Telomeres are specialized nucleoprotein structures that are localized at the end of chromosomes and formed by thousands of noncoding repetitive sequences of DNA associated with six proteins, the shelterin complex. Recent studies have related telomere length (TL) to longevity (Brown et al., 2012) and welfare (Bateson, 2016; Beloor et al., 2010) in farm animals. In cattle, studies into TL have focused mainly on the Holstein Friesian breed (HFR) (Ilska-Warner et al., 2019; Seeker et al., 2018b) and blood matrix (Seeker et al., 2018a), providing little information about their ratio in other cattle breeds (Tiley et al., 2010) and different biological matrix (Iannuzzi et al., 2020; Laubenthal et al., 2016).

We focused our study on Agerolese (AGR) cattle, an autochthonous Italian breed characterized by good productive longevity due to the early age of culling in this breed. Telomere length provides information about the productive lifespan and the quality of farm management, complying with the ‘One Health’ approach. This study evaluated telomere length in Agerolese cattle, an autochthonous dairy breed characterized by a long productive lifespan (13 years). Multiplex quantitative PCR estimated telomere length in DNA extracted from blood and milk matrices. Interestingly, the results showed longer telomeres in Agerolese (compared to the Holstein Friesian cattle control group), with a negative correlation between telomere length and increasing age and a synchronous trend between blood and milk samples, with a positive correlation between them.

**KEYWORDS**
Agerolese cattle, animal welfare biomarker, blood and milk samples, quantitative PCR, telomere length
breed (HFR) at the same age (24–78 months) and physiological stage (first lactation).

The HFR was chosen for comparison because it is the most studied and widespread breed of cattle. The study was approved by the Ethical Animal Care and Use Committee of the University of Naples Federico II (Prot. Nr. PG/2019/0104896), with informed consent from the cows’ owners. Healthy cattle were split into two groups according to their breed: 49 AGR (25–78 months) and 52 HFR (26–73 months). Milk (somatic cell [SC] count <200,000 cells/ml) and blood samples were collected simultaneously during the spring season at the same lactation stage (100 days postpartum). We also included a group of 42 older AGR (AGRe, 96–158 months) for comparison with the AGR group (Supporting Information) to evaluate TL in older cattle.

According to their manufacturer’s protocol, genomic DNA was isolated from blood (leukocytes) samples using the Wizard Genomic DNA Purification Kit (Promega). In contrast, genomic DNA from milk (SCs) samples was isolated using the QIAamp Fast DNA Stool Mini Kit (QIAGEN). DNA yield and purity were measured on a NanoDrop ND-1000 spectrophotometer (Thermo Scientific) for each sample. Before the quantitative polymerase chain reaction (qPCR) analysis, all the genomic DNA samples were diluted at 10 ng/μl in nuclease-free water and stored at −20°C. DNA showed the following requirements: yield >30 ng/μl, 260/280 ratio >1.7, and 260/230 ratio >1.8. All methods for monochrome multiplex qPCR followed the guidelines and regulations for qPCR experiments (Bustin et al., 2009) and TL measurement using qPCR approaches (Lindrose et al., 2021; Nettle et al., 2019). The total reaction mix per well (20μl) contained SYBR Green Supermix (Biorad), genomic DNA, and forward and reverse primer sequences for telomere and β-globin (defined as SCG), following Cawthon (2009). Each sample was assayed in triplicates (intra-assay) in three different runs (inter-assay) with negative control (NTC), and a standard curve for each primer evaluated the amplification efficiency and linearity. The quantification cycle (Cq) values presented a standard deviation (SD) <0.25, and relative TL was calculated using the ΔΔCt method with the Pfaffl correction (Pfaffl, 2001). More details are reported in the Supporting Information section.

IBM SPSS analyzed the results for the Windows software package version 22.0 (SPSS Inc.). Skewness test, kurtosis test, Z value and Shapiro test were performed to verify parameters distribution (parametric/non-parametric) and data are presented as medians and interquartile ranges. Since the data were distributed in a nonparametric way, we compared independent groups using the nonparametric Mann–Whitney test for continuous variables. Spearman’s test was used to assess correlation among RLTL, RMCTL, and age (Figure 1, Figures S1 and S2). Statistical significance was set at p ≤ 0.05.

Blood and milk matrices were compared for the first time, both in the AGR and HFR groups; a strong positive correlation between RMCTL and RLTL was found, both in groups at the first lactation (AGR; r = 0.9; p < 0.0001; Figure 1a. HFR; r = 0.7; p < 0.0001; Figure 1b) and in the two entire groups (AGR; r = 0.9; p < 0.0001; Figure S1a. HFR; r = 0.7; p < 0.0001; Figure S1b). One probable reason for this result could be the kind of cells present in both matrices. Milk SCs (lymphocytes, exfoliated epithelial cells, neutrophils, and macrophages) are usually present at low levels, varying with the health status and breed of the animal. SCs in milk are a typical physiological finding necessary for the regeneration of normal epithelia. The mammary epithelial cells are lined by blood vessels absorbing several milk precursors from blood and they are responsible for milk synthesis and release into the alveolar lumen (Alhussien & Dang, 2018). For this reason, any TL variation in the blood is also reflected in milk, as shown in our data (Table 1 and Table S1). In this way, the milk sample represents a suitable alternative, which is easier and less stressful to collect than blood samples from dairy animals.

**Figure 1** Relative milk cell telomere length (RMCTL) and relative leukocyte telomere length (RLTL) correlations in the Agerolese (a) and Holstein Friesian (b) groups at the first lactation. Spearman’s correlation analysis shows a significant strong positive correlation between RMCTL vs. RLTL in Agerolese (r = 0.9; p < 0.0001) and in Holstein Friesian (r = 0.7; p < 0.0001).
The TL comparison analysis between the AGR and HFR groups showed a statistically significant difference. RLTL and RMCTL always resulted higher in AGR than in the HFR groups at the first lactation (RLTL $p<0.01$; RMCTL $p<0.01$; Table 1) and also when the two entire groups were compared (RLTL $p<0.05$; RMCTL $p<0.01$; Table S1). Tiledi et al. (2010) studied TL in two Italian cattle breeds (Chianina and Maremmana) used for meat production and characterized by high longevity and range breeding; longer telomeres were observed in Maremmana than in Chianina. Interestingly, the selection history of Maremmana (the breed with longer telomeres) is much more similar to that of AGR (reblood with individuals of other breeds for the heads’ contraction in recent years) than HFR. Furthermore, the AGR breed is also characterized by high genome stability, probably due to the environment and mild zootechnical selective pressure (Ciotola et al., 2005). Both AGR and HFR were healthy, sampled simultaneously, and reared in the same area and under similar conditions, confirming that telomeres are also determined by heritable quantitative traits derived from both genetic and epigenetic mechanisms (Eisenberg & Kuzawa, 2018). Furthermore, we noticed that relative TL in AGR is naturally longer than in HFR at the same age and lactation stages (Table 1).

The correlations of RLTL were negative regarding the age of the animal (reported in months) in the AGR group compared with AGRRe, showing a significant negative regular correlation between RLTL vs. age in months ($r = -0.5$; $p<0.0001$; Figure S2). Before our study, only Miyashita et al. (2002) studied TL, by terminal restriction fragment, in leukocytes from 50 Japanese black cattle sampled one time (age range 0–18 years), showing the decline in telomere lengths with age. In line with Miyashita et al. (2022), our results demonstrated that RLTL reported an age-dependent decline (Figure S2) not linked to a specific pathology, as reported in humans, but to senescence. This is probably due to better maintenance of telomerase expression and the lack of age-related diseases (including cancer, atherosclerosis, autoimmune disorders, obesity, chronic obstructive pulmonary disease, diabetes, hematological disorders, and neurodegenerative diseases) often associated with the increased speed of telomere shortening in elderly humans (Kordinas et al., 2016). Sometimes, it is difficult to separate the effects of chronic and/or age-related diseases from normal ageing. For this reason, TL can be considered a relevant biomarker of farm animals’ general state, revealing potentially risky diseases and environmental stress (Iannuzzi et al., 2019; Seeker et al., 2021) due to the lack of age-related pathologies.

Still, we observe that the TL in the HFR breed, whose median age at culling is 6 years, is significantly lower than in the AGR breed, supporting the hypothesis that a higher TL may indicate a longer productive lifespan.

To summarize, our results revealed a higher TL in a long-living autochthonous cattle breed (Agerolose). Since TL heritability has been shown (Seeker et al., 2018a), and a longer TL is observed in a cattle breed with a longer productive lifespan, the inclusion of this parameter in the breeding selection plans could be interesting. Moreover, this study shows that milk, a biological matrix that is easier to collect from dairy animals than blood, can be used to evaluate telomere length.

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**CONFLICT OF INTEREST**
The authors declare that no competing interests exist.

**DATA AVAILABILITY STATEMENT**
The data that support the findings of this study are available from the corresponding author, [AI], upon reasonable request.

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**SUPPORTING INFORMATION**

Additional supporting information can be found online in the Supporting Information section at the end of this article.

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