Intratumor heterogeneity of k-ras and p53 mutations among human colorectal adenomas containing early cancer

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The molecular pathways and the timing of genetic events during human colorectal carcinogenesis are still not fully understood. We have addressed the intratumor heterogeneity of the mutational status of the k-ras oncogene and of the p53 oncosuppressor gene during the adenoma–carcinoma sequence by investigating 26 human colorectal adenomas containing early cancer. An intratumor comparative analysis was obtained among the adenomatous and carcinomatous component pairs. Additionally, we have analyzed 17 adenomas having cancer in the near vicinity.

The adenomatous components of the adenomas containing early cancer and the adenomas having cancer in the near vicinity had comparable frequencies for k-ras mutations (28 and 47%) but different for p53 mutations (52 and 7%, p-value = 0.01). Interestingly, the adenomatous and carcinomatous components of the adenomas containing early cancer were rarely heterogeneous for the k-ras mutational status (only in 13% of the cases) but were characterized by heterogeneity of the p53 status in 59% of the cases (p-value < 0.01). In addition, the mutations of p53 for the adenomatous components of the adenomas containing early cancer were statistically significantly associated with severe dysplasia (p-value = 0.01).

Intratumor homogeneity of k-ras status during the human colorectal adenoma–carcinoma sequence suggests that the role of k-ras is more related to tumor initiation than to tumor progression. On the contrary, intratumor heterogeneity of p53 mutations indicates that the type of the p53 mutations may also be relevant for selection and expansion of new subclones leading to tumor progression.

Keywords: Oncogenes, tumor suppressor genes, colorectal tumor progression, molecular biology

1. Introduction

The vast majority of human sporadic colorectal cancers are thought to derive from adenomas, through histologically well-defined steps characterized by increasing degrees of dysplasia. From the genetic point of view, colorectal cancer development and progression is considered to be due to the accumulation of several genetic events. Among these, inactivation of the APC oncosuppressor gene, mutations of the k-ras protooncogene, mutations and deletions of the p53 and DCC oncosuppressor genes, and still loss or inactivation of other oncosuppressor genes, as indicated by studies addressing loss of heterozygosity and cytogenetic alterations [3,4,6,9,20,26,27,34,38].

The incidence of k-ras mutations in human colon aberrant crypt foci, which are considered to be early precursors of adenomas, has been reported to be up to 85% [31,32,37,40]. In contrast, k-ras mutations in sporadic adenomas and adenocarcinomas were reported to be only at 30–50% incidence, with a trend of higher incidence of mutations among the adenomas [9,38]. Thus, a k-ras mutation present in either aberrant crypt foci or in adenomas would not seem to provide a definitive growth advantage to colonic epithelial cells. This hypothesis for a role of k-ras in initiation more than in progression was further tested in the present...
study by comparing the adenomatous and carcinomato-
tous components of human colorectal adenomas con-
taining early cancer and adenomas having cancer in the
near vicinity.

Ras proteins in structurally altered forms, due to
the gene mutations, are thought to lead to mitogenic
signals without ongoing stimulation by their normal
upstream regulators, but their precise role in human
colorectal cancer genesis and progression is still not
fully understood [3,19]. Recently, new functions of the
Ras proteins were suggested with the discovery of a
plethora of new effector pathways and different cel-

The p53 oncosuppressor gene was found to be rarely
mutated among human colorectal adenomas with mild
or moderate dysplasia, though mutations were fre-
quent in adenomas with severe dysplasia and adeno-
carcinomas [9]. The mutations of p53, disseminated
along the five most conserved exons (exons 5a, 5b,
6, 7, and 8), were reported to interfere with its con-
trol functions. The p53 gene product represents an im-
portant cell cycle checkpoint associated to cell block
in the G1 phase to drive cells either to DNA repair
or apoptosis. These very important functions of p53
at the crossroads of proliferation, apoptosis and aneu-
plody have been extensively investigated and are the
subject of several excellent reviews [22,24,25]. Thus,
when p53 mutations occur in human colorectal ade-
nomas, it is likely that other genetic alterations may
accumulate and lead to definitive malignant progres-
sion. Also, the degree of cell heterogeneity is expected
to be high at these stages, due to selection and ex-
pansion of new subclones with novel genetic changes.
New studies are needed to assess these hypotheses fur-
ther. In the present study, we aimed to investigate the
intratumor heterogeneity of k-ras mutations among the
adenomatous and carcinomatous components of the
human sporadic colorectal adenomas containing
early cancer. These relatively rare lesions should
provide a unique model to contribute to a better under-
standing of the colorectal adenoma–canceroma transi-
tion.

2. Materials and methods

2.1. Study population and histological analysis

The study was carried out on 11 females and 16
males, in the age-range 45–86 (median age 66.5 years),
who did not have a history of familial adenomatous
polyposis. We have considered two categories of col-
orectal adenomas, i.e., 26 adenomas containing early
cancer (A/C) and 17 adenomas having cancer in the
near vicinity (A+C). Table 1 reports the list of the cases
with site, size, type, and dysplasia. The site was in the
right bowel (including caecum, ascending colon, hep-
atic flexure and transverse colon) for 21 adenomas and
in the left bowel (including splenic flexure, descend-
ing colon, sigmoid colon and rectum) for 22 cases. The
size of the adenomas ranged from 3.5 to 80 mm (11
were smaller than 10 mm, 16 were 10–20 mm, and
16 were greater than 20 mm). Considering the adeno-
matous components of the adenomas containing early
cancer, the histological type was tubular for 17 cases
and tubulovillous or villous for 25. In one case this in-
formation was missing. Dysplasia was graded accord-
ing to Konishi and Morson [23] and mild and moder-
ate dysplasia cases were taken together as one group
\( n = 28 \) versus severe dysplasia \( n = 15 \).

Using a hematoxylin–eosin stained section as his-
totopographic reference, samples were taken by hand
with a scalpel blade from selected prevalent areas with
homogeneous grades of dysplasia. Areas with preva-
ent connective tissue, inflammatory cells, and normal
epithelial cells were discarded as much as possible.

2.2. k-ras mutation spectrum analysis

Extracted DNA from peripheral blood lymphocytes
from healthy donors was used as wild type k-ras codon
12 GGT-gly and codon 13 GGC-gly controls. Addi-
tionally, extracted DNA from 6 different colon cancer
cell lines was used as control for known k-ras muta-
tions as previously described [15]. Extraction of high
molecular weight genomic DNA was performed ac-
cording to standard methods [35] and amplified by
PCR as previously described [14]. Oligonucleotide 20-
mers panel (TIB MOLBIOL, Advanced Biotechnology
Center, Genoa, Italy) including k-ras codon 12 and 13
wild type sequences, all possible mutations of codon
12, and the AGC and GAC mutations of codon 13 were
5'-end labeled by phosphorylation with \([\gamma^{32}\text{P}]\text{ATP}
\) (Amersham International, Buckinghamshire, UK), ac-
 accordance with the standard method [35]. Dot blot and se-
quence specific oligonucleotide hybridization was per-
formed as usual [14].
Table 1
Site, size, type, and dysplasia of 26 adenomas containing early cancer (A/C) and 17 adenomas having cancer in the near vicinity (A+C)

| N. | Category | Site | Size (mm) | Type | Dysplasia |
|----|----------|------|-----------|------|-----------|
| 1  | A/C      | L    | 11        | T    | S         |
| 2  | A/C      | L    | 30        | TV   | S         |
| 3  | A/C      | L    | 32        | T    | MM        |
| 4  | A/C      | L    | 29        | TV   | S         |
| 5  | A/C      | L    | 20        | TV   | MM        |
| 6  | A/C      | R    | 75        | V    | MM        |
| 7  | A/C      | R    | 26        | TV   | MM        |
| 8  | A/C      | R    | 60        | TV   | MM        |
| 9  | A/C      | L    | 20        | TV   | S         |
| 10 | A/C      | R    | 80        | TV   | S         |
| 11 | A/C      | L    | 20        | TV   | MM        |
| 12 | A/C      | R    | 18        | T    | S         |
| 13 | A/C      | R    | 40        | TV   | MM        |
| 14 | A/C      | R    | 10        | T    | S         |
| 15 | A/C      | L    | 15        | T    | S         |
| 16 | A/C      | R    | 50        | TV   | MM        |
| 17 | A/C      | R    | 10        | T    | S         |
| 18 | A/C      | L    | 10        | TV   | S         |
| 19 | A/C      | L    | 80        | –    | MM        |
| 20 | A/C      | R    | 20        | TV   | MM        |
| 21 | A/C      | R    | 35        | TV   | S         |
| 22 | A/C      | L    | 20        | TV   | S         |
| 23 | A/C      | L    | 55        | TV   | MM        |
| 24 | A/C      | L    | 3.5       | T    | MM        |
| 25 | A/C      | L    | 75        | V    | MM        |

| 1  | A+C      | L    | 5         | TV   | MM        |
| 2  | A+C      | L    | 15        | T    | MM        |
| 3  | A+C      | R    | 5         | T    | MM        |
| 4  | A+C      | R    | 29        | TV   | MM        |
| 5  | A+C      | R    | 30        | TV   | MM        |
| 6  | A+C      | R    | 24        | TV   | S         |
| 7  | A+C      | L    | 10        | T    | MM        |
| 8  | A+C      | L    | 9         | TV   | MM        |
| 9  | A+C      | L    | 11        | T    | MM        |
| 10 | A+C      | L    | 11        | T    | MM        |
| 11 | A+C      | R    | 10        | V    | MM        |
| 12 | A+C      | R    | 15        | T    | S         |
| 13 | A+C      | R    | 16        | TV   | MM        |
| 14 | A+C      | L    | 20        | T    | MM        |
| 15 | A+C      | L    | 18        | T    | MM        |
| 16 | A+C      | R    | 9         | T    | MM        |
| 17 | A+C      | R    | 8         | T    | MM        |

Right colon (R) includes caecum, ascending colon, hepatic flexure, and transverse colon; Left colon (L) includes splenic flexure, descending and sigmoid colon, and rectum. T = tubular type; TV = tubulovillous type; V = villous type. MM = mild and moderate.

2.3. p53 mutation analysis

Starting from about 400 ng of extracted DNA (see above), the genomic region encompassing exons 5–8 was first amplified with GC-clamped ampliprimers as previously described [7]. The PCR products obtained were analyzed by Denaturing Gradient Gel Electrophoresis (DGGE) as previously detailed [7]. After electrophoresis, the gels were stained for 20 min in 0.5 μg/ml ethidium bromide, visualized using an ultraviolet transilluminator and photographed using Polaroid type 57 films.

2.4. Statistical analysis

Contingency tables (Chi-square and Fisher exact test) were used to evaluate the associations among k-ras and p53 status with site, size, type and dysplasia of the adenomas. Statistical significance was set at p-value = 0.05 [10].

3. Results

The material for the study was composed of 26 human colorectal adenomas containing early cancer and 17 adenomas having cancer in the near vicinity. Table 1 reports the list of the cases together with site of origin, size, type, and degree of dysplasia.

The adenomatous components of the adenomas containing early cancer showed a statistically significantly higher frequency of severe dysplasia than the adenomas with cancer in the near vicinity (50% versus 12%; p-value = 0.02; Table 2). The adenomatous components of the adenomas with early cancer also showed a slightly higher proportion of cases with a villous pattern (69% versus 41%; p-value = 0.06) and were more often with size greater than 20 mm (50% versus 17%; p-value = 0.06) than the adenomas with cancer in the near vicinity, but the differences were not statistically significant. The sites of origin of the adenomas containing early cancer and the adenomas having cancer in the near vicinity were, instead, comparable.

Analyses of k-ras and p53 mutations for both categories of adenomas were performed after PCR amplification of the extracted genomic DNA at exon 1 codons 12–13 for k-ras and for exons 5–8 for p53, respectively. Figures 1 and 2 show examples of the k-ras and p53 mutations obtained, respectively, by sequence specific oligonucleotide hybridization and DGGE. The cases with k-ras mutations are for the 6 possible mutations
Table 2
Comparison of site, size, type, and dysplasia grade for 26 colorectal adenomas containing early cancer (A/C) and 17 adenomas having cancer in the near vicinity (A+C). Type and dysplasia of A/C lesions refer to their adenomatous components (A).

| Site  | Size (mm) | Type | Dysplasia |
|-------|-----------|------|-----------|
|       | ≤10       | >10 and ≤20 | >20     | T  | TV+V | MM | S  |
| A/C   | 12 | 14 (54%) | 4 | 9 | 13 (50%) | 7 | 18 (69%) | 13 | 13 (50%) |
| A+C   | 9  | 8 (47%)  | 7 | 7 | 3 (17%)  | 10| 7 (41%)  | 15 | 2 (12%)  |

**p-values**
- \( p = 0.76 \)
- \( p = 0.06 \)
- \( p = 0.02 \)

Right colon (R) includes caecum, ascending colon, hepatic flexure and transverse colon; Left colon (L) includes splenic flexure, descending colon, sigmoid colon and rectum. T = tubular type; TV = tubulovillous type; V = villous type; MM = mild and moderate dysplasia; S = severe dysplasia.

in codon 12 (AGT-ser, TGT-cys, CGT-arg, GAT-asp, GCT-ala, GTT-val) and the 2 most common mutations in codon 13 (AGC-ser, GAC-asp) as shown in Fig. 1 (from 2nd top left rectangle to 9th bottom right rectangle). Identical k-ras mutations for the adenomatous and carcinomatous components of 4 adenomas containing early cancer (these cases are listed in Tables 1 and 3 under the cases N. 4, 14, 21 and 26) corresponding to the pairs of spots GAT (b5–b6), GTT (c1–c2), GTT (d5–d6), and GAC (a5–a6) may be observed (see also the legend to the figure for more details).

Mutations of p53 within exons 5 to 8 were detected by comparison of the electrophoresis migration patterns with respect to the genomic DNA extracted from negative and positive controls. The presence of a four-band pattern in the case of heterozygous mutation is due to the simultaneous presence of wild type and mutated alleles (as in the examples of Fig. 2 for lanes A, H, and I of exon 8) that generate two heteroduplex combinations (wild type/mutated) and two homoduplex combinations (wild type/wild type and mutated/mutated; see also the legend to the figure for more details).

Table 3 shows the k-ras and p53 mutation data obtained for the adenomatous and carcinomatous components of the adenomas containing early cancer. The adenomatous components had frequencies for k-ras and p53 mutations of 28% (7/25) and 52% (12/23), respectively. The corresponding frequencies for the carcinomatous components were 28% (7/25) for k-ras and 64% (16/25) for p53. One may also notice that 2 different mutations were present for k-ras in the carcinomatous component of adenoma N. 20 and for p53 in the carcinomatous components of adenomas N. 10 and 13.

Component pairs were obtained in 24 out of 26 cases for k-ras and in 22 cases for p53. While the adenomatous and carcinomatous components rarely presented differences for k-ras, heterogeneity of the p53 status was commonly detected. Overall, intratumor k-ras heterogeneity between the adenomatous and carcinomatous components was present only in the pair cases N. 7, 17 and 24. Table 4 reports in detail the heterogeneous cases for both k-ras and p53 genetic status.
Fig. 1. Examples of k-ras mutation spectrum analysis for the adenomatous (A) and carcinomatous (C) components of human colorectal adenomas containing early cancer (A/C, as listed in Tables 1 and 3). Wild type k-ras results in codons 12 and 13, which correspond in both cases to the aminoacid glycine, are shown in the top-left rectangle (wt 12–13 GGT-gly, GGC-gly). Peripheral blood lymphocytes from a healthy donor were used as a k-ras wild type control (spot f6). The pairs of spots (a1–a2), (a3–a4), (a5–a6), (b1–b2), and so on up to (e3–e4) correspond to the wild type cases listed in Tables 1 and 3 under N. 24, 25, 26, 22, 23, 5, 14, 2, 8, 12, 13, 21, 16 and 1. An additional wild type case (the A component of case N. 6) corresponds to spot e5. The light grey spots g1–g6 correspond to 6 cell lines (respectively, A549, SW837, NIH3T3, SKLU-1, SW480 and DLD1) which had specific k-ras mutations in at least one allele. The cases with mutations are shown in the other rectangles: the 6 possible mutations in codon 12 (AGT-ser, TGT-cys, CGT-arg, GAT-asp, GCT-ala, GTT-val) and the 2 most common mutations in codon 13 (AGC-ser, GAC-asp). Specific k-ras mutation controls were obtained using mutated cell lines: the A549 cell line with heterozygous AGT mutation (g1); the SW837 cell line with heterozygous TGT mutation (g2); the NIH3T3 cell line with transfected CGT human mutated k-ras (g3); the SKLU-1 cell line with heterozygous GAT mutation (g4); the SW480 cell line with homozygous GTT mutation (g5); the DLD-1 cell line with heterozygous GAC mutation (g6) [15]. Identical k-ras mutations for the adenomatous and carcinomatous components of 4 human colorectal adenomas containing early cancer (these cases are listed in Tables 1 and 3 under the cases N. 4, 14, 21 and 26) corresponding to the pairs of spots GAT (b5–b6), GTT (c1–c2), GTT (d5–d6), and GAC (a5–a6) may be observed.
Fig. 2. PCR-based DGGE analysis examples of exon 8 of the p53 gene. A positive control obtained from the p53 mutated cell line SW480 mixed with normal lymphocyte is shown in lane A. A negative control obtained from normal lymphocytes DNA [7] is shown in lane O. DGGE variants corresponding to the adenomatous and carcinomatous components of the colorectal adenoma containing early cancer listed in Tables 1 and 3 under N. 22 are shown in lanes H and I.

### Table 4
Heterogeneity for k-ras and p53 genetic status between adenomatous (A) and carcinomatous (C) component pairs for human colorectal adenomas containing early cancer (A/C)

| Homogeneous classes | Heterogeneous classes | Heterogeneous cases |
|---------------------|-----------------------|---------------------|
|                     | wt/wt | m/m | wt/m | m/wt | m/m' | 3/24 (13%) |
| k-ras status in A/C component pairs | 16 | 5 | 1 | 2 | 0 | 3/24 (13%) |
| p53 status in A/C component pairs | 5 | 4 | 5 | 3 | 5 | 13/22 (59%) |

wt = wild type; m, m' = mutations with m ≠ m'. Read wt/wt as wild type in both A and C component pairs. (See raw data in Table 3.)

### Table 5
Associations of k-ras and p53 mutational status with site, size, type and dysplasia among human colorectal adenomas containing early cancer

| Site | Size (mm) | Type | Dysplasia |
|------|-----------|------|-----------|
|      | ≤10 | >10 and ≤20 | >20 | T | TV+V | MM | S |
| k-ras |      |      |      |      |      |      |      |
| wild type | 9 | 9 | 4 | 7 | 6 | 7 | 11 | 7 | 11 |
| mutated | 3 | 4 | 0 | 2 | 5 | 0 | 7 | 5 | 2 |
| p-value | p = 0.13 | p = 0.2 | p = 0.2 | p = 0.2 | p = 0.7 | p = 0.97 | p = 0.07 | p = 0.012 |
| p53 |      |      |      |      |      |      |      |
| wild type | 6 | 5 | 2 | 4 | 5 | 4 | 7 | 8 | 3 |
| mutated | 7 | 2 | 4 | 6 | 3 | 9 | 2 | 10 | 10 |

Right colon (R) includes caecum, ascending colon, hepatic flexure and transverse colon; Left colon (L) includes splenic flexure, descending colon, sigmoid colon and rectum. T = tubular type; TV = tubulovillous type; V = villous type. MM = mild and moderate dysplasia; S = severe dysplasia.

One may see that there are three types of heterogeneity: the cases within the (wt/m) class with wild type status for the adenomatous component and mutation for the carcinomatous component; the cases within the opposite (m/wt) class; and the cases in which the adenomatous and carcinomatous components had different mutations (m/m’ class). In the case of k-ras, only 3 out of 24 cases were heterogeneous (13%), while heterogeneity was present for p53 in 13 out of 22 cases (59%) (p-value = 0.002).

Among the adenomas containing early cancer, we have also analyzed the possible associations of k-ras and p53 mutational status within the adenomatous component with adenoma site, size, type, and dysplasia (Table 5). We found that 83% of the p53 mutated cases had severe dysplasia, providing a statistically signifi-
Table 6
Analysis of k-ras and p53 mutations for 17 human colorectal adenomas having cancer in the near vicinity

| N. | Category | k-ras  | p53   |
|----|----------|--------|-------|
|    |          | A      | C     | A     | C     |
| 1  | A+C      | GAT    | GTT   | –     | Ex.7  |
| 2  | A+C      | wt     | GTT   | Ex.7  | Ex.5B |
| 3  | A+C      | TGT    | –     | wt    | –     |
| 4  | A+C      | TGT    | wt    | wt    | wt    |
| 5  | A+C      | TGT    | wt*   | wt    | wt*   |
| 6  | A+C      | TGT    | wt    | wt    | wt    |
| 7  | A+C      | wt     | wt*   | wt    | wt*   |
| 8  | A+C      | TGT    | wt*   | wt    | wt*   |
| 9  | A+C      | wt     | wt    | wt    | wt    |
| 10 | A+C      | GAC    | GTT   | –     | wt    |
| 11 | A+C      | GAT    | AGT   | –     | wt    |
| 12 | A+C      | wt     | wt    | wt    | wt    |
| 13 | A+C      | GAC    | GAT/GAC | wt   | Ex.6  |
| 14 | A+C      | wt     | GAT/GAC | wt  | Ex.6* |
| 15 | A+C      | wt     | GAT/GAC | wt  | Ex.6* |
| 16 | A+C      | wt     | wt    | wt    | Ex.8  |
| 17 | A+C      | wt     | wt*   | wt    | Ex.8* |

Multiple adenomas had the same cancer in common to be recognised by a* (adenomas N. 4, 5, 6, 7 and 8; adenomas 13, 14 and 15; adenomas 16 and 17). When sufficient material was not available, a dotted line was drawn instead of the data.

Significantly association between p53 mutations and severe dysplasia (p = 0.01). We also observed an increase of k-ras mutations among the adenomatous components having tubulo-villous and villous type but the association was not statistically significant (p = 0.1).

Table 6 shows the k-ras and p53 mutation analysis data obtained for 17 adenomas having cancer in the near vicinity and the corresponding cancers. Incidences of k-ras and p53 mutations for the adenomas were respectively 47% (8/17) and 7% (1/14), while for the carcinomas we obtained 55% (5/9) for k-ras and 44% (4/9) for p53.

4. Discussion

Human colorectal adenomas containing early cancer are relatively rare lesions that should provide the opportunity to observe some of the critical genetic events driving the transition from a preneoplastic to a neoplastic state. Pilot observations, done almost 12 years ago using this same human system, indicated that the adenoma–carcinoma transition was characterized by mutations of the p53 oncosuppressor gene [38]. A later model proposed that development of colorectal cancer was due to the cumulative effect of p53 with mutations of other genes like APC, k-ras, and DCC, rather than to an exact sequence of changes [9]. Recent understanding of these gene functions, however, stressed that in the original model of Vogelstein, contrary to the original proposal, the order of p53 mutations seems to be essential [3].

In the present study, we compared the frequencies of k-ras and p53 gene mutations in adenomas containing early cancer and determined their degree of intratumor heterogeneity among the adenomatous and carcinomatous components. This type of analysis should provide information on the relative timing and role of these mutations in association with a definitive growth advantage and selection of the progressing subclones. In fact, if a mutation is generated or selected late, when the adenoma was already formed, one should expect heterogeneity among the adenomatous and carcinomatous components.

Interestingly, mutations of the p53 gene were present in the carcinomatous components of the adenomas containing early cancer in approximately 50% of the cases that were not detected in the adenomatous components of the same adenomas. In addition, the p53 mutations within the adenomatous components were found to be statistically significantly associated with severe dysplasia. In contrast, the intratumor heterogeneity of the k-ras oncogene status between adenomatous and carcinomatous component pairs of the adenomas containing early cancer was relatively rare (in approximately 10% of the cases).

In the present study, we have also compared the incidence of mutations of the k-ras and p53 genes of adenomas containing early cancer with those of adenomas having cancer in the near vicinity. We found that the incidence of k-ras mutations was slightly higher in the adenomas having cancer in the near vicinity (47%) than in the adenomatous or carcinomatous component pairs of the adenomas containing early cancer (28% in both cases), but that the difference in the present series of cases was not statistically significant. On the contrary, the incidences of p53 mutations of the adenomas containing early cancer (52 and 64%, respectively, for the adenomatous and carcinomatous components) were statistically significantly higher than the incidence of p53 mutations for the adenomas having cancer in the near vicinity (7%).

The present findings reinforce the previous conclusions on the role of k-ras and p53 mutations in colorectal tumor progression seen earlier in the literature [9, 38]. Additionally, the present study demonstrates the
existence of genetic heterogeneity of p53 mutations and the high homogeneity of k-ras mutations during the adenoma–cancerina sequence. We suggest that in a number of adenomas containing early cancer the type of mutations, in particular, of the p53 tumor supressor gene, may also be relevant for tumor progression. In addition, the high intratumor heterogeneity of p53 mutations in a large number of cases suggests late occurrence of the p53 mutations. Thus, the present data suggest that the colorectal adenoma–cancerina transition is dependent on the order of the k-ras and p53 mutations and that p53 mutations are later decisive players at this transition. In other words, it appears that the different biological roles of k-ras and p53 in human colorectal adenomas containing early cancer are reflected by their timing and therefore by their degree of heterogeneity. In this respect, k-ras mutations, which are mainly intratumor homogeneous between the adenomatous and carcinomatous components, appear as early events that do not have the potential to drive the adenoma–cancerina transition. This also appears to be the case for human aberrant crypt foci, which are putative precursor lesions of the adenomas, for which k-ras mutations were recently detected in up to 80–90% of the cases [31,32,37,40].

The functional roles of the ras and p53 genes in colorectal cancer development and progression have been the subject of excellent studies, that were also extensively reviewed in the literature, but they are still not fully understood [3,9,19]. In particular, new functions of the Ras and P53 proteins associated to proliferation, apoptosis, and chromosomal stability became recently evident with the discovery of a plethora of new effector pathways and different cellular mechanisms [1,5,8,18,21,22,25,36,39].

Recently, we have also demonstrated that mutations of p53 and k-ras genes in human colorectal cancer were associated with DNA aneuploidy as assessed by flow cytometry [7,13–16] and that the results obtained for k-ras in the adenomas were confirmed by other studies using in vitro models in which a mutated k-ras was transfected [17,28,30].

In the present study we did not perform a correlative analysis of k-ras and p53 mutations with flow cytometric DNA aneuploidy due to the fact that DNA aneuploidy in colorectal adenomas is often near-diploid [12,13] and that paraffin embedded material does not allow great sensitivities with this technique [33]. Other techniques, like FISH and CGH, may be more suitable toward these aims [26,34].

In conclusion, the present study indicates that while the k-ras genetic status was mainly homogeneous within the adenomatus and carcinomatus components of adenomas with early cancer, the p53 genetic status, in contrast, was highly heterogeneous. These findings confirm the relevant role of p53 mutations as a late event in colorectal carcinogenesis for the acquisition of malignancy and stress the specificity of the mutations. The role of k-ras remains less clear, though it appears more related to tumor initiation and maintenance than to tumor progression.

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