Identification of novel genes involved in light-dependent CRY degradation through a genome-wide RNAi screen

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Circadian clocks regulate many different physiological processes and synchronize these to environmental light:dark cycles. In Drosophila, light is transmitted to the clock by a circadian blue light photoreceptor CRYPTOCHROME (CRY). In response to light, CRY promotes the degradation of the circadian clock protein TIMELESS (TIM) and then is itself degraded. To identify novel genes involved in circadian entrainment, we performed an unbiased genome-wide screen in Drosophila cells using a sensitive and quantitative assay that measures light-induced degradation of CRY. We systematically knocked down the expression of ∼21,000 genes and identified those that regulate CRY stability. These genes include ubiquitin ligases, signal transduction molecules, and redox molecules. Many of the genes identified in the screen are specific for CRY degradation and do not affect degradation of the TIM protein in response to light, suggesting that, for the most part, these two pathways are distinct. We further validated the effect of three candidate genes on CRY stability in vivo by assaying flies mutant for each of these genes. This work identifies a novel regulatory network involved in light-dependent CRY degradation and demonstrates the power of a genome-wide RNAi approach for understanding circadian biology.

[Keywords: RNAi; circadian rhythms; clock proteins; degradation; photoreceptors]

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The molecular circadian clock consists of feedback loops in which specific clock proteins regulate transcription of their own mRNAs, thereby maintaining cycles of gene expression. The major loop in Drosophila is comprised of the PERIOD (PER) and TIMELESS (TIM) proteins, both of which are expressed cyclically and rhythmically regulate transcription by inhibiting the activity of transcriptional activators CLOCK (CLK) and CYCLE (CYC) [Ashmore and Sehgal 2003; Stanewsky 2003; Hardin 2006]. The cycling of PER and TIM is essential for circadian rhythms and is thought to provide timekeeping cues for the organism. Consistent with the idea that levels of PER–TIM constitute time-of-day signals, light sets the time of the clock by altering levels of these molecules. Thus, entrainment of the molecular clock to light is mediated by light-induced degradation of the TIM protein. In response to light, TIM is targeted to the ubiquitin–proteasome pathway by an E3 ubiquitin ligase complex containing the F-box protein JETLAG (JET) [Naidoo et al. 1999; Koh et al. 2006]. The ubiquitin–proteasome pathway may also affect light-independent PER–TIM degradation, which maintains cycling under free-running conditions [Grima et al. 2002; Ko et al. 2002]. Degradation of TIM in response to light, as well as in free-running conditions, is affected by its phosphorylation state [Zeng et al. 1996; Naidoo et al. 1999; Grima et al. 2002; Yuan et al. 2005].

Photic signals are transmitted to TIM by a dedicated circadian photoreceptor, CRYPTOCHROME (CRY) [Cashmore 2003; Lin and Todo 2005]. After signaling to TIM, CRY is itself degraded by the proteasome pathway [Lin et al. 2001]. However the role of light-dependent CRY degradation in regulating circadian behavior and resetting is not clear. According to one report, clock neuron-specific overexpression of CRY increases circadian photo sensitivity [Emery et al. 2000], while another study found that overexpression of CRY reduces photo sensitivity [Ishikawa et al. 1999]. A CRY mutant (CRY<sup>−</sup>) lacking the C-terminal domain required for the regula-
tion of stability is able to transmit photic signals at low light intensity but is unstable and entrainment-defective at normal light intensity; thus, the requirement of light-induced CRY degradation for entrainment is not directly addressed [Busza et al. 2004]. Regardless of the role for light-induced degradation of CRY, it is clear that activation of CRY by light, which leads to its degradation, is essential for entrainment of the clock. Interestingly, CRY activation involves intramolecular electron transfer and presumably subsequent conformational changes, so cellular redox status also regulates the transfer of photic information and CRY stability [Lin et al. 2001].

In addition to its role in entrainment, CRY functions as an integral component of peripheral clocks found outside the brain in Drosophila [Ivanchenko et al. 2001; Krishnan et al. 2001; Collins et al. 2006]. Likewise in mammals, the two CRY orthologs are essential components of both central and peripheral clocks, where they, rather than TIM, act as partners of PER [Kume et al. 1999; Shearman et al. 2000]. Importantly, both CRYs in mammals are expressed with a robust circadian rhythm, suggesting that they are central timekeeping molecules. Thus, the identification of molecules that regulate CRY is an important objective, since it should reveal mechanisms that sustain free-running clock function as well as those that entrain the clock.

In the current study, we sought to identify novel genes involved in the photic input pathway in Drosophila, using light-induced CRY degradation as an end point assay. We performed a genome-wide RNAi screen in an S2 cell culture system and identified 190 candidate genes involved in the regulation of CRY in response to light. Most of these genes are components of the proteasome, ubiquitin ligases, signal transduction molecules, or redox molecules. We validated the effects of the selected candidates on CRY degradation through Western blot analysis. To further understand their role in the circadian light entrainment pathway, we examined their effect on TIM degradation in response to light. Using mutant flies, we obtained in vivo validation for the function of an E2–E3 ubiquitin ligase, Bruce, a dual specific phosphatase, slingshot (ssh), and a novel HECT domain-containing E3 ligase, CG17735, in the regulation of CRY. Since the screen identified several ubiquitin ligases that specifically affect CRY degradation without affecting TIM degradation, we propose that once CRY is activated by light, it targets TIM for degradation, following which the pathways for CRY and TIM degradation are distinct.

**Results**

A high-throughput assay for light-dependent CRY degradation in Drosophila Schneider 2 (S2) cells

We previously reported that CRY is degraded by light in Drosophila S2 cells [Lin et al. 2001]. This suggested an assay for a high-throughput cell-based genomic screen. To develop a robust, high-throughput assay for determining CRY stability in S2 cells, we generated N-terminal or C-terminal luciferase fusion proteins of CRY and monitored luciferase activity driven by these constructs in the presence and absence of light. As seen for CRY itself, luciferase activity from each of the fusion proteins decreased in a light-dependent manner [Fig. 1A; data not shown]. Incubation with a proteasome inhibitor, MG132, blocked this light-dependent CRY-luc degradation [Fig. 1A]. We also quantified protein levels by Western blot analysis and observed a corresponding decrease in the levels of the CRY-luc fusion proteins [data not shown]. However, the luciferase assay was more sensitive and amenable to high-throughput screening. As a control, we generated a luciferase fusion protein of an inactive form of CRY, CRYb, which has a point mutation (D410N) in the flavin-binding pocket that renders it insensitive to light [Stanewsky et al. 1998; Ceriani et al. 1999; Busza et al. 2004]. Consistent with other assays indicating lack of light responsiveness of the CRYb protein, the CRYb-luc fusion protein was insensitive to light [Fig. 1B].

We next optimized conditions for RNA-mediated interference using various S2 cell derivatives. Since other high-throughput screens in Drosophila have used S2R+ cells with dsRNA libraries plated in 384-well plates, we needed to develop a robust CRY degradation assay under these conditions. We optimized assay conditions in S2R+ cells using dsRNA against proteasome components Prosβ2 and Dms13. Dms13 [also called Rpn11] is a component of the 19S regulatory lid complex, and Prosβ2 is an essential component of the 20S proteolytic core complex. dsRNA against each of these proteasome components significantly blocked light-dependent CRY degradation. On the other hand, dsRNAs against Slimb, an E3 ligase for PER [Ko et al. 2002], or morgue, a putative SCF component that may be involved in the circadian entrainment pathway [Murad et al. 2007], had no effect [Fig. 1C]. The robustness of the assay was tested in a miniscreeen in which we examined the effect of dsRNAs against a random collection of 380 genes on light-dependent CRY degradation. dsRNA against the proteasome component Prosβ2 was used as a positive control. In order to quantify the effects of the dsRNAs we determined Z-scores as a measure of the effect on CRY degradation [see the Materials and Methods for details regarding Z-scores]. The Z-score for the Prosβ2 dsRNA indicated that the efficiency of CRY degradation was reduced such that it was 6.2 standard deviations away from the mean [Fig. 1D], reflecting the sensitivity and robustness of this assay. As expected, most of the dsRNAs did not score positive, but dsRNAs against four genes showed small effects, including one against a mono-oxygenase in the ubiquitination pathway [CG7277] that gave a Z-score of less than −2 [Fig. 1D]. Similar results were observed in a duplicate experiment [data not shown]. dsRNA against thread, which is essential for S2 cell survival, promoted light-dependent CRY degradation [Z-score +3.2] [Fig. 1D], suggesting that dsRNAs that promote apoptosis and cell death may indirectly promote CRY-luc reporter degradation. Since it is very difficult to distinguish the effect of dsRNAs that affect cell health from those that actively promote CRY degradation, we confined our analy-
Figure 1. Development of a primary screen to identify novel components involved in light-dependent CRY degradation. (A) Light-dependent degradation of a CRY-luc fusion protein: S2R+ cells were transfected with expression constructs for a Cry-luc fusion protein and Actin-Renilla luciferase. Cells were exposed to light in the presence of DMSO (control) or proteasome inhibitor MG132 (100 µM) for the indicated times, and luciferase activity was monitored. Luciferase activity was normalized to that of cells kept in darkness, set as 1. Average values from three samples are plotted, and the error bars depict standard error of the mean [SEM]. The results are representative of two independent experiments. Light-dependent degradation of the CRY-luc fusion protein was blocked by proteasome inhibitor MG132. (B) A single point mutation in the flavin-binding pocket of CRY, CRYb, renders it insensitive to light. S2R+ cells were transfected with expression constructs for CRY-luc or CRYb-luc fusion protein in 384-well plates and assayed for luciferase activity after light exposure for 6 h. Luciferase activity from three samples is plotted relative to the CRY activity in transfected cells kept in the dark [set to 100%]; error bars represent SEM. In response to light exposure, activity of the CRY-luc fusion protein was reduced by 83% (P-value = 2 × 10⁻⁶), while activity of the CRYb-luc fusion protein was unchanged. Similar results were obtained in three experiments with different cell densities. (C) RNAi of 26S proteasome components blocks light-dependent CRY degradation. S2R+ cells were cotransfected with the CRY-luc expression construct and dsRNAs, as indicated, in 384-well plates. Act-Renilla luc was included as a transfection control. Ninety-six hours post-transfection, cells were exposed to light and luciferase activity was measured as indicated above. Relative luciferase activity from three samples is plotted; error bars represent SEM. The value obtained with dsRNA against gfp in the dark was set as 100% and used for normalization. dsRNA against cry, which dramatically reduced luciferase activity (data not shown), was used as a positive control. RNAi of the 26S proteasome core components Prosβ2 and DmS13 (also called as Rpn11) significantly blocked light-dependent CRY-luc degradation. Similar results were obtained in two independent experiments. (D) A pilot screen demonstrates the validity of the CRY-luc assay. The light-dependent CRY-luc degradation assay was performed in S2R+ cells with 380 randomly picked dsRNAs from the genomic collection. dsRNA against the proteasome component Prosβ2 was used as a positive control; thread dsRNA was included as a control to verify the efficiency of RNAi. Normalized luciferase levels were compared between the dark- and light-treated plates. Z-scores, standard deviations from the plate mean (see the Materials and Methods for details), are plotted. While dsRNA against Prosβ2 significantly blocked light-dependent CRY-luc degradation (Z-score = −6.2), most dsRNAs had no effect. Similar results were obtained in two independent experiments.

sis to dsRNAs that block light-dependent CRY degradation (i.e., Z-score less than −2). Our ability to identify ubiquitin–proteasome components from a random selection of dsRNAs validated our assay, indicating that it was capable of identifying novel genes involved in regulating CRY stability.

Genome-wide RNAi screen for components involved in the light-dependent CRY degradation pathway

Having developed a robust assay, a genome-wide RNAi library was screened in duplicate using S2R+ cells to identify dsRNAs that block light-dependent CRY degradation. Analysis of the Z-scores from duplicate assays showed that the results were reproducible, with good correlation between the two experiments [Fig. 2A]. The comparison of dark and light samples identified dsRNAs that inhibited CRY degradation and we selected those with a Z-score of less than or equal to −2. In addition, those dsRNAs that elevated CRY-luc levels under light conditions, with a Z-score of 2 or more, were also scored as positives. Of the 21,000 dsRNAs in the library, after filtering out dsRNAs harboring homology with multiple repeat elements with predicted off-target effects [Kulkarni et al. 2006], the primary screen identified 303 annotated genes that inhibited CRY degradation [Fig. 2B; Supplemental Table S1].

To confirm our results, we conducted further analysis with the positives identified in the primary screen. Of the 303 candidates, we were able to synthesize dsRNA against 287 genes and confirmed 173 (60%) as positive by rescreening them using the CRY-luc light response assay. Functional annotation and clustering analysis identified a high enrichment of proteasome complex genes [EASE score 10⁻¹⁴] (Hosack et al. 2003). Notably, we identified 17 genes encoding proteasome components, including Prosβ2 and DmS13. Among other proteasome components, we identified seven of the eight AAA-ATPases from the 19S base complex. These AAA-ATPases are essential for substrate anchoring, unfolding, and the gating of the 20S proteolytic core.

Apart from components of the core–proteasome complex, we isolated five genes encoding components of the ubiquitin-conjugating complex. This type of complex includes E3 ligases that perform an essential/rate limiting role in targeting specific proteins for ubiquitination and
proteasome degradation. We identified three novel HECT or RING domain-containing E3 ligases (CG17735, CG11321, and CG5604), as well as a composite E2–E3 ligase, Bruce, which contains both ubiquitin-conjugating enzyme activity and a putative substrate-binding domain.

Previous studies have indicated a role for electron transfer and redox in light-dependent CRY degradation (Lin et al. 2001). Our screen identified eight genes involved in regulating cellular redox status, including Glutathione S-transferase (GstE7), Thioredoxin (Txl), lactate/malate dehydrogenase (ImpL3), and bioppterin-binding thyrosine hydroxylase (TH) gene pale (ple). We also identified 13 genes involved in signal transduction, of which three are kinases and two are phosphatases. Given earlier studies suggesting a role of tyrosine kinases in light entrainment (Naidoo et al. 1999), it is interesting that the two phosphatases (slingshot and CG7556) are predicted to recognize phospho-tyrosine residues.

Effects of candidate genes on light-dependent CRY and TIM degradation

To confirm the effect of the candidate dsRNAs on CRY degradation and to place these molecules in the circadian entrainment pathway, we extended our analysis for a select set of 22 genes. These genes included ubiquitin ligases, redox molecules, and signal transduction molecules that significantly blocked light-dependent CRY degradation (Table 1; Supplemental Table S1). Three genes involved in the 26S proteasome complex were also included. After light activation, CRY binds TIM and then both proteins are targeted for proteasome-mediated degradation (Ceriani et al. 1999; Lin et al. 2001). To investigate if the candidates identified by our screen also block light-dependent TIM degradation, we cotransfected S2R+ cell with V5 tagged cry along with tim. An expression construct for JET, a recently identified E3 ligase for TIM, was also included in the assay since this is required for the TIM response to light in S2 cells. Following light exposure, TIM and CRY levels were examined by Western blot analysis (Fig. 3). For ease of comparison, relative Z-scores were calculated and plotted for CRY and TIM degradation (Fig. 3C). dsRNAs against all three core components of the proteasome included in the list of 22 genes (Rpt1, Rpn6, and Pros/H9252) significantly inhibited light-dependent CRY degradation (Fig. 3). RNAi of Rpn6, the component of the proteasome lid complex, was most effective in blocking light-dependent CRY degradation, and it also affected TIM degradation. However, dsRNAs against Rpt1 and Prosβ2 were ineffective against TIM degradation. The role of the 26S proteasome complex in regulating light-dependent TIM degradation has been well documented (Naidoo et al. 1999; Koh et al. 2006). It is possible that when E3 ligase (JET) levels are in excess, TIM degradation is accomplished by limiting amounts of the proteasome complex. In general, our results show that proteasome-mediated degradation is essential for CRY and TIM degradation.

The selected 22 genes included five that regulate elec-
electron transfer or redox status of proteins. Previous studies using small molecule regulators of cellular redox suggested that electron transfer plays an important role in the light-dependent activation of CRY (Lin et al. 2001; Froy et al. 2002). However the molecules involved in regulating this process have not been identified. Our re-screen included three thioredoxin domain-containing proteins (GstE7, Txl, and CG11790) that are small disulfide-containing proteins that regulate redox status of many proteins (Maher 2006). RNAi-mediated knockdown of all three genes (GstE7, Txl, and CG11790) blocked light-dependent CRY degradation (Fig. 3), thus strongly supporting the role of redox proteins in this process. Since electron transfer is involved in the light-dependent activation of CRY (Lin et al. 2001; Froy et al. 2002), this step should precede TIM degradation. As expected, all these redox genes also blocked light-dependent TIM degradation (Fig. 3C). RNAi-mediated knockdown of a cytochrome P450 gene, Cyp49a1, also blocked CRY and TIM degradation. In addition, knockdown of a TH gene Pale (Ple), involved in catecholamine biosynthesis and pigmentation of the cuticle, inhibited CRY and TIM degradation, suggesting a novel role for TH in the circadian light entrainment pathway.

We also examined the effects of RNAi against three kinases and two phosphatases identified in the screen. As noted above, tyrosine phosphorylation has been previously implicated in light-dependent TIM degradation. RNAi knockdown of a dual specific phosphatase ssh (Niwa et al. 2002) blocked light-dependent CRY degradation but had little effect on TIM degradation. SSH is involved in actin filament reorganization and is activated by phosphotyrosininositol 3'-kinase (PI3'-kinase) in an insulin-dependent manner (Huang et al. 2006). RNAi knockdown of three kinases identified in the screen had a moderate effect in blocking the degradation of CRY/TIM. Nipped-A, a kinase involved in Notch signaling, and Stretchin-MLCK (Strn-Mlck), a myosin light chain kinase, blocked both CRY and TIM degradation. Knockdown of a TGFβ-activated kinase, Tak, and of a peptido-glycan receptor, PGRP-SA, both of which are involved in the IMD immune pathway in Drosophila, moderately inhibited CRY degradation. The effect of knocking down IMD pathway components Tak and PGRP-SA was restricted to CRY degradation; they did not affect TIM, suggesting that components of this pathway may be specifically utilized for the regulation of CRY. In addition to these kinase/phosphatases, dsRNA against an SH2 domain-containing RAS-GTPase, vacuolar peduncle (vap), blocked both CRY and TIM degradation.

We also retested a number of components known to be involved in protein degradation pathways. RNAi against an SCF adaptor gene, skpB, had a strong effect on both CRY and TIM degradation. The role of the SCF complex in light-dependent TIM degradation has been previously described (Koh et al. 2006); the current results indicate that it may also be required for CRY degradation. On the other hand, a putative SUMO protease (CG10107) blocked light-dependent TIM degradation but did not affect TIM degradation. In follow-up studies, we assayed for SUMOylation of CRY in S2R+ cells but were unable to detect this, suggesting that CRY is not directly regulated by SUMOylation (data not shown). Thus, other components of the CRY degradation complex may be regulated by SUMOylation.

Finally, our list of 22 genes included two HECT domain-containing E3 ligases (CG17735 and CG5604) and a RING domain-containing E3 ligase (CG11321). While neither affected TIM degradation, RNAi against CG17735 in particular had a profound effect on CRY degradation. However, of all the ubiquitin ligases tested

| Gene | Class | Putative functional domain | Z-score |
|------|-------|----------------------------|---------|
| prosB2 | Proteasome | Multispecific proteases of the proteasome | -12.0452 |
| Rpt1 | Proteasome | P-loop-containing nucleotide triphosphate hydrolase | -32.1033 |
| Rpn6 | Proteasome | Tetratricopeptide repeat (TPR); Rpn6 | -39.5271 |
| Cyppy9A1 | Redox | E-class P450 group I, cytochrome P450 enzyme | -5.05589 |
| CG17790 | Redox | Thioredoxin-like, CG11790, thioredoxin | -5.24895 |
| GstE7 | Redox | Thioredoxin-like, CG17531, glutathione S-transferases | -11.8823 |
| Txl | Redox | Thioredoxin, thioredoxin-like, Txl | -37.5307 |
| ple | Redox | Biotin-dependent aromatic amino acid hydroxylase | -45.4888 |
| CG7556 | Signal transduction, Phosphatase | (Phosphotyrosine protein) phosphatases II; CG7556 | -22.4196 |
| vap | Signal transduction | Src homology 2 (SH2) domain, | -22.3501 |
| Nipped-A | Signal transduction, kinase | Phosphatidylinositol 3- and 4-kinase, ARM repeat | -8.82319 |
| Strn-Mlck | Signal transduction, kinase | Serine/threonine protein kinase, tyrosine kinase catalytic domain | -12.2444 |
| ssh | Signal transduction, | Phosphatase | Tyrosine-specific protein phosphatase, dual specificity | -5.49466 |
| CG17735 | Ubiquitin ligase | ARN repeat; CG17735 | -2.33618 |
| Bruce | Ubiquitin ligase | Ubiquitin-conjugating enzymes, inhibitor of apoptosis (IAP) repeat | -5.63309 |
| CG11321 | Ubiquitin ligase | RING domain, ubiquitin ligase activity | -5.91971 |
| CG5604 | Ubiquitin ligase | ARN repeat; HECT domain, ubiquitin-protein ligase E3a | -9.54632 |
| skpB | Ubiquitin ligase | POZ domain, skpB, Skp1-Skp2 dimerization domains, skpB | -23.3075 |
| CG10107 | Ubiquitin-like | SUMO/Sentrin/Ub1-specific protease | -5.123 |

*Table 1. List of genes selected for secondary screening*

The functional annotation and Z-scores from the reconfirmation analysis using the CRY-luc degradation assay are listed.
However, the magnitude of the effect on TIM was less (Fig. 3A, B). It also partially blocked TIM degradation, ubiquitin-conjugating enzyme), a composite E2–E3 ligase, Bruce; the dual specific phosphatase, ssh; and the HECT domain-containing E3 ligase CG17735. All these candidates had a strong effect on CRY degradation in the cell culture studies. BRUCE/apollon/Bric6 is a large (528 kD) composite E2–E3 ligase with a BIR domain, a hallmark of inhibitor of apoptosis proteins [IAPs] (Bartke et al. 2004; Hay and Guo 2006). Indeed, BRUCE plays an anti-apoptotic role by inhibiting Reaper- and Grim-mediated apoptosis (Vernooy et al. 2002). Given the strong effect of BRUCE RNAi in blocking CRY degradation in S2 cells, we were interested in testing the phenotypes of Bruce mutations in flies.

We focused these studies on a line containing a piggyBac transposon insertion, Bruce<sup>coos984</sup> (referred to as Bruce<sup>Rb</sup>), in the coding region [exon 8] of the Bruce gene (Supplemental Fig. S1A). To assess the effects of this transposon insertion on Bruce mRNA levels, we performed RT–qPCR [Supplemental Fig. 1B]. Using a probe 3′ to the insertion site, we could not detect any transcript indicating that the piggyBac insertion results in premature termination of the Bruce transcript [Supplemental Fig. S1]. This would result in a truncated protein lacking the critical C-terminal UBC domain.

We next evaluated the role of BRUCE in regulating CRY and TIM stability following an acute light pulse at night. In the Bruce<sup>Rb</sup> line, we found that CRY levels were elevated under baseline conditions, and although degradation occurred in response to a 5-min or 60-min pulse, relatively high levels remained [Fig. 4A]. These results suggested that the major effect of Bruce knockdown is to increase the overall stability of CRY. To further investigate the role of BRUCE in regulating CRY stability, we evaluated CRY levels in a 12:12 light/dark (LD) cycle. In wild-type [control] flies, the light sensitivity of CRY results in an oscillation of protein levels in LD cycles, with higher levels expressed during the night. In Bruce<sup>Rb</sup> mutants, CRY levels were elevated in the dark and CRY protein persisted following the transition into the light period [Fig. 4B]. Since CRY does not cycle in constant darkness [Emery et al. 1998], indicating that its cyclic expression in LD cycles is driven entirely by its response to light, its incomplete degradation in an LD cycle in Bruce mutants supports the idea that light-dependent degradation of CRY in flies is affected by loss of Bruce. We note that the cell culture assay involves a 6-h exposure to light, and so it may be somewhat analogous to an LD cycle, thereby accounting for the robust effects of Bruce knockdown in this assay. In Bruce flies, levels of CRY were even higher during the day than at night,

**Figure 3.** Effect of select candidates on light-dependent CRY and TIM degradation in S2R<sup>+</sup> cells. (A) Western blot analysis showing effects of primary screen positives on light-dependent CRY and TIM degradation. S2R<sup>+</sup> cells were transfected with dsRNA and then either exposed to light for 2 h or maintained in constant darkness [black bar]. CRY [top] and TIM [middle] were visualized via Western analysis using V5 and TIM antibodies, respectively. (Bottom) Endogenous HSP-70 levels were used as a loading control. (B) Quantification of the effects of different dsRNAs on CRY degradation. CRY levels were quantified, and differences in average intensities from two experiments were normalized to the dark values and plotted. Error bars represent SEM. Similar results were obtained by using a myc-tagged CRY [data not shown]. (C) Quantification of the effects of different dsRNAs on CRY degradation. Relative light-dependent TIM degradation was measured by similar quantification of the TIM band. Based on these values, a relative TIM or CRY degradation score [see the Materials and Methods] was obtained and plotted.

In the secondary screen, dsRNA against Bruce [for baculovirus inhibitor of apoptosis repeat [BIR]-containing ubiquitin-conjugating enzyme], a composite E2–E3 ligase, was most effective in blocking CRY degradation [Fig. 3A,B]. It also partially blocked TIM degradation. However, the magnitude of the effect on TIM was less than that on CRY. Interestingly, Bruce RNAi also affected baseline levels of CRY when it was fused to a tag other than the one used in Figure 3, suggesting that BRUCE may affect overall CRY levels under some conditions [data not shown].

**In vivo validation of candidate genes that affect CRY degradation**

In order to validate results of the cell culture screen in vivo, we examined flies mutant for each of three candidate genes: the E2–E3 ubiquitin ligase, Bruce; the dual specific phosphatase, ssh; and the HECT domain-containing E3 ligase CG17735. All these candidates had a strong effect on CRY degradation in the cell culture studies.

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Figure 4. Effect of BRUCE on light-dependent CRY and TIM degradation. (A) Bruce mutants have elevated CRY levels that persist even after prolonged light exposure. A representative Western blot image of light-dependent CRY and TIM degradation in yellow white (control) or BrucexRB mutants following an acute light pulse [5 min] or prolonged light pulse [60 min] as compared with the dark control (D). High levels of CRY protein were observed in BrucexRB mutants, and CRY was not efficiently degraded even after a prolonged light exposure. (B) Western blot analysis of CRY and TIM levels in control and BrucexRB mutants in 12:12 light/dark (LD) cycle. Fly heads collected during the light phase (open bar) or dark phase (dark bar) were probed for TIM, CRY, and HSP-70 [loading control]. In control flies, CRY levels are high at late night (ZT20), but in BrucexRB mutants, CRY levels are low at this time point. Also the overall CRY levels are elevated by more than fivefold in BrucexRB mutants. TIM levels are low during the day and high at night in both wild-type and BrucexRB mutant flies. (C,D) Quantification of light-dependent CRY and TIM degradation in Bruce mutants. CRY and TIM levels were quantified in the samples described in A and normalized to levels of the respective protein in the control line maintained in the dark. HSP70 levels were determined to control for loading. BrucexRB mutants show significantly elevated CRY levels when compared with wild-type siblings and Bruce heterozygotes [P < 0.008]. Light-dependent degradation of CRY was also reduced in BrucexRB mutants [in the data shown here, levels of CRY were significantly reduced in the heterozygous sibling [P < 0.01] but not in the mutant [P = 0.1]]. Statistical analysis was conducted using ANOVA followed by a post-hoc Tukey HSD test. An average of four independent experiments is shown.

perhaps because of the higher levels of cry mRNA in the daytime. We infer that normally light-induced CRY degradation negates any influence of cry mRNA cycling on protein expression, but in Bruce mutants, the attenuated response to light unmasks such regulation. The effects of BRUCE on CRY levels were supported by analysis of a fly line carrying an RNAi construct that knocks down expression of Bruce specifically in the eye (data not shown).

The cell culture results suggested that BRUCE may have a minor effect on TIM; however in BrucexRB mutants, the regulation of TIM was largely unaffected, either following an acute light pulse or under LD cycles [Fig. 4A,B]. Thus both cell culture and fly experiments show that BRUCE specifically regulates CRY stability. Since some degradation of CRY occurs following acute light exposure in Bruce mutants, we suggest that there are redundant mechanisms for regulating light-dependent CRY degradation. In fact, the screen identified a novel E3 ligase, one of which, CG17735, we further characterized [see below]. We also assayed behavioral rhythms of BrucexRB mutant and found that many of the flies either lacked rhythms altogether or had weak rhythms [data not shown]. This effect on rhythms was not accompanied by any major defects in the clock neurons [Supplemental Fig. S2] or in the molecular oscillations of clock proteins in LD cycles or constant darkness [Fig. 4B, data not shown]. However, this weak rhythmicity, which may or may not map to Bruce, made it difficult to assess the resetting of behavioral rhythms in these flies by light.

To assess the contribution of the HECT domain-containing novel E3 ligase CG17735 to light-dependent CRY degradation, we generated two independent fly lines [541 and 302] in which CG17735 expression was knocked down through a microRNA [mir6.1]-based RNAi approach described previously [Supplemental Fig. S3A; Chen et al. 2007]. We expressed the UAS-mir 6.1-CG17735 construct under the control of a tim-Gal4 driver so as to drive expression in all clock cells and verified through qPCR that levels of CG17735 mRNA were knocked down [Supplemental Fig. S3B]. In CG17735 knockdown flies, CRY degradation, but not TIM, was reduced, in response to an acute light pulse [Fig. 5A]. We note that it did not completely block CRY degradation by light; this could be either because the knockdown was not complete or because other proteins can also mediate light-dependent CRY degradation. Thus, Bruce may regulate overall CRY stability while CG17735 regulates CRY degradation following acute light exposure.

Interestingly, free-running rhythms of CG17735 knockdown flies had periods that were considerably longer than those of controls: ~27 h as compared with 24 h in controls [Supplemental Fig. S3C]. This difference in period shifted the phase response curve [PRC] of these flies such that a delay, rather than an advance, occurred in response to a light pulse at ZT20 [Supplemental Fig. S3D]. However, at ZT22 an advance was seen in both control and experimental flies. Thus, overall the CG17735 knockdown flies did not display major defects in the behavioral response to light.

Finally, we determined the effects of a mutation in the dual specific phosphatase, ssh, on light-dependent CRY stability. Null mutations of ssh are lethal [Niwa et al. 2002], and so we characterized a heteroallelic combination of two hypomorphic mutations, ssh1−6 and ssh31, that disrupts ssh expression [Niwa et al. 2002]. Following an acute light pulse at ZT20, higher levels of residual
CRY were observed in the ssh hypomorphic mutant (Fig. 6A). However after prolonged light exposure (60 min), CRY degradation was equivalent to that in controls (data not shown). The response of TIM to light was normal (Fig. 6). We also examined CRY and TIM levels in a 12:12 LD cycle. As in Bruce mutants CRY levels were elevated in the dark, and increased residual CRY was detected following transition into light at ZT2 (Fig. 6B), indicating that light-dependent degradation is impaired under these conditions. Free-running behavioral rhythms were normal, and their response to light was largely intact in the ssh hypomorphic mutants (Supplemental Fig. 4). These in vivo results support the cell culture data showing that CG17735, ssh and Bruce affect CRY stability in response to light.

Discussion

Using a luciferase-based CRY degradation assay, we employed a genome-wide RNAi knockdown approach to identify genes required for the CRY light response. A subset of these genes was further validated by secondary assays. To place these genes in the context of the circadian light entrainment pathway, we determined whether they are required for the degradation of the TIM clock protein by light. In addition, we investigated the role of two ubiquitin ligases (CG17735 and BRUCE) and a dual specific phosphatase (SSH) on light-dependent CRY degradation in vivo. Utilizing this unbiased approach, we identified several new genes, including protein kinases, phosphatases, molecules involved in regulating cellular redox, and ubiquitin ligases, none of which were previously identified as regulators of circadian rhythms. Further, our in vivo analysis validates the cell-based screen and demonstrates novel roles for the composite E2–E3 ligase BRUCE, the E3 ligase CG17735, and the phosphatase SSH in regulating CRY stability.

CRY contains a flavin and a pterin-binding pocket, and intramolecular electron transfer has been reported in its activation by light (Lin et al. 2001; Froy et al. 2002). In fact, if the cellular redox system is compromised, light-induced CRY degradation is impaired (Lin et al. 2001). However, the identity of redox molecules that participate in the circadian light response has remained largely unknown. Here we report the identification of three thioredoxin domain-containing redox molecules (GstE7, Txl, and CG11790) and a cytochrome p450 CYP49a1 that modulate light-dependent CRY and TIM degradation. These results suggest that cellular redox status and electron transfer modulate light-dependent activation of CRY, which in turn affects subsequent transmission of the light signal to TIM and the degradation of CRY itself.

The screen also identified a novel role for the TH gene, ple, in regulating light-dependent CRY and TIM degradation. TH catalyzes a rate limiting step in the synthesis of L-Dopamine [DA] and requires the cofactor tetrahydrobiopterin (BH4). S2 cells have been used successfully to demonstrate the role of DA signaling in FGFR receptor endocytosis and cytoskeleton rearrangement (Hsouna et al. 2007); thus, they may express components...
of the dopamine pathway. The mechanism of action of TH and DA signaling in light-dependent CRY degradation is not clear. However, TH has been implicated in oxidative mechanisms that lead to the production of free radicals, which may underlie its effects on CRY degradation (Adams et al. 1997).

A number of signal transduction molecules involved in the insulin signaling and IMD pathways were also identified by the screen. A phosphatase SSH appears to be of particular importance for the regulation of CRY. Knockdown of ssh blocks light-dependent CRY degradation in cultured cells and also affects CRY stability in flies. However, it has little effect on TIM degradation. Thus, dephosphorylation of CRY, or a component required for CRY degradation, by SSH is important for regulating CRY stability. Currently, we presume that dephosphorylation of CRY, or a component related to it, is not clear.

Although we identified various components of the CRY degradation complex, the importance of light-dependent CRY degradation to circadian resetting behavior is not clear. Bruce mutants demonstrate a strong defect in CRY degradation; however, due to the weak behavioral rhythms in these mutants, the effect on circadian photic sensitivity could not be addressed. Also we note that light-dependent CRY degradation, although reduced, still persists in BruceRB mutants, suggesting that it is regulated by redundant pathways. The two mutants assayed ssh and CG17735 did not completely abolish CRY degradation either; in this case, it may be because neither is a null mutant. Interestingly, TIM...
degradation was not affected in any of these mutants, suggesting that they specifically affect CRY degradation. TIM is targeted for light-dependent degradation by an F-box-containing protein, JET, which has minimal effect on CRY degradation [Koh et al. 2006]. This result again supports the existence of two distinct destruction complexes that regulate CRY and TIM degradation. However, some aspects of the degradation pathways may be similar. The demonstration that SkpB, a component of the SCF complex, is critical for both CRY and TIM degradation suggests that some components are shared between the two destruction complexes. In addition, components required for the activation of CRY are presumably also required for the TIM light response. It is likely that other classes of molecules identified in the screen, such as those that affect redox status, are involved in CRY activation and therefore also affect TIM and the behavioral response. Further analysis of these molecules in clock neurons should address the role of redox and CRY degradation in circadian photosensitivity.

Materials and methods

Expression constructs and cell culture

The pIZ-luc-cry plasmid was generated by fusing the luciferase (luc) gene with cry. The coding region of luc was amplified by PCR using Pfu polymerase [New England Biolabs] from pGL3-Basic [Promega], and cloned in-frame with wild-type cry or cry carrying a point mutation, cry<sup>R</sup>, as an N-terminal or C-terminal fusion in pIZ-V5 [Lin et al. 2001]. In another plasmid, a myc tag sequence was added to the N terminus of cry or cry<sup>R</sup> cDNA (Busza et al. 2004), and a stop codon was introduced at the end of cry and cloned into pIZ-V5/His A [Invitrogen]. Insect cell expression constructs for jet, tim were described previously [Ceriani et al. 1999; Koh et al. 2006]. A Remilla luciferase gene expressed under the control of Actin5C or RNA polIII promoter was described previously [Darlington et al. 1998; Nybakken et al. 2005]

S2R<sup>+</sup> cells from the Drosophila RNAi Screening Center (DRSC; http://flyrnai.org) were grown in Schneider's medium (DRSC; http://flyrnai.org) were grown in Schneider's medium (DRSC; http://flyrnai.org) were grown in Schneider's medium [pGL3-Basic (Promega), and cloned in-frame with wild-type cry or cry<sup>R</sup> cDNA (Busza et al. 2004), and a stop codon was introduced at the end of cry and cloned into pIZ-V5/His A [Invitrogen]. Insect cell expression constructs for jet, tim were described previously [Ceriani et al. 1999; Koh et al. 2006]. A Remilla luciferase gene expressed under the control of Actin5C or RNA polIII promoter was described previously [Darlington et al. 1998; Nybakken et al. 2005]

SIR<sup>+</sup> cells from the Drosophila RNAi Screening Center (DRSC; http://flyrnai.org) were grown in Schneider's medium containing 10% FBS, split at 1:8 dilution, and maintained at 25°C. Transfection was performed using Effectene [Qiagen] with expression constructs according to the manufacturer's instructions. Thirty-six hours post-transfection, cells were exposed to bright light in the presence of DMSO or the proteasome inhibitor MG132. All cells were harvested and processed for Western blot analysis or luciferase assay.

RNA-mediated interference

For RNAi experiments performed independently of the DRSC, DNA fragments 600–800 base pairs [bp] in length, containing various coding regions of Prosβ2, morgue, Dms13, or Slimb, were amplified with primers containing T7 promoters at both the 5′ and 3′ ends, and dsRNA was generated by T7 transcription and annealed as described previously [Sathyaranayanan et al. 2004]. Prosβ2 was amplified using the following primers: 5′-ACTGC CGCAAGATCCATTAC-3′ and 5′-GTGGAGGTCC GGTCTTTGAA-3′; morgue was amplified using the following primers: 5′-AGACATCAGC TCTTTGAGG-3′ and 5′-CAGC CGA TACCAGT TTTGAT-3′; and Dms13 was amplified using 5′-CTGCTACTGCTTTGACGTCGCTCATTGAGACGG-3′ and 5′-ACAGTGTCATTGTGATCGACGACACCTGGGG-3′. A T7 promoter sequence [GAATTAATACGACCTCACTATAGG GAGA] was added to the 5′ end of both forward and reverse primers.

Primary screen and analysis

Two sets of 58 plates [384-well] containing 0.08 µg of dsRNA per well were provided by the DRSC. The primary screen was transfection-based, utilizing a dual luciferase assay. The N-terminal luc fusion construct of cry, pIZ-luc-cry [12 ng per well], was mixed with pAct-Renilla-luc [9 ng per well], and a lipid complex was prepared with Effectene [300 µL per plate] and enhancer [250 µL per plate] in Buffer EC [Qiagen]. The DNA–effectene mix was transferred to the screening plate by using multidrop 384 and mixed gently. Cells [2.6 × 104 per well] in complete medium were added to the plate. Plates were wrapped in foil and incubated in dark for 4 d at 25°C in a humidified container. On the fifth day, one set of plates was exposed to white light for 12 h, while the other was wrapped in two layers of foil and placed adjacent to the light-exposed plates. Firefly and Renilla luciferases assays were performed using the Dual-glo luciferase assay [Promega] according to the manufacturer’s instructions. All cell culture medium was aspirated off the cells, an equal volume of Dual-Glo Luciferase reagent was mixed with complete medium, and 40 µL of the mix were dispensed using a MicroFill [BioTek]. Luciferase levels were read using an Analyst HT plate reader [Molecular Dynamics]. Subsequently, Renilla luciferase activity was measured. Each receptacle plate was assigned a number and a bar code for automatic identification by the plate reader. The transient transfection efficiency was estimated by utilizing a GFP expression construct. About 25%–30% of the cells were GFP-positive at the end of the assay [data not shown].

To identify genes involved in regulating light-dependent CRY-luc degradation, the luciferase activity was normalized to Renilla luciferase values to give a normalized luciferase score. The normalized luciferase scores were compared between light [L] and dark [D] samples, and the relative light-dependent change in luciferase level was calculated [D – L/D]. Using the relative light-dependent change in luciferase levels, a Z-score for each well was determined based on the interior-well plate average and interior-well plate standard deviation [as described in Nybakken et al. 2005]. Z-scores are a measure of how different a particular value is from the mean in terms of the number of standard deviations from that mean. The interior wells are all wells except those on the outer edges of the plate, which, we found empirically, it was best not to include in these calculations due to occasional edge effects found in some plates. Z-scores from duplicate screens were averaged, and the average Z-score was used to identify hits. The artificial change in normalized luciferase scores arising from a decrease in renilla luciferase levels observed at the edges of the plates or in one of the two replicate plates was eliminated by comparing dark and light levels of luciferase. Most of the Heidelberg collection [plates 35–54] did not target legitimate genes [many target noncoding RNA] and were not included in the second replicate. Due to a jam in the MicroFill, addition of luciferase reagent to most of the wells in plate numbers 45 and 53 was affected, and thus these results were eliminated from analysis. Among the two replicates experiments, results that were found to be specific replicate showed less variability. In order to get maximum coverage, positives [± 2 Z-score] from the second replicate were also included in the secondary screen.

From the primary screen, positives corresponding to annotated genes were selected for reconfirmation. Hits that were predicted to target a number of genes [off-targets] were eliminated. The remaining positives from the primary screen were cherry-picked, and DNA templates were amplified by PCR us-
ing T7 primers. The length of the amplified product was checked on an agarose gel and purified using Montage PCR cleanup kit (Millipore). dsRNA was generated by T7 transcription (Sathyanarayanan et al. 2004). Reconfirmation of the hits was performed by utilizing the primary screen assay. pIZ-locquency and pAct-remilla luciferase constructs and dsRNA were cotransfected into S2R+ cells as described above. dsRNA against gfp was utilized as a negative control, and Pros82 was utilized as a positive control. Here the Z-scores were calculated utilizing GFP values as a mean ($n$ = 3).

**Western blot analysis**

Cells (S2R+) were cotransfected with 320 ng of dsRNA and 25 ng of pLI-cry-V5 (or pLI-myc-cry) along with 50 ng of pAct-tim and 50 ng of pFlag-jet in a 1:2 well plate. Five days post-transfection, one set of sample was exposed to light while the other was kept in the dark for 2 h. Cells were washed with PBS and processed for Western blot analysis as described in Sathyanarayanan et al. (2004). The blots were probed with anti-V5 antibody (Invitrogen), anti-TIM antibody (UPR8), and anti-HSP70 antibody (Sigma). A myc epitope tag mouse monoclonal antibody (Santa Cruz Biotechnologies) was utilized to probe for myc-tagged CRY (1:1000). The blots were quantified using a Kodak Image Station, and images were captured by using a Leica confocal microscope. Hemispheres were examined per condition. Immunofluorescent images were obtained with a Leica confocal microscope.

**Fly lines, locomotor activity measurements, and statistics**

miRNA-based CG17735 RNAi lines 541 and 302 were generated by cloning a hairpin targeting CG17735 into the pUAST plasmid; Kent Nybakken for the PolIII-rluc plasmid; members of the DRSC for providing the genome-wide RNAi library and for help with RNAi screening, Patrick Emery for the anti-CRY antibody, Tadashi Uemura for soh fly stocks; Yanshan Fang for the pIZ-myc-cry plasmid; Kent Nybakken for the PolIII-rluc plasmid; members of the Perrimon laboratory for help with the assay set-up, and Kyunghue Koh and Susan Harbison for help with data analysis. S.S. and X.Z. were supported in part by NIH grant NS048471.

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Identification of novel genes involved in light-dependent CRY degradation through a genome-wide RNAi screen

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