Genetic deletion of microRNA-22 blunts the inflammatory transcriptional response to status epilepticus and exacerbates epilepsy in mice

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Abstract

MicroRNAs perform important roles in the post-transcriptional regulation of gene expression. Sequencing as well as functional studies using antisense oligonucleotides, indicate important roles for microRNAs during the development of epilepsy through targeting transcripts involved in neuronal structure, gliosis and inflammation. MicroRNA-22 (miR-22) has been reported to protect against the development of epileptogenic brain networks through suppression of neuroinflammatory signalling. Here, we used mice with a genetic deletion of miR-22 to extend these insights. Mice lacking miR-22 displayed normal behaviour and brain structure and developed similar status epilepticus after intraamygdala kainic acid compared to wildtype animals. Continuous EEG monitoring after status epilepticus revealed an accelerated and exacerbated epilepsy phenotype whereby spontaneous seizures began sooner, occurred more frequently and were of longer duration in miR-22-deficient mice. RNA sequencing analysis of the hippocampus during the period of epileptogenesis revealed a specific suppression of inflammatory signalling in the hippocampus of miR-22-deficient mice. Taken together, these findings indicate a role for miR-22 in establishing early inflammatory responses to status epilepticus. Inflammatory signalling may serve anti-epileptogenic functions and cautions the timing of anti-inflammatory interventions for the treatment of status epilepticus.

Introduction

Prolonged or repeated seizures are damaging to the brain and can establish lasting states of hyperexcitability that produce recurrent spontaneous seizures (epilepsy [1, 2]). Gene expression programmes drive many of the changes underlying network reorganisation in epileptogenesis, including neurodegeneration, astrogliosis, microgliosis, aberrant neurogenesis and restructured local and distant neuronal contacts, among other changes [3-5]. A number of post-transcriptional mechanisms fine-tune the gene expression landscape. Small noncoding RNAs called microRNAs (miRNA) negatively regulate gene expression by sequence-specific targeting of protein-coding transcripts [6]. Binding of a miRNA to a complementary sequence in a target mRNA is mediated by Argonaute proteins leading to degradation of the mRNA or translational inhibition, thereby lowering protein levels in cells [7]. Individual miRNAs are capable of regulating gene networks by interacting with multiple targets or by suppressing transcriptional controllers [6, 8]. miRNAs are essential for brain development and function, with multiple miRNAs enriched in specific cell types where they regulate differentiation, structure and neurophysiological properties [9, 10].

Expression of various miRNAs is dysregulated in experimental models of status epilepticus and in resected brain tissue from patients with drug-resistant temporal lobe epilepsy [11-13]. This is predicted to exert an important influence on the gene expression landscape. Indeed, functional studies in animals, mainly using antisense oligonucleotides (ASOs) termed antagonirs, have demonstrated that targeting miRNAs can alter evoked and spontaneous seizures and neuropathological outcomes including neuronal loss and gliosis [11]. miR-22-3p (hereafter miR-22), a conserved miRNA that is expressed throughout the body, including the brain [14, 15], was previously identified among upregulated miRNAs within the mouse hippocampus contralateral to the epileptogenic zone in the intraamygdala kainic acid model of status epilepticus [16]. Inhibiting miR-22 using intracerebroventricular injection of an ASO increased the frequency of spontaneous seizures, indicating a protective role in the model [16]. Inhibition of miR-22 was also associated with increased astrogliosis [16]. The P2X7 receptor was identified as a miR-22 target and co-injection of a P2X7 receptor antagonist mitigated the epilepsy phenotype in mice given the ASO inhibitor of miR-22 [16]. Over-expression of miR-22 has been reported to protect in models of brain injury and additional targets of miR-22 have been identified [17, 18].

ASOs produce only transient and incomplete knockdown of RNAs and have the potential to modulate targets with closely-related sequences [19]. Accordingly, the role of miR-22 in epileptogenesis remains incompletely resolved. Mice with a genetic ablation of miR-22 (miR-22−/−) were recently developed [20]. The mice were viable and developed normally but displayed increased sensitivity to haemodynamic stress, consistent with roles for miR-22 in the heart [20]. Accordingly, we sought to evaluate epilepsy development after status epilepticus in
mice lacking miR-22. We characterised the morphology of the hippocampus, basic behaviour, tracked the development of epilepsy using continuous electroencephalogram (EEG) recordings and explored how miR-22 ablation affected gene expression. Our findings confirm that miR-22 serves an important role in protecting against epileptogenic injury from status epilepticus and reveal an unexpected role in promoting an inflammatory transcriptional landscape.

## Results

### Characterisation of mice lacking miR-22

We first analysed the expression of miR-22 and other miRNAs in the hippocampus of wildtype, heterozygous (miR-22\(^{+/-}\)) and knockout (miR-22\(^{-/-}\)) mice at three months of age. This confirmed a dose-dependent reduction in miR-22 levels consistent with heterozygous and full (homozygous) deletion of miR-22 (Figure 1A). Since miR-22 is expressed in both neurons and glia [16], we explored whether loss of miR-22 results in compensatory changes to other miRNAs enriched in these cell types. Analysis of hippocampal levels of a selection of miRNAs including neuronal (miR-124-3p, miR-134-5p), astrocyte (miR-146a-5p, miR-29a) and microglial (miR-342, miR-150) miRNAs associated with roles in epilepsy [21-23] revealed normal levels of all tested miRNAs in the hippocampus of mice lacking miR-22 (Figure 1B-F and Supplementary Data S1A). Analysis of the hippocampus from nine months old miR-22\(^{+/-}\) and miR-22\(^{-/-}\) mice also revealed no differences in levels of these miRNAs compared to wildtype (Supplementary Data S1B-G). Thus, loss of miR-22 does not result in compensatory changes to these cell-enriched miRNAs.

We also investigated the behaviour of miR-22-deficient mice. Mice lacking miR-22 did not display any locomotor impairments in the Rotarod test (Figure 1G). Furthermore, miR-22\(^{-/-}\) mice performed similarly to wildtype mice in the elevated plus maze, an assay for anxiety-like behaviour (Figure 1H). Sub-group analysis of male and female mice revealed there were no significant sex differences in any of these tests.

Analysis of hippocampal tissue sections stained for Nissl (Figure 2A) or the neuronal marker NeuN (Figure 2B) revealed normal gross brain structure in three months old mice lacking miR-22. Staining and counts of glial fibrillary acidic protein (GFAP) positive cells, a marker of astrocytes (Figure 2C), and Iba1, a marker of microglia (Figure 2D), were normal in miR-22\(^{-/-}\) mice (and Supplementary Data S2). The morphology of these cells was not quantified. Staining and counts of parvalbumin inhibitory interneurons did not differ between genotypes (Figure 2E and Supplementary Data S2). Inspection of brains from older, nine months old heterozygous and homozygous miR-22-deficient mice also revealed no differences in staining of the same neuronal and glial histological markers (Supplementary Data S3).

### Status epilepticus in mice lacking miR-22\(^{-/-}\)

Wildtype, miR-22\(^{+/-}\) and miR-22\(^{-/-}\) mice were next equipped for acute EEG recordings to assess seizure severity during status epilepticus triggered by intraamygdala kainic acid. This is a well-characterised model of prolonged seizures that recruits limbic circuits and causes damage to the ipsilateral hippocampus. Mice develop spontaneous recurrent seizures after a short latency period of 3 – 5 days [24].

Status epilepticus after intraamygdala microinjection of kainic acid was similar between wildtype, miR-22\(^{+/-}\) and miR-22\(^{-/-}\) mice (Figure 3A). Analysis of electrographic seizure activity and EEG parameters over the period between kainic acid and administration of anticonvulsant (to curtail seizures and reduce morbidity and mortality) revealed no difference amongst the groups or between sexes for EEG amplitude and total power (Figure 3B, C). EEG parameters (amplitude and total power) also remained similar between groups after anticonvulsant administration (Figure 3D, E).

Analysis of brains obtained 72 hours after status epilepticus revealed similar neuronal damage and glial responses between wildtype, miR-22\(^{+/-}\) and miR-22\(^{-/-}\) mice (Figure 3F-I and Supplementary Data S4). This included loss of neurons within the ipsilateral CA3 subfield of the hippocampus (Figure 3F and Supplementary
Data S4A). Staining for gliosis using GFAP and Iba1 was also similar between groups (Figure 3G, H and Supplementary Data S4B, C). Parvalbumin-positive interneuron staining was also not different between genotypes (Figure 3I and Supplementary Data S4D). Neuron counts within the injected amygdala were also similar between genotypes (Supplementary Data S4E). Taken together, these results indicate that genetic deletion of one or both copies of miR-22 did not affect acute electrographic and histopathological outcomes of status epilepticus in mice.

**Accelerated and exacerbated epilepsy phenotype in mice lacking miR-22**

Next, wildtype and miR-22 mice were equipped with implantable EEG telemetry units to record the long-term consequences of miR-22 deletion. Heterozygous mice were not included in further studies. After recovery from the surgery and induction of status epilepticus, mice were monitored 24 hours per day for 14 days after status epilepticus.

All mice lacking miR-22 developed their first spontaneous seizures on the second day after status epilepticus, compared to wildtype mice of which only a single animal presented their first spontaneous seizure on the second day (Figure 4A). All other wildtype mice developed their first spontaneous seizures from the third day onwards, consistent with the known course of epilepsy in the model [24] (Figure 4A). A zero-inflated Poisson regression model was used to analyse the incidence of seizures. Statistical analysis showed that the incidence rate ratio of seizures was five times higher in the miR-22 mice compared to wildtype animals. The average number of seizures per day in wildtype mice was 2.47 (range 1-31). In contrast, the average number of seizures per day in miR-22 mice was 16 (range 1-138) (Figure 4A, B). Statistical analysis also showed that miR-22 mice presented on average significantly longer seizures (18.4 seconds) compared to wildtype mice (11.9 seconds) (Figure 4C, E). Furthermore, as expected, the total number of seizures in miR-22 over the 14 days period was higher when compared to wildtype mice (Figure 4D). Notably, however, this higher rate occurred largely during the first 10 days of monitoring, with rates during the last few days similar between mice.

Analysis of brain tissue sections after epilepsy monitoring revealed similar overall histological changes. This included a macroscopic lesion comprising neuron loss and gliosis within the ipsilateral CA3 subfield (Figure 5A, B). Counts of GFAP (Figure 5C, D) and parvalbumin-positive cells (Figure 5G, H) were similar between wildtype and miR-22 mice (and Supplementary Data S5). In contrast, there was a small but significantly higher number of Iba1-stained microglia in miR-22 mice at the end of monitoring (Figure 5E, F and Supplementary Data S5).

**RNA sequencing analysis of miR-22 reveals down-regulation of inflammatory signalling pathways during epileptogenesis**

Last, we sought insights into the molecular mechanism underlying the exacerbated epilepsy phenotype in miR-22 mice. Previous studies identified the P2X7 receptor as a key target of miR-22, indicating a role in suppressing inflammation [16]. RNA sequencing was performed to identify dysregulated protein-coding mRNAs within the hippocampus 24 h after status epilepticus in wildtype and miR-22 mice. This captures the gene expression landscape immediately before the emergence of the exacerbated epilepsy phenotype. We prioritised our bioinformatics search on mRNAs with shared functions and displaying small fold changes relevant to how miRNAs affect levels of their targets, to identify biologically relevant pathways dysregulated by loss of miR-22 [9] [8]. This revealed a set of 18 genes in inflammatory signalling-enriched pathways that exhibited small differences in gene expression levels between the wildtype and miR-22 mice after status epilepticus. Remarkably, the transcript levels of all the genes were downregulated in the miR-22-deficient mice (Figure 6A). A number of significantly enriched gene ontology (GO) terms of transcription factors regulating all these down-regulated immune-associated transcripts is shown in Figure 6B, including 93 transcription repressors and corepressors. Among these (co)-repressors, 25 (27%) of them are putative targets of miR-22. There was an extensive overlap among the immune-associated targets of these transcriptional (co)-repressors (Figure 6C), indicating these immune-associated genes are tightly controlled by miR-22 through the simultaneous manipulation of multiple transcriptional (co)-factors. However, expression of the individual repressors was not statistically different in miR-22 deficient mice, perhaps due to their low expression abundance and/or small magnitude of changes in expression levels (Figure 6D).
Here we show that genetic ablation of miR-22 results in a temporally-restricted, accelerated and exacerbated epilepsy phenotype in a mouse model of status epilepticus. Unexpectedly, the transcriptional landscape generated by loss of miR-22 featured extensive downregulation of inflammatory signalling pathways. These findings suggest miR-22 may activate as well as suppress neuroinflammation. Together, this extends the evidence for miRNA roles in the pathogenesis of epilepsy, imposes limits on the therapeutic potential of targeting miR-22, and cautions the delivery or timing of anti-inflammatory interventions for the treatment of status epilepticus.

There is increasing evidence that miRNAs regulate the gene expression landscape during and after epileptogenic injuries [11, 13]. Here, we focused on miR-22 which is an abundant miRNA expressed in multiple tissues and cell types. Previous studies suggested miR-22 is protective in models of brain injury [17, 18], and ASO targeting demonstrated that inhibiting miR-22 exacerbated epilepsy, an effect due in part to de-repression of the P2X7 receptor [16]. Targeting using ASOs, however, produces only incomplete and temporary knockdown of miRNAs, leaving questions about the significance and potential therapeutic value of this miRNA for epilepsy. We addressed this limitation by studying the effect of genetic deletion of miR-22 on epilepsy phenotypes and molecular outcomes of status epilepticus.

The main finding in the present study was that genetic ablation of miR-22 results in exacerbated epilepsy in mice. Specifically, spontaneous seizures emerged earlier, occurred more frequently and were longer in animals lacking miR-22 in the intraamygdala kainic acid model. We can exclude this being due to a difference in the initial precipitating injury because, consistent with targeting miR-22 using ASOs [16], genetic deletion of miR-22 did not affect the status epilepticus. This is the first report showing deletion of a single miRNA in a model of status epilepticus results in an altered epilepsy phenotype. The finding supports a role for miR-22 in suppressing hyper-excitability in response to an epilepsy-precipitating injury [16, 25]. The accelerated and exacerbated phenotype in miR-22−/− mice matches the overall findings from studies that used ASOs to reduce miR-22 levels in the same model [16]. However, the number of seizures recorded in miR-22-deficient mice in the present study is approximately double the number in the ASO study. This likely reflects the partial and transient reduction in miR-22 levels achieved using ASOs and is evidence of a gene dose-dependent effect of the loss of miR-22 accelerating and exacerbating the epilepsy phenotype. It is noteworthy that the exacerbated epileptic phenotype in miR-22−/− mice was time-restricted and did not persist for the full two-week recordings. This indicates the most important influence of miR-22 in this model is during the epileptogenic phase or during the emergence of first spontaneous seizures. The seizure rates in miR-22−/− mice appeared to normalise later in the second week of recordings. Future studies could explore if this is permanent and due to a compensatory change within the hippocampus of miR-22−/− mice. For example, via neurophysiological adjustments, which could be assessed by molecular or electrophysiological studies around the time when seizure rates normalise. Since spontaneous seizures were more frequent and lasted longer in miR-22-deficient mice, miR-22 may have targets that influence icetogenesis and seizure termination, the mechanisms of which remain incompletely understood [26]. More broadly, the findings may be clinically relevant since miR-22 is expressed within the hippocampus of patients with drug-resistant epilepsy [25].

The observation that genetic deletion of miR-22 does not affect susceptibility to status epilepticus is consistent with our observation that miR-22−/− mice lack any obvious brain abnormalities and the known viability of the mice from their original description [20]. Indeed, neuronal and glial numbers and distribution appeared normal in naïve miR-22−/− mice and animals behaved normally. Thus, despite its relative abundance, miR-22 is probably dispensable for normal brain organisation. This contrasts with the overt brain phenotypes of mice lacking certain other brain-enriched miRNAs which include gross brain development abnormalities, neuronal migration defects and epilepsy [27, 28]. The result is consistent with ASO targeting of miR-22 which did not alter the duration or severity of status epilepticus in the same model [16]. Histopathological outcomes after status epilepticus including neuronal loss and gliosis were also equivalent between miR-22−/− and wildtype mice and only a modest
difference in microgliosis was observed in the hippocampus at the end of epilepsy monitoring. This is consistent with animals having experienced an equivalent status epilepticus and with the similar final rates of spontaneous seizures between groups at the end of recordings. These findings differ somewhat from those reported when miR-22 was blocked using an ASO [16]. In that study, increased spontaneous seizures occurred later and in association with increased astrogliosis. This discrepancy may arise from cellular responses to transient, uneven and incomplete reduction of miR-22 levels using ASOs compared to complete deletion in the knockout mice. As miR-22⁻/⁻ mice experienced more spontaneous seizures than ASO-treated mice, microglial rather than astroglial increases may be a cellular correlate of the cumulative or overall burden of spontaneous seizures in this model. This would be consistent with microglia responses and recent seizures in humans [29], the use of microglia-based brain imaging as epilepsy biomarkers [30, 31] and experimental evidence that microglia changes are epileptogenic [32].

The present study used RNA sequencing to define the transcriptional landscape that may be driving the altered epilepsy phenotype in mice lacking miR-22. A number of known miR-22 targets have been linked to epilepsy, including the purinergic P2X7 receptor that drives neuroinflammation through releasing IL1β [16]. Unexpectedly, RNA sequencing analysis of the hippocampus revealed that the gene expression landscape of mice lacking miR-22 is enriched with downregulated transcripts linked to inflammation. This included multiple transcripts involved in interleukin, cytokine, TNF and NF-kB signalling, all contributing to the regulation of neuroinflammatory responses. This finding reveals a more complex and nuanced function of this miRNA, regulating the outcome of status epilepticus by both strongly suppressing individual pro-inflammatory targets such as the P2X7 receptor as well as smaller effects across multiple transcripts that promote inflammation. This is consistent with the dual and opposing roles that have emerged for other brain enriched miRNAs [22] and updates concepts of how miRNAs regulate brain excitability and epilepsy [11].

Why did we see extensive down-regulation of inflammatory transcripts in miR-22⁻/⁻ mice? Because miRNAs largely function as negative regulators of gene expression [6, 8], loss of a miRNA will typically de-repress and upregulate targets. The RNA sequencing findings can be explained if one or more miR-22 target is a transcriptional repressor which upon de-repression by deletion of miR-22 then shuts down genes under its control. Our bioinformatics analysis identified multiple potential transcriptional repressors suggesting that the downregulation of inflammatory pathways could be mediated by de-repression of a transcriptional silencer. Notably, transcriptional silencers of inflammation have previously been identified for miR-22 including Hdac4 [33]. Although Hdac4 was not found among the increased genes in miR-22⁻/⁻ mice, the present study identified several other plausible candidates. Future studies will be needed to identify which such miR-22 targets target the transcriptional control elements of the down-regulated gene networks and whether this is causally involved epileptogenesis.

The present findings challenge the pathogenicity of neuroinflammation in epileptogenesis. Certainly, prolonged or excessive cytokine signalling and neuroinflammation is a causal mechanism in the development and maintenance of the epileptic state [5]. But if epilepsy develops earlier and is more severe in mice with a subdued inflammatory transcriptional landscape then the neuroinflammatory response to status epilepticus, at least during the early period, is not maladaptive or pro-epileptogenic. This supports other evidence that early coordinated inflammatory signalling is a necessary and protective response to epileptogenic brain insults [5, 34]. Inflammatory signalling serves various protective functions including clearance of cellular debris after injury and prompting glial responses that contribute to repair [35]. This may complicate therapeutic strategies. Treatment with anti-inflammatory drugs, if mistimed or aligned with specific early phases, could lead to exacerbation of epileptogenesis. The finding that loss of miR-22 results in an accelerated epileptogenesis suggests delivery or over-expression of miR-22 might be a potential therapeutic strategy. This could be achieved using miRNA mimics, synthetic double-stranded molecules or other approaches [17, 18]. Notably, intracerebral injection of mimics for miR-22 and other inflammation-linked miRNAs has been reported to reduce seizures in the present model [16, 36].

There are caveats to consider in the present study. Foremost, we used a constitutive knockout mouse to study the effects of miR-22 deletion [20]. This is limiting because miR-22 is expressed in multiple cell types in the brain and the amounts of miR-22 expressed by cells differ between neurons and glia [16]. It will be important to
understand whether the pathways controlled by miR-22 during epileptogenesis are cell type-specific. That is, does miR-22 exert its inflammatory actions in all or specific cells? The use of inducible knockout lines would allow the cell-specific and timed deletion of miR-22 from neurons and glia. While we did not observe differences in status epilepticus in the intraamygdala kainic acid model, miR-22-deficient mice may display altered vulnerability to other chemoconvulsants or epileptogenic injuries. Moreover, neuronal microstructure or subtle cellular dysorganisation may have been missed presently and recent work has shown that inhibiting miR-22 after status epilepticus produces changes to dendritic branching patterns in new hippocampal neurons [25]. Finally, a limit of the present study is that we did not perform a recovery experiment. It would be interesting in future studies to determine if the knockout phenotype we observed can be recovered or obviated by the reintroduction of miR-22. For example, testing if delivery of a mimic [16, 36] increases early inflammatory signalling and reduces the excess of spontaneous seizures in miR-22−/− mice subject to status epilepticus.

In summary, the present study extends the evidence that miR-22 serves an important role in countering the epileptogenic process triggered by status epilepticus. Genetic deletion of miR-22 results in a temporally-restricted, accelerated and exacerbated epilepsy phenotype in the intraamygdala kainic acid model of status epilepticus and is associated with large-scale reduction in the transcriptional inflammatory response. The findings indicate early, protective effects of inflammatory responses during epileptogenesis and suggest miR-22 or the pathways under its control could be potential targets for the development of disease-modifying treatments.

**Materials And Methods**

**Breeding and identification of miR-22 knockout mice:**

Mice carrying a knockout (null) mutation of mir-22 were purchased from the Jackson laboratory (Mir22tm1.1Arod, Stock No: 018155) [37]. Mice were mated in trios (one male and two females). For breeding, either a single heterozygous male was mated with two homozygous mutant female mice or a single homozygous male mated with two heterozygous females. Wild type mice were also mated in trios. Mice were housed in a climate-controlled biomedical facility on a 12-hour light/dark cycle with food and water provided ad libitum. Primers for genotyping were (Forward (common), 5’ TGG GAC TTG GGT TCT ACA CC 3’; Reverse (wildtype), 5’ TCC TAA AAG GAA GGG GAG GA 3’; Reverse (miR-22 mutant), 5’ TGC TTT AGG TGG AGG GAA AG 3’.

**Status epilepticus and epilepsy monitoring:**

Status epilepticus was induced by unilateral intraamygdala microinjection of kainic acid, as described [16, 24]. Briefly, animals were anesthetized with isoflurane, placed in a stereotaxic frame and maintained normothermic by means of a feedback-controlled heat blanket. A craniotomy was performed and a guide cannula positioned over the surface of the dura. For acute EEG recordings, mice were equipped with three skull-mounted screw electrodes (Bilaney Consultants, U.K.) and connected to a Grass 40 channel lab-based digital EEG. For long-term epilepsy monitoring, mice were equipped with an EEG telemetry device (Model: F20-EET, Data Systems International) inserted subcutaneously and connected to skull screws. After baseline EEG recordings, a cannula was inserted to inject kainic acid (0.3 μg/0.2 μl in phosphate-buffered saline; PBS) or PBS into the basolateral amygdala nucleus. After 40 minutes, all mice received an intraperitoneal injection of lorazepam (8 mg/kg) to curtail seizures and reduce morbidity and mortality. EEG recordings during acute status epilepticus were analysed as before [23], including total power, amplitude and number of electrographic seizures. For long-term EEG monitoring, the number and duration of spontaneous seizures were manually scored, with epileptic seizures being defined as high-amplitude (>2X baseline) polyspike discharges of ≥10 seconds duration [16]. Mice were killed by pentobarbital overdose and perfused with PBS. Brains were then either microdissected and frozen or retained whole for sectioning.

**Rotarod and elevated plus maze**
The rotarod test was used to assess motor coordination and balance. After a habituation trial day, mice were placed on a horizontally oriented, rotating suspended cylinder and the length of time that mice stayed on the rotating rod was measured during a gradual acceleration from 0 to 40 rpm over 120 seconds. The average time each mouse spent on the accelerating rotating rod was calculated. The elevated plus maze apparatus comprised two open arms (25 cm length x 5 cm width) and two closed arms (25 cm length x 5 cm width x 16 cm height walls to enclose the arms) placed in the centre of the testing room, 50 cm above the floor. The test consisted of a single trial lasting five minutes. Mice were individually placed in the centre facing one of the open arms. The number of entries into the open and closed arms and the time spent was recorded. These parameters were used to calculate the index of anxiety-like behaviour.

**Histopathology**

Immunohistochemistry was performed as described [16], using post-fixed fresh frozen or paraformaldehyde-perfused sections. Sections were blocked in goat serum followed by incubation with primary antibodies against NeuN, glial fibrillary acidic protein (GFAP), ionized calcium binding adaptor molecule 1 (Iba1) or parvalbumin (all from Cell Signaling Technology). Negative controls were included that omitted the primary antibody. Slides were washed then incubated with secondary antibodies, rinsed again and mounted. Positive cells were quantified and were the average of two adjacent sections. Neuronal damage was assessed based on a graded five-point system under 40X magnification: level 0, no damage; level 1, 0–25%; level 2, 25–50%; level 3, 50–75%; level 5, 75% cell loss.

**Analysis of miRNA expression**

Expression of individual miRNAs was performed using Taqman individual miRNA assays as described [16]. Levels of miR-22 and a selection of miRNAs enriched in neurons (miR-124a, miR-134), microglia (miR-342, miR-150) and astrocytes (miR-146a, miR-29a) was based on in situ hybridisation and in vitro cell culture studies [21-23, 38]. Five microliters of RNA (500 ng/µl) was mixed with RT buffer, RNAase inhibitor, multiscribe reverse transcriptase enzyme, dNTPs and the microRNA specific RT primer (Applied Biosystems) and placed in a thermocycler. Tested miRNAs were: miR-124a, miR-134, miR-150, miR-29a, miR-146a, miR-342. The generated cDNA was diluted, mixed with TaqMan Fast Universal PCR Master Mix and miRNA-specific PCR primers and ran in triplicate and analysed using the QuantStudio™ 12K Flex PCR system as described [16].

**RNA sequencing preparation and analysis**

Wildtype and miR-22-/- mice (n = 4 each) were subjected to status epilepticus as described above. Twenty-four hours later, mice were euthanised and the ipsilateral hippocampus was obtained. Total RNA was extracted and library preparation was performed using an Illumina TruSeq Stranded mRNA Sample Prep Kit (poly-A enrichment). The generated RNA-Seq data were deposited in the Sequence Read Archive under GEO accession number GSE147466. Follow-up analysis including QC, alignment (reference genome GRCh38, Annotation_Ensembl70), mapping and raw analysis was performed by a service provider (Exiqon A/S, Vedbaek, Denmark). On average 41.3 million reads were obtained for each sample and the average genome mapping rate was 94.4%. Numbers of identified genes ranged from 21,260 to 22,315.

To reflect the mostly subtle effects of miRNAs on gene expression, only genes with small fold changes (less than two times, p < 0.01) were included in the pathway enrichment analysis. A total of 67 genes were searched against Reactome [39] and KEGG databases [40] using ReactomePA [41] and EnrichR [42]. Genes involved in immune-associated pathways of the top five most significantly (p < 0.05) enriched pathways from each analysis were selected and combined to analyse further. Potential transcription factors for these genes were retrieved from Tf2DNA database [43].

To identify targets of mmu-miR-22, experimentally validated targets were retrieved from miRTarBase Release 7.0[44], TarBase v.8[45] and miRecords[46] while predicted targets were retrieved from TargetScan Release 7.2 [47] and miRDB Version 6.0 [48] and processed as described previously [49], with some modifications. Briefly, prediction scores of TargetScan targets were rescaled between 0 and 1 while those of miRDB targets were rescaled between 0.5 and 1 (since original miRDB database excluded all targets with scores < 50). Then, any targets with rescaled prediction scores < 0.5 were removed from further analysis. All data processing, analyses
and visualisation were performed using R and bash scripts in Unix working environment.

**Data analysis**: Data are presented as mean ± SEM. Comparison of data was performed using ANOVA with appropriate post hoc test. EEG variables (e.g., total power, frequency) were analysed using the parametric Student's t-test, considering $P < 0.05$ as significant. For the long-term experiments, Zero-inflated Poisson regression was used to compare daily seizure counts between treatment groups. Robust variance estimation was used to correct for the effects of measurements within animals. Significance was accepted when $p < 0.05$.

**Supplementary Information**

Supplementary information accompanies this paper (see Supplementary File).

**Abbreviations**

ANOVA, Analysis of variance; ASO, Antisense oligonucleotide; EEG, Electroencephalogram; GFAP, Glial fibrillary acidic protein; GO, Gene ontology; PCR, Polymerase chain reaction; miRNA, MicroRNA; RNA, Ribonucleic acid; SEM, standard error of the mean; wt, wildtype

**Declarations**

**Ethics approval**

All procedures were performed in accordance with the guidelines of the Directive 2010/63/EU of the European Parliament and Council and the principles of 3R’s. All animal procedures were reviewed and prior approved by the Research Ethics Committee of the Royal College of Surgeons in Ireland (REC #842) under project license from the Health Products Regulatory Authority (HPRA), Dublin, Ireland (AE19127/P001).

**Consent for publication**

Not applicable

**Availability of data and materials**

The dataset(s) supporting the conclusions of this article are included within the article and additional supplementary files and in the GEO submission.

**Competing interests**

The authors declare that they have no competing interests.

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**Authors’ contributions**
LFAS, CRR and DCH conceived and designed the study. LFAS, CRR performed experiments in mice with assistance from MF, FRR, RRG, TE, ASR and EL. LFAS performed histology with assistance from EL. NTN performed bioinformatics analysis. GPB performed molecular studies. RMC performed statistical analysis. DCH wrote the manuscript. All authors approved the final version of the manuscript.

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References

1. Dingledine R, Varvel NH, Dudek FE: When and how do seizures kill neurons, and is cell death relevant to epileptogenesis? Adv Exp Med Biol 2014, 813:109-122.
2. Goldberg EM, Coulter DA: Mechanisms of epileptogenesis: a convergence on neural circuit dysfunction. Nat Rev Neurosci 2013, 14:337-349.
3. Johnson MR, Behmoaras J, Bottolo L, Krishnan ML, Pernhorst K, Santoscoy PL, Rossetti T, Speed D, Srivastava PK, Chadeau-Hyam M, Hajji N, Dabrowska A, Rotival M, Razzaghi B, Kovac S, Wanisch K, Grillo FW, Slaviero A, Langley SR, Shkura K, Roncon P, De T, Mattheisen M, Niehusmann P, O'Brien TJ, Petrovski S, von Lehe M, Hoffmann P, Eriksson J, Coffey AJ, Cichon S, Walker M, Simonato M, Danis B, Mazzuferi M, Foerch P, Schoch S, De Paola V, Kaminski RM, Cunliffe VT, Becker AJ, Petretto E: Systems genetics identifies Sestrin 3 as a regulator of a proconvulsant gene network in human epileptic hippocampus. Nat Commun 2015, 6:6031.
4. Pitkanen A, Lukasiuk K: Mechanisms of epileptogenesis and potential treatment targets. Lancet Neurol 2011, 10:173-186.
5. Vezzani A, Balosso S, Ravizza T: Neuroinflammatory pathways as treatment targets and biomarkers in epilepsy. Nat Rev Neurol 2019, 15:459-472.
6. Bartel DP: Metazoan MicroRNAs. Cell 2018, 173:20-51.
7. Gebert LFR, MacRae IJ: Regulation of microRNA function in animals. Nat Rev Mol Cell Biol 2018.
8. Ebert MS, Sharp PA: Roles for microRNAs in conferring robustness to biological processes. Cell 2012, 149:515-524.
9. Lim LP, Lau NC, Garret-Engele P, Grimson A, Schelter JM, Castle J, Bartel DP, Linsley PS, Johnson JM: Microarray analysis shows that some microRNAs downregulate large numbers of target mRNAs. Nature 2005, 433:769-773.
10. Nowakowski TJ, Rani N, Golkaram M, Zhou HR, Alvarado B, Huch K, West JA, Leyrat A, Pollen AA, Kriegstein AR, Petzdold LR, Kosik KS: Regulation of cell-type-specific transcriptomes by microRNA networks during human brain development. Nat Neurosci 2018, 21:1784-1792.
11. Henshall DC, Hamer HM, Pasterkamp RJ, Goldstein DB, Kjems J, Prehn JH, Schorge S, Lamottke K, Rosenow F: MicroRNAs in epilepsy: pathophysiology and clinical utility. Lancet Neurol 2016, 15:1368-1376.
12. Srivastava PK, Roncon P, Lukasiuk K, Gorter JA, Aronica E, Pitkanen A, Petretto E, Johnson MR, Simonato M: Meta-analysis of microRNAs dysregulated in the hippocampal dentate gyrus of animal models of epilepsy. eNeuro 2017, 4.
13. Lovisari F, Simonato M: Gene networks and microRNAs: Promises and challenges for treating epilepsies and their comorbidities. Epilepsy Behav 2019:106488.
14. Lagos-Quintana M, Rauhut R, Yalcin A, Meyer J, Lendeckel W, Tuschl T: Identification of tissue-specific microRNAs from mouse. Curr Biol 2002, 12:735-739.
15. Ludwig N, Leidinger P, Becker K, Backes C, Fehlmann T, Pallasch C, Rheinheimer S, Meder B, Stahler C, Meese E, Keller A: Distribution of miRNA expression across human tissues. Nucleic Acids Res 2016, 44:3865-3877.
16. Jimenez-Mateos EM, Arribas-Blazquez M, Sanz-Rodriguez A, Concannon C, Olivos-Ore LA, Reschke CR, Mooney CM, Mooney C, Lugara E, Morgan J, Langa E, Jimenez-Pacheco A, Silva LF, Mesuret G, Boison D, Miras-Portugal MT, Letavic M, Artalejo AR, Bhattacharya A, Diaz-Hernandez M, Henshall DC, Engel T:
microRNA targeting of the P2X7 purinoceptor opposes a contralateral epileptogenic focus in the hippocampus. *Sci Rep* 2015, 5:17486.

17. Jovicic A, Zaldivar Jolissaint JF, Moser R, Silva Santos Mde F, Luthi-Carter R: MicroRNA-22 (miR-22) overexpression is neuroprotective via general anti-apoptotic effects and may also target specific Huntington's disease-related mechanisms. *PLoS One* 2013, 8:e54222.

18. Yu H, Wu M, Zhao P, Huang Y, Wang W, Yin W: Neuroprotective effects of viral overexpression of microRNA-22 in rat and cell models of cerebral ischemia-reperfusion injury. *J Cell Biochem* 2015, 116:233-241.

19. Levin AA:Treating Disease at the RNA Level with Oligonucleotides. *N Engl J Med* 2019, 380:57-70.

20. Gurha P, Abreu-Goodger C, Wang T, Ramirez MO, Drumond AL, van Dongen S, Chen Y, Bartonicek N, Enright AJ, Lee B, Kelm RJ, Jr., Reddy AK, Taffet GE, Bradley A, Wehrens XH, Entman ML, Rodriguez A: Targeted deletion of microRNA-22 promotes stress-induced cardiac dilation and contractile dysfunction. *Circulation* 2012, 125:2751-2761.

21. Aronica E, Fliuter K, Iyer A, Zurolo E, Vreijling J, van Vliet EA, Baayen JC, Gorter JA: Expression pattern of miR-146a, an inflammation-associated microRNA, in experimental and human temporal lobe epilepsy. *Eur J Neurosci* 2010, 31:1100-1107.

22. Brennan GP, Dey D, Chen Y, Patterson KP, Magnetta EJ, Hall AM, Dube CM, Mei YT, Baram TZ: Dual and opposing roles of microRNA-124 in epilepsy are mediated through inflammatory and NRSF-dependent gene networks. *Cell Rep* 2016, 14:2402-2412.

23. Jimenez-Mateos EM, Engel T, Merino-Serras P, McKiernan RC, Tanaka K, Mouri G, Sano T, O'Tuathaigh C, Waddington JL, Prenter S, Delanty N, Farrell MA, O'Brien DF, Conroy RM, Stallings RL, Defelipe J, Henshall DC: Silencing microRNA-134 produces neuroprotective and prolonged seizure-suppressive effects. *Nat Med* 2012, 18:1087-1094.

24. Mouri G, Jimenez-Mateos E, Engel T, Dunleavy M, Hatazaki S, Paucard A, Matsushima S, Taki W, Henshall DC: Unilateral hippocampal CA3-predominant damage and short latency epileptogenesis after intramygdala microinjection of kainic acid in mice. *Brain Res* 2008, 1213:140-151.

25. Beamer EH, Jurado-Arjona J, Jimenez-Mateos EM, Morgan J, Reschke CR, Kenny A, de Leo G, Olivos-Ore LA, Arribas-Blazquez M, Madden SF, Merchant-Rubira J, Delanty N, Farrell MA, O'Brien DF, Avila J, Diaz-Hernandez M, Miras-Portugal MT, Artalejo AR, Hernandez F, Henshall DC, Engel T: MicroRNA-22 controls aberrant neurogenesis and changes in neuronal morphology after status epilepticus. *Front Mol Neurosci* 2018, 11:442.

26. Blauwblomme T, Jiruska P, Huberfeld G: Mechanisms of ictogenesis. *Int Rev Neurobiol* 2014, 114:155-185.

27. Shibata M, Nakao H, Kiyonari H, Abe T, Aizawa S: MicroRNA-9 regulates neurogenesis in mouse telencephalon by targeting multiple transcription factors. *J Neurosci* 2011, 31:3407-3422.

28. Tan CL, Plotkin JL, Veno MT, von Schimmelmann M, Feinberg P, Mann S, Handler A, Kjems J, Surmeier DJ, O'Carroll D, Greengard P, Schaefer A: MicroRNA-128 governs neuronal excitability and motor behavior in mice. *Science* 2013, 342:1254-1258.

29. Morin-Brureau M, Milior G, Royer J, Chali F, LeDuigou C, Savary E, Blugeon C, Jourdren L, Akbar D, Dupont S, Navarro V, Baulac M, Bielle F, Mathon B, Clemenceau S, Miles R: Microglial phenotypes in the human epileptic temporal lobe. *Brain* 2018, 141:3343-3360.

30. Koepp MJ, Arstad E, Bankstahl JP, Dedeurwaerdere S, Friedman A, Potschka H, Ravizza T, Theodore WH, Baram TZ: Neuroinflammation imaging markers for epileptogenesis. *Epilepsia* 2017, 58 Suppl 3:11-19.

31. Janssen B, Vugts DJ, Wilkinson SM, Ory D, Chalson S, Hoozemans JJM, Schuit RC, Beaino W, Kooijman EJM, van den Hoek J, Chishty M, Domene A, Van der Perren A, Villa A, Maggi A, Molenaar GT, Funke U, Shevchenko RV, Baekelandt V, Bormans G, Lammertsma AA, Kassiou M, Windhorst AD: Identification of the allosteric P2X7 receptor antagonist [(11)C]SMW139 as a PET tracer of microglial activation. *Sci Rep* 2018, 8:6580.

32. Zhao X, Liao Y, Morgan S, Mathur R, Feustel P, Mazurkiewicz J, Qian J, Chang J, Mathern GW, Adamo MA, Ritaccio AL, Grunenthal M, Zhu X, Huang Y: Noninflammatory Changes of Microglia Are Sufficient to Cause Epilepsy. *Cell Rep* 2018, 22:2080-2093.

33. Lu W, You R, Yuan X, Yang T, Samuel EL, Marcano DC, Sikkema WK, Tour JM, Rodriguez A, Kheradmand F, Corry DB: The microRNA miR-22 inhibits the histone deacetylase HDAC4 to promote T(H)17 cell-dependent emphysema. *Nat Immunol* 2015, 16:1185-1194.
34. Roth TL, Nayak D, Atanasijevic T, Koretsky AP, Latour LL, McGavern DB: Transcranial amelioration of inflammation and cell death after brain injury. Nature 2014, 505:223-228.
35. Allan SM, Tyrrell PJ, Rothwell NJ: Interleukin-1 and neuronal injury. Nat Rev Immunol 2005, 5:629-640.
36. Iori V, Iyer AM, Ravizza T, Beltrame L, Paracchini L, Marchini S, Cerovic M, Hill C, Ferrari M, Zucchet M, Molteni M, Rossetti C, Brambilla R, Steve White H, D'Incagli M, Aronica E, Vezzani A: Blockade of the IL-1R1/TLR4 pathway mediates disease-modification therapeutic effects in a model of acquired epilepsy. Neurobiol Dis 2017, 99:12-23.
37. A. R: Direct Data Submission 2012/05/07 MGI Direct Data Submission (MGI: J:183005). 2012.
38. Jovicic A, Roshan R, Moisoi N, Pradervand S, Moser R, Pillai B, Luthi-Carter R: Comprehensive expression analyses of neural cell-type-specific miRNAs identify new determinants of the specification and maintenance of neuronal phenotypes. J Neurosci 2013, 33:5127-5137.
39. Jassal B, Matthews L, Viteri G, Lorente P, Sidiropoulos K, Cook J, Gilleispie M, Hauw R, Loney F, May B, Milacic M, Rothfels K, Sevilla C, Shorser S, Varusai T, Weiser J, Wu G, Stein L, Hermjakob H, D'Eustachio P: The reactome pathway knowledgebase. Nucleic Acids Res 2020, 48:D498-D503.
40. Kanehisa M, Goto S: KEGG: kyoto encyclopedia of genes and genomes. Nucleic Acids Res 2000, 28:27-30.
41. Yu G, He QY: ReactomePA: an R/Bioconductor package for reactome pathway analysis and visualization. Mol Biosyst 2016, 12:477-479.
42. Kuleshov MV, Jones MR, Rouillard AD, Fernandez NF, Duan Q, Wang Z, Koplev S, Jenkins SL, Jagodnik KM, Lachmann A, McDermott MG, Monteiro CD, Gundersen GW, Ma'ayan A: Enrichr: a comprehensive gene set enrichment analysis web server 2016 update. Nucleic Acids Res 2016, 44:W90-97.
43. Pujato M, Kieken F, Skiles AA, Tapinos N, Fiser A: Prediction of DNA binding motifs from 3D models of transcription factors; identifying TLX3 regulated genes. Nucleic Acids Res 2014, 42:13500-13512.
44. Chou CH, Shrestha S, Yang CD, Chang NW, Lin YL, Liao KW, Huang WC, Sun TH, Tu SJ, Lee WH, Chiew MY, Tai CS, Wei TY, Tsai TR, Huang HT, Wang CY, Wu HY, Ho SY, Chen PR, Chuang CH, Hsieh PJ, WuYS, Chen WL, Li MJ, Wu YC, Huang XY, Ng FL, Buddhakosai W, Huang PC, Lan KC, Huang CY, Weng SL, Cheng YN, Liang C, Hsu WL, Huang HD: miRTarBase update 2018: a resource for experimentally validated microRNA-target interactions. Nucleic Acids Res 2018, 46:D296-D302.
45. Karagkouni D, Paraskevopoulos MD, Chatzopoulos S, Vlachos IS, Tastsoglou S, Kanellos I, Papadimitriou D, Kavakiotis I, Maniou S, Skoupos G, Vergoulis T, Dalamagas T, Hatzigeorgiou AG: DIANA-TarBase v8: a decade-long collection of experimentally supported miRNA-gene interactions. Nucleic Acids Res 2018, 46:D239-D245.
46. Xiao F, Zuo Z, Cai G, Kang S, Gao X, Li T: miRecords: an integrated resource for microRNA-target interactions. Nucleic Acids Res 2009, 37:D105-110.
47. Agarwal V, Bell GW, Nam JW, Bartel DP: Predicting effective microRNA target sites in mammalian mRNAs. Elife 2015, 4.
48. Liu W, Wang X: Prediction of functional microRNA targets by integrative modeling of microRNA binding and target expression data. Genome Biol 2019, 20:18.
49. Raoof R, Bauer S, El Naggar H, Connolly NMC, Brennan GP, Brindley E, Hill T, McArdle H, Spain E, Forster RJ, Prehn JHM, Hamer H, Delanty N, Rosenow F, Mooney C, Henshall DC: Dual-center, dual-platform microRNA profiling identifies potential plasma biomarkers of adult temporal lobe epilepsy. EBioMedicine 2018, 38:127-141.

Figures

A 2.0

B 2.0
miRNA expression and behaviour in miR-22-deficient mice. (A-E) Graphs show relative expression of miR-22 and a selection of other brain cell type-enriched miRNAs in miR-22 mutant mice. (A) Graph confirms gene dose-related reductions of miR-22 in the hippocampus of miR-22+/- and miR-22-/- compared to wildtype (wt) mice. (B-E). Expression of neuron-enriched miRNAs (miR-124a and -134), microglial (miR-150, miR-342) and astrocyte (miR-29a) were unaltered in miR-22 mutant mice. (F, G) Graphs show miR-22-deficient mice perform normally in tests of coordination (rotarod) and anxiety (elevated plus maze). *P < 0.05 and ***P < 0.01 (n = 6/group; 3 males and 3 females).
Hippocampal histology in naïve mice lacking miR-22. Representative photomicrographs showing morphology of the hippocampus of wildtype (wt) and miR-22-deficient mice (miR-22+/-, miR-22-/-). Sections were stained with markers of (A) nissl, (B) NeuN (neurons), (C) GFAP (astrocytes), (D) Iba1 (microglia) and (E) parvalbumin (interneurons). No differences were found for any parameter (n = 6/group).
Status epilepticus and acute histological outcomes in miR-22-deficient mice. (A) Representative recordings of electrographic seizures (pseudocolour transforms of original EEG) during status epilepticus in mir-22+/− and miR-22−/− mice compared to wildtype controls. (B-E) Graphs show EEG total power and amplitude during the 40 min period from kainic acid administration to lorazepam and the 60 min period thereafter. Blue dots indicate male mice, red dots indicate female mice (n = 7-10/group). (F-I). Representative photomicrographs of the hippocampus of mice 72 h after status epilepticus. There were no differences in acute neuronal loss, astrogliosis, microgliosis or interneuron (parvalbumin) staining.
Days after status epilepticus

Seizure duration (s)

Days after status epilepticus

Total seizure count

0.5 mV
30 sec
Figure 4

Epilepsy phenotype in miR-22-deficient mice. (A) Graphic shows individual mouse (1-6) daily seizure counts and summative graphs showing spontaneous seizures during 14 days recording in male wildtype (wt) and miR-22/- mice. (B) Graph showing numbers of spontaneous recurrent seizures during the 14 days recording in which miR-22/- mice display an elevated number of spontaneous seizures compared to wildtype mice. (C) Average time in seizures per day was increased in mice lacking miR-22 and average spontaneous seizure duration is longer in miR-22/- mice. (D) Total seizure counts for the two groups over the 14 days monitoring. *P < 0.05 (n = 6/group). (E) Representative traces of spontaneous seizures captured by telemetry EEG in a wildtype and miR-22/- mouse. Note the longer duration of seizure typical to miR-22-deficient mice.
Figure 5

Histological findings after epilepsy monitoring in mice lacking miR-22. Representative photomicrographs and graphs of cell counts, showing histological outcomes in wildtype (wt) and miR-22-deficient mice at the end of 14 days epilepsy monitoring. (A, B) Neuronal loss, (C, D) astrogliosis, (E, F) microgliosis and (G, H) interneuron counts. Note, most cell markers were similar but microglial staining was increased in miR-22-deficient mice. *P < 0.05 (n = 6/group).
RNA sequencing reveals downregulated inflammatory transcriptional landscape in miR-22-deficient mice. (A)
Volcano plot based on RNA sequencing, shows that miR-22 deficiency in mice resulted in the decreased expression of 18 immune-associated genes (labelled with names) in the hippocampus 24 h after status epilepticus. (B) Bar plot shows the numbers of genes in different classes of 437 transcription factors identified for the 18 immune-associated genes, including 93 transcription repressors and co-repressors (21.3%). (C) Bar plot showing the numbers of the immune-associated genes targeted by each of the 25 miR-22 mediated transcriptional repressors. Each of these repressors regulates several the immune-associated targets. (D) Volcano plot showing gene expression, magnitude of changes and statistical significance of differential expression analysis of the 25 transcriptional repressors. None of the repressors passed the statistical significance threshold (p value < 0.01) due to small magnitudes of changes in gene expression and/or relatively low expression abundance.

**Supplementary Files**

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