Research Article

In Vitro Neural Differentiation of Bone Marrow Mesenchymal Stem Cells Carrying the FTH1 Reporter Gene and Detection with MRI

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Magnetic resonance imaging (MRI) based on the ferritin heavy chain 1 (FTH1) reporter gene has been used to trace stem cells. However, whether FTH1 expression is affected by stem cell differentiation or whether cell differentiation is affected by reporter gene expression remains unclear. Here, we explore the relationship between FTH1 expression and neural differentiation in the differentiation of mesenchymal stem cells (MSCs) carrying FTH1 into neuron-like cells and investigate the feasibility of using FTH1 as an MRI reporter gene to detect neurally differentiated cells. By inducing cell differentiation with all-trans retinoic acid and a modified neuronal medium, MSCs and MSCs-FTH1 were successfully differentiated into neuron-like cells (Neurons and Neurons-FTH1), and the neural differentiation rates were (91.56 ± 7.89)% and (92.23 ± 7.64)%, respectively. Neuron-specific markers, including nestin, neuron-specific enolase, and microtubule-associated protein-2, were significantly expressed in Neurons-FTH1 and Neurons without noticeable differences. On the other hand, FTH1 was significantly expressed in MSCs-FTH1 and Neurons-FTH1 cells, and the expression levels were not significantly different. The R2 value was significantly increased in MSCs-FTH1 and Neurons-FTH1 cells, which was consistent with the findings of Prussian blue staining, transmission electron microscopy, and intracellular iron measurements. These results suggest that FTH1 gene expression did not affect MSC differentiation into neurons and was not affected by neural differentiation. Thus, MRI reporter gene imaging based on FTH1 can be used for the detection of neurally differentiated cells from MSCs.

1. Introduction

Mesenchymal stem cells (MSCs) exhibit pluripotency and have been extensively applied in preclinical and clinical studies of many types of human diseases in recent years [1–4]. In particular, studies on the application of MSCs in neurological diseases are a hotspot [5–8]. The common neurological diseases are mainly caused by loss or damage of neurons or glial cells. The proliferation and neural differentiation potentials of stem cells can be harnessed to promote the regeneration of nervous tissues to achieve the purpose of organ or tissue repair [9, 10].

During the process of stem cell transplantation therapy, real-time dynamic monitoring of the distribution, migration, proliferation, and differentiation of transplanted cells should be performed. At present, imaging methods for cell tracing mainly include optical imaging [11], nuclear medicine imaging [12], and magnetic resonance imaging (MRI) [13, 14].
2. Materials and Methods

2.1. Acquisition and Identification of MSCs. All animal protocols performed in this study were approved by the Animal Care and Use Committee of Chongqing Medical University, and the procedures were conducted in compliance with the National Institutes of Health Guidelines. All efforts were made to minimize animal suffering.

Four-week-old Sprague-Dawley (SD) rats (40-60 g) were used for isolation and culture of MSCs according to previous methods [21]. Briefly, the long bones of four limbs of rats were isolated. The bone marrow cavity was washed with Dulbecco’s Modified Eagle’s Medium (DMEM; Gibco, Grand Island, NY, USA), and cells were centrifuged and resuspended in the washing solution. Cells were cultured in DMEM/F12 supplemented with 10% fetal bovine serum (FBS; HyClone, Logan, UT, USA), 100 units/ml penicillin, and 100 g/ml streptomycin at 37°C under 5% CO2. Third generation cells with excellent growth status were collected, digested in 0.25% trypsin, centrifuged at 4°C (1000 r/min, 5 min), washed with phosphate-buffered saline (PBS) thrice, and stored for future use.

For identification of stem cells, the mononuclear antibodies CD29, CD90, CD34, and CD45 (Beyotime, Nanjing, Jiangsu, China) were separately added to the cells and incubated in the dark for 30 min. Cells were washed with PBS thrice, resuspended in 200 μl of PBS, detected, and analyzed using a flow cytometer.

2.2. Construction of the Recombinant Lentivirus Carrying FTH1. The cDNA sequence of the FTH1 gene (accession number BC000857) was synthesized (primers: forward, AACCGTCAGATCGCACCACGTAGACGA-CGCCGTCACCCTC; reverse, TCTTTGTAACCATGAA-TTCGCTTTCATTATCACTGTCTC). After polymerase chain reaction (PCR) amplification, the DNA fragment was double digested with EcoRI and AgeI, purified, and ligated to the empty lentiviral vector CMV-MCS-3Flag-Puro containing the CMV promoter to obtain the recombinant lentiviral plasmid CMV-FTH1-3Flag-Puro (pLV-FTH1). After PCR and DNA sequencing, pLV-FTH1, the packaging plasmid pHelper 1.0, and the envelope plasmid pHelper 2.0 were cotransfected into 293T cells (Jikai Gene Technology, Shanghai, China) to obtain the recombinant lentivirus carrying the FTH1 gene, which was referred to as LV-FTH1. The virus titer was determined using enzyme-linked immunosorbent assay (ELISA).

2.3. Reporter Gene Transduction and Cell Identification. To determine the optimal multiplicity of infection (MOI), lentivirus at various MOIs (0, 5, 10, 20, 30, 50) was used to infect the MSCs. After 48 h of infection, the infection efficiency was detected by flow cytometry. The cell viability after 48 h of infection at different MOIs was also evaluated using Cell Counting Kit-8 (CCK-8, Jikai Gene Technology, Shanghai, China) assays. Briefly, 10 μl of CCK-8 reagent was added to each group of cells and incubated for 2 h. The optical density (OD) values (OD450 and OD650) were measured using an optical density scanner. The final OD value was calculated as follows: OD = OD450 - OD650.

When the cell confluence reached 30-40%, the lentivirus LV-FTH1 at the optimal MOI was added. After 24 h, the culture medium was replaced, and cells were continuously cultured. After 48 h, 4 μg/ml of puromycin was added for selection for 7 d. The MSCs carrying FTH1 that contained the drug-resistant gene were obtained and were named MSCs-FTH1.

A total of 1×10⁵ MSCs-FTH1 cells were collected. An equal number of cells transduced with the empty virus (MSCs-LV) and MSCs not transduced with virus were used as controls. FTH1 gene expression was detected using Western blot. The total cellular protein in the 3 groups was extracted using a cellular total protein reagent kit (Beyotime, Nanjing, Jiangsu, China). The protein concentration was measured using the bichinchoninic acid (BCA; Beyotime, Nanjing, Jiangsu, China) method. After SDS-polyacrylamide gel electrophoresis (SDS-PAGE; Beyotime, Nanjing, Jiangsu, China), proteins were transferred onto a polyvinylidene fluoride (PVDF) membrane (Jikai Gene Technology, Shanghai, China). The membrane was blocked in 5% bovine serum albumin (BSA,
Sigma-Aldrich, St. Louis, MO, USA) for 1h and incubated with the primary rabbit anti-FTH1 monoclonal antibody (1:1000, Abcam, Cambridge, MA, UK) or mouse anti-β-actin monoclonal antibody (1:1000, Abcam, Cambridge, MA, UK) at 4°C overnight. The membrane was then incubated with the secondary goat anti-rabbit (1:5,000) or goat anti-mouse (1:1000) antibody (Genscript, Nanjing, Jiangsu, China) for 2h. Finally, the results were developed using the Enhanced Chemiluminescence (ECL) Reagent Kit (Beyotime, Nanjing, Jiangsu, China). To further confirm the expression of FTH1, the tag protein Flag in 3 types of cells was also detected via Western blotting with the primary mouse anti-Flag antibody (1:2000) and the secondary goat anti-mouse antibody (1:2,000) (Jikai Gene Technology, Shanghai, China).

In addition, the expression of the Flag protein was detected using immunofluorescence. Cells were fixed in 4% paraformaldehyde for 20 min, permeabilized in 1% Triton X-100 solution at room temperature for 10 min, and blocked in 5% BSA at room temperature for 1h. Then, the cells were incubated with the primary antibody (mouse anti-Flag 1:200) and the secondary antibody (anti-mouse Cy-3 conjugated, 1:1000), counterstained with 4,6-diamidino-2-phenylindole (DAPI; Beyotime, Nanjing, Jiangsu, China), and observed under a fluorescence microscope (TE2000-S, Nikon, Tokyo, Japan).

2.4. In Vitro Differentiation of MSCs into Neurons and Identification. MSCs-FTH1 cells with excellent growth were inoculated into a 24-well plate. When the cell confluence reached 70-80%, 1 mmol/L all-trans retinoic acid (ATRA; Beyotime, Nanjing, Jiangsu, China) was added for preinduction for 24h. The original culture medium was discarded. Cells were washed with PBS thrice, and modified neuronal medium (MNMs, DMEM/F12/1.6% DMSO/160 μM butyraldehyde/hydroxyanisole/20 mM KCl/1.6 mM valproic acid/8 μM forskolin/0.8 M hydrocortisone/4 g/ml insulin) was added for induction for 24 h [32]. The obtained cells were neuron-like cells and were named Neurons-FTH1. MSCs were induced into neuron-like cells using the above method and were named Neurons and used as the control.

The expression of neuron-specific markers nestin, neuron-specific enolase (NSE), and microtubule-associated protein (MAP-2) in Neurons-FTH1 and Neurons was detected using immunofluorescence. Cells were fixed in 4% paraformaldehyde for 20 min, permeabilized in 1% Triton X-100 at room temperature for 10 min, and blocked in 5% BSA at room temperature for 1h. Cells were incubated with the primary mouse anti-NSE monoclonal antibody, rabbit anti-nestin monoclonal antibody, or rabbit anti-MAP2 monoclonal antibody (1:200, Proteintech, IL, USA) at 4°C overnight. Next, cells were incubated with the appropriate secondary goat anti-mouse or goat anti-rabbit DyLight 488- or DyLight 594-conjugated antibody (1:1000, Proteintech, IL, USA), counterstained with DAPI, and observed under a fluorescence microscope (TE2000-S, Nikon, Tokyo, Japan).

In the NSE immunofluorescence image, 10 fields were randomly selected to count the NSE positive and axon forming cells. Its ratio with the total number of cells was the differentiation rate. In addition, the length of axons of differentiated cells was measured using ImageJ software (National Institutes of Health, Bethesda, Maryland, USA). The measurement was independently thrice, and each measurement was repeated thrice.

The expression of nestin, NSE, and MAP-2 before and after cell differentiation was quantitatively measured using quantitative real-time PCR (qRT-PCR). Total cellular RNA was extracted using TRIzol (Invitrogen, Carlsbad, CA, USA). RNA was reversely transcribed into cDNA using the PrimeScript RT Reagent Kit (TaKaRa, Tokyo, Japan). Using each synthesized cDNA as a template, PCR was performed with the following primers: Nestin: 5'-AGCTGCGGACACCTC-AAGATG-3' and 5'-AGGGAGTTGGGCTCAG GAC-3', NSE: 5'-TCGCCACATTGCTCAACT-3' and 5'-AACTCA-GAGGCAGCCACATC-3', and MAP2: 5'-AATCAGCTC-TGGCCTCCAGT-3' and 5'-AGTGGGTGTGATGACCAC-AC-3'. A total of 30 cycles was performed.

2.5. Western Blot Detection of FTH1 Expression before and after Cell Differentiation. Western blotting was performed to detect FTH1 expression in MSCs and MSCs-FTH1 before induction and in Neurons and Neurons-FTH1 after 48 h of induction. The total cellular protein was extracted, and the protein concentration was determined using the BCA method. After SDS-PAGE, proteins were transferred onto a PVDF membrane. The membrane was blocked in 5% BSA for 1h, incubated with the primary anti-β-actin antibody or the primary anti-FTH1 antibody at 4°C overnight, and the secondary antibody for 2h. Finally, the results were developed using ECL. Each group of experiments was repeated thrice.

2.6. Detection of Cell Viability before and after Neural Differentiation. MSCs and MSCs-FTH1 before differentiation and Neurons and Neurons-FTH1 after differentiation were cultured for 72 h in the presence or absence of 500 μM ferric ammonium citrate (FAC; Beyotime, Nanjing, Jiangsu, China). The effect of FTH1 expression and/or iron ions on cell viability was detected using a CCK-8 assay. Each group had 5 replicated wells. Briefly, 10 μl of CCK-8 reagent was added to each well. The plates were gently mixed and incubated for 2h. The OD values (OD_{450} and OD_{550}) of each well were measured using an optical density scanner. The final optical density value of each well was calculated as follows: OD=OD_{450}−OD_{550}. Each well was plated in triplicate.

2.7. In vitro MR Imaging of Cells. The four groups of cells (MSCs, MSCs-FTH1, Neurons, and Neurons-FTH1) were cultured in the presence of 500 μM FAC for 72h. A total of 1×10^6 cells were collected, washed with PBS thrice, resuspended, and transferred into a 0.5-ml PCR tube. The tube was left standing for 30 min to form the cell pellet at the bottom of the tube. Cells were scanned in an Achieva 3.0 T MRI scanner (Philips Medical Systems, Eindhoven, Netherlands) using the knee joint coil. The scanning sequences and parameters were as follows: T2WI: TR=2000 ms, TE=65 ms; GRE sequence: TR=2000 ms, TE: 13, 26, 39, 52, 65, and 78 ms, FOV=120×120 mm, matrix=512×512, and slice thickness=1 mm. The R2 artificial color map (R2 map) was obtained using the R2 star software.
in the postprocessing workstation. Five different regions of interest (ROIs, 3.0 mm²) were selected for measuring R₂
values.

2.8. Detection of Intracellular Iron

Prussian Blue Staining. Cells were fixed in 4% paraformaldehyde for 20 min and stained with the Prussian blue staining
reagent (mixture of 2% potassium ferrocyanide and 2% HCl
at an equal volume) at room temperature for 30 min and
nuclear fast red staining solution for 2-3 min. After washing
with ddH₂O, cells were mounted in neutral balsam and
observed under a light microscope.

Transmission Electron Microscopy (TEM). Cells were fixed
in 2.5% glutaraldehyde solution and 1% osmic acid for
30 min and stained in 0.5% uranyl acetate overnight. Cells
were dehydrated in a series of ethanol gradations, immersed
and embedded into the mixture of the equal volume of
100% acetone and the EP812 embedding solution, ultra-thin
sectioned at 100nm, and observed under a Hitachi-7500
transmission electron microscope (Hitachi, Tokyo, Japan).

Detection of Intracellular Iron Content. A total of 1×10⁵
cells from each group were centrifuged, and the pellet was dried at
110°C overnight. Cells were digested in 500 µl of a perchloric
acid-nitric acid mixture (1:3) at 60°C for more than 3 h to
completely dissolve the cells. The iron ion concentration was
measured using an atomic absorption spectrophotometer.
The results were expressed as pg/cell. Each sample was
measured 3 times.

2.9. Statistical Methods. All data were expressed as X±SD.
The comparison of the difference in iron content between the
groups was performed using Student’s t test. The statistical significance was determined by p-values. A p-value of <0.05
was considered statistically significant.

3. Results

3.1. Identification of MSCs. The positive rates of the surface
markers of the 3rd generation MSCs detected by flow cytometry were 96.7% and 99.2% for CD29 and CD90, respectively;
however, the CD34 and CD45 positive rates were only 0.2% and 0.1%, respectively (Figure 1).

3.2. Transduction of MSCs with the FTH1 Gene and Identification. The optimal MOI was determined by the infection
efficiency and cell viability. When the MOI increased, the infection efficiency increased. When the MOI reached 10,
the infection efficiency reached 78.5% (Figure 2(a)). CCK-8
assay results showed that the cell viability did not decrease
significantly in the groups treated at a MOI ≤ 10 but decreased
significantly in the groups with a MOI ≥ 20 compared with the
control (P<0.05) (Figure 2(b)). Considering cell safety, the optimal MOI was determined to be 10, although the infection
efficiency continued to increase when the MOI exceeded 10.
Using this protocol, MSCs carrying the reporter gene FTH1
were successfully obtained.

Western blotting results revealed that MSCs transduced with lentiviruses carrying the FTH1 gene (MSCs-TFH1) exhibited a positive band at 21 KDa, which was consistent with
the theoretical size of the FTH1 protein. The positive
band was not observed in the MSCs and MSCs-LV in the control
groups (Figure 3(a)). Western blotting of the tag protein
Flag also showed a positive band near FTH1 (Figure 3(b)),
which was of the expected molecular weight of the recombinant FTH1 (21 KDa) and Flag (1 KDa). Immunofluorescence revealed that the Flag protein was expressed in
MSCs-TFH1 and MSCs-LV but not in MSCs (Figure 3(c)). The above results confirmed that MSCs were
successfully transduced with FTH1.

3.3. Morphological Observation and Quantitative Analyses of MSCs before and after Neural Differentiation. Before differenti-
ation induction, MSCs and MSCs-FTH1 exhibited a flat
or spindle shape and did not have retraction. After ATRA
preinduction and MNM induction for 24 h, cell morphology
exhibited significant changes. Most cells had enhanced transparency. In addition, the cytoplasm shrank to the nuclear
center, and cells had long and thin processes that exhibited bipolar or multipolar expansion to the surroundings, formed
secondary or even multiple levels of processes, and connected to adjacent cells (Figure 4(a)). The neural differentiation
rates of MSCs and MSCs-TFH1 were (95.6±8.7)% and
(92.23±7.64)%, respectively, and these cell types did not
exhibit significant differences (P<0.05) (Figure 4(b)).

The mean axonal length of Neurons was (97.13±10.89), which was
not significantly different from Neurons-FTH1 (102.24±12.61)
(P>0.05) (Figure 4(c)).

3.4. Expression of Neuron-Specific Markers before and after Neural Differentiation. Immunofluorescence results showed
that undifferentiated MSCs and MSCs-FTH1 did not express
euron-specific markers. After cells were induced to differ-
tiate into Neurons and Neurons-FTH1, strong positive
expression of NSE, Nestin, and MAP-2 was detected (Fig-
ure 4(a)).

qRT-PCR quantitative detection of neuron-specific markers
before and after cell differentiation revealed that nestin,
NSE, and MAP-2 were not expressed in MSCs and MSCs-
FTH1 before differentiation. After differentiation induction,
nestin, NSE, and MAP-2 were obviously expressed in Neu-
rons and Neurons-FTH1, and the difference between these two
groups was not significantly different (P>0.05) (Figures 5(b)
and 5(c)).

3.5. FTH1 Expression in MSCs before and after Differentiation.
Western blot results revealed that MSCs and Neurons
that did not contain the FTH1 gene did not exhibit a target band,
Figure 1: Detection of surface markers of MSCs using flow cytometry. The MSCs specific markers, CD29 and CD90, had strong positive expression, while CD34 and CD45 did not have obvious positive expression.

whereas both MSCs-FTH1 and Neuron-FTH1 exhibited an obvious target band (Figure 6(a)). The gray density analysis results revealed that the gray density ratio between MSCs-FTH1 and Neurons-FTH1 did not exhibit a significant difference (P > 0.05) (Figure 6(b)). These results indicate that FTH1 gene expression was not significantly altered after MSCs were induced to differentiate into neuron-like cells.

3.6. The Effect of FTH1 Expression on Cell Viability. CCK-8 detection results showed that the OD values (OD\textsubscript{450} - OD\textsubscript{650}) of the 4 groups of cells (MSCs, MSCs-FTH1, MSCs/FAC, and MSCs-FTH1/FAC) before differentiation were 1.07±0.11, 1.02±0.09, 1.05±0.08, and 1.04±0.10, respectively; P > 0.05 (Figure 7(a)). The OD values of the 4 groups of cells (Neurons, Neurons-FTH1, Neurons/FAC, and Neurons-FTH1/FAC) after differentiation were 0.98±0.08, 1.06±0.10, 1.02±0.09, and 1.05±0.07, respectively; P > 0.05 (Figure 7(b)). These results suggested that FTH1 gene expression and 500 μM FAC both did not have significant effects on cell viability before and after differentiation.

3.7. In vitro MRI Findings. MRI scanning was performed on MSCs-FTH1, Neurons-FTH1, MSCs, and Neurons after 72 h of culture in the presence of 500 μM FAC. The results showed that the precipitation of T\textsubscript{2} WI signals in MSCs-FTH1 and Neurons-FTH1 significantly decreased compared with MSCs and Neurons (Figure 8(a)). The R2 map quantitative measurement showed that the R2 values of MSCs-FTH1 and Neurons-FTH1 significantly increased compared with MSCs and Neurons-FTH1, and the difference between MSCs-FTH1 and Neurons-FTH1 was not significant (Figures 8(b) and 8(c)). These results confirmed that the iron transport function of FTH1 was clear and was not affected by neural differentiation.

3.8. The Iron Accumulation Effect of FTH1. The Prussian blue staining results showed that after 4 groups of cells were
cultured in the presence of FAC for 3 d, the cytoplasm of MSCs-FTH1 and Neurons-FTH1 exhibited more blue particles, whereas MSCs and Neurons in the control group did not exhibit obvious blue particles (Figure 9(a)). The transmission electron microscopy results were consistent with the Prussian blue staining results. The cytoplasm of MSCs-FTH1 and Neurons-FTH1 contained black electron-dense particles, whereas the cytoplasm of MSCs and Neurons in the control group did not contain electron-dense particles (Figure 9(b)). The atomic absorption spectrophotometer detection results revealed that the iron levels in MSCs-FTH1 and Neurons-FTH1 were 0.87±0.08 pg/cell and 0.91±0.13 pg/cell, respectively, and there was no significant difference (P>0.05). However, the levels were significantly increased compared with those in MSCs (0.13±0.01 pg/cell) and Neurons (0.91±0.13 pg/cell) (P<0.05) (Figure 9(c)).
Figure 3: FTH1 and Flag-tag expression in MSCs. (a) Detection of the FTH1 gene in MSCs via Western blot. MSCs-FTH1 exhibited a positive protein band at 21 KDa, which was consistent with the theoretical size of the FTH1 protein. The positive band was not observed in MSCs and MSCs-LV in the control group. (b) Detection of the Flag-tag in MSCs via Western blot. A positive band near FTH1 was observed, which was of the expected molecular weight of the recombinant FTH1 (21 KDa) and Flag (1 KDa) proteins. (c) Detection of the Flag protein in MSCs using immunofluorescence. Red fluorescence was observed in MSCs-FTH1 and MSCs-LV but not in MSCs. These results confirmed that transduction with TFH1 was successful.
Figure 4: Differentiation of MSCs into neuron-like cells. (a) Before induction, MSCs and MSCs-FTH1 exhibited a flat or spindle shape and no refractive index. After induction, cells had enhanced transparency and thin and long processes that exhibited bipolar or multiple-polar expansion and connected to adjacent cells. (b) The neural differentiation rates between MSCs and MSCs-FTH1 did not exhibit significant differences ($P > 0.05$). (c) The mean axonal lengths between Neurons and Neurons-FTH1 did not exhibit significant differences ($P > 0.05$).

4. Discussion

The application of the MRI reporter gene imaging strategy depends on whether the reporter gene can be persistently and stably expressed in cells [30, 31]. This study successfully induced the differentiation of MSCs carrying the FTH1 reporter gene into neuron-like cells. The results demonstrated that the reporter gene was clearly expressed before and after differentiation and induced the iron accumulation effect, which could be displayed by a clinical 3.0T MRI scanner. These results provide important foundations for further use of the MRI reporter gene imaging for long-term tracing of the biological behaviors of MSCs after transplantation.

Safety is a prerequisite for the application of the MRI tracing strategy for stem cells [33, 34]. Previous studies demonstrated that although the use of SPIO labeling did not affect the proliferation activity of stem cells, it might reduce the osteogenic differentiation ability [35]. For the FTH1 reporter gene used as an MRI tracer, whether its expression affects stem cell differentiation and the effect of stem cell differentiation on the expression of the reporter gene should be confirmed. This study used a lentiviral vector to
transduce the reporter gene FTH1 into MSCs. Under certain induction conditions, MSCs were successfully differentiated into neuron-like cells. Regardless of the morphological observation or quantitation of the differentiation efficiency and axonal lengths, MSCs with or without FTH1 transduction did not exhibit significant differences. The quantitative detection of several neuron-specific markers also did not reveal differences between these two groups. These results indicated that transduction with FTH1 did not have significant effects on the neural differentiation potential of MSCs. On the other hand, although differentiated neuron-like cells exhibit changes in morphology and various specific markers, FTH1...
Figure 6: FTH1 expression in cells before and after differentiation. (a) Western blot results revealed that MSCs and Neurons did not exhibit a target band, whereas MSCs-FTH1 and Neurons-FTH1 both exhibited a significant target band. (b) The gray density analysis results showed that the gray density ratio between MSCs-FTH1 and Neurons-FTH1 did not exhibit significant differences (P>0.05).

Figure 7: Detection of cell viability using CCK-8. (a) The OD values (OD_{450}-OD_{650}) among 4 groups of cells (MSCs, MSCs-FTH1, MSCs/FAC, and MSCs-FTH1/FAC) before differentiation did not have significant difference, P>0.05. (b) The OD values among 4 groups of cells (Neurons, Neurons-FTH1, Neurons/FAC, and Neurons-FTH1/FAC) after differentiation did not exhibit significant differences, P>0.05.

expression in cells was normal, and the expression levels did not exhibit significant differences compared with those before differentiation. These results suggest that the neural differentiation of stem cells also did not have significant effects on the expression of the reporter gene.

The greatest advantage of FTH1 reporter gene imaging involves the use of a lentiviral vector to transduce cells with the gene and to integrate the gene into the cell genome. Through the expression of the reporter gene, cells continuously accumulate iron to achieve the purpose of the long-term continuous tracing of cells. This overcomes the deficiency of the direct SPIO labeling method given that the iron level gradually decreases in cells as a result of division and proliferation. Previous studies demonstrated that MRI detection of SPIO-labeled stem cells in vitro and in vivo is limited to within 4 w and 2 w, respectively [19, 21]. Because iron in cells is not stable, and cells can be engulfed by macrophages, iron-induced MRI signal changes cannot truly reflect the number of cells [36, 37]. Although the iron transporting efficiency of FTH1 is relatively low, and the MRI signal changes are weak, the signals are stable and sustained. Therefore, theoretically, the strength of MRI signals can indirectly determine the number of transplanted cells [38]. The results of this study revealed that significant signal changes in 10^6 FTH1-transduced cells could be discovered by 3T MRI, and the iron levels and MRI signal changes in cells before and after differentiation did not exhibit significant changes. These results indicated that the iron transporting


The effect of FTH1 was clear and stable. The sensitivity of MRI in cell imaging is not only associated with cell labeling methods and cell number but is also associated with the magnetic field intensity and imaging sequences [29, 30]. When the magnetic field intensity is increased, the MRI sensitivity is increased. The clinical 3T MRI used in this study could detect signal changes in $1 \times 10^6$ cells. We speculate that MRI with increased field strength can detect fewer cells with more sensitivity. It is generally considered that the $T_2^*$ sequence is more sensitive in the detection of the ferromagnetic material than $T_2$. However, the former has the amplification effect and is easy to produce artifacts. In addition, its signal range and strength typically cannot truly reflect the number and distribution of cells [39]. Thus, we adopted the $T_2$WI sequence in this study.

Currently, the effect of FTH1 gene expression and iron accumulation on cell viability remains controversial [40–43]. Some studies revealed that excessive expression of the FTH1 gene in cells results in the uptake of excessive unstable iron, which might induce the Fenton effect to cause inorganic oxidation of intracellular carboxylic acids, alcohols, and esters to damage cells [40, 41]. Other studies considered that the FTH1 gene catalyzes the reaction between stable divalent iron ions and intracellular hydroxyl radicals to produce ferric ions to attenuate the cytotoxic function of the Fenton effect induced by excessive uptake of exogenous iron ions [42, 43]. The inconsistency of these results might be associated with the experimental cell types and the concentration and time of supplemented iron. This study detected the cell viability before and after differentiation to determine the effect of two factors, namely, FTH1 gene expression and/or addition of iron ions, on cell viability. The results demonstrated that cell viability between MSCs and MSCs-FTH1 before neural differentiation and between Neurons and Neurons-FTH1 after differentiation both did not exhibit significant differences. In addition, regardless of before or after differentiation, the cell viability between these two groups of cells with or without the addition of 500 μM FAC also did not exhibit significant differences, indicating that these two factors, namely, FTH1 gene expression and/or the addition of iron ions, did not have significant effects on cell viability.

This study confirmed the feasibility of using FTH1 in the imaging of neuronal differentiation of stem cells. However, the study has the following limitations. First, this experiment is limited to in vitro studies. To validate the iron accumulation function of FTH1, a certain concentration of FAC was added to the culture medium. The actual iron accumulation function of FTH1 in the body mainly depends on the iron uptake from surrounding tissues. Compared with the simple and controllable in vitro environment, the in vivo environment is more complex. Under this condition, some issues still await further in vivo experiments for confirmation, including whether the endogenous iron level can meet the requirement of MRI imaging, whether gene expression and iron accumulation in cells affect cell differentiation, and whether the reporter gene in cells can be continuously expressed after differentiation. Next, during the actual application in transplanted cell tracing, both stem cells and differentiated cells exhibited reporter gene expression and reduced MRI signals. Hence, simply relying on the MRI reporter gene imaging cannot distinguish cells before and after differentiation. Therefore, how to use MRI to determine whether stem cells have already been successfully differentiated into specific cells in vivo, i.e., using MRI to monitor the occurrence of the differentiation event of stem cells, has become a difficult problem in this research field and is a future direction of our work.

5. Conclusions

This study successfully differentiated MSCs carrying the FTH1 gene into neuron-like cells. FTH1 gene expression neither affected the differentiation of MSCs into neurons nor was affected by neural differentiation. The iron accumulation effect was sufficient to induce changes in $T_2$ MRI signals. These results suggested that MRI reporter gene imaging based on FTH1 could be used for the detection of neurally differentiated cells from MSCs.

Data Availability

The data are available from the corresponding author upon request.
Figure 9: The iron transport effect of FTH1. (a) Prussian blue staining: there was an increased number of blue particles in the cytoplasm of both MSCs-FTH1 and Neurons-FTH1, whereas no obvious blue particles were observed in MSCs and Neurons in the control group. (b) Transmission electron microscopy: the cytoplasm of MSCs-FTH1 and Neurons-FTH1 contained black electron-dense particles (arrows), whereas the cytoplasm of MSCs and Neurons in the control group did not contain electron-dense particles. (c) Atomic absorption spectrophotometer detection revealed that the iron levels between MSCs-FTH1 and Neurons-FTH1 were not significantly different (P > 0.05), but the levels were increased compared with MSCs and Neurons (P<0.05).

Conflicts of Interest

The authors declare that they have no conflicts of interest to disclose.

Authors’ Contributions

Tong Mu and Yong Qin contributed equally to this work. All authors were involved in the study and approved its final version.

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