PD-L1 Expression Is Associated With VEGFA and LADC Patients’ Survival

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Objectives: To elucidate the relationship between VEGFA and PD-L1 expression in lung adenocarcinoma (LADC).

Methods: PD-L1 and VEGFA expression were determined by immunohistochemistry with H-score on formalin-fixed paraffin-embedded resected LADC specimens of 129 cases.

Results: High PD-L1 expression in 53 (41.1%) patients, high VEGFA expression in 65 (50.4%), and co-expression in 18 (14.0%) were observed. Inverse correlation between expression of PD-L1 and VEGFA was found ($P = 0.002, r = -0.274$). VEGFA and PD-L1 expression were not significantly associated with the clinicopathological features. High PD-L1 expression was significantly associated with all patients’ poor progression-free survival and overall survival in a univariate analysis, but there was no significantly association with high VEGFA expression and prognosis. Co-expression of PD-L1 and VEGFA exhibited a worst overall survival compared to negative groups ($P = 0.005$).

Conclusions: These findings indicate that high PD-L1 expression could impact both poor overall survival and progression-free survival in patients with resected LADC. Co-expression of PD-L1 and VEGFA may be considered as an important prognostic factor for patients with resected lung adenocarcinoma.

Keywords: PD-L1, VEGFA, overall survival, progression-free survival, lung adenocarcinoma

INTRODUCTION

Lung cancer is the most common and deadliest cancer worldwide (1). Non-small-cell lung cancer (NSCLC) accounts for approximately 80% of all patients, in which lung adenocarcinoma (LADC) is the most common. Although targeted drugs for driver gene mutations and immunotherapy in clinic have obtained a clinical benefit, prognosis for patients with LADC still remains unsatisfied (2). To improve the efficacy, it is of great importance to find ideal biomarkers to predict the progressive disease and overall survival of LADC patients.

In the last decade, inhibitors of PD-1 and its ligand PD-L1 were introduced into clinic. As a cell surface protein, PD-L1 is wildly expressed in variety of cells (such as immune cells, endothelial cells and tumor cells) and suppress immune-response lead by lymphocytes (3). Increased PD-L1 enhances tumor immune evasion and has been reported to be associated with a poor survival in various malignancies including lung cancer (4). With the development of anti-PD-1/PD-L1 inhibitors (such as nivolumab, pembrolizumab, ipilimumab), some clinical studies (5–8) have demonstrated excellent efficacy in patients with LADC, even in some with EGFR or ALK tyrosine kinase inhibitors (TKIs) resistance. Furthermore, it was found that high PD-L1 expression was closely correlated to a higher likelihood of objective response (OR) of anti-PD-1/PD-L1 in many studies (9, 10).
Like PD-L1, vascular endothelial growth factor (VEGF) is known as an immunosuppressive factor and plays an important role in tumor immune through promoting a tumor immunosuppressive microenvironment (11, 12). VEGFA, an important angiogenic factor in VEGF family, can exert an important negative impact on immune cells' development and function (8). Some studies indicated that PD-L1 expression was significantly correlated with VEGF in clear cell renal carcinoma (13) and classical Hodgkin lymphoma (11). Additionally, overexpression of VEGFA has been confirmed in a majority of patients with NSCLC, some studies reported that there was a positive correlation between tumor vascularization and poor disease-free survival (DFS) or overall survival (OS) (14, 15).

Few studies reported that the connection between PD-L1 and VEGFA expression in patients with resected LADC. In the present study, we retrospectively reviewed the data of patients with resected tissue of LADC and investigated the relationship between PD-L1 and VEGFA expression to further explore their potential efficacy-predictive value in anti-angiogenic therapy with immunotherapy and their prognostic significance.

**MATERIALS AND METHODS**

**Patients and Samples**

We retrospectively reviewed 129 patients with LADC (56 with wild type, 56 with EGFR mutations, 13 with KRAS mutations, 4 with ALK mutations) surgically treated in Tianjin Medical University Cancer Institute and Hospital between December 2011 and September 2016. The postoperative follow-up period ranged from 21 to 79 months (median: 47 months). The patients' inclusion criteria included: (1) complete following-up date; (2) without neoadjuvant therapy before surgery; Regarding to preoperative staging assessment, all patients underwent positron emission tomography-computed tomography (PET-CT) or enhanced chest CT, contrast-enhanced computed tomography (ECT) and brain magnetic resonance imaging (MRI) if they were necessary. Twelve patients with stage IVA refer to incidental pleural metastases detected at surgery and 20 patients with stage III were found incidental N2 nodes during surgery. The clinical data included age at diagnosis, sex, smoking history, gene mutation status, histological subtypes, clinical stage, and postoperative chemotherapy; all parameters were retrieved based on electronic medical records. Progression-free survival (PFS) was defined as the interval between the date of resection and the date of disease progression or death. The follow-up for patients' survival with documented disease progression were censored on the last follow-up date. Overall survival (OS) was defined as the interval between the date of resection and either death due to any cause or the last follow-up. All clinical information was obtained from medical records. The study was approved by the Tianjin Medical University Cancer Institute and Hospital’s Ethics Committee and informed consent was obtained from all patients in verbal form.

**Immunohistochemical Staining**

Four micrometer thickness of tumor sections were deparaffinized and rehydrated. Antigen retrieval was performed at 130°C for 2 min, using EDTA solution (PH 11.0) for PD-L1, or citrate buffer (PH 6.0) for VEGFA. After blocking with 3% hydrogen peroxide and normal goat serum, the sections were incubated with primary antibody against PD-L1/CD274 antibody (66248-1-lg, Proteintech, USA) at dilution 1:1,200 or anti-VEGFA antibody (ab1316, Abcam, USA) at dilution 1:300 at 4°C overnight.
followed by EIVISON plus (kit-9903, MXB, China), DAB kit (ZL1-9019, ZSGB-BIO, China) per the manufacturers protocol were used for coloration. Finally, the tumor sections were stained with hematoxylin and evaluated under a light-field microscope.

**Image Analysis**

To measure the PD-L1 and VEGFA expression by IHC, stained sections were digitally analyzed at x400 resolution using an Olympus BX-UCB. PD-L1 and VEGFA expression were scored by H-score system. An H-score (range, 0–300) was calculated as the sum of the product of the highest intensity of staining (0, negative; 1, weak positive; 2, moderate positive; 3, strong positive) and percentage of tumor cells positive (0–100%; with any intensity of positive staining) (12). Two clinical pathologists assessed the intensity of the immunostaining on each section independently and in a blinded manner. According to previous

| TABLE 2 | Relationship between VEGFA/PD-L1 expression and the clinical characteristics in 129 patients with LADC. |
|----------|-------------------------------------------------------------------------------------------------|
| Characteristics | PD-L1 negative, n (%) | PD-L1 positive, n (%) | P-value | VEGFA negative, n (%) | VEGFA positive, n (%) | P-value |
| Gender | | | | | | | |
| Male | 37 (48.7) | 24 (45.3) | 27 (42.2) | 34 (52.3) | 0.703 | 0.250 |
| Female | 39 (51.3) | 29 (54.7) | 37 (57.8) | 31 (47.7) | | |
| Age | | | | | | | |
| <65 | 61 (80.3) | 43 (81.1) | 53 (82.8) | 51 (78.5) | 0.902 | 0.532 |
| ≥65 | 15 (19.7) | 10 (18.9) | 11 (17.2) | 14 (21.5) | | |
| Smoking history | | | | | | | |
| Yes | 36 (47.4) | 21 (60.4) | 27 (41.3) | 30 (46.2) | 0.383 | 0.690 |
| No | 40 (52.6) | 32 (39.6) | 37 (58.7) | 35 (53.8) | | |
| Stage | | | | | | | |
| I | 45 (59.2) | 28 (52.8) | 35 (54.7) | 38 (58.5) | 0.867 | 0.733 |
| II | 11 (14.5) | 8 (15.2) | 11 (17.2) | 8 (12.3) | | |
| III | 13 (17.1) | 12 (22.6) | 11 (17.2) | 14 (21.5) | | |
| IV | 7 (9.2) | 5 (9.4) | 7 (10.9) | 5 (7.7) | | |
| Histological types | | | | | | | |
| Lepidic predominate | 29 (38.1) | 18 (34.0) | 25 (39.1) | 22 (33.8) | 0.529 | 0.350 |
| Acinar predominate | 26 (34.2) | 15 (28.3) | 15 (23.4) | 26 (40.0) | | |
| Papillary/micropapillary predominate | 10 (13.2) | 13 (24.5) | 13 (20.3) | 10 (15.4) | | |
| Solid predominate | 5 (6.6) | 2 (3.8) | 4 (6.3) | 3 (4.6) | | |
| Other | 6 (7.9) | 5 (9.4) | 7 (10.9) | 4 (6.2) | | |
| T | | | | | | | |
| T1 | 42 (55.3) | 30 (56.6) | 40 (62.5) | 32 (49.2) | 0.051 | 0.281 |
| T2 | 28 (36.8) | 12 (22.7) | 16 (25.0) | 24 (37.0) | | |
| >T2 | 6 (7.9) | 5 (9.4) | 7 (10.9) | 4 (6.2) | | |
| N | | | | | | | |
| N0 | 50 (65.8) | 36 (67.9) | 43 (67.2) | 43 (66.2) | 0.721 | 0.700 |
| N1 | 6 (7.9) | 2 (3.8) | 5 (7.8) | 3 (4.6) | | |
| N2 | 20 (26.3) | 15 (28.3) | 16 (25.0) | 19 (29.2) | | |
| M | | | | | | | |
| M0 | 68 (89.5) | 49 (92.5) | 58 (90.6) | 59 (90.8) | 0.760 | 1.000 |
| M1 | 8 (10.5) | 4 (7.5) | 6 (9.4) | 6 (9.2) | | |
| Gene status | | | | | | | |
| WT | 33 (43.4) | 23 (43.4) | 28 (43.8) | 28 (43.1) | 0.081 | 0.417 |
| EGFR | 29 (38.2) | 27 (50.9) | 30 (48.9) | 26 (40.0) | | |
| Other mutations | 14 (18.4) | 3 (5.7) | 6 (9.3) | 11 (16.9) | | |
| Postoperative therapy | | | | | | | |
| Chemotherapy/chemoradiotherapy | 28 (36.8) | 18 (34.0) | 22 (34.4) | 24 (36.9) | 0.893 | 0.472 |
| Chemotherapy+TKI/TKI | 3 (3.9) | 4 (7.5) | 2 (3.1) | 5 (7.7) | | |
| Non-treatment | 45 (59.3) | 31 (58.5) | 40 (62.5) | 36 (55.4) | | |
| VEGFA | | | | | | | |
| H-score ≤50 | 29 (38.2) | 35 (66.0) | | | | |
| H-score >50 | 47 (61.8) | 18 (34.0) | | | | |
literatures (16), we set the cut-off value at 100, i.e., H-score >100 as a PD-L1 overexpression group (generally 2+ intensity in >50% of tumor cells); when considering the VEGFA expression (17), we set the cut-off value at 50, i.e., H-score >50 as a VEGFA overexpression group (generally 2+ intensity in >25% of tumor cells) (Figure 1).

**Statistical Analysis**

Statistical analyses were performed using SPSS v.21 (IBM Corp, Armonk, NY, USA) and survival curve was performed with GraphPad Prism 6 (USA, GraphPad Software). The relationships between PD-L1/VEGFA expression and clinical features were compared using Fisher’s exact test; the linear relationship between PD-L1 and VEGFA expression was determined using Pearson’s test. Patients’ survival was estimated by the Kaplan-Meier method and compared using the log-rank test. Multivariate and univariate analysis were performed using the cox proportional hazards regression model to calculate the hazard ratio (HR) and 95% CI. Two-tailed \( P < 0.05 \) was considered statistically significant.

**RESULTS**

**Correlation of PD-L1 and VEGFA Expression With Clinic-Pathological Features**

Baseline clinic-pathological characteristics of 129 LADC cases were summarized in Table 1. 61 (52.7%) patients were male; 25 (19.4%) patients were above age of 65; 57 (44.2%) patients were smokers; 92 (71.3%) patients were diagnosed as stage I/II; 41 (31.8%) cancer were characterized as acinar adenocarcinoma; high PD-L1 expression in 53 (41.1%) patients; high VEGFA expression in 65 (50.4%); 56 (43.4%) patients had EGFR mutations and 17 (12.2%) patients had other mutations including KRAS mutations and ALK mutations; 29 (38.2%) cases were in VEGFA- & PD-L1- group, 47 (61.8%) were in VEGFA+ & PD-L1- group, 35 (66.0%) were in VEGFA- & PD-L1+ group and 18 (34.0%) were in VEGFA+ & PD-L1+ group. No significant correlation of the PD-L1 or VEGFA expression and gender, clinical stage, age, histological subtypes, TNM, gene mutations, or smoking status were found (all \( P > 0.05 \)) (Table 2).

However, we found that expression of VEGFA was negatively correlated with expression of PD-L1 \( (P = 0.002, r = -0.274) \) (Supplement Figure 1).

**Correlation of PD-L1 and VEGFA Expression With Patients’ Survival**

PD-L1 and VEGFA expression were measured in 129 patients wherein 53 patients (41.1%) had high expression PD-L1 (H-score >100) and 65 patients (50.4%) had high expression VEGFA (H-score >50). In all patients, PD-L1\text{high} group showed a significant negative impact on the OS \( (P = 0.024) \) and PFS \( (P = 0.018) \) (Figure 2); however, high VEGFA expression was no significantly correlated with unfavorable OS \( (P = 0.190) \) and PFS \( (P = 0.325) \) in all patients (Supplement Figure 2).

Therefore, we further investigated the association between the expression of PD-L1 and survival in different subgroups. Firstly, we found high PD-L1 expression was not significantly correlated with adverse survival in patients of clinical stage I/II, in contrast, significantly correlated with poor OS \( (P < 0.010) \) and PFS \( (P < 0.001) \) in stage III/IV. Secondly, same result was

![Images of immunohistochemical staining for PD-L1 and VEGFA expression in LADC. The staining strength of PD-L1 expression: (A) negative; (B) weak positive; (C) moderated positive; (D) strong positive; (E) VEGFA negative expression; (F) VEGFA positive expression; scale bar: 100 µm.](image-url)
found in patients with acinar adenocarcinoma, i.e., high PD-L1 expression was significantly correlated to an adverse PFS ($P < 0.001$), but not to poor OS ($P = 0.129$) or PFS ($P = 0.291$) in patients with non-acinar adenocarcinoma (Figure 3). Thirdly, we found high PD-L1 expression was no significantly correlated with OS ($P = 0.698$) and PFS ($P = 0.142$) in all
gene mutations patients (EGFR, ALK, or KRAS). Because it was insufficient to analyze the relationship between PD-L1 expression and prognosis in only 17 (12.2%) patients with KRAS or ALK mutations, correlation between PD-L1 expression and survival in KRAS or ALK mutations group was not analyzed. However, we found that PD-L1 high expression has no significant impact on OS ($P = 0.721$) and PFS ($P = 0.653$) in patients with EGFR mutations; in wild type (WT) patients, PD-L1$^{high}$ expression caused a poor OS ($P < 0.010$) and PFS ($P = 0.023$) compared to PD-L1$^{low}$ cases (Figure 2).

Finally, we found that high PD-L1 expression was as a poor prognostic role for PFS ($P = 0.036$) and OS ($P = 0.004$) in VEGFA$^+$ group, but not a prognostic factor for OS ($P = 0.165$) and PFS ($P = 0.078$) in VEGFA$^-$ group. Concordantly, we found that VEGFA$^+$&PD-L1$^+$ group had the worst OS ($P = 0.005$) and PFS ($P = 0.034$) compared to other groups.
FIGURE 4 | Kaplan–Meier curves showing OS (A,C) and PFS (B,D) of different groups (VEGFA+ or VEGFA-) with high and low of PD-L1 expression, showing OS (E) and PFS (F) in patients with PD-L1–&VEGFA–, PD-L1+&EVGFA–, PD-L1–&VEGFA+ and PD-L1+&VEGFA+ expression.

In a univariate analysis on the all patients, six clinical characteristics were determined as unfavorable prognostic factors for PFS: clinical stage III/IV [HR = 6.552 (95% CI 3.852–11.145), \( P < 0.0001 \)], high PD-L1 expression [HR = 3.143 (95% CI 1.106–8.64), \( P = 0.019 \)], advanced TNM (>T2; >N0; M1; all \( P < 0.001 \)) and co-expression of PD-L1 and VEGFA [HR = 2.907 (95% CI 1.082–4.067), \( P = 0.028 \)]. Those factors were also determined as unfavorable prognostic factors for OS. In a multivariate analysis, co-expression of VEGFA and PD-L1 was a poor prognostic factor for OS [HR = 3.230 (95% CI 1.388–7.518), \( P = 0.007 \)] and high PD-L1 expression was an unfavorable prognostic factor for PFS [HR = 2.163 (95% CI 1.124–4.163, \( P = 0.021 \)], but only clinical stage III/IV was adverse factor for both OS [HR = 2.657 (95% CI 1.085–6.504, \( P = 0.032 \)] and PFS [HR = 4.064 (95% CI 1.815–9.097, \( P = 0.001 \)] (Table 3).

DISCUSSION

Recently, some clinical trials (6, 9) of PD-1/PD-L1 inhibitors have demonstrated durable clinical benefit in many patients. Several studies reported that the expression of PD-L1 is closely related to EGFR (18) mutations, KRAS (19) mutations, smoking history...
TABLE 3 | Univariate and multivariate cox analysis of factors for progression-free survival and overall survival in patients with LADC.

| Variable                                | Overall survival |       | Progression-free survival |       |
|-----------------------------------------|------------------|-------|---------------------------|-------|
|                                         | HR               | 95% CI| P-value                   | HR    | 95% CI| P-value |
| **UNIVARIATE COX ANALYSIS**             |                  |       |                           |       |       |
| Gender (female vs. male)                | 1.451            | 0.819–2.571 | 0.202 | 1.199 | 0.716–2.007 | 0.490 |
| Age (≤65 vs. >65)                       | 1.461            | 0.744–2.870 | 0.271 | 1.065 | 0.634–1.790 | 0.812 |
| Smoking history (no vs. yes)            | 1.286            | 0.729–2.267 | 0.385 | 1.046 | 0.543–2.018 | 0.892 |
| T factor (T ≤2 vs. T >2)                | 3.735            | 1.997–6.985 | <0.0001 | 3.778 | 2.073–6.855 | <0.001 |
| N factor (N0 vs. >N0)                   | 2.798            | 1.580–4.955 | <0.0001 | 3.674 | 2.178–6.194 | <0.001 |
| M factor (M0 vs. M1)                    | 3.254            | 1.568–6.750 | 0.002 | 3.710 | 1.863–7.391 | <0.001 |
| Gene mutations (WT vs. EGFR)            | 1.099            | 0.662–1.942 | 0.745 | 1.277 | 0.761–2.141 | 0.354 |
| Histological subtypes (acinar vs. non-acinar) | 0.599            | 0.337–1.063 | 0.113 | 0.706 | 0.544–0.916 | 0.089 |
| Stage (II/III vs. IV)                   | 4.801            | 2.698–8.542 | <0.0001 | 6.552 | 3.852–11.145 | <0.0001 |
| Postoperative therapy (else vs. adjuvant chemotherapy) | 0.917            | 0.492–1.709 | 0.785 | 1.036 | 0.587–1.828 | 0.903 |
| VEGFA expression (=50 vs. >50)          | 1.470            | 0.823–2.624 | 0.193 | 1.297 | 0.711–2.183 | 0.327 |
| PD-L1 expression (=100 vs. >100)        | 1.924            | 1.080–3.427 | 0.026 | 3.143 | 1.106–8.164 | 0.019 |
| Co-expression (else vs. PD-L1+/VEGFA+)   | 3.114            | 1.564–6.201 | 0.001 | 2.907 | 1.082–4.067 | 0.028 |
| **MULTIVARIATE COX ANALYSIS**           |                  |       |                           |       |       |
| T factor                                | 2.313            | 1.130–4.733 | 0.022 | 1.389 | 0.694–2.778 | 0.353 |
| N factor                                | 1.546            | 0.720–3.316 | 0.263 | 1.806 | 0.919–3.550 | 0.086 |
| M factor                                | 1.730            | 0.756–3.956 | 0.194 | 1.842 | 0.738–3.655 | 0.224 |
| Stage                                   | 2.657            | 1.065–6.504 | 0.032 | 4.064 | 1.815–9.097 | 0.001 |
| PD-L1 expression                        | 1.494            | 0.735–3.036 | 0.267 | 2.163 | 1.124–4.163 | 0.021 |
| Co-expression                           | 3.230            | 1.388–7.518 | 0.007 | 1.960 | 0.621–2.980 | 0.443 |

Statistically significant p-values (p < 0.05) are shown in bold.

(19) and advanced tumor stage (18, 20), but we failed to define a significant correlation between PD-L1 expression and these clinical features in LADC, which is concordant with the results of other studies (20, 21). In addition, some studies suggested that VEGFA expression has a significant association with EGFR mutations, advanced clinical stage and lymph node metastasis (21). Similar to PD-L1 expression, in our patient cohort we didn’t find a significant relationship between VEGFA expression and any clinical variables. Possibly due to the variety of cut-off value of PD-L1 and VEGFA positive expression in cancer, results of our study are different with that from previous some studies (positive PD-L1 ever defined as 1, 5, 10, or even 50%) (22, 23).

In the present study, we analyzed the correlation of PD-L1 and VEGFA expression and patients’ prognosis. There was no significant correlation between VEGFA expression and survival found, as similar as the results in other studies (24, 25). But when considering PD-L1, we found a significant association between PD-L1 expression and prognosis. When analyzed in different subgroups, there was no significant association between PD-L1 expression and OS or PFS in patients with EGFR mutations; however, high expression PD-L1 has a significant correlation to poor OS and PFS in patients with wild type. These findings suggested that PD-L1 expression may be not a prognostic factor for survival in patients with EGFR mutations. A meta-analysis of three clinical trials (CheckMate057, POPLAR, and KEYNOTE-010) also found that patients with EGFR mutations did not represent a survival benefit from anti-PD-1/PD-L1 therapy compared with other treatment (26). Additionally, in patients with acinar adenocarcinoma and clinical stage III/IV, high expression of PD-L1 was associated with unfavorable OS and PFS, and other studies (27, 28) have found similar evidences.

The interesting finding from our study is that PD-L1 expression inversely related to the expression of VEGFA. About this issue, the traditional view is that VEGFA high expression will lead to vascular abnormalities, further aggravation of hypoxia and the activation of HIF-1a pathway in tumor tissues, which could result in increased expression of PD-L1 on tumor cells. According to this viewpoint there seems to be a close positive association between PD-L1 and VEGF expression in tumor tissues. Several studies also found that VEGF expression is positive associated with PD-L1 expression in clear cell renal cell carcinoma (13) and classical Hodgkin lymphoma (11). Nevertheless, other studies found that PD-L1 protein expression measured by IHC is inversely correlation with the mRNA expression of VEGFA in patients with clear cell renal cell carcinoma (29) and not associated with VEGF gene expression and patients’ survival in angiosarcoma (30). In the previous study, no explanation of potential reason of such contradictory events and any report to observe the association among them in LADC were made. Here we suppose that a relatively normal vasculature is formed in tumor tissue with low expression of VEGFA (23, 31), which may lead to more immune cells infiltrating into tumor tissues, causing increased responding PD-L1 expression on tumor cells, therefore showing...
an opposite relationship between PD-L1 and VEGFA expression. Accordingly, some studies found that after blocking VEGFA, the expression of PD-L1 in tumor tissues will be upregulated in the preclinical model (32). Similarly, upregulation of PD-L1 has been found on both endothelial cells and tumor cells following treatment with anti-VEGFR2 therapy (33), which confirms the necessity of combination with anti-angiogenic and immunotherapy. It also explains from profile that a relationship between VEGFA and PD-L1 expression was not positively correlated. Of course, this is only an assumption and the underlying mechanisms need further to study.

Finally, we found that only 14.0% (18/129) patients revealed VEGFA+&PD-L1+, far fewer than the other types, but these patients had the worst prognosis compared to other groups. As far as we know, no study has reported this phenomenon in LADC, so we proposed another hypothesis: some LADC tumor cells in patients with co-expression of VEGFA and PD-L1 may have strong ability to evade from the attacks by immune cells, so even if few immune cells infiltrate into tumor tissue, through abnormal vasculature caused by high-expressed VEGFA, they can also cause "high-response" tumor cells to express high PD-L1. This explanation indicated that co-expression of PD-L1 and VEGFA may be as a predictor for high recurrent risk and poor prognosis. For these patients, the combination of anti-VEGF and anti-PDL1 could be an interesting treatment strategy. It also provides a theoretical possibility for screening optimal population to combination of anti-VEGFA and anti-PD-1/PD-L1 therapy. We also analyzed the relationship between co-expression of PD-L1 and VEGFA and clinical features, but failed to found a significant correlation between them and clinical features maybe due to only 18 patients with co-expression of PD-L1 and VEGFA.

Our study still has several limitations: 1. A similar limitation with other studies is the lack of standardized cut-off value of PD-L1 and VEGFA expression; 2. The amount of patient samples collected retrospectively were relatively small; 3. It should be emphasized that it is the initial and immature study to explore the correlation of VEGFA and PD-L1 expression in LADC; we demonstrated the correlation between them, but the underlying mechanisms is still unclear.

CONCLUSION

High expression PD-L1 is a poor factor on PFS and OS in patients with WT, clinical stage III/IV or acinar adenocarcinoma, but has no significant impact on patients with early stage and EGFR mutations. Expression of VEGFA is negatively correlated with the expression of PD-L1, but patients with co-expression of PD-L1 and VEGF will lead to significantly poor prognosis than negative ones. Our study also provides the theoretical possibility to screen optimal population of combination with anti-PD-L1/PD-1 and anti-VEGF therapy in LADC.

AUTHOR CONTRIBUTIONS

SL: design of study and manuscript preparation; YJ: data statistics; TQ and KL: critical revision of the manuscript.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fonc.2019.00189/full#supplementary-material

REFERENCES

1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. (2018) 68:394–424. doi: 10.3322/caac.21492
2. Ettinger DS, Wood DE, Aisner DL, Akerley W, Bauman J, Chirieac LR, et al. Non-small cell lung cancer, version 5.2017. NCCN clinical practice guidelines in oncology. J Natl Compr Canc Netw. (2017) 15:504–35. doi: 10.6004/jnccn.2017.0050
3. Blank C, Gajewski TF, Mackensen A. Interaction of PD-L1 on tumor cells with PD-1 on tumor-specific T cells as a mechanism of immune evasion: implications for tumor immunotherapy. Cancer Immunol Immunother. (2003) 54:307–14. doi: 10.1007/s00262-004-0593-x
4. Patel SP, Kurzrock R. PD-L1 expression as a predictive biomarker in cancer immunotherapy. Mol Cancer Ther. (2015) 14:847–56. doi: 10.1158/1535-7163.MCT-14-0983
5. Qiao M, Jiang T, Ren S, Zhou C. Combination strategies on the basis of immune checkpoint inhibitors in non-small-cell lung cancer: where do we stand? Clin Lung Cancer. (2018) 19:1–11. doi: 10.1016/j.cllc.2017.06.005
6. Herbst RS, Baas P, Kim DW, Felip E, Pérez-Gracia JL, Han JY, et al. Pembrolizumab versus docetaxel for previously treated, PD-L1-positive, advanced non-small-cell lung cancer (KEYNOTE-010): a randomised controlled trial. Lancet. (2016) 387:1540–50. doi: 10.1016/s0140-6736(15)02181-7
7. Reed M, Rodriguez-Abreu D, Robisson AG, Hui R, Cossii T, Fulop A, et al. Pembrolizumab versus chemotherapy for PD-L1-positive non-small-cell lung cancer. N Engl J Med. (2016) 375:1823–33. doi: 10.1056/NEJMoa1606774
8. Huang Y, Goel S, Duda DG, Fukumura D, Jain RK. Vascular normalization as an emerging strategy to enhance cancer immunotherapy. Cancer Res. (2013) 73:2943–8. doi: 10.1158/0008-5472.CAN-12-4354
9. Herbst RS, Soria JC, Kowanetz M, Fine GD, Hamid O, Gordon MS, et al. Predictive correlates of response to the anti-PD-L1 antibody MPDL3280A in cancer patients. Nature. (2014) 515:563–7. doi: 10.1038/nature14011
1. Tumeh PC, Harvie CL, Yearley JH, Shintaku IP, Taylor EJ, Robert L, et al. PD-1 blockade induces responses by inhibiting adaptive immune resistance. *Nature*. (2014) 515:568–71. doi: 10.1038/nature13954

2. Koh YW, Han JH, Yoon DH, Suh C, Huh J. PD-L1 expression correlates with VEGF and microvessel density in patients with uniformly treated classical Hodgkin lymphoma. *Ann Hematol*. (2017) 96:1883–90. doi: 10.1007/s00277-017-3115-6

3. Howitt BE, Sun HH, Roemer MG, Kelley A, Chapuy B, Aviki E, et al. Genetic basis for PD-L1 expression in squamous cell carcinomas of the cervix and vulva. *JAMA Oncol*. (2016) 2:518–22. doi: 10.1001/jamaoncol.2015.6326

4. Shin SJ, Jeon YK, Kim PJ, Cho YM, Koh J, Chung DH, et al. Clinico-pathologic analysis of PD-L1 and PD-L2 expression in renal cell carcinoma: association with oncogenic proteins status. *Ann Surg Oncol*. (2016) 23:694–702. doi: 10.1245/s10434-015-4903-7

5. Zhu CQ, Shih W, Ling CH, Tsao MS. Immunohistochemical markers of prognosis in non-small cell lung cancer: a review and proposal for a multiphasic approach to marker evaluation. *J Clin Pathol*. (2006) 59:790–800. doi: 10.1136/jcp.2005.031351

6. Salgia R. Prognostic significance of angiogenesis and angiogenic growth factors in nonsmall cell lung cancer. *Cancer*. (2011) 117:3889–99. doi: 10.1002/cncr.25935

7. Yang CY, Lin MW, Chang YL, Wu CT, Yang PC. Programmed cell death-ligand 1 expression is associated with a favourable immune microenvironment and better overall survival in stage I pulmonary squamous cell carcinoma. *Eur J Cancer*. (2016) 57:91–103. doi: 10.1016/j.ejca.2015.12.033

8. Liontos M, Trigka EA, Korkolopoulou P, Tzannis K, Lainakis G, Koutsoukos K, et al. Expression and prognostic significance of VEGF and mTOR pathway proteins in metastatic renal cell carcinoma patients: a prognostic immunohistochemical profile for kidney cancer patients. *World J Urol*. (2017) 35:411–9. doi: 10.1007/s00345-016-1890-7

9. Yang CY, Lin MW, Chang YL, Wu CT, Yang PC. Programmed cell death-ligand 1 expression is associated with a favourable immune microenvironment and better overall survival in stage I pulmonary squamous cell carcinoma. *Eur J Cancer*. (2016) 57:91–103. doi: 10.1016/j.ejca.2015.12.033

10. Calles A, Liao X, Sholl LM, Rodig SJ, Freeman GJ, Butaney M, et al. Combined antiangiogenic and anti-PD-L1 therapy stimulates tumor immunity through HEV formation. *Sci Transl Med*. (2017) 9:eaak9679. doi: 10.1126/scitranslmed.aak9679

11. Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest. Copyright © 2019 Liu, Qin, Jia and Li. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.