Genetic Evidence That Captured Retroviral Envelope syncytins Contribute to Myoblast Fusion and Muscle Sexual Dimorphism in Mice
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Abstract

Syncytins are envelope genes from endogenous retroviruses, “captured” for a role in placentation. They mediate cell-cell fusion, resulting in the formation of a syncytium (the syncytiotrophoblast) at the fetomaternal interface. These genes have been found in all placental mammals in which they have been searched for. Cell-cell fusion is also pivotal for muscle fiber formation and repair, where the myotubes are formed from the fusion of mononucleated myoblasts into large multinucleated structures. Here we show, taking advantage of mice knocked out for syncytins, that these captured genes contribute to myoblast fusion, with a >20% reduction in muscle mass, mean muscle fiber area and number of nuclei per fiber in knocked out mice for one of the two murine syncytin genes. Remarkably, this reduction is only observed in males, which subsequently show muscle quantitative traits more similar to those of females. In addition, we show that syncytins also contribute to muscle repair after cardiotoxin-induced injury, with again a male-specific effect on the rate and extent of regeneration. Finally, ex vivo experiments carried out on murine myoblasts demonstrate the direct involvement of syncytins in fusion, with a >40% reduction in fusion index upon addition of siRNA against both syncytins. Importantly, similar effects are observed with primary myoblasts from sheep, dog and human, with a 20–40% reduction upon addition of siRNA against the corresponding syncytins. Altogether, these results show a direct contribution of the fusogenic syncytins to myogenesis, with a demonstrated male-dependence of the effect in mice, suggesting that these captured genes could be responsible for the muscle sexual dimorphism observed in placental mammals.
Author Summary

Syncytins are “captured” genes of retroviral origin, corresponding to the fusogenic envelope gene of endogenized retroviruses. They are present in all placental mammals in which they have been searched for, where they play an essential role in placentation via their cell-cell fusion activity. Here we show that they also contribute to myoblast fusion and muscle formation in development and repair after injury, using both in vivo knock-out mouse models and ex vivo primary myoblast cell cultures from several mammals, including humans, carnivores and ruminants. Interestingly, the effects observed in mice are sex-dependent, thus suggesting that the added “collateral” effect of syncytins on myogenesis could be responsible for the muscle sexual dimorphism observed in placental mammals.

Introduction

Syncytins are “captured” genes of retroviral origin that correspond to the envelope gene of ancestrally endogenized retroviruses (reviewed in [1]). These genes encode fusogenic proteins that are involved in the formation by cell-cell fusion of the placental syncytiotrophoblast in eutherian mammals and marsupials [2–5]. Furthermore, genetically modified mice, knocked out for their two syncytin genes—i.e. syncytin-A and syncytin-B [6]—proved to be deficient in placenta development, with altered structures of the materno-fetal interface resulting, in the case of Syncytin-A, in the death of the embryo at mid-gestation [7, 8]. It thereafter turned out that syncytins can be found in all placental mammals in which they had been searched for, with independently captured syncytins found in all major clades of placental mammals, including the Euarchontoglires (primates, rodents, lagomorpha), Laurasiatherians (ruminants, carnivores), Afrotherians (tenrec), and even the marsupials (opossum). This has led to the hypothesis that these genes, which are absolutely required for placentation, as shown by the knock-out mice experiments, are most likely responsible for the emergence of placental mammals from egg-laying animals. Analysis of the conservation of these “new” genes clearly indicated that they have been subjected to purifying selection in the course of evolution, as expected for any bona fide cellular gene. Of note, cell-cell fusion is a basic phenomenon for all species, from drosophila to humans, where such events are involved in numerous processes including, in addition to placentation in mammals, macrophage fusion for osteoclast formation, and most importantly myoblast fusion for myogenesis during development and for muscle regeneration after injury (reviewed in ref. [9–13]). Although syncytins are essentially expressed in the placenta trophoblast cells, low level expression can be detected in other cells such as fusing macrophages [14] and myoblasts [15], and we therefore tested, using genetically modified mice, whether syncytins could participate in muscle formation, thus revealing a “collateral” effect of the “primary” capture of these genes for placentation.

Myoblast fusion is a complex and tightly controlled process required for the formation of the skeletal muscle fibers (reviewed in refs. [9, 10, 16]). The fusion process is highly cell-type specific to ensure that fusogenic myoblasts do not form syncytia with non-muscle cells. The molecular mechanisms that coordinate myoblast fusion remain incompletely understood, although several partners of myoblast fusion have already been identified. These include numerous proteins involved in cell-cell adhesion, as well as proteins involved in actin and lipid dynamics that allow cytoskeletal and membrane remodeling for cell fusion. Furthermore, a muscle-specific membrane protein called myomaker that is transiently expressed during myogenesis and is both necessary and sufficient to promote myoblast fusion in vivo and in vitro has recently been identified [17, 18]. Although this protein alone is not sufficient to induce the
fusion of non-myoblast cells such as fibroblasts, myomaker is likely to be a major player of the membrane fusion process; yet its partner (either a receptor or a bona fide fusogenic ligand) remains to be identified. Altogether, the numerous studies carried out on myoblast fusion have led to the conclusion that these processes involve numerous partners and pathways. As syncytins have been shown to be bona fide fusogens, we investigated their involvement in the overall myogenesis process.

Results

Syncytin-B knockout male mice present a delay in muscle development

As previously reported, whereas deletion of both syncytin genes, i.e. syncytin-A and syncytin-B leads to embryonic lethality, knockout of syncytin-B alone allows for surviving progeny to be obtained. Yet, syncytin-B null (SynB−/−) neonates display a sex-independent growth retardation characterized by a reduction of body weight of 18% when compared to wild-type neonates from the same litter [8]. This difference was attributed to associated placental malformations and was still apparent in 6–8 week old male mice but absent in female mice [8]. In order to clarify whether the observed decrease in body weight of male mice could be attributed to a reduction in muscle mass, we measured, in adult 12 week old mice, both the body weight (Fig 1A and 1B) and the mass of several skeletal muscles, located in the hind limbs of the mouse: two fast-twitch muscles, i.e. Tibialis Anterior (TA) and Extensor Digitorum Longus (EDL), and one slow-twitch muscle the Soleus (SOL) (Fig 1C). Measurements were performed on F1 mice, either wild-type or SynB−/−, from at least four different crosses between heterozygous SynB+/− mice, in order to minimize possible lineage effects and compare genetically related individuals. As shown in Fig 1A and 1C, adult SynB−/− and WT female mice show no differences in body weight and muscle mass (17 wt and 13 SynB−/− females). However, adult SynB−/− male mice show a significant reduction in body mass (12%; p<0.01, Mann and Whitney test; 16 wt and 16 SynB−/− males). This decrease is associated with a reduction in muscle mass as compared to wild type male mice (15%; p<0.05, Mann and Whitney test), which is observed independently of the crossing (S1A Fig). We also quantified, in the corresponding animals, as an internal control, the mass of two organs (heart and kidney), and the length of two bones (tibia and femur), with no significant differences seen between SynB−/− and WT mice (Fig 1D). These last results suggest that the reduction in body weight in adult SynB−/− male mice is mainly due to a decrease in muscle mass (which accounts for 35–40% of the body weight). In addition, these decreases appear to affect only skeletal muscles, where fusion processes take place. Altogether these results suggest that Syncytin-B takes part into muscle development in male mice.

Syncytin-B knockout male mice exhibit a reduction in myofiber size and myonuclei number per fiber

To better characterize the role of syncytin-B in muscle development, morphological analyses of SynB−/− and WT muscles (see scheme in S2 Fig) were performed on cryosections of TA, EDL and SOL muscles, respectively. We first quantified the cross-sectional area (CSA) of the entire muscle, which was found to be reduced for the three muscles in SynB−/− males as compared to WT males (Fig 2A; * p<0.05, ** p<0.01, Mann and Whitney test). The alteration in muscle CSA was observed independently of the crossing (four crosses at least) (S1B Fig). As expected, in females, which did not show a reduction in muscle mass, no differences in CSA were observed between SynB−/− and wild-type mice. The number of fibers per muscle and the individual myofiber CSA, both parameters that contribute to muscle CSA, were also measured, as illustrated in Fig 2. Equivalent fiber numbers were observed between SynB−/− and wild-type
male mice (Fig 2B), but a significant reduction in myofiber CSA was observed in SynB−/− male mice (Fig 2C and 2D; \( p < 0.05 \), \( p < 0.01 \), Mann and Whitney test). The alteration in myofiber mean area was observed independently of the crossing (three to four crosses) (S1C Fig). Finally, we quantified the number of myonuclei per fiber, in knockout and wild-type mice (Fig 3).

Transversal sections of the three muscles TA, EDL and SOL were immunolabeled with a dystrophin antibody to outline the sarcolemma, and were stained with 4',6'-diamidino-2-phenylindole (DAPI) to visualize the nuclei. Nuclei located inside the dystrophin-labeled sarcolemma were identified as myonuclei and counted (Fig 3B). Accordingly, a decrease of approximately 30% in the myonuclei number was observed in muscles from SynB−/− male mice as compared to control males (Fig 3A; \( p < 0.05 \), \( p < 0.01 \), Mann and Whitney test), strongly suggestive of a defect in myoblast fusion. This difference was not observed in SynB−/− females in comparison to wild-type. As variations in nuclear size and shape may affect myonuclear count [19], we measured the nuclear length of myonuclei on longitudinal sections. No differences were observed between either WT or SynB−/−, male or female mice (S3A and S3B Fig). As expected,
the resulting number of myonuclei per mm of fiber (S3C–S3E Fig) showed a decrease in muscles from SynB−/− males as compared to control males, whereas no significant differences were observed in females. Since myoblast fusion and accumulation of myonuclei contribute to mammalian myofiber development and growth [20–22] the data also strongly suggest that the reduced myofiber CSA observed in SynB−/− male mice, which have an unaltered myofiber number, mainly results from a defect in myoblast fusion to myofibers.

**Syncytin-B plays a role during muscle regeneration in male mice**

Muscle regeneration is a physiological process that recapitulates, through the differentiation and fusion of satellite cell descendants, all the phases of myogenesis [16]. We therefore investigated the requirement for syncytin-B during muscle regeneration. We first analyzed, by in situ hybridization, the expression of syncytin-B upon muscle injury. Muscle injury was induced by a single injection of cardiotoxin into the TA muscle of 8 week old female and male mice. Injection of PBS in the contralateral muscle was used as a control. Muscles were collected after 4
days of regeneration (see scheme in Fig 4M). Hematoxylin, eosin and safran staining (HES) of the muscle sections (Fig 4A), shows normal myofibers after PBS injection, with myonuclei located at the myofiber periphery. In contrast, following cardiotoxin injection, similar damage of the muscle section could be observed (delineated by black arrows, Fig 4D and 4G) in male and female mice. During muscle regeneration, the damaged myofibers are destroyed and replaced by small newly centro-nucleated fibers, typically representative of muscle regeneration. These new fibers observed in male as in female mice, are illustrated in the enlarged Fig 4J for a male mouse. Specific digoxigenin-labeled antisense probes were then synthetized for the detection of syncytin-B, and the corresponding sense probes were used as negative controls. As further shown in Fig 4E and 4K, labeling was only observed with the antisense probe for syncytin-B, and only in males. Closer examination of the labeling further shows that it is restricted to regions of muscle regeneration (Fig 4D and 4E) where myofibers are centro-nucleated (Fig 4J and 4K). In situ hybridization of syncytin-B shows conclusively that this gene is expressed in the regenerative muscle in males, strongly suggesting that the encoded protein could play a role in this process specifically in males.

To characterize further the role of the Syncytin-B protein in this process, SynB−/− male and female mice (as well as control wild-type mice) were analyzed after cardiotoxin injection as above, and muscle regeneration was quantified at day 14. Histological sections of the muscle fibers were performed as above, and the mean myofiber CSA measured. As illustrated in Fig 4N, quantification of the myofiber CSA in SynB−/− male mice in comparison to wild-type shows a delay at day 14, suggesting that Syncytin-B plays a role during muscle regeneration (Fig 4N; p<0.05, Mann and Whitney test) after cardiotoxin treatment. In the case of female mice, no differences could be detected in the myofiber CSA between SynB−/− and wild-type, suggesting that Syncytin-B plays a role during muscle regeneration also in a sex-dependent manner (Fig 4N).

**Syncytin-A and -B are involved in myoblast fusion**

To characterize further the role of the syncytin genes in myoblast fusion, we isolated mouse primary satellite cells and quantified the expression of both syncytin-A and -B in proliferating...
myoblasts, as well as during the process of cell differentiation and fusion (day 0, day 2 and day 4, see Fig 5A). Both genes were detected by quantitative RT-PCR and were found to be upregulated after 2–4 days of myogenic differentiation and myoblast fusion (when myotubes could be clearly observed), whereas the level of the housekeeping gene remained stable. These data suggest that both syncytin genes could be involved in this process. To investigate further their role in myoblast fusion, we knocked-down syncytin-A and -B expression by RNA-mediated interference (Fig 5B). After two days of differentiation, silencing of both syncytin genes resulted in a very significant inhibition of myoblast fusion as illustrated in Fig 5C, with a close to 50% decrease in the fusion index (Fig 5D). Analysis of the nuclei distribution in myoblast cells revealed a significantly higher number of mononucleated cells in cells transfected with either...
syncytin-A or syncytin-B siRNAs in comparison to cells transfected with a control siRNA (Fig 5E; * p<0.05, ** p<0.01, Student’s t-test). In addition, a significantly higher number of cells with >4 nuclei was observed in control cells in comparison to cells transfected with syncytin-A or syncytin-B siRNAs, with no myotube containing >8 nuclei in both cases. Combination of the two syncytin siRNAs had no additive effect (Fig 5D) suggesting that, at least ex vivo, the two syncytins are acting on similar—and most probably interactive—steps of myoblast fusion. Of note, reduction in fusion index mediated by siRNA targeted to syncytins remains smaller than that with a siRNA targeted to the myomaker gene (ref. 17 and Fig 5D), consistent with the notion that syncytins are only “add-on” contributors to the basal fusogenic activity associated with the non-syncytin genes. Yet both syncytins are able to trigger myoblast cell-cell fusion. This is illustrated in Fig 6, where transfection of C2C12 myoblasts with expression vectors for either syncytin-A or syncytin-B clearly triggered the fusion of myoblasts in their proliferative state (Fig 6A), and also enhanced the fusion normally triggered upon differentiation of the cells (Fig 6B and 6C). Altogether, the ex vivo data corroborate the in vivo data on syncytin-B, and further suggest that syncytin-A could also contribute to myoblast fusion.
The role of syncytin genes in myoblast fusion extends to other mammals

To determine if the effects observed in the mouse can be extrapolated to other placental mammals, we established primary cultures of myoblasts from the muscles of mammals where the cognate syncytins have been characterized. Accordingly, we obtained viable and fusion-prone myoblasts from human (primate) (Fig 7A), sheep (ruminant) (Fig 7B) and dog (carnivore) (Fig 7C). As illustrated in Fig 7, left panels, induction of each cognate syncytin gene could be observed during the process of cell differentiation and fusion, as similarly observed for the murine myoblasts (Fig 5), whereas the level of the housekeeping genes remained stable. Experiments were then carried out as above for the mouse myoblast cells, but using siRNAs specific for each cognate syncytin (i.e., syncytin-1 and -2, syncytin-Rum1 and syncytin-Car1 respectively). Accordingly, transfection of the cells with siRNAs specific for each cognate syncytin clearly resulted in a significant (>20%) reduction in the measured fusion index (Fig 7, middle panels), also visible in the reduction in the number of large myotubes (Fig 7, right panels).
Discussion

In this study, we address for the first time the role of the two murine syncytin genes (syncytin-A and -B), in myogenesis, both in vivo using a syncytin-B knockout mouse model, and ex vivo using primary cell cultures derived from isolated muscle satellite cells. We show that both syncytin genes are expressed ex vivo during the fusion of myoblast cells and that syncytin-B is expressed in regenerative fibers in vivo following cardiotoxin injury. Using syncytin-A and -B siRNAs, we demonstrate ex vivo that both syncytin genes contribute to myoblast cell fusion. In addition, using a knockout mouse model for syncytin-B, we show that impairment of syncytin-B leads to a reduction in muscle mass, that largely contributes to the observed reduction in body weight. This effect is male-specific, being not observed in females, that provide a clear-cut internal control. This decreased mass correlates with an observed reduction in several muscle parameters, including the muscle cross-sectional area, the individual myofiber cross-sectional

Fig 7. Role of syncytins in myoblasts from human, sheep and dog. Left panels: level of expression of the human syncytin-1 and -2 (A), the ruminant syncytin-Rum1 (B) and the carnivore syncytin-Car1 (C) genes (arbitrary units, A. U.) during proliferation and differentiation of primary myoblasts from human, sheep and dog, as determined by quantitative RT-PCR and normalized by the beta2-microglobulin, SDHA and PPIA housekeeping genes, respectively; the fold-change in gene expression was expressed relative to the values of proliferating myoblasts, arbitrarily settled to one. Middle and right panels: impact of siRNA-mediated knockdown of syncytins on ex vivo cell-cell fusion of myoblasts from human, sheep and dog, respectively. Cells were transfected with siRNA (control or specifically designed against the corresponding syncytins), and differentiation induced 2 days later. Following 2 days of differentiation, cells were fixed in 4% PFA, stained with an anti-desmin antibody (muscle cell marker), and nuclei were counterstained with DAPI. The fusion index (number of nuclei in cells with >2 nuclei per total number of nuclei) was determined (middle panels). Representative images of desmin-immunolabeled and DAPI-stained myoblasts transfected with either control or syncytin-specific siRNAs, respectively, are shown (right panels). Data are the mean ± SEM (at least three independent experiments; *p<0.05, **p<0.01, Student’s t-test).

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area, and most importantly the number of nuclei per muscle fiber. Furthermore, we show that the total number of fibers per muscle is not altered in syncytin-B knockout mice, suggesting that syncytin-B activity is essentially involved in myofiber growth and not in the initiation of myogenesis. The reduction in the number of nuclei per myofiber would be consistent with such a role and with the fusogenic activity demonstrated in the ex vivo experiments. Finally, using a model of muscle regeneration after injury by cardiotoxin injection, we show that impairment of syncytin-B results in a delay in muscle regeneration, again only observed in males, consistent with the specific induction of syncytin-B expression observed in wild-type regenerating male myofibers. To summarize, the present data indicate that syncytin-B (and possibly syncytin-A, see below), participates in myoblast fusion, and contributes to muscle growth and regeneration in the mouse, in a sex-dependent manner.

This series of results raise several issues. One of the most obvious questions is related to the function of syncytin-A. Indeed, our cell fusion assays performed in C2C12 cells and in primary myoblasts strongly suggest that both syncytins have closely related activities, with similar effects of synA and synB siRNAs on myoblast fusion. This would fit with the known properties of these two genes, which have very similar activities at the placental level, each one being involved in the formation of a definite syncytiotrophoblast layer at the feto-maternal interface [8]. However a definitive characterization of the role of syncytin-A in myogenesis will require its conditional knock-out, specifically in the muscle, because homozygous null placentae are embryonic lethal. Along these lines, a mouse model obtained by crossing a lox-syncytin-A knock-in mouse with a Pax7-Cre-ERT2 mouse [23] is being developed, which should allow to generate a muscle-specific knock-out of syncytin-A, thus circumventing the detrimental effect of syncytin-A inactivation in placentation.

Another important issue concerns the generality of the contribution of syncytin genes to myogenesis. We have clearly demonstrated that not only the murine syncytins but also syncytins from ruminants, carnivores and humans contribute to myoblast fusion, at least in ex vivo assays. Similar conclusions were reached for human myoblasts [15, 24], where either siRNAs or an antibody targeting Syncytin-1 inhibit cell fusion of cultured myoblasts. Although the physiological relevance of such assays can be questioned, they nevertheless strongly suggest that the effects observed for the murine genes might extend to all species possessing a syncytin. Indeed, in the present study, we tested all four species with identified syncytins for which we succeeded in establishing primary myoblasts prone to cell fusion (we could not isolate myoblasts from the rabbit, in which we had previously characterized syncytin-Ory1, [25]), and in all four cases the results were identical, with inhibition of fusion by the corresponding syncytin gene-specific siRNA. It is therefore tempting to speculate that syncytins, which have been captured in all placental mammals most probably primarily for their placental function, all contribute, as observed here in vivo in the mouse, to myogenesis. However, this hypothesis will remain difficult to demonstrate or invalidate unless, for instance, myoblast fusion is found to be independent of syncytin, in a species where the gene is present. At the molecular level, the myoblast fusion process is still an unresolved question, although it is clear that it involves a wide panel of genes (for a review see ref. [9, 12]). Some have been shown to be required for myogenesis in vivo (via the use of KO mice), such as the recently identified myomaker gene [17, 18], the contributing galectin gene [26], cell adhesion/signal transduction proteins [27] or a series of actin cytoskeletal remodeling factors (reviewed in [9, 28]). Yet, in all cases none of the identified genes are truly autonomous fusogens, but rather seem to be either co-factors, yet absolutely required for myogenesis, or partners of a still to be identified “universal” fusogen. Our data on myomaker transfection of C2C12 myoblasts (Fig 6) do not provide evidence for a direct fusogenic activity in the proliferating state (at variance with syncytin A and B). This result is consistent either with myomaker not being a bona fide fusogen or its fusion partner only being expressed upon myoblast
differentiation. In the case of the presently identified syncytins, these are bona fide fusogens, a function inherited from the envelope gene of the ancestral retroviruses from which they were captured. Their contribution to myoblast fusion, as well as to cytotrophoblast or macrophage fusion, could in that case be directly due to their fusogenic activities.

An additional important question arises from the unexpected result on the sex-dependent effect of syncytin-B in vivo, in myogenesis after birth and muscle regeneration after injury. The data clearly indicate that the syncytin-B effect is essentially an “add-on” specific to the male, not taking place in the female, and resulting in an increased number of nuclei per muscle fiber (consequently associated with an increased muscle mass) and an increased rate of muscle regeneration after injury. Then a question arises as to the molecular origin of this uncovered male-dependence. Among the possible mechanisms, a hormone-dependent regulation of syncytin expression would of course fit the data, and could be analyzed by searching for binding sites within the syncytin gene promoter/enhancer for hormone–dependent factors. Along these lines, it is clear that testosterone and the androgen receptor (AR) would be appropriate effectors, which could be tested by hormone injection to wild-type mice, with or without prior castration, or even in mice knocked-out for the AR gene [29]. It is also possible that factors known to act as transcription factors in the placenta (among others GCM1 [30, 31]) could also act as syncytin activators in the muscle. Experiments are now in progress to tentatively elucidate these points. It is also noteworthy that testosterone and/or sustained physical activity (the latter known to generate microlesion-and-repair cycles) have been reported to result in muscle mass increase [24, 32–34], with, in some cases, indications for an increase in myonuclei number per fiber [35, 36]. Moreover, one of these studies shows that long-term endurance exercise in male humans results in an increase in Syncytin-1 expression, among other genes, in muscle biopsies [24]. It will be interesting to carry out a similar study on female humans to determine whether syncytin-1 induction is male-specific, or not. Indeed, such sex-specific effects could be a hint for a generalized male-dependence of syncytin expression and role in myogenesis among placental mammals. If so, it is likely that syncytins could then be responsible for the muscle sexual dimorphism observed in mammals, where in most cases the male has a higher body and muscle mass than the female.

To conclude, we propose as a working model that syncytins have been captured primarily as a placental gene, but that some expression in the muscle is taking place (possibly as a consequence of regulatory processes and transcription factors shared by placenta and muscle), and that a “collateral” effect of the capture of such genes— with a clear-cut fusogenic activity— is an add-on contribution to myogenesis in males of placental mammals.

**Methods**

**Animal model, ethical agreement**

Targeted mutagenesis of syncytin-B gene, located on chromosome 14, was described previously [8]. Briefly, the mouse syncytin-B ORF (carried by a single 1.8-kb exon) was deleted by homologous recombination using a strategy based on the Cre/LoxP recombination system for generating KO mice. Male and female mice were on a mixed 129/Sv C57BL/6 background. They were housed under controlled conditions at 24°C, and were allowed free access to food and water in full compliance with the French government animal welfare policy. All the experiments were approved by the Ethics Committee of the Gustave Roussy Institute.

**Experimentally induced muscle regeneration**

Before manipulations mice were anaesthetized with an intraperitoneal injection of 0.1 ml per 10 g body weight of a solution containing 1 mg/ml xylazine (Bayer) and 10 mg/ml ketamine
(Merial) per mouse. Then, to induce muscle injury, 35 μl of Cardiotoxin (Ctx) (12 μM; Latoxan) were injected in a single injection into Tibialis Anterior (TA) muscle of 8 week old mice. Muscle regeneration was followed at different times depending on the experiments.

Histological and immunofluorescence analyses
Mouse Tibialis anterior (TA), Extensor Digitorum Longus (EDL) and Soleus (SOL) muscles were dissected, weighed and frozen in isopentane/liquid nitrogen baths. Cryopreserved tissues were then cut in 10 μm cryosections. Cryosections were cut in the same central region of the muscle for all analyzed animals. Muscle cryosections were fixed in 4% paraformaldehyde on ice for 8 min. Tissue sections were then saturated with 5% Goat serum 2% BSA solution for 30 min. Overnight immunolabeling for dystrophin (Leica Biosystems) and laminin (Sigma) was then performed at 4°C. Nuclei were counterstained with 4',6'-diamidino-2-phenylindole (DAPI) for 15 min at room temperature. Myofiber cross-sectional area (CSA) was measured from stained slides from randomly chosen fields using the ImageJ software (at least 150 myofibers were measured per condition).

In situ hybridization (ISH)
Freshly collected muscles were fixed in 4% (wt/vol) paraformaldehyde and embedded in paraffin. Serial sections (7 μm) were used for ISH. For the syncytin-B gene, three PCR-amplified syncytin-B fragments of 414, 511, and 370 bp, respectively (primers listed in S1 Table) were cloned into pGEM-T Easy (Promega). In vitro synthesis of the antisense and sense riboprobes was performed with SP6 or T7 RNA polymerase and digoxigenin 11-UTP (Roche Applied Science) after cDNA template amplification. Sections were processed, hybridized at 42°C overnight with the pooled riboprobes, and incubated further overnight at 4°C with alkaline phosphatase-conjugated anti-digoxigenin antibody Fab fragments (Roche Applied Science). Staining was achieved with the nitroblue tetrazolium (NT) and 5-bromo-4-chloro-3-indolyl phosphate (BCIP) phosphatase alkaline substrates as indicated by the manufacturer (Roche Applied Science).

Primary muscle cell culture and fusion assay
Mouse primary myoblasts (female mice) were isolated from TA muscles of 4 week old mice by enzymatic digestion as described [37]. Myoblasts were grown on collagen-coated dishes in Ham’s F10 (Life technologies) growth medium supplemented with 20% fetal bovine serum (FBS), bFGF (10 ng/ml, Invitrogen), penicillin (100 U/ml) and streptomycin (100 mg/ml). For differentiation, myoblasts were plated at 10⁵ cells per well in collagen-coated 24-well dishes and differentiation was induced by switching the growth medium to the differentiation medium DMEM (Life technologies) with 2% horse serum (Biowest) for different times, the day of the induction being referred to as D0.

Dog (male) and sheep (female) myoblasts were obtained from biopsies of biceps femoris. The biopsy was cut into small pieces, treated with collagenase (Worthington; 5 ml of collagenase per g of tissue) and incubated for 1 h at 37°C. Using a syringe and 18G needle, the mix was dissociated and filtered sequentially through 100 and 40 μm sieves. After centrifugation, the cellular pellet was suspended and in the case of dog myoblasts, cells were seeded in Myo1 medium (Hyclone) containing 20% FBS, gentamycin (25 μg/ml, Sigma), bFGF (10 ng/ml, Invitrogen) and dexamethasone (1 μM, Mylan). Sheep myoblasts were cultivated on collagen-coated dishes in DMEM medium (Life technologies) with 20% FBS, bFGF (10 ng/ml, Invitrogen), penicillin (100 U/ml) and streptomycin (100 mg/ml). For differentiation, growth medium
was replaced with DMEM medium (Life technologies) supplemented with 2% horse serum (Life technologies).

Primary human myoblasts (female) were cultivated in proliferation medium (4 vol. of DMEM, 1 vol. of 199 medium, 20% FBS, gentamycin (50 mg/ml)). Differentiation was induced by replacing the proliferation medium by DMEM supplemented with insulin (10 mg/ml).

For desmin immunolabeling, cells were fixed for 10 min with 4% formaldehyde and rinsed with PBS. Cells were permeabilized for 10 min with PBS-0.1% Triton (Sigma) and blocked with PBS-5% BSA (Sigma) for 45 min before incubation with antibodies. Primary mouse anti-desmin anti-human antibodies (clone D33, Dako) were incubated overnight at 4°C and revealed using secondary Alexa 488 goat anti-mouse antibodies (life technologies) for 1h. Nuclei were stained with DAPI (4',6' diamidino-2-phenylindol) for 15 min at room temperature.

The fusion index was calculated as the number of nuclei within cells with ≥2 nuclei at day 3 of differentiation (at least 600 nuclei were counted for each condition).

**RNA interference**

RNA interference was performed using Lipofectamine RNAi-MAX reagent (Invitrogen) according to the manufacturer recommendations. Cells were transfected 2 days before induction of differentiation. All siRNAs, including the siRNA control oligonucleotides, were synthesized by Dharmaco or Eurogentec. SiRNA sequences are listed in S1 Table.

**C2C12 cell culture**

The C2C12 mouse myoblast cell line (ATCC: CRL1772) (female mice) was grown in DMEM (Dulbecco’s Modified Eagle medium, life technologies) supplemented with 10% fetal calf serum (FCS), 100 U/ml penicillin and 100 μg/ml streptomycin. Differentiation was induced by switching the growth medium to differentiation medium (DMEM with 2% horse serum (Bio-west)) for four days.

**Plasmids and cell transfection**

The GFP expression vector peGFP-C3 was purchased from BD Biosciences and the murine myomaker gene in pCMV-SPORT6.1 from Dharmacon. The control (phCMV-none) and syncytin-A and –B expression vectors were previously described [6]. Transfection of the C2C12 cells was performed using Lipofectamine LTX (ThermoFischer) according to the manufacturer’s protocol.

**Quantitative real-time RT-PCR**

Total RNAs were extracted from mouse, sheep and human myoblast cells using Trizol according to the manufacturer’s instructions (Sigma Aldrich). Total RNAs were extracted from dog myoblasts using the nucleospin RNA XS kit (Macherey Nagel) according to the manufacturer instructions. The amount of extracted RNAs was evaluated using a NanoDrop spectrophotometer (Thermo Scientific, Wilmington, USA). Syncytin mRNA expression was determined by Real-time RT-PCR (RT-qPCR). Reverse transcription was performed on 500 ng of DNase-treated RNAs (Ambion) according to the manufacturer’s instructions. Quantitative PCR was performed on 5 μL of diluted (1:5) cDNA in a final volume of 25 μL using the SYBR-Green PCR Master Mix (Applied Biosystems) in an ABI PRISM 7000 sequence detection system. The parameters used were as follows: 2 min incubation at 50°C, 5 min at 95°C, followed by 40 cycles of repeated incubations at 95°C for 10 s and 60°C for 30 s. For each cDNA sample, duplicates were analyzed and data normalized to the housekeeping genes listed in S1 Table.
Statistical analysis

Histological analysis of the muscle phenotype in 12 week old mice and the muscle regeneration in 8 week old mice was performed on at least 150 myofibers randomly chosen from three different tissue sections. The number of myonuclei was estimated by counting the number of myonuclei in at least 300 myofibers randomly chosen from three different tissue sections per condition. Statistical analyses were performed using the Mann-Whitney test. A p value <0.05 was considered as significant (* p<0.05, ** p<0.01). When the number of samples was <4, Student’s t-test was used and differences were considered significant when p <0.05 (* p<0.05, ** p<0.01). For the *ex vivo* fusion assays, according to the number of samples, a Mann-Whitney test (n = 4, for mouse and sheep myoblast cells) or a Student’s t-test (n = 3, for C2C12, human and dog myoblast cells) was used and differences were considered significant when p <0.05 (* p<0.05, ** p<0.01).

Supporting Information

S1 Fig. Muscle mass, muscle CSA and myofiber area of individual 12 week old SynB−/− and WT mice with mention of the cross from which they originate. Individual EDL muscle mass (A), muscle CSA (B) and myofiber area (C) from wild-type (+/+) and SynB KO (-/-), male (m) and female (f) mice. The crosses from which the mice originate were labelled by 4 to 5 different markers (blue square, green triangle, yellow circle, dark blue cross, red star). The values represented by the same marker in WT and SynB KO mice in the same graph correspond to that of individual mice from the same cross. The mean muscle mass, muscle CSA and myofiber area are indicated by horizontal black lines.

(TIF)

S2 Fig. Simplified scheme of skeletal muscle structure.

(TIF)

S3 Fig. Myonuclear length and number of myonuclei per mm of myofiber in 12 week old SynB−/− mice. (A) Measure of the average myonuclear length of TA muscles from WT (+/+), SynB−/− (-/-), male (m) and female (f) mice. Data are the mean ± SEM (at least 60 myonuclei measured for each type) (B) Anti-dystrophin and DAPI labelling of 10 μm longitudinal sections of optimum cutting temperature (OCT)-frozen TA muscles from 12 week old WT and SynB−/− male mice (scale bar: 10 μm). (C-E) Quantification of the myonuclei number per mm of fiber length in SOL, EDL and TA muscles of WT and SynB−/− male and female mice. Data are the mean ± SEM (4–6 mice analyzed per sex and per genotype; * p<0.05, ** p<0.01, Mann and Whitney test).

(TIF)

S1 Table. List of primers.

(DOCX)

S1 File. Supplementary Methods.

(DOCX)

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Author Contributions

Conceived and designed the experiments: FR NR AB CV VM VG NBG SC LT JD AD TH.

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