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Review Article

The IL-2 Defect in Systemic Lupus Erythematosus Disease Has an Expansive Effect on Host Immunity

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1. Introduction

IL-2 is a multifunctional cytokine primarily produced by T cells and is necessary for T cell activation, proliferation, and contraction. It has been reported that production of IL-2 is decreased in patients with systemic lupus erythematosus (SLE) and this defect affects multiple aspects of host immunity [1]. SLE is a complex autoimmune disease characterized by autoantibody production and widespread inflammation. Patients with this chronic disease cycle between periods of controlled disease and disease flare involving various organs and tissue.

Defective IL-2 production is one of many factors involved in the immune dysregulation responsible for this disease. As reported in patients, an IL-2 defect is observed in various SLE murine models and murine models have provided much information about the relationship between IL-2 and SLE. Furthermore, other IL-2 family members (IL-15 and IL-21) have been shown to be dysregulated in SLE disease. It has been found that the transcriptional regulators responsible for the transcription or suppression of IL-2 production are imbalanced in SLE T cells [2] and this explains the reduced IL-2 levels found in SLE patients.

One of the consequences of decreased IL-2 production is a reduction in the number of regulatory T cells (T_{reg}) in these patients. T_{reg} cells control expansion of autoreactive T cells and therefore are important in inhibiting autoimmunity. The decreased IL-2 production in SLE patients affects additional immune functions other than T cell proliferation. Activation-induced cell death (AICD), an IL-2 regulated process which is necessary for the down-regulation of expanded T cell clones, is defective in SLE T cells [3, 4]. IL-2 is also necessary for CD8+ T cell effector functions and the development of CD8+ T cell memory [5]. A defect in CD8+ T cell cytotoxic function leads to an increased chance of intracellular infection which could be deadly for SLE patients.

2. Defective IL-2 Production in SLE

It has long been known that IL-2 production is defective in SLE patients. Early in vitro studies examined SLE T cell response to mitogens. T cells were stimulated with the mitogen phytohemagglutinin (PHA) and it was found that they were unable to produce high levels of IL-2; a greater defect was observed in T cells isolated from patients with active disease [6]. Interestingly, it was observed that PHA-activated SLE T cells respond to exogenous IL-2 stimulation, indicating that IL-2R expression is normal on these cells [7]. Furthermore, it has been suggested that the IL-2R is upregulated on T cells from SLE patients with active disease as compared to those with inactive disease [8]. However, it seems that the IL-2R found on these SLE T cells is not
the high affinity receptor and therefore these cells are not optimally responsive to exogenous IL-2 [9].

Why do SLE T cells produce less IL-2? IL-2 production is upregulated through an autocrine (or paracrine) loop therefore it would seem logical that a defect in the T cell receptor (TCR) would result in decreased transcription of IL-2. Indeed, one of the TCR components, the CD3ζ receptor (TCR) would result in decreased transcription of IL-2; therefore it would seem logical that a defect in the T cell is upregulated through an autocrine (or paracrine) loop [10].

SLE is the MRL/lpr mouse strain. These mice carry the lpr mutation in the apoptotic protein Fas. They spontaneously develop lupus-like disease beginning at twelve weeks of age and subsequently display skin lesions, lymphadenopathy, autoantibody production, and proteinuria and often succumb to death from glomerulonephritis between seventeen (female) to twenty-two weeks (male) of age. These mice exhibit lymphoproliferation by three months of age which is not commonly seen in patients with SLE. It was found that IL-2 production and responsiveness is decreased in MRL/lpr mice and as mice age (and develop severe disease), this defect becomes more pronounced [15]. Furthermore, as described for humans, mitogenic or antigenic stimulation of MRL/lpr T cells results in weak IL-2 production [15–17].

BXSB/Yaa mice develop lupus in male mice due to the Y-linked autoimmune accelerator (Yaa) gene. Similar to the MRL/lpr mice, the male BXSB/Yaa mice develop lupus rapidly and fifty percent will succumb to glomerulonephritis by five months of age [18]. Like the MRL/lpr mice, male BXSB mice display a defective IL-2 mitogenic response as early as six weeks of age [15].

The NZB × NZW F1 murine model develops lupus at a slower rate than the other strains mentioned. The female NZB × NZW F1 mice will succumb to disease at approximately eight months of age. Therefore, it is not surprising that NZB × NZW F1 T cells do not show a decreased mitogenic response until 7.5 months, as these mice develop lupus-like disease slower than the other two strains [15]. Amongst these commonly studied murine models of SLE, the defect in IL-2 production and responsiveness is similar to that observed in humans.

4. IL-2/IL-2R Deficient Mice

Much of what is known about IL-2 regulation of the immune response has been realized from studies using the IL-2 or IL-2R knockout mice. Mice lacking IL-2 or IL-2R initially appear normal, but by four weeks of age they develop spontaneous autoimmune disease. The early disease displayed by the IL-2−/− mice is characterized by splenomegaly, lymphadenopathy, and severe anemia. If they survive beyond nine weeks of age, they will develop irritable bowel disease leading to death [19, 20]. Similarly, IL-2Rβ−/− mice are runted by four weeks of age and develop splenomegaly, lymphadenopathy, and anemia caused by autolysis. It has also been noted that these mice have increased serum IgG1 levels and high concentrations of autoantibodies, but they do...
not develop irritable bowel disease. Nevertheless, these mice only survive about twelve weeks before succumbing to severe autoimmunity [21].

It has more recently been discovered that the lymphoproliferation observed in the IL-2/IL-2R−/− mice is due to the greatly reduced number of regulatory T cells (Treg) found in the periphery [22]. Similarly, regulatory T cells are decreased in SLE patients (discussed in Section 6). Unlike the knockout mice, SLE patients do not exhibit extreme lymphoproliferation but they do have some, albeit reduced, levels of IL-2. Nevertheless, IL-2/IL-2R deficient mice present with many of the other characteristics observed in SLE patients, such as autoantibody production, lymphadenopathy, and decreased Treg cells.

### 5. Other IL-2 Family Members and SLE

Since IL-2 is integral to maintaining a healthy immune system, it is not surprising there is some redundancy of IL-2 function in host immunity. The IL-2 superfamily consists of IL-2, IL-15, and IL-21 (Figure 2). The IL-2R and IL-15R share two of their three receptor chains (IL-2Rβ and γc) while the IL-21R shares the common gamma chain (γc) with the others and binds a unique receptor chain (IL-21R). All three cytokine receptors are associated with JAK1 and JAK3, but IL-2 and IL-15 signal primarily through STAT5, whereas IL-21 primarily signals through STAT3 homodimers.

![Figure 2: IL-2 superfamily. IL-2, IL-15, and IL-21 are closely related in receptor binding, and function. IL-2 and IL-15 bind trimeric receptors and two of the three chains (IL-2Rβ, γc) are the same, while each cytokine binds a unique third component of the receptor (IL-2α or IL-15α, respectively). The dimeric IL-21R consists of the γc chain, shared with IL-2R and IL-15R, and a unique chain, IL-21R. All three cytokines utilize the Janus kinases signaling molecules JAK1 and JAK3. IL-2 and IL-21 can activate STAT1/3/5 while IL-15 is only known to activate STAT5. While IL-2 and IL-21 can activate multiple STAT family members, IL-2 primarily signals through STAT3 homodimers.](image)

IL-2 superfamily. IL-2, IL-15, and IL-21 are closely related in receptor binding, and function. IL-2 and IL-15 bind trimeric receptors and two of the three chains (IL-2Rβ, γc) are the same, while each cytokine binds a unique third component of the receptor (IL-2α or IL-15α, respectively). The dimeric IL-21R consists of the γc chain, shared with IL-2R and IL-15R, and a unique chain, IL-21R. All three cytokines utilize the Janus kinases signaling molecules JAK1 and JAK3. IL-2 and IL-21 can activate STAT1/3/5 while IL-15 is only known to activate STAT5. While IL-2 and IL-21 can activate multiple STAT family members, IL-2 primarily signals through STAT3 homodimers.

levels of IL-15, regardless of disease state, and this may contribute to the pathogenesis of disease [24, 25]. IL-15 has also been observed in increased levels in synovial joints and shown to play a role in arthritic inflammation due to activation of synovial T cells [26].

IL-21 is produced by T cells and it affects the function of T, NK, NKT, and B cells [27]. IL-21 enhances NK and CD8+ T cell cytotoxicity, and overexpression of IL-21 results in an increase in memory CD8+ T cells [28]. IL-21 production is increased in some murine models of SLE [29] and particular polymorphisms of the IL-21 gene have been described in SLE patients [30]. It has been shown in the BXSB/Ya murine model that IL-21 is integral to SLE development as deletion of the IL-21R in these mice prevents disease progression [31]. While the function of IL-21 on T or NK cells may be redundant with IL-2 or IL-15, IL-21 uniquely promotes activation and plasma cell differentiation of peripheral B cells [32]. Patients with SLE have decreased IL-21R expression in their peripheral B cells and this is associated with nephritis and high levels of autoantibodies [33]. Additionally, IL-21 is also necessary for activation-induced death of expanded B cells [29, 32]. While these studies suggest a role for IL-15 and IL-21 in SLE pathogenesis, the significance of these cytokines in human SLE disease is not clear and warrants further study.

### 6. Regulatory T Cells (Treg) and SLE

Regulatory T cells (Treg) are necessary for maintaining tolerance to self antigens and they are able to do this by suppressing self-reactive T cells. Natural Treg cells develop in the thymus, but induced Treg cells can be derived from a subset of activated CD4+ T cells in the periphery. Development and homeostasis of the CD4+CD25+Foxp3+ Treg cell subset is dependent on IL-2. Mice lacking the Treg specific transcription factor Foxp3 completely lack Treg cells and are severely ill due to systemic autoimmunity. Decreased IL-2 leads to a decrease in Treg population with concurrent autoimmunity due in part to excessive lymphoproliferation. It should be noted that Treg cells are not completely dependent on IL-2 for development as evidenced by the low numbers of Treg cells present in IL-2/IL-2R knockout mice [34].

Treg cells are decreased in SLE and there is evidence to suggest that the number of Treg cells in SLE patients may inversely relate to the severity of disease [38]. It was observed that in pediatric SLE patients, those with active disease had fewer Treg cells and this inversely correlated with the Systemic Lupus Erythematosus Disease Activity Index (SLEDAI) score and serum anti-double-stranded DNA levels [39]. It has been reported (in adult samples) that expression of FOXP3 is decreased in Treg cells from SLE patients with active disease in Treg population with concurrent autoimmunity due in part to excessive lymphoproliferation. It should be noted that Treg cells are not completely dependent on IL-2 for development as evidenced by the low numbers of Treg cells present in IL-2/IL-2R knockout mice [34].

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observed in SLE patients, a definitive link between these two observations has not been made.

7. Activation-Induced Cell Death (AICD)

While IL-2 is necessary for T cell activation, it is also needed following clonal expansion to control lymphoproliferation of a T cell clone. Following antigen stimulation through the TCR, a specific clonal T cell population will expand to eliminate the foreign object. Once the antigen is no longer present, the process of AICD will destroy the excess effector cells that were expanded to fight the antigen. IL-2 is required for this controlled apoptotic mechanism, in conjunction with costimulation through CD95, TNFR1, and other costimulatory molecules [35, 36]. Furthermore, IL-2 regulates AICD in CD8+ T cells by downregulating IL-2R as this receptor chain is associated with the survival of the cell [37]. It has been found that SLE patients are more resistant to AICD due to decreased levels of intracellular TNFα and upregulation of COX2 [3, 4].

8. IL-2 and Cytotoxicity

IL-2 is necessary for the development of CD8+ T cell/NK cell cytotoxicity. These cell types are important for fighting infection, and cytotoxic T lymphocytes (CTLs) cells are crucial for the destruction of virally infected cells. Some insight into the role of IL-2 in CTL development comes from a derivative of the IL-2Rβ−/− mouse that has thymic transgenic expression of IL-2Rβ which results in mice lacking the characteristic autoimmunity observed in IL-2Rβ−/− mice, and contains CD8− T cell cytotoxicity [43, 44]. Additional support for the IL-2 requirement in CTL expansion and activity is apparent if we look at infection data from the IL-2−/− mice. Following viral infection of IL-2−/− mice with lymphocytic choriomeningitis virus (LCMV), a T cell response could be detected but it was significantly decreased, suggesting that IL-2 plays an important role in the expansion of virus-specific CTLs [45]. These observations indicate that IL-2 signaling is necessary for the development of cytotoxicity.

IL-2 upregulates the costimulatory molecule CD70 which is necessary for CD8 expansion. It has been demonstrated that when CD70 is blocked and IL-2 is administered, CTL expansion does not occur [46]. CD8+ T cells expressing the co-stimulatory molecule CD27 preferentially secrete IL-2 in response to antigen stimulation [47] and it has recently been shown that CD27-driven IL-2 production in virally infected mice stimulates CD8+ T cell survival in an autocrine manner [48]. These data demonstrate an important role for co-stimulatory molecules working in concert with IL-2 during CTL activation.

It is not clear how the IL-2 defect in SLE disease affects CTL development and function. Thus far, the data has been conflicting. While there is evidence suggesting a cytotoxic defect in some patients [41, 42], there is also data that asserts that perforin and granzyme B are upregulated in patients with active SLE disease [49]. As seen in many aspects of SLE disease, there may be heterogeneity amongst patients. A CD8+ T cell defect in SLE patients would be relevant because this may contribute to the increased mortality from infections reported in these patients.

9. Conclusions

IL-2 is important for regulating T cell immunity and therefore affects many aspects of immune responses. Not only is IL-2 dysregulated in SLE, but the IL-2 family members IL-15 and IL-21 are also expressed aberrantly. The decreased production of IL-2 in SLE patients most likely contributes to various immune defects such as decreased Treg production, decreased AICD, and potentially decreased CTL responses (Table 1). Interestingly, the decreased FOXP3 expression in SLE Treg cells is reversible which may be exploited for therapeutic development. A potential decrease in CTL activity is of utmost importance in SLE as decreased cytotoxicity would make patients more susceptible to intracellular infection. There is evidence that SLE patients have a higher incidence of Epstein Barr virus (EBV) infection [50] and it has been reported that SLE patients have forty times higher viral load and a greater repertoire of antigen-specific B cells for EBV viral proteins [51]. IL-2 deficiency affects multiple regulatory pathways in the host and in the case of SLE this contributes to the multifaceted dysregulation of the immune response.

| Table 1: Consequences of decreased IL-2 production in SLE disease. |
|---------------------------------------------------------------|
| (i) Decreased activation-induced cell death (AICD) [3, 4, 35–37]. |
| (ii) Decreased number and function of Treg cells [38–40]. |
| (iii) May lead to decreased cytotoxic activity [41, 42]. |

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