Phenotypes and personalized medicine in the acute respiratory distress syndrome

Michael A. Matthay, Yaseen M. Arabi, Emily R. Siegel, Lorraine B. Ware, Lieuwe D. J. Bos, Pratik Sinha, Jeremy R. Beitler, Katherine D. Wick, Martha A. Q. Curley, Jean-Michel Constantin, Joseph E. Levitt, and Carolyn S. Calfee

Abstract

Although the acute respiratory distress syndrome (ARDS) is well defined by the development of acute hypoxemia, bilateral infiltrates and non-cardiogenic pulmonary edema, ARDS is heterogeneous in terms of clinical risk factors, physiology of lung injury, microbiology, and biology, potentially explaining why pharmacologic therapies have been mostly unsuccessful in treating ARDS. Identifying phenotypes of ARDS and integrating this information into patient selection for clinical trials may increase the chance for efficacy with new treatments. In this review, we focus on classifying ARDS by the associated clinical disorders, physiological data, and radiographic imaging. We consider biologic phenotypes, including plasma protein biomarkers, gene expression, and common causative microbiologic pathogens. We will also discuss the issue of focusing clinical trials on the patient’s phase of lung injury, including prevention, administration of therapy during early acute lung injury, and treatment of established ARDS. A more in depth understanding of the interplay of these variables in ARDS should provide more success in designing and conducting clinical trials and achieving the goal of personalized medicine.

Keywords: Acute respiratory distress syndrome, Acute lung injury, Sepsis, Pulmonary edema, COVID-19, Phenotype, Precision medicine

Introduction

The acute respiratory distress syndrome (ARDS) is defined clinically by the onset of acute respiratory failure with hypoxemia and bilateral pulmonary infiltrates on chest imaging that cannot be primarily attributed to volume overload, left ventricular dysfunction or chronic lung disease. However, these relatively simply clinical criteria do not capture the complexity and diversity of ARDS [1]. The syndromic definition of ARDS encompasses different clinical disorders associated with ARDS, a range of pulmonary physiologic abnormalities, varying chest radiographic abnormalities, variability in biologic pathways of injury as reflected by plasma protein biomarkers and gene expression, diverse microbiologic etiologies, and an evolution over time ranging from early development of acute lung injury in spontaneously breathing patients to ventilated patients who meet the Berlin criteria of ARDS.

The theme of this article is to consider the potential for classifying ARDS into phenotypes on the basis of clinical, physiologic, radiologic, and biologic criteria that may lead to more targeted therapies that could improve clinical outcomes. For example, ARDS from pneumonia may be caused by a variety of pathogens. Identifying the causative microbiologic agent can lead to specific treatment that may be effective, as has been shown in the coronavirus disease 2019 (COVID-19) pandemic with the recent evidence that anti-viral therapy with remdesivir...
reduces morbidity [2]. As another example of how subdividing ARDS may be useful, patients with more moderate to severe hypoxemia (P/F < 150 mmHg) benefit from prone positioning [3]. One of the central challenges facing the field in ARDS is whether or how to incorporate our expanded knowledge of ARDS phenotypes into future clinical trials and practice, with the hope that this approach could identify subsets that will be more responsive to specific treatments beyond the current supportive care therapies that have improved clinical outcomes. We will also consider time dependent factors in the evolution of acute lung injury and ARDS, since this variable is one determinant of which therapies might be more effective.

The reader is also referred to a related article in this issue of Intensive Care Medicine that considers approaches to the design of ARDS trials based on prognostic and predictive enrichment strategies that identifies some approaches that are based on physiologic, systemic, radiographic or biologic criteria [4].

Clinical categories
The development of ARDS is associated with a wide range of infectious and non-infectious clinical conditions (Fig. 1). Sepsis, primarily due to pneumonia, accounts for the majority of cases among both adults and children, though nonpulmonary sepsis (e.g., due to urinary tract infection, bowel perforation, or other causes) may

---

**Take-home message:**
Integration of clinical, physiologic, radiographic, microbiologic and biologic variables can provide pathways for defining phenotypes and testing therapeutics in future clinical trials that can lead to a more personalized approach for therapies in ARDS.

---

**Fig. 1** Some recognized etiologies of ARDS. Circle size represents approximate relative frequency, although we do not have enough information regarding frequency for this figure to be an accurate estimate for COVID-19 viral pneumonia. Clinical disorders associated with ARDS include drug-induced ARDS and ARDS following major surgical procedures such as cardiopulmonary bypass.
also lead to ARDS. Aspiration, trauma, and blood product transfusions are also common etiologies [5–8]. More recent clinical categories of ARDS include E-cigarette or vaping associated lung injury (EVALI) and the pandemic of SARS-CoV-2 related ARDS [9–11]. There are other clinical disorders associated with ARDS including smoke inhalation, acute exacerbations of interstitial lung disease and primary graft dysfunction following lung transplantation [12]. Negative pressure pulmonary and re-expansion pulmonary edema are often thought of as causes of ARDS, but they are primarily a form of hydrostatic pulmonary edema and not acute lung injury [13, 14].

Clinical categorization remains an important anchoring framework for both the management and prognosis of ARDS. For example, the presence of sepsis not only guides antimicrobial strategy, but also is associated with more severe illness and worse outcomes [15] and has informed clinical trial design by enrolling patients with a common risk factor [16, 17]. Whether or not shock is present offers further specificity to clinical classification, although the presence or absence of shock does not readily distinguish patients identified by biologic factors [18, 19]. Clinical categories of ARDS classified as direct (e.g., pneumonia, aspiration, or ischemia–reperfusion injury) or indirect (e.g., non-pulmonary sepsis, multiple transfusions, pancreatitis) differ in clinical predictors of mortality and in their biomarker profiles; however, there is as yet no evidence that treatment responses consistently differ on this basis [20, 21].

Mortality from ARDS overall has decreased since the syndrome was first described [22, 23]; however, mortality continues to differ by clinical risk factor. In 2001, one study noted that the mortality of patients in a clinical trial of low tidal volume ventilation ranged from 43% in patients with sepsis to 11% in those who had sustained major trauma [24]. Data from 2006–2014 showed a similar mortality rate for sepsis starting at 46% and declining to 40% [25]. Of note, the mortality rate for trauma started at above 20% with an upward trend over the study interval. This lack of improvement in mortality in the trauma group was confirmed in a recent study comparing data from 1990 through 2009 [26]. Mortality in this study was also above 20%, though much of this may be attributable to traumatic brain injury [27]. The incidence of transfusion associated lung injury (TRALI) has significantly decreased with the discovery that fresh frozen plasma from female patients was a major factor in TRALI pathogenesis [28, 29]. Recent studies suggest that the risk of developing ARDS is greater with direct than indirect injury [30], and although predictors of mortality differ between direct and indirect ARDS, overall mortality rates are likely similar [20, 30]. In addition, one recent study from two institutions reported that attributable mortality in patients who developed ARDS from sepsis was primarily related to the initial severity of hypoxemia (PaO2/ FiO2 < 100 mmHg) [31].

The different clinical categories associated with ARDS illustrate the complexity of ARDS in terms of pathogenesis and also highlight how personalized medicine approaches will need to consider how the associated clinical category influences therapy. For example, patients with COVID-19 ARDS probably benefit from the antiviral therapy, remdesivir, but additional data are needed to confirm whether mechanically ventilated patients truly benefit [2]. Survival in these patients is improved by the use of the anti-inflammatory therapy dexamethasone [32], in contrast to the uncertain benefit of corticosteroids in unselected cohorts of patients with ARDS who do not share a common clinical risk factor [33, 34]. This discrepancy illustrates the potential value of focusing on distinct ARDS phenotypes based on clinical risk factor—that is, the signal for corticosteroid benefit is much clearer in COVID-19 ARDS than it has been in 50 years of study in non-COVID-19 ARDS. However, even the role of corticosteroids in COVID-19 will require more study to understand when dexamethasone may be harmful or beneficial [35] and will need re-evaluation in conjunction with new therapies that may show efficacy as well, including potentially monoclonal antibodies, immunomodulatory agents, and new anti-viral therapies.

**Physiologic categories**

In adult ARDS, the Berlin definition categorizes mild ARDS as a PaO2/FiO2 of 201–300 mmHg, moderate as PaO2/FiO2 of 101–200 mmHg, and severe as PaO2/FiO2 of equal to or less than 100 mmHg (Table 1) [36]. Criteria for moderate and severe ARDS also require invasive mechanical ventilation with positive end-expiratory pressure (PEEP) of 5 cmH2O or greater, while in mild ARDS, the PEEP requirement can be met noninvasively. In a patient-level meta-analysis, mortality increased from 27% to 32% to 45% for mild to moderate to severe ARDS [36, 37]. Several recent trials have used PaO2/FiO2 for prognostic enrichment, with PaO2/FiO2 eligibility thresholds of no more than 150 or 200 being most common [3, 38–40]. PaO2/FiO2 has important limitations, however, including being heavily influenced by ventilator settings, especially PEEP [41, 42].

Higher pulmonary dead-space fraction and surrogate indices such as ventilatory ratio (VR) also correlate with ARDS mortality [43–46]. VR is defined as [minute ventilation (mL/min) × PaCO2 (mmHg)]/[predicted body weight × 100 × ideal PaCO2 (mmHg)], where ideal PaCO2 is 37.5 mmHg. Whether dead-space fraction, VR, or other physiologic measures are useful to guide
therapeutic selection is not well explored but has been advocated for consideration [47, 48]. Several investigators have advocated categorizing ARDS by respiratory mechanics [49–51]. Among the most promising measures are airway driving pressure (plateau airway pressure minus PEEP) [51] and transpulmonary pressure (airway minus pleural pressure) [52–54]. Both have intuitive appeal for quantifying overdistension and, for transpulmonary pressure, risk of end-expiratory airway closure and lung collapse [40, 55]. Respiratory mechanics have value for guiding mechanical ventilation [56, 57], but whether they are useful for phenotyping ARDS for interventions unrelated to ventilator-induced lung injury (VILI) is less clear. One study did find that improved oxygenation in response to PEEP identified patients with better survival [58], but this strategy still requires prospective study.

Pediatric critical care clinicians have long recognized the inadequacies of adult based operational definitions for ARDS when applied to children [59, 60]. First, pediatric risk factors, etiologies, pathophysiology, and outcomes differ [61, 62]. Second, PaO₂ is measured less often, making the calculation of PaO₂/FiO₂ more challenging, since invasive arterial sampling is used more sparingly in children. Third, dissimilar ventilator modes are commonly used in children, and the oxygenation index (OI) ((FiO₂ × mean airway pressure (Paw) × 100)/PaO₂) or oxygen saturation index (OSI) ((FiO₂ × Paw × 100)/SpO₂) are primarily used to describe hypoxemia [63, 64]. The 2015 pediatric ARDS (PARDS) consensus [65] definition excludes infants with perinatal related lung disease, allows for unilateral or bilateral infiltrates, and requires OI or OSI to assess oxygenation during invasive ventilation, but criteria for edema origin and timing are similar to adult ARDS. The consensus definition delineates mild, moderate and severe PARDS based on OI or OSI for patients on mechanical ventilation, with no stratification for patients on non-invasive ventilation (Table 1).

The ongoing PROSpect trial (NCT03896763) is a two-by-two factorial, response-adaptive, randomized controlled clinical trial of supine vs. prone positioning and conventional mechanical ventilation vs. high-frequency oscillatory ventilation being conducted at 60 PICUs internationally. Up to 1000 participants will be enrolled and adaptive randomization will begin after data is collected for 400 subjects and will repeat every 100 patients thereafter. Any group demonstrating inferiority at a pre-planned adaptive randomization update analysis will be eliminated to increase allocation to the superior group(s). The trial also includes sampling of plasma biomarkers which will make it potentially possible to identify treatment responsive subsets within each of the four treatment groups or within the PARDS patients overall that are enrolled in the trial.

### Chest radiography
Radiographic methods have been used for identifying morphological phenotypes of ARDS that may respond differently to ventilator strategies and for assessing ARDS.

#### Table 1 Definitions and severity classification for ARDS and PARDS

| Criteria | ARDS* | PARDSb |
|----------|-------|--------|
| **Timing** | Within 7 days of known clinical insult or new or worsening respiratory symptoms | Within 7 days of known clinical insult |
| **Origin of edema** | Respiratory failure not fully explained by cardiac failure or fluid overload. Objective assessment needed to exclude hydrostatic edema if no risk factor present | Respiratory failure not fully explained by cardiac failure or fluid overload |
| **Chest imaging** | Bilateral opacities not fully explained by effusions, lobar/lung collapse, or nodules | New infiltrate(s) consistent with acute pulmonary parenchymal disease |
| **Age** | Adult, no age cutoff specified | Perinatal related lung disease is excluded. No age cutoff specified |
| **Oxygenation and severity** | | |
| **Noninvasive** | Included in mild ARDS | Full face-mask bi-level ventilation or CPAP ≥ 5 cmH₂O with PaO₂/FiO₂ ≤ 300 or SpO₂/FiO₂ ≤ 264 |
| Mild | 200 < PaO₂/FiO₂ ≤ 300 with PEEP or CPAP ≥ 5 cmH₂O | 4 ≤ OI < 8, or 5 ≤ OSI ≤ 7.5 |
| Moderate | 100 < PaO₂/FiO₂ ≤ 200 with PEEP ≥ 5 cmH₂O | 8 ≤ OI ≤ 16, or 7.5 ≤ OSI ≤ 12.3 |
| Severe | 100 < PaO₂/FiO₂ ≤ 100 with PEEP ≥ 5 cmH₂O | OI ≥ 16, or OSI ≥ 12.3 |

ARDS, acute respiratory distress syndrome; CPAP, continuous positive airway pressure; OI, oxygenation index; OSI, oxygen saturation index; PARDS, pediatric acute respiratory distress syndrome; PEEP, positive end-expiratory pressure

* ARDS Berlin Definition [36]

b Pediatric Acute Lung Injury Consensus Conference definition [65]. Special considerations addressed for cyanotic heart disease, chronic lung disease, and left ventricular dysfunction

---

ARDS, acute respiratory distress syndrome; CPAP, continuous positive airway pressure pressure; OI, oxygenation index; OSI, oxygen saturation index; PARDS, pediatric acute respiratory distress syndrome; PEEP, positive end-expiratory pressure

* ARDS Berlin Definition [36]

b Pediatric Acute Lung Injury Consensus Conference definition [65]. Special considerations addressed for cyanotic heart disease, chronic lung disease, and left ventricular dysfunction
severity. Studies of the morphologic characteristics of radiographic lung infiltrates on computed tomography (CT) have identified two subgroups termed focal and non-focal ARDS that appear to have distinct lung physiology [66, 67]. Patients with non-focal ARDS have diffuse alveolar opacities on chest CT, whereas patients with focal ARDS have focal loss of aeration predominantly in the lower lobes and dependent regions. Based on the observation that the lungs of patients with non-focal ARDS are significantly more recruitable than lungs of patients with focal ARDS, it is postulated that these radiographic subgroups may benefit from different ventilator strategies. Specifically, non-focal ARDS may benefit from a higher PEEP strategy with recruitment maneuvers to maximize recruitment of diffusely non-aerated lung. By contrast, focal ARDS may benefit from a lower PEEP strategy that minimizes overdistension of less affected regions of the lungs combined with proning to improve matching of ventilation to perfusion.

To test this hypothesis, European investigators designed the LIVE trial [68], a randomized clinical trial that compared personalization of ventilator settings according to CT assessment of lung morphology vs. standard 6 mL/kg tidal volume, lower PEEP ventilation in 420 patients with moderate-to-severe ARDS. In the personalized ventilator arm, patients with non-focal ARDS received a tidal volume of 6 mL/kg, recruitment maneuvers and high PEEP, while those with focal ARDS received a tidal volume of 8 mL/kg, low PEEP, and prone position. In an intention-to-treat analysis, there was no difference in 90-day mortality between the groups. However, misclassification of patients as having focal or non-focal ARDS occurred in 21% of patients, highlighting the difficulty of prospective assessment of ARDS phenotypes by CT. In a subgroup analysis, the 90-day mortality of the misclassified patients was significantly higher in the personalized group than in the standard ventilation group. This finding suggests that although tailoring mechanical ventilation to lung morphology might decrease mortality in correctly classified patients, misalignment of mode of mechanical ventilation and morphologic phenotype in ARDS may be particularly deleterious.

Plain chest radiographs have been studied as a tool to assess the extent of pulmonary edema in patients with ARDS and are more readily obtained than chest CTs in the critically ill. One group of investigators developed the Radiographic Assessment of Lung Edema (RALE) score to quantify both the extent and density of alveolar opacities by quadrant of the chest radiograph [69]. The RALE score provides a semiquantitative assessment of the extent of radiographic edema that has been validated against gravimetric measurement of lung edema in explanted lungs from organ donors. In patients with ARDS enrolled in a randomized trial of conservative vs. liberal fluid management [70], higher RALE scores were independently associated with lower PaO2/FiO2 and worse survival. Conservative fluid management significantly decreased RALE score over 3 days compared with liberal fluid management. Two subsequent studies have confirmed that both the baseline RALE score and the change in RALE over time are associated with clinical outcomes in ARDS [71, 72]. Given the ready availability of chest radiographs, the RALE score may have value as a method for prognostic enrichment in clinical trials, in particular for trials that specifically target resolution of pulmonary edema, and could be complementary to assessment of lung mechanics at the bedside. How the RALE score relates to CT assessment of diffuse vs. focal ARDS, and associated implications for personalizing treatment, remains unknown.

**Recent advances in protein biomarkers**

Several recent discoveries in protein biomarkers enable a better understanding of the pathophysiology of ARDS, improve prognostication and offer a possible route to precision-based care and assignment of phenotypes. Protein biomarkers in ARDS are most frequently studied in blood (usually plasma) or the air spaces of the lung (usually bronchoalveolar lavage fluid, BALF). Theoretically, BALF offers the advantage of studying injured lungs directly. Variance in specimen dilution secondary to exudative edema and/or acquisition procedures limits the standardisation of protein quantification in BALF [73]. Furthermore, the need for fiberoptic bronchoscopy also limits the routine clinical measurements of BALF protein biomarkers in ARDS [74, 75]. Less invasive sample acquisition using blind lavage or tracheal aspirates has been described; their relevance to corresponding BALF samples is uncertain [76]. In a recent innovation, investigators have studied proteins that were captured in the heat moisture exchanger filter in ventilated patients with ARDS; protein concentrations were reasonably well correlated with undiluted edema fluid samples (r2 0.849–0.951) [77]. Further studies are needed to validate this non-invasive approach to study airspace biomarkers.

While most protein biomarkers have been confined exclusively to the research realm, alveolar type III procollagen peptide (PCP-III) is a compelling BALF protein candidate that has been associated with pathophysiology and outcomes in ARDS [78, 79]. In an observational study of 32 patients, alveolar PCP-III was highly sensitive (0.90) and specific (0.92) for diagnosing fibroproliferation in ARDS [80]. In a subsequent study, the same cut-off was associated with fibroproliferative changes on lung CT [81]. Further studies are underway to determine if PCP-III can be used to guide and test interventions with
corticosteroids in ARDS (NCT#03371498). In COVID-19, several routinely measured serum biomarkers, such as ferritin, CRP, and d-dimer, have been associated with prognosis. Currently, it is unclear whether these acute-phase proteins are merely indicators of disease severity or offer opportunity for actionable interventions and/or are mechanistically informative. Further studies are needed to elucidate the precise role of these markers both in COVID-19 and non-COVID-19 ARDS.

Translation of biomarkers into the clinical domain has been challenging, in part due to the pursuit of a single discriminating biomarker that is continuously distributed in a heterogeneous clinical syndrome [82]. Investigators are increasingly applying machine learning algorithms to identify biologically discrete clusters of ARDS using multiple protein biomarkers [83]. A series of studies applying latent class analysis (LCA) used a combination of clinical data and plasma biomarkers to identify phenotypes in ARDS [18, 84–86]. Using this approach, hypo- and hyper-inflammatory phenotypes of ARDS were consistently identified across five RCTs, and these phenotypes exhibited divergent biological characteristics, clinical features and outcomes. The hyper-inflammatory phenotype was associated with exaggerated inflammation with elevated plasma levels of IL-8, IL-6, and sTNFR-1. Levels of PAI-1 were also higher in this phenotype, whereas protein C was lower. Mortality at day 90 was approximately twofold higher in the hyper-inflammatory phenotype across all cohorts. Importantly, in three of the trials, differential treatment responses were observed in the phenotypes with improved outcomes in the treatment arm for patients in the hyperinflammatory subgroup [18, 84, 85]. None of the original trials showed a mortality benefit [70, 87, 88]. In related work, a group of Dutch investigators applied cluster analysis to plasma protein biomarker data from ARDS patients and identified two phenotypes, which they termed “reactive” and “uninflamed”; these phenotypes have prognostic value as well, though how they correspond to the LCA-phenotypes remains unknown [89].

One challenge of implementing these approaches clinically is the difficulty of identifying these phenotypes at the bedside. LCA models are complex, comprising 30–40 variables that are not amenable for prospective use. To address this challenge, investigators have developed 3-variable (IL-8, bicarbonate and protein C) and 4-variable (addition of vasopressor) parsimonious models that accurately classified the two LCA phenotypes in an independent validation cohort (area under the curve: 0.94 and 0.95, respectively) [86]. Importantly, when one of the ancillary parsimonious models was applied to the HARP-2 study, a randomized clinical trial that tested the efficacy of simvastatin vs. placebo, a survival benefit was observed in
the hyperinflammatory phenotype at day 28 (Fig. 2). The inability to quantify protein biomarkers rapidly at the bedside is another challenge. Recently, in a preliminary report in patients with COVID-19 ARDS, the same investigators quantified IL-6 and sTNFR-1 using a novel point-of-care assay (Evidence Multistat Analyser; Randox Laboratories, Country Antrim, UK) [90]. The assay results were used to assign ARDS phenotype based on the parsimonious model, demonstrating the feasibility of phenotype-classification in real-time. In summary, these studies indicate that the application of biomarkers for clinical trials and/or decision-making may soon be plausible.

**Gene expression**

Gene expression from leukocytes can be used to acquire a snapshot image of the immune response. An advantage of transcriptomics over protein biomarkers is that nucleic acid-based arrays cover tens of thousands of genes and, therefore, provide a more comprehensive overview than the measurement of a limited number of proteins. The major disadvantage is that relative quantification of messenger RNA does not provide direct information on a number of transcribed proteins and may, therefore, not directly reflect the functional landscape in organ injury. In addition, it is challenging to translate gene expression to a point of care assay. Peripheral blood transcriptomics has been used to subphenotype patients with sepsis resulting in the identification of two to four subphenotypes with clear differences in immune exhaustion, innate immune response, T-cell balance and coagulation [91–94].

In ARDS, such an approach has not been tried. Gene expression profiles of peripheral leukocytes were, however, compared between ARDS patients who were classified as having a “reactive” or “uninflamed” subphenotype based on a set of 20 plasma biomarkers of inflammation, coagulation and endothelial injury, as described above [95]. Approximately 30% of all genes were differentially expressed between these subphenotypes. Genes associated with neutrophil activation were enriched in the “reactive” subphenotype, which is consistent with the activated innate immune response suggested by increased interleukin-8 and interleukin-6 protein concentrations in this subgroup. However, there was low agreement between subphenotype allocation based on plasma proteins and sepsis subphenotypes informed by blood leukocyte expression profiles. It should be noted that any measurement in blood reflects systemic host response, which is influenced by several processes other than ARDS, as lung injury often does not develop in isolation (Fig. 1). Thus, another study involving 16 patients with ARDS focused on paired measurements of gene expression of alveolar macrophages (AMs) and peripheral blood monocytes (PBMs) [96]. This research group found highly divergent patterns of expression between AMs and PBMs. Enrichment of immunoinflammatory gene sets in AMs was associated with better clinical outcomes, but with worse clinical outcomes in PBMs. Further studies of both the lung and the systemic circulation in ARDS patients will likely provide more insights into the mechanisms of both lung and non-pulmonary organ injury that are relevant for classical ARDS and COVID-19 ARDS (Fig. 3).

**Microbiology**

Bacterial and viral infections of the lung are the most common etiologies associated with ARDS [97]. The causative pathogens differ from ARDS secondary to community-acquired pneumonia (CAP) or hospital-acquired ventilator-associated pneumonia (HAP/VAP) [98, 99]. Pathogens differ also according to age and the presence of co-morbid conditions, prior colonization, exposure history and vaccination status. The treatment of a specific infection is a prerequisite for standard of care and would seem to be a necessary part of any precision medicine approaches in ARDS. However, making the diagnosis of specific infections in ARDS can be challenging and involves standard and new methods and may inform specific phenotype designations in the future.

The typical diagnostic approach for ARDS secondary to CAP or HAP/VAP includes blood cultures and Gram stain and culture of respiratory specimens (expectorated sputum or tracheal aspirate), ideally obtained before commencement of antimicrobial therapy [98, 99]. However, the yield of traditional cultures for detecting organisms is generally low (30–50%) and varies according to organism, receipt of prior antimicrobial therapy, and setting [98].

The use of invasive respiratory sampling (ie, bronchoalveolar lavage [BAL], protected specimen brush [PSB]) and blind bronchial sampling (ie, mini-BAL) leads to higher yield in identifying pathogens compared to traditional microbiologic tests of tracheal aspirate or sputum examination. However, even with invasive respiratory sampling, a high percentage of patients with ARDS remain with no identified organisms [97]. In addition, studies on VAP did not demonstrate an outcome benefit on the routine use of invasive sampling with quantitative cultures compared to noninvasive sampling with quantitative cultures. Invasive respiratory sampling should be performed in immunocompromised patients, in the presence of unexplained, persistent or worsening infiltrates.

Nucleic acid detection tests (NATs), such as polymerase chain reaction (PCR), are becoming available to
detect several pathogens including *Mycoplasma* spp., *Chlamydia pneumoniae* and *Bordetella pertussis* as well as common bacteria such as *Staphylococcus aureus* and *Streptococcus pneumoniae*. In one study, molecular testing for 26 bacterial and viral pathogens of single lower respiratory tract specimens achieved pathogen detection in 87% of CAP patients compared with 39% with culture-based methods [100]. Of these, 78% had a bacterial pathogen detected by PCR but only 32% were culture-positive [100]. Legionella and Pneumococcal Urinary Antigen can be used in patients with ARDS due to CAP [98].

With the increasing use of NATs, viral pathogens are increasingly detected among critically ill adult patients with respiratory illness, with a reported prevalence between 17% to 53% of patients [101]. Detecting viruses in respiratory specimens does not always establish causality with ARDS [102]. However, it is generally believed that most respiratory viruses may cause ARDS, especially in the elderly, the immunocompromised, patients with other co-morbid conditions, and occasionally in previously healthy individuals [103]. While the role of routine testing of PCR panels for respiratory viruses has not been established, testing for influenza with PCR is recommended when influenza is circulating in the community; this test is preferred over other rapid influenza diagnostic tests (i.e., antigen test) [98]. Other uncommon but important endemic pathogens should also be considered in the presence of history of travel, residence in certain geographic locations or history of specific exposures (Table 2).

Next generation sequencing (NGS) techniques have increased our ability to detect pathogens and possibly to detect novel pathogens [104–108]. At present they have limited clinical use, but they have the potential to change our understanding of ARDS epidemiology. Next-generation sequencing techniques may in the future enable pathogen identification using blood samples without the need for invasive respiratory sampling [109, 110].

In the last two decades, several ARDS pandemics have been due to emerging viral pathogens. The 2009 H1N1 pandemic was caused by a novel influenza A virus (H1N1pdm09 virus), and since then, the virus has circulated seasonally around the world. Novel coronavirus infections have been responsible for three pandemics causing Severe Acute Respiratory Syndrome (SARS), Middle East Respiratory Syndrome (MERS) and the ongoing...
COVID-19. The latter syndrome is caused by the SARS-CoV-2, which has 75–80% genomic similarity to the severe acute respiratory syndrome coronavirus (SARS-CoV), 50% to the Middle East Respiratory syndrome coronavirus (MERS-CoV), and 96% to a bat coronavirus [111–113]. COVID-19 was first reported in December 2019, in Wuhan, Hubei Province, China [111, 114, 115]. It then spread worldwide, infecting over 54 million people to date to become the largest pandemic in modern history.

### Table 2  Common, pandemic, and endemic pathogens responsible for ARDS and their features and risk factors

| **Common pathogens** | **Features and risk factors** |
|----------------------|------------------------------|
| *Streptococcus pneumoniae, Staphylococcus aureus, Haemophilus influenzae, Moraxella catarrhalis, Legionella pneumophila, Chlamydia pneumonia, Mycoplasma pneumonia* | Community-acquired pneumonia<br>Co-morbid conditions*<br>Influenza active in community (*Streptococcus pneumoniae, Staphylococcus aureus, Haemophilus influenzae*)<br>Cruise ships or resorts (*Legionella pneumophila*) |
| *Pseudomonas aeruginosa, Klebsiella pneumoniae, Escherichia coli, Acinetobacter species, Enterobacter species* | Hospital-acquired pneumonia<br>Co-morbid conditions*<br>Mechanical ventilation |
| Anaerobes | Aspiration |
| *Influenza A or B virus* | Influenza A can cause pandemics<br>Co-morbid conditions*<br>Influenza active in community |
| *Picornaviruses (rhinovirus, enterovirus), Human coronaviruses (229E, NL63, OC43, HKU1), Respiratory syncytial virus, Human metapneumovirus, Parainfluenza (1–4), Adenoviruses* | Frequently detected in critically ill patients with ARDS<br>May cause disease in the elderly and in patients with co-morbid conditions* |
| *Pneumocystis jirovecii* | HIV infection or other causes of immunosuppression |

**Pandemic pathogens**

| **SARS-CoV-2** | Pandemic situation<br>Exposure to patient with known COVID-19<br>Residence or travel in an area with active COVID-19 |

**Endemic pathogens**

| **Bacillus anthracis** (anthrax) | In context of bioterrorism |
| **Yersinia pestis** (pneumonic plague) | Residence in rural areas in Madagascar<br>Camping, hunting or contact with rodents |
| **Francisella tularensis** (tularemia) | Exposure to rabbits |
| **Chlamydia psittaci** (psittacosis) | Exposure to birds and poultry/poultry market |
| **Coxiella burnetti** (Q fever) | Exposure to farm animals or parturient cats |
| **Mycobacterium tuberculosis** | Residence in tuberculosis endemic areas<br>Co-morbid conditions: Alcoholism, injection drug use, HIV infection, immunosuppression |
| **Avian influenza A/H5N1, A/H5N6, A/H7N9 and other subtypes** | Residence or travel to Southeast and East Asia<br>Exposure to birds and poultry/poultry market |
| **MERS-CoV** | Residence or travel to the Arabian Peninsula<br>Exposure to dromedary camel |
| **Hantaviruses (e.g., Sin Nombre, Andes)** | Residence or travel to Western and Southwestern United States<br>Exposure to rodent excretions |
| **Measles virus** | Incomplete vaccination |
| **Human adenovirus type 55 (HAdV-55)** | Residence or travel to Southeast and East Asia |
| **Varicella-zoster virus** | Pregnancy<br>Immunosuppression |
| **Cytomegalovirus** | Immunosuppression |
| **Blastomyces dermatitidis, Histoplasma capsulatum, Coccidioides immitis** | Residence or travel to Ohio and Mississippi River valleys and Great Lakes (*Blastomyces dermatitidis, Histoplasma capsulatum*) or Western and southwestern United States (*Coccidioides immitis*)<br>Outdoor activities in wooded areas (*Blastomyces dermatitidis*) |
| **Plasmodium falciparum** | Residence or travel to malaria endemic areas |

*Co-morbid conditions include alcoholism, COPD, aspiration, pregnancy, bronchiectasis, injection drug use, HIV infection, and immunosuppression
Timing of interventions
Clinical trials have established the benefit of lung protective ventilation and a conservative strategy of fluid management in the treatment of ARDS [57, 70]. Pharmacologic therapies have failed to improve survival [17, 116–119], although there is evidence that dexamethasone is beneficial for COVID-19 ARDS and may have benefit in classical non-COVID-19 ARDS [32, 34, 120]. Given the lack of effective treatments once lung injury has progressed to ARDS, the approach to ARDS management has been reframed to also incorporate identification and treatment of patients at risk for ARDS or in the early stages of lung injury. There has been a shift towards categorizing patients into one of three stages of lung injury – presence of risk factors such as sepsis or pneumonia, early acute lung injury, and ARDS – and modifying their treatment accordingly (Fig. 4). Personalized approaches to ARDS trials or treatment will need to consider the timing of the intervention, as the clinical and biological phenotype of ARDS evolves rapidly over the first few hours to days of illness.

Prevention of lung injury
Since it was first hypothesized that ARDS could be prevented, some studies focused on preventing lung injury in at-risk patients. In theory, prevention might work if the therapeutic had a low risk of harm and might actually reduce the development of ARDS. It was thought that there might be a role for aspirin in preventing ARDS [121], but a phase 2b trial found that the use of aspirin did not reduce the risk of ARDS [122]. A recent randomized controlled trial showed no benefit of early vitamin D3 supplementation among critically ill, vitamin D-deficient patients [123]. The NHLBI-funded Prevention and Early Treatment of Acute Lung Injury (PETAL) network is currently enrolling patients with septic shock in a randomized phase 3 trial, Crystalloid Liberal or Vasopressors Early Resuscitation in Sepsis (CLOVERS), testing the impact of a restrictive fluids strategy (early vasopressors followed by rescue fluids) compared to a liberal fluid strategy (early fluids followed by rescue vasopressors) in patients with sepsis-induced hypotension (NCT03434028) on the development of ARDS and 28-day mortality. Another trial that may launch in the near future is Acetaminophen and Ascorbate in Sepsis:
Targeted Therapy to Enhance Recovery (ASTER) for testing the effects of acetaminophen and high dose intravenous vitamin C on both pulmonary and non-pulmonary organ dysfunction in early sepsis (NCT04291508). This trial may help to identify specific sepsis responsive phenotypes prior to the development of ARDS. For example, the trial will measure plasma free hemoglobin to test if it will enrich for the development of more organ failure, including ARDS, from early sepsis. Also, if acetaminophen shows efficacy especially in patients with elevated plasma hemoglobin, then this assay could be incorporated as a method for predictive enrichment in future trials that target the pro-oxidant effects of plasma free hemoglobin.

Treatment of early acute lung injury
Another growing interest has been to prevent progression of acute lung injury in patients who do not yet meet criteria for ARDS. Since the initial lung injury prevention score (LIPS) study, investigators have tried to identify early acute lung injury prior to the onset of respiratory failure that requires positive pressure ventilation [124]. A three-component early acute lung injury score incorporating oxygen requirement, respiratory rate, and immune suppression accurately identified patients who progressed to ARDS with an area under the curve of 0.86, a similar performance to the LIPS. Patients received one point for an oxygen requirement > 2–6 L/min or 2 points for an oxygen requirement > 6 L/min, one point each for a respiratory rate > 30/min and one point for immune suppression [125]. In this study, the median time of progression from meeting criteria for early acute lung injury to ARDS was 20 h, suggesting a window for intervention. In line with the concept of selecting patients in early sepsis at risk for ARDS by respiratory as well as hemodynamic criteria, the above discussed ASTER trial will enroll patients who require ≥ 6 L nasal oxygen or vasopressors for shock.

In 2015 a multicenter French trial compared high-flow nasal oxygen, non-invasive face mask ventilation, and standard oxygen therapy in patients in the emergency department with acute hypoxemic respiratory failure, primarily from pneumonia. The results indicated that high-flow nasal oxygen reduced the rate of endotracheal intubation in patients who presented with PaO2/FiO2 ≤ 200 mm Hg and reduced mortality in the entire cohort of 310 acutely hypoxemic patients with PaO2/FiO2 ≤ 300 mm Hg [126]. This was a pivotal trial that has changed clinical practice worldwide for critically ill patients. This is also an excellent example of how to test new interventions in the early phases of acute lung injury prior to the development of ARDS by Berlin criteria.

On a smaller scale, a Phase 2a randomized trial demonstrated the safety and feasibility of early administration of a combination of an inhaled corticosteroid and beta agonist in 61 patients at high risk of ARDS [127]. Treated patients demonstrated greater longitudinal improvement in SpO2/FiO2. An ongoing clinical trial, ARrest RESpiratory Failure From PNEUMONIA (ARREST PNEUMONIA), is testing the effects of a combination of an inhaled corticosteroid and a beta agonist in preventing progression to acute respiratory failure among patients who present with pneumonia and hypoxemia (NCT04193878). This therapeutic approach would likely have low value for established ARDS based on prior negative trials, but has more promise in the early phase of acute lung injury. Thus, personalized medicine needs to consider the optimal timing to test a new intervention.

Treatment of ARDS
There has also been promising new work to test therapies for patients with established ARDS. A phase 2b trial of high dose vitamin C in sepsis-induced ARDS demonstrated a significant difference in the primary outcome of Sequential Organ Failure Assessment score at 96 h [128]. In addition, the Vitamin C treated patients had a significant reduction in 28-day mortality and an increase in ICU-free days, and hospital-free days [16].

To enrich prognostically for the potential efficacy of neuromuscular blockade, the ACURASYS phase 3 trial required patients to be enrolled with moderate to severe hypoxemia (PaO2/FiO2 < 150 mmHg) and the results showed a lower mortality in the patients treated with neuromuscular blockade [38]. This prospective trial was designed on the basis of a phase 2 trial that suggested the value of this cut-off for a phase 3 trial [129]. However, the positive results of the phase 3 trial were not widely adopted and, therefore, a new trial, Reevaluation Of Systemic Early Neuromuscular Blockade (ROSE), was carried out to reassess this therapy. This phase 3 trial enrolled 1006 patients and found that there was no mortality benefit to early continuous neuromuscular blockade compared to usual care with lighter sedation goals in patients with moderate-to-severe ARDS [39]. The different results in the ROSE trial might be explained by the higher PEEP strategy, lighter sedation in the control group, and earlier enrollment in the more recent trial than in the ACURASYS trial [38, 39]. These two trials emphasize how phenotypes for personalized medicine can be selected on the basis of a physiologic abnormality, in this case the severity of arterial hypoxemia, when the goal is to test the therapy in a patient population most likely to respond to the treatment. In the future, prognostic enrichment for trials such as these may be amplified by including measures of lung compliance, radiographic
extent of pulmonary edema, and plasma biologic markers as well. As discussed below, these trials did not include patient groups to prove that non-responders would be those with a P/F > 150 mmHg, but they nevertheless did prospectively identify an ARDS population that might respond more effectively to the proposed therapy.

Conclusions and future challenges
Each of the sections in this review has identified several factors that contribute to heterogeneity in patients with ARDS. These variables include the associated clinical risk factor (pneumonia vs. non-pulmonary sepsis vs. trauma) or the combination of contributing clinical factors (blood transfusions, shock), the degree of physiologic lung injury graded by hypoxemia or impaired carbon dioxide excretion, variability in the extent of pulmonary opacities and lung edema on chest imaging, the range of biologic profiles as indicated by plasma biomarkers and gene expression in the blood, and the diverse number of pathogens that can initiate ARDS, including bacteria, viruses, fungi and parasites. How to best integrate these various dimensions of complexity into trials testing novel therapies for ARDS is understandably challenging, especially when one must consider whether the therapy will be tested in the early phase of lung injury prior to positive pressure ventilation, as was the case with high flow nasal oxygen, or once ARDS is established as in trials of neuromuscular blockade.

Importantly, different approaches to “personalization” will likely be required for different treatments; the key question will be how to best select potential patients for the therapeutic at hand. In many cases, it will not be feasible to determine the best personalization method in advance; particularly for trials of novel pharmacotherapies, counter-intuitive results may emerge [130], and/or our understanding of biology in specific phenotypes may be incomplete or frankly incorrect [131]. Instead, trials in broader ARDS populations will often need to be conducted first, with pre-specified plans for stratified analyses by phenotype, followed by dedicated smaller trials in phenotypes that seem to respond. Such an approach is consistent with US FDA guidance for enrichment in clinical trials, which stipulates that it is important to identify both responders and non-responders within a given condition before concluding that a therapy has benefits for a particular subgroup.

Clearly some therapies have been successful in randomized clinical trials without focusing on specific ARDS sub-groups. Lung protective ventilation [57] and fluid conservative therapy [70] were effective when applied to most patients with ARDS. However, these successes are the exception rather than the rule, and new approaches are increasing the feasibility of testing new therapies in a more personalized approach. For example, recent evidence indicates success with real time measurements of two plasma biomarkers in ARDS, making it more feasible to incorporate sub-phenotypes in clinical trials [90]. Physiologic variables such as the ventilatory ratio could be used to select patients for trials that had a high likelihood to improve pulmonary blood flow and reduce pulmonary dead space, similar to what was achieved in a trial of activated protein C for ARDS from non-sepsis causes in which pulmonary dead space was reduced with this anti-coagulant and anti-inflammatory therapy [132]. This approach could be used to assess the effects of therapies that target lung vascular injury and neutrophil endothelial complexes and reduce microvascular injury and enhance capillary blood flow to the lung [133, 134].

The current global pandemic of ARDS related to SARS-CoV-2 demonstrates the potential of a more focused approach to ARDS clinical trials. Corticosteroids appear to be effective for patients with COVID-19 associated ARDS, in contrast to many prior studies of corticosteroids for ARDS which had conflicting findings, demonstrating the value of enriching for a specific clinical phenotype [32]. In the future, progress with specific biologic therapeutics for ARDS such as statins or other anti-inflammatory treatments could benefit from enrichment with some of the variables considered in this review to maximize their chance of efficacy.

Author details
1 Department of Anesthesia, University of California San Francisco, San Francisco, CA, USA. 2 Cardiovascular Research Institute, University of California, San Francisco, USA. 3 Division of Pulmonary and Critical Care Medicine, Department of Medicine, University of California, San Francisco, USA. 4 King Saud Bin Abdulaziz University for Health Sciences and King Abdullah International Medical Research Center, Riyadh, Saudi Arabia. 5 Division of Allergy, Pulmonary and Critical Care, Vanderbilt University Medical Center, Nashville, TN, USA. 6 Department of Respiratory Medicine, Amsterdam University Medical Centers, Location AMC, University of Amsterdam, Infection and Immunity, Amsterdam, The Netherlands. 7 Department of Anesthesiology, Washington University, Saint Louis, MO, USA. 8 Division of Pulmonary, Allergy, and Critical Care Medicine, Center for Acute Respiratory Failure, Columbia University College of Physicians and Surgeons, New York, NY, USA. 9 School of Nursing, University of Pennsylvania, Philadelphia, PA, USA. 10 Department of Anesthesia and Critical Care, La Pitie Salpetriere Hospital, University Paris-Sorbonne, Paris, France. 11 Department of Medicine, Stanford University, Stanford, CA, USA.

Acknowledgements
This work was funded by the National Institutes of Health (Grant numbers HL123004, HL134828, STL1TR001871-05, HL14722, K23 HL133489, HL145506, UH3HL141736, HL140026, ST32GM08440-24, and HL126176).

Compliance with ethical standards
Conflicts of interest
The authors have no conflicts of interest for this manuscript.

Publisher's Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.
References

1. Matthay MA, Zemans RL, Zimmerman GA, Arabi YM, Beitler JR, Mercat A, Hemledge M, Randolph AG, Calfee CS (2019) Acute respiratory distress syndrome. Nat Rev Dis Primers 5:18

2. Beigel JH, Tomashek KM, Dodd LE, Mehta AK, Zingman BS, Kalil AC, Redd SC, Barzaoui G, Ansari A, El-Ramli M, Diaz M, Khairullah A, Koral L, Pique J, Reilly JP, Bellamy S, Shashaty MG, Galante R, Meyer NJ, Lanken PN, Kaplan S, Holena DN, May AK, Atmosphere LB, Christie JD (2017) Protein bio-synthesis in severe acute respiratory distress syndrome. N Engl J Med 366:2159–2166

3. Ware LB, Matthay MA, Mebazaa A (2020) Designing an ARDS trial for 2020 and beyond. Intensive Care Med. https://doi.org/10.1007/s00134-020-06232-x

4. Bellini G, Laffey JG, Pham T, Fan E, Brochard L, Esteban A, Gattinoni L, Khemani RG, Smith L, Lopez-Fernandez YM, Kwok J, Morzov R, Klein MJ, Vossoughi S, Gorlin J, Kessler DA, Hillyer CD, Van Buren NL, Jimenez A, Toy P, Gajic O, Bacchetti P, Looney MR, Gropper MA, Hubmayr R, Sheu CC, Gong MN, Zhai R, Bajwa EK, Chen F, Thompson BT, Christiani DC (2010) Clinical characteristics and outcomes of sepsis-related vs non-sepsis-related ARDS. Chest 138:S59–S67

5. Fowler AA III, Truwit JD, Hite RD, Morris PE, DeWilde C, Priday A, Fisher B, Thacker LR II, Natarajan R, Brophy DF, Sculthorpe R, Nanchal R, Syed A, Sturgill J, Martin GS, Sevansky J, Kashiouaris M, Hamman S, Egan KR, Hastings A, Spencer W, Trench S, Mehri O, Bindas J, Duggal A, Graf J, Zellner S, Yanny L, McPolin C, Hollrith T, Kramer D, Ojiole C, Damm T, Cassity E, Wieliczko A, Halquist M (2019) Effect of vitamin C infusion on organ failure and biomarkers of inflammation and vascular injury in patients with sepsis and severe acute respiratory failure: the CITRIS-ALI randomized clinical trial. JAMA 322:1261–1270

6. The National Heart, Lung, and Blood Institute ARDS Clinical Trials Network (2014) Rosuvastatin for sepsis-associated acute respiratory distress syndrome. N Engl J Med 370:2191–2200

7. Calfee CS, Delucchi K, Parsons PE, Thompson BT, Ware LB, Matthay MA, Network NA (2014) Subphenotypes in acute respiratory distress syndrome: latent class analysis of data from two randomised controlled trials. Lancet Respir Med 2:611–620

8. Reilly JP, Bellamy S, Shashaty MG, Galante R, Meyer NJ, Lanken PN, Kaplan S, Holena DN, May AK, Atmosphere LB, Christie JD (2014) Heterogeneous phenotypes of acute respiratory distress syndrome after major trauma. Ann Am Thorac Soc 11:728–736

9. Luo L, Shaver CM, Zhao Z, Koyama T, Calfee CS, Bastarache JA, Ware LB (2017) Clinical predictors of hospital mortality differ between direct and indirect ARDS. Chest 151:755–763

10. Calfee GR, Janz DR, Bernard GR, May AK, Kangelaris KN, Matthay MA, West JB, Zellner S, Torres A, Laffey JG, Hong Y (2019) Declining mortality in patients with acute respiratory distress syndrome: an analysis of the acute respiratory distress syndrome network trials. Crit Care Med 47:315–323

11. Esirmer D, Thompson T, Hudson LD, Luce JM, Hayden D, Schoenfeld D, Matthay MA, Acute Respiratory Distress Syndrome Network (2001) Efficacy of low tidal volume ventilation in patients with different clinical risk factors for acute lung injury and the acute respiratory distress syndrome. Am J Respir Crit Care Med 164:231–236

12. Ewuroke E, Major MJ, Gilbert McClinn LI (2018) National incidence rates for acute respiratory distress syndrome (ARDS) and ARDS cause-specific factors in the United States (2006–2014). J Crit Care 47:192–197

13. Birken DR, Halvachizadeh S, Pape HC, Pfeifer R (2020) Mortality of adult respiratory distress syndrome in trauma patients: a systematic review over a period of four decades. World J Surg 44:2243–2254

14. Eberhard LW, Morabito DJ, Matthay MA, Mackersie RC, Campbell AR, Marks JD, Atwood JA, Pittet JF (2000) Initial severity of metabolic acidosis contributes to the pathogenesis of human re-expansion pulmonary edema. Chest 118:559–567

15. Lu K, Stoller JK, Mekala SR, Johns J, Calfee CS, Ewigman B, Mehta A, Wittich GR, May AK, Wang Q, Nishimura J, Arora A, Phillips DKW, Mehta R, Kau AL, Nunn JF, G.Choi WS, Liu T, Duggal A, Zheng S, Knetemann RB, Young HI, DeStefano S, Saha SK, Ghusinga V, Ranganathan H, Blom N, Dickson LT, Paganin V, Macdougall C, Wang G, Sturdivant D, Gelfond J, Burt KM, Shovlin CL, Smith JC, Zhang J, Schrier RW, Mehta A, Kreller K, Zellner S, Torres A, Vosburgh K, Lavoie K, Weger W, Zayed H, Trinh V, Bajwa EK, Chen F, Thompson BT, Christiani DC (2010) The influence of infection sites on development and mortality of ARDS. Intensive Care Med 36:963–970

16. Lewis SR, Pritchard MW, Thomas CM, Smith AF (2019) Pharmacological agents for adults with acute respiratory distress syndrome. Cochrane Database Syst Rev 7:CD004477
34. Villar J, Ferrando C, Martínez D, Ambros A, Munoz T, Soler JA, Aguilar G, Alba F, Gonzalez-Higuera E, Conesa LA, Martín-Rodriguez C, Diaz-Dominguez FJ, Serna-Grande P, Rivas R, Ferreiras J, Belda J, Capilla L, Tallett A, Anon JM, Fernandez RL, Gonzalez-Martín JM (2020) Desmethylamfetamine treatment for the acute respiratory distress syndrome: a multicentre, randomised controlled trial. Lancet Respir Med 8:267–276

35. Mattey MA, Wick KD (2020) Corticosteroids, COVID-19 pneumonia and acute respiratory distress syndrome. J Clin Investig. https://doi.org/10.1172/JCI143331

36. ARDS Definition Task Force, Ranieri VM, Rubenfeld GD, Thompson BT, Ferguson ND, Caldwell E, Fan E, Camporota L, Slutsky AS (2012) Acute respiratory distress syndrome—The Berlin definition. JAMA 307:2562–2533

37. Ferguson ND, Fan E, Camporota L, Antonelli M, Anzueto A, Beale R, Brochard L, Brower R, Esteban A,Gattinoni L, Rhodes A, Slutsky AS, Vincent JL, Rubenfeld GD, Thompson BT, Ranieri VM (2012) The Berlin definition of ARDS: an expanded rationale, justification, and supplementary material. Intensive Care Med 38:1573–1582

38. Papazian L, Forel JM, Gacouin A, Penot-Ragon C, Perrin G, Loundou A, Jaber S, Arnal JM, Perez D, Sibbeyhyan JM, Constantin JM, Courant P, Lefant JY, Guerin C, Prat G, Morange S, Roch A, ACURASYS Study Investigators (2010) Neuromuscular blockers in early acute respiratory distress syndrome. N Engl J Med 363:1107–1116

39. The National Heart, Lung and Blood Institute PETAL Clinical Trials Network (2019) Early neuromuscular blockade in the acute respiratory distress syndrome. JAMA 321:846–857

40. Beitler JR, Sarge T, Banner-Goodspeed VM, Gong MN, Cook D, Novack V, Loring SH, Talmor D, EPVent-2 Study Group (2019) Effect of titrating positive end-expiratory pressure (PEEP) with an esophageal pressure-guided strategy via an empirical high PEEP-Fio2 strategy on death and days free from mechanical ventilation among patients with acute respiratory distress syndrome: a randomized clinical trial. JAMA 321:846–857

41. Villar J, Perez-Mendez L, Blanco J, Anon JM, Blanch L, Belda J, Santos-Bouza A, Fernandez RL, Kacmarek RM, Spanish Initiative for Epidemiology, Stratification, and Therapies for ARDS (SIESTA) Network (2013) A universal definition of ARDS: the PaO2/Fio2 ratio under a standard ventilatory setting—a prospective, multicenter validation study. Intensive Care Med 39:583–592

42. Villar J, Perez-Mendez L, Lopez J, Belda J, Blanco J, Saralegui I, Suarez-Sipmann F, Lopez J, Libulbo S, Kacmarek RM, HELP Network (2007) An early PEEP/Fio2 trial identifies different degrees of lung injury in patients with acute respiratory distress syndrome. Am J Respir Crit Care Med 176:795–804

43. Nuckton TJ, Alonso JA, Kallet RH, Daniel BM, Pittet JF, Daniel BM, Pittet JF, Matthay MA (2006) Esophageal and transpulmonary pressures in acute respiratory distress syndrome. Am J Respir Crit Care Med 176:795–804

44. Nuckton TJ, Alonso JA, Pittet JF, Matthay MA (2004) Prognostic value of the pulmonary dead-space fraction during the first 6 days of acute respiratory distress syndrome. Am J Respir Crit Care Med 176:795–804

45. Maze M, Huth SJ, Gaudreault D, Hynan LS, Huter JW, Forster AR, Gurosko KS, Hoff DL, Kim HS, Mehta S, Phillips BH, Pagan SJ, Parry AR, Remick GD, Ryan JL, Rubenstein MJ, Schmidt RA, Silverstein PD, Soltesz M, Tang VC, Varney B, Wells CA, Wilson WR, Wood KE, Ye CX, Younès A, Zwerling C, Zhang Y, Zvezdoski LJ, Dorey R, Nuckton TJ, Gaudreault D (2012) An early PEEP/Fio2 trial identifies different degrees of lung injury in patients with acute respiratory distress syndrome: proceedings from the pediatric acute respiratory distress syndrome incidence and epidemiology (PARDIE) V1 Investigators and the Pediatric Acute Lung Injury Consensus Conference Group (2015) Pediatric acute respiratory distress syndrome: definition, incidence, and epidemiology.
programs are distinct in acute respiratory distress syndrome. Am J Respir Crit Care Med 197:528–532
97. Kao KC, Chiu LC, Hung CY, Chang CH, Huang CC, Hu HC (2017) Coinfection and mortality in pneumonia-related acute respiratory distress syndrome patients with bronchoalveolar lavage: a prospective observational study. Shock 47:615–620
98. Metlay JP, Waterer GW, Long AC, Anzueto A, Brozek J, Coles G, Cooley LA, Dean NC, Fine MJ, Flanders SA, Griffin MR, Metersky ML, Musher DM, Restrepo MI, Whitney CG (2019) Diagnosis and treatment of adults with community-acquired pneumonia. An official clinical practice guideline of the American Thoracic Society and Infections Diseases Society of America. Am J Respir Crit Care Med 200:e45–e67
99. Kalic AC, Metersky ML, Compas M, Muscedere J, Sweeney DA, Palmer Gadsby NJ, Russell CD, McHugh MP, Mark H, Conway Morris A, Lauren-Arbabi YM, Fowler R, Hayden FG (2020) Critical care management of severe pneumonia: a review. The Lancet 395:119–125
100. Hasvold J, Sjoding M, Pohl K, Cooke C, Hyzy RC (2016) The role of biomarkers in respiratory infections, using metagenomic sequencing. PLoS ONE 7:e30875
101. Fischer N, Rohde H, Indenbirken D, Gunther T, Reumann K, Lutgether M, Leo S, Gaïa N, Ruppé E, Emonet S, Girard M, Lazarevic V, Schrenzel J, Madi N, Al-Nakib W, Mustafa AS, Habibi N (2018) Metagenomic analysis of viral diversity in respiratory samples from patients with respiratory tract infections, using metagenomic sequencing. PLoS ONE 7:e30875
102. Lindberg AM, Persson B, Allander T, Andersson B (2012) Characterization of the viral microbiome in patients with severe lower respiratory tract infections, using metagenomic sequencing. PLoS ONE 7:e30875
103. Lysholm F, Wetterbom A, Lindau C, Darban H, Bjerkner A, Fahlander K, Langelier C, Fung M, Caldera S, Deiss T, Lyden A, Prince BC, Serpa PH, Belzer A, Bolourchi S, Caldera S, Fung M, Jauregui A, Malcolm K, Lyden A, Khan L, Vessel K, Quan J, Zinter M, Chiu CY, Chow ED, Wilson J, Miller A, Grundhoff P (2014) Rapid metagenomic diagnostics for suspected outbreak of severe pneumonia. Emerg Infect Dis 20:1072–1075
104. Hasvold J, Spinding M, Pohli K, Cooke C, Hyzy RC (2016) The role of human metagenomics in the critically ill adult patient. J Crit Care 31:233–237
105. Fischer N, Rohde H, Indenbirken D, Gunther T, Reumann K, Lutgether M, Leo S, Gaïa N, Ruppé E, Emonet S, Girard M, Lazarevic V, Schrenzel J (2017) Detection of Bacterial Pathogens from Broncho-Alveolar Lavage by Next-Generation Sequencing. Int J Mol Sci. 18(9):2011. https://doi.org/10.3390/ijms18092011
106. Madi N, Al-Nakib W, Mustafa AS, Habibi N (2018) Metagenomic analysis of viral diversity in respiratory samples from patients with respiratory tract infections, using metagenomic sequencing. PLoS ONE 7:e30875
107. Lysholm F, Wetterbom A, Lindau C, Darban H, Bjerkner A, Fahlander K, Langelier C, Fung M, Caldera S, Deiss T, Lyden A, Prince BC, Serpa PH, Belzer A, Bolourchi S, Caldera S, Fung M, Jauregui A, Malcolm K, Lyden A, Khan L, Vessel K, Quan J, Zinter M, Chiu CY, Chow ED, Wilson J, Miller A, Grundhoff P (2014) Rapid metagenomic diagnostics for suspected outbreak of severe pneumonia. Emerg Infect Dis 20:1072–1075
108. Hasvold J, Spinding M, Pohli K, Cooke C, Hyzy RC (2016) The role of human metagenomics in the critically ill adult patient. J Crit Care 31:233–237
109. Fischer N, Rohde H, Indenbirken D, Gunther T, Reumann K, Lutgether M, Leo S, Gaïa N, Ruppé E, Emonet S, Girard M, Lazarevic V, Schrenzel J (2017) Detection of Bacterial Pathogens from Broncho-Alveolar Lavage by Next-Generation Sequencing. Int J Mol Sci. 18(9):2011. https://doi.org/10.3390/ijms18092011
110. Madi N, Al-Nakib W, Mustafa AS, Habibi N (2018) Metagenomic analysis of viral diversity in respiratory samples from patients with respiratory tract infections, using metagenomic sequencing. PLoS ONE 7:e30875
111. Lysholm F, Wetterbom A, Lindau C, Darban H, Bjerkner A, Fahlander K, Langelier C, Fung M, Caldera S, Deiss T, Lyden A, Prince BC, Serpa PH, Belzer A, Bolourchi S, Caldera S, Fung M, Jauregui A, Malcolm K, Lyden A, Khan L, Vessel K, Quan J, Zinter M, Chiu CY, Chow ED, Wilson J, Miller A, Grundhoff P (2014) Rapid metagenomic diagnostics for suspected outbreak of severe pneumonia. Emerg Infect Dis 20:1072–1075
112. Hasvold J, Spinding M, Pohli K, Cooke C, Hyzy RC (2016) The role of human metagenomics in the critically ill adult patient. J Crit Care 31:233–237
113. Lysholm F, Wetterbom A, Lindau C, Darban H, Bjerkner A, Fahlander K, Langelier C, Fung M, Caldera S, Deiss T, Lyden A, Prince BC, Serpa PH, Belzer A, Bolourchi S, Caldera S, Fung M, Jauregui A, Malcolm K, Lyden A, Khan L, Vessel K, Quan J, Zinter M, Chiu CY, Chow ED, Wilson J, Miller A, Matthay MA, Pollard KS, Calfee CS, DeRisi JL (2018) Systematic analysis of viral diversity in respiratory samples from patients with respiratory tract infection diagnosis in critically ill adults. Proc Natl Acad Sci USA 115:E21353–E21362
114. Hong DK, Blauwkamp TA, Kertesz M, Becovici S, Tsuang C, Banaei N (2018) Liquid biopsy for infectious diseases: sequencing of cell-free plasma to detect pathogen DNA in patients with invasive fungal disease. Diagn Microbiol Infect Dis 92:210–213
115. Langelier C, Kalantr KL, Moazed F, Wilson MR, Crawford ED, Deiss T, Belzer A, Bolouchi S, Caldera S, Fung M, Jauregui A, Malcom K, Lyden A, Khan L, Vessel K, Quan J, Zinter M, Chiu CY, Chow ED, Wilson J, Miller A, Matthay MA, Pollard KS, Christenson S, Calfee CS, DeRisi JL (2018) Integrating host response and unbiased microbe detection for lower respiratory tract infection diagnosis in critically ill adults. Proc Natl Acad Sci USA 115:E21353–E21362
116. Hong DK, Blauwkamp TA, Kertesz M, Becovici S, Tsuang C, Banaei N (2018) Liquid biopsy for infectious diseases: sequencing of cell-free plasma to detect pathogen DNA in patients with invasive fungal disease. Diagn Microbiol Infect Dis 92:210–213
117. Langelier C, Kalantr KL, Moazed F, Wilson MR, Crawford ED, Deiss T, Belzer A, Bolouchi S, Caldera S, Fung M, Jauregui A, Malcom K, Lyden A, Khan L, Vessel K, Quan J, Zinter M, Chiu CY, Chow ED, Wilson J, Miller A, Matthay MA, Pollard KS, Christenson S, Calfee CS, DeRisi JL (2018) Integrating host response and unbiased microbe detection for lower respiratory tract infection diagnosis in critically ill adults. Proc Natl Acad Sci USA 115:E21353–E21362
118. Hong DK, Blauwkamp TA, Kertesz M, Becovici S, Tsuang C, Banaei N (2018) Liquid biopsy for infectious diseases: sequencing of cell-free plasma to detect pathogen DNA in patients with invasive fungal disease. Diagn Microbiol Infect Dis 92:210–213
119. Langelier C, Kalantr KL, Moazed F, Wilson MR, Crawford ED, Deiss T, Belzer A, Bolouchi S, Caldera S, Fung M, Jauregui A, Malcom K, Lyden A, Khan L, Vessel K, Quan J, Zinter M, Chiu CY, Chow ED, Wilson J, Miller A, Matthay MA, Pollard KS, Christenson S, Calfee CS, DeRisi JL (2018) Integrating host response and unbiased microbe detection for lower respiratory tract infection diagnosis in critically ill adults. Proc Natl Acad Sci USA 115:E21353–E21362
127. Festic E, Carr GE, Cartin-Ceba R, Hinds RF, Banner-Goodspeed V, Bansal V, Asuni AT, Talmor D, Rajagopalan G, Frank RD, Gajic O, Matthay MA, Levitt JE (2017) Randomized clinical trial of a combination of an inhaled corticosteroid and beta agonist in patients at risk of developing the acute respiratory distress syndrome. Crit Care Med 45:798–805
128. Fowler AA 3rd, Fisher BJ, Kashouriis MG (2020) Vitamin C for sepsis and acute respiratory failure-reply. JAMA 323:792–793
129. Gainnier M, Roch A, Forel JM, Thirion X, Arnal JM, Donati S, Papazian L (2004) Effect of neuromuscular blocking agents on gas exchange in patients presenting with acute respiratory distress syndrome. Crit Care Med 32:113–119
130. Meyer NJ, Reilly JP, Anderson BJ, Palakhappha JA, Jones TK, Dunn TG, Shashaty MG, Feng R, Christie JD, Opal SM (2018) Mortality benefit of recombinant human interleukin-1 receptor antagonist for sepsis varies by initial interleukin-1 receptor antagonist plasma concentration. Crit Care Med 46:21–28
131. Sinha P, Matthay MA, Calfee CS (2020) Is a "Cytokine Storm" Relevant to COVID-19? JAMA Intern Med 180:1152–1154
132. Liu KD, Levitt J, Zhuo H, Kallet RH, Brady S, Steingrub J, Tidswell M, Siegel MD, Soto G, Peterson MW, Chesnutt MS, Phillips C, Weinacker A, Thompson BT, Eisner MD, Matthay MA (2008) Randomized clinical trial of activated protein C for the treatment of acute lung injury. Am J Respir Crit Care Med 178:618–623
133. Barnes BJ, Adrover JM, Baxter-Stoltzfus A, Borczuk A, Cools-Lartigue J, Crawford JM, Dassler-Plenker J, Guerci P, Huynh C, Knight JS, Loda M, Looney MR, McAllister F, Rayes R, Renaud S, Rousseau S, Salvatore S, Schwartz RE, Spicer JD, Yost CC, Weber A, Zuo Y, Egeblad M (2020) Targeting potential drivers of COVID-19: Neutrophil extracellular traps. J Exp Med 217
134. Ackermann M, Verleden SE, Kuehnel M, Haverich A, Welte T, Laenger F, Vanstapel A, Werlein C, Stark H, Tzankov A, Li WW, Li VW, Mentzer SJ, Jonigk D (2020) Pulmonary vascular endotheliitis, thrombosis, and angiogenesis in Covid-19. N Engl J Med 383:120–128