Canadian Arctic Contaminants and Their Effects on the Maternal Brain and Behaviour: A Scoping Review of the Animal Literature

Claire Fong-McMaster 1, Sandra Konji 1, Amanda Nitschke 1 and Anne TM Konkle 1,2,3,*

1 Interdisciplinary School of Health Sciences, University of Ottawa, Ottawa, ON K1N 6N5, Canada; cfong006@uottawa.ca (C.F.-M.); sandrakonji@gmail.com (S.K.); anits104@uottawa.ca (A.N.)
2 School of Psychology, University of Ottawa, Ottawa, ON K1N 6N5, Canada
3 University of Ottawa Brain and Mind Research Institute, University of Ottawa, Ottawa, ON K1H 8M5, Canada
* Correspondence: Anne.konkle@uottawa.ca

Received: 15 January 2020; Accepted: 31 January 2020; Published: 2 February 2020

Abstract: Background: Environmental toxicants such as methylmercury, polychlorinated biphenyls, and organochlorine pesticides are potentially harmful pollutants present in contaminated food, soil, air, and water. Exposure to these ecologically relevant toxicants is prominent in Northern Canadian populations. Previous work focused on toxicant exposure during pregnancy as a threat to fetal neurodevelopment. However, little is known about the individual and combined effects of these toxicants on maternal health during pregnancy and post-partum. Methods: A scoping review was conducted to synthesize the current knowledge regarding individual and combined effects of methylmercury, polychlorinated biphenyls, and organochlorine pesticides on maternal care and the maternal brain. Relevant studies were identified through the PubMed, Embase, and Toxline databases. Literature involving animal models and one human cohort were included in the review. Results: Research findings indicate that exposures to these environmental toxicants are associated with neurochemical changes in rodent models. Animal models provided the majority of information on toxicant-induced alterations in maternal care behaviours. Molecular and hormonal changes hypothesized to underlie these alterations were also addressed, although studies assessing toxicant co-exposure were limited. Conclusion: This review speaks to the limited knowledge regarding effects of these persistent organic pollutants on the maternal brain and related behavioural outcomes. Further research is required to better comprehend any such effects on maternal brain and behaviour, as maternal care is an important contributor to offspring neurodevelopment.

Keywords: maternal behaviour; maternal brain; pregnancy; postpartum; methylmercury; polychlorinated biphenyls; organochlorine pesticides; persistent organic pollutants

1. Introduction

Environmental pollution is a global problem associated with many adverse health outcomes [1–3]. Organic and inorganic pollutants contaminate soil, air, water, and food in many urban and rural communities [4–7]. Of particular concern are polychlorinated biphenyls (PCBs) and organochlorine pesticides (OCPs), which are two classes of industrial chemicals that persist in the environment due to their stable chemical structure and long half-life [8,9]. Mercury is another potent environmental contaminant that exerts toxic effects on a variety of vital organs [10]. Due to these concerns, international agreements have been established to limit the production and usage of these chemicals [11,12].
1.1. Methylmercury

Mercury is naturally found in three predominant forms: elemental, organic, and inorganic mercury [13]. Organic methylmercury (MeHg) is the most toxic of all forms. Humans are primarily exposed through consumption of contaminated fish and marine mammals, particularly large long-lived predators at the top of the food web [3,14]. Ingested MeHg is absorbed via the digestive tract and can accumulate in the brain throughout life [15]. As a result of increasing fish consumption, chronic low-level dietary intake of MeHg has become more prevalent and as such may pose a significant toxicological risk [16].

1.2. Polychlorinated Biphenyls

PCBs are contaminants that were widely used in industrial machinery such as transformers, capacitors, hydraulic fluids, and lubricants until the 1970s [17]. This class of contaminants includes 209 chemical congeners that differ in the extent and pattern of chlorination of the biphenyl structure [18]. An increase in chlorine substitutions further decreases the aqueous solubility and degradability of PCB compounds. Thus, the structural composition has direct effects on the environmental fate and toxicity of each congener [19].

Although most industrialized countries have banned their production and use, PCBs remain widely dispersed in the environment due to their bioaccumulation in food webs [20]. Atmospheric deposition of PCBs is higher in the Arctic region compared to lower latitudes [21]. Generally, food samples from the northern hemisphere contain greater PCBs compared to the southern hemisphere [22]. Exposure to PCBs most often results from ingestion of contaminated foods such as fish, meat, and milk [23].

1.3. Organochlorine Pesticides

OCPs are a group of chlorinated compounds that were extensively used in the agricultural industry between the 1950s and 1970s [24,25]. These are a large group of compounds that consist of aliphatic and aromatic cyclical structures with multiple chlorine substitutions [25]. OCPs persist in the environment due to their stability and slow biodegradation [26]. OCPs such as hexachlorocyclohexane and endosulfan are found in high concentrations in Asia, while varied levels are observed across North America [27]. Similar to PCBs, air–seawater monitoring suggests the net deposition of OCPs in the Arctic region [28].

Since the 1970s, widespread use of OCPs has been drastically restricted because of concerns about their environmental persistence, bioaccumulation, and potential to cause adverse effects in humans [29]. Dietary consumption of fatty foods such as fish is now the main source of exposure [24] due to their bioaccumulation in these animals.

1.4. Contaminant Mixtures

Complex combinations of environmental toxicants are prevalent in the Canadian Arctic. As a result, individuals are typically exposed to multiple pollutants over time [30]. The Canadian Arctic region in particular is vulnerable to the aforementioned contaminants due to long-range transport and oceanic currents, which drive their accumulation in this region’s environment and biota [31]. The lipophilic nature and persistence of MeHg, PCBs, and OCPs allow for their bioaccumulation in high trophic level species including whales, walruses, seals, and fish [30,32–35]. Northern Aboriginal populations, over 56,000 individuals from Labrador, Northwest Territories, Nunavik, Nunavut, and Yukon, have a heavy dietary reliance on these food sources [36,37]. As such, they are exposed to high levels of complex toxicant combinations. Biomonitoring studies have indicated that contaminant burden is higher in populations that consume large amounts of traditional foods from the marine environment than in those that do not [38–40]. For example, PCB and OCP levels measured in breast milk of Nunavik Inuit mothers were nearly ten-fold higher than those found in Southern
Canada mothers [39,41–43]. Similarly, almost 25% of Aboriginal peoples of Northern and Eastern Inuit communities had MeHg levels above 20 ppm in blood (6 ppm in hair samples), the acceptable limit determined by the World Health Organization at the time of this study [44].

To investigate the possible health effects of simultaneous exposure to multiple pollutants in the Canadian Arctic communities, the Northern Contaminant Program was developed in 1991 under the Ministry of Indigenous and Northern Affairs Canada [30]. Work under this program sought to better understand the consequences of exposure to a mixture of these toxicants. As such, Health Canada developed the Northern Contaminant Mixture (NCM) to be used for testing in animal models [45]. The NCM was formulated to comprise the 27 most abundant environmental contaminants: 14 PCB congeners, 12 OCPs and MeHg, which have been detected in the blood profiles of 159 mothers residing in the Canadian Arctic [45,46].

1.5. Maternal Brain and Behaviour

Pregnancy modifies physiological and neuroendocrine processes, and the resulting behavioural adaptations allow the postpartum female to effectively care for her young [47]. An increased ratio of estradiol to progesterone and increased prolactin and oxytocin are hormonal events associated with late pregnancy and parturition. These parameters increase in rats to provide initial activation of the maternal neural circuitry and maternal behaviour [48,49]. However, this hormonal influence is transient, which is why sensory experiences acquired through mother–pup interactions are essential for the continuance of maternal responsiveness [50,51].

Many researchers have explored the impact of the abovementioned pollutants on the neurological health of offspring [52–55]. Considering the sensitive period of in utero development, studies have focused on developmental outcomes following toxicant exposure. Multiple epidemiological and experimental studies have identified damaging effects of prenatal toxicant exposure leading to cognitive dysfunction and behavioural alterations [56]. Conversely, little research has sought to explore contaminant effects on mothers during pregnancy and post-partum. Given the mother’s importance in caring for offspring once they are born, any changes in brain plasticity during this period may influence the quality of care she provides to her offspring. The quality of maternal care is paramount during offspring neurodevelopment [57–59]. Thus, this paper aims to provide a scoping review of the literature in an attempt to better understand the effects of maternal exposure to MeHg, OCPs, or PCBs on maternal health and behaviour.

2. Materials and Methods

A scoping review was conducted to explore the literature in this growing research field. This study was conducted according to the five-stage scoping review framework described by Arksey and O’Malley [60].

2.1. Research Question

This review was guided by the research question “What are the individual and combined effects of MeHg, PCB, and OCP exposure on maternal behaviour and the maternal brain?”

2.2. Identifying Relevant Studies

Comprehensive searches of the Medline, Embase, and Toxline electronic databases were conducted between July 11 and 13 2019. These databases were selected to encompass both disciplines of interest—maternal health and environmental toxicology. Articles published between 2000 and 2019 were considered for this review, as the increase in developmental toxicology studies involving maternal brain and behavioural assessments is relatively recent [61,62]. While there are potential confounding factors of epidemiological data, studies with human cohorts and experimental animal models were included due to the limited number of toxicology studies looking at maternal endpoints. [63]. The search strategy and keywords were developed with the assistance of a Health Sciences librarian from
the University of Ottawa. Keywords consisted of terms associated with the maternal exposure period, brain outcomes and toxicants, such as pregnancy, perinatal, maternal, gestation, brain, behaviour, MeHg, PCB, OCP, and northern contaminant mixture. Both keywords and subject headings were used for the Medline and Embase searches. Complete search strategy details are shown in Tables 1 and 2. All citations were imported to the Covidence reference management software (Veritas Health Innovation, Melbourne, Australia) and duplicate articles were immediately removed.
Table 1. Keywords and subject headings used for Medline and Embase searches.

| Concept 1: Population | Keywords | Subject Headings (MeSH) | Subject Headings (Emtree) |
|-----------------------|----------|-------------------------|----------------------------|
|                       | pregnancy.ti,ab,kw. | pregnancy OR perinatal care OR maternal exposure OR perinatal period | pregnancy OR perinatal period OR perinatal care OR maternal care OR maternal exposure |
|                       | pregnant.ti,ab,kw. |                         |                            |
|                       | perinatal.ti,ab,kw. |                         |                            |
|                       | maternal.ti,ab,kw. |                         |                            |
|                       | postpartum.ti,ab,kw. |                         |                            |
|                       | antenatal.ti,ab,kw. |                         |                            |
|                       | gestation*.ti,ab,kw. |                         |                            |

| Concept 2: Exposure | Keywords | Subject Headings (MeSH) | Subject Headings (Emtree) |
|---------------------|----------|-------------------------|----------------------------|
|                     | methylmercury.ti,ab,kw. | methylmercury OR polychlorinated biphenyl OR Aroclor OR Aroclor 1242 OR Aroclor 1260 OR Aroclor 1254 OR Aroclor 1248 OR organochlorine insecticide OR “1,1 dichloro 2,2 bis(4 chlorophenyl)ethane” OR “1,1 dichloro 2,2 bis(4 chlorophenyl)ethylene” OR “1,1,1 trichloro 2 (2 chlorophenyl) 2 (4 chlorophenyl)ethane” OR 1,2 dichlorobenzene OR 1,4 dichlorobenzene OR aldrin OR alpha hexachlorocyclohexane OR beta hexachlorocyclohexane OR campheclor OR chlordane OR chlordene OR chlordanes OR chlordecone OR chloroform OR dichlorodiphenyldichloroethane OR dichlorodiphényl dichloroéthanes OR dichlorodiphenyl dichloroethanes OR dichlorodiphenyl dichloroethanes OR chlorobenzilate OR chloropicrin OR chlorothalonil OR dacthal OR dicofol OR dichlorodiphenyl dichloroethanes OR dichlorodiphenyl dichloroethanes OR dichlorodiphenyl dichloroethanes OR dichlorodiphenyl dichloroethanes OR dichlorodiphenyl dichloroethanes |
|                     | MeHg.ti,ab,kw. |                         |                            |
|                     | polychlorinated biphenyl*.ti,ab,kw. |                         |                            |
|                     | PCB*.ti,ab,kw. |                         |                            |
|                     | Aroclor*.ti,ab,kw. |                         |                            |
|                     | kanechlor*.ti,ab,kw. |                         |                            |
|                     | clophen*.ti,ab,kw. |                         |                            |
|                     | phenoclor*.ti,ab,kw. |                         |                            |
|                     | pyralene*.ti,ab,kw. |                         |                            |
|                     | fenclor*.ti,ab,kw. |                         |                            |
|                     | sovol*.ti,ab,kw. |                         |                            |
|                     | chlorfen*.ti,ab,kw. |                         |                            |
|                     | delor*.ti,ab,kw. |                         |                            |
|                     | methylmercury compounds OR polychlorinated biphenyls OR Aroclors OR hydrocarbons, chlorinated OR aldrin OR chlordane OR chlordanes OR chlordecone OR chloroform OR ddt OR dichlorodiphényl dichloroéthanes OR dichlorodiphenylether OR dichloroéthanes OR chlorobenzilate OR chloropicrin OR chlorothalonil OR dacthal OR dicofol OR dichlorodiphenyl dichloroethanes OR dichlorodiphenyl dichloroethanes OR dichlorodiphenyl dichloroethanes OR dichlorodiphenyl dichloroethanes OR dichlorodiphenyl dichloroethanes |
| Keywords Subject Headings (MeSH) | Subject Headings (Emtree) |
|---------------------------------|--------------------------|
| organochlorine pesticide*.ti,ab,kw. |                           |
| aldrin*.ti,ab,kw.                |                           |
| chlordane*.ti,ab,kw.            |                           |
| chlordan*.ti,ab,kw.             |                           |
| chlordan*.ti,ab,kw.             |                           |
| chlordecone*.ti,ab,kw.          |                           |
| chloroacetate*.ti,ab,kw.        |                           |
| chlorobenzene*.ti,ab,kw.        |                           |
| chlorofluorocarbon*.ti,ab,kw.   |                           |
| chloroform*.ti,ab,kw.           |                           |
| chloromethane*.ti,ab,kw.        |                           |
| DDT*.ti,ab,kw.                  |                           |
| dichlorodiphenyldichloroethylene*.ti,ab,kw. |                     |
| dichlorodiphenyldichloroethane*.ti,ab,kw. |                   |
| dieldrin*.ti,ab,kw.             |                           |
| endrin*.ti,ab,kw.               |                           |
| ethyl chloride*.ti,ab,kw.       |                           |
| ethylene dichloride*.ti,ab,kw.  |                           |
| heptachlor*.ti,ab,kw.           |                           |
| hexachlorocyclohexane*.ti,ab,kw. |                          |
| methoxychlor*.ti,ab,kw.         |                           |
| methyl chloride*.ti,ab,kw.      |                           |
| methylene chloride*.ti,ab,kw.   |                           |
| mirex*.ti,ab,kw.                |                           |
| tetrachloroethylene*.ti,ab,kw.  |                           |
| toxaphene*.ti,ab,kw.            |                           |
| Trichloroepoxypropane*.ti,ab,kw.|                           |
| Trichloroethylene*.ti,ab,kw.    |                           |
| Trichloroethylene*.ti,ab,kw.    |                           |
| northern contaminant mixture*.ti,ab,kw. |                      |
| NCM*.ti,ab,kw.                  |                           |
| (north* adj3 (pollutant* or toxicant* or contaminant*).ti,ab,kw. |         |

Concept 3: Outcome

| Keywords Subject Headings (MeSH) | Subject Headings (Emtree) |
|---------------------------------|--------------------------|
| brain*.ti,ab,kw.                | behavior OR maternal behavior OR brain |
| behavior?.ti,ab,kw.             | behavior OR maternal behavior OR brain |
Table 2. Keywords used for Toxline searches.

| Concept 1: Population | Keywords |
|-----------------------|----------|
| pregnancy OR pregnant OR perinatal OR maternal OR postpartum OR antenatal OR gestation OR gestational |

| Concept 2: Exposure | Keywords |
|---------------------|----------|
| methylmercury OR MeHg OR polychlorinated biphenyl OR pcb OR Aroclor OR kanechlor OR clophen OR phenoclor OR pyralene OR fenclor OR sovol OR chlorfen OR delor OR organochlorine pesticide OR aldrin OR chlordane OR chlordan OR chlordane OR chloracetate OR chlorobenzene OR chlorofluorocarbon OR chloroform OR chloromethane OR ddt OR dichlorodiphenyl dichloroethylene OR dichlorodiphenyldichloroethane OR dichloroethylene OR dieldrin OR endrin OR ethyl chloride OR ethylene dichloride OR heptachlor OR hexachlorocyclohexane OR methoxychlor OR methylychloride OR methylene chloride OR mirex OR tetrachloroethylene OR toxaphene OR trichloroepoxypropane OR trichloroethylene OR trichloroethylene OR northern contaminant mixture OR ncm OR northern pollutant OR northern toxicant OR northern contaminant |

| Concept 3: Outcome | Keywords |
|--------------------|----------|
| behavior OR behaviour OR brain |

2.3. Selection of Studies

All studies included in the analysis: (1) involved a human cohort or animal model; (2) included direct or indirect exposure to MeHg, PCB congeners, OCPs, or co-exposure to any of the three toxicants; (3) included maternal behavioural or molecular assessments of the maternal brain; (4) were primary source literature. Studies were excluded from the analysis if the full text was unavailable.

Titles and abstracts were first screened to eliminate articles irrelevant to the research objective. Full-text articles for the remaining studies were reviewed to determine eligibility for inclusion in the scoping review. Articles were screened by one reviewer (C.F.M.), and any uncertainties in study selection were discussed with the principal investigator (A.T.M.K.).

2.4. Charting the Data

Nineteen included studies were reviewed and information from each article was abstracted, including: year of study, toxicant of interest, objective, study design, maternal subjects, sample size, treatment groups, exposure route, exposure period, behavioural findings, and neurochemical findings.

2.5. Collating, Summarizing, and Reporting the Results

Data extracted from the full-text review were organized using a Microsoft Excel spreadsheet (Microsoft Corporation, Redmond, WA, USA). Studies included in the analysis are shown in Table 3.

3. Results

A total of 1194 studies were identified after de-duplication for possible inclusion in the review. A total of 1138 studies were excluded based on the criteria for inclusion listed in Section 2.3. After title and abstract screening, 56 studies remained that were included in the full text review. Eighteen articles met the criteria for inclusion in the scoping review. One study further analysed original data from a previous study, which was subsequently included. Full details of the scoping review process are displayed in Figure 1. All studies used experimental animal models, except for one human cohort study. Many studies reported both maternal and offspring outcomes. For the purpose of this review, only the maternal findings are presented in Table 3; please refer to this table for additional details regarding each study.
3.1. Effects of Methylmercury on the Maternal Brain or Behaviour

3.1.1. Maternal Behaviour

In one study, Weston et al. (2014) used rats to monitor maternal behaviours, including passive nursing, arched back nursing, blanket nursing, pup licking and grooming, pup licking, no contact and no contact resting following the administration of MeHg [64]. Mothers (dams) were separated into treatment groups exposed to 0, 0.5, or 2.5 ppm MeHg drinking water. This was administered two to three weeks prior to breeding until offspring weaning. The authors did not report the amount of water consumed in each experimental group, making it difficult to deduce the actual dose consumed. Changes in maternal behaviour attributed to MeHg exposure alone were limited, with no significant group differences [64]. As such, this study suggests a limited effect of MeHg on maternal rodent behaviour.

In a second rodent study, rat dams were exposed to MeHg at 0 or 0.5 mg/kg body weight/day and retinyl palmitate (Vitamin A), either alone or in combination throughout gestational day (GD) 0 to postnatal day (PND) 21 [65]. Similar to the Weston et al. study, no differences in nursing and pup retrieval behaviours were observed between control and treated groups [65]. However, changes in redox parameters in the maternal brain were observed, as described in Section 3.1.2 below.

Another study using a rodent model investigated different speciations of MeHg and their effects on behaviour [66]. Mouse dams were exposed to 0, 1.5, or 4.5 mg/kg methylmercury chloride (MeHgCl) or methylmercury cysteine (MeHgCys) diets ad libitum from six weeks prior to mating until two weeks following birth [66]. Based on the measured feeding rates, mice in the low dose MeHg groups consumed 223–250 µg/kg, and the high dose consumed 596–629 µg/kg body weight per day of MeHgCys or MeHgCl. The high MeHgCl-exposed group exhibited significantly decreased exploratory behaviour compared to the control group. In the elevated plus maze, this group displayed an increased latency to move from the center section compared to the high MeHgCys diet and control group [66]. The elevated plus maze is widely used to assess anxiety-like behaviour in rodent models [67]. This study suggests that high dose MeHgCl may cause anxiety-like behaviours in rodent dams.
Lastly, a recent study using an avian model explored the effects of MeHg on avian parental behaviour such as nest building, incubation behaviour and provisioning behaviour [68]. Zebra finches were exposed to 0 or 1.2 ppm wet weight MeHg through lifetime dietary exposure. MeHg exposed pairs spent less time constructing nests and built lighter nests, but both variables were also influenced by male age and mass [68]. Control pairs had a greater proportion of successful nest-building trips (pieces of hay brought to the nest compared to number of attempts), but did not differ in amount of hay compared to MeHg-treated finches [68]. As such, the authors suggest a potential compensatory effect of more nest-building trips [68].

3.1.2. Maternal Brain

In the MeHg and retinyl palmitate (Vitamin A) study, hippocampal catalase and glutathione peroxidase activity were also investigated. These two enzymes have critical functions in reducing oxidative damage by detoxication of hydrogen peroxide [69]. Hippocampal catalase activity was reduced in both the MeHg and VitA groups but not the combination treatment. Glutathione peroxidase activity also decreased in MeHg-treated rats, as well as in the combination treatment group [65]. In the prefrontal cortex, total reduced thiol content, a key indicator of redox status, was significantly increased in the MeHg–VitA group [65,70]. No significant redox profile changes were observed in the olfactory bulbs of treated dams compared to control dams [65]. These results collectively suggest a potential adaptive response by which the increase in total thiol content acts to decrease the toxic effects of MeHg. Conversely, decreases in both catalase and glutathione peroxidase activity suggest a toxic effect of MeHg on the maternal hippocampal region. Further evaluation of the transcript and protein levels of catalase and glutathione peroxidase would clarify these findings.

Another study examined the effects of MeHg exposure on glutamatergic homeostasis and oxidative stress in the cerebellum of mice [71]. Exposed dams received drinking water ad libitum with 15mg/L of MeHg from PND1 to PND21. The estimated daily dose of MeHg was 8.25 mg/kg body weight based on the liquid intake per day. No differences in cerebellar glutamate uptake, levels of total sulfhydryl groups, nonprotein sulfhydryl groups, and nonprotein hydroperoxide were observed between control and MeHg-exposed dams [71]. Cerebellar catalase activity showed no difference between groups, while glutathione peroxidase activity significantly decreased in MeHg-exposed dams [71]. Thus, this study suggests a slight neurotoxic effect of MeHg, as decreases in glutathione peroxidase activity limit the brain tissue antioxidant capacity, which is consistent with previous findings [72].

3.2. Effects of PCBs on the Maternal Brain or Behaviour

3.2.1. Maternal Behaviour

Two studies investigated the effects of PCB 77 (CASRN 32598-13-3) on maternal behaviour. PCB 77 is reported in a wide range of aquatic and mammalian species, and the high toxicity of PCB 77 is attributed to its coplanar structure [73,74]. One study found that dams administered 4mg/kg PCB 77 (s.c.) daily from GD6 to GD18, spent significantly more time in the nest compared to the control group and more time licking and grooming their pups than the control dams or those having received 2 mg/kg/day of PCB 77 [75]. Although the proportion of time nursing was unaffected by the PCB treatments, there was a statistically significant difference in the proportion of total time nursing in the high-crouch posture specifically, between the exposed and control groups [75]. Both PCB-treated groups showed significantly less high-crouch nursing compared to the control group, with no significant difference between the two PCB doses [75].

The second study used a cross-fostering design to explore direct and indirect effects of PCB 77 exposure on maternal behaviour [76]. Dams were exposed to corn oil or 2 mg/kg PCB 77 bodyweight/daily (s.c.) from GD16 to GD18 and pups were cross-fostered or raised by their birth mothers, which resulted in four treatment groups: (1) PCB-exposed dams and their pups or PCB-exposed dams and cross-fostered PCB-exposed pups (data combined), (2) PCB-exposed dams and vehicle
(oil)-treated pups, (3) oil-treated dams and PCB-exposed pups, (4) oil-treated dams and their pups or oil-treated dams and cross-fostered oil-exposed pups (data combined). When the data from all PCB groups (1, 2, and 3) were combined, it was shown that dams spent significantly more time on the nest compared to the vehicle-only control group [76]. As well, pup grooming and number of nursing bouts were increased in dams from the PCB treatment groups. There were no differences in amount of time nursing between the PCB groups and oil groups, but PCB groups displayed the high-crouch nursing posture significantly less than the oil-only group [76].

Another study investigated maternal PCB exposure using an avian model with exposure from one month prior to pairing, lasting until the hatching of eggs [77]. Adult captive kestrel pairs were administered Aroclors 1248, 1254, and 1260 commercial mixtures consisting of multiple PCB congeners [78]. Birds consumed day-old cockerels injected with the Aroclor mixture, thereby intaking 5-7 µg/g body weight PCBs daily. During the incubation period, 8% of PCB-exposed pairs abandoned their clutches prior to hatching compared to 0% of the control pairs, a difference reported to have a medium effect size [77].

Dover et al. (2015) used a mixture of PCB 47 (CASRN 2437-79-8) and PCB 77 (equal parts) at 25 mg/kg wet weight dietary exposure from GD0 to parturition to examine effects on maternal behaviour and underlying molecular mechanisms in rat dams [79]. PCB 47 is a non-coplanar congener which is less toxic but more frequently identified in environmental samples than PCB 77 [73]. Although the authors provided values for food intake, estimated PCB exposure from food intake was not reported. The proportion of time spent in low-crouch nursing posture and high-crouch nursing posture significantly increased compared to control dams on PND4 and PND6, respectively [79]. No effects of treatment were found for the remaining maternal behaviours assessed, including active nursing, pup licking, maternal auto-grooming, time off nest, and resting nursing. Nest building was assessed with results showing that PCB-exposed dams used more nesting strips on GD20 compared to the control group, but the overall quality of nest did not differ significantly between groups [79].

3.2.2. Maternal Brain

From the PCB 47 and PCB 77 exposure study, analysis of the maternal hypothalamus revealed an increased expression of the oxytocin receptor (OXTR) gene in the PCB-treated dams that fostered PCB-exposed pups and cross-fostered control pups. This receptor and the oxytocin ligand play a key role in mediating the effects of estrogen on the initiation of maternal behaviour [80,81]. Hypothalamic Cyp1a1 expression did not differ between groups [79]. The CYP1A1 enzyme belongs to the cytochrome P450 (CYP450) family of enzymes, which metabolize xenobiotic substances and certain endogenous compounds [82,83]. As such, this study suggests that PCB exposure affects OXTR expression, which in turn may affect maternal behaviour.

Another study examined CYP1A1/2 AND CYP1B1/2 protein expression following PCB exposure. Rat dams were administered a mixture of PCB 138 (CASRN 35065-28-2), 153 (CASRN 35065-27-1), 180 (CASRN 35065-29-3), and 126 (CASRN 35465-28-8) from GD15 to GD19 at 0 or 10 mg/kg/day (s.c.) [84]. PCB 138, 153, and 180 are highly abundant noncoplanar congeners, whereas PCB 126 is less abundant but highly toxic [73,85–87]. PCB exposure did not induce higher CYP1A or CYP2B expression compared to the control dams, as determined by protein analysis in total brain samples [84]. Similar to the Dover et al., this study reports no changes in CYP450 metabolism of PCBs in the maternal brain.

Honma et al. (2009) used the PCB 153 congener to study alterations in neurotransmitter levels and their metabolites [88]. Dams were administered PCB 153 at 0, 16, or 64 mg/kg body weight by daily oral gavage treatment through GD10 to GD16. Multiple brain regions were analysed including the occipital cortex and hippocampus, which displayed significant decreases in dopamine (DA), DOPAC, and homovanillic acid (HVA) levels for each PCB-treated group compared to the control [88]. In the striatum, HVA levels decreased significantly in the higher dose PCB group. In the hypothalamus, HVA and HVA/DA ratios decreased significantly in the high dose PCB group, while serotonin levels increased significantly in the same group. In the medulla oblongata, DA levels were significantly
decreased in the high dose PCB group [88]. Many other neurotransmitter levels and ratios were altered but did not reach statistical significance.

3.3. Effects of OCPs on the maternal Brain or Behaviour

3.3.1. Maternal Behaviour

Three studies investigating OCP exposure focused on maternal behaviour outcomes. Matsuura et al. conducted a reproductive toxicity study using lindane, a pesticide which has been banned for agricultural use [89]. Rat dams were given a diet including 0, 10, 60, or 300 ppm lindane for 10 weeks before mating until PND21. Based on daily food intake, the 10 ppm group consumed $0.573 \pm 0.0328$ and $1.525 \pm 0.075$ mg lindane/kg body weight per day during the gestational and lactational periods, respectively. The 60 ppm group consumed $3.389 \pm 0.167$ and $8.941 \pm 0.677$, while the 300 ppm lindane group consumed $16.55 \pm 0.95$ and $45.21 \pm 3.54$ mg/kg body weight per day during the gestational and lactational periods, respectively. Lack of retrieval behaviour and consequential litter loss were observed in one 300 ppm lindane-exposed dam [90]. Other than the one case, lindane exposure did not affect any maternal behaviours, including lactation, nest building, and cannibalism [90].

Another study using methoxychlor, a synthetic OCP, examined the effects of maternal exposure from GD11 to GD17 [91]. Doses of methoxychlor at 0, 20, 200, or 2000 µg/kg body weight/day were administered to dams by oral administration from a modified syringe. Compared to the control dams, dams exposed to the lowest methoxychlor dose spent less time nursing, less time in the nest, more time eating and resting outside the nest during the dark period [91]. Within-group post hoc analysis revealed early onset decline in maternal behaviour of the methoxychlor-exposed dams. Compared to PND2, control dams spent less time nursing and in the nest from PND11 onwards while the lowest dose methoxychlor group showed these behavioural changes from PND4 onwards. Similarly, control dams increased time eating and resting at PND15, and the lowest dose methoxychlor group displayed these increases at PND5 for eating and PND7 for resting [91].

One study in this scoping review assessed maternal toxicant exposure in humans [92]. Four assessments were used to evaluate maternal psychopathologies including the Brief Symptom Inventory (BSI), Postpartum Bonding Questionnaire (PBQ), Mother to Infant Bonding Scale (MIBS), and Edinburgh Postnatal Depression scale. High scores on these assessments suggest maternal psychopathologies or infant bonding issues. Breast milk was analysed at the eighth month postpartum for 12 OCPs. Of the 12 OCPS, heptachlor epoxide levels positively correlated with PBQ scores, MIBS scores, and three indexes of the BSI, including the global severity index, positive symptom total index, and positive symptom distress index [92]. As well, five subscales of the BSI correlated positively with heptachlor epoxide levels, specifically somatization, depression, anxiety, hostility, and phobic anxiety [92]. Note that this study was included even though our search was not specific to maternal psychopathologies; a search with keywords specific to maternal psychopathologies may have yielded additional results.

3.3.2. Maternal Brain

No data were found pertaining to the effects of OCP exposure on the maternal brain.

3.4. Effects of Toxicant Co-Exposure on the Maternal Brain or Behaviour

3.4.1. Maternal Behaviour

One study conducted in Scandinavia exposed mouse dams to an environmentally relevant mixture of 29 organic pollutants, including multiple PCB congeners, OCPs, brominated compounds, and perfluorinated compounds [93]. Dams were exposed to 0, 5000, or 100,000 times the estimated daily intake for humans through dietary feed. Exposure began when the dams were young pups, from weaning through the duration of their pregnancies to project completion. The open field test was used to examine anxiety-like behaviours and locomotion. Exposure to the POP mixture at either dose
had no effect on the behavioural endpoints for dams, including time spent within the different zones, total distance moved, or velocity [93]. Note that brominated or perfluorinated compounds may have impacted any effects of the PCBs or OCPs as they were not studied in isolation.

Another study investigated toxicant co-exposure using Glaucous gull pairs in two different Norwegian breeding regions [94]. Blood concentrations of multiple toxicants including 8 PCB congeners, \( p,p' \)-DDE, HCB and oxychlordane were measured, and avian parental behaviours were assessed. PCB concentrations in parental pairs were significantly related to the proportion of time away from the nest when not incubating. As well, increased PCB concentrations were related to the number of absences from the nest [94]. These data were later reanalysed and both PCB and oxychlordane blood concentrations were significantly and positively correlated with time away from the nest when not incubating [95]. No significant effects of \( p,p' \)-DDE or HCB were reported [95].

3.4.2. Maternal Brain

Two studies used rodent models to identify potential neurotoxic effects of co-exposure to MeHg and PCB 153 on the cholinergic system. In the first study, rat dams were exposed to 0, 0.5, or 1.0 mg/kg MeHg body weight per day alone or in combination with PCB 153 treatment at 20 mg/kg/day [96]. MeHg treatment spanned GD7 to PND7, while PCB was administered from GD10 to GD16. These dosages and exposure periods followed those used in previous studies demonstrating neurochemical and behavioural changes in adult rats [97–100]. Dams from the higher exposure MeHg group, the PCB group, and both co-exposed groups each had significant increases in muscarinic receptor (MR) density in the cerebral cortex compared to the control group [96]. In the cerebellum, MR density significantly increased in the high dose MeHg group, while PCB 153 exposure resulted in significantly decreased MR density compared to the control group. Both co-exposure groups had significant decreases in MR density similar to the PCB group [96]. No significant changes in MR density were observed for the low dose MeHg-exposed group in the cerebral cortex or cerebellum. The hippocampal and striatal brain regions did not express any changes in MR density following any treatment. Treatment did not affect the MR dissociation constant in any brain area [96].

Roda et al. [101] used the same dosing regime as the above study to further investigate the potential role of alterations in the cholinergic systems as biomarkers for MeHg and PCB-associated neurotoxicity. In this experiment, dams exposed to the higher dose of MeHg expressed a significant increase in cerebellar MR density [101] (Table 3). Exposure to the lower dose of MeHg, PCB 153, and either MeHg dose in combination with PCB 153 did not result in significant MR density changes. Again, MR dissociation constants did not differ between groups. As well, monoamine oxidase B activity did not differ between the treatment groups and the control group [101].
| Reference Number | Study | Toxicant(s) of Interest | Objective | Study Design | Maternal Subjects | Toxicant Treatment Group(s) | Exposure Route | Exposure Period | Behavioural Findings | Neurochemical Findings |
|------------------|-------|-------------------------|-----------|--------------|------------------|-----------------------------|----------------|-----------------|----------------------|----------------------|
| [65]             | Espitia-Pérez et al., 2018 | MeHg | Examine the effects of MeHg and VitA co-exposure on pregnant and lactating Wistar rats to evaluate behavioural and biochemical changes in brains of the dams and their offspring | Experimental design | Wistar rats N = 30 | 0.5mg/kg body weight/day MeHg | Oral gavage treatment | GD0 to PND21 | No differences in nursing and pup retrieval behaviours were observed between treated dams and controls. | Hippocampal catalase activity was reduced in both the MeHg and VitA groups, while glutathione peroxidase activity decreased in both the MeHg and MeHg-VitA-treated groups. In the prefrontal cortex, total reduced thiol content significantly increased in the MeHg-VitA group. No significant redox profile changes were observed in the olfactory bulbs of treated dams. |
| [93]             | Hudecova et al., 2018 | PCBs: 28, 52, 101, 118, 138, 153, 180 OCPs: p,p'-DDE, HCB, α-chlordane, oxychlordane, trans-nonachlor, α-HCH, β-HCH, γ-HCH, dieldrin 7 BFRs 6 PFAAs | Determine whether a POP mixture relevant to human exposure levels affects basal corticosterone levels, anxiety-like behavior, and locomotor activity in female mice and their offspring | Experimental design | 129:CS7BL/6F1 hybrid female mice N = 47 | 5000 or 100,000x EDI toxicant mixture | Dietary exposure | From weaning prior to mating until project completion | Exposure to the POP mixture at either dose had no effect on the endpoints of the open field behavioural test for dams including: time spent within the different zones, total distance moved, or velocity. | NR |
| [68]             | Chin et al., 2017 | MeHg | Determine the effects of MeHg exposure on avian parental behavior and reproductive success in zebra finches | Experimental design | Zebra finch pairs N = 87 (N = 73 initiated nests) | 1.2ppm MeHg wet weight | Dietary exposure | From in ovo through maturity | MeHg-exposed pairs spent less time constructing and built lighter nests (both influenced by male age and mass). Control pairs had greater efficiency in bringing hay to the nest, but did not differ in amount of hay compared to MeHg-treated finches, suggesting a compensatory effect of more trips made by the MeHg-treated finches. | NR |
| [79]             | Dover et al., 2015 | PCBs: 47 and 77 | Examine possible molecular mechanisms underlying changes in maternal care behaviour due to PCB exposure | Experimental design | Sprague-Dawley rats N = 11 | 25mg/kg wet weight PCB 47 and 77 | Dietary exposure | GD0 to PND0 | PCB altered nest building and maternal care behaviours. Specifically, there was a significant increase in time spent in low crouch and high crouch nursing posture on PND4 and PND6 respectively. | Molecular analysis revealed an increased OXTR expression in the hypothalamus of dams exposed to PCBs. |
Table 3. Continued.

| Reference Number | Study | Toxicant(s) of Interest | Objective | Study Design | Maternal Subjects | Toxicant Treatment Group(s) | Exposure Route | Exposure Period | Behavioural Findings | Neurochemical Findings |
|------------------|-------|-------------------------|-----------|--------------|------------------|---------------------------|----------------|-----------------|-----------------------|------------------------|
| [92]             | Yalçın et al., 2015 | OCPs: α-HCH, β-HCH, Y-HCH, aldrin, dieldrin, heptachlor, heptachlor epoxide, α-endosulfan, β-endosulfan, trans-chlordane, cis-chlordane, DDT | Assess detectable OCPs in maternal breast milk to evaluate the relation between OCPs and maternal psychopathologies | Correlational design | Human mothers N=75 | NR NR NR | Heptachlor epoxide levels were positively associated with PBQ scores, MIBS scores, and three indexes of the maternal BSI (the global severity index, positive symptom total index and positive symptom distress index) and five subscales of the maternal BSI (depression, hostility, anxiety, phobia, and somatic symptoms). | NR |
| [64]             | Weston et al., 2014 | MeHg | Examine the effects of MeHg and prenatal stress on maternal and infant behaviour and neurochemical markers | Experimental design | Long-Evans rats N = 66 (N = 24 for behavioural testing) | 0.5 or 2.5 ppm MeHg | Drinking water | 2 to 3 weeks before breeding to post-weaning | Changes in maternal behavior, attributed to MeHg exposure alone, were extremely limited. | NR |
| [101]            | Roda et al., 2012 | MeHg PCB 153 | Evaluate brain and lymphocyte muscarinic receptors and cerebral monoamine oxidase-B activity as potential biomarkers for assessing exposure to environmental toxicants | Experimental design | Sprague-Dawley rats N = 12 per set of experiment | 0.5 or 1mg/kg body weight/day MeHg and/or 20mg/kg/day PCB 153 | Drinking water (MeHg) Oral gavage treatment (PCB 153) | GD7 to PND07 (MeHg) GD10 to GD16 (PCB 153) | Cerebellar muscarinic receptor density increased (87%) with exposure to the higher MeHg dose, while no changes were observed in the lower MeHg dose, PCB dose or PCB coexposed group. The muscarinic receptor (MR) cerebellar dissociation constants were not altered in any of the treatment groups. Cerebellar MAO-B activity did not differ between any treatment group and the control. | NR |
| [84]             | Bonfanti et al., 2009 4 | PCBs: 138, 153, 180 and 126 | Investigate PCB disposition in two maternal and fetal rat organs for toxic implications | Experimental design | Sprague-Dawley rats N=10 | 10mg/kg body weight/day PCB 138, 153, 180, 126 mixture | Subcutaneous injection | GD15 to GD19 | CYP1A and CYP2B levels were determined in maternal brains. | NR |
| [66]             | Glover et al., 2009 | MeHg | Compare the effects of MeHgCl and MeHgCys on the accumulation, brain gene expression, and behavior of mice | Experimental design | Balb/c mice N=32 (N=31 due to MeHgCys adverse toxicity) | 1.5 or 4.5 mg/kg MeHgCys or MeHgCl | Dietary exposure | 6 weeks prior to mating through to 2 weeks following birth | High MeHgCl diet group exhibited reduced exploratory behaviour compared to the control, and increased latency to move compared to the control and high MeHgCys exposure groups. | NR |
Table 3. Cont.

| Reference Number | Study          | Toxicant(s) of Interest | Objective                                                                 | Study Design | Maternal Subjects | Toxicant Treatment Group(s) | Exposure Route | Exposure Period | Behavioural Findings | Neurochemical Findings |
|------------------|----------------|-------------------------|---------------------------------------------------------------------------|--------------|-------------------|-----------------------------|----------------|-----------------|----------------------|------------------------|
| [88]             | Honma et al., 2009 | PCB 153                | Investigate the effects of PCB administration on cerebral neurotransmitters and related substances in rat dams and offspring | Experimental design | Crj:CD(SD)ICR rats N=30 | 16 or 64 mg/kg/weight/day PCB 153 | Oral gavage treatment | GD10 to G16 | NR | In the occipital cortex, DA, DOPAC, and HVA levels decreased in both PCB-treated groups. In the hippocampus, DA, DOPAC, and HVA levels decreased by 30% and 40% in the 16 mg/kg/bw and 64 mg/kg/bw, respectively. In the striatum, HVA decreased significantly in the 64 mg/kg/bw group. In the hypothalamus, HVA and HVA/DA ratios decreased significantly in the 64 mg/kg/bw day group. SHT increased significantly in the same group. In the medulla oblongata, DA level decreased significantly in the 64 mg/kg/bw group. |
| [96]             | Coccini et al., 2006 | MeHg PCB 153           | Determine whether MeHg and PCB 153 alter the MRs in the cerebral cortex, cerebellum, hippocampus and striatum | Experimental design | Sprague-Dawley ratsN=24 per set of experiment | 0.5 or 1.0mg/kg/day MeHg and/or 20mg/kg/day PCB 153 | Drinking water (MeHg) Oral gavage (PCB 153) | GD7 to PND7 (MeHg) GD20 to GD16 (PCB 153) | NR | Cerebral cortex MR density increased for 1.0 mg/kg/day MeHg, PCB 153, and both MeHg+PCB153 treatments groups (60% MeHg group, 47% PCB group, 45% 1.0 MeHg/kg/day+PCB153, 42% 0.5MeHg/kg/day+PCB153). Treatment with MeHg at the higher dose resulted in increased cerebellar MR density (87%), while PCB 153 exposure resulted in significantly decreased MR density (27%). Both combined exposure groups resulted in MR density similar to the PCB-exposed group. The lower MeHg dose did not cause any changes in MR density in the cerebellum of dams. In the hippocampus and striatum, no MR density changes were observed following any treatment or combination of treatments. In all brain areas, the dissociation constant values for MR were not altered. |
| Reference Number | Study | Toxicant(s) of Interest | Objective | Study Design | Maternal Subjects | Toxicant Treatment Group(s) | Exposure Route | Exposure Period | Behavioural Findings | Neurochemical Findings |
|------------------|-------|------------------------|-----------|-------------|------------------|--------------------------|----------------|----------------|----------------|---------------------|
| [95] Bustnes et al., 2005 | PCBs: 99, 118, 138, 153, 170, and 180, 3 OCPs: Oxychlordane, p,p'-DDE, HCB | Analyse four fitness components (time spent away from nest, early chick growth and return rate) in relation to blood residues of PCBs, OCPs in Glaucous gulls | Correlational design | Glaucous gulls N=16 | NR | NR | Lifetime | PCBs and oxychlordane were positively and significantly related to time spent away from the nest site when not incubating. DDE and HCB levels had no effect on this trait. | NR |
| [62] Cummings et al., 2005 | PCB 77 | Differentiate between direct and indirect effects of PCB exposure on maternal behaviour | Experimental design | Long-Evans rats N=36 | 2mg/kg body weight/day PCB77 | Subcutaneous injection | GD6 to GD18 | Dams exposed to PCBs during pregnancy spent more time on the nest and more time grooming (and licking) the pups, when compared to control dams. | NR |
| [90] Matsuura et al., 2005 | OCP: Y-HCH (lindane) | Assess the endocrine disruption activity and toxicity of lindane using additional toxicological and behavioral endpoints | Experimental design | C57CD(SD)IGS female ratsN=24 | 10, 60, or 300 ppm lindane | Dietary exposure | 10 weeks before mating until PND21 | Lindane exposure did not affect any maternal behaviours including lactation, nest building and cannibalism. Lack of retrieval behaviour and consequential litter loss was observed in one 300 ppm lindane-exposed dam. | NR |
| [75] Simmons et al., 2005 | PCB 77 | Investigate the effects of PCB exposure on the behavior of dams as they rear exposed litters | Experimental design | Long-Evans rats N=21 (N=19 due to failed delivery of 2 dams) | 2mg/kg body weight/day or 4mg/kg/body weight/day PCB 77 | Subcutaneous injection | GD6 to GD18 | PCB 77 exposure resulted in changes in maternal behaviour including: an increase in time spent on the nest, increase in licking and grooming of the offspring, and a decrease in the display of the high-crouch nursing posture. | NR |
| [71] Manfroi et al., 2004 | MeHg | Investigate the effects of lactational MeHg exposure on neurotoxicity and glutamatergic transmission | Experimental design | Swiss Albino mice N=14 | 15mg/l MeHg | Drinking water | PND1 to PND21 | MeHg exposure did not alter glutamate uptake in cerebellar slices of dams. Cerebellar levels of total and nonprotein sulfhydryl groups and nonprotein hydroperoxide did not differ between control and MeHg-treated dams. MeHg exposure inhibited the activity of cerebellar glutathione peroxidase but had no effects of cerebellar catalase activity. | NR |
| [77] Fernie et al., 2003 | PCBs: Aroclors 1248, 1254, 1260 | Identify short and long-term abnormal development and behavior of American kestrels through all stages of the breeding season from parental PCB exposure | Experimental design | American kestrel pairs N=50 | 5-7µg/g body weight/day Aroclor 1248, 1254, 1260 mixture | Dietary exposure | 1 month prior to pairing until anticipated hatching of eggs | 8% of PCB-exposed pairs abandoned their clutches prior to hatching. There were no incidences of altered incubation behavior in the PCB-exposed pairs of the next breeding season. | NR |
Table 3. Cont.

| Reference Number | Study | Toxicant(s) of Interest | Objective | Study Design | Maternal Subjects | Toxicant Treatment Group(s) | Exposure Route | Exposure Period | Behavioural Findings | Neurochemical Findings |
|------------------|-------|-------------------------|-----------|--------------|-------------------|-----------------------------|----------------|-----------------|---------------------|------------------------|
| [91]             | Palanza et al., 2002 | OCP: methoxychlor | Investigate the effects of maternal exposure to methoxychlor on behaviour responses of dams and their offspring | Experimental design | CD-1 mice N=72-84 treated (N=64-80 for data analysis) | 20, 200, or 2000 µg/kg body weight/day methoxychlor | Modified oral gavage treatment | GD11 to GD17 | Dams exposed to the low dose of methoxychlor spent lower amounts of time in the nest, nursing and more time eating and resting outside the nest during the dark period. | NR |
| [94]             | Bustnes et al., 2001 | PCBs: 28, 101, 99, 118, 138, 153, 170 and 180 | Investigate the effects of PCB contamination on nesting behaviour in Glaucous gulls | Correlational design | Glaucous gulls N=16 | NR | NR | Lifetime | Time away from the nest and proportion of time absent from the nest was significantly related to PCB concentration in blood. | NR |

NR: not reported; Note: subgroups of the maternal subjects were used in multiple studies as indicated by the sample sizes in brackets. 1: Relative amounts of the 29 toxicants are based on the estimated daily intake of Scandinavians. 2: Equal parts PCB 47 and 77 were added to the chow diet. 3: Equal concentration of PCB 138, 153, and 180 were included in the PCB mixture, and PCB 126 was added at a 1:10000 ratio. 4: Equal weight of Aroclors 1248, 1254, and 1260 were added to frozen cockerel diet.
4. Discussion

This scoping review demonstrated the limited number of studies investigating behavioural and neurochemical changes resulting from maternal toxicant exposure. Limited changes in maternal behaviour were reported for MeHg-treated dams, while many behavioural changes were observed in maternal PCB exposure studies. Animal studies investigating OCPs focused on behavioural assessments in which effects were observed at the lowest and highest doses of two different OCPs. One human study showed a positive correlation between OCP levels and maternal psychopathology assessments. Studies involving co-exposure to multiple toxicants were limited in behavioural findings, with the exception of the correlational avian research study.

Two maternal MeHg exposure studies described change in redox status of the brain, where both increases and decreases were documented. Changes in neurotransmitter levels, gene expression, and protein expression in the brain were reported in three PCB exposure studies. Two co-exposure studies described alterations to the cholinergic system in multiple brain regions, and there were no studies that reported effects of OCPs on neurochemical measures in this review. Note that to our knowledge, no studies have investigated the effects of the mixture considered the Northern Contaminant Mixture on maternal behaviour or related modifications to the brain.

4.1. Changes to Maternal Behaviour

4.1.1. Rodent Maternal Care Behaviours

Rat dams exposed to PCB 77 at a higher dose (4mg/kg bw/day) spent more time licking and grooming the pups when compared to dams not exposed to PCBs [75]. Similar findings were shown in a cross-foster design study whereby combined data from PCB-exposed rat dams (2 mg/kg bw/day) rearing PCB-exposed pups and non-exposed dams rearing PCB-exposed pups displayed increases in time grooming as well as in licking and nursing bouts [76]. Recent literature using a glyphosate-based herbicide (Roundup) presents similar increases in maternal licking behaviour [102]. Conversely, maternal bisphenol A exposure has shown significant reductions in licking and grooming behaviour [103]. Maternal behaviours including licking and grooming have been shown to be stable across litters, so changes in these behaviours may function to mediate harmful effects of early environmental stressors [57].

The same PCB 77 groups above also spent more time on the nest [75,76]. On the contrary, results from an OCP exposure study showed that mouse dams exposed to low dose methoxychlor spent less time in the nest [91]. Mouse dams treated with bisphenol A have similarly shown increases in time spent out of their nest [104]. Total amounts of maternal care in the methoxychlor study were not different between treatment groups, but alterations in the onset and decline of maternal behaviours were observed.

PCB groups in both studies showed significantly less high-crouch nursing [75,76]. Contradicting results were shown with a dietary exposure study using PCB 47 and 77, as a proportion of time spent in high-crouch nursing posture increased on PND6 [79]. As the authors noted, these results could be attributable to procedural differences and different mechanisms of toxicity of the two PCB congeners.

4.1.2. Rodent Maternal Exploratory Behaviours

One study found effects of MeHg on exploratory behaviour, as the highest MeHgCl dietary dose group showed reduced exploratory behaviour compared to MeHgCys and the vehicle-treated control [66]. MeHgCl is commercially available and commonly used in neurotoxicology studies, but MeHgCys has been shown as the dominant chemical form in fish tissue, to which humans are exposed through consumption [105]. As MeHgCl is more hydrophobic than other MeHg forms, this may cause differential toxic properties and may limit environmental relevance [105]. From studies included in this scoping review, all but two studies used MeHgCl in their treatment protocol [66,68].
4.1.3. Avian Parental Behaviours

Treatment with MeHg was explored in an avian model using zebra finches. Findings showed alterations in nest-building behaviour, a key component of avian paternal behaviour [106]. As well, a compensatory effect was suggested because MeHg-exposed birds had fewer successful nest-building trips but did not differ in the amount of hay brought to the nest building area, per hour. Behavioural data in this study were analysed in reproductive pairs, as zebra finches exhibit a biparental care strategy [107]. Interspecies comparison is limited due to different parental strategies, but these behavioural changes should be noted.

Another avian study found that PCB-exposed American kestrels abandoned their clutch more than control parents [77]. Incubation behaviour is a critical component of offspring success in this species as optimal development of an embryo occurs within a small temperature range [108,109]. Incubation behaviour has been associated with increases in serum prolactin level [110], which may contribute to the molecular pathways underlying these behavioural differences [111]. Similarly, disruptions to endocrine signalling may contribute to the correlations observed between toxicant blood concentrations of Glaucous gulls and non-incubating time away from the nest, as suggested by the authors [94,95]. As well, they note neurological disruptions as a potential cause of these behavioural changes [94,95].

4.1.4. Human Maternal Psychopathologies

In human mothers, heptachlor blood concentrations were associated with maternal psychopathological assessments, including correlations with depression and anxiety measures. Maternal anxiety and depression have been associated with reduced infant care, so heptachlor exposure may have negative effects on maternal care [112,113].

4.2. Neurochemical Changes in the Maternal Brain

4.2.1. Redox Activity

Changes in hippocampal catalase and glutathione peroxidase were observed in one MeHg exposure study, along with changes in thiol content in the prefrontal cortex [65]. Similarly, another study showed reduced cerebellar glutathione peroxidase activity following MeHg exposure [71]. The cellular mechanisms underlying MeHg neurotoxicity are not fully understood, but evidence suggests the electrophilic properties of MeHg allow for interactions with key nucleophilic groups including thiols and selenols. Furthermore, these interactions may disrupt the activity of many important metabolic proteins and receptors involved in antioxidant defense mechanisms [114].

4.2.2. OXTR Gene Expression

Oxytocin modulates the onset of maternal behaviour, which is mediated by receptor binding [115,116]. Dover et al. reported elevation of OXTR expression in PCB-exposed mice. This is consistent with the behavioural findings where high-crouch licking posture was increased on PND6. High-crouch posture is associated with optimal milk letdown, a key component of maternal care [117].

4.2.3. P450 Protein Expression

One study observed no change in hypothalamic Cyp1a1 expression following PCB 47 and 77 treatment [79]. Cyp1a1/2 and Cyp1b1/2 protein analysis was conducted in another PCB mixture study and expression levels were comparable between control and treated dams [84]; however, caution must be expressed with respect to the implication of these results given that the route of administration was a subcutaneous one and thus, unlikely a natural route of exposure to these substances.
4.2.4. Muscarinic Receptor Density

Maternal rat dams treated with higher doses of MeHg showed increase in MR density in the cerebral cortex and cerebellum, while PCB-exposed dams showed an increase in cerebral cortex MR density but a decrease in the cerebellum [96]. Co-exposed groups had similar regional increases and decreases as the PCB-only exposed group. In a second study using the same dosing regimen, a higher dose of MeHg increased cerebellar MR density, but limited changes were observed with the other treatment groups [101].

4.2.5. Neurotransmitter Levels

Decreases in dopamine and its metabolites were observed in multiple brain regions [88] following PCB exposure. In the same study, serotonin levels significantly increased in the hypothalamus. Both dopamine and serotonin are involved in molecular pathways underlying maternal behaviour, [118,119] so changes in levels of these neurotransmitters may have consequential effects on maternal care.

4.3. Limitations and Implications

4.3.1. Toxicant Inclusion

This scoping review focused on exposure to MeHg, PCBs, and OCPs, as these toxicants are the most abundant in the Northern Arctic region. As such, keywords were used to identify studies that assessed exposure to these three chemicals classes. The specificity of these keywords may have missed relevant studies, such as avian studies measuring mercury levels, which are a reliable proxy for MeHg levels [120].

4.3.2. Differentiating Direct and Indirect Effects of Exposure

Maternal behaviour can be affected by direct exposure to environmental toxicants [75,91]. Pup behaviour can also have direct effects on maternal responsiveness and behaviour [121]. Results from this scoping review show multiple neurochemical changes following toxicant exposure. As well, one cross-fostering designed study found significant increases in maternal behaviours when data from PCB-exposed dams raising PCB-exposed pups or control pups, and control dams raising PCB-exposed pups were combined [76]. These data suggest both direct effects on maternal behaviour and pup-mediated effects, which have been shown in other toxicant exposure studies [122]. Research protocols should continue to employ cross-fostering designs to gain insight into direct and pup-interaction behavioural changes.

4.3.3. Environmental Relevance

Humans are exposed to multiple toxicants through dietary and environmental sources [30]. Only six of 16 experimental studies assessed toxicant co-exposure. While it is crucial to understand molecular changes underlying single toxicant exposure, these studies lack insight into potential synergistic, additive, or antagonistic effects of environmentally relevant co-exposure. Future studies should aim to include co-exposure groups for highly abundant toxicants.

5. Conclusions

This scoping review gives insight into the multitude of effects associated with maternal toxicant exposure. Varied behavioural effects were identified following MeHg, PCB, or OCP treatment in avian and rodent models. The neurochemical pathways so far shown to be involved in mediating the effects of toxicant exposure include the oxytocin, serotonin, and dopamine signalling pathways, antioxidants, and muscarinic receptors. Future environmental toxicant research needs to characterize the potential harmful or adaptive responses to toxicant exposure involving neuromodulator signalling pathways and the role of antioxidants in these. Detailed mechanistic studies investigating these
pathways and maternal behavioural endpoints are necessary, given the critical intersection between maternal behaviour and offspring development, with an outlook toward sequential pregnancy and trans-generational consequences. Findings from this scoping review may help guide research and inform policy decisions in this field.

Conflicts of Interest: The authors declare no conflict of interest.

References
1. Ahlborg, U.G.; Brouwer, A.; Fingerhut, M.A.; Jacobson, J.L.; Jacobson, S.W.; Kennedy, S.W.; Kettrup, A.A.; Koeman, J.H.; Poiger, H.; Rappe, C. Impact of polychlorinated dibenzo-p-dioxins, dibenzofurans, and biphenyls on human and environmental health, with special emphasis on application of the toxic equivalency factor concept. *Eur. J. Pharm.* 1992, 228, 179–199. [CrossRef]
2. Longnecker, M.P.; Rogan, W.J.; Lucier, G. The human health effects of DDT (dichlorodiphenyltrichloroethane) and PCBS (polychlorinated biphenyls) and an overview of organochlorines in public health. *Annu. Rev. Public Health* 1997, 18, 211–244. [CrossRef]
3. Hong, Y.S.; Kim, Y.M.; Lee, K.E. Methylmercury exposure and health effects. *J. Prev. Med. Public Health* 2012, 45, 353–363. [CrossRef]
4. Jones, K.C.; De Voogt, P. Persistent organic pollutants (POPs): State of the science. *Environ. Pollut.* 1999, 100, 209–221. [CrossRef]
5. Thornton, I. Metal contamination of soils in urban areas. In *Soils in the Urban Environment*; Blackwell: Oxford, UK, 1991; pp. 47–75.
6. Mage, D.; Ozolins, G.; Peterson, P.; Webster, A.; Orthofer, R.; Vandeweerd, V.; Gwynne, M. Urban air pollution in megacities of the world. *Atmosph. Environ.* 1996, 30, 681–686. [CrossRef]
7. Wong, F.; Robson, M.; Diamond, M.; Harrad, S.; Truong, J. Concentrations and chiral signatures of POPs in soils and sediments: A comparative urban versus rural study in Canada and UK. *Chemosphere* 2009, 74, 404–411. [CrossRef] [PubMed]
8. Fowler, S.W. Critical review of selected heavy metal and chlorinated hydrocarbon concentrations in the marine environment. *Mar. Environ. Res.* 1990, 29, 1–64. [CrossRef]
9. Sinkkonen, S.; Paasivirta, J. Degradation half-life times of PCDDs, PCDFs and PCBs for environmental fate modeling. *Chemosphere* 2000, 40, 943–949. [CrossRef]
10. Rice, K.M.; Walker Jr, E.M.; Wu, M.; Gillette, C.; Blough, E.R. Environmental mercury and its toxic effects. *J. Prev. Med. Public Health* 2014, 47, 74–83. [CrossRef]
11. United Nations Environment Programme. Minamata Convention on Mercury—Text and Annexes. 2013. Available online: http://www.mercuryconvention.org/Portals/11/documents/Booklets/Minamata%20Convention%20on%20Mercury_booklet_English.pdf (accessed on 5 July 2019).
12. United Nations Environment Programme. Stockholm Convention on Persistent Organic Pollutants—Texts and Annexes. 2018. Available online: http://chm.pops.int/TheConvention/Overview/TextoftheConvention/tabid/2232/Default.aspx (accessed on 5 July 2019).
13. Clarkson, T.W. The three modern faces of mercury. *Environ. Health Persp.* 2002, 110 (Suppl. 1), 11–23. [CrossRef]
14. Rice, G.; Ambrose, R.; Bullock, O.; Swartout, J. Mercury Study Report to Congress. Volume 3. Fate and Transport of Mercury in the Environment; Environmental Protection Agency: Research Triangle Park, NC, USA, 1997. Available online: https://www.epa.gov/sites/production/files/2015-09/documents/volume3.pdf (accessed on 5 July 2019).
15. National Research Council. *Toxicological Effects of Methylmercury*; National Academy Press: Washington, DC, USA, 2000. Available online: https://www.ncbi.nlm.nih.gov/books/NBK225778/ (accessed on 5 July 2019).
16. World Health Organization. Methylmercury. 1990. Available online: https://apps.who.int/iris/bitstream/handle/10665/38082/9241571012_eng.pdf (accessed on 5 July 2019).
17. Nisbet, I.C.; Sarofim, A.F. Rates and routes of transport of PCBs in the environment. *Environ. Health Persp*. 1972, I, 21–38. [CrossRef] [PubMed]
18. Faroon, O.; Olson, J.N. Toxicological Profile for Polychlorinated Biphenyls (PCBs). 2000. Available online: https://www.atsdr.cdc.gov/toxprofiles/tp17.pdf (accessed on 8 July 2019).
19. Giesy, J.P.; Kannan, K. Dioxin-like and non-dioxin-like toxic effects of polychlorinated biphenyls (PCBs): Implications for risk assessment. *Crit. Rev. Toxicol.* 1998, 28, 511–569. [CrossRef]

20. Jensen, A.A. Polychlorobiphenyls (PCBs), polychlorodibenzo-p-dioxins (PCDDs) and polychlorodibenzofurans (PCDFs) in human milk, blood and adipose tissue. *Sci. Total Environ.* 1987, 64, 259–293. [CrossRef]

21. Gioia, R.; Lohmann, R.; Dachs, J.; Temme, C.; Lakaschus, S.; Schulz-Bull, D.; Hand, I.; Jones, K.C. Polychlorinated biphenyls in air and water of the North Atlantic and Arctic Ocean. *J. Geophy. Res. Atmosp.* 2008, 113. [CrossRef]

22. Kalantz, O.; Alcock, R.E.; Johnston, P.; Santillo, D.; Stringer, R.; Thomas, G.; Jones, K. The global distribution of PCBs and organochlorine pesticides in butter. *Environ. Sci. Technol.* 2001, 35, 1013–1018. [CrossRef] [PubMed]

23. Ribas-Fitó, N.; Sala, M.; Kogevinas, M.; Sunyer, J. Polychlorinated biphenyls (PCBs) and neurological development in children: A systematic review. *J. Epidem. Commun. Health* 2001, 55, 537–546. [CrossRef]

24. Centers for Disease Control and Prevention. *Fourth National Report on Human Exposure to Environmental Chemicals; Centers for Disease Control and Prevention*: Atlanta, GA, USA, 2009. Available online: https://www.cdc.gov/exposurerreport/pdf/fourthreport.pdf (accessed on 8 July 2019).

25. Shen, L.; Wania, F. Compilation, evaluation, and selection of physical–chemical property data for organochlorine pesticides. *J. Chem. Eng. Data* 2005, 50, 742–768. [CrossRef]

26. Jayaraj, R.; Megha, P.; Sreedev, P. Organochlorine pesticides, their toxic effects on living organisms and their fate in the environment. *Interd. Toxical.* 2016, 9, 90–100. [CrossRef]

27. Shunhirasingham, C.; Oyiliagu, C.E.; Cao, X.; Gouin, T.; Wania, F.; Lee, S.-C.; Pozo, K.; Harner, T.; Muir, D.C. Spatial and temporal pattern of pesticides in the global atmosphere. *J. Environ. Monit.* 2010, 12, 1650–1657. [CrossRef]

28. Lohmann, R.; Gioia, R.; Jones, K.C.; Nizzetto, L.; Temme, C.; Xie, Z.; Schulz-Bull, D.; Hand, I.; Morgan, E.; Jantunen, L. Organochlorine pesticides and PAHs in the surface water and atmosphere of the North Atlantic and Arctic Ocean. *Environ. Sci. Technol.* 2009, 43, 5633–5639. [CrossRef]

29. Reid, A.; Callan, A.; Stasinska, A.; Heyworth, J.; Phi, D.T.; Odland, J.O.; Hinwood, A. Maternal exposure to organochlorine pesticides in Western Australia. *Sci. Total Environ.* 2013, 449, 208–213. [CrossRef] [PubMed]

30. Donaldson, S.G.; Van Oostdam, J.; Tikhonov, C.; Feeley, M.; Armstrong, B.; Ayotte, P.; Boucher, O.; Bowers, W.; Chan, L.; Dallaire, F.; et al. Environmental contaminants and human health in the Canadian Arctic. *Sci. Total Environ.* 2010, 408, 5165–5234. [CrossRef] [PubMed]

31. Barrie, L.A.; Gregor, D.; Hargrave, B.; Lake, R.; Muir, D.; Shearer, R.; Tracey, B.; Bidleman, T. Arctic contaminants: Sources, occurrence and pathways. *Sci. Total Environ.* 1992, 122, 1–74. [CrossRef]

32. Arctic Monitoring and Assessment Programme. AMAP assessment 2011: Mercury in the Arctic. Arctic Monitoring and Assessment Programme (AMAP), 2011. Available online: https://www.amap.no/documents/download/989/inline (accessed on 20 July 2019).

33. Atwell, L.; Hobson, K.A.; Welch, H.E. Biomagnification and bioaccumulation of mercury in an arctic marine food web: Insights from stable nitrogen isotope analysis. *Canad. J. Fisher. Aqua.* 1998, 55, 1114–1121. [CrossRef]

34. Korhonen, P.; Virtanen, M.; Schultz, T. Bioenergetic calculation of mercury accumulation in fish. *Water Air Soil Pollut.* 1995, 80, 901–904. [CrossRef]

35. Thomas, D.J.; Tracey, B.; Marshall, H.; Norstrom, R.J. Arctic terrestrial ecosystem contamination. *Sci. Total Environ.* 1992, 122, 135–164. [CrossRef]

36. Kuhnlein, H.V.; Chan, H.M. Environment and contaminants in traditional food systems of northern indigenous peoples. *Annu. Rev. Nutr.* 2000, 20, 595–626. [CrossRef]

37. van Oostdam, J.; Donaldson, S.G.; Feeley, M.; Arnold, D.; Ayotte, P.; Bondy, G.; Chan, L.; Dewailly, E.; Furgal, C.; Kuhnlein, H. Human health implications of environmental contaminants in Arctic Canada: A review. *Sci. Total Environ.* 2005, 351, 165–246. [CrossRef]

38. Arctic Monitoring and Assessment Programme. AMAP assessment 2002: Human Health in the Arctic. Arctic Monitoring and Assessment Programme (AMAP), 2002. Available online: https://www.amap.no/documents/download/181/inline (accessed on 20 July 2019).
39. Arctic Monitoring and Assessment Programme. AMAP assessment report: Arctic pollution issues. Arctic Monitoring and Assessment Programme (AMAP), 1998. Available online: https://www.amap.no/documents/doc/amap-assessment-report-arctic-pollution-issues/68 (accessed on 20 July 2019).

40. Laird, B.D.; Goncharov, A.B.; Chan, H.M. Body burden of metals and persistent organic pollutants among Inuit in the Canadian Arctic. Environ. Int. 2013, 59, 33–40. [CrossRef]

41. Dewailly, E.; Ayotte, P.; Bruneau, S.; Laliberté, C.; Muir, D.C.; Norstrom, R.J. Inuit exposure to organochlorines through the aquatic food chain in arctic quebec. Environ. Health Persp. 1993, 101, 618–620. [CrossRef]

42. Dewailly, E.; Nantel, A.; Bruneau, S.; Laliberté, C.; Ferron, L.; Gingras, S. Breast milk contamination by PCDDs, PCDFs and PCBs in Arctic Quebec: A preliminary assessment. Chemosphere 1992, 25, 1245–1249. [CrossRef]

43. Dewailly, E.; Nantel, A.; Weber, J.P.; Meyer, F. High levels of PCBs in breast milk of Inuit women from arctic Quebec. Bull. Environ. Cont. Toxicol. 1989, 43, 641–646. [CrossRef] [PubMed]

44. Wheatley, B.; Paradis, S. Exposure of Canadian aboriginal peoples to methylmercury. Water Air Soil Pollut. 1995, 80, 3–11. [CrossRef]

45. Chu, I.; Bowers, W.J.; Caldwell, D.; Nakai, J.; Wade, M.G.; Yagminas, A.; Li, N.; Moir, D.; El Abbas, L.; Håkansson, H.; et al. Toxicological effects of in utero and lactational exposure of rats to a mixture of environmental contaminants detected in Canadian Arctic human populations. J. Toxicol. Environ. Health A 2008, 71, 93–108. [CrossRef]

46. Muckle, G.; Ayotte, P.; Dewailly, E.E.; Jacobson, J.L. Prenatal exposure of the northern Québec Inuit infants to environmental contaminants. Environ. Health Persp. 2001, 109, 1291–1299. [CrossRef]

47. Pereira, M.; Morrell, J.I. Functional mapping of the neural circuitry of rat maternal motivation: Effects of site-specific transient neural inactivation. J. Neuroendocrinol. 2011, 23, 1020–1035. [CrossRef]

48. Numan, M. Hypothalamic neural circuits regulating maternal responsiveness toward infants. Behav. Cogn. Neurosci. Rev. 2006, 5, 163–190. [CrossRef]

49. Bridges, R.S. Biochemical Basis of Parental Behavior in the Rat. In Advances in the Study of Behavior; Academic Press: New York, NY, USA, 1996; Volume 25, pp. 215–242. [CrossRef]

50. Magnusson, J.E.; Fleming, A.S. Rat pups are reinforcing to the maternal rat: Role of sensory cues. Psychobiology 1995, 23, 69–75. [CrossRef]

51. Olazábal, D.E.; Pereira, M.; Agrati, D.; Ferreira, A.; Fleming, A.S.; González-Mariscal, G.; Lévy, F.; Lucion, A.B.; Morrell, J.I.; Numan, M.; et al. Flexibility and adaptation of the neural substrate that supports maternal behavior in mammals. Neurosci. Biobehav. Rev. 2013, 37, 1875–1892. [CrossRef]

52. Elabbas, L.E.; Finnilä, M.A.; Herlin, M.; Stern, N.; Trossvik, C.; Bowers, W.J.; Nakai, J.; Tuukkanen, J.; Heimeier, R.A.; Åkesson, A. Perinatal exposure to environmental contaminants detected in Canadian Arctic human populations changes bone geometry and biomechanical properties in rat offspring. J. Toxicol. Environ. Health Part. A 2011, 74, 1304–1318. [CrossRef]

53. Gill, S.; Bowers, W.J.; Nakai, J.S.; Yagminas, A.; Mueller, R.; Pulido, O. Effects of environmentally relevant mixtures of persistent organic pollutants on the developmental neurobiology in rats. Toxicol. Pathol. 2013, 41, 38–47. [CrossRef]

54. Kyriklaki, A.; Vafeiadi, M.; Kampouri, M.; Koutra, K.; Roumeliotaki, T.; Chalkiadaki, G.; Anousaki, D.; Rantakokko, P.; Kiviranta, H.; Fthenou, E. Prenatal exposure to persistent organic pollutants in association with offspring neuropsychological development at 4 years of age: The Rhea mother-child cohort, Crete, Greece. Environ. Int. 2016, 97, 204–211. [CrossRef] [PubMed]

55. Pelletier, G.; Masson, S.; Wade, M.J.; Nakai, J.; Alwis, R.; Mohottalage, S.; Kumarathasan, P.; Black, P.; Bowers, W.J.; Chu, I.; et al. Contribution of methylmercury, polychlorinated biphenyls and organochlorine pesticides to the toxicity of a contaminant mixture based on Canadian Arctic population blood profiles. Toxicol. Lett. 2009, 184, 176–185. [CrossRef] [PubMed]

56. Seegal, R.F. Epidemiological and laboratory evidence of PCB-Induced neurotoxicity. Crit. Rev. Toxicol. 1996, 26, 709–737. [CrossRef]

57. Champagne, F.A.; Francis, D.D.; Mar, A.; Meaney, M.J. Variations in maternal care in the rat as a mediating influence for the effects of environment on development. Physiol. Behav. 2003, 79, 359–371. [CrossRef]

58. Lupien, S.J.; McEwen, B.S.; Gunnar, M.R.; Heim, C. Effects of stress throughout the lifespan on the brain, behaviour and cognition. Nat. Rev. Neurosci. 2009, 10, 434–445. [CrossRef]
59. Meaney, M.J. Maternal care, gene expression, and the transmission of individual differences in stress reactivity across generations. *Annu. Rev. Neurosci.* 2001, 24, 1161–1192. [CrossRef]

60. Arksey, H.; O’Malley, L. Scoping studies: Towards a methodological framework. *Int. J. Soc. Res. Method* 2005, 8, 19–32. [CrossRef]

61. Catanese, M.C.; Suvorov, A.; Vandenberg, L.N. Beyond a means of exposure: A new view of the mother in toxicology research. *Toxicol. Res.* 2015, 4, 592–612. [CrossRef]

62. Cummings, J.; Clemens, L.; Nunez, A. Mother counts: How effects of environmental contaminants on maternal care could affect the offspring and future generations. *Front. Neuroendocrinol.* 2010, 31, 440–451. [CrossRef]

63. Debes, F.; Budtz-Jørgensen, E.; Weihe, P.; White, R.F.; Grandjean, P. Impact of prenatal methylmercury exposure on neurobehavioral function at age 14 years. *Neurotoxicol. Teratol.* 2006, 28, 536–547. [CrossRef] [PubMed]

64. Weston, H.I.; Sobolewski, M.E.; Allen, J.L.; Weston, D.; Conrad, K.; Pelkowski, S.; Watson, G.E.; Zareba, G.; Cory-Slechta, D.A. Sex-dependent and non-monotonic enhancement and unmasking of methylmercury neurotoxicity by prenatal stress. *Neurotoxicology* 2014, 41, 123–140. [CrossRef] [PubMed]

65. Espitia-Pérez, P.; Albino, S.M.; Espitia-Pérez, L.; Brango, H.; da Rosa, H.; Silveira, A.K.; Moraes, D.P.; Cerveira, C.; Mingori, M.; Ribeiro, C.T. Neurobehavioral and oxidative stress alterations following methylmercury and retinyl palmitate co-administration in pregnant and lactating rats and their offspring. *Neurotoxicology* 2018, 69, 164–180. [CrossRef] [PubMed]

66. Glover, C.N.; Zheng, D.; Jayashankar, S.; Sales, G.D.; Hogstrand, C.; Lundebye, A.-K. Methylmercury speculation influences brain gene expression and behavior in gestationally-exposed mice pups. *Toxicol. Sci.* 2009, 110, 389–400. [CrossRef]

67. Walf, A.A.; Frye, C.A. The use of the elevated plus maze as an assay of anxiety-related behavior in rodents. *Nat. Protoc.* 2007, 2, 322–328. [PubMed]

68. Chin, S.Y.; Hopkines, W.A.; Cristol, D.A. Mercury alters initiation and construction of nests by zebra finches, but not incubation or provisioning behaviors. *Ecotoxicology* 2017, 26, 1271–1283. [CrossRef]

69. Day, B.J. Catalase and glutathione peroxidase mimics. *Biochem. Pharmacol.* 2009, 77, 285–296. [CrossRef]

70. Baba, S.P.; Bhatnagar, A. Role of thiols in oxidative stress. *Curr. Opin. Toxicol.* 2018, 7, 133–139. [CrossRef]

71. Manfroi, C.; Cereser, V.; Abreu, F.; Oliveira, A.; Bizarro, L.; Rocha, J.; Frizzo, M.; Souza, D.; Farina, M. Maternal milk as methylmercury source for suckling mice: Neurotoxic effects involved with the cerebellar glutamatergic system. *Toxicol. Sci.* 2004, 81, 172–178. [CrossRef]

72. Franco, J.L.; Posser, T.; Dunkley, P.R.; Dickson, P.W.; Mattos, J.J.; Martins, R.; Bainy, A.C.; Marques, M.R.; Dafre, A.L.; Farina, M. Methylmercury neurotoxicity is associated with inhibition of the antioxidant enzyme glutathione peroxidase. *Free Rad. Biol. Med.* 2009, 47, 449–457. [CrossRef]

73. McFarland, V.A.; Clarke, J.U. Environmental occurrence, abundance, and potential toxicity of polychlorinated biphenyl congeners: Considerations for a congener-specific analysis. *Environ. Health Persp.* 1989, 81, 225–239. [CrossRef] [PubMed]

74. Simmons, S.; Cummings, J.; Clemens, L.; Nunez, A. Exposure to PCB 77 affects the maternal behavior of rats. *Physiol. Behav.* 2005, 84, 81–86. [CrossRef] [PubMed]

75. Simmons, S.; Cummings, J.; Clemens, L.; Nunez, A. A cross-fostering analysis of the effects of PCB 77 on the maternal behavior of rats. *Physiol. Behav.* 2005, 85, 83–91. [CrossRef]

76. Fernie, K.; Bortolotti, G.; Smits, J. Reproductive abnormalities, teratogenicity, and developmental problems in American kestrels (Falco sparverius) exposed to polychlorinated biphenyls. *J. Toxicol. Environ. Health Part A* 2003, 66, 2089–2103. [CrossRef]

77. Albro, P.W.; Corbett, J.T.; Schroeder, J.L. Quantitative characterization of polychlorinated biphenyl mixtures (aroclor® 1248, 1254 and 1260) by gas chromatography using capillary columns. *J. Chromatogr. A* 1981, 205, 103–111. [CrossRef]

78. Dover, E.N.; Mankin, D.E.; Cromwell, H.C.; Phuntumart, V.; Meserve, L.A. Polychlorinated biphenyl exposure alters oxytocin receptor gene expression and maternal behavior in rat model. *Endoc. Disrup.* 2015, 3, e979681. [CrossRef]
80. De Kloet, E.R.; Voorhuis, T.A.; Elands, J. Estradiol induces oxytocin binding sites in rat hypothalamic ventromedial nucleus. *Eur. J. Pharmacol.* **1985**, *118*, 185–186. [CrossRef]

81. Pedersen, C.A. Oxytocin control of maternal behavior. Regulation by sex steroids and offspring stimuli. *Annu. N. Y. Acad. Sci.* **1997**, *807*, 126–145. [CrossRef]

82. Ferguson, C.S.; Tyndale, R.F. Cytochrome P450 enzymes in the brain: Emerging evidence of biological significance. *Trend. Pharmacol. Sci.* **2011**, *32*, 708–714. [CrossRef]

83. Hedlund, E.; Gustafsson, J.A.; Warner, M. Cytochrome P450 in the brain, a review. *J. Neurochem.* **2001**, 2, 245–263. [CrossRef]

84. Bonfanti, P.; Colombo, A.; Villa, S.; Comelli, F.; Costa, B.; Santagostino, A. The effects of accumulation of an environmentally relevant polychlorinated biphenyl mixture on cytochrome P450 and P-glycoprotein expressions in fetuses and pregnant rats. *Chemosphere* **2009**, *75*, 572–579. [CrossRef] [PubMed]

85. Ramos, L.; Hernandez, L.M.; Gonzalez, M.J. Variation of PCB congener levels during lactation period and relationship to their molecular structure. *Arch. Environ. Contam. Toxicol.* **1997**, *33*, 97–103. [CrossRef] [PubMed]

86. Bachour, G.; Failing, K.; Georgii, S.; Elmadfa, I.; Brunn, H. Species and organ dependence of PCB contamination in fish, foxes, roe deer, and humans. *Arch. Environ. Contam. Toxicol.* **1998**, *35*, 666–673. [CrossRef] [PubMed]

87. Chu, S.; Covaci, A.; Schepens, P. Levels and chiral signatures of persistent organochlorine pollutants in human tissues from Belgium. *Environ. Res.* **2003**, *93*, 167–176. [CrossRef]

88. Honma, T.; Suda, M.; Miyagawa, M.; Wang, R.-S.; Kobayashi, K.; Sekiguchi, S. Alteration of brain neurotransmitters in female rat offspring induced by prenatal administration of 16 and 64 mg/kg of 2, 2′, 4, 4′, 5, 5′-hexachlorobiphenyl (PCB153). *Indus. Health* **2009**, *47*, 11–21. [CrossRef]

89. Ashton, M.; Kantai, T.; Kohler, P.; Roemer-Mahler, A.; Templeton, J. Summary of the fourth conference of the parties to the stockholm convention on persistent organic pollutants: 4–8 May 2009. Available online: https://enb.iisd.org/download/pdf/enb15174e.pdf (accessed on 23 July 2019).

90. Matsuura, I.; Saitoh, T.; Tani, E.; Wako, Y.; Iwata, H.; Toyota, N.; Ishizuka, Y.; Namiki, M.; Hoshino, N.; Tsuchitani, M.; et al. Evaluation of a two-generation reproduction toxicity study adding endpoints to detect endocrine disrupting activity using lindane. *J. Toxicol. Sci.* **2005**, *30*, S15–S161. [CrossRef]

91. Palanza, P.; Morellini, F.; Parmigiani, S.; Vom Saal, F. Ethological methods to study the effects of maternal exposure to estrogenic endocrine disrupters: A study with methoxychlor. *Neurotoxicol. Teratol.* **2002**, *24*, 55–69. [CrossRef]

92. Yalçın, S.S.; Örün, E.; Yalçın, S.; Aykut, O. Organochlorine pesticide residues in breast milk and maternal psychopathologies and infant growth from suburban area of Ankara, Turkey. *Int. J. Environ. Health Res.* **2015**, *25*, 364–372. [CrossRef]

93. Hudecova, A.M.; Hansen, K.E.; Mandal, S.; Berntsen, H.F.; Khezri, A.; Bale, T.L.; Fraser, T.W.; Zimmer, K.E.; Ropstad, E. A human exposure based mixture of persistent organic pollutants affects the stress response in female mice and their offspring. *Chemosphere* **2018**, *197*, 585–593. [CrossRef]

94. Bustnes, J.; Bakken, V.; Erikstad, K.; Mehlum, F.; Skaare, J. Patterns of incubation and nest-site attentiveness in the species and organ dependence of PCB contamination in fish, foxes, roe deer, and humans. *Arch. Environ. Contam. Toxicol.* **2001**, *38*, 364–372. [CrossRef] [PubMed]

95. Bustnes, J.O.; Miland, Ø.; Fjeld, M.; Erikstad, K.E.; Skaare, J.U. Relationships between ecological variables and four organochlorine pollutants in an artic glaucous gull (Larus hyperboreus) population. *Environ. Pollut.* **2005**, *136*, 175–185. [CrossRef] [PubMed]

96. Coccini, T.; Randine, G.; Castoldi, A.F.; Grandjean, P.; Ostendorp, G.; Heinzow, B.; Manzo, L. Effects of developmental co-exposure to methylmercury and 2, 2′, 4, 4′, 5, 5′-hexachlorobiphenyl (PCB153) on cholinergic muscarinic receptors in rat brain. *Neurotoxicology* **2006**, *27*, 468–477. [CrossRef]

97. Coccini, T.; Randine, G.; Candura, S.M.; Nappi, R.E.; Prockop, L.D.; Manzo, L. Low-level exposure to methylmercury modifies muscarinic cholinergic receptor binding characteristics in rat brain and lymphocytes: Physiologic implications and new opportunities in biologic monitoring. *Environ. Health Persp.* **2000**, *108*, 29–33. [CrossRef] [PubMed]

98. Gimenez-Llort, L.; Ahlborn, E.; Dare, E.; Vahter, M.; Ögren, S.-O.; Ceccatelli, S. Prenatal exposure to methylmercury alters locomotor activity of male but not female rats. *Experim. Br. Res.* **1997**, *117*, 428–436. [CrossRef] [PubMed]
100. Schantz, S.L.; Moshtaghian, J.; Ness, D.K. Spatial learning deficits in adult rats exposed to ortho-substituted PCB congeners during gestation and lactation. *Fundam. Appl. Toxicol.* 1995, 26, 117–126. [CrossRef]

101. Roda, E.; Manzo, L.; Coccini, T. Application of neurochemical markers for assessing health effects after developmental methylmercury and PCB coexposure. *J. Toxicol.* 2012, 2012. [CrossRef]

102. Dechartres, J.; Pawluski, J.L.; Gueguen, M.M.; Jablaoui, A.; Maguin, E.; Rhimi, M.; Charlier, T.D. Glyphosate and glyphosate-based herbicide exposure during the peripartum period affects maternal brain plasticity, maternal behaviour and microbiome. *J. Neuroendocrinol.* 2019, e12731. [CrossRef]

103. Della Seta, D.; Minder, I.; Dessi-Fulgheri, F.; Farabollini, F. Bisphenol-A exposure during pregnancy and lactation affects maternal behavior in rats. *Brain Res. Bull.* 2005, 65, 255–260. [CrossRef]

104. Palanza, P.L.; Howdeshell, K.L.; Parmigiani, S.; vom Saal, F.S. Exposure to a low dose of bisphenol A during fetal life or in adulthood alters maternal behavior in mice. *Environ. Health Persp.* 2002, 110 (Suppl. 3), 415–422. [CrossRef]

105. Harris, H.H.; Pickering, I.J.; George, G.N. The chemical form of mercury in fish. *Science* 2003, 301, 1203. [CrossRef]

106. Collia, N.E.; Collia, E.C. *Nest Building and Bird Behavior*; Princeton University Press: Princeton, NJ, USA, 1984.

107. Cockburn, A. Prevalence of different modes of parental care in birds. *Proc. Royal Soc. B Biol. Sci.* 2006, 273, 1375–1383. [CrossRef]

108. Buntin, J.D. Neural and Hormonal Control of Parental Behavior in Birds. In *Advances in the Study of Behavior*; Academic Press: New York, NY, USA, 1996; Volume 25, pp. 161–213. [CrossRef]

109. Webb, D. Thermal tolerance of avian embryos: A review. *Condor* 1987, 89, 874–898. [CrossRef]

110. Sockman, K.W.; sharp, P.J.; Schwabl, H. Orchestration of avian reproductive effort: An integration of the ultimate and proximate bases for flexibility in clutch size, incubation behaviour, and yolk androgen deposition. *Biol. Rev.* 2006, 81, 629–666. [CrossRef]

111. Angelier, F.; Wingfield, J.C.; Tartu, S.; Chastel, O. Does prolactin mediate parental and life-history decisions in response to environmental conditions in birds? A review. *Horm. Behav.* 2010, 58, 754–761. [CrossRef] [PubMed]

112. Gelfand, D.M.; Teti, D.M. The effects of maternal depression on children. *Clin. Psychol. Rev.* 1990, 10, 329–353. [CrossRef]

113. Britton, J.R. Pre-discharge anxiety among mothers of well newborns: Prevalence and correlates. *Acta Paediatr.* 2005, 94, 1771–1776. [CrossRef] [PubMed]

114. Farina, M.; Ashner, M.; Rocha, J.B. Redox State in Mediating Methylmercury Neurotoxicity. In *Methylmercury and Neurotoxicity*; Springer: Boston, MA, USA, 2012; pp. 101–125.

115. Gimpl, G.; Fahrenholz, F. The oxytocin receptor system: Structure, function, and regulation. *Physiol. Rev.* 2001, 81, 629–683. [CrossRef] [PubMed]

116. Pedersen, C.A.; Prange, A.J. Induction of maternal behavior in virgin rats after intracerebroventricular administration of oxytocin. *Proc. Nat. Acad. Sci. USA* 1979, 76, 6661–6665. [CrossRef] [PubMed]

117. Lincoln, D.; Hentzen, K.; Hin, T.; Van der Schoot, P.; Clarke, G.; Summerlee, A. Sleep: A prerequisite for ultrasonic vocalizations of the rat pup. *Experim. Br. Res.* 1980, 38, 151–162. [CrossRef] [PubMed]

118. Pedersen, C.A.; Prange, A.J. Induction of maternal behavior in virgin rats after intracerebroventricular administration of oxytocin. *Proc. Nat. Acad. Sci. USA* 1979, 76, 6661–6665. [CrossRef] [PubMed]

119. Strathearn, L. Maternal neglect: Oxytocin, dopamine and the neurobiology of attachment. *J. Neuroendocrin.* 2011, 23, 1054–1065. [CrossRef] [PubMed]

120. Hartman, C.A.; Ackerman, J.T.; Herzog, M.P. Mercury exposure and altered parental nesting behavior in a wild songbird. *Environ. Sci. Technol.* 2019, 53, 5396–5405. [CrossRef] [PubMed]

121. Hashimoto, H.; Saito, T.R.; Furudate, S.; Takahashi, K.W. Prolactin levels and maternal behavior induced by ultrasonic vocalizations of the rat pup. *Exp. Anim.* 2001, 50, 307–312. [CrossRef]

122. Cox, K.H.; Gatewood, J.D.; Howeth, C.; Rissman, E.F. Gestational exposure to bisphenol A and cross-fostering affect behaviors in juvenile mice. *Horm. Behav.* 2010, 58, 754–761. [CrossRef]