Degradation of GRK2 and AKT is an early and detrimental event in myocardial ischemia/reperfusion

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ABSTRACT

Background: Identification of signaling pathways altered at early stages after cardiac ischemia/reperfusion (I/R) is crucial to develop timely therapies aimed at reducing I/R injury. The expression of G protein-coupled receptor kinase 2 (GRK2), a key signaling hub, is up-regulated in the long-term in patients and in experimental models of heart failure. However, whether GRK2 levels change at early time points following myocardial I/R and its functional impact during this period remain to be established.

Methods: We have investigated the temporal changes of GRK2 expression and their potential relationships with the cardioprotective AKT pathway in isolated rat hearts and porcine preclinical models of I/R.

Findings: Contrary to the maladaptive up-regulation of GRK2 reported at later times after myocardial infarction, successive GRK2 phosphorylation at specific sites during ischemia and early reperfusion elicited GRK2 degradation by the proteasome and calpains, respectively, thus keeping GRK2 levels low during early I/R in rat hearts. Concurrently, I/R promotes decay of the prolyl-isomerase Pin1, a positive regulator of AKT stability, and a marked loss of total AKT protein, resulting in an overall decreased activity of this pro-survival pathway. A similar pattern of concomitant down-modulation of GRK2/AKT/Pin1 protein levels in early I/R was observed in pig hearts. Calpain and proteasome inhibition prevents GRK2/Pin1/AKT degradation, restores bulk AKT pathway activity and attenuates myocardial I/R injury in isolated rat hearts.

Interpretation: Preventing transient degradation of GRK2 and AKT during early I/R might improve the potential of endogenous cardioprotection mechanisms and of conditioning strategies.
1. Introduction

Acute myocardial infarction (MI) as a result of coronary artery occlusion is a major cause of death and disability [1,2]. Prompt restoration of blood flow by means of thrombolytic or primary percutaneous coronary intervention (PCI) is key for reducing infarct size and mortality. However, the process of restoring blood flow can itself trigger additional myocardial damage, termed reperfusion injury. Despite advances in the protocols allowing rapid and effective reperfusion in MI patients, therapies adequately targeting reperfusion injury remain elusive and constitute one of the top unmet needs in cardiology [2,7,8]. Brief cycles of myocardial I/R applied either before (ischemic preconditioning) or immediately after reflow (ischemic post-conditioning) in experimental models can reduce infarct size by activating different combinations of endogenous cardioprotective pathways [1,2,4,9]. Activation of kinases such as AKT and ERK1/2, important components of the so-called reperfusion injury salvage kinase pathway (RISK) pathway, plays a key cardioprotective role, and modulation of this pathway by endogenous mediators or pharmacological agents has been postulated as a relevant strategy to prevent I/R injury [1,4,10,11]. Given the limited efficacy of these current therapeutic strategies, it is crucial to gather information about the status of key cardiac signaling pathways at early stages after I/R to identify potential new targets for intervention.

G protein-coupled receptor kinase 2 (GRK2) is a central regulator of beta-adrenergic and many other G protein-coupled receptors (GPCRs) involved in cardiovascular physiopathology. In addition, data in different cell types put forward GRK2 as a key regulatory node in non-GPCR pathways via the modulation of insulin and growth factor signaling, the PI3K/AKT route, MAPK cascades, NO bioavailability or mitochondrial function, which are also instrumental in cardiac function and dysfunction (reviewed in [12–17]). In particular, myocardial GRK2 has been reported to participate in apoptotic pathways after I/R by mechanisms involving its mitochondrial targeting [18,19]. Maladaptive increased GRK2 expression has been described in the failing human heart of patients and in experimental models of HF due to chronic hypertensive or ischemic disease, and its genetic ablation or inhibition has been reported to be cardioprotective in the long-term in animal models by reducing adverse post-infarction remodeling, purportedly as a result of the integrated actions of GRK2 in myocardial contractile function and cardiac metabolism (reviewed in [15,20–23]). However, whether GRK2 protein levels are altered at early time points following myocardial I/R, the molecular mechanisms involved and the impact of such changes in GRK2-related signaling networks has not been fully investigated.

2. Materials and methods

2.1. Experimental protocol of ischemia/reperfusion in rats

The experimental procedures conformed to the Guide for the Care and Use of Laboratory Animals published by the United States National Institute of Health (NIH Publication Eighth Edition, 2011) and were approved by the Research Commission on Ethics of the Hospital Vall d’Hebron. Male Sprague-Dawley rats (Charles River, Barcelona, Spain) weighing 250 to 300 g were anaesthetized with sodium pentobarbital (100 mg/kg). Hearts were removed, mounted onto a Langendorff apparatus, and perfused with a modified Krebs-Henseleit bicarbonate buffer as previously described. Hearts were perfused normoxically for 60 min (Nx group) or for 20 min and then subjected to 40 min of global ischemia followed by reperfusion of different duration (I/R group). Ischemic...
preconditioning protocol consisted of two cycles of 5 min of ischemia and 5 min of reperfusion applied before index ischemia (PreCo group). Ischemic postconditioning was achieved with a previously established protocol commenced immediately after ischemia consisting of 6 cycles of 10-second reperfusion-10-second occlusion (PostCo group) [24–26]. In additional groups of hearts, the membrane-permeable calpain inhibitor SNJ-1945 (Senju Pharmaceutical Co,Ltd) at 10 μM [27], the proteosome inhibitor bortezomib (Cell Signaling Technology) at 1 μM and 10 μM, the PI3K inhibitor LY-294002 (10 μM, Sigma-Aldrich), or the indicated combinations of SNJ-1945; LY-294002 and 10 μM bortezomib or their vehicle (0.01% DMSO) were added to the perfusion media during the 10 min prior to 40 min of ischemia and the first 10 min of reperfusion.

The selective contractile inhibitor blebbistatin (10 μM, Merck) was added during the first 5 min of reperfusion in all the experiments involving ischemia/reperfusion, except in those experiments specifically aimed to assess cell death and infarct size. Blebbistatin prevents cell death by reducing the mechanical stress caused by the excessive and irreversible contractile contractive activation occurring at the onset of reperfusion. It was used to discard the possibility that any variation on the measured parameters was a mere consequence of differences in cell death associated with sarcolemmal disruption [26].

2.2. Experimental protocol of ischemia/reperfusion in pigs

Transient myocardial ischemia in pigs was performed as previously described [28]. Farm pigs (25–30 kg) were pre-medicated with tiletamine–zolazepam (4–6 mg/kg, IM) and xylazine (1–2 mg/kg, IM), anaesthetised with propofol-lipuro 1% (1–2.5 mg/kg, IV, followed by continuous infusion at 11 mg/kg/h) and fentanyl (5 mg/kg, IV, followed by continuous infusion at 3–6 mg/kg/h), and mechanically ventilated. A mid-sternotomy was performed, and the left anterior descending (LAD) coronary artery was dissected free at its midpoint. Lead II of ECG, left ventricular (LV) pressure and LV dP/dt, coronary bloodflow, and regional myocardial function were continuously recorded. LAD coronary artery was occluded for 40 min followed by 5 min or 120 min of reperfusion. At the end of the experiment, animals were sacrificed by a pentobarbital over-dose (100 mg/kg, IV) and myocardial tissue samples from both, the control region and the area at risk were quickly excised and frozen in liquid nitrogen.

2.3. Myocardial tissue processing

Myocardial rat or porcine tissue was minced and homogenized in 4 vol (v:v) 20 mm Tris–HCl (pH 7.5), 5 mm EDTA, 5 mm EGTA, and protease inhibitors. The homogenate was centrifuged (2000 × g, 5 min, 4 °C) to obtain a clarified post-nuclear supernatant designated as crude cytoplasmic fraction (SB1). The pellet was rinsed in the same buffer (1/5 of the initial volume) supplemented with 0.5% NP40, extensively vortexed and centrifuged (850 × g, 10 min, 4 °C) after 15 min ice-cold incubation to obtain a NP40-extracted nuclear fraction (SB2). For determination of GRK2 levels in some conditions, crude cytoplasmic fractions were previously enriched in GRK2 protein by incubating these fractions in liquid nitrogen.

Myocardial tissue was lysed in immunoprecipitation buffer (50 mM HEPES pH 7.5, 150 mM NaCl, 1% Triton, 10% Glycerol, 10 mM NaF, 1 mM sodium orthovanadate, plus protease inhibitors). Upon centrifugation (15,000 × g, 10 min), supernatants were incubated with a specific anti-Pin1 polyclonal antibody (Upstate Biotechnology or Millipore). Immune complexes were resolved in 15% SDS/PAGE and transferred to nitrocellulose membranes. After incubation with antibody anti-Acetyl lysine (Clone 4G12, Millipore), blots were stripped and reproved with a polyclonal antibodies directed against the immunoprecipitated Pin1 [30]. The amount of Pin1 acetylation was normalized to the amount of the immunoprecipitated protein, and data were represented as the fold over control normoxic conditions [29,30].

2.4. Western blot and dot blot analysis

Proteins were separated by SDS-PAGE for Western blot analysis or applied directly on nitrocellulose membranes in a single spot for immunodetection (dot-blots) as previously described [29]. Protein bands were detected by the chemiluminescence method (ECL, Amersham Pharmacia Biotech) or with the Odyssey Imaging Systems (LI-COR Biosciences). Bands were quantified by laser-scanner densitometry with a Biorad GS-700 scanner or by the software included in the Odyssey Infrared Imaging System. The rabbit polyclonal antibodies raised against phospho-Ser/Thr-PKA substrate, phospho-Thr/Thr-AKT substrate, pSer473-AKT and AKT and the mouse monoclonal anti-Acetylated-Lysine antibody were all from Cell Signaling Technologies. Rabbit primary antibodies raised against ERK1/2, Lamin B1 and PKA catalytic subunit were from Santa Cruz Biotechnology Inc. (Santa Cruz, CA, USA). Mouse anti-PKA RI-subunit was from BD Transduction Laboratories. Anti-Pin1 rabbit polyclonal and GAPDH mouse monoclonal (6C5) antibodies were provided by Upstate Biotechnology and Abcam, respectively. Anti-pSer670-GRK2 and anti-pSer685-GRK2 polyclonal antibodies were from Biosource International and from Immunoway. Anti-acetyl-K40-tubulin and anti-tubulin antibodies were purchased from Sigma. GRK2 protein was immunodetected with a rabbit polyclonal antibody Ab C-15 (sc-562) (Santa Cruz). Vertical dotted lines in western blots indicate juxtaposed lanes that come from the same gel but were non-adjacent.

2.5. Determination of Pin1 acetylation

Myocardial tissue was lysed in immunoprecipitation buffer (50 mM HEPES pH 7.5, 150 mM NaCl, 1% Triton, 10% Glycerol, 10 mM NaF, 1 mM sodium orthovanadate, plus protease inhibitors). Upon centrifugation (15,000 × g, 10 min), supernatants were incubated with a specific anti-Pin1 polyclonal antibody (Upstate Biotechnology or Millipore). Immune complexes were resolved in 15% SDS/PAGE and transferred to nitrocellulose membranes. After incubation with antibody anti-Acetyl lysine (Clone 4G12, Millipore), blots were stripped and reproved with a polyclonal antibodies directed against the immunoprecipitated Pin1 [30]. The amount of Pin1 acetylation was normalized to the amount of the immunoprecipitated protein, and data were represented as the fold over control normoxic conditions [29,30].

2.6. Cell culture and transfection and protein degradation assays

HEK293 were obtained from the American Type Culture Collection (ATCC) and maintained in DMEM supplemented with 10% (v/v) foetal bovine serum (FBS) at 37 °C in a humidified 5% CO2 atmosphere. Transient transfections with the indicated combinations of cDNA were performed using the Lipofectamine/Plus method, following manufacturer’s instructions.

Metabolic labelling and pulse-chase experiments were performed as described [31]. HEK293 cells previously co-transfected with PKA catalytic subunit or empty vector and GRK2wt, GRK2-K220R or GRK2-S685A constructs were labeled by keeping the cells for 2 h in methionine and cysteine-free Dulbecco’s modified Eagle’s medium (DMEM) and then incubated for 15–30 min in DMEM supplemented with 250 μCi/ml of [35S]methionine and [35S]cysteine labeling mixture (NEN Life Science Products). The plates were washed with phosphate-buffered saline and chased for the indicated times in DMEM plus 10% fetal bovine serum. The proteolysis inhibitors lactacystin (Calbiochem, La Jolla, CA), ALLN (Sigma), or PD150606 (Calbiochem), the PKA activator Forskolin blue with 100 mM maleimide) or reducing SDS sample buffer (i.e. supplemented with 5% β-mercaptoethanol).
Results

GRK2 was quantified by laser densitometry analysis and data were previously validated [32]. Immunoprecipitates were resolved by SDS/PAGE and transferred to PVDF membranes to be treated with a specific rabbit polyclonal antibody AbFP1 and probed with phosphosite-specific antibodies. Data were normalized to total GRK2 levels, GAPDH expression was used as loading control. In all panels, data are mean ± SEM.

2.7. Quantification of cell death

Lactate dehydrogenase (LDH) activity was spectrophotometrically measured in the coronary effluent throughout the reperfusion period. After 60 min of reperfusion, heart slices were incubated at 37°C for 10 min in 1% 2,3,5-triphenyltetrazolium chloride and imaged under white light to outline the area of necrosis as previously described [25]. Previous studies demonstrated that in the Langendorff rat model, reperfusion for 60 min is sufficient for acute assessment of infarct size [26,33].

2.8. Statistical analysis

Data analysis was performed using GraphPad Prism for Windows. Means between groups were compared by 2-way or one-way ANOVA with Bonferroni’s or Tukey’s post hoc test, respectively, or with unpaired Student’s t-test as indicated in the Figure legends. All results are expressed as mean ± SEM.

3. Results

3.1. GRK2 is degraded in ischemia and reperfusion conditions by different mechanisms

Potential changes in GRK2 levels were analyzed during myocardial ischemia and the early time points following reperfusion, by using a well-established experimental model of cardiac ischemia/reperfusion (IR) in isolated rat hearts [24,25,34]. Notably, 40 min of ischemia promoted a marked decay in cardiac GRK2 protein levels compared to control conditions, and GRK2 down-regulation was maintained after 30 min of reperfusion, when a circa 50% decrease compared to normoxic GRK2 expression was noted (Fig. 1A). Neither pre- nor post-conditioning treatments, performed as described previously [24,25], were able to prevent the ischemia-promoted GRK2 down-regulation. These treatments did not fully restore GRK2 protein levels during the 30 min of reperfusion, although slightly higher kinase levels at early (preconditioning) or later (post-conditioning) reperfusion times were observed. No overt changes in the subcellular distribution of the kinase were observed in either I or in I/R experimental conditions (Fig. S1), suggesting that the rapid down-modulation of GRK2 levels was related to protein degradation.

GRK2 can be rapidly degraded by the proteasome, in a process dependent on its previous phosphorylation at the S670

Fig. 1. Ischemia, reperfusion and conditioning treatments trigger marked and dynamic changes in cardiac GRK2 protein levels and phosphorylation status. Isolated rat hearts were exposed to 40 min of ischemia alone 40 min before metabolic labeling and maintained during the chase period. At different chase times (1–3 h), cells were harvested using radioimmune precipitation assay lysis buffer (20 mm Tris-HCl, pH 7.5, 150 mm NaCl, 1% sodium deoxycholate, 0.5% Nonidet P-40, 0.1% SDS, with a mixture of protease inhibitors). GRK2 protein was immunoprecipitated with a specific rabbit polyclonal antibody AbFP1 and probed with phosphosite-specific antibodies. Data were normalized to total GRK2 levels, GAPDH expression was used as loading control. In all panels, data are mean ± SEM, n = 3–4 rats per condition. In all panels, data were analyzed by comparing the different experimental situations (I/R, I/R-PreCo and I/R-PosCo) to the normoxic condition [2-way ANOVA followed by Bonferroni’s post-hoc test, †p < .05; ‡p < .01; §§p < .001]. In addition, we compared I/R pre-Co and I/R post-Co conditions versus I/R alone (*p < .05; **p < .01) or between conditioning situations (#p < .05; †††p < .001) [1-way ANOVA and Tukey’s post hoc test]. Representative blots are shown.
residue [35,36]. Interestingly, a dramatic increase of GRK2 S670-phosphorylation status was noted in ischemic conditions (Fig. 1B), a pattern consistent with the involvement of the S670-GRK2 phosphorylation/proteasome pathway in kinase downmodulation during this period. However, S670 phosphorylation rapidly returned to normoxic levels upon reperfusion in all conditions tested, suggesting that additional mechanisms were involved in maintaining GRK2 protein levels low during the initial phase of reperfusion.

We searched for other regulatory post-translational modifications during this period. GRK2 is phosphorylated by PKA on S685, a residue highly conserved across species, leading to increased phosphorylation during this period. GRK2 is phosphorylated by PKA on S685, suggesting that additional mechanisms were involved in maintaining GRK2 protein levels low during the initial phase of reperfusion.

The pattern of total and phospho-S685 GRK2 levels during reperfusion suggested that this phosphorylation might help to keep GRK2 downregulated. Pulse-chase experiments in HEK-293 cells, a well-established model for characterizing GRK2 degradation mechanisms [20,35,40], indicated that forskolin (FO), a direct stimulator of adenylyl cyclase activity, and thus of the PKA pathway, increased the basal turnover of GRK2 (Fig. 3A), as well as triggered phosphorylation of GRK2 on S685 (Fig. S2). Interestingly, FO also enhanced the degradation of the catalytically-inactive GRK2-K220R mutant (Fig. 3A), previously reported to be impaired in GPCR-induced proteasome-mediated degradation [41]. This suggested that the GRK2-destabilizing effect of FO involved alternative proteolytic pathways.

It is worth noting that the fostering effect of FO in GRK2 degradation was fully prevented upon addition of a PKA inhibitor (Fig. S2B), whereas over-expression of the catalytic subunit of PKA enhanced the decay of both wild-type GRK2 and the K220R mutant (Fig. 3B), confirming the involvement of PKA downstream of FO in the modulation of GRK2 stability. Moreover, a GRK2-S685A mutant (unable to be phosphorylated by PKA) displayed a stability similar to WT GRK2 in basal conditions, but lacked the PKA-stimulated component of GRK2 degradation, demonstrating that phosphorylation on S685 was required for PKA activity-induced GRK2 downregulation (Fig. 3C).

Consistent with the involvement of alternative PKA-dependent proteolytic pathways, the proteasome inhibitor lactacystin fully inhibited the basal degradation of wt GRK2, whereas only partially attenuated GRK2 proteolysis in the presence of FO stimulation (Fig. 3D). On the other hand, PKA-triggered GRK2 degradation was completely abrogated upon addition of broad spectrum (ALLN, Fig. 3D)

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**Fig. 2.** Cardiac ischemia/reperfusion induces rapid and transient oxidation-dependent activation of PKA. (A) Global PKA activity was assessed in lysates from normoxic or reperfused isolated rat hearts (40 min of sustained ischemia followed by the indicated reperfusion times) by dot-blot, using a pan-specific phospho-substrate antibody that broadly detects phosphorylated proteins by PKA as indicated in Methods. GAPDH expression was used as loading control. Data are mean ± SEM, n = 3–4 rats per condition. **p < .01; ***p < .001 compared to I/R 30 min [1-way ANOVA and Tukey’s post hoc test]. A representative dot-blot is shown. (B) I/R promotes oxidation-related dimerization of the regulatory PKA-RIα subunit in parallel to global PKA activation. Lysates as in panel A were probed with a specific PKA-RI antibody that recognizes both monomers and dimers of this regulatory subunit linked by disulfide bond formation. The dimer formation detected in non-reducing SDS-PAGE resolving conditions was fully reversed in the presence of 2-mercaptoethanol (reducing SDS-PAGE). The expression of the PKA catalytic subunit and of GAPDH was used as loading controls. Data are mean ± SEM, n = 3–4 rats per condition. *p < .05 for the indicated comparisons [Student’s t-test]. Representative blots are shown.
or specific (PD150606, Fig. S2C) inhibitors of calpains, a family of Ca2+ -dependent cysteine proteases reportedly over-activated in myocardial reperfusion, leading to myocardial injury through the proteolysis of different proteins [42-44], suggesting that calpain-mediated proteolysis of phospho-S685-GRK2 might take place in early I/R conditions.

Overall, our data suggested that dynamic and sequential phosphorylation of GRK2 at S670 and S685 during ischemia and in early reperfusion would trigger GRK2 degradation by the proteasome and via calpains, respectively, thus maintaining low GRK2 protein levels during such initial period after heart damage.

3.2. AKT degradation takes place in early I/R resulting in decreased overall activity of this pathway

We investigated whether such transient GRK2 downmodulation was related to changes in other key cardiac signaling pathways. Total ERK1/2 protein levels were not significantly altered during I/R or upon pre- or post-conditioning (Fig. S3A), indicating that I/R does not promote a general decay in signaling proteins. Interestingly however, total AKT protein levels were markedly diminished by ischemia and maintained low during the first 30 min of reperfusion (Fig. 4A), with a pattern similar to that detected for GRK2. Moreover, AKT and GRK2 protein levels displayed a similar trend of changes upon pre- conditioning, which led to a less marked AKT decrease after ischemia and at very early reperfusion times (Fig. 4A). As for GRK2, no manifest changes in the nuclear/cytoplasmic distribution of AKT were observed in either I or in I/R experimental conditions (Fig. S3B), suggesting the occurrence of protein degradation processes.

Such pattern of decreased AKT protein levels during I/R was puzzling, since the cardio-protective and pro-survival AKT pathway is considered to be robustly enhanced in these conditions to counterbalance I/R-promoted injury, as inferred from enhanced AKT phosphorylation at activating residues [1,4,11]. In our experimental setting, we also detected such marked increase of stimulated (phosphor-S473) AKT during reperfusion and upon pre- or post-conditioning conditions (Fig. 4B). However, the concurrent alteration in total AKT protein levels appears to have been disregarded or not addressed in detail in other studies. Our data point to a more complex scenario in which I/R triggers both AKT protein downmodulation and activation of the remaining kinase. Notably, when the “global” or bulk AKT activity was tested in cardiac extracts using a pan-AKT substrate antibody, we observed a marked decrease throughout the I/R period considered (Fig. 4C), with a similar pattern upon post- or pre-conditioning, although the latter condition allowed slightly higher substrate phosphorylation at 5 min after reperfusion. Our data indicated that hyper-activation of the remaining AKT during reperfusion might not be able to fully compensate for the decay in protein levels, leading to a clear over-
all downmodulation of the "catalytic potency" of this key cardioprotective pathway in early I/R contexts, what would counteract the beneficial effects of treatments specifically aimed at enhancing the AKT activation status.

In search of molecular mechanisms underlying AKT protein decay and potentially connecting transient GRK2 and AKT downmodulation, we focused on the prolyl-isomerase Pin1, a central player in the control of folding, activity, and stability of proteins [45,46]. Pin1 is a critical positive factor in the regulation of AKT stability in cancer cells by binding to activated AKT and preserving the protein from degradation by the proteasome [46,47]. Pin1 has also been described as a relevant factor in cardiac hypertrophy, directly binding to AKT and MEK and fostering hypertrophic signaling [48]. Interestingly, we found a clear reduction of Pin1 levels after ischemia and during early reperfusion times (Fig. 5A), with a pattern resembling that of AKT protein, suggesting a link between Pin1 loss and decreased AKT stability. A similar pattern of Pin1 changes was evident in nuclear and cytoplasmic cellular fractions, consistent with Pin1 degradation events (Fig. 5C).

Interestingly, GRK2 is a regulator of Pin1 stability and functionality in other cell types [30]. GRK2 activates HDAC6, a cytoplasmic lysine deacetylase targeting tubulin and many other proteins, thus triggering de-acetylation of Pin1 at a specific residue, thereby favoring Pin1 protein stabilization and stimulation of prolyl-isomerase activity [30,49]. We thus assessed whether Pin1 acetylation might be triggered upon GRK2 downregulation during I/R. This treatment induced a marked increase in Pin1 protein acetylation, which peaked at 5 min of reperfusion (Fig. 5B). A similar pattern was detected for the acetylation levels of the canonical HDAC6 substrate tubulin (Fig. 5C), strongly suggesting an impaired HDAC6 activation by GRK2 in I/R conditions.

Importantly, a comparable pattern of concurrent down-modulation of GRK2, AKT, and Pin1 protein levels in early stages of myocardial reperfusion was also detected in porcine hearts subjected to in situ transient ischemia. The levels of these proteins in the area at risk were significantly decreased at 5 min of reperfusion with respect to control area of the same animal (Fig. 6A–D). After 120 min of reperfusion, total AKT and Pin1 protein expression values were still slightly lower than those of the matched control area, whereas a marked GRK2 down-regulation remained. In line with the mechanistic role of PKA in promoting GRK2 down-modulation observed in the rat model, increased levels of PKA-phosphorylated substrates were detected at 120 min of reperfusion (Fig. 5E). Of note, although an enhanced AKT phosphorylation status was apparent at these I/R stages (Fig. 6C) as previously reported in such conditions [50,51], the global or bulk capacity of active AKT to phosphorylate its targets was markedly reduced (Fig. 6E), akin to the observed decline in total AKT protein levels and in line with our data in the isolated rat heart model (see Fig. 4). Although whether such early and dynamic changes in GRK2/AKT/Pin1 levels take place in patients during cardiac I/R cannot be directly investigated, the data obtained in this relevant translational large animal model supports the potential clinical relevance of these alterations.

3.3. Combined use of proteasome and calpain inhibitors prevents GRK2/Pin1/AKT degradation and attenuates I/R myocardial injury

Overall, our data suggested a complex interconnection among GRK2, Pin1 and AKT in early cardiac I/R. Changes in the phosphorylation status of GRK2 would trigger transient reduction of kinase levels by the combined action of proteasome and calpain path-
Fig. 5. Ischemia/reperfusion alters the protein levels and the acetylation status of the prolyl-isomerase Pin1. (A) Total content of Pin1 protein was analyzed with a specific anti-Pin1 antibody in the indicated conditions. GAPDH expression was used as loading control (B). The acetylation status of Pin1 in the same experimental conditions was determined with a pan anti-Acetyl lysine antibody after immunoprecipitation of total Pin1 from myocardial lysates as detailed in Methods. The amount of Pin1 acetylation was normalized to the amount of the immunoprecipitated protein. (C) Ischemia-reperfusion causes strong hyper-acetylation of myocardial α-tubulin. The levels of acetylated and total α-tubulin in the indicated conditions were analyzed with site-specific anti-acetyl-K40-tubulin and total anti-tubulin antibodies. Total α-tubulin and GAPDH served as loading controls. In all panels, normalized data were represented as fold-change with respect to the normoxic situation, and are the mean ± SEM, n = 3–4 rats per condition. *p < .05, **p < .01, ***p < .001 compared to normoxia [1-way ANOVA and Tukey’s post hoc test]. Representative blots are shown.

Fig. 6. Concurrent down-modulation of GRK2, AKT, and Pin1 protein levels in early stages of reperfusion in porcine hearts. Samples from non-ischemic (controls) or from the ischemic area at risk of lysates obtained from porcine hearts after 40 min of ischemia and 5 or 120 min of reperfusion as detailed in Methods were analyzed with specific antibodies for GRK2 (A), total AKT (B) AKT phosphorylated at Ser473 (C) or total Pin1 levels (D). Data were normalized by GAPDH (panels A, B, D) or total AKT (panel C) loading. (E) Global activity of AKT towards its substrates was assessed in pig cardiac lysates from the indicated conditions by dot-blot, using a pan phospho-AKT substrate-specific antibody, and data normalized by GAPDH loading. In all panels, normalized data were represented as fold-change with respect to control situation (mean value of myocardial control area at 5 min of I/R) and are mean ± SEM, n = 3–5 pigs per condition. *p < .05; **p < .01; ***p < .001 compared with the conditions indicated with the lines [1-way ANOVA and Tukey’s post hoc test]. Representative blots are shown.
ways. GRK2 degradation would favor Pin1 proteolysis by altering its acetylation status, which in turn would facilitate AKT degra-
dation, leading to impaired overall AKT catalytic ability despite hyper-activation of remaining protein, resulting in reduced global capacity to counteract cardiac injury.

We therefore tested the effect of the administration of the clinically used proteasome inhibitor Bortezomib (Bz) and/or the cal-
pain inhibitor SNJ-1945 (SNJ) on cardiac GRK2, Pin1 and AKT levels in isolated hearts subjected to 40 min of ischemia and reper-
fusion for 10 min. A slight significant reduction of AKT degra-
dation was observed in the presence of Bz, and a tendency was de-
tected for GRK2 and AKT protection upon administration of SNJ, but only the combined treatment with Bz and SNJ fully prevented the downregulation of GRK2, Pin1 and AKT in such conditions (Fig. 7A-C), allowing the maintenance of levels close to those detected in normoxic conditions.

Importantly, the administration of protea-
some or calpain inhibitors or their combined delivery significantly rescued bulk AKT functionality, as assessed by the overall phos-
phorylation of AKT substrates during reperfusion (Fig. 7D). This is consistent with the fact that these treatments tend to protect AKT protein from degradation while preserving kinase activation triggered by reperfusion, as indicated by AKT phospho-S473 status (see representative blot in Fig. 7C), thus allowing a normalization of pAKT/AKT ratios. Moreover, the administration of the SNJ calpain inhibitor or the combined delivery of SNJ and the protea-
some inhibitor Bz prior to a protocol of 40 min of ischemia fol-
lowed by 60 min of reperfusion significantly decreased infarct size and LDH release. Of note, this effect was not observed in the pres-
ence of a PI3K inhibitor, indicating that calpain and proteasome inhibitors exert its protective action by mechanism involving the PI3K/Akt pathway (Fig. 8A). Overall, our results postulate that pre-
venting GRK2/Pin1/AKT degradation at early times in I/R may help to counteract myocardial injury.

4. Discussion

Our data reveal several unforeseen alterations taking place in relevant cardiac signaling pathways during the early stages of I/R in rat and porcine experimental models that might provide a ra-

tionale for therapeutic strategies aimed at fostering cardioprotec-
tion mechanisms. First, we uncover that, contrary to the reported up-regulation of cardiac GRK2 taking place at later times after MI [21–23], successive GRK2 phosphorylation at specific sites during ischemia and in early reperfusion would sequentially elicit GRK2 degradation by the proteasome and calpains, respectively, thus keeping GRK2 protein levels low during early I/R. Second, we unveil a marked and previously uncharacterized decrease of AKT protein levels during this period, likely related to concurrent changes in the Prolyl isomerase Pin1 (a known AKT stabilizer), which results in an overall down-modulation of this key card-

ioprotective pathway in early I/R contexts, despite the high acti-
vation status of the remaining AKT upon reperfusion. Third, we show that hindering GRK2/Pin1/AKT degradation by the combined administration of proteasome and calpain inhibitors can attenuate I/R-myocardial injury in isolated rat heart experimental models (scheme in Fig. 8B).

The prevailing view to date is that GRK2 levels are in-
creased in the heart of chronic HF patients with dilated or ischemic cardiomyopathy, as a result of sympathetic nervous system hyperactivity, leading to enhanced GRK2 mRNA expression. Such augmented GRK2 levels may initially help the my-

ocardium to counterbalance the beta-adrenergic overdrive and re-
duce the risk of tachyarrhythmia, but persisting high GRK2 activ-
ity is maladaptive, resulting in GPCR desensitization and down-
regulation, defective contractility, insulin resistance, mitochon-
drial dysfunction and apoptosis, paving the way to HF [13,21,
23].
Strikingly, we find that following myocardial I/R, GRK2 protein levels markedly decrease during the ischemic phase and are kept low during the early phase of reperfusion. Our data suggest that enhanced Ser670-GRK2 phosphorylation during ischemia would trigger its degradation by the proteasome, whereas phosphorylation at Ser685 taking place in early reperfusion would favor calpain-mediated GRK2 proteolysis. Of note, these phosphorylation sites are highly conserved across species, (murine, dog, pig and humans), consistent with preserved modulatory mechanisms.

To our knowledge, the pattern of GRK2 protein levels at such initial times of I/R has not been previously described. In murine models, cardiac GRK2 protein upregulation has been reported as early as 3 and 7 days after acute myocardial injury [13,52]. However, in line with our results, in a canine model of MI GRK2 levels were decreased in the subepicardial border and the infarct zone at 6 and 24h after ligation of the coronary artery, without apparent changes in mRNA levels, in a process blocked by proteasome inhibitors. The fact that GRK2 protein levels remained low during early reperfusion in the absence of enhanced Ser670 phosphorylation suggested the involvement of an alternative degradation pathway. We find that GRK2 is rapidly and transiently phosphorylated by PKA upon intracellular pH normalization, since during ischemia ongoing acidosis inhibits these proteases [42,44].

Upon phosphorylation at Ser670 GRK2 can be rapidly degraded by the proteasome [31,35], and GPCR stimulation facilitates β-arrestin-dependent ERK1/2 activation and subsequent phosphorylation of GRK2 at this residue [35]. Thus, the observed burst in phospho-Ser670 GRK2 after ischemia points to the occurrence of a very active proteasome-dependent degradation of the kinase in this condition. Although an ischemia-triggered increase in GRK2 transcription and/or translation cannot be ruled out, a previous report showed that during stop-flow and low flow ischemia in the isolated perfused rat heart GRK2 mRNA even increased after 20 min of ischemia and then returned to baseline after 40 min of ischemia onset [58], further supporting a predominant role for GRK2 proteolysis in this context.

GRK2 is phosphorylated at Ser670 by several kinases in a context- and stimulus-dependent way, including ERK1/2 [49,59,60], p38MAPK [61] or Cdk2 [31]. Cdk2 is highly activated during myocardial ischemia [10,62] and CDK2-mediated GRK2 S670 phosphorylation can trigger proteasome-dependent GRK2 degradation in a GPCR-independent way [31], suggesting that this regulatory axis would be active during the ischemic phase.

The fact that GRK2 protein levels remained low during early reperfusion in the absence of enhanced Ser670 phosphorylation suggested the involvement of an alternative degradation pathway. We find that GRK2 is rapidly and transiently phosphorylated by PKA at Ser670 upon reperfusion, and that PKA stimulation can trigger calpain-dependent GRK2 proteolysis in a cellular model. Consistent with sequential proteasome and calpain GRK2 degradation during I/R, the combined inhibition of these proteolytic pathways completely prevents GRK2 protein decay in vivo.

Calpains are ubiquitous Ca2+ - dependent cysteine proteases activated in pathological conditions associated with Ca2+ overload. Calpains are thus strongly activated at the onset of myocardial reperfusion, contributing to myocardial injury through the proteolysis of key structural membrane, cytoskeletal components and regulatory enzymes [42,44,63]. Notably, PKA-phosphorylated GRK2 would only be targeted by calpains during the reperfusion phase upon intracellular pH normalization, since during ischemia ongoing acidosis inhibits these proteases [42].

However, the functional relationships among PKA stimulation, calpain activation status and GRK2 degradation are not straightforward and may involve additional regulatory layers. While phosphorylation of GRK2 by PKA switches on degradation by calpain, PKA has been reported to inhibit calpains upon βAR-triggered cAMP stimulation, and such mechanism appears to participate in the molecular mechanisms of cardioprotection upon ischemic preconditioning [64]. This apparent paradox may be reconciled by the

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**Fig. 8.** Cardiac ischemia-reperfusion injury is attenuated by combined-inhibition of proteasome and calpain activities. (A) Isolated rat hearts were treated during the 10 min prior to ischemia (40 min) and the first 10 min of a total period of 60 min reperfusion with the indicated combinations of the calpain inhibitor SNJ-1945 (SNJ, 10 μM), the proteasome inhibitor Bortezomib (BZ) (10 nM) and the PI3K inhibitor LY-294002 (10 μM). After the reperfusion period, total LDH released during reperfusion (expressed as units of activity released per gram of dry weight during the first 60 min of reperfusion, U/g dw/60 min) and infarct size (expressed as the percentage of ventricular mass in the different experimental groups) were determined as detailed in Methods. Data are the mean ± SEM, n = 6–9 rats per condition. *p < .05 vs untreated I/R control group [1-way ANOVA and Tukey’s post-hoc test]. (B) Scheme of the molecular mechanisms underlying GRK2/Pin1/AKT degradation during myocardial ischemia and early reperfusion. See text for details.
existence of compartmentalized pools of these proteins and of dis-
tinct localization and activation mechanisms of PKA isofoms via
regulatory anchoring proteins termed AKAPs [65]. In the vicinity of
the plasma membrane, βAR-dependent production of cyclic-AMP
reportedly promotes activation of AKAP79-bound type II PKA II
leading to phosphorylation of GRK2 [37] and potentially also cal-
pains, which subsequent inhibition would partially protect sub-
strates such as fodrin and receptor-bound GRK2 from proteoly-
sis in such locations. On the other hand, we find that reperfu-
sion, in parallel to increased global PKA activity and S685-GRK2
phosphorylation, enhances the oxidation-dependent dimerization
status of PKA-Rlo, an event leading to PKA activation independ-
ently of cAMP and driving its localization at the myofila-
ment compartment [39,66], where a significant pool of GRK2 protein
is present via association with sarcomeric a-actinin [67]. We pos-
tulate that PKA-mediated GRK2 phosphorylation at such location
would particularly favor its subsequent degradation by nearby cal-
pains, less likely to be targeted there by the reported βAR-
PKA inhibitory phosphorylation mechanism. Interestingly, at early
reperfusion times after preconditioning we observed comparatively
higher GRK2 levels and decreased S685 phosphorylation status,
consistent with diminished oxidation-dependent PKA activation
in such conditions due to decreased ROS production [68], despite en-
hanced βAR-PKA stimulation also taking place in such conditions
[64].

The cardio-protective and pro-survival AKT pathway is consid-
ered to be up-regulated during reperfusion to compensate I/R-
promoted injury, and enhanced stimulatory AKT phosphorylation
is found in such conditions. Activation of the Akt cascade is cen-
tral to several conditioning strategies and plays a relevant cardio-
protective role within the reperfusion injury salvage kinase path-
way (RISK) pathway, whereas inhibitors of the upstream kinase
PI3K block conditioning and cardioprotection [1,4,10,11]. However,
we uncover that a concurrent and marked reduction in total AKT
protein levels is triggered by ischemia and maintained during the
first 30 min of reperfusion, resulting in an overall decrease of
global AKT activity in cardiac extracts. The fact that such changes
in total Akt levels at early times of I/R has not been investigated
in detail before may be explained by the circumstance that in
this type of studies total AKT levels are usually employed as load-
ing controls to normalize the pAkt phosphorylation status, with-
out direct comparison to other loading controls that would allow
assessing possible changes in Akt protein levels (for instance see
[11,51,69–71] However, a few reports do show an apparent de-
crease in total Akt levels compared to normoxia in I/R-related con-
ditions [26,72,73] although this fact or the mechanisms involved
were not discussed in detail.

Our results indicate that hyper-stimulation of the remaining
AKT enzyme during reperfusion might not be able to fully com-
pensate for the decay in protein levels taking place during early
I/R. Such Akt protein down modulation would thus likely coun-
teract the beneficial effects of therapeutic approaches specifically
aimed at fostering Akt stimulation, suggesting that preventing
GRK2/Pin1/Akt degradation would safeguard global Akt catalytic
and foster the efficacy of cardioprotective drugs or conditioning
strategies (see section below).

We postulate that GRK2-related changes in the Prolyl isomerase
Pin1 may provide functionally link altered GRK2 and AKT protein
levels during I/R. AKT undergoes proteosomal degradation in dif-
f erent cell types, and phosphorylation by mTORC2 at S473 can de-
crease AKT stability in addition to allowing full kinase activation
[47,74]. Of note, Pin1, a known modulator of protein folding, ac-
tivity and stability [45] and a relevant player in cardiac hypertro-
phy [48] and cardiac calcium handling [75], reportedly acts as a
key positive factor in the regulation of AKT stability in cancer cells
by binding to Ser473-activated AKT and protecting it from degra-
dation by the proteasome, so decreased Pin1 expression facilitates
AKT degradation [46,47]. Pin1 can also be degraded by the pro-
teaseome [76,77], and decreased expression takes place in liver I/R
by unknown mechanisms [78]. GRK2 controls Pin1 stability and
functionality in cancer cells by modulating its acetylation status
via HDAC6, so decreased GRK2 expression fosters Pin1 acetylation
and favors its degradation [30]. Consistently, we find that cardiac
GRK2 downregulation during I/R correlates with a marked reduc-
tion in total Pin1 levels, along with its enhanced acetylation. Such
decrease in this AKT stabilizing factor would in turn facilitate the
observed concurrent AKT degradation.

Supporting a potential clinical relevance of our findings, a pat-
tern of concomitant down-modulation of GRK2/AKT/Pin1 protein
levels in early stages of cardiac I/R was also observed in pigs,
a very relevant large animal model used in translational studies
[28,51,79]. One limitation of our study is the relatively low number
of animals per condition, arising from the logistic complexity of
this type of experiments, including comparison of expression levels
during myocardial ischemia and different early time points follow-
ning reperfusion, as well as pre- or post-conditioning conditions
in the rat model. However, the fact that the key changes reported are
observed at different time points, in independent experiments in
other rat cohorts using calpain and proteasome inhibitors and in a
large animal pig model support the main conclusions of this work.

Consistent with this notion of related proteolysis events of
GRK2 (via both proteasome and calpains), Pin1 and AKT (mostly
via proteasome) during I/R, the combined inhibition of these
degradation pathways prior to I/R fully prevents the degradation
of GRK2, Pin1 and AKT, rescues global AKT functionality in rat
cardiac extracts, and significantly decreases infarct size and LDH re-
lease. Both calpain [42–44] and the proteasome [80,81] have been
reported to target a variety of cardiac substrates, including com-
ponents of several signaling cascades. The susceptibility of specific
substrates for degradation may vary depending on their subcel-
lular localization, whereas different proteasome inhibitors inhibit
cardiac proteasome subtypes to a different extent [82]. Thus, it is
likely that the combined effect of SNJ-1945 and bortezomib may
alter the levels of other cardiac signaling pathways in addition to
the GRK2/Pin1/AKT axis. However, the fact that the protective ef-
cfect of these inhibitors in I/R-induced cardiac damage is lost in the
presence of an inhibitor of the PI3K/Akt pathway strongly suggests
that preventing GRK2/Pin1/AKT degradation in cardiac cell types at
early times in I/R and the parallel rescue of global AKT functionality
play a relevant role in helping reduce myocardial injury.

Our observation that AKT protein levels fall during early I/R,
resulting in an overall decrease of the functionality of this key car-
dioprotective pathway despite the stimulated status of the remain-
ing AKT upon reperfusion puts forward a strong impediment for
 treatment approaches aimed only to enhance AKT activation sta-
tus, as is the case for a variety of conditioning and pharmacologi-
cal strategies [1,4,9–11], and suggest that treatments using a com-
bination of calpain and proteasome inhibitors at the time of flow
restoration in order to inhibit GRK2 and AKT degradation would
foster the efficacy of endogenous cardioprotection, cardioprotective
drugs or conditioning strategies.

In addition to helping preserve global AKT functionality, shield-
ing GRK2 from degradation during early reperfusion might also
protect against malignant ventricular tachyarrhythmias [53–55]
and adrenergic-mediated myocardial injury [38]. However, given
the reported increase in GRK2 expression at later time points after
MI and their overall maladaptive role in cardiac remodeling
and progression to HF [12,13,15,16,23], the therapeutic win-
dow for strategies aimed at decreasing GRK2 degradation should
be carefully established. In mouse models, genetic GRK2 ablation
10 days after MI [83] or treatment with the GRK2 catalytic in-
hibitor paroxetine initiated 2 weeks after MI [22] improved cardiac
function and reduced the adverse remodeling of ischemic nature. In this regard, permanent down-regulation of GRK2 in cardiogenic expression of GRK2 at the onset of MI aggravates the injury caused by I/R, reportedly related to pro-apoptotic signaling triggered by enhanced targeting of GRK2 to the mitochondrial compartment, likely initiated during the ischemic phase of MI injury, as GRK2 mitochondrial shuttling depends on S670 phosphorylation, which is induced by ischemia. Although such data are consistent with a maladaptive role of high GRK2 levels in the long term, these models do not recapitulate the dynamic changes in GRK2 levels occurring at different phases of ischemia/reperfusion/remodeling and do not rule out a protective role for timely prevention of GRK2 degradation at early points of reperfusion. Interestingly, adenosinergic delivery or cardiac transgenic expression of the C-terminal region of GRK2 (βARKct construct, aa 495–689) has a clear cardio-protective effect in both HF and acute MI experimental settings in murine or porcine preclinical models, by mechanisms involving prevention of the interaction and activation of GRK2 by Gβγ subunits released upon GPCR activation and interference with GRK2 phosphorylation at Ser670 and maladaptive targeting to mitochondria (reviewed in [13,21,23]). In other cell types, expression of βARKct also inhibits phosphorylation of endogenous GRK2 at Ser-670 by CDK2 and Pin1/GRK2 association, thus preventing down-regulation of endogenous GRK2 during cell cycle progression [31]. It is thus tempting to suggest that in the cardiac context an additional therapeutic effect of the ectopic expression of βARKct (which bears both the Ser670 and Ser685-related GRK2 phosphodegrons, highly conserved across species) would be to compete with endogenous GRK2 in the phosphorylation by the respective kinases and the action of the degradation machinery during I/R. Therefore, βARKct would contribute to the stabilization of the full-length endogenous GRK2 protein and thus of its cardioprotective Pin1 and AKT partners in the early phase after acute myocardial injury, while still acting as a modulator of GRK2 subcellular distribution and as a potent inhibitor of GRK2 functionality once the maladaptive increase in GRK2 takes place at later stages.

In sum, our data uncover dynamic changes in the central GRK2 and AKT cardiac signaling nodes at early stages after I/R, and suggest that the timely use of calpain and proteasome inhibition in these contexts may be an amenable way to reinforce therapeutic strategies aimed at reducing I/R injury.

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Author contributions

PP and JI designed and performed experiments, prepared figures, analyzed data, wrote and revised manuscript, ARS provided cardiac pig samples and analyzed data, PR performed experiments, analyzed data and prepared figures, DG-D and FM Jr., provided study concept and design, interpreted experiments, analyzed data, supervised the study, wrote and revised manuscript.

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Declarations of competing interest

The authors have nothing to disclose.

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.ebiom.2019.09.019.

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