Significantly Lower Case-fatality Ratio of Coronavirus Disease 2019 (COVID-19) than Severe Acute Respiratory Syndrome (SARS) in Hong Kong—A Territory-Wide Cohort Study

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Background. The case-fatality ratios (CFR) of coronavirus disease 2019 (COVID-19) and severe acute respiratory syndrome (SARS) appeared to differ substantially. We aimed to compare the CFR and its predictors of COVID-19 and SARS patients using a territory-wide cohort in Hong Kong.

Methods. This was a territory-wide retrospective cohort study using data captured from all public hospitals in Hong Kong. Laboratory-confirmed COVID-19 and SARS patients were identified. The primary endpoint was a composite endpoint of intensive care unit admission, use of mechanical ventilation, and/or death.

Results. We identified 1013 COVID-19 patients (mean age, 38.4 years; 53.9% male) diagnosed from 23 January to 14 April 2020 and 1670 SARS patients (mean age, 44.4 years; 44.0% male) from March to June 2003. Fifty-five (5.4%) COVID-19 patients and 432 (25.9%) SARS patients had reached the primary endpoint in 30 days. By 30 June 2003, 286 SARS patients had died (CFR, 17.1%). By 7 June 2020, 4 COVID-19 patients had died (CFR, 0.4%). After adjusting for demographic and clinical parameters, COVID-19 was associated with a 71% lower risk of primary endpoint compared with SARS (adjusted hazard ratio, 0.29; 95% confidence interval, 0.21–0.40; \( P < .0001 \)). Age, diabetes mellitus, and laboratory parameters (high lactate dehydrogenase, high C-reactive protein, and low platelet count) were independent predictors of the primary endpoint in COVID-19 patients, whereas use of antiviral treatments was not associated with primary endpoint.

Conclusions. The CFR of COVID-19 was 0.4%. Age and diabetes were associated with worse outcomes, whereas antiviral treatments were not.

Keywords. SARS-CoV-2; COVID-19; nCoV; death; pneumonia.

In late 2002 to mid-2003, an outbreak of severe acute respiratory syndrome (SARS), a newly emerged infectious disease caused by a novel coronavirus SARS-coronavirus (CoV), occurred in southern China [1]. SARS posed an enormous global public health threat in 2003 because 8098 patients were infected worldwide, resulting in 774 deaths in 26 countries, with an overall case fatality ratio (CFR) of 9.6% [2].

Another novel zoonotic coronavirus hit the world in late 2019 [3]. This novel virus, with the official name of SARS-CoV-2, is responsible for causing coronavirus disease 2019 (COVID-19), which has affected more than 180 countries and 30 territories in all 7 continents of the world. COVID-19 has resulted in more than 8.86 million cases and 465 000 deaths worldwide as of 22 June 2020, with a CFR of 5.3% [4]. The CFR related to COVID-19 appeared to be lower than that of SARS.

Because SARS-CoV-2 is closely related to SARS-CoV [5], the exact reasons for the difference in CFRs between COVID-19 and SARS remained obscure. Moreover, because the number or proportion of infections that remained undiagnosed was unknown and varied widely across countries because of differences in testing strategies, the true CFR for COVID-19 was uncertain. With the evolving pandemic, the prediction of CFR and the risk factors of death are important for distribution of healthcare resources. In this study, we aimed to compare the CFR and its predictors of COVID-19 and SARS patients using a territory-wide cohort in Hong Kong.

METHODS

Setting and Study Design
We performed a territory-wide retrospective cohort study using data from the Clinical Data Analysis and Reporting System
Conserved region in the E gene of SARS-CoV and SARS-CoV-2. The reverse transcription PCR (RT-PCR) assay was used to detect a SARS-CoV-2 via RT-PCR. Antigen tests, were performed as appropriate. A real-time reverse transcription PCR (RT-PCR) assay was performed regularly as clinically indicated. Microbiological tests. Systemic corticosteroids were not given routinely, except for selected patients (eg, those with refractory shock). Patients were discharged when they improved clinically and when 2 consecutive clinical specimens tested negative for SARS-CoV-2 via RT-PCR.

**Clinical Evaluation**

The clinical evaluation of SARS patients in 2003 was described in detail in our previous publications [11, 12]. All COVID-19 patients were admitted to medical wards or ICUs with isolation facilities. Initial investigations included a complete blood count (with a differential count), clotting profile (prothrombin time, activated partial-thromboplastin time, international normalized ratio), and serum biochemical measurements (including electrolytes, renal and liver biochemistries, C-reactive protein, lactate dehydrogenase, glucose, and procalcitonin). These laboratory assessments and chest radiography scans were performed regularly as clinically indicated. Microbiological workup, including sputum and blood bacterial culture, nasopharyngeal aspirate for respiratory viruses and atypical pathogens, and urine for Streptococcus pneumoniae and Legionella antigen tests, were performed as appropriate. A real-time reverse transcription PCR (RT-PCR) assay was used to detect a conserved region in the E gene of SARS-CoV and SARS-CoV-2.

**Clinical Management of COVID-19 Patients**

Supportive therapy, including supplemental oxygen, intravenous fluid, vasopressor support, mechanical ventilation, and renal replacement therapy, were given as appropriate. Patients were started on lopinavir-ritonavir (200 mg/50 mg twice daily) monotherapy or in combination with ribavirin (400 mg twice daily) and/or interferon-beta-1b, for up to 14 days, according to local interim guidelines, if antiviral therapy was considered appropriate. Antibacterial therapy was given if bacterial coinfections were suspected clinically or confirmed by microbiological tests. Systemic corticosteroids were not given routinely, except for selected patients (eg, those with refractory shock). Patients were discharged when they improved clinically and when 2 consecutive clinical specimens tested negative for SARS-CoV-2 via RT-PCR.
Continuous variables were expressed in mean ± standard deviation or median (interquartile range [IQR]), as appropriate, while categorical variables were presented as number (percentage). Qualitative and quantitative differences between subgroups were analyzed by χ² or Fisher’s exact tests for categorical parameters and Student t test or Mann-Whitney U test for continuous parameters, as appropriate.

Missing data were assumed missing at random and replaced with substituted values by multiple imputation by chained equations to create 20 complete data sets after the first 10 burn-in iterations [14]. The variables (percentage of missing data) included in the imputation model were age, sex, hemoglobin (1.6%), white blood cell (1.7%), platelet (1.7%), alanine aminotransferase (1.8%), alkaline phosphatase (1.8%), albumin (1.8%), total bilirubin (1.8%), international normalized ratio (25.3%), creatinine (1.8%), urea (1.8%), sodium (2.9%), potassium (2.9%), C-reactive protein (17.0%), lactate dehydrogenase (LDH) (4.1%), and comorbidities at baseline (Supplementary Table 4). The primary endpoint measure and the corresponding Nelson-Aalen estimator of the cumulative hazard at the time of primary endpoint or censoring were also included in the imputation model [15]. Imputed values were constrained within plausible ranges.

Cumulative probabilities of the primary endpoint were estimated by Kaplan-Meier method with 95% confidence interval (CI); log-rank test was used to compare the cumulative probability in COVID-19 and SARS patients. On univariate and multivariable analysis, hazard ratios and adjusted hazard ratios (aHRs) with 95% CI were estimated with the Cox proportional hazard model. We included the following covariates: COVID-19 versus SARS, age, sex, presence of comorbidities, laboratory parameters including baseline complete blood count, liver and renal functions, C-reactive protein, and LDH. The overall coefficient estimates and standard errors were computed by combining the estimates obtained on each individual multiple imputation data set using Rubin’s rules [16]. On multivariable analysis, backward elimination was performed by repeated use of Rubin’s rules to select important covariates [17]. Schoenfeld’s global test was used to test the proportional hazards assumption, which did not detect any significant violations. All statistical tests were 2-sided. Statistical significance was taken as P < .05.

RESULTS

Demographic Characteristics
We identified 1013 COVID-19 patients (all COVID-19 patients reported to the Department of Health from 23 January to 14 April 2020) and 1670 SARS patients (95.2% of all reported SARS patients) from March to June 2003. Among COVID-19 patients, 705 (69.6%) were imported cases and 222 patients (21.9%) were secondary cases of imported and local cases, respectively [18]. The number of COVID-19 patients with coexisting conditions were: diabetes mellitus in 80 (7.9%), cardiovascular disease in 145 (14.3%), chronic liver disease in 37 (3.7%), respiratory disease in 11 (1.1%), and kidney disease in 8 (0.8%). One of them was a healthcare worker. At baseline, compared with patients with SARS, patients with COVID-19 were younger, more likely to be male, and had a lower prevalence of various comorbidities, including cardiovascular diseases and diabetes (Table 1 and Supplementary Table 5).

Hematologic and Biochemical Findings
The initial blood count showed similar prevalence of leukopenia in in 88 (9.0%) of COVID-19 and in 160 (9.6%) of SARS patients, whereas fewer COVID-19 patients had moderate lymphopenia (22.6% vs 55.6%) and thrombocytopenia (11.7% vs 29.4%) on presentation. Prothrombin time remained normal in most cases (Table 1 and Supplementary Table 5).

Serum chemical values were normal in the majority of COVID-19 patients. They had lower serum creatinine level, lower LDH level, and lower C-reactive protein level than SARS patients. The results of laboratory tests performed on presentation are listed in Table 1. The results in a single imputation data set are shown in Supplementary Table 6.

Microbiologic and Virologic Findings
The prevalence of viral coinfections was similar in COVID-19 and SARS patients, whereas bacterial coinfections were less common among COVID-19 patients (Table 1). COVID-19 patients had higher prevalence of rhinovirus/enterovirus, but lower prevalence of influenza A and B. The most common bacterial pathogens isolated from respiratory tract of COVID-19 patients were Staphylococcus aureus and Hemophilus influenzae (Table 2).

Pharmacological Treatment for COVID-19 and SARS Patients
Of the 1013 COVID-19 patients, 372 (36.7%) had received antibiotics, 592 (58.4%) lopinavir-ritonavir, 519 (51.2%) ribavirin, 315 (31.1%) interferon beta, 42 (4.1%) corticosteroid therapy (4 received pulse methylprednisolone), and 2 (0.2%) IVIG. Of the 1670 SARS patients, 1554 (93.1%) had received antibiotics, 1423 (85.2%) ribavirin, 110 (6.6%) lopinavir-ritonavir, 1336 (80.0%) corticosteroid therapy (972 received pulse methylprednisolone), and 74 (4.4%) IVIG (Table 1).

Clinical Outcomes
Fifty-five (5.4%) COVID-19 patients and 432 (25.9%) SARS patients had reached the primary endpoint in 30 days, respectively. Among the 1013 COVID-19 patients, 53 (5.2%) were admitted to the ICU, all because of respiratory failure. Mechanical ventilatory support was required in 22 patients (2.2%). The clinical characteristics of these patients are summarized in Supplementary Table 7. By 7 June 2020, 4 patients had died (CFR, 0.4%); a total of 1006 (99.3%) patients had been
Table 1. Baseline Clinical Characteristics Before Multiple Imputation of Patients With SARS-CoV-2 Infection/COVID-19 or SARS-CoV Infection/SARS

| Baseline clinical characteristics | All N = 2683 | COVID-19 N = 1013 | SARS N = 1670 | P Value |
|----------------------------------|-------------|-------------------|---------------|---------|
| Age, y                           | 42.2 ± 19.5 | 38.4 ± 17.7       | 44.4 ± 20.1   | <.0001  |
| Male, n (%)                      | 1280 (47.7) | 546 (53.9)        | 734 (44.0)    | <.0001  |
| Comorbidities, n (%)             |             |                   |               |         |
| Cardiovascular diseases          | 539 (20.1)  | 145 (14.3)        | 394 (23.6)    | <.0001  |
| Hypertension                     | 493 (18.4)  | 138 (13.6)        | 355 (21.3)    | <.0001  |
| Ischemic heart disease           | 61 (2.3)    | 6 (0.6)           | 55 (3.3)      |         |
| Cardiac dysrhythmias             | 93 (3.5)    | 10 (1.0)          | 83 (5.0)      | <.0001  |
| Heart failure                    | 56 (2.2)    | 2 (0.2)           | 56 (3.4)      |         |
| Digestive diseases               | 272 (10.1)  | 42 (4.1)          | 230 (13.9)    | <.0001  |
| Peptic ulcer                     | 47 (1.8)    | 1 (0.1)           | 46 (2.8)      | <.0001  |
| Chronic liver disease            | 179 (6.7)   | 37 (3.7)          | 142 (8.5)     | <.0001  |
| Liver failure, cirrhosis, or cirrhotic complications | 8 (0.3) | 0 (0) | 8 (0.5) | .028 |
| Biliary disease                  | 24 (0.9)    | 3 (0.3)           | 21 (1.3)      | .010    |
| Gastrointestinal hemorrhage      | 49 (1.8)    | 1 (0.1)           | 48 (2.9)      | <.0001  |
| Diabetes mellitus                | 357 (13.3)  | 80 (7.9)          | 277 (16.6)    | <.0001  |
| Malignant tumor                  | 75 (2.8)    | 13 (1.3)          | 62 (3.7)      | <.0001  |
| Nervous system diseases          | 130 (4.8)   | 13 (1.3)          | 117 (7.0)     | <.0001  |
| Cerebrovascular events           | 87 (3.2)    | 8 (0.8)           | 79 (4.7)      | <.0001  |
| Other nervous system diseases*   | 83 (3.1)    | 5 (0.5)           | 78 (4.7)      | <.0001  |
| Respiratory disease*             | 121 (4.5)   | 11 (1.1)          | 110 (6.6)     | <.0001  |
| Kidney disease                   | 61 (2.3)    | 8 (0.8)           | 53 (3.2)      | <.0001  |
| Human immunodeficiency virus infection | 5 (0.2) | 3 (0.3) | 2 (0.1) | .37 |
| Follow-up duration, d            | 21 (14–28)  | 21 (13–29)        | 21 (15–28)    | .62     |
| Laboratory results               |             |                   |               |         |
| Creatinine, µmol/L              | 77 (64–92)  | 71 (60–84)        | 80 (67–97)    | <.0001  |
| Urea, mmol/L                    | 4.7 ± 4.0   | 4.1 ± 1.5         | 5.1 ± 4.9     | <.0001  |
| Sodium, mmol/L                  | 136.6 ± 4.2 | 138.8 ± 3.0       | 135.4 ± 4.2   | <.0001  |
| Potassium, mmol/L               | 3.8 ± 0.5   | 3.9 ± 0.4         | 3.8 ± 0.6     | <.0001  |
| Albumin, g/L                    | 39.7 ± 5.4  | 41.5 ± 4.9        | 38.6 ± 5.4    | <.0001  |
| ALT, U/L                        | 22 (15–37)  | 22 (15–34)        | 23 (15–39)    | .14     |
| ALP, U/L                        | 64 (52–83)  | 62 (51–75)        | 67 (53–90)    | <.0001  |
| AST, U/L                        | 26 (20–38)  | 26 (21–37)        | 26 (19–39)    | .37     |
| GGT, U/L                        | 46 (28–89)  | 41 (26–62)        | 58 (30–129)   | .00033  |
| Total bilirubin, µmol/L          | 8.9 ± 8.7   | 8.1 ± 4.7         | 9.4 ± 10.3    | <.0001  |
| Total protein, g/L              | 73.7 ± 6.5  | 74.8 ± 5.5        | 73.0 ± 7.0    | <.0001  |
| Haptoglobin, g/L                | 2.2 ± 1.3   | 2.6 ± 1.3         | 2.1 ± 1.2     | .14     |
| LDH, U/Lc                       | 233 (176–355) | 183 (157–225) | 300 (208–423) | <.0001  |
| CRP, mg/dL                      | 3.0 ± 5.2   | 1.4 ± 3.6         | 4.2 ± 5.8     | <.0001  |
| ESR, mm/h                       | 29.6 ± 29.1 | 25.2 ± 23.6       | 31.7 ± 31.3   | <.0001  |
| Prothrombin time, s             | 11.9 ± 3.4  | 12.1 ± 1.1        | 11.9 ± 3.9    | .014    |
| International normalized ratio  | 1.1 ± 0.3   | 1.1 ± 0.1         | 1.1 ± 0.3     | .80     |
| Hemoglobin, g/dL                | 13.4 ± 1.8  | 13.9 ± 1.5        | 13.0 ± 1.8    | <.0001  |
| WCC, x10³/L                     | 6.4 ± 3.1   | 5.7 ± 2.0         | 6.7 ± 3.5     | <.0001  |
| WCC < 3.5 x 10³/L, n (%)         | 248 (9.4)   | 88 (9.0)          | 160 (9.6)     | .63     |
| Lymphocyte, x10³/L              | 1.2 ± 0.7   | 1.5 ± 0.7         | 1.0 ± 0.7     | <.0001  |
| Lymphocyte < 1 x 10³/L, n (%)    | 1146 (43.4) | 219 (22.6)        | 927 (55.6)    | <.0001  |
| Monocyte, x10³/L                | 0.5 ± 0.3   | 0.5 ± 0.2         | 0.5 ± 0.3     | .21     |
| Neutrophil, x10³/L              | 4.5 ± 2.9   | 3.7 ± 1.8         | 5.1 ± 3.2     | <.0001  |
| Eosinophil, x10³/L              | 0.05 ± 0.1  | 0.07 ± 0.1        | 0.04 ± 0.2    | <.0001  |
| Platelet, x10³/L                | 208.1 ± 82.0 | 228.3 ± 75.2 | 196.2 ± 83.5  | <.0001  |
| Platelet < 150 x 10³/L, n (%)   | 603 (22.9)  | 114 (11.7)        | 489 (29.4)    | <.0001  |
| Fasting glucose, mmol/L         | ...         | 5.9 ± 1.8         | ...           | ...     |
| Random glucose, mmol/L          | ...         | 6.1 ± 2.4         | ...           | ...     |
| Ferritin, pmol/L                | ...         | 658 (260–1528)    | ...           | ...     |
| Procalcitonin, ng/mL            | ...         | 0.2 ± 2.0         | ...           | ...     |
Table 1. Continued

| Baseline clinical characteristics | All N = 2683 | COVID-19 N = 1013 | SARS N = 1670 | PValue |
|---------------------------------|-------------|------------------|--------------|--------|
| Coinfections during follow-up, n (%)\a |             |                  |              |        |
| Viral                           | 59 (2.2)    | 22 (2.2)         | 37 (2.2)     | .94    |
| Bacterial                       | 139 (5.2)   | 19 (1.9)         | 120 (7.2)    | <.0001 |
| Hypoxemia during follow-up, n (%)| 401 (14.9)  | 53 (5.2)         | 348 (20.8)   | <.0001 |
| NSAID during follow-up, n (%)    | 116 (4.3)   | 38 (3.8)         | 78 (4.7)     | .26    |

\aRespiratory system disease was defined by ICD-9-CM diagnosis codes for pneumonia other than SARS-related pneumonia (ICD-9-CM codes: 480–487.0) in previous 3 months, chronic obstructive pulmonary disease and allied conditions (ICD-9-CM codes: 490–496), pneumoconioses and other lung diseases due to external agents (ICD-9-CM codes: 500–508) in previous 3 months, and other diseases of respiratory system (ICD-9-CM codes: 510–519) in previous 3 months.

\bOther nervous system disease was defined by ICD-9-CM diagnosis codes for inflammatory diseases of the central nervous system (ICD-9-CM codes: 320–327), hereditary and degenerative diseases of the central nervous system (ICD-9-CM codes: 330–337), and other disorders of the central nervous system (ICD-9-CM codes: 340–345).

\cPresented in median (interquartile range).

\dBased on nasopharyngeal-aspirate reverse transcription-polymerase chain reaction and/or serology, sputum culture, and blood culture.

discharged [18]. By 7 June 2020, among the 53 patients admitted to the ICU, 1 (1.9%) remained in the ICU, 2 (3.8%) had been discharged from the ICU but remained in the hospital, 48 (90.6%) had been discharged from the hospital, and 2 (3.8%) had died.

Of the 1670 SARS patients, 333 (19.9%) were admitted to the ICU within 30 days. Invasive mechanical ventilatory support was required in 61 (3.7%) patients, whereas 43 (2.6%) required noninvasive ventilator support. By 30 June 2003, 286 patients had died (CFR, 17.1%). Among those admitted to the ICU, 136 (40.8%) patients had died. Supplementary Table 8 shows the comparisons between COVID-19 and SARS patients who developed the primary endpoint.

Factors Predictive of ICU Admission, Invasive Mechanical Ventilation, and Death in COVID-19 Patients

Supplementary Table 9 shows the comparisons between patients who developed and did not develop the primary endpoint. Univariate analysis showed that SARS, advanced age, male gender, comorbidities, laboratory parameters, coinfections, and treatment used for the viral infections were significant predictive factors for the primary endpoint (Table 3). On multivariable analysis, COVID-19 was associated with 71% lower risk of primary endpoint compared with SARS (aHR 0.29; 95% CI, 0.21–0.40; P < .0001; Figure 1). Other factors that were predictive of an adverse outcome included advanced age (aHR per year 1.01; 95% CI, 1.01–1.02; P = .00015), male (aHR 1.24; 95% CI, 1.02–1.50; P = .027), diabetes mellitus (aHR 2.14; 95% CI, 1.74–2.63; P < .0001), and bacterial or viral coinfection (aHR 1.74; 95% CI, 1.36–2.22; P < .0001) (Table 3).

Factors Predictive of ICU Admission, Mechanical Ventilation, and Death in COVID-19 Patients

Five predictive factors were identified for adverse outcome on multivariable analysis: age (aHR 1.02; 95% CI, 1.00–1.04; P = .040), diabetes mellitus (aHR 3.21; 95% CI, 1.72–6.00; P = .00026), baseline LDH level (aHR per 100 U/L 1.48; 95% CI, 1.15–1.91; P = .0027), C-reactive protein (aHR 1.07; 95% CI, 1.02–1.12; P = .0065), and platelet count (aHR 0.995; 95% CI, .991–1.000; P = .031) (Table 4).

DISCUSSION

This is the first report to compare the clinical outcomes of COVID-19 patients and SARS patients with detailed patient-level data. Among our cohort of the first 1013 COVID-19 patients in Hong Kong, the CFR was 0.4%, and 5% had ICU admission or death within 30 days of hospital admission. Among

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patients admitted to the ICU, 4% died. Age, diabetes, and biochemical laboratory parameters were predictive of adverse outcomes, whereas none of the antiviral treatments were associated with clinical outcomes. COVID-19 was associated with 71% lower risk of adverse outcomes compared with SARS.

In our territory-wide cohort of COVID-19 patients in Hong Kong, 5% required ICU care and CFR was 0.4%. At the time of our analysis, 99.3% of our patients had already been discharged from the hospital or had died. Therefore, our observed CFR should be close to the true CFR of COVID-19 patients, with minimal risk of biases from preferential selection of severe cases and delayed reporting of deaths, as in studies performed in the early phase of novel disease epidemics [21].

Observed and estimated CFRs of COVID-19 across different populations around the world have varied greatly. Among the first 82,719 laboratory-confirmed cases in China, the overall CFR was 5.65% [22]. However, the CFR was much higher in Hubei province (5.9%–7.7%) than in other provinces outside Hubei (0.86%–0.98%) [22, 23]. The average CFR in Italy until March 2020 was 7.5%, with CFRs in different regions ranging from 3.1% to 16.7% [24]. Until early April 2020, the CFR in the United States was 3.2%, ranging from 0.7% to 5.7% among different states [25].

There are several reasons for the large differences between CFRs observed in different countries or cities. Accessibility to medical care and national strategies for testing and case identification are possibly the major factors causing differences in reported CFRs. Countries with scarce resources in proactive contact tracing and identification of milder or asymptomatic cases would inevitably see a falsely low denominator in estimating the CFR, leading to a falsely high CFR [26, 27]. As explained previously, our observed CFR likely reflected closely the true CFR of all asymptomatic and symptomatic patients with COVID-19. Another situation in which a complete dataset was available for all diagnosed patients was from the Diamond Princess cruise ship, in which an age-adjusted CFR of 0.5% was observed, which was very similar to our observation [26].

Another likely reason for the much lower CFR in Hong Kong than that in many other countries was the lower incidence of COVID-19 and the lower burden on surge capacity of our healthcare system. Italy, for example, reported a linear negative correlation between CFR and ICU admission rate among different provinces, indicating higher mortality being associated with absence of ICU care because of the operational capacity of ICUs being exceeded [24]. Mortality among patients admitted to ICU in other cities ranged from 26% to 62% early in the pandemic [28–30]. In our cohort, among those admitted to ICU, only 42% received mechanical ventilation and the CFR was only 4%, implying that patients who were less critically ill were admitted to the ICU for close monitoring. These findings highlight the importance of public health measures to "flatten the curve" in preventing depletion of hospital resources and direct impact on patient mortality [19, 31].

Differences in host determinants of cellular entry and other molecular mechanisms of pathogenesis may result in variations in genetic susceptibility among different ethnic groups to this infection [32], although large variations in CFRs observed

### Table 2. Nasopharyngeal-Aspirate (NPA), Sputum, and Blood Sample Results

|                     | COVID-19 N = 1013 | SARS N = 1670 | P Value |
|---------------------|-------------------|---------------|---------|
| Nasopharyngeal-aspirate RT-PCR and/or serology |                     |               |         |
| Influenza A         | 0 (0%)            | 20 (1.2%)     | .00047  |
| Influenza B         | 0 (0%)            | 10 (0.6%)     | .017    |
| Parainfluenza viruses 1–3 | 5 (0.5%) | 5 (0.3%) | .52 |
| Respiratory syncytial virus | 4 (0.4%) | 1 (0.1%) | .071 |
| Rotavirus           | 0 (0%)            | 0 (0%)        | -       |
| Adenovirus          | 6 (0.6%)          | 4 (0.2%)      | .19     |
| Rhinovirus/enterovirus | 13 (1.3%) | 0 (0%)      | <.0001  |
| Metapneumovirus     | 1 (0.1%)          | 0 (0%)        | .38     |
| Sputum culture      |                   |               |         |
| Haemophilus influenza | 5 (0.5%)   | 24 (1.4%)     | .022    |
| Haemophilus parainfluenza | 0 (0%) | 2 (0.1%) | .53 |
| Klebsiella pneumonia | 1 (0.1%)  | 8 (0.5%)      | .17     |
| Klebsiella species  | 0 (0%)            | 17 (1.0%)     | .0013   |
| Streptococcus pneumonia | 0 (0%) | 10 (0.6%) | .017   |
| Streptococcus—Group G | 0 (0%)     | 1 (0.1%)   | 1.00    |
| Methylillin-resistant Staphylococcus aureus (MRSA) | 0 (0%) | 40 (2.4%) | <.0001 |
| Staphylococcus aureus | 6 (0.6%) | 10 (0.6%) | .98    |
| Coagulase –ve Staphylococci | 0 (0%) | 2 (0.1%) | .53    |
| Blood culture       |                   |               |         |
| Klebsiella pneumonia | 0 (0%)          | 1 (0.1%)      | 1.00    |
| Staphylococcus aureus | 0 (0%) | 1 (0.1%) | 1.00 |
| Staphylococcus albus | 0 (0%) | 6 (0.4%) | .089 |
| Staphylococcus epidermidis | 0 (0%) | 2 (0.1%) | .53 |
| Staphylococcus haemolyticus | 0 (0%) | 1 (0.1%) | 1.00 |
| Staphylococcus hominis | 1 (0.1%) | 0 (0%) | .38 |
| Coagulase –ve Staphylococcus | 1 (0.1%) | 0 (0%) | .38 |

Qualitative and quantitative differences between subgroups were analyzed by χ2 or Fisher’s exact tests for categorical parameters, as appropriate.

Abbreviations: COVID-19, coronavirus disease 2019; RT-PCR, reverse transcription-polymerase chain reaction; SARS, severe acute respiratory syndrome.
within the same country could not be explained by genetic factors alone [22, 24].

Practice in prescribing medications with presumed antiviral activity varies greatly among different countries. However, to date, although antivirals like remdesivir or interferon beta, have been shown to reduce time to recovery and viral clearance [33, 34], there is no antiviral agent proven to reduce mortality in randomized clinical trials. In our own cohort, use of lopinavir-ritonavir, ribavirin, and interferon therapies was not associated with a reduced risk of adverse clinical outcomes both in univariate and multivariable analyses. Therefore, the use or nonuse of various antiviral agents is unlikely to cause significant differences in CFRs among different countries.

Patients with COVID-19 had a 71% lower risk of adverse clinical outcomes than patients with SARS in 2003 in Hong Kong. Differences in host characteristics partly accounted for this difference because patients with COVID-19 were generally younger and had fewer comorbidities. However, because the association of SARS with adverse clinical outcomes persisted after adjustment of host characteristics and use of antiviral and steroid treatment, higher virulence of SARS-CoV than SARS-CoV-2 is the most possible reason for the lower CFR observed in COVID-19. Although 1.2% of COVID-19 patients were asymptomatic and 81% had mild infections [35], asymptomatic or subclinical infections were rare in SARS, as shown in seroprevalence studies [36]. Genomic differences, particularly amino acid substitutions concentrated in 2 nonstructural proteins and spike protein, might explain the differences in pathogenicity between the 2 viruses [37, 38]. Moreover, Hong Kong had been the major epicenter of the SARS outbreak in 2003 [1]. This explained the similar number of COVID-19 and SARS patients in our cohort, despite the global number of COVID-19 patients far exceeding that of SARS. The huge burden on hospital, and particularly ICU, resources thus partly explains the higher CFR of SARS patients in Hong Kong (17.0%), compared with the global CFR (9.5%) in 2003 [27].

We have identified older age, diabetes, higher LDH, higher C reactive protein, and lower platelet count as independent predictors of adverse outcomes among COVID-19 patients. All of these variables have been identified as risk factors for mortality or adverse outcomes in other cohorts [39–42]. Nevertheless, different host characteristics and laboratory parameters were identified from various cohorts around the world as predictors of adverse outcomes. These differences may possibly be due to variations in disease severity and ethnic differences. For

| Parameters                                      | Univariate Analysis | Multivariable Analysis |
|-------------------------------------------------|---------------------|------------------------|
|                                                 | HR (95% CI)         | P Value                | aHR (95% CI)   | P Value                |
| COVID-19 (vs SARS)                              | 0.21 (1.16–28)      | <.0001                 | 0.32 (2.33–44) | <.0001                 |
| Age, y                                          | 1.03 (1.03–1.04)    | <.0001                 | 1.01 (1.00–1.02) | .00022                |
| Male                                           | 1.38 (1.15–1.64)    | .00047                 | 1.23 (1.02–1.49) | .030                  |
| Hemoglobin                                      | 0.85 (1.81–88)      | <.0001                 | ...             | ...                   |
| White blood cell                                | 1.10 (1.08–1.11)    | <.0001                 | ...             | ...                   |
| Platelet                                        | 0.997 (1.996–998)   | <.0001                 | 0.997 (1.996–998) | <.0001                |
| Alanine aminotransferase                        | 1.002 (1.001–1.003) | <.0001                 | ...             | ...                   |
| Albumin                                         | 0.90 (1.89–91)      | <.0001                 | 0.97 (95–99)    | .0011                 |
| Total bilirubin                                 | 1.01 (1.01–1.02)    | <.0001                 | 0.99 (1.98–1.00) | .029                  |
| International normalized ratio                  | 1.56 (1.29–1.90)    | <.0001                 | ...             | ...                   |
| Creatinine                                      | 1.002 (1.001–1.002) | <.0001                 | ...             | ...                   |
| C-reactive protein                              | 1.08 (1.07–1.08)    | <.0001                 | 1.04 (1.03–1.06) | <.0001                |
| Lactate dehydrogenase (per 100 U/L)             | 1.21 (1.19–1.24)    | <.0001                 | 1.11 (1.08–1.15) | <.0001                |
| Neutrophil-to-lymphocyte ratio                  | 1.06 (1.06–1.07)    | <.0001                 | 1.02 (1.01–1.03) | .032                  |
| Circulatory system disease                      | 3.55 (2.97–4.24)    | <.0001                 | ...             | ...                   |
| Digestive system disease                        | 1.89 (1.49–2.38)    | <.0001                 | ...             | ...                   |
| Diabetes mellitus                               | 4.30 (3.58–5.18)    | <.0001                 | 2.13 (1.74–2.61) | <.0001                |
| Malignant tumor                                 | 2.83 (2.02–3.97)    | <.0001                 | ...             | ...                   |
| Nervous system disease                          | 1.91 (1.41–2.60)    | <.0001                 | 0.48 (0.34–0.68) | <.0001                |
| Respiratory disease                             | 2.33 (1.73–3.13)    | <.0001                 | ...             | ...                   |
| Chronic kidney disease                          | 3.31 (2.33–4.72)    | <.0001                 | ...             | ...                   |
| Bacterial or viral coinfection                  | 3.17 (2.51–4.00)    | <.0001                 | 1.71 (1.34–2.20) | <.0001                |
| Hypoxia during follow-up                        | 2.52 (2.08–3.06)    | <.0001                 | ...             | ...                   |
| Lopinavir-ritonavir during follow-up            | 0.33 (1.25–44)      | <.0001                 | ...             | ...                   |
| Ribavirin during follow-up                      | 0.78 (1.64–95)      | .013                   | 0.44 (1.35–56)  | <.0001                |
| Steroid during follow-up                        | 1.45 (1.21–1.74)    | <.0001                 | ...             | ...                   |

Abbreviations: aHR, adjusted hazard ratio; CI, confidence interval; COVID-19, coronavirus disease 2019; HR, hazard ratio; SARS, severe acute respiratory syndrome.
example, although all confirmed patients were hospitalized in Hong Kong, fewer than 40% of all confirmed patients were hospitalized in a state in United States, and disease severity varies between ethnic groups [40].

In particular, diabetes carried a 3-fold higher risk of adverse clinical outcomes in COVID-19 in our cohort and was an independent predictor of adverse outcomes in both COVID-19 and SARS. Among 18 571 COVID-19 patients in the United States, diabetes was more prevalent in those requiring ICU admission (32%) and hospitalization (24%) than those managed as outpatients (6%) [43]. In a cohort of 1099 COVID-19 patients in China, diabetes was also more prevalent in those with ICU admission, mechanical ventilation, or death (27% vs 6%) [44]. Glycosylated hemoglobin had a linear correlation with inflammation, hypercoagulability, and hypoxia in COVID-19 patients [45]. Diabetes is associated with impaired innate immunity, and pneumonia is an increasingly important cause of mortality in patients with diabetes [46]. Diabetes predisposed patients to cardiac and renal injuries, which were common complications in 45%–71% and 57%–88% of patients with severe or fatal disease, respectively [47, 48]. Diabetes also induces expression of angiotensin-converting enzymes in lung, liver, and heart tissues. Activation of angiotensin 1 and 2 receptors in diabetes may enhance pro-inflammatory cytokine responses, thereby increasing the risk of acute respiratory distress syndrome in COVID-19. Effective control of glucose and blood pressure in COVID-19 patients may help to reduce local inflammatory response and dampen the acute effects of viral infection [49].

Our study has provided important outcome data that facilitate the clinical management, quarantine arrangement, and the resource allocation amid the COVID-19 outbreak [50]. The established risk factors, namely advanced age and presence of comorbidities, facilitate early identification of population at risk, so that such people should strictly adopt social distancing or even staying at home [51, 52]. We are still in the middle of an ongoing outbreak worldwide and we have to identify hospitalized patients at risk of deterioration as soon as possible based on these risk factors [39].

The strength of our study includes a territory-wide cohort that covers about 90% of the inpatient service and essentially all the SARS and COVID-19 cases in Hong Kong. Data from real-life cohorts represent a wider spectrum of patients such that the findings from real-life cohorts are thus more readily applicable to routine clinical practice. Our study has a few limitations. First, the mean age of COVID-19 patients in our cohort was generally younger than other reported hospitalized cohorts [39, 40]. This was largely because of the large proportion of imported cases, who were younger patients involved in international travelling. Therefore, our results may not be extrapolated to regions with predominantly local transmission. Second, we missed 85 of 1755 (4.8%) of SARS patients in 2003 because of diagnosis coding. Nonetheless, we believe missing less than 5% of the patients does not have major impact on the findings because the proportion of deaths in our cohort (286/1670; 17.1%) was consistent with what was reported officially in 2003 (299/1755; 17.0%). Third, COVID-19 and SARS patients might have been different in terms of the baseline clinical characteristics (eg, age, gender, comorbidities) such that our study might be subjected to confounding as in other observational studies. Therefore, we applied multivariable adjustment on important baseline characteristics. Fourth, missing data on laboratory measurements might lead to biases as in other retrospective studies, though these biases can partially be compensated for by our respectable cohort size. Missing data were rare for common laboratory parameters because they are regularly checked in our routine clinical practice. Yet, some less common laboratory parameters, such as troponin, ferritin, or procalcitonin, might not be checked for every single patient because of minor variations of clinical practice in different hospitals. Multiple imputation with 20 imputed data sets was used to reduce the possible selection bias resulting from missing data [53]. Fifth, ascertainment bias may affect the reliability of the study because of inaccurate entry of certain diagnosis codes for comorbidities, namely diabetes mellitus and cardiovascular disease. We minimized this bias by including laboratory as well as medication data for certain diagnoses (diabetes mellitus, hypertension).

In conclusion, CFR of COVID-19 observed in Hong Kong was 0.4%. COVID-19 was associated with an ~71% lower risk of
Table 4. Univariate and Multivariable Analysis with Cox Proportional Hazard Model on Factors Associated with Primary Endpoint (A Composite Endpoint of Intensive Care Unit Admission, Use of Invasive Mechanical Ventilation, and Death) Among COVID-19 Patients After Multiple Imputation

| Parameters                           | Univariate Analysis | Multivariable Analysis |
|--------------------------------------|---------------------|------------------------|
|                                      | HR (95% CI)         | PValue                 | aHR (95% CI)         | PValue                 |
| Age, y                               | 1.05 (1.04–1.07)    | <.0001                 | 1.02 (1.00–1.04)     | .040                   |
| Male                                 | 1.63 (1.94–2.85)    | .084                   | ...                  | ...                    |
| Hemoglobin                           | 0.88 (1.74–1.03)    | .12                    | ...                  | ...                    |
| White blood cell                     | 1.14 (1.02–1.27)    | .023                   | ...                  | ...                    |
| Platelet                             | 0.99 (1.99–1.00)    | .0065                  | 0.995 (0.991–1.000)  | .031                   |
| Alanine aminotransferase             | 1.01 (1.00–1.02)    | .0030                  | ...                  | ...                    |
| Albumin                              | 0.87 (0.83–0.90)    | <.0001                 | ...                  | ...                    |
| Total bilirubin                      | 1.02 (1.96–1.07)    | .31                    | ...                  | ...                    |
| International normalized ratio       | 2.14 (1.46–9.97)    | .33                    | ...                  | ...                    |
| Creatinine                           | 1.02 (1.01–1.03)    | .00020                 | ...                  | ...                    |
| C-reactive protein                   | 1.14 (1.11–1.17)    | <.0001                 | 1.07 (1.02–1.12)     | .0065                  |
| Lactate dehydrogenase (per 100 U/L) | 2.17 (1.34–3.52)    | .0031                  | 1.48 (1.15–1.91)     | .0027                  |
| Neutrophil-to-lymphocyte ratio       | 1.24 (1.18–1.30)    | <.0001                 | ...                  | ...                    |
| Circulatory system disease           | 5.62 (3.31–9.54)    | <.0001                 | ...                  | ...                    |
| Digestive system disease             | 3.48 (1.58–7.70)    | .0020                  | ...                  | ...                    |
| Diabetes mellitus                    | 10.06 (5.90–17.15)  | <.0001                 | 3.21 (1.72–6.00)     | .00026                 |
| Malignant tumor                      | 2.88 (1.70–11.83)   | .14                    | ...                  | ...                    |
| Nervous system disease               | 1.41 (1.20–10.19)   | .73                    | ...                  | ...                    |
| Respiratory disease                  | 5.90 (1.84–18.89)   | .0028                  | ...                  | ...                    |
| Chronic kidney disease               | 5.47 (1.33–22.46)   | .018                   | ...                  | ...                    |
| Bacterial or viral coinfection       | 1.41 (1.44–4.52)    | .56                    | ...                  | ...                    |
| Hypoxia during follow-up             | 2.22 (1.95–5.17)    | .066                   | ...                  | ...                    |
| Lopinavir-ritonavir during follow-up | 0.95 (1.56–1.62)    | .65                    | ...                  | ...                    |
| Ribavirin during follow-up           | 1.11 (1.65–1.88)    | .71                    | ...                  | ...                    |
| Interferon beta during follow-up     | 1.14 (1.66–1.99)    | .64                    | ...                  | ...                    |
| Steroid during follow-up             | 4.02 (1.90–8.51)    | .0003                  | ...                  | ...                    |

Abbreviations: aHR, adjusted hazard ratio; CI, confidence interval; COVID-19, coronavirus disease 2019; HR, hazard ratio.

adverse clinical outcomes compared with SARS. Patients with risk factors, namely advanced age and presence of diabetes, would be at much higher risk of death and ICU admission. In view of the ongoing outbreak worldwide, we have to identify patients at risk of deterioration as soon as possible based on these risk factors. Health authorities should allocate adequate resources, in particular intensive care facilities, based on the trajectories of the numbers of confirmed cases and well ahead to avoid collapse of the healthcare systems.

Supplementary Data

Supplementary materials are available at Clinical Infectious Diseases online. Consisting of data provided by the authors to benefit the reader, the posted materials are not copypedited and are the sole responsibility of the authors, so questions or comments should be addressed to the corresponding author.

Notes

Author contributions. All authors were responsible for the study concept and design. G. W., T. Y., Y.-K. T., and G. L. were responsible for the acquisition and analysis of data, had full access to all of the data in the study, and take responsibility for the integrity of the data and the accuracy of the data analysis. All authors were responsible for the interpretation of data, the drafting, and critical revision of the manuscript for important intellectual content.

Potential conflicts of interest. G. L. has served as an advisory committee member for Gilead, Merck, and GSK; speaker for Merck and Gilead; and received research grant from Gilead, Merck, and GSK. T. Y. has served as an advisory committee member and a speaker for Gilead Sciences. V. W. has served as an advisory committee member for 3V-BIO, AbbVie, Allergan, Boehringer Ingelheim, Echosens, Gilead Sciences, Intercept, Janssen, Novartis, Novo Nordisk, Perspectum Diagnostics, Pfizer, TARGET-NASH, and Terns, and a speaker for Bristol-Myers Squibb, Echosens, Gilead Sciences, and Merck; he has also received a research grant from Gilead Sciences. H. C. is an advisor for AbbVie, Aptorum, Arbutus, Hepion, Intellia, Janssen, Gilead, GSK, GRAIL, Medimmune, Merck, Roche, Vaccitech, VenatoRx, and Vir Biotechnology, and a speaker for Mylan, Gilead, and Roche. D. H. has served as an advisory committee member for Roche. G. W. has served as an advisory committee member for Gilead Sciences; as a speaker for AbbVie, Abbvie, Bristol-Myers Squibb, Echosens, Furui, Gilead Sciences, Janssen, and Roche; and received research grant from Gilead Sciences. All other authors declare that they have no competing interests. All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

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