Plasma soluble factor following two decades prolonged suppressive antiretroviral therapy in HIV-1-positive males
A cross-sectional study

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Abstract
Acute human immunodeficiency virus (HIV) infection is associated with a marked induction of several pathways that are linked to inflammation and CD4+ T-cell depletion. Many of these processes do not fully resolve on short-term combination antiretroviral therapy (cART) (<5 years), despite complete and durable suppression of viremia. The effects of long-term (>15 years) successful antiretroviral therapy (ART) and the linkage between levels of biomarkers remain unclear. Therefore, the present study aims to assess the host plasma proteome in a well-defined clinical material from HIV-1-positive male patients on successful long-term ART (>15 years) and compared them with age-matched healthy controls and treatment-naive male patients with viremia in a cross-sectional manner.

Plasma samples were obtained from 3 categories of age-matched HIV-1-positive male patients on long-term successfully (ART, n=10) with a median (Interquartile range, IQR) of 19 (17–20) years, treatment-naive patients with viremia (VP, n=14), and HIV-1-negative persons (HC, n=11). Plasma proteome was analyzed using the proximity extension assay targeting 92 factors. Statistical analyses were performed with GraphPad Prism v7, R-packages, and Qlucore Omics Explorer v3.2. Functional enrichment analyses was performed by Kyoto Encyclopedia of Genes and Genomes (KEGG), and interactions of specific molecules were identified using Path Designer integrated into Ingenuity Pathway Analysis (IPA).

Group wise comparison identified 53 soluble factors, which differed between the groups \(P<.05\). Cluster analysis identified 13 discrete soluble factors (CD8A, CRTAM, CXCL13, EGF, CD5, CD40, CXCL9, Gal-1, IL12RB1, KLRD1, PD-1, CASP-8 and TNFRSF9) between the studied groups (adjusted \(P<.001\)). The long-term successfully ART-treated individuals clustered and networked with the HC while VPs clustered separately. All of the proinflammatory cytokines and chemokines were normalized back to levels of healthy controls in long-term successfully ART-treated individuals, but not the levels of KLRD1 and PGDFB.

skL1RD1 that is involved in the regulation of natural killer cell (NK) mediated cytotoxicity, failed to be restored to the level of HIV-negative individuals despite successful long-term ART. Additional analysis of NK cells along with T-cell subsets can provide insights into the long-term effects of ART on the immune system.

Abbreviations: ANG-1 = angiopoietin-1, ANG-2 = angiopoietin-2, ANOVA = analysis of variance, cART = combination antiretroviral therapy, CASP-8 = caspase 8, CCL13 = chemokine (C-C motif) ligand 13, CCL19 = chemokine (C-C motif) ligand 19/macrophage inflammatory protein-3-beta, CCL23 = chemokine (C-C motif) ligand 23/macrophage inflammatory protein 3, CCL4 = chemokine (C-C motif) ligand 4/macrophage inflammatory protein-1 beta, CD27 = cluster of differentiation 27, CD4 = cluster of differentiation 4, CD40 = cluster of differentiation 40/tumor necrosis factor receptor super family member 5, CD40-L = cluster of differentiation 40 ligand/tumor necrosis factor super family member 5, CD8A = cluster of differentiation 8a, CRTAM = cytotoxic and regulatory T cell molecule, CSF-1 = macrophage colony-stimulating factor 1, CXCL1 = chemokine (C-X-C motif) ligand 1, CXCL11 = Chemokine (C-X-C motif) ligand 11, CXCL13 = chemokine (C-X-C motif) ligand 13/B lymphocyte chemoattractant, CXCL5 = chemokine (C-C motif) ligand 5, CXCL9 = chemokine (C-C motif) ligand 9, DCN = decorin, EGF = epidermal growth factor, FDR = false discovery rate, Galectin-L = Galectin-1, Gal-9 = Galectin-9, HC = healthy control, HIV-1 = human immunodeficiency virus type 1, HLA = human leukocyte antigen, HO-1 = heme oxygenase 1, IL = interleukin, IL12RB1 = interleukin 12 receptor subunit beta 1, IPA = Ingenuity Pathway Analysis, IQR = interquartile range, KEGG = Kyoto Encyclopedia of Genes and Genomes, KLRD1 = killer cell lectin-like receptor subfamily D, member 1, NK = natural killer cell, NPX

Editor: Alejandro Vallejo.
This study was funded by Swedish Research Council (2017-01330), Stockholm County Council (ALF 20160074), and Jonas Söderquist’s Stipendium for Experimental Virology and Immunology Research-2016 to UN.
The authors report no conflicts of interest.

Supplemental Digital Content is available for this article.
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Medicine (2018) 97:5(s09759)
Received: 8 November 2017 / Received in final form: 10 January 2018 / Accepted: 11 January 2018
http://dx.doi.org/10.1097/MD.000000000009759

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1. Introduction

Acute HIV infection is associated with a distinct induction of several pathways that are linked to inflammation, CD4+ T-cell depletion, and the establishment of the viral reservoir. HIV-induced dysregulation of the inflammatory network is multidimensional based on the immune status of the patients, any co-infections, and microbial translocation. The effect of combination antiretroviral therapy (cART) on mortality and morbidity of HIV-infected patients is remarkable. However, the life spans of also well-treated patients are shorter than those of matched HIV-negative controls. This is to a large extent due to an increased rate and deaths in comorbidities and effect of ART such as cardiovascular events, kidney disease, and others. This could be a result of persistent immune activation despite successful ART, which is poorly understood. Most of the studies have been conducted on small numbers of soluble biomarkers or analyzed only a shorter duration of therapy, given the technological challenges and limited availability of a long-term quality patients’ follow-up. The effects of successful long-term ART and the cross-talk between the soluble biomarkers and subsequent altered immunological pathways remain unclear and could provide useful insights into disease progression and pathogenesis. With the advancement of high-throughput technologies, it is now possible to analyze a large panel of soluble factors simultaneously. Proximity extension assay (PEA) technology (Olink Bioscience AB, Uppsala, Sweden) is one of these high-throughput multiplex immunoassays and measures 92 soluble factors simultaneously using only 1 μL plasma. The method has high sensitivity and specificity and can detect low abundant proteins and be used in wellness study and other diseases including cardiovascular, inflammatory disease, metabolic disorders, and so on to identify novel biomarkers. Therefore, the aim of the present study is to assess the host plasma proteome in a well-defined clinical material from HIV-1-positive male patients on successful long-term ART (>15 years) and compared them with age-matched healthy controls and treatment-naive male patients with viremia.

2. Materials and methods

2.1. Patients

Cross-sectional plasma samples were obtained from 3 categories of age-matched HIV-1-positive male patients on ART (n = 10) with a median (IQR) of 19 (17–20) years, treatment-naive patients with viremia (VP, n = 14) and HIV-1-negative persons (HC, n = 11). The patients were selected based on the clinical data obtained from the Swedish InfCareHIV cohort who were attending the Infectious Disease Clinic at Karolinska University Hospital, Stockholm. The inclusion criteria for the successful long-term ART was male with at least 10 years of suppressive therapy with not more than 1 viral blips (viral load <100 copies/mL). The 2 decades long clinical data were obtained from the prospective Swedish InfCareHIV cohort. Samples were collected between June 2015 and April 2016. Clinical data included were between January 1982 and June 2017.

2.2. Plasma proteome

Plasma samples were analyzed using the PEA and the Olink Immuno-oncology panel (Olink Bioscience AB). This panel includes 92 proteins. The protein analysis is reported as normalized protein expression levels (NPX), which are Ct values normalized by the subtraction of values for extension control, as well as interplate control; the scale is shifted using a correction factor (normal background noise) and reported in log2 scale.

2.3. Functional enrichment and interactome analysis

Functional enrichment was performed by Kyoto Encyclopedia of Genes and Genomes (KEGG), and interactions of specific molecules were identified using Path Designer integrated into QIAGEN’s Ingenuity Pathway Analysis (IPA, QIAGEN Redwood City, www.qiagen.com/ingenuity).

2.4. Statistical analysis and visualization

Given the low number of samples, we applied non-parametric test using GraphPad Prism v7. Group-wise comparison was performed using Mann–Whitney U test for continuous variables and chi-square test for discrete variables. The multi-group analysis was performed by Kruskal–Wallis test. Change in CD4 T-cell count from the start of therapy and at the time of sampling was performed using Wilcoxon matched-pairs signed-rank test. The explorative analysis was carried out in Qucore Omics Explorer version 3.2. Multi-group comparison was performed using analysis of variance (ANOVA) at false discovery rate (FDR) adjusted P(q) <.05. Cluster analysis (k-means) was performed at stricter FDR <0.001 using ANOVA. Venn diagram was created in InteractiVenn. The differential profile (heatmap) of the soluble factors (proteome) was analyzed using Qucore Omics Explorer version 3.2. CIRCOS plot was used to visualize the circular plot.

2.5. Ethical considerations

The study is approved by regional ethics committees of Stockholm (2013/1944–31/4). All participants have given informed consent.

3. Results

3.1. Patient characteristics

Patients clinical and demographic characteristics are given in Table 1. All the patients are male with a median age of 50 years. There was a significant difference in HIV-1 seropositivity.
between the individuals of VP and ART group (P < .001). The median (IQR) seropositivity year in the patients with long-term ART was 1995 (1991–1997). There was a significant difference in nadir CD4+ T-cell count (324 vs. 135; P < .001) and CD4+ T-cell count at the time of sample collection (390 vs 520; P = .008) between the VP and ART group of patients. The median (IQR) duration of ART in the long-term ART group was 19 years (17–20) with median (IQR) of 16 (15–18) years of suppressive therapy (viral load below detection level). The gain in the CD4+ T-cell count was statistically significant across the group (P = .002) (Fig. 1A). The 2-decade-long viral load count is given in Figure 1B. None of the patients had any co-infection at the time of sampling.

3.2. Plasma proteome profiling

Group-wise comparison identified 53 soluble factors which differed between the groups (P < .05; Mann–Whitney U test), of which 47 factors were different between HC and VP, 45 between long-term ART and VP, and 3 between HC and ART (Fig. 2A and Supplementary file 1, http://links.lww.com/MD/C95). Among those factors, 5 factors were unique between ART and VP (CCL13, CCL4, CXCL1, CD4, and CSF-1), while 7 factors were unique between HC and VP (DCN, PIGF, HO-1, CCL23, TNFRSF21, CXCL11, and VEGFR-2), and only 1 secretory factor between HC and ART, namely angiopoietin-2 (ANG-2). The 2 chemokines CCL4 and CCL13 were significantly elevated in ART-treated patients compared to VP, and even a trend was observed compared to HC (Fig. 2B). In patients on long-term suppressive ART, all of the pro-inflammatory molecules examined (IL-7, IL-12, and soluble IL receptor IL12RB1) went back to levels of healthy controls. Among the 29 cytokines tested, 93% (27/29) of them went back to physiological levels in the long-term ART group, but not the levels of the non-cytokine molecules soluble killer cell lectin-like receptor subfamily D member 1 (KLRD1, also CD94) (P = .02) and platelet-derived growth factor subunit B (PDGFB) (P = .0485) (Fig. 2B). ANG-2, an endothelial activation marker, did also not normalize to the healthy state following long-term suppressive therapy.

To further dissect which biomarkers can differentiate the groups, cluster analysis (k-means) was performed at significance level FDR < .05. Cluster analyses identified higher levels of

![Figure 1](https://example.com/figure1.png)

**Figure 1.** Longitudinal clinical follow-up of ART patients. (A) CD4+ T-cell count at the start of therapy and at the time of sampling. (B) Viral load.
CCL13, SDF-1, TWEAK, CXCL5, CD40-L, CASP8, TNFRSF12A, DCN, and TNFRSF21 in ART and HC, while CXCL11, Gal-9, IL-12, CXCL13, CXCL10, CCL19, KLRD1, CD8A, IL12RB1, CD27, Granzyme-H, and Granzyme-A in VP (Fig. 3A). The patients with successful long-term ART clustered and networked with HC, while VP grouped separately. We further used a stringent statistical significance to find the precise network and potential markers between the VP and ART/HC groups. Cluster and network analysis identified 13 soluble factors (CD8A, CRTAM, CXCL13, EGF, CD5, CD40, CXCL9, Gal-1, IL12RB1, KLRD1, PD-1, CASP-8, and TNFRSF9) which were significantly discrete at false discovery rate (FDR) adjusted to \( P(q) < 0.001 \) using ANOVA (Fig. 3B).

### 3.3. Functional enrichment analysis

We performed functional enrichment analysis by Kyoto Encyclopedia of Genes and Genomes (KEGG) using 53 proteins (defined by Mann–Whitney U test significant differences between groups). In total, 32 pathways are enriched with FDR < 0.05 (Supplementary data file 2, http://links.lww.com/MD/C94). Among these pathways, cytokine–cytokine receptor interaction (FDR = \( 3.62 \times 10^{-29} \)) and chemokine signaling pathway (FDR = \( 1.26 \times 10^{-10} \)) included together 29 soluble factors. Apart from that, antigen processing and presentation (FDR = \( 0.007 \)), and natural killer cell mediated cytotoxicity (FDR = \( 0.037 \)) are enriched. The CIRCOS plot indicates the difference of soluble marker levels between groups and subsequent signaling pathways.

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**Figure 2.** Level of plasma soluble factor. (A) Venn diagram of plasma soluble factors statistically difference \( P < .05 \) of the NPX value in a case–control manner. The sum of the numbers in each large circle represents the total number of statistically different proteins among various combinations (ART vs HC, HC vs VP and ART vs VP). The overlapping part of the circles represents common proteins between combinations. (B) Bean plot is indicating the level of the selected soluble factor. \( P < .05 \) is marked with * while \( P < .001 \) with **. ART = antiretroviral therapy, HC = healthy control, VP = viremic progressors.

**Figure 3.** The plasma proteome profiles using hierarchical clustering. (A) Cluster analysis based on the soluble factors at FDR < 0.05. Heatmap shows fold change +1.5 (red) to −1.5 (white). (B) Cluster (k means) and network analysis with FDR < 0.001, FDR = false discovery rate.
the molecules are involved in (Fig. 4A). There are several factors, which are present in more than 1 pathway. Further Ingenuity Pathway analysis (IPA) identifies that the expression of KLRD1 is regulated by proinflammatory molecules (IL15, IL21, IL2, IL4, IL12 complex, etc) and it regulates the tumor necrosis factor (TNF) and cytokine signaling pathways (Fig. 4B).

4. Discussion

Our study investigated the levels of soluble biomarkers in patients with nearly 2 decades successful ART with the highest number of proteins studied so far. Very long-term ART normalizes the level of most cytokines and chemokines explored to the degree of healthy individuals. However, some biomarkers, like KLRD1 and ANG-2, do not go back to healthy physiological levels indicating that immunological events still take place in HIV-1-infected patients despite long-term suppressive ART.

In concordance with other studies, elevations of pro-inflammatory cytokines and chemokines were seen in VP. As observed earlier, the 2 chemokines CCL4 and CCL13 were significantly elevated in ART-treated patients compared to VP,
Angiopoietin-2 binds to the cell surface receptor for angiopoietin-1 (ANG-1), namely TEK/TIE2, and thus, modulates ANG-1 signaling. An earlier study in females from Kenya observed, that increased ANG-2 plasma levels in chronic HIV-1 infection decrease after ART.[26] A higher level of ANG-2 is also associated with higher mortality.[27] However, in our study with male populations both in VP and ART, the median plasma level is lower than in HC, with statistical significance between ART and HC group. The study in Kenya also detected significant associations between the use of oral contraceptive pills and higher plasma ANG-2 levels in pregnancy.[26] They also observed that estrogen stimulates ANG-2 mRNA expression.[27] We, therefore, conclude that endothelial activation marker ANG-2 was lower in a male with advanced HIV infection and had no effect on ART initiation, though it does not increase to a healthy status.

The study has limitations that merit comments. First, the numbers of patients were relatively low. This is mainly because of limited numbers of HIV-infected individuals with very long-term suppressive therapy for whom adequate clinical and demographic information is available. These groups of patients were identified from nearly 10,000 patients who got treatment care in Sweden and availability of the plasma samples. Second, we only looked into 92 plasma soluble factors. However, to best of our knowledge, this is the most substantial amount of markers that have been studied to date.

In conclusion, this is the first study, which investigated the levels of soluble biomarkers in patients with nearly 2 decades successful ART with the highest number of proteins studied so far. Very long-term ART normalizes the level of most cytokines and chemokines explored to the degree of healthy individuals. However, some biomarkers do not go back to healthy physiological levels indicating that immunological events still take place in HIV-1-infected patients, despite long-term suppressive ART. Future analyses of cellular subsets other than T-lymphocyte populations, like NK cells, are likely to help us gain further insights into the long-term restoration of the immune system by ART.

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