Aptamers as a Sensitive Tool to Detect Subtle Modifications in Therapeutic Proteins

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Abstract
Therapeutic proteins are derived from complex expression/production systems, which can result in minor conformational changes due to preferential codon usage in different organisms, post-translational modifications, etc. Subtle conformational differences are often undetectable by biochemical methods but can sometimes profoundly impact the safety, efficacy and stability of products. Numerous bioanalytical methods exist to characterize the primary structure of proteins, post-translational modifications: protein-substrate/protein/protein interactions and functional bioassays are available for most proteins that are developed as products. There are however few analytical techniques to detect changes in the tertiary structure of proteins suitable for use during drug development and quality control. For example, x-ray crystallography and NMR are impractical for routine use and do not capture the heterogeneity of the product. Conformation-sensitive antibodies can be used to map proteins. However the development of antibodies to represent sufficient epitopes can be challenging. Other limitations of antibodies include limited supply, high costs, heterogeneity and batch to batch variations in titer. Here we provide proof-of-principle that DNA aptamers to thrombin can be used as surrogate antibodies to characterize conformational changes. We show that aptamers can be used in assays using either an ELISA or a label-free platform to characterize different thrombin products. In addition we replicated a heat-treatment procedure that has previously been shown to not affect protein activity but can result in conformational changes that have serious adverse consequences. We demonstrate that a panel of aptamers (but not an antibody) can detect changes in the proteins even when specific activity is unaffected. Our results indicate a novel approach to monitor even small changes in the conformation of proteins which can be used in a routine drug-development and quality control setting. The technique can provide an early warning of structural changes during the manufacturing process that could have consequential outcomes downstream.

Introduction
Therapeutic proteins now represent a significant segment of the pharmaceutical industry [1] and include some of the most innovative products which are on the cutting edge of clinical care. This class of therapeutics is clearly different from small molecule synthetic drugs. Protein-drugs are 100 to 1,000 times more expensive than small molecule drugs but offer the potential for a more selective delivery of therapeutic effects. For example, x-ray crystallography and NMR are impractical for routine use and do not capture the heterogeneity of the product. Conformation-sensitive antibodies can be used to map proteins. However the development of antibodies to represent sufficient epitopes can be challenging. Other limitations of antibodies include limited supply, high costs, heterogeneity and batch to batch variations in titer. Here we provide proof-of-principle that DNA aptamers to thrombin can be used as surrogate antibodies to characterize conformational changes. We show that aptamers can be used in assays using either an ELISA or a label-free platform to characterize different thrombin products. In addition we replicated a heat-treatment procedure that has previously been shown to not affect protein activity but can result in conformational changes that have serious adverse consequences. We demonstrate that a panel of aptamers (but not an antibody) can detect changes in the proteins even when specific activity is unaffected. Our results indicate a novel approach to monitor even small changes in the conformation of proteins which can be used in a routine drug-development and quality control setting. The technique can provide an early warning of structural changes during the manufacturing process that could have consequential outcomes downstream.
primary and secondary structures) with a high degree of accuracy and sensitivity. Significant improvements in mass spectrometry over the last decade allow the determination of variations in post-translational modifications. Similarly the biophysicist's toolkit offers a choice of technologies to quantify protein-protein and protein-substrate interactions. Moreover robust assays to measure the biochemical activity of most protein products have been well established. A significant gap, however, remains in monitoring the tertiary and quaternary structures of proteins during drug development and manufacture [9]. Techniques currently used to determine the structures of proteins such as X-ray crystallography and NMR do not lend themselves to routine use during the manufacturing process. Issues of cost, time and technical skills aside these techniques fail to capture the heterogeneity of the product, which is a hallmark of biotherapeutics. It has, for example, been estimated that as many as 10^6 possible product-variants and impurities for a monoclonal antibody product can occur [2].

Epitope mapping using antibodies is one of the few methods currently available to monitor the conformation of a protein product [10–12]. In recent years synthetic nucleic acid reagents, called aptamers have emerged as surrogates to antibodies and appear to be particularly suited for bioanalytical applications [13]. Aptamers are engineered through repeated rounds of in vitro selection, SELEX (systematic evolution of ligands by exponential enrichment) to bind virtually any molecular target such as small molecules, proteins, nucleic acids, and even cells, tissues and organisms. The in vitro selection process confers significant advantages to aptamers over antibodies. The three topical thrombins currently marketed in the US were not developed as biosimilars, nonetheless these products are used for the same indications and are generally regarded as interchangeable in a clinical setting [14]. However, these products are derived from different sources and have different manufacturing processes [15] and offer a good real life model system to evaluate the panel of aptamers for discriminating between different potential conformations of similar proteins. Here we present proof-of-principle that aptamers can be used to characterize conformational epitopes on therapeutic proteins. We have used six anti-human thrombin aptamers to characterize epitopes on three thrombin products currently marketed. The aptamers clearly distinguish between the human and bovine thrombin products. However the two human thrombin products (one purified from human plasma and the other manufactured by recombinant DNA technology) show comparable binding to all six aptamers. We also heat treated the human thrombin products using a procedure similar to that resulting in enhanced immunogenicity in Factor VIII (FVIII) [16,17]. This treatment did not affect thrombin activity or the binding to an anti-thrombin antibody. The panel of anti-thrombin aptamers however detected significant alterations in the conformation of thrombin epitopes.

### Results

Using aptamers as surrogate antibodies in an ELISA assay

We identified six anti-thrombin DNA aptamers previously described to bind different thrombin epitopes with varying affinities [18,19]. The affinities of these anti-thrombin aptamers for thrombin have been determined using different assays and there have been some discrepancies (see [18–20]). For consistency of results it is very important that the conditions for the generation of aptamers be rigorously adhered to. Moreover the assay we developed uses biotinylated aptamers, which could affect the affinity. The primary sequences of the six polynucleotides used in this study are depicted in Table 2 and the conditions for generating the aptamers are given in the Materials and Methods. The aptamers were first evaluated using a sandwich ELISA. This assay was standardized using an anti-thrombin monoclonal capture antibody and a biotinylated polyclonal anti-thrombin detector antibody (Fig. 1A). In modifying this assay, we used the biotinylated anti-thrombin aptamers in lieu of the detector antibody (Fig. 1B). Both assays use the same monoclonal antibody for capture and identical reagents, NutraAvidin-HRP and TMB. Using recombinant thrombin expressed in CHO cells (Reco-throm), we demonstrate that the limit of detection for thrombin is approximately 10 ng/ml using either the antibody or the aptamer (Figs. 1 C&D). We therefore used a fixed concentration of the thrombin (5 µg/ml) to determine the relative affinities of the six aptamers used in this study (Fig. 2 and Table 3). The assay is robust, reproducible and the six aptamers bind to the thrombin with apparent K_d values ranging from 0.38±0.01 nM to 2.75±0.36 nM (Table 3). Prior to each experiment the oligonucleotides were denatured and then allowed to refold under controlled conditions (see Materials and Methods for details) to form the aptamers. The SD thus represents not only the variation in the assay per se but also the variability in process of generating the aptamers.

Comparing the binding of aptamers to thrombin products

There are three topical thrombin products in the market. Prior to 2007 the only topical thrombin available was one derived from bovine plasma (Thrombin JMI), since then a human-plasma derived thrombin (Evithrom) and a human recombinant thrombin (Recothrom) have been approved [14]. Bovine thrombin is produced by extracting prothrombin from bovine plasma. After activation to thrombin, the product undergoes a chromatographic

### Table 1. Analytical techniques used to characterize protein therapeutics.

| Composition, primary structure | Peptide mapping (mass spectrometry), peptide mass fingerprint (mass spectrometry), amino acid sequencing |
|--------------------------------|----------------------------------------------------------------------------------------------------------|
| Higher-order structure         | Spectroscopy, thermal stability testing                                                                   |
| Conformation                   | ELISA, epitope mapping                                                                                   |
| Post-translational modifications (e.g., glycosylation) | Mass spectrometry, ion-exchange chromatography                                                          |
| Size, detection of aggregates | Gel electrophoresis, size-exclusion chromatography, analytical ultracentrifugation                      |
| Binding                        | Cell assays, spectroscopy, SPR, ELISA                                                                 |
| Biological activity            | Cell assays, animal models                                                                               |

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purification process that includes ion exchange and viral filtration [21]. Human thrombin is derived from human plasma [22] while human recombinant thrombin is produced via recombinant DNA technology from genetically modified Chinese hamster ovary (CHO) cells [23]. Studies have indicated that the efficacies of the human thrombins are comparable to that of the bovine thrombin which has been used in the clinic for decades [24,25]. Fig. 3A shows a colloidal blue stained gel with the three thrombin products. All three products are highly purified, however the major protein component of the plasma derived human product is human serum albumin (HSA) which is used as a stabilizing agent [22]. No proteins are added to the bovine thrombin or the human recombinant thrombin drug-products [21,23]. Despite these differences in manufacturing processes and excipients, the binding of the Aptamer 30-8 to the two human thrombin products is almost indistinguishable (Fig. 3B). Furthermore when we compared the kinetics of the binding of all six aptamers to the two human thrombin products we found an extremely high correlation ($r = 0.996$, $P < 0.05$) (Fig. 3C). Interestingly none of the six aptamers binds to the bovine thrombin (data not given).

**Aptamers can detect subtle conformational changes in human thrombin products**

There are many instances where a small change in manufacturing process significantly impacts a protein therapeutic (see [2] for numerous examples). For example, a manufacturing change involving heat treatment of FVIII resulted in a significant increase

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**Figure 1. ELISA based binding assays using aptamers as surrogate antibodies.** (A & B) Illustrations showing assay platforms using either polyclonal antibody (A) or aptamers (B) to detect thrombin. Both assays use an identical anti-thrombin monoclonal antibody as the capture antibody and identical reagents for detection (NutAvidin-HRP and TMB). Assays used in Fig. 6 however use a polyclonal antibody as the capture antibody, one of four monoclonal antibodies as detectors and anti-mouse-HRP as a secondary antibody (C) ELISA assay showing the limit of detection for purified human thrombin using the antibody based assay illustrated in (A). (D) ELISA assay showing the limit of detection for purified human thrombin using the aptamer based assay illustrated in (B). doi:10.1371/journal.pone.0031948.g001
in the frequency of inhibitors even though other product parameters (such as specific activity) remained unaffected [26]. It was subsequently determined that the heat treatment likely resulted in subtle alterations in the conformation leading to the exposure of some epitopes [17]. We therefore investigated whether a comparable treatment of the human thrombins would affect the binding of the panel of aptamers used in this study. Evithrom and Recothrom were incubated at 60 °C for a maximum of 17 h in the presence of 60% sucrose buffer as has been described previously (see Materials and Methods and [16] for details). Consistent with previous studies [16] we found that heating in the presence of 60% sucrose afforded protection and there was no significant effect on the specific activity of either product (Fig. 4A). On the other hand incubation at 60 °C in PBS alone resulted in both protein degradation (Fig. 4B) as well as complete abrogation of thrombin activity (data not given). Similarly the antibody based binding assay (see Fig. 1A for assay format) showed no detectable difference in the binding of the polyclonal antibody, to thrombin following up to 17 h of incubation at 60 °C in the presence of 60% sucrose buffer (Fig. 4C). On the other hand the positive control (heat treatment in the presence of PBS alone) results in antibody binding being almost complete abolished. The aptamer based assay shows time-dependent changes in the binding kinetics when the thrombin is incubated in the presence of sucrose (Fig. 4D). We compared the effect of heat treatment of both the plasma derived and recombinant thrombin on the binding of all six aptamers (Fig. 5). The binding of five out of six aptamers showed a statistically significant decrease in binding compared to untreated

**Table 2. Sequences of aptamers used in this study.**

| Aptamer | Sequence |
|---------|----------|
| 30-14   | 5’AGATGCCTGCTGAGCATGCTGTTAGGGTTAGGGATGGTACGGGTAAGACTTTTGTGGCGG’3 |
| 30-16/27| 5’TACCGTGGTGAGGGGAAAGTTGGAGTTGTA’3 |
| 30-38/27| 5’ACCCGTGGTAGGATGGGATGGGATGG’3 |
| 60-18/38| 5’CAGTCCGAGGCTAGGGCTAGGCTAGGCTAAGTGGTGGAA |
| G15D    | 5’GGTGGTGCTCGG’3 |
| 30-8    | 5’AGATGCCTGTGGAGCATGCTGAATAGGTAGGGTGAGATGGTACGGGTAAGACTTTTGTGGCGG’3 |

Figure 2. Affinities of the six aptamers used in this study determined using the aptamer based ELISA. A fixed concentration of thrombin (Evithrom), 150 nM was captured by the anti-thrombin monoclonal antibody, 5 μg/ml. Binding of increasing concentrations of the six anti-thrombin aptamers (A, Aptamer 30-14, B, Aptamer 30-8, C, Aptamer 60-18/38, D, Aptamer 30-16/27, E, Aptamer 30-38/27 and F, Aptamer G15D) was determined using the assay depicted in Fig. 1B. Data presented shows the mean (± SD) of three experiments where the aptamers were generated independently for each experiment. doi:10.1371/journal.pone.0031948.g002
recombinant thrombin. However, the binding of only two aptamers was affected when the plasma derived thrombin preparation was heat treated.

We evaluated four commercially available anti-thrombin monoclonal antibodies for their capacity to distinguish between control and heat-treated thrombin (Fig. 6). In the experiment depicted in Fig. 3B a monoclonal antibody was used to capture thrombin and a polyclonal antibody used as the detector. In the ELISA assays shown in Fig. 6, a sheep anti-human thrombin polyclonal was used to capture the thrombin and a panel of mouse anti-thrombin monoclonal antibodies was used to detect the protein. The monoclonal antibody EST-2 recognizes thrombin alone as well as the thrombin-antithrombin complex [27]. The EST-4 on the other hand binds thrombin but not the thrombin-antithrombin complex and it has been suggested that this monoclonal antibody recognizes an epitope near the active site [27]. Both these antibodies were obtained by the fusion of α-thrombin immunized spleen cells with a myeloma cell line. We also used two additional mouse anti-thrombin antibodies (sc-59717 and AH-5020) [28,29] raised against purified full-length native human thrombin in these assays (Fig. 6C&D). We selected monoclonal antibodies that were raised using protein molecules rather than peptides as these are more likely to be conformation sensitive. However all the four monoclonal antibodies showed comparable binding kinetics for the control and heat treated thrombin. In fact the monoclonal antibody sc-59717 continued to bind denatured thrombin (heated at 60°C for 17 h in PBS).

Using aptamers in label free characterization of the human thrombin products

ELISA based assays are simple, and do not require much specialized equipment or skills and have found wide application in research, clinical laboratories as well as during drug development. These assays however require capture and detection antibodies, are subject to wash steps, multiple incubations, and serial dilutions for each sample; factors that affect precision, accuracy and turn around time. Moreover, in recent years label free technologies have been found to be advantageous over methods where the kinetic constants are derived indirectly and are assay dependent [30]. Several technologies are available that permit the label-free determination of kinetic constants including the association rate constant (kₐ), dissociation rate constant (k₈), and equilibrium dissociation constant (Kₐ). Here we utilized BioLayer Interferometry, to measure in real-time the aptamer-thrombin interactions using streptavidin. In this assay, the biotinylated aptamer is bound to the biosensor and the rate of binding to thrombin was found to be concentration dependent as has been shown previously for protein-protein interactions such as those between an antibody and its ligand [30]. We used this data for base-line correction and to obtain the kₐ and k₈ and Kₐ values using the Octet Software (data not given). Using the recombinant human thrombin, Recothrom as an example we found that the K₈ values obtained in the label-free assay correlate (r = 0.842, P<0.05) with the apparent Kₐ estimated using the ELISA based assay (Fig. 7A). A similar correlation was obtained when we compared the KD values from the label free assay with the Kₐ values from the ELISA based assay (data not given). The advantage of using the k₈ values is that while the apparent Kₐ and KD determinations are concentration dependent and require accurate estimations of protein and nucleotide the k₈ is independent of the concentration of the ligand. This can be useful as the concentration of samples can change during processing. The heat treatment described in the

| Table 3. Kinetic parameters for the binding of aptamers to Recothrom. |
|-------------------|-------------------|
| Aptamer           | Vₐₚₜₓ             | Apparent Kₐ |
| 60-18/38          | 1.58±0.01         | 1.48±0.05   |
| 30-14             | 1.38±0.05         | 2.75±0.36   |
| 30-8              | 1.29±0.01         | 0.58±0.01   |
| 30-16/27          | 0.819±0.0196      | 1.54±0.42   |
| 30-38/27          | 0.18±0.03         | 1.28±0.03   |
| G15D              | 0.68±0.02         | 1.48±0.15   |

Figure 3. Comparing three topical thrombin products using binding assays with aptamers as detector reagents in an ELISA format. (A) The three topical thrombin products were electrophoresed on a 12% NuPAGE gel and stained using Colloidal Blue. The thrombin and HSA (used as an exipient) bands are indicated with red and blue arrows respectively. (B) Kinetics of binding of the Aptamer 30-8 to the purified human thrombin products Recothrom (squares) and Evithrom (circles). (C) The affinities (apparent Kₐ) of all six aptamers to both Recothrom and Evithrom were determined as shown in (B). The apparent Kₐ values for Recothrom (X-axis) were plotted against those for Evithrom (Y-axis) and were shown to be highly correlated (r = 0.996, P<0.05). doi:10.1371/journal.pone.0031948.g003
previous section involves dilution and incubation in the 60% sucrose medium. We therefore compared the change in the dissociation rate constants (k_{dis}) for the six aptamers following heat treatment of Recothrom (Fig. 7B). A statistically significant change in the k_{dis} is observed in 4 of the 6 aptamers as a consequence of heat treatment, consistent with our results using the ELISA based assay (Fig. 5).

Discussion

In this study we characterize aptamers as surrogate antibodies for the identification of changes in the conformation of a protein during the manufacturing process and during storage. This is important because the inability to detect changes in the conformation of a protein-product can have consequences even if the activity is unaffected. For example even subtle changes in conformation can result in new epitopes on the protein, leading to immunogenicity [31]. Immunogenicity is currently a significant impediment to the successful development of therapeutic proteins, as the development of inhibitory antibodies can reduce the efficacy and sometimes even result in life threatening safety issues [32]. Similarly, comparison of biosimilars to reference standards using bioanalytical techniques is a key component of the licensure of such products [5]. Here we have used a panel of aptamers to compare three different thrombin products vis-à-vis the conformational epitopes.

The affinity of a panel of six anti-thrombin aptamers for the protein was determined using an ELISA like assay which is illustrated in Fig. 1A. A single monoclonal antibody was used to capture the thrombin molecules which were then detected using either a polyclonal anti-thrombin antibody or one of the six anti-thrombin aptamers. In another set of experiments, the anti-thrombin polyclonal antibody was used to capture the thrombin molecules and the binding kinetics of four anti-thrombin monoclonal antibodies were evaluated. By standardizing the assay we were able to obtain excellent signal to noise ratio for all six aptamers, comparable to that for the antibody based assay. Moreover the limit of detection for thrombin is also comparable regardless of whether an aptamer or antibody was used and we obtained consistent results for binding kinetics where the percent CV ranged from 9 to 14% with an average of 12.4% (Table 3).

Both the polyclonal detector antibody and all six aptamers bound the Evithrom with high affinity (Fig. 2 & 3B) but showed
almost no binding to Thrombin JMI. Both these thrombin products are purified from plasma but the former is of human origin and the latter of bovine origin. This demonstrates significant specificity as the bovine and human thrombins are 89% homologous and show 81% sequence identity [33]. Moreover in bioassays as well as clinical studies the bovine and human thrombins have been shown to exhibit comparable activity. A comparison of the binding affinities of the aptamers for Evithrom and Recothrom (Fig. 3C) showed a very high correlation (r = 0.996, P < 0.05). Evithrom and Recothrom are both human thrombin products, however while the former is purified from plasma the latter is a recombinant protein expressed in CHO cells [22,23]. This result would suggest that these two products while obtained by two very different processes have remarkably similar conformations given that the aptamers are extremely sensitive to subtle changes (Figs. 4D, 5 & 7B).

We have alluded to a case study from the 1990s where an additional step of pasteurization was used to inactivate non-lipid-enveloped viruses in the manufacture of FVIII led to the development of inhibitory antibodies at a very high frequency leading to the withdrawal of the product from the market [26]. Subsequent studies have suggested that the change in the manufacturing process exposed sites in the C2 domain of the FVIII leading to immunogenicity [17]. This case vividly illustrates the significance of the need for techniques that can routinely detect subtle changes in conformation because not all changes affect product activity in in vitro assays. When we used a similar heat-treatment for the two human thrombin products we found no significant change in activity even after 17 h (Fig. 4A). Nor could assays using the antibody-based ELISA platform detect any changes in the affinity of the anti-thrombin antibody (Figs. 4C & 6) following heat-treatment when the protein is protected with 60% sucrose. As a control we demonstrate that heat-treatment in the absence of sucrose completely abolishes antibody binding (Fig. 4C & 6) in all but one of the antibodies screened. Aptamers on the other hand show small but significant differences in binding during heat-treatment of Recothrom in the presence of 60% sucrose and these changes are time dependent (Fig. 4D). It is interesting to note (a) that the magnitude of change varies for different aptamers and (b) the binding of only some aptamers is affected (Fig. 5A). This suggests that the aptamers recognize different epitopes or conformations on the protein. We obtained similar results in a label-free platform using BioLayer Interferometry (Fig. 7B). This panel of aptamers also showed alterations in binding to Evithrom following heat treatment, except that the binding of fewer aptamers is affected (Fig. 5B). It is noteworthy that all the aptamers affected by the heat-treatment of Evithrom are also affected in Recothrom. It is plausible that the high concentration of HSA in the Evithrom provides additional protection to the protein during heat treatment. Taken together these results suggest that a suitable panel of aptamers can be used to detect small changes in protein conformation not detectable by other methods.

Figure 5. Heat treatment of human thrombins in the absence and presence of 60% sucrose. The two purified human topical thrombin products were heated as described in Fig. 3. Control samples were heated in for 17 h at 60°C in PBS (in the absence of 60% sucrose). Binding of either the antibody or each of the six aptamers was monitored to (A) Recothrom or (B) Evithrom at time 0 and 17 h. The percent of the binding at time 0 (mean ± SD, n = 3) is represented. The aptamers used are shown on the figure.

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We used a panel of six aptamers in this study; the nature, number and choice of antibodies/aptamers is likely to be an important factor in the success of epitope mapping for detecting subtle changes in the conformation of individual protein products. Depending on the complexity of a therapeutic protein the panel of aptamers used to map a protein can readily be expanded, the same is not true of antibodies. In addition protein-specific aptamers are selected from a very large pool of conformations [34–36]. Consequently a larger repertoire of aptamers representing more epitopes can be generated for a target protein. Aptamers are also likely to be more useful in a quality control environment [37–41]. In addition, we determined in this study that the presence of excipients (including proteins other than the active ingredient) has no significant effect on the binding of aptamers to the thrombins. This observation could potentially be of consequence in comparing biosimilars to their reference standards. The primary hurdle for the developer of any biosimilar is that there is normally no direct access to originator companies’ proprietary data. Thus, the developer of a biosimilar has to retrieve the reference protein as a finished drug product, purify the drug substance and reverse engineer the process [42]. This in turn has raised concerns that the reverse engineered ‘reference standard’ could itself be altered from the original active ingredient manufactured by the innovator. As aptamer binding is unaffected by the excipients, the assays described here provide a means of comparing the active ingredient in the finished drug product with the reverse engineered reference standard.

This study demonstrates that aptamers can be used to characterize changes in the tertiary structure of proteins. This has potential applications as an analytical test during the development of therapeutic proteins and to monitor the effect of manufacturing changes and storage. A range of biochemical and biophysical tests are available to characterize protein-drugs, however analytical methods to rapidly and routinely monitor subtle changes in the tertiary structure represent a critical lacuna. The need for such methods is particularly acute in the development of biosimilars. Here we have provided proof-of-principle that aptamers are stable reagents that can be used to design sensitive and robust analytical tests to detect small changes in the conformation of therapeutic proteins.
Materials and Methods

Materials

Recombinant human thrombin was from Zymogenetics (Seattle, WA), and plasma-derived human thrombin was from Omrix (Israel). Aptamers were synthesized by the standard phosphoramidite method on an ABI model 394 oligonucleotide synthesizer (Facility for Biological Research, CBER, FDA), purified by reversed phase HPLC and lyophilized. Aptamers were used either unmodified or in the biotinylated forms as indicated in the text. Table 2 lists the aptamers used in the study.

Antibody based binding assay

ELISA plates (MAxisorp, Nunc) were coated with 50 μl mouse anti-human thrombin monoclonal antibody (Hemtech AHT-5020) diluted to 3.9 μg/ml in coating buffer (50 mM Na2CO3, pH 9.6), and incubated overnight at 4°C. Plates were washed in PBST and blocked for 1 hr at 37°C with 300 μl per well of 2% (W/V) BSA in Tris-NaCl pH 7.6 (TSTA). After washing, plates were incubated with 5 μg/ml of the indicated thrombin preparation diluted in PBST (50 μl, in duplicate) for 1 hr at 37°C. Plates were washed 4 times with PBST and incubated with biotinylated sheep anti-human thrombin polyclonal antibody (50 μl, Hemtech PAHT-S-B) diluted 1:10000 in dilution buffer for 1 hr at RT. After washing, plates were incubated with 50 μl of HRP-conjugated anti-mouse antibody (Rockland) diluted 1:10000 in PBST for 1 hr at 37°C. Finally, plates were washed 4 times with PBST, and the chromogenic reaction was started using 100 μl TMB. Reaction was stopped with 100 μl 0.25 M H2SO4 and absorbance was measured at 450 nm.

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Figure 7. Estimating the binding of aptamers to thrombin in a label free assay. The binding of aptamers to Recothrom was monitored in real time using BLI and the kinetic parameters obtained using the Octet data analysis software version 7.0. (A) Correlation between Kds values obtained in the label-free assay with the apparent Kds estimated using the ELISA based assay (r = 0.842, P<0.05). (B) Effect of heat-treatment of human thrombins on the kdis for all 6 aptamers. Recothrom was heat-treated as described in Fig. 3 and the kdis determined following heat treatment for 0 and 17 h. The percent of the binding at time 0 is represented. Data presented is the mean (± SD) of three independent experiments.

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Stability Assay

Recombinant and plasma derived thrombin were diluted to 30 μg/ml in sucrose buffer (60% w/v sucrose, 0.1 M glycine, pH 7.5). Control samples were diluted accordingly in PBS. Samples (1.4 ml) were incubated at 60°C for 0, 1 and 17 hr and maintained at −80°C until analyzed.

Activity assay

Two-fold dilution of calibrator thrombin (25 μl, Thrombinscope, Netherlands) in the range of 1000-0.1 nM or unknown samples were added to 96-well plates pre-loaded with streptavidin (SA) biosensor tips (Fortebio, Inc., Menlo Park, CA, USA). The assays were maintained at a temperature of and the speed of 30°C and 1000 rpm respectively. Streptavidin-coated biosensor tips were pre-wet for 15 min. Then the tips were loaded with 250 nm–500 nM of biotinylated aptamer for 5 min. The typical resulting capture levels of eight biosensors were in the range of 0.4–0.5 nm.

Determining the Binding Affinity of aptamers using Bio-Layer Interferometry

Bio-Layer Interferometry (BLI), a label-free technology, was used for measuring the binding of aptamers with thrombin samples. The affinity measurements were performed with Fortebio Octet RED96 equipped with streptavidin (SA) biosensor tips (ForteBio, Inc., Menlo Park, CA, USA). The assays were monitored continuously at 37°C by automatic reading every 30 sec up to 30 min using fluorescence reader (Tecan) with an excitation wavelength of 380 nm and an emission wavelength of 430 nm. Thrombin concentrations were expressed as the rate development of fluorescence intensity [arbitrary fluorescence units (FU)], calculated for every reading (FU min⁻¹).

The association (k_on) and dissociation (k_off) were then established by dipping the biosensors for 10 mins in various concentrations of thrombin samples dispensed in 96-microwell plates (Fisher Scientific, Turnbury Drive, Hanover Park, IL, USA) at a volume of 200 μl per well. Data were processed and analysed using the Octet data analysis software version 7.0 (ForteBio). The binding profile of each aptamer was shown as “nm” shift. This shift is a comparison of the shift in the interference patterns of light reflected from a reference layer within the biosensor versus molecules as the bind to the biosensor tip. The results were summarized as KD which was calculated from “KD = k_on/k_off”, where k_on is the ‘on rate’ or association and k_off is the ‘off rate’ or dissociation.

Statistical analysis

Stability experiment was conducted in three independent repeats. Statistical analysis was carried out using repeated analysis of variance (ANOVA) performed with GraphPad InStat 3 software. Differences were considered significant when P was <0.05.

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Disclaimer

The findings and conclusions in this article have not been formally disseminated by the Food and Drug Administration and should not be construed to represent any Agency determination or policy.

Author Contributions

Conceived and designed the experiments: RZ BG ZES. Performed the experiments: RZ WC GSP. Analyzed the data: RZ WC ZES. Wrote the paper: RZ ZES.

References

1. Beley MJ, Harris LM, Das RR, Cherkow J (2006) Biesimilars: initial excitement gives way to reality. Nat Rev Drug Discov 5: 535–536.
2. Rathore AS, Ford NE, Steiner W (1997) Analyte specific aptamers: issues, developments, and challenges. Trends Biotechnol 25: 698–705.
3. Rathore AS, Winkle H (2009) Quality by design for biopharmaceuticals. Nat Biotechnol 27: 26–34.
4. Schellekens H (2004) When biotech proteins go off-patent. Trends Biotechnol 22: 406–410.
5. Anonymous (2006) Guidelines on similar biological medicinal products containing biotechnology-derived proteins as active substance. European Medicines Agency.
6. Kozloski S, Woodcock J, Midthun K, Sherman RR (2011) Developing the nation’s biosimilars program. N Engl J Med 365: 385–388.
7. Schellekens H, Moores E (2010) Clinical comparability and European biosimilar regulations. Nat Biotechnol 28: 20–31.
8. Roger SD (2010) Biosimilars: current status and future directions. Expert Opin Biol Ther 10: 1011–1018.
9. Chirino AJ, Mire-Shiah A (2004) Characterizing biological products and assessing comparability following manufacturing changes. Nat Biotechnol 22: 1303–1391.
10. Wanebo J, Lohfeld J, Hyden B, Uhlen M, Stahl S (2000) Octet mapping of antibodies using bacterial surface display. Nat Methods 5: 1001–1005.
11. Barlow DJ, Edwards MS, Thornton JM (1986) Continuous and discontinuous protein antigenic determinants. Nature 322: 747–748.
12. Coates RE, Corte E, Trumonao A (1995) Mapping epitopes on protein surfaces. Biopolymers 36: 675–680.
13. Tombelli S, Mascini M (2009) Aptamers as molecular tools for bioanalytical characterization. Comb Chem High Throughput Screen 12: 791–800.
31. Schellekens H (2005) Factors influencing the immunogenicity of therapeutic proteins. Nephrol Dial Transplant 20: 3–9.
32. Shankar G, Pendley C, Stein KE (2007) A risk-based bioanalytical strategy for the assessment of antibody immune responses against biological drugs. Nat Biotechnol 25: 553–561.
33. Schoenecker JG, Johnson RK, Fields RG, Lesher AP, Domzalski T, et al. (2003) Relative purity of thrombin-based hemostatic agents used in surgery. J Am Coll Surg 197: 580–590.
34. Carothers JM, Oestruch SC, Davis JH, Szostak JW (2004) Informational complexity and functional activity of RNA structures. J Am Chem Soc 126: 5130–5137.
35. Ruigrok VJ, Levisson M, Eppink MH, Smidt H, van der Oost J (2011) Alternative affinity tools: more attractive than antibodies? Biochem J 436: 1–13.
36. Jhaveri S, Rajendran M, Ellington AD (2000) In vitro selection of signaling aptamers. Nat Biotechnol 18: 1293–1297.
37. Iliuk AB, Hu L, Tao WA (2011) Aptamer in bioanalytical applications. Anal Chem 83: 4440–4452.
38. Soukup GA, Breakey RR (1999) Nucleic acid molecular switches. Trends Biotechnol 17: 469–476.
39. Famulok M (2003) Allosteric aptamers and aptazymes as probes for screening approaches. Curr Opin Mol Ther 7: 137–143.
40. Rentmeister A, Mayer G, Kuhn N, Famulok M (2007) Conformational changes in the expression domain of the Escherichia coli thiM riboswitch. Nucleic Acids Res 35: 5713–5722.
41. Vinkenborg JL, Karnowski N, Famulok M (2011) Aptamers for allosteric regulation. Nat Chem Biol 7: 519–527.
42. Schneider CK, Kalinke U (2008) Toward biosimilar monoclonal antibodies. Nat Biotechnol 26: 985–990.