Formation of pre-metastatic bone niche in prostate cancer and regulation of traditional Chinese medicine

Chiwei Chen, Renlun Huang, Jianfu Zhou, Lang Guo and Songtao Xiang*

The Second Clinical College of Guangzhou University of Chinese Medicine, Guangdong Provincial Hospital of Chinese Medicine, Guangzhou, China

Prostate cancer with bone metastasis has a high cancer-specific mortality. Thus, it is essential to delineate the mechanism of bone metastasis. Pre-metastatic niche (PMN) is a concept in tumor metastasis, which is characterized by tumor-secreted factors, reprogramming of stromal cells, and immunosuppression by myeloid-derived suppressor cells (MDSC), which is induced by bone marrow-derived cells (BMDC) in the target organ. However, PMN does not explain the predilection of prostate cancer towards bone metastasis. In this review, we discuss the initiation of bone metastasis of prostate cancer from the perspective of PMN and tumor microenvironment in a step-wise manner. Furthermore, we present a new concept called pre-metastatic bone niche, featuring inherent BMDC, to interpret bone metastasis. Moreover, we illustrate the regulation of traditional Chinese medicine on PMN.

KEYWORDS
pre-metastatic niche, bone metastasis, tumor microenvironment, prostate cancer, Traditional Chinese medicine

1 Introduction

Prostate cancer (PCa) has gradually become a major threat to humans. Recent reports reveal that 1.6 million people were diagnosed with PCa and 366 thousand people succumb to it each year (Pernar, Ebot, Wilson, & Mucci, 2018). PCa metastasis occurs predominantly in the bone, which is associated with high mortality (Mazzone et al., 2018; Riihimäki, Thomsen, Sundquist, Sundquist, & Hemminki, 2018), thereby...
suggesting the need for its mechanistic evaluation. Recent studies have focused on understanding the role of the pre-metastatic niche (PMN) and tumor micro-environment (TME) in cancer metastasis and have made a groundbreaking revelation about the mechanism of metastasis. However, little is known about specific bone metastasis, especially in PCa, because the PMN theory may not completely explain the metastatic tendency of PCa to the bone. Moreover, the bone TME seems more complicated than other secondary tumor TMEs. Therefore, we proposed a secondary concept of PMN: pre-metastatic bone niche (PMBN) to illustrate the mechanism of bone metastasis and bone TME. This review aims to explore the difference between PMN and PMBN and provide an insight into the formation of PMBN and the mechanism of bone metastasis in PCa.

### 2. Formation of the pre-metastatic niche

The classical hypothesis regarding metastasis is “seeds” (cancer cells) and the “soil” (host micro-environment) theory (Paget, 1889; Peinado et al., 2017). A hundred years later, this hypothesis has been further explained by the concept of PMN (Kaplan et al., 2005). With time, the theory of PMN has developed, where the formation of PMN takes place in three steps: first, migration of tumor-secreted extracellular vesicles and non-vesicular tumor-secreted factors to the future metastatic organ. Secondly, reprogramming of stromal cells occurs in the metastatic organ. Thirdly, recruitment of vascular endothelial growth factor receptor (VEGFR+) hematopoietic stem/progenitor cells (HSPC) that differentiate into myeloid-derived suppressor cells (MDSC) to exert immunosuppressive effects (Kaplan et al., 2005; Koh & Kang, 2012; Chin & Wang, 2016; Zheng et al., 2020). PMN can provide a hotbed for metastatic cells. However, different tumors display varying propensities to target organs. According to a recent report, 50% of breast cancer, 44–90% of pancreatic cancer, and 35–55% of colorectal cancer are known to migrate to the liver. About 30–60% of breast cancer migrated to bone, and surprisingly, 68–80% of PCa migrated to bone (Zhuyan et al., 2020). Till date, the mechanisms of specific metastasis are poorly understood. As the tumor micro-environment and the immune environment have been explored, the mechanism of specific metastasis could be explained in this aspect, especially the bone metastasis of PCa.

#### 2.1 Pre-metastatic niche vs. pre-metastatic bone niche

PMN has been detected in the lung and liver, but is poorly explored in bone. However, the components of PMN like bone marrow-derived cells (BMDC)/HSPC are components of the stromal niche and hence do not require reprograming like other PMN. Thus, we put forth a concept of PMBN to explain the specific metastasis, particularly in PCa. We discuss the differences in the sources and functions of components between PMN and PMBN (Table 1).

### 2.2 The two subtypes of myeloid-derived suppressor cells

MDSCs are a heterogeneous population of immature myeloid cells (IMCs), which are the precursors of dendritic cells (DCs), macrophages, and granulocytes. They have the ability to significantly inhibit immune cell responses and negatively regulate immunity. MDSCs are mainly composed of two subtypes, including polymorphonuclear myeloid-derived suppressor cells (PMN-MDSCs) and monocytic myeloid-derived suppressor cells (M-MDSCs) (Li et al., 2021). Mice and human MDSCs have different cell surface markers (Gabrilovich, 2017). In mice, PMN-MDSCs and M-MDSCs can be respectively defined as CD11b+Ly6G-Ly6Chigh and CD11b+Ly6G-Ly6Chig (Gabrilovich, 2017). Similarly, PMN-MDSCs and M-MDSCs can be respectively defined as CD11b+CD14−CD15+CD66b+ and CD11b+CD14+HLA-DR−CD15+ in human peripheral blood mononuclear cells (PBMC) (Gabrilovich, 2017). Increasing evidence shows that M-MDSCs and PMN-MDSCs achieve immunosuppressive effects through different mechanisms. PMN-MDSCs highly express arginine (Arg1) and reactive oxygen species (ROS), while M-MDSCs highly express nitric oxide (NO) and nitric oxide synthase (iNOS), all of which mainly inhibit the function of T cells (Gabrilovich, 2017). Notably, current research suggests that PMN-MDSCs are the main subtype responsible for promoting prostate cancer metastasis. For example, one study has found that PMN-MDSCs are more abundant in bone metastases than in primary prostate cancer (Wen et al., 2020).

#### 2.3 Steps for the formation of the pre-metastatic bone niche

##### 2.3.1 Step 1: Tumor-derived soluble factors and tumor-derived exosomes are induced by hypoxia and inflammation

TDSF such as interleukin (IL)-10, vascular endothelial growth factor (VEGF), transforming growth factor-beta (TGF-β), and soluble human leukocyte antigens (HLA) molecules have several important immune modulatory functions like immune escape, immunosurveillance, and subduing the immune cells’ functions (Allard et al., 2011; Deepak & Acharya, 2010; Packard, Lee, Remold-O’Donnell, & Komoriya, 1995; Shimabukuro-Vornhagen et al., 2012). As the first step of the formation of PMBN, the role in bone environment and MDSC has been
Quantitative mass spectrometry. They found that integrins, an essential component in PMN, representing the cell adhesion receptor proteins in exosomes, were the deciding factor for metastatic organotropism. They showed that ITGa6β4 and ITGa6β1 expressed by lung-tropic exosomes migrated to the lung microenvironment and that pancreatic-tropic exosomes expressing ITGa6β5 preferred liver niches (Hoshino et al., 2015; Y. Liu & Cao, 2016). However, integrins expressed by TDEs migrating to the bone still need to be explored.

Hypoxia and inflammation driven by PCs are the two major causes for the secretion of TDSF and tumor-derived exosomes. Tumor promotes progression and metastasis potentially through exosome driven by hypoxia which may be mediated by hypoxia inducible factor-1α (King et al., 2012). Meanwhile, the data in PCs cells was more representative. Recent data showed that PCs derived exosomes promoted invasiveness and stemness under hypoxia, which in turn promoted the activity of matrix metalloproteinases (MMPs) (Deep et al., 2020). Interestingly, the expression of MMP is one of the features of PMN; thus, the hypoxia-tumor derived exosomes axis plausibly remodels distant PMN (Deep et al., 2020). Moreover, a large number of exosomes are secreted by PCs cells, which is a mechanism to remove metabolic waste and promote the survival of PCs under chronic hypoxia (Panigrahi et al., 2018). Other molecules induced by hypoxia, including lysyl oxidase, annexin A1, and PIM kinases (Erl et al., 2009; Bizzarro et al., 2017; Toth et al., 2019), also regulate the invasion of PMN. Inflammation is not only induced but is also a part of the TME, which is rich in proinflammatory cytokines and cells. These promote metastasis in several ways, such as inducing TDSF, which can potentially alter myelopoiesis (Ugel, De Sanctis, Mandruzzato, & Bronte, 2015). In PCs cells, TDSF such as indoleamine 2,3-dioxygenase and IL-6 have already been reported to mediate PCs morbidity. Elicitation of PCs-TDSFs by inflammatory factors like interferon gamma (IFN-γ) and TNF-α might be responsible for rendering a tumor untreatable (Banzola et al., 2018). Another TDSF called sHLLA-E can be upregulated by IFN-γ, TNF-α and IFN-α (Allard et al., 2011). Further, IL-6 secretion regulates the epithelial-

### TABLE 1 The difference of source and function of components between PMN and PMBN.

| Components                      | PMN                          | PMBN                          | Ref.                                |
|--------------------------------|------------------------------|-------------------------------|------------------------------------|
| BMDC/HSPC                      | Migrate from bone            | Inherent                      | Kaplan et al. (2005); Kaplan, Psaela et al. (2006) |
| CXCL12                         | Secreted by stromal cells, fibroblasts and epithelial cells | Secreted by stromal cells, fibroblasts, epithelial cells, osteoblast, osteoclast | Ahmadzadeh et al. (2015); Meng, Xue, & Chen (2018) |
| extracellular matrix           | Collagens, proteoglycans, laminins, fibronectin, matricellular-associated proteins | Collagens, proteoglycans, laminins, fibronectin, matricellular-associated proteins, osteopontin | Moun et al. (2014); Garland et al. (2016); Kae et al. (2019) |
| bone morphogenetic protein (BMP) | Inhibits breast cancer cells at lung metastatic sites | Promote invasive properties in prostate cancer at bone metastatic sites | Graham et al. (2010); Gao et al. (2012) |
| Parathyroid hormone-related protein | None                         | Upregulating the expression of integrin | Shen & Faljoo (2003) |
| bone-stored growth factors     | None                         | Includes insulin-like growth factors, TGF-β | Yoneeda (2011); Xie, Ling, van Dam, Zhou, & Zhang (2018); Hiraga (2019) |
mesenchymal transition (EMT) and homing of tumor cells to the bone (Nguyen, Li, & Tewari, 2014).

2.3.2 Step 2: Migration of metastasis-initiating cells

MICs, stimulated by TDFs and TDEs, are recruited to the bone metastatic lesions, preparing to form the PMBN. They display long-term self-renewal, specific driver mutations, and high cellular plasticity (Celià-Terrassa & Kang, 2016). Although tumor metastasis is prevalent in clinics, dissemination of the tumor to a secondary lesion at a molecular level is difficult. Metastasis is a difficult process with a huge rate of attrition; thus, only about 1% of disseminated tumor cells (DTCs) are estimated to metastasize successfully (Luzzi et al., 1998; Cameron et al., 2000). MICs are somewhat like cancer stem cells that maintain their stemness and self-renewal. However, their most prominent feature is dormancy. The dormant MIC travels to distant metastatic target organs, like bone, becomes a part of PMN or PMBN, and when the PMN/PMBN matures, the MIC reinvades and exerts its malignant potential. However, MIC in dormancy still plays a critical role, especially in bone. Bone marrow contains hematopoietic niches that also play an important part in bone metastasis, especially in the dormant stage of MIC. Mesenchymal and endothelial cells contribute to a microenvironment called hematopoietic niches (Aurrand-Lions & Mancini, 2018), which is a protective site for tumor cells. Cancer cells survive in a dormant state by utilizing hematopoietic niches (Mukaida, Zhang, & Sasaki, 2020). PMN is suggested to be created or conditioned by early DTC which might affect metastasis development even through dormancy (Sosa, Bragado, & Aguirre-Ghiso, 2014). When MICs migrate to the bone marrow, mesenchymal cells form heterotypic adherens junction with MICs and promote outgrowth. MICs also use secreted and membrane-bound vascular cell adhesion molecule 1 (VCAM1) to recruit pre-osteoclasts, which can further promote osteolytic invasion of indolent bone micro-metastasis by connecting α4β1-positive osteoclast progenitors (Lu et al., 2011; Celià-Terrassa & Kang, 2016). Besides, PCa-induced osteoblast activity increases receptor activator of nuclear factor kappa-B ligand (RANKL), release of parathyroid hormone, and promotes osteoclast activity (Casimiro, Ferreira, Mansinho, Alho, & Costa, 2016). The above findings demonstrate that PMBN MICs enhance the “vicious cycle” of bone metastasis, which is another difference between PMBN and PMN.

2.3.3 Step 3: Activation of chemokine-chemokine receptor signaling

2.3.3.1 Chemokine (C-X-C motif) ligand 12/receptor 4 axis in bone

Stromal derived factor (SDF)-1, also called CXCL12, is highly expressed within the fully formed PMN (Kaplan et al., 2006a) and whose secretion is an important event for PMN. Here, we discuss some reports on CXCL12/CXCR4 in bone metastasis of PCa. CXCL12, secreted by osteoblast, endothelial cells, and mesenchymal stem cells, plays a vital role in the formation of the bone metastatic niche (Ahmadzadeh et al., 2015). CXCL12 is known to be more highly expressed in metastatic lesions than in normal tissues of PCa (Sun et al., 2003). Chemokines recruit cells with corresponding receptors by generating a concentration gradient (Jin, Xu, & Hereld, 2008). PCa were found to express CXCR4 (Sun et al., 2003), and the CXCL12 secreted by osteoblast and osteoclast then recruits the PCa cells to the PMBN. This may be a possible mechanism for cancer cells’ “homing” to the bone and may be responsible for their growth in selected organs (Sun et al., 2003). CXCL12 levels were found to be high in the pelvis,ibia,femur,liver, and adrenal/ Kidneys compared with those in the lungs, tongue, and eye (Sun et al., 2005). Interestingly, tumorderived CXCL8 and phosphatase and tensin homolog (PTEN)-deficient cancer cells increase their sensitivity and reactivity to stromal chemokines by upregulating the expression of receptors in cancer cells and inducing stromal chemokine synthesis (Maxwell, Neisen, Messenger, & Waugh, 2014). PTEN is a regulator of CXCL12/CXCR4 and its loss leads to the activation of both Akt1 signaling and MMP9 expression. This promotes the expression of CXCL12/CXCR4, which in turn regulates the metastasis and invasion of PCa. Moreover, Akt1 overexpression reversed the osteoclerotic phenotype to an osteolytic phenotype and promoted intra-osseous tumor growth (Chinni et al., 2006; Conley-LaComb et al., 2013).

2.3.3.2 Tumor-associated macrophages/chemokine (C-C motif) ligand 5 and tumor-associated macrophages/CXCL1 axis in bone

Chemokines make a great contribution to the PMN/PMBN. Here, we discuss two important axes in the bone microenvironment. TAMs are immune cells that have anti-tumor properties. However, the M2 phenotype of TAMs reveals a contrasting function, promoting tumor angiogenesis and metastasis (S. Lin et al., 2017). A study shows that consumption of M2 TAMs disrupted lung PMN and prevented metastasis (Chen et al., 2017b). TAMs function by secreting chemokines such as CCL5 and CXCL1. A recent study showed that TAMs/CXCL1 signaling could enhance breast cancer metastasis (N. Wang et al., 2018) and stimulate the recruitment of HSPCs and their differentiation into MDSCs, further promoting the formation of PMN (S. Wang et al., 2020; Zheng et al., 2020). However, TAMs/CXCL1 has not been reported in bone metastasis. Surprisingly, CCL5 was found to be a critical chemokine in gastric cancer, colorectal cancer, and breast cancer (An et al., 2019; Ding et al., 2016; S. Zhang et al., 2018). TGF-β signaling is an important pathway regulating the bone micro-environment and crosstalks with several pathways associated with tumor invasion (we will discuss the TGF-β later). CCR5, the cognate receptor of CCL5, was shown to be regulated by TGF-β signaling (S. Lin et al., 2017). Also, THP1-derived TAMs co-injection with cancer cells increased the bone...
metastasis of PCa xenografts, while CCL5 knockdown partly abrogated it (R. Huang et al., 2020).

### 2.3.3.3 Angiopoietin-1/tie-2 axis in bone

Ang-1 and its receptor Tie-2 are one of the significant cytoactive molecules found within bone marrow, primary lesion and the PMN (Kaplan et al., 2006b). Tie-2 expression was found to be higher in PCa cell lines, which are capable of migrating to the bone. The Tie-2 high PCa cells displayed more adherence than the Tie-2 low PCa cell population to both osteoblasts and endothelial cells, and these cells also had a high expression of cancer stem cells (CSCs). However, knockdown of the Ang-1 led to suppression of CSCs. More importantly, only the Tie-2 high but not the Tie-2 low cells developed metastasis in vivo (K. D. Tang et al., 2016). Moreover, ng-1 mRNA expression was not observed in bone, lymph node or liver metastasis but was observed in bone marrow cells (Morrissey et al., 2008), suggesting that PCa cells move to PMBN through the Ang-1/Tie-2 axis.

### 2.3.4 Step 4: Changes in extracellular matrix for preparing pre-metastatic bone niche

ECM changes are symbolic pre-metastatic changes in the target organ (Zhuyan et al., 2020) and contain comprehensive components such as collagens, proteoglycans, laminins, fibronectin, and matricellular-associated proteins (Mouw, Ou, & Weaver, 2014; Kai, Drain, & Weaver, 2019). When it comes to PMN, hypoxia, chemokines, and TDSF, as discussed previously, induce ECM to change to a form with increased stiffness and tensile strength (Garland, Erler, & Cox, 2016). Furthermore, lysyl oxidase-mediated collagen crosslinking creates a fibrotic microenvironment supporting metastatic growth (Gartland et al., 2013). Notably, each organ has its own PMN and changes in ECM, but when it comes to PMN and bone, the whole organ (bone) is an enormous “organized mesh” containing a tremendous amount of ECM, which consists of type I collagen, fibrous proteins, and non-collagenous proteins predominantly (Garland et al., 2016). This is another characteristic of PMBN which is different from PMN, giving an insight into why so many tumors have a predilection for bone metastasis. An enzyme called lysyl oxidase is known to post-translationally modify collagen and elastin in the ECM, thereby catalyzing the covalent crosslinking of collagen fibers associated with bone metastasis (Garland et al., 2016). An in vivo experiment showed that tumor bearing mice displayed increased bone loss and formed focal osteolytic lesions over time before metastasizing, and these changes were lysyl oxidase (LOX)-dependent (Garland et al., 2016). Thus, during PMBN formation, bone is osteolytic, far before the time that cancer cells home to the bone. This is yet another feature of PMBN. Another important ECM component in the bone is osteopontin, which is associated with malignant transformation and acts as a paracrine and autocrine mediator of PCa growth and progression (Thalmann et al., 1999). As stated previously, the bone marrow microenvironment contains intricate components including ECM, cytokines and chemokines regulate the hematopoietic progenitor cells’ proliferation and differentiation, which was a core step of PMBN.

### 2.3.5 Step 5: Recruitment of VEGFR1+ bone marrow-derived cells in bone

Firstly, VEGFR was found to have a high expression in PCa. More importantly, its expression was elevated at sites of bone metastasis compared to the original prostate tumor. VEGF interacting with VEGFR regulated adhesive and migratory properties of the cancer cells (J. Chen, De, Brainard, & Byzova, 2004). As discussed about PMN formation above, HSPC expressing VEGF1 and bone BMDC colonize pre-metastatic sites before tumor cells (Kaplan et al., 2005). However, the reason for the recruitment of VEGFR+ BMDC or VEGFR+ HSPC to be considered as a symbolic event of PMN is not known (Kaplan, Rafii, & Lyden, 2006; Zheng et al., 2020)? The following reasons can be considered: regular T cells and MDSC, especially those differentiated from HSPC, are functional types of BMDC (Koh & Kang, 2012). Purified HSPC in vivo differentiated into MDSC in early metastatic sites of tumor-bearing mice and promoted tumors metastasis (Giles et al., 2016). The MDSC in the PMN provided a microenvironment suitable for cancer cells by enhancing immunosuppression, leaking vasculature, and collagen restructuring in the PMN by suppressing T cells, generating a lot of NO, arginase1 (Arg-1) and immunosuppressive cytokines, and promoting regular T cell expansion (Y. Wang, Ding, Guo, & Wang, 2019). A recent study reported that, during tumor progression, MDSC contributed to PMN formation by upregulating MMP-9 expression (J. Zhang et al., 2020). The ability to suppress immune cells is an important standard that is used to define MDSC (Bronte et al., 2016). Recruitment of HSPC as a symbolic event for PMN, which has been proven by several cancers like breast and colorectal cancer (Psaila, Kaplan, Port, & Lyden, 2006; C. Zhang et al., 2014). As for bone metastasis, distant primary tumor drives the expansion of HSPC within the bone marrow and their mobilization to the bloodstream (Giles et al., 2016). That means they don’t even need to move to the bloodstream and site to the PMN, because the bone is already its PMN, we call that PMBN. The bone marrow is an inherent store of HSPC. Besides, metabolic conditions in niches such as calcium concentrations mediated HSPC retention within, but not homing to, the endosteal niche (Kucia et al., 2005). It is not surprising that many cancers show a proclivity to establish themselves in the bone marrow (Kaplan, Psaila, et al., 2006). Recently, a new study demonstrated a previously unidentified role for perivascular cells in PMN formation, which is a new mechanism for PMN. The author showed that perivascular cells lost the expression of traditional vascular smooth muscle cells to build the PMN by stimulating the tumor-secreted factors and genetic activation of Klf4 (Murgai et al., 2017).
2.3.6 Step 6: Integrin plays a role in the last step of the pre-metastatic bone niche

Here, we illustrate some data about integrin in the bone environment of PCa. Bone-metastatic castration-resistant PCa is lethal and shows drug resistance. One of the mechanisms is the integrin α6β1-mediated adhesion to laminin (Toth et al., 2019). Adhesion capability of PCa cells to bone marrow endothelial cells was enhanced through upregulation of integrin-α4 expression, concurrent with transcriptionally activated NF-κB (Zhao et al., 2020). Parathyroid hormone-related protein, upregulates the expression of α1, α5, α6, and β4 integrin subunits and plays a vital role in the development of bone metastasis (Shen & Falzon, 2003). In other cell lines, integrin β1 has high expression in hepatocellular carcinoma with stiffness substrates and co-regulates with the INK/c-JUN signaling pathway in upregulating LOXL2, MMP9, fibronectin production, CXCL12 expression and BMDC recruitment, which account for PMN formation (Wu et al., 2018). However, the positive outcome of α4 integrin which is most relative with PMN. It has been reported that fibroblasts contributed to breast cancer bone metastasis by mediating CCL4/CCR5 axis (Sasaki et al., 2016). Upregulation of fibronectin activated by fibroblasts is one of the earliest changes observed in future metastatic niches (Kaplan et al., 2005), and then integrin α4β1 (VLA4) carrying VEGFR1+ HSPC attached to the upregulating fibronectin. From the figure of Kaplan, VLA4 connected with HSPC to fibroblasts and that was the last step of the PMN formation (Kaplan, Rafii, et al., 2006). Those series events associated with VLA4 are similar to their function within the bone marrow (Kaplan, Psaila, et al., 2006). It seems like the PMN try to provide a tumor-friendly environment by imitation from bone marrow environment and PMBN is an integration of bone marrow environment. Consequently, we know from above that PMBN is the initial PMN. In PCa cells line, VLA4 is not the only key for connection, integrin α6β1, integrin α1, α5, α6, and β4 had their contribution to the bone metastasis. VLA4+VEGFR+ HSPC attaching fibronectin completely is a symbolic event of the maturation of PMBN (Figure 1).

2.4 From pre-metastatic bone niche to mature bone metastasis

2.4.1 Activation of migration of Metastasis-initiating cells

Reactive oxygen-generating enzyme (Nox1) increases tumorigenicity of prostate epithelial cell line. Importantly, Nox1 could significantly upregulate VEGF mRNA, induce VEGFR1, VEGFR2 and MMPs (Arbiser et al., 2002), which is...
important for PMN and MIC. Nox1 may activate MIC by triggering the angiogenic switch. Early DTCs contributed to the PMN and the later ones account for the escape from dormancy to promote metastasis (Sosa et al., 2014). For example, tumor cells could escape from dormancy by upregulating and activating VCAM1 (Lu et al., 2011). Periostin, as a cancer-promoting factor, is induced by endothelial tip cells, also promoting dormancy escape (Malanchi et al., 2011; Ghajar et al., 2013). β1 integrins are indispensable to prevent dormancy onset and critical for proliferation of micro-metastatic cancer cells (Shibue & Weinberg, 2009; Sosa et al., 2014). Gaining the function of EMT and MET is also a typical event of MICs’ activation. MICs are a type of stem-like cell that increases expression of EMT, stemness of stem cells and pro-survival (Lawson et al., 2015). When they arrive at the secondary organ, a reversed process: MET is required (Tsai, Donaher, Murphy, Chau, & Yang, 2012). Recent research found that low-burden cells expressed high-level dormancy genes such as CDKN1B, CHEK1, TGFBR3 and TGFBI2 while highly-burden metastatic cells rarely expressed dormancy-associated genes, whereas highly expressed cell-cycle promoting genes such as CDK2, MYC, MMP1, and CD24, which were accounted for dormancy escape (Lawson et al., 2015). We could imply that during the formation of PMBN, the surrounding MIC belong to low-burden cells which show the stemness however, when the PMBN is completely finish, those MIC turn to more heterogeneous. Energy metabolism is another aspect to elucidate the activation of MIC. A study demonstrated that in brain metastatic breast cancer cells, tumors could obtain energy in multiple ways in order to reinitiate MIC proliferation (Celià-Terrassa & Kang, 2016; E. I. Chen et al., 2007). A high plasticity in energy substrate metabolism in PCa maybe another energy mentalism pathway to active MIC as previous reported (Aguilar et al., 2016). Other mechanism such as high expression of VCAM1 induced by inflammatory, inducing reactivation of growth by stromal niche also account for the activation of MIC (Lu et al., 2011; Giancotti, 2013). Intriguingly, bone morphogenetic protein (BMP) could promote invasive properties in PCa at bone metastatic sites (Graham, Agrawal, & Abdel-Mageed, 2010), but the BMP inhibitor Coco reactivates breast cancer cells at lung metastasis. It seems that BMP plays an opposite role in PMBN and PMN (Gao et al., 2012).

2.4.2 Disruption of balance between osteoblast and osteoclast

The balance comes from a classical theory of bone metastasis called the “vicious cycle theory,” which demonstrates a series of molecular events. In brief, osteoclast-stimulating factors, such as PTHrP, promote osteoblasts to secrete RANKL, which in turn promotes differentiation of osteoclast precursor cells into mature osteoclasts (Mundy, 2002; Hiraga, 2019). Later, osteoclasts secrete TGF-β to promote PCa invasion and migration. Surprisingly, a lot of insights go into exploring PCa with bone metastasis. First comes the regulation of micro-RNA. Three studies showed that micro-RNA (miR-141-3p, miR-210-3p, miR-133a-3p, miR-204-5p) affected bone metastasis by activating NF-κB and PI3K/AKT signaling (S. Huang et al., 2017; Ren et al., 2017; Y. Tang et al., 2018; Wa et al., 2019; Ye et al., 2017). Micro-RNAs (microRNA-124, miR-133b, miR-505-3p, miR-19a-3p) regulate the bone metastasis via activating TGF-β signaling pathway (Coniglio, 2018; S. Huang et al., 2018; Y. Tang et al., 2019; Wa et al., 2018). Other factors such as lnc-RNA, HIFTM3, PICK1 were also mediate TGF-β signaling pathway (Y. Dai et al., 2017; Lang et al., 2020; X. Liu et al., 2019). TGF-β signaling is an important signaling/factor for bone metastasis in PCa. To echo the PMBN, those symbolic factors of PMBN indeed play a role in bone metastasis. For example, VEGF contributes to PCa-induced osteoblastic activity in vivo (Kitagawa et al., 2005); MMP2 is upregulated to promote PCa bone metastasis (Dutta et al., 2014; Chen et al., 2017a). Other factors, like interferon regulatory factor 7, stimulate oncostatin M or even (m6A) mRNA methylation, regulate the bone metastasis of PCa. Here, we highly focus on the specific factors of bone: bone-stored growth factors. Osteoclastic bone resorption followed by the release of bone-stored growth factors (Yoneda, Hashimoto, & Hiraga, 2003; Yoneda, 2011) such as insulin-like growth factors and TGF-β therefore, provides fertile soil for metastatic cancer cells. Bone-derived IGF-1 connected with bone and metastasized the tumor cells via IGF-IR/Akt/NF-κB signaling, while BMP9 was able to inhibit the migration involving SDF-1/CXCR4/P13K pathway, which was associated with PMBN regulation (Hiraga et al., 2012; W. Wang et al., 2015).

2.4.3 Osteoblastic bone metastasis in pre-metastatic bone niche

Osteoprotegerin (OPG)/RANKL/RANK axis mediated osteolytic bone metastasis is common in a lot of cancer bone metastasis, but there is a consensus that osteoblastic bone metastasis is dominant in PCa (Berruti et al., 2001). OPG is an important cytokine to prevent pre-osteoclast from becoming osteoclast by acting as a decoy receptor for RANKL. A meta-analysis found that OPG was highly expressed in PCa with bone metastasis (Zang et al., 2015), suggesting its importance in mediating osteoblastic bone metastasis. Dickkopf-1 (DKK-1)/Wnt signaling, endothelin-1 (ET-1) and BMP are also very important in regulating osteoblastic bone metastasis. In TME, PCa cells highly express DKK-1 to promote cancer proliferation before bone metastasis. However, when the PMBN is accomplished, PCA cells rarely express DKK-1 (Aufderklamm et al., 2018), which subsequently activates.
Wnts’ osteoblastic activity (Schwaninger et al., 2007; Hall, Daignault, Shah, Pienta, & Keller, 2008). Moreover, endothelin 1, elicits pleiotropic effects on the microenvironment, expressed by PCA cells, suppresses the function of DKK-1 so as to enhance the activation of Wnt signaling (Rosanò, Spinella, & Bagnato, 2013). BMP, which induces MIC to activate cancer cells in PMN to mature PMBN, also plays a role in promoting osteoblastic activity (J.Dai et al., 2005). Osteoblast enhanced VCAM-1 expression in PCa cells and subsequently promoted the adherence of cancer cells to osteoblasts (Chang et al., 2018) and surprisingly, this process was corroborated with the formation of PMBN.

2.5 Traditional Chinese medicine and Pre-metastatic niche

The theoretical basis of treatment using Traditional Chinese Medicine (TCM) is the Yin-Yang theory. It is important for TCM to maintain the Yin-Yang balance in the body. Tumor cells are taken for Yang as invasion and spread, while immune cells are taken for Yin on account of clearing. If immune cells cannot clear tumor cells, it will cause an imbalance of Yin-Yang in the body, leading to disease progression. Notably, MDSCs are the main cells that make up PMBN and are capable of forming a pre-metastatic immunosuppressive microenvironment (Zheng et al., 2020). Fortunately, TCM with multi-targeted efficacy can exert anti-tumor effects by regulating MDSCs. For example, cinnamaldehyde (CA), an important component of cinnamon, can enhance the immune killing effect of PCa by inhibiting myeloid-derived suppressor cells (MDSCs) (Han et al., 2020). Accumulated evidence indicates that TCM plays a pivotal role in regulating the pre-metastatic niche and suppressing tumor metastasis. Thus, PMN is an integral process involving multiple organs, multiple cells, and multiple cytokines. Coincidentally, TCM has a holistic view of diagnosis and treatment. In the past few years, a large number of studies have shown that TCM can suppress tumor metastasis by inhibiting the formation of PMN in breast cancer (Tian et al., 2020; Zheng et al., 2020), gastric cancer (Zhu, Zhou, Xu, & Wu, 2017) and colorectal cancer (C. Chen
Therefore, summarizing the functions of TCM herbs that can inhibit the formation of PCa PMN is beneficial to extend the clinical application of TCM.

Several studies have demonstrated that natural phytochemicals extracted from TCM herbs show great advantages in the control of tumor metastasis via inhibiting the formation of PMN. Active components extracted from TCM herbs such as bufalin, which were obtained from the ChanSu skin and parotid venom glands, can inhibit bone metastasis of PCa (J. J. Zhang et al., 2019). Celastrol, one of the active components of Tripterygium wilfordii, can inhibit the bone metastasis of PCa cells by inhibiting the VEGF pathway of bone marrow-derived endothelial progenitor cells (RM-EPCs) (Kuchta et al., 2017). However, these two studies only preliminary explored the inhibition of bone metastasis of TCM herbs, and have not thoroughly investigated the mechanism of inhibiting bone metastasis of PCa. The balance of osteogenesis and osteoclast is an important regulatory mechanism for the formation of PMN in bone metastasis (Furesi, Rauner, & Hofbauer, 2021), and the RANKL pathway is closely related to osteogenesis (Portal-Núñez et al., 2017). Perilla aldehyde (PAH), one of the active components of the TCM herb Perilla, is widely used and has important anticancer activity. A study has found that PAH can inhibit the formation of an osteoclast pre-metastatic niche by inhibiting the RANKL pathway and ultimately inhibit the bone metastasis of PCa (Z. Lin et al., 2022). In addition, Aldehydic components of Cinnamon bark ultimately extract can also suppress RANKL-induced osteoclastogenesis by down-regulating the expression of transcription factor NFATc1 (Tsuji-Naito, 2008).

However, there is still a lack of in-depth research on the regulation of PCa bone pre-metastatic niche by TCM. In addition to osteoclastogenesis, some immunosuppressive cells such as MDSC and Treg play an important role in the formation of the PCa bone pre-metastatic niche (Cheng & Wang, 2021). There are some exciting progresses in the field of breast cancer pre-metastatic niche. For example, Wang et al. found that XIAOPI formula, a TCM herb composed of multiple prescriptions, can inhibit the pre-metastatic niche formation in breast cancer via the suppressing function of TAMs (Zheng et al., 2020). TCM treasure trove. On the basis of illuminating the formation of the PCa bone metastasis niche, in-depth research on the mechanism of TCM will help us to expand the clinical application of TCM in the treatment of patients with PCa bone metastasis (Table 2).

### 3 Conclusion

The concept of PMBN may be convenient for the illustration of the predilection of PCa bone metastasis. In brief, after stimulation by TDSF, MICs will migrate to the target niche and activate chemokines, which further induces the BMDC to MDSC in bone. Fibronectin, integrin, and VEGFR promote the recruitment and fusion of PMBN. As BMDC comes from the bone, the formation of PMBN is easier than any other PMN. After maturation of PMBN, MICs will activate in bone and induce tumor cell metastasis from primary prostate to bone, which affects the balance between osteoblast, and osteoclast thereby leading to bone metastasis. Moreover, TCM has potential regulation in PMBN. However, PMBN is just a hypothesis for now and further basic research is necessary to determine the cause of the predilection of PCa bone metastasis.

### Author contributions

SX conceived and revised the article. CC and RH wrote and revised the article. JZ and LG participated in the work of reference selection.

### Funding

This work was supported by the National Natural Science Foundation of China (No. 81774067), the Specific Research Fund for TCM Science and Technology of Guangdong Provincial Hospital of Chinese Medicine (No. YN2016M03, KY2017I01), the Medical Science and Technology Research Fund of Guangdong Province (No. A2020239), the Guangdong-Hong Kong collaborative innovation projects from Guangdong Provincial Hospital of Chinese Medicine (No. YN2018HK02), and the Department of Science and Technology of Guangdong Province (No. 2017A050506042).

### Acknowledgments

We would like to thank Editage (www.editage.cn) and Zhaoqiang Lie for English language editing.

### Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

### Publisher’s note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors, and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.
Osteoblast-secreted WISP-1 promotes adherence of prostate cancer cells to bone via the up-regulation of matrix metalloproteinase-2 through down-regulating miR-376c expression. J. Hematol. Oncol. 10 (1), 33. doi:10.1186/1756-8722-10-33

Chen, X. W., Yu, T. J., Zhang, J., Li, Y., Chen, H. L., Yang, G. F., et al. (2017b). CYP1A2 in tumor-associated macrophages promotes pre-metastatic niche formation and metastasis. ONCOGÉNE 36 (35), 5045–5057. doi:10.1038/onc.2017.118

Cheng, X., and Wang, Z. (2021). Immediate modulation of metastatic niche formation in the bone. Front. Immunol. 12, 765994. doi:10.3389/fimmu.2021.765994

Chen, A. R., and Wang, S. E. (2016). Cancer tills the premetastatic field. Mechanistic basis and clinical implications. Clin. Cancer Res. 22 (15), 3732–3733. doi:10.1158/1078-0432.Ccr-16-0028

Chinni, S. R., Sivalogan, S., Dong, Z., Filho, J. C., Deng, X., Bonfil, R. D., et al. (2006). CXCL12/CXCR4 signaling activates akt-1 and MMP-9 expression in prostate cancer cells. The role of bone microenvironment-associated CXCL12. PROSTATE 66 (1), 32–48. doi:10.1002/pros.20318

Coniglio, S. J. (2018). Role of tumor-derived chemokines in osteolytic bone metastasis. Front. Endocrinol. 9, 313. doi:10.3389/fendo.2018.00313

Conley-LaCombi, M. K., Saliganan, A., Kandagatha, P., Chen, Y. Q., Cher, M. L., Chinni, S. R., et al. (2012). PTEN loss mediated Akt activation promotes prostate tumor growth and metastasis via CXCL12/CXCR4 signaling. Mol. Cancer 12 (1), 85. doi:10.1186/1476-4598-12-85

Cox, T. R., Bird, D., Baker, M. A., Barker, H. E., Ho, M. W., Lang, G., et al. (2013). LOX-mediated collagen crosslinking is responsible for fibrosis-enhanced metastasis. Cancer Res. 73 (6), 1721–1732. doi:10.1158/0008-5472.CAN-12-2233

Cramer, C. A., Ahn, B. J., Han, S. I., and Parsa, A. T. (2012). Soluble factors secreted by glioblastoma cell lines facilitate recruitment, survival, and expansion of regulatory T cells: Implications for immunotherapy. Neuro. Oncol. 14 (5), 584–595. doi:10.1177/1368952812457907

Dai, J., Keller, J., Zhang, J., Lu, Y., Yao, Z., Keller, E. T., et al. (2005). Bone morphogenetic protein-6 promotes osteoblastic prostate cancer bone metastases through a dual mechanism. Cancer Res. 65 (18), 8274–8285. doi:10.1158/0008-5472.Can-05-1891

Deepak, P., and Acharya, A. (2010). Anti-tumor immunity and mechanism of immunosuppression mediated by tumor cells. Role of tumor-derived soluble factors and cytokines. Int. Rev. Immunol. 29 (4), 421–458. doi:10.1080/08993600903561018

Ding, H., Zhao, L., Dai, S., Li, L., Wang, F., Shan, B., et al. (2016). CCL5 secreted by tumor associated macrophages may be a new target in treatment of gastric cancer. Biomed. Pharmacother. 77, 142–149. doi:10.1016/j.biopha.2015.12.004

Dutta, A., Li, J., Lu, H., Akech, J., Pratap, J., Wang, T., et al. (2014). Integrin αvβ5 promotes an osteolytic program in cancer cells by upregulating gastric cancer. Biomed. Pharmacother. 77, 142–149. doi:10.1016/j.biopha.2015.12.004

Dutta, A., and Wang, C. (2011). The immunoregulatory mechanisms of carcinoma for its survival and development. J. Exp. Clin. Cancer Res. 30 (1), 12. doi:10.1186/1756-9966-30-12

Dutta, A., Li, J., Lu, H., Akech, J., Pratap, J., Wang, T., et al. (2014). Integrin αvβ5 promotes an osteolytic program in cancer cells by upregulating MMP2. Cancer Res. 74 (5), 1598–1608. doi:10.1158/0008-5472.CAN-13-1796

Erfurter, P. –, Bennetworth, K. L., Cox, T. R., Lang, G., Bird, D., Koong, A., et al. (2009). Hypoxia-induced hsa-lyso oxidase is a critical mediator of bone marrow cell recruitment to form the premetastatic niche. CANCER Cell. 15 (7), 35–44. doi:10.1016/j.ccr.2008.11.012

Furest, G., Rauner, M., and Hofbauer, L. C. (2021). Emerging players in prostate cancer-bone niche communication. Trends Cancer 7 (2), 112–121. doi:10.1016/j.trecan.2020.09.006

Gabrilovich, D. I. (2017). Myeloid-derived suppressor cells. Cancer Immunol. Res. 5 (1), 3–8. doi:10.1158/2326-6066.Cir-16-0297

Gao, H., Chakraborty, G., Lee-Lim, A. P., Mo, Q., Deck, M., Venica, A., et al. (2012). The BMP inhibitor Cocos reactivates breast cancer cells at lung metastatic sites. Cellf. 150 (4), 764–779. doi:10.1016/j.cell.2012.06.035

Gartland, A., Erl, E. T., and Cox, T. R. (2016). The role of lysyl oxidase, the extracellular matrix and the pre-metastatic niche in bone metastasis. J. Bone Oncol. 5 (3), 100–103. doi:10.1016/j.jbono.2016.04.001

Ghajar, C. M., Penalado, H., Mor, H., Mateu, I. R., Evans, K. J., Brazier, H., et al. (2013). The perivascular niche regulates breast tumour dormancy. Nat. Cell. Biol. 15 (7), 807–817. doi:10.1038/ncl2767
et al. (2018). miR-130a and miR-145 reprogram Gr-1(+)CD11b(+) myeloid cells via activating TGF-β signaling, interfering the Glutathione-associated mitochondria function. *Cancer Sci.* 101 (1), 103–111. doi:10.1111/csc.12806

Han, L., Mei, J., Ma, J., Wang, F., Gu, Z., Li, J., et al. (2020). Cinnamaldehyde induces endogenous apoptosis of the prostate cancer-associated fibroblasts via interfering the Glutathione-associated mitochondria function. *Med. Oncol.* 37 (10), 91. doi:10.1007/s12032-020-01417-2

Hiraga, T. (2019). Bone metastasis: Interaction between cancer cells and bone microenvironment. *J. Oral Biol.* 61 (2), 95–98. doi:10.1016/j.job.2019.02.002

Hiraga, T., Myoui, A., Hashimoto, N., Sasaki, A., Hata, K., Morita, Y., et al. (2012). Bone-derived IGf mediates crosstalk between bone and breast cancer cells in bone metastases. *Cancer Res.* 72 (16), 4238–4249. doi:10.1158/0008-5472.Can-11-3061

Hoshino, A., Costa-Silva, B., Shen, T. L., Rodrigues, G., Hashimoto, A., Tasic Mark, M., et al. (2015). Tumour exosome integrins determine organotropism metastasis. *NATURE* 527 (7578), 329–335. doi:10.1038/nature15576

Huang, R., Wang, S., Wang, N., Zheng, Y., Zhou, J., Yang, B., et al. (2020). CCL5 derived from tumor-associated macrophages promotes prostate cancer stem cells and metastasis via activating β-catenin/STAT3 signaling. *Cell. Death Dis.* 11 (4), 234. doi:10.1038/s41419-020-2435-y

Huang, S., Wa, Q., Pan, J., Peng, X., Ren, D., Huang, Y., et al. (2017). Downregulation of miR-141-3p promotes bone metastasis via activating NEK-x signaling in prostate cancer. *J. Exp. Clin. Cancer Res.* 36 (1), 173. doi:10.1186/s13046-017-0465-7

Huang, S., Wa, Q., Pan, J., Peng, X., Ren, D., Li, Q., et al. (2018). Transcriptional downregulation of miR-15b by REST promotes prostate cancer metastasis to bone via activating TGF-β signaling. *Cell. Death Dis.* 9 (7), 779. doi:10.1038/s41419-018-0807-3

Ishii, H., Vodnala, S. K., Achyut, B. R., So, J. Y., Hollander, M. C., Greten, T. F., et al. (2015). Single-cell analysis reveals a stem-cell program in human metastatic prostate cancer cells. *Nature* 526 (7578), 750–754. doi:10.1038/nature15576

Janowska-Wieczorek, A., et al. (2005). Traf-dependent signaling promotes prostate cancer bone metastasis. *Cancer Res.* 65 (12), 5628–5633. doi:10.1158/0008-5472.CAN-04-3574

Kauci, M., Reca, R., Mieku, K., Waneckz, J., Wojaskowski, W., Janowska-Wieczorek, A., et al. (2005). Trafficking of normal stem cells and metastasis of cancer stem cells involve similar mechanisms: Pivotal role of the SDF-1-CXCR4 axis. *STEM CELLS* 23 (7), 879–894. doi:10.1634/stemcells.2004-0342

Koh, B. L., and Kang, Y. (2012). The pro-metastatic role of bone marrow-derived suppressor cells. *Frontiers in Pharmacology* 3, 11. doi:10.3389/fphar.2012.00031

Kuczti, A., Jais, R., Red, C. M., Evans, J. D., Murgia, M., Vicicco, Y., Highfill, S. L., et al. (2016). Activation of hematopoietic stem/progenitor cells promotes immunosuppression within the pre-metastatic niche. *Cancer Res.* 76 (6), 1335–1347. doi:10.1158/0008-5472.CAN-15-0204

Lynch, M. E., Chiou, A. E., Lee, M. J., Marcott, S. C., Polamraju, P. V., Lee, Y., et al. (2016). Three-dimensional mechanical loading modulates the osteogenic response of mesenchymal stem cells to tumor-derived soluble signals. *Tissue Eng. Part A* 22 (15–16), 1006–1015. doi:10.1089/ten.TEA.2016.0153

Malanchi, A., Santamaria-Martínez, A., Susanto, E., Peng, H., Lehr, H. A., Delaloye, J. F., et al. (2011). Interactions between cancer stem cells and their niche govern metastatic colonization. *NATURE* 478 (7379), 85–89. doi:10.1038/nature10094

Meng, W., Xue, S., and Chen, Y. (2018). The role of CXCL12 in tumor microenvironment. *Gastroenterol. Res.* 15 (1), 105–110. doi:10.1008/gire.2017.10.015

Morales, J. K., Kmiecik, M., Knutsen, K. L., Bear, D. H., and Manjhi, M. H. (2010). GM-CSF is one of the main breast tumor-derived soluble factors involved in the differentiation of CD11b-Gr1- bone marrow progenitor cells into myeloid-suppressor cells. *Breast Cancer Res. Treat.* 123 (1), 39–49. doi:10.1007/s10549-009-0662-8

Morrissery, C., True, L. D., Roudier, M. P., Coleman, I. M., Hawley, S., Nelson, P. S., et al. (2018). Location of metastases in contemporary prostate cancer patients affects cancer-specific mortality. *Clin. Gynecol. Cancer* 17 (5), 376–384. e371. doi:10.1016/j.clgc.2018.05.016

Mou, J. K., Ou, G., and Weaver, V. Y. (2014). Extracellular matrix assembly: A multiscale deconstruction. *Nat. Rev. Mol. Cell. Biol.* 15 (12), 771–785. doi:10.1038/nrm3902

Page dimensions: 595.3x841.9
Yoneda, T., Hashimoto, N., and Hiraga, T. (2003). Bisphosphonate actions on cancer. *Calcif. Tissue Int.* 73 (4), 315–318. doi:10.1007/s00223-002-0025-x

Yoneda, T. (2011). Mechanism and strategy for treatment of cancer metastasis to bone. *Gan Kagaku Ryoho.* 38 (6), 877–884.

Zang, L., Ma, M., Hu, J., Qiu, H., Huang, B., Chu, T., et al. (2015). The effects of lung and prostate cancer bone metastasis on serum osteoprotegerin levels: A meta-analysis. *Sci. Rep.* 5, 18324. doi:10.1038/srep18324

Zhang, C., Zhou, C., Wu, X. J., Yang, M., Yang, Z. H., Xiong, H. Z., et al. (2014). Human CD133-positive hematopoietic progenitor cells initiate growth and metastasis of colorectal cancer cells. *CARCINOGENESIS* 35 (12), 2771–2777. doi:10.1093/carcin/bgu192

Zhang, J., Han, X., Shi, H., Gao, Y., Qiao, X., Li, H., et al. (2020). Lung resided monocytic myeloid-derived suppressor cells contribute to premetastatic niche formation by enhancing MMP-9 expression. *Mol. Cell. Probes* 50, 101498. doi:10.1016/j.mcp.2019.101498

Zhang, J. J., Zhou, X. H., Zhou, Y., Wang, Y. G., Qian, B. Z., He, A. N., et al. (2019). Bufalin suppresses the migration and invasion of prostate cancer cells through HOTAIR, the sponge of miR-520b. *Acta Pharmacol. Sin.* 40 (9), 1228–1236. doi:10.1038/s41401-019-0234-8

Zhao, Z., Li, E., Luo, L., Zhao, S., Liu, L., Wang, J., et al. (2020). A PSCA/PGRN-NF-κB-Integrin-α4 Axis promotes prostate cancer cell adhesion to bone marrow endothelium and enhances metastatic potential. *Mol. Cancer Res.* 18 (3), 501–513. doi:10.1158/1541-7786.Mcr-19-0278

Zheng, Y., Wang, N., Wang, S., Yang, B., Situ, H., Zhong, L., et al. (2020). XIAOPI formula inhibits the pre-metastatic niche formation in breast cancer via suppressing TAMs/CXCL1 signaling. *Cell Commun. Signal.* 18 (1), 48. doi:10.1186/s12964-020-0520-6

Zhuyan, J., Chen, M., Zhu, T., Bao, X., Zhen, T., Xing, K., et al. (2020). Bone niche is critical steps to tumor metastasis: alterations of tumor microenvironment and extracellular matrix in the formation of pre-metastatic and metastatic niche. *Cell Biosci.* 10, 89. doi:10.1186/s13578-020-00453-9

Zhang, S., Zhong, M., Wang, C., Xu, Y., Gao, W. Q., Zhang, Y., et al. (2018). CCL5-deficiency enhances intratumoral infiltration of CD8(+) T cells in colorectal cancer. *Cell Death Dis.* 9 (7), 766. doi:10.1038/s41419-018-0796-2

Zhao, Z., Li, E., Luo, L., Zhao, S., Liu, L., Wang, J., et al. (2020). A PSCA/PGRN-NF-κB-Integrin-α4 Axis promotes prostate cancer cell adhesion to bone marrow endothelium and enhances metastatic potential. *Mol. Cancer Res.* 18 (3), 501–513. doi:10.1158/1541-7786.Mcr-19-0278

Zheng, Y., Wang, N., Wang, S., Yang, B., Situ, H., Zhong, L., et al. (2020). XIAOPI formula inhibits the pre-metastatic niche formation in breast cancer via suppressing TAMs/CXCL1 signaling. *Cell Commun. Signal.* 18 (1), 48. doi:10.1186/s12964-020-0520-6

Zhu, X., Zhou, Y., Xu, Q., and Wu, J. (2017). Traditional Chinese medicine Jiujpi Bushen therapy suppresses the onset of pre-metastatic niche in a murine model of spontaneous lung metastasis. *Biomed. Pharmacother.* 86, 434–440. doi:10.1016/j.biopha.2016.12.013

Zhuyan, J., Chen, M., Zhu, T., Bao, X., Zhen, T., Xing, K., et al. (2020). Bone niche is critical steps to tumor metastasis: alterations of tumor microenvironment and extracellular matrix in the formation of pre-metastatic and metastatic niche. *Cell Biosci.* 10, 89. doi:10.1186/s13578-020-00453-9