Genetic Analysis of Collagen Q: Roles in Acetylcholinesterase and Butyrylcholinesterase Assembly and in Synaptic Structure and Function

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Abstract. Acetylcholinesterase (AChE) occurs in both asymmetric forms, covalently associated with a collagenous subunit called Q (ColQ), and globular forms that may be either soluble or membrane associated. At the skeletal neuromuscular junction, asymmetric AChE is anchored to the basal lamina of the synaptic cleft, where it hydrolyzes acetylcholine to terminate synaptic transmission. AChE has also been hypothesized to play developmental roles in the nervous system, and ColQ is also expressed in some AChE-poor tissues. To seek roles of ColQ and AChE at synapses and elsewhere, we generated ColQ-deficient mutant mice. ColQ<sup>−/−</sup> mice completely lacked asymmetric AChE in skeletal and cardiac muscles and brain; they also lacked asymmetric forms of the AChE homologue, butyrylcholinesterase. Thus, products of the ColQ gene are required for assembly of all detectable asymmetric AChE and butyrylcholinesterase. Surprisingly, globular AChE tetramers were also absent from neonatal ColQ<sup>−/−</sup> muscles, suggesting a role for the ColQ gene in assembly or stabilization of AChE forms that do not themselves contain a collagenous subunit. Histochemical, immunohistochemical, toxicological, and electrophysiological assays all indicated absence of AChE at ColQ<sup>−/−</sup> neuromuscular junctions. Nonetheless, neuromuscular function was initially robust, demonstrating that AChE and ColQ do not play obligatory roles in early phases of synaptogenesis. Moreover, because acute inhibition of synaptic AChE is fatal to normal animals, there must be compensatory mechanisms in the mutant that allow the synapse to function in the chronic absence of AChE. One structural mechanism appears to be a partial ensheathment of nerve terminals by Schwann cells. Compensation was incomplete, however, as animals lacking ColQ and synaptic AChE failed to thrive and most died before they reached maturity.

Key words: acetylcholine • acetylcholinesterase • butyrylcholinesterase • collagen • neuromuscular junction

Acetylcholinesterase (AChE)<sup>1</sup> is concentrated in the synaptic cleft of the skeletal neuromuscular junction. A cetylcholine released from motor nerve terminals is rapidly inactivated by the synaptic AChE, thereby limiting the action of the neurotransmitter. The importance of the enzyme is demonstrated by the effects of blocking its activity. Inhibition of AChE, for example by nerve gas, causes prolonged activation of acetylcholine receptors (AChRs), leading to desensitization of the receptors, respiratory paralysis, and death. Partial inhibition of AChE, for example by overexposure to insecticides, results in excessive influx of calcium through the AChR’s ion channel, which leads to local necrotic myopathy (Leonard and Salpeter, 1979; Wecker et al., 1986). On the other hand, inhibitors of AChE are therapeutically useful to patients with diseases of inadequate neurotransmission in which levels of AChRs are decreased (such as acquired autoimmune myasthenia gravis; Engel, 1994a) or insufficient acetylcholine is released (such as familial infantile myasthenia; Engel, 1994b).

Athough the function of AChE is apparently simple, its patterns of structure and distribution are remarkably com-

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<sup>1</sup>Abbreviations used in this paper: AChE, acetylcholinesterase; AChR, acetylcholine receptor; BuChE, butyrylcholinesterase; ColQ, collagen Q; DS, detergent-soluble; ES cells, embryonic stem cells; HS, high-salt; LS, low-salt; mepps, miniature endplate potentials; PRAD, proline-rich attachment domain; VVA-B4, Vicia villosa agglutinin-B4.
plex. First, the transcript of the AChE gene is subject to alternative splicing and its protein products can associate covalently with each other and with noncatalytic subunits (for review see Assoulié et al., 1993, 1998). Second, the oligomers can occur in cytoplasmic, membrane-bound and extracellular matrix-associated forms (for review see Assoulié and Bon, 1982; Toutsant and Assoulié, 1988). Third, AChE is found not only at neuromuscular junctions, but also at cholinergergic interneuronal synapses, in some noncholinergergic neurons, and in some nonneural cells such as erythrocytes (Assoulié et al., 1993). Fourth, a related enzyme, butyrylcholinesterase (BuChE), capable of hydrolyzing acetylcholine, is expressed in some of the same sites as AChE , including the neuromuscular junction (for review see Chatonnet and Lockridge, 1989). Based in part on their complex patterns of expression in embryos and adults, AChE and BuChE have been hypothesized to play a variety of noncholinergergic and even noncatalytic roles in embryos and adults (for review see Layer and Willbold, 1995).

Much of the AChE at the neuromuscular junction occurs in forms that have been called asymmetric. A symmetric forms of AChE were initially identified in Torpedo and Electrophorus electric organ; they were defined and distinguished from the remaining (globular) forms by virtue of their anomalous sedimentation properties in sucrose density gradients. The asymmetry was shown to reflect the association of catalytic AChE subunits with a tail that was rod-shaped, collagenase-sensitive, and rich in amino acids characteristic of collagens (hydroxyproline and hydroxylysine; Lwebuga-Mukasa et al., 1976; Rosenberry and Richardson, 1977). Three asymmetric forms of AChE were characterized, in which one, two, or three tetramers of catalytic subunits (now called AChE (α) ) were disulfide-bonded to a single collagenous triple helical tail. These forms are called A α, A β, and A γ, to indicate the total number of catalytic subunits per oligomer (Assoulié and Bon, 1982).

Hall (1973) showed that asymmetric forms of AChE were selectively associated with synapse-containing regions of rodent skeletal muscle, whereas globular forms were more uniformly distributed. Later, it was found that synaptic AChE was stably associated with the basal lamina that runs between the motor nerve terminal and the postsynaptic membrane at the neuromuscular junction (M-C Mahan et al., 1978; Sanes and Hall, 1979). The collagen tail of asymmetric AChE is likely to be critical for anchoring the enzyme to the basal lamina, perhaps by association with proteoglycans (Bon et al., 1978; Vigny et al., 1983; Brandan et al., 1985; Deprez and Inestrosa, 1995; Rossi and Rotundo, 1996; Rotundo et al., 1997). However, the relationship of asymmetric AChE to the synapse is not completely understood in that some synaptic AChE may be globular (Anglister et al., 1994) and some asymmetric AChE is found extrasympathetically (Carson et al., 1979; Younkin et al., 1982; Sketelj and Brzin, 1985).

Structural and functional analyses of the asymmetric and synaptic forms of AChE were delayed by the difficulty of isolating its collagenous subunit. Recently, however, cDNA s encoding an AChE-associated collagenous subunit called Q (ColQ) were molecularly cloned from rat muscle (Krejci et al., 1997), based on homology to a previously isolated Q subunit from Torpedo (Krejci et al., 1991). A nitrotydes to recombinant ColQ recognize native asymmetric AChE and BuChE, and coexpression of cDNA s encoding ColQ and AChE (α) generates asymmetric AChE in heterologous cells (Krejci et al., 1997). A assembly of asymmetric AChE involves interaction of a proline-rich attachment domain (PRA D) in ColQ with a transphosphampphilic tetramerization domain in AChE (α) (Bon et al., 1997; Krejci et al., 1997; Simon et al., 1998). A alternatively spliced products of the ColQ gene that encode a PRA D but no collagenous domain have been described; they may organize other AChE isoforms or interact with other proteins. In fact, ColQ RNA is expressed in many tissues with little or no asymmetric AChE, suggesting that it may have additional roles (Krejci et al., 1997).

Here, we have used homologous recombination in embryonic stem (ES) cells to inactivate the ColQ gene in mice, thereby allowing us to assess roles of the ColQ protein in vivo. Using a combination of biochemical, histological, and electrophysiological methods, we answered the following questions. Does asymmetric AChE in muscle require ColQ protein for assembly or accumulation? Are only asymmetric forms of AChE ColQ-dependent? How much of the AChE concentrated at the neuromuscular junction is ColQ-dependent? Does asymmetric BuChE require the ColQ gene product? How does loss of ColQ affect the structure and function of the neuromuscular junction? Is AChE or ColQ required for proper neuromuscular development? Is ColQ required for the structure or function of nonmuscle tissues?

**Materials and Methods**

**Generation of Mutant Mice**

Genomic clones containing the ColQ gene were isolated by screening a 129sv strain mouse genomic library (Stratagene) with cDNA s encoding rat ColQ (Krejci et al., 1997). For targeting vector PRAD1 (Fig. 1 a), a 0.6-kb EcoR I-HindII fragment containing the exon encoding the PRA D domain as well as adjacent intronic sequences was replaced by a PGK -neo cassette (Tybulewicz et al., 1991). For targeting vector PRAD2 (Fig. 1 a), a 0.6-kb HindII fragment containing the PRA D exon was replaced by a cassette containing neo plus the Escherichia coli lacZ gene, for monitoring gene expression. The PRA D2 vector also contained a diphtheria toxin-A gene for negative selection (Yagi et al., 1990). Both constructs were transfected into R1 ES cells (Nagy et al., 1993) by electroporation. Homogenic recombinants were identified by PCR and confirmed by probing genomic Southern blots with an 0.8-kb fragment of the ColQ gene that was entirely external to the targeting vector (Fig. 1 a). Chimeric mice from one PRA D1 and two PRA D2 ES cell clones gave rise to heterozygous and then homozygous mutant mice. The PRA D2 vector contained a lacZ gene, but no β-galactosidase activity was detected in heterozygotes, either histochemically or immunohistochemically.

**Histology**

For immunohistochemistry, sternomastoid and tibialis anterior muscles were embedded in Tissue-Tek OCT compound (Sakura Finetek USA), frozen in liquid nitrogen-cooled isopentane, and sectioned at 10 μm in a cryostat. Sections were blocked for 1 h with 2% BSA and 5% normal goat serum in PBS, incubated with primary antibodies for 1–2 h, washed, and then reincubated with secondary antibodies for another 1–2 h. Antibodies to the following antigens were used: ColQ (Krejci et al., 1997), AChE (a gift of Tereone Rosenberg, May Clinic), α-sarcoglycan, β-dystroglycan, utrophin (Novocastra Laboratories Ltd.), rapsyn (Phillips et al., 1991), and agrin (a gift of Z. Hall, University of California, San Francisco). FITC-conjugated Vicia villosa agglutinin-B4 (VVA-B4) was obtained from Sigma Chemical Co. and rhodamine-α-bungarotoxin was obtained from Molecular Probes.
Cholinesterase activity was detected by the histochemical method of Karnovsky and Roots (1964) on sections fixed with 2% paraformaldehyde in PBS. To distinguish AChE from BuChE, the reaction mixture was supplemented with 10^{-4} M tetraisopropylpyrophosphoramide (iso-OMPA), a selective inhibitor of BuChE, or with 5 \times 10^{-5} M 1,5-bis (4-allyldimethylammoniumphenyl)-pentan-3-one dibromide (BW284c51), a selective inhibitor of AChE (Austin and Berry, 1953). Where indicated, sections were treated with 1% Triton X-100 for 60 min at room temperature or highly purified bacterial collagenase (800 U/ml, type VII; Sigma Chemical Co.) for 60 min at 37°C before fixation and staining.

For electron microscopy, mice were perfused with lactated Ringer’s solution followed by 4% paraformaldehyde and 4% glutaraldehyde in 100 mM cacodylate buffer, pH 7.2. Sternomastoid muscles were dissected and fixed overnight at 4°C. The endplate-rich region of the muscle was refixed in 1% OsO₄ in cacodylate buffer, dehydrated, and embedded in Araldite. Thin sections were stained with uranyl acetate and lead citrate.

### Biochemistry

Mice were killed and tissues were homogenized in a glass Potter homogenizer in 1 ml of low-salt (LS) buffer, which contained 50 mM Tris, pH 7.0, 40 mM MgCl₂, 1% Triton X-100, 2 mM benzamidine, 40 μg/ml leupeptin, and 25 μg/ml pepstatin. The homogenates were centrifuged for 20 min at 60,000 rpm, the supernatant was removed, and the pellet was homogenized in a second aliquot of LS buffer and centrifuged again. The two resulting supernatants comprised the detergent-soluble (DS) extracts. The pellet was then homogenized with 300 μl of high-salt (HS) buffer, which consisted of LS buffer plus 0.8 M NaCl. A fiber centrifugation, the supernatant (HS extract) and the pellet were both saved.

Samples of the DS and HS extracts were analyzed by sedimentation in 5–20% sucrose gradients containing 50 mM Tris, pH 7.0, 40 mM MgCl₂, 1% Triton X-100, and 0.4 M NaCl. Centrifugation was in an SW41 rotor for 16 h at 39,000 rpm. A bout 75 fractions of 130 μl were collected from the bottom of the tube. 60-μl aliquots were transferred to a microtiter plate. Cholinesterase activity was assayed colorimetrically, using acetylcholine as substrate (Ellman et al., 1961). To assay AChE, iso-OMPA (10^{-2} M) was included to inhibit BuChE. To assay BuChE, BW284c51 (10^{-6} M) was included to inhibit AChE. Alkaline phosphatase (6.1 S) and β-galactosidase (16 S) from E. coli were included as internal sedimentation standards, and their activity profiles were used to establish a linear relationship between the fraction numbers and Svedberg units (Simon et al., 1998). All reagents were from Sigma Chemical Co.

### Electrophysiology

ColQ^{+/−} mice or control littermates were killed and immediately exsanguinated. Hemi-diaphragms with their associated phrenic nerves were isolated. The two hemi-diaphragms were separated and each was mounted in Figure 1. Generation of a ColQ mutant. (a) The targeted alleles. The top line shows schematic of the ColQ protein, including the aminoterminal PRAD domain that links ColQ to the catalytic subunit of AChE. The second line shows gene structure, with exons indicated by dark boxes and introns by lines. The third and fourth lines show the PRAD1 and PRAD2 vectors, in which the exon encoding the PRAD domain was deleted. The fifth and sixth lines, labeled recombinant, show predicted structures of the mutant alleles. Location of probe used for Southern blots is indicated. (b) Southern blots of wild-type ES cells and homologous recombinant PRAD1 and PRAD2 clones. In the recombinants, the 3.2-kb EcoRI fragment from the wild-type gene is shifted to 4.6 kb. (c) PCR assay used to genotype ColQ mutants. (d) Weight of ColQ^{−/−} mice (bottom line) and littermates (top line). Mutant homozygotes are smaller than littermates by P5, and weigh less than half as much as littermates by P20. Bars show mean ± SEM, n = 4–14.
Results

Generation of a ColQ Mutant

Clones encoding the ColQ gene were isolated from a genomic library and used to produce two targeting vectors, PRA D1 and PRA D2 (Fig. 1 a). Both vectors deleted the PRA D domain of ColQ, which forms the binding site for the catalytic (A ChE T) subunits of A ChE (B on et al., 1997). Both targeting vectors gave rise to homologous recombinant ES cells, germline chimeras, heterozygotes, and homozygotes (Fig. 1, b and c). Phenotypes of PRA D1 and PRA D2 homozygotes were identical, so both are called PRAD homozygotes (Fig. 1, b and c). Phenotypes of PRAD1 and PRAD2 heterozygotes and were externally indistinguishable from heterozygotes, and distribution (see below) but, as expected, bore no ColQ (Fig. 2, b and b’). No ColQ transcripts were detectable in PRAD1 or PRAD2 mice.

ColQ Mutants Lack Synaptic A ChE

Sections of skeletal muscle from ColQ /− mice and littermates were doubly stained with an antiserum to recombinant ColQ plus rhodamine-α-bungarotoxin, which binds to A Ch Rs and thereby marks synaptic sites. In controls, ColQ was concentrated at synaptic sites and was undetectable extrasynaptically (Fig. 2, a and a’). In ColQ /− mutants, A ChR-rich synaptic sites were nearly normal in size, shape, and distribution (see below) but, as expected, bore no ColQ (Fig. 2, b and b’).

The ColQ subunit is believed to anchor A ChE to the basal lamina at the neuromuscular junction. To obtain definitive evidence on this point, we stained sections of mutant and control muscle with antibodies specific for the A ChE catalytic subunit. This antibody recognizes both of the known alternatively spliced products (T and H) of the mammalian A ChE gene (D uval et al., 1992). A ChE was abundant at all synaptic sites in controls, but undetectable in homozygotes (Fig. 2, c and d).

This result suggested that ColQ is required for accumulation of all synaptic A ChE. An alternative explanation, however, was that the association of A ChE with synaptic membranes or matrix had merely been weakened in the absence of its collagenous tail, in which case A ChE might have been present at synapses in vivo but lost during preparation of sections. Therefore, we performed additional tests to seek A ChE at synaptic sites in ColQ /− muscle in situ. First, we injected fasciculin-2, a selective inhibitor of A ChE (R odriguez-Ithurralde et al., 1983) into ColQ /− mice and their littermates. This inhibitor was chosen because it penetrates the central nervous system poorly, and is therefore believed to exert its effects predominantly on peripheral A ChE. Fasciculin-2 (3–6 µg/g, injected intraperitoneally) caused tremor followed by flaccid paralysis in controls. In mutants, in contrast, effects of fasciculin were minor, and injected animals were reflexive and capable of righting. Second, we recorded mepps from muscle fibers of ColQ /− mice and littermate controls (Fig. 3). The decay times of mepps were variable but on average slower in homozygotes than in controls (data not shown), presumably because A ChE terminates neurotransmitter action at normal neuromuscular junctions (W hathey et al., 1979), whereas diffusion does so in ColQ /− muscle. More importantly, addition of fasciculin greatly slowed the decay of mepps in normal muscle, but had no significant effect on the time course of mepps in ColQ /− muscle (from 3.57 ± 0.18 to 3.47 ± 0.07 ms in four experiments; P > 0.2). Together, these toxicological and physiological tests support the idea that ColQ /− mutants lack functional synaptic A ChE.

ColQ Is Required for Assembly of All Asymmetric and Some Globular A ChE in Muscle

We used sucrose density gradient centrifugation to assess
Figure 3. Mepps recorded in muscle fibers from 40-d-old ColQ<sup>+/−</sup> (top) and ColQ<sup>−/−</sup> (bottom) mice. A edition of the AChE inhibitor fasciculin-2 (320 nM) to the bath increased the amplitude and prolonged the decay time constant of mepps in control muscle but had no significant effect on the size or shape of mepps in ColQ<sup>−/−</sup> muscle. Each trace represents the average of 50 mepps. The decay phases of averaged mepps were fitted by a single exponential from 10% to 90% of the maximal amplitude (vertical lines). The difference in mepp amplitudes between ColQ<sup>+/−</sup> and ColQ<sup>−/−</sup> muscles is due to differences in muscle fiber diameter and input resistance.

the molecular forms of AChE present in ColQ<sup>−/−</sup> muscles. Globular forms are extracted into detergent-containing buffers at low ionic strength, whereas asymmetric (collagen-tailed) forms are extracted only at high ionic strength (Bon et al., 1978). Therefore, we first homogenized muscles with detergent-containing LS buffers, then reincubated the pellet with HS buffers to selectively solubilize asymmetric forms.

Control (ColQ<sup>+/−</sup> and ColQ<sup>−/−</sup>) muscles contain three asymmetric forms, corresponding to a single helical trimer of ColQ chains attached to one, two, or three tetramers of catalytic subunits (A<sub>4</sub>, A<sub>8</sub>, and A<sub>12</sub>, respectively; Massoulié and Bon, 1982). In contrast, no asymmetric forms were detectable in ColQ<sup>−/−</sup> muscle (Fig. 4, b and c). Because some asymmetric AChE is difficult to solubilize from tissue (Rossi and Rotundo, 1964), we also assayed the pellet remaining after HS and detergent extraction. AChE activity in insoluble fractions was 14% of that in the HS extract for ColQ<sup>+/−</sup> muscles (n = 4), and 30% of that in the HS extract for ColQ<sup>−/−</sup> muscles. However, AChE activity was undetectable in the insoluble fraction from ColQ<sup>−/−</sup> muscles. Similar results were obtained with muscles from neonates (P0) and at P20. Thus, all asymmetric AChE in muscle requires ColQ for its assembly and/or accumulation.

Surprisingly, loss of AChE in ColQ<sup>−/−</sup> mice was not limited to the asymmetric forms. At birth, three major peaks of AChE activity were distinguishable in detergent extracts of control muscle, corresponding to amphiphilic monomers and dimers (G<sub>3</sub><sup>a</sup> and G<sub>2</sub><sup>a</sup>) and nonamphiphilic tetramers (G<sub>4</sub><sup>ab</sup>; Massoulié et al., 1998). G<sub>3</sub><sup>a</sup> and G<sub>2</sub><sup>a</sup> were retained in neonatal ColQ<sup>−/−</sup> muscles, but the G<sub>4</sub><sup>ab</sup> peak was almost entirely absent (Fig. 4 a). In addition, the pool of globular tetramers extracted by HS was absent from ColQ<sup>−/−</sup> neonates (Fig. 4 b). At P20, however, amphiphilic and nonamphiphilic tetramers (G<sub>4</sub><sup>a</sup> and G<sub>3</sub><sup>ab</sup>) as well as G<sub>3</sub><sup>a</sup> and G<sub>2</sub><sup>a</sup> forms were present in ColQ<sup>−/−</sup> muscles (data not shown). Thus, ColQ-independent mechanisms for assembly or retention of G<sub>4</sub> do exist (see Discussion).

ColQ Is Required for Assembly of Asymmetric BuChE

BuChE is homologous to AChE, hydrolyzes acetylcholine, and is present at the neuromuscular junction (Chatonnet and Lockridge, 1989; Chapron et al., 1997). Like AChE, BuChE catalytic monomers assemble into both globular and asymmetric multimers, and the asymmetric multimers bear a collagenase-sensitive subunit (Vigny et al., 1978; Toutant et al., 1985). To determine whether ColQ serves as the collagen tail for BuChE, we assayed succrose density gradients in the presence of a selective inhibitor of AChE, BW284c51. A symmetric and globular forms of BuChE were detectable in control muscle (Fig. 4, d and e). As reported previously, S values of BuChE differed from those of AChE (Vigny et al., 1978; Krejci et al., 1997), providing assurance that the activity seen in the presence of BW284c51 did not reflect incomplete inhibition of AChE. Moreover, activity was inhibited by the selective inhibitor of BuChE, iso-OMP A (data not shown). Globular isoforms of BuChE were retained in ColQ<sup>−/−</sup> mutants, but no asymmetric BuChE was detectable (Fig. 4, d and e). Thus, the ColQ gene appears to encode the collagenous tails of both AChE and BuChE.

Lacking antibodies to BuChE, we used the histochemical stain of Karnovsky and Rootes (1964) to localize BuChE in control and mutant muscles. This method relies on hydrolysis of the acetylcholine analogue, acetylthiocholine, which is cleaved by both AChE and BuChE. A<sub>4</sub><sup>a</sup> reaction product was readily detectable at ColQ<sup>−/−</sup> synaptic sites, although levels were substantially reduced compared with controls (Fig. 5, a and b). The AChE inhibitor BW284c51 had no effect on the staining of ColQ<sup>−/−</sup> synaptic sites, but decreased staining in controls to approximately the level seen in mutants (Fig. 5, c and d). This result provides independent evidence that AChE is depleted from ColQ<sup>−/−</sup> neuromuscular junctions. In contrast, the BuChE inhibitor iso-OMPA decreased staining of control synaptic sites only slightly, but completely abolished staining of ColQ<sup>−/−</sup> synaptic sites (Fig. 5, e and f). Thus, the synaptic acetylcholine-hydrolyzing activity in mutants is largely or entirely BuChE. Reaction product was undetectable in controls even after prolonged incubation when a mixture of BW284c51 plus iso-OMPA was added with the substrate, indicating that AChE and BuChE together account for most or all of the ACh-hydrolyzing activity at control synapses (Fig. 5, g and h).

To obtain information on the molecular forms and cellular localization of synaptic BuChE, we treated cross-sections in various ways then stained them in the presence of BW284c51. In both control and ColQ<sup>−/−</sup> muscle, reaction product was tightly associated with synaptic sites (Fig. 5, i and j). Pretreatment of sections with either collagenase, which releases matrix-associated enzyme, or with the detergent Triton X-100, which releases membrane bound or intracellular enzyme, reduced staining of synaptic sites in control muscles (Fig. 5, k and m). Combined treatment
with collagenase and Triton X-100 removed all detectable BuChE (Fig. 5, o and p). Thus, both matrix- and membrane-associated BuChE are present at control synapses. In ColQ<sup>-/-</sup> muscle, collagenase treatment had no effect on the level of BuChE (Fig. 5 l) but detergent treatment abolished all staining (Fig. 5 n), confirming the absence of matrix-associated BuChE and showing that residual activity is membrane-associated. Denervation, which causes loss of nerve terminals and rapid remodeling of terminal Schwann cells, led to complete loss of this membrane-associated BuChE from ColQ<sup>-/-</sup> synapses but had little effect on controls (Fig. 5, q and r). Together, these results suggest that both membrane- and matrix-associated forms of BuChE are normally concentrated at the neuromuscular junction. The BuChE of the synaptic cleft is predominantly matrix-associated, asymmetric, and requires ColQ

![Figure 4](image-url)

Figure 4. Molecular forms of cholinesterase in neonatal ColQ mutant muscles. DS and HS pools of cholinesterase, which correspond to globular and asymmetric forms, respectively, were separated on sucrose gradients. Fractions were incubated in the presence of the BuChE inhibitor iso-OMPA (a and b) or the AChE inhibitor BW284c51 (d and e), to selectively assay AChE or BuChE, respectively. a–e show profiles from representative gradients. In each case, ColQ<sup>-/-</sup>, ColQ<sup>+/+</sup>, and ColQ<sup>+/+</sup> littermates were assayed in parallel, and with identical incubation times. High background levels in e reflect low specific activity, which necessitated overnight incubation of samples. c shows average AChE activities from all experiments, expressed as total activity in International Units (IU) per muscle. In the absence of ColQ, muscles are devoid of asymmetric forms of AChE and BuChE. Levels of G<sub>4</sub>AChE are also dramatically reduced. At P20, asymmetric AChE and BuChE were also completely absent in ColQ<sup>-/-</sup>, but G<sub>4</sub>AChE was present.
Collagen Q Mutant Lacks Synaptic Acetylcholinesterase

Figure 5. Sections of muscle from P20 ColQ+/− and ColQ−/− littermates stained for cholinesterase activity by the Karnovsky method. a–h are longitudinal sections; i–r are cross-sections. (a and b) Levels of cholinesterase activity are much higher at control than ColQ−/− endplates. (c, d, i, and j) BW284c51 (BW), a selective inhibitor of AChE, has no effect on synaptic cholinesterase activity in ColQ−/− muscle, but decreases activity in controls to a level equivalent to that in the mutant. (e and f) Iso-OMP A, a selective inhibitor of BuChE, has only a slight effect on cholinesterase activity in controls but abolishes all activity in mutants. (g and h) No activity is detectable in either muscle in the presence of both BW284c51 and iso-OMP A. These results indicate that both AChE and BuChE are present at control endplates, but only BuChE at ColQ−/− endplates. (k and l) Incubation of sections with collagenase to release asymmetric enzyme before staining for BuChE (BW + ) reduces activity in controls but has no effect at ColQ−/− synapses. (m and n) After incubation with Triton X-100 to release membrane-bound enzyme, some BuChE persists at control synapses but is abolished at ColQ−/− synapses. (o and p) No BuChE activity is detectable in controls or mutants after sequential treatment with Triton and collagenase. (q and r) 3 d after nerve section, when nerve terminals have degenerated, BuChE persists at control synaptic sites but is lost from mutant synaptic sites. Bar, 20 μm.

Synaptic Structure Is Abnormal in ColQ Mutants

To assess the size and shape of neuromuscular junctions in ColQ−/− muscle, thick longitudinal sections were stained with rhodamine-α-bungarotoxin. In controls, the AChR-rich membrane is a roughly circular plaque at birth, then passes through a perforated-plaque stage (Steinbach, 1981) and eventually matures into a pretzel-like array of distinct AChR-rich branches by P20 (Fig. 6 a). The precise geometry of branches is unique at each synaptic site, but the general pattern is stereotyped. In mutant homozygotes, in contrast, synaptic geometry was variable. Some synaptic sites were smaller than controls but roughly normal in appearance (Fig. 6 b), some retained the immature appearance characteristic of ~1-wk-old controls (Fig. 6 c), and some appeared fragmented (Fig. 6 d). A approximately 40% of the neuromuscular junctions were normal in geometry, another 40% were fragmented, and the remaining 20% were immature.

Electron microscopy revealed two structural abnormalities at synaptic sites in ColQ−/− muscles. First, the cytoplasm beneath the postsynaptic membrane was often ridged with holes, consistent with a localized degenerative response (Fig. 7 b). Such local degeneration has been observed after acute inhibition of AChE (Laskowski et al., 1977), and is believed to result from entry of Ca2+ through excessively activated AChRs (Leonard and Salpeter, 1979). Consistent with this interpretation, signs of necrosis were seldom observed in nonsynaptic regions of ColQ−/− muscle, and no signs of muscle fiber degeneration and regeneration (such as central nuclei, which are diagnostic of immature or regenerated fibers) were detectable. Second, nerve terminals were sometimes partially enwrapped by processes of Schwann cells (Fig. 7 d). Such enwrapping has been seen in several pathological situations, and can occur after muscle damage, or as a direct consequence of abnormalities in the synaptic cleft (Duchên et al., 1974; Irrmanová, 1975; Noakes et al., 1995; Patton et al., 1998). Because Schwann cell processes are impermeable to neurotransmitter, their intrusion into the synaptic cleft might represent one adaptive mechanism by which ColQ−/− synapses limit activation of AChRs.

Interestingly, the incidence of subsynaptic necrosis and Schwann cell enwrapping of nerve terminals varied as a function of age. At P20, nearly two-thirds of synaptic sites...
encountered (80/125) showed clear signs of necrosis. By 6 mo of age, however, <5% of synaptic profiles (2/71) showed necrosis, and many were indistinguishable from controls (compare Fig. 7, a and c). This difference raises the possibility that damage occurring early is repaired and that compensatory mechanisms, such as Schwann cell enwrapping, prevent damage from recurring. Indeed, the incidence of nerve terminals partially or completely enwrapped by Schwann cell processes increased somewhat, from one-third (42/125) at P20 to over one-half (39/71) at 6 mo of age. This difference suggests either that Schwann cell enwrapping is progressive, or that individuals with more complete enwrapping are more likely to survive. Necrosis and Schwann cell wrapping were seen in only 1 and 4 of 83 control endplates, respectively.

Finally, we stained sections of muscle from ColQ−/− mice and littermate controls with a panel of antibodies to proteins present at synaptic sites. These included the transmembrane proteins α-sarcoglycan and β-dystroglycan, which are present throughout the muscle fiber but concentrated in the postsynaptic membrane; rapsyn and utrophin, which are selectively associated with the subsynaptic cytoplasm; and agrin, which is concentrated in synaptic basal lamina (Sanes and Lichtman, 1999). The distribu-
tion of all of these markers was qualitatively similar in control and homozygous mutant mice (Fig. 8, a–j).

We also stained sections with the lectin VVA-B4. VVA-B4 binds to glycoconjugates that terminate with a \(\beta\)-N-acetylgalactosaminy1 residue and selectively stains the synaptic cleft at the neuromuscular junction (Scott et al., 1988). We showed previously that VVA-B4 binds to asymmetric but not globular forms of AChE in muscle, but presented indirect evidence that other VVA-B4-binding proteins were also present in the synaptic cleft (Scott et al., 1988; Martin and Sanes, 1995). VVA-B4 stained synaptic sites nearly as intensely in ColQ\(^{-/-}\) muscle as in control muscle (Fig. 8, k and l), providing direct evidence for the existence of additional synaptic VVA-B4-binding moieties.

**ColQ Is Required for Assembly of All Asymmetric AChE**

A symmetric forms of AChE are most abundant in skeletal muscle, but are also present in several other tissues (Tou-
the neuromuscular junction. Specifically, we have been able to answer the seven questions posed in the Introduction.

First, no asymmetric AChE is present in muscles that lack ColQ, presumably because the ColQ protein forms the collagen tail that endows asymmetric forms with their distinguishing feature. This result was expected but not inevitable. For example, ColQ might have been only one of multiple collagenous subunits, a possibility suggested by immunological studies showing a structural change in the collagen tail during posthatching development in birds (Tsim et al., 1988b). Alternatively, a synaptic collagen that normally plays other roles (Miner and Sanes, 1994, 1996) might have been capable of compensating for lack of ColQ in mutants.

Second, muscles from newborn ColQ−/− mice lack not only all asymmetric AChE, but also the nonamphiphilic tetrameric form of AChE, G4na. This observation was unexpected because the G4na molecules have been assumed to be homotetramers of AChE catalytic subunits. Possible explanations for this result include the following. G4na molecules may be derived from collagen-tailed molecules by proteolysis of the collagen tail, either in vivo or after extraction. A assembly of the tetramer might require transient association of AChET with ColQ. A assembly might require an alternatively spliced product of the ColQ gene that contains the amino-terminal PRAD domain, but not the collagenous domain. Such alternatively spliced products do in fact occur (Krejci et al., 1997), but have not yet been characterized. Some of the G4na may comprise tetramers associated with a single full-length collagen chain rather than a triple helix; such molecules sediment around 10S (Krejci, E., unpublished results). In any event, muscles of ColQ−/− mice did contain both amphiphilic and nonamphiphilic tetramers (G4a and G4na) at P20. These tetramers might have been assembled without any organizer or might contain an alternate organizing subunit such as the still incompletely characterized P subunit (Massoulié et al., 1993). Thus, although the PRAD of ColQ can induce tetramerization of AChE (Bon et al., 1997; Simon et al., 1998), and may do so in vivo at P0, there are clearly additional PRAD-independent mechanisms for forming tetramers.

Third, no detectable AChE is present at synaptic sites in ColQ−/− muscles. Histochemical, immunochemical, toxicological, and electrophysiological assays all gave similar results in this regard. The lack of synaptic AChE could reflect loss of collagen-tailed AChE and/or loss of G4 forms. We favor the interpretation that all synaptic AChE is collagen-tailed because all of the histochemically detectable AChE can be removed by collagenase treatment of sections (data not shown) or tissue (Hall and Kelly, 1971). In addition, all globular forms (G1a, G3a, G4a, and G4na) are present in ColQ−/− mice at P20, yet synaptic AChE is absent at all ages tested. Globular AChE is associated with isoamylase basal lamina from frog muscles (Anglister et al., 1994), but this may result from the alterations produced in the experimental system of muscle and nerve degeneration.

Fourth, ColQ is required for assembly or accumulation of asymmetric BuChE as well as AChE. In birds, collagen-tailed molecules containing both AChE and BuChE subunits have been detected in muscles of hatchlings (Tsim et al., 1988a), suggesting that both enzymes use a common tail. Likewise, Krejci et al. (1997) showed that antibodies to ColQ interact with both AChE and BuChE as assessed by sucrose density centrifugations. Taken together with these biochemical data, our genetic results strongly support the conclusion that the ColQ gene produces the collagen tails for both AChE and BuChE.

Fifth, the function of the neuromuscular junction is impaired in the absence of ColQ. However, the defects are not as severe as might be expected, considering the dramatic effects of acutely administering extremely selective blockers of peripheral AChE such as fasciculin. In particular, movements of ColQ−/− mice are relatively normal, while fasciculin-treated normal mice are paralyzed. Moreover, the postsynaptic necrosis observed in ColQ−/− weanlings, a likely result of excessive calcium influx through AChRs (Leonard and Salpeter, 1979), is largely absent from those mutants that survive to later ages. These observations suggest that neuromuscular junctions compensate for the absence of AChE. Compensation does not appear to involve persistence or upregulation of BuChE: asymmetric BuChE is lost in the mutant and the BuChE inhibitor, iso-OMPA, does not affect the time course of mepps.

Figure 9. ColQ is required for assembly of asymmetric forms of AChE in heart and brain. HS extracts of heart (a) or brain (b) of 2-mo-old mice were fractionated on sucrose gradients and assayed for AChE. Only portions of the gradient corresponding to 8–20S material are shown. No asymmetric AChE was detected in ColQ−/− tissue.
Instead, ultrastructural studies suggest a simple structural mechanism to limit the amount of neurotransmitter that reaches the postsynaptic membrane: invasion of the synaptic cleft by Schwann cell processes at ColQ−/− synapses. Enwrapping of nerve terminals by Schwann cell processes is, in fact, actively regulated by another component of synaptic basal lamina, laminin-11 (Patton et al., 1998). ColQ might exert a similar effect. In addition, compensatory changes in quantal size and quantal content may occur; such homeostatic changes have been documented in response to several chemical or genetic perturbations of the efficacy of neuromuscular transmission in both vertebrates and invertebrates (reviewed by Landmesser, 1998). Currently, we are using electrophysiological methods to seek such alterations in ColQ−/− mice, with the aim of learning how the synapse combines structural and physiological compensations to adapt to the challenge posed by AChE deficiency.

Sixth, initial formation of the neuromuscular junction proceeds in the absence of ColQ and AChE. A ChE promotes the extension of neurites from cultured motoneurons by a noncatalytic mechanism (Sternfeld et al., 1998; see also Small et al., 1995), and overexpression of AChE in transgenic frogs and mice perturbs formation of neuromuscular junctions in vivo (Seidman et al., 1994; Andres et al., 1997; Sternfeld et al., 1998). Our results imply that the developmental roles of ColQ and AChE at the synapse are either subtle or masked by compensatory mechanisms. However, it remains possible that expression of AChE at other sites is crucial for synaptogenesis. Moreover, at later stages, there are clear defects in the maturation of ColQ−/− neuromuscular junctions: some remain immature in geometry and others appear fragmented. These abnormalities could reflect a direct requirement for AChE or ColQ in synaptic maturation or maintenance, but might also be secondary consequences of subsynaptic necrosis or alterations in activity levels. Further experiments will be required to distinguish these alternatives.

Seventh, ColQ is required for assembly or accumulation of asymmetric AChE in all tissues tested, but tissues other than skeletal muscle are not obviously defective in structure or function. However, it is premature to conclude that ColQ plays no roles in these tissues. For example, asymmetric AChE is concentrated in synaptic layers of the retina (Ress et al., 1996); analyses of these synapses have not yet been undertaken. In addition, it is tempting to speculate that in AChE-poor tissues, ColQ or other PRA D-containing products of the ColQ gene serve to anchor proteins other than AChE to membranes or matrix. Identification of such proteins will guide the search for subtype phenotypes in ColQ−/− mice.

Finally, it is interesting to note the parallels between ColQ−/− mice and humans with a AChE-deficiency syndrome (CMS-Ic). This syndrome was described by Engel et al. (1977) over 20 years ago, and was reported just this year to result from defects in the human COLQ gene (Donger et al., 1998; Ohno et al., 1998). Some patients appear to be protein nulls, while others have mutations in the carboxy terminus of ColQ that permit formation of asymmetric AChE but prevent its stable association with the basal lamina of the synaptic cleft. AII of the COLQ patients are severely myasthenic, as are the ColQ−/− mice, making the mice a valid model for assessing pathogenic mechanisms and therapeutic interventions. We thank Mia Nichol, Carol Borgmeyer, Jimmy Gross, and Sherry Weng for assistance.

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