The Endothelium, A Protagonist in the Pathophysiology of Critical Illness: Focus on Cellular Markers

Sabrina H. van Ierssel,1,2 Philippe G. Jorens,1,2 Emeline M. Van Craenenbroeck,3,4 and Viviane M. Conraads3,4

1 Department of Critical Care Medicine, Antwerp University Hospital (UZA), University of Antwerp (UA), Wilrijkstraat 10, 2650 Edegem, Belgium
2 Department of Internal Medicine, Antwerp University Hospital (UZA) and University of Antwerp (UA), Wilrijkstraat 10, 2650 Edegem, Belgium
3 Laboratory of Cellular and Molecular Cardiology, Antwerp University Hospital (UZA), University of Antwerp (UA), Wilrijkstraat 10, 2650 Edegem, Belgium
4 Department of Cardiology, Antwerp University Hospital (UZA), University of Antwerp (UA), Wilrijkstraat 10, 2650 Edegem, Belgium

Correspondence should be addressed to Sabrina H. van Ierssel; sabrina.vanierssel@ua.ac.be

Received 5 November 2013; Revised 18 February 2014; Accepted 4 March 2014; Published 1 April 2014

Academic Editor: Iveta Bernatova

Copyright © 2014 Sabrina H. van Ierssel et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The endothelium is key in the pathophysiology of numerous diseases as a result of its precarious function in the regulation of tissue homeostasis. Therefore, its clinical evaluation providing diagnostic and prognostic markers, as well as its role as a therapeutic target, is the focus of intense research in patients with severe illnesses. In the critically ill with sepsis and acute brain injury, the endothelium has a cardinal function in the development of organ failure and secondary ischemia, respectively. Cellular markers of endothelial function such as endothelial progenitor cells (EPC) and endothelial microparticles (EMP) are gaining interest as biomarkers due to their accessibility, although the lack of standardization of EPC and EMP detection remains a drawback for their routine clinical use. In this paper we will review data available on EPC, as a general marker of endothelial repair, and EMP as an equivalent of damage in critical illnesses, in particular sepsis and acute brain injury. Their determination has resulted in new insights into endothelial dysfunction in the critically ill. It remains speculative whether their determination might guide therapy in these devastating acute disorders in the near future.

1. Introduction

The endothelium forms the inner layer of blood and lymphatic vessels [1, 2]. Besides its mere role as a barrier between blood and tissue, the endothelial cell layer displays a myriad of physiological functions. Integrity of the endothelium is required for adequate deliverance of oxygen and nutrition to tissue and the migration of blood cells. Furthermore it plays a central role in coagulation and fibrinolysis, it regulates vascular tone and the formation of new blood vessels. As such the endothelium is a key regulator of homeostasis, for which continuous interaction with its environment is crucial. Its importance in the pathophysiology of not only cardiovascular, but also inflammatory and malignant diseases, is increasingly recognized.

The clinical evaluation of the endothelium has been thwarted by its location at the inner side of the vessels. The growing interest in the endothelium as a central player in numerous diseases has stimulated the development of a multitude of new circulating markers and in vivo evaluation techniques [1].

In this review we will focus on critically ill patients with sepsis and acute brain injury, both devastating conditions seen frequently in the intensive care unit. Sepsis and acute brain injury are characterized by secondary complications, that is, multiorgan failure and cerebral ischemia, respectively, which have enormous impact on outcome. Vascular dysregulation and endothelial dysfunction play a central role herein. As such, markers of endothelial dysfunction are of potential interest in determining prognosis.
2. Enumeration of Circulating Cellular Markers of Endothelial Dysfunction

During the last two decades cellular markers of endothelial repair and damage have emerged as potential noninvasive candidates for functional evaluation of the endothelium. In this overview, we highlight the role of endothelial progenitor cells (EPC) as a marker of endothelial repair and endothelial microparticles (EMP) as a measure for endothelial damage. We will briefly discuss their methods of detection. For thorough discussion on these matters we refer to recently published reviews [3–7].

Endothelial progenitor cells originate from the bone marrow and can differentiate into mature endothelial cells [3, 8]. In situations of ischemia and in case of inflammation, EPC repair damaged endothelium and help in creating capillary networks, in a direct and paracrine fashion [9]. Several humoral factors are implicated in their mobilization, differentiation, and homing such as vascular endothelial growth factor (VEGF), granulocyte macrophage colony forming factor, stromal derived factor 1α (SDF1α), erythropoietin (EPO), amongst others [10]. Despite a multitude of papers published on EPC in various diseases, their definition remains a point of debate. The confusion on EPC definitions originates from the various techniques used for their detection having poor inter-method agreement (different types of cell culture techniques and flow cytometry) [3, 4, 11]. In cell culture techniques we discriminate two types, that is, short-term culture identifying early outgrowth EPC and long-term cultures isolating truly proliferating cells with endothelial fate [3, 4]. Since the first rather identifies hematopoietic cells involved in angiogenesis and the last are very elaborate, long-term cultures up to 30 days necessitating large amounts of blood and resulting in low colony counts, flow cytometry is at this moment the preferred technique for their detection in clinical studies. However there is a lack of any specific phenotypic marker for EPC to use in flow cytometry. Asahara et al. were the first to describe putative EPC, and they used a combination of CD34, a hematopoietic and progenitor cell marker, and flk-1/KDR, a receptor for VEGF important for homing of EPC and expressed on endothelial cells [8]. Both markers, however, are rather aspecific and as such are also expressed on mature circulating endothelial cells. For this reason Peichev et al. added CD133, a stem cell marker to better differentiate true EPC [12]. A drawback of using these triple positive cells, as circulating marker of endothelial function, is that their number is so low that enumeration becomes less reliable [3]. Furthermore it has been shown that these cells do develop into hematopoietic and not endothelial colonies [13]. EPC defined as CD34 and KDR positive cells have been most widely evaluated in clinical studies and have proven to be implicated in angiogenesis and endothelial repair in vivo [4, 9, 14]. Hence our research group prefers to use these cells as markers of endothelial repair, keeping in mind that this is a heterogeneous group of cells possessing an overlapping phenotype with endothelial cells and hematopoietic progenitors [6].

Endothelial microparticles (EMP) originate through vesiculation of the endothelial cell membrane upon cell activation, damage, or apoptosis [38]. EMP are membrane particles smaller than 1 μm, which contain oxidized phospholipids and proteins characteristic of endothelial cells. Surface antigens vary with the microparticle generating process; CD31+, CD105+, and Annexin V+ EMP are generated mainly during apoptosis, while CD62E, CD54, and CD106 expression are mostly seen when E are released upon activation [39, 40]. For their detection flow cytometry is the mainly used and mostly studied technique [41]. It has been shown that preanalytical and analytical heterogeneity amongst various research groups has led to differing results [38, 41, 42]. At this moment efforts are made for analytical and preanalytical standardization for flow cytometric detection of microparticles [43, 44]. Another difficulty for the use of EMP as biomarker in the critically ill is the possible interference with lipid-rich solutions [45]. The use of propofol and total parenteral nutrition in these patients could lead to secondary lipid accumulation negatively influencing the number of microparticles detected by flow cytometry. EMP are increasingly used as a marker of endothelial damage, especially in cardiovascular disorders, but growing evidence also indicates that EMP have an important modulating role in inflammation, coagulation, and vascular function [5, 38].

Multiparameter analysis for the evaluation of endothelial function is emerging as a valuable ex vivo tool for assessment of endothelial function [46], in addition to its potential to further unravel the pathophysiology of endothelial disruption in several disease conditions [7].

3. The Endothelium in Sepsis: The Orchestrator of Organ Failure

In one of four patients hospitalized at the intensive care unit severe sepsis is the reason for admission [47, 48]. Sepsis is defined as the systemic inflammatory response syndrome to an infection [49]. It is a devastating disorder that can progress to severe sepsis with the development of organ dysfunction, septic shock when hypotension is unresponsive to fluid resuscitation, and eventually multiorgan failure and death [49]. These stages of severity form a continuum in which patients evolve during their disease and treatment. The chance of survival decreases with the progression of the sepsis syndrome over this continuum. Hospital mortality in sepsis varies between 14 and 45% in Europe [47, 48]. Despite important advances in microbiological and supportive therapy, mortality has only slightly improved during the last decades [47]. Organ failure is the major cause of death in sepsis patients [50]. This is further supported by the finding that the number of organs that failed correlates with short-term mortality [51] and that the therapeutic improvement of organ failure early in sepsis improves survival [52].

3.1. Endothelial Function in Sepsis. The pathophysiology of sepsis is complex. Being multifactorial and heterogeneous among patients is two of the main characteristics of sepsis [53]. Sepsis is caused by a systemic maladaptive response of the host to an invading microorganism. Under normal conditions, infection triggers a local inflammatory reaction associated with an antiinflammatory response, local activation of the coagulation process together with a systemic acute
In infection, inflammation, and coagulation, endothelial dysfunction plays a central role in orchestrating both the physiologic and pathologic responses leading to the conquest of the infection in one case and the derailment and lead to sepsis in others. The exact factors leading to sepsis are not completely understood but are host and microorganism dependent. The endothelium has a central position in orchestrating both the physiologic and pathologic host response to infection due to its regulation of cellular permeability, coagulation, and vascular blood flow [54].

In the development of distant organ dysfunction, macro- and microvascular dysfunction play an important role. Macrovascular dysfunction during sepsis constitutes two major effects: hyperdynamic shock due to hypovolemia caused by venous and arterial vasodilation at the macrovascular level and capillary leak and cardiac depression [55]. On the other hand, it has been shown that tissue hypoperfusion remains despite normalization of macrocirculatory derangements, underlining the importance of additional microvascular derangements and mitochondrial dysfunction in sepsis [56, 57]. At microvascular level there is heterogeneity in flow, stopped flow, and decreased density of perfused capillaries [56]. As such, in sepsis the microcirculation is unable to adequately regulate microvascular perfusion to meet local oxygen demand.

The endothelium is a key component in the development of these macro- and microcirculatory disturbances in sepsis (see Figure 1). Activation of the endothelium leads to a procoagulant and proinflammatory condition, a disrupted barrier and an abnormal vascular tone [2]. In sepsis there is a direct destruction of the endothelial barrier [2, 58], and an increased amount of circulating endothelial cells has been shown in patients with septic shock [59]. The vasomotor regulation is also hampered in sepsis. More in particular there is an imbalance between vasodilator and vasoconstrictor signaling molecules leading to an impaired vasomotor tone. Despite increased concentration of circulating catecholamines in sepsis there is a decreased vascular response to these factors [60]. On the other hand, a disturbed endothelial mediated vasodilation has also been shown at macrovascular and microvascular level [61–64].

Evaluating circulating endothelial markers in patients with sepsis has evolved from circulating endothelial adhesion molecules to cellular markers of endothelial repair and damage.

### 3.2. Endothelial Progenitor Cells in Sepsis

Several groups have investigated EPC in sepsis but their role has not yet been unequivocally defined [15–22] (see Table 1). Observational studies found an increased percentage of circulating EPC enumerated by flow cytometry in highly selected patients with sepsis [15, 19, 20], while experimental studies, that is, the administration of LPS in healthy volunteers and a MODS model in pigs showed a decreased number [17, 18]. At our own center, we found a decreased absolute number of EPC in a heterogeneous group of severe sepsis patients compared to healthy volunteers [22]. Furthermore, while Becchi et al. found an increased number of EPC in severe sepsis versus sepsis patients, Rafat et al. found a positive correlation between EPC number and survival [15, 20]. Our data are in line with the last findings, with lower numbers of absolute EPC in patients with increasing sequential organ failure assessment (SOFA) score, a measure of severity of organ failure, during the first week after sepsis. Differences in study population, expressing results as percentage of peripheral blood mononuclear cells (PBMC) (which are decreased in sepsis) versus absolute numbers; methodology (isolated PBMC versus whole blood) and the different phenotypes studied can explain these opposing findings. Despite these contradictory findings, the functional impairment of EPC seems indisputable in sepsis [16–19, 22]. All studies, case-control and experimental, describe decreased proliferative or migratory capacities of EPC [16, 17, 19, 21, 22]. Data on the exact role of EPC in vascular repair during sepsis are scarce. Lam et al., for example, showed that EPC transplantation in a rabbit ARDS model decreased endothelial dysfunction, maintained the alveolocapillary membrane, and reduced inflammation [65]. As mentioned before, numerous humoral factors influence EPC mobilization, differentiation, and homing; therefore, EPC are an important therapeutic target to stimulate endothelial repair in sepsis. Several therapeutic strategies that focus on sepsis-related endothelial dysfunction have been shown to influence EPC [66], for example, statins, shown to increase EPC number and ameliorate their functional capacity, that is, decreasing senescence and improving proliferation [67, 68].

### 3.3. Endothelial Microparticles in Sepsis

Several studies, case-control human studies as well as animal models, have explored EMP in sepsis, with differing results (see Table 2) [22, 23, 25–27, 29]. EMP were found to be increased in patients with sepsis by some research groups [25, 26, 29], while others found a decreased or equal number [22, 23].

---

**Figure 1:** Endothelial dysfunction in sepsis. In sepsis the reaction that has the aim of containing the infection derails and leads to a proinflammatory, procoagulant situation and endothelial dysfunction, finally resulting in the development of organ failure.
| Study group       | Study type                  | Phenotype EPC                                                                 | Main findings                                                  |
|------------------|-----------------------------|------------------------------------------------------------------------------|-----------------------------------------------------------------|
| Becchi et al. [15] | Case-control sepsis (n = 24) | CD34+ KDR+ in isolated PBMC                                                   | (i) Increased % EPC the first 72 h after sepsis                 |
|                  |                             | CD34+ KDR+ CD133+ in isolated CD34+ cells                                     | (ii) EPC severe sepsis ≫ sepsis                                 |
| Cribbs et al. [16] | Case-control sepsis (n = 86) | CFU-EPC                                                                      | (i) Decreased CFU-EPC in sepsis                                |
|                  |                             |                                                                              | (ii) Inversely associated with SOFA                             |
| Luo et al. [17]   | MODS model in pigs (n = 20)  | CD133+ CD34+ in WB CFU-EPC                                                    | Decreased EPC, CFU-EPC and migratory function in MODS          |
|                  |                             | Migration to VEGF                                                            |                                                                  |
| Mayr et al. [18]  | LPS in healthy volunteers (n = 32) | CD34+ KDR+ CD133+ EPC in WB CFU-EPC                                            | (i) Decrease in EPC number with nadir at 6 h post LPS           |
|                  |                             |                                                                              | (ii) Decreased CFU-EPC nadir 4 h after LPS                     |
| Patschan et al. [19] | Case-control sepsis (n = 40) | KDR+ CD133+ in isolated PBMC CFU-EPC                                         | (i) Increased % EPC in sepsis                                  |
|                  |                             |                                                                              | (ii) Decreased CFU-EPC in sepsis                               |
| Rafat et al. [20] | Case-control sepsis (n = 32) | CD34+ KDR+ CD133+ in isolated PBMC                                          | (i) Increased % EPC in sepsis                                  |
|                  |                             |                                                                              | (ii) Lower % EPC in nonsurvivors                               |
| Schlichting et al. [21] | Case-control severe sepsis (n = 18) | CFU-EPC                                                                      | No difference                                                  |
|                  |                             |                                                                              |                                                                  |
| van Ierssel et al. [22] | Case-control severe sepsis (n = 30) | CD34+ KDR+ in WB Migration to SDF-1α and VEGF                                  | (i) Decreased absolute number                                 |
|                  |                             |                                                                              | (ii) Decreased migratory capacity                                |
|                  |                             |                                                                              | (iii) Impending organ dysfunction the first week was associated with lower EPC and a trend to impaired migration |

CFU-EPC: EPC colony forming units; EPC: endothelial progenitor cells; LPS: lipopolysaccharides; MODS: multiorgan dysfunction syndrome; PBMC: peripheral blood mononuclear cells; SDF-1α: stromal derived factor 1α; SOFA: sequential organ failure assessment; VEGF: vascular endothelial growth factor; WB: whole blood.

These inconsistent results may result from a lack of pre- and analytical standardization of microparticle (MP) detection, the different phenotypes studied, and differences in study population. In contrast to the interpretation in cardiovascular diseases, where an elevation of EMP is considered a marker of endothelial dysfunction, the number of EMP is positively related to survival and inversely correlated with the SOFA-score in patients with sepsis [29]. Since it is becoming more and more clear that microparticles are more than simple markers of endothelial damage or activation, their interpretation as marker of endothelial dysfunction is less unambiguous. As such it has been shown that the general pool of MP in septic patients is protective against vascular hyporeactivity in vitro, increasing the response to 5-HT in vitro while not affecting endothelium-dependent vasodilation [25]. Mortaza et al., on the other hand, found that injection of MP from septic rats induced vasodilatory shock in healthy animals [24]. MP also have been implicated in hypercoagulability and inflammation [23, 26, 29]. Finally Pérez-Casal et al. found increased numbers of MP bearing endothelial protein C receptor (EPCR) of endothelial and monocytic origin in patients treated with recombinant protein C [28]. These MP decreased apoptosis and reduced permeability in endothelial cells in an APC dependent way, a confirmation of earlier in vitro findings [69]. At this moment the knowledge on EMP functions in sepsis is too scarce to clarify their role in the development of organ failure.

4. The Endothelium as Key Player in Secondary Cerebral Ischemia after Acute Brain Injury

Acute brain injury, more in particular subarachnoid hemorrhage (SAH) and traumatic brain injury (TBI), is devastating neurological events which have an important socioeconomic impact. The development of secondary cerebral ischemia is an important prognostic factor in both SAH and TBI [70–72]. It develops in 8–12% and 20–30% of patients after TBI and SAH, respectively, mostly within the first 2 weeks after the insult [70, 71, 73, 74].

4.1. Endothelial Function and Secondary Cerebral Ischemia after Subarachnoid Bleeding. In SAH the concepts of delayed cerebral ischemia (DCI) and cerebral vasospasm have been well studied and clearly defined (see Figure 2) [75]. While previously macrovascular cerebral vasospasm was thought key for the development of DCI, it is now accepted to be a multifactorial process of which the exact underlying mechanisms are not yet completely unraveled [75]. As such it has been repetitively shown that macrovascular vasospasms are not a condition sine qua non to develop DCI, and on the other hand not all vasospasms will lead to the development of DCI [74]. Other mechanisms such as microvascular dysfunction, disturbed autoregulation, thromboembolism, and cortical spreading depression have been implicated in
### Table 2: Overview endothelial microparticles in sepsis.

| Study group                        | Study design                              | Detection method   | Phenotype EMP             | Main findings                                                                 |
|------------------------------------|-------------------------------------------|--------------------|----------------------------|-------------------------------------------------------------------------------|
| Joop et al. [23]                   | Case-control MODS and sepsis (n = 9)      | Flow cytometry     | CD62E+/- Annexin V+        | (i) Lower number CD62E+ EMP                                                   |
|                                    |                                            | isolated MP frozen samples | CD62E+/- Annexin V+        | (ii) Unchanged number CD144+ EMP                                               |
| Mortaza et al. [24]                | Rat cecal ligation and puncture model     | Flow cytometry     | CD54+/ Annexin V+         | (i) Unchanged EMP in sepsis                                                    |
|                                    |                                            | PFP                |                            | (ii) Septic MP caused vasoplegic shock in healthy rats                         |
| Mostefai et al. [25]               | Case-control sepsis (n = 36) mouse model: injection of septic MP | Flow cytometry     | CD146+                     | (i) Increased EMP in sepsis                                                    |
|                                    |                                            | PFP                |                            | (ii) Septic MP induced increased responsiveness to vasoconstrictors in aortic rings |
| Nieuwland et al. [26]              | Case-control meningococcal sepsis (n = 7) | Flow cytometry     | CD62E+/- Annexin V+        | Nonsignificant increase in sepsis                                               |
|                                    |                                            | isolated MP        |                            |                                                                               |
| Ogura et al. [27]                  | Case-control severe SIRS (n = 28, sepsis = 12) | Flow cytometry     | CD54+CD31+ EMP            | EMP increased in sepsis                                                        |
|                                    |                                            | PRP                |                            |                                                                               |
| Pérez-Casal et al. [28]            | Case control study of APC treated sepsis patients | Flow cytometry     | CD13+ EPCR+                | Increased CD13+ EPCR+ MP                                                       |
|                                    |                                            | isolated MP        |                            |                                                                               |
| Soriano et al. [29]                | Case control severe sepsis (n = 35)       | Flow cytometry     | CD31+ CD42b−              | (i) EMP higher in severe sepsis                                                |
|                                    |                                            | PPP                |                            | (ii) EMP higher in survivors                                                   |
|                                    |                                            |                    |                            | (iii) Negative correlation with SOFA on D2 and D3                               |
| van Ierssel et al. [22]            | Case-control severe sepsis (n = 26)       | Flow cytometry     | CD31+ CD42b−              | Unchanged number of EMP versus healthy controls                               |
|                                    |                                            | PPP                |                            |                                                                               |

EMP: endothelial microparticles; MODS: multiorgan dysfunction syndrome; MP: microparticle; PFP: platelet free plasma; PPP: platelet poor plasma; PRP: platelet rich plasma; SOFA: sequential organ dysfunction assessment.

**Figure 2**: Endothelial dysfunction in SAH. SAH: subarachnoid hemorrhage. In subarachnoid hemorrhage, the development of delayed cerebral ischemia is a multifactorial process in which besides macrovascular vasospasm; thromboembolism, disturbed autoregulation, microvascular dysfunction, and cortical spreading depression are involved. Endothelial dysfunction is a key factor in the development of these processes. It is not clarified yet if local and general inflammation are causal factors or bystanders in the development of secondary ischemia.

4.2. **Endothelial Function and Secondary Ischemia after Traumatic Brain Injury.** In traumatic brain injury (TBI), on the other hand, the concept of posttraumatic cerebral ischemia is less well studied and understood. This can be explained by the fact that patients with TBI are a very heterogeneous group and that besides the primary cerebral injury other extra-cerebral processes may cause secondary damage [70, 71, 85]. The mechanisms involved are mechanical compression,
hypotension, direct vascular injury, thromboembolism, and posttraumatic cerebral vasospasm; moreover, distinguishing these is difficult (see Figure 3). The appearance of posttraumatic cerebral vasospasms has been related to the presence of traumatic subarachnoid blood but has also been reported in the absence of a traumatic SAH [86]. These findings suggest that besides the mechanisms important in the development of vasospasm after spontaneous SAH, other processes are involved after TBI, such as direct stretching and mechanical irritation [88]. Furthermore the relation between cerebral vasospasm and the development of secondary ischemia is still a point of debate, and there are only few prospective studies on this matter [86, 87]. Besides macrovascular changes, the microvasculature is also involved. As such in animal experiments diffuse loss of microvasculature networks and capillary density after TBI were found [88]. Increased VEGF expression, indicating a possible role for neovascularization [88], and impaired cerebral endothelium-dependent cerebral vascular responses have also been documented [89].

4.3. Endothelial Progenitor Cells after Acute Brain Injury. Until now research on markers of endothelial function in SAH and TBI has mostly focused on circulating endothelial adhesion molecules and markers of endothelial activation [90–92], both of which are increased in patients developing secondary cerebral ischemia. Endothelial progenitor cells show a biphasic response after traumatic brain injury; after an initial decrease they peak 7 days after the insult (see Table 3) [31]. Furthermore they have been associated with an improved outcome after TBI [32]. In patients with cerebral aneurysm a decreased number of EPC also has been shown, possibly related to patients’ risk factors (e.g., smoking and hypertension) [34]. Our group also enumerated EPC in patients with SAH and TBI and confirmed the finding of a decreased number of EPC initially after the insult. (van Ierssel S.H., unpublished results) publication) Furthermore an impaired functional capacity of EPC was seen [30, 34]. After endovascular coiling of ruptured aneurysm there is a rapid increase of EPC with a peak at 14 days after rupture [33]. At this moment, we are not aware of studies on the relation between EPC and DCI or posttraumatic cerebral ischemia. The exact role of EPC in vascular repair after acute brain injury has not been studied yet. In a rat model of traumatic brain injury Wang et al. looked at the role of atorvastatin [93]. They found an increased number of EPC and enhanced cerebral angiogenesis, together with an improved functional outcome in treated rats. These results again show the importance of EPC as a possible therapeutic target.

4.4. Endothelial Microparticles in Acute Brain Injury. Few researchers have looked at the evolution of endothelial

| Study group               | Study type                          | Phenotype EPC                  | Main findings                                                                 |
|--------------------------|-------------------------------------|--------------------------------|-------------------------------------------------------------------------------|
| Liang et al. [30]        | Case-control unruptured intracranial aneurysm (n = 24) | CFU-EPC Migration to VEGF     | Decreased proliferative and migratory capacity of EPC                        |
| Liu et al. [31]          | Case-control TBI (n = 29)           | CD34+ CD133+ in isolated PBMC  | Decreased EPC in TBI, steady increase from day 5–7 with peak day 7           |
| Liu et al. [32]          | Case-control TBI (n = 84)           | CD34+ CD133+ in isolated PBMC  | (i) Decreased EPC 24–48 h after TBI, increase to day 7 (ii) Non-survivors lower EPC |
| Wei et al. [33]          | Case-control ruptured cerebral aneurysm (n = 14) | CD34+ CD133+ Isolated PBMC    | (i) Decreased number of EPC in patients (ii) Increase after coiling with a peak at day 14 |
| Wei et al. [34]          | Case-control cerebral aneurysm (n = 56, ruptured n = 35) | CD34+ CD133+ CD34+ KDR+ in isolated PBMC Migration to VEGF | (i) Both EPC phenotypes reduced in cerebral aneurysm (ii) Impaired migration and increased of EPC in cerebral aneurysm |

EPC: endothelial progenitor cells; PBMC: peripheral blood mononuclear cells; VEGF: vascular endothelial growth factor; TBI: traumatic brain injury.
Table 4: Overview on endothelial microparticles in acute brain injury, SAH, and TBI.

| Study group       | Study population       | Detection method     | Phenotype EMP                     | Main findings                                                                 |
|-------------------|------------------------|----------------------|-----------------------------------|-------------------------------------------------------------------------------|
| Lackner et al.    | Case-control spontaneous SAH (n = 20) | Flow cytometry plasma | CD105+/Annexin V+ or −          | (i) Increased number of all EMP phenotypes studied in SAH versus healthy       |
|                   |                        |                      | CD62E+/Annexin V+ or −          | (ii) In patients with Doppler detected cerebral vasospasm increased          |
|                   |                        |                      | CD54+/Annexin V+ or −          | CD105+/Annexin V+ and CD62E+/Annexin V+                                       |
|                   |                        |                      | CD106+/Annexin V+ or −         | (iii) CD105+/Annexin V+ associated with cerebral infarction                   |
| Morel et al. [36] | Case-control TBI (n = 16) | Capture technique PFP and CSF | Annexin V+ CD31+ | (i) Increased MP number in plasma and CSF at D0, decreased D3, D5, D10       |
|                   |                        |                      |                                  | (ii) High proportion of EMP                                                   |
| Sanborn et al.    | Case-control SAH (n = 22) | Flow cytometry Frozen plasma samples | CD146+/Annexin V+ | (i) Elevated EMP after SAH, and remained high until D10                      |
|                   |                        |                      |                                  | (ii) Negative correlation EMP and infarction at D14                            |

CSF: cerebral spinal fluid; EMP: endothelial microparticles; MP: microparticle, PFP: Platelet free plasma; SAH: subarachnoid hemorrhage; TBI: traumatic brain injury.

microparticles after SAH and TBI (see Table 4) [35–37]. They all found an increased number of EMP after acute brain injury, in line with the common use of EMP as markers of endothelial damage [38]. With regard to the development of secondary ischemia, the relation seems more ambiguous. While Lackner et al. found an increased number of CD105+/Annexin+ EMP early after the insult, Sanborn et al. found a decreased number of CD146+ EMP at Day 1 in patients developing DCI [35, 37]. The different populations that were studied can explain these opposing results, as well as the variable pre- and analytical methods used and the variances in phenotype of EMP studied. At this moment there are no data available on the exact functional role of EMP after acute brain injury.

5. Conclusion

The endothelium seems to be a central actor in the development of organ failure in sepsis and secondary ischemia after acute brain injury, as illustrated here for SAH and TBI. The exact role of markers of repair (EPC) and injury (EMP), however, needs further clarification. Nevertheless the importance of both organ failure and secondary ischemia in the prognosis of these devastating disorders explains the craving for adequate prognostic and therapeutic clues and hence the interest in the endothelium makes common sense.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgments

This work was supported by a grant from the Research Foundation-Flanders, Belgium, to Sabrina H. van Ierssel and Viviane M. Conraads. In between the review process and the resubmission, the senior author of this manuscript passed away. Prof. V. Conraads, MD, PhD significantly contributed to this review and approved the final version that was submitted initially. Therefore our colleague’s name is retained as the last author, in order to show great respect for her previous achievements in this field and her contribution to this review.

References

[1] J. E. Deanfield, J. P. Halcox, and T. J. Rabelink, “Endothelial function and dysfunction: testing and clinical relevance,” Circulation, vol. 115, no. 10, pp. 1285–1295, 2007.
[2] W. C. Aird, “The role of the endothelium in severe sepsis and multiple organ dysfunction syndrome,” Blood, vol. 101, no. 10, pp. 3765–3777, 2003.
[3] G. P. Fadini, D. Losordo, and S. Dimmeler, “Critical reevaluation of endothelial progenitor cell phenotypes for therapeutic and diagnostic use,” Circulation Research, vol. 110, no. 4, pp. 624–637, 2012.
[4] M. C. Yoder, “Human endothelial progenitor cells,” Cold Spring Harbor Perspectives in Medicine, vol. 2, no. 7, Article ID a006692, 2012.
[5] F. Dignat-George and C. M. Boulanger, “The many faces of endothelial microparticles,” Arteriosclerosis, Thrombosis, and Vascular Biology, vol. 31, no. 1, pp. 27–33, 2011.
[6] E. M. van Craenenbroeck, A. H. van Craenenbroeck, S. van Ierssel et al., “Quantification of circulating CD34+/KDR+/CD45dim endothelial progenitor cells: analytical considerations,” International Journal of Cardiology, vol. 167, no. 5, pp. 1688–1695, 2013.
[7] D. Burger and R. M. Touyz, “Cellular biomarkers of endothelial health: microparticles, endothelial progenitor cells, and circulating endothelial cells,” Journal of the American Society of Hypertension, vol. 6, no. 2, pp. 85–99, 2012.
[8] T. Asahara, T. Murohara, A. Sullivan et al., “Isolation of putative progenitor endothelial cells for angiogenesis,” Science, vol. 275, no. 5302, pp. 964–967, 1997.
[70] H.-L. Tian, Z. Geng, Y.-H. Cui et al., "Risk factors for posttraumatic cerebral infarction in patients with moderate or severe head trauma," Neurosurgical Review, vol. 31, no. 4, pp. 431–436, 2008.

[71] I. Tawil, D. M. Stein, S. E. Mirvis, and T. M. Scalea, "Posttraumatic cerebral infarction: incidence, outcome, and risk factors," Journal of Trauma—Injury, Infection and Critical Care, vol. 64, no. 4, pp. 849–853, 2008.

[72] M. D. I. Vergouwen, D. Ildigwe, and R. L. MacDonald, "Cerebral infarction after subarachnoid hemorrhage contributes to poor outcome by vasospasm-dependent and -independent effects," Stroke, vol. 42, no. 4, pp. 924–929, 2011.

[73] N. K. de Rooij, J. P. Greving, G. J. Rinkel, and C. J. Frijns, "Early prediction of delayed cerebral ischemia after subarachnoid hemorrhage: development and validation of a practical risk chart," Stroke, vol. 44, no. 5, pp. 1288–1294, 2013.

[74] R. J. Brown, A. Kumar, R. Dhar, T. R. Sampson, and M. N. Diringer, "The relationship between delayed infarcts and angiographic vasospasm after aneurysmal subarachnoid hemorrhage," Neurosurgery, vol. 72, no. 5, pp. 702–708, 2013.

[75] M. D. I. Vergouwen, M. Vermeulen, J. van Gijn et al., "Definition of delayed cerebral ischemia after aneurysmal subarachnoid hemorrhage as an outcome event in clinical trials and observational studies: proposal of a multidisciplinary research group," Stroke, vol. 41, no. 10, pp. 2391–2395, 2010.

[76] J. P. Dreier, J. Wotizik, M. Fabricius et al., "Delayed ischaemic neurological deficits after subarachnoid haemorrhage are associated with clusters of spreading depolarizations," Brain, vol. 129, no. 12, pp. 3224–3237, 2006.

[77] J. M. K. Lam, P. Smelevski, M. Czosnyka, J. D. Pickard, and P. J. Kirkpatrick, "Predicting delayed ischemic deficits after aneurysmal subarachnoid hemorrhage using a transient hyperemic response test of cerebral autoregulation," Neurosurgery, vol. 47, no. 4, pp. 819–826, 2000.

[78] M. D. I. Vergouwen, M. Vermeulen, B. A. Coert, E. S. G. Stroes, and Y. B. W. E. M. Roos, "Microthrombosis after aneurysmal subarachnoid hemorrhage: an additional explanation for delayed cerebral ischemia," Journal of Cerebral Blood Flow and Metabolism, vol. 28, no. 11, pp. 1761–1770, 2008.

[79] M. Sabri, J. Ai, K. Lakovic, J. D’Abbondanza, D. Ildigwe, and R. L. MacDonald, "Mechanisms of microthrombi formation after experimental subarachnoid hemorrhage," Neuroscience, vol. 224, pp. 26–37, 2012.

[80] G. W. Britz, J. R. Meno, I.-S. Park et al., "Time-dependent alterations in functional and pharmacological arteriolar reactivity after subarachnoid hemorrhage," Stroke, vol. 38, no. 4, pp. 1329–1335, 2007.

[81] E. Kóniewska, R. Michalik, J. Rafałowska et al., "Mechanisms of vascular dysfunction after subarachnoid hemorrhage," Journal of Physiology and Pharmacology, vol. 57, no. 11, pp. 145–160, 2006.

[82] K. W. Park, C. Metais, H. B. Dai, M. E. Comunale, and F. W. Sellke, "Microvascular endothelial dysfunction and its mechanism in a rat model of subarachnoid hemorrhage," Anesthesia and Analgesia, vol. 92, no. 4, pp. 990–996, 2001.

[83] S. Park, M. Yamaguchi, C. Zhou, J. W. Calvert, J. Tang, and J. H. Zhang, "Neurovascular protection reduces early brain injury after subarachnoid hemorrhage," Stroke, vol. 35, no. 10, pp. 2412–2417, 2004.

[84] Y. Kubo, K. Ogasawara, S. Kakino et al., "Serum inflammatory adhesion molecules and high-sensitivity C-reactive protein correlates with delayed ischemic neurologic deficits after subarachnoid hemorrhage," Surgical Neurology, vol. 69, no. 6, pp. 592–596, 2008.

[85] A. Server, R. Dullerud, M. Haakonsen, P. H. Nakstad, U. L.-H. Johnsen, and B. Magnaes, "Post-traumatic cerebral infarction: neuroimaging findings, etiology and outcome," Acta Radiologica, vol. 42, no. 3, pp. 254–260, 2001.

[86] S. S. Armin, A. R. T. Colohan, and J. H. Zhang, "Vasospasm in traumatic brain injury," Acta Neurochirurgica, Supplementum, vol. 104, no. 13, pp. 421–425, 2008.

[87] M. Taneda, K. Kataoka, F. Akai, T. Asai, and I. Sakata, "Traumatic subarachnoid hemorrhage as a predictable indicator of delayed ischemic symptoms," Journal of Neurosurgery, vol. 84, no. 5, pp. 762–768, 1996.

[88] E. Park, J. D. Bell, I. P. Siddiq, and A. J. Baker, "An analysis of regional microvascular loss and recovery following two grades of fluid percussion trauma: a role for hypoxia-inducible factors in traumatic brain injury," Journal of Cerebral Blood Flow and Metabolism, vol. 29, no. 3, pp. 575–584, 2009.

[89] H. A. Kontos and E. P. Weï, "Endothelium-dependent responses after experimental brain injury," Journal of Neurotrauma, vol. 9, no. 4, pp. 349–354, 1992.

[90] M. D. I. Vergouwen, K. Bakhtiari, N. Van Geloven, M. Vermeulen, Y. B. Roos, and J. C. M. Meijers, "Reduced ADAMTS13 activity in delayed cerebral ischemia after aneurysmal subarachnoid hemorrhage," Journal of Cerebral Blood Flow and Metabolism, vol. 29, no. 10, pp. 1734–1741, 2009.

[91] C. J. M. Frijns, R. Fijnheer, A. Algra, J. A. van Mourik, J. van Gijn, and G. J. E. Rinkel, "Early circulating levels of endothelial cell activation markers in aneurysmal subarachnoid haemorrhage: associations with cerebral ischaemic events and outcome," Journal of Neurology, Neurosurgery and Psychiatry, vol. 77, no. 1, pp. 77–83, 2006.

[92] J. J. Nissen, D. Mantle, B. Gregson, and A. D. Mendelow, "Serum concentration of adhesion molecules in patients with delayed ischaemic neurological deficit after aneurysmal subarachnoid haemorrhage: the immunoglobulin and selectin superfamilies," Journal of Neurology Neurosurgery and Psychiatry, vol. 71, no. 3, pp. 329–333, 2001.

[93] B. Wang, L. Sun, Y. Tian et al., "Effects of atorvastatin in the regulation of circulating EPCs and angiogenesis in traumatic brain injury in rats," Neurological Sciences, vol. 319, no. 1-2, pp. 117–123, 2012.