Atherosclerosis is responsible for coronary artery disease (CAD), which remains the leading cause of morbidity and mortality worldwide.1 One of the first steps in the atherosclerotic process is the deposition and retention of atherogenic lipoprotein particles in a susceptible coronary arterial vessel wall, followed by a reactive inflammatory process, smooth muscle cell proliferation, fibrosis, and calcification.2 On the other hand, lipoproteins associated with reverse cholesterol transport are able to remove excess cholesterol from macrophages in atherosclerotic plaques, providing an atheroprotective mechanism.

Lipoprotein particles are composed of lipid components such as cholesterol, cholesteryl-esters, phospholipids, and apoproteins.
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triglycerides and protein components such as apoprotein B (apoB), apoprotein A (apoA), apoprotein C, and apoprotein E. The central principle of the atherogenic dyslipidemia paradigm is that apoB-containing particles are atherogenic because of the physical binding of apoB to proteoglycans in the vessel wall. In contrast, apoAI-containing high-density lipoprotein (HDL) particles are atheroprotective by removing cholesterol from macrophages in the vessel wall and by preventing low-density lipoprotein (LDL) oxidation and the maladaptive inflammatory response. ApoAI is primarily synthesized in the liver and intestines and is secreted into circulation, where it undergoes a series of remodeling steps facilitated by a number of enzymes such as plasma lipid transfer protein, lecithin-cholesterol acyltransferase, and cholesterol ester transfer protein). Native apoAI gradually acquires phospholipid and becomes pre-β1-HDL, whereas further acquisition of cholesterol and apoAI facilitates remodeling to initially smaller, alpha-migrating α4-HDL. α4-HDL matures to more lipid-rich and larger α3-, α2-, and α1-HDL particles (Figure 1). The large, cholesterol-rich α2- and α1-HDL particles can unload excess cholesterol in the liver through the scavenger receptor–B1 pathway (Figure 1).

HDL subpopulations can be quantitatively measured and characterized by native 2D gel electrophoresis, immunodetection, and image analysis. Certain subpopulations have been shown to be associated with either lower or higher risk of coronary heart disease based on angiographic and outcomes studies.

Coronary atherosclerotic plaque can be characterized invasively by intravascular ultrasound (IVUS) with radio-frequency backscatter analysis (IVUS/VH). More recently, contrast-enhanced coronary computed tomography angiography (CTA) has been introduced as a noninvasive means for this purpose. Over past years, our laboratory has developed and validated a standardized, 3-dimensional quantitative method for CTA-based coronary plaque quantification and published its reproducibility as well as its accuracy.

However, the relationship between quantitative, 3-dimensional coronary plaque measurements by IVUS/VH and CTA and precise lipoprotein and HDL subpopulation measurements has not previously been evaluated. Therefore, the objective of our study was to study the association between circulating apoB- and apoAI-containing lipoprotein particles and quantitative coronary plaque composition as measured by IVUS/VH and CTA. We hypothesized that apoB-containing lipoprotein particles are associated with a higher-risk phenotype, whereas adequate maturation or dynamic remodeling of apoAI-containing HDL particles is associated with a lower-risk plaque phenotype, as measured by IVUS/VH and CTA.

Methods

General Study Design

This was a prospective, investigator-initiated, single-center study approved by the Institutional Review Board of Piedmont Healthcare; all patients provided written informed consent. The overall study design has been published previously. Briefly,
enrollment criteria were (1) being male or female 18 to 70 years old; (2) having no prior known obstructive CAD; (3) having symptoms suggestive of myocardial ischemia or a high probability of CAD; and (4) having ≥1 “intermediate” coronary artery stenosis present, defined by either CTA (40% to 70% luminal stenosis by CTA criteria) or invasive x-ray angiography (40% to 70% luminal stenosis by angiographic criteria), and lacking obstructive stenosis (>70% luminal stenosis) in the 4 major epicardial arteries. For the entire study, all patients underwent biomarker testing and CTA and IVUS/VH at baseline and then biomarker testing and CTA 12 months later. Blood samples were drawn either at the time of the invasive angiogram or immediately prior to CTA. For the purposes of the current analysis, we analyzed associations between biomarkers, CTA, and IVUS/VH at the baseline examination.

**CTA Image Acquisition**

All image acquisition was performed on a 32×2 CT system (Siemens Somatom 64; Erlangen, Germany) using our institutional imaging protocols as published before.11 Contrast-enhanced coronary artery CTA was performed during end-expiratory breath hold using retrospective ECG gating. Oral and intravenous metoprolol were administered as needed to keep the heart rate <60 beats per minute. After noncontrast localization image acquisition, a test bolus of 20 mL iodinated contrast (Visipaque, GE Amersham Health) was administered at a rate of 3 to 5 mL/s to determine the delay until the arrival of the contrast in the ascending aorta for optimal opacification of the coronary arteries. Coronary arterial image acquisition was performed using 60 to 80 mL of intravenous contrast followed by 30 mL of normal saline flush. Acquisition parameters included 32×2 detector rows, 0.6-mm collimation, gantry rotation of 330 ms, pitch of 0.24, tube voltage of 120 kV, and tube current of 800 to 950 mA. Images were reconstructed in 0.6-mm axial slices using 3 different reconstruction algorithms in 10 phases of the cardiac cycle.

**CTA Image Analysis**

Selected for image analysis in each patient was a target vessel segment that contained the preidentified intermediate coronary artery lesion (40% to 70% luminal stenosis) in the preenrollment CTA or x-ray angiography examination, as previously described.8,11 Starting and termination points of the target segment were precisely defined based on predeﬁned intermediate coronary artery stenosis calculations.

Within the vessel wall area of the target vessel segment, plaque composition was quantiﬁed based on predefined Hounsﬁeld unit (HU) thresholds. Plaque was classiﬁed as calciﬁed with a density >150 HU. Noncalciﬁed plaque (NCP) was divided into 2 categories: NCP with attenuation values <30 HU were classiﬁed as low-density NCP and NCP with attenuation values in the range of 31 to 150 HU was classiﬁed as high-density NCP. The total volume and the percentage of each plaque component were determined.

**IVUS/VH Image Acquisition**

After intracoronary injection of nitroglycerin (mean total dose per case, 561.5 μg; range, 0 to 1800 μg) and after placing a guiding catheter in the target coronary artery, a 3.2F, 20-mHz ultrasound catheter (Eagle Eye; Volcano Inc, Rancho Cordova, CA) was inserted and was advanced at least 2 cm beyond the most distal portion of the target lesion. Automated pullback was performed at a rate of 0.5 mm/s (R-100; Volcano Inc, Rancho Cordova, CA). The electrocardiographic signal was simultaneously recorded for the reconstruction of the radiofrequency backscatter information using In-Vision Gold (Volcano Inc, Rancho Cordova, CA).

**IVUS/VH Image Analysis**

Deidentiﬁed IVUS/VH data sets were transferred to a dedicated workstation and were analyzed using dedicated software (pcVH 3.0.394; Volcano Inc, Rancho Cordova, CA) by an experienced cardiologist (G.V.). The luminal boundary and the external elastic lamina were contoured in a semiautomatic
fashion in each frame. On the basis of a previously validated algorithm, the software classified each pixel as dense calcium (white), fibrous tissue (green), fibrofatty tissue (light green), and necrotic core (red). Total volume and percentages of each of the 4 components were measured in the study segment. Furthermore, we calculated the volume and percentages of all noncalcified plaque components (sum of necrotic core, fibrofatty tissue, and fibrous tissue).

**Lipoprotein Measurements**

Blood was collected from all participants at either the time of the index CTA examination or the invasive angiographic procedure, whichever occurred first. Plasma samples were separated by low-speed centrifugation, aliquoted, and frozen at −80°C within an hour. All samples were analyzed as a single batch in a core laboratory (Boston Heart Laboratory, Framingham, MA) to maintain consistency. Total cholesterol, LDL-C, HDL-C, and triglycerides were measured with standard enzymatic methods and apoB, apoAI, and Lp(a) by immunoturbimetric methods. HDL subpopulations, including pre-

Statistical Analysis

Continuous variables are expressed as mean±standard deviation and median (interquartile range). Pearson correlation coefficient (r) was determined using standard methods. Univariate associations between lipoprotein measurements and plaque parameters were estimated using linear regression models. Unless otherwise noted, relationships between lipoprotein measurements and plaque parameters were approximately linear. Adjusted associations were estimated using multivariable linear regression models that included age, sex, BMI, diabetes, and medication usage. For all analyses, P≤0.05 was considered statistically significant, with no adjustment for multiple comparisons.

**Results**

**General Demographic Parameters**

Overall, 60 patients were enrolled in the study (Table 1). In the entire cohort, mean age was 60.4±7.2 years; 60% were male. Although greater than 70% of patients enrolled had cardiovas-

| Characteristic       | Frequency, Value, or IQR |
|----------------------|--------------------------|
| Subjects             | n=60                    |
| Age, mean±SD         | 60.4±7.2                |
| Men (%)              | 36 (60%)                |
| Ethnicity            |                         |
| White                | 55 (91.7%)              |
| African American     | 5 (8.3%)                |
| Risk factors         |                         |
| Hypertension         | 50 (83.3%)              |
| Dyslipidemia         | 58 (96.7%)              |
| Tobacco use          | 35 (58.3%)              |
| Diabetes mellitus    | 44 (73.3%)              |
| Framingham Risk Score (10-year risk; mean±SD) | 8.2±6.1 |
| Symptoms             |                         |
| Chest pain           | 27 (45%)                |
| Shortness of breath  | 16 (26.7%)              |
| Medications          |                         |
| Aspirin              | 43 (71.7%)              |
| Beta-blocker         | 30 (50%)                |
| Angiotensin-converting enzyme inhibitor | 18 (30%) |
| Angiotensin receptor blocker | 6 (10%) |
| Nitrate              | 6 (10%)                 |
| Statin               | 40 (66.7%)              |
| Cholesterol absorption inhibitor | 15 (25%) |
| Niacin               | 9 (15%)                 |
| Fibric acid derivative | 3 (5%)               |
| Laboratory values    |                         |
| Total cholesterol, mg/dL | 154.6±47.9 (124 to 172) |
| ApoB, mg/dL          | 78.4±29.8 (59.7 to 91.5) |
| LDL cholesterol, mg/dL | 87.5±37.4 (59.2 to 107.1) |
| sd-LDL-C, mg/dL      | 14.8±10.8 (6.8 to 18.6) |
| ApoAI, mg/dL         | 141.3±31.6 (126.3 to 154.3) |
| HDL cholesterol, mg/dL | 37.7±10.9 (30.7 to 41.9) |
| Triglycerides, mg/dL | 127.3±60.9 (79.7 to 164) |

HDL indicates high-density lipoprotein; IQR, interquartile range; LDL, low-density lipoprotein; SD, standard deviation; sd-LDL, small, dense low-density lipoprotein.
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Table 2. ApoA-Containing HDL Subclass Distribution in Patients at Baseline

| HDL Particle | Absolute Value, SD, and IQR (mg/dL) | Percentage, SD, and IQR |
|--------------|-----------------------------------|------------------------|
| pre-β1-HDL   | 18.8±7.9 (12.4 to 23.4)           | 13.0±4.2 (10 to 15.7) |
| pre-β2-HDL   | 2.0±1.4 (0.9 to 2.6)              | 1.4±1.0 (0.6 to 1.9)  |
| pre-α1-HDL   | 2.9±3.0 (1.0 to 3.7)              | 1.9±1.9 (0.7 to 2.4)  |
| α1-HDL       | 13.2±9.2 (6.7 to 18.1)            | 8.9±5.3 (5.1 to 11.9) |
| pre-α2-HDL   | 5.1±3.1 (3.1 to 6.6)              | 3.5±1.8 (2.5 to 4.6)  |
| α2-HDL       | 48.5±14.5 (38.6 to 62)            | 34.5±5.7 (30.2 to 37.2)|
| pre-α3-HDL   | 2.9±1.8 (1.8 to 3.2)              | 2.1±1.4 (1.3 to 2.4)  |
| α3-HDL       | 33.7±9.3 (26.8 to 41.5)           | 24.5±7.9 (18.4 to 28.2)|
| pre-α4-HDL   | 1.1±0.7 (0.6 to 1.5)              | 0.8±0.5 (0.4 to 0.9)  |
| α4-HDL       | 12.9±4.2 (9.7 to 15.5)            | 9.3±3.0 (7 to 11)     |

HDL indicates high-density lipoprotein; IQR, interquartile range; SD, standard deviation.

Biomarkers and Plaque Characteristics by CTA

Plaque geometrical and compositional parameters in the overall CTA cohort are shown in Table 3 and Figure 2A and 2B. Results of unbiased heat-map analysis are shown in Figure 3A. In general, apoB-related measurements and apoB-containing particles were associated with a shift in plaque composition from a lower proportion of calcified to a higher proportion of noncalcified plaque components (Table 4). In univariate analysis, apoB was associated with less calcified plaque (r=-0.33, P=0.02), more high-density NCP (r=0.31, P<0.03), and more low-density NCP (r=0.29, P=0.04); see Figure 4. sd-LDL-C was associated with a smaller percentage of calcified plaque (r=-0.30, P=0.03) and a higher percentage of high-density NCP (r=0.30, P=0.03), whereas triglycerides were associated with a higher percentage of low-density NCP (r=0.34, P=0.01).

Smaller, lipid-poor HDL particles (pre-α4-HDL, pre-α3-HDL, and α3-HDL) were associated with a lower proportion of calcified and a higher proportion of noncalcified plaque components (for high-density NCP%: pre-α3-HDL, r=0.40, P<0.01; pre-α4-HDL, r=0.29, P=0.04; for low-density NCP%: α3-HDL%, r=0.32, P=0.02; Table 4; Figure 5).

Larger, lipid-rich HDL particles (α2-HDL and α1-HDL) were associated with a lower proportion of low-density NCP% (α2-HDL%, r=-0.34, P=0.02; α1-HDL%, r=-0.28, P=0.05). Also, overall HDL-C measurements were associated with a lower proportion of low-density NCP on univariate analysis (r=-0.34, P=0.01; Table 4; Figure 5).

Finally, pre-β2-HDL particles were associated with lower volume of calcified plaque (r=-0.29, P=0.04) and less stenosis (minimal luminal diameter, r=-0.30, P=0.03; percent diameter stenosis, r=-0.30, P=0.03) see Table 5 and Figure 6. Importantly, several of the lipoproteins remained significant in the multivariable analysis after adjusting for age, sex, BMI, diabetes, and medications (Table 4).

Table 3. CTA Characteristics of Coronary Atherosclerotic Plaques

| CTA Parameter   | Value, SD, IQR |
|-----------------|----------------|
| MLD, mm         | 1.6±0.6 (1.2 to 1.9) |
| % Diameter stenosis | 47.1±16.7 (34.5 to 61.5) |
| MLA, mm²        | 5.5±2.5 (3.7 to 6.7) |
| Area stenosis, % | 49.9±18.0 (38.5 to 60.5) |
| Lesion length, mm | 27.9±14.2 (18.9 to 35.3) |
| Lesion plaque volume, mm³ | 234.0±132.1 (138.5 to 294.5) |
| Calcified plaque volume, mm³ | 75.1±70.2 (19.7 to 130.8) |
| High-density NCP volume, mm³ | 139.7±73.7 (93.4 to 170.2) |
| Low-density NCP volume, mm³ | 19.2±12.7 (11.4 to 22) |
| Calcified plaque, % | 28.9±17.8 (12.6 to 43.5) |
| High-density NCP, % | 62.6±16.1 (50.5 to 77.1) |

CTA indicates computed tomography angiography; IQR, interquartile range; MLA, minimal lumen area; MLD, minimal luminal diameter; NCP, noncalcified plaque; SD, standard deviation.

Biomarkers and Plaque Characteristics by IVUS/VH

Plaque geometrical and compositional parameters in the overall IVUS/VH cohort are shown in Table 5. Results of unbiased heat-map analysis are shown in Figure 3B. Of the apoB-containing particles, Lp(a) was associated with larger plaque volume by IVUS/VH both in univariate analysis (r=0.42, P=0.01) and in multivariable analysis (β=2.7, P=0.03), mostly because of higher volume of fibrous tissue (r=0.38, P=0.02; β=1.0, P=0.08). In addition, LDL-C was also associated with a higher volume of fibrous tissue (r=0.34, P=0.04; Table 6).

Smaller, lipid-poor HDL particles (α4-HDL and α3-HDL) were associated with higher plaque volume (α4-HDL%, univariate r=0.41, P=0.01; multivariable β=26.4, P=0.02; α3-HDL%, r=0.37, P=0.03). Percentage of α4-HDL was associated with higher volume of dense calcium (r=0.37, P=0.03), higher volume of fibrous tissue (r=0.34, P=0.04), and higher volume of necrotic core (univariate r=0.46, P=0.01; multivariable β=5.1, P=0.02). Percentage of α3-HDL was...
associated with higher volume of necrotic core ($r=0.37$, $P=0.03$; Table 6; Figure 5).

On the other hand, larger, lipid-rich HDL particles (pre-$\alpha$2-HDL, $\alpha$2-HDL, pre-$\alpha$1-HDL, and $\alpha$1-HDL particles) were associated with lower plaque volume (Table 6; Figure 5), mostly because of a lower volume of dense calcium (pre-$\alpha$2-HDL, $r=-0.33$, $P=0.05$), lower volume of fibrous tissue (pre-$\alpha$1-HDL, univariate $r=-0.39$, $P=0.02$; multivariable $\beta=-11.9$, $P<0.01$; $\alpha$1-HDL, $r=-0.37$, $P=0.03$), fibrofatty tissue (pre-$\alpha$1-HDL, $r=-0.34$, $P=0.05$), and, importantly, lower volume of necrotic core ($\alpha$1-HDL, $r=-0.42$, $P<0.01$; pre-$\alpha$2-HDL, $r=-0.38$, $P=0.02$; $\alpha$2-HDL, $r=-0.35$, $P=0.04$; and pre-$\alpha$1-HDL, $r=-0.34$, $P=0.04$).

Of the $\beta$-migrating HDL particles, pre-$\beta$2-HDL particles were associated with a shift in plaque composition to a lower proportion of necrotic core (univariate $r=-0.44$, $P<0.01$;
Figure 2. Example of CTA and IVUS/VH analysis of coronary arterial lesions. Long-axis (A) and short-axis (B) views of coronary plaques by CTA, demonstrating the lumen (light green), high-density noncalcified plaque (blue), and low-density noncalcified plaque (red). Corresponding long-axis (C) and short-axis (D) views of the same lesion by IVUS/VH, demonstrating fibrous tissue (dark green), fibrofatty tissue (light green), necrotic core (red), and calcium (white). CTA indicates computed tomography angiography; IVUS/VH, intravascular ultrasound virtual histology.

multivariable $\beta=-1.6$, $P=0.03$) and dense calcium (univariate $r=-0.44$, $P<0.01$; multivariable $\beta=-1.6$, $P<0.03$) to a higher proportion of fibrous tissue (univariate $r=0.53$, $P<0.01$; multivariable $\beta=3.47$, $P<0.01$). Finally, pre-$\beta$-2-HDL particles were associated with a lower percentage of area stenosis ($r=-0.32$, $P=0.03$) and a lower percentage of diameter stenosis ($r=-0.30$, $P=0.05$); see Table 6 and Figure 6. Importantly, several of the lipoproteins remained significant in the multivariable analysis after adjusting for age, sex, BMI, diabetes, and medications (Table 4).

Discussion

In the current study, we used a unique, standardized, quantitative CTA postprocessing method that we have validated in the past both for reproducibility,8,10 and for accuracy.11 Similarly, we used an IVUS/VH method that has also been validated in the past by our laboratory11 and by other laboratories.12 We also measured lipids and lipoproteins, including sd-LDL-C and the apoA1-containing HDL subpopulations with a previously validated methodology.3–5

The present study has 2 significant aspects. First, circulating levels of lipoprotein particles were mostly associated with noncalcified plaque, particularly low-density-NCP on CTA, which is the presumed lipid-rich component.11,13–15 This is important because it provides external validation that noncalcified regions of atherosclerotic plaques may be directly related to deposition of circulating lipoproteins. Second, the results of CTA-based and IVUS/VH-based plaque characteristics were essentially identical, providing further external validation of the findings.

Our study has 4 important findings. First, apoB-containing lipoprotein particles were associated with a higher proportion of noncalcified and a lower proportion of calcified plaque components. On CTA, sd-LDL-C was associated with a higher proportion of high-density NCP and triglycerides with a higher proportion of low-density NCP, and on IVUS, Lp(a) was associated with a higher plaque volume, mostly because of more fibrous tissue. Second, small, dense, lipid-poor apoA-containing HDL particles were associated with a higher proportion of noncalcified and a lower proportion of calcified components. On CTA, pre-$\alpha$-4-HDL and pre-$\alpha$-3-HDL were associated with a higher proportion of noncalcified and a lower proportion of calcified components. On CTA, pre-$\alpha$-4-HDL and pre-$\alpha$-3-HDL were associated with a larger plaque volume because of a higher volume of dense calcium ($\alpha$-4-HDL), a higher volume of fibrous tissue ($\alpha$-4-HDL), and importantly, a higher volume of necrotic core ($\alpha$-4-HDL, $\alpha$-3-HDL). Third, large, lipid-rich apoA-containing HDL particles were associated with less low-density NCP on CTA and with lower plaque volume on IVUS because of a lower volume of dense calcium (pre-$\alpha$-2-HDL), a lower volume of fibrous tissue (pre-$\alpha$-1-HDL, $\alpha$-1-HDL), a lower volume of fibrofatty tissue (pre-$\alpha$-1-HDL), and importantly, a lower volume of necrotic core (pre-$\alpha$-2-HDL, $\alpha$-2-HDL, pre-$\alpha$-1-HDL, $\alpha$-1-HDL). Finally, pre-$\beta$-2-HDL particles were associated with less calcification and less stenosis; on IVUS, pre-$\beta$-2-HDL was associated with a lower proportion of dense calcium and necrotic core and a higher proportion of fibrous tissue.

Our working model of HDL maturation/remodeling is summarized in Figure 1. The major apolipoprotein component of HDL, apoA1, is synthesized in the liver and in the small intestine and is secreted to the circulation in a lipid-free form. ApoA1 gains some phospholipids in the circulation and turns into pre-$\beta$-1-HDL. Pre-$\beta$-1-HDL is capable of promoting cholesterol efflux from resident macrophages in atherosclerotic plaques via the ABCA1 pathway, resulting in the formation of $\alpha$-4-HDL particles, which are still poorly lipidated, discoidal particles.16 Through multiple steps, several enzymes including lecithin-cholesterol acyltransferase, lipoprotein lipase (LPL),

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Figure 3. Heat plots illustrating the unbiased grouping of biomarkers in relation to plaque composition by CTA (A) and IVUS/VH (B). On CTA (A), there are 4 major terms. Term 1 contains small HDL particles and is associated with more low-density NCP and more stenosis. Term 2 contains apoA, HDL-C, and large HDL particles and is associated with less low-density NCP and more calcification and less stenosis. Term 3 contains apoB-containing particles and small HDL particles and is associated with more low-density NCP and less calcification. Term 4 contains large HDL particles and is associated with less low-density NCP, less calcified plaque, and less stenosis. On IVUS/VH (B), there are 2 major terms. Term 1 contains apoB-containing particles and small HDL particles and is associated with more necrotic core, larger plaque, and more stenosis. Term 2 contains apoA, HDL-C, and large HDL particles and is associated with less necrotic core and less calcium, more fibrous tissue, and smaller plaque. AS indicates area stenosis; CAP, calcified plaque; CTA, computed tomography angiography; DC, dense calcified; DS, diameter stenosis; FF, fibrofatty; FI, fibrous tissue; HDL, high-density-lipoprotein; HD-NCP, high-density noncalcified plaque; IVUS/VH, intravascular ultrasound virtual histology; LDL, low-density lipoprotein; LD-NCP, low-density noncalcified plaque; Lp(a), lipoprotein a; MLA, minimal lumen area; MLD, minimal lumen diameter; NC, necrotic core; NCP, noncalcified plaque; TC, total cholesterol; TG, triglycerides; sd-LDL, small dense low-density lipoprotein.

hepatic lipase (HL), cholesterol ester transfer protein, and plasma lipid transfer protein transform α-4 HDL particles into larger, more lipiddated, spherical HDL particles (α3-, α2-, and α1-HDL with increasing size, respectively). Intermediate-size α3-HDL particles are able to promote more cell-cholesterol efflux via the ABCG1 pathway. The 2 largest HDL subpopulations, α1- and α2-HDL, are able to promote bidirectional cell-cholesterol flux via the scavenger receptor–B1 pathway. Whether these large HDL particles are cholesterol donors or acceptors depends on the lipid status of the macrophages. Cholesterol-loaded cells donate cholesterol to these HDL particles, whereas cholesterol-depleted cells acquire cholesterol from them. Cholesterol uptake by αβ1-HDL particles via the ABCA1 pathway is the first step and selective cholesterol download by α1-HDL and α2-HDL particles via the liver scavenger receptor–B1 pathway the last step in direct reverse cholesterol transport.

In this context, our study found that higher levels of smaller, immature HDL particles (pre-α4-HDL, α4-HDL, pre-α3-HDL, and α3-HDL) were associated with a higher risk plaque phenotype both by CTA and IVUS/VH, as evidenced by more NCP, a higher plaque volume, and more necrotic core, whereas the larger, more “mature” forms of HDL (pre-α2-HDL, α2-HDL, pre-α1-HDL, and α1-HDL) were associated with a less high-risk phenotype, as evidenced by less low-density NCP, a lower plaque volume, and less necrotic core. Finally, higher levels of pre-β2-HDL were associated with less stenosis and less necrotic core. Efficient reverse cholesterol transport is associated with the removal of more cholesterol from the vessel wall, leaving behind less lipid-rich and more calcified plaque. On the other hand, inefficient HDL maturation and/or HDL remodeling, marked by accumulation of the smaller, lipid-poor (α3- and α4-HDL) and low levels of the larger, lipid-rich (α1-HDL and α2-HDL) HDL subclasses, are associated with decreased reverse cholesterol transport and more lipid-rich atherosclerotic plaque.

Although the present study is the first to implement previously validated, quantitative plaque phenotyping with a combination of CTA and IVUS/VH and detailed lipoprotein analysis, such as 2D gel for HDL phenotyping, our findings are consistent with previously published data. Cheng et al showed that increasing levels of LDL-C were associated with more noncalcified and less calcified plaque on CTA, similar to our findings. Furthermore, Shiga et al showed that lower HDL levels were associated with more CTA-defined CAD in both statin-naive and statin-treated patients.

With regard to the role of HDL subclasses, our study is consistent with previously published data and expands on...
Figure 4. Correlation between CTA-derived low-density noncalcified plaque, apoB, and HDL-C. Increasing levels of apoB are associated with a shift from calcified (A and C) to more low-density noncalcified (B and D) plaque. Conversely, increasing levels of HDL-C are associated with less low-density noncalcified plaque (B and D). CTA indicates computed tomography angiography; HDL, high-density lipoprotein cholesterol; LD-NCP, low-density noncalcified plaque.

Figure 5. Correlation of small, dense, lipid-poor versus large, lipid-rich HDL particles and low-density noncalcified plaque on CTA and necrotic core on IVUS/VH. Increasing levels of small, dense, cholesterol-poor α3-HDL particles are associated with increasing amounts of low-density noncalcified plaque (B and E) and less calcified plaque (A) by CTA and with increasing amounts of necrotic core by IVUS/VH (C and D). Conversely, increasing levels of larger, cholesterol-rich α1-HDL particles are associated with less low-density noncalcified plaque by CTA (A and F) and less necrotic core by IVUS/VH (G and H). CTA indicates computed tomography angiography; HDL, high-density lipoprotein; IVUS/VH, intravascular ultrasound virtual histology; LD-NCP, low-density noncalcified plaque; NC, necrotic core.
those findings. Invasive coronary angiographic data from the HDL Atherosclerosis Treatment Study showed in 123 subjects that simvastatin-niacin treatment was associated with an increase in α1- and pre-α1-HDL and a decrease in pre-β1-HDL and α3-HDL, and these changes were associated with less angiographic atherosclerosis progression. These findings are consistent with our findings of α1- and pre-α1-HDL associated with more benign and pre-β1-HDL and α3-HDL with less benign plaque phenotype by both CTA and IVUS/VH. Furthermore, Asztalos et al demonstrated in the Framingham Offspring Study that high levels of α1 and low levels of pre-β1- and α3-HDL were associated with less coronary heart disease, consistent with our findings. Finally, Asztalos et al also examined the association of HDL subpopulations with coronary events in the VA-HIT study in 398 subjects with and in 1907 subjects without recurrent events, showing that α1- and α2-HDL were associated with less and α3-HDL with more coronary heart disease events, consistent with our findings. These results taken together indicate that an inefficient remodeling of HDL particles may be associated with the removal of less cholesterol from atherosclerotic plaques, leaving behind more lipid-rich plaques, resulting in more progression of atherosclerosis and more cardiovascular events.

Our findings are also consistent with results from a recent publication by Khera et al, demonstrating that cholesterol efflux capacity, measured by a validated ex vivo essay, was an independent predictor of carotid artery wall thickness as measured by carotid intima-media thickness. Furthermore, efflux capacity was also an independent predictor of coronary disease status by invasive coronary angiography. Importantly, although efflux capacity was associated with apoAI and HDL-C levels, it remained an independent predictor even after adjusting for apoAI and HDL-C levels. Whereas high pre-β1 is a good acceptor of cholesterol from cells in ex vivo experiments, high pre-β1 level in vivo is a marker of inadequate HDL maturation and remodeling. Moreover, high

| IVUS/VH Parameter                  | Value, SD, IQR |
|-----------------------------------|----------------|
| Segment length, mm                | 63.0±18.9 (52.4 to 79) |
| Plaque volume, mm³                | 433.6±178.1 (350.3 to 518.4) |
| Plaque burden, %                  | 46.9±8.1 (43 to 51.8) |
| MLD, mm                           | 1.76±0.42 (1.5 to 2.0) |
| Dense calcium volume, mm³         | 35.4±28.9 (15.4 to 48.3) |
| Fibrous tissue volume, mm³        | 125.6±74.1 (56 to 144) |
| Fibrofatty tissue volume, mm³     | 22.2±17.0 (10 to 26.8) |
| Necrotic core volume, mm³         | 62.2±35.8 (31.2 to 91) |
| Dense calcium, %                  | 13.8±7.5 (7.9 to 19.1) |
| Fibrous tissue, %                 | 52.2±10.0 (46.8 to 58.7) |
| Fibrofatty tissue, %              | 9.2±5.5 (5.3 to 10.8) |
| Necrotic core, %                  | 24.8±6.4 (21.3 to 28.9) |

IQR indicates interquartile range; IVUS/VH, intravascular ultrasound virtual histology; MLD, minimal luminal diameter; SD, standard deviation.

**Figure 6.** Pre-β2-HDL and plaque composition by CTA and IVUS/VH. On CTA, increasing levels of pre-β2-HDL particles are associated with less calcified plaque (A vs D and B) and smaller-diameter stenosis (A vs D and C). On IVUS/VH, increasing levels of pre-β2-HDL particles are associated with less dense calcium (E vs H and F) and less necrotic core (E vs H and G). CAP indicates calcified plaque; CTA, computed tomography angiography; DC, dense calcium; DS, diameter stenosis; HDL, high-density-lipoprotein; IVUS/VH, intravascular ultrasound virtual histology; LD-NCP, low-density noncalcified plaque; NC, necrotic core.
Table 6. Correlation Analysis Between Lipoproteins and IVUS-Derived Quantitative Coronary Atherosclerotic Plaque Parameters.

| IVUS/VH Parameters | Biomarker | Univariate r Value | Univariate P Value | Multivariable β-Coefficient | Multivariable P Value | Adjusted P Value |
|-------------------|-----------|--------------------|--------------------|------------------------------|-----------------------|------------------|
| **Plaque volume** | Lp(a)     | 0.42               | 0.01               | 2.7                          | 0.03                  | 0.03             |
|                   | α4-HDL%   | 0.41               | 0.01               | 26.4                         | 0.02                  | 0.02             |
|                   | α3-HDL%   | 0.37               | 0.03               | 8.8                          | 0.07                  | 0.07             |
|                   | Pre-α2-HDL| −0.33              | 0.05               | −33.4                        | 0.02                  | 0.02             |
|                   | α1-HDL%   | −0.41              | 0.01               | −12.1                        | 0.03                  | 0.03             |
|                   | α1-HDL    | −0.44              | <0.01              | −7.6                         | 0.03                  | 0.03             |
|                   | pre-α1-HDL| −0.45              | <0.01              | −47.0                        | <0.01                 | <0.01            |
|                   | Pre-α1-HDL| −0.48              | <0.01              | −33.4                        | <0.01                 | <0.01            |
| **Dense calcium volume** | α4-HDL% | 0.37               | 0.03               | 2.76                         | 0.15                  | 0.15             |
|                   | Pre-α2-HDL| −0.39              | 0.02               | −3.7                         | 0.13                  | 0.13             |
| **Fibrous tissue volume** | Lp(a) | 0.38               | 0.02               | 1.0                          | 0.08                  | 0.08             |
|                   | LDL-C     | 0.34               | 0.04               | 1.36                         | <0.01                 | <0.01            |
|                   | α4-HDL%   | 0.34               | 0.04               | 10.2                         | 0.04                  | 0.04             |
|                   | α1-HDL%   | −0.37              | 0.02               | −4.8                         | 0.05                  | 0.05             |
|                   | Pre-α1-HDL| −0.37              | 0.03               | −17                          | 0.01                  | 0.01             |
|                   | α1-HDL    | −0.37              | 0.03               | −2.8                         | 0.06                  | 0.06             |
|                   | Pre-α1-HDL| −0.39              | 0.02               | −11.9                        | <0.01                 | <0.01            |
| **Fibrofatty tissue volume** | Pre-α1-HDL% | −0.33              | 0.05               | −4.0                         | 0.01                  | 0.01             |
|                   | Pre-α1-HDL| −0.34              | 0.05               | −2.74                        | 0.01                  | 0.01             |
| **Necrotic core volume** | α4-HDL% | 0.46               | <0.01              | 5.1                          | 0.02                  | 0.02             |
|                   | α3-HDL%   | 0.37               | 0.03               | 1.22                         | 0.21                  | 0.21             |
|                   | Pre-α1-HDL| −0.34              | 0.04               | −4.01                        | 0.06                  | 0.06             |
|                   | α2-HDL    | −0.35              | 0.03               | −0.5                         | 0.30                  | 0.30             |
|                   | Pre-α2-HDL| −0.38              | 0.02               | −5.0                         | 0.08                  | 0.08             |
|                   | α1-HDL%   | −0.39              | 0.01               | −2.0                         | 0.06                  | 0.06             |
|                   | α1-HDL    | −0.42              | <0.01              | −1.3                         | 0.06                  | 0.06             |
| **Dense calcium, %** | Pre-α2-HDL| −0.33              | 0.05               | −0.5                         | 0.42                  | 0.42             |
|                   | Pre-β2-HDL%| −0.38              | 0.02               | −3.2                         | 0.01                  | 0.01             |
|                   | pre-β2-HDL| −0.44              | <0.01              | −2.2                         | 0.01                  | 0.01             |
| **Fibrous tissue, %** | pre-β2-HDL% | 0.53               | <0.01              | 3.47                         | <0.01                 | <0.01            |
|                   | pre-β2-HDL| 0.47               | <0.01              | 5.02                         | <0.01                 | <0.01            |
| **Necrotic core, %** | A2-HDL% | −0.27              | 0.11               | −0.21                        | 0.32                  | 0.32             |
|                   | Pre-β2-HDL%| −0.37              | 0.03               | −2.0                         | 0.06                  | 0.06             |
|                   | Pre-β2-HDL| −0.44              | <0.01              | −1.6                         | 0.03                  | 0.03             |
| **MLA** | Pre-α2-HDL| −0.33              | 0.05               | −0.31                        | 0.05                  | 0.05             |
|                   | Pre-α2-HDL%| −0.34              | 0.04               | −0.46                        | 0.04                  | 0.04             |
| **%AS** | Pre-β1-HDL| −0.32              | 0.03               | −0.59                        | 0.11                  | 0.11             |
| **%DS** | Pre-β1-HDL| −0.30              | 0.05               | −0.39                        | 0.15                  | 0.15             |

Adjusted P values represent adjusting for age, sex, BMI, diabetes, and medication use. One multivariable model was fit for each biomarker separately. AS indicates area stenosis; BMI, body mass index; DS, diameter stenosis; HDL, high-density lipoprotein; IVUS/VH, intravascular ultrasound virtual histology; LDL-C, low-density lipoprotein cholesterol; Lp(a), lipoprotein a; MLA, minimal lumen area.

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In conclusion, our study showed a significant association between plaque characteristics and coronary stenosis by both CTA and IVUS/VH, whereas efficient remodeling of HDL particles was associated with a less high-risk phenotype and, ultimately, with less luminal stenosis with higher levels of pre-β2-HDL. Determination of more detailed lipoprotein parameters such as apoB, sd-LDL-C, and HDL particles by 2D gel electrophoresis may provide incremental clinical value beyond the measurements of the standard fasting lipid profile.

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