Influence of Different Vegetable Oils on In Vitro Ruminal Fermentability and Nutrient Digestibility in Ettawah Crossbred Goat

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Abstract. This research was aimed to determine the effect of supplementation of vegetable oil (corn oil, palm oil and kapok seed oil) on in vitro ruminal fermentability and nutrient digestibility. Experiment design used was Randomized Block Design with four treatments and four replicates according to rumen fluid sampling time. The four treatments were R0 = feed without oil supplementation; R1 = R0 + 5% corn oil; R2 = R0 + 5% palm oil; and R3 = R0 + 5% kapok seed oil. The result showed that supplementation of 5% vegetable oil did not affect the rumen pH, A/P ratio, and the efficiency of energy conversion. Compared to control, the 5% supplement increased total VFA, acetate, propionate, butyrate, methane and NH3. However, the 5% supplement resulted in a lower protozoa population, microbial protein, in vitro dry matter digestibility (IVDMD), organic matter digestibility (IVOMD), and crude fiber digestibility (IVCFD) than those of control. Population of protozoa decreased until 58.76% for R1; 66.89% for R2; and 43.33% for R3. It can be concluded that 5% vegetable oil supplementation decreased protozoa population but increased the production of VFA and NH3. 5% kapok seed oil produced the highest of total VFA, acetate, propionate, butyrate and NH3 among other treatments.

Keywords: ruminal fermentability, vegetable oil, nutrient digestibility, in vitro

Introduction

Dairy cattle, especially during lactation, need high energy feed because they need two to three-fold nutrient supplies for body maintenance, milk synthesis, and post-partum tissue recovery (Khotijah et al., 2017). Fat is an important energy source for dairy cattle feed because it contains high calorie; fat energy is twice as much of carbohydrate or protein, and it produces lower heat increment (Wina and Susana, 2013; Diapari et al., 2017). Another benefit is for rumen defaunating agent. Fat can decrease protozoa population so that bacterial activity could increase the production of volatile fatty acid (VFA). Wibowo et al (2012) stated that up to 5% fat incorporated in ruminal diet did not interfere ruminal metabolism. However, high fatty feed would render biohydrogenation process in the rumen.

Vegetable oils from different source have different fatty acids composition (Titi and Fataftah, 2013) and is expected to produce different result in ruminal responses. Some vegetable oils for supplement in goat ration are

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corn oil, palm oil and kapok seed oil, with different fatty acids composition and market availability. Corn oil are easy to get and contains plenty unsaturated long chain fatty acids. The major fatty acids in corn oil is linoleic acids (C18:2) up to 53.99% (Giron et al., 2016). Palm oil is cheap and highly available and composed mainly of palmitic acid (C16) 32-37% and oleic acid (C18:1) 46.14% (Sitoresmi et al., 2009). Kapok seed oil contains high unsaturated long chain fatty acids such as palmitic acid (C16) 23.62%, oleic acid (C18:1) 24.30%, and linoleic acid (C18:2) 43.68% (Widiyanto et al., 2016).

According to Giron et al. (2016), supplementing 4% corn oil in the dairy cows increased milk production and milk fat concentration, decreased the VFA molar production without interfere the molar ratio of acetate, propionate ad butyrate. Supplementation of 5% palm oil increased the growth and efficiency of feed conversion in weaning lamb (Dutta et al., 2008). Meanwhile, kapok seed oil (up to 5%) supplemented to sheep did not affect the cellulolytic enzyme activity in the rumen (Widiyanto et al., 2010).

Accordingly, it is important to investigate the effect of different vegetable oils on ruminal fermentability, nutrient digestibility and energy efficiency based on the fatty acids. This in vitro study aimed to examine the potency of vegetable oils (corn oil, palm oil and kapok seed oil) to provide energy for lactating goat diet to optimize the rumen fermentability and nutrient digestibility.

Material and Methods

Feed preparation

Feed was made of forage and concentrates in 40:60 ratio. Forage consisted of corn straw and Calliandra calothyrsus, while the concentrate was made of rice bran, pollard, cassava waste meal, soybean meal, and molasses. The rations were formulated with 65-69% Total Digestible Nutrient (TDN) and 14% Crude Protein (CP). The ration treatments were R0 = control (ration without supplemented oil), R1 = R0 + 5% corn oil, R2 = R0 + 5% palm oil, and R3 = R0 + 5% kapok seed oil. Feed composition and nutrients contents of rations are presented in Table 1.

The evaluated parameters were rumen pH, protozoa population, microbial protein, ammonia concentration, VFA, acetate/propionate ration, methane energy, efficiency of energy conversion, in vitro dry matter digestibility (IVDMD), organic matter digestibility (IVOMD), and crude fiber digestibility (IVCFD).

Experimental procedures

In vitro rumen fermentation

The samples of each experimental feed were incubated in vitro with buffered rumen fluid mixture following the procedure by Tilley and Terry (1963). Rumen fluid was collected before morning feeding from a rumen fistulated Ettawah Crossbred goat fed on a diet of forage and concentrate mixture following the ration for in vitro substrate. The experiment was conducted in the Faculty of Animal and Agricultural Sciences, Diponegoro University, Semarang. Before use, rumen fluid was filtered using cheese cloth and placed in insulated flasks under anaerobic conditions. About 500 mg feed was inserted into a fermentation tube, added with 40 ml McDougall buffer solution, 10 ml rumen fluid and shaken with CO2 for 30 seconds. The tube was sealed with a rubber cap and placed in a waterbath at 39°C for 4 hour fermentation. Afterwards, the rubber cap was opened, and the tube was placed in iced water. The fermentation fluid was drawn to analyze protozoa population, and the supernatant was sampled to analyze rumen pH, microbial protein, and ammonia and VFA concentration.
Table 1. Composition and Nutrient Contents of Rations

| Feed Stuffs (% DM)          | Composition       |
|----------------------------|-------------------|
|                            | R0    | R1    | R2    | R3    |
| Corn straw                 | 30    | 28    | 28    | 28    |
| Calliandra calothyrsus     | 10    | 10    | 10    | 10    |
| Rice bran                  | 17    | 16    | 16    | 16    |
| Pollard                    | 19    | 18    | 18    | 18    |
| Cassava waste meal         | 13    | 13    | 13    | 13    |
| Soybean meal               | 8     | 7     | 7     | 7     |
| Molasses                   | 3     | 3     | 3     | 3     |
| Corn oil                   | 0     | 5     | 0     | 0     |
| Palm oil                   | 0     | 0     | 5     | 0     |
| Kapok seed oil             | 0     | 0     | 0     | 5     |
| **Total**                  | **100**| **100**| **100**| **100**|

| Nutrient contents (%)      |       |       |       |       |
|----------------------------|-------|-------|-------|-------|
| Ash                        | 10.28 | 10.03 | 10.03 | 10.03 |
| Crude protein (CP)         | 15.15 | 14.69 | 14.69 | 14.69 |
| Crude fiber (CF)           | 25.05 | 24.56 | 24.56 | 24.56 |
| Ether extract (EE)         | 2.67  | 7.60  | 7.60  | 7.60  |
| Nitrogen free extract (NFE)| 46.77 | 45.55 | 45.55 | 45.55 |
| **Total Digestible Nutrient (TDN)** | 60.41 | 68.93 | 68.93 | 68.93 |

R0: controlled ration, R1: R0 + 5% corn oil, R2: R0 + 5% palm oil, R3: R0 + 5% kapok seed oil.

Protozoa population was count using a counting chamber according to Ogimoto and Imai (1981). A total of 1 ml methyl blue formal saline solution (MFS = formaldehyde 37%, methyl blue, NaCl and aquadest) was added to 1 ml of rumen fluid. The mixture was directly enumerated using a counting chamber under the microscope with 40-time magnification.

The fermentation fluid was centrifuged at 3000 rpm for 15 minutes, and the supernatant was used to measure rumen pH using pH meter (ATC), analyses of microbial protein and ammonia concentration using a spectrophotometer, and individual VFA concentration using a Gas Chromatography. Methane gas emission and energy efficiency were calculated using a method by Orskov and Ryle (1990).

**Nutrient digestibility determination**

In vitro dry matter, organic matter and crude fiber digestibility were identified by inserting 500 mg feed to the fermentation tube and then added with 40 ml McDougall buffer solution, 10 ml rumen fluid, and shaken with CO2 for 30 seconds. The mixture was subsequently incubated for 48 h, then centrifuged at 3000 rpm / 15 min and added with 50 ml pepsin HCl solution. The mixture was further incubated for 48 h, and filtered using a vacuum pump and Whatman no. 41 filter paper. The samples were used to determine IVDMD, IVOMD and IVCFD based on Tilley and Terry (1963).

**Data analysis**

The collected data were subject to analysis of variance (ANOVA) in a randomized block design. Any significant difference across treatments was further tested by Duncan test (Steel and Torrie, 1994).

**Results and Discussion**

**Ruminal Fermentability**

**pH**

The data on in vitro rumen fermentability are presented in Table 2. Supplementing 5% vegetable oil did not affect rumen pH (P>0.05). It showed that the treatments did not interfere the rumen fermentation process. According to Adeyemi et al. (2015), rumen pH is not affected by the increasing amount of 0%, 4% and 8% of mixed 20% palm oil and 80% canola oil. Supplementation up to 4% of mustard, groundnut, sunflower, sesame, soybean and
rice bran oil with substrate concentrate and hay in 60:40 ratio did not affect rumen pH (Roy et al., 2017). Khotijah et al. (2017) stated that 6% supplementation of sunflower oil in a substrate containing 70:30 ratio of concentrate and Brachiaria humidicola did not affect the change in rumen pH. Measurement of pH is an important factor for measuring the rumen health due to the sensitivity of cellulose bacterial on pH changes. Ruminal pH value ranged from 6.86 to 6.99 which was still optimum for microbial growth. The optimal value of pH is in range 6.4 - 6.8 for supporting cellulolytic bacterial activity to digest the fiber content of feed (Khaing et al., 2016; Arief et al., 2016).

**Protozoa Population**

Protozoa population was significantly affected (P<0.05) by 5% vegetable oils supplementation. The mean total protozoa decreased after the addition of corn oil, palm oil and kapok seed oil compared to control. Vegetable oil contains unsaturated long chain fatty acid that could inhibit protozoa growth. The fatty acid coated the protozoa’s cell membrane which subsequently interfered lipolysis activity of protozoa and kill the protozoa (Tamminga and Doreau, 1991). Some studies also documented the depression in protozoa population by the addition of vegetable oil. According to Sitoresmi et al. (2009), total protozoa ranged from 23.82 to 30.16 x 10³/ml with the addition of coconut, palm and sunflower oil. Protozoa population also decreased by the addition of 6% sunflower oil (Khotijah et al., 2017). Supplementation of vegetable oil into the ration could decrease total protozoa and increase bacterial activity to produce VFA as an energy supply for ruminant (Dijkstra et al., 2000). Supplementation of palm oil (39.8% palmitic acid and 46.14% oleic acid) was the most significant in decreasing total protozoa; however, the result was not significantly different from corn oil (containing 11.8% palmitic acid, 10.66% oleic acid and 53.99% linoleic acid) and kapok seed oil (containing 23.63% palmitic acid, 24.30% oleic acid and 43.68% linoleic acid). It showed that both saturated and unsaturated long chain fatty acids in the vegetable oil had the same effect in decreasing the protozoa numbers.

**Volatile Fatty Acid (VFA)**

The production of acetate, propionate, butyrate and total VFA in the rumen fluid was significantly affected (P<0.05) by the supplementation of vegetable oil. The highest production of acetate was in the supplementation of 5% kapok seed oil, followed by palm oil and corn oil; these results were significantly higher (P<0.05) than those of the control. Similarly, Morsy et al. (2015) reporting a higher acetic acid when lactating goat ration was supplemented with sunflower oil and sunflower seed compared to control feed. The higher acetate may due to cellulolytic bacterial activity. Hungate (1966) stated that acetate production from ruminal fermentation of fibrous feed was affected by cellulolytic bacterial such as Ruminococcus, Butyrivibrio, and Bactroides. The highest production of propionate, butyrate and

| Fatty Acid (%) | Corn Oil* | Palm Oil** | Kapok Seed Oil*** |
|---------------|-----------|------------|-------------------|
| Lauric acid (C12:0) | - | 0.06 | - |
| Palmitic acid (C16:0) | 11.80 | 39.8 | 23.62 |
| Stearic acid (C18 :0) | 5.18 | 4.4 | 2.38 |
| Oleic acid (C18 :1) | 20.66 | 46.14 | 24.30 |
| Linoleic acid (C18 :2) | 53.99 | 0.72 | 43.68 |
| Linolenic acid (C18 :3) | 1.01 | - | 2.92 |

Source: Giron et al. (2016)*, Sitoresmi et al. (2009) and Koushki et al. (2015)**, Widiyanto et al. (2016)***
Table 3. Effect of vegetable oil supplementation on ruminal fermentability

| Parameters                          | Control    | Corn Oil   | Palm Oil   | Kapok S. Oil |
|------------------------------------|------------|------------|------------|--------------|
| pH                                 | 6.86 ± 0.21| 6.94 ± 0.10| 6.95 ± 0.16| 6.99 ± 0.10  |
| Protozoa (x10^3/ml)                | 99.44 ± 32.38a| 40.83 ± 15.27ab| 32.78 ± 14.64b| 56.11 ± 18.63b|
| VFA (Mm)                           |            |            |            |              |
| Acetate                            | 46.88 ± 5.52b| 60.07±3.20c| 61.10±6.28a| 68.82±7.58a  |
| Propionate                         | 20.91 ± 1.80b| 22.82±1.55b| 23.41±2.85b| 27.44±3.72a  |
| Butyrate                           | 19.73 ± 3.92b| 19.91±2.84b| 17.28±2.33b| 29.37±3.86a  |
| A/P Ratio                          | 2.26 ± 0.32b| 2.64±0.23b| 2.63±0.34b| 2.52±0.25b   |
| Total VFA                          | 87.51±8.23b| 102.80±5.72a| 101.78±9.02b| 125.62±10.76a|
| Methane*                           | 28.08±4.45b| 34.28±2.18b| 33.33±3.24b| 42.23±4.42a  |
| Energy efficiency (%)              | 77.19 ± 1.01| 76.00±0.62| 75.97±0.93| 76.47±0.58   |
| NH3 (mg/dl)                        | 19.69 ± 0.39b| 22.35±1.91ab| 20.23±1.71b| 24.97±2.30a  |
| MicrobProt (mg/ml)                 | 12.01 ± 1.97a| 10.11±1.87ab| 9.39±0.86b| 8.84±0.75b   |
| IVDMD (%)                          | 61.06 ± 3.11a| 57.19±3.30b| 56.82±3.20b| 57.18±1.83b  |
| IVOMD (%)                          | 62.16 ± 3.31a| 56.34±2.88b| 55.89±2.35b| 55.81±1.14b  |
| IVCFD (%)                          | 74.46 ± 3.04a| 45.74±8.08c| 61.46±2.38b| 48.76±6.89c  |

Values with different superscripts in the same column indicate significant difference (P<0.05)

Total VFA was in supplementation of 5% kapok seed oil, while 5% corn and palm oil were not different from control (P>0.05). 5% kapok seed oil produced the highest total VFA because of the accumulation of a higher acetate, propionate and butyrate than the other treatments. The acids accumulated because the unsaturated fatty acid in kapok seed oil endured lipolysis and biohydrogenation process, then fermented into VFA. Jarvis et al. (2009) stated that supplying vegetable oil into the ruminant ration would trigger lipolysis and biohydrogenation process, producing long chain fatty acids, glycerol, and galactose which would be fermented into acetate, propionate and butyrate.

The result showed that acetate/propionate (A/P) ratio was not different (P>0.05) across treatments. The A/P ratio with the addition of vegetable oil ranged from 2.26 to 2.64. The lower the A/P ratio, the higher proportion of propionate. Accordingly, energy efficiency for metabolism also increased because the lipolysis process produced glycerol and transformed into propionate. Khaing et al. (2016) stated that propionate is the precursor of the gluconeogenesis process which could increase glucose synthesis as an energy source for ruminant. This result was lower than A/P ratio in supplementation of canola, sunflower and castor oil within 2.8 – 3.2 (Maia et al., 2012) or mixed canola and palm oil in a range of 2.71 – 3.61 (Adeyemi et al., 2015).

**Methane and Energy Efficiency**

The result showed that supplementation of 5% vegetable oil significantly affected (P<0.05) methane production. Methane produced from ration + 5% kapok seed oil was the highest compared to corn oil, palm oil and control treatments because kapok seed oil helped produced higher acetate and butyrate. Martin et al. (2008) stated that production of acetate and butyrate trigger the formation of hydrogen (H2) that would be utilized by methanogenic bacterial to produce methane, hence methane accumulation was higher.

The efficiency of energy conversion (EEC) was not affected (P>0.05) by the supplementation of vegetable oil. The EEC value, as well as the A/P ratio in this study, was not different across treatments. The lower A/P ratio reflected a higher efficiency of energy conversion from hexose to VFA. Rasmussen and
Harisson (2011) stated that the lower A/P ratio indicated a higher efficiency of energy.

**NH<sub>3</sub>**

The mean concentration of NH<sub>3</sub> was significantly affected (P<0.05) by the addition of vegetable oil. The highest concentration was obtained from the 5% kapok seed oil. The result of this study was higher (15.3 to 17.4 mg/dl) than that of addition of canola, sunflower and castor oil (Maia et al., 2012) and in addition of blended palm oil and canola oil on 0 – 8% which ranged from 11.03 – 12.5 mg/dl (Adeyemi et al., 2015). Leng (1990) reported that 10-20 mg/dl NH<sub>3</sub> was optimum to digest fibrous feed by rumen microbes. Furthermore, Mc Donald et al. (2002) stated that the optimum NH<sub>3</sub> concentration for microbial protein synthesis was 6 - 21 mMol or about 8.4 – 29.4 mg/dl. The average value of NH<sub>3</sub> in this study was 19.69 – 24.97 mg/dl, indicating a sufficient level for amino acids synthesis by rumen microbes.

**Microbial protein**

Microbial protein significantly decreased (P<0.05) by the vegetable oil supplementation. The highest and lowest microbial protein in this study was found in the supplementation of corn oil (10.11 mg/ml) and kapok seed oil (8.84 mg/ml), respectively, due to fatty acids contained in the vegetable oil. Long chain fatty acids tend to inhibit the rumen microbes’ growth, including bacteria and protoza, which could depress microbial protein synthesis. In general, long chain fatty acids are detrimental to rumen microbial activity, and worse to protoza (Dijkstra et al., 2000). The average microbial protein in this research was higher than 0.39-0.41 mg/ml produced by 7.5% supplementation of coconut oil, palm oil and sunflower seed oil supplementation (Sitoresmi et al., 2009) and 3.8-5.1 mg/ml from 3% soybean oil (H-L Mao et al., 2010). Supplementation of 5% corn oil, palm oil and kapok seed oil did not affect microbial protein and remained in normal range.

According to Orskov (1992), the precursors for microbial protein synthesis were carbon chain, energy and NH<sub>3</sub>. Carbon chain and energy could be obtained from VFA, while NH<sub>3</sub> is the source for N. The optimum microbial protein production was obtained from 80-160 mM VFA and 8.4 – 29.4 mg/dl NH<sub>3</sub> (Mc Donald et al., 2002). This study showed that supplementation of 5% vegetable oil increased VFA to 125.42 mM and NH<sub>3</sub> to 24.97 mg/dl; therefore, it was a viable precursor for microbial protein synthesis.

**In vitro digestibility of dry matter, organic matter and crude fiber**

The result of nutrient digestibility analysis is presented in Table 3. Vegetable oil supplementation significantly decreased (P<0.05) in vitro digestibility of dry matter (IVDMD) (56.82 to 61.06%) and organic matter (IVOMD) (48.62 to 56.00%). The declining digestibility may due to unsaturated fatty acids effect in the supplemented vegetable oil involving the cytotoxic effect of the unsaturated fatty acid with free carboxyl group. This compound could interfere the function of cell bio-membrane, such as the uptake of amino acids and protoplasmic energy metabolism. Another contributing factor was the depleting of the essential mineral availability for rumen microbe, because the fatty acid bond those elements (Widiyanto et al., 2007).

The result showed that IVCFD was significantly different (P<0.05) across treatments. Supplementation of corn oil and kapok seed oil resulted in a lower IVCFD than that of palm oil and control because corn oil and kapok seed oil had a higher unsaturated long chain fatty acid (LCFA) than palm oil, namely 74.65% vs. 67.98% vs. 46.86% (long chain UFA), respectively. Vegetable oil rich in unsaturated LCFA would decrease more crude fiber digestibility. Dijkstra et al. (2000) stated that fiber degradation in the rumen was reduced when the amount of unsaturated LCFA
in the diet was raised. This phenomenon could occur due to the inhibition of fibrolytic bacteria activity with the unsaturated LCFA. According to Messana et al. (2013), the unsaturated LCFA could inhibit fibrolytic bacteria activity, thereby reducing fiber digestibility.

Conclusions
Supplementation of 5% vegetable oil tend to decrease the nutrient digestibility without interfering the ruminal fermentability. Supplementation of 5% kapok seed oil resulted in the highest total VFA and NH3 compared to corn oil and palm oil.

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