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Kdo Hydrolase Is Required for Francisella tularensis Virulence and Evasion of TLR2-Mediated Innate Immunity

Nihal A. Okan, Sabina Chalabaev, Tae-Hyun Kim, Avner Fink, Robin A. Ross, Dennis L. Kasper

Department of Microbiology and Immunobiology, Harvard Medical School, Boston, Massachusetts, USA

N.A.O. and S.C. contributed equally in this work.

ABSTRACT The highly virulent Francisella tularensis subsp. tularensis has been classified as a category A bioterrorism agent. A live vaccine strain (LVS) has been developed but remains unlicensed in the United States because of an incomplete understanding of its attenuation. Lipopolysaccharide (LPS) modification is a common strategy employed by bacterial pathogens to avoid innate immunity. A novel modification enzyme has recently been identified in F. tularensis and Helicobacter pylori. This enzyme, a two-component Kdo (3-deoxy-D-manno-octulosonic acid) hydrolase, catalyzes the removal of a side chain Kdo sugar from LPS precursors. The biological significance of this modification has not yet been studied. To address the role of the two-component Kdo hydrolase KdhAB in F. tularensis pathogenesis, a ΔkdhAB deletion mutant was constructed from the LVS strain. In intranasal infection of mice, the ΔkdhAB mutant strain had a 50% lethal dose (LD50) 2 log10 units higher than that of the parental LVS strain. The levels of the proinflammatory cytokines tumor necrosis factor alpha (TNF-α) and interleukin-1β (IL-1β) in bronchoalveolar lavage fluid were significantly higher (2-fold) in mice infected with the ΔkdhAB mutant than in mice infected with LVS. In vitro stimulation of bone marrow-derived macrophages with the ΔkdhAB mutant induced higher levels of TNF-α and IL-1β in a TLR2-dependent manner. In addition, TLR2−/− mice were more susceptible than wild-type mice to ΔkdhAB bacterial infection. Finally, immunization of mice with ΔkdhAB bacteria elicited a high level of protection against the highly virulent F. tularensis subsp. tularensis strain Schu S4. These findings suggest an important role for the Francisella Kdo hydrolase system in virulence and offer a novel mutant as a candidate vaccine.

IMPORTANCE The first line of defense against a bacterial pathogen is innate immunity, which slows the progress of infection and allows time for adaptive immunity to develop. Some bacterial pathogens, such as Francisella tularensis, suppress the early innate immune response, killing the host before adaptive immunity can mature. To avoid an innate immune response, F. tularensis enzymatically modifies its lipopolysaccharide (LPS). A novel LPS modification—Kdo (3-deoxy-D-manno-octulosonic acid) saccharide removal—has recently been reported in F. tularensis. We found that the ΔkdhAB mutant was significantly attenuated in mice. Additionally, the mutant strain induced an early innate immune response in mice both in vitro and in vivo. Immunization of mice with this mutant provided protection against the highly virulent F. tularensis subsp. tularensis strain Schu S4. Thus, our study has identified a novel LPS modification important for microbial virulence. A mutant lacking this modification may be used as a live attenuated vaccine against tularemia.

Francisella tularensis, one of the deadliest respiratory pathogens in existence, is considered a potential biological weapon because it is readily aerosolized and exhibits a high degree of infectivity and lethality in humans. An attenuated live vaccine strain (LVS) has been developed but has not been licensed as a vaccine because of an incomplete understanding of the basis for its attenuated virulence and associated side effects. There has been a widespread research effort aimed at elucidating Francisella pathogenesis and identifying components for rational vaccine design. F. tularensis is a successful pathogen that can evade and suppress innate immune responses. After intranasal infection of mice, F. tularensis initially colonizes the lung, infecting several cell types but failing to stimulate proinflammatory cytokine production for the first 2 to 3 days (1–5). This initial delay in inflammation contributes to the ability of F. tularensis to replicate in the lung, spread into systemic organs, and cause sepsis leading to the host’s death (4). To suppress the early proinflammatory cytokine response, F. tularensis employs a variety of strategies, such as avoiding disclosure of itself to the innate immune system (6, 7).

Lipopolysaccharide (LPS) of F. tularensis plays a significant role in evasion of host immune responses (8–11). LPS, the major component of Gram-negative bacterial outer membranes (12, 13), itself has three components: (i) lipid A, a glucosamine-based glycolipid; (ii) a core oligosaccharide ketosidically linked to lipid A by an eight-carbon sugar, 3-deoxy-D-manno-octulosonic acid (Kdo); and (iii) the O-antigen polysaccharide comprising mult-
ple repeating units of a complex tetrasaccharide. The lipid A of many bacterial species such as *Escherichia coli* is a powerful stimulant of Toll-like receptor 4 (TLR4)-mediated innate immune responses. In contrast, *F. tularensis* lipid A triggers no TLR-mediated innate response because of a number of key structural modifications (9); specifically, despite initial similarity to *E. coli* lipid A, the lipid A precursor of *F. tularensis* undergoes dephosphorylation, deacetylation, and carbohydrate incorporation that render the molecule undetectable by the TLR4 receptor. Differences in the length of the fatty acid chain of lipid A (16 to 18 carbons for parental *LVS* as opposed to 12 to 14 carbons for *E. coli*) further enhance the ability of *F. tularensis* to evade TLR4-mediated innate immunity. These modifications contribute to bacterial virulence (8, 10).

A novel LPS-modifying two-component KdhAB Kdo hydrolase enzyme complex has recently been identified in *F. tularensis* (14, 15). A core LPS precursor containing two Kdo residues is first synthesized. One Kdo in the backbone of the core polysaccharide is ketosidically linked to lipid A and glycosidically linked to the remainder of the core polysaccharide, and the second Kdo residue is glycosidically attached as a side chain. KdhAB removes the side chain Kdo, leaving an LPS molecule containing only one Kdo residue (Fig. 1) (14–16). The KdhAB Kdo hydrolase is a two-component enzyme composed of a sialidase-like protein (the catalytic subunit) and a small inner membrane protein (perhaps a membrane-anchoring subunit) (15). This novel enzyme complex has been reported in two other pathogens, *Helicobacter pylori* and *Legionella pneumophila* (14, 17, 18). Its biological significance—in particular, its role in pathogenicity—has not yet been investigated. Here, we show that the *F. tularensis* ΔkdhAB LVS mutant is attenuated in a mouse model of infection and stimulates—a TLR2-dependent manner—a stronger proinflammatory cytokine response than the parental LVS. Furthermore, immunization of mice with the ΔkdhAB mutant provides significant protection against fully virulent *F. tularensis* type A strain Schu S4.

**RESULTS**

The *F. tularensis* LVS ΔkdhAB mutant is attenuated in mice. To fully evaluate the importance of the KdhAB Kdo hydrolase in the virulence of *F. tularensis* LVS, *kdhA* and *kdhB* genes were deleted by homologous recombination in an LVS background, giving rise to the ΔkdhAB mutant. Previously described analysis by high-performance anion-exchange chromatography (HPAEC) (14) confirmed that, unlike the LPS of parental LVS, which has a single Kdo residue, the LPS of the ΔkdhAB mutant has two Kdo residues (Fig. 1; see Table S1 in the supplemental material).

To evaluate the role of Kdo hydrolase activity in *F. tularensis* virulence, we infected BALB/cByJ mice with graded doses of LVS or the ΔkdhAB mutant via the intranasal route. All mice challenged with 105 CFU of LVS succumbed to the infection within 9 days (Fig. 2A). In contrast, all mice challenged with a similar dose of the ΔkdhAB mutant survived (Fig. 2A). At a dose of 103 CFU of the ΔkdhAB mutant, 17% of the infected mice survived (Fig. 2A).

The observed phenotypes in the ΔkdhAB mutant were successfully complemented with the pkdAB plasmid carrying the *kdhA* and *kdhB* genes. LPS analysis showed that the complemented strain had only a single Kdo residue in its core LPS (see Table S1 in the supplemental material). In addition, complementation of ΔkdhAB restored virulence to the wild-type (WT) level. The survival rates of BALB/cByJ mice (10 mice in each group) 28 days after an intranasal challenge with 5 × 104 *F. tularensis* LVS or a ΔkdhAB mutant harboring either a vector control (pFNLTP6) or a plasmid carrying *kdhA* and *kdhB* genes (pkdAB) were as follows: 0% for mice infected with LVS, 100% for mice infected with the ΔkdhAB mutant carrying pFNLTP6, and 20% for mice infected with the ΔkdhAB mutant carrying pkdAB. These data indicate that Kdo hydrolase activity is important for the virulence of *F. tularensis* LVS.

We also examined the ability of the ΔkdhAB mutant to colonize the lung and to disseminate into other organs after intranasal challenge of mice. Six days after infection with 106 bacteria, both parental LVS and its ΔkdhAB mutant had reached a similar level of colonization in the lungs, with counts of 3 × 106 CFU/g and 107 CFU/g, respectively (Fig. 2B). However, the numbers of ΔkdhAB mutant bacteria in spleen, liver, and blood were 1.5 to 3 logarithmic units lower than the numbers of LVS bacteria (Fig. 2B) (Student’s *t* test, *P < 0.05). Overall, these results indicate that the ΔkdhAB mutant is deficient in its ability to disseminate systemically.

To determine whether the observed defect in dissemination is related to bacterial growth kinetics, growth rates were compared under different conditions. There was no difference in growth between LVS and the ΔkdhAB mutant when bacteria were cultured in broth medium or in J774A.1 macrophage-like and MH-S alveolar macrophage-like cell lines (see Fig. S1A and S1B in the supplemental material). Moreover, the ΔkdhAB mutant was not sensitive to complement-mediated killing (Fig. S1C). Therefore, the defect of the ΔkdhAB mutant in dissemination to systemic organs was not due to a growth defect or to serum sensitivity.

The cytokine response to the *F. tularensis* LVS ΔkdhAB mutant is enhanced. Within the first day after infection, mutants of *F. tularensis* that cannot modify their lipid A elicit a stronger proinflammatory response than parental LVS did (8, 19). Since the KdhAB hydrolase is involved in the modification of the LPS core oligosaccharide but not in the modification of lipid A, we compared the proinflammatory responses to LVS and the ΔkdhAB mutant. To this end, mice were challenged intranasally with 103 CFU of either LVS or the ΔkdhAB mutant, and the levels of tumor necrosis factor alpha (TNF-α) and interleukin-1β (IL-1β) in samples of bronchoalveolar lavage (BAL) fluid were determined in an enzyme-linked immunosorbent assay (ELISA). Consistent with previous reports (6), the levels of TNF-α and IL-1β in...
Characterization of kdhAB in Francisella tularensis

BAL fluid 24 h after inoculation of LVS bacteria were not different from those in control mice given phosphate-buffered saline (PBS) (Fig. 3A, left panel). In contrast, the levels of TNF-α and IL-1β were significantly higher (~2-fold) in mice infected by the ΔkdhAB mutant than in mice infected by LVS or control mice given PBS (Fig. 3A, left panel). The stronger proinflammatory response was not due to differences in bacterial burden, as the numbers of ΔkdhAB and LVS bacteria in the lungs were similar (Fig. 3A, right panel). Therefore, in contrast to LVS, the ΔkdhAB mutant elicited an early proinflammatory response that was detected 24 h after infection.

To test whether the in vivo cytokine response also differs in ex vivo macrophages, bone marrow-derived macrophages (BMMs) from C57BL/6J mice were infected at different multiplicities of infection (MOIs) with either LVS or the ΔkdhAB mutant. Supernatants were collected after 24 h, and secretion of TNF-α and IL-1β was measured by ELISA. At all MOIs tested, TNF-α secretion by BMMs stimulated by the ΔkdhAB mutant was higher by a factor of at least 2 than that by BMMs stimulated by LVS (Fig. 3B, left panel). Results were similar for IL-1β secretion (Fig. 3B, right panel). Consistent with the cytokine secretion profile, the expression of TNF-α and IL-1β genes was higher in ΔkdhAB mutant-stimulated BMMs than in LVS-stimulated BMMs (Fig. 3C).

The Francisella tularensis LVS ΔkdhAB mutant activates the TLR2 signaling pathway. To investigate the basis for the enhanced innate immune response to the ΔkdhAB mutant, we assessed the responses of murine macrophage-like cell lines in which various TLR signaling pathways were inactivated. Cells were infected with either LVS or the ΔkdhAB mutant at an MOI of 100 and lysed 24 h later for measurement of IL-1β. Cell activation was calculated as the IL-1β ratio in infected cells versus uninfected cells. LVS bacteria elicited a proinflammatory response in WT, Unc93B−/−, and TLR4−/− cells, but not in MyD88−/− or TLR2−/− cells (Fig. 3A). These data are in agreement with previous reports that the immune response to LVS is mediated through TLR2 signaling (3, 20). Interestingly, the ΔkdhAB mutant showed a similar pattern of activation, a result indicating that TLR2—but not TLR3, -4, -7, or -9—mediates the innate immune response to ΔkdhAB mutant bacteria. Moreover, the proinflammatory response was significantly greater in ΔkdhAB mutant-infected cells than in LVS-infected cells (Fig. 4A). Control experiments confirmed that WT cells and knockout cells for Unc93B (required for TLR3, TLR7, and TLR9 signaling), MyD88, TLR4, and TLR2 responded to various TLR agonists as expected (data not shown).

To assess whether ΔkdhAB mutant-mediated enhancement of the proinflammatory cytokine response is likely to be due to an increase in nuclear factor κB (NF-κB) activation, we used HEK293 cells expressing TLR2 (HEK-TLR2) or TLR4 and MD2 (HEK-TLR4/MD2). The stable cell lines were transiently transfected with the reporter plasmid pNFκB-luc, which expresses a firefly luciferase gene under control of an NF-κB promoter, and were incubated with LVS or ΔkdhAB organisms. Reporter activity in HEK-TLR4/MD2 cells incubated with bacteria of either strain did not differ (Fig. 4B). On the other hand, reporter activity increased slightly but significantly in HEK-TLR2 cells stimulated with the ΔkdhAB mutant over that in HEK-TLR2 cells stimulated with ΔkdhAB mutant alone does not trigger TLR2 signaling (9). To test this hypothesis, we assessed purified LPS preparations from each strain for their ability to activate NF-κB in HEK-TLR2 cells. LPS from LVS and the ΔkdhAB mutant showed similar levels of NF-κB reporter activity, which was also similar to that elicited by whole LVS organisms (Fig. 4B). Additionally, BMMs did not respond to LPS from either LVS or the ΔkdhAB mutant by secretion of TNF-α and IL-1β when the purified LPS was tested at concentrations of 50, 250, 1,250, and 10,000 ng/ml (see Fig. S2 in the supplemental material). Taken together, these results show that the ΔkdhAB mutant elicits stronger NF-κB activation in a TLR2-dependent manner, with a consequently greater proinflammatory response. However, the greater proinflammatory response induced by the mutant LPS probably is not due to activation of TLR2 by the mutant LPS per se, as the LPS of the ΔkdhAB mutant alone does not trigger TLR2 signaling in vitro.

To further investigate the mechanism for enhanced TLR2 signaling by the ΔkdhAB mutant, we evaluated bacterial membrane integrity. A leaky cell membrane caused by ΔkdhAB mutation would potentially release TLR2 ligands into the culture supernatant to be detected by host cells. Thus, the growth behaviors of LVS and the ΔkdhAB mutant were tested in the presence of SDS and EDTA, agents that inhibit growth of bacterial cells with defective outer membranes. The similar growth characteristics of the two
Enhanced proinflammatory cytokine response to the \textit{F. tularensis} LVS \textit{ΔkdhAB} mutant strain. (A) BALB/cByJ mice \((n = 3)\) were challenged intranasally with LVS bacteria \((10^3 \text{ CFU})\), \textit{ΔkdhAB} bacteria \((10^3 \text{ CFU})\), or PBS (control). BAL fluid was collected 24 h after infection, and the levels of TNF-\(\alpha\) and IL-1\(\beta\) were measured by ELISA (left panel). The bacterial burden in lung was measured 24 h after infection (right panel). (B) Levels of TNF-\(\alpha\) and IL-1\(\beta\) released by \textit{ex vivo} BMMs after stimulation with LVS or \textit{ΔkdhAB} bacteria (MOI of 10, 20, or 50). Culture supernatants were analyzed by ELISA 24 h after infection. (C) TNF-\(\alpha\) and IL-1\(\beta\) gene expression in BMMs infected with LVS or \textit{ΔkdhAB} bacteria (MOI of 10). Gene expression was measured by real-time PCR, normalized with RP II gene expression, and reported as relative expression compared with that in uninfected macrophages. All data are presented as mean plus SD (error bars) values \((n = 3)\).
have similar lipid A mass values (Fig. 6B). Although the ΔkdhAB mutation does not affect the total LPS level or the characteristic banding pattern of LPS, we thought it is possible that the additional Kdo—a negatively charged sugar—in the ΔkdhAB mutant alters the conformation of LPS molecules, thereby making other surface constituents more accessible to TLR2 receptors. To test this hypothesis, we evaluated the binding of FopA antibodies to the FopA surface protein in vitro. Live bacteria were incubated with mouse antibodies to FopA, and surface-bound antibodies were analyzed by Western blotting. Consistent with a previous study (24), fully formed LPS significantly reduced the binding of antibodies to the LVS bacterial surface, whereas ΔwbtA mutant LPS, which is devoid of O polysaccharide, did not (Fig. 5B). Strikingly, substantial binding of FopA antibodies to the ΔkdhAB mutant bacterial surface was found, and this result suggested that a TLR2 ligand present on the bacterial cell surface would be potentially exposed for detection by host immune-cell TLR2 receptors. The accessibility of surface proteins might well explain the enhanced TLR2 signaling activity caused by the ΔkdhAB mutation.

**Biological impact of TLR2 activation by the ΔkdhAB mutant.** We next addressed the biological importance of the TLR2-mediated response to ΔkdhAB bacteria in a mouse infection model. WT C57BL/6, TLR2−/−, and TLR4−/− mice were infected with various doses of the ΔkdhAB mutant via the intranasal route and monitored for survival. Most strikingly, at a dose of 10^4 CFU, all WT and TLR4−/− mice survived, while no TLR2−/− mice survived (Table 1). The results suggest that TLR2-mediated responses play a significant role in the survival of mice infected with the *F. tularensis* ΔkdhAB mutant strain. Thus, the association of Kd-hAB Kdo hydrolase with enhanced TLR2-mediated inflammation points to a critical role for this enzyme complex in the virulence of LVS.

**Immunization with *F. tularensis* LVS ΔkdhAB mutant protects against the highly virulent *F. tularensis* Schu S4 strain.** Immunization with LVS is protective against the fully virulent *F. tularensis* type A strain Schu S4 in a mouse model of infection, and this protection involves TLR2-dependent innate immunity (25–27). However, even in low numbers, LVS retains significant virulence in mice. Our findings that the ΔkdhAB mutant is further attenuated and elicits enhanced TLR2 signaling led us to investigate the protective capacity of this mutant strain as a live cell vaccine. To this end, BALB/cByJ mice were immunized with three intranasal doses of LVS, the ΔkdhAB mutant, or (as a control) PBS. Four weeks after the last dose, the mice were challenged via the intranasal route with either LVS at a dose of 10^4 CFU (10 × LD50) or the Schu S4 strain at a dose of 10 CFU (10 × LD50). All mice that had been immunized with LVS survived LVS challenge, whereas all control mice that had received PBS succumbed within 9 days (Fig. 7A). The ΔkdhAB mutant was also fully protective against LVS challenge (Fig. 7A). Furthermore, as expected, LVS-immunized mice survived the fully virulent Schu S4 challenge, while all PBS-treated control mice succumbed within 7 days (Fig. 7B). Strikingly, mice immunized with the ΔkdhAB mutant were well protected; 90% of these mice survived an otherwise lethal challenge with Schu S4. Histologic examination of the livers from infected mice revealed that ΔkdhAB mutant-immunized mice had much less tissue necrosis after challenge with Schu S4 (Fig. 7C) than did PBS-treated control mice. The protection conferred by the ΔkdhAB mutant was also reflected by lower bacterial loads in the livers, spleens, and lungs from mutant-immunized
mice (Fig. 7D) than in these tissues from PBS-treated control mice. The Schu S4 bacterial loads in organs were similar in LVS- and ΔkdhAB mutant-immunized mice. Finally, the levels of Francisella-specific IgG antibody were slightly higher in the sera of ΔkdhAB mutant-immunized mice compared to LVS-immunized mice (Fig. 7E).

**DISCUSSION**

Evasion of innate immunity is a key virulence factor for a number of pathogens, including *F. tularensis* (6, 7). To avoid detection by pattern recognition receptors such as TLRs, *F. tularensis* modifies its lipid A through the action of enzymes such as 4′-phosphatase, galactosamine transferase, and mannosyl transferase (8, 10). A novel LPS modification was hypothesized in *F. tularensis* and *H. pylori*; the removal of side chain Kdo from Kdo2-lipid A precursors by a Kdo hydrolase (16, 17). Such a modification was recently confirmed, and the genes coding for a two-component Kdo hydrolase were identified in *F. tularensis* and *H. pylori* (14, 15, 17, 18). The biological significance of the Kdo hydrolase, which is also expressed by *L. pneumophila* (14), remained to be addressed. By screening an *F. tularensis* subsp. *novicida* transposon mutant library, Weiss and colleagues showed that a kdhB transposon mutant was attenuated in mice, but no further characterization has been reported (28). Here we demonstrate that the two-component Kdo hydrolase KdhAB is important in the pathogenicity of *F. tularensis* LVS bacteria and in the organism’s avoidance of the TLR2-mediated immune response. We further show that a Kdo hydrolase-deficient mutant is protective against the fully virulent *F. tularensis* type A strain Schu S4. Such protective capacity is obviously a key feature to be hoped for in an effective vaccine strain.

The ΔkdhAB deletion mutant, unable to remove side chain Kdo from its LPS (Fig. 1), was impaired in its ability to kill mice after intranasal challenge (Fig. 2). This attenuation was not due to a growth defect or to sensitivity to complement-mediated killing. In *H. pylori*, Kdo hydrolase inactivation has downstream effects on subsequent steps in LPS biosynthesis, reducing the efficiency of lipid A modification enzymes 4′-phosphatase and 3′-O-deacylase and the amount of fully extended O antigen (17). Both 4′-phosphatase and O antigen are required for full virulence of *F. tularensis* in mice (10, 19, 29). However, Kdo hydrolase inactivation in *F. tularensis* LVS did not affect lipid A or O-antigen biosynthesis (Fig. 6). Therefore, virulence attenuation was not related to a defect in 4′-phosphatase activity or O-antigen assembly.

The avoidance of the TLR response by modifications in lipid A structure has been described in a number of pathogens, including members of the genera *Salmonella*, *Yersinia*, *Pseudomonas*, and *Francisella* (8, 10, 30–32). These modifications include lipid A deacylation, dephosphorylation, and glycosylation. We thus hypothesized that *F. tularensis* removes side chain Kdo from LPS core oligosaccharide to avoid the triggering of innate immunity. In agreement with this hypothesis, the ΔkdhAB mutant stimulated a stronger TLR2-mediated proinflammatory response in macrophage monolayers than the parental LVS did (Fig. 3 and 4). In *vivo*, ΔkdhAB bacteria induced a proinflammatory response in the lungs 24 h after intranasal challenge, while LVS bacteria did not (Fig. 3A, left panel). Attenuation of the virulence of ΔkdhAB bacteria may be related to this early innate immune response. Indeed, several studies have shown that eliciting an early inflammatory response in mice by priming the animals with an agonist of TLR3, TLR4, or TLR9 enhances survival to *F. tularensis* infection and decreases organ bacterial burdens (33–35).

Recent studies have indicated that Kdo residues contribute to proinflammatory responses to LPS (36–38). Therefore, the absence of a side chain Kdo in the ΔkdhAB mutant, with direct recognition of this modified LPS by the TLR2 signaling pathway, has emerged as a potential mechanism for the observed phenotype. However, our *in vitro* studies suggest that this is not the case, as purified LPSs from the ΔkdhAB mutant and LVS elicit similarly weak NF-κB activation (Fig. 4B). Another possible explanation is that the structural alteration of LPS exposes one or more TLR2 ligands present on the bacterial surface. The electrostatic charge of LPS is important for the folding of certain outer membrane proteins (39–42), and Kdo—as a negatively charged sugar—contributes to the overall charge of LPS. Thus, the presence of an additional Kdo residue in ΔkdhAB mutant LPS may alter the conformation of the outer membrane, exposing TLR2 ligands such as lipoproteins (43). In support of this idea, we showed that FopA surface protein is more accessible to antibodies in the ΔkdhAB mutant than in parental LVS (Fig. 5B).

LVS is an attenuated *F. tularensis* subsp. *holarctica* strain (type B) that was empirically derived in the former Soviet Union and developed as a live attenuated vaccine against the highly virulent *F. tularensis*. However, the LVS-based vaccine remains unlicensed in the United States and several other nations for several reasons, including the following: (i) an incomplete understanding of the genetic basis for its attenuation; (ii) untoward reactions; and (iii)
concerns about possible reversion back to the virulent form. The mechanisms of LVS-mediated protection against tularemia involve TLR2-dependent innate immunity (25–27). Therefore, an LVS mutant that exhibits attenuated virulence in mice and more strongly elicits activation of TLR2 signaling might prove to be a safer and more effective vaccine candidate than the original LVS. Here we show that the \( \Delta kdhAB \) mutant may be such a vaccine candidate: its virulence is attenuated, its ability to activate TLR2 is enhanced, and its protective efficacy against \( F. \) tularensis type A strain Schu S4 is similar to that of LVS. Mutants unable to modify lipid A by carbohydrate incorporation were attenuated in mice, activated proinflammatory responses, and were protective against \( F. \) tularensis subsp. novicida (8). However, protection against the highly virulent \( F. \) tularensis type A strain Schu S4 has not been investigated. Few previously assessed mutants have been shown to induce protective immunity against type A strains (for a review, see reference 44). These mutants have had deletions in genes including \( \text{FTT}1103 \), \( \text{FTT}0918 \), \( \text{sodB} \), \( \text{wzy} \), and \( \text{ggt} \) (45, 46). We report a new mutation that is relevant for tularemia vaccine design.

Herein we demonstrate that the two-component Kdo hydrolase KdhAB is involved in the virulence of \( F. \) tularensis LVS. A \( \Delta kdhAB \) mutant is attenuated in mice, elicits a stronger proinflammatory cytokine response in a TLR2-dependent manner, and induces protective immunity against the highly virulent Schu S4 strain of \( F. \) tularensis. Our data suggest that a novel LPS modification—the removal of side chain Kdo—is important in the evasion of innate immunity by \( F. \) tularensis. Kdo hydrolase is also produced by two other pathogens, \( H. \) pylori and \( L. \) pneumophila (14, 15, 17, 18). It will be particularly interesting to study the role of this enzyme in the virulence of these organisms.

**MATERIALS AND METHODS**

**Ethics statement.** All work with animals was conducted according to the recommendations in the Guide for the Care and Use of Laboratory Animals (47). The protocol was approved by the Standing Committee on Animals of the Harvard Medical Area (protocol 04723).

**Bacterial strains and plasmids.** Francisella tularensis LVS (kindly provided by Karen Elkins, U.S. Food and Drug Administration, Bethesda, MD) was grown in cysteine heart agar supplemented with 1% hemoglobin (CHAH) for 72 h at 37°C and 5% CO\(_2\), in modified Muller-Hinton broth (Becton, Dickinson, Franklin Lakes, NJ) supplemented with ferric pyrophosphate and IsoVitaleX (Becton, Dickinson, Franklin Lakes, NJ), or in tryptic soy broth (Difco, Detroit, MI). Escherichia coli strain DH5\(_\alpha\) was used for general cloning and was grown either in Luria-Bertani (LB) broth or on LB agar plates. When appropriate, ampicillin (100 \( \mu \)g/ml) and kanamycin (30 \( \mu \)g/ml) were added to broth medium, and kanamycin (10 \( \mu \)g/ml) was added to CHAH.

**Construction of the \( F. \) tularensis LVS \( \Delta kdhAB \) deletion mutant.** The \( \Delta kdhAB \) mutant was constructed by allelic exchange (48) by a procedure described previously (14). The whole \( kdhA \) gene was deleted. As the last

**TABLE 1** Survival of WT C57BL/6, TLR2\(^{−/−}\), and TLR4\(^{−/−}\) mice 21 days after an intranasal challenge with \( F. \) tularensis \( \Delta kdhAB \) mutant

| Challenge dose (CFU) | Survival rate (%) of mice: |
|---------------------|----------------------------|
|                     | C57BL/6 \((n = 10)\) | TLR2\(^{−/−}\) \((n = 10)\) | TLR4\(^{−/−}\) \((n = 5)\) |
| 10\(^2\)            | 100                       | 100                       | 100                       |
| 10\(^3\)            | 100                       | 40                        | 100                       |
| 10\(^4\)            | 100                       | 0                         | 100                       |
eight bases of \(kdhA\) overlap with \(kdhB\), the first eight bases of \(kdhB\) were also deleted. The following primers were used: 5'-ACCCCCGGGCACTTT CGTGCTTAGATATCT-3' (XmaI site underlined) and 5'-TGTGGTAC CAGTTTTAAAAATTAAGTAAGT-3' (KpnI site underlined) to amplify the 1-kb upstream region; 5'-ACCGGTACCTAATTATAACCTTTTGAC AAC-3' (KpnI site underlined) and 5'-TGTAATCTCGAAGTTAGGTG ATATGATTGC-3' (EcoRI site underlined) to amplify the 1-kb downstream region.

For complementation studies, plasmid \(pkdhAB\) encoding \(kdhAB\) genes under a \(groEL\) promoter was constructed. In brief, a DNA sequence containing open reading frames of both \(kdhA\) and \(kdhB\) genes was amplified by PCR with primers 5'-TGGAATTCATGAAACATAAACTAAAGC TAG-3' (EcoRI site underlined) and 5'-TCCTCGAGTTAGTAATATAT AATCTATCTT-3' (XhoI site underlined). After restriction endonuclease digestion, the \(kdhAB\) DNA fragment was ligated into the EcoRI and XhoI sites of plasmid pTH17 (49).

**Bacterial DNA/RNA preparation.** Plasmids were introduced into \(F. tularensis\) LVS by electroporation, and miniplasmids for \(E. coli\) DH5\(\alpha\) and \(F. tularensis\) LVS were prepared with a QIAprep spin miniprep kit according to the manufacturer’s instructions (Qiagen, Venlo, Nether-
lands). Restriction enzymes and DNA-modifying enzymes were purchased from New England Biolabs. DNA ligations and transformations of chemically competent E. coli were performed by standard protocols (50).

**LPS purification.** LPS was purified from *F. tularensis* WT and ΔkdhAB strains as described elsewhere (49). A modification of the hot phenol-water method was used (51).

**Mouse infection and immunization studies.** Male BALB/cByJ, C57BL/6, TLR2−/−, and TLR4−/− specific-pathogen-free mice (6 to 8 weeks old; Jackson Laboratory, Bar Harbor, ME) were housed in an animal facility at Harvard Medical School. All procedures conformed to protocols approved by the Institutional Animal Care and Use Committee. Bacterial virulence was assessed by intranasal challenge of BALB/cByJ mice (*n* = 6) with 10⁹ CFU of *F. tularensis* LVS bacteria or with 10⁸, 10⁹, or 10³ CFU of ΔkdhAB bacteria (inoculum volume, 50 µl per mouse). Mice were lightly anesthetized with isoflurane (Baxter, Deerfield, IL). The importance of the TLR2 signaling pathway in mouse survival after intranasal challenge with ΔkdhAB mutant bacteria (10⁶, 10⁷, or 10⁸ CFU) was assessed in C57BL/6 WT, TLR2−/−, and TLR4−/− mice. In these instances, survival was monitored over a 26-day period unless otherwise indicated.

For immunization studies, groups of BALB/cByJ mice (*n* = 10) were immunized by the intranasal route with three doses of bacteria at 2-week intervals; the doses were 10, 10, and 10⁵ CFU for *F. tularensis* LVS and 10⁷, 10⁸, and 10⁶ CFU for the ΔkdhAB mutant. A mock immunization group that received PBS served as a control. Four weeks after the last dose, mice were challenged intranasally with either 10⁴ CFU of LVS (10 × 10⁶ CFU) or 10⁶ CFU of strain Schu S4 (10 × 10⁶ CFU). Mice were challenged with the *F. tularensis* Schu S4 strain at the animal biosafety level 3 (ABS3) core facility at Harvard Medical School. Titers of serum antibody to LPS were measured by ELISA as described previously (26).

**Organ bacterial burden and histopathology.** BALB/cByJ mice were challenged intranasally with 10⁷ CFU of *F. tularensis* LVS or ΔkdhAB bacteria. Six days after infection, mice were killed, blood was collected, and tissues were weighed, hand-mashed, and homogenized with a Stomacher 80-paddle action blender (Seward, Port Saint Lucie, FL). Serial 10-fold dilutions were prepared with sterile 1× PBS supplemented with 2% fetal bovine serum. A 10-µl volume of each dilution was plated onto CHAH plates at a density of 5 × 10⁵ cells/well, and the plates were incubated overnight at 37°C in 5% CO₂. The macrophages were then infected with mid-log *F. tularensis* LVS or ΔkdhAB bacteria at an MOI of 100. At 2 h after infection, the cells were washed three times with Dulbecco’s PBS and lysed in 60 µl of lysis buffer (kit BFI14100; R&D Systems). IL-1β in lysates was measured with an ELISA Duoset kit (R&D Systems).

**Murine macrophage-like cell line infection.** WT, Un93B−/−, MyD88−/−, TLR4−/−, and TLR2−/− murine macrophage-like cell lines, kindly provided by Douglas T. Golenbock (University of Massachusetts Medical School, Worcester, MA), were seeded in 24-well plates at a density of 5 × 10⁵ cells/well, and the plates were incubated overnight at 37°C in 5% CO₂. The macrophages were then infected with mid-log *F. tularensis* LVS or ΔkdhAB bacteria at an MOI of 100. After 2 h, cells were washed three times with Dulbecco’s PBS and lysed in 60 µl of lysis buffer (kit BFI14100; R&D Systems). IL-1β in lysates was measured with an ELISA Duoset kit (R&D Systems).

**NF-κB reporter luciferase assay.** Stable HEK293 cell lines expressing TLR2 receptor were kindly provided by Douglas T. Golenbock. HEK-TLR2 and HEK-TLR4/MD2 cells were seeded into 12-well tissue culture plates at a density of 3 × 10⁵ cells/well. The following day, cells were transiently transfected with luciferase reporter genes; Genejuice (EMD Chemicals, Darmstadt, Germany) was used per the manufacturer’s recommendations. To assess NF-κB activation, an NF-κB luciferase reporter gene (consisting of an artificial promoter NF-κB luciferase reporter gene, phRL-TK (Promega, Fitchburg, WI); phRL-TK was used to normalize transfection efficiency. The following day, the cells were stimulated as indicated in Results. During stimulation with LPFs from *F. tularensis* LVS or ΔkdhAB bacteria, the mixture contained soluble CD14 recombinant protein (5 µg/ml) (ProSci Inc., Poway, CA). After 24 h of stimulation, the cells were lysed in passive lysis buffer (Promega), and reporter gene activity was measured with a plate reader luminescence detector (Victor2; PerkinElmer Life Sciences) and the Dual-Luciferase reporter assay system (Promega).

**Surface antigen-binding assay.** Evaluation of FopA antibody binding to the bacterial surface was performed as described previously (24), with some modifications. In brief, cultures of WT *F. tularensis* LVS, ΔkdhAB, and ΔwbtA bacteria were grown overnight in brain heart infusion (BHI) broth. Cultures were diluted in fresh BHI medium to an optical density at 620 nm (OD₆₂₀) of 0.07, and bacteria were further grown to an OD₆₂₀ of 0.5. After the bacteria were harvested, 6 × 10⁹ bacteria from each strain (in a 50-µl volume) were mixed with monoclonal FopA antibodies (final concentration, 0.1 mg/ml) (54) in PBS containing 0.1% glucose and 0.5% bovine serum albumin (BSA). After incubation for 1 h at 37°C, bacteria were assayed with ELISA Duoset kits (R&D Systems). For measurements of gene expression, BMMs were seeded in 6-well plates at a density of 2 × 10⁵ cells/well, and the plates were incubated at 37°C in 5% CO₂ overnight. The macrophages were then infected with mid-log *F. tularensis* LVS or ΔkdhAB bacteria at an MOI of 10. At various time points, cells were lysed in 0.4 ml Trizol (Invitrogen), and RNA was isolated according to the manufacturer’s instructions. Reverse transcription was performed with a cScript Flex cDNA synthesis kit (Quanta BioSciences, Gaithersburg, MD) according to the manufacturer’s instructions. Real-time PCR was conducted with Perfecta SYBR green supermix (Quanta Biosciences) in a Bio-Rad iCycler iQ system (45 cycles, with 1 cycle consisting of 15 s at 95°C, 45 s at 60°C, and 30 s at 72°C). Gene expression was normalized to the expression of the RNA polymerase II (RP II) gene, which is considered a stably expressed housekeeping gene (52). The following primer sets were used: for TNF-α, sense (5’-TATGG CTCAGGGTCCAACCTC-3’) and antisense (5’-CTCCTCTTTGGCAGACT CAGG-3’); for IL-1β, sense (5’-GGGCCTCAAAGGAAAGAATC-3’) and antisense (5’-TACCAGTGGGGAACCTCCTG-3’); and for RP II, sense (5’-GCACCAGCTCAATGATAT-3’) and antisense (5’-CTGGCGGTT GACCCCATGAC-3’). Relative gene expression was calculated by the ΔΔC_T method (53) and reported as fold induction over the uninfected control level (mean ± standard deviation [SD] values for samples from three independent experiments).

**Characterization of kdhAB in Francisella tularensis.**
REFERENCES
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SUPPLEMENTAL MATERIAL
Outer membranes from F. tularensis LVS and ΔkdhAB bacteria were iso-
ated, resolved by Tricine SDS-PAGE, and stained with zinc as previously
described (14). Lipid A from LVS and ΔkdhAB bacteria was isolated and
analyzed by negative-ion matrix-assisted laser desorption ionization—
time of flight (MALDI-TOF) mass spectrometry (14).
Data analysis. Data are expressed as mean ± standard deviation (SD)
values. Unpaired Student’s t test was used for statistical analysis. A P value
of <0.05 was considered significant.
SUPPLEMENTAL MATERIAL
Supplemental material for this article may be found at http://mbio.asm.org
/lookup/suppl/doi:10.1128/mBio.00638-12/-/DCSupplemental.
Figure S1, EPS file, 0.8 MB.
Figure S2, JPEG file, 0.5 MB.
Figure S3, JPEG file, 0.5 MB.
Table S1, DOCX file, 0.1 MB.
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REFERENCES
1. Bosio CM, Bielefeldt-Ohmann H, Belisle JT. 2007. Active suppression
of the pulmonary immune response by Francisella tularensis Schu. J. Immunol. 178:4358–4367.
2. Hall JD, Woolard MD, Gunn BM, Craven RB, Taft-Benitez S, Frelinger
JA, Kawula TH. 2008. Infect. Immun. 76:5843–5852.
3. Malik M, Bakshi CS, Sahay B, Shah A, Lotz SA, Sellati TJ. 2006. Toll-like
receptor 2 is required for control of pulmonary infection with Francisella
Klebsiella pneumoniae-induced, antibody-mediated protection against
Francisella tularensis. Infect. Immun. 74:3657–3662.
4. Mares CA, Ojeda SS, Morris EG, Li Q, Teale JM. 2009. Initial delay in the
immune response to Francisella tularensis is followed by hyporeactivecyto-
kinemia characteristic of severe sepsis and correlating with upregulation and
release of damage-associated molecular patterns. Infect. Immun. 77:536–541.
5. Katz J, Zhang P, Martin M, Vogel SN, Michalek SM. 2005. Toll-like
receptor 2 is required for inflammatory responses to Francisella tularensis
LVS. Infect. Immun. 73:2809–2816.
6. Russo TA, Beanan JM, Olson R, MacDonald U, Cope JJ. 2007. Capsular
polysaccharide and the O-specific antigen impede antibody binding: a
potential obstacle for the successful development of an extraintestinal
pathogenic Escherichia coli vaccine. Vaccine 27:388–395.
7. Schneider MC, Exley RM, Ram S, Sim RB, Tang CM. 2007. Interactions
between Neisseria meningitidis and the complement system. Trends Microbiol.
15:233–240.
8. Wu MF, Yang CY, Lin TL, Wang JT, Yang JL, Wu SH, Hu BS, Chou
TY, Tsai MD, Lin CH, Hsieh SL. 2009. Humoral immunity against
capsule polysaccharide protects the host from maga+ Klebsiella
pneumoniae-induced lethal disease by evading Toll-like receptor 4 signa-
l. Infect. Immun. 77:615–621.
9. Zarrrella TM, Singh A, Bitsaksitis C, Rahman T, Sahay B, Feustel PJ,
Charbit A. 2007. Role of the wrt locus of Francisella tularensis in lipopoly-
saccharide O-antigen biogenesis and pathogenicity. Infect. Immun. 75:
536–541.
10. Pinkham JT, Henninger SJ, Chalabaev S, Kasper DL. 2012. Genetic
modification of the O-polysaccharide of Francisella tularensis results
in an avirulent live attenuated vaccine. J. Infect. Dis. 205:1056–1065.
11. Kirmajonjeswara GS, Olimos S, Bakshi CS, Metzger DW. 2008. Humoral
and cell-mediated immunity to the intracellular pathogen Francisella
tularensis. Immunol. Rev. 225:244–255.
12. Weiss DS, Brotcke A, Henry T, Margolis JJ, Chan K, Monack DM. 2007.

In vivo negative selection screen identifies genes required for Francisella virulence. Proc. Natl. Acad. Sci. U. S. A. 104:6037–6042.

29. Sebastian S, Dillon ST, Lynch JG, Blalock LT, Balon E, Lee KT, Comstock LE, Conlan JW, Rubin EJ, Tzianabos AO, Kasper DL. 2007. A defined O-antigen polysaccharide mutant of Francisella tularensis live vaccine strain has attenuated virulence while retaining its protective capacity. Infect. Immun. 75:2391–2402.

30. Ernst RK, Adams KN, Moskowitz SM, Kraig GM, Kawasaki K, Stead TM, Trent MS, Miller SI. 2006. The Psedomonas aeruginosa lipid A deacylase: selection for expression and loss within the cystic fibrosis airway. J. Bacteriol. 188:191–201.

31. Kawahara K, Tsukano H, Watanabe H, Lindner B, Matsuura M. 2002. Modification of the structure and activity of lipid A in Yersinia pestis lipopolysaccharide by growth temperature. Infect. Immun. 70:4092–4098.

32. Kawasaki K, Ernst RK, Miller SI. 2004. 3-O-deacylation of lipid A by PagL, a PhoP/PhoQ-regulated deacylase of Salmonella typhimurium, modulates signaling through Toll-like receptor 4. J. Biol. Chem. 279:20044–20048.

33. Pyles RB, Jezek GE, Eaves-Pyles TD. 2010. Toll-like receptor 3 agonist protection against experimental Francisella tularensis respiratory tract infection. Infect. Immun. 78:1700–1710.

34. Rozak DA, Gelhaus HG, Smith M, Zadeh M, Huzella L, Waag D, Adamovicz JJ. 2010. Cpg oligodeoxyribonucleotides protect mice from Burkholderia pseudomallei but not Francisella tularensis Schu S4 aerosols. J. Immunol. Based Ther. Vaccines 8:2.

35. West TE, Pelletier MR, Majure MC, Lembo A, Hajjar AM, Skerrett SJ. 2008. Inhalation of Francisella novicida delta mglA causes replicative infection that elicits innate and adaptive responses but is not protective against invasive pneumonic tularemia. Microbes Infect. 10:773–780.

36. Gaedwad J, Zhang Y, Zhang W, Reeves J, Wolfert MA, Boons GI. 2010. Differential induction of innate immune responses by synthetic lipid A derivatives. J. Biol. Chem. 285:29375–29386.

37. Muroi M, Tanamoto K. 2002. The polysaccharide portion plays an indispensable role in Salmonella lipopolysaccharide-induced activation of NF-kappaB through human Toll-like receptor 4. Infect. Immun. 70:6043–6047.

38. Zughaier SM, Tseng YL, Zimmer SM, Datta A, Carlson RW, Stephens DS. 2004. Neisseria meningitidis lipooligosaccharide structure-dependent activation of the macrophage CD14/Toll-like receptor 4 pathway. Infect. Immun. 72:371–380.

39. Brandenburg K, Andrá J, Müller M, Koch MH, Garidel P. 2003. Physicochemical properties of bacterial glycopolymers in relation to bioactivity. Carbohydr. Res. 338:2477–2489.

40. de Cock H, Brandenburg K, Wiese A, Holst O, Seydel U. 1999. Nonlamellar structure and negative charges of lipopolysaccharides required for efficient folding of outer membrane protein PhoE of Escherichia coli. J. Biol. Chem. 274:5114–5119.

41. Kramer RA, Brandenburg K, Vandeputte-Ruten L, Werkhoven M, Gros P, Dekker N, Emond MR. 2002. Lipopolysaccharide regions involved in the activation of Escherichia coli outer membrane protease OmpT. Eur. J. Biochem. 269:1746–1752.

42. Snyder S, Kim D, McIntosh TJ. 1999. Lipopolysaccharide bilayer structure: effect of chemotype, core mutations, davalent cations, and temperature. Biochemistry 38:10758–10767.

43. Thakran S, Li H, Lavine CI, Miller MA, Bina JE, Bina XR, Re F. 2008. Identification of Francisella tularensis lipoproteins that stimulate the Toll-like receptor (TLR) 2/TLR1 heterodimer. J. Biol. Chem. 283:3751–3760.

44. Conlan JW. 2011. Tularemia vaccines: recent developments and remaining hurdles. Future Microbiol. 6:391–405.

45. Ireland PM, Lebutt H, Thomas RM, Oyston PC. 2011. A Francisella tularensis SCHU S4 mutant deficient in gamma-glutamyl transferase activity induces protective immunity: characterisation of an attenuated vaccine candidate. Microbiology 157:3172–3179.

46. Qin A, Scott DW, Thompson JA, Mann BJ. 2009. Identification of an essential Francisella tularensis subsp. tularensis virulence factor. Infect. Immun. 77:152–161.

47. National Research Council. 2011. Guide for the care and use of laboratory animals, 8th ed. National Academy Press, Washington, DC.

48. Lauriano CM, Barker JR, Nano FE, Arulanandam BP, Klose KE. 2003. Allelic exchange in Francisella tularensis using PCR products. FEMS Microbiol. Lett. 229:195–202.

49. Kim TH, Sebastian S, Pinkham JT, Ross RA, Blalock LT, Kasper DL. 2010. Characterization of the O-antigen polymerase (Wzy) of Francisella tularensis. J. Biol. Chem. 285:27839–27849.

50. Sambrook J, Fritsch EF, Maniatis T. 1989. Molecular cloning: a laboratory manual, 2nd ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.

51. Westphal O, Jann K. 1965. Bacterial lipopolysaccharides. Extraction with phenol-water and further applications of the procedure. Methods Carbohydr. Chem. 5:83–91.

52. Radoníč A, Thulke S, Mackay IM, Landt O, Siegert W, Nitsche A. 2004. A defined Francisella tularensis strain missing a gammaglutamyl transferase gene shows impaired virulence. Infect. Immun. 72:283–290.

53. Livak KJ, Schmittgen TD. 2001. Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) method. Methods 25:402–408.

54. Savitz AG, Mena-Taboada P, Monsalve G, Benach JL. 2009. Francisella tularensis infection-derived monoclonal antibodies provide detection, protection, and therapy. Clin. Vaccine Immunol. 16:414–422.

55. Vinogradov E, Perry MB, Conlan JW. 2002. Structural analysis of Francisella tularensis lipopolysaccharide. Eur. J. Biochem. 269:6112–6118.