Marine Alga Sargassum Horneri Component And Bone Homeostasis: Role In Osteoporosis Prevention

Yamaguchi M*

Division of Diabetes, Endocrinology and Metabolism, Department of Medicine, Baylor College of Medicine, Houston, TX 77030, USA.

Abstract

Bone homeostasis is maintained through a balance between osteoblastic bone formation and osteoclastic bone resorption. Aging induces bone loss due to decreased osteoblastic bone formation and increased osteoclastic bone resorption. Osteoporosis, which is a major public health problem, has been a worthy of notice in maintaining bone health and prevention of bone loss with increasing age. Nutritional factors may play a role in the prevention of bone loss with aging. Among marine algae of Undaria pinnatifida, Sargassum horneri, Eisenia bicyclis, Cryptonemia cernitzaiana, Gelidium amansii, and Ulva pertusa Kjellman which were gathered seasonally, Sargassum horneri (S. horneri) was found to have a unique anabolic effect on bone components. S. horneri extract had a stimulatory effect on osteoblastic bone formation and an inhibitory effect on osteoclastic bone resorption in vitro, thereby increasing bone mass. The intake of S. horneri extract caused a preventive effect on bone loss in animal models for osteoporosis and in healthy human. S. horneri extract, which is a functional food, may be usefulness as an osteogenic factor in preventing osteoporosis in human subjects.

Key Words: S. horneri extract, Osteoblastic bone formation, Osteoclastic bone resorption, Osteoporosis

Introduction

Bone is a dynamic tissue that preserves skeletal size, shape, and structural integrity and to regulate mineral homeostasis. Bone homeostasis is maintained through a balance between osteoblastic bone formation and osteoclastic bone resorption. Aging and numerous pathological processes induce decrease in bone formation and increase in bone resorption, leading to osteoporosis, a devastating bone disease [1]. Osteoporosis, which is induced with decrease in bone mass, is widely recognized as a major public health problem. The most dramatic expression of the disease is represented by fractures of the proximal femur for which the number increases as the population ages [2].

Nutritional factors may have the potential effect to prevent bone loss with increasing age. There is growing evidence that the supplementation of nutritional and food factors may have the preventive effect on bone loss that is induced in animal model of osteoporosis and in human subjects [3-6]. Functional factors in food and plants, which regulate on bone homeostasis, have been to be worthy of notice in maintaining of bone health and prevention of bone loss with increasing age [7-13].

The effect of marine algae on bone metabolism was not determined so far. Undaria pinnatifida, Sargassum horneri, Eisenia bicyclis, Cryptonemia cernitzaiana, Gelidium amansii, and Ulva pertusa Kjellman are applied as food materials. Among these marine algae, Sargassum horneri (S. horneri) was found to have a unique anabolic effect on bone components [14]. S. horneri extract has been found to have a stimulatory effect on osteoblastic bone formation and an inhibitory effect on osteoclastic bone resorption in vitro, thereby increasing bone mass.

This review has been written to outline the recent advances that have been made concerning the anabolic effect of S. horneri extract on bone metabolism and will discuss the usefulness of this functional food factor in prevention of osteoporosis.

Cellular regulation of bone homeostasis

Bone homeostasis is skillfully regulated through the functions of osteoblasts and osteoclasts, which are major cells in bone tissues [16-20]. Osteoclasts, which develop from hematopoietic progenitors, are recruited to the site and excavate the calcified matrix. In the physiologic process of bone turnover, a resorptive stimulus firstly triggers recruitment of osteoclasts to a site on the bone surface. During the initiation phase of bone resorption, osteoclasts arising from local mesenchymal stem cells assemble at the bottom of the cavity and bone formation begins. After the resorbed lacunar pit is filled with new osteoid, osteoblasts become flatter and less active, with the final newly remodeled bone surface lined by flat lining cells. As bone formation progresses,
some osteoblasts are entombed within the matrix as osteocytes but the majority dies by apoptosis. When mechanical forces are reduced, for example in weightlessness, osteocytes die by apoptosis. This event appears to act as a beacon for osteoclast recruitment and generation of a new basic multicellular unit, which in turn replaces the old bone containing dead osteocytes with new bone containing viable osteocytes. Remodeling of cancellous bone begins with the retraction of lining cells that cover the bone surface [19,20].

Bone acts as major storage site for growth factors, which are produced by osteoblasts, diffuse into newly deposited osteoid and are stored in the bone matrix including insulin-like growth factors (IGF-1 and II), transforming growth factor-β1 (TGF-β1), platelet-derived growth factor (PDGF), or bone morphologic proteins (BMPs) [21,22]. These bone-derived factors, which can be liberated during subsequent periods of bone resorption, act in an autocrine, paracrine, or delayed paracrine fashion in the local microenvironment of the bone surface.

This process of bone remodeling to make bone unique among organs and tissues, and it also adds so many levels of complexity, with respect to interactions along the remodeling sequence by systemic influences (hormones), stress action on trabecular and cortical systems (physical activity/weight bearing), growth factors and cytokines produced by the bone cells which act locally on their own cell types and on the other bone cell types, or factors that come from nearby cells present in the marrow tissues.

**S. horneri** extract stimulates osteoblastic bone formation

Marine algae *S. horneri* was seasonally gathered from the coast at Shimoda (Shizuoka Prefecture, Japan) and Miyako (Iwate Prefecture, Japan), and it was freeze-dried and powdered [14]. The gathered fresh marine algae were homogenized in distilled water and were centrifuged at 5500 g in a refrigerated centrifuge for 10 min. The 5500 g supernatant fraction was pooled for freeze-drying. The powder of the water-solubilized extract was dissolved in ice-cold distilled water for use in the experiments. The water-solubilized extract from *S. horneri* was purified through the method of membrane fractionation to collect active component of various molecular weights.

The direct effect of *S. horneri* extract on bone formation and mineralization has been examined using MC3T3 preosteoblastic cells in vitro [15]. The preosteoblastic cells were differentiated into mineralizing osteoblasts in vitro in the presence of absence of *S. horneri* extract in mineralizing medium for 21 days and stained for calcium deposition with Alizarin Red-S. *S. horneri* extract (10 and 25 μg/ml) was found to potentely enhance mineralization in MC3T3 cells [15].

The *S. horneri* extract (25 - 100 μg/ml of medium) did not have a significant effect on cell number of MC3T3-E1 cells with culture. *S. horneri* extract did not appear to mediate direct toxic effects on the cultures as cells were proliferated robustly over the culture period, and were still alive and visibly attached to the plate at the end of the experiment as compared with those of control group.

Bone morphogenic proteins (BMPs) such as BMP-2 are anabolic agents that signal through the Smad signaling pathway [22]. The effect of *S. horneri* extract on basal and BMP-2-induced Sma activation has been shown using a Smad 4-luciferase reporter responsive to all Smad species [15]. *S. horneri* extract did not have a direct effect on basal Smad activation. *S. horneri* extract (50 or 100 μg/ml) significantly enhanced Smad-activation induced by BMP-2. Also, *S. horneri* extract (25 -100 μg/ml) significantly enhanced Smad-activation induced by TGF-β1 [15].

NF-κB activation is a potent inhibitor to osteoblast differentiation, and TNF-α-induced NF-κB activation leads to Smad suppression in MC3T3 osteoblast precursors [17,18]. Whether *S. horneri* extract is able to prevent TNF-α-induced NF-κB activation in MC3T3 osteoblast precursors was examined [15]. *S. horneri* extract (25 – 100 μg/ml of medium) significantly prevented TNF-α-induced NF-κB activation, although *S. horneri* extract did not have a direct effect on basal NF-κB activation.

Thus, *S. horneri* extract has been shown to stimulate osteoblastic differentiation and mineralization *in vitro*. TGF-β1- and BMP-2-induced activation of Smad signaling respectively plays an important role in the early commitment and differentiation of osteoblasts [22]. *S. horneri* extract was found to enhance BMP-2- or TGF-β1-induced Smad activation with the dose-dependency [15]. *S. horneri* extract-induced Smad enhancement may be an important to stimulate osteoblastic differentiation and mineralization.

NF-κB signaling has been shown to downregulate osteoblast differentiation [23,24]. One major mechanism appears to involve the intersection of NF-κB with the Smad signaling pathway [23]. NF-κB signaling in osteoblasts intersects and disrupts Smad signaling by promoting production of Smad7, an inhibitor of TGF-β1- and BMP-induced R-Smad activation [25]. TNF-α further antagonizes BMP signaling by upregulating Smad ubiquitination regulatory factor 1 (Smurf1), promoting proteasomal degradation of bone morphogeneic signaling proteins [26]. Multiple suppressors of NF-κB activation are capable of rescuing the inhibitory effect of TNF-α on BMP-2 and/or TGF-β1-induced Smad activation [23]. *S. horneri* extract was found to suppress TNF-α-induced NF-κB activation in preosteoblastic MC3T3-E1 cells. This finding provides a possible mechanism by which *S. horneri* extract stimulates osteoblastic bone formation.

The anabolic effect of *S. horneri* extract on bone tissues has been found. Rat femoral-metaphyseal tissues were cultured in a medium containing water-solubilized extract (25 and 50 μg/ml) obtained from *U. pinnatifida*, *S. horneri*, *E. bicyclis*, or *C. myctiziana* in vitro [14]. The bone calcium content was significantly elevated in the presence of *S. horneri* extract (25 and 50 μg/ml). No effect was seen in the extracts of other marine algae. In addition, water suspensions (5%) of marine alga powder were orally administered once daily for 7 days. Bone calcium content was significantly increased after the administration of *U. pinnatifida*, *S. horneri*, *E. bicyclis*, or *C. myctiziana* [14].

Bone morphogenic proteins (BMPs) such as BMP-2 are anabolic agents that signal through the Smad signaling pathway [22]. The effect of *S. horneri* extract on basal and BMP-2-induced Sma activation has been shown using a Smad 4-luciferase reporter responsive to all Smad species [15]. *S. horneri* extract did not have a direct effect on basal Smad activation. *S. horneri* extract (50 or 100 μg/ml) significantly enhanced Smad-activation

**S. horneri** extract inhibits osteoclastic bone resorption

Bone morphogenic proteins (BMPs) such as BMP-2 are anabolic agents that signal through the Smad signaling pathway [22]. The effect of *S. horneri* extract on basal and BMP-2-induced Sma activation has been shown using a Smad 4-luciferase reporter responsive to all Smad species [15]. *S. horneri* extract did not have a direct effect on basal Smad activation. *S. horneri* extract (50 or 100 μg/ml) significantly enhanced Smad-activation induced by BMP-2. Also, *S. horneri* extract (25 -100 μg/ml) significantly enhanced Smad-activation induced by TGF-β1 [15].

NF-κB activation is a potent inhibitor to osteoblast differentiation, and TNF-α-induced NF-κB activation leads to Smad suppression in MC3T3 osteoblast precursors [17,18]. Whether *S. horneri* extract is able to prevent TNF-α-induced NF-κB activation in MC3T3 osteoblast precursors was examined [15]. *S. horneri* extract (25 – 100 μg/ml of medium) significantly prevented TNF-α-induced NF-κB activation, although *S. horneri* extract did not have a direct effect on basal NF-κB activation.

Thus, *S. horneri* extract has been shown to stimulate osteoblastic differentiation and mineralization *in vitro*. TGF-β1- and BMP-2-induced activation of Smad signaling respectively plays an important role in the early commitment and differentiation of osteoblasts [22]. *S. horneri* extract was found to enhance BMP-2- or TGF-β1-induced Smad activation with the dose-dependency [15]. *S. horneri* extract-induced Smad enhancement may be an important to stimulate osteoblastic differentiation and mineralization.

NF-κB signaling has been shown to downregulate osteoblast differentiation [23,24]. One major mechanism appears to involve the intersection of NF-κB with the Smad signaling pathway [23]. NF-κB signaling in osteoblasts intersects and disrupts Smad signaling by promoting production of Smad7, an inhibitor of TGF-β1- and BMP-induced R-Smad activation [25]. TNF-α further antagonizes BMP signaling by upregulating Smad ubiquitination regulatory factor 1 (Smurf1), promoting proteasomal degradation of bone morphogeneic signaling proteins [26]. Multiple suppressors of NF-κB activation are capable of rescuing the inhibitory effect of TNF-α on BMP-2 and/or TGF-β1-induced Smad activation [23]. *S. horneri* extract was found to suppress TNF-α-induced NF-κB activation in preosteoblastic MC3T3-E1 cells. This finding provides a possible mechanism by which *S. horneri* extract stimulates osteoblastic bone formation.

The anabolic effect of *S. horneri* extract on bone tissues has been found. Rat femoral-metaphyseal tissues were cultured in a medium containing water-solubilized extract (25 and 50 μg/ml) obtained from *U. pinnatifida*, *S. horneri*, *E. bicyclis*, or *C. myctiziana* in vitro [14]. The bone calcium content was significantly elevated in the presence of *S. horneri* extract (25 and 50 μg/ml). No effect was seen in the extracts of other marine algae. In addition, water suspensions (5%) of marine alga powder were orally administered once daily for 7 days. Bone calcium content was significantly increased after the administration of *U. pinnatifida*, *S. horneri*, *E. bicyclis*, or *C. myctiziana* [14]. Also, bone alkaline phosphatase activity, which is an enzyme for calcification [27,28], was significantly enhanced with the administration of *S. horneri* or *G. amansii*. Thus, *S. horneri* extract has a unique anabolic effect on bone calcification *in vitro* and *in vivo*. This was the first time finding. The effects of *S. horneri* extract in increasing calcium content, alkaline phosphatase activity, and deoxyribonucleic acid (DNA) content in the femoral-diaphyseal and -metaphyseal tissues in vitro was completely abolished in the presence of cycloheximide, an inhibitor of protein synthesis [29]. DNA may be an index of cell number in bone tissues [16,21]. The anabolic effect of *S. horneri* extract may be resulted from newly synthesized protein components.
S. horneri extract has been shown to suppress osteoclastic bone resorption. To examine the effect of S. horneri extract on osteoclast formation, RAW264.7 osteoclast precursors were differentiated into mature osteoclasts by stimulation with RANKL in the presence or absence of S. horneri extract with the dose range of 5 to 100 μg/ml of medium [15]. The S. horneri extract had no effect on cell number of preosteoclasts (RAW267.4 cells) with culture. S. horneri extract (100 μg/ml) did not have a toxicity to the precursors suppressing their proliferation over 7 days of culture [15]. S. horneri extract (25 – 100 μg/ml) significantly suppressed osteoclast formation induced by RANKL [15].

The NF-κB signal transduction pathway is essential for the formation of osteoclasts [17,18]. The effect of S. horneri extract on NF-κB activation by RANKL in osteoclast precursors has been examined. RAW 264.7 cells were transfected with an NF-κB reporter and stimulated with RANKL to induce NF-κB activity in the presence or absence of S. horneri extract (5 – 100 μg/ml) [15]. S. horneri extract did not have a significant effect on basal NF-κB activity [15]. However, RANKL-induced increase in NF-κB activity was significantly blunted in the presence of S. horneri extract [15].

S. horneri extract with dose-dependency, which did not have cell toxicity, was found to have a suppressive effect on RANKL-stimulated osteoclastogenesis [15]. RANKL, the key osteoclastogenic cytokine, is central to formation of osteoclasts, the cells that resorb bone, through NF-κB signaling [17,18]. S. horneri extract suppressed RANKL-induced NF-κB activation in osteoclast precursor cells [15]. This finding coincided with the observation that S. horneri extract suppresses RANKL-induced osteoclastogenesis [15]. S. horneri extract may inhibit osteoclastogenesis through suppression of NF-κB activation.

The effect of S. horneri on mineralization in MC3T3-E1 cells with culture for 21 days was observed at 10 μg/ml [15]. However, the effects of S. horneri on Smad activity in MC3T3-E1 cells and on NF-κB activity in RAW 264.7 cells with culture for 24 hours were observed at 25 μg/ml [15]. The effects of S. horneri on osteoclastogenesis in RAW 264.7 cells with culture for 6 days were also revealed at 25 μg/ml [15]. The effect of S. horneri on osteoblastogenesis and osteoclastogenesis may be observed with lower doses in the culture for longer periods.

The effects of S. horneri extract on bone resorption using femoral tissues in vitro have been examined [30]. Parathyroid hormone (PTH)- and prostaglandine E2 (PGE2) are known to induce osteoclastic bone resorption [31-33]. PTH- and PGE2-induced decreases in bone calcium content were completely inhibited after culture with water-solubilized extracts (10, 25, and 50 μg/ml) [30]. Also, the extract completely inhibited the PTH- or PGE2-induced increase in medium glucose consumption and lactic acid production by bone tissues [30]. Moreover, S. horneri extracts blocked the PTH-induced increase in acid phosphatase activity in the diaphyseal and metaphyseal tissues [30]. These findings indicate that the water-solubilized extracts of S. horneri have a direct inhibitory effect on bone resorption in tissue culture in vitro.

S. horneri extract possesses a potent Smad activation and anti-NF-κB activity and may have promise for development into an antosteoporotic material capable of promoting new bone formation while simultaneously reducing bone resorption. S. horneri extract may be usefulness as a tool in the prevention of osteolysis with various pathophysiologic states. The mechanism by which S. horneri stimulates bone mass is summarized in Fig. 1.

Characterization of active component in S. horneri extract

Figure 1: S. horneri regulates bone homeostasis. S. horneri component stimulates osteoblastic bone formation and suppresses osteoclastic bone resorption, thereby increasing bone mass. S. horneri component enhances bone growth factors (TGF-β1 and BMP-2)-induced Smad activation in osteoblasts and suppresses TNF-α- and RANKL-enhanced NF-κB activation in preosteoclasts.
S. horneri extracts has been found to stimulate osteoblastic bone formation [14,15,29] and to suppress osteoclastic bone resorption [30]. Those active components have been found to be present in S. horneri extract obtained from various coasts in both Japan (Shimoda and Iwate, Japan) and China [34]. The active component of S. horneri extract in stimulating bone calcification has been found to be near molecular weight (MW) 3000 [15,34]. Meanwhile, the active component of S. horneri extract in inhibiting osteoclastic cell formation was less than MW 3000 and over MW 50000 [15,34]. These components were stable under heat treatment. It has been speculated that the active component in stimulating bone calcification is a chemical but not peptide, and that the component over MW 50000 in inhibiting bone resorption may be a polysaccharide. These active components obtained from the coasts of Iwate (Japan) or China showed an identical molecular weight. The active component, which stimulates osteoblastogenesis and suppresses osteoclastogenesis, was found to be present in the components less than MW 3000 of S. horneri extract using preosteoblastic cells and RAW267.4 cells in vitro [15]. The active component that stimulates osteoblastic bone formation and suppresses osteoclastic bone resorption may be identical.

We found the existence of 4 chemicals in S. horneri components (less than MW 3000) using the analysis with liquid chromatography mass spectrophotometry system (LCMS-IT-TOF; Shimadzu, Kyoto, Japan). These chemicals were identified as 1,3,5-tris(oxolan-2-ylmethyl)-1,3,5-triazinan (MW 339), 5-phenyl-2-[2-(5-pentyltetrazol-2-yl)ethyl]tetrazole (MW 318), 3-(hexadecylamino)propane-1,2-diol (MW 316), and 2-(2-hydroxyethyl-tridecyl-aminoo)ethanol (MW 288). These chemicals may have an effect on osteoblastogenesis and/or osteoclastogenesis, although their biological effects on bone cells remain to be elucidated. It is possible that combination of these compounds has an anabolic effect on bone.

Anabolic effects of S. horneri extract on bone in vivo

The anabolic effects of S. horneri extract on bone components in the femoral tissues of young and aged rats in vivo have been demonstrated [35]. Calcium content, alkaline phosphatase activity, and DNA content in the femoral-diaphyseal and metaphyseal tissues of young male (4-week-old) rats were significantly increased after the administration of S. horneri extract (25, 50, and 100 mg/kg) for 7 days [35]. Moreover, these bone components in the femoral-diaphyseal and metaphyseal tissues of aged female (50-week-old) rats were significantly increased after the administration of S. horneri extract (100 mg/kg) for 14 days, suggesting a preventive effect on bone loss with increasing age [35].

The intake of S. horneri extract has a preventive effect on bone loss in a pathophysiologic state has also been shown [36]. Diabetes has been shown to induce bone loss [37-39]. Streptozotocin (STZ) induces decrease in insulin secretion in pancreatic cells and causes type 1 diabetes. The oral administration of S. horneri extract (100 mg/kg body weight) to STZ (60 mg/kg body weight)-diabetic rats was found to have a preventive effect on bone loss with diabetes in vivo [36]. This finding suggests that the dietary intake of S. horneri extract has a preventive effect on bone loss in the pathophysiologic state. When the femoral tissues obtained from STZ-diabetic rats were cultured in medium containing S. horneri extract solution, the femoral calcium content and alkaline phosphatase activity were significantly increased in vitro [36]. Alkaline phosphatase is related to bone calcification [27,28]. S. horneri extract has a stimulatory effect on bone formation [14,15,29] and an inhibitory effect on bone resorption in vitro [15,30]. Presumably, the preventive effect of S. horneri extract administration on diabetes-induced bone loss may be related to a direct action of the active component of S. horneri extract.

Interestingly, the oral administration of S. horneri extract (100 mg/kg body weight) to STZ-diabetic rats has been found to have a significant preventive effect on the decrease in body weight and the increase in serum glucose and triglyceride levels induced in the diabetic state [36]. This was a novel finding. The intake of S. horneri extract has a partial restorative effect on serum biochemical finding with diabetes in vivo.

Thus, the intake of S. horneri extract has been demonstrated to have preventive effects on bone loss, hyperglycemia, and hyperlipidemia in STZ-diabetic rats [36]. The active component of S. horneri extract in preventing bone loss induced with diabetic state may be identical to the component that prevents an elevation of serum glucose and triglyceride levels with diabetes. This remains to be elucidated.

Supplemental intake of S. horneri extract has an anabolic effect on bone metabolism in human subjects

The supplemental intake of S. horneri extract has been shown to have an anabolic effect on bone metabolism in human subjects [40]. Study has been undertaken to determine the effect of supplemental intake of the water-solubilized S. horneri extract on circulating bone metabolic markers in healthy human [40]. Thirty-six volunteers, aged 20-60 years (16 men and 20 women), were enrolled. Volunteers were divided into three groups; placebo tablet without S. horneri extract (5 men and 7 women), tablet containing S. horneri extract at 300 mg/day (6 men and 7 women) or 900 mg/day (5 men and 6 women) [40]. Placebo or S. horneri extract tablet was ingested once a day for 4 or 8 weeks [40]. The intake of dietary S. horneri extract (900 mg/day) for 8 weeks did not have a significant alteration in other biochemical markers for the metabolic function of organs, suggesting that the intake does not have toxic effects in humans [40].

Bone-specific alkaline phosphatase [41] and γ-carboxylated osteocalcin [42] are serum bone markers of bone formation, and bone tartrate-resistant acid phosphatase (TRACP) [43] and N-telopeptides of type I collagen [44] are markers of bone resorption. Serum bone-specific alkaline phosphatase or γ-carboxylated osteocalcin concentration was not significantly changed after the intake of S. horneri extract (300 or 900 mg/day) for 4 or 8 weeks [40]. Serum bone tartrate-resistant acid phosphatase (TRACP) activity was significantly decreased after the intake of S. horneri extract (300 or 900 mg/day) for 8 weeks [40]. Serum N-telopeptides of type I collagen concentration was significantly decreased after the intake of S. horneri extract (900 mg/day) for 8 weeks [40]. Meanwhile, serum calcium, inorganic phosphorus, and other biochemical findings were not changed after the intake of S. horneri extract (300 or 900 mg/day) for 4 or 8 weeks [40]. Thus, the prolonged intake of S. horneri extract has been found to have inhibitory effects on bone resorption in humans.

S. horneri extract has been shown to have a stimulatory effect on osteoblastic bone formation and an inhibitory effect on osteoclastic bone resorption. It has been speculated that the supplemental intake of S. horneri extract reveals suppressive effects on...
bone resorption, and later it exhibits stimulatory effects on bone formation in humans, thereby increasing bone mass. The supplemental intake of \textit{S. horneri} extract may have a preventive effect on bone loss with increasing age and post-menopausal women.

**Conclusion**

Aging and numerous pathological processes induce decrease in bone formation and increase in bone resorption, leading to osteoporosis, which is induced with decrease in bone mass. Osteoporosis is widely recognized as a major public health problem. Functional food factors may have the potential effect to prevent bone loss with increasing age. There is growing evidence that the supplementation of food factors may have the preventive effect on bone loss that is induced in animal model of osteoporosis and in human subjects. Among various marine algae which are applied to food, \textit{S. horneri} extract has been found to have a unique anabolic effect on bone mass using stimulating osteoblastic bone formation and depressing osteoclastic bone resorption. The intake of \textit{S. horneri} extract has usefulness in prevention of bone loss with increasing age and pathologic state. This biomedical food material may play a role in maintaining of bone health and preventing of osteoporosis.

**Author Contribution**

Masayoshi Yamaguchi contributed to the design and conduct of the study, collection, analysis, and interpretation of data, and manuscript writing.

**Acknowledgement**

\textit{S. horneri} extract was supplied through Masaaki Yokoyama and Toru Matsumoto, Biomaterial Department, Muruhachi Muramatsu, Inc., Yaizu, Japan.

**References**

[1]. Weitzmann MN, Pacifici R (2006) Estrogen deficiency and bone loss: an inflammatory tale. J Clin Invest 116: 1186-1194
[2]. Johnell O, Kanis JA (2006) An estimate of the worldwide prevalence and disability associated with osteoporotic fractures. Osteoporos Int 17: 1726-1732
[3]. Bonjour J-P, Schurch M-A, Rizzoli R (1996) Nutritional aspects of hip fracture. Bone 18: 1395-1445
[4]. Yamaguchi M (2006) Regulatory mechanism of food factors in bone metabolism and prevention of osteoporosis. Yakugaku Zasshi 126:117-1137
[5]. Yamaguchi M (2010) Nutritional Factors and Osteoporosis Prevention. Nova Science Publishers, Inc., New York
[6]. Yamaguchi M (2010) Role of nutritional zinc in the prevention of osteoporosis. Mol Cell Biochem 338: 241-254
[7]. Yamaguchi M (2002) Isoflavone and bone metabolism: Its cellular mechanism and preventive role in bone loss. J Health Sci 48: 209-222
[8]. Yamaguchi M (2012) Nutritional factors and bone homeostasis: Synergistic effect of zinc and genistein osteostasis. Mol Cell Biochem 366:201-223
[9]. Yamaguchi M (2012) Role of beta-cryptoxanthin in bone homeostasis. J Biomedical Sci, 19:36-48
[10]. Yamaguchi M, Hamamoto R, Uchiyama S, Ishiyama K (2008) Effects of flavonoid on calcium content in femoral tissue culture and parathyroid hormone-stimulated osteoclastogenesis in bone marrow culture in vitro. Mol Cell Biochem 303: 83-88
[11]. Yamaguchi M, Uchiyama S, Nakagawa T (2007) Preventive effects of bee pollen \textit{Cistus ladaniferus} extract on bone loss in ovariectomized rats in vivo. J Health Sci 53: 571-575
[12]. Yamaguchi M, Ma ZJ, Suzuki T (2005) Anabolic effect of wasabi leafstalk (\textit{Wasabia Japonica} MATSUM.) extract on bone components in the femoral-diaaphyseal and -metaphyseal tissues of aged female rats in vitro and in vivo. J Health Sci 49: 123-128
[13]. Yamaguchi M (2012) Bone anabolic effect of wasabi leafstalk components: its related phytochemicals p-hydroxybenzoyl acid prevents osteoporosis. Open Access Scientific Reports 1:194. doi:10.4172/scientificreports.19
[14]. Yamaguchi M, Hachiya S, Hirotsuka S, Suzuki T (2001) Effect of marine algae extract on bone calcification in the femoral-metaphyseal tissues of rats: Anabolic effect of \textit{Sargassum horneri}. J Health Sci 47: 533-538
[15]. Yamaguchi M, Matsutomo T (2012) Marine algae \textit{Sargassum horneri} bioactive factor stimulates osteoclastogenesis and suppresses osteoclastogenesis in vitro. OA Biorechemistry 1:3
[16]. Parfitt AM (1990) Bone-forming cells in clinical conditions. In: Hall BK, editor. Bone Volume 1. The Osteoblast and Osteocyte. Boca Raton, FL: Taylor & Francis Press and CRC Press, pp351-429
[17]. Zaidi M, Blair HC, Moonga BS, Abe E, Huang C L-H (2003) Osteoclastogenesis, bone resorption, and osteoblast-based therapeutics. J Bone Miner Res 18: 599-609
[18]. Chambers TJ, Fuller K (2011) How are osteoclasts induced to resorb bone? Ann N Y Acad Sci 1240:1-6
[19]. Baron R, Vigary A, Horowitz M (1984) Lymphocytes, macrophages and the regulation of bone remodeling. Bone Miner Res 2:175-243
[20]. Raggatt LJ, Partridge C (2010) Cellular and molecular mechanisms of bone remodeling. J Biol Chem 285: 25103-25108
[21]. Canalis E, McCarty T, Centrella M (1988) Growth factors and the regulation of bone remodeling. J Clin Invest 81: 277-281
[22]. Chen G, Deng C, Li YP (2012) TGF-B and BMP signaling in osteosteatosis and bone formation. J Int Biol Sci 8:272-288
[23]. Li Y, Li A, Strait K, Zhang H, Nanes MS, Weitzmann MN (2007) Endogenous TNFalpha lowers maximum peak bone mass and inhibits osteoblastic Sma activation, through NF-kappaB. J Bone Miner Res 22:646-655
[24]. Chang J, Wang Z, Tang E, Fan Z, McAuley L, Franceschi R, Guan K, Krebsbach PH, Wang CY (2009) Inhibition of osteoblastic bone formation by nuclear factor-kappaB. Nat Med 15:682-9
[25]. Elsevra RA, Schwarz EM, Ziczik MJ, O’Keefe RJ, Drissi H, Rosier RN (2006) Smad7 mediates inhibition of Sos2 osteosarcoma cell differentiation by NF-kappaB. Exp Cell Res 312:40-50
[26]. Guo R, Yamashita M, Zhang Q, Zhou Q, Chen D, Reynolds DG, Awad HA, Yanosko L, Zhao L, Schwarz EM, Zhang YE, Boyce BF, Xing L (2008) Ubiquitin ligase Smurf1 mediates tumor necrosis factor-induced systemic bone loss by promoting proteasomal degradation of bone morphogenetic signaling proteins. J Biol Chem 283:23084-92
[27]. Lian JB, Stein GS, Canalis E, Roby PG, Boskey AL (1999) Bone formation: Osteoblast lineage cells, growth factors, matrix proteins, and the mineralization process. In: Primer on the Metabolic Bone Diseases and Disorders of Mineral Metabolism (Favus, M. J., Eds.), 4th edition, Lippincott Williams & Wilkins Press, New York pp 14-29
[28]. Yehoy, DA, Zhang J, Thraillkill KM, Arthur JM, Quaries LD (1994) Role of serum in the developmental expression of alkaline phosphatase of MC3T3-E1 osteoblasts. J Cell Physiol 158: 467-475
[29]. Uchiyama S, Yamaguchi M (2002) Stimulatory effect of \textit{Sargassum horneri} extract on bone formation in rat femoral-diaphyseal and -metaphyseal tissues in vitro. J Health Sci 48: 148-153
[30]. Uchiyama S, Yamaguchi M (2002) Inhibitory effect of marine alga \textit{Sargassum horneri} extract on bone resorption in tissue culture in vitro. J Health Sci 48: 154-160
[31]. Klein-Nulend J, Fall PM, Raiz LG (1990) Comparison of the effects of synthetic human parathyroid hormone (PTH)-(1-34) related peptide of magnanuine and bovine PTH-(1-34) on bone formation and resorption in organ culture. Endocrinology 126: 223-227
[32]. Graves J III, Jikla R L (1990) Comparison of bone and parathyroid hormone as stimulants of osteoclast development and activity in calvarial cells cultures from normal and osteopetrotic (mi/mi) mice. J Cell Physiol 145: 102-109
[33]. Klein DC, Raizs LG (1970) Stimulation of bone resorption in tissue culture. Endocrinology 86: 1436-1440
[34]. Uchiyama S, Hashizume, M., Hokari, Y., Nakagawa, T., Igarashi, A. & Yamaguchi, M. (2004). Characterization of active component in marine alga \textit{Sargassum horneri} extract in stimulating bone calcification in vitro. J Health Sci 50: 634-639
[35]. Uchiyama S, Yamaguchi M (2001) Anabolic effect of marine alga \textit{Sargassum horneri} extract on bone components in the femoral-diaphyseal and -metaphyseal tissues of young and aged rats in vivo. J Health Sci., 48: 325-330
[36]. Uchiyama S, Yamaguchi M (2003) Preventive effect of marine alga \textit{Sargassum horneri} extract on bone loss in streptozotocin-diabetic rats in vivo. J Health Sci 49: 149-155
[37]. Hui S L., Epstein S, Johanston C C (1985) A prospective study of bone mass in patients with type I diabetes. J Clin. Endocrinol Metab 60: 74-80
[38]. McNair P (1988) Bone mineral metabolism in human type I diabetes melilitus. Dan Med Bull 35: 109-121
[39]. Leslie WD, Rubin MR, Schwartz AV, Kanis JA (2012) Type 2 diabetes and bone. J Bone Miner Res 27:2331-2337
[40]. Matsutomo T, Hokari Y, Hashizume M, Yamaguchi M (2008) Effect of \textit{Sargassum horneri} extract on circulating bone metabolic markers: Supplemental intake has an effect in healthy humans. J Health Sci 54: 50-55
[41]. Levy JR, Murray E, Manolagas S, Olefsky JM (1986) Demonstration of insulin receptors and modulation of alkaline phosphatase activity by insulin

*International Journal of Food Science, Nutrition and Dietetics, 2013 ©*
in rat osteoblastic cells. Endocrinology 119: 1786-1792

[42]. Price PA (1985) Vitamin K-dependent formation of bone gla protein (osteocalcin) and its function. Vitam Horm 42: 65-108

[43]. Hallen JM, Alaralo SL, Suminen H, Cheng S, Jänekila A., &Vaananen, H K (2000) Tartrate-resistant acid phosphatase 5b: A novel serum marker of bone resorption. J Bone Miner Res 15: 1337-1345

[44]. Clements JD, Herrick MV, Singer FR, Eyre DR (1997) Evidence that serum NTx (collagen-type I N-telopeptides) can act as an immunochemical marker of bone resorption. Clin Chem 43: 2058-2063