Modern concept of biological identification of selenoproteins

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Introduction

Implementation of the influence of the microelement selenium on biological systems is mediated by the inclusion of this element to the aminoacid residue of selenocysteine, which is an integral part of all selenoproteins. About 25 types of selenoproteins are known today, most of them are regulators of oxidative processes (Addinsall et al., 2018) and redox homeostasis (Wrobel et al., 2016). High metabolic activity of selenoproteins, their participation in the pathogenetic links of processes such as oxidative stress (Zachara, 2015), carcinogenesis (Guerrero et al., 2014; Short & Williams, 2017; Bertz et al., 2018), neurodegeneration (Pitts et al., 2014; Bark et al., 2014; Bouldhazar et al., 2016), insulin resistance (Prevost et al., 2013; Mita et al., 2017), myopathy (Moghadaszadeh et al., 2013), and others underscores the constant interest of the scientific community in the issues of the disclosure of mechanisms for the realization all biological functions of these compounds. (Brigelius-Flohé, 2015). In addition to the generally known glutathione peroxidase, which performs the catalytic function in the reduction of hydroperoxides and lipidperoxides, other selenoproteins have been partially investigated. Thioredoxin reductase is one of them, it exists in three forms TrxR1, TrxR2, TGR (cytosolic, mitochondrial, and in the testicles, respectively). The thioredoxin system is extremely important for mammals, as evidenced by the early embryonic mortality of mice that lacked any of the isoforms (Bobba et al., 2014). Another enzyme including selenocysteine residue, iodothyronine deiodinase, plays an important role in the activation and inactivation of thyroid hormones (Hernandez & Stohn, 2018). Selenoprotein synthetase is a mammalian selenium enzyme that provides a synthesis of other selenocompounds (Na et al., 2018; Tobe & Mihara, 2018). Methionine-R-sulfoxide reductase participates in protection against oxidative stress (Gladyshev, 2014; Kim et al., 2014). There is a genetic nomenclature of selenoproteins. By the last nomenclature, all proteins that include selenocysteine are divided into proteins with a known function. These are thioredoxin reductase 1, thioredoxin reductase 2, and thioredoxin reductase 3; glutathione peroxide 1, glutathione peroxide 2, glutathione peroxide 3, glutathione peroxide 4 and glutathione peroxide 6; iodothyronine deiodinase 1, iodothyronine deiodinase 2 and iodothyronine deiodinase 3; methionine sulfoxide reductase B1, and selenoprotein synthetase 2. Designation of selenoproteins with unknown and unexplored functions by the proposed nomenclature have the following abbreviations; SELENOP (sele-nopein K, SELK), SELENON (sele-nopein N, SEPN1, SELN), SELENOO (sele-nopein O, SELO), SELENOV (sele-nopein P, SEPO, SEPP1, SELP), SELENO (sele-nopein S, SELS, SEPS1, VIMP), SELENOT (sele-nopein T, SELT), SELENOL (sele-nopein V, SELV), and SELENOW (sele-nopein W, SELW, SEPW1). This system was agreed upon by the HUGO Genes
Commodity Committee (Gladyshev et al., 2016). This article is an attempt to systematize the general information of biological effects and the role of newly identified selenoproteins with unexplored functions in pathophysiological and biochemical processes. The data, published in recent years, were analyzed. The main task of the work is to find out promising directions in the issue of authentication of selenoproteins, which in turn will help determine the vectors of subsequent studies of this group of proteins.

**General characteristics of selenoproteins**

Trace element selenium (Se) is part of the polypeptide chain of 21 natural amino acids selenocysteine (Sec), which encoded UGA codon. Inclusion Sec in any protein defines the affinity of the last one to selenoproteins. Selenoproteins have definite functions, and their synthesis is dependent on the specific co-factors and dietary Se. It is known that many pathological conditions are associated with changes in the elaboration of selenoproteins and their activity. Emerging evidence points out that selenium deficiency or mutation and polymorphism in genes of selenoproteins is part of the pathogenesis of many diseases, including cardiovascular disorders, immune dysfunction, cancer, muscular and skeletal defects, endocrine dysfunction and neurological maladies (Zoidis et al., 2018).

Selenoproteins with unknown and unexplored functions can be divided into groups according to:

- Localization:
  - in the endoplasmic reticulum membrane (Sel N, SelK, SelT);
  - in the endoplasmic reticulum lumen (SelF, SelR, SelM, SelI);
  - in the nucleus (SelH, SelR or methionine-R-sulfoxyd
    rucase 1);
  - in the cytoplasm (SelP, SelW, SelR, SelI);
  - location is unknown (SelO, SelU, SelV, Fig. 1).

- Biofunctional role:
  - regulatory of oxidative stress;
  - mediators of carcinogenesis;
  - providers of insulin resistance and glucose tolerance;
  - coordinators of redox homeostasis;
  - contractors of neurodegeneration;
  - contractors of myopathy.

**Selenoproteins of the endoplasmic reticulum membrane**

**Selenoprotein T (SELENOT).** Selenoprotein T (SelT) belongs to the thiorodoxin group of selenoproteins and is regulated by a neuropeptide pituitary polypeptide. Almost every tissue highly expresses SelT during rat embryogenesis. Subsequently, its levels gradually decrease during maturation and adult tissues lose this selenoprotein. In the brain, SelT is highly produced in neuroblasts in different areas, but not determined in mature neurons, except rostral migratory streams of astrocytes and Bergman cells. At the same time, some endocrine tissues continue the expression of SelT throughout life. Among them are: the pituitary gland, the thyroid gland or the testes. SelT expression is detected in chromophagic cells of the pars distalis of the pituitary gland, but in the tests it was observed in spermatogenic cells and Leydig cells only. Additionally, partial hepatectomy exerts SelT expression in liver cells during regeneration. Consequently, it can be concluded that embryogenesis, maturing and regeneration is characterized by SelT induction (Tanguy et al., 2011).

Finding high levels of SelT in pancreatic tissue prompted some researchers to study this selenoprotein in β-cell-knockout on gene SelT. Glucose administration shows the violation of tolerance to it in mutant animals and insufficiency of insulin production. Morphometric studies show the quantity of Langerhans islands increases while their size reduces. All the above suggests that SelT is localized in the β- and δ-cells of the pancreas and is involved in glucose homeostasis (Prevost et al., 2013).

Immunofluorescence and immunogold transmission electron microscopy methods revealed that SelT is produced by all endocrine cells – location is unknown (SelO, SelU, SelV, Fig. 1).

**Selenoprotein K (SELENOК)**

Today it is known that selenoprotein K is located in the ER and maintains a normal physiological state of skeletal muscle. Research has shown that Se deficiency reduces the expression of selenoprotein K and causes oxidative stress, ER stress, and apoptosis in chicken muscles (Fan et al., 2018).

The development of melanoma, according to research, requires the expression of selenoprotein K, which is necessary for the Ca^{2+} influx into the cancerous cells. Malignant cells show a strong dependence on the flow of calcium. In one experiment, CRISPR/Cas9 was used to form the selenoprotein K deficiency in human melanoma cells. This led to a decrease in the Ca^{2+} flux and impaired function of the specialized receptor (IP3R), which inhibited proliferation, invasion, and cell migration. Consequently, the growth and metastatic potential of the tumours depend on the selenoprotein K synthesis in the cancer cells (Marciel et al., 2018).

According to other data, selenoprotein K can act as a tumour suppressor in human choriocarcinoma cells, negatively regulating the expression of the beta-subunit of human chorionic gonadotrophin by signalling pathways ERK, p38 MAPK and Akt. All the above suggests that selenoprotein K may become a new target for human choriocarcinoma in vitro (Li et al., 2018).

Selenoprotein K (SelK) is a membrane protein that provides antioxidant protection, calcium regulation in the cell. It is a part of the protein degradation pathogenetic link related with the ER. The activity of low-speed peroxidase is regulated by SelK but within the range of other peroxidases. In particular, SelK reduces hydrophobic substrates, such as phospholipid hydroperoxides, thereby preserving membrane integrity. Hence, SelK can take part in membrane reconstruction. Disel

**Selenoprotein N (SELENON).** SelN gene damage causes the myopathy associated with this gene, characterized by muscle weakness, spinal rigidity, and respiratory insufficiency. In SelN-knockout mice under normal conditions, the histology of the muscles remains normal, but in skeletal muscle, after detecting oxidative stress, significant fine damage can occur. Ryanodine receptor calcium release channels (RyR) reveal a weaker sensibility to caffeine in SelN deficient myofibrils, displaying the possible role of SelN in the RyR regulation. The SelN deficiency also leads to the pathological development of the lungs, which is characterized by expanded alveoli. This is associated with impaired of tissue elasticity and enhancement in quasi-static lung ductility. This conclusion increases the possibility that the respiratory syndrome observed in patients with SEPN1 mutations may have a primary pulmonary component, in addition to the weakness of the respiratory muscles (Moghaddazadeh et al., 2013).

Pathological changes in the SelN gene cause a group of muscle violations. In this case, the axial muscles are affected. Involvement of the SelN in Ca^{2+} and redox homeostasis was demonstrated in some studies, but how it participates in skeletal muscle physiology remains unclear. To find out the SelN function in vivo, a group of researchers created a model of SelN knockout mice. These mice are characterized by usual parameters of maturing and morphology, they had the appearance of wild mice. Usually, SelN knockout mice display trivial defects only of muscle microstructure and contractile features. Forced training by swimming contributes to the development of specific phenotype ma-
manifestations characterized by limited mobility and rigidity of the body as well as progressive curvature of the spine and an overwhelming change in muscularity characterized by limited mobility and rigidity of the body as well as excessive consumption of selenium increased the release of Ca\(^{2+}\), Ca\(^{2+}\)-calmodulin, myosin light chain phosphorylation, and may be associated with the transport of Ca\(^{2+}\). However, selenocysteine is present in it, and selenoprotein M can also function as a regulator of oxidative processes, since its decreased or increased expression, regulated by dietary selenium, changes the redox homeostasis (Guaniniello et al., 2014). The interaction of some selenoproteins is integrative. Thus, selenoprotein P (SelP-H) and mutant selenoprotein M (SelM) are able to bind ions of transition metal and modulate Zn\(^{2+}\)-mediated amyloid-β (Aβ) aggregation, production of ROS and neurotoxicity. Aggregation and cytotoxicity of amyloid-β (Aβ) peptide with transition metal ions in neuronal cells are engaged in the progression of Alzheimer’s disease. The binding of Aβ to Zn\(^{2+}\) almost completely suppressed the fibrillation of Aβ, which could be significantly restored by SelP-H and SelM, which was observed by fluorescence and electron transmission microscopy. Both SelP-H and SelM inhibit Zn\(^{2+}\)-Aβ-induced neurotoxicity and intracellular synthesis of ROS in living cells. Studies have shown that SelP and SelM can play a greatly role in regulating the oxidation-reducing balance, as well as metallic homeostasis (Du et al., 2013).

Expression of SelM is observed in the genesis of the axial skeleton of chickens and rodents. It was most pronounced in osteoblasts, tendons and bones (Grosch et al., 2013). SelM expression is also fixed in embryonic human kidney cells. Full-size SelM prevents aggregation of Aβ (β-amyloid, an appropriate Alzheimer’s disease protein) by counteracting the oxidative stress which occurs in oligomeric cells (Chen et al., 2013).

Common proteins have been extracted to find out the general changes in the all kidney proteins of transgenic rodents characterized by excess SelM expression. Synthesis of protein was greater in transgenic rodents than in the control (LAP3, BAAP21, CRP2, CD73, PDGF D, KIAA143, PRP5S2, ZFP313, HSP-60, and N-WASP), while other proteins were diminished (ALKDH3, MCP-3, STC-1). Selenium administration promoted amplified production levels of the five highly expressed proteins in control mice, while transgenic animals did not undergo changes (Goo et al., 2013).

Changes in global protein synthesis were measured in the cerebral cortex in transgenic rats that expressed SelM and non-transgenic rats.

1) Comparison of transgenic with non-transgenic animals revealed high parameters of enzyme activity for antioxidant protein in the cerebral cortex.

2) Rise in activity of these enzymes stimulates depletion of the total antioxidant content and activity of γ-glutamylcysteine transaminase in transgenic rats.

3) Five proteins were amplified, and three of them were reduced as a result of excessive SelM expression.

4) Among the five upregulated proteins, two of them were enlarged in both groups after addition of selenium, while the eukaryotic factor for initiating 4H (eIF-4H) and lactate dehydrogenase B (LDH-B) increased or decreased under the same conditions.

5) Three downregulated proteins did not cause considerable changes in expression after the administration of selenium (Kim et al., 2014).

Selenoprotein M is highly produced in the human brain, but the biological effect and molecular mechanism remain obscure. For verification of SelM mechanisms of action, a mutant selenoprotein M was created. Then it was discovered that the new interactive SelM protein is galecit-1 (Gal-1). Gal-1 plays a crucial role in preventing neurodegeneration and promoting neuroprotection in the brain, the interaction between SelM and Gal-1 demonstrates a new direction in the study of the biological function of SelM in the human brain (Qiao et al., 2013).

SelF deficiency induces a decrease in the content of glutathione peroxidase and the activity of catalase; elevated levels of malondialdehyde and reduced expression of selenoprotein RNA (mRNA) and an essential reduction in SelM protein in the brain (Huang et al., 2016).

**Selenoprotein S (SELENOS).** The next selenoprotein related with the ER is selenoprotein S, which, by cooperation with another membrane protein (valosin-containing protein), provides the incorporation of selenoprotein K into the ER. The main evidence of this is that interaction between selenoprotein K and valosin-containing protein does not occur in cells deprived of selenoprotein S, while selenoprotein S interacts with valosin-containing protein, regardless of the presence or absence of selenoprotein K. Expression of selenoprotein S and K rises according to stress of the ER (Lee et al., 2015).

The expression and location of SelS, its function and regulatory factors in the intact and inflammatory intestine epithelium have been defined. SelS was elevated in inflamed ileal tissues in patients with Crohn’s disease. The same elevation was observed in mice models of enterocolitis. SelS expression was not connected with the differentiation...
of enterocytes, but increased in response to the supplementation of selenium and after correction of ER stress by an inducer of tunicamycin. Therefore SelS can be considered as a new marker for endoplasmic reticulum intestinal stress (Speckmann et al., 2014).

To study the role of SelS in the adjustment of ER stress, the interaction of selenoprotein with the membrane protein p97 on mouse neuroblastoma cells and human embryonic kidney cells were explored. SelS expression level was always enhanced when there was an ER induction of stress (Lee et al., 2014).

Selenoprotein S (SelS), as one of the coordinators of the inflammatory process, responds to ischemia. Recently, the exploration of transitory cerebral ischemia aftermaths was conducted. A decline in SelS synthesis in the ischemic cell was observed 3–7 days after occlusion, but in the zones adjacent to hypoxic site there was an elevation in SelS expression corresponding to reactive angiogenesis (Liu et al., 2013).

Cell culture has shown that SelS overexpression mitigates cytotoxicity and apoptosis induced by ochratoxin, as well as increases glutathione levels and reduces ROS, overwhelms ochratoxin-induced phosphorylation. Conversely, the SelS knockdown reduces glutathione levels, enhances the generation of ROS and ochratoxin-induced phosphorylation to the same extent as cytotoxicity and apoptosis (Gan et al., 2017).

Selenoprotein S is highly expressed in skeletal muscle. The depression of the SelS gene greatly exacerbates the inflammatory profile of fast muscle fibers, which are generally more susceptible to degeneration in dystrophy (Wright et al., 2017).

Enhanced SelS expression increases the levels of nitric oxide and endothelial nitric oxide synthase in endothelial cells treated by the tumour necrosis factor (TNF-α). A study was conducted on the human umbilical vein endothelial cells culture (HUVECs). Moreover, overexpression of SelS keeps out the TNF-α-induced adhesion of THP-1 cells to endothelial cells. Also, SelS overexpression regulates TNF-α-induced inflammatory factors, including interleukin-1β, interleukin-6, interleukin-8, and monocytic chemotactic protein-1. Conversely, the SelS knockout of siRNA leads to an increased TNF-α-induced injury in HUVECs. Thus, SelS protects endothelial cells from TNF-α-induced dysfunction (Cui et al., 2018).

Atherosclerotic vascular lesions may be accompanied by calcification and differentiation of osteoblast. Osteoblast differentiation and calcification, which are induced by lipopolysaccharides (LPS) or TNF-α were significantly aggravated with SelS knockdown in vascular smooth muscle cells. The SelS knockdown also exacerbates the LPS-induced increase in the expression of proinflammatory cytokines TNF-α and interleukin-6, as well as increased expression of stress markers of ER. Thus, it can be assumed that SelS can suppress inflammation-induced calcification of smooth myocytes in vessels due to suppression of signaling pathways of nuclear factors and endoplasmic reticulum stress (Ye et al., 2018).

Selenoprotein F (SELENOF). The selenoprotein F or 15 kDa is localized in the lumen of the ER and takes part in quality controlling of protein compaction. The RNA-induced deficiency of Sel F leads to inhibition of cell proliferation, whereas cell growth is restored after removal of the knockdown inducer. SelF deficient cells are blocked in the G1 phase and show signs of ER stress. In addition, the Sel F deficiency leads to the displacement of the adhesive contacts of the cells to the periphery of the basal part, and also diminishes the migration and cellular invasive power. But all these changes are reversible and depend on the state of Sel F. Thus, it can be noted that Sel F plays a crucial role in adjustment of the G1 cell cycle phase as well as cell mobility (Bang et al., 2015). Selenoprotein F shows the ability to react with an enzyme retinol dehydrogenase that catalyzes the recovery of trans-retinol (vitamin A). At the same time there is a blockade of this enzyme. Consequently, the overexpression of selenoprotein F leads to a decline in the retinol synthesis (Tian et al., 2018).

In the study of ultramicroscopic features in Sel F deficient cells, most of them exhibited the formation of membrane vesicles. Accumulation of bubbles modifies the shape of cells from a flat spindles to a spherical one. In such cells, actin filibrns are displaced to the periphery, overlapping α-tubulin. These morphological alterations are invertebrable, and inhibited by inhibitors of Rho-associated protein kinase. Sel F deficient cells are nonapoptotic and show a clear localization of F-actin and α-tubulin, as opposed to typical apoptotic blebbing cells. Thus, it can be argued that Sel F regulates the pathway that counteracts the RhoA / ROCK / MLC-dependent non-apoptotic bubble formation (Bang et al., 2015).

**Nuclear selenoproteins**

Selenoprotein H (SELENOH). Selenoprotein H is a newly identified selenoprotein, which is a nucleolar oxidoreductase. According to some authors, SelH regulates redox homeostasis and suppresses DNA damage (Cox et al., 2016). Selenoprotein H defends cells from aging through the effect on oxidative stress by supporting the genome (Wu et al., 2014). Human cells, in which the SelH expression was generated, were exposed to glutamate, which is known for its pathogenicity and its ability to cause cell death mediated by mitochondrial damage. The results of the study showed that cytotoxicity of glutamate is bound to an increase in the production of ROS, an imbalance in the dynamics of mitochondria and autophagy. These changes disappeared, and cellular integrity was restored by excessive SelH expression (Ma et al., 2017).

It is believed that SelH is involved in redox homeostasis, as well as teratogenesis. Expression of SelH is elevated in tumour tissue, in undifferentiated epithelial cells of the gastrointestinal tract of mice. The suppression of Sel H expression by knockdown reduces cell differentiation and enhances proliferation and migration. Moreover, SelH knockdown cells are more capable of creating tumour colonies and xenografts. However, they show a more accelerated cell cycle. The foregoing indicates SelH is the leading regulator of cell cycle progression and prevents uncontrolled proliferation. SelH expression is consistently dependent on Se, so the effect of selenium addition on cancer initiation and progression is probably due to SelH (Hertz et al., 2018).

Selenoprotein R (SELENOR). Selenoprotein R (SelR), known as methionine sulfoxidredactase, is an enzyme whose role was studied in the SelR-knockdown epithelial cells of the eye lens, against the background of galactose-induced apoptosis. The results have shown both d-galactose and the SelR-gene knockdown independently induce oxidative stress. But the effect of galactose on cells under the SelR knockdown induces an even greater elevation in glucose-regulated protein levels, and lowers mitochondrial membrane potential, which is accompanied by the liberation of mitochondrial cytochrome. At the same time, the percentage of cells with apoptosis significantly increases. Thus, SelR can protect the mitochondria of lens epithelial cells against oxidative stress, and weaken apoptosis in these cells (Dai et al., 2016).

The antioxidant properties of SelR are elucidated in a number of studies. Thus, SelR-deficient mice exhibit significantly greater sensitivity to acetaminophen-induced hepatocyte damage than mice without deficiency. This is confirmed by necrotic lesions in the central areas of the liver lobes, as well as laboratory data (Kim et al., 2017; Singh et al., 2017).

Similar changes were observed in the kidney model. The SelR knockdown in mouse kidney cells causes increase in cisplatin-induced lesions. Cisplatin induces swelling, loss of cristae and fragmentation of mitochondria with an increase in peroxide lipid edema, especially in SelR-deficient kidneys compared to kidneys with normal SelR expression (Noh et al., 2017).

MsrB1 (or SelR) is highly expressed in immunoreactive macrophages and contributes to the realization of cellular and organismal immune responses. MsrB1 controls immune responses by facilitating the detection of anti-inflammatory cytokines in macrophages. MsrB1 dependent reduction of oxidized methionine in proteins may constitute an unrecognised regulatory mechanism underlying immunity and inflammatory disease, and a new target for clinical applications (Lee et al., 2017).

Decreased expression of MsrB3 (methionine sulfoxide reductase B3) promote apoptosis of cancer cells by the mitochondrial pathway and leads to the undoing of cancer cells. The deficiency of MsrB3 greatly increased the ER stress, which led to apoptosis. In addition, depletion of MsrB3 activates the proapoptotic Bim molecule, which is essential for ER-stressed apoptosis. The deficiency of MsrB3 increases the level of cytosolic calcium, suggesting that overexpression of MsrB3 leads to violations of calcium homeostasis in ER, which, as a result, causes endoplasmic reticulum stress. MsrB3 plays a decisive role in cancer cell apo-
Selenoprotein P (SELENOP). According to published data, circulatory selenoprotein P (SePP) is linked to bone metabolism, but its function for bone balance is not completely known. Some researchers report that selenoprotein R is an important transporter of Se to the bones. Se is necessary for normal osteogenesis, which is confirmed by the data on Kashin-Beck disease, endemic Se-dependent osteoarthritides (Pitschmann et al., 2014).

One study examined the role of selenoprotein P (SelP) in recirculatory and ischemic alterations. The study was conducted in SelP-knockout mice group and in wild mice. Both groups were exposued to 30-minute ischemia, followed by 24-hour reperfusion. The area of myocardial infarction, which was evaluated by Evans coloration, was significantly less in the SelP-knockout mice group than in the wild mice. There was also a significant increase in the size of the heart in mice that excessively expressed SelP in the liver compared to other mice in the modeling of ischemia-reperfusion. These data indicate that SelP inhibition protects the heart against damage from ischemia-reperfusion (Chadani et al., 2018).

Glucose administration increases the expression of genes and the transcriptional activity of SelP in cultured hepatocytes. Physiological SelP concentrations inhibit cell proliferation stimulated by vascular endothelial growth factor (VEGF), tubular formation and migration in the endothelium of the human umbilical vein. SelP inhibits the generation and phosphorylation of VEGF-induced reactive oxygen species and extracellular signaling regulated kinase in the endothelium of the human umbilical vein. Healing of the wound is disturbed in SelP expressing mice, while healing in SelP-knockout mice was improved (Ishikura et al., 2014).

According to some researchers, selenoprotein P (SelP), a liver secretary protein, causes resistance to insulin. Using sequential gene expression analysis (SAGE) and DNA chip methods, it was found that levels of SelP mRNA correlate with insulin resistance in humans. The introduction of purified SelP disrupts insulin signaling and regulates glucose metabolism in both hepatocytes and myocytes. Conversely, both knock-out and interference mediated elimination of SelP mRNA improved system sensitivity to insulin and glucose tolerance in mice. SelP's metabolic action is mediated, at least in part, by inactivation of adenosine monophosphate-activated protein kinase (AMPK). Thus, these results demonstrate the role SelP plays in the regulation of glucose metabolism and insulin sensitivity, suggesting that SelP may be a therapeutic target for type 2 diabetes (Misu et al., 2010).

Distinct clinical investigations have proved the linkage of selenoprotein P and insulin resistance. The researches compared serum SelP content of one hundred people with different glucose sensitivity states. In addition, of some cardiac metabolic risk factors with SelP were evaluated. Among them were the intima-media thickness of the carotid artery, insulin resistance and high sensitivity of the C-reactive protein. In the serum of type 2 diabetes persons and prediabetes patients, concentration of SelP is significantly higher, and it is lower in patients with normal glucose tolerance and gradually decreasing. The same differences of SelP in blood were observed in patients with overweight and normal weight, respectively. Strong correlation was detected by Spearman’s partial correlation analysis between serum SelP and most risk factors such as insulin resistance, body mass index, systolic blood pressure, hemoglobin, glucose, aspartate aminotransferases, triglycerides, waist circumference. Consequently, glucose tolerant persons serum SelP parameters were increased and related to some cardiac metabolic factors such as atherosclerosis, insulin resistance, and inflammation (Yang et al., 2011).

Selenoprotein P (SelP) is defined as hepatokin and promotes type 2 diabetes resistance. The depression of SelP activity may improve glucose metabolism. The monoclonal antibody AE2 with neutralizing activity against SelP was elaborated, administration of which to mice significantly improved glucose intolerance and insulin resistance. Moreover, the excessive administration of SelP significantly reduces the level of insulin in the pancreas and glucose-induced insulin secretion, which improves when AE2 is administered. The mapping of the epitope shows that AE2 recognizes the SelP region of a person adjacent to the first histidine-rich area. The anti-SelP polyclonal antibody improved glucose intolerance and insulin secretion in the diabetes model (Mita et al., 2017).

Sel-P deficient mice exhibit a “super-sustainability” phenotype after training, as well as enhanced production of ROS, phosphorylation of protein kinase and activation of the proliferative receptor with peroxide. However, the addition of N-acetylcysteine antioxidant reduces the production of ROS and endurance in SelP deficient mice (Misu et al., 2017).

There are data about the interaction of SelP with tubulin in human embryonic brain cells, as evidenced by the analysis of fluorescence resonance energy transfer (FRET) and co-immunoprecipitation reactions (Du et al., 2014).

Selenoprotein W (SELENOW). The study of selenoprotein W, in most cases, was performed on the model of chickens. So, it is well-known that excess Ca²⁺ is present in Se deficiency, but the mechanisms of this phenomenon are not sufficiently studied. That is why this process was studied in the sample of chicken embryo and SelW knockdown myoblasts. It was found that Se deficiency induces typical muscle damage that is accompanied by a Ca²⁺ disorder, oxidative stress, which damages the ultrastructure of the sarcoplasmic reticulum and mitochondria; reduction of Ca²⁺ channels, decrease of levels of selenium-containing enzymes SERCA, SLCA, CACNAIS, ORAI, STIM1, TRPC1, and TRPC3. Similar changes are also observed in the case of modeling of SelW knockout myoblasts (Yao et al., 2016).

The effect of selenoprotein W on lymphoid tissues and cultivated spleen leukocytes was studied in the inflammatory process of chickens. The use of the selenogenic deficiency diet effectively reduces the expression of SelW mRNA and induces a considerable increase in glutathione peroxidase-2 (Gpx2), glutathione reductase, selenoprotein W, selenoprotein H and selenoprotein W and glutathione peroxidase-2 and thiorredoxin reductase, selenoprotein W, selenoprotein H and selenoprotein F on models of various cell lines. The most effective in increasing all biomarkers were selenium sodium and methyl selenium cholesterol, while other compounds had only minor effects (Kipp et al., 2013).

Selenoproteins with the least studied functions: V (SELENOV), I (SELENOI), U (SELENOU), O (SELENOO). The recently discovered selenoprotein U is characterized by considerable autophagosome formation and lysosome degradation with intact cytoskeleton in Sertoli cells that are exhausted or deprived of this protein. These data indicate that the deposition of selenoprotein U causes autophagy and reduces the expression of important growth factors in Sertoli cells by way of signaling pathway disturbances. Generally, selenoprotein U is important for the survival and functioning of the Sertoli cells (Sattar et al., 2018).

The functions of most selenoproteins are not yet identified. Among these, there is also selenoprotein V. SelV expression reaches its secretory peak during puberty, while progressive decline of expression is revealed in adult mice. SelV also has activity of glutathione peroxidase and thio-
redox reductase (Varlamova et al., 2015). SelV (Selenoprotein V) has a thioerdioxin-like folding and a conservative motif (CXXU, where C is cysteine, U-selenocysteine) in its catalytic center; it belongs to the fraction of redox proteins whose participants are engaged in oxidative-reduction reactions. The SelV redox protein can interact with the O-linked N-acetylglucosamine transferase (OGT) and proteins belonging to the ASB family: Asb-17 and Asb-9. Specificity of SelV interactions with OGT and Asb-9, but not with Asb-17, is confirmed by immunoprecipitate. Additionally, Selv mRNA expression has been shown in later stages of spermatogenesis, as well as during puberty and reproduction of rats (Varlamova et al., 2012).

The function of SelO has not been clarified, however, there are reports of the effects of SelO on chondrocyte differentiation, notably the increase of the mRNA levels and SelO expression when chondrogenic induction of ATDC5 cells. The silencing of the SelO gene leads to inhibition of chondrogenic differentiation, which is accompanied by the accumulation of several cartilaginous glycosaminoglycans and decreased activity of alkaline phosphatase in selenium deficiency cells. The inhibition of proliferation is also due to delay in the progression of the cell cycle. The deficiency of SelO stimulates the extinction of chondrocytes by apoptosis (Yan et al., 2016).

The intracellular localization of SelI human selenoprotein and the levels of gene expression in various human tumour cell lines are determined. SelI protein was found in the nucleus, cytoplasm and endoplasmic reticulum, and absent in the nucleolus (Varlamova et al., 2017). For the convenience of perception, all the information is provided in Table 1.

### Table 1

| Name of selenoprotein | Localization | Biological effects | Knockdown effects on the corresponding gene (exclusion of the gene) |
|----------------------|--------------|--------------------|------------------------------------------------------------------|
| SELENOF              | Lumen of endoplasmic reticulum | Quality control of protein compaction regulates the G1 period of the cell cycle, as well as cellular mobility. It inhibits retinol dehydrogenase, which reduces the production of retinol | It inhibits cell proliferation, causes cells blockade in the G1 period, endoplasmic reticulum stress, displacement of cells adhesive contacts, accumulation of membrane nonapoptotic vesicles |
| SELENOH (nuclear oxide reductase) | Nucleus | Involved in redox regulation, testogogenesis (elated in tumours), regulator of cell cycle progression, prevents uncontrolled proliferation. With excessive expression, it exhibits antioxidative properties. Protects DNA from damage | Reduces cell differentiation and increase proliferation and migration |
| SELENOI              | Cytoplasm, nucleus, endoplasmic reticulum | Expressed in tumours of different human cell lines Regulates Ca2+ flux in cancer cells, provides antioxidative protection, calcium regulation in the cells, and the pathway for degradation of the protein associated with the endoplasmic reticulum. Affects the physiological state of the muscles. Reduces phospholipid hydroperoxides (antioxidant function) | Unknown |
| SELENOK              | Endoplasmic reticulum membrane | Involved in testogogenesis (increased expression in liver tumours). Transportation of Ca2+ affects redox homeostasis | Provides reduction of Ca2+ in cancer cells and impaired function of the specialized receptor (IP3R), which inhibits proliferation, invasion and cell migration |
| SELENOH              | Lumen of endoplasmic reticulum | Involved in testogogenesis, (increased expression in liver tumours). | Unknown |
| SELENON              | Endoplasmic reticulum membrane | Participates in calcium and redox homeostasis, but mechanisms of regulation of skeletal, cardiac muscles, smooth muscles are unknown | Demonstrates limitation of mobility and rigidity of the body, as well as the progressive curvature of the spine and the predominant change in para-axial muscles (signs of myopathy) |
| SELENOO              | Unknown | Affects the differentiation of chondrocytes | Stimulates inhibition of chondrocytes differentiation, delayed progression of the cell cycle, loss of chondrocytes |
| SELENOE              | Cytoplasm, secretion by hepatocytes | Regulation of glucose metabolism and insulin sensitivity contributes to the development of insulin resistance in type 2 diabetes | Provides improvement of systemic insulin sensitivity and glucose tolerance in mice, formation of the "super-sustainability" phenotype, increasing the production of reactive oxygen species, reducing the area of myocardial infarction |
| SELENO (methionine-R-sulfoxidase) | Nucleus, cytoplasm | It can protect mitochondria from oxidative stress, and weaken apoptosis in cells | Reduces the level of glutathione, enhances the production of reactive oxygen species, aggravates the cytotoxicity and apoptosis. Exacerbates the inflammatory profile of fast muscle fibers, differentiation, and calcification of osteoblasts |
| SELENO               | Lumen of endoplasmic reticulum | Endoplasmic reticulum stress marker, regulator of inflammatory processes | Induces oxidative stress |
| SELENOS              | Endoplasmic reticulum membrane | It is a subunit of the protein oligosacchaeidine transferase complex. Provides the regulatory function of the endocrine glands, the control of glucose homeostasis. It is expressed by cells in the embryogenesis, during the maturation of tissues, and regeneration | Violation of glucose tolerance, deficiency of insulin products, rapid and severe Parkinson-like and motor defects |
| SELENOT              | Unknown | Shows signs of belonging to the system of redox regulation, taking part in oxidative-reduction reactions. Maximum expression in animals is found during puberty, in adults is reduced | Considerable autophagy and lysosomal degradation with the intact cytoskeleton in Sertoli cells |
| SELENOU              | Unknown | The least studied. Can regulate the expression of important growth factors in Sertoli cells | Unknown |
| SELENOW              | Cytoplasm | Regulator Ca2+ releasing due to the effect on its channels in the sarcoplasmic reticulum of myosinplast | Ca2+ releasing, typical muscle damage, decreased cell viability, accelerated apoptosis and increased sensitivity to H2O2 |

### Conclusions

The identification of selenoproteins remains one of the promising directions of modern science. This is due, as shown by the analysis of recent studies, to the active involvement of this family of proteins in the most crucial pathophysiological processes of living systems. Some mechanisms of participation of selenoproteins in the oxidative balance, redox homeostasis, carcinogenesis, neurodegeneration and myodystrophic disorders have been discovered. Methods have been developed to demonstrate the expression of certain selenoproteins in different tissues, to identify the role of these proteins in various signaling pathways. However, many questions regarding the verification of biological functions remain open. This creates interest for specialists in many areas in further studies of selenoproteins.

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