VEGF-A isoforms program differential VEGFR2 signal transduction, trafficking and proteolysis

Gareth W. Fearnley¹, Gina A. Smith¹, Izma Abdul-Zani¹, Nadira Yuldasheva², Nadeem A. Mughal¹, Shervanthi Homer-Vanniasinkam³, Mark T. Kearney², Ian C. Zachary⁴, Darren C. Tomlinson⁵, Michael A. Harrison⁶, Stephen B. Wheatcroft² and Sreenivasan Ponnambalam¹,∗

ABSTRACT
Vascular endothelial growth factor A (VEGF-A) binding to the receptor tyrosine kinase VEGFR2 triggers multiple signal transduction pathways, which regulate endothelial cell responses that control vascular development. Multiple isoforms of VEGF-A can elicit differential signal transduction and endothelial responses. However, it is unclear how such cellular responses are controlled by isoform-specific VEGF-A–VEGFR2 complexes. Increasingly, there is the realization that the membrane trafficking of receptor–ligand complexes influences signal transduction and protein turnover. By building on these concepts, our study shows for the first time that three different VEGF-A isoforms (VEGF-A₁₆₅, VEGF-A₁₂₁ and VEGF-A₁₄₅) promote distinct patterns of VEGFR2 endocytosis for delivery into early endosomes. This differential VEGFR2 endocytosis and trafficking is linked to VEGF-A isoform-specific signal transduction events. Disruption of clathrin-dependent endocytosis blocked VEGF-A isoform-specific VEGFR2 activation, signal transduction and caused substantial depletion in membrane-bound VEGFR1 and VEGFR2 levels. Furthermore, such VEGF-A isoforms promoted differential patterns of VEGFR2 ubiquitylation, proteolysis and terminal degradation. Our study now provides novel insights into how different VEGF-A isoforms can bind the same receptor tyrosine kinase and elicit diverse cellular outcomes.

KEY WORDS: VEGF-A, VEGFR2, Endothelial, Trafficking

INTRODUCTION
Vascular endothelial growth factor A (VEGF-A) is a soluble ligand that is essential for mammalian development and function (Carmeliet et al., 1996; Ferrara et al., 1996; Koch et al., 2011). VEGF ligands bind to a receptor tyrosine kinase (RTK) subfamily termed vascular endothelial growth factor receptors (VEGFR1, 2 and 3), which regulate many aspects of vascular and lymphatic development (Koch et al., 2011; Smith et al., 2015). The VEGF family member VEGF-A binds a major receptor and membrane glycoprotein (VEGFR2) expressed on endothelial cells and such interactions facilitate signal transduction events that control different aspects of vascular physiology. This VEGFR2 RTK is a major regulator of new blood vessel sprouting i.e. angiogenesis (Carmeliet, 2005; Koch and Claesson-Welsh, 2012).

Human VEGF-A is encoded by the VEGFA gene on locus 6p21.3 and contains at least eight exons and seven introns. The VEGF4 primary RNA transcript undergoes alternative splicing to produce seven pro- and one anti-angiogenic isoforms of VEGF-A (Harper and Bates, 2008). However, the reasons for this VEGF-A isoform complexity and its conservation in mammalian species is unclear. In general, work in this field has focused on the VEGF-A₁₆₅ isoform that is secreted by most animal cells and tissues; nonetheless, it is clear that other VEGF-A isoforms elicit important and distinct biological responses from endothelial cells (Harper and Bates, 2008; Smith et al., 2015). The VEGF-A₁₆₅ isoform programs sequential steps in VEGFR2 tyrosine phosphorylation, ubiquitylation, trafficking and proteolysis (Bruns et al., 2010; Horowitz and Seerapu, 2012), linked to downstream signal transduction events that trigger pro-angiogenic outcomes such as cell proliferation, migration, tubulogenesis, vascular permeability and leukocyte recruitment (Fearnley et al., 2014a; Koch et al., 2011). Furthermore, VEGF-A isoforms differentially promote VEGFR2-dependent signal transduction and cellular responses (Fearnley et al., 2015, 2014a; Kawamura et al., 2008b; Pan et al., 2007). However, the underlying mechanism(s) by which VEGF-A isoforms act are still unclear, although VEGF-A isoform-specific binding is implicated in recruiting a co-receptor called neuropilin 1 (NRP1) (Ballmer-Hofer et al., 2011; Harper and Bates, 2008; Herzog et al., 2011; Kawamura et al., 2008a,b; Pan et al., 2007; Tillo et al., 2015). This membrane receptor can bind both VEGF-A₁₆₅ and VEGF-A₁₂₁ but only VEGF-A₁₆₅ is believed to form a trimeric complex with VEGFR2 and NRP1 (Koch et al., 2011).

The role of membrane trafficking in regulating receptor-ligand function is becoming increasingly apparent (Bruns et al., 2010; Horowitz and Seerapu, 2012). For example, VEGF-A₁₆₅-stimulated signal transduction requires co-ordination of VEGFR2 tyrosine kinase activation with residence at different locations within the endocytic pathway e.g. plasma membrane and endosomes (Gourlaouen et al., 2013; Jopling et al., 2009; Koch et al., 2014; Lanahan et al., 2013, 2010, 2014; Manickam et al., 2011; Nakayama et al., 2013; Yamada et al., 2014; Zhang et al., 2013). Plasma membrane VEGFR2 activation promotes recruitment of phospholipase Cγ1 thus stimulating phosphatidylinositol-4,5-bisphosphate (PIP₂) hydrolysis to generate inositol-1,4,5-trisphosphate (IP₃) and diacylglycerol (DAG): these molecules act as second messengers that promote cytosolic calcium ion flux and protein kinase C activation respectively (Meyer et al., 2003; Takahashi and Shibuya, 1997; Wong and Jin, 2005). However,
VEGF-A-stimulated activation of the MAP kinase pathway is linked to VEGFR2 residence in early endosomes (Bruns et al., 2010; Jopling et al., 2009; Lampugnani et al., 2006; Lanahan et al., 2010).

An important question is whether VEGF-A isoforms have the capacity to differentially 'program' VEGFR2 trafficking and turnover that subsequently impacts on signal transduction and endothelial cell responses. By combining biochemical and cell biological approaches, our study finds that three different VEGF-A isoforms (VEGF-A_{165}, VEGF-A_{121} and VEGF-A_{145}) stimulate different patterns of VEGFR2 phosphorylation and internalization into early endosomes, which subsequently impact on downstream signal transduction events. Furthermore, such activated VEGFR2 polypeptides exhibit distinct patterns of ubiquitylation and proteolysis. Our work now shows that VEGF-A isoform-specific programming of VEGFR2 function is dependent on a combination of post-translational modifications linked to residence time within different compartments along the endocytic route.

RESULTS

VEGF-A isoforms promote differential signal transduction and endothelial responses

VEGF-A binding to VEGFR2 activates multiple signal transduction pathways (e.g. ERK1/2, Akt and p38 MAPK) with evidence of VEGF-A isoform-specific cellular responses (Fearnley et al., 2015, 2014a; Kawamura et al., 2008b; Pan et al., 2007). Such intracellular signaling is dependent on VEGFR2 tyrosine phosphorylation on cytoplasmic residues such as Y951, Y1054, Y1059, Y1175 and Y1214 (Koch et al., 2011; Smith et al., 2016). To test the idea that VEGF-A isoforms trigger differential VEGFR2 activation, we monitored the presence of such VEGFR2 phosphotyrosine-epitopes in human umbilical vein endothelial cells (HUVECs) in response to stimulation with different VEGF-A isoforms (1.25 nM; 0-20 min) using immunoblot analysis (Fig. 1A). Quantification of these immunoblot data revealed that these three VEGF-A isoforms had differential capacities to promote the appearance of the VEGFR2-pY1175 epitope (Fig. 1B). However, another VEGFR2 phosphotyrosine epitope, pY1214, showed relatively similar profiles in response to VEGF-A isoform stimulation (Fig. 1C). Surprisingly, we discovered that significant levels of VEGFR2-pY1214 already existed in non-stimulated endothelial cells; furthermore, there was a relatively modest ~2-fold rise in VEGFR2-pY1214 levels in response to any of the VEGF-A isoforms tested (Fig. 1A,C). Notably, the kinetics of VEGFR2-pY1214 levels displayed a more sustained profile (Fig. 1C), suggesting a more long-lived regulatory function.

Such findings raised the question whether other VEGFR2 phosphotyrosine epitopes exhibit distinct or different kinetic profiles. To evaluate this aspect, we monitored the appearance and kinetics of the VEGFR2-pY951, VEGFR2-pY1054 and VEGFR2-pY1059 epitopes using site-specific antibodies (Fig. 1A). All three VEGFR2 phosphotyrosine epitopes exhibited similar kinetics and profiles in response to stimulation with the VEGF-A_{165} isoform (Fig. S1A-C). Further analysis of these VEGFR2 phosphotyrosine epitopes in response to VEGF-A_{121} and VEGF-A_{145} isoform stimulation revealed ~2-5-fold reduced signals with subtle profile differences (Fig. S1A-C). To examine signaling events downstream of VEGFR2-pY1175, we analyzed the phosphorylation and subsequent activation of Akt and ERK1/2 (Koch et al., 2011) (Fig. 1D). We found that the VEGF-A_{165} isoform promoted the highest increase in either Akt (Fig. 1E) or ERK1/2 (Fig. 1F) phosphorylation in comparison to the other VEGF-A isoforms. However, we noted that although VEGF-A_{121} was more effective (than VEGF-A_{145}) in elevating VEGFR2-pY1175 (Fig. 1B) and Akt-pS473 (Fig. 1E) levels, both VEGF-A_{121} and VEGF-A_{145} caused relatively similar levels of phospho-ERK1/2 (Fig. 1F). One possible explanation for such effects is that VEGF-A isoform-specific programming of VEGFR2 phosphotyrosine epitopes is not the sole event(s) in controlling downstream signal transduction and subsequent cellular response(s).

In endothelial cells, VEGF-A-stimulated signal transduction through the Akt and ERK1/2 pathways is linked to increased cell permeability and proliferation respectively (Koch et al., 2011; Lal et al., 2001; Pedram et al., 1998; Six et al., 2002; Takahashi et al., 2001). To determine whether VEGF-A isoform-specific signal transduction differentially regulates such cellular responses, endothelial cells were subjected to VEGF-A isoform stimulation prior to the assessment of endothelial cell permeability and proliferation (Fig. 1G,H). VEGF-A_{165} and VEGF-A_{121} stimulation promoted ~1.8-fold increase in endothelial monolayer permeability, and these effects were significantly higher than that observed upon VEGF-A_{145} stimulation (Fig. 1G). Analysis of endothelial cell proliferation upon VEGF-A_{165} isoform stimulation revealed an ~2-fold increase, which was consistently higher (~25-30%) than that observed upon either VEGF-A_{121} or VEGF-A_{145} stimulation (Fig. 1H). Additionally, VEGF-A_{121} and VEGF-A_{145} were comparable in their capacity to promote endothelial cell proliferation (Fig. 1H). We also compared the ability of these VEGF-A isoforms to stimulate endothelial cell migration, which revealed that all three VEGF-A isoforms caused ~2-fold increase in cell migration with no isoform-specific effects (Fig. S2A,B). Other unique VEGF-A-stimulated cellular responses include the endothelial capacity to form tubules (tubulogenesis) or sprouts from arterial slices. Here, stimulation with the VEGF-A_{165} isoform was clearly the most effective at promoting endothelial tubulogenesis (Fig. S2C,D) and aortic sprouts (Fig. S2E,F) compared to the other two VEGF-A isoforms. Taken together, such data supports the idea that unique features encoded by each VEGF-A isoform enables the programming of specific patterns of VEGFR2 activation linked to downstream signal transduction events, which regulate endothelial cell responses.

VEGF-A isoforms cause differential plasma membrane-to-endosome trafficking of VEGFR2

Recent studies have highlighted the importance of plasma membrane-to-endosome trafficking in regulating VEGFR2 activation and downstream signal transduction (Gaengel and Betsholtz, 2013; Gourlaouen et al., 2013; Lanahan et al., 2013, 2010; Nakayama et al., 2013). Our studies revealed that VEGF-A isoforms have unique properties to stimulate VEGFR2 activation and downstream signal transduction (Fig. 1); therefore, we hypothesized that different VEGF-A isoforms could differentially regulate VEGFR2 endocytosis. To test this idea, we assessed these VEGF-A isoforms for their effects on the pools of VEGFR2 located at the cell surface versus internal compartments using cell surface biotinylation, affinity enrichment and quantitative immunoblotting (Fig. 2A-C). Furthermore, we monitored the presence of VEGFR2-pY1175 within the biotinylated VEGFR2 pool at the cell surface (Fig. 2A). Quantification of these immunoblot data revealed that as expected, all three VEGF-A isoforms promoted a significant rise in cell surface VEGFR2-pY1175 levels, peaking at either 5 min (VEGF-A_{165} and VEGF-A_{121}) or 15 min (VEGF-A_{145}) respectively (Fig. 2B). Interestingly, cell surface VEGFR2-pY1175 levels were significantly decreased after 30 min of VEGF-A_{165} or VEGF-A_{145}
stimulation (Fig. 2B). In contrast, VEGF-A121 caused a more long-lived pool of VEGFR2-pY1175 at the cell surface that persisted for at least 30 min (Fig. 2B). Both VEGF-A165 and VEGF-A145 produced similar kinetics of VEGFR2 endocytosis at the plasma membrane; however, VEGF-A121-stimulated internalization of VEGFR2 was clearly slower (Fig. 2C). These data suggest that these VEGF-A isoforms have differential abilities in not only promoting VEGFR2 activation but also causing plasma membrane-to-endosome trafficking.

To assess the localization of VEGFR2 in intracellular compartments in response to VEGF-A isoform stimulation, we used quantitative immunofluorescence microscopy to monitor VEGFR2 accumulation within early endosomes (Fig. 2D). By evaluating VEGFR2 co-distribution with an early endosome marker (EEA1), we discovered that either VEGF-A165 or VEGF-A145 caused a similar ~30% increase in VEGFR2 accumulation within this compartment (Fig. 2E). In contrast, VEGF-A121 stimulation did not cause a significant increase in VEGFR2 co-distribution with EEA1 (Fig. 2E). Taken together, these data strongly support the view that VEGF-A isoforms have distinct properties in programming VEGFR2 plasma membrane-to-endosome trafficking.

Fig. 1. VEGF-A isoforms promote differential VEGFR2 phosphorylation and downstream signal transduction. (A) Endothelial cells were stimulated with either VEGF-A165, VEGF-A121 or VEGF-A145 (1.25 nM) for 2.5, 5, 10 or 20 min before lysis and processing for immunoblot analysis using site-specific phospho-antibodies against VEGFR2. (B,C) Quantification of VEGFR2-pY1175 (B) and VEGFR2-pY1214 (C) levels upon VEGF-A isoform stimulation. (D) Endothelial cells were stimulated with either VEGF-A165, VEGF-A121 or VEGF-A145 (1.25 nM) for 5, 15, 30 or 60 min before lysis and processing for immunoblot analysis of signal transduction. (E,F) Quantification of Akt-pS473 (E) and ERK1/2-pT202/pY204 (F) levels upon VEGF-A isoform stimulation. (G,H) Endothelial cells were seeded into cellular assays to assess endothelial cell permeability by measuring trans-endothelial electrical resistance (TEER) (G) or proliferation (H) upon control (Con) or VEGF-A165 (165), VEGF-A121 (121) or VEGF-A145 (145; 1.25 nM) stimulated conditions for 4 or 24 h respectively. Error bars indicate ±s.e.m. (n≥4). *P<0.05, **P<0.01, ***P<0.001.
VEGF-A isoforms program differential VEGFR2 ubiquitylation and proteolysis

VEGF-A binding to VEGFR2 triggers distinct patterns of ubiquitylation, proteolysis and clearance within the endosome-lysosome system (Bruns et al., 2010, 2012; Ewan et al., 2006; Smith et al., 2016). From our findings that VEGF-A isoforms had distinct properties in regulating VEGFR2 endocytosis at the plasma membrane, we predicted that VEGF-A-stimulated VEGFR2 degradation was isoform-specific. We confirmed that this was indeed true: VEGF-A121 showed negligible effects on VEGFR2
degradation (Fig. 3A,B); however, VEGF-A165 or VEGF-A145 showed a similar ~30% reduction in total VEGFR2 levels after 120 min (Fig. 3B).

Previous studies have shown that VEGFR2 undergoes 26S proteasome-dependent proteolysis (~30 min post-stimulation) resulting in the appearance of ~160 kDa VEGFR2-derived proteolytic fragment (Bruns et al., 2010). In this context, one possibility was that VEGF-A isoforms programmed different patterns of VEGFR2 proteolysis. To test this idea, we subjected endothelial cells to VEGF-A isoform stimulation for 30 or 120 min and assessed VEGFR2 proteolysis by monitoring the presence of this 160 kDa proteolytic fragment using immunoblotting.
VEGFR2 proteolysis (Fig. 3E). We found that VEGF-A165 stimulation promoted complexes for their ubiquitylation status via immunoblotting isoforms stimulation, immunoisolated VEGFR2 and probed these this idea, we subjected endothelial cells to different VEGF-A properties of each VEGF-A isoform to program distinct patterns of VEGFR2 ubiquitylation, which is implicated in targeting VEGFR2 does not immediately correlate with a reduction in proteolysis in endosomes. VEGF-A-stimulated VEGFR2 activation triggers increased receptor ubiquitylation, which is implicated in targeting VEGF2 for proteolysis and degradation (Bruns et al., 2010; Ewan et al., 2006; Zhang et al., 2010). One possible explanation for the unique properties of each VEGF-A isoform to program distinct patterns of VEGFR2 proteolysis and degradation could be through programming specific patterns of VEGFR2 ubiquitylation. To test this idea, we subjected endothelial cells to different VEGF-A isoforms stimulation, immunoisolated VEGFR2 and probed these complexes for their ubiquitylation status via immunoblotting (Fig. 3E). We found that VEGF-A165 stimulation promoted maximal VEGFR2 ubiquitylation (Fig. 3E) corresponding to ~3.5-fold increase (Fig. 3F). This ubiquitylation signal was significantly greater than that caused by either VEGF-A121 or VEGF-A145 stimulation (Fig. 3E,F). Interestingly, although VEGF-A121 stimulation caused significant VEGFR2 ubiquitylation with a different kinetic profile (Fig. 3F), VEGFR2 degradation was greatly reduced (Fig. 3B). Contrastingly, a further finding is that although VEGF-A145 promoted significant VEGFR2 degradation (Fig. 3B), it did not cause a corresponding increase in VEGFR2 ubiquitylation (Fig. 3E). Based on these data, we conclude that VEGFR2 ubiquitylation is not a prerequisite for degradation but could be required for proteosome-regulated VEGFR2 cleavage on endosomes.

**Disruption of clathrin-dependent VEGFR2 trafficking, results in the loss of isoform-specific signal transduction**

Clathrin is an important structural protein that regulates clathrin-dependent endocytosis at the plasma membrane (Robinson, 2015). This endocytic route is further accessed by VEGFR2 complexes upon VEGF-A stimulation (Bruns et al., 2010; Ewan et al., 2006; Lampugnani et al., 2006). We hypothesized that targeted disruption of clathrin-dependent endocytosis of VEGFR2 would perturb VEGF-A isoform-specific signal transduction. To test this idea, we depleted clathrin heavy chain (CHC17) levels in endothelial cells using duplex siRNAs directed at the mRNA. Such treatment caused a substantial reduction in CHC17 protein levels (Fig. 4A). Unexpectedly, depletion of CHC17 and perturbation of clathrin-dependent endocytosis caused a substantial (>2-fold) decrease in VEGFR2-pY1175 and VEGFR2-pY1214 levels caused by stimulation with any of the VEGF-A isoforms (Fig. 4A–C). This reduced activation was also evident upon monitoring VEGF-A isoform-mediated activation of either Akt (Fig. 4A,D) or ERK1/2 enzymes (Fig. 4A,E). One major consequence occurring upon CHC17 depletion is a decrease in steady-state VEGFR2 levels (Fig. 4A). Hence, one likely explanation for the reduction in VEGFR2 activation and downstream signal transduction upon clathrin depletion is a corresponding and substantial depletion of VEGFR2 levels.

One question that was raised is whether other VEGFR2-regulated signal transduction pathways are also affected by depletion of CHC17 and perturbation of clathrin-dependent endocytosis. Therefore, since different VEGF-A isoforms also cause differential activation of p38 MAPK (Fearnley et al., 2015, 2014a; Kawamura et al., 2008a), we analyzed effects of CHC17 depletion on this signal transduction pathway (Fig. 4F). Surprisingly and in contrast to previous findings, CHC17 depletion did not significantly affect the activation of the p38 MAPK pathway (Fig. 4F). Quantification of these data showed no significant differences in VEGF-A isoform-specific p38 MAPK activation between control or CHC-17-depleted conditions (Fig. 4G). Notably, the relative magnitude of VEGF-A isoform-specific p38 MAPK activation was also retained (Fig. 4G), in spite of the substantial reduction in VEGFR2 levels upon CHC17 depletion (Fig. 4F). One conclusion is that clathrin-dependent endocytosis regulates VEGF-A isoform-specific VEGFR2-mediated turnover and signal transduction via Akt and ERK1/2 signal transduction pathways; however, VEGF-A-stimulated p38 MAPK signal transduction likely involves a VEGFR2 pool linked to a different membrane trafficking pathway.

To evaluate the effects of perturbation of clathrin-dependent endocytosis on the turnover of different membrane proteins, we depleted clathrin heavy chain (CHC17) and examined the levels of VEGF-R2, VEGFR1 and NRP1 using immunoblotting (Fig. 5A). As controls, we evaluated the turnover of the endosome-associated transferrin receptor (TIR) and the trans-Golgi network marker (TGN46) which are known to undergo recognition by clathrin-associated machinery, endocytosis and delivery to endosomes (Banting et al., 1998; Huang et al., 2004; Murphy et al., 2008; Owen and Evans, 1998). Quantification of these data showed that CHC17 depletion (~70% reduction) caused ~60% reduction in VEGFR2 and ~40% reduction in VEGFR1 levels (Fig. 5B). Interestingly, although CHC17 depletion caused a reduction in membrane VEGFR1 levels, soluble VEGFR1 levels were not significantly affected (Fig. 5A,B). Furthermore, depletion of CHC17 and perturbation of clathrin-dependent endocytosis did not affect the turnover of TIR, NRP1 and TGN46 (Fig. 5A,B). One conclusion from these findings is that clathrin-dependent endocytosis regulates the turnover of mature membrane-bound VEGFR2 and VEGFR1 proteins. However, the turnover of other membrane proteins such as NRP1, TIR and TGN46 are not affected by blocking clathrin-dependent endocytosis.

**DISCUSSION**

Our study shows, for the first time, that different VEGF-A isoforms have unique properties in programming VEGFR2 endocytosis, ubiquitylation and proteolysis. In our model, three different VEGF-A isoforms can bind with similar affinity to the extracellular domain of this RTK (VEGFR2) but differentially program the cytoplasmic domain to acquire post-translational modifications, leading to specific patterns of trafficking and proteolysis (Fig. 6). We now suggest that activated VEGFR2 signified by acquisition of the pY1175 epitope undergoes endocytosis and delivery to early endosomes; such trafficking is essential for VEGF-A isoform-specific activation of the Akt and ERK1/2 signal transduction pathways (Fig. 6). This type of signal transduction is an essential feature of how different VEGF-A isoforms regulate the endothelial response that is central to the control of vascular physiology (Fig. 6).

Key lines of evidence support our conclusion that VEGF-A isoforms differentially stimulate VEGFR2 endocytosis and delivery to early endosomes (Fig. 6). Ligand-dependent VEGFR2 endocytosis was elevated in endothelial cells stimulated with either VEGF-A165 or VEGF-A145. In contrast, VEGF-A121 displayed negligible effects in promoting VEGFR2 endocytosis. One possible explanation lies in VEGF-A isoform-specific
recruitment of VEGF co-receptors such as the neuropilins (NRP1, NRP2), which are implicated in regulating VEGFR signal transduction and trafficking (Ballmer-Hofer et al., 2011; Fearnley et al., 2014a; Herzog et al., 2011; Lanahan et al., 2013; Pan et al., 2007; Zachary, 2014). It has been proposed that both VEGF-A165 and VEGF-A121 can bind NRP1; however, only VEGF-A165 is believed to form a heteromeric complex with VEGFR2 and NRP1 (Koch et al., 2011). Thus differential NRP1 recruitment into a VEGFR2–VEGF-A complex depending on the VEGF-A isoform involved could be a mechanism of programming signaling and trafficking outcomes.

VEGFR2 trafficking is an essential regulatory component of VEGF-A165-stimulated signaling events, with evidence for different signal transduction pathways associated with the plasma membrane...
and early endosomes (Gaengel and Betsholtz, 2013; Gourlaouen et al., 2013; Lanahan et al., 2013, 2010; Nakayama et al., 2013). Our study further extends such ideas as depletion of clathrin heavy chain (CHC17) and perturbation of clathrin-dependent endocytosis impaired Akt and ERK1/2 activation but did not affect the p38 MAPK pathway. Surprisingly, clathrin heavy chain depletion caused ∼60% reduction in steady-state VEGFR2 levels but an explanation for this remarkable effect is not clear. One possibility is that multiple plasma membrane-associated VEGFR2 pools are linked to different trafficking pathways that impact on VEGFR2 recycling and turnover. Here, perturbation of clathrin-dependent endocytosis increases VEGFR2 proteolysis and turnover: increased plasma membrane VEGFR2 accumulation could result with association with clathrin-independent endocytosis pathways in endothelial cells such as caveolae (Mukherjee et al., 2006) and micropinocytosis (Muro et al., 2004). In contrast, a separate VEGFR2 pool that exhibits different long-lived plasma membrane dynamics couples VEGF-A binding to signal transduction via the p38 MAPK pathway. VEGFR2 has been previously documented to be also associated with clathrin-independent endocytic routes (Labrecque et al., 2003). In contrast, a separate VEGFR2 pool that exhibits different long-lived plasma membrane dynamics couples VEGF-A binding to signal transduction via the p38 MAPK pathway. VEGFR2 has been previously documented to be also associated with clathrin-independent endocytic routes (Labrecque et al., 2003). A logical conclusion is that VEGFR2 ubiquitylation is a prerequisite for proteasome-regulated VEGFR2 cleavage on endosomes rather than lysosome-mediated terminal degradation (Fig. 6). This proposed mechanism is further strengthened by a recent study, which shows that VEGFR2 de-

**Fig. 5. Depletion of clathrin heavy chain reduces basal VEGFR2 and mature VEGFR1 levels.** (A) Endothelial cells were subjected to scrambled (Scr) or clathrin heavy chain (CHC17)-specific siRNA duplexes, prior to cell lysis and processing for immunoblot analysis. (B) Quantification of basal VEGFR2, VEGFR1, NRP1, transferrin receptor (TfR), TGN46 and CHC17 levels upon depletion of clathrin heavy chain (CHC17). Error bars indicate ±s.e.m. (n=3). ***P<0.001, ****P<0.0001.
ubiquitylation is functionally coupled to different proteolytic steps on early endosomes (Smith et al., 2016). The selective proteolytic cleavage of the activated and ubiquitylated VEGFR2 complex can modulate communication to the ERK1/2 and Akt signal transduction pathways (Bruns et al., 2010). Thus VEGF-A isoform-specific VEGFR2 proteolysis and/or terminal degradation could be a mechanism to program signal transduction downstream of VEGFR2 in order to control diverse cellular responses.

An important finding in this study is that VEGF-A isoforms cause differential phosphorylation of tyrosine residues within the VEGFR2 cytoplasmic domain. A commonly postulated model is that ligand-induced RTK dimerization enables one RTK polypeptide to trans-phosphorylate the other 'partner' within the complex (Endres et al., 2014; Kovacs et al., 2015; Lemmon and Schlessinger, 2010; Lemmon et al., 2014). This is also postulated to occur here with VEGF-A-stimulated VEGFR2 dimerization promoting trans-autophosphorylation of multiple tyrosine residues. We now find that different VEGF-A isoforms promote distinct patterns of phosphorylation on VEGFR2 cytoplasmic residues Y951, Y1054, Y1059 and Y1175. One unexpected finding is the discovery that a phosphotyrosine epitope, VEGFR2-pY1214, is already present at significant levels in resting or non-stimulated cells. In contrast to other VEGFR2 phosphotyrosine epitopes analyzed in this study, VEGF-A stimulation only caused a modest twofold increase in VEGFR2-pY1214 levels with no evident isoform-specific effects. One possible explanation is that the VEGFR2-pY1214 epitope has a regulatory function(s) in the quiescent RTK state. This idea is supported by transgenic mouse studies where the VEGFR2-Y1175F germline mutation causes embryonic lethality and resembles VEGFA (+/−) mice; however, the VEGFR2-Y1214F mutant mice are viable and fertile (Sakurai et al., 2005). Interestingly, the presence of the VEGFR2-pY1214 epitope is linked to p38 MAPK activation (Koch et al., 2011; Lamalice et al., 2006); however, this view can be challenged by the finding that the VEGF-A165, VEGF-A121 and VEGF-A145 isoforms all cause comparable VEGFR2-pY1214 levels but promote
different levels of p38 MAPK activation. One explanation for this discrepancy is that differential NRP1 recruitment to the VEGFR2–VEGF-A complex is also linked to p38 MAPK activation and blood vessel sprouting (Kawamura et al., 2008a). Thus the VEGF-A_{165} isoform-specific recruitment of NRP1 could account for its capacity to promote increased p38 MAPK signal transduction. We conclude that the two VEGFR2 phosphotyrosine-epitopes, pY1175 and pY1214, have different functional roles within the VEGFR2 complex and act by integrating different aspects of signal transduction, post-translational modifications and trafficking.

An emerging view is based on VEGF-A isoforms having unique properties in programming VEGFR2 endocytosis, phosphorylation, ubiquitylation, proteolysis and terminal degradation in lysosomes (Fig. 6). We now provide an integrated mechanism to explain how different VEGF-A isoforms regulate endothelial cell responses such as cell permeability and proliferation. Our study now provides novel insights into how multiple VEGF-A isoforms bind to the same RTK, yet elicit diverse biochemical and membrane trafficking outcomes that influence the cellular response. A future challenge will be to identify specific cytoplasmic factors that regulate the differences in VEGF-A isoform-mediated VEGF2 trafficking and turnover. Such findings will provide a platform towards new ways of manipulating endothelial cell function in health and disease.

**MATERIALS AND METHODS**

**Antibodies and growth factors**

Antibodies: goat-anti-VEGFR1 (#AF321), goat-anti-VEGFR2 (#AF357), rabbit-anti-phospho-VEGFR2-Y1214 (#AF1766) (R&D Systems, Minneapolis, MN, USA), rabbit-anti-Akt (#9272S), rabbit-anti-phospho-Akt (S473; #4060B), rabbit-anti-ERK1/2 (#9102S), mouse-anti-phospho-ERK1/2 (T202/Y204; #9106S), rabbit-anti-neuropilin 1 (NRP1; #3725S), rabbit-anti-phospho-VEGFR2-Y951 (#4991S), rabbit-anti-phospho-VEGFR2-Y1059 (#3817S), rabbit-anti-phospho-VEGFR2-Y1175 (#2478S; Cell Signaling Technology, Danvers, MA, USA), rabbit-anti-phospho-VEGFR2-Y1054 (Clone D1W; #04-894; Merck Millipore, Watford, UK), mouse-anti-α-tubulin (Clone DM1A; #T6199; Sigma Aldrich, Poole, UK), mouse-anti-PECAM-1 (CD31; #sc-65260; Santa Cruz Biotechnology, Dallas, TX, USA), mouse-anti-ubiquitin (FK2; #14220; Caymen Chemical, MI, USA), mouse-anti-clathrin heavy chain antibody (X22; #ab2731; Abcam, Cambridge, UK), sheep-anti-NGN4 (#AHP509GT; AbD Serotec, Oxford, UK). Endothelial cell growth medium (ECGM) was from PromoCell (Heidelberg, Germany). Recombinant human VEGF-A_{165} was from Genentech Inc. (San Francisco, CA, USA), both VEGF-A_{121} and VEGF-A_{145} was from Promocell. Endothelial cells were seeded at 3×10^4 cells/well into an 8 µm pore size Transwell filter inserted into a 24-well plate (BD Biosciences, Oxford, UK) in ECGM and cultured until a confluent monolayer was formed. Transwells containing endothelial cells were then washed twice, transferred to a fresh well containing 500 µl MCDB131+0.2% (w/v) BSA and starved in 400 µl MCDB131+0.2% (w/v) BSA (added to the top chamber) for 2 h. After 2 h (t=0 h) the trans-endothelial electrical resistance (TEER) across each monolayer was measured using a MILLICELL-ERS TEER machine (Merck Millipore). Following which, 100 µl of MCDB131+0.2% (w/v) BSA containing the desired VEGF-A isoform was added to the upper chamber. After a further 4 h (t=4 h) TEER across each monolayer was measured again and the relative increase in permeability (corresponding to a decrease in electrical resistance across the endothelial monolayer) was calculated.

**Tubulogenesis assay**

Primary human foreskin fibroblasts (Promocell) were cultured until confluent in 48-well plates in Q333 fibroblast growth media (PA Laboratory, Pasching, Austria). 7500 endothelial cells were then seeded per well onto the fibroblasts monolayer in a 1:1 mixture of Q333 and ECGM and left to acclimatize for 24 h. Media was then aspirated and replaced with fresh ECGM+VEGF-A isoform (1.25 nM) as desired; media was replaced every 2-3 days for seven days. Co-cultures were then fixed in 200 µl 10% (w/v) formalin for 20 min and blocked in 5% (w/v) BSA for 20 min. Immunofluorescence microscopy using an EVOS-fl inverted digital imaging microscope (Mannedorf, Switzerland) was performed to examine tubulogenesis.

**Cell culture and immunoblotting analysis**

Endothelial cells were seeded per well of a 96-well plate and left to acclimatize in ECGM overnight. Media was aspirated and cells starved in MCDB131+0.2% BSA (w/v) for 2 h. Cells were stimulated with VEGF-A isoforms (1.25 nM) in 100 µl total volumes for 24 h. 10 µM BrdU was added per well 20 h post-stimulation. A cell proliferation ELISA (Roche Diagnostics, Mannheim, Germany) was performed according to manufacturer’s instructions. The color change was developed using 3,3′,5,5′-tetramethylbenzidine solution and the reaction quenched with 1 M H₂SO₄. Absorbance was measured at 450 nm using a Tecan Sunrise multiwavelength 96-well plate reader (Mannedorf, Switzerland).

**Assessment of endothelial monolayer permeability using trans-endothelial electrical resistance (TEER)**

Human endothelial cells were seeded at 5×10^4 cells/well into a 0.4 µm pore size Transwell filter inserted into a 24-well plate (BD Biosciences, Oxford, UK) in ECGM and cultured until a confluent monolayer was formed. Transwells containing endothelial cells were then washed twice, transferred to a fresh well containing 500 µl MCDB131+0.2% (w/v) BSA and starved in 400 µl MCDB131+0.2% (w/v) BSA (added to the top chamber) for 2 h. After 2 h (t=0 h) the trans-endothelial electrical resistance (TEER) across each monolayer was measured using a MILLICELL-ERS TEER machine (Merck Millipore). Following which, 100 µl of MCDB131+0.2% (w/v) BSA containing the desired VEGF-A isoform was added to the upper chamber. After a further 4 h (t=4 h) TEER across each monolayer was measured again and the relative increase in permeability (corresponding to a decrease in electrical resistance across the endothelial monolayer) was calculated.

**Cell migration assay**

Endothelial cells were seeded at 3×10^4 cells/well onto a 8 µm pore size Transwell filter inserted into a 24-well plate (BD Biosciences) in MCDB131+0.2% (w/v) BSA. MCDB131+0.2% (w/v) BSA containing the desired concentration of VEGF-A was added to the lower chambers to stimulate cell migration. Cells were allowed to migrate for 24 h before being fixed and stained with 0.2% (w/v) crystal violet in 20% (v/v) methanol. Non-migrated cells were removed from the upper chamber using a moist cotton bud; chambers were rinsed using double-distilled water. Three to five random fields were imaged per Transwell filter and the average number of migratory cells calculated.

**Biology Open**
5 M NaOH. 55 μl of this embedding matrix was pipetted per well into a 96 well plate and aortic ring submerged within. Plates were incubated at room temperature for 15 min before incubation at 37°C for 90 min. 150 μl OptiMEM containing 2.5% (v/v) FCS and penicillin-streptomycin was added per well with appropriate VEGF-A isoform (1.25 nM). Aortic rings were incubated at 37°C for 5 days with a media change on day 3. Wells were washed with 150 μl PBS containing 2 mM CaCl₂ and 2 mM MgCl₂, and fixed in 4% formalin for 30 min. The collagen was permeabilized with three 2 mM CaCl₂ and 0.25% (v/v) Triton X-100 before incubation for 2 h with PBS. Cells were lysed in 500 μl RIPA buffer for 1 h at 4°C. Lysates were cleared by centrifugation at 16,000 × g for 5 min. 50 μl of 50:50 Protein G-sepharose slurry (Invitrogen) and allowed to settle at room temperature for 15 min before being returned to the incubator. After 6 h total incubation, media was replaced for ECGM. Cells were allowed to recover for 72 h prior to treatment or processing for analysis. Scrambled siRNA was purchased as a siGENOME SMARTpool from Dharmacon (GE Healthcare, Buckinghamshire, UK).

Cell surface biotinylation
Endothelial cells were stimulated (1.25 nM VEGF-A isoform in MCDB131 + 0.2% (v/v) BSA) before washing twice with ice-cold PBS and incubation with 0.5 mg/ml EZ-Link Sulfo-NHS-LC-Biotin (Thermo Fisher) in PBS containing 2 mM MgCl₂ and 2 mM CaCl₂ for 30 min at 4°C. Biotinylation was quenched by washing twice with ice-cold TBS followed by washing twice with ice-cold PBS. Cells were lysed in 500 μl RIPA buffer for 1 h at 4°C. Lysates were cleared by centrifugation at 16,000 g for 30 min at 4°C. Equivalent protein amounts were incubated with 35 μl neutravidin-agarose beads (Thermo Fisher) overnight at 4°C. Beads were pelleted by brief centrifugation, supernatant removed and beads washed four times with 500 μl ice-cold RIPA buffer. 50 μl of 2× SDS-PAGE sample buffer was added and proteins eluted via heating at 92°C for 10 min before SDS-PAGE and immunoblotting.

Immunofluorescence analysis
For immunofluorescence analysis, endothelial cells were serum starved in MCDB131+0.2% (v/v) BSA for 2 h before being stimulated for 30 min with desired VEGF-A isoform (1.25 nM). Endothelial cells were fixed and processed as previously described (Bruns et al., 2010). Images were acquired either using a wide-field deconvolution microscope DeltaVision (Applied Precision Inc., Issaquah, USA). Relative VEGFR2 co-distribution was quantified using Image J (NIH, Bethesda, USA).

VEGFR2 ubiquitylation analysis
Endothelial cells were stimulated (1.25 nM VEGF-A isoform in MCDB131+0.2% (v/v) BSA; 2 wells per condition) before washing twice with ice-cold PBS and lysed in RIPA buffer (150 mM NaCl, 50 mM Tris-HCl pH 7.4, 0.1% (w/v) SDS, 0.5% (w/v) sodium deoxycholate, 2 mM EDTA, 1% (v/v) NP-40, 50 mM NaF) with freshly added 1 mM phenylmethylsulfonyl fluoride (PMSF) and 10 mM iodoacetamide, and incubated for 5 min on ice at 4°C. Lysates were cleared by centrifugation at 16,000 g for 30 min at 4°C. Equivalent protein concentrations of supernatant were incubated with VEGFR2 antibody for 2 h at 4°C. 35 μl of 50:50 Protein G-sepharose slurry (Millipore, Watford, UK) was added and incubated overnight at 4°C. Beads were pelleted by brief centrifugation, supernatant removed and beads washed four times with 500 μl ice-cold RIPA buffer. 50 μl of 2× SDS-PAGE sample buffer was added and proteins eluted via heating at 92°C for 10 min before SDS-PAGE and immunoblotting. VEGFR2 ubiquitylation was monitored using mouse anti-FK2 antibody which detects both poly- and mono-ubiquitylation.

Lipid-based transfection of siRNA duplexes
Cells were transfected with siRNA duplexes using lipofectamine RNAiMAX (Invitrogen). Per well of a 6-well plate, 15 μl of 2 μM siRNA duplexes was added to 481 μl of serum/antibiotic-free OptiMEM (Invitrogen) and allowed to settle at room temperature for 5 min. 4 μl of lipofectamine was then added and the mixture was inverted briefly and incubated at room temperature for 20 min. HUVECs were seeded at 2.5×10⁵ cells/ml in a 1 ml volume of OptiMEM, followed by immediate dropwise addition of the siRNA/lipofectamine mixture. Cells were left at room temperature for 30 min before being returned to the incubator. After 6 h total incubation, media was replaced for ECGM. Cells were allowed to recover for 72 h prior to treatment or processing for analysis. Scrambled siRNA was purchased as a siGENOME SMARTpool from Dharmacon (GE Healthcare, Buckinghamshire, UK). clathrin heavy chain (CHC17) siRNA was from Ambion (Life Technologies, Paisley, UK).

Statistical analysis
This was performed using a one-way analysis of variance (ANOVA) followed by Tukey’s post-hoc test or two-way ANOVA followed by Bonferroni multiple comparison test using GraphPad Prism software (La Jolla, CA, USA). Significant differences between control and test groups were evaluated with P values less than 0.05 (*), 0.01 (**), 0.001 (***), and 0.0001 (****) indicated on the graphs. Error bars in graphs and histograms denote s.s.c.m. (standard error of mean).

Acknowledgements
The authors would like to thank the members of the Endothelial Cell Biology Unit for their help and advice.

Competing interests
The authors declare no competing or financial interests.

Author contributions
G.W.F., S.B.W. and S.P., conception and design, analysis and interpretation of data; G.W.F. and N.A.M., acquisition of data, contributed essential data and reagents; G.W.F., G.A.S., I.A.-Z., N.Y., S.H.-V., M.T.K., I.C.Z., D.C.T., M.A.H., S.B.W. and S.P., drafting and revising the article.

Funding
This work was supported by a Heart Research UK PhD studentship (TRP11/11 to G.W.F.), a Brunel Government PhD studentship (I.A.-Z.), British Heart Foundation PhD studentship [FS/12/20/29462 to G.A.S.], Circulation Foundation George Davies Surgeon Scientist Award (N.A.M.), British Heart Foundation project grant [PG/14/54/30539 to M.T.K. and S.B.W.] and BHF programme grants [RG/15/7/31521 to M.T.K.] and [RG/11/11/29050 to I.C.Z.]. S.B.W. is the recipient of European Research Council Fellowship [ERC-2012-StG 310747-BP-CardIO].

Supplementary information
Supplementary information available online at http://bio.biologists.org/lookup/suppl/doi:10.1242/bio.017434/-/DC1

References
Baker, M., Robinson, S. D., Lechtiert, T., Barber, P. R., Tavora, B., D’Amico, G., Jones, D. T., Vojnovic, B. and Hodivala-Dilke, K. (2012). Use of the mouse aortic ring assay to study angiogenesis. Nat. Protoc. 7, 89-104.

Ballmer-Hofer, K., Andersson, A. E., Ratcliffe, L. E. and Berger, P. (2011). Neuropilin-1 promotes VEGFR-2 trafficking through Rab11 vesicles thereby specifying signaling output. Blood 118, 816-826.

Banting, G., Maile, R. and Roquemore, E. P. (1998). The steady state distribution of humTN46 is not significantly altered in cells defective in clathrin-mediated endocytosis. J. Cell Sci. 111, 3451-3458.

Bruns, A. F., Herbert, S. P., Odel, A. F., Jolping, H. M., Hooper, N. M., Zachary, I. C., Walker, J. H. and Ponnambalam, S. (2010). Ligand-stimulated VEGFR2 signaling is regulated by co-ordinated trafficking and proteolysis. Traffic 11, 161-174.

Bruns, A. F., Yuldasheva, N., Latham, A. M., Bao, L., Pellet-Macy, C., Frankel, P., Stephen, S. L., Howell, G. J., Wheatcroft, S. B., Kearney, M. T. et al. (2012). A heat-shock protein-10 axis regulates VEGFR2 proteolysis, blood vessel development and repair. PLoS ONE 7, e48539.

Carmeliet, P. (2005). Angiogenesis in life, disease and medicine. Nature 438, 323-336.

Carmeliet, P., Ferreira, V., Breier, G., Pollefeyt, S., Hickens, L., Gertsenstein, M., Fahrig, M., Vandenheoock, A., Harpal, K., Eberhardt, C. et al. (1996).
Abnormal blood vessel development and lethality in embryos lacking a single VEGF allele. Nature 380, 435-439.

Cohen, T., Gitay-Goren, H., Sharon, R., Shibuya, M., Halaban, R., Levi, B. Z. and Neufeld, G. (1995). VEGF121, a vascular endothelial growth factor (VEGF) isoform lacking of heparin binding ability, requires cell-surface heparan sulfates for efficient binding to the VEGF receptors of human melanoma cells. J. Biol. Chem. 270, 11322-11326.

Endres, N. F., Barros, T., Cantor, A. J. and Kuriyan, J. (2014). Emerging concepts in the regulation of the EGF receptor and other receptor tyrosine kinases. Trends Biochem. Sci. 39, 435-446.

Ewan, L. C., Jopling, H. M., Jia, H., Mittar, S., Bagherzadeh, A., Howell, G. J., Walker, J. H., Zachary, I. C. and Ponnambalam, S. (2006). Intrinsinc tyrosine kinase activity is required for vascular endothelial growth factor receptor 2 ubiquitination, sorting and degradation in endothelial cells. Traffic 7, 1270-1282.

Fearnley, G. W., Odell, A. F., Latham, A. M., Mughal, N. A., Bruns, A. F., Bursenka, N. J., Zachary, I., Hollman, M. C., Wheatcroft, S. B. et al. (2014a). VEGF-A isoforms differentially regulate ATF-2-dependent VCAM-1 gene expression and endothelial-leucocyte interactions. Mol. Biol. Cell. 25, 2509-2521.

Fearnley, G. W., Smith, G. A., Odell, A. F., Latham, A. M., Wheatcroft, S. B., Harrison, M. A., Tomlinson, D. C. and Ponnambalam, S. (2014b). Vascular endothelial growth factor a-stimulated signaling from endosomes in primary endothelial cells. Meth. Enzymol. 535, 265-292.

Fearnley, G. W., Smith, G. A., Harper, S. J., Bates, D. O. and Claesson-Welsh, L. (2011). Signal transduction by vascular endothelial growth factor receptors. Cold Spring Harb. Perspect. Med. 2, 3638-3649.

Huang, F., Khvorova, A., Marshall, W. and Sorkin, A. (2008). NRP1 in regulation of VEGF-induced activation of p38MAPK and VEGF receptor-2 due to lack of coreceptor binding and deficient regulation of membrane recycling. Ponnambalam, S. (2009). Rab GTPase regulation of VEGFR2 trafficking and angiogenesis by Golgi endocytic trafficking regulates arterial morphogenesis. Dev. Cell. 18, 713-724.

Kawamura, H., Zhang, X., Fujin, A., Zhuang, Z., Rivera-Molina, F., Speichinger, K., Prahst, C., Zhang, J., Wang, Y., Davis, G. et al. (2013). The neuropilin 1 cytoplasmic domain is required for VEGF-A-dependent arteriogenesis. Dev. Cell 25, 156-168.

Kawamura, H., Chaudhary, C., Zhang, X., Zhuang, Z., Eichmann, A. and Simons, M. (2014). PTP1b is a physiologic regulator of vascular endothelial growth factor signaling in endothelial cells. Circulation 130, 902-909.

Koch, S., Tugues, S., Li, X., Gualandi, L. and Claesson-Welsh, L. (2015). Emerging concepts on the regulation of the EGF receptor and other receptor tyrosine kinases. Trends Biochem. Sci. 40, 743-756.

Koval, M. and Muzykantov, V. (2004). Endothelial endocytic pathways: gates for vascular drug delivery. Curr. Vasc. Pharmacol. 2, 281-299.

Lamalice, L., Houle, F. and Huot, J. (2006). Phosphorylation of Tyr 1214 within VEGF-R2 triggers the recruitment of Nick and activation of Fyn leading to SAPK2/p38 activation and endothelial cell migration in response to VEGF. J. Biol. Chem. 281, 34009-34020.

Lampugnani, M. G., Orsengino, F., Gagliani, M. C., Tacchetti, C. and Dejana, E. (2006). Vascular endothelial cadherin controls VEGFR-2 internalization and signaling from intracellular compartments. J. Cell Biol. 174, 993-1004.

Lanahan, A. A., Hermans, M. J., Shaik, S., Nucera, C., Inuzuka, H., Gao, D., Garnaas, M., Frechette, G., Harris, L., Meth. Enzymol.

Lanahan, A. A., Zhuang, Z., Wang, Y., Davis, G. et al. (2013). The neuropilin 1 cytoplasmic domain is required for VEGF-A-dependent arteriogenesis. Dev. Cell 25, 156-168.

Lanahan, A. A., Dech, D., Dubrac, A., Zhang, J., Zhuang, Z. W., Eichmann, A. and Simons, M. (2014). PTP1b is a physiologic regulator of vascular endothelial growth factor signaling in endothelial cells. Circulation 130, 902-909.

Lanahan, A. A., Schlessinger, J. and Ferguson, K. M. (2014). The EGFR family: not so prototypical receptor tyrosine kinases. Cold Spring Harb. Perspect. Biol. 6, a020768.

Lemmon, M. A., Schlessinger, J. (2010). Cell signaling by receptor tyrosine kinases. Cell 141, 1117-1134.

Lemmon, M. A., Schlessinger, J. and Ferguson, K. M. (2014). The EGFR family: not so prototypical receptor tyrosine kinases. Cold Spring Harb. Perspect. Biol. 6, a020768.

Morgan, D. C., Stowers, T. A., Verma, V. N., Srivastava, A., Chang, B. and Liu, X. (2014). Essential role for endocytosis in the growth factor-stimulated activation of ERK1/2 in endothelial cells. J. Biol. Chem. 288, 7467-7480.

Murphy, J. E., Vohra, R. S., Dunn, S., Holloway, Z. G., Monaco, A. P., Hommer-Vanlaarsinkam, S., Walker, J. H. and Ponnambalam, S. (2008). Oxidised LDL internalisation by the LOX-1 scavenger receptor is dependent on a novel cytoplasmic motif and is regulated by dynamin-2. J. Cell Sci. 121, 2136-2147.

Nakayama, M., Nakayama, A., van Lessen, M., Yamamoto, H., Hoffmann, S., Drexler, H. C. A., Itoh, N., Hirose, T., Breier, G., Westever, D. et al. (2013). Spatial regulation of VEGF receptor endocytosis in angiogenesis. Nat. Cell Biol. 15, 233-243.

Owen, D. J. and Evans, P. R. (1998). A structural formulation for the recognition of tyrosine-based endocytic signals. Science 282, 1327-1332.

Pan, Q., Chattergy, A., Liu, K., Wang, Y., Liu, F. X., Bae, J. H., Hong, S. et al. (2012). Scf(beta-TRCP) suppresses VEGFR2 receptor tyrosine kinase endocytosis and regulates endothelial cell migration and sprouting. J. Biol. Chem. 287, 14170-14181.

Pedram, A., Razandi, M. and Levin, E. R. (2008). Extracellular signal-regulated protein kinase-Jun kinase cross-talk regulates vascular endothelial cell growth factor-induced endothelial cell proliferation. J. Biol. Chem. 283, 26722-26728.

Pedram, A., Razandi, M. and Levin, E. R. (2008). Extracellular signal-regulated protein kinase-Jun kinase cross-talk regulates vascular endothelial cell growth factor-induced endothelial cell proliferation. J. Biol. Chem. 283, 26722-26728.

Pedram, A., Razandi, M. and Levin, E. R. (2008). Extracellular signal-regulated protein kinase-Jun kinase cross-talk regulates vascular endothelial cell growth factor-induced endothelial cell proliferation. J. Biol. Chem. 283, 26722-26728.

Robinson, M. S. (2015). Forty years of clathrin-coated vesicles. Traffic 16, 1210-1238.

Sakurai, Y., Ohigmot, K., Kataoka, Y., Yoshida, N. I. and Shirabe, M. (2005). Essential role of Flik-1 (VEGF receptor 2) tyrosine residue 1173 in vasculogenesis in mice. Oncogene 24, 4373-4384.
factors requires co-ordinated signal transduction, trafficking and proteolysis. 

Smith, G. A., Fearnley, G. W., Abdul-Zani, I., Wheatcroft, S. B., Tomlinson, D. C., Harrison, M. A. and Ponnambalam, S. (2016). VEGFR2 trafficking, signaling and proteolysis is regulated by the Ubiquitin Isopeptidase USP8. Traffic 17, 53-65.

Takahashi, T. and Shibuya, M. (1997). The 230 kDa mature form of KDR/Flk-1 (VEGF receptor-2) activates the PLC-gamma pathway and partially induces mitotic signals in NIH3T3 fibroblasts. Oncogene 14, 2079-2089.

Takahashi, T., Yamaguchi, S., Chida, K. and Shibuya, M. (2001). A single autophosphorylation site on KDR/Flk-1 is essential for VEGF-A-dependent activation of PLC-gamma and DNA synthesis in vascular endothelial cells. EMBO J. 20, 2768-2778.

Tillo, M., Erskine, L., Cariboni, A., Fantin, A., Joyce, A., Denti, L. and Ruhrberg, C. (2015). VEGF189 binds NRP1 and is sufficient for VEGF/NRP1-dependent neuronal patterning in the developing brain. Development 142, 314-319.

Wong, C. and Jin, Z.-G. (2005). Protein kinase C-dependent protein kinase D activation modulates ERK signal pathway and endothelial cell proliferation by vascular endothelial growth factor. J. Biol. Chem. 280, 33262-33269.

Xu, D., Fuster, M. M., Lawrence, R. and Esko, J. D. (2011). Heparan sulfate regulates VEGF165- and VEGF121-mediated vascular hyperpermeability. Journal Biol. Chem. 286, 737-745.

Yamada, K. H., Nakajima, Y., Geyer, M., Wary, K. K., Ushio-Fukai, M., Komarova, Y. and Malik, A. B. (2014). KIF13B regulates angiogenesis through Golgi to plasma membrane trafficking of VEGFR2. J. Cell Sci. 127, 4518-4530.

Zachary, I. (2014). Neuropilins: role in signalling, angiogenesis and disease. Chem. Immunol. Allergy 99, 37-70.

Zhang, Z., Neiva, K. G., Lingen, M. W., Ellis, L. M. and Nör, J. E. (2010). VEGF-dependent tumor angiogenesis requires inverse and reciprocal regulation of VEGFR1 and VEGFR2. Cell Death Differ. 17, 499-512.

Zhang, X., Lanahan, A. A. and Simons, M. (2013). VEGFR2 trafficking: speed doesn’t kill. Cell Cycle 12, 2163-2164.