Prognostic role of circulating tumor cells in patients with EGFR-mutated or ALK-rearranged non-small cell lung cancer

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Keywords
ALK; circulating tumor cells; EGFR; molecular targeted therapy; non-small cell lung cancer.

Abstract

Background: Circulating tumor cell (CTC) counts at baseline and follow-up are an independent prognostic factor in patients receiving standard chemotherapy for non-small cell lung cancer (NSCLC). This study further explored the role of CTCs in EGFR-mutated and ALK-rearranged NSCLC patients administered targeted therapies as first-line treatment.

Methods: CTCs were enumerated with a novel high-efficiency detection method from the blood of 43 patients with EGFR-mutated or ALK-rearranged NSCLC at baseline and at disease-progression. Patients were stratified into favorable and unfavorable groups with baseline CTC counts of < 8 or ≥ 8 CTCs/3.2 mL, respectively.

Results: A total of 76.7% of the patients were positive for ≥ 2 CTCs /3.2 ml blood at baseline. The median progression-free survival (PFS) and overall survival (OS) rates of the favorable compared to the unfavorable group were longer (11.6 vs. 8.5 months, \( P = 0.004 \) for PFS; 21.0 vs. 17.7 months, \( P = 0.013 \) for OS). Multivariate analysis demonstrated that baseline CTC count was a strong predictor of PFS (hazard ratio 2.835; 95% confidence interval 1.240–6.483; \( P = 0.014 \)) and OS (hazard ratio 3.317; 95% confidence interval 1.360–8.092; \( P = 0.008 \)).

Conclusion: Baseline CTC count could be a predictive biomarker for EGFR-mutated and ALK-rearranged NSCLCs, which allows for better guidance and monitoring of patients over the course of molecular targeted therapies.

Introduction

Non-small cell lung cancer (NSCLC) is one of the most common cancers in the world, with increasing incidence in recent decades.1 Recent advances in molecular therapies targeted to specific oncogenic driver alterations, such as EGFR mutations and ALK rearrangements, have greatly improved the quality of life and long-term prognosis of patients with advanced NSCLC.2,3 However, more precise regimens involve greater complexity in terms of decision-making. A biomarker for better personalization and monitoring of patient treatment would thus be of great potential clinical utility.

It is widely recognized that circulating tumor cells (CTCs) are of great prognostic value as a tumor biomarker for metastatic breast, colorectal, and prostate cancers.4-6 We recently demonstrated that CTC counts at baseline and CTC count changes during treatment could also serve as independent predictive and prognostic markers for NSCLC patients receiving first-line chemotherapy.7,8 However, previous analyses were limited to patient subgroups without oncogenic alterations, or patients with positive EGFR mutations/ALK rearrangements administered chemotherapy as first-line therapy. Little is known about the prevalence and clinical significance of CTCs in EGFR-mutant and ALK-rearranged NSCLC.

The aims of this study were to further explore the clinical value of CTC in patients with EGFR mutations or ALK alterations receiving targeted therapy as first-line treatment,
and to evaluate the relevance of CTC-related characteristics in this oncogene-addicted group of NSCLC patients.

**Methods**

**Study design**

Patients with histologically proven advanced (stage IIIb or IV) NSCLC who attended the Peking Union Medical College Hospital (PUMCH) between October 2013 and September 2015 were included in this prospective study. Adult males and females with a sensitizing \( \text{EGFR} \) mutation or \( \text{ALK} \) rearrangement in primary tumor tissue and no prior history of treatment with \( \text{EGFR} / \text{ALK} \)-targeted agents were eligible for the study. Previous surgical and adjuvant chemotherapy was permitted one year after treatment for recurrent NSCLC. All patients received \( \text{EGFR} \) or \( \text{ALK} \) targeted therapy as first-line treatment. Peripheral blood samples (3.2 mL) were collected for CTC analysis at baseline and at disease progression. All of the samples were collected under protocols approved by the PUMCH Ethics Committee and written informed consent from the patients.

Tumor assessments were performed using a computed tomography scan and/or magnetic resonance imaging every six or eight weeks, and disease status was evaluated using Response Evaluation Criteria in Solid Tumors (RECIST). Progression-free survival (PFS) and overall survival (OS) were measured from the date of informed consent to the date of disease progression or death. Patients who were progression-free and alive at the time of final analysis were censored.

**Circulating tumor cell (CTC) collection and analysis**

Circulating tumor cells were enumerated and identified using a novel technique that combines subtraction enrichment, leukocyte common antigen (CD45) immunostaining, and fluorescence in situ hybridization. The technical details, including sensitivity, accuracy, linearity, and reproducibility, are described in our previous reports.\(^7,8\)

Herein, \( \geq 2 \) CTCs per 3.2 mL of blood was classified as CTC-positive, and a cut-off threshold of 8 CTCs per 3.2 mL of blood was selected to stratify patients into favorable (\(< 8 \) CTCs per 3.2 mL of blood) and unfavorable (\( \geq 8 \) CTCs per 3.2 mL of blood) prognostic groups, based on preliminary studies.\(^7,8\)

**Statistical analysis**

Differences between groups were tested using a chi-square test or with a Mann–Whitney \( U \) test in the case of continuous variables. Survival curves were plotted using the Kaplan–Meier method and compared by log-rank test. Cox proportional hazards regression analysis was used to estimate univariate and multivariate hazard ratios for PFS and OS. Statistical analysis was carried out using SPSS version 19.0 (IBM Corp., Armonk, NY, USA) and a two-tailed \( P \) value of \(< 0.05\) was considered statistically significant.

**Results**

**Patient characteristics**

A total of 43 patients were enrolled in this study. The clinicopathological characteristics of the patients at the time of study entry are presented in Table 1. The histologic type in all 43 patients was lung adenocarcinoma. \( \text{EGFR} \) mutations were detected in 36 patients (L858R point mutations \( [n = 11] \) and exon 19 deletions \( [n = 25] \)) using an amplification-refractory mutation system (AmoyDx, Xiamen, China), while seven patients were identified as harboring \( \text{ALK} \) rearrangements using the Ventana immunohistochemistry platform (Roche, Basel, Switzerland). All patients received molecular targeted agents as first-line therapy: 28 cases received gefitinib, 8 icotinib, and 7 crizotinib.

**CTC count at baseline**

Overall, 33 patients (76.7\%) had positive CTC counts at baseline, with \( \geq 2 \) CTCs per 3.2 mL of blood (range 0–8) being positive. The median CTC count was 2.5 (range 0.1–72). The median survival time for the overall study population was 17.5 months (range 1–48 months).

| Characteristics | \( n \) | Proportion (%) |
|-----------------|-------|---------------|
| **Age**         |       |               |
| \(< 60\)        | 26    | 60.5          |
| \(\geq 60\)     | 17    | 39.5          |
| **Gender**      |       |               |
| Male            | 15    | 34.9          |
| Female          | 28    | 65.1          |
| **Smoking history** |     |               |
| Yes             | 14    | 32.6          |
| No              | 29    | 67.4          |
| **ECOG**        |       |               |
| 0               | 33    | 76.7          |
| 1               | 10    | 23.3          |
| **TNM status**  |       |               |
| IIIb            | 6     | 14.0          |
| IV              | 37    | 86.0          |
| **Tumor size**  |       |               |
| \(> 3 \text{ cm}\) | 23    | 53.5          |
| \(\leq 3 \text{ cm}\) | 20    | 46.5          |
| **Mutation status** |   |               |
| \(\text{EGFR}\) | 36    | 83.7          |
| \(\text{ALK}\)  | 7     | 16.3          |

ECOG, Eastern Cooperative Oncology Group; TNM, tumor node metastasis.
0–50). In total, 21% of patients had an unfavorable CTC count (≥ 8 CTCs) at baseline, while 79% had a favorable count (< 8 CTCs): 48.8%, ≤ 3; 58.1%, ≤ 4; 62.8%, ≤ 5; and 69.8%, ≤ 6. No significant associations were observed between baseline CTC counts and clinicopathological characteristics, including age, gender, smoking history, Eastern Cooperative Oncology Group performance status, tumor stage, tumor size, mutant status (EGFR or ALK), the number of metastases, and metastatic organs (i.e. the bone, liver, and adrenal gland), as shown in Table 2.

**Prognostic significance of CTC count at baseline**

At the final analysis on 30 March 2017, 34 patients had experienced disease progression, 25 patients had died, and 5 patients were lost to follow-up. The median OS and PFS rates of all patients were 20.2 and 10.0 months, respectively.

In CTC univariate analysis, the median OS rate for patients with a favorable CTC count (< 8 CTCs) at baseline was 21.0 months compared to 17.7 months for patients with unfavorable counts (≥ 8 CTCs): log-rank test \( P = 0.013 \) (Fig 1); Cox proportional hazards regression: hazard ratio (HR) 2.739, 95% confidence interval (CI) 1.194–6.283; \( P = 0.017 \) (Table 3). Consistent with the results of our previous studies, the median PFS rate of patients in the favorable group was also significantly longer than the rate in the unfavorable group: 11.6 vs. 8.5 months, log-rank test \( P = 0.004 \) (Fig 2); Cox proportional hazards regression: HR 3.084; 95% CI 1.357–7.011; \( P = 0.007 \) (Table 3).

In univariate analyses, age and a smoking history were significantly correlated to OS, while a smoking history and Eastern Cooperative Oncology Group performance status were significantly associated with PFS (Table 3). These clinical factors were then included in multivariate forward stepwise Cox proportional hazards regression analyses. The baseline CTC count remained an independent prognostic factor for OS and PFS (HR 3.317, 95% CI 1.360–8.092, \( P = 0.008 \) for OS; HR 2.835, 95% CI 1.240–6.483, \( P = 0.014 \) for PFS) (Table 3, Table S1).

### Table 2 Relationship between circulating tumor cell count and clinicopathological characteristics

| Variable                  | \( n \) | \( P \) | Z Statistics |
|---------------------------|--------|--------|--------------|
| Age                       | 0.726  | −0.350 |              |
| < 60                      | 26     |        |              |
| ≥ 60                      | 17     |        |              |
| Gender                    | 1.000  | 0.000  |              |
| Male                      | 15     |        |              |
| Female                    | 28     |        |              |
| Smoking history           | 0.167  | −1.383 |              |
| Yes                       | 14     |        |              |
| No                        | 29     |        |              |
| ECOG                      | 0.750  | −0.318 |              |
| 0                         | 33     |        |              |
| 1                         | 10     |        |              |
| TNM status                | 0.069  | −1.817 |              |
| IIIb                      | 6      |        |              |
| IV                        | 37     |        |              |
| Tumor size                | 0.059  | −1.887 |              |
| > 3 cm                    | 23     |        |              |
| ≤ 3 cm                    | 20     |        |              |
| Mutation status           | 0.246  | −1.159 |              |
| EGFR                      | 36     |        |              |
| ALK                       | 7      |        |              |
| Liver metastases          | 0.784  | −0.274 |              |
| Yes                       | 4      |        |              |
| No                        | 39     |        |              |
| Bone metastases           | 0.990  | −0.012 |              |
| Yes                       | 22     |        |              |
| No                        | 21     |        |              |
| Adrenal metastases        | 0.950  | −0.063 |              |
| Yes                       | 4      |        |              |
| No                        | 39     |        |              |
| No. of metastatic sites   | 0.149  | −1442  |              |
| < 2                       | 27     |        |              |
| ≥ 2                       | 16     |        |              |

ECOG, Eastern Cooperative Oncology Group; TNM, tumor node metastasis.
Changes in CTC count during follow-up

In the current study, the CTC counts of 29 patients were collected and analyzed at the time of disease progression, and changes in the CTC count from baseline in these patients are shown in Figure 3. The CTC count was increased in 14 patients, decreased in 13, and stable in 2. The median CTC counts were 6 (range 0–50) and 7 (range 0–38) CTCs/3.2 mL of blood at baseline and at disease progression, respectively. Further analysis showed no significant difference between CTC counts at these two time-points ($P = 0.932$).

Discussion

In recent years, CTCs have emerged as an important tumor biomarker for a wide range of human cancers.4–11 Our previous reports of CTC analyses in advanced NSCLC mainly focusing on patients without oncogenic mutations

Table 3 Univariate and multivariate Cox proportional hazards regression analysis for prediction of OS and PFS

| Variable                        | OS (95.0% CI) | P   | PFS (95.0% CI) | P   |
|---------------------------------|---------------|-----|---------------|-----|
| Age: < 60 vs. ≥ 60              | 0.479 (0.189–1.210) | 0.120 | 0.435 (0.205–0.922) | 0.030 |
| Gender: female vs. male         | 1.437 (0.623–3.312) | 0.395 | 1.620 (0.805–3.261) | 0.176 |
| Smoking history: yes vs. no     | 0.344 (0.146–0.808) | 0.014 | 0.314 (0.143–0.689) | 0.004 |
| ECOG PS: 0 vs. 1                | 0.294 (0.087–0.994) | 0.049 | 0.478 (0.206–1.107) | 0.085 |
| Tumor stage: II vs. IV          | 1.274 (0.378–4.294) | 0.696 | 1.342 (0.470–3.829) | 0.582 |
| Tumor size: < 3 vs. ≥ 3 cm      | 1.523 (0.674–3.440) | 0.312 | 2.065 (1.000–4.267) | 0.050 |
| Mutation status: EGFR vs. ALK   | 1.547 (0.519–4.609) | 0.434 | 1.930 (0.780–4.777) | 0.155 |
| Liver metastasis: yes vs. no    | 0.491 (0.144–1.669) | 0.254 | 1.222 (0.371–4.022) | 0.742 |
| Bone metastasis: yes vs. no     | 0.737 (0.330–1.646) | 0.456 | 0.761 (0.387–1.497) | 0.429 |
| Adrenal metastasis: yes vs. no  | 3.121 (0.419–23.258) | 0.267 | 1.300 (0.397–4.260) | 0.665 |
| Baseline CTCs: < 8 vs. ≥ 8      | 2.739 (1.194–6.283) | 0.017 | 3.084 (1.357–7.011) | 0.007 |

Stepwise multivariate Cox proportional hazards regression analysis

| Variable                        | OS (95.0% CI) | P   | PFS (95.0% CI) | P   |
|---------------------------------|---------------|-----|---------------|-----|
| Smoking history: yes vs. no     | 0.227 (0.087–0.593) | 0.022 | 0.301 (0.135–0.669) | 0.003 |
| ECOG PS: 0 vs. 1                | 0.216 (0.058–0.805) | 0.022 | —             | —   |
| Baseline CTCs: < 8 vs. ≥ 8      | 3.317 (1.360–8.092) | 0.008 | 2.835 (1.240–6.483) | 0.014 |

Bold value indicates $P<0.05$ are statistically significant. CI, confidence interval; CTCs, circulating tumor cells; ECOG PS, Eastern Cooperative Oncology Group performance status; HR, hazard ratio; OS, overall survival; PFS, progression-free survival.
rearranged NSCLCs.\textsuperscript{12} The current study extends our previous analyses to patients harboring \textit{EGFR} mutations or \textit{ALK} rearrangements, and yielded remarkably consistent results showing poorer PFS and OS rates in patients with unfavorable compared to favorable baseline CTC counts.

To our knowledge, this study provides the first complete evidence of the prognostic value of baseline CTC counts for predicting PFS and OS rates in patients with positive \textit{EGFR}/\textit{ALK}. To date few reports have investigated the prognostic significance of CTCs in \textit{EGFR}-mutated or \textit{ALK}-rearranged NSCLCs.\textsuperscript{12-15} Yang \textit{et al.} performed similar CTC analyses using the CellSearch system (Johnson & Johnson, Brunswick, NJ, USA) and reported PFS values of 11.1 versus 6.8 months (\textit{P} = 0.009) for \textit{EGFR}-mutated NSCLC patients with < 5 CTCs compared to those with ≥ 5 CTCs at baseline.\textsuperscript{12} Their multivariate analysis indicated that baseline CTC number was the most significant prognostic factor of PFS (HR 8.635, 95\% CI 2.341–15.613; \textit{P} < 0.001). He \textit{et al.} reported that a higher baseline CTC count was significantly associated with inferior PFS (5.6 vs. 11.5 months; \textit{P} = 0.005) and OS (18.3 vs. 22.8 months; \textit{P} = 0.010) rates in advanced NSCLC patients with \textit{EGFR} mutations.\textsuperscript{13} These findings in \textit{EGFR}-mutated NSCLC patients correspond to the results of our study.

Furthermore, we enrolled patients with \textit{ALK}-rearranged NSCLC, and no significant difference was observed between \textit{EGFR}-mutated and \textit{ALK}-rearranged NSCLC patients. However, another study of \textit{ALK}-rearranged NSCLC patients did not confirm the prognostic role of baseline CTC count. Pailler \textit{et al.} enrolled 41 \textit{ALK}-rearranged NSCLC patients treated with crizotinib, and evaluated CTC levels with aberrant \textit{ALK}-fluorescence in situ hybridization patterns at baseline and after two months of crizotinib treatment.\textsuperscript{14} They found no significant relationship between baseline CTC levels and PFS (data not shown). Discrepancies may have arisen as a result of targeting different CTC populations and NSCLC oncogenic mutations. We examined 34 cases with \textit{EGFR} mutations, but only seven cases with \textit{ALK} rearrangement in the final analysis of prognostic significance. Larger samples are needed to elucidate more fully the association between baseline CTC counts and \textit{ALK}-rearranged NSCLC. Nonetheless, with respect to the remarkable difference in survival outcomes between unfavorable and favorable CTC groups of advanced NSCLC patients, it is critical that this novel biomarker be carefully balanced when evaluating patient prognosis in future clinical trials.

In this prospective study, we examined CTC levels at two time-points: baseline and disease progression. No significant increase in CTC level at disease progression was observed (increase/stable, \textit{n} = 16; decrease, \textit{n} = 13). Similarly, Pailler \textit{et al.} also reported no significant difference between CTC levels at these two time points.\textsuperscript{14} However, they did report a significant association between a decrease in CTC level with \textit{ALK}-copy number gain after crizotinib treatment and longer PFS (HR 4.485, 95\% CI 1.543–13.030; \textit{P} = 0.006). Punnoose \textit{et al.} conducted a study of 41 patients treated with erlotinib and pertuzumab and reported that increasing CTC levels during treatment were closely associated with shorter PFS (\textit{P} = 0.006) and a better RECIST response (\textit{P} = 0.019).\textsuperscript{15} Previous studies have demonstrated that a dynamic change in CTC count during treatment may serve as a predictive biomarker of chemotherapy response.\textsuperscript{16,17} Unfortunately, in the present study we did not monitor changes in CTC levels over the course of treatment as it was difficult to keep patients in the study. Further analysis of changes in CTC levels can be expected from future large-scale prospective studies.

We performed CTC analysis using the Cyttel method (Cyttel, Beijing, China), which is independent to the epithelial cell adhesion molecule (EpCAM) method and thus obtains higher efficiency than the CellSearch system.\textsuperscript{18} A threshold of eight CTCs was chosen to define unfavorable and favorable prognostic groups, and was validated as a suitable CTC threshold for EpCAM-independent CTC detection methods. The CTC detection rates in this study were consistent with those previously reported by studies of NSCLC patients without mutations. No correlations were found between baseline CTC counts and clinicopathological characteristics in \textit{EGFR}-mutated or \textit{ALK}-rearranged NSCLC patients.\textsuperscript{7,19}

The main limitations of this study were the inadequate sample size of \textit{ALK}-rearranged cases and a lack of analysis of other oncogenes, such as \textit{KRAS}, \textit{BRAF}, and \textit{c-MET}. CTC detection during treatment would have yielded further valuable information. Nonetheless, our data provides a basis for future studies.

In conclusion, our data demonstrated the prognostic significance of the presence and characterization of CTCs in \textit{EGFR}-mutated and \textit{ALK}-rearranged NSCLC patients treated with targeted therapies. The baseline CTC count remains an important prognostic factor contributing to risk stratification and individualized treatment for advanced NSCLC patients. Further studies are warranted to validate these results.

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Disclosure

No authors report any conflict of interest.

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Supporting Information

Additional Supporting Information may be found in the online version of this article at the publisher’s website.

Table S1 Stepwise multivariate Cox analysis of overall survival (OS) and progression-free survival (PFS).