Thymocyte selection-associated high mobility group box gene (TOX) is aberrantly over-expressed in mycosis fungoides and correlates with poor prognosis

The Harvard community has made this article openly available. Please share how this access benefits you. Your story matters

Citation
Huang, Yuanshen, Ivan V. Litvinov, Yang Wang, Ming-Wan Su, Ping Tu, Xiaoyan Jiang, Thomas S. Kupper, Jan P. Dutz, Denis Sasseville, and Youwen Zhou. 2014. “Thymocyte selection-associated high mobility group box gene (TOX) is aberrantly over-expressed in mycosis fungoides and correlates with poor prognosis.” Oncotarget 5 (12): 4418-4425.

Citable link
http://nrs.harvard.edu/urn-3:HUL.InstRepos:12785931

Terms of Use
This article was downloaded from Harvard University's DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA
Thymocyte selection-associated high mobility group box gene (TOX) is aberrantly over-expressed in mycosis fungoides and correlates with poor prognosis

Yuanshen Huang1,2,3,*, Ivan V. Litvinov4,*, Yang Wang5,*, Ming-Wan Su1,2, Ping Tu5, Xiaoyan Jiang3, Thomas S. Kupper6, Jan P. Dutz2,7, Denis Sasseville4 and Youwen Zhou1,2,7

1 Molecular Medicine Lab and Chieng Genomics Centre, Vancouver Coastal Health Research Institute, Vancouver, BC, Canada
2 Department of Dermatology and Skin Science, University of British Columbia, Vancouver, BC, Canada
3 Terry Fox Laboratory, British Columbia Cancer Agency, and Department of Medical Genetics, University of British Columbia, Vancouver, BC, Canada
4 Division of Dermatology, McGill University Health Center, Montreal, QC, Canada
5 Department of Dermatology and Venerology, Peking University First Hospital, Beijing, China
6 Department of Dermatology, Brigham and Women’s Hospital, Harvard Skin Disease Research Center, Harvard University, Boston, MA, USA
7 Dermatologic Oncology Program, British Columbia Cancer Agency, Vancouver, BC, Canada

* These authors contributed equally to this work

Correspondence to: Youwen Zhou, email: youwen.zhou@ubc.ca
Denis Sasseville, email: denis.sasseville@mcgill.ca

Keywords: TOX, mycosis fungoides, cutaneous T cell lymphoma, marker, prognostic factor

Received: April 2, 2014 Accepted: May 27, 2014 Published: May 28, 2014

This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

ABSTRACT

Mycosis fungoides (MF) often mimics the common chronic inflammatory skin diseases and is difficult to be diagnosed with certainty, partly because of the lack of well-characterized molecular markers. Previously, we discovered that TOX, a key T cell development regulator, was aberrantly over-expressed in early stage MF. In the current multi-center study involving two independent patient cohorts, we determined the prevalence of TOX over-expression in the full spectrum of MF skin biopsies, and tested if TOX expression levels correlated with long term clinical outcomes. We examined TOX expression levels in 113 MF biopsies. We found that the MF biopsies expressed higher TOX mRNA than the controls in both cohorts (17.9 fold in cohort 1, P = 0.002; 5.8 fold in cohort 2, P < 0.0001). In addition, thicker skin lesions such as plaques and tumors expressed even higher TOX levels than thinner patches. Further, TOX over-expression differentiated MF from the controls (area under the curve [AUC]=0.87, P < 0.0001). Finally, high TOX mRNA levels correlated with increased risks of disease progression (P = 0.003) and disease-specific mortality (P = 0.008). In conclusion, TOX may be a useful marker for improving MF diagnosis and prognostication.

INTRODUCTION

The incidence of cutaneous T cell lymphoma (CTCL) is 6.4 per million and rising, making it the second most common extranodal non-Hodgkin lymphomas [1,4]. Characterized by clonal accumulation of post-thymic T cells residing in the skin [2], CTCL represents a group of diseases that are heterogeneous in clinical presentation and prognosis. The most common variants are mycosis fungoides (MF) and Sézary syndrome (SS), two distinct but related diseases, together representing more than 80% of CTCL[2-4]. In the early disease stages, which
can last several years, MF presents as flat erythematous skin patches resembling benign inflammatory dermatoses (BID), such as allergic contact dermatitis, eczema, or psoriasis; whereas in the later stages, MF cells gradually form plaques or tumors, and may disseminate to the lymph nodes and internal organs. The cancerous cells in some patients may appear in the peripheral blood, a hallmark of the leukemic stage of CTCL[2].

At present, it is difficult to accurately diagnose MF. The current approach combines clinic-pathological features in three categories: compatible clinical presentations, compatible pathologic features, and presence of T cell receptor (TCR) gene clonal rearrangements (International Society of Cutaneous Lymphomas ISCL Criteria, 2005) [5]. However, these features are not universally or exclusively present in MF. This is particularly true for early stage MF (eMF), which has clinical, histological, and even molecular (including TCR clonality) resemblance to common benign inflammatory skin diseases [5-8]. Further, eMF skin lesions contain only a small number of malignant cells admixed with a large number of reactive CD4+ T cells. As a result, diagnostic delays in MF are common, which can occasionally exceed a decade.

Another challenge in MF management is to predict long term clinical outcome. While most patients with eMF have a life span similar to that of healthy individuals, 9% of MF patients with limited patch and/or plaque disease, as well as 24% of patients with extensive patches or plaques, will experience disease progression. MF patients with end-stage disease have a high mortality [9, 10]. It is therefore critical to identify MF patients with high risk of disease progression or disease-specific mortality. However, few robust prognostic markers exist for MF, which hinders the accurate prediction of disease prognosis.

In a recent study focusing on gene markers of eMF, we identified 19 genes that were highly enriched in eMF skin biopsies, compared with benign control biopsies. The most enriched gene in eMF biopsies is TOX[11], a critical nuclear factor regulating thymocyte lineage commitment. The aberrant TOX expression in MF was subsequently confirmed in early as well as more advanced disease of MF by in a small patient population (N=15)[12]. These studies raised the possibility that TOX could be used as a marker for MF diagnosis and prognostication.

The aim of the current study is to test this possibility. We examined the expression level of TOX in 113 MF skin biopsies from two independent MF cohorts, including 59 patients for whom long term clinical outcome data, such as disease progression and survival, are available. We found that TOX was aberrantly over-expressed in the majority of MF skin lesions in both cohorts, and that higher TOX expression levels correlated with increased risks of disease progression and disease-specific mortality.

**RESULTS**

**Demographics of study subjects**

A total of 149 individuals contributed skin biopsies, including 113 with MF (summarized in Table 1), 25 with BID, and 11 with healthy skin (HS).

**TOX is ectopically over-expressed in MF skin biopsies**

First, TOX mRNA expression in skin biopsies was assessed in cohort 1 patients, using HS and BID biopsies as the controls. Since the levels of TOX mRNA expression (mean ± standard error of the mean) of BID were not significantly different from those of HS (3.77 ± 0.94
versus 3.15 ± 1.24, *P* = 0.70), they were combined to form the non-MF controls (3.58 ± 0.75). Compared with this, MF cohort 1 had a 17.9 fold increase in TOX gene expression (64.21 ± 18.69, **P** = 0.002) (Figure 1A).

To evaluate if the TOX over-expression in MF skin biopsies is the result of simple increase of CD4+ T cells in MF skin biopsies compared with benign controls, TOX mRNA level was normalized to CD4 mRNA level in the same skin biopsies. In Figure 1B, TOX mRNA to CD4 mRNA ratios were also much higher (8.7 fold) in MF samples, compared with non-MF controls (0.542 ± 0.182 versus 0.062 ± 0.014, *P* = 0.011).

Next, to test if TOX expression varies with tumor burden, TOX mRNA levels were plotted according to lesional morphological types, such as patches, plaques, and tumors. As shown in Figure 1C, skin samples from thicker skin lesions of MF, including plaques and tumors, expressed higher levels of TOX (74.91 ± 29.72 in plaques and 101.3 ± 37.23 in tumors), compared with thinner patches (18.31 ± 4.10, *P* = 0.018). Similarly, TOX/CD4 ratios displayed an increase in thicker lesions (0.348 ± 0.065 in plaques and 1.870 ± 1.145 in tumors), compared with thinner lesions (0.216 ± 0.055 in patches, *P* = 0.088, Figure 1D).

To further confirm the TOX mRNA expression increase in MF skin lesions, an independent MF cohort (cohort 2) was analyzed. As shown in Figure 1A, TOX mRNA in cohort 2 was also much higher than that in non-MF controls (20.64 ± 3.90 versus 3.58 ± 0.75, *P* < 0.0001).

**TOX protein is detected in the CD4+ T cell nuclei in various MF skin lesions and its levels are in parallel with lesional thickness**

To test if TOX protein can be detected in the CD4+ T cells of MF lesions, we performed immunofluorescence (IF) staining on patch, plaque, and tumor MF biopsies, using benign chronic dermatitis (CD) lesions as the controls. In line with the TOX mRNA expression results, although the CD skin biopsies contained numerous CD4+ T cells, few of them had detectable TOX protein expression. In contrast, strong nuclear staining of TOX was detected in the CD4+ T cells of MF lesions. In addition, thicker MF skin lesions (plaque and tumor types) displayed stronger TOX staining than patch lesions (Figure 2).

**High TOX mRNA levels differentiate MF from non-MF skin biopsies**

We next applied receiver operating characteristic (ROC) curve to evaluate if increased TOX mRNA levels

---

**Figure 1:** TOX mRNA levels are increased in MF skin biopsies. (A) TOX mRNA in MF samples compared with BID and HS. ***, *P* = 0.002, ***, **P* < 0.0001. (B) The ratios of TOX mRNA/CD4 mRNA in samples of MF, BID and HS. **, *P* = 0.011. (C) TOX mRNA in different types of MF lesions. *, *P* = 0.018. (D) The ratios of TOX mRNA/CD4 mRNA in different types of MF lesions. Horizontal bars denote the mean and standard error of mean for each sample type analyzed. Two tailed t tests were used for comparison. NS=not significant.
could differentiate MF from non-MF controls. As shown in Figure 3, **TOX** mRNA levels had good discriminatory power for MF, demonstrated by an area under the curve (AUC) value of 0.87 (95% confidence interval (CI) = 0.81-0.94, \( P < 0.0001 \), Figure 3A). When cutoff was set at 2.99, **TOX** mRNA levels had 90.3% sensitivity and 75.0% specificity for MF.

Given that early MF is most difficult to diagnose in the clinical practice, ROC curve was further plotted in stage-I MF (N=73). **TOX** mRNA levels had an AUC of 0.85 (95% CI = 0.77-0.93, \( P < 0.0001 \), Figure 3B) for stage-I MF, with a sensitivity of 86.3% and a specificity of 75.0% at the cutoff of 2.99.

**High **TOX **mRNA levels define a group of MF patients with increased risks of disease progression and disease-specific mortality**

To test if **TOX** expression in MF skin biopsies influences disease progression and disease-specific mortality, we evaluated MF cohort 2, for whom clinical outcome data are available for up to 6 years (median follow up time = 45 months). Kaplan-Meier curves showed that high **TOX** mRNA levels in skin biopsies were associated with much increased likelihood of disease progressing into a higher clinical stage during the follow up period \( (P = 0.003, \text{Figure 4A}) \). Meanwhile, those cases with low **TOX** expression had very low tendency to progress clinically during the follow up period. Through multivariate testing, we found that high **TOX** expression remained significantly associated with disease progression, after adjusting for stage, age, and sex (hazard ratio (HR) = 2.62, \( P = 0.028 \), Table 2).

### Table 2: Multivariate analyses of prognostic factors in MF disease progression and disease-specific mortality

| Prognostic factor | Progression | Disease-specific mortality |
|------------------|-------------|----------------------------|
|                  | HR          | 95.0% CI                   | P   | HR          | 95.0% CI                   | P   |
| **TOX**  | 2.62        | 1.1 to 6.2               | 0.028 | 6.16        | 1.2 to 30.8              | 0.027 |
| Stage           | 3.57        | 1.6 to 7.9               | 0.002 | 5.31        | 1.2 to 23.5              | 0.028 |
| Age             | 1.02        | 1.0 to 1.0               | 0.246 | 1.00        | 1.0 to 1.0               | 0.989 |
| Sex             | 1.02        | 0.5 to 2.3               | 0.960 | 0.55        | 0.1 to 2.1               | 0.386 |

**Figure 2: Ectopic **TOX **protein is detected in CD4+ T lymphocytes in MF skin lesions, but absent in BID.** BID (shown here is CD) and MF skin biopsies were stained with antibodies against **TOX** (red, Alexa Fluor® 594) and CD4 (green, Alexa Fluor® 488). DAPI was used to stain the nuclei of cells. Insets: magnification from representative areas. Bars=20 μm.

**Figure 3: Increased **TOX **mRNA levels differentiate MF from non-MF biopsies.** (A) ROC analysis on the MF biopsies as a whole (N=113) and non-MF skin biopsies (N=36). (B) ROC analysis on the stage-I MF biopsies (N=73) and non-MF skin biopsies (N=36). AUC=Area under the curve.
In addition, high TOX mRNA levels were strongly associated with increased disease-specific mortality ($P = 0.008$, Figure 4B) in MF patients. Multivariate analysis showed that high TOX mRNA expression was an independent predictor of worse survival after adjusting for other variables (HR = 6.16, $P = 0.027$, Table 2), including clinical stage, age and sex.

Collectively, these findings supported that high TOX mRNA levels defined a subset of MF patients with increased risks of disease progression and disease-specific mortality.

DISCUSSION

The earliest evidence of aberrant TOX expression in MF came from our previous transcriptome analysis focusing on eMF, which showed that eMF CD4$^+$ T cells, but not CD4$^+$ T cells in BID, over-expressed TOX at both mRNA level and protein level[11].

TOX encodes a nuclear protein of the high-mobility group (HMG) box superfamily. It contains a DNA-binding domain, which allows it to regulate transcription by modifying local chromatin structure and modulating the formation of multi-protein complexes [13, 14]. In mouse models, TOX was reported to be essential for the proper development of CD4$^+$ thymocyte [13, 15]. Tightly controlled in a stage-specific manner in developing thymocytes, TOX is down-regulated before T cells exit the thymus, and remains suppressed in the peripheral lymphoid tissues [13]. Despite its rapid induction by pre-TCR and TCR signalling in immature thymocytes, TOX is not induced by TCR signalling in mature T cells[13]. Hence its up-regulation in eMF lesional CD4$^+$ T cells is unlikely to be a result of T cell activation. Rather, this unusual observation led us to propose that TOX is a disease marker with diagnostic and/or prognostic value for MF. This study tested and confirmed our hypothesis.

Our findings not only confirmed TOX expression in eMF, but also demonstrated that TOX ectopic expression is a common feature across the entire spectrum of MF. Moreover, high TOX mRNA levels effectively differentiate MF, both as a whole and for stage-I MF alone, from benign inflammatory or healthy skin. Finally, high TOX expression strongly correlated with worse prognosis, including increased disease progression and disease-specific mortality.

MF clinically resembles the far more common, but clinically benign skin inflammatory diseases such as CD, psoriasis, and cutaneous reactions to drugs. Differentiating MF from these benign conditions is difficult, largely due to the lack of well-defined molecular markers in the clinical setting. Although CD2, CD3, CD5, and CD7 deficiency is included in ISCL criteria to define early MF, the loss of CD2, CD3, and/or CD5 in T cells is only 10% sensitive, despite its 100% specificity. CD7 deficiency is about 40% sensitive and 80% specific in general [5]. Therefore better markers with higher sensitivity and specificity are needed. Although a small number of markers have been reported for MF skin biopsies, including loss of CD13, ectopic expression of BLK gene, microRNAs (including miR-155, miR-203 and miR-205) [16-18], BCL7A loss [19, 20], enhanced AHI1 [21], and CD158K/KIR3DL2 in transformed advanced MF [22], few of these markers were tested in multicenter studies, or used in a clinical setting.

Similarly, achieving accurate prognostication of MF patients is challenging clinically. Several molecular markers have been described to be of potential prognostic value. For MF prognosis, persistence of the same CD4$^+$ T cell clone over time in skin biopsies correlated with an aggressive disease course[23]. Loss of BCL7A expression predicted aggressive disease course in patients with early stage CTCL [20]. Specifically for tumor stage MF, presence of chromosomal alterations on 9p21, 8q24, 10q26qter and 1q21-1q22 often indicates a poor prognosis [24, 25]. The clinical utility of these markers needs further evaluation in larger studies.

Figure 4: Higher TOX mRNA levels correlate with worse clinical outcome. (A) Kaplan-Meier plot showing the relationship between TOX mRNA levels and disease progression in MF patients. (B) The relationship between TOX mRNA levels and disease-specific mortality in MF patients.
In light of these, the current multicenter study consisting of patients with diverse ethnic origins demonstrated considerable potential of TOX to improve management for MF. Kaplan-Meier curve analysis demonstrated that MF lesions, including early MF lesions, that contain no or low TOX mRNA expression had little tendency of disease progression or MF-related death. TOX as a marker can identify these low-risk patients for whom conservative management perhaps would be adequate rather than being subjected to more toxic treatments, such as topical nitrogen mustard, or Carmustine. In contrast, MF patients with high levels of TOX expression may benefit from early and more aggressive treatment to prevent disease progression and to reduce MF-related mortality.

Despite the predictive potential of TOX for MF demonstrated in this study, caution needs to be taken while interpreting its clinical usefulness. Due to the rarity of this disease, only a moderate sample size of 113 can be reached from three study centres from three countries. Additional confirmation in other centres is warranted to uncover the true clinical relevance of TOX for MF diagnosis and prognostication.

Notwithstanding this limitation, the multicenter nature of our study underscores the consistency of TOX up-regulation in patients from diverse ethnic and geographic backgrounds. Furthermore, TOX as a marker is highly robust and user-friendly, since it can be detected by a number of routine dermatopathologic tools, such as qPCR, IF, and immunohistochemistry staining.

Given TOX’s critical regulatory role in CD4+ T cell development and its aberrant over-expression in the majority of MF skin lesion, it is highly likely that TOX activation plays a pathogenic role in MF. Further studies are therefore warranted to evaluate if TOX ectopic expression contributes to the development of MF. If proven to be true, TOX may emerge as a novel therapeutic target for MF in the future.

In summary, TOX ectopic expression is readily and frequently detected in the malignant CD4+ T cells in MF skin biopsies, including the most challenging eMF. Moreover, increased TOX expression levels define a group of MF patients with increased risk of disease progression and disease-specific mortality. Therefore, characterization of TOX expression status of MF patients may be valuable not only for diagnostic confirmation but also for guiding MF management in the future.

MATERIALS AND METHODS

Study subject recruitment

With approval from institutional clinical ethics board, this study was conducted according to Declaration of Helsinki Principles, and all participants has provided their written informed consent. Two MF cohorts were examined in this study (Table 1). Cohort 1 was prospectively recruited from Vancouver Canada and Beijing China from 2008-2012, including 54 subjects with MF (cohort 1, N=54) recruited from the Skin Lymphoma Clinics of British Columbia Cancer Agency (BCCA, Vancouver, Canada, N=26), and Peking University First Hospital (Beijing, China, N=28). In addition, 25 individuals with BID (9 psoriasis, 13 CD, and 3 pityriasis rubra pilaris) and 11 volunteers with HS were recruited from the outpatient dermatology clinic of University of British Columbia (UBC, Vancouver, Canada). Cohort 2 of MF patients (cohort 2, N=59) was collected from Harvard University with institutional approval and had been well-characterized previously[20, 21, 26, 27]. The diagnosis and clinical staging were established according to the diagnostic criteria of CTCL[7].

Preparation of clinical samples and analyses of TOX mRNA and protein

Obtaining skin biopsies, RNA extraction, and quantitative reverse transcription-polymerase chain reaction (qPCR) were performed using protocols that were previously described[11]. Beta actin (ACTB) mRNA was used as the internal control. The results were expressed as copies of TOX mRNA per 1000 copies of ACTB mRNA. In addition, TOX protein in MF and control skin biopsies was analyzed by IF using specific antibodies against TOX (rabbit anti-TOX polyclonal antibody, Sigma-Aldrich, St. Louis, MO) and human CD4 protein (mouse monoclonal anti-human CD4 antibody, Dako, Denmark), as previously described[11]. Cell nuclei were counterstained with 4’-6-diamidino-2-phenylindole (DAPI). The slides were visualized under a Zeiss AxioVert 200M inverted fluorescence microscope (Carl Zeiss AG, Jena, Germany). Representative sections of the micrographs were obtained and processed with the Zeiss AxioVision 4.8 image acquisition and processing software (Carl Zeiss AG). Brightness and contrast were adjusted consistently across all images.

Statistical analyses

SPSS 14 (Chicago, IL), GraphPad Prism 5.00 (San Diego, CA), and X-Tile (New Haven, CT) were used for statistical analyses. P values < 0.05 were considered to be statistically significant.

Continuous variables were compared by two-tailed t tests. ROC method was used to analyze the sensitivity and specificity of TOX mRNA levels to differentiate MF from BID or HS skin biopsies.

X-Tile software, a marker cutpoint analysis tool developed by Camp RL et al[28], was applied to determine the optimal cut-point for TOX expression.
level as 8.2 (Miller-Siegmund corrected P value=0.027). In our analysis of disease progression and disease-specific mortality, TOX high and TOX low groups were defined by the expression levels higher or lower than 8.2, respectively.

MF patients were considered to experience disease progression when disease progressed to more advanced clinical stages (i.e. beyond stage I) and/or death due to MF during the follow up period[20]. Individuals with multiple progressions (n > 1) were counted as n progression events. Survival time was defined as the duration from the date of sample collection to death.

Disease progression and disease-specific mortality rates were assessed using the Kaplan-Meier curves. Univariate analysis using the log-rank test was first used to detect any association between TOX mRNA level and disease progression and disease-specific mortality. Multivariate analyses using COX proportional hazards regression were then used to evaluate the following prognostic factors: stage at diagnosis (stages I and II versus stages III and IV), TOX mRNA level, age, and sex.

CONFLICT OF INTEREST

The authors disclose no potential conflicts of interest.

ACKNOWLEDGMENTS

We thank Lixin Xia, Richard Yu, and Guohong Zhang for their excellent technical assistance. This work was supported by grants from Canadian Institutes of Health Research (to Zhou Y), Canadian Dermatology Foundation (to Zhou Y, Litvinov IV, and Sasseville D), Fonds de la recherche en santé du Québec (FRSQ) (to Zhou Y, Litvinov IV, and Sasseville D), National Natural Science Foundation of China (81072233 and 81201228) (to Wang Y, and Tu P). Huang Y is a Canadian Institutes of Health Research-Skin Research Training Center Scholar, and a Vanier Canada Graduate Scholar.

REFERENCES

1. Criscione VD and Weinstock MA. Incidence of cutaneous T-cell lymphoma in the United States, 1973-2002. Arch Dermatol. 2007; 143(7):854-859.
2. Girardi M, Heald PW and Wilson LD. The pathogenesis of mycosis fungoides. N Engl J Med. 2004; 350(19):1978-1988.
3. Porcu P and Wong HK. We should have a dream: unlocking the workings of the genome in cutaneous T-cell lymphomas. Clin Lymphoma Myeloma. 2009; 9(6):409-411.
4. Mao X, Orchard G, Mitchell TJ, Oyama N, Russell-Jones R, Vermeer MH, Willemze R, van Doorn R, Tensen CP, Young BD and Whittaker SJ. A genomic and expression study of AP-1 in primary cutaneous T-cell lymphoma: evidence for dysregulated expression of JUNB and JUND in MF and SS. J Cutan Pathol. 2008; 35(10):899-910.
5. Pimpinelli N, Olsen EA, Santucci M, Vonderheide E, Haeffner AC, Stevens S, Burg G, Cerroni L, Dreno B, Glusac E, Guitar J, Heald PW, Kempf W, Knobler R, Lessin S, Sander C, et al. Defining early mycosis fungoides. J Am Acad Dermatol. 2005; 53(6):1053-1063.
6. Glusac EJ. Criterion by criterion, mycosis fungoides. Am J Dermatopathol. 2003; 25(3):264-269.
7. Olsen E, Vonderheide E, Pimpinelli N, Willemze R, Kim Y, Knobler R, Zackheim H, Duvic M, Estrach T, Lambsg, Wood G, Dummer R, Ranki A, Burg G, Heald P, Pittelkow M, et al. Revisions to the staging and classification of mycosis fungoides and Sezary syndrome: a proposal of the International Society for Cutaneous Lymphomas (ISCL) and the cutaneous lymphoma task force of the European Organization of Research and Treatment of Cancer (EORTC). Blood. 2007; 110(6):1713-1722.
8. Keehn CA, Belongie IP, Shistik G, Fenske NA and Glass LF. The diagnosis, staging, and treatment options for mycosis fungoides. Cancer Control. 2007; 14(2):102-111.
9. Kim YH, Chow S, Varghese A and Hoppe RT. Clinical characteristics and long-term outcome of patients with generalized patch and/or plaque (T2) mycosis fungoides. Arch Dermatol. 1999; 135(1):26-32.
10. Kim YH, Bishop K, Varghese A and Hoppe RT. Prognostic factors in erythrodermic mycosis fungoides and the Sezary syndrome. Arch Dermatol. 1995; 131(9):1003-1008.
11. Zhang Y, Wang Y, Yu R, Huang Y, Su M, Xiao C, Martinka M, Dutz JP, Zhang X, Zheng Z and Zhou Y. Molecular markers of early-stage mycosis fungoides. J Invest Dermatol. 2012; 132(6):1698-1706.
12. McGirt LY, Adams CM, Baerenwald DA, Zwerner JP, Zic JA and Eischen CM. miR-223 Regulates Cell Growth and Targets Proto-Oncogenes in Mycosis Fungoides/Cutaneous T-Cell Lymphoma. J Invest Dermatol. 2014; 134(4):1101-1107.
13. Wilkinson B, Chen JY, Han P, Rufner KM, Goularte OD and Kaye J. TOX: an HMG box protein implicated in the regulation of thymocyte selection. Nat Immunol. 2002; 3(3):272-280.
14. Aliahmad P, Seksenyana and Kaye J. TOX in the immune system. Curr Opin Immunol. 2012; 24(2):173-177.
15. Aliahmad P and Kaye J. Development of all CD4 T lineages requires nuclear factor TOX. J Exp Med. 2008; 205(1):245-256.
16. Bernier C, Nguyen JM, Quereux G, Renault JJ, Bureau B and Dreno B. CD13 and TCR clone: markers of early mycosis fungoides. Acta Derm Venereol. 2007; 87(2):155-159.
17. Krejsgaard T, Vetter-Kaucek CS, Woettmann A, Kneitz H, Erikse K, Lovato P, Zhang Q, Wasik MA, Geisler C,
Ralfkiaer E, Becker JC and Odum N. Ectopic expression of B-lymphoid kinase in cutaneous T-cell lymphoma. Blood. 2009; 113(23):5896-5904.

18. Ralfkiaer U, Hagedorn PH, Bangsgaard N, Lovendorf MB, Ahler CB, Svensson L, Kopp KL, Vennegaard MT, Lauenborg B, Zibert JR, Krejsgaard T, Bonefeld CM, Sokilde R, Gjerdrum LM, Labuda T, Mathiesen AM, et al. Diagnostic microRNA profiling in cutaneous T-cell lymphoma (CTCL). Blood. 2011; 118(22):5891-5900.

19. Carbone A, Bernardini L, Valenzano F, Bottilo I, De Simone C, Capizzi R, Capalbo A, Romano F, Novelli A, Dallapiccola B and Amerio P. Array-based comparative genomic hybridization in early-stage mycosis fungoides: recurrent deletion of tumor suppressor genes BCL7A, SMAC/DIABLO, and RHOF. Genes Chromosomes Cancer. 2008; 47(12):1067-1075.

20. Litvinov IV, Zhou Y, Kupper TS and Sasseville D. Loss of BCL7A expression correlates with poor disease prognosis in patients with early-stage cutaneous T-cell lymphoma. Leuk Lymphoma. 2013; 54(3):653-654.

21. Litvinov IV, Kupper TS and Sasseville D. The role of AHI1 and CDKN1C in cutaneous T-cell lymphoma progression. Exp Dermatol. 2012; 21(12):964-966.

22. Ortonne N, Le Gouvello S, Tabak R, Marie-Cardine A, Setiao J, Berrehar F, Nghe-Tang A, Martin N, Bagot M and Bensussan A. CD158k/KIR3DL2 and NKp46 are frequently expressed in transformed mycosis fungoides. Exp Dermatol. 2012; 21(6):461-463.

23. Vega F, Luthra R, Medeiros LJ, Dunmire V, Lee SJ, Duvic M and Jones D. Clonal heterogeneity in mycosis fungoides and its relationship to clinical course. Blood. 2002; 100(9):3369-3373.

24. van Doorn R, van Kester MS, Dijkman R, Vermeer MH, Mulder AA, Szuha K, Knijnenburg J, Boer JM, Willemze R and Tensen CP. Oncogenomic analysis of mycosis fungoides reveals major differences with Sezary syndrome. Blood. 2009; 113(1):127-136.

25. Salgado R, Servitje O, Gallardo F, Vermeer MH, Ortiz-Romero PL, Karpova MB, Zipser MC, Muniesa C, Garcia-Muret MP, Estrach T, Salido M, Sanchez-Schmidt J, Herrera M, Romagosa V, Suela J, Ferreira BI, et al. Oligonucleotide array-CGH identifies genomic subgroups and prognostic markers for tumor stage mycosis fungoides. J Invest Dermatol. 2010; 130(4):1126-1135.

26. Shin J, Monti S, Aires DJ, Duvic M, Golub T, Jones DA and Kupper TS. Lesional gene expression profiling in cutaneous T-cell lymphoma reveals natural clusters associated with disease outcome. Blood. 2007; 110(8):3015-3027.

27. Litvinov IV, Jones DA, Sasseville D and Kupper TS. Transcriptional profiles predict disease outcome in patients with cutaneous T-cell lymphoma. Clin Cancer Res. 2010; 16(7):2106-2114.

28. Camp RL, Dolled-Filhart M and Rimm DL. X-tile: a new bio-informatics tool for biomarker assessment and outcome-based cut-point optimization. Clin Cancer Res. 2004; 10(21):7252-7259.