Genetic Distances of Paralichthys olivaceus Populations Investigated by PCR

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ABSTRACT: The author carried out PCR-based genetic platform to investigate the hierarchical polar dendrogram of Euclidean genetic distances of one bastard halibut population, particularly for Paralichthys olivaceus, which was further connected with those of the other fish population, by involving with the precisely designed oligonucleotide primer sets. Eight oligonucleotides primers were used generating excessively altering fragments, ranging in size of DNA bands from larger than approximately 100 bp to less than 2,000 bp. As regards average bandsharing value (BS) results, individuals from Hampyeong population (0.810) displayed lower bandsharing values than did individuals from Wando population (0.877). The genetic distance between individuals approved the existence of close relationship in the cluster II. Relatively, individuals of one bastard halibut population were fairly related to that of the other fish population, as shown in the polar hierarchical dendrogram of Euclidean genetic distances. The points of a noteworthy genetic distance between two P. olivaceus populations demonstrated this PCR procedure is one of the quite a few means for individuals and/or populations biological DNA investigates, for species security and proliferation of bastard halibut individuals in coastal region of the Korea.

Key words: Euclidean genetic distances, Polar dendrogram, Bastard halibut population

INTRODUCTION

Paralichthys olivaceus is commercially important teleost species, belonging to family Paralichthyidae, order Pleuronectiformes, broadly distributed on the seashore of the Yellow Sea, southern sea and the Jeju Island of Korea, Chinese sea and Japanese sea. In the environment, the fishes inhabit the benthic flats consisting of a lot of sand and slime. Like other fishes, the rate at which the fish grows, is greatly influenced by water quality. The outer body color of this fish is yellowish and/or body color is black and grey. The color of the abdomen is yellowish brown or light white. Mainly, there are marked shifts of the fish weight, size, color and shape in P. olivaceus in keeping with the ecological surroundings of habitat such as prey, rock crystal, water temperature, feed and harsh period. The bastard halibut is environmentally and biologically very important fishes in the Korea. However, this kind of finfish, which are well-known important environmentally (Bae et al., 2017), physiologically (Kim et al., 2018), histopathologically (Kim et al., 2017), as well as aquaculturally (Lee & Yoo, 2016) are not genetically and/or molecular-biologically studied comparable other fishes. There is a necessity to understand the genetic traits and composition of this finfish population in order to evaluate exactly the patent genetic significance. PCR-based molecular research...
METHODS AND MATERIALS

PCR analysis was accomplished on DNA samples extracted from a total of 22 individuals using eight oligonucleotides primers. DNA extraction should be performed along with the separation and extraction methods (Yoon & Kim, 2004). 600 µL of chloroform was added to the mixture and then inverted (no phenol). After quite a few washing, the lysis buffer I (155 mM NH₄Cl; 10 mM KHCO₃; 1 mM EDTA) was augmented to samples, and the mixture tubes were gently upset. The precipitates obtained were centrifuged and suspended with lysis buffer II (10 mM Tris-HCl, pH 8.0; 10 mM EDTA; 100 mM NaCl; 0.5% SDS) and added 15 µL proteinase K solution (10 mg/mL). After incubation, there was added 300 µL of 3 M NaCl and gently pipetted for a few of min. Added not phenol, 600 µL of chloroform were added to the mixture and then inverted. Ice-cold 70% EtOH was added, and then the samples were centrifuged at 19,621 g for 5 min to extract the DNA from the lysates. The concentration of the extracted genomic DNA was measured with the optical density (OD) at 260 nm by a spectrophotometer (Beckman Coulter, Buckinghamshire, UK). The DNA pellets were then incubation-dried for more than 12 hours, maintained at −70°C until needed and then dissolved in the distilled water. The DNA amplification was performed in 25 µL containing 10 ng of template DNA, 20 µL premix (Bioneer Co., Daejeon, Korea) and the 1.0 unit primer. Amplification products were separated by 1.4% agarose (Bioneer Co., Daejeon, Korea) gel electrophoresis with TBE (90 mM Tris, pH 8.5; 90 mM borate; 2.5 mM EDTA). The 100 bp DNA ladder (Bioneer Co., Daejeon, Korea) was used as DNA molecular weight marker. The agarose gels electrophoresed were stained with ethidium bromide (Song & Yoon, 2013). The electrophoresed agarose gels were illuminated by ultraviolet rays, and photographed using a photomaton direct copy system (PECA Products, Beloit, WI, USA). The oligonucleotides primer were acquired from Operon Technologies, USA. OPA-02 (5’-TGCCGAGCTG-3’), OPA-07 (5’-GAAACGGGTG-3’), OPA-18 (5’-AGGTGACCGT-3’), OPA-20 (5’- GTTGCGATCC-3’), OPB-08 (5’-GTCCACACGG-3’), OPB-09 (5’-TGGGGGACTC-3’), OPB-15 (5’-GGAGGGTGTT-3’), and OPB-17 (5’-AGGGACACGAG-3’) were displayed to yield the bandsharing values and genetic distances of the two bastard halibut populations. PCR was carried out using programmable DNA Thermal Cycler Cycler (MJ Research Inc., Waltham, MA, USA). Similarity matrix including bandsharing values between dissimilar individuals in the two P. olivaceus populations, was generated allowing formula of Jeffreys and Morton (1987) and Yoke-Kqueen and Radu (2006). A hierarchical clustering tree was accumulated using similarity matrices to yield a dendrogram, which was supported by the Systat version 10 (SPSS Inc., Chicago, IL, USA).

RESULTS AND DISCUSSION

In this study, the bandsharing value, which is based on the presence or absence of amplified fragments, was utilized to calculate similarity indices in two bastard halibut populations, as demonstrated in Table 1. Here, the complexity of the banding patterns varied dramatically be-
between the primers from the two finfish populations. The similarity matrix, which was based on the average bandsharing value of all the samples, ranged from 0.677 to 0.987 in the Hampyeong population and 0.757–0.980 to the Wando population. The bandsharing value between individuals no. 08 and no. 09 within the Hampyeong population (P. olivaceus) was 0.987, which was the highest value identified among the two populations. As regards average bandsharing value (BS) results, individuals from Hampyeong population (0.810) displayed lower bandsharing values than did individuals from Wando population (0.877). The average bandsharing value reported by this study is similar to the value ported for Spanish barbell species (0.71–0.81) (Callejas & Ochando, 1998). The average bandsharing value recorded in our study is also higher than the average value between the bullhead population (0.504±0.115) (Yoon & Kim, 2004). In the present study, the hierarchical polar dendrogram obtained by the eight oligonucleotides primers designates two genetic clusters: cluster

| Table 1. Trigonal similarity matrix containing bandsharing values calculated using Nei and Li’s index of the similarity of two bastard halibut (P. olivaceus) populations from Hampyeong and Wando, respectively |
|-----------------------------------------------|
| **Bandsharing values of Hampyeong population** | **Bandsharing values of Wando population** |
| 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 | 17 | 18 | 19 | 20 | 21 | 22 |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| 1 | - | 0.831 | 0.816 | 0.831 | 0.76 | 0.738 | 0.801 | 0.713 | 0.724 | 0.794 | 0.740 | 0.659 | 0.560 | 0.550 | 0.605 | 0.603 | 0.623 | 0.564 | 0.568 | 0.538 | 0.592 | 0.549 |
| 2 | - | 0.847 | 0.838 | 0.859 | 0.800 | 0.82 | 0.774 | 0.810 | 0.786 | 0.731 | 0.562 | 0.567 | 0.591 | 0.613 | 0.654 | 0.642 | 0.611 | 0.609 | 0.569 | 0.635 | 0.625 |
| 3 | - | 0.906 | 0.785 | 0.768 | 0.802 | 0.719 | 0.751 | 0.707 | 0.677 | 0.572 | 0.572 | 0.532 | 0.512 | 0.546 | 0.609 | 0.582 | 0.548 | 0.548 | 0.570 | 0.557 |
| 4 | - | 0.805 | 0.807 | 0.818 | 0.759 | 0.793 | 0.771 | 0.757 | 0.52 | 0.553 | 0.504 | 0.488 | 0.522 | 0.559 | 0.537 | 0.501 | 0.496 | 0.519 | 0.511 |
| 5 | - | 0.918 | 0.86 | 0.891 | 0.831 | 0.789 | 0.751 | 0.551 | 0.554 | 0.544 | 0.546 | 0.596 | 0.614 | 0.565 | 0.586 | 0.523 | 0.584 | 0.580 | 0.589 |
| 6 | - | 0.918 | 0.907 | 0.867 | 0.785 | 0.767 | 0.554 | 0.528 | 0.536 | 0.548 | 0.590 | 0.554 | 0.535 | 0.510 | 0.473 | 0.528 | 0.519 |
| 7 | - | 0.845 | 0.84 | 0.827 | 0.761 | 0.504 | 0.506 | 0.468 | 0.483 | 0.544 | 0.537 | 0.516 | 0.515 | 0.477 | 0.536 | 0.526 |
| 8 | - | 0.987 | 0.907 | 0.828 | 0.528 | 0.554 | 0.517 | 0.532 | 0.562 | 0.556 | 0.560 | 0.560 | 0.557 | 0.498 | 0.554 | 0.592 |
| 9 | - | 0.894 | 0.815 | 0.566 | 0.541 | 0.53 | 0.545 | 0.576 | 0.568 | 0.573 | 0.569 | 0.511 | 0.567 | 0.557 |
| 10 | - | 0.921 | 0.613 | 0.575 | 0.567 | 0.621 | 0.618 | 0.610 | 0.610 | 0.611 | 0.587 | 0.635 | 0.622 |
| 11 | - | 0.568 | 0.545 | 0.575 | 0.624 | 0.589 | 0.580 | 0.556 | 0.556 | 0.557 | 0.578 | 0.568 |
| 12 | - | 0.823 | 0.757 | 0.824 | 0.770 | 0.784 | 0.808 | 0.764 | 0.844 | 0.839 | 0.792 |
| 13 | - | 0.870 | 0.885 | 0.896 | 0.889 | 0.878 | 0.853 | 0.858 | 0.852 | 0.895 |
| 14 | - | 0.922 | 0.879 | 0.896 | 0.863 | 0.891 | 0.818 | 0.859 | 0.824 |
| 15 | - | 0.951 | 0.930 | 0.891 | 0.899 | 0.880 | 0.867 | 0.849 |
| 16 | - | 0.980 | 0.935 | 0.923 | 0.878 | 0.889 | 0.878 |
| 17 | - | 0.955 | 0.943 | 0.899 | 0.909 | 0.898 |
| 18 | - | 0.937 | 0.892 | 0.808 | 0.908 |
| 19 | - | 0.900 | 0.940 | 0.943 |
| 20 | - | 0.887 | 0.911 |
| 21 | - | 0.915 |
| 22 | - | |

| Table 2. Multiple calculations of average bandsharing values (mean±SE) between two bastard halibut populations were generated in keeping with the bandsharing values and similarity matrix. |
|-----------------------------------------------|
| **Population** | **Hampyeong** | **Wando** |
| Hampyeong | 0.810±0.009<sup>a</sup> | 0.560±0.004<sup>b</sup> |
| Wando | - | 0.877±0.007<sup>a</sup> |

<sup>a,b</sup> Values with different superscript are significantly different, p<0.05. Each value is a result of three different experiments.
Fig. 1. Hierarchical polar dendrogram of genetic distances obtained from two *Paralichthys olivaceus* populations.

The relatedness between dissimilar individuals of two bastard halibut populations from cluster I (HAMPYEONG 01, 02, 03, 04, 05, 06, 07, 08, 09, 10, and 11) and cluster II (WANDO 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, and 22) generated according to the bandsharing values and similarity matrix.

The genetic distance between individuals approved the existence of close relationship in the cluster II. The values of the pairwise comparisons of unbiased genetic distance between the populations of the Indian major carp (*Catla catla*) from the combined data for the four primers, ranged from 0.025 to 0.052 (Islam et al., 2005). They reported that the Padma and the Jamuna populations were separated from each other with the lowest genetic distance (D=0.025). From what has been said above, the prospective of this research method in determining the diagnostic markers for the breed, stock, species, genus and geographic population identification in teleost (Mamuris et al., 1999; Diaz-Jaimes & Uribe-Alcocer, 2003), in shellfish (Tassanakajon et al., 1998; McCormack et al., 2000; Oh & Yoon, 2014) and in livestock (Jeffreys & Morton, 1987; Gwakisa et al., 1994) has also been established. The points of a significant
genetic distance between two \( P. \) olivaceus populations demonstrated this PCR means is one of the several devices for individuals and/or populations biological DNA investigates.

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**REFERENCES**

Bae SH, Kim KW, Kim SK, Kim JH, Kim JH (2017) Lethal toxicity and hematological changes exposed to nitrate in flatfish, *Paralichthys olivaceus* in biofloc and seawater. Environ Biol Res 35:373-379.

Callejas C, Ochando MD (1998) Identification of Spanish barbel species using the RAPD technique. J Fish Biol 53:208-215.

Chenyambuga SW, Hanotte O, Hirbo J, Watts PC, Kemp SJ, Kifaro GC, Gwakisa PS, Petersen PH, Rege JEO (2004) Genetic characterization of indigenous goats of sub-Saharan Africa using microsatellite DNA markers. Asian-Australas J Anim Sci 17:445-452.

Diaz-Jaimes P, Uribe-Alcocer M (2003) Allozyme and RAPD variation in the eastern Pacific yellowfin tuna (*Thunnus albacares*). Fish Bull 101:769-777.

Gwakisa PS, Kemp SJ, Teale AJ (1994) Characterization of zebu cattle breeds in Tanzania using random amplified polymorphic DNA markers. Anim Genet 25:89-94.

Islam MS, Ahmed ASI, Azam MS, Alam MS (2005) Genetic analysis of three river populations of *Catla catla* (HAMILTON) using randomly amplified polymorphic DNAs markers. Asian-Australas J Anim Sci 18:453-457.

Jeffreys AJ, Morton DB (1987) DNA fingerprints of dogs and cats. Anim Genet 18:1-15.

Kim SR, Ko SM, Choi H, Park JJ (2017) The first report of marine leech, *Austrobdella* sp. parasited on the wild flounder, *Paralithys olivaceus* and histopathological characteristics of the host. J Fish Mar Sci Edu 29:1394-1404.

Kim JY, Sung GH, Lim JJ, Suo SA, Cho YR, Kim JH (2018) Effects of exposure to hexavalent chromium on hematological parameters and plasma components in flatfish, *Paralichthys olivaceus*. Korean J Environ Biol 36:124-130.

Lee HY, Yoo HK (2016) Effects of various diets on growth and body composition of juvenile olive flounder, *Paralichthys olivaceus*. Korean J Ichthyol 28:200-206.

Mamuris Z, Stamatis C, Bani M, Triantaphyllidis C (1999) Taxonomic relationships between four species of the Mullidae family revealed by three genetic methods: Allozymes, random amplified polymorphic DNA and mitochondrial DNA. J Fish Biol 55:572-587.

McCormack GP, Powell R, Keegan BF (2000) Comparative analysis of two populations of the brittle star *Amphiura filiformis* (Echinodermata: Ophiuroidae) with different life history strategies using RAPD markers. Mar Biotechnol 2:100-106.

Muchmore ME, Moy GW, Swanson WI, Vacquier VD (1998) Direct sequencing of genomic DNA for characterization of a satellite DNA in five species of Eastern Pacific abalone. Mol Mar Biol Biotechnol 7:1-6.

Oh H, Yoon JM (2014) Genetic distances of three mollusk species investigated by PCR analysis. Dev Reprod 18:43-49.

Partis L, Wells RJ (1996) Identification of fish species using random amplified polymorphic DNA (RAPD). Mol Cell Probes 10:435-441.

Song YJ, Yoon JM (2013) Genetic differences of three
Pollicipes mitella population identified by PCR analysis. Dev Reprod 17:199-205.

Tasanakajon A, Pongsomboon S, Jarayabhand P, Klinbunga S, Boonsaeng V (1998) Genetic structure in wild populations of black tiger shrimp (Penaeus monodon) using randomly amplified polymorphic DNA analysis. J Mar Biotechnol 6:249-254.

Yoon JM, Park HY (2002) Genetic similarity and variation in the cultured and wild crucian carp (Carassius carassius) estimated with random amplified polymorphic DNA. Asian-Australas J Anim Sci 15:470-476.

Yoon JM, Kim JY (2004) Genetic differences within and between populations of Korean catfish (S. asotus) and bullhead (P. fulvidraco) analysed by RAPD-PCR. Asian-Australas J Anim Sci 17:1053-1061.

Yoke-Kqueen C, Radu S (2006) Random amplified polymorphic DNA analysis of genetically modified organisms. J Biotechnol 127:161-166.

Zhou L, Wang Y, Gui JF (2000) Analysis of genetic heterogeneity among five gynogenetic clones of silver crucian carp, Carassius auratus gibelio Block, based on detection of RAPD molecular markers. Cytogenet Cell Genet 88:133-139.