Tephritid-microbial interactions to enhance fruit fly performance in sterile insect technique programs

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Abstract

Background: The Sterile Insect Technique (SIT) is being applied for the management of economically important pest fruit flies (Diptera: Tephritidae) in a number of countries worldwide. The success and cost effectiveness of SIT depends upon the ability of mass-reared sterilized male insects to successfully copulate with conspecific wild fertile females when released in the field.

Methods: We conducted a critical analysis of the literature about the tephritid gut microbiome including the advancement of methods for the identification and characterization of microbiota, particularly next generation sequencing, the impacts of irradiation (to induce sterility of flies) and fruit fly rearing, and the use of probiotics to manipulate the fruit fly gut microbiota.

Results: Domestication, mass-rearing, irradiation and handling, as required in SIT, may change the structure of the fruit flies’ gut microbial community compared to that of wild flies under field conditions. Gut microbiota of tephritids are important in their hosts’ development, performance and physiology. Knowledge of how mass-rearing and associated changes of the microbial community impact the functional role of the bacteria and host biology is limited. Probiotics offer potential to encourage a gut microbial community that limits pathogens, and improves the quality of fruit flies.

Conclusions: Advances in technologies used to identify and characterize the gut microbiota will continue to expand our understanding of tephritid gut microbial diversity and community composition. Knowledge about the functions of gut microbes will increase through the use of gnotobiotic models, genome sequencing, metagenomics, metatranscriptomics, metabolomics and metaproteomics. The use of probiotics, or manipulation of the gut microbiota, offers significant opportunities to enhance the production of high quality, performing fruit flies in operational SIT programs.

Keywords: Tephritidae, SIT, Gut microbiota, Gut microbiome, Host-microbe interaction, Insect microbial symbiosis, Microbial symbiont, Probiotics, Mass-rearing

Background

Worldwide, fruit flies (Tephritidae) annually cause substantial damage to horticultural crops, and limit domestic and international trade. Some of the most economically important tephritids include the Mediterranean fruit fly (Ceratitis capitata), oriental fruit fly (Bactrocera dorsalis) and Queensland fruit fly (Bactrocera tryoni). Sterile Insect Technique (SIT) is currently employed in a number of countries to prevent, suppress, contain or eradicate targeted pest species, including tephritid fruit flies [1]. SIT is most successful in an area wide - integrated pest management (AW-IPM) scenario, or geographic isolation [2, 3], and when used in conjunction with other management techniques [4, 5]. The success of SIT depends on irradiated sterile male insects effectively locating, attracting and successfully copulating with wild females [6]. This approach has several advantages including that it is sustainable, has low impact on the environment, does not involve insecticides, and is target-specific.

Fruit fly domestication, irradiation, mass-rearing and handling reduce the fitness, performance and longevity
of flies used in SIT programs, thereby reducing the effectiveness of SIT and its cost-benefit ratio [7–9], Behavioural and physiological changes of mass-reared sterile males, such as changes in mating time and duration, ability to join leks, courtship rituals, pheromone production and attractiveness compared to wild fertile males, dramatically affect copulatory success with wild females [8, 10]. Post-mating factors, such as ejaculate transfer and the inability to prevent re-mating, also influence copulatory success [11]. To overcome the typically low copulatory success of sterile males, a larger number of sterile flies are released, relative to the number of wild flies in the field [10, 12], resulting in high mass-rearing costs. Understanding the biology, ecology and behaviour of fruit flies and the effects of domestication, mass-rearing, handling and sterilization of target pest species allows optimization, and improves the cost, efficiency and effectiveness of SIT.

The gut microbiome greatly influences insect health and homeostasis [13, 14]. The symbiotic association of tephritids with bacteria has been recognized for over a century [15], but our appreciation of the importance and complexity of tephritid-microbial symbiont interactions has increased considerably over the last 35 years. Studies removing, or significantly reducing tephritid gut microbiota through antibiotics indicate that microbiota can positively influence various aspects of tephritid biology, such as nitrogen metabolism, longevity, reproduction, fecundity and overcoming phenolic fruit compounds [16–20]. For example, in contrast to antibiotic-fed (asymbiotic) adult olive fly (Bactrocera oleae), untreated flies were able to utilize inaccessible sources of nitrogen, and bacteria assisted in the provision of missing essential nutrients to the host [20]. Offspring of antibiotic-fed field caught B. oleae females failed to complete larval development in unripe olives unlike larvae of untreated females; however, both were able to complete development in ripe olives. Therefore, it was postulated that symbiotic bacteria help overcome the phenolic compounds in unripe olives [19]. A less intuitive example was found for C. capitata. Adults of this species treated with antibiotics and fed a sugar-only adult diet had significantly increased longevity compared to non-antibiotic-treated flies on the same diet; however, the same effects were not seen when the flies were fed a full adult diet (sugar and yeast hydrolysate) [17]. The authors suggested that the antibiotics may be aiding the immune system against non-beneficial gut microbiota of nutritionally stressed flies [17]. Further, an important trait that maintains gut microbiota in flies is their transmission across generations. Female tephritids coat the egg surface with bacteria prior to, or during oviposition, which aids larval development [21–25]. Fitt and O’Brien [26] found surface sterilization of eggs significantly reduced larval weight (3 mg) at 10 days, while larvae from eggs that were not surface sterilized grew normally, weighing about 15 mg. Studies adding symbiotic bacteria to artificial larval diets significantly improved the development and fitness of domesticated fruit flies [26–28]. Thus, the tephritid-microbe symbiotic fly is their transmission across generations. Female tephritids coat the egg surface with bacteria prior to, or during oviposition, which aids larval development [21–25]. Fitt and O’Brien [26] found surface sterilization of eggs significantly reduced larval weight (3 mg) at 10 days, while larvae from eggs that were not surface sterilized grew normally, weighing about 15 mg. Studies adding symbiotic bacteria to artificial larval diets significantly improved the development and fitness of domesticated fruit flies [26–28]. Thus, the tephritid-microbe symbiotic relationships are very intricate and of significant ecological and evolutionary importance. Increasing our knowledge of these relationships may identify ways to enhance performance of insects that are mass-reared for SIT programs.

Our review focuses exclusively on tephritid gut symbionts, excluding intracellular endosymbionts, such as Wolbachia, which may also be detected in insect gut microbiome studies [29]; however, a previous study suggested that fewer tephritid species than expected harbour Wolbachia [30]. While previous review papers have mostly focused on specific tephritid species [31, 32], or progress in understanding the function of tephritid gut microbiota [33, 34], our review examines recent progress on methods and identification of tephritid microbial symbionts, the impact of the domestication process and irradiation on tephritid-microbial symbiont associations and the use of probiotics to manipulate the fruit fly gut microbiota and consequently gut health.

**Tephritid gut microbiota**

**Influence of methodology and sampling design**

Current characterization techniques of tephritid gut microbial communities have advantages and limitations. Culture-dependent approaches select for microbes capable of growing under culturing conditions, with a large number of bacterial diversity still unculturable. Molecular methods enable the detection of both culturable and unculturable bacteria, rare bacteria and other difficult to culture microorganisms. Molecular approaches used in tephritid gut microbiome studies have targeted the 16S rRNA gene, and are rapidly expanding our knowledge of tephritid gut bacteria. Indeed, sequencing of 16S rRNA gene amplicons from DNA extracted from oesophageal bulbs of B. oleae, enabled the identification of the unculturable symbiont “Candidatus Erwinia dacicola” [35] that assists larvae developing in unripe olives to overcome the plant’s chemical defense mechanism [19].

Tephritid 16S rRNA gene NGS microbiome studies provide a more comprehensive view of fruit fly gut bacterial communities than earlier methods; however, in general each microbiome study employing NGS needs to be interpreted with some caution [36]. For example, 16S rRNA gene amplicon NGS of wild and laboratory-reared tephritids (larvae and adults) have found up to 24 operational taxonomic units (OTUs) at 97% sequence similarity [19, 20, 22, 37] (Table 1). These studies indicate that the tephritid microbiome is low in diversity, similar to that of Drosophila [43, 44]. However, two studies have reported much higher numbers of OTUs (97% similarity) when studying the gut microbiome of tephritid fruit fly
| Study                   | Fly Species | Wild (i.e. field) or Domesticated (i.e. laboratory) | Life Stage | Tissue                | Sequencing Method | Primers | Pipeline | Number of Samples | Number of Reads after Quality Control | OTUs (97% similarity) |
|------------------------|-------------|---------------------------------------------------|------------|-----------------------|-------------------|---------|----------|-------------------|---------------------------------------|----------------------|
| Morrow et al. [37]     | B. tryoni   | laboratory adult whole flies                      |            |                       | 454 Pyrosequencing | 341f 806r | QIIME    | 3 pools of 8      | 3019, 5994, 5991                       | 10,714               |
|                        | B. tryoni   | field (citrus) adult whole flies                  |            |                       |                   |         |          | 1 pool of 8       | 6133                                  | 16                   |
|                        | B. neohumeralis | laboratory adult whole flies                |            |                       |                   |         |          | 3 pools of 8      | 4761, 6741, 6344                       | 6, 18, 14            |
|                        | B. jarvisi  | laboratory adult whole flies                      |            |                       |                   |         |          | 1 pool of 8       | 13,200                                | 7                    |
|                        | B. cacuminata | laboratory adult whole flies                  |            |                       |                   |         |          | 1 pool of 8       | 8202                                  | 4                    |
|                        | B. cacuminata | field (wild tobacco) adult whole flies            |            |                       |                   |         |          | 1 pool of 8       | 7357                                  | 8                    |
|                        | C. capitata | laboratory adult whole flies                      |            |                       |                   |         |          | 1 pool of 8       | 7134                                  | 1                    |
|                        | D. pornia   | laboratory adult whole flies                      |            |                       |                   |         |          | 1 pool of 8       | 8022                                  | 17                   |
| Aharon et al. [22]     | C. capitata | field (apricot) adult midgut                     |            |                       | 454 Pyrosequencing | 926f 1392r | Mothur   | 5 individual adults | 5000-12000                           | 5-23                 |
|                        | C. capitata | field (apricot) larval midgut                    |            |                       |                   |         |          | 15 pooled as 3 samples | 1700-3200                           | 7-13                 |
| Andongma et al. [38]   | B. dorsalis | field (healthy fruit) eggs                       |            | whole egg             | 454 Pyrosequencing | 27f 533r | Mothur   | approx. 50 insects pooled as above | Total reads 46332 (lowest number of reads per sample was 5967) | 76                   |
|                        | B. dorsalis | field (healthy fruit) first instar whole larva    |            |                       |                   |         |          | as above          | 77                                    | 77                   |
|                        | B. dorsalis | field (healthy fruit) third instar whole larva    |            |                       |                   |         |          | as above          | 81                                    | 81                   |
|                        | B. dorsalis | field (fallen fruit) third instar whole gut (proventriculus to rectum) | | | | | | | | |
|                        | B. dorsalis | field (3rd instar larva collected from fallen fruit allowed to pupate in a laboratory) pupae whole pupa (without puparium) | | | | | | | | |
|                        | B. dorsalis | field (ME traps) adult (F)                       |            | whole gut (proventriculus to rectum) | 454 Pyrosequencing | 27f 533r | Mothur   | 4 individuals (ripe olives) | as above | 59 |
|                        | B. dorsalis | field (protein traps) adult (M)                  |            | whole gut (proventriculus to rectum) | 454 Pyrosequencing | 27f 533r | Mothur   | 5 individuals (unripe olives) | as above | 54 |
| Ben-Yosef et al. [19]  | B. oleae    | field (unripe and ripe ‘Souri’ olives) third instar gastric caeca at the proximal section of the midgut | | | | | | | | |
|                        | B. oleae    | field (ovipositing in unripe and ripe ‘Souri’ olives) adult (F) midgut and esophageal bulb | | | | | | | | |

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| Study                | Fly Species | Life Stage | Tissue | Sequencing Method | Primers | Pipeline | Number of Samples | Number of Reads after Quality Control | OTUs (97% similarity) |
|----------------------|-------------|------------|--------|-------------------|---------|----------|-------------------|---------------------------------------|-----------------------|
| Ben-Yosef et al. [20] | B. oleae    | field (green “Manzanillo” olives) | third instar | midgut and esophageal bulb | 454 Pyrosequencing | 92f 1392r | Mothur | 5 individuals | 8351 - 15,098 sequences per sample | 1-2 per sample |
|                      |             | field (soil under olive trees) | pupae | midgut and esophageal bulb |         |          | 5 individuals | 6596,18,335 sequences per sample | 1-3 per sample |
|                      |             | field collected pupae eclosed in cage | adult (F) | midgut and esophageal bulb |         |          | 5 individuals | 8344 - 12,599 sequences per sample | 1-2 per sample |
| Wang et al. [39]     | B. minax    | field (citrus) | adult (F) | intestine | 454 Pyrosequencing | 343f 798r | DOTUR | 3 pools of 15 | 7857 | 319 |
|                      |             | field (citrus) | adult (F) | ovaries | 454 Pyrosequencing | 343f 798r | DOTUR | 3 pools of 15 | 8124 | 415 |
|                      |             | field (citrus) | adult (M) | intestine | 454 Pyrosequencing | 343f 798r | DOTUR | 3 pools of 15 | 7353 | 322 |
|                      |             | field (citrus) | adult (M) | testes | 454 Pyrosequencing | 343f 798r | DOTUR | 3 pools of 15 | 8957 | 389 |
| Yong et al. [40]     | B. carambolae | field (ME trap) | adult (M) | whole fly | Illumina MiSeq | 341f | MEGAN5 | 4 individuals | 1,561,203 – 2,077,403 | 44-75 genera |
|                      | B. dorsalis | field (ME trap) | adult (M) | whole fly | 518r |         | 2 individuals | 1,584,084 – 1,607,064 | 55-75 genera |
| Ventura et al. [41]  | A. ludens   | field (bitter orange) | third instar | whole gut | 454 Pyrosequencing | 8f 556r | QIIME | 30 insects pooled | 110,073 reads | 72 |
|                      | A. obliqua  | field (CeraTrap*) | adult | whole gut | 454 Pyrosequencing | 8f 556r | QIIME | 30 insects pooled | 110,073 reads | 72 |
|                      | A. serpentina | field (mamey sapote) | third instar | whole gut | 454 Pyrosequencing | 8f 556r | QIIME | 30 insects pooled | 110,073 reads | 72 |
|                      | A. striata  | field (guava) | adult | whole gut | 454 Pyrosequencing | 8f 556r | QIIME | 30 insects pooled | 110,073 reads | 72 |
| Malacrinò et al. [42] | C. capitata | field (orange) | first instar | whole larvae | Illumina MiSeq | 515f 806r | QIIME | 15 individuals | not specified | a total of 3, 169 |
|                      |             | field larvae pupate in laboratory (orange) | third instar | whole larvae | 454 Pyrosequencing | 92f 1392r | Mothur | 5 individuals | not specified | a total of 1, 118 |
|                      |             | from field collected larvae (orange) | pupae | whole pupae | 454 Pyrosequencing | 92f 1392r | Mothur | 5 individuals | not specified | a total of 1, 118 |
|                      |             | field (fig) | third instar | whole larvae | 454 Pyrosequencing | 92f 1392r | Mothur | 5 individuals | not specified | a total of 1, 118 |
Table 1 Summary of methodologies employed and results (reads and OTUs) of tephritid NGS microbiome studies (Continued)

| Study     | Fly Species | Wild (i.e. field) or Domesticated (i.e. laboratory) | Life Stage | Tissue       | Sequencing Method | Primers | Pipeline | Number of Samples | Number of Reads after Quality Control | OTUs (97% similarity) |
|-----------|-------------|-----------------------------------------------------|------------|--------------|-------------------|---------|----------|-------------------|---------------------------------------|----------------------|
| field     | (prickly pear) | third instar                                        | whole larvae | as above     | as above          |         |          |                   |                                       |                      |
| field     | (peach)     | third instar                                        | whole larvae | as above     | as above          |         |          |                   |                                       |                      |
| field     | (cherimoya) | third instar                                        | whole larvae | as above     | as above          |         |          |                   |                                       |                      |
| field     | (orange fruits) | third instar                                       | whole larvae | as above     | as above          |         |          |                   |                                       |                      |

*a* methyl eugenol  
*b* average calculated from all field samples, i.e. both larvae and adults  
*c* >10 reads per OTU and OTU clustered at 98% similarity
samples; up to 322 OTUs for Bactrocera minax [39] and up to 81 OTUs for B. dorsalis [38] within a life stage time point. These large numbers of OTUs may, for example, be due to the number of samples pooled (50 samples were pooled in Andongma et al. [38]), quality trimming and/or clustering algorithms. Differences also appear to arise based on whether OTUs with low read numbers were discarded. For example Ben-Yosef et al. [19] removed OTUs with less than 10 sequences. No such restrictions were put on the total number of OTUs from various life stages reported in Andongma et al. [38]; however, employing the same criteria would result in a reduction in the total number of OTUs from combined life stages studied from 172 to 42. It is unclear whether OTUs with low read numbers were also removed from Wang et al. [39] and whether possible erroneous OTUs due to sequencing artefacts were removed from the pyrosequencing data; such erroneous OTUs were removed in Morrow et al. [37]. Nonetheless, discounting low prevalence organisms may also be risky, as microbes at low titers may be overlooked [45]. Furthermore, the percentage of sequence similarity used to define OTUs can alter the taxonomic microbiome profile. For example at > 97% similarity, larvae and adult C. capitata shared a dominant OTU, but this was not true when OTUs were called at > 98% similarity [22]. In regard to taxonomic resolution, the region of the 16S rRNA gene sequenced and the length of sequences obtained using NGS technologies is another factor that can confound analyses [46–50]. No two tephritid NGS microbiome studies have followed the same sequencing and analytical approaches (Table 1), which can complicate comparisons between studies, thus clear archiving of sequence data and reporting of downstream processing of the data (e.g. scripts) are critical.

Very few common or ‘core’ bacteria at the genus or species level have been identified in tephritid gut microbiome studies. “Ca. E. dacicola” (Enterobacteriaceae) and Acetobacter tropicalis (Acetobacteraceae) have been identified as prevalent and possible ‘core’ bacteria in B. oleae; however, recent NGS studies of gut microbiota in B. oleae have failed to detect A. tropicalis in the samples analyzed [19, 20], possibly due to sampling of different host populations. The identity of core bacteria has probably also been overlooked as often tephritid gut microbiome studies have only analyzed pooled or small numbers (fewer than seven) individual samples, such as Andongma et al. [38], Morrow et al. [37], Ventura et al. [41], Wang et al. [39], Ben-Yosef et al. [19], Ben-Yosef et al. [20] and Yong et al. [40]. Furthermore, analysis of single pools of samples does not provide any information about diversity within a population. An exception is the C. capitata microbiome study by Malacrinò et al. [42], where 15 or more individuals per life stage were analyzed; however, whether any core bacteria were identified was not discussed. Increased studies on the bacterial diversity within and between populations can provide insight into the environmental influences on tephritids.

Tephritid bacterial communities

To date, the majority of studies investigating tephritid gut bacterial communities have focused on adults. Bacteria of tephritid larvae and changes across tephritid ontogeny have been characterized in few studies [19, 22, 38, 42, 51]. Bacterial complexity is lower at larval and pupal stages, but increases during the adult stage [22, 51], and likely reflects that the larval stage is naturally confined to a single fruit. There does not appear to be major differences in the bacterial classes or families present in the larval and the adult stage [22, 38]; however, relative abundances of bacterial families may shift with development [38]. This suggests that adult flies acquire microbiota in the larval and early teneral stages, although changes between life stages may be more pronounced when looking at the bacterial genus and species levels. Unfortunately, in many studies the short NGS reads combined with the polyphyly of Enterobacteriaceae has limited the resolution of taxa to these levels when analyzing them across developmental stages [22]. Current laboratory-based evidence suggests that once acquired, tephritid gut microbiota may remain relatively stable throughout adult fly development. The same bacterial species were still recoverable from a B. tryoni population 13 days after the bacteria were fed to the flies [52]. Furthermore, fluorescently labelled Enterobacter agglomerans and Klebsiella pneumoniae fed to adult C. capitata remained detectable in three successive generations of adult flies [21].

The majority of bacteria associated with tephritids belong to the phyla Proteobacteria or Firmicutes, with the most abundant and prevalent from only a few families. Studies of culturable and non-culturale bacteria of field collected tephritids revealed that Enterobacteriaceae are dominant in the vast majority of tephritids, including C. capitata [22, 37, 51, 53–57], Anastrepha spp. [41, 58], Bactrocera spp. [23, 26, 35, 37, 39, 40, 52, 59–69], Rhagoletis spp. [70, 71], and others. Further, Enterobacteriaceae dominate the bacteria vertically transferred from adult tephritid females to larvae, via coating of the egg surface with bacteria prior to, or during oviposition [21–25]. Morphological characteristics and behaviour of fruit flies, which contribute to both vertical and horizontal transmission of Enterobacteriaceae, suggests that these bacteria play an important role in fruit fly development and physiology.

Known functions of tephritid gut bacteria within the Enterobacteriaceae family include diazotrophy and pectinolysis [20, 22, 51, 53, 72], and the break-down of
chemical host plant defenses [19] and insecticides [73]. However, there does not appear to be a common species or genus within the Enterobacteriaceae family that is consistently found in the studied tephritids or even within a fruit fly species, with the exception of “Ca. E. dacicola”, which is prevalent in all wild B. oleae. This phenotypic plasticity of gut microbiota could indicate that a number of bacteria can perform similar roles, which are conserved at higher taxonomic levels, and are interchangeable, thereby allowing tephritids to adapt to diverse diets, and changing bacterial communities.

Other commonly reported Proteobacteria belong to the families Pseudomonaceae and Acetobacteraceae. Pseudomonaceae are present in a number of tephritid species. For example, *Pseudomonas* constitutes a minor but stable community within the gut of *C. capitata*; however, at high densities *Pseudomonas aeruginosa* significantly reduces *C. capitata* longevity [54]. Therefore, the role of *Pseudomonas* spp. in tephritids remains unclear. The acetic acid bacteria *A. tropicalis* was reported as a major symbiont in *B. oleae* via a specific end-point PCR, but, as mentioned earlier, has not been detected in *B. oleae* 16S rRNA gene amplicon NGS studies [19, 20]. Acetobacteraceae have also been reported at low levels in other adult tephritids, but were highly abundant in a single pool of adult female *Dirioxa pornia* [36], a tephritid species with a particular ecological niche, infesting and developing in damaged and fermenting fallen fruit. Apart from research into *A. tropicalis* in *B. oleae*, very little attention has been given to the presence of acetic acid bacteria in tephritids, even though such bacteria are frequently reported as symbionts of insects that have a sugar-based diet within the orders Diptera (including *Drosophila* fruit fly species), Hymenoptera and Hemiptera [74].

Firmicutes constitute part of the microbiota of most adult *Bactrocera* spp. studied to date. Bacteria of the order Bacillales have been reported in *Bactrocera zonata* [68], and in *B. oleae* [75], and bacteria of the order Lactobacillales have been identified in *B. tryoni* [37, 64, 65], *B. minax* [39], *Bactrocera cucuminata* [64], *Bactrocera neohumeralis* [37], *B. oleae* [75] and *B. dorsalis* [38, 62]. Firmicutes have not frequently been reported for *C. capitata*, although *Leuconostoc* were recently detected in the *C. capitata* NGS microbiome study by Malacrinò et al. [42]. Lactobacillales were more common in laboratory-reared than field collected *Bactrocera* spp. flies [37]. Most Firmicutes stain Gram positive, and Gram positive bacteria are known to possess a number of mechanisms that increase their survival in acidic environments [76]. This could increase their tolerance of the low pH of larval diets, and, therefore, be carried on to the adult stage. In addition, some lactic acid bacteria are known to produce antimicrobial peptides [77], which may influence the presence of other bacteria in the diet and gut. The function of lactic acid bacteria in tephritids remains unknown.

**Fruit fly rearing in an artificial environment impacts on gut microbiota**

Fruit flies reared in an artificial environment are not exposed to bacteria typically found in their natural habitat, including microbes that could confer fitness benefits. Artificial tephritid adult diets used for mass-rearing (colony maintenance, not pre-release diets) normally only comprise sugar and yeast hydrolysate; while larval diets typically comprise a bulking agent, yeast, carbohydrates (in the form of sugar or other carbohydrates either added, or within the bulking agent) and antimicrobial agents, such as antifungal and antibacterial agents [78]. While the antimicrobial agents and pH of the larval diet reduce the possibility of contamination with detrimental microorganisms, they may also reduce the opportunities for horizontal transmission of beneficial microbes. Similarly, egg collection methods that rely on water as a transfer medium, and handling methods (e.g. bubbling at temperatures to induce female mortality; required for temperature sensitive lethal strains to produce male only flies under SIT programs), may allow the wider spread of pathogenic bacteria across cultures, and also reduce the vertical transmission of beneficial microorganisms from the adult through to the larval stage.

Consequently, tephritid rearing can change gut microbial communities by reducing bacterial diversity relative to field-collected specimens [19, 24, 37], altering the relative abundance of particular microbes [56] and promoting the acquisition of bacterial species not commonly found in field flies [19, 37]. Mass-reared larvae also have a lower bacterial load than their wild counterparts; larvae from mass-reared olive flies developing in olives have a comparable bacterial load to larvae from field-collected olive flies treated with antibiotics [19]. In addition, olive flies fed an artificial diet have been shown to specifically lack the bacterial symbiont “Ca. E. dacicola”, found in wild flies [59], while artificially reared olive flies fed on olives retain the symbiont [19]. This bacterium allows larvae to develop in unripe olives by counteracting the effects of the phenolic glycoside oleuropein [19]. Although this function is no longer necessary for olive flies not reared on olives, “Ca. E. dacicola” can also accelerate larval development, perhaps through the provision of nitrogen [19]. In contrast, mass-reared adult female olive fly guts were dominated almost exclusively by *Providencia* spp. [19]. Similarly, while *Pseudomonas* spp. occur at only low levels in field collected *C. capitata* (~0.005% of total gut bacteria) [54], they can constitute more than 15% of the total gut bacterial population of mass-reared adult Vienna 8 *C. capitata* [56]. The relative
abundance of Enterobacteriaceae in laboratory-reared adult B. tryoni colonies was reduced compared to field collected B. tryoni; however, only three pools of laboratory-reared B. tryoni from different populations were compared to just one pool of field collected B. tryoni, and only females were analyzed [37]. Laboratory rearing also influences the abundance of lactic acid bacteria, such as Lactococcus, Vagococcus and Enterococcus in some Bactrocera laboratory-adapted flies, which do not tend to be present in high densities in wild flies [37].

The gut microbiota of fruit flies also become very similar and ‘streamlined’ when maintained on the same diet within a location. Adult B. tryoni, sourced from different locations maintained on the same larval and adult diets, in the same laboratory, possessed similar microbiota [37]. Indeed, similar bacteria where also identified from B. neohumeralis laboratory-adapted colonies, which were established 3 years apart but rear within the same facility [37]. Interestingly, the gut microbiome profile of B. neohumeralis differed between populations reared in different laboratories, suggesting an environmental influence on the bacteria associated with artificially reared-adult fruit flies. Identifying the factors driving changes in tephritid gut microbiota, such as age, diet, environment and genetics, is important to identify ways to minimise, or even avoid, unwanted microbial changes, and optimise the gut ecology of mass-reared tephritids.

When domesticated flies are stressed due to nutrition, overcrowding, increased waste products, exposure to larger densities of particular bacteria and genetic changes, this could influence fly susceptibility to pathogens. For example, Serratia marcescens is pathogenic to Rhagoletis pomonella [79] and to Drosophila melanogaster [80, 81]. Lloyd et al. [69] found that Enterobacteriaceae, such as Klebsiella, Erwinia and Enterobacter, were frequently cultured from field collected B. tryoni, while S. marcescens and Serratia liquefaciens were dominant in laboratory flies, which may have been introduced by artificially reared-adult fruit flies. Identifying the factors driving changes in tephritid gut microbiota, such as age, diet, environment and genetics, is important to identify ways to minimise, or even avoid, unwanted microbial changes, and optimise the gut ecology of mass-reared tephritids.

The general hypothesis is that microbial diversity contributes to healthier flies and that observed taxonomic differences in artificially reared flies result in less resilience and increased sensitivity to environmental changes due to decreased bacterial diversity and perhaps decreased functional diversity. Little is known about the relationship between the structure of fruit fly bacterial communities and functional diversity and the impact of taxonomic differences at the functional level. Analytical approaches such as metagenomics, metatranscriptomics, metabolomics and metaproteomics will facilitate significant progress in this area as they will permit better characterization of microbial communities, their function and contribution to host development, fitness and performance.

**Effect of irradiation**

Fruit flies to be released in SIT operations are typically sterilized as pupae using gamma irradiation [84]. Lauzon and Potter’s [85] comparison of irradiated versus non-irradiated C. capitata and A. ludens midguts using electron microscopy showed that irradiation has an effect on both the gut microbiota and the development of the midgut epithelium. Transmission electron microscope
measurements of tephritid fruit fly fitness and performed positive outcomes for the host (Fig. 1).

Results. Substantial changes are not always observed (disbiotics, added to tephritid diets, on the host, with mixed bacterial supplements, more recently referred to as probiotics, to domesticated tephritids may provide increased or even additional benefits. Therefore, any probiotic study needs to be well replicated, or a sufficient number of samples included due to the complexity of such studies. In addition, any trade-offs (if observed) need to be assessed, for example, against improved mating performance, as to their importance in SIT effectiveness.

The addition of symbiotic bacteria to the larval and adult fruit fly diets changes the structure of fruit fly gut bacterial communities (Fig. 1). Indeed, adding a probiotic supplement cocktail containing *Klebsiella pneumoniae*, *Enterobacter* sp. and *Citrobacter freundii* to the *C. capitata* larval diet simultaneously increased the number of Enterobacteriaceae in the larval and adult gut and reduced the number of *Pseudomonas* spp. present at both the larval and adult stages [27]. Similarly, feeding *Klebsiella oxytoca* to adult Vienna 8 strain *C. capitata* increased the abundance of *K. oxytoca* in the gut, and reduced the number of *Pseudomonas*, *Morganella* and *Providencia* spp. [56]. It is hypothesized that the gut Enterobacteriaceae community of *C. capitata* can control the density of bacteria that are harmful in high abundance, such as *Pseudomonas aeruginosa* [54].

The majority of tephritid probiotic studies have involved the addition of bacteria to the adult diet, and while, the observed impacts on the host have been variable, the positive impacts are encouraging for their potential application in SIT programs (Fig. 1). Sterile male *C. capitata* fed a sugar diet enriched with *K. oxytoca* compared to flies fed a sugar only diet, showed increased on immediate benefits. Therefore, it is possible that other impacts, such as changes in the expression of host immune response genes, and genes involved in signalling and/or metabolism, have been overlooked. Negative impacts have been observed in probiotic fed adult *B. oleae*, where a reduction in longevity has been observed; however, whether this appears to be influenced by the diet the adult flies are feeding on (i.e. sugar versus sugar and protein diet) or the bacteria remains unclear [99]. The bacterial species fed to the adult fly can also influence longevity [90]. Thus, benefits provided by probiotics are not always consistent between studies, most likely due to the complexity of tephritid-bacteria interactions. Further, other factors are likely to influence results including variations in experimental design, probiotic supplements tested and their delivery (dose, mode), experimental conditions, traits measured on varying life stages, irradiated or non-irradiated flies, pre-existing microbiota in experimental flies, diet (nutritional value, antimicrobials, agar versus granular), rearing environment, age and genetic diversity of experimental colonies. As the wild tephritid gut microbiome is often comprised of diverse microbiota, it is feasible that the addition of more than one probiotic candidate, i.e. bacterial blends/consortiums, to domesticated tephritids may provide increased or even additional benefits. Therefore, any probiotic study needs to be well replicated, or a sufficient number of samples included due to the complexity of such studies. In addition, any trade-offs (if observed) need to be assessed, for example, against improved mating performance, as to their importance in SIT effectiveness.

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mating competitiveness in both laboratory and field cages, reduced female remating (presumably in a laboratory setting), and increased survival under starvation in the laboratory [92]. Similarly, a mating advantage was conferred in laboratory studies of *C. capitata* fed *Enterobacter agglomerans* and *K. pneumoniae* in a yeast-enhanced agar compared to non-bacterial inoculated yeast-enhanced agar, but no significant effect was observed with a sugar-yeast or sugar-reduced yeast granular diet [94]. Conversely, mating competitiveness studies in field cages only found significantly more matings (with wild/F1-F15 laboratory-reared flies) than the control when the flies were fed yeast-reduced sugar granulate diet [94]. While mating was not assessed, Meats et al. [93] detected no evidence of either, *K. oxytoca* or *K. pneumoniae* added to the adult *B. tryoni* diet (paste of sugar and autolysed yeast) impacting on egg production regardless of whether the fly generation was F0-F20; however, as expected (presumably due to laboratory adaptation) regardless of bacterial supplementation, egg
production increased as the fly generation increased. The addition of *Pseudomonas putida* to the sugar diet of *B. oleae* increased female fecundity compared to females fed a sugar only diet [99]. However, *P. putida* added to a complete diet (comprising of sugar and hydrolysed brewer’s yeast) had no significant effect on fecundity compared to the same diet without added *P. putida* [99]. These studies indicate that bacteria contribute to fly nutrition, although not exclusively (see next paragraph). It is possible that when the flies are provided with a nutritionally balanced diet, i.e. the amount of yeast, providing fatty acids, amino acids and vitamins, is adequate, the effect of a probiotic supplement is minimal, but this would be dependent on the influence of nutrition on the trait being measured. Thus, the role of the gut microbiome may have largely been underestimated in nutritional studies, and it is possible that through adding bacterial supplements to the fruit fly diet, the amount of yeast required could be reduced. Further, other components of the gut microbiome, such as yeasts, which can also contribute to the host nutrition, have largely been overlooked until recently [100].

Several studies have investigated the impact of feeding autoclaved bacteria, which by definition are not classed as a probiotic, to tephritids [28, 56, 92]. Autoclaved *Enterobacter* sp. added to the *C. capitata* larval diet significantly reduced egg to adult developmental time [28]. This study suggests that bacterial mass and/or bacterial substrates can have a positive nutritional effect on immature *C. capitata*. However, studies comparing the use of autoclaved bacteria to live bacteria show that the contribution of live bacteria to the host is greater than just the nutritional value of dead bacteria themselves and what they produce in culture. The addition of autoclaved *K. oxytoca* to the *C. capitata* adult pre-release sugar/sucrose only diet did not improve mating performance [92] or mating latency [56], in contrast to a diet supplemented with live *K. oxytoca*. The nutritional benefits observed when using an autoclaved, or live culture may be due to metabolites produced by the bacteria; it is not known what metabolites are being produced by tephritid gut bacteria and what effect they have on the gut microbiome and the host. In *Drosophila*, the metabolite acetate, a product of pyrroloquinoline quinone–dependent alcohol dehydrogenase (PQQ-ADH) by the commensal gut bacterium, *Acetobacter pomorum*, modulates insulin/insulin-like growth factor signalling, which is important for normal larval development [101]. Gut microbiota and their metabolites will be an exciting area of research to follow in the future, particularly with the development of tools such as metabolomics.

Although only a few studies have investigated the effects of adding probiotic supplements to the larval diet, the results have revealed a number of benefits. Addition of *Enterobacter* spp., *K. pneumoniae* and *C. freundii* to the wheat bran larval diet increased pupal weight of *C. capitata* [28, 56, 92]. *Enterobacter* sp. to the larval carrot-based diet improved egg-pupal and egg-adult recovery of *C. capitata* [28, 56, 92]. Reduced developmental time is a considerable advantage in mass-rearing facilities leading to cost savings and increased production. The benefits observed at the larval stage could have flow-on effects to the pupae and to adult morphology, fitness and performance. Thus, there is a need to increase our understanding of the influence of each life stage on successive stages and generations, particularly considering the vertical transmission of microbiota.

The presence of beneficial microbes in the larval diet may allow a reduction in the added amount of antimicrobials. Some yeasts possess antagonistic properties against undesirable bacteria [102]. Four studies have cultured yeasts from field collected tephritid fruit fly larvae (*B. tryoni* and *Anastrepha mucronota*) indicating that they consume yeasts while feeding within fruit [100, 103–105]. Thus, the incorporation of live yeasts, rather than pasteurized yeasts, for example, into the larval diet may be a way to reduce the amount of antimicrobials in the diet and warrants further testing. The interaction between bacteria and yeasts in the gut is an unexplored area in tephritid fruit fly research.

The development of a tephritid gnotobiotic model system that allows the addition and manipulation of flies, which have either developed under axenic conditions, or for which all present microbiota are known, would enable the better examination and verification of host-microbe relationships. Surface sterilization of eggs would remove the transmission of gut microbiota transferred with the egg during oviposition, and the larvae that emerge can then be used in an axenic system. This would help avoid non-microbial effects that could derive from the use of antibiotics, such as effects on mitochondrial respiration [106].

**Conclusion**

While significant progress has been made towards the taxonomic characterisation and profiling of gut microbial populations in tephritids, there are still considerable gaps in our knowledge of tephritid-bacteria interactions. Improvements in NGS technologies and bioinformatics, in combination with decreased costs, will improve our knowledge of gut microbial diversity and potentially identify further key bacterial and other microbial symbionts. However, the largest unknown factors remain with
the functional roles of the microbial symbionts. Use of gnotobiotic models, genome sequencing, metagenomics, metatranscriptomics, metabolomics and metaproteomics will help in defining precise roles of gut microbes. To maintain tephritid-microbial symbiont interactions during the mass-rearing process, we need to understand how such interactions evolve and how both irradiation [107] and the domestication process [108], including diet, disrupts the relationship and associated bacterial functions. This will inform the development of ways to encourage, maintain or introduce symbiotic microbes in the rearing process to produce better performing, and cost-effective flies for SIT programs. Microbial symbionts, whether through the administration of larval and/or adult probiotics, or the maintenance of a healthy gut microbiome through dietary and environmental manipulation, may well be the next major improvement to fruit fly mass-rearing.

Abbreviations

AW-IPM: Area wide - integrated pest management; GSS: Genetic sexing strain; NGS: Next-generation sequencing; OTU: Operational taxonomic unit; PQQ-ADH: Pyrroloquinoline quinone–dependent alcohol dehydrogenase; SIT: Sterile insect technique

Acknowledgements

We thank Cheryl Jenkins for comments on an earlier version of the manuscript. We thank Anne Johnson for formatting sections of this work.

About this supplement

This article has been published as part of BMC Microbiology Volume 19 Supplement 1, 2019: Proceedings of an FAO/IAEA Coordinated Research Project on Use of Symbiotic Bacteria to Reduce Mass-rearing Costs and Increase Mating Success in Selected Fruit Pests in Support of SIT Application: microbiology. The full contents of the supplement are available online at https://bmcmicrobiol.biomedcentral.com/articles/supplements/volume-19-supplement-1.

Authors’ contributions

AD drafted and wrote the manuscript with input from OR, MR, & TC. LS developed Fig. 1 with input from all authors. All authors read and approved the final manuscript.

Funding

This project has been funded by Hort Innovation using the summer fruit industry levy with co-investment from NSW Department of Primary Industries and funds from the Australian Government as part of the SITplus initiative. Hort Innovation is the grower owned, not-for-profit research and development corporation for Australian horticulture.

Availability of data and materials

Not applicable.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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Published: 24 December 2019

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