HIV, immune activation and salt-sensitive hypertension (HISH): a research proposal

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Abstract

Objective: The objective of this study is to quantify and compare the effect of excess dietary salt on immune cell activation and blood pressure in HIV versus HIV negative individuals.

Results: Salt-sensitivity is associated with increased immune cell activation in animal studies. This concept has not been tested in people living with HIV. This study will therefore add more information in elucidating the interaction between HIV infection and/or anti-retroviral therapy (ART), immune-activation/inflammation and hypertension.

Keywords: Hypertension, Immune activation, Salt-sensitivity, HIV

Introduction

Salt-sensitive hypertension is defined as a change in blood pressure (BP) greater than 10% or > 5 mmHg in response to either increased or reduced salt intake [1, 2]. This phenomenon is more common in individuals with higher BP, blacks, elderly, and is seen in certain comorbid conditions such as chronic kidney disease, diabetes mellitus or metabolic syndrome [3]. Salt has been implicated to initiate an inflammatory process that could result in hypertension [4–6]. Moreover, inflammation-driven salt accumulation is evident on sites of infection and/or inflammation [7].

Antiretroviral (ART) treated People living with HIV (PLWH) are prone to development of hypertension and elevated BP [8–10]. One of the reasons for this could be the interaction of a generalized systemic suboptimal immune activation and inflammation that persists, traditional risk factors including dietary salt intake, HIV particles and ART [11, 12]. However, the effect of salt on BP and immune activation has not been tested in PLWH and dietary assessments are never part of routine assessment in the clinical setting especially in Africa hence the need for this proposed study.

Study objectives:

1. To determine the prevalence of salt-sensitive hypertension in PLWH and compare with HIV negative population
2. To quantify and compare the effect of excess dietary salt on immune cell activation and BP in ART-treated normotensive, ART-treated hypertensive, HIV negative hypertensive and HIV negative normotensive individuals.

Main text

Methods

Study site, design and patients

We propose to conduct a three-week prospective study at Livingstone Central Hospital (LCH) involving PLWH attending routine ART and HIV negative controls from volunteer health workers who will be matched for age, sex, body mass index (BMI), and waist circumference (WC). LCH is the largest referral hospital in Southern Province of Zambia that offers ART and general medical services to the community with approximately 3776 PLWH enrolled in ART.

Eligibility criteria

Inclusion criteria The study cohort will include all adults (aged 18 and above) who will be required to verbally consent and sign a consent form and should be attending medical clinic for both general clinics and ART. Four groups will be established for comparison namely:
1. ART treated normotensive individuals
2. ART treated hypertensive individuals
3. Hypertensive individuals who are HIV negative
4. Health controls.

Exclusion criteria  Exclusion from the study will be based on existence of co-morbidities such as diabetes mellitus and cancer, and also those with existing and recent past opportunistic infections, syphilis, hepatitis C and B virus infection and tuberculosis infection; and sick patients (clients seeking healthcare due to an illness rather than routine ART clinic reviews) will be excluded. Those with recent and current alcohol consumption and smoking status will also be excluded from the study.

Study variables
Response variables  Primary: Salt-sensitivity of BP

Secondary: Markers of antigen presenting cell activation CD80, CD83, CD86, CD69 and IsoLGs, pro-inflammatory markers such as interleukin 17A (IL-17A), IL-6, tumor necrosis factor alpha (TNF-α), interferon gamma (IFN-γ) and intermediate monocyte subset (CD14\(^{++}\)CD16\(^{+}\)) which are raised in inflammation.

Explanatory variables  Traditional risk factors: age, sex, WC, BMI, waist-to-hip ratio (WHR), sedentary lifestyle, physical activity, fruit and vegetable intake, hours of sleep.

Clinical risk factors: BP, Electrocardiogram (ECG) parameters, lipid profile, Urine and blood electrolyte (sodium, potassium and chloride), Urine macro- and micro-analysis, ART regimen, duration on ART, adherence to ART, family history of hypertension and diabetes risk, duration of hypertension, medical history.

Immune related factors: Immune status (CD4 and HIV RNA viral load absolute counts), soluble CD14 (sCD14), Complete blood count, C-reactive protein (CRP).

Sample size
For aim 1, OpenEpi online software (Kelsey’s method) will be used to compute a total sample size of 236 (118 hypertensive versus 118 normotensive in 1:1 ratio each between PLWH and HIV negative groups) for salt-sensitive hypertension with the following inputs: 95% significance level at 80% power; Estimated percent of salt-sensitive hypertension in the hypertensive and normotensive population was 50% and 25% respectively [13–15].

For aim 2, G*Power [16, 17] version 3.1.9.4 will be employed to calculate a total sample size of 48 (12 for each group). The input assumptions are detailed in Table 1.

Table 1  Sample size determination

| Options             | Pillai V, O'Brien-Shieh Algorithm |
|--------------------|----------------------------------|
| Analysis            | A priori: Compute sample Size    |
| Input               | Effect size f(\(v\)) 1           |
| \(\alpha\) err prob| 0.05                             |
| Power (1 - \(\beta\) err prob) | 0.80 |
| Number of groups    | 4                                |
| Number of measurements | 3                             |
| Output              | Non-centrality parameter \(\lambda\) 22 |
| Critical F          | 2.8477260                        |
| Numerator df        | 6.0                              |
| Denominator df      | 14.0                             |
| Sample size         | 11 per group                     |
| Lost to follow (10%) | 1 per group                     |
| Total sample size   | 48                               |
| Actual power        | 0.825                            |
| Pillai V            | 1.0                              |

Sampling
For selection of hypertensive and normotensive study participants, simple random sampling will be used assisted by an online random number generator. PLWH will be randomly selected during their routine medical visits.

In order to create equal groups in the strata, usage of randomization blocked in blocks of size four and six will be employed.

Procedure for aim 1  In order to determine the prevalence of salt-sensitive hypertension in PLWH and HIV negative population, we will conduct salt-sensitive analyses as described below:

Salt sensitivity analysis  We will employ a modification of the procedure described by He et al. [18] as shown in Fig. 1. Briefly, in the 1st week (7 days), participants will be requested to avoid consuming processed foods and adding salt to their food. Participants will be required to record their daily diet. In the 2nd week, they will be given the World health organisation (WHO) recommended low salt (2.3 g), and in the 3rd week, high salt (9 g/day). The changes for low salt (2.3 g NaCl/day) on BP will be calculated as previously described [18]. BP will be monitored three times each day and on the last day of each phase a 24-h BP using an ambulatory BP monitoring device will be employed to assure that BP does not exceed 180/110 mmHg, in which case the protocol will be discontinued for safety.

Sodium excretion  We will assess sodium excretion in urine samples collected over 24 h period as previously described by Rakova et al. [19]. Briefly, a 24-h urine will be collected by participants in aliquots, transported to
the laboratory and sodium measured. An average sodium excretion will be calculated and used as a surrogate of dietary intake.

Procedure for aim 2 Blood measurements will be conducted three times (Fig. 1) at baseline (day 7 before low salt intake is commenced), day 14 and on day 21 (last day of high salt intake). Heparinized blood specimens will be collected to compare IsoLGs, inflammatory markers (IL-17A, IL-6, TNF-α, IFN-γ) and immune activation markers (CD80, CD86, CD83) in all four groups. Daily BPs will be collected and a 24-h BP on the last day of each phase. Laboratory procedures and flow cytometry analysis are elaborated in Additional file 1.
Statistical analysis
We will use FlowJo software (Tree Star, Inc.) for flow cytometry data analysis. STATA version 15 or SPSS version 22 and Graph pad prism version 8 will be used for statistical inferences and to determine if dietary salt is associated with inflammation and/or immune activation in HIV. Analysis of variance or the Kruskal–Wallis test to compare pro- and anti-inflammatory cytokines and cells, activation markers and hypertension status in all four groups will be employed followed by the Dunnett post hoc test for pairwise comparisons or the Tukey test to compare all the groups with the control (HIV negative normotensive) group. We will also compare the means of the markers of immune cell activation between salt-sensitive and salt-resistant subjects, dichotomized by a drop of $\geq 10$ mmHg in SBP owing to salt loading, as described above. For this purpose, we will carry out unpaired t-tests for normally distributed data or Mann–Whitney tests for non-normally distributed data.

Regression models will be used to examine the relationship between outcome variables and selected determinant factors, to determine the impact of each variable on the outcomes and to control for confounding.

Discussion
Murine model and few human studies have shown that excess dietary salt induces BP elevation and activates cells of the innate and adaptive immune system [14, 20]. The interaction between dietary salt, BP and immune activation is a fairly new concept with recent evidence that salt can accumulate in skin without commensurate water retention and induce inflammation [1]. Furthermore, high consumption of dietary salt correlates positively with BP and is associated with the development or exacerbation of hypertension [21–23].

Previous studies have found that dendritic cells (DCs) accumulate isolevuglandin (IsoLG)-protein adducts during hypertension [4]. IsoLGSs are highly reactive products of oxidation of fatty acids that rapidly adduct to lysines on proteins and their accumulation is associated with DC activation [4]. Recent studies have established that elevated Na⁺ is a potent stimulus for IsoLG-protein adduct formation in murine DCs [20]. Na⁺ enters DCs through amiloride sensitive transporters. Intracellular Na⁺ is exchanged for calcium (Ca²⁺) via the Na⁺/Ca²⁺ exchanger. Ca²⁺ activates protein kinase C (PKC) which in turn phosphorylates the NADPH oxidase subunit p47phox. This leads to activation of the NADPH oxidase, increased superoxide ($O_2^-\cdot$) and IsoLG-protein adduct formation [20]. Adoptive transfer of salt-exposed DCs primes hypertension in response to a sub-pressor dose of angiotensin II and IsoLG-protein adduct formation is absent in mice lacking the NADPH oxidase and pharmacological scavenging of IsoLGSs prevents DC activation, hypertension and end-organ damage [4, 20] (Fig. 2).

Fig. 2  HIV and Salt-sensitive hypertension working Hypothesis. We hypothesize that HIV infection and treatment can lead to increased accumulation of sodium in tissues. This can activate the immune system leading to hypertension. Sodium enters antigen presenting cells and is exchanged for calcium through the sodium hydrogen exchanger 1. Calcium activates protein kinase C (PKC), which then activates NADPH oxidase by phosphorylating its subunit p47phox. This leads to increased production of superoxide with subsequent formation of immunogenic isolevuglandin (IsoLG)-protein adducts. IsoLGSs activate DCs which in turn promote T cell proliferation and production of cytokines that contribute to salt-sensitive hypertension [4]
These studies in mice suggest that the immune system can exhibit salt-sensitivity, however the role of Na+ in activating human antigen presenting cells such as monocytes, and the interplay between elevated Na+ and salt sensitivity, has not been defined in the context of HIV. Sodium intake is positively associated with BP and accounts for much of the age-related increase in BP [24, 25]. Independent of other factors, excess intake of salt is associated with an increased risk of stroke [26, 27], cardiovascular disease and other adverse outcomes including death [28]. However, the effects of salt on BP may vary among individuals due to salt-sensitivity and salt resistance [26, 29]. This study will identify patients with or without salt-sensitive hypertension among HIV-negative and HIV-positive participants and examine levels of IsoLG-protein adducts and the activation markers of monocytes.

There are no studies known to us that have studied salt-sensitive hypertension and immune activation in PLWH. With this novel perspective, it is expected that PLWH may exhibit increased salt-sensitivity and immune activation compared to HIV negative individuals. This study has the potential to create points of intervention that will improve the management of hypertension and prevent dietary salt-related hypertension in future studies. Moreover, there is evidence that in the general population of Zambia, salt consumption exceeds that recommended by WHO [22]. This study is therefore, of clinical interest.

**Strengths of the study**

1. This is likely, the first study known to us that will explain or report a possible relationship between HIV, immune activation and salt-sensitive hypertension.
2. Generally, salt consumption in Africa and Zambia is higher than recommended. With increasing incidence of hypertension, knowledge about a possible interaction in this research is of clinical interest.

**Limitations**

- Sodium excretion exhibits circaseptan rhythm and infra-radian rhythm depending on salt intake, increases or decreases in intake affects BP. However, a 24-h urine recovers about 95% of dietary salt intake.
- Though there are no established criteria for salt-sensitivity, we employ already published methods with maximal controls to avoid possible confounders. Information generated will provide baseline data for hypotheses generation for future planned studies.
- The underlying factors associated with HIV, immune activation and salt-sensitive hypertension are multifactorial, many factors not studied are likely to confound the results of the study. However, the experimental nature of the study will minimize confounding.
- This proposed study cannot determine causality and exhaust the mechanism and possible interactions between salt, immune system and HIV. Hence, further studies are warrantable.

**Additional file**

Additional file 1. Laboratory procedures.

**Abbreviations**

AMBP: ambulatory blood pressure; ART: antiretroviral therapy; BMI: body mass index; BP: blood pressure; CRP: C-reactive protein; DC: dendritic cell; DMSO: dimethyl sulfoxide; ECG: electrocardiogram; GM-CSF: granulocyte macrophage stimulating factor; HISH: HIV; Immune-activation and salt-sensitive hypertension study; FMD: flow minus one; HEPES: 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid; HIV: human immunodeficiency virus; IFN-γ: interferon gamma; IL: interleukin; IsoLG: isolevuglandins; LCH: Livingstone Central Hospital; MIFlowCyt: Minimum Information about a Flow Cytometry Experiment; PBMC: peripheral blood mononuclear cells; PLWH: people living with HIV; PMA: phorbol-12-myristate 13-acetate; PKC: protein kinase C; RPMI: Roswell Park Memorial Institute; sCD14: soluble CD14; TNF-α: tumor necrosis factor alpha; UVP: University of Zambia Vanderbilt partnership; WB: whole blood, WC: waist circumference; WHO: World Health Organizations; WHR: waist-to-hip ratio.

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**Authors’ contributions**

SKM and AK conceived the study. SKM, BMH, SN, GK, DCH, WM, JRK, AK and SMM contributed to the writing of the manuscript. SKM is the principal investigator and guarantor. All authors provided feedback. All authors read and approved the final manuscript.

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**Availability of data and materials**

All data generated or analyzed during this study are included in this published article. For other data, these may be requested through the corresponding author.

**Ethics approval and consent to participate**

Ethical approval was obtained from the University of Zambia Health Sciences Research Ethics Committee (UNZAHSREC) (Assurance no. FWA00026270, IRB no. 00011000) on the 2nd April 2019. Permission to conduct the study was granted by the Livingstone Central Hospital Administration. All participants will be asked to consent by signing a consent form before being included in the study. All data collected will be identified and used for research purposes only.
Consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

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References
1. Kirabo A. A new paradigm of sodium regulation in inflammation and hypertension. Am J Physiol Regul Integr Comp Physiol. 2017;313:R706–10.
2. Elijovich F, Weinberger MH, Anderson CAM, Appel LJ, Bursztyn M, Cook NR, et al. Salt sensitivity of blood pressure: a scientific statement from the American Heart Association. Hypertens Dallas Tex. 1979;2016(68):e7–46.
3. Whelton PK, Carey RM, Aronow WS, Casey DE, Collins KJ, Dennison Himmelfarb C, et al. ACC/AHA/ACP/AAPA/ABC/ACPAM/AGS/APhA/ASH/ASPC/NMA/PCNA Guideline for the Prevention, Detection, Evaluation, and Management of High Blood Pressure in Adults: executive summary. A Report of the American College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines. Hypertens Dallas Tex. 2017;1979:2017.
4. Kirabo A, Fontana V, de Faria APC, Loperena R, Galindo CL, Wu J, et al. DC isoformaldehyde-modified proteins activate T cells and promote hypertension. J Clin Invest. 2014;124:4642–56.
5. McMaster WG, Kirabo A, Madhur MS, Harrison DG. Inflammation, immunity, and hypertensive end-organ damage. Circ Res. 2015;116:1022–33.
6. Zhang J, Crowley SD. Role of T lymphocytes in hypertension. Curr Opin Pharmacol. 2015;21:14–9.
7. Schatz V, Neubert P, Schröder A, Binger K, Gebhard M, Müller DN, et al. Elementary immunology. Na+ as a regulator of immunity. Pediatr Nephrol Berl Ger. 2017;32:201–10.
8. Xu Y, Chen X, Wang K. Global prevalence of hypertension among people living with HIV: a systematic review and meta-analysis. J Am Soc Hypertens JASH. 2017;11:530–40.
9. Palacios R, Santos J, García A, Castells E, González M, Ruiz J, et al. Impact of highly active antiretroviral therapy on blood pressure in HIV-infected patients. A prospective study in a cohort of naive patients. HIV Med. 2006;7:10–5.
10. Tripathi A, Jerrell JM, Skelton TN, Nickels MA, Duffus WA. Incidence of primary hypertension in a population-based cohort of HIV-infected compared with non-HIV-infected persons and the effect of combined antiretroviral therapy. J Am Soc Hypertens. 2015;9:351–7.
11. Wada NJ, Jacobson LP, Margolick JB, Breen EC, Macatangay B, Penugonda S, et al. The effect of HAART-induced HIV suppression on circulating markers of inflammation and immune activation. AIDS Lond Engl. 2015;29:463–71.
12. Mohan T, Bhatnagar S, Gupta DL, Rao DN. Current understanding of HIV-1 and T-cell adaptive immunity: progress to date. Microb Pathog. 2014;73:60–9.
13. He FJ, MacGregor GA. Reducing population salt intake worldwide: from evidence to implementation. Prog Cardiovasc Dis. 2010;52:363–82.
14. He FJ, Li J, MacGregor GA. Effect of longer term modest salt reduction on blood pressure: cochrane systematic review and meta-analysis of randomised trials. BMJ. 2013;346:f1325.
15. Aburto NJ, Ziolkovska A, Hooper L, Elliott P, Cappuccio FP, Meerpohl JJ. Effect of lower sodium intake on health: systematic review and meta-analyses. BMJ. 2013;346:f1326.
16. Faul F, Edfelder E, Buchner A, Lang A-G. Statistical power analyses using G’S Power 3.1: tests for correlation and regression analyses. Behav Res Methods. 2009;41:1149–60.
17. Faul F, Edfelder E, Lang A-G, Buchner A. G*Power 3: a flexible statistical power analysis program for the social, behavioral, and biomedical sciences. Behav Res Methods. 2007;39:175–91.
18. He J, Gu D, Chen J, Jaquish CE, Rao DC, Hixson JE, et al. Gender differences in blood pressure responses to dietary sodium intervention in the ginseng study. J Hypertens. 2009;27:48–54.
19. Rakova N, Jütten K, Dahlmann A, Schroder A, Linz P, Kopp C, et al. Long-term space flight simulation reveals infradian rhythmicity in human Na(+) balance. Cell Metab. 2013;17:235–31.
20. Barbaro NR, Foss JD, Krystalh DO, Tsya N, Kumaesan S, Xiao L, et al. Dendritic cell aminolide-sensitive channels mediate sodium-induced inflammation and hypertension. Cell Rep. 2017;21:1009–20.
21. Ware LJ, Charlton K, Schutte AE, Cockeran M, Naidoo N, Kowal P. Associations between dietary salt, potassium and blood pressure in South African adults: WHO SAGE Wave 2 Salt & Tobacco. Nutr Metab Cardiovasc Dis NMCD. 2017;27:784–91.
22. Oelke ND, Rush KL, Goma FM, Barker J, Marck P, Pedersen C. Understanding perceptions and practices for zambian adults in western province at risk for hypertension: an exploratory descriptive study. Glob J Health Sci. 2016;8:248–59.
23. Azinge EC, Sofola OA, Silva BO. Relationship between salt intake, salt-taste threshold and blood pressure in Nigerians. West Afr J Med. 2011;30:373–6.
24. Mente A, O’Donnell MJ, Rangarajan S, McQueen MJ, Poirier P, Welgosz A, et al. Association of urinary sodium and potassium excretion with blood pressure. N Engl J Med. 2014;371:601–11.
25. Takase H, Sugiuara T, Kimura G, Ohhe N, Dohi Y. Dietary sodium consumption predicts future blood pressure and incident hypertension in the Japanese normotensive general population. J Am Heart Assoc. 2015;4:e001959.
26. Strazzullo P, D’Elia L, Kandala N-B, Cappuccio FP. Salt intake, stroke, and cardiovascular disease: meta-analysis of prospective studies. BMJ. 2009;339:b4567.
27. Whelton PK. Sodium, potassium, blood pressure, and cardiovascular disease in humans. Curr Hypertens Rep. 2014;16:465.
28. Whelton PK, Appel LJ, Sacco RL, Anderson CAM, Antman EM, Campbell N, et al. Sodium, blood pressure, and cardiovascular disease: further evidence supporting the American Heart Association sodium reduction recommendations. Circulation. 2012;126:2880–90.
29. Sanaa H, Jones JE, Jose PA. Genetics of salt-sensitive hypertension. Curr Hypertens Rep. 2011;13:55–66.

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