Friend of Prmt1, FOP is a Novel Component of the Nuclear SMN Complex Isolated Using Biotin Affinity Purification

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Abstract

SMN (survival motor neuron protein) complexes are essential for the biogenesis of uridine-rich small nuclear ribonucleoproteins (UsnRNPs). During the biogenesis, the SMN complexes bound to UsnRNPs are transported from the cytoplasm to the nucleus, and moved to Cajal body (bodies)/Gems (Cajal/Gems) where the SMN complexes-UsnRNPs are subjected to additional chemical modifications and dissociated to the SMN complexes and the mature UsnRNPs. Although the mature UsnRNPs are assembled into spliceosome with newly transcribed pre-mRNA in the perichromatin fibrils at the chromatin, the role of the dissociated nuclear SMN complexes remains undetermined. In this study, we identified Friend of Prmt1 (FOP; chromatin target of Prmt1, CHTOP; C1orf77) as a novel component of the nuclear SMN complexes by the biotin affinity purification, coupled with the mass spectrometry-based protein identification. FOP was associated with SMN, Gemines 2, 3, 4, 6, and 8, unrip, and fragile X mental retardation 1 protein (FMR1), as well as with U6 and U6 snRNAs in the nucleus, but not with Sm proteins, Gemins5, collin, and U1 and U2snRNAs. Using the quantitative proteomic method with SILAC coupled with RNA interference, we also showed that FOP is required for the association of the SMN complexes with hnRNPs, histone proteins, and various RNA-binding proteins. It is reported that FOP localizes mainly in the nuclear speckles, binds chromatin, and plays a role in mRNA transcriptional regulation. Our present data suggest that the nuclear SMN complex containing FOP participates in the process of mRNA post-transcriptional regulation.

Keywords: SMN; Gemin; C1orF77; Chromatin target of PrMT1; CHTOP; SRAG; LC-MS/MS; SILAC

Abbreviation: CHTOP: Chromatin Target of Prmt1; Collin: Collin p80; FMR1: Fragile X Mental Retardation 1 Protein; FOP: Friend of Prmt1; GARN: Glycine-Ariginine-Rich; LC: Liquid Chromatography; MS: Mass Spectrometry; Prmt: Protein Arginine Methyl Transferase; SILAC: Stable Isotope Labeling using Amino Acids in Cell Culture; SMA: Spinal Muscular Atrophy; SMN: Survival of Motor Neuron; SRAG: Small Protein Rich in Arginine and Glycine; unrip: uninteracting Protein; UsnRNP: Uridine-rich Small Nuclear Ribonucleoprotein

Introduction

SMN is linked to one of the most common inheritable causes of childhood mortality, spinal muscular atrophy (SMA) [1], a neuromuscular disorder characterized by loss of motor neurons, which results from low levels of, or loss-of-function mutations in the SMN protein [2,3]. The SMN gene is duplicated as an inverted repeat on chromosome 5. Only mutations in the telomeric copy, SMN1, give rise to SMA [4]; another centromeric copy, SMN2, expresses a functionally defective form of the protein that lacks the carboxyl terminus encoded by exon 7, but its mutations are not related to SMA.

Human SMN protein is a 294-amino-acid polypeptide that forms complexes with at least eight proteins; Gemin2 (SIP1), Gemin3 (DEAD box RNA helicase 20, DDX20), Gemin4, Gemin5 (a WD-repeat protein), Gemin6, Gemin7, Gemin8 and unrip (unr interacting protein, STRAP). The SMN complexes promote assembly of UsnRNP core complex consisting seven different Sm proteins, and U1, U2, U4, U5, or U7 snRNAs [5]; i.e., the SMN complexes assist the ordered assembly of the seven Sm proteins as three sub-complexes, B–D–D3, D1–D2 and E–F–G Sm onto the Sm sequence of UsnRNAs [6]. Protein arginine methyl transferase (Prmt) 5 promotes this assembly, in which it catalyzes symmetrical arginine methylation of some of the Sm core proteins [7]. Indeed, SMN interacts with a dimethylarginine residue on some of the Sm core proteins present in the UsnRNP core complex [8]. Gemin5 and unrip also participate in this UsnRNP assembly, and are detached from the SMN complexes-UsnRNPs at later stages of the biogenesis in the nucleus [9-11]. The assembled cytoplasmic UsnRNPs are transported into the nucleus, and are targeted to Cajal/Gems (splicing snRNPs) or to the histone locus body (U7 snRNP) that constitutes a part of Cajal/Gems. Collin p80 (collin) is required for the localization of SMN complexes in the Cajal/Gems through direct interaction of SMN with a symmetrically dimethylated arginine in a RG domain of collin [12,13]. Collin also interacts with Sm proteins in UsnRNPs through its C-terminal domain, and the binding is not

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affected by dimethylation [14]. Sm proteins and UsnRNPs are probably released from the SMN complexes upon the binding with coflin in Cajal/Gems [15], and are then transferred into splicing speckles [16]. Dissociation of Gemin5 may also promote the release of UsnRNPs from SMN complexes in Cajal/Gems, since Gemin5 is required for the initial binding of snRNA to the SMN complexes in the cytoplasm [17]. The UsnRNPs that are released from the Cajal/Gems then receive additional proteins from the nuclear speckles. The final assembly of the spliceosome from its component UsnRNPs (and other proteins) takes place in situ on nascent pre-mRNAs at the site of transcription that occurs at the outer edge of the nuclear speckle, where all or almost all introns are removed in conjunction with or soon after the transcription [18]. The nuclear speckles are rich in spliced mRNAs and various pre-mRNA metabolic factors, such as spliceosome assembly factor SC35, and thus, are believed to be the places that complete mRNA maturation, release for mRNA export with some recycling factors, and recycle splicing complex including UsnRNPs [19]. Although it has been well documented that improper process of mRNAs impedes transit of the mRNAs through the nuclear speckle [20,21], the mechanism how the mRNAs are retained in recycle splicing complex and impeded the transit remains undetermined. The splicing complexes including UsnRNPs that are recycled after each round of splicing into the cytoplasm, must pass again through the Cajal/Gems for re-assembly before they can take part in a further rounding of splicing [22-24].

In this study, we isolated SMN complexes present in the nucleus and identified Friend of Prmt1, FOP as a novel component of the nuclear SMN complexes. As noted by alternative names of FOP such as SRAG (small protein rich in arginine and glycine) [25] and CHTOP (Chromatin target of Prmt1) [26], human FOP has a central glycine-arginine-rich (GAR) domain containing 26 RG/GR repeats that is recognized and methylated with Prmt1, localized mainly in the nuclear speckles and tightly associated with facultative chromatin in interphase cells [26]. The C-terminus of FOP also harbors a duplication of the sequence LDXXLDAYM (where “X” is any amino acid) [26]. FOP is expressed widely among tissues and cell lines [25] and is highly conserved in all vertebrates, while no orthologs was identified for FOP is expressed widely among tissues and cell lines [25], and is highly conserved in all vertebrates, while no orthologs was identified.

**Materials and Methods**

**Reagents and antibodies**

Antibodies against SMN (2B1), Gemin2 (2E17), Gemin3 (12H12), Gemin4 (17D10), Gemin6 (1F8), Gemin8 (2E12), and Unrip (3G6) were generous gifts from Dr. Gideon Dreyfuss. Anti-Gemin5 was obtained from Bethyl Laboratories. Antibody to fibrillarin, lamin B, p32, and FMRI were obtained from Santa Cruz. Anti-FLAG-M2, anti-collin p80, and anti-FLAG-M2-conjugated agarose beads were obtained from Sigma-Aldrich. Antibody to FOP (KT64) was prepared as described [26]. Anti-GAPDH was obtained from Ambion. Anti-Prmt1 was obtained from Upstate. HRP conjugated anti-mouse IgG was obtained from Cell Signaling Technology. HRP–conjugated anti-rabbit IgG were obtained from Sigma-Aldrich. All general reagents were purchased from Wako Pure Chemical, Kanto Chemical, or Nacalai Tesque.

**Cell culture**

Flip-In T-Rex 293 (293T Rex) cells, 293EBNA cells, HEK293 cells, and 293T cells were cultured in Dulbecco’s modified Eagle’s medium (DMEM, Sigma-Aldrich, St. Louis, MO) (final concentration of glucose in medium was 4.5 mg/l for 293T Rex cells, and 1.0 mg/l for HEK293, 293EBNA and 293T cells) supplemented with 10% heat-inactivated fetal bovine serum (Biowest LLC, Miami, FL), containing streptomycin (0.1 mg/ml, WAKO Pure Chemicals, Osaka, Japan), and penicillin G (100 U/ml, WAKO Pure Chemicals) at 37°C under 5% CO₂ in air.

**Construction of epitope-tagged expression plasmids**

RT-PCR was performed with the Super Script II kit (Invitrogen) using total mRNA prepared from HEK293 cells. cDNAs encoding SMN (NM_000344) was amplified by PCR using primer sets 5’-GAAGAAGGATCCGAGGTACCCGACCATGAGC-3’ and 5’-CTTACTCGAGAGTTTCTGGATG-3’. The PCR products were cloned into the BamHI/Xho I site of pcDNA3.1(+). The cloned cDNAs were subcloned into pcDNA3.1(+)-DAP (encoding biotinylation sequence and FLAG tag) and an additional N-terminal epitope tag 6×histidine) [29]. The DAP-tag fused SMN (HB-SMN-F) coding sequence was excised with Bgl II/Apa I and ligated into the Bgl II/Apa I site of pcDNA5/FRT for inducible expression (HB-SMN1-F-pcDNA5/FRT). cDNA encoding FOP (NM_015607) was amplified by PCR using primer sets 5’-TATAGGATCCCATGGCTGCACAGTCAGCGCCG-3’ and 5’-TATACCTCGAGACTAATCATTGGTTTCGGGATCTGT-3’ for the construction of HB-FOP-pcDNA5/FRT, or 5’-TATAGGATCCCATGGCTGCACAGTCAGCGCCG-3’ and 5’-TATACCTCGAGACTAATCATTGGTTTCGGGATCTGT-3’ for the construction of HB-SMN1-F-pcDNA5/FRT. The PCR products were cloned into the BamHI/Xho I site of pcDNA5/FRT for inducible expression, or into the Kpn I/Xho I site of pcDNA5/FRT (Invitrogen) for FLAG-FOP-pcDNA5/FRT. All cloned cDNAs were verified by DNA sequencing.

**Construction of cell lines expressing HB-SMN-For HB-FOP**

The cell lines expressing HB-SMN-For HB-FOP were established using an Flip-In T-Rex Expression System (Invitrogen). Flip-In T-Rex 293 (293T Rex) cells were cultured in a well of a 24-well plate, and in an approximately 70% conflueny, 293T Rex was transfected with 1 µl Lipofectamine2000, containing 0.25 µg pOG44 (Invitrogen), and 0.25 µg of HB-SMN1-F-pcDNA5/FRT, or HB-FOP-pcDNA5/FRT, orFLAG-FOP-pcDNA5/FRT. The clonal cells were selected in the culture medium containing 100 µg/ml hygromycin B (Invitrogen).

**Cellular fractionation and preparation of nuclear extract**

Sub-confluent 293T Rex cells expressing HB-SMN-F or HB-FOP stably, 293EBNA cells, or 293T cells were collected, lysed with buffer A (16.7 mM Tris-HCl (pH 7.4), 50 mM NaCl, 1.67 mM MgCl₂, 1 mM PMSF, 2 µg/ml aprotinin, 2 µg/ml pepstatin A and 2 µg/ml leupeptin) containing 0.1% Triton X-100, incubated for 5 min on ice, and centrifuged at 1,000×g for 5 min. The supernatant was collected as the cytoplasmic extract (Cytoplasm). To fully remove the cytoplasmic constituents, the pellet was suspended again with buffer B and centrifuged at 1,000×g for 5 min. The supernatant was collected as the nuclear pellet. To prepare the nuclear extract under low salt condition, the nuclear pellet was lysed with buffer B (50 mM Tris-HCl pH 7.4, 150 mM NaCl, 5 mM MgCl₂, 1 mM PMSF, 2 µg/ml aprotinin, 2 µg/ml pepstatin A and 2 µg/ml leupeptin) containing 0.5% IGEPAL CA-630 for 10 min at 4°C and centrifuged for 20 min.

**References**

[1] Izumikawa K, Ishikawa H, Yoshikawa H, Terukina G, Miyazawa N, et al. (2013) Friend of Prmt1, FOP is a Novel Component of the Nuclear SMN Complex Isolated Using Biotin Affinity Purification. J Proteomics Bioinform 7: 002. doi:10.4172/jpb.S7-002
at 20,000 xg. The supernatant was used as the nuclear extract under the low salt condition (Nuc: Low salt). To prepare the nuclear extract under high salt condition, the nuclear pellet was lysed with buffer C (50 mM Tris-HCl pH 7.4, 400 mM NaCl, 5 mM MgCl₂, 1 mM PMSF, 2 µg/ml aprotinin, 2 µg/ml pepstatin A and 2 µg/ml leupeptin) containing 0.5% IGEPA LC-630 for 10 min at 4°C and centrifuged for 20 min at 20,000 xg. The supernatant was used as the nuclear extract under the high salt condition (Nuc: high salt). To prepare total cell extract, the collected cells were lysed with 1 x SDS sample buffer, sonicated twice for 30 sec using a Bioruptor (Cosmo Bio, Tokyo, Japan) at its highest setting, and then centrifuged at 20,000 xg for 30 min. The supernatant was used as total cell extract (TCE). An equal aliquot of each extract was subjected to western blot analysis. The nuclear extract under the high salt condition was subjected to the following biotin affinity pull down or immune precipitation.

**Western blot analysis**

Proteins were separated by SDS-PAGE and electrophoretically transferred to a PVDF membrane (Millipore, Billerica, MA). The membrane was blocked with 5% non-fat dried skim milk in TBS for 1 h at 25°C, and incubated with the appropriate primary antibody at 4°C overnight. After washing with TBS containing 0.1% (w/v) Tween 20 (TBST) for 10min three times, the membranes were incubated with a secondary antibody conjugated with horseradish peroxidase (HRP) for 1 h, and then washed with TBST for 10 min three times. To detect biotin-tag of HB-SMN-F or HB-FOP, HRP-conjugated streptavidin (Thermo Scientific) was used. Chemiluminescent-stained proteins were detected by using LAS4000 image analyzer (Fujifilm).

**Biotin affinity purification method**

The nuclear extract under the high salt condition was prepared as described in Cellular fractionation and preparation of nuclear extract. 4 mg of the nuclear extract was incubated with 30 µl of Neutr Avidin Agarse Resins (Thermo Scientific) for 4 h at 4°C and washed five times with 1 ml of buffer C. HB-tagged protein complexes were eluted with 150 µl of Protein-RNA extraction buffer (7 M urea, 350 mM NaCl, 1% SDS, 10 mM Tris-HCl(pH8.0), 10 mM EDTA, 2% 2-mercaptoethanol) for 5 min. The eluted protein complexes containing RNAs were separated from Neutr Avidin Agarse Resins by centrifugation at 1,000 xg for 5 min at 4°C, and subjected to Phenol-chloroform extraction; namely, 150 µl of the eluates were added to 150 µl of Phenol-chloroform (pH8.0) (Ambion) and 15 µl of 3 M sodium acetate (pH5.2), and after vigorous mixing for 20 min at 25°C, the organic phase containing proteins and the aqueous phase containing RNAs were separated by centrifugation at 20,000 xg for 20 min at 4°C. Proteins from organic phase were precipitated with 700 µl of isopropanol and following centrifugation at 20,000 xg for 10 min at 4°C. RNAs from aqueous phase were collected by isopropanol precipitation. The precipitated proteins or RNAs were washed independently with 75% ethanol, dried up, and subjected to SDS-PAGE or denaturing urea PAGE, respectively. For liquid chromatography (LC)-tandem mass spectrometry (MS/MS) analysis of the HB-FOP complex, the precipitated proteins were dissolved with 5 µl of dissolution buffer (8 M Urea, 50 mM Tris-HCl (pH 8.0), and 1% 2-mercaptoethanol), mixed well, diluted to ten times with 50 µM of ammonium hydrogen carbonate and 5 mM of calcium chloride, and subjected to the digestion with 200 ng of trypsin for at 37°C for 6 hr. The resulting trypsin digest solution containing peptides adding 0.1% formic acid was analyzed using a nanoscale LC-MS/MS system.

**Protein identification by LC-MS/MS**

Pulled-down proteins were separated on a gel by SDS-PAGE; the gel was cut into some pieces, each of which was subjected to in-gel digestion with trypsin [30], and the resulting peptides were analyzed using a nanoflow LC-MS/MS system with qua- drople-time-of-flight 2 hybrid mass spectrometer (Q-Tof2, Micromass, Wythenshawe, UK) as described [31,32]. The peptide mixture was applied to a Mightsily- RP-18 (3 µm particle, Kanto Chemical, Osaka, Japan) fritless column (45 mm×0.150 mm i.d.) and separated using a 0 - 40% gradient of acetonitrile containing 0.1% formic acid over 80 min at a flow rate of 50 nl/min. Eluted peptides were sprayed directly into Q-Tof2. The peptides were detected in the MS mode to select a set of precursor ions for a data-dependent, collision-induced dissociation (CID) MS/MS analysis, and every 4 s the four ions having the greatest signal intensity were subjected to the MS/MS analysis. The MS/MS signals were acquired by MassLynx version 3.5 (Micromass, UK) and converted to text files by ProteinLynx software (Micromass). The database search was performed by MASCOT version 2.3.2 (Matrix Science Ltd., London, UK) against the UniProt human databases version SwissProt_2013_08, 540732 sequences, with the following parameters: fixed modification; carbamidemethyl (Cys), variable modifications: oxidation (Met), N-acetylation, pyroglutamine; maximum missed cleavages: 2; peptide mass tolerance: 200 ppm; MS/MS tolerance: 0.5 Da. The criteria were based on the vendor’s definitions (Matrix Science, Ltd.). Furthermore, we set more strict criteria for protein assignment: more than two peptides with p value<0.05 were considered as ‘hit’.

**Denaturing urea PAGE of RNAs**

The dried up RNA from the pulled-down protein complexes was resolved well in 100% formamide, and heated at 65°C for 5 min and cooled immediately into ice. Then one-fourth of volume of 50 mM EDTA (pH8.0) containing loading dye was added to RNAs in formamide and used as a RNA sample. RNA was separated on 8% (w/v) polyacrylamide gels containing 8 M urea and 0.5×TBE (45 mM Tris, 32.3 mM boric acid, 1.25 mM EDTA, pH 8.3) with 0.5×TBE as a running buffer, stained with SYBR Gold (Invitrogen, Carlsbad, CA) for 10 min, and visualized with LAS4000 image analyzer.

**In gel RNase digestion for LC-MS/MS analysis**

In gel RNase digestion was done principality by the method described [33]. Briefly, gel pieces containing RNA bands were excised from gels, cut into small pieces, and dried under vacuum. The gel pieces were digested with 15 µl of 2 ng/µl RNase T1, with incubation at 37°C for 1 hr. The nuclease fragments were extracted from the gel using 100 µl of RNase-free water, passed through a centrifugal filter unit with a polyvinylidene fluoride membrane (Ultrafree-MC, Millipore, Billerica, MA), and then 5 µl of 2 M triethylammonium acetate (pH 7.0) was added before LC-MS analysis.

**LC-MS/MS Apparatus for RNA Analysis**

The LC system used was essentially as described [34], consisting of a nanoflow pump (LC Assist, Tokyo, Japan) that delivers solvent to the fritless spray tip electrospray ionization column and a ReNCon gradient device. The column was prepared with a fused-silica capillary (150 µm i.d.×375 µm o.d.) using a laser puller (Sutter Instruments Co., Novato, CA) and was slurry-packed with reversed-phase material (Develosil C30-UG-3, particle size 3 µm, Nomura Chemical, Aichi, Japan) to a length of 50 mm. High voltage for ionization in negative mode (~1.4 kV) was applied to the metal union, and the eluate from RPLC was...
sprayed on-line to an LTQ-Orbitrap hybrid mass spectrometer (model XL, Thermo Fisher Scientific, San Jose, CA). Reverse phase separation of oligoribonucleotides was performed at flow rate of 100 nl/min using a 30-min linear gradient from 10 to 40% methanol in 10 mM triethylammonium acetate (pH 7.0). The mass spectrometer was operated in a data-dependent mode to automatically switch between Orbitrap-MS and linear ion trap-MS/MS acquisition as described [35]. Survey full scan MS spectra (from m/z 500 to 1,500) were acquired in the Orbitrap with resolution R=30,000 (after accumulation to a target value of 500,000 ions in the linear ion trap). The most intense ions (up to four, depending on signal intensity) were sequentially isolated for fragmentation in the linear ion trap using CID at a target value of 30,000 ions. An MS scan was accumulated for 2 s and an MS/MS scan for 3 s. The resulting fragment ions were recorded in the linear ion trap with a high-scan rate ‘Enhanced’ mode. Target ions selected for MS/MS were dynamically excluded for 60 s. General mass spectrometric conditions were as follows: electrospray voltage, 1.4 kV; no lock mass option; with sheath and auxiliary gas flow; normalized collision energy, 35% for MS/MS. Ion selection threshold was 10,000 counts for MS/MS. An activation q-value of 0.25 and activation time of 30 ms were applied for MS/MS acquisitions. LC was performed at a flow rate of 100 nl/min using a 30-min linear gradient from 10% to 40% methanol in 10 mM triethylammonium acetate (pH 7.0). The mass spectrometer was operated in a mode to automatically switch between Orbitrap-MS and linear ion trap–MS/MS acquisition as described [35].

Database search and interpretation of MS/MS Spectra of RNA

We used the software, Ariadne [36], for database searches for RNA identification. Ariadne searched DNA/RNA sequence database using MS/MS data of an RNase T1 digest of an RNA sample. Databases used were an in-house small RNA database, which was constructed from the cDNA of less than 1000 bases in NCBI GenBank (http://www.ncbi.nlm.nih.gov/genbank/), and EMBL sequence database (http://www.ebi.ac.uk/embl/), which contained more than 1,500 entries, and the genome database of Homo Sapiens (reference assembly GRCh37 obtained from ftp://ftp.ncbi.nlm.nih.gov/genomes/H_sapiens/). The following search parameters were used: the maximum number of missed cleavages was set at 1; the maximum number of variable modifications for a nucleotide was 1; The precursor mass tolerance of ± 20 ppm and MS/MS tolerance of ± 750 ppm were allowed; The significance thresholds for both the nucleotide and RNA identifications were set at 5%; The nucleotide lengths considered by the software were from 4 to 19. The original MS/MS spectra for identified nucleotides by Ariadne searches were carefully inspected by at least two MS specialists; Nucleotides with less than 50% sequence coverage were eliminated from the identifications; the length of an identified RNA estimated by PAGE and that from the database was used to confirm RNA identification. After the RNA identification, unidentified MS/MS spectra were manually inspected to identify unidentified nucleotides of the RNA such as those having lengths shorter than 3 nt or longer than 20 nt. The manual identification criteria were as follows: mass differences for nucleotides were within 20 ppm; product ions should cover more than half of the sequence of a candidate nucleotide.

Immunoprecipitation of endogenous SMN complexes

Mouse monoclonal anti-SMN antibody bound to Dynabeads were prepared by incubating 3 µg of the antibody with 15 µl of protein G Dynabeads (Invitrogen, Carlsbad, CA) in PBS containing 0.01% Triton X-100 for 30 min at 25°C, and were washed with PBS containing 0.01% Triton X-100 three times. The antibody-bound Dynabeads were incubated with 2 mg of the nuclear extract from 293EBNA cells for 4 h at 4°C and washed five times with 1 ml of buffer C. SMN binding proteins were eluted by 1X SDS sample buffer, and subjected to the following western blot analysis. For SILAC analysis, anti-SMN bound beads was made using Dynabeads Co-Immunoprecipitation Kit (Invitrogen) according to the manufacturer’s instructions. Briefly, 20 µg of anti-SMN antibody was bound to 5 mg of Dynabeads, Epoxy M-270. FLAG-tagged protein was pulled down using 15 µl of anti-FLAG M2 agarose beads (SIGMA). The cytoplasmic extract or nuclear extract was incubated with FLAG-M2 affinity Agarose Resins (SIGMA) for 2 h at 4°C. After washing five times with buffer B or buffer C, FLAG-protein complexes were eluted with 300 µg/ml of FLAG peptide or 1X SDS sample buffer, and subjected to the following SDS-PAGE and silver staining or western blot analysis.

Northern blot analysis

The separated RNAs by denaturing urea PAGE were transferred to a Hybond N+ membrane. The membrane was dried and UV cross-linked using UV-cross linker (FUNAKOSHI) at 120 ml/cm², and subsequently hybridized to biotin-labeled DNA probes at 42°C overnight in Perfect Hyb Plus hybridization buffer (Sigma) according to the manufacturer’s instructions. The hybridized membrane was washed sequentially with 2X SSC containing 0.1% SDS for 5 min at 5°C, 0.5X SSC containing 0.1% SDS for 20 min at 42°C, and 0.1X SSC containing 0.1% SDS for 20 min at 25°C. The hybridized RNA was detected using a Chemiluminescent Nucleic Acid Detection Kit (Thermo Fisher Scientific) according to the manufacturer’s instructions. RNAs were detected by using LAS4000 image analyzer and quantified by Multi Gauge software. The sequence of oligonucleotides used as probes were noted; 5'-GTATCCTCCCCCTGCGAACAGTAT-3' for U1 probe, 5'-TACCTGAAATGGCTGCATGCGT-3' for U2 probe, 5'-ATCATCAATGGCTGACGGCAGTTG-3' for U3 probe, 5'-GACTATATTGCAAGTCGTCACGGC-3' for U4 probe, 5'-GACATCAGCTTATCCCTCTTCACCG-3' for U5 probe, and 5'-ACGAATTGCGTGTCATCCTTGCG-3' for U6 probe.

Ribonuclease treatment of the pulled-down protein complexes

The pulled-down complexes bound to the agarose resins described above were incubated with 50 mM Tris-HCl pH 8.0, 150 mM NaCl, 5 mM MgCl2, containing 10 µg/ml RNase A for 30 min at 37°C, washed three times with 1 ml of buffer C and eluted with Protein-RNA extraction buffer. The eluted proteins and RNAs were separated as described above, and subjected to the following western blot analysis and northern blot analysis, respectively.

Immunofluorescence staining

The FLAG-FOP inducibly expressed cells were grown on collagen-coated culture slides and FLAG-FOP was expressed by the treatment with 100 ng/ml of doxycycline for 24 h. To visualize nuclei, the cells were washed with PBS followed by permeabilization with PBS containing 0.1% (w/v) Triton X-100 for 10 min at room temperature. After washing twice with PBST, the cells were fixed with 4% paraformaldehyde in PBS for 10 min at room temperature and washed twice with PBST. The cells were then blocked with 3% (w/v) nonfat dried milk in PBS for 1 h, and incubated with anti-FLAG rabbit polyclonal antibody (SIGMA) and anti-SMN antibody (2B1) as the primary antibodies for more than 2 h. The cells were washed three times and visualized using appropriate fluorescently labeled secondary antibodies.
times in PBST for 10 min and then incubated with FITC-conjugated anti-rabbit IgG goat antibody (American Qualex) and Cy-conjugated anti-mouse IgG sheep antibody (SIGMA) as the secondary antibodies for 1 h at room temperature. The cells were washed three times in PBST for 10 min and then mounted with VECTASHIELD (Vector Laboratories) containing 4,6-diamidino-2-phenylindole (DAPI). The resulting cells were examined with an Axiovert 200 M microscope (Carl Zeiss, Germany).

RNA interference

293T (2×10^6 cells) were cultured in 60-mm Petri dishes until they reached 70% confluency. 293T cells were transfected with 15 µl of Lipofectamine RNAiMax and 300 pmol of stealth small interfering RNA (siRNA) or control scramble RNA (scRNA).

The following stealth RNAs were used to knockdown the transcripts of FOP as noted: 5'-AUACCUUAACGAACCGGGUUGUUAGC-3' and 5'-GCUAACAAACCGGUUCGUAAGGUAU-3' for scRNA, and 5'-AAUCAUUGUUUUGCGGUGCUGUCUG-3' and 5'-AGACAGAUCGGGAAACCAUGAUU-3' for siRNA.

Quantitative MS with SILAC

293T cells were grown in SILAC DMEM (Thermo Scientific) supplemented with either "light" L-13C6,14N4 lysine-2HCl (L-Lys) and L-13C6,14N4 arginine-HCl (L-Arg) or "heavy" L-13C6,14N4 lysine-2HCl (H-Lys) and L-13C6,14N4 arginine-HCl (H-Arg). For full incorporation of the "light" or "heavy" amino acids, cells were grown for at least six passages. Subsequently, 2×10^6 cells cultured in DMEM with L-Lys and L-Arg, or H-Lys and H-Arg were transfected with FOP scRNA and siRNAs described above, respectively. After being cultured in 96 hour, each of the si/scRNA transfected cells was harvested and immunoprecipitated with anti-SMN antibody as described above. Their immunoprecipitants were eluted with 1x SDS sample buffer, respectively, combined and subjected to SDS-PAGE followed by staining with colloidal blue staining kit (Invitrogen). For quantitative MS, the gel stained with colloidal blue was cut into 16 pieces, each of which was subjected to in-gel digestion with trypsin and the resulting peptides were analyzed using a nanoflow LC-MS/MS system with LTQ-Orbitrap hybrid mass spectrometer (model XL, Thermo Fisher Scientific, CA, USA). The mass spectrometer was operated in a data-dependent mode to automatically switch between Orbitrap-MS and linear ion trap-MS/MS acquisition. Survey full scan mass spectra (from m/z 450 to 1500) acquired in the Orbitrap with resolution r=15,000 (an AGC target value of 500,000 counts). The most intense ions (top ten, depending on signal intensity) were sequentially isolated for fragmentation. The resulting fragment ions were recorded in the linear ion trap with a normal mode (Scan Rate, 16666Da/sec). General mass spectrometric conditions were as follows: electrospray voltage, 1.6 kV; normalized collision energy, 35% for MS/MS. Ion selection threshold was 10,000 counts for MS/MS. The resulting raw files were processed and analyzed using Proteome Discoverer version 1.3 (Thermo Fisher Scientific) with default parameters, except for the labels where lysine 6 and arginine 10 were selected. Searches were conducted within MASCOT version 2.3.2 against UniProt human database version SwissProt_2013_08, 540732 sequences and a reverse database, with the following parameters: fixed modification; carbamidemethyl (Cys), variable modifications: oxidation (Met), N-acetylation, pyroglutamine, K+6 and R+10; maximum missed cleavages: 2; peptide mass tolerance 15 ppm; MS/MS tolerance: 0.8 Da. We selected the candidate peptides with probability-based Mowse scores that exceed their threshold indicating a significant homology (p < 0.05), and referred to them as “hits.” The criteria were based on the vendor’s definitions. Furthermore, we set more strict criteria for protein assignment: more than two peptides with p value<0.05 were considered as ‘hit’. Protein ratios were calculated based on vendor’s default definition. Only protein groups with at least three SILAC counts were considered for further analysis.

Results

The nuclear SMN associates with Friend of Prmt1 (FOP)

Although the roles and the components of SMN complexes present in the cytoplasm have been well established, those in the nucleus are unknown except that coilin and fibrillarin were associated with SMN in the nucleus [12,13,37]. We, therefore, tried to isolate the SMN complexes present in the nucleus, and first constructed 293T-Rex cell line, which has a single-copy transgene encoding HB-SMN-F [29] (Figure 1A). The cytomegavirus promoter was used to drive stable expression of the transgene, which was integrated at a common locus in the 293TRex genome [38]. As most SMN complex population

![Figure 1: Biotin affinity purification method isolates efficiently the nuclear SMN complexes.](image-url)
existed in the cytoplasm, we first separated the HB-SMN-F expressing 293T cells into the cytoplasmic and the nuclear fractions by cell fractionation method with the lysis buffer containing 150 mM NaCl (low salt buffer). Under this low salt condition, about 80% of the SMN population in the total cell extract was recovered as the SMN complexes from the cytoplasmic fraction by a pull down method using anti-FLAG M2 antibody beads (Figures 1B and 1C) [30-32], while we detected only a small amount of the nuclear HB-SMN-F (Figure 1B). To improve the yield of the nuclear SMN, we used lysis buffer containing 400 mM NaCl (high salt buffer), and found that this high salt buffer increased the yield of the nuclear HB-SMN-F about three folds of that extracted with low salt buffer (Figure 1B). Our initial attempts to extract the nuclear SMN complex from anti-FLAG M2 antibody beads with the high salt buffer were unsuccessful (data not shown), probably because the interaction of anti-FLAG M2 antibody and the FLAG epitope was inhibited under high salt condition. Therefore, the biotin affinity purification procedure presented here is essential for efficient recoveries of the nuclear HB-SMN-F and its associated proteins (Figures 1B and 1C). The subsequent MS-based protein identification showed that the isolated nuclear SMN complexes contained Gemins 2, 3 (DDX20), 4, and 8, endogenous SMN, and fibrillarin that is reported to associate with SMN (Figure 1C and Supplementary Table 1) [39]. In addition, the MS-based method identified Friend of Prmt1, FOP, with the protein score 113 and matched sequence 2, as a potential component of the nuclear SMN complexes (Figure 1D and Supplementary Table 1).

FOP is a component of the nuclear SMN complexes that lack SmB/B’ and SmD1

We validated the results of MS-based analyses by western blotting; the method clearly detected Gemins 2, 3, 4, and 8, endogenous SMN, fibrillarin, and FOP (Figure 2A). The method also detected Gemin6 and unrip, the known components of SMN complexes, as well as collin, which is a marker of Cajal/Gems in the nuclear HB-SMN-F complexes (Figure 2A). Interestingly, SmB/B’ and SmD were absent in the isolated nuclear SMN complexes (Figure 2A). We considered that FOP is a novel de facto component of the SMN complexes present specifically in the nucleus for the additional evidences: (1) the endogenous nuclear SMN complexes prepared by immunoprecipitation using anti-SMN antibody contained FOP, as well as the several known components, Gemins 2, 3, 4, and endogenous SMN (Figure 2B); (2) the SMN complexes isolated from the nuclear fraction prepared by centrifugal cell fractionation technique contained FOP, whereas the equivalent complexes isolated from the cytoplasmic fraction did not (data not shown); (3) the pulled-down protein mixture prepared from the nuclear fraction using HB-SMN-F as affinity bait contained FOP, whereas the equivalent mixture prepared from the cytoplasmic fraction did not (Figure 2C).

We also recovered several small RNA components from the nuclear SMN complexes isolated under the high salt condition (Figure 2D), and identified the RNAs by the LC-MS/MS system equipped with a genome-oriented database searching engine Ariadne [33,35,36]. This analysis identified a number of known RNA components of the SMN complexes, U1, U3, and U4 snRNAs (Figure 2D, supplementary Table 2), showing that our preparation of the nuclear SMN complexes in fact contained UsnRNAs. However, the result suggest that the UsnRNAs are present in only small population of the isolated nuclear SMN complexes because the nuclear SMN complexes lack SmB/B’ and SmD (Figure 2A).

The FOP-associating SMN complexes do not contain U1 and U2 snRNAs

To confirm the association of FOP with the nuclear SMN complexes further, we performed a reverse pull-down-assay using FOP associating protein(s) as bait(s). We prepared the high salt nuclear extract of 293T cells expressing HB-FOP, where the DNA fragment-encoding FOP fused to HB-tag towards amino-terminus was integrated into the 293T genome (Figure 3A). Like the nuclear SMN, the expressed HB-FOP was extracted more efficiently under the high salt condition than the low salt condition (Figure 3B). This is consistent with the report of van Dijk et al. [26] which demonstrated that FOP was associated with chromatin, localized mainly in facultative heterochromatin region, and thus was extracted mostly under high salt conditions. Biotin affinity

![Figure 2](image-url)
Figure 3: FOP-associating nuclear SMN complex lacks Gemin5 and coilin as well as U1 and U2 snRNAs. (A) Schematic representation of FOP tagged with DAP lacking FLAG (HB-FOP). (B) 293TRex cells stably expressing HB-FOP (HB-FOP cells) were fractionated into cytoplasmic extract (Cyto), cytoplasm wash fraction (Wash), and nuclear extract at low salt condition (Nuc: Low salt) or at high salt condition (Nuc: high salt) (see Material and methods). They were analyzed by western blot method with the antibodies against Lamin B and GAPDH, respectively. HB-FOP was detected with HRP-conjugated streptavidin. Parental 293TRex cells were used as a control (293TRex cells). (C) HB-FOP complex was pulled down from the nuclear extract of HB-FOP cells or 293TRex cells with avidin-fixed beads (Biotin), separated by SDS-PAGE and visualized by silver staining. The protein band of HB-FOP was assigned based on its mobility expected from the amino acid sequence of HB-FOP on SDS-PAGE gel. Molecular weights of the marker proteins are indicated at left. (D) The pulled-down HB-FOP complex were analyzed by western blot with the antibodies indicated at the right. HB-FOP was detected with HRP-conjugated streptavidin. Total RNA (1 µg) was used as input. (E) The RNAs extracted from the HB-FOP complex were analyzed by northern blot with the biotin labeled DNA probes complementary to U1-U6 sn(o)RNAs, respectively. (F) The pulled-down HB-FOP complex was analyzed by western blotting with the antibodies against proteins indicated at the right with (+) or without (-) RNase A treatment. U6 snRNA was detected by northern blot analysis. (G) 293TRex cells inducibly expressing FLAG-fused-FOP (FLAG-FOP cells) were subjected to immunocytochemical analysis with the antibody against FLAG (green) or endogenous SMN (red) after the removal of the cytoplasmic proteins by permeabilization with Triton. DAPI staining (blue) shows the nucleus. Scale bar: 10 μm.
purification and MS-based analyses showed that HB-FOP was associated with SMN, Gemins 2, 3, 4, and 8, and unrip (Figure 3C and Table 1 and Supplementary Table 3). The analyses also identified Fragile X mental retardation 1 protein (FMR1), which was reported to associate with SMN [40] (Table 1, and supplementary Table 3). The other proteins identified include protein arginine N-methyltransferases 1 (Prmt1) and 5 (Prmt5) both of which were reported to be the FOP binding proteins (26), the complement component 1 Q subcomponent-binding protein (p32), lamin-B receptor, protein arginine U5 snRNP 200 kDa helicase, 116 kDa U5 snRNP component, heterogeneous nuclear RNP (hnRNP) C1/C2, hnRNP H, nuclear RNA helicase 2, ATP-dependent RNA helicase DDX39 (BAT1-associating protein), Cip1-interacting zinc finger protein, zinc finger CCCH domain-containing protein 14 and others (supplementary Table 1 and Supplementary Table 3).

Western blot analysis using available antibodies confirmed minimally the presence of SMN, Gemins 2, 3, 4, 6, and 8, unrip, p32, Prmt1, and FMR1 in the HB-FOP associating complexes (Figure 3D). The isolated HB-FOP associating complexes contained neither Sm B/B’ and Sm D, nor Gemin5 and collin (Figure 3D). HB-FOP was associated with U4, U5, and U6 snRNAs but not with U1, U2, and U3 snRNAs as revealed by northern blot analysis (Figure 3E). These data suggest that FOP is associated with SMN complexes that were separated from the major U1 and U2 snRNPs containing Sm proteins in Cajal/Gems. We treated the HB-FOP associating complexes with RNase A and found that SMN and FOP were associated with each other independently of RNA, whereas the RNase treatment reduced U6 snRNA in the complexes (Figure 3F).

In addition, we examined whether SMN and FOP were co-localized in the nucleus by immunocytochemical analysis. Because SMN mainly localizes in the cytoplasm, we stained SMN and FOP in the nucleus with their corresponding antibodies after the removal of the cytoplasmic proteins by permeabilization with 0.1% Triton, and showed that the two proteins were co-localized in several foci in the nucleus (Figure 3G), supporting their specific in vivo association.

FOP is required for the association of the nuclear SMN complexes with hnRNPs, histones and RNA-binding proteins

To gain insight into the functional relationship between FOP and the nuclear SMN complexes, we examined the effect of FOP knockdown on protein components associated with the nuclear SMN by SILAC MS analysis. We cultured scRNA- and siRNA-treated cells with light and heavy amino acids, and fractionated into the cytoplasmic and nuclear extracts by cell fractionation method, respectively. Each of the nuclear extracts was then subjected to immunoprecipitation with the nuclear extracts by cell fractionation method, respectively. Each of the nuclear extracts was then subjected to immunoprecipitation with the antibody against GAPDH and FOP. The isolated HB-FOP associating complexes with RNase A and found that SMN and FOP were associated with each other independently of RNA, whereas the RNase treatment reduced U6 snRNA in the complexes (Figure 3F). In addition, we examined whether SMN and FOP were co-localized in the nucleus by immunocytochemical analysis. Because SMN mainly localizes in the cytoplasm, we stained SMN and FOP in the nucleus with their corresponding antibodies after the removal of the cytoplasmic proteins by permeabilization with 0.1% Triton, and showed that the two proteins were co-localized in several foci in the nucleus (Figure 3G), supporting their specific in vivo association.

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### Table 1: Identification of proteins associated with the nuclear HB-FOP by LC-MS/MS analysis.

The Nuclear HB-FOP -associated proteins contain SMN complex proteins. Proteins are classified into functional groups based on SwissProt classification. Proteins were identified by Q-Tof2 as described in Supplementary Tables II. Proteins detected in mock are not listed in this table. We added GEMIN6 to the identification list by the confirmation of its association with HB-FOP using western blot analysis.

| UniProt Accession | Protein Name | Gene symbol | MW [Da] | Protein Score | Match of sequences |
|-------------------|--------------|-------------|---------|---------------|--------------------|
| **Bait**          |              |             |         |               |                    |
| CHTOP_HUMAN       | Chromatin target of PRMT1 protein | CHTOP | 26380 | 709 | 8 |
| **SMN binding proteins** |            |             |         |               |                    |
| SMN_HUMAN         | Survival motor neuron protein | SMN | 32285 | 255 | 3 |
| GEMI2_HUMAN       | Gem-associated protein 2 | GEMIN2 | 32021 | 102 | 2 |
| DDX20_HUMAN       | Probable ATP-dependent RNA helicase DDX20 | DDX20 | 92981 | 293 | 9 |
| GEMI4_HUMAN       | Gem-associated protein 4 | GEMIN4 | 121728 | 207 | 5 |
| GEMI6_HUMAN       | Gem-associated protein 6 | GEMIN6 | 18933 | 41 | 1 |
| GEMI8_HUMAN       | Gem-associated protein 8 | GEMIN8 | 28904 | 54 | 2 |
| STRAP_HUMAN       | Serine-threonine kinase receptor-associated protein | STRAP | 38756 | 121 | 4 |
| FRM1_HUMAN        | Fragile X mental retardation protein 1 | FRM1 | 71473 | 265 | 9 |
| **Protein arginine N-methyltransferases (PRMTs) and related proteins** | | | | | |
| ANM1_HUMAN        | Protein arginine N-methyltransferase 1 | PRMT1 | 42059 | 648 | 8 |
| ANMS5_HUMAN       | Protein arginine N-methyltransferase 5 | PRMT5 | 73322 | 90 | 4 |
| **Zinc finger proteins** | | | | | |
| KHDR1_HUMAN       | KH domain-containing, RNA-binding, signal transduction-associated protein 1 | KHDR1 | 48311 | 290 | 7 |
| ZC3HE_HUMAN       | Zinc finger CCCH domain-containing protein 14 | ZC3HE | 83793 | 210 | 5 |
| ZC11A_HUMAN       | Zinc finger CCCH domain-containing protein 11A | ZC11A | 89931 | 66 | 2 |
| **Transcription related proteins** | | | | | |
| CI21_HUMAN        | C21-interacting zinc finger protein | CI21 | 101066 | 396 | 8 |
| P53_HUMAN         | Cellular tumor antigen p53 | TP53 | 44196 | 216 | 8 |
| ERH_HUMAN         | Enhancer of rudimentary homolog | ERH | 12422 | 180 | 3 |
| TR150_HUMAN       | Thyroid hormone receptor-associated protein 3 | TRHAP3 | 108658 | 85 | 2 |
| SAFB2_HUMAN       | Scaffold attachment factor B2 | SAFB2 | 107921 | 78 | 3 |
| YLPM1_HUMAN       | YLP motif-containing protein 1 | YLPM1 | 220077 | 53 | 2 |
| MATR3_HUMAN       | Matrin-3 | MATR3 | 95078 | 52 | 2 |
| **snRNA binding proteins** | | | | | |
| US51_HUMAN        | 116 kDa U5 small nuclear ribonucleoprotein component | EFTUD2 | 110336 | 105 | 3 |
| RU17_HUMAN        | U1 small nuclear ribonucleoprotein 70 kDa | SNRNP70 | 51583 | 74 | 2 |
| **mRNA splicing factors** | | | | | |
| C1GBP_HUMAN       | Complement component 1 Q subcomponent-binding protein, mitochondrial | C1GBP | 31742 | 591 | 5 |
| IF4A3_HUMAN       | Eukaryotic initiation factor 4A-III | EIF4A3 | 47126 | 227 | 2 |
| PABP2_HUMAN       | Polyadenylate-binding protein 2 | PABPN1 | 32843 | 161 | 2 |
| SRSF6_HUMAN       | Serine/arginine-rich splicing factor 6 | SRSF6 | 39677 | 74 | 2 |
| **IntrRNPs and RNA helicases** | | | | | |
| HNRH1_HUMAN       | Heterogeneous nuclear ribonucleoprotein H | HNRNPH1 | 49484 | 142 | 3 |
| HNRPC_HUMAN       | Heterogeneous nuclear ribonucleoproteins C1/C2 | HNRNPC | 33707 | 143 | 3 |
| HNRPF_HUMAN       | Heterogeneous nuclear ribonucleoprotein F | HNRNPF | 45985 | 113 | 2 |
| DX39A_HUMAN       | ATP-dependent RNA helicase DDX39A | DDX39A | 49611 | 78 | 2 |
| **Chaperons & Cytoskeletons** | | | | | |
| LBR_HUMAN         | Lamin-B receptor | LBR | 71067 | 1134 | 14 |
| ACTA_HUMAN        | Actin, aortic smooth muscle | ACTA2 | 42381 | 97 | 4 |
| HSP71_HUMAN       | Heat shock 70 kDa protein 1A/1B | HSPA1A | 70294 | 743 | 13 |
| HSP7C_HUMAN       | Heat shock cognate 71 kDa protein | HSPA8 | 71082 | 371 | 9 |
| **Others**        | | | | | |
| NPM_HUMAN         | Nucleophosmin | NPM1 | 32726 | 127 | 3 |
| PNMA2_HUMAN       | Paraneoplastic antigen Ma2 | PNMA2 | 41711 | 117 | 3 |
| VIME_HUMAN        | Vimentin | VIM | 53676 | 54 | 2 |

Table 1: Identification of proteins associated with the nuclear HB-FOP by LC-MS/MS analysis. The Nuclear HB-FOP-associated proteins containing SMN complex are indicated. Proteins are classified into functional groups based on SwissProt classification. Proteins were identified by Q-ToF2 as described in Supplementary Tables II. Proteins detected in mock are not listed in this table. Uniprot accession, protein name, gene symbol as well as molecular weight are shown. We added GEMIN6 to the identification list by the confirmation of the association of GEMIN6 with HB-FOP using western blot analysis.
suggest that FOP recruits many hnRNP complexes or RNA binding proteins onto the nuclear SMN complexes. On the other hand, the knockdown increased the association of SMN complexes with ATP or GTP binding proteins including Guanine nucleotide-binding protein subunit beta-2-like 1, ADP/ATP translocase 1 etc. (Supplementary Table 4), suggesting that FOP suppresses the binding of those proteins to the SMN complexes.

Discussion

We summarized the strategy of the present study in Figure 5. In this study, we successfully isolated the nuclear SMN complexes by a combination of high salt extraction and biotin affinity purification methods, and found that FOP is a novel component of the nuclear SMN complexes (Step 1 in Figure 5). Our data suggest that the nuclear SMN complexes isolated using HB-SMN-F as affinity bait contain at least two distinct SMN complexes; one contains coilin in addition to Gemins and unrip (Figurere 2A), and another contains FOP but does not contain coilin, U1 and U2 snRNAs (Figure 3D and 3E). The former nuclear SMN complex probably corresponds to that present in Cajal/Gems, because the complex contains coilin, which is presumably required to dissociate the SMN complex from U1 and U2 snRNPs in the Cajal/Gems [12-15,17]. Our results also suggest that the use of HB-FOP as affinity bait isos preferentially the latter SMN complex(FOP-SMN complex) (Step 2 in Figure 5). In addition, the quantitative analysis of the components of the endogenous nuclear SMN complexes before and after the knockdown of FOP suggests that the FOP-SMN complex is retained on chromatin [26] possibly via the association with histones (Step 3 in Figure 5). FOP is also required for the association of SMN complexes with a number of hnRNPs and RNA binding proteins (Step 3 in Figure 5). Based on these data and the previous reports [26], we propose that FOP-SMN complex has a role in retaining spliced and/or unspliced mRNAs in near the transcription sites on chromosome. This does not conflict to the previous notion that FOP has a critical role in specific mRNA transcriptional regulation and mRNA export [27,28]. We do not exclude, however, other roles of FOP-SMN complex, such that it may prevent improperly processed mRNAs from transit through the nuclear speckle [20,21], or that FOP may keep the nuclear SMN complexes near the transcription sites on chromosome until it is used for recycling UsnRNPafter splicing [22-24].

Thus, for the first time in our knowledge, our present study identifies the FOP-SMN complexes that are separated from the mature U1 and U2 snRNPs in the nucleus, and provides its potential role in post-transcriptional gene regulation; i.e., FOP bridges between the nuclear SMN complexes and the region near the transcription sites on chromosome via the association with histones, and possibly retains spliced and/or unspliced mRNAs via the association with a number of hnRNPs and RNA-binding proteins. It has been reported that loss of Cajal/Gems is a cellular hallmark of SMA and amyotrophic lateral sclerosis (ALS) [41,42]. Since the formation of FOP-SMN complex is probably related to or coupled with the events taken place in Cajal/Gems, the detailed analysis of FOP-SMN complex may open up a novel molecular mechanism by which a common biochemical pathway leads to SMA and ALS.

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