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The Role of Adenosine Signaling in Headache: A Review

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Abstract: Migraine is the third most prevalent disease on the planet, yet our understanding of its mechanisms and pathophysiology is surprisingly incomplete. Recent studies have built upon decades of evidence that adenosine, a purine nucleoside that can act as a neuromodulator, is involved in pain transmission and sensitization. Clinical evidence and rodent studies have suggested that adenosine signaling also plays a critical role in migraine headache. This is further supported by the widespread use of caffeine, an adenosine receptor antagonist, in several headache treatments. In this review, we highlight evidence that supports the involvement of adenosine signaling in different forms of headache, headache triggers, and basic headache physiology. This evidence supports adenosine A2A receptors as a critical adenosine receptor subtype involved in headache pain. Adenosine A2A receptor signaling may contribute to headache via the modulation of intracellular Cyclic adenosine monophosphate (cAMP) production or 5′ AMP-activated protein kinase (AMPK) activity in neurons and glia to affect glutamatergic synaptic transmission within the brainstem. This evidence supports the further study of adenosine signaling in headache and potentially illuminates it as a novel therapeutic target for migraine.

Keywords: migraine; headache; adenosine; adenosine receptors; mitochondria; astrocytes; trigeminal

1. Introduction

Adenosine is a purine nucleoside that plays a critical role in numerous cellular and molecular functions throughout the brain such as metabolism, cell signaling, purinergic neuronal signaling, and inflammation [1]. Not surprisingly, adenosine is appreciated for its complex role in many nervous system disorders such as alcoholism, epilepsy, traumatic brain injury, ischemia, anxiety, Alzheimer’s disease, Parkinson’s, and brain cancer [2–8]. Although progress has been made in understanding adenosine’s role in certain pain states, less is known about how it is involved in migraine [9].

Current treatment options for headache and migraine include analgesics, triptans, ergots, behavioral intervention, and nerve stimulation, while new treatments, such as monoclonal antibodies against Calcitonin gene-related peptide (CGRP) or the CGRP receptor and CGRP inhibitors, are under development. Some patients who are non-responders to these treatments choose to undergo surgical implantation of invasive nerve stimulators to relieve pain [10]. A component of some of these treatment options is caffeine, a non-specific adenosine receptor antagonist [11]. Combining caffeine with other treatments can improve pain relief by as much as 40% in patients [12–16]. Caffeine was
originally included in many migraine treatments due to the previous assumption that migraine was a vascular disorder. Now that migraine is thought to be a neurological and not a vascular disorder, caffeine’s common inclusion in migraine treatments suggests an alternative, non-vascular mechanism of action behind its efficacy in migraine patients. In this review, we present mounting evidence that demonstrates adenosine’s role in migraine and headache, revealing it as a potential therapeutic target for migraine and headache treatment.

2. Modeling Headache and Migraine

Headache is described as pain located above the orbitomeatal line that transects the head, approximately from the top of the eyes to the back of the skull. Migraine is a disabling neurological disorder most often characterized by the accompanying headache, but it is much more than just an episodic headache as it includes many other symptoms such as phonophobia, photophobia, nausea, and visual auras during the prodromal phase in a subset of patients. Thirty-six million individuals in the United States suffer from migraine, with a cost-estimate of $50 billion each year [17]. The World Health Organization has classified it as the third most common disease worldwide [18]. Fourteen-million migraine patients are diagnosed as chronic migraineurs, described as an individual who experiences a migraine at least fifteen days out of every month for at least three months [17].

The headache phase of migraine is thought to be caused by activation of dural C and Aδ nociceptors of the trigeminal ganglion [19]. However, no direct evidence for this exists due to the difficulty of studying the contribution of meningeal afferents to headache in humans. However, functional magnetic resonance imaging data demonstrating increased activity in the trigeminal ganglion during migraine as well as other human studies suggest that headache pain is transmitted via these nociceptors [20,21]. Trigeminal afferent fibers project centrally to second order neurons of the trigeminal nucleus caudalis (TNC) in the brainstem. It is thought that peripheral and central changes occur after repeated episodic nociceptor activation that sensitizes this trigeminal system, exacerbating the perception of migraine pain in secondary or referred areas [22]. Chronic migraine most likely involves the development of chronic peripheral and central sensitization from repeated attacks [23,24].

We have previously developed two behavioral animal models of chronic trigeminal allodynia that feature numerous migraine-like characteristics to allow for the investigation of migraine pathophysiology [25–31]. The first model, designated as the inflammatory stimulation model (IS rats), was developed by simulating the putative dural nociceptor activation thought to occur during a headache attack by infusing an inflammatory soup (1-mM histamine, serotonin, bradykinin, and 0.1-mM prostaglandin E2 in phosphate-buffered saline (PBS), pH 7.4) onto the dura through an affixed cannula 3 ×/ week for a total of 12 infusions [26]. Each infusion causes the development of trigeminal sensitivity in the periorbital region, as measured with von Frey hairs, which recovers within 2–3 h. Following approximately the fifth infusion, the animals’ baseline thresholds begin to also decrease (i.e., the animal is still experiencing trigeminal sensitivity before the following infusion is performed). This repeated inflammatory dural stimulation then induces a steady-state of chronic trigeminal sensitivity that outlasts the final infusion for months.

The second model, called the spontaneous trigeminal allodynia (STA) in rats, is currently the only model of primary trigeminal allodynia in animals [32]. Naturally born with this trait, no surgical procedure or pharmacological treatment is needed to produce an allodynic state that is confined to the trigeminal system. These animals are not genetically modified, but were instead discovered and bred to create a colony of STA rats. The gene or genes responsible for this trait are currently unknown. Similar to human migraineurs who experience a combination of headache-free and headache days with fluctuating intensity, these animals feature episodically fluctuating trigeminal sensitivity. Both models of headache share symptomology similar to migraine in that they experience phonophobia, sensitivity to migraine triggers, similar efficacious responses to migraine treatments, and trigeminal sensitivity (although expressed differently—IS rats with stable chronic trigeminal sensitivity and STA rats with episodically fluctuating chronic trigeminal sensitivity) [25,27–29]. These animal models provide an
experimental means to determine elements that are key to migraine pathophysiology and to screen potential therapeutic treatments.

3. Adenosine Receptor Signaling in the Nervous System

The physiological role of adenosine was first appreciated in 1929 when it was described for its ability to modulate the beat and conductance of cardiac muscle fibers [33]. Adenosine is now well-accepted as a neuromodulator with extracellular levels of it derived from several cellular sources within the central nervous system, including multiple glia and neuronal cell types [34–37]. Some evidence even suggests it may be directly packaged and released as a neuro/glio-transmitter in a Soluble NSF Attachment Protein Receptor (SNARE)- and stimulus-dependent manner [38–40]. Similar to a neuro/glio-transmitter, there are distinct mechanisms for the removal of adenosine from the extracellular space to deactivate its effects, such as reuptake via equilibrative nucleoside transporters (ENTs) or metabolism to inosine by adenosine deaminase (ADO) [41,42]. Three primary mechanisms are responsible for the production/release of extracellular adenosine: equilibrative transport of adenosine out of the cell through equilibrative nucleoside transporters (ENTs) when intracellular levels are high [43], metabolism of released adenosine triphosphate (ATP) to adenosine by subsequent hydrolysis steps with extracellular \( 5' \) ectonucleotidases (i.e., ATP to adenosine diphosphate (ADP) to adenosine monophosphate (AMP) to adenosine) [44], or activity-dependent mechanisms during gliotransmission [38–40].

There are four known adenosine receptor subtypes. These purinergic G protein-coupled receptors each modulate cyclase activity in alternate ways [1]. Adenosine \( A_{2A} \) and \( A_{2B} \) receptors are coupled to the \( G_s \) alpha subunit, stimulating adenylate cyclase activity, while adenosine \( A_1 \) and \( A_3 \) receptors are coupled to the \( G_i \) alpha subunit, inhibiting adenylate cyclase activity [45]. Control of adenylate cyclase activity with these receptors allows adenosine to fine-tune intracellular cAMP levels to control a large host of cellular mechanisms including resting potentials, metabolism, calcium channel phosphorylation, and gene expression [46]. Each adenosine receptor subtype has been reported to be expressed throughout the central and peripheral nervous system. Adenosine \( A_1 \) receptors are the most abundant adenosine receptors in the brain and are found in the neocortex, cerebellum, hippocampus, and the dorsal horn [45,47]. Adenosine \( A_{2A} \) receptors are more widely distributed throughout the entire brain and the peripheral nervous system; they are found on pre- and postsynaptic nerve terminals, astrocytes, and the blood-brain barrier [45,48,49]. Adenosine \( A_{2B} \) receptors are less studied but are present on many cell types including astrocytes [50–52]. Adenosine \( A_3 \) receptor expression has been reported throughout the brain with a particular abundance in the hippocampus [53–56]. With this wide expression of multiple adenosine receptor subtypes throughout the brain, it is thought that adenosine subtype-specific physiological processes are achieved by their different affinities for adenosine: \( A_1 \), 3–30 nM; \( A_{2A} \), 1–20 nM; \( A_{2B} \), 5–20 \( \mu \)M; \( A_3 \), >1 \( \mu \)M [57]. Thus, localized modulation of adenosine concentrations would dictate subtype specificity.

Basal extracellular adenosine levels have been estimated in rats with microdialysis and range between 30–970 nM depending on the area of the brain [57]: striatum (80 nM) [58], cortex (120 nM) [59], basal forebrain (30 nM) [60], hippocampus (850 nM) [61], and thalamus (970 nM) [62]. Microdialysis, however, is heavily dependent on the time point at which the samples are collected. Insertion of the microdialysis probe causes damage that leads to a large increase in extracellular neurotransmitter levels. An acclimation period of at least two hours is needed to establish a baseline before measuring neurotransmitter concentrations [29,63]. This is seen in the above estimates in the hippocampus and thalamus which are taken prior to the 2 h-post implantation time point. In fact, other forms of adenosine measurement such as the collection of whole brain samples or cerebral spinal fluid and in vitro pharmaco-dynamic measurement of adenosine suggests lower basal adenosine levels of 50–200 nM [57]. Although these estimates of basal adenosine levels are high enough to activate \( A_1 \) and \( A_{2A} \) receptors, it is important to note that these techniques do not allow for the measurement
of adenosine concentration in the synaptic cleft or at the site of the receptor which may be highly regulated by ENTs and adenosine deaminase.

4. The Role of Adenosine in Pain

The investigation of adenosine’s potential as a therapeutic has revealed its complex role in pain. Adenosine signaling can be anti- or pro-nociceptive depending on the location (central or peripheral), form of pain (acute or chronic), and the adenosine receptor subtype targeted [64,65].

Adenosine A$_1$ receptor agonists are anti-nociceptive [66]. Adenosine A$_1$ receptor activation in peripheral nerve terminals with N6-cyclopentyladenosine (CPA) prevented prostaglandin E$_2$-induced mechanical allodynia in the hindpaw of rats [67]. Adenosine A$_1$ receptor agonists, R-phenylisopropyl-adenosine and N-ethylcarboxamide-adenosine, decreased formalin-induced hyperalgesia in rats (as measured by total licking time during a five minute period) [68]. Mice genetically engineered to lack adenosine A$_1$ receptors had enhanced thermal hyperalgesia and lost the analgesic effect of intrathecal adenosine injection [69]. Administration of CPA, the adenosine A$_1$ receptor agonist, decreased thermal hyperalgesia in normal rats and decreased both mechanical and thermal hyperalgesia in nerve injured rats. These analgesic effects were reversed with administration of the adenosine A$_1$ receptor antagonist, 8-cyclopentyl-1,3-dipropylxanthine (DPCPX) [70]. Adenosine A$_1$ receptors have been found to play a vital role in the adenosine-induced antinociceptive effects of acupuncture in mice where adenosine A$_1$ knock-out mice did not experience the anti-nociceptive effects of acupuncture in an inflammatory or neuropathic pain model [71]. Intrathecal injection of an adenosine A$_1$ agonist in humans decreased postoperative pain [72]. Interestingly, intrathecal injection of adenosine (i.e., non-subtype specific action) prevents post-operative pain but causes headache, suggesting that other adenosine receptor subtypes may play an opposing role in headache when compared to other forms of pain [73,74].

Adenosine A$_2A$ receptor agonists are pro-nociceptive [66]. Adenosine injection into the hindpaw of rats induced a dose-dependent increase in mechanical hyperalgesia in a cAMP-mediated manner. This was prevented by the administration of the adenosine A$_2A$ receptor antagonist, PD 081360-0002. In the same study, a hyperalgesic response similar to adenosine injection was produced with the injection of adenosine A$_2A$ receptor agonists, 5′-(N-ethyl)-carboxamido-adenosine or 2-phenylamino-adenosine [67]. The adenosine A$_2A$ receptor agonist, 2-p-(2-carboxyethyl) phenethylamino-5′-N-ethylcarboxamido adenosine hydrochloride (CGS-21680), enhanced formalin-induced pain in rats while the adenosine A$_2A$ antagonist, 3,7-dimethyl-1-propargylxanthine (DMPX), decreased formalin-induced pain [75]. Another adenosine A$_2A$ agonist, 2-(2-aminoethylamino)-carbonylethylphenoylethylamino-adenosine (APEC), enhanced formalin induced pain in mice [68]. Microdialysis in the rat hindpaw demonstrated that subcutaneous formalin induces a dose-dependent increase in adenosine production, further suggesting adenosine involvement in formalin-induced pain [76]. An adenosine A$_2A$ knock-out mouse had a decreased thermal pain response, suggesting that the presence of adenosine A$_2A$ receptors promotes thermal pain [77]. These knock-out mice also feature increased mechanical hyperalgesia in response to inflammatory-induced pain [78].

Less is known about the specificity of adenosine A$_2B$ and A$_3$ receptors in pain. Adenosine A$_2B$ receptor activation, increasing intracellular cAMP levels similar to adenosine A$_2A$ receptor activation, appear to be pro-nociceptive. Administration of multiple adenosine A$_2B$ antagonists prevented thermal-induced pain in mice [79]. Mechanical and thermal pain was enhanced in an adenosine deaminase (ADO) knock-out mouse, leading to system-wide adenosine elevation. This pain was prevented pharmacologically with the adenosine A$_2B$ antagonist, PSB-1115 (1-propyl-8-p-sulfophenulxanthine). It was also prevented in mice lacking both ADO and adenosine A$_2B$ receptors, suggesting that adenosine A$_2B$ receptors are pronociceptive [80]. PSB-1115 treatment also decreased pain-related behaviors during the early and late phases of formalin-induced pain, suggesting peripheral and central activity of adenosine A$_2B$ receptors in pain [81]. In the same study, adenosine A$_3$ receptors also appeared to be pro-nociceptive. An adenosine A$_3$
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receptor antagonist, PSB-10, prevented the second phase of formalin-induced pain. Interestingly, it had no effect on the first phase suggesting central, but not peripheral, activity of adenosine A3 receptors during inflammatory pain. This pro-nociceptive effect of adenosine A3 receptor activation is interesting since it inhibits adenylate cyclase activity similar to that of adenosine A1 receptors, which are anti-nociceptive. This suggests mechanisms independent of adenylate cyclase activity that are alternatively mediated by adenosine A1 and A3 receptor activation. Contrary to this, however, central administration of an adenosine A3 agonist (MRS5698) was analgesic in a rat model of neuropathic pain [54]. Another A3 receptor agonist, IB-MECA (1-Deoxy-1-[6-[((3-Iodophenyl)methyl)amino]-9H-purin-9-yl]-N-methyl-β-D-ribofuranuronamide), also prevented the development of paclitaxel-induced neuropathic pain [54]. This may highlight a differential role of adenosine A3 receptors in acute (formalin) and persistent (neuropathic) pain states.

These studies suggest that adenosine receptors associated with the Gs alpha subunit (A2A and A2B) are generally pro-nociceptive, while those associated with the Gi alpha subunit (A1 and A3) are generally anti-nociceptive. This is likely due to their effects on intracellular adenylate cyclase activity and cAMP levels that have been implicated in neuropathic and inflammatory pain [82].

5. The Role of Adenosine in Headache Pathophysiology

Numerous key physiological changes that have been observed in chronic headache patients and animal models of trigeminal allodynia can be modulated by adenosine. This suggests that adenosine signaling may contribute to or sustain these changes.

Mitochondrial dysfunction has been hypothesized to play a critical role in migraine. Histological studies have revealed in migraine patients ragged-red fibers and cytochrome-c-oxidase-negative fibers in skeletal muscles [83,84]. Decreased electron transport chain complex activity and metabolic abnormalities are seen in migraine patients [85,86]. We recently identified a decrease in mitochondrial spare respiratory capacity of brain sections from the TNC of the IS and STA rat models [27,87]. Mitochondrial dysfunction can affect the production of extracellular adenosine. The formation of extracellular adenosine is common in tissue hypoxia and is thought to be due to decreased mitochondrial activity such as that seen in the TNC of our models [88]. Artificially inducing mitochondrial dysfunction in isolated rat hepatocytes by treatment with the ATP synthase inhibitor, oligomycin, decreases intracellular ATP levels while increasing AMP levels [89]. Similar to adenosine production from the residual AMP molecule derived from acetate utilization in astrocytes by 5′-nucleotidases (5′NT), this increase in AMP within the cell could be a source for adenosine production when mitochondrial dysfunction is present. 5′NT activity can be enhanced during hypoxia or when ADP levels are high, increasing the production of adenosine [90,91]. ADP also increases the affinity of 5′NT for AMP, potentially facilitating the production of adenosine [92]. Mitochondrial dysfunction and adenosine signaling could affect pain via activation of AMPK, a protein kinase heavily associated with pain plasticity [93]. Extracellular adenosine, increases in the AMP:ATP ratio during hypoxia, and inhibition of 5′NT all enhance AMPK activity [94–97]. It is possible that mitochondrial dysfunction within trigeminal pain processing brain regions exacerbates adenosinergic mechanisms and AMPK activity to produce headache.

The inflammatory mechanisms thought to be involved in migraine can also be influenced by adenosine signaling. We recently found that the key component to dural-stimulated inflammatory-induced trigeminal sensitivity in rats is prostaglandin E2 (PGE2) [28]. The activation of PGE2 receptors increases intracellular cAMP levels, which is the common signaling pathway for adenosine A2A and A2B receptor activation. In fact, adenosine and PGE2 receptors have been shown to have synergistic effects on the suppression of the immune response of T cells [98]. PGE2 and adenosine both increase sodium channel sensitivity in neonatal rat dorsal root ganglion neurons via cAMP mediated mechanisms [99,100]. It was recently found that PGE2 may initiate its painful effects directly by increasing extracellular adenosine concentrations through PKC-mediated mechanisms [101].
The roles of glia have become more appreciated in the migraine field with their activation states potentially affecting headache [102]. Activated microglia and astrocytes could affect pain states by releasing neuroexcitatory or neuroinflammatory mediators associated with headache or even by modulating glutamatergic signaling within the brainstem [103–106]. Adenosine signaling can contribute to astrogliosis via adenosine A₂ receptor activation but can also decrease astrogliosis via adenosine A₁ receptor signaling [107,108]. Adenosine has profound effects on astrocytic regulation of extracellular glutamate levels. Activation of adenosine A₂A receptors decrease the activity of the Na/K ATPase which powers the ionic gradients required for proper glutamate transport into astrocytes through GLT1, which is responsible for deactivation and regulation of extracellular glutamate in the synaptic cleft [49,109]. This modulation is important because extracellular glutamate levels are highly associated with pain. We have found an enhanced increase in extracellular glutamate within the TNC following treatment with GTN in IS rats that can be prevented by the use of headache treatments such as noninvasive vagus nerve stimulation [26,28,29]. Similar to the TNC, modulation of glutamate regulation in the dorsal horn is associated with other forms of chronic pain [110]. Adenosine may contribute to the dysregulation of extracellular glutamate within the TNC, contributing to the development of headache.

Activation of microglia and astrocytes also modulate blood-brain barrier (BBB) permeability which has been associated with migraine patients [111–114]. Breakdown of the BBB has been hypothesized to play a key role in migraine. Serum levels of S100B, a marker for astrocyte activation and a candidate marker for changes in BBB permeability, are increased during and following migraine attacks [115]. MMP-9 and MMP-2, matrix metalloproteinases that digest endothelial cell basal lamina and cause BBB breakdown, are elevated in the serum of migraine patients [116–119]. White matter lesions in MRI images of headache patient brains have also been proposed as focal disruptions of the BBB [118,120,121]. Adenosine signaling can modulate the blood-brain barrier via changes in endothelial cells, as measured by the entrance of intravenously administered macromolecules into the brain [122]. A₁ or A₂A knock-out mice feature a decrease in BBB permeability [122]. A non-selective adenosine receptor agonist decreases transendothelial electrical resistance, increases actinomyosin stress fiber formation, and alters tight junction molecules [122]. The Federal Drug Administration-approved adenosine A₂A agonist, Lexiscan, also increased permeability in an in vitro primary human BBB system [123]. This was further confirmed where a series of novel adenosine A₂A nanoagonists increased BBB permeability as measured by increased delivery of multiple drugs to the brain [124]. It is possible that repeated aberrant adenosine signaling during migraine attacks could cause long-lasting changes to BBB permeability that may contribute to headache persistence by allowing the entry of normally excluded factors into the brainstem that enhance pain hypersensitivity [125].

Furthermore, an adenosine A₂A receptor gene haplotype was found to be associated with migraine with aura [126]. The authors of this study ascribe this to adenosine’s regulation of the calcitonin gene-related peptide (CGRP), a neuropeptide that plays an integral role in migraine. An adenosine A₁ receptor agonist decreased the release of CGRP and trigeminal activity in cats while an A₂A receptor agonist facilitated CGRP’s effect on synaptic transmission [127,128]. Interestingly, electrical stimulation of the trigeminal ganglion increased mRNA and protein levels of both CGRP and adenosine A₂A receptors while decreasing mRNA and protein levels of adenosine A₁ receptors [129]. Cortical spreading depression, thought to be the cause of the aura in migraine, also induces adenosine accumulation in mice [130].

Phonophobia and photophobia are also key characteristics of migraine. Adenosine plays a significant role in both auditory and visual sensory system processing. Adenosine A₁ receptor activation inhibits light-induced responses of intrinsically photosensitive retinal ganglion cells (ipRGCs), the non-image forming cell type that plays a critical role in exacerbating headache pain with light [131,132]. This suggests that A₁ activation may even decrease the effects of photophobia. Furthermore, adenosine signaling plays a critical role in cochlear blood flow and the auditory system and has been investigated for its therapeutic ability to treat hearing loss [133,134].
6. The Role of Adenosine in Common Forms of Headache

Adenosine may play a critical role in multiple forms of headache. This observation is primarily derived from the effect of caffeine, a non-specific adenosine receptor antagonist, to relieve headache pain. In fact, a core component of many abortive headache treatments (Excedrin, Vivarin, Midol, Anacin, Migergot, and Fiorinal) is caffeine. The reasoning to include caffeine was based on the previous hypothesis that migraine was a vascular disorder and the pulsating pain associated with it was due to vasodilation of intra- or extracranial arteries [135]. Adenosine, a known vasodilator, was targeted with caffeine to decrease vasodilation. Although caffeine was found to be efficacious in headache, it is now thought that migraine headache is a neurological disorder, so caffeine’s effect as a therapeutic likely does not involve modulation of vasodilation.

Although caffeine may induce its effects on pain through other targets, clinical evidence exists for adenosine’s role in headache. Papaverine and Sildenafil, adenosine reuptake inhibitors used clinically for heart attacks, and Dipyridamole, an adenosine deaminase inhibitor used to prevent thrombosis, act by increasing extracellular adenosine levels in the brain. Interestingly, each of these drugs can also cause headache in patients [136,137]. Patients undergoing spinal anesthesia treatment to prevent pain often experience a post-operative headache. Caffeine by itself is often prescribed for treatment of this post-dural puncture headache [138]. Additionally, migraine patients without aura experience a surge in serum-adenosine levels during migraine attacks. This surge in adenosine also increases platelet uptake of serotonin, potentially accounting for the low levels of serotonin observed in migraine patients [139,140]. This process was pharmacologically determined to work through adenosine A<sub>2</sub> receptors [140]. The primary mediator of adenosine receptor intracellular signaling, cAMP, which is stimulated during adenosine A<sub>2A</sub> and A<sub>2B</sub> receptor activation, is also suggested to play a direct role in migraine. Cilostazol, an inhibitor of phosphodiesterase type 3 which selectively degrades cAMP, produced headache in both healthy adults and migraineurs [141,142]. Below, we present numerous forms of headache and the evidence supporting the involvement of adenosine in each.

6.1. Delayed Ethanol-Induced Headache

Delayed ethanol-induced headache (DEIH or hangover headache) is similar to migraine headache in that it often features throbbing pain with phono- and photophobia. We previously identified that acetate, an ethanol metabolite, was the key element behind ethanol-induced trigeminal sensitivity in the IS rat model of trigeminal pain that features migraine-like characteristics [25]. In fact, clinical observations identified that the inclusion of sodium acetate as a buffer during kidney dialysis in diabetic patients also causes headache [143]. Acetate has profound effects on the formation of extracellular adenosine with many neuronal consequences of ethanol thought to be induced via acetate-mediated adenosinergic mechanisms [2]. In the brain, acetate is utilized almost exclusively by astrocytes as an energy source, resulting in a residual AMP molecule that is readily converted to adenosine [2,144]. This process is thought to be the mechanism behind the acetate-induced adenosine-mediated effects of ethanol. In humans, serum-adenosine levels rise following ethanol consumption. In wild-type rats, ethanol perfusion in the basal forebrain with microdialysis increases extracellular adenosine levels [60,144–146]. The headache associated with acetate is potentially due to acetate’s effect on adenosine concentrations in the brain. Not surprisingly, caffeine is widely used to treat DEIH in humans, suggesting adenosine as a key aspect to the pain phase of DEIH [147]. We also found that caffeine treatment in the IS rats prevented ethanol-induced trigeminal sensitivity [25]. Caffeine’s efficacy in the treatment of DEIH suggests adenosine receptor signaling as a critical component to the pain phase of ethanol ingestion.

6.2. Caffeine Withdrawal Headache

Caffeine withdrawal headaches are very common and occur when individuals decrease their daily consumption of products containing caffeine, such as coffee or tea. The headache is relieved within
one hour after re-exposure to caffeine [17]. Since basal adenosine signaling is essential for proper neuronal function, chronic exposure to caffeine likely has significant compensatory effects on adenosine signaling to sustain basal adenosine function [148]. During chronic adenosine receptor antagonism with daily caffeine use, this essential basal adenosine signal could be sustained through up-regulation of adenosine receptors, increasing extracellular adenosine content, or modulation of downstream mediators of adenosine receptor signaling. Compensatory mechanisms such as these are likely at play due to the clinical observation that caffeine tolerance quickly develops in humans [149]. In fact, chronic exposure to caffeine in zebrafish increased expression of adenosine $A_1$ and $A_{2A}$ receptors, but not $A_{2B}$ and $A_3$ receptor subtypes [150]. Chronic exposure to caffeine in rats increased $A_1$ receptor expression in the hippocampus [151]. Chronic exposure to caffeine in mice induces a dose-dependent increase in $A_{2A}$ receptor expression in the striatum and shifted low-affinity $A_1$ receptors to a high affinity state [152]. Chronic caffeine intake also increases plasma adenosine levels [153]. Each of these seemingly compensatory changes caused by chronic caffeine exposure could increase the adenosine signal in the absence of caffeine during the withdrawal period, contributing to caffeine withdrawal headache that is relieved by re-exposure to caffeine.

### 6.3. Cluster Headaches

Cluster headaches are a rare but intensely painful headache that is instantly debilitating [154]. One common treatment method for these attacks is the inhalation of pure oxygen [155]. Decreased oxygen supply has been associated with an increase in adenosine levels [156,157]. The effect of pure oxygen to relieve this pain may be directly tied to a reduction in adenosine levels. Platelet aggregation is impaired in cluster headache patients and could be a sign that adenosine levels are higher in these individuals since adenosine has been shown to dose-dependently inhibit platelet aggregation in humans [158,159]. Therefore, cluster headache patients may experience greater basal levels of adenosine that could contribute to the pain phase of this disorder. Increasing oxygen content could ameliorate this by decreasing adenosine content to relieve pain.

### 6.4. Sleep-Wake Cycle Related Headaches

Some forms of headache such as migraine, cluster headache, and the hypnic headache are associated with the sleep-wake cycle [17,160,161]. Adenosine plays a critical role in circadian rhythms [162]. Extracellular adenosine accumulates during sleep deprivation to promote sleep and decreases during sleep [162,163]. Caffeine is used as a stimulant by acting as an adenosine receptor antagonist to decrease adenosine’s sleep-promoting activities [162]. These forms of headache may occur due to the daily modulation of adenosine receptor expression within the brain or adenosine metabolism enzymes [164,165]. Upon waking when adenosine levels begin to rise again, there may be a partial imbalance in the adenosine receptor signal output or changes in receptor subtype-specific output, contributing to headache via adenosinergic mechanisms. Interestingly, there is even a high prevalence of sleep disorders that afflict cluster headache patients [166]. These patients may have aberrantly high levels of adenosine due to lack of sleep. Similar to cluster headaches, caffeine is used to treat hypnic headaches and has been found to be the most effective treatment option for these patients [167].

### 6.5. Post-Traumatic Headache

Post traumatic headache occurs in nearly all patients who experience mild or severe traumatic brain injury (TBI) [168]. This headache can develop further into chronic post traumatic headache. Clinical studies have found that interstitial brain adenosine levels increase during oxygen desaturation events following TBI and are thought to contribute to secondary events following TBI [169]. In the context of TBI, adenosine is thought to have a neuroprotective effect following injury and thus, use of adenosine receptor agonists are currently being investigated for TBI treatment [6]. Although adenosine
may serve a role in these neuroprotective processes, they may contribute to the headache or chronic headache that develops following injury.

6.6. Menstrual Migraine

Menstrual migraine occurs exclusively during the first two days of menstruation in women [17]. During this time, women experience a sudden decrease in estrogen and progesterone levels [170]. Estradiol, an estrogen receptor-alpha ligand, was found to increase adenosine A1 receptor expression in a breast cancer cell line, suggesting that estrogen receptor activation can modulate adenosine receptor expression and activity [5]. Ovariectomized rats that have systemically depleted estrogen levels, also have a four-fold decrease in adenosine A2A receptors [171]. In fact, birth controls that contain estrogen are also associated with headache [172]. Progesterone and estradiol were shown to increase adenosine’s depressant actions in rat cerebral cortical neurons, suggesting that these hormones could increase the adenosine signal [173]. Rats treated with progesterone were found to have lower adenosine deaminase levels, the enzyme responsible for breaking down adenosine, suggesting increased adenosine concentrations in response to progesterone treatment [174]. Although these data would suggest that estrogen or progesterone withdrawal at the start of menses would decrease adenosine levels, they demonstrate the critical cross-talk between sex hormones and the adenosinergic system which may contribute to menstrual migraine.

7. Adenosine and Common Headache Triggers

Headache triggers increase the probability a headache will develop in a migraine patient within 24 hours. Triggers can be dietary (alcohol, aspartame, cheese), environmental (perfumes, the *Umbellularia californica* tree, pollution), physical (extraneous activity, inactivity, exposure to intense light), or even psychological (anxiety, stress). Adenosine is involved in a number of these common headache triggers and may act as a unifying mediator for the induction of headache by them.

Glyceryl trinitrate (GTN) is a common chemical headache trigger that induces an extended headache period in chronic migraine patients, but only a short-lived headache in normal individuals [175,176]. In the IS rat model, GTN induces an extended period of trigeminal sensitivity and increased extracellular glutamate in the TNC [26,28]. GTN can potentiate the effects of adenosine on platelet aggregation, and is thought to occur through modulation of adenosine’s intracellular downstream effector molecule, cAMP [177]. Through this mechanism, GTN could potentially enhance the adenosine signal even when adenosine levels or adenosine receptor expression are not modified. GTN also has multiple effects on mitochondrial function, an organelle that has been suggested to play a role in the development of migraine [27,178,179]. GTN treatment was found to decrease the activity of complex I in the electron transport chain within isolated rat aortas, decreasing oxygen consumption [180]. During periods of decreased mitochondrial function, extracellular levels of adenosine are known to increase [88,181–183]. It could be through this mechanism that GTN induces trigeminal pain.

The headache tree, or *Umbellularia californica*, is known to induce severe headache attacks following inhalation of its vapors [184]. Studies in isolated rat trigeminal ganglion neurons revealed that the monoterpene ketone, umbellulone, is the chemical substance responsible for this tree’s effect on headache and that it occurs by selectively activating transient receptor potential ankyrin 1 (TRPA1) channels in the trigeminal system to induce aberrant activity that is interpreted as headache [184]. Other environmental stimuli such as cigarette smoke, chlorine, formaldehyde, and pollution are also thought to induce headache via their activation of TRPA1 or other TRP channel subtypes such as transient receptor potential vanilloid 1 (TRPV1) or transient receptor potential menthol 8 (TRPM8) [185]. Similar to adenosine A2A and A2B receptors, these channels increase calcium conductance via increases in cAMP intracellular concentrations. Adenosine A1 and A3 receptors reduce cAMP levels, decreasing calcium conductance [1]. Furthermore, cAMP can actively inhibit TRP channel activity [186]. In fact, caffeine inhibits human TRPA1 channels through unknown mechanisms [187]. There may be a
converging modulation of intracellular cAMP levels by both TRPA1 and adenosine receptors during pain processing. Furthermore, the activation of TRPA1 channels on human odontoblasts, cells of neural crest origin, stimulates the release of ATP which can then be quickly metabolized to adenosine [188]. Similarly, TRPV4 activation can cause ATP release in primary urothelial cells [189]. ATP has also been shown to increase capsaicin-evoked currents in TRPV1 expressing HEK293 cells, suggesting that ATP or adenosine following extracellular ATP metabolism can interact with and modulate TRPV1 activity [190]. This mechanism of ATP release and metabolism to adenosine following TRP channel activation likely also occurs in the brain and may serve as a conduit between TRP channels and headache in various environmental triggers.

Anxiety and stress is a very common headache trigger. Interestingly, adenosine plays a vital role in anxiety [3,8]. Clinical and rodent data support the concept that adenosine can be both anxiogenic and anxiolytic, with A2A receptors playing a critical role [8]. In support of adenosine being anxiogenic, an adenosine A2A receptor antagonist reduced anxiety and stress in rodents that was caused by maternal separation or chronic unpredictable stress [191,192]. Although a genetic polymorphism of the A2A receptor gene is associated with individuals diagnosed with panic disorders, the effects of these polymorphisms on receptor function have not been investigated [193,194]. In support of adenosine being anxiolytic, A1 or A2A knock-out mice feature anxiety-like behaviors [77,195]. Furthermore, inducing stress in rats enhanced the expression of adenosine A2A receptors while the overexpression of adenosine A2A receptors decreased anxiety in an open field test [196]. Although not fully understood, the critical role of adenosine A2A receptors in both anxiety and headache is interesting because it provides a potential common target that may play a role in both.

8. Conclusions

The role of adenosine in different forms of headache, headache triggers, and basic headache physiology suggests it as a core component to headache pain. Although adenosine signaling can produce differential effects on pain, depending on the receptor subtype activated, adenosine A2A receptors are likely the critical subtype involved in headache pain. Adenosine signaling may initiate headache pain by modulation of intracellular cAMP production or AMPK activity that can change neuronal conductance within critical trigeminal pain processing brain regions. It may also directly contribute to pain by modulating extracellular glutamate within the synaptic cleft of tripartite synapses in the TNC or through modulation of the BBB and enhancement of gliosis. Migraineurs may be more susceptible to these adenosinergic impacts on headache due to the potential presence of mitochondrial dysfunction which could facilitate the production of excess levels of adenosine. If adenosine plays a role in headache triggers and various types of headache, it is interesting to note that systemic changes in adenosine signaling produce headache as opposed to pain across the entire body. This suggests something unique about headache pathophysiology that makes the trigeminal system more susceptible to the effects of adenosine on pain.

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References
1. Jacobson, K.A.; Gao, Z.-G. Adenosine receptors as therapeutic targets. *Nat. Rev. Drug Discov.* 2006, 5, 247–264. [CrossRef] [PubMed]
2. Carmichael, F.J.; Orrego, H.; Israel, Y. Acetate-induced adenosine mediated effects of ethanol. *Alcohol Alcohol. Suppl.* 1993, 2, 411–418. [PubMed]
3. Correa, M.; Font, L. Is there a major role for adenosine A2A receptors in anxiety? *Front. Biosci. J. Virtual Libr.* 2008, 13, 4058–4070. [CrossRef]

4. Gomes, C.V.; Kaster, M.P.; Tomé, A.R.; Agostinho, P.M.; Cunha, R.A. Adenosine receptors and brain diseases: Neuroprotection and neurodegeneration. *Biochim. Biophys. Acta* 2011, 1808, 1380–1399. [CrossRef] [PubMed]

5. Lin, Z.; Yio, P.; Reierstad, S.; O’Halloran, M.; Coon, V.J.S.; Pearson, E.K.; Mutlu, G.M.; Bulun, S.E. Adenosine A1 receptor, a target and regulator of estrogen receptoralpha action, mediates the proliferative effects of estradiol in breast cancer. *Oncogene* 2010, 29, 1114–1122. [CrossRef] [PubMed]

6. Lusardi, T. Adenosine Neuromodulation and Traumatic Brain Injury. *Curr. Neuropharmacol.* 2009, 7, 228–237. [CrossRef] [PubMed]

7. Rahman, A. The Role of Adenosine in Alzheimer’s disease. *Curr. Neuropharmacol.* 2009, 7, 207–216. [CrossRef] [PubMed]

8. Yamada, K.; Kobayashi, M.; Kanda, T. Involvement of adenosine A2A receptors in depression and anxiety. *Int. Rev. Neurobiol.* 2014, 119, 373–393. [PubMed]

9. Sollevi, A. Adenosine for pain control. *Acta Anaesthesiol. Scand. Suppl.* 1997, 110, 135–136. [CrossRef] [PubMed]

10. Schwedt, T.J.; Green, A.L.; Dodick, D.W. Occipital Nerve Stimulation for Migraine: Update from Recent Multicenter Trials. *Prog. Neurol. Surg.* 2015, 29, 117–126. [PubMed]

11. Cupini, L.M.; Calabresi, P. Medication-overuse headache: Pathophysiological insights. *J. Headache Pain* 2005, 6, 199–202. [PubMed]

12. Derry, C.J.; Derry, S.; Moore, R.A. Caffeine as an analgesic adjuvant for acute pain in adults. *Cochrane Database Syst. Rev.* 2014, 12, CD009281.

13. Hughes, J.R. Clinical importance of caffeine withdrawal. *N. Engl. J. Med.* 1992, 327, 1160–1161. [CrossRef] [PubMed]

14. Ward, N.; Whitney, C.; Avery, D.; Dunner, D. The analgesic effects of caffeine in headache. *Pain* 1991, 44, 151–155. [PubMed]

15. Olesen, J.; Burstein, R.; Ashina, M.; Tfelt-Hansen, P. Origin of pain in migraine: evidence for peripheral sensitisation. *Lancet Neurol.* 2009, 8, 679–690. [CrossRef]

16. Noseda, R.; Burstein, R. Migraine pathophysiology: Anatomy of the trigeminovascular pathway and associated neurological symptoms, CSD, sensitization and modulation of pain. *Pain* 2013. [CrossRef] [PubMed]

17. Strassman, A.M.; Raymond, S.A.; Burstein, R. Sensitization of meningeal sensory neurons and the origin of headaches. *Nature* 1996, 384, 560–564. [CrossRef] [PubMed]

18. Olesen, J.; Burstein, R.; Ashina, M.; Tfelt-Hansen, P. Origin of pain in migraine: evidence for peripheral sensitisation. *Lancet Neurol.* 2009, 8, 679–690. [CrossRef]

19. Noseda, R.; Burstein, R. Migraine pathophysiology: Anatomy of the trigeminovascular pathway and associated neurological symptoms, CSD, sensitization and modulation of pain. *Pain* 2013. [CrossRef] [PubMed]

20. Burstein, R.; Jakubowski, M. Unitary hypothesis for multiple triggers of the pain and strain of migraine. *J. Comp. Neurol.* 2005, 493, 9–14. [CrossRef] [PubMed]

21. Malick, A.; Burstein, R. Peripheral and central sensitization during migraine. *Funct. Neurol.* 2000, 15, 28–35. [PubMed]

22. Maxwell, C.R.; Spangenberg, R.J.; Hoek, J.B.; Silberstein, S.D.; Oshinsky, M.L. Acetate Causes Alcohol Hangover Headache in Rats. *PLoS ONE* 2010, 5, e15963. [CrossRef] [PubMed]

23. Oshinsky, M.L.; Gomonchareonsiri, S. Episodic dural stimulation in awake rats: a model for recurrent headache. *Headache* 2007, 47, 1026–1036. [CrossRef] [PubMed]
27. Fried, N.T.; Moffat, C.; Seifert, E.L.; Oshinsky, M.L. Functional mitochondrial analysis in acute brain sections from adult rats reveals mitochondrial dysfunction in a rat model of migraine. *Am. J. Physiol. Cell Physiol.* 2014, 307, C1017–C1030. [CrossRef] [PubMed]

28. Oshinsky, M.L.; Murphy, A.L.; Hekierski, H.; Cooper, M.; Simon, B.J. Noninvasive vagus nerve stimulation as treatment for trigeminal allodynia. *Pain* 2014, 155, 1037–1042. [CrossRef] [PubMed]

29. Oshinsky, M.L.; Luo, J. Neurochemistry of trigeminal activation in an animal model of migraine. *Headache* 2006, 46, S39–44. [CrossRef] [PubMed]

30. Vermeer, L.M.M.; Gregory, E.; Winter, M.K.; McCarson, K.E.; Berman, N.E.J. Exposure to Bisphenol A Exacerbates Migraine-Like Behaviors in a Multibehavior Model of Rat Migraine. *Toxicol. Sci.* 2014, 137, 416–427. [CrossRef]

31. Wieseler, J.; Ellis, A.; Sprunger, D.; Brown, K.; McFadden, A.; Mahoney, J.; Rezvani, N.; Maier, S.F.; Watkins, L.R. A novel method for modeling facial allodynia associated with migraine in awake and freely moving rats. *J. Neurosci. Methods* 2010, 185, 236–245. [CrossRef] [PubMed]

32. Oshinsky, M.L.; Sanghvi, M.M.; Maxwell, C.R.; Gonzalez, D.; Spangenberg, R.J.; Cooper, M.; Silberstein, S.D. Spontaneous trigeminal allodynia in rats: A model of primary headache. *Headache* 2012, 52, 1336–1349. [CrossRef] [PubMed]

33. Drury, A.N.; Szent-Györgyi, A. The physiological activity of adenine compounds with especial reference to their action upon the mammalian heart. *J. Physiol.* 1929, 68, 213–237. [CrossRef] [PubMed]

34. Brundege, J.M.; Dunwiddie, T.V. Modulation of excitatory synaptic transmission by adenosine released from single hippocampal pyramidal neurons. *J. Neurosci. Off. J. Soc. Neurosci.* 1996, 16, 5603–5612.

35. Lovatt, D.; Xu, Q.; Liu, W.; Takano, T.; Smith, N.A.; Schnerrmann, J.; Tieu, K.; Nedergaard, M. Neuronal adenosine release, and not astrocytic ATP release, mediates feedback inhibition of excitatory activity. *Proc. Natl. Acad. Sci. USA* 2012, 109, 6265–6270. [CrossRef] [PubMed]

36. Manzoni, O.J.; Manabe, T.; Nicoll, R.A. Release of adenosine by activation of NMDA receptors in the hippocampus. *Science* 1994, 265, 2098–2101. [CrossRef] [PubMed]

37. Martin, E.D.; Fernández, M.; Perea, G.; Pascual, O.; Haydon, P.G.; Araque, A.; Ceña, V. Adenosine released by astrocytes contributes to hypoxia-induced modulation of synaptic transmission. *Glia* 2007, 55, 36–45. [CrossRef] [PubMed]

38. Nadjar, A.; Blustein, T.; Aubert, A.; Laye, S.; Haydon, P.G. Astrocyte-derived adenosine modulates increased sleep pressure during inflammatory response. *Glia* 2013, 61, 724–731. [CrossRef] [PubMed]

39. Wall, M.; Dale, N. Activity-Dependent Release of Adenosine: A Critical Re-Evaluation of Mechanism. *Curr. Neuropharmacol.* 2008, 6, 329–337. [CrossRef] [PubMed]

40. Zorec, R.; Araque, A.; Carmignoto, G.; Haydon, P.G.; Verkhratsky, A.; Parpura, V. Astroglial excitability and gliotransmission: An appraisal of Ca2+ as a signalling route. *Int. J. Mol. Sci.* 2012, 307, E39–44. [CrossRef] [PubMed]

41. Sheth, S.; Brito, R.; Mukherjea, D.; Rybak, L.P.; Ramkumar, V. Adenosine Receptors: Expression, Function and Regulation. *Int. J. Mol. Sci.* 2014, 15, 2024–2052. [CrossRef] [PubMed]

42. Zimmermann, M.A.; Tak, E.; Ehrentraut, S.F.; Kaplan, M.; Giebler, A.; Weng, T.; Choi, D.-S.; Blackburn, M.R.; Kam, I.; Eltzschig, H.K.; et al. Equilibrative nucleoside transporter (ENT)-1-dependent elevation of extracellular adenosine protects the liver during ischemia and reperfusion. *Hepatol. Baltim. Md.* 2013, 58, 1766–1778. [CrossRef] [PubMed]

43. McGaraughty, S.; Cowart, M.; Jarvis, M.F.; Berman, R.F. Anticonvulsant and antinociceptive actions of novel adenosine kinase inhibitors. *Curr. Top. Med. Chem.* 2005, 5, 43–58. [CrossRef] [PubMed]

44. Zimmermann, H. Extracellular metabolism of ATP and other nucleotides. *Naunyn. Schmiedebergs Arch. Pharmacol.* 2000, 362, 299–309. [CrossRef] [PubMed]

45. Sachdeva, S.; Gupta, M. Adenosine and its receptors as therapeutic targets: An overview. *ASN Neuro* 2012, 4, e00080. [CrossRef] [PubMed]

46. Castro, L.R.V.; Guiot, E.; Polito, M.; Paupardin-Tritsch, D.; Vincent, P. Decoding spatial and temporal features of neuronal CAMP/PKA signaling with FRET biosensors. *Biotechnol. J.* 2014, 9, 192–202. [CrossRef] [PubMed]

47. Poulsen, S.A.; Quinn, R.J. Adenosine receptors: new opportunities for future drugs. *Bioorg. Med. Chem.* 1998, 6, 619–641. [CrossRef]

48. Fredholm, B.B.; Cunha, R.A.; Svenningsson, P. Pharmacology of adenosine A2A receptors and therapeutic applications. *Curr. Top. Med. Chem.* 2003, 3, 413–426. [CrossRef] [PubMed]
49. Matos, M.; Augusto, E.; Agostinho, P.; Cunha, R.A.; Chen, J.-F. Antagonistic Interaction between Adenosine A2A Receptors and Na+/K+-ATPase-a2 Controlling Glutamate Uptake in Astrocytes. J. Neurosci. 2013, 33, 18492–18502. [CrossRef] [PubMed]

50. Cacciari, B.; Pastorin, G.; Bolcato, C.; Spalluto, G.; Bacilieri, M.; Moro, S. A2B adenosine receptor antagonists: recent developments. Mini Rev. Med. Chem. 2005, 5, 1053–1060. [CrossRef] [PubMed]

51. Eusemann, T.N.; Willmroth, F.; Fiebich, B.; Biber, K.; van Calker, D. Adenosine Receptors Differentially Regulate the Expression of Regulators of G-Protein Signalling (RGS) 2, 3 and 4 in Astrocyte-Like Cells. PLoS ONE 2015, 10, e0134934. [CrossRef] [PubMed]

52. Rosi, S.; McGann, K.; Hauss-Wegrzyniak, B.; Wenk, G.L. The influence of brain inflammation upon neuronal adenosine A2B receptors. J. Neurochem. 2003, 86, 220–227. [CrossRef] [PubMed]

53. Dunwiddie, T.V.; Diao, L.; Kim, H.O.; Jiang, J.-L.; Jacobson, K.A. Activation of Hippocampal Adenosine A3 Receptors Produces a Desensitization of A1 Receptor-Mediated Responses in Rat Hippocampus. J. Neurosci. 1997, 17, 607–614. [PubMed]

54. Little, J.W.; Ford, A.; Symons-Liguori, A.M.; Chen, Z.; Doyle, T.; Xie, J.; Luongo, L.; Tosh, D.K.; Maione, S.; et al. Endogenous adenosine A3 receptor activation selectively alleviates persistent pain states. Brain 2015, 138 Pt 1, 28–35. [CrossRef] [PubMed]

55. Livingston, M.; Heaney, L.G.; Ennis, M. Adenosine, inflammation and asthma—A review. Inflamm. Res. 2004, 53, 171–178. [CrossRef]

56. Rivkees, S.A.; Thevananther, S.; Hao, H. Are A3 adenosine receptors expressed in the brain? Neuroreport 2000, 11, 1025–1030. [CrossRef] [PubMed]

57. Latini, S.; Pedata, F. Adenosine in the central nervous system: Release mechanisms and extracellular concentrations. J. Neurochem. 2001, 79, 463–484. [CrossRef] [PubMed]

58. Ballarin, M.; Fredholm, B.B.; Ambrosio, S.; Mahy, N. Extracellular levels of adenosine and its metabolites in the striatum of awake rats: inhibition of uptake and metabolism. Acta Physiol. Scand. 1991, 142, 97–103. [CrossRef] [PubMed]

59. Pazzagli, M.; Corsi, C.; Latini, S.; Pepeu, G. In vivo regulation of extracellular adenosine levels in the cerebral cortex by NMDA and muscarinic receptors. Eur. J. Pharmacol. 1994, 254, 277–282. [CrossRef] [PubMed]

60. Dobolyi, A.; Reichart, A.; Szikra, T.; Nyitrai, G.; Kékesi, K.A.; Juhász, G. Sustained depolarisation induces changes in the extracellular concentrations of purine and pyrimidine nucleosides in the rat thalamus. Neurochem. Int. 2000, 37, 71–79. [CrossRef]

61. Sawynok, J. Adenosine receptor activation and nociception. Eur. J. Pharmacol. 1998, 347, 1–11. [CrossRef]

62. Sawynok, J.; Liu, X.J. Adenosine in the spinal cord and periphery: Release and regulation of pain. Prog. Neurobiol. 2003, 69, 313–340. [CrossRef]

63. Ferrari, S.; Diamond, I.; Goldberg, S.R.; Yao, L.; Hourani, S.M.O.; Huang, Z.L.; Urade, Y.; Kitchen, I. Adenosine A2A receptors in ventral striatum, hypothalamus and nociceptive circuitry. Implications for drug addiction, sleep and pain. Prog. Neurobiol. 2007, 83, 332–347. [CrossRef] [PubMed]

64. Taiwo, Y.O.; Levine, J.D. Direct cutaneous hyperalgesia induced by adenosine. Neuroscience 1990, 38, 757–762. [CrossRef]

65. Karlsten, R.; Gerdh, T.; Post, C. Local antinociceptive and hyperalgesic effects in the formalin test after peripheral administration of adenosine analogues in mice. Pharmacol. Toxicol. 1992, 70, 434–438. [CrossRef]

66. Johansen, J.P.; Fields, H.L.; Manning, B.H. The affective component of pain in rodents: Direct evidence for a contribution of the anterior cingulate cortex. Proc. Natl. Acad. Sci. USA 2001, 98, 8077–8082. [CrossRef] [PubMed]
70. Gong, Q.-J.; Li, Y.-Y.; Xin, W.-J.; Wei, X.-H.; Cui, Y.; Wang, J.; Liu, Y.; Liu, C.-C.; Li, Y.-Y.; Liu, X.-G. Differential effects of adenosine A1 receptor on pain-related behavior in normal and nerve-injured rats. *Brain Res.* 2010, 1361, 23–30. [CrossRef] [PubMed]

71. Goldman, N.; Chen, M.; Fujita, T.; Xu, Q.; Peng, W.; Liu, W.; Jensen, T.K.; Pei, Y.; Wang, F.; Han, X.; et al. Adenosine A1 receptors mediate local anti-nociceptive effects of acupuncture. *Nat. Neurosci.* 2010, 13, 883–888. [CrossRef] [PubMed]

72. Gordh, T.; Karlsten, R.; Kristensen, J. Intervention with spinal NMDA, adenosine, and NO systems for pain modulation. *Ann. Med.* 1995, 27, 229–234. [CrossRef] [PubMed]

73. Eisenach, J.C.; Hood, D.D.; Curry, R. Phase I Safety Assessment of Intrathecal Injection of an American Formulation of Adenosine in Humans. *J. Am. Soc. Anesthesiol.* 2002, 96, 24–28. [CrossRef]

74. Eisenach, J.C.; Curry, R.; Hood, D.D. Dose Response of Intrathecal Adenosine in Experimental Pain and Allodynia. *J. Am. Soc. Anesthesiol.* 2002, 97, 938–942. [CrossRef]

75. Doak, G.J.; Sawynok, J. Complex role of peripheral adenosine in the genesis of the response to subcutaneous formalin in the rat. *Eur. J. Pharmacol.* 1995, 281, 311–318. [CrossRef]

76. Liu, X.J.; White, T.D.; Sawynok, J. Potentiation of formalin-evoked adenosine release by an adenosine kinase inhibitor and an adenosine deaminase inhibitor in the rat hind paw: a microdialysis study. *Eur. J. Pharmacol.* 2000, 408, 143–152. [CrossRef]

77. Ledent, C.; Vaugeois, J.M.; Schiffmann, S.N.; Pedrazzini, T.; El Yacoubi, M.; Vanderhaeghen, J.J.; Costentin, J.; Heath, J.K.; Vassart, G.; Parmentier, M. Aggressiveness, hypoalgesia and high blood pressure in mice lacking the adenosine A2a receptor. *Nature* 1997, 388, 674–678. [PubMed]

78. Li, L.; Hao, J.X.; Fredholm, B.B.; Schulte, G.; Wiesenfeld-Hallin, Z.; Xu, X.J. Peripheral adenosine A2A receptors are involved in carrageenan-induced mechanical hyperalgesia in mice. *Neuroscience* 2010, 170, 923–928. [CrossRef] [PubMed]

79. Abo-Salem, O.M.; Hayallah, A.M.; Bilkei-Gorzo, A.; Filipek, B.; Zimmer, A.; Müller, C.E. Antinociceptive Effects of Novel A2B Adenosine Receptor Antagonists. *J. Pharmacol. Exp. Ther.* 2004, 308, 358–366. [CrossRef] [PubMed]

80. Hu, X.; Adebiyi, M.G.; Luo, J.; Sun, K.; Le, T.-T.T.; Zhang, Y.; Wu, H.; Zhao, S.; Karmouty-Quintana, H.; Liu, H.; et al. Sustained Elevated Adenosine via ADORA2B Promotes Chronic Pain through Neuro-immune Interaction. *Cell Rep.* 2016, 16, 106–119. [CrossRef] [PubMed]

81. Bilkei-Gorzo, A.; Abo-Salem, O.M.; Hayallah, A.M.; Michel, K.; Müller, C.E.; Zimmer, A. Adenosine receptor subtype-selective antagonists in inflammation and hyperalgesia. *Naunyn. Schmiedebergs Arch. Pharmacol.* 2008, 377, 65–76. [CrossRef] [PubMed]

82. Wang, H.; Xu, H.; Wu, L.-J.; Kim, S.S.; Chen, T.; Koga, K.; Descalzi, G.; Gong, B.; Vadakkan, K.I.; Zhang, X.; et al. Identification of an Adenylyl Cyclase Inhibitor for Treating Neuropathic and Inflammatory Pain. *Sci. Transl. Med.* 2011, 3, 65ra3. [CrossRef] [PubMed]

83. Uncini, A.; Lodì, R.; Di Muzio, A.; Silvestri, G.; Servidei, S.; Lugaresi, A.; Iotti, S.; Zaniol, P.; Barbiroli, B. Abnormal brain and muscle energy metabolism shown by 31P-MRS in familial hemiplegic migraine. *J. Neurol. Sci.* 1995, 129, 214–222. [CrossRef]

84. Welch, K.M.; Levine, S.R.; D’Andrea, G.; Schultz, L.R.; Helpner, J.A. Preliminary observations on brain energy metabolism in migraine studied by in vivo phosphorus 31 NMR spectroscopy alterations in a migraine patient. *J. Neurol. Sci.* 1991, 104, 182–189. [CrossRef]

85. Sangiorgi, S.; Mochi, M.; Cortelli, P.; Monari, L.; Pierangeli, G.; Montagna, P. Abnormal platelet mitochondrial function in patients affected by migraine with and without aura. *Cephalalgia Int. J. Headache* 1994, 14, 21–23. [CrossRef] [PubMed]

86. Eltzschig, H.K.; Thompson, L.F.; Karhausen, J.; Cotta, R.J.; Ibla, J.C.; Robson, S.C.; Colgan, S.P. Endogenous adenosine produced during hypoxia attenuates neutrophil accumulation: coordination by extracellular nucleotide metabolism. *Blood* 2004, 104, 3986–3992. [CrossRef] [PubMed]
89. Cannon, J.R.; Harvison, P.J.; Rush, G.F. The effects of fructose on adenosine triphosphate depletion following mitochondrial dysfunction and lethal cell injury in isolated rat hepatocytes. *Toxicol. Appl. Pharmacol.* 1991, 108, 407–416. [CrossRef] [PubMed]

90. Bak, M.I.; Ingwall, J.S. Acidosis during ischemia promotes adenosine triphosphate resynthesis in postischemic rat heart. In vivo regulation of 5′-nucleotidase. *J. Clin. Investig.* 1994, 93, 40–49. [CrossRef] [PubMed]

91. Bak, M.I.; Ingwall, J.S. Regulation of cardiac AMP-specific 5′-nucleotidase during ischemia mediates ATP resynthesis on reflow. *Am. J. Physiol.* 1998, 274, C992–C1001. [PubMed]

92. Darvish, A.; Metting, P.J. Purification and regulation of an AMP-specific cytosolic 5′-nucleotidase from dog heart. *Am. J. Physiol.* 1993, 264, H1528–H1534. [PubMed]

93. Price, T.J.; Dussor, G. AMPK: An emerging target for modification of injury-induced pain plasticity. *Neurosci. Lett.* 2013, 557 Pt A, 9–18. [CrossRef] [PubMed]

94. Aymerich, I.; Foufelle, F.; Ferré, P.; Casado, F.J.; Pastor-Anglada, M. Extracellular adenosine activates AMP-dependent protein kinase (AMPK). *J. Cell Sci.* 2006, 119, 1612–1621. [CrossRef] [PubMed]

95. Kulkarni, S.S.; Karlsson, H.K.R.; Szekeres, F.; Chibal, A.V.; Krook, A.; Zierath, J.R. Suppression of 5′-nucleotidase enzymes promotes AMP-activated protein kinase (AMPK) phosphorylation and metabolism in human and mouse skeletal muscle. *J. Biol. Chem.* 2011, 286, 34567–34574. [PubMed]

96. Plaideau, C.; Liu, J.; Hartlieb-Geschwindner, J.; Bastin-Coyette, L.; Bontemps, F.; Oscarsson, J.; Hue, L.; Rider, M.H. Overexpression of AMP-metabolizing enzymes controls adenine nucleotide levels and AMPK activation in HEK293T cells. *FASEB J.* 2012, 26, 2685–2694. [CrossRef] [PubMed]

97. Wang, W.; Yang, X.; Silanes, I.L.; de; Carling, D.; Gorospe, M. Increased AMP:ATP Ratio and AMP-activated Protein Kinase Activity during Cellular Senescence Linked to Reduced HuR Function. *J. Biol. Chem.* 2003, 278, 27016–27023. [CrossRef] [PubMed]

98. Mandapathil, M.; Szczepanski, M.J.; Szajnik, M.; Ren, J.; Jackson, E.K.; Johnson, J.T.; Gorelik, E.; Lang, S.; England, S.; Bevan, S.; Docherty, R.J. PGE2 modulates the tetrodotoxin-resistant sodium current in neonatal rat dorsal root ganglion neurones via the cyclic AMP-protein kinase A cascade. *J. Physiol.* 1996, 495 Pt 2, 429–440. [CrossRef] [PubMed]

99. Kidd, B.L.; Urban, L.A. Mechanisms of inflammatory pain. *Br. J. Anaesth.* 2001, 87, 3–11. [CrossRef] [PubMed]

100. Ferrari, L.F.; Levine, E.; Levine, J.D. Role of a novel nociceptor autocrine mechanism in chronic pain. *Eur. J. Neurosci.* 2013, 37, 1705–1713. [CrossRef] [PubMed]

101. Bartley, J. Could glial activation be a factor in migraine? *Med. Hypotheses* 2009, 72, 255–257. [CrossRef] [PubMed]

102. Beschorner, R.; Simon, P.; Schauer, N.; Mittelbronn, M.; Schluesener, H.J.; Trautmann, K.; Dietz, K.; Meyermann, R. Reactive astrocytes and activated microglial cells express EAAT1, but not EAAT2, reflecting a neuroprotective potential following ischaemia. *Histopathology* 2007, 50, 897–910. [CrossRef] [PubMed]

103. Vermeiren, C.; Najimi, M.; Vanhoutte, N.; Tilleux, S.; de Hemptinne, I.; Maloteaux, J.-M.; Hermans, E. Acute up-regulation of glutamate uptake mediated by mGluR5a in reactive astrocytes. Regulation by protein kinase C and cell activation. *Front. Cell. Neurosci.* 2013, 7, 251. [CrossRef] [PubMed]

104. Gosselin, R.-D.; Meylan, P.; Decosterd, I. Extracellular microvesicles from astrocytes contain functional glutamate transporters: Regulation by protein kinase C and cell activation. *Brain Sci.* 2017, 7, 30.
109. Matos, M.; Augusto, E.; Santos-Rodrigues, A.D.; Schwarzschild, M.A.; Chen, J.-F.; Cunha, R.A.; Agostinho, P. Adenosine A2A receptors modulate glutamate uptake in cultured astrocytes and gliosomes. Glia 2012, 60, 702–716. [CrossRef] [PubMed]

110. Tao, Y.-X.; Gu, J.; Stephens, R.L. Role of spinal cord glutamate transporter during normal sensory transmission and pathological pain states. Mol. Pain 2005, 1, 30. [CrossRef] [PubMed]

111. Abbott, N.J.; Rönnbäck, L.; Hansson, E. Astrocyte–endothelial interactions at the blood–brain barrier. Nat. Rev. Neurosci. 2006, 7, 41–53. [CrossRef]

112. da Fonseca, A.C.C.; Matias, D.; Garcia, C.; Amaral, R.; Geraldo, L.H.; Freitas, C.; Lima, F.R.S. The impact of microglial activation on blood-brain barrier in brain diseases. Front. Cell. Neurosci. 2014, 8, 362. [CrossRef] [PubMed]

113. DosSantos, M.F.; Holanda-Afonso, R.C.; Lima, R.L.; DaSilva, A.F.; Moura-Neto, V. The role of the blood-brain barrier in the development and treatment of migraine and other pain disorders. Front. Cell. Neurosci. 2014, 8, 302. [CrossRef] [PubMed]

114. Harper, A.M.; MacKenzie, E.T.; McCulloch, J.; Pickard, J.D. Migraine and the blood-brain barrier. Lancet 1977, 1, 1034–1036. [CrossRef]

115. Teepker, M.; Munk, K.; Mylius, V.; Haag, A.; Möller, J.C.; Oertel, W.H.; Schepelmann, K. Serum concentrations of s100b andNSE in migraine. Headache 2009, 49, 245–252. [CrossRef] [PubMed]

116. Gao, X.; Qian, J.; Zheng, S.; Changyi, Y.; Zhang, J.; Ju, S.; Zhu, J.; Li, C. Overcoming the blood-brain barrier for delivering drugs into the brain by using adenosine receptor nanoagonist. ACS Nano 2014, 8, 3678–3689. [CrossRef] [PubMed]

117. Gao, H.-M.; Li, L.; Zhang, K.-L.; Chen, X.-H.; Tian, S.-Q.; Zhang, Z.-L. Impact of migraine attacks on the blood-brain barrier. Mol. Pain 2005, 1, 30. [CrossRef] [PubMed]

118. Gupta, V.K. CSD, BBB and MMP-9 elevations: animal experiments versus clinical phenomena in migraine. J. Clin. Investig. 2000, 113, 1447–1455. [CrossRef]

119. Gursoy-Ozdemir, Y.; Qiu, J.; Matsuoka, N.; Bolay, H.; Bermpohl, D.; Jin, H.; Wang, X.; Rosenberg, G.A.; Lo, E.H.; Moskowitz, M.A. Cortical spreading depression activates and upregulates MMP-9. Mol. Pain 2014, 10, 2559–2561. [PubMed]

120. Kruit, M.C.; van Buchem, M.A.; Launer, L.J.; Terwindt, G.M.; Ferrari, M.D. Migraine is associated with an increased risk of deep white matter lesions, subclinical posterior circulation infarcts and brain iron accumulation: the population-based MRI CAMERA study. Cephalalgia Int. J. Headache 2010, 30, 129–136. [CrossRef] [PubMed]

121. Kruit, M.C.; van Buchem, M.A.; Launer, L.J.; Terwindt, G.M.; Ferrari, M.D. MRI findings in migraine. Rev. Neurol. (Paris) 2005, 161, 661–665. [CrossRef]

122. Carman, A.J.; Mills, J.H.; Krenz, A.; Kim, D.-G.; Bynoe, M.S. Adenosine Receptor Signaling Modulates Permeability of the Blood–Brain Barrier. J. Neurosci. 2011, 31, 13272–13280. [CrossRef] [PubMed]

123. Kim, D.-G.; Bynoe, M.S. A2A Adenosine Receptor Regulates the Human Blood–Brain Barrier Permeability. Mol. Neurobiol. 2015, 52, 664–678. [CrossRef] [PubMed]

124. Gao, X.; Qian, J.; Zheng, S.; Changyi, Y.; Zhang, J.; Ju, S.; Zhu, J.; Li, C. Overcoming the blood-brain barrier for delivering drugs into the brain by using adenosine receptor nanoagonist. ACS Nano 2014, 8, 3678–3689. [CrossRef] [PubMed]

125. Beggs, S.; Liu, X.J.; Kwan, C.; Salter, M.W. Peripheral nerve injury and TRPV1-expressing primary afferent C-fibers cause opening of the blood-brain barrier. Mol. Pain 2010, 6, 74. [CrossRef] [PubMed]

126. Hohoff, C.; Marziniak, M.; Lesch, K.-P.; Deckert, J.; Sommer, C.; Mössner, R. An Adenosine A2A Receptor Gene Haplotype is Associated with Migraine With Aura. Cephalalgia 2007, 27, 177–181. [CrossRef] [PubMed]

127. Goadsby, P.J.; Hoskin, K.L.; Storer, R.J.; Edvinsson, L.; Connor, H.E. Adenosine A1 receptor agonists inhibit trigeminovascular nociceptive transmission. Brain 2002, 125, 1392–1401. [CrossRef] [PubMed]

128. Sebastião, A.M.; Macedo, M.P.; Ribeiro, J.A. Tonic activation of A2A adenosine receptors unmasks, and of A1 receptors prevents, a facilitatory action of calcitonin gene-related peptide in the rat hippocampus. Br. J. Pharmacol. 2000, 129, 374–380. [CrossRef] [PubMed]

129. Lu, W.; Li, B.; Chen, J.; Su, Y.; Dong, X.; Su, X.; Gao, L. Expression of calcitonin gene-related peptide, adenosine A2a receptor and adenosine A1 receptor in experiment rat migraine models. Biomed. Rep. 2016, 4, 379–383. [CrossRef] [PubMed]
130. Lindquist, B.E.; Shuttleworth, C.W. Spreading depolarization-induced adenosine accumulation reflects metabolic status in vitro and in vivo. *J. Cereb. Blood Flow Metab.* 2014, 34, 1779–1790. [CrossRef] [PubMed]

131. Noseda, R.; Kainz, V.; Jakubowski, M.; Gooley, J.J.; Saper, C.B.; Dígre, K.; Burstein, R. A neural mechanism for exacerbation of headache by light. *Nat. Neurosci.* 2010, 13, 239–245. [CrossRef] [PubMed]

132. Sodhi, P.; Hartwick, A.T.E. Adenosine modulates light responses of rat retinal ganglion cell photoreceptors through cAMP-mediated pathway. *J. Physiol.* 2014, 592, 4201–4220. [CrossRef] [PubMed]

133. Vlajkovic, S.M.; Housley, G.D.; Thorne, P.R. Adenosine and the Auditory System. *Curr. Neuropsychopharmacol.* 2009, 7, 246–256. [CrossRef] [PubMed]

134. Wei, C.J.; Li, W.; Chen, J.-F. Normal and abnormal functions of adenosine receptors in the central nervous system revealed by genetic knockout studies. *Biochim. Biophys. Acta* 2013, 1834, 1358–1379. [CrossRef] [PubMed]

135. Demarquay, G. A causative role of vasodilation in migraine? No. *Rev. Neurol. (Paris)* 2014, 170, 490–494. [CrossRef] [PubMed]

136. Kruuse, C.; Thomsen, L.L.; Jacobsen, T.B.; Olesen, J. The Phosphodiesterase 5 Inhibitor Sildenafil Has No Effect on Cerebral Blood Flow or Blood Velocity, but Nevertheless Induces Headache in Healthy Subjects. *J. Cereb. Blood Flow Metab.* 2002, 22, 1124–1131. [CrossRef] [PubMed]

137. Theis, J.G.W.; Deichsel, G.; Marshall, S. Rapid development of tolerance to dipyridamole-associated headaches. *Br. J. Clin. Pharmacol.* 1999, 48, 750–755. [CrossRef] [PubMed]

138. Guo, S.; Olesen, J.; Ashina, M. Phosphodiesterase 3 inhibitor cilostazol induces migraine-like attacks via cyclic AMP increase. *Brain Sci.* 2014, 137, 2951–2959. [CrossRef] [PubMed]

139. Hamel, E. Serotonin and migraine: Biology and clinical implications. *Cephalalgia Int. J. Headache* 2007, 27, 1293–1300. [CrossRef] [PubMed]

140. Guieu, R.; Devaux, C.; Henry, H.; Bechis, G.; Mallet, D.; Sampieri, F.; Juin, M.; Gola, R.; Rochat, H. Adenosine and migraine. *Can. J. Neurol. Sci. J. Can. Sci. Neurol.* 1998, 25, 55–58. [CrossRef] [PubMed]

141. Birk, S.; Kruuse, C.; Petersen, K.A.; Tfelt-Hansen, P.; Olesen, J. The headache-inducing effect of cilostazol in human volunteers. *Cephalalgia Int. J. Headache* 2006, 26, 1304–1309. [CrossRef] [PubMed]

142. Guo, S.; Olesen, J.; Ashina, M. Phosphodiesterase 3 inhibitor cilostazol induces migraine-like attacks via cyclic AMP increase. *Brain J. Neurol.* 2014, 137, 2951–2959. [CrossRef] [PubMed]

143. Diamond, S.M.; Henrich, W.L. Acetate dialysate versus bicarbonate dialysate: A continuing controversy. *Am. J. Kidney Dis. Off. J. Natl. Kidney Found.* 1987, 9, 3–11. [CrossRef] [PubMed]

144. Jiang, L.; Gulanski, B.I.; De Feyter, H.M.; Weinzimer, S.A.; Pittman, B.; Guidone, E.; Koretski, J.; Harman, S.; Petrakis, I.L.; Krystal, J.H.; et al. Increased brain uptake and oxidation of acetate in heavy drinkers. *J. Clin. Invest.* 2013, 123, 1605–1614. [CrossRef] [PubMed]

145. Jelski, W.; Grochowska-Skiba, B.; Szmitkowski, M. Alcohol dehydrogenase and the metabolism of ethanol in the brain. *Postepy Hig. Med. Do´sw. Online* 2007, 61, 226–230. [PubMed]

146. Waniek, M.; Caffarelli, A.M.; Caffarelli, A.M. Caffeine and adenosine. *J. Alzheimers Dis.* 2006, 77, 246–256. [PubMed]

147. Swift, R.; Davidson, D. Alcohol hangover: Mechanisms and mediators. *Hear. Res.* 2010, 260, 81–88. [CrossRef] [PubMed]

148. Wei, C.J.; Li, W.; Chen, J.-F. Normal and abnormal functions of adenosine receptors in the central nervous system revealed by genetic knockout studies. *Biochim. Biophys. Acta* 2011, 1808, 1358–1379. [CrossRef] [PubMed]

149. Ribeiro, J.A.; Sebastião, A.M. Caffeine and adenosine. *J. Alzheimers Dis.* 2010, 20, S3–S15. [PubMed]

150. Capiotti, K.M.; Menezes, F.P.; Nazario, L.R.; Pohlmann, J.B.; de Oliveira, G.M.T.; Fazenda, L.; Bogo, M.R.; Bonan, C.D.; Da Silva, R.S. Early exposure to caffeine affects gene expression of adenosine receptors, DARPP-32 and BDNF without affecting sensibility and morphology of developing zebrafish (Danio rerio). *Neurotoxicol. Teratol.* 2011, 33, 680–685. [CrossRef] [PubMed]

151. Johansson, B.; Ahlberg, S.; van der Ploeg, I.; Brené, S.; Lindefors, N.; Persson, H.; Fredholm, B.B. Effect of long term caffeine treatment on A1 and A2a adenosine receptor binding and on mRNA levels in rat brain. *Neurotoxicol. Teratol.* 2011, 33, 4201–4220. [CrossRef] [PubMed]

152. Traversa, U.; Rosati, A.M.; Florio, C.; Vertua, R. Effects of chronic administration of adenosine antagonists on adenosine A1 and A2a receptors in mouse brain. *Vivo Athens Greece* 1994, 8, 1073–1078.
153. Conlay, L.A.; Conant, J.A.; deBros, F.; Wurtman, R. Caffeine alters plasma adenosine levels. *Nature* 1997, 389, 136. [CrossRef] [PubMed]
154. Fischera, M.; Marziniak, M.; Gralow, I.; Evers, S. The incidence and prevalence of cluster headache: a meta-analysis of population-based studies. *Cephalalgia Int. J. Headache* 2008, 28, 614–618. [CrossRef] [PubMed]
155. Martelletti, P. Cluster headache management and beyond. *Expert Opin. Pharmacother.* 2015, 16, 1411–1415. [CrossRef] [PubMed]
156. Petersen, A.M.; Gleeson, T.T.; Scholnick, D.A. The effect of oxygen and adenosine on lizard thermoregulation. *Physiol. Biochem. Zool.* 2003, 76, 339–347. [CrossRef] [PubMed]
157. Rubio, R.; Berne, R.M.; Bockman, E.L.; Curnish, R.R. Relationship between adenosine concentration and oxygen supply in rat brain. *Am. J. Physiol.* 1975, 228, 1896–1902. [PubMed]
158. D’Andrea, G.; Granella, F.; Cadaldini, M. Platelet aggregation profiles in cluster headache. *Headache* 2003, 43, 272–275. [CrossRef] [PubMed]
159. Paul, S.; Feoktistov, I.; Hollister, A.S.; Robertson, D.; Biaggioni, I. Adenosine inhibits the rise in intracellular calcium and platelet aggregation produced by thrombin: Evidence that both effects are coupled to adenylate cyclase. *Mol. Pharmacol.* 1990, 37, 870–875. [PubMed]
160. Holle, D.; Naegel, S.; Obermann, M. Hypnic headache. *Cephalalgia Int. J. Headache* 2013, 33, 1349–1357. [CrossRef] [PubMed]
161. Singh, N.N.; Sahota, P. Sleep-related headache and its management. *Curr. Treat. Options Neurol.* 2013, 15, 704–722. [CrossRef] [PubMed]
162. Antle, M.C.; Steen, N.M.; Mistlberger, R.E. Adenosine and caffeine modulate circadian rhythms in the Syrian hamster. *Neuroreport* 2001, 12, 2901–2905. [CrossRef] [PubMed]
163. Schwartz, J.R.; Roth, T. Neurophysiology of Sleep and Wakefulness: Basic Science and Clinical Implications. *Curr. Neuropharmacol.* 2008, 6, 367–378. [CrossRef] [PubMed]
164. Landolt, H.-P. Sleep homeostasis: A role for adenosine in humans? *Biochem. Pharmacol.* 2008, 75, 2070–2079. [CrossRef] [PubMed]
165. Mackiewicz, M.; Nikonova, E.V.; Zimmerman, J.E.; Galante, R.J.; Zhang, L.; Cater, J.R.; Geiger, J.D.; Pack, A.I. Enzymes of adenosine metabolism in the brain: Diurnal rhythm and the effect of sleep deprivation. *J. Neurochem.* 2003, 85, 348–357. [CrossRef] [PubMed]
166. Barloese, M.C.J. Neurobiology and sleep disorders in cluster headache. *J. Headache Pain* 2015, 16, 562. [CrossRef] [PubMed]
167. Liang, J.-F.; Wang, S.-J. Hypnic headache: A review of clinical features, therapeutic options and outcomes. *Cephalalgia Int. J. Headache* 2014, 34, 795–805. [CrossRef] [PubMed]
168. Langlois, J.A.; Rutland-Brown, W.; Wald, M.M. The epidemiology and impact of traumatic brain injury: a brief overview. *J. Head Trauma Rehabil.* 2006, 21, 375–378. [CrossRef] [PubMed]
169. Bell, M.J.; Robertson, C.S.; Kochanek, P.M.; Goodman, J.C.; Gopinath, S.P.; Carrillo, J.A.; Clark, R.S.; Marion, D.W.; Mi, Z.; Jackson, E.K. Intersitial brain adenosine and xanthine increase during jugular venous oxygen desaturations in humans after traumatic brain injury. *Crit. Care Med.* 2001, 29, 399–404. [CrossRef] [PubMed]
170. Silberstein, S.D. Sex hormones and headache. *Rev. Neurol. (Paris)* 2000, 156, 4530–4541. [CrossRef]
171. Rose’ Meyer, R.B.; Mellick, A.S.; Garnham, B.G.; Harrison, G.J.; Massa, H.M.; Griffiths, L.R. The measurement of adenosine and estrogen receptor expression in rat brains following ovariectomy using quantitative PCR analysis. *Brain Res. Brain Res. Protoc.* 2003, 11, 9–18. [CrossRef]
172. Edlow, A.G.; Bartz, D. Hormonal Contraceptive Options for Women With Headache: A Review of the Evidence. *Rev. Obstet. Gynecol.* 2010, 3, 55–65. [PubMed]
173. Phillips, W.; Michell, A.; Pruess, H.; Barker, R.A. Animal models of neurodegenerative diseases. *Methods Mol. Biol.* 2009, 549, 137–155. [CrossRef]
174. PEÜE, S.; ERKUTLU, I.; KURTUL, N.; ALPETEKIN, M.; TAN, U. Effects of progesterone on total brain tissue adenosine deaminase activity in experimental epilepsy. *Int. J. Neurosci.* 2009, 119, 204–213. [CrossRef] [PubMed]
175. ASHINA, M.; SIMONSEN, H.; BENDTSSEN, L.; JENSEN, R.; OLESSEN, J. Glyceryl trinitrate may trigger endogenous nitric oxide production in patients with chronic tension-type headache. *Cephalalgia Int. J. Headache* 2004, 24, 967–972. [CrossRef] [PubMed]
176. Hsi, D.H.; Roshandel, A.; Singh, N.; Szombathy, T.; Meszaros, Z.S. Headache response to glyceryl trinitrate in patients with and without obstructive coronary artery disease. *Heart* 2005, 91, 1164–1166. [CrossRef] [PubMed]

177. Anfossi, G.; Massucco, P.; Piretto, V.; Mularoni, E.; Cavalot, F.; Mattioli, L.; Trovati, M. Glyceryl trinitrate enhances the adenosine-induced inhibition of platelet responses: A mechanism potentially involved in the in vivo anti-aggregating effects of organic nitrates. *Clin. Exp. Pharmacol. Physiol.* 1995, 22, 803–811. [CrossRef] [PubMed]

178. Gori, T.; Daiber, A.; Di Stolfo, G.; Sicuro, S.; Dragoni, S.; Lisi, M.; M’nzel, T.; Forconi, S.; Parker, J.D. Nitroglycerine causes mitochondrial reactive oxygen species production: In vitro mechanistic insights. *Can. J. Cardiol.* 2007, 23, 990–992. [CrossRef] [PubMed]

179. Montagna, P.; Sacquegna, T.; Martinelli, P.; Cortelli, P.; Bresolin, N.; Moggio, M.; Baldrati, A.; Riva, R.; Lugaresi, E. Mitochondrial abnormalities in migraine. Preliminary findings. *Headache* 1988, 28, 477–480. [CrossRef] [PubMed]

180. Garcia-Bou, R.; Rocha, M.; Apostolova, N.; Herance, R.; Hernandez-Mijares, A.; Victor, V.M. Evidence for a relationship between mitochondrial Complex I activity and mitochondrial aldehyde dehydrogenase during nitroglycerin tolerance: Effects of mitochondrial antioxidants. *Biochim. Biophys. Acta* 2012, 1817, 828–837. [CrossRef] [PubMed]

181. Duley, J.A.; Garrick, D.P.; Pratt, D.A. Raised Plasma Adenosine Associated with Chronic Fatigue Syndrome. Available online: http://informahealthcare.com/doi/abs/10.1300/J092v07n03_07 (accessed on 2 August 2013).

182. Koos, B.J. Adenosine A2a receptors and O2 sensing in development. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 2011, 301, R601–R622. [CrossRef] [PubMed]

183. Watanabe, F.; Kamiike, W.; Nishimura, T.; Hashimoto, T.; Tagawa, K. Decrease in mitochondrial levels of adenine nucleotides and concomitant mitochondrial dysfunction in ischemic rat liver. *J. Biochem. (Tokyo)* 1983, 94, 493–499. [CrossRef]

184. Nassini, R.; Materazzi, S.; Vriens, J.; Prenen, J.; Benemei, S.; Siena, G.D.; la Marca, G.; Andrè, E.; Preti, D.; Avonto, C.; et al. The “headache tree” via umbellulone and TRPA1 activates the trigeminovascular system. *Brain* 2012, 135, 376–390. [CrossRef] [PubMed]

185. Benemei, S.; Cesaris, F.D.; Fusi, C.; Rossi, E.; Lupi, C.; Geppetti, P. TRPA1 and other TRP channels in migraine. *J. Headache Pain* 2013, 14, 71. [CrossRef] [PubMed]

186. Wicher, D.; Agricola, H.-J.; Schönherr, R.; Heinemann, S.H.; Derst, C. TRPgamma channels are inhibited by cAMP and contribute to pacemaking in neurosecretory insect neurons. *J. Biol. Chem.* 2006, 281, 3227–3236. [CrossRef] [PubMed]

187. Nagatomo, K.; Kubo, Y. Caffeine activates mouse TRPA1 channels but suppresses human TRPA1 channels. *Proc. Natl. Acad. Sci. USA* 2008, 105, 17373–17378. [CrossRef] [PubMed]

188. Egbuniwe, O.; Grover, S.; Duggal, A.K.; Mavroudias, A.; Yazdi, M.; Renton, T.; Di Silvio, L.; Grant, A.D. TRPA1 and TRPV4 activation in human odontoblasts stimulates ATP release. *J. Dent. Res.* 2014, 93, 911–917. [CrossRef] [PubMed]

189. Mochizuki, T.; Sokabe, T.; Araki, I.; Fujishita, K.; Shibasaki, K.; Uchida, K.; Naruse, K.; Takeda, M.; Tominaga, M. The TRPV4 Cation Channel Mediates Stretch-evoked Ca2+ Influx and ATP Release in Primary Urothelial Cell Cultures. *J. Biol. Chem.* 2009, 284, 21257–21264. [CrossRef] [PubMed]

190. Tominaga, M.; Wada, M.; Masu, M. Potentiation of capsaicin receptor activity by metabotropic ATP receptors as a possible mechanism for ATP-evoked pain and hyperalgesia. *Proc. Natl. Acad. Sci. USA* 2001, 98, 6951–6956. [CrossRef] [PubMed]
193. Deckert, J.; Nöthen, M.M.; Franke, P.; Delmo, C.; Fritze, J.; Knapp, M.; Maier, W.; Beckmann, H.; Propping, P. Systematic mutation screening and association study of the A1 and A2A adenosine receptor genes in panic disorder suggest a contribution of the A2A gene to the development of disease. *Mol. Psychiatry* **1998**, *3*, 81–85. [CrossRef] [PubMed]

194. Hamilton, S.P.; Slager, S.L.; De Leon, A.B.; Heiman, G.A.; Klein, D.F.; Hodge, S.E.; Weissman, M.M.; Fyer, A.J.; Knowles, J.A. Evidence for genetic linkage between a polymorphism in the adenosine 2A receptor and panic disorder. *Neuropsychopharmacology* **2004**, *29*, 558–565. [CrossRef] [PubMed]

195. Giménez-Llort, L.; Fernández-Teruel, A.; Escorihuela, R.M.; Fredholm, B.B.; Tobeña, A.; Pekny, M.; Johansson, B. Mice lacking the adenosine A1 receptor are anxious and aggressive, but are normal learners with reduced muscle strength and survival rate. *Eur. J. Neurosci.* **2002**, *16*, 547–550. [CrossRef] [PubMed]

196. Coelho, J.E.; Alves, P.; Canas, P.M.; Valadas, J.S.; Shmidt, T.; Batalha, V.L.; Ferreira, D.G.; Ribeiro, J.A.; Bader, M.; Cunha, R.A.; et al. Overexpression of Adenosine A2A Receptors in Rats: Effects on Depression, Locomotion, and Anxiety. *Front. Psychiatry* **2014**, *5*, 67. [CrossRef] [PubMed]