GTF2I Mutation in Thymomas: Independence From Racial-Ethnic Backgrounds. An Indian/German Comparative Study

Deepali Jain1*, Prerna Guleria1, Varsha Singh1, Rajinder Parshad2, Sunil Kumar3, Timo Gaiser4, Katrin S. Kurz4,6, German Ott4,6, Stefan Porubsky4,7, Gerhard Preissler8, Christian G. Sauer4, Sebastian Schölch9,10, Philipp Ströbel11, Thomas Hielscher12, Alexander Marx4 and Zoran V. Popovic4*

1Department of Pathology, All India Institute of Medical Sciences, New Delhi, India, 2Department of Surgery, All India Institute of Medical Sciences, New Delhi, India, 3Surgical Oncology, All India Institute of Medical Sciences, New Delhi, India, 4Institute of Pathology, University Medical Centre Mannheim, University of Heidelberg, Mannheim, Germany, 5Dr. Margarete Fischer-Bosch Institute for Clinical Pharmacology, Stuttgart, Germany, 6Department of Clinical Pathology, Robert-Bosch-Krankenhaus, Stuttgart, Germany, 7Institute of Pathology, University Hospital, Johannes-Gutenberg University Mainz, Mainz, Germany, 8Department of Thoracic Surgery, Schillerhöhe Clinics, Robert-Bosch-Krankenhaus, Gerlingen, Germany, 9Department of Surgery, University Medical Centre Mannheim, University of Heidelberg, Mannheim, Germany, 10Junior Clinical Cooperation Unit Translational Surgical Oncology, German Cancer Research Center (DKFZ), Heidelberg, Germany, 11Institute of Pathology, University Medical Center Göttingen, University of Göttingen, Göttingen, Germany, 12Department of Biostatistics, German Cancer Research Center (DKFZ), Heidelberg, Germany

Thymomas are the most frequent adult mediastinal cancers. Their etiology is unknown and their pathogenesis poorly understood. Racial, ethnic and environmental factors influence tumorigenesis in many cancers, but their role in thymomas remains unclear to date. In this study that included pretreatment thymoma cases from India and Germany (n = 37 and n = 77, respectively) we compared i) the prevalence of the thymoma-specific chromosome 7 c.74146970T > A mutation of the GTF2I gene in type A and AB thymomas; ii) epidemiological features; and iii) the frequency of myasthenia gravis (MG). Due to a known predominance of GTF2I mutation in A and AB histotypes, we included only a marginal number of type B thymomas as a control group in both cohorts. While the distribution of histological types between the cohorts was similar (p = 0.1622), Indian patients were strikingly younger (p < 0.0001; median age 50 vs. 65 years) and showed significantly lower tumour stage (Masaoka-Koga stage I) at primary diagnosis (p = 0.0005) than the German patients. In patients with known MG status (n = 17 in Indian and n = 25 in German cohort), a clear trend towards more frequent MG was observed in the Indian group (p = 0.0504; 48 vs. 82%). The prevalence of the GTF2I mutation (analysed in n = 34 Indian and n = 77 German patients) was identical in the two cohorts. We conclude that racial-ethnic and environmental factors do not significantly influence the most common molecular feature of thymomas but may have an impact on the timing of clinical presentation.

Keywords: epidemiology, thymoma, GTF2I mutation, myasthenia gravis, racial-ethnic factors
**BACKGROUND**

Thymic epithelial tumours (TETs) are the most common tumours arising in the anterior mediastinum in adults, comprising mainly thymomas and the rarer and more aggressive thymic carcinomas (1). The focus here is on thymomas that are subdivided into type A, AB, B1, B2, and B3 and rare other subtypes according to histological criteria of the WHO classification. Studies on the genomic landscape of TETs are limited. Nevertheless, the few studies conducted have demonstrated a genetic variability between the various histologic types thereby indicating that the subtypes are not only morphologically but also molecularly distinct (2–4). So much so that some authors have even identified molecular subtypes of TETs and have associated them with clinical phenotypes and outcomes (5, 6). Among various other genomic alterations, the most common one is the point mutation in chromosome 7 c.7414G>T > A of the general transcription factor II-I (GTF2I) gene that has been identified to be specific to thymic epithelial tumours, predominantly in type A and AB thymomas and associated with a more favourable prognosis (7, 8). Its prevalence in type B1-B3 thymomas and thymic carcinomas is much lower, with decreasing frequency as the histology progresses to a more aggressive subtype (7). Since the mutation has not been widely studied with respect to different racial and ethnic backgrounds, we aimed to identify its prevalence in two populations from different regions of the world and identify any geographical variability if present in the molecular profile of thymomas. With this objective in mind, we carried out a retrospective analysis, to assess the frequency of GTF2I mutation in Indian and German thymomas (mainly type A and AB) and to correlate it with clinical features.

**MATERIALS AND METHODS**

The study was carried out as collaboration between the departments of pathology at the University Medical Centre Mannheim, Heidelberg University, Mannheim, Germany and the All India Institute of Medical Sciences, New Delhi. A cohort of Indian (n = 37) and German (n = 77) thymomas enriched for A and AB histotypes with a primary goal to address the GTF2I mutation frequency was retrieved from the departmental archives. Pretreatment surgical resections were performed in the period 2008–2018 and included no recurrent tumor or metastatic tissue. The clinical details of these cases were obtained from case files. Hematoxylin-eosin stained slides were reviewed by two experts in the field (AM and DJ) in terms of histotype (1) and Masaoka-Koga stage (9) that allows for a more detailed description of early invasion than the TNM system (see Discussion). Primers for Exon 15 of the GTF2I gene were designed using National Center for Biotechnology Information (NCBI) and Primer 3 (v.0.4.0) software.

**DNA Isolation and Quantification**

Genomic DNA was isolated from formalin-fixed, paraffin embedded (FFPE) samples using 50-µm-thick sections of the tumour according to the manufacturer’s protocol using ReliaPrep gDNA Tissue Miniprep system (cat. No. A2050, Promega, United States). Only paraffin blocks where cancer occupied >50% of the section were selected for DNA extraction. The isolated DNA was subjected to qualitative and quantitative assessment via spectrophotometry-based absorbance studies using NanoDrop, Biodrop Resolution, Cambridge, United Kingdom. Polymerase Chain Reaction (PCR) of GTF gene (exon 15) was carried out in a 25 µL total volume, containing 80 ng genomic DNA, 10 µM respective primers and using GoTaq colorless master mix (cat. No. M7142, Promega, United States).

**Polymerase Chain Reaction and Sanger Sequencing**

The PCR program was set as: HotStarsTaq activation at 95°C for 5 min, followed by 40 cycles, each having denaturation at 95°C for 30 s, annealing at 55°C for 1 min and extension at 72°C for 45 s, and the final extension was 72°C for 10 min. The quality of the PCR product was assessed using electrophoresis by resolving a part of the PCR amplified product (3 µl) on 1.5% agarose gel for all considered samples. Clear band of expected size was observed in all samples which corresponded to the size of the desired amplicons. The PCR product was then cleaned up manually using polyethylene glycol (PEG) purification methods. The PEG purified PCR product was used for sequencing PCR with a total reaction volume of 10 µl. The sequencing PCR was set as: initial denaturation at 95°C, 5 min, followed by 35 cycles, each having denaturation at 95°C for 10 s, annealing at 55°C for 5 s and extension at 60°C for 4 min. The sequencing PCR product was cleaned up using 125 µM EDTA, 100% ethanol, and 3 M sodium citrate. Post-clean up, DNA was reconstituted in HiDi and incubated at 95°C for 5 min for denaturation and immediately put on ice to cool the PCR product. After this snap chilling, the samples were proceeded for sequencing. The sequencing was done using both forward and reverse primers (GTF2I_F: 5’-ATC CGGTACCCCTTTTTTC-3’, GTF2I_R: 5’-AGACAAAGGTTCC AACAGG-3’) for greater accuracy and the results were analysed using SeqMan II software (DNASTAR).

**Immunohistochemistry**

For immunohistochemical staining of representative thymoma cases, paraffin embedded tissue was cut to 1 µm sections. The staining was performed using an automated slide preparation system (Benchmark Ultra, Roche Tissue Diagnostics, Ventana, Tucson, AZ, United States), a Ventana OptiView DAB IHC Detection Kit (Ventana, Tucson, AZ, United States) and commercially available monoclonal antibodies for CD20 (Clone L26, DCS, Hamburg, Germany), TdT (Clone SEN28, Leica Biosystems, Buffalo Grove, IL, USA) and CK19 (Clone RCK108, Agilent, Santa Clara, CA. United States).

**Statistical Analyses**

Primary endpoint of this study was the GTF2I mutational status. Subgroup analyses and analysis of MG status were exploratory. Associations between clinico-pathological and molecular factors and geographical origin were assessed with Fisher’s exact test for categorical factors and with Wilcoxon test for age. Age-adjusted
effects of clinical parameters were tested with the likelihood-ratio test of logistic regression models for origin between a model with predictors clinical parameter and age and a model with predictor age only. Heterogeneity of association between origin and mutational status across subgroups was analysed with the likelihood-ratio test between a model with origin and subgroup factor as predictors and a model including interaction term of both predictors. 

\[ p \] -values below 0.05 were considered significant. Statistical software R 4.0 was used to carry out analyses.

**RESULTS**

**Cohort Data and Histological Distribution**

A total of 138 resection specimens were retrieved of which 37 were Indian cases and 101 were German. Among these, 114 cases were thymomas consisting of 23 (20.2%) type A thymomas, 52 (45.6%) type AB, 17 atypical A/AB (14.9%) and 22 (19.3%) type B1-B3 thymomas. German thymoma cases \((n = 77)\) included 33 female (42.9%) and 44 male (57.1%) patients in the age range 31–85 years (median age 65 years). Masaoka-Koga stage I was reported in 16/77 cases (20.8%), stage II in 38 cases (49.4%), stage III in 9 cases (11.7%) and stage IV in 14 cases (18.2%). The German cohort included 15 (19.5%) type A thymomas, 31 (40.3%) type AB, 15 atypical A/AB (19.5%) and 16 (20.8%) type B1-B3 thymomas. The remaining 24 German cases comprised 18 thymic carcinomas, 2 cases of sclerosing mediastinitis, 2 metaplastic thymomas, 1 thymolipoma and 1 thymic hyperplasia with lymphoepithelial sialadenitis (LESA)-like features \((10)\).

All the 37 Indian cases included in this study were thymomas [8 type A (21.6%), 21 type AB (56.8%), 2 atypical A/AB (5.4%) and 6 type B1-B3 thymomas (16.2%)]. Among the 37 Indian cases, there was an equal gender distribution (19 cases or 51.4% were females). The age range in the Indian group was 20–73 years with a median age of 50 years; hence, the Indian cohort was significantly younger than the German counterpart \((p < 0.0001, Table 1; Figure 1)\). Notably, even after excluding type B thymomas, this striking age difference remained in separate analyses of histotypes A and AB \((p = 0.00097\) and \(p = 0.0068\), respectively; Supplementary Table S1). In both cohorts there were no significant differences between the age distribution of patients with and without MG. Masaoka-Koga stage distribution was as follows: 24 (64.9%) stage I, 9 (24.3%) stage II and 4 cases (10.8%) were in stage III.

| TABLE 1 | Contingency table of clinico-pathological factors between origins (for gender, MG status, Masaoka-Koga stage, GTF2I mutation status, histological type and age). |
|----------|----------------------------------|-----------------|-----------------|-----------------|
| Level    | German                          | Indian          | \(p\)            | Age-adjusted \(p\) |
| N        | —                               | 77             | 37              | —               |
| Gender (%) | Female                        | 33 (42.9)        | 19 (51.4)        | 0.4270          | 0.20          |
|          | Male                           | 44 (57.1)        | 18 (48.6)        | —               | —             |
| MG Status (%) \(a\) | No                            | 13 (22.0)        | 3 (7.6)          | 0.0504          | 0.13          |
|          | MG                             | 12 (48.0)        | 14 (52.4)        | —               | —             |
| Masaoka-Koga stage (%) | I                             | 16 (20.8)        | 24 (64.9)        | <0.0001         | 0.0003        |
|          | II                             | 38 (49.4)        | 9 (24.3)         | —               | —             |
|          | III/IV                         | 23 (29.9)        | 4 (10.8)         | —               | —             |
| Mutation status (%) \(b\) | WT                            | 28 (36.4)        | 12 (33.3)        | 1.00            | 0.26          |
|          | L404H                          | 49 (63.6)        | 22 (66.7)        | —               | —             |
| Histological type (%) | A                             | 15 (19.5)        | 8 (21.6)         | 0.1622          | 0.048         |
|          | AB                             | 31 (40.3)        | 21 (56.8)        | —               | —             |
|          | atypical A/AB                  | 15 (19.5)        | 2 (5.4)          | —               | —             |
|          | B                              | 16 (20.8)        | 6 (16.2)         | —               | —             |
| Age [median (IQR)] | —                              | 65.00 (53.00, 72.00) | 50.00 (37.00, 55.00) | <0.0001         | —             |

\(MG\)—myasthenia gravis; IQR—interquartile Range.

\(a\)MG status was known in 25 German and 17 Indian cases.

\(b\)GTF2I Mutation was analysed in 77 German and 34 Indian cases.
Within defined WHO thymoma histotypes, we could not observe morphologic variations that could be assigned as specific to the Indian or German cohort, as confirmed by two experts in the field (AM and DJ). Representative histological and immunohistological features of thymoma cases included in this study are illustrated in Figure 2.

Also, no significant difference regarding the distribution of thymoma histotypes was evident \( (p = 0.16; \text{Table } 1) \). In age-adjusted analysis, there was a difference \( (p = 0.048) \), because age and histotype are strongly associated. On the other hand, we could detect a highly significant difference with respect to disease stage at clinical presentation, showing a clear majority of Indian patients at Masaoka-Koga Stage I (64.9% of all Indian thymoma cases) in comparison to only 20.8% of German patients at this early stage \( (p < 0.0001; \text{Table } 1) \). In our histotype A- and AB-dominated cohorts, this difference was most prominent for type AB \( (p = 0.006) \) followed by type A thymomas \( (p = 0.03; \text{Tables } 2 \text{ and } 3) \).

### Association With Myasthenia Gravis
In the German cohort, follow up information on myasthenia gravis status was available in 25/77 thymomas (of which 19 were

### Table 2
Contingency table of clinico-pathological factors between origins (for histological type A).

| Level                  | German | Indian | \( p \)  |
|------------------------|--------|--------|--------|
| N                      | 15     | 8      |        |
| Gender (%)             |        |        |        |
| Female                 | 6 (40.0) | 6 (75.0) | 0.193 |
| Male                   | 9 (60.0) | 2 (25.0) |        |
| MG Status (%)          |        |        |        |
| No                     | 2 (100.0) | 1 (14.3) | 0.083 |
| MG                     | 0 (0.0) | 6 (85.7) |        |
| Masaoka-Koga stage (%) |        |        |        |
| I                      | 3 (20.0) | 6 (75.0) | 0.032 |
| II                     | 6 (40.0) | 2 (25.0) |        |
| III/IV                 | 6 (40.0) | 0 (0.0) |        |
| Mutation status (%)    |        |        |        |
| WT                     | 6 (40.0) | 1 (12.5) | 0.345 |
| L404H                  | 9 (60.0) | 7 (87.5) |        |
| Age [median (IQR)]     | 70.00 (66.00, 73.50) | 50.00 (41.50, 56.75) | 0.001 |

MG—myasthenia gravis; IQR—Interquartile Range.
type AB, 2 type A and 4 atypical A/AB) and 12 patients had myasthenia gravis (48% of patients with available information on myasthenia status). All tumours arising in myasthenia-positive German patients were diagnosed as type AB thymoma in Masaoka-Koga stage I or II (TNM: pT1a stage I).

Myasthenic status was known in 17 Indian patients, of these 14 had MG (82.3%) of which 6 (42.9%) had type A, 5 (35.7%) type AB thymomas and 3 cases (21.4%) type B thymomas. The single myasthenic patient with an atypical type A thymoma had recurrence of disease, and another patient with a “conventional” type A thymoma died due to myasthenic crisis. High risk features seen in the atypical A thymoma were increased atypia of the epithelial cells along with frequent mitotic activity and areas of necrosis.

Hence, a clear tendency towards significantly more frequent MG could be seen in the Indian cohort (82 vs. 48%; \( p = 0.0504; \) **Table 1**). When accounting for age differences between cohorts, no significant effect remained (\( p = 0.13; \) **Table 1**). No significant difference in association between geographical origin and MG status could be observed between age groups (\( p = 0.69; \) **Figure 3**).

**TABLE 3** | Contingency table of clinico-pathological factors between origins (for histological type AB).

| Level       | German | Indian | \( p \) |
|-------------|--------|--------|--------|
| Gender (%)  |        |        |        |
| Female      | 13 (41.9) | 11 (52.4) | 0.573 |
| Male        | 18 (58.1) | 10 (47.6) | —      |
| MG Status (%) |       |        |        |
| No          | 7 (36.8) | 0 (0.0)  | 0.272 |
| MG          | 12 (63.2) | 5 (100.0) | —      |
| Masaoka-Koga stage (%) | | | |
| I           | 11 (35.5) | 15 (71.4) | 0.006 |
| II          | 19 (61.3) | 4 (9.0)   | —      |
| III/IV      | 1 (3.2)  | 2 (9.5)   | —      |
| Mutation status (%) | | | |
| WT          | 2 (8.5)  | 4 (22.2)  | 0.175 |
| L404H       | 29 (93.5) | 14 (77.8) | —      |
| Age [median (IQR)] | | | 0.007 |
| —           | 59.00 (50.00, 70.50) | 50.00 (41.00, 55.00) | |

MG—myasthenia gravis; IQR—interquartile Range.

**FIGURE 3** | Forest plot of logistic regression models for association between origin and MG status in age subgroups. “IA \( p \)” gives test on interaction/heterogeneity between subgroups.
Intriguingly, all German patients with MG had thymomas of AB histotype, compared with only 35.7% of Indian patients (Tables 4 and 5).

The analysis of clinico-pathological factors between MG and non-MG patients (pooled German and Indian groups) showed significant differences in Masaoka-Koga stage between the two groups. Patients with MG presented at earlier stage (100% MG versus 56.2% non-MG patients at Masaoka-Koga stage I or II; \( p = 0.0013; \) Table 6).

**GTF2I Mutation Status**

Out of the 114 thymomas, 3 Indian cases were excluded from mutational studies since the extracted DNA was of insufficient quality. Overall, mutation rates were similar in both cohorts (64 vs. 65%, \( p = 1.00 \), age-adjusted \( p = 0.26 \); Table 1). As expected, in general there was a strong association between histotypes A and AB and GTF2I mutation (\( p < 0.0001 \); Table 7). Out of a total of 89 type A/AB thymomas (including atypical forms), 71 (79.8%) were positive for the GTF2I mutation, including 78.6% (22/28) Indian and 80% (49/61) German cases. Overall, 20.2% (18/89) of types A/AB were negative for the mutation. Among the type B thymomas, overall prevalence of the mutation was 9.1% (2/22) of which all Indian B histotypes (\( n = 6 \)) were negative, whereas 2 of 16 (12.5%) German B thymomas were positive, hence without significant difference (\( p > 0.05 \)). In 25 German cases with known myasthenia gravis status, the GTF2I mutation was evident in 11/12 patients with myasthenia gravis (91.7%) and 10/13 myasthenia-free patients (76.9%). Out of 14 Indian cases with MG, 10 (71.4%) showed the GTF2I mutation; 13 patients (33.3%) without myasthenia harboured the mutation. There was no significant association of the GTF2I mutational status with the prevalence of MG overall and in both cohorts. No mutation was found in non-thymomatous samples, including 18 thymic carcinomas. Association between origin and mutational status was not significantly different between clinical subgroups (Figure 4).

**DISCUSSION**

The potential impact of racial-ethnic, and environmental factors on the pathogenesis of thymoma has rarely been addressed. A study of the International Thymic Malignancy Interest Group (ITMIG) with over 4,000 thymomas from Asia, Europe and the Americas reported geographic differences for the prevalence of WHO thymoma histotypes between continents (11). That study did not include patients from India, i.e., a population with distinct heterogeneous germ line differences (12) and environmental features, including specific dietary habits (13). Thus, the present study is the first to address such
differences through the comparison of Indian and German thymoma patients with regard to the prevalence of the thymoma-specific GTF2I mutation, clinical manifestation, paraneoplastic myasthenia gravis and histology. In the Indian cohort, we observed a significantly younger age at the time point of surgery, a higher prevalence of thymoma-associated MG, and a lower Masaoka-Koga (but not TNM-based) tumour stage. By contrast, gender and WHO histotype distribution, GTF2I mutation rates, and absent association between the presence of MG and GTF2I mutational status were similar in both cohorts.

As to the higher proportion of MG (+) cases among the Indian thymomas compared to the German and a previous world-wide cohorts (5, 6), referral bias needs to be taken into consideration, since the 82% MG positivity rate in the Indian cohort is much higher than any other percentage reported previously in larger series (14, 15). Furthermore, striking differences of MG frequencies in cohorts from different institutions in the same country were reported in meta-analyses and thought to reflect referral bias (14). Referral bias also could have a bearing on the lower tumour stage observed in the Indian cohort, since MG (+) thymomas may be detected earlier than MG (−) thymomas through myasthenic rather than local symptoms that are typical of larger thymomas in MG (−) patients (16). Indeed, also in our study, the different nature of the Indian and German institutes should be taken into consideration: while Indian patients come from an institute that is super-specialized in neurosciences, the major focus of the German referral center is thymic pathology. This fact may contribute to the striking difference in MG prevalence between the two groups and should be appreciated as one of the limitations of the study. Therefore, we are reluctant to attribute the lower tumour stage in Indian patients to racial-ethnic differences. By contrast, the median age difference of 15 years between Indian and German thymoma patients (50 versus 65 years) is intriguing, since such a young age, for a cohort that is dominated by type A and AB thymomas is
among the lowest reported to date (14, 15). However, a comparably young age was found in another Asian cohort from Korea (17). This observation needs validation in a larger and MG-unbiased cohort of Indian thymoma patients. If confirmed, a younger manifestation age may hint to an increased risk for thymoma and thymoma-associated MG development in Indians. In fact, the proportion of thymoma patients among MG patients was found to be higher in some Asian countries compared to Northern European countries (18-20).

Since gender and WHO histotype distribution as well as GTF2I mutation rates were similar in the Indian and German cohort and in line with a previous world-wide study (6), racial-ethnic, environmental and lifestyle factors appear not to have a major impact on the molecular mechanism(s) driving the development of thymomas. A similar conclusion was recently drawn in relation to thymic squamous cell carcinomas in which age-related rather than racial-ethnic and environmental factors appear to be related to oncogenesis (6, 21).

Our study has some limitations: First, different sample size of the two cohorts together with the overall low number of patients with known MG status in this retrospective analysis preclude the drawing of definite conclusions. Second, the very high prevalence of MG patients in the Indian cohort largely precludes the detection of any differences between Indian and German non-MG patients. Third, since the total mutational burden of thymomas is extremely low (6), the prevalence of the only common and unique mutation, GTF2I, was the sole readout to study molecular differences between Indian and German thymoma patients. Therefore, it will be interesting to see, whether multiomics analyses can still identify molecular thymoma features that segregate with racial-ethnic and environmental differences. Fourth, since the GTF2I mutation is largely restricted to type A and AB thymomas, the two patients cohorts were equally and strongly skewed towards an over-representation of these two histologies and are thus not representative regarding the histotype distribution (22). Therefore, future comparative studies between racially and ethnically disparate groups should comprise type B thymomas and thymic carcinomas as well. Finally, we had no information on the patients’ specific racial-ethnic and dietary settings, precluding a distinction between potential genetic and environmental reasons for the observed differences. These caveats underline the necessity of comprehensive molecular and epidemiological studies of larger and more representative cohorts of Indian and German thymoma patients to identify genetic and environmental risk factors for the development of thymomas and their complications.

**FIGURE 4 |** Bar chart of mutation rates between origin in the full cohorts and in subgroups.
In summary, comparing rather unique cohorts of Indian and German thymoma patients, we could not identify significant differences regarding WHO histological types, gender and GTF2I mutation status, implying that there are no differences in molecular pathogenesis of thymoma between the two countries. Nonetheless, the significantly younger age of Indian thymoma patients is a first hint that genetic or environmental factors could contribute to the timing and overall risk of thymoma development.

**DATA AVAILABILITY STATEMENT**

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found below: https://www.ncbi.nlm.nih.gov/genbank/, BankIt2452830 GTF2I MZ015743.

**ETHICS STATEMENT**

The studies involving human participants were reviewed and approved by the Ethic Committee of the University Medical Centre Mannheim, University of Heidelberg and All India Institute of Medical Sciences, New Delhi Institution Ethics Committee. Written informed consent for participation was approved by the Ethic Committee of the University Medical Institute Ethics Centre Mannheim, University of Heidelberg and All India Institution Ethics Committee. Written informed consent for participation was not required for this study in accordance with the national legislation and the institutional requirements.

**REFERENCES**

1. Travis WD, Brambilla E, Nicholson AG, Yatabe Y, Austin JHM, Beasley MB, et al. The 2015 World Health Organization Classification of Lung Tumors. *J Thorac Oncol* (2015) 10(9):1243–60. doi:10.1097/JTO.0000000000000639
2. Badve S, Goswami C, Gökmen–Polar Y, Nelson RP, Henley J, Miller N, et al. Molecular Analysis of Thymoma. *PLoS One* (2012) 7(8):e42669. doi:10.1371/journal.pone.0042669
3. Enkner F, Pichhofe B, Zaharie AT, Krunic M, Holper TM, Janik S, et al. Molecular Profiling of Thymoma and Thymic Carcinoma: Genetic Differences and Potential Novel Therapeutic Targets. *Pathol Oncol Res* (2017) 23(3):551–64. doi:10.1007/s12553-016-0144-8
4. Girard N, Shen R, Guo T, Heguy A, Riely GJ, Zakowski MF, et al. The 2015 World Health Organization Classification of Lung Tumors. *Clin Cancer Res* (2017) 23(11):2935–49. doi:10.1158/1078-0432.ccr-16-2805
5. Lee H-S, Jang H-J, Shah R, Yoon D, Hamaqi M, Wald O, et al. Genomic Analysis of Thymic Epithelial Tumors Identifies Novel Subtypes Associated with Distinct Clinical Features. *Clin Cancer Res* (2018) 24(11):3065–79. doi:10.1158/1078-0432.ccr-17-1522
6. Radovich M, Pickering CR, Felau I, Ha G, Zhang H, Jo H, et al. The Integrated Genomic Landscape of Thymic Epithelial Tumors. *Cancer Cell* (2018) 33(2):244–e10. doi:10.1016/j.ccell.2018.01.003
7. Feng Y, Lei Y, Wu X, Huang Y, Rao H, Zhang Y, et al. GTF2I Mutation Frequently Occurs in More Indolent Thymic Epithelial Tumors and Predicts Better Prognosis. *Lung Cancer* (2017) 110:48–52. doi:10.1016/j.lungcan.2017.05.020
8. Petroni I, Meltzer PS, Kim I-K, Lucchi M, Park K-S, Fontanini G, et al. A Specific Missense Mutation in GTF2I Occurs at High Frequency in Thymic Epithelial Tumors. *Nat Genet* (2014) 46(8):844–9. doi:10.1038/ng.3016
9. Dettetbeck FC, Nicholson AG, Kondo K, Van Schil P, and Moran C. The Masaoka-Koga Stage Classification for Thymic Malignancies: Clarification and Definition of Terms. *J Thorac Oncol* (2011) 6(7 Suppl. 3):S1710–6. doi:10.1097/JTO.0b013e31821e8c4f
10. Porubsky S, Popovic ZV, Badve S, Banz Y, Berezowska S, Borchert D, et al. Thymic Hyperplasia with Lymphoepithelial Sialadenitis (LESA)-Like Features: Strong Association with Lymphomas and Non-myasthenic Autoimmune Diseases. *Cancers (Basel)* (2021) 13(2). doi:10.3390/cancers13020315
11. Weiss C-A, Yao X, Deng Y, Dettetbeck FC, Marino M, Nicholson AG, et al. The Impact of Thymoma Histotype on Prognosis in a Worldwide Database. *J Thorac Oncol* (2015) 10(2):367–72. doi:10.1097/JTO.0000000000000393
12. Reich D, Thangaraj K, Patterson N, Price AL, and Singh L. Reconstructing Indian Population History. *Nature* (2009) 461(7263):489–94. doi:10.1038/nature08365
13. Bhupathiraju SN, Guasch-Ferré M, Gadgil MD, Newgard CB, Bain JR, Muehlbauer MJ, et al. Dietary Patterns among Asian Indians Living in the United States Have Distinct Metabolomic Profiles that Are Associated with Cardiometabolic Risk. *J Nutr* (2018) 148(7):1150–9. doi:10.1093/jn/nny1074
14. Dettetbeck FC. Clinical Value of the WHO Classification System of Thymoma. *Ann Thorac Surg* (2006) 81(6):2328–34. doi:10.1016/j.athoracsur.2005.11.067
15. Ruffini E, Guerrera F, Brunelli S, Passani S, Pellicano D, Thomas P, et al. Report from the European Society of Thoracic Surgeons Prospective Thymic Database 2017: a Powerful Resource for a Collaborative Global Eort to Manage Thymic Tumours. *Eur J Cardiothorac Surg* (2019) 55(4):601–9. doi:10.1093/ejcts/ezys448
16. Ströbel P, Bauer A, Puppe B, Kraushaar T, Krein A, Toyka K, et al. Tumor Recurrence and Survival in Patients Treated for Thymomas and Thymic Squamous Cell Carcinomas: a Retrospective Analysis. *Ico* (2004) 22(8):1501–9. doi:10.1200/jco.2004.10.113

**AUTHOR CONTRIBUTIONS**

DJ, AM, and ZP initiated and designed the study and wrote the manuscript. PG helped in writing the manuscript and collating data. VS helped in mutation analysis. TH analysed data and provided critical feedback. RP, SK, KK, GO, GP, and SS contributed clinical data. TG, SP, CS, SS, and PS participated in data interpretation. All authors helped shape the manuscript and approved the final version.

**FUNDING**

We acknowledge financial support by the Open Access Publishing Fund of Ruprecht-Karls-Universität Heidelberg.

**CONFLICT OF INTEREST**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**SUPPLEMENTARY MATERIAL**

The Supplementary Material for this article can be found online at: https://www.por-journal.com/articles/10.3389/pore.2021.1609858/full#supplementary-material

Pathology & Oncology Research 9 August 2021 | Volume 27 | Article 1609858
17. Kim DJ, Yang WI, Choi SS, Kim KD, and Chung KY. Prognostic and Clinical Relevance of the World Health Organization Schema for the Classification of Thymic Epithelial Tumors. *Chest* (2005) 127(3):755–61. doi:10.1378/chest.127.3.755

18. Boldingh MI, Maniaol A, Brunborg C, Dekker L, Lipka A, Niks EH, et al. Prevalence and Clinical Aspects of Immigrants with Myasthenia Gravis in Northern Europe. *Muscle Nerve* (2017) 55(6):819–27. doi:10.1002/mus.25408

19. Huang X, Liu WB, Men LN, Feng HY, Li Y, Luo CM, et al. Clinical Features of Myasthenia Gravis in Southern China: a Retrospective Review of 2,154 Cases over 22 Years. *Neurol Sci* (2013) 34(6):911–7. doi:10.1007/s10072-012-1157-z

20. Murai H, Yamashita N, Watanabe M, Nomura Y, Motomura M, Yoshikawa H, et al. Characteristics of Myasthenia Gravis According to Onset-Age: Japanese Nationwide Survey. *J Neurol Sci* (2011) 305(1-2):97–102. doi:10.1016/j.jns.2011.03.004

21. Yamada Y, Simon-Keller K, Belharazem-Vitacolononna D, Bohnenberger H, Kriegsmann M, Kriegsmann K, et al. A Tuft Cell-like Signature Is Highly Prevalent in Thymic Squamous Cell Carcinoma and Delineates New Molecular Subsets Among the Major Lung Cancer Histotypes. *J Thorac Oncol* (2021). doi:10.1016/j.jtho.2021.02.008

Copyright © 2021 Jain, Guleria, Singh, Parshad, Kumar, Gaiser, Kurz, Ott, Porubsky, Preissler, Sauer, Schölch, Ströbel, Hielscher, Marx and Popovic. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.