Review

Therapeutic Potential of Leaves from *Fridericia chica* (Bonpl.) L. G. Lohmann: Botanical Aspects, Phytochemical and Biological, Anti-Inflammatory, Antioxidant and Healing Action

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Abstract: Plants of the species *Fridericia chica* (Bonpl.) L. G. Lohmann (Bignoniaceae), which are widely distributed in Brazil and named crajiru in the state of Amazonas, are known in folk medicine as a traditional medicine in the form of a tea for the treatment of intestinal colic, diarrhea, and anemia, among other diseases. The chemical analysis of extracts of the leaves has identified phenolic compounds, a class of secondary metabolites that provide defense for plants and benefits to the health of humans. Several studies have shown the therapeutic efficacy of *F. chica* extracts, with antitumor, antiviral, wound healing, anti-inflammatory, and antioxidant activities being among the therapeutic applications already proven. The healing action of *F. chica* leaf extract has been demonstrated in several experimental models, and shows the ability to favor the proliferation of fibroblasts, which is essential for tissue repair. The anti-inflammatory activity of *F. chica* has been clearly demonstrated by several authors, who suggest that it is related to the presence of 3-deoxyanthocyanidins, which is capable of inhibiting pro-inflammatory pathways such as the kappa B (NF-kB) nuclear transcription factor pathway. Another important effect attributed to this species is the antioxidant effect, attributed to phenolic compounds interrupting chain reactions caused by free radicals and donating hydrogen atoms or electrons. In conclusion, the species *Fridericia chica* has great therapeutic potential, which is detailed in this paper with the objective of encouraging new research and promoting the sum of efforts for the inclusion of herbal medicines in health systems around the world.

Keywords: *Fridericia chica*; phytochemistry; anti-inflammatory; antioxidant; wound healing

1. Introduction

*Fridericia chica* (Bonpl.) L. G. Lohmann (Figure 1) is a liana or creeper that belongs to the family Bignoniaceae, which comprises about 120 genera and 800 species [1]. The largest number of species of this family is found mainly in tropical and subtropical regions, with Brazil and the African continent as the two major centers of geographical distribution. In Brazil, the species of the family Bignoniaceae do not have a unique habitat and are widely distributed in the region of the legal Amazon, as well as throughout the southern region of the country as far down as Rio Grande do Sul. In addition, plants of this family can be found...
in the Cerrado and Atlantic Forest biomes [2–4]. *F. chica* was first described by Cronquist [5] as *Arrabidae chica*, belonging to the division Magnoliophyta, class Magnoliopsida, subclass Asteridae, order Scrophulariales, family bignoniaceae, and genus *Arrabidae*. However, its classification has been altered due to recent taxonomic changes, such as the inclusion of several species of the genus *Arrabidae* in the genus *Fridericia*. This change has affected the classification of its varieties *A. chica* var. *acutifolia* (DC.) Bureau, *A. chica* var. *angustifolia* Bureau & K. Schum, *A. chica* var. *cuprea* Bureau & K. Schum, *A. chica* var. *thyrsoidea* (DC.) Bureau and *A. chica* var. *viscida* Donn.Sm [6].

![Figure 1](image-url)

**Figure 1.** *Fridericia chica* (Bonpl.) L. G. Lohmann: specimen and presentations forms. (A) Arboreal specimen of adult *F. chica* in situ in the city of Manaus, Amazonas Brazil. (B) Dried leaves of *F. chica* as marketed in the city of Manaus, Brazil. (C) Tea obtained from the dried leaves of *F. chica* and its main compounds responsible for the characteristic color. Photos: Author.

Botanically, *Fridericia chica* are described as woody, shrub-like, or arboreal plants, as well as climbing plants, with that leaves measure between 18–20 cm in length when mature. They have opposite crossed leaves, compound bi- or trifoliate, however, with the terminal foliole modified in tendrils in the elevated part of the branches. The folioles have an oblong-lanceolate shape, cartaceous, with an obtuse base, acute apex and herbaceous consistency. The surface of the folioles is smooth, and the rib is of the peninerveal type, i.e., the secondary ribs branch from the main rib [3,7–9]. The flowers are campanulate, resemble the shape of a bell, and have pink or violet coloration [10,11].

*F. chica* occurs in Central American countries, the Caribbean, and mainly in South American countries, such as Guyana and French Guiana, and especially in Brazil. In the latter it has wide geographical distribution, with confirmed occurrence in all regions and phytogeographic domains, including the Amazon, the Caatinga, the Cerrado, the Atlantic Forest, and the Pantanal biomes. With regard to the type of vegetation, *F. chica* can be found in floodplains or forests of various types, such as riparian, flooded forests, and terra firma, among others [6].

### 2. Revision Strategy

An extensive literature review was carried out using different scientific electronic sources, including databases such as Scifinder, Pubmed, Scopus, Web of Science, and Google Scholar. The study databases included original papers published in peer reviewed journals, books, dissertations, and theses, and all data of scientific interest written or translated into English published prior to July 2022 was considered. The keywords “*Arrabidae chica*” and “*Fridericia chica*” alone and in combination with “inflammation”, “inflammatory”, “antioxidant”, “wound healing”, and “healing”. In addition, the names of all phytochemical compounds were used in the search.

### 3. Ethnobotany

In Brazil, *F. chica* is popularly known as cajuru, carajuru, carajunu, carajuru, chica, china, cipó-cruz, cipó-pau, coá-piranga, cajiru, cajiru, cuica, guajuru, guajuru-piranga,
guarajuru, oajuru, oajuru-piranga, paripari, pariri, and piranga, among others [2,6,10]. Its mature leaves, collected from plants ranging from 50 cm to 2 m, are used in the form of tea (dried form), which has an astringent function and can be used against intestinal colic, diarrhea, anemia, uterine inflammation, hemorrhages, leukemia, jaundice, albuminuria, for vaginal hygiene via bathing, or in the form of tincture for topical use directly on skin lesions or even in ointments, creams [10,12], and in the form of soap with an antiacne effect [13]. It is used as an anti-inflammatory, antioxidant, antidiabetic medicine, and a disinfectant [3,10,14,15].

4. Chemical Composition

Chemically, *F. chica* is rich in polyphenols, especially flavonoids. Among the flavonoids, derivatives of the class of anthocyanins stand out, which in this plant are abundant in the form of 3-deoxyanthocyanins. These are involved in plant growth and development, including UV protection, stimulation of pollination and seed dispersal, and as a defense mechanism [16]. The 3-deoxyanthocyanins are the substances that give the tea its red color, especially the main compounds carajurin (1) and carajurone (2) [17], and to a lesser extent four other substances, 3’-hydroxy-carajurone (3), 3’-hydroxy-carajurin (4), 6,7,3’,4’-tetrahydroxy-5-methoxyflavylum (5), and 6,7,4’-trihydroxy-5-methoxyflavylum (6) [18]. Substances 1 and 2 were the first to be described in this species.

In addition to anthocyanidins, other flavonoids (aglycones and glycosides) have been isolated and/or characterized using high-performance liquid chromatography coupled to mass spectrometry (HPLC-MS) analysis, among them the flavones carajuflavone (7), luteolin (8) [13], chrysoeriol (9) [19], 4’-hydroxy-3,7-dimethoxyflavone (10) [20], 5,7-dimethoxy-4′-hydroxyflavone (11) [19], acacetin (12) [17], isoscutellarein (13), scutellarein (14), 6-hydroxyluteolin (15), hispidulin (16), apigenin (17) [21], thevetiaflavone (18) [13], cirsimarin (19) [19], apigenin 7-glucuronide (20), scutellarin (21) [22], chrysoeriol-O-glucoside (22) [19]. Flavonols are a more restricted group, being so far characterized as quercetin-O-gallate (23), kaempferol (24), isorhamnetin (25) hyperin 6”-gallate (26), quercetin-O-glucoside (27), and isorhamnetin-3-O-glucoside (28) [19]. The biflavonoid amentoflavone (29) and the flavans-3-ols catechin (30) and epicatechin (31) [19] are other flavonoid derivatives identified in *F. chica* (Figure 2).

With respect to other classes of natural products, the characterization of terpenes, including carotenoids, and other classes such as fatty acids and their derivatives, tocopherols and alkaloids, stands out. The diterpene phytol (32), the triterpene squalene (33), and the steroidal β-sitosterol (34) [23] are described as structures of a terpenic nature of up to 30 carbons described in *F. chica*. On the other hand, a number of tetraterpenes have been identified using HPLC-MS, among them the main compounds β-carotene (35), (all-E)-zeaxanthin (36), (all-E)-lutein (37), (all-E)-violaxathin (38), (13Z)-violaxathin (39), and (all-E)-luteoxanthin (40) [22]. In addition to antioxidant carotenoids, tocochromanol α-tocopherol (41) has been described as well [23]. Additionally, the identification of the fatty acid ethyl ester ethyl palmitate (42) has been reported [23], as has the alkaloid pheophorbide α (43) [24]. In Table 1, the 43 compounds already identified in *F. chica* and their biological activities already studied from these molecules alone (isolated from *F. chica* extract or other sources) are shown. It is worth noting that despite the tea being the most popular use of *F. chica*, phytochemical studies were focused on hydroalcoholic and/or organic extracts, possibly aiming for the enhancement of extraction yields. Moreover, tea preparations may contain the most polar compounds, specifically the positively charged 3-deoxyanthocyanins, information that is corroborated by the bright red color of the dried leaf infusions. Thus, the composition of the extract is of great importance, as these compounds can exert pharmacological effects with summative, potentiating or deleterious characteristics (Figure 3).
| Compound No. | Name                      | Source               | Isolation/Detection | Biological Activity * |
|--------------|---------------------------|----------------------|---------------------|-----------------------|
| **Anthocyanins** |                           |                      |                     |                       |
| 1            | Carajurin                 | leaves, MeOH         | Isolation [17]      | Anti-inflammatory [17] |
| 2            | Carajurone                | leaves, MeOH         | Isolation [17]      | N.A.                  |
| 3            | 3’-Hydroxy-carajurone     | leaves, MeOH         | Isolation [17]      | N.A.                  |
| 4            | 3’-Hydroxy-carajurone     | leaves, MeOH         | Isolation [17]      | N.A.                  |
| 5            | 6,7,3’,4’-Tetrahydroxy-5’-methoxyflavilium | leaves, DCM fraction | Isolation [18]      | N.A.                  |
| 6            | 6,7,4’-Trihydroxy-5’-methoxyflavilium | leaves, DCM fraction | Isolation [18]      | N.A.                  |
| **Flavones** |                           |                      |                     |                       |
| 7            | Carajuflavone             | leaves, AcOEt fraction | Isolation [1]      | N.A.                  |
| 8            | Luteolin                  | leaves, AcOEt fraction | Isolation [1]      | Anti-inflammatory [25] |
|              |                           |                      | HPLC-MS detection   | Anti-inflammatory [27] |
| 9            | Chrysoeriol               | leaves, 70% EtOH     | HPLC-MS detection   | Anti-inflammatory [26] |
| 10           | 4’-Hydroxy-3,7-dimethoxyflavone | leaves, EtOH         | Isolation [20]      | N.A.                  |
| 11           | 5’,7-dimethoxy-4’-hydroxyflavone | leaves, 70% EtOH     | HPLC-MS detection   | N.A.                  |

Figure 2. Polyphenols (1–31) of *F. chica*.

Table 1. Phytochemical components of *F. chica* extracts and their biological activities.

1. **Anthocyanins**
2. **Flavones**
| Compound No. | Name               | Source                 | Isolation/Detection                  | Biological Activity * |
|-------------|--------------------|------------------------|--------------------------------------|-----------------------|
| 12          | Acacetin           | leaves, MeOH           | Isolation [17]                       | Anti-inflammatory [29]|
|             |                    |                        |                                      | Antioxidant [30]      |
| 13          | Isoscutellarein    | leaves, 90% EtOH       | HPLC-MS detection [21]               | Antioxidant [31]      |
| 14          | Scutellarein       | leaves, 90% EtOH       | HPLC-MS detection, isolation [21]   | Anti-inflammatory [32,33]|
|             |                    |                        |                                      | Antioxidant [34]      |
| 15          | 6-Hydroxyluteolin  | leaves, 90% EtOH       | HPLC-MS detection [21]               | N.A.                  |
| 16          | Hispidulin         | leaves, 90% EtOH       | HPLC-MS detection [21]               | Anti-inflammatory [35,36]|
|             |                    |                        |                                      | Antioxidant [37]      |
| 17          | Apigenin           | leaves, 90% EtOH       | HPLC-MS detection, isolation [21]   | Anti-inflammatory [38,39]|
|             |                    |                        |                                      | Antioxidant [40]      |
|             |                    |                        |                                      | Healing [41]          |
| 18          | Thevetiaflavone    | leaves, AcOEt fraction | Isolation [1]                        | Antioxidant [42]      |
| 19          | Cirsimarin         | leaves, 70% EtOH       | HPLC-MS detection [19]               | Anti-inflammatory [43]|
|             |                    |                        |                                      | Antioxidant [44]      |
|             |                    |                        |                                      | Healing [45]          |
| 20          | Apigenin 7-glucuronide | leaves, 80% MeOH     | HPLC-MS detection [22]               | Anti-inflammatory [46]|
|             |                    |                        |                                      | Antioxidant [47]      |
| 21          | Scutellarin        | leaves, 80% MeOH       | HPLC-MS detection [22]               | Anti-inflammatory [48]|
|             |                    |                        |                                      | Antioxidant [34]      |
|             |                    |                        |                                      | Healing [49]          |
| 22          | Chrysoeriol-O-glucoside | leaves, 70% EtOH     | HPLC-MS detection                   | N.A.                  |

**Flavonols**

| 23          | Quercetin-O-gallate | leaves, 70% EtOH       | HPLC-MS detection [19]               | N.A.                  |
| 24          | Kaempferol          | leaves, 70% EtOH       | HPLC-MS detection [19]               | Anti-inflammatory [50,51]|
|             |                    |                        |                                      | Antioxidant [31]      |
|             |                    |                        |                                      | Healing [52]          |
| 25          | Isorhamnetin       | leaves, 70% EtOH       | HPLC-MS detection [19]               | Anti-inflammatory [53]|
|             |                    |                        |                                      | Antioxidant [54]      |
| 26          | Hyperin 6″-gallate  | leaves, 70% EtOH       | HPLC-MS detection [19]               | Anti-inflammatory [55]|
|             |                    |                        |                                      | Antioxidant [55]      |
| 27          | Quercetin-O-glucoside | leaves, 70% EtOH     | HPLC-MS detection [19]               | Antioxidant [56]      |
| 28          | Isorhamnetin-3-O-glucoside | leaves, 70% EtOH     | HPLC-MS detection [19]               | Antioxidant [57]      |

**Flavone dimer**

| 29          | Amentoflavone      | leaves, 70% EtOH       | HPLC-MS detection [19]               | Anti-inflammatory [58]|
|             |                    |                        |                                      | Antioxidant [59]      |

**Flavan-3-ols**

| 30          | Catechin           | leaves, 70% EtOH       | HPLC-MS detection [19]               | Anti-inflammatory [60]|
|             |                    |                        |                                      | Antioxidant [61]      |
| 31          | Epicatechin        | leaves, 70% EtOH       | HPLC-MS detection [19]               | Anti-inflammatory [62]|
|             |                    |                        |                                      | Antioxidant [63]      |
| Compound No. | Name       | Source                   | Isolation/Detection       | Biological Activity * |
|-------------|------------|--------------------------|---------------------------|-----------------------|
| **Terpenes**|            |                          |                           |                       |
| 32          | Phytol     | leaves, 70% EtOH, hexane fraction | GC-MS detection [23]     | Anti-inflammation [64]  
|             |            |                          |                           | Antioxidant [65]       |
| 33          | Squalene   | leaves, 70% EtOH, hexane fraction | GC-MS detection [23]     | Antioxidant [66]       
|             |            |                          |                           | Anti-inflammation [67]  |
| 34          | β-Sitosterol| leaves, 70% EtOH, hexane fraction | GC-MS detection [23]     | Anti-inflammation [68]  |
| 35          | β-Carotene | leaves, acetone          | HPLC-MS detection [22]   | Anti-inflammation [69]  
|             |            |                          |                           | Antioxidant [69]       
|             |            |                          |                           | Healing [70]           |
| 36          | (all-E)-Zeaxanthin| leaves, acetone     | HPLC-MS detection [22]   | Anti-inflammation [71]  
|             |            |                          |                           | Antioxidant [72]       |
| 37          | (all-E)-Lutein| leaves, acetone       | HPLC-MS detection [22]   | Anti-inflammation [73]  
|             |            |                          |                           | Antioxidant [74]       |
| 38          | (all-E)-Violaxanthin| leaves, acetone  | HPLC-MS detection [22]   | Anti-inflammation [75]  
|             |            |                          |                           | Antioxidant [75]       |
| 39          | (13Z)-Violaxanthin| leaves, acetone | HPLC-MS detection [22]   | Antioxidant [76]        |
| 40          | (all-E)-Luteoxanthin| leaves, acetone | HPLC-MS detection [22]   | N.A.                   |
| **Tocochromanol** |      |                          |                           |                       |
| 41          | α-Tocopherol| leaves, 70% EtOH, hexane fraction | GC-MS detection [23]     | Antioxidant [77]       
|             |            |                          |                           | Healing [78]           |
| **Fatty acid** |            |                          |                           |                       |
| 42          | Ethyl palmitate | leaves, 70% EtOH, hexane fraction | GC-MS detection [23]     | Anti-inflammation [79]  |
| **Alkaloid** |            |                          |                           |                       |
| 43          | Pheophorbide a | leaves, 90% EtOH | Isolation [24]           | Anti-inflammation [80]  
|             |            |                          |                           | Antioxidant [81]       
|             |            |                          |                           | Healing [82]           |

* Biological activities concern anti-inflammatory, antioxidant, and healing activity described in previous articles using the isolated components obtained from *F. chica* or other plants. N.A., not available.
Figure 3. Terpenes (32–40), tocochromanol (41), fatty acid (42), and alkaloid (43) of *F. chica*.

5. Pharmacological Properties

Historically, plants have been an important source of active ingredients for drug development, and scientific research has been directed towards the prospect of new drugs based on natural products [83]. Thus, the connection between plants and health is responsible for the beginning of a new generation of therapy that integrates drugs derived from plants or the use of plants themselves or their parts [84]. The public health system in Brazil offers twelve herbal medicines as treatment options, among them several with anti-inflammatory potential, such as *Aloe vera*, *Schinus terebenthifolius*, and *Uncaria tomentosa* [85,86].

The use of plants as a medicine occurs in many traditional societies [87]; therefore, it is important to stimulate scientific studies that prove existing popular knowledge about plants and their effectiveness in the treatment of diseases [88]. In this sense, several studies have been carried out with different extracts of *F. chica* leaves in order to prove the effectiveness of the plant for the therapeutic purposes for which it is popularly used (Table 2).
Table 2. Pharmacological properties of extracts of *Fridericia chica*.

| Authors | Type of Study | Action | Etiological Agent | Plant Material/Part | Treatment |
|---------|---------------|--------|-------------------|---------------------|-----------|
| [20]    | In vitro study | Antifungal and antiprotozoal | Trichophyton mentagrophytes (fungus) and *Trypanosoma cruzi* | Ethanolic extract of leaves | 4 mg/mL (trypanocide); 3.125 mg/mL (fungicide) |
| [89]    | In vitro study | Antiprotozoal | *Leishmania amazonensis* and *Leishmania infantum* | Hexanic leaf extract | 37.2 µg/mL (*L. amazonensis*); 18.6 µg/mL (*L. infantum*) |
| [90]    | In vitro and in vivo study | Antiprotozoal and healing | *Leishmania amazonensis*; Swiss Webster mouse | Ethanolic extract of leaves and fractions | Leishmanicidal effect: 60–155.9 µg/mL; healing effect: 10 mg/g |
| [24]    | In vitro study | Antiprotozoal | *Trypanosoma cruzi* | Hydroethanolic extract of leaves and fractions | 24.8 µg/mL–213 µg/mL (hydroethanolic extract); 2.3 µg/mL–10 µg/mL (feoforbide-a) |
| [91]    | In vivo Study | Antimicrobial | *Helicobacter pylori* 43,504 and *Enterococcus faecalis* 29,212 | Hydroethanolic extract of leaves | 12.5 µg/mL (*H. pylori*); 100 µg/mL (*E. faecalis*) |
| [92]    | In vitro study | Antimicrobial | *Staphylococcus* sp. | Hydroalcoholic extract | 250 µg gallic acid equivalent (GAE)/mL–MIC; 1000 µg GAE/mL–MBC |
| [93]    | In vitro study | Antimicrobial | *Candida* sp. | Dichloromethane extract of leaves | 0.007–0.03 mg/mL |
| [94]    | In vitro study | Antiviral | aMPV cepa SHS/669/03 | Ethanolic extract of leaves | 2.5 µg/mL |
| [95]    | In vitro study | Antiviral | Human Herpes Virus type 1 (HHV-1); murine Encephalomyocarditis virus (EMCV); Vaccinia Virus strain Western Reserve (VACV-WR) | Ethanolic extract of leaves | EC₅₀: 245.7 µg/mL (*HHV-1*); 86.3 µg/mL (VACV-WR) |
| [96]    | In vivo study | Antitumor | Solid Ehrlich tumor | Ethanolic extract and aqueous extract of leaves | Oral administration for 16 weeks: extract at a dose of 300 mg/kg; 7.12-dimethyl-1,2-benzanthracene (DMBA) associated with vincristine 250 µg/mL |
| [97]    | In vivo study | Antitumor | 7,12-dimethyl injection-induced breast cancer-1,2-benzanthracene (DMBA) | Hydroalcoholic extract of leaves | |
| [98]    | In vitro study | Antitumor and fibroblast proliferation | Human tumor cell lines: MCF-7 (breast), NCI-ADR/RES (ovary with multiple drug resistance phenotype), UACC-62 (melanoma), NCI-hs60 (lung), PC-3 (prostate), HT29 (colon), OVCAR-05 (ovary), 786-O (kidney) and K562 (leukemia). Fibroblasts: obtained from 3T3 mice | Crude leaf extracts (without and with enzyme treatment) | 0.25; 2.5 and 25 µg/mL without enzymatic treatment (fibroblast proliferation); 7.4 and 8.7 µg/mL with enzymatic treatment (cytostatic effect for UACC-62–melanoma lineage) |
| [99]    | Survey of use by health professionals | Anti-inflammatory | Oral diseases | Leave | Tea (no dose determined) |
| [19]    | In vivo study | Analgesic and anti-inflammatory | Osteoarthritis induced by sodium monoiodoacetate | Hydroalcoholic extract of leaves | Oral administration s. i.d. for 25 days: 50 mg/kg, 150 mg/kg, 450 mg/kg |
### Table 2. Cont.

| Authors | Type of Study | Action | Etiological Agent | Plant Material/Part | Treatment |
|---------|---------------|--------|-------------------|---------------------|-----------|
| [100]   | Survey of studies with herbal medicines | Anti-acne | Does not apply | Not mentioned | Not informed |
| [101]   | Survey of traditional use | Treatment of skin irritation and healing | Measles and smallpox | Leaves | Infusion and bath (measles and smallpox); leaves macerated and applied to the affected area (lesions) |
| [102]   | Review | Treatment of tuberculosis-related symptoms | Mycobacterium tuberculosis | Not specified | Not specified |
| [103]   | Survey of traditional use | Anemia and weakness | Does not apply | Leaves | Not specified |
| [104]   | In vivo study | Antihypertensive | Does not apply | Hydroalcoholic extract of leaves | Oral administration of the extract at doses of 100 mg/kg; 250 mg/kg and 500 mg/kg |
| [105]   | Survey of traditional use | (1) Anemia, weakness, restoration facial color in malaria patients; (2) ovarian cysts, cystitis, hepatitis, liver, diarrhea; (3) flu, cough, anemia; (4) aids getting pregnant, ulcers, (5) vaginal itching | Does not apply | Leaves | (1) maceration or tea; (2) tea or infusion; (3) syrup; (4) bottled; (5) bath. No dose determined |
| [21]    | Ex vivo study | Photoprotection | Does not apply | Various parts | Topical application of nonionic cream with 2.5% ethylacetate fraction and 2.5% hexane fraction |
| [106]   | In vitro study | Photosensitization | MCF-7 cells of human breast adenocarcinoma | Extract nanoemulsion produced from aerial parts | CC50: 1.3 µg ACE/mL |
| [107]   | In vitro study | Anti-hepatic | Does not apply | Leaves | 0.25-1.25 mg/mL |
| [108]   | In vivo study | Anti-hepatic | Carbon tetrachloride | Ethanolic extract of leaves | 300, 500 and 600 mg/kg |
| [109]   | In vitro study | Antioxidant | Free Radical DPPH (1,1-diphenyl-2-picrylhydrazyl) | Ethanolic extract of leaves and fractions | 5, 10, 25, 50, 125 and 250 µg/mL in ethanol |
| [14]    | In vitro study | Antioxidant | Free Radical DPPH (1,1-diphenyl-2-picrylhydrazyl) | Methanolic extract of leaves | 0.25; 2.5; 25 and 250 µg/mL |
Among biological effects from *F. chica* leaf extracts, anti-inflammatory, antioxidant, and healing properties are well evidenced from previous studies (Table 2), and result from the leaves’ phytochemical composition. Studies have shown that the compounds identified in *F. chica* leaf extracts, as well as in several other plants, are responsible for the above-mentioned pharmacological activities (Table 1) when evaluated in isolation (Table 1).

5.1. Healing Activity

The tissue repair process is divided into three successive and overlapping phases that present specific characteristics: (1) the inflammatory phase, (2) the proliferative phase, and (3) the remodeling phase. In the inflammatory phase, there is an increase in vascular permeability, which promotes chemotaxis and the consequent intense migration of leukocytes, mainly neutrophils and macrophages, to the site of tissue damage in response to the release of pro-inflammatory chemical mediators. The proliferative phase is characterized by the reconstitution of epithelial tissue, angiogenesis and migration of endothelial cells, granulation tissue formation, and extracellular matrix deposition, mainly collagen produced by activated fibroblasts. In the remodeling phase, the initial deposited collagen (type III) is degraded by collagenases and reabsorbed, and the gradual replacement of type III collagen by type I collagen occurs until the proportion found in healthy tissue is reached, providing reorganization of the extracellular matrix and marking the success of tissue repair [110,111].

Several herbal medicines have been explored in order to evaluate their action on tissue healing, mainly due to their lower cost and fewer observed adverse effects [112]. Tissue regeneration studies with *F. chica* leaves have addressed the action of different extracts and formulations in both in vitro and in vivo experiments using experimental animals in order to investigate different phases of the healing process. Among the main experimental procedures in the bioprospecting of new components with healing potential, cell proliferation assays using fibroblasts and the production of extracellular matrix components such as collagen are considered highly reliable models. Fibroblasts are connective tissue cells that are essential for the formation of the dermis and are responsible for structural firmness. After injury, fibroblasts proliferate and migrate to the injured site, where they produce a large amount of extracellular matrix material, such as type I and III collagen, which helps to isolate and repair the injured tissue. The use of a methanolic extract of *F. chica* leaves stimulates the growth of fibroblasts in vitro in a concentration-dependent manner and increases collagen production [14].

From this tissue regeneration property, several in vivo studies with experimental animals have been developed to evaluate the healing potential of *F. chica* in different tissue types. In treatment of gastric ulcers induced by ethanol and indomethacin in Wistar/Uni rats, administration of the hydroalcoholic extract of *F. chica* leaves reduced about 60% of lesions [113]. The methanolic extract administered orally was able to reduce up to 96% of the ulcerative effect of ethanol in a murine model [14]. The ethanol-induced model is associated with gastric mucosal injury induced by oxidative stress (via alcohol); thus, the gastroprotective effect of *F. chica* may occur via antioxidant action. In skin lesions of Swiss mice, treatment with ethanolic extract of *F. chica* leaves has not yet shown promising results. During 21 days of observation, it was found that the extract did not accelerate total tissue regeneration and presented a profile that was similar to the control group [90]. Other studies have investigated the effect of *F. chica* leaves on the healing process of tendon injuries in rats. Observed results included an increase in the total collagen content on the seventh and twenty-first day of the healing process, an increase in the synthesis of collagenase type IV (matrix metalloproteinase-2, an enzyme that participates in the extracellular matrix remodeling process), and better organization of collagen fibers. The treated groups presented a lower inflammatory reaction, which promoted the recovery of the injured animals’ ability to walk [114,115].

In addition to the studies developed in different tissues, investigations of the healing properties of *Fridericia chica* have used different types of formulations for administration of the plant extract and for carrying out these studies. The choice of formulation is ex-
tremely important, as it influences the stability, distribution, solubility, bioavailability, and protection of the active compound. Topical formulations such as cream and gel containing *F. chica* leaf extract are being developed for the treatment of skin lesions [90,116]. For the treatment of gastric ulcers, a nanoformulation of *F. chica* extract has been elaborated in chitosan triphosphate-based nanoparticles for oral administration. Nanoformulation was responsible for a greater fibroblast proliferation effect, and the anti-ulcerogenic effect of *F. chica* leaf extract in a nanoformulation was more efficient compared to hydroalcoholic extract [113].

From the 43 compounds identified from *F. chica* leaves, seven molecules (comprising the classes of flavones, flavonols, terpenes, tochromanols, and alkaloids) have been described to present healing properties when evaluated alone. Among them, kaempferol (compound 24—Table 1) is a flavonol which has been described as improving repair of diabetic excisional and nondiabetic incisional wounds in rats, working as an effective topical wound healing agent [52]. Another component present in *F. chica* leaf extract in considering amount is α-tocopherol (compound 41, Table 1), belonging to the tocochromanol class. Shuid and colleagues [78] evaluated the effects of α-tocopherol supplementation on osteoporotic fracture healing in a rat model, showing an improvement in fracture healing associated with the antioxidatant property. Other compounds have been shown to induce healing activity, such as apigenin (compound 17, Table 1), with anti-hyaluronidase and anti-collagenase activities [41], scutellarin (compound 21, Table 1) with angiogenic properties [49], and pheophorbide a (compound 43, Table 1), which decreased the length of the inflammation stage [82].

5.2. Anti-Inflammatory Activity

Inflammation is defined as a protective response of the host to various interactions with the external environment. The assembly of this response requires the participation of cells and molecules, including macrophages and neutrophils, along with cytokines, such as interleukin (IL)-1β and IL-6 [117–121]. These cells recognize and discriminate stimuli as damage-associated molecular patterns (DAMPs) and pathogen-associated molecular patterns (PAMPs) through pattern recognition receptors (PRR) [122]. The inflammatory response, despite being an effective defense mechanism, can occur in an exacerbated or perpetuated manner for a longer period of time than is necessary, causing important tissue damage and contributing to the development of diseases such as rheumatoid arthritis [123,124].

The participation of inflammation in the pathophysiology of multiple diseases stimulates the constant development of anti-inflammatory drugs, and nature represents an important source of plants with therapeutic potential that needs to be explored [125,126]. Plants such as *Aloe vera* have demonstrated efficacy in controlling the inflammatory response and are considered by the Brazilian public health system as a treatment option [127]. Flavonoids, a class of secondary metabolites belonging to the group of phenolic compounds, are anti-inflammatory agents and can act to block arachidonic acid cascade through inhibition of the COX and lipoxygenase pathways [128,129]. The species *Fridericia chica* is used in traditional medicine as an anti-inflammatory agent, which has been confirmed in several studies using different extracts prepared from the leaves of this plant [15,17,130].

The chemical characterization of extracts of *F. chica* leaves has allowed the identification of 3-deoxyanthocyanidins, to which are attributed the greater part of the anti-inflammatory effect of this plant [4,17,18]. In 2001, Zorn et al. [17] demonstrated that both the lipophilic extract of *Fridericia chica* leaves (200 mg/mL) and one of its isolated components (carajurine) at a concentration of 500 µM had the ability to completely inhibit nuclear factor kappa B (NF-κB), which is responsible for the transcription of genes that encode several pro-inflammatory mediators and which participates in the pathogenesis of diseases such as multiple sclerosis [131–133]. Several authors have suggested that other unidentified substances present in the extract of *F. chica* leaves may contribute to the anti-inflammatory activity of this species [17].
Several models of inflammation have been used for the study of anti-inflammatory drugs, one of which is the experimental model of inflammation induced by snake venom. Its pathogenesis may involve the participation of the enzyme phospholipase A$_2$, which stimulates the release of arachidonic acid [134,135]. In 2009, Oliveira et al. [15] evaluated the anti-inflammatory effect of an aqueous extract of the leaves of _Fridericia chica_ in mice inoculated with the venom of the snakes _Bothrops atrox_ and _Crotalus durissus rurima_, administered via the subcutaneous and intraperitoneal routes at different times. After treatment with the extract, it was possible to observe that it showed inhibitory activity, especially in the processes of myocytolysis and granulocyte migration, thus resulting in a decrease in inflammation. This effect is attributed to substances present in the extract, flavonoids among them.

The effectiveness of _Fridericia chica_ leaves as an anti-inflammatory agent has been demonstrated in other models, such as inflammatory angiogenesis, which was induced in a murine model by a sponge implant (polyether-polyurethane) and in the proliferative process of human tumor cell lines in vitro, in which Michel et al. (2015) [130] observed a reduction in the amount of neutrophils at the site of the sponge implants and a great decrease in angiogenesis after treatment with aqueous and ethanol extracts. Flavone escutelarin derivatives, found in the hydroethanolic extract of _Fridericia chica_ leaves, have been associated with a reduction of parameters linked to inflammatory response in the model of intestinal mucositis induced by 5-fluorouracil in a study by [136].

Among phytochemical compounds present in _F. chica_ leaf extracts, 25 have been described as presenting anti-inflammatory behavior (Table 1). The anthocyanins comprise the class of most abundant compounds in _F. chica_ leaves, with carajurin composing one of the major molecules found in extract of the leaves [17]. Carajurin (compound 1, Table 1) is capable of inhibiting transcription factor NF-κB, an important signaling pathway associated with pro-inflammatory response [17]. _F. chica_ leaf extracts consist of a high range of flavonoid compounds, such as flavones and flavonols (Table 1). Regarding this class, the literature on their biological potentials is extensive, with a total of 14 flavonoids from _F. chica_ leaf extracts having been shown to suppress inflammation through different cell signaling pathways. An example is luteolin (compound 8, Table 1), a flavone which has been shown to interact with several inflammatory targets such as matrix metalloprotease 9 (MMP9) and mitogen-activated protein kinase 1 (MAPK1) [25]. Chrysoeriol (compound 9, Table 1), another flavone, ameliorates TPA-induced ear edema in mice, and its inhibition of JAK2/STAT3 and IκB/p65 NF-κB pathways is involved in the anti-inflammatory effects [27]. Hispidulin, a flavone found in a number of plants, among them in _F. chica_, was evaluated by Yu et al. (2020) [36] in relation to protection against neuroinflammation using the immortalized cell line of murine microglial BV2 (BV2 cells). Hispidulin has been shown to inhibit NO and ROS production, which suppressed the expression of the inflammation-related enzymes iNOS and COX-2 [137] in a dose-dependent manner. Corroborating these data, Torres et al. (2018) [138] showed a decrease in the enzymatic activity of lipooxygenase in the presence of ethanolic extract of _F. chica_ leaves. They observed that treatment with hispidulin in the presence of LPS inhibited the production of TNF-α, IL-6, IL-1β, and PGE2 in a dose-dependent manner. In addition, hispidulin showed activity in reducing NF-κB, and decreased the levels of phosphorylation of IκB kinase in BV2 cells. Based on the above examples, there is other evidence of the effect of the species _F. chica_ leaves in the modulation of the cytokine profile, among them that demonstrated by Lima (2020) [139], which suggests an increase in the production of IL-10 and a decrease in the production of IL-1β in RAW 264.7 cells activated with LPS in the presence of a hydroethanolic extract of _F. chica_. The authors attributed this effect to 5 O-methylescutelarin, a derivative of the flavone escutelarin. The aforementioned studies corroborate the therapeutic potential described in traditional medicine for _F. chica_.

Despite being encountered in minor proportions in _F. chica_, terpenes may play roles in its biological activities, including in anti-inflammatory response. β-carotene (compound 35, Table 1) was capable of inhibiting LPS-stimulated Cox2, Nos2, and Tnalpha gene expression.
from macrophages in vitro [69]. Another terpene, phytol (compound 32, Table 1), has been described as inducing anti-inflammatory activity in vitro and in vivo, with higher potency in the presence of others non-steroidal anti-inflammatories [64]. Among fatty acids, ethyl palmitate (compound 42, Table 1) reduced plasma levels of tumor necrosis factor-α (TNF-α) and IL-6, decreased NF-κB expression in liver and lung tissues, and ameliorated histopathological changes in an LPS-induced endotoxemia rat model [79]. The alkaloid pheophorbide a (compound 43, Table 1), was found to inhibit nitric oxide production and suppress the expression of iNOS proteins in LPS-stimulated macrophages [80].

5.3. Antioxidant Activity

The term oxidative stress, first proposed in 1985, refers to the imbalance between the production of oxidizing and antioxidant molecules; it favors oxidation reactions and compromises cell signaling and redox control [140–142]. The aforementioned changes result in excessive production of free radicals, which can contribute to the generation of reactive oxygen species, thus resulting in structural damage to biological systems [143–145]. Reactive species are defined as atoms, molecules, or ions resulting from oxygen, usually with a high reactivity capacity, which can cause damage to cellular constituents, such as lipid peroxidation and oxidation of DNA bases [146–149]. Free radicals and other reactive species are essential for cellular homeostasis, but when generated excessively they contribute to cellular aging and the development of diseases, as is observed in the development of hemolytic anemia due to deficiency of the enzyme glucose-6-phosphate dehydrogenase. In addition, oxidative stress is part of the pathophysiology of several chronic degenerative diseases, including cancer, diabetes, and hypertension [150–156].

Antioxidants are compounds that are capable of slowing, preventing, or removing damage caused by redox imbalance, and thereby regulating the development of oxidative stress [157]. The large number of adverse effects observed with the use of synthetic antioxidants have driven the search for other alternatives, among them substances isolated from natural products [158–160]. The main antioxidants present in plants are phenolic compounds, which represent a family of secondary metabolites with the property of interrupting chain reactions caused by free radicals by donating hydrogen atoms or electrons [161–163]. To this large group of compounds belong several of the flavonoids, which are present in medicinal plants such as the species F. chica [20,21,91,164–166].

As already mentioned, flavonoids are phenolic compounds which have a great antioxidant potential. Among them are flavones, the antioxidant capacity of which is related to the presence of free hydroxyl groups in their A and B rings [161,162]. Luteolin (compound 8—Table 1), one of the most abundant flavones, was identified by Do Amaral et al. (2012) [116], who evaluated the antioxidant activity of the ethanolic extract of F. chica leaves in the presence of the free radical DPPH (2,2-diphenyl-1-picrylhydrazil) using Ginkgo bibola as a positive control and demonstrated that the antioxidant capacity of F. chica extract is superior. This antioxidant effect was attributed mainly to the flavones luteolin and apigenin. In 2015, Gemelli et al. [168] confirmed the presence of luteolin in the aqueous extract of F. chica
leaves and demonstrated its antioxidant potential by DNA damage assay in CHO cells (hamster ovary cells) [109,168].

The leaves of Fridericia chica have a high flavones content along with the presence of apigenin (compound 17, Table 1), with high antioxidant activity [40], as evidenced by Siraichi et al. in 2013 [21], confirming what had already been proposed by Do Amaral et al. in 2012 [109]. Other important flavone identified in the extract of F. chica leaves are scutellarin (compound 21, Table 1) and scutellarein (compound 14, Table 1), which are present in other medicinal plants such as Scutellaria barbata and S. lateriflora. Scutellarin is a glucuronid form of scutellarein, which means that although both molecules present antioxidant activity, the aglycone form (scutellarein) has a stronger activity [34]. The research of Campos de Siqueira et al. (2019) [22], who identified carotenoids such as β-carotene and α-carotene in a hydrometanolic extract of F. chica, was not the first time that escutelarin was identified in the species F. chica, as in 2013 Siraichi et al. [21] attributed the antioxidant effect of the species to the presence of this flavone [21,22,109].

Terpenes are well known antioxidant agents, and have been shown to provide relevant protection under oxidative stress conditions. The terpene zeaxanthin (compound 36, Table 1) protects against chronic eye and cardiovascular diseases due its antioxidant property by directly quenching reactive oxygen species (ROS) and by facilitating glutathione synthesis [72]. Violaxanthin (compound 38, Table 1) possesses potent lipid peroxidation inhibitory activity [76]. Other compounds from different classes, such as α-Tocopherol and Pheophorbide a (compounds 41 and 43, respectively, in Table 1) have shown antioxidant activity as well, and are present in others plant species [77,81].

Martins et al., in 2016 [169] and Ribeiro et al., working in 2018 [170], demonstrated the antioxidant capacity of a hydroethanolic extract of Fridericia chica in the presence of both the free radical DPPH and the oxidative damage induced by ultraviolet radiation. The use of several methodologies to evaluate the effects of F. chica extracts for the control of oxidative stress allowed for a more complete view of this mechanism. Other authors, such as Torres et al. in 2018 [138] and Teixeira et al. in 2017 [171], have included other free radical neutralization assays, such as ABTS (2,2’-Azino-bis (3-ethylbenzothiazoline-6-sulfonic acid) and FRAP (ferric reducing antioxidant power), thus reaffirming the biological effects of F. chica [138,169–171].

In 2009, the Brazilian Ministry of Health published a list of 71 species that are part of the Report on Medicinal Plants of Interest to the SUS (RENISUS); F. chica is among them due to the large amount of scientific evidence pointing to its different biological effects [172,173]. One of the most important biological effects of this species is the antioxidant effect, which can contribute to the control and prevention of degenerative diseases, as already mentioned. Encouraging the development of research related to F. chica is be essential for its inclusion in the list of herbal medicines prescribed within the Brazilian health services, through which twelve herbal medicines are already available that are part of the National List of Essential Medicines (RENAME) [127].

6. Conclusions

The World Health Organization (WHO) has encouraged the inclusion of medicinal plants in primary health care networks, and in Brazil their use is regulated by the National Health Surveillance Agency (ANVISA). The public health system of Brazil has offered herbal medicines since 2007, and 12 herbal medicines have been made available in the public network by the Ministry of Health (MS). The species Fridericia chica (Bonpl.) L. G. Lohmann is traditionally used in folk medicine, especially in the Amazon region, and is part of the 71 species of interest to the SUS (Brazilian public health system). Among these species, Aloe vera, Schinus terebenthifolius, and Uncaria tomentosa are used as commercial herbal pharmaceutical formulations in the Brazilian Health System. Schinus terebenthifolius has healing and anti-inflammatory action and is used as an antiseptic topical for gynecological applications, available in the forms of cream and ovules. Uncaria tomentosa is indicated as an assistant in cases of arthritis and osteoarthritis thanks its anti-inflammatory and
immunomodulatory activity, and is available in capsule, pill, and gel forms. *Aloe vera* is indicated for the topical treatment of first and second degree burns and as an adjunct in cases of psoriasis vulgaris, and is available in cream form [85]. Therefore, the present review indicates the potential inclusion of *F. chica* on the list of herbal medicines prescribed within the scope of Brazilian Health System services, as the extract can be used in different pharmacological formulations such as topical or oral administrations in order to treat diseases associated with inflammatory response and wounded tissue.

Scientific evidence indicates the different biological effects of *F. chica*, which proves the therapeutic effect of this species in folk medicine, which is explained especially by the presence of a wide range of biologically active compounds that are known from other plant species. The present review has shown that from all compounds identified in *F. chica* leaf extract, over 70% are known to exert anti-inflammatory, antioxidant, and/or healing properties based on previous studies. Therefore, the pharmacological properties of *F. chica* leaf extract are directly associated with the phytochemical composition, with different molecules working in association in order to observe the conclusive therapeutical potential. Although carajurin is found as the major component of the extract with anti-inflammatory activity, its synergism with other less abundant components is important to the overall pharmacological properties of the extract. Therefore, the proportion of every molecule within the extract and its biological potency should be more closely investigated. It is of great importance to encourage more studies related to the chemical profile, standardization of extracts, and proof of pharmacological activities of *F. chica*. Concerning the traditional use of *F. chica* leaf extract as tea, more studies should be performed in order to evaluate the phytochemical composition of aqueous infusions, considering that almost all studies on leaves have used extract compositions with other types of solvents. New research needs to be developed in order to add scientific evidence that favors the use of this and other medicinal species, both in Brazil and in other regions of the world.

The scientific investigations cited in this study contribute to the construction of arguments, providing a solid basis to attribute anti-inflammatory, antioxidant, and healing properties to *Fridericia chica*, thus demonstrating the scientific bases that justify its use in traditional medicine.

**Author Contributions:** M.A.S., A.D.d.S.J.B. and D.C.d.M.S. conceived the main idea and designed the topics. A.D.d.S.J.B., D.C.d.M.S. and R.D.U. conducted the bibliography search. A.D.d.S.J.B., D.C.d.M.S., R.D.U., F.C.M.C., W.M.M., F.M.A.d.S., H.H.F.K., A.L.B. and M.A.S. wrote the review. A.D.d.S.J.B., D.C.d.M.S., H.H.F.K. and M.A.S. designed the figures and tables of this review. F.C.M.C., W.M.M., F.M.A.d.S., H.H.F.K., A.L.B. and M.A.S. revised and edited the manuscript. All authors have read and agreed to the published version of the manuscript.

**Funding:** The following agencies founded the present work: Fundação de Amparo à Pesquisa do Estado do Amazonas (FAPEAM) for the funding, CT&I Priority Areas Call FAPEAM 010/2021 for MAS and HHFK, and HHFK’s research under the Universal Call FAPEAM-006/2019. HHFK and WMM acknowledge FAPEAM for funding via the calls PAPAC 005/2019, PRO-ESTADO-002/2008; 007/2018; 005/2019 and POSGRAD 2020-2021.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Not applicable.

**Acknowledgments:** The authors would also like to thank Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) for the payment of scholarships to MAS (No. 163843/2020-1), WM (No. 309207/2020-7) and HK (No. 305942/2020-4).

**Conflicts of Interest:** The authors declare no conflict of interest.
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