**Mycobacterium tuberculosis** Type VII Secreted Effector EsxH Targets Host ESCRT to Impair Trafficking

Alka Mehra1,*, Aleena Zahra1, Victor Thompson1, Natalie Sirisaengtaksin2, Ashley Wells1, Maura Porto1, Stefan Köster1, Kristen Penberthy1, Yoshihisha Kubota2, Amelie Dricot3,4, Daniel Rogan1, Marc Vidal3,4, David E. Hill2,4, Andrew J. Bean2,5, Jennifer A. Philips1*

1 Division of Infectious Diseases, Department of Medicine, Department of Pathology and Department of Microbiology, New York University School of Medicine, New York, New York, United States of America, 2 Department of Neurobiology and Anatomy, and Graduate School of Biomedical Sciences, The University of Texas Health Science Center at Houston, Houston, Texas, United States of America, 3 Center for Cancer Systems Biology (CCSB) and Department of Cancer Biology, Dana-Farber Cancer Institute, Boston, Massachusetts, United States of America, 4 Department of Genetics, Harvard Medical School, Boston, Massachusetts, United States of America, 5 Division of Pediatrics, The University of Texas M.D. Anderson Cancer Center, Houston, Texas, United States of America

**Abstract**

*Mycobacterium tuberculosis* (MtB) disrupts anti-microbial pathways of macrophages, cells that normally kill bacteria. Over 40 years ago, D’Arcy Hart showed that MtB avoids delivery to lysosomes, but the molecular mechanisms that allow MtB to elude lysosomal degradation are poorly understood. Specialized secretion systems are often used by bacterial pathogens to translocate effectors that target the host, and MtB encodes type VII secretion systems (TSSSs) that enable mycobacteria to secrete proteins across their complex cell envelope; however, their cellular targets are unknown. Here, we describe a systematic strategy to identify bacterial virulence factors by looking for interactions between the MtB secretome and host proteins using a high throughput, high stringency, yeast two-hybrid (Y2H) platform. Using this approach we identified an interaction between EsxH, which is secreted by the Esx-3 TSSS, and human hepatocyte growth factor-regulated tyrosine kinase substrate (Hgs/Hrs), a component of the endosomal sorting complex required for transport (ESCRT). ESCRT has a well-described role in directing proteins destined for lysosomal degradation into intraluminal vesicles (ILVs) of multivesicular bodies (MVBs), ensuring degradation of the sorted cargo upon MVB-lysosome fusion. Here, we show that ESCRT is required to deliver MtB to the lysosome and to restrict intracellular bacterial growth. Further, EsxH, in complex with EsxG, disrupts ESCRT function and impairs phagosome maturation. Thus, we demonstrate a role for a TSSS and the host ESCRT machinery in one of the central features of tuberculosis pathogenesis.

**Citation:** Mehra A, Zahra A, Thompson V, Sirisaengtaksin N, Wells A, et al. (2013) Mycobacterium tuberculosis Type VII Secreted Effector EsxH Targets Host ESCRT to Impair Trafficking. PLoS Pathog 9(10): e1003734. doi:10.1371/journal.ppat.1003734

**Editor:** Christopher M. Sassetti, University of Massachusetts, United States of America

**Received** April 8, 2013; **Accepted** September 12, 2013; **Published** October 31, 2013

**Copyright:** © 2013 Mehra et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Funding:** This work was supported by grants from the NIH/NIAID (AI087682), the Doris Duke Charitable Foundation, the Infectious Disease Society of America (IDSA), and the Michael Saperstein Medical Scholars Research Fund to JAP. It was supported by NIH (HG004233 and HG001715), Ellison Foundation (Boston, MA), and Institute Sponsored Research funds from the Dana-Farber Cancer Institute Strategic Initiative to MV and DEH. Potts Memorial Foundation provided support to AM, IDSA provided support to MP, and the American Society of Microbiology supported AZ. http://www.ddcf.org http://www.idsociety.org/Index.aspx http://www.ellisonfoundation.org http://www.asm.org http://www.dana-farber.org Pott’s and Michael Saperstein Fund: URL not available. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing Interests:** The authors have declared that no competing interests exist.

* E-mail: Jennifer.Philips@nymc.org

† These authors contributed equally to this work.

**Introduction**

An important virulence property of *Mycobacterium tuberculosis* (MtB)- the causative agent of the disease tuberculosis- is its ability to avoid delivery to the lysosome. It has long been appreciated that MtB alters phagosome maturation, such that internalized bacteria are not transported to the lysosome but instead reside in an early endosome-like compartment [1,2]. The MtB-induced block in phagosome-lysosome fusion has been attributed to a wide array of lipid and protein effectors [3,4] but the mechanism remains poorly understood. More recently, the ability of MtB to permeabilize the phagosomal membrane, which allows bacterial products and in some cases intact bacteria to access the cytosol, has been described [5–9]. The TSSS Esx-1 and its secreted effectors, EsxA/ESAT-6 and EsxB/CFP-10, are critical for this process. Esx-1 has been investigated intensively because its absence in the vaccine strain *Mycobacterium bovis*-BCG (BCG) largely accounts for attenuation of that strain [8–10]. MtB encodes five loci resembling Ess-1 (Ess-1-Ess-5), as well as 11 tandem pairs of proteins similar to EssA and EssB (EssA-EssW), but their cellular targets, if any, are unknown [11]. Ess-3 plays a role in iron acquisition in MtB, as well as in a non-pathogenic strain, *Mycobacterium smegmatis* (Msmeg) [12,13]. Ess-3 is a focus of vaccine efforts because it secretes EsxG/ TB9.8 and EssH/TB10.4, which are highly antigenic [14,15], and because introduction of the MtB ESX-3 locus into an Msmeg strain lacking the endogenous ESX-3 region generates highly protective immunity [16]. The ESX-5 locus is required for transport of proteins with conserved proline-glutamic acid (PE) and proline-proline-glutamic acid (PPE) motifs [17,18] and modulates macrophage responses [19]. Thus, TSSSs and their putative effectors appear to be important in virulence and modulating host cells, however, their mechanism of action and molecular targets are unclear.
Author Summary

*Mycobacterium tuberculosis* (Mtbc) causes the disease tuberculosis, one of the world's most deadly infections. The host immune system cannot eradicate Mtbc because it grows within macrophages, cells that normally kill bacteria. One of the intracellular survival strategies of Mtbc is to avoid delivery to lysosomes, a phenomenon described over 40 years ago, but for which the mechanism and molecular details remain incomplete. Mtbc possess specialized secretion systems (Type VII secretion systems; TSSS) that transfer particular proteins out of the bacteria, but how these proteins promote infection is not well understood. In this study, we used a high stringency yeast two-hybrid system to identify interactions between secreted effectors from Mtbc and human host factors. We identified ninety-nine such interactions and focused our attention on the interaction between EsxH, secreted by Esx-3, a TSSS of Mtb, and Hrs, a component of the host ESCRT machinery. We provide evidence that Mtbc EsxH directly targets host Hrs to disrupt delivery of bacteria to lysosomes. Thus, this study demonstrates the role of a TSSS effector and the ESCRT machinery in what is one of the central features of tuberculosis pathogenesis, thereby providing molecular insight into why humans can't clear Mtbc infection.

Here, we show that EsxG and EsxH from Mtbc, but not the Msmeg homologs, target the host factor, Hrs. Hrs is a component of the ESCRT machinery, a group of four protein complexes (ESCRT-0 to ESCRT-III) composed of cytosolic components that are sequentially recruited to the endosomal membrane. The ESCRT machinery has a well-described role in directing cargo destined for degradation into intraluminal vesicles of multivesicular bodies (MVBs) that fuse with lysosomes [20,21]. We show that ESCRT is also required to deliver Mtbc to the lysosome and to restrict intracellular bacterial growth. EsxH, in complex with EsxG, is able to disrupt ESCRT function and impair phagosome maturation.

Results

High throughput identification of Mtbc secretome-human host interactions

We used a systematic strategy to identify secreted bacterial virulence factors by looking for interactions with host proteins using a high throughput, high stringency, yeast two-hybrid (Y2H) platform [22]. First, we curated the literature to define the Mtbc secretome. Thirty-eight publications predicted 718 secreted proteins based on presence in culture filtrate (CF), ability to cause secretion of an assayable protein, bioinformatic criteria, or detailed study (see Text S1 for additional details). In order to prioritize open reading frames (ORFs) for screening, we imposed a number of criteria, such as excluding proteins with multiple transmembrane spanning domains (see Text S1 for additional details). In addition, since the starting list of putative secreted proteins might contain proteins that are not actually secreted, we attempted to eliminate ORFs that were likely to be inaccurately classified as secreted. One way in which this can happen is if cytoplasmic proteins appear in CF due to bacterial lysis. In order to minimize the contribution of such proteins, we did not include ORFs that were annotated in Tuberculist (http://tuberculist.epfl.ch/) as being involved in lipid metabolism, information pathways (which contains proteins involved in replication, transcription and translation), or intermediary metabolism and respiration, since most of these are likely involved in basic, intrinsic bacterial processes, and hence, many may be misclassified. To avoid removing true secreted proteins, ORFs were not de-prioritized if they had a possible signal sequence or there were data supporting their role during infection. In doing so, we removed many proteins that were found in CF in a single study, and hence may be misclassified (see Text S1 for details). From the final list, 339 sequence validated secretome ORFs were provided by Pathogen Functional Genomic Resource Center (PFGRC; Dataset S1). Because many secreted proteins play an intrinsic role in the bacterial lifecycle, we anticipated that only a small fraction would interact with human proteins. Thus, to estimate a false positive hit rate of our system, we included sixty ORFs that are not likely to be secreted to serve as controls (see Text S1 for details; Dataset S2).

In order to evaluate their performance in Y2H protein-protein interaction (PPI) mapping, we tested the 399 Mtbc ORFs expressed as Gal4-DNA binding domain (DB) fusions for pair wise interactions with the same 399 Mtbc ORFs expressed as Gal4-activation domain (AD) fusions. From the ~160,000 combinations queried, we found 14 unique PPIs (Table S1). The rate of interactions is as high as in human ORFeome mapping [22], exceeding the stochastic false positive rate of the Y2H platform by fifteen-fold [23]. Half of the interactions were between proteins belonging to the WXG100 family (EsxA-EsxW). These proteins are approximately 100 amino acids in length and have a characteristic hairpin bend formed by a Trp-Xaa-Gly (W-X-G) motif. Mtbc encodes 11 tandem pairs of such proteins, which are thought to function as secreted heterodimers. Heterodimer formation is proposed to be limited to interactions between genome pairs or very closely related family members [24,25], and the interactions we detected by Y2H exhibit this specificity. Six of the remaining seven PPIs involved homotypic interactions; for example, bacterioferritin (BfrB) was found to interact with itself, consistent with the proposal that it assembles into 24-subunit oligomers [26].

After ensuring the high performance of Mtbc ORFs in Y2H PPI mapping, we looked for interactions between the Mtbc secreteme and ~12,000 human ORFs, testing approximately 4 million interactions. From the secreteme collection, we identified 99 PPIs between 53 Mtbc proteins and 63 human proteins (Dataset S3). The number of Mtbc proteins exhibiting an interaction with a human protein was approximately two-fold higher for the secreteme collection compared to the non-secreted control set (53 out of 339 versus 5 out of 60). We analyzed the collection to determine whether PPIs were enriched for subsets of Mtbc proteins (Table S2). We observed that the sixteen Esx proteins included in the collection were significantly more likely to interact with human ORFs than were controls (p = 0.0087). The finding that Esx proteins were enriched for interactions may reflect that this group of proteins plays an important role in virulence, or could mean that these proteins, which usually form a heterodimer, are prone to aberrant interactions when they are expressed without their binding partner.

It is difficult to gauge the success of the screen based upon known interaction between Mtbc proteins and cytosolic human protein because so few are known. Included in our screening set were, PtpA, which has been shown to interact with Vps33B and the H subunit of the human v-ATPase [27,28], LpdC, which interacts with coronin 1 [29], and NdKα, which interacts with Rab5 and Rab7 [30]. We did not identify these known interactions, however, the screen was not performed to saturation and the Y2H platform can detect ~20% of well-validated interacting pairs [31]. We did identify an interaction between PtpA and Ligand of Numb protein X (LINX1), a RING finger-type...
E3 ligase that contains 4 PDZ domains and plays a scaffolding role in diverse cellular pathways. Several other secretome ORFs also interacted with LNX1, suggesting that LNX1 might be modulated by Mtb, or LNX1 might regulate the function or stability of certain Mtb effectors.

When we examined hit rates based upon the functional category of the Mtb ORFs, the category with the greatest enrichment was cell wall and cell processes, which contains the Esx family members. The intermediary metabolism and respiration category exhibited a hit rate similar to the control collection, consistent with the idea that most of these ORFs do not function in host interactions as their annotation suggests. Interestingly, one of the two proteins that did exhibit an interaction in this category is Zmp1, a zinc metalloprotease which has been shown to inhibit the inflammasome and impair phagosome maturation but whose cellular target is unknown [32]. Zmp1 interacted with KCTD6b, a BTB/POZ domain containing protein that can function as a Cullin3 (Cul3) adaptor [33]. Cul3 has recently been shown to be a regulator of endo-lysosomal trafficking, suggesting that Zmp1 may impair phagosome maturation by acting on KCTD6-Cul3 [34].

To evaluate the human targets of the Mtb proteins, we searched the STRING database (http://string.embl.de/) for each of the proteins’ functional associations. The STRING database predicts protein-protein interactions based upon physical and functional associations, such as available high-throughput data, co-expression, genomic context, and text mining of available literature. Using a medium confidence value to define protein-protein interactions, there were significantly more interactions (n = 16) observed for the human targets than would be predicted by chance (p = 8.3 x 10^{-15}). We identified proteins involved in host immunity to bacterial infection, such as Ndp52 [35,36], Taxlpb1 [37], and STAT3 [38], however, human targets of the Mtb proteins were not significantly enriched for annotated pathways in the Kyoto Encyclopedia of Genes and Genomes (KEGG) when corrected for multiple testing, which may reflect the limited number of human targets found.

**ESCRT is required for restricting intracellular growth and trafficking of slow growing mycobacteria**

We focused on the interaction between EsxH and Hrs because TSSSs, which secrete Exs proteins, are clearly important in virulence but the function of their secreted effectors is largely unknown. In addition, our existing data supported the idea that the ESCRT machinery is important in controlling bacterial replication. Hrs, which plays a central role in the assembly of the initial ESCRT components on endosomes, is recruited to mycobacterial phagosomes [39], and we had previously shown in an RNAi screen in *Drosophila* that ESCRT restricts the intracellular growth of rapidly growing mycobacteria [10,41]. Control of bacterial replication appears to be particularly sensitive to ESCRT perturbation, because, in addition, when we screened 6500 siRNA pools in RAW 264.7 (RAW) macrophages for their ability to confer enhanced intracellular growth of Msmeg, we found that the two strongest hits were Rab7, known to be involved in late endosome-lysosome fusion, and Tsg101, an ESCRT-I component that is recruited to endosomes by Hrs [data not shown]. Hrs was also identified in this screen, although previously we had found no effect with Hrs silencing, which we now attribute to insufficient protein depletion [41]. In the RAW cell RNAi screen that identified Hrs, we used Ambion Silencer siRNA pools, whereas previously we used a Dharmacon siGENOME pool to deplete Hrs [41]. To clarify the discrepancy, we tested a third pool (Dharmacon ON-TARGETplus), which, like the Ambion pool, conferred enhanced growth to Msmeg. We tested the individual Dharmacon ON-TARGETplus siRNAs and found that 2 of 4 targeting Hrs resulted in depletion of Hrs protein, enhanced the growth of Msmeg, and altered trafficking, whereas the other two had no effect (Figure S1 and data not shown). Thus, one possibility is that Mtb secretes EsxH, which binds Hrs and impairs ESCRT function, thereby promoting intracellular bacterial growth. To determine whether ESCRT restricts growth of Mtb, we depleted Hrs and Tsg101 and examined the intracellular growth of Mtb in RAW macrophages. We found no significant effect of silencing on bacterial uptake (data not shown), however when we assessed bacterial colony forming units (CFU) two day post-infection, we observed enhanced intracellular survival of Mtb in cells depleted of Hrs or Tsg101, similar to what was seen with Rab7 silencing (Figure 1A). Intracellular growth of BCG in bone marrow-derived macrophages (BMDMs) was even more strongly effected (Figure S2). Thus, Hrs restricts growth of slow growing and virulent mycobacteria.

ESCRT targets certain cell surface receptors and biosynthetic cargo to lysosomes [42]. Thus, ESCRT might restrict intracellular bacterial growth by governing bacterial trafficking and/or lysosomal content. We examined the localization of Mtb relative to Transferrin Receptor (TIR), a marker of early and recycling endosomes, and LAMP1, a marker of late endosomes and lysosomes using automated image analysis (Figures S3A, S3B). In cells depleted of Tsg101, Hrs, or Rab7, we observed diminished co-localization between Mtb and LAMP1 and a concomitant increase in co-localization of Mtb with TIR compared to control cells 24 hours post-infection (hpi) (Figure 1B), suggesting decreased Mtb delivery to degradative compartments. Similarly, in cells infected with BCG there was diminished co-localization with LysoTracker, which accumulates in the acidic environment of the lysosome, and enhanced co-localization with TIR (Figure 1B). Thus, Hrs and Tsg101, like Rab7, are required for bacterial trafficking. To verify that bacterial viability correlates with low LAMP1 and LysoTracker co-localization and with high TIR co-localization, we compared the cellular localization of viable bacteria to total bacteria. We identified metabolically active BCG 24 hpi based upon their ability to induce expression of GFP from a tetracycline-inducible promoter (BCG-tet-GFP) and compared their intracellular localization to the BCG strain that constitutively express GFP (BCG-GFP). Whereas there was a wide range in intensities of associated LAMP1 and LysoTracker with BCG-GFP, metabolically active bacteria were found almost exclusively in phagosomes with minimal acidification, little co-localization with LAMP1, and enhanced TIR co-localization at 48 hpi (Figure S3C). Thus, impaired bacterial trafficking to a late endosomal or lysosomal compartment underlies the failure to control mycobacterial replication in ESCRT-depleted cells, although altered lysosomal content may also contribute.

**The Mtb EsxG EsxH heterodimer binds Hrs**

Pathogenic mycobacteria arrest phagosome maturation in evolutionarily diverse cells. Supporting the idea that EsxH might play a role in inhibition of bacterial degradation, we observed that EsxH interacts with human, mouse, and zebrafish orthologs of Hrs, suggesting that it recognises a conserved structural feature of Hrs (Figure 2A). Orthologs of EsxH are found widely in mycobacteria, including in the non-pathogen Msmeg. If EsxH prevents phagosome-lysosome fusion by impairing Hrs function, we anticipate that would be a feature specific to EsxH from pathogenic mycobacteria. To test this, we cloned the EsxH ortholog from Msmeg (MSMEG_0621; EsxH*), which encodes a protein 75% identical to EsxH from Mtb (hereafter referred to as
EsxH_Mt interacted with EsxG_Mt, demonstrating that the protein was functional in the Y2H assay, it interacted poorly with Hrs (Figure 2A), consistent with the notion that the interaction of EsxH_Mt with Hrs contributes to virulence.

EsxH_Mt forms a heterodimer with EsxG_Mt, composed of a four-helix bundle with flexible N- and C-terminal arms from both proteins that coordinate zinc and contribute to a cleft that has been predicted to mediate a PPI [43]. To determine whether Hrs interacts with the heterodimer, we used a fusion protein in which EsxG_Mt and EsxH_Mt were expressed as a single polypeptide that preserves the folded structure of the native heterodimer [44]. This fusion protein interacted with Hrs (Figure 2B). Deletion of the first five amino acids of EsxH_Mt (EsxG_Mt-EsxH_Mt-AN5) weakened its interaction with Hrs (Figure 2B). These data show that Hrs can interact with EsxH_Mt when it is complexed to EsxG_Mt, and suggest that the conformation of the amino terminal arm of EsxH_Mt is important. To further test whether the structure of the N- and C-termini are important, we mutated His-14, His-70, and His-76 Glu-77. These residues contribute to zinc binding, and His-76 is also part of the predicted cleft. We mutated them to Ala, with the exception of His-76, which we changed to Arg because this is found in EsxH_Mt. While His-14 and H70R did not have a detectable effect, when His-76 and Glu-77 were both changed to Ala, the interaction between EsxH_Mt and Hrs was impaired, although EsxH_Mt H76A-E77A still interacted with EsxG_Mt (Figure 2A). To verify that EsxH_Mt binds Hrs, we purified the EsxG_Mt, EsxH_Mt heterodimer from E. coli [44] and Hrs from baculovirus [45]. Hrs bound EsxG_Mt, EsxH_Mt in a saturable manner, exhibiting stoichiometric binding with a Kd of ~5 μM (Figure 2C and 2D). We conclude that Hrs interacts with the EsxG_Mt, EsxH_Mt heterodimer, and the interaction likely involves the N- and C-terminal arms of EsxH_Mt.

**EsxG_Mt and EsxH_Mt Disrupt ESCRT function in mammalian cells**

To determine whether EsxH_Mt interacts with Hrs and alters ESCRT function in mammalian cells, we expressed EsxH_Mt–FLAG in HEK293 cells. EsxH_Mt was not detectable unless we co-expressed EsxG_Mt (Figure 3A, compare lanes 1 and 3; see Figure S4 for quantification); its abundance was also increased slightly by overexpression of Hrs (compare lane 1’ with 2’ and lane 5 with 6). When expressed alone, EsxH_Mt could be stabilized by MG132, likely because it is not properly folded without EsxG_Mt and hence is subject to proteasome-mediated degradation (Figure 3A compare lanes 1 and 5). To determine if there was an interaction between EsxG_Mt–EsxH_Mt and Hrs, we performed co-immunoprecipitation experiments in cells co-transfected with EsxG_Mt, EsxH_Mt, and Hrs-myc. Hrs was immunoprecipitated with an antibody directed against the myc-tag, and we found that EsxH_Mt was co-immunoprecipitated (Figure 3B). No EsxH_Mt was co-immunoprecipitated when an isotype control antibody was used, and as expected, EsxH_Mt and EsxH_Mt-H76A-E77A were impaired in the interaction with Hrs (Figure 3B and 3C). Interestingly, the co-immunoprecipitation of EsxH_Mt and Hrs could only be detected when cells were pre-treated with MG132. Thus, one possibility is that the EsxG_Mt–EsxH_Mt heterodimer is polyubiquitinated and degraded by the proteasome. In the presence of MG132, the polyubiquitinated species might accumulate, allowing us to detect an interaction between Hrs and an ubiquitinated species of EsxH_Mt, since Hrs contains an ubiquitin interacting motif (UIM) domain. Arguing against this possibility, when EsxH_Mt was co-expressed with EsxH_Mt there was little, if any, effect of MG132 on EsxH_Mt protein levels (Figure 3A, compare lanes 3 and 7, Figure S4, and Figure S5). In addition, when we examined mono- and polyubiquitinated proteins using the FK2 antibody, inhibition of the proteasome with MG132 caused the accumulation of high molecular weight proteins as anticipated. However, there was no difference seen in the quantity or mobility of EsxH_Mt (Figure S3). Further, when we mapped the region of Hrs required for the interaction with EsxG_Mt–EsxH_Mt in the Y2H assay, the UIM domain was not required. Amino acids 398–630, which contain a coiled-coil region, were sufficient to mediate the interaction (Figure 2E). We verified that the C-terminal half of Hrs was sufficient to mediate an interaction by co-immunoprecipitation (Fig. 3D). In summary, these data show that EsxG_Mt stabilizes EsxH_Mt in the mammalian cytosol and that the heterodimer can bind the C-terminus of Hrs.

To test the hypothesis that the EsxH_Mt interaction with Hrs disrupts ESCRT, we examined the effect of EsxG_Mt, EsxH_Mt on epidermal growth factor (EGF) and epidermal growth factor receptor (EGFR) degradation (all performed without MG132 treatment). Upon binding ligand, EGFR is internalized and transferred into ILVs by ESCRT so that it can be degraded upon MVB-lysosome fusion. To determine whether EsxH_Mt interferes with this process, we transfected EsxG_Mt and EsxH_Mt or vector control into HEK293 cells and examined EGFR levels 90 min after EGF treatment. We found that EGFR levels decreased in control cells. In cells co-expressing EsxG_Mt and EsxH_Mt, there was a 63±8% (n = 3) increase in the fraction of EGFR that remained undegraded (Figure 3E), similar to what has been seen with Hrs depletion [46]. In contrast, co-expression of EsxG_Mt and EsxH_Mt had no detectable effect on EGFR degradation (Figure 3E). We observed similar results when we used fluorescent EGF to examine trafficking in A549 cells using fluorescence microscopy. As expected, cells depleted of Hrs showed enhanced EGF fluorescence due to impaired degradation. We observed a similar decreased degradation in cells expressing EsxG_Mt and EsxH_Mt (Figure 3F and G). In contrast, expression of EsxG_Mt, EsxH_Mt or EsxG_Mt, EsxH_Mt-H76A-E77A had little effect on EGF degradation (Figure 3G and 3H). Thus, EsxH_Mt in complex with EsxG_Mt is sufficient to inhibit EGF and EGFR degradation, an activity that correlates with its binding to Hrs. EsxH_Mt, from the non-pathogenic species, does not have this property.

**EsxG_Mt and EsxH_Mt arrest phagosome maturation**

Because Esx-3 is essential for Mtb growth, we examined the effect of overexpressing EsxH_Mt on bacterial trafficking. First, we wanted to determine whether EsxG_Mt, EsxH_Mt could confer a block in maturation to phagosomes containing Msmeg. However, when we expressed EsxG_Mt, EsxH_Mt–FLAG under control of the
Figure 2. EsxH<sub>Mt</sub> binds Hrs. (A) Gal4 DNA-binding domain (DB) fusions of EsxH<sub>Mt</sub>, EsxH<sub>Ms</sub>, or mutant EsxH<sub>Mt</sub> were tested for Y2H interactions with Gal4 activation-domain (AD) fusions of EsxG<sub>Mt</sub>, human (Hs), mouse (Mm), or zebrafish (Dr) Hrs. (B) Y2H interaction between indicated DB and AD constructs. Hrs is human. EsxG<sub>Mt</sub>-EsxH<sub>Mt</sub>-DB did not interact with EsxG<sub>Mt</sub>-AD or EsxH<sub>Mt</sub>-AD, presumably because of the intramolecular interaction in the DB construct. (C) Increasing amounts of Hrs were incubated with a constant amount of immobilized EsxG<sub>Mt</sub>-EsxH<sub>Mt</sub> and bound fraction examined.
EsxHMt-DB was tested in the Y2H for interactions with human Hrs-AD deletion constructs. The domain structure of Hrs is indicated.

combination of EsxGMt EsxHMt overexpression and ESCRT-siRNA-mediated silencing of Hrs, Tsg101, and Rab7, and the localization with TfR with the strain overexpressing EsxHMt-FLAG, compared to a strain transformed with vector control manipulation alone (Figure 4B). An Mtb strain that expressed EsxGMt EsxHMt-FLAG, did not block LAMP1 co-localization, but the mutant protein also failed to be secreted (Figure 4A, 4F). Mtb did secrete EsxGMt EsxHMt-FLAG, which, unlike EsxGm, EsxHMt-FLAG, did not block LAMP1 co-localization (Figure 4A and 4F). We conclude that EsxGMt, EsxHMt, but not EsxGm, EsxHMt, can prevent lysosomal trafficking during infection, most likely reflecting the ability of EsxHMt to bind Hrs and impair ESCRT activity.

Discussion

We used high throughput Y2H interactome mapping to identify interactions between secreted Mtb proteins and human proteins, identifying 99 new potential interactions. We made use of a large body of literature that has attempted to catalogue the secretome of Mtb. Our study is subject to the uncertainty around the definition of the Mtb secretome. For example, proteins can be in the culture filtrate due to bacterial lysis, rather than secretion, and bioinformatics predictions may be inaccurate. In addition, many secreted proteins play an intrinsic role in the bacterial lifecycle and are unlikely to make a biologically meaningful interaction with host proteins. Thus, to estimate a false positive hit rate of our system, we included a non-secreted control collection. We found approximately two-fold enrichment in the rate of interactions comparing the secretome collection to the control collection, suggesting that true interactions were identified, but that there also may be a relatively high rate of "pseudo-interactions," which may be valid biophysiologically but never occur in vivo because the involved proteins are separated spatially or temporally. In addition, the interactome list is by no means complete. We did not screen the entire putative secretome, but rather imposed criteria to try to arrive at a set that was enriched for true secreted proteins likely to play a role in virulence. In addition, the screen was not performed to saturation, and only a fraction of verifiable interactions can be detected by a single method to detect PPIs [31]. Therefore, the list is not comprehensive and likely contains false-positives, but given the paucity of data on host-pathogen interactions in Mtb, it has likely significantly expanded the known Mtb-human protein-protein interactome. It represents a resource for investigators working on Mtb; the confirmation and significance of such interactions will require further validation.

Interactome mapping provides an unbiased strategy to identify host-pathogen interactions for pathogens in which genetic strategies are limited, and it can be complemented by depletion studies in host cells. Such physical interaction mapping can identify redundant or essential factors that may be missed using genetic approaches. For example, in the case of EsxHMt, its importance may have been unrecognized in previous genetic approaches to identify Mtb virulence factors because of redundancy within this large gene family, the existence of additional mechanisms to modulate phagosome maturation, and the essentiality of the Exs-3 system.

This Y2H screen and our previous genome-wide RNAi screen in Drosophila pointed to the importance of the ESCRT machinery in mycobacterial pathogenesis. Here, we show that the ESCRT machinery is important in restricting the intracellular growth of pathogenic mycobacteria, which likely reflects a role of ESCRT in trafficking bacteria to the lysosome, although the effects of ESCRT on endo-lysosomal content and signaling pathways may also play a role. In addition, by modulating the ESCRT machinery, Mtb might alter antigen presentation or exosome formation [42,47].

Further work is required to understand exactly how EsxHMt impairs ESCRT function. We envision that EsxHMt inhibits ESCRT on or near the mycobacterial phagosome, where its local concentration would be highest, as opposed to globally disrupting ESCRT. The C-terminal half of Hrs, which we showed binds Hrs, has previously been shown to be involved in the interactions with Tsg101 [48,49], STAM [50], and SNAP-25 [45]. Thus, one possibility is that EsxHMt interferes with these associations. The relatively low affinity measured in vitro between EsxHMt and Hrs (~5 μM) may be sufficient to disrupt Hrs interactions with other host proteins, as the interactions of Hrs with many of its binding partners are of low affinity [45,48,51,52]. For example, HIV Gag recruits Tsg101 to sites of viral budding by binding the Tsg101 UEV domain with an even lower affinity (Kd = 21–50 μM) [48,53]. There may also be a particular form of Hrs or EsxHMt, that exist in vivo in macrophages that exhibits higher affinity. For example, Hrs interacts with the endosomal membrane, engages in numerous protein-protein interactions, and is modified by phosphorylation and ubiquitination, none of which occur when the affinity is measured with recombinant protein. Interestingly MG132, which is known to alter ESCRT activity [54,55], enhanced our ability to detect an interaction between EsxGm-EsxHMt and Hrs in co-immunoprecipitation experiments in HEK293 cells. One explanation for the requirement of MG132 to detect the Hrs-EsxHMt interaction by co-immunoprecipitation may be related to the observation that MG132 impairs ESCRT-dependent trafficking [54,55]. Thus, it is possible that MG132 stabilizes the interaction between EsxHMt and Hrs by altering ESCRT, although other potential mechanisms could be envisioned. Even in the absence of MG132, EsxGMt-EsxHMt inhibits ESCRT function. Therefore, we speculate that EsxHMt preferentially binds to a form of Hrs that exists transiently in cells, a form that is stabilized by MG132. Once bound to Hrs, EsxHMt could interact with other host proteins that modify Hrs or ESCRT components.

Hrs is one of several host factors that Mtb likely target to create a protected niche [3,4]. Lipoamide dehydrogenase (LpdC) is thought to prevent phagosome-lysosome maturation by retaining the host factor, corinon 1 [29,56]. PtpA, a secreted tyrosine phosphatase, may directly exclude the vacuolar-H^{+}ATPase during infection, impairing acidification and phagosome maturation [27,28,57], while nucleoside diphosphate kinase A (NdkA) targets Rab7 activation [30,58]. In addition, there is less phosphatidylinositol 3-phosphate on the mycobacterial phagosome than latex bead phagosomes, which may reflect the activity of the secreted lipid phosphatase, SapM [59,60]. This leads to impaired recruitment of Hrs [39]. Thus, Hrs activity could be inhibited by Coomassie blue. (D) Average binding (n = 3) was fitted with the Hill function, revealing a Hill coefficient of ~1.7 and a Kd of 5.4 μM. (E) EsxGMt–EsxHMt–DB was tested in the Y2H for interactions with human Hrs-AD deletion constructs. The domain structure of Hrs is indicated. doi:10.1371/journal.ppat.1003734.g002
Figure 3. EsxG<sub>Mt</sub> and EsxH<sub>Mt</sub> interact with Hrs and disrupt ESCRT function in mammalian cells. (A) EsxH<sub>Mt</sub>-FLAG, EsxG<sub>Mt</sub>-His, and Hrs-myc expressed in HEK293 cells. DMSO or MG132 were added 3 h prior to protein harvest and samples were analyzed by western blotting. Lanes 1’ and 2’ are identical to 1 and 2 except that twice the amount of protein was loaded. Quantification from three independent experiments is shown in Figure S4. (B) Immunoprecipitation (IP) of Hrs using antibody recognizing myc tag or isotype control from HEK293 cells expressing Hrs-myc and either EsxG<sub>Mt</sub>-His or EsxH<sub>Mt</sub>-FLAG. MG132 was used as pre-treatment. Western blot of IP and input were probed with antibodies as indicated. (C) IP of Hrs-myc in HEK293 cells with antibody recognizing myc tag or isotype control from HEK293 cells expressing Hrs-myc, EsxG<sub>Mt</sub>-His, and either EsxH<sub>Mt</sub>-FLAG or EsxH<sub>Mt</sub>-H76A-E77A-FLAG. MG132 was used as pre-treatment. Western blot of IP and input were probed with antibodies as indicated. (D) IP of C-terminal fragment of Hrs (amino acids 398–777) using antibody recognizing V5-tag or isotype control from HEK293 cells expressing Hrs-398–777-V5, EsxG<sub>Mt</sub>-His, and EsxH<sub>Mt</sub>-FLAG. MG132 was used as pre-treatment. Western blot of IP and input were probed with antibodies as indicated. (E) HEK293 cells transfected with indicated plasmids were incubated with EGF for 0 or 90 min prior to western analysis. (F–H) A549 cells transfected with plasmids or siRNAs were imaged 90 min after incubation with Alexa-488 EGF. In F, white lines indicate cell borders. (G) and (H), MFI
on mycobacterial phagosomes by two synergistic mechanisms: impaired recruitment and direct targeting by EssHM. How the activities of these various bacterial effectors are coordinated, whether they are required in concert or function in different cell types or at different time points post-infection, has not been explored. In order to evaluate the relative contribution of the EssHM-Hrs interaction to trafficking and intracellular survival during infection, we will have to identify mutations in EssHM that disrupt its binding to Hrs, but that do not interfere with its secretion from Mtb or disrupt bacterial iron acquisition.

It was surprising to us that it was possible to alter Mtb trafficking by overexpressing EssGM, EssHM, if Mtb normally expresses a "sub-optimal" amount to maximally alter phagosome maturation. Similarly, ESCRT and Rab7 appear to be sub-maximally inhibited, as further impairing their function by RNAi-mediated silencing also enhances the block in phagosome-lysosome fusion. Given that over-expression of EssHM by Mtb caused a greater effect on trafficking than Hrs silencing (Fig. 4B), EssHM may have additional cellular targets involved in cellular trafficking as well. One explanation for the observation that additional EssHM can further impair trafficking is that this reflects in vitro growth conditions, whereas, during infection in vivo, EssHM levels may be higher. An additional possibility is that EssGM, EssHM production is finely tuned to balance an opposing effect that is detrimental to the bacteria. For example, EssGM and EssHM generate prominent T cell responses, [14,15]. In addition, we found that although the Mtb strain that overexpresses EssGM, EssHM exhibited diminished co-localization with LAMP1 and enhanced co-localization with TIR, there was no difference in intracellular growth for this strain relative to control (Figure S7). Thus, overexpression of EssGM, EssHM while promoting trafficking, might come with an opposing intracellular fitness cost for bacteria.

In summary, our studies demonstrate that Mtb adapted Ess-3, an ancient microbial system for iron acquisition, to alter host cell physiology. Analogously, Ess-1, which is important for conjugation in Msmeg [61], mediates important host interactions that are critical for virulence, including permeabilizing the mycobacterial phagosome and altering phagosome maturation [5-9,62]. Thus, the duplication and adaptation of TSSs to new functions appears to be a particularly important evolutionary path to virulence in Mtb. The relatively low affinity between Hrs and its endogenous binding partners may have made it particularly susceptible to manipulation by diverse pathogens, from enveloped viruses to intracellular bacteria. The observation that macrophase control of infection is especially sensitive to ESCRT inhibition suggests ESCRT is a likely target of additional pathogens as well.

Materials and Methods
Detailed methods, including description of Y2H interaction mapping, plasmids, siRNAs, and Hill plot analysis, are provided in Text S1.

Tissue culture conditions
RAW264.7 and HEK293 cells were grown in Dulbecco’s Modified Eagle Medium (DMEM; Gibco), 20 mM HEPES, 2 mM L-glutamine, and 10% heat inactivated fetal bovine serum (hiFBS; Invitrogen). BMDMs were isolated from C57BL/6 mice as described [63] Penicillin/Streptomycin (Gibco), added for passaging, was omitted during infections. A549 cells were grown in RPMI 1640 Medium (Gibco), 2 mM L-glutamine, 1× Non-essential Amino Acids (Cellgro), and 10% hiFBS. Cells were grown at 37°C with 5% CO₂ atmosphere. siRNAs were transfected with Hiperfect (Qiagen). Plasmids were transfected into HEK293 cells with Effectene (Qiagen) and A549 cells with Lipofectamine 2000 (Invitrogen).

Bacterial strains and growth conditions
*M. tuberculosis* H37Rv, *M. bovis*-BCG, and *M. smegmatis* mc²155 were grown at 37°C to log phase in Middlebrook 7H9 media with 0.05% Tween 80, BBL, Middlebrook OADC Enrichment, and 0.2% glycerol. Plasmids were selected with 50 μg/ml kanamycin or hygromycin depending upon the resistance marker. To generate EssG, EssHFLAG and EssGM, EssHMFLAG for overexpression in mycobacteria, EssG-EssH was amplified from BCG (the EssG-EssH region is 100% identical between BCG and Mtb) and Msmeg genomic DNA, respectively, using primers described in Text S1. The PCR products were cloned into pSYMP under control of the hsp60 promoter [64].

Intracellular bacterial growth assay
RAW cells were seeded one day before infection or they were transfected with siRNAs two days prior to infection with a single cell suspension of Mtb (MOI=1−2.5), obtained as previously described [40]. The cells were extensively washed and lysed with 0.2% Triton X-100 3 hpi or 2d later and serial dilutions were plated on 7H10 or 7H11. CFU were calculated 15 to 21 d later.

Lysosomal trafficking assay
RAW cells were transfected with siRNAs for two days and then infected with a single cell suspension of BCG or Mtb (MOI=20) for 3 h, then washed extensively. Cells were fixed 24 hpi with 4% formaldehyde/PBS for BCG and with 1% paraformaldehyde/PBS overnight for Mtb and immunostained for LAMP1 (Abcam) or TIR (Invitrogen). For LysoTracker (Invitrogen) staining, unfixed RAW cells were incubated with 200 nM LysoTracker, washed twice in PBS, and visualized. Images were captured using the Nikon Eclipse TiE/B automated fluorescent microscope with Photometrics HQ@ Monochrome digital camera. 60 x z-stack images were acquired, deconvoluted, and analyzed using NIS-Elements DUO software (see Fig. S3 for details). Contrast was not altered prior to automated image analysis; for reproduced images, alterations were applied equally to all samples.

Recombinant protein binding assay
His-tagged EssGM-EssHM was purified as described in Text S1. Prior to inclusion of recombinant proteins in binding reactions they were centrifuged at 100,000 × g for 30 min to remove aggregated protein. To determine whether EssGM, EssHM binds to Hrs in a direct and saturable manner, 1.0 μg EssGM, EssHM-6XHis was bound to Ni-NTA beads and incubated with increasing amounts of purified, soluble Hrs (0−4 μg) in 20 mM HEPES (pH, 7.4), 150 mM KCl, and 0.05% Tween-20, with protease inhibitors (10 mM leupeptin, 1 μg/mL pepstatin, 0.3 mM aprotinin, and 1.74 μg/mL PMSF) for 1 h at 4°C. Beads were washed in PBST (0.1 M PBS 0.05% Tween-20) with 10 mM imidazole. Bound Hrs was analyzed by SDS-PAGE and
Co-immunoprecipitation and Western blotting

Cellular lysates were prepared in RIPA buffer with Halt Protease Inhibitor Cocktail (Thermo Scientific) and 10 μM N-ethylmaleimide (Sigma) and analyzed by western blotting. The antibodies used for western analysis are: actin (clone C4/ MAB1501, Millipore), Hrs (M79/sc-30221, Santa Cruz Biotechnology), Rab7 (117, Abcam), FLAG (F7425, Sigma), EGFR (#42678, Cell Signaling), and FK2 (Millipore). For co-immunoprecipitation, HEK293 cells transfected with Hrs-myc and Esx expression plasmids were treated with 20 μM MG132 (Calbiochem) for 3 h prior to mechanical lysis and incubated with Dynabeads Protein G (Novex, Life technologies) pre-bound to isotype control antibody (sc-2025, Santa Cruz Biotechnology), anti-myc antibody (sc-40/9E10, Santa Cruz Biotechnology), or anti-V5 antibody (Invitrogen), and bound proteins were analyzed by western blotting.

EGFR and EGF degradation assays

Two days after transfection, HEK293 cells were incubated in serum-free DMEM and treated with 100 ng/ml of recombinant human EGF (rh-EGF, R&D Systems) essentially as described [65]. Cells were harvested immediately prior to addition of EGF and 90 min later and EGFR analyzed by western blotting. EGFR trafficking was assessed similarly to described [66]. Two days after transfection of A49 cells with siRNA or DNA, cells were incubated with serum-free RPMI before addition of 25 μg/ml Alexa Fluor 488-EGF (Invitrogen) in EGF uptake media (RPMI, 2% BSA, 20 mM HEPES) at 4 ºC for 1 h. Cells were washed to remove unbound ligand, incubated at 37 ºC for 90 min, and examined by immunofluorescence microscopy.

Mycobacterial secretion of EsxH

To analyze secretion of EsxHFLAG or EsxHFLAG from Mtb and Msmeg, strains were grown to mid-log phase, washed with PBS, and inoculated into Sauton’s media. In Sauton’s media, they were grown to reach log phase (overnight in the case of Msmeg and for two days in case of Mtb). Thereafter, mycobacteria were pelleted by centrifugation. The supernatants were filtered through 0.22 μM filters followed by precipitation with 12% trichloroacetic acid. The precipitate was washed with ice-cold acetone, air dried, and resuspended in SDS sample buffer. The bacterial pellets were lysed by bead beating in lysis buffer (50 mM Tris-HCl pH 7.5, 5 mM EDTA, 0.6% SDS, 10 mM NaH2PO4, and protease inhibitor) with 0.1 mm zirconia/silica beads (BioSpec Products, Inc.). SDS-sample buffer was added, followed by boiling at 95°C for 5 min. Antibody to the pyruvate dehydrogenase E2 component sucB (Rv2215/dlaT) [67], a cytosolic protein, was used as a loading control and to indicate the degree of bacterial lysis. (B) MFI of phagosomal LAMP1 24 hpi in RAW cells treated with siRNAs and infected with Mtb containing EsxGM-EsxHM (red) or vector control (black); data points are the mean fluorescence intensity (MFI) around at least 70 phagosomes for each condition; p<0.0001 between the two Mtb strains for all conditions, unpaired Student’s t-test. (C) Composite images of cells infected with Mtb with autofluorescence of Mtb (blue) and LAMP1 (red). Regions indicated by yellow circles are shown in higher magnification in adjacent panels. (D) MFI of phagosomal LAMP1 24 hpi in RAW cells treated with siRNAs and infected with Mtb containing EsxGM-EsxHM (red) or vector control (black); data points are the mean fluorescence intensity (MFI) around at least 50 phagosomes for each condition; p<0.0001 between the two Mtb strains for all conditions, except Rab7 (p = 0.0005), unpaired Student’s t-test. (E) Composite images of cells infected with Mtb with autofluorescence of Mtb (blue) and LAMP1 (red). Regions indicated by yellow circles are shown in higher magnification. (F) MFI of phagosomal LAMP1 in RAW cells infected with Mtb containing the indicated plasmids 24 hpi. Bars show mean +/- SEM. ***p<0.0001, unpaired Student’s t-test.

doi:10.1371/journal.ppat.1003734.g004

Figure 4. EsxGM-EsxHM arrests phagosome maturation. (A) H377rv transformed with empty vector, EsxGM-EsxHM-FLAG, EsxGM-FLAG, EsxHM-FLAG or EsxGM-FLAG, H76A-E77A-FLAG were analyzed for the presence of EsxH in the pellet and culture filtrate (CF). DiaT (Rv2215), a cytosolic protein, was used as a loading control and to indicate the degree of bacterial lysis. (B) MFI of phagosomal LAMP1 24 hpi in RAW cells treated with siRNAs and infected with Mtb containing EsxGM-EsxHM (red) or vector control (black); data points are the mean fluorescence intensity (MFI) around at least 70 phagosomes for each condition; p<0.0001 between the two Mtb strains for all conditions, unpaired Student’s t-test. (C) Composite images of cells infected with Mtb with autofluorescence of Mtb (blue) and LAMP1 (red). Regions indicated by yellow circles are shown in higher magnification in adjacent panels. (D) MFI of phagosomal LAMP1 24 hpi in RAW cells treated with siRNAs and infected with Mtb containing EsxGM-EsxHM (red) or vector control (black); data points are the mean fluorescence intensity (MFI) around at least 50 phagosomes for each condition; p<0.0001 between the two Mtb strains for all conditions, except Rab7 (p = 0.0005), unpaired Student’s t-test. (E) Composite images of cells infected with Mtb with autofluorescence of Mtb (blue) and LAMP1 (red). Regions indicated by yellow circles are shown in higher magnification. (F) MFI of phagosomal LAMP1 in RAW cells infected with Mtb containing the indicated plasmids 24 hpi. Bars show mean +/- SEM. ***p<0.0001, unpaired Student’s t-test.

Supporting Information

Dataset S1  Secretome collection screened.

(XLS)

Dataset S2  Control collection screened.

(XLS)

Dataset S3  Y2H interactions between Mtb secretome and human ORFs.

(XLS)

Figure S1 siRNA-mediated depletion of Hrs and Rab7.

(A) RAW264.7 (RAW) cells were treated with 50 nM ON-TARGETplus individual siRNAs (#9–#12) targeting Hrs or control for 2 d. (B) RAW cells were treated with increasing concentration of siRNA#9 targeting Hrs for 2 or 5 days. (C) A549 cells treated with 50 nM Hrs siRNAs (#12) or control for 2 d. (A)–(C) Western blotting with antibody recognizing Hrs was used to assess silencing. (D) RAW cells were treated with 30 nM siRNA targeting Rab7 or control. Silencing was assessed 2 d later by western blotting using an antibody recognizing Rab7. (E) RAW cells treated with control siRNA (siCON) or siRNA targeting Hrs (#9 or #12) for 2 d were examined by immunofluorescence using antibodies against Hrs, shown in red, and ubiquitinated proteins (FK2) in green.

(TIF)

Figure S2 siRNAs targeting Hrs and Rab7 enhance the intracellular survival of BCG in BMDMs. 4×10⁶ BMDMs were transfected with 50 nM siRNA pools targeting Hrs (ONTARGETplus) or Rab7 (siGENOME) 6–10 d after harvest. 3 d later, they were infected with BCG (MOI of 2 to 5). CFU were enumerated 2 days post-infection and are normalized to the average number of CFU in control wells from two independent experiments. Results reflect the mean +/- SEM. *p<0.05; **p<0.01, unpaired Student’s t-test.

(TIF)

Figure S3 Automated image analysis of phagosome maturation

(A) For quantifying the degree of co-localization between bacteria and cellular markers or Lysotracker, images were background subtracted and analyzed using the Binary Operation Analysis within NIS Elements Software. Bacteria were selected in the green channel. The region the software has selected that corresponds to the bacteria is shown in red in the second panel. That region was expanded (dilate binary) and then eroded and a binary operation was performed to generate a “donut” in the region surrounding the bacteria. The region of interest (ROI) is shown in purple. The mean fluorescence intensity (MFI) in the
ROI was determined for the cellular marker. Bacteria were analyzed from at least three fields per sample per experiment. We confirmed that automated quantification closely paralleled manual quantification and visual scoring by a blinded observer. To further validate the automated analysis, we verified enhanced LAMP1 co-localization in macrophages pre-treated with IFN-γ, which promotes phagosome maturation [61]. RAW cells treated with control siRNA (siCON) were either pre-treated with IFN-γ or solvent control 24 hours prior to infection with Mtb-GFP. In IFN-γ pre-treated macrophages there is a significant shift in LAMP1 co-localization around bacterial phagosomes 24 hpi. Data points are the MFI of LAMP1 around bacteria; bars show mean +/− SEM; p<0.0001. (C) Co-localization of LAMP1, Lysotracker, and TIR with metabolically active BCG compared to co-localization with total BCG. RAW cells were treated with control siRNA (siCON) and infected with BCG constitutively expressing GFP (BCG-GFP) or BCG expressing GFP under a tetracycline inducible promoter (BCG-tet-GFP). AnTc was added 24 hpi to induce expression of GFP. Because it takes >12 h for the strain to become detectably GFP positive, co-localization between BCG-tet-GFP and LAMP1, LysoTracker, or TIR was measured at 48 hpi. For the BCG-GFP strain, LAMP1 and TIR were examined at 48 hpi and LysoTracker at 24 hpi. Data points are the MFI around bacteria; bars show mean +/− SEM; p value of Student’s t-test; ns-not significant. Whiskers reflect the minimum and maximum data points, while the cross bars show the median. (TIF)

Figure S4 Quantification of EsxHMs-FLAG in transfected HEK293 cells. EsxHMs was co-transfected with vector control, EsxGMs, or Hrs as indicated. Prior to protein harvest, cells were treated with DMSO or MG132. EsxHMs-FLAG levels were quantified from at least three independent experiments using ImageJ software. *p<0.05; **p<0.01, unpaired Student’s t-test; ns-not significant. Whiskers reflect the minimum and maximum data points, while the cross bars show the median. (TIF)

Figure S5 Treatment with MG132 does not result in higher molecular weight forms of the EsxH proteins. HEK293 cells were transfected with plasmids as indicated. Cells were either treated with DMSO or MG132 prior to protein harvest. Lysates were examined for mono- and polyubiquitinated proteins using the FK2 antibody. The EsxH proteins were visualized using the FLAG antibody. No differences were seen in the mobility of EsxHMs, EsxHMs, or EsxHMs-H76A/E77A in the presence of MG132. (TIF)

References

1. Armstrong JA, Hart JD (1971) Response of cultured macrophages to Mycobacterium tuberculosis, with observations on fusion of lysosomes with phagosomes. J Exp Med 134: 713–740.
2. Russell DG (2001) Mycobacterium tuberculosis: here today, and here tomorrow. Nat Rev Mol Cell Biol 2: 569–577.
3. Flanagan RS, Costo G, Grinstein S (2009) Antibacterial mechanisms of phagocytes and bacterial evasion strategies. Nat Rev Microbiol 7: 355–366.
4. Philips JA, Ernst JD (2012) Tuberculosis pathogenesis and immunity. Annu Rev Pathol 7: 353–384.
5. van der Wel N, Hara D, Houben D, Fuitama D, van Zon M, et al. (2007) M. tuberculosis and M. leprae translocate from the phagolysosome to the cytosol in murine and zebrafish cDNA, respectively. The NYU RNAi core, in particular, C. Yun and R. Dasgupta, provided assistance. We thank M. Carr (University of Leicester), H. Darwin, V. Torres, J. Ernst (NYU SOM), and their laboratory members, in particular L. Desvignes (NYU SOM), and members of the Philips lab for assistance and helpful discussions. J. Ernst, J. Wei (Harvard Medical School), and M. Chheda (Memorial Sloan-Kettering Cancer Center) provided comments on the manuscript.

Author Contributions
Conceived and designed the experiments: JAP AM AZ VT NS AW MP KP AD DR VK SK. Performed the experiments: JAP AM AZ VT NS AW MP KP AD DR VK SK. Analyzed the data: JAP AJB AM AZ VT NS AW MP KP AD DR VK SK. Contributed reagents/materials/analysis tools: DEH MV. Wrote the paper: JAP AM.
23. Venkatesan K, Rual JF, Vazquez A, Stelzl U, Lemmens I, et al. (2009) An
20. Katzmann DJ, Odorizzi G, Emr SD (2002) Receptor downregulation and
19. Abdallah AM, Savage ND, van Zon M, Wilson L, Vandenbroucke-Grauls CM,
17. Abdallah AM, Verboom T, Weerdenburg EM, Gey van Pittius NC, Mahasha
14. Hervas-Stubbs S, Majlessi L, Simsova M, Morova J, Rojas MJ, et al. (2006) High
37. Shembade N, Harhaj NS, Liebl DJ, Harhaj EW (2007) Essential role for
35. Watson RO, Manzanillo PS, Cox JS (2012) Extracellular M. tuberculosis DNA
33. Lange S, Perera S, Teh P, Chen J (2012) Obscurin and KCTD6 regulate cullin-
30. Sun J, Wang X, Lau A, Liao TY, Bucci C, et al. (2010) Mycobacterial nucleoside
28. Bach H, Papavinasasundaram KG, Wong D, Hmama Z, Av-Gay Y (2008)
25. Lightbody KL, Bighari D, Waters LC, Carey G, Bailey MA, et al. (2008) Molecular
23. Venkatesan K, Rual JF, Vazquez A, Stelzl U, Lemmens I, et al. (2009) An
22. Rual JF, Venkatesan K, Hao T, Hirozono-Kishikawa T, Driot A, et al. (2005)
20. Katzmann DJ, Odorizzi G, Emr SD (2002) Receptor downregulation and
19. Abdallah AM, Savage ND, van Zon M, Wilson L, Vandenbroucke-Grauls CM,
17. Abdallah AM, Verboom T, Weerdenburg EM, Gey van Pittius NC, Mahasha
14. Hervas-Stubbs S, Majlessi L, Simsova M, Morova J, Rojas MJ, et al. (2006) High
37. Shembade N, Harhaj NS, Liebl DJ, Harhaj EW (2007) Essential role for
35. Watson RO, Manzanillo PS, Cox JS (2012) Extracellular M. tuberculosis DNA
33. Lange S, Perera S, Teh P, Chen J (2012) Obscurin and KCTD6 regulate cullin-
30. Sun J, Wang X, Lau A, Liao TY, Bucci C, et al. (2010) Mycobacterial nucleoside
28. Bach H, Papavinasasundaram KG, Wong D, Hmama Z, Av-Gay Y (2008)
25. Lightbody KL, Bighari D, Waters LC, Carey G, Bailey MA, et al. (2008) Molecular
23. Venkatesan K, Rual JF, Vazquez A, Stelzl U, Lemmens I, et al. (2009) An
22. Rual JF, Venkatesan K, Hao T, Hirozono-Kishikawa T, Driot A, et al. (2005)
20. Katzmann DJ, Odorizzi G, Emr SD (2002) Receptor downregulation and
19. Abdallah AM, Savage ND, van Zon M, Wilson L, Vandenbroucke-Grauls CM,
17. Abdallah AM, Verboom T, Weerdenburg EM, Gey van Pittius NC, Mahasha
14. Hervas-Stubbs S, Majlessi L, Simsova M, Morova J, Rojas MJ, et al. (2006) High
37. Shembade N, Harhaj NS, Liebl DJ, Harhaj EW (2007) Essential role for
35. Watson RO, Manzanillo PS, Cox JS (2012) Extracellular M. tuberculosis DNA
33. Lange S, Perera S, Teh P, Chen J (2012) Obscurin and KCTD6 regulate cullin-
30. Sun J, Wang X, Lau A, Liao TY, Bucci C, et al. (2010) Mycobacterial nucleoside
28. Bach H, Papavinasasundaram KG, Wong D, Hmama Z, Av-Gay Y (2008)
25. Lightbody KL, Bighari D, Waters LC, Carey G, Bailey MA, et al. (2008) Molecular
23. Venkatesan K, Rual JF, Vazquez A, Stelzl U, Lemmens I, et al. (2009) An
22. Rual JF, Venkatesan K, Hao T, Hirozono-Kishikawa T, Driot A, et al. (2005)
20. Katzmann DJ, Odorizzi G, Emr SD (2002) Receptor downregulation and
19. Abdallah AM, Savage ND, van Zon M, Wilson L, Vandenbroucke-Grauls CM,