A BDNF loop-domain mimetic acutely reverses spontaneous apneas and respiratory abnormalities during behavioral arousal in a mouse model of Rett syndrome

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ABSTRACT
Reduced levels of brain-derived neurotrophic factor (BDNF) are thought to contribute to the pathophysiology of Rett syndrome (RTT), a severe neurodevelopmental disorder caused by loss-of-function mutations in the gene encoding methyl-CpG-binding protein 2 (MeCP2). In MeCP2 mutant mice, BDNF deficits have been associated with breathing abnormalities, a core feature of RTT, as well as with synaptic hyperexcitability within the brainstem respiratory network. Application of BDNF can reverse hyperexcitability in acute brainstem slices from MeCP2-null mice, suggesting that therapies targeting BDNF or its receptor, TrkB, could be effective at acute reversal of respiratory abnormalities in RTT. Therefore, we examined the ability of LM22A-4, a small-molecule BDNF loop-domain mimetic and TrkB partial agonist, to modulate synaptic excitability within respiratory cell groups in the brainstem nucleus tractus solitarius (nTS) and to acutely reverse abnormalities in breathing at rest and during behavioral arousal in MeCP2 mutants. Patch-clamp recordings in MeCP2-null brainstem slices demonstrated that LM22A-4 decreases excitability at primary afferent synapses in the nTS by reducing the amplitude of evoked excitatory postsynaptic currents and the frequency of spontaneous and miniature excitatory postsynaptic currents. In vivo, acute treatment of MeCP2-null and -heterozygous mutants with LM22A-4 completely eliminated spontaneous apneas in resting animals, without sedation. Moreover, we demonstrate that respiratory dysregulation during behavioral arousal, a feature of human RTT, is also reversed in MeCP2 mutants by acute treatment with LM22A-4. Together, these data support the hypothesis that reduced BDNF signaling and respiratory dysfunction in RTT are linked, and establish the proof-of-concept that treatment with a small-molecule signaling and respiratory dysfunction in RTT are linked, and establish the proof-of-concept that treatment with a small-molecule structural mimetic of a BDNF loop domain and a TrkB partial agonist can acutely reverse abnormal breathing at rest and in response to behavioral arousal in symptomatic RTT mice.

KEY WORDS: MeCP2, Brain-derived neurotrophic factor (BDNF), Respiration, Brainstem, Arousal

INTRODUCTION
Complex respiratory disturbances, including atypical respiratory pauses and apneas, are a prominent feature of Rett syndrome (RTT), severely impacting health and quality of life (Katz et al., 2009; Ramirez et al., 2013). The severity of breathing dysfunction in individuals with RTT is strongly influenced by behavioral state: respiratory phenotypes worsen when patients are agitated and improve when patients are relaxed or sleeping (Katz et al., 2009; Ramirez et al., 2013). Increasing evidence from mouse models suggests that respiratory dysfunction in RTT is associated with a more excited default state in the brainstem respiratory network, including in cell groups involved in respiratory pattern generation (preBotzinger complex) and modulation [nucleus Koelliker-Fuse, the nucleus locus coeruleus and the nucleus of the solitary tract (nTS)] (Stettner et al., 2007; Katz et al., 2009; Ramirez et al., 2013). Recent findings of respiratory hyperreflexia in MeCP2 mutant mice (Roux et al., 2008; Voiturou et al., 2009) are consistent with network hyperexcitability, particularly in the nTS, which is the principal relay for primary afferent inputs to respiratory reflex pathways. In particular, hyperexcitability within lateral subnuclei of the MeCP2 mutant nTS, where pulmonary stretch receptors form the first synapse in the Hering-Breuer reflex (HBR) pathway (Kubin et al., 2006), would be expected to decrease the activation threshold for the inspiratory off-switch and thereby promote the generation of apneas in RTT. These data suggest that therapeutic strategies aimed at restoring normal sensory gating in nTS by reducing synaptic hyperexcitability might ameliorate abnormal breathing in MeCP2 mutant mice.

Brain-derived neurotrophic factor (BDNF), which is in deficit in RTT (Li and Pozzo-Miller, 2014; Katz, 2014), normally modulates excitability at primary afferent synapses in the nTS by inhibiting postsynaptic responses to glutamatergic excitation (Balkowiec and Katz, 2002). Consistent with these findings, short-term exposure to exogenous BDNF reduces synaptic hyperexcitability in the nTS in brainstem slices prepared from MeCP2-null mice (Kline et al., 2010), raising the possibility that restoration of BDNF-TrkB signaling might acutely improve respiratory control in RTT. However, BDNF itself does not have favorable drug-like characteristics, i.e. limited half-life and poor blood-brain barrier penetration, thus motivating the search for alternative approaches to enhancing BDNF-TrkB signaling in RTT (Ogier et al., 2007; Deogracias et al., 2012; Johnson et al., 2012; Schmid et al., 2012). In the present study, we used in vitro electrophysiology and in vivo plethysmography to examine the acute effects of LM22A-4, a small-molecule non-peptide BDNF loop-domain mimetic (Massa et al., 2010), on synaptic hyperexcitability in respiratory subnuclei of the nTS, as well as on abnormal breathing at rest and during behavioral arousal in MeCP2 mutant mice. We previously found that chronic daily administration of a low dose of LM22A-4 [50 mg/kg body weight,
intrapertoneally (i.p.) reverses deficits in TrkB signaling in the brainstem in MeCP2 mutant mice and improves resting breathing frequency (Schnait et al., 2012). In the present study, we unexpectedly found that acute administration of LM22A-4 results in complete reversal of spontaneous apneas, a key feature of breathing dysfunction in RTT. In addition, the present study reveals a potential synaptic mechanism for this acute effect by demonstrating that LM22A-4 decreases synaptic excitability in brainstem respiratory nuclei in MeCP2-null mice. Moreover, we demonstrate that respiratory dysregulation during behavioral arousal, a feature of human RTT, is also reversed in MeCP2 mutants by acute treatment with LM22A-4. Our results are consistent with the hypothesis that deficits in BDNF-TrkB signaling contribute to breathing abnormalities in RTT and support the possibility that BDNF loop-domain mimetics could be useful in the acute treatment of respiratory dysfunction, including dysregulated responses to behavioral arousal, in RTT patients.

RESULTS

LM22A-4 reduces synaptic excitation in the lateral, ventrolateral and interstitial subnuclei of MeCP2-null mice

We previously showed that MeCP2-null mice exhibit synaptic hyperexcitability within respiratory cell groups in the nTS, including in the lateral, ventrolateral and interstitial subnuclei (referred to here collectively as lnTS). Therefore, we decided to use the lnTS as a model system to test the ability of LM22A-4 to reduce synaptic hyperexcitability in the brainstem respiratory network of MeCP2-null mice. Male null mice, rather than female heterozygotes, were used for these experiments because the heterozygotes are mosaic for wild-type (WT) and MeCP2 mutant cells, which cannot be distinguished in the in vivo slice preparation.

Synaptic hyperexcitability in the MeCP2-null InTS

Intracellular recordings of evoked, spontaneous and miniature excitatory postsynaptic currents (eEPSC, sPSC and mEPSC, respectively) from lnTS second-order neurons were performed to confirm and define in more detail the hyperexcitability phenotype previously observed in MeCP2 nulls (Nulls) (Kline et al., 2010; Kron et al., 2012). Indeed, upon 0.5 Hz train stimulation, sPSC amplitudes were significantly larger in the Null InTS compared with WT InTS (representative trace in Fig. 1A; WT, 266.0±18.1 pA, n=48; Null, 430.2±43.0 pA, n=71, P<0.01, unpaired Student’s t-test). Similarly, the instantaneous frequency of spontaneous synaptic currents was significantly higher in the Nulls, resulting in a strong trend towards a higher number of events within a 2-minute period as well (representative trace in Fig. 1B, sPSC frequency, WT, 10.4±1.3 Hz, n=57; Null, 21.8±1.5 Hz, n=53, P<0.05; number of events: WT, 599±84.5; Null, 772±68.9, P=0.07), with no significant effect of genotype on sPSC amplitudes. In addition, the frequency and amplitude distribution plots for mEPSCs in the presence of tetraodotxin (TTX, 0.5 μM) and bicuculline (10 μM) showed significant right shifts in the Nulls as compared with WT, indicating higher mEPSC frequency and amplitude (Null, n=13; WT, n=11; Kolmogorov-Smirnov test, P<0.05; Fig. 1C). We did not detect genotype differences in membrane resistance (R膜) and membrane capacitance (C膜); however, capacitance (C膜) was significantly reduced in the Nulls compared with WT (C膜 WT, 34.5±1.8 μF; Null, 29.6±1.1, P<0.05).

LM22A-4 acutely reduces synaptic hyperexcitability in the Null InTS

In slices from Null mice, bath-application of LM22A-4 (5 μM) significantly reduced eEPSC amplitudes in second-order lnTS neurons compared with pre-drug control recordings within 20 minutes (Fig. 2A, pre-drug control, 466.0±44.7 pA, LM22A-4, maximum effect, 377.4±35.0 pA, n=18, P<0.001, paired t-test). Specifically, eEPSC amplitudes decreased on average by 11.3±3.9% after 5-10 minutes of drug exposure (P<0.01 repeated measures (RM)-ANOVA), by 13.6±3.0% after 10-15 minutes, and by 17.1±4.0% after 15-20 minutes (P<0.001 RM-ANOVA). In addition, LM22A-4 caused a significant reduction in sPSC frequency as indicated by the left shift of the cumulative frequency distribution (Fig. 2B, P<0.05). As with eEPSC amplitudes, this effect was also time-dependent (average reduction in sPSC frequency by 21.8±4.5% after 8-10 minutes, by 24.6±3.1% after 13-15 minutes, and by 28.0±6.3% after 18-20 minutes of drug exposure, P<0.001 RM-ANOVA). Unlike eEPSC amplitudes, mean sPSC amplitudes were not affected by LM22A-4 (pre-drug control, 33.7±2.5 pA, LM22A-5, maximum effect, 33.7±2.7 pA, n.s.). In addition, LM22A-4 reduced both the frequency and, to a lesser degree, the amplitude of mEPSCs in the Nulls, reflected in left-shifts of frequency and amplitude distribution plots (n=13; Kolmogorov-Smirnov test, P<0.05; Fig. 2C). In contrast to Nulls, LM22A-4 had no significant effect on WT eEPSC amplitudes and mean sPSC frequency and amplitude (supplementary material Fig. S1A, RM-ANOVA, n.s.). Analyses of mEPSC frequency and
amplitude distribution curves in the presence and absence of LM22A-4 revealed virtually identical distributions in Wt cells (supplementary material Fig. S1B).

LM22A-4 effects are blocked by K252a, a non-selective inhibitor of receptor tyrosine kinases
Although LM22A-4 has previously been shown to selectively activate TrkB in vitro (Massa et al., 2010), this is the first study in which LM22A-4 has been applied to a slice preparation to test its effects on synaptic activity. Therefore, we next investigated whether or not prior application of K252a, a non-selective inhibitor of receptor tyrosine kinases, would reduce or eliminate the LM22A-4-mediated reduction of synaptic excitability in Nulls. Indeed, in the presence of K252a (100 nM), the effects of LM22A-4 on eEPSC amplitudes were abolished (Fig. 3A). Analyses of sPSC and mEPSC frequency distribution curves before and after LM22A-4 application in the presence of K252a revealed virtually overlapping distributions despite small shifts in some regions that reach statistical significance (Fig. 3B,C).

LM22A-4 acutely reverses apneic breathing in Mecp2 heterozygotes and nulls
Our electrophysiology data demonstrated that, in Mecp2-null mice, LM22A-4 acutely reduces excitability at primary afferent synapses within the lnTS, a region that is crucial for regulation of the inspiratory off-switch (Kubin et al., 1985). Therefore, we next sought to determine whether or not acute treatment with LM22A-4 in vivo would ameliorate the apnea phenotype in Mecp2-null (Null) and -heterozygous (Het) mice, both genotypes exhibited qualitatively similar breathing phenotypes, which were more severe in the Nulls. For each gender, animals were divided into four groups, i.e. female Wt saline, n=15; female Wt LM22A-4, n=14; female Het saline, n=18; female Het LM22A-4, n=17 and male Wt saline, n=11; male Wt LM22A-4, n=7; male Null saline, n=6; male Null LM22A-4, n=5 (female mice were 13-15 weeks of age and male mice were 6-7 weeks of age). Analysis of quiet breathing in female Wt and Het mice revealed that saline-treated Hets exhibited approximately twice as many apneas per minute as saline-treated Wt animals (Fig. 4A,B). However, LM22A-4 treatment reversed the Het apnea phenotype, restoring the number of apneas to female Wt levels [Fig. 4Bi; apneas/minute: Wt saline, 0.45±0.05; Wt LM22A-4, 0.44±0.09; Het saline, 0.34±0.10; Het LM22A-4, 0.47±0.06, P=0.001 (** in Fig. 4Bi) versus all other groups, ANOVA], even at doses as low as 5 mg/kg body weight (supplementary material Fig. S2). LM22A-4 had no effect on the number of apneas in Wt animals, and apnea length, respiratory frequency, inspiratory time, expiratory time and total breath duration were not affected by LM22A-4 treatment in animals of...
either genotype (not shown). Saline-treated male Nulls exhibited approximately four times as many apneas per minute as saline-treated Wt, and, as in HetS, LM22A-4 treatment restored the number of apneas to Wt levels [Fig. 4Bii; apneas/minute: Wt saline, 0.42±0.06; Wt LM22A-4, 0.30±0.07; Null saline, 1.63±0.24; Null LM22A-4, 0.78±0.22, *P*<0.001 (*** in Fig. 4Bii) versus all other groups, ANOVA].

To determine whether or not LM22A-4 treatment had prolonged effects on the apnea index, plethysmographic recordings were obtained from a subset of female Wt and Het mice 24 hours after LM22A-4 injection. These experiments demonstrated that the number of apneas per minute in LM22A-4-treated Hets had returned to the level of saline-treated Hets within 24 hours (apneas/minute: Wt saline, 0.56; Het saline, 1.15; Het LM22A-4, 1.52, *P*<0.01 versus all other groups, ANOVA). To test the hypothesis that this abnormal arousal phenotype is linked to reduced TrkB signaling, 6- to 8-week-old Wt and Null mice were treated with either saline or LM22A-4 (150 mg/kg body weight, i.p.) 1 hour prior to auditory stimulation. In Wt animals, we saw no effect of saline or drug treatment on the respiratory response to the auditory pulse. However, in Nulls, LM22A-4 treatment completely abolished the abnormal respiratory response to auditory stimulation, whereas saline injection
breathing and abnormal respiratory responses to sensory arousal. Our data support a link between reduced BDNF signaling in subnuclei in the nTS and the apnea phenotype characteristic of RTT. The present results demonstrate the reversibility of synaptic and behavioral phenotypes associated with abnormal breathing in Mecp2 mutants, including other BDNF-responsive cell populations in the brainstem respiratory network that regulate respiratory pattern generation, including the preBötzinger complex (Thoby-Brion et al., 2003) and nucleus Kolliker-Fuse (Kron et al., 2007).

**DISCUSSION**

The present results demonstrate the reversibility of synaptic and behavioral phenotypes associated with abnormal breathing in symptomatic Mecp2 mutant mice, a model of RTT. Specifically, we found that LM22A-4, a BDNF loop-domain mimetic and TrkB partial agonist, reduces synaptic hyperexcitability within respiratory subnuclei in the nTS in vitro, and acutely eliminates apneic breathing and abnormal respiratory responses to sensory arousal following systemic administration in vivo.

**Acute reversal of apneas during resting ventilation**

Our data support a link between reduced BDNF signaling in Mecp2 mutants and the apneic breathing phenotype characteristic of RTT. Moreover, these data are consistent with previous results demonstrating that synaptic hyperexcitability in nTS is associated with reduced levels of BDNF (Wang et al., 2006; Oger et al., 2007; Kline et al., 2010) and can be reversed by the application of exogenous BDNF in isolated brainstem slices (Kline et al., 2010). We hypothesize, therefore, that the ability of LM22A-4 to reduce or eliminate apneas in Mecp2 mutants is directly related to its ability to reduce synaptic excitability in nTS and thereby restore normal sensory gating of the inspiratory off-switch, possibly by enhancing input-output coupling in the HBR pathway (Diugrás et al., 2013). The present findings do not rule out the possibility that LM22A-4 might also act at sites other than the nTS to improve breathing in Mecp2 mutants, including other BDNF-responsive cell populations in the brainstem respiratory network that regulate respiratory pattern generation, including the preBötzinger complex (Thoby-Brion et al., 2003) and nucleus Kolliker-Fuse (Kron et al., 2007).

We found that hyperexcitability at primary afferent synapses in the nTS in Mecp2-null mice is characterized by an increase in the frequency of spontaneous and miniature excitatory currents and an increase in the amplitude of currents evoked by primary afferent stimulation, suggesting both pre- and postsynaptic effects, respectively, of MeCP2 loss. Moreover, both of these synaptic phenotypes are reduced by application of LM22A-4. The reduction in eEPSC and mEPSC amplitudes following LM22A-4 treatment parallels previous results obtained with exogenous BDNF (Kline et al., 2010) and is consistent with a role for BDNF in modulating excitatory transmission at primary afferent synapses in the nTS by inhibiting postsynaptic AMPA receptors (Balkowiec et al., 2000). Although underlying mechanisms remain to be defined, a similar role for acute TrkB activation in inhibiting glutamatergic transmission has been described at excitatory synapses onto parvalbumin-positive cortical interneurons (Jiang et al., 2004). Given previous evidence that BDNF-TrkB signaling can positively or negatively regulate AMPA receptor trafficking to the plasma membrane (Rutherford et al., 1997; Nansawa-Saito et al., 2002; Caldeira et al., 2007; Li and Keifer, 2008; Keifer and Zheng, 2010; Reimers et al., 2014), reduced AMPA receptor availability might underlie the inhibitory effect of LM22A-4 on postsynaptic excitability in the mutant nTS. In addition, the reduction in pEPSC frequency following LM22A-4 application indicates a presynaptic effect that is most likely mediated by an indirect pathway involving TrkB-expressing interneurons, because primary afferent inputs to the nTS do not express TrkB by 5 weeks of age (Kline et al., 2010).
Acute reversal of abnormal respiratory responses to behavioral arousal

As we observed for apneic breathing in resting animals, acute treatment with LM22A-4 completely reverses the abnormal respiratory response to auditory arousal in Mecp2 mutants. It is possible that both of these treatment effects result from reduced excitability within the respiratory network in mutants. However, it is also possible that LM22A-4 acts at mid- and/or forebrain sites that are important for behavioral state-dependent modulation of breathing, including the periaqueductal gray, amygdala and limbic cortices (Keay et al., 1988; Brandão et al., 1993; Nalivaiko et al., 2011).

In light of the fact that breathing dysfunction in RTT patients improves with relaxation or sleep (Weese-Mayer et al., 2008; Katz et al., 2009; Ren et al., 2012; Ramirez et al., 2013), we considered the possibility that, rather than, or in addition to, any direct effects on respiratory control per se, LM22A-4 might also improve breathing by inducing sedation. Indeed, other studies have shown that apneic breathing in RTT mouse models is reduced by drugs with sedating or anxiolytic effects, including the GABA uptake inhibitor NO-711 (Abdala et al., 2010), the GABAA partial agonist L338,417 (Abdala et al., 2010), the 5-HT1A antagonist 8-OH-DPAT (Abdala et al., 2010), the benzodiazepines midazolam (Voiuron and Hilaire, 2011) and diazepam (Abdala et al., 2010), the norepinephrine uptake inhibitor desipramine (Roux et al., 2007; Zanella et al., 2008), and the corticotropin-releasing hormone receptor 1 antagonist antalarmin (Ren et al., 2012). However, in the present study, open-field testing revealed no detectable differences in behaviors that are sensitive to sedation, including velocity of movement and total distance traveled, between saline- and drug-treated Hets. Similarly, drug-treated Hets showed no increase in the total duration of quiet breathing periods compared with their saline-treated counterparts. On the basis of these data, we conclude that sedation cannot explain the reversal of apneic breathing or normalization of the respiratory response to sensory arousal following treatment of Mecp2 mutants with LM22A-4.

Other strategies for overcoming functional deficits associated with reduced BDNF-TrkB signaling in RTT include enhancing the levels of endogenous BDNF with ampakines (Ogier et al., 2007), which stimulate activity-dependent BDNF expression by enhancing AMPA receptor function (Lynch et al., 2008) or fingolimod (Deogracias et al., 2012), a sphingosine-1 phosphate receptor modulator. One potential limitation of these other approaches is that BDNF activates receptors other than TrkB, including p75. The properties of binding to p75 and/or functioning as a full TrkB ligand might play an important role in unwanted pleiotropic effects of elevated BDNF levels (Longo and Massa, 2013). LM22A-4, on the other hand, does not bind p75 and, in addition, does not fully compete for BDNF activation of TrkB (Massa et al., 2010). Thus, although LM22A-4 increases TrkB phosphorylation and activates downstream signaling pathways in multiple, but not all, models in vivo and in vitro (Kajjya et al., 2014; Massa et al., 2010, Han et al., 2012; Schmid et al., 2012).
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22-week-old Het mice were used for experiments. All experimental procedures used for analysis. Quiet breathing was defined as periods when the animal breathed at a rate of 50 breaths/minute. Only episodes of quiet breathing totaling at least 10 minutes were included for analysis. The animal recording chamber ensured continuous inflow of fresh air (1 L/minute). Following the initial period of quiet breathing, an episode of quiet breathing was considered to have ended when the animal began moving or when the breathing rate increased above 50 breaths/minute. The time course of LM22A-4 effects on breathing was determined by recording breathing traces during and after exposure to a single 50-ms broad-spectrum auditory pulse (80 dB white noise). The acoustic stimulus was presented through two speakers (Advent) placed outside the plethysmographic chamber and the intensity level was calibrated using a sound meter (AML-925A-PC; Med Associates, VT) placed inside the chamber. Animals were injected with saline or LM22A-4 as described above and reflex testing was performed once the animals exhibited at least 2 minutes of quiet breathing (approximately 1 hour after being placed in the plethysmographic chamber).

Orienting reflex testing

Plissotomographic recordings were obtained as described above, but during and after exposure to a single 50-ms broad-spectrum auditory pulse (80 dB white noise). The acoustic stimulus was presented through two speakers (Advent) placed outside the plethysmographic chamber and the intensity level was calibrated using a sound meter (AML-925A-PC; Med Associates, VT) placed inside the chamber. Animals were injected with saline or LM22A-4 as described above and reflex testing was performed once the animals exhibited at least 2 minutes of quiet breathing (approximately 1 hour after being placed in the plethysmographic chamber).

Acoustic startle testing

The acoustic startle response was measured using a San Diego Instruments startle response recording system (San Diego, CA). Mice were placed inside a small-sized, non-restrictive, cylindrical Plexiglas recording chamber [3.5” (L) x 1.1” (ID)] on a accelerometer platform. Following a 5-minute acclimation period inside the recording chamber with a constant 70 dB background white noise, a 40-ms white-noise stimuli of different intensities (70, 74, 78, 82, 86, 90, 100, 110 and 120 dB) were delivered four times each in a random order with random intervals. The maximum accelerometer response amplitude measured within 40 ms after the acoustic stimulus was defined as the acoustic startle response (Vmax). Vmax measurements obtained from each of the four repeated trials of the same stimulus intensity were averaged.

Open-field testing

Locomotor activity was measured while the mice were in an open field consisting of a 40×40 cm box located in a dimly lit room. Using EthoVision XT 5.0 software, the field was digitally subdivided into a 20×20 cm center area and a periphery. The periphery was further divided into middle (inner 10 cm) and outer (outer 10 cm) areas. 1 hour after injection with LM22A-4 or saline, animals were placed in the open field and allowed to explore the enclosure freely for 15 minutes. During this period we measured total distance moved, velocity, angular velocity, rearing and heading to determine basic locomotor activity, as well as frequency and duration in the center, periphery and outer areas to evaluate thigmotaxis. Additionally, data were collected into three 5-minute bins and the distance moved during each of these three periods was recorded to evaluate habituation differences across groups. Values for the middle and outer sections were added together to calculate the center-periphery ratio.

Electrophysiology

Slice preparation

To study acute effects of LM22A-4 on synaptic transmission in the nTS, horizontal brainstem slices were prepared from 5- to 7-week-old Null and Het male mice as previously described (Chen et al., 2001; Kline et al., 2002, Kline et al., 2010). In brief, mice were deeply anesthetized by inhalation of isoflurane and then decapitated. Brains were removed from the skull and placed for 2-5 minutes in ice-cold, low Ca+2 artificial cerebrospinal fluid (ACSF) containing (mM): NaCl, 125; KCl, 3; NaH2PO4, 1.2; CaCl2, 1; MgSO4, 1.2; MgCl2, 2; NaHCO3, 25; D-glucose, 10 and L-aspartic acid 0.4, equilibrated to pH 7.4 with 95% O2 / 5% CO2. Brainstem were
dissected, glued on the mounting platform of a vibratome (Leica, VT 1000S), and horizontal sections containing the nTS, including a long segment of the TS, were cut at 220-250 μm. Slices were then transferred to recording ACSF (mM: NaCl, 125; KCl, 3; NaH2PO4, 1.2; CaCl2, 1.2; NaHCO3, 25; D-gucose, 10 and L-aspartic acid, 0.4, equilibrated to pH 7.4 with 95% O2/5% CO2) at ~32°C and allowed to recover from the procedure for at least 30 minutes before recordings.

Recordings
Slices were placed into the recording chamber, held in place with a nylon-wired grid and superfused with recording ACSF at 30-32°C at a flow rate of 4-5 ml/min. For stimulation of presynaptic inputs to nTS neurons, a concentric bipolar stimulation electrode (Frederic Haer) was placed on the TS, the medullary tract containing the central axons of primary afferent inputs to the nTS, its rostral to recording sites. Patch pipettes were pulled from thick-walled borosilicate glass capillaries and filled with intracellular solution (mM: K+glucosolate, 130; NaCl, 10, EGTA, 11, CaCl2, 1, HEPES, 10, MgCl2, 1, MgATP 2; NaGTP 0.2), and had resistances between 4 and 9 MΩ. Recordings were made lateral to the TS at the level of, and caudal to, the obex, which includes the interstitial, lateral and ventrolateral subnuclei [referred to as lateral nTS (lnTS)], the central targets of pulmonary stretch receptors (Kubin et al., 2006). Neurons were visualized with an upright Olympus microscope (BX51W1IF). To mimic afferent sensory input, the TS was stimulated at 0.5 Hz. Only neurons receiving monosynaptic input on 0.5 Hz TS stimulation (20 sweeps), defined as a low jitter of latency of the TS was stimulated at 0.5 Hz. Only neurons receiving monosynaptic input (<250 μs) were considered. From these neurons, eEPSCs, sPSCs and mEPSCs were recorded in the whole-cell current-clamp configuration at a holding potential of ~60 mV. To obtain lengthy eEPSC data during drug application, the TS was stimulated continuously at 0.5 Hz in these experiments. Neurons with a resting membrane potential more positive than ~40 mV upon breakthrough were rejected. Series resistance (Ra) was compensated (80%), and only neurons with RaR/Th were used. Signals were amplified (Axopatch 200B, Axon Instruments), filtered at 2 kHz and digitized at 10 kHz.

Control and drug recordings were performed in n=47 mice (Mecp2Δ) and n=30 mice (WT). For control measurements, one to four cells were recorded per slice. Because of potential long-lasting and non-reversible drug effects (M.S. and D.M.K.), analyzed the data; and M.K., M.L., F.L. and D.M.K. were responsible for the study design and data interpretation, and wrote the manuscript.

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Supplementary material
Supplementary material available online at http://dmm.biologists.org/lookup/suppl/dmm.016030/-DC1

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