The Effect of the Aquatic Extract of Stevia on the MDA Level and Catalase Activity in the Testicular Tissue of Streptozotocin-Nicotinamide-Induced Diabetic Rats

Fatemeh Gholizadeh,1 Pooneh Mokarram1,*, Sanaz Dastgheib1 and Zahra Rahpeima1

1Department of Biochemistry, School of Medicine, Shiraz University of Medical Sciences, Shiraz, Iran

*Corresponding author: Department of Biochemistry, School of Medicine, Shiraz University of Medical Sciences, Shiraz, Iran. Tel/Fax: +98-7132303029, Email: mokaram2@gmail.com

Received 2017 August 29; Revised 2018 June 27; Accepted 2018 June 30.

Abstract

Background: It is believed that diabetes has undesirable impacts on the function of the reproductive system in men like decreasing plasma testosterone levels and sperm count and making apoptosis in the testicular germ cells. There are some chemical drugs for improving the complications, but it has been proven that chemicals have many side effects in the patients. Hence, herbal medicines due to anti-oxidant properties, less cost, and fewer side effects can be beneficial substitutions for chemical drugs. Nowadays, Stevia rebaudiana Bertoni as a sweet plant has shown to be effective in ameliorating diabetic problems. This experimental study investigated the effects of stevia on suppressing oxidative stress in Streptozotocin-Nicotinamide (STZ-NA)-induced diabetic rats’ testes.

Methods: In the present study, adult Wistar rats were separated into three diabetic groups and two sham groups each containing 10 rats: 1. normal group receiving 1 mL of water; 2. diabetic control group receiving 1 mL of water; 3. and 4. diabetic groups treated with the aquatic extracts of stevia (400 mg/kg) and metformin (500 mg/kg), respectively (treated four weeks after confirming diabetes for 14 days); and 5. healthy group treated with the aquatic extract of stevia (400 mg/kg). Finally, after six weeks, the rats were anesthetized, blood samples were collected from the heart for evaluating serum glucose, and testis tissues were separated for measuring Malondialdehyde (MDA) level and Catalase (CAT) activity. Statistical analysis was carried out using one-way analysis of variance (ANOVA), followed by Tukey post hoc multiple-comparison test and data were presented as the mean ± standard error of the mean (SEM).

Results: This study documented that stevia significantly declined serum glucose (P < 0.001) and MDA level (P = 0.008) and increased catalase activity (P = 0.005) in the treated groups (metformin and stevia) compared to the diabetic groups.

Conclusions: The upshot of all, stevia, due to its antioxidant nature, could improve oxidative stress in the testis tissue of diabetic rats.

Keywords: Diabetes Mellitus, Nicotinamide-Streptozotocin, Oxidative Stress, MDA, Catalase Activity, Testicular Tissue, Stevia

1. Background

Diabetes mellitus is a serious metabolic disorder with functional and structural impacts (1). Oxidative stress has a fundamental role in the progression of diabetes complications (2). Within and following diabetes, non-absorbed glucose in fat tissue and muscles leads to the elevation of glucose concentration in the blood. The excessive glucose increases the oxidant product and devastates the antioxidant defense system (3).

Reduced CAT activity and increased MDA levels are considered as indicators of oxidative stress (4, 5). CAT is one of the effective enzymes known to protect cells from hydrogen peroxide generated within the cells (6) and MDA is a toxic product of polyunsaturated fatty acid peroxidation (7).

It is believed that diabetes has undesirable impacts on the function of the reproductive system in men like decreasing plasma testosterone levels and sperm count and making apoptosis in the testicular germ cells (8). Besides, diabetes in the testicular tissue of rats brings about hypox spermatogenesis changing in the dynamics of testicular microvascular blood flow, endocrine signaling, and male infertility by increasing lipid peroxidation and ROS production (9).

The testicular tissue because of several reasons is extremely predisposed to the activity of oxidative stress and free radicals due to containing high levels of unsaturated
fatty acids, high rate of cell division, high rate of cell competition for oxygen, and low oxygen pressure due to weakened vessels. Additionally, since the body’s antioxidant system is not capable of invalidating all free radicals, antioxidant supplements are suggested to reduce the adverse impacts of oxidative stress, elevate spermatogenesis, and enhance fertility (10).

Synthetic antioxidants are often associated with undesirable side effects and high cost for patients; hence, herbal plants are alternative therapies because of anti-oxidant properties, low-cost treatment, reliability, and fewer side effects. *Stevia rebaudiana* Bertoni as an herbal medicine is a sweet and nutrient-rich plant, and it is effective in the regulation of blood glucose. The sweet taste of Stevia leaf is due to the presence of steviol glycosides in its leaves (10). Stevia and its glycosides have therapeutic effects against several diseases such as cancer, diabetes mellitus, inflammation, obesity, and tooth decay (11-15).

Since the control of carbohydrate intake and compliance with a low-carbohydrate diet are important for successful remedy of diabetes, consuming Stevia as an impressive plant in diabetes treatment is affordable (16). Clinical trial studies have proven that the leaf extract of *Stevia rebaudiana* decreases blood glucose in diabetic patients (16, 17). Another study showed that stevioside prevents the absorption of glucose in the duodenum of rats (18).

It has been well documented that the aquatic extract of stevia decreases the serum level of Interleukin-6 as an inflammatory factor, in STZ-NA-induced diabetic rats (19). Moreover, a recent study has shown that stevia by increasing the insulin level exerts profitable anti-hyperglycemic effects via a PPAR-dependent mechanism and antioxidant activities in pancreatic tissues could be effective in diabetic rats (20).

Nowadays, it is believed that one of the adverse impacts of diabetes on the male reproductive system is increasing the MDA level and decreasing the catalase activity (9). Since stevia has an antioxidant nature and the effects of stevia on oxidative stress in the testicular tissue of diabetic rats are unclear, the current study focused on stevia for the treatment of diabetes by diminishing oxidative stress.

2. Methods

2.1. Chemicals

STZ (Streptozotocin), NA (Nicotinamide), TBA (thiobarbituric acid), and TEP (1, 1’, 3, 3’-teta ethoxy propane) were provided by Sigma (Germany). In addition, metformin was supplied by Sigma (USA); Stevia leaves were purchased from Golsaran company, Rasht, Iran.

2.2. Animals

Adult male Wistar rats (4 months-old, 200 - 250 g) were procured from Shiraz University of Medical Science. All rats were acclimatized to new environmental conditions for one week, prior to initiation of the experiment. The rats were kept under standard laboratory conditions of light/dark cycle (12/12 hours), fed with standard diet, chow diet (Pars Dam Co, Tehran, Iran), and water ad libitum. The experimental rats were divided into 5 groups each containing 10 rats. The study was conducted in accordance with the principles of institutional animal ethics.

2.3. Experimental Design

Diabetes was induced chronically by an intraperitoneal injection of NA (Nicotinamide, 120 mg/kg b. wt., dissolved in normal saline) 15 minutes before the single-dose administration of STZ (Streptozotocin, 60 mg/kg b. wt., freshly prepared in citrate buffer, pH 4.5). Following the diabetes induction, blood was drawn from the tail vein of the experimental rats for determining their fasting blood glucose levels. Rats with fasting blood glucose levels of more than 250 mg/dL were considered diabetic and selected for the treatments using metformin (500 mg/kg) and aquatic extract of stevia (400 mg/kg). Within six weeks, our cohort comprised the following groups: 1. Group A: water, 1 mL (Control) and 2. Group B: water, 1 mL (diabetic control). Four weeks after the induction of diabetes, the diabetic rats were randomly divided into two groups: 3. Group C and 4. Group D: diabetic groups treated with the aquatic extract of stevia (400 mg/kg) and metformin (500 mg/kg), respectively; these groups received the treatment by gavage in a single dose every morning for two weeks; and 5. Group E: healthy group treated with the aquatic extract of stevia (400 mg/kg).

2.4. Preparation of Aquatic Extract of Stevia

Stevia leaves were purchased from Golsaran Company (Rasht, Iran) from March 10 to 29. The dried leaves were finely powdered with the help of a grinder. In the next step, the powder was extracted in a Soxhlet apparatus and evaporated to dryness under decreased pressure in a vacuum rotary evaporator. The extract was air-dried up to solid to obtain a semi-solid mass. Next, 100 g of the powder extract suspension was mixed in 1200 mL of distilled water and kept in dark for 24 hours. Subsequently, the solution was filtered and the extract was evaporated into a rotary evaporator at the temperature range of 40 - 50°C. To confirm that the extraction is dry and has no humidity, it was placed in a vacuum desiccator for 24 hours; hence, the efficiency was calculated (20).
2.5. Glucose Measurement

The serum glucose level was measured after six weeks of treatment by using prestige instrument (Hitachi, Japan) and diagnostic colorimetric kits (BioSystem, Spain).

2.6. Preparation of Homogenate from Testicular Tissue

Testes were separated, frozen, and rapidly homogenized in 5 mL cold-normal saline with the aid of homogenizer (Potter Elvehjem, Bodine Electric, Chicago, IL, USA). The homogenate was filtered and then centrifuged at 10,000 g for 1 hours at 4°C. The supernatant obtained was used for the estimation of total protein, MDA, and CAT activity.

2.7. Determination of Malondialdehyde (MDA)

The MDA level was measured using Hagar et al.’s method (21). In short, since aldehyde is not stable, MDA was supplied as an acetal derivative. Within the acid incubation step at 95°C, the acetal form (TEP or 1, 1', 3, 3'-tetra ethoxy propane) was hydrolyzed to form MDA. The TEP standard was supplied as a 10 mM stock solution in Tris-HCl and diluted 1/500 (v/v) in water. Finally, MDA concentrations were detected according to a standard curve at 532 nm.

2.8. Determination of Catalase (CAT) Activity

Catalase (EC 1.11.1.6) was measured according to the Aebi’s procedure based on the content of H2O2 that decomposed (22). In short, 66.7 mmol/L phosphate buffer (pH = 7.0), 20 ul of pancreas supernatant, and 355 ul of distilled water were added to 75 ul of H2O2 (120 mmol/L) and the utilization of H2O2 was followed spectrophotometrically at 240 nm for 3 minutes at 25°C. The catalase activity was expressed as µmol H2O2 consumed/min per mg testis supernatant protein using a molar extinction coefficient of 43.6 L/mol per cm for H2O2.

2.9. Statistical Analysis

The results were stated as the mean ± standard error of the mean (SEM). One-way analysis of variance (ANOVA) followed by Tukey’s post hoc multiple-comparison tests using SPSS (Version 16; SPSS, Chicago, IL, USA) was performed on the data of biochemical variables to investigate the between-group differences. The values of P < 0.05 were considered statistically significant.

3. Results

The present study describes that stevia has an influential effect on decreasing blood glucose and oxidative stress via reducing the MDA and increasing the CAT activity. The mean value of body weight, serum levels of glucose, and antioxidant status of the testis tissues in different experimental groups are shown in Table 1.

This Table shows the increasing effect of both drugs (groups C and D) on weight compared to group B, increasing CAT activity and decreasing FBS in treatment groups, and a non-significant difference in the CAT activity and MDA level between groups A and E.

3.1. Body Weight

Body weight significantly decreased (P = 0.001) in the diabetic group (group B) in comparison with the control group (group A). In addition, in comparison with the diabetic group, body weight significantly increased in both diabetics treated with stevia (group C, P = 0.03) and diabetics treated with metformin (group D, P = 0.002) and metformin could decrease body weight more than stevia did. Between the healthy stevia (group E) and control groups, there was no significant difference (Table 1).

3.2. Serum Glucose

The level of serum FBS in the diabetic group (group B) in comparison with the control group (group A) significantly increased (P < 0.001). In both of the diabetic group treated with stevia and the metformin group (groups C and D, respectively) FBS significantly decreased (P < 0.001), but the effect of metformin in decreasing FBS was significantly more than the effect of stevia (P = 0.018). Moreover, there were non-significant differences between control and healthy stevia groups (Table 1).

3.3. Testes Catalase Activity

As shown in Table 1 and Figure 2, it has been shown that the activity of catalase in the homogenate of testes significantly decreased in diabetic rats (group B) in comparison with controls (group A) (P = 0.031). On the other hand, the activity of the enzyme increased in the groups treated with stevia or metformin (groups C and D) compared to the diabetic rats (P = 0.005, P = 0.016, respectively); the effect of stevia was better than the effect of metformin, but the differences were not significant.

3.4. Testes Lipid Peroxidation

In Table 1 and Figure 2, it has been shown that the amount of MDA significantly increased in the diabetic group compared to the control group (P < 0.001). In addition, in both groups of treated with stevia and treated with...
Table 1. Body Weight, Serum Levels of Glucose and Antioxidant Status of the Testis Tissues in Different Experimental Groups (N = 12)(router)

| Experimental Groups | Control (A) | Diabetic (B) | Diabetic Stevia (C) | Diabetic Metformin (D) | Healthy Stevia (E) |
|---------------------|-------------|-------------|---------------------|------------------------|-------------------|
| Body weight, gr     | 284.3 ± 12.47** | 204.7 ± 8.804** | 261.5 ± 22.21** | 283.5 ± 6.796** | 269.6 ± 14.54**  |
| Serum FBS, mg/dL    | 76.83 ± 3.89** | 507.3 ± 18.34** | 231.0 ± 61.59** | 88.83 ± 10.04** | 76.0 ± 5.04**    |
| Catalase activity, IU/mg protein | 31.89 ± 4.78** | 9.14 ± 1.02** | 37.3 ± 5.87** | 34.08 ± 7.24** | 36.07 ± 4.53**   |
| MDA, nmol/mg protein | 0.036 ± 0.006** | 0.115 ± 0.005** | 0.071 ± 0.008** | 0.055 ± 0.006** | 0.03 ± 0.008**   |

*a Values are expressed as mean ± SEM.

**A, Compared with diabetic groups. B, Compared with the control groups. *P < 0.05, **P < 0.01.

Figure 1. The effect of treatment with the aquatic extract of stevia on catalase activity. Data are presented as mean ± standard error of the mean (SEM). *P < 0.05, **P < 0.001 for significant changes compared to control.

Figure 2. The effect of treatment with the aquatic extract of stevia on the MDA level. Data are presented as mean ± standard error of the mean (SEM). (*P < 0.05, **P < 0.001) for significant changes compared to control.

metformin, the amount of MDA significantly decreased, respectively (P = 0.001, P < 0.001). Moreover, the effect of metformin was better than the effect of stevia but not significantly. Stevia had no significant adverse impact on the healthy group.

4. Discussion

The present study indicates that stevia can significantly reduce serum glucose, decline MDA level, and increase catalase activity in the testicular tissue of STZ-NA-induced diabetic rats. Indeed, these results may be due to the beneficial anti-diabetic impacts of stevia as an herbal medicine. This study showed that stevia had no any side effects; in our previous studies, this result had also been proven (19, 20).

The induction of this type of diabetes was done by the injection of NA (120 mg/kg b. wt.) 15 minutes before STZ administration (60 mg/kg b. wt.) to benefit from the effect of NA in protecting β-cells against STZ (23).

According to one of our studies conducted on STZ-NA-induced diabetic rats, after treating diabetic rats with stevia for 28 days, it was observed that stevia could decrease serum glucose but metformin was more effective than stevia in decreasing serum glucose (19). Thus, our previous outcome was in the same line and approved our present study that stevia has significant effects on decreasing serum glucose (P < 0.001). In our another study, it has been documented that stevia in STZ-induced diabetic rats after 28 days had dramatic impacts on pancreatic β-cells and raised the insulin level and diminished blood glucose by increasing PPARγ expression. PPARγ could control the level of blood glucose in diabetic rats by inducing the insulin secretion (20). Besides, in some investigations, it was revealed that stevioside not only boosted insulin secretion but also diminished gluconeogenesis in diabetic rats;
therefore, serum glucose dropped (24). To sum up, stevia and its ingredients could decrease serum glucose in diabetic rats (24-27).

Hyperglycemia and insulin deficiency conditions can induce excessive reactive oxygen species (ROS) and oxidative stress in the testicular tissue (28). ROS and oxidative stress can cause functional disorders in testes and boost insulin deficiency (29, 30). Excessive free radicals or ROS lead to the depletion of the antioxidant enzymes, the drop of insulin deficiency (29, 30). Excessive free radicals or ROS lead to the depletion of the antioxidant enzymes, the drop of insulin deficiency (29, 30).

The depletion of the antioxidant enzymes, the drop of insulin deficiency (29, 30). Excessive free radicals or ROS lead to the depletion of the antioxidant enzymes, the drop of insulin deficiency (29, 30). Excessive free radicals or ROS lead to the depletion of the antioxidant enzymes, the drop of insulin deficiency (29, 30). Excessive free radicals or ROS lead to the depletion of the antioxidant enzymes, the drop of insulin deficiency (29, 30).

In 2012, Sharma et al. proved that in alloxan diabetes. Since stevia due to its antioxidant features has a specific effect on declining the amount of MDA and raising the CAT activity in the supernatant of diabetic rats’ testes, stevia might be effective in improving the complications of diabetes in the male reproductive system.

Acknowledgments

The present manuscript was extracted from a thesis written by Fatemeh Gholizadeh and financially supported by Shiraz University of Medical Sciences, Grant No. 95-11292.

References

1. Meyer KA, Kushner LH, Jacobs DR Jr, Slavin J, Sellers TA, Folsom AR. Carbohydrates, dietary fiber, and incident type 2 diabetes in older women. Am J Clin Nutr. 2000;71(4):921-30. doi: 10.1093/ajcn/71.4.921. [PubMed: 1078498].

2. Wolff SP. Diabetes mellitus and free radicals. Free radicals, transition metals and oxidative stress in the aetiology of diabetes mellitus and complications. Br Med Bull. 1993;49(3):542-52. doi: 10.1093/oxfordjournals.bmb.a072637. [PubMed: 8226029].

3. Shivanna N, Naika M, Khanum F, Kaul VK. Antioxidant, anti-diabetic and renal protective properties of Stevia rebaudiana. J Diabetes Complications. 2013;27(3):431-3. doi: 10.1016/j.jdiacomp.2012.10.001. [PubMed: 2340951].

4. Roman-Ramos R, Flores-Saenz JL, Partida-Hernandez G, Lara-Lemus A, Alarcon-Aguilar F. Experimental study of the hypoglycemic effect of some antidiabetic plants. Arch Invest Med (Mex). 1991;21(1):87-93. [PubMed: 1819989].

5. Pooya S, Jalali MD, Jazzayeri AD, Saedisomeili A, Eshraghian MR, Toorang F. The efficacy of omega-3 fatty acid supplementation on plasma homocysteine and malondialdehyde levels of type 2 diabetic patients. Nutr Metab Cardiovasc Dis. 2010;20(5):326-31. doi: 10.1016/j.numecd.2009.04.002. [PubMed: 19470799].

6. Mates JM, Sanchez-Jimenez F. Antioxidant enzymes and their implications in pathophysiological processes. Front Biosci. 1999;4:D339-45. [PubMed: 10075544].

7. Rosato R, Ciccone G, Bo S, Pagano GF, Merletti F, Gregori D. Evaluating cardiovascular mortality in type 2 diabetes patients: an analysis based on competing risks Markov chains and additive regression models. J Eval Clin Pract. 2007;13(3):422-8. doi: 10.1111/j.1365-2753.2006.00732.x. [PubMed: 17588809].

8. Hajizadeh MR, Eftekhar E, Zal F, Jafari A, Mostafavi-Pour Z. Mulberry leaf extract attenuates oxidative stress-mediated testosterone depletion in streptozotocin-induced diabetic rats. Iran J Med Sci. 2014;39(2):123-9. [PubMed: 24644381]. [PubMed Central: PMC3957011].

9. Shirilatha B; Muralidhara. Occurrence of oxidative impairments, response of antioxidant defences and associated biochemical perturbations in male reproductive milieu in the Streptozotocin-diabetic rat. Int J Androl. 2007;30(6):508-18. doi: 10.1111/j.1365-2605.2007.00748.x. [PubMed: 17573857].

10. Bridel M, Lavielle R. Sur le principe sucre des feuilles de kaa-he-e (Stevia rebaudiana B). Acad Sci Paris Compt Rendus. 1931;192(112):3-5.
11. Boonkaewwan C, Burodom A. Anti-inflammatory and immunomodulatory activities of stevioside and steviol on colonic epithelial cells. J Sci Food Agric. 2013;93(15):3820–5. doi:10.1002/jsfa.6287. [PubMed: 23794454].

12. Boonkaewwan C, Toskulkao C, Vongsakul M. Anti-Inflammatory and Immunomodulatory Activities of Stevioside and Its Metabolite Steviol on THP-1 Cells. J Agric Food Chem. 2006;54(3):785–9. doi:10.1021/jf0523465. [PubMed: 16448183].

13. Jayaraman S, Manoharan MS, Illanchezian S. In-vitro antimicrobial and antitumor activities of Stevia rebaudiana (Asteraceae) leaf extracts. Trop J Pharm Res. 2008;7(4):1434-9. doi:10.4314/tjpr.v7i4.14700.

14. Cardello HM, Da Silva MA, Damasio MH. Measurement of the relative sweetness of stevia extract, aspartame and cyclamate/saccharin blend as compared to sucrose at different concentrations. Plant Foods Hum Nutr. 1999;54(2):199–30. doi:10.1023/A:1008344420339. [PubMed: 10646559].

15. Grenby TH. Update on low-calorie sweeteners to benefit dental health. Int Dent J. 1991;41(4):127–24. [PubMed: 1917078].

16. Gregersen S, Jeppesen PB, Holst J]., Hermansen K. Antihyperglycemic effects of stevioside in type 2 diabetic subjects. Metabolism. 2004;53(1):73–6. doi:10.1016/j.metabol.2003.07.013. [PubMed: 14681845].

17. Oviedo C, Franciani G, Moreno R, Maas L. Action hipoglucemiante de la Stevia rebaudiana Bertoni. Seventh Congress of the International Diabetes Federation. 1970.

18. Maier V, Hermann K, Kienle U. Isolated perfused gut: absorption of glucose is inhibited by steviol, a degradation product of the sweetener stevioside. Diabetes. 2003;52:554.

19. Bayat E, Dastghib S, Edgar S, Mokarram P. Effect of the aquatic extract of stevia on the serum level of interleukin-6 in streptozotocin-diabetic rats. PPARgamma-dependent Regulation or Antioxidant Potential. Avicenna J Med Biotechnol. 2016;8(2):65–74. [PubMed: 2714265]. [PubMed Central: PMC4842244].

20. Hagar HH, El Etter E, Arafa M. Taurine attenuates hypertension and renal dysfunction induced by cyclosporine A in rats. Clin Exp Pharmacol Physiol. 2006;33(3):189–96. doi:10.1111/j.1440-1681.2006.03454.x. [PubMed: 16487261].

21. Goth L. A simple method for determination of serum catalase activity and revision of reference range. Clin Chim Acta. 1991;196(2-3):343–51. doi:10.1016/0009-8981(91)90067-M. [PubMed: 2927980].

22. Szudeksi T. Streptozotocin-nicotinamide-induced diabetes in the rat. Characteristics of the experimental model. Exp Biol Med (Maywood). 2012;237(5):481-90. doi:10.1058/ebmh.2012.018372. [PubMed: 22691973].

23. Chen TH, Chen SC, Chan P, Chu YL, Yang HY, Cheng JT. Mechanism of the hypoglycemic effect of stevioside, a glycoside of Stevia rebaudiana. Planta Med. 2000;66(2):108-13. doi:10.1055/s-2000-67775. [PubMed: 15729617].

24. Sumon MH, Mostofa M, Jahan MS, Kayesh MEH, Haque MA. Comparative efficacy of powdered form of stevia (Stevia rebaudiana Bertoni) leaves and glimepiride in induced diabetic rats. Bangl J Vet Med. 2008;6(2):211-5.

25. Benford D, Hill MF, Schlatter J, DiNovi M. Stevial glycosides (addendum). Safety evaluation of certain food additives. 2009. 183 p.

26. Saravanan R, Vengathash babu K, Ramachandran V. Effect of Rebaudioside A, a diterpenoid on glucose homeostasis in STZ-induced diabetic rats. JPhytoBioChem. 2012;6(8):421-31. doi:10.1007/s10525-012-0156-0. [PubMed: 22374587].

27. Alves MG, Martins AD, Rato L, Moreira PL, Socorro S, Oliveira PF. Molecular mechanisms beyond glucose transport in diabetes-related male infertility. Biochim Biophys Acta. 2013;1832(5):826–35. doi:10.1016/j.bbadis.2013.01.010. [PubMed: 23348098].

28. Adewole SO, Caxton-Martins EA, Salako AA, Doherty OW, Naicker T. Effects of oxidative stress induced by streptozotocin on the morphology and trace minerals of the testes of diabetic wistar rats. Pharmacologyonline. 2007;2:478-97.

29. Sexton WJ, Jarow JP. Effect of diabetes mellitus upon male reproductivity function. Urology. 1997;49(4):508-11. doi:10.1016/S0090-4295(96)00573-0. [PubMed: 911668].

30. Del Rio D, Stewart AJ, Pellegrini N. A review of recent studies on malondialdehyde as toxic molecule and biological marker of oxidative stress. Nutr Metab Cardiovasc Dis. 2005;15(4):316-28. doi:10.1016/j.numecd.2005.05.003. [PubMed: 16054557].

31. White CR, Brock TA, Chang LY, Crapo J, Briscoe P, Ku D, et al. Synergistic effects of oxidative stress induced by streptozotocin on the morphological and physiologic response of Leydig cell function in the rat. Proc Natl Acad Sci U S A. 1994;91(1):1044-8. doi:10.1073/pnas.91.1.1044.

32. Benitez A, Perez Diaz J. Effect of streptozotocin-diabetes and insulin treatment on regulation of Leydig cell function in the rat. Horm Metab Res. 1985;17(1):5-7. doi:10.1055/s-1985-131433. [PubMed: 2988762].

33. Kakkar R, Kalra J, Mantha SV, Prasad K. Lipid peroxidation and activity of antioxidant enzymes in diabetic rats. Mol Cell Biochem. 1995;151(2-3):233-9. doi:10.1007/BF03223333. [PubMed: 8509756].

34. Kim IS, Yang M, Lee OH, Kang SN. The antioxidant activity and the bioactive compound content of Stevia rebaudiana water extracts. JWT - Food Sci Technol. 2011;44(5):5:328-32. doi:10.1016/j.jwt.2010.12.003.

35. Singh S, Garg V. Antidiabetic, antisyphilidymic and antioxidant potential of methanolic root extract of Stevia rebaudiana (Bertoni) on alloxan induced diabetic. J Pharm Pharm Sci. 2014;17(3):1859-72.