Quantitative DNA methylation analyses reveal stage dependent DNA methylation and association to clinico-pathological factors in breast tumors

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Abstract

Background: Aberrant DNA methylation of regulatory genes has frequently been found in human breast cancers and correlated to clinical outcome. In the present study we investigate stage specific changes in the DNA methylation patterns in order to identify valuable markers to understand how these changes affect breast cancer progression.

Methods: Quantitative DNA methylation analyses of 12 candidate genes \textit{ABCB1}, \textit{BRCCA1}, \textit{CDKN2A}, \textit{ESR1}, \textit{GSTP1}, \textit{IGF2}, \textit{MGMT}, \textit{HMLH1}, \textit{PPP2R2B}, \textit{PTEN}, \textit{RASSF1A} and \textit{FOXC1} was performed by pyrosequencing a series of 238 breast cancer tissue samples from DCIS to invasive tumors stage I to IV.

Results: Significant differences in methylation levels between the DCIS and invasive stage II tumors were observed for six genes \textit{RASSF1A}, \textit{CDKN2A}, \textit{MGMT}, \textit{ABCB1}, \textit{GSTP1} and \textit{FOXC1}. \textit{RASSF1A}, \textit{ABCB1} and \textit{GSTP1} showed significantly higher methylation levels in late stage compared to the early stage breast carcinoma. Z-score analysis revealed significantly lower methylation levels in DCIS and stage I tumors compared with stage II, III and IV tumors. Methylation levels of \textit{PTEN}, \textit{PPP2R2B}, \textit{FOXC1}, \textit{ABCB1} and \textit{BRCA1} were lower in tumors harboring \textit{TP53} mutations than in tumors with wild type \textit{TP53}. \textit{Z}-score analysis showed that \textit{TP53} mutated tumors had significantly lower overall methylation levels compared to tumors with wild type \textit{TP53}. Methylation levels of \textit{RASSF1A}, \textit{PPP2R2B}, \textit{GSTP1} and \textit{FOXC1} were higher in \textit{ER} positive vs. \textit{ER} negative tumors and methylation levels of \textit{PTEN} and \textit{CDKN2A} were higher in \textit{HER2} positive vs. \textit{HER2} negative tumors. \textit{Z}-score analysis also showed that \textit{HER2} positive tumors had significantly higher \textit{z}-scores of methylation compared to the \textit{HER2} negative tumors. Univariate survival analysis identifies methylation status of \textit{PPP2R2B} as significant predictor of overall survival and breast cancer specific survival.

Conclusions: In the present study we report that the level of aberrant DNA methylation is higher in late stage compared with early stage of invasive breast cancers and DCIS for genes mentioned above.

Keywords: Breast cancer, DNA methylation, Methylation index, Stage, \textit{TP53}
Background

Breast cancer is the most common form of malignant disease in women worldwide and also the principal cause of death from cancer among women globally. In Norwegian women breast cancer accounts for approximately 23% of all cancers and ~800 women die as a result of the disease every year. Breast cancer is a heterogeneous disease with distinct histopathological, genetic and epigenetic characteristics. Epigenetic regulation is critical for normal growth and development and provides a layer of transcriptional control. Epigenetic alterations which occur in transformed cells involve changes in DNA methylation including global hypomethylation and focal hypermethylation, histone modifications and nucleosomal remodeling [1]. Epigenetic changes are considered to be an early event in tumor development and one of the hallmarks of cancer [2]. The degree of DNA methylation in the promoter region of tumor suppressor genes, transcription factors and drug response genes may play a role in the initiation of cancer, tumor progression and response to treatment. Identification of early epigenetic changes in breast cancer might give valuable markers for early detection and contribute to the understanding of how these changes affect the progression of the disease and prognosis for the patient. We previously described treatment-specific DNA methylation patterns in 432 CpGs in the promoter regions of 14 genes in samples from 75 patients with locally advanced breast cancer treated with doxorubicin [3]. Further, we identified four novel genes (ABCB1, FOXC1, PPP2R2B and PTEN) that were found to be already aberrantly methylated in DCIS (Ductal carcinoma in situ), a pre-invasive stage of breast cancer [4] and that were also found to be methylated in the locally advanced breast cancers [3]. These findings raised the question if DNA methylation patterns evolve between the different stages of breast cancers and pre-neoplastic lesions or are similar and independent of tumor stage. In the present study, using the same candidate gene approach as previously described [3,4], we performed a large-scale analysis of 12 candidate genes and determined quantitatively the DNA methylation patterns of the 12 genes in 238 breast cancer patients of all stages from early premalignant DCIS to advanced metastatic disease. In addition to the analysis of stage dependent DNA methylation patterns, associations between additional clinico-pathological factors of breast cancer such as grade and ER status and DNA methylation patterns of these 12 genes were assessed.

Methods

Patient material

A total of 238 samples were included in present study: 75 patients with locally advanced breast cancer that were enrolled in a prospective study evaluating predictive factors for response to doxorubicin [3] and 35 patients treated with 5-FU and Mitomycin for locally advanced breast cancer [5], patients in both cohorts were admitted to the Haukeland University Hospital in Norway between 1991 and 2001; 57 samples from a series of 212 breast cancers samples collected from Ullevål University Hospital (Norway) between 1990 and 1994 [6]; 71 tumors from a population-based cohort including 854 women diagnosed between 1986 and 2004 in Uppsala, Sweden with either: a) pure ductal carcinoma in situ of the breast (DCIS), b) an invasive breast cancer, 15 mm or less, without an in situ component or c) a mixed lesion, i.e., a lesion with both an invasive- and an in situ component [4]. Clinical and molecular characteristics of the tumors are given in Table 1. DNA from six normal breast tissues was included to identify the DNA methylation baseline in normal tissues. Normal breast tissue was obtained from women who underwent a biopsy of the mammary gland because of mammographic screening and for whom histology confirmed the presence of only normal tissue. All patients had given informed consent, and the project was approved by the local ethical committee.

Methylation assays

DNA concentrations were determined using the Quant-iT™ dsDNA broad range assay kit (Invitrogen, Cergy Pontoise, France) and normalized to a concentration of 50 ng/μl. One μg of DNA was bisulphite converted using the MethylEasy™ HT Kit for Centrifuge (Human Genetic Signatures, North Ryde, Australia) according to the manufacturer's instructions. Quantitative DNA methylation analysis of the bisulphite treated DNA was performed by pyrosequencing or - in case of several sequencing primers - by serial pyrosequencing [7]. Oligonucleotides for PCR amplification and pyrosequencing (Additional file 1) were synthesized by Biotez (Buch, Germany) [3]. In the present study, same candidate gene approach was used as previously described [3,4] with the difference in number of covered CpGs (205 in our case) because of absence of variability. These genes were initially selected on the following basis: previous reports of DNA methylation in breast tumors or at least breast cancer cell lines (ABCB1 [8], BRCA1 [9], CDKN2A [9], ESR1 [10], GSTP1 [11], IGF2 [12], MGMT [9], MLH1 [9], PPP2R2B [13], PTEN [14], RASSF1A [15]) and genes displaying variation in breast cancer gene expression profiles (FOXC1 [16]).

Statistical analysis

The average value of methylation for all CpGs in a target region was calculated for each sample and each gene. Although there was some stochastic variation between different CpG positions, the overall methylation level was quite constant and CpG positions were highly correlated in the analyzed regions. A sample was considered 1)
hypermethylated if the percentage of DNA methylation was higher than the sum of two times the standard deviation and mean of the normal samples, 2) normal-like methylation if the% DNA methylation was in range of two times the standard deviation +/− mean of the normal sample and 3) hypomethylated if% DNA methylation was lower than two times standard deviation – mean of normal sample. The aggregated quantitative DNA methylation data is presented in Table 2. Differences in the presence of methylation were determined by a two-sided Fisher test (for variables with two categories) and Chi-squared tests for variables with three or more categories. Odds ratio and 95% confidence intervals were calculated for two-categorical variables. Differences in the distribution of methylation were assessed by the non-parametric Mann–Whitney test (on parameters with 2 categories) or the Kruskal-Wallis test analysis on parameters with more than two categories. All obtained p-values were corrected with the Bonferroni correction method in which the p-values are multiplied by the number of comparisons. The methylation index of samples (Z-score) was calculated as: (methylation level of each sample – mean

### Table 1 Clinical characteristics of the analyzed samples (Continued)

| Molecular subtypes | N(%) |
|--------------------|------|
| Luminal A          | 52(30.8%) |
| Luminal B          | 34(20.1%) |
| ERRB2              | 29(17.2%) |
| Basal              | 37(21.9%) |
| Normal             | 17(10%)  |
| Unknown            | 69     |

Unknown data was excluded from total when the percentage was calculated.

### Table 2 Quantitative methylation data

| Gene       | Mean of samples | SD of samples | Mean of normal | SD of normal |
|------------|-----------------|---------------|----------------|--------------|
| BRCA1      | 85.36           | 11.32         | 85.48          | 6.15         |
| RASSF1A    | 30.92           | 17.34         | 3.37           | 0.99         |
| PTEN       | 5.15            | 4.07          | 2.83           | 1.28         |
| PPP2R2B    | 10.83           | 9.80          | 3.18           | 1.17         |
| CDKN2A     | 4.68            | 4.64          | 3.58           | 1.08         |
| MLH1       | 2.94            | 3.85          | 2.72           | 0.47         |
| MGMT       | 4.21            | 3.53          | 4.48           | 1.11         |
| ABCB1      | 16.18           | 17.25         | 2.63           | 1.23         |
| KSF2       | 38.28           | 9.13          | 40.7           | 2.41         |
| GSTP1      | 17.18           | 17.27         | 3.73           | 1.01         |
| FOXC1      | 13.41           | 13.23         | 3.6            | 1.98         |
| ESR1       | 2.92            | 2.50          | 4.43           | 2.94         |
The sum for the 12 genes was calculated giving one single value (Z-score) for each sample. The false discovery rate was not considered in this study due to small number of genes which were tested. All calculations were performed using Statistical Package for Science version 18.

Univariate, Kaplan-Meier analyses and the log-rank test for each parameter for single gene was performed to

| GENE      | Methylation status | Normal | % Sample number | DCIS | % Sample number | Invasive | % Sample number | I | % Sample number | II | % Sample number | III | % Sample number | IV | % Sample number |
|-----------|-------------------|--------|-----------------|------|-----------------|----------|-----------------|---|-----------------|----|-----------------|-----|-----------------|----|-----------------|
|           |                   | N      |                 |      |                 |          |                 |   |                 |    |                 |     |                 |    |                 |
| ABCB1     | Hypomethylated    | 0      | 0               | 0    | 0               | 0        | 0               |   |                 |    |                 |     |                 |    |                 |
|           | Normal like       | 6      | 100             | 15   | 57.7            | 33       | 52.4            | 6  | 30              | 31  | 36              | 6   | 30              |    |                 |
|           | Hypermethylated   | 0      | 11              | 42.3 | 20              | 30       | 47.6            | 14 | 70              | 57  | 64              | 14  | 70              |    |                 |
| BRCA1     | Hypomethylated    | 0      | 0               | 0    | 2               | 3        | 10              | 10 | 12.8            | 1   | 5.9             |     |                 |    |                 |
|           | Normal like       | 6      | 100             | 26   | 100             | 63       | 97              | 18 | 90              | 68  | 87.2            | 16  | 94.1            |    |                 |
|           | Hypermethylated   | 0      | 0               | 0    | 0               | 0        | 0               |    |                 |    |                 |     |                 |    |                 |
| CDKN2A    | Hypomethylated    | 0      | 0               | 0    | 0               | 0        | 0               |    |                 |    |                 |     |                 |    |                 |
|           | Normal like       | 6      | 100             | 26   | 100             | 60       | 92.3            | 18 | 90              | 61  | 67.8            | 14  | 70              |    |                 |
|           | Hypermethylated   | 0      | 0               | 0    | 5               | 7.7      | 2               | 10 | 29              | 32.2| 6               | 30  |                 |    |                 |
| ESR1      | Hypomethylated    | 0      | 0               | 0    | 0               | 0        | 0               |    |                 |    |                 |     |                 |    |                 |
|           | Normal like       | 6      | 100             | 26   | 100             | 63       | 97              | 19 | 95              | 63  | 98.4            | 19  | 100             |    |                 |
|           | Hypermethylated   | 0      | 0               | 0    | 2               | 3        | 1               | 5  | 1               | 1.6 | 0               |     |                 |    |                 |
| FOXC1     | Hypomethylated    | 0      | 0               | 0    | 0               | 0        | 0               |    |                 |    |                 |     |                 |    |                 |
|           | Normal like       | 6      | 100             | 6    | 23              | 34       | 52.3            | 7  | 35              | 40  | 46.5            | 10  | 47.6            |    |                 |
|           | Hypermethylated   | 0      | 20              | 77   | 31              | 47.7     | 13              | 65 | 46              | 53.5| 11              | 52.4|                 |    |                 |
| GSTP1     | Hypomethylated    | 0      | 0               | 3.8  | 0               | 0        | 0               |    |                 |    |                 |     |                 |    |                 |
|           | Normal like       | 6      | 100             | 10   | 38.5            | 27       | 41.5            | 5  | 25              | 33  | 5               | 5   | 25              |    |                 |
|           | Hypermethylated   | 0      | 15              | 57.7 | 38              | 58.5     | 15              | 75 | 59              | 67  | 15              | 75  |                 |    |                 |
| IGF2      | Hypomethylated    | 0      | 0               | 11   | 42.3            | 25       | 38.4            | 6  | 30              | 34  | 39.1            | 9   | 47.4            |    |                 |
|           | Normal like       | 6      | 100             | 14   | 53.8            | 32       | 49.2            | 9  | 45              | 32  | 36.8            | 5   | 26.3            |    |                 |
|           | Hypermethylated   | 0      | 0               | 1    | 3.9             | 8        | 12.4            | 5  | 25              | 21  | 24.1            | 5   | 26.3            |    |                 |
| MGMT      | Hypomethylated    | 0      | 0               | 0    | 13              | 20.3     | 13              | 65 | 16              | 24.2| 8               | 44.4|                 |    |                 |
|           | Normal like       | 6      | 100             | 25   | 96.1            | 50       | 78.1            | 5  | 25              | 35  | 53.1            | 5   | 27.7            |    |                 |
|           | Hypermethylated   | 0      | 1               | 3.9  | 1                | 11.6     | 2               | 10 | 15              | 22.7| 5               | 27.7|                 |    |                 |
| MLH1      | Hypomethylated    | 0      | 0               | 4    | 15.4            | 12       | 18.5            | 6  | 32              | 4   | 4.5             | 3   | 17.6            |    |                 |
|           | Normal like       | 6      | 100             | 21   | 80.8            | 47       | 72.3            | 10 | 53              | 68  | 77.3            | 13  | 76.5            |    |                 |
|           | Hypermethylated   | 0      | 1               | 3.8  | 6               | 9.2      | 3               | 16 | 16              | 18.2| 1               | 5.9 |                 |    |                 |
| PPP2R2B   | Hypomethylated    | 0      | 0               | 0    | 0               | 0        | 0               |    |                 |    |                 |     |                 |    |                 |
|           | Normal like       | 6      | 100             | 7    | 26.9            | 15       | 24.6            | 6  | 35              | 40  | 30.8            | 3   | 16.7            |    |                 |
|           | Hypermethylated   | 0      | 19              | 73.1 | 46              | 75.4     | 11              | 65 | 45              | 69.2| 15              | 83.3|                 |    |                 |
| PTEN      | Hypomethylated    | 0      | 0               | 0    | 0               | 0        | 0               |    |                 |    |                 |     |                 |    |                 |
|           | Normal like       | 6      | 100             | 21   | 80.8            | 56       | 86.2            | 13 | 65              | 23  | 27.7            | 9   | 45              |    |                 |
|           | Hypermethylated   | 0      | 5               | 19.2 | 9               | 13.8     | 7               | 35 | 60              | 72.3| 11              | 55  |                 |    |                 |
| RASSF1A   | Hypomethylated    | 0      | 0               | 2    | 7.7             | 2        | 3.1             | 1  | 0               |     |                 |    |                 |    |                 |
|           | Normal like       | 6      | 100             | 2    | 7.7             | 10       | 15.4            | 2  | 11              | 3   | 4.8             | 1   | 5.6             |    |                 |
|           | Hypermethylated   | 0      | 22              | 84.6 | 53              | 81.5     | 17              | 90 | 60              | 95.2| 17              | 94.4|                 |    |                 |

Samples were considered as hypermethylated if the% DNA methylation was higher than the sum of two times the standard deviation and mean of the normal samples and hypomethylated if% DNA methylation was lower than two times the standard deviation - mean of the normal samples.
investigate which genes and parameters affect survival. Further, multivariate, the Cox proportional hazard model was used to identify independent prognostic markers for all genes and from all clinical parameters: age, stage, tumor size and grade, lymph node status, TPS3 mutation status, ER, PR status, T status. Methylation status of all genes was treated as continuous and categorical. We constructed possible model candidates using all combinations of given variables. To select the best-fitted model from all candidates, we evaluated the Akaike Information Criterion (AIC) [17]. AIC gives an evaluation for model selection, which is modified by a penalty increasing with the number of variables of the model. Analyzing all combinations of given variables we selected the model that fitted best to the data indicated by a minimal value for the AIC.

Results

Methylation analysis and correlation with clinico-pathological parameters: stage and grade

A total of 48790 epigenotypes were generated through analyses of 205 CpGs in 12 genes (ABCB1 (20 CpGs), BRCA1 (19 CpGs), CDKN2A (28 CpGs), ESR1 (21 CpGs), FOXC1 (9 CpGs), GSTP1 (21 CpGs), IGF2 (17 CpGs), MGMT (9 CpGs), MLH1 (16 CpGs), PPP2R2B (14 CpGs), PTEN (19 CpGs) and RASSF1A (12 CpGs)). Six normal samples were used to estimate the normal-like methylation levels for all analyzed genes. Our analysis showed that five genes ABCB1, FOXC1, GSTP1, PPP2R2B and RASSF1A were the most frequently hypermethylated genes in all invasive samples as well as in the DCIS samples. PTEN was hypermethylated in invasive cancer of stage II, III and IV and MGMT was hypomethylated in invasive tumors of stage II, III and IV. CDKN2A had a normal-like methylation level in a high percentage of the DCIS samples and early stage tumors. In late stage tumors CDKN2A showed higher percentage of hypermethylated samples compared to the early stage tumors and the DCIS. The methylation levels of ESR1 and MLH1 were normal-like both in the DCIS and the invasive tumors and BRCA1 had a normal-like methylation level in almost all the DCIS and all invasive tumors (Table 3).

Significant differences in methylation levels between the DCIS and invasive stage II tumors were observed for six genes RASSF1A, CDKN2A, MGMT, ABCB1, GSTP1 and FOXC1 (p = 0.008, p = 0.005, p = 0.003, p = 0.006, p = 0.010, p = 0.010 respectively). RASSF1A, ABCB1 and GSTP1 showed significantly higher quantitative methylation levels in late stage compared to the early stage breast carcinoma. The most significant differences in methylation levels for these three genes were between stage I and III (p = 0.001, p = 0.022, p = 0.019 respectively) and between stage I and IV (p = 3.3e-6, p = 0.030 and p = 0.014 respectively). PTEN and CDKN2A methylation levels were low and increased in late stage III and IV (p = 0.003, p = 0.004 between stage I and III, p = 0.018 and p = 0.003 between stage I and IV), while MGMT methylation levels were low and appear to decrease with tumor stage (p = 4.5e-4 between stage I and IV) (Figure 1). After correction for multiple testing (Bonferroni correction), differences in methylation levels for RASSF1A between stage I and III and stage I and IV remained significant. Absolute differences in mean methylation levels for RASSF1A were higher than 10%. For PTEN, CDKN2A, MGMT methylation levels between stage I and
III and stage I and IV remained significant after correction. Differences in methylation levels between the DCIS and invasive stage II tumors for MGMT also reached statistical significance after correction. Absolute differences in mean methylation levels between different stages for PTEN, CDKN2A, MGMT were less than 3% even though they remained significant after Bonferroni correction. We next combined all the methylation data into a single variable, the methylation index, to investigate further, how methylation levels change during progression of breast cancer. At the same time we wanted to investigate the presence of general pattern which might be more robust than single genes. As shown in Figure 2, we observed significantly lower methylation levels in DCIS and stage I samples compared to stage II, III and IV samples (p = 3e-7). Significant differences in methylation levels were observed between the normal breast tissue and stage II, III and IV tumors (p = 0.001, p = 0.006, p = 0.009 respectively). Normal breast tissue showed lower levels of methylation compared with tumors mentioned above. There was no significant difference observed between the normal samples and the DCIS and stage I tumors. Also

Figure 2 Boxplots illustrating significant association between methylation status and tumor stage and grade. (a) Significantly lower methylation levels in DCIS and stage I samples compared to stage II, III and IV samples was observed. (b) Also grade 1 tumors had significantly lower methylation index compared with grade 2 and grade 3 tumors.
grade 2 and 3 tumors had significantly higher Z-scores than grade 1 tumors (p = 0.022). Methylation index analysis showed that Luminal A and Luminal B tumors had significantly higher Z-scores than Basal-like tumors (p = 0.007).

Correlation with TP53 mutations and hormone receptor status

We compared the DNA methylation profiles with the TP53 mutations status and found that tumors with TP53 mutations had significantly lower DNA methylation levels then tumors with TP53 wild type in RASSF1A, PTEN, PPP2R2B, FOXC1, ABCB1 and BRCA1 (p = 0.028, p = 0.031, p = 0.002, p = 0.017, p = 0.010, p = 0.001 respectively). After Bonferroni correction DNA methylation levels in PPP2R2B and BRCA1 were still significantly lower in tumors with TP53 mutations. Significant associations with (ER) estrogen receptor status were observed for RASSF1A, PPP2R2B, GSTP1 and FOXC1 methylation levels (p = 0.004, p = 0.012, p = 0.012, p = 0.032, respectively). HER2 receptor status was associated with RASSF1A, PTEN, MGMT, CDKN2A and ESR1 (p = 0.023, p = 3.6e-7, p = 1.1e-8, p = 5.8e-9, p = 0.017) and after Bonferroni correction PTEN, MGMT and CDKN2A remained significant. No significant association with (PR) progesterone receptor status was observed (Table 4). Mann–Whitney test revealed that ER and HER2 negative tumors had lower methylation levels compared with ER and HER2 positive tumors for all studied genes. Interestingly HER2 negative tumors had higher methylation levels of ESR1 compared with HER2 positive tumors (p = 0.017). Z-score analysis showed that TP53 mutated tumors had significantly lower overall methylation levels compared to tumors with wild type TP53. Also HER2 positive tumors had significantly higher z-scores of methylation compared to the HER2 negative tumors (Figure 3). There was no significant association with PR and ER status.

Survival analysis

To investigate which parameters contribute to differences in survival we applied: 1) univariate analysis using Kaplan-Meier modeling and the log-rank test for each gene and each clinical parameter and 2) multivariate analysis using Cox hazard proportional model to all variables. To select the best-fitted model from all model candidates, we evaluated the Akaike Information Criterion [17].

Univariate survival analysis identified methylation status of PPP2R2B as significant predictor of overall survival. As expected, grade, estrogen receptor status, TP53 status and stage also appeared as significant predictors of survival. The Kaplan-Meier plot (Figure 4) showed a significant difference in survival between hypermethylated and normal-like samples for PPP2R2B (p = 0.012) indicating that patients with hypermethylated genes had better survival. Breast cancer specific survival was significantly improved in patients with hyper-methylated promoters for PPP2R2B (p = 0.012).

Further, multivariate survival analysis was performed in order to investigate if any of the methylation markers were independent prognostic markers, using both, categorical and continuous methylation data. We constructed possible model candidates using all relevant parameters and methylation data. Using calculated AIC, the best model that fits to the survival data was assessed. For categorical methylation data AIC identified a model explaining survival, which included the methylation status of IGF2, GSTP1, estrogen receptor status, TP53, N status and stage. The estimated coefficients, the hazard ratio, the p-values, and the 95% confidence intervals of the hazard ratio were summarized in Table 5. For continuous methylation data the best model explaining survival included TP53, T status, N status, estrogen receptor status and methylation status of IGF2 and GSTP1. The estimated coefficients, the hazard ratio, the p-values, and the 95% confidence intervals of the hazard ratio were summarized in Table 6. For both, categorical and continuous methylation data IGF2 and GSTP1 (p = 0.009 and p = 0.014) were significant together with ER status (p = 0.008), TP53 status (p = 6.6e-5) and N status (p = 0.047). For these models, we also described statistical significance for the likelihood ratio, Wald and log-rank tests in the Tables 5 and 6.

Discussion

The aim of this study was to quantitatively determine the methylation levels in the promoter region of 12 genes in breast cancer patients of all stages and to investigate stage specific changes in tumors. In addition, we wanted to evaluate the association between clinico-pathological
factors (ER, HER2 status, grade) including survival and methylation levels of these genes.

In the present study, five genes, \textit{ABCB1}, \textit{FOXC1}, \textit{GSTP1}, \textit{PPP2R2B} and \textit{RASSF1A} were hypermethylated already in early stage breast cancer (stage I and II). For all five genes hypermethylation was also detected in DCIS suggesting that inactivation of these genes is a frequent event in the process of mammary tumorigenesis. We found that \textit{RASSF1A} was hypermethylated in approximately 85% of all invasive tumors and DCIS, and our results are in agreement with such a high incidence of \textit{RASSF1A} methylation [18-20]. \textit{RASSF1A} is a putative tumor-suppressor gene. It belongs to an increasing list of tumor suppressor genes that are frequently inactivated by promoter methylation rather than by somatic mutations [21]. Since we detected a constant hypermethylation of \textit{RASSF1A} in all of the different stages of the breast carcinomas we can suggest that hypermethylation of \textit{RASSF1A} is an early event during breast cancer pathogenesis and also the main mechanism of inactivation. In our study, \textit{GSTP1} was found to be hypermethylated in different stages of breast carcinomas, for early stages (I and II) our results are in agreement with previous reports [22,23]. The frequency of \textit{GSTP1} promoter hypermethylation in stage III and IV (around 70%) was found to be higher than reported previously [23,24]. It is known that \textit{GSTP1} plays a role in detoxification of potential carcinogens and that loss of the expression of \textit{GSTP1} will lead to DNA damage of breast cells and they will be more easily exposed to carcinogens [25]. Loss of \textit{GSTP1} expression and its

\begin{figure}[h]
\centering
\includegraphics[width=\textwidth]{figure3.png}
\caption{Boxplots illustrating significant association between methylation status and HER2 and TP53 status. (a) TP53 mutated tumors had significantly lower overall methylation levels compared to tumors with wild type TP53. (b) HER2 positive tumors had significantly higher methylation index compared to the HER2 negative tumors.}
\end{figure}
potential role in breast carcinogenesis was observed in high proportion of breast tumors [23]. It appears that promoter hypermethylation is associated with loss of GSTP1 expression [26]. Important question is when the promoter hypermethylation of the GSTP1 gene starts to play a role in tumor progression? We found that already in DCIS there is a high proportion of hypermethylated GSTP1 (58%), which indicates that GSTP1 promoter hypermethylation is an early event in breast carcinogenesis. From our analysis we observed that hypermethylation of ABCB1, PPP2R2B and FOXC1 is also an early event in breast carcinogenesis since our results indicate high level of hypermethylation of these genes already in DCIS which was reported before [4]. We found frequencies of methylation for CDKN2A MGMT and MLH1 similar to previously published reports [27,28]. According to our results, we could suggest that hypermethylation of CDKN2A is possible event leading to its inactivation in late stage breast cancers. PTEN is a tumor suppressor gene. Its product PTEN protein works as a negative regulator of the Akt pathway, leading to suppression of apoptosis and increasing cell survival [29]. We suggest here that epigenetic silencing of PTEN might be an early event in initiation of cancer and also the mechanism of its inactivation. Next, combining all the methylation data into a single variable, the methylation index, we investigate how methylation levels change during progression of breast cancer. Our analysis showed that the methylation pattern of all genes included in this study is changing during breast cancer progression. DCIS and stage I tumors had similar

**Table 5 Multivariate survival analysis - categorical methylation data**

| Covariate | Baseline | Coefficient(b_i) | HR(exp(b_i)) | p-value | 95.0%CI for Exp(B) |
|-----------|----------|------------------|--------------|---------|-------------------|
| TP53      | TP53 wild type | −2.332           | 0.097        | 6.6e-5  | (0.030-0.305)     |
| ER        | ER positive | 1.481            | 4.401        | 0.008   | (1.460-13.269)    |
| Stage II  | Stage I   | 2.260            | 9.584        | 0.013   | (1.593-57.667)    |
| Stage III | Stage I   | −0.011           | 0.988        | 0.992   | (0.088-11.019)    |
| N1        | N0        | −6.362           | 0.529        | 0.338   | (0.143-1.947)     |
| N2        | N0        | 1.673            | 5.332        | 0.047   | (1.019-27.885)    |
| IGF2      | Normal like IGF2 | 1.597           | 4.940        | 0.009   | (1.472-16.581)    |
| GSTP1     | Normal like GSTP1 | −1.621          | 0.197        | 0.014   | (0.053-0.729)     |
| PPP2R2B   | Normal like PPP2R2B | −0.972          | 0.378        | 0.085   | (0.125-1.143)     |
| CDKN2A    | Normal like CDKN2A | −0.839          | 0.432        | 0.133   | (0.144-1.294)     |

The variables that the minimum AIC selected. (p-values: 3.4e-7 for likelihood ratio test, 3.4e-4 for Wald test, 3.2e-6 for log-rank test). Positive hazard ratios indicate an increased risk of dying from breast cancer and are calculated for the different covariates.

Figure 4 Kaplan-Meier plots of overall survival for patients with normal-like or hyper methylated PPP2R2B promoter. The p value was calculated using a long-rank test.
methylation levels with no significant difference. Stage II tumors showed the most significant difference in methylation levels when compared with DCIS and stage I and then in stage III and IV methylation levels were lower but still significantly higher than in DCIS and stage I. It remains unclear how this process of dramatic change in stage II is achieved and driven further during breast cancer progression. To our knowledge only one group investigate stage dependent DNA methylation in breast cancer using DCIS and all four stages of invasive tumors [30]. They identified 33 cancer specific genes that were either highly methylated in early stage breast cancer or showed stage dependent methylation pattern, lower methylation frequency in early stage breast cancers and a higher methylation frequency in late stage breast cancers. None of our selected genes were methylated in this study. Additional studies of the relationship between DNA methylation in tumors and tumor stages are necessary.

In the present study we showed the associations between DNA methylation levels of candidate genes and the TP53 mutation status, estrogen receptor status, and HER2 status. The TP53 tumor suppressor gene has a central role in cell cycle regulation, DNA repair and apoptosis, and a large number of reports have discussed the important role of TP53 alterations in breast cancer. Also, a number of studies have shown that breast tumors with TP53 mutations are strongly associated with poor prognosis and lacking methylation in a number of regulatory genes [31,32]. Additionally, studies on different expression subtypes in breast cancer showed that different subtypes have a different underlying biology reflected in methylation and is strongly influenced by TP53 mutation status. It was shown that basal-like tumors are TP53 mutated and unmethylated [6,33]. In present study we have identified 26.6% of breast tumors with TP53 mutations and significantly lower levels of DNA methylation in RASSF1A, PTEN, PPP2R2B, FOXC1, ABCB1 and BRCA1 compared to tumors with wild type TP53. Since in our study we had low number of basal-like tumors (37 out of 238) we cannot confirm association between TP53 mutated and unmethylated tumors and basal-like but most of the basal-like samples were TP53 mutated and had lowest methylation levels compared to other subtypes (data not shown). The status of ER and HER2 have been recognized as important prognostic factors in patients with breast cancer, in addition to a predictive marker for the response to treatment with endocrine and trastuzumab therapy. Identification of genes with subtype-specific methylation revealed that RASSF1A and GSTP1 were highly methylated in Lum B tumors [33]. These two genes were reported previously to be significantly more methylated in ER- positive than ER-negative tumors [34]. In the present study we showed the same trend in ER-positive tumors compared to ER-negative for four genes and RASSF1A and GSTP1 were among them. The same group reported that HER2 positive tumors had higher methylation level for these two genes compared to HER2-negative tumors, which is again in accordance with our study. Furthermore, in a study on methylation in breast cancer and breast cancer molecular subtypes it was shown that RASSF1A is hypermethylated in HER2 positive breast tumors (ERBB2 and luminal B) [35]. In our study RASSF1A was hypermethylated in ERBB2 and luminal B tumors (data not shown). Taken all together, these results, suggest that methylation plays a significant role in the different breast tumor phenotypes.

We report here for the first time the PPP2R2B methylation status as significant predictor for breast cancer survival as well as for overall survival. PPP2R2B is a candidate tumor suppressor gene and it was shown that changes in DNA methylation of this gene contribute to its expression [36]. Further, multivariate analysis showed that IGF2 and GSTP1 were independent prognostic markers. Recently, it has been shown that the absence of GSTP1 protein expression correlate with promoter

### Table 6 Multivariate survival analysis - continuous methylation data

| Covariate | Baseline | Coefficient (b_i) | HR(exp(b_i)) | p-value | 95.0% CI for Exp(B) |
|-----------|----------|------------------|-------------|---------|-------------------|
| GSTP1     | Normal like GSTP1 | −0.066          | 0.935       | 4e-4    | (0.902-0.970)     |
| IGF2      | Normal like IGF2  | 0.079           | 1.082       | 0.002   | (1.031-1.136)     |
| MLH1      | Normal like MLH1  | −0.552          | 0.575       | 0.135   | (0.278-1.187)     |
| ER        | ER positive      | 1.659           | 5.256       | 0.002   | (1.804-15.317)    |
| TP53      | TP53 wild type   | −2.169          | 0.114       | 0.002   | (0.028-0.450)     |
| N 1       | N0               | 1.684           | 5.391       | 0.022   | (1.265-22.975)    |
| N 2       | N0               | 1.706           | 5.511       | 0.032   | (1.151-26.371)    |
| T 2       | T1               | 1.937           | 6.942       | 0.035   | (1.143-42.157)    |
| T3        | T1               | 0.050           | 1.051       | 0.960   | (0.143-7.716)     |
| T4        | T1               | 1.125           | 3.081       | 0.258   | (0.437-21.692)    |

The variables that the minimum AIC selected. (p-values: 1.2e-7 for likelihood ratio test, 5.8e-4 for Wald test, 6.7e-6 for log-rank test).

Positive hazard ratios indicate an increased risk of dying from breast cancer and are calculated for the different covariates.
hypermethylation and with improved survival in invasive breast cancer samples [37]. Hypermethylation of GSTP1 is a well established biomarker for hormone dependent cancers. Our previous analyses suggest that methylation of GSTP1 in locally advanced breast cancer patients treated with doxorubicin was associated to survival [3]. However, GSTP1 had no effect on treatment response.

Conclusions
Here we report aberrant methylation levels of ABCB1, FOXC1, GSTP1, PPP2R2B and RASSF1A in DCIS and stage I-IV providing evidence that suggests that changes in methylation level is an early event and may also be important in progression to later stages of breast cancer. We also report that methylation levels of important breast cancer genes are associated to hormone receptor status and TP53 mutation status suggesting mechanisms of deactivation of tumor suppressor genes in breast cancer. Further studies are necessary to identify which methylation gene profiles are of predictive and which of prognostic value.

Additional file

Additional file 1: PCR and pyrosequencing primers. Sequences of primers used for amplification and pyrosequencing reactions, Genbak accession numbers and nucleotides (Nt) corresponding to the amplified fragments as well as the annealing temperatures for the respective PCR amplifications. CpGs are numbered in the order of appearance from the 5’ end of an amplification product. Y = pyrimidine.

Competing interests
The authors declare that they have no competing interests.

Authors’ contribution
JK performed data analyses and wrote the manuscript. ED performed laboratory experiments. TF, HE and HS were involved in the statistical analyses. PEL and FW were responsible for the patient cohorts and IB for the laboratory experiments. TF, HE and HS were involved in the statistical analyses. PEL and FW were responsible for the patient cohorts and IB for the laboratory experiments. TF, HE and HS were involved in the statistical analyses. PEL and FW were responsible for the patient cohorts and IB for the laboratory experiments.

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