A Middle East systematic review and meta-analysis of prevalence and antibiotic susceptibility pattern in MRSA *Staphylococcus aureus* isolated from patients with cystic fibrosis

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**Abstract**

**Background:** This study aimed to determine the prevalence and antibiotic resistance patterns in *Staphylococcus aureus* isolated from patients with cystic fibrosis in Middle Eastern countries.

**Methods:** A systematic search was conducted in the PubMed, Web of Science (ISI), and Scopus databases for studies presenting the prevalence of MRSA strains, antibiotic resistance pattern in *S. aureus* strains isolated from patients who suffered from cystic fibrosis in Middle Eastern countries from 1999 to 10 June 2020. The following terms were used; prevalence, antibiotic resistance, antimicrobial drug resistance, drug resistance, *Staphylococcus aureus*, *S. aureus*, *Methicillin-resistant Staphylococcus aureus*, MRSA, cystic fibrosis, CF, and the Middle East. The meta-analysis was performed using Comprehensive Meta-analysis software (Version 3.3.070).

**Results:** Patients' age ranged from 1.6 to 18 years. Females were more than males. The prevalence of *S. aureus* was varied between 5.6 and 77.8%. The prevalence of *S. aureus* was varied between 5.6 and 77.8% in different countries. The combined prevalence of *S. aureus* in Middle East countries from 1999 to 2020 was reported by 40.9% (95% CI 29.6–53.1). The pooled prevalence of MRSA was reported at 18.6% (95% CI 1.1–82.6), $Z = 0.9$, $I^2 = 98.6$, $Q = 146.7$. The highest combined resistance in *S. aureus* strains was reported to Penicillin G (94%), followed by Ciprofloxacin (54.9%).

**Conclusion:** Regarding a quite prevalence of *S. aureus* and an intermediate prevalence of MRSA in CF patients, preventive measures and health policies should be implemented in the Middle East area to prevent the spread of infections caused by MRSA strains in CF patients.

**Keywords:** Cystic fibrosis, Patient, Middle East, *Staphylococcus aureus*, Antibiotic resistance

**Background**

Cystic fibrosis (CF) is an inherited multisystem dysfunction determined by abnormalities in exocrine gland function. A mutation in the cystic fibrosis conductance regulator (CFTR) gene sited on chromosome 7 in humans is the cause of cystic fibrosis (CF) [1]. Chronic sinopulmonary disease, pancreatic exocrine impairment, elevated sweat chloride, and male infertility are common disorders that resulted from CF [2]. Cystic fibrosis is the most frequent, lethal, genetic mutation which affects more than 70,000 people globally [3, 4]. In recent decades, advancements in disease management and new therapeutic developments have
led to improved patients’ survival, with surviving into the late thirties, as around 50% of all persons with CF are now 18 years of age or older [2].

Significant advances have been made in the lives of people with CF over the past six decades, which was once a life-threatening disease of infants and young children. However, life expectancy for people with CF has gradually increased. However, the disease still affects the survival and quality of people’s lives and imposes a heavy health burden on patients and their families [5]. According to reports, most patients with CF acquire pathogens from the environment, especially medical settings when they stay in these centers for a long time [1].

Staphylococcus aureus and Pseudomonas aeruginosa, Achromobacter xylosoxidans, Burkholderia cepacia complex, and Stenotrophomonas maltophilia are the most common isolates retrieved from CF patients [6]. S. aureus is one of the first respiratory colonizers in people with CF [7]. This microorganism causes opportunistic infections in people with underlying diseases such as CF [8, 9]. MRSA; a type of bacteria that causes an infection that does not respond to common antibiotics, including methicillin, amoxicillin, and penicillin as opposed to methicillin-susceptible Staphylococcus aureus (MSSA) [10]. The findings suggest that not only is it difficult to treat infections caused by MRSA strains, but it is also now known that MRSA infection may exacerbate lung function [11, 12].

The Middle East region is a vast area with several countries of different ethnicities, races, cultures, and climatic diversity [13]. The local prevalence in countries in the Middle East area is changing, due to the outline of new strains with the intercontinental exchange of several clones [14]. Several studies from this region reported a prevalence rate between 10 and 35% [15–17]. MRSA is endemic in this district, and this causes an increase in the risk of domestic and global transmission [14].

Therefore, due to the widespread prevalence of S. aureus in cystic fibrosis patients, the determination of the prevalence of this bacterium and its antibiotic resistance pattern is of special importance. So, this study aimed to determine the prevalence of MRSA strains, antibiotic resistance patterns in S. aureus strains isolated from patients with cystic fibrosis in Middle Eastern countries.

**Material and methods**

**Literature search**

A systematic search was conducted in PubMed, Web of Science (ISI), and Scopus databases for studies presenting the prevalence of MRSA strains, antibiotic resistance pattern in S. aureus strains isolated from patients with CF in some Middle Eastern countries from 1999 to 10 June 2020. The following search terms were used: prevalence, antibiotic resistance, antimicrobial drug resistance, drug resistance, Staphylococcus aureus, S. aureus, Methicillin-resistant Staphylococcus aureus, MRSA, cystic fibrosis, CF, and the Middle East, Iran, Palestine, Kuwait, Qatar, Emirate United Arab, Saudi Arabia, Lebanon, Turkey, Yemen, Oman, Bahrain, Egypt, Iraq, Jordan, Syria, and Cyprus.

Search strategy in PubMed was as follows; (prevalence [MeSH Terms]) AND (Drug Resistance [MeSH Terms] OR Antimicrobial Drug Resistance [MeSH Terms] OR Antibiotic Resistance [MeSH Terms]) AND (Staphylococcus aureus [MeSH Terms] OR S. aureus [MeSH Terms]), AND (Methicillin-resistant Staphylococcus aureus [MeSH Terms] OR MRSA [MeSH Terms]), AND (cystic fibrosis [MeSH Terms] OR CF [Title/Abstract]), AND (Middle East [MeSH Terms]). The bibliographic section of pertinent studies was also hand-searched to recognize further potentially eligible articles. Articles published in languages other than English were not evaluated.

**Inclusion and exclusion criteria**

Study inclusion criteria were cross-sectional and cohort studies that presented the prevalence of S. aureus, antibiotic resistance pattern, MRSA prevalence in CF patients from the Middle East were included. Also, articles that use standardized tests to determine antibiotic susceptibility were included. Studies before 1999, studies other than Middle East countries, studies with missed data, unclear data, abstracts, conferences, case reports, editorials, meetings, and reviews were excluded. As well, studies were written in languages other than English and also published before 1999 were deleted.

**Quality assessment**

Selection bias was assessed with the Critical Appraisal Skills Programme (CASP) checklist for cross-sectional studies (www.casp-uk.net). For each study, 10 questions were asked. So, the questions that were answered "yes" were given a score of 1, and if there was "no answer or doubts," the score was 0. According to this scoring system, studies were divided into three categories: poor (1–4), intermediate (6–8), and strong (>8). In the end, poor studies were eliminated (Additional file 1).

**Data extraction**

Two investigators independently reviewed the selected articles and extracted the pertinent data regarding the characteristics of each study. Data were first author, location (country), study's time, publication time, sample size, age, genus (male, female), and cystic fibrosis prevalence (n).
Statistical data analysis
Meta-analysis was performed using Comprehensive meta-analysis software (Version 3.3.070). The prevalence was calculated by 95% confidence intervals (CIs). Statistical heterogeneity between studies was assessed using Cochran’s Q-test ($p < 0.05$ was defined to indicate the presence of heterogeneity) and the $I^2$ (for assessing the degree of heterogeneity). The random-effect model was used because there was significant statistical heterogeneity between the studies. Also, publication bias was checked by Egger’s regression asymmetry test and Funnel plot. As well, subgroup analysis was done for MRSA strains and antibiotic resistance patterns.

Results
Study selection and characteristics of included studies
A total of 1003 studies were recognized in the primary search. After assessing titles, abstracts and full-texts, 12 studies met eligibility criteria for inclusion (Fig. 1). The frequency of studies from different countries was as follows; Turkey ($n = 3$), Iran ($n = 3$), Qatar ($n = 4$), Egypt ($n = 1$), and the United Arab Emirates ($n = 1$). Patients’ age ranged from 1.6 to 18 years. Females were more than males. The most samples used were sputum, deep pharyngeal swabs, and Bronchoalveolar lavage (BAL) (Table 1).

Overall effects
As it is observed in Table 1 and Fig. 2, the prevalence of $S. aureus$ was varied between 5.6 and 77.8% in different countries included in this review. In general, the combined prevalence of $S. aureus$ in some Middle East countries from 1999 to 2020 was reported by 40.9% (95% CI 29.6–53.1). Findings from selected studies showed that apart from Egypt, there is no significant difference in the prevalence of $S. aureus$ in Middle Eastern countries and they are almost in the same range (Fig. 2). Also, comparing the prevalence of $S. aureus$ based on...
| First Author | Study time | Publication | Settings | sample size | s. aureus (n) | Genus (%) | Age | Samples types | MRSA (n) |
|--------------|------------|-------------|----------|-------------|---------------|-----------|-----|---------------|---------|
| Nobandegani [30] | 2011–12 | 2016 | Iran | 172 | 93 | 60 | 40 | 5.9 ± 0.43 years | Sputum samples, an oropharyngeal (OP) swab | 74 |
| Khanbabaee [31] | 2004–2010 | 2012 | Iran | 129 | 12 | 60.5 | 39.5 | 75.57 (±6.2.48) months | Sputum samples or pharyngeal swabs | – |
| Aghamohammadi [32] | 2014–2015 | 2018 | Iran | 174 | 28 | 59 | 41 | 7.1 ± 5.7 y | – | – |
| Wahab [33] | – | 2014 | Qatar | 26 | 11 | 54 | 46 | Median 15.5 years | Sputum, deep pharyngeal swabs, BAL | – |
| Thomas [34] | 2010–2017 | 2019 | Qatar | 23 | 15 | – | – | – | – |
| Elshafie [35] | 2002–2003 | 2007 | Qatar | 113 | 30 | 60.5 | 39.5 | 10 years (1.6–18 years) | Deep oropharyngeal swabs, sputum | – |
| Wahab [20] | 2002–2003 | 2004 | Qatar | 36 | 28 | 60.5 | 39.5 | 10 years (1.6–18 years) | Sputum or oropharyngeal samples | – |
| Pakasticali [36] | 2013–2014 | 2016 | Turkey | 84 | 60 | 60.7 | 39.3 | 11.5 years | Throat specimen, sputum BAL | 2 |
| Yurdakul [37] | 2003–2010 | 2012 | Turkey | 604 | 325 | 37.5 | 62.5 | – | Deep throat swabs or sputum samples | 24 |
| Yagci [3] | 2007–2008 | 2013 | Turkey | 248 | 123 | 52 | 48 | 10.4 years (range: 1–58 years) | Sputum, deep throat swab | – |
| Frossard [38] | 1994–1996 | 1999 | UAE | 15 | 7 | 40 | 60 | 5.4–3.5 and 1.0–1.1 years | – | – |
| El-Falaki [39] | 2010–2012 | 2014 | Egypt | 36 | 2 | 61 | 39 | 3.91 ± 4.18 years | Sputum | – |

UAE United Arab Emirates
the year of study showed that the prevalence of this microorganism hasn't changed much since the beginning (1999) until now (2020), and its isolation from cystic fibrosis patients has been constant (Table 1).

Heterogeneity and publication bias
Regarding data achieved in the present study, heterogeneity was seen (Q = 185.9 and Z = 1.4, and I² = 94). Visual survey of Funnel plot showed the bias in the studies. Egger's linear regression test was performed to further investigate this subject. According to the findings, there was no publication bias among studies due to p = 0.31 (Fig. 3).
Subgroup analysis

Prevalence of MRSA

As summarized in Table 2, the pooled prevalence of MRSA strains was reported at 18.6% (95% CI 1.1–82.6), \( Z = 0.9, I^2 = 98.6, Q = 146.7 \).

Antibiotic resistance pattern

The pooled prevalence of resistance for each antibiotic was calculated. All isolates were resistant against Cefoxitin and Oxacillin, that’s why they all were MRSA. The highest combined resistance of *S. aureus* strains was reported against Penicillin G, followed by Ciprofloxacin with resistance rates 94% (95% CI 70.1–99.1), and 54.9% (95% CI 2.5–98.3), respectively. All strains displayed susceptibility to Tigecycline, Clarithromycin, Telithromycin, Teicoplanin, Linezolid, and Vancomycin.

Discussion

Bacteria as the most important pathogens still play a significant role in aggravating lung complications in patients with cystic fibrosis [18]. Many organisms isolated from the sputum of CF patients are normal flora of the nose (*S. aureus*) or opportunistic pathogens such as *P. aeruginosa* which is a common environmental organism [19]. Due to the importance of *S. aureus* especially MRSA isolates in CF patients, in the current review, the prevalence of this microorganism and its antibiotic resistance pattern was investigated. In our study, the prevalence of *S. aureus* in different countries varied between 5.6 and 77.8%. In general, the combined prevalence of *S. aureus* in some Middle East countries from 1999 to 2020 was reported by 40.9%. Also, the pooled prevalence of MRSA strains was reported at 18.6%. So, based on data obtained from the current meta-analysis, *S. aureus* was isolated at a high rate from CF patients. Finding *S. aureus* in the lower respiratory tract certainly indicates a pathological situation that has never been adequately investigated [20, 21]. Our results are in accordance with reports from the USA, where the US Patient Registry Annual Data Report presented the prevalence of *S. aureus* 70.3% in CF patients [22]. Also, accordingly, the report of the prevalence among European countries was varied between 60 and 75% [23–25].

According to the results of included studies, there is no significant difference in the prevalence of *S. aureus* in Middle Eastern countries except Egypt and they all are almost in the same range. Also, comparing the prevalence of *S. aureus* based on the year of study showed that the prevalence of this microorganism

| Table 2 | Overall effects and subgroup analysis for antibiotic resistance pattern in *S. aureus* recovered from CF patients |
| --- | --- |
| **Subgroups** | **Number of studies** | **Heterogeneity test** | **Egger’s test** | **Random model** |
| | | **Prevalence (95% CI) (%)** | **Z** | **P** | **Q** | **P** | **I^2** | **T** | **P** |
| *S. aureus* | 12 | 40.9% (29.6–53.1) | 1.4 | 0.1 | 185.9 | 0.00 | 94 | 1 | 0.31 |
| **Subgroup analysis** | | | | | | | | | |
| MRSA | 3 | 18.6% (1.1–82.6) | 0.9 | 0.3 | 146.7 | 0.00 | 98.6 | 0.07 | 0.95 |
| **Subgroup analysis for antibiotic resistance pattern** | | | | | | | | | |
| Amikacin | 3 | 21.5 (12.2–38.3) | 2.3 | 0.00 | 23 | 0.00 | 86 | 1 | 0.34 |
| Cephaloxin | 3 | 12.3 (3.1–22.4) | 3.1 | 0.21 | 44.1 | 0.00 | 73 | 0.1 | 0.22 |
| Chloramphenicol | 3 | 9.7 (3.3–32.2) | 21 | 0.00 | 117 | 0.00 | 89 | 2.3 | 0.11 |
| Ciprofloxacin | 4 | 54.9 (2.5–98.3) | 0.1 | 0.9 | 33.9 | 0.00 | 97 | 0.12 | 0.33 |
| Clindamycin | 4 | 17.9 (12.6–24.9) | 7.1 | 0.00 | 1.2 | 0.00 | 19.8 | 0.12 | 0.14 |
| Gentamycin | 4 | 10.8 (6.2–18.3) | 6.7 | 0.00 | 2 | 0.15 | 51.6 | 3.2 | 0.2 |
| Linezolid | 5 | 0 (0.1–1.2) | 1 | 0.00 | 2 | 1 | 0.00 | 0.01 | 0.1 |
| Vancomycin | 5 | 0 (0.1–1.2) | 1 | 0.00 | 2.1 | 1 | 0.00 | 0.01 | 0.1 |
| Trimethoprim-Sulfamethoxazole | 5 | 17 (8.6–30.9) | 3.9 | 0.00 | 5.4 | 0.06 | 63.4 | 5.1 | 0.12 |
| Penicillin G | 4 | 94 (70.1–99.1) | 2.8 | 0.005 | 5.7 | 0.016 | 82.6 | 0.1 | 0.21 |
| Tetraacycline | 4 | 22 (8.6–46) | 2.2 | 0.025 | 6.6 | 0.01 | 84.8 | 1.3 | 0.01 |
| Teicoplanin | 3 | 0 (0.1–1.2) | 1 | 0.00 | 2 | 1 | 0.00 | 0.01 | 0.1 |
| Telithromycin | 3 | 0 (0.1–1.2) | 1 | 0.00 | 2 | 1 | 0.00 | 0.01 | 0.1 |
| Cefoxitin | 3 | 100 (99.9–100) | 1 | 0.001 | 3.6 | 0.001 | 89.3 | 0.1 | 0.32 |
| Clarithromycin | 3 | 0 (0.1–1.2) | 1 | 0.00 | 2 | 1 | 0.00 | 0.01 | 0.1 |
| Oxacillin | 3 | 100 (99.9–100) | 1 | 0.001 | 3.6 | 0.001 | 88.2 | 0.1 | 0.32 |
| Tigecycline | 3 | 0 (0.1–1.2) | 1 | 0.00 | 2 | 1 | 0.00 | 0.01 | 0.1 |
has not changed much since the beginning (1999) until now (2020), and its isolation from cystic fibrosis patients has been constant. The stability of prevalence of *S. aureus* is possibly owing to improved awareness and infection prevention and control strategies [22]. If we look at it from another angle, the lack of new molecular techniques and the constant use of phenotypic methods or the lack of proper health policies in this area may have caused the prevalence of these microorganisms in these patients over the years to be almost constant which this requires a change in health policy attitudes. We reported the prevalence rate about of 18.6% of MRSA, similarly, USA Patient Registry Annual Data Report showed a prevalence rate of around 25% for MRSA [22], and Argentina (25.9%) [26]. But in contrast to our study, this rate was lower in Poland (6%) [27] and Germany (4%) [28]. We surely know MRSA strains will bring up more problems for patients with CF [2].

In the present systematic review and meta-analysis, the highest combined resistance in *S. aureus* strains was reported against Penicillin G, followed by Ciprofloxacin with resistance rates of 94%, and 54.9%, respectively. Therefore, according to results achieved, most antibiotics used in included studies except Penicillin G and Ciprofloxacin were effective against *S. aureus* isolates. This must be taken into consideration in line with our results, a study conducted by Cafiso et al. All strains were susceptible to Linezolid and Tigecycline [8]. Regarding the identification of the clinical significance of *S. aureus* pulmonary infection particularly MRSA in CF and the restricted data available to guide present therapeutic regimes [2, 29], in the current review, most antibiotics especially Tigecycline, Clarithromycin, Telithromycin, Teicoplanin, Linezolid, and Vancomycin showed a good impact on *S. aureus* isolates. Therefore, it can use the mentioned antibiotics in MRSA pulmonary infection in CF patients in the Middle East area.

The main limitation of the present study is that we search only studies published in English and other languages such as Persian, Turkish, and Arabic that didn’t include. Also, the sample size used here was small and the number of studies enrolled unfortunately did not cover all Middle Eastern countries.

In summary, our review reported a high prevalence of *S. aureus* and an intermediate prevalence of MRSA strains in pulmonary specimens achieved from CF patients. The prevalence in most of the countries included in this study was almost the same and showed a steady trend within a few years from 1999 to 2020. Due to these facts, it is necessary to use new molecular methods to identify this microorganism.

**Conclusion**

Preventive measures and health policies should be implemented in the Middle East region to prevent the spread of infection caused by this microorganism, specifically MRSA in CF patients.

**Abbreviations**

ISI: Web of Science; CF: Cystic fibrosis; CFTR: Cystic fibrosis conductance regulator; MSSA: Methicillin-susceptible staph; CASP: Critical Appraisal Skills Programme; CI: Confidence intervals.

**Supplementary Information**

The online version contains supplementary material available at https://doi.org/10.1186/s41043-022-00305-x.

**Additional file 1.** PRISMA 2020 Checklist for reporting systematic reviews.

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**Author contributions**

YN and AFA conducted data extraction, analysis, and wrote the first draft of the paper; PA and MRT conducted data Extraction; PS, EN and AK designed the study, analyses, and drafting of the article. All authors contributed to discuss results and writing the manuscript. All authors read and approved the final manuscript.

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**Availability of data and materials**

All data used and analyzed during this study are based on an original article and is available from the corresponding author, by request.

**Declarations**

**Ethics approval and consent to participate**

Not applicable.

**Consent for publication**

Not applicable.

**Competing interests**

The authors declare that they have no competing interests.

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**References**

1. Sousa AM, Pereira MO. *Pseudomonas aeruginosa* diversification during infection development in cystic fibrosis lungs-a review. Pathogens. 2014;3(3):680–703. https://doi.org/10.3390/pathogens3030680.
2. Jennings MT, Riekert KA, Boyle MP. Update on key emerging challenges in cystic fibrosis. Med Princ Pract. 2014;23(3):393–402. https://doi.org/10.1159/000357646.

3. Bernardy EE, Petit RA, Moller AG, McAdam AJ, Priebe GP, et al. Whole-genome sequences of Staphylococcus aureus isolates from cystic fibrosis lung infections. Microbiol Resour Announc. 2019;3(8):e01564–e1618. https://doi.org/10.1128/MRA.01564-18.

4. Lavelle GM, White MM, Browne N, McElvaney NG, Reeves EP. Animal models of cystic fibrosis pathology: phenotypic parallels and divergences. BioMed Res Int. 2016. https://doi.org/10.1155/2016/5258727.

5. Yagci S, Hasicelik G, Dogru D, Ozcelik U, Sener B. Prevalence and genetic diversity of Staphylococcus aureus small-colony variants in cystic fibrosis patients. Clin Microbiol Infect. 2013;19(1):77–84. https://doi.org/10.1111/j.1469-0691.2013.03742.x.

6. Stone A, Saiman L. Update on the epidemiology and management of Staphylococcus aureus, including methicillin-resistant Staphylococcus aureus. In patients with cystic fibrosis. Curr Opin Pulm Med. 2007;13(3):515–21. https://doi.org/10.1097/MCP.0b013e3282efbbac.

7. Cafiso V, Bertuccio T, Spina D, Campanile F, Bongiorno D, Santagati M, et al. Methicillin resistance and vancomycin heteroresistance in Staphylococcus aureus in cystic fibrosis patients. Eur J Clin Microbiol Infect Dis. 2010;29(12):1277–85. https://doi.org/10.1007/s10096-010-1000-5.

8. Goodrich JS, Sutton-Shields TN, Kerr A, Wedd JP, Miller MB, Gilligan CM. Acquisition and adaptation of the airway microbiota in the early life of cystic fibrosis patients. J Cyst Fibros. 2011;10(1):122–8. https://doi.org/10.1016/j.jcf.2010.11.014.

9. Thomas M, Raja M, Albakri M, Najim M, Chandra P, Allangawi M. CT score and its correlation with lung function and microbiology of adult patients with cystic fibrosis with predominant 11.234V genotype in Qatar. Qatar Med J. 2020. https://doi.org/10.5539/qmj.2020.4.

10. Elshafie SS, Wahab AA, Al Janahi I. Antimicrobial resistance of bacterial strains isolated from respiratory tract of cystic fibrosis patients with CFTR 11243V mutation. J Pediatr Infect Dis. 2007;2(12):039–43.

11. Frossard P, Hertecant J, Bossaert Y, Dawson K. Genotype-phenotype characterization of Panton-Valentine leukocidin-positive community-acquired methicillin-resistant Staphylococcus aureus strains isolated from respiratory tract of cystic fibrosis patients with CFTR 11243V mutation. J Pediatr Infect Dis. 2012;2(16):122–8.

12. El-Shafie SS, Wahab AA, Al Janahi I. Antimicrobial resistance of bacterial strains isolated from respiratory tract of cystic fibrosis patients with CFTR 11243V mutation. J Infect Dis. 2012;206(12):1941–9. https://doi.org/10.1093/infdis/jis535.

13. Khaledi A, Khademi F, Esmaeili E, Esmaeili S, Rostami H. The role of HPaA protein as candidate vaccine against Helicobacter pylori. Der Pharma Chem. 2016;6(3):235–7.

14. Alexander B, Petren E, Rizvi S, Fink A, Ostrenga J, Sewall A, et al. Cystic fibrosis. Lancet Respir Med. 2014;2(10):999–1006. https://doi.org/10.1016/S2213-2600(14)00337-6.

15. Gemmell CG, Edwards DI, Fraise AP, Gould FK, Ridgway GL, Warren RE. National survey of molecular epidemiology of Staphylococcus aureus colonization in Belgian cystic fibrosis patients. J Antimicrob Chemother. 2007;5(59):893–9. https://doi.org/10.1093/jac/dkm037.

16. Desmet K, Ortuño-Ortín I, Wacziarg R. Culture, ethnicity, and diversity. Clin Microbiol Rev. 2002;2(15):194–222. https://doi.org/10.1128/CMR.15.2.194-222.2000.

17. Khaledi A, Khademi F, Esmaeili E, Esmaeili S, Rostami H. The role of HPaA protein as candidate vaccine against Helicobacter pylori. Der Pharma Chem. 2016;6(3):235–7.

18. Goodrich JS, Sutton-Shields TN, Kerr A, Wedd JP, Miller MB, Gilligan CM. Acquisition and adaptation of the airway microbiota in the early life of cystic fibrosis patients. J Cyst Fibros. 2011;10(1):122–8. https://doi.org/10.1016/j.jcf.2010.11.014.

19. Marshall B, Fink A, Loeffler D, Elbert A, O’Neill T, Rush T, et al. Patient registry annual data report. Cystic Fibrosis Foundation; 2016.

20. Wahab AA, Al Janahi I. Antimicrobial resistance of bacterial strains isolated from respiratory tract of cystic fibrosis patients with CFTR 11243V mutation. J Pediatr Infect Dis. 2007;2(12):039–43.

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