Fully-automated radiosynthesis of the amyloid tracer $[^{11}\text{C}]$ PiB via direct $[^{11}\text{C}]$CO$_2$ fixation-reduction

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Abstract

**Background:** The β-amyloid radiotracer $[^{11}\text{C}]$ PiB is extensively used for the Positron Emission Tomography (PET) diagnosis of Alzheimer’s Disease and related dementias. For clinical use, $[^{11}\text{C}]$ PiB is produced using the $^{11}$C-methylation method ([$^{11}\text{C}]$ Methyl iodide or [$^{11}\text{C}]$ methyl triflate as $^{11}$C-methylation agents), which represents the most employed $^{11}$C-labelling strategy for the synthesis of $^{11}$C-radiopharmaceuticals. Recently, the use of direct $[^{11}\text{C}]$CO$_2$ fixation for the syntheses of $^{11}$C-tracers has gained interest in the radiochemical community due to its importance in terms of radiochemical versatility and for permitting the direct employment of the cyclotron-produced precursor $[^{11}\text{C}]$CO$_2$.

This paper presents an optimised alternative one-pot methodology of $[^{11}\text{C}]$CO$_2$ fixation-reduction for the rapid synthesis of $[^{11}\text{C}]$ PiB using an automated commercial platform and its quality control.

**Results:** $[^{11}\text{C}]$ PiB was obtained from a $(25.9 \pm 13.2)\%$ (Average ± Variation Coefficient, $n = 3$) (end of synthesis, decay corrected) radiochemical yield from trapped $[^{11}\text{C}]$CO$_2$ after 1 min of labelling time using PhSiH$_3$ / TBAF as the fixation-reduction system in Diglyme at 150 °C. The radiochemical purity was higher than 95% in all cases, and the molar activity was $(61.4 \pm 1.6)$ GBq/μmol. The radiochemical yield and activity (EOS) of formulated $[^{11}\text{C}]$ PiB from cyclotron-produced $[^{11}\text{C}]$CO$_2$ was $(14.8 \pm 12.1)\%$, decay corrected) and $9.88$ GBq ($\pm 6.0\%$), respectively. These are higher values compared to that of the $^{11}$C-methylation method with $[^{11}\text{C}]$CH$_3$OTf ($\sim 8.3\%$).

**Conclusions:** The viability of the system PhSiH$_3$ / TBAF to efficiently promote the radiosynthesis of $[^{11}\text{C}]$ PiB via direct $[^{11}\text{C}]$CO$_2$ fixation-reduction has been demonstrated. $[^{11}\text{C}]$ PiB was obtained through a fully automated radiosynthesis with a satisfactory yield, purity and molar activity. According to the results, the one-pot methodology employed could reliably yield sufficiently high tracer amounts for preclinical and clinical use.

**Keywords:** $[^{11}\text{C}]$ PiB, $[^{11}\text{C}]$CO$_2$ fixation-reduction, Automated radiosynthesis, PhSiH$_3$ / TBAF, Alzheimer’s disease, PET

Introduction

The compound 2-(4′-N-[$^{14}\text{C}$]methylaminophenyl)-6-hydroxybenzothiazole, also known as $[^{11}\text{C}]$6-OH-BTA-1 or $[^{11}\text{C}]$ Pittsburgh Compound B ($[^{11}\text{C}]$PiB), has long been recognised as a potent PET radiotracer for beta-amyloid (Aβ) plaque imaging in the brains of patients with Alzheimer’s Disease (AD) and other forms of dementia (Engler et al.
[11C] PiB still remains the gold standard for amyloid imaging in AD diagnosis due to its high affinity for Aβ plaques ($K_d = 1.4 \text{ nM}$) (Mathis et al. 2003), fast uptake and low non-specific binding.

The original radiosynthesis of [11C] PiB was performed by Mathis et al. (2003). It consisted of the $^{11}\text{C}$-$N$-methylation of the precursor 2-(4′-aminophenyl)-6-methoxy-methoxybenzothiazole (6-MOMO-BTA-0) with [11C] methyl iodide ([11C]CH$_3$I), followed by an acidic deprotection of the methoxymethyl group. The use of the more reactive $^{11}$C-methylating agent [11C] methyl trifluoromethanesulfonate ([11C]CH$_3$OTf) (Holschbach and Schüller 1993; Jewett 1992) over the unprotected precursor 2-(4′-N-aminophenyl)-6-hydroxybenzothiazole (6-OH-BTA-0) has permitted a direct and efficient $^{11}$C-$N$-methylation, as was demonstrated by Wilson and co-workers (Wilson et al. 2004) and Solbach and co-workers (Solbach et al. 2005) (Scheme 1).

During the last decade, additional optimised radiosyntheses of [11C] PiB have been published in the literature. (Philippe et al. 2011; Coliva et al. 2015; Boudjemeline et al. 2017) Although these radiosyntheses proved to be reliable and easily adaptable to a GMP compliant production for patients, all of them use the secondary precursors [11C] methyl iodide or triflate. The preparation of these secondary precursors from cyclotron-produced [11C]CO$_2$ is always time and activity consuming. (Långström et al. 1999). The overall RCY of any [11C] PiB radiosynthesis, and subsequently the total activity available for patients, is directly affected by these losses of radioactivity during the preparation of [11C]CH$_3$I or [11C]CH$_3$OTf. In this sense, any approach capable of eliminating the steps of synthesis of

![Scheme 1](image-url)
the $^{11}$C-methylating agents would be advantageous for a better overall performance in terms of the RCY and the available activity for PET scans.

$[^{11}C]CO_2$ is an attractive starting material for radiolabelling because it is produced directly from the cyclotron with a good yield and with a high $A_m$. The use of $[^{11}C]CO_2$ via the so-called ‘fixation’ to synthesise $^{11}$C-ureas, $^{11}$C-carbamates, $^{11}$C-oxazolidinones, $^{11}$C-carboxilic acids and $^{11}$C-amides is well-documented in the literature, and it is regarded as ‘a renaissance of PET radiochemistry’ (Rotstein et al., 2013, and references therein). The use of CO$_2$ as a C$_1$ building block for the methylation of amines has been recognised and demonstrated (Jacquet et al. 2012, 2013; Li et al. 2013; Das et al. 2014). These approaches have been translated to the $^{11}$C radiochemistry field by Liger and co-workers (Liger et al. 2015), which constitutes the first experience of using $[^{11}C]CO_2$ fixation-reduction for the catalytic methylation of amines. In particular, Liger and co-workers used the approach of Jacquet and co-workers (Jacquet et al. 2013), adapting it to an ‘one-pot’ procedure involving PhSiH$_3$, 1,3-bis (2,6-diisopropylphenyl)-1,3-dihydro-2H-imidazol-2-ylidene (IPr, a $N$-heterocyclic carbene) and ZnCl$_2$ in Diglyme at 150 °C. The authors were able to synthesise a series of aromatic and aliphatic amines with an RCY ranging from 24% to 60% (decay corrected, from trapped $[^{11}C]CO_2$) and also applied the methodology for $[^{11}C]$ PiB preparation. $[^{11}C]$ PiB was produced after 20 min of incubation at 150 °C (50 min overall time), yielding 2.1 GBq of formulated radiopharmaceuticals (RCY = 38% based on trapped $[^{11}C]CO_2$ and decay corrected, Scheme 2). Although this work demonstrated the considerable potential of the direct $[^{11}C]$ methylation of amines from $[^{11}C]CO_2$ fixation-reduction, some weaknesses in this methodology when applied to the radiosynthesis of $[^{11}C]$ PiB still remains, such as low molar activity ($A_m$, 15 GBq/μmol), long reaction times, carbene stability, metal-catalysed $N$-methylation and few or no description of quality control specifications for the so-prepared batches of radiopharmaceuticals.

With an ageing population, there is an increasing demand for $[^{11}C]$ PiB for clinical applications, and therefore the possibility of applying a methodology of $[^{11}C]CO_2$ fixation-reduction for the routine preparation of $[^{11}C]$ PiB to increase its radiochemical yield and thus perform more PET studies for each batch of radiopharmaceuticals was devised. In this context, attention was focused on the strategy recently developed by Liu and co-workers (Liu et al. 2016) for the reductive functionalisation of amines with $[^{11}C]CO_2$.
CO₂. In their work, the authors explored the utility of the formylation and methylation of amines with CO₂ using hydrosilanes / TBAF as the catalytic system.

This approach employed mild reaction conditions because it is a metal-free protocol, and the use of the unstable N-heterocyclic carbene was avoided. The radiochemical translation of this methodology was proposed for the radiosynthesis of amines based on radiopharmaceutical interest in particular [¹¹C] PiB (Scheme 2).

The aim of this work was to assess the applicability of the PhSiH₃ / TBAF mediated reductive incorporation of [¹¹C]CO₂ for the radiosynthesis of the β-amyloid tracer [¹¹C]PiB. A fully automated, GMP adaptable, fast radiosynthesis of [¹¹C] PiB by means of [¹¹C]CO₂ fixation-reduction was developed employing the commercially available platform GE TRACERlab® FX C Pro and its quality control.

Materials and methods
Chemicals and materials
All chemicals and reagents used in this work were commercially available products and were used without further purification. Anhydrous solvents MeCN (99.8%), DMF (99.8%), DMSO (99.9%) and bis (2-methoxyethyl) ether (Dyglime, 99.5%) were purchased from Sigma-Aldrich and stored and handled under inert atmosphere. Tetrabutylammonium fluoride (TBAF) 1.0 M in THF and PhSiH₃ (97%) were acquired from Aldrich, stored in a desiccator and handled in an inert atmosphere (N₂). 6-OH-BTA-0 was obtained from Siquimia. 4-Toluidine was purchased from Fluka. 6-OH-BTA-0, 6-OH-BTA-1 and 6-(MeO)-BTA-0 were used as analytical standards and were purchased from ABX (GmbH). N-methyl-4-toluidine, N-formyl-4-toluidine and 2-(4′-N-[¹¹C]formamidophenyl)-6-hydroxybenzothiazole (PiB N-formyl derivative) were synthesised using standard organic chemistry procedures described in the literature (Vogel 1956; Shekhar et al. 2009). Absolute Ethanol (99.8%) was purchased from Merck. Saline and Water for Injection (sterile, USP grade) were acquired from Farmaco Uruguayo. Sep-Pak® C₁₈ light cartridges were purchased from Waters and were preconditioned with ethanol (5 mL) followed by water for injection (10 mL) and air (5 mL). Millex® GV sterile filters (0.22 μm, PVDF, 33 mm) were purchased from Millipore. The semipreparative HPLC column used for [¹¹C] PiB purification was a Luna® 5 μm C₁₈(2) 100 Å 250 × 10 mm column (Phenomenex). The analytical HPLC column used for both model amine (4-toluidine) ¹¹C-methylation and [¹¹C] PiB radiosynthesis was a Nucleodur 100–5 C₁₈-ec 250 × 4.6 mm column (Macherey-Nagel). The analytical GC column was a DB-WAX that was 30-m in length, 0.53-mm in diameter and 1.00-mm in film thickness (Agilent).

Instruments
[¹¹C]CO₂ was produced by the ¹⁴N(p,α)¹¹C nuclear reaction in a PETtrace™ 800 16.5 MeV cyclotron (GE Healthcare). A high-performance target was used for [¹¹C]CO₂ production. The target content was a mixture of N₂ and 1.0% O₂ (Praxair). To assess the best labelling conditions, ~ 18.5 GBq of [¹¹C]CO₂ (50 μA, 3 min) were used, and ~ 185 GBq (70 μA, 35 min) were used for complete radiosynthesis (labelling, purification and formulation).

Radiosyntheses were carried out using a TRACERlab® FX C PRO module (GE Healthcare) (see Additional file 1: Figure S1). All valves of the TRACERlab® module were controlled according to the pre-programmed time intervals (time lists) to transfer the reagents from one
part to another part of the instruments. Helium pressure was used to transfer the reagents. The transfer of the radioactivity was traced and recorded with an inbuilt radioactivity detector. A by-pass of the iodination loop was made to redirect the purified [11C]CO₂ towards the reaction vessel (reactor). A pre-injection vial (10 mL) was installed before the injection loop, controlled by valve 10 and pressurised with helium from valve 19. The purpose of this vial was to collect the reactor content and the portions of rinse solvent (acetone) added from Vial 3 before the loading of the HPLC loop.

HPLC analyses were performed with a Shimadzu UFLC equipped with UV and a gamma detector (Lablogic Flow RAM HPLC detector). The GC analyses of ethanol, residual reagents and residual solvents were carried out using a Shimadzu GC-2010 Plus equipped with an FID detector. The gamma spectrometry was performed using a 1023-channel Ortec multichannel analyser with a 1” × 1” NaI (TI) crystal. The activity measurements were performed using a Capintec CRC 25 ionisation chamber.

**General procedure for the ¹¹C-methylation of amines with [¹¹C]CO₂ using PhSiH₃/TBAF**

**Preparation of the module**

Prior to any radiosynthetic procedure, the molecular sieves (MS, 4 Å, 60–80 mesh) column of the TRACERlab™ FX C Pro was heated to 350 °C under helium flow (40 mL/min) for 15 min and then cooled to room temperature under a helium atmosphere. Simultaneously, the reactor was rinsed with acetone, flushed with helium and further dried under a vacuum to 100 °C for 30 min. The reactor was then cooled to 25 °C and was kept in a positive helium atmosphere (> 300 kPa) until its use in the radiosynthetic experiment.

**Flushing of the target and lines**

To improve Aₘᵣ, a protocol described by our group (Savio et al. 2012) was followed. In short, the content of the target was delivered to the molecular sieves column at 350 °C under flowing helium (40 mL/min) to decrease the amount of unlabelled CO₂ and to send it to waste (‘cold flush’). Immediately before beginning the irradiation for the radiosynthesis, the target was bombarded at 70 μA for 5 min, and its content was directly sent to waste (‘hot flush’).

**Preparation of the reagent solution**

PhSiH₃ was added slowly over a solution of the amine (4-toluidine or 6-OH-BTA-0) in the desired solvent (0.5 mL) in an inert atmosphere. The solution was vigorously mixed using a vortex agitator, and then TBAF 1.0 M in THF was carefully added over the mixture. The evolution of gas and changes in colour were usually observed during this step. The resulting solution was mixed using a vortex agitator, taken by a syringe, loaded into the reactor of the TRACERlab™ module and sparged with helium (40 mL/min) for 5 min, ideally no more than 5 min before the end of bombardment (EOB).

**Optimisation of N-[[¹¹C-methyl]-4-toluidine**

The cyclotron produced [¹¹C]CO₂ (EOB Activity: A₀) was sent to the module and trapped in the MS column at room temperature for further purification. The delivery duration was approximately 3 min. The MS column was then heated to 350 °C to desorb purified [¹¹C]CO₂, which was transferred under a helium stream (15 mL/min) to
the reactor where the amine solution was placed. Trapping was performed at room temperature. The trapped $^{11}\text{C}\text{CO}_2$ activity was monitored and registered. Once trapping was complete (a maximum activity $A_T$ is reached), the reactor was sealed, and the solution was heated to the chosen temperature. For evaluating the losses of $^{11}\text{C}\text{CO}_2$ during the heating step, ‘start of labelling activity’ ($A_{\text{SOL}}$) was registered once the temperature reached the desired value. Likewise, ‘end of labelling activity’ ($A_{\text{EOL}}$) was registered once the labelling time was finished. The solution was cooled to approximately 70 °C and was diluted with 0.5 mL of the same solvent used in the radiolabelling step. The solution was collected in a vial, its activity ($A_{\text{VIAL}}$) was measured and radio HPLC analyses were performed to determine the relative radiochemical proportion of the expected species.

Automated radiosynthesis of $^{11}\text{C}\text{PiB}$

The same protocol described was conducted using 6-OH-BTA-0 as the precursor amine. Once the labelling step was finished, the solution was cooled and sent to the pre-injection vial. The reactor was then rinsed with acetone (1 mL) from Vial 3, combined with the reaction crude and injected into the HPLC. The separation of $^{11}\text{C}\text{PiB}$ was achieved using MeCN:H$_2$O (50:50 v/v) at a flow rate of 4 mL/min (t$_{R}$: 8.5–9.5 min). The fraction containing $^{11}\text{C}\text{PiB}$ was collected over 50 mL of water for injection and then passed through a Sep-Pak$^\text{®}$ C18 light cartridge. The excess HPLC solvent was washed with water for injection (10 mL). $^{11}\text{C}\text{PiB}$ was eluted from the SPE cartridge with 0.9 mL of absolute ethanol and collected over 5 mL of preloaded saline. In addition, 4 mL of saline were used to rinse the SPE cartridge. The solution of formulated $^{11}\text{C}\text{PiB}$ was filtered through a 0.22 μm sterilising filter. The total time of radiosynthesis was about 32 min (since EOB) or 25 min (since the end of $^{11}\text{C}\text{CO}_2$ trapping).

Physicochemical quality control

Radiochemical purity (RCP) was determined using analytical radio-HPLC. An isocratic condition with a CH$_3$COONH$_4$ / CH$_3$COOH buffer 0.1 M, pH = 5.0 and MeCN (40:60 v/v) at a flow rate of 1.2 mL/min was used for 4-toluidine. For $^{11}\text{C}\text{PiB}$, an isocratic condition with H$_2$O and MeCN (50:50 v/v) at a flow rate of 1.2 mL / min was used. RCP was calculated considering the area of the peak corresponding to the desired analyte in relation to the sum of the areas of all peaks. The identity of radioactive products was confirmed by co-elution with the non-radioactive standard compounds. UV detection was 270 nm for 4-toluidine and 340 nm for PiB. $A_m$ was calculated considering the activity of $^{11}\text{C}\text{PiB}$ EOS x RCP in relation to the molar amount of PiB in the sample.

Residual solvents and reagents (such as acetone, acetonitrile, Diglyme and PhSiH$_3$) and ethanol were analysed by gas chromatography (GC) in accordance with USP general chapter < 467>. The temperature programme for GC runs was a gradient of 40 °C hold for 2 min, 1 °C/min to 44 °C, 20 °C/min to 200 °C and 200 °C hold for 2 min (total time of 15 min) with helium (11.3 mL/min) as the carrier gas.

The appearance of the solution was checked by visual inspection. The pH level was determined using a calibrated pH-meter. Radionuclidic purity was assessed by recording the corresponding gamma spectrum, and radionuclidic identity was assessed by measuring the physical half-life.
Results and discussion

Optimisation of $N$-$[^{11}C$-methyl]-4-toluidine

To assess the potential of the $[^{11}C]CO_2$ reduction-fixation methodology with PhSiH$_3$ / TBAF, the research began by selecting the primary aromatic amine 4-toluidine (4-methylaniline) as a model substrate. The fixation-reduction reaction in different solvents, temperatures and varying amounts of amine substrates and TBAF catalysts was studied using an automated commercial platform TRACERlab™ FX C PRO. The aim was to maintain an excess of PhSiH$_3$ (0.6 mmol), which remained constant during the test runs, as well as the total volume of the solution (0.5 mL). Data collected for the radiosynthesis using the reactivity model are summarised in Table 1.

The reactivity of the system was evaluated through the radiochemical yield (RCY) of $N$-$[^{11}C$-methyl]-4-toluidine ($[^{11}C]_2$). The formation of $[^{11}C]_2$ was observed in practically all conditions where the capture of $[^{11}C]CO_2$ was considerable. Based on the work of Liu and co-workers (Liu et al. 2016), the experimental conditions began with using MeCN as a solvent. Thus, in MeCN at 80 °C, a radiochemical yield of 14% was obtained for $[^{11}C]_2$ (entry 1, Table 1). An increase in the amount of the amine 1 precursor led to an improvement in the radiochemical yields of $[^{11}C]_2$ (entries 2 and 3). Similar

Table 1 Direct $^{11}C$-methylation of 1 with $[^{11}C]CO_2$

| Entry | 1 [mmol] | Solvent | Temperature (°C) | Time (min) | $[^{11}C]CO_2$ trapping efficiency (% dc) | Radiochemical yield ($[^{11}C]_2$) (%) | Radiochemical yield $[^{11}C]_3$ (%) |
|-------|----------|---------|-----------------|------------|---------------------------------|--------------------------------|--------------------------------|
| 1     | 0.01     | MeCN    | 80              | 5          | 92                              | 12                              | 8                               |
| 2     | 0.05     | MeCN    | 80              | 5          | 84                              | 27                              | 3                               |
| 3     | 0.10     | MeCN    | 80              | 5          | 88                              | 35                              | 3                               |
| 4     | 0.05     | MeCN    | 100             | 5          | 48                              | 37                              | 0                               |
| 5     | 0.05     | MeCN    | 100             | 10         | 70                              | 27                              | 6                               |
| 6     | 0.05     | MeCN    | 100             | 5          | 60                              | 6                               | 3                               |
| 7     | 0.10     | MeCN    | 100             | 5          | 65                              | 42                              | 0                               |
| 8     | 0.05     | DMSO    | 50              | 5          | 89                              | 3                               | 14                              |
| 9     | 0.05     | DMSO    | 100             | 5          | 76                              | 48                              | 0                               |
| 10    | 0.05     | DMSO    | 120             | 5          | 81                              | 63                              | 0                               |
| 11    | 0.05     | DMSO    | 150             | 5          | 82                              | 71                              | 0                               |
| 12     | 0.01     | DMSO    | 100             | 5          | 2                               | 0                               | 0                               |
| 13     | -        | DMSO    | -              | -          | 67                              | -                               | -                               |
| 14     | -        | DMSO    | 150             | 5          | 63                              | -                               | -                               |
| 15     | 0.05     | DMSO    | 100             | 5          | 80                              | 15                              | 26                              |
| 16     | 0.05     | DMSO    | 100             | 5          | 4                               | 0                               | 0                               |

[a] Experiments were performed using approximately 18.5 GBq of $[^{11}C]CO_2$ as a starting activity from the cyclotron; [b] Radiochemical yield (% dc) was estimated as the actual final activity of $[^{11}C]_2$ or $[^{11}C]_3$ in relation to trapped $[^{11}C]CO_2$ (AT) (decay corrected); [c] TBAF (0.05 mmol) [d] without TBAF [e] blank experiments in DMSO
radiolabelling results were obtained when both the temperature and the reaction time were increased (entries 4 and 5). Nonetheless, under these conditions, the RCY observed was lower than 45% in all cases (entries 1–7, Table 1).

Next, DMSO and DMF were selected as solvents for radiolabelling. The choice was inspired by a recent work in which these solvents promoted the N-Formylation of amines using carbon dioxide and phenylsilane under mild conditions (Lv et al. 2016). Thus, these solvents might play a key role in promoting the CO₂ dissolution-insertion and the subsequent interaction of amines with the phenylsilane complexes towards the N-methylation reaction. When DMSO was used as a radiolabelling solvent at 150 °C for 5 min, the desired N-methylamine [¹¹C]2 was formed with a high radiochemical yield (71%, entry 11, Table 1), presumably due to the higher temperature employed. Indeed, a drop in the RCY for [¹¹C]2 in DMSO was observed as the labelling temperature decreased (entries 8–11, Table 1). It should be noted that the presence of TBAF proved to be essential because the amine [¹¹C]2 was not obtained in its absence (entry 12, Table 1), which is in accordance with the work of Liu and co-workers (Liu et al. 2016). Two blank experiments were carried out in DMSO to determine which species formed in the absence of amine. When [¹¹C]CO₂ was merely collected in a DMSO solution of PhSiH₃/TBAF (entry 13, Table 1), the formation of a single hydrophilic product eluting at tᵣ = 1.68 min was observed, which was presumed to be [¹¹C]HCOO⁻. Upon heating this solution for 5 min at 150 °C (entry 14, Table 1), the predominant radioactive compound eluted at tᵣ = 2.18 min. Based on the proposed reaction mechanism, it was hypothesised that this product could be the key formoxysilane (3) intermediate (See Additional file 1: Scheme S1) because it was the main radiochemical impurity observed in DMSO when amine was present in the labelling crude.

Finally, a significantly lower radiochemical yield was observed when DMF was tested as a reaction solvent compared to DMSO (entry 15 vs entry 9, Table 1). In some cases, the presence of N-[¹¹C]-4-tolylformamide ([¹¹C]3) was detected. This is in agreement with the proposed reaction mechanism in which a N-¹¹C-formamide would be a precursor for the N-¹¹C-methyl amine (Additional file 1: Scheme S1). Thus, when DMF was used as a solvent at 100 °C, ([¹¹C]3) was obtained with an RCY of 26% (entry 15, Table 1), whereas when using MeCN and DMSO as solvents at 80 and 50 °C, [¹¹C]3 was formed in 8 and 14% of RCY, respectively (entries 1 and 8, Table 1).

Another important parameter of the [¹¹C]CO₂ fixation methodology is the trapping efficiency of the reagent solution. This was evaluated as the relationship between maximum trapped [¹¹C]CO₂ activity (A₁) and [¹¹C]CO₂ EOB theoretical activity (A₀), and it was decay corrected. The trapping of [¹¹C]CO₂ was dramatically affected by the amount of TBAF in the solution. When no TBAF was present, the trapping efficiency of [¹¹C]CO₂ was less than 5% (entries 12 and 16, Table 1). This is consistent with the mechanism in which TBAF was necessary to form the adduct [PhSiH₃F⁻] that would have been responsible for solubilising the [¹¹C]CO₂ in the form of [¹¹C]HCOO⁻ (See Additional file 1: Scheme S1); however, increasing the amount of TBAF to 0.05 mmol did not appear to be favourable for a more efficient trapping (60%, entry 6, Table 1). All the solvents used showed good performance for trapping [¹¹C]CO₂ at room temperature when 0.01 mmol of TBAF was used independently from the amine concentration. Nevertheless, when no amine was added (blank runs in DMSO), a small decrease in the trapping efficiency was observed (entries 13 and 14, Table 1).
No losses of radioactivity due to the volatilisation of $[^{11}\text{C}]\text{CO}_2$ or other radioactive derivatives were observed during the radiolabelling step, which indicated that the $^{11}\text{C}$ species were quantitatively solubilised into the solution (Fig. 1). The losses of radioactivity were evaluated based on the relationship between $A_{\text{EOL}}/A_{\text{SOL}}$ and were decay corrected. In all cases, this relationship was approximately 100%.

**Radiosynthesis of $[^{11}\text{C}]\Pi\text{B}$**

In accordance with the encouraged results obtained for $N$-$[^{11}\text{C}-\text{methyl}]-4$-toluidine, DMSO was initially chosen for attempting the labelling of $[^{11}\text{C}]\Pi\text{B}$ with $[^{11}\text{C}]\text{CO}_2$, starting from 0.02 mmol (5.0 mg) of the precursor 6-OH-BTA-0. As shown in Table 2, labelling time varied (entries 1–5, Table 2), and it was found that at 2.5 min, the reductive incorporation of $[^{11}\text{C}]\text{CO}_2$ into $[^{11}\text{C}]\Pi\text{B}$ transpired with good RCY and radiochemical purity (RCP) (74% and 65%, respectively); even at 1.0 min, both parameters were promising (62% and 61%, respectively). To decrease the amount of 6-OH-BTA-0 to a value closer to those used for the $[^{11}\text{C}]\text{CH}_3\text{OTf}$ labelling methodology, 0.01 mmol (2.5 mg) and 0.005 mmol (1 mg) were used. Lowering the mass of 6-OH-BTA-0 was detrimental for the RCY as well as for RCP when 1 mg was employed (entry 6, Table 2); however, decreasing the molar amount of the precursor proved to be advantageous in terms of $A_m$ (entries 2, 7 and 6, Table 2) (as defined in Coenen et al. 2017).

These results could be correlated with a higher degree of reaction between increasing concentrations of the precursor and environmental CO$_2$ to obtain PiB (considering an excess of environmental CO$_2$ that dilutes $[^{11}\text{C}]\text{CO}_2$ during the experiments). In this context, a further flushing of the lines and the target lowered the concentration of unlabelled PiB, but the total concentration was still high for the purposes (entry 8, Table 2). The elevated values in the concentrations of PiB led to the assumption that other phenomena could be added to the incorporation of environmental CO$_2$, and in this sense, a contribution of a methyl group from the DMSO used as a solvent was
suspected. Indeed, as Jiang and co-workers (Jiang et al. 2014) reported, DMSO can be used as a methylating agent for amines in the presence of HCOOH at 150 °C. In this system, the presence of unlabelled formoxysilane (PhSiH₂OCHO) could lead to the activation of DMSO in the form of a methylmethylenesulphonium cation, which could act as the methylating agent depicted by the authors (as shown in Additional file 1: Scheme S2). To demonstrate the contribution of the methyl group from DMSO, two [11C]CO₂ blank experiments were carried out (entries 9 and 10, Table 2). As expected, a higher concentration of PiB was observed in the presence of DMSO and PhSiH₃/TBAF compared to that obtained when no PhSiH₃/TBAF were added to the DMSO solution.

In view of these assumptions, it is proposed that the use of an alternative solvent with a high boiling point might be favourable for improving the Aₘ of [11C]PiB. Indeed, the work of Liger and co-workers employed Diglyme as a reaction solvent for the direct [11C]methylation of amines from [11C]CO₂ (Liger et al. 2015). When using Diglyme in a [11C]CO₂ blank experiment carried out in the presence of PhSiH₃/TBAF, the concentration of PiB decreased considerably with respect to the corresponding experiment with DMSO (entry 11, Table 2).

Similar results were obtained in the presence of [11C]CO₂ in Diglyme at 150 °C for 2.5 min, decreasing the concentration of PiB to 105 μg/mL (entry 12). The RCP and RCY of the [11C] PiB achieved under these conditions were 57% and 64%, respectively. Further increasing the labelling time to 5 min allowed for obtaining a higher RCY and RCP for [11C] PiB, and the highest Aₘ achieved (entry 13, Table 2). Furthermore, the [11C]CO₂ trapping efficiency was 83% for this condition.

### Table 2: Optimisation of reaction conditions for radiolabelling [11C] PiB from direct [11C]CO₂ incorporation

| Entry | 6-OH-BTA-0 [mmol] | Solvent | Time (min) | [11C]CO₂[a] trapping efficiency (%), dc | RCP (%) of [11C]PiB | RCY (%), dc | PiB Concentration (μg/mL) | Aₘ (GBq/μmol) |
|-------|--------------------|---------|------------|----------------------------------------|---------------------|-------------|--------------------------|---------------|
| 1     | 0.02               | DMSO    | 1          | 53                                     | 61                  | 62          | -                        | -             |
| 2     | 0.02               | DMSO    | 2.5        | 52                                     | 65                  | 74          | 1511                     | 0.3           |
| 3     | 0.02               | DMSO    | 5.0        | 59                                     | 65                  | 68          | -                        | -             |
| 4     | 0.02               | DMSO    | 7.5        | 46                                     | 69                  | 55          | -                        | -             |
| 5     | 0.02               | DMSO    | 10         | 55                                     | 65                  | 50          | -                        | -             |
| 6     | 0.005              | DMSO    | 2.5        | 63                                     | 32                  | 33          | 83                       | 2.5           |
| 7     | 0.01               | DMSO    | 2.5        | 70                                     | 67                  | 48          | 761                      | 0.9           |
| 8     | 0.01               | DMSO    | 2.5        | 67                                     | 44                  | 51          | 333                      | 1.5           |
| 9     | 0.01               | DMSO    | 2.5        | -                                      | -                   | -           | 375                      | -             |
| 10    | 0.01               | DMSO    | 2.5        | -                                      | -                   | -           | 44                       | -             |
| 11    | 0.01               | dyglime | 2.5        | -                                      | -                   | -           | 56                       | -             |
| 12    | 0.01               | dyglime | 2.5        | 40                                     | 64                  | 57          | 105                      | 4.4           |
| 13    | 0.01               | dyglime | 5.0        | 83                                     | 68                  | 72          | 69                       | 16.2          |

[a] Experiments were performed using approximately 18.5 GBq of [11C]CO₂ as a starting activity from the cyclotron; [b] 'cold' and 'hot' flushes of targets and lines; [c] solution sparged with He (5 min, 40 mL/min); [d] without [11C]CO₂, [e] without [11C]CO₂ and without PhSiH₃/TBAF, [f] n = 3
It is important to highlight that the corresponding products of \( N \)-formylation and \( O \)-methylation, 2-(4′-\( N \)-[\( ^{11} \)C]formamidophenyl)-6-hydroxybenzothiazole and 2-(4′-amino-phenyl)-6-\( O \)\( ^{11} \)C] metoxybenzothiazole, respectively, were not observed under the conditions assayed (data not shown).

Full radiosynthesis and quality control of \(^{11} \)C\)PiB

Based on the results of the previous experiments, the complete radiosynthesis of \(^{11} \)C\) PiB in Diglyme was tested at 150 °C for 5 min using the starting activities of \(^{11} \)C\)CO\(_2\) in the range of 37 to 185 GBq. In general, it was observed that RCY dramatically dropped after the semipreparative HPLC separation, and it was found that this was due to a serious decrease in the radiochemical purity of the \(^{11} \)C\) PiB formed under these conditions. Indeed, highly lipophilic impurities were found to elute after passing a low polarity mixture through the chromatographic column, such as MeOH:THF (50:50 v/v) (see Additional file 1). Furthermore, an abrupt drop in the trapping efficiency of \(^{11} \)C\)CO\(_2\) was noted when 185 GBq of \(^{11} \)C\)CO\(_2\) was used as the starting activity. Nevertheless, it was possible to considerably improve the radiochemical yields and molar activities by means of decreasing both the molar amount of PhSiH\(_3\) and the radiolabelling time. The RCY of 28% and 26%, the \( A_m \) of 52.4 and 61.4 GBq/\( \mu \)mol and an RCP higher than 95% for \(^{11} \)C\) PiB were obtained when 0.30 mmol of PhSiH\(_3\) and labelling times of 2.5 and 1.0 min, respectively, were employed (Figs. 2 and 3). Three consecutive experiments were performed using the optimised radiolabelling condition of 1 min, and physicochemical quality control was performed. An RCP higher than 95% was verified for all the batches (Table 3). In all cases, molar activities were in compliance with the release criteria (> 30 GBq/\( \mu \)mol). Residual solvents were found to be present in quantities below their respective limits, and Diglyme or PhSiH\(_3\) were not detected in the formulated radiopharmaceutical.

To evaluate the applicability of this new approach for the radiosynthesis of \(^{11} \)C\) PiB, the classical methodology of nucleophilic \(^{11} \)C-methylation used in the radiopharmacy laboratory was compared with this novel radiosynthesis from direct \(^{11} \)C\)CO\(_2\).
incorporation. The $^{11}$C-methylation with $[^{11}$C]$\text{CH}_3\text{OTf}$, as reported by Phillipe and co-workers (Phillipe et al., Philippe et al. 2011), is currently used, which allows for obtaining an average activity of $[^{11}$C] PiB (EOS) of about 3.14 GBq (RCY 7%, dc) and an $A_m$ of 471 GBq/umol from ~160 GBq of cyclotron-produced $[^{11}$C]$\text{CO}_2$ ($N = 57$, all productions of the last two years) (Fig. 4). This new approach allows for obtaining a higher activity (more than two-fold) of EOS $[^{11}$C] PiB with a similar overall radiosynthesis time using the same automated module with minimal modifications. As was expected due to

![Image](image_url)

**Fig. 3** Analytical radio-HPLC for formulated $[^{11}$C]$\text{PiB}$: upper; UV (340 nm); lower; gamma

| Parameter                              | Release criteria         | 1      | 2      | 3      |
|----------------------------------------|--------------------------|--------|--------|--------|
| Visual inspection                      | Clear; colourless        | Pass   | Pass   | Pass   |
| pH                                     | 4.5–8.5                  | 5.5    | 5      | 5.2    |
| Radiochemical identity                 | Relative retention time: | 1.02   | 1.01   | 1.00   |
|                                        | 0.9–1.1                  |        |        |        |
| Radiochemical Purity                   | $\geq$ 95%               | 100    | 98.6   | 98.1   |
| PiB concentration ($\mu$g/mL)          | No limit established     | 4.61   | 4.15   | 4.06   |
| Molar Activity                         | $\geq$ 30 GBq/umol       | 60.3   | 62.3   | 61.6   |
| Precursor concentration ($\mu$g/mL)    | No limit established     | 10.25  | 3.06   | 1.63   |
| Total impurities concentration ($\mu$g/mL) | No limit established | 3.18   | 2.08   | 1.50   |
| Residual solvents analysis             | Acetone: < 0.5%          | 0.006  | 0.003  | 0.056  |
|                                        | Ethanol: < 10%           | 7.09   | 7.57   | 7.67   |
|                                        | Acetonitrile < 0.04%     | 0.004  | 0.008  | 0.005  |
|                                        | Diglyme: < 0.04%         | No detected | No detected | No detected |
|                                        | PhSiH$_3$: < 0.04%       | No detected | No detected | No detected |
| Radionuclides identity ($t_{1/2}$)    | 19.9–20.9 min            | 20.4   | 20.1   | 20.6   |
| Radionuclides purity                   | > 99.5% gamma emission at 511 keV | Ok     | Ok     | Ok     |

**Table 3** Physicochemical quality control parameters for three consecutive batches of $[^{11}$C]$\text{PiB}$ (labelling time 1 min)
isotopic dilution with environmental CO₂, the direct use of [¹¹C]CO₂ affected the Aₐᵈ, though to a degree compatible with the specifications.

**Conclusion**

It has been demonstrated that the direct [¹¹C]CO₂ fixation-reduction for the radiosynthesis of [¹¹C] PiB can be achieved using the PhSiH₃ / TBAF system. To obtain knowledge related to the radiochemical nature of the methodology and thus to produce [¹¹C] PiB with good and reproducible RCY and Aₐᵈ, the influence of physical and radiochemical parameters was investigated. Higher radiochemical yields and activities (EOS) of formulated [¹¹C] PiB from cyclotron-produced [¹¹C]CO₂ were obtained compared to that of the [¹¹C]-methylation method using PhSiH₃ / TBAF as a fixation-reduction system in Diglyme at 150 °C for 1 min.

Based on these results, a rapid one-pot methodology for the radiosynthesis of [¹¹C] PiB by means of the direct use of the primary precursor [¹¹C]CO₂, was developed by employing an automated commercial platform along with a physicochemical quality control proposed for its analysis. In this context, the study indicates the advantages of the unique published work in the application of a direct [¹¹C]CO₂ fixation-reduction methodology for the radiochemical productions of [¹¹C] PiB, especially in terms of reaction conditions (carbene- and metal-free), Aₐᵈ and overall radiosynthesis time.
Additional file

**Additional file 1:** Semipreparative HPLC chromatogram for \([^{11}C] \)PiB. (DOCX 735 kb)

Abbreviations

6-OH-BTA-0: 2-(4′-N-aminophenyl)-6-hydroxybenzothiazole; AD: Alzheimer’s Disease; Am: Molar activity; dc: decay corrected; Diglyme: Bis (2-methoxyethyl) ether; EOB: End of bombardment; EOS: End of synthesis; GC: Gas chromatography; GMP: Good manufacture practice; HPLC: High-performance liquid chromatography; ndc: no decay corrected; PET: Positron emission tomography; PhSiH3: Phenylsilane; PiB: Pittsburgh compound B (2-(4′-N-methylaminophenyl)-6-hydroxybenzothiazole); QC: Quality control; RCP: Radiochemical purity; RCY: Radiochemical yield; TBAF: Tetrabutylammonium fluoride

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Authors’ contributions

PB designed the methodology, performed the radiolabelling and optimisation of the radiosynthesis and was a major contributor in writing the manuscript. WP designed the methodology, discussed the results, contributed to the manuscript revision and approved its final version. ES approved the methodology design, contributed to the discussion of the results, the manuscript revision and its approval. All authors read and approved the final manuscript.

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References

Boudjemeline M, Hopewell R, Racon PL, Jolly D, Hamnami I, Villeneuve S, Kostikov A. Highly efficient solid phase supported radiosynthesis of \([^{11}C] \) PiB using tC18 cartridge as a “3-in-1” production entity. J Label Compd Radiopharm. 2017;60(14):632–8.

Coenen HH, Gee AD, Adam M, Antoni G, Cutler CS, Fujibayashi Y, et al. Consensus nomenclature rules for radiopharmaceutical chemistry—Setting the record straight. Nuc Med Biol. 2017;55:v–xi.

Coliva A, Monterisi C, Apollaro A, Gatti D, Penso M, Gianoli L, et al. Synthesis optimization of 2-(4-N-[11C] methylaminophenyl)-6-hydroxybenzothiazole ([11C] PiB), β-amloid PET imaging tracer for Alzheimer’s disease diagnosis. Appl Radiat Isot. 2015;105:66–71.

Das S, Bobbink FD, Laurenczy G, Dyson PJ. Metal-free catalyst for the Chemoselective methylation of amines using carbon dioxide as a carbon source. Angew Chem Int Ed. 2014;53(47):12876–9.

Engler H, Santillo AF, Wang SX, Lindau M, Savitcheva I, Nordberg A, Lannfelt L, Långström B, Kilander L. In vivo amyloid imaging with PET in frontotemporal dementia. Eur J Nucl Med Mol Imaging. 2008;35(1):100–6.

Herholz K, Carter SF, Jones M. Positron emission tomography imaging in dementia. Br J Radiol. 2007;80(special issue_2):S160–7.

Jacquet O, Das Neves Gomes C, Epiftr inventive M, Cantat T. Recycling of carbon and silicon wastes: room temperature formylation of N–H bonds using carbon dioxide and polyethyleneoxide. J Am Chem Soc. 2012;134(6):2934–7.

Jacquet O, Gomes CDN, Cantat T. CO 2 as a C 1-building block for the catalytic methylation of amines. Chem Sci. 2013;4(5):2127–31.

Jewett DM. A simple synthesis of \([^{11}C] \) methyl triflate. Int J Radi Appl Instrum A. Appl Radiat Isot. 1992;43(1):1383–5.

Jiang X, Wang C, Wei Y, Xue D, Liu Z, Xiao J. A general method for N-methylation of amines and nitro compounds with Dimethylsulfoxide. Chem Eur J. 2014;20(1):58–63.

Klunk WE, Engler H, Nordberg A, Wang Y, Blomqvist G, Holt DP, et al. Imaging brain amyloid in Alzheimer’s disease with Pittsburgh Compound-B. Ann Neurol. 2004;55(3):306–19.
Långström B, Kihlberg T, Bergström M, Antoni G, Björkman M, Forngren BH, et al. Compounds labelled with short-lived beta (+)-emitting radionuclides and some applications in life sciences. The importance of time as a parameter. Acta Chem Scand (Copenhagen, Denmark: 1989). 1999;53(9):651–69.

Li Y, Fang X, Junge K, Beller M. A general catalytic methylation of amines using carbon dioxide. Angew Chem. 2013;125(36):9747–50.

Liger F, Ejstjouls T, Cadarossanesaib F, Tourvielle C, Le Bars D, Billard T. Direct [11C] methylation of amines from [11C] CO2 for the synthesis of PET radiotracers. Eur J Org Chem. 2015;2015(29):6434–8.

Liu XF, Ma R, Qiao C, Cao H, He LN. Fluoride-catalyzed methylation of amines by reductive functionalization of CO2 with Hydroxilanes. Chem Eur J. 2016;22(46):16489–93.

Lv H, Xing Q, Yue C, Lei Z, Li F. Solvent-promoted catalyst-free N-formylation of amines using carbon dioxide under ambient conditions. Chem Commun. 2016;52(39):6945–6.

Mathis CA, Wang Y, Holt DP, Huang GF, Debonath ML, Klink WE. Synthesis and evaluation of 11C-labeled 6-substituted 2-arybenzothiazoles as amyloid imaging agents. J Med Chem. 2003;46(13):2740–54.

Nordberg A. PET imaging of amyloid in Alzheimer’s disease. Lancet Neurol. 2004;3(9):519–27.

Nordberg A. Amyloid plaque imaging in vivo: current achievement and future prospects. Eur J Nucl Med Mol Imaging. 2008;35(1):46–50.

Philippe C, Haeusler D, Mitterhauser M, Ungerboeck J, Viemstein H, Dudczak R, Wadsak W. Optimization of the radiosynthesis of the Alzheimer tracer 2-4-N-[11C] methylaminophenyl-6-hydroxybenzothiazole ([11C] PIB). Appl Radiat Isot. 2011;69(9):1212–7.

Rabinovici GD, Jagust WJ. Amyloid imaging in aging and dementia: testing the amyloid hypothesis in vivo. Behav Neurol. 2009;2(1), 2:117–28.

Rotstein BH, Liang SH, Holland JP, Collier TL, Hooker JM, Wilson AA, Vaisdev N. 11 CO 2 fixation: a renaissance in PET radiochemistry. Chem Commun. 2013;49(50):5621–9.

Savio E, Garcia O, Trindade V, Buccino P, Giglio J, Balter H, Engler H. Improving production of 11C to achieve high specific labelled radiopharmaceuticals. AIP Conf Proc. 2012;1509:185–189.

Shekhar AC, Kumar AR, Sathalia G, Paul VL, Sridhar M, Rao PS. Facile N-formylation of amines using Lewis acids as novel catalysts. Tetrahedron Lett. 2009;50(50):7099–101.

Solbach C, Uebele M, Reischl G, Machulla HJ. Efficient radiosynthesis of carbon-11 labelled uncharged Thioflavin T derivatives using [11C] methyl triflate for β-amyloid imaging in Alzheimer’s disease with PET. Appl Radiat Isot. 2005;62(4):591–5.

Furniss BS, Hannaford AJ, Rogers V, Smith PWG, Tatchell AR. Vogel’s Textbook of Practical Organic Chemistry, Including Quotative Organic Analysis. ELBS; 1978.

Wilson AA, Garcia A, Chestakova A, Kung H, Houle S. A rapid one-step radiosynthesis of the β-amyloid imaging radiotracer N-methyl-[11C]-2-(4′-methylaminophenyl)-6-hydroxybenzothiazole ([11C]-6-OH-BTA-1). J Label Compd Radiopharm. 2004;47(10):679–82.