Novel brain PET imaging agents: Strategies for imaging neuroinflammation in Alzheimer’s disease and mild cognitive impairment

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Alzheimer’s disease (AD) is a devastating neurodegenerative disease with a concealed onset and continuous deterioration. Mild cognitive impairment (MCI) is the prodromal stage of AD. Molecule-based imaging with positron emission tomography (PET) is critical in tracking pathophysiological changes among AD and MCI patients. PET with novel targets is a promising approach for diagnostic imaging, particularly in AD patients. Our present review overviews the current status and applications of in vivo molecular imaging toward neuroinflammation. Although radiotracers can remarkably diagnose AD and MCI patients, a variety of limitations prevent the recommendation of a single technique. Recent studies examining neuroinflammation PET imaging suggest an alternative approach to evaluate disease progression. This review concludes that PET imaging towards neuroinflammation is considered a promising approach to deciphering the enigma of the pathophysiological process of AD and MCI.

KEYWORDS
Alzheimer’s disease, positron emission tomography, neuroinflammation, mild cognitive impairment, review

1 Introduction

Alzheimer’s disease (AD) is a neurodegenerative disease with a concealed onset and continuous progress. It is a severe risk factor that is threatening the health and life of elderly individuals (1, 2). AD is known to accumulate two different insoluble protein aggregates. During the prodromal phase of AD, also called mild cognitive impairment (MCI), patients have a receding performance of cognitive domains (3). The neuropathological biomarkers for AD and MCI are β-amyloid (Aβ) plaques and intracellular tau neurofibrillary tangles (NFTs) (4, 5). Aβ cascade hypothesis is one of the dominant hypotheses of AD pathogenesis (6). It
refers to the Aβ patch as the initial factor causing excessive phosphorylation of tau protein, microglial activation, neurotransmitter disorders, oxidative stress, and neuro-pathological changes (7). However, AD pathological changes may have been maintained for decades before symptoms occur. When the exact diagnosis has been made with the above approaches, patients may present with irreversible nervous system damages. Therefore, developing an early, non-invasive, and accurate diagnostic method could be a research hotspot.

The central nervous system (CNS) degeneration and disease neuropathology predate AD and MCI (8). One dominant theory indicates that the excess generation or disordered clearance of Aβ is the dominant event that initiates AD pathogenesis. However, about 30% of healthy individuals with Aβ deposition are without clinical symptoms. Nevertheless, most clinical studies revealed that high Aβ levels are associated with the severity of cognitive impairment. The hyperphosphorylated tau-based NFT pathology is positively associated with the severity of cognitive symptoms (9). However, anti-amyloid interventions have been reported to depict limited effects in clinical trials. Because of poor understanding of the pathogenesis, the clinical diagnosis of AD mainly depends on detailed medical history, imaging, and neuropsychological scale assessment. Therefore, besides the traditional biomarkers of AD pathology, more targets require further investigation in exploring AD.

Positron emission tomography (PET) is a neuroimaging approach to evaluate the molecular processes in the brain, effective and accurate for diagnostic purposes, clinical strategy planning, and assessing disease progression (10, 11). PET radioligands can bind to responding targets, including receptors, transporters, or enzymes. Furthermore, tracer binding or uptake degrees can serve as a quantitative neuropathology approach. With the advance of PET technology, targeted molecular probes toward Aβ deposition, tau protein, and neuroinflammation have been extensively developed and utilized in the clinical management of AD. PET biomarkers have also been recommended to ameliorate diagnostic accuracy for AD and MCI, including cerebral metabolism or Aβ deposition on amyloid PET (12). However, the rate of cerebral metabolism cannot explicitly elucidate potential neuropathology. Although measurement of amyloid and neuroinflammation through PET tracers provides excellent insight into the underlying process of AD and MCI, the latter has not been implemented in clinical practice. The present review describes the associated trials of PET imaging targeting neuroinflammation within AD and MCI.

2 AD and neuroinflammation

Currently, neuroinflammation in AD has been well illustrated (13). In the initial phase of neuroinflammation, immune cells aim to ameliorate neuronal injury. However, abnormal inflammatory responses involving prolonged microglial activation presented adverse effects and exacerbated neurodegeneration (14). According to histopathological studies, activated microglia localize to Aβ plaques and NFTs, which attached significant attention to the role of neuroinflammation in the AD process (15). Microglia are dominant immune cells in the central nervous system (CNS), essential in maintaining hemostasis and secreting inflammatory factors (16, 17). Under the hemostatic condition, microglia present M1-like status, while under pathological conditions, they convert to an M2 state (18). M1 microglia are pro-inflammatory and produce reactive oxygen species (ROS) to remove the foreign substance and trigger neuroinflammation, while M2 microglia present an anti-inflammatory function to protect the neurons. Additionally, reactive astrocyte is also a critical member of neuroinflammation, which precipitates both Aβ and tau and is closely linked to microgliosis (19, 20). Astrocytes can be classified into A1 and A2 subtypes based on their phenotype and genetic expression profiles (21). A1 astrocyte secretes and produces various inflammatory factors and neurotoxins, whereas A2 astrocyte produces neurotrophic substances (22, 23).

Neuroinflammation is considered to have an essential role in AD. It has been demonstrated that the persistent accumulation of Aβ levels in AD patients leads to activated neuroinflammation and elevated ROS, which induces cell death through apoptosis (7). Moreover, neuroinflammation initiated by infection has been revealed to exacerbate the tau pathological process in transgenic rodent AD models. Because elderly individuals are more prone to infections, the elevated Aβ burden with activated neuroinflammatory insult may accelerate the progression of neurodegeneration in patients at risk for AD (24). According to the above discussion, it is urgent to make international efforts to develop novel radiotracers for imaging neuroinflammation in AD patients with PET. Therefore, the following sections will briefly review the different strategies within advanced PET radiotracers targeting the potential molecules in the neuroinflammation process.

3 Application of PET imaging agents for neuroinflammation

The AD-associated neuroinflammatory biomarkers are divided into (1) enzymes or signal molecules, including 18-kDa transporters (TSPO), monoamine oxidase B (MAO-B), imidazoline binding sites I2 (I1BS), epoxidation enzyme, and arachidonic acid (AA); (2) G protein-coupled receptors, including purine P2X7 and P2Y receptors, and type 2 cannabinoid receptors (CB2R). Therefore, the corresponding PET tracers are developed and will be summarized in the following section.
### 3.1 TSPO PET radiotracers

It is reported that TSPO presents a variety of cellular functions, including cholesterol transport, inflammatory responses, and hormone synthesis. However, its exact role in the brain immune reaction is not fully understood. Under the hemostatic condition, the expression of TSPO maintains a low level in microglia within CNS, while after neuroinflammation, abnormal activation of microglia is associated with a high level of TSPO. Furthermore, upregulation of TSPO and activation of microglia colocalize spatially after a neurotoxic intervention was established through immunohistochemistry staining, suggesting that TSPO can detect activated microglia and can be a novel approach to measure neuroinflammation. Therefore, combining imaging agents with TSPO can serve as microglia activation and neuroinflammation biomarkers. $^{11}$C-PK11195 presents a high affinity to TSPO, the first classical probe used for PET neuroinflammation imaging. Accumulative evidence of PET images revealed that the region of the cingulate, temporo-parietal cortex, and amygdala in patients with AD presented a high level of $^{11}$C-PK11195 absorption compared to healthy individuals. However, the low sensitivity, low bioavailability, high rate of nonspecific binding, and short half-life defects limited the application of $^{11}$C-PK11195 in patients with AD (25–27). Therefore, the second- and third-generation imaging agents, including the $^{11}$C-DPA-713, $^{18}$F-DPA-714, $^{18}$F-GE-180, and $^{11}$C-ER176, exert higher affinity, and brain uptake rate can detect TSPO in neuroinflammation in low expression.

#### 3.1.1 First generation of TSPO PET radiotracers: $^{11}$C-PK11195

The dominant first generation of TSPO PET radiotracers was the $^{11}$C-PK11195, which is accompanied by either the racemic mixture or the active R-enantiomer. It has been considered the most widely used radiotracer for PET imaging in brain tissue. However, due to the intrinsic properties of the compound and complexity of carbon-11 radiolabeling within a short period of 20 min, the development of this technique for exploring neuroinflammation was impeded. Due to the poor blood–brain barrier (BBB) penetration and low brain uptake, $^{11}$C-PK11195 presents a poor signal-to-noise ratio. Moreover, $^{11}$C-PK11195 has various shortcomings, including low bioavailability and nonspecific binding, which limit its ability to determine subtle changes in TSPO expression in brain tissues (25, 27, 28). Therefore, the above difficulties challenge its application in clinical management and require further development of novel TSPO PET radiotracers.

#### 3.1.2 Second generation of TSPO PET radiotracers

A wide range of second-generation TSPO PET radiotracers have emerged, which present a higher affinity to TSPO and better characteristics. Several radiotracers have already been widely used in patients, especially in AD. According to a previous study, $^{11}$C-PBR28 presents a higher specific signal towards microglial activity than $^{11}$C-PK11195 (29). Additionally, $^{11}$C-DPA-713 showed better sensitivity in the healthy brain than $^{11}$C-PK11195 in measuring increased TSPO expression in the brain (30). Moreover, $^{11}$C-DPA-713 determined increased TSPO density in widespread brain regions in AD patients than $^{11}$C-PK11195 (31). The evaluations of TSPO radioligands, including $^{11}$C-DPA-713, $^{18}$F-DPA-714, and $^{11}$C-PK11195, have been compared in acute neuroinflammation rat models (32). The results indicated that $^{18}$F-DPA-714 had the highest ipsilateral-to-contralateral uptake ratio and had a better binding potential than $^{11}$C-DPA-713 and $^{11}$C-PK11195 (31). Moreover, the ligand $^{11}$C-DAA1106 has demonstrated a higher affinity than $^{11}$C-PK11195 to activate microglia in various neurological disorders (33, 34). The development of novel molecules with higher affinity, greater bioavailability, and the possibility of radiolabel with $^{18}$F facilitates the easier development of inflammation imaging. Currently, a couple of radioligands labeled with $^{18}$F have also been established. Among these, $^{18}$F-FEDAA1106 presents a higher affinity to TSPO than DAA1106. Accumulated evidence has revealed the higher brain uptake of $^{18}$F-FEDAA1106 than $^{11}$C-PK11195 or $^{11}$C-DAA1106 (35). Moreover, $^{18}$F-FEMPA could be a suitable PET radiotracer for TSPO (36). It has been described that $^{18}$F-FEPPA is associated with higher affinity towards TSPO, higher brain penetration, and better pharmacokinetics than the first generation of TSPO PET radiotracers (37).

#### 3.1.3 Third generation of TSPO PET radiotracers

A wide range of third-generation TSPO tracers, including $^{18}$F-GE-180 (R, S)-$^{18}$F-GE-387, $^{11}$C-ER176, $^{11}$C-CB184, $^{11}$C-CB190, $^{11}$C-N-MPB, and $^{18}$F-LW223, have been well established. A couple of clinical trials have compared the binding properties, brain uptake, and performance of TSPO radiotracers. The measurement of microglial activation through $^{18}$F-GE-180 was more sensitive than that by $^{18}$F-PBR06. A wide range of studies have reported on the value of $^{18}$F-GE-180 in assessing microglial activity in different rodent models. Furthermore, rising levels of $^{18}$F-GE-180 uptake indicate elevated microglial activation in patients with AD, semantic dementia, and MCI. However, another study on mouse stroke models suggested that $^{11}$C-DPA-713 PET presented higher accuracy and sensitivity on microglial activation measurement than $^{18}$F-GE-180. Moreover, a more favorable brain entrance property of $^{11}$C-PBR28 has also been reported compared to $^{18}$F-GE-180. $^{11}$C-ER176 has been revealed with a higher binding affinity than $^{11}$C-PK11195, $^{11}$C-PBR28, and $^{11}$C-DPA-71. A clinical trial of $^{11}$C-ER176 PET for accessing microglia activation in AD and MCI patients is still ongoing (NCT03744312).
Based on the evidence, the dominant concern for TSPO ligands is the cellular location of the signal. The different binding sites on glial and vascular TSPO have been reported for several TSPO ligands. The high-glial-TSPO-selectivity and polymorphism-sensitive ligand $^{18}$F-FEBMP has been developed. It presented a higher contrast to neuroinflammation than $^{11}$C-PK11195 in the PS19 tauopathy mouse model. Further studies assessing the binding selectivity to TSPO polymorphism among different generations of TSPO radiotracers are highly required.

3.2 TSPO PET imaging of neuroinflammation in AD

The following section will summarize the clinical effects secured from the clinical trials of PET imaging toward neuroinflammation among AD and MCI patients. An in-depth understanding of the underlying mechanisms of AD neuroinflammation will lead to novel therapeutic approaches to monitor disease progression using TSPO PET imaging.

3.2.1 First generation of TSPO PET imaging in AD

The radiotracer has been examined in accumulative studies involving AD and MCI patients (Table 1). The first clinical trial focusing on $^{11}$C-PK11195 failed to detect TSPO binding related to microglial activation in patients with mild to moderate dementia (51). Another AD study also reported a similar result, which suggested that microglial activation presented in later stages of AD or $^{11}$C-PK11195 is insensitive in mild to moderate AD (40). A recent study reveals only a small cluster of significantly elevated $^{11}$C-PK11195 binding in occipital lobes in AD dementia patients without any difference between clinically stable prodromal AD patients and those who progressed to dementia (43). The rising $^{11}$C-PK11195 brain uptake among AD patients was observed in two studies: (1) higher uptake was found in the frontal and right mesotemporal regions using SPECT (52); (2) higher uptake was seen in the frontal, temporal, occipital, and striatum using PET (39). Additionally, elevated regional $^{11}$C-PK11195 binding is observed in the entorhinal, temporoparietal, and cingulate cortex in patients having mild and early AD. The results were consistent with another study showing increased $^{11}$C-PK11195 signals (38, 53).

Moreover, two other longitudinal studies provided the course of neuroinflammation in AD via $^{11}$C-PK11195. The first study revealed an increase in radiotracer signals in AD patients (44). On the contrary, another longitudinal study illustrated the evolution of $^{11}$C-PK11195 in eight MCI patients, four of whom revealed negative amyloid imaging. In contrast, a longitudinal reduction of microglial activation was observed in this population (47). The small sample sizes, different methods, and first-generation TSPO radiotracer limits could elaborate on these contradictory results.

3.2.2 Second generation of TSPO PET imaging in AD

The shortcomings of the second generation of TSPO radiotracers were improved with the development of advanced-generation TSPO ligands with an enhanced signal-to-noise ratio and higher binding affinity compared to $^{11}$C-PK11195 (54). Several studies assessed neuroinflammation with the measurement of radiotracer retention in AD patients through these advanced ligands (Table 2). This section will summarize the second-generation TSPO radiotracers widely used in AD or MCI. Elevated region-specific TSPO binding signals in a wide range of cortical areas are more illustrated in patients with AD than in normal individuals. The temporal pattern of neuroinflammation over the course of AD has also been well-characterized through longitudinal investigations (55, 60, 62). The role of neuroinflammation is revealed by a large longitudinal study with the administration of $^{18}$F-DPA-714 (60, 61). Participants with MCI and higher initial TSPO binding indicate a slower rate of decline due to dementia than those with lower initial TSPO binding. Combined with the other $^{11}$C-PK11195 studies, the proposal of a dual peak hypothesis of neuroinflammation in AD has been well-established (47). The hypothesis illustrated that neuroinflammation in the early phase of MCI patients is protective and beneficial to AB removal, while activated neuroinflammation in the later phase is detrimental. Notably, the results from TSPO imaging studies could reflect different PET signals to explore the association between TSPO expression and clinical outcomes. Similar to $^{11}$C-PK11195, enhanced TSPO signals are related to impairments in cognition and memory, visuospatial and language ability, executive functioning, dementia severity, and brain atrophy. Further research and clinical trials are urgently required when illustrating regional uptake patterns of TSPO ligands in MCI. Some studies indicated that striking patterns of high cortical tracer retention, especially in the temporal lobe, have been observed compared to healthy controls (47, 63, 65).

Nevertheless, it is necessary to understand the limitation of second-generation TSPO radiotracers, particularly sensitivity to the single-nucleotide polymorphism (SNP) of the TSPO gene. Genetic variation in the SNP rs6917 leads to different binding affinities in AD or MCI. Elevated region-specific TSPO binding signals in a wide range of cortical areas are more illustrated in patients with AD than in normal individuals. The temporal pattern of neuroinflammation over the course of AD has also been well-characterized through longitudinal investigations (55, 60, 62). The role of neuroinflammation is revealed by a large longitudinal study with the administration of $^{18}$F-DPA-714 (60, 61). Participants with MCI and higher initial TSPO binding indicate a slower rate of decline due to dementia than those with lower initial TSPO binding. Combined with the other $^{11}$C-PK11195 studies, the proposal of a dual peak hypothesis of neuroinflammation in AD has been well-established (47). The hypothesis illustrated that neuroinflammation in the early phase of MCI patients is protective and beneficial to AB removal, while activated neuroinflammation in the later phase is detrimental. Notably, the results from TSPO imaging studies could reflect different PET signals to explore the association between TSPO expression and clinical outcomes. Similar to $^{11}$C-PK11195, enhanced TSPO signals are related to impairments in cognition and memory, visuospatial and language ability, executive functioning, dementia severity, and brain atrophy. Further research and clinical trials are urgently required when illustrating regional uptake patterns of TSPO ligands in MCI. Some studies indicated that striking patterns of high cortical tracer retention, especially in the temporal lobe, have been observed compared to healthy controls (47, 63, 65).

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3.3 Other radiotracers targeting neuroinflammation

It is imperative to develop other novel and effective imaging radiotracers to measure neuroinflammation with higher specificity and affinity since TSPO is not specifically located in microglia.
Therefore, promising targets should present precise localization in neuroinflammation and particular ligands to enable imaging measurement (68–71). Further work is required to explore novel targets for the activated microglia and their ability to phagocytose Aβ. Transcriptional profiling of human microglia in plaque-associated and parenchymal tissues could decipher changes in whole tissue RNA, leading to novel targets identified and prioritized for further investigation (72). In the following section, we summarized the other neuroinflammation-targeted ligands for imaging neuroinflammation.

### 3.3.1 CSF1R PET imaging

Colony-stimulating factor 1 receptor (CSF1R) is dominantly expressed in the microglia, macrophages/monocytes, and dendritic cells in the brain parenchyma. CSF1R presents an essential role in microglia growth, proliferation, and survival. Previous studies have determined that the growth factors of colony stimulating factor-1 (GCSF1) and interleukin-34 (IL-34) are endogenous ligands toward CSF1R (73). It has also been demonstrated that the upregulation of CSF1R is responsible for injury and AD-related neuropathology (74, 75). Due to the role

| Radiotracer | Included individuals | Conclusion | Year | Author |
|-------------|----------------------|------------|------|--------|
| $^{11}$C-PK11195 | 8 AD patients and 15 normal individuals | Elevated levels of radiotracer level were observed in brain areas in AD patients. Uptake in the left inferior temporal lobe differentiated AD patients with a sensitivity of 75%. | 2001 | Cagnin (38) |
| $^{11}$C-PK11195 | 13 AD patients and 10 normal individuals | Areas in frontal temporal, parietal, and occipital association cortex showed increased radiotracer uptake in AD patients than controls. | 2008 | Edison (39) |
| $^{11}$C-PK11195 | 6 AD patients, 6 MCI patients, and 5 normal individuals | No statistic difference in TSPO binding was observed when comparing the AD with controls in any brain region. | 2009 | Wileey (40) |
| $^{11}$C-PK11195 | 14 MCI patients and 10 normal individuals | Frontal cortical regions presented higher TSPO binding in MCI patients compared to controls. | 2009 | Okello (41) |
| $^{11}$C-PK11195 | 11 AD patients and 10 normal individuals | Higher $^{11}$C-PK11195 retention was observed in medial frontal, parietal, and left temporal cortical areas in AD patients compared to controls. Additionally, uptake in the left anterior cingulate, left precentral, left hippocampus, and left medial frontal cortex presented negative relationship with cognitive performance. | 2011 | Yokokura (42) |
| $^{11}$C-PK11195 | 19 AD patients, 10 MCI patients, and 21 normal individuals | The bilateral occipital cortex is the only brain region assessed with a statistical difference between AD patients and controls, while no such differences were found when comparing MCI patients to controls. | 2013 | Schuitmaker (43) |
| $^{11}$C-PK11195 | 8 AD patients and 14 normal individuals | Increased microglial tracer uptake in frontal, parietal, occipital, temporal cortical areas, and striatum and hippocampus was observed in AD patients compared to controls. | 2015 | Fan (44) |
| $^{11}$C-PK11195 | 10 AD patients, 10 MCI patients, and 16 normal individuals | Cortical retention of $^{11}$C-PK11195 in the occipital lobe, temporal lobe, hippocampus, and parahippocampus was higher in AD patients compared to controls. Additionally, temporal, frontal, orbital, straight, parietal gyrus, insula, putamen, and occipital lobe presented higher $^{11}$C-PK11195 retention in MCI compared to controls. | 2015 | Fan (45) |
| $^{11}$C-PK11195 | 8 AD patients and 8 normal individuals | $^{11}$C-PK11195 uptake in the areas of medial temporal regions and the hippocampus in AD was negatively related to hippocampal volume. | 2016 | Femminella (46) |
| $^{11}$C-PK11195 | 8 AD patients, 8 MCI patients, and 14 normal individuals | MCI patients showed reductions in $^{11}$C-PK11195 uptake in the region of temporal, occipital, parietal, cingulate cortex, and the hippocampus after 14 months, while AD patients showed an increase in microglial activation than controls. | 2017 | Fan (47) |
| $^{11}$C-PK11195 | 42 MCI patients and 10 normal individuals | In amyloid positive MCI subjects, TSPO binding was elevated in frontal, parietal, and lateral temporal regions compared to controls. Moreover, positive correlation was observed between the results of $^{11}$C-PK11195 and $^{11}$C-PiB in frontal, temporal, and parietal brain areas. | 2017 | Parbo (48) |
| $^{11}$C-PK11195 | 16 AD and MCI patients, and 13 normal individuals | Areas within the occipital, parietal, temporal cortex, and medial temporal regions showed increased radiotracer uptake in the AD and MCI combined group compared to controls. | 2018 | Passamonti (49) |
| $^{11}$C-PK11195 | 6 AD patients, 20 MCI patients, and 20 normal individuals | In the areas of frontal, posterior cingulate, parahippocampal, lateral and posterior temporal cortex, precuneus, and hippocampus, increased TSPO binding was observed in MCI patients compared to controls. | 2018 | Parbo (50) |

TABLE 1  Studies examining regional brain uptake via first-generation TSPO tracers in AD and MCI.
of CSF1R, a novel CSF1R-targeting radiotracer $^{11}$C-CPPC was developed. In animal acute inflammation models, the encephalomyelitis model of multiple sclerosis, and APPswi with cerebral Ab pathology, increased microglial levels of CSF1R have been captured through this radiotracer. Moreover, a recent immunochemical evidence, COX-1 and COX-2 are located in both microglia and neuron in the CNS. Various radiotracers showed specificity toward COX-1 (80, 86). A few studies demonstrated that $^{11}$C-KTP-Me harbors a greater BBB entrance and selective sensitivity towards COX-1 (82, 86). A positive correlation between $^{18}$F-FEPPA binding and $^{11}$C-PiB can be observed in aMCI in the hippocampus. A few studies demonstrated that $^{11}$C-KTP-Me harbors a greater BBB entrance and selective sensitivity towards COX-1 (80, 86). Moreover, clinical trials with $^{11}$C-KTP-Me revealed an elevated brain signal in AD patients compared to normal individuals. $^{11}$C-KTP-Me accumulation can be seen in activated microglia surrounding Aβ plaques within the frontal cortex and hippocampus. Similar in vivo studies can also be observed in APPswi (Tg2576) mice compared to wild-type mice (80, 81). Additionally, previous studies illustrated that both $^{11}$C-Ps1 (COX-1 PET imaging) and $^{11}$C-MCI (COX-2 PET imaging) radiotracers showed specific detection patterns after LPS-

### TABLE 2: Studies examining regional brain uptake via advanced TSPO tracers in AD and MCI.

| Radiotracer   | Included individuals | Conclusion                                                                                           | Year  | Author          |
|---------------|----------------------|------------------------------------------------------------------------------------------------------|-------|-----------------|
| $^{11}$C-PBR28 | 19 AD patients, 10 MCI patients, and 13 normal individuals | Areas of prefrontal, inferior parietal, temporal, precuneus, posterior cingulate, occipital, hippocampus, and entorhinal cortex presented higher $^{11}$C-PBR28 binding in AD patients compared to controls, while no such difference was observed in MCI patients. | 2013  | Kreisl (55)     |
| $^{11}$C-PBR28 | 25 AD patients, 11 MCI patients, and 21 normal individuals | Areas of temporal and parietal brain presented higher $^{11}$C-PBR28 uptake in AD patients compared to controls, while no such difference was observed in MCI patients. | 2015  | Lyoo (56)       |
| $^{11}$C-PBR28 | 14 AD patients and 8 normal individuals | Areas of inferior parietal lobule, occipital cortex, precuneus, entorhinal cortex, hippocampus, inferior, and middle temporal cortex presented higher $^{11}$C-PBR28 binding in AD patients compared to controls. Annual increase in radiotracer binding was also observed in AD patients. | 2016  | Kreisl (57)     |
| $^{11}$C-PBR28 | 13 MCI patients and 9 normal individuals | Higher radiotracer uptake in the temporal lobe, post-cingulate cortex, thalamus, medial temporal lobe, hippocampus, amygdala, and cerebellum can be observed in MCI patients than controls. | 2018  | Fan (58)        |
| $^{11}$C-PBR28 | 16 AD patients, 16 MCI patients, and 19 normal individuals | Positive correlations were found between $^{11}$C-PBR28 and amyloid retention on $^{18}$F-flutemetamol and tau aggregation measured by $^{18}$F-AV-1451. | 2018  | Dani (59)       |
| $^{11}$C-DPA-713 | 17 AD patients and 22 normal individuals | $^{11}$C-DPA-713 presented higher accuracy in TSPO binding than $^{11}$C-PK11195 in AD patients, and demonstrated an inverse relationship with cognition. | 2017  | Yokoura (31)    |
| $^{18}$F-DPA-714 | 64 AD patients and 32 normal individuals | Areas of precuneus, parietal, temporal cortex, and medium and posterior cingulate presented higher $^{18}$F-DPA-714 uptake in AD patients compared to controls. | 2016  | Hamelin (60)    |
| $^{18}$F-DPA-714 | 52 AD patients and 17 normal individuals | Areas of temporal and parietal brain presented higher tracer retention in AD patients relative to controls. Annual increases of 12.2% were observed for AD patients. | 2018  | Hamelin (61)    |
| $^{11}$C-C | 10 AD patients and 10 normal individuals | Areas of cerebellum, prefrontal cortex, parietal cortex, temporal cortex, occipital cortex, anterior cingulate cortex, and striatum presented higher $^{11}$C-DAA1106 binding in AD patients compared to controls. | 2008  | Yasuno (62)     |
| DAA1106 | 10 AD patients, 7 MCI patients, and 10 normal individuals | Areas of striatum, lateral temporal, parietal, and anterior cingulate cortex presented increased $^{11}$C-DAA1106 binding in AD patients compared to controls. | 2012  | Yasuno (63)     |
| $^{18}$F-FEPPA | 21 AD patients and 21 normal individuals | Areas of temporal, frontal, parietal, and occipital cortical regions, and the hippocampus presented increased $^{18}$F-FEPPA retention in AD patients compared to controls. | 2015  | Surdijan (64)   |
| $^{18}$F-FEPPA | 11 MCI patients and 14 normal individuals | A positive correlation between $^{18}$F-FEPPA binding and $^{11}$C-PiB can be observed in aMCI in the hippocampus. | 2017  | Knezevic (65)   |
| $^{18}$F-FEMPA | 10 AD patients and 7 normal individuals | Areas of medial temporal, lateral temporal, posterior cingulate cortex, putamen, caudate, thalamus, and cerebellums presented increased $^{18}$F-FEMPA uptake in AD patients compared to controls. | 2015  | Varrone (36)    |
| $^{18}$F-GE-180 | 6 AD patients and 7 normal individuals | Cerebellum is a suitable pseudo-reference region for PET imaging of AD by $^{18}$F-GE-180. No significant increases in $^{18}$F-GE-180 binding in the frontoparietal VOIs of patients with AD when compared to the healthy controls. | 2021  | Vettermann (66) |

3.3.2 COX1 and COX2 PET imaging

Cyclooxygenase (COX) plays an essential role in the generation of prostaglandin H2, the substrate for prostaglandins and thromboxanes. There are two isoforms of COX, COX-1 and COX-2, determined to present a critical role in neuroinflammation and links to various neurodegenerative diseases, especially AD. According to the results of immunochemical evidence, COX-1 and COX-2 are located in both microglia and neuron in the CNS. Various radiotracers for COX-1 and COX-2 have been well established, such as $^{18}$F-TMI, $^{18}$F-triacoxib, $^{11}$C-rofecoxib, $^{11}$C-KTP-Me, $^{11}$C-Ps13, and $^{11}$C-MCI (79–86). A few studies demonstrated that $^{11}$C-KTP-Me harbors a greater BBB entrance and selective sensitivity towards COX-1 (80, 86). Moreover, clinical trials with $^{11}$C-KTP-Me revealed an elevated brain signal in AD patients compared to normal individuals. $^{11}$C-KTP-Me accumulation can be seen in activated microglia surrounding Aβ plaques within the frontal cortex and hippocampus. Similar in vivo studies can also be observed in APPswi (Tg2576) mice compared to wild-type mice (80, 81). Additionally, previous studies illustrated that both $^{11}$C-Ps1 (COX-1 PET imaging) and $^{11}$C-MCI (COX-2 PET imaging) radiotracers showed specific detection patterns after LPS-
induced neuroinflammation in monkey brain and human inflammatory tissues (82, 83).

3.3.3 P2X7R and P2Y12R PET imaging

The upregulated levels of the purinergic P2X7 receptor (P2X7R) can activate neuroinflammation, especially in M1 microglia. P2X7R had various biological functions, including inflammasome activation, cytokine secretion, T lymphocyte differentiation, and cell death (87). Microglia monitors through P2Y12R-dependent junctions are associated with mitochondrial activity in neurons (88). Brain injuries altered somatic junctions that induced P2Y12R-dependent neuroprotective effects through calcium load and functional connectivity in neurons (89, 90). Based on the immunohistochemical staining results, levels of P2Y12R were decreased in the brains of AD patients (91). Accumulative evidence has revealed that various P2X7R-targeting radiotracers have been developed currently, such as 11C-GSK1482160, 11C-JNJ-47965567, 18F-JNJ-64413739, 11C-JNJ-54173717, 11C-SMW139, and 18F-PTTP (92–98). A previous study overexpressed human P2X7R in a rat model by rAAV3lag-hP2X7R and demonstrated that 11C-SMW139 had higher affinity and specificity to the P2X7R (99). Moreover, brains of AD patients revealed higher 11C-SMW139 binding compared to healthy individuals through autoradiography, consistent with histological staining results (99). An ongoing clinical trial applied 11C-SMW139 as a PET imaging radiotracer toward neuroinflammation in Parkinson’s disease (PRI-PD: 2018-000405-23). The other probes are P2Y12R-based radiotracers, including 11C-AZD1283, 11C-P2Y12R-ant, and 11C-Cou, assessed among animal models (94, 100, 101). A previous study has demonstrated that the P2Y12 radiotracer, 11C-AZD1283, distinctly responds to tau and amyloid deposits. The levels of P2Y12R binding increase in APP23 and APPNL-F mice (101). However, 11C-AZD1283 PET imaging showed no signal in the wild-type mouse brain.

Additionally, two other radiotracers, 11C-P2Y12R and 11C-Cou, exerted sufficient brain uptake, high affinity, and promising results in experimental autoimmune encephalomyelitis and stroke models, detecting anti-inflammatory microglia (48) (94, 100).

3.3.4 MAO-B PET imaging

In several clinical trials, 11C-deuterium-L-deprenyl (DED) MAO-B inhibitors have also been applied in PET imaging toward neuroinflammation. It has been proved that early astrocytosis can be measured via 11C-DED in sporadic and autosomal dominant AD patients and amyloidosis mouse models (102–110). In addition, 18F-fluorodeprenyl-D2 showed favorable kinetic properties and ameliorated affinity in MAO-B imaging (111). However, the technical problems of irreversible inhibitors impede the accuracy of imaging. Therefore, several reversible-binding inhibitors have been developed and validated, including 11C-Cou, 11C-SL25.1188, and 11C-SMBT-1 (69, 112, 113). Among the advanced PET imaging based on reversible-binding inhibitors, a specific elevated regional retention of 11C-SMBT-1 within the cortical and hippocampal regions can be seen in patients with AD compared to healthy individuals (114).

3.3.5 I2BS PET imaging

I2BS is located on both monoamine oxidases A (MAO-A) and B (MAO-B), which is another novel target for PET imaging toward neuroinflammation (115–118). 11C-FTIMD presented the specific binding to I2BS in the monkey brain (119). A previous study has revealed that the activated astroglia determined using 11C-BU99008 PET in the early period of Parkinson’s disease is responsible for α-synuclein accumulation (116). A previous in vitro study demonstrated that 3H-BU99008 revealed high specificity in brain tissues from AD patients and colocalized with glial fibrillary acidic protein staining of astrocytes (120). Moreover, a clinical study reported that an increasing 3H-BU99008 binding signal could be detected in the brain of patients with AD compared to healthy individuals. Similar results were reported in the cortical region via 11C-BU99008, consistent with the high cerebral Aβ load evaluated by 18F-florbetaben in MCI and AD patients (121). A previous study demonstrated that increased 11C-BU99008 signaling could be detected in earlier stages with low Aβ loads. At the same time, reduced astrocytosis can be observed in the advanced stages with a more significant Aβ load and atrophy (122).

3.3.6 CB2R PET imaging

Cannabinoid type 2 receptor (CB2R) is a member of the endogenous cannabinoid system. CB2R has a low concentration in the brain during healthy conditions. However, Aβ deposition activates microglia leading to the high expression of CB2R, another widely considered dominant marker of AD neuroinflammation. Various PET radiotracers with higher affinity towards CB2R have been well established. Among the developed CB2R imaging agents, 11C-NE40 can specifically and reversibly bind to CB2R. Previous studies have depicted that the uptake of 11C-NE40 was reduced in the brain of AD patients due to loss of neuron-based CB2R expression, contrary to the expectations from preclinical studies. This inconsistency may be due to the low expression of CB2R and insufficient selectivity for CB2R and CB1R (123). The other CB2R agonists with high affinity are under exploration, including 11C-MA2, 18F-MA3, or 18F-RS126 (124).

3.3.7 11C-AA PET imaging

AA, a kind of n-6 polyunsaturated fatty acid and an essential component of the metabolic network of inflammation, is abundant in the brain parenchyma and participates in cell signal transduction. Microglia in the CNS release the inflammatory cytokines and bind
to receptors on the surface of astrocytes, leading to the secretion of phospholipase and cytoplasmic phosphatase and mediated AA release. Therefore, AA detection can serve as an indirect marker of microglial activation (125). Based on the above results, 11C-AA and PET imaging can determine brain phospholipase activity. By injecting 11C-AA, the regional brain incorporation coefficients and metabolic loss of AA in the brain can be determined (125). Therefore, elevated signals of 11C-AA could evaluate the disordered metabolism of AA due to neuroinflammation.

3.3.8 Nicotinic acetylcholine receptors

It has been demonstrated that nicotinic acetylcholine receptors (nAChRs) were closely related to neuroinflammation. Moreover, the ligand 2-18F-A85380 (2-FA) towards nAChR has also presented similar patterns of uptake with 11C-PK11195 in activated microglia and astrocytes (126). Additionally, 18F-flutabine has been established with more favorable kinetic profile, which leads to a better understanding of nAChRs in neuroinflammation (126, 127). The homomeric nAChRs are colocaled in neuritic plaques in patients’ brain with AD, and Aß1–42 has been reported to bind to the α7 nAChRs with high affinity (128). α7 nAChRs are also strongly expressed on astrocytes and microglia, the activation of which has been shown to suppress inflammatory processes (128). Recently, several new compound-based bio-tracers, such as 18F-ASEM and 18F-DBT-10, presented more promising results (129).

4 Methodological issues in radiotracers quantification by PET

A variety of quantification methods have been established due to the tracers. It appears that it is a great challenge for quantification by PET and that various factors should be taken into account. (1) Genetic polymorphism and affinity: The main limitation of the second-generation TSPO radiotracers is their sensitivity to a polymorphism of the TSPO gene. This polymorphism leads to differential affinity of these ligands to TSPO (67). (2) Arterial plasma input function: The kinetics model analyses are based on the input function, which requires the relative traumatic placement of an arterial catheter and the development of radioanalytical methods to accurately identify the plasma metabolite fractions (56). (3) Reference region definition: The reference region should be devoid of specific ligand binding to the target and only share the same free and non-specific binding with the region expressing the target and remain unaffected by the disease.

5 Conclusion

The interplay between amyloid, tau, and neuroinflammation is a brand new area of investigation that has only recently become possible through the development of an expanded repertoire of PET tracers. Therefore, we reviewed various strategies for PET imaging for neuroinflammation and summarized the dominant results from previous clinical trials on AD patients. There are a variety of approaches explaining the exploration of neuroinflammation. Although determining neuroinflammation by PET imaging in AD is widely used, there is a conflict on whether neuroinflammation is beneficial or detrimental to the evolution of the symptoms, neuronal injury, and cognitive deficits. This review also highlighted are a number of areas of uncertainty and various radiotracers’ limitations. The challenges of radiotracer establishment and accurate binding quantification, the complicated neuroinflammation in the brain tissue of AD patients, and its extensive roles in different stages have also been emphasized. PET imaging towards neuroinflammation is a potential approach to deciphering the pathophysiological processes of AD patients. It clarifies the links between amyloid/tau pathologies and neuroinflammation, influencing different clinical symptoms and pathophysiological progression. More efforts are also required to improve the approaches to determine the binding signal from the PET imaging based on the current radiotracers and develop novel radiotracers. Moreover, deciding on a consensus to standardize the PET data analysis is critical. Given the proposal that the role of neuroinflammation in AD pathogenesis changes over the disease course, this is a prerequisite to conducting multi-tracer longitudinal studies based on multiple centers and elucidating the multifaced role of neuroinflammation in AD and MCI patients.
Microglial activation correlates with hypometabolism in Alzheimer’s disease. *Eur J Nucl Med Mol Imaging* (2011) 38:343–51. doi: 10.1007/s00259-010-1612-0

Schuettemaker A, Krogholler MA, Boelddal R, van der Fler W, Klot RW, van der Doef TF, et al. Microglial activation in Alzheimer’s disease. *Ann (R) [11C]PK11195 position emission tomography study*. *Neurobiol Aging* (2013) 34:128–36. doi: 10.1016/j.neurobiolaging.2012.04.021

Fan Z, Okello AA, Brooks DJ, Edison P. Longitudinal influence of microglial activation and amyloid on neuronal function in Alzheimer’s disease. *Brain: J Neurol* (2015) 138:3665–98. doi: 10.1093/brain/awv349

Fan Z, Aman Y, Ahmed I, Chatlet G, Landeau B, Ray-Chaudhuri K, et al. Influence of microglial activation on neuronal function in Alzheimer’s and Parkinson’s disease dementia. *Alzheimer’s Dement* (2015) 11:608–621.e607. doi: 10.1016/j.jalz.2015.05.004

Feminidilla GD, Nisan S, Atkinson R, Edison P, et al. Does microglia-mediated influence hippocampal volume and neuronal function in Alzheimer’s disease and Parkinson’s disease dementia? *J Alzheimers Dis* (2016) 51:1275–89. doi: 10.3233/jad-150827

Fan Z, Brooks DJ, Okello A, Edison P. An early and late peak in microglial activation in Alzheimer’s disease trajectory. *Brain: J Neurol* (2017) 140:792–803. doi: 10.1093/brain/awx120

Passamonti L, Rodriguez PV, Hong YT, Allinson KSJ, Bevan-Jones WR, Williamson D, et al. (11)C]PK11195 binding in Alzheimer disease and progressive supranuclear palsy. *Neurology* (2018) 90:e1949–96. doi: 10.1227/wnl.0000000000005610

Parbo P, Ismail R, Hansen KV, Amidi A, Mårup FH, Grottap H, et al. Brain inflammation accompanies amyloid in the majority of mild cognitive impairment cases due to Alzheimer’s disease. *Brain: J Neurol* (2017) 140:2004–11. doi: 10.1093/brain/awx120

Kreisl WC, Kim MJ, Coughlin JM, Henter ID, Innis RB. PET imaging of neuroinflammation in neurodegenerative disorders. *Lancet Neurol* (2020) 19:940–50. doi: 10.1016/S1474-4422(20)30346-2

Narayanaswami V, Drake LR, Brooks AF, Meyer JH, Hode S, Kilbourn MR, et al. Classics in neuroimaging: Development of PET tracers for imaging monoamine oxidases. *ACS Chem Neurosci* (2019) 10:1867–71. doi: 10.1021/acschemneuro.9b00081

Janssen B, Mach RH. Development of brain PET imaging agents: Strategies for imaging neuroinflammation in Alzheimer’s disease. *Prog Mol Bio Chem Sci* (2019) 16:371–99. doi: 10.1016/j.pmbs.2019.04.005

Janssen B, Vugs DJ, Windhorst AD, Mach RH. PET imaging of microglial activation beyond targeting TSPO. *Molecules* (2018) 23(3):251–61. doi: 10.3390/ molecules23030357

Kreisl WC, Kim MJ, Coughlin JM, Henter ID, Innis RB. PET imaging of neuroinflammation in neurodegenerative disorders. *Lancet Neurol* (2020) 19:940–50. doi: 10.1016/S1474-4422(20)30346-2

Huang 10.3389/fimmu.2022.1010946
PET tracer of microglial activation. Identi – 17:300. doi: 10.1186/s12974-020-01962-7

(18)F-PTTP for differentiation of lung tumor from in – 259. doi: 10.1186/s12974-017-1034-z

PET imaging of P2X(7)R in the experimental autoimmune encephalomyelitis study. doi: 10.2967/jnumed.118.216747

Horssen J, et al. Purinergic receptors P2Y12R and P2X7R: Potential targets for PET radioligands for PET imaging of cyclooxygenase-2 in an ischemic neuronal injury. In vivo – 87. doi: 10.1002/glia.23097

Characterization of (11)C-GSK1482160 for targeting the P2X7 receptor as a methyl ester.

receptor in infection and in – 570. doi: 10.1093/brain/awz260

Loss of Brain: J Neurol – 87. doi: 10.1002/glia.23097

Radioligands for PET imaging of cyclooxygenase-2 in an ischemic neuronal injury. In vivo – 87. doi: 10.2967/jnumed.110.084046

Madsen A, Strabburger H, Ayata P, Chen X, Nair A, Igeami A, et al. Negative feedback control of neuronal activity by microglia. Nature – 560:847–23. doi: 10.1038/s41586-020-2777-8

Mödlner A, Huang H, Radke J, Stenzel W, Priller J. P2Y12 receptor is expressed on microglia under physiological conditions throughout development and is sensitive to neuronal inflammation. Glia – 65:375–87. doi: 10.1002/glia.23097

Han J, Liu H, Liu C, Jin H, Perlmutt JS, Egan TM, et al. Pharmacologic characterizations of a P2X7 receptor–specific radioligand. J Nucl Med – 38:372–82. doi: 10.2967/jnum.111.090560

Terrio PR, Meyer JA, Peters JS, Riley AA, McCarthy BP, Gao M, et al. Characterization of (11)C-GSK1482160 for targeting the P2X7 receptor as a biomarker for neuronal inflammation. J Nucl Med – 57:48–65. doi: 10.2967/jnumed.116.183514

Beano W, Janssen B, Kooij G, van der Pol SMA, van Het Hof B, van – 61:604–7. doi: 10.2967/jnumed.119.331985

Beano W, Janssen B, Kooijman E, Vugts DJ, Wilkinson SM, Ory D, Chalon S, Hoozemans JJM, et al. Comparison of early-phase 11C-Deuterium-l-Deprenyl and 11C-deprenyl and ³H-PIB autoradiography show different laminar distributions of amyloid and fibrillary β-amyloid in Alzheimer brain. J Nucl Neuroim – 101. doi: 10.2967/jnumed.110.084046

Olsen M, Aguilar X, Selhin D, Fang XT, Antoni G, Erlandsson A, et al. Astroglial responses to amyloid beta-preservation in a mouse model of Alzheimer’s disease. Mol Imaging Biol – 20:605–14. doi: 10.1007/s11307-011-1153-2

Nag S, Fazio P, Lehmann I, Kettschaunag H, Heinrich T, Theile A, et al. In vivo and In vitro characterization of a novel MAO-b inhibitor radioligand, 18F-labeled deuterated fluoroepinephrine. J Nucl Med – 56:315–20. doi: 10.2967/jnumed.115.161083

Moriguchi S, Wilson AA, Mäer L, Rusjan PM, Vaszek N, Kish SJ, et al. Astrocyte function and glucose metabolism in autosomal dominant Alzheimer’s disease. JAMA Psychiatry – 7. doi: 10.1001/jamapsychiatry.2019.0044

Drake LR, Brooks AF, Mufarrej AJ, Pham JM, Koepp RA, Shao X, et al. Deuterium kinetic isotope effect studies of a potential in vivo metabolic trapping agent for monooamine oxide b. ACS Chem Neurosci – 9:3024–7. doi: 10.1021/acschemneuro.8b00103

Wilson H, Dervenoulas G, Pagana G, Tyackle RJ, Polychronis S, Myers J, et al. Midazolam 2 binding sites reflecting astroglia pathology in parkinson’s disease: an in vivo11C-BU99008 PET study. Brain Neuroim – 142:3116–28. doi: 10.1093/brain/aww290

Tsuyuki RJ, Myers JM, Verkman KA, Mack I, Turton S, Pasquier J, et al. Evaluation of (11)C-BU99008, a PET ligand for the Immunozone(2) binding site in human brain. J Nucl Med – 59:1597–607. doi: 10.2967/jnumed.18.210809

Tesson F, Lemoine I, Gillberg PG, Bogdanicov N, Nennesmo I, Saint-Aubert L, Viitanen M, et al. Amyloid, tau, and astrocyte pathology in autosomal-dominant Alzheimer’s disease variants. AJNParc and PSEN1DE96 Mol Psychiatry – 26:5609–19. doi: 10.1038/s41380-020-0187-2

Manuele G, Gillberg PG, Bergfors A, Yu W, N, Sennestam I, et al. ‘H- deprenyl and ‘H-PIB autoradiography show different laminar distributions of astroglia and fibrillary β-amyloid in Alzheimer brain. J Nucl Neuroim – 118:1090. doi: 10.1186/s12974-2016-00740-8

Kim MJ, Eldridge M, Lehmann ML, Frankland M, Liow JS, et al. Astrocytosis precedes amyloid plaque deposition in Alzheimer APP/PS1 transgenic mouse brain: A correlative positron emission tomography and in vitro imaging study. Eur J Nucl Med Mol Imaging – 42:1119–32. doi: 10.1007/s00259-015-3047-0

Vilaplana E, Rodriguez-Vieitez F, Ferreira D, Montal V, Almkvist O, Wall A, et al. Cortical microstructural correlates of astrocytosis in autosomal-dominant Alzheimer disease. Neurology – 2018;94:e2026–36. doi: 10.1212/wnl.0000000000004905

Schröder C, Fischle W, Kuhlman J, Judge JH, et al. Characterization of the regional binding distribution of amyloid PET tracer flortetaben and the glia tracers deprenyl and PK11195 in autopsy Alzheimer’s brain tissue. J Alzheimers Dis – 2021. doi: 10.2333/jad-201344

Zrzavy T, Hametner S, Wimmer I, Butovsky O, Weiner HL, Lassmann H. Immunohistochemistry and IF analysis of new molecular targets for PET imaging of the microglial anti-inflammatory response. Frontiers in Immunology – 11:5527. doi: 10.3389/fimmu.2020.010946
121. Calsolaro V, Matthews PM, Donat CK, Livingston NR, Femminella GD, Guedes SS, et al. Astrocyte reactivity with late-onset cognitive impairment assessed in vivo using (11)C-BU99008 PET and its relationship with amyloid load. *Mol Psychiatry* (2021) 26:5848–55. doi: 10.1038/s41380-021-01193-z

122. Livingston NR, Calsolaro V, Hinz R, Nowell J, Raza S, Gentleman S, et al. Relationship between astrocyte reactivity, using novel (11)C-BU99008 PET, and glucose metabolism, grey matter volume and amyloid load in cognitively impaired individuals. *Mol Psychiatry* (2022) 27:2019–29. doi: 10.1038/s41380-021-01429-y

123. Ahmad R, Postnov A, Bormans G, Versijpt J, Vandenbulcke M, Van Laere K. Decreased in vivo availability of the cannabinoid type 2 receptor in alzheimer’s disease. *Eur J Nucl Med Mol Imaging* (2016) 43:2219–27. doi: 10.1007/s00259-016-3457-7

124. Slavik R, Müller Herde A, Haider A, Krämer SD, Weber M, Schibli R, et al. Discovery of a fluorinated 4-oxo-quinoline derivative as a potential positron emission tomography radiotracer for imaging cannabinoid receptor type 2. *J Neurochem* (2016) 138:874–86. doi: 10.1111/jnc.13716

125. Esposito G, Giovacchini G, Lione JS, Bhattacharjee AK, Greenstein D, Schapiro M, et al. Imaging neuroinflammation in alzheimer’s disease with radiolabeled arachidonic acid and PET. *J Nucl Med* (2008) 49:1414–21. doi: 10.2967/jnumed.107.049619

126. Albrecht DS, Granziol C, Hooker JM, Loggia ML. In vivo imaging of human neuroinflammation. *ACS Chem Neurosci* (2016) 7:470–83. doi: 10.1021/acschemneuro.6b00056

127. Lagarde J, Sarazin M, Chauviré V, Stankoff B, Kas A, Lacomblez L, et al. Cholinergic changes in aging and Alzheimer disease: An [18F]-F-A-85380 exploratory PET study. *Alzheimer Dis Assoc Disord* (2017) 31:8–12. doi: 10.1097/ wad.0000000000000163

128. Wang HY, Lee DH, D’Andrea MR, Peterson PA, Shank RP, Reitz AB. Beta-Amyloid(1-42) binds to alpha7 nicotinic acetylcholine receptor with high affinity: implications for alzheimer’s disease pathology. *J Biol Chem* (2000) 275:5626–32. doi: 10.1074/jbc.275.8.5626

129. Hillmer AT, Li S, Zheng MQ, Schuememann M, Lin SF, Nabulsi N, et al. PET imaging of t(7) nicotinic acetylcholine receptors: A comparative study of [(18)F]ASEM and [(18)F]FDRT-10 in nonhuman primates, and further evaluation of [(18)F]ASEM in humans. *Eur J Nucl Med Mol Imaging* (2017) 44:1042–50. doi: 10.1007/s00259-017-3621-8