Role of Microbial Agents in Pulmonary Fibrosis

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Pulmonary fibrosis is a form of lung disease that develops due to aberrant wound-healing following repeated alveoli injury in genetically susceptible individuals, resulting in chronic inflammation, excess deposition of the extracellular matrix components, mainly collagen, and scarring of lung tissue. In addition to irradiation, environmental agents such occupational inhalants, and chemotherapeutic agents, microbial agents also play a role in the etiology of the disease. While viruses have received the most attention, emerging evidence suggest that bacteria and fungi also play a part in the etiology of pulmonary fibrosis. Furthermore, successful use of antibiotics, antiviral and antifungal drugs in several studies to attenuate fibrosis progression is also an indication of microbial involvement in the pathogenesis of the disease and could be a promising therapeutic modality for treating pulmonary fibrosis initiated or exacerbated by infectious agents.

INTRODUCTION

Pulmonary fibrosis (PF\textsuperscript{†}) is a form of progressive lung disease that belongs to a large family of lung diseases called interstitial lung diseases [1]. The most common type of PF is idiopathic pulmonary fibrosis (IPF), and its cause is unknown. IPF is the most severe forms of interstitial lung disease that result in PF, with an annual incidence of 6.8–8.8 per 100,000 and 16.3–17.4 per 100,000 populations, using narrow and broad case definitions respectively in the U.S. [2]. PF develops as a result of a repetitive injury to the alveolar epithelium or endothelium, which triggers elements of the innate and adaptive immune system to restore the tissue architecture of the damaged tissue. Inflammatory mediators such as the profibrotic cytokine transforming growth factor-be-

\textsuperscript{†}Abbreviations: PF, Pulmonary Fibrosis; IPF, Idiopathic Pulmonary Fibrosis; AE-IPF, Acute Exacerbated Idiopathic Pulmonary Fibrosis; BAL, Bronchoalveolar Lavage; NETs, Neutrophil Extracellular Traps.

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propranolol), and antibiotics (nitrofurantoin, ethambutol) as well as exposure to animal proteins, molds, and bacteria can also cause PF [2,5]. In addition, medical conditions such as gastrointestinal flux disease, autoimmune diseases (rheumatoid arthritis and systemic lupus erythematosus), sarcoidosis, and certain muscle diseases (such as dermatomyositis, polymyositis, and the anti-synthetase syndrome), can also lead to scarring of the lungs [6-9].

Several studies and reviews support a genetic basis for the development of PF. Familial PF occurs when two or more members of the same biological family are affected by the PF, and approximately 10 to 15 percent of those with an “idiopathic” form of PF have another family member affected by the disease [10-14]. Mutations and polymorphisms in several genes such as telomerase reverse transcriptase gene (TERT), and telomerase RNA component gene (TERC) have been shown to increase susceptibility to IPF [10,11,14-17]. Current therapy for the management of IPF include the use of immunosuppressants (prednisone), chemotherapy drugs (cyclophosphamide), anti-fibrotic drugs (Pirfenidone), proton pump inhibitors, oxygen therapy, and even lung transplant surgery for the management of IPF [18].

In this review, we report experimental findings from several studies with the goal of identifying the role of microbial infectious agents in PF. We provide evidence of antimicrobial drug efficacy in several cases of PF and suggest future experiments that may shed more light on the role of infectious agents in the pathogenicity of PF.

MICROBIAL AGENTS AND PULMONARY FIBROSIS

Accumulating evidence suggests that infectious microbial agents (viral, fungal, and bacterial) may play a role in PF. While several studies have identified the presence of infectious agents in induction and exacerbation of PF using animal models, there is a paucity of literature regarding microbial induction of IPF in patients [19-22]. Studies employing antimicrobials such as antivirals, antibiotics, and antifungals also show great promise for the treatment of IPF and strengthen the connection between microbial agents and IPF [20,23-26]. Figure 1 highlights evidence available to support microbial involvement in PF.

VIRUSES AND PULMONARY FIBROSIS

IPF has been suggested to have a viral etiology based on the presence of viral signatures in the lungs of IPF patients, and the observation that IPF patients showed improvement when treated with antivirals [26,27]. Viruses detected in IPF patients include members of the Human Herpes Viruses (HHVs) family, such as Epstein-Barr virus (EBV), cytomegalovirus (CMV), Herpes simplex virus (type-1, -6, -7, -8) and Kaposi’s sarcoma herpesvirus [28]. HHVs can induce endoplasmic reticulum stress and apoptosis in epithelial cells in vitro, and the murine herpesvirus type 68, which is very similar to HHVs, can cause IPF by upregulating up-regulating TGF-β [29-31]. All these could be mechanisms of viral involvement in the pathogenesis of IPF.

EBV has received the most attention in relation to IPF. IPF patients have both deoxyribonucleic acid (DNA) and protein of EBV in their bronchoalveolar lavage fluid and lung tissue [32-34]. Tang et al., tested for the presence of eight HHVs in lung specimens of IPF patients and reported that EBV was present in 97 percent of the IPF patients, and 36 percent in the control. Other viruses identified were CMV and HHV-8 [27,35]. Rearrangement in EBV genome associated with productive EBV replication was found in lung tissue biopsies of 61 percent of EBV DNA-positive IPF patients [35]. Also, EBV latent membrane protein 1 is associated with rapid disease progression in IPF [21]. Interestingly although murine herpesvirus type 68 did not induce PF in murine models of PF, it enhanced the progression of fibrosis when administered with fibrotic stimuli [36-38]. Therefore, it can be inferred that viruses may be involved in the progression rather than initiation of IPF.

However, there are conflicting reports that negate the existence of EBV in cases of IPF [39,40]. Most of the studies on EBV and IPF are retrospective and show association and not causation. Also, since infection is a potential complication of therapy, there is also a high prevalence of EBV DNA in the general population it remains unclear how EBV plays a role in IPF.

Cytomegalovirus (CMV) is also present in cases of PF [41,42]. A study found elevated levels (80 percent) of CMV antibodies (anti-CMV) and other HHVs in PF patients compared with 30 percent in the control group [41].

Other HHVs such as HHV-1 and HHV-6 have been shown to have a link to PF. HHV-1 was present in 9 percent of IPF patients and 10 percent in fibrotic idiopathic interstitial pneumonia bronchoalveolar lavage fluid, whereas the control group was negative for the virus [43]. A test for Human Herpes Virus 6 (HHV-6) was 83.3 percent positive in of IPF patients, compared to 30 percent control patients also; HHV-6B antigens were present in mononuclear cells of IPF lung tissue [44]. Herpes viral infections may also contribute to the development of pulmonary hypertension in IPF patients [45].

Variable results have arisen from studies on the relationship between HCV and IPF. An immunological study conducted to evaluate the prevalence of serum antibodies in Japanese IPF patients to HCV showed that 28 percent of IPF patients were seropositive to HCV in contrast to 3.6 percent of the control population [46]. One Italian
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**Figure 1. Evidence of Microbial Involvement in Pulmonary Fibrosis.** (A) Induction of pulmonary fibrosis by infectious agents using animal models (B) Presence of infectious agents or their products in cases of PF (C) Exacerbation of PF by infectious agents (D) Success of antimicrobial drug therapy in cases of PF (E) Secondary infection of microbial etiology can induce PF (F) Other evidence.

PF: Pulmonary Fibrosis, HCV: Hepatitis C Virus, EBV: Epstein-Barr Virus, CMV: Cytomegalovirus, HHV: Human Herpes Virus, TTV: Torque-Teno Virus, RA: Rheumatoid Arthritis, MCTD: Mixed Connective Tissue Disease, PM: Polymyositis, DM: Dermatomyositis.

A study showed a higher incidence of IPF in an HCV-positive group compared to an HBV group [47]. HCV infection can also cause active alveolitis, which can lead to IPF [48]. However, HCV is also prevalent in other forms of non-fibrotic respiratory diseases, and some studies have found no correlation between HCV and IPF [49,50].

Adenovirus, particularly its early region 1 protein, is of interest in lung disease due to its ability to upregulate TGF-β1 expression in bronchiolar epithelial cells and transformation of lung epithelial cells to express mesenchymal markers [51]. In a recent study, intratracheal instillation of high dose adenoviral vectors induced an inflammatory response, lung injury, and PF in a dose-dependent manner [22]. However, contradicting studies show no adenoviral antibodies in the sera of IPF patients, and treatment of patients with corticosteroid may encourage the prevalence of adenovirus [52]. Additional studies employing a larger patient population size is required to quantify messenger RNA in infected lung tissues.

Torque-Teno Virus (TTV), a circular single-stranded virus initially associated with post-transfusion hepatitis virus infection is significantly more common in IPF patients with acute exacerbation than stable controls [53,54]. Investigators detected TTV DNA in 36.4 percent of IPF patients evaluated, and associated TTV with lower survival rate among the TTV-positive IPF patients evaluated, compared to the TTV-negative IPF patients [55]. In a more recent study by the same group, they suggest that TTV DNA titer may reflect the immunosuppressive state of the host due to treatment [56].

Taken together, the presence of significantly higher viral signatures in patients with IPF may be an indication that viruses may play a role in the pathogenesis of IPF. However, the small sample size of patients and short du-
ration of these studies makes it difficult to draw any bold conclusions. More experiments with adequate controls need to be carried out to elucidate the role that viruses play in cases of IPF.

**BACTERIA AND PULMONARY FIBROSIS**

One of the earliest evidence of microbial involvement in interstitial lung disease came from studies, which showed that a glycolipid cell-wall component of mycobacteria, trehalose dimycolate induced interstitial pneumonitis and alveolar hemorrhages in mice, with the help of T-lymphocytes [57]. Following this discovery, numerous studies have implicated bacteria in the etiology of PF. A study conducted to culture bronchoalveolar lavage fluid from patients with Granulomatosis with polyangiitis using IPF patients as a control, revealed bacterial colonization of the lungs with species such as *Pseudomonas aeruginosa*, *Streptococcus pneumonia*, *Moraxella catarrhalis*, and *Proteus mirabilis* in 8 of the 22 IPF patients sampled [58]. Another study found an increase in bacterial burden in BAL of IPF patients particularly with species such as *Haemophilus*, *Streptococcus*, *Neisseria*, and *Veillonella* which serve as a prediction of decline in lung function and death [59]. *Haemophilus influenza* and *Pneumocystis jirovecii* bacteria can also cause chronic pulmonary disease by stimulating inflammation and activating macrophages [19,60].

Two independent lung fibrosis models show that *S. pneumonia* triggers progression of PF through the action of its pore-forming cytotoxin pneumolysin (ply). Interestingly, antibiotic treatment with clarithromycin or amoxicillin at 24 h and 48 h post-infection led a significant decrease in lung hydroxyproline contents, comparable with mock-infected mice thereby abolishing the infection-induced fibrosis progression [20].

However, it can be argued that since IPF is linked with increased aspiration and gastroesophageal reflux, the increased bacterial burden in IPF patients compared to healthy control groups could be a result of microaspiration, or lack of the airway mucin gene promoter polymorphism, which is also associated with bacterial burden in BAL [61-66]. In addition, others show using an established murine model of lung fibrosis that *P. aeruginosa* did not exacerbate bleomycin-induced fibrosis [67].

It remains unclear whether a bacterial infection is a direct cause of IPF, a secondary infection, or only an exacerbating agent, particularly because patients with IPF are susceptible to developing bacterial pneumonia [68].

**FUNGI AND PULMONARY FIBROSIS**

Inhalation of fungal material can lead to lung fibrosis by infecting the pulmonary tissue, lung cavity, or by triggering an immune reaction. *Paracoccidiodes brasiliensis*, a fungus involved in the etiology of paracoccidioidomycosis (PCM), can also induce experimental PF [69,70]. PCM is characterized by a chronic inflammatory response, and neutrophil infiltration, leading to lung fibrosis and loss of lung function in most patients [71]. A study conducted on the development of fibrosis in a model of experimental chronic pulmonary PCM, employing a combined antifungal (Itraconazole) and immunomodulatory (Pentoxifylline) therapy revealed that administration of the combined therapy in advanced stage of the disease resulted in a significant reduction of granulomatous inflammation and PF, compared with the results of classical antifungal therapy using itraconazole alone [23]. In addition, Arias and colleagues show that depletion of neutrophils in PCM promotes the resolution of PF and inflammation [72]. The fungus *Aspergillus* has also been reported to colonize the parenchymal lung cavity in cases of IPF [73]. Untreated chronic pulmonary aspergillosis caused by the fungus *A. fumigatus* can progress to chronic fibrosing pulmonary aspergillosis, and this can result in chronic scarring of the lungs [74-76].

**SUCCESS OF ANTIMICROBIALS AND VACCINATIONS IN MURINE MODELS OF PULMONARY FIBROSIS**

Various microbial pathogens such as *S. pneumoniae*, *P. acnes*, *P. brasiliensis*, and proteins from *Mycobacteria* have been utilized in animal models to induce PF [20,57,69,77]. Several studies have also reported the success of antimicrobials such as ganciclovir, valacyclovir, clarithromycin, amoxicillin, azithromycin, co-trimoxazole (sulfamethoxazole, and trimethoprim), and a combined therapy of antifungal (Itraconazole) and immunomodulatory (Pentoxifylline), in decreasing the microbial load, thereby significantly reducing lung fibrosis by reducing hydroxyproline levels, restrictive lung function pattern and improving the quality of life of IPF patients [20,23-27]. Others show that protein-based vaccination of mice with established lung fibrosis with the non-cytotoxic *S. pneumonia* pneumolysin derivative (PdB) provides protection against pneumolysin-induced fibrosis exacerbation [20].

It is noteworthy that antibiotics in addition to their traditional antimicrobial effects also possess immunomodulatory and anti-inflammatory properties [25,78]. Macrolide antibiotics such as clarithromycin, azithromycin, and erythromycin can prevent the production of pro-inflammatory cytokines and immune mediators [79-81].

Therefore, additional studies need to be carried out to elucidate the precise mechanism of action underlying the therapeutic effects of antibiotics in treatment of PF.
A. Microbial Agent implicated in Pulmonary Fibrosis

**Viruses**

| Virus                      | References |
|---------------------------|------------|
| Hepatitis C Virus (HCV)   | [46-50]    |
| Epstein-Barr Virus (EBV)  | [21,27,32-35,54] |
| Cytomegalovirus (CMV)     | [27,41,42] |
| Human Herpes Virus 1 (HHV-1) | [43]         |
| Human Herpes Virus 6 (HHV-6) | [44]         |
| Human Herpes Virus 7 (HHV-7) | [27]         |
| Human Herpes Virus 8 (HHV-8) | [27]         |
| Adenovirus                | [22,51,52] |
| Torque-Teno Virus (TTV)   | [53-56]    |
| Murine gammaherpes virus type 68 (MHV-68) | [29,36-38,67,95] |
| Parainfluenza Virus       | [54]       |
| Parvovirus B19            | [42]       |

**Bacteria**

| Bacteria                     | References |
|------------------------------|------------|
| Pseudomonas aeruginosa       | [58]       |
| Streptococcus pneumonia      | [20,58]    |
| Moraxella catarrhalis        | [58]       |
| Proteus mirabilis            | [58]       |
| Propionibacterium acnes      | [77]       |
| Staphylococcus aureus        | [28]       |
| Pneumocystis jiroveci        | [60]       |
| Haemophilus influenza        | [19]       |
| Mycobacteria                 | [57]       |

**Fungi**

| Fungi                        | References |
|------------------------------|------------|
| Aspergillus fumigatus        | [73]       |
| Paracoccidiodes brasiliensis | [69-71]    |

B. Antimicrobial Studies on infectious Agents involved in Pulmonary Fibrosis

**Antivirals**

| Antivirals              | Findings                                                                 |
|-------------------------|---------------------------------------------------------------------------|
| Ganciclovir             | 2-week course of ganciclovir may attenuate disease progression in a subgroup of advanced IPF patients [26]. |
| Valacyclovir            | Treatment with this antiviral led to a decrease in sputum viral load in lungs of patients with IPF [27]. |
| Cidovir                 | Antiviral treatment administered to symptomatic animals, improved survival from 20 to 80% compared with untreated symptomatic animals, but lung fibrosis persisted in 60% of the mice [96]. |

**Antibiotics**

| Antibiotics              | Findings                                                                 |
|--------------------------|---------------------------------------------------------------------------|
| Clarithromycin           | Antibiotic treatment with clarithromycin or amoxicillin led to significantly decreased lung hydroxyproline contents, thereby in blocking S. pneumoniae-induced fibrosis exacerbation in mice [20]. |
| Amoxicillin              | Antibiotic treatment with clarithromycin or amoxicillin led to significantly decreased lung hydroxyproline contents, thereby in blocking S. pneumoniae-induced fibrosis exacerbation in mice [20]. |
PULMONARY FIBROSIS CAUSED AS A COMPLEXIFICATION OF A DISEASE WITH MICROBIAL ETIOLOGY

PF may occur as a secondary complication due to diseases of microbial etiology. Such diseases include hypersensitivity pneumonitis (also called “hot tub lung”), fibrosing mediastinitis, sarcoidosis, and untreated tuberculosis [82-86]. Autoimmune diseases with infectious etiology such as scleroderma, systemic lupus erythematosus, rheumatoid arthritis, and mixed connective tissue diseases can also induce PF [87-92].

Other evidence such as increased in activated neutrophils in BAL fluid from IPF patients associated with early mortality, and raised plasma concentrations of alpha-defensins could indicate the presence of microbial agents [93,94]. Table 1 highlights all studies on microbial agents and antimicrobial therapy in PF.

CONCLUSION

Evidence from the literature show that infectious pathogens exist in some cases of PF, can induce and exacerbate the disease, and antimicrobials may be useful in improving the quality of life of patients with PF. Experimental animal models of PF show that infectious agents play a role in the initiation of PF, however, whether this is true in patients is yet to be reported. The possibility of utilizing antimicrobials as a potential therapeutic strategy administered early in the course of the disease to improve the quality of life needs to be critically explored. More studies need to be designed and executed with a larger population size to investigate the efficacy of antimicrobials to either halt or to slow the progression of PF.

Taken together these studies provide evidence to show that infectious agents can either induce or exacerbate PF, however, additional studies to assess the viability of infectious agents after induction of PF will delineate the role of antimicrobials in the treatment of PF.

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