RESEARCH ARTICLE

Association between NF-κB polymorphism and age-related macular degeneration in a high-altitude population

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Abstract

Objective
To investigate the association between the nuclear factor kappa B (NF-κB) gene polymorphism and age-related macular degeneration (AMD) in a high-altitude population.

Methods
Fifty-five patients with AMD and 57 control subjects were recruited from the Qinghai Provincial People’s Hospital, China. Genomic DNA was extracted from the blood sample of each participant. Four NF-κB polymorphisms (rs3774959, rs3774932, rs3774937, and rs230526) were genotyped using a MassARRAY system. The genotype and allele frequencies were compared between the case and control groups using the chi-squared test or Fisher’s exact test.

Results
There was no significant difference in sex, age, hypertension, diabetes, blood lipid level or smoking and drinking status between the AMD and control groups (P > 0.05). The genotype distributions of four NF-κB polymorphisms were in accordance with Hardy-Weinberg equilibrium in the control group (P > 0.05). The frequencies of genotype AA of rs3774932 and genotype CC of rs3774937 were nominally significantly higher in the AMD group than in the control group (P = 0.046 and 0.023, respectively), although these associations did not survive the Bonferroni correction (corrected P > 0.05). Genotype distributions of rs3774959 and rs230526 were not significantly different between the two groups (P = 0.08 and 0.16, respectively). No significant difference in the allele frequencies of the four polymorphisms was found between the AMD and control groups (P > 0.05).

Conclusions
Genotype AA of rs3774932 and genotype CC of rs3774937 in NF-κB might be risk factors for AMD.
Introduction

Age-related macular degeneration (AMD) is one of the most common irreversible blinding eye diseases, which seriously affects patients’ quality of life. The disease affects tens of millions of people all over the world [1]. In China, the prevalence of AMD ranged from 2.44% in people aged 45–49 years to 18.98% in people aged 85–89 years [2]. The development of AMD is related to many factors, such as age, sex, race, eating habits, obesity, and sun exposure, but the specific etiology is not clear [3]. Among various factors affecting the pathogenesis of AMD, mitochondrial damage in the retinal pigment epithelium (RPE) is an important cause of RPE dysfunction [4]. Studies have shown that mitochondrial damage in the RPE is closely related to inflammation, autophagy, and apoptosis [5–7]. In recent years, results from many studies have suggested that genetic factors may play an important role in the development of AMD. Studies have shown that the heritability of AMD can be as high as 71% [8], which indicates that genetic factors are closely related to the development of AMD. Over 50% of the heritability of AMD has been explained by two major genes (CFH and ARMS2/HTRA1), making it one of the most well-defined genetically complex disorders [9]. A recent genome-wide association study (GWAS) on 43,566 subjects identified 52 independently associated variants spanning 34 loci for AMD [10].

The nuclear factor kappa B (NF-κB) family of transcription factors plays a pivotal role in regulating inflammatory response, immune function, and malignant transformation [11]. In addition, NF-κB affects the expression of genes for cell [6] differentiation, proliferation, and survival in almost all multicellular organisms [12]. There is also a significant correlation between NF-κB and autophagy [13]. NF-κB is composed of a group of homodimer and heterodimer protein complexes, and p50/p65/p53 heterodimer complex is the most common complex [14]. Results from studies have suggested that the NF-κB gene plays a role in the occurrence of lung cancer, colorectal cancer, breast cancer, and other cancers [15–18]. Some studies have shown that the NF-κB gene has strong correlation with inflammation and autophagy, while inflammation and autophagy are important pathogenesis of AMD [19–23]. A recent genome-wide meta-analysis identified novel loci associated with AMD, including C4BPA-CD55, ZNF385B, ZBTB38, and NFKB1 [24].

However, researchers have not investigated the association between the NF-κB gene and the AMD risk in a Chinese population. In the present study, four single nucleotide polymorphisms (SNPs) of NF-κB (rs3774959, rs3774932, rs3774937, and rs230526) were selected for genotyping and their associations with the risk of AMD were analyzed in a high-altitude Chinese population.

Methods

Study participants

Fifty-five patients with AMD were and 57 control participants were recruited from the Qinghai Provincial People’s Hospital, China from December 2016 to December 2019. This study was conducted in accordance with the tenets of the Declaration of Helsinki and has been approved by the local hospital ethics committee (approval number 2017–21). Written informed consent was obtained from all participants.

According to the Preferred Practice Pattern Guidelines: Age-related Macular Degeneration [25] and the "Chinese clinical diagnosis and treatment pathway for age-related macular degeneration" [26], AMD was diagnosed when the presence of one or more of the following criteria was met: 1) medium-sized hyaline warts (>63 μm in diameter); 2) RPE abnormalities such as hypopigmentation, pigment proliferation, migration, and metaplasia; and 3) any of the
following characteristics: retinal pigment epithelium map atrophy, choroidal neovascularization (exudative), polypoid choroidal vasculopathy, and retinal hemangioma hyperplasia.

The inclusion criteria for AMD patients were as follows: 1) long-term (>20 years) residence at altitudes >2,000 m; 2) age >40 years; and 3) diagnosed with AMD based on the diagnostic criteria. The exclusion criteria for AMD patients were as follows: 1) late vitelliform macular degeneration; 2) choroidal neovascularization with high myopia; 3) Stargardt disease; 4) retinal vascular occlusion; 5) chorioretinitis; 6) diabetic retinopathy; 7) hypertensive retinopathy; and 8) other eye diseases.

All control participants were >40 years of age and diagnosis of AMD and other fundus diseases had been ruled out by fundus examination. Participants with other diseases such as high myopia and glaucoma were also excluded. In addition, those with serious systemic diseases such as hypertension, diabetes, renal insufficiency, blood diseases, and benign or malignant tumors were excluded.

Sample collection and DNA extraction
Two milliliters of peripheral blood was collected from all participants. After anticoagulant treatment, the samples were frozen at −8°C before use. Genomic DNA was extracted using xxx. DNA quality was assessed using the NanoDrop 2000 spectrophotometer (Thermo Fisher Scientific, Waltham, MA, USA).

SNP genotyping
Single nucleotide polymorphisms (SNPs) were genotyped using a MassARRAY system (Sequenom, San Diego, CA, USA). Single-base extension primers were designed and synthesized by the Sangon Biotech (Shanghai, China). The sequences of the primers are listed in Table 1.

The PCR reaction was performed using a MassARRAY mass spectrometer (Sequenom, San Diego, CA). Genotypes were called using the MassARRAY RS1000 software (Sequenom).

Statistical analysis
SPSS statistical software (version 25.0; IBM, Armonk, NY, USA) was used for statistical analysis. The numerical data were expressed as mean ± standard deviation and compared between the case and control groups using the Student’s t test. The categorical data were expressed as n (%) and compared between the two groups using the chi-squared test. Hardy-Weinberg equilibrium was tested for genotype distributions using the chi-squared test. Genotype and allele frequencies were compared between the case and control groups using the chi-squared test or Fisher’s exact test. We corrected for multiple tests using the Bonferroni method. A P < 0.05 was considered statistically significant. Power analysis was performed using the Genetic Power Calculator [27].

Table 1. Sequences of the PCR primers used in this study.

| SNP      | Forward primer                  | Reverse primer                  | Amplicon (bp) |
|----------|---------------------------------|---------------------------------|---------------|
| rs3774959| 5'-AGTAACACACCATAGGGCAGTAAACG-3' | 5'-TGACTGATGAGATACGGGGCTA-3'    | 184           |
| rs3774932| 5'-TCTAGCAGAAATCCCCAAACTGAAAT-3' | 5'-ATTCAGAAGTCCACATTATCGTAC-3'  | 332           |
| rs3774937| 5'-CATTAATGCATAGGGTCTCTCTCT-3'  | 5'-AATCGTCTCAAA1ACCTGTCTCAG-3'  | 242           |
| rs230526 | 5'-GCTTGGCAGACAGCAATTAA-3'      | 5'-TACGGGCAAAAGGTACATAAC-3'     | 307           |

PCR, polymerase chain reaction; SNP, single nucleotide polymorphism.

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Results

Demographic and clinical features of patients with AMD and controls

The AMD group consisted of 28 males and 27 females, with an average age of 66.5 (±11.8) years. The control group had 29 males and 28 females, with an average age of 65.5 (±11.5) years. There was no significant difference in sex, age, hypertension, diabetes, blood lipid level, or smoking or drinking status between the AMD group and the control group (P > 0.05; Table 2).

Genotype distributions of \(\text{NF-}\kappa\text{B}\) gene polymorphisms in patients with AMD and controls

Genotype distributions of all four SNPs followed Hardy-Weinberg equilibrium in the control group (P > 0.05). The frequencies of genotype AA of rs3774932 and genotype CC of rs3774937 were nominally significantly higher in the AMD group than in the control group (P = 0.046 and 0.023, respectively), although these associations did not survive the Bonferroni correction (corrected P > 0.05). Genotype distributions of rs3774959 and rs230526 were not significantly different between the two groups (P = 0.08 and 0.16, respectively; Table 3).

Allele distributions of \(\text{NF-}\kappa\text{B}\) gene polymorphisms in patients with AMD and controls

There was no significant difference in allele frequencies of four polymorphisms between the AMD and control groups (P > 0.05; Table 4).

Table 2. Demographic and clinical features of patients with AMD and controls.

| Feature          | AMD (n = 55) | Controls (n = 57) | P  |
|------------------|-------------|-----------------|----|
| Female (%)       | 27 (49.1)   | 28 (49.1)       | 0.99|
| Age (years)      | 66.5 ± 11.8 | 65.5 ± 11.5     | 0.13|
| Hypertension (%) | 21 (38.2)   | 19 (33.3)       | 0.69|
| Diabetes (%)     | 7 (12.7)    | 9 (15.8)        | 0.98|
| Hyperlipidemia (%) | 9 (16.4)   | 7 (12.3)        | 0.54|
| Smoking (%)      | 4 (7.3)     | 6 (10.5)        | 0.55|
| Drinking (%)     | 4 (7.3)     | 6 (10.5)        | 0.55|

Table 3. Genotype distributions of \(\text{NF-}\kappa\text{B}\) gene polymorphisms in patients with AMD and controls.

| SNP         | Allele | Group | Genotype, n (%) | P  |
|-------------|--------|-------|----------------|----|
| rs3774959   | G/A    | AMD   | 22 (0.400)      | 1/1|
|             |        |       | 21 (0.382)      | 1/2|
|             |        |       | 12 (0.218)      | 2/2|
| rs3774932   | A/G    | AMD   | 21 (0.382)      | 1/1|
|             |        |       | 19 (0.345)      | 1/2|
|             |        |       | 15 (0.273)      | 2/2|
| rs3774937   | T/C    | AMD   | 29 (0.527)      | 1/1|
|             |        |       | 15 (0.273)      | 1/2|
|             |        |       | 11 (0.200)      | 2/2|
| rs230526    | G/A    | AMD   | 19 (0.345)      | 1/1|
|             |        |       | 23 (0.418)      | 1/2|
|             |        |       | 13 (0.236)      | 2/2|

AMD, age-related macular degeneration; SNP, single nucleotide polymorphism.

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Discussion

AMD is a complex, highly inherited multifactorial disease caused by the interaction of genetic and environmental risk factors [28]. In this study, we compared the frequencies of NF-κB gene polymorphisms between AMD cases and controls to explore the correlation between the NF-κB gene and AMD. The allele frequencies of the four SNPs analyzed in this study are comparable to the corresponding allele frequencies in East Asians reported in the public gnomAD database [29] (Table 4). Our results showed that the frequencies of genotype AA of rs3774932 and genotype CC of rs3774937 were nominally significantly higher in the AMD group than in the control group (Table 3), although these associations did not survive the Bonferroni correction (corrected $P > 0.05$). These findings suggest that individuals carrying genotype AA of rs3774932 or genotype CC of rs3774937 may have a higher risk of developing AMD.

Silico analyses suggest that SNP rs3774932 could change the Nkx2 and SIX5 motifs of the NF-κB protein, while SNP rs3774937 could change the DMRT1, LUN-1, and YY1 motifs of the NF-κB protein [30]. Functional studies such as the ChIP assay to identify the binding activity of these genotypes or luciferase reporter assay to test the function of these polymorphisms, especially the NF-κB binding site activity in genotype AA of rs3774932 or genotype CC of rs3774937, would be helpful to further elucidate the potential effects of these genotypes on gene transcriptional regulation and expression, consequently affecting the NF-κB pathway activation and/or susceptibility to AMD pathology.

This study had several limitations. First, the power of this study to detect the association between the NF-κB gene polymorphisms and AMD was limited due to small sample sizes. We estimated that this study had a power of 0.78 to detect the association at OR = 2.0 and a prevalence of 7.11% for people aged 65–69 years [2]. Therefore, the negative associations we observed after correcting for multiple tests in this study did not disprove potential real associations between the NF-κB gene polymorphisms and AMD. Second, the use of 40 years of age as an inclusion criterion for AMD cases and controls may have compromised the representativeness of the samples in this study. Although AMD can be diagnosed as early as 35 years, most AMD cases are diagnosed at age 60 years and older [2]. In addition, since control participants younger than 60 years may develop AMD in their later life, the age cut-off may have introduced selection bias in this study, resulting in lower power to detect real associations. Third, even though one of the strengths of this study is that all patients with AMD were long-term residents (>20 years) at a high altitude (over 2,000 m), which is a unique population that has been under-represented in genetic studies of AMD, the lack of comparison to non-high-altitude patients in this study made it impossible to conclude that the risk alleles are associated with AMD in the high-altitude population alone.

### Table 4. Genotype distributions of NF-κB gene polymorphisms in patients with AMD and controls.

| SNP       | Allele | AMD group | Control group | $P$  | OR   | 95% CI    |
|-----------|--------|-----------|---------------|------|------|-----------|
| rs3774959 | A      | 45 (0.409) | 35 (0.307)    | 0.11 | 1.56 | [0.90, 2.71] |
|           | G      | 65 (0.591) | 79 (0.693)    |      |      |           |
| rs3774932 | A      | 61 (0.555) | 61 (0.535)    | 0.77 | 1.08 | [0.64, 1.83] |
|           | G      | 49 (0.445) | 53 (0.465)    |      |      |           |
| rs3774937 | C      | 37 (0.336) | 32 (0.281)    | 0.37 | 1.30 | [0.74, 2.29] |
|           | T      | 73 (0.664) | 82 (0.719)    |      |      |           |
| rs230526  | A      | 49 (0.445) | 43 (0.377)    | 0.30 | 1.33 | [0.78, 2.26] |
|           | G      | 61 (0.555) | 71 (0.623)    |      |      |           |

AMD, age-related macular degeneration; CI, confidence interval; OR, odds ratio; SNP, single nucleotide polymorphism.

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In conclusion, the results of this study suggest that the AA genotype at rs3774932 and the CC genotype at rs3774937 in the NF-\(k\)B gene may be risk factors for the development of AMD. This is the first study that indicates an association between these two NF-\(k\)B variants and the risk of AMD. Whether this association exists in non-Chinese populations is worth further study.

**Supporting information**

S1 File. (PDF)

**Author Contributions**

Data curation: Yan Xin.

Formal analysis: Yan Xin.

Funding acquisition: Li Ling, Guan Ruijuan.

Methodology: Guan Ruijuan.

Software: Yan Xin.

Supervision: Kang Zefeng, Li Ling, Guan Ruijuan.

Writing – original draft: Yan Xin.

Writing – review & editing: Yan Xin.

**References**

1. Fine SL, Berger JW, Maguire MG, Ho AC. Age-related macular degeneration. N Engl J Med. 2000; 342: 483–492. https://doi.org/10.1056/NEJM200002171734207 PMID: 10675430.

2. Song P, Du Y, Chan KY, Theodoratou E, Rudan I. The national and subnational prevalence and burden of age-related macular degeneration in China. J Glob Health. 2017; 7(2): 020703. https://doi.org/10.7189/jogh.07.020703 PMID: 29302323.

3. Sivaprasad S, Bailey TA, Chong VNH. Bruch’s membrane and the vascular intima: Is there a common basis for age-related changes and disease? Clin Exp Ophthalmol. 2005; 33(5): 518–523. https://doi.org/10.1111/j.1442-9071.2005.01074.x PMID: 16181282.

4. Nashine S, Kenney MC. Effects of mitochondrial-derived peptides (MDPs) on mitochondrial and cellular health in AMD. Cells. 2020; 9(5): 1102. https://doi.org/10.3390/cells9051102 PMID: 32365540.

5. Datta S, Cano M, Ebrahimi K, Wang L, Handa JT. The impact of oxidative stress and inflammation on RPE degeneration in non-neovascular AMD. Prog Retin Eye Res. 2017; 60: 201–218. https://doi.org/10.1016/j.preteyeres.2017.03.002 PMID: 28336424.

6. Wang S, Wang X, Cheng Y, Ouyang W, Sang X, Liu J, et al. Autophagy dysfunction, cellular senescence, and abnormal immune-inflammatory responses in AMD: From mechanisms to therapeutic potential. Oxid Med Cell Longev. 2019; 2019: 3632169. https://doi.org/10.1155/2019/3632169 PMID: 31249643.

7. Chang C-C, Huang T-Y, Chen H-Y, Huang T-C, Lin L-C, Chang Y-J, et al. Protective effect of melatonin against oxidative stress-induced apoptosis and enhanced autophagy in human retinal pigment epithelium cells. Oxid Med Cell Longev. 2018; 2018: 9015765. https://doi.org/10.1155/2018/9015765 PMID: 30174783.

8. Klaver CCW, Wolfs RCW, Assink JJM, van Duijn CM, Hofman A, de Jong PTVM. Genetic risk of age-related maculopathy: Population-based familial aggregation study. Arch Ophthalmol. 1998; 116(12): 1646–1651. https://doi.org/10.1001/archophthalmol.116.12.1646 PMID: 9869796.

9. DeAngelis MM, Owen LA, Morrison MA, Morgan DJ, Li M, Shakoor A, et al. Genetics of age-related macular degeneration (AMD). Hum Mol Genet. 2017; 26(R1): R45–R50. https://doi.org/10.1093/hmg/ddx226 PMID: 28854576.
10. Fritsche LG, Igl W, Bailey JNC, Grassmann F, Sengupta S, Bragg-Gresham JL, et al. A large genome-wide association study of age-related macular degeneration highlights contributions of rare and common variants. Nat Genet. 2016; 48(2): 134–143. https://doi.org/10.1038/ng.3448 PMID: 26691988.

11. Cancer Genome Atlas Research Network. Comprehensive genomic characterization defines human glioblastoma genes and core pathways. Nature. 2008; 455(7216): 1061–1068. https://doi.org/10.1038/ nature07385 PMID: 18772890. Erratum in: Nature. 2013;494(7438): 506.

12. Hayden MS, Ghosh S. NF-κB, the first quarter-century: Remarkable progress and outstanding questions. Genes Dev. 2012; 26(3): 203–234. https://doi.org/10.1101/gad.183434.111 PMID: 22302935.

13. Véquaud E, Sévénon C, Louisouan D, Engelhart L, Campone M, Juin P, et al. YM155 potently triggers cell death in breast cancer cells through an autophagy-NF-κB network. Oncotarget. 2015; 6(15): 13476–13486. https://doi.org/10.18632/oncotarget.3638 PMID: 25974963.

14. Grüssel T, Busch W. Experimentelle untersuchungen zur wirkung der peressigsäure auf das endometrium des rindes [Experimental studies of the effect of peracetic acid on the endometrium of cattle]. Tierarztl Prax. 1997; 25(1): 28–34. PMID: 9157627.

15. Tang X, Sun L, Wang G, Chen B, Luo F. RUNX1: A regulator of NF-κB signaling in pulmonary diseases. Curr Protein Pept Sci. 2019; 19(2): 172–178. https://doi.org/10.2174/1389203718666171009111835 PMID: 28990531.

16. De Simone V, Franzé E, Ronchetti G, Colantoni A, Fantini MC, Di Fusco D, et al. Th17-type cytokines, IL-6 and TNF-α synergistically activate STAT3 and NF-κB to promote colorectal cancer cell growth. Oncogene. 2015; 34(27): 3493–3503. https://doi.org/10.1038/onc.2014.286 PMID: 25174402.

17. Dolcet X, Llobet D, Pallares J, Matias-Guiu X. NF-κB in development and progression of human cancer. Virchows Arch. 2005; 446(5): 475–482. https://doi.org/10.1007/s00428-005-1264-9 PMID: 15856292.

18. Khongthong P, Roseweir AK, Edwards J. The NF-KB pathway and endocrine therapy resistance in breast cancer. Endocr Relat Cancer. 2019; 26(6): R369–R380. https://doi.org/10.1530/erc-19-0087 PMID: 32013374.

19. Wei Q, Tu Y, Zuo L, Zhao J, Chang Z, Zou Y, et al. MiR-345-3p attenuates apoptosis and inflammation caused by oxidized low-density lipoprotein by targeting TRAF6 via TAK1/p38/NF-κB signaling in endothelial cells. Life Sci. 2020; 241: 117142. https://doi.org/10.1016/j.lfs.2019.117142 PMID: 31825793.

20. Chen C, Luo F, Liu X, Lu L, Xu H, Yang Q, et al. NF-κB-regulated exosomal miR-155 promotes the inflammation associated with arsenite carcinogenesis. Cancer Lett. 2017; 388: 21–33. https://doi.org/10.1016/j.canlet.2016.11.027 PMID: 27913196.

21. Mitchell JP, Carmody RJ. NF-κB and the transcriptional control of inflammation. Int Rev Cell Mol Biol. 2018; 335: 41–84. https://doi.org/10.1016/bs ircmb.2017.07.007 PMID: 29305014.

22. Peng X, Wang Y, Li H, Fan J, Shen J, Yu X, et al. ATG5-mediated autophagy suppresses NF-κB signaling to limit epithelial inflammatory response to kidney injury. Cell Death Dis. 2019; 10(4): 253. https://doi.org/10.1038/s41419-018-01483-7 PMID: 30874544.

23. Yi W, Wen Y, Tan F, Liu X, Lan H, Ye H, et al. Impact of NF-κB pathway on the apoptosis-inflammation-autophagy crosstalk in human degenerative nucleus pulposus cells. Aging. 2019; 11(17): 7294–7306. https://doi.org/10.18632/aging.102266 PMID: 31518335.

24. Han X, Gharahkhani P, Mitchell P, Liew G, Hewitt AW, MacGregor S. Genome-wide meta-analysis identifies novel loci associated with age-related macular degeneration. J Hum Genet. 2020; 65(8): 657–665. https://doi.org/10.1038/s10038-020-0750-x PMID: 32277175.

25. American Academy of Ophthalmology Retina/Vitreous Panel. Preferred Practice Pattern Guidelines: Age-Related Macular Degeneration. San Francisco, CA: American Academy of Ophthalmology; 2014. Available from: www.aao.org/ppp.

26. Ferris FL, Wilkinson CP, Bird A, Chakravarthy U, Chew E, Csaky K, et al. Clinical classification of age-related macular degeneration. Ophthalmology. 2013; 120(4): 844–851. https://doi.org/10.1016/j. ophtha.2012.10.036 PMID: 23332590.

27. Purcell S, Cherny SS, Sham PC. Genetic Power Calculator: design of linkage and association genetic mapping studies of complex traits. Bioinformatics. 2003; 19(1): 149–150. https://doi.org/10.1093/ bioinformatics/19.1.149 PMID: 12499305.

28. Roubeix C, Sahel J-A, Guillonneau X, Delarasse C, Sennlaub F. Sur les origines inflammatoires de la DMLA [On the inflammatory origins of AMD]. Med Sci. 2020; 36(1): 886–892. https://doi.org/10.1051/ medsci/20200159 PMID: 33026331.

29. Karczewski KJ, Francioli LC, Tiao G, Cummings BB, Alfoöd J, Wang Q, et al. The mutational constraint spectrum quantified from variation in 141,456 humans. Nature. 2020; 581(7809): 434–443. https://doi.org/10.1038/s41586-020-2308-7 PMID: 32461654
30. Ward LD, Kellis M. HaploReg: a resource for exploring chromatin states, conservation, and regulatory motif alterations within sets of genetically linked variants. Nucleic Acids Res. 2012; 40(Database issue): D930–D934. https://doi.org/10.1093/nar/gkr917 PMID: 22064851.