Basic techniques in endoscopic ultrasound-guided fine needle aspiration for solid lesions: Adverse events and avoiding them

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ABSTRACT
Endoscopic ultrasound (EUS) guided fine-needle aspiration (FNA) is often the preferred technique for tissue acquisition in the diagnosis of suspected intrathoracic and intraabdominal pathology. Although EUS FNA is a safe and accurate procedure, it has been associated with a low risk of adverse events. The unique properties of the echoendoscope and its ability to acquire tissue outside of the gastrointestinal lumen impart risks that are not associated with routine endoscopic procedures. In this review, we discuss the risk of perforation related to the echoendoscope itself and adverse events related to FNA of solid masses including infections, bleeding, pancreatitis and pancreatic duct leak, bile duct leak and tumor seeding. We also provide tips on how to avoid the most common adverse events related to EUS-FNA.

Key words: Bleeding, infection, pancreatitis, perforation

INTRODUCTION
Since the introduction of endoscopic ultrasound (EUS)-guided fine-needle aspiration (FNA) in the early 1990s, the technique has emerged as a method of choice for tissue acquisition in the diagnosis of a broad spectrum of intraabdominal and intrathoracic malignancies.[1,2] Virtually any lesion or organ that lies in close proximity to the gastrointestinal (GI) tract can be accessed with a FNA needle. However, the utility of any tissue sampling method is measured not only by the ability to access a site and achieve an accurate diagnosis, but also by its safety profile. In this review, we discuss the adverse events that may be encountered when performing EUS-FNA of solid lesions and provide tips that may help avoid or minimize the risk of their occurrence.

TIPS TO AVOIDING ADVERSE EVENTS
The subsequent discussion will focus on various adverse events with tips provided to help avoid each specific event. However, there are general measures that may be taken to reduce the risk of all adverse events and should be considered prior to EUS. Endosonographers must be mindful of the indications and impact of performing FNA on clinical care. There must be the potential for the findings from both the EUS and FNA to impact patient management. Certainly, other factors such as the clinical need to obtain a tissue diagnosis must be carefully weighed against the risk of the procedure, including the presence and severity of comorbidities. These factors impact decisions...
regarding the need and timing of EUS-FNA. The lack of a sound justification for EUS, likely indicates an unnecessary FNA as well. Periprocedural planning is also key to optimize exam safety. Much can often be learned from thoroughly reviewing the clinical records, imaging studies and in particular prior EUS exams. As a part of pre-procedural planning, ensuring adequate sedation, through either monitored anesthesia care or general anesthesia, helps avoid inadvertent or uncontrolled patient movement that may increase the risk of complications. It is also important to consider the lesion to be sampled. For instance, the clinical consequence of an adverse event secondary to sampling a perigastric lymph node likely differs from those that occur when sampling a pancreatic or mediastinal mass. Finally, knowing when to ask for help can save both you and the patient an undesired outcome. Knowing one’s own experience and expertise is important as is relying on other endoscopists, interventional radiologists, or surgeons within a given institution or referral center when assistance is needed.

PERFORATION

Luminal perforation during EUS most often results from the echoendoscope itself. Compared with the standard upper endoscope, the echoendoscope is several millimeters wider (9.9-9.8 mm vs. 12.4-14.6 mm, respectively). In general, echoendoscopes have a longer non-flexible region between the tip of the scope and the bending region.[3] In addition, the degree of angulation that can be achieved within the distal segment of echoendoscopes is less than that of standard endoscopes.[4] These properties, along with the side-viewing nature of echoendoscopes makes intubation and scope advancement around regions of angulation more difficult and increase the risk of perforation. The risk of perforation likely varies based on the particular echoendoscope. For instance, echoendoscopes with longer and less angulated transducers are often larger in caliber and have a longer non-flexible distal region which is necessary to house the increased number of cable components. Although such instruments may offer enhanced imaging, they also can also negatively impact the physical properties of the echoendoscope.

Cervical esophageal perforation

Cervical esophageal perforation was reported in 16 of 43,852 upper GI EUS examinations (0.03%) performed by 86 members of the American Endosonography Club who participated in a survey.[5] One patient died due to the perforation (0.002% mortality), two patients required surgical intervention, while the remaining 13 patients were treated conservatively with or without percutaneous drainage. The majority (94%) of patients were elderly (age ≥ 65 years) and 94% of the cases occurred using the radial echoendoscope. 9 (56%) perforations occurred in the hands of an endosonographer who had less than 1 year of experience, suggesting that inexperience may increase the risk of perforation. A significant proportion (44%) of patients had a history of difficult intubation during prior endoscopies or had large cervical osteophytes (19%) that were thought to contribute to the perforation. Other perceived contributing factors included frail patients, inadvertent patient movement during intubation and forceful blind intubation.

A prospective study of 4894 upper GI EUS examinations performed by one experienced endosonographer documented 3 (0.06%) cervical esophageal perforations, all which occurred using a linear echoendoscope.[6] All three perforations were recognized at the time of intubation and the most common finding was a yellow area in the posterior hypopharynx, reflecting the echoendoscope entering the mediastinum. The patients all underwent surgical repair with complete recovery without subsequent swallowing difficulty. In contrast to the prior study, the perforations did not occur early in the endosonographers experience, but after the first 2500 procedures. In addition, cervical osteophytes were not felt to be a contributing factor. However, the patients were retrospectively noticed to have a short and webbed-appearing neck that was thought to contribute to the perforation.

Lower esophageal perforation

Luminal obstruction secondary to a malignant or benign esophageal stricture was historically viewed as a risk factor for perforation. Malignant tumor obstruction may prohibit passage of the echoendoscope and limit T and N staging in approximately 30% of patients.[7] In a prospective study of 3324 patients who underwent EUS at a single center, five patients developed an esophageal perforation (0.15%), which accounted for half of all adverse events.[8] Two patients had prior dilation using a 15 mm through-the-scope balloon immediately preceding the EUS examination. One patient who did not undergo prior dilation required surgical intervention, three recovered after stenting and conservative treatment and one patient did not require any endoscopic or surgical therapy.
Three studies have shown that esophageal dilation up to 14-18 mm is safe prior to EUS with no perforations occurring in the cumulative 137 patients who required dilation.[7,9,10] Dilation allowed appropriate EUS staging to be completed and in one study, EUS upstaged the tumor in 19% of patients who required dilation.[10]

**Duodenal perforation**

Duodenal perforation is more common than esophageal perforation and in one retrospective study was reported in 0.022% (19 of 85,084 upper EUS examinations) when compared to 0.009% of esophageal perforations (N = 8).[3,11] Factors felt to have contributed to duodenal perforation included duodenal diverticula (N = 4), stenosis (N = 3), ulceration (N = 1), scarring (N = 1) and acute pancreatitis (N = 1). In another study, 2 of 233 patients (0.86%) who underwent EUS-FNA for presumed pancreatic cancer experienced a duodenal perforation that required surgical intervention.[12] Death has also been reported following duodenal perforation. A study by Carrara et al. describe one death attributed to duodenal perforation in a patient with duodenal infiltration of a neuroendocrine tumor.[13] 7 of 8 deaths recorded in a multicenter Israeli study of 20,000 EUS were attributed to duodenal perforations using a linear echoendoscope.[14] In this study, duodenal diverticula were thought to contribute to 4 (57.1%) of the complications and endosonographer inexperience (defined as <300 total procedures) was implicated in five cases (71.4%).

**Colorectal perforation**

No study has been reported to date evaluating the risk of colorectal perforation during lower GI EUS.

**Tips – to avoid perforation**

It is important to have an understanding of any anatomic features that may predispose to perforation including a Zenker's diverticulum, hiatal hernia, duodenal diverticulum, malignant or benign stenosis, or surgically altered anatomy. For instance, knowledge of a large duodenal diverticulum may lead to caution when retracting the echoendoscope within the duodenum, because this maneuver typically leads to some scope advancement that may be potentially problematic. Knowledge regarding any prior difficult exams is also helpful in planning future exams. Initial intubation and scope advancement, particular around angulated regions, are also facilitated by scope lubrication and use of a partially inflated balloon.[15] In our practice, we also limit dilation to 15 mm, which nearly uniformly allows echoendoscope passage and limits the risk of perforation. One must also question the need to traverse some malignant strictures given that the vast majority of truly obstructing esophageal cancers is an advanced stage without the potential for EUS findings to impact patient care.

**INFECTIONOUS ADVERSE EVENTS**

**Bacteremia and fever**

Four studies have assessed the risk of bacteremia following EUS with or without FNA[16-19] with each finding the overall rate of bacteremia comparable to routine endoscopic procedures. Three studies evaluating upper GI EUS-FNA included a total of 202 patients; each study excluded patients who required antibiotic prophylaxis according to the American Heart Association or American Society of Gastrointestinal Endoscopy guidelines.[16-18] 12 patients (5.9%) were found to have contaminants (coagulase-negative Staphylococcus or Corynebacterium F1) on blood cultures taken 5-60 min after EUS-FNA. True bacteremia was diagnosed in five patients (2.5%) with Streptococcus viridans (N = 3), hemolytic Streptococcus group F (N = 1) and an unidentified Gram-negative bacillus (N = 1). No patient in any study developed signs or symptoms of infection. Janssen et al. compared the risk of bacteremia in EUS with and without FNA and found no significant difference (2%, 95% confidence interval [CI]: 0-4.8% in those with diagnostic EUS only vs. 4%, 95% CI: 0-9.6% in those with EUS-FNA).[18] Potential risk factors including the procedure duration, maximal size of the target lesion, number of needle passes and depth of needle penetration were similar between those who did and did not develop bacteremia.[17,18]

Fever has been reported in up to 2% of patients with pancreatic cancer who underwent EUS-FNA.[20,21] The only patient (out of 44) who developed fever in a study from Chang et al. underwent EUS-FNA of both a solid pancreatic mass and an adjacent pancreatic cystic lesion.[20] In another study, two patients who developed fever (total N = 355; 0.56%) were treated with intravenous (IV) antibiotics without clinical sequela. One of these patients had cystic spaces within the biopsied pancreatic mass. A meta-analysis that evaluated the adverse events associated with EUS-FNA identified only four patients who developed fever among 7337 EUS-FNA exams of solid pancreatic masses (0.05%).[22] Only one study has examined the risk of bacteremia following lower GI EUS in 100 patients.[19] Blood
cultures were taken immediately prior to any procedure, after the flexible sigmoidoscopy or radial echoendoscope exam and 15 min after EUS-FNA. Contaminants were found in four patients, whereas two patients had bacteremia with *Bacteroides fragilis* or *Gemella morbillorum*. In the patient with the *Bacteroides* bacteremia, only the 2nd set of blood cultures taken after the flexible sigmoidoscopy and/or radial EUS exam was positive, while in the patient with *Gemella* bacteremia the final blood culture returned positive. An updated prospective evaluation of 563 patients who underwent lower GI EUS-FNA at our center will show that five patients developed low-grade fever without clinical significance. Of note, there was no patient overlap between our two lower GI EUS-FNA studies.

**Other infections**

Most infectious adverse events are associated with EUS-FNA of cystic lesions, rather than solid masses or lymph nodes. When excluding cystic pancreatic lesions, a meta-analysis evaluating adverse events associated with EUS-FNA reported infectious adverse events in only 3 of 10,032 EUS-FNA (0.03%) exams.[22] The same meta-analysis identified two patients who developed infectious adverse events among 909 EUS-FNA exams (0.22%) of cystic pancreatic lesions.

Two case reports highlight the risk of EUS-FNA mediastinal lymph node induced mediastinal abscess formation in patients with metastatic testicular teratoma and metastatic hepatocellular carcinoma. The first event was detected 24 h after the procedure and required percutaneous drainage followed by thoracotomy.[23] The second patient presented with dysphagia 3 weeks later and required EUS-guided drainage.[24] To subsequently close the iatrogenic esophagomediastinal fistula, the endoscopist placed 2 band ligators at the lateral margins of the fistula which were fastened together using an Endoloop. Both patients experienced a full and otherwise uneventful recovery following therapy. Infectious mediastinitis occurred 2 days after EUS-FNA of a subcarinal lymph node in a patient with small cell lung cancer.[25] The patient had a chest computed tomography (CT) scan that demonstrated air bubbles within the subcarinal region. The patient responded well to conservative therapy with IV antibiotics. A patient with suspected metastatic colon cancer to an aortopulmonary window lymph node developed chest pain, fevers and leukocytosis 5 days after EUS-FNA.[26] A chest CT scan revealed inflammatory changes abutting the T5-T7 vertebral bodies and blood cultures were positive for *G. morbillorum*. A magnetic resonance imaging obtained 6 weeks after the EUS showed diskitis and osteomyelitis involving T6-T7. The patient was treated with IV ceftriaxone for 12 weeks, followed by amoxicillin for several months until the symptoms resolved. No prophylactic antibiotic was given in any of the cases where mediastinal infections occurred after lymph node EUS-FNA. One case of infectious endocarditis with *Streptococcus salivarius* has been described following EUS-FNA of a malignant mediastinal lymph node.[27]

EUS trucut biopsy (TCB) of submucosal tumors (SMT) has also been associated with infectious adverse events. Two infectious adverse events (3.9%) occurred in 49 patients undergoing EUS TCB of a gastric SMT.[28] One patient with a hepatocellular carcinoma invading the gastric wall developed streptococcal sepsis requiring a 7 day hospitalization, complicated by *Clostridium difficile* colitis. The other patient developed an abscess around a large gastrointestinal stromal tumor (GIST) that required surgical drainage. A 49-year-old patient who underwent 7 EUS-FNA passes using a 22-gauge needle of a duodenal mass was given prophylactic antibiotics for a suspected infectious granulomatous infection based on FNA results.[29] 6 weeks after completing the course of antibiotics, the patient developed fever and epigastric pain. Abdominal CT scan revealed a large peri-duodenal mass containing air-fluid levels and the abscess resolved with IV antibiotics and endoscopic drainage. EUS-FNA was repeated 2 weeks later allowing diagnosis of a duodenal GIST for which the patient underwent pancreaticoduodenectomy.

No infectious adverse events occurred in our prior prospective study on EUS-FNA of rectal and perirectal lesions (*N* = 100) or within our most recent prospective study (*N* = 563).[19] Two of 69 patients who underwent EUS-FNA of extramural pelvic masses developed an abscess.[30] Both these patients had cystic pelvic lesions, one received prophylactic antibiotic and both required percutaneous drainage. In patients with solid pelvic masses in this series, no abscesses or other infectious adverse events were diagnosed.

**Tips – to avoid infectious adverse events**

Current guidelines do not recommend using prophylactic antibiotics in average risk patients undergoing EUS-FNA of solid masses or lymph nodes as the risk of infectious adverse events is not higher than other routine endoscopies.[31] One must
consider the relative risk versus perceived need for antibiotics when sampling lesions that are partly cystic. Although there are insufficient data to verify the need we often administer antibiotics whenever the FNA needle traverses the bile duct, pancreatic duct, or major vascular structure. In addition, while data refute the need for antibiotics for lower GI EUS-FNA, the studies were conducted in centers that routinely administer a full colon prep. It is unclear whether antibiotics are necessary among patients whose colon prep is deemed inadequate.

BLEEDING ADVERSE EVENTS

Intraluminal hemorrhage

Intraluminal bleeding is typically self-limited without clinical implications [Figure 1]. This adverse event has been reported in 1-4.4% of EUS-FNA of solid pancreatic masses.\(^{32-34}\) Gress et al., in their study, the two cases of intraluminal bleeding occurred with radial echoendoscopes.\(^ {32}\) One patient had several blood clots 10-15 mm away from the FNA site that resolved spontaneously with no clinical consequence other than an extended recovery time. The second patient developed extraluminal bleeding and is described below. The four patients who developed bleeding in the Voss et al., study and two patients from the Eloubeidi et al., study did not have clinically apparent bleeding after the procedure and did not require blood transfusions.\(^ {33,34}\) One patient in a prospective study of the adverse events of EUS died from a massive GI hemorrhage, of which the source could not be identified on autopsy.\(^ {8}\)

Five patients (0.44%) had intraluminal bleeding following EUS-FNA of 1135 SMT in a multicenter study.\(^ {35}\) One patient had severe bleeding following FNA of a duodenal GIST which required packed red blood cell transfusions. The other four patients required endoscopic therapy of the bleeding site. Three patients had EUS-FNA performed while on antithrombotic agents, none of whom experienced bleeding adverse events.

Extraluminal hemorrhage

Extraluminal bleeding [Figure 2] was found in 3 of 227 patients (1.3%) with normal coagulation studies who underwent EUS-FNA at a single institution.\(^ {36}\) Despite the use of Doppler to identify intervening vascular structures, extraluminal bleeding occurred during FNA of a pancreatic tail mass, periesophageal lymph node in a patient with esophageal cancer and cystic pancreatic lesion. In every case the extraluminal bleeding appeared as a hypoechoic region in proximity to the FNA. Their practice was to apply tamponade for 15-20 min using the echoendoscope balloon and tip deflection. They noted that the hypoechoic region stabilized in size within that time and no clinical sequela (i.e., decreasing hematocrit or symptom development) occurred in any patient.

One patient in the Gress et al., study described above experienced a peripancreatic bleed, which eventually lead to death.\(^ {32}\) After two FNA passes were obtained, significant bleeding developed in the vicinity of the FNA. Epinephrine was injected and the procedure was aborted before adequate samples could be obtained. Approximately 12 h later, the patient became hypotensive with a 3 g/dL drop in hemoglobin. Esophagogastroduodenoscopy (EGD) failed to identify the bleeding source. A bleeding scan

![Figure 1](image1.png)

**Figure 1.** Ampullary bleeding following endoscopic ultrasound-guided fine-needle aspiration of a pancreatic mass. (a) Hemosuccus pancreaticus with blood extruding from the papilla; (b) Treatment of the ampullary bleed with pancreatic duct stenting

![Figure 2](image2.png)

**Figure 2.** Extraluminal bleeding after endoscopic ultrasound (EUS)-guided fine-needle aspiration of a gastrointestinal stromal tumor (GIST). (a) Endoscopic view of a gastric GIST; (b) EUS Doppler imaging reveals intralesional vessels; (c) New hyperechoic area signaling blood can be seen lying in proximity to the needle tract; (d) Increasing size of the hyperechoic area, indicating continued bleeding, within the GIST
and later an angiogram localized the bleed to an inferior pancreaticoduodenal artery pseudoaneurysm, which was embolized. The patient developed respiratory failure and died 3 days later. There was a question of whether the patient may have formed the pseudoaneurysm secondary to the pancreatic cancer itself or whether there was sufficient time to develop as direct consequence of the FNA.

**Tips – to avoid bleeding adverse events**

The technique of EUS-FNA may impact the risk of bleeding. It is thought by some that larger blood vessels lie within the center of a solid mass. If true, then one may diminish the risk of bleeding by initially targeting the periphery of the mass and only move centrally if inadequate samples are obtained. Doppler imaging may clarify the need within individual lesions. In addition, since extraluminal bleeding likely occurs from traversing blood vessels that run along the surface of the lesion, some suggest to limit the number of needle punctures made to the surface of the solid lesion to decrease the risk of bleeding adverse events. One must also be sure that smaller and/or easily compressible blood vessels (e.g., varices) are not within the needle path. Such vessels are easily compressed by the echoendoscope and flow may even be obscured during Doppler imaging. Furthermore, when we unintentionally, or intentionally, traverse a blood vessel during EUS-FNA, care is taken to avoid torquing the echoendoscope as this may lacerate the blood vessel wall and increase the risk of bleeding.

Patient characteristics can also decrease the risk of bleeding. Bleeding following EUS-FNA or TCB was seen in 0% (0/26) of patients taking aspirin or non-steroidal anti-inflammatory drugs (NSAIDs), 33.3% (2/6) patients taking prophylactic doses of low-molecular weight heparin (LMWH) and 3.7% of patients taking neither (7/190; *P* = 0.023). The mean number of FNA passes, use of suction, bloody specimens, or cellular yield did not significantly differ between the patients taking and not taking the anticoagulants. Therefore it is recommended that in patients undergoing EUS-FNA of solid lesions, aspirin or NSAIDs may be continued, but warfarin, heparin/LMWH, thienopyridines (clopidogrel, prasugrel) and other anticoagulants (fondaparinux) should be discontinued. Furthermore, many recommend a platelet count >50,000/mm³ and international normalized ratio <1.5 prior to proceeding with EUS-FNA.

**INFLAMMATORY ADVERSE EVENTS**

**Pancreatitis**

EUS-FNA of a pancreatic mass carries the risk of pancreatitis [Figure 3] as the needle often traverses normal pancreatic parenchyma and/or ducts to reach the target lesion. Acute pancreatitis has been reported in up to 2% of pancreatic EUS-FNA. The needle gauge and type of pancreatic lesion has not been shown to increase the risk of pancreatitis. The risk of pancreatitis following EUS-FNA is lower than the 3% risk of severe pancreatitis previously reported with percutaneous pancreatic FNA.

Gress et al. in their study reported two cases of acute pancreatitis developing in a cohort of 100 patients. Both patients each had two prior episodes of pancreatitis and were undergoing EUS-FNA of a mass in either the head or body of the pancreas. Their last episode of pancreatitis occurred at 6 and 8 weeks prior to the EUS. In this study, three additional patients had elevated pancreatic enzymes without clinical symptoms of pancreatitis 2 h after the procedure. As is true following endoscopic retrograde cholangiopancreatogram (ERCP), there is no clinical utility for checking serum pancreatic enzyme levels in asymptomatic patients following EUS-FNA. In a multicenter United States survey, pancreatitis occurred in 14 of 4909 (0.29%) EUS-FNA of solid pancreatic masses. The targeted lesion was located in the head for majority of the cases (N = 12), whereas the lesion was located in the body and tail in one patient each. Half of the patients had benign FNA cytopathology results, despite the overall smaller cohort of patients with benign pancreatic disease, suggesting that the risk of pancreatitis is higher in these patients who...
are undergoing the EUS for benign reasons such as recurrent acute pancreatitis. One patient with multiple other comorbidities died from a pulmonary embolism.

In a systematic review and meta-analysis of EUS adverse events, acute pancreatitis occurred in 36 of 8246 patients (0.44%) undergoing EUS-FNA of pancreatic solid or cystic lesions.\(^{[22]}\) Of these, 27 were mild (hospitalized for 2-3 days), 6 moderate (hospitalized for 4-10 days) and 3 were severe (hospital stay >10 days, presence of necrosis, abscess, or pseudocyst, or required percutaneous drainage or surgery). One patient with severe pancreatitis died.

**Pancreatic duct leak (without pancreatitis)**

Although rare, there are three case reports describing a pancreatic duct leak following EUS-FNA.\(^{[45-47]}\) A 71-year-old woman underwent EUS-FNA of a pancreatic tail cyst, which was complicated by a loculated pancreatic fluid collected with an amylase content of 7809 U/L that required percutaneous drainage.\(^{[45]}\) The patient underwent pancreatic duct stenting via ERCP and the fluid collection resolved. She was found to have a tubulovillous adenoma with high-grade dysplasia involving the ampulla, which was thought to contribute to the pancreatic duct leak. The second case was of a 67-year-old woman with metastatic carcinoid who underwent EUS-FNA with a 25-gauge needle of a pancreatic neck mass which was complicated by pancreatic ascites.\(^{[46]}\) Diagnostic and therapeutic paracentesis revealed pancreatic fluid with an amylase of 2480 U/L. The patient underwent ERCP with pancreatic duct stenting and the ascites resolved within several months. The final case described a 60-year-old man who underwent three passes using a 22-gauge needle of a pancreatic neck mass resulting in a dilated duct, but also larger side branches, in particular those located upstream from a mass resulting in a dilated duct that contains pancreatic fluid under high pressure that may extravasate from the needle track.

**Bile peritonitis or cholangitis**

Biliary adverse events following EUS-FNA may occur if the bile duct is inadvertently punctured. Bile peritonitis occurred in a 54-year-old man with a 2 cm pancreatic head lesion causing obstructive jaundice who underwent EUS-FNA with a 22-gauge needle.\(^{[48]}\) 2 days after EUS, the patient developed ascites, a fever and physical exam evidence of peritonitis. Endoscopic and percutaneous attempts to drain the bile duct were unsuccessful, which lead to laparotomy during which biliary ascitic fluid was found along with a 1 mm perforation of the bile duct. He underwent surgical drainage with resolution of his jaundice, but was subsequently found to have metastatic pancreatic cancer.

Cholangitis leading to death was reported in one patient following EUS-FNA of a liver lesion in a patient with a suspected occluded biliary stent.\(^{[49]}\) The overall adverse event rate of hepatic mass EUS-FNA in the study was 4% (6 of 167 procedures), with death (\(N = 1\)), bleeding (\(N = 1\)), fever (\(N = 2\)) and abdominal pain (\(N = 2\)) reported. This risk is higher than the risk associated with FNA of other lesions, which was confirmed by a meta-analysis that found hepatic EUS-FNA had the second highest morbidity rate (2.33%) only exceeded by FNA of ascites.\(^{[22]}\)
Tips – to avoid bile peritonitis or cholangitis
To avoid adverse events related to the biliary system, one should ensure that the FNA needle does not transect the bile duct. In particular, the endosonographer should be cautious in performing FNA around an obstructed bile duct as the only biliary adverse events reported in the literature were in patients with an inadequately drained biliary system. It is sometimes difficult to identify the bile duct as it courses through a pancreatic mass given the resulting narrowing that results.

MISCELLANEOUS ADVERSE EVENTS

Tumor seeding
The enhanced diagnostic capability of EUS-FNA must be balanced against the risk of tumor seeding; also sometimes referred to as needle tract seeding or implantation metastasis.

The reported risk of clinically evident tumor seeding following EUS-FNA of all sites is only 1/10,000-40,000.[50,51] However, there are many limitations in assessing the incidence of tumor seeding and the cited rates likely greatly underestimate the occurrence. The inevitable mortality among patients with unresectable cancers makes it difficult to document tumor seeding as patients typically succumb before developing clinical evidence of tumor seeding. In addition, for patients undergoing attempted curative resection, pre-operative biopsy likely deposits cancer cells that are undetected within the surgical specimen or deposited outside the field of resection. The resulting occult reservoir of tumor cells may lead to disease progression that is falsely attributed to incomplete resection or tumor recurrence rather than growth of residual needle tract cancer cells. Limited data suggest that tumor seeding correlates with important clinical outcomes including tumor stage, prognosis, resectability, resection margin status, recurrence and survival.[32-62] Others report a lack of correlation between tumor seeding and clinical outcomes.[83-86]

Until date, tumor seeding has been reported in six patients following EUS-FNA.[67-72] FNA of four pancreatic adenocarcinomas of the pancreatic body or tail resulted in posterior gastric body metastases at a median of 23 (range: 21-36) months after partial pancreatectomy.[67,70] Tumor seeding following EUS-FNA of a pancreatic head mass has not been reported, as the area that the needle traverses the duodenum is resected during pancreaticoduodenectomy and therefore seeding is likely not discernible. Shah et al. in their study reported a patient who developed of gastric wall metastases following EUS-FNA of metastatic melanoma to the perigastric lymph nodes 6 months prior.[71] The remaining patient underwent EUS-FNA of a malignant mediastinal lymph node secondary to metastatic gastric cancer.[72] The patient’s lymph node responded well to chemotherapy therefore he eventually underwent a partial gastrectomy. At 18 months after surgery the patient developed mid-esophageal metastases located in the region of prior EUS-FNA. In all these six patients, except the first, a 22-gauge needle was used to perform a median of 2 (range: 1-5) passes.

The potential for tumor cell displacement during FNA was recently demonstrated.[73] In a prospective study of 140 patients undergoing EUS, the luminal fluid that is routinely aspirated through the accessory channel was submitted for cytological analysis. Luminal fluid cytology was positive for malignancy in 48% of patients with a luminal cancer, which may be expected given the inherent tumor shedding that occurs within the GI lumen. More concerning was the detection of positive cytology within post-FNA luminal fluid in 3 of 26 patients (11.5%) with pancreatic cancer. In patients with extraluminal cancers such as pancreatic cancer, we would not anticipate finding malignant cells within the GI luminal fluid. This finding suggests the process of FNA may withdraw malignant cells from a pancreatic cancer into the gut. This is likely an analogous method by which needle tract seeding occurs.

This hypothesis is supported by a study that examined the rate of newly occurring peritoneal carcinomatosis following EUS versus percutaneously guided FNA among matched cohorts with pancreatic cancers.[74] One patient developed peritoneal carcinomatosis in the EUS-FNA group compared to seven patients in the percutaneous FNA group (2.2% vs. 16.3%; P < 0.025). This study suggests a difference in tumor seeding rates between techniques and a potentially greater risk of seeding than previously recognized. Similarly, a recent meta-analysis of eight published series identified tumor seeding in 2.7% of patients following biopsy of hepatocellular carcinoma.[75]
some centers, because of the potential for tumor seeding and associated worse outcomes, patients with CCA are denied transplantation if they have undergone FNA of their primary tumor. Furthermore, the endoscopist should ensure that the targeted lesion is located as close to the echoendoscope transducer as safely possible to limit the amount of tissue traverse by the needle during FNA.

**Pneumoperitoneum and pneumothorax**

A case of pneumoperitoneum without overt perforation was described in a 60-year-old female with a pancreatic head mass that underwent three FNA passes from the duodenum using a 22-gauge needle.\(^6\) 6 days later, the patient on staging CT scan was found to have massive pneumoperitoneum. The patient was asymptomatic, treated conservatively with antibiotics with resolution of the pneumoperitoneum within 2.5 weeks. The authors proposed that the air used for insufflation entered the peritoneum through a pre-existing mucosal disruption or the high intraluminal pressure allowed air to dissect through perivascular or perineural sheaths.

Although not reported in the literature, we experienced a case of pneumothorax following EUS-FNA performed for esophageal cancer staging [Figure 4].

**Esophageal fistula**

Stigt et al. reported a case of a 54-year-old female with metastatic lung cancer treated with chemotherapy who underwent EUS-FNA of a paraesophageal lymph node metastasis.\(^7\) She received bevacizumab 13 and 8 days prior to EUS. A chest scan performed at 4 and 8 weeks revealed a progressively enlarging air space in the region of the paraesophageal lymph node. EGD showed two elongated fissures with overlying blood clots. During aspiration, air bubbles could be seen arising from the fissures. The patient was treated with tube feeds and antibiotics. The authors recommend against performing EUS-FNA in patients who are taking bevacizumab, which may have delayed the wound healing process.

Another case of a mediastinal-esophageal fistula occurred in a patient who had weight loss and mediastinal lymphadenopathy. EUS-FNA of a subcarinal lymph node was performed with four passes using a 22-gauge needle. Cultures from the FNA aspirate diagnosed the patient with tuberculosis. To evaluate new onset dysphagia, the patient underwent a thoracic CT scan that showed multiple esophageal submucosal air collections. He was started on tuberculosis medications with complete resolution of the fistula.

**SUMMARY**

EUS-FNA is a generally safe and effective modality for tissue acquisition in the diagnosis of GI and non-GI pathology. However, adverse events do develop and having a thorough understanding of the breadth and rates of adverse events facilitates patient counseling. Steps may be taken to minimize the risk, but when they occur early recognition is vital to allow timely intervention and improved outcomes.

**REFERENCES**

1. Vilmann P, Jacobsen GK, Henriksen FW, et al. Endoscopic ultrasonography with guided fine needle aspiration biopsy in pancreatic disease. *Gastrointest Endosc* 1992;38;172-3.
2. Wiersma MJ, Hawes RH, Tao LC, et al. Endoscopic ultrasonography as an adjunct to fine needle aspiration cytology of the upper and lower gastrointestinal tract. *Gastrointest Endosc* 1992;38;35-9.
3. Jenssen C, Alvarez-Sánchez MV, Napoléon B, et al. Diagnostic endoscopic ultrasonography: Assessment of safety and prevention of complications. *World J Gastroenterol* 2012;18;4659-76.
4. ASGE Standards of Practice Committee, Early DS, Acosta RD, et al. Adverse events associated with EUS and EUS with FNA. *Gastrointest Endosc* 2013;77;839-43.
5. Das A, Sivak MV Jr, Chak A. Cervical esophageal perforation during EUS: A national survey. *Gastrointest Endosc* 2001;53;599-602.
6. Eloubeidi MA, Tamhane A, Lopes TL, et al. Cervical esophageal perforations at the time of endoscopic ultrasound: A prospective evaluation of frequency, outcomes, and patient management. *Am J Gastroenterol* 2009;104;53-6.
7. Pfau PR, Ginsberg GG, Lew RJ, et al. Esophageal dilation for endosonographic evaluation of malignant esophageal strictures is safe and effective. *Am J Gastroenterol* 2000;95;2813-5.
8. Mortensen MB, Fristrup C, Holm FS, et al. Prospective evaluation of patient tolerability, satisfaction with patient information, and
complications in endoscopic ultrasonography. *Endoscopy* 2005:37; 146-53.

9. Kallimanis GE, Gupta PK, al-Kawas FH, et al. Endoscopic ultrasound for staging esophageal cancer, with or without dilation, is clinically important and safe. *Gastrointest Endosc* 1995:41;540-6.

10. Wallace MB, Hawes RH, Sahai AV, et al. Dilatation of malignant esophageal stenosis to allow EUS-guided fine-needle aspiration: Safety and effect on patient management. *Gastrointest Endosc* 2000:51;309-13.

11. Jennisen C, Faiss S, Nürnberg D. Complications of endoscopic ultrasound and endoscopic ultrasound-guided interventions - results of a survey among German centers. *Z Gastroenterol* 2008:46;1177-84.

12. Raut CP, Grau AM, Staerkel GA, et al. Frequency of major complications in endoscopic ultrasound-guided fine needle aspiration of the pancreas: Evaluation of pancreatitis as a complication. *Gastrointest Endosc* 1997;45; 387-93.

13. Eloubeidi MA, Chen VK, Eltoum IA, et al. Endoscopic ultrasound-guided fine needle aspiration biopsy of a posterior mediastinal metastatic teratoma. *Endosc* 2005:37;283-90.

14. Affi A, Vaquiez-Sequeiros E, Norton ID, et al. Acute extraluminal hemorrhage associated with EUS-guided fine-needle aspiration: Frequency and clinical significance. *Gastrointest Endosc* 2001:53;221-5.

15. Boustière C, Veitch A, Vanbiervelt G, et al. Endoscopy and antiplatelet agents. European Society of Gastrointestinal Endoscopy (ESGE) Guideline. *Endoscopy* 2011:43;445-61.

16. Polkowski M, Larghi A, Weynand B, et al. Learning, techniques, and complications of endoscopic ultrasound (EUS)-guided sampling in gastroenterology: European Society of Gastrointestinal Endoscopy (ESGE) Technical Guideline. *Endoscopy* 2012:44;190-206.

17. Mueller PR, Miketic LM, Simeone JF, et al. Severe acute pancreatitis after percutaneous biopsy of the pancreas. *AJR Am J Roentgenol* 1988:151;493-4.

18. Babich JP, Bonasera RJ, Klein J, et al. Pancreatic ascites: Complication after endoscopic ultrasound-guided fine needle aspiration of a pancreatic cyst. *Endoscopy* 2009:41 Suppl 2,E211-2.

19. Reddymasu S, Oropeza-Vail MM, Williamson S, et al. Pancreatic leak after endoscopic ultrasound guided fine needle aspiration managed by transpapillary pancreatic duct stenting. *JOP* 2011:12;489-90.

20. Chung KH, Ryu JK, Oh HS, et al. Pancreatic pseudocyst after endoscopic ultrasound-guided fine needle aspiration of pancreatic mass. *Clin Endosc* 2012:45;431-4.

21. Chen HY, Lee CH, Hsieh CH. Bile peritonitis after EUS-guided fine-needle aspiration. *Gastrointest Endosc* 2002:56;864-7.

22. Polkowski M, Gerke W, Jarosz D, et al. Diagnostic yield and safety of endoscopic ultrasound-guided trucut [corrected] biopsy in patients with gastric submucosal tumors: A prospective study. *Endoscopy* 2009:41;329-34.

23. DeWitt J, Al-Haddad M, Fogel E, et al. Endoscopic transduodenal drainage of an abscess arising after EUS-FNA of a duodenal GI stromal tumor. *Gastrointest Endosc* 2009:70;185-9.
52. Doi S, Yasuda I, Iwashita T, et al. Needle tract implantation on the esophageal wall after EUS-guided FNA of metastatic mediastinal lymphadenopathy. Gastrointest Endosc 2008;67:988-90.
53. Ito Y, Asahi S, Matsuzuka F, et al. Needle tract implantation of follicular neoplasm after fine-needle aspiration biopsy: Report of a case. Thyroid 2006;16:1059-62.
54. Suzuki K, Takamochi K, Funai K, et al. Needle tract implantation clearly visualized by computed tomography following needle biopsy of malignant mesothelioma. Eur J Cardiothorac Surg 2006;29:1051.
55. Fowler N, Asatiani E, Cheson B. Needle tract seeding after bone marrow biopsy in non-Hodgkin lymphoma. Leuk Lymphoma 2008:49:156-8.
56. Chang S, Kim SH, Lim HK, et al. Needle tract implantation after percutaneous interventional procedures in hepatocellular carcinomas: Lessons learned from a 10-year experience. Korean J Radiol 2008;9:268-74.
57. Rowe LR, Mulvihill SJ, Emerson L, et al. Subcutaneous tumor seeding following needle core biopsy of hepatocellular carcinoma. Diagn Cytopathol 2007;35:717-21.
58. Liu YW, Chen CL, Chen YS, et al. Needle tract implantation of hepatocellular carcinoma after fine needle biopsy. Dig Dis Sci 2007;52:228-31.
59. Sakata J, Shirai Y, Wakisaka T, et al. Catheter tract implantation metastases associated with percutaneous biliary drainage for extrahepatic cholangiocarcinoma. World J Gastroenterol 2005;11:7024-7.
60. Gerhards MF, Gonzalez DG, ten Hoopen-Neumann H, et al. Prevention of implantation metastases after resection of proximal bile duct tumours with pre-operative low dose radiation therapy. Eur J Surg Oncol 2000;26:480-5.
61. Lundstedt C, Stridbeck H, Andersson R, et al. Tumor seeding occurring after fine-needle biopsy of abdominal malignancies. Acta Radiol 1991:32:518-20.
62. Smith EH. Complications of percutaneous abdominal fine-needle biopsy. Review. Radiology 1991:178:253-8.
63. Castells A, Boix L, Bessa X, et al. Detection of colonic cells in peripheral blood of colorectal cancer patients by means of reverse transcriptase and polymerase chain reaction. Br J Cancer 1998;78:1366-72.
64. Wyld DK, Selby P, Perren TJ, et al. Detection of colorectal cancer cells in peripheral blood by reverse-transcriptase polymerase chain reaction for cytokeratin 20. Int J Cancer 1996;69:278-82.
65. Soeth E, Röder C, Juhl H, et al. The detection of disseminated tumor cells in bone marrow from colorectal-cancer-patients by a cytokeratin-20-specific nested reverse-transcriptase-polymerase-chain reaction is related to the stage of disease. Int J Cancer 1996:69:278-82.
66. Nakamura T, Yasumura T, Hayashi K, et al. Immunocytochemical detection of circulating esophageal carcinoma cells by immunomagnetic separation. Anticancer Res 2000;20:4739-44.
67. Mori M, Mimori K, Ueo H, et al. Clinical significance of molecular detection of carcinoma cells in lymph nodes and peripheral blood by reverse transcription-polymerase chain reaction in patients with gastrointestinal or breast carcinomas. J Clin Oncol 1998;16:128-32.
68. Miyazono F, Natsuue S, Takao S, et al. Surgical maneuvers enhance molecular detection of circulating tumor cells during gastric cancer surgery. Ann Surg 2001:233:189-94.
69. Z‘raggen K, Centeno BA, Fernandez-del Castillo C, et al. Biological implications of tumor cells in blood and bone marrow of pancreatic cancer patients. Surgery 2001:129:537-46.
70. Hardingham JE, Kotasek D, Sage RE, et al. Detection of circulating tumor cells in colorectal cancer by immunobead-PCR is a sensitive prognostic marker for relapse of disease. Mol Med 1995;1:789-94.
71. Funaki NO, Tanaka J, Imamura M. Quantitative analysis of alpha-fetoprotein mRNA in circulating peripheral blood of patients with hepatocellular and alpha-fetoprotein-producing gastric carcinomas. Life Sci 1998:62:1973-84.
72. Uchikura K, Takao S, Nakajo A, et al. Intraoperative molecular detection of circulating tumor cells by reverse transcription-polymerase chain reaction in patients with biliary-pancreatic cancer is associated with hematogenous metastasis. Ann Surg Oncol 2002:9:364-70.
73. Koike M, Hibi K, Kasai Y, et al. Molecular detection of circulating esophageal squamous cell cancer cells in the peripheral blood. Clin Cancer Res 2002:8:2879-82.
74. Piva MG, Navaglia F, Basso D, et al. CEA mRNA identification in peripheral blood is feasible for colorectal, but not for gastric or pancreatic cancer staging. Oncology 2000:59:323-8.
75. Yeh KH, Chen YC, Yeh SH, et al. Detection of circulating cancer cells by nested reverse transcription-polymerase chain reaction of cytokeratin-19 (K19) – Possible clinical significance in advanced gastric cancer. Anticancer Res 1998:18:1283-6.
76. Bessa X, Elizalde JI, Boix L, et al. Lack of prognostic influence of circulating tumor cells in peripheral blood of patients with colorectal cancer. Gastroenterology 2001:120:1084-92.

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