Review

Non-Steroidal Anti-Inflammatory Drugs in the Carcinogenesis of the Gastrointestinal Tract

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Abstract: It is estimated that underlying infections and inflammatory responses are linked to 15–20% of all deaths from cancer worldwide. Inflammation is a physiologic process in response to tissue damage resulting from microbial pathogen infection, chemical irritation, and/or wounding. Tissues injured throughout the recruitment of inflammatory cells such as macrophages and neutrophils, generate a great amount of growth factors, cytokines, and reactive oxygen and nitrogen species that may cause DNA damage that in turn predisposes to the transformation from chronic inflammation to neoplasia. Cyclooxygenase (COX), playing a key role in cell homeostasis, angiogenesis and tumourigenesis, may represent the link between inflammation and cancer. Currently COX is becoming a pharmacological target for cancer prevention and treatment.

Keywords: cyclooxygenase-2; non-steroidal anti-inflammatory drugs; esophageal cancer; gastric cancer; colorectal cancer

1. Introduction

It was in 1863 that Rudolf Virchow indicated the link between cancer and inflammation on the basis of observations that tumours often arose at sites of chronic inflammation and that inflammatory cells were present in samples from tumours [1].
With time, many epidemiologic evidences have supported Virchow’s hypothesis showing that chronic inflammatory diseases are frequently associated with increased risk of cancers [1–3]. Currently it is estimated that underlying infections and inflammatory responses are linked to 15–20% of all deaths from cancer worldwide [4]. For instance, the risk of colorectal cancer (CRC) was 10-fold greater if linked with an inflammatory bowel disease, such as ulcerative colitis and Crohn’s disease [5,6]. The risk of esophageal, gastric and pancreatic cancer may be increased by inflammatory diseases, such as esophagitis with Barrett’s metaplasia, chronic atrophic gastritis with intestinal metaplasia and chronic pancreatitis respectively [7,8]. Furthermore, the cancer risk appears to be positively associated with the severity and duration of inflammatory diseases [9].

But the question now is how does chronic inflammation develop into tumours and which are the driving mediators in this process. Inflammation is a physiologic process in response to tissue damage resulting from microbial pathogen infection, chemical irritation, and/or wounding [2]. Tissues injured throughout the recruitment of inflammatory cells such as macrophages and neutrophils, generate a great amount of growth factors, cytokines, and reactive oxygen and nitrogen species that may cause DNA damage [3,10]. If the inflammatory process is activated persistently it may lead to continuous tissue damage and an altered microenvironment that in turn sustains cell proliferation and predisposes to the transformation from chronic inflammation to neoplasia [1].

The chronic inflammation microenvironment is predominated by macrophages [10]. Macrophages generate a great amount of inflammatory mediators which react with DNA and cause mutations in proliferating epithelial and stroma cells [11,12]. Macrophages and T lymphocytes may release tumour necrosis factor-α (TNF-α) and macrophage migration inhibitory factor to exacerbate DNA damage [13]. Macrophages, neutrophils, eosinophils, dendritic cells, mast cells, and lymphocytes are also found to be key components in the epithelial-originated tumours [14]. Indeed, inflammatory cells act as tumour promoters in inflammation associated cancers. Tumour-associated macrophages (TAM) are a major component of the infiltrate of most, if not all, tumours [15]. TAM derive from circulating monocytic precursors, and are directed into the tumour by chemoattractant cytokines called chemokines. Many tumour cells also produce cytokines called colony-stimulating factors that prolong survival of TAM. Furthermore, TAM also produce growth and angiogenic factors as well as protease enzymes which degrade the extracellular matrix. Hence, TAM can stimulate tumour-cell proliferation, promote angiogenesis, and favor invasion and metastasis [16,17]. Cytokines, including interleukins (IL), TNF-α, growth factors and colony-stimulating factors, are secreted or membrane-bound molecules that play a regulatory role in the growth, differentiation, and activation of immune cells [18]. Cytokine signaling could contribute to the progression of tumours in two aspects: the stimulation of cell growth and differentiation and the inhibition of apoptosis of altered cells at the inflammatory site [18]. Chemokines include the largest family of cytokines. Most tumours produce chemokines of the two major groups, α (or CCX) and β (or CC) [19,20]. In the inflammation process, chemokines, usually induced by cytokines, are major soluble regulators that control the directional migration of leukocytes to the inflammatory site. It is well established that chemokines are involved in the promotion of cancer. Moreover, chemokines also facilitate tumour invasion and metastasis in various cancer types and the balance between chemokines with proangiogenic and angiostatic activities is critical in regulating angiogenesis. Mechanistically, chemokines may contribute to tumour invasion and
metastasis by mediating the directional migration of tumour cells to specific distal organs via circulation in a similar manner to its control of leukocyte migration [21–24].

Therefore, the relationship between cancer and inflammation is not simple but rather it is a complicated pathologic processes under the control of many driving forces. To address the details of development of inflammation-associated cancers, and further the transition from inflammation to cancer, it is necessary to investigate the specific roles of key regulatory molecules involved in this process. Here we centre the attention on cyclooxygenase (COX), which plays a key role in cell homeostasis, angiogenesis and tumourigenesis. The function of COX in linking inflammation to cancer is now becoming the target of intense investigation and starting to have implications for cancer prevention and treatment.

2. Cyclooxygenase-2

Cyclooxygenase is a rate-limiting enzyme in the synthesis of prostaglandins (PGs). It catalyses the conversion of arachidonic acid to PGG2, then to PGH2 which is subsequently converted to various physiologically active prostanoids, including PGE2, PGD2, PGF2a, PGI2 (prostacyclin) and thromboxane A2 (TXA2) by the relevant enzymes in a variety of cell types [25,26]. The COX enzyme exists in three isoforms, commonly referred to as COX-1, COX-2, and COX-3 [27,28]. COX-1 is expressed constitutively in many tissues and mediates the "housekeeping" functions such as cytoprotection of gastric mucosa, regulation of renal blood flow and platelet aggregation. In contrast, COX-2 is not detected in most normal tissues, but its expression may be induced mainly at sites of inflammation in response to inflammatory stimuli including pro-inflammatory cytokines such as IL-1α/β, interferon-γ (IFN-γ), TNF-α and growth factors [27]. COX-3, a novel COX-1 splice variant (now called COX-1b) has been identified in canine tissue (most abundant in cerebral cortex) as an acetaminophen-sensitive isoform. However, the implication of this splice variant in humans is still not known [28].

The stimulation of COX-2 expression in Src-transformed fibroblasts, endothelial cells and monocytes treated with the tumour promoter tetradecanoylphorbol acetate or lipopolysaccharide led to the notion that COX-2 is an inducible enzyme that produces prostaglandins during inflammatory and tumourigenic settings [29]. A review article on this issue showed that COX-2 is up-regulated from 51% up to 100% of the tumours by Northern blot, reverse transcription polymerase chain reaction, immunoblotting and immunohistochemistry [30].

The up-regulation of COX-2 results in an increased synthesis of PGs. Prostaglandins exert their effects locally in both autocrine and paracrine patterns. In particular PGE-related to COX-2 up-regulation appeared strongly involved in the carcinogenic process. PGE2 effects are mediated by a family of G-protein-coupled receptors, namely, EP1, EP2, EP3, and EP4 [31]. In some cell types, nuclear peroxisome proliferator-activated receptors (PPAR) are also involved in mediating the PG effects [32]. In a recent study in CRC cells, PGE2 promoted cell growth and motility via the EP4 receptor by activation of the phosphatidylinositol 3-kinase (PI3K)/protein kinase B (Akt/PKB) pathway. Interestingly, EP2 and EP3 were also expressed in the CRC cells and their binding affinities to PGE2 are similar to EP4 [33].
The only other COX-2 derived PG implicated in oncogenesis is TxA2, which was reported to promote tumour growth and angiogenesis [34]. Moreover, it has been reported that TxA2 synthase inhibitor blocks colorectal carcinoma liver metastasis in an in vitro study [35]. COX-2 is involved in the carcinogenesis process throughout cellular proliferation, antiapoptotic activity, angiogenesis, and immune response.

2.1. Proliferation and apoptosis

Prostaglandins stimulate proliferation of different cell lines derived from gastrointestinal tract such as colonic, intestinal, gastric and esophageal cell lines. COX-2 derived PGE2 promotes human cancer cell growth by autoregulation of COX-2 expression, which depends primarily on PGE2 induced activation of the Ras-MAPK pathway [36].

Overall data from literature show that COX-2 inhibits apoptosis through three different pathways: the Bcl-2 mediated pathway, the nitric oxide pathway, and that of ceramide [37]. The role of COX-2 in preventing apoptosis is likely mediated by COX-2 derived PGE2, which attenuates cell death induced by the COX-2 selective inhibitor SC-58125 [38]. PGE2 induces antiapoptotic protein expression such as Bcl-2 and increases nuclear factor kappa B (NF-κB) transcriptional activity, which is a key antiapoptotic mediator [39].

COX-2-derived PGs regulate programmed cell death and reduce the apoptotic rate via inhibition of the mitochondrial apoptotic pathway characterized by reduced cytochrome C release, attenuated activation of caspase-9 and -3 and up-regulation of bcl-2 [36]. Additionally, increased prostanoid generation due to COX-2 overexpression specifically inhibits Fas-mediated apoptosis [40]. These findings have stimulated great interest in identifying COX-2 as a target for modulating apoptosis.

In vivo, both non-selective and selective COX-2 inhibitors stimulate apoptosis in APC-deficient cells that have not yet undergone malignant transformation. This is also seen clinically in familial adenomatous polyposis (FAP) patients treated with sulindac and in experimental studies of ApcMin mice and rats exposed to chemical carcinogens [41–45]. Non-selective COX-2 inhibitors lose their ability to inhibit chemically induced tumours when polyps undergo malignant transformation. In contrast, selective COX-2 inhibitors stimulate apoptosis and suppress growth in many carcinomas, including cultured human cancers of the stomach [46], esophagus [47], colon [48], and pancreas [49].

2.2. Angiogenesis

The formation of new blood vessels by angiogenesis to provide adequate blood supply is a key requirement for the growth of many tumours. While normal blood vessels express the COX-1 enzyme, new angiogenic endothelial cells express the inducible COX-2. Overexpression of COX-2 in CRC cells induces the production of angiogenic factors such as vascular endothelial growth factor (VEGF), basic fibroblast growth factor (bFGF), IL-8, TNFα, platelet derived growth factor (PDGF) and PGs. Several reports demonstrated that PGE2 up-regulates VEGF in cultured human fibroblasts and increases VEGF and bFGF expression through stimulation of ERK2/JNK1 signaling pathways in endothelial cells [50–52]. Interestingly, VEGF and bFGF induce COX-2 and subsequent PGs production in endothelial cells, suggesting that the effects of PGE2 on regulation of VEGF and bFGF are likely amplified through a positive feedback loop [53]. More recently, PGE2 has been shown to regulate angiogenesis
through the modulation of a chemokine receptor signalling by modulating bFGF-induced chemokine receptor-4, that is important for microvessel assembly in vivo, and inducing the pro-angiogenic chemokine CXCL-1 in vivo [54]. PGE2 may also stimulate the transcription of the hypoxia inducible factor-1 (HIF-1) and work concomitantly with the hypoxic tumour microenvironment to orchestrate the process of angiogenesis [55].

The contribution of COX-2 at multiple points in the angiogenetic cascade makes it an ideal target in the pharmacological inhibition. Both non-selective and selective COX-2 inhibitors inhibit angiogenesis through a combined inhibition of angiogenic growth factors production, response to angiogenic factor and impairment of endothelial cell survival and migration. Inhibition of COX-2 activity in endothelial cells by COX-2 inhibitors resulted in a diminished integrine αvβ3-dependent activation Cdc42 and Rac, two members of the Rho family of GTPases that regulate cytoskeletal organization and cell migration, resulting in FGF-2-induced angiogenesis in vivo [56].

2.3. Immune response

The tumour microenvironment is predominantly shifted from a Th1 to a Th2 dominant immune response [57]. PGE2 has been shown to down-regulate Th1 cytokines (TNF-α, IFN-γ and IL-2) and up-regulate Th2 cytokines such as IL-4, IL-6 and IL-10 [58,59]. Moreover, PGE2 can modulate immune function through inhibiting dendritic cell differentiation and T cell proliferation and suppressing the antitumour activity of natural killer cells and macrophages [58]. In addition, PGE2 up-regulates the complement regulatory protein decay accelerating factor which results in blocking the complement C3 into two active compounds, C3a and C3b in CRC cells [60]. This ability of PGE2 to suppress these immune responses may allow tumour cells to escape immunosurveillance, adding to the already countless roles of the COX-2/PGE2 pathway during tumour development. COX-2 selective inhibitors restore the tumour induced imbalance between Th1 and Th2 and promote antineoplastic responses in lung cancer and metastatic spread of CRC [61,62]. These findings led to extensive efforts to understand how PGE2 can regulate immunosuppression.

2.4. COX inhibitors and cancer chemoprevention

Non-steroidal anti-inflammatory drugs (NSAIDs) block both COX-1 and COX-2 isoenzymes. In addition to the beneficial effects on the treatment of pain and inflammation, the use of NSAIDs is linked to other beneficial effects in the prevention and treatment of gastrointestinal tract tumours. The National Cancer Institute has tested many NSAIDs, such as ibuprofen, indomethacin, ketoprofen, piroxicam, and sulindac for chemopreventive activity [63]. However, the prolonged use of these compounds is limited by gastric (bleeding and ulcers) and kidney toxicity [64–67]. In addition, the use of steroidal antiinflammatory or glucocorticoids (e.g. dexamethasone) which can also inhibit COX-2 [68,69], is limited in the chemoprevention setting because of long-term adverse effects (adrenal cortical suppression). Recently, selective COX-2 inhibitors (valdecoxib, rofecoxib, celecoxib, and others yet in development) has been developed to minimize gastrointestinal toxicity because of the relative paucity of COX-2 expression in the gastrointestinal tract and the relative abundance of COX-2 expression in inflamed and painful tissues. However, selective inhibition of COX-2 might increase the
risk for thrombotic cardiovascular events, due to a relative reduction in endothelial production of prostacyclin, while leaving the platelet production of TXA2 intact [70,71].

More long-term data are needed to fully evaluate the extent to which these important adverse side effects may be offset by other beneficial effects of NSAIDs and selective COX-2 inhibitors in cancer chemoprevention.

2.5. Esophageal carcinogenesis

Esophageal adenocarcinoma (EAC) is generally considered to develop from gastroesophageal reflux disease through Barrett’s esophagus (BE). Over the past years, accumulating evidence has been obtained suggesting that increased COX-2 expression could be responsible for chronic inflammation related esophageal cancer promotion. Indeed, the incidence of COX-2 protein expression gradually increases with the development of esophageal lesions, from 75% in metaplasia, to 83% in low-grade dysplasia and up to 100% in high-grade dysplasia and EAC [72]. Shirvani et al. demonstrated that the expression of COX-2 increases parallel to the grade of dysplasia observed in BE [73]. Moreover, the same group demonstrated in an ex vivo model that both gastric acid and bile significantly elevated the expression of COX-2 [73]. Zhang et al. using a duodenogastroesophageal reflux model including unconjugated and conjugated bile acids, reported increased COX-2 expression in the esophageal mucosa followed by PGE2 production. Further, increased PG synthesis caused stimulation of cell proliferation and contributed to the development of dysplasia in Barrett's epithelium [74]. A recent metanalysis by Abnet et al. found a significant reduction in the incidence of esophageal cancer in aspirin or non-aspirin NSAID users (OR 0.64; 95% CI, 0.52–0.79 and OR 0.65; 95% CI, 0.50–0.85 respectively) [75].

Experimental models have demonstrated reduced expression of an apoptosis ligand, Fas (CD-95), in esophageal dysplastic and malignant tissues [76]. This reduced expression is linked to overexpression of COX-2, which in turn down-regulates the expression of Fas ligand [40]. A mechanism for the inhibitory effects of aspirin and NSAIDs on the occurrence of EAC could be the induction of apoptosis by COX-2 inhibition. Overexpression of COX-2 also was associated with increased levels of bcl-2, a proapoptotic protein that induced resistance to apoptosis [77]. Therefore, selective COX-2 inhibitors may up-regulate the expression of Fas receptors on the cell surface in subjects with Barrett dysplasia and have an inhibiting role in esophageal carcinogenesis by influencing apoptosis and cellular proliferation. Finally, COX-2 expression might be a prognostic marker in patients with Barrett's adenocarcinoma, as expression of COX-2 correlates with patient survival. Further support for the role of COX-2 derived PGs in the carcinogenesis emerged from the animal study by Buttar et al. in which both non-selective COX inhibitor (sulindac) and selective COX-2 blocker (MF tricyclic) significantly attenuated the incidence of Barrett adenocarcinoma [78]. Moreover, Kaur et al. studied the effect of COX-2 inhibitor (rofecoxib) on the marker of cell proliferation PCNA. In BE the authors found a significant increased PCNA expression as compared to normal esophageal mucosa. In addition, therapy with rofecoxib caused a significant inhibition of cell proliferation as evidenced by the decreased PCNA expression. Rofecoxib therapy led also to significant down-regulation of COX-2 expression in the Barrett's epithelium [79]. The suppressive effects of a COX-2 inhibitor, NS398, on the epithelium of BE have been demonstrated in two independent in vitro studies [80,81]. An increase
in apoptosis and a suppression of cell proliferation are supposed to be responsible for the inhibition of cancer cells. Furthermore, in a study carried out using a carcinogen-induced rodent model, selective COX-2 inhibitors have been reported to prevent the development of esophageal cancer. *N*-nitrosomethylbenzylamine-induced esophageal tumourigenesis in rats was prevented by the administration of another selective COX-2 inhibitor, JTE-522 [82]. Nevertheless, a Chemoprevention Barrett's Esophagus Trial (CBET) started in 2003 as a phase IIb, multicenter, randomized, double-masked, placebo-controlled study of celecoxib in patients with Barrett's dysplasia failed to prevent progression of Barrett's dysplasia to cancer. However, the apparent inability of celecoxib, compared with placebo, to decrease the percentage of samples with dysplasia is probably due to the several limitations of the study (previous diagnosis of displasia but no evidence of displasia at enrollment, imperfect biopsy sampling, natural reversion of dysplasia without any intervention, inadequate utilization of dysplasia grading as predictor of cancer because of the low intra- and interobserver agreement among pathologists) [83].

2.6. Gastric carcinogenesis

Gastric cancer (GC) is one of the most frequent malignancies worldwide [84]. The development of GC, at least of intestinal type, occurs on the basis of atrophy-metaplasia-dysplasia-sequence [85]. This multistep process is triggered by *Helicobacter pylori* (*H. pylori*) infection [85]. Indeed, the colonization of gastric mucosa with this bacterium causes a chronic inflammatory reaction with increased production of proinflammatory cytokines and generation of reactive oxygen species [86]. Interestingly, the presence of *H. pylori* also correlates with an up-regulation of the expression of COX-2 mRNA and PGE2 in GC cell lines [87].

Normal gastric mucosa scarcely expresses COX-2, but the expression of COX-2 and the production of eicosanoids (especially PGE2) increases through the multistep process of gastric carcinogenesis [88,89]. Ristimaki *et al.* described for the first time in 1997 an elevated expression of COX-2 in GC [90]. Since then, numerous studies have reported the relationship between COX-2 expression and gastric carcinogenesis. Sun *et al.* by immunohistochemistry reported a progressive positive rate of COX-2 from superficial gastritis, to gastric atrophy, intestinal metaplasia, dysplasia, and cancer (10.0%, 35.7%, 37.8%, 41.7%, and 69.5%, respectively) [91]. The COX-2 expression is more frequent in intestinal type than in diffuse type GC [92,93], and it also correlates with tumour size, depth of invasion, lymph node metastasis, lymphatic invasion, clinical stage, and prognosis [94–98]. This suggests that COX-2 expression may be an early event in gastric carcinogenesis process even if, the precise mechanisms leading to the overexpression of COX-2 are still not fully understood. However, there is evidence that proinflammatory cytokines and different gastric mucosal growth factors such as transforming growth factor alpa (TGFα) or hepatocyte growth factor (HGF) or finally gastrin could be involved in this process [99].

Previous studies demonstrated an increased gastrin level in the GC tissue. Gastrin is a potent stimulator of HGF expression and possesses also anti-apoptotic capabilities by inducing the antiapoptotic- proteins Bcl-2 and surviving [100,101]. The importance of gastrin and its precursor progastrin in mediating of COX-2 dependent gastric carcinogenesis was demonstrated in humans with GC treated with COX-2 inhibitor rofecoxib [102]. Treatment of GC patients with rofecoxib (50 mg/day)
resulted in a significant decrease in plasma and tumour contents of both progastrin and gastrin levels, and this was accompanied by the increased expression of proapoptotic proteins such as Bax and caspase-3 with a concomitant reduction in Bcl-2 and survivin expression. The blockade of COX-2 was also associated with a decrease in the serum level of proinflammatory cytokines IL-8 and TNFα being also involved in the gastric carcinogenesis [103].

Experimental evidence has shown that COX-2 influences key cellular events, including apoptosis, cell cycle control, cell proliferation, and angiogenesis [50,77,104–106]. Selective COX-2 inhibitors (NS-398 and JTE-522), indomethacin, and aspirin can suppress cell replication, induce apoptosis, and reduce epidermal growth factor in gastric carcinoma cell lines (KATO III) [107–109]. Nam et al. examined the effect of nimesulide on gastric carcinogenesis using an N-methyl-N-nitrosourea (NMU)-induced and an \textit{H pylori}-infected mouse model, demonstrating that gastric tumours developed in 68.8% of mice given both MNU and \textit{H pylori}, whereas the tumour incidence in the mice receiving nimesulide in addition to MNU and \textit{H pylori} was 27.8% [110]. More recently COX-2 was proven to have a strong relationship with gastric tumourigenesis in a study using transgenic mice. In the transgenic model expressing both COX-2 and microsomal prostaglandin E synthase (mPGES)-1, the animals developed inflammation-associated hyperplastic gastric tumours in the proximal glandular stomach [111]. In addition, NS-398 treatment for four weeks completely suppressed the gastric hypertrophy, thereby reducing the mucosal thickness in the same model [112].

Epidemiologic studies also have shown a decreased frequency of GC in people who take NSAIDs, Several case-control and cohort studies on NSAIDs use in gastric carcinoma have demonstrated a chemopreventive effect of NSAIDs [113–115]. Coogan et al. found that regular NSAID use (at least 4 days a week for >3 months) reduced the risk of GC in a hospital-based-case-control study of 254 patients (OR 0.3; 95% CI, 0.1–0.6). The protective effect was more pronounced among those patients using NSAIDs continually for >5 years (OR 0.2; 95% CI, 0.1–0.7) than for those using NSAIDs for <5 years (OR 0.4; 95% CI, 0.1–0.9) [114]. In a large cohort study of 635,031 subjects followed over 6 years, the American Cancer Society demonstrated that regular exposure to aspirin (>16 times/month) exerted a protective effect against GC; aspirin users were found to have approximately 50% the risk of GC compared with nonusers (OR = 0.53; 95% CI, 0.34–0.81) [93]. A recent metaanalysis by Abnet et al. found a significant reduction in the incidence of GC in aspirin or non-aspirin NSAID users (OR 0.74; 95% CI, 0.64–0.87 and OR 0.79; 95% CI, 0.71–0.89 respectively) [75].

This evidence suggests that inhibition of COX-2 may be an attractive target for treatment and prevention of GC. However, upper gastrointestinal bleeding is a common side-effect of aspirin therapy, so co-administration of aspirin and proton-pump inhibitors is an attractive option in this setting, and is currently being studied in the AspECT study of esomeprazole and aspirin in patients with Barrett’s esophagus [116].

2.7. Colorectal carcinogenesis

Colorectal cancer is one of the most popular cancers in westernized countries [117]. CRC develops in a stepwise manner from aberrant crypts to adenomas, with increasing grade of dysplasia and finally to cancer. According to this adenoma-carcinoma sequence model, carcinogenesis proceeds through the
accumulation of series of epigenetic and genetic alterations involving several tumour-suppressor genes (i.e., APC and p53) and oncogenes (i.e., k-ras) [118,119]. Among these COX-2 oncogene has been most intensively elucidated in both basic and clinical research due to its pathogenetic implication.

In normal human epithelium, COX-2 generally is down-regulated and is not expressed in the gastrointestinal tract. Dubois et al. were the first to report increased expression of COX-2 in CRC [120]. Their original observation was followed by several reports that confirmed increased COX-2 expression in this setting. An epoch-making paper was published by Oshima et al. in 1996 about the contribution of COX-2 to carcinogenic sequence in Wnt/Apc/Tcf pathway. They induced COX-2 mutations in APCΔ716 knockout mice, which led to the development of numerous polyps in the intestine. In COX-2−/− APCΔ716 and COX-2+/− APCΔ716 mice, the number of polyps dramatically decreased by 86% and 66%, in comparison to that in the littermate COX-2+/+ APCΔ716 mice [121].

Many studies have demonstrated that COX-2 is expressed early during the adenoma-carcinoma sequence, suggesting COX-2 should be in first line linked to the colorectal carcinogenesis. COX-2 expression is up-regulated by approximately 50% in colorectal adenoma [122] and 85–90% in CRC [105]. COX-2 overexpression appears to be associated with both the histological type and the location of the tumours. Overexpression was less prominent in tumours with signet cell morphology and was found more frequently in rectal carcinoma compared to carcinoma at others sites in the colon [123,124]. Furthermore the overexpression of COX-2 in CRCs appears to be associated with the genetic and epigenetic make-up of the tumours being significantly lower in proximal carcinomas that has the micro satellite instability (MSI) phenotype [123].

COX-2 is also expressed in the stromal compartment of adenomatous polyps and of invasive carcinomas both in experimental animal models and in humans. These stromal cells have the morphological and immunohistochemical characteristics of inflammatory cells [125–127].

Transfection of human CRC cells with a COX-2 expression vector resulted in increased invasiveness and activation of matrix metalloproteinases compared to the parental cell line. COX-2 overexpressing cells also produced proangiogenic factors, stimulated endothelial migration and tube formation and produced the proangiogenic factor VEGF [128].

In chemically (1,2-dimethylralazine -DMH- and azoxymethane -AOM-), induced CRC in rat, the inhibition of PGs by non-selective NSAIDs and selective COX-2 inhibitors significantly reduced formation of aberrant crypts and development of adenomas and CRC [129]. Moreover, coxibs (rofecoxib) and non-selective NSAID (sulindac) significantly reduced the number and size of intestinal polyps in the mice with dysfunctional APC gene (APCD716 mice) [130]. Jacoby et al. by using the Min mice model showed that celecoxib decreased not only tumour size but also caused a decrease in the size of established polyps in the regression study [131].

In human CRC cell lines, HCA-7, which express high levels of COX-2 protein constitutively, and HCT-116 cells, which lack COX-2 protein, studies were conducted to investigate the relationship between inhibition of intestinal cancer growth and selective inhibition of the COX-2 pathway. Treatment of nude mice implanted with HCA-7 cells with a selective COX-2 inhibitor (SC-58125) reduced tumour formation by 85–90%. Colony formation of cultured HCA-7 cells also was inhibited by SC-58125. On the other hand, SC-58125 had no effect on HCT-116 implants in nude mice or colony formation in culture [132]. In addition Chan et al. found that regular use of aspirin appears to reduce the risk of CRCs that overexpress COX-2 (RR 0.64; 95% CI, 0.52–0.78) but not the risk of
CRCs with weak or absent expression of COX-2 (RR 0.94; 95% CI, 0.73–1.26) [133]. This evidence suggests that a correlation may exist between inhibition of CRC cell growth and selective inhibition of the COX-2 enzyme.

In a National Cancer Institute-sponsored double-blind, placebo-controlled trial, celecoxib helped to reduce the number of colon polyps that occurred in patients with FAP. In this study, 77 patients were randomly assigned to treatment with celecoxib (100 or 400 mg twice/day) or placebo for 6 months. After 6 months, the patients receiving celecoxib 400 mg twice/day had a 28.0% reduction in the mean number of colorectal polyps (p = 0.003) and a 30.7% reduction in the polyp size (p = 0.001), as compared with reductions of 4.5% and 4.9%, respectively, in the placebo group. The occurrence of adverse events was similar among the groups [134]. The results of the study led to the approval of celecoxib by the United States Food and Drug Administration (FDA) as an adjunct to usual care for patients with FAP. In another placebo-controlled study, rofecoxib given daily at a dose 25 mg, significant decreased the size and number of rectal polyposis in patients with FAP after 9 months [135]. Finally, Phillips et al. showed a significant reduction in duodenal polyps in patients with FAP treated with selective COX-2 inhibitor [136].

The data obtained from FAP patients encouraged the conduction of further studies in patients with sporadic adenomas and CRC. Baron et al. investigated the adenoma recurrence in 1121 patients with a history of sporadic colorectal adenomas randomized to receive placebo (n = 372), 81 mg aspirin (n = 377) or 325 mg of aspirin (n = 375) daily. Relative risks for advanced lesions were 0.59 (0.38–0.92) in the 81 mg group and 0.83 (0.55–1.23) in 325 mg group as compared to placebo. Surprisingly, the lower aspirin dose had stronger chemopreventive effect that the higher one. However, the assessment of possible chemopreventive effect of aspirin on colorectal carcinogenesis was limited by the short follow-up time of the study [137]. A recent prospective cohort study involving 1,279 subjects (549 who regularly used aspirin, 730 who did not use aspirin) with a diagnosed CRC, followed up to 12 years has shown a lower risk of CRC specific and overall mortality in aspirin users vs non-users (HR 0.71; 95% CI, 0.53-0.95). [138]. The Approve trial, a randomized multicenter, placebo controlled, double blind trial to investigate whether the chronic use of the coxib (rofecoxib 25 mg daily) would reduce the adenoma recurrence in 2,586 patients with a history of colorectal adenomas. Therapy with rofecoxib was associated with a significant reduction in adenoma number and size. Unfortunately, an increase in rofecoxib associated cardiovascular adverse events beginning at 18 months was also noted, which led to early study termination [139]. Similarly, Bertagnolli et al. in a five-years efficacy and safety analysis of the adenoma prevention with celecoxib trial, found an inhibitory effect of celecoxib in colorectal adenoma formation but they reported an elevated risk for cardiovascular and thrombotic adverse events [6% (RR, 1.6; 95% CI, 1.0–2.5) and 7.5% (RR, 1.9; 95% CI, 1.2–3.1) in celecoxib 200 and 400 mg twice daily users, respectively compared to 3.8% in placebo group [140].

The role of COX-2 inhibitors has been investigated in the treatment of advanced human CRC. The 14-day therapy with celecoxib (200 mg/day) caused a significant decrease in the progastrin and gastrin levels in the CRC tissue as well as significant decrease in the survivin expression [141]. Based upon these results it has been hypothesized that celecoxib therapy could contribute to the treatment of CRC via suppression of the anti-apoptotic proteins and reduction in progastrin-promoted tumour growth.

Since the overexpression of COX-2 in tumour may counteract the efficacy of cytotoxic chemotherapy due to the apoptosis resistance, the combination of chemotherapy with coxibs seems to be an attractive
strategy to enhance the antitumour activity. Until now, the number of clinical studies in which rofecoxib was administered with chemotherapy in patients with CRC is very limited. Beccera et al. reported a phase II study in which rofecoxib was administered in combination with 5-fluorouracil and leucovorin in patients with metastatic CRC. The study was terminated when it was noted an increased toxicity (upper gastrointestinal bleeding, stomatitis, thrombocytopenia, diarrhea) in patients treated with chemotherapy and rofecoxib [142]. The addition of COX-2 inhibitor to the chemotherapy did not increase the efficacy of the antitumour activity of the chemotherapy. Despite these disappointing results, further studies with chemotherapy and COX-2 inhibitors will be needed to determine whether specific COX-2 therapy is able to improve patient outcome with a reasonable safety profile.

Finally, there are some groups postulating that both COX-isoforms are involved in the intestinal tumourigenesis. Chulada et al. demonstrated that deficiency of either COX-1 and COX-2 caused similar reduction in intestinal tumourigenesis in Min mice having a mutation in the APC gene and spontaneously developing intestinal adenomas. Furthermore, both COX-isoforms contributed to PGE2 production in polyps [143]. Finally, the inhibitory effect of non-selective NSAIDs and coxibs was demonstrated in xenograft mice models in which CRC cell lines are injected and form tumours with metastasis [62].

3. Conclusions

Mounting evidence gained from studies with cancer cell lines, mouse models and a number of clinical trials with both non-selective and selective COX-2 inhibitors support the notion of an important role for the COX-PG pathway in the development of gastrointestinal tumours. However, the mechanisms of the anti-tumoural action of the COX-2 inhibitors still remain to be defined and may vary from agent to agent and tumour to tumour.

In vitro studies have shown a mixture of COX-related mechanisms in controlling proliferation and apoptosis balance. Animal model studies are often performed with much higher pharmacological doses than those clinically achievable. Human observational studies are prevalently of the case-control type and often suffer from inadequate sample size to avoid a type II statistical error. In addition, more studies are needed to define the lowest effective dose, the age at which to initiate therapy, the optimum treatment duration and the subpopulations for which the benefits of chemoprevention outweigh the risks of adverse side effects. Furthermore, due to the high cost of these new agents, cost-effectiveness analyses must be carried out to optimize the allocation of resources. The cumulative probability of developing a lesion from birth to 80 years of age is less than 4% thus, in the general population, more than 95% of people treated prophylactically with COX-2 inhibitors will not benefit.

Understanding the molecular mechanisms of COX-2 and its downstream targets will help to identify specific molecular targets for developing new drugs which target this pathway, however, until now inconsistency still exists regarding the specific role of COX-2 in linking inflammation and cancer.

4. Future Directions

There are many reasons to be optimistic that tumour formation mediated by abnormal COX-2 activity can be effectively and safely targeted by chemoprevention regimens. Available anti-COX-2 agents vary substantially in their activities, with different degrees of COX-1 versus COX-2 selectivity,
differences in antitumour efficacy, and differences in toxic effects. In the balance of factors that maintain control over the highly reactive prostaglandin system, small differences in agent selectivity, dose, drug metabolism, and patient reactivity or predisposition to toxic effects could be important. Presently, we have little information about the relation between these important variables and agent efficacy and toxic effects. The rapid development of safe and effective inhibitors targeting individual COX enzymes could not only dramatically improve our understanding of the function of COX-2, but also result in discovery of COX independent functions of NSAIDs, providing important hints for future drug design.

An exciting, novel concept in cancer chemoprevention may be the use of combination therapy, which may allow dose reduction (and hence decreased systemic bioavailability) of NSAIDs or coxibs when combined with other anti-cancer agents, e.g., epidermal growth factor receptor inhibitors. Alternatively, other steps in PG biosynthesis and signalling represent potential targets. Indeed, pharmacological inhibitors of PGE2-EP receptors, which have anti-neoplastic activity, have been generated. Development of specific inhibitors for individual enzymes and receptors will be dependent on better understanding of the roles of particular PGs and their signaling receptors in health and disease. Finally, inducible expression of COX-2 is tightly controlled at the transcriptional and translational level in a cell-specific manner. Therefore, targeting mechanisms controlling neoplastic COX-2 regulation in stromal and epithelial elements of tumours may provide an alternative in tumour-specific COX-2 inhibition, avoiding the side effects related to systemic COX-2 inhibition.

In the light of an innovative alternative to these pharmacological approaches, it has been proposed the possible strategy to achieve a strong and selective inhibition of COX-2 enzyme by using the mechanism of RNA Interference targeted against its mRNA. Anti-COX-2 RNA molecules can be generated in CRC cells from short hairpin RNA precursors, delivered in vitro by a retroviral expression system, and induce a significant and stable silencing of overexpressed COX-2 in human CRC cells. As a safer alternative to viral approach, nonpathogenic bacteria (E. coli) can be engineered to invade eukaryotic cells. Moreover, the involvement of micro-RNAs in COX-2 posttranscriptional regulation opens up the possibility to exploit an endogenous silencing mechanism to knockdown overexpressed COX-2. Thus, these recent strategies disclose new challenging perspectives for selectively inhibiting COX-2 enzyme.

References

1. Balkwill, F.; Mantovani, A. Inflammation and cancer: back to Virchow? Lancet 2001, 357, 539-545.
2. Philip, M.; Rowley, D.A.; Schreiber, H. Inflammation as a tumour promoter in cancer induction. Semin. Cancer Biol. 2004, 14, 433-439.
3. Coussens, L.M.; Werb, Z. Inflammation and cancer. Nature 2002, 420, 860-867.
4. Colotta, F.; Allavena, P.; Sica, A.; Garlanda, C.; Mantovani, A. Cancer-related inflammation, the seventh hallmark of cancer: links to genetic instability. Carcinogenesis 2009, 30, 1073-1081.
5. Itzkowitz, S.H.; Yio, X. Inflammation and cancer. IV. Colorectal cancer in inflammatory bowel disease: the role of inflammation. Am. J. Physiol. Gastrointest. Liver Physiol. 2004, 287, G7-17.
6. Seril, D.N.; Lia, J.; Yang, G.Y.; Yang, C.S. Oxidative stress and ulcerative colitis associated carcinogenesis: studies in humans and animal models. Carcinogenesis 2003, 24, 353-362.
7. Macarthur, M.; Hold, G.L.; El-Omar, E.M. Inflammation and cancer. II. Role of chronic inflammation and cytokine polymorphisms in the pathogenesis of gastrointestinal malignancy. *Am. J. Physiol. Gastrointest. Liver Physiol*. 2004, 286, G515-G520.

8. Whitcomb, D.C. Inflammation and cancer V. Chronic pancreatitis and pancreatic cancer. *Am. J. Physiol. Gastrointest. Liver Physiol*. 2004, 287, G315-G319.

9. Cordon-Cardo, C.; Prives, C. Commentary: at the crossroads of inflammation and tumourigenesis. *J. Exp. Med*. 1999, 190, 1367-1370.

10. Nathan, C. Points of control in inflammation. *Nature* 2002, 420, 846-852.

11. Maeda, H.; Akaike, H. Nitric oxide and oxygen radicals in infection, inflammation, and cancer. *Biochemistry* 1998, 63, 854-865.

12. Fulton, A.M.; Loveless, S.E.; Heppner, G.H. Mutagenic activity of tumour-associated macrophages in Salmonella typhimurium strains TA98 and TA100. *Cancer Res*. 1984, 44, 4308-4311.

13. Pollard, J.W. Tumour-educated macrophages promote tumour progression and metastasis. *Nat. Rev. Cancer* 2004, 4, 71-78.

14. Coussens, L.M.; Werb, Z. Inflammatory cells and cancer: think different! *J. Exp. Med*. 2001, 193, F23-26.

15. Mantovani, A.; Bottazzi, B.; Colotta, F.; Sozzani, S.; Ruco, L. The origin and function of tumour-associated macrophages. *Immunol. Today* 1992, 13, 265-270.

16. Mantovani, A.; Bussolino, F.; Dejana, E. Cytokine regulation of endothelial cell function. *FASEB J.* 1992, 6, 2591-2599.

17. Coussens, L.M.; Tinkle, C.L.; Hanahan, D.; Werb, Z. MMP-9 supplied by bone marrow-derived cells contributes to skin carcinogenesis. *Cell* 2000, 103, 481-490.

18. Dranoff, G. Cytokines in cancer pathogenesis and cancer therapy. *Nat. Rev. Cancer* 2004, 4, 11-22.

19. Negus, R.P.M.; Stamp, G.W.H.; Relf, M.G.; Burke, F.; Malik, S.T.; Bernasconi, S.; Allavena, P.; Sozzani, S.; Mantovani, A.; Balkwill, F.R.. The detection and localization of monocyte chemoattractant protein-1 (MCP-1) in human ovarian cancer. *J. Clin. Invest*. 1995, 95, 2391-2396.

20. Luboshits, G.; Shina, S.; Kaplan, O.; Engelberg, S.; Nass, D.; Lifshitz-Mercer, B.; Chaitchik, S.; Keydar, I.; Ben-Baruch, A.. Elevated expression of the CC chemokine regulated on activation, normal T cell expressed and secreted (RANTES) in advanced breast carcinoma. *Cancer Res.* 1999, 59, 4681-4687.

21. Daniel, D.; Meyer-Morse, N.; Bergsland, E.K.; Dehne, K.; Coussens, L.M.; Hanahan, D. Immune enhancement of skin carcinogenesis by CD4+ T cells. *J. Exp. Med*. 2003, 197, 1017-1028.

22. Wilson, J.; Balkwill, F. The role of cytokines in the epithelial cancer microenvironment. *Semin. Cancer Biol.* 2002, 12, 113-120.

23. Ardestani, S.K.; Inserra, P.; Solkoff, D.; Watson, R.R. The role of cytokines and chemokines on tumour progression: a review. *Cancer Detect. Prev.* 1999, 23, 215-225.

24. Hanahan, D.; Weinberg, R.A. The hallmarks of cancer. *Cell* 2000, 100, 57-70.

25. Herschman, H.R. Prostaglandin synthase 2. *Biochim. Biophys. Acta* 1996, 1299, 125-140.

26. Diaz, B.L.; Arm, J.P. Phospholipase A2. *Prostag. Leukotr. Ess.* 2003, 69, 87-97.

27. Reddy, S.T.; Herschman, H.R. Ligand-induced prostaglandin synthesis requires expression of the TIS10/PGS-2 prostaglandin synthase gene in murine fibroblasts and macrophages. *J. Biol. Chem.* 1994, 269, 15473-15480.
28. Chandrasekharan, N.V.; Dai, H.; Roos, K.L.; Evanson, N.K.; Tomsik, J.; Elton, T.S.; Simmons, D.L. COX-3, a cyclooxygenase-1 variant inhibited by acetaminophen and other analgesic/antipyretic drugs: cloning, structure, and expression. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 13926-13931.

29. Fosslien, E. Biochemistry of cyclooxygenase COX-2 inhibitors and molecular pathology of COX-2 in neoplasia. *Crit. Rev. Clin. Lab. Sci.* **2000**, *37*, 431-502.

30. Saukkonen, K.; Rintahaka, J.; Sivula, A.; Buskens, C.J.; Van Rees, B.P.; Rio, M.C.; Haglund, C.; van Lanschot, J.J.; Offerhaus, G.J.; Ristimaki, A. Cyclooxygenase-2 and gastric carcinogenesis. *APMIS* **2003**, *111*, 915-925.

31. Hata, A.N.; Breyer, R.M. Pharmacology and signaling of prostaglandin receptors: multiple roles in inflammation and immune modulation. *Pharmacol. Ther.* **2004**, *103*, 147-166.

32. Kota, B.P.; Huang, T.H.; Roufogalis, B.D. An overview on biological mechanisms of PPARs. *Pharmacol. Res.* **2005**, *51*, 85-94.

33. Leone, V.; di Palma, A.; Ricchi, P.; Acquaviva, F.; Giannouli, M.; Di Prisco, A.M.; Iuliano, F.; Acquaviva, A.M. PGE2 inhibits apoptosis in human adenocarcinoma Caco-2 cell line through Ras-PI3K association and cAMP-dependent kinase A activation. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2007**, *293*, G673-681.

34. Pradono, P.; Tazawa, R.; Maemondo, M.; Tanaka, M.; Usui, K.; Saijo, Y.; Hagiwara, K.; Nukiwa, T. Gene transfer of thromboxane A(2) synthase and prostaglandin I (2) synthase antithetically altered tumour angiogenesis and tumour growth. *Cancer Res.* **2002**, *62*, 63-66.

35. Yokoyama, I.; Hayashi, S.; Kobayashi, T.; Negita, M.; Yasutomi, M.; Uchida, K.; Takagi, H. Prevention of experimental hepatic metastasis with thromboxane synthase inhibitor. *Res. Exp. Med.* **1995**, *195*, 209-215.

36. Wang, D.; Buchanan, F.G.; Wang, H.; Dey, S.K.; DuBois, R.N. Prostaglandin E2 enhances intestinal adenoma growth via activation of the Ras-mitogen-activated protein kinase cascade. *Cancer Res.* **2005**, *65*, 1822-1829.

37. Cao, Y.; Prescott, S.M. Many actions of cyclooxygenase-2 in cellular dynamics and in cancer. *J. Cell. Physiol.* **2002**, *190*, 279-286.

38. Sheng, H.; Shao, J.; Morrow, J.D.; Beauchamp, R.D.; DuBois, R.N. Modulation of apoptosis and Bcl-2 expression by prostaglandin E2 in human colon cancer cells. *Cancer Res.* **1998**, *58*, 362-366.

39. Poligone, B.; Baldwin, A.S. Positive and negative regulation of NF-kappa B by COX-2: roles of different prostaglandins. *J. Biol. Chem.* **2001**, *276*, 38658-38664.

40. Nzeako, U.C.; Guicciardi, M.E.; Yoon, J.H.; Bronk, S.F.; Gores, G.J. COX-2 inhibits Fas-mediated apoptosis in cholangiocarcinoma cells. *Hepatology* **2002**, *35*, 552-559.

41. Pasricha, P.J.; Bedi, A., O'Connor, K.; Rashid, A.; Akhtar, A.J.; Zahurak, M.L.; Piantadosi, S.; Hamilton, S.R.; Giardiello, F.M. The effects of sulindac on colorectal proliferation and apoptosis in familial adenomatous polyposis. *Gastroenterology* **1995**, *109*, 994-998.

42. Boolbol, S.K.; Dannenberg, A.J.; Chadburn, A.; Martucci, C.; Guo, X.J.; Ramonetti, J.T.; Abreu-Goris, M.; Newmark, H.L.; Lipkin, M.L.; De Cosse, J.J.; Bertagnolli, M.M. Cyclooxygenase-2 overexpression and tumour formation are blocked by sulindac in a murine model of familial adenomatous polyposis. *Cancer Res.* **1996**, *56*, 2556-2560.
43. Mahmoud, N.N.; Bilinski, R.T.; Churchill, M.R.; Edelmann; W.; Kucherlapati; R.; Bertagnolli, M.M. Genotype-phenotype correlation in murine Apc mutation: differences in enterocyte migration and response to sulindac. *Cancer Res.* 1999, 59, 353-359.

44. Mahmoud, N.N.; Dannenberg, A.J.; Mestre, J.; Bilinski, R.T.; Churchill, M.R.; Martucci, C.; Newmark, H.; Bertagnolli, M.M. Aspirin prevents tumours in a murine model of familial adenomatous polyposis. *Surgery* 1998, 24, 225-231.

45. Samaha, H.S.; Kellogg, G.J.; Steele, V.; Rao, C.V.; Reddy, B.S. Modulation of apoptosis by sulindac, curcumin, phenylethyl-3-methylcafflate, and 6-phenylethyl isothiocyanate: apoptotic index as a biomarker in colon cancer chemoprevention and promotion. *Cancer Res.* 1997, 57, 1301-1305.

46. Uefuji, K.; Ichikura, T.; Shinomiya, N.; Mochizuki, H. Induction of apoptosis by JTE-522, a specific cyclooxygenase-2 inhibitor, in human gastric cancer cell lines. *Anticancer Res.* 2000, 20, 4279-4284.

47. Li, M.; Lotan, R.; Levin, B.; Tahara, E.; Lippman, S.M.; Xu, X.C. Aspirin induction of apoptosis in esophageal cancer: a potential for chemoprevention. *Cancer Epidemiol. Biomarkers Prev.* 2000, 9, 545-549.

48. Grösch, S.; Tegeder, I.; Niederberger, E.; Bräutigam, L.; Geisslinger, G. COX-2 independent induction of cell cycle arrest and apoptosis in colon cancer cells by the selective COX-2 inhibitor celecoxib. *FASEB J.* 2001, 15, 2742-2744.

49. Molina, M.A.; Sitja-Arnau, M.; Le moine, M.G.; Frazier, M.L.; Sinicrope, F.A. Increased cyclooxygenase-2 expression in human pancreatic carcinomas and cell lines: growth inhibition by nonsteroidal anti-inflammatory drugs. *Cancer Res.* 1999, 59, 4356-4362.

50. Tsuji, M.; Kawano, S.; Tsuji, S.; Sawaoka, H.; Hori, M.; DuBois, R.N. Cyclooxygenase regulates angiogenesis induced by colon cancer cells. *Cell* 1998, 93, 705-716.

51. Trompezinski, S.; Pernet, I.; Schmitt, D.; Viac, J. UV radiation and prostaglandin E2 up-regulate vascular endothelial growth factor (VEGF) in cultured human fibroblasts. *Inflamm. Res.* 2001, 50, 422-427.

52. Pai, R.; Szabo, I.L.; Soreghan, B.A.; Atay, S.; Kawanaka, H.; Tarnawski, A.S. PGE(2) stimulates VEGF expression in endothelial cells via ERK2/JNK1 signaling pathways. *Biochem. Biophys. Res. Commun.* 2001, 286, 923-928.

53. Kage, K.; Fujita, N.; Oh-hara, T.; Ogata, E.; Fujita, T.; Tsuruo, T. Basic fibroblast growth factor induces cyclooxygenase-2 expression in endothelial cells derived from bone. *Biochem. Biophys. Res. Commun.* 1999, 254, 259-263.

54. Salcedo, R.; Zhang, X.; Young, H.A.; Michael, N.; Wasserman, K.; Ma, W.H.; Martins-Green, M.; Murphy, W.J.; Oppenheim, J.J. Angiogenic effects of prostaglandin E2 are mediated by up-regulation of CXCR4 on human microvascular endothelial cells. *Blood* 2003, 102, 1966-1977.

55. Kaidi, A.; Quatrough, D.; Williams, A.C.; Paraskeva, C. Direct transcriptional up-regulation of cyclooxygenase-2 by hypoxia-inducible factor (HIF)-1 promotes colorectal tumour cell survival and enhances HIF-1 transcriptional activity during hypoxia. *Cancer Res.* 2006, 66, 6683-6691.

56. Dormond, O.; Foletti, A., Paroz; C., Rüegg, C. NSAIDs inhibit alpha V beta 3 integrin-mediated and Cdc42/Rac-dependent endothelial-cell spreading, migration and angiogenesis. *Nat. Med.* 2001, 7, 1041-1047.
57. Knutson, K.L.; Disis, M.L. Tumour antigen-specific T helper cells in cancer immunity and immunotherapy. *Cancer Immunol. Immunother.* 2005, 54, 721-728.

58. Harris, S.G.; Padilla, J.; Koumas, L.; Ray, D.; Phipps, R.P. Prostaglandins as modulators of immunity. *Trends Immunol.* 2002, 23, 144-150.

59. Della Bella, S.; Molteni, M.; Compasso, S.; Zulian, C.; Vanoli, M.; Scorza, R. Differential effect of cyclo-oxygenase pathway metabolites on cytokine production by T lymphocytes. *Prostaglandins Leukot. Essent. Fatty Acids* 1997, 56, 177-184.

60. Holla, V.R.; Wang, D.; Brown, J.R.; Mann, J.R.; Katkuri, S.; DuBois, R.N. Prostaglandin E2 regulates the complement inhibitor CD55/decay-accelerating factor in colorectal cancer. *J. Biol. Chem.* 2005, 280, 476-483.

61. Stolina, M.; Sharma, S.; Lin, Y.; Dohadwala, M.; Gardner, B.; Luo, J.; Zhu, L.; Kronenberg, M.; Miller, P.W.; Portanova, J.; Lee, J.C.; Dubinett, S.M.. Specific inhibition of cyclooxygenase 2 restores antitumour reactivity by altering the balance of IL-10 and IL-12 synthesis. *J. Immunol.* 2000, 164, 361-370.

62. Yao, M.; Kargman, S.; Lam, E.C.; Kelly, C.R.; Zheng, Y.; Luk, P.; Kwong, E.; Evans, J.F.; Wolfe, M.M. Inhibition of cyclooxygenase-2 by rofecoxib attenuates the growth and metastatic potential of colorectal carcinoma in mice. *Cancer Res.* 2003, 63, 586-592.

63. Seibert, K.; Zhang, Y.; Leahy, K.; Hauser, S.; Masferrer, J.; Perkins, W.; Lee, L.; Isakson, P. Pharmacological and biochemical demonstration of the role of cyclooxygenase 2 in inflammation and pain. *Proc. Nat. Acad. Sci. USA* 1994, 91, 12013-12017.

64. Singh, G.; Ramey, D.R.; Morfeld, D.; Fries, J.F. Comparative toxicity of non-steroidal anti-inflammatory agents. *Pharmacol. Ther.* 1994, 62, 175-191.

65. Bjarnason, I.; Macpherson, A.J.S. Intestinal toxicity of non-steroidal anti-inflammatory drugs. *Pharmacol. Ther.* 1994, 62, 145-157.

66. Allison, M.C.; Howatson, A.G.; Torrance, C.J.; Lee, F.D.; Russell, R.I. Gastrointestinal damage associated with the use of non-steroidal anti-inflammatory drugs, *N. Engl. J. Med.* 1992, 327, 749-754.

67. Langman, M.J.S.; Weil, J.; Wainwright, P.; Lawson, D.H.; Rawlins, M.D.; Logan, R.F.A.; Murphy, M.; Vessey, M.P.; Colin-Jones, D.G. Risks of bleeding peptic ulcer associated with individual non-steroidal anti-inflammatory drugs. *Lancet* 1994, 343, 1075-1078.

68. Masferrer, J.L.; Seibert, K. Regulation of prostaglandin synthesis by glucocorticoids. *Receptor* 1994, 4, 25-30.

69. Masferrer, J.L.; Reddy, S.T.; Zweifel, B.S.; Seibert, K.; Needleman, P.; Gilbert, R.S.; Herschman, H.R. *In vivo* glucocorticoids regulate cyclooxygenase-2 but not cyclooxygenase-1 in peritoneal macrophages. *J. Pharmacol. Exp. Ther.* 1994, 270, 1340-1344.

70. FitzGerald, G.A. Coxibs and cardiovascular disease. *N. Engl. J. Med.* 2004, 351, 1709-1711.

71. Topol, E.J. Failing the public health: rofecoxib, Merck, and the FDA. *N. Engl. J. Med.* 2004, 351, 1707-1709.

72. Morris, C.D.; Armstrong, G.R.; Bigley, G.; Green, H.; Attwood, S.E. Cyclooxygenase-2 expression in the Barrett’s metaplasia-dysplasia-adenocarcinoma sequence. *Am. J. Gastroenterol.* 2001, 96, 990-996.
73. Shirvani, V.N.; Ouatu-Lascar, R.; Kaur, B.S.; Omary, M.B.; Triadafilopoulos, G. Cyclooxygenase 2 expression in Barrett's esophagus and adenocarcinoma: Ex vivo induction by bile salts and acid exposure. *Gastroenterology* 2000, 118, 487-496.

74. Zhang, F.; Altorki, N.K.; Wu, Y.C.; Soslow, R.A.; Subbaramaiah, K.; Dannenberg, A.J. Duodenal reflux induces cyclooxygenase-2 in the esophageal mucosa of rats: evidence for involvement of bile acids. *Gastroenterology* 2001, 121, 1391-1399.

75. Abnet, C.C.; Freedman, N.D.; Kamangar, F.; Leitzmann, M.F.; Hollenbeck, A.R.; Schatzkin, A. Non-steroidal anti-inflammatory drugs and risk of gastric and oesophageal adenocarcinomas: results from a cohort study and a meta-analysis. *Br. J. Cancer* 2009, 100, 551-557.

76. Hughes, S.J.; Nambu, Y.; Soldes, O.S.; Hamstra, D.; Rehmtulla, A.; Iannettoni, M.D.; Orringer, M.B.; Beer, D.G. Fas/APO-1 (CD95) is not translocated to the cell membrane in esophageal adenocarcinoma. *Cancer Res.* 1997, 57, 5571-5578.

77. Tsujii, M.; DuBois, R.N. Alterations in cellular adhesion and apoptosis in epithelial cells overexpressing prostaglandin endoperoxide synthase 2. *Cell* 1995, 83, 493-501.

78. Buttar, N.S.; Wang, K.K.; Leontovich, O.; Westcott, J.Y.; Pacifico, R.J.; Anderson, M.A.; Krishnadath, K.K.; Lutzke, L.S.; Burgart, L.J. Chemoprevention of esophageal adenocarcinoma by COX-2 inhibitors in an animal model of Barrett's esophagus. *Gastroenterology* 2002, 122, 1101-1112.

79. Kaur, B.S.; Khamnehei, N.; Iravani, M.; Nambrum, S.S.; Lin, O.; Triadafilopoulos, G. Rofecoxib inhibits cyclooxygenase 2 expression and activity and reduces cell proliferation in Barrett's esophagus. *Gastroenterology* 2002, 123, 60-67.

80. Souza, R.F.; Shewmake, K.; Beer, D.G.; Cryer, B.; Spechler, S.J. Selective inhibition of cyclooxygenase-2 suppresses growth and induces apoptosis in human esophageal adenocarcinoma cells. *Cancer Res.* 2000, 60, 5767-5772.

81. Buttar, N.S.; Wang, K.K.; Anderson, M.A.; Dierkhising, R.A.; Pacifico, R.J.; Krishnadath, K.K.; Lutzke, L.S. The effect of selective cyclooxygenase-inhibition in Barrett’s esophagus epithelium: an in vitro study. *J. Natl. Cancer Inst.* 2002, 94, 422-429.

82. Li, Z.; Shimada, Y.; Kawabe, A.; Sato, F.; Maeda, M.; Komoto, I.; Hong, T.; Ding, Y.; Kaganoi, J.; Imamura, M. Suppression of N-nitrosomethylbenzylamine (NMBA)-induced esophageal tumourigenesis in F344 rats by JTE-522, a selective COX-2 inhibitor. *Carcinogenesis* 2001, 22, 547-551.

83. Heath, E.I.; Canto, M.I.; Piantadosi, S.; Montgomery, E.; Weinstein, W.M.; Herman, J.G.; Dannenberg, A.J.; Yang, V.W.; Shar, A.O.; Hawk, E.; Forastiere, A.A. Chemoprevention for Barrett's Esophagus Trial Research Group. Secondary chemoprevention of Barrett's esophagus with celecoxib: results of a randomized trial. *J. Natl. Cancer Inst.* 2007, 99, 545-557.

84. Parkin, D.M.; Bray, F.; Ferlay, J.; Pisani, P. Global cancer statistics, 2002. *CA Cancer J. Clin.* 2005, 55, 74-108.

85. Correa, P. Human gastric carcinogenesis: a multistep and multifactorial process – First American Cancer Society Award Lecture on Cancer Epidemiology and Prevention. *Cancer Res.* 1992, 52, 6735-6740.

86. Hofman, P.; Waidner, B.; Hofman, V.; Bereswill, S.; Brest, P.; Kist, M. Pathogenesis of *Helicobacter pylori* infection. *Helicobacter* 2004, 9 (Suppl. 1), 15-22.
87. Romano, M.; Ricci, V.; Memoli, A.; Tuccillo, C.; Di Popolo, A.; Sommi, P.; Acquaviva, A.M.; Del Vecchio Blanco, C.; Bruni, C.B.; Zarrilli, R. Helicobacter pylori up-regulates cyclooxygenase-2 mRNA expression and prostaglandin E2 synthesis in MKN 28 gastric mucosal cells in vitro. *J. Biol. Chem.* 1998, 273, 28560-28563.

88. Nardone, G.; Rocco, A.; Vaira, D.; Staibano, S.; Budillon, A.; Tatangelo, F.; Sciulli, M.G.; Perna, F.; Salvatore, G.; Di Benedetto, M.; De Rosa, G.; Patrignani, P. Expression of COX-2, mPGE-synthase1, MDR-1 (P-gp), and Bcl-xl: a molecular pathway of *H pylori*-related gastric carcinogenesis. *J. Pathol.* 2004, 202, 305-312.

89. Sung, J.J.; Leung, W.K.; Go, M.Y.; To, K.F.; Cheng, A.S.; Ng, E.K.; Chan, F.K. Cyclooxygenase-2 expression in Helicobacter pylori-associated premalignant and malignant gastric lesions. *Am. J. Pathol.* 2000, 157, 729-735.

90. Ristimaki, A.; Honkanen, N.; Jankala, H.; Sipponen, P.; Harkonen, M. Expression of cyclooxygenase-2 in human gastric carcinoma. *Cancer Res.* 1997, 57, 1276-1280.

91. Sun, W.H.; Yu, Q.; Shen, H.; Ou, X.L.; Cao, D.Z.; Yu, T.; Qian, C.; Zhu, F.; Sun, Y.L.; Fu, X.L.; Su, H. Roles of *Helicobacter pylori* infection and cyclooxygenase-2 expression in gastric carcinogenesis. *World J. Gastroenterol.* 2004, 10, 2809-2813.

92. Saukkonen, K.; Nieminen, O.; van Rees, B.; Vilikki, S.; Harkonen, M.; Juhola, M.; Mecklin, J.P.; Sipponen, P.; Ristimaki, A. Expression of cyclooxygenase-2 in dysplasia of the stomach and in intestinal-type gastric adenocarcinoma. *Clin. Cancer Res.* 2001, 7, 1923-1931.

93. Yamagata, R.; Shimoyama, T.; Fukuda, S.; Yoshimura, T.; Tanaka, M.; Munakata, A. Cyclooxygenase-2 expression is increased in early intestinal-type gastric cancer and gastric mucosa with intestinal metaplasia. *Eur. J. Gastroenterol. Hepatol.* 2002, 14, 359-363.

94. Uefuji, K.; Ichikura, T.; Mochizuki, H. Expression of cyclooxygenase-2 in human gastric adenomas and adenocarcinomas. *J. Surg. Oncol.* 2001, 76, 26-30.

95. Xue, Y.W.; Zhang, Q.F.; Zhu, Z.B.; Wang, Q.; Fu, S.B. Expression of cyclooxygenase-2 and clinicopathologic features in human gastric adenocarcinoma. *World J. Gastroenterol.* 2003, 9, 250-253.

96. Murata, H.; Kawano, S.; Tsuji, S.; Tsuji, M.; Sawaoka, H.; Kimura, Y.; Shiozaki, H.; Hori, M. Cyclooxygenase-2 overexpression enhances lymphatic invasion and metastasis in human gastric carcinoma. *Am. J. Gastroenterol.* 1999, 94, 451-455.

97. Yamamoto, H.; Itoh, F.; Fukushima, H.; Hinoda, Y.; Imai, K. Overexpression of Cyclooxygenase-2 protein is less frequent in gastric cancers with microsatellite instability. *Int. J. Cancer.* 1999, 84, 400-403.

98. Joo, Y.E.; Oh, W.T.; Rew, J.S.; Park, C.S.; Choi, S.K.; Kim, S.J. Cyclooxygenase-2 expression is associated with well-differentiated and intestinal-type pathways in gastric carcinogenesis. *Digestion* 2002, 66, 222-229.

99. Konturek, P.C.; Kania, J.; Kukharsky, V.; Ocker, S.; Hahn, E.G.; Konturek, S.J. Influence of gastrin on the expression of cyclooxygenase-2, hepatocyte growth factor and apoptosis-related proteins in gastric epithelial cells. *J. Physiol. Pharmacol.* 2003, 54, 17-32.

100. Konturek, P.C.; Konturek, S.; Sulekova, Z.; Meixner, H.; Bielanski, W.; Starzynska, T.; Karczewska, E.; Marlicz, K.; Stachura, J.; Hahn, E.G. Expression of hepatocyte growth factor,
transformation growth factor alpha, apoptosis related proteins Bax and Bcl-2, and gastrin in human gastric cancer. *Aliment. Pharmacol. Ther.* **2001**, *15*, 989-999.

101. Konturek, P.C.; Konturek, S.J.; Bielanski, W.; Karczewska, E.; Pierzchalski, P.; Duda, A.; Starzynska, T.; Marlicz, K.; Popiela, T.; Hartwich, A.; Hahn, E.G. Role of gastrin in gastric carcinogenesis in Helicobacter pylori infected humans. *J. Physiol. Pharmacol.* **1999**, *50*, 857-873.

102. Konturek, S.J.; Konturek, P.C.; Bielanski, W.; Karczewska, E.; Zuchowicz, M.; Hartwich, A.; Rehfeld, J.F.; Goetze, J.P.; Hahn, E.G. Serum progastrin and its products, gastric acid secretion and serum pepsinogen I in gastric cancer. *Digestion* **2003**, *68*, 169-177.

103. Konturek, P.C.; Konturek, S.J.; Bielanski, W.; Kania, J.; Zuchowicz, M.; Hartwich, A.; Rehfeld, J.F.; Hahn, E.G. Influence of COX-2 inhibition by rofecoxib on serum and tumour progastrin and gastrin levels and expression of PPAR gamma and apoptosis-related proteins in gastric cancer patients. *Dig. Dis. Sci.* **2003**, *48*, 2005-2017.

104. Fosslien, E. Molecular pathology of cyclooxygenase-2 in neoplasia. *Ann. Clin. Lab. Sci.* **2000**, *30*, 3-21.

105. Koki, A.T.; Leahy, K.M.; Masferrer, J.L. Potential utility of COX-2 inhibitors in chemoprevention and chemotherapy. *Expert Opin. Investig. Drugs* **1999**, *8*, 1623-1638.

106. Fosslien, E. Review: molecular pathology of cyclooxygenase-2 in cancer-induced angiogenesis. *Ann. Clin. Lab. Sci.* **2001**, *31*, 325-348.

107. Fujiwara, Y.; Tarnawski, A.; Fujiwara, K.; Arakawa, T.; Kobayashi, K. Inhibitory effects of indomethacin on growth and proliferation of gastric carcinoma cells KATO III. *J. Physiol. Pharmacol.* **1993**, *44*, 147-153.

108. Sawaoka, H.; Kawano, S.; Tsuji, S.; Gunawan, E.S.; Takei, Y.; Nagano, K.; Hori, M. Cyclooxygenase-2 inhibitors suppress the growth of gastric cancer xenografts via induction of apoptosis in nude mice. *Am. J. Physiol.* **1998**, *274*, G1061-G1067.

109. Fujiwara, Y.; Schmassmann, A.; Arakawa, T.; Halter, F.; Tarnawski, A. Indomethacin interferes with epidermal growth factor binding and proliferative response of gastric KATO III cells. *Digestion* **1995**, *56*, 364-369.

110. Nam, K.T.; Hahn, K.B.; Oh, S.Y.; Yeo, M.; Han, S.U.; Ahn, B.; Kim, Y.B.; Kang, J.S.; Jang, D.D.; Yang, K.H.; Kim, D.Y. The selective cyclooxygenase-2 inhibitor nimesulide prevents Helicobacter pyloriasociated gastric cancer development in a mouse model. *Clin. Cancer Res.* **2004**, *10*, 8105-8113.

111. Oshima, H.; Oshima, M.; Inaba, K.; Taketo, M.M. Hyperplastic gastric tumours induced by activated macrophage in COX-2/mPGES-1 transgenic mice. *EMBO J.* **2004**, *23*, 1669-1678.

112. Miwa, K.; Hasegawa, H.; Fujimura, T.; Matsumoto, H.; Miyata, R.; Kosaka, T.; Miyazaki, I.; Hattori, T. Duodenal reflux through the pylorus induces gastric adenocarcinoma in the rat. *Carcinogenesis* **1992**, *13*, 2313-2316.

113. Thun, M.J.; Namboodiri, M.M.; Calle, E.E.; Flanders, W.D.; Heath, C.W., Jr. Aspirin use and risk of fatal cancer. *Cancer Res.* **1993**, *53*, 1322-1327.

114. Coogan, P.F.; Rosenberg, L.; Palmer, J.R.; Strom, B.L.; Zauber, A.G.; Stolley, P.D.; Shapiro, S. Nonsteroidal anti-inflammatory drugs and risk of digestive cancers at sites other than the large bowel. *Cancer Epidemiol. Biomarkers Prev.* **2000**, *9*, 119-123.
115. Farrow, D.C.; Vaughan, T.L.; Hansten, P.D.; Stanford, J.L.; Risch, H.A.; Gammon, M.D.; Chow, W.H.; Dubrow, R.; Ahsan, H.; Mayne, S.T.; Schoenberg, J.B.; West, A.B.; Rotterdam, H.; Fraumeni, J.F., Jr.; Blot, W.J. Use of aspirin and other nonsteroidal anti-inflammatory drugs and risk of esophageal and gastric cancer. Cancer Epidemiol. Biomarkers Prev. 1998, 7, 97-102.

116. Das, D.; Chilton, A.P.; Jankowski, J.A. Chemoprevention of oesophageal cancer and the AspECT trial. Recent Results Cancer Res. 2009, 181, 161-169.

117. Jemal, A.; Siegel, R.; Ward, E.; Murray, T.; Xu, J.; Smigal, C.; Thun, M.J. Cancer statistics, 2006. CA Cancer J. Clin. 2006, 56, 106-130.

118. Fearon, E.R.; Vogelstein, B. A genetic model for colorectal tumourigenesis. Cell 1990 61, 759-767.

119. Sinicrope, F.A.; Gill, S. Role of cyclooxygenase-2 in colorectal cancer. Cancer Metastasis Rev. 2004, 23, 63-75.

120. DuBois, R.N.; Radhika, A.; Reddy, B.S.; Entingh, A.J. Increased cyclooxygenase-2 levels in carcinogen-induced rat colonic tumours. Gastroenterology 1996, 110, 1259-1262.

121. Oshima, M.; Dinchuk, J.E.; Kargman, S.L.; Oshima, H.; Hancock, B.; Kwong, E.; Trzaskos, J.M.; Evans, J.F.; Taketo, M.M. Suppression of intestinal polyposis in Apc delta716 knockout mice by inhibition of cyclooxygenase 2 (COX-2). Cell 1996, 87, 803-809.

122. Eberhart, C.E.; Coffey, R.J.; Radhika, A.; Giardiello, F.M.; Ferrenbach, S.; DuBois, R.N. Up-regulation of cyclooxygenase 2 gene expression in human colorectal adenomas and adenocarcinomas. Gastroenterology 1994, 107, 1183-1188.

123. Karnes, W.E., Jr.; Shattuck-Brandt, R.; Burgart, L.J.; DuBois, R.N.; Tester, D.J.; Cunningham, J.M.; Kim, C.Y.; McDonnell, S.K.; Schaid, D.J.; Thibodeau, S.N. Reduced COX-2 protein in colorectal cancer with defective mismatch repair. Cancer Res. 1998, 58, 5473-5477.

124. Dimberg, J.; Samuelsson, A.; Hugander, A.; Soderkvist, P. Differential expression of cyclooxygenase 2 in human colorectal cancer. Gut 1999, 45, 730-732.

125. Chapple, K.S.; Cartwright, E.J.; Hawcroft, G.; Tisbury, A.; Bonifer, C.; Scott, N.; Windsor, A.C.; Guillou, P.J.; Markham, A.F.; Coletta, P.L.; Hull, M.A. Localization of cyclooxygenase-2 in human sporadic colorectal adenomas. Am. J. Pathol. 2000, 156, 545-553.

126. Shattuck-Brandt, R.L.; Varilek, G.W.; Radhika, A.; Yang, F.; Washington, M.K.; DuBois, R.N. Cyclooxygenase 2 expression is increased in the stroma of colon carcinomas from IL-10 (−/−) mice. Gastroenterology 2000, 118, 337-345.

127. Tomozawa, S.; Tsuno, N.H.; Sunami, E.; Hatano, K.; Kitayama, J.; Osada, T.; Saito, S.; Tsuruo, T.; Shibata, Y.; Nagawa, H. Cyclooxygenase-2 overexpression correlates with tumour recurrence, especially haematogenous metastasis, of colorectal cancer. Br. J. Cancer 2000, 83, 324-328.

128. Tsujii, M.; Kawano, S.; DuBois, R.N. Cyclooxygenase-2 expression in human colon cancer cells increases metastatic potential. Proc. Natl. Acad. Sci. USA 1997, 94, 3336-3340.

129. Reddy, B.S.; Hirose, Y.; Lubet, R.; Steele, V.; Kelloff, G.; Paulson, S.; Seibert, K.; Rao, C.V. Chemoprevention of colon cancer by specific cyclooxygenase-2 inhibitor, celecoxib, administered during different stages of carcinogenesis. Cancer Res. 2000, 60, 293-297.
130. Oshima, M.; Murai, N.; Kargman, S.; Arguello, M.; Luk, P.; Kwong, E.; Taketo, M.M.; Evans, J.F. Chemoprevention of intestinal polyposis in the Apcdelta716 mouse by rofecoxib, a specific cyclooxygenase-2 inhibitor. *Cancer Res.* 2001, 61, 1733-1740.

131. Jacoby, R.F.; Seibert, K.; Cole, C.E.; Kellogg, G.; Lubet, R.A. The cyclooxygenase-2 inhibitor celecoxib is a potent preventive and therapeutic agent in the min mouse model of adenomatous polyposis. *Cancer Res.* 2000, 60, 5040-5044.

132. Sheng, H.; Shao, J.; Kirkland, S.C.; Isakson, P.; Coffey, R.J.; Morrow, J.; Beauchamp, R.D.; DuBois, R.N. Inhibition of human colon cancer cell growth by selective inhibition of cyclooxygenase-2. *J. Clin. Invest.* 1997, 99, 2254-2259.

133. Chan, A.T.; Ogino, S.; Fuchs, C.S. Aspirin and the risk of colorectal cancer in relation to the expression of COX-2. *N. Engl. J. Med.* 2007, 356, 2131-2142.

134. Steinbach, G.; Lynch, P.M.; Phillips, R.K.; Wallace, M.H.; Hawk, E.; Gordon, G.B.; Wakabayashi, N.; Saunders, B.; Shen, Y.; Fujimura, T.; Su, L.K.; Levin, B. The effect of celecoxib, a cyclooxygenase-2 inhibitor, in familial adenomatous polyposis. *N. Engl. J. Med.* 2000, 342, 1946-1952.

135. Higuchi, T.; Iwama, T.; Yoshinaga, K.; Toyooka, M.; Taketo, M.M.; Sugihara, K. A randomized, doubleblind, placebo-controlled trial of the effects of rofecoxib, a selective cyclooxygenase-2 inhibitor, on rectal polyps in familial adenomatous polyposis patients. *Clin. Cancer Res.* 2003, 9, 4756-4760.

136. Phillips, R.K.; Wallace, M.H.; Lynch, P.M.; Hawk, E.; Gordon, G.B.; Saunders, B.P.; Wakabayashi, N.; Shen, Y.; Zimmerman, S.; Godio, L.; Rodrigues-Bigas, M.; Su, L.K.; Sherman, J.; Kellogg, G.; Levin, B.; Steinbach, G.; FAP Study Group. A randomised, double blind, placebo controlled study of celecoxib, a selective cyclooxygenase 2 inhibitor, on duodenal polyposis in familial adenomatous polyposis. *Gut* 2002, 50, 857-860.

137. Baron, J.A.; Cole, B.F.; Sandler, R.S.; Haile, R.W.; Ahnen, D.; Bresalier, R.; McKeown-Eyssen, G.; Summers, R.W.; Rothstein, R.; Burke, C.A.; Snover, D.C.; Church, T.R.; Allen, J.I.; Beach, M.; Beck, G.J.; Bond, J.H.; Byers, T.; Greenberg, E.R.; Mandel, J.S.; Marcon, N.; Mott, L.A.; Pearson, L.; Saibil, F.; van Stolk, R.U. A randomized trial of aspirin to prevent colorectal adenomas. *N. Engl. J. Med.* 2003, 348, 891-899.

138. Chan, A.T.; Ogino, S.; Fuchs, C.S. Aspirin use and survival after diagnosis of colorectal cancer. *JAMA* 2009, 302, 649-658.

139. Baron, J.A.; Sandler, R.S.; Bresalier, R.S.; Quan, H.; Riddell, R.; Lanas, A.; Bolognese, J.A.; Oxenius, B.; Horgan, K.; Loftus, S.; Morton, D.G. Approve Trial Investigators. A randomized trial of rofecoxib for the chemoprevention of colorectal adenomas. *Gastroenterology* 2006, 131, 1674-1682.

140. Bertagnolli, M.M.; Eagle, C.J.; Zauber, A.G.; Redston, M.; Breazna, A.; Kim, K.; Tang, J.; Rosenstein, R.B.; Umar, A.; Bagheri, D.; Collins, N.T.; Burn, J.; Chung, D.C.; Dewar, T.; Foley, T.R.; Hoffman, N.; Macrae, F.; Pruitt, R.E.; Saltzman, J.R.; Salzberg, B.; Sylvestrowicz, T.; Hawk, E.T. Adenoma Prevention with Celecoxib Study Investigators. Five-year efficacy and safety analysis of the Adenoma Prevention with Celecoxib Trial. *Cancer Prev. Res.* 2009, 4, 310-321.
141. Konturek, P.C.; Rembiasz, K.; Burnat, G.; Konturek, S.J.; Tusinela, M.; Bielanski, W.; Rehfell, J.; Karcz, D.; Hahn, E. Effects of cyclooxygenase-2 inhibition on serum and tumour gastrins and expression of apoptosis-related proteins in colorectal cancer. *Dig. Dis. Sci.* **2006**, *51*, 779-787.

142. Becerra, C.R.; Frenkel, E.P.; Ashfaq, R.; Gaynor, R.B. Increased toxicity and lack of efficacy of Rofecoxib in combination with chemotherapy for treatment of metastatic colorectal cancer: A phase II study. *Int. J. Cancer* **2003**, *105*, 868-872.

143. Chulada, P.C.; Thompson, M.B.; Mahler, J.F.; Doyle, C.M.; Gaul, B.W.; Lee, C.; Tiano, H.F.; Morham, S.G.; Smithies, O.; Langenbach, R. Genetic disruption of Ptgs-1, as well as Ptgs-2, reduces intestinal tumourigenesis in Min mice. *Cancer Res.* **2000**, *60*, 4705-4708.

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