LC03 DSM 27537, *L. salivarius* LS03 DSM 22776, lupeol, and *Echinacea* extract versus placebo, probiotics, or botanical extracts alone over 8 weeks of therapy in subjects with mild to moderate facial acne. It was possible to observe a significant improvement in inflammatory and noninflammatory signs of acne since 4 weeks of treatment, and this effect was further improved after 8 weeks of treatment. The observed effects may be due firstly to the anti-inflammatory and antimicrobial effects of probiotics, especially *L. salivarius* LS03 DSM 22776 [18]. A general higher improvement in acne symptoms was reported in subjects who took a daily oral dose of the complete dietary supplement. There was a greater than 56% reduction in the number of total facial lesions, expressed as GAGS score, after 4 and especially 8 weeks of supplementation with the dietary supplement versus placebo and single-component dietary supplement. A significant reduction also in erythema and desquamation score, porphyrins, and sebum production was reported.

Most interestingly, the dietary supplements containing both probiotics and botanical extracts acted on microbial dysbiosis by controlling the growth of *C. acnes* and *S. aureus* and...
simultaneously stimulating the growth of the most beneficial *S. epidermidis*.

In addition to clinical improvement, this study also reported good compliance with the treatment by the enrolled subjects.

Some limitations apply to the study such as its short-duration intervention, single site, relatively small population, and the possibility that noninflammatory lesions may resolve on their own.

**Fig. 5** Relative abundance of main bacterial species on the face of enrolled subjects by RT qPCR. Log relative abundance of **A** *C. acnes*, **B** *S. aureus*, and **C** *S. epidermidis* in group (I) placebo, group (II) study agent, group (III) botanical extracts, and group (IV) probiotics after 4 and 6 weeks of treatment. Values are presented as mean ± SEM, in duplicate. Asterisks indicate a significant difference to the control (*p* < 0.05; **p** < 0.01)

**Fig. 6** Representative photographic evaluation of Primak Integratore acne in volunteer of group II. Notes: **A** Before treatment. **B** After 4 weeks of treatment
Given that, to date, there are no published studies on the synergistic effects of probiotics, lupeol, and *Echinacea* on acne treatment, based on the results of this study, the combination of the above active ingredients may be a reasonably safe and compliant complementary treatment in mild to moderate acne.

**CONCLUSION**

The results from this study suggest that the administration of a dietary supplement containing *B. breve* BR03, *L. casei* LC03, *L. salivarius* LS03, lupeol, and *Echinacea* extract in subjects with mild to moderate facial acne is safe, well tolerated, and able to reduce total facial lesion count (GAGS score) versus placebo and probiotics and botanical extracts alone after 8 weeks of administration. Secondary endpoints analysis shows that the treatment significantly reduced erythema, desquamation, porphyrins, sebum, and microbial dysbiosis. Thus, the combination of effective active ingredients in a single dietary supplement seems to be a promising optional treatment to be used as an adjuvant for the treatment of inflammatory acne as well as for control of acne-prone skin. Larger randomized, placebo-controlled trials may substantiate our findings.

**ACKNOWLEDGEMENTS**

The authors thank the participants of the study for their cooperation.

**Funding.** This research and the journal’s Rapid Service Fee was funded by Giuliani S.p.A.

**Authorship.** All named authors meet the International Committee of Medical Journal Editors (ICMJE) criteria for authorship for this manuscript, take responsibility for the integrity of the work as a whole, and have given final approval for the version to be published.

**Author Contributions.** F.R. and D.P. conceptualized the study. D.P., L.M., and A.M. contributed to methodology. D.P. worked on the formal analysis. D.P. and F.R. carried out the investigation. D.P., M.P., A.A., L.M., and A.M. wrote the original draft. D.P. and F.R. reviewed and edited the manuscript. F.R. supervised the study. F.R. and G.G. acquired the funding.

**Disclosure.** Fabio Rinaldi serve as a consultant for Giuliani S.p.A., Daniela Pinto, Laura Marotta and Antonio Mascolo are employed by Giuliani S.p.A., Giammaria Giuliani is in the board of director of Giuliani S.p.A., Marco Pane and Angela Amoruso have nothing to disclose.

**Compliance with Ethics Guidelines.** This study was approved by the Independent Ethical Committee for Clinical not Pharmacological Investigation of Genoa, Italy and was in accordance with the ethical standards of the Declaration of Helsinki of 1964.

**Data Availability.** The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

**Open Access.** This article is licensed under a Creative Commons Attribution-NonCommercial 4.0 International License, which permits any non-commercial use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article’s Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by-nc/4.0/.
REFERENCES

1. Heng AHS, Chew FT. Systematic review of the epidemiology of acne vulgaris. Sci Rep. 2020;10:5754. https://doi.org/10.1038/s41598-020-62715-3.

2. Rocha MA, Bagatin E. Adult-onset acne: prevalence, impact, and management challenges. Clin Cosmet Investig Dermatol. 2018;11:59–69. https://doi.org/10.2147/CCID.S137794 (Published 2018 Feb 1).

3. Hay RJ, Johns NE, Williams HC, Bolliger IW, Dellavalle RP, Margolis DJ, et al. The global burden of skin disease in 2010: an analysis of the prevalence and impact of skin conditions. J Investig Dermatol. 2014;134(6):1527–34.

4. Perkins AC, Maglione J, Hillebrand GG, Miyamoto K, Kimball AB. Acne vulgaris in women: prevalence across the life span. J Womens Health. 2012;21(2):223–30.

5. Kang S, Cho S, Chung JH, et al. Inflammation and extracellular matrix degradation mediated by activated transcription factors nuclear factor-kappaB and activator protein-1 in inflammatory acne lesions in vivo. Am J Pathol. 2005;166:1691–9.

6. Layton AM. Optimal management of acne to prevent scarring and psychological sequelae. Am J Clin Dermatol. 2001;2:135–41.

7. Kirck LH. Re-evaluating treatment targets in acne vulgaris: adapting to a new understanding of pathophysiology. J Drugs Dermatol. 2014;13(s7-60).

8. Fabbrocini G, Cacciapuoti S, Masara A, Tedeschi A, Dall’Oglio F, Micali G. Linee Guida SIDEMAST per il trattamento dell’acne lieve e moderata. Pacini Editore Medicina eds. Pisa; 2015:1–12.

9. Kumasaka BH, Odland PB. Acne vulgaris. Topical and systemic therapies. Postgrad Med. 1992;92(5):181–3. https://doi.org/10.1080/00325481.1992.11701492 (PMID: 1409171).

10. Zaenglein AL, Pathy AL, Schlosser BJ, Alikhan A, Baldwin HE, Berson DS, Bowe WP, Graber EM, Harper JC, Kang S, Keri JE, Leyden JJ, Reynolds RV, Silverberg NB, Stein Gold LF, Tollefson MM, Weiss JS, Dolan NC, Sagan AA, Stern M, Boyer KM, Bhushan R. Guidelines of care for the management of acne vulgaris. J Am Acad Dermatol. 2016;74(5):945–73.e33.

11. Kelhäla HL, Aho VTE, Fyrqurist N, Pereira PAB, Kubin ME, Paulin L, Palatsi R, Auvinen P, Tasanen K, Lauerma A. Isotretinoin and tetracycline treatments modify the skin microbiota in acne. Exp Dermatol. 2018;27(1):30–6. https://doi.org/10.1111/exd.13397 (Epub 2017 Sep 14. PMID: 28636791).

12. Lee YB, Byun EJ, Kim HS. Potential role of the microbiome in acne: a comprehensive review. J Clin Med. 2019;8(7):987. https://doi.org/10.3390/jcm8070987 (Published 2019 Jul 7).

13. Dréno B, Dagnelie MA, Khannari A, Corvec S. The skin microbiome: a new actor in inflammatory acne. Am J Clin Dermatol. 2020;21(Suppl 1):18–24. https://doi.org/10.1007/s40257-020-00531-1 (PMID: 32910436; PMCID: PMC7584556).

14. Goodarzi A, Mozafarpoor S, Bodaghabadi M, Mohamadi M. The potential of probiotics for treating acne vulgaris: a review of literature on acne and microbiota. Dermatol Ther. 2020;33(3):e13279. https://doi.org/10.1111/dth.13279 (Epub 2020 Apr 7. PMID: 32266790).

15. Yu Y, Dunaway S, Champer J, Kim J, Alikhan A. Changing our microbiome: probiotics in dermatology. Br J Dermatol. 2020;182(1):39–46.

16. Hacini-Rachinel F, Gheit H, Le Luduec J-B, Dif F, Nancey S, Kaiserlian D. Oral probiotic control skin inflammation by acting on both effector and regulatory T cells. PLoS One. 2009;4:e4903.

17. Muizzuddin N, Maher W, Sullivan M, Schnittert S, Mammonde T. Physiological effect of a probiotic on skin. J Cosmet Sci. 2012;63(6):385–95.

18. Deidda F, Amoroso A, Nicola S, Graziano T, Pane M, Mogna L. New approach in acne therapy: a specific bacteriocin activity and a targeted anti IL-8 property in just 1 probiotic strain, the L. salivarius LS03. J Clin Gastroenterol. 2018;52 Suppl 1. In: Proceedings from the 9th probiotics, prebiotics and new foods, nutraceuticals and botanicals for nutrition & human and microbiota health meeting, held in Rome, Italy from September 10 to 12, 2017, pp S78–S81. https://doi.org/10.1097/MCG.0000000000001053 (PMID: 29782471).

19. Clark AK, Haas KN, Sivamani RK. Edible plants and their influence on the gut microbiome and acne. Int J Mol Sci. 2017;18(5):1070.

20. Magin PJ, Adams J, Heading GS, Pond DC, Smith W. Complementary and alternative medicine therapies in acne, psoriasis, and atopic eczema: results of a qualitative study of patients’ experiences and perceptions. J Altern Complement Med. 2006;12(3):451–7.

21. Soleymani S, Farzazi MH, Zargaran A, Niknam S, Rahimi R. Promising plant-derived secondary metabolites for treatment of acne vulgaris: a mechanistic review. Arch Dermatol Res. 2020;312(1):5–23. https://doi.org/10.1007/s00403-
22. Sharma N, Palia P, Chaudhary A, Shalini, Verma K, Kumar I. A review on pharmacological activities of lupeol and its triterpene derivatives. JDDT. 2020;10(5):325–332. http://jddtonline.info/index.php/jddt/article/view/4280. Accessed 8 Jul 2021.

23. Kwon HH, Yoon JY, Park SY, Min S, Kim YI, Park JY, Lee YS, Thiboutot DM, Suh DH. Activity-guided purification identifies lupeol, a pentacyclic triterpene, as a therapeutic agent multiple pathogenic factors of acne. J Investig Dermatol. 2015;135(6):1491–500. https://doi.org/10.1038/jid.2015.29 (PMID:25647437).

24. Sharma M, Schoop R, Suter A, Hudson JB. The potential use of Echinacea in acne: control of Propionibacterium acnes growth and inflammation. Phytother Res. 2011;25(4):517–21. https://doi.org/10.1002/ptr.3288 (Epub 2010 Sep 9. PMID:20830697).

25. Xu Z, Wang Z, Yuan C, Liu X, Yang F, Wang T, et al. Dandruff is associated with the conjoined interactions between host and microorganisms. Sci Rep. 2016;6:24877. https://doi.org/10.1038/srep24877 (PMID:27172459).

26. Clavaud C, Jourdain R, Bar-Hen A, Tichit M, Bouchier C, Pouradier F, et al. Dandruff is associated with disequilibrium in the proportion of the major bacterial and fungal populations colonizing the scalp. PLoS One. 2013;8(3):e58203. https://doi.org/10.1371/journal.pone.0058203 (Erratum in: PloS One 2013;8(10). PMID:23483996).

27. Pasonen-Seppanen S, Karvinen S, Torronen K, Hytinen JM, Jokela T, Lammi MJ. EGF upregulates whereas TGF-beta downregulates the hyaluronan synthases Has2 and Has3 in organotypic keratinocyte cultures: correlations with epidermal proliferation and differentiation. J Invest Dermatol. 2003;120:1038–44.

28. Jung GW, Tse JE, Guiha I, Rao J. Prospective, randomized, open-label trial comparing the safety, efficacy, and tolerability of an acne treatment regimen with and without a probiotic supplement and minocycline in subjects with mild to moderate acne. J Cutan Med Surg. 2013;17(2):114–22. https://doi.org/10.2310/7750.2012.12026 (PMID:23582165).

29. Barnes J, Anderson LA, Gibbons S, Phillipson JD. Echinacea species (Echinacea angustifolia (DC.) Hall. Echinacea pallida (Nutt.) Nutt. Echinacea purpurea (L.) Moench: a review of their chemistry, pharmacology and clinical properties. J Pharm Pharmacol. 2005;57:929–54.

30. Azad MAK, Sarker M, Wan D. Immunomodulatory effects of probiotics on cytokine profiles. Biomed Res Int. 2018. https://doi.org/10.1155/2018/8063647.