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A Point Mutation in the Hair Cell Nicotinic Cholinergic Receptor Prolongs Cochlear Inhibition and Enhances Noise Protection

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The transduction of sound in the auditory periphery, the cochlea, is inhibited by efferent cholinergic neurons projecting from the brainstem and synapsing directly on mechanosensory hair cells. One fundamental question in auditory neuroscience is what role(s) this feedback plays in our ability to hear. In the present study, we have engineered a genetically modified mouse model in which the magnitude and duration of efferent cholinergic effects are increased, and we assess the consequences of this manipulation on cochlear function. We generated the Chrna9L9T line of knockin mice with a threonine for leucine change (L9T) at position 9 of the second transmembrane domain of the α9 nicotinic cholinergic subunit, rendering α9-containing receptors that were hypersensitive to acetylcholine and had slower desensitization kinetics. The Chrna9L9T allele produced a 3-fold prolongation of efferent synaptic currents in vitro. In vivo, Chrna9L9T mice had baseline elevation of cochlear thresholds and efferent-mediated inhibition of cochlear responses was dramatically enhanced and lengthened: both effects were reversed by strychnine blockade of the α9×10 hair cell nicotinic receptor. Importantly, relative to their wild-type littermates, Chrna9L9T/α9T mice showed less permanent hearing loss following exposure to intense noise. Thus, a point mutation designed to alter α9×10 receptor gating has provided an animal model in which not only is efferent inhibition more powerful, but also one in which sound-induced hearing loss can be restrained, indicating the ability of efferent feedback to ameliorate sound trauma.

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Introduction

In bringing information about the world to an individual, sensory systems perform a series of common functions. Each system responds with some specificity to a stimulus, and each one employs some specialized receptor cells at the periphery to translate specific stimuli into electrical signals that all neurons can use. That initial electrical event begins the process by which the central nervous system constructs an orderly representation of, for example, sounds, odors, tastes, and objects. Thus, basic sound detection begins when sound waves strike the eardrum, which transmits that physical stimulus to the organ of Corti within the cochlea, the sensory epithelium of the mammalian inner ear. Here, the primary receptor cells known as inner hair cells (IHCs) transform the information into electrical signals that are sent to the central nervous system by the auditory nerve [1]. However, unlike vision, touch, and the chemical senses, sound processing is modulated by efferent signals that travel in reverse, from the brain back to the inner ear [2]. One fundamental question in auditory neuroscience is what role(s) this feedback plays in our ability to hear.

The medial olivocochlear (MOC) efferents (Figure 1A) originate in the medial portion of the superior olivary complex and project to outer hair cells (OHCs; Figure 1B) of the organ of Corti, where large synaptic contacts are formed [2]. In contrast to IHCs, OHCs act as biological motors to amplify the sound-evoked motions of the organ of Corti through a type of somatic electromotility generated by a specialized membrane protein known as prestin. Activation of the MOC pathway, either by sound or by shock trains delivered to the bundle at the floor of the IVth ventricle (Figure 1A), reduces cochlear sensitivity through the action of the neurotransmitter acetylcholine (ACh) on nicotinic re-
Author Summary

Nicotinic cholinergic receptors are essential to higher order brain function. Structurally, these operate through a myriad of ligand-gated pentameric arrangements of different homologous subunits. Here, we report progress in understanding the structural properties of a neuronal nicotinic receptor at the synapse. Receptors assembled from two nicotinic cholinergic subunits (α9 and α10) serve exclusively at the synapse between central nervous system descending fibers and cochlear hair cells. This enabled us to show direct causality between a point mutation of the α9 subunit, and predicted alterations in the synaptic strength in sensory hair cells of the cochlea of α9 point mutant mice. Furthermore, this single mutation results in profound enhancement of central nervous system feedback to the cochlea. And finally, as a consequence, mutant mice possessing this altered receptor have substantially improved resistance to traumatic sound. Thus, central neuronal feedback on cochlear hair cells provides an opportunity to define one specific role that nicotinic receptors can play in the nervous system, enabling study from biophysical to behavioral levels and promoting a target for the prevention of noise-induced hearing loss.

Figure 1. Schematics Showing the Central Origin (A) and Peripheral Projections (B) of the MOC Fibers and the Cholinergic Synapse onto OHCs in the Mature Organ of Corti (C)

MOC efferent neurons are located in the superior olivary complex of the brainstem and project to the cochlea, where they make direct synaptic contacts at the base of the OHCs. At this synapse, ACh is released. It binds to α9x10 receptors present at the OHCs, leading to Ca2+ influx and the subsequent opening of small conductance Ca2+-activated K+ SK2 channels, thus leading to hyperpolarization of hair cells [3,16,17]. (Figure 1C).

In the present study, we have generated a genetically modified mouse model in which the magnitude and duration of the MOC efferent effect is increased, and assessed the consequences of this manipulation on auditory thresholds and susceptibility to noise-induced hearing loss. We have substituted a threonine for a leucine at position 9T (L9T) of the second transmembrane domain of the α9 subunit and examined the neurotransmitter responses, synaptic efficiency, and cochlear function of Chrna9<sup> wt/L9T </sup>, Chrna9<sup> L9T/wt</sup>, and Chrna9<sup>L9T/T9L9T</sup> mice. We show that the L9T mutation produced an increase in sensitivity to ACh, and decreased rates of ACh-induced desensitization. More significantly, synaptic currents were dramatically prolonged in the OHCs of Chrna9<sup>L9T/T9L9T</sup> mice. Consistent with these effects, Chrna9<sup>L9T/T9L9T</sup> mice had elevated acoustic thresholds, and shock-evoked MOC activation produced both enhanced and prolonged cochlear suppression. This enhanced efferent inhibitory drive attenuated sound-induced, permanent acoustic injury. Our results establish efferent feedback inhibition's function in preventing acoustic trauma, and provide preliminary characterization of a mouse model to explore additional physiological functions of MOC innervation. In addition, it motivates further exploration of the efferent synapse as a model system to study fast neurotransmission mediated by a neuronal nAChR.

Results

The targeting construct and generation of Chrna9L9T mutant mice are described in the Materials and Methods section and Figure 2. Both Chrna9<sup> wt/L9T </sup>and Chrna9<sup>L9T/T9L9T</sup> mutant mice were viable through adulthood, fertile, showed the expected gender and Mendelian genotype ratios, and exhibited no overt behavioral phenotype. Gross morphology of the cochlear duct was normal as seen in light microscopic evaluation of plastic sections of osmium-stained cochleae.
As assessed by quantitative real-time PCR (RT-PCR), the expression level of cochlear nAChR $\alpha_9$ subunit mRNA was similar in Chrna9wt/wt and Chrna9L9T mice (Table S1). In addition, the expression levels of genes encoding proteins known to localize to the olivocochlear (OC) synaptic complex (for example, the nAChR $\alpha_{10}$ subunit and SK2 channel) were not changed in mutant mice (Table S1). Lastly, the electrophysiology of IHCs and OHCs was normal, as indicated by similar voltage-dependent K$^+$ currents (Figure S2). Hair cell membrane area measured by capacitance was identical across phenotypes (Chrna9wt/wt: 8.3 ± 0.3 pF, n = 5 animals, 13 OHCs; Chrna9L9T: 8.1 ± 0.2 pF, n = 8 animals, 19 OHCs, measured at P10–P11).

Hypersensitive and Slowly Desensitizing nAChRs in Mutant Mice

In Xenopus oocytes, $\alpha_9\alpha_{10}$ nAChRs assembled from mutant $\alpha_9\beta_9\gamma_9\delta_9T$ subunits exhibit an increased sensitivity to ACh, a reduced desensitization rate, and an increased channel mean open time [18]. To determine whether similar changes occurred in hair cells of L9T mutant mice, we examined the response of neonatal IHCs to exogenous ACh. IHC nAChRs, which are expressed transiently before the onset of hearing, are of the $\alpha_9\alpha_{10}$ subtype and are functionally indistinguishable from those that arise later in OHCs [19,20].

Figure 2. Genetic Engineering and Genotyping of Chrna9L9T Mutant Mice

(A) Gene structures and restriction maps for the targeting construct, wild-type, and recombinant alleles (before and after removal of the neomycin resistance cassette) are shown. Black boxes represent the exons of the Chrna9 gene. The introduction of the mutation is shown by an asterisk (*). (B) An autoradiograph of a genomic Southern blot using DNA prepared from wild-type (left lane) and successfully targeted ES cells (right lane) and hybridized to a $^{32}$P-labeled DNA probe prepared from the 962-bp SacI-KpnI fragment shown. Wild-type (WT) ES cell DNA yield a fragment of 13,800 bp, whereas ES cells that have undergone homologous recombination yield a 7,300-bp fragment. (C) Routine genotyping of neo-deleted Chrna9L9T mutant mice performed by PCR from isolated tail biopsy tissue genomic DNA samples and amplimers flanking the lox P footprint (white box). A representative gel indicating the results for wild-type, heterozygous, and homozygous knockin mice is shown. The PCR fragment length for the wild-type Chrna9 allele is 203 bp, and 269 bp for the mutant allele. MW, molecular weight. (D) A PCR fragment using amplimers flanking the L9T mutation (Figure 1A) was obtained from genomic DNA isolated from tail biopsies and sequenced to check the integrity of the mutation in each breeding pair. A representative electropherogram indicating the results for wild-type and homozygous knockin mice is shown. The wild-type sequence at nucleotide position 210, corresponding to the codon at position 9 in the second transmembrane domain, is TTG, encoding leucine; the mutant sequence is ACG, encoding threonine.

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Since recordings are more frequently and reliably obtained from neonatal IHCs than OHCs, the former provided the bulk of the data presented. As shown below, a subset of our findings has been repeated in OHCs of Chrna9L9T mutant mice.

As observed in Figure 3A, IHCs of Chrna9\textsuperscript{wt/\textsuperscript{wt}} and Chrna9\textsuperscript{L9T/L9T} mice responded to ACh at P6–P10. In each case, ACh dose-response curves of IHCs were left-shifted when compared to wild type, with a concomitant decrease in the half maximal effective concentration (EC\textsubscript{50}). The EC\textsubscript{50} was significantly different (p < 0.01) between homozygous mutant and wild-type IHCs (Chrna9\textsuperscript{wt/\textsuperscript{wt}}: EC\textsubscript{50} 90.8 ± 18.2 μM, Hill coefficient 1.6 ± 0.1, n = 4 animals, 7 cells; Chrna9\textsuperscript{L9T/L9T}: EC\textsubscript{50} 57.2 ± 6.5 μM, Hill coefficient 2.2 ± 0.1, n = 4 animals, 9 cells; and Chrna9\textsuperscript{L9T/\textsuperscript{wt}}: EC\textsubscript{50} 41.7 ± 5.7 μM, Hill coefficient 2.1 ± 0.2, n = 4 animals, 7 cells). During prolonged (>60 s) application of 1 mM ACh at −90 mV (Figure 3B), the current was entirely inward since the ACh response was recorded in isolation from the SK2 component (see below). The evoked membrane current in wild-type IHCs decayed with a complex time course. In IHCs of both Chrna9\textsuperscript{wt/\textsuperscript{wt}} and Chrna9\textsuperscript{L9T/L9T} mice, ACh-evoked membrane current was more than that seen in wild-type mice (i.e., showed reduced desensitization, as previously shown with recombinant α9\textsubscript{2}α10 nAChRs expressed in oocytes [18]). The fraction of current remaining after a 30-s ACh application was 0.29 ± 0.04 in Chrna9\textsuperscript{wt/\textsuperscript{wt}} (n = 10 animals, 15 cells), 0.45 ± 0.05 in Chrna9\textsuperscript{L9T/\textsuperscript{wt}} (n = 9 animals, 12 cells, p < 0.05), and 0.48 ± 0.06 in Chrna9\textsuperscript{L9T/L9T} (n = 6 animals, 10 cells, p < 0.05).

**Prolonged Synaptic Currents in Mutant IHCs**

Following efferent transmitter release in vitro, spontaneous inhibitory postsynaptic currents (sIPSCs) can be measured in neonatal IHCs. As determined for MOC synapses located on OHCs, the IHC inhibitory postsynaptic currents (IPSCs) result from ion flux through α9\textsubscript{2}α10 nAChRs and calcium-activated SK2 channels [19,20], (Figure 1C). To only measure currents through α9\textsubscript{2}α10 nAChRs, responses were recorded using pipettes filled with KCl-BAPTA (thereby preventing calcium from activating SK2 channels) and 5 mM apamin (an SK channel blocker) in the extracellular solution. As shown in Figure 4A and detailed in Table 1, these “nAChR-only” sIPSCs of mutant mice were dramatically prolonged compared to wild type. There was a 6-fold increase in the τ\textsubscript{decay}, no difference in τ\textsubscript{rise}, and a 3-fold increase in the duration at half amplitude (halfwidth). In addition, the sIPSCs in Chrna9\textsuperscript{L9T/L9T} IHCs had significantly smaller amplitudes than those of wild-type IHCs. Despite the amplitude reduction, overall charge transfer (as indicated by the area value) of the “nAChR-only” synaptic current was more than doubled in Chrna9\textsuperscript{L9T/L9T} when compared to Chrna9\textsuperscript{wt/\textsuperscript{wt}} IHCs.

When sIPSCs were recorded with KCl-EGTA in the pipette, currents were inward at −90 mV and outward at −40 mV (Figure 4B). As noted for wild-type IHCs ([19,20] and present results), the polarity shift of the sIPSC indicates that SK2 channel gating occurs in the Chrna9L9T mutant mice. At −90 mV, sIPSCs of Chrna9\textsuperscript{L9T/L9T} mice were dramatically prolonged compared to Chrna9\textsuperscript{wt/\textsuperscript{wt}} with a 4- and 3-fold increase in the τ\textsubscript{decay} and halfwidth, respectively. At −40 mV, τ\textsubscript{decay} and halfwidth were 3-fold greater than in wild type, and a slight increase in τ\textsubscript{rise} also was observed. The peak current at −40 mV, carried by SK2 channels, was equivalent in Chrna9\textsuperscript{L9T/L9T} and Chrna9\textsuperscript{wt/\textsuperscript{wt}} IHCs. Nonetheless, the smaller amplitude nAChR-only current observed in IHCs of mutant mice activates SK2 channels as effectively as in wild-type IHCs. Indeed, the area under the Chrna9\textsuperscript{L9T/L9T} SK2 current was nearly three times greater than that of the Chrna9\textsuperscript{wt/\textsuperscript{wt}} SK2 current. Presumably, this enhancement results from the increased total calcium ion flux during the prolonged activity of the nAChRs in mutant mice.

As shown in Figure 4C, synaptic activity evoked by 40 mM K\textsuperscript{+} in IHCs was reversibly blocked by the α9\textsubscript{2}α10 blocker strychnine (300 nM) [13], indicating that the synaptic activity observed in Chrna9\textsuperscript{L9T/L9T} was in fact mediated through mutant α9\textsubscript{2}α10 nAChRs.

**Prolonged Synaptic Currents and Coupling to SK2 Channels in Mutant OHCs**

Since hair cell sIPSCs were infrequently observed, synaptic currents and the effects of exogenous ACh were measured in P10–P11 OHCs using high-potassium buffer (40 mM K\textsuperscript{+} plus
EGTA, minus apamin) to depolarize the efferent terminals (Figure 5A and 5B). Under these recording conditions, the synaptic waveform includes current through both the nAChR and associated SK2 channels (entirely inward at \( V_{\text{hold}} \) of \(-90\) mV since \( 40\) mM K\(^+\) shifts the equilibrium potential of this cation to \(-33\) mV). Analysis of waveforms revealed a 4- and 3.5-fold increase in \( \tau_{\text{decay}} \) and halfwidth, respectively, and a 0.6-fold reduction in amplitude (Figure 5C and Table 2) in \( \text{Chrna9}^{L9T/L9T} \) IPSCs compared to \( \text{Chrna9}^{wt/wt} \), much as was observed in IHCs.

Despite a drop in amplitude, the overall charge transfer (as indicated by the area value) was 2-fold larger in \( \text{Chrna9}^{L9T/L9T} \) compared to \( \text{Chrna9}^{wt/wt} \) OHCs.

Under these recording conditions, synaptic currents reflect ion flux through both nAChRs and associated SK2 channels.

### Table 1. Synaptic Currents in IHCs from \( \text{Chrna9}^{wt/wt} \) and \( \text{Chrna9}^{L9T/L9T} \) Mice

| Currents | nAChR | nAChR + SK2 |
|----------|-------|-------------|
| \( V_{\text{hold}} \) | \(-82\) mV | \(-40\) mV | \(-90\) mV |
| Genotype | \( \text{wt/wt} \) | \( \text{L9T/L9T} \) | \( \text{wt/wt} \) | \( \text{L9T/L9T} \) | \( \text{wt/wt} \) | \( \text{L9T/L9T} \) |
| Amplitude (pA) | 18.4 ± 0.8 | 11.1 ± 1.1 \( ^{*} \) | 28.2 ± 1.1 | 30.2 ± 1.3 | 28.9 ± 1.9 | 16.7 ± 0.9 |
| \( \tau_{\text{rise}} \) (ms) | 0.8 ± 0.1 | 0.8 ± 0.2 | 8.6 ± 1.2 | 18.8 ± 1.3 \( ^{*} \) | 5.4 ± 1.0 | 9.3 ± 1.9 |
| \( \tau_{\text{decay}} \) (ms) | 27.9 ± 1.0 | 186 ± 26 \( ^{c} \) | 52.9 ± 10.1 | 170 ± 37 \( ^{a} \) | 53.7 ± 5.4 | 222 ± 47 \( ^{b} \) |
| Halfwidth (ms) | 25.0 ± 1.2 | 79.2 ± 10.3 \( ^{b} \) | 66.5 ± 2.8 | 194 ± 23 \( ^{b} \) | 70.3 ± 6.0 | 235 ± 51.1 \( ^{b} \) |
| Area (pC) | 0.55 ± 0.09 | 1.1 ± 0.1 \( ^{*} \) | 2.0 ± 0.1 | 5.7 ± 1.4 \( ^{*} \) | 1.4 ± 0.1 | 3.9 ± 1.0 \( ^{*} \) |
| Number of animals | 2 | 3 | 3 | 2 | 3 | 4 |
| Number of cells | 4 | 4 | 3 | 2 | 6 | 5 |
| Number of events | 1125 | 141 | 83 | 55 | 277 | 141 |

\( ^{a} p < 0.05. \)
\( ^{b} p < 0.01. \)
\( ^{c} p < 0.001. \)

\( \text{wt} \), wild type.

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Figure 4. Prolonged Synaptic Currents in IHCs Are Blocked by Strychnine

(A) Superimposed representative traces from individual cells of nAChR-only sIPSCs in IHCs from a \( \text{Chrna9}^{wt/wt} \) (27 events) and a \( \text{Chrna9}^{L9T/L9T} \) (27 events) mice at a \( V_{\text{hold}} \) (\( V_{h} \)) of \(-82\) mV.

(B) Superimposed representative traces from individual cells of nAChR+SK synaptic currents in IHCs at a \( V_{\text{hold}} \) of \(-90\) mV from a \( \text{Chrna9}^{wt/wt} \) (84 events) and a \( \text{Chrna9}^{L9T/L9T} \) (16 events), and of \(-40\) mV from a \( \text{Chrna9}^{wt/wt} \) (54 events) and a \( \text{Chrna9}^{L9T/L9T} \) (14 events). The representative traces shown are the average of the different events in one cell (number of cells per experiment indicated in Table 1).

(C) Representative traces (\( n = 4 \) IHCs, 2 animals) of synaptic currents evoked by \( 40\) mM external K\(^+\) at \( V_{\text{hold}} \) of \(-90\) mV in \( \text{Chrna9}^{L9T/L9T} \). The blow-ups show IPSCs, which disappear in the presence of 300 nM strychnine.

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The presence of an ACh-evoked SK2 current was demonstrated directly by application of ACh to Chrna9L9T/L9T OHCs held at different membrane potentials. As shown in Figure 5D, ACh responses were outward at −40 mV, confirming the activation of potassium currents in mutant OHCs.

Increased Cochlear Thresholds in Mutants Can Be Reversed with Strychnine

We examined cochlear responses in Chrna9L9T mutant mice via auditory brainstem responses (ABRs) and distortion product otoacoustic emissions (DPOAEs). Wave 1 of the ABR, which is recorded from scalp electrodes in response to short tone pips, represents the summed activity of cochlear nerve fibers projecting from IHCs (Figure 1B) to the cochlear nucleus, the first central nucleus in the ascending auditory pathway [21]. The DPOAEs are sounds created within the cochlea, amplified by the action of OHCs and propagated through the middle ear back to the ear canal, where they can be measured with a microphone [22]. Given that the generation of DPOAEs requires neither IHCs nor cochlear nerve fibers [23], a comparison of ABRs and DPOAEs can provide insight into the locus of any dysfunction.

As shown in Figure 6A, mean ABR thresholds were elevated by 5–15 dB in both Chrna9wt/L9T (F(1,55) = 4.30, p = 0.043) and Chrna9L9T/L9T (F(1,79) = 7.87, p = 0.06) when compared to Chrna9wt/wt mice. DPOAE thresholds (Figure 6B) were also elevated by 5–15 dB in both Chrna9wt/L9T (F(1,53) = 14.82, p < 0.001) and Chrna9L9T/L9T (F(1,77) = 9.20, p = 0.003). Threshold shifts were larger at frequencies above 16 kHz and were similar in magnitude whether measured by ABRs or DPOAEs, suggesting that all dysfunction can be explained by changes in OHC contributions to cochlear amplification.

Since MOC activity in vivo decreases OHC-based amplification of cochlear responses [2], we asked whether the increased cochlear thresholds might arise from enhanced synaptic currents through mutant α9-containing channels in response to the normal low level of spontaneous MOC activity [24]. To test this hypothesis, we injected mice with strychnine (30 mg/kg, intraperitoneally [i.p.]), a potent blocker of α9α10 channels [13,25]. Indeed, as shown in Figure 7, strychnine improved acoustic thresholds in Chrna9L9T/L9T mice.
mice by almost 9 dB, thus restoring the baseline thresholds to that seen in the wild type.

**Shock-Evoked OC Suppression Is Slowed, Enhanced, and Prolonged in Mutants**

Activation of MOC efferents normally decreases the OHC contribution to cochlear amplification. Thus, the electrical stimulation of the MOC fibers at the floor of the IVth ventricle (Figure 1A, black arrow) results in a decrease in the amplitude of the DPOAEs [25]. To assess MOC function in vivo, DPOAEs were measured before, during, and after a 70-s train of shocks to the olivocochlear bundle. In wild-type mice, DPOAE suppression is seen immediately after shock-train onset (Figure 8A, black arrowhead) and then adapts to a steady state, which is maintained throughout the shock epoch (Figure 8A and 8B). In *Chrna9L9* mutant mice, suppression had a much slower onset (Figure 8A, white arrowhead), but continued to grow during the shock train (Figure 8A and 8B) until responses disappeared into the noise floor. To determine the full suppression magnitude, we raised the level of the acoustic stimulus (Figure 8C): peak suppression in wild-type mice by almost 10 min after the end of the shock train, with a slow approximately 5 dB. In homozygous mutants, suppression persisted for almost 10 min after the end of the shock train, with a slow recovery to baseline (Figure 8B). Moreover, there is a prominent “overshoot,” which, except for a slower time course, is reminiscent of the post-shock enhancement seen in wild-type mice [25]. As shown in Figure 8A and 8B, responses in *Chrna9L9* were intermediate between those of homozygous mutant and wild-type mice.

As with wild-type mice [25], suppression could be blocked by strychnine (10 mg/kg), leaving only the postshock enhancement (Figure 9A, grey arrowhead), suggesting that the increased suppression in *Chrna9L9* mutants was due to the enhanced activity of *a9* receptors. With a lower dose of strychnine (3 mg/kg), suppression in *Chrna9L9* resembled the normal *Chrna9* response (i.e., reduced suppression magnitude but with rapid onset and decay; Figure 9B, grey arrowhead).

**Discussion**

The present study demonstrates that substitution of a single amino acid in transmembrane region 2 of the *a9* subunit (*L9*T) enhances nAChR function in cochlear hair cells (i.e., decreased ACh EC50 and slowed receptor desensitization rate), increases synaptic efficacy, and dramatically strengthens cochlear suppression in vivo. More importantly, it shows that the MOC efferent system inhibits cochlear...
sensitivity and protects the inner ear from acoustic injury. Thus, the efferent pathway provides a promising target for pharmacological prevention of inner ear pathologies derived from acoustic injury, such as hearing loss and tinnitus.

The Hair Cell as a Model System to Study the Nicotinic Synapse

Understanding neurotransmission mediated by neuronal nicotinic receptors is a fundamental challenge in neuroscience, since decline, disruption, or alteration of nicotinic cholinergic mechanisms in the central nervous system contributes to dysfunctions such as epilepsy, schizophrenia, Parkinson’s disease, autism, dementia with Lewy bodies, Alzheimer’s disease, and addiction [10]. Although fast, direct nicotinic synaptic activity drives neurotransmission in autonomic ganglia, only rare cases of fast nicotinic transmission mediated by neuronal nAChRs have been reported in the mammalian brain. Because cholinergic neurons in the brain are usually loosely distributed and often sparsely innervate broad areas, it is experimentally difficult to stimulate a large number of those neurons and to record selectively from their postsynaptic targets. Indeed, it is likely that authentic fast nicotinic transmission is present at low densities in more neuronal areas than the few that have been reported [10].

Furthermore, the variable transcription of genes coding for nAChR subunits and the possible combinatorial assembly of these subunits produces a wide structural diversity of receptor types. This imposes additional challenges for studying native synaptic neuronal nAChRs [31].

Thus, the cholinergic synapse between MOC terminals and cochlear hair cells provides a valuable model for the study of fast neurotransmission mediated by nAChRs of known composition [12,13]. Targeted mutations can be introduced and the phenotypic consequences can be analyzed at the synaptic, whole-organ, and systems level. When analyzed at the level of nAChR function, the α9 L9′T mutant mice reproduce what has been previously described for the recombinant receptor expressed in oocytes, i.e., a decrease in EC50 for ACh and a reduced desensitization kinetics [18]. Both these effects probably derive from the fact that the L9′ position of nAChRs is critical for channel gating [32] and that hydrophilic substitutions at this position lead to increased open times [18,33]. Moreover, these changes in channel gating properties translate into increased synaptic efficacy, as seen from the prolonged synaptic currents observed in IHCs and OHCs of mutant mice. In addition, the inhibitory signature of the efferent synapse is conserved, since the nAChR currents remain coupled to the associated SK2 currents, which were also substantially prolonged. In fact, the increase in overall charge transfer during sIPSCs between wild-type and knockin mice was more pronounced when the secondary SK2 currents were measured, than for the nAChR-only currents (Table 1), pointing to an additional amplified step, possibly calcium-induced calcium release from the nearby synaptic cistern [34]. Finally, the finding that the L9′T mutation does not lead to hair cell death, is distinct from effects of similar mutations in α4 and α7 nAChR subunits, which lead to death of substantia nigra dopaminergic neurons [35] and apoptotic cell death throughout the somatosensory cortex [36], respectively, most likely due to Ca2⁺ excitotoxicity. The observation that α9α10 receptors are indeed highly permeable to Ca2⁺ [37,38] points toward an efficient Ca2⁺ buffering system in hair cells. In fact, proteinaceous calcium buffers (e.g., parvalbumin-β) are expressed in OHCs at high levels, similar to those found in skeletal muscle [39].

Functional Consequences of Enhanced nAChR Gating

The main peripheral effect of the MOC activity is to inhibit cochlear responses by decreasing the gain of the cochlear amplifier [2]. MOC neurons comprise the effector arm of a sound-evoked negative feedback system that, in a quiet environment, normally has little effect on cochlear sensitivity,
since MOC neurons have low levels of spontaneous activity and do not respond to sound until levels exceed threshold by 15–20 dB [24]. The fact that the baseline threshold elevation in \(L^9T\) mutants can be “rescued” via strychnine, the potent \(\alpha9\alpha10\) nAChR blocker, suggests that it arises from an enhancement of cholinergic effects on OHCs. Therefore, compared to \(ChRNA9\) and \(ChRNA10\) knockouts, in which baseline cochlear thresholds were normal [40,41], the present knockin strategy reveals cholinergic MOC effects under resting conditions. This baseline inhibition could arise from an increased sensitivity of mutant receptors to normal low levels of spontaneous ACh release from MOC terminals or to the increased probability of spontaneous channel openings of \(L^9T\) mutant receptors in the absence of ACh [18]. The appearance of spontaneous channel openings, as described for the mutant receptors [18], may also explain how low-dose strychnine can speed the onset kinetics of the in vivo response in mutant mice (Figure 9B), i.e., by restoring mutant channels to the closed state, from which rapid ACh-mediated channel openings can occur.

Hair cell recordings from the \(L^9T\) mutant mice in the present study show (1) ACh-evoked currents with a greatly reduced desensitization rate, and (2) spontaneous miniature synaptic currents with slower activation and decay kinetics. These changes in ACh response kinetics provide likely explanations for some aspects of the electrically evoked suppression of DPOAEs. The doubling of onset time for the ACh-evoked SK2 current in mutant hair cells (Table 1) must contribute to the slowed onset of MOC-mediated suppression in vivo, by reducing linear summation of postsynaptic effects. However, the slowing of suppression onset is so dramatic (<1 s in wild type vs. >20 s in mutant mice) that other possible factors, e.g., alterations of release probability or slowed facilitation in the efferent terminals [42], might also contribute to this delayed time course.

In addition to rising more slowly, MOC-mediated cochlear suppression was also larger and longer lasting in \(L^9T\) mutant mice. The normal adaptation of suppression during continuous MOC activation was absent in the presence of slowly desensitizing \(\alpha9\alpha10\) mutant receptors, consistent with the idea that receptor desensitization is a key factor in the decay of the wild-type response after shock-train onset. However, the dramatic prolongation of cochlear suppression after shock-train offset (500 s in mutants vs. <5 s in wild types) is too large to be explained only by the 2- to 3-fold prolongation of synaptic currents observed in the mutant hair cells. Another contributing factor may be the observation that choline, the metabolite produced when acetylcholinesterase degrades ACh released at the synapse, is a full agonist of the mutant \(\alpha9\alpha10\) receptor, and only a weak partial agonist of the wild-type receptor [18]. Thus, in vivo, the suppression can only decay after choline is taken up by the synaptic terminal or otherwise diffuses away. Treatment with a low dose of the \(\alpha9\alpha10\) antagonist strychnine (3 mg/kg) supports this interpretation. This treatment completely abolished the prolonged postshock suppression in mutant mice (and unmasked a fast onset). Since choline has 4-fold lower affinity for \(\alpha9\alpha10\) mutant receptors than ACh [18], a low concentration of strychnine may be differentiating the effects of the two agonists (i.e., blocking the effects of the agonist with a lower potency, choline, but leaving unchanged the effects of the high-affinity agonist, ACh), as well as modulating the resting level of desensitization. Beyond the possibility of metabolite activation, extended inhibitory effects could involve more than changes in membrane conductance. For example, it has been shown that exposure to ACh alters the stiffness and motility of OHCs isolated from the gerbil cochlea over tens of minutes [43]. Stiffness and motility of OHCs depends, at least in part, on the motor protein prestin, a molecule that has selectively evolved in mammals to subserve somatic electromotility and amplification [44,45]. Thus, it remains to be determined whether prolonged Ca\(^{2+}\) influx through mutated \(\alpha9\alpha10\) receptors leads to changes in prestin structure or function resulting in prolonged MOC efferent effects.

The presence of mutant \(\alpha9\alpha9\) receptors, designed to increase the magnitude of MOC effects on OHCs, also increased the protective action of the MOC system in vivo.
receptors. The further observation that in the reducing mechanical vibration of the sensory epithelium, or effects of a decrease in electromotility. protective action, rather than the hyperpolarization-induced magnitude of MOC-mediated DPOAE suppression, suggests reduction of vibration amplitude as the underlying mechanism, producing only reversible changes, argues against a general threshold shifts, and not in less traumatic exposures enough to produce irreversible damage and permanent threshold shifts, and not in less traumatic exposures producing only reversible changes, argues against a general reduction of vibration amplitude as the underlying mechanism. Most of the permanent threshold elevation underlying this type of noise exposure arises from damage to the hair cell stereocilia bundles, including disarray loss and/or fusion of these modified microvilli [50], which house the mechanoelectric transduction channels. Further insight into the mechanisms linking calcium entry through nAChRs and protection from acoustic injury will require a clearer delineation of the molecular events leading up to this type of noise-induced stereocilia damage.

**Conclusions**

This work shows that a point mutation in the hair cell’s nAChR produces dramatic prolongation of efferent MOC inhibitory effects at both cellular and systems levels. This alteration provides enhanced protection from permanent acoustic trauma, indicating that cholergic synaptic feedback is not only necessary, but sufficient for this effect. In addition, the enhanced activity of the modified nAChR revealed a novel tonic inhibitory effect, raising baseline acoustic thresholds over those of wild-type littermates, confirming the inhibitory nature of the MOC efferent system. Thus, this n9L9’T knockin presents new insights into the cellular mechanisms of cholergic inhibition, as well as a promising model in which to probe the functional role of MOC efferents. In addition, the hair cell’s efferent synapse, much like the neuromuscular junction, can provide insights into cholinerigic signaling, and promises to be an equally informative model for studying activity-dependent synaptic function.

**Materials and Methods**

Genetic engineering and genotyping of Chrna9L9’T knockin mutant mice. A HindIII-NotI (the latter derived from the construction of the library) restriction endonuclease fragment of approximately 9,300 bp encoding Chrna9 exons 1–4 (see GenBank accession number NT_039305.7) and Figure 2A) was obtained from a mouse strain 129S4/SvJae genomic DNA library (kindly provided by Dr Bernhard Bettler, University of Basel) and subcloned into the vector pKO-Select DT (Lexicon Genetics). A 2-kbp neomycin resistance cassette flanked by two loxP sites (loxP-neo-loxP) was inserted in the Ncol restriction site located between exon 3 and 4 and the Chrna9L9’T mutation (*) introduced via site-directed mutagenesis using the QuickChange Site-Directed Mutagenesis kit (Stratagene) and amplimers A9sense (5’-CTTCTGGGAGTGACCATCCTAacGGG-CATGACTGTTCTTCTCCGAGGCT-3’) and A9antisense (5’-GCTGAAATA-CAGTCATGCGGTAGTATGTCACTGACGAGGCT-3’). This targeting vector was used to electroporate 129S4/SvJae embryonic stem (ES) cells, and homologous recombinants were obtained following gentamicin (G418) selection and Southern blot hybridization analyses. Genomic DNA was purified from G418-resistant ES cell clones, digested with HindIII and KpnI, electrophoresed on 0.8% agarose, and hybridized to a 32P-labeled DNA 962-bp SacII-KpnI fragment probe prepared from the DNA fragment shown in Figure 2A. Based on the sequence of the mouse Chrna9 subunit gene (see GenBank accession number NT_039305.7), wild-type ES cell DNA yielded a fragment of 13,800 bp, whereas ES cells that have undergone homologous recombination yielded a 7,300-bp fragment (see Figure 2B). Transfection of the linearized targeting vector into murine ES cells resulted in the insertion of the L9’T mutation into the n9 nAChR subunit. The frequency of homologous recombination events in ES cells was 42%. Six independent ES cell lines carrying the mutation were used to electroporate wildtype 129S4/SvJae embryonic stem cells expressing the transgene and were then implanted into pseudopregnant females. Chimeric male progeny mice were backcrossed to strain C57BL/6j females, and agouti coat color was used to assess the germline status of the targeted males. Founder males were backcrossed to wild-type 129S4/SvJae females and heterozygous N1 females mated to c-rex-expressing transgenic males (FVBN-Tg[Elag-cre]C579J, stock 003154; Jackson Laboratory) to remove the “floxed” neomycin resistance cassette. The excision of the neo cassette and subsequent segregation and loss of the c-rex transgene were monitored via PCR. The single loxp site footprint
and flanking regions of the targeted allele in neo-deleted mice were confirmed by DNA sequencing.

The *Chrin9* mutant allele has been maintained in congenic FVB.129P2-Pde6b-/* Tyrch-ch AntJ stock (number 004828; Jackson Laboratory) strain. C57BL/6j mice develop a marked and progressive late-onset hearing loss characterized by cochlear degeneration. The background hearing loss by the age of 8 months, however, the *Pde6b* allele avoids blindness due to retinal degeneration typical of the FVB strain. All experiments reported in this paper were performed using neo-deleted *Chrin9*/*Chrin9*, or *Chrin9*/*Chrin9* mutant mice backcrossed with congenic FVB.129P2-Pde6b-/* Tyrch-ch AntJ stock for four to five generations (i.e., N4–N5).

Routine genotyping of *Chrin9* mice was performed using tail biopsy tissue DNA samples (Wizard Genomic DNA Purification kit; Promega), amplifiers A9L0GP1 (5'-TAC TGG CTA TCG TCC AGA CAG AGC-3') and A9LOXP2 (5'-AGG AGC GAG CAG AGG TCC TAA TAA TTA GAA AGT CTC-3'), and Fast Forward PCR Kit with buffer D (Epicentre) as described by the manufacturer. PCR cycle parameters were: 95 °C, 0.5 min; 55 °C, 1.0 min; and 72 °C, 2 min for a total of 35 cycles. Reaction products were electrophoresed on 1.5% agarose, stained with ethidium bromide, and photographed. The PCR fragment length for the wild-type *Chrin9* allele is 203 bp and 269 bp for the mutant allele (see Figure 2C). In each breeding pair, the PCR fragment obtained with amplifiers L9T5 and A9LOXP2 was scored.

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Electrophysiological recordings from cochlear hair cells. Mice were sacrificed by decapitation. All experimental protocols were carried out in accordance with the American Veterinary Medical Associations’ AVMA Guidelines on Euthanasia (June 2007). Apical turns of the organ of Corti were exposed via a wedge-shaped incision made in the right external auditory meatus. The mouse was placed on ice during surgery. Following aseptic procedures, the tympanic membrane was removed and the round window was accessed near the tail. ABRs were evoked with a 200 Hz click (120 dB SPL). For ABR recordings, needle electrodes were inserted at vertex and pinna, with a ground near the tail. ABRs were recorded and analyzed using the program Harmony (Dyad, 1988). ABR amplitudes were measured with simultaneous shocks to the right ear, with a Knowles miniature microphone (EK5103) to record ear-canal sound pressure. Stimuli were generated digitally, while ear-canal sound pressure was measured with a calibrated IEA92A (Harvard Apparatus, USA) or a Bruel and Kjaer (B&K) DAS 2000 sound level meter.

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other reagents were from Sigma Chemical. EGTA and Na2ATP were dissolved at the moment of preparing the intracellular solutions.

Animal welfare. All experimental protocols were carried out in accordance with the National Institutes of Health guide for the care and use of laboratory animals as well as Instituto de Investigaciones en Ingeniería Genética y Biología Molecular (INGEBI), Tufts University, and Massachusetts Eye and Ear Infirmary Institutional Animal Care and Use Committee (IACUC) guidelines, and best practice procedures.

Supporting Information
Figure S1. Morphology of the Cochlear Duct Is Normal in Mutant Ears

Gross histological assessment via plastic sections of osmium-stained Chrna9\textsuperscript{T/L9} cochleae showed no morphological changes when compared to Chrna9\textsuperscript{wt/wt}. Sections through the middle turn are shown. Black and grey arrows in Chrna9\textsuperscript{T/L9} indicate OHCs and IHCs, respectively. Histological procedures were performed as described in Text S1.

Found at doi:10.1371/journal.pbio.1000018.s001 (2.38 MB TIF).

Figure S2. Voltage-Dependent K\textsuperscript{+} Currents from IHCs and OHCs (A) Voltage-dependent currents obtained in P6–P10 IHCs by holding the cell at –80 mV and stepping the voltage to 80 mV in 10-mV increments (200-ms duration). Representative traces are shown for the three genotypes as well as for averaged steady state current-voltage curves (Chrna9\textsuperscript{9TG/TG}: n = 10 animals, 13 cells; Chrna9\textsuperscript{9TG/9TG}: n = 9 animals, 13 cells; and Chrna9\textsuperscript{9TG/10TG}: n = 14 animals, 17 cells). (B) Same as in (A), but for P10–P14 OHCs (Chrna9\textsuperscript{9TG}: n = 2 animals, 3 cells; and Chrna9\textsuperscript{9TG/10TG}: n = 6 animals, 8 cells).

Found at doi:10.1371/journal.pbio.1000018.s002 (225 KB TIF).

Figure S3. Dose-Response Curve for Strychnine Blockade of Cholinergic OC Effects on DPOAE Thresholds Each point shows data from different animals extracted from runs such as those shown in Figure 8 (f2 + f1). All data were collected 60 min after strychnine injection. Although thresholds in wild-type mice slightly rise with strychnine dosage, thresholds in knockout mice improve by as much as 8 dB.

Found at doi:10.1371/journal.pbio.1000018.s003 (117 KB TIF).

References
1. Hudspeth A (1997) How hearing happens. Neuron 19: 947–950.
2. Guinan JJ (1996) Physiology of olivocochlear efferents. In: Dallos P, Popper AN, Fay RR, editors. The cochlea. New York: Springer-Verlag. pp. 435–502.
3. Fuchs P (1996) Synaptic transmission at vertebrate hair cells. Curr Op Pharmacol 6: 514–519.
4. Dolan DF, Nuttall AL (1988) Masked cochlear whole-nerve response intensity functions altered by electrical stimulation of the crossed olivocochlear bundle. J Acoust Soc Am 83: 1081–1086.
5. Kawase T, Delgutte B, Liberman MC (1993) Anti masking effects of the olivocochlear reflex. II. Enhancement of auditory-nerve response to masked tones. J Neurophysiol 70: 2533–2549.
6. Winslow RL, Sachs MB (1988) Single-tone intensity discrimination based on auditory-nerve rate responses in backgrounds of quiet, noise, and with stimulation of the crossed olivocochlear bundle. Hear Res 35: 165–189.
7. Delano PH, Elgueta D, Hamame CM, Robles I (2007) Selective attention to visual stimuli reduces cochlear sensitivity in chinchillas. J Neurosci 27: 4146–4153.
8. Oatman LC (1976) Effects of visual attention on the intensity of auditory evoked potentials. Exp Neurol 51: 41–53.
9. Rajan R, Johnstone BM (1988) Binaural acoustic stimulation exercises improve by as much as 8 dB.
10: a determinant of nicotinic cholinergic receptor function in mammalian vestibular and cochlear mechanosensory hair cells. Proc Natl Acad Sci U S A 98: 3501–3506.

Table S1. Analysis of Gene Expression by Quantitative PCR

| Chrna9: gene encoding the α9 nACHR subunit; | Chrna10: gene encoding the α10 nACHR subunit; | Kenm2: gene encoding the SK2 channel; | Ryrl: gene encoding the ryanodine receptor 1; | Ryrl2: gene encoding the ryanodine receptor 2; | Ryrl3: gene encoding the ryanodine receptor 3; | Kenm11: gene encoding the calcium-activated potassium channel (BK) β1-subunit; | Cachnl: gene encoding the voltage-gated calcium channel α subunit CaV1.3; | Myos7a: gene encoding myosin VIIa; | Cy2c: threshold cycle of sample; | Cy2c: threshold cycle of Myos7a. Results shown are the mean ± S.E.M of three experiments per group, each performed in triplicates. The protocol for quantitative PCR was as indicated in Text S2. |

Found at doi:10.1371/journal.pbio.100018.st001 (88 KB DOC).

Text S1. Histology of the Cochlear Duct

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Text S2. Quantitative PCR

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Author contributions. JT, SFM, JAB, EK, DEV, JB, MCL, PAF, and ABE conceived and designed the experiments. JT, SFM, JS, and JB performed the experiments. JT, SFM, EK, DEV, JB, MCL, PAF, and ABE analyzed the data. SFM, MCL, PAF, and ABE wrote the paper.

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Plazas PV, Katz E, Gomez-Casati ME, Bouzat C, Elgoyhen AB (2005) Stoichiometry of the α9α10 nicotinic cholinergic receptor. J Neurosci 25: 10965–10972.

Lustig LR, Peng H, Hiel H, Yamamoto T, Fuchs P (2001) Molecular cloning and mapping of the human nicotinic acetylcholine receptor α10 (CHRNA10). Genomics 73: 272–283.

Dulon D, Luo L, Zhang C, Ryan AF (1998) Expression of small-conductance calcium-activated potassium channels (SK) in outer hair cells of the rat cochlea. Eur J Neurosci 10: 981–985.

Oliver D, Klocker N, Schuck J, Baukovzit R, Rupperrtersberg JP, et al. (2000) Gating of Ca\textsuperscript{2+}-activated K\textsuperscript{+} channels controls fast inhibitory synaptic transmission at auditory outer hair cells. Neuron 26: 595–601.

Plazas PV, De Rosa MJ, Gomez-Casati ME, Vertbisky M, Weinstaub N, et al. (2005) Key roles of hydrophobic rings of TM2 in gating the alpha9alpha10 nicotinic cholinergic receptor. Br J Pharmacol 145: 963–954.

Glowczewski E, Fuchs P (2000) Cholinergic synaptic inhibition of inner hair cells in the neonatal mammalian cochlea. Science 288: 2366–2368.

Gomez-Casati ME, Fuchs PA, Elgoyhen AB, Katz E (2005) Biophysical and pharmacological characterization of nicotinic cholinergic receptors in cochlear inner hair cells. J Physiol 566: 103–118.

Melcher JR, Kiang NY (1996) Generators of the brainstem auditory evoked potential in cat. III: Identified cell populations. Hear Res 93: 52–71.

Kemp DT (1986) Otoacoustic emissions, travelling waves and cochlear mechanisms. Hear Res 22: 95–109.

Liberman MC, Chesney CP, Kujawa SG (1997) Effects of selective inner hair cell loss on DPOAE and CAP in carboplatin-treated chinchillas. Aud Neurosci 3: 255–288.

Liberman MC (1988) Physiology of cochlear efferent and afferent neurons: direct comparisons in the same animal. Hear Res 34: 179–191.

Maison SF, Vetter DE, Liberman MC (2007) A novel effect of cochlear efferents: in vivo response enhancement does not require α9 cholinergic receptors. J Neurophysiol 97: 3289–3297.

Maison SF, Laubke AE, Liberman MC, Jau J (2002) Efferent protection from acoustic injury is mediated via α9 nicotinic acetylcholine receptors on outer hair cells. J Neurosci 22: 10883–10896.
27. Rajan R (1988) Effect of electrical stimulation of the crossed olivocochlear bundle on temporary threshold shifts in auditory sensitivity. I. Dependence on electrical stimulation parameters. J Neurophysiol 60: 549–568.
28. Rajan R (1988) Effect of electrical stimulation of the crossed olivocochlear bundle on temporary threshold shifts in auditory sensitivity. II. Dependence on the level of temporary threshold shifts. J Neurophysiol 60: 569–579.
29. Saunders JC, Dear SP, Schneider ME (1985) The anatomical consequences of acoustic injury: a review and tutorial. J Acoust Soc Am 78: 853–860.
30. Nordmann AS, Bohme BA, Harding GW (2000) Histopathological differences between temporary and permanent threshold shift. Hear Res 139: 13–30.
31. McGeehe D, Role L (1995) Physiological diversity of nicotinic acetylcholine receptors expressed by vertebrate neurons. Annu Rev Physiol 57: 521–546.
32. Miyazawa A, Fujiyoshi Y, Unwin N (2003) Structure and gating mechanism of the acetylcholine receptor pore. Nature 425: 949–955.
33. Labarca C, Nowak MW, Zhang H, Tang L, Deshpande P, et al. (1995) Channel gating governed symmetrically by conserved leucine residues in the M2 domain of nicotinic receptors. Nature 376: 514–516.
34. Lioudyno M, Hiel H, Kong JH, Katz E, Waldman E, et al. (2004) A ''synaptoplasmic cistern'' mediates rapid inhibition of cochlear hair cells. J Neurosci 24: 11160–11164.
35. Labarca C, Schwarz J, Deshpande P, Schwarz S, Nowak M, et al. (2001) Point mutant mice with hypersensitive alpha4 nicotinic receptors show dopaminergic deficits and increased anxiety. Proc Natl Acad Sci U S A 98: 2786–2791.
36. Orr-Uttreger A, Broide R, Kasten M, Dang H, Dani J, et al. (2000) Mice homozygous for the L250T mutation in the alpha7 nicotinic acetylcholine receptor show neuronal apoptosis and die within 1 day of birth. J Neurochem 74: 2154–2166.
37. Katz E, Verbitsky M, Rothlin C, Vetter D, Heinemann S, et al. (2000) High calcium permeability and calcium block of the alpha9 nicotinic acetylcholine receptor. Hear Res 141: 117–128.
38. Weisstaub N, Vetter D, Elgoyhen A, Katz E (2002) The alpha9alpha10 nicotinic acetylcholine receptor is permeable to and is modulated by divalent cations. Hear Res 167: 122–135.
39. Hackney CM, Mahendrasingam S, Penn A, Fettiplace R (2003) The concentrations of calcium buffering proteins in mammalian cochlear hair cells. J Neurosci 25: 7867–7875.
40. Vetter D, Lieberman M, Mann J, Barhanin J, Boulter J, et al. (1999) Role of alpha9 nicotinic ACh receptor subunits in the development and function of cochlear efferent innervation. Neuron 25: 93–103.
41. Vetter DE, Katz E, Maisson SF, Taranda J, Turcan S, et al. (2007) The alpha10 nicotinic acetylcholine receptor subunit is required for normal synaptic function and integrity of the olivocochlear system. Proc Natl Acad Sci U S A 104: 20594–20599.
42. Goutman JD, Fuchs PA, Glowatzki E (2005) Facilitating efferent inhibition of inner hair cells in the cochlea of the neonatal rat. J Physiol 566: 49–59.
43. Srikai I, He DZ, Dallos P (1996) Effect of acetylcholine and GABA on the transfer function of electromotility in isolated outer hair cells. Hear Res 95: 87–99.
44. Dallos P (2008) Cochlear amplification, outer hair cells and prestin. Curr Opin Neurobiol.
45. Franchini LF, Elgoyhen AB (2006) Adaptive evolution in mammalian proteins involved in cochlear outer hair cell electromotility. Mol Phylogenet Evol 41: 622–635.
46. Sridhar TS, Brown MC, Sewell WF (1997) Unique postsynaptic signaling at the hair cell efferent synapse permits calcium to evoke changes on two time scales. J Neurosci 17: 428–437.
47. Sridhar TS, Liberman MC, Brown MC, Sewell WF (1995) A novel cholinergic “slow effect” of efferent stimulation on cochlear potentials in the guinea pig. J Neurosci 15: 3667–3678.
48. Cooper NP, Guinan JJ Jr (2003) Separate mechanical processes underlie fast and slow effects of medial olivocochlear efferent activity. J Physiol 548: 307–312.
49. Maisson SF, Parker LL, Young L, Adelman JP, Zuo J, et al. (2007) Overexpression of SK2 channels enhances efferent suppression of cochlear responses without enhancing noise resistance. J Neurophysiol 97: 2930–2936.
50. Wang Y, Hirose K, Liberman MC (2002) Dynamics of noise-induced cellular injury and repair in the mouse cochlea. J Assoc Res Otolaryngol 3: 248–268.
51. Kros C, Ruppersberg JP, Rusch A (1998) Expression of a potassium current in outer hair cells during development of hearing in mice. Nature 394: 281–284.