Introduction

Apart from well-known bacterial methods of mutations, the horizontal genetic transfer of genetic material within microbes plays a crucial role in the emergence of novel antibiotic resistance genes and their dissemination noticed worldwide. Rapid and widespread appearance of multidrug resistance patterns observed in closely as well as distantly related bacterial species are due to presence of mobile genetic elements, more specifically integrons.

Integrons are able to capture and exchange genes in a small mobile elements called cassettes by a process of site-specific recombination. They have been assigned to at least four different classes of mobile genetic elements, more specifically integrons. Class 1 integrons usually consist of 5’-conserved segment (5’CS), 3’-CS and gene cassettes in 3’CS, but we also observe 14/69 isolates that showed amplification for 5’CS-3’CS region but did not show presence of integrons in order to illustrate the location of antibiotic resistance genes (especially $\beta$-lactam). The genotyping was done by RAPD so as to find out genetic relatedness among isolates. 55.20% (69/125) isolates were found positive for integrons. 41 isolates showed single amplification band for CS region, 20 showed two bands, 4 showed three bands and four isolates showed multiple banding patterns. Sul-1 was reported to be present in 3’CS, but we also observe 14/69 isolates that showed amplification for 5’CS-3’CS region but did not show presence of Sul-1 (when detected by PCR). Out of 109 E. coli isolates, 91 could be typed by RAPD, while 18 were found untypable. Among 91 E. coli isolates, 33 were grouped in 15 clusters while the remaining 58 isolates showed unique banding patterns indicating genetic un-relatedness. Among 16 K. pneumoniae isolates, 14 were typed by RAPD and 2 isolates were found untypable. The higher rate of resistance to several classes of $\beta$-lactam antibiotics in integron-positive isolates is probably attributable to the association of $\beta$-lactamase genes with integron-carrying plasmids and hence suggests that antibiotic drug resistance is transmitting through these mobilizing agents. As evident from RAPD-typing, most patients in our hospital were infected with different clades of organisms, thereby demonstrating clonal diversity among isolates suggesting horizontal transmission of $bla$ genes.
1610 isolates were identified as susceptibility testing during a period of 18 months and out of these, 1412 samples were subjected to the department of microbiology, J. N. Medical College for routine culture and bacterial isolates.

Materials and Methods

It has been reported in the literature that integrons can be categorized as classic and non-classic integrons. Classic class 1 integrons contain an integrase gene (intI1) in their 5’CS and qacEΔ1-Sul-1 (encoding resistance to quaternary ammonium compounds and sulfonamides respectively) in their 3’CS region [2]. Moreover, non-classic class 1 integrons lack 3’CS region from the normal integron structure composition [3-5]. The Sul-1 gene was found typically associated with class 1 integrons [2]. Sul-2 gene was reported to be presenting adjacent to streptomycin resistance genes [6], whereas Sul-3 gene has been recently illustrated to be linked to non-classic integrons [3,4,7].

While antibiotic resistance is a great emerging health issue, it is important to study the spread of antibiotic resistance genes to understand the relationship between resistance gene pool, its mobilization through transposons and integrons and dissemination among closely related as well as completely unrelated bacterial species.

Antimicrobial Susceptibility of integron-positive and integron-negative E. coli and Klebsiella pneumoniae isolates.

| Antimicrobial Agents | Integron-positive (n=69) | Integron-negative (n=56) |
|----------------------|--------------------------|-------------------------|
| β-Lactams            |                          |                         |
| Cefoperazone         | 91.43%                   | 76.36%                  |
| Cefixime             | 91.43%                   | 87.27%                  |
| Cefotaxime           | 95.71%                   | 92.73%                  |
| Ceftriaxime          | 78.57%                   | 83.64%                  |
| Cefpime              | 90.00%                   | 96.36%                  |
| Cefprome             | 65.71%                   | 61.82%                  |
| Aztreonam            | 78.57%                   | 72.73%                  |
| Amikacin             | 90.00%                   | 92.73%                  |
| Imipenem             | 0.0%                     | 0.0%                    |
| Fluoroquinolones     |                          |                         |
| Gatifloxacin         | 87.14%                   | 83.64%                  |
| Ofloxacin            | 88.57%                   | 85.45%                  |
| Aminoglycosides      |                          |                         |
| Gentamicin           | 61.43%                   | 49.09%                  |
| Amikacin             | 47.14%                   | 29.09%                  |

Characterization of integrons

Cefoxitin-resistant E. coli and K. pneumoniae isolates that were previously characterized for the presence of bla<sub>ampC</sub> genes [9] were now looked for the presence of integrons in order to illustrate the location of antibiotic resistance genes (especially bla<sub>ampC</sub>). Presence of integrons was demonstrated by PCR amplification, using primer set 5CS-F1 (5’-ATG TTA CGG AGC AGG GC-3’) and 3CS-R (5’-GGA ATT CGA CCT GAT AGT TTG GCT GTG-3’) as forward and reverse primers respectively. PCR reaction mixture was prepared in a total volume of 25 μL comprising of 0.05 μL each of primers 5CS-F1 and 3CS-R, 12.5 μL master mixture, 9.9 μL DNase/RNase free distil water and 2.5 μL of template DNA. The reaction mixture so prepared was dispensed in PCR tube and placed in MJ-mini Bio-Rad thermal cycler (Bio-Rad, USA). PCR cycling conditions consist of an initial denaturation step at 95 ºC for 3 min, followed by 30 cycles of 95 ºC for 1 min, 58 ºC for 1 min, 72 ºC for 2 min, and a final elongation step at 72 ºC for 10 min.

Genetic relatedness of cefoxitin-resistant isolates done by RAPD-PCR

RAPD-typing was done by using primer ERIC-2b (5’-AAG TAA GTG ACT GGG GTG AGG C-3’) [10]. Reaction mixture was prepared by mixing 12.5 μL of master mix, 10.25 μL of distilled water, 0.25 μL of primer and 2.0 μL of template DNA. The PCR amplification was attained by placing reaction mixture containing tubes in MJ-mini Bio-Rad thermal cycler (Bio-Rad, USA). PCR cycling conditions consist of an initial denaturation step at 94 ºC for 2 min, followed by 35 cycles of 94 ºC for 1 min, 55 ºC for 1 min, 72 ºC for 4 min, and a final elongation step at 72 ºC for 8 min. The results were analyzed by using Bio-Rad Gel documentation system (Bio-Rad, USA) and clustering was performed by Quantity One software provided by gel documentation system. Isolates of same species were typed in same batch and isolates of different wards were compared together to identify clonal spread.

Results

Antimicrobial susceptibility

All the integron-positive isolates were characterized by resistance to more than one antimicrobial agent tested as compared to integron-negative isolates. The range of the proportions of resistance for integron-positive isolates and integron-negative isolates. The range of the proportions of resistance for integron-positive isolates and integron-negative isolates for the third-generation cephalosporins (ceftriaxone, cefotaxime, ceftazidime) was 78.57%-95.71% and 83.64%–96.36% resistant, respectively. Moreover, all the isolates were found susceptible to imipenem. Details are shown in Table 1.

Characterization of integrons

41 isolates showed single amplification band when PCR was performed to amplify CS region. Out of these, three isolates showed amplification at 400 bp (lowest one), 11 isolates showed the ampiclon at 800 bp, while three isolates showed the amplification of product at 900 bp. A major part of isolates, comprising of 24 isolates showed the amplification product at 1500 bp. It was the most common ampiclon noted in our isolates. Details are shown in Table 2.

Out of three isolates that showed the ampiclon at lowest position, two isolates were found negative for Sul-1 gene. One isolate was found to carry bla<sub>ampC</sub> only along with insertion sequence IS26. The
other two isolates harbor bla<sub>ampC</sub>, bla<sub>CTX-M</sub>, bla<sub>TEM</sub> and bla<sub>SIV</sub>. As far as mobilizing elements are concerned (IS26, ISEcp1, and ORF513), the Sul-1-harbouring isolates also carried IS26 while the other one do not bear any of the insertion sequence.

Among 11 isolates showing amplification for CS region at 800 bp, nine isolates were noticed to harbor Sul-1 gene and two were found negative for the same. When analyzed for bla genes, three isolates were noted to carry combination of bla<sub>ampC</sub>, bla<sub>CTX-M</sub>, bla<sub>TEM</sub>, and bla<sub>SIV</sub>, but these three isolates carry different array of mobile genetic element. Sul-1 was noticed to be present in all three isolates, ORF513 was observed in two, and ISEcp1 was found to be present in one isolate. IS26 was also noticed in one isolate. The combination of bla<sub>ampC</sub>, bla<sub>CTX-M</sub> and bla<sub>TEM</sub> was observed in two isolates, where, IS26, ISEcp1, Sul-1, and ORF513 were present in one and the other three mobilizing elements except ISEcp1 were noticed in another isolate. Similarly, bla<sub>ampC</sub>, bla<sub>CTX-M</sub> and bla<sub>SIV</sub> combination was noticed in two isolates where ORF513 was noticed to be absent in one isolate while rest of the three mobilizing elements were observed in both isolates. The combination of bla<sub>ampC</sub> and bla<sub>CTX-M</sub> was observed in two isolates and Sul-1 was found absent in both isolates. Single isolate that harbors only bla<sub>ampC</sub> was found to carry all the four mobilizing elements. One isolate shows the amplification for CS region, but carries only ORF513 and rest of the genes including bla genes and mobile genetic elements (screened in the present study) were found absent.

Three isolates showed CS amplification band at 900 bp, out of which Sul-1 was noted only in one isolate. Sul-1-positive isolate harbors all the four bla genes studied i.e., bla<sub>ampC</sub>, bla<sub>CTX-M</sub>, bla<sub>TEM</sub> and bla<sub>SIV</sub>. While, among mobilizing elements ORF513 and ISEcp1 were present while IS26 was found absent. In the remaining two isolates, one carries only bla<sub>ampC</sub> along with ORF513, ISEcp1, and IS26. The other Sul-1-negative isolate carries combination of bla<sub>ampC</sub>, bla<sub>CTX-M</sub>, and bla<sub>SIV</sub> whilst other mobile genetic elements were absent.

The most common amplicon that was observed in amplification of Conserved Segment (CS) region was of 1500 bp, and was noticed in 24 isolates. Sul-1 gene was observed in eighteen isolates and three isolates were noticed negative for it. Twelve isolates were found to carry combination of bla<sub>ampC</sub> and bla<sub>CTX-M</sub> genes and three out of them showed the absence of Sul-1 gene. A total of five isolates showed the same array of bla genes and mobilizing element i.e., bla<sub>ampC</sub>, bla<sub>CTX-M</sub>, Sul-1, ORF513, and ISEcp1. Two isolates showed the combination of bla<sub>ampC</sub>, bla<sub>CTX-M</sub>, and IS26. A single isolate was noticed that possess the combination of all four mobile genetic elements along with bla genes, i.e., the combination of bla<sub>ampC</sub>, bla<sub>CTX-M</sub>, Sul-1, ORF513, and ISEcp1. Two isolates showed the presence of ISEcp1 as mobilizing agents. Among the three Sul-1-negative isolates, one isolate was found devoid of all mobilizing elements studied and carries only bla<sub>ampC</sub> and bla<sub>CTX-M</sub>. In the remaining two isolates, one showed only IS26 as insertion sequence that can mobilize antibiotic resistance genes, while other one bears ORF513 and ISEcp1. The second most common combination of bla genes observed was that of bla<sub>ampC</sub>, bla<sub>CTX-M</sub>, and bla<sub>SIV</sub>. All the four isolates demonstrating this combination of genes, showed similar pattern of mobilizing elements, i.e., the combination of Sul-1 and ISEcp1 was noticed along with bla<sub>ampC</sub>, bla<sub>CTX-M</sub> and bla<sub>SIV</sub>. Combination of bla<sub>ampC</sub>, bla<sub>CTX-M</sub>, bla<sub>TEM</sub> along with Sul-1, ORF513, and ISEcp1 was noticed in three isolates. The combination of all four bla genes was noticed in two isolates and they both displayed combination of mobilizing elements different from each other. One isolate showed combination of Sul-1, ORF513, and ISEcp1 while other one showed IS26, Sul-1, ORF513 and ISEcp1 as mobile elements.

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**Table 2: Genetic organization of integron-positive isolates of E. coli and Klebsiella pneumonia.**

| Amplicon size | Genes | Number of isolates |
|---------------|-------|--------------------|
| Single band   |       |                    |
| 400 bp        | bla<sub>ampC</sub>, bla<sub>CTX-M</sub>, bla<sub>TEM</sub>, bla<sub>SIV</sub>, Sul-1, IS26 | 3 |
| 800 bp        | bla<sub>ampC</sub>, bla<sub>CTX-M</sub>, bla<sub>TEM</sub>, bla<sub>SIV</sub>, Sul-1, ORF513, ISEcp1, IS26 | 11 |
| 900 bp        | bla<sub>ampC</sub>, bla<sub>CTX-M</sub>, bla<sub>TEM</sub>, bla<sub>SIV</sub>, Sul-1, ORF513, ISEcp1, IS26 | 3 |
| 1500 bp       | bla<sub>ampC</sub>, bla<sub>CTX-M</sub>, bla<sub>TEM</sub>, bla<sub>SIV</sub>, Sul-1, ORF513, ISEcp1, IS26 | 24 |
| Two bands     | bla<sub>ampC</sub>, bla<sub>CTX-M</sub>, bla<sub>TEM</sub>, bla<sub>SIV</sub>, Sul-1, ORF513, ISEcp1, IS26 | 20 |
| Three bands   | bla<sub>ampC</sub>, bla<sub>CTX-M</sub>, bla<sub>TEM</sub>, bla<sub>SIV</sub>, Sul-1, ORF513, ISEcp1, IS26 | 4 |
| Multiple bands| bla<sub>ampC</sub>, bla<sub>CTX-M</sub>, bla<sub>TEM</sub>, bla<sub>SIV</sub>, Sul-1,ORF513, ISEcp1, IS26 | 4 |

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**Figure 1(a):** 2% agarose gel showing single amplicon observed in amplification of CS region.

**Figure 1(b):** Agarose gel showing the amplification pattern of single, double and multiple amplicons noticed in amplification of CS region.

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Sixty nine isolates showed amplification when PCR was performed for CS region and out of them twenty isolates showed two bands. Interestingly one isolate that do not harbor any of bla genes, showed two bands in amplification PCR performed for CS region and ORF513 was noted. Remaining isolate showed the combination of all four mobile elements i.e. IS\textsubscript{Ecp1}, Sul-1, and IS\textsubscript{526} was noticed.

Three isolates were found carrying the combination of bla\textsubscript{ampC}, bla\textsubscript{CTX-M}, bla\textsubscript{TEM}, and bla\textsubscript{SHV}, however, combination of mobile genetic elements varied. Combination of Sul-1 and IS\textsubscript{Ecp1} was observed in one isolate while in other ORF513 replaces IS\textsubscript{Ecp1} (combination of Sul-1 and ORF513 was noted). Remaining isolate showed the combination of all four mobile elements i.e. IS\textsubscript{Ecp1}, Sul-1, ORF513, and IS\textsubscript{526} was noticed.

Sixty nine isolates showed amplification when PCR was performed for CS region and out of them twenty isolates showed two bands. Interestingly one isolate that do not harbor any of bla genes, showed two bands in amplification PCR performed for CS region and it carried Sul-1 gene also thereby indicating the presence of Class 1 integron. Figure 1(a) and 1(b) shows the banding pattern observed in amplification of CS region.

### Table 3: bla gene composition in various clusters of E. coli and K. pneumoniae isolates.

| RAPD profile | No. of Isolates | Wards         | bla genes                                      |
|--------------|-----------------|---------------|------------------------------------------------|
| EC1          | 3               | Surgery       | \(\text{bla}_{\text{ampC}}; \text{bla}_{\text{CTX-M}}; \text{bla}_{\text{TEM}}; \text{bla}_{\text{SHV}}\) |
| EC2          | 2               | Surgery       | \(\text{bla}_{\text{ampC}}; \text{bla}_{\text{CTX-M}}; \text{bla}_{\text{TEM}}; \text{bla}_{\text{SHV}}\) |
| EC3          | 3               | Surgery       | \(\text{bla}_{\text{ampC}}; \text{bla}_{\text{CTX-M}}; \text{bla}_{\text{TEM}}; \text{bla}_{\text{SHV}}\) |
| EC4          | 2               | Surgery       | \(\text{bla}_{\text{ampC}}; \text{bla}_{\text{CTX-M}}; \text{bla}_{\text{TEM}}; \text{bla}_{\text{SHV}}\) |
| EC5          | 2               | Surgery       | \(\text{bla}_{\text{ampC}}; \text{bla}_{\text{CTX-M}}; \text{bla}_{\text{TEM}}; \text{bla}_{\text{SHV}}\) |
| EC6          | 2               | Surgery       | \(\text{bla}_{\text{ampC}}; \text{bla}_{\text{CTX-M}}; \text{bla}_{\text{TEM}}; \text{bla}_{\text{SHV}}\) |
| EC7          | 2               | Surgery       | \(\text{bla}_{\text{ampC}}; \text{bla}_{\text{CTX-M}}; \text{bla}_{\text{TEM}}; \text{bla}_{\text{SHV}}\) |
| EC8          | 2               | Surgery       | \(\text{bla}_{\text{ampC}}; \text{bla}_{\text{CTX-M}}; \text{bla}_{\text{TEM}}; \text{bla}_{\text{SHV}}\) |
| EC9          | 2               | Gynaecology   | \(\text{bla}_{\text{ampC}}; \text{bla}_{\text{CTX-M}}; \text{bla}_{\text{TEM}}; \text{bla}_{\text{SHV}}\) |
| EC10         | 2               | Gynaecology   | \(\text{bla}_{\text{ampC}}; \text{bla}_{\text{CTX-M}}; \text{bla}_{\text{TEM}}; \text{bla}_{\text{SHV}}\) |
| EC11         | 2               | Gynaecology   | \(\text{bla}_{\text{ampC}}; \text{bla}_{\text{CTX-M}}; \text{bla}_{\text{TEM}}; \text{bla}_{\text{SHV}}\) |
| EC12         | 2               | Gynaecology   | \(\text{bla}_{\text{ampC}}; \text{bla}_{\text{CTX-M}}; \text{bla}_{\text{TEM}}; \text{bla}_{\text{SHV}}\) |
| EC13         | 3               | Medicine      | \(\text{bla}_{\text{ampC}}; \text{bla}_{\text{CTX-M}}; \text{bla}_{\text{TEM}}; \text{bla}_{\text{SHV}}\) |
| EC14         | 2               | Medicine      | \(\text{bla}_{\text{ampC}}; \text{bla}_{\text{CTX-M}}; \text{bla}_{\text{TEM}}; \text{bla}_{\text{SHV}}\) |
| EC15         | 2               | Medicine & Paediatrics | \(\text{bla}_{\text{ampC}}; \text{bla}_{\text{CTX-M}}; \text{bla}_{\text{TEM}}; \text{bla}_{\text{SHV}}\) |
| KP1          | 2               | Surgery       | \(\text{bla}_{\text{ampC}}; \text{bla}_{\text{CTX-M}}; \text{bla}_{\text{TEM}}; \text{bla}_{\text{SHV}}\) |
| KP2          | 2               | Surgery       | \(\text{bla}_{\text{ampC}}; \text{bla}_{\text{CTX-M}}; \text{bla}_{\text{TEM}}; \text{bla}_{\text{SHV}}\) |

### Epidemiological typing of E. coli and K. pneumoniae isolates

Out of 109 E. coli isolates, 91 could be typed by RAPD, while 18 were found untypable. Among 91 E. coli isolates, 33 were grouped in 15 clusters (EC1 to EC15) while the remaining 58 isolates showed unique banding patterns indicating genetic unrelatedness. Maximum clustering was noticed in isolates obtained from surgery ward (EC1 to EC8). Surprisingly, no cluster was noticed in isolates obtained from orthopaedics ward. Figure 2(a-d) shows the RAPD patterns of E. coli isolates. bla gene composition of the isolates grouped in clusters was variable (Table 3), however, occurrence of \(\text{bla}_{\text{ampC}}\) was observed in all clusters except one isolate that harbours combination of \(\text{bla}_{\text{TEM}}\) & \(\text{bla}_{\text{SHV}}\). Among 16 K. pneumoniae isolates, 14 were typed by RAPD and 2 isolates were found untypable. Figure 3 shows the RAPD profile.

![Figure 2a (i) and (ii): RAPD profile of E. coli isolates obtained from Surgery wards.](image-url)
of *K. pneumoniae* isolates obtained from various wards. Out of 14 isolates, 4 were grouped in two clusters (KP1 & KP2) and 10 displayed unique banding patterns. Figure 4 shows the representative clusters of *E. coli* and *K. pneumoniae* isolates.

**Discussion**

The beneficial aspects of the introduction of antibiotics in the treatment of human infectious diseases is hampered by the emergence of bacterial resistance has become an ever-increasing problem that threatens the clinical usefulness of these drugs. Class 1 integrons has been identified as the primary source of antimicrobial resistance genes and is suspected to serve as reservoirs and exchange platform of resistance genes in a variety of Gram-negative bacteria. [11].

It has been reported that the antibiotic resistance was noticeably higher in class 1 integron-positive strains of *P. aeruginosa* as compared to those of class 1 integron-negative strains [12]. Recently [13] reported presence of integron in 53.9% Enterobacterial isolates. However, we have observed quite high percentage of β-lactam resistance in integron-negative isolates also, indicating that probably these genes are located on plasmids. More integron-harboring isolates were found resistant to fluoroquinolones than integron-negative isolates as previously reported by [14]. This may be due to the presence of integrons, which increase the mutation rate of the host cell [15], or the presence of genes on integrons that code for reduced membrane permeability or enhanced efflux [16].

The prevalence of antibiotic resistance is mainly due to the horizontal transfer of antibiotic-resistance genes, expressed by mobile genetic elements such as plasmids and transposons. One way for the spread of antibiotic resistance-encoding genes involves integrons. Class 1 integrons were significantly associated with resistance to all the antimicrobials tested. However, integron-positive strains were significantly more common among isolates showing resistance to β-lactam antibiotics. The higher rate of resistance to several classes of β-lactam antibiotics in integron-positive isolates is probably attributable to the association of β-lactamase genes with integron-carrying plasmids. Many β-lactamase genes within the integron have been reported, like *bla*<sub>CMY</sub>, *bla*<sub>CTX-M</sub>, and *bla*<sub>OXA</sub> [17,18]. Moreover, integrons were also reported to be associated with fluoroquinolone resistance [19]. We noticed occurrence of integrons (as detected by amplifying 5'CS-3'CS region) in 55.20% enterobacterial isolates. However, the presence of integrons from community and nosocomial environment has been frequently reported [20-22]. Only 22% and 15% of isolates were found to carry Class 1 integrons in a Swiss [23] and Australian study [24] respectively. Shaheen, et al. reported the occurrence of Class 1 and class 2 integrons in 27% and 2% isolates respectively [25]. Bhattacharjee, et al. observed that among 63 ESBL-producing *K. pneumoniae* isolates, 58 (92%) carried Class 1 integrons and 6 isolates were found to harbour Class1 as well as Class 2 integron [26].

*Sul-1* was reported to be present in 3’CS [27], but we also observe 14 isolates (out of 69) that showed amplification for 5’CS-3’CS region but did not show presence of *Sul-1* (when detected by PCR). Similar type of results was observed by Jin & Ling [28]. It was suggested by Jin & Ling that using sulfamethoxazole resistance as a marker for integrons
could either misses the isolates or overestimates the prevalence of integrons. Insertion of class 1 integrons. Deletion/replacement of the typical 3'CS. According to Saenz, et al. [32], some markers (absence of qacE1-Sul-1 and the presence of qacH, Sul3 & cmlA genes) could be used to envisage the presence of non-classic integrons. Deletion/replacement of the typical qacE1-Sul-1 sequence may result in the integration of more stable and efficiently expressed genes [33]. Laverstein-van Hall, et al. [34] noticed a significant association of multiple resistance genes with the presence of an integron and that the transfer of integron among bacterial species plays an important role in the development of multiple antibiotics resistance.

In our study isolates, single band was observed at variable position when PCR was performed by using primers specific for 5’CS and 3’CS. Similar type of results were observed by Peng, et al. [35] as they reported amplicons ranging from 0.7 to 3.0 kb obtained by PCR amplification of gene cassette region by using 5’CS-3’CS primers. The resistance genes of 1.0 kb (aadA7), 1.6 kb (dfrA12-orfF-aadA2), and 2.4 kb (folA-catB3-aadA5) class 1 integrons have also been reported [36,37]. It has been suggested that the deletion in the 3’CS segment or acquisition of different antibiotic resistance genes results in differences observed in molecular weight of amplicons.

It was evident from RAPD-typing that most patients in our hospital were infected with different clades of organisms, thereby demonstrating clonal diversity among isolates suggesting horizontal transmission of bla genes. RAPD-typing demonstrated that untypability of isolates is increasing. However, few isolates from gynaecology, surgery and orthopedics wards displayed similar banding patterns. Most probably, the same clone is circulating in these wards as the above described wards are sharing the same building block in our hospital and the chances of cross contamination increases.

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Figure 4: Representative Clusters of E. coli and K. pneumoniae isolates obtained from various wards.
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