Intubated patients developing tracheobronchitis or pneumonia have distinctive complement system gene expression signatures in the pre-infection period: A pilot study

I. Martin-Loeches, E. Papiol, R. Almansa, G. López-Campos, J.F. Bermejo-Martin, J. Rello

Critical Care Centre, Corporació Sanitaria Parc Taulí-Hospital de Sabadell - CIBERES, Sabadell, Spain
Critical Care Department, Vall d’Hebron University Hospital, VHIR, CIBERES, Universitat Autonoma de Barcelona, Spain
Microarrays and Medical Bioinformatics (MBM), Infection & Immunity Unit, Hospital Clínico Universitario-IÉSCYVL, Valladolid, Spain
Medical Bioinformatics Department, Institute of Health ''Carlos III'', Madrid, Spain

Received 5 August 2011; accepted 15 October 2011
Available online 1 February 2012

KEYWORDS
Ventilator associated pneumonia;
Gene expression;
Ventilator associated tracheobronchitis;
Genes;
Sepsis;
RNA;
Biomarker;
VAP;
VAT

Abstract
Introduction: It remains unknown why some intubated patients remain infection-free while others develop tracheobronchitis (VAT) or pneumonia (VAP).
Objective: To identify and compare VAP/VAT gene expression “signatures” using genome-wide oligonucleotide microarrays.
Material and methods: A prospective translational study of gene expression profiles of VAP and VAT groups was carried out, establishing comparisons in both pre-infection and infection phases. Pathway and functional analyses were performed with Ingenuity Pathway Analysis (IPA). Data analysis and hierarchical clustering of the genes involved in the signalling pathways expressed differentially in the two groups were performed with GeneSpring GX 11.0.
Results: Eight patients developing respiratory infections (3 VAP and 5 VAT) after 4 days of mechanical ventilation were assessed. Comparison of gene expression profiles in the pre-infection period revealed 5595 genes expressed differentially between VAP and VAT (p < 0.01, fold change >2). Comparative IPA analysis identified a significant depression of the complement system signalling pathway in the VAP group, affecting the classical pathway along with the final common pathway (p < 0.05). In addition, the cAMP and calcium signalling pathways were also significantly depressed in the VAP group during the pre-infection phase also.
Conclusion: Intubated patients complicated with pneumonia developed immune impairment in the pre-infection period, manifesting as a relatively lower expression of genes involved in the complement system that differed from patients developing tracheobronchitis. These findings suggest that a significant proportion of VAP episodes cannot be prevented, but might be treatable through pre-emptive therapy.
© 2011 Elsevier España, S.L. and SEMICYUC. All rights reserved.

* Corresponding author.
E-mail address: drmartinloeches@gmail.com (I. Martin-Loeches).

2173-5727/ - see front matter © 2011 Elsevier España, S.L. and SEMICYUC. All rights reserved.
doi:10.1016/j.medin.2011.10.009
Los pacientes intubados que presentan traqueobronquitis o neumonía tienen patrones distintivos de expresión genética del sistema del complemento en el periodo previo a la infección: un estudio piloto

Resumen

Introduction: Seguimos sin saber por qué algunos pacientes intubados no sufren infecciones mientras que otros presentan traqueobronquitis (TAV) o neumonía (NAV). Objetivo: Identificar y comparar los patrones de la expresión genética de la NAV/TAV usando micromatrizes multigenéticas oligonucleotídicas. Material y métodos: Se realizó un estudio aplicado prospectivo de los patrones de la expresión genética de los grupos con NAV y TAV, estableciendo comparaciones tanto en la fase previa a la infección como en la fase infecciosa. Se realizaron análisis de vías y funcionales con Ingenuity Pathway (IPA). Los análisis de datos y el agrupamiento jerárquico de los genes implicados en las vías de señalización expresados de manera diferenciada en ambos grupos se realizaron con GeneSpring GX 11.0. Resultados: Se evaluaron ocho pacientes que presentaron infecciones respiratorias (3 NAV y 5 TAV) después de 4 días con la ventilación mecánica. La comparación de los patrones de la expresión genética durante el periodo previo a la infección reveló 5,595 genes expresados de forma diferenciada entre la NAV y la TAV (p < 0.01, cambio múltiplo > 2). Los análisis comparativos de los IPA identificaron una depresión importante de la vía de señalización del sistema del complemento en el grupo con NAV, que afectó a la vía clásica además de a la vía común final (p < 0.05). Por otra parte, el monofosfato ciclico de adenosina y las vías de señalización del calcio también se vieron muy deprimidos en el grupo con NAV durante la fase previa a la infección. Conclusión: Los pacientes intubados que presentaron complicaciones por la neumonía desarrollaron un trastorno inmunitario durante el periodo previo a la infección, que se manifestó como una expresión relativamente menor de genes implicados en el sistema del complemento diferente de la de los pacientes que presentaron traqueobronquitis. Estos resultados sugieren que no se puede evitar una importante proporción de los casos de NAV, aunque podrían ser tratables mediante un tratamiento preventivo.

© 2011 Elsevier España, S.L. y SEMICYUC. Todos los derechos reservados.

Introduction

Respiratory infections including pneumonia and tracheobronchitis remain the leading nosocomial infections in the ICU. Ventilator-associated tracheobronchitis (VAT) is manifested by purulent respiratory secretions. Differentiating VAT from ventilator-associated pneumonia (VAP) is mainly based on presence/absence of radiographic opacities, but the potential continuum of VAT to VAP is unknown. The incidence of VAT has been reported to be more common in surgical patients. Some reports suggest that patients with VAT also have increased length of ICU stay and a prolonged need for mechanical ventilation.

To gain a better understanding of the molecular mechanisms that underlie the lung phenotypes in VAT vs VAP, we sought to identify and compare their gene expression "signatures" using genome-wide oligonucleotide microarrays. We hypothesized that VAP/VAT would exhibit different gene expression signatures which could help to distinguish between the two conditions and would also shed light on their pathogenesis. We also expected that specific signatures could lead to the identification of new biomarkers to aid in the diagnosis and classification of these diseases.

Methods

Study design

This is a cohort study including ICU patients with VAP/VAT prospectively recorded. The protocol was approved by the ethical committee and informed consent was obtained from the next of kind prior to enrolment in the study. All patients enrolled were recorded anonymously in the registry. Immunocompetent patients who were intubated and ventilated for more than 48 h were eligible for the study and were followed up for a week. Patients, who developed nosocomial pneumonia before or after the occurrence of VAT, were excluded. Patients with chronic respiratory failure, patients who were not ventilated or ventilated for less than 48 h, patients who received only noninvasive pressure ventilation, patients with tracheostomy at ICU admission, patients who had been treated with corticosteroids/immunosuppressive drugs and patients who were immunocompromised were not eligible and not included in the present study.

VAT was defined as the presence of all of the following in a patient endotracheally intubated and receiving mechanical ventilation for >48 h: body temperature > 38.3 °C or <36.0 °C, new or increased purulent tracheal secretions,
positive culture of tracheal secretions at a concentration of ≥10^5 cfu/ml, and no new or progressive infiltrate on portable chest radiograph. VAP was defined as the presence of a new or progressive pulmonary infiltrate and two of the following: temperature >38.3 °C or <36.0 °C, leukocyte count >12,000/mm³ or <4000/mm³, or purulent tracheal secretions. The diagnosis of VAP was considered to be microbiologically confirmed if either BAL or ETA cultures had significant growth. The presence or absence of a new or progressive radiographic infiltrate was based on the interpretation of the chest radiograph by board certified radiologists who were blinded to the study. According to the American-European consensus conference, a diagnosis of ARDS was assigned by the following criteria: acute onset; arterial hypoxemia with PaO₂/FIO₂ lower than 200 (regardless of PEEP level); bilateral infiltrates seen on chest radiograph; pulmonary artery occlusive pressure lower than 18 mm Hg or no clinical evidence of left atrial hypertension; and compatible risk factors.

All classifications, including the radiographs and laboratory data used in their determinations, were prospectively reviewed by one of the investigators (I.M.L.) and confirmed by a second investigator (J.R.). ETAs were only obtained when there was suspicion of either VAT or VAP. Specimens were obtained by nurses using a deep tracheal suctioning technique to obtain a specimen for culture. All ETAs and BAL cultures were processed using quantitative methods as previously described.

Blood samples were taken daily during the study period. Samples for microarray analysis were distributed in two groups, based on the presence of ventilator-associated tracheobronchitis (VAT) or ventilator-associated pneumonia (VAP). Basic investigators remained blind to the final diagnosis. Attending to the timing of infection, two moments of disease evolution were established: pre-infection, defined as the period with absence of positive results in the microbiological test (from day 0 to 5) and infection, defined as the period with positive results in the microbiological test (from day 5 to 7). Gene expression profiles were compared between VAT and VAT groups in both pre-infection and infection phases.

Total RNA was extracted from blood samples using the PAXgene Blood RNA System (PreAnalytiX, Hombrechtikon, Switzerland). RNA was quantified by spectrometry (NanoDrop ND1000, NanoDrop Technologies, Wilmington, DE, USA) and quality confirmed by RNA 6000 Nano Bioanalyzer (Agilent Technologies, Palo Alto, CA, USA) assay. Up to 1750 ng of RNA sample was concentrated with the RNeasy MinElute Cleanup kit (Qiagen, Hilden, Germany). RNA was eluted with 10 μl of RNase-free H₂O. 300 ng of purified total RNA were used to produce Cyanine 3-CTP-labeled cRNA using the Quick Amp Labeling kit (Agilent p/n 5190-0442) according to the manufacturer’s instructions. Following ‘One-Color Microarray-Based Gene Expression Analysis’ protocol Version 5.7 (Agilent p/n 4140-90040), 3 μg of labeled cRNA was hybridized with Whole Human Genome Oligo Microarray Kit (Agilent p/n G2519F-014850) containing 41,000+ unique human genes and transcripts. Arrays were scanned in an Agilent Microarray Scanner (Agilent G2565BA) according to the manufacturer’s protocol and data extracted using Agilent Feature Extraction Software 9.5.3 following the Agilent protocol GE1-v5.95_Feb07 and the QC Metric Set GE1_QCMT_Jan08.

Microarray data analysis

Data analysis was performed in Microarrays & Medical Bioinformatics (MBM), Infection & Immunity Unit, Hospital Clínico Universitario-IECSYL, Valladolid, Spain and Medical Bioinformatics Department, Institute of Health ‘‘Carlos III’’, Madrid, Spain by using GeneSpring GX 11.0 software. The original data were cleaning and normalized using the robust multichip average (RMA) algorithm consists in three steps: background correction, quantile normalization and expression calculation. Subsequent to logarithms transformation, baseline transformation of the data was performed using the median of all samples.

Before statistical analyses all microarrays were subjected to quality and filtering criteria. Quality of the microarray data was assessed on principal component analysis (PCA) plots. All the 53 arrays passed these criteria and were included in the analyses.

Student’s t tests (GeneSpring GX11.0) were used to identify genes differentially expressed between the VAT and the VAT groups in the pre-infection and infection periods at the level of significance p < 0.01.

Pathways and functional analyses were generated using Ingenuity Pathway Analysis 8.5 (IPA) (Ingenuity Systems, Redwood City, CA) software. For IPA analyses, only significantly differentially datasets obtained in the GeneSprin analyses were analyzed for identification of those canonical pathways differentially expressed between comparison groups. Hierarchical clustering of the genes participating of the signaling pathways differentially expressed between groups was performed by using GeneSpring GX 11.0.

Results

Eight mechanically ventilated patients were enrolled: five who developed VAT and three VAP. Ventilator-associated infection was diagnosed in all patients after the 4th day. None had received antibiotics before ICU admission or received selective digestive decontamination, and all had started empirical antibiotic therapy when infection was suspected. All but one patient who developed VAP were male. The mean age was 55.2 ± 22.1 years. Subjects were critically ill, with a mean ± SD APACHE III score at ICU admission of 75 ± 27. Descriptive data of the cohort are shown in Table 1 (additional material available online Table 1).

The number of samples analyzed in each phase was: in the pre-infection period: 20 samples from 5 VAT patients and 12 samples from 3 VAP patients; in the infection period: 15 samples from 5 VAT patients and 9 samples from 3 VAP patients. To determine the potential influence of quantitative changes of leukocytes subpopulations on our results, we compared the average concentration (cells/mm³) of lymphocytes, neutrophils, eosinophils, basophils and monocytes between VAT and VAP groups both in the pre-infection and infection periods (Table 2) and found no statistical differences.
Figure 1  Complement system gene expression analysis. (a) IPA canonical pathway modelling of the complement system pathway in the pre-infection phase. Those genes with relative lower expression in VAP vs VAT are depicted in green. (b) One-way hierarchical clustering of those genes selected by IPA (red, upregulated; blue, downregulated) in the pre-infection and infection phases in VAP and VAT.
Comparison of VAP/VAT gene expression "signatures" using genomewide oligonucleotide microarrays

Table 1: Clinical characteristics of the patients included according to the diagnosis onset.

| Diagnosis onset (days) | Group | BMI (kg/m²) | Age (years) | Sex | APACHE II score | SAPS II | Diagnosis | Pathogen |
|-----------------------|-------|-------------|-------------|-----|----------------|---------|-----------|----------|
| 5                     | VAT   | 31.57       | 42          | Male| 4              | 36      | Stroke    | N/A      |
| 6                     | VAT   | 27.48       | 49          | Male| 5              | 38      | Multiple traumas | S. pneumoniae |
| 7                     | VAT   | 27.77       | 77          | Male| 15             | 72      | Stroke    | H. influenzae |
| 5                     | VAT   | 35.66       | 59          | Male| 2              | 10      | Stroke    | H. ariaeus |
| 7                     | VAT   | 35.66       | 77          | Male| 10             | 52      | Stroke    | N/A      |
| 7                     | VAP   | 35.66       | 60          | Male| 10             | 52      | Stroke    | N/A      |
| 5                     | VAP   | 20.22       | 25          | Female| 10           | N/A     | Stroke    | N/A      |
| 7                     | VAP   | 27.33       | 77          | Male| 10             | 52      | Stroke    | N/A      |
| 7                     | VAP   | 27.33       | 77          | Male| 10             | 52      | Stroke    | N/A      |
| 7                     | VAP   | 20.22       | 25          | Female| 10           | N/A     | Stroke    | N/A      |
| 7                     | VAP   | 27.33       | 77          | Male| 10             | 52      | Stroke    | N/A      |

Table 2: Median concentration and interquartile range (cells/mm³) of monocytes, neutrophils, lymphocytes, eosinophils and basophils between VAT and VAP groups both in the pre-infection and post-infection periods.

| Pathogen | Median | IQR |
|----------|--------|-----|
| Monocytes | VAT pre-infection | 592.8 | 457.6 |
| Neutrophils | VAT pre-infection | 7833.9 | 3614.2 |
| Lymphocytes | VAT pre-infection | 1001.3 | 552.5 |
| Eosinophils | VAT pre-infection | 22.1 | 58.4 |
| Basophils | VAT pre-infection | 31.7 | 29.9 |

Microarrays analysis

Comparison of gene expression profiles in the pre-infection period revealed 5595 genes differentially expressed between VAP and VAT (p value <0.01, fold change >2). IPA analysis identified a significant depression of the complement system pathway in the VAT group, affecting to the classical pathway along with the final common pathway (p < 0.05) (Fig. 1). Genes found to be differentially expressed in this route were C6, C9, C1QA, C1QB, C1QC, C1S, C3AR1, C4B, C4BPB, CD55, CD59, CR1, and MASP1 (Table 2). In addition, the cAMP and the calcium signalling pathways were also significantly depressed in the VAT group (Fig. 1, Suppl. Table 1) during the pre-infection phase also.

On the other hand, comparison of gene expression profiles during the infection period showed 8724 genes differentially expressed between VAP and VAT (p value <0.01, fold change >2). Although not significant differences were found between VAP and VAT for the complement signalling pathway, as showed by IPA, expression of 8 genes participating of this route were significantly depressed in the VAT group compared to the VAT one (Fig. 1). As in the previous phase, IPA evidenced a comparative depression of the cAMP signalling pathway in the VAT group (Suppl. Table 1).
Discussion

This is the first study to compare gene expression profiles of critically ill patients suffering from VAP and VAT. Gene expression profiles in the pre-infection period identified a significant depression of the complement system pathway in the VAP group compared to the VAT group, affecting to the classical pathway along with the final common pathway. This suggests that intubation by itself is associated with different degrees of immunocompromise that facilitates different infectious complications.

Surveillance cultures every 48–72 h in patients with ARDS, using either a protected specimen brush or BAL, have shown that tracheobronchitis can occur before the onset of VAP with tracheal colonization in only 56% of patients and was not predictive of VAP developing. The range of time from colonization to the development of VAP was 2–6 days. Conceivably, then, an as yet unidentified intermediate process may allow the bacterial burden to develop VAT before VAP, and this intermediate period may constitute an excellent time window in which to halt the progression.

Other data in mechanically ventilated patients suggest a similar dichotomy between upper and lower airway colonization patterns. As the upper airways have contact with pathogenic microbes, immune recognition principles have to be tightly controlled.

In our study we mainly observe a suppression of genes involved in the classical pathway and converge for common pathway of complement activation. Complement activation is a key component for maintaining internal inflammatory homeostasis. The complement system is a set of more than thirty proteins which serves as an important effector arm in the immune defense and contributes to the first-line host defense. It comprises an essential link between the innate and adaptive immune system, as its proteins identify invading bacteria and label them for phagocytosis or lysis.

Monocytes actively produce proteins of the complement system. In consequence, altered numbers of monocytes in peripheral blood could contribute to explain the observed differences in the gene expression profiles between groups. To determine the potential influence of quantitative changes of leukocytes subpopulations on our results, we compared the average concentration of lymphocytes, neutrophils, eosinophils, basophils and monocytes between VAT and VAP groups both in the pre-infection and infection periods and revealed no significant differences between groups. This supports that relative depression in the expression values of the genes associated to complement function in the VAP group compared to VAT in the pre-infection period should be potentially due to functional changes induced by the disease, and not to quantitative changes in leukocyte subpopulations.

Comparing the gene expression profiles of VAP and VAT, the CAMP signalling pathways were significantly depressed in the VAP group. During the development of the immune systems, CAMP regulates cell proliferation, differentiation and apoptosis. Recent studies have demonstrated that CAMP is necessary for T cell activation. In contrast, other studies have shown it to be a potent immunosuppres-

down-regulated the stimulatory molecules CD80 and CD86. Another pathway with differential gene expression was the calcium signalling pathway. Calcium is involved many biological and immune system functions such as T lymphocyte activation, the immunological synapse (IS) between T cells and antigen-presenting cells and phagocytosis. However, the depression of the CAMP and Ca pathway genes is difficult to interpret as almost every cell uses these pathways at the initiation of different signalling pathways and this effect should be elucidated in future studies.

In our study, patients who developed VAP had lower gene expression in the complement signalling pathways. In recent years, immunodulatory therapies in sepsis were driven by the assumption that the adverse outcomes were related to an overly exuberant inflammatory response. However, recent evidence on cytokine production in response to bacterial antigens, lymphocyte proliferation in response to recall antigens, and new antigen presentations as markers of the immune function has been published. Immunoanalysis should be taken into account as a continuum along which patients with VAP presented a less compartmentalised infection.

The present study has some strengths and weaknesses. First, a control population of patients matched by severity and diagnoses who underwent ventilation for equivalent times but did not develop VAP or VAT would be advisable. Unfortunately, lack of financial support precludes microarray analysis of controls. Therefore, our findings constitute a pilot study and needs to be validated before generalization, because selection bias cannot be excluded.

Second, VAP does not have gold standard. Indeed, there was a possible overlap in clinical signs between VAT and VAP, and ICU chest X-rays have limitations in sensitivity to identify opacities. Indeed, as Kollef suggested, episodes of VAT might constitute only airways colonization, or perhaps small pneumonia in some patients. CT scan was not implemented for diagnosis because this technique is expensive and is not part of standard care; in addition, transfers to CT scan in intubated patients present safety concerns. Moreover, other opacities such as atelectasis or ARDS cannot be ruled out. Indeed, Craven et al. have acknowledged this potential bias. The third major limitation of our study is that we only followed patients until infection onset. Cobb et al. reported that the onset of an infection-specific transcriptional program may precede the clinical diagnosis in patients who developed VAP based on the Riboleukogram. The main aim of that study was to determine whether VAP/VAT exhibits different gene expression signatures that might explain divergent pathways leading to VAT/VAP as a continuum between bronchitis and pneumonia in mechanically ventilated patients. Fourth, our findings cannot be generalized to pneumonia developing after one week of ventilation, but 80% of episodes of VAP present during the first 10 days. Finally, the sample size is relatively small and gene expression profiles between Gram-positive and Gram-negative pathogens could not be determined, but the number of samples obtained was sufficient to generate statistically significant differences with a large number of genes being functionally involved in the immune response. This is the first study to investigate the biological plausibility between VAT and VAP.
In summary, this study suggests that intubated patients with VAP show a relative depression of the genes involved in the complement system, cAMP, and calcium signalling pathways which all play a key role in the immune response to bacteria. The relative depression of these routes may lead to an impaired immunocompetence status, and contributes to explain why patients with VAP exhibit more inflammation and worse outcomes than those with VAT. Therefore, therapy of VAP should be more aggressive than VAT. Our findings suggest that VAP does not have an abrupt onset and risk of VAP might be anticipated by identification of the signature that precedes pneumonia, facilitating preemptive therapy. Moreover, this translational study offers insight to improve the understanding of pathogenesis of VAP vs VAT, with implications on prevention. Indeed, early development of immunocompromise in the post-intubation period suggests that not all episodes of VAP are preventable and "pneumonia zero" is not a realistic objective.

Conflict of interest
The authors declare that they have no conflict of interest.

Acknowledgments
We are indebted to Michael Maudsley (Universitat de Barcelona) for editorial assistance and language editing. We thank H. Goossens, Belgium, for critical review of the manuscript. This work was supported by CIBERes nodo 18 and ERMO07/050.

Appendix A. Supplementary data
Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.jmedin.2011.10.009.

References
1. Rello J, Ausina V, Castella J, Net A, Prats G. Nosocomial respiratory tract infections in multiple trauma patients. Influence of level of consciousness with implications for therapy. Chest. 1992;102:525–9.
2. Craven DE, Hjalmarson KI. Ventilator-associated tracheobronchitis and pneumonia: thinking outside the box. Clin Infect Dis. 2010;51 Suppl. 1:559–66.
3. Craven DE. Ventilator-associated tracheobronchitis (VAT): questions, answers, and a new paradigm? Crit Care. 2008;12:157.
4. Dallas J, Skrupky L, Abebe N, Boyle 3rd WA, Kollef MH. Ventilator-Associated Tracheobronchitis (VAT) in a mixed surgical and medical ICU population. Chest. 2010.
5. Nseir S, Di Pompeo C, Pronnier P, Beague S, Onimus T, Saulnier F. Nosocomial tracheobronchitis in mechanically ventilated patients: incidence, aetiology and outcome. Eur Respir J. 2002;20:1483–93.
6. Nseir S, Favory R, Jozefowicz E, Decamps F, Dewavrin F, Brunin G, et al., VAT Study Group. Antimicrobial treatment for ventilator-associated tracheobronchitis: a randomized, controlled, multicenter study. Crit Care. 2008;12:R62.
7. American Thoracic Society, Infectious Diseases Society of America. Guidelines for the management of adults with hospital-acquired, ventilator-associated, and healthcare-associated pneumonia. Am J Respir Crit Care Med. 2005;171:388–416.
8. Bernard GR, Artigas A, Brigham KL, Carlet J, Falke K, Hudson L. Report of the American-European consensus conference on ARDS: definitions, mechanisms, relevant outcomes and clinical trial coordination. The Consensus Committee. Intensive Care Med. 1994;20:225–32.
9. Kollef MH, Bock KR, Richards RD, Hears M. The safety and diagnostic accuracy of minibranchioalveolar lavage in patients with suspected ventilator-associated pneumonia. Ann Intern Med. 1995;122:743–8.
10. Delclaux C, Roupie E, Blot F, Brochard L, Lemaire F, Brun-Buisson C. Lower respiratory tract colonization and infection during severe acute respiratory distress syndrome: incidence and diagnosis. Am J Respir Crit Care Med. 1997;156:1092–8.
11. Ferreira VP, Pangburn MK, Cortés C. Complement control protein factor H: the good, the bad, and the inadequate. Mol Immunol. 2010;47:2187–97.
12. Morris Jr JA, Francois C, Olson PK, Cotton BA, Summar M, Jenkins JM, et al. Genetic variation in complement component 2 of the classical complement pathway is associated with increased mortality and infection: a study of 627 patients with trauma. J Trauma. 2009;66:1265–70.
13. Laarmann A, Milder F, van Strijp J, Rooijakkers S. Complement inhibition by gram-positive pathogens: molecular mechanisms and therapeutic implications. J Mol Med. 2010;88:115–20.
14. Walport MJ. Complement. First of two parts. N Engl J Med. 2001;344:1058–66.
15. Duan B, Davis R, Sadat EL, Collins J, Sternweiss PC, Yuan D, et al. Distinct roles of adenylyl cyclase VII in regulating the immune responses in mice. J Immunol. 2010;185:315–44.
16. Conche C, Boulla G, Trautmann A, Randriamampita C. T cell adhesion primes antigen receptor-induced calcium responses through a transient rise in adenosine 3′,5′-cyclic monophosphate. Immunity. 2009;30:33–43.
17. Fassbender M, Herter S, Holttappels R, Schild H. Correlation of dendritic cell maturation and the formation of aggregates of poly-ubiquitinated proteins in the cytosol. Med Microbiol Immunol. 2008;197:185–9.
18. Varga Z, Hajdu P, Panyi G. Ion channels in T lymphocytes: an update on facts, mechanisms and therapeutic targeting in autoimmune diseases. Immunol Lett. 2010;130:19–25.
19. Kummerow C, Junker K, Kruse K, Rieger H, Quintana A, Hoth M. The immunological synapse controls local and global calcium signals in T lymphocytes. Immunol Rev. 2009;231:132–47.
20. Korade-Mirnics Z, Corey SJ. Src kinase-mediated signaling in leukocytes. J Leukoc Biol. 2000;68:603–13.
21. Kollef MH. Review of recent clinical trials of hospital-acquired pneumonia and ventilator-associated pneumonia: a perspective from academia. Clin Infect Dis. 2010;51 Suppl. 1:529–35.
22. Craven DE, Chroneneu A, Zias N, Hjalmarson KI. Ventilator-associated tracheobronchitis: the impact of targeted antibiotic therapy on patient outcomes. Chest. 2009;135:521–8.
23. Cobb JP, Moore EE, Hayden DL, Minei JP, Cuschieri J, Yang J, et al. Validation of the riboleukogram to detect ventilator-associated pneumonia after severe injury. Ann Surg. 2009;250:531–9.
24. McDunn JE, Husain KD, Polpitiya AD, Burykin A, Ruan J, Li Q, et al. Plasticity of the systemic inflammatory response to acute infection during critical illness: development of the riboleukogram. PLoS One. 2008;3:e1564.
25. Kollef MH, Afessa B, Anzueto A, Veremakis C, Kerr KM, Margolis BD, et al., NASCENT Investigation Group. Silver-coated endotracheal tubes and incidence of ventilator-associated pneumonia: the NASCENT randomized trial. JAMA. 2008;300:805–13.
26. Koulenti D, Lisboa T, Brun-Buisson C, Krueger W, Macor A, Sole-Violan J, et al., EU-VAP/CAP Study Group. Spectrum of practice in the diagnosis of nosocomial pneumonia in patients requiring mechanical ventilation in European intensive care units. Crit Care Med. 2009;37:2360–8.