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Data Article

Data supporting the investigation of interaction of biologically relevant proteins with ZnO nanomaterials: A confounding factor for in vitro toxicity endpoints

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A B S T R A C T

Test materials, like manufactured nanomaterials (MN), may interact with serum proteins, interleukins (IL) and lactate dehydrogenase (LDH) and cause measurement artefacts as a result of e.g., physical adsorption and electrostatic forces, and/or interaction with dissolved species or conditional chemical changes during testing. In this article, data are given on the zeta-potentials of two manufactured ZnO nanomaterials (NM-110 and NM-111) dispersed in 0.05% w/v Bovine Serum Albumin (BSA) water batch dispersions and in Ham’s F12 nutrient mixture added Fetal Bovine Serum (FBS), penicillin, and streptomycin and particle free mediums (cHam’s F12). Data on the Zeta-potential and the iso-electrical point of lactate hydrogenase in pure Ham’s F12 nutrient mixture is also provided. The percentage of added IL-6, IL-8 and LDH remaining after 24-h incubation in cHam’s F12 are given as function of MN concentrations. Finally data from thermodynamic chemical reaction modeling of changes in pH and Zn-speciation during dissolution of ZnO or dissolved ZnCl2 additions to Ham’s F12 using Geochemist Workbench® are given. For further information, data interpretation and discussion please refer to the DOI of original article: https://doi.org/10.1016/j.tiv.2018.12.016.

Abbreviations: BSA, bovine serum albumin; DLS, dynamic light scattering; DMEM, Dulbecco’s modified egle’s medium; FBS, fetal bovine serum; ILs, interleukins; LDH, lactate dehydrogenase; MN, manufactured nanomaterials; OD, optical density.

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research article “Interaction of biologically relevant proteins with ZnO nanomaterials: a confounding factor for in vitro toxicity endpoints” (E. Da Silva et al. 2019).

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1. Data

Table 1 lists the zeta-potentials of NM-110 and NM-111 (uncoated and coated ZnO manufactured nanomaterials, respectively [1,2]) in cHAM's F12 test medium over time (0, 30 and 60 minutes) at 10,
Table 1
Zeta potential of NM-110 and NM-111 in cHAMs F12 as function of MN concentration and time as well as in the cHAMs F12 and the 0.05% BSA-water batch dispersion with and without NM-110 and NM-111.

| Test material          | MN concentration (µg/mL) | Time (min) | Zeta potential (mV, mean ± sd) |
|------------------------|--------------------------|------------|-------------------------------|
| NM-110 in cHAMs F12    | 10                       | 0          | 0.08 ± 1.18                   |
|                        | 10                       | 30         | 0.06 ± 1.72                   |
|                        | 10                       | 60         | −0.41 ± 0.90                  |
|                        | 320                      | 0          | −0.47 ± 0.87                  |
|                        | 320                      | 30         | −0.79 ± 0.65                  |
|                        | 320                      | 60         | −0.10 ± 1.30                  |
|                        | 640                      | 0          | 0.31 ± 0.93                   |
|                        | 640                      | 30         | 0.99 ± 1.01                   |
|                        | 640                      | 60         | 0.22 ± 1.03                   |
| NM-111 in cHAMs F12    | 10                       | 0          | −11.20 ± 1.03                 |
|                        | 10                       | 30         | −12.82 ± 0.47                 |
|                        | 10                       | 60         | −13.10 ± 0.80                 |
|                        | 320                      | 0          | −12.92 ± 0.74                 |
|                        | 320                      | 30         | −13.62 ± 0.35                 |
|                        | 320                      | 60         | −14.15 ± 0.93                 |
|                        | 640                      | 0          | −13.13 ± 0.71                 |
|                        | 640                      | 30         | −13.77 ± 0.98                 |
|                        | 640                      | 60         | −14.32 ± 1.04                 |
| cHam’s F12             | —                        | 0          | −10.44 ± 2.00                 |
|                        | —                        | 30         | −12.43 ± 1.01                 |
|                        | —                        | 60         | −12.00 ± 1.10                 |
| NM-110 in 0.05% BSA    | 2560                     | 0          | −0.02 ± 0.39                  |
|                        | 2560                     | 30         | −0.17 ± 0.32                  |
|                        | 2560                     | 60         | −0.16 ± 0.34                  |
| NM-111 in 0.05% BSA    | 2560                     | 0          | 0.07 ± 0.38                   |
|                        | 2560                     | 30         | −0.04 ± 0.29                  |
|                        | 2560                     | 60         | 0.01 ± 0.27                   |
| 0.05% BSA              | —                        | 0          | −20.70 ± 0.14                 |
|                        | —                        | 30         | −23.25 ± 1.77                 |
|                        | —                        | 60         | −24.40 ± 1.13                 |

320 and 640 µg ZnO/mL and the 0.05% w/v BSA water batch dispersions with 2560 µg ZnO/mL. The zeta potentials of the particle free mediums are shown for reference. The pH values in the dispersions were close to neutral (NM-110: pH = 7.0 ± 0.3; NM-111: pH = 6.8 ± 0.3) during measurements.

Table 2 lists the percentage of the initially added levels of IL-6 (500 pg/mL), IL-8 (4000 pg/mL) and LDH (100 ng/mL) after incubation with NM-110 and NM-111 at eight different doses (0–640 µg/mL) in cHam’s F12: The LDH data (OD_{LDH}) are given as the optical density for LDH expressed as a percentage of optical density of the medium reference OD_{ref} without MN. IL-6 and IL-8 concentrations are given as the percent of the initially dosed concentrations.

Fig. 1 shows the zeta-potential data as function of pH and the iso-electrical point of LDH at pH = 4.67 (vertical line, triplicates) in pure Ham’s F12 nutrient mixture. A concentration of 10000 ng LDH/mL was used to allow good zeta-potential measurements. 1 M HCl and 1 M NaOH were used for pH-titrations.

Fig. 2 shows the pH-variation as function of the amount of ZnO (Fig. 2A and B) and ZnCl₂ (Fig. 2C and D) reacted and as function of Zn-concentrations in Ham’s F12 nutrient mixture calculated using the React program in the Geochemist Workbench® [3]. The pH may increase to approximately 9.5 during dissolution of only 10 µg ZnO/mL (Fig. 2A and B). Opposite, a slight reduction in pH from pH 7.4 to pH 6.7 occurs with increased amounts of dissolved ZnCl₂ (Fig. 2C and D).

Fig. 3 shows the data from calculations of the Zn speciation in Ham’s F12 nutrient mixture during dissolution of ZnO and ZnCl₂. More than 99.3% (by weight) of the dissolved Zn will occur as Zn²⁺ during dissolution of ZnO (Fig. 3A). Slightly less (>95.8% by weight) will be Zn²⁺ during dissolution of ZnCl₂ (Fig. 3B).
2. Experimental design, materials and methods

Two ZnO MN test materials; NM-110 (uncoated ZnO) and NM-111 (tri-ethoxy-silane-coated ZnO) and ZnCl₂ originating from the from the Sponsorship Programme for the Testing of Manufactured Nanomaterials organized by the OECD Working Party on Manufactured Nanomaterials [4] were used here. The main research article [1] is referred to for compiled data on the ZnO MN. ZnCl₂ was purchased from Merck (EMSURE® ACS, Reag. Ph Eur, catalogue no. 108816).

### Table 2
LDH, IL-6 and IL-8 levels after incubation with NM-110 and NM-111 for 24 h at 37°C.

| ZnO MN  | MN concentration (μg/mL) mean ± sd | LDH (% OD_ref) | IL-6 (% initial concentration) | IL-8 (% initial concentration) |
|--------|-----------------------------------|----------------|-------------------------------|-------------------------------|
| NM-110 | 0                                 | 100 ± 0.8      | 100 ± 2.4                     | 100 ± 1.8                     |
|        | 10                                | 40.5 ± 6.9a    | 100.2 ± 2.8                   | 96.8 ± 2.2                    |
|        | 20                                | 34.2 ± 5.6a    | 98.9 ± 3.7                    | 97.9 ± 2.8                    |
|        | 40                                | 31.5 ± 3.9a    | 100.2 ± 6.3                   | 97.0 ± 4.2                    |
|        | 80                                | 30.1 ± 3.4a    | 100.6 ± 5.1                   | 98.6 ± 6.8                    |
|        | 160                               | 28.3 ± 2.7a    | 97.8 ± 5.2                    | 95.4 ± 4.0                    |
|        | 320                               | 27.6 ± 2.6a    | 98.5 ± 4.0                    | 96.7 ± 3.6                    |
|        | 640                               | 26.5 ± 1.5a    | 96.8 ± 4.4                    | 95.4 ± 6.8                    |
| NM-111 | 0                                 | 100 ± 0.5      | 100 ± 2.1                     | 100 ± 2.1                     |
|        | 10                                | 47.2 ± 9.7a    | 98.9 ± 4.3                    | 96.8 ± 5.5                    |
|        | 20                                | 35.9 ± 4.3a    | 99.2 ± 9.1                    | 95.8 ± 7.4                    |
|        | 40                                | 32.3 ± 3.0a    | 99.1 ± 6.5                    | 100.6 ± 9.8                   |
|        | 80                                | 30.3 ± 3.4a    | 100.3 ± 6.0                   | 99.2 ± 12.0                   |
|        | 160                               | 28.0 ± 3.3a    | 96.1 ± 9.3                    | 98.9 ± 13.4                   |
|        | 320                               | 26.4 ± 3.0a    | 96.5 ± 8.2                    | 101.7 ± 9.8                   |
|        | 640                               | 25.4 ± 3.4a    | 97.2 ± 6.6                    | 107.7 ± 15.9                  |

sd = standard deviation, obtained from three independent experiments with two replicates each.

a Denotes a statistically significant difference with the baseline (ANOVA, p-value <0.001).

![Fig. 1. Zeta potential of LDH (10000 ng/mL) in cHam's F12 versus pH. The vertical line represents the isoelectric point where the surface charge is neutral.](image-url)
Fig. 2. Change in pH in Ham’s F12 nutrient mixture at 37 °C and CO2 balanced atmosphere using an initial pH 7.4 and an oxygen fugacity of 0.2 at different conditions. A. pH-change during simulated reaction with up to 320 mg/mL ZnO. B. pH-change as a function of measured Zn concentration due to dissolution of ZnO. C. pH-change during simulated reaction with up to 640 mg/mL ZnCl2. D. pH-change as a function of the resulting Zn concentration (<307 mg/mL) due to dosing of up to 640 mg/mL ZnCl2. For all plots, calculations were made using the React and Gtplot Apps in Geochemist’s Workbench® version 11 [3].

Fig. 3. Change in Zn-species concentrations as function of dissolved Zn-concentrations in Ham’s F12 nutrient mixture at 37 °C and CO2 balanced atmosphere using an initial pH 7.4 and an oxygen fugacity of 0.2. A. Zn-species concentrations during simulated dissolution of up to 320 μg/mL ZnO. B. Zn-species concentrations during simulated dissolution with up to 640 μg/mL ZnCl2. D. For all plots, calculations were made using the React and Gtplot Apps in Geochemist’s Workbench® version 11 [3].
Batch dispersions of the test materials were produced by dispersing 2.56 mg/mL in sterile filtered 0.05% (v/v) BSA water (Nanopure Diamond UV) by probe-sonication as described in the NANO-GENOTOX batch dispersion protocol [5]. Sonication was performed using a Branson Sonifier S-450D (Branson Ultrasonics Corp., Danbury, CT, USA) operated for 16 min at 400 W and 10% amplitude using a 13 mm disruptor horn. For further details please see Jensen et al. [5].

The complete test medium (cHam’s F12) consisted of Ham’s F12 nutrient mixture (ThermoFisher Scientific, Hvidovre, Denmark) with 1% v/v penicillin/streptomycin (10000 U/mL and 10 mg/mL respectively; In Vitro A/S, Fredensborg, Denmark) and 10% v/v of inactivated Fetal Bovine Serum (USA grade; In Vitro A/S, Fredensborg, Denmark). For interaction studies cHam’s F12 was added 500 pg/mL IL-6 standard (National Institute for Biological Standards and Controls, Hertfordshire, UK), 4000 pg IL-8 standard (National Institute for Biological Standards and Controls, Hertfordshire, UK; final concentration), and 100 ng/mL rabbit muscle LDH (Sigma-Aldrich, Brøndby, Denmark). Please see Da Silva et al. [1] for further details.

Zeta potential measurements were completed using a Zetasizer Nano ZS (Malvern Instruments Ltd., Malvern, United Kingdom) controlled by Zetasizer software v. 7.11 and 7.12. For determination of the iso-electrical point, the pH was adjusted using 1 M HCl, (grade AVS TITRINORM Reagent Ph.Eur. chapter 4.2.2, USP, NF, VWR International S.A.S., 42250 Briane, France) and 1M NaOH, (grade AVS TITRINORM Reagent Ph.Eur. chapter 4.2.2, USP, NF, VWR International S.A.S., 42250 Briane, France) dosed using the MPT-2 Titrator (Malvern Instruments Ltd., Malvern, United Kingdom). Please see Da Silva et al. [1] for further details.

IL-6, IL-8 and LDH levels in cHam’s F12 with and without test materials were determined after 24 h incubation in darkness at 37 °C and a 5% CO2 atmosphere with a 95% relative humidity in a cell incubator (CelCulture® CO2 Incubator, ESCO Medical, Egaa, Denmark) maintaining a temperature of 37 °C and a 5% CO2 atmosphere with a 95% relative humidity in the dark without shaking. Suspensions were collected, centrifuged and prepared for spectrophotometric analysis of interleukine (BD Pharmingen kit, Cat. No. 555220 and 555244 respectively; BD Biosciences, Lyngby, Denmark) and LDH (Roche Diagnostics GmbH, Roche Applied Science, Mannheim, Germany) using an ELISA reader as described in Da Silva et al. [1].

Thermodynamic chemical reaction modeling of dissolved Zn concentration associated effect on pH and Zn-ion speciation after addition of ZnO and ZnCl2 was completed using the Geochemist Workbench® v. 11.0 [3]. Please see Da Silva et al. [1] for further details on medium composition and conditions defined for generating the data.

Statistical analysis, tables and graphs were made using R (version 3.4.3) and Gt-plot in Geochemist Workbench® v. 11.0. Statistical significance was set at p-value < 0.05. Each treatment was repeated in duplicate and each experiment replicated three times.

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Transparency document

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