Lung involvement in childhood onset granulomatosis with polyangiitis

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Abstract
Granulomatosis with polyangiitis is an ANCA-associated systemic vasculitis with a low incidence in the pediatric population. Lung involvement is a common manifestation in children affected by granulomatosis with polyangiitis, both at disease’s onset and during flares. Its severity is variable, ranging from asymptomatic pulmonary lesions to dramatic life-threatening clinical presentations such as diffuse alveolar haemorrhage. Several radiologic findings have been described, but the most frequent abnormalities detected are nodular lesions and fixed infiltrates. Interstitial involvement, pleural disease and pulmonary embolism are less common. Histology may show necrotizing or granulomatous vasculitis of small arteries and veins of the lung, but since typical features may be patchy, the site for lung biopsy should be carefully chosen with the help of imaging techniques such as computed tomography. Bronchoalveolar lavage is helpful to confirm the diagnosis of alveolar haemorrhage. Pulmonary function tests are frequently altered, showing a reduction in the diffusion capacity for carbon monoxide, which can be associated with obstructive abnormalities related to airway stenosis. Nodular lung lesions tend to regress with immunosuppressive therapy, but lung disease may also require second line treatments such as plasmapheresis. In cases of massive diffuse alveolar haemorrhage, ventilator support is crucial in the management of the patient.

Keywords: Granulomatosis with polyangiitis, Wegener granulomatosis, Childhood, Lung, Pulmonary

Background
Granulomatosis with polyangiitis (GPA), formerly known as Wegener’s granulomatosis [1], is an idiopathic vasculitis of medium and small arteries, characterized by necrotizing granulomatous inflammation of the respiratory tract with necrotizing, pauci-immune glomerulonephritis; vasculitis frequently involves also other organs. GPA is a member of a family of vasculitides associated with positive anti-neutrophil cytoplasmic antibodies (ANCA). In GPA, the pattern of autoantibody staining of ethanol-fixed neutrophils is typically cytoplasmic (c-ANCA), rather than perinuclear (p-ANCA), due to the presence of antibodies against proteinase 3, which is a constituent of the azurophilic granules of the neutrophil [2, 3].

The family of ANCA-associated vasculitides (AAV) also includes eosinophilic granulomatosis with polyangiitis (EGPA, previously known as Churg-Strauss syndrome) and microscopic polyangiitis (MPA). The clinical spectrum of vasculitides is broad; the lack of pathognomonic features and the presence of overlapping clinical manifestations makes diagnosis challenging, especially in discriminating one form from another. In 1990 the American College of Rheumatology (ACR) published the classification criteria for seven vasculitides, including GPA and EGPA [4–7]. However, because of the poor performance of ACR criteria in classifying children with vasculitis, the EULAR/PRINTO/PRES criteria were developed using pediatric data. The EULAR/PRINTO/PRES criteria for GPA (Table 1) showed improved sensitivity compared to the adult-based ACR criteria (93% vs 83%) [8–10]. However, both classification criteria did not include MPA, which had its specific definition thanks to the Chapel Hill Consensuses Conference (CHCC) [11]. It is therefore possible that cases classified as GPA according to ACR criteria may be described as MPA using the CHCC definition. In 2007 the European Medicines Agency (EMA) endorsed a classification algorithm to classify patients with a mutually exclusive diagnosis (EGPA, GPA, MPA, or polyarteritis nodosa) [12]. The EMA classification algorithm showed to...
Lung involvement in childhood-onset GPA
Prevalence and clinical features
Lung involvement is a frequent feature of GPA. Main features of lung involvement in children with GPA are displayed in Table 2. X-ray evidence of fixed pulmonary

| First author and year of publication | Number of patients | Median age at diagnosis (range) | Female | Lung involvement (%) | Hemoptysis/Alveolar haemorrhage | Dyspnea | Chronic cough | Pleural effusion/thickening | Lung nodules or cavity | Lung infiltrate |
|-------------------------------------|--------------------|--------------------------------|--------|----------------------|-------------------------------|---------|---------------|-----------------------------|----------------------|-----------------|
| Tahghighi 2013 [23]                | 11                 | 11 (6–15)                      | 5      | 7 (63.6%)            | 2                             | 2       | N.R.          | 1                           | N.R.                 | N.R.            |
| Iudici 2015 [25]                   | 25                 | 14 (2–17)                      | 18     | 17 (68%)             | 3                             | 3       | N.R.          | 1                           | N.R.                 | 7               |
| Sacri 2015 [26]                    | 28                 | 12.8 (10,1–14,8)               | 21     | 19 (67.8%)           | 12                            | N.R.    | N.R.          | 1                           | 7                    | 16              |
| Bohm 2014 [30]                     | 56                 | 38 (78,5%)                     | 14/55  | 7                    | 17                            | 26/55   | 2             | 4                           | 2                    | 2               |
| Kosalka 2014 [31]                  | 9                  | N.R.                           | 6      | 8 (88,9%)            | 4                             | N.R.    | 2             | 4                           | N.R.                 | 16              |
| Cabral 2016 [32]                   | 183                | 14 (2–18)                      | 113    | 136 (74,3%)          | 68                            | 15      | 99            | 25                          | 97                   | 64              |
| Betostotsky 2002 [33]              | 17                 | N.R.                           | 13     | 14 (82,3%)           | 3                             | 4       | 9             | N.R.                        | 2                    | 2               |
| Akikusa 2007 [22]                  | 25                 | 14,5 (8,7–17,1)                | 20     | 21 (84%)            | 12                            | N.R.    | 2             | 13                          | 6                    | N.R.            |
| Anilkumaran 2011 [27]              | 7                  | N.R.                           | 5      | 5 (71,4%)            | 2                             | N.R.    | N.R.          | N.R.                        | N.R.                 | N.R.            |
| Wong 1998 [28]                     | 12                 | N.R.                           | 8      | 7 (58,3%)            | 3                             | N.R.    | N.R.          | N.R.                        | N.R.                 | N.R.            |

N.R. not reported
infiltrates, nodules, or cavitations for more than 1 month essentially precludes a diagnosis of MPA according to the EMA algorithm to classify vasculitis [12]. MPA is a necrotizing vasculitis with few or no immune deposits, predominantly affecting small vessels (capillaries, venules, or arterioles) and more rarely small- and medium-sized arteries [11]. The limited available literature suggests that genetic, pathophysiological and prognostic differences exist between MPA and GPA, but distinguishing between these diseases is challenging because of overlapping clinical features. The absence of granulomata and sparing of the upper respiratory tract are features of MPA, in which necrotizing glomerulonephritis and pulmonary capillaritis are common. These features help to distinguish MPA from GPA [11].

Lung disease is frequently the presenting clinical manifestation of GPA, however it can sometimes develop later during the disease course. Pulmonary involvement is frequent during disease’s flares, also in patients who did not present lung disease at GPA onset [22]. In the ARChiVe (A Registry for Children with Vasculitis) cohort including 65 children affected by GPA, shortness of breath was reported in 58.5% of cases and chronic cough in 52.3% [20]. In a subsequent study on the ARChiVe cohort, including 183 patients with childhood-onset GPA, pulmonary involvement was detected in 74% of patients. Overall, 54% of patients presented chronic cough, while only 8% showed wheezing or expiratory dyspnea. Massive haemoptysis or alveolar haemorrhage was reported in 42% of patients. Chest imaging was abnormal in 89% of patients, showing nodules in 54% of cases, fixed pulmonary infiltrates in 36% and cavitations in 21%; other findings such as fibrosis, septal thickening and pneumothorax were detected in less than 10% of cases. Pleurisy affected 14% of patients: 12% of patients had respiratory failure and 22% required supplemental oxygen. In 31 patients lung biopsy was performed: in 77% of cases biopsy findings confirmed the presence of vasculitis or were consistent with features of vasculitis [32].

In the meta-analysis published by Iudici et al., lower respiratory tract involvement has a prevalence of 61% (95% confidence interval [CI] 48–74%), with haemoptysis/alveolar haemorrhage occurring in 16% of patients and lung nodules identified in 10% of patients [18]. It should be noted that Rottem et al. reported that 41% of radiographic abnormalities found in patients with childhood GPA were observed in absence of symptoms. [21]

Manifestations of lung involvement during flares are variable, but in a pediatric series of 25 patients, alveolar haemorrhage was reported to be the least common condition observed [22]. Considering mortality in childhood GPA, together with infections, respiratory complications (including pulmonary haemorrhage and chronic lung disease) were the most common causes of death [25, 32].

Nodules
Nodular lesions are the most common pulmonary radiologic finding in GPA, in both adults and children [22, 32, 39]. Their presence is not necessarily associated with clinical signs and symptoms. [21] They consist of granulomatous inflammation and necrosis. In children both multiple and single nodular lesions have been described; on chest radiography, their size may vary from 1 to 4 cm [29, 39–41]. In adult patients, nodules are usually multiple but in a number not greater than ten, with a size ranging from 1 to 10 cm [39]. Computed tomography (CT) shows also smaller nodules and can detect intranodular cavitation, that cannot be seen on chest x rays. Without treatment, nodules increase in number and size and tend to cavitate.

Pulmonary nodules usually regress with therapy in weeks or months, but local scarring is possible. Persistent nodules may indicate active disease, but could also be due to cicatricial residues in patients with inactive disease [39, 42]. The absence of regression or the extension of the lesion in a patient who is receiving immunosuppressive treatment may also be caused by a bacterial superinfection [43].

Diffuse alveolar haemorrhage
Diffuse alveolar haemorrhage is a life-threatening condition which presents with haemoptysis, anemia, and diffuse alveolar infiltrates leading to respiratory failure. The clinical triad is often incomplete, in particular hemoptysis may be absent in about a third of cases [44]. Hypocomplementemia at the time of diagnosis was recognized as a risk factor for alveolar haemorrhage [45].

Chest radiography shows a bilateral alveolar filling pattern, involving perihilar and basal lung fields. Bronchial fibroscopy shows a peripheral diffuse bleeding, and is required to exclude a localized cause of hemorrhage. Bronchoalveolar lavage (BAL) allows detection of blood red cells if the haemorrhage is acute or siderophages if it is chronic; furthermore, it is useful in ruling out infections, that may be suspected also due to the presence of constitutional symptoms such as fever [39]. The underlying histologic findings of diffuse alveolar haemorrhage are neutrophilic infiltration and fibrinoid necrosis of the alveolar and capillary walls [46].

A study in the adult population showed that in patients with alveolar haemorrhage due to AAV, the degree of hypoxemia upon presentation was the most important predictor of respiratory failure; a higher disease activity score, a higher percentage of neutrophils in the BAL fluid and higher C reactive protein levels were also associated with respiratory failure [47].

Other manifestations
Pulmonary fibrosis is uncommon in patients with antineutrophil cytoplasmic antibodies-associated vasculitis...
and mainly occurs in microscopic polyangiitis in association with anti-MPO antibodies; nonetheless, it was described in adult patients affected with GPA [48–51] and also in pediatric patients [32]. Interstitial involvement in a treated patient should induce the suspicion of a iatrogenic pneumopathy [39, 52].

Pleural involvement is present in a minority of patients, and may consist of pleural thickening, pneumothorax or pyopneumothorax, which can be caused by the rupture of a peripheral cavitated lesion into the pleural cavity.

It should be remembered that GPA is associated with an increased risk of venous thromboembolic events in adults and children: although in a small proportion of cases, also pulmonary embolic disease has been described in patients with childhood onset GPA, both at disease presentation and during follow-up [22, 53, 54].

Necrotizing granulomas not only alter the normal alveolar architecture progressively impairing the respiratory function, but may also cause bronchial stenosis. Multilevel airway stenosis was described in about one third of adult patients, in particular 17% had mid/distal tracheal stenosis and 11% had bronchial stenosis [55]. In children affected by GPA, tracheobronchial stenosis was detected in 17% of cases [35].

Histological findings
The simplest approach to collect tissue for histological analysis is transbronchial biopsy, but blind transbronchial biopsy of lung tissue is rarely informative because of the focal distribution of the inflammatory lesions in GPA. Open-lung biopsy has a higher accuracy, but also more significant morbidity [56–58]. Computed tomography may help in choosing the best site for tissue sampling, in order to increase the yield of transbronchial biopsy; lesions with little necrotic component should be preferred. In adults, image-guided automated core needle biopsy was even used to obtain histological proof of disease activity and guide therapy [59].

Necrotizing or granulomatous vasculitis may be found in small arteries and veins of the lung. The lumen of the vessels may be occluded by inflammatory infiltrate or by a thrombus [60]. Granulomas show central necrosis surrounded by histiocytes, lymphocytes and giant cells; also eosinophils may be present but in small proportion. In some cases nonspecific inflammation may be found [17].

Histology in pulmonary haemorrhage generally shows pulmonary capillaritis, which consists of interstitial neutrophil predominant infiltration, fibrinoid necrosis of the alveolar and capillary walls and leukocytoclasis, with subsequent loss of the integrity of the alveolar capillary membrane [46].

Radiologic findings
The typical lower respiratory tract involvement of GPA is demonstrated by radiologic findings of lung nodules, cavities or fixed infiltrates. In adults the most common radiological presentation is represented by multiple nodules randomly distributed throughout the lung, of variable size as mentioned above. They range from well to poorly defined and may be distributed along bronchovascular bundles. In approximately a half of cases, some of the nodules demonstrate cavitation, which is best seen by CT [61, 62].

Presence of nodules was the most frequent observation also in pediatric patients (Fig. 1) [22, 32]. However, in a small cohort of children, diffuse interstitial and alveolar opacities attributable to pulmonary hemorrhage were more common than nodular lesions [41]. Both nodules and regions of consolidation can be surrounded by haemorrhage, which in some cases dominates the radiographic appearance. Unfortunately, the radiographic appearance of GPA is very variable and not specific, making a diagnosis by imaging alone often difficult [39].

Conventional chest X rays can easily detect nodules and masses, fixed infiltrates, cavitations, pleural effusions and pneumothoraces, while small nodules, linear opacities, focal infiltrates and fluffy perivascular densities are best identified using high-resolution computed tomography (HRCT) [17] (Fig. 2). Bilateral perihilar and basal infiltrates of alveolar haemorrhage may resemble pulmonary oedema, but differential diagnosis can be based

Fig. 1 Chest X ray in a 16-year old girl affected by granulomatosis with polyangiitis (GPA). The image shows multiple nodular lesions together with diffuse opacities in the lower and middle regions of the lung.
on the absence of cardiomegaly and signs of venous hypertension. Following acute diffuse alveolar haemorrhage, a radiographic pattern of septal thickening, known as crazy-paving, can appear [62].

The most common HRCT abnormalities are lung nodules, usually multiple and bilateral which tend to increase with disease progression. They can range from few millimeters to >10 cm in diameter and become cavitated. Cavities are usually thick walled and characterized by an irregular inner margin and absent calcification. The nodular lesions are often related to the vessels, and they tend to involve mainly the subpleural regions but have no predilection for the upper or lower lung zones. Haemorrhage may result in a HRCT finding consisting of an edge of ground glass opacity surrounding the lung lesion, known as halo sign [62, 63]. HRCT could be used also to identify active disease: ground-glass opacities, cavitations and masses measuring more than 3 cm tend to correspond to active disease; while non-cavitated small nodules and linear opacities can also represent cicatrical lesions [64].

18-F-fluorodeoxyglucose positron emission tomography/computed tomography scans have been used to evaluate global disease distribution and to identify active lesions suitable for biopsy [65, 66].

**Pulmonary function tests**

When tested for pulmonary function, patients affected by GPA most commonly show airflow obstruction, often in association with a reduced diffusion capacity of the lung for carbon monoxide. Reduction of lung volumes has also been described [67, 68]. In a pediatric series, of the 35 patients who performed pulmonary function tests, 27 had abnormal results, with obstructive and restrictive abnormalities detected in the same proportion of cases [20]. Sixty-seven patients of the ARCHiVe cohort performed pulmonary function tests, which were abnormal in 61% of cases [32]. Since pulmonary involvement in GPA may be initially asymptomatic, it should be investigated not only radiographically, but also with pulmonary function testing. The first sign of pulmonary haemorrhage may be an alteration in the diffusion capacity of the lung for carbon monoxide [21, 57, 69]. In patients with subglottic stenosis, the flow-volume curve shows a flattening in both inspiratory and expiratory phase, consistent with an extrathoracic airway obstruction [70].

**Differential diagnosis**

In the presence of lung disease, concomitant upper airway and kidney involvement can guide the clinician towards GPA diagnosis. Other conditions associated with pulmonary-renal syndrome, such as Goodpasture’s syndrome, polyarteritis nodosa, systemic lupus erythematosus or mixed connective tissue disease, should be excluded. Laboratory evidence of positive ANCA, in particular c-ANCA or PR3-ANCA, and a compatible kidney biopsy, are supportive findings of GPA diagnosis. Lung involvement of ulcerative colitis associated with ANCA positivity may also mimic GPA [71].

Considering nodular lung lesions, neoplastic disease and sarcoidosis should also be suspected. In children, also chronic granulomatous disease has to be considered [72]. Other causes of alveolar haemorrhage described in the pediatric population are idiopathic pulmonary hemosiderosis and childhood onset systemic lupus erythematosus [73–75].

Another important differential diagnosis is with infectious processes, in particular mycobacterial, micotic or helminthic infections [76]. Diffuse alveolar haemorrhage has been described as a consequence of pulmonary histoplasmosis [77]. It should also be remembered that lung disease in GPA patients may also be due to concomitant infections, in particular *Pneumocystis jirovecii* infection in the immunocompromised patient [78, 79].

**Treatment**

Treatment requires a remission induction phase, followed by maintenance therapy. Standard treatment of GPA has primarily consisted of glucocorticoids and cyclophosphamide. Several other immunosuppressive agents have been used, in monotherapy or in combination with glucocorticoids, including methotrexate, azathioprine, mycophenolate, cyclosporine, colchicines, etanercept, infliximab,
adalimumab, rituximab. Remission induction is mainly based on the use of glucocorticoids and cyclophosphamide; cyclophosphamide can be administered either orally or with intravenous pulses and should be withdrawn when remission is achieved. Remission maintenance therapy, which should last at least 18–24 months, is usually based on the introduction of azathioprine or methotrexate, with concomitant tapering of glucocorticoids [18, 19, 21, 80–84].

Other therapeutic strategies have been used in severe and refractory cases. Intravenous immunoglobulin administration has been used as adjunctive therapy in AAV patients with refractory or relapsing disease, even if 11.9% of patients presented serious adverse events [85]. In severe cases of alveolar haemorrhage, together with aggressive immunosuppressive therapy, plasmapheresis has been used [86]. The use of plasma exchange in patients with creatinine levels > 5.8 mg/dl was proved to be more effective than intravenous methylprednisolone in preventing end-stage renal disease at 12 months, but no difference was observed in survival rates and incidence of adverse events [87]. Even if plasma exchange is usually reserved for refractory disease and life-threatening conditions, it has been proposed also for induction of remission in less severe cases [88]. The PEXIVAS study will better define the benefits of plasma exchange in AAV. [89] Kidney involvement may require dialysis and kidney transplant [90].

So far, no clinical trial has been conducted in the pediatric population and no pediatric specific recommendations are available. Pediatric guidelines for GPA will be hopefully developed by the SHARE project, whose aim is to provide recommendations for the care of children and young adults with rheumatic diseases, including vasculitis [91]. At the moment, therapeutic management is guided by data extrapolated from adult studies. Guidelines for the treatment of AAV in adult patients have recently been published [92, 93].

Of note, both British and EULAR recommendations include rituximab, an anti-CD20 monoclonal antibody, which has proved to be a promising new therapeutic option for AAV both in remission induction and in remission maintenance [92–96]. The RAVE trial, which included AAV patients older than 15 years, showed that rituximab was not inferior to cyclophosphamide in achieving remission induction and was more effective for relapsing disease; better response to rituximab was observed in anti-PR3 positive patients [94]. Rituximab should be considered as a therapeutic option for remission induction also in children, since steroid sparing in pediatric age is mandatory and cyclophosphamide avoidance is especially desirable in young people at risk of infertility. In a pediatric series, 10 patients with primary systemic vasculitis, including 4 children with GPA, were treated with rituximab with a decrease in disease activity and in corticosteroid dose. Of 10 patients receiving rituximab, 3 presented adverse events: one patient with unclassified vasculitis had mild headache with second rituximab infusion; one patient with GPA developed paronychia 2 months after receiving rituximab treatment; one patient with GPA treated with both infliximab and rituximab presented, respectively 9 and 7 months after drug discontinuation, a Pseudomonas urinary tract infection and concurrent pneumonia, while he was also being administered concurrent immunosuppressive therapy with cyclosporine A for renal transplant [97]. At the moment, as long-term safety data on the use of rituximab in pediatric vasculitis are still missing, rituximab should be prescribed carefully, in refractory cases. A phase IIa international open-label trial is currently recruiting patients to evaluate the safety and pharmacokinetics of rituximab in children with severe GPA or MPA (PePRS study, NCT01750697), but mechanical ventilation due to alveolar hemorrhage represents an exclusion criterion for participation in the study. It should be noted that also the RAVE trial excluded patients with alveolar haemorrhage requiring mechanical respiratory assistance upon enrollment [94].

A retrospective study analyzing a cohort of patients with alveolar haemorrhage secondary to AAV, including also patients needing mechanical ventilation, showed that complete remission by 6 months was achieved in a higher proportion of cases with rituximab than with cyclophosphamide, even though the use of rituximab was not associated with a higher long-term survival rate. The same study could not demonstrate plasma exchange efficacy in addition to the standard remission induction therapy [47].

Lung disease can also necessitate the use of supportive measures. Pulmonary haemorrhage may require ventilator support in an intensive care unit setting or even extracorporeal membrane oxygenation [20, 98]. Akikusa and colleagues reported a median duration of intubation of 8 days in the subgroup of children requiring ventilation [22].

Airway lesions rarely respond satisfactorily to systemic therapy with cyclophosphamide, but rituximab appeared to be a more efficient therapy for tracheobronchial lesions [35, 99, 100]. In the presence of subglottic or tracheal involvement, surgical intervention is often needed to maintain a patent airway [101–103].

**Conclusion**

Lung involvement is a common manifestation in children affected by GPA. Its severity is variable, ranging from asymptomatic pulmonary lesions to dramatic life-threatening clinical presentations such as diffuse alveolar haemorrhage. Several radiologic findings have been
described, but the most frequent abnormalities detected are nodular lesions and fixed infiltrates. Nodular lung lesions tend to regress with immunosuppressive therapy, but lung disease may also require second line treatments. Rituximab may represent a promising treatment option also in pediatric patients, even if its efficacy and safety in children should be better studied. In cases of massive diffuse alveolar haemorrhage, ventilator support is crucial in the management of the patient.

**Abbreviations**

ACR: American College of Rheumatology; CI: Confidence interval; CT: Computed tomography; GPA: Granulomatosis with polyangiitis; MPA: Microscopic polyangiitis

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**Competing interests**

The authors declare that they have no competing interests.

**Consent for publication**

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**References**

1. Falk RJ, Gross WL,Guillevin L, Hoffman GS, Jayne DR, Jennette JC, Kallenberg CG, Luqmani R, Mahr AD, Matteson EL, et al. Granulomatosis with polyangiitis (Wegener’s): an alternative name for Wegener’s granulomatosis. Arthritis Rheum. 2011;63:863–4.

2. Comarmond C, Cacoub P. Granulomatosis with polyangiitis (Wegener’s): clinical aspects and treatment. Autoimmun Rev. 2014;13:1121–5.

3. Niles JL, McCluskey RT, Ahmad MF, Arnaout MA. Wegener’s granulomatosis autoantigen is a novel neutrophil serine proteinase. Blood. 1989;74:1888–93.

4. Hunder GG, Arend WP, Bloch DA, Calabrese LH, Fauci AS, Fries JF, Leavitt RY, Lie JT, Lightfoot Jr JW, Masi AT, et al. The American College of Rheumatology 1990 criteria for the classification of vasculitis. Introduction. Arthritis Rheum. 1990;33(8):1065–7.

5. Leavitt RY, Fauci AS, Bloch DA, Michel BA, Hunder GG, Arend WP, Calabrese LH, Fries JF, Lie JT, Lightfoot Jr JW, et al. The American College of Rheumatology 1990 criteria for the classification of Wegener’s granulomatosis. Arthritis Rheum. 1990;33(11):101–7.

6. Masi AT, Hunder GG, Lie JT, Michel BA, Bloch DA, Arend WP, Calabrese LH, Edworthy SM, Fauci AS, Leavitt RY, et al. The American College of Rheumatology 1990 criteria for the classification of Churg-Strauss syndrome (allergic granulomatosis and angitis). Arthritis Rheum. 1990;33(9):1094–100.

7. Bloch DA, Michel BA, Hunder GG, McShane DJ, Arend WP, Calabrese LH, Edworthy SM, Fauci AS, Fries JF, Leavitt RY, et al. The American College of Rheumatology 1990 criteria for the classification of vasculitis. Patients and methods. Arthritis Rheum. 1990;33(8):1068–73.

8. Ozen S, Ruperto N, Dillon MJ, Bagga A, Barron K, Davin JC, Kawasaki T, Lindsay C, Petty RE, Prieur AM, et al. Eular/pres endorsed consensus criteria for the classification of childhood vasculitides. Ann Rheum Dis. 2006;65:936–41.

9. Ozen S, Pistorio A, Iuson SM, Bakkaloglu A, Herlin T, Brik R, Buoncappaglia A, Lazar C, Bilge I, Uzel Y, et al. Eular/pres endorsed consensus criteria for Henoch-Schönlein purpura, childhood polyarteritis nodosa, childhood Wegener granulomatosis and childhood Takayasu arteritis: Ankara 2008. Part II: Final classification criteria. Ann Rheum Dis. 2010;69:798–806.

10. Ruperto N, Ozen S, Pistorio A, Dolezalova P, Brogan D, Cabaš D, Cuttica R, Khrubchanadi R, Lloyd D, O’Neill K, et al. Eular/pres criteria for Henoch-Schönlein purpura, childhood polyarteritis nodosa, childhood Wegener granulomatosis and childhood Takayasu arteritis: Ankara 2008. Part I: Overall methodology and clinical characterisation. Ann Rheum Dis. 2010;69:790–7.

11. Jennette JC, Falk RJ, Andassy K, Bacon PA, Chung J, Gross WL, Hagen EC, Hoffman GS, Hunder GG, Kallenberg CG, et al. Nomenclature of systemic vasculitides. Proposal of an international consensus conference. Arthritis Rheum. 1994;37(2):187–92.

12. Watts RA, Lane S, Helslisk T, Hauser T, Hellmich B, Koldingsnes W, Mahr A, Segelmark M, Cohen-Tervaert JW, Scott D. Development and validation of a consensus methodology for the classification of the anca-associated vasculitides and polyarteritis nodosa for epidemiological studies. Ann Rheum Dis. 2007;66:222–7.

13. Uribe AG, Huber AM, Kim S, O’Neill KM, Wahezi DM, Abramson L, Baszis K, Benseler SM, Bowyer SL, Campbell S, et al. Increased sensitivity of the European medicines agency algorithm for classification of childhood granulomatosis with polyangiitis. J Rheumatol. 2012;39(8):1687–97.

14. Watts RA, Lane SE, Bentham G, Scott DG. Epidemiology of systemic vasculitis: a ten-year study in the united kingdom. Arthritis Rheum. 2000;43:414–9.

15. Cotech MF, Hoffman GS, Yerg DE, Kaufman GI, Targonski P, Kaslow RA. The epidemiology of Wegener’s granulomatosis. Estimates of the five-year period prevalence, annual mortality, and geographic disease distribution from population-based data sources. Arthritis Rheum. 1996;39:897–92.

16. Grisaru S, Yuen GW, Miettunen PM, Hamiwka LA. Incidence of Wegener’s granulomatosis in children. J Rheumatol. 2010;37:440–2.

17. Cabral DA, Morisitha K. Antineutrophil cytoplasmic antibody associated vasculitis. In: Petty RE, Laxer R, Lindley C, Wedderburn L, editors. Textbook of pediatric rheumatology. 7th ed. Philadelphia: Saunders; 2016. p. 484–98.

18. Ludici M, Quartier P, Terrier B, Mouton L, Guillen L, Puechel X. Childhood-onset granulomatosis with polyangiitis and microscopic polyangiitis: systematic review and meta-analysis. Orphanet J Rare Dis. 2016;11:141.

19. Stegmayr BG, Goethefs L, Malmer B, Muller Wielfeld DE, Nilsson K, Sundelin B. Wegener granulomatosis in children and young adults. A case study of ten patients. Pediatr Nephrol. 2000;14:208–13.

20. Cabral DA, Uribe AG, Benseler S, O’Neill KM, Hashkes P.J, Higgins G, Zeff AS, Lovell DJ, Kingsbury DJ, Stevens A, et al. Classification, presentation, and initial treatment of Wegener’s granulomatosis in childhood. Arthritis Rheum. 2010;62:440–24.

21. Kallenberg CG, Luqmani R, Mahr AD, Matteson EL, et al. Granulomatosis with polyangiitis (Wegener’s): an alternative name for Wegener’s granulomatosis. Arthritis Rheum. 2010;62:440–24.

22. Neil KM, Hashkes PJ, Higgins G, Zeff AS, Lovell DJ, Kingsbury DJ, Stevens A, et al. Classification, presentation, and initial treatment of Wegener’s granulomatosis in childhood. Arthritis Rheum. 2009;60:3413–24.

23. Rottem M, Fauci AS, Hallahan CW, Kerr GS, Lebovics R, Leavitt RY, Hoffman GS. Wegener granulomatosis in children and adolescents: clinical presentation and outcome. J Pediatr. 1993;122:26.

24. Akkus OA, Schneider R, Harvey EA, Hebet D, Thornton PS, Laxer RM, Silverman ED. Clinical features and outcome of pediatric Wegener’s granulomatosis. Arthritis Rheum. 2007;57:837–44.

25. Tajighighi F, Moradinejad MH, Aghighy Y, Shari R, Raeeskarami SR, Salehzadeh F, Javadi V, Ziaee V. Evaluation of 10-year experience of Wegener’s granulomatosis in iranian children. ISRN Rheumatol. 2013;2013:794928.

26. Gajic-Veljic M, Nikolic M, Peco-Antic A, Bogdanovic R, Andrejevic S, Bonacik-Nikolic B. Granulomatosis with polyangiitis (Wegener’s granulomatosis) in children: Report of three cases with cutaneous manifestations and literature review. Pediatr Dermatol. 2013;30(2):e37–42.
71. dos Santos CE, Dal Pizzolo V, Araujo SC, Rachid Filho A, Marques FM. C-anca associated vasculitides in patients with ulcerative colitis: case report. Rev Bras Reumatol. 2013;53:41–3.

72. Mahdaviani SA, Mehrian P, Najafi A, Khalilzadeh S, Estampanah S, Nasri A, Karam MB, Rezaei N, Velayati AA. Pulmonary computed tomography scan findings in chronic granulomatous disease. Allergol Immunopathol (Madr). 2014;42:444–4.

73. Castelazz L, Patria MF, Frati G, Esposito AA, Esposito S. Iatridosip pulmonary haemosiderosis in paediatric patients: How to make an early diagnosis. Ital J Pediatr. 2016;42:86.

74. Kimura D, Shah S, Briceno-Medina M, Sathanandam S, Haberman B, Zhang J, Myers K, Kumar TS, Knott-Craig C. Management of massive diffuse alveolar hemorrhage in a child with systemic lupus erythematosus. J Intensive Care. 2015;3:10.

75. Singla S, Canter DL, Vecchi TJ, Muscal E, DeGuzman M. Diffuse alveolar hemorrhage as a manifestation of childhood-onset systemic lupus erythematosus. Hosp Pediatr. 2016;6:496–500.

76. Fauci AS, Haynes BF, Katz P, Wolff SM. Wegener granulomatosis: prospective clinical and therapeutic experience with 85 patients for 21 years. Ann Intern Med. 1983;98:76–85.

77. Grover K, Zainah N, Bhatnagar S, Stein T. A rare case of diffuse alveolar hemorrhage secondary to acute pulmonary histoplasmosis. Case Rep Infect Dis. 2015;2015:821749.

78. Grewal P, Brassard A. Fact or fiction: does the non-hiv/aids immunosuppressed patient need pneumocystis jiroveci pneumonia prophylaxis? An updated literature review. J Cutan Med Surg. 2009;13:309–12.

79. Croxton C, Holle JU, Gross WL. Value of anti-infective chemoprophylaxis in pediatric patients: How to make an early diagnosis. J Laryngol Otol. 2014;128:831.

80. Yates M, Watts RA. ANCA-associated vasculitis. Clin Med (Lond). 2017;17(1):60–4.

81. Wegener granulomatosis. Etanercept Trial (WGET). Control Clin Trials. 2002;23:450–61.

82. Pinheiro JD, Marques FM, Pires AL, Silva KS, Santos G, et al. Prospective study of TNFalpha blockade with infliximab in anti-neutrophil cytoplasmic antibody-associated systemic vasculitis. J Am Soc Nephrol. 2004;15(3):717–21.

83. Laurino S, Chaudhry A, Booth A, Conte G, Jayne D. Prospective study of TNFalpha blockade with adalimumab in ANCA-associated systemic vascular involvement. Nephrol Dial Transplant. 2010;25(10):3307–14.

84. Crick E, Machelart J, Lazaro E, Kahn JE, Cohen-Aubart F, Martin T, Mania A, Hatron PY, hayem G, Blanchard-Delaunay C, et al. Intravenous immunoglobulin as an immunomodulating agent in antineutrophil cytoplasmic antibody-associated vasculitides: A French nationwide study of ninety-Two patients. Arthritis Rheumatol. 2016;68(3):702–12.

85. Nguyen T, Martin MK, Indikovs A. Plasmapheresis for diffuse alveolar hemorrhage in a patient with wegener’s granulomatosis: case report and review of the literature. J Clin Apher. 2005;20:230–4.

86. Jayne DR, Gaskin G, Rassmannen N, Abramowicz D, Pez JP, Guillevin L, Cottin V, Cordier JF, Guillevin L. Tracheobronchial stenoses in granulomatosis with polyangiitis (wegener’s): a report on 26 cases. Medicine (Baltimore). 2015;94:e1088.

87. Jordan ND, Verma H, Siddiqui A, Morrison GA, D’Cruz DP. Morbidity and mortality associated with subglottic laryngotracheal stenosis in granulomatosis with polyangiitis (wegener’s granulomatosis): a single-centre experience in the united kingdom. J Laryngol Otol. 2014;128:831–7.

88. Croxton C, Holle JU, Gross WL. Value of anti-infective chemoprophylaxis in pediatric patients: How to make an early diagnosis. J Laryngol Otol. 2014;128:831.

89. Walsh M, Merkel PA, Pez JP, Guillevin L, Pusey CD, de Zuyts JA, Ives N, Clark WF, Quellen K, PEIXIS Investigators, et al. Plasma exchange and glucocorticoid dosing in the treatment of anti-neutrophil cytoplasm antibody associated vasculitides (PEIXIS): protocol for a randomized controlled trial. Trials. 2013;14:73.

90. Hruskova Z, Stel VS, Jayne D, Aasard K, De Meester J, Ekstrand A, Eller K, Hagf JG, Hotma A, Mantz Jemeris C, et al. Characteristics and outcomes of granulomatosis with polyangiitis (Wegener) and microscopic polyangiitis requiring renal replacement therapy: results from the European renal association-European dialysis and transplant association registry. Am J Kidney Dis. 2015;66:613–20.

91. Wulftraat NM, Vastert B. Time to share. Pediatr Rheumatol Online J. 2013;11:5.

92. Nittasuri E, Carruthers D, Chakravarty K, D’Cruz D, Harper L, Jayne D, Luqmani R, Mills J, Mooney J, Venning M, et al. BSR and BHRP Standards, Guidelines and Audit Working Group. BSR and BHRP guideline for the management of adults with ANCA-associated vasculitis. Rheumatology (Oxford). 2014;53(12):2306–9.

93. Yates M, Watts RA, Bajema IM, Cid MC, Crestani B, Hauser T, Hellmich B, Holle JU, Lauaran M, Little MA, et al. EULAR/ERA-EDTA recommendations for the management of ANCA-associated vasculitis. Ann Rheum Dis. 2016;75(9):1583–94.

94. Stone JH, Merkel PA, Speira R, Sxe P, Langford CA, Hoffman GS, Kallenberg CG, St Clair EW, Turkewicz A, Tchao NK, et al. Rituximab versus cyclophosphamide for ANCA-associated vasculitis. N Engl J Med. 2010;363(3):221–32.

95. Jones RB, Tervaert JW, Hauser T, Luqmani R, Morgan MD, Peh CA, Savage CO, Segelmark M, Sesar V, van Paasen P, et al. Rituximab versus cyclophosphamide in ANCA-associated renal vasculitis. N Engl J Med. 2010;363(3):211–20.

96. Guillevin L, Pagnoux C, Karas A, Khoatra C, Aumître O, Cohen P, Maurier F, Debaux O, Ninet J, Gamber P, et al. Rituximab versus azathioprine for maintenance in ANCA-associated vasculitis. N Engl J Med. 2014;371(19):1771–86.

97. Bethehoui D, Melo M, Marks SQ, Tullus K, Sills J, Cleary G, Dolezalova P, Ozon S, Pilkington C, Woo P, et al. Biologic therapy in primary systemic vasculitis of the young. Rheumatology (Oxford). 2009;48(8):978–86.

98. James KE, Xiao R, Merkel PA, Weiss PF. Clinical course and outcomes of childhood-onset granulomatosis with polyangiitis. Clin Exp Rheumatol 2016.

99. Hoffman GS, Thomas-Golbanov CK, Chan J, Alok LM, Blachar I. Treatment of subglottic stenosis, due to Wegener’s granulomatosis, with intrathecal corticosteroids and dialation. J Rheumatol. 2003;30:107–21.

100. Girard C, Charles P, Terrier B, Rousonne G, Cohen P, Pagnoux C, Cottin V, Cordier JF, Guillevin L. Tracheobronchial stenoses in granulomatosis with polyangiitis (wegener’s): a report on 26 cases. Medicine (Baltimore). 2015;94:e1088.

101. Jordan ND, Verma H, Siddiqui A, Morrison GA, D’Cruz DP. Morbidity and mortality associated with subglottic laryngotracheal stenosis in granulomatosis with polyangiitis (wegener’s granulomatosis): a single-centre experience in the united kingdom. J Laryngol Otol. 2014;128:831–7.

102. Lee PY, Adli EA, irace A, Neff L, Son MB, Lee EY, Perez-Atyade A, Rahr B. The presentation and management of granulomatosis with polyangiitis (Wegener’s Granulomatosis) in the pediatric airway. Laryngoscopy. 2017;127:233–40.

103. Fijolek J, Wiatr E, Gawryluk D, Martusewicz-Boros MM, Orlowski TM, Dziedzic D, Polubicz-Kowacka M, Onisz K, Langfort R, Roszkowski-Sz K. Intratracheal dialtion-injection technique in the treatment of granulomatosis with polyangiitis patients with subglottic stenosis. J Rheumatol. 2016;43(11):2042–8.