Complete Genome Sequence of *Paenibacillus* sp. Strain VT 400, Isolated from the Saliva of a Child with Acute Lymphoblastic Leukemia

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We report here the complete genome sequence of spore-forming *Paenibacillus* sp. strain VT 400, isolated from the saliva of a child with acute lymphoblastic leukemia. The genome consists of 6,986,122 bp, with a G+C content of 45.8%. It possesses 5,777 predicted protein-coding genes encoding multidrug resistance transporters, virulence factors, and resistance to chemotherapeutic drugs.

The genus *Paenibacillus* comprises aerobic or facultatively anaerobic, rod-shaped, immotile or motile (via peritrichous flagella), endospore-forming bacteria. Species of this genus have been isolated from soil, water, plants, milk, and other media (1–3). *Paenibacillus* spp. were not known to cause human disease until recent reports showed the possible involvement of *P. alvei*, *P. thiominolyticus*, and *P. putida* in human diseases (4–6).

*Paenibacillus* sp. strain VT 400 was isolated from the saliva of a pediatric patient with acute lymphoblastic leukemia. Complete sequencing of its 16S rRNA gene showed 99% similarity with that of *Paenibacillus amylolyticus*, a pectinase producer found in the larval hindgut of the fly. To date, *P. amylolyticus* has not been detected in humans. Biochemical characterization and matrix-assisted laser desorption ionization–time of flight (MALDI-TOF) mass spectrometry have revealed differences between *Paenibacillus* sp. strain VT 400 and other *P. amylolyticus* strains.

Whole-genome sequencing was performed using the Illumina HiSeq 2500 sequencer, according to the manufacturer’s instructions (Illumina GA IIx; Illumina, CA). *De novo* assembly was performed with the SPAdes genome assembler software, version 3.5.0 (7). The assembly resulted in 116 contigs with 125-fold average coverage.

The draft genome of *Paenibacillus* sp. strain VT 400 consists of 6,986,122 bp, with a G+C content of 45.8%. The genome was annotated using the NCBI Prokaryotic Genome Annotation Pipeline (8). It contains 100 RNA genes, 50 rRNA and two noncoding RNA operons, and 5,777 protein-coding sequences (CDSs). The analysis revealed genes encoding resistance to chemotherapeutic drugs, like tunicamycin and bleomycin (9, 10). Furthermore, genes of multidrug resistance (MDR) efflux pumps (ABC transporter, a member of the multidrug and toxin extrusion [MATE] family of MDR proteins, and an MDR transporter) and genes encoding resistance to antibiotics, including vancomycin (*vanH*, *vanX*, *vanW*, and *vanZ*), fosmidomycin, tetracycline (*tetA*, *tetB*), as well as beta-lactamase genes from superfamilies I to III, were identified. The genome contains genes encoding known virulence factors, like hemolysin D and flagellar and sporulation proteins (11).

In comparison with the genome of *P. amylolyticus* FSL-R5-192 (the phylogenetically closest organism), that of *Paenibacillus* sp. strain VT 400 is smaller (6,986,122 versus 7,083,071 bp) and contains fewer protein-coding genes (5,777 versus 6,163), although the two genomes have a similar G+C content (45.8%). According to DNA-DNA hybridization (DDH) prediction based on genome BLAST distance phylogeny (GBDP), this pair of genomes (*Paenibacillus* sp. strain VT 400 and *P. amylolyticus* FSL-R5-192) has a DDH value of 74.40%, as calculated in the Genome-to-Genome Distance Calculator (GGDC) Web server (GGDC 2.0). This value is below the threshold of 79% for genomes belonging to different subspecies (12, 13).

The most variable regions between two strains were found in the coding regions of transcriptional regulators, RNA polymerase, DNA polymerase, DNA topoisomerase III, endonuclease IV, and spor germination-related genes, which had only 24 to 38% similarity; some genes responsible for antibiotic resistance are absent in *P. amylolyticus* FSL-R5-192.

The complete genome sequence of *Paenibacillus* sp. strain VT 400 will help determine the role of *Paenibacillus* species in human diseases and provide insights into the composition of microbial flora in patients with hematological malignancies.

**Nucleotide sequence accession numbers.** This complete genome sequencing project has been deposited in GenBank under the accession no. LELF0000000. The version described in this paper is the first version, LELF01000000.

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REFERENCES

1. Rybakova D, Wetzlinger U, Müller H, Berg G. 2015. Complete genome sequence of \textit{Paenibacillus polymyxa} strain SB3-1, a soilborne bacterium with antagonistic activity toward plant pathogens. Genome Announc 3(2):e00052-15. http://dx.doi.org/10.1128/genomeA.00052-15.

2. De Souza R, Sant’Anna FH, Ambrosini A, Tadra-Sfeir M, Faoro H, Pedrosa FO, Souza EM, Passaglia LM. 2015. Genome of \textit{Pseudomonas} sp. FeS53a, a putative plant growth-promoting bacterium associated with rice grown in iron-stressed soils. Genome Announc 3(2):e00248-15. http://dx.doi.org/10.1128/genomeA.00248-15.

3. Spence R, Demchick P, Hornitzky M, Pharo H, Peacock L, McFadden A, Stone M. 2013. Surveillance of New Zealand apiaries for \textit{Paenibacillus alvei}. N Z Entomol 36:82–86.

4. Kim KK, Lee KC, Yu H, Ryoo S, Park Y, Lee JS. 2010. \textit{Paenibacillus sputi} sp. nov., isolated from the sputum of a patient with pulmonary disease. Int J Syst Evol Microbiol 60:2371–2376. http://dx.doi.org/10.1099/ijs.0.017137-0.

5. Padhi S, Dash M, Sahu R, Panda P. 2013. Urinary tract infection due to \textit{Paenibacillus alvei} in a chronic kidney disease: a rare case report. J Lab Physicians 5:133–135. http://dx.doi.org/10.4103/0974-2727.119872.

6. Ouyang J, Pei Z, Lutwick L, Dalal S, Yang L, Cassai N, Sandhu K, Hanna B, Wieczorek RL, Bluth M, Pincus MR. 2008. \textit{Paenibacillus thiaminolyticus}: a new cause of human infection, inducing bacteremia in a patient on hemodialysis. Ann Clin Lab Sci 38:393–400.

7. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prijibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. http://dx.doi.org/10.1089/cmb.2012.0021.

8. Tatusova T, DiCuccio M, Badretdin A, Chetverinin V, Ciufo S, Li W. 2013. Prokaryotic Genome Annotation Pipeline. In Beck J, Benson D, Coleman J, Hoepfner M, Johnson M, Magloff D, Mirzachi I, Morris R, Ostell J, Pruitt K, Rubinstein W, Sayers E, Sirotkin K, Tatusova T (ed), The NCBI handbook, 2nd ed. National Center for Biotechnology Information, Bethesda, MD.

9. Calle Y, Palomares T, Castro B, del Olmo M, Bilbao P, Alonso-Varona A. 2000. Tunicamycin treatment reduces intracellular glutathione levels: effect on the metastatic potential of the rhabdomyosarcoma cell line 54MH. Chemotherapy 46:408–428.

10. Kang H, Kim TJ, Kim WY, Choi CH, Lee JW, Kim BG, Bae DS. 2008. Outcome and reproductive function after cumulative high-dose combination chemotherapy with bleomycin, etoposide and cisplatin (BEP) for patients with ovarian endodermal sinus tumor. Gynecol Oncol 111:106–110. http://dx.doi.org/10.1016/j.ygyno.2008.05.033.

11. Slamti L, Lereclus D. 2002. A cell-cell signaling peptide activates the PlcR virulence regulon in bacteria of the \textit{Bacillus cereus} group. EMBO J 21:4550–4559. http://dx.doi.org/10.1093/emboj/cdf450.

12. Auch AF, von Jan M, Klenk H-P, Göker M. 2010. Digital DNA-DNA hybridization for microbial species delineation by means of genome-to-genome sequence comparison. Stand Genomic Sci 2:117–134. http://dx.doi.org/10.4056/sigs.531120.

13. Chun J, Rainey FA. 2014. Integrating genomics into the taxonomy and systematics of the \textit{Bacteria} and \textit{Archaea}. Int J Syst Evol Microbiol 64:316–324. http://dx.doi.org/10.1099/ijs.0.054171-0.