Crocin protects the renal tubular epithelial cells against high glucose-induced injury and oxidative stress via regulation of the SIRT1/Nrf2 pathway

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Abstract

Objective(s): Renal tubular damage is critical pathological feathers of diabetic nephropathy (DN). This study aimed to explore the protective activity and related mechanisms of crocin in renal epithelial cell injury induced by high glucose.

Materials and Methods: Renal tubular epithelial HK-2 cells were cultured with D-glucose to establish an in vitro DN model. Cell viability was evaluated by CCK-8 assay. Apoptosis was detected by Annexin V-FITC kit. Oxidative stress was evaluated by colorimetry. RT-qPCR was carried out to determine the mRNA expressions of NF-E2-related factor 2 (Nrf2) and its pathway genes. Western blot was applied to determine the protein expressions of Nrf2 and related proteins.

Results: High glucose (5.5, 30, and 50 mM D-glucose) decreased cell viability at 72 hr, which was attenuated by crocin (25 and 50 μM). Crocin also attenuated the high glucose (30 mM D-glucose) induced apoptosis of HK-2 cells, decreased MDA content, and increased SOD activity in culture media. Crocin increased mRNA levels of Nrf2, HO-1, and NQO1. Moreover, crocin increased protein expressions of Nrf2, Sirtuin 1 (SIRT1), and p-Akt (Ser473). Inhibition of Nrf2 using siRNA, and inhibitors of SIRT1 (nicotinamide, NAM, 20 μM) and PI3K/Akt (LY294002, 50 μM) all attenuated the protective effect of crocin. Nrf2 siRNA and NAM also partially attenuated the inhibitory effect on oxidative stress and increase in the Nrf2 protein by crocin treatment.

Conclusion: Crocin protects renal epithelial cells against injury induced by high glucose, and the mechanism is associated with partial activation of the SIRT1-Nrf2 pathway.

Introduction

Diabetic nephropathy (DN) is a progressive microvascular complication of diabetes mellitus and is one of the leading causes of DN patients (1). The pathology of DN is characterized by basement membrane thickening, extracellular matrix accumulation, and subsequent albuminuria (2). DN is driven by numerous factors, among which hyperglycemia alters cellular signaling and metabolism, contributes to glomerular hyperfiltration (3), increases advanced glycation end products (AGEs) (4), oxidative stress (5), and inflammation (6).

Oxidative stress is considered an important pathological process that links hyperglycemia to vascular complications through changing renal metabolism and hemodynamics, thereby leading to adverse effects on kidney tissues (7). Excessive oxidative stress could offset the endogenous antioxidant defense system, and then oxidize various biomolecules like DNA, proteins, carbohydrates, and lipids (8). Hyperglycemia aggravates oxidative stress through various mechanisms (9), which involves the progression of diabetic complications (10).

Crocin is one major pharmacologically active constituent of saffron (Crocus sativus L.). Crocin has demonstrated various pharmacological effects, including anti-inflammatory, anticonvulsant, and anti-tumor activities (11), and it is reported to inhibit cisplatin-induced oxidative stress in rats (12). Current studies have shown that crocin exhibits significant antioxidant activity and scavenges ROS (13), and this may be related to its endogenous antioxidant enzymatic activities (14).

In this study, we established an in vitro injury model using HK-2 human renal tubular epithelial cells, explored the protective effect of crocin on high glucose-induced injury, and evaluated cell viability, apoptosis, oxidative stress, and related molecular mechanisms.

Materials and Methods

Cell culture

The HK-2 cells were purchased from the Chinese academy of sciences. HK-2 cells were cultured in a low glucose DMEM medium (5.5 mM D-glucose) with 10% FBS and penicillin/streptomycin (HyClone). Cells were cultured in a 5% CO₂ incubator at 37 °C.

Experiment grouping

The HK-2 cells were seeded in 96 well culture plates (2×10⁴ cells per well, in 100 μl DMEM medium). When cells grew to 80% confluence, they were divided into the following groups: (1) Control group: cells were incubated with only 5.5 mM D-glucose (normal glucose); (2) High-glucose group: cells were incubated with 5.5 mM D-glucose plus 30 or 50 mM D-glucose; (3) Crocin group: cells were incubated with 25 or 50 μM crocin plus 30 or 50 mM D-glucose; (4) Nicotinamide group: cells were incubated with 20 μM nicotinamide plus 30 or 50 mM D-glucose; (5) Sirtuin 1 group: cells were incubated with 20 μM nicotinamide plus 30 or 50 mM D-glucose, and 20 μM Sirtuin 1 inhibitor; (6) PI3K/Akt group: cells were incubated with 50 μM LY294002 plus 30 or 50 mM D-glucose, and 20 μM Sirtuin 1 inhibitor; (7) Combined group: cells were incubated with 25 or 50 μM crocin plus 30 or 50 mM D-glucose, 20 μM nicotinamide plus 30 or 50 mM D-glucose, and 20 μM Sirtuin 1 inhibitor.

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glucose group: cells were incubated with D-glucose at 15, 30, and 50 mM for up to 72 hr; (3) Crocin group: cells were incubated with 30 mM D-glucose and crocin at 5, 10, 25, 50, and 100 μM for 48 and 72 hr.

**Cell viability assay**

After various treatments, HK-2 cells were added with CCK-8 solution (10 μl; Beyotime, Shanghai, China), and were incubated at 37°C for 2 hr. The absorbance at 450 nm was measured using a microplate reader.

**Apoptosis assay**

The HK-2 cells were seeded in 24-well plates (3×10^4 cells/well) and cultured in a medium containing 30 mM D-glucose, followed by treatment with crocin 25 and 50 μM for 72 hr. Cells were collected and washed with cold PBS and were incubated with 5 μl Annexin V-FITC and 5 μl PI in the dark for 15 min. Apoptosis was assessed by a flow cytometer (Becton Dickinson, San Francisco, CA, USA).

**Oxidative stress test**

Following the treatment, HK-2 cells were lysed using the RIPA lysis buffer. The supernatant was collected for the experiments. The oxidative and antioxidant proteins were measured by spectrophotometry. (Nanjing Jiancheng Bioengineering Research Institute, Nanjing, China). The determination of malondialdehyde (MDA) (cat no. S0131S) and superoxide dismutase (SOD) activity (cat no. S0109) were investigated by commercial assay kits (Beyotime, Jiangsu, China). Eventually, MDA activity was measured with a microplate reader at 532 nm absorbance and expressed as μmol/l. SOD activity was measured at 520 nm absorbance and expressed as U/L.

**siRNA transfection**

The Nrf2 small interference RNA (siRNA) was designed and synthesized by Shanghai GenePharma Co., Ltd, using the following primer: 5′-CAC ACT GGA TCA GAC AGG AGG ATA T-3′ for transfection. Following the manufacturer's instructions, the siRNA was transfected into the HK-2 cells with Lipofectamine® 2000 transfection kit (Invitrogen, USA). Briefly, the 5×10^4 HK-2 cells were seeded into the 6-well plates in a complete medium and transfected with high glucose and crocin and used for the experiments.

**RT-qPCR**

Total RNA was extracted by Trizol agent and was used to generate cDNA by the Superscript III enzyme (Life Technologies). RT-qPCR was carried out using SYBR® Premix Ex Taq™ II (cat. no. RR820L; Takara Bio, Inc.). The PCR conditions were as follows: denaturation at 94°C for 5 min; followed by 40 cycles at 94°C for 5 sec and 60°C for 60 sec. PCR primers for Nrf2, HO-1, NQO1, and GAPDH were listed in Table 1. Reaction systems contained 2 μl template cDNA, 10 μl SYBR-Green PCR master mix, 1 μl forward and reverse primers, and 7.2 μl deionized water. PCR results were quantified by using the 2−ΔΔCt method.

**Western blotting**

Total protein was extracted from HK-2 cells using RIPA buffer. Protein samples (50 μg) were subjected to 12% SDS-PAGE and then transferred to a PVDF membrane (Millipore, Bedford, USA) for detection with appropriate antibodies. Membranes were blocked with 5% nonfat dry milk in TBST buffer and treated with primary antibodies against human Nrf2 (#12721), SIRT1 (#9475), and p-Akt1 (Ser473) (#4060) (all 1:1000 dilutions; Cell Signaling Technology, USA) at 4°C overnight, washed and then incubated with HRP-linked secondary antibody (1:1000 dilution) for 1 hr at room temperature. A chemiluminescent detection system (ECL, UK) was utilized for visualization. The gray value of the target band was normalized to that of β-actin.

**Statistical analysis**

Data from at least three independent experiments were presented as means ± standard deviation (SD), and analyzed by SPSS 20.0 statistical software (SPSS Inc., Chicago, IL, USA). Unpaired Student's t-test or ANOVA was used to analyze the differences between two or more groups. P<0.05 was considered as statistical significance.

**Results**

**High glucose decreased cell viability of high glucose-induced HK-2 cells**

HK-2 cells were incubated with D-glucose (5.5, 15, 30, or 50 mM) for 72 hr. D-glucose treatment at 15, 30, and 50 mM significantly reduced cell viability (Figure 1a). Then HK-2 cells were incubated with 5.5 and 30 mM D-glucose for different time points. Compared to cells with normal glucose, 30 mM D-glucose markedly reduced viability at 48 and 72 hr (Figure 1b). We then examined the toxicity of crocin at various concentrations. Crocin had little effect on cell viability between 10, 25, and 50 μM at 48 and 72 hr. However, crocin at 100 μM dramatically reduced cell viability at 48 and 72 hr (Figures 1c, 1d). Thus, we chose crocin at 25 and 50 μM for further experimental analysis.

**Effect of crocin on cell viability and apoptosis in high glucose-induced HK-2 cells**

HK-2 cells were incubated with 30 mM D-glucose, followed by crocin treatment (25 and 50 μM) for 48 and 72 hr. Crocin significantly attenuated the D-glucose-
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induced decrease in viability at 48 and 72 hr (P<0.05) (Figure 2a). In order to explore whether apoptosis mediates reduced cell survival by crocin, Annexin V/PI staining was performed. High glucose (30 mM D-glucose) markedly induced apoptosis at 72 hr, and crocin at 25 and 50 μM both significantly attenuated the increase in apoptotic rate induced by high glucose (Figures 2b, 2c).

**Crocin activated Nrf2 pathway and inhibited oxidative stress in high glucose-induced HK-2 cells**

HK-2 cells were incubated with 30 mM D-glucose and crocin (25 and 50 μM) for 72 hr co-culture. High glucose markedly reduced the mRNA and protein levels of Nrf2. Co-treatment with crocin increased mRNA expressions of Nrf2 (Figure 3a), HO-1 (Figure 3b), and NQO1 (Figure 3c). Then Western blotting was performed and protein expression of Nrf2 was also increased by crocin (Figure 3d). We then investigated the effect of crocin on oxidative stress. Crocin markedly reversed 30 mM D-glucose increase in MDA content and decrease SOD activity in HK-2 cells (Figures 3e, 3f).

**Crocin protected HK-2 cells against high glucose-induced injury via the SIRT1-Nrf2 pathway**

Western blot was carried out to measure the protein levels of SIRT1 (a) and p-Akt1 (b). HK-2 cells were pre-incubated with Nrf2 siRNA, SIRT1 specific inhibitor nicotinamide (NAM, 20 μM), or with LY294002 (a specific inhibitor of PI3K/Akt1, 50 μM) before crocin treatment.
In this study, we found that crocin attenuated the high glucose-induced reduction in cell viability. Crocin also increased mRNA levels of Nrf2, HO-1, and NQO1, and suppressed oxidative stress in high glucose-induced HK-2 cells. Crocin increased protein expressions of SIRT1 and p-Akt (Ser473), and inhibition of SIRT1 (nicotinamide, NAM, 20 μM) and PI3K/Akt (LY294002, 50 μM) all attenuated the protective effect of crocin on high glucose-induced HK-2 cells. Therefore, the protective effect of crocin might be related to the modulation of Nrf2, SIRT1, and Akt molecules.

Crocin showed suppressive effects on high glucose-induced toxicity and oxidative injury. These results support the concept that the high glucose is a potent inducer of oxidative stress, and thereby initiates and aggravates DN [9]. Our results are consistent with previous reports that crocin relieved DN and improved renal function in diabetic rats through inhibition of oxidative stress and inflammation (15, 16). Moreover, crocin also protects podocytes from high glucose-induced oxidative injury (17). Our study provides the same protective effect on renal tubular cells. Recently, changes in the renal tubules, are increasingly implicated in the development and progression of diabetic kidney disease (18). The renal tubules play the same important roles as glomeruli and podocytes in the pathogenesis of DN (19), and tubular injury has been postulated as a critical contributor to the early DN (20). Thus, our study provides another target tissue to elucidate the roles and mechanisms of crocin in DN.

This study showed that crocin activated Nrf2 and downstream antioxidant response in combating high glucose-induced injury. Nrf2 is an endogenous antioxidant defense system that maintains cellular homeostasis under stress conditions. Nrf2 could slow down the progression of DN (21). High glucose treatment inhibited Nrf2 expression in renal tubular epithelial cells, and overexpression of Nrf2 reduced renal cell damage in diabetes (22). Our study adds crocin as a new Nrf2 activator in high glucose-induced renal epithelial cells, which is supported by other reports that crocin modulates Nrf2 in hepatic ischemia/reperfusion injury (23) and cigarette smoke-induced lung injury (24).

Our study showed that crocin increased SIRT1 protein, which was essential for the protective effect of crocin in high glucose-induced injury. Sirtuin 1 (SIRT1) modulates NAD(+) coenzymes and maintains cellular energetic metabolism and oxidative state. SIRT1 deficits are associated with diabetes mellitus and kidney diseases. SIRT1 promoted the survival of kidney cells and protected against apoptosis in renal tubules (25). In renal tubular injury induced by hyperglycemia, SIRT1 activation could attenuate renal tubular injury through inhibiting apoptosis (26), which is consistent with our results that SIRT1 inhibition abolished the protective effect of crocin. Our study also showed that SIRT1 might lie upstream of Nrf2, as SIRT1 inhibition abolished the increase of Nrf2 protein by crocin. High glucose could induce injury of renal epithelial cells via activation of the SIRT1/NF-kB/Nrf2 pathway (27). The SIRT1/Nrf2 pathway was also activated by crocin and mediated alleviation of myocardial ischemia/reperfusion-induced injury (28). Our study firstly reported SIRT1/Nrf2 activation by crocin in high glucose-induced renal epithelial cells, and provided experimental data for further investigation on mechanisms of crocin in DN.

In the present study, crocin enhanced p-Akt protein, which was essential for protective effect on high glucose-induced injury. Akt1 is a signal protein that modulates many cellular processes such as glucose uptake, cell survival, and angiogenesis in the kidney, and its activation showed protective effects in DN (29), and also inhibited high glucose-induced apoptosis of renal epithelial cells (30). This can explain how Akt1 inhibition could abolish high glucose-induced cellular injury by high glucose. Our results are in accordance with other reports that crocin activated Akt1 in microglial cells of diabetic retinopathy (31) and myocardial ischemia/reperfusion injury (32). In these reports, Akt1 activation was associated with induction of autophagy, which protected podocyte function and showed a preventive effect on DN progression (33). However, it remains unclear about the role of autophagy in the protective effect of crocin on high glucose-induced renal tubular injury. Moreover, in our study Akt1 inhibition could not attenuate oxidative stress and only partially attenuated the increase in Nrf2 protein. This indicates that Akt1 activation by crocin may further activate pathways other than Nrf2, especially in cellular survival.

Discussion

In this study, we found that crocin attenuated the high glucose-induced reduction in cell viability. Crocin also increased mRNA levels of Nrf2, HO-1, and NQO1, and suppressed oxidative stress in high glucose-induced HK-2 cells. Crocin increased protein expressions of SIRT1 and p-Akt (Ser473), and inhibition of SIRT1 (nicotinamide, NAM, 20 μM) and PI3K/Akt (LY294002, 50 μM) all attenuated the protective effect of crocin on high glucose-induced HK-2 cells. Therefore, the protective effect of crocin might be related to the modulation of Nrf2, SIRT1, and Akt molecules.

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Conclusion

Crocin attenuates high glucose-induced oxidative injury in renal tubular epithelial cells, which is related to the SIRT1-Nrf2 pathway. Crocin may be a promising therapeutic agent of DN, especially through targeting renal tubules against oxidative stress initiated by high glucose.

Acknowledgment

Not applicable.

Authors’ Contributions

JZ performed experiments and wrote the manuscript; XZ, HZ, and JW performed experiments and collected data; JM analyzed the data and revised the manuscript; MG designed and supervised the study. All authors have read and approved the manuscript.

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Conflicts of Interest
The authors declared no conflicts of interest with other people or organizations.

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