"HLA-G 3’UTR gene polymorphisms and rheumatic heart disease: a familial study among South Indian population"

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Abstract

**Background:** Rheumatic heart disease (RHD) is an autoimmune disease where cross reactive CD4⁺ T cells are involved in the pathogenesis of valvular damage. Human Leukocyte Antigen-G (HLA-G), an immunosuppressive molecule playing a crucial role in the inhibition of T cell response is associated with the pathogenesis of various autoimmune and inflammatory diseases. Genetic polymorphisms within the 3' untranslated region (UTR) of HLA-G influences its expression and thus disease pathogenesis. Hence, the present study aims to unravel the association of 14 bp Ins/Del (rs66554220) and +3142 C/G (rs1063320) polymorphisms in 3’ UTR of HLA-G with RHD.

**Methods:** This familial study consists of 99 RHD families (99 RHD patients, 140 parents and 126 healthy siblings). The 14 bp Ins/Del and +3142 C/G polymorphisms were evaluated by PCR using sequence specific primers and its transmission disequilibrium (TD) was tested by TD test in 70 trio families.

**Results:** The frequency of +3142 C/C genotype was high in patients with combined valvular lesions (CVL) (OR = 5.88; pc = 0.012) and pooled RHD patients (P: OR = 2.76; p = 0.043; pc = 0.076) when compared to healthy siblings. Under the additive (OR = 5.50; pc = 0.026) and recessive genetic model (OR = 5.88; pc = 0.012), the +3142 C/C genotype was significantly associated with CVL in patients.

**Conclusion:** The results suggest that the +3142 C/C genotype may be associated with minor risk for the development of RHD and is more likely to influence the severity of the disease.

**Keywords:** Human leukocyte antigen G, Rheumatic fever, Rheumatic heart disease, Polymorphism, Autoimmunity

Background

RHD is an autoimmune disease occurring as a result of group A β-hemolytic Streptococcus (GAS) infection followed by rheumatic fever (RF) [1]. Molecular mimicry between streptococcal M protein and human cardiac proteins leads to the development of RHD in a susceptible host [2]. RHD causes major cardiac illness, subsequently leading to high morbidity and mortality among children in developing countries and also in certain indigenous populations within developed countries. Globally, over 15 million cases of RHD have been reported and its prevalence in India ranges from 2–2.5 million [3, 4]. It is an inflammatory condition resulting in chronic valvular damage specifically mediated by cellular immune response. The aberrant immune response against host cardiac proteins may account for the progressive valvular damage [5–9].

HLA-G is a non-classical class-I HLA molecule associated with anti-inflammatory and immuno-modulatory properties [10, 11] which interacts with inhibitory receptors (ILT2/ILT4/KIR2DL4) present on various immune cells. It inhibits proliferation of B cells, T cells and natural killer cells; and also induces regulatory T cells (T-reg) [12–15]. The polymorphic 3’UTR of HLA-G plays a vital role in the regulation of HLA-G expression. The 14bp Ins/Del polymorphism regulates HLA-G expression by influencing the splicing pattern and stability of mRNA. Whereas, the +3142 C/G polymorphism enhances the affinity for micro RNAs and downregulate the expression of HLA-G [16–19]. Thus, both the polymorphisms influence...
HLA-G expression and are associated with various autoimmune and inflammatory diseases [20, 21]. Several studies have demonstrated the genetic predisposition to RHD, especially those genes which are involved in immune response [22]. However, the role of HLA-G in the pathogenesis of RHD has not been reported yet. Hence, the aim of this study is to evaluate the possible role of HLA-G gene polymorphisms with particular importance to 14bp Ins/Del (rs66554220) and +3142C/G (rs1063320) variants towards the development of RHD in South Indian population.

Methods
Study population
This family based retrospective study consisted of 365 individuals (99 RHD patients, 140 parents and 126 healthy siblings, which includes 70 trios), recruited from the Institute of Child Health and Research Centre, Government Rajaji Hospital, Madurai, India. The study was approved by ethics committee of the participating institutions and was carried out in accordance with the declaration of Helsinki. Patients with ARF were diagnosed based on modified Jones criteria. They clinically present with migrating polyarthritis, carditis and chorea. Based on echocardiographic data, the patients were classified into three subgroups: (i) rheumatic fever (RF) - patients without residual valvular disease; (ii) mitral valvular lesions (MVL) - patients with mitral valvular lesions only; (iii) combined valvular lesions (CVL) - patients with multiple valvular lesions along with MVL. Further, the severity of valve damage was assessed by investigations like echo cardiogram showing Mitral/Aortic/Tricuspid valve morphology, color flow doppler MR/AR/TR jet, vena contracta width, effective regurgitant orifice area, left atrium (LA) and left ventricle (LV) size, pulmonary arterial pressure, pulmonary vein flow and hepatic vein flow. In addition, regurgitation is clinically diagnosed by prominent pulsations of the jugular veins, systolic pulsations of the liver and a blowing holosystolic murmur at the lower left sternal border which increases in intensity during inspiration and expiration for TR and MR respectively. Currently, all the patients in the study group are in quiescent phase, under penicillin (tablet) prophylaxis against rheumatic fever recurrence. Patients with carditis were treated with steroids (oral prednisone 2mg/Kg) initially and subsequently treated with aspirin tablets. Antifailure drugs like frusemide, digoxin and enalapril were given appropriately to the patients with congestive cardiac failure. None of the patients underwent surgery for valvular heart disease in this study group. Siblings, free from autoimmune and cardiac diseases were considered as controls. Like the patients, the siblings are under follow-up at regular intervals. None of the siblings developed any symptoms of ARF like, polyarthritis, carditis, chorea, erythema marginatum and subcutaneous nodules during the follow-up period. The siblings are advised to be under regular follow-up. Blood samples (2.5 ml) were collected from all the participating individuals with their consent. For children (<18 years), the consent was duly signed by their parents.

Genotyping of HLA-G gene polymorphisms
Genomic DNA was extracted from the blood samples by salting out method [23]. HLA-G 14 bp Ins/Del and +3142 C/G polymorphisms were examined by direct PCR DNA amplification method [24]. Further, validation of 14 bp Ins/Del polymorphism was also performed by PCR-SSP [25].

Statistical analysis
Hardy-Weinberg equilibrium (HWE), TDT, linkage disequilibrium (LD) and haplotype analysis was performed using Haplovie v4.2 [26]. The HLA-G allelic and genotypic frequencies of patients and siblings were compared using the Fisher’s exact test or Pearson’s χ² test with Yates correction using Epi Info v7. The odds ratio (OR) with 95% confidence interval (CI) was calculated and the p < 0.05 was considered statistically significant.

Results
Characteristics of the study population
This familial study included 99 RHD patients (52 males, 47 females, mean age 11.2 ± 2.5 years), 126 healthy siblings (75 males and 51 females, mean age 12.2 ± 4.3 years) and 140 parents (70 mothers and 70 fathers). The demographic and clinical characteristics of the study subjects is represented in Table 1. In this study group, migrating polyarthritis was present in 80 patients and carditis was seen in 56 patients. In addition, chorea was present in 7 patients while erythema marginatum and subcutaneous nodules were not seen even in a single patient. Out of 99 patients, 10 (10.1%) patients had RF, 67 (67.7%) had MVL and 22 (22.2%) had CVL.

Transmission disequilibrium test (TDT)
In the present study, 70 trio families were included for TDT analysis (Table 2). No Mendelian error was observed and the families were in HWE expectations. In single marker analysis, Del allele (32 T vs 28 UT; p = 0.606) and G allele (27 T vs 23 UT; p = 0.572) were overtransmitted, however no statistical significance was observed. Likewise, the haplotype analysis also showed no significance among Ins/G (28 T vs. 31 UT; p = 0.697), Del/G (28.5 T vs. 20.5 UT; p = 0.255) and Del/C haplotypes (23.5 T vs. 27.5 UT; p = 0.578).
Linkage disequilibrium analysis

Pairwise linkage disequilibrium analysis was carried out for the two polymorphic sites of the HLA-G gene in the study population. The haplotype analysis predicted a single block of high LD in healthy siblings (D’ = 1, LOD = 10.57 and r² = 0.38), RHD patients (D’ = 1, LOD = 10.56 and r² = 0.32) and Trios (D’ = 0.966; LOD = 17.82; r² = 0.285) between HLA-G 14 bp Ins/Del and +3142 C/G polymorphism (Additional file 1: Figure S1).

Genotype and Allele distribution in RHD patients and healthy siblings

The genotype and allele frequencies of 14bp Ins/Del and +3142 C/G polymorphisms for pooled (P) RHD patients and healthy siblings were shown in Table 3. The data was further stratified based on gender (male (M) and female (F)). There were no significant differences observed for the genotype and allele frequencies of these two polymorphisms between RHD patients and healthy siblings.

In 14 bp Ins/Del polymorphism, an increased frequencies of Ins/Ins genotype (OR = 2.86; p < 0.05) and Ins allele (OR = 1.14; p = 0.766) were observed in female RHD patients. Moderately increased frequencies were noticed in Del/Del genotype (P: OR = 1.37; p = 0.312) and Del allele (P: OR = 1.28; p = 0.236) among pooled RHD patients. The subgroup analysis showed increased frequencies of Del/Del genotype in CVL (OR = 1.73; p = 0.363) and MVL patients (OR = 1.39; p = 0.383) (Table 4). In addition, increased frequency of Del allele was observed in MVL (OR = 3.0; p = 0.184) and CVL patients (OR = 1.25; p = 0.607).

The +3142 C/C genotype frequency was relatively high in pooled RHD patients (P: OR = 2.76; p = 0.043; p = 0.076). Significantly increased frequency of +3142 C/C genotype was found in CVL patients (OR = 5.88; p = 0.012) when compared to healthy siblings (Table 4). In the genetic model analysis, the additive (OR = 5.50; p = 0.026) and recessive pattern (OR = 5.88; p = 0.012) of +3142 C/C genotype was inherited significantly among CVL patients (Table 5) when compared to healthy siblings. The +3142 C/C genotype frequency was higher in male RHD patients (M: OR = 3.68; p = 0.112) when compared to female RHD patients (F: OR = 1.94; p = 0.604).

The three possible haplotypes observed in the present study were Ins/G, Del/G and Del/C. The frequency distribution of these haplotypes among the study groups did not show statistical significance (Table 3). However, increased frequency of Del/C haplotype was observed in pooled RHD (P: OR = 1.31; p = 0.228) and CVL patients (OR = 1.78; p = 0.130) (Table 4) while Ins/G haplotype was found to be high among female RHD patients (F: OR = 1.14; p = 0.766).
Table 3  HLA-G genotype, allele and haplotype frequencies in patients and healthy siblings

|                  | Pooled Male |                      | Pooled Female |                      | Male |                      | Female |                      |
|------------------|-------------|----------------------|---------------|----------------------|------|----------------------|--------|----------------------|
|                  | Patients (99) n (%) | Healthy siblings (126) n (%) | OR (95% CI) p c | Patients (52) n (%) | Healthy siblings (75) n (%) | OR (95% CI) p c | Patients (47) n (%) | Healthy siblings (51) n (%) | OR (95% CI) p c |
| HLA-A*11:01      | 16 (16.2)   | 27 (21.4)            | 0.71 (0.36-1.40) | 0.4084               | 9 (17.3) | 24 (32.0)            | 0.44 (0.18-1.04) | 0.0987               | 7 (14.9) | 3 (5.9) | 2.86 (0.69-11.77) | 0.2418 |
| HLA-A*11:02      | 35 (35.3)   | 36 (28.6)            | 1.37 (0.78-2.40) | 0.3128               | 18 (34.6) | 19 (25.3)            | 1.53 (0.71-3.32) | 0.3755               | 17 (36.2) | 17 (33.3) | 1.17 (0.51-2.68) | 0.8791 |
| HLA-A*11:05      | 48 (48.5)   | 63 (50.0)            | 0.94 (0.55-1.59) | 0.9272               | 25 (48.1) | 32 (42.7)            | 1.24 (0.61-2.53) | 0.6734               | 23 (48.9) | 31 (60.8) | 0.62 (0.28-1.38) | 0.3296 |
| HLA-DRB1*08:01   | 118 (59.6)  | 135 (53.6)           | 1.28 (0.88-1.86) | 0.2368               | 61 (58.7) | 70 (46.7)            | 1.62 (0.98-2.70) | 0.0797               | 57 (60.6) | 65 (63.7) | 0.88 (0.49-1.56) | 0.7657 |
| HLA-DRB1*15:02   | 63 (31.8)   | 66 (26.2)            | 2.76 (0.99-7.63) | 0.0763*              | 7 (13.5) | 3 (4.0)              | 3.68 (0.90-14.98) | 0.1121               | 5 (10.6) | 3 (5.9) | 1.94 (0.44-8.62) | 0.6041 |
| HLA-DQB1*02:03   | 48 (48.5)   | 66 (52.4)            | 0.86 (0.50-1.51) | 0.6556               | 27 (51.9) | 44 (58.7)            | 0.76 (0.37-1.55) | 0.5680               | 21 (44.7) | 22 (43.1) | 0.96 (0.48-2.37) | 0.9602 |
| HLA-DRB1*03:01   | 39 (39.4)   | 54 (42.8)            | 0.87 (0.51-1.48) | 0.6985               | 18 (346) | 28 (37.3)            | 0.87 (0.41-1.82) | 0.8556               | 21 (44.7) | 26 (51.0) | 0.81 (0.37-1.78) | 0.7431 |
| HLA-DRB1*03:04   | 63 (31.8)   | 66 (26.2)            | 1.31 (0.87-1.98) | 0.2280               | 32 (30.8) | 34 (22.7)            | 1.52 (0.86-2.66) | 0.1927               | 31 (33.0) | 32 (31.4) | 0.98 (0.59-1.96) | 0.9302 |
| HLA-DRB1*03:06   | 135 (68.2)  | 186 (73.8)           | 0.76 (0.50-1.15) | 0.2280               | 72 (69.2) | 116 (77.3)           | 0.66 (0.37-1.16) | 0.1927               | 63 (67.0) | 70 (68.6) | 0.93 (0.51-1.69) | 0.9302 |
| HLA-DQB1*02:01   | 80 (40.4)   | 117 (46.4)           | 0.78 (0.54-1.40) | 0.2368               | 43 (41.3) | 80 (53.3)            | 0.62 (0.37-1.02) | 0.0797               | 37 (39.4) | 37 (36.3) | 1.14 (0.64-2.03) | 0.7657 |
| HLA-DQB1*06:02   | 55 (27.8)   | 69 (27.4)            | 1.02 (0.67-1.55) | 0.9898               | 29 (27.9) | 36 (24.0)            | 1.22 (0.69-2.16) | 0.5813               | 26 (27.6) | 33 (23.3) | 0.80 (0.43-1.48) | 0.5756 |
| HLA-C*01:02      | 63 (31.8)   | 66 (26.2)            | 1.31 (0.87-1.98) | 0.2280               | 32 (30.8) | 34 (22.7)            | 1.49 (0.85-2.62) | 0.2148               | 31 (33.0) | 32 (31.4) | 1.11 (0.61-2.01) | 0.8567 |

OR—odds ratio, CI—confidence interval, Pc—Yates corrected p value

*χ² = 6.3692, uncorrected p = 0.0434
**Discussion**

RHD is an autoimmune disease characterised by persistent inflammatory reaction towards valvular tissue. HLA-G has an immunoregulatory function and could play a vital role in the pathogenesis of immune-mediated diseases, including RHD. To our knowledge, this is the first study to investigate the association of HLA-G 14 bp Ins/Del and +3142 C/G polymorphisms with the pathogenesis of RHD. Hence, we validate our findings with previous reports on various autoimmune diseases such as systemic lupus erythematosus (SLE), rheumatoid arthritis (RA), type 1 diabetes (T1D), idiopathic dilated cardiomyopathy (IDC), and juvenile idiopathic arthritis (JIA) [27–38].

In the present study, the TDT analysis showed that the Del allele was overtransmitted in RHD patients. In addition, the Del/Del genotype and Del allele were overrepresented in pooled RHD patients when compared to healthy siblings. This is in agreement with the previous studies carried out on autoimmune diseases such as T1D, IDC and JIA [33–35]. Increased frequencies of Ins/Ins genotype and Ins allele were observed in female RHD patients, which differ from the previous finding of increased frequency of Del allele among female JIA patients [35]. Increased frequencies of Ins allele and Ins/Ins genotypes were documented in SLE patients [27, 28, 36, 37], however no association of HLA-G 14 bp Ins/Del polymorphism was reported in SLE [31, 38] and RA patients [32, 35].

In this study, the TDT analysis revealed that the +3142 C/G genotype as risk factor in CVL patients

**Table 4** HLA-G genotype, allele and haplotype frequencies in patient sub groups

|                           | Healthy Siblings (126) n (%) | RF 10 n (%) | OR (95% CI) p c | MVL 67 n (%) | OR (95% CI) p c | CVL 22 n (%) | OR (95% CI) p c |
|---------------------------|-----------------------------|-------------|----------------|--------------|----------------|--------------|----------------|
| **14 bp Ins/Del Genotypes** |                             |             |                |              |                |              |                |
| Ins/Ins                   | 27 (21.4)                   | 2 (20.0)    | 0.92 (0.18-4.57) | 0.7680       | 9 (13.4)       | 0.57 (0.25-1.29) | 0.2446       | 5 (22.7)       | 1.08 (0.36-3.19) | 0.8854       |
| Del/Del                   | 36 (28.6)                   | 2 (20.0)    | 0.62 (0.13-3.09) | 0.8294       | 24 (35.8)      | 1.39 (0.74-2.62) | 0.3829       | 9 (40.9)       | 1.73 (0.68-4.40) | 0.3630       |
| Ins/Del                   | 63 (50.0)                   | 6 (60.0)    | 1.5 (0.40-5.57)  | 0.7792       | 34 (50.8)      | 1.03 (0.57-1.86) | 0.9581       | 8 (36.4)       | 0.57 (0.22-1.46) | 0.3421       |
| **Alleles**               |                             |             |                |              |                |              |                |
| Ins                       | 117 (46.4)                  | 10 (50.0)   | 1.15 (0.46-2.86) | 0.9399       | 52 (38.8)      | 0.73 (0.48-1.12) | 0.1837       | 18 (40.9)      | 0.80 (0.42-1.53) | 0.6070       |
| Del                       | 135 (53.6)                  | 10 (50.0)   | 0.87 (0.35-2.15) | 0.9399       | 82 (61.2)      | 1.37 (0.89-2.09) | 0.1837       | 26 (59.1)      | 1.25 (0.65-2.40) | 0.6070       |
| **+3142 C/G Genotypes**   |                             |             |                |              |                |              |                |
| C/C                       | 6 (4.8)                     | 0           | 0               | 0.9250       | 7 (10.5)       | 2.33 (0.75-7.25) | 0.2306       | 5 (22.7)       | 5.88 (1.62-21.39) | 0.0116 a |
| G/G                       | 66 (52.4)                   | 4 (40.0)    | 0.61 (0.16-2.25) | 0.6705       | 34 (50.7)      | 0.94 (0.52-1.69) | 0.9481       | 10 (45.5)      | 0.76 (0.30-1.89) | 0.7124       |
| C/G                       | 54 (42.8)                   | 6 (60.0)    | 2.0 (0.54-7.44)  | 0.4714       | 26 (38.8)      | 0.84 (0.46-1.55) | 0.6962       | 7 (31.8)       | 0.62 (0.24-1.63) | 0.4618       |
| **Alleles**               |                             |             |                |              |                |              |                |
| C                         | 66 (26.2)                   | 6 (30.0)    | 1.21 (0.44-3.27) | 0.9136       | 40 (29.9)      | 1.20 (0.75-1.90) | 0.5174       | 17 (38.6)      | 1.78 (0.91-3.46) | 0.1300       |
| G                         | 186 (73.8)                  | 14 (70.0)   | 0.83 (0.30-2.24) | 0.9136       | 82 (61.2)      | 0.83 (0.52-1.33) | 0.5174       | 27 (61.4)      | 0.56 (0.29-1.10) | 0.1300       |
| **Haplotypes**            |                             |             |                |              |                |              |                |
| Ins/G                     | 117 (46.4)                  | 10 (50.0)   | 1.15 (0.46-2.86) | 0.9399       | 52 (38.8)      | 0.73 (0.48-1.12) | 0.1837       | 18 (40.9)      | 0.80 (0.42-1.53) | 0.6070       |
| Del/G                     | 69 (27.4)                   | 4 (20.0)    | 0.66 (0.21-0.205) | 0.6941       | 42 (31.3)      | 1.21 (0.76-1.91) | 0.4835       | 9 (20.5)       | 0.68 (0.31-1.49) | 0.4372       |
| Del/C                     | 66 (26.2)                   | 6 (30.0)    | 1.21 (0.44-3.27) | 0.9136       | 40 (29.9)      | 1.19 (0.75-1.91) | 0.5174       | 17 (38.6)      | 1.78 (0.91-3.46) | 0.1300       |

RF- Rheumatic Fever, MVL-Mitral Valvular Lesions, CVL-Combined Valvular Lesions, CI-confidence interval, OR-odds ratio, p c-Yates corrected p value

- Bold - significant p value (<0.05)
- a–χ² = 6.3692, Fisher Exact P (P f) = 0.0117

**Table 5** Analysis of Genotype as risk factor in CVL patients

| Study models | OR (95% CI) | p c |
|--------------|------------|-----|
| CC + CG vs GG | 1.32 (0.53-3.28) | 0.7124 |
| CC vs GG     | 0.92 (0.43-1.93) | 0.0261 |
| CC vs GG + CG | 5.88 (1.62-21.39) | 0.0016 |
| CG vs CC + GG | 0.62 (0.24-1.63) | 0.4618 |

CVL- Combined Valvular Lesions, CI-confidence interval, OR odds ratio, p c-Yates corrected p value

- a-Dominant Effect
- b-Additive Effect
- c-Recessive Effect
- d-Co-dominant Effect
analysis revealed that +3142 C/C genotype was significantly associated with risk for CVL. Likewise, the +3142 C allele was associated with risk for clinical subtypes of SLE patients in Japanese population [27]. In contrast, the +3142 G/G genotype and G allele were associated with susceptibility to SLE [28, 29] and RA [30] in Brazilian populations. However, no association of +3142 C/G polymorphism was observed in Brazilian and South Indian RA patients [31, 32].

Our results did not reveal any significant difference in the haplotype frequencies of Ins/G, Del/G and Del/C in the study population. However, increased frequencies of Del/C haplotype was observed in pooled RHD patients and among the patient subgroups. Conversely a previous study reported the association of Del/C haplotype with protection to SLE [29] and RA [30]. Several studies reported that Del/C haplotype is associated with high level of HLA-G expression while Ins/G haplotype is associated with low level of HLA-G expression [39]. In addition, there might be other polymorphic loci in the 3’UTR and promoter region with strong LD which could regulate the HLA-G expression [40, 41] and play a conspicuous role in the development and progression of RHD. Several studies showed contradictory results regarding the influence of HLA-G 3’UTR polymorphisms on HLA-G expression and pathogenesis of various autoimmune and inflammatory diseases [27–38, 42–49]. Thus it is hard to speculate the association of HLA-G with the pathogenesis of RHD. The limitation of the present study in ascertaining the definite role of HLA-G in RHD could be due to the lack of soluble HLA-G quantification, thereby causing difficulty to correlate the influence of 3’UTR polymorphisms on HLA-G expression.

**Conclusion**

In conclusion, in this pilot study we analysed for the first time the association of HLA G 3’prime UTR polymorphism in the development of RHD and the results suggest that the +3142 C/C genotype may influence the development of RHD and subsequent severity of the disease. While, the 14bp Ins/Del polymorphism is not associated with RHD risk in South Indian population. However, further investigation by increasing the sample cohort with diverse populations are indispensable to elucidate the significance of HLA-G in the etiopathogenesis of RHD.

**Additional file**

**Additional file 1: Figure S1.** Pairwise linkage disequilibrium based on 2 HLA-G Polymorphisms using Haploview 4.2. Red squares represent high pair-wise linkage disequilibrium. The numbers in the individual square are D’ multiplied by 100. A. Healthy siblings B. RHD patients C. Trio families. (DOC 301 kb)

**Abbreviations**

CI: Confidence intervals; CVL: Combined Valvular Lesions; GAS: Group A β-hemolytic Streptococcus; HLA-G: Human Leukocyte Antigen-G; HWE: Hardy-Weinberg equilibrium; IDC: Idiopathic dilated cardiomyopathy; JIA: Juvenile idiopathic arthritis; LD: Linkage disequilibrium; MVL: Mitral valvular lesions; OR: Odds ratio; RA: Rheumatoid arthritis; RF: Rheumatic fever; RHD: Rheumatic heart disease; SLE: Systemic lupus erythematosus; T1D: Type 1 diabetes; TDT: Transmission disequilibrium test; T-reg: Regulatory T cells; UTR: Untranslated region

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**Availability of data and material**

All data generated or analysed during this study are included in this published article [and its supplementary information files].

**Authors’ contributions**

PM collected samples and data, performed genotyping, performed statistical analysis, analysed and interpreted the data, and wrote the manuscript. ES participated in the design of the study, participated in samples and data collection, revised the manuscript. SS participated in samples and data collection, revised the manuscript. MS the corresponding author, did conception, designed and coordinated the study, analysed and interpreted the data, revised the manuscript. All authors read and approved the final manuscript.

**Competing interests**

The authors declare that they have no competing interests.

**Consent for publication**

Not applicable.

**Ethics approval and consent to participate**

The study was approved by ethics committees of Government Rajaji hospital, Madurai and Madurai Kamaraj University, Madurai. All individuals were recruited after obtaining informed consent. For children (<18 years), the consent was duly signed by their parents or guardians.

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