Polysaccharides obtained from mycelia of *Cordyceps militaris* attenuated doxorubicin-induced cytotoxic effects in chemotherapy

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Abstract

Objectives: *Fungus Cordyceps militaris* has been used as a herbal tonic in traditional Chinese medicine, which could be surface liquid-cultured for *mycelia* production. To evaluate the potential of polysaccharides obtained from *mycelia of Cordyceps militaris* (PS-MCM) for attenuation of side-effects of chemotherapy.

Methods: Doxorubicin was used to induce cytotoxicity in THP-1 monocytes and EL-4 T cells, and the effects of PS-MCM on cell viability and cytokine production were detected on doxorubicin-treated THP-1 and EL-4 cells.

Results: PS-MCM reduced doxorubicin-induced cell death and promoted cell proliferation in THP-1 and EL-4 cells. Moreover, PS-MCM elevated the level of cytokines associated with immune-modulation of THP-1 and EL-4 cells.

Conclusion: These findings indicate that PS-MCM has potential for development as a functional food to counteract side effects of chemotherapy.

Keywords: *Cordyceps militaris*, traditional Chinese medicine, doxorubicin, chemotherapy, immunotherapy.

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Introduction

Combined immunotherapy and chemotherapy is one of the most efficient strategies for treating cancer. It is important to design an effective treatment program which can optimize immunotherapy and chemotherapy to diminish side effects. Clinical evidence shows that eliminating cancer via the immune system is promising and will propel more research in immunotherapy. Therefore, many studies have been done to use the most appropriate
strategies to meet different performance objectives. The use of complementary and alternative medicine has become a popular option to support the conventional therapy in many areas. For example, many herbal formulas and remedies based on the traditional Chinese medicine are well accepted among cancer patients with Chinese background.

The fungus Cordyceps has been used as a herbal tonic in traditional Chinese medicine for over 300 years. Numerous studies have recently reported the anticancer activity of *C. sinensis*, against B16 melanoma, leukemia, thyroid carcinoma, hepatocellular carcinoma, and renal cancer cells. *Cordycepin* is a crucial bioactive compound derived from *C. militaris*. The inhibitory effect of cordycepin on cancer was diminished by adenosine deaminase catalysis in vivo. We have demonstrated the anti-oral cancer activity of water extract isolated from the mycelia of surface liquid-cultured *C. militaris*.14 In addition, polysaccharides were isolated from the mycelium of submerged-fermented *C. militaris* by liquid static culture, which have been shown to stimulate immune activity via promotion of lymphatic spleen cell proliferation and cytokines production.16

The chemotherapeutic drug doxorubicin has been found to cause cell toxicity of normal immunocytes during chemotherapy, leading to loss of immune activity and other side-effects. Moreover, a healthy immune system was deemed irrelevant for treating cancer in the context of chemotherapy. The immune cells interact with the cancer cells and the immune system is fully capable of killing tumor cells. However, immune cells have trouble to recognizing them due to tumor-induced immune suppression. Evidence from a recent study has shown that immune-stimulatory activity mediated by herbs could attenuate side-effects caused by chemotherapy. Therefore, the aim of this study was to evaluate the potential of polysaccharides obtained from the mycelia of surface liquid-cultured *C. militaris* (PS-MCM) on immune-stimulatory activity against side-effects of chemotherapy.

**Materials and methods**

**Sample isolation**

*C. militaris* (BCRC34380) was obtained from the Bioresource Collection and Research Center (Hsinchu, Taiwan). The liquid seed culture was inoculated with a mycelium mat from a stock slant and cultured in a 500mL conical flask containing 200 mL of the medium (10 g/L peptone, 30 g/L glucose, 0.5 g/LMgSO4, 0.5 g/L K2HPO4, and 0.5 g/L KH2PO4) at 25°C on a rotary shaker at 150 rpm for 7 days for maintenance. The cultured *mycelia* were collected and freeze dried, and the *mycelium* powder was dissolved in water and heated for 60 minutes at 60°C to extract intracellular polysaccharides by alcohol precipitation.

**Cell culture**

THP-1, a human monocytic leukemia cell line, was obtained from the Bioresource Collection and Research Center (BCRC 60430; Food Industry Research and Development Institute, Hsinchu, Taiwan). Cells were maintained in RPMI-1640 medium supplemented with 10% FBS, streptomycin (100 mg/mL), penicillin (100 U/mL), 50 μM β-mercaptoethanol, and 5.5 mM D-glucose in a 5% CO2 incubator at 37°C. EL-4 murine T-lymphoma cells were purchased from the Bioresource Collection and Research Center (Hsinchu, Taiwan). EL-4 cells were cultured in RPMI-1640 medium supplemented with 10% heat-inactivated FBS. Confluent cells were sub-cultured at a ratio of 1:3 in a 10 cm dish, and media were changed twice a week. Cells were cultured at 37°C in a humidified atmosphere of 5% CO2.

**Cell viability**

THP-1 and EL-4 cells were treated by doxorubicin with various polysaccharides obtained from the *mycelia* of surface liquid-cultured *C. militaris* (PS-MCM) for 24 hours. Cell viability was measured using the MTT reduction assay. Incubation was terminated by media aspiration and addition of 5 mg/mL of MTT working solution to each well. Formazane formation was terminated after 3 hours by removal of the MTT solution. Subsequently, appropriate amounts of DMSO were added to each well to solubilize the formazane. The formazane-containing samples were transferred to a new 96-well plate and the absorbance at 590 nm was measured with a spectrophotometer (U-2000, Hitachi, Tokyo, Japan).

**Cytokines assay**

Quantification of cytokines was carried out by ELISA kits. After 24 hours of incubation, the culture medium of control and treated cells was collected, centrifuged, and stored at −80°C until tested. The levels of interleukin (IL)-1beta, tumor necrosis factor (TNF)-alpha, interferon (IFN)-gamma, IL-6, and IL-4 released into culture medi-
um were determined with ELISA kits from eBioscience (San Diego, CA, USA).

**Statistical analysis**

Experimental results were analyzed in triplicates and expressed as means ± standard deviation (SD). The results were subjected to one-way analysis of variance (ANOVA) and Duncan's multiple range tests and the significance of differences between sample means was calculated; and p≤0.05 was considered significant.

**Results**

Effect of PS-MCM on cell viability of THP-1 monocytes

The data showed that biomass of mycelium from *C. militaris* was significantly increased in porcine liver extracts medium when compared with the basal medium or the de-fat soybean extracts medium by surface liquid cultivation (Supplemental Figure 1A). Moreover, we evaluated polysaccharides production of surface liquid-cultured *C. militaris* by different substrates, including polysaccharides obtained from mycelia of *C. militaris* grown in basal medium, polysaccharides obtained from mycelia of *C. militaris* grown in porcine liver extracts medium, and polysaccharides obtained from mycelia of *C. militaris* grown in de-fat soybean medium. Results are shown in Supplemental Figure 1B. The ability of polysaccharide production by *C. militaris* was greater in basal medium than porcine liver extracts medium and de-fat soybean medium, revealing that protein and nitrogen resource may suppress the production of polysaccharides. There were no significant differences in DPPH and ABTS free radicals scavenging activity, and chelating activity among the groups (Supplemental Figure 2A and 2B).

![Figure 1. Effects of PS-MCM on cell proliferation in THP-1 cells for 24 h. Results are expressed as mean ± SD (n = 3)](image)

The THP-1 monocytes were treated with various doses (50, 100, and 250 μg/mL) of PS-MCM isolated from mycelia of *C. militaris* grown in basal medium for 24 hours and the cell viability were determined by MTT assay. The results showed that BM PS-MCM promoted cell proliferation of THP-1 monocytes in a dose-dependent manner (Figure 1).

Doxorubicin is a drug used in chemotherapy. The THP-1 monocytes were treated with various doses of doxorubicin for 24 hours, and cell viability was measured by MTT assay. As shown in Figure 2, when compared to the control group, doxorubicin (0.5, 1, 2.5, and 5 μM, respectively) treatment significantly resulted in cell toxicity in THP-1 monocytes, and the cell viabilities were 95.8, 74.9, 61.3, and 23.9%, respectively. Some 2.5 μM of doxorubicin was used to induce cell toxicity for suppressing immune activity of the cells, and the protective effects of PS-MCM on doxorubicin-treated THP-1 monocytes against side effects in chemotherapy were investigated.
As shown in Figure 3, when compared with the blank group, PS-MCM treatment significantly increased the cell proliferation of THP-1 monocytes, whereas 2.5 μM of doxorubicin treatment markedly decreased cell viability. Moreover, PS-MCM treatment attenuated doxorubicin-induced toxic effects in THP-1 monocytes.

Effects of PS-MCM on immune regulation in doxorubicin-treated THP-1 monocytes

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**Figure 2.** Inhibitory effects of doxorubicin on cell viability of THP-1 cells for 24 h. Results are expressed as mean ± SD (n = 3).

**Figure 3.** Effects PS-MCM combined with doxorubicin on cell viability of THP-1 cells for 24 h. Results are expressed as mean ± SD (n = 3). Significant difference was shown by different letters (p < 0.05). PS: PSMCM; D: doxorubicin.
The levels of IL-1beta, TNF-alpha, IFN-gamma, and IL-6 were detected after treating with PS-MCM or/and doxorubicin in THP-1 monocytes. The results displayed in Table 1 show that, 250 μg/mL of PS-MCM treatment efficiently promoted the production of cytokines in THP-1 monocytes when compared to the blank group. Reciprocal results were obtained with doxorubicin treatment. None the less, treating with PS-MCM potentially reversed the immune-stimulatory activity in doxorubicin-induced THP-1 monocytes for the generation of L-1beta, TNF-alpha, IFN-gamma, and IL-6.

**Effect of PS-MCM on cell viability of EL-4 T cells**

The EL-4 T cells were treated with various doses (50, 100, and 250 μg/mL) of PS-MCM for 24 hours and the cell viability were determined by MTT assay. The results indicated that PS-MCM promoted cell proliferation of EL-4 T cells in a dose-dependent manner (Figure 4).

**In addition, The EL-4 T cells were treated with various doses of doxorubicin for 24 hours, and the cell viability was measured by MTT assay. As shown in Figure 5, when compared with the control group, doxorubicin significantly caused cell toxicity in EL-4 T cells. Some 2.5 μM of doxorubicin was used to induce cell toxicity for suppressing immune activity of the cells, and the protective effects of PS-MCM on doxorubicin-treated EL-4 T cells against side effects in chemotherapy were investigated. The results in Figure 6 show that, PS-MCM significantly increased cell proliferation of EL-4 T cells when compared with the blank group, and 2.5 μM of doxorubicin markedly suppressed cell viability, whereas PS-MCM attenuated doxorubicin-induced toxic effects in EL-4 T cells.**

**Discussion**

THP-1 cells have been extensively used to study func-
tion, mechanism, signaling pathways and nutrient or drug transport of monocytes/macrophages. THP-1 cells can accurately mimic human monocytes and can differentiate into macrophages if properly incubated with the appropriate stimuli. Due to their uniform genetic background, THP-1 cells are frequently used as a model system for the study of monocytes abolishing donor variability. This cell line has also been employed to study cardiovascular diseases, for dermatological tests of new cosmetics, and to analyze the effect of food-derived products in the immune response. Our data showed that PS-MCM exerted M1-like induction in monocytes, inducing cytokines production such as IL-1β and IFN-γ (Table 1), and attenuated cytotoxic effects caused by chemotherapeutic drug doxorubicin in monocytes (Figure 1; Figure 2; Figure 3). Recently, polysaccharides have been reported to display immune stimulatory effects via inducing cytokines production in THP-1 monocytes. Cytokines IL-1β, TNF-alpha, and IFN-γ have been found to act as immune regulators and suppress cancer cells. Dox can cause a severe decrease in the number of white blood cells, red blood cells, and platelets in cancer patients. The accumulation of Dox in white blood cells has been investigated, and this accumulation in monocytes was greater than granulocytes and lymphocytes, suggesting that Dox resulted in cytotoxic effects and immune-suppression in monocytes. Taken together, our results suggest that PS-MCM has potential to be used as a novel agent for immunotherapy in cancer patients by up-regulating monocytes survival and cytokines secretion.

Conclusion

Here we found that PS-MCM increased the cell proliferations of monocytes and T cells and promoted immune activity through cytokines regulation. It is considered that PS-MCM could be developed as functional foods for attenuating side effects in cancer patients with chemotherapy.

Conflict of interest

The authors declare that there are no conflicts of interest regarding the publication of this paper.

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