Lipid Accumulation Product Is Associated with Insulin Resistance, Lipid Peroxidation, and Systemic Inflammation in Type 2 Diabetic Patients

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\textbf{Background:} Lipid accumulation product (LAP) is a novel biomarker of central lipid accumulation related to risk of diabetes and cardiovascular disease. In this study, we assessed the association of LAP with glucose homeostasis, lipid and lipid peroxidation, and subclinical systemic inflammation in diabetic patients.

\textbf{Methods:} Thirty-nine male and 47 female type 2 diabetic patients were assessed for anthropometrics and biochemical measurements. LAP was calculated as \([\text{waist circumference (cm)} - 65] \times [\text{triglycerides (mmol/L)}]\) in men, and \([\text{waist circumference (cm)} - 58] \times [\text{triglycerides (mmol/L)}]\) in women. Associations of LAP with fasting glucose, insulin, insulin resistance index, lipid and lipoprotein levels, malondialdehyde, and high-sensitive C-reactive protein (hs-CRP) were assessed.

\textbf{Results:} Mean age and LAP index were 53.6±9.6 and 51.9±31.2 years, respectively. After adjustments for age, sex and body mass index status, a significant positive correlation was observed between LAP index and fasting glucose \((r=0.39, P<0.001)\), and homeostasis model assessment of insulin resistance \((r=0.31, P<0.05)\). After additional adjustment for fasting glucose levels, antidiabetic and antilipidemic drugs, the LAP index was also correlated to total cholesterol \((r=0.45, P<0.001)\), high density lipoprotein cholesterol (HDL-C) levels \((r=-0.29, P<0.05)\), triglycerides to HDL-C ratio \((r=0.89, P<0.001)\), malondialdehyde \((r=0.65, P<0.001)\), and hs-CRP levels \((r=0.27, P<0.05)\).

\textbf{Conclusion:} Higher central lipid accumulation in diabetic patients was related to higher insulin resistance, oxidative stress and systemic inflammation.

\textbf{Keywords:} Diabetes mellitus, type 2; Lipid accumulation product; Subclinical inflammation; Oxidative stress

\textbf{INTRODUCTION}

Lipid accumulation product (LAP) index, a newly developed biomarker of central lipid accumulation, has been proposed as an accurate and independent indicator of the risk of insulin resistance, metabolic syndrome, type 2 diabetes and cardiovascular disease [1-3]. LAP, which is estimated based on the combination of waist circumference (WC) and triglyceride levels, and is compared to anthropometric measures, including body mass index (BMI), WC, and waist to hip ratio, has recently been con-
considered a better predictor of all-cause and cardiovascular mortality as well as diabetes development in different ages and ethnic populations [4-6]. Since LAP was developed taking into account both triglyceride levels and WC, it is suggested that this index has a stronger correlation with visceral adiposity, higher levels of lypolysis and adipocytokines including interleukin-6, and plasminogen activator inhibitor-1 [7]. Recent studies report that higher LAP is related to abnormal glucose homeostasis and insulin resistance, as well as elevated alanine aminotransferase, an indicator of the hepatic feature of metabolic syndrome, in apparently healthy individuals [8,9]. Higher LAP was also found to be related to lower levels of sex-hormone-binding globulin and higher free androgen index as potential mediators of cardiovascular disease [10].

Despite data available regarding the association of LAP and cardiometabolic risk factors in healthy populations, little is known concerning LAP and the metabolic status of diabetic patients. Our primary focus in this study was to assess whether LAP index could be related to glucose homeostasis parameters, lipid and lipoprotein levels, lipid peroxidation, and subclinical systemic inflammation in type 2 diabetic patients.

METHODS

Study population
This study was conducted from April 2012 to January 2013. Men and women, aged 25 to 60 years, with a clinical diagnosis of type 2 diabetes for at least 1 year, were recruited from the Iran Diabetes Society and the endocrine clinic of Taleghani Medical Center. Patients were excluded from the study if they had severe impairment of cardiac, hepatic or renal function, gestation or lactation and if they used insulin injection or consumed dietary supplements. Finally, 86 of the initially eligible patients, were included in the study. Written informed consent was obtained from all participants. Ethics approval for the trial was obtained from the Ethical Committee of the Research Institute for Endocrine Sciences, Shahid Beheshti University of Medical Sciences.

Demographics, anthropometrics, and clinical measurement
Trained interviewers collected information using the pretested questionnaires. Information on age, educational levels, medical history and medications, duration of diabetes and oral anti-diabetic drugs, was collected. Anthropometric measurements were assessed by trained staff. Weight was measured to the nearest 100 g using digital scales, while the subjects were minimally clothed, without shoes. Height was measured to the nearest 0.5 cm, in a standing position without shoes, using a tape measure. BMI was calculated as weight (kg), divided by the square of the height (m²). WC was measured to the nearest 0.1 cm, midway between the lower border of the ribs and the iliac crest at the widest portion, over light clothing, using a soft measuring tape, without any pressure to the body.

Biochemical measurement
For all biochemical measurements, 12-hour fasting blood samples were collected into tubes containing 0.1% Ethylenediaminetetraacetic acid disodium salt dihydrate and were centrifuged at 4°C and 500 g for 10 minutes to separate plasma. Fasting serum glucose was measured by the enzymatic colorimetric method using a glucose oxidation kit (Pars Azmun Co., Tehran, Iran). Serum insulin concentrations were measured using an enzyme-linked immunosorbent assay (ELISAs) kit (Merckodia, Uppsala, Sweden). Insulin resistance was estimated using the homeostasis model assessment of insulin resistance (HOMA-IR) index, which is defined as fasting plasma insulin (mU/L) multiplied by the fasting plasma glucose (mmol/L) divided by 22.5.

Serum total cholesterol and triglyceride levels were measured by enzymatic colorimetric analysis with cholesterol esterase/cholesterol oxidase and glycerol phosphate oxidase, respectively (Pars Azmun Co.). High density lipoprotein cholesterol (HDL-C) was measured by the immunoturbidimetry method after precipitation of apo B-containing lipoproteins with phosphotungstic acid (Pars Azmun Co.). Low density lipoprotein-cholesterol was calculated from serum total cholesterol, triglycerides and HDL-C, according to the Friedewald equation.

Serum high-sensitive C-reactive protein (hs-CRP; pg/mL) concentration was measured using the ELISA kit (Diagnostics Biochem Canada Inc., Thames Centre, Ontario, Canada). Serum malondialdehyde (MDA) was measured spectrophotometrically by the thiobarbituric acid reactive substances (TBARs) assay kit (Cayman Chemical Inc., Ann Arbor, MI, USA). Inter- and intra-assay coefficients of variations of all assays were <5%.

LPA index, a novel measure of central lipid accumulation and predictor of metabolic syndrome and cardiovascular disease, was calculated as \[\text{WC (cm)} - 65 \times \frac{\text{triglycerides (mmol/L)}}{2.2} \] in men, and \[\text{WC (cm)} - 58 \times \frac{\text{triglycerides (mmol/L)}}{2.2} \] in women [11,12].


Statistical methods
The Kolmogorov-Smirnov test was used to test for normal distributions. If the variable was not normally distributed, log-
arithm of the skewed variable was entered in the models. The
LAP index was categorized into quartiles (<24.5, 24.5 to 43.9,
44 to 65.5, and >65.5) metabolic parameters of the patients
were compared across the quartile categories using the general
linear models with adjustments for age and gender. A partial
correlation test with adjustments for age, sex, antidiabetic and
antilipidemic drugs, BMI status (≤24.9, 25 to 29.9, ≥30) and
fasting glucose levels was used to clarify the association of
LAP index and BMI with glucose homeostasis parameters,
lipid and lipoprotein levels, MDA, and hs-CRP. To better esti-
mate the association of LAP and the mentioned parameters,
linear regression curve estimation analysis was also conducted
and significant associations were presented as plots. Statistical
analysis was performed with SPSS version 16.0 (SPSS Inc.,
Chicago, IL, USA). A P<0.05 was considered significant.

RESULTS
The mean age of participants was 53.6±9.6 years, and 45%
were men. The mean of LAP index was 22±3.8, 35±3.7, 56±
4.1, and 93±3.7 in the 1st, 2nd, 3rd, and 4th quartile categories
of LAP, respectively. Table 1 presents the characteristics, bio-
chemical and anthropometric values of the participants across
quartile categories of LAP index. Participants in the highest
quartile of LAP also had higher BMI. A significant increasing
trend of serum fasting glucose, insulin, insulin resistance in-
dex, and total cholesterol levels was observed across increas-
ing LAP. Mean levels of HDL-C significantly decreased across
increasing LAP quartiles. Compared to the lowest quartile cat-
egory of LAP index, participants in the highest had higher tri-
glyceride/HDL-C ratios, MDA and hs-CRP levels. Partial cor-
relation coefficients of LAP index and BMI with metabolic
parameters are presented in Table 2. After adjustments for age,
sex and BMI status, significant positive correlations were ob-
served between LAP index and fasting glucose (r=0.39,
P<0.001), and LAP index and HOMA-IR (r=0.31, P<0.05).

Table 1. Demographics, Anthropometric Measurements, and Cardiometabolic Risk Factors of the Patients by Categories of LAP Index

| LAP index | <25.4 (n=22) | 25.4-43.9 (n=21) | 43.9-65.5 (n=21) | >65.5 (n=22) | P valuea |
|-----------|-------------|-----------------|-----------------|-------------|---------|
| Age, yr   | 50.3±8.5    | 57.0±10.5       | 54.3±10.8       | 56.1±9.7    | 0.22    |
| Male, %   | 47.6        | 45.5            | 45.0            | 38.0        | 0.55    |
| Antidiabetic drugs | 18 | 20 | 19 | 21 | 0.65 |
| Antilipidemic drugs | 17 | 14 | 16 | 17 | 0.85 |
| Body mass index, kg/m² | 26.5±1.2 | 26.9±1.3 | 33.3±1.2 | 34.7±1.4 | 0.001 |
| Waist circumference, cm | 91±2.1 | 95±2.2 | 104±2.3 | 105±2.1 | 0.001 |
| Fasting serum glucose, mg/dL | 143±13.6 | 157±13.4 | 146±14.6 | 192±13.1 | 0.051 |
| Fasting serum insulin, mU/L | 6.80±0.88 | 6.49±0.86 | 9.43±0.91 | 9.63±0.88 | 0.017 |
| HOMA-IR   | 2.12±0.51   | 2.84±0.51      | 3.42±0.55      | 4.77±0.49   | 0.014 |
| Total cholesterol, mg/dL | 143±7.5 | 153±7.3 | 160±8.0 | 182±7.1 | 0.001 |
| Triglycerides, mg/dL | 66±12.2 | 96±11.9 | 113±13.0 | 193±11.6 | 0.001 |
| HDL-C, mg/dL | 30.7±0.74 | 30.6±0.73 | 29.0±0.76 | 27.8±0.74 | 0.021 |
| LDL-C, mg/dL | 109±6.7 | 109±6.7 | 108±7.2 | 115±6.5 | 0.84 |
| Triglyceride/HDL-C ratio | 2.23±0.43 | 3.29±0.42 | 3.96±0.45 | 6.93±0.41 | 0.001 |
| Lipid accumulation index | 22±3.8 | 35±3.7 | 56±4.1 | 93±3.7 | 0.001 |
| Malondialdehyde, μmol/L | 5.14±0.45 | 6.13±0.43 | 6.13±0.45 | 9.36±0.45 | 0.001 |
| hs-CRP, ng/mL | 2.29±0.50 | 2.07±0.49 | 3.83±0.51 | 3.44±0.50 | 0.001 |

Values are expressed as mean±SEM.
LAP, lipid accumulation product; HOMA-IR, homeostatic model assessment of insulin resistance; HDL-C, high density lipoprotein cholesterol; LDL-C, low density lipoprotein cholesterol; hs-CRP, high-sensitive C-reactive protein.
*P values compared the mean values across quartile categories of LAP using age- and sex-adjusted analysis of covariance.
LAP has previously been reported as a predictor of diabetes, metabolic syndrome, and cardiovascular disease [2-6]. The odds ratio of diabetes in subjects with high LAP was 7.40 (95% confidence interval [CI], 5.10 to 10.75) and 19.09 (95% CI, 6.57 to 55.50) in Japanese men and women, respectively [1]. Some previous studies indicated that, compared to other anthropometric measures such as WC and BMI, LAP could be considered a better predictor of diabetes development and cardiovascular disease risk [4-6]. A recent cross-sectional study on 2,524 nondiabetic Chinese subjects showed that, compared to BMI and WC, LAP had a greater impact on the insulin resistance index [13]. In an analysis conducted in the third National Health and Nutrition Examination Survey, LAP had better correlation with cardiovascular risk factors, including lipid risk variables, uric acid concentration, and heart rate, among US adults compared to BMI [3]. A possible explanation for these observations may be that the two components of LAP, abdominal fat and triglyceride concentrations, have greater physiological correlations with lipid and lipoprotein metabolism, as well as lipoprotein particle size, compared to BMI (describing lipid over-accumulation) [3]. In our study, compared to BMI, LAP had greater correlation with fasting serum glucose, lipid and lipoprotein parameters, and lipid peroxidation index. BMI rather than LAP was correlated with serum insulin and hs-CRP; the association of both BMI and LAP with HOMA-IR were similar.

In young healthy Korean women, higher LAP was also related to higher postprandial glucose levels, insulin response and homeostatic assessment model of insulin resistance [8]. Polycystic ovary syndrome patients in the higher quartile of LAP, had a risk of impaired glucose tolerance of 41.81 (95% CI, 5.52 to 316.54) [14]. An 11-year follow-up of nondiabetic patients showed that visceral adiposity, but not abdominal subcutaneous fat, directly measured by computed tomography (CT) scan as the volume of intra-abdominal fat at the umbilicus level, was an independent predictor of insulin resistance [15]. In the current study, the insulin resistance index in patients with higher visceral lipid accumulation was twice as high (4.77 ± 0.49 vs. 2.12 ± 0.51, in the first and fourth quartiles, respectively). Some possible mechanisms have been proposed regarding the association of visceral fat and insulin resistance; first, visceral fat, compared to subcutaneous fat, has a higher rate of lipolysis and subsequently, could produce a higher free fatty acid load, which leads to fat accumulation in the liver and induces insulin resistance; second, adipocytokines derived from the visceral fat may be responsible for in-

### Table 2. The Correlation of LAP Index and BMI with Glucose Homeostasis Parameters, Lipid and lipoprotein Levels, Lipid Peroxidation and Systemic Inflammation

| Variable                          | Partial correlation $r$ ($P$ value) |
|-----------------------------------|------------------------------------|
| Fasting serum glucose             | 0.39 (0.001)                       |
| Fasting serum insulin             | 0.15 (0.24)                        |
| HOMA-IR                           | 0.31 (0.014)                       |
| Total cholesterol                 | 0.45 (0.001)                       |
| HDL-C                             | -0.29 (0.021)                      |
| LDL-C                             | 0.02 (0.85)                        |
| Triglycerides/HDL-C ratio         | 0.89 (0.001)                       |
| Malondialdehyde                   | 0.65 (0.001)                       |
| hs-CRP                            | 0.27 (0.032)                       |

LAP, lipid accumulation product; BMI, body mass index; HOMA-IR, homeostatic model assessment of insulin resistance; HDL-C, high density lipoprotein cholesterol; LDL-C, low density lipoprotein cholesterol; hs-CRP, high-sensitive C-reactive protein.

Additionally adjusted for fasting glucose levels, antidiabetic and antilipidemic drugs.
Another finding of this study was the strong association of LAP with total cholesterol, HDL-C levels and triglyceride/HDL-C ratio, independent risk factors of cardiovascular disease. LAP has been correlated with total cholesterol ($r=0.498, P<0.001$), and HDL-C ($r=-0.319, P=0.026$) [10]. In a previous study, visceral adipose tissue assessed by CT scan was significantly related to apolipoprotein B ($\beta=1.33, P=0.001$) and HDL-C ($\beta=-1.89, P=0.004$) [11]. Another study also showed that visceral fat accumulation was correlated with...
apolipoprotein B ($r=0.26, P<0.05$), and HDL-C ($r=-0.26, P<0.05$) [12]. Abnormal levels of adipocytokines caused by higher levels of visceral adiposity, including decreased levels of adiponectin and increased levels of visfatin, have been suggested as mediators of dyslipidemia [17]. Triglyceride/HDL-C ratio in patients with higher LAP was more than 3-fold (6.93±0.41 vs. 2.23±0.43, in the first and fourth quartiles, respectively); this ratio is directly related to lipoprotein particle size and the risk of atherosclerosis [18,19].

In the current study, LAP was strongly correlated with MDA, an important biomarker of lipid peroxidation and oxidative stress, independent of age, gender, and fasting glucose levels. Although the association between LAP and oxidative stress parameters has not yet been determined, previous studies have reported that higher visceral adiposity induced oxidative stress and lipid peroxidation. In healthy men and women, TBARs as biomarkers of systemic oxidative stress were positively related to visceral adipose tissue and development of subclinical atherosclerosis [20]. Visceral fat was also correlated with serum TBARs/cholesterol ratio ($r=0.541, P<0.001$) in patients with metabolic syndrome [21]. Moreover, visceral fat was reported as a significant determinant of expression of genes related to oxidative stress [22].

A moderately significant correlation between LAP and hs-CRP ($r=0.25, P=0.007$), an indicator of subclinical systemic inflammation, was also observed in this study. A similar association was recently reported in postmenopausal women ($r=0.315, P=0.042$) [10]. It is well known that obesity and increased visceral adipocytes contribute to increased levels of several inflammatory proteins such as CRP, interleukine-6, plasminogen activator inhibitor-1, P-selectin, vascular cell adhesion molecule 1, fibrinogen, and $\alpha_1$-acid glycoprotein [23, 24]. In a prospective cohort, a 6-year follow-up of middle-aged individuals showed that increases in visceral adiposity, measured by CT scan, were associated with increased levels of CRP ($r^2=17.9\%$) [25].

Although, previous studies have indicated that LAP index is correlated to some cardiometabolic risk factors, to our knowledge these associations in diabetic patients are reported for the first time in this study. Moreover the correlation between LAP and oxidative stress in diabetic patients has not been previously reported. There were some limitations which might be considered important in the current study: cross-sectional setting and small sample size. Also, some potentially confounding variables including duration of diabetes, levels of glycosylated hemoglobin, and chronic vascular complications in diabetic patients have been not considered in the analysis. In addition, the validity of HOMA-IR as an insulin resistance parameter has not validated in the patients.

In conclusion, LAP showed strong associations with glucose hemostasis parameters, lipid and lipoprotein levels, atherosclerotic lipid parameters, lipid peroxidation, and a subclinical inflammatory marker in type 2 diabetic patients. It is plausible to suggest that LAP may be a useful and simple clinical marker for assessment of cardiometabolic risk factors in type 2 diabetic patients. Further studies of longer durations are recommended to better estimate the power of LAP in the prediction of diabetes complications.

CONFLICTS OF INTEREST

No potential conflict of interest relevant to this article was reported.

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