The role of source delocalization in the development of morphogen gradients

Hamid Teimouri and Anatoly B Kolomeisky
Department of Chemistry and Center for Theoretical Biological Physics, Rice University, Houston, Texas, 77005, USA
E-mail: tolya@rice.edu

Keywords: morphogen gradient, stochastic models, reaction-diffusion processes

Abstract
Successful biological development via spatial regulation of cell differentiation relies on the action of multiple signaling molecules that are known as morphogens. It is now well-established that signaling molecules create non-uniform concentration profiles, morphogen gradients, that activate different genes, leading to patterning in the developing embryos. The current view of the formation of morphogen gradients is that it is a result of complex reaction-diffusion processes that include the strongly localized production, diffusion and uniform degradation of signaling molecules. However, multiple experimental studies also suggest that the production of morphogen in many cases is delocalized. We develop a theoretical method that allows us to investigate the role of the delocalization in the formation of morphogen gradients. The approach is based on discrete-state stochastic models that can be solved exactly for arbitrary production lengths and production rates of morphogen molecules. Our analysis shows that the delocalization might have a strong effect on mechanisms of the morphogen gradient formation. The physical origin of this effect is discussed.

1. Introduction

One of the most important fundamental biological processes is a formation of a multi-cellular organism from a set of genetically identical cells. The central role in this process is played by morphogens, which are signaling molecules that control the fate of biological cells [1–9]. It is now widely accepted that these signaling molecules develop non-uniform concentration profiles, called morphogen gradients, that interact with embryo cells. The idea behind the morphogen gradients is that different concentrations of signaling molecules turn on different genes, leading to complex patterning observed in developed multi-cellular systems [1, 3, 4]. A significant progress in understanding of morphogen gradients and how they function has been achieved in recent years when many quantitative investigations have appeared [9–20]. However, despite these advances, mechanisms of the formation of signaling molecules profiles remain not fully understood [7, 8].

Several theoretical ideas were presented to explain the complex processes associated with the development of morphogen gradients [7, 8]. The dominating view is based on a so-called synthesis-diffusion-degradation (SDD) model [11, 13, 24]. According to this approach, the production of signaling molecules starts at the specific localized region of the embryo, then signaling molecules diffuse along the cells. In addition, with equal probability they can be degraded after binding to specific receptors on the cells. As a result, exponentially decaying concentration profiles are developed at long times. Since qualitatively similar behavior is observed in many experimental systems, the SDD models have been widely utilized for understanding morphogen gradients [11–16, 21, 24].

The majority of investigations of morphogen gradients formation that use the SDD models postulate that the signaling molecules are produced from a sharply localized source [25–28, 30–33]. However, experimental observations suggest that in many biological systems the production region of the morphogens is delocalized [11]. Morphogens are protein molecules that are synthesized from the corresponding RNA molecules. So the production of these signaling molecules to a large degree is determined by the distribution of the corresponding RNA molecules. For one of the most intensely studied systems, the formation of bicoid morphogen in an early Drosophila embryo, it is...
known that the maternal RNA molecules are distributed over the region of size 30–50 μm, while the total length of the embryo is of order of 400 μm [11]. Obviously, in this case the production area cannot be defined as sharply localized. This raises many queries about the role of the source production in the developing morphogen gradients. Specific questions include: why is the synthesis of signaling molecules delocalized, why is it not produced over the whole embryo, and how does the morphogen gradient depend on the spatial distribution of the source and on the synthesis rate? At the same time, although some of these issues were discussed, a comprehensive theoretical analysis of the delocalization of signaling molecules synthesis is not available [21, 22, 23, 25–28].

Experimental investigations suggest that specific distribution of maternal RNA molecules within cells controls the synthesis of morphogen molecules [29]. Three distinct mechanisms of RNA localization have been identified, including the protection from RNA degradation over the specific regions, diffusion of RNA molecules coupled with entrapment at specific locations and directed transport by motor proteins along cytoskeleton filaments [29]. The first mechanisms leads to mostly uniform production distribution of signaling molecules over finite regions, while the second and the third mechanisms produce mostly exponential source distributions.

In this paper, we present a theoretical investigation on the role of source production in the development of morphogen gradients. A theoretical approach for analyzing the formation of signaling molecules concentration profiles with arbitrary delocalization length and production rates is developed. Our method is based on discrete-state stochastic models that can be explicitly solved for arbitrary sets of parameters. We investigate several possible cases of the formation of signaling molecules concentration profiles to analyze the role of the synthesis of the signaling molecules. It is shown that the production might have a strong impact on the development of morphogen gradients.

The paper is organized as follows. In section 2, a general discrete-state stochastic SDD model with an arbitrary source distribution is presented and analyzed. In section 3, three different distributions are explicitly analyzed as illustrative examples. In this section the stationary profiles and local accumulation times (LATs) are calculated. Finally, in section 4, the important biophysical implications of the presented results are discussed and conclusions are given.

2. Theoretical method

We start our analysis by considering a general discrete-state stochastic SDD model in one-dimension as illustrated in Figure 1. Our system is semi-infinite. It is assumed that the synthesis of the morphogen particles is taking place only in the interval consisting of L sites: see figure 1. Inside the source region, signaling molecules are produced at any site \( m \) (0 ≤ \( m \) ≤ \( L \)) with a corresponding rate \( Q_{in} \), and the total production rate is equal to \( Q = \sum_{m} Q_{in} \). From any site morphogens can jump to nearest neighbors left or right sites with a rate \( u \). The particles might be degraded at any site with a rate \( k \). One can define a function \( P(n, t; m) \) as the probability to find the morphogen at the site \( n \) at time \( t \) if the particle can only be produced at the site \( m \) (0 ≤ \( m \) ≤ \( L \)). The temporal evolution of this probability is governed by the following master equations:

\[
\frac{dP(n, t; m)}{dt} = Q_{in} \delta_{n,n} + u[P(n - 1, t; m)] + P(n + 1, t; m)] - (2u + k)P(n, t; m) \tag{1}
\]

for \( n > 0 \), and

\[
\frac{dP(0, t; m)}{dt} = Q_{in} \delta_{m,1} + uP(1, t; m)] - (u + k)P(0, t; m) \tag{2}
\]

for \( n = 0 \). Here we use the fact that \( \delta_{n,n} = 1 \) for \( m = n \), and it is zero otherwise.

At long times we have \( \frac{dP(n, t; m)}{dt} = 0 \), and these equations can be solved analytically, yielding the following stationary probability functions

\[
P_{\infty}^{(1)}(n; m) = \left( Q_{in} \left[ \begin{array}{c} k + \sqrt{k^2 + 4uk} \\ k + \sqrt{k^2 + 4uk} \\
\end{array} \right] x^{m-n} \\
\left( k + \sqrt{k^2 + 4uk} \right)^{x^{m}} \right) \tag{3}
\]

for 0 ≤ \( n \) ≤ \( m \), and

\[
P_{\infty}^{(n)}(n; m) = \left( Q_{in} \left[ \begin{array}{c} k + \sqrt{k^2 + 4uk} \\ k + \sqrt{k^2 + 4uk} \\
\end{array} \right] x^{n-m} \\
\left( k + \sqrt{k^2 + 4uk} \right)^{x^{n}} \right) \tag{4}
\]

for \( m \leq n \). In these expressions, the subscript < (>) corresponds to the case of \( n \leq m \) (\( n > m \)), and a parameter \( x (0 < x < 1) \) is defined as
\[ x = \left( 2u + k - \sqrt{k^2 + 4uk} \right)/(2u). \] (5)

This approach is very useful since it allows us to obtain the total concentration profiles from all producing sites using a kind of a superposition principle. In other words, the total probability \( P(n, t) \) of finding the particle at site \( n \geq 0 \) at time \( t \) can be expressed as a sum of the probabilities \( P(n, t; m) \) with production at the specific site \( m \) \((0 \leq m \leq L)\). This is because the synthesis processes at each site are independent of each other. The general equations for concentration profiles are given by

\[
P(n, t) = \begin{cases} 
\sum_{m=0}^{n} P_n(n, t; m) + \sum_{m=n+1}^{L} P_n(n, t; m), & \text{for } 0 \leq n \leq L; \\
\sum_{m=0}^{L} P_n(n, t; m), & \text{for } L \leq n.
\end{cases}
\] (6)

The explicit expressions can be easily obtained by employing equations (3) and (4).

One of the main goals of morphogen gradients is to transfer the information. Most probably, it can be done well if the system is close to the stationary conditions. Then the important characteristics of morphogen gradients are times needed to achieve the steady state at specific spatial locations. These times are known as LAT, and a theoretical framework for computing these quantities has been developed recently [25]. It can be done by utilizing local relaxation functions which are defined as

\[ R(n, t) = \frac{P(n, t) - P^i(n)}{P(n, t = 0) - P^i(n)} = 1 - \frac{P(n, t)}{P^i(n)}. \] (7)

The physical meaning of these functions is that they represent the relative distance to the stationary state: at \( t = 0 \) the distance is one, while at steady state it is equal to zero. The explicit formulas for the local accumulation time can be derived then via Laplace transformations of the local relaxation function, \( \bar{R}(n, s) = \int_{0}^{\infty} R(n, t)e^{-st}dt \) [25]

\[ t(n) = -\int_{0}^{\infty} \frac{\partial R(n, t)}{\partial t} e^{-st}dt = \bar{R}(n, s = 0). \] (8)

### 3.1. Single localized source

As a first example, we start with the case when the source of signaling molecules is localized at the site \( m = m' \)

\[ Q_m = Q\delta(m - m'). \] (9)

From equations (3) and (4) we directly obtain

\[ P_{m'}^i(n; m') = \frac{Qx^n}{k + \sqrt{k^2 + 4uk}} \times \left[ \left( k + \sqrt{k^2 + 4uk} \right)x^{-n} + (-k + \sqrt{k^2 + 4uk})x^n \right]/\left( \sqrt{k^2 + 4uk} \right). \] (10)

For \( m' = 0 \), as expected, we recover the results obtained earlier [31]

\[ P^i(n; 0) = \frac{2Qx^n}{k + \sqrt{k^2 + 4uk}}. \] (12)

These expressions indicate that the signaling molecules profiles are exponentially decaying functions of the distance from the source for \( n > m' \), and the decay length, \( \lambda = 1/\ln x \), is independent of the production rate \( Q \). The resulting morphogen gradients are presented in figures 2 and 3 for the case of \( m' = 0 \) with various diffusion and degradation rates.

From relations (7) and (8) the explicit expressions for the LATs can be evaluated. It is found that LAT are exponentially decaying functions of the distance from the source for \( n > m' \), and the decay rate \( \lambda = 1/\ln x \), is independent of the production rate \( Q \). The resulting morphogen gradients are presented in figures 2 and 3 for the case of \( m' = 0 \) with various diffusion and degradation rates.

From relations (7) and (8) the explicit expressions for the LATs can be evaluated. It is found that LAT are exponentially decaying functions of the distance from the source for \( n > m' \), and the decay rate \( \lambda = 1/\ln x \), is independent of the production rate \( Q \). The resulting morphogen gradients are presented in figures 2 and 3 for the case of \( m' = 0 \) with various diffusion and degradation rates.

### 3.3. Illustrative examples

Our approach allows us to analyze the formation of morphogen gradients for arbitrary length of the production region and for arbitrary production rates. To explain it better, we illustrate the method by doing explicit calculations for three different production scenarios, which might be relevant for the morphogen gradients formation in real cellular conditions.
Again, for the source localized at the origin, \( m' = 0 \), our results reproduce the already known expression [31]

\[
\tau_{\text{s}}(m_0, m') = \frac{1}{\sqrt{k^2 + 4uk}} \\
\times \left[ n + \frac{2u + k + \sqrt{k^2 + 4uk}}{k + \sqrt{k^2 + 4uk}} \right] \\
\times \left[ m' \left( \left( x^m - x^{-m} \right) \right) / \left( x^{-m} \left( k + \sqrt{k^2 + 4uk} \right) \right) \\
+ x^m \left( -k + \sqrt{k^2 + 4uk} \right) \right] \\
+ \frac{2uk}{k^2 + 4uk} \left[ \left( x^m - x^{-m} \right) / \left( x^{-m} \left( k + \sqrt{k^2 + 4uk} \right) \right) \right] \right].
\] (13)

LAT for the formation of morphogen gradients with \( m' = 0 \) for various diffusion and degradation rates are presented in figures 4 and 5. One can see that the LATs for the sharply localized source linearly grow with the distance from the source. This can be explained by invoking the idea that the degradation acts as an effective potential that pushes the signaling molecules away from the source [31], leading to effectively driven diffusion of morphogens in the system.

### 3.2. Uniformly distributed production over the finite interval

In another example, we consider uniformly distributed production of signaling molecules along a finite interval of length \( L \). This distribution can be represented as follows.

\[
\frac{1}{\sqrt{k^2 + 4uk}} \times \left[ \frac{1}{k + \sqrt{k^2 + 4uk}} \right].
\] (15)
In this case, the synthesis rates are the same for all production sites. Applying equation (6), one can obtain explicit expressions for the stationary profiles. They are different depending on the position of the lattice site with respect to the production region. Inside the production area we have

\[ Q_n = \begin{cases} \frac{Q}{L + 1}, & 0 \leq m \leq L; \\ 0, & m > L. \end{cases} \quad (16) \]

The maximal value of the concentration is achieved for \( n = 0 \)

\[ P^{(i)}_{\infty}(n = 0) = \frac{2Q(-1 + x^{L+1})}{(x - 1)(k + \sqrt{k^2 + 4uk})(L + 1)}. \quad (19) \]

The concentration profiles normalized by this maximal value are presented in figures 2 and 3.

A stationary-state behavior of the morphogen gradients with the uniform production along the finite interval can be generally described in the following way. The concentration of signaling molecules is large and almost constant in the production region, and then it exponentially decays outside of the synthesis area. The transition between these two regimes depends on the parameters of the system. Decreasing the diffusion and/or increasing the degradation rate makes this crossover sharper. In addition, the transition is more abrupt when the production length \( L \) is
much larger than the decay length \( \lambda \) (see figure 2 where \( L = 10\lambda \) is utilized), while for comparable \( L \) and \( \lambda \) (see figure 3 where \( L = \lambda \) is utilized) the concentration profile decays more smoothly. It is interesting to analyze the behavior of the profiles in the asymptotic limit of very large production intervals, \( L \to \infty \),

\[
P^{(i)}(n, L \to \infty) \approx \frac{Q(1 + x)}{L(1 - x)\sqrt{k^2 + 4uk}}. \tag{20}
\]

This corresponds to the case when the morphogens are synthesized all over the embryo with the same production rates. As expected, the concentration profile becomes uniform, and this is the reason why there is no dependence on the position \( n \) in this equation. Obviously, the morphogen gradient cannot be established in such systems, and this is not a realistic situation for biological development. This might be the reason why the production region does not occupy the whole embryo.

As was discussed above, the LATs for uniform production along the finite intervals can be found from equations (7) and (8). LAT for sites inside the production region are given by

\[
t_c(n) = \frac{1}{\sqrt{k^2 + 4uk}} \times \left[ n + \frac{2u + k + \sqrt{k^2 + 4uk}}{k + \sqrt{k^2 + 4uk}} \right] + \frac{1}{\sqrt{k^2 + 4uk}} \left[ \left( nx^{-n} - (1 - n)x^{1-n} \right) + (L - 2n + 1)x^{L-2n+1} \right] \left( k + \sqrt{k^2 + 4uk} \right)
+ Lx^{L+1} \left( -k + \sqrt{k^2 + 4uk} \right) \left( x - x^{-n} - x^{1-n} + x^{L-2n+1} \right)
+ \left( k + \sqrt{k^2 + 4uk} \right) - (1 - x^{L+1})
+ \left( -k + \sqrt{k^2 + 4uk} \right) + \frac{2uk}{k^2 + 4uk}
\left[ \left( -x + x^{-n} + x^{1-n} - x^{L-2n+1} - 1 + x^{L+1} \right) / \left( k + \sqrt{k^2 + 4uk} \right)
- (1 - x^{L+1}) \left( -k + \sqrt{k^2 + 4uk} \right) \right]
\]
The results for LATs are presented in figures 4 and 5. For the uniform production along the finite interval the LAT usually consist of two parts. Inside the production area LAT is almost constant as a function of the distance from the origin. Solid curves correspond to the single localized source at $m' = 0$. Dashed curves correspond to uniform production rates along the finite interval. Dotted curves correspond to exponentially decaying production rates along the semi-infinite interval. $L = \lambda = \lambda$ is assumed for the single localized and uniform productions, while $\lambda = \lambda$ and $L \to \infty$ are assumed for the exponentially distributed productions. (a) Fast degradation rates with $k = 1, u = 0.01$; (b) comparable diffusion and degradation rates with $k = u = 1$; and (c) fast diffusion rates with $k = 1, u = 100$. 

$$t_c(n) = \frac{1}{\sqrt{k^2 + 4uk}} \left[ n + \frac{2n + k + \sqrt{k^2 + 4uk}}{k + \sqrt{k^2 + 4uk}} \right] + \frac{L}{\sqrt{k^2 + 4uk}} \left[ x^{-L} \left( k + \sqrt{k^2 + 4uk} \right) - x^{-L-1} \left( k + \sqrt{k^2 + 4uk} \right) \right] + x^{L+1} \left( -k + \sqrt{k^2 + 4uk} \right)/$$

while for the outside region we have

$$t_c(n) = \frac{1}{\sqrt{k^2 + 4uk}} \left[ \left( \frac{1}{x - 1} \right) \left( k + \sqrt{k^2 + 4uk} \right) - x^{-L} \left( k + \sqrt{k^2 + 4uk} \right) \right] + \frac{L}{\sqrt{k^2 + 4uk}} \left[ x^{-L} \left( k + \sqrt{k^2 + 4uk} \right) - x^{-L-1} \left( k + \sqrt{k^2 + 4uk} \right) \right] + x^{L+1} \left( -k + \sqrt{k^2 + 4uk} \right)/$$

+ \left( \frac{2uk}{k^2 + 4uk} \right) \left[ x^{-L} - n - x^{-L-1} - 1 \right] + \left( \frac{1}{x - 1} \right) \left( k + \sqrt{k^2 + 4uk} \right) - x^{L+1} \left( -k + \sqrt{k^2 + 4uk} \right)/$

Figure 5. Local accumulation times as a function of the distance from the origin. Solid curves correspond to the single localized source at $m' = 0$. Dashed curves correspond to uniform production rates along the finite interval. Dotted curves correspond to exponentially decaying production rates along the semi-infinite interval. $L = \lambda = \lambda$ is assumed for the single localized and uniform productions, while $\lambda = \lambda$ and $L \to \infty$ are assumed for the exponentially distributed productions. (a) Fast degradation rates with $k = 1, u = 0.01$; (b) comparable diffusion and degradation rates with $k = u = 1$; and (c) fast diffusion rates with $k = 1, u = 100$.
the distance from the origin. At the same time, outside of the production area LAT is linearly growing. Again, arguments using the role of the degradation as an effective potential can be employed [31]. Outside of the synthesis region, the degradation creates the potential that moves particles away from the production area. This effective potential leads to strong biased diffusion and consequently linearly growing LATs. This behavior is very similar to the case of the single localized source discussed above. Inside the synthesis region the dynamics is different. The degradation is compensated by synthesis and diffusion from neighboring cells, leading to mostly uniform accumulation times.

3.3. Exponentially distributed production along the semi-infinite interval

In the final example, we consider the exponentially distributed production along the interval of length L. In this case, the synthesis rates can be written as

\[ Q_m = \frac{Q(1 - z)z^m}{1 - z^{L+1}}, \]

where \( 0 \leq m \leq L \) and a parameter \( z \) (0 < z < 1) is introduced to characterize the exponential distribution of synthesis rates. One can see that the relation \( \sum_{m=0}^{L} Q_m = Q \) is satisfied. We can also define a decay length \( \lambda_s \) for this exponential distribution of the production rates

\[ \lambda_s = -1/\ln z. \]

Detailed calculations of the stationary profiles for exponentially distributed production rates and for arbitrary length of the source region are given in the appendix. Here we report the final results for the case of the semi-infinite source interval, i.e., in the limit of \( L \to \infty \)

\[ P(n) = \frac{Q(1 - z)}{\sqrt{k^2 + 4uk}} \times \left[ \frac{(x^{n+1} - x^n - z^n + 1)}{x^n (1 - xz)} \right] + \frac{Q(1 - z)}{\sqrt{k^2 + 4uk}} \left[ \frac{(-k + \sqrt{k^2 + 4uk}) x^n}{k + \sqrt{k^2 + 4uk} (1 - xz)} \right]. \] (25)

The maximal value of the concentration profile is achieved again for \( n = 0 \)

\[ P(n = 0) = \frac{2Q(1 - z)}{k + \sqrt{k^2 + 4uk}}. \] (26)

It is interesting to consider a special limiting case of this problem when the diffusion along the lattice cells is much faster than the degradation rate, \( u \gg k \), while the decay length \( \lambda_s \) for the exponential production rates is also large. This corresponds to a continuum limit of our problem, and it was already discussed in the literature [25–28]. In this limit, we have the following asymptotic relations for important parameters \( x \) and \( z \)

\[ x \approx 1 - \frac{1}{\lambda_s}, \] (27)

\[ z \approx 1 - \frac{1}{\lambda_s}. \] (28)

Using these results, one can easily show that the equation (25) reduces to the following form

\[ P(n) = \frac{Q(z^2)}{u(\lambda_s^2 - \lambda_s^2)} \left( \lambda_s e^{-n/\lambda_s} - \lambda_s e^{-n/\lambda_s} \right). \] (29)

This is exactly the concentration profile obtained previously in the continuum SDD model with the exponentially decaying distribution of the production rates [28].

The normalized concentration profiles for exponentially decaying production along the semi-infinite interval (\( L \gg 1 \)) are plotted in figures 2 and 3. For all sets of the parameters these profiles are monotonically decreasing functions of the distance from the origin, although the decay is slower for larger diffusion rates and/or for smaller degradation rates. This allows one to increase significantly the range of morphogen action, as compared to systems with the sharply localized source, while still keeping the concentration profile of signaling molecules to be non-uniform. As expected, accelerating the exponential decay of production rates (lowering of \( \lambda_s \)) diminishes this effect as one can see by comparing figures 2 and 3. As expected, for small \( \lambda_s \) the concentration profiles of morphogen gradients with exponentially distributed productions become similar to the morphogen gradients with the single localized source.

The calculations for LATs for exponentially decaying synthesis of signaling molecules and for arbitrary production lengths \( L \) are also given in the Appendix. We are interested in the large \( L \) values that correspond to the semi-infinite interval. In this case, we obtain the following expression for LAT

\[ t(n) = \frac{n}{\sqrt{k^2 + 4uk}} \left[ \left( -k + \sqrt{k^2 + 4uk} \right) \left( x^{n+1} - z x^n \right) + k + \sqrt{k^2 + 4uk} \right] \times \left( x^{n+1} - z x^n \right) + 2k \left( z^{n+1} - z x^n \right) + \frac{2nk}{k^2 + 4uk} \left( z^{n+2} - z x^{n+1} \right) \]
\[-x^2z^{n+1}/\left( (k + \sqrt{k^2 + 4uk})(x^{n+1}) - z^{n+2} - z^{n+1} + x^2z^{n+1} + (k + \sqrt{k^2 + 4uk})(x^{n+1} - zx^n) \right) + \frac{1}{\sqrt{k^2 + 4uk}} \times (x^{n+1} - 2zx^{n+2} + 2x^2z^{n+1} + (k + \sqrt{k^2 + 4uk})(x^{n+1} - zx^{n+2} - z^{n+1}) + x^2z^{n+1} + (k + \sqrt{k^2 + 4uk})(x^{n+1} - zx^n)) \right] + \frac{1}{\sqrt{k^2 + 4uk}} \left[ \frac{2u + k + \sqrt{k^2 + 4uk}}{k + \sqrt{k^2 + 4uk}} \right] + \frac{xz}{1 - xz} = \frac{x}{x - z}. \]

(30)

The results for LAT for the exponentially distributed production of the morphogens are illustrated in figures 4 and 5. In all cases the LATs increase with the distance from the origin, as was found for other production scenarios of morphogen gradients considered in this work. The LAT growth is mostly linear since our arguments about the effective potential due to degradation can still be applied. But what is different from other production mechanisms is the fact that these times are usually much smaller, especially at large distances from the origin (larger than \( L \) or \( \lambda_1 \)). This can be understood if we recall that main contribution to the LATs at given spatial position is due to the first-passage (arrival) times to this location starting from the source region [31]. In the exponentially distributed production of signaling molecules along the semi-infinite interval the source is everywhere, so that the contribution of the arrival times is mostly negligible. This significantly lowers the time to reach the stationary-state concentration at the given spatial location.

4. Summary and conclusions

We developed a theoretical framework for investigating the role of synthesis of signaling molecules in the formation of concentration profiles that are critically important in the biological development. Our analysis is based on discrete-state stochastic models for complex reaction-diffusion processes associated with the formation of morphogen gradients. It allows us to test the effect of the production in systems with arbitrary source lengths and synthesis rates by calculating stationary profiles and the times to achieve the stationary concentrations at specific locations. By analyzing several different systems we found that the spatial distributions of the sources and the production speeds have a strong effect in the development of morphogen gradients.

To understand the role of the production one might recall that there are two main requirements for successful function of the morphogen gradients in the biological systems [7]. The first one is to deliver the information to as many as possible embryo cells about their future fates. This can be done if observable concentrations of signaling molecules can be found as far as possible from the source. The second function is to ensure that different genes can be controllably turned on in the neighboring embryo cells. This can be accomplished by producing sharp boundaries in the concentration profiles of signaling molecules at specific locations. Our analysis suggests that the morphogen gradients produced with the single localized sources generally are not able to satisfy these requirements. At the same time, the morphogen gradients developed from the delocalized sources with various synthesis rates are capable to do all these tasks successfully.

Furthermore, the presented theoretical method provides a simple physical-chemical explanation on the role of delocalizations in the formation of signaling molecules concentration profiles. The delocalization effectively leads to faster diffusion along the productions regions, and it also shortens the arrival times to specific locations. As a result, the range of morphogen gradients with delocalized sources increases, while the times to reach the stationary states at specific locations become smaller. All of these properties make the morphogen gradients more efficient and robust. At the same time, increasing the production area to the whole embryo is not reasonable since it will be difficult to sustain the non-uniform concentration profiles.

Although our theoretical approach gives a clear physical picture on the effect of production in the development of morphogen gradients, one should note that our model is oversimplified. It neglects many crucial factors and processes in the biological development such as a three-dimensional nature of the embryo, inhomegeneity and complex cooperative mechanisms in the degradation, as well as variable mechanical responses of the embryo cells. It will be critically important to test the proposed theoretical picture in experimental studies and in more advanced theories.

Acknowledgments

The work was supported by grants from the Welch Foundation (C-1559), from the NSF (Grant CHE-1360979) and by the Center for Theoretical Biological Physics sponsored by the NSF (Grant PHY-1427654).
Appendix

In this appendix we present detailed calculations of the stationary profiles and LATs for the exponential distribution of production rates. Using a superposition principle, the total stationary profile can be written as

\[
P_{\xi}(n) = \frac{x^n}{\sqrt{k^2 + 4uk (k + \sqrt{k^2 + 4uk})}} \times \left[ \left( k + \sqrt{k^2 + 4uk} \right) \sum_{m=0}^{\infty} Q_m x^{-m} \right. \\
+ \left. \left( -k + \sqrt{k^2 + 4uk} \right) ^n \right]
\]

for the sites inside the production area. For the sites outside of the production we have

\[
P_{\xi}(n) = \frac{x^n}{\sqrt{k^2 + 4uk (k + \sqrt{k^2 + 4uk})}} \times \left[ \left( k + \sqrt{k^2 + 4uk} \right) \sum_{m=0}^{l} Q_m x^{-m} \right. \\
+ \left. \left( -k + \sqrt{k^2 + 4uk} \right) \sum_{m=0}^{l} Q_m x^{-m} \right]
\]

The summations over \( m \) for different intervals can be performed in the following way

\[
\sum_{m=0}^{L} Q_m x^{-m} = \frac{Q(1 - z)}{1 - \frac{xz^{L+1}}{1 - x}} \left[ 1 - \frac{(xz)^{L+1}}{1 - xz} \right], \quad (33)
\]

\[
\sum_{m=0}^{L} Q_m x^{-m} = \frac{Q(1 - z)}{1 - \frac{xz^{L+1}}{1 - x}} \left[ \frac{x - z (z/x)^L}{x - z} \right], \quad (34)
\]

\[
\sum_{m=0}^{n} Q_m x^{-m} = \frac{Q(1 - z)}{1 - \frac{xz^{L+1}}{1 - x}} \left[ \frac{x - z (z/x)^n}{x - z} \right], \quad (35)
\]

\[
\sum_{m=n+1}^{l} Q_m x^{-m} = \frac{Q(1 - z)}{1 - \frac{xz^{L+1}}{1 - x}} \left[ \frac{(xz)^{n+1} - (xz)^{L+1}}{1 - xz} \right]. \quad (36)
\]

Substituting these expressions into equations (31) and (32), we obtain for the sites inside the source region

\[
P_{\xi}^i(n) = \frac{Q(1 - z)}{\sqrt{k^2 + 4uk (1 - z)}} \times \left[ \left( -k + \sqrt{k^2 + 4uk} \right) \frac{(x^{n+1} - xz^{n+2} - \frac{z^{L+1}x^{L+2+n}}{1-xz})}{(x-z)(1-xz)} \right]
\]

while for the outside region we have

\[
P_{\xi}^o(n) = \frac{Q(1 - z)}{\sqrt{k^2 + 4uk (1 - z)}} \times \left[ \left( -k + \sqrt{k^2 + 4uk} \right) \frac{(x^{n+1} - xz^n)}{(x-z)(1-xz)} \right]
\]

The corresponding expressions for LATs for the source region can be written as

\[
t_{\xi}(n) = \frac{n}{\sqrt{k^2 + 4uk}} \times \left[ \left( -k + \sqrt{k^2 + 4uk} \right) \frac{\Phi_{\xi}(n)}{(k + \sqrt{k^2 + 4uk})} \right]
\]
\[
\Delta_\varphi(n) = \left( k + \sqrt{k^2 + 4uk} \right)\Phi_\varphi(n) + \left( -k + \sqrt{k^2 + 4uk} \right) \left( x^{n+1} \left( 1 - (xz)^{l+1} \right) - (xz^n) (xz)^{l+1} \right) / \left( \Delta_\varphi(n) \right)
\]

where another set of auxiliary functions is introduced

\[
\Delta_\varphi(n) = \left( k + \sqrt{k^2 + 4uk} \right) \left( x^{n+1} - xz^{n+2} - z^{L+1}x^{L-n+1} + z^{L+2}x^{L-n+1} \right)
\]

Taking the limit of \( L \to \infty \) in equations (37) and (43) we obtain the expressions for the stationary-state profiles and the LATS used in the main text of the paper.

**References**

[1] Martinez-Arias A and Stewart A 2002 Molecular Principles of Animal Development (New York: Oxford University Press)

[2] Lodish H, Berk A, Kaiser C A, Krieger M, Scott M P, Bretscher A, Ploegh H and Matsudaira P 2007 Molecular Cell Biology, 6th edn. (New York: Freeman)

[3] Wollpert L 1998 Principles of Development (New York: Oxford University Press)

[4] Wollpert L 1969 Positional information and the spatial pattern of cellular differentiation J. Theor. Biol. 25 1

[5] Crick F H 1970 Diffusion in embryogenesis Nature 225 420

[6] Kerszberg M and Wolpert L 1998 Mechanisms for positional signalling by morphogen transport: a theoretical study J. Theor. Biol. 191 103

[7] Lande R A 2007 Morphozone: reimagining the morphogen gradient Cell 128 245

[8] Kornberg T B 2012 The imperatives of context and contour for morphogen dispersion Biophys J. 103 2252

[9] Rogers K W and Schier A F 2011 Morphogen gradients: from generation to interpretation Annu. Rev. Cell Dev. Biol. 27 377

[10] Tabata T and Takei Y 2004 Morphogens, their identification and regulation Development 131 703

[11] Porcher A and Dostatni N 2010 The bicoid morphogen system Curr. Biol. 20 R249

[12] Gregor T, Wieschaus E F, McGregor A P, Bialek W and Tank D W 2007 Stability and nuclear dynamics of the bicoid morphogen gradient Cell 130 141

[13] Kicheva A, Pantazis P, Bollenbach T, Kalaidzidis Y, Bittig T, Julicher F and Goulian M 2007 Kinetics of morphogen gradient formation Science 315 521
[14] Zhou S, Lo W C, Suhalim J L, Digman M A, Grattom E, Nie Q and Lander A D 2012 Free extracellular diffusion creates the Dpp morphogen gradient of the Drosophila wing disc *Current Biology* 22 668–75

[15] Yu S R, Burkhart M, Nowak M, Ries J, Petrasek Z, Scholpp S, Schwille P and Brand M 2009 Fgf8 morphogen gradient forms by a source–sink mechanism with freely diffusing molecules *Nature* 461 533

[16] Entchev E V, Schwabedissen A and Gonzales-Gaitan M 2000 Gradient formation of the TGF-beta homolog Dpp *Cell* 103 981

[17] Müller P, Rogers K W, Jordan B M, Lee J S, Robson D, Ramanathan S and Schier A F 2012 Differential diffusivity of Nodal and Lefty underlies a reaction-diffusion patterning system *Science* 336 721

[18] Spirov A, Fahmy K, Schneider M, Frei E, Noll M and Baumgartner S 2009 Formation of the bicoid morphogen gradient: an mRNA gradient dictates the protein gradient *Development* 136 605–14

[19] Drocco J A, Grimm O, Tank D W and Wieschaus E 2011 Measurement and perturbation of morphogen lifetime: effects on gradient shape *Biophys. J.* 101 1807

[20] Little S C, Tkacik G, Kneeland T B, Wieschaus E F and Gregor T 2011 The formation of the Bicoid morphogen gradient requires protein movement from anteriorly localized mRNA *PLoS Biol.* 9 e1000596

[21] Grimm O, Coppy M and Wieschaus E 2009 Modelling the Bicoid gradient *Development* 137 2253

[22] Dalessi S, Neves A and Bergmann S 2012 Modeling morphogen gradient formation from arbitrary realistically shaped sources *J. Theor. Biol.* 294 130

[23] Deng J, Wang W, Lu L J and Ma J 2010 A two-dimensional simulation model of the Bicoid gradient in Drosophila PLoS Biol. 5 e10275

[24] Drocco J A, Wieschaus E F and Tank D W 2012 The synthesis-diffusion-degradation model explains Bicoid gradient formation in unfertilized eggs *Phys. Biol.* 9 055004

[25] Berezhkovskii A M, Sample C and Shvartsman S Y 2010 How long does it take to establish a morphogen gradient? *Biophys. J.* 99 1–59

[26] Berezhkovskii A M 2011 Ordinary differential equation for local accumulation time *J. Chem. Phys.* 135 074112

[27] Berezhkovskii A M and Shvartsman S Y 2011 Physical interpretation of mean local accumulation time of morphogen gradient formation *J. Chem. Phys.* 135 154115

[28] Berezhkovskii A M, Sample C and Shvartsman S Y 2011 Formation of morphogen gradients: local accumulation time *Phys. Rev. E* 83 051906

[29] Müller P, Rogers K W, Yu S R, Brand M and Schier A F 2013 Morphogen transport *Development* 140 1621

[30] Medioni C, Mowry K and Bess F 2012 Principles and roles of mRNA localization in animal development *Development* 139 3263

[31] Kolomeisky A B 2011 Formation of a morphogen gradient: acceleration by degradation *J. Phys. Chem. Lett.* 2 1502

[32] Teimouri H and Kolomeisky A B 2014 Development of morphogen gradient: the role of dimension and discreteness *J. Chem. Phys.* 140 085102

[33] Gordon P V, Muratov C B and Shvartsman S Y 2013 Local accumulation times for source, diffusion, and degradation models in two and three dimensions *J. Chem. Phys.* 138 104121