Review
Extracellular Vesicles, Inflammation, and Cardiovascular Disease

Akbarshakh Akhmerov 1,* and Tanyalak Parimon 2

1 Smidt Heart Institute, Cedars-Sinai Medical Center, Los Angeles, CA 90048, USA
2 Women’s Guild Lung Institute, Pulmonary and Critical Care Division, Department of Medicine, Cedars-Sinai Medical Center, Los Angeles, CA 90048, USA; tanyalak.parimon@cshs.org
* Correspondence: akbarshakh.akhmerov@cshs.org

Abstract: Cardiovascular disease is a leading cause of death worldwide. The underlying mechanisms of most cardiovascular disorders involve innate and adaptive immune responses, and extracellular vesicles are implicated in both. In this review, we describe the mechanistic role of extracellular vesicles at the intersection of inflammatory processes and cardiovascular disease. Our discussion focuses on atherosclerosis, myocardial ischemia and ischemic heart disease, heart failure, aortic aneurysms, and valvular pathology.

Keywords: extracellular vesicles; exosomes; microvesicles; inflammation; innate immunity; adaptive immunity; cardiovascular disease; cardiac disease; heart disease; myocardium

1. Introduction
Cardiovascular disease is a leading cause of morbidity and mortality, accounting for one-third of global deaths in 2019 [1]. Collectively, atherosclerosis, ischemic heart disease, heart failure, aortopathy, and valvular pathology make up most of these deaths (Figure 1). Inflammation plays a significant role in the pathogenesis of these cardiovascular conditions [2–4], and anti-inflammatory therapies have demonstrated benefits in several clinical trials [5,6]. The underlying mechanisms of inflammation in cardiovascular disease are multifaceted, but an emerging body of evidence implicates extracellular vesicles (EVs) as key mediators.

EVs are nano-sized, membrane-bound vesicles released by nearly all cells. EVs are extremely heterogeneous but can be broadly classified based on their size and biogenesis [7]. Exosomes are typically 50–150 nm in size, derived through an endosomal pathway, and released upon exocytosis of multivesicular bodies [8–10]. Microvesicles (also referred to as ectosomes or microparticles) are typically 100–1000 nm in size and are assembled at and released from the plasma membrane [10,11]. Similarly, apoptotic bodies are >100 nm in size but are formed during apoptosis. EVs are crucial in cell-to-cell communication and mediate their effects through bioactive cargo, including proteins, RNA, and lipids. The composition of EVs, however, may change depending on the physiologic state of the parent cell.

The role of EVs in inflammatory pathways and their involvement in cardiovascular disease have been independently discussed in other reviews [12–15]. Here, we review findings that implicate EVs at the intersection of inflammation and cardiovascular disease. As such, we highlight the inextricable interplay between EVs, inflammation, and cardiovascular disease (Graphical Abstract).
2. The Role of EVs and Inflammation in Specific Cardiovascular Disorders

2.1. The Role of EVs and Inflammation in Atherosclerosis

Atherosclerosis is the accumulation of fibrofatty material within an arterial wall, which can narrow or occlude the vessel lumen, impede blood flow, and ultimately result in myocardial infarction, stroke, or peripheral arterial disease [16]. The clinical sequelae of atherosclerosis (e.g., coronary artery disease, stroke) are the leading cause of death worldwide [1]. Although the pathogenesis of atherosclerosis is multifactorial, inflammation plays a central role [17]. Indeed, atherosclerosis is often characterized as a chronic inflammatory disorder.

Atherosclerosis is initiated by endothelial activation and dysfunction, with subsequent accumulation of lipid content (low-density lipoprotein, LDL) in the subendothelial layer of the artery. The lipid material undergoes oxidation and further modifications that propagate inflammatory responses driven by innate and adaptive immunities [16,17]. EVs have been isolated from intimal lesions and are a major determinant of atherosclerotic plaque progression [18]. Plaque analysis in humans showed that most EVs originate from leukocytes, with macrophages accounting for the majority (29%), followed by lymphocytes (15%) and granulocytes (8%) [19]. Smooth muscle cells, platelets, adipocytes, and endothelial cells, however, are equally important sources of EVs [18]. Regardless of their origin, EVs interact with immune cell mediators at all stages of disease progression (Figure 2) [20].
During the initial stages of atherogenesis, plaque EVs transfer the intercellular adhesion molecule 1 (ICAM-1) to endothelial cells, thereby promoting monocyte adhesion and recruitment [21]. Similarly, monocyte-derived EVs induce the expression of ICAM-1 and chemokine C–C motif chemokine ligand 2 (CCL2) by activating the nuclear factor-κB (NF-κB) pathway within endothelial cells [22]. Neutrophil-derived EVs also facilitate recruitment, by enhancing transendothelial migration of monocytes through upregulation of CCL2, ICAM-1, and the vascular cell adhesion molecule 1 (VCAM-1). The proposed mechanism involves activation of the NF-κB pathway by EV-delivered miR-155 [23]. Among non-leukocytes, activated platelets release EVs that facilitate monocyte recruitment by transferring the chemokine RANTES (Regulated upon Activation, Normal T Cell Expressed and Presumably Secreted) to the endothelium, and by transferring a platelet-adhesion molecule GPIbα directly to monocytes [24,25]. In addition, platelet-derived EVs promote neutrophil adhesion to endothelial cells through P-selectins [26]. In the setting of inflammation, adipocyte-derived EVs upregulate the expression of VCAM-1 on endothelial cells and enhance leukocyte attachment [27]. Finally, EVs released from endothelial cells pretreated with reactive oxygen species stimulate endothelial cells to bind monocytes [28]. Taken together, EVs released from activated leukocytes, platelets, endothelial cells, and adipocytes promote inflammatory infiltration. It is important to note, however, that most of these studies were performed in vitro, and further investigation is needed to firmly establish these mechanistic links.

Beyond the recruitment of leukocytes, EVs play an important role in plaque maturation. The polarization of macrophages and the progression of macrophages to foam cells are important examples. Following LDL deposition in the subendothelial region, reactive oxygen species convert LDL to oxidized LDL, which, in turn, leads to monocyte-to-macrophage differentiation [20,29]. Further uptake of oxidized LDL by macrophages via scavenger receptors leads to the formation of lipid-containing foam cells. Platelet-derived EVs promote phagocytosis of oxidized LDL by macrophages, thereby enhancing foam cell formation [30]. Adipose-derived EVs also enhance foam cell formation, by inhibiting cholesterol efflux from macrophages. Furthermore, these adipose-derived EVs polarize macrophages toward the classic pro-inflammatory M1 phenotype [31]. Similarly, endothelial EVs have the ca-
pacity to polarize macrophages, either toward the M1 phenotype or the anti-inflammatory M2 phenotype, depending on environmental stimuli [20]. Endothelial cells exposed to oxidized LDL secrete EVs containing metastasis-associated lung adenocarcinoma transcript 1 (MALAT1), which promotes M2 polarization [32]. Other studies, however, demonstrated that endothelial cells exposed to oxidized LDL secrete EVs containing miR-155, which shift the phenotype toward the M1 phenotype [33]. Given the potentially competing mechanisms, the overall effect of endothelium-derived EVs on macrophage polarization requires further investigation.

The other important protein involved in LDL metabolism is heparan sulfate proteoglycan (HSPG), which offers a protective role in atherosclerosis. It is a key regulator of LDL retention in endothelial cells and immune cells (macrophages and neutrophils), given its constitutive expression in those cells [34]. HSPG halts the atherosclerotic process by binding to a proliferation-inducing ligand (APRIL), and serum levels of APRIL are associated with cardiovascular mortality in atherosclerotic patients [35]. In addition, HPSG facilitates endocytosis and uptake of LDLRQ722*-containing EVs, improving LDL clearance and preventing atherosclerotic plaque formation [36]. Overall, the presence of HPSG appears beneficial in atherosclerotic blood vessels. On the hand, proteoglycans in the extracellular matrix of the vascular wall may contribute to the pathogenesis of atherosclerosis, but it is unclear whether this process is mediated by EVs [37].

Another crucial component of atherosclerotic plaque progression is calcification, and EVs mechanistically contribute to this process [38]. Analysis of murine and human plaques showed that EVs derived from pro-inflammatory macrophages promote intimal microcalcification [39,40]. Furthermore, enrichment of these EVs with the calcium-binding protein S100A9 facilitates the development of calcification. Smooth-muscle-cell-derived EVs are equally important in promoting microcalcification. The reader is referred elsewhere for a thorough review of these mechanisms [20]. In conclusion, the interplay between EVs derived from leukocytes, platelets, endothelial cells, adipocytes, and smooth muscle cells is central to the pathogenesis of atherosclerosis, and mechanistic insights into this interplay have important therapeutic implications. When atherosclerotic disease progresses, the sequelae include myocardial infarction (MI) and ischemic heart disease.

2.2. The Role of EVs and Inflammation in Myocardial Infarction and Ischemic Heart Disease

Ischemic heart disease is a leading cause of mortality worldwide, with an estimated prevalence of 197 million cases in 2019 [41]. Inflammation and fibrosis play crucial, interconnected roles in the pathophysiology of MI and ischemic heart disease. EVs derived from cardiomyocytes, endothelial cells, and immune cells influence inflammatory and fibrotic responses and can therefore serve as potential diagnostic, prognostic, and therapeutic agents.

Following an acute MI, rapid humoral and cell-mediated responses ensue [42]. In preclinical models, infiltrating monocytes that subsequently differentiate into macrophages are the major cell mediators [43]. The release of EVs within the myocardium appears to correspond to the inflammatory influx, with a significant increase in EVs at 15–24 h following MI [44]. These EVs originate locally from cardiomyocytes and endothelial cells and promote the release of chemokines and inflammatory cytokines from infiltrating monocytes [44]. Similarly, cardiomyocyte-derived EVs modulate the inflammatory phenotype of macrophages, depending on the state of donor cells (i.e., ischemic versus non-ischemic) [45]. The crosstalk between cardiomyocytes also extends beyond the local milieu of the heart (Figure 3). Following acute MI, myocardial micro-RNAs (miRNAs) are selectively shuttled by EVs to peripheral organs. In mice, miR-1, miR-208, and miR-499 are transported by EVs to the bone marrow, where they downregulate the expression of CXC chemokine receptor 4 (CXCR4) in mononuclear cells, thereby allowing mobilization into the circulation [46]. Likewise, endothelial-derived EVs are increased after acute MI and trigger mobilization and activation of monocytes from the spleen [47]. Thus, EVs derived from cardiomyocytes and endothelial cells recruit peripheral mononuclear cells and alter their phenotype. Furthermore, the number of EVs closely correlates with the extent of
myocardial injury [47], suggesting a potential diagnostic and prognostic value of EVs in MI. Interestingly, EVs derived from endothelial cells overexpressing Krüppel-like factor 2 (KLF2) inhibit the recruitment of Ly6<sup>high</sup> monocytes and attenuate ischemia–reperfusion injury following MI [48]. EVs released by cardiac stromal-progenitor cells in mice, rats, and humans also have immunomodulatory effects through macrophage polarization in models of MI, highlighting their therapeutic potential [49–51].

![Figure 3. The role of EVs and inflammation in myocardial infarction.](image)

Importantly, EVs derived from non-cardiomyocytes can modulate the inflammatory and fibrotic responses after MI. In innate immunity, macrophages release various non-coding RNA species that target cardiac fibroblasts. For example, EVs containing the circular RNA circUbe3a exacerbate myocardial fibrosis by altering cardiac fibroblast proliferation, migration, and phenotype (Figure 3) [52]. Activated macrophages also transfer miR-155-laden EVs to cardiac fibroblasts, suppressing their proliferation, promoting inflammation, and ultimately leading to cardiac rupture [53]. Additionally, the transfer of miR-155-containing EVs from macrophages to endothelial cells exerts an anti-angiogenic effect, further exacerbating ischemic injury [54]. On the other hand, EVs derived from cells of innate immunity can have salutary effects in acute MI. Following MI, EVs from dendritic cells mediate the activation of CD4<sup>+</sup> T cells, and thereby improve cardiac function after MI [55]. Therapeutic modulation of T cells in MI has recently gained considerable interest [56–58]. Using regulatory T cells (Tregs) and their EVs for such modulation is a promising strategy. Indeed, EVs released by cardiac stromal-progenitor cells have been shown to potentiate Tregs by increasing their proliferation and IL-10 production, with downstream cardioprotective effects in models of inflammation [59]. Thus, EVs have diagnostic, prognostic, and therapeutic potential in models of MI and ischemic heart disease. Further translational studies are needed to potentiate their clinical value. So far, we have demonstrated that EVs play a significant role in the pathophysiology of atherosclerosis, which, untreated, can lead to ischemic heart disease. Ischemic heart disease, in turn, can progress to ischemic cardiomyopathy and heart failure.

2.3. The Role of EVs and Inflammation in Heart Failure

Heart failure (HF) is a clinical syndrome caused by structural and/or functional abnormalities of the heart [60]. With a global prevalence of 64.3 million in 2017, HF is a major public health concern [61,62]. In the United States, the prevalence of HF is projected to increase to >8 million by 2030 [63]. The proportion of HF patients with reduced ejection fraction (<40%) ranges from 36% to 81%, depending on the region and registry used [62]. The proportion of HF patients with preserved ejection fraction (≥50%) ranges from 16% to 52%, but the prevalence appears to be increasing [64]. Both the left and the right heart...
can fail [65], and the causes can be broadly classified as either ischemic or non-ischemic. The mechanisms behind heart failure are varied and depend on the underlying cause. Nevertheless, EVs have been broadly implicated in several aspects of HF pathophysiology, including those that involve chronic inflammation.

Circulating EVs and their cargo miRNAs have been identified as potential biomarkers and drivers of HF [66–69]. A subset of these EVs originate from the inflammatory processes that accompany the initial insult and may contribute to the progression of HF. In animal models of MI, there is an increase in circulating EVs, carrying IL-1α, IL-1β, and RANTES (Figure 4). The reduction in these pro-inflammatory EVs (originating from M1 macrophages) results in improved cardiac function [70]. Notably, the interaction between cardiomyocytes and macrophages is bidirectional. For instance, in animal models of cardiac hypertrophy, EVs from hypertrophic cardiomyocytes activate macrophages by transferring miR-155 (Figure 4) [71]. This activation triggers the release of pro-inflammatory cytokines, IL-6 and IL-8, from macrophages.

Because inflammation and adverse cardiac remodeling are interrelated [72], the role of fibroblast-derived EVs is equally important. In vitro, fibroblasts stimulated with TNF-α release EVs containing miR-27a, miR-28-3p, and miR-34a, which dysregulate the nuclear factor erythroid 2–related factor 2 (Nrf2) pathway that ordinarily prevents oxidative injury and protects against adverse cardiac remodeling and dysfunction (Figure 4) [73]. Cardiac fibroblasts also secrete EVs containing miR-21* and miR-27a*, which induce cardiac hypertrophy [74–77]. Cardiomyocytes, in turn, secrete miR-217-containing EVs that enhance fibroblast proliferation and drive cardiac hypertrophy [67]. In the context of ischemia, long non-coding RNAs in cardiomyocyte-derived EVs interact with fibroblasts and increase the expression of profibrotic genes [78]. Therefore, the underlying mechanisms in heart failure due to ischemic and hypertrophic etiologies are an active interplay between macrophages, cardiomyocytes, and fibroblasts.

![Figure 4. The role of EVs and inflammation in heart failure. Interaction between cardiomyocytes (CM), macrophages (Mφ), and fibroblasts (FB) through EVs in the pathogenesis of heart failure.](image-url)
2.4. The Role of EVs and Inflammation in Aneurysmal and Valvular Pathology of the Aorta

The prevalence of thoracic aortic aneurysms (TAA) (>5 cm), incidentally noted on CT imaging performed for screening, ranges from 0.2% to 0.3% [63]. The overall prevalence of abdominal aortic aneurysms (AAA) ranges from 4% to 8%, but the prevalence of AAA ≥ 5.5 cm ranges from 0.4% to 0.6% [79–83]. Aortic aneurysms in younger individuals are typically due to hereditary or familial syndromes (e.g., Marfan syndrome, Loeys-Dietz syndrome, bicuspid aortic valve disease), while in older individuals the aneurysms are degenerative and share many risk factors with atherosclerosis [84]. Despite differences in the pathophysiology between TAA and AAs, EVs are clinically relevant in both as diagnostic and prognostic biomarkers and provide important insight into the underlying mechanisms.

Proteomic analysis of human plasma identified differential profiles in patients with aortic aneurysms, including proteins that drive inflammatory processes [85]. Platelets are activated in both TAA and AAs [86,87], and EVs derived from these platelets contribute to the pathogenesis of aortic aneurysms. For example, increased ficolin-3 levels are noted in platelet-derived EVs in patients with aortic aneurysms, compared with healthy controls, and are associated with aneurysm progression [88]. Ficolin-3 is one of the recognition molecules within the lectin pathway that activates the complement system, which, in turn, mobilizes innate immunity [89]. EVs from leukocytes are also identified in aortic aneurysms. Macrophage-derived EVs are found in murine and human adventitia from atherosclerotic tissue. In vitro, these EVs induce the expression of matrix metalloproteinase-2 within vascular smooth muscle cells via the JNK and p38 pathways [90]. Neutrophil-derived EVs may also play a role in aneurysm progression. EVs bearing neutrophil markers are identified within the intraluminal thrombus of aortic aneurysms. These EVs carry active proteases ADAM10 and ADAM17, which contribute to aortic wall degradation [91]. Finally, the interaction between adaptive and innate immunities must be considered in aneurysm development. Activated T lymphocytes within vascular lesions are noted to have increased pyruvate kinase muscle isozyme 2 (PKM2) expression [92]. Importantly, these lymphocytes release EVs that are rich in polyunsaturated-fatty-acid-containing phospholipids and are taken up by macrophages. The activated lymphocyte-derived EVs then promote macrophage migration, lipid peroxidation, and iron accumulation, which drive aortic aneurysm progression. Corollary findings are noted in patients with AAs, highlighting the translational value of these findings [92]. Thus, inflammation plays a prominent role in aortic aneurysm development, especially AAs. EV interactions with non-immune mediators, however, are equally important in understanding the pathophysiology of aortic aneurysms. Specifically, vascular smooth muscle cell biology is central; the reader is referred elsewhere for focused reviews on this topic [84,93].

The prevalence of valvular disease depends on severity. Mild valvular disease is present in 51% of individuals ≥ 65 years of age, and moderate or severe disease is present in 6.4% [63]. The prevalence also varies based on specific valvular pathology, with aortic valve stenosis (AS) noted in 4.3% of individuals ≥ 70 years of age [63]. Chronic inflammation, infiltration of mononuclear cells, and activated EVs are all implicated in AS [94–96]. In a study of 22 patients with severe AS, leukocyte-derived EVs, platelet-derived EVs, and endothelial-derived EVs were all elevated, compared with healthy controls. Furthermore, these elevations were associated with monocyte activation [97]. Indeed, calcified aortic valves in humans have increased infiltration of macrophages that colocalize with metalloproteinases, which are involved in AS pathogenesis [98]. Macrophages may also contribute to valvular microcalcification through mechanisms discussed above in atherosclerotic disease [39,40,99]. Non-leukocyte EVs (e.g., endothelial cells) also play a critical role in aortic valve pathology, and the reader is referred to previous reviews for more details regarding interactions between EVs, valve endothelial cells, and valve interstitial cells [100,101]. The role of proteoglycans as a nidus of calcium accumulation in matrix vesicles is one such example. In conclusion, both aneurysmal and valvular pathologies of the aorta involve complex interactions between leukocytes, non-inflammatory cells, and EVs derived from...
both. Further investigation of these interactions and their mechanisms will yield clinically relevant data.

3. Conclusions

Inflammation is involved in nearly all cardiovascular conditions, contributing to the pathogenesis at multiple stages of the disease. An increasing body of literature implicates EVs derived from leukocytes and non-leukocytes in this pathophysiology. In this review, we summarize the mechanistic role of EVs at the intersection of inflammatory processes and cardiovascular disease. In atherosclerosis, EVs are involved in the recruitment of monocytes, polarization of macrophages, and microcalcification, which together contribute to plaque formation, maturation, and destabilization. Following the natural progression of untreated atherosclerosis, we then describe the role of EVs in ischemic heart disease, where EVs interact locally in the heart and peripherally in the bone marrow and spleen to recruit and modulate mononuclear cells. Following the progression of untreated cardiac ischemia, we discuss heart failure, where EVs derived from inflammatory cells, cardiomyocytes, and fibroblasts shape cardiac remodeling. Finally, we briefly discuss the role of EVs in aortic aneurysmal and valvular pathology, where EVs are implicated in inflammatory processes that influence proteinase activity and calcification. The importance of EVs, however, extends beyond their role in inflammatory processes. Throughout the text, we refer the reader to other sources that discuss EV involvement in non-inflammatory mechanisms of cardiovascular disease. It is important to note that much of the EV data summarized in this review (and others) are derived from preclinical models, often from isolated in vitro assays and small animal models. Furthermore, much of the data were generated before the minimal information for studies of extracellular vesicles guidelines were established by the ISEV. Given various isolation methods (e.g., differential centrifugation, density gradient centrifugation, size exclusion chromatography, ultrafiltration, immunoprecipitation, immunocapture, precipitation, and others), variable yields and purities, lack of universally accepted EV-specific markers, and diverse bioactive cargo, further rigorous studies are needed to strengthen the translational potential of the findings presented in this review. Nevertheless, EVs are promising biomarkers for diagnostic and prognostic applications.

Finally, our discussion centered primarily on the diagnostic and prognostic value of EVs, with little mention of therapeutics. Understanding the mechanisms and contribution of EVs to cardiovascular pathology can in principle identify novel therapeutic targets. For instance, biogenesis and uptake of EVs is facilitated by diverse but targetable pathways (e.g., heparan sulfate proteoglycan pathway), which can be modulated for therapeutic purposes. Furthermore, EVs derived from various sources have emerged as promising cell-free therapeutic agents [102–104]. As reviewed previously [104], the potential advantages of EVs over cell therapy include EV stability, relative non-immunogenicity, and capacity for pre- and post-isolation modification. Therefore, EVs and their cargo will play an important role in the development of cell-free cardiovascular therapeutics.

Author Contributions: Conceptualization, original draft preparation, revisions, and funding, A.A. and T.P. All authors have read and agreed to the published version of the manuscript.

Funding: A.A. was supported by the National Institutes of Health (HL116273-07) and the Jack and Gitta Nagel and Family Endowed Cardiac Surgery Fellowship. T.P. was funded by the National Institutes of Health (K08HL141590).

Acknowledgments: Graphics created with BioRender.com.

Conflicts of Interest: The authors declare no conflict of interest.

References
1. WHO. Cardiovascular Diseases (CVDs) Fact Sheet. 2021. Available online: https://www.who.int/news-room/fact-sheets/detail/cardiovascular-diseases-(cvds) (accessed on 18 May 2022).
2. Lancellotti, P.; Marechal, P.; Donis, N.; Oury, C. Inflammation, cardiovascular disease, and cancer: A common link with far-reaching implications. Eur. Heart J. 2019, 40, 3910–3912. [CrossRef] [PubMed]
3. Ridker, P.M.; Cushman, M.; Stampfer, M.J.; Tracy, R.P.; Hennekens, C.H. Inflammation, aspirin, and the risk of cardiovascular disease in apparently healthy men. N. Engl. J. Med. 1997, 336, 973–979. [CrossRef] [PubMed]

4. Willerson, J.T.; Ridker, P.M. Inflammation as a cardiovascular risk factor. Circulation 2004, 109, II2–II10. [CrossRef] [PubMed]

5. Ridker, P.M.; Everett, B.M.; Thuren, T.; MacFadyen, J.G.; Chang, W.H.; Ballantyne, C.; Fonseca, F.; Nicolau, J.; Koenig, W.; Anker, S.D.; et al. Anti-Inflammatory Therapy with Canakinumab for Atherosclerotic Disease. N. Engl. J. Med. 2017, 377, 1119–1131. [CrossRef] [PubMed]

6. Everett, B.M.; Cornel, J.H.; Lainscak, M.; Anker, S.D.; Abbate, A.; Thuren, T.; Libby, P.; Glynn, R.J.; Ridker, P.M. Anti-Inflammatory Therapy With Canakinumab for the Prevention of Hospitalization for Heart Failure. Circulation 2019, 139, 1289–1299. [CrossRef]

7. Thery, C.; Witwer, K.W.; Aikawa, E.; Alcaraz, M.J.; Anderson, J.D.; Andriantsitohaina, R.; Antoniou, A.; Arab, T.; Archer, F.; Atkin-Smith, G.K.; et al. Minimal information for studies of extracellular vesicles 2018 (MISEV2018): A position statement of the International Society for Extracellular Vesicles and update of the MISEV2014 guidelines. J. Extracell. Vesicles 2018, 7, 1535750. [CrossRef]

8. Denzer, K.; van Eijik, M.; Kleijmeer, M.J.; Jakobson, E.; de Groot, C.; Geuze, H.J. Follicular dendritic cells carry MHC class II-expressing microvesicles at their surface. Trends Cell Biol. 2015, 25, 364–372. [CrossRef]

9. Cocucci, E.; Meldolesi, J. Ectosomes and exosomes: Shedding the confusion between extracellular vesicles. Trends Cell Biol. 2014, 14, 195–208. [CrossRef] [PubMed]

10. Jansen, F.; Nickenig, G.; Werner, N. Extracellular Vesicles in Cardiovascular Disease: Potential Applications in Diagnosis, Prognosis, and Epidemiology. Circ. Res. 2017, 120, 1649–1657. [CrossRef] [PubMed]

11. Voukalis, C.; Shantsila, E.; Lip, G.Y.H. Microparticles and cardiovascular diseases. Ann. Med. 2019, 51, 193–223. [CrossRef] [PubMed]

12. Libby, P.; Buring, J.E.; Badimon, L.; Hansson, G.K.; Deanfield, J.; Bittencourt, M.S.; Tokgozoglu, L.; Lewis, E.F. Atherosclerosis. Nat. Rev. Dis. Primers 2019, 5, 56. [CrossRef]

13. Konkoth, A.; Saraswat, R.; Dubrou, C.; Sabatier, F.; Leroyer, A.S.; Lacroix, R.; Duchez, A.C.; Dignat-George, F. Multifaceted role of neutrophil microvesicles in the pathogenesis of atherosclerosis. Arterioscler. Thromb. Vasc. Biol. 2020, 40, 3097–3106. [CrossRef] [PubMed]

14. Rautou, P.E.; Leroyer, A.S.; Ramkhelawon, B.; Devue, C.; Duflaut, D.; Vion, A.C.; Nalboga, G.; Castier, Y.; Lesche, G.; Lehoux, S.; et al. Microparticles from human atherosclerotic plaques promote endothelial ICAM-1-dependent monocyte adhesion and transendothelial migration. Circ. Res. 2011, 108, 335–343. [CrossRef]

15. Legein, B.; Temmerman, L.; Biessen, E.A.; Lutgens, E. Inflammation and immune system interactions in atherosclerosis. Cell Mol. Life Sci. 2013, 70, 3847–3869. [CrossRef]

16. Legein, B.; Temmerman, L.; Biessen, E.A.; Lutgens, E. Inflammation and immune system interactions in atherosclerosis. Cell Mol. Life Sci. 2013, 70, 3847–3869. [CrossRef]

17. Gomez, I.; Ward, B.; Souilhol, C.; Recarti, C.; Ariaans, M.; Johnston, J.; Burnett, A.; Mahmoud, M.; Luong, L.A.; West, L.; et al. Neutrophil microvesicles drive atherosclerosis by delivering miR-155 to atheroprone endothelium. Nat. Commun. 2020, 11, 214. [CrossRef] [PubMed]

18. Libby, P.; Buring, J.E.; Badimon, L.; Hansson, G.K.; Deanfield, J.; Bittencourt, M.S.; Tokgozoglu, L.; Lewis, E.F. Atherosclerosis. Nat. Rev. Dis. Primers 2019, 5, 56. [CrossRef]

19. Kauravi, S.J.; Harrison, P.; Rainger, G.E.; Nash, G.B. Ability of Platelet-Derived Extracellular Vesicles to Promote Neutrophil-Endothelial Cell Interactions. Inflammation 2019, 42, 290–305. [CrossRef] [PubMed]

20. Wadey, R.M.; Connolly, K.D.; Mathew, D.; Walters, G.; Rees, D.A.; James, P.E. Inflammatory adipocyte-derived extracellular vesicles promote leukocyte attachment to vascular endothelial cells. Atherosclerosis 2019, 283, 19–27. [CrossRef]
28. Huber, J.; Vales, A.; Mitulovic, G.; Blumer, M.; Schmid, R.; Witztum, J.L.; Binder, B.R.; Leitinger, N. Oxidized membrane vesicles and blebs from apoptotic cells contain biologically active oxidized phospholipids that induce monocyte-endothelial interactions. *Arterioscler. Thromb. Vasc. Biol.* 2002, 22, 101–107. [CrossRef]

29. Yu, X.H.; Fu, Y.C.; Zhang, D.W.; Yin, K.; Tang, C.K. Foam cells in atherosclerosis. *Clin. Chim. Acta* 2013, 424, 245–252. [CrossRef]

30. Feng, C.; Chen, Q.; Fan, M.; Guo, J.; Liu, Y.; Ji, T.; Zhu, J.; Zhao, X. Platelet-derived microparticles promote phagocytosis of oxidized low-density lipoprotein by macrophages, potentially enhancing foam cell formation. *Ann. Transl. Med.* 2019, 7, 477. [CrossRef]

31. Xie, Z.; Wang, X.; Liu, X.; Du, H.; Sun, C.; Shao, X.; Tian, J.; Gu, X.; Wang, H.; Tian, J.; et al. Adipose-Derived Exosomes Exert Proatherogenic Effects by Regulating Macrophage Cell Fusion and Polarization. *J. Am. Heart Assoc.* 2018, 7, e007442. [CrossRef]

32. Huang, C.; Han, J.; Wu, Y.; Li, S.; Wang, Q.; Lin, W.; Zhu, J. Exosomal MALAT1 derived from oxidized low-density lipoprotein-treated endothelial cells promotes M2 macrophage polarization. *Mol. Med. Rep.* 2018, 18, 509–515. [CrossRef] [PubMed]

33. He, S.; Wu, C.; Xiao, J.; Li, D.; Sun, Z.; Li, M. Endothelial extracellular vesicles modulate the macrophage phenotype: Potential implications in atherosclerosis. *Scand. J. Immunol.* 2018, 87, e12648. [CrossRef] [PubMed]

34. Gerdts, P.L.S.M.; Esko, J.D. The heparan sulfate proteoglycan stress on hyperlipidemia and atherosclerosis. *Matrix Biol.* 2018, 71–72, 262–282. [CrossRef] [PubMed]

35. Tsiantoulas, D.; Eslami, M.; Obermayer, G.; Clement, M.; Smeets, D.; Mayer, F.J.; Kiss, M.G.; Enders, L.; Weißler, J.; Göderle, L. APRIL limits atherosclerosis by binding to heparan sulfate proteoglycans. *Nature* 2021, 597, 92–96. [CrossRef]

36. Zhou, Y.; Xie, Q.; Pan, S.; Wu, J.; Wang, X.; Cao, Z.; Wang, M.; Zha, L.; Zhou, M.; Li, Q.; et al. Small extracellular vesicles containing LDLRQ722* protein reconstructed the lipid metabolism via heparan sulfate proteoglycans and clathrin-mediated endocytosis. *Clin. Transl. Med.* 2022, 12, e773. [CrossRef]

37. Wight, T.N. A Role for proteoglycans in vascular disease. *Matrix Biol.* 2018, 71–72, 396–420. [CrossRef]

38. Rogers, M.A.; Aikawa, E. Cardiovascular calcification: Artificial intelligence and big data accelerate mechanistic discovery. *Nat. Rev. Cardiol.* 2019, 16, 261–274. [CrossRef]

39. New, S.E.; Goettsch, C.; Aikawa, M.; Marchini, J.F.; Shibasaki, M.; Yabusaki, K.; Libby, P.; Shanahan, C.M.; Croce, K.; Aikawa, E. Macrophage-derived matrix vesicles: An alternative novel mechanism for microcalcification in atherosclerotic plaques. *Circ. Res.* 2013, 113, 72–77. [CrossRef] [PubMed]

40. Kawakami, R.; Katsuki, S.; Travers, R.; Romero, D.C.; Becker-Greene, D.; Passos, L.S.A.; Higashi, H.; Blaser, M.C.; Sukhova, G.K.; Buttigieg, J.; et al. s100A9-RAGE Axis Accelerates Formation of Macrophage-Mediated Extracellular Vesicle Microcalcification in Diabetes Mellitus. *Arterioscler. Thromb. Vasc. Biol.* 2020, 40, 1838–1853. [CrossRef]

41. Roth, G.A.; Mensah, G.A.; Johnson, C.O.; Addolorato, G.; Ammirati, E.; Baddour, L.M.; Barengo, N.C.; Beaton, A.Z.; Benjamin, E.J.; Benziger, C.P.; et al. Global Burden of Cardiovascular Diseases and Risk Factors, 1990–2019: Update From the GBD 2019 Study. *J. Am. Coll. Cardiol.* 2020, 76, 2982–3021. [CrossRef]

42. Frangogiannis, N.G.; Smith, C.W.; Entman, M.L. The inflammatory response in myocardial infarction. *Cardiovasc. Res.* 2002, 53, 31–47. [CrossRef]

43. Nahrendorf, M.; Pittet, M.J.; Swirski, F.K. Monocytes: Protagonists of infarct inflammation and repair after myocardial infarction. *Circulation* 2012, 121, 2437–2445. [CrossRef] [PubMed]

44. Loyer, X.; Zlatanova, I.; Devue, C.; Yin, M.; Howangyin, K.Y.; Zlatanov, I.; Kheloufi, M.; Vilar, J.; Zannis, K.; et al. Like Factor 2-overexpressing endothelial cells attenuate myocardial ischemia-reperfusion injury by preventing Ly6C(high) monocyte recruitment. *Theranostics* 2020, 10, 11562–11579. [CrossRef] [PubMed]

45. Almeida Paiva, R.; Martins-Maques, T.; Jesus, K.; Ribeiro-Rodrigues, T.; Zuzarte, M.; Silva, A.; Reis, L.; da Silva, M.; Pereira, P.; Góes, M.; et al. Ischaemia alters the effects of cardiomyocyte-derived extracellular vesicles on macrophage activation. *J. Cell Mol. Cardiol.* 2023, 123, 1137–1151. [CrossRef]

46. Cheng, M.; Yang, J.; Zhao, X.; Zhang, J.; Zeng, Q.; Yu, Y.; Yang, L.; Wu, B.; Yi, G.; Mao, X.; et al. Circulating myocardial microRNAs from infarcted hearts are carried in exosomes and mobilise bone marrow progenitor cells. *Nat. Commun.* 2019, 10, 959. [CrossRef] [PubMed]

47. Akbar, N.; Digby, J.E.; Cahill, T.J.; Tavare, A.N.; Corbin, A.L.; Saluja, S.; Dawkins, S.; Edgar, L.; Rawlings, N.; Ziberna, K.; et al. Endothelium-derived extracellular vesicles promote splenic monocyte mobilization in myocardial infarction. *JCI Insight* 2017, 2, e93344. [CrossRef]

48. Qiao, S.; Zhang, W.; Yin, Y.; Wei, Z.; Chen, F.; Zhao, J.; Sun, X.; Mu, D.; Xie, J.; Xu, B. Extracellular vesicles derived from Kruppel-Like Factor 2-overexpressing endothelial cells attenuate myocardial ischemia-reperfusion injury by preventing Ly6C(high) monocyte recruitment. *Theranostics* 2020, 10, 11562–11579. [CrossRef] [PubMed]

49. Camblin, L.; de Couto, G.; Ibrahim, A.; Echavez, A.K.; Valle, J.; Liu, W.; Kreke, M.; Smith, R.R.; Marban, L.; Marban, E. Y RNA fragment in extracellular vesicles confers cardioprotection via modulation of IL-10 expression and secretion. *EMBO Mol. Med.* 2017, 9, 337–352. [CrossRef]

50. de Couto, G.; Gallet, R.; Cambier, L.; Jaghatspayan, E.; Makkar, N.; Dawkins, J.F.; Berman, B.P.; Marban, E. Exosomal MicroRNA Transfer Into Macrophages Mediates Cellular Postconditioning. *Circulation* 2017, 136, 200–214. [CrossRef]

51. Akherov, A.; Marban, E. COVID-19 and the Heart. *Circ. Res.* 2020, 126, 1443–1455. [CrossRef]
52. Wang, Y.; Li, C.; Zhao, R.; Qiu, Z.; Shen, C.; Wang, Z.; Liu, W.; Zhang, W.; Ge, J.; Shi, B. CircUbe3a from M2 macrophage-derived small extracellular vesicles mediates myocardial fibrosis after acute myocardial infarction. *Theranostics* 2021, 11, 6315–6333. [CrossRef] [PubMed]

53. Wang, C.; Zhang, C.; Liu, L.; Xi, A.; Chen, B.; Li, Y.; Du, J. Macrophage-Derived mirc-155-Containing Exosomes Suppress Fibroblast Proliferation and Promote Fibroblast Inflammation during Cardiac Injury. *Mol. Ther.* 2017, 25, 192–204. [CrossRef] [PubMed]

54. Liu, S.; Chen, J.; Shi, J.; Zhou, W.; Wang, L.; Fang, W.; Zhong, Y.; Chen, X.; Chen, Y.; Sabri, A.; et al. M1-like macrophage-derived exosomes suppress angiogenesis and exacerbate cardiac dysfunction in a myocardial infarction microenvironment. *Basic Res. Cardiol.* 2020, 115, 22. [CrossRef]

55. Liu, H.; Gao, W.; Yuan, J.; Wu, C.; Yao, K.; Zhang, L.; Ma, L.; Zhu, J.; Zou, Y.; Ge, J. Exosomes derived from dendritic cells improve cardiac function via activation of CD4(+) T lymphocytes after myocardial infarction. *J. Mol. Cell Cardiol.* 2016, 91, 123–133. [CrossRef]

56. Wang, Y.; Dembowsky, K.; Chevalier, E.; Stuve, P.; Korf-Klingebiel, M.; Lochner, M.; Napp, L.C.; Frank, H.; Brinkmann, E.; Kanwischer, A.; et al. C-C Motif Chemokine Receptor 4 Blockade Promotes Tissue Repair After Myocardial Infarction by Enhancing Regulatory T Cell Mobilization and Immune-Regulatory Function. *Circulation* 2019, 139, 1798–1812. [CrossRef]

57. Feng, G.; Baiqat, G.; Ma, P.; Koenig, A.; Bredemeyer, A.; Lokshina, I.; Lai, L.; Forster, I.; Leuschner, F.; Kreisel, D.; et al. CCL17 Aggravates Myocardial Injury by Suppressing Recruitment of Regulatory T Cells. *Circulation* 2022, 145, 765–782. [CrossRef]

58. Xia, N.; Lu, Y.; Gu, M.; Li, N.; Liu, M.; Jiao, J.; Zhang, Z.; Li; J.; Li, D.; Tang, T.; et al. A Unique Population of Regulatory T Cells in Heart Potentiates Cardiac Protection From Myocardial Infarction. *Circulation* 2020, 142, 1956–1973. [CrossRef]

59. Akhmerov, A.; Rogers, R.; de Couto, G.; Valle, J.; Li, L.; Ibrahim, A.; Sanchez, L.; Zhang, R.; Lin, Y.N.; Liu, W.; et al. Regulatory T cell activation, proliferation, and reprogramming induced by extracellular vesicles. *J. Heart Lung Transpl.* 2021, 40, 1387–1395. [CrossRef]

60. Bozkurt, B.; Coats, A.J.S.; Tsutsui, H.; Abdelhamid, C.M.; Adamopoulos, S.; Albert, N.; Anker, S.D.; Atherton, J.; Bohm, M.; Butler, J.; et al. Universal definition and classification of heart failure: A report of the Heart Failure Society of America, Heart Failure Association of the European Society of Cardiology, Japanese Heart Failure Society and Writing Committee of the Universal Definition of Heart Failure: Endorsed by the Canadian Heart Failure Society, Heart Failure Association of India, Cardiac Society of Australia and New Zealand, and Chinese Heart Failure Association. *Eur. J. Heart Fail.* 2021, 23, 352–380.

61. GBD 2017 Disease and Injury Incidence and Prevalence Collaborators. Global, regional, and national incidence, prevalence, and years lived with disability for 354 diseases and injuries for 195 countries and territories, 1990–2017: A systematic analysis for the Global Burden of Disease Study 2017. *Lancet* 2018, 392, 1789–1858. [CrossRef]

62. Savarese, G.; Becher, P.M.; Lund, L.H.; Seferovic, P.; Rosano, G.M.C.; Coats, A. Global burden of heart failure: A comprehensive and updated review of epidemiology. *Cardiovasc. Res.* 2022, 1–16. [CrossRef] [PubMed]

63. Tsao, C.W.; Aday, A.W.; Almarzooq, Z.I.; Alonso, A.; Beaton, A.Z.; Bittencourt, M.S.; Boehme, A.K.; Buxton, A.E.; Carson, A.P.; Commodore-Mensah, Y.; et al. Heart Disease and Stroke Statistics-2022 Update: A Report From the American Heart Association. *Circulation* 2022, 145, e153–e639. [CrossRef] [PubMed]

64. Owari, T.E.; Hodge, D.O.; Herges, R.M.; Jacobsen, S.J.; Roger, V.L.; Redfield, M.M. Trends in prevalence and outcome of heart failure with preserved ejection fraction. *N. Engl. J. Med.* 2006, 355, 251–259. [CrossRef] [PubMed]

65. Akhmerov, A.; Ramzy, D. Mechanical Circulatory Support in Right Ventricular Failure. *Interv. Cardiol. Clin.* 2021, 10, 185–194. [CrossRef]

66. Matsumoto, S.; Sakata, Y.; Suna, S.; Nakatani, D.; Usami, M.; Hara, M.; Kitamura, T.; Hamasaki, T.; Nanto, S.; Kawahara, Y.; et al. Circulating p53-responsive microRNAs are predictive indicators of heart failure after acute myocardial infarction. *Circ. Res.* 2013, 113, 322–326. [CrossRef]

67. Nie, X.; Fan, J.; Li, H.; Yin, Z.; Zhao, Y.; Dai, B.; Dong, N.; Chen, C.; Wang, D.W. miR-217 Promotes Cardiac Hypertrophy and Dysfunction by Targeting PTEN. *Mol. Ther. Nucleic Acids* 2018, 12, 254–266. [CrossRef]

68. Goren, Y.; Kushnir, M.; Zafirri, B.; Tabak, S.; Lewis, B.S.; Amir, O. Serum levels of microRNAs in patients with heart failure. *Eur. J. Heart Fail.* 2012, 14, 147–154. [CrossRef]

69. Qiao, L.; Hu, S.; Liu, S.; Zhang, H.; Ma, H.; Huang, K.; Li, Z.; Su, T.; Vandergriff, A.; Tang, J.; et al. microRNA-21-5p dysregulation in exosomes derived from heart failure patients impairs regenerative potential. *J. Clin. Investig.* 2019, 129, 2237–2250. [CrossRef]

70. Biemmi, V.; Milano, G.; Ciullo, A.; Cervio, E.; Burrello, J.; Dei Cas, M.; Paroni, R.; Tallone, T.; Mocetti, T.; Pedrazzini, G.; et al. Inflammatory extracellular vesicles prompt heart dysfunction via TLR4-dependent NF-kappaB activation. *Theranostics* 2020, 10, 2773–2790. [CrossRef]

71. Yu, H.; Qin, L.; Peng, Y.; Bai, W.; Wang, Z. Exosomes Derived From Hypertrophic Cardiomyocytes Induce Inflammation in Macrophages via mirc-155 Mediated MAPK Pathway. *Front. Immunol.* 2020, 11, 606045. [CrossRef]

72. Lindner, D.; Zietsch, C.; Tank, J.; Sossalla, S.; Fluschnik, N.; Hinrichs, S.; Maier, L.; Poller, W.; Blankenberg, S.; Schultheiss, H.P.; et al. Cardiac fibroblasts support cardiac inflammation in heart failure. *Basic Res. Cardiol.* 2014, 109, 428. [CrossRef]

73. Tian, C.; Gao, L.; Zimmerman, M.C.; Zucker, I.H. Myocardial infarction-induced microRNA-enriched exosomes contribute to cardiac Nrf2 dysregulation in chronic heart failure. *Am. J. Physiol. Heart Circ. Physiol.* 2018, 314, H928–H939. [CrossRef] [PubMed]

74. Bang, C.; Batkai, S.; Dangwal, S.; Gupta, S.K.; Foinquinos, A.; Holzmann, A.; Just, A.; Remke, J.; Zimmer, K.; Zeug, A.; et al. Cardiac fibroblast-derived microRNA passenger strand-enriched exosomes mediate cardiomyocyte hypertrophy. *J. Clin. Investig.* 2014, 124, 2136–2146. [CrossRef]
75. Tian, C.; Hu, G.; Gao, L.; Hackfort, B.T.; Zucker, I.H. Extracellular vesicular MicroRNA-27a* contributes to cardiac hypertrophy in chronic heart failure. *J. Mol. Cell Cardiol.* 2020, 143, 120–131. [CrossRef] [PubMed]

76. Han, C.; Yang, J.; Sun, J.; Qin, G. Extracellular vesicles in cardiovascular disease: Biological functions and therapeutic implications. *Pharmacol. Ther.* 2022, 233, 108025. [CrossRef] [PubMed]

77. Ailawadi, S.; Wang, X.; Gu, H.; Fan, G.C. Pathologic function and therapeutic potential of exosomes in cardiovascular disease. *Biochim. Biophys. Acta* 2015, 1852, 1–11. [CrossRef] [PubMed]

78. Kenneweg, F.; Bang, C.; Xiao, K.; Boulanger, C.M.; Loyer, X.; Mazlan, S.; Schroen, B.; Hermans-Beijnsberger, S.; Foinquinos, A.; Hirt, M.N.; et al. Long Noncoding RNA-Enriched Vesicles Secreted by Hypoxic Cardiomyocytes Drive Cardiac Fibrosis. *Mol. Ther. Nucleic Acids* 2019, 18, 363–374. [CrossRef]

79. Ashton, H.A.; Buxton, M.J.; Day, N.E.; Kim, L.G.; Marteau, T.M.; Scott, R.A.; Thompson, S.G.; Walker, N.M.; Multicentre Aneurysm Screening Study Group. The Multicentre Aneurysm Screening Study (MASS) into the effect of abdominal aortic aneurysm screening on mortality in men: A randomised controlled trial. *Lancet* 2002, 360, 1531–1539.

80. Norman, P.E.; Jamrozik, K.; Lawrence-Brown, M.M.; Le, M.T.; Spencer, C.A.; Tuohy, R.J.; Parsons, R.W.; Dickinson, J.A. Population based randomised controlled trial on impact of screening on mortality from abdominal aortic aneurysm. *BMJ* 2004, 329, 1259. [CrossRef]

81. Lindholt, J.S.; Juul, S.; Fasting, H.; Henneberg, E.W. Screening for abdominal aortic aneurysms: Single centre randomised controlled trial. *BMJ* 2005, 330, 750. [CrossRef]

82. Ashton, H.A.; Gao, L.; Kim, L.G.; Druce, P.S.; Thompson, S.G.; Scott, R.A. Fifteen-year follow-up of a randomized clinical trial of ultrasonographic screening for abdominal aortic aneurysms. *Br. J. Surg.* 2007, 94, 696–701. [CrossRef] [PubMed]

83. Von Allmen, R.S.; Powell, J.T. The management of ruptured abdominal aortic aneurysms: Screening for abdominal aortic aneurysm and incidence of rupture. *J. Cardiovasc. Surg.* 2012, 53, 69–76.

84. Martin-Ventura, J.L.; Roncal, C.; Orbe, J.; Blanco-Colio, L.M. Role of Extracellular Vesicles as Potential Diagnostic and/or Therapeutic Biomarkers in Chronic Cardiovascular Diseases. *Front. Cell Dev. Biol.* 2022, 10, 813885. [CrossRef]

85. Martinez-Pinna, R.; Gonzalez de Peredo, A.; Monsarrat, B.; Burlet-Schiltz, O.; Martin-Ventura, J.L. Label-free quantitative proteomic analysis of human plasma-derived microvesicles to find protein signatures of abdominal aortic aneurysms. *Proteom. Clin. Appl.* 2014, 8, 620–625. [CrossRef]

86. Touat, Z.; Lepage, L.; Ollivier, V.; Nataf, P.; Hvass, U.; Labreuche, J.; Jandrot-Perrus, M.; Michel, J.B.; Jondeau, G. Dilation-valve calcification inversely correlates with osteoporotic bone remodelling: A role for inflammation. *Br. J. Surg.* 2012, 99, 120–131. [CrossRef] [PubMed]

87. Touat, Z.; Ollivier, V.; Dai, J.; Huisse, M.G.; Bezeaud, A.; Sebbag, U.; Palombi, T.; Rossignol, P.; Meilhac, O.; Guillin, M.C.; et al. Inflammation is associated with the remodeling of calcific aortic valve disease. *J. Mol. Cell Cardiol.* 2016, 94, 575–585. [CrossRef] [PubMed]

88. Fernandez-Garcia, C.E.; Burillo, E.; Lindholt, J.S.; Martinez-Lopez, D.; Pilely, K.; Mazzeo, C.; Michel, J.B.; Egido, J.; Garred, P.; Blanco-Colio, L.M.; et al. Association of ficolin-3 with abdominal aortic aneurysm presence and progression. *J. Thromb. Haemost.* 2017, 15, 575–585. [CrossRef]

89. Munthe-Fog, L.; Hummelshoj, T.; Honore, C.; Madsen, H.O.; Permin, H.; Garred, P. Immunodeficiency associated with FCN3 mutation and ficolin-3 deficiency. *N. Engl. J. Med.* 2009, 360, 2637–2644. [CrossRef]

90. Wang, Y.; Jia, L.; Xie, Y.; Cai, Z.; Liu, Z.; Shen, J.; Lu, Y.; Wang, Y.; Su, S.; Ma, Y.; et al. Involvement of macrophage-derived exosomes in abdominal aortic aneurysm development. *Atherosclerosis* 2019, 289, 64–72. [CrossRef] [PubMed]

91. Folkesson, M.; Li, C.; Frebelius, S.; Swedenborg, J.; Wagstater, D.; Williams, K.J.; Eriksson, P.; Roy, J.; Liu, M.L. Proteolytically active ADAM10 and ADAM17 carried on membrane microvesicles in human abdominal aortic aneurysms. *Thromb. Haemost.* 2015, 114, 1165–1174.

92. Dang, G.; Li, T.; Yang, D.; Yang, G.; Du, X.; Yang, J.; Miao, Y.; Han, L.; Ma, X.; Song, Y.; et al. T lymphocyte-derived extracellular vesicles aggravate abdominal aortic aneurysm by promoting macrophage lipid peroxidation and migration via pyruvate kinase muscle isoforme 2. *Redox Biol.* 2022, 50, 102257. [CrossRef] [PubMed]

93. Potsophsakul, P.; Furmanik, M.; Forsythe, R.; Dweck, M.; Schurink, G.W.; Natour, E.; Reutelingsperger, C.; Jacobs, M.; Mees, B.; Schurgers, L. Role of Vascular Smooth Muscle Cell Phenotypic Switching and Calcification in Aortic Aneurysm Formation. *Arterioscler. Thromb. Vasc. Biol.* 2019, 39, 1363–1368. [CrossRef] [PubMed]

94. Hjortnaes, J.; Butler, J.; Figueiredo, J.L.; Riccio, M.; Kohler, R.H.; Kozloff, K.M.; Weissleder, R.; Aikawa, E. Arterial and aortic valve calcification inversely correlates with osteoporotic bone remodelling: A role for inflammation. *Eur. Heart J.* 2010, 31, 1975–1984. [CrossRef] [PubMed]

95. Cui, L.; Rashdan, N.A.; Zhu, D.; Milne, E.M.; Ajuh, P.; Milne, G.; Helfrich, M.H.; Lim, K.; Prasad, S.; Lerman, D.A.; et al. End stage renal disease-induced hypercalcemia may promote aortic valve calcification via Annexin VI enrichment of valve interstitial muscle isozyme 2. *Thromb. Haemost.* 2020, 120, 2985–2995. [CrossRef] [PubMed]

96. Cote, N.; Mahmut, A.; Bosse, Y.; Couture, C.; Page, S.; Trahan, S.; Boulanger, M.C.; Fournier, D.; Fibaro, P.; Mathieu, P. Inflammation is associated with the remodeling of calcic aortic valve disease. *Inflammation* 2013, 36, 573–581. [CrossRef] [PubMed]

97. Diehl, P.; Nagy, F.; Sossong, V.; Helbing, T.; Beyersdorf, F.; Olschewski, M.; Bode, C.; Moser, M. Increased levels of circulating microparticles in patients with severe aortic valve stenosis. *Thromb. Haemost.* 2008, 99, 711–719.
98. Kaden, J.J.; Dempfle, C.E.; Grobholz, R.; Fischer, C.S.; Vocke, D.C.; Kilic, R.; Sarikoc, A.; Pinol, R.; Hagl, S.; Lang, S.; et al. Inflammatory regulation of extracellular matrix remodeling in calcific aortic valve stenosis. *Cardiovasc. Pathol.* 2005, 14, 80–87. [CrossRef]

99. Blaser, M.C.; Aikawa, E. Roles and Regulation of Extracellular Vesicles in Cardiovascular Mineral Metabolism. *Front. Cardiovasc. Med.* 2018, 5, 187. [CrossRef]

100. Jansen, F.; Xiang, X.; Werner, N. Role and function of extracellular vesicles in calcific aortic valve disease. *Eur. Heart J.* 2017, 38, 2714–2716. [CrossRef]

101. Di Vito, A.; Donato, A.; Presta, I.; Mancuso, T.; Brunetti, F.S.; Mastroroberto, P.; Amorosi, A.; Malara, N.; Donato, G. Extracellular Matrix in Calcific Aortic Valve Disease: Architecture, Dynamic and Perspectives. *Int. J. Mol. Sci.* 2021, 22, 913. [CrossRef] [PubMed]

102. Barile, L.; Moccetti, T.; Marban, E.; Vassalli, G. Roles of exosomes in cardioprotection. *Eur. Heart J.* 2017, 38, 1372–1379. [CrossRef] [PubMed]

103. Marban, E. The Secret Life of Exosomes: What Bees Can Teach Us About Next-Generation Therapeutics. *J. Am. Coll. Cardiol.* 2018, 71, 193–200. [CrossRef] [PubMed]

104. Marban, E. A mechanistic roadmap for the clinical application of cardiac cell therapies. *Nat. Biomed. Eng.* 2018, 2, 353–361. [CrossRef] [PubMed]