Bench-to-bedside review: Citrate for continuous renal replacement therapy, from science to practice

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Introduction

Anticoagulation is required to prevent clotting in the extracorporeal circuit during continuous renal replacement therapy (CRRT). Heparin is still the most commonly used anticoagulant. However, heparins increase the risk of bleeding, especially in critically ill patients. Evidence has accumulated that regional anticoagulation of the CRRT circuit with citrate is feasible and safe. Compared to heparin, citrate anticoagulation reduces the risk of bleeding and requirement for blood products, not only in patients with coagulopathy, but also in those without. Metabolic complications are largely prevented by the use of a strict protocol, comprehensive training and integrated citrate software. Recent studies indicate that citrate can even be used in patients with significant liver disease provided that monitoring is intensified and the dose is carefully adjusted. Since the citric acid cycle is oxygen dependent, patients at greatest risk of accumulation seem to be those with persistent lactic acidosis due to poor tissue perfusion. The use of citrate may also be associated with less inflammation due to hypocalcemia-induced suppression of intracellular signaling at the membrane and avoidance of heparin, which may have proinflammatory properties. Whether these beneficial effects increase patient survival needs to be confirmed. However, other benefits are the reason that citrate should become the first choice anticoagulant for CRRT provided that its safe use can be guaranteed.

Mechanism of regional anticoagulation with citrate

Citrate acts by chelating calcium and therefore inhibits the clotting cascade at several levels. It provides regional anticoagulation, virtually restricted to the extracorporeal circuit. Citrate systems work on simple and shared modalities: a) pre-filter infusion of citrate, which chelates ionized calcium (ionCa²⁺; aiming for an ionCa²⁺ concentration <0.35 mmol/L); b) a replacement infusion of calcium at the end of the extracorporeal circuit or via a separate venous access to correct for the calcium loss into filtrate or calcium-free dialysate; and c) preferable use of calcium-free dialysis or filtration fluids. Some authors report the use of calcium-containing solutions. However, when used in hemodialysis or pre-dilution
hemofiltration, a higher citrate concentration in the filter (mmol citrate/L blood flow) is required to attain a similar degree of hypocalcemia in the circuit [3-8], and when administered for post-dilution in the venous chamber, the risk of local clotting is increased [9,10].

Whereas citrate causes a dose-dependent decrease of ionCa\(^{2+}\) concentration, the relationship between ionCa\(^{2+}\) concentration and the degree of anticoagulation is more complex. Citrate has almost no anticoagulant effect when ionCa\(^{2+}\) levels remain >0.50 mmol/l [11] or >0.56 mmol/L [12]. However, clotting times steeply increase when ionCa\(^{2+}\) levels decrease further. Below 0.25 [11] or 0.33 mmol/L [12] there is near total inhibition of coagulation.

Anticoagulation can be monitored by measuring post-filter ionCa\(^{2+}\) with adjustment of citrate dose according to the desired calcium target. Other protocols are less complex and use a fixed citrate dose proportional to blood flow targeting a citrate concentration in the filter of about 3 mmol/L [13,14].

Citrate is partially removed by filtration or dialysis. Its sieving coefficient is about one for both dialysis and hemofiltration [8]. The removed fraction varies between 0.20 and 0.50 depending on the relationship between blood and effluent flow and on CRRT modality. Citrate requirements are lower with dialysis because, to achieve similar effluent doses, dialysis is possible with lower blood flows. Furthermore, dialysis removes a higher fraction of the infused citrate due to the higher ratio of effluent to blood flow [15]. In case of hemofiltration, removal approaches filtration fraction. The remaining citrate enters the patient where coagulation is restored because a) the circuit blood is diluted in the patient's circulation and b) citrate is rapidly metabolized in the mitochondrial citric acid cycle in liver, kidney and muscle, liberating the bound calcium. As a result, citrate has anticoagulant properties only within the circuit and not in the patient.

**Citrate solutions and acid base effects**

There are numerous citrate solutions for daily practice. The choice depends on local availability, preferences regarding CRRT modality (pre- or post-dilution, diffusion versus convection), legislation and available software. None of the solutions have proven clinical superiority, but the individual type of solution can have different metabolic consequences [13,14].

In principle, citrate is infused either as a separate tri-sodium citrate (TSC) [9,16-20], acid-citrate-dextrose (ACD) solution [4-7,21,22], or as a component of an isotonic citrate in saline with [23] or without citric acid [18] or of a balanced pre-dilution replacement fluid [24-29] (Table 1). Although citrate is primarily used for extracorporeal anticoagulation, it has profound effects on acid-base balance as well. Anticoagulant and acid-base effects are not directly related. The degree of anticoagulation depends on citrate dose and hypocalcemia (see above), while the effect on acid base state depends on metabolism of citrate and the 'apparent strong ion difference' of the solution (SIDa = [Na\(^{+}\)+K\(^{+}\)+Ca\(^{2+}\)+Mg\(^{2+}\)] - [Cl\(^{-}\} + citrate\(^{3-}\)]) [30], that is, the type of cations opposing the citrate anion. For example, only two thirds of the cations in ACD solution are strong ions (that is, Na\(^{+}\)). The buffer strength of ACD is therefore lower than that of the pure TSC solution. Within the body, citrate is rapidly metabolized and the effective SID (SIDE) of the solution is [Na\(^{+}\)+K\(^{+}\)+Ca\(^{2+}\)+Mg\(^{2+}\)] - [Cl\(^{-}\]). Thus, the alkalizing effect of the citrate solution depends on the metabolism of citrate. In clinical practice, the citrate dose is primarily titrated according to its anticoagulant effects (hypocalcemia). Therefore, the buffer strength can be best compared between solutions when expressing SIDe per mmol citrate (Table 1).

The accompanying dialysis or replacement solutions should compensate for the sodium load and buffer strength of the citrate solution used and for the loss of calcium and magnesium. For this reason, the composition of the dialysis or replacement fluids and their metabolic side effects differ in the various citrate anticoagulation protocols [13,14]. Depending on the modality used, metabolic acidosis can be corrected by increasing citrate dose, effluent dose (removal of metabolic acids) or bicarbonate replacement, and metabolic alkalosis vice versa. It should be noted that when using the isotonic balanced citrate-containing pre-dilution replacement solution, anticoagulant dose and CRRT dose (effluent flow) cannot be adjusted separately [29,31]. When using a low-bicarbonate dialysis solution, acidosis is corrected by increasing citrate dose and decreasing dialysis flow [19]. To date, no studies have compared the different modalities on clinical endpoints.

**Clinical benefits of citrate**

Clinical benefits of citrate are primarily related to less bleeding, a better circuit survival and lower requirement for blood products. A recent meta-analysis, including 6 randomized controlled trials (RCTs) with a total of 488 patients and data on 658 circuits, with a focus on safety and efficacy of citrate anticoagulation, found a longer circuit survival time and a reduced risk of bleeding [32]. Control anticoagulation used in the six different studies was unfractionated heparin [10,17,20], low molecular weight heparin [9] or regional heparinization [33]. Metabolic derangements were similar to control anticoagulation and could be controlled easily. Notably, in the largest clinical trial, citrate anticoagulation was better tolerated than heparin [9]. It should also be noted that patients with an increased bleeding risk - that is, those who are likely to benefit most from citrate
anticoagulation were not included in the randomized studies. Similarly, patients with liver failure were excluded and therefore results do not apply to this population either. In contrast to the above-mentioned meta-analysis, a repeat meta-analysis including the same six RCTs concluded that the efficacy of citrate and heparin for CRRT was similar, but citrate decreased the risk of bleeding with no significant increase in the incidence of metabolic alkalosis [34]. The main reason for the discrepant results related to circuit life was the different way the authors handled the study by Betjes and colleagues [10], which did not report interquartile ranges of circuit survival. Zhang and colleagues [32] estimated the survival times by scaling the Kaplan-Meier curve. In contrast, Wu and colleagues [34] did not include this study in their circuit survival analysis.

Most of the studies on citrate anticoagulation are too small to evaluate patient outcome [13]. However, the largest (single center) trial (performed by one of the authors), including 200 critically ill patients, unexpectedly showed a 15% absolute increase in 3-month survival using an intention to treat analysis [9]. Among the higher proportion of surviving patients, there was a trend towards better renal recovery with more patients free from chronic dialysis in the citrate group ($P = 0.08$). Of note, the benefit of citrate on survival could not be fully explained by less bleeding. Although citrate did not perform worse in any post hoc subgroup, it appeared particularly beneficial in surgical patients, younger patients, patients with sepsis and in patients with more organ failure, suggesting either a protective role of citrate or harmful effects of heparin during inflammation. A subsequent multi-centre RCT did not confirm the survival benefit with citrate [35]. However, this trial was smaller, included a younger patient population with less severe organ failure, used a different citrate protocol, and had a short follow-up period and a high proportion of drop outs.

### Bioenergetic consequences

Potential sources of CRRT-derived energy consist of citrate, glucose (in ACD) and lactate. Their respective caloric equivalents are 2.48 kJ (0.59 kcal), 3.06 kJ (0.73 kcal) and 1.37 kJ (0.33 kcal) per millimole. Net energetic gain depends on the dose infused and the amount removed by CRRT. The dose of citrate infused during hemodialysis is lower compared to hemofiltration, not because of a better clearance with dialysis (sieving coefficients approach 1 for both), but because hemodialysis is feasible with lower blood flow rates [15]. Table 2 shows an estimate of the daily energy delivery when using different modalities and citrate solutions for CRRT at a dose of 2 L/h. During continuous venovenous hemodialysis (CVVHD), energy delivery is lowest with TSC as citrate source and during pre-dilution CVVH using an isotonic citrate-containing replacement fluid. ACD-A, acid citrate dextrose; Ca, citric acid; NaCl, saline; SIDe, effective strong ion difference; TSC, trisodium citrate.

### Table 1. Composition and buffer strength of the different citrate solutions

|                | Separate citrate solutions | Citrate-buffered pre-dilution replacement fluid |
|----------------|---------------------------|-----------------------------------------------|
|                | TSC | TSC + Ca | ACD-A | TSC + Ca + NaCl | TSC + NaCl | Citrate in balanced fluid |
| Na+, mmol/L    | 210-3,000 | 1,352 | 224 | 136 | 140 | 140-159 |
| H+, mmol/L (% of cations) | None | 148 | 115 (33) | 6 | None | None |
| Citrate3-, mmol/L | 68-1,000 | 450 (+50)a | 113 | 10 (+2)a | 18 or 23 | 13.3-20 |
| Glucose, mmol/L | None | None | 139 | None | None | ±5 |
| K+, mmol/L | None | None | None | None | None | 0-3 |
| Mg2+, mmol/L | None | None | None | None | None | 0-0.75 |
| Cl−, mmol/L | None | None | None | None | None | 0-0.75 |
| SIDe3, mmol/L | 3,000 | 1,352 | 224 | 30 | 54 | 33-54 |
| SIDe per mmol citrate | 3 | 2.7 | 2.0 | 3 | 3 | 2.25-3 |

*aThe 50 citrate ions are from citric acid. *The two citrate ions are from citric acid. *SIDe: effective strong ion difference as calculated after metabolism of citrate: ($Na^++K^++Ca^{2+}+Mg^{2+}$) - (Cl−). ACD-A, acid citrate dextrose; Ca, citric acid; NaCl, saline; SIDe, effective strong ion difference; TSC, trisodium citrate.
mitochondrial ATP and redox state under conditions of reperfusion after hypoxia [42] or limited substrate availability [43]. Lactate enters the mitochondria, where it is converted to pyruvate generating ATP, or enters the liver for gluconeogenesis. Third, glycolysis requires an initial investment of ATP before more ATP is produced [44]. Therefore, when energy stores are depleted, citrate and lactate may be preferable over glucose [36-38]. On the other hand, using ACD as citrate source means that 90 to 150 g of glucose is infused per day, because the solution provides 1.23 mmol glucose for each millimole of citrate. This degree of energy delivery may pose an unnecessary strain, especially when using ACD in combination with lactate-buffered replacement fluids [7]. It is important to be aware of the metabolic consequences of any type of fluid used and to adjust nutritional intake when necessary [45].

### Citrate and inflammation

**Hypocalcemia and reduction of inflammation**

Activation of neutrophils and platelets and the subsequent release of mediators from intracellular granules are regulated by cytosolic Ca^{2+}, which acts as an intracellular messenger [46-49]. Cytosolic Ca^{2+} is maintained through mobilization from intracellular stores and an influx of extracellular Ca^{2+} via plasma membrane channels. Therefore, citrate-induced changes of the extracellular calcium concentration may have a direct effect on intracellular Ca^{2+} signaling. Hypocalcemia in the filter not only down-regulates membrane-induced inflammation [50-53], but seems to reduce systemic inflammation as well [54,55]. Several pro-inflammatory actions of neutrophils are calcium-dependent. Among them are superoxide generation, adhesion, degranulation, phospholipase-A2 activation and interleukin-8 synthesis [46]. We stimulated isolated leukocytes under different extracellular calcium conditions with and without citrate, and found that inhibition of the cytosolic Ca^{2+} increase after stimulation with N-formyl-L-methionyl-L-leucyl-L-phenylalanine (fMLP) depends on the degree of extracellular hypocalcemia and not on the presence of citrate. Typical results of a single experiment are presented in Figure 1. The effects of citrate on complement activation, especially seen when using the old unsubstituted

| CRRT settings | TSC solution | ACD solution | Balanced solution |
|---------------|--------------|--------------|------------------|
| QB (ml/minute)|              |              |                  |
| Citrate target (mmol/L QB) | 4 | 4 | 4 |
| Citrate dose (mmol/h) | 36 | 24 | 36 |
| CRRT dose (ml/h) | 2,000 | 2,000 | 2,000 |
| Removal Fraction | 0.22 | 0.33 | 0.22 |
| Delivery to the patient | | | |
| Citrate (mmol/h) | 28 | 16 | 28 |
| kcal/h | 14 | 8 | 14 |
| Glucose (mmol/h) | 69 | 40 | 69 |
| kcal/h | 34 | 20 | 34 |
| Lactate (mmol/h) | 70 | 70 | 70 |
| kcal/h | 23 | 23 | 23 |
| Total energy excluding lactate (kcal/24 h) | 343 | 196 | 946 |
| (kJ/24 h) | 1,667 | 952 | 4,196 |
| Total energy including lactate (kcal/24 h) | 897 | 750 | 1,501 |
| (kJ/24 h) | 3,968 | 3,254 | 6,497 |

*Predilution dose is corrected for loss due to dilution. Caloric equivalents per millimole: citrate 2.48 kJ (0.59 kcal), glucose 3.06 kJ (0.73 kcal) and lactate 1.37 kJ (0.33 kcal). ACD, acid citrate dextrose; CRRT, continuous renal replacement therapy; CVVH, continuous venovenous hemofiltration; CVVHD, continuous venovenous hemodialysis; QB, blood flow; TSC, trisodium citrate.*

Table 2. Globally estimated total energy delivery for three common CRRT settings, postdilution CVVH and CVVHD using different citrate solutions with or without lactate replacement fluids, and predilution CVVH at a CRRT dose of 2 L/h.
cuprophone membranes, are not uniform; some studies found no effect [47,52,55], whereas others reported suppression of the complement cascade [50]. Complement activation and associated neutropenia seem to be mediated by a different membrane receptor, not related to degranulation.

Avoidance of the non-anticoagulant effects of heparins on inflammation

In addition to their anticoagulant effects, heparins have both pro- and anti-inflammatory effects elicited by their binding to numerous proteins and cells. The balance of these effects likely depends on the site of heparin binding, on heparin dose and on the presence, type and severity of sepsis [56-58] and may be beneficial [59-61], neutral [62], but totally unpredictable and potentially deleterious [58,63-67]. Anti-inflammatory effects may be due to inhibition of the formation of thrombin. In addition, heparins may attenuate neutrophil-induced endothelial damage by inhibiting leukocyte adherence to the endothelial cells as well as mast-cell activation by prevention of intracellular calcium release [56]. Pro-inflammatory effects may primarily be due to the binding and inhibition of endothelial antithrombin, thereby preventing local prostacyclin formation (summarized in [2]) and hampering the microcirculation [66,67]. Furthermore, during sepsis, heparin may contribute to the inactivation of antithrombin by elastase on vascular surfaces where neutrophils are sequestered [68]. Second, heparin can mobilize inflammatory mediators, such as myeloperoxidase, lactoferrin, elastase and platelet factor-4, not only from circulating neutrophils, platelets and cells adhered to and activated by the dialysis membrane, but also from heparan sulfate like substances on the endothelial surface [64,65]. Third, heparin binds to lipopolysaccharide-binding protein, and this process facilitates the transfer of lipopolysaccharide to the CD-14 receptor and augments endotoxin-induced activation of monocytes [58,67]. Finally, heparin avidly binds to discrete domains released from the nucleus onto the membrane of apoptotic or necrotic cells. These heparin-binding sites on apoptotic cells signal phagocytotic clearance and heparin may thus actually delay such clearance [63]. Therefore, by its binding to numerous proteins and cells, heparin induces unpredictable effects on many body functions and some of these may be harmful, especially during inflammation.

Risks and limitations of citrate

Intoxication

The main risk of citrate anticoagulation is the unintended infusion of large amounts of citrate into the patient’s circulation, which can lead to severe hypocalcemia, hypotension due to decreased myocardial contractility and vascular tone, and eventually cardiac arrest. This potentially severe adverse event can be instantaneously counteracted by calcium infusion. Unintended continued citrate infusion during the change of bags is usually not associated with clinical side effects, mainly because this amount is relatively low. However, the nursing staff should be aware of this potential risk. Fortunately, integrated software takes care of discontinuation of the citrate pump when bags are changed.

Accumulation

The main limitation of citrate anticoagulation is accumulation as a result of reduced mitochondrial citrate metabolism. Reduced metabolism is seen in patients with liver failure due to decompensated chronic liver disease and also in those with ischemic hepatitis and poor muscle perfusion as seen in prolonged cardiogenic shock. The citrate molecule itself is not toxic, but the symptoms of citrate accumulation are due to secondary hypocalcemia and acidosis. Monitoring of the patient’s Ca$^{2+}$ is therefore crucial. Ionized hypocalcemia is the most sensitive indicator of citrate accumulation [69]. While decreasing Ca$^{2+}$, citrate accumulation concomitantly increases total calcium concentration, due to an increase in citrate-bound calcium as well as calcium supplementation in response to ionized hypocalcemia. The total to ionized calcium ratio, therefore, is a useful marker to detect citrate accumulation [3,69,70] and seems the most specific [69]. A rise in total to ionized calcium ratio >2.25 should trigger the clinician to consider citrate accumulation. A recent prospective observational study in 208 critically ill
medical patients receiving CRRT with regional citrate anticoagulation for acute kidney injury found that a ratio ≥2.4 independently predicted a 33.5-fold increase in 28-day mortality [71]. Failure to metabolize citrate therefore seems to indicate a high risk of dying. Citrate accumulation due to metabolic failure can lead to metabolic acidosis. However, measurement of the anion gap is not helpful in this setting, mainly because citrate accumulation due to metabolic failure tends to occur in situations where lactic acid accumulates too, that is, in the context of advanced liver disease or tissue hypoxia. In contrast, increased citrate infusion in patients with adequate metabolism (that is, in the context of inadvertent protocol error or after polytransfusion) can lead to metabolic alkalosis.

Management of citrate accumulation includes a) decreasing both citrate flow and blood flow if feasible, b) increasing citrate clearance (effluent flow), c) reducing or discontinuing citrate flow at the cost of anticoagulant activity (when citrate is part of the pre-dilution replacement, reduction of citrate dose is not an option, because the dose of anticoagulation and CRRT are coupled, and citrate should be replaced by bicarbonate with or without heparin), d) administration of intravenous calcium to correct hypocalemia, and e) titrated replacement of bicarbonate to correct acidosis.

**Other electrolyte disturbances**

Systemic hypocalcemia during citrate anticoagulation may also be due to insufficient replacement - for example, when calcium replacement is lower than calcium loss. In that case, both total and ionized calcium decrease. The risk of calcium loss across the membrane is particularly high when calcium-free dialysate or replacement fluids are used. Although it may be rational to replace the lost amount of calcium, calcium replacement dose is generally adjusted according to actual systemic Ca²⁺ concentrations, which additionally reflect trans-cellular shifts, such as the influx of extracellular Ca²⁺ upon inflammatory stimuli [46,48]. Furthermore, the optimal ionCa²⁺ concentration during critical illness is not known. Critically ill patients often have low ionCa²⁺ concentrations [72] and supplementation of calcium might be harmful [73]. An alternative target for calcium replacement might be normalization of the PTH concentration. Citrate also binds to magnesium and this may cause hypomagnesemia due to increased magnesium loss. The use of highly concentrated trisodium citrate solutions may lead to hypernatremia if the sodium content of the replacement solutions is not proportionally reduced.

**Citrate in liver failure**

The metabolism of citrate is diminished in patients with liver failure [74,75]. Unfortunately, citrate clearance cannot be reliably predicted from standard liver function tests. However, adjustment of dose and intensified monitoring of ionized calcium levels seem to allow the safe use of citrate in patients with decompensated cirrhosis. Two recent studies showed the feasibility of citrate anticoagulation in patients with liver failure receiving treatment with a molecular absorbent recirculating system (MARS) [76,77]. In the observational study, the median citrate infusion rate was 3.1 mmol/L blood flow and median duration of treatment was 20 h [76]. Although the total to ionized calcium ratio increased significantly, treatment was well tolerated. The second study was a randomized cross-over study comparing citrate anticoagulation with no anticoagulation. Out of 27 sessions, 4 had to be terminated prematurely but all 4 were in the no anticoagulation group [77]. The use of citrate was associated with a lower ionized calcium concentration, albeit without adverse events. It should be noted that the MARS treatments are intermittent. However, in a large prospective observational study including 133 patients treated with citrate-based continuous venovenous hemodialysis for 72 hours, citrate anticoagulation was well tolerated in 86 patients with liver dysfunction as defined by a bilirubin >2 mg/dl [78]. Only 2% of patients developed an increased total to ionized calcium ratio (≥2.5).

**Citrate in shock with lactate acidosis**

Citrate metabolism occurs primarily in liver and muscle. A high lactate concentration in patients with shock at the start of CRRT should raise awareness of the risk of citrate accumulation, because this may indicate mitochondrial dysfunction. Nevertheless, clinical practice shows that a considerable proportion of patients with shock do tolerate citrate anticoagulation, especially those with septic shock and high lactate levels if circulation improves. In the randomized controlled trial by one of the authors, only one patient developed signs of citrate accumulation [9]. However, citrate is likely to accumulate in patients with persistent severe heart failure, ischemic hepatitis and poor muscle perfusion, because the Krebs cycle only operates under aerobic conditions. In these cases, intensified monitoring of the total to ionized calcium ratio and acid base balance is advised (2-hourly at start), and citrate infusion should be reduced or discontinued when calcium ratio increases above 2.25 to 2.5.

**Effects on bone metabolism**

Systemic hypocalcemia is a potent stimulus for PTH secretion. In normal subjects, a decrease in serum ionized calcium of as little as 0.025 mmol/L leads to release of preformed PTH within minutes, followed by an increased production of biologically active PTH. The
The immediate effect of PTH is to mobilize calcium from skeletal stores. In patients with normal kidney function, PTH also increases tubular calcium reabsorption and stimulates the conversion of 25-hydroxyvitamin D3 to 1,25-dihydroxyvitamin D3. In critically ill patients with acute kidney injury, the renal effects of PTH may be negligible, but concern about the potential harmful effects on bone metabolism has been expressed [79]. Three studies in critically ill patients with acute kidney injury reported a negative calcium balance and significant rise in intact PTH levels during citrate-based CRRT when aiming for systemic ionized calcium levels between 0.8 and 1.1 mmol/L [79-81]. The exact significance of PTH secretion during critical illness remains uncertain but severe bone reabsorption has been described during prolonged citrate-based CRRT. Adequate calcium replacement may be crucial for preventing bone loss.

Strategies to increase the safety of citrate

The full advantage of citrate can only be realized if its risks are well appreciated and controlled. Safe introduction of citrate starts with the choice of a well-designed and flexible protocol with proven efficacy, adjusted to the preferences for CRRT modality and dose, and availability of fluids and devices. Strict adherence to the protocol and its algorithms can prevent metabolic derangement. Safe implementation of citrate requires focused training of all staff involved, the availability of ionized calcium measurement 24 h a day and attention to detail. The risk of citrate intoxication can be avoided by using CRRT machines with integrated software that interrupt citrate infusion when the blood pump stops and also by preventing the unintended mix-up of citrate bags and crystalloid infusion bags by markedly different labeling. Intensified monitoring of ionized calcium is needed in patients with risk of accumulation. Patient safety can be improved by ‘pop-up’ alerts in the patient data management system, continuous monitoring of citrate and ionized calcium concentrations, and computerized algorithms predicting systemic and post-filter total or ionized calcium concentrations [82-84].

Evidence is accumulating that regional anticoagulation of the CRRT circuit with citrate is feasible and safe in critically ill patients. Compared to heparin, citrate anticoagulation reduces the risk of bleeding and associated blood transfusion, not only in patients with an increased risk of bleeding but also in those without. Metabolic complications depend on the type of fluids used and are largely prevented by the use of a strict protocol, training and integrated citrate software. Recent studies indicate that citrate can even be used in patients with (acute-on-chronic) liver failure when monitoring is intensified and dose is carefully adjusted. Patients at greatest risk of accumulation are those with persistent poor tissue perfusion and lactic acidosis, since citrate metabolism is oxygen dependent. The use of citrate may additionally be associated with less inflammation due to hypocalcemia at the membrane and also by avoiding the therapeutic use of heparin. Whether these beneficial effects increase patient survival needs to be confirmed. However, its other benefits are reason enough [85] to make citrate the first choice anticoagulant for CRRT provided that its safe use can be guaranteed.

Abbreviations

ACD, acid-citrate-dextrose; CRRT, continuous renal replacement therapy; CVVH, continuous venovenous hemofiltration; CVVHD, continuous venovenous hemodialysis; MARS, molecular absorbent recirculating system; PTH, parathyroid hormone; RCT, randomized controlled trial; SID, strong ion difference; TSC, trisodium citrate.

Competing interests

None of the authors has a financial or non-financial competing interest.

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