The COVID-19 Vaccine in Clinical Trials: Where Are We Now?

Hu-Dachuan Jiang1, Jing-Xin Li2, Peng Zhang2, Xiang Huo3*, Feng-Cai Zhu1,4,∗

1 School of Public Health, Southeast University, Nanjing 210009, China;
2 Vaccine Clinical Evaluation Department, Jiangsu Provincial Center for Disease Control and Prevention, Nanjing 210009, China;
3 Food Safety and Evaluation Department, Jiangsu Provincial Center for Disease Control and Prevention, Nanjing 210009, China;
4 NHC Key Laboratory of Enteric Pathogenic Microbiology, Jiangsu Provincial Center for Disease Control and Prevention, Nanjing 210009, China

Abstract
The coronavirus disease 2019 (COVID-19) pandemic caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) continues to scale up around the world. The development of an effective COVID-19 vaccine is of utmost importance. Most vaccine designs can be classified into three camps: protein based (inactivated vaccines, protein subunit, VLP and T-cell based vaccines), gene based (DNA or RNA vaccines, replicating or non-replicating viral/bacterial vectored vaccines), and a combination of both protein-based and gene-based (live-attenuated virus vaccines). Up to now, 237 candidate vaccines against SARS-CoV-2 are in development worldwide, of which 63 have been approved for clinical trials and 27 are evaluated in phase 3 clinical trials. Six candidate vaccines have been authorized for emergency use or conditional licensed, based on their efficacy data in phase 3 trials. This review summarizes the strengths and weaknesses of the candidate COVID-19 vaccines from various platforms, compares, and discusses their protective efficacy, safety, and immunogenicity according to the published clinical trials results.

Keywords: COVID-19; Efficacy; Immunogenicity; Safety; SARS-CoV-2; Vaccine clinical trial

Introduction
Coronavirus disease 2019 (COVID-19) pandemic caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) continues to scale up around the world. Over 105 million COVID-19 cases and 2.3 million deaths have been reported globally till 6 February, 2021, according to WHO.11 Approximately 40% to 45% of those infected with SARS-CoV-2 will remain asymptomatic12,13 and most people (about 80%) can recover from the disease without treatment.14,15 Thus, the actual number of COVID-19 infected cases is supposed to be much higher than what have been reported.16,17 Nevertheless, the level of antibody seropositivity in the general population was still low,18 indicating that most of the population in the world remain susceptible.

Most patients displayed an antibody response after infection with SARS-CoV-2,17 and reinfection incidents with valid evidence were few.19 Convalescent plasma transfusion has been indicated as an effective therapy against COVID-19.10 These evidences together highlighted the necessity and feasibility of COVID-19 vaccine development.

The SARS-CoV-2 genome contains four main structural proteins: the spike (S), membrane (M), envelope (E) and nucleocapsid (N) protein. The main target for antigen epitopes of COVID-19 vaccine is S protein:11 the S1 domain, which contains the receptor-binding domain (RBD) for the host cell receptor angiotensin-converting enzyme-2 (ACE2),12 the N-terminal domain (NTD), which has been proven as another site with potent neutralizing activity,13-15 and the S2 domain containing the fusion peptide.16,17 Currently, 237 candidate vaccines against SARS-CoV-2 are in development worldwide according to the survey of WHO.18 Among them, 63 vaccines have been approved for clinical trials and 27 are evaluated in phase 3 clinical trials. Up to now, six COVID-19 vaccines, including two mRNA vaccines, two inactivated vaccines, and two viral-vectored vaccines, have been authorized for emergency use or conditional licensed in some countries or regions, based on their efficacy data in phase 3 trials.18

Platforms for COVID-19 candidate vaccines
Currently, COVID-19 candidate vaccines can be classified into three camps: protein based (inactivated vaccines, protein subunit, VLP and T-cell based vaccines), gene based (DNA or RNA vaccines, replicating or non-replicating viral/bacterial vectored vaccines), and a combination of both protein-based and gene-based (live-attenuated virus vaccines).19 Most of the COVID-19 candidate vaccines underdevelopment belong to the first two camps.

Protein based vaccines
Many of the vaccines in clinical use today fall into this category. This approach utilizes the entire or a part of the pathogen as
antigen to elicit protective immune responses. This type of vaccine is generally well tolerated, safe and can be used to most people, even the elderly or people with immunodeficiency.[20,21] Besides, the immunogenicity of vaccines from protein-based platform were stable, fluctuates very little due to the stability of the protein content. At the time of writing, the number of protein subunit vaccines is the most among the candidate vaccines being clinically studied (n=21). There are ten inactivated vaccines evaluated in clinical trials, and six of which have been evaluated at phase 3 study.

In the production of inactivated vaccines, conserving the viral antigen of high quality is the key to induce protective immunity.[21] In general, inactivated vaccines are highly immunogenic. However, in the case of COVID-19, the inactivated vaccine’s immunogenicity could be jeopardized by the immune evasion capacity of SARS-CoV-2, such as “Glycan shield” and the lying-down of RBD.[22–25] In addition, non-neutralizing epitopes contained in the inactivated whole virus might be able to induce high level of non-neutralizing antibodies, which have the potential to cause antibody dependent enhancement (ADE).

Protein subunit or VLP designed vaccines may address the concern of ADE, by removing as much non-neutralizing epitopes as possible, however, this would be accompanied by a long production time.[26] Furthermore, protein subunit vaccine can hardly induce cellular immune response with the traditional adjuvant, and thus an appropriate adjuvant may in need.[27]

**Gene based vaccines**

Gene based vaccines have the potential to elicit broad immune responses and are easier to achieve mass production compared with protein-based vaccines.[28] To date, there are 11 non-replicating viral vectored vaccines (4 at phase 3), five replicating viral vectored vaccines, seven RNA-based vaccines (3 at phase 3), and eight DNA-based vaccines (3 at phase 3) are evaluated in clinical trials.

The RNA or DNA-based vaccine could be swiftly advanced towards the targeted antigen and be manufactured massively. Therefore, even it’s a relative new platform for vaccine development, and never been approved for marketing before this pandemic, this platform is expected to contribute to accelerating COVID-19 vaccine development. However, the efficacy of an RNA or DNA-based vaccine in human strongly depends on its formulation and the delivery system for introducing the target genes into cells, which may vary a lot in different individuals.

Viral vectored vaccines deliver the target gene into the cells for compilation and expression through an infectious attenuated vector virus. For replication-incompetent vectored vaccines, the pre-existing immunity to the vectors could affect the efficiency of deliver significantly, with reduced the vaccine-induced immune responses in those with pre-exposure to vector at baseline. While, using replication-competent adenovirus as a vector would raise a safety concern that the vector virus may recombine or revert to a parental or wild-type phenotype at a low frequency, or cause clinical infections in some immunocompromised populations.[29]

Though most of the viral vectored vaccines underdevelopment are administrate intramuscularly, there is a potential advantage for this type vaccine to be is given by inhalation or intranasal administration. Intranasal vaccination is supposed to provide a better protection compared to subcutaneous inoculation in terms of respiratory pathogens, due to the ability of inducing high level of specific IgA antibodies.[30] Some vaccines that induce mucosal immunity against SARS-CoV-1 and MERS-CoV have succeeded in producing IgA in the respiratory tract, preventing corresponding viral dissemination to the lung.[31] As ACE2 expression is abundant in nasal epithelial cells of human upper respiratory tract, and nasal goblet cells and ciliated cells might be the initial infection sites of SARS-CoV-2.[18,23] Four intranasal vaccines were developed and evaluated in clinical trials.[18]

**The results of clinical trials on COVID-19 vaccines**

Up to now, a total of ten vaccine candidates have reported the results of human clinical trials, including four non-replicating viral vectored vaccines, four inactivated vaccines, two mRNA vaccines and two protein subunit vaccines [Table 1].

**Safety**

The accumulated safety data from clinical trials shows that candidate COVID-19 vaccines from different platforms are generally safe and tolerable, but with distinct safety profiles. Viral vectored vaccines and mRNA vaccines were associated with increased adverse reactions including fever, fatigue, headache, myalgia compared with inactivated vaccines. Significantly, people aged 18 to 55 reported more adverse reactions than that in elderly.

Around 22% of the recipients of chimpanzee adenovirus-vectored vaccine (ChAdOx1 nCoV-19) reported severe adverse reactions in phase 1 clinical trial, in which participants received only single dose, in phase 2/3 trial however, all people administrated two doses, none of whom reported adverse reactions. Besides, all the vaccines of the recombinant adenovirus type 26 vector vaccine and recombinant adenovirus type 5 vector vaccine (rAd26-S+rAd5-S nCoV-19) had fever.[34–37]

Of note, mRNA-1273 caused pain in all vaccine recipients aged 18 to 55 and severe adverse reaction was reported by 6.7% participants between the ages of 18 to 55 years, and 5.3% participants over 55 years old.[38,39] For another mRNA vaccine BNT162b2, 8.0% participants between the ages of 18 to 55 years reported severe adverse reaction.[40]

Protein subunit (Matrix-M1 adjuvanted NVX-CoV2373 and SCB-2019) also induced some adverse reactions, especially fatigue and pain, and 11.4% of the recipients of NVX-CoV2373 developed severe adverse reactions.[41,42]

Comparing to above mentioned vaccines, alum-adjuvanted inactivated vaccines and INO-4800 generated the least adverse reactions with only a small proportion of participants having mild to moderate adverse reactions, probably because of the limited capacity of producing cellular immunity.[43–44] However, it is worth noting that some individuals had abnormal increase of blood glucose, or facial neuritis after vaccination. The relationship between these abnormal changes and inactivated vaccines is uncertain yet, which needs to be further investigated [Figure 1; Supplementary Table 1; http://links.lww.com/IDI/A1].

**Immunogenicity**

Humoral immune responses in terms of S- or RBD-binding antibodies measured by glycoprotein-specific enzyme-linked immunosorbent assay (ELISA), and neutralizing antibodies (Nab) to live SARS-CoV-2 virus or pseudo virus were measured in clinical trials.
| Vaccine platform | Vaccine | Phase | Age  | Study design                        | Location       | Sample size | Number of doses | Schedule | Administration | Doses           |
|------------------|---------|-------|------|-------------------------------------|----------------|-------------|-----------------|----------|----------------|-----------------|
| Non-replicating viral vectored | Ad5-vectored COVID-19 | 2     | ≥18  | Randomised, double-blind, placebo-controlled | Wuhan, China  | 508         | 1               | –        | IM             | 0.5 or 1 × 10^{11} VP |
|                   | ChAdOx1 nCoV-19 | 1/2   | 18–55| Randomised, single-blind, placebo-controlled | UK             | 1077        | 0 or 1          | 0, 28 days | IM             | 0.5 × 10^{11} VP   |
|                   | rAd26-S+rAd5-S COVID-19 | 1/2   | 18–60| Open, non-randomised                | Moscow, Russia | 76          | 2               | 0, 21 days | IM             | 1 × 10^{11} VP     |
|                   | Ad26.COV2.S | 1/2   | 18–55| Randomised, multi-center, placebo-controlled | US             | 1045        | 0 or 1          | 0, 56 days | IM             | 0.5 or 1 × 10^{11} VP |
| Inactivated       | Inactivated whole-virus COVID-19 vaccine | 1, 2  | 18–59| Randomised, single-blind, placebo-controlled | Henan, China | 224         | 2               | 0, 14 or 0, 21 days | IM | 2.5, 5, 10 μg |
|                   | BBIBP-CorV | 1, 2  | ≥3   | Randomised, single-blind, placebo-controlled | Henan, China | 448         | 2               | 0, 14 or 0, 21 or 0, 28 days | IM | 2, 4, 8 μg |
|                   | CoronaVac  | 2     | 18–59| Randomised, double-blind, placebo-controlled | Jiangsu, China | 600        | 2               | 0, 14 days or 0, 28 days | IM | 3, 6 μg |
|                   | BBV152    | 1, 2  | 12–65| Double-blind, randomised, controlled | India          | 755         | 2               | 0, 14 days | IM             | 3, 6 μg          |
| mRNA             | mRNA-1273 | 1     | 18–55| Dose-escalation, open-label          | Seattle and Atlanta, US | 45          | 2               | 0, 28 days | IM             | 25, 100, 250 μg   |
|                   | mRNA-1273 | 1     | ≥56  | open-label, dose-ranging            | US             | 120         | 2               | 0, 28 days | IM             | 25, 100 μg       |
|                   | BNT162b1  | 1/2   | 18–55| Randomised, dose-escalation, single-blind, placebo-controlled | Germany | 45          | 2               | 0, 28 days | IM             | 10, 30, 100 μg   |
| DNA              | INO-4800  | 1     | 18–55| open-label, multi-center            | US             | 20          | 2               | 0, 4 weeks | ID             | 1.0, 2.0 mg      |
| Protein subunit  | INO-Cov2373 | 1/2   | 18–59| Open, non-randomised                | Australia      | 76          | 2               | 0, 21 days | IM             | 5, 25 μg         |
|                   | SGB-2019  | 1     | 18–54, 55–75 | Randomised, double-blind, placebo-controlled | Australia    | 150         | 2               | 0, 21 days | IM             | 3, 9, 30 μg      |

ID: Intradermal; IM: Intramuscular; NA: No data; VP: Virus particle.
Protein subunit (NVX-CoV2373 and SCB-2019) induced the strongest antibodies levels with the GMT of 3906 and 1810, respectively, by microneutralization assay (MNA).\(^{[41,42]}\) mRNA vaccines also generated good specific antibodies. GMTs of Nab were 654.3, 361 and 102.3 elicited by mRNA-1273 (by PRNT80), BNT162b2 (by MNA50).\(^{[38,39]}\) The humoral immunity elicited by non-replicating viral vectored vaccines, inactivated vaccines and DNA vaccine was comparable, and was minor than that of other types of vaccines. The detected GMTs of Nab ranged from 27.6 (GMTs by MNA50) to 300 (GMTs by PRNT50) for inactivated vaccines and DNA vaccine, and ranged from 18.3 (GMTs by MNA50) to 827 (median by MNA50) for non-replicating viral vectored vaccines.\(^{[35–37,43–47]}\) The Nab titers of 827 was induced by Ad26.COV2.S, evidently higher than other non-replicating viral vectored vaccines.\(^{[48]}\)

Since there are no standardized methods for these serological tests, these published data of the antibody tests in clinical trials are impossible to compare across the different studies. Nevertheless, a panel of convalescent serum from COVID-19 patients was provided as active competitors in most studies, and we can get some comparative information from these data.

The mRNA vaccine BNT162b2 showed better humoral responses, 1.7 to 4.6 times higher for Nab results compared with convalescent patients after vaccination, and Ad26.COV2.S also generated ELISA antibodies and Nab titers of 1.9 and 1.0 times higher than human convalescent serum. The rAd26-S+rAd5-S, mRNA-1273 and INO-4800 vaccination group presented a higher ELISA titres or Nab results than detected in convalescent patients. The NVX-CoV2373 and SCB-2019 vaccine induced approximately four times greater than that in outpatients for Nah, and also resulted in similar GMT levels of ELISA antibodies and Nab compared with hospitalized patients. The Nab titres after vaccination of inactivated vaccine were lower in participants than that was detected in convalescent serum from patients who has previously had COVID-19.

To sum up, the protein subunit vaccine performed notably best, followed by the mRNA vaccines and the non-replicating Chimpanzee adenovirus and Ad26 vectored vaccines, which induced relatively high humoral immune response than inactivated vaccines, DNA vaccine and Ad5-vectored vaccines. The differences in disease severity, age, and sampling time points post-infection could affect the level of antibody titer of the panel of convalescent serum, so the comparisons still have some uncertainty.

According to the reported data in terms of the Nab, two doses administration were preferred for most of the candidate COVID-19 vaccines, in order to induce more satisfying antibody responses than one shot did. Nevertheless, the two viral vectored vaccines (Ad26 and Ad5) were evaluated in phase 3 trials with one shot regimen for efficacy estimation, expecting to generate acceptable protection for COVID-19 after one dose.

As for cellular immunity, the protein subunit vaccine (NVX-CoV2373), one of the mRNA vaccines (mRNA-1273), Ad26.COV2.S and SCB-2019 induced significant CD4+ T-cell responses, especially Th1, and three non-replicating viral vectored vaccines (ChAdOx1 nCoV-19, Ad5-vectored and rAd26-S+rAd5-S COVID-19 vaccine) and DNA vaccine induced significant interferon-γ response. Whereas, T-cell response induced by inactivated vaccines was relatively weak [Figure 2; Supplementary Table 2; http://links.lww.com/IDI/A1].

**Protective efficacy**

Either laboratory-confirmed COVID-19 or laboratory-confirmed SARS-CoV-2 infection is an acceptable primary endpoint for a COVID-19 vaccine efficacy trial, and the estimate of the primary
efficacy endpoint should be at least 50% for a placebo-controlled efficacy trial. In addition, Food and Drug Administration requests that a total of five or more severe COVID-19 cases should be observed in the placebo group, so as to assess the efficacy and VED sufficiently. Currently, the interim analysis results of the efficacy data from the phase 3 clinical trials of eight COVID-19 vaccines have been reported and shown efficacies ranged from 50.4% to 95% against COVID-19. However, there are many factors influencing the efficacy estimation, such as primary endpoints, case monitoring system, and the definition of confirmed cases, all of which are different used in each vaccine. BNT162b2 was 95% effective in preventing confirmed symptomatic COVID-19 with onset at least 7 days after the second dose in participants, and similar efficacy (generally 90 to 100%) was observed across subgroups defined by age, sex, ethnicity, baseline body-mass index, and the presence of coexisting conditions. In this efficacy trials, a total of 170 cases were occurred, only eight in vaccine group, and ten severe cases occurred after the first dose, nine in placebo group and one in BNT162b2 group. The efficacy of mRNA-1273 was 94.1% in preventing a first occurrence of symptomatic COVID-19 with onset at least 14 days after the second injection in per-protocol population, with 196 cases being observed (11 in vaccine group), of which ten were severe COVID-19 cases in the placebo group. In terms of secondary analyses, including analyses in participants who had evidence of SARS-CoV-2 infection at baseline, and analyses in participants aged over 65 years old, the efficacy was similar. As for SPUTNIK V, PCR confirmed COVID-19 from day 21 after receiving the first dose occurred in 62 placebo recipients including 10 moderate or severe cases and in 16 vaccine recipients, so the efficacy was shown 91.6%. An interesting phenomenon happened to the ChAdOx1 nCoV-19 in phase 3 study, in which the primary outcome was virologically confirmed, symptomatic COVID-19 after last vaccination. In participants who received two standard doses, vaccine efficacy was 62.1% and in participants who received a low dose followed by a standard dose, efficacy was 90.0%. All the cases hospitalized for COVID-19 or severe cases were occurred in the control arm. Also, Janssen announced the efficacy results of 66% in preventing confirmed moderate to severe COVID-19 14 days after single dose. The most immunogenic vaccine, Novavax, demonstrated 89.3% efficacy, based on 62 cases, of which 56 cases (including one severe case) were observed in the placebo group. Two inactivated vaccines were shown the efficacy of 79.3% and 91.3% (in Turkey), 50.4% (in Brazil), respectively. More details are shown in Table 2.

Outlook

The most efficient approach to halt the pandemic is to achieve herd immunity with a valid COVID-19 vaccine. The history of vaccine development tells us that not all the vaccine candidates would succeed, particularly for a novel emerging respiratory virus. Thus, the more candidates we test, the bigger chance we gain to have safe and efficacious vaccines against COVID-19. Up to now, at least nine candidate COVID-19 vaccines from different platforms evaluated in clinical trials, appeared to be safe, and able to elicit significant immune responses. Of them, eight COVID-19 vaccines had their preliminary efficacy data being released, meeting the minimum requirement of 50%
efficacy, and were authorized for emergency use or conditional licensed in some countries or regions. Since a massive immunization campaign of the effective vaccine is implementing, we are expecting a slowing down of the COVID-19 epidemic later this year. However, there are still a lot scientific questions remain to be answered, including durability of vaccine-induced immune responses, safety and rare severe adverse reactions, vaccine effectiveness and its clinical evaluation and correlates of protection.

The duration of the antibodies following natural infection with SARS-CoV-2 has not been fully understood, although some researches demonstrated that the specific antibody level could be stable for at least 3 to 4 months. Some experts believe that a vaccine could provide stronger and more durable immune response than a natural infection. One reason is that the vaccine could be designed to contain highly-concentrated antigens, which could give an answer to the duration of vaccine-induced immunity.

Another highly controversial issue about the safety of the COVID-19 vaccine is ADE. ADE is commonly identified in vivo or animal models, which mainly occurs in flavivirus, coronavirus, respiratory virus and arthropod-borne viruses, but the fact in humans can be exemplified only in dengue viruses with clinical, epidemiological, biological, or pathological evidence. To address this issue, systematic evaluation endpoints have been put forward to measure ADE, including Nab versus binding antibodies (low Nab titer, low ratio of Nab to total binding antibody, low affinity of IgG antibody binding to RBD receptor), cellular immunity (low CD4+ but high CD8+ proliferative responses, CD4 T-cell responses biased toward expression of Th2 cytokines), inflammatory reactions (IL-1, IL-6, IL-8, TNF, IFN-I increased) and immunopathology (eosinophilic, Th2 cytokines IL4, IL5, IL10, IL13 increased). The lack of a standardized serum antibody and available evidence on immunoassays being correlated to functional/neutralization assays or to clinical protection is another hurdle in clinical trials to evaluate the immunogenicity of vaccine candidates. In recent published clinical studies, researchers tend to compare Nab levels between vaccines and convalescent COVID-19 patients. However, antibody response varies by time and between convalescent patients, with severe or older patients having higher antibody titers. Hence, it is important to clearly state the sampling time and clinical severity of convalescent patients in clinical studies.

In addition to humoral immunity, cellular immunity and local mucosal immunity play important roles in protection against SARS-CoV-2 infection as well. Mucosal immunity is critical in the prevention of respiratory infection, such as influenza, respiratory syncytial virus and pneumococcal. A single mucosal inoculation of Ad5-nCoV could induce better protection

### Table 2: The efficacy results from published clinical studies

| Vaccine                | Number of vaccines | Targeted dose | Immunization procedure | Time for observation | Number of COVID-19 cases | Estimate of efficacy | Severe COVID-19 cases |
|------------------------|--------------------|---------------|------------------------|----------------------|-------------------------|----------------------|-----------------------|
| BNT162b2               | 43,661             | 30 μg         | 0-21 days apart        | 7 days after two doses | 170 (8 in vaccine group) | 95%                  | 10 (1 in vaccine group) |
| mrNA-1273              | 30,000             | 100 μg        | 0-28 days apart        | 14 days after two doses | 196 (11 in vaccine group) | 94.1%                | 30                    |
| ChAdOx1 nCoV-19        | 22,690             | 2.55 \times 10^{10} / 5 x 10^{15} vp | 0-28 days apart | 14 days after two doses | 131 (30 in vaccine group) | low dose+standard dose: 90%; Two standard dose: 62%; Overall: 70% | 10 cases hospitalised (2 severe cases) all in placebo group |
| Ad26.COV2.S            | 34,000             | 5 \times 10^{10} vp | 0-21 days apart | 14 days after two doses | NA | 66% | NA |
| SPUTNIK V              | 22,714             | 1 \times 10^{11} vp | 0-21 days apart | 14 days after first dose | 78 (16 in vaccine group) | 91.6% | 20 |
| Inactivated whole-virus nCoV-19 vaccine | 60,000             | 5 μg         | 0-21 days apart        | 14 days after two doses | NA | 79.3% | NA |
| CoronaVac              | 7371               | 3 μg         | 0-14 days apart        | 14 days after two doses | NA | 91.25% (in Turkey); 50.38% (in Brazil) | NA |
| Novavax                | 15,000             | 5 μg         | 0-21 days apart        | 7 days after two doses | 62 (56 in vaccine group) | 89.3% | 1 in placebo group |

NA: Not available.
than intramuscular vaccination for the upper and lower respiratory tracts against SARS-CoV-2 challenge in mice and ferrets. These findings suggest that a COVID-19 vaccine that can not only induce humoral immunity but also a protective T cell response and mucosal immunity may maximize the protection against SARS-CoV-2, which should be investigated in future studies.

Mutations in spike protein, especially the RBD predicting conformational changes in the S1 domain, may compromise the efficacy of vaccines. Unfortunately, there has been evidence that the neutralization of antibody generated by two RNA vaccines (mRNA-1273 and BNT162b2) might be reduced caused by some variants, but whether the vaccine protective efficacy being influenced should be further investigated, and a close monitor on the viral evolution should be continued and amplified.

The granting of emergency use designation to candidate vaccines and licensed vaccines being available raise some issues. In some countries and regions with intensive COVID-19 vaccination campaign, a randomized, double-blind placebo-controlled phase 3 efficacy clinical trial is tough to develop and maintain. Investigators might be requested to unmask trial subjects to guarantee that those who received placebo are offered or actively seek approved candidate vaccines. Also, risk-benefit profile of normal placebo-controlled trial will be unacceptable, and the compliance of the trial can be impacted by drop-outs or “contamination”, alternative strategies to evaluate those vaccines are needed. Head-to-head comparative design, stepped-wedge design and cross-over design are suggested as alternative study design to avoid ethical issue as all people in the trial are offered protective vaccine, but at a considerable cost to efficiency and benefit. If possible, a serological correlate of protection and an immunological surrogate endpoint are expected to be justified by scientific evidence.

At the present, vaccine efficacy results are just the relatively short-term data, and the durability of protection needs to be observed. Also, the number of severe cases observed in trials were still limited, in order to obtain a solid vaccine efficacy for severe cases, more severe COVID-19 cases need to be captured in the continuing surveillance of phase 3 trials. Furthermore, the protective efficacy data of the inactivated vaccines is mainly in 18 to 59 adults, and more data of other populations should be collected to support the vaccine efficacy. Since the emergency use authorization and conditional licensure are not full licensures, head-to-head comparative design, stepped-wedge design and cross-over design are suggested as alternative study design to avoid under-ascertainment in infectious disease datasets: a comparison of methods. BMC Public Health 2014;14:147.

References

[1] World Health Organization. Coronavirus disease (COVID-19) situation report; 2021. Available from: https://www.who.int/emergencies/diseases/novel-coronavirus-2019/situation-reports. Accessed February 6, 2021.

[2] Jeyanathan M, Afkhami S, Fiona S, et al. Immunological considerations for COVID-19 vaccine candidates. Nat Rev Immunol 2020;20(10):615–632. doi: 10.1038/s41577-020-00434-6.

[3] Oran DP, Topol EJ. Prevalence of asymptomatic SARS-CoV-2 infection: a narrative review. Ann Intern Med 2020;173(3):362–367. doi: 10.7326/0003-4819-170-5-202003021-00526.

[4] World Health Organization. What happens to people who get seriously ill? Available from: https://www.who.int/news-room/q-a-detail/q-a-corona-viruses. Accessed October 15, 2020.

[5] Cheryl LG, Marie JM, Dietrich P, et al. Measuring underreporting and under-ascertainment in infectious disease datasets: a comparison of methods. BMC Public Health 2014;14:147. doi: 10.1186/1471-2458-14-147.

[6] Russell TW, Golding N, Hellewell J, et al. Reconstructing the early global dynamics of under-ascertained COVID-19 cases and infections. BMC Med 2020;18(1):332. doi: 10.1186/s12916-020-01790-9.

[7] European Centre for Disease Prevention and Control. Immune responses and immunity to SARS-CoV-2. Available from: https://www.ecdc.europa.eu/en/covid-19/latest-evidence/immune-responses. Accessed October 15, 2020.

[8] Anand S, Montez-Rath M, Han J, et al. Prevalence of SARS-CoV-2 antibodies in a nationwide sample of patients on dialysis in the USA: a cross-sectional study. Lancet 2020;396(10259):1335–1344. doi: 10.1016/S0140-6736(20)30229-2.

[9] Iwasaki A. What reinfections mean for COVID-19. Lancet Infect Dis 2021;21(1):3–5. doi: 10.1016/S1473-3099(20)30783-0.

[10] Liu SIH, Lin HM, Bame I, et al. Comoviral plasma treatment of severe COVID-19: a propensity-score-matched control study. Nat Med 2020;26(11):1708–1713. doi: 10.1038/s41591-020-1088-9.

[11] Watanabe Y, Allen JD, Wrapp D, et al. Site-specific glycan analysis of the SARS-CoV-2 spike. Science 2020;369(6501):330–333. doi: 10.1126/science.abb9983.

[12] Hoffmann M, Kleine-Weber H, Schroeder S, et al. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. Cell 2020;181(2):271–280. doi: 10.1016/j.cell.2020.02.052.

[13] Zhang L, Cao L, Gao XS, et al. A proof of concept for neutralizing antibody-guided vaccine design against SARS-CoV-2. bioRxiv doi: https://doi.org/10.1101/2020.09.23.309294.

[14] Chi X, Yan R, Zhang J, et al. A neutralizing human antibody binds to the N-terminal domain of the Spike protein of SARS-CoV-2. Science 2020;369(6504):650–655. doi: 10.1126/science.abe9592.

[15] Liu L, Wang P, Nair MS, et al. Potent neutralizing antibodies against multiple epitopes on SARS-CoV-2 spike. Nature 2020;584(7821):450–456. doi: 10.1038/s41586-020-2571-7.

[16] Wrapp D, Wang N, Corbett KS, et al. Cryo-EM structure of the 2019-nCoV spike in the prefusion conformation. Science 2020;367(6483):1260–1263. doi: 10.1126/science.abb2307.

[17] Walls AC, Park YJ, Tortorici MA, et al. Structure, function, and antigenicity of the SARS-CoV-2 spike glycoprotein. Cell 2020;181(2):281–292. doi: 10.1016/j.cell.2020.02.058.

[18] World Health Organization. Draft landscape of COVID-19 candidate vaccines. Available from: https://www.who.int/publications/m/item/draft-landscape-of-covid-19-candidate-vaccines. Accessed January 29, 2021.

[19] Abbasi J. COVID-19 and mRNA vaccines—first large test for a new approach. JAMA 2020;324(12):1125–1127. doi: 10.1001/jama.2020.16866.

[20] Vartak A, Suchek SJ. Recent advances in subunit vaccine carriers. Vaccines (Basel) 2016;4(2):12. doi: 10.3390/vaccines4020012.

[21] Delrieu I, Verzele D, Madder A, et al. Inactivated virus vaccines from chemistry to prophylaxis: merits, risks and challenges. Expert Rev Vaccines 2012;11(6):695–719. doi: 10.1586/erv.12.38.

[22] Watanabe Y, Bowden TA, Wilson IA, et al. Exploitation of glycosylation in enveloped virus pathobiology. Biochim Biophys Acta Gen Subj 2019;1863(10):1480–1497. doi: 10.1016/j.bbadgen.2019.05.012.

[23] Vankadari N, Wilce JA. Emerging COVID-19 coronavirus: glycan shield and structure prediction of spike glycoprotein and its interaction with human CD26. Emerg Microbes Infect 2020;9(1):601–619. doi: 10.1016/j.evimi.2020.01.002.

[24] Shang J, Wan YS, Luo C. Cell entry mechanisms of SARS-CoV-2. Proc Natl Acad Sci U S A 2020;117(21):11727–11734. doi: 10.1073/pnas.2003138117.

Author Contributions

Feng-Cai Zhu, Xiang Huo and Jing-Xin Li contributed to critical review and revision of the manuscript. Hu-Dachuan Jiang drafted of the manuscript. Peng Zhang contributed to the literature search.

Conflicts of Interest

None.
Amanat F, Krammer F. SARS-CoV-2 vaccines: status report. Immunity 2020;52(4):583–589. doi: 10.1016/j.immuni.2020.03.007.

Karch CP, Burkhard P. Vaccine technologies: from whole organisms to rationally designed protein assemblies. Biochem Pharmacol 2016;120:1–14. doi: 10.1016/j.bcp.2016.05.001.

Rauch S, Jasny E, Schmidt KE, et al. New vaccine technologies to combat outbreak situations. Front Immunol 2018;9:1963. doi: 10.3389/fimmu.2018.01963.

U.S. Department of Health and Human Services Food and Drug Administration. Chemistry, manufacturing, and control (CMC) information for human gene therapy Investigational New Drug applications (INDs), guidance for industry. Available from: https://www.fda.gov/vaccines-blood-biologics/guidance-compliance-regulatory-information-biologics/biologics-guidances. Accessed September 20, 2020.

Halifa SA, Gauthier L, Arpin D, et al. Nanoparticle-based vaccines against respiratory viruses. Front Immunol 2019;10:22. doi: 10.3389/fimmu.2019.00022.

Fierros LM, Silva IG, Mendoza SR. Development of SARS-CoV-2 vaccines. Biotechnol J 2020;15(20):e1900421. doi: 10.1002/biot.201900421.

Sadoff J, Gubarev AV, Shukarev G, et al. Updated interim results of a Phase 1–2a trial of Ad26.COV2.S Covid-19 vaccine. N Engl J Med 2021;384:384. doi: 10.1056/NEJMoa2034201. Online ahead of print.

World Health Organization. WHO target product profiles for COVID-19 vaccines. Available from: https://www.who.int/who-documents-database/who-target-product-profiles-for-covid-19-vaccines. Accessed October 2, 2020.

Food and Drug Administration. Development and licensure of vaccines to prevent COVID-19 guidance for industry. Available from: https://www.fda.gov/regulatory-information/search-fda-guidance-documents/development-and-licensure-vaccines-prevent-covid-19. Accessed October 2, 2020.

World Health Organization. WHO target product profiles for COVID-19 vaccines. Available from: https://www.who.int/who-documents-database/who-target-product-profiles-for-covid-19-vaccines. Accessed October 2, 2020.

Sadoff J, Gubarev AV, Shukarev G, et al. Updated interim results of a Phase 1–2a trial of Ad26.COV2.S Covid-19 vaccine. N Engl J Med 2021;384:384. doi: 10.1056/NEJMoa2034201. Online ahead of print.

World Health Organization. WHO target product profiles for COVID-19 vaccines. Available from: https://www.who.int/who-documents-database/who-target-product-profiles-for-covid-19-vaccines. Accessed October 2, 2020.
[64] Halstead SB, Chow JS, Marchette NJ. Immunological enhancement of dengue virus replication. Nature 1973;243:24–26.

[65] Sridhar S, Luedtke A, Langevin E, et al. Effect of dengue serostatus on dengue vaccine safety and efficacy. N Engl J Med 2018;379(4):327–340. doi: 10.1056/NEJMo1800820.

[66] Wilder-Smith A, Ooi EE, Horstick O, et al. Dengue. Lancet 2019;393(9918):350–363. doi: 10.1016/S0140-6736(18)32560-1.

[67] Moore JP, Klasse PJ. COVID-19 vaccines: “warp speed” needs mind melds, not warped minds. J Virol 2020;94(17):e01083-e1120. doi: 10.1128/JVI.01083-20.

[68] Chuang AT, Garcia-Carreras B, Hitchings MDT, et al. A systematic review of antibody mediated immunity to coronaviruses: antibody kinetics, correlates of protection, and association of antibody responses with severity of disease. medRxiv 2020;2020.04.14.20065771.

[69] Chandrasekhar A, Liu J, Martinot AJ, et al. SARS-CoV-2 infection protects against rechallenge in rhesus macaques. Science 2020;369(6533):812–817. doi: 10.1126/science.abc4776.

[70] Yu J, Tostanoski LH, Peter L, et al. DNA vaccine protection against SARS-CoV-2 in rhesus macaques. Science 2020;369(6533):806–811. doi: 10.1126/science.abc6284.

[71] Holmgren J, Czerkinsky C. Mucosal immunity and vaccines. Nat Med 2003;11(4 Suppl):S45–S53. doi: 10.1038/nm1213.

[72] Mazzur NL, Higgins D, Nunes MC, et al. The respiratory syncytial virus vaccine landscape: lessons from the graveyard and promising candidates. Lancet Infect Dis 2018;18(10):e295–e311. doi: 10.1016/S1473-3099(18)30292-5.

[73] Wu S, Zhong G, Zhang J, et al. A single dose of an adenovirus-vectorized vaccine provides protection against SARS-CoV-2 challenge. Nat Commun 2020;11(1):4081. doi: 10.1038/s41467-020-17972-1.

[74] Singh PK, Kulsum U, Rufai SB, et al. Mutations in SARS-CoV-2 leading to antigenic variations in spike protein: a challenge in vaccine development. J Lab Physiol 2020;12(2):154–160. doi: 10.1056/S040-1715790.

[75] Becerra-Flores M, Cardozo T. SARS-CoV-2 viral spike G614 mutation exhibits higher case fatality rate. Int J Clin Pract 2020;74(8):e13525. doi: 10.1111/ijs.p.13525.

[76] Kober B, Fischer WM, Gnanakaran S, et al. Spike mutation pipeline reveals the emergence of a more transmissible form of SARS-CoV-2. bioRxiv 2020, 2020.05.05. doi: https://doi.org/10.1101/2020.04.29.069054.

[77] Wang ZJ, Schmidt F, Wesblum Y, et al. mRNA vaccine-elicited antibodies to SARS-CoV-2 and circulating variants. bioRxiv, 2021;2021.01.30. doi: https://doi.org/10.1101/2021.01.15.426911.

[78] World Health Organization Solidarity VaccinesTrial Expert Group COVID-19 vaccine trials should seek worthwhile efficacy. Lancet 2020;396(10253):741–743. doi: 10.1016/S0140-6736(20)31823-3.

[79] Cohen J, Kupferschmidt K. Infectious diseases. Ebola vaccine trials raise ethical issues. Science 2014;346(6207):289–290. doi: 10.1126/science.346.6207.289.

[80] Krause PR, Fleming TR, et al. WHO Ad Hoc Expert Group on the Next Steps for Covid-19 Vaccine Evaluation: placebo-controlled trials of Covid-19 vaccines – why we still need them. N Engl J Med 2020;384(2):e2. doi: 10.1056/NEJMep2033338.

Edited By Haijuan Wang and Wei Zhao

How to cite this article: Jiang HD, Li JX, Zhang P, et al. The COVID-19 vaccine in clinical trials: Where are we now? Infect Dis Immun 2021;1(1):43–51. doi: 10.1097/IDI.0000000000000003