Neurodegeneration caused by LRRK2-G2019S requires Rab10 in select dopaminergic neurons

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Abstract

Inherited mutations in the LRRK2 protein are the commonest known cause of Parkinson’s, but the molecular link from increased kinase activity to pathological neurodegeneration remains to be determined. *In vitro* (biochemical and cell culture) assays led to the hypothesis that several Rab GTPases might be LRRK2 substrates. Here we show that Rab10 potently modifies LRRK2-G2019S mediated electrophysiological responses in an *in vivo* screen, in which each Rab was overexpressed in *Drosophila* dopaminergic neurons. We therefore tested the effect of Rab10 loss of function on three LRRK2-G2019S phenotypes (vision, movement and sleep) that rely on dopaminergic circuits in both flies and mammals. The knock-out of Rab10 *in vivo* fully rescues the reduced responses induced by dopaminergic LRRK2-G2019S in visual and motor (reaching, proboscis extension) assays, but the sleep phenotype is unaffected. We show that Rab10 is expressed in dopaminergic (tyrosine hydroxylase positive) neurons controlling vision and proboscis movement, but undetectable in those controlling sleep, indicating that anatomical and physiological patterns of Rab10 are related. Our results support the idea that LRRK2 phosphorylates separate targets in distinct neurons and confirm that one degenerative pathway starts with Rab10. Although Rab3 is another putative substrate of LRRK2, it shows no synergy with G2019S and localises to a different subset of dopaminergic neurons from Rab10. We propose that variations in Rab expression may contribute to differences in the rate of neurodegeneration seen in different dopaminergic nuclei in Parkinson’s.
Significance Statement

A key question in Parkinson’s is why dopamine neurons die particularly fast in some parts of the substantia nigra. We focused on the commonest Parkinson’s-related mutation, LRRK2-G2019S. *In vitro* assays suggested that neurodegeneration may start by LRRK2-G2019S increasing phosphorylation of Rab10. We found Rab10 in fly dopamine neurons in visual and motor pathways, but not in the sleep system. Rab10 knock-out rescues G2019S-induced visual and movement degeneration, leaving sleep dysfunction unaffected. Thus, LRRK2 activates at least two pathways, one Rab10-dependent, leading to neurodegeneration *in vivo*. Rab3 is found in a different subset of dopaminergic neurons and shows no synergy with LRRK2-G2019S. We propose that variations in Rab expression contribute to differences in neurodegeneration seen in Parkinson’s.
Inherited mutations in the LRRK2 (Leucine-rich-repeat kinase2) protein are the most common known cause of Parkinson’s. A single amino-acid change, G2019S, increases LRRK2 kinase activity (1), leading to a toxic cascade that kills dopaminergic neurons, but the main steps in this pathological signalling pathway remain to be determined. A number of studies highlighted a diverse range of >30 proteins that might be phosphorylated by LRRK2, suggesting it is a generalised kinase (2). However, several research teams have recently reported that LRRK2 is a more specific kinase, phosphorylating Rab GTPases (3–8). However, it is not clear which of the more than 60 Rabs are actually phosphorylated in vivo. Here, we use a Drosophila model to address the link from LRRK2 to Rabs physiologically in dopaminergic neurons, as these are lost in Parkinson’s.

Rabs are a plausible LRRK2 substrate leading to neurodegeneration, as they act as molecular switches interacting with a range of proteins (GEFs, GAPs and GDI) regulating supply and delivery of cargo to membranes. Indeed many of the 66 Rabs in humans have been linked to neurodegenerative disorders (9). Mutations in Rabs 29 and 39 cause Parkinson’s (10–12).

Biochemically, about 8 seem to be directly phosphorylated by LRRK2 [Rabs 3, 5, 8, 10, 12, 29, 35 and 43 (3)]. In mammals, analysis of the role of the Rabs is complex because individual Rabs may have similar, or even compensatory functions, which may differ by tissue (8, 13). The situation is simpler in the fly, because there are fewer Rabs - only 23 mammalian orthologs.

Furthermore, flies provide the well-developed GAL4-UAS system for neuron-specific gene manipulation, with the Tyrosine Hydroxylase (TH) GAL4 specific for dopaminergic neurons. This permits us to analyse the effects of Rab10 manipulation in three systems: vision, a reaching movement task (proboscis extension) and sleep. We selected these systems as they are dopaminergic in both flies and mammals, and because the physiological and degenerative consequences of LRRK2 manipulations can be accurately quantified using robust, highly sensitive assays (14–16).
For vision, we use the Steady State Visual Evoked Potential (SSVEP) assay based on that developed to analyse visual problems in humans. SSVEP measures the output of the photoreceptors and second-order lamina neuron. It provides a much more sensitive assay than conventional flash electroretinograms with less impact from movement or electrical interference. We found that a number of Parkinson’s- mimic flies showed increased activity in the lamina neurons at 1 day old, including those expressing LRRK2-G2019S in their dopaminergic neurons using the Tyrosine Hydroxylase GAL4 (THG2 flies) (15, 17). Old THG2 flies show loss of visual response, which is accelerated by a mild visual stress, achieved by keeping the flies in a ‘disco-chamber’ where the light is turned off every ~2 s (18). Our proboscis extension assay was developed because a single dopaminergic neuron, TH-VUM, modulates the extension reflex (19) and we found that THG2 flies show bradykinesia and tremor (14). Sleep – defined as a 5-minute period of inactivity – is routinely monitored by the Drosophila activity monitor, and THG2 flies show aberrant sleep in 12:12 Light-Dark (LD) cycles.

Using these tools we tested if the LRRK2 ⇔ Rab pathway(s) reported in tissue culture also occur in vivo, focusing on functional outputs of dopaminergic neurons. If so, is LRRK2 specific to one or two Rabs, or is LRRK2 a more generalized kinase? Is any one Rab essential?

We determined that Rab10 has the strongest synergy with LRRK2-G2019S, Rab3 the weakest, and that each cluster of dopaminergic neurons expresses a different subset of Rabs. Dopaminergic Rab10 is essential for THG2 visual and reaching phenotypes, but sleep phenotypes are independent of Rab10. This is correlated to high levels of Rab10 in the dopaminergic neurons associated with vision and proboscis extension, whereas Rab10 was not detected in dopaminergic neurons linked to sleep.

**Results**
A Visual Expression screen identifies that Rab10 has the strongest genetic interaction with LRRK2; Rab3 the weakest

In young flies where Tyrosine Hydroxylase GAL4 is used to express UAS-LRRK2-G2019S (THG2) the visual response in lamina neurons is much increased (15). This led us to screen flies with each Rab ± G2019S and record their visual response by SSVEP.

We found that dopaminergic expression of most Rabs (TH > Rab) phenocopies THG2 (Fig. S1G), with a strong increase in visual response. This is true for both the photoreceptors (mean response 1.95 x THG2), and the lamina neurons (11.0 x THG2). For most Rabs, co-expression with LRRK2-G2019S (THG2 > Rab) further enhances the visual response (photoreceptor 2.8 x; lamina neuron 14.5 x THG2; Fig. S1G, Table S1).

We next test if any Rab disrupts the neuronal feedback circuit between photoreceptor and lamina neurons. Normally, as the photoreceptor response increases, so does the lamina neuron response. This relationship is apparent throughout our screen (Fig. 1A), being remarkably similar in young (day 1) and older (day 7) flies. However, there is a one marked outlier, Rab10, where the lamina neuron response at day 1 is ~5 times the value expected from the regression, and at day 7 is substantially below the line. Thus, in young THG2 > Rab10 flies there is much greater neuronal activity than expected, but in 1-week old THG2 > Rab10 flies we observe reduced activity, suggesting neurodegeneration has begun.

Next, we tested if there is genetic interaction between Rabs and LRRK2-G2019S. For some Rabs (10, 14, 27, 26) expression of both G2019S and the Rab in dopaminergic neurons leads to a big increase in the lamina neuron response (Fig. 1B). Interestingly, these Rabs have little effect when expressed alone. The converse is also true: for the Rabs with the biggest effect (3, 32, 1), adding G2019S has no further effect.
We wanted to examine which factors controlled this synergy. A number of hypotheses have been put forward in the LRRK2/Rab literature. First, LRRK2-G2019S is thought to phosphorylate only some Rabs \textit{in vitro}, possibly due to a preference for Thr over Ser (20). Neither \textit{in vitro} evidence for phosphorylation of the Rab by LRRK2, nor the amino-acid at the active site affects the regression (Fig. S2A,B). Nor is the relationship affected by the phylogenetic similarity of the Rabs (Fig. S2C). However, Rabs previously linked to Parkinson’s (21) have a stronger response to G2019S than others (Fig. S2D). Indeed, the Rab furthest above the regression line is one that causes Parkinson’s, \textit{Rab39} (10).

We now ask if the synergy seen with joint expression of \textit{Rab10} and G2019S could arise from phosphorylation of fly Rab10 by \textit{LRRK2-G2019S}. The sequence of human and fly Rab10 is highly conserved, both where LRRK2 and a phospho-antibody to hRab10 bind, so that the antibody recognises fly pRab10 (Fig. S3). When G2019S is expressed pan-neuronally, a stronger pRab10 signal is seen than when a kinase dead form of \textit{LRRK2} (K1906M-G2019S) is expressed. Thus, we conclude that \textit{Rab10} shows the strongest synergy with \textit{LRRK2-G2019S} in our visual electroretinogram assay, most likely due to direct phosphorylation by LRRK2. Contrastingly, Rab3 expression exhibits no synergistic interactions. This is an important difference, because both Rab10 and Rab3 are reported to be phosphorylated by LRRK2 \textit{in vitro}.

\textit{A novel Rab10 null}

Since \textit{Rab10} was the top hit in our overexpression assay, we wanted to test if reduction of \textit{Rab10} affected retinal degeneration. We generated a \textit{Rab10} null by deleting the coding region from just after the start of the GTPase domain to the end of the protein using CRISPR/Cas9 (Fig. S3). The deletion includes the LRRK2 binding site. The null mutant (\textit{Rab10}⁻) is viable, unlike that of the mouse (22).
We tested for a Rab10\textsuperscript{−} phenotype in visual assays. When kept in the dark at 22 °C, Rab10\textsuperscript{−} flies had a normal flash electroretinogram (Fig. S4). However, flies kept in the dark at 29 ° and tested in the more sensitive SSVEP assay have a larger neuronal response than their matched control (Rab10\textsuperscript{−}; TH and TH/+; Fig. S5); those Rab10\textsuperscript{−}; TH flies kept in the disco-chamber to accelerate neurodegeneration have much worse visual response than controls (Fig. 2D). Dopaminergic expression of Rab10 RNAi does not affect the vision of flies kept in the dark (Fig. S5) but does accelerate the loss of lamina neuron function in the disco chambers (Fig. 2D, compare TH; Rab10 RNAi with TH/+). These experiments suggest dopaminergic Rab10 is important for maintenance of visual response to light in aged flies.

Global or dopaminergic reduction in Rab10 rescues G2019S-induced anatomical deficits

The key question is how Rab10 reduction affects G2019S-induced neurodegeneration. We started by measuring retinal structure anatomically, using the ‘Deep-Pseudo-Pupil’ (DPP) assay, comparing flies raised in the dark those kept in a disco-chamber to accelerate neurodegeneration (Fig 2A). Dark-reared flies show a DPP, since the normal retinal structure focuses light from below towards the observer through 7 ommatidia (23), (Fig. 2Ci). Disco flies expressing LRRK2-G2019S in the dopaminergic neurons (THG2) lose pigment and their DPP by 10 days, with disorganized patterns of light and dark across the surface of the eye (Fig. 2Ci). The lack of DPP in disco-reared THG2 flies indicates that light is scattered randomly within the eye since the internal retinal structure is lost. However, in the Rab10 null (Rab10\textsuperscript{−}; THG2), the DPP is preserved, showing Rab10 knock-out rescues the G2019S-induced loss of retinal structure.

As global Rab10 knock-out protects the structure of the eye from G2019S-induced degeneration, we next asked if dopaminergic reduction of Rab10 is sufficient to rescue the structural deficit. We used the deGradFP technique...
(Fig. 2B) to achieve dopamine-specific Rab10 knock-down. For this, we took flies in which GFP has been inserted at the start of the Rab10 gene (Rab10 GFP) (24). We depleted Rab10 GFP protein specifically in dopaminergic neurons by targeted expression of a GFP-targeted nanobody (vhhGFP). This marks GFP-proteins for proteolysis (25, 26). Flies with Rab10 GFP, nanobodies and G2019S show normal pigmentation and DPP whether kept in the dark or in a disco-chamber. This shows that depletion of Rab10 protein is sufficient to protect the flies from G2019S-induced degeneration. As a control, flies with the nanobody and G2019S, but with wild-type Rab10, lose pigmentation and DPP, as the nanobody does not bind to the normal Rab10.

These experiments suggest that knock-down of Rab10 in dopaminergic neurons plays a role in protecting against the LRRK2-G2019S induced neurodegeneration.

Global or dopaminergic reduction in Rab10 rescues G2019S-induced visual function

To consolidate the anatomical results, we tested the electrophysiological response of the eyes. All dark-reared flies show a strong lamina neuron signal – at least >50% of the control (Fig. S5). The response is less in flies with multiple transgenes due to their darker eyes. Disco-reared flies show strong effects of genotype: the THG2 genotypes in three control backgrounds all have very poor vision, < 10 % of dark-reared progeny of the same cross (Fig. 2D, magenta bars). In the global Rab10\(^-\) null background, this response is fully rescued (Fig. 2D compare green and red bars). Thus, both electrophysiological response and structural observation of the DPP indicate that Rab10 null rescues the effect of dopaminergic LRRK2-G2019S.

Since global knock-out of Rab10 rescued the G2019S phenotype, we next asked if dopaminergic-neuron-specific knock-down of Rab10 is enough to rescue the visual deficits induced by G2019S in disco-chamber flies. The first strategy was to use a Rab10 specific RNAi driven by TH-GAL4. In these flies (THG2; Rab10 RNAi Fig. 2D, yellow bar) the lamina neuron response is
completely rescued. To confirm that Rab10 knock-down rescues G2019S mediated degeneration, we used the flies with Rab10 GFP and depleted the mRNA or protein using GFP RNAi or vhhGFP expression. These flies (Rab10 GFP; THG2; GFP RNAi or Rab10 GFP; THG2; vhhGFP Fig. 2D brown bars) show much larger lamina neuron visual responses than the corresponding controls with wild-type Rab10 where there is no GFP, and so the Rab10 is fully functional. Thus, all three strategies indicate that dopaminergic Rab10 is essential for the G2019S-induced loss of physiological response, as well as anatomical degeneration.

**Rab10 reduction ameliorates a movement LRRK2-G2019S phenotype**

The role of Rab10 in a visual phenotype lead us to ask if Rab10 is essential for all LRRK2-G2019S phenotypes. Dopaminergic signalling is important in movement, including a reaching movement used to obtain food, the Proboscis Extension Response (PER, Fig. 3A). Expression of G2019S in a single dopaminergic neuron, TH-VUM (19) results in movement deficits – akinesia (loss of the PER, Proboscis Extension Response), slower response (bradykinesia) and tremor (14). When the PER is elicited by an optogenetic stimulus, or by sucrose application, dopaminergic knock-down of Rab10 fully rescues the G2019S-induced akinesia (Fig. 3B, Fig.S6). Additionally, Rab10 reduction ameliorates bradykinesia, reverting the slow extension seen in G2019S flies (Fig. 3C). This was not due to a deficit in extending the proboscis as all genotypes extended their proboscis to the same extent (Fig. 3Ci).  

Dopaminergic expression of Rab10 phenocopies G2019S expression, reducing the proportion of flies that respond (Fig. S6).

These experiments show that Rab10 reduction ameliorates movement, as well as visual deficits induced by LRRK2-G2019S.

**Rab10 reduction fails to ameliorate a ‘sleep’ LRRK2-G2019S phenotype**
Dopaminergic neurons also influence circadian rhythms and ‘sleep/wake’ patterns (27, 28). At least three clusters of dopaminergic cells (PAM, PPL1, PPM3) contribute to this phenotype, though the exact pathways have yet to be fully determined. In 12:12 light on:off cycles, THG2 flies show a marked sleep deficit (Fig.4). In the dark, they spend less than 50% of the time ‘asleep’ than control flies do, but in the light they sleep more. Manipulation of Rab10 has no effect on sleep (Fig. 4) but gives subtle changes with the lights off responses of these flies (Fig. S7A). Dopamine level has little effect on circadian period (29) and neither LRRK2-G2019S nor Rab10 manipulations affects the circadian period of flies in constant darkness (Fig. S7B).

Thus, we find that Rab10 reduction does not rescue the sleep phenotype, unlike the visual and proboscis extension phenotypes.

*Rab10 is only found in a subset of dopaminergic neurons*

To examine the overall distribution of Rab10, we used Rab10-GAL4 (30) to express a nuclear RFP, and α-TH antibody to mark the dopaminergic neurons. We found that many, but not all dopaminergic neurons were Rab10 positive (Fig. 5A). Rab10 positive neurons include the small MC neurons in the optic lobes (Fig. 5Aii), the TH-VUM neuron (that controls the PER) and DADN neurons (control leg movements) (Fig. 5Aiii). Coincidence was also observed in some PAL, PPM3, PPL2, and T1 neurons. However, other neurons, notably those in the PPL1, PPM2 and PAM clusters did not show Rab10 driven RFP. The PPL1 and PAM neurons have been linked to sleep/wake patterns (27, 28). To confirm this data, we adopted an intersectional strategy, based on split GFP, whereby dopaminergic *TH-LexA* and *Rab10-GAL4* were used to deliver the two halves of GFP. Only neurons which are both dopaminergic and Rab10+ will generate GFP and fluoresce. This delivered the same pattern of coincidence (Fig. 3Aiv). A different subset of dopaminergic neurons is Rab3 positive (Fig. S8).
We next asked if Rab10 was indeed phosphorylated in any dopaminergic neurons. In the fly brain, the phospho-Rab10 antibody marks 3 clusters of neurons along the dorsal edge of the brain: a lateral pair and a medial cluster (Fig. S8A,B, white arrows). Also stained are a pair or ventral neurons, close (but not identical) to the dopaminergic TH-VUM and DADN (Fig. S8A,B, yellow arrows). None of these phospho-Rab10 cells were marked by the α-TH antibody used to identify dopaminergic neurons. No cells were strongly marked in the Rab10− flies (Fig. S8C, one cell was weakly stained, grey arrow), showing the specificity of the antibody.

Thus, we conclude that some dopaminergic neurons natively express Rab10, notably those concerned with vision and Proboscis Extension - while in others the level of Rab10 is too low to be detected – those controlling ‘sleep’. Only a few of the Rab10+ neurons show high levels of phospho-Rab10.

**Discussion**

Our key observation is that knock-out of Rab10 in vivo fully rescues the reduced responses induced by dopaminergic LRRK2-G2019S in visual and motor (reaching, proboscis extension) assays, but the sleep phenotype is unaffected. Rab10 is expressed in dopaminergic neurons controlling vision (MC) and proboscis movement (TH-VUM), but undetectable in those controlling sleep, so that the anatomical and physiological patterns of Rab10 are related. Our results support the idea that LRRK2 phosphorylates separate targets in distinct neurons.

The existence of Rab10 in the tyrosine hydroxylase positive neurons controlling vision and proboscis movement argues that LRRK2 might indeed phosphorylate Rab10 directly. Thus our in vivo results both support the in vitro (biochemical and cell culture) data in which LRRK2 directly phosphorylates Rab10 (3–8), but also suggest a separate pathway for LRRK2-G2019S action. Further, while Rab3 is also phosphorylated by LRRK2 in vitro
(3), it did not synergise with LRRK2-G2019S in the visual assay, and was not detected in the MC neurons.

In the visual system, we confirmed dopaminergic rescue of both anatomical and physiological deficits using three different genetic strategies. In the akinesia movement assay, dopaminergic Rab10 knock-out (two strategies) rescues the G2019S deficit. In both systems, rescue is back to wild-type levels, identifying Rab10 as the key component. However, we have noted that in human peripheral mononuclear blood cells, LRRK2 phosphorylates both Rab10 and Rab12 (6). Since the fly has no Rab12, with Rab10 being the closest homolog, we cannot exclude a G2019S ⇔ Rab12 relationship in mammalian dopaminergic neurons.

The effects of LRRK2-G2019S and Rab10 on vision and proboscis extension could be accounted for by a reduction in dopamine release. Dopamine inhibits photoreceptors (31) and excites proboscis extension (19). This fits with the observation, in worm, that Rab10 is required for release of transmitter from Dense Core Vesicles (DCVs) (32) and we note that dopamine is packaged in DCVs. In mammals, Rabs 10, 14 facilitate delivery of GLUT4 vesicles to the plasma membrane in adipocytes (13). However, a developmental effect cannot be excluded as Rab10 modulates neuronal growth (34, 35).

Expression screen highlights synergy between Rab10 and G2019S

A wider view of possible LRRK2 ⇔ Rab pathways is provided by the visual screen. All Rabs, either by themselves or when co-expressed with G2019S, increase the visual response. One possible physiological mechanism is that Rab overexpression reduces retinal dopamine release onto the photoreceptors. This will increase the amplitude and speed of the photoreceptor response (31). Dopamine may also affect the lamina neurons, and third order MC cells, but this remains to be determined. It is also possible
that the Rabs modulate the release of co-transmitters or growth factors from
dopaminergic neurons.

Our screen placed the Rabs along a spectrum, ranging from those with a
strong synergy with G2019S to those which had a strong effect when
expressed by themselves.

Rabs 10, 14 and 27 have the strongest synergy with G2019S, though by
themselves they have little effect on visual sensitivity. These three Rabs are
best known for their roles in exocytosis. However, another function for Rab10
is suggested by the stand-out, large lamina neuron: photoreceptor response
ratio when G2019S and Rab10 are expressed together. This might the role of
Rab10 in endocytosis (36–39). Here, the effects of Rab10 are mediated through
the EHBP1-EHD2 complex. In the follicle cells of Drosophila, ehbp1 expression
and knockdown phenocopy Rab10 manipulations (40), while EHBP1 was also
identified by a systematic proteomic analysis as indirectly phosphorylated by
LRRK2 in HEK293 cells (3) and a lysosomal assay (41).

Among the Rabs which have strong visual effect when expressed in
dopaminergic neurons, but show little synergy with G2019S are 1, 3, 5, 6 and
11. Rab3 is notable for its role in synaptic transmission, while the other Rabs
here play roles in the fly optic system in intracellular cycling [see for review
(42)]. Indeed, Rab3 was at the opposite end of the Rab spectrum from Rab10 –
this is notable because in vitro mammalian cell assays have highlighted
similar roles of Rabs 3 and 10 in lysosome exocytosis, (43, 44), as well the
observation that both phosphorylated in HEK cells (3). On explanation for the
discrepancy between the in vivo and in vitro assays is that Rab3 is localised to
synapses, possibly far from LRRK2 at the trans-Golgi network (7).

Our observation that every Rab seems to have some effect on dopaminergic
signalling in the visual system goes some way to explain why many studies of
individual Rabs have demonstrated effects with LRRK2; Rab3a (45); Rab5
(46); Rab7 (47); Rab29 (11). Although cellular studies support an binding of
Rab29 to LRRK2 (48), the closest fly homolog (Rab32) shows little synergy with G2019S in our screen.

The availability of Rab transgenic flies facilitates screening in *Drosophila*. Screens have identified key roles for Rab2 in muscle T-tubule development (49); Rabs 2, 7, 19 in loss of huntingtin (50), 1, 5, 7, 11 and 35 in the *Drosophila* renal system (51), Rab32 in lipid storage (52) and Rab39 in tracheal formation (53). The varied outcomes of these screens indicate the validity of the LRRK2-G2019S ⇔ Rab10 relationship reported here.

*Each dopaminergic neuron has its own palette of Rab expression*

Finally, we note that not all dopaminergic neurons are equally susceptible in Parkinson’s. A long-standing observation is that the dopaminergic neurons in the VTA (ventral tegmental area) do not degenerate in the same way as those in the *substantia nigra*. More particularly, even within the *substantia nigra* there is a range of outcomes, with dopaminergic neurons in the pars compacta dying more than those in the dorsal and lateral zones (54). The same is true for fly neurons: the neurons in the PPM1 and PPM2 degenerate more than those in other clusters (though no data is available for the MC and TH-VUM neurons which are our focus). Previously, faster neurodegeneration has been ascribed to increased cytosolic dopamine levels (55), to intracellular effects of glutamate (56), to increased calcium influx (57), through more action potentials (58), or to longer axons with more synapses (59). It has not escaped our notice that faster degeneration in some neurons may be the result of their different palettes of Rab proteins.

**Materials and methods**

*Flies* (*Drosophila melanogaster*) were raised and manipulated according to standard fly techniques. Data in Figs. 1 and S1 is from female flies; all other data from male flies.
A new Rab10 was generated by CRISPR-mediated mutagenesis by WellGenetics Inc. using modified methods by (60). A 1644-bp region, spanning from +57 nt relative to ATG of Rab10 to +70 nt relative to the 1st nt of Stop of Rab10, was replaced by a STOP-RFP knock-in cassette, deleting most of the coding region.

Briefly, a pair of gRNAs (upstream: CTGATCGGTGATTCAGGAGT, downstream: GAACGGGGCGTGGTTTGGCC) was cloned into a U6 promotor plasmid. For repair, a STOP-RFP cassette containing 3xSTOP codon and 3xP3-RFP and 1-kb homology arms were cloned into a pUC57-Kan donor plasmid. Wild-type w1118 embryos were microinjected with Rab10-targeting gRNAs and hs-Cas9 supplied in DNA plasmids, together with the donor plasmid. Flies carrying the positive selection marker 3xP3-RFP were validated by genomic PCR and sequencing. For the upstream PCR, primers were designed at 5’ outside of upstream homology arm (Fwd primer 5’- GATGTTCAGGCGATTCTGGT) and at 3xP3 promotor (Rev primer 5’- CGAGGGTTCGAAATCGATAA). For the downstream PCR primers were designed at alpha-tubulin 3’ UTR, which is used for STOP-RFP transcript stability (Fwd primer 5’- AACGCAAGCAAATGTGTCAG) and at 3’ outside of downstream homology arm (Rev primer 5’- CGATGATTGTTTGCATTG).

Fly stocks are listed in Table S6.

**Visual assays:** On the day of emergence, flies were placed in the dark or in disco-chambers at 29 °C. 1 day or 1 week old flies were prepared for SSVEP (Steady State Visual Evoked Potential) measurements as described (Fig. S1) (15). The same protocol was used, except that stimuli were generated, and responses recorded by an Arduino Due system instead of a PC. Data was analysed in Matlab and R. Full code at [https://github.com/wadelab/flyCode](https://github.com/wadelab/flyCode). DPP (Deep-Pseudo-Pupil) assays were performed on a Zeiss Stemi microscope with AxioCamERc5s camera.
Western Blots were made using standard protocol and antibodies: α-phospho-Rab10 (Abcam: ab230261, 1:1000) with α-synaptotagmin 91 as a loading control (61). Control Rab10 samples were derived from female Wistar rats (Charles River UK), maintained in accordance with the UK Animals (Scientific Procedures) Act (1986). Animals were euthanised via exposure to CO2 in a rising concentration, followed by cervical dislocation in accordance with Home Office guidelines and with approval of the Biology Ethics Committee (University of York). The brain was dissected into PBS and 10 mg of tissue lysed for the blots (62).

Immunocytochemistry was performed as described recently (14). Tyrosine hydroxylase was detected with Mouse anti TH Immunostar (22941, 1:1000), and phospho-Rab10 (Abcam: ab230261, 1:250).

Akinesia was recorded from 1 week old flies, kept in the dark at 29 °C. Flies were restrained as described (14) and starved at 25 °C for 3 hours before being offered a droplet of 100 mM sucrose three times. Each response was scored Yes/No and the median response for each sample used. χ² with post-hoc tests used the R package ‘fifer’.

Bradykinesia was assessed using an optogenetic stimulus. Flies were fed retinal (1 mM) pipetted onto the surface of their food for 1 week in the dark at 29 °C. They were restrained at 25 °C for 3 hours before the proboscis extension responses were observed with a Grasshopper 3 (Point Grey) camera mounted on a Zeiss Stemi microscope at 200 frames/second. A single flash was delivered from a ThorM470L3 LED, driven at 8 V for 7 ms. The stimulus was transcoded by LexOpReachR expressed in the Gr5a neurons. The area of the video occupied by the proboscis was automatically analysed by python code https://github.com/biol75/PER, and the statistics in R.

Circadian activity patterns were recorded with a TriKinetics DAM system, in dim light at 29 °C and analysed using the ShinyR-DAM code (63) using the branch at https://karolcichewicz.shinyapps.io/ShinyR-DAM_3_1_Beta/.
Statistics were calculated in R, with the mean ± SE reported by error bars or median ± interquartile range in box plots. Post-hoc tests were calculated for χ² using the fifer package and for ANOVA, the Dunnett test.
Figure Legends

Fig. 1. Expression screen highlights **Rab10 with the strongest synergy with LRRK2-G2019S, and Rab3 as the weakest**. Each **Rab** was expressed in dopaminergic neurons (using TH-GAL4) by itself (**TH > Rab**), or along with **G2019S (THG2 > Rab)** and the visual response measured after 1 day or 7 days in the dark. A. **Standout role of Rab10**. The response is split into two components, representing the photoreceptors and lamina neurons (inset blue and purple). The increase in lamina neuron response is highly correlated with the response of the photoreceptors, with one outlying exception, **Rab10** at 1 day. B. **Rab10 has the strongest synergy with G2019S**. Relationship of **Rab** and **G2019S** showing their inverse relationship. Rabs (3, 32, 1) which have a big effect on vision when expressed on their own have little further consequence when **G2019S** is also expressed; but other Rabs (10, 27, 14, 26) which have little visual impact on their own have a strong synergy with **G2019S**. Details in Figs. S1, S2 and Table S1.

Fig. 2. **Accelerated visual decline due to LRRK2-G2019S is rescued by dopaminergic knock-down of Rab10**. A. Accelerated visual decline was achieved by keeping flies in disco-chambers (18) compared with those in the dark. B. Destruction of Rab10 GFP protein in the deGradFP technique by dopaminergic expression (**TH-GAL4**) of a nanobody targeted at GFP (**vhhGFP**), which sends Rab10 GFP protein to the proteasome. C. **Anatomically, dopaminergic Rab10 knock-down rescues the loss of eye structure**. All flies kept in the dark for 10 days have strongly pigmented eyes, with a marked DPP (Deep Pseudo-Pupil (23)). This shows that the internal, reflective structure of the eye is intact. Flies with dopaminergic expression of **LRRK2-G2019S (THG2)** kept in disco chambers lose their pigmentation (indicative of lysosomal dysfunction) and have no DPP. This is rescued in the **Rab10 null (Ci: Rab10−/−; THG2)**, or by knockdown of Rab10 protein in dopaminergic neurons using the deGradFP technique (Cii: **Rab10 GFP; THG2**;
Physiologically, dopaminergic Rab10 knock-down rescues the neuronal vision. THG2 flies in disco chambers lose their physiological response after 7 days (magenta bars). This is rescued by global knock-out of Rab10, (Rab10−; THG2 orange bar) or by dopaminergic knock-down of Rab10 by RNAi (THG2; Rab10RNAi yellow bar). Rescue is confirmed using flies in which Rab10 had been replaced with Rab10 GFP using RNAi targeted at Rab10-GFP mRNA or using deGradFP mediated knockdown of Rab10 protein (Rab10 GFP; THG2; GFP or Rab10 GFP; THG2; vhhGFP brown bars). Control experiments show that Rab10 knock-out (Rab10−; TH) or dopaminergic knock-down (TH > Rab10 RNAi) accelerates the loss of visual function. This is particularly marked for the Rab10−; TH flies, as these have a stronger visual response than the other genotypes when kept in the dark. The lamina neuron response of dark-reared flies and the neuron / photoreceptor relationship is in Fig. S5, with exact genotypes in Table S2.

Fig 3. Rab10 knock-down rescues proboscis reaching deficits induced by dopaminergic expression of LRRK2-G2019S. A. To reach for food or liquid, the fly extends its proboscis in response to a stimulus to sensory neurons on the legs. B. Dopaminergic reduction in Rab10 rescues the akinesia (less frequent response) of flies expressing G2019S in their dopaminergic neurons.

C. Dopaminergic reduction in Rab10 rescues the bradykinesia (slower response) of flies expressing G2019S in their dopaminergic neurons. Ci. Raw traces; Cii. mean data. To respond to the optogenetic stimuli all flies use LexA/Op Gr5a>ReachR. Exact genotypes in Table S3.

Fig 4. Rab10 knock-down does not rescue sleep deficits induced by dopaminergic expression of LRRK2-G2019S. A. THG2 flies show increased sleep during the day, and reduced sleep at night. Neither reduced nor increased expression of Rab10 affects the daily pattern (Ai), with summary data in Aii. Sleep is defined as periods of inactivity longer than 5 min. Actograms from these flies are shown in Fig. S5. Exact genotypes in Table S4.

Fig. 5. Rab10 and Rab3 are located in different subsets of the dopaminergic neurons. A. Not all dopaminergic neurons, identified by a cytosolic α-
tyrosine hydroxylase antibody (α-TH, green), are indicated by Rab10-GAL4 expression of nuclear RFP (magenta). Rab10 is detected in the dopaminergic MC neurons (in the optic lobe) (Aii) and in the TH-VUM (controls proboscis extension response (19)) and DADN neurons (control leg movements (64)) (Aiii). B. An intersectional strategy to express both halves of split GFP in cells using TH-LexA and Rab10-GAL4 (magenta) confirms that not all the dopaminergic neurons (green) are positive for Rab10. White arrows indicate cells marked with GFP and α-TH, including the TH-VUM and DADN neurons; green arrows cells which are dopaminergic but not Rab10. C. Summary of the dopaminergic neurons coincident with (i) Rab10 and (ii) Rab3. Diagram and nomenclature after (65, 66). Antisera and exact genotypes in Table S5.
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neuronal increase due to G2019S (as % of TH > Rab)

photoreceptor increase due to G2019S (% of TH > Rab)

Rab1 Rab2 Rab3 Rab4 Rab5 Rab6 Rab7 Rab8 Rab9 Rab10 Rab11 Rab14 Rab18 Rab19 Rab21 Rab23 Rab26 Rab27 Rab30 Rab32 Rab35 Rab39 Rab40

Synergistic effect of G2019S (% of TH > Rab)

Effect of Rab (% of wildtype)

photoreceptors

lamina neurons

A

B

day 1
day 7

0 100 300 400 500 600 700 800

day 1
day 7

0 100 300 400 500 600 700 800 900 1000

day 1
day 7

0 100 300 400 500 600 700 800 900 1000

A dark disco 24/7

B

THG2/+

C

THG2/+

D (disco)

remaining lamina neuron response

| Treatment                  | Remaining Neuron Response (% of dark flies) |
|---------------------------|--------------------------------------------|
| TH+ Rab10−, TH          | 50 ± 12                                    |
| TH−; Rab10 RNAi         | 25 ± 8                                     |
| THG2+ THG2, Rab10 RNAi  | 50 ± 11                                    |
| THG2+; GFP RNAi         | 50 ± 12                                    |
| Rab10+; THG2, vhhGFP    | 25 ± 10                                    |
| Rab10+; THG2, vhhGFP    | 25 ± 10                                    |

Significance levels:

- * p < 0.05
- ** p < 0.01
- *** p < 0.001
B CiA

ReachR

THG2; Gr5a>ReachR N=28
THG2; Gr5a>ReachR N=24
THG2; Rab10RNAi; Gr5a>ReachR N=13

Ci

proboscis extension (% of control)

Gr5a>ReachR N=28
THG2; Gr5a>ReachR N=24
THG2; Rab10RNAi; Gr5a>ReachR N=13

mean time to 75% extension (s)

Gr5a>ReachR
THG2; Gr5a>ReachR
THG2; Rab10RNAi; Gr5a>ReachR

Cii

**

0.25

0.2

0.15

0.1

0.05

0

time (s)

0

0.2

0.4

0.6

0

0.2

0.4

0.6

proboscis extension (% of control)

Gr5a>ReachR N=28
THG2; Gr5a>ReachR N=24
THG2; Rab10RNAi; Gr5a>ReachR N=13

mean time to 75% extension (s)

Gr5a>ReachR
THG2; Gr5a>ReachR
THG2; Rab10RNAi; Gr5a>ReachR

**
A: Rab10
Ai: whole CNS

B: Rab10
B: whole CNS

Ci: Rab10

Cii: Rab3