Histology of Interstitial Lung Disease in Common Variable Immune Deficiency

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Interstitial lung disease (ILD) is an important non-infectious complication in several primary immune deficiencies. In common variable immune deficiency (CVID) it is associated with complex clinical phenotypes and adverse outcomes. The histology of ILD in CVID is heterogeneous and mixed patterns are frequently observed within a single biopsy, including non-necrotising granulomatous inflammation, lymphoid interstitial pneumonitis, lymphoid hyperplasia, follicular bronchiolitis, organizing pneumonia, and interstitial fibrosis; ILD has to be differentiated from lymphoma. The term granulomatous-lymphocytic interstitial lung disease (GLILD), coined to describe the histopathological findings within the lungs of patients with CVID with or without multisystem granulomas, is somewhat controversial as pulmonary granulomata are not always present on histology and the nature of infiltrating lymphocytes is variable. In this mini review we summarize the literature on the histology of CVID-related ILD and discuss some of the factors that may contribute to the inter- and intra-patient variability in the histological patterns reported. Finally, we highlight areas for future development. In particular, there is a need for standardization of histological assessments and reporting, together with a better understanding of the immunopathogenesis of CVID-related ILD to resolve the apparent heterogeneity of ILD in this setting and guide the selection of rational targeted therapies in different patients.

Keywords: common variable immune deficiency, interstitial lung disease, histology review, literature analysis, primary immune deficiencies

INTRODUCTION

Common variable immune deficiency (CVID) is the most common of the primary immunodeficiency (PID) syndromes with a prevalence of 1 in 25,000 and 50,000, depending on the population (1, 2). It is characterized by low serum levels of IgG, IgA, and/or IgM, and poor specific antibody production (3). There is no definitive diagnostic test, so diagnosis requires the exclusion of secondary hypogammaglobulinaemia, combined immune defects, and, where appropriate, Mendelian disorders (4, 5). Up to 70% of patients suffer with variable non-infectious complications reflecting broader immune dysregulation, including autoimmunity, most commonly autoimmune cytopenias; lymphocytic infiltration and/or granulomatous inflammation.
which can affect the lungs, gastrointestinal tract, spleen, skin or liver; or malignancy, in particular lymphoma (6, 7). Importantly, while bacterial infections are significantly reduced by adequate replacement therapeutic IgG, disease-related complications are not, but are associated with substantially increased mortality (7–9).

Respiratory tract pathology is a major contributor to impaired quality-of-life (10). Bacterial sinopulmonary infections are often the presenting feature, most frequently caused by *Haemophilus influenzae* or *Streptococcus pneumoniae* (11, 12). Recurrent and/or severe lower respiratory tract infections, particularly pneumonia, lead to bronchiectasis with an overall estimated prevalence of 30–35% among CVID patients, which, when present in isolation, does not contribute to increased mortality (8, 11–14). Interstitial lung disease (ILD), on the other hand, probably occurs due to immune dysregulation and/or viral infection rather than as a consequence of bacterial infection (7, 15, 16), and occurs alongside other disease-related complications, and shortens survival (7–9, 16). More rarely, the lungs can be the location for extranodal lymphomas, particularly B-cell non-Hodgkin’s lymphomas or MALToma (7, 17–20).

**INTERSTITIAL LUNG DISEASE IN COMMON VARIABLE IMMUNE DEFICIENCY**

**Clinical Significance of CVID-Related ILD**

ILD is among the more frequent non-infectious complications of CVID, reported in 15%–60% of patients (7, 9, 14, 21–23). Clinical symptoms and high-resolution computed tomography (HRCT) findings of ILD can appear before or after CVID diagnosis (24, 25). The pathogenesis of CVID-related ILD is presumed to be unrelated to bacterial infections because it can be seen in the absence of bronchiectasis and is not significantly associated with a history of pneumonia (21). Patients with ILD have distinct clinical and immunological phenotypes in keeping with immune dysregulation, in contrast to those without ILD or those with bronchiectasis alone (6, 9, 14, 16, 21, 26, 27). Furthermore, there is no current histological or molecular evidence for chronic bacterial, EBV or CMV viral infections as triggers for inflammation (16, 28–30), though granulomas in other PIDs, such as those with DNA repair defects, show evidence of vaccine derived rubella virus (31). Other related complications, including splenomegaly, autoimmune cytopaenias, persistent lymphadenopathy and lymphoproliferation, but not necessarily granulomata, occur more frequently in patients with CVID-related ILD, supporting at least a role for intrinsic immune dysregulation driving these varied features (6, 9, 16, 21, 27, 32, 33).

Since CVID-related ILD causes significant morbidity, can be progressive and contributes to mortality, there is urgent need for effective treatments (8, 9, 34, 35). Because the mechanisms underlying CVID ILD have not been elucidated, immunosuppressive treatments have been tried with varying success, including corticosteroids, cyclosporin, methotrexate, sirolimus, cyclophosphamide, hydroxychloroquine, anti-TNF agents, mycophenolate mofetil, abatacept, rituximab and azathioprine (16, 34, 36–38). Corticosteroids are often used first-line, however, response may be short-lived or incomplete, there are significant side effects associated with protracted use and a proportion of patients are refractory (16, 34, 36, 39). Success with Rituximab, both in combination with azathioprine or mycophenolate mofetil, and as monotherapy, has been reported although controlled trials and long-term outcome data are lacking (40–43). Elevated levels of B-cell activating factor (BAFF), a cytokine that promotes the maturation and survival of B-cells, within the serum and lungs of patients with CVID-related ILD levels drives B-cell hyperplasia and may account for disease progression in a small proportion of patients (15) with invasive B cells in inappropriate germinal centers (28, 44).

**Nomenclature**

Various terminologies are used for CVID-related ILD, reflecting a lack of consensus regarding the naming of this complication and its heterogeneous nature (45). Lymphoid interstitial pneumonitis was first reported in patients with antibody deficiency in 1973 (46). Since then, various histopathological entities have been reported within lung biopsies of CVID ILD patients, from those caused by polyclonal lymphocytic inflammation to well-formed granulomata, organizing pneumonia, or pulmonary fibrosis, often with mixed pathology within individual patient biopsies (7, 9, 16, 27, 33, 35, 44). “Granulomatous-lymphocytic interstitial lung disease” (GLILD), first proposed in 2004, is often used as an overarching term to describe CVID ILD with lymphocytic infiltrates and/or granulomata (9, 45). However, the accuracy of this term has been called into question. Since not all patients have pulmonary granulomata, it does not fully capture the heterogeneity of the histopathology and similar histological patterns fulfilling a GLILD diagnosis are found in non-CVID PIDs (33, 47).

**Investigations for CVID-Related ILD**

Non-invasive investigations for CVID-related ILD include elevated serum IgM, decreased class-switched memory B-cells and absolute/relative numerical abnormalities of T-cell populations (15, 16, 34, 35, 48). Alongside rising IgM levels, BAFF, soluble IL-2 receptor and β2microglobulin have also been proposed as serum biomarkers for disease activity (15, 34, 49). Lung function tests, particularly the diffusion capacity for carbon monoxide (DLCO), are useful in monitoring for disease progression and response to treatment, but can lack the sensitivity required for diagnosis, particularly early in the disease course (14, 28, 34, 35, 37). HRCT is highly sensitive for the detection of CVID ILD, including at an early stage before symptoms or abnormal pulmonary function have developed (14, 33, 34). Radiographic findings are mixed and include lymphadenopathy, ground glass opacification, nodularity, septal thickening and consolidation (21, 33, 50). The use of CT combined with positron emission technology (PET) has also been reported as useful to identify sites of active disease, guide biopsy sampling, and monitor response to treatment (41). In selected cases, particularly, but not restricted to, pediatric presentations, genetic testing may be warranted. For example, patients with mutations in *CTLA4*, *LRBA*, *TACI*, *KMT2D*, *XIAP*, *RAG1*, and *NFKB1* have been found within so called “CVID”
HISTOLOGICAL PATTERNS OF ILD IN CVID

The histological abnormalities reported in CVID ILD vary and overlap extensively. Similar patterns can also be found in numerous other lung diseases, making diagnosis challenging (44). Using a similar structure as Rao et al. (44), we summarize the commonly reported lung biopsy findings, each of which we discuss in turn (Table 1).

Granulomata

The granulomata reported in CVID ILD can vary from poorly-to well-circumscribed, with an apparent predilection for the former (28, 33, 44). Non-infectious CVID granulomatous lung disease shares some similar histological features with sarcoidosis and hypersensitivity pneumonitis; thus, clinical and radiological correlation is important in distinguishing these conditions (44, 62). “Poorly-formed granulomata” have been found within areas of pulmonary lymphoid hyperplasia and are difficult to define, as these are very subjective; additionally, granulomata can be found throughout the lung parenchyma (28, 44). It is worth re-emphasizing that granulomata are not reported in all cases of CVID-related ILD, with frequencies ranging from 0-94% depending on the individual study (Table 1) (7, 33, 44, 47). This suggests that there may be more than one pathological process in CVID-ILD (33, 47) and that the generalized use of overarching term “GLILD” to refer to all CVID-related ILD can be misleading.

Pulmonary Lymphoid Hyperplasia

Lymphoid proliferation has been designated as the “cardinal” feature of CVID ILD, and different patterns of pulmonary lymphoid hyperplasia (PLH) have been described, including follicular bronchiolitis, lymphocytic interstitial pneumonitis (LIP), lymphocytic infiltrates, and nodular lymphoid hyperplasia (28, 38, 40, 44, 47). In one case series where severity was assessed, PLH tended to be toward the moderate to severe end of the spectrum,

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**TABLE 1 | Histological lung biopsy findings from common variable immune deficiency (CVID) patients reported in the literature.**

| Publication (Ref) | Number of CVID patients with lung biopsies | Granulomata (%) | Pulmonary Lymphoid Hyperplasia | Organizing pneumonia | Pulmonary Fibrosis | Remodeling |
|-------------------|-------------------------------------------|-----------------|-------------------------------|---------------------|-------------------|------------|
| Rao et al. (44)   | 16                                        | 15 (93%)        | 16 (100%)                     | NS                  | NS                | 14 (87%)   |
| Patel et al. (33) | 19                                        | 1 (5%)          | 11 (58%)                      | 7 (37%)             | NS                | 6 (32%)    |
| Maglione et al. (21) | 12                                      | 3 (25%)        | 4 (33%)                       | 2 (17%)             | 4 (33%)           | 3 (16%)    |
| Larsen et al. (47) | 34                                        | 23 (68%)        | 12 (35%)                      | 22 (65%)            | 10 (29%)          | 25 (71%)   |
| Verbsky et al. (61) | 34                                        | 31/34 (91%)    | NS                            | 33/34 (97%)         | 33/34 (97%)       | 30/34 (88%) |

Only publications with sufficient histological detail were included; single case histories or small studies (less than 10) are not included. Rao et al. (44) and Patel et al. (33) reported their findings in similar terms, but these varied in other publications. Efforts were made to group similar findings on the basis on similar histological terms in these instances. Where detail for a given finding was not specified (NS), this is also indicated. *Where the inclusion of previously published cases in a paper could not be completely excluded. ** On CT not reported on histology.
with peribronchiolar and interstitial lymphocytic inflammation (44). These patterns often occur together and are rarely found in isolation (33, 44). Follicular bronchiolitis and/or LIP are found in around half of the cases reviewed (Table 1), and this is also in keeping with a recent review where 20/46 patients had some form of lymphoid infiltration, though not always specified (7).

Organizing Pneumonia
Organizing pneumonia (OP), intra-alveolar buds of granulation tissue with myofibroblasts and connective tissue, is reported in a substantial number of histological specimens, although to varying degrees between studies (Table 1). Cryptogenic organizing pneumonia (COP) is also found in CVID patients and is an important differential diagnosis when OP is the predominant finding on biopsy (40, 44). However, Rao et al. demonstrated the potential for misdiagnosis of CVID ILD when isolated COP was found on limited biopsy samples obtained by bronchoscopy.

OP can have many aetiologies. Larsen et al. reported that in their cohort OP was accompanied by a “dense lymphoid infiltrate”, which was not seen in biopsies from other causes of OP (47). Therefore, in their cohort of 34 patients with CVID and 4 with IgAD, these authors suggest that the combination of these two findings should suggest CVID or IgA deficiency rather than another etiology.

The lack of overlap between OP and pulmonary fibrosis (1/19 cases) in our cases might indicate separate pathological entities; however, significant overlap was described by Rao et al. (11/16 cases) (33, 44), who suggested evolving pathology.

Pulmonary Fibrosis
Pulmonary fibrosis is described in a quarter of CVID ILD cases (Table 1); however, similar to OP, one case series accounts for most of these cases (44), where the majority of patients had some degree of fibrosis. In contrast, Ho et al. found 6.3% of cases where “extensive pulmonary fibrosis” was the “predominant” finding at the time of biopsy; however, it was not reported whether it was a feature in other biopsies to a lesser degree (7).

Interstitial fibrosis in CVID ILD together with lymphoproliferation may resemble some of the patterns of idiopathic interstitial pneumonia, particularly if significant fibrosis (44). Only two studies looked specifically for architectural remodeling, and one of these found this to be associated with significant interstitial fibrosis (33, 44). The presence of fibrosis is a poor prognostic factor; prospective clinical studies are needed to justify earlier treatment (33).

Immunohistochemistry
Immunohistochemical staining of the lymphocytic infiltrate has produced discordant findings in the cases where it has been performed. CD20+ B-cells were found in a small proportion of cases, in follicles with T-cells circumscribing them, although T-cells are also reported more diffusely and in areas without B-cells (28, 33, 44). Rao et al. found a predominance of CD4+ T-cells within lymphoid infiltrates and also observed the presence of B-cell follicles surrounded by CD4+ T-cells (44). We recently reported a predominance of T-cells in most cases (Figure 1A), either CD4+ or CD8+; only 1 of 6 had germinal centers within B-cell follicles (Figure 1B) (33). Maglione et al. reported actively proliferating germinal centers in some of their patients with B-cell follicles (28). It is important to differentiate these from pulmonary MALToma, as found in two patients in the Oxford series (33).

We suggested that since the predominant T-cells were either CD4+ or CD8+, this pointed to different pathological entities (33). Chase et al. hypothesized that the inflammatory infiltrate, including B- and T-cells, might contribute to progressive ILD and pulmonary fibrosis, something that therapy directed against B- and T-cells might possibly prevent (40). Similarly, Maglione et al. suggested B-cells may be responsible for leukocyte accumulation in their role as antigen presenting cells and producers of chemokines and/or cytokines, making them a therapeutic target (28).

ADDRESSING THE HETEROGENEITY OF HISTOPATHOLOGICAL FINDINGS CVID-RELATED ILD
There is a large amount of histopathological heterogeneity in biopsies from CVID-related ILD cases, both from one patient to the next, as well as between different case reports (Table 1). We discuss possible reasons for this in respect to the underlying pathophysiology, the patient populations reported, and factors relating to obtaining and interpreting lung biopsies.

Pathophysiology: A Spectrum of Disease, Separate Diseases, or a Shared Endpoint for Several Diseases?
Since the pathophysiology of CVID ILD is unknown, it is not surprising that there is no explanation for the degree of heterogeneity in the histology (33, 44). CVID-related ILD (or GLILD) was originally defined as a “conglomeration of pulmonary histopathologic abnormalities seen in a subset of patients with CVID (44). The divergent findings may represent a “spectrum” of a single disease (44) or several different pathologies, in addition to the primary antibody deficiency. Another hypothesis is that CVID ILD represents a common “pulmonary reaction pattern” (or “morphological common endpoint”) not only for CVID but also for other PIDs in which similar clinical, radiographical, and histological features have been described (44, 47). None of these hypotheses are mutually exclusive; it may be that the small numbers and the absence of international standardization frustrate the recognition of distinct pathological patterns.

Patient Populations
Geography may influence the variability observed, with different genetic influences in particular populations. It is interesting that three of the large CVID-related ILD case series, one from the UK and two from the USA, show the most divergence, despite a conscious effort on the part of the former to adhere to similar definitions used previously.
Differences in clinical practice, including diagnosis, cannot be totally discounted. Some series are restricted to patients with spontaneous (non-familial) CVID in adults and others include patients diagnosed in childhood. Since no diagnostic details are given, the exclusion of combined immune deficiencies involving T-cell immunity as well as B-cell failure (5), or known mutations in monogenic disease (e.g. CTLA4, LRBA, KMT2D, XIAP, RAG1, NFKB1) (34, 39, 51–57, 63) is unclear.

Biopsy-Related Factors: Technique, Timing, Treatment, and Interpretation

The method by which a biopsy has been obtained may have a significant impact on the clinical conclusions reached (61). Given that several different biopsy techniques have been used across the cases reported, this may be a contributing factor to some of the variation between cases, though in almost all series so far, imaging was used to obtain the biopsy.

A further consideration is the timing of the biopsy with respect to disease progression but most patients do not undergo repeat biopsies. It is likely that once pulmonary fibrosis and possibly organizing pneumonia are present that these may progress (33).

Another potential contributing factor is whether the biopsy was performed prior to or following corticosteroid or immunosuppressive treatment. These drugs could plausibly alter the patterns observed or mask them entirely, particularly those related to inflammation. While some authors have clearly documented when such drugs were used before biopsies were performed (33), this is not always the case, so firm conclusions cannot be drawn.

In the absence of standardized reporting, reading of the biopsy adds a great deal of potential for variation to be introduced. Although some authors have tried to mirror the approach pioneered by others and/or have a second, independent pathologist review the histology, some degree of both intra- and inter-operator variability is inevitable when faced with an uncommonly encountered pathological entity (33, 40).

CONCLUSIONS AND FUTURE DIRECTIONS

In summary, there is considerable heterogeneity in the histopathological findings both within individual patients,
AUTHOR CONTRIBUTIONS

FD and DM prepared the first draft of the manuscript. All authors contributed to editing of subsequent versions and reviewed and authorized the final version. HC and SP played a supervisory role. All authors contributed to the article and approved the submitted version.

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REFERENCES

1. Shillitoe B, Bangs C, Guzman D, Gennery AR, Longhurst HJ, Slatter M, et al. The United Kingdom Primary Immunodeficiency (UKPID) registry 2012 to 2017. Clin Exp Immunol (2018) 192(3):284–91. doi: 10.1111/cei.13125

2. Westh L, Mogensen TH, Dalgaard LS, Bernth Jensen JM, Katzenstein T, Hansen AE, et al. Identification and Characterization of a Nationwide Danish Adult Common Variable Immunodeficiency Cohort. Scand J Immunol (2017) 85(6):450–61. doi: 10.1111/sj.i.12551

3. Seidel MG, Kindle G, Gathmann B, Quinti I, Buckland M, van Montfrans J, et al. The European Society for Immunodeficiencies (ESID) Registry Working Definitions for the Clinical Diagnosis of Inborn Errors of Immunity. J Allergy Clin Immunol Pract (2019) 7(6):1763–70. doi: 10.1016/j.jaip.2019.02.004

4. Bertinchamp R, Gerard L, Boutboul D, Malphettes M, Fieschi C, Oksenhendler E, et al. Exclusion of Patients with a Severe T-Cell Defect Improves the Definition of Common Variable Immunodeficiency. J Allergy Clin Immunol Pract (2016) 4(6):1147–57. doi: 10.1016/j.jaip.2016.07.002

5. Chapel H. Common Variable Immunodeficiency Disorders (CVID) - Diagnoses of Exclusion, Especially Combined Immune Defects. J Allergy Clin Immunol Pract (2016) 4(6):1158–9. doi: 10.1016/j.jaip.2016.09.006

6. Chapel H, Lucas M, Lee M, Bjorkander J, Webster D, Grimbacher B, et al. Common variable immunodeficiency disorders: division into distinct clinical phenotypes. Blood (2008) 112(2):277–86. doi: 10.1182/blood-2007-11-124545

7. Ho HE, Cunningham-Rundles C. Non-infectious Complications of Common Variable Immunodeficiency: Updated Clinical Spectrum, Sequelae, and Insights to Pathogenesis. Front Immunol (2020) 11:149. doi: 10.3389/fimmu.2020.00149

8. Resnick ES, Mosher EL, Godbold JH, Cunningham-Rundles C. Morbidity and mortality in common variable immune deficiency over 4 decades. Blood (2012) 119(7):e1650–7. doi: 10.1182/blood-2011-09-377945

9. Bates CA, Ellison MC, Lynch DA, Cool CD, Brown KK, Routes JM. Granulomatous-lymphocytic lung disease shortens survival in common variable immunodeficiency. J Allergy Clin Immunol (2004) 114(2):415–21. doi: 10.1016/j.jaci.2004.05.057

10. Hurst JR, Workman S, Garcha DS, Seneviratne SL, Haddock JA, Grimbacher B. Activity, severity and impact of respiratory disease in primary antibody deficiency syndromes. J Clin Immunol (2014) 34(4):68–75. doi: 10.1007/s10875-013-9942-x

11. Oksenhendler E, Gerard L, Fieschi C, Malphettes M, Moullet G, Jaussaud R, et al. Infections in 252 patients with common variable immunodeficiency. Clin Infect Dis (2008) 46(10):1547–54. doi: 10.1086/587669

12. Sperlich JM, Grimbacher B, Workman S, Haque T, Seneviratne SL, Burns SO, et al. Respiratory Infections and Antibiotic Usage in Common Variable Immunodeficiency. J Allergy Clin Immunol Pract (2018) 6(1):139–68. doi: 10.1016/j.jaip.2017.05.024

13. Ramzi N, James M, Bakhtiyari M, Rafifemanesh H, Zaialldan H, Tavakol M, et al. Bronchiectasis in common variable immunodeficiency: A systematic review and meta-analysis. Pediatr Pulmonol (2020) 55(2):292–9. doi: 10.1002/ppul.24599

14. Maarschalk-Ellerbroek LJ, de Jong PA, van Montfrans JM, Lammers JW, Bloem AC, Hoepelman AI, et al. CT screening for pulmonary pathology in common variable immunodeficiency disorders and the correlation with clinical and immunological parameters. J Clin Immunol (2014) 34(6):642–54. doi: 10.1007/s10875-014-0068-6

15. Maglione PJ, Gyimesi G,Cols M, Radigan L, Ko HM, Weinberger T, et al. BAFF-driven B cell hyperplasia underlies lung disease in common variable immunodeficiency. JCI Insight (2019) 4(5). doi: 10.1172/jci.insight.122728

16. Schussler E, Beasley MB, Maglione PJ. Lung Disease in Primary Antibody Deficiencies. J Allergy Clin Immunol Pract (2016) 4(6):1039–52. doi: 10.1016/j.jaip.2016.08.005

17. Reichenberger F, Wyser C, Gonon M, Cathomas G, Tamm M. Pulmonary mucosa-associated lymphoid tissue lymphoma in a patient with common variable immunodeficiency syndrome. Respiration (2001) 68(1):109–12. doi: 10.1159/000050475

18. Aghamohammadi A, Parvaneh N, Tirgari F, Movahedi M, Ghargozliou M, et al. Lymphoma of mucosa-associated lymphoid tissue in common variable immunodeficiency. J Hematol Am Soc Hematol Educ Program (2012) 2012:301. doi: 10.1182/asheducation.V2012.1.301.3798316

19. Cunningham-Rundles C, Cooper DL, Duffy TP, Strauchen J, Cunningham-Rundles C. The many faces of common variable immunodeficiency: A systematic review and meta-analysis. Am J Hematol (2002) 69(3):171–8. doi: 10.1002/ajh.10050

20. Cunningham-Rundles C. The many faces of common variable immunodeficiency. Hematol Am Soc Hematol Educ Program (2012) 2012:301–5. doi: 10.1182/asheducation.V2012.1.301.3798316

21. Maglione PJ, Overyer JR, Radigan L, Bagiella E, Cunningham-Rundles C. Pulmonary radiologic findings in common variable immunodeficiency: clinical and immunological correlations. Ann Allergy Asthma Immunol (2014) 113(4):452–9. doi: 10.1016/j.anai.2014.04.024

22. Verma N, Grimbacher B, Hurst JR. Lung disease in primary antibody deficiency. Lancet Respir Med (2015) 3(8):651–60. doi: 10.1016/S2213-2600(15)00202-7
31. Buchbinder D, Hauck F, Albert MH, Rack A, Bakhtiar S, Shcherbina A, et al. Predictors of granulomatous lymphocytic interstitial lung disease in common variable immunodeficiency. Allergy Asthma Immunol (2017) 118(5):614–20. doi: 10.1016/j.jai.2017.02.004

32. Bondioni MP, Paolini MV, Fernández Romero DS. Lung disease in patients with common variable immunodeficiency. Allergologia Immunopathol (2020) 48(1):25–30. doi: 10.1007/s10875-019-00708-2

33. Patel S, Anzilotti C, Lucas M, Moore N, Chapel H. Interstitial lung disease in common variable immunodeficiency. Ann Allergy Asthma Immunol (2017) 118(5):614–20. doi: 10.1016/j.jai.2017.02.004

34. Baumann U, Routes JM, Soler-Palacin P, Jolles S. The Lung in Primary Immune Deficiency Disorders (CVID): a case report. Immunodeficiency. A Multicenter Observational Study. J Allergy Clin Immunol Pract (2017) 5(4):938–45. doi: 10.1016/j.jaip.2017.01.021

35. Park JH, Levinson AL. Granulomatous-lymphocytic interstitial lung disease (GLILD) in patients with common variable immunodeficiency (CVID). J Clin Immunol (2013) 33(1):84–95. doi: 10.1007/s10875-012-9778-9

36. Boursiquot JN, Gerard L, Malphettes M, Fieschi C, Galicier L, Boutboul D, et al. Interstitial Lung Disease Frequently Precedes CVID Diagnosis. J Clin Immunol (2013) 33(1):84–95. doi: 10.1007/s10875-012-9778-9

37. Hurst JR, Verma N, Lowe D, Baxendale HE, Jolles S, Kelleher P, et al. Bronchopulmonary Lesions in Common Variable Immunodeficiency: A Spectrum of Pulmonary Histopathologic Lesions in Common Variable Immunodeficiency–histologic and immunohistochemical analyses of 16 cases. Hum Pathol (2015) 46(10):1306–14. doi: 10.1016/j.humpath.2015.05.011

38. Wehr C, Kivioja T, Schmitt C, Ferry B, Witte T, Eren E, et al. The EUROdast trial: defining subgroups in common variable immunodeficiency. Blood (2018) 131(1):77–85. doi: 10.1182/blood-2007-06-091744

39. Vitale J, Conovers KD, Goretzke S, Guzman M, Noyes B, Parkar N, et al. Serum IL-12 and soluble IL-2 receptor levels as possible biomarkers of granulomatous and lymphocytic interstitial lung disease in common variable immunodeficiency: a case report. J Allergy Clin Immunol Pract (2015) 3(2):273–6. doi: 10.1016/j.jaip.2014.09.019

40. Bousiquot JN, Gerard L, Malphettes M, Fieschi C, Galicier L, Boutboul D, et al. Granulomatous disease in CVID: retrospective analysis of clinical characteristics and treatment efficacy in a cohort of 59 patients. J Clin Immunol (2013) 33(1):84–95. doi: 10.1007/s10875-012-9778-9

41. Leeflang MM, El-Shanawany T, Williams P, Marshall C, et al. FDG PET-CT imaging of therapeutic response in granulomatous lymphocytic interstitial lung disease (GLILD) in common variable Immunodeficiency (CVID). J Clin Exp Immunol (2017) 187(1):138–45. doi: 10.1111/cei.12856

42. Buchbinder D, Baker R, Lee VN, Ravel J, Zhang Y, McEehwey E, et al. Identification of patients with RAG mutations previously diagnosed with common variable immunodeficiency disorders. J Clin Immunol (2015) 35(2):119–24. doi: 10.1007/s10875-014-0211-5
57. Tuijnenburg P, Lango Allen H, Burns SO, Greene D, Jansen MH, Staples E, et al. Loss-of-function nuclear factor κB subunit 1 (NFKB1) variants are the most common monogenic cause of common variable immunodeficiency in Europeans. J Allergy Clin Immunol (2018) 142(4):1285–96. doi: 10.1016/j.jaci.2018.01.039

58. Schwab C, Gabrysch A, Olbrich P, Patino V, Warnatz K, Wolff D, et al. Phenotype, penetrance, and treatment of 133 cytotoxic T-lymphocyte antigen 4-insufficient subjects. J Allergy Clin Immunol (2018) 142(6):1932–46. doi: 10.1016/j.jaci.2018.02.055

59. Lo B, Zhang K, Lu W, Zheng L, Zhang Q, Kanellopoulou C, et al. AUTOIMMUNE DISEASE. Patients with LRBA deficiency show CTLA4 loss and immune dysregulation responsive to abatacept therapy. Science (2015) 349(6246):436–40. doi: 10.1126/science.aaa1663

60. Gregersen S, Holm AM, Fevang B, Ueland T, Sikkeland LI, Aalokken TM, et al. Lung disease, T-cells and inflammation in common variable immunodeficiency disorders. Scand J Clin Lab Invest (2013) 73(6):514–22. doi: 10.3109/00365513.2013.819523

61. Verbisky JW, Hintermeyer MK, Simpson PM, Feng M, Barbeau J, Rao N, et al. Rituximab and antimetabolite treatment of granulomatous and lymphocytic interstitial lung disease in common variable immunodeficiency. J Allergy Clin Immunol (2020) S0091-6749(20)31069-1. doi: 10.1016/j.jaci.2020.07.021

62. Bouvy D, Mouthon L, Brillet PY, Kambouchner M, Ducroix JP, Cottin V, et al. Granulomatosis-associated common variable immunodeficiency disorder: a case-control study versus sarcoidosis. Eur Respir J (2013) 41 (1):115–22. doi: 10.1183/09031936.00189011

63. Sood AK, Funkhouser W, Handly B, Weston B, Wu EY. Granulomatous-Lymphocytic Interstitial Lung Disease in 22q11.2 Deletion Syndrome: A Case Report and Literature Review. Curr Allergy Asthma Rep (2018) 18(3):14. doi: 10.1007/s11882-018-0769-7

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