Ex vivo study of human visceral nociceptors

Cian McGuire,1 George Boundouki,2 James R F Hockley,2 David Reed,2 Vincent Cibert-Goton,1 Madusha Peiris,2 Victor Kung,1 John Broad,1 Qasim Aziz,2 Christopher Chan,1 Shafi Ahmed,1 Mohamed A Thaha,1 Gareth J Sanger,1 L Ashley Blackshaw,2 Charles H Knowles,1 David C Bulmer1

ABSTRACT
Objective The development of effective visceral analgesics free of deleterious gut-specific side effects is a priority. We aimed to develop a reproducible methodology to study visceral nociception in human tissue that could aid future target identification and drug evaluation.

Design Electrophysiological (single unit) responses of visceral afferents to mechanical (von Frey hair (VFH) and stretch) and chemical (bradykinin and ATP) stimuli were examined. Thus, serosal afferents (putative nociceptors) were used to investigate the effect of tegaserod, and transient receptor potential channel, vanilloid 4 (TRPV4) modulation on mechanical responses.

Results Two distinct afferent fibre populations, serosal (n=23) and muscular (n=21), were distinguished based on their differences in sensitivity to VFH probing and tissue stretch. Serosal units displayed sensitivity to key algic ergic mediators, bradykinin (6/14 units tested) and ATP (4/10), consistent with a role as polymodal nociceptors, while muscular afferents are largely insensitive to bradykinin (0/11) and ATP (1/10). Serosal nociceptor mechanosensitivity was attenuated by tegaserod (−20.8 ±6.9%, n=6, p<0.05), a treatment for IBS, or application of HC067047 (−34.9±10.0%, n=7, p<0.05), a TRPV4 antagonist, highlighting the utility of the preparation to examine the mechanistic action of existing drugs or novel analgesics. Repeated application of bradykinin or ATP produced consistent afferent responses following desensitisation to the first application, demonstrating their utility as test stimuli to evaluate analgesic activity.

Conclusions Functionally distinct subpopulations of human visceral afferents can be demonstrated and could provide a platform technology to further study nociception in human tissue.

INTRODUCTION
Abdominal pain is a common presenting symptom of GI disease. For many patients, this is a chronic problem without any clear underlying pathology. Treating chronic abdominal pain is clinically challenging due to the lack of efficacy or presence of gut-specific side effects of many analgesics. As a result, abdominal pain is a significant factor in the long-term morbidity associated with many GI diseases, impacting negatively on many quality of life indicators such as fatigue, sleep and depression.1,2 Conditions characterised by chronic pain are consistently identified as a major burden on the healthcare system3 underpinning the need to develop new, safe and effective treatments from both clinical and socioeconomic perspectives.

Significance of this study
What is already known on this subject?
▸ Dysregulation of peripheral afferent sensitivity is an important mechanism in chronic visceral pain; however, current research in this field has been largely limited to animal models.
▸ Preliminary studies have demonstrated the feasibility of recording from human GI afferents in vitro.
▸ Pilot data suggest that subpopulations of visceral afferents may exist in the human gut.

What are the new findings?
▸ Our studies define the presence of functionally distinct subpopulations of human visceral afferents comparable to those seen in animal studies. Importantly, these include a population of polymodal nociceptors that preferentially respond to algogenic stimuli and are located in the serosa.
▸ Human serosal visceral nociceptor mechanosensitivity is attenuated by treatment with the transient receptor potential channel, vanilloid 4 (TRPV4) antagonist (HC067047), highlighting the therapeutic potential of TRPV4 blockade for the treatment of visceral pain.
▸ Human visceral nociceptor mechanosensitivity is also attenuated by tegaserod suggesting that its efficacy for the treatment of pain in IBS may be mediated through the reduction of visceral nociceptor mechanosensitivity.

How might it impact on clinical practice in the foreseeable future?
▸ Data from our studies will allow a greater understanding of visceral nociceptor function in health and disease.
▸ The presented methodology provides a platform for the identification of novel therapeutic targets and evaluation of novel putative visceral analgesics.
▸ There is an approach to basic research on visceral pain that reduces the need for the use of animal experimentation.
One approach to analgesic development has been to target sensory nerves called nociceptors that transduce painful or noxious stimuli from the periphery to the central nervous system. This approach has therapeutic potential as evidenced by the effectiveness of local anesthetics, which inhibit peripheral input into the pain pathway. A goal of research in this area has been to identify mechanisms specific to nociceptor activation that can reproduce the efficacy of local anesthetics without the side effects associated with pan-sodium channel blockade. In particular, the identification of the transient receptor potential channel (TRP) family of ion channels as key transducers of noxious stimuli such as heat, cold, and pressure, and as downstream effectors of receptor activation by inflammatory mediators such as bradykinin, ATP or prostaglandins, has been the focus of substantive investigation.

More recently, the identification of causative loss-of-function or gain-of-function mutations in specific sodium channel subtypes (Nav1.7, 1.8, 1.9) selectively expressed in sensory nerves with clinical pain phenotypes supports the concept that targeting nociceptor-specific mechanisms can inhibit pain without affecting other sensory modalities including touch.

A limitation to research in this area has been our lack of specific knowledge of human nociceptor function leading to reliance on data from model organisms, principally rodent and guinea pig. This is particularly true for visceral nociceptors where current gold standard approaches to studying human nociceptors such as microneurography or the use of native/ resected human bowel can be obtained on a frequent basis from consenting patients undergoing surgery as part of their standard clinical treatment for GI disease. We therefore sought to develop a methodology to study human nociceptor activity ex vivo using resected human bowel.

Visceral nociceptors have been extensively characterised in rodent by their sensitivity to noxious mechanical stimuli (eg, compression of receptive fields with von Frey hair (VFH) filaments, tissue stretch or high pressure distension), ischaemic and hypoxic conditions and algogenic mediators (eg, bradykinin and ATP). This contrasts with other gut afferent populations that are sensitive to innocuous levels of stretch or light mucosal stroking and which respond to physiological stimuli associated with normal movements of the bowel. Visceral nociceptors have been further characterised in model species, including rodent and guinea pig, into serosal, mesenteric and submucosal subclasses based on the location of their receptive fields in the wall of the gut or mesentery of flat-sheet ex vivo colonic preparations. Pilot data in human tissue suggest similar subpopulations may exist in man.

To identify and characterise nociceptors in human tissue, we assessed electrophysiological responses of discriminated visceral afferent units to the application of mechanical (VFH probing, tissue stretch and mucosal stroking) and chemical (bradykinin and ATP) stimuli to their receptive fields. Further, we examined the effect of tegaserod, a clinically effective treatment of pain in IBS, and transient receptor potential channel, vanilloid 4 (TRPV4) antagonism, a high value target for the development of novel visceral analgesics in IBS.

Finally, we sought to develop a chemosensitivity paradigm in which the efficacy of other novel analgesics could be tested against noxious inflammatory mediators.

**METHODS**

All human tissue was collected and used with the approval of the East London and the City HA Local Research Ethics Committee (NREC 10/H0703/71). Resected human ileum, colon, rectum and appendix were collected after written consent from patients undergoing elective surgery as part of their standard clinical treatment at the Barts Health NHS Trust (London, UK). All tissues were obtained from a histopathologist following pathological examination and used either on the day of collection (n=57) or after overnight (12–15 hours) cold storage (4°C) in carbogenated (95% O2, 5% CO2) Krebs buffer (n=39) (see online supplementary methods).

**Electrophysiology**

Tissues were transferred to a bespoke rectangular recording chamber (100 mm (length)×60 mm (width)×20 mm (depth)), with a cylindrical base (Dow Corning, UK) and pinned flat with serosal facing up (figure 1A). Tissues were superfused with carbogenated Krebs buffer at a rate of 6 mL/min maintained at 32–34°C supplemented with atropine (10 μM) and nifedipine (10 μM) to prevent smooth muscle contractility. Mesenteric nerve bundles were dissected and recorded using suction electrodes as previously described.

**Experimental protocols**

**Characterisation of visceral nociceptors**

Flat-sheet preparations were assessed for mechanosensitivity by serosal probing with a blunt cotton bud, longitudinal and/or circumferential stretch of the tissue with forcep, and in some preparations mucosal stroking with a metal rod. If focal receptive fields were identified, stimulus-response curves were generated using a range of increasing strength VFH probes (0.02, 0.04, 0.07, 0.16, 0.4, 1, 2 and 4 g; 3×3 s probe; at 3 s intervals). The response of mechanosensitive units to the bath application of algogenic mediators bradykinin and or ATP was then assessed.

**Effects of existing and novel therapeutic treatments for IBS on visceral nociceptor mechanosensitivity**

To confirm the stability of repeated VFH probing, time-matched control experiments were performed. In these experiments, three sets (3×3 s) of VFH probes, separated by 5 min intervals, were performed prior to a 5 min administration of either Krebs buffer or dimethyl sulfoxide (DMSO) (0.1%) and repeated for up to an hour thereafter. A similar protocol was then used to determine the effects TRPV4 modulation on visceral nociceptor mechanosensitivity by applying the agonist GSK1016790A (20 mL 10 μM) followed by the antagonist HC067047 (20 mL 100 μM) or in separate studies the 5-hydroxytryptamine (5-HT) receptor 4 partial agonist tegaserod (100 mL 30 μM) (see online supplementary methods).

**Chemosensitivity of putative nociceptors**

In tissues where reproducible mechanosensitivity could not be determined, chemosensitivity to noxious inflammatory mediators was examined. In initial studies, bradykinin, ATP, 5-HT, histamine, prostaglandin E2 (PGE2) or capsaicin, were superfused sequentially into the tissue bath to determine the responsiveness of each preparation. Next, the response to repeat applications of bradykinin, ATP, 5-HT, capsaicin or histamine was examined in separate experiments. A small number of preparations were tested for mechanosensitivity before and after the application of bradykinin to test for the recruitment of silent afferents. Finally, specific bradykinin or ATP receptor agonists (B1 receptor agonist...
Sar-[D-Phe8]-des-Arg9-Bradykinin, P2X receptor agonist α, β-methylene ATP) and antagonists (B1 receptor antagonist R715, B2 receptor antagonist HOE140, adenosine receptor antagonist CGS15943, P2×2/3,3 receptor antagonist RO4) were used to examine the pharmacology of the responses to these mediators. Post hoc analysis were performed to examine the effect of cold storage, age, gender and tissue region, on mechanosensitivity (VFH probing) and chemosensitivity (bradykinin and ATP), in macroscopically normal tissue from surgical resections performed for the treatment of bowel cancer. Additionally, the effects of inflammatory disease were examined by comparing responses with those obtained from inflamed tissue obtained from resections performed for the surgical treatment of IBD, Crohn’s disease and UC.

Data analysis
Data analysis has been described previously.22 Neuronal firing rates were examined offline using data analysis software in Spike 2 (CED, UK). Data sets were analysed using the appropriate parametric and non-parametric tests; paired and unpaired t-tests, Fisher’s exact tests and Pearson’s and Spearman’s correlations (see online supplementary methods). Data are expressed as mean±SEM, and statistical significance was set at p<0.05.

Drugs
Drugs in powder form were solubilised as per the manufacturer’s recommendations, aliquoted and frozen at −20°C until required. Aliquots were diluted in Krebs buffer to final working concentrations. Bradykinin, GSK1016790A, capsaicin, 5-HT, histamine, adenosine and ATP were obtained from Sigma-Aldrich (St Louis, Missouri, USA). HC067047, tegaserod, HOE140, R715, CGS15943, α, β-methylene ATP, Sar-[D-Phe8]-des-Arg9-bradykinin and PGE2 were purchased from Tocris Bioscience (Bristol, UK). RO4 was a gift from Neusentis (Cambridge, UK).

RESULTS
Characterisation of mechanosensitive units
Forty-six mechanosensitive units were identified in 37/97 pieces of tissue recorded (online supplementary table S1). Of these,
two clear subtypes of units could be determined. One subtype found in 23/46 units tested responded to low weight (threshold $\leq 0.6$ g, 100% of units responded at 0.6 g) VFH probing of the serosa but not tissue stretch or mucosal stroking, suggesting localisation within the serosal layer (figure 1B–E). The other subtype found in 21/46 units tested were responsive to either circular or longitudinal tissue stretch (13/18 responding to both) and only responded to high weight VFH probing, if at all (threshold $> 0.6$ g, only 40% of units responded at 4 g max weight tested) suggesting they were located in deeper tissue layers, probably muscle (see figure 1B, D, E and online supplementary tables S2 and S3). These were termed serosal and muscular afferents, respectively, in keeping with nomenclature adopted in animal studies. One of the 21 muscular units also responded to mucosal stroking fulfilling the criteria for muscular-mucosal units. An additional 2 units had receptive fields in the mesentery fulfilling the criteria for mesenteric units. The mesentery was not routinely probed due to the danger of disturbing the recording electrode positioned in the mesentery.

Characterisation of mechanosensitive units by response to application of algogenic mediators

The potential role of human serosal and muscular units in nociception was examined by testing their responsiveness to prototypic algogenic mediators, bradykinin and ATP. Robust responses were observed to bradykinin in 6/14 serosal units tested and ATP in 4/10 units tested (figure 1F). By contrast, no muscular unit responded to bradykinin (0/11) and only one muscular unit responded to ATP (1/10), indicating that serosal but not muscular units were likely to function as visceral nociceptors (figure 1F). Consistent with a role as nociceptors the majority of serosal units (13/23) were silent at rest displaying no ongoing discharge. By contrast, a significantly greater proportion of muscular units displayed spontaneous activity (17/21, p<0.05), and this ongoing activity was significantly greater than that displayed by serosal nociceptors (firing rate muscular 4.1 $\pm 0.8$ spikes/s vs serosal 0.9 $\pm 0.2$ spikes/s, p<0.01, figure 2). Mesenteric units also responded to bradykinin and ATP indicative of a role in transducing noxious stimuli.

Additionally, we found evidence for the presence of a ‘silent’ nociceptive population that only displayed mechanosensitivity to VFH probing following application of bradykinin (n=2, figure 3 and online supplementary table S4).

Effect of repeat testing and vehicle

We examined the stability of serosal nociceptor responses to VFH probing and application of vehicle (0.1% DMSO or Krebs buffer, n=5). Responses to VFH probing were comparable with baseline (100$\pm 9.9$%), following vehicle application, for example, 5 min (109.6$\pm 9.4$%, p>0.05), 10 min (114.1$\pm 9.4$%, p>0.03) and 15 min (106.6$\pm 11.3$%, p>0.03, figure 4Ai, Aii and online supplementary table S3) postvehicle, and remained stable for a further 30 min afterwards (45 min; 84.9$\pm 7.2$%, p>0.03), total test period 45 min.

Effect of TRPV4 ligands on visceral nociceptor mechanosensitivity

Compelling data from animal studies demonstrate a role for TRPV4 in serosal nociceptor mechanosensitivity suggesting that TRPV4 antagonists could be effective treatments of visceral pain. To investigate this further, we examined the effects of TRPV4 ligands on human serosal nociceptor mechanosensitivity. Pretreatment with the TRPV4 agonist GSK1016790A had no effect on mechanosensitivity (n=6, figure 4Bi, Bii and online supplementary table S6), but significantly increased baseline activity in 3/8 units tested by $\geq 50$% (see online supplementary figure S1 and table S7). Strikingly, application of the TRPV4 antagonist HC067047 (at concentrations that attenuates mouse mechanosensitivity, online supplementary figure S2) significantly reduced human visceral nociceptor mechanosensitivity ($-34.9 \pm 10.0\%$, n=7, p<0.05), indicating that TRPV4 antagonists may have utility in the treatment of human visceral pain (see figure 4Ci, Cii and online supplementary table S8).

Effect of tegaserod on visceral nociceptor mechanosensitivity

We examined the effects of tegaserod, a clinically effective treatment of abdominal pain in IBS, on serosal nociceptor mechanosensitivity. Tegaserod reduces pain scores in patients with IBS and rectal sensitivity to distension in healthy subjects indicating an inhibitory effect on pain processing. Animal studies suggest this effect is mediated through a reduction in visceral afferent activity. Tegaserod significantly reduced human nociceptor mechanosensitivity ($-20.8 \pm 6.9\%$, n=6, p<0.05) suggesting that nociceptor inhibition may contribute to the clinical effects of tegaserod IBS (see figure 4Di, Dii and online supplementary table S9).

Chemosensitivity in putative visceral nociceptors

Finally, chemosensitivity was assessed in the remaining preparations using bradykinin, ATP, capsaicin, 5-HT, histamine and PG$E_2$ as prototypic algogenic or disease mediators. At the concentrations tested, the frequency of preparations responding to each mediator ranged from $\sim 40\%$ to 80% (figure 5). Greater afferent discharge was observed to mediators typically associated with the direct production of pain (bradykinin, ATP and
capsaicin) as opposed to mediators associated with nociceptor sensitisation (histamine and PGE2) (see figure 5 and online supplementary table S10). In preparations where single units could be clearly discriminated, 8/15 units tested responded to multiple chemical mediators with 4/8 of these units responding to four or more mediators, highlighting the presence of a specific subpopulation of afferents with polymodal sensitivity.

**Repeat administration**

To determine which mediators might be suitable for future interventional studies, we examined the effect of repeated mediator applications. The response to the first application of bradykinin or ATP was greater than subsequent applications, as previously reported. However, after this initial desensitisation, more consistent responses were obtained to the second and third applications of bradykinin (second 65.2±9.3% vs third 61.1±9.9%, of the response to the first application, n=6; online supplementary table S11) or ATP (second 53.8±12.3% vs third 47.7±10.1% of the response to the first application, n=4, p>0.05, figure 6A, B and online supplementary table S12). Responses to a second application of histamine (1/2, online supplementary table S13) or 5-HT (2/2; online supplementary table S14) were greatly reduced. Preparations did not respond to a third application of these mediators (figure 6C, D). Capsaicin (10 µM) produced a marked inhibition of ongoing nerve activity following initial application.
application. Lower concentrations of capsaicin (100 nM) were also tested. This concentration did not inhibit baseline activity, however, only 9 out of 46 units tested responded to two applications of capsaicin, and the second response to capsaicin typically showed marked desensitisation. Responses to PGE2 were considered to be of insufficient magnitude to be of utility in an interventional paradigm.

**Mediator pharmacology**

We explored the pharmacology of the bradykinin and ATP activation of human visceral afferents. Pretreatment with the selective B2 receptor antagonist HOE140 significantly attenuated the afferent response to bradykinin, while treatment with the selective B1 antagonist R715 had no effect (second bradykinin additions: control 65.2±9.3% vs HOE140 300 nM 27.2±6.5% (n=6, p<0.05; online supplementary table S15) versus HOE140 1 mM 9.3±8.6% (n=4, p<0.05; online supplementary table S16) versus R715 81.6±14.9% (n=6, p>0.05; online supplementary figure S17 and figure 7A,B). The B1 receptor agonist Sar-[(D-Phe)6]-dev-Arg8-bradykinin had no effect on human afferent activity (0/14 preparations tested, online supplementary figure S3A).
For ATP, pretreatment with the pan-adenosine receptor antagonist CGS15943 or the P2X<sub>3</sub> antagonist RO4 did not significantly alter the afferent response to ATP (second ATP additions: control 53.8±12.3% vs CGS15943 81.8±10.6%) (n=6, p>0.05, online supplementary table S18) versus RO4 100.7±39.7% (n=3, p>0.05; online supplementary table S19 and figure 7C, D). However, it should be noted that the P2X receptor agonist α,β-methylene ATP can activate human afferents (1/3 preparations tested, online supplementary figure S3C, D).

Additionally, in one unit which displayed stable responses to repeated capsaicin administration, incubation with the transient receptor potential channel vanilloid 1 antagonist ABT-102 abolished the response to a third application of capsaicin. This unit demonstrated washout to a fourth application of capsaicin (see online supplementary figure S4).

**Post hoc analysis**

Finally, we performed a post hoc analysis of single unit responses to VFH probing, bradykinin and ATP to confirm the viability of preparations following overnight cold storage; and elucidate differences based on age, gender or inflammatory disease. These differences were examined for different regions of the gut where sufficient data were available. Afferent responses were comparable following cold storage, with no significant difference seen in the mechanosensitivity or chemosensitivity of stored tissues compared with tissue used immediately (table 1 and online supplementary table S20). Further data analysis did not reveal any significant difference in responses based on gender (table 2 and online supplementary table S20), age (see figure 8 and online supplementary table S20), tissue region (see table 3 and online supplementary table S20) or between normal tissue, and tissues obtained from patients with inflammatory disease (see table 4 and online supplementary table S22).

**DISCUSSION**

We present a comprehensive investigation, demonstrating the feasibility of studying human nerve endings in situ. Our approach measures action potential firing, the propagating signal in nociceptors, as opposed to other surrogate markers of nerve activation (calcium fluxes or transmitter release). Additionally, we retain the structural complexity of the nerve terminal architecture in conjunction with local cellular interactions, thereby providing an extension and complementary approach to existing cell-based methodologies such as use of isolated human dorsal root ganglia neurons. 35 36

We have characterized two functional subtypes of human visceral afferent fibres innervating the bowel in detail. These are readily distinguished in a manner analogous to widely used classifications in rodent nerves. 14 Specifically, muscular and serosal subtypes defined by the sensitivity of the former but not the latter to tissue stretch, and a clear separation in the activation threshold to VFH probing between the two subtypes. For example, serosal units have low intensity VFH thresholds consistent with a superficial location of their receptive fields in the serosa, while muscular units have a higher threshold suggestive of a receptive field located deeper in the muscle layers. We also provide evidence for other functional subtypes, namely mesenteric, silent nociceptors and muscular-mucosal afferents suggesting that the modality of sensory signalling from the bowel is functionally conserved from rodent to human. In addition, serosal but not muscular afferents were responsive to algogenic mediators, and possessed a paucity of spontaneous activity suggesting that serosal units were most likely nociceptors, and...
hence transmit pain from the viscera in response to tissue damaging stimuli. By contrast, muscular units, being responsive to low threshold stretch of the bowel, are more likely to transduce physiological stimulation, for example, the passage of bolus through the gut.

We evaluated the effect of existing and potential therapeutic treatments for visceral pain on mechanosensitivity to VFH probing. In keeping with the rodent literature, the application of TRPV4 agonists or antagonists enhanced or inhibited human visceral nociceptor activity, respectively. Combined with studies showing endogenous lipid mediators, which stimulate TRPV4, are elevated in IBS, our data suggest that TRPV4 antagonists could be clinically effective in IBS. Further studies are needed to confirm these initial findings, particularly using

| Table 1 The effect of overnight cold storage on visceral afferent mechanosensitivity and chemosensitivity |
|---------------------------------------------------------------------------------------------------------------|
| Mechanosensitivity                                                                                     |
|                                                                                                               |
| **No storage**                                                                                             |
|                                                                                                               |
| Firing rate spikes 2 s⁻¹ probe (proportion responders)                                                        |
|                                                                                                               |
| VFH (g)                                                                                                     |
| All tissues                                                                                                 |
| 0.07                                                                                                        |
| 0.4                                                                                                         |
| 2                                                                                                            |
|                                                                                                               |
| Sigmoid colon                                                                                               |
| 0.07 (6/10) (60%)                                                                                           |
| 0.4 (9/10) (90%)                                                                                            |
| 2 (25.7±3.6 (9/9) (100%)                                                                                  |
| Rectum                                                                                                      |
| 7.8±3.3 (2/3) (67%)                                                                                         |
| 15.7±6.4 (3/3) (100%)                                                                                      |
| 24.2±5.4 (3/3) (100%)                                                                                      |
|                                                                                                               |
| **Cold storage**                                                                                            |
|                                                                                                               |
| Firing rate spikes 2 s⁻¹ probe (proportion responders)                                                        |
|                                                                                                               |
| VFH (g)                                                                                                     |
| All tissues                                                                                                 |
| 0.07                                                                                                        |
| 0.4                                                                                                         |
| 2                                                                                                            |
|                                                                                                               |
| Sigmoid colon                                                                                               |
| 9.0±2.3 (5/5) (80%)                                                                                        |
| 23.2±3.3 (4/5) (80%)                                                                                       |
| 29.6±4.9 (5/5) (100%)                                                                                      |
| Rectum                                                                                                      |
| 7.8±3.3 (2/3) (67%)                                                                                         |
| 15.7±6.4 (3/3) (100%)                                                                                      |
| 24.2±5.4 (3/3) (100%)                                                                                      |
|                                                                                                               |
| Bradykinin                                                                                                  |
|                                                                                                               |
| Proportion responders (spikes 2 s⁻¹)                                                                          |
|                                                                  | Proportion responders (spikes 2 s⁻¹)                                                                          |
| All tissues                                                                                                 |
| 19/26 (73%)                                                                                                 |
| 51.7±10.5                                                                                                   |
| 5/13 (39%)                                                                                                  |
| 37.7±8.1NS                                                                                                  |
| Sigmoid colon                                                                                               |
| 8/13 (62%)                                                                                                  |
| 63.1±19.2                                                                                                   |
| 4/10 (40%)                                                                                                  |
| 32.3±8.4NS                                                                                                  |
| Rectum                                                                                                      |
| 4/6 (67%)                                                                                                   |
| 34.4±18.6                                                                                                   |
| 1/1 (100%)                                                                                                  |
| 54                                                                                                           |
| ATP                                                                                                          |
|                                                                                                               |
| Proportion responders (spikes 20 s⁻¹)                                                                         |
|                                                                  | Proportion responders (spikes 20 s⁻¹)                                                                         |
| All tissues                                                                                                 |
| 14/14 (100%)                                                                                               |
| 30.5±8.0                                                                                                   |
| 9/12 (75%)                                                                                                  |
| 30.2±7.8NS                                                                                                  |
| Sigmoid colon                                                                                               |
| 8/8 (100%)                                                                                                  |
| 32.4±10.9                                                                                                   |
| 6/8 (75%)                                                                                                   |
| 33.2±12.7NS                                                                                                 |
| Rectum                                                                                                      |
| 3/3 (100%)                                                                                                  |
| 31.3±18.6                                                                                                   |
| 1/1 (100%)                                                                                                  |
| 20                                                                                                           |

Table illustrating single unit responses to VFH probing at 0.07, 0.4 and 2 g, bradykinin or ATP in tissues used on the day of operation or following cold storage. The data were analysed for tissue from cancer resections only, and presented for all tissue regions studied, sigmoid colon only and rectum only. Mean±SEM; p>0.05; paired t-test. NS, not significant; VFH, von Frey hair.

| Table 2 The effect of gender on visceral afferent mechanosensitivity and chemosensitivity |
|---------------------------------------------------------------|
| Mechanosensitivity                                             |
|                                                              |
| **Male**                                                                                                      |
|                                                              |
| Firing rate spikes 2 s⁻¹ probe (proportion responders)        |
|                                                              |
| VFH (g)                                                                                                       |
| All tissues                                                                                                    |
| 0.07                                                                                                           |
| 0.4                                                                                                            |
| 2                                                                                                              |
|                                                              |
| Sigmoid colon                                                                                                 |
| 11.8±1.4 (4/9) (44%)                                                                                        |
| 18.1±3.4 (8/9) (89%)                                                                                        |
| 21.5±4.5 (7/7) (100%)                                                                                       |
| Rectum                                                                                                        |
| 15 (1/1) (100%)                                               |
| 26 (1/1) (100%)                                               |
| 33 (1/1) (100%)                                               |
|                                                              |
| **Female**                                                                                                    |
|                                                              |
| Firing rate spikes 2 s⁻¹ probe (proportion responders)        |
|                                                              |
| VFH (g)                                                                                                       |
| All tissues                                                                                                    |
| 0.07                                                                                                           |
| 0.4                                                                                                            |
| 2                                                                                                              |
|                                                              |
| Sigmoid colon                                                                                                 |
| 11.5±2.0 (2/6) (33%)                                                                                         |
| 19.1±4.7 (5/6) (83%)                                                                                        |
| 21.4±5.4 (5/5) (100%)                                                                                       |
| Rectum                                                                                                        |
| 3/3 (100%)                                                                                                   |
| 31.3±18.6                                                                                                   |
| 1/1 (100%)                                                                                                   |
| 20                                                                                                           |
| Bradykinin                                                                                                    |
|                                                              |
| Proportion responders (spikes 20 s⁻¹)                                                                         |
|                                                                  | Proportion responders (spikes 20 s⁻¹)                                                                         |
| All tissues                                                                                                    |
| 15/23 (65%)                                                                                                  |
| 58.5±12.0                                                                                                   |
| 9/16 (56%)                                                                                                   |
| 30.2±5.9NS                                                                                                   |
| Sigmoid colon                                                                                                 |
| 8/15 (53%)                                                                                                   |
| 69.6±22.1                                                                                                   |
| 4/8 (50%)                                                                                                    |
| 36.9±7.3NS                                                                                                   |
| Rectum                                                                                                        |
| 3/3 (100%)                                                                                                   |
| 52.6±20.9                                                                                                   |
| 2/4 (50%)                                                                                                    |
| 16.9±12.3                                                                                                    |
| ATP                                                                                                           |
|                                                              |
| Proportion responders (spikes 20 s⁻¹)                                                                         |
|                                                                  | Proportion responders (spikes 20 s⁻¹)                                                                         |
| All tissues                                                                                                    |
| 17/18 (94%)                                                   |
| 35.4±6.4                                                                                                    |
| 6/8 (75%)                                                                                                    |
| 13.8±3.2NS                                                                                                   |
| Sigmoid colon                                                                                                 |
| 12/13 (92%)                                                   |
| 35.3±8.4                                                                                                    |
| 2/3 (67%)                                                                                                    |
| 10.4                                                                                                          |
| Rectum                                                                                                        |
| 2/2 (100%)                                                                                                   |
| 43.9±23.9                                                                                                   |
| 2/2 (100%)                                                                                                   |
| 13.0±6.4                                                                                                    |

Table illustrating single unit responses to VFH probing at 0.07, 0.4 and 2 g, bradykinin or ATP in tissues from male or female patients. The data were analysed for tissue from cancer resections only, and presented for all tissue regions studied, sigmoid colon only and rectum only. Mean±SEM; p>0.05; paired t-test. NS, not significant; VFH, von Frey hair.
TRPV4 ligands from alternative chemotypes to support selectivity of action over other TRP family members. We also demonstrated an inhibitory effect of tegaserod on human visceral nociceptor mechanosensitivity illustrating how our studies provide information on the reverse translation of clinically effective drugs, in addition to supporting rodent data suggesting a peripheral site of action of tegaserod. Chemosensitivity was assessed to a range of noxious, inflammatory mediators in preparations in which mechanosensitive units could not be identified. We chose to examine responses to capsaicin and bradykinin as prototypic noxious mediators that elicit pain following local injection in humans. ATP was examined, due to its release from the viscera following inflammation or distension and algesic effects following dermal injection in humans, while 5-HT, PGE2 and histamine were selected due to their increased production in the bowel of patients with IBS and clinical efficacy of agents modulating their pharmacology. The goal of these studies was to evaluate the effect of each mediator on visceral afferent signalling, and develop an alternative experimental protocol which could be used to investigate the analgesic potential of novel therapeutic approaches. We showed a subpopulation of fibres are sensitive to a range of mediators, indicating that drugs which block convergent points in the activation of nociceptors by multiple algogenic mediators are needed. We also examined the stability of repeated mediator application, highlighting the suitability of bradykinin and ATP for use as test stimuli. Afferent responses to capsaicin, histamine and 5-HT showed progressive desensitisation to repeat application, and capsaicin also inhibited ongoing nerve activity. The inhibitory effect of capsaicin is consistent with its clinical use in topical creams, which treat pain by desensitising nociceptors. By contrast, the pathophysiological implications of desensitising responses to 5-HT and histamine is unclear given the clinical efficacy of 5-HT3 antagonists and histamine H1-receptor antagonists in IBS, and may be a feature of this experimental system.

Further investigation confirmed that bradykinin stimulates human visceral afferent activity via B2 receptors consistent with rodent data. While responses to repeated application of ATP were not sensitive to blockade with a selective P2X2/3 antagonist (RO4) or pan-P1 adenosine receptor antagonist.

---

**Table 3** The effect of tissue region on visceral afferent mechanosensitivity and chemosensitivity

| Mehanosensitivity VFH (g) | All tissues | Sigmoid colon | Rectum |
|--------------------------|-------------|---------------|--------|
|                          | Firing rate spikes 2 s⁻¹ probe | Proportion responders | Firing rate spikes 2 s⁻¹ probe | Proportion responders | Firing rate spikes 20 s⁻¹ probe | Proportion responders |
| 0.07                     | 8.8±1.8     | 3/13 (23%)    | 9.1±1.6 | 4/8 (50%) | 7.8±7.3 | 2/8 (25%)     |
| 0.4                      | 17.8±2.7    | 12/13 (92%)   | 20.6±3.7 | 7/8 (88%) | 15.7±5.2 | 3/13 (100%)  |
| 2                        | 23.7±3.3    | 11/11 (100%)  | 25.4±4.6 | 7/11 (100%) | 24.2±4.4 | 3/13 (100%)  |
| Chemosensitivity         |             |               |         |            |         |            |
| Bradykinin               | 49.1±8.6    | 24/26 (92%)   | 57.7±14.7 | 12/13 (92%) | 38.3±14.9 | 3/3 (100%)   |
| ATP                      | 30.3±5.4    | 22/23 (92%)   | 32.8±7.9 | 14/16 (88%) | 28.5±13.5 | 2/2 (100%)   |

Table illustrating single unit responses to VFH probing at 0.07, 0.4 and 2 g, bradykinin or ATP in tissues from different tissue regions. The data were analysed for tissue from cancer resections only. Mean±SEM; p<0.05; paired t-test. NS, not significant; VFH, von Frey hair.
(CGS15493), suggesting the response to ATP in our experimental paradigm was largely driven by P2Y receptor activation, consistent with recent findings in mouse and human tissue. Importantly, our work does not preclude the activation of human visceral afferents by P2X$_{2,3}$ or P1 receptors. We have found excitatory responses to the P2X$_{2,3}$ agonist, α,β-methylene ATP (see online supplementary figure S3B–D), highlighting the redundancy and complexity of purinergic signalling.

Post hoc analysis of mechanosensitivity and chemosensitivity demonstrated stability to overnight cold storage of tissue, suggesting that tissue could be transported over distance to recipient laboratories. Additionally, no significant differences were seen in responses analysed by age, gender or tissue region, suggesting that the function of individual fibres may not be greatly affected by these variables. This does not exclude the possibility that differences exist. For example, the tissue region used may be important as the anatomical innervation by vagal, splanchnic and pelvic afferent nerves is markedly different between regions of the gut and functional responses to different stimuli can vary. However, we applied specific stimuli that preferentially targeted spinal nociceptors consistent with a lack of regional differences in responses. In addition, the recent demonstration of reduced visceral afferent sensitivity to noxious stimuli in aged mice, and reduced afferent activity with age in human tissue, highlights the need for further studies on visceral nociception in human tissue.

A further consideration with the use of human tissue is the diversity of patients’ backgrounds. This will result in greater variability of responses compared with tissue from experimental animals, making small changes difficult to see without large sample numbers. Our tissue is obtained from patients with disease, which may polarise patient demographics compared with the populace as a whole. This is particularly apparent in the age and gender of the patients, for example, the majority of our normal tissues came from male patients aged between 50 and 70 years, and our female tissues comes from postmenopausal women.

Finally, our findings in diseased human tissue (IBD) demonstrate how this approach will provide insight into the pathophysiology of naturally occurring disease. Our data set does not support a marked change in sensitivity of visceral afferents in IBD, although clearly more work is needed to understand if this is related to disease or an effect of patient treatments, for example, the use of steroids or antitumour necrosis factor antibodies which could have an effect on afferent signalling.

The use of human visceral nociceptor recordings has great future potential enabling further investigation of the physiology of human nociceptor stimulus transduction, the pharmacology of nociceptor signalling and as a translational platform for the validation and identification of future novel visceral analgesics. As highlighted, a pragmatic approach needs to be taken when using human tissue, particularly when interpreting negative findings; however, there is clear value in using human tissue to study specific, well-designed, research questions on human visceral nociceptor function.

**Correction notice** This article has been corrected since it published Online First. The joint author statement has been added.

**Contributors** Study concept and design (CMcG, DR, CHK, AB, DCB); funding and supervision (DCB, CHK, QA, AB); tissue collection (CMcG, GB, DR, VK, JB, MP, CC, SA, MAT, GS); identification of suitable patients and gathering of patient details (CMcG, GB, VK); data acquisition and analysis (CMcG, DR, VCG, AB, DCB); manuscript preparation (CMcG, DCB, JRFH, VK, DR, VCG, AB, CHK). CMcG is funded by the Dr Hadwen Trust and did not participate in experiments involving animals, or cells or tissues from animals or from human embryos. All authors approved the final version of the manuscript.

**Funding** This work was supported by the Dr Hadwen Trust for Human Research (CMcG), The Royal College of Surgeons (GB), The Biotechnology and Biological Sciences Research Council (BB/153236/1, JRFH), Canadian Association of...
Hypoxia, bradykinin, and prostaglandins stimulate visceral afferents signal different qualities of colonic stimuli in mice. Neuroungastroenterol Motil 2011;23:75–81.

Vehe GN, Robinson ME, Vase L et al. Reversal of visceral and cutaneous hyperalgesia by local rectal anesthesia in irritable bowel syndrome (IBS) patients. Pain 2014;155:1962–75.

Serra J. Mizonurogenesis: an opportunity for translational drug development in neuropathic pain. Neurosci Lett 2010;470:155–7.

Wainger BJ, Buttermore ED, Oliveria JT, et al. Modeling pain in vitro using nociceptor neurons reprogrammed from fibroblasts. Nat Neurosci 2015;18:17–24.

Corveo F, Connell LA. Distribution of somatic and visceral primary afferent fibers within the thoracic spinal cord of the cat. J Comp Neurol 1984;220:88–98.

Brierley SM, Jones RC III, Xu L et al. Activation of splanchnic and pelvic colonic afferents signal different qualities of colonic stimuli in mice. Neuroungastroenterol Motil 2005;17:854–62.

Brierley SM, Carter R, Jones W III, et al. Differential chemosensory function and receptor expression of splanchic and pelvic colonic afferents in mice. J Physiol (Lond) 2005;567:267–81.

Brierley SM, Page AJ, Hughes PA, et al. Selective role for TRPV4 ion channels in visceral sensory pathways. Gastroenterology 2008;134:2059–69.

Ray BS, Neill CL. Abdominal Visceral Sensation in Man. Ann Surg 1947;126:709–23.

Longhurst JC, Dittman LE, Ordway GA et al. Sensitization of visceral afferents to bradykinin in rat urinary bladder contraction and hyperactivity. Part I. J Pharmacol Exp Ther 2008;326:432–42.

Eveerts W, Zhen X, Ghosh D, et al. Inhibition of the cation channel TRPV4 improves bladder function in mice and rats with cyclophosphamide-induced cystitis. Proc Natl Acad Sci USA 2010;107:19084–9.

Raiheli M, Någel Å, Zofp, et al. Plasma histamine levels (H1) during adjunctive H1-receptor antagonist treatment with loratadine in patients with active inflammatory bowel disease (IBD). Inflamm Res 2010;59(Suppl 2):S257–8.

Bernstein JE, Bickers DR, Dahl MV, et al. Treatment of chronic postherpetic neuralgia with topical capsaicin. A preliminary study. J Am Acad Dermatol 1987;17:93–6.

Wouters MM, Balemans D, Van Wanrooy S, et al. Histamine receptor H1-mediated sensitization of TRPV1 mediates visceral hypersensitivity and symptoms in patients with irritable bowel syndrome. Gastroenterology 2016;150:875–87.e9.

Camiller M, Mayer EA, Drossman DA, et al. Improvement in pain and bowel function in female irritable bowel patients with alosetron, a 5-HT3 receptor antagonist. Aliment Pharmacol Ther 1999;13:1149–59.

Maubach KA, Grundy D. The role of prostaglandins in the bradykinin-induced activation of somatic afferents of the rat jejunum in vitro. J Physiol (Lond) 1999;515:277–85.

Hockley JR, Tranter MM, McGuire C, et al. P2Y receptors sensitize mouse and human colonic nociceptors. J Neurosci 2016;36:2364–76.

Blackshaw LA, Brooks SI, Grundy D, et al. Sensory transmission in the gastrointestinal tract. Neurogastroenterol Motil 2007;19:1–19.

Keating C, Nocchi L, Yu Y, et al. Ageing and gastrointestinal sensory function: altered colonic mechanosensory and chemosensory function in the aged mouse. J Physiol (Lond) 2016;594:4549–64.

Keating C, Grundy D. Ageing and gastrointestinal sensory function. Adv Exp Med Biol 2016;891:71–4.

Yu Y, Daly DM, Adam II, et al. Interplay between mast cells, enterochromaffin cells, and sensory signaling in the ageing human bowel. Neurogastroenterol Motil 2016.

Róka R, Alt-Belpaomo A, Salvador-Carter C, et al. Oxydisepsin prevents visceral hyperalgesia but not colonic permeability increase induced by luminal protease-activated receptor-2 agonist in rats. Gut 2007;56:1071–8.

Hughes AM, Harrington AM, Castro J, et al. Sensory neuro-immune interactions differ between irritable bowel syndrome subtypes. Gut 2013;62:1456–65.