A TILLING Platform for Functional Genomics in *Brachypodium distachyon*

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**Abstract**

The new model plant for temperate grasses, *Brachypodium distachyon* offers great potential as a tool for functional genomics. We have established a sodium azide-induced mutant collection and a TILLING platform, called “BRACHYTIL”, for the inbred line Bd21-3. The TILLING collection consists of DNA isolated from 5530 different families. Phenotypes were reported and organized in a phenotypic tree that is freely available online. The tilling platform was validated by the isolation of mutants for seven genes belonging to multigene families of the lignin biosynthesis pathway. In particular, a large allelic series for *BdCOMT6*, a caffeic acid O-methyl transferase was identified. Some mutants show lower lignin content when compared to wild-type plants as well as a typical decrease of syringyl units, a hallmark of COMT-deficient plants. The mutation rate was estimated at one mutation per 396 kb, or an average of 680 mutations per line. The collection was also used to assess the Genetically Effective Cell Number that was shown to be at least equal to 4 cells in *Brachypodium distachyon*. The mutant population and the TILLING platform should greatly facilitate functional genomics approaches in this model organism.

**Introduction**

*Brachypodium distachyon* is a valuable model plant for economically important temperate grasses such as wheat, barley and oat [1]. Attractive features are the small size, simple culture conditions and the small (272 Mb), diploid (2n = 10) and fully sequenced genome [2]. *B. distachyon* is currently being used as a model for the study of domestication in grasses [1,2,3,4,5], plant pathogen interactions, root and culm development, biomass production and cell wall biosynthesis. Important progress has been made in the development of efficient transformation protocols [6,7,8] and sequence-indexed T-DNA insertion collections [9]: the BrachyTAG collection at the John Innes Centre (5000 lines) ([10], http://www.brachytag.org/) and the USDA Brachypodium Genome Resources collection (8491 lines) [11]. To extend the panel of resources for functional genomics in this species, we have developed a mutagenized population and a TILLING (Targeting Induced Local Lesion IN Genome) platform. Chemically-induced mutants are complementary to insertion mutants in several aspects: (i) mutation rates are higher and hence screens can be done on smaller mutant populations, (ii) single base pair changes, as opposed to insertion mutants, more likely yield allelic series of partial loss-of-function, conditional or gain-of-function mutants and (iii) somaclonal variation is avoided in the absence of an *in vitro* culture step that is required for T-DNA-induced mutagenesis [12,13]. An efficient method to identify new alleles for target genes is TILLING. This method developed a decade ago has been successfully applied to many plant species [14,15]. In the TILLING method, seeds are mutagenized, the resulting M1 plants are self-fertilized, and the M2 generation of individuals is used to prepare DNA samples for mutational screening, while seeds of the M3 families can be stored and distributed. DNA samples are pooled and subjected to gene-specific PCR. The amplification products are incubated with an endonuclease that preferentially cleaves mismatches in heteroduplexes between wild type and mutant. Upon detection of a mutation in a pool, the individual DNA samples are similarly screened to identify the plant carrying the mutation. TILLING populations have been established for grasses including wheat, sorghum, barley, rice and oat [15,16,17,18,19,20,21,22,23] but to our knowledge not yet for *Brachypodium distachyon*. Here we report on the development of a mutagenized population and a TILLING platform and demonstrate the efficiency of the TILLING method with the identification of a series of allelic mutants for an O-methyl transferase, involved in the lignification of the internodes.
Materials and Methods

Plant Material and Growth Conditions

*Brachypodium distachyon* (L.) Beauv. inbred line Bd21-3 was kindly provided by John Vogel. Bd21-3 seeds were grown in a greenhouse under long-day conditions (18 h light, 400 watt sodium lamps). Day and night temperatures were 23°C and 18°C, respectively. The relative humidity was about 60%. Plants were grown in soil (one-liter pots) and watered twice a week.

Chemical Mutagenesis

For the production of mutants, dry seeds were pre-soaked in distilled water for 2 h. Portions of 5000 seeds were then suspended in 200 mL of fresh sodium azide (NaN₃) solution diluted in phosphate buffer (0.1 M, pH 3) for 2 h under the hood and with gentle shaking. The seeds were washed 3 times in water for 1 h and then kept at +4°C for 72 h before sowing in pots. For establishment of the kill curve, 500 seeds were mutagenized with 0.5, 1, 1.5, 3 or 10 mM NaN₃.

Genomic DNA Extraction and Pooling

Four M₂ plants per family were grown for one month in a greenhouse. DNA was extracted from 3 cm-long portions of the median foliar part. The collected samples were pooled and placed in 96-well plates containing 2 steel beads per well. Samples were lyophilized and ground using a bead mill. Genomic DNAs were isolated using the DNeasy 96 Plant Kit (Qiagen, Hilden, Germany). All genomic DNAs were both quantified on a 1% agarose gel with λ DNA (Invitrogen, Carlsbad, CA, USA) as a concentration reference using a NanoDrop spectrophotometer 2000 c (Thermo Fisher Scientific, MA, USA). DNA concentration was normalized to 6 ng·µL⁻¹ and pooled eightfold in a 96-well format.

PCR Amplification and Detection of Mutations

DNA amplification is based on nested-PCR. The first PCR amplification is a standard PCR reaction with target-specific primers and 10 ng of *Brachypodium* genomic DNA. One µl of the first PCR product served as a template for the second nested PCR amplification, with a combination of specific primers carrying M13 tail and M13 universal primers, M13F700 (5’-CAC-GACGTTGTAAAACGAC-3’) and M13R800 (5’-GGATAACATTTTCAGACAGG-3’); labelled at the 5’end with infra-red dyes IRD700 and IRD800 (LI-COR®, Lincoln, Nebraska, USA) respectively. Mutation detection was carried out as described previously except for the second PCR. This PCR was carried out using 0.05 µM of specific primers carrying M13 tail and 0.1 µM of M13 universal primers. The identity of the mutations was determined by sequencing.

Sequence Analysis Tools

The CODDLE software (Codons Optimized to Discover Deleterious Lesions, http://www.proweb.org/coddle/) was used to identify regions of the target gene in which G/C to A/T transitions are most likely to result in deleterious effects on the protein. The PARSESNP software (Project Aligned Related Sequences and Evaluate SNPs, http://www.proweb.org/parsesnp/) was used to illustrate the distribution of mutations within the gene and to indicate the nature of each single mutation. The SIFT software (Sorting Intolerant from Tolerant, http://sift.jcvi.org/www/SIFT_seq_submit2.html) was used to predict the impact of the mutation on the protein. Multiple sequence alignment of full-length protein sequences was performed with ClustalW software (http://www.ebi.ac.uk/Tools/clustalw2).

Phylogenetic Tree and 3D-structure Prediction

Protein sequences of COMT from several vascular plants were identified by protein blast (http://www.ncbi.nlm.nih.gov/and
http://www.greenphyl.org) and aligned with Clustaw2 (pairwise alignment) at http://www.ebi.ac.uk/Tools/msa/clustalw2. Phylogenetic analysis was performed with TreeTop [http://www.genebee.msu.su/services/plhree_reduced.html]. Cluster algorithm was used and 100 bootstrap were done. Final construction of tree was made with TREX [Treeview Newick Viewer option] at http://www.trex.uqam.ca/. Bootstrap values are expressed in percentages and placed at nodes of the trees. The 3D-structure of the BdCOMT6 protein was predicted from the crystallized ryegrass enzyme [24] with the molecular modeling program Geno3D [http://geno3d-pbil.ibcp.fr/] and UCSF Chimera software [http://www.cgl.ucsf.edu/chimera/].

**Lignin Analyses**

All reagents and solvents were high-quality grade commercial reagents employed without further purification. Dried mature stems (3 month old) from wild-type (WT) plants or from azygous (mutagenized plants segregating the WT allele for the studied locus) or homozygous mutant plants were ground to 0.5 mm before exhaustive extraction with water, then ethanol in a Soxhlet apparatus. The lignin content was measured on the recovered extractive-free samples and by the Klason method [25]. Lignin structure was evaluated by thioacidolysis performed from 10 to 20 mg of extractive-free sample put together with 0.5 mg of C21 internal standard in 10 mL of reagent (dioxane/ethanethiol 9/1 V/V, containing 0.2 M BF3 etherate) and incubated at 100°C for 4 h. After the reaction, the lignin-derived monomers were extracted with methylene chloride as previously described [26], the combined organic extracts were concentrated to about 2 mL and then 4 μL of the sample were silylated by 100 μL BSTFA and 5 μL pyridine before injection onto a DB1 supelco capillary columns (carrier gas helium, constant flow rate 1 mL/min) operating from 40 to 180°C at +30°C/min, then 180 to 260°C at +2°C/min and combined with an ion trap mass spectrometer (Varian Saturn2100) operating in the electron impact mode (70 eV), with ions detected on the 50–600 m/z range. The surface area of the p-hydroxyphenyl (H), guaiacyl (G), syringyl (S) and 5-hydroxyguaiacyl (5-OH G) monomers were measured on specific ion chromatograms, at m/z 239, 269, 299, and 357 respectively.

**Results and Discussion**

**Production of a Mutagenized Collection**

When grown under long-day conditions in our greenhouses (18 h light, Figure 1A), the life cycle was about 12 weeks. Sodium azide (NaN3) was used to mutagenize *B. distachyon* Bd21-3 seeds. This compound is commonly used for mutagenesis of grasses [15] and the frequency of chromosome breakage is relatively low. To establish a dose-response kill curve, we determined the fraction of M1 seeds that germinated after imbibition with increasing NaN3 concentrations (Figure 1B). The lowest germination frequency was 28% for 10 mM NaN3. The frequency of albino M2 seedlings (Figure 1C) and miniature/dwarf plants (Figure 1D) also showed a dose-dependent increase with for instance a maximum frequency of 3.3% and 1.3% albinos respectively for 10 mM and 3 mM NaN3 (Figure 1D). We concluded that 3 mM and 10 mM NaN3 provided a good compromise combining high mutation rate and

**Figure 2. Examples of phenotypes detected in the Brachypodium mutant collection.** A. Spikelets of wild-type plants. B–D Aborted or abnormal spikelets in mutants. E. Ligule of wild-type plant. F. Abnormal ligule in mutant. G, H. Mis-shaped leaves in mutants. I. Crumple stem in mutant. J. Coloured stems in mutant. K. Advenitious roots in mutant. L. Curved stems in mutant. doi:10.1371/journal.pone.0065503.g002

**Table 1. Types of phenotypes observed on 565 phenotyped lines (3360 plants).**

| Major category | Phenotype sub-category | Number of families affected | % of families affected |
|----------------|------------------------|----------------------------|-----------------------|
| Dead           | Not emerged or early death | 340                        | 15.4                  |
| Plant development | Delay Plant development | 87                          | 15.4                  |
| Plantlet       | Albino                 | 20                         | 3.5                   |
| Leaf           | Size                   | 129                        | 22.8                  |
|                | Color                  | 121                        | 21.4                  |
|                | Appearance             | 125                        | 22.1                  |
|                | Shape and arrangements | 9                          | 1.6                   |
| Stem           | Shape                  | 112                        | 19.8                  |
|                | Color                  | 19                         | 3.3                   |
| Node           | Shape                  | 1                          | 0.2                   |
| Spike          | Flowering time         | 52                         | 9.2                   |
|                | Reproductive organs    | 15                         | 2.6                   |
|                | Spike color            | 62                         | 11.1                  |
| Plant architecture | Architecture    | 394                        | 69.7                  |
|                | Height and appearance  | 146                        | 25.8                  |
|                | Tillers                | 70                         | 12.4                  |
|                | Branching type         | 47                         | 8.3                   |

*phenotypes are available at http://urgv.evry.inra.fr/UTILLdb. doi:10.1371/journal.pone.0065503.t001
sufficient germination frequency. These concentrations were used to generate the mutagenized population of 5530 individual families.

Plant Phenotyping

The phenotypes (shape, color and size) of a subset of mutants were recorded, using an ontology developed for sorghum [27], at three developmental stages: Germination/seedling stage, vegetative stage (before flowering) and reproductive stage (after flowering). Phenotypes were organized in a phenotypic tree available at http://urgv.evry.inra.fr/UTILLdb. Figure 2 and Table 1 illustrate the most striking phenotypes of greenhouse-grown mutants.

TILLING Genes Involved in Lignin Biosynthesis

*B. distachyon* is increasingly used as a model for bioenergy grasses [28,29]. Lignins are the main obstacle in the enzymatic conversion of cellulose into fermentable sugars. We therefore chose to screen the mutant collection for genes involved in lignin biosynthesis as a validation of the tilling method [29]. We focused on seven genes belonging to multigene families that are potentially involved in monolignol biosynthesis (Table 2): one caffeic acid O-methyltransferase gene (*Bradi3g16530*), two laccase genes (*Bradi1g74320*, *Bradi1g66720*), one 4-coumarate:coenzyme A ligase gene (*Bradi3g05750*) and three cytochrome P450 genes (*Bradi2g31510*, *Bradi2g53470*, *Bradi3g43160*).

We identified a total of 128 mutations. The nucleotide changes induced by NaN₃ are mainly G/A and C/T substitutions similar to those induced by ethyl methane sulfonate (EMS) for instance [30,31]. Only 9% other mutations were detected (Table 3). We identified 63% non-synonymous mutations (including 53% and 5% inducing amino acid changes or stop codons, respectively). One line showed a mutation in a splicing region. Extrapolating from this small set of genes, we calculated an average mutation rate of one in around 400 kb (Table 2) or 700 mutations per genome for a genome size of 272 Mb [2]. This frequency of induced mutations is similar to that found in EMS-mutagenized rice (1/294 kb, [23]) or in barley treated with NaN₃ (1/374 kb, [22]) or EMS (1/500 kb, [20]).

We also used our mutagenized population to estimate the ‘Genetically Effective Cell Number (GECN)’. This is the number of cells within the shoot meristem of the embryo that will contribute to the seed output. The GECN is usually estimated by determining the proportion of mutant seedlings in M₂ families [32,33]. We took advantage of our sequencing data on M₂ families from individual M₁ plants segregating mutations in 4 genes. In 584 lines from 30 different families (between 9 and 49 M₂ independent lines were analyzed per family) we detected 38

**Table 2.** Tilled genes and mutation frequency in the mutant collection with 5530 M₂ families screened.

| Target names | Accession number | Amplicon size (bp) | GC content (%) | Identified mutants | Mutation frequency |
|--------------|------------------|--------------------|----------------|--------------------|--------------------|
| COMT         | Bradi3g16530     | 1245               | 65             | 23                 | 1/300 kb           |
| Laccase      | Bradi1g74320     | 2372               | 43             | 17                 | 1/771 kb           |
| Laccase      | Bradi1g66720     | 2607               | 51             | 18                 | 1/801 kb           |
| 4CL          | Bradi3g05750     | 735                | 68             | 17                 | 1/239 kb           |
| C4H          | Bradi2g31510     | 643                | 65             | 14                 | 1/254 kb           |
| C4H          | Bradi2g53470     | 728                | 63             | 12                 | 1/335 kb           |
| C4H          | Bradi3g43160     | 829                | 61             | 27                 | 1/170 kb           |
| TOTAL        |                  | 9159               | 59             | 128                | 1/396 kb           |

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**Table 3.** Frequencies of induced mutations types in tilled gene-coding regions.

| Accession number | Missense* | Nonsense* | Splicing* | Silent* | Unusual nucleic transition* |
|------------------|-----------|-----------|-----------|---------|----------------------------|
| COMT             | Bradi3g16530 | 13        | 0         | 0       | 10 | 0 |
| Laccase          | Bradi1g74320 | 8         | 3         | 0       | 6  | 0 |
| Laccase          | Bradi1g66720 | 8         | 0         | 1       | 5  | 3 |
| 4CL              | Bradi3g05750 | 13        | 0         | 0       | 4  | 4 |
| C4H              | Bradi2g31510 | 10        | 1         | 0       | 3  | 1 |
| C4H              | Bradi2g53470 | 6         | 1         | 0       | 4  | 1 |
| C4H              | Bradi3g43160 | 13        | 1         | 0       | 13 | 2 |
| TOTAL            |            | 71        | 6         | 1       | 45 | 11 |
| TOTAL (%)        |            | 57.7      | 4.8       | 0.8     | 36.7 | 8.6 |

* nucleic acid transition is a non-synonymous mutation and induce amino acid change in the translated protein.

nucleic acid transition produces a stop codon and may induce a truncated protein.

nucleic acid transition is located in splicing motif.

nucleic acid transition induces a synonymous mutation and then no change in the translated protein.

other nucleic acid transition than guanine to adenine and cytosine to thymine.

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homozygous mutations corresponding to a ratio of 1:15.3 (= 38/584). The 1:15 ratio corresponds to a GECN of 4. This is higher than in Arabidopsis (GECN = 2) but in accordance with the number found in other grass species [33,34].

Identification of a Caffeic acid O-methyltransferase (COMT) Gene Potentially Involved in Internode Lignification

*B. distachyon* lignins, like those in other grasses, are mainly composed of guaiacyl (G) and syringyl (S) units, with low amounts of p-hydroxyphenyl (H) units [35] [36]. These H, G and S lignin units respectively originate from the three monolignols, namely p-coumaryl, coniferyl, and sinapyl alcohols that differ only in the degree of methoxylation of the phenolic ring [37]. The main role of the COMT involved in lignification is the methylation of 5-hydroxyconiferaldehyde to produce sinapaldehyde, which is reduced by another enzyme to sinapyl alcohol, the precursor of S lignin units. The COMT enzyme belongs to the S-Adenosyl methionine (SAM)-dependent O-methyltransferases. It is active as a homodimer and does not need any metal ion as cofactor. The hallmark of transgenic or mutant angiosperms with strongly repressed COMT activity is a reduction of the amount of S lignin units (or of the S/G ratio) together with the appearance of easily detectable amounts of 5-hydroxyguaiacyl (5-OH G), which is present only in trace amounts in the WT [38] [39,40,41,42]. Another trait of some COMT-mutant grass lines, referred to as the brown-midrib mutants, is a lower lignin level resulting in a higher
enzymatic degradability [43], and reduced levels of p-coumaric acid (CA) ester-linked to the cell walls [44].

To unambiguously identify the *B. distachyon* COMT gene specifically involved in lignification, protein sequences of orthologs in several species were BLASTed onto the *B. distachyon* predicted proteome sequence. Eight proteins were identified: BdCOMT1 (Bradi1g14870), BdCOMT2 (Bradi2g02380), BdCOMT3 (Bradi2g02390), BdCOMT4 (Bradi2g19830), BdCOMT5 (Bradi2g19850), BdCOMT6 (Bradi3g16530), BdCOMT7 (Bradi3g55890), BdCOMT8 (Bradi4g20020). We performed a phylogenetic analysis with the most exhaustive list of encoded OMT proteins found in *Oryza sativa*, *Arabidopsis thaliana* and *Zea mays* genomes [45,46,47] (Figure 3, Information S1). In addition, we added to the phylogenetic analysis the protein sequences for which biological data (transgenics or mutants) support indisputably a role of the corresponding protein in lignification of several grass (maize, ryegrass, tall fescue, switchgrass, sorghum) and poplar [24,39,42,48,49,50,51]. *AtOMT1* was clearly identified by our group as a unique gene involved in sinapyl alcohol biosynthesis in Arabidopsis since the knockout mutant displays a lignin devoid of S units [52]. Therefore, *AtOMT1* is a reference model for dicot COMT proteins as well as maize ZmCOMT1 (BM3) is a reference for grasses [49]. The phylogenetic analysis shows that both dicot proteins (*AtOMT1*, *PtOMT*) cluster together whereas four members of BdCOMTs (BdCOMT3, BdCOMT2, BdCOMT1 and BdCOMT6) are grouped with the genuine grass COMT proteins, BdCOMT6 being the closest ortholog (figure 3).

![Figure 4. Klaon lignin (KL) level of extractive-free mature stems of *Bd4142*, *Bd4604* and *Bd5139* lines as compared to control samples. Control samples are either wild-type (Bd21-3) or agzyous plants grown together with the corresponding mutants. The KL level is expressed as weight percentage of the extractive-free sample. Data are means and SD from 4 or 5 plants analyzed per line. Asterisks indicate significant difference compared to the control (ANOVA value at P<0.05).
![Figure 4](http://www.plosone.org/article-pdf/10.1371/journal.pone.0065503.g004)

Table 4. Allelic series of mutations in *BdCOMT6* gene identified by TILLING.

| Nucleic acid transition* | Amino acid substitution* | SIFT* | Family name |
|-------------------------|--------------------------|-------|-------------|
| G37A                    | Asp13Asn                 | 0.18  | 5645        |
| G117A                   | Leu39Leu                 | -     | 5714        |
| C498T                   | Asp166Asp                | -     | 5588        |
| G588A                   | Glu196Glu                | -     | 8240        |
| C600T                   | Tyr200Tyr                | -     | 5827        |
| G616A                   | Gly206Ser                | 0.01  | 6840        |
| G638A                   | Gly213Asp                | 0.26  | 3380        |
| G638A                   | Gly213Asp                | 0.26  | 3380        |
| G672A                   | Gly224Gly                | -     | 3725        |
| G708A                   | Gly236Gly                | -     | 5338        |
| G721A                   | Gly241Arg                | 0.02  | 4688        |
| G737A                   | Gly246Asp                | 0     | 4604        |
| G737A                   | Gly246Asp                | 0     | 4604        |
| C762T                   | Pro254Pro                | -     | 5115        |
| G767A                   | Gly256Ser                | 0     | 5139        |
| G767A                   | Gly256Asp                | 0     | 7549        |
| C840T                   | Cys280Cys                | -     | 5348        |
| C854T                   | Pro285Lys                | 0.02  | 7391        |
| G969A                   | Gly323Gly                | -     | 3730        |
| G976A                   | Gly326Lys                | 0     | 4142        |
| G1013A                  | Gly338Asp                | 0.14  | 5200        |
| G1063A                  | Ala355Thr                | 0.01  | 4927        |
| G1071A                  | Glu357Glu                | -     | 185         |

*Position of transition in mutants are relative to the starting ATG on the coding sequence.
*Position of substitution in mutants are relative to the starting methionine of the encoded protein.

Numbers are predictive score from the SIFT software (http://sift.bii.a-star.edu.sg/).

Mutations in *BdCOMT6* Affect the Lignification of Mature Stems

We identified 25 lines, corresponding to 22 different mutations, among which eleven missense mutations cause changes in the BdCOMT6 amino acid sequence (Table 4). No induced stop codon mutation were identified. A first analysis using the SIFT software (using 0.05 as significant threshold) indicated that the substitutions caused by the mutations in lines *Bd4880, Bd4688, Bd4604, Bd7480, Bd5139, Bd7549, Bd7391, Bd4142* and *Bd4927* may partially or totally disrupt the COMT activity. Among them, mutations in lines *Bd5139* and *Bd7549* were found redundant, as well as those in lines *Bd7480* and *Bd4604*, thereby reducing the number of mutations potentially affecting COMT activity to seven.

We next isolated homozygous lines for the all *comt* mutants, except for *Bd7480*. Indeed, this line failed to produce viable seeds at heterozygous stage. All genotyped plants were indistinguishable from WT plants when grown in the greenhouse except for homozygous *Bd211*, which was dwarfed. The dwarfism presumably is independent from the mutated COMT allele since *Bd3380*, which carries the same mutations was not dwarfed (Table 4). We therefore excluded *Bd211* from the subsequent analyses. We next studied the lignin composition in mature stems using thioacidolysis (reviewed in [53]). Thioacidolysis identifies H, G and S thioethylated monomers from arylglycerol-β-ether-linked H, G and S units. In addition, it allows the identification of 5-OH G thioacidolysis monomers in higher amounts, compared to the trace amounts detected in the control lines (Table 5). Together with the increased frequency of 5-
Table 5. Relative frequency (% molar) of \( p \)-hydroxyphenyl H, guaiacyl G, syringyl S and 5-hydroxyguaiacyl 5-OH G monomers released by thioacidolysis of mature and extractive-free stems from control (Ctrl) and mutant \( Bd4142 \), \( Bd4604 \), \( Bd5139 \) lines.

| Culture        | Sample (n°)  | % H (SD) | % G (SD) | % S (SD) | % 5-OH G (SD) | S/G molar ratio (SD) |
|----------------|--------------|----------|----------|----------|---------------|----------------------|
| 1              | Azygous Control (4) | 5.9 (0.3) | 37.9 (1.9) | 55.7 (1.0) | 0.5 (0.0)     | 1.47 (0.15)           |
|                | \( Bd4142 \) (4) | 6.2 (0.5) | 46.5 (3.1)* | 45.3 (3.3)* | 2.0 (0.3)*    | 0.98 (0.14)*          |
| 2              | Wild-type Control (2) | 3.5 (0.3) | 30.8 (2.3) | 65.4 (2.7) | 0.2 (0.0)     | 2.14 (0.25)           |
|                | \( Bd4604 \) (2) | 3.2 (0.0) | 38.4 (0.3)* | 55.1 (3.3)* | 3.3 (0.3)*    | 1.44 (0.02)*          |
| 3              | Wild-type Control (5) | 3.5 (0.5) | 31.4 (2.0) | 64.8 (2.4) | 0.3 (0.1)     | 2.08 (0.22)           |
|                | \( Bd5139 \) (5) | 4.0 (0.6) | 42.4 (0.4)* | 49.5 (1.4)* | 4.1 (1.5)*    | 1.17 (0.04)*          |

Ctrl (azygous line for \( Bd4142 \) and wild-type line for \( Bd4604 \) and \( Bd5139 \)) and corresponding mutant samples were recovered from plants grown together and in identical conditions. Asterisks indicate significant differences compared to the corresponding Ctrl (ANOVA, value at \( P < 0.01 \)).

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Figure 5. Structural representation of the amino acid substitution of \( BdCOMT6 \) protein in the line \( Bd4142 \). A. 3-D model of wild-type protein highlighting three amino acids important for proper enzymatic activities. B. 3-D model of the protein in \( Bd4142 \) line in which Glu-326 is substituted by Lys-326. SAH, S-Adenosyl-L-homocysteine is shown to illustrate the proximity of the substituted amino acid in the substrat binding pocket.

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OH G thioacidolysis monomers, we found that S monomer levels were substantially reduced (by 30 to 40% of the control value). In contrast, H monomers were obtained as minor components (3–6% range) and no substantial difference between mutants and controls was detected. Based on the S levels or S/G thioacidolysis ratio suggests, we hypothesize that COMT activity is lower in Bd5139 and Bd4412 compared to Bd4604. This hypothesis was further supported by the lignin level of the extractive-free stems as measured by the Klason method. Compared to the corresponding control, this level was reduced by 15%, 10% and 0% in Bd5139, Bd4412 and Bd4604 respectively (Figure 4).

To further investigate how amino acid substitutions may affect BdCOMT6 activity, we used the 3-D structure, determined for the closely related (90% amino acid sequence identity) Lolium perenne BdCOMT6 activity, we used the 3-D structure, determined for the O-dependent COMT enzymes belong to the plant type-1 family of SAM-OMT, LpOMT [24] to compare WT with mutant proteins. Both COMT enzymes belong to the plant type-1 family of SAM-dependent O-methyltransferases, have 360 amino acid residues and possess an auxiliary N-terminal domain that may functions in homodimerization [55,24]. The Bd4412 mutation induces a Gla-326-Lys substitution (Figure 5). Gla-326 is thought to be one of the catalytic bases that activate the hydroxyl group of the substrate/lignin [24], whereby Gla-326 provides the hydrogen bond acceptor in an interaction with the His-266 and contributes indirectly to the deprotonation of the phenolic substrate [24]. The Gla-326-Lys substitution, inverts the charge of the aminoacid and therefore is expected to alter proper protein activity and explain the severe reduction of S units detected in this line (Table 5).

The substitution observed in lines Bd5139 and Bd7549 is a glycine to aspartic acid at position 256, which is part of a loop facing a β-sheet carrying an amino acid involved in the substrate binding. It is worth noting that two residues (Phe-250 and Asp-248) in the vicinity of Gla-256 are also placed in this loop and are essential for stabilisation of the ligand (SAM/SAH). We speculate that the size and the charge modifications in this mutant could impact the β-sheet and destabilize the SAM binding site, disturbing its function as methyl donor and in consequence reduce the activity of the protein.

Finally, Bd4604 carries a Gla-246-Asp substitution. This residue is located at the periphery of the protein but still in the SAM/SAH binding domain and adjacent to a α-helical layer and a β-sheet both involved in SAM/SAH binding site conformation. The lateral chain of the aspartic acid in the mutant may turn towards residues involved in the binding of the cofactor. The size and charge differences may modify this site thus hampering the binding of SAH and the activity of the protein.

In conclusion, the three mutant lines that have an altered lignin content and/or composition share at least a mutation located in the vicinity of the SAM/SAH binding and catalytic domain. It is worth noting that the three COMT mutants do not show coloured leaf veins as observed in the brown-midrib mutants of maize, pearl millet or sorghum [56]. In addition, the levels of S units is still high in the mutants (as revealed by the 45 to 55% of S thioacidolysis monomers). This result suggests that enzymatic methoxylation at C5 of the phenolic ring of monogluin is still present in these lines. It remains to be shown whether it is due to residual enzymatic activity of the mutated proteins as shown in a similar allelic series in sorghum [57]. Finally, no grass COMT-deficient mutants or transgenic lines described so far are completely devoid of S units, in contrast to what has been reported in dicots, for example in the Arabidopsis Actom1 mutant [52]. This observation suggests that an alternative pathway may produce S units in grasses.

Conclusions
We have generated a large collection of chemically-induced mutants useful for forward genetics in B. distachyon. A subset of the phenotypes can be consulted at http://urgv.evry.inra.fr/UTILLdb. In addition, BRACHYTIL provides and efficient platform for reverse genetics. This study illustrates the power of this approach by the isolation of an allelic series for BdCOMT6 involved in monogluin biosynthesis. Next generation sequencing techniques will now greatly accelerate reverse genetics approaches in this collection.

Supporting Information
Information S1 amino acid sequences of putative COMT proteins used for phylogenetic analysis in Figure 3.

(DOCX)

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Author Contributions
Conceived and designed the experiments: RS AB LJ HH. Performed the experiments: MD OD SHYK YW MBY SA LC FL. ER NO CT. Analyzed the data: RS MD. Contributed reagents/materials/analysis tools: VB. Wrote the paper: RS CL HH.

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