High-throughput sequencing data of soil bacterial communities from Tweefontein indigenous and commercial forests, South Africa

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ABSTRACT

In this report, the high-throughput sequencing data of soil bacterial communities from indigenous and commercial forests in Tweefontein, South Africa are presented. These data were collected to study the influence of land-use change on soil bacterial diversity and community structure in forests. Illumina Miseq sequencing of 16S rRNA gene amplicon was carried out on soils sampled from Tweefontein commercial (TC) and indigenous (TI) forests in South Africa. The metagenome contained 101,938 sequences with 46,709,377 bp size and 57% G + C content in TI and 91,160 sequences with 41,707,827 bp size and 57% G + C content in TC. Metagenome sequence information are available at NCBI under the Sequence Read Archive (SRA) database with accession numbers SRR8134476 (TI) and SRR8135323 (TC). Taxonomic hits distribution from Metagenomic Rast Server (MG-RAST) analysis of the TI sample revealed the dominance of the phyla Acidobacteria (21.61%), Actinobacteria (18.23%) and Verrucomicrobia (16.78%). Predominant genera were Candidatus Koribacter (12.82%), Candidatus Solibacter (11.74%) and Chthoniobacter (9.36%). MG-RAST assisted analysis of TC sample also detected the dominance of Actinobacteria (23.62%) along with Verrucomicrobia (21.92%) and Acidobacteria (20.74%). Predominant genera were Chthoniobacter...
Candidatus Solibacter (24.94%), Candidatus Solibacter (16.74%) and Candidatus Koribacter (9.39%) which play vital ecological functions in forest ecosystems.

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1. Data

The dataset contains raw sequencing data acquired through the 16S rRNA amplicon sequencing of Tweefontein indigenous (TI) and commercial forest (TC) soils from South Africa. The data files (reads in FASTQ format) were deposited at NCBI SRA database under project accession numbers SRR8134476 (TI) and SRR8135323 (TC). Data about the diversity and structure of bacterial communities of Tweefontein indigenous (TI) and commercial (TC) forest soils are presented in Figs. 1 and 2 respectively.

2. Experimental design, materials and methods

In this dataset, soil samples were collected from Tweefontein indigenous forest and the adjacent Tweefontein commercial forest (−24°58′S, 30.48′E and 1239.47 m above mean sea level) in July 2016 during winter. The indigenous forest covers an area of 10,484.09 ha while the commercial forest covers
5965.84 ha. The commercial forest is presently on second rotation (one rotation = 30 years) and sustainable forest management is practiced. This plantation has been FSC certified for the past 20 years [1]. The indigenous forest and commercial plantation are about 2 km apart. Ten soil cores (2 cm in diameter and 10 cm in depth) were collected within multiple tree rows at various points within the sampling sites. These cores were then pooled together and homogenized into a composite sample per site. After sampling, the soil samples were preserved temporarily in cooler boxes filled with ice and conveyed to the laboratory where they were stored in a fridge at a temperature of 4°C for 2 weeks. Thereafter, metagenomic DNA extraction was performed using the PowerSoil® DNA isolation kit (MoBio Laboratory, CA, USA) according to the manufacturer's instructions. NGS was done using Illumina Miseq at Molecular Research LP, Shallowater, TX, USA. Quality and quantity of extracted DNA were analysed by NanoDrop ND-2000 and Qubit. The 16S rRNA libraries were prepared from the QC passed DNA samples using the PCR primers 515F (5′- AATGATACGGCGACCAAGCTATGCTAATT GT GTGCCAGCMGCCGCGGTAA – 3′) and 806R (5′- CAACGAGAAGACGGTACGAGAT TCCGTTCCTCC AGTCAGTCAG CC GGACTACHVGGGTWTCTAAT – 3′) with standard Illumina barcodes and adapters. The amplicons were further purified using Ampure XP beads. The barcoded libraries were validated by Agilent DNA 1000 Bioanalyser and quantified using Qubit DNA BR reagent assay. The quantified libraries were pooled and sequenced using MiSeq. Raw sequences from Illumina Miseq were processed and analysed using MG-RAST server v4.0.3 (http://metagenomics.anl.gov/) [2]. Raw data were
uploaded as FASTQ files after demultiplexing of paired-end reads. Reads generated after quality processing and deduplication by MG-RAST pipeline analysis were subjected to taxonomic analysis. MG-RAST pipeline made available an estimation of bacterial abundances present in Tweefontein indigenous and commercial forests and based on this, an evaluation was done to appraise the bacterial diversity within the samples.

3. Nucleotide sequence accession number

Sequences used for the compilation of this data have been deposited in the Sequence Read Archive (SRA) of the National Center for Biotechnology Information (NCBI) under the bioproject numbers SRR8134476 (TI) and SRR8135323 (TC).
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Conflict of Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

References

[1] A.E. Amoo, O.O. Babalola, Impact of land use on bacterial diversity and community structure in temperate pine and indigenous forest soils, Diversity 11 (2019) 217.

[2] F. Meyer, D. Paarmann, M. D'Souza, R. Olson, E.M. Glass, M. Kubal, T. Paczian, A. Rodriguez, R. Stevens, A. Wilke, The metagenomics RAST server—a public resource for the automatic phylogenetic and functional analysis of metagenomes, BMC Bioinf. 9 (2008) 386.