INTRODUCTION

Preterm infants need nutritional and medical requirements in accordance with the physiologic maturity at birth. The main targets physicians reach for are maintaining optimal postnatal corporal and cerebral growth. In fact, there are only a few approaches to improving the outcomes of infants in a pathogen-rich and nutrient-poor neonatal intensive care unit environment. In this pilot study, we hypothesize that synbiotics may enhance brain growth, which is reflected indirectly by an increase in head circumference through several signalling molecules.

Methods: A pilot study was conducted in preterm infants with a gestational age of ≤32 weeks and a birth weight of ≤1500 grams at neonatal intensive care unit of Uludag University Medical Faculty (NICU) for one-year period. Following the randomization of the infants, a prepared commercial synbiotic solution containing multi-combined probiotics and prebiotics was administered enterally to the study group.

Results: The odds of a patient having a lower body weight and head circumference below the 10th percentile were significantly lower in the probiotic group (p=0.001, p=0.03, respectively). Moreover, the infants in the synbiotics group had a more optimal head circumference (between the 50th and 90th percentiles, p=0.001).

Conclusions: Our results show that if we can maintain optimal gut microbiota, we might achieve better neuro-development via the beneficial effects of synbiotics on cytokines, neurotransmitters, and the cellular immunity of the nervous system. Further investigational models are needed to demonstrate the beneficial effects of synbiotics on the central nervous system.

KEYWORDS: Neurodevelopment, Newborn, Synbiotics.
and a reduction in pro-inflammatory cytokines. Probiotics are becoming more important for preterm newborn nutrition since these infants have frequent gastrointestinal and systemic complications because of the altered microbiota. Currently, probiotic use in the neonatal period for preterm infants is generally accepted as a regular element of optimal nutrition.

During the last trimester of pregnancy, important phases of brain growth and maturation occur. Both white and grey matter structures undergo a dramatic increase in volume, with the cerebellum and cortical grey matter showing the highest growth rates. Maturation of myelin-producing cells, development of axons, and proliferation and migration of neurons to the cerebral cortex take place in the period between 24 and 40 weeks of gestation. It is also known that bidirectional signalling exists between the gastrointestinal tract and the brain at the gut-brain axis, which is vital for controlling homeostasis and regulated at the neuronal, hormonal, and immunological levels. This bilateral interaction may adjust brain function and development with pro- and anti-inflammatory cytokines, chemokines, immune cells, and hormones. In this pilot study, we hypothesize that synbiotics may enhance brain growth, which is reflected indirectly by an increase in head circumference through these signalling molecules.

**METHODS**

A pilot study was conducted on preterm infants with a gestational age of ≤32 weeks and a birth weight of ≤1500 grams at the neonatal intensive care unit of Uludag University Medical Faculty for one-year period. The infants with detected chromosomal abnormalities, previous gastrointestinal system surgery, and infants with a severe sepsis episode were excluded from the study. Alternate randomization was used to enrol the infants in the study; following the randomization of the infants, a prepared commercial synbiotic solution containing the multi-combined probiotics of Lactobacillus rhamnosus (4.1x10⁸ cfu), Lactobacillus casei (8.2x10⁸ cfu), Lactobacillus plantarum (4.1x10⁸ cfu), and Bifidobacterium animalis (4.1x10⁸ cfu) (NBL Probiotic®, Nobel, Istanbul, Turkey) together with 383 mg of fructooligosaccharides and 100 mg of galactooligosaccharides as the prebiotic content was administered enterally to the study group. All infants’ demographic and clinical characteristics were recorded such as birth weight, gestational week, head circumferences, gender, antenatal steroids, mode of delivery and feeding characteristics.

Minimal enteral feeding was started on the first day of life with the infant’s mother’s milk for all patients, and formula feeding was preferred when mother’s milk was not available. The synbiotic supplement was started along with the first enteral feeding and continued until the patient was discharged. The infants who developed abdominal distension, gastric residual, or vomiting during feedings were categorized as having episodes of feeding intolerance and their feeding was suspended. Body weights and head circumferences of the subjects were recorded on a weekly basis on standardized charts. The study was approved by Uludag University Clinical Research Ethics Board. Written consent was obtained from preterm infants’ parents.

Statistical analyses of the data were performed using the Statistical Package for the Social Sciences (SPSS), version 16.0.1 (SPSS Inc., Chicago, IL, USA). All the continuous values were presented as median and mean ± standard deviation, where suitable. The categorical values were presented as numbers and percentages. Chi-square analysis or Fisher’s exact test was used to compare the categorical variables among groups. The Mann-Whitney U-test was used to compare nonparametric variables. Logistic regression analysis was used to identify the factors affecting the outcome. Statistical significance was set at p<0.05.

**RESULTS**

A total of 139 patients were included in the study. Of the 110 infants finally analysed, 64% (n=70) were in the group that received probiotics (Group-1), and 36% (n=40) were in the control group (Group-2). Patients were randomized in a 2:1 ratio to receive probiotics or not (control group). The demographic and clinical characteristics of the groups are summarized in Table-I. All infants started to be fed within the first hour of life with own mother’s milk as a study protocol. If mother’s milk was not available, preterm formula was given. There was found to be no significant difference between the groups (p=0.108). In Group-1, the initiation of synbiotic supplementation ranged between the 2nd and 7th days postnatal, with a mean of 4.3±1.5 days; patients in Group-1 had a mean of 36.5±12.6 days of synbiotic use. Multivariate analysis was carried out
for head circumference of our patients in accordance with birth weight and gestation weeks. Examining birth weight and birth head circumference of groups was not given significant difference. The odds of a patient having a lower body weight and a head circumference below the 10th percentile were significantly lower in the synbiotic group (p=0.001, p=0.03; respectively). Moreover, the infants in the probiotic group had a more optimal body weight and head circumference (between the 50th and 90th percentiles, p=0.001). The feeding characteristics of the study infants are summarized in Table-II. Adverse effect was not observed due to synbiotic supplement in patients.

**DISCUSSION**

As recent advances in the science of neonatology have increased the survival of the tiniest of

| Table-I: Demographic and clinical characteristics of the infants. |
|---------------------------------------------------------------|
| **Group 1** (Probiotic group) *(n=70)*                          |
| **Group 2** (Control group) *(n=40)*                           |
| **P**                                                          |
|----------------------------------------------------------------|
| Gestational age (weeks)                                        |
| 29.7 ± 1.9                                                    |
| 29.3 ± 1.7                                                    |
| 0.3                                                           |
| Birth weight (gram)                                           |
| 1228 ± 257                                                    |
| 1228 ± 249                                                    |
| 0.9                                                           |
| Gender, male, n (%)                                           |
| 45 (64)                                                       |
| 19 (47)                                                       |
| 0.1                                                           |
| Antenatal steroid, n (%)                                      |
| 33 (47)                                                       |
| 21 (52)                                                       |
| 0.7                                                           |
| Cesarean-section birth, n (%)                                  |
| 58 (82)                                                       |
| 31 (77)                                                       |
| 0.6                                                           |

| Table-II: Weight gain and head circumference at birth and discharge. |
|-----------------------------------------------------------------------|
| **Group 1** Probiotic *(n=70)*                                        |
| **Group 2** Control *(n=40)*                                          |
| **P**                                                                 |
|-----------------------------------------------------------------------|
| **n** **%** **n** **%** **n** **%**                                    |
| < 10 **p**                                                             |
| 10 14.3 9 22.5                                                        |
| 10-50 **p**                                                          |
| 45 64.3 24 60.0                                                      |
| 0.5                                                                  |
| 50-90 **p**                                                          |
| 15 21.4 7 17.5                                                       |
| < 10 **p**                                                             |
| 14 20.0 6 15.0                                                       |
| 10-50 **p**                                                          |
| 41 58.6 29 72.5                                                      |
| 0.3                                                                  |
| 50-90 **p**                                                          |
| 15 21.4 5 12.5                                                       |
| < 10 **p**                                                             |
| 19 27.1 20 50.0                                                      |
| 10-50 **p**                                                          |
| 40 57.1 18 45.0                                                      |
| 0.03                                                                 |
| 50-90 **p**                                                          |
| 11 15.7 2 5.0                                                       |
| < 10 **p**                                                             |
| 23 32.9 27 67.5                                                      |
| 10-50 **p**                                                          |
| 33 47.1 12 30.0                                                      |
| 0.001                                                                |
| 50-90 **p**                                                          |
| 14 20.0 1 2.5                                                       |

**Feeding characteristics**

| Total duration of TPN use, days                                    |
|------------------------------------------------------------------|
| 13 ± 5                                                           |
| 22 ± 11                                                          |
| 0.001                                                             |
| Time to full enteral feeding, days                               |
| 14 ± 3                                                           |
| 21 ± 8                                                           |
| <0.001                                                            |
| Feeding intolerance, n (%)                                       |
| 24 (34.3)                                                       |
| 31 (79.5)                                                       |
| <0.001                                                            |

TPN: Total parenteral nutrition.
infants, the importance of optimal growth and neurodevelopment has also gained paramount importance. One of the main aims of neonatologists is sustaining this postnatal growth as it would have progressed in the intrauterine period and maintaining optimum neurodevelopment since the time spent in the hospital is the most critical period of postnatal growth. In this pilot study, we used multivariate analysis for head circumference of our patients in accordance with birth weight and gestation weeks. We aimed to show that very low birth weight infants had a shorter time until full enteral feedings, better weight gain, and a larger head circumference at the time of discharge when administered combined multi-strain probiotic and prebiotic supplements, and we demonstrate that synbiotic preparations have beneficial effects on these parameters, the most important of which is the more optimal head circumference growth in.

In preterm infants, microbiota are different from those of healthy term infants, with a gastrointestinal system that is dominated by Proteobacteria initially and a deficiency of detectable Bifidobacterium and Lactobacillus. Development of gut microbiota is affected by the mode of delivery, colonization from the surrounding environment, use of antibiotics, and feeding characteristics. Breastmilk is a rich source of mutual microorganisms, which are necessary for the maintenance of healthy microbiota. Besides regulating microbiota directly and indirectly, breastfeeding is a source of multiple neuroactive agents or their precursors such as tryptophan and can also affect the central nervous system. In animal studies, administration of Bifidobacterium infants enhanced the plasma concentration of tryptophan, suggesting that normal microbiota can affect the precursor pool for serotonin (5-HT). In addition to optimal development of the neuroendocrine system, head growth is a proxy measure of brain growth, which can be assessed by means of magnetic resonance imaging. In our study, we demonstrate that infants on synbiotics needed less time to reach full enteral feedings, which was in accordance with the previous literature. However, we believe our novel finding that the infants on synbiotics had a larger head circumference upon discharge from the hospital deserves attention.

The growth of Bifidobacterium, which may decrease the load of potentially pathogenic micro-organisms in the sequence in the gut. In a recent study, despite supplementation of prebiotics, there was no improvement in neuro-developmental outcomes, and lower Bifidobacteria colonization correlated with lower neuro-developmental outcomes in preterm infants. This bifidogenic effect contributes to the survival of probiotic micro-organisms in the host and regulates an immunologic interaction in the neuronal network. Our synbiotic combination also contained a prebiotic compound, which might further explain the improved neurological development of premature infants, reflected by an optimal head circumference.

Nowadays, it is well known that normal gut microbiota are important for the optimal neurological and behavioural development of the human organism. In recent reviews, effects of antimicrobial properties of different probiotics on gut immune system and homeostasis by enhancing the gut epithelial barrier and modulating mucus production indicated. However, it is not well established which nutrients and to what extent nutrient components have an effect on neurological maturation. Our results show that if we can maintain optimal gut microbiota, we might achieve better neuro-development via the beneficial effects of synbiotics on cytokines, neurotransmitters, and the cellular immunity of the nervous system. Further investigational models are needed to demonstrate the beneficial effects of synbiotics on the central nervous system.

Limitations of the study: First one was limited number of patients while second was not assessing the correlation between head circumference and neurological development at two years of age monitoring of our patients. However, we are evaluating our study in this respect.

Source of funding: None.

Conflict of Interest Statement: None.

REFERENCES
1. Belfort MB, Rifas-Shiman SL, Sullivan T, Collins CT, McPhee AJ, Ryan P, et al. Infant growth before and after term: Effects on neurodevelopment in preterm infants. Pediatr 2011;128(4):899-906. doi: 10.1542/peds.2011-0282.
2. Martin CR, Walker WA, Probiotics: Role in pathophysiology and prevention in necrotizing enterocolitis. Semin Perinatol 2008;32(2):127-137. doi: 10.1053/j.semperi.2008.01.006.
3. Collado MC, Cernada M, Neu J, Perez-Martinez G, Gormaz M, Vento M. Factors influencing gastrointestinal tract and microbiota immune interaction in preterm infants. Pediatr Res. 2015;77(6):726-731. doi: 10.1038/pr.2015.54.
4. Chauhan M, Henderson G, McGuire W. Enteral feeding for very low birth weight infants: Reducing the risk of necrotising enterocolitis. Arch Dis Child Fetal Neonatal Ed. 2008;93(2):F162-166. doi: 10.1136/adc.2007.115824.
5. AlFaleh K, Anabrees J. Probiotics for prevention of necrotizing enterocolitis in preterm infants. The Cochrane Database of Syst Rev. 2014;(4):CD005496. doi: 10.1002/14651858.CD005496.
6. Clouchoux C, Guizard N, Evans AC, du Plessis AJ, Limperopoulos C. Normative fetal brain growth by quantitative in vivo magnetic resonance imaging. Am J Obstet Gynecol 2012;206(2):173 e1-8. doi: 10.1016/j.ajo.2011.10.002.
7. Huppi PS, Warfield S, Kikinis R, Barnes PD, Zientara GP, Jolesz FA, et al. Quantitative magnetic resonance imaging of brain development in premature and mature newborns. Ann Neurol 1998;43(2):224-235. doi: 10.1002/ana.410430213.
8. Volpe JJ. Brain injury in premature infants: A complex amalgam of destructive and developmental disturbances. Lancet Neurol 2009;8(1):110-124. doi: 10.1016/S1474-4422(08)70294-1.
9. Cryan JF, O'Mahony SM. The microbiome-gut-brain axis: From bowel to behavior. Neurogastroenterol Motil 2011;23(3):187-192. doi: 10.1111/j.1365-2982.2010.01664.x.
10. Keunen K, van Elburg RM, van Bel F, Benders MJ. Impact of nutrition on brain development and its neuroprotective implications following preterm birth. Pediatr Res. 2015;77(1-2):148-155. doi: 10.1038/pr.2014.171.
11. Barrett E, Guinane CM, Ryan CA, Dempsey EM, Murphy BP, O’Toole PW, et al. Microbiota diversity and stability of the preterm neonatal ileum and colon of two infants. Microbiologyopen. 2013;2(2):215-225. doi: 10.1002/mbo3.64.
12. Clarke G, O’Mahony SM, Dinan TG, Cryan JF. Priming for health: Gut microbiota acquired in early life regulates physiology, brain and behaviour. Acta Paediatr. 2014;103(8):812-819. doi: 10.1111/apa.20164.
13. Sudo N, Chida Y, Aiba Y, Sonoda J, Oyama N, Yu XN, et al. Postnatal microbial colonization programs the hypothalamic-pituitary-adrenal system for stress response in mice. J Physiol. 2004;558(PT 1):263-275. doi: 10.1113/jphysiol.2004.063388.
14. Bravo JA, Forsythe P, Chew MV, Escaravage E, Savignac HM, Dinan TG, et al. Ingestion of Lactobacillus strain regulates emotional behavior and central GABA receptor expression in a mouse via the vagus nerve. Proc Natl Acad Sci USA. 2011;108(38):16050-16055. doi: 10.1073/pnas.1102999108.
15. Cheong JL, Hunt RW, Anderson PJ, Howard K, Thompson DK, Wang HX, et al. Head growth in preterm infants: Correlation with magnetic resonance imaging and neurodevelopmental outcome. Pediatr. 2008;121(6):e1534-1540. doi: 10.1542/peds.2007-2671.
16. Kapiki A, Costalos C, Oikonomidou C, Triantafyllidou A, Loukatou E, Pertrohilou V. The effect of a fructo-oligosaccharide supplemented formula on gut flora of preterm infants. Early Hum Dev. 2007;83(5):335-339. doi: 10.1016/j.earlhumdev.2006.07.003.
17. Van den Berg JP, Westerbeek EA, Broring-Starre T, Garssen J, van Elburg RM. Neurodevelopment of preterm infants at 24 months after neonatal supplementation of a prebiotic mix: A randomized trial. J Pediatr Gastroenterol Nutr. 2016;63(2):270-276. doi: 10.1097/MPG.0000000000001148.
18. Jeurink PV, van Esch BC, Rijnierse A, Garsen J, Knippels LM. Mechanisms underlying immune effects of dietary oligosaccharides. Am J Clin Nutr. 2013;98(2):572S-577S. doi: 10.3945/ajcn.112.038596.
19. Cong X, Henderson WA, Graf J, McGrath JM. Early life experience and gut microbiome: The brain-gut-microbiota signalling system. Adv in Neonatal Care: 2015;15(5):314-323; quiz E1-2. doi: 10.1097/ANC.0000000000000191.
20. La Fata G, Weber P, Mohajeri MH, Probiotics and the Gut Immune System: Indirect Regulation. Probiotics Antimicrob Proteins. 2018;10(1):11-21. doi: 10.1007/s12602-017-9322-6.

Authors’ Contribution:

IGV: Concepted the original idea, initial and final draft, data supervision, writing and critical review of the manuscript. NK: Designed the study, data analysis and interpretation. HO: Did data analysis and interpretation. OB: Designed tables with text data. PD: Did data collection and added references text.

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