High-Quality Draft Genome Sequence of the *Xanthomonas translucens* pv. cerealis Pathotype Strain CFBP 2541

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*Xanthomonas translucens* pv. cerealis is the causal agent of bacterial leaf streak on true grasses. The genome of the pathotype strain CFBP 2541 was sequenced in order to decipher mechanisms that provoke disease and to elucidate the role of transcription activator-like (TAL) type III effectors in pathogenicity.

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**Wheat** and other small grain cereals are major crops worldwide and are considered important 4F (food, feed, fiber, and fuel) plants. In human consumption, wheat ranks as the second most-produced crop plant after rice, and wheat is grown on more land area than any other commercial crop (see [http://faostat3.fao.org/home/E](http://faostat3.fao.org/home/E)).

*Xanthomonas translucens* pv. cerealis has been found on crops, like wheat (*Triticum* spp.), barley (*Hordeum* spp.), and rye (*Secale cereale*) (1–3), and it also naturally occurs on smooth brome grass and quack grass (4). Bacterial leaf streak caused by strains of *X. translucens* (5) is the most common bacterial disease of wheat. As a seed-borne disease, it is a constraint for international germplasm exchange (6). The symptoms include translucent stripes at the leaf blade at the early infection state, which later develop into elongated water-soaked lesions, as well as the production of exudates at late infection state (7). While most plant-pathogenic xanthomonads studied thus far belong to the group II clade, the strains of *X. translucens* belong to the group I clade, which also includes the species *Xanthomonas albilineans*, *Xanthomonas yatcinthii*, *Xanthomonas sacchari*, and *Xanthomonas theicola* (8).

Pathotype strain CFBP 2541 (LMG 679, NCPPB 1944) was isolated from *Bromus inermis* in the United States in 1941. We tested this strain on barley (*Hordeum vulgare* L. Morex and Betzes) and wheat (*Triticum aestivum* L. Alondra) under laboratory conditions. Strong symptoms were obtained with the “Morex” and “Alondra” plants, while “Betzes” remained symptomless.

To obtain new insights into the molecular determinants provoking disease or resistance, we sequenced strain CFBP 2541 using the Illumina HiSeq 2000 platform (GATC Biotech, Germany). The shotgun sequencing yielded 59,447,151 read pairs (263,377,209 100-bp paired-end reads, with an insert size of 250 bp, and 33,109,942 50-bp mate-pair reads, with an insert size of 3 kb). A combination of Velvet (9), SOAPdenovo, and SOAPGapCloser (10) yielded 31 contigs >500 bp (N50, 1,399,657 bp), with the largest contig being 1,809 kb, for a total assembly size of 4,515,938 bp, corresponding to 1,926× coverage.

The genome was found to encode a noncanonical hypersensitive response and pathogenicity (Hrp) type III protein secretion system, the genetic organization of which differs from that of clade II xanthomonads, as previously reported for *X. translucens* pv. graminis strain Xtg29 (11). In contrast to strain Xtg29, however, the genome assembly of strain CFBP 2541 indicated the presence of two type III transcription activator-like (TAL) effector genes (12, 13), which was supported by Southern blot hybridization. Since tal genes are notoriously difficult to be assembled from short reads due to their repetitive nature, we sequenced the tal genes upon PCR amplification. Surprisingly, the two genome-assembled tal genes turned out to be correctly assembled, probably due to the very high coverage and a significant number of single-nucleotide polymorphisms (on average, 1 per 10 bp) that distinguish all individual repeats from each other. This information opens the way for studying the role of tal genes in the pathogenicity of *X. translucens*.

**Nucleotide sequence accession numbers.** This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession no. JWHD00000000. The version described in this paper is the first version, JWHD01000000.

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Strain CFBP 2541 is available at the CIRM-CFBP, French Collection for Plant-Associated bacteria (http://www6.inra.fr/cirm_eng/CFBP-Plant-Associated-Bacteria).
REFERENCES
1. Rademaker JL, Norman DJ, Forster RL, Louws FJ, Schultz MH, de Bruijn FJ. 2006. Classification and identification of Xanthomonas translucens isolates, including those pathogenic to ornamental asparagus. Phytopathology 96:876–884. http://dx.doi.org/10.1094/PHYTO-96-0876.
2. Wagoner P, Schauer A. 1990. Intermediate wheatgrass as a perennial grain crop, p 143–145. In Janick J, Simon JE (ed), Advances in new crops. Timber Press, Portland, OR.
3. Bragar C, Singer E, Alizadeh A, Vauterin L, Maraite H, Swings J. 1997. Xanthomonas translucens from small grains: diversity and phytopathological relevance. Phytopathology 87:1111–1117. http://dx.doi.org/10.1094/PHYTO.1997.87.11.1111.
4. Fang CT, Allen ON, Riker AJ, Dickson JG. 1950. The pathogenic, physiological and serological reactions of the form species of Xanthomonas translucens. Phytopathology 40:44–64.
5. Vauterin L, Hoste B, Kersters K, Swings J. 1995. Reclassification of Xanthomonas. Int J Syst Bacteriol 45:472–489. http://dx.doi.org/10.1099/00207713-45-3-472.
6. Duveiller E, Bragar C, Rudolph K, Fucikovsky L. 1997. The bacterial diseases of wheat: concepts and methods of disease management, p 78. International Maize and Wheat Improvement Center, Estado de México, Mexico.
7. Smith EF. 1917. A new disease of wheat. J Agric Res 10:51–54.
8. Young JM, Park DC, Shearman HM, Fargier E. 2008. A multilocus sequence analysis of the genus Xanthomonas. Syst Appl Microbiol 31:366–377. http://dx.doi.org/10.1016/j.syapm.2008.06.004.
9. Zerbino DR, Birney E. 2008. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. Genome Res 18:821–829. http://dx.doi.org/10.1101/gr.074492.107.
10. Luo R, Liu B, Xie Y, Li Z, Huang W, Yuan J, He G, Chen Y, Pan Q, Liu Y, Tang J, Wu G, Zhang H, Shi Y, Liu Y, Yu C, Wang B, Lu Y, Han C, Cheung DW, Yiu SM, Peng S, Xiaoqian Z, Liu G, Liao X, Li Y, Yang H, Wang J, Lam TW, Wang J. 2012. SOAPdenovo2: an empirically improved memory-efficient short-read de novo assembler. GigaScience 1:18. http://dx.doi.org/10.1186/2047-217X-1-18.
11. Wichmann F, Vorhölter FJ, Hersemann L, Widmer F, Blom J, Niehaus K, Reinhard S, Conradin C, Kolliker R. 2013. The noncanonical type III secretion system of Xanthomonas translucens pv. graminis is essential for forage grass infection. Mol Plant Pathol 14:576–588. http://dx.doi.org/10.1111/mpp.12030.
12. Bragar C, Vérardier V, Maraite H. 1995. Genetic diversity among Xanthomonas campestris strains pathogenic for small grains. Appl Environ Microbiol 61:1020–1026.
13. Boch J, Bonas U. 2010. Xanthomonas AvrBs3 family-type III effectors: discovery and function. Annu Rev Phytopathol 48:419–436. http://dx.doi.org/10.1146/annurev-phyto-080508-081936.