Long-term stocking practices threaten the original genetic diversity of the southernmost European populations of Atlantic salmon *Salmo salar*

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ABSTRACT: Many Atlantic salmon *Salmo salar* populations in Europe are threatened by previous stocking with foreign hatchery strains. Temporal patterns of genetic characteristics of salmon from northern Spain, the southernmost European populations, were compared before and after species decline and heavy stocking with specimens from northern Europe. Eleven microsatellite loci were analysed in archival (scales from 1958–1960) and contemporary (2007–2008) samples from the River Sella. Temporal analyses revealed a similar heterozygosity between archival and contemporary samples, despite a drastic decrease in population abundance, while the contemporary sample showed a higher allelic richness due to the occurrence of foreign alleles. Considering only the alleles with at least 4% frequency in the archival sample, 2 alleles exclusive to the River Sella were absent in the contemporary sample, and 14 alleles showed a decrease of at least 4% frequency. Four alleles common in Scotland showed a high occurrence in the contemporary sample, so they are good candidates as markers of introgression of foreign genes. The heavy stocking with non-native Scottish broodstocks between 1970 and 1990 caused the introgression found in the contemporary sample when compared with the pristine population. An abrupt decrease was evident when the estimates of effective number of breeders were adjusted to take into account overlapping generations (*N*ₐₛₐₐ₈), effective population size (*N*ₑₐₛₐ₈) estimated from *N*ₐₛₐ₈, and number of breeders estimated using the sibship assignment method (*N*ₛᵢᵦ). The very low effective size values found in the contemporary sample, together with the detrimental synergy between genetic drift and high rates of introgression, represent a severe risk for the conservation of native salmon.

KEY WORDS: Conservation genetics · Effective population size · Foreign stocking · Genetic variability · Introgression · Population genetics · Salmonidae

1. INTRODUCTION

Preventing the loss of genetic diversity is essential for the conservation and long-term preservation of species, because high genetic diversity is a requisite for evolutionary adaption to ongoing rapid environmental change (Schindler et al. 2010). Increasing temperatures due to global warming influence the distribution and variation of genetic diversity within and among natural populations (Jonsson & Jonsson 2009, Hórreo et al. 2014a, Almodóvar et al. 2019). Global warming affects the gene flow across populations because it reduces connectivity; this anthropogenic alteration is more relevant in peripheral populations because the loss of their genetic diversity makes them more prone to local extinctions due to
Atlantic salmon *Salmo salar* is an anadromous species with a strong homing behaviour, which makes it especially vulnerable to habitat degradation in its natal streams, mainly due to dams, water extraction and pollution (Almodóvar et al. 2019). Human-induced changes in water temperature or quality, overfishing, and alterations in fluvial connectivity may result in steep reductions in populations as well as modifications of population structure (Hórreo et al. 2014a). Another consequence of returning to their natal rivers is the minimal genetic flow among local populations, so salmon that inhabit neighbouring rivers are to a large degree reproductively isolated. This isolation facilitates the genetic differentiation among wild Atlantic salmon populations (Ayllón et al. 2006) and creates the potential for local adaptation (Ciborowski et al. 2007).

In recent decades, Atlantic salmon has suffered a severe decline, becoming extinct in many rivers, with this decline being particularly alarming in southern Europe (Campos et al. 2008). One of these rivers is the River Sella, which supports one of the largest salmon populations of Spain. Catches of returning salmon by recreational fishing started to decline in the mid-1970s and reached the lowest levels in the last decade (Nicola et al. 2018). This trend has been observed throughout northern Spain, including in the nearest neighbours of the Sella, where the conservation status of Atlantic salmon is considered as non-vulnerable in only 4 out of 32 salmon rivers (WWF 2001). Additionally, this decline has been accompanied by alterations in the biology of the species in northern Spain, such as a reduction in the average size and age of returning adults, a delay in river entry, changes in genetic structure and reduction of the river area used by the species (A. Almodóvar, unpubl. data). This raises an obvious conflict in salmon conservation, and the urgent need to negotiate the trade-off between the risk of losing the genetic diversity of local populations and the exploitation of fisheries and the species’ habitat.

Different efforts have been made to stop the decline of Atlantic salmon populations, such as stocking, native supportive breeding and habitat restoration. However, these actions should be preceded by an accurate genetic study of the populations. For example, stocking may initially increase the local genetic diversity but the introduction of a foreign genome into a wild native population could damage local adaptations (García de Leániz et al. 2007).

The salmon stocking programs in Spain, based on native fish, began in the early twentieth century. In the study area, all breeders came from local rivers, including the River Sella, and the alevins used for stocking were reared in local fish farms and released annually in all salmon rivers (between 100,000 and 300,000 alevins). However, over time these practices changed, and heavy stockings of salmon imported from northern Europe were performed in Spanish rivers between the 1970s and 1990s, although they failed in their primary purpose of increasing the census population size (Ciborowski et al. 2007, Campos et al. 2008). Fertilized eggs were imported mainly from Scotland to supplement rivers in the study area. They were reared in fish farms until the alevin stage, when they were released (between 20,000 and 100,000) annually at similar densities in all rivers.

The use of native supportive breeding to preserve genetic diversity and possible local adaptations of populations has been recognized worldwide. However, as has previously been seen, this action can cause both genetic and diversity losses if it is not well monitored and planned (Hórreo et al. 2012).

Some authors have described the loss of genetic integrity in native Atlantic salmon populations due to the introgression produced by farmed escapees in the wild and by the practice of stocking with foreign specimens (Perrier et al. 2013a,b, Le Cam et al. 2015, Glover et al. 2012, 2017, Lehnert et al. 2019, Sylvestre et al. 2019). Cross-breeding of stocked individuals with natives should reduce the genetic integrity of the wild population and, if the pressure of invasion continues, there should be a decrease in general population fitness (Sylvestre et al. 2019). The loss of native alleles throughout generations also implies the appearance of less adapted individuals. This process of replacement of native alleles may have contributed to the current population declines, since natural selection operates against introgressed individuals (Almodóvar et al. 2006). In the case of Atlantic salmon there are not enough long-term comparative studies, since temporal analyses usually include short time intervals (Ayllón et al. 2006).
The primary aim of the present work was to assess and quantify any changes in the genetic diversity of the Atlantic salmon population from the River Sella from the 1950s until the 2000s using microsatellite loci. The uniqueness of this temporal comparison is the use of archival scales collected before the stocking practices began, which provide an exceptional source to infer the demographic history of wild salmon populations in its southernmost distribution over 50 yr. Temporal analyses with microsatellite loci have been commonly employed to obtain information about temporal genetic variation because they allow identification of human impacts on populations in terms of introgression, structure and genetic diversity, as well as variations in the effective population size (e.g. Hórreo et al. 2011b, 2014a). Three questions were tested with respect to the results of stocking practices: (1) Has the stocking of foreign specimens changed the genetic variability parameters of Atlantic salmon populations? (2) Is there any loss of native alleles? (3) Is there any significant decline in demographic parameters such as the effective number of breeders (Nb), effective population size (Ne) and their ratio to census size (Nc)?

2. MATERIALS AND METHODS

2.1. Study population

The River Sella is among the most important basins for salmon fishing in northern Spain and supports one of the largest Atlantic salmon Salmo salar populations in the Iberian Peninsula. It is a Site of Community Importance (Code ES1200032) within the Habitats Directive (92/43/EEC), and contributes to the conservation of European freshwater ecosystems. The River Sella is almost free from human alterations, whereas in other southern rivers, multiple anthropogenic stressors can confound or mask interpretations of the natural changes salmon populations may be experiencing.

The analysis includes a sample of the archival River Sella population, represented by dried scales from 172 Atlantic salmon returned after 2 winters at sea (MSW), caught by recreational fishing and collected by the National Inland Fishing and Hunting Service of the Ministry of Agriculture of Spain in 1958 to 1960 (hereafter referred to as ‘archival sample’). In addition, 146 contemporary samples of adipose fin clips of returning adults collected in 2007 and 2008 (hereafter referred to as ‘contemporary sample’) were included for comparing with the archival data.

A baseline of Atlantic salmon microsatellites recently developed for the Eastern Atlantic (SALSEA, Gilbey et al. 2018) was used to identify the possible introgression of northern European fish currently present in the contemporary sample. We exclusively used the baseline for Atlantic salmon from Scottish origin (hereafter referred to as ‘Scottish broodstocks’), because most individuals used for massive stocking in the River Sella shared this origin.

2.2. Molecular analyses

Genomic DNA was extracted from archival scales using the QIAamp DNA Mini Kit (QIAGEN, IZASA). DNA from contemporary samples was extracted from fin clips with the Chelex resin protocol (Estoup et al. 1996). The quality and concentration of DNA was determined by spectrophotometry and was verified by 0.8% agarose gel electrophoresis. Eleven microsatellite loci were analysed: Ssa85, Ssa197, Ssa202 (O’Reilly et al. 1996), SSOSL85, SSOSL311, SSOSL417 (Slettan et al. 1995), SSOSL438 (Slettan et al. 1996), SsaF43 (Sánchez et al. 1996), SSspG7, SSsp1605 and SSsp2210 (Paterson et al. 2004). For each locus, polymerase chain reaction (PCR) was performed in a volume of 25 µl. PCR reactions contained 1.5–2.5 mM MgCl₂, 1× Mg-free PCR buffer, 200 µM of dNTPs, 0.16 mM of each primer, 1–1.25 U of HotSplit DNA polymerase (Biotools) and 50–100 ng genomic DNA template. Specific annealing temperatures for the loci were 42°C (SSsp1605), 50°C (SSOSL85, SSOSL311), 53°C (SSOSL417, SSOSL438), 55°C (SsaF43, SSspG7) and 58°C (Ssa85, Ssa197, Ssa202, SSsp2210).

The amplifications were performed using the following conditions depending on the primers: 95°C for 5 min, 30–40 cycles of 95°C for 20–40 s, 42°C up to 58°C for 20–40 s and 72°C for 20–40 s, with a final extension at 72°C for 10 min. PCR products were visualized on an ABI PRISM 3730 sequencer (Applied Biosystems), and allele scoring was performed manually with the Peak ScannerTM Software v1.0 (Applied Biosystems). All procedures with archival and contemporary samples were kept separate and performed at different times. The preparation of the buffer solutions, DNA extraction and PCR amplification were all performed in a laminar flow hood. The work area and the equipment were cleaned with UV light followed by ethanol before work on the next step was started, and strict cleaning procedures were respected by all staff. Negative and positive controls were applied in all steps of the extraction and genotyping process. Finally, multiple extractions and PCR
from the same archival and contemporary samples were undertaken at different times.

### 2.3. Data analyses

MICRO-CHECKER v2.2.3 (Van Oosterhout et al. 2004) was used to assess the frequency of null alleles and scoring errors due to stuttering or large allelic drop-out. The combined use of 2 or 3 methods is the best strategy for minimizing the false-positive and false-negative rates (Dąbrowski et al. 2014). For this reason, 2 different analysis methods were used in addition to test for the presence of null alleles: CERVUS v3.0.3 (Kalinowski et al. 2007) and ML-NullFreq (Kalinowski & Taper 2006).

Tests for Hardy-Weinberg equilibrium and linkage disequilibrium were carried out for each locus and sample with the software GENEOPOP v4.1 (Rousset 2008). Allele frequencies, number of alleles, observed ($H_o$) and expected ($H_e$) heterozygosity were calculated at the population level with GENETIX v4.05.2 (Belkhir et al. 2004). FSTAT v2.9.3 (Goudet 1995) was used to compute allelic richness ($A_R$) as well as the genetic differentiation ($F_{ST}$) between temporal samples (10,000 permutations). Differences in heterozygosity and allelic richness between archival and contemporary samples were analysed by chi-square test using STATISTICA v8.0. Estimation of Wright’s fixation indices for the deviation of intra-temporal samples from Hardy-Weinberg expectations for heterozygote disequilibrium ($F_{IS}$) were estimated using GENETIX v4.05.2 software. Locus by locus analysis of molecular variance (AMOVA) between temporal samples was performed using ARLEQUIN v3.5 (Excoffier & Lischer 2010).

Full-sib relationships in archival and contemporary samples were estimated using COLONY v2.0.6.5 (Jones & Wang 2010) assuming a Full Likelihood (FL) method with female and male polygamy allowed. An empirical range of genotyping error rates (0.4 to 3.2% across loci) was used for each population (see Table A1 in the Appendix).

Evidence for a demographic bottleneck was examined using 2 approaches. First, heterozygosity excess was analysed in the archival and contemporary samples using BOTTLENECK v1.2.02 (Piry et al. 1999), assuming a 2-phase mutation model (TPM) with percentages of 70, 80 and 90% in the stepwise mutation model, a variance of 12 among multiple steps and 10,000 iterations. Significance of heterozygous excess was statistically tested with a 1-tailed Wilcoxon’s signed-rank test. In addition, we estimated the ratio of the number of alleles to the range in allele size ($M$-ratio) using the M_P_VAL software as described by Garza & Williamson (2001). This was compared to a critical value of $M$ ($M_c$) from a theoretical population in mutation-drift equilibrium, using the CRITICAL_M software (Garza & Williamson 2001), assuming pre-bottleneck effective population sizes of 50, 100, 500 and 1000 and a mutation rate ($\mu$) of $5 \times 10^{-4}$. Following the recommendations of Peery et al. (2012), the proportion of 1-step mutations ($p_g$) was set to 0.22 and the mean size of non-1-step mutations ($\Delta g$) to 3.1.

To assess introgression of Scottish broodstocks in our archival and contemporary Sella samples, the estimation of individual admixture coefficients was performed using the Bayesian clustering method implemented in STRUCTURE v2.3.4 (Pritchard et al. 2000). STRUCTURE analyses were performed for 1 to 10 clusters ($K$) with 10 replicates for each simulated cluster (burn-in period of 250,000 steps and 1,000,000 Markov chain Monte Carlo [MCMC] sweeps) and applying the admixture model with correlated allele frequencies. Optimal $K$ was determined using STRUCTURE HARVESTER v0.6.94 (Earl & von Holdt 2012). Replicates were aggregated using CLUMP v1.1.2 (Jakobsson & Rosenberg 2007) and graphically displayed with DISTRUCT v1.1 (Rosenberg 2004). The threshold values used to assign individuals to parental or admixed classes were derived from the results obtained for Scottish broodstocks. Thus, individuals with $Q$-values between 0 and 0.190 were classified as Scottish broodstock and those with values between 0.191 and 1 as native River Sella, while individuals with $Q$-values between 0.191 and 0.810 were classified as introgressed individuals.

The power of our microsatellite loci for assigning analysed samples to parental or admixed classes was evaluated using 3 metrics after Våhå & Primmer (2006): efficiency, accuracy and overall performance. To this end, 400 genotypes of parental individuals (200 archival River Sella and 200 Scottish broodstock), 200 backcrosses between parental individuals from both origins (F1), 200 backcrosses of F1 (F2) and 200 backcrosses of F1 with each parental individual were generated in HYBRIDLAB v1.0 (Nielsen et al. 2006). Simulated individuals were analysed using STRUCTURE v2.3.4, with the a priori assumption of 2 groups ($K = 2$) corresponding to native River Sella and Scottish broodstocks. Ten replicates, an admixture model with correlated allele frequencies and a burn-in period of 250,000 steps and 1,000,000 MCMC sweeps were performed. These simulated individuals were classified as parental or admixed classes (F1, F2 and backcrosses
of F1) according to the threshold Q-value of 0.810 to estimate the efficiency, accuracy and overall performance of these assignments.

Principal coordinates analysis (PCoA) was computed to explore the allelic divergence among populations in the multivariate space using GenAlEx v6.5 (Peakall & Smouse 2012). PCoA was computed from the genetic distance calculated across multiple loci for each pair of individuals in the GenAlEx software.

Estimates of census size (\(N_c\)) for the archival (1950s period) and contemporary (2000s period) samples were determined by the harmonic mean from 10 years of catch records of anadromous adults from the River Sella collected by local management authorities. These estimates were corrected considering that catches throughout the fishing period may vary between 20 and 40% of the total salmon population inhabiting a river (Ribeiro et al. 2008). Effective number of breeders over a reproductive year (\(N_b\)) was calculated for each period (1950s and 2000s) using the linkage disequilibrium (LD) method implemented in NeEstimator v.2.1 (Do et al. 2014). A minimum allele frequency cut-off value of 0.02 was employed and 95% confidence intervals were obtained using the jack-knife method. This approach is based on LD between alleles from neutral loci unlinked within populations with random mating and implements a bias correction under a wide range of sample sizes. This method has shown consistent values across different demographic scenarios and has been shown to be the best method to estimate the effective population size in populations with both low to no migration and small effective population sizes (Gilbert & Whitlock 2015), and missing data adjustment. Considering a situation of overlapping generations, \(N_b\) estimates can be biased. Despite this, the method developed by Waples et al. (2014) was applied using 2 simple life history traits to adjust genetic estimates of \(N_b\) for correcting biases due to age structure. This \(N_b\) was adjusted using the ratio between adult life span (AL) and age at maturity (\(\alpha\)) using the following equation:

\[
N_{bAdj} = \frac{N_bLD}{1.103 - 0.245 \times \log(AL/\alpha)} \tag{1}
\]

\(N_{bAdj}\) can be a more easily quantifiable and useful tool for managers but remains less used than Ne (Perrier et al. 2016). Adjusted age at spawning has been estimated to be between 2.65 and 3.04 yr in northern Iberian rivers (Consuegra et al. 2005). Considering this range, an average age at maturity of 2.8 yr was assumed for the River Sella. The AL value was calculated as described by Waples et al. (2014) using a maximum breeding age (\(\omega\)) of 5 yr for Atlantic salmon. \(N_{eAdj}\) was calculated using the equation proposed by Waples et al. (2014):

\[
N_{eAdj} = \frac{N_{bAdj}}{0.485 + 0.758 \times \log(AL/\alpha)} \tag{2}
\]

In addition, effective population size \(N_{bSIB}\) was estimated using the sibship assignment method implemented in COLONY v2.0.6.5 (Jones & Wang 2010), assuming a random mating population and a 'Known Ne' prior. The 'Known Ne' prior includes an estimated sex ratio of 1.1 (Álvarez et al. 2010) and an estimated Ne of 10% of \(N_c\), assuming a ratio for Ne/Ne of 0.1 (Verspoor et al. 2007). \(N_{bLD}\) and \(N_{bSIB}\) estimates were carried out after removing the individuals assigned as admixed in the contemporary samples. The effective size ratios \(N_{bSIB}/N_c\) and \(N_{eAdj}/N_c\) were also calculated following Perrier et al. (2016).

### 3. RESULTS

#### 3.1. Genetic variation and introgression analysis

Eleven microsatellite loci were analysed in archival sample of Atlantic salmon *Salmo salar*, with an average successful amplification ranging from 88 to 99% and a variation between PCR replicates of 96 to 100% depending on the locus. The loci SSOSL438, SsaF43 and Ssa85 proved difficult to amplify in the archival sample, with no allelic information in more than 40% of the individuals. Thence, these 3 loci were excluded from the analyses and were not amplified in the contemporary sample.

Degradation and the small quantities of DNA in the archival sample could have caused some allelic dropouts, but MICRO-CHECKER analyses (at the Bonferroni confidence level) did not exhibit any evidence of null alleles, large allelic dropout or stuttering. The results from CERVUS showed similar results for this sample, with an intermediate null allele frequency (\(F_{null} < 0.05\)). However, the analysis of ML-NullFreq software indicated a null allele frequency for loci SSOSL417 and SSspG7 that was higher and significant (\(F_{null} ≥ 0.02, p < 0.05\)) in the archival sample. For true null alleles, consistency among different methods and temporal populations is expected (Dąbrowski et al. 2014). Thus, the null alleles observed did not justify the elimination of any locus from the data analysis.

A total of 92 alleles from the archival sample and 116 from the contemporary sample were observed in the 8 loci analysed. The number of alleles per locus ranged from 6 to 15 in the archival sample, and from 8 to 23 in the contemporary one (Table 1). The esti-
A decrease was found at the temporary sample. Similarly, a marked temporal sample was high and significant over time.

An abrupt increase in the allele number was observed over time in the contemporary sample. Likewise, a marked increase in the allele number was observed over time in the contemporary sample.

Heterozygosity was high and stable and did not differ between temporal samples (p > 0.05), despite a drastic decrease in population abundance, as inferred from the pattern of declining rod catches (Fig. 1). However, the number of alleles showed significant temporal heterogeneity between archival and contemporary samples (p < 0.01, average A: 11.5 in the archival and 14.5 in the contemporary sample). Likewise, a slight tendency of increasing allelic richness was observed over time in the River Sella (average A_R: 11.7 in the archival and 14.3 in the contemporary sample). These consistent differences in allelic richness were mainly due to the abrupt increase in the allele number at the SSOSL311 locus in the contemporary sample. Similarly, a marked decrease was found at the SSOSL85 locus over time.

Genetic differentiation between the temporal samples was high and significant over loci (average \( \Phi_{ST} = 0.013 \), p < 0.001). Locus-by-locus AMOVA revealed that this differentiation was significant for 6 loci (Table 1).

We analysed the degree of association between the allele frequencies of the archival and contemporary samples, which, as expected, was very high and significant (r = 0.928, r^2 = 0.86, p < 0.001, n = 132). It was also significant, but lower, between the archival sample and Scottish broodstocks (r = 0.571, r^2 = 0.33, p < 0.001, n = 69), while the degree of association was

![Fig. 1. Long-term changes in total annual catch of Atlantic salmon (number of fish) in the River Sella from 1949 to 2019. The Mann-Kendall test was used to identify a significant negative trend (dotted line S = −0.432, p < 0.001)](image)

| Locus          | Ssa197 | SSOSL85 | SSOSL311 | SSOSL417 | SSsp2210 | SSspG7 | SSsp1605 | Ssa202 |
|----------------|--------|---------|----------|----------|----------|--------|----------|--------|
| **Archival sample** |        |         |          |          |          |        |          |        |
| N              | 172    | 165     | 166      | 151      | 171      | 162    | 157      | 153    | 149    |
| A              | 11.500 | 10      | 14       | 10       | 15       | 15     | 15       | 15     | 6      |
| A_R            | 11.658 | 13.127  | 12.543   | 9.596    | 12.968   | 8.038  | 13.674   | 5.267  | 8.000  |
| H_e            | 0.753  | 0.860   | 0.820    | 0.796    | 0.777    | 0.768  | 0.789    | 0.553  | 0.661  |
| H_o            | 0.736  | 0.842   | 0.813    | 0.768    | 0.760    | 0.765  | 0.777    | 0.542  | 0.618  |
| F_{IS}         | 0.026  | 0.023   | 0.011    | 0.039    | 0.024    | 0.006  | 0.018    | 0.024  | 0.072  |
| HWE            | ns     | ns      | ns       | ns       | ns       | ns     | ns       | ns     | ns     |
| **Contemporary sample** |        |         |          |          |          |        |          |        |
| N              | 146    | 146     | 149      | 151      | 18       | 147    | 143      | 144    | 146    |
| A              | 14.500 | 19      | 12       | 23       | 18       | 10     | 16       | 8      | 10     |
| A_R            | 14.283 | 16.239  | 10.713   | 19.381   | 16.762   | 9.559  | 13.041   | 7.287  | 8.989  |
| H_e            | 0.799  | 0.864   | 0.787    | 0.883    | 0.870    | 0.783  | 0.768    | 0.647  | 0.786  |
| H_o            | 0.789  | 0.863   | 0.729    | 0.879    | 0.875    | 0.759  | 0.811    | 0.628  | 0.767  |
| F_{IS}         | 0.016  | 0.004   | 0.078    | 0.007    | -0.002   | 0.034  | -0.052   | 0.034  | 0.028  |
| HWE            | ns     | ns      | *        | ns       | ns       | ns     | ns       | ns     | ns     |
| \( \Phi_{ST} \) | 0.013*** | 0.004*  | 0.016**  | 0.025***  | 0.021***  | 0.005* | 0.004  | 0.006  | 0.023** |

*p < 0.05, **p < 0.01, ***p < 0.001
higher between the contemporary sample and Scottish broodstocks \(r = 0.657, r^2 = 0.43, p < 0.001, n = 69\), which may be indicative of a higher level of introgression in the contemporary sample.

The variation (increase or decrease) of allele frequencies between archival and contemporary samples was significantly associated with the difference between allele frequencies originally present in the River Sella and in Scottish broodstocks \(r = 0.646, r^2 = 0.42, p < 0.001, n = 69\). That is, increases were positively correlated with relatively high allele frequencies in Scottish broodstocks, while decreases were correlated with relatively high allele frequencies in the River Sella.

The contemporary sample showed evidence of genetic erosion due to disappearance of native alleles that were present in the archival sample. Regarding the most frequent alleles in the archival sample (i.e. with at least 4% frequency; Table 2), SSOSL85*194 and SSspG7*110 were not found in the contemporary sample.

Furthermore, 14 alleles of the 8 loci analysed (with at least 4% frequency) showed a decrease in the contemporary sample (range: 4.12–15.24%): Ssa197*211, SSOSL85*190, SSOSL85*218, SSOSL85*220, SSOSL311*125, SSOSL311*147, SSOSL417*157, SSOSL417*171, SSsp2210*126, SSsp2210*162, SSspG7*118, SSsp1605*222, Ssa202*240, and Ssa202*244.

In contrast, 3 new alleles were detected in the contemporary sample (with at least 4% frequency) that had not been observed in the archival sample: SSOSL311*123, SSOSL417*179 and SSsp2210*158.

Likewise, the comparison of allele frequencies between archival and contemporary samples showed an increase higher than 4% (range: 4.66–8.78%) in 10 alleles of 6 loci: SSOSL85*182, SSOSL85*196, SSOSL85*222, SSOSL311*145, SSO SL417*201, SSOSL417*203, SSsp2210*130, SSsp1605*242, Ssa202*236 and Ssa202*268. Four of these alleles were also present in relatively high frequencies and were widely distributed in Scotland according to the SALSEA baseline: SSsp2210*130 (37.16% of mean frequency in Scotland), SSsp1605*242 (24.97%), Ssa202*236 (16.22%) and Ssa202*268 (5.87%). Therefore, they are good indicators of introgression of Scottish non-native alleles in native populations.

The level of population genetic differentiation \(F_{ST}\) between temporal samples (archival vs. contemporary) was significant \(F_{ST} = 0.011, p < 0.001\) and consistent with the results observed in allelic diversity.

The BOTTLENECK test did not detect significant excess heterozygosity for the TPM model with percentages of 70, 80 and 90% in the stepwise mutation model \(p > 0.05\); Table 3). Similarly, \(M\)-ratio values were not significantly lower than the simulated value of \(Mc\) under mutation-drift equilibrium for the lower

| Locus       | Allele   | Archival sample \(\bar{x}\) | Contemporary sample \(\bar{x}\) | \(\Delta\) | Direction of change | Scottish broodstocks \(\bar{x}\) |
|-------------|----------|-----------------------------|-----------------------------|----------|---------------------|-----------------------------|
| SSsp1605    | *222     | 58.75                       | 50.34                       | −8.41    | ↓                   | 20.26                       |
|             | *242     | 3.75                        | 8.62                        | +4.87    | ↑                   | 24.97                       |
| Ssa197      | *211     | 16.97                       | 7.88                        | −9.09    | ↓                   | 6.80                        |
| Ssa202      | *236     | 5.26                        | 14.04                       | +8.78    | ↑                   | 12.52                       |
|             | *240     | 33.55                       | 24.66                       | −8.89    | ↓                   | 15.05                       |
|             | *244     | 46.71                       | 33.22                       | −13.49   | ↓                   | 15.63                       |
|             | *268     | 5.26                        | 13.01                       | +7.75    | ↑                   | 5.87                        |
| SSOSL85     | *190     | 19.88                       | 15.36                       | −4.52    | ↓                   | 11.24                       |
|             | *194     | 5.72                        | 0.00                        | −5.72    | Lost                |                             |
|             | *196     | 2.11                        | 10.36                       | +8.25    | ↑                   |                             |
|             | *218     | 5.42                        | 0.36                        | −5.06    | ↓                   |                             |
|             | *220     | 8.73                        | 2.14                        | −6.59    | ↓                   |                             |
|             | *222     | 2.41                        | 8.57                        | +6.16    | ↑                   |                             |
| SSOSL311    | *123     | 0.00                        | 8.87                        | +8.87    | ↑                   |                             |
|             | *125     | 35.10                       | 19.86                       | −15.24   | ↓                   |                             |
|             | *145     | 14.24                       | 19.50                       | +5.26    | ↑                   |                             |
|             | *147     | 14.90                       | 5.32                        | −9.58    | ↓                   |                             |
| SSOSL417    | *157     | 38.60                       | 23.96                       | −14.64   | ↓                   |                             |
|             | *171     | 16.37                       | 7.29                        | −9.08    | ↓                   |                             |
|             | *179     | 0.00                        | 5.56                        | +5.56    | New                 |                             |
|             | *201     | 4.39                        | 9.72                        | +5.33    | ↑                   |                             |
|             | *203     | 2.63                        | 7.29                        | +4.66    | ↑                   |                             |
| SSspG7      | *110     | 4.78                        | 0.00                        | −4.78    | Lost                | 0.08                        |
|             | *118     | 18.79                       | 14.34                       | −4.45    | ↓                   | 11.24                       |
| SSsp2210    | *126     | 20.68                       | 16.21                       | −4.47    | ↓                   | 9.97                        |
|             | *130     | 16.67                       | 22.07                       | +5.40    | ↑                   | 37.16                       |
|             | *158     | 0.00                        | 7.24                        | +7.24    | New                 | 1.87                        |
|             | *162     | 8.95                        | 4.83                        | −4.12    | ↓                   | 6.91                        |
and upper values of pre-bottleneck effective population size values (p > 0.05; Table 3). Neither test showed any old or recent bottlenecks in the archival or the contemporary sample.

Full-sib relationship analyses detected 166 and 79 families in archival and contemporary samples, respectively. Archival family samples were frequently composed of 1 individual, except 1 family of 3 individuals and 4 of 2. In the contemporary sample, a similar result was observed, with 1 family of 3 individuals and 10 of 2 individuals.

The assignment efficiency and accuracy of the 5 microsatellite loci in simulated individuals were high, with values ranging from 95 to 100%. When a threshold Q-value of 0.810 was used, efficiency and accuracy decreased by 2% in backcrosses of F1 with archival River Sella parental individuals and by 5% in backcrosses of F1 with parental Scottish broodstocks. These backcrosses were responsible for the observed decreases in overall performance of 3 and 10%, respectively. Therefore, based on the above and the results obtained for the Scottish broodstock baseline, a threshold of Q > 0.810 can be used to identify individuals to parental or introgressed classes in nature populations.

The STRUCTURE analysis indicated that the most likely number of clusters was K = 2 using the Evanno approach. Examination of individual admixture proportions revealed that one cluster corresponded to the archival Sella sample and the other to the Scottish broodstocks. In addition, admixture between the archival Sella sample and Scottish broodstocks was evident in the contemporary Sella sample (Fig. 2). Results from the admixture analysis were strongly consistent across runs and did not show individual changes in the observed Q-values and classes assigned (native, Scottish broodstock or introgressed individuals). The archival sample (native) and Scot-

| Sample            | Heterozygosity excess p-values | M-ratio | Mc         |
|-------------------|-------------------------------|---------|------------|
|                   | 70% SMM 80% SMM 90% SMM       | Ne = 50 | Ne = 100   |
| Archival sample   | 0.844 0.973 0.986             | 0.758   | 0.750      |
| Contemporary sample| 0.963 0.990 0.998            | 0.833   | 0.774      |

Table 3. Heterozygosity excess assuming a 2-phase mutation model with percentages of 70, 80 and 90% in the stepwise mutation model (SMM), M-ratio and critical value of M (Mc) assuming pre-bottleneck effective population sizes (Ne) of 50, 100, 500 and 1000 for the archival and contemporary Atlantic salmon samples from the River Sella.

Fig. 2. STRUCTURE plot for the archival and contemporary samples of the River Sella and putative exotic Scottish broodstock strain Atlantic salmon under the assumption of 2 genetic units (K = 2). Vertical bars represent proportions of membership of each individual to each of the 2 clusters.
tish broodstocks showed a high percentage of membership within these 2 clusters (Fig. 2), with average Q-values of 0.989 and 0.979, respectively. Thus, all individuals in the archival Sella sample were non-admixed, but 55 out of the 146 individuals in the contemporary sample were identified as introgressed from Scottish broodstocks (38%) with Q-values between 0.436 and 0.809, thereby providing strong evidence of admixture. Furthermore, 33% of introgressed individuals from the contemporary Sella sample showed admixture proportions exceeding 10%.

The PCA showed a clear separation between the archival Sella sample and Scottish broodstocks, with the contemporary Sella sample being placed between both of them (Fig. 3). These results were consistent with the STRUCTURE analysis and, although the 3 samples showed intermixing with each other, the analyses mostly separated the archival sample (with higher intrapopulation diversity and absence of introgression) from the admixed contemporary sample.

3.2. Estimates of census size, effective number of breeders, effective population size and their ratio to census size

\( N_c \) values revealed the strong decrease in this parameter in the River Sella over the 5 compared decades (Table 4). Our demographic analyses and interpretations were robust to delete-one jack-knifing of each of our analysed loci. The effective number of breeders estimated using the linkage disequilibrium method (\( N_{bLD} \)) also showed the same decreasing tendency (Table 4). The \( N_{bAdj} \) values were nearly 8% lower than the non-adjusted ones and varied between 812 and 180. The same decreasing pattern was found in \( N_{eAdj} \) but the values were significantly smaller than the census size calculated from the rod catches.

The contribution of mature parr to reproduction was assessed considering a mean of 8 mature male parr per spawning female, in accordance with field studies from Iberian populations (Martínez et al. 2000). The contribution of mature male parr was considered to increase \( N_e \) by 1.5 to 2.2 times in Iberian populations. However, these appear to be maximum values because individual variations in reproductive success were not considered (Garant et al. 2000).

We also estimated \( N_{bSIB} \) with the sibship method (Wang 2009), assuming random mating given that our samples are in Hardy-Weinberg equilibrium. The ob-

| Sample          | Caught | \( N_c \) (20–40%) | \( N_{bLD} \) (CI 95%) | \( N_{bSIB} \) (CI 95%) | \( N_{bAdj} \) (CI 95%) | \( N_{eAdj} \) (CI 95%) | \( N_{bAdj}/N_c \) (20–40%) | \( N_{eAdj}/N_c \) (20–40%) |
|-----------------|--------|------------------|-----------------------|-----------------------|-----------------------|-----------------------|--------------------------|--------------------------|
| Archival sample | 1204   | 3010–6021        | 884 (377–∞)          | 486 (392–619)         | 812 (346–∞)          | 1555 (654–∞)         | 0.13–0.27                | 0.26–0.51                |
| Contemporary    | 266    | 664–1329         | 176 (115–502)        | 62 (43–92)            | 180 (106–464)        | 341 (200–877)         | 0.14–0.27                | 0.23–0.46                |
tain values were significant lower than those obtained with the LD method, probably due to the fact that the LD method underestimates Ne in large populations when sampling effort is limited, which could be our case, since its accuracy relies on the proportion of the population sampled (Ferchaud et al. 2016). Nevertheless, both estimates showed a similar and very pronounced decline (77 to 88%) in the effective population size over time; this decline coincides with the decrease observed in salmon catches.

The estimated ratio between \( N_{b_A} \) and \( N_c \) did not show any temporal variation, with a mean value of 0.20. Likewise, the estimated ratio of \( \frac{N_{e_A}}{N_c} \), based only on adult censuses without taking migration into account, did not change between temporal samples (0.26 to 0.51).

4. DISCUSSION

This study encompasses one of the largest temporal analyses of Atlantic salmon from the Iberian Peninsula to date. Archival scales of Atlantic salmon were collected in the 1950s prior to stocking practices, and we have used them as source material for a temporal comparison with contemporary samples from the 2000s to assess the relative status of current salmon populations in the River Sella, one of the rivers with a more pristine habitat within the southernmost distribution of the species. The Atlantic salmon population from the River Sella was recipient of 687100 foreign alevis introduced from Scotland between 1981 and 1992 (Hórreo et al. 2011a, 2014b).

Our results indicate a deterioration of the River Sella population, since 3 main changes can be observed: (1) the loss of native and distinctive alleles, (2) introgression in the contemporary River Sella sample, and (3) a reduction in the effective number of breeders and effective population size. The very low values of effective size in the contemporary population, together with the deleterious synergistic effects between genetic drift and high introgression rates, emerge as a severe threat to the long-term conservation of the native salmon population. We hypothesise that these genetic modifications are largely attributable to long-term stocking practices. The introduction of foreign stocks of salmon into the affected rivers was intended to counteract human impacts and the rapid decline of populations (Campos et al. 2008). We can now say that these measures, which lacked sufficient genetic knowledge, not only failed in their purpose of increasing population numbers but also introduced a new element of genetic change to populations: the presence of non-adapted individuals with new alleles (Ciborowski et al. 2007, Le Cam et al. 2015).

4.1. Genetic variation and introgression analysis

Loss of original alleles is not commonly reported in Atlantic salmon populations, but, for instance, the nearly extinct population in the River Skjern (Denmark) showed a significant decrease in the number of alleles over 60 yr (Nielsen et al. 1997). Moreover, Ribeiro et al. (2008) found a loss of 7 alleles over 5 yr in the River Eo, northern Spain, which they attributed to immigration, sampling strategy or, less likely, mutation events. However, a similar analysis carried out in several northwestern Spanish rivers, comparing scale collections from 1950–1960 and modern samples from 1997–2004 did not find any loss of original alleles (Saura et al. 2006), and the authors concluded that average heterozygosity and allelic richness were very similar in the modern and old samples. In contrast, our study that considers a 50 yr lapse of time seems to provide good evidence of such a change in the stocked populations of the River Sella: the loss of 2 formerly frequent alleles (5.72 and 4.78% frequency in the archival sample), a large decrease (range: 4.12–15.24%) in the frequency of 14 alleles (originally occurring with at least 4% frequency), the occurrence of 3 new alleles (at least 4% frequency) in the contemporary sample, and a large increase (range: 4.66–8.78%) in the frequency of 10 alleles already present in the past.

Domestic Atlantic salmon strains were introduced into the Sella population as shown by the 38% putative non-native Scottish broodstock in the contemporary sample compared to the archival non-introgressed one. This indicates that foreign individuals released over decades, who succeeded in introgression with local salmon, introduced new alleles into the natural populations (see also Martínez et al. 2001, Hórreo et al. 2014b). Furthermore, stocking with juvenile salmon born in hatcheries may alter life-history traits of adaptive significance (Le Cam et al. 2015), as their relative reproductive success is nearly half that of wild-born fish (Milot et al. 2013).

Perrier et al. (2013b) found a 10 to 25 times lower survival of stocked fish relative to wild specimens, which might suggest that genetic introgression due to stocking practices may not be a relevant problem. Anyway, stocked precocious males always have a great opportunity to elevate maladaptive gene flow, even if they do not survive to the post-smolt stage.
However, the genetic deterioration found here in the River Sella rules out the idea suggested by Perrier et al. (2013b), because any introgression of foreign genetic material causing a reduction in genetic diversity and/or a loss of unique genetic combinations can be a serious threat, since salmon populations are characterized by strong adaptations to local conditions (García de Leániz et al. 2007).

Our results provide evidence that heterozygosity was high and stable and did not differ between temporal samples. However, the contemporary sample presented a higher allelic richness due to the occurrence of new foreign alleles. These 2 temporal samples showed high polymorphism values and no signs of old or recent bottlenecks.

In COLONY analyses, the archival sample showed 166 full-sib families of typically 1 individual, with the exception of 1 family of 3 individuals and 4 of 2. Likewise, the contemporary sample was composed of 79 full-sib families of generally 1 or 2 individuals.

In conclusion, the increase in allelic richness over time in the River Sella, the loss or decrease in frequency of native alleles, and the appearance or increase in frequency of other alleles could be the result of a genetic introgression process due to the stocking made in the 1980s, mainly with Scottish salmon. Similar results have been found in studies with microsatellites in nearby rivers, such as the Esva and Cares (Ayllón et al. 2006), and in more remote locations, such as the River Nivelle in France (Martínez et al. 2001) and other French rivers (Perrier et al. 2013a). Other genetic markers such as mitochondrial DNA have also revealed an increase in genetic diversity in salmon populations from the Iberian Peninsula due to the introduction of foreign haplotypes (Ciborowski et al. 2007).

4.2. Estimates of census size (Nc) and effective number of breeders (Nb), effective population size (Ne) and their ratio to Nc

The present study also represents an attempt to document the long-term possible variation in number of breeders, effective population size and their ratio to census size in Atlantic salmon, before and after the abrupt decline of the population.

The NbLD values decreased by 78% over time (884 in the archival and 176 in the contemporary sample) (Table 4). These values are within the wide range observed in Canadian rivers (42–3784) by Palstra et al. (2009), Johnstone et al. (2013) and Perrier et al. (2016). NbSIB also showed an abrupt change in our 50 yr analysis (486 in the archival and 62 in the contemporary sample), with very low values in the contemporary sample and a decrease of 87%. Similarly, low values were recently found in the River Nivelle, France (23–76, in 2004–2014) by Bacles et al. (2018), and in 10 Canadian rivers (46–186, in 2010–2014) by Ferchaud et al. (2016). Moreover, NbSIB estimates in our study may be biased towards low values due to overlapping generations. An alternative explanation already put forward for Atlantic salmon (Johnstone et al. 2013) might be that the sibship method implemented in COLONY is more likely to be biased when sample sizes are small. This may result in an underestimation of Nb values when studying large populations with small sample sizes that do not accurately reflect the entire sibship network (Wang 2016). However, the estimated NeAdj for each year seems to be a better estimate of the LD within the cohort. In addition, to avoid bias due to overlapping generations using the LD method, the estimates of Nb and Ne have been corrected according to Waples et al. (2013a), incorporating 3 life-history traits of the Sella population.

After adjusting NbLD to consider overlapping generations, NbAdj showed the same downward trend (812 in the archival and 180 in the contemporary sample). The NeAdj estimated from NbLD in our 50 yr analysis also showed an abrupt change (1535 in the archival and 341 in the contemporary sample), with a decrease of 78%. The NeAdj estimates in the present study are within the range observed by other authors in Atlantic salmon populations in 10 Canadian rivers (124–2976; Ferchaud et al. 2016), and also compare well with Ne values in Canada (443–1697; Palstra et al. 2007), Russia (26–8318; Ozerov et al. 2012), France (48–1289; Perrier et al. 2013a) and Spain (37–96; Hórreo et al. 2011b).

The contemporary sample showed a relatively low NeAdj, but this was higher than the proposed specific minimum threshold for Atlantic salmon (Ne = 95). Moreover, the value was also higher than the proposed threshold above which short-term inbreeding depression can be minimized (Ne = 100; Frankham et al. 2014). Therefore, our findings could suggest that negative inbreeding effects may not be a major threat to this population. However, the contemporary sample had an NeAdj estimate under the threshold where no assurance can be given that the evolutionary potential is preserved in the long term (Ne = 1000; Frankham et al. 2014).

The NeAdj/Nc ratio (0.23–0.51) estimates obtained in this study did not show temporal variation. The NeAdj/Nc ratio values in the River Sella are in agree-
ment with those found by Consuegra et al. (2005) (0.18−0.54) in 4 contemporary samples from northern Spain.

The lack of relationship between the Ne\textsubscript{Adj}/Nc ratio and Nc found in the River Sella could be explained by the fact that mechanisms of genetic compensation may counteract Ne\textsubscript{Adj} reductions and thus increase Ne\textsubscript{Adj}/Nc (Palstra & Ruzzante 2008). This genetic compensation can be considered to produce a buffering effect against the loss of genetic diversity at low Nc.

Similarly, the Nb\textsubscript{Adj}/Nc ratio (0.13−0.27) did not show any significant temporal variation, despite the sharp decline in Nc. Moreover, the contribution to reproduction of fish that may not have been considered while calculating Nc may produce a similar Nb\textsubscript{Adj}/Nc ratio while Nc decreases. In southern European populations, where environmental conditions are particularly limiting for salmon, species can make use of alternative mating strategies to increase their reproductive success. In Atlantic salmon, this alternative strategy is specific to males, which can reach maturity and spawn in freshwater without having migrated to sea. Saura et al. (2008) observed that mature parr can increase the effective population size up to 2 or 3 times. A possible explanation is that warmer temperatures allow a faster growth in body size and weight, and this is correlated with reproductive success (Martínez et al. 2000, García-Vázquez et al. 2001). These precocious mature parr can contribute 60% of the total paternity (Saura et al. 2008) and the participation of male parr throughout the generations creates a buffer effect on the population size, compensating a skewed sex ratio between adults (Johnstone et al. 2013). Consequently, if the relative contribution of mature male parr to Nb\textsubscript{Adj} rises in smaller populations or during years with reduced numbers of anadromous fish returning to the river, the Nb\textsubscript{Adj}/Nc ratio is expected to rise or not change while Nc decreases. Moreover, compensation mechanisms by which the Nb\textsubscript{Adj}/Nc ratio decreases less than would be expected during years with reduced Nc may be of prime importance in a conservation context.

4.3. Conclusions

An Atlantic salmon population from one of the main rivers in their southernmost distribution has suffered a substantial change over the last 5 decades, with low Ne and Nb and a steep population decline. Whether by direct impacts such as stocking practices and overfishing, or by indirect effects such as climate change, anthropogenic influence is the main threat for the species. It is difficult to assess which factor is the most influential or how they will shape populations in the future, but what seems clear is that the biology of this species is changing to adapt to the harsh conditions that man has imposed in such a short evolutionary period. However, current genetic status in the River Sella presents a relevant genetic diversity with a higher number of alleles than in the past and a similar H\textsubscript{e} level. Small populations are expected to suffer under the detrimental effects of genetic drift because it leads to loss of genetic diversity. Despite the appearance of higher allelic richness, it is important to determine where those alleles came from and why native alleles have been replaced or have disappeared. The foreign stocks introduced over decades are one of the main reasons for these allele frequency alterations, as well as the complex shifting in the life-history traits of the species. Mature parr may contribute to the increase in genetic variability and enlarge the effective population size.

In summary, Atlantic salmon is a resilient species trying to survive in the adverse conditions that man has imposed over decades. Stocking has been pernicious, and its effects add to those of climate change, being the main human pressures affecting salmon biology and the integrity of the species’ genome. Our findings in southern Europe could be extended to other areas where declining salmon populations have been managed through similar stocking practices.

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Appendix.

Table A1. Genotype error rates and their 95% confidence interval (error rate, CI 95%) of each locus for the archival and contemporary Atlantic salmon samples from the River Sella

| Locus | Archival sample | Contemporary sample |
|-------|----------------|---------------------|
| Ssa197 | 0.009 (0.000−0.029) | 0.004 (0.000−0.018) |
| Ssa202 | 0.029 (0.009−0.166) | 0.013 (0.001−0.050) |
| SSOSL83 | 0.005 (0.000−0.014) | 0.032 (0.002−0.090) |
| SSOSL311 | 0.022 (0.000−0.025) | 0.009 (0.000−0.026) |
| SSOSL417 | 0.007 (0.000−0.039) | 0.007 (0.000−0.026) |
| SSspG7 | 0.004 (0.000−0.052) | 0.004 (0.000−0.014) |
| SSsp1605 | 0.013 (0.000−0.085) | 0.016 (0.000−0.033) |
| SSsp2210 | 0.001 (0.000−0.014) | 0.027 (0.000−0.054) |