Draft Genome Sequence for *Caulobacter* sp. Strain OR37, a Bacterium Tolerant to Heavy Metals

Sagar M. Utturkar, Annette Bollmann, Ryann M. Brzoska, Dawn M. Klingeman, Slava E. Epstein, Anthony V. Palumbo, Steven D. Brown

Graduate School of Genome Science and Technology, University of Tennessee, Knoxville, Tennessee, USA; Department of Biology, Northeastern University, Boston, Massachusetts, USA; Department of Microbiology, Miami University, Oxford, Ohio, USA; Biosciences Division, Oak Ridge National Laboratory, Oak Ridge, Tennessee, USA.

*Caulobacter* sp. strain OR37 belongs to the class *Alphaproteobacteria* and was isolated from subsurface sediments in Oak Ridge, TN. Strain OR37 is noteworthy due to its tolerance to high concentrations of heavy metals, such as uranium, nickel, cobalt, and cadmium, and we present its draft genome sequence here.

*Caulobacter* spp. are Gram-negative bacteria that undergo asymmetric cell division and differentiation to produce two types of daughter cells: a motile swarmer cell and a division-compatible stalked cell (1, 2). Mobile swarmer cells are able to exploit new niches and eventually develop a tubular stalk that aids in surface attachment and nutrient uptake (3). The unique cell division pattern and ease of manipulation make *Caulobacter* spp. a simple single-celled model system to study cell cycle progression events. *Caulobacter* spp. are able to grow in low-nutrient environments, utilize dilute carbon sources, and degrade aromatic ring compounds (4) and have also been shown to possess surprisingly high tolerance to uranium (5, 6).

*Caulobacter* sp. strain OR37 was isolated from a site located at the Field Research Center (FRC) in Oak Ridge, TN (7), that is contaminated with radionuclides and heavy metals. Metagenomic, proteomic, and other functional approaches have been used to gain insights into the complex biogeochemical processes at contaminated sites (8, 9, 10, 11). Targeted efforts have aimed to bring formerly uncultivated organisms in culture and explore their functional roles, abilities, and adaptations to different environments (12, 13). We have recently sequenced the genome for *Microbacterium laevaniformans* strain OR221, a bacterium tolerant to metals, nitrate, and low pH that was also isolated from the FRC site (14). Strain OR37 was notable because it was tolerant to the highest tested concentrations of uranium (>200 μM), nickel (>500 μM), cobalt (>50 μM), and cadmium (>50 μM), while being inhibited only by the lowest pH and the highest concentration of nitrate (>250 mM) (7). We generated a draft genome sequence for *Caulobacter* sp. strain OR37 to gain insights into its physiology.

Draft genome sequence data for the strain OR37 were generated using a hybrid approach of 454 FLX (15) and Illumina MiSeq (16) technologies essentially as described previously (17). The 454 data consisted of 442,553 reads and generated 104,442,508 bp. Quality-trimmed Illumina data (CLC Genomics Workbench, version 5.5.1) consisted of 6,006,496 reads with an average length of 126 bp. Trimmed Illumina reads were assembled with CLC Genomics Workbench, and consensus sequences were distributed into 1.5-kbp overlapping fake reads using the fb_dice.pl script from the FragBlast module (http://www.clarkfrancis.com/codes/fb_dice.pl). The Newbler application (version 2.6, 454; Life Sciences) was used to generate a hybrid assembly from shredded Illumina reads and 454 data. The hybrid assembly consisted of 70 large (≥500 bp) contigs, with a total genome size of 4.4 Mb. The average contig size is 63,292 bp, with the largest contig being 604,995 bp, and the genome has an overall estimated G+C content of 67.7%.

The draft genome was annotated at the Joint Genome Institute (JGI) Integrated Microbial Genomes (IMG) database and comparative analysis system (18), which predicted 4,138 candidate protein-encoding gene models for *Caulobacter* sp. strain OR37. A number of heavy metal-translocating P-type ATPases, heavy metal efflux pumps, and metal ion efflux proteins were predicted, which might contribute toward its heavy metal resistance phenotype. This draft genome sequence will enable comparative genomics analysis and help to identify the genes responsible for heavy metal tolerance.

**Nucleotide sequence accession numbers.** This Whole-Genome Shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession no. APMP00000000. The version described in this paper is the first version, accession no. APMP00000000.

**ACKNOWLEDGMENTS**

For strain and DNA requests, please contact A.B.

Support for this work was provided by the U.S. Department of Energy Office of Science (BER) and DOE grant DE-FG02-04ER63782 to S.E.E. The Oak Ridge National Laboratory is managed by UT-Battelle, LLC, for the U.S. Department of Energy under contract DE-AC05-00OR22725.

Received 19 April 2013  Accepted 26 April 2013  Published 6 June 2013

Citation Utturkar SM, Bollmann A, Brzoska RM, Klingeman SE, Palumbo AV, Brown SD. 2013. Draft genome sequence for *Caulobacter* sp. strain OR37, a bacterium tolerant to heavy metals. Genome Announc. 1(3):e00322-13. doi:10.1128/genomeA.00322-13.

Copyright © 2013 Utturkar et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to Steven D. Brown, brownsd@ornl.gov, or Annette Bollmann, bollmaa@miamioh.edu.

© 2013 Utturkar et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

© 2013 Utturkar et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.
REFERENCES

1. Martin ME, Brun YY. 2000. Coordinating development with the cell cycle in Caulobacter. Curr. Opin. Microbiol. 3:589–595.

2. Stovepoindexter JL, Cohen-Bazire G. 1964. The fine structure of stalked bacteria belonging to the family Caulobacteraceae. J. Cell Biol. 23: 587–607.

3. Wagner JK, Brun YY. 2007. Out on a limb: how the Caulobacter stalk can boost the study of bacterial cell shape. Mol. Microbiol. 64:28–33.

4. Nierman WC, Feldblum TV, Laub MT, Paulsen IT, Nelson KE, Eisen JA, Heidelberg JF, Alley MR, N. Maddock JR, Potocka I, Nelson WC, Newton A, Stephens C, Phadke ND, Ely B, DeBoy RT, Dodson RJ, Durkin AS, Gwinn ML, Haft DH, Kolonay JF, Smit J, Graven MB, Khouri H, Shetty J, Berry K, Utterback T, Tran K, Wolf A, Vamathevan J, Ermolaeva M, White O, Salzberg SL, Venter JC, Shapiro L, Fraser CM, Eisen J. 2001. Complete genome sequence of Caulobacter crescentus. Proc. Natl. Acad. Sci. U. S. A. 98:4136–4141.

5. Hillson NJ, Hu P, Andersen GL, Shapiro L. 2007. Caulobacter crescentus as a whole-cell uranium biosensor. Appl. Environ. Microbiol. 73: 7615–7621.

6. Hu P, Brodie EL, Suzuki Y, McAdams HH, Andersen GL. 2005. Whole-genome transcriptional analysis of heavy metal stresses in Caulobacter crescentus. J. Bacteriol. 187:8437–8449.

7. Bollmann A, Palumbo AV, Lewis K, Epstein SS. 2010. Isolation and physiology of bacteria from contaminated subsurface sediments. Appl. Environ. Microbiol. 76:7413–7419.

8. Hemme CL, Deng Y, Gentry TJ, Fields MW, Wu L, Barua S, Barry K, Tringe SG, Watson DB, He Z, Hazen TC, Tiedje JM, Rubin EM, Zhou J. 2010. Metagenomic insights into evolution of a heavy metal-contaminated groundwater microbial community. ISME J. 4:660–672.

9. Lloyd JR, Renshaw JC. 2005. Bioremediation of radioactive waste: radionuclide-microbe interactions in laboratory and field-scale studies. Curr. Opin. Biotechnol. 16:254–260.

10. Waldron PJ, Wu L, Van Nostrand JD, Schadt CW, He Z, Watson DB, Jardine PM, Palumbo AV, Hazen TC, Zhou J. 2009. Functional gene array-based analysis of microbial community structure in groundwaters with a gradient of contaminant levels. Environ. Sci. Technol. 43: 3529–3534.

11. Wilkins MJ, Verberkmoes NC, Williams KH, Callister SJ, Mouser PJ, Elizaint H, N’Guessan AI, Thomas BC, Nicora CD, Shah MB, Abraham P, Lipton MS, Lovley DR, Hettich RL, Long PE, Banfield JF. 2009. Proteogenomic monitoring of Geobacter physiology during stimulated uranium bioremediation. Appl. Environ. Microbiol. 75:6591–6599.

12. Handelsman J. 2004. Metagenomics: application of genomics to uncultured microorganisms. Microbiol. Mol. Biol. Rev. 68:669–683.

13. Rappé MS, Giovannoni SJ. 2003. The uncultured microbial majority. Annu. Rev. Microbiol. 57:369–394.

14. Brown SD, Palumbo AV, Panikov N, Ariyawansa T, Klingeman DM, Johnson CM, Land ML, Utturkar SM, Epstein SS. 2012. Draft genome sequence for Microbacterium laevaniformans strain OR221, a bacterium tolerant to metals, nitrate, and low pH. J. Bacteriol. 194:3279–3280.

15. Margulies M, Egholm M, Altman WE, Attiya S, Bader JS, Bemben LA, Berka J, Braverman MS, Chen YJ, Chen Z, Dewell SB, Du L, Fierro JM, Gomes XY, Godwin BC, He W, Helgesen S, Ho CH, Izryk GP, Jando SC, Alenquer ML, Jarvie TP, Jirage KB, Kim JB, Knight JR, Lanza JR, Leamon JH, Lefkowitz SM, Lei M, Li J, Lohman KL, Lu H, Makhljani VB, Mckade KE, Mckenna MP, Myers EW, Nickerson E, Nobile JR, Plant R, Puc BP, Ronan MT, Roth GT, Sarkis GJ, Simons JF, Simpson JW, Srinivasan M, Tartar KR, Tomas A, Vogt KA, Volkmer GA, et al. 2005. Genome sequencing in microfabricated high-density picolitre reactors. Nature 437:376–380.

16. Loman NJ, Misra RV, Dallman TJ, Constantinidou C, Gharbia SE, Wain J, Pallen MJ. 2012. Performance comparison of benchtop high-throughput sequencing platforms. Nat. Biotechnol. 30:434–439.

17. Elkins JG, Lochner A, Hamilton-Brehm SD, Davenport KW, Podar M, Brown SG, Land ML, Hauger JI, Klingeman DM, Ramon B, Goodwin LA, Tapia R, Meinder L, Detter JC, Bruce DC, Han CS, Palumbo AV, Cottingham RW, Keller M, Graham DE. 2010. Complete genome sequence of the cellulolytic thermophile Caldicellulosiruptor obsidiansis OB47T. J. Bacteriol. 192:6099–6100.

18. Markowits VM, Chen IM, Palaniappan K, Chu K, Szeto E, Grechkin Y, Ratner A, Jacob B, Huang J, Williams P, Huntemann M, Anderson I, Mavromatis K, Ivanova NN, Kyrpides NC. 2012. IMG: the integrated microbial genomes database and comparative analysis system. Nucleic Acids Res. 40:D115–D122.