BAFF is produced by astrocytes and up-regulated in multiple sclerosis lesions and primary central nervous system lymphoma

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We report that B cell–activating factor of the tumor necrosis factor (TNF) family (BAFF) is expressed in the normal human brain at ~10% of that in lymphatic tissues (tonsils and adenoids) and is produced by astrocytes. BAFF was regularly detected by enzyme-linked immunosorbent assay in brain tissue lysates and in normal spinal fluid, and in astrocytes by double fluorescence microscopy. Cultured human astrocytes secreted functionally active BAFF after stimulation with interferon-γ and TNF-α via a furin-like protease–dependent pathway. BAFF secretion per cell was manifold higher in activated astrocytes than in monocytes and macrophages. We studied brain lesions with B cell components, and found that in multiple sclerosis plaques, BAFF expression was strongly up-regulated to levels observed in lymphatic tissues. BAFF was localized in astrocytes close to BAFF-R–expressing immune cells. BAFF receptors were strongly expressed in situ in primary central nervous system (CNS) lymphomas. This paper identifies astrocytes as a nonimmune source of BAFF. CNS–produced BAFF may support B cell survival in inflammatory diseases and primary B cell lymphoma.
sils or adenoids (Fig. 2). Similar results were obtained with PPIA (Fig. 2) or with GAPDH (not depicted) as housekeeping gene.

The transcript level of BAFF was quantified in a series of other tissues. BAFF expression was detectable in heart, skeletal muscle, pancreas, kidney, skin with adipose tissue, and liver at similar levels as in the CNS (i.e., in the range of 0.03–0.1% PPIA). In the lung, the BAFF levels were slightly higher, reaching 0.18% PPIA. Although the present paper identifies astrocytes as BAFF producers in the CNS (Figs. 3 and 4), further studies might establish whether in other tissues resident macrophages (particularly in lung) or parenchymal cells are responsible for local BAFF transcription.

An alternative splice variant lacking exon 3 (ΔBAFF) has been identified in mouse and human myeloid cells (12). We detected ΔBAFF, which was less abundant than the full-length isoform, in all brain specimens as well as in lymphatic tissue (Fig. 1).

BAFF-expressing cells had the morphology of astrocytes. They were found in three normal brains in the parenchyma and in the perivascular area both in the gray and in the white matter (Fig. 3, A–D). BAFF+ cells displayed long processes that extended to the perivascular area (Fig. 3 A).

In tonsils and adenoids, BAFF was detected on a few scattered cells without a clear distribution pattern. The frequency of BAFF-labeled cells differed considerably in different tonsil specimens. The labeled cells appeared in germinal centers (Fig. 3 O), T cell areas, and in some specimens also in the connective tissue (not depicted).

BAFF was detected in brain tissue by ELISA at a concentration of ~140 pg/mg protein lysate, whereas ~5 ng/mg
was observed in tonsils/adenoids. The amount of BAFF detected in normal brain tissue cannot be explained by blood contamination because in lysates of whole blood the BAFF concentration was ~10-fold lower than in brain tissue lysates.

10 paired noninflammatory serum/CSF samples were analyzed by ELISA. BAFF concentrations ranged from 153 to 845 pg/ml in CSF (mean ± SD: 483 ± 263 pg/ml) and from 2,257 to 6,197 pg/ml in serum (4,268 ± 1,295 pg/ml). No significant correlation was found between BAFF values in serum and CSF (R = 0.35, P = 0.31). The relatively high level of BAFF in CSF (on average 10% of the serum concentration) and the lack of correlation between serum and CSF content suggest that the BAFF present in the CSF is at least in part produced locally.

The functional relevance of the constitutive BAFF production in the CNS is presently unknown. Although no neurological deficits of BAFF-deficient mice have been described yet, this does not exclude subtle functional changes in the CNS.

**BAFF production by cultured human astrocytes**

Cultured human astrocytes constitutively contained a low level of BAFF transcripts (Fig. 4 A), which was increased by IFN-γ, to a lesser extent by TNF-α or IL-1β, but not by IL-10 (Fig. 4 B), GM-CSF, or IL-4 (not depicted). The strongest induction was seen with a combination of IFN-γ and TNF-α (Fig. 4 B). The amount of BAFF secreted by astrocytes paralleled the induction of transcription (Fig. 4 C). An inhibitor of furin-like proteases, decanoyl-Arg-Val-Lys-Arg-chloromethylketone, reduced the amount of secreted BAFF by 96% (unpublished data), indicating that astrocytes use furin-like proteases to release BAFF as do granulocytes (3). The biological activity of the astrocyte-produced BAFF was verified in a B cell proliferation assay (Fig. 4 E).

The level of BAFF released per cell after stimulation was much greater in astrocyte cultures than by activated monocytes (50 times) or macrophages (5 times; Fig. 4 D), which released in our study similar BAFF levels as described previously (2). To exclude the possibility that the BAFF levels in monocyte and macrophage cultures were artificially lowered by paracrine BAFF consumption of B cells, PBMCs were depleted of B cells before negatively selecting monocytes. FACS analysis showed that, in the purified monocyte cultures, essentially no BAFF binding cells could be detected.

**BAFF and BAFF receptors in MS lesions**

The transcript level of BAFF was ~10-fold higher in MS lesions as compared with normal CNS (P < 0.01), reaching approximately the same level as in lymphatic tissues (Fig. 2).

Immunostaining for BAFF was performed on 11 brain specimens from four MS patients. In each specimen, BAFF was immunolocalized on astrocytes (Fig. 3, E–N). In acute MS lesions, BAFF localized to astrocytes outside of the inflammatory cuffs (Fig. 3 M). In chronic MS lesions, numerous BAFF+ cells with the morphology of astrocytes were detected in both the perivascular area and in the parenchyma (Fig. 3, H–L and N). Double labeling for glial fibrillary acidic protein (GFAP) and BAFF identified BAFF-expressing cells as astrocytes in both acute (Fig. 3, E–G) and chronic MS lesions (Fig. 3, H–J). Confocal analysis revealed a dense network of BAFF+ astrocytes in the perivascular area and in the parenchyma of MS lesions (Fig. 3, H–J). Although the most sensitive staining was observed with the BAFF–specific mAb B1A3.2 (Fig. 3, A–C, E, G, H, J, K, M, and N), a similar astrocytic staining was observed with the BAFF-specific...
mAb A9C9.1 in normal brain (Fig. 3 D) and MS lesions (Fig. 3 L). BAFF-R was detected on a subset of inflammatory cells in the vicinity of BAFF-expressing astrocytes, mainly in the perivascular area (Fig. 3 P).

Our histological observation of enhanced BAFF expression on astrocytes in MS lesions is in harmony with the in vitro induction of BAFF on astrocytes by inflammatory cytokines. However, it should be noted that the histological detection of BAFF is presumably insensitive. For example, in mice, BAFF staining was unsuccessful (13), and in tonsils we detected BAFF only on a few scattered cells (Fig. 3 O) despite its abundant transcription. This indicates that, by immunohistochemical staining, the number and cell types expressing BAFF might be underestimated. Because macrophages are abundant in MS lesions and produce BAFF in vitro, the elevated BAFF expression in MS lesions might be due to BAFF produced by both astrocytes and macrophages. Our in vitro findings that appropriately stimulated astrocytes produce five times more BAFF per cell than highly activated macrophages might explain why in MS lesions BAFF is immunolocalized preferentially to astrocytes.

Our findings suggest the following scenario: BAFF produced by astrocytes promotes the survival of BAFF-R–expressing B cells in MS brains, allowing the persistence and clonal expansion of B cells in the CNS (7–9, 11). The oligoclonal Ig in the CSF are remarkably stable for many years (6), consistent with the long-term survival of local plasma cells. Recently, it has been shown that BCMA, a receptor for BAFF and APRIL, is critical for the long-term survival of plasma cells (14). Locally produced BAFF might contribute to plasma cell survival in the CNS. Indeed, our quantitative PCR indicated that BCMA was up-regulated in MS brain specimens (Fig. 5). Our conclusions are consistent with two animal studies. First, the normal brain microenvironment supports development of antigen-directed humoral immunity in rodents (15). Second, increased BAFF gene expression was detected in the CNS of animals with experimental autoimmune encephalomyelitis, although in this paper, neither BAFF-producing nor BAFF receptor–expressing cells were described previously (16).

Local BAFF production within MS lesions might also promote T cell activation because BAFF has also been recognized as a costimulator for T cells (17). The elevated expression of BAFF in MS lesions was not reflected by an increase of BAFF in the CSF (reference 18 and unpublished data). This resembles observations with CCL-2/MCP-1. This chemokine is constitutively present in the CSF and it is abundantly expressed in MS lesions, but its concentration in the CSF is even reduced in MS (19). Presumably, the ratio of production and consumption by inflammatory cells determines the CSF level.

BAFF and BAFF receptors in primary CNS lymphomas

Primary CNS lymphomas are extranodal lymphomas that are confined to the CNS in the absence of systemic disease. Primary CNS lymphomas are typically of B cell origin for still unknown reasons (5).

Although BAFF receptors had not been analyzed in primary CNS lymphomas before, we report that in these CNS tumors, the transcript levels of all three BAFF receptors greatly exceeded that in normal brain and reached the same level as in lymphatic tissue (Fig. 5). Immunohistochemical analysis detected BAFF-R in primary CNS lymphomas rather homogeneously (Fig. 3 Q). A clear BAFF-R staining was observed on tumor cells in six out of seven specimens of the CNS lymphomas. Expression of the three BAFF receptors in individual tumor samples was heterogeneous (Fig. 5). The PCR data paralleled the histological data. The tumor sample with the lowest BAFF-R transcript level was the only one of the seven specimens without a clear BAFF-R staining. This tumor showed the highest TACI expression and the second highest BCMA level of the specimens analyzed (Fig. 5).

BAFF was transcribed in each of the five analyzed tumor specimens, in three of them at a similar level as in normal CNS, and in two of them at an elevated level (Fig. 2). The CNS lymphoma tissue we investigated in this paper did not contain parenchyma surrounding the tumors. However,
based on our findings in normal brain and in inflammatory MS lesions, it seems plausible to assume that BAFF released by resident astrocytes supports the survival of BAFF-receptor–expressing B cell lymphomas, which are otherwise found in lymph nodes where BAFF is abundant. Primary CNS lymphomas reside at least partially behind an intact blood–brain barrier (5), implicating the importance of the CNS-derived BAFF as a survival factor. This local supply of BAFF together with the shielding of the brain microenvironment from the immune system might provide a favorable basis for tumor development. Recent animal studies are consistent with this conclusion; in the absence of TNF-α, BAFF–overexpressing mice develop lymphomas (20), showing that BAFF in combination with a reduced tumor immunosurveillance facilitates lymphoma development. Our findings have therapeutic implications by suggesting that it might be beneficial to inhibit the local availability of BAFF in and around the CNS lymphoma.

In conclusion, we have extended the spectrum of BAFF sources to astrocytes and demonstrate that BAFF is constitutively present in the CNS. In MS lesions, BAFF expression was elevated to levels of lymphatic tissues. Local BAFF production in the CNS might promote B cell survival in MS and primary CNS lymphoma.

MATERIALS AND METHODS

Tissue specimens, CSF, and serum. Control tissue specimens were obtained from autopsy material of persons who had died of acute accidents: normal brain including gray and white matter of the cerebrum and cerebellum (17 tissue blocks from four adult persons), heart, skeletal muscle, lung, liver, skin with adipose tissue, pancreas, and kidney. Tonsils and adenoids were obtained after operations. Eight brain lesions of six MS patients were examined by RT-PCR. One case was published (case 2 in reference 21), the other specimens were obtained from the Institute of Neurology in London (courtesy of L. Cuzner and J. Newcombe). They were classified as acute based on the presence of oil red O positive macrophages and perivascular infiltrates. Frozen brain specimens from 11 blocks of four MS patients were examined immunohistologically: two blocks from an acute lesion (patient 1; case 2 in reference 21), the other specimens were obtained from the Institute of Neurology in London (courtesy of L. Cuzner and J. Newcombe). They were classified as acute based on the presence of oil red O positive macrophages and perivascular infiltrates. Frozen brain specimens from 11 blocks of four MS patients were examined immunohistologically: two blocks from an acute lesion (patient 1; case 2 in reference 21), the other specimens were obtained from the Institute of Neurology in London (courtesy of L. Cuzner and J. Newcombe). They were classified as acute based on the presence of oil red O positive macrophages and perivascular infiltrates.

Cell culture. Human astrocytes of embryonic origin were devoid of microglial cells or macrophages (22). These cells were stimulated with 10 or 25 ng/ml TNF-α, 100 U/ml IFN-γ (Roche), 50 ng/ml IL-1β, and 100 ng/ml IL-10 (R&D Systems).

For the analysis of the biological activity of secreted BAFF, astrocytes were switched to serum-free medium (Pan-Biotech) and stimulated with IFN-γ + TNF-α for 3 d. The supernatant was concentrated by 200-fold and a B cell proliferation assay was done essentially as described previously (17).

Monocytes were isolated by negative selection from CD19-depleted PBMCs (both obtained from Milteny Biotech), differentiated to macrophages with 20 ng/ml M-CSF for a period of 9–11 d (2), and stimulated in new medium. BAFF-binding cells were identified by myc-BAFF, biotinylated anti-myc mAb, and streptavidin-PE. Cells were stimulated with 300 U/ml IFN-γ + 25 ng/ml TNF-α or 300 ng/ml LPS for 48 h.

PCR. Quantitative PCR and the identification of BAFF isoforms was performed as described in the supplemental material (available at http://www.jem.org/cgi/content/full/jem.20041674/DC1).

BAFF ELISA. The anti-human BAFF mAb clone B4H7.2 was used for coating, the biotin-labeled anti-human BAFF clone A9C9.1 was used for detection, and recombinant human BAFF was used as a standard (all obtained from Biogen Idec Inc.). To determine whether this ELISA can be used to detect BAFF content in tissue lysates, 400 pg/ml BAFF was spiked in a 1:50 and 1:20 dilution of the RIPA lysis buffer; >95% of the spiked BAFF was detected. Protein lysates of brain specimens and whole blood were obtained using RIPA buffer and measured in a dilution of at least 1:40.

Immunohistochemistry. Mabs recognizing BAFF (B1A3.2 and A9C9.1), BAFF-R (9–1; 5–10 μg/ml), and IgG1 control were used. The mAb B1A3.2 revealed a good staining with 20 μg/ml, the strongest BAFF-expressing cells could still be seen with 10 μg/ml. mAb A9C9.1 was used at 35 μg/ml. Stainings were made on PFA or formalin–fixed cryosections with the peroxidase antiperoxidase method or the avidin–biotin complex system (DakoCytomation). The autopic specimens of patient 4, which had been fixed in PFA before freezing, were processed as described previously (10).

For double immunofluorescence staining, mouse anti–BAFF mAb B1A3.2 and rabbit anti–GFAP were applied simultaneously, followed by donkey anti–mouse Alexa Fluor 488 or donkey anti–mouse biotin plus Cy2-conjugated streptavidin, and donkey anti–rabbit Alexa Fluor 594. Confocal images were taken on a TCS NT laser scanning microscope (Leica).

Online supplemental material. Materials and methods for the quantitative PCR and the identification of BAFF isoforms. Online supplemental material is available at http://www.jem.org/cgi/content/full/jem.20041674/DC1.

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