Novel Insights into Crystal-Induced Kidney Injury

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Abstract

Background: The entity of crystal nephropathies encompasses a spectrum of different kidney injuries induced by crystal-formed intrinsic minerals, metabolites, and proteins or extrinsic dietary components and drug metabolites. Depending on the localization and dynamics of crystal deposition, the clinical presentation can be acute kidney injury, progressive chronic kidney disease, or renal colic. Summary: The molecular mechanisms involving crystal-induced injury are diverse and remain poorly understood. Type 1 crystal nephropathies arise from crystals in the vascular lumen (cholesterol embolism) or the vascular wall (atherosclerosis) and involve kidney infarcts or chronic ischemia, respectively. Type 2 crystal nephropathies arise from intratubular crystal deposition causing obstruction, interstitial inflammation, and tubular cell injury. NLRP3 inflammasome and necroptosis drive renal necroinflammation in acute settings. Type 3 is represented by crystal and stone formation in the draining urinary tract, i.e., urolithiasis, causing renal colic and chronic obstruction. Key Messages: Dissecting the types of injury is the first step towards a better understanding of the pathophysiology of crystal nephropathies. Crystal-induced activation of the inflammasome and necroptosis, crystal adhesion, crystallization inhibitors, extratubulation, and granuloma formation are only a few of certainly many involved pathomechanisms that deserve further studies to eventually form the basis for innovative cures for these diseases.

Introduction

A substantial number of kidney diseases involve crystalline microparticles that contribute to mechanical obstructions, local intrarenal inflammation, and tissue injury. Crystalline nephropathies can be divided into three subgroups [1]: renal ischemic due to vascular calcifications or crystal embolism (type 1), intra- and extratubular crystalline precipitates causing tubular injury (type 2), and nephrolithiasis causing obstructive nephropathy (type 3) (Fig. 1). Traditionally, research on the pathophysiology of crystalline nephropathies has focused on the physicochemical properties of urinary minerals, supersaturation, and crystal growth inhibition. Accumulating data now enrich these concepts by mechanisms of intrarenal crystal adhesion, crystal-induced inflammation, and cell death that link local crystal deposits to a large
number of different diseases in and outside the kidney referred to as “crystallopathies” [2]. Here, we briefly summarize the clinical presentations and pathophysiological mechanisms of such crystalline nephropathies.

**Type 1 Crystalline Nephropathy: Renal Cholesterol Embolism**

Cholesterol crystal embolism (CCE) occurs when emboli of cholesterol crystals (CCs) arising from the rupture of an atherosclerotic plaque from aortic or other major arteries block smaller arteries (150–200 μm in diameter), which may affect kidneys, skin, gastrointestinal system, eyes, muscles, bones, brain, nerves, internal organs, and limbs [3]. Since kidneys are located near the abdominal aorta, they are the most frequent target organ of CCE [3]. However, diagnosis of renal CCEs is difficult and controversial and often based on the presence of a precipitating event, renal failure, and peripheral CCE. Clinically, renal CCE can be acute, subacute, or chronic [3].

A critical factor in the development of renal CCEs is the formation of an atherosclerotic lesion. Spontaneous or iatrogenic rupture of plaques in the thoracic aorta or renal arteries results in showers of CCs, which obstruct the smaller arteries. In the kidney, obstruction of arcuate and interlobar arteries by CCs causes cortical and medullary infarcts [1]. CCs can also induce either passive (physical) or regulated necrosis of the endothelial cells. They are reported to induce endothelial cell injury via complement activation [4]. Furthermore, they kill tubular epithelial cells by activating receptor-interacting serine/threonine-protein kinase 3 (RIPK3) and mixed lineage kinase domain-like (MLKL) [5, 6]. Activation of MLKL...
## Table 1. Acute or chronic crystal-induced kidney diseases

| Type | Renal damage | Disease | Crystal (source of crystal-forming factor) |
|------|--------------|---------|------------------------------------------|
| Type 1 (vascular) | AKI | cholesterol embolism | cholesterol |
| | CKD | atherosclerotic renal artery stenosis | cholesterol, apatite, CaP |
| Type 2 (tubular) | AKI | diet-induced crystal nephropathy | CaOx monohydrate (whewellite) |
| | | | CaOx dihydrate (weddellite) |
| | | | CaP-, CaOx-, and vitamin C-rich foods and drinks |
| | | drug-induced crystal nephropathy | uric acid |
| | | tumor lysis syndrome | drug crystals |
| | | drug induced crystalluria | CaP |
| | | acute phosphate nephropathy | |
| | | enteric crystal nephropathy | CaOx monohydrate (whewellite) (bariatric surgery, IBD, celiac disease) |
| | | short bowel syndrome | CaOx dihydrate (weddellite) |
| | | myogen crystal nephropathy | myoglobin (rhabdomyolysis) |
| | | bilirubin crystal nephropathy (bile cast nephropathy) | bile pigment |
| | | paraprotein-induced crystal nephropathy (light-chain cast nephropathy) | monoclonal light chains |
| CKD | CAKUT | genetic forms of RTA | brushite, CaCO₃, CaOx, CaP, hydroxyapatite, struvite |
| | | genetic forms of hyperuricosuria | uric acid |
| | | primary hyperoxaluria | CaOx monohydrate (whewellite), CaOx dihydrate (weddellite) |
| | | cystinosis | cystine |
| | | APRT deficiency | adenine |
| | | genetic diseases that cause nephrocalcinosis | CaP, calcium pyrophosphate |
| | | Dent disease, Lowe syndrome, idiopathic infantile hypercalcinemia and hypercalciuria | |
| | | diet- or drug-induced crystal nephropathy | uric acid (diets rich in sodium, fructose, sucrose) |
| | | tophaceous gout | uric acid (diets low in potassium, calcium, phytate, fluid administration as part of Mesoamerican nephropathy treatment) |
| | | immunoglobulin-induced crystal nephropathies | monoclonal light chains |
| | | light-chain cast nephropathy, proximal tubulopathy with or without Fanconi syndrome, crystalglobulinemia, crystal-storing histiocytosis | |
| | | chronic glomerular crystal nephropathies | monoclonal and polyclonal IgG |
| | | cryoglobulinemia | monoclonal light chains |
| | | crystalglobulinemia | cystine |
| | | cystinosis | |

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by RIPK3 results in disruption of the plasma membrane and a regulated form of necrosis, which is termed necroptosis [7]. In addition, CCs can directly induce inflammation. For example, during the development of atherosclerosis, macrophages engulf small CCs, which activate the NLRP3 inflammasome, and thus, set up IL-1β-dependent inflammation [8]. The binding of CCs to the human macrophage-inducible C-type lectin receptors on the surface of macrophages and dendritic cells induces secretion of proinflammatory cytokine tumor necrosis factor (TNF) and macrophage inflammatory protein 2 [9]. CCs can also activate Syk and PI3K in human macrophages and dendritic cells and induce IL-1α/β [10]. However, the contribution of CCs to inflammation and necrosis in CCE remains obscure to date, mostly owing to the lack of suitable animal models of the disease. Nevertheless, researchers have tried to mimic atheroembolic renal diseases in rabbits and rats by injecting human atherosclerotic material or CCs [11, 12]. These studies identified early neutrophilic reactions followed by recruitment of macrophages in response to the atherosclerotic material or CCs. All these inflammatory events culminate in the formation of giant cell granulomas in the lumen, luminal thrombosis, and long-term obstruction because of crystal emboli or fibrotic narrowing of the lumen leading to downstream ischemic injury in the kidneys.

### Type 2 Crystalline Nephropathy: Intratubular Crystal Formation

The glomerular ultrafiltrate can be enriched with minerals, proteins, or drug metabolites, especially in states of volume depletion fostering supersaturation. Acute supersaturation can induce a sudden onset of crystal formation, leading to acute episodes of crystal-induced tubular cell injury, interstitial inflammation, and impairment of renal function, i.e., acute kidney injury (AKI) (Table 1). In contrast, long-standing moderate supersaturation can rather promote crystal formation over extended periods of time, tubule obstruction, and tissue remodeling, i.e., chronic kidney disease (CKD) (Table 1). For example, monoclonal gammopathies can be associated with accelerated filtration of immunoglobulin light chains, some of which form crystals inside the lumen of the proximal tubules, depending on their specific physiochemical properties [13]. Other options are precipitation as β-sheets and amyloid fibril deposition often leading to nephrotic syndrome [13, 14]. Occasionally, light-chain crystals also precipitate in intrarenal mononuclear phagocytes causing so-called cryoglobulinemia or crystal-storing histiocytosis [13, 14]. Chemotherapy for leukemia can induce tumor lysis syndrome, a condition characterized by uric acid release from dying blood cells and monosodium urate (MSU) crystal precipitates inside the tubular lumen.

| Type          | Renal damage | Disease                                                                 | Crystal (source of crystal-forming factor) |
|---------------|--------------|------------------------------------------------------------------------|------------------------------------------|
| Type 3        | colic        | monogenic crystal nephropathies (CAKUT)                                 | brushite, CaCO₃, CaOx, CaP, hydroxyapatite, struvite |
|               |              | genetic forms of RTA                                                   | CaOx monohydrate (whewellite), CaOx dihydrate (weddelite) |
|               |              | primary hyperoxaluria                                                 | cysteine                                  |
|               |              | cystinosis                                                            | adenine                                   |
|               |              | APRT deficiency                                                       | uric acid, CaOx, CaP                      |
|               |              | genetic forms of hyperuricosuria and hypercalciuria (as listed above) |                                          |
|               |              | diet- or drug-induced crystal nephropathy                             | CaOx, CaP (diets rich in oxalate, sodium, fructose, vitamin C, sucrose; diets low in potassium, calcium, phytate) |
|               |              | hyperuricosuric syndromes                                             | uric acid                                 |
|               |              | drug crystalluria                                                     | drug crystals                             |
|               |              | enteric oxalosis                                                      | CaOx monohydrate (whewellite)             |
|               |              | short bowel syndrome                                                  | CaOx dihydrate (weddelite) (bariatric surgery, IBD, celiac disease) |

| Type 3 (uroliithiasis) | colic | monogenic crystal nephropathies (CAKUT) | brushite, CaCO₃, CaOx, CaP, hydroxyapatite, struvite |
|                       |       | genetic forms of RTA | CaOx monohydrate (whewellite), CaOx dihydrate (weddelite) |
|                       |       | primary hyperoxaluria | cysteine |
|                       |       | cystinosis | adenine |
|                       |       | APRT deficiency | uric acid, CaOx, CaP |
|                       |       | genetic forms of hyperuricosuria and hypercalciuria (as listed above) | |

AKI, acute kidney injury; APRT, adenine phosphoribosyl transferase; CaCO₃, calcium carbonate; CAKUT, congenital anomalies of the kidney and the urinary tract; CaOx, calcium oxalate; CaP, calcium phosphate; CKD, chronic kidney disease; IBD, inflammatory bowel disease; RTA, renal tubular acidosis.
Obstruction of distal tubules and collecting ducts lead to AKI, a condition addressed by hydration, alkalization, and preventive uric acid breakdown with recombinant uricase, an enzyme clearing the metabolite substrate of such crystal plugs.

Patients with advanced liver disease retain high levels of bile salts in the circulation as clinically evident by severe jaundice. Once filtered in the kidney, bile salts can precipitate and form casts that obstruct nephrons. In addition, bile salts and bilirubin elicit direct toxic effects on renal tubular cells. Bile cast nephropathy can be a contributing factor in so-called “hepatorenal syndrome” [15]. Other gastrointestinal disorders such as celiac disease, inflammatory bowel disease, or a previous intestinal bypass surgery can be associated with oxalate nephropathy [16]. All of these conditions are associated with increased intestinal oxalate absorption implying hyperoxalemia and hyperoxaluria. Inside the nephron, oxalate does not crystallize up to the distal tubule, the site of calcium secretion. Microparticle formation, crystal adhesion to the luminal tubular cell membranes, and crystal growth can affect nephron function and cause AKI or CKD [16]. Nephrocalcinosis is also a manifestation of numerous rare genetic metabolic or renal disorders as listed in Table 1 [1, 14, 17].

Many mechanisms contribute to AKI associated with rhabdomyolysis. Muscle cell injury follows a release of myoglobin that is freely filtered and can precipitate inside renal tubules, especially during hypovolemia, often coinciding in earthquake or car crash victims [18]. Dehydration is also a common cofactor in crystal-induced AKI related to exposures to drugs, toxins, or dietary components that tend to crystallize in the urine. Oxalate-rich food products as a cause of secondary hyperoxaluria and acute oxalate nephropathy include spinach, nuts, black tea, rhubarb, and certain fruits, especially when intestinal abnormalities increase oxalate uptake. Vitamin C supplements or accidental polyethylene glycol ingestion trigger acute oxalate nephropathy because oxalate is their major downstream water-soluble metabolite [19, 20]. Sodium phosphate, a component of oral bowel purgatives often used before colonoscopy, can cause massive intratubular precipitation of calcium phosphate crystals, leading to acute phosphate nephropathy [21]. Finally, sulfamethoxazole, indinavir, aciclovir, and other drugs can cause acute crystalline kidney injury, especially when high doses coincide with low fluid intake [22].

Pathophysiology of AKI

Supersaturation of urine because of either dehydration-induced concentration or extensive exposure to crystal-forming substances results in crystallization of solutes in renal tubules [20]. These deposited crystals then contribute to kidney injury by inducing direct and indirect cytotoxicity as well as by setting up an auto-amplification loop of necroinflammation [23–25].

Direct Crystal Cytotoxicity. Although tubular cells surrounding crystal deposits often die by necrosis, the underlying mechanisms of such cytotoxicity effect remained ambiguous. It has been proposed that tubular cells try to digest smaller crystals in lysosomes after phagocytosis. During this process, calcium-containing crystals release free calcium into the cytosol and thus engage the calpain-mediated cellular necrosis pathways [1, 2]. Conversely, indigestible lysosomal crystals can destabilize lysosomal membranes, release lysosomal enzymes into cytosol causing cell stress to engage autophagic tubular cell death. Some of the lysosomal enzymes – e.g., cathepsin B – are reported to activate necroptosis by cleaving the necrosis inhibitor protein RIPK1 in immune cells [26]. In fact, crystals of calcium oxalate, calcium phosphate, calcium pyrophosphate, cysteine, cholesterol, and MSU induced tubular cell necroptosis in vitro by engaging RIPK3 and pseudokinase MLKL [5, 6] (Fig. 2). Moreover, both chemical inhibition as well as deficiency of promoters of necroptosis – for example, Ripk3 and Mlkl deficiency – protected mice from acute oxalate nephropathy [5]. On the contrary, deficiency of inhibitors of necroptosis – for example, repulsive guidance molecule b aggravated acute oxalate nephropathy [27]. During rhabdomyolysis-induced AKI, tubular cytotoxicity because of heme crystals also involved the RIPK3-MLKL pathway. In addition, oxalate and folic acid crystals induced ferroptosis of tubular cells in vivo, resulting in AKI [28, 29]. However, the ability of oxalate and folic acid crystals to induce ferroptosis in vitro remains obscure to date.

Indirect Crystal Cytotoxicity. Crystals can also trigger necrosis of tubular cells indirectly via necroinflammation [23] (Fig. 2). Crystal-induced cellular necrosis leads to the release of danger-associated molecular patterns, high-mobility group protein 1, histones, mitochondrial DNA, demethylated DNA and RNA, ATP, uric acid, and double-stranded DNA into the extracellular compartment [23]. These factors may induce regulated necrosis of neighboring cells by engaging death receptors on their surface [7, 23]. Moreover, because of their strong basic charge, extracellular histones disrupt plasma membranes of neighboring live cells and thus aggravate renal injury [30]. Furthermore, a broad range of crystals of different size and shape are reported to induce neutrophil necroptosis [31, 32]; however, whether crystal-induced neutro-
phil necroptosis contributes to crystalline nephropathies remains unexplored to date.

**Crystal-Induced Renal Inflammation.** As mentioned earlier, crystal-induced cellular necrosis leads to the release of danger-associated molecular patterns, alarmins, histones, proteases, etc., which can also trigger inflammation via activating the Toll-like receptors (TLRs). Tubular cell necrosis promotes leukocyte influx in the kidney as well as complement activation, thus enhancing renal necroinflammation [24]. In addition, crystals also activate the NLRP3 inflammasome in renal phagocytes and induce secretion of mature IL-1β during crystalline nephropathy [33]. A variety of crystals share similar mechanisms of NLRP3 inflammasome activation – e.g., phagocytosis, reactive oxygen species production, cathepsin B release, and potassium efflux [33, 34]. Further, mature IL-1β sets up subsequent inflammation via the IL-1 receptor [33]. Some crystals can also induce inflammation via other pathways in addition to inflammasome activation. For example, direct membrane binding of MSU crystals induces cell surface lipid sorting and activation of a tyrosine protein kinase Syk in a receptor-independent manner [34].

Activation of NLRP3 inflammasomes induces cell death, which is termed pyroptosis [7]. Although crystals do not induce pyroptosis of tubular cells directly [5, 6], the chances of tubular cell death via pyroptosis cannot be ignored in crystalline nephropathies. Some inflammar-
tory cytokines (e.g., TNFa) drive necroptosis of tubular cells to enhance oxalate crystal-induced AKI [5]. Multiple cathepsins are also suggested to regulate crystal-induced cell death in an inflammasome-independent manner [35]. Furthermore, physical obstruction of the tubular lumen by crystals themselves or necrotic cell debris contributes to a decline in renal function in crystalline nephropathy [1]. All these processes constitute the auto-amplification loop of necroinflammation [7, 23]. Interestingly, an inhibitory C-type lectin receptor (Clec12a) was recently identified as an inhibitory receptor for MSU that regulates inflammation in response to cell death [36], though its contribution to the suppression of crystal-induced renal inflammation remains to be studied.

**Pathophysiology of CKD**

Crystalline CKD, also referred to as kidney stone disease, if often a result of genetic disorders that enhance the synthesis of certain minerals or metabolites so much that their concentration in urine reaches supersaturation, leading to their crystallization (Table 1). The term nephrocalcinosis refers only to the presence of calcium phosphate and calcium oxalate crystals in the kidney. However, kidney stones comprise crystals of uric acid and adenine, in addition to calcium-containing crystals.

**Sites of Intrarenal Crystallization.** (a) The proximal tubule. It is the most preferred site of crystallization in the kidney owing to its native function. Proximal tubules reabsorb most of the glomerular filtrate, and thus, concentrate urine, which favors supersaturation and crystallization of minerals, metabolites, small proteins such as light chains, etc. (b) The thick ascending loop of Henle. It reabsorbs chloride ions from the urine, leading to urine acidification, favoring crystallization of lithogenic solutes such as uric acid, and secretes calcium ions into urine, favoring crystallization of calcium-containing solutes such as calcium phosphate and oxalate. In addition, it secretes uromodulin, which enhances cast formation in association with light chains, necrotic debris, as well as myoglobin [1]. (c) The medullary collecting ducts. They passively excrete urea in the urine, and therefore, are the preferential sites of uric acid crystallization in the kidney [37]. (d) The interstitial compartment. Extratubularation of intratubular crystal plugs leads to crystal granuloma in the renal interstitial compartment [38]. The presence of crystal granulomas is a characteristic feature of certain forms of crystalline CKD, e.g., urate nephropathy.

**Mechanisms of Crystal-Induced Inflammation.** Similar to crystalline AKI, crystal deposits in crystalline CKD also activate the NLRP3 inflammasome and induce inflammation in the kidney. Deficiency of inflammasome components, NLRP3, and ASC in mice reduced renal inflammation during progressive renal failure in nephrocalcinosis-related CKD [39, 40]. Furthermore, chemical inhibition of NLRP3 using a specific inhibitor – CP-456773 or β-hydroxybutyrate – reduced renal inflammation and progressive decline in renal function in murine nephrocalcinosis-related CKD [40, 41]. Inflammasome activation leads to secretion of mature IL-1β and IL-18 that contribute to renal inflammation. Surprisingly, IL-1 receptor inhibition using anakinra has no effect on progressive decline in renal function in murine nephrocalcinosis-related CKD [40, 41], suggesting a contribution of inflammasome-independent functions of NLRP3, which involve transforming growth factor beta receptor signaling-mediated renal fibrosis [42]. Indeed, chemical inhibition of NLRP3 by CP-456773 during oxalate and adenine nephropathy or by β-hydroxybutyrate during oxalate nephropathy reduced intrarenal fibrosis in murine crystalline CKD models [40, 41]. In addition, our own data demonstrated that β-hydroxybutyrate induced M2 macrophage phenotype shift and also contributed to a decline in renal inflammation and preserved renal function during chronic oxalate nephropathy without affecting intrarenal crystal deposition [40].

Macrophages, which often have a giant cell-like appearance, border the extratubular crystal plaques in the interstitial compartment, leading to the term “crystal granuloma” [1]. They are thought to contribute to the removal of crystal deposits via phagocytosis and digestion [1]. Consistently, colony-stimulating factor suppressed renal crystal deposition by inducing M2 macrophages in the kidneys of hyperoxaluric mice [43]. However, the molecular mechanisms that trigger the formation of crystal granulomas in crystalline CKD are unknown to date.

Furthermore, the procedure of crystallization itself has immunostimulatory effects – e.g., high concentrations of uric acid activate TLRs (TLR4) and induce expression of inflammasome components [34], whereas oxalate ions induce expression of chemokines and osteopontin, which regulate inflammation [44]. Moreover, crystal plugs adhere to the tubular lumen and cause obstruction, leading to declined renal function. Crystal adhesion to tubular epithelium is regulated by several surface molecules such as CD44, annexin II, osteopontin, phosphatidylserine, hyaluronan, as well as TNF receptor signaling [45–47].
Type 3 Crystalline Nephropathy: Urolithiasis

Nephrolithiasis, ureterolithiasis, or cystolithiasis can cause nephron loss, renal inflammation, and fibrosis as a manifestation of obstructive nephropathy. In the North American population, 7–11% are affected by urolithiasis, with a preponderance of male individuals; however, CKD as an obvious consequence of urolithiasis is recognized only in a small percentage of stone formers [45]. Many factors contribute to urolithiasis, but hereditary metabolic diseases associated with an increased production of cysteine, uric acid, or oxalate are paradigmatic examples illustrating the pathogenesis of crystalluria and stone formation (Table 1). Variants in genes encoding for mineral or metabolite transporters that promote supersaturation of minerals or metabolites in the urine are another common cause of urolithiasis [17]. In addition, diets rich in fructose or oxalate, together with low fluid intake or excessive sweating or diarrhea, can have synergistic effects on microcrystal and stone formation [45, 48].

Urolithiasis most commonly presents as renal colic or a sudden onset of crescendo and decrescendo in flank pain or the inguinal region. Oliguria is absent unless bilateral urolithiasis causes simultaneous obstruction [49]. Stone passage or removal relieves pressure and pain. Most frequently, stones suddenly mobilize from calyces of the renal pelvis, but the triggering events remain largely unknown. During stone passage, the rough surface of the calculus, together with the mechanical forces of ureteric peristalsis, can cause mucosal injury, bleeding, and swelling at the site of obstruction. Despite the alarming clinical presentation, renal colic is not necessarily associated with long-term impairments of urinary tract function unless chronic obstruction leads to obstructive nephropathy and irreversible nephron loss.

Calcium formation often initiates at Randall plaques inside the renal pelvis, i.e., interstitial apatite plaques formed along thin parts of the loops of Henle [46]. Randall plaques are a preferred nidus site for calcium oxalate stones. The plaque-stone interface is a multilayer ribbon consisting of matrix molecules such as Tamm-Horsfall urinary glycoprotein and calcium oxalate crystals. Indeed, although uromodulin inhibits luminal crystal formation, it is a regular component of interstitial crystal plaques. Sites of brushite and hydroxyapatite stone formation in the renal pelvis are plugs inside Bellini ducts and inner medullary collecting ducts formation. Such plugs occur upon ileostomy, small bowel resection, or bariatric surgery as well as cystinuria, primary hyperparathyroidism, or primary hyperoxaluria [50].

In healthy kidneys, crystal nucleation does not necessarily lead to crystal attachment or growth [46]. Indeed, epithelial injury or absence of crystallization inhibitors enhances crystal adhesion to the tubular epithelium [38, 46]. For example, own experiments suggest that the receptors of TNFs mediate the adhesion of calcium oxalate crystals to the tubular epithelium by inducing the expression of crystal adhesion molecules at the luminal surface [47]. However, the process of adhesion and growth of calcium oxalate stones at Randall plaques and plugs obstructing the ducts of Bellini remains poorly understood. Some factors include plug composition, urine flow, and mineral supersaturation [38, 46].

Summary

Crystals contribute to kidney injury involving a broad variety of different pathomechanisms. Vascular manifestations (type 1) involve calcium phosphate deposits inside the vascular wall or CC deposits in atheromatous plaques that both can contribute to renal ischemia by contributing to renal artery stenosis or by cholesterol embolism. Supersaturation of minerals or certain proteins can promote crystallization inside the tubular compartment and cause tubular obstruction and tubular epithelial cell injury up to necrosis. These processes trigger inflammation, interstitial edema, and eventually renal fibrosis (type 2). Nephro- and urolithiasis present as renal colic or obstructive nephropathy, which when chronic can cause extensive nephron damage. The molecular mechanisms that link crystal formation to adhesion, growth, renal inflammation, and renal cell death remain largely to be discovered. There is hope that a better understanding of the underlying pathophysiology will help develop new cures for patients with crystalline nephropathies.

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Conflict of Interest Statement

The authors have no conflicts of interest to disclose.
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