Introduction:

Helminths, protozoa and ectoparasites could resist many types of drugs by different mechanisms [1-11]. In our previous paper [12], we reviewed the medicinal plants possessed antiparasitic, antiprotozoal, molluscicidal and insecticidal activity. These included Achillea santolina, Ailanthus altissima, Allium species, Ammi majus, Antirrhinum majus, Apium graveolens, Arachis hypogaea, Artemisia campestris, Arundo donax, Asclepias curassavica, Ballota nigra, Bauhinia variegate, Betula alba, Bidens tripartite, Brassica nigra, Bryophyllum calycinum, Caccinia crassifolia, Caesalpinia crista, Calendula officinalis, Calotropis procera, Canna indica, Capparis spinosa, Carum carvi, Cassia occidentalis, Celosia cristata and Chenopodium album. This review was designed as a second part of medicinal plants exerted antiparasitic, antiprotozoal, molluscicidal and insecticidal activity.

Keywords: antiparasitic, antiprotozoal, molluscicidal, insecticidal, medicinal plants, herbs.

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Review Article

Antiparasitic, antiprotozoal, molluscicidal and insecticidal activity of medicinal plants (part 2) – plant based review

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doi: 10.21276/sajp.2016.5.6.1

Scholars Academic Journal of Pharmacy (SAJP)
ISSN 2320-4206 (Online)
ISSN 2347-9531 (Print)
©Scholars Academic and Scientific Publisher
(An International Publisher for Academic and Scientific Resources)
www.saspublisher.com

Didymyris tenuifolia, Florida, Georgia, Guatemala, Hungary, India, Iran, Ireland, Iran, Jamaica, Madagascar, Mexico, Morocco, New Zealand, Nigeria, Palestine, Persia, Peru, Philippines, Puerto Rico, United States, St. Helena, Spain, Sudan, Sweden, Switzerland, Trinidad, Turkey, Russia, South Africa and Zimbabwe [39-40].

The plant products were available for use even though the chemistry of active ingredients was unknown. In 1924, German chemist Herman Staudinger (1881-1965) and a Croatian scientist, Lavoslav Ružička (1887-1976), winner of the 1939 Nobel Prize in Chemistry, identified chemical structure of the active pyrethrin ingredients, pyrethin I and pyrethrin II [41].

Pyrethrin is mainly concentrated in the flower heads, 93.7 % of pyrethrin is accumulated in achenes, and minor quantities in disc florets (2.0 %), ray florets (2.6 %), and receptacles (2.6 %). The term pyrethrin is
referred to the six insecticide active ingredients: pyrethrins I, cinerin I, jasminol I, pyrethrin II, cinerin II and jasminol II. Pyrethrins I, cinerin I and jasminol I are closely related insecticidal esters of chrysanthemic acid, while pyrethrin II, cinerin II and jasminol II are insecticidal esters of pyrethric acid. The three chrysanthemic acid esters are commonly referred as pyrethrins I, and pyrethric acid esters as pyrethrins II. Among these compounds pyrethrin I and pyrethrin II are the most predominant and active [42-45].

It is used for the control of a wide range of insects and mites in public health and on domestic and farm animals and for the control of chewing and sucking insects and spider mite on fruit, vegetables, field crops, ornamentals, glasshouse crops, and house plants. Pyrethrin also acts as a repellent. It has a quick knock-down effect on a wide range of insect species, causing paralysis within a few minutes. It acts as contact poison and affects the central nervous system of insects. The commonly accepted mechanism of action of pyrethroids is the prolongation of the open state of voltage-dependent sodium channels in nervous tissue. These altered sodium channels result in repetitive firing or depolarizing block of the neuron, depending on how long the channel open state is prolonged. Other channel and receptor systems in neuronal tissues have been proposed to play a role in the generation of compound-specific clinical symptoms in mammals, including calcium channels and GABA<sub>A</sub> receptors [43, 46].

In studying the mechanism of biosynthesis of pyrethrins, the results of experiments using <sup>13</sup>C-labeled glucose as the biosynthesis precursor indicated that the acid and alcohol moieties are biosynthesized via the 2-C-methyl-D-erythritol 4-phosphate (MEP) and oxylipin pathways, respectively. Further study on the effects of wound-induced signals in leaves showed that biosynthesis is enhanced in response to both volatile and nonvolatile signals [47]. In other study the acid was found produced by the same route, but the authors mentioned that the alcohol moiety was possibly biosynthesized from linoleic acid [48].

Tissue cultures of *Chrysanthemum cinerariaefolium* were used to study the production of pyrethrin insecticides, and their precursor chrysanthemic acid. Callus cultures and root-differentiated cultures did not contain pyrethrins whereas shoot differentiated callus was found to produce the pyrethrins. Chrysanthemic acid was isolated by extraction from callus cultures, and feeding <sup>14</sup>C-labelled chrysanthemic acid to a cell suspension of *Chrysanthemum cinerariaefolium* established that the acid accumulates largely as a glucoside ester [49].

The main use of pyrethrum at present is in household formulations to kill houseflies, cockroaches and mosquitoes. Insects react very quickly when dosed with pyrethrum, and this quick knock-down effect is a very valuable property for household insecticides as the user sees the onset of paralysis within 2–3 minutes. Pyrethrum acts on a very wide spectrum of insect species – more so than with most individual synthetic insecticides. It is also a very powerful repellent to mosquitoes. The process of piercing the skin using saliva injected as a lubricant, finding and piercing a blood capillary followed by sucking of blood into the insect is a very precise and complicated process and coordination is lost when even a few molecules of pyrethrum are present. Synergised pyrethrum formulations are particularly effective against the mosquitoes and midges, which bite humans, and to date there have been no reports of resistance developing in these species to this biopesticide. Pyrethrins are very quickly degraded in sunlight leaving no toxic residues and are ideal for use as pre-harvest sprays to remove insect pests on edible crops – up to 24 hours before harvest [50-51].

The efficacies of pyrethrum and albendazole against experimental sheep gastrointestinal nematode infection were compared. Sheep were infected orally with 10,000 larvae (*Haemonchus* spp. (60.1%), *Oesophagostomum* spp. (13.9%), *Trichostrongylus* spp. (13.2%), *Cooperia* spp. (8.3%), *Nematodirus* spp. (3.5%), *Strongyloides* spp. (0.8%) and *Ostertagia* spp. (0.2%). Faecal egg count reduction in albendazole-treated sheep was 100% by day 4 following treatment, compared to 37.03%, 31.3%, 38.9% and 51.8% on days 4, 6, 8 and 10 in pyrethrum treated sheep. These effects were statistically significant on days 8 and 10 post-treatment (p< 0.05) [52].

*Chichorium intybus*

It appeared that the animals grazing on chicory have a lower incidence of gastrointestinal nematode infestations, the total number of abomasal helminths was found to be lesser in the lambs grazing on this plant. The anthelmintic activity of the plant attributed to the condensed tannins and sesquiterpene lactones [53-54]. The effects of condensed tannins (CT) and an extract containing crude sesquiterpene lactones (CSL) from chicory (*Chichorium intybus*) on the motility of the first-(L1) and third-stage (L3) larvae of deer lung worm *Dictyocaulus viviparus* and the L3 larvae of gastrointestinal nematodes was studied in vitro, using the larval migration inhibition (LMI) assay. The CT and CSL had a profound effect on the motility of the larvae displayed by their ability to inhibit larval passage through nylon mesh sieves. Incubation of lungworm L1 larvae in rumen fluid (collected from deer fed pasture) containing 100, 400 and 1000 microg CT/ml, inhibited 12, 28 and 41% of the larvae from passing through the sieves, respectively, while the incubation of L3 larvae with rumen fluid (pH 6.6) containing the same concentrations inhibited 26, 37 and 67% of L3 larvae from passing through the sieves, respectively. Gastrointestinal larvae seem more susceptible to CT than lungworm larvae especially at
higher concentrations. CT inhibited 27, 56 and 73% of
gastrointestinal larvae from passing through the sieves
when used at a concentration of 100, 400 and 1000
microg/ml, respectively. CT were more effective
(P<0.001) at reducing the motility of lungworm L1 and
L3 larvae when added to the rumen fluid than when
added to the abomasal fluid (pH 3.0). Addition of 2
microg polyethylene glycol/microg CT eliminated the
inhibitory effect of CT against L1 and L3 lungworm
and L3 gastrointestinal larvae in both fluids, indicating
that this extract was not affected by the pH of the fluid,
and was more effective against L3 than L1 lungworm
larvae. Condensed tannins appeared to be more
effective than CSL at inactivating L1 and L3 lungworm
and L3 gastrointestinal larvae in rumen fluid, but CSL
were particularly effective against L3 lungworm larvae
in abomasal fluid [55-56].

To determine whether the individual sesquiterpene lactone compounds of Cichorium intybus
[lactucin (LAC), 8-deoxylactucin (DOL), and
lactucopicrin (LPIC)] differ in anthelmintic activity,
their effects were studied on the hatching of a
predominantly Haemonchus contortus egg population.
The dominant constituents in the Puna and Forage Feast
extracts were DOL and LAC, respectively; LPIC
concentrations in the two extracts were similar. Extracts
from both cultivars inhibited egg hatching at all
concentrations tested (P<0.001), but there were
significant differences in egg responses to the two
extracts (P<0.001). With Puna, egg hatching decreased
sharply in a linear fashion when the combined LAC,
DOL, and LPIC concentrations ranged from 0 to 5.0
mg/ml. A biphasic effect on egg hatching occurred with
the Forage Feast extract. The fraction of eggs that
hatched decreased gradually to 65% as the
sesquiterpene lactone concentrations increased from 0
to 6.7 mg/ml. Treatment with higher concentrations
resulted in a sharp decline in egg hatchability.
Concentrations of sesquiterpene lactones required for
50% lethality were determined by probit dose-effect
analysis to be 2.6 mg/ml (95% confidence interval: 2.4-
2.8 mg/ml) for the Puna extract and 6.4 mg/ml (95%
confidence interval: 5.9-7.2mg/ml) for the Forage Feast
extract (P<0.0001). These concentrations provided 1.3
and 1.5mg/ml of DOL and 0.8 and 3.9 mg/ml of LAC
for Puna and Forage Feast extracts, respectively.
However, the results showed that LAC has minimal
effect on egg hatching [57].

The bitter compounds in the plant, namely,
lactucin, lactucopicrin, and the guaianolide
sesquiterpenes, isolated from aqueous root extracts of
chicory were concluded to be the antimalarial
components of the plant. Lactucin and lactucopicrin
completely inhibited the HB3 clone of strain Honduras-
1 of Plasmodium falciparum at concentrations of 10 and 50
μg/ml, respectively [58-59].

Citrullus colocynthis
Albinio mice were intraperitoneally infected with 100 X 10⁸ promastigotes of Leishmania donovani
(MHOM/ IQ/ 982/BRCI) strain. The inoculation of
albinio mice caused elevation of liver and spleen weight
after 7-15 days. The mice treated with 20-100 mg/kg from Citrullus colocynthis showed decreased average
liver and spleen weight in comparison to the positive
control. The most important histopathological results in
the positive control included scattered necrosis,
lymaphatic infiltration, proliferation of macrophages and
a variable number of leishmanian bodies were observed.
80-100 mg/kg of Citrullus colocynthis return liver
section to normal histology [60-61].

Larvicidal activity of crude hexane, ethyl
acetate, petroleum ether, acetone, and
methanol extracts of the leaf of five species
cucurbitaceous plants, Citrullus colocynthis,
Coccinia indica, Cucumis sativus, Momordica
charantia, and Trichosanthes anguina, were
tested against the early fourth instar larvae of Aedes
aegypti L. and Culex quinquefasciatus (Say) (Diptera:
Culicidae). The larval mortality was observed after 24 h
of exposure. All extracts showed moderate larvicidal
effects; however, a high larval mortality was found in petroleum ether extract of
Citrullus colocynthis against the larvae of A. aegypti
(LC₅₀=74.57 ppm) and against lymphatic filariasis
vector, C. quinquefasciatus (LC₅₀=88.24ppm) [62].

The larvicidal activity of crude acetone,
hexane, ethyl acetate, methanol, and petroleum ether
effects of the leaf of Citrullus colocynthis (Linn.)
Schrad were assayed for their toxicity against the early
fourth instar larvae of Culex quinquefasciatus (Diptera:
Culicidae). The larval mortality was observed after 24 h
exposure. The highest larval mortality was found in
whole plant petroleum ether extract of Citrullus
colocynthis. Bioassay-guided fractionation of petroleum
ether extract led to the separation and identification of
fatty acids; oleic acid and linoleic acid were isolated
and identified as mosquito larvicidal compounds.
Oleic and linoleic acids were quite
potent against fourth instar larvae of Aedes aegypti L.
(their LC₅₀ were 8.80, 18.20 and LC₅₀=35.39, 96.33 ppm
respectively), Anopheles stephensi Liston (LC₅₀ 9.79,
11.49 and LC₅₀ 37.42, 47.35 ppm respectively), and
Culex quinquefasciatus Say (LC₅₀ 7.66, 27.24 and LC₅₀
30.71, 70.38 ppm respectively) [63].

Methylene chloride, n-hexane, chloroform and
ethanol extracts of Citrullus colocynthis fruits were
tested against Aphis craccivora. The highest
insecticidal effect (LC₅₀: 11003 ppm) was obtained
from the ethanol extract. The residue remaining after
evaporation of ethanol extract was re-extracted by
different solvents with increasing polarity. Each fraction was tested against *Aphis craccivora*. The butanol extract showed the maximum insecticidal effect. The effective compound was identified as 2-O-β-D-glucopyranosylcurcurbitacin E [64].

*Citrullus colocynthis* was evaluated as new therapeutic approach for scorpion envenomation mainly *Androctonus australis* hector venom (Aah). Local action (paw edema) and systemic effects (inflammatory, metabolic parameters, oxidative stress and hyperglycemia) were studied in pretreated mice with *Citrullus colocynthis* (50 mg/kg), 30 min before injection of sublethal dose of *Androctonus australis* hector venom (10 µg/20 g). Results showed that injected *Citrullus colocynthis* extract before envenomation is able to protect animals against the toxicity of the venom. It significantly reduced paw edema, cell migration, exudation, hyperglycemia, and MDA. *Citrullus colocynthis* decreased also some inflammatory markers (MPO and EPO activities, CRP and C3) and maintain the level of CPK, ASAT and ALAT. *Citrullus colocynthis* appeared to be a potential tool that can reduce pathophysiological effects induced after envenomation (inflammation and oxidative stress) [65].

**Citrus species**

Methanolic extract of *Citrus medica* was evaluated for anthelmintic activity against Indian adult earthworm *Pheretima posthuma*. Various concentrations of extract were tested and results were expressed in terms of time for paralysis and time for death of worms. Piperazine citrate (10 mg/ml) was used as a reference standard and distilled water as a control group. Dose dependent anthelmintic activity was possessed by the methanolic extract of *Citrus medica* [66].

Petroleum ether extracts of *Citrus medica* leaves also possessed dose dependant anthelmintic activity against the Indian adult earthworms (*Pheretima posthumad*). The effect which could be attributed to inhibition of glucose uptake in the parasites and depletion of its glycogen synthesis. It also activated nicotinic cholinergic receptor in the worms resulting in either persistent depolarization or hyperpolarization [67].

The anthelmintic activity of petroleum ether extract of the peels of *Citrus sinensis* was studied against Indian adult earthworms, *Pheretima posthuma*, it exhibited a dose dependent inhibition of spontaneous motility (paralysis), and evoked responses to pin-prick, and the effects were comparable with that of piperazine citrate [68].

The effect of aqueous extract of this *Citrus medica* on viability of the protoscolices of *Echinococcus granulosus* in vitro, in a concentration of 90 mg/ml, the aqueous extract was effective in killing all protoscolices after four days of incubation [69].

Alcoholic extracts of the rind of *Citrus medica* showed *in vitro* anthelmintic activity against human *Ascaris lumbricoides* [70].

The larvicidal potential of hexane and petroleum ether extracts of *Citrus limetta* peels was assessed against dengue fever vector, *Aedes aegypti*, and malarial vector, *Anopheles stephensi*, by evaluating the toxicity effects on early fourth instars. Both the extracts were found effective against both the species. The bioassay with hexane extracts resulted in LC₅₀ values of 132.45 and 96.15 ppm against *A. stephensi* and *A. aegypti*, respectively; while the petroleum ether extracts from the *C. limetta* peels showed LC₅₀ values of 244.59 and 145.50 ppm, respectively. It revealed that the hexane extracts possessed 1.9-fold more larvicidal potential against *A. stephensi* and 1.5-fold more efficacy against *A. aegypti* as compared to the extracts obtained using petroleum ether as solvent. The data further revealed that the extracts were 1.4-1.7 times more effective against *A. aegypti* as compared to *A. stephensi* [71].

The mosquito repellent activity of extracts from Peels of five citrus fruit species, *Citrus sinensis*, *Citrus limonum*, *Citrus aurantifolia*, *Citrus reticulata* and *Citrus vitis*, were studied using five different concentrations, 5%, 10%, 15%, 20% and 25% (volume by volume). Topical application of the extract concentrations on human volunteers revealed that 20% and 25% repelled mosquitoes 2 hours and 5 hours, respectively. Short-lived and mild skin itching and sneezing reactions were observed as side effects [72].

**Clerodendrum inerme**

Leaf extracts were evaluated for their nematicidal efficacy against root-knot nematodes. In the juvenile mortality assay against egg masses, leaf extracts of *C. inerme* significantly inhibited the development [73-74].

The aqueous extract of *Clerodendron inerme* (*C. inerme*) plant leaves was evaluated against laboratory strain *Aedes aegypti* larvae. The extract elucidated 100% inhibition of adult emergence at 2% concentration of extract, and concentrations above 4% led to prolongation of larval developmental period without moulting leading to death during larval stage. Mortality during larval stage was found to be dose-dependent elucidating 100% mortality at 16% concentration. It is apparent that the extract interferes in the developmental process affecting larval developmental period and disruption of larval-pupal moul [75].

Laboratory and field investigations have been made to evaluate the combined effect of *Clerodendron*
inerme and Acanthus ilicifolius on three species of mosquito vectors, Anopheles stephensi, Aedes aegypti and Culex quinquefasciatus. Different concentrations of Clerodendrum inerme and Acanthus ilicifolius have been tested on the various stages of species of mosquito vectors. They were active against different larval stages of mosquitoes. The lethal effect on mosquito larvae may be due to the active plant compounds on the gut lining of the mosquito larvae. The larval density was decreased after the treatment with the Clerodendron inerme extracts at the breeding sites (drinking water and ditches water) [76].

The dry powder of Clerodendrum inerme leaves was tested (10 to 60 mg) against freshly moulted fourth instar larvae of dengue mosquito vector Aedes aegypti. The results revealed that there was no larval mortality in the treated larvae and they moulted to pupae after 60h from the start of the experiment and the process was completed by 72h. Control larvae also required 60–72h to pupate. There were no visible behavioural changes in the treated larvae, except for the fact that they were not as active as those of control ones after 24h of treatment. During pupal stage also, the pupae in treated flasks were not as active as control groups. Flasks containing 40, 50 and 60 mg powder showed pupal mortality after about 18-20h. At the end of 72h, the percent pupal mortality in the same treated groups was 48, 74 and 96 respectively. Flasks containing 20 and 30 mg of powder exhibited less than 10% pupal mortality. In order to determine the quantity of powder required to cause larval mortality, the quantity of powder was increased from 100 to 200 mg with 20 mg increment between the treatments. The results showed dose-dependent larval mortality. As much as 85% larval mortality was seen when the powder quantity was increased to 160 mg. It was further noted that the fourth instar larvae that moulted to pupae died during the early pupal stage. The final analysis of results revealed 100% mortality in all the experimental flasks, which included larval as well as pupal mortality. Microscopic examination of dead larvae revealed that the larval cuticle had started sclerotization, which appeared to be a characteristic feature of the pupal cuticle. The dead pupae on the other hand, showed less sclerotization of the cuticle compared to untreated ones, and in majority of the pupae, the head capsule remained attached to the pupal head [77]. It was stated that petroleum ether extract of Clerodendrum inerme gave 3h protection against mosquitoes at 9% concentration [78].

The Petroleum ether, Chloroform, Ethyl acetate, Ethanol and water fractions of the powdered leaves of Clerodendrum inerme were tested for their efficacy against the stored grain insect pest Corcyra cephalonica (Stainton) (Lepidoptera Pyralidae). Seven different doses (0.05, 0.1, 0.15, 0.5, 1.0, 1.5, and 2.0 g) per 20.0 g of rice were tested against this common insect pest of rice to evaluate their effect on its life cycle and mortality. Three higher doses were further tested for their effect on physiological parameters like total haemocyte count (THC), total protein content and glycogen level along with starved insects. C. inerme exhibited biopesticidal activity as evidenced by the high mortality rate in treated insects. There was also a significant reduction in the THC (39-53%), protein (30-38%) and glycogen (40-61%) content in C. inerme treated larvae with respect to their controls [79].

The efficacy of Clerodendron inerme leaf extract was evaluated against Pieris brassicae. Larva, pupa and adult of P. brassicae have been treated with the aqueous extract of C. inerme leaf of different concentration. The results show that extract was quite effective against all the three stages in general, and pupa in particular. A typical extract with 12.5% concentration showed a mortality rate of 20% for larvae which rises to 55% for pupa. The mortality rate generally increases with increase in the concentration, reached to its maximum at 10% to 17.5% of concentration and then decreased or became constant for different developmental stages [80].

Clitoria ternatea

The ethanolic extract of Clitoria ternatea (100mg/ml) bring paralysis within 15-20 min and bring death within 28-30 min to the Indian earthworm Pheretima posthuma [81-82]. However, the anthelmintic activity of ethanolic extracts of flowers, leaves, stems and roots of Clitoria ternatea were also evaluated on adult Indian earthworms Pheretima posthuma. Results showed that roots of the Clitoria ternatea took less time to paralyze and death of the earthworms. Roots were further extracted successively with petroleum ether, chloroform, ethyl acetate and methanol and these extracts were screened for anthelmintic activity. Results showed that methanol extract of Clitoria ternatea root is the more potent [83].

The in vitro comparative study of anthelmintic activity of aqueous and ethanolic extracts of leaves of Clitoria ternatea was carried out against Eisenia fetida at three different concentrations (100, 50, 25 mg/ml). The study involved the determination of time of paralysis and time of death of the worms. At the concentration of 100 mg/ml both the ethanolic and the aqueous extracts showed very significant anthelmintic activities as compared to the standard drug, levamisole (0.55 mg/ml). In case of aqueous extract the time of paralysis and death time was observed as 18 ± 1.57 min and 53.33 ± 0.33 min, and in case of ethanolic extracts 12.33 ± 0.80 min and 32.33 ± 0.71min respectively [84].

The mosquito larvicidal activity of Clitoria ternatea was investigated against three major mosquito vectors Aedes aegypti, Culex quinquefasciatus, and Anopheles stephensi. Among the methanol extracts of Clitoria ternatea leaves, roots,
flowers, and seeds, the seed extract was effective against the larvae of all the three species with LC₅₀ values 65.2, 154.5, and 54.4 ppm, for A. stephensi, A. aegypti, and C. quinquefasciatus, respectively. Among three tested plant species, Clitoria ternatea was showing the most promising mosquito larvicidal activity [85].

Corchorus capsularis

The mosquitocidal activities of Corchorus capsularis against a common malarial vector, Anopheles stephensi and a dengue vector Aedes aegypti was studied. The larvicidal activity exerted by ethyl acetate was more prominent than acetone and methanol extracts in all concentrations tested against Ae. aegypti larvae. Evaluation of the lethal concentration values (LC₅₀ and LC₉₀) of acetone, ethyl acetate and methanol extract of the plant against An. stephensi and Ae.aegypti revealed that LC₅₀ of 197.34ppm and LC₉₀ of 358.59ppm was recorded for acetone extract against the An. stephensi; furthermore, the larvae of Ae. aegypti showed the LC₅₀ and LC₉₀ values of 222.45 and 383.06ppm respectively, with the treatment with the acetone extract of Corchorus capsularis. Minimum LC₅₀ values were observed among the experimental larval groups treated with methanol extract of Corchorus capsularis were 176.19ppm and 182.06ppm against An. stephensi and Ae. aegypti respectively. With regard to the ovicidal activity of acetone, ethyl acetate and methanol extract, it was apparent that 300 -450 ppm concentrations resulted with no hatchability on An. stephensi and 375-450pp concentrations in Ae. aegypti. The authors referred to the possible utilization of Corchorus capsularis to control mosquito menace to a greater extent [86].

The efficacy of emulsified petroleum ether extract of Corchorus capsularis seed was studied against three stored product pests (Callosobruchus chinensis, Sitophilus oryzae L. and Tribolium castaneum Herbst) in adult phase. The residual film technique method was conducted to determine the LC₅₀ value of the mentioned plant extract against three stored product pests. LD₅₀ (μg /cm) of Corchorus capsularis against C. chinensis was 74.26 (50.26 - 109.74) after 24 hrs and 6.67 (0.49 - 90.07) after 48hrs. LD₅₀ against S. oryzae was 84.61 (61.98-115.50) after 24 hrs and 32.87 (16.03-67.39) after 48hrs. While, LD₅₀ against T. castaneum was 547.08 (477.38 - 626.97) after 24 hrs and 452.51 (380.30 - 538.42) after 48hrs [87].

However, On the other hand, in studying of the role of jute leaf (Corchorus capsular) phytochemicals on feeding, growth and reproduction of Diacrisia cassinum Kollar (Lepidoptera: Arctiidae), it appeared that the larval and post larval developmental duration was shorter on mature jute leaf fed insects whereas adult longevity was higher in it (P < 0.05) relative to young and senescent leaf fed insects. Fecundity of D. castagnetum was also highest on mature leaves followed by young and senescent leaves. The growth and development of D. cassinum were related to the nutrient content relative to the secondary metabolites of these three types of jute leaves. Higher levels of nutritional factors (total carbohydrates, proteins, lipids, nitrogen and amino acids including water content) and lower levels of anti-nutritional factors (secondary metabolites) in mature jute leaves have influenced lower developmental time along with higher growth rate, fecundity and accumulated survivability of D. cassinum than the young and senescent leaves [88-89].

Cordia myxa

The anti-leishmanial activity of the mucilage extract of Cordia myxa was examined against promastigotes of L. infantum (MCAN/IR/96/LON49) and L. major (MRHO/IR/75/ER) (1x10⁵ cells/ml) They were seeded in a 96-well microtiter plate, in the presence of the serial concentrations (0, 0.61, 1.22, 2.44, 4.88, 9.75, 19.5, 39, 78, and 156 mg/ml) of the extract and then incubated at 24°C, for 72 hours. Antileishmanial activity was assayed by light microscopy and (3-(4,5-dimethylthiazol-2-yl)-2,5 diphenyl tetrazolium bromide) MTT method. The concentration inhibiting parasite growth by 50% (IC₅₀ value) was calculated with a sigmoid dose-response curve. Mucilage extract of Cordia myxa was active against promastigotes form of L. major and L. infantum, with an IC₅₀ of 26 ± 2.2 mg/ml and an IC₅₀ of 35 ± 2.2 mg/ml, respectively. The survival percentage of L. major and L. infantum promastigotes after 72 hours treatment appeared concentration dependent. Percentage of survival Leishmania major after 72 hours reached 17.68% in a concentration of 156 mg/ml, while the percentage of survival of L. infantum promastigotes after 72 hours reached 16.68% in a concentration of 156 mg/ml [90-91].

Cordia myxa were tested for antiplasmodial activity. Antimalarial effects were quantified with respect to inhibition of parasite growth, as measured by the production of Plasmodium lactate dehydrogenase. Alkaloids extract of Cordia myxa showed good antiplasmodial activity, IC₅₀ was 6.2 μg/ml, while dichloromethane extract of Cordia myxa showed moderate antiplasmodial activity IC₅₀ was 4.2 μg/ml, followed by aqueous and methanol extracts [92].

The crude alkaloid compounds for Cordia myxa leaves was tested against Culex pipines at (10, 7.5, 5, 2.5, 0) mg /ml. It possessed significant effect on some biological aspects of Culex pipines. The results showed that of eggs and larval stages (1⁰, 2⁰, 3⁰, 4⁰) was (13.38 , 0, 0, 0, 0) respectively in 10 mg /ml. At the same concentration, it also reduced productivity from 320 egg / female to 0 egg / female [93].
Coriandrum sativum

Commercial essential oils from 28 plant species were tested for their nematicidal activities against the pine wood nematode, Bursaphelenchus xylophilus. The best nematicidal activity against B. xylophilus was achieved with essential oils of coriander (Coriandrum sativum) [94].

In vitro anthelmintic activities of crude aqueous and hydro-alcoholic extracts of the seeds of Coriandrum sativum were investigated on the egg and adult nematode parasite Haemonchus contortus. The aqueous extract of Coriandrum sativum was also investigated for in vivo anthelmintic activity in sheep infected with Haemonchus contortus. Both extracts of Coriandrum sativum inhibited hatching of eggs completely at a concentration less than 0.5 mg/ml. ED₅₀ of aqueous extract of Coriandrum sativum was 0.12 mg/ml while that of hydroalcoholic extract was 0.18 mg/ml. There was no statistically significant difference between aqueous and hydroalcoholic extracts (p>0.05). The hydroalcoholic extract showed better in vitro activity against adult parasites than the aqueous one. For the in vivo study, sheep were artificially infected with Haemonchus contortus, crude aqueous extract of Coriandrum sativum was given at 0.45 and 0.9 g/kg dose levels. Efficacy was tested by faecal egg count reduction (FECR) and total worm count reduction (TWCR). On day 2 post treatment, significant FECR was detected in groups treated with higher dose of Coriandrum sativum (p<0.05) and albendazole (p<0.001). Significant (p<0.05) TWCR was detected only for higher dose of Coriandrum sativum compared to the untreated group. Reduction in male worms was higher than female worms. Treatment with both doses of Coriandrum sativum did not help the animals to improve or maintain their PCV, while those treated with albendazole showed significant increase in PCV (p<0.05) [95].

The antiparasitic efficacy of Coriandrum sativum essential oils was studied by two in vitro assays on Haemonchus contortus using egg hatch test (EHT) and larval development test (LDT). Coriandrum sativum essential oils exhibited a dose-dependent effect in the EHT, inhibiting 81.2% of H. contortus larvae hatching, at a concentration of 2.5 mg/ml. The effective concentration to inhibit 50% (EC₅₀) of egg hatching was 0.63 mg/ml. In LDT, Coriandrum sativum at concentration of 10 mg/ml inhibited 97.8% of H. contortus larval development [96].

The in vitro effect of fractions from Coriandrum sativum (coriander) on promastigotes and amastigotes of L. infantum was studied in addition to its toxicity against the murine monocytic cells RAW 264.7. All fractions were effective against L. infantum promastigotes and did not differ from the positive control pentamidine (p>0.05). However, the Coriandrum sativum methanol fraction, was the most effective against amastigotes and did not differ from the positive control amphotericin B (p>0.05) [97].

The biological activity of essential oil of Coriandrum sativum seeds was tested against adult Tribolium confusus Duval (Coleoptera: Tenebrionidae) and Callosobruchus maculatus F. (Coleoptera: Bruchidae) in a series of laboratory experiments. The mortality of 1-7 day old adults of the insect pests increased with concentration from 43 to 357 μl/l air and with exposure time from 3 to 24 h. In the probit analysis, LC₅₀ values showed that C. maculatus (LC₅₀ = 1.54 μl/l air) was more susceptible than T. confusus (LC₅₀ = 318.02 μl/l air) to seed essential oil of Coriandrum sativum [98].

The essential oil (EO) of the fruits of Coriandrum sativum was evaluated for its larvicidal and repellent activities against Aedes albopictus Skuse (Diptera: Culicidae). Coriandrum sativum EO exerted toxic activity against A. albopictus larvae: LC₅₀ was 421 ppm, while LC₉₀ was 531.7 ppm. Repellence trials highlighted that Coriandrum sativum EO was a good repellent against A. albopictus, RD₉₀ was 0.0001565 μl/cm² of skin, while RD₉₀ was 0.002004 μl/cm². At the highest dosage (0.2 μl/cm² of skin), the protection time achieved with Coriandrum sativum essential oil was higher than 60 min [99].

The leaf oil had significant toxic effects against the larvae of Aedes aegypti with an LC₅₀ value of 26.93 ppm and an LC₉₀ value of 37.69 ppm, and the stem oil has toxic effects against the larvae of A. aegypti with an LC₅₀ value of 29.39 ppm and an LC₉₀ value of 39.95 ppm [100].

The seed oil had significant toxic effects against the larvae of Aedes aegypti with an LC₅₀ value of 21.55 ppm and LC₉₀ value of 38.79 ppm. The major components in the essential oil of coriander play an important role as immunotoxicity on the A. aegypti [101].

Coronilla scorpioides

Screening of Ae. aegyptii larvicidal activity of 110 selected Egyptian plants proved that Coronilla scorpioides exhibited highest larvicidal activity, calculated as 22.53 ± 2.01 mg% for aqueous extracts and 18.53 ± 1.95 mg% for methanol extract [102].

Coronilla varia

A group of 3-nitropropanoyl-D-glucopyranosases was also isolated from active fractions of the crude extracts of the root. These compounds were toxic when administered orally to 3rd instar Costelytra zealandica larvae [103].

Crocus sativus

The effectiveness of Crocus sativus and its apoptotic activity against Leishmania major...
promastigotes was studied using MTT assay to find viability of L. major promastigotes and the results were explicated as IC₅₀ (50% inhibitory concentration). ED₉₀ (50% effective doses) for L. major amastigotes were also analyzed. Annexin-V FLUOS staining was performed to study the cell death properties of saffron by using FACS analysis. Qualitative analysis of the DNA fragmentations was accomplished by agarose gel electrophoresis, and light microscopy was used to observe morphological changes of promastigotes. The results revealed that L. major promastigotes and amastigotes are sensitive to saffron at different concentrations and time dependent manner, with apoptotic features including DNA laddering, cytoplasmic shrinkage, and externalization of phosphatidylserine. IC₅₀ and ED₉₀ of this extract after 48 h of incubation was 0.7 and 0.5 mg/ml respectively [104-105].

Safranal isolated from Coccus sativus extract exhibited insecticidal and pesticidal effect. This fact could present saffron as safe and effective herbal insecticide and pesticide which was more environment friendly than other synthetic insecticides [106].

**Cuminum cuminum**

The electrophysiological, behavioural (repellency, irritancy) and toxic effects of the essential oils was studied against Anopheles gambiae strain (Kisumu). Aldehydes elicited the strongest responses and monoterpenes the weakest responses in electroantennogram (EAG) trials. However, EAG responses did not correlate consistently with results of behavioral assays. In behavioral and toxicity studies, several of the single compounds exhibited repellency, irritancy or toxicity in An. gambiae; however, the activity of essential oils did not always correlate with activity expected from the major components. The biological activity of essential oils appeared complex, suggesting interactions between individual compounds and the insect. Data also indicated that the three effects appeared independent, suggesting that repellency mechanism(s) may differ from mechanisms of irritancy and toxicity [107-108].

Fumigant activity of essential oil vapours distilled from cumin was recorded against the eggs of two stored-product insects, the confused flour beetle, Tribolium confusum, and the Mediterranean flour moth, Ephesia kuehniella. The exposure to vapours of essential oils resulted in 100% mortality of the eggs at a concentration of 98.5 μl cumin essential oil/l air [109-110].

**Cupressus sempervirens**

The ethanol extract of the powdered cones of Cupressus sempervirens, collected from Oxford, Mississippi, exhibited potent antiparasitic activities. Bioassay-guided fractionation using a centrifugal preparative thin-layer chromatography led to isolation of many diterpenes, 6-deoxytaxodione (11-hydroxy-7, 9(11), 13-abietatrien-12-one), taxodione, ferruginol and sugiol. 6-deoxytaxodione (11-hydroxy-7, 9(11), 13-abietatrien-12-one) and taxodione, displayed potent anti-leishmanial activity with half-maximal inhibitory concentration (IC₅₀) values of 0.077 μg/ml and 0.025 μg/ml, respectively, against Leishmania donovani promastigotes, compared to those of the standard anti-leishmanial drugs, pentamidine (IC₅₀ 1.62 μg/ml) and amphotericin B (IC₅₀ 0.11 μg/ml) [111-112].

Ethanolic, acetone and petroleum ether extracts of leaves from the Egyptian Cupressus sempervirens were tested against 3rd instar larvae of the mosquito Culex pipiens. The obtained results indicated that petroleum ether extracts were more efficient than ethanolic and acetone extracts. The toxicity, based on LC₅₀ values, were: ethanol (LC₅₀ 263.6 ppm) > acetone extract (LC₅₀ 104.3 ppm) > petroleum ether extract (LC₅₀ 37.8 ppm). A remarkable reduction in both the pupation percent and adult emergence was obtained. Moreover, all extracts exerted a delayed toxic effect on the pupae and adults after treatment of larvae. Furthermore, various degrees of morphogenic abnormalities were observed in the immature and adult stages [113].

**Cymbopogon schoenanthus**

The anthelmintic potential of Cymbopogon schoenanthus essential oil was evaluated in lambs experimentally infected with Haemonchus contortus. Two-month-old lambs with mean body weight (BW) of 22.5 kg were experimentally infected with a multidrug-resistant Haemonchus contortus strain. Infected animals were dosed orally with Cymbopogon schoenanthus essential oil. Eighteen animals were allocated into three groups of six animals, and each received one of the following treatments: Group 1 - control (10 ml of water), Group 2 - Cymbopogon schoenanthus essential oil (180 mg/kg bw); and Group 3 - Cymbopogon schoenanthus essential oil (360 mg/kg bw). Animals received the oil once a day for 3 consecutive days. Lambs were evaluated clinically for blood biochemistry before and at 1, 5, 10, 15, 20 days after treatment. No statistically significant reduction in fecal egg count, packed cell volume or total worm count was observed after treatments. Also, no statistical difference among group means for blood levels of urea, creatinine, albumin, alkaline phosphatase, aspartate aminotransferase and gamma glutamyl transferase was found. Larval development assay (LDA) and egg hatch assay (EHA) were performed from feces of treated animals at 1, 5, 10 and 15 days after essential oil administration. An inhibition in LDA was observed 1 day after the 3-day treatment in larvae from feces of animals treated with 360 mg/kg essential oil [113].

Cymbopogon schoenanthus essential oils were evaluated against developmental stages of trichostrongyliids from sheep naturally infected (95%
**Haemonchus contortus** and 5% *Trichostrogyulus* spp.) using egg hatch assay (EHA), larval development assay (LDA), larval feeding inhibition assay (LFIA), and larval exsheathment assay (LEA). *Cymbopogon schoenanthus* essential oil showed a good activity against ovine trichostrongylids. It had LC_{50} value of 0.045 mg/ml in EHA, 0.063 mg/ml in LDA, 0.009 mg/ml in LFIA, and 24.66 mg/ml in LEA [114].

The insecticidal activity of crude essential oil extracted from *Cymbopogon schoenanthus* and its main constituent (piperitone), was assessed on different developmental stages of *Callosobruchus maculatus*. Piperitone was more toxic to adults with LC_{50} value of 1.6 microl/l vs. 2.7 microl/l obtained with the crude extract. Piperitone inhibited the development of newly laid eggs and of neonate larvae, but was less toxic than the crude extract to individuals developing inside the seeds [115].

*Cymbopogon schoenanthus* essential oils from Benin Republic in west Africa displayed about 100% mortality rate against adult *Anopheles gambiae* [116].

The efficacies of essential oils of nine plant species, which were traditionally used to avoid mosquito bites in Benin, were investigated. These oils were tested on susceptible “kisunu” and resistant “ladji-Cotonou” strains of *Anopheles gambiae*. The results showed that *Cymbopogon schoenanthus* was a potential promising plant sources alternative to pyrethroids, for the control of the *Anopheles malaria* vector in Benin. The efficacy of essential oil was possibly attributed to its chemical composition in which major and/or minor compounds have been shown insecticidal activities on various pests and disease vectors such as *Anopheles* [117].

The effect of camelgrass (*Cymbopogon schoenanthus*) oil on *Anopheles* mosquito and its larvae was tested to evaluate its repellency property. Different quantity of the oil extract viz: 10ml, 5ml and 1ml was introduced into two set of twelve beakers each containing twenty larvae and adult mosquito. Mortality rate was recorded at certain time interval. Application of the oil extract on adult mosquitoes larva caused 100% mortality. The maximum mortality time taken was 15 minutes for the adult mosquito and 18 minutes for the larvae. The minimum mortality time taken was 3 minute. The rapid mortality recorded in respect to both larvae and adult of anopheles mosquito indicated high insecticidal and larvicidal properties of the chemical compounds present in the oil of the grass species [118].

The efficacy of 3% citronella candles and 5% citronella incense were evaluated in protecting subjects from bites of *Aedes* spp under field conditions. The study was conducted in a deciduous woodlot in Guelph, Ontario, Canada. Eight subjects, dressed identically, were assigned to one of 8 positions on a grid within the study area. Two citronella candles, 2 citronella incense, 2 plain unscented candles, or no candles (i.e., nontreated controls) were assigned to 2 positions on the grid each evening. Subjects conducted 5-min biting counts at each position and performed 16 biting counts per evening. On average, subjects received 6.2 ± 0.4, 8.2 ± 0.5, 8.2 ±0.4, and 10.8 ± 0.5 bites/ 5 min at positions with citronella candles, citronella incense, plain candles, and no candles, respectively. Although significantly fewer bites were received by subjects at positions with citronella candles and incense, than at non-treated locations, the overall reduction in bites provided by the citronella candles and incense was only 42.3 and 24.2%, respectively [119].

The insecticidal properties of the aerial part of *Cymbopogon schoenanthus* was studied experimentally. Cabbage plants were sprayed with the aqueous extracts of *Cymbopogon schoenanthus* leaves as treatment, and the damage levels of *Plutella xylostella* was assessed. *In vitro*, the emulsified essential oil concentrations were used in a contact test on the larvae in order to assess the mortality effects. The larvae survival time was only 22 seconds with *Cymbopogon schoenanthus* emulsified oil treatment (2 g/l), whilst it exceeded 44,100 seconds (over 12 hours) for the dimethoate. The nutrition test showed that at 48 h period, a significant effectiveness against larvae was observed with emulsified oil treatment 2 g/l (60% mortality) versus 10% of mortality for dimethoate. The authors concluded that *Cymbopogon schoenanthus* can validly be used as alternative in *P. xylostella* management. The results of the field experiments showed no significant difference between the treatments and the control in terms of marketable cabbages harvested [120].

**Cynodon dactylon**

Anthelmintic activity of petroleum ether, methanol, and water extracts of *Cynodon dactylon* was evaluated on adult Indian earthworm *Pheretima posthuma* with the using of albendazole as a standard drug. The aqueous extract of *Cynodon dactylon* exerted anthelmintic activity in comparison with the standard drug [121].

The of mosquito repellents activity of volatile oils of *Cynodon dactylon* was studied against (A. aegypti). The distillates of the fruits of *Cynodon dactylon* was effective for 3 hours. The mixture of *C. papaya* and *Cynodon dactylon* was effective for 2.5 hours compared to that of *C. papaya* (2.5 hours) alone or *Cynodon dactylon* (1.5 hours) alone [122].

**Cyperus rotundus**

Hexane extract of tuber of plant *Cyperus rotundus* was tested for repellent activity against mosquito vector *Anopheles culicifacies*, *Anopheles stephensi* and *Culex quinquefasciatus*. Results showed that the tuber extracts were effective
for repellency of the entire mosquito vector even at a low dose [123].

_Cyperus rotundus_ was more effective insecticidal than carbamate and has almost the same efficacy as that of organophosphate. Result showed that all the test ants died after 10s, while organophosphate ranked second with 9 ants dead after 10s, and the carbamate ranked third with seven ants dead after 12s [124].

The ovicidal and larvicidal efficacy of essential oils of the tubers of _Cyperus rotundus_ was studied on eggs and fourth instar larvae of _Aedes albopictus_. The eggs and larvae were exposed to serial concentration of the oils ranging from 5-150 ppm and observed for 24 h. Oils showed remarkable ovicidal and larvicidal activities indicated by EC50 values of <5 ppm and LC50 values of <20 ppm [125].

Activity-guided investigation of _Cyperus rotundus_ tubers led to the isolation of patchoulolene, carphyllene alpha-oxide, 10,12-peroxycalamenene and 4,7-dimethyl-1-tetralone. The antimalarial activities of these compounds were in the range of EC50 10−4 to10−6 M, with the novel endoperoxide sesquiterpene, 10,12-peroxycalamenene, exhibiting the strongest effect at EC50 2.33 × 10−6 M [126].

**Conclusion:** The paper reviewed the antiparasitic, antiprotozoal, molluscicidal and insecticidal effects of the medicinal plants to open the door for their utilization in medical applications as a result of effectiveness and safety.

**References:**
1. Al-Snafi AE. Study of drugs prescribing pattern of specialists and general practitioners in Tikrit city . Med. J. Tikrit Univ. 1997, 3, 12-17 .
2. Kadir, MA , Al- Snafi, AE and Farman, NA .Comparison between the efficacy of sulphur and garlic in treatment of scabies. Med. J. Tikrit Univ. 1999, 5, 122-125 .
3. Farjou, IB, and Al – Snafi, AE . A novel treatment for hepatic hydatid cysts, combination of therapy of praziquantel and methotrexate with Albendazole . J. Fac. Med. (Baghdad) 2000, 42(3) 570-578 .
4. Kadir, MA. and Al-Snafi, AE . Epidemiology of scabies in Tuz district, J. Fac. Med. (Baghdad) 2000, 42(2) 321-329 .
5. Farjou, IB and Al-Snafi, AE. Some protozocidical alternatives to formalin for E. granulosus hydatid cysts in man J. Fac. Med. 2001, 43(1) 81-85 .
6. Al-Gazi ZS, Al-Snafi AE, Al-Abady FA. Effect of toxoplasmosis and/or its treatment (sulpadazine and pyrimetamine) on female rats reproductive performance. Indian Journal of Pharmaceutical Science & Research 2016; 6(1): 35-40.
7. Al-Ghezy ZS, Al-Abady FA, Al-Snafi AE. Effect of toxoplasmosis and its treatments on male rats reproductive functions Asian Journal of Pharmaceutical Science & Technology 2016; 6(2): 82-88.
8. Al-Ghezy ZS, Al-Abady FA, Al-Snafi AE. Histological effects of toxoplasmosis and its treatments on male and female rats. American Journal of Pharmacy & Health Research 2016; 4(4): 40-52.
9. Al-Snafi, AE., Kdir , MA., and Al-Batat, H.A . Evaluation of 5 days single and combined treatment of intestinal amebiasis . J Tikrit univer 2001; 7(2): 158-162.
10. Al-Snafi AE. Antimicrobial drugs. Al Diaa Publication house, Iraq 2013.
11. Al- Snafi AE. Pharmacology and therapeutics. Al Diaa Publication house, Iraq 2013.
12. Al-Snafi AE. Therapeutic properties of medicinal plants: a review of their antiparasitic, antiprotozoal, molluscicidal and insecticidal activity (part 1). J of Pharmaceutical Biology 2015; 5(3): 203-217.
13. Al-Snafi AE. Chemical constituents and pharmacological activities of Milfoil (Achillea santolina) - A Review. Int J Pharm Tech Res 2013, 5(3): 1373-1377.
14. Al-Snafi AE. The pharmacological importance of _Ailanthus altissima_ - A review. International Journal of Pharmacy Review and Research 2015; 5(2):121-129
15. Al-Snafi AE. Pharmacological effects of _Allium_ species grown in Iraq. An overview. International Journal of Pharmaceutical and health care Research 2013;1(4):132-147.
16. Al-Snafi AE. Chemical constituents and pharmacological activities of _Ammi majus_ and _Ammi visnaga_. A review. International Journal of Pharmacy and Industrial Research 2013;3 (3):257-265.
17. Al-Snafi AE. The pharmacological Importance of _Antirrhinum majus_ - A review. Asian J of Pharm Sci & Tech 2015; 5(4): 313-320.
18. Al-Snafi AE. The Pharmacology of _Apium graveolens_. - A review. International Journal for Pharmaceutical Research Scholars 2014; 3(1-1): 671-677.
19. Al-Snafi AE. Chemical constituents and pharmacological activities of _Arachis hypogaea_. – A review. International Journal for Pharmaceutical Research Scholars 2014; 3(1-1): 615-623.
20. Al-Snafi AE. The pharmacological importance of _Artemisia campestris-_ A review. Asian Journal of Pharmaceutical Research 2015;5(2): 88-92.
21. Al-Snafi AE. The constituents and biological effects of _Arundo donax_- A review. International Journal of Phytopharmacy Research 2015; 6(1): 34-40.
22. Al-Snafi AE. Chemical constituents and pharmacological effects of _Asclepias curassavica_ –
Al-Snafi AE., Sch. Acad. J. Pharm., June 2016; 5(6):194-207

A review. Asian Journal of Pharmaceutical Research 2015; 5(2): 83-87.
23. Al-Snafi AE. The Pharmacological Importance of *Ballota nigra* – A review. Ind J of Pharm Sci & Res 2015; 5(4): 249-256.
24. Al-Snafi AE. The Pharmacological importance of *Bauhinia variegata*. A Review. International Journal of Pharma Sciences and Research 2013; 4(12): 160-164.
25. Al-Snafi AE. The medical importance of *Betula alba* - An overview. Journal of Pharmaceutical Biology 2015; 5(4): 254-256.
26. Al-Snafi AE. Chemical constituents and pharmacological importance of *Bidens tripartitus* - A review. Ind J of Pharm Sci & Res 2015; 5(4): 257-263.
27. Al-Snafi AE. The pharmacological importance of *Brassica nigra* and *Brassica rapa* grown in Iraq. J of Pharm Biology 2015; 5(4): 240-253.
28. Al-Snafi AE. The Chemical constituents and pharmacological effects of *Bryophyllum calycinum*. A review. Journal of Pharma Sciences and Research 2013; 4(12): 171-176.
29. Al-Snafi AE. Encyclopedia of the constituents and pharmacological effects of Iraqi medicinal plants. Rigi Publication 2015
30. Al-Snafi AE. Pharmacology and medicinal properties of *Caesalpinia crista* - An overview. International Journal of Pharmacy 2015; 5(2): 71-83.
31. Al-Snafi AE. The chemical constituents and pharmacological effects of *Calendula officinalis* - A review. Indian Journal of Pharmaceutical Science & Research 2015; 5(3): 172-185.
32. Al-Snafi AE. The constituents and pharmacological properties of *Calotropis procera* - An Overview. International Journal of Pharmacy Review & Research 2015; 5(3): 259-275.
33. Al-Snafi AE. Bioactive components and pharmacological effects of *Canna indica* - An Overview. International Journal of Pharmacology and toxicology 2015; 5(2):71-75.
34. Al-Snafi AE. The chemical constituents and pharmacological effects of *Capparis spinosa* - An overview. Indian Journal of Pharmaceutical Science and Research 2015; 5(2): 93-100.
35. Al-Snafi AE. The chemical constituents and pharmacological effects of *Carum carvi* - A review. Indian Journal of Pharmaceutical Science and Research 2015; 5(2): 72-82.
36. Al-Snafi AE. The therapeutic importance of *Cassia occidentalis* - An overview. Indian Journal of Pharmaceutical Science & Research 2015; 5 (3): 158-171.
37. Al-Snafi AE. The chemical constituents and pharmacological importance of *Celosia cristata* – A review. J of Pharm Biology 2015; 5(4): 254-261.
38. Al-Snafi AE. The chemical constituents and pharmacological effects of *Chenopodium album* - An overview. International J of Pharmacological Screening Methods 2015; 5(1): 10-17.
39. Gnadinger CB. Pyrethrum flowers supplement 1936-1945. McLaughlin Gormley King Co. Published: Minneapolis 1945.
40. Greenhill M. Pyrethrum production: Tasmanian success story. Chronica Horticulturae 2007;47(3): 5-8.
41. Coomber HE. The chemical evaluation of pyrethrum flowers. Pyrethrum Post 1948; 1 (1): 16-19.
42. Essig K and Zhao ZJ. Method Development and Validation of a High-Performance Liquid Chromatographic Method for Pyrethrum Extract. J Chromatogr Sci 2001; 39 (4): 473-480.
43. Casida JE and Quistad GB. Pyrethrum flowers: production, chemistry, toxicology, and uses. Oxford University Press, New York 1995.
44. Essig K and Zhao ZJ. Preparation and characterization of a Pyrethrum extract standard. LC/GC 2001; 19(7): 722-730.
45. Godin PJ, Sleeman RJ, Snarey M and Thain EM. The jasmonins, new insecticidally active constituents of *Chrysanthemum cinerariaefolium*. VIS J Chem Soc C 1966: 332-334.
46. Gridsa M, Carovic-Stamkko K, Kolak I and Satovic Z. Morphological and biochemical diversity of Dalmatian Pyrethrum (*Tanacetum cinerariaefolium* (Trevir.) Sch. Bip.). Agriculturae Conspectus Scientificus 2009;74(2):73-80.
47. Matsuda K. Pyrethrin biosynthesis and its regulation in *Chrysanthemum cinerariaefolium*. Top Curr Chem 2012;314:73-81.
48. Matsuda K, Kikuta Y, Haba A, Nakayama K, Kasuda Y, Hatanaka A and Komai K. Biosynthesis of pyrethrin I in seedlings of *Chrysanthemum cinerariaefolium*. Phytochemistry 2005; 66(13):1529-1535.
49. Kueh JS, Mackenzie IA and Pattenden G. Production of chrysanthemic acid and pyrethrins by tissue cultures of *Chrysanthemum cinerariaefolium*. Plant Cell Rep 1985; 4(3):118-119.
50. Pyrethrum, Antonia Glynne-Jones of the Gooddeed Chemical Company in Aylesbury, UK, describes a unique biopesticide. Pesticide Outlook, http://www. researchinformation.co.uk/pest/2001/B108601B.PDF (October 2001).
51. Casida JE (ed). Pyrethrum, the natural insecticide. Academic Press 1973.
52. Mbaria JM, Maitho TE, Mitema ES and Muchiri DJ. Comparative efficacy of pyrethrum marc with albendazole against sheep gastrointestinal nematodes. Trop Anim Health Prod 1998;30(1):17-22.
53. Miller MC, Duckett SK and. Andrae JG. The effect of forage species on performance and gastrointestinal nematode infection in lambs. Small Ruminant Research 2011; 95(2-3): 188-192.
54. Marley CL, Cook R, Keatinge R, Barrett J and Lampkin HN. The effect of birdsfoot trefoil (Lotus corniculatus) and chicory (Cichorium intybus) on parasite intensities and performance of lambs naturally infected with helminth parasites. Veterinary Parasitology 2003; 112(1-2): 147-155.

55. Molan AL, Duncan AJ, Barry TN and McNabb WC. Effects of condensed tannins and crude sesquiterpene lactones extracted from chicory on the motility of larvae of deer lung worm and gastrointestinal nematodes. Parasitol Int 2003;52(3):209-218.

56. Ali Esmaal Al-Snafi. Medical importance of Cichorium intybus – A review IOSR Journal Of Pharmacy 2016; 6(3): 41-56.

57. Foster JG, Cassida KA and Turner KE. In vitro analysis of the anthelmintic activity of chicory (Cichorium intybus L.) sesquiterpene lactones against a predominantly Haemonchus contortus egg population. Vet Parasitol 2011;180(3-4):298-306.

58. Leecqrc E. Determination of lactucin in roots of chicory (Cichorium intybus L.) by high-performance liquid chromatography. Journal of Chromatography A 1984; 283: 441-444.

59. Bischoff TA, Kelley CJ, Karchesy Y, Laurantos M, Nguyen-Dinh P and Arefi AG. Antimalarial activity of lactucin and lactucopirin: sesquiterpene lactones isolated from Cichorium intybus L. Journal of Ethnopharmacology 2004; 95(2-3):455–457.

60. Al-Harmni KIF, Rahemo ZIF and Al-Khan HIA. The effect of two fruit extracts and drugs on the liver of albino mice with induced leishmaniasis. Trends in Parasitology Research 2012; 1(3): 16-26.

61. Ali Esmaal Al-snafi. Chemical constituents and pharmacological effects of Citrus colocynthis - A review. IOSR Journal Of Pharmacy 2016; 6(3): 57-67.

62. Rahuman AA and Venkatesan P. Larvicidal efficacy of five cucurbitaceous plant leaf extracts against mosquito species. Parasitol Res 2008; 103(1):133-139.

63. Rahuman A, Venkatesan P and Gopalakrishnan G. Mosquito larvicidal activity of oleic and linoleic acids isolated from Citrus colocynthis (Linn.) Schrad. Parasitol Res 2008;103(6):1383-1390.

64. Torkey HM, Abou-Yousef HM, Abdel Azeiz, AZ and Farid HEA. Insecticidal effect of curcubitacin E glycoside isolated from Citrullus colocynthis against Aphis craccivora. Australian Journal of Basic and Applied Sciences 2009; 3(4): 4060-4066.

65. Fatima LD and Mohamed K. Phytotherapy as new approach to treat scorpion envenomation: Experimental study. IJPSR 2014; 5(5): 1682-1692.

66. Kabra AO, Bairagi GB, Mandade RJ and Sakarkar DM. Comparative studies on anthelmintic activity of Citrus medica L. and Ocimum basilicum L. International Journal of Pharmaceutical Sciences Review and Research 2011; 8(1): 6-8.

67. Bairagi GB, Kabra AO and Mandade RJ. Anthelmintic activity of Citrus medica L. leaves in Indian adult earthworm. International Journal of PharmTech Research 2011; 3(2):664-667.

68. Rajarajan AT, Vijayasree VG, Kenichi W, Kumar SV, Narasimman G and kumar SS. Anthelmintic and antimicrobial properties of peels of Citrus sinensis. Pharmacologyonline 2009; 1: 363-368.

69. Muala BA, Muhii RR and Jaber MA. Effect of the aqueous extract of Citrus medica on protozoocides and Gram positive and Gram negative bacteria in vitro. Al-Nahrain Univ Journal 2008; 11(2): 16-19.

70. Raj RK. Screening of indigenous plants for anthelmintic action against human Ascaris lumbricoides: Part-II. Indian J Physiol Pharmacol 1974;18(2):129-131.

71. Kumar S, Warikoo R, Mishra M, Seth A and Wahab N. Larvicidal efficacy of the Citrus limetta peel extracts against Indian strains of Anopheles stephensi Liston and Aedes aegypti L. Parasitol Res 2012;111(1):173-178.

72. Effiom OE, Avoaja DA and Ohaer, C C. Mosquito repellent activity of phytochemical extracts from peels of Citrus fruit species. Global Journal of Science Frontier Research 2012; 12(1): 1-8.

73. Chedekal AN. Effect of four leaf extracts on egg hatching and juvenile mortality of root knot nematode Meloidogyne incognita. International Journal of Advanced Life Sciences 2013; 6(1):68–74.

74. Al-Snafi AE. Chemical constituents and pharmacological effects of Clerodendrum inerme- A review. SMU Medical Journal 2016; 3(1): 129-153.

75. Patil PB, Kallapur YL and Holihosur SN. Evaluation of Clerodendron inerme Gaertn. plant extract against Aedes aegypti L. mosquito. International Journal of Natural Products Research 2013; 2(2): 36-38

76. Kovenden K and Murugan K. Effect of medicinal plants on the mosquito vectors from the different Agroclimatic regions of Tamil Nadu, India. Advances in Environmental Biology 2011; 5(2): 335-344.

77. Patil PB, Holihosur SN and Kallapur VL. Efficacy of natural product, Clerodendron inerme against dengue mosquito vector Aedes aegypti. Current Science 2006; 90(8): 1064-1066.

78. Venkatachalam MR and Jebanesan A. Screening of repellent activity of certain plants of Tamil Nadu. India. Convergence 2001; 3(1):39-43.

79. Morya K, Pillai S and Patel P. Effect of powdered leaves of Lantana camara, Clerodendrum inerme and Citrus limon on the rice moth, Coccrya cephalonica. Bulletin of Entomology 2010; 63 (2): 183-189.

80. Arya MC, Kadabinakatti SK and Kumar R. Study of efficacy of Clerodendron inerme Gaertn. leaf
extract against Pieris brassicae (Linnaeus). Nat Sci 2014; 12(9): 22-24.
81. Shekhawat N and Vijayvergia R. Anthelmintic activity of extracts of some medicinal plants. International Journal of Computational Science and Mathematics 2011;3(2):183-187.
82. Al-Snafi AE. Pharmacological importance of Clitoria ternatea – A review IOSR Journal Of Pharmacy 2016; 6(3): 68-83.
83. Nirmal SA, Bhalke RD, Jadhav RS and Tambe VD. Anthelmintic activity of Clitoria ternatea. Pharmacologyonline 2008; 1: 114-119.
84. Salhan M, Kumar B, Tiwari P, Sharma P, Sandhar HK and Gautam M. Comparative anthelmintic activity of aqueous and ethanolic leaf extracts of Clitoria ternatea. Int J Drug Dev & Res 2011; 3 (1):68-69.
85. Mathew N, Anitha MG, Bala TS, Sivakumar SM, Narmadha R and Kalyanasundaram M. Larvicidal activity of Saraca indica, Nyctanthes arbor-tristis, and Clitoria ternatea extracts against three mosquito vector species. Parasitol Res 2009; 104(5):1017-1025.
86. Elangovan A, Dhanasekaran S, Anandan A, Krishnappa K, Gokulakrishnan J and Elumalai K. Mosquitoicidal activities of Corchorus capsularis L ( Malvaceae) against a common malaria vector, Anopheles stephensi (Liston) and a dengue vector Aedes aegypti (L) (Diptera: Culicidae). International Journal of Recent Scientific Research 2012; 3(6): 564-568.
87. Talukder D and Khanam LAM. Toxicity of four plant based products against three stored product pests. J Bio Sci 2009; 17: 149-153.
88. Roy N. Role of Chorchorus capsularis phytochemicals on the feeding dynamics of Diacrisia casignetum Kollar (Lepidoptera: Arctiidae). Journal of Entomology and Zoology Studies 2014; 2 (4): 227-236.
89. Al-Snafi AE. The contents and pharmacological importance of Corchorus capsularis- A review. IOSR Journal of Pharmacy 2016; 6(6): 58-63.
90. Saki J, Khademvatan S, Pazyar N, Eskandari A, Tamoradi A and Nazari P. In vitro activity of Cordia myxa mucilage extract against Leishmanina major and L. infantum promastigotes. Jundishapur J Microbiol 2015;8(3):e19640.
91. Al-Snafi AE. The Pharmacological and therapeutic importance of Cordia myxa- A review. IOSR Journal of Pharmacy 2016; 6(6): 47-57.
92. Sanon S, Gansane A, Ouattara LP, Traore A, Ouedraogo IN, Tiono A, Taramelli D, Basilico N and Sirima SB. In vitro antiplasmodial and cytotoxic properties of some medicinal plants from western Burkina Faso. Afr J Lab Med 2013;2(1)
93. Rashid YD,Al-Zubaidi FSH and Zaidan HK. Effect of crude alkaloid extract of the leaves of C. myxa on some biological aspects of Culex pipienes. Babylon Scientific Journal of Pure and Applied Sciences 2015; 23(1):55-62.
94. Kim J, Seo SM, Lee SG, Shin SC and Park IK. Nematicidal activity of plant essential oils and components from coriander (Coriandrum sativum), Oriental sweetgum (Liquidambar orientalis), and valerian (Valeriana wallichii) essential oils against pine wood nematode (Bursaphelenchus xylophilus). J Agric Food Chem 2008; 56(16): 7316-7320.
95. Eguale T, Tilahun G, Debelaa A, Feleke A and Makonnen E. In vitro and in vivo anthelmintic activity of crude extracts of Coriandrum sativum against Haemonchus contortus. J Ethnopharmacol 2007;110(3):428-433.
96. Macedo IT, de Oliveira LM, Camurça-Vasconcelos AL, Ribeiro WL, dos Santos JM, de Morais SM, de Paula HC and Bevilaqua CM. In vitro effects of Coriandrum sativum, Tagetes minuta, Alpinia zerumbet and Lantana camara essential oils on Haemonchus contortus. Rev Bras Parasitol Vet 2013; 22(4): 463-469.
97. Rondon FC, Bevilaqua CM, Accioly MP, Morais SM, Andrade-Junior HF, Machado IK, Cardoso RP, Almeida CA, Queiroz-Junior EM and Rodrigues AC. In vitro effect of Aloe vera, Coriandrum sativum and Ricinus communis fractions on Leishmania infantum and on murine monocytic cells. Vet Parasitol 2011; 178(3-4):235-240.
98. Khani A and Rahdari T. Chemical composition and insecticidal activity of essential oil from Coriandrum sativum seeds against Tribolium confusum and Callosobruchus maculatus. ISRN Pharm 2012;2012:263517. doi: 10.5402/2012/ 263517.
99. Benelli G, Flamini G, Fiore G, Cioni PL and Conti B. Larvicidal and repellent activity of the essential oil of Coriandrum sativum L. (Apiaceae) fruits against the filariasis vector Aedes albopictus Skuse (Diptera: Culicidae). Parasitol Res 2013; 112(3): 1155-1161.
100. Chung IM, Ahmad A, Kim EH, Kim SH, Jung WS, Kim JH, Nayem A and Nagella P. Immunotoxicity activity from the essential oils of coriander (Coriandrum sativum) seeds. Immunopharmacol Immunotoxicol 2012; 34(3):499-503.
101. Chung IM, Ahmad A, Kim SJ, Naik PM and Nagella P. Composition of the essential oil constituents from leaves and stems of Korean Coriandrum sativum and their immunotoxicity activity on the Aedes aegypti L. Immunopharmacol Immunotoxicol 2012;34(1):152-156.
102. El-Hela AA, Abdel-Hady NM, Dawoud GT, Hamed AM and Morsy TA. Phenolic content, antioxidant potential and Aedes aegyptii ecological friend larvicidal activity of some selected Egyptian plants. J Egypt Soc Parasitol 2013; 43(1): 215-234.
103. Gnanasunderam C, Greenfield WJ, Williams EM and Wright HJ. Toxicity of nitro compounds from
Lotus pedunculatus to grass grub (Costelytra zealandica) (Coleoptera: Scarabaeidae). Journal of Chemical Ecology 1984; 10(1): 81-93.

104. Yousefi E, Eskandari A, Gharavi MJ and Khademvatan S. In vitro activity and cytotoxicity of Crocus sativus extract against leishmaniasis major (MRHO/IR/75/ER). Infect Disord Drug Targets 2014; 14(1): 56-60.

105. Al-Snafi AE. The pharmacology of Crocus sativus- A review. IOSR Journal of Pharmacy 2016; 6(6): 8-38.

106. Leffingwell JC. Saffron. Leffing Well Reports 2002; 2 (5):1-3.

107. Deletre E, Chandre F, Williams L, Duménil C, Menut C and Martin T. Electro-physiological and behavioral characterization of bioactive compounds of the Thymus vulgaris, Cymbopogon winterianus, Cuminum cyminum and Cinnamomum zeylanicum essential oils against Anopheles gambiae and prospects for their use as bednet treatments. Parasit Vectors 2015; 8: 316.

108. Al-Snafi AE. The pharmacological activities of Cuminum cyminum - A review. IOSR Journal of Pharmacy 2016; 6(6): 46-65.

109. Clark L. Dermal contact repellants for starlings: foot exposure to natural plant products. Journal of Wilderness Management 1998; 61(4): 1352–1358.

110. Tunc I, Berger BM, Erler F and Dagli F. Ovicidal activity of essential oils from five plants against two stored product insects. Journal of Stored Product Research 2000; 36: 161–168.

111. Zhang J, Rahman AA, Jain S, Jacob MR, Khan SI, Tekwani BL and Ilias M. Antimicrobial and antiparasitic abietane diterpenoids from Cupressus sempervirens. Neuropsychiatric Disease and Treatment 2012; 2:1-6.

112. Al-Snafi AE. Medical importance of Cupressus sempervirens- A review. IOSR Journal of Pharmacy 2016; 6(6): 66-76.

113. EL-Sheikh TMY, Hassan MI, Moselhy WA, Amer MS and Shehata AZ. Evaluation of the biological activity of some Cupressus sempervirens (Cupressaceae) extracts against the mosquito vector Culex pipiens L. (Diptera: Culicidae). Egypt Acad J biol Sci 2011; 4 (1): 33-48.

114. Katiki LM, Chagas AC, Bizzo HR, Ferreira JF and Amarante AF. Anthelminthic activity of Cymbopogon martini, Cymbopogon schoenanthus and Mentha piperita essential oils evaluated in four different in vitro tests. Vet Parasitol 2011; 183(1-2): 103-108.

115. Ketho G. Comparative effects of Cymbopogon schoenanthus essential oil and piperitone on Callosobruchus maculatus development. Fitoterapia 2006; 77(7-8): 506-510.

116. Nonviho G, Wotto VD, Noudogbessi J, Avlessi F, Akogbeto M and Sohoundholoue DC. Original research paper insecticidal activities of essential oils extracted from three species of poaceae on Anopheles gambiae spp, major vector of malaria. Sci Study Res 2010; 11: 411-420.

117. Bossou AD, Mangelinkx S, Yedomonhan H, Boko PM, Akogbeto MC, Kimpe ND, Avlessi F and Sohoundholoue DCK. Chemical composition and insecticidal activity of plant essential oils from Benin against Anopheles gambiae (Giles). Parasites & Vectors 2013; 6: 337-353.

118. Musa AR, Aleiro BL, Aleiro AA and Tafinta YI. Larvicidal and insecticidal effect of camelgrass (Cymbopogon schoenanthus) oil on Anopheles mosquito. Annals of Biological Sciences 2014; 2 (1):19-22.

119. Lindsay RL, Surgeoner GA, Heal JD and Gallivan GJ. Evaluation of the efficacy of 3% citronella candles and 5% citronella incense for protection against field populations of Aedes mosquitoes. J Am Mosq Control Assoc 1996; 12(2 Pt 1): 293-294.

120. Laba B, Nenonéné AY, Adjrha Y, Koba K, Poutouli W and Sandra K. Efficacy of Cymbopogon schoenanthus L. Spreng (Poaceae) extracts on diamondback moth damaging cabbage. J Biofertil Biopestici 2012; 3: 3-6.

121. Abhishek B and Thakur A. Anthelminthic activity of Cynodon dactylon. Journal of Pharmacognosy and Phytochemistry 2012; 1(3): 1-3.

122. Kazembe T and Makusha C. Evaluation of mosquito repellencies of Capsicum frutescens, Carica papaya and Cynodon dactylon extracts and extract mixtures. Bull Environ Pharmacol Life Sci 2012; 1(7): 34-40.

123. Singh SP, Raghavendra K and Dash AP. Evaluation of hexane extract of tuber of root of Cyperus rotundus Linn (Cyperaceae) for repellency against mosquito vectors. J Parasitol Res 2009; 1: 1-5.

124. Solita ES and Castor L. Phytocidal and pesticidal properties of barsanga (Cyperus rotundus Linn.). JPAIR Multidiscip J 2011; 6: 197-214.

125. Vivek K, Bhat SK. Ovicidal and larvicidal activities of Cyperus giganteus Vahl and Cyperus rotundus Linn essential oils against Aedes albopictus (Skuse). Natural Product Radioactive 2008; 7(5): 416-419.

126. Thebtaranonth C, Thebtaranonth Y, Wanauppathunkul S and Yuthavong Y. Antimalarial sesquiterpenes from tubers of Cyperus rotundus: structure of 10,12-peroxycalamene, a sesquiterpene endoperoxide. Phytochemistry 1995; 40:125-128.