"Candidatus Rickettsia kellyi," India

Jean-Marc Rolain,* Elizabeth Mathai,† Hubert Lepidi,* Hosaagrahara R. Somashekar,† Leni G. Mathew,† John A.J. Prakash,† and Didier Raoult*

We report the first laboratory-confirmed human infection due to a new rickettsial genotype in India, "Candidatus Rickettsia kellyi," in a 1-year-old boy with fever and maculopapular rash. The diagnosis was made by serologic testing, polymerase chain reaction detection, and immunohistochemical testing of the organism from a skin biopsy specimen.

Human rickettsioses are infections of emerging importance in India, where increasing numbers of cases among residents and travelers have been reported recently (1,2). Nevertheless, these diseases are not well described in the literature and, to date, only serologic evidence of rickettsial infections has been reported, including murine typhus, scrub typhus, and unidentified spotted fever group (SFG) rickettsiosis (1,3,4). Moreover, the results of serologic testing are presumptive and should be interpreted with caution. SFG rickettsiosis is seldom diagnosed in India, probably because of a low index of suspicion and a relative lack of diagnostic facilities. Specific diagnostic methods are needed to identify unexpected SFG agents either by polymerase chain reaction (PCR) or by culture (5).

Few reports of rickettsioses in children from southern India have been reported. Here we report the case of a 1-year-old boy with a new SFG rickettsiosis characterized by a maculopapular rash on the palms and soles. The diagnosis was confirmed by serologic testing, molecular detection, and immunohistochemical testing of a skin biopsy specimen. We propose the name "Candidatus Rickettsia kellyi" in honor of Professor Patrick Kelly, who has greatly contributed to the current knowledge of rickettsiae throughout the world.

The Study

A 1-year-old boy from Thiruppathur, Tamil Nadu, India, was brought for treatment; he had exhibited fever for 10 days and a maculopapular rash on the face and chest that had spread rapidly to the trunk and limbs. The rash was also on his palms and soles. No tick bite was noted. A skin biopsy was taken from a maculopapular lesion. Laboratory tests showed a leukocyte count of 15,300/mm³, hemoglobin level of 9.2 g/dL, and normal platelet count, and normal cerebrospinal fluid was seen by lumbar puncture. The patient was given doxycycline syrup and cefotaxime because the diagnosis was not definitive and the boy was very ill. The boy responded dramatically and eventually recovered. Results of conventional culture of cerebrospinal fluid, skin biopsy specimens, and blood culture were negative.

With Weil-Felix agglutination assay, the serum sample taken at admission was weakly positive with OX-2 antigen (titer 40) and negative for OXK and OX-19 antigens, giving presumptive evidence of a rickettsial infection. DNA was extracted from the skin biopsy specimen and used as a template in 2 previously described PCR assays that targeted a portion of the rickettsial ompA gene as well as a portion of the rickettsial gltA gene, ompB gene, and sca4 genes (6,7). Amplification products of the expected size were obtained from this extract but not from any concurrently processed control materials. The most closely related rickettsial species was found to be R. honei with pairwise nucleotide sequence homologies of 92.3% for ompA, 99.2% for gltA, 94.6% for ompB, and 99.1% for sca4. Histopathologic testing of the skin biopsy specimen showed a leukoclastic vasculitis, and immunohistochemical testing by using a rabbit polyclonal antibody directed against SFG rickettsiae showed positive result (Figure 1).

For specific microimmunofluorescence assay, a panel of 13 rickettsial antigens, including SFG rickettsiae (R. conorii subsp. indica, R. japonica, R. honei, R. helvetica, R. slovaca, AT1 Rickettsia, R. felis, “R. heilongjiangensis”), typhus group rickettsiae (R. typhi), Orientia tsutsugamushi (strains Gilliam, Kato, Karp, and Kawasaki),
Anaplasma phagocytophilum, Ehrlichia chaffeensis, and Coxiella burnetii, was used as previously described (8,9). The serum sample from the patient at the acute stage of illness was weakly positive for all SFG rickettsia with a negative immunoglobulin G (IgG) titer and a IgM titer from 64 to 128 according to the species tested. Serum was also positive by Western blot analysis as previously reported (8), but the species remained undetermined.

Conclusions

Clinical and laboratory data for this patient suggest that he had an SFG rickettsial infection that was confirmed by 3 different testing methods: serologic, immunohistochemical, and molecular based. Although no tick bite and no eschar were noted, the infection could have been acquired from any of a wide variety of arthropods in this area. In India, serologic evidence of human SFG rickettsioses has been found (1,3,10), but the epidemiology of etiologic agents is deduced only by serologic testing performed by using known rickettsial antigens. Our case, to the best of our knowledge, is the first human SFG rickettsiosis case diagnosed in India that was laboratory confirmed by using specific and direct detection of a rickettsial strain. Moreover, according to genetic guidelines for the classification of rickettsial isolates (6), our rickettsial strain found in the skin biopsy specimen belongs to a new species. Unfortunately, because the skin biopsy specimen was stored in alcohol, culture and complete phenotypic description of this isolate were not possible. The most closely-related rickettsial strain, according to genetic guidelines, was R. honeï as shown in the phylogenetic tree (Figure 2). R. honeï is the etiologic agent of Flinders Island (Australia) spotted fever, which was isolated from the blood of 2 patients in 1993 (11,12). The main reservoir of R. honeï was later determined to be Aponomma hydrosauri, a reptile tick (13). The pathogenicity of the original isolate of R. honeï (Thai tick typhus strain TT-118) for humans has not yet been confirmed, but it is possibly responsible for SFG human rickettsiosis in Thailand (14). Thus, genotyping of these strains is needed to better understand the epidemiology of SFG rickettsiosis in Asia. Further studies are needed to isolate and establish this new pathogenic SFG rickettsial strain from humans to confirm our case report. Moreover, tick species prevalent in this area of South Asia should be tested to find the rickettsial reservoir and increase understanding of the epidemiology of this rickettsial infection. New pathogens remain to be discovered in India, and new rickettsial diseases represent a challenge.

The partial ompA gene sequence of “Candidatus Rickettsia kellyi” has been deposited in the GenBank data library under accession no. DQ080005. GenBank accession nos. were TTU59726 for gltA, AF123724 for ompB, and AF163004 for sca4.

Dr Rolain is a researcher at the Unité des Rickettsies, the national reference center for rickettsiosis and WHO collaborative center in Marseille, France. The laboratory studies emerging and reemerging bacteria and arthropodborne diseases.

References

1. Murali N, Pillai S, Cherian T, Raghupathy P, Padmini V, Mathai E. Rickettsial infections in South India—how to spot the spotted fever. Indian Pediatr. 2001;38:1393–6.
2. Rahman A, Tegnell A, Vene S, Giesecke J. Rickettsioses in Swedish travellers, 1997–2001. Scand J Infect Dis. 2003;35:247–50.
3. Mathai E, LLOYD G, Cherian T, Abraham OC, Cherian AM. Serological evidence for the continued presence of human rickettsioses in southern India. Ann Trop Med Parasitol. 2001;95:395–8.
4. Mathai E, Rolain JM, Verghese GM, Abraham OC, Mathai M, et al. Outbreak of scrub typhus in southern India during the cooler months. Ann N Y Acad Sci. 2003;990:359–64.
5. Raoult D. A new rickettsial disease in the United States. Clin Infect Dis. 2004;38:812–3.
6. Fournier PE, Dumler JS, Greub G, Zhang J, Wu Y, Raoult D. Gene sequence-based criteria for identification of new rickettsia isolates and description of Rickettsia heliogiangensis sp. nov. J Clin Microbiol. 2003;41:5456–65.
7. Fournier PE, Zhu Y, Ogata H, Raoult D. Use of highly variable intergenic spacer sequences for multiplex typing of Rickettsia conorii strains. J Clin Microbiol. 2004;42:5757–66.
8. Parola P, Miller RS, McDaniel P, Telford SR III, Rolain JM, Wongsrichanalai C, et al. Emerging rickettsioses of the Thai-Myanmar border. Emerg Infect Dis. 2003;9:592–5.
9. Zhu Y, Fournier PE, Eremeeva M, Raoult D. Proposal to create sub-species of Rickettsia conorii based on multi-locus sequence typing and an emended description of Rickettsia conorii. BMC Microbiol. 2005;5:11.
10. Sundhindra BK, Vijayakumar S, Katty KA, Tholpadi SR, Rajan RS, Mathai E, et al. Rickettsial spotted fever in Kerala. Natl Med J India. 2004;17:51–2.
11. Graves SR, Stewart L, Stenos J, Stewart RS, Schmidt E, Hudson S, et al. Spotted fever group rickettsial infection in South-Eastern Australia: isolation of rickettsiae. Comp Immun Microbiol Infect Dis. 1993;16:223–33.

Figure 2. Phylogenetic tree of rickettsiae, including “Candidatus Rickettsia kellyi,” obtained by comparison of partial sequences of ompA with the parsimony method.
12. Stenos J, Roux V, Walker D, Raoult D. *Rickettsia honei* sp. nov., the aetiological agent of Flinders Island spotted fever in Australia. Int J Syst Bacteriol. 1998;48:1399–1404.

13. Stenos J, Graves S, Popov VL, Walker DH. *Aponomma hydrosauri*, the reptile-associated tick reservoir of *Rickettsia honei* on Flinders Island, Australia. Am J Trop Med Hyg. 2003;69:314–7.

14. Graves S, Stenos J. *Rickettsia honei*: a spotted fever group rickettsia on three continents. Ann N Y Acad Sci. 2003;990:62–6.

Address correspondence: Didier Raoult, Unité des Rickettsies, Faculté de Médecine, 27, Boulevard Jean Moulin, 13385 Marseille CEDEX 5, France; fax: 33-491-38-77-72; email: Didier.Raoult@medecine.univ-mrs.fr