Serological testing for SARS-CoV-2 in Bosnia and Herzegovina: first report

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Abstract

Introduction: Serological detection of SARS-CoV-2-specific immunoglobulins G (IgG) and M (IgM) antibodies is becoming increasingly important in the management of the COVID-19 pandemic.

Methods: We report the first results of COVID-19 serological testing in Bosnia and Herzegovina on 2841 samples collected and analysed in 2 medical institutions in Sarajevo. Antibody detection was performed using commercially available kits.

Results: In the first cohort, 43 IgM-positive/IgG-negative and 16 IgM-positive/IgG-positive individuals were detected, corresponding to 3.41% of participants having developed antibodies. In the second cohort, 4.28% participants were found to be IgM-negative/IgG-positive.

Conclusions: Our results suggest the need for population-wide serological surveying in Bosnia and Herzegovina.

Key words: COVID-19, immunoglobulins G, immunoglobulins M, molecular testing, serological testing.

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is causing a novel COVID-19 infection [1, 2]. While the standard diagnostic test is molecular analysis using qRT-PCR, serological testing for COVID-19 is becoming increasingly important in research community and immunity surveillance efforts. Serological testing is performed on whole blood, serum, or plasma samples in a relatively simple and rapid procedure, requiring less expertise and simpler laboratory settings compared to molecular methods. Tests are designed to detect either total immunoglobulins (Ig) or to differentiate between immunoglobulins G (IgG) and immunoglobulins M (IgM) fractions [3, 4]. Previous studies have reported sensitivity and specificity of 77.3% and 100% for IgM, respectively, and 83.3% and...
95% for IgG, respectively [5]. Serological assays directed towards SARS-CoV-2 are based on detection of antibodies against a 2-subunit viral spike (S) protein, with S1 being responsible for receptor binding and S2 for fusion [6, 7].

The aim of this preliminary study is to offer the first report on serological testing for SARS-CoV-2 in Bosnia and Herzegovina (B&H).

Methods. Sample collection and analysis in the first cohort. This study was conducted in accordance with the Helsinki Declaration for research involving human subjects. All patient samples were collected during routine laboratory operations and anonymized prior to manuscript preparation.

A total of 1720 whole blood and/or pharyngeal swab samples were collected and tested for IgM/IgG antibodies, as well as for viral RNA at the Dr. Abdulah Nakaš General Hospital in Sarajevo, B&H in April and May 2020. All samples were collected as a part of the hospital triage procedure. Blood samples were collected in a tube containing an anticoagulant and were tested immediately. Additionally, 131 nasopharyngeal and oropharyngeal swab samples were taken from a subset of the study population for qRT-PCR testing, according to patients’ clinical profiles and based on the recommendation from the attending medical doctor. For serological testing, the Artron One-Step Novel Coronavirus (COVID-19) IgM/IgG Antibody Test Kit (Artron Laboratories, Burnaby, BC, Canada) was used. For molecular testing, RNA was extracted using QIAamp Viral RNA Mini Kit (Qiagen, Hilden, Germany), followed by RNA quantification using a Qubit fluorometer (Thermo Fisher Scientific, Waltham, MA, USA). Two thermal cyclers were used for viral RNA amplification, namely QuantStudio™ 7 Flex Real-Time PCR System and QuantStudio 5 Dx Real-Time PCR (qPCR) System (Thermo Fisher Scientific, Waltham, MA, USA). Diagnovital® 2019-nCoV Real-Time PCR Kit (RTA Laboratories, Gebze/Kocaeli, Turkey) was used in amplification reactions. Internal control was HEC (human extraction control), which was amplified with every sample. In addition, each run had manufacturer-provided positive and negative controls, as well as an extraction negative control, which was treated as a sample.

Sample collection and analysis in the second cohort. A total of 1121 individuals were tested for COVID-19-specific IgG and IgM antibodies at the Eurofarm Centre Laboratory in Sarajevo between 21 April 2020 and 17 July 2020. Venous blood samples were collected in a plain tube without anticoagulant. All collected serum samples were immediately processed with the Dynamiker Biotechnology Co. rapid test (Tianjin, China) to detect SARS-CoV-2 IgG/IgM antibodies. The test was internally validated in the laboratory on 5 PCR-positive patients (day 21 after detection) and 5 healthy PCR-negative volunteers.

Statistical analysis. All analyses were performed using Excel 2010 (Microsoft Corporation, Redmond, USA). The exact test of goodness-of-fit was used to determine the effect of age on the frequency of COVID-19-positive individuals, in addition to comparing the frequency of individuals who were most probably still actively infected (IgM-positive, IgG-negative) and individuals at a later stage of the infection (IgM-positive, IgG-positive). Furthermore, we determined the effect of a reported underlying condition and sex on the frequency of IgG-positive cases. The 2-sample t-test was used to compare the mean age among female and male individuals.

Results. A total of 59 (3.41%) out of 1720 individuals were confirmed to be positive for COVID-19 antibodies in the first cohort. The mean age of individuals who tested positive was 61.46 years. The number of individuals who were older than 50 years (86.44%) was significantly higher than the number of individuals who were younger than 50 years (13.56%, \( p = 9.05 \times 10^{-9} \)). In addition, the mean age of female participants (66.57 years), who comprised 50% of the individuals who tested positive for COVID-19, was found to be significantly higher than the mean age of male participants (56.37, \( p = 8.5 \times 10^{-10} \)). None of the reported symptoms were shown to have been in correlation with either sex or age of the participants (data not shown) (Table I).

Out of 59 antibody-positive individuals, 43 (72.88%) tested positive for IgM but negative for IgG, indicating that these individuals were most probably still actively infected [5, 8]. The remaining 16 (27.11%) individuals tested positive for both antibodies, indicating either a later stage of active infection or convalescence [5, 8] (Table II). The number of individuals that were IgM-positive, IgG-negative was significantly higher

| Table I. Clinical and demographic characteristics of serologically positive patients from Sarajevo, B&H |
|-------------------------------------------------|
| Total serologically positive 107 |
| Female 50 |
| Male 54* |
| Sore throat 0 |
| Cough 4 |
| Fever 5 |
| Muscle pain 1 |
| Short breath 4 |
| Traveling 2 |
| Contact with risk group 52 |

**There were cases where sex was unknown.**
The prevalence of asymptomatic infections. Our early results indicate that 3.41% of study participants from Dr. Abdulah Nakaš General Hospital of Sarajevo and 4.28% of individuals tested in the Eurofarm Centre Laboratory in Sarajevo had developed SARS-CoV-2-specific antibodies. Additional molecular detection of viral RNA in the first set of samples showed that it was detected in one IgM-positive/IgG-negative sample and was not detected in 11 samples of IgM-positive/IgG-negative and 11 samples that were IgM-positive/IgG-positive. On the other hand, 5 out of 48 (10.4%) IgM-negative/IgG-positive participants from the Eurofarm Centre Laboratory set were previously confirmed to be PCR-positive through the governmental testing procedure, implying that serological testing correctly showed previous infection with SARS-CoV-2. These results further emphasize different roles of serological and molecular assays for SARS-CoV-2 antibodies and RNA, respectively, as well as the importance of time of sampling with respect to the infection start time.

The IgM/IgG antibodies for SARS-CoV-2 were found to develop 6–15 days post disease onset [9–13]. The presence of antibodies was detected in < 40% patients within 1 week of infection onset, and rapidly increased to 100% (total antibodies), 94.3% (IgM), and 79.8% (IgG) from day 15 after onset [11]. Previous studies suggested that the fraction of IgM-positive patients was decreasing as the time passing since infection confirmation [14]. Other reports found that an antibody response is displayed between days 10 and 21 after the infection onset, with evidence that the detection in some cases can take 28 days or more, while in a small number of cases IgM and/or IgG antibodies were not detected at all during the study. It is also necessary to note that the virus-specific IgG is significantly lower in asymptomatic than in symptomatic individuals in the acute phase [9]. This issue is even more pronounced considering that 40–45% of COVID-19 cases appear asymptomatic [15].

Table II. An overview of serological testing results from 2 laboratories in Sarajevo, B&H

| Variable                  | Dr. Abdulah Nakaš General Hospital | Eurofarm Centre Laboratory |
|---------------------------|------------------------------------|----------------------------|
| Total tested              | 1720                               | 1121                       |
| IgG+/IgM+                 | 16                                 | 0                          |
| IgG+/IgM–                 | 43                                 | 0                          |
| IgG+/IgM–                 | 0                                  | 48                         |
| IgG–/IgM+                 | 1661                               | 1073                       |

Table III. Comparison of serological vs. molecular testing of 131 selected samples from Dr. Abdulah Nakaš General Hospital of Sarajevo

| Variable                  | PCR positive | PCR negative |
|---------------------------|--------------|--------------|
| IgM+/IgG–                 | 2            | 106          |
| IgM+/-IgG+                | 1            | 11           |
| IgM+/-IgG+                | 0            | 11           |
| IgM–/IgG+                 | 0            | 0            |
stage of the COVID-19 pandemic. For this reason, we were unable to perform routine and repeated molecular and serological test monitoring of all participants, but the selection of patients for both types of analyses was made upon the decision of the attending medical doctor.

In conclusion, this study represents the very first results of a large-scale serological testing effort in B&H conducted on 2841 samples in 2 laboratories in Sarajevo during early stage (spring 2020) of the COVID-19 pandemic in B&H. The obtained results suggested a relatively small percentage of COVID-19 serologically positive individuals within the Bosnian-Herzegovinian population. We obtained 3.41% of IgM-positive individuals in the first, as well as 4.28% IgG-positive/IgM-negative in the second, Sarajevo study group. Serological testing is expected to play an important role in future when it comes to analysis of the extent of COVID-19 epidemics in B&H, as well as to follow the dynamics of infection and frequency of seropositive individuals. For such efforts, future optimization of existing procedures and sampling methodology will be required.

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Conflict of interest

The authors declare no conflict of interest.

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