Structural Alteration of Gut Microbiota during the Amelioration of Human Type 2 Diabetes with Hyperlipidemia by Metformin and a Traditional Chinese Herbal Formula: a Multicenter, Randomized, Open Label Clinical Trial

Xiaolin Tong, Jia Xu, Fengmei Lian, Xiaotong Yu, Yufeng Zhao, Lipeng Xu, Menghui Zhang, Xiyan Zhao, Jian Shen, Shengping Wu, Xiao Yan Pang, Jiaxing Tian, Chenhong Zhang, Qiang Zhou, Linhua Wang, Bing Pang, Feng Chen, Zhiping Peng, Jing Wang, Zhong Zhen, Hao Fang, Min Li, Limei Chen, Liping Zhao

Guang’annmen Hospital, China Academy of Chinese Medical Sciences, Beijing, People’s Republic of China

State Key Laboratory of Microbial Metabolism, School of Life Sciences and Biotechnology, Shanghai Jiao Tong University, Shanghai, People’s Republic of China

Graduate School, Beijing University of Chinese Medicine, Beijing, People’s Republic of China

Shanghai Centre for Systems Biomedicine, Shanghai Jiao Tong University, Shanghai, People’s Republic of China

ABSTRACT

Accumulating evidence implicates gut microbiota as promising targets for the treatment of type 2 diabetes mellitus (T2DM). With a randomized clinical trial, we tested the hypothesis that alteration of gut microbiota may be involved in the alleviation of T2DM with hyperlipidemia by metformin and a specifically designed herbal formula (AMC). Four hundred fifty patients with T2DM and hyperlipidemia were randomly assigned to either the metformin- or AMC-treated group. After 12 weeks of treatment, 100 patients were randomly selected from each group and assessed for clinical improvement. The effects of the two drugs on the intestinal microbiota were evaluated by analyzing the V3 and V4 regions of the 16S rRNA gene by Illumina sequencing and multivariate statistical methods. Both metformin and AMC significantly alleviated hyperglycemia and hyperlipidemia and shifted gut microbiota structure in diabetic patients. They significantly increased a coabundant group represented by Blautia spp., which significantly correlated with the improvements in glucose and lipid homeostasis. However, AMC showed better efficacies in improving homeostasis model assessment of insulin resistance (HOMA-IR) and plasma triglyceride and also exerted a larger effect on gut microbiota. Furthermore, only AMC increased the coabundant group represented by Faecalibacterium spp., which was previously reported to be associated with the alleviation of T2DM in a randomized clinical trial. Metformin and the Chinese herbal formula may ameliorate type 2 diabetes with hyperlipidemia via enriching beneficial bacteria, such as Blautia and Faecalibacterium spp.

IMPORTANCE

Metabolic diseases such as T2DM and obesity have become a worldwide public health threat. Accumulating evidence indicates that gut microbiota can causatively arouse metabolic diseases, and thus the gut microbiota serves as a promising target for disease control. In this study, we evaluated the role of gut microbiota during improvements in hyperglycemia and hyperlipidemia by two drugs: metformin and a specifically designed Chinese herbal formula (AMC) for diabetic patients with hyperlipidemia. Both drugs significantly ameliorated blood glucose and lipid levels and shifted the gut microbiota. Blautia spp. were identified as being associated with improvements in glucose and lipid homeostasis for both drugs. AMC exerted larger effects on the gut microbiota together with better efficacies in improving HOMA-IR and plasma triglyceride levels, which were associated with the en-
richment of Faecalibacterium spp. In brief, these data suggest that gut microbiota might be involved in the alleviation of diabetes with hyperlipidemia by metformin and the AMC herbal formula.

**KEYWORDS** clinical trial, gut microbiota, hyperlipidemia, metformin, traditional Chinese medicine, type 2 diabetes

**Type 2 diabetes mellitus (T2DM), a global epidemic disease with a rapid growth in rates of incidence and prevalence, affects people from all income levels worldwide** (1). It is a metabolic disease characterized by insulin resistance and β-cell dysfunction (2) associated with inflammation caused by overnutrition and other environmental factors (3–5). The intestinal microbiota has been proposed to play a vital role in the pathophysiology of obesity, T2DM, and other metabolic diseases (6–8). Transplantation of the gut microbiota from the obese human gut can transfer the obese phenotypes to germfree mice, indicating the causative role of gut microbiota in the development of obesity and metabolic diseases (7). Moreover, an opportunistic pathogen, *Enterobacter cloacae* B29, isolated from a diabetic obese human’s gut, can induce obesity and insulin resistance in germfree mice (9). These findings disclosed that gut microbiota can causatively arouse insulin resistance, obesity, and T2DM. Furthermore, a more recent study showed that dietary modulation of gut microbiota contributes to alleviation of both genetic and simple obesity in children (10). Therefore, targeting the gut microbiota with prebiotics, probiotics, diets, and drugs would be a novel therapeutic approach to ameliorate diabetes, hyperlipidemia, and other metabolic diseases.

Metformin is one of the most widely used oral antihyperglycemic drugs recommended as the first-line remedy for the treatment of diabetes. It also shows potential applications in the treatment or prevention of hyperlipidemia (11, 12) and cardiovascular diseases (13). Numerous studies showed that metformin might function by inhibition of hepatic gluconeogenesis via activation of the AMP-activated protein kinase (AMPK) signaling pathway (14, 15). However, in contrast to oral dosing, intravenously administered metformin does not improve glucose metabolism (16), indicating that gut microbiota might be the target. A more recent study demonstrated that metformin enriched the *Akkermansia* sp. population in high-fat-diet (HFD)-induced hyperglycemic and hyperlipidemic mice. Moreover, an inoculum of *Akkermansia* administered to diabetic mice exhibited a similar glucose-lowering efficacy to metformin, suggesting that alteration of gut microbiota is a new mechanism of metformin’s therapeutic effect in treatment of diabetes (17). However, our study showed that metformin significantly increased short-chain fatty acid (SCFA)-producing bacteria such as *Blautia* spp. and decreased *Akkermansia* spp., together with the amelioration of obesity in HFD-induced rats (18). As for in humans, when using metagenomic data from a multicountry T2DM metagenomic data set, univariate tests showed a significant increase in *Escherichia* spp. and a reduced abundance of *Intestinibacter* spp. in T2DM metformin-treated samples (19). However, this was the analysis of a collected microbial data set rather than a metformin-mediated gut microbiota shift analysis in a randomized clinical trial. Therefore, investigations with randomized clinical trials are needed to better understand metformin’s effect on gut bacteria in humans.

Traditional Chinese medicine (TCM) is a form of polypharmacy that has been developed and advocated for the treatment of many diseases for over 2,500 years in China. Our previous study suggested that gut microbiota might be involved in the effect of a standardized traditional herbal formula, GQD, which indicated that TCM can also modulate gut microbiota structure as a potential mechanism for improvement of diseases (20). An herbal formula consisting of eight herbs (AMC) was specifically developed for type 2 diabetic patients with hyperlipidemia. The herbs used in this formula, such as *Rhizoma Anemarrhenae* (21), *Momordica charantia* (22, 23), *Coptis chinensis* (24), *Aloe vera* (25, 26), and red yeast rice (27, 28), have profound effects in ameliorating blood glucose and lipid levels in various clinical trials and animal experiments. In addition, this herbal formula has been used in clinical application for more
than 10 years, which displayed satisfying efficacy. AMC contains the herb *Coptis chinensis*, whose major component, berberine, has been clinically demonstrated to be able to alleviate diabetes with hyperlipidemia, possibly via increasing SCFA-producing bacteria as shown in rats with HFD-induced obesity (29). We hypothesized that gut microbiota might also be involved in the efficacy of AMC.

To test the hypothesis that alteration of gut microbiota may be involved in the alleviation of T2DM with hyperlipidemia by metformin and AMC, 450 patients with T2DM and hyperlipidemia were randomly assigned to either a metformin- or AMC-treated group. After 12 weeks of treatment, 100 patients were randomly selected from each group and assessed for clinical improvements. The effects of the two drugs on the intestinal microbiota were evaluated by analyzing the V3 and V4 regions of the 16S rRNA gene by Illumina sequencing and multivariate statistical analysis.

**RESULTS**

**Alleviation of hyperglycemia and hyperlipidemia by metformin and AMC.** We analyzed the clinical data from 200 participants as shown in Table S2 in the supplemental material. The baseline variables showed no significant differences between the two groups. After 12 weeks of treatment, both metformin and AMC significantly improved fasting blood glucose (FBG), glycosylated hemoglobin (HbA1c), 2-h postprandial blood glucose (PGB), and homeostatic model assessment of β-cell function (HOMA-β). However, only AMC significantly improved homeostatic model assessment of insulin resistance (HOMA-IR), although no significant difference was obtained between AMC and metformin by analysis of covariance (ANCOVA). There were no significant differences between the two treatment groups in the alleviation of diabetes, except that insulin resistance was only significantly improved by AMC (Fig. 1a to e; Table S2). Besides, the AMC herbal formula significantly reduced total plasma cholesterol, triglyceride, and low-density lipoprotein cholesterol (LDL-c) levels, body weight, body mass index (BMI), and waist and hip circumferences of diabetic patients with hyperlipidemia (Fig. 1f to h; see Fig. S2a to d and Table S2 in the supplemental material). These parameters were also significantly ameliorated by metformin, except for triglyceride levels. However, metformin significantly increased diastolic blood pressure unexpectedly, which was significantly higher than that by AMC according to ANCOVA (Fig. S2e and f).

**Alteration of gut microbiota structure by metformin and AMC.** To analyze the structural changes of gut microbiota in T2DM patients treated with metformin and the AMC herbal formula, we performed Illumina sequencing of the V3 and V4 regions of the 16S rRNA gene using fecal samples collected at weeks 0 and 12 in the metformin and AMC treatment groups. Altogether, we got 6,216,877 high-quality sequences and 507 operational taxonomic units (OTUs) from 372 samples, with an average of 16,712 ± 3,663 reads. (One sample was excluded in later analysis because only 1,891 reads were obtained.) The diversity of the gut microbiota community showed that metformin significantly increased Simpson’s diversity index, while AMC significantly decreased rarefaction and Chao1 richness estimates, indicating that metformin significantly increased diversity, whereas AMC significantly reduced it (see Fig. S3 in the supplemental material). Principal-component analysis (PCA) and principal-coordinate analysis (PCoA) based on Bray-Curtis distance showed that both metformin and AMC significantly changed the gut microbiota structure of T2DM patients (Fig. 2). As for the PCoA results based on weighted and unweighted UniFrac distances and abundance-weighted and binary Jaccard distances, they displayed similar values, although the effect of metformin did not reach a significant level (see Fig. S4 in the supplemental material). Furthermore, comparing the changes of five distances before and after treatments using the Mann-Whitney U test, AMC exerted a significant larger modulating effect on the gut microbiota structure (see Fig. S5 in the supplemental material).

**CAGs associated with the improvements in glucose and lipid homeostasis.** To find the key phylotypes that correlated with the efficacy of metformin and the AMC herbal formula, coabundant network analysis was used in this study. The 214 core OTUs
which existed in >20% samples) were clustered into 24 coabundant groups (CAGs) by SparCC correlation and permutational multivariate analysis of variance (PERMANOVA). Twenty-one CAGs containing OTUs with \( R \) values of >0.4 were displayed in the coabundant network (Fig. 3a).

In the metformin treatment group, four CAGs were increased by metformin and three were significantly decreased (Fig. 3b; see Table S3 in the supplemental material). Among the four CAGs enriched by metformin, CAG21 showed significant inverse correlations with HbA1c, and CAG25 with FBG, 2-h PBG, and LDL-c (Fig. 4a). CAG21 consisted of 10 OTUs: two OTUs belong to *Clostridium* XIVa and one to each of the following genera: *Erysipelotrichaceae incertae sedis*, *Escherichia/Shigella*, *Fusobacterium*, *Flavonifractor*, *Lachnospiraceae*, *Lachnospiracea incertae sedis*, and *Clostridium* XVIII and IV. CAG25 contained four OTUs from *Blautia* and one from *Anaerostipes*. Among the three CAGs inhibited by metformin, CAG7 and CAG8 were significantly correlated with the alleviation of hyperglycemia (Fig. 4a). CAG7 and -8 showed significantly negative correlations with HOMA-IR. CAG7 contained three OTUs from *Bacteroides* and one from *Parabacteroides*; CAG8 consisted of three OTUs from *Bacteroides*, two OTUs from *Alistipes*, and one OTU each from *Oscillibacter* and un-*Ruminococcaceae*, respectively. To sum up, metformin significantly enriched CAG21 and CAG25 and inhibited CAG7 and CAG8, which were significantly correlated with the amelioration of hyperglycemia and hyperlipidemia.
After the AMC herbal formula treatment, six CAGs were increased and five were decreased (Fig. 3b; Table S3). Among the six CAGs enriched by AMC herbal formula, CAG5, CAG13, CAG24, and CAG25 showed significant positive correlations with the improvement of glucose and lipid homeostasis (Fig. 4b). CAG5 was inversely correlated with HOMA-IR, which contained two OTUs from *Megamonas*. CAG13 has the highest relative abundance, with OTU2 from *Faecalibacterium* accounting for over 50%. It showed significant negative correlations with HbA1c, FBG, 2-h PBG, HOMA-IR, and triglyceride levels, which consisted of two OTUs from *Roseburia*, and one OTU each from *Faecalibacterium*, *Gemmiger*, *Coprococcus*, and un-identified Lachnospiraceae, respectively. CAG24 also showed inverse correlations with FBG, 2-h PBG, HOMA-IR, and triglyceride levels, which contained two OTUs from un-identified Lachnospiraceae and one OTU from *Ruminococcus* 2 and *Lachnospiraceae incertae sedis*, respectively. CAG25 was inversely correlated with HOMA-IR and triglyceride levels, and it contained four OTUs from *Blautia* and one from *Anaerostipes*, as described above. Taken together, AMC herbal formula significantly enriched CAG5, CAG13, CAG24, and CAG25, which showed significant correlations with the improvements of hyperglycemia and hyperlipidemia.

We compared the key CAGs that responded to metformin and the AMC herbal formula and tried to find the common CAGs shared by the two drugs. These CAGs might be related to the alleviation of diabetes rather than simply responding to the drug treatment. Among the eight CAGs that were associated with the efficacy of metformin or the AMC herbal formula, CAG25, represented by the OTU from *Blautia*, was identified in both drug treatment groups.

**DISCUSSION**

To our knowledge, this is the first randomized clinical trial evaluating a traditional Chinese herbal formula’s efficacy in treating T2DM with hyperlipidemia and obesity using metformin as a positive control. In the 12 weeks of clinical trial, both drugs significantly ameliorated hyperglycemia and hyperlipidemia, but only AMC significantly improved triglyceride levels. This is consistent with a meta-analysis of randomized controlled clinical trials in which metformin had no intrinsic effect on triglyceride levels.
FIG 3 Alterations of coabundant groups by metformin and AMC herbal formula treatments. (a) Coabundant network analysis. Each circle represents one OTU. Circles with the same color are from the same coabundant group (CAG). Circle size represents the average abundance of each (Continued on next page)
in T2DM patients (30). To sum up, AMC displayed a slightly better effect in the alleviation of hyperglycemia and hyperlipidemia as only AMC significantly improved diabetic patients’ HOMA-IR and plasma triglyceride levels.

In this clinical trial, an alteration of gut microbial structure by metformin in humans was observed. Metformin treatment significantly inhibited two functional groups, namely, CAG7 and CAG8, which contained potential pathogen-like genera, including Alistipes, Oscillibacter, and Bacteroides. In addition, the reductions in the two CAGs significantly correlated with the improvement of glucose homeostasis by metformin. Members of Alistipes were positively associated with the colonic tumorigenesis in gnotobiotic mice into which were transplanted the gut microbiota from colorectal cancer patients (31). Furthermore, elevation of several OTUs from Alistipes was correlated with a greater frequency of pain in pediatric patients with irritable bowel syndrome (32). As for Oscillibacter, transferring the gut microbiota of obesity-prone rats harboring specific species from Oscillibacter and Clostridium clusters IV to germfree mice replicated the obese phenotypes and associated disordered signaling pathways.

**FIG 3** Legend (Continued)

OTU in each group. (b) Relative abundance of altered CAGs by metformin and/or the AMC herbal formula. The relative abundances of CAGs are expressed as means ± standard deviations (SD). ***, P < 0.001, **, P < 0.01, and *, P < 0.05, by Wilcoxon’s signed-rank test.
(33). These results suggest that inhibition of potential pathogen-like bacteria might be involved in the glucose-lowering effect of metformin.

The present study observed the increase in *Blautia* spp., a population of SCFA producers in the metformin-treated group, in agreement with our previous animal study (18). Interestingly, we observed a decrease in *Akkermansia* in T2DM patients after metformin treatment. This finding confirmed our results in Wistar rats with HFD-induced insulin resistance and obesity treated with metformin (18), but it was contrary to the previous study that metformin specifically increased *Akkermansia* spp. and thus improved glucose homeostasis in mice with HFD-induced obesity (17). This might be attributed to the differences in strain-specific functions. To further dissect the gut microbiota-mediated mechanism of metformin in the later study, we first tried to identify the key population that responds to metformin in a randomized clinical trial. AMC increased functional groups CAG5 and CAG13, which were significantly correlated with the improvements in glucose and lipid homeostasis. They contained OTUs from *Faecalibacterium, Roseburia, Gemmiger, Coprococcus,* and *Megamonas. Faecalibacterium* and *Roseburia* are two genera containing butyrate-producing bacteria (34, 35). *Faecalibacterium prausnitzii* is an anti-inflammatory (36) and functionally important bacterium (37). Several recent studies showed that *F. prausnitzii* and its supernatant prevented the physiological damage to the intestine induced by 2,4,6-trinitrobenzenesulfonic acid (TNBS) (36, 38) or dextran sodium sulfate (DSS) (39) in mice, possibly through the production of butyrate, salicylic acids (40), or an anti-inflammatory molecule (41). The gut metagenome analysis showed that T2DM patients had lower levels of butyrate-producing bacteria, such as *Roseburia* and *F. prausnitzii*, in the cohorts of Chinese diabetics and European diabetic women (42–44). It has been reported that a higher abundance of *F. prausnitzii* was linked with the reduction of a low-grade inflammatory state during the alleviation of obesity and diabetes in patients receiving Roux-en-Y gastric bypass (RYGB) surgery (45). Moreover, according to our previous study, an increase in *F. prausnitzii* was associated with the alleviation of diabetes by another Chinese herbal formula, GQD, in a double-blinded, placebo-controlled clinical trial (20). *Faecalibacterium* was also recovered after the treatment of AMC in this study, indicating that *Faecalibacterium* spp. might be important in the efficacy of the TCM formula in the improvements in diabetes and obesity.

*Roseburia, Gemmiger,* and *Coprococcus* enriched by the AMC herbal formula in T2DM patients are also reported to have beneficial influences. Besides, restoration of *Roseburia* spp. by chitin-glucan fiber was related with alleviations of obesity, diabetes, and hepatic steatosis in mice fed with HFD (46). These results showed that enrichment of SCFA-producing bacteria might be associated with the amelioration of hyperglycemia and hyperlipidemia by AMC.

Notably, CAG25 dominated by OTUs from *Blautia* was significantly increased by both drugs and correlated with the improvements in glucose and lipid homeostasis. *Blautia* is a group of bacteria containing various acetate and butyrate producers (47, 48). More importantly, *Blautia* spp. were also reported to be enriched by metformin during the amelioration of obesity and insulin resistance in rats (18). It has been reported that 16 children with type 1 diabetes possessed a significantly lower abundance of *Blautia coccoides* with respect to 16 healthy children by PCR-denaturing gradient gel electrophoresis (DGGE) and real-time quantitative PCR (qPCR) (49). Moreover, according to the gut microbial structure analysis of 64 patients receiving blood/marrow transplantation, a larger amount of anti-inflammatory *Blautia* spp. was associated with a reduction in mortality caused by acute graft-versus-host disease after the surgery. This result was further confirmed in another independent cohort with 51 patients enrolled from the same institution (50). These results suggest that *Blautia* spp. might be a common target for the management of diabetes with hyperlipidemia.

Compared with metformin, AMC exerted a much stronger modulatory effect on the gut microbiota together with better efficacy in improving HOMA-IR and triglyceride levels. This might be attributed to the effects of multiple chemical components in the
AMC, such as mangiferin and berberine. In a randomized, double-blind clinical trial, mangiferin treatment significantly improved serum levels of triglycerides, free fatty acids, and HOMA-IR in overweight patients with hyperlipidemia (S1), but the pharmacological mechanism of mangiferin has not been elucidated yet. Another study showed that gut microbiota might play an important role in it despite its poor absorption in the gastrointestinal tract. Bacteroides sp. strain MANG might be responsible for the pharmacological effects of mangiferin via converting it into its aglycone and norathyriol (S2). Berberine was reported to significantly improve hyperglycemia and hyperlipidemia in a multicenter, randomized, double-blind, and placebo-controlled clinical trial (24). In addition, berberine prevented and alleviated HFD-induced obesity and insulin resistance in rats through enrichment of SCFA producers such as Blautia spp. (18, 29). These findings implied that mangiferin and berberine might be the major active ingredients in AMC that shifted the gut microbiota.

Conclusion. Taken together, these data suggest that metformin and the Chinese herbal formula may ameliorate type 2 diabetes with hyperlipidemia by enriching beneficial bacteria, such as Blautia and Faecalibacterium spp.

MATERIALS AND METHODS

Study design. This study was a multicenter, randomized, positive-control, and open label clinical trial, including a 4-week washout period and a 12-week treatment period. It was performed according to the Declaration of Helsinki (2008) and approved by the Ethics Committee of Guang’anmen Hospital of the China Academy of Chinese Medical Sciences (approval no. eighty-sixth 2011). All participants signed informed consent forms before enrolling in the study (Clinicaltrials.gov registration no. NCT01471275). The inclusion and exclusion criteria are stated in Text S1 in the supplemental material.

In the initial screening, a 75-g oral glucose tolerance test (OGTT) was used for diagnosis of diabetes, and plasma triglyceride levels, waist circumference, and hip circumference were used for diagnosis of hyperlipidemia. After a 4-week washout period, 450 patients with T2DM and hyperlipidemia were enrolled in the clinical trial. They were randomized into two treatment groups, receiving either metformin or AMC. Randomization was performed centrally and stratified in the proportion of 1:1 for the two treatment groups through the Interactive Web Response System by a third party not involved in data management and statistical analysis. After a 12-week treatment, 100 patients were randomly chosen from each treatment group. The random assignment was generated by the computer program SAS 9.3, which was also conducted by independent statisticians who did not participate in data acquisition and evaluation. All of the recruited patients followed advice on diabetic and low-fat diets and exercises for diabetic patients as instructed by the Chinese Diabetes Society.

Drug administration. The TCM formula in our study was the AMC herbal formula composed of 8 herbs, namely, Rhizoma Anemarrhenae, Momordica charantia, Coptis chinensis, Salvia miltiorrhiza, red yeast rice, Aloe vera, Schisandra chinensis, and dried ginger. Herbs were all provided by Jiangyin Tianjiang Pharmaceutical Co., Ltd., and quality controlled. All the herbs were processed into boil-free granules (see Table S1 in the supplemental material). The inclusion and exclusion criteria are stated in Text S1 in the supplemental material. Table S1 in the supplemental material lists the amount of granular extracts of each herb in 1 unit of AMC formula the patients had daily. Patients mixed the eight granular extracts and took them like coffee twice daily.

Patients in the positive-control group took metformin hydrochloride tablets, which were all provided by Beijing Liling Hengtai Pharmaceutical Co., Ltd., and quality controlled (approval no. H11021560). They took metformin tablets 0.25 g/time and 3 times/day orally after meals.

Study evaluation and outcomes. The primary efficacy outcomes were glycosylated hemoglobin (HbA1c), triglyceride (TG), weight, and waist circumference. Secondary efficacy outcomes were fasting blood glucose (FBG), 2-h postprandial blood glucose (2-h PBG), serum insulin, total cholesterol, high-density lipoprotein cholesterol (HDL-c), and low-density lipoprotein cholesterol (LDL-c). The adverse events and the occurrence of hypoglycemia were recorded. FBG, weight, and waist circumference were detected every 4 weeks; other assessments were performed before and after the treatment. Fecal samples were collected by using InviMag stool collection tubes with DNA stabilizer (Invitex, Germany) for gut microbiota analysis before and after the treatment.

DNA extraction and V3 and V4 regions of 16S rRNA gene sequencing. Total genomic DNA of fecal samples was extracted by the InviMag stool DNA kit (Invitex, Germany) as previously described (20). The sequencing library of the V3 and V4 regions of the 16S rRNA gene was prepared following the protocol from Illumina, Inc., with minor modification (S3). Before the sequencing, 40% PhiX (PhiX control kit v3; Illumina, Inc.) was added to the pooled library to serve as an internal control. The pooled library was sequenced on the Illumina MiSeq System using paired-end 300-bp reads and the MiSeq v3 reagent kit.

Bioinformatics analysis of sequencing data. High-quality sequencing data extraction was performed as follows (S4). (i) Both ends of the forward and reverse reads were truncated at the base, where the Phred quality score was no more than 20. (ii) If the forward and reverse reads had a minimum 50-bp-length overlap, they could be merged into a complete read. (iii) These reads were not kept unless they were longer than 399 bp and the expected errors were less than 0.5. Operational taxonomic units
(OTUs) were delineated at the cutoff of 97% using the USEARCH v7.0.1090 pipeline. The protocol can be found on the website http://drive5.com/usearch/manual/uparse_pipeline.html. The detailed procedures were stated in our previous publication (53). α- and β-diversity analyses were performed using QIME v1.8.0 (55). To normalize read depth, all samples were randomly subsampled to 5,000 reads/sample. Shannon’s index, Simpson’s index, rarefaction, and Chao1 estimates were evaluated. A normalized and log2-transformed OTU abundance table was used for the β-diversity analysis, including principal-component analysis (PCA) and principal-coordinate analysis (PCoA) based on Bray-Curtis, UniFrac, and Jaccard distances. The taxonomy of each OTU was assigned online by RDP Classifier (RDP Naive Bayesian rRNA Classifier version 2.10) with a bootstrap cutoff of 80%.

Coabundant network analysis. In ecologic systems, species with similar requirements and functions would constitute an ecologic guild to adapt to the environments. Species from the same guild were shown to be coabundant. In this study, coabundant network analysis was used to find the key phylotypes associated with the improvements in glucose and lipid homeostasis. The correlations between the core OTUs (which existed in more than 20% samples) were calculated with the SparCC algorithm (bootstrap value, 100) (56). The 214 core OTUs were clustered into 24 coabundant groups (CAGs) by using the ward algorithm and PERMANOVA (999 permutations; P < 0.001) based on the SparCC correlation coefficient matrix with R version 3.1.3. The 21 CAGs with an R coefficient higher than 0.4 were visualized into a network with cytoscape v3.1.1 (http://www.cytoscape.org/). The abundance of each CAG was the sum of relative abundances of the OTUs in this CAG.

Statistical analysis. With regard to the demographic and clinical parameter, the two-tailed t test was used for paired analysis (before and after intervention). ANCOVA was used to compare treatment groups with baseline values as covariates. Missing data were handled using the last observation carried forward method. As for the gut microbiota data, Wilcoxon’s signed-rank test was used for comparison of paired samples, and the Mann-Whitney U test was used for independent samples.

Accession number(s). The sequence data have been uploaded to the Sequence Read Archive database under accession no. SRP126564.

SUPPLEMENTAL MATERIAL
Supplemental material for this article may be found at https://doi.org/10.1128/mBio.02392-17.

TEXT S1, DOCX file, 0.1 MB.
FIG S1, TIF file, 1.5 MB.
FIG S2, TIF file, 0.4 MB.
FIG S3, TIF file, 0.6 MB.
FIG S4, TIF file, 0.2 MB.
FIG S5, TIF file, 0.6 MB.
TABLE S1, DOCX file, 0.1 MB.
TABLE S2, DOCX file, 0.1 MB.
TABLE S3, DOCX file, 0.1 MB.

ACKNOWLEDGMENTS
This work was supported by the National Natural Science Foundation of China (no. 81273720, 31330005, and 81401141) and the Scientific Specialized Program of Traditional Chinese Medicine of China (no. 201007004).

F.L., X.T., and L.Z. designed the study. J.X. and L.Z. drafted and revised the manuscript. None of the authors reports a conflict of interest.

REFERENCES
1. Aguirre F, Brown A, Cho NH, Dahlgquist G, Dodd S, Dunning T, Hirst M, Hwang C, Magliano D, Patterson C, Scott C, Shaw J, Soltesz G, Usher-Smith J, Whiting D. 2013. IDF diabetes atlas, 6th ed. International Diabetes Federation, Basel, Switzerland.
2. Kahn SE, Cooper ME, Del Prato S. 2014. Pathophysiology and treatment of type 2 diabetes: perspectives on the past, present, and future. Lancet 383:1068–1083. https://doi.org/10.1016/S0140-6736(13)62154-6.
3. Donath MY, Shoelson SE. 2011. Type 2 diabetes as an inflammatory disease. Nat Rev Immunol 11:98–107. https://doi.org/10.1038/nri2925.
4. Hotamisligil GS. 2008. Inflammation and endoplasmic reticulum stress in obesity and diabetes. Int J Obes 32(Suppl 7):S52–S54. https://doi.org/10.1038/ijo.2008.238.
5. Shoelson SE, Goldfine AB. 2009. Getting away from glucose: fanning the flames of obesity-induced inflammation. Nat Med 15:373–374. https://doi.org/10.1038/nm0409-373.
6. Zhao L. 2013. The gut microbiota and obesity: from correlation to causality. Nat Rev Microbiol 11:639–647. https://doi.org/10.1038/nrmicro3089.
7. Turnbaugh PJ, Ley RE, Mahowald MA, Magrini V, Mardis ER, Gordon JI. 2006. An obesity-associated gut microbiome with increased capacity for energy harvest. Nature 444:1027–1031. https://doi.org/10.1038/nature05414.
8. Vijay-Kumar M, Aitken JD, Carvalho FA, Cullender TC, Mwangi S, Srivastava S, Sitaraman SV, Knight R, Ley RE, Gewirtz AT. 2010. Metabolic syndrome and altered gut microbiota in mice lacking Toll-like receptor 5. Science 328:228–231. https://doi.org/10.1126/science.1179721.
Microbiota Effects on Hyperglycemia and Hyperlipidemia

9. Fei N, Zhao L. 2013. An opportunistic pathogen isolated from the gut of an obese human causes obesity in germfree mice. ISME J 7:880–884. https://doi.org/10.1038/ismej.2013.151.

10. Zhang C, Yin A, Li H, Wang R, Wu G, Shen J, Zhang M, Wang L, Hou Y, Ouyang H, Zhang Y, Zheng Y, Wang J, Li X, Wang Y, Zhang F, Zeng B, Li W, Wen F, Zhao Y, Pang X, Zhang X, Fu H, Chen F, Zhao N, Hamaker BR, Bridgewater LC, Weinkove D, Clement K, Dore J, Holmes E, Xiao H, Zhao G, Yang S, Bork P, Nicholson JK, Wei H, Tang H, Zhang X, Zhao L. 2015. Dietary modulation of gut microbiota contributes to alleviation of both genetic and simple obesity in children. EBioMedicine 2:968–984. https://doi.org/10.1016/j.ebiom.2015.07.007.

11. Pruski M, Krysiak R, Okopien B. 2009. Pleiotropic action of short-term metformin and fenofibrate treatment, combined with lifestyle intervention, in type 2 diabetic patients with mixed dyslipidemia. Diabetes Care 32:1421–1424. https://doi.org/10.2337/dc09-1132.

12. Luong DO, Oster R, Ashraf AP. 2015. Metformin treatment improves weight and dyslipidemia in children with metabolic syndrome. J Pediatr Endocrinol Metabol 28:649–655. https://doi.org/10.1515/jped-2014-0201.

13. Cittadini A, Napoli R, Monti MG, Rea D, Longobardi S, Netti PA, Walser M, Samà A, Almargenti G, Isagardi J, Saccà L. 2012. Metformin prevents the development of chronic heart failure in the SHRf rat model. Diabetes 61:944–953. https://doi.org/10.2337/db11-1132.

14. He X, Sabet D, Miller R, Sun X, Hussain MA, Radovick S, Wondisford FE. 2009. Metformin and insulin suppress hepatic gluconeogenesis through phosphorylation of CREB binding protein. Cell 137:635–646. https://doi.org/10.1016/j.cell.2009.03.016.

15. Zhou G, Myers R, Li Y, Chen Y, Shen X, Fenyk-Melody J, Wu M, Ventre J, Pruski M, Krysiak R, Okopien B. 2009. Pleiotropic action of short-term metformin and fenofibrate treatment improves glucose homeostasis in diet-induced obese mice. Sci Rep 5:14405. https://doi.org/10.1038/srep014405.

16. Bonora E, Cigolini M, Bosello O, Zancanaro C, Capretti L, Zavaroni I, Xue Z, Pang X, Zhao L, Tong X. 2015. Structural modulation of gut microbiota during alleviation of type 2 diabetes with metformin treatment. Gut 63:727–735. https://doi.org/10.1136/gutjnl-2013-308339.

17. Han J, Yang N, Zhang F, Zhang C, Liang F, Xie W, Chen W. 2015. Role of AMP-activated protein kinase in mechanism of metformin action. J Clin Invest 125:1170–1174. https://doi.org/10.1172/JCI72079.

18. Bonora E, Cigolini M, Bosello O, Zancanaro C, Capretti L, Zavaroni I, Xue Z, Pang X, Zhao L, Tong X. 2015. Structural modulation of gut microbiota during alleviation of type 2 diabetes with metformin treatment. Gut 63:727–735. https://doi.org/10.1136/gutjnl-2013-308339.

19. Ren J, Li W, Yan F, Zhao Y, Peng X, Zhang X, Wu H, Zhao N, Li W, Yan F, Zhao Y, Pang X, Zhang X, Wu H, Zhao N, Hamaker BR, Bridgewater LC, Weinkove D, Clement K, Dore J, Holmes E, Xiao H, Zhao G, Yang S, Bork P, Nicholson JK, Wei H, Tang H, Zhang X, Zhao L. 2015. Dietary modulation of gut microbiota contributes to alleviation of both genetic and simple obesity in children. EBioMedicine 2:968–984. https://doi.org/10.1016/j.ebiom.2015.07.007.

20. Pruski M, Krysiak R, Okopien B. 2009. Pleiotropic action of short-term metformin and fenofibrate treatment, combined with lifestyle intervention, in type 2 diabetic patients with mixed dyslipidemia. Diabetes Care 32:1421–1424. https://doi.org/10.2337/dc09-1132.

21. Han J, Yang N, Zhang F, Zhang C, Liang F, Xie W, Chen W. 2015. Role of AMP-activated protein kinase in mechanism of metformin action. J Clin Invest 125:1170–1174. https://doi.org/10.1172/JCI72079.
