Case Report:

SURVIVAL OF A CORONAVIRUS DISEASE-2019 (COVID-19) PATIENT WITH ACUTE RESPIRATORY DISTRESS SYNDROME (ARDS) IN DR. SOETOMO HOSPITAL, SURABAYA, INDONESIA

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ABSTRACT

An outbreak of coronavirus disease 2019 (COVID-19) caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) that began in Wuhan, China has spread rapidly in multiple countries of the world and has become a pandemic. Currently, there is no vaccine or specific antiviral for COVID-19. A study reported 7.3% of critical patients admitted to ICU, 71% of them required mechanical ventilation, and 38.5% of them were survived. Herein, we reported a 54-year-old man with Acute Respiratory Distress Syndrome (ARDS) of COVID-19 who survived the disease. Real-time reverse transcriptase-polymerase chain reaction (RT-PCR) assay of nasopharyngeal and oropharyngeal swabs were positive for SARS-CoV-2. Diagnosis of ARDS was also according to clinical symptoms, laboratory, chest radiograph, and chest CT scan. Alcaligenes faecalis and Candida albicans were also identified from sputum culture. Treatment for this patient was causal and supportive therapy, including antibiotic, antiviral, and antifungal therapy according to the culture results, fluid resuscitation, and oxygen supply from the mechanical ventilator. This patient was survived and discharged on hospital day-29. A fibrosis in parenchyma pulmonary and sensory peripheral neuropathy occurred after survived from ARDS. Monitoring of clinical, laboratory, and chest radiograph were continued after the patient discharged from the hospital. This case highlights the importance of early diagnosis and effective treatment to the care of COVID-19 patient.

Keywords: COVID-19; Acute Respiratory Distress Syndrome

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ABSTRAK

Wabah penyakit coronavirus 2019 (COVID-19) disebabkan oleh severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) yang pertama dilaporkan di Wuhan, Cina telah menyebabkan dampak di berbagai negara di dunia dan telah menjadi pandemi global. Belum ada vaksin atau antivirus khusus untuk COVID-19 yang ditemukan hingga saat ini. Sebuah studi melaporkan terdapat 7,3% pasien kritis yang dirawat di ICU, 71% dari mereka membutuhkan ventilator, dan 38,5% dari mereka selamat. Kami melaporkan seorang pria berusia 54 tahun dengan Acute Respiratory Distress Syndrome (ARDS) COVID-19 yang sembuh dari penyakit tersebut. Uji real-time reverse transcriptase-polymerase chain reaction (RT-PCR) sampel nasofaring dan oropharingeal menunjukkan hasil positif SARS-CoV-2. Diagnosis ARDS juga sesuai dengan gejala klinis, laboratorium, fototoraks, dan CT scan. Alcaligenes faecalis dan Candida albicans juga dideteksi dari sputum. Pengobatan untuk pasien ini adalah terapi kausal dan suportif, termasuk terapi antibiotik, antivirus, dan antifungal berdasarkan hasil kultur, reusitasi cairan, dan pasokan oksigen dari ventilator. Pasien ini sembuh dan diperbolehkan pulang hari ke-29. Fibrosis pada parenkim paru dan neuropati perifer sensorik terjadi setelah sembuh dari ARDS. Pemantasan klinis, laboratorium, dan fototoraks dilanjutkan setelah pasien kembali dari rumah sakit. Kasus ini menunjukkan pentingnya diagnosis dini dan pengobatan yang efektif untuk perawatan pasien COVID-19.

Kata kunci: COVID-19; Acute Respiratory Distress Syndrome

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INTRODUCTION

On December 2019, several pneumonia cases of unknown etiology have been reported in Wuhan, Hubei province, China, with clinical presentations greatly resembling viral pneumonia (WHO 2020, Huang et al 2020). Deep sequencing analysis from lower respiratory tract samples indicated a novel coronavirus and the term 2019 novel coronavirus (2019-nCoV) was used by the World Health Organization (WHO) referring to this novel coronavirus (Huang et al 2020). The WHO announced the official name for the disease as coronavirus disease 2019 (COVID-19) and the official name for the virus was severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) (WHO 2020).

Currently, there is no vaccine or specific antiviral for COVID-19. A better understanding of COVID-19 management including supportive therapy was needed. Several reports have described the epidemiology and clinical features of COVID-19 cases. More specifically, through a detailed and comprehensive strategy, this report described the symptoms, biological abnormalities, radiographic abnormalities, and treatment of a case of COVID-19 with Acute Respiratory Distress Syndrome (ARDS) in Surabaya, Indonesia.

CASE REPORT

A 54-year-old man presented with viral pneumonia and respiratory failure on March 10, 2020. The patient was a nonsmoker. He had a long history of cough and there was no history of diabetes mellitus, hypertension, asthma, and tuberculosis. On admission to a private hospital, the patient reported a history of a one-week dry cough, headache, myalgia, and dyspnea followed with chest pain on illness day 7 (Table 1). The physical examination revealed a body temperature of 37.3°C, blood pressure of 144/88 mmHg, pulse rate of 107 beats per minute, respiratory rate of 30 breaths per minute, and oxygen saturation of 95%. Clinical laboratory examination was done on the 1st day of hospitalization. It showed a low lymphocyte of 13.5 % and a high C-reactive protein of 242.7 mg/L (Table 2). First examination of chest X-ray and chest CT scan on the same day. Chest X-ray showed dominant bilateral opacity in the 2/3 lower lung, partially homogenous and patchy (Fig. 1). Chest CT scan showed extensive confluent ground glass opacity in the right superior and medial lobe, left superior lobe and lingual (dominantly in the peripheral zone), followed with consolidation in the bilateral right inferior lobe as seen in Fig. 2.

Fig. 1. Chest X-Ray, 10 March 2020 (Illness Day 7, Hospital Day 1).
Initial blood gas analysis of this patient showed severe hypoxemia (PaO2/FiO2 = 68.39). According to the Berlin definition, this was defined as severe oxygenation (PaO2/FiO2 = 100 mmHg with PEEP = 5 cm H2O) (ARDS Definition Task Force 2012). Starting on the first hospitalization, the patient received a combination of antibiotics consisted of meropenem injection 1g every 8 hours, and Levofloxacin injection 750 mg every 24 hours, as well as supporting therapy. On the 2nd day of hospitalization (day 8 of illness), he reported a heavier shortness of breath. The patient was then transferred to Dr. Soetomo Hospital, one of the referral hospitals, to be treated in the ICU isolation room. Specimens such as blood, sputum, nasopharyngeal, and oropharyngeal swab specimens were to identify SARS-CoV-2, bacterial, and fungal culture examinations. The patient was also supported by O2 NRM 15 lpm and then replaced by O2 ventilator PEEP 8, FiO2 50% on the following day. Diagnosis of ARDS was established according to acute onset, severe hypoxemia (PaO2/FiO2 < 200) and diffuse bilateral pulmonary infiltrate on a chest X-ray or CT Scan Thorax.

The patient’s nasopharyngeal and oropharyngeal swabs tested positive for SARS-CoV-2 by real-time reverse transcriptase-polymerase chain reaction (RT-PCR) assay. The general condition of the patient was weak with GCS of 456. Vital signs of body temperature and SO2 were in normal ranges (36.9 0C and 95%, respectively), while other vital signs were out of normal ranges (blood pressure of 133/88 mmHg, pulse rate of 107 beats per minute, and respiratory rate of 30 times per minute). On hospital day 8, vital signs showed 162/98 mmHg of blood pressure, 121 beats per minute of pulse rate, and 24 times per minute of respiratory rate. Added therapy during hospitalization were including Chloroquine for 10 days, Isoprenosin 500 mg every 8 hours given via nasogastric tube (NGT) for 7 days, 2 tablets of Lopinavir/Ritonavir every 12 hours for 7 days. Chloroquine and Lopinavir/Ritonavir were given based on the patient’s severity/worsening clinical status.

Alcaligenes faecalis and Candida albicans were isolated from the sputum culture on hospital day 8. Alcaligenes faecalis was susceptible to Piperacillin-Tazobactam and Candida albicans was susceptible to Fluconazole sensitive. Meropenem injection and Levofloxacin were discontinued after given for 8 days and replaced with Piperacillin-Tazobactam pump 4.5 gr every 6 hours, then Fluconazole was started with the initial dose of 1x400 mg and followed by 1x200 mg every 24 hours. The patient experienced septic shock and hypoalbuminemia (2.9 g/dL) on hospital day 10 or illness day 16. Other than NGT diet of 200 ml every 4 hours and supported with O2 ventilator PEEP 6, FiO2 55%, Triofusin IVFD E1000 500 ml in 24 hours, albumin infusion, and Norepinephrine pump starting from 50 nano.

General condition on day 6 to 9 of hospitalization (day 12 to 15 of illness) was shown with GCS of 111 (on sedation), and on day 10 to of hospitalization (day 16 to of illness) was shown with GCS of 1x1 (intubated). The following RT-PCR for SARS-CoV-2 showed negative on illness day 16 (hospital day 10). Serial chest X-ray on hospital day 6 to hospital day 22 showed a slow improvement (Fig. 3). On hospital day 16 (illness day 22), the clinical condition of the patient was improved, the dyspnea was getting better but still experienced an infrequent cough and weaning ventilator was done. Oxygen supply was replaced with a simple mask. Piperacillin-Tazobactam was given for 10 days, while Fluconazole was given for 3 weeks.
| Day of illness | Pre-hosp | D 1 | D 2 | D 3 | D 4 | D 5 | D 6 | D 7 | D 8 | D 9 | D 10 | D 11 | D 12 | D 13 | D 14 | D 15 | D 16 |
|---------------|---------|-----|-----|-----|-----|-----|-----|-----|-----|-----|------|------|------|------|------|------|------|
| Lab           |         |     |     |     |     |     |     |     |     |     |      |      |      |      |      |      |      |
| Private Hospital |       | 37.3 | 37.2 | 37.5 | 37.1 | 37.2 | 36.9 | 36.9 | 36 | 36.6 | 36.6 | 37.4 | 36.8 | 36.5 | 36.5 | 36.8 | 36.8 |
| Dr. Soetomo Hospital |      |      |      |      |      |      |      |      |      |      |      |      |      |      |      |      |      |
| Fever (°C)    |         |     |     |     |     |     |     |     |     |     |      |      |      |      |      |      |      |
| Subjective fever |      | 37.3 | 37.2 | 37.5 | 37.1 | 37.2 | 36.9 | 36.9 | 36 | 36.6 | 36.6 | 37.4 | 36.8 | 36.5 | 36.5 | 36.8 | 36.8 |
| Dry cough     |         |     |     |     |     |     |     |     |     |     |      |      |      |      |      |      |      |      |
| Dyspnea       |         |     |     |     |     |     |     |     |     |     |      |      |      |      |      |      |      |      |
| Myalgia       |         |     |     |     |     |     |     |     |     |     |      |      |      |      |      |      |      |      |
| Headache      |         |     |     |     |     |     |     |     |     |     |      |      |      |      |      |      |      |      |
| Respiratory failure |       | 1   | 1   | 1   | 1   | 1   | 1   | 1   | 1   | 1   | 1   | 1   | 1   | 1   | 1   | 1   | 1   |
| Sensory peripheral neuropathy |     |     |     |     |     |     |     |     |     |     |      |      |      |      |      |      |      |      |
| Date (March 2020) |     | 4-5 | 6-9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 | 17 | 18 | 19 | 20 | 21 | 22 | 23 | 24 | 25 |
### Table 2. Clinical Laboratory Results

| Measure                                      | Reference Range | ID-7 | ID-10 | ID-12 | ID-15 | ID-18 | ID-21 | ID-24 | ID-27 | ID-2 Apr |
|----------------------------------------------|-----------------|------|-------|-------|-------|-------|-------|-------|-------|---------|
| White blood cell (10^3/µL)                  | 4.8-10.8        | 5.2  | 9.36  | 11.69 | 11.96 | -     | 15.96 | 7.75  | 7.68  |         |
| Lymphocyte (%)                               | 19-48           | 13.5 | 5.8   | 3.7   | 4.0   | -     | 6.2   | 9     | 15.9  |         |
| Platelet (10^3/µL)                           | 150-450         | 233  | 317   | 110   | 108   | -     | 254   | 353   | 486   |         |
| Hemoglobin (g/dL)                            | 12-16           | 17.5 | 16.7  | 15.8  | 15.8  | -     | 15.3  | 14.1  | 14.7  |         |
| Hematocrit (HCT) (%)                         | 37.7-53.7       | 49.3 | 45.9  | 46.9  | 47.9  | -     | 45.2  | 40.4  | 42.8  |         |
| Sodium (Na) (mmol/l)                         | 136-145         | -    | 132.6 | 138   | 133   | 137   | 143   | -     | -     |         |
| Potassium (K) (mmol/l)                       | 3.5-5.1         | -    | 3.79  | 4.5   | 4.3   | 3.4   | 3.4   | -     | -     |         |
| Glucose (mg/dL)                              | 100-125         | 97   | -     | 140   | 126   | 218   | 92    | -     | -     |         |
| Blood urea nitrogen (mg/dL)                  | 7-18            | 18   | 10    | 13    | 18    | 24    | 30    | -     | -     |         |
| Creatinine (mg/dL)                           | 0.6-1.3         | 1.0  | 0.6   | 0.7   | 0.8   | 1.0   | 0.86  | -     | -     |         |
| Albumin (g/dL)                               | 3.4-5           | 3.6  | 3.4   | 3.0   | 3.0   | 3.3   | 3.2   | -     | -     |         |
| SGOT (U/L)                                   | 0-50            | 71.9 | 57.2  | 58    | 29    | -     | 66    | -     | -     |         |
| SGPT (U/L)                                   | 0-50            | 40.1 | 40.3  | 87    | 36    | -     | 107   | -     | -     |         |
| Procalcitonin (ng/ml)                        | <0.05           | 0.78 | 4.06  | -     | -     | -     | -     | 0.05  | 0.02  |         |
| C-reactive protein (mg/L)                    | <6              | 242.7| 94.5  | -     | -     | -     | 13.8  | -     | -     |         |

Note: ID = Illness Day, HD = Hospital Day.

### Table 3. Blood Gas Analysis

| Hospital (Day) | D  | D  | D  | D  | D  | D  | D  | D  | D  | D  | D  | D  |
|----------------|----|----|----|----|----|----|----|----|----|----|----|----|
| Day of illness | 2  | 4  | 6  | 8  | 9  | 10 | 11 | 12 | 13 | 14 | 15 |
| pH             | 7.53| 7.56| 7.41| 7.53| 7.14| 7.42| 7.24| 7.33| 7.42| 7.48| 7.53| 7.47|
| pCO₂           | 26.9| 31.1| 33  | 63  | 83  | 57  | 57  | 55  | 49  | 37  | 47  | 42  |
| pO₂            | 55.4| 79.4| 91  | 93  | 112 | 80  | 89  | 99  | 96  | 74  | 78  | 71  |
| HCO₃            | 23 | 28.3| 20.9| 31  | 28.3| 37  | 24.4| 29  | 31.8| 27.6| 39.3| 30.6|
| BE             | 2.2 | 3.3 | -3.7| 4.6 | -0.7| 12.5| -3.0| 3.1 | 7.3 | 4.1 | 16.6| 6.9 |
| SO₂            | 92.6| 97.5| 95  | 96  | 97  | 96  | 95  | 97  | 98  | 96  | 97  | 95  |
| PaO₂/ FiO₂      | 68.39| 115.07| 182 | -   | 224 | 145 | 148 | 15 | 240 | 185 | 195 | 180 |
| A-aDO₂         | 488.5| 373.7| 219 | 185 | 246 | - | 553 | 260.1 | - | 165 | 148.4| 161.7|

March 2020

| Date | 11 | 13 | 15 | 16 | 17 | 18 | 19 | 20 | 21 | 22 | 23 | 24 |

### Table 4. Results of Real-Time Polymerase Chain Reaction Testing for SARS-CoV-2

| Specimen | Illness Day 11 | Illness Day 16 | Illness Day 20 |
|----------|----------------|----------------|----------------|
| Hospital Day 5 | Positive      | Negative       | Negative       |
| Hospital Day 10 | Negative      | Positive       | Negative       |
| Hospital Day 14 | Negative      | Positive       | Negative       |

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Table 5. Given Supportive Therapy for Patient

| Day of illness | Hospital (Day) |
|----------------|----------------|
|                | D 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 |
| Therapy        |                |
| IVFD           |                |
| O₂ NRM 15 ml   |                |
| O₂ ventilator  |                |
| O₂ simple mask 6 lpm |                |
| NE 50 nano     |                |
| NE 25 nano     |                |
| Meropenem inj 1gr |                |
| Levofloxacin 750 mg |                |
| Piperacillin-tazobactam pump: 4.5 gr |                |
| Isoprenosin 500 mg |                |
| Lopinavir/ritonavir 2x2 |                |
| Chloroquine    |                |
| Fluconazole 1x400 mg |                |
| Fluconazole 1x200 mg |                |
| Methylcobalt 500 mg |                |
| Alinamin F     |                |
| Provelyn 75 mg  |                |

| Date           | March 2020 |
|----------------|------------|
|                | 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 |
Fig. 3. Serial Chest Radiograph Showed A Slow Improvement (A-I). A. 15 March 2020 (Illness Day 12, Hospital Day 6); B. 16 March 2020 (Illness Day 13, Hospital Day 7); C. 19 March 2020 (Illness Day 16, Hospital Day 10); D. 27 March 2020 (Illness Day 24, Hospital Day 18); E. 30 March 2020 (Illness Day 27, Hospital Day 19); F. 2 April (Illness Day 30, Hospital Day 22).

Evaluation of chest CT scan of the patient on 7 April 2020 (illness day 35, hospital day 29) showed fibrosis in the lung peripheral basal and peripheral apex, dominantly in the posterior side of the right lung and inferior lobe of the left lung. Minimal ground glass opacity appeared on the posterior side of the right lung. A left bronchial retraction with minimal hilar retraction was seen. Trachea looked patent. Overall, minimal pneumonia was seen in the anterior segment of the superior lobe of the right lung, extensive fibrosis accompanied by predominantly ground glass opacity in the bilateral peripheral lung base (Fig. 4). On this evaluation day of chest CT scan, the patient’s vital signs were stable and light mobility without an oxygen supply.

Fig. 4. Serial of Chest CT Scan, 7 April 2020 (Illness Day 35, Hospital Day 29)
DISCUSSION

This patient was discharged on hospital day 29 (7 April 2020). A lung fibrosis was shown by chest CT scan (Fig. 4). Chang et al (2018) reported a pneumonia patient with persistent lung fibrosis and new onset of ground glass opacity (GGO). After antibiotics treatment, the patient was discharged and was followed up at the outpatient clinic. The patient has received a short course of corticosteroid treatment (prednisolone 10 mg) daily for 3 months. After 7 months of illness, the chest CT showed resolution of the GGO and a significant improvement in lung fibrosis. One year after the illness, his pulmonary function was normal: the forced vital capacity 80%, forced expiratory volume in 1 s 86%, diffusing capacity for carbon monoxide 61%, and diffusing capacity divided by the alveolar volume 80%. He returned to his normal daily activities (Chang et al 2018).

A study in China reported of the 710 patients with SARS-CoV-2 pneumonia, 52/710 (7.3%) were critical patients treated in the ICU. Of the 52 critical patients, 32/52 (61.5%) died, 20/52 (38.5%) survived, 35/52 (67%) had ARDS, and 37/52 (71%) required mechanical ventilation (Yang et al 2020). In our hospital, there were 14 critical patients treated in the ICU. All critical patients had ARDS (100%). Of the 14 critical patients, 4/14 (28.5%) patients still receive ventilator, 3 (21.5%) patients survived, 1 (7%) patient is weaning ventilator (low care), and 6/14 (43%) died with causa mortis of acute kidney injury (AKI) and septic shock.

The most symptoms of survivors among critical patients with COVID-19 are fever (100%), cough (75%), dyspnea (60%), myalgia (10%), malaise (20%), arthralgia (5%), chest pain (5%), headache (5%), and vomiting (5%) (Yang et al 2020). Fever is often the major and initial symptom of COVID-19, which can be accompanied by no symptom or other symptoms such as dry cough, shortness of breath, muscle ache, dizziness, headache, sore throat, rhinorrhea, chest pain, diarrhea, nausea, and vomiting. Some patients experienced dyspnea and/or hypoxemia one week after the onset of the disease (Guan et al 2020). The early clinical and laboratory findings of COVID-19 pneumonia are fever, dry cough, and fatigue with normal WBC count, reduced lymphocyte count, and elevated C-reactive protein level (Han et al 2020). Our case presented symptoms of fever, dry cough, headache, myalgia and experienced dyspnea followed with chest pain on illness day 7. Laboratory findings are normal WBC count, low lymphocyte, high procalcitonin, and high C-reactive protein.

Comorbidities in COVID-19 patients were diabetes mellitus, hypertension with complications of acute respiratory distress syndrome (ARDS), acute kidney injury, cardiac injury, liver dysfunction, gastrointestinal hemorrhage, hospital-acquired pneumonia, bacteremia, and urinary tract infection (Yang et al 2020, Guan et al 2020). Bacteremia and fungal coinfection were found only in non survived patients. Klebsiella pneumonia, Pseudomonas aeruginosa, Serratia marcescens, Aspergillus flavus, and Aspergillus fumigates were identified from respiratory tract secretions, and Candida albicans was identified in the urine culture (Yang et al 2020). Our case only had comorbidity of chronic bronchitis. Alcaligenes faecalis and Candida albicans were identified from sputum culture.

A study of 52 critical patients with COVID-19 reported 33 (63.5%) patients were treated with a high-flow nasal cannula, 37 (71%) with mechanical ventilation, 6 (11.5%) with prone position ventilation, 6 (11.5%) with extracorporeal membrane oxygenation (ECMO) (Yang et al 2020), while our case received supplemental oxygen of NRM, mechanical ventilator, and a simple mask.

Antiviral agents consisted of oseltamivir, ganciclovir, and lopinavir were given to 18 (35%), 14 (27%), and 7 (13.5%) patients, respectively (Yang et al 2020), while our case received lopinavir/ritonavir and chloroquine as antiviral treatment. Yang et al (2020) reported 49 (94%) patients received antibacterial agents, while our case received empirical antibiotics of levofloxacin and meropenem. After culture result was obtained, piperacillin-tazobactam was given as definitive antibiotic therapy according to the culture result (Yang et al 2020). Yang et al (2020) did not report any patient received antifungal therapy (Yang et al 2020), while our case received fluconazole according to the culture result showed Candida albicans. The detail of supportive therapy for our case was presented in Table 5.

As shown in Table 4, the initial nasopharyngeal and oropharyngeal swabs obtained from this patient on illness day 11 (hospital day 5) were positive for SARS-CoV-2. The specimens tested negative for SARS-CoV-2 on illness day 16 (hospital day 10), while Holshue et al (2020) reported the initial specimens obtained on illness day 4 tested positive for SARS-CoV-2. On illness day 12, the oropharyngeal swab tested negative for SARS-CoV-2. The nasopharyngeal swab remained positive for SARS-CoV-2 on illness day 12, but showed decreasing levels of virus. The patient received acetaminophen, ibuprofen, and guaifenesin for symptom management. Vancomycin and cefepime were given as antibiotics. Remdesivir was given as antiviral (Holshue et al 2020).
The early CT findings of COVID-19 are patchy ground glass opacity with or without consolidation involving multiple lobes, mainly in the peripheral zone, accompanied by halo sign, vascular thickening, crazy paving pattern, or air bronchogram sign (Han et al 2020), the most common patterns on chest CT were ground-glass opacity (56.4%) and bilateral patchy shadowing (51.8%) (Guan et al 2020). Abnormalities in chest radiography of COVID-19 patients featured by bilateral patchy shadows or ground glass opacity in the lungs. Patients often develop atypical pneumonia, acute lung injury, and acute respiratory distress syndrome (ARDS). When ARDS happens, uncontrolled inflammation, fluid accumulation, and progressive fibrosis severely compromise the gas exchange (Yi et al 2020). Chest radiograph of our case showed dominant bilateral opacity in the 2/3 lower lung, partially homogenous and patchy. Chest CT scan showed extensive confluent ground glass opacity in the right superior and medial lobe, left superior lobe, and lingual (dominantly in the peripheral zone), followed with consolidation in the bilateral right inferior lobe.

The long-term sequelae in patients with severe pneumonia, although not available for the present, could be the fibrotic changes such as reticulation, interlobular septal thickening, and traction bronchiectasis, which are usually seen in the fibrotic phase of lung injury (Kim et al 2020). At the end of treatment, our case’s chest CT scan showed fibrosis in the peripheral basal and peripheral apex, dominantly in the posterior side of the right lung lobe and inferior side of the left lung lobe. Minimal ground glass opacity was present in the posterior side of the right lung lobe. Minimal hiliar retraction was also presented.

Our case reported sensory peripheral neuropathy symptoms of SARS-CoV-2 infection (felt after awake post-extubation). Troyer et al (2020) stated that following past influenza pandemics and CoV outbreaks, such complications have been described over highly variable periods, from weeks following acute respiratory symptoms in the case of neuromuscular and demyelinating processes, to decades after in-utero exposure to viral infection in the case of schizophrenia onset. Whether recovered SARS-CoV-2 patients will exhibit an increased incidence of delayed neurologic sequelae is currently unknown and remain to be seen over the next several months to years (Troyer et al 2020).

CONCLUSION

In conclusion, our case survived from COVID-19 after experienced ARDS. Many factors may play a role in this survival case, including age, little comorbid, and the implementation of early supportive therapy, early initial antibiotic and on time use of the mechanical ventilator. Rapid diagnosis, wise use of diagnosis tools, and effective treatment including antibiotic, antiviral, antifungal, and supportive therapy were needed to manage this patient in the pandemic area.

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