Strict major histocompatibility complex molecule class-specific binding by co-receptors enforces MHC-restricted αβ TCR recognition during T lineage subset commitment

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Since the discovery of co-receptor dependent αβ TCR recognition, considerable effort has been spent on elucidating the basis of CD4 and CD8 lineage commitment in the thymus. The latter is responsible for generating mature CD4 helper and CD8αβ cytotoxic T cell subsets. Although CD4$^+$ and CD8$^+$ T cell recognition of peptide antigens is known to be MHC class II- and MHC class I-restricted, respectively, the mechanism of single positive (SP) thymocyte lineage commitment from bipotential double-positive (DP) progenitors is not fully elucidated. Classical models to explain thymic CD4 vs. CD8 fate determination have included a stochastic selection model or instructional models. The latter are based either on strength of signal or duration of signal impacting fate. More recently, differential co-receptor gene imprinting has been shown to be involved in expression of transcription factors impacting cytotoxic T cell development. Here, we address commitment from a structural perspective, focusing on the nature of co-receptor binding to MHC molecules. By surveying 58 MHC class II and 224 MHC class I crystal structures in the Protein Data Bank, it becomes clear that CD4 cannot bind to MHC I molecules, nor can CD8αβ or CD8αα bind to MHC II molecules. Given that the co-receptor delivers Lck to phosphorylate exposed CD3 ITAMs within a peptide/MHC (pMHC)-ligated TCR complex to initiate cell signaling, this strict co-receptor recognition fosters MHC class-restricted SP thymocyte lineage commitment at the DP stage even though both co-receptors are expressed on a single cell. In short, the binding preference of an αβ TCR for a peptide complexed with an MHC molecule dictates which co-receptor subsequently binds, thereby supporting development of that subset lineage. How function within the lineage is linked further to biopotential fate determination is discussed.

Keywords: TCR, co-receptor, lineage commitment, structure, thymocyte development

CO-RECEPTORS: THEIR HISTORY AND FUNCTION

Two major subsets of human T lymphocytes were distinguished in the 1980s by surface expression of CD8 and CD4 as defined by monoclonal antibodies. These were shown to represent cytotoxic and helper T lymphocytes, respectively (1–4). Analysis of T cell clones revealed that CD8$^+$ T cells were MHC class I restricted whereas CD4$^+$ T cells were MHC class II restricted in their TCR recognition. The involvement of CD4 and CD8 in antigen-specific T cell recognition, despite their invariant structures, suggested that CD4 and CD8 might function as co-receptors in cognate recognition (4). In such a model, the co-receptors interact with conserved segments of MHC molecules (CD8 with MHCI and CD4 with MHCII) whereas the αβ TCR recognizes a specific peptide bound to a polymorphic segment of the same MHC molecule. Subsequent structural studies over the last two decades validated this hypothesis, revealing the bidentate interaction of a TCR and a co-receptor with the peptide/MHC (pMHC) in a trimolecular complex (5–10).

Given that recent reviews have highlighted detailed structures of the co-receptors (11, 12), these shall not be reviewed herein. Instead, features relevant to the biologic function of co-receptors in the mature peripheral T cell and thymocyte compartments are highlighted.

The CD8 transmembrane co-receptor is encoded by two distinct genes: CD8α and CD8β. Each consists of a single Ig-like domain followed by a lengthy stalk region of 30–50 residues with multiple O-glycosylated adducts, a TM helix, and a short cytoplasmic tail [reviewed in Ref. (13)]. The CD8α but not CD8β tail binds to Lck, essential for T cell signaling. Both CD8αα homodimers and CD8αβ heterodimers are found on the surface of lymphocytes, with the CD8αβ heterodimer being the dominant isoform expressed on CTL. (14). The CD8αα isoform is expressed on γδ T cells, some NK cells, and a subset of intraepithelial lymphocytes (15). By contrast to CD8, CD4 comprises four Ig-like domains in tandem with a short stalk region and TM helix, but its cytoplasmic
tail also binds Lck. In fact, a zinc clasp tethers Lck to the cytoplasmic tail of both CD4α and CD8α [Ref. (16) and references therein]. The major function of the co-receptor in T cell-mediated adaptive responses is to deliver Lck into the TCR-pMHC interacting system so that exposed ITAM(s) on one or more of the CD3 tails can be phosphorylated on tyrosine residues. This phosphorylation allows Zap-70 recruitment and the remainder of downstream signaling apparatus to assemble (17).

The affinity of CD4 for pMHCII is extremely weak (200 μM or higher) (18) and that of CD8αβ for pMHC1 only slightly stronger (6, 19). By contrast, ~1 μM affinities of TCR-pMHC interactions are not uncommon (20). Thus, the half-life of TCR-pMHC is ~1000 times longer than that of CD4-pMHC; with little binding to pMHC contributed by the co-receptor ectodomain per se. The bidentate interaction of TCR and CD8αβ with a single agonist pMHC has been studied by a micropipette adhesion assay (21). Kinetic analysis reveals a two stage cooperative process with the first stage representing TCR dominant binding to pMHC. The second stage binding, delayed by 1 s, is Src-tyrosine kinase-dependent (i.e., presumably Lck) resulting in the CD8αβ co-receptor binding to the TCR-engaged pMHC molecule. This time delay may relate to a required reorientation of the pMHC upon TCR ligation to foster CD8 binding and/or other intracellular molecular events. This ordered and cooperative trimeric interaction favors agonist ligands and synergistically augments the bidentate binding to pMHC in turn linked to T cell signaling. Although not formally studied as yet, it appears likely that a similar cooperativity in binding processes will be operative for CD4 and TCR.

**THYMOCYTE DEVELOPMENT**

αβTCR signaling is essential for adaptive immune responses as well as thymocyte differentiation (22–28). However, in early thymic development, the first major checkpoint is referred to as β-selection. There, TCRβ chain gene rearrangement occurs in the absence of TCRA chain gene rearrangements such that an expressed TCRβ chain associates with the invariant pre-Tα (pTα), forming a pre-TCR on the cell surface (28). Signaling through the pre-TCR terminates subsequent TCRβ locus rearrangements, rescues cells from apoptosis and induces massive proliferation. This signaling process enables CD4+ CD8α− double-negative (DN) cells to differentiate into CD4+CD8+ double-positive (DP) cells, facilitating TCRA gene rearrangements and thus generating a large αβTCR repertoire among DP thymocytes (28). DP thymocytes undergo positive selection and negative selection following self-MHC interaction via their αβTCR. During positive selection, DP cells mature into single positive (SP) CD4+ or CD8+ thymocytes and may egress into the thymic medulla. After 2 weeks or so of maturation therein, these SP cells migrate out to the periphery (29, 30). During negative selection, strongly self-reactive thymocytes undergo cell death. Those apoptotic thymocytes are purified from the T lineage repertoire [reviewed in Ref. (31–33)].

One remarkable feature of thymic development is the presence of the numerically major subpopulation of DP thymocytes expressing both CD4+ and CD8+ co-receptors. This DP phenotype is absent from human and murine peripheral T lineages (34). As Lck is critical for TCR activation and physically associated with both CD4+ and CD8α and DP thymocytes express both of these co-receptors, it follows that co-receptor-based pMHC interaction will impose MHC-restricted recognition on the mature αβTCR repertoire. This was recently tested in an elegant experiment with knockout mice lacking both co-receptors and MHC class I and MHC class II proteins. In such “Quattro” knockout mice, Lck is not sequestered away from the TCR so that TCR functions in the absence of co-receptor engagement. Hence, TCR signaling is mediated through non-MHC-ligands. Such αβTCRs possess antibody-like recognition specificities (35, 36).

While the CD4 ectodomain is a rigid concanamer of four Ig-like domains with no known developmentally regulated glycosylation and hence, “tuneability,” the structural features of CD8 are quite different (13). The combined presence of O-glycans and prolines in the CD8 subunit stalk suggests that these membrane connectors adopt extended and somewhat stiff conformations, similar to that observed for leukosialin and mucins, nonetheless with some conformational flexibility (37, 38). This CD8αβ co-receptor flexibly likely accommodates to the variability of TCR pMHC docking onto pMHC [reviewed in Ref. (11)]. In addition, developmentally regulated glycosylation of the CD8α stalk modulates pMHC binding (13). Immature DP thymocytes bind MHC more avidly than CD8 SP thymocytes. This differential binding is governed by developmentally programed O-glycan modification of several CD8 stalk threonine residues proximal to the CD8β headpiece and controlled by ST3Gal-I sialyltransferase (13, 39). ST3Gal-I specifically localizes to the medulla of the thymus where SP thymocytes reside. ST3Gal-I induction and attendant core 1 sialic acid addition to CD8α on mature thymocytes decreases CD8αβ-MHC1 avidity by altering CD8αβ domain-domain association and/or orientation. Hence, glycans on the CD8β stalk appear to modulate the ability of the distal binding surface of the dimeric CD8 globular head domains to clamp MHC. The DP stage facilitates efficient elimination by negative selection of autoreactive TCR specificities through this enhanced co-receptor function working in tandem with highly specific TCR-pMHC1 triggered apoptosis (24). Once a thymocyte has differentiated to the CD8 SP stage, however, CD8αβ O-glycan sialylation reduces the strength of the CD8αβ co-receptor interaction with pMHC1, mandating a greater requirement for TCR-pMHC1 interaction to achieve a subsequent activation threshold in mature T lineage cells (13). At the DP thymocyte stage, the higher avidity CD8αβ interaction with pMHC1 will further enforce MHC-restricted αβTCR recognition via Lck delivery.

**LINEAGE FATE**

The differentiation of DP thymocytes into CD4+ or CD8+ SP thymocytes is a fundamental example of bipotential cell fate determination. How this process is controlled during positive selection events in the thymus has been the subject of intense scrutiny [reviewed in Ref. (40)]. A stoichiastic selection model that postulates that co-receptor gene expression occurs randomly has been largely discredited (41). Instructional models postulating that TCR signals during positive selection specifically direct termination of irrelevant co-receptors have been controversial. One early strength of signal model was popular because the CD4 cytoplasmic tail manifest stronger affinity for Lck than that of CD8α, engendering TCR/CD4 co-engagement signals stronger than that of TCR/CD8.
However, genetic manipulation of ITAMs within the TCR complex did not change CD4 vs. CD8 lineage choice (44). A more recent variant of the instructional model is a duration of γ signaling with more recent common cytokine receptor incorporation (45). The kinetic signaling model proposed by Singer et al. incorporates aspects of this signaling duration model in conjunction with more recent common cytokine receptor γ-chain (γC) signaling biology (40, 46).

In the kinetic signaling model, positive selection and lineage choice are sequential events. Positive selection induces all DP thymocytes, regardless of TCR specificity, to transiently terminate CD8 gene expression resulting in CD4+ CD8dim thymocytes, in part by suppressing E8βIII Cdh enhancer activity (47). Persistent TCR engagement blocks IL-7 signaling and fosters CD4+ SP development with loss of CD8. In contrast, disruption of TCR engagement permits IL-7 signaling and results in “co-receptor reversal” with CD8 SP development [reviewed in Ref. (47)]. IL-7 signaling induces RUNX3 transcription that initiates CD8 expression via the E8βIII Cdh enhancer and also activates the cytotoxic gene transcriptome in CD8 T cells. The Cdh gene locus cis elements control thymocyte lineage fate with respect to the cytotoxic vs. helper lineage program (47).

The above thymocyte biology now needs to be interpreted in a structural context as relates to co-receptor and pMHC interactions in order to understand bipotential DP thymocyte cell fate determination in the DP to SP developmental transition. Here we offer insights into this process with respect to MHC-restricted lineage development and linkage to mature T cell function.

CONSERVED GEOMETRY OF MHCI α2/β2 DOMAINS FOR CD4-BINDING

Structures of CD4/MHCI as well as CD8/MHCI complexes have been solved. These reveal two very different binding modes. CD4 solely uses its N-terminal domain for insertion into a hydrophobic cave-like structure formed by the two membrane-proximal domains of the MHCI molecule (α2/β2). In contrast, CD8 uses its two dimeric Ig-like domains to clamp onto the CD loop of the MHCI α3 domain in a fashion similar to an antigen (5–7).

We have systematically surveyed herein 58 MHCI structures and 224 MHCII structures in the protein data bank (PDB). The structure of human CD4 in complex with HLA-DR1 (PDB code 3SSL) was taken as a reference (48). All other MHC structures were overlaid onto HLA-DR1 within this 3SSL structure. The superposition was based on the α2 domain in the case of MHCI molecules or the corresponding β2M domain in the case of MHCII molecules. Figure 1 depicts this overlay of selected MHC molecules onto HLA-DR1 in the CD4/HLA-DR1 complex. The representative MHCI molecules from human or mouse all preserve an angle between the two membrane-proximal α2/β2 domains that can readily accommodate CD4-binding (Figure 1A). On the contrary, Figures 1B,C show two selected MHCII molecules overlaid, which have largest (1AGF) or smallest (1QVO) opening angle between respective membrane-proximal β2M/α3 domains. Clearly the MHCI molecules’ geometry appears to prevent CD4-binding.

We have superimposed a representative group of MHCII molecules onto mouse 1-Aβ in the complex structure of D10 TCR/1-Aβ (1D9K) based on the two membrane-proximal α2/β2 domains, several of which are pictured in Figure 1A. A larger set includes human HLA-DR1/TCR complex (1FYT), HLA-DRA/TCR complex (1YMM), HLA-DR2a/TCR complex (1ZGL), HLA-DQ8 (1JK8) as well as mouse 1-Aβ/TCR complex (1U3H), I-Ek (1IEA), and CD4/I-Ak complex (1JL4). The RMSD value for 178 overlaid Ca atoms ranges from 0.63 to 1.1 Å, indicating that the two membrane-proximal domains as a structural unit have a conserved relative angle. The same calculation was carried out for MHCI molecules. A representative group of MHCI molecules was superimposed onto H2-Kb in the H2-Kb/CD8α complex structure (1BQH) based on the two membrane-proximal β2M/α3 domains. These include human HLA-A (1HH), HLA-B (1AGB), HLA-C (1IM9), HLA-A/TCR complex (1A07), HLA-A2/CD8α complex (1AKJ), as well as mouse H2-Kb/TCR complex (2CKB) and mouse H2-Db (1QLF). The RMSD value for about 180 Ca atoms ranges from 0.73 to 1.49 Å, indicating that the two membrane-proximal domains as a structural unit also have a relatively fixed angle. Notably, the RMSD value for MHCI is slightly larger than that of MHCI, consistent with previous observation that MHCI α3 domain has a small angular variation relative to the rest of MHCI molecule (5). We further compared the separation angles of the two membrane-proximal domains between the two classes of MHC molecules. We took the class II molecule HLA-DQ8 (1JK8) and class I molecule H2-Kb in its CD8 complex (1BQH) for comparison. If the MHCI β2 domain is superimposed onto the MHCI α3 domain, the rest of the MHCI molecule has about a 38° rotation relative to the counterpart of MHCI. This manifests as a larger opening between α2 and β2 domains in MHCI compared to between β2M and α3 in MHC, a geometric requirement for CD4-binding as shown in Figure 1B. In that figure, MHCI α2 domain and MHCI β2M domain were overlaid in the back. To accommodate CD4, the red-colored front β2 domain of MHCI molecule DR1 appears to “swing” leftwards compared to its counterparts α3 domain of MHCI molecules HLA-A (in yellow) and HLA-B (in blue).

Figure 2 shows the conserved domain-domain interface interaction in the MHCI1 molecules HLA-DQ8 (1JK8). Figure 2A is the overview of the structure, whereas the Figure 2B is a zoomed-in view of the α2/β2 domain interface revealing atomic interaction in detail. One critical residue is an invariant Trp153 of the β2 domain (Trp153β) that forms a hydrogen bond to the main chain carbonyl oxygen of Glu30α from α1 domain. In addition, the indole ring of this Trp153β is surrounded by three conserved hydrophobic residues (Phe26α, Leu45α, and Phe48a) of the α1 domain. Since Trp153β sits on the DE loop, which is itself rigidified by an internal hydrogen-bond network within the loop (shown in blue broken lines), this network may play a key role in fixing the β2 domain’s relative position. Also seen in Figure 2 is an invariant Tyr150α of the α2 domain that forms a hydrogen bond to the main chain carbonyl oxygen of Asp152β, just next to Trp153β. This Tyr150α is located at the E strand of α2 domain’s framework, and along with other interaction shown in Figure 2 constrains the α2 domain

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FIGURE 1 | The CD4 co-receptor binds to class II but not class I MHC molecules. (A) Superposition of selected MHCII molecules onto the structure of the MHCII molecule DR1 in complex with the N-terminal two-domain of CD4 (PDB code: 3S5L). The superposition is based on the α2 domain. In all cases, the MHCII molecules can accommodate the CD4 N-terminal domain for binding. (B) Two MHC I molecules are superimposed onto the DR1-CD4 complex structure. Here the β2M domain of MHC I was overlaid on the α2 domain of DR1 molecule. These two selected MHC I molecules have the largest (1AGF, in blue) and smallest (1QVO, in yellow) opening angle, respectively, between the two membrane-proximal domains, α3 and β2M. Compared with the MHCII DR1-CD4 complex, they do not have a sufficiently large opening angle to accommodate CD4. (C) Close-up of (B). so that it maintains a relatively fixed orientation with respect to the β2 domain. These interface interactions ensure that the α2/β2 domains are positioned suitably for insertion of the N-terminal domain of CD4 during binding.

Among class II MHC-like molecules, it is interesting to note that the HLA-DM (1HDM) is structurally distinct from other class II MHC molecules (49) (Figure 3). HLA-DM has a C′ strand instead of a D-strand in its β2 domain. In all other MHCII molecules, the D strand plays a critical role in forming a mini-β sheet with the CD4 C′′ strand upon binding (7). Additionally, HLA-DM has a distinct AB loop protruding out from its β2 domain that would prevent CD4-binding. DM is not expressed as a cell surface molecule, instead functioning intracellularly to facilitate peptide loading into other MHCII molecules. It would appear that HLA-DM is without any CD4-binding features, consistent with its intracellular role.

THE INVARIANT Gln226 OF MHC I MOLECULES IS THE DETERMINATE RESidue FOR CD8-BINDING

Figure 4A is the overview of the structure of mouse CD8αβ heterodimer in complex with MHC I molecule H-2Dd (9). The overall structure is in a good agreement with previously published complex structures including the mouse CD8αa/H-2Kβ (50) and human CD8αa/HLA-A2 (5) as well as mouse CD8αa homodimer in complex with non-classic MHCIIb molecule TL (51). That CD8αβ/H-2Dd complex interaction is clearly distinct from CD4-binding to an MHCII molecule shown in Figure 1. The major CD8-binding site is the CD loop of MHC I molecule’s α3 domain, most notably containing Q226. Importantly, this α3 domain is the sole contact region for the CD8αβ co-receptor, essential for cytotoxic T cell function (9).

Of note, there is a very similar CD loop in the β2 domain of MHCII molecules (7). However, what makes MHC I molecules unique with respect to CD8-binding is the invariant Gln226 at the tip of their α3 domain CD loop. The functional importance of this Gln226 within the CD loop was first reported in 1990 (52). Later, it was further found that this Gln226, but not acidic residues on the CD loop was critically required (53). Figure 4B is a detailed view of the Gln226 contact area in the structure of the CD8αβ/H-2Dd (3DMM). The very tip amide nitrogen forms bifurcated hydrogen bonds to the Ser37 side chain of the CD8α subunit (in yellow) and the main chain oxygen of CD8β subunit’s Pro102 (in gold). Interestingly, we have noticed that these hydrogen bonds pull the side chain of Gln226 into an extended conformation such that the elongated aliphatic portion of this Gln226 side chain packs onto the aromatic ring of CD8α Tyr55. This hydrophobic interaction conceivably contributes a significant amount of binding energy. Remarkably the Ser37 and Tyr55 are conserved only in CD8α, but not in CD8β subunit, which
FIGURE 2 | Conserved inter-domain interface in the MHCII molecule HLA-DQ8 (1JK8) that maintains the inter-domain orientation for CD4-binding. (A) The overall view of HLA-DQ8. The gold color indicates the α chain whereas the cyan color defines the β chain. (B) Zoomed-in view of detailed inter-domain interaction involving particularly W153 at the β chain’s DE loop (in yellow) and Y150 on the α chain’s E strand (in light brown).

FIGURE 3 | Superposition of the HLA-DM structure onto the DR1-CD4 complex. The superposition is based on the α2 domain. HLA-DM’s α2 domain is in green, whereas the β2 domain is in cyan. Note that DM’s β2 domain does not have a D-strand for pairing to CD4’s C-strand necessary for docking, a key feature of CD4-binding to all MHCII molecules. In addition, DM’s AB loop protrudes, thereby colliding with CD4.

structurally positions CD8α subunit at the bottom and CD8β unit on the top shown in the Figure 4. The MHCII CD loop was noticed to have an extensive internal hydrogen-bond network to rigidify the loop, supporting the Gln226 for binding (50). These CD8-binding structural features observed in CD8αβ/H-2Dd are shared in all known CD8αα/MHCI complex structures. Upon CD8 binding to MHCII, the MHCII molecules’ buried surface area for structures of HLA-A2/CD8αα, H-2Kd/CD8αα, TL/CD8αα and H-2Dd/CD8αβ is 1074, 1557, 1645, and 781 Å², among which a single Gln226 residue alone contributes 16, 11, 10, and 23%, respectively. Interestingly for the H-2Dd/CD8αβ complex, since only the MHCII α3 domain is involved in binding, the buried surface area is smallest but with Gln226 makes the highest contribution.

In summary, whereas MHCII molecules have their α2/β2 domains arranged in a conserved and opened conformation suitable for CD4 N-terminal domain insertion for binding, MHC molecules are unique in having a key invariant Gln226 at the tip of their α3 domain’s CD loop. The latter is well-positioned to extend into the crevice formed between two CD8 subunits for specific interactions. These observations apply to the 58 MHCII and 224 MHC structures of both human and murine origin examined.

SEQUENTIAL αβTCR–pMHC INTERACTION AND CO-RECEPTOR DELIVERY OF Lck IS LINKED TO CD4/8 GENE IMPRINTING

The above structural scrutiny leads us to propose the following heuristic. On developing DP thymocytes, there are clonally distributed MHCI- or MHCII-restricted TCRs that can potentially interact with the antigen presenting platforms of MHCI (α1α2 domains) and MHCII (α1β1 domains) molecules extending from the stromal cell surface. The interacting domains of both TCR and MHC molecules have no intrinsic structural differences that distinguish the two classes of MHC interactions; nor are TCR complex components distinct in overall structural features vis-à-vis the individual subunits (11). It is difficult, therefore, to conceive that such TCR-pMHC interactions would direct a differential switch-off of either CD4 or CD8 co-receptors. Instead, the preference of the TCR for a specific pMHC dictates which co-receptor then enters into the preformed TCR-pMHC
complex. During positive selection, DP thymocytes first become intermediate cells (CD4⁺CD8low). If during positive selection, one MHCII-restricted TCR on the CD4⁺CD8low cell surface encounters an MHCII molecule that presents a self-peptide on the stromal cell, the “snug” fit of CD4 into the MHCII molecule’s α2β2 domains on the cell surface as described above brings Lck into close proximity with exposed CD3 cytoplasmic tail ITAMs in the TCR complex. If successful ITAM phosphorylation and downstream activation persists, transcriptional factors like GATA3 and Th-POK are induced and that induction promotes CD4⁺ T cell differentiation by preventing Cd4 gene silencing (40). This successful signaling eventually terminates any CD8 expression, and the thymocyte differentiates into CD4⁺ SP cell. If during positive selection an MHCI-restricted TCR on one CD4⁺CD8low thymocyte surface encounters an MHCI that presents a self-peptide, then CD4 fails to contact the MHCI molecule. Consequently, TCR signaling is disrupted with loss of GATA3 and Th-POK transcription. Instead, IL-7R signaling is initiated to up-regulate RUNX3, thereby silencing Cd4 and activating Cd8 expression. CD8 co-receptor function is operative. As RUNX3 is also linked to the cytotoxic program, this dichotomy normally ensures that CD8 SP development is coupled to the CTL program.

As both human and mouse TCRs manifest ~10-fold greater 3-D binding affinities for pMHCI compared to pMHCII cognate ligands (54, 55), the relatively persistent interaction of class II MHC-restricted TCRs with pMHCI ligands in the thymus as postulated above may appear paradoxical. However, recent studies show that TCR-triggered interaction with pMHC is force dependent, dramatically modifying TCR-pMHCI bond lifetimes (56, 57). Such bond lifetime alteration is not predicted by 3-D affinity measurements. Furthermore, the trimolecular TCR-pMHCI-co-receptor interaction likely alters the landscape of bond lifetimes further, particularly under a force load, as would occur when thymocytes scan thymic epithelial cells. That said, other differences in CD4 vs. CD8 lineage-specific death rates at the DP thymocyte branch point imply additional biological parameters modulating developmental progression (58).

CONCLUDING REMARKS
Since the introduction of TCR transgenic mouse studies, it has been clear that the interaction of the TCR in a DP thymocyte with a specific thymic pMHC molecule determines differentiation of that DP thymocyte to either a CD4 or CD8 SP thymocyte (59). The paradox that there were no overall architectural distinctions between TCRs that recognize peptides bound to MHCI vs. bound to MHCII molecules as revealed by structural analysis made this finding enigmatic [reviewed in Ref. (11)]. From our extensive structure data survey, it is now clear that the ability of CD4 to bind pMHCII and CD8 to bind to pMHCI molecules is inviolate. TCR recognition of antigen is MHC-restricted due to the requirement for co-receptor-mediated delivery of Lck to the TCR-pMHC complex during positive selection. Transient loss of CD8 expression that accompanies positive selection of all DP thymocytes supports a prolonged CD4 co-receptor dependent interaction of class II MHC-restricted TCRs on thymocytes with pMHCII on stromal cells. By comparison, the corresponding CD8 co-receptor dependent interaction of class I MHC-restricted TCRs with pMHCI is attenuated. In the latter case, TCR-pMHCI signaling is disrupted, IL-7R signaling permitted and consequently, RUNX3 triggers the cytotoxic transcriptome program. Thus, the specificity for CD4 vs. CD8 SP lineage commitment is coupled to the co-receptor based on a given TCR’s pMHC class preference. In contrast, effector function is linked to cytokine receptor-mediated signaling events that control downstream cellular transcriptional programs.

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Co-receptors’ role in lineage commitment

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