Larvicidal Efficacy and GC-MS Analysis of Hyptis suaveolens Leaf Extracts against Anopheles Species

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Authors’ contributions

This work was carried out in collaboration among all authors. Author YDD designed the study, managed the literature research and wrote the first draft of the manuscript. Authors CNA and DDP supervised and edited the work. Author AO performed the statistical analysis. Author FA played a role in GC-MS aspect of the work. Authors STJ and IRJ assisted in the lava scouting. Authors IRJ and SWK they played a role in the aspect of the phytochemical screening and author PML participated in the lava collection. All authors read and approve the final manuscript.

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ABSTRACT

Mosquitoes are the most important group of insects in terms of public health importance, which transmit serious human diseases. The continuous application of synthetic insecticides to control these mosquitoes causes development of resistance in vector species, and an adverse effect on environmental quality and non-target organisms including human health. Therefore, the use of active toxic agents from plant extracts as alternative mosquito control strategy cannot be over emphasized, as these are non-toxic, easily available at affordable prices, biodegradable and show...
The primary public health intervention for reducing malaria transmission at the community level is through vector control and it is the only approach that can be effective in large-scale areas, especially in urban and rural settings. It is now being reconsidered as a complementing intervention to current priorities such as insecticide-treated nets (ITNS) and access to early diagnosis and prompt treatment [12]. The control of immature aquatic stages of *Anopheles* mosquitoes may have particular promise in urban settings where large numbers of people can be protected in a relatively small area and rural settings with focal, seasonal breeding sites [13].

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intervention that can reduce malaria transmission from very high levels to close to zero [14]. Many approaches have been developed to prevent mosquito menace. One of such strategies has been mainly based on the use of synthetic insecticides to interrupt the disease transmission cycle by either targeting the mosquito larvae at breeding sites through spraying of stagnant water, by killing or repelling the adult mosquitoes [15]. Even though these are effective, they created many problems such as toxicity to human beings, harm to non-target population, long persistence in environment and entry in the food chain [16]. As a result of these problems, it is necessary to develop environmentally safe, biodegradable, economical and indigenous method for control of vector that can be used with minimum care by individuals and communities [17]. Plant products with potentials to act as insecticides or repellent can play an important role in the interruption of transmission of mosquito borne disease at the individual as well as community level [18]. The secondary metabolites in different plants make up a vast repository of compounds with a wide range of biological activities [19]. It is in view of an increasing interest in developing plant based insecticides that the present study was undertaken.

Hyptis suaveolens is a medium aromatic weedy shrub found in the tropics and sub-tropical region [20]. The plant has been reported to possess repellent activity against insect pests of stored grains [21]. Methanolic extract of Hyptis suaveolens possess insecticidal activity against Sitophilus maculates [22]. Fumes of the dried leaves are also used locally to repel mosquitoes and to control insect pests of stored grains. It was also reported by [23] that placing H. suaveolens branches or whole plants in houses was one of the most effective methods to repel malaria vector An. gambiae s.s Giles. Therefore, the crude leaf extracts of Hyptis suaveolens has been investigated in this study for its larvicidal potency against malaria vectors, Anopheles species.

2. MATERIALS AND METHODS

2.1 Study Area

The study was carried out at the Research Laboratory of Zoology Department, University of Jos, Plateau State, Nigeria.

2.2 Procedures of Plant Collection and Extraction

Mature fresh leaves of Hyptis suaveolens were collected in May, 2016 at Rafin Saniye village in the Southern part of Pankshin (Latitude 9° 20’ 0” North, and longitude 9° 27’ 0” East) Local Government Area of Plateau state, Nigeria. The plant leaves were air-dried under shade in Zoology Department Research Laboratory; University of Jos, Nigeria at room temperature for 14days. The plant specimen was identified by a taxonomist Mr. Joseph Azila, at the Herbarium of Federal College of Forestry, Jos, Nigeria.

The plant extract was prepared by maceration method in which the dried leaves of the plant was ground with mortar and pestle and then sieved to get fine powder. 250 g of the plant powder was weighed into 500 ml conical flasks and soaked in 200 ml of methanol. This was left to stand overnight (24hours) and shaked for 3hours on a mechanical shaker. The content was filtered using a non-absorbent cotton wool on a Buchner funnel-flask using a vacuum pump. The residue was subjected to several parts of rinsing and filtration with fresh solvent to attain some level of maceration. The filtrate was evaporated to dryness using a rotary evaporator and a drying cabinet. The extract was then transferred into a sample container and preserved in the refrigerator. Same procedure was applied for the aqueous extract using distilled water as the extracting solvent and was concentrated to dryness using a water bath at control temperature of 60°C.

2.3 Phytochemical Screening

The phytochemical screening was carried out as described by [24,25]. The procedures are briefly described below.

2.3.1 Test for tannins

About 2 ml of the extract was stirred with 2 ml of distilledwater and few drops of FeCl₃ solution were added. The formation ofa green precipitate was an indication for the presence of tannins.

2.3.2 Test for saponins

5 ml of extract was shaken vigorously with 5 ml of distilledwater in a test tube and warmed. The
formation of stable foam wastaken as an
dication for the presence of saponins.

2.3.3 Test for flavonoids

To 1 ml of extract was added 1 ml of 10% lead
acetatesolution. The formation of a yellow
precipitate was taken as a positive test for
flavonoids.

2.3.4 Test for terpenoids

2 ml of the extract was dissolved in 2 ml of
chloroform and evaporated to dryness. 2 ml of
concentrated sulphuric acid was then added and
heated for about 2 min. A greyish colour
indicates the presence of terpenoids.

2.3.5 Tests for steroids

A red colour produced in the lower chloroform
layer when 2 ml of extract was dissolved in 2 ml
of chloroform and 2 ml concentrated sulphuric
acid added indicates the presence of steroids.

2.3.6 Test for alkaloids

3 ml of the extract was stirred with 3 ml of 1%
HCl on a steambath. Mayer’s and Wagner’s
reagents were then added to the mixture.
Turbidity of the resulting precipitate was taken as
evidence for the presence of alkaloids.

2.3.7 Tests for carbohydrates

To 3 ml of the extract was added about 1 ml of
iodinesolution. A purple colouration at the
interphase indicates the presence of carbohydrates.

2.3.8 Tests for glycosides

Salkowski’s test: 2 ml of the extract was
dissolved in 2 ml of chloroform. 2 ml of sulphuric
acid was added carefully and shakengently. A
reddish brown colour indicates the presence of a
steroidal ring (that is, a glycone portion of
glycoside).

2.3.9 Test for resin

2 ml of the extract and 2 ml of acetic anhydride
were mixed together. A drop of concentrated
sulphuric acid was added to the mixture. A purple
or violet colour indicates the presence of resin.

2.3.10 Test for balsams

2 ml of the extract and 2 ml ethanol were mixed
together and two drops of alcoholic ferric chloride
solution was added. A dark green colouration
indicates the presence of balsams.

2.4 Larvae Collection

Anopheles larvae were collected from stagnant
water at the edges of river Timshing in Rafin
saniye village of Pankshin Local Government
Area of Plateau State, Nigeria using a dipper.
The larval stage of Anopheles species were
distinguished from those of Culex species using
morphological differences [26].

2.5 Gas Chromatography and Mass
Spectrometry (GC – MS) Analysis

The GC-MS analysis of the methanol extracts of the
H. suaveolens was carried out on GCMS-QP
2010 plus, at National Research Institute for
Chemical Technology (NARICT) Zaria, Nigeria.
REF TEK column was used for the analysis. The
sample was injected with split less mode. Helium
gas was used as a carrier gas. The database
used for the identification of the compounds was
National Institute Standard and Technology MS
library.

2.6 Preparation of the Concentrations of
the Extracts and Larvicidal Bioassay

A Range Finding Test (RFT) was conducted to
obtain definitive toxic concentration for bioassay
[27]. Five concentrations (200, 400, 600, 800 and
1000 mg/L) of the leaf extracts with control as
0.00 mg/L were prepared.

The larval bioassay was carried out according to
the WHO standard procedure [28]. The tests
were conducted in plastic containers. From the
extracts, concentrations of 200, 400, 600, 800 and
1000 mg/L were prepared. The fourth instars
larvae of Anopheles species were placed in each
test solution using a dropper to study the
larvicidal property as per the following
procedures. Twenty healthy fourth larval instars
of Anopheles species were released in each
concentration of both treatment and the
replicates with extract solution and control
experiments without extracts were run in parallel.
The larvae in each solution were then left for
24hours and 48hours, the numbers of dead
larvae were counted after 24hours and 48hours
of exposure and the percentage mortality were
recorded from the average of three replicates.
Larvae were considered dead if appendages did
not move when probed with needle. Larvae
incapable of rising to the surface or not showing
the characteristic diving reaction when water was disturbed, were considered moribund and added to the dead larvae for calculation of percentage mortality.

2.7 Calculation of Mortality Rate

The mortality rate was calculated using the formula:
\[
\text{Observed mortality} = \frac{\text{Total number of dead larvae}}{\text{Total number of larvae used}} \times 100 \quad \text{equation(1)}
\]

2.8 Interpretation of Susceptibility Test Results

The interpretation was based on WHO bioassay protocol [29] which states that:

- Mortality in the range of 98 – 100% indicates susceptibility.
- Mortality in the range of 80 – 97% is suggestive of the existence of resistance and further investigation is needed.
- Mortality less than 80% indicates resistance.

2.9 Statistical Analysis

Data obtained were analyzed using R Console software (Version 3.2.2). Proportions of mortality rate of Anopheles species larvae in relation to concentrations of aqueous and as well as methanol extracts were compared using Pearson's Chi-square test. It was also used to compare proportions of mortality rate of Anopheles species larvae between extracts for each concentration. The average larval mortality data were subjected to probit analysis for calculating LC50. Results with P-values < 0.05 were considered to be statistically significant.

3. RESULTS

3.1 Phytochemical Screening of H. suaveolens from Methanol and Aqueous Extracts

Phytochemical screening analysis showed that H. suaveolens has seven (7) active ingredients from both methanol and aqueous extracts with the exception of saponins and alkaloids (Table 1). Terpenoids was strongly positive in methanol extract while flavonoid was strongly positive in aqueous extract. Cardiac glycosides was at trace level in methanol extract but not detected in aqueous extract. Resins were at positive level in aqueous extract but not detected in methanol extract. Steroids were at trace level in both methanol and aqueous extracts.

3.2 Mortality Rate of Anopheles species Larvae a Cross the Concentrations of Extracts of H. suaveolens in Relation to Time

3.2.1 Aqueous extract

The control had no dead Anopheles species larvae at both 24 hours and 48 hours of the bioassay whereas the aqueous extracts treatments showed varying levels of mortality across concentrations (Table 2). At the end of 24hrs and 48hours exposure time respectively, mortality rate increased in ascending order of concentrations. The 1000 mg/L concentration recorded the highest mortality rate of Anopheles species larvae of 95% followed by 800 mg/L with 80%, 600 mg/L had 63.5%, 400 mg/L showed 55% while 200 mg/L was 25% .The LC50 of aqueous extract obtained at the end of 48 hours exposure time of the assay was 323.5 mg/L. There was significant difference in the mortality rate of Anopheles species larvae across concentrations of aqueous extract (\(\chi^2 = 116.8, \text{df} = 5, P < 0.05\)).

The Anopheles larvae were tolerant (mortality rate <80%) to concentrations of aqueous extract between 200 to 600 mg/L while they showed possible tolerance (mortality rate 80% to 97%) between 800 to 1000 mg/L concentrations.

3.2.2 Methanol extract

The control had no dead Anopheles species larvae at both 24 hours and 48 hours of the bioassay whereas the methanol extract treatments showed varying levels of mortality
across concentrations (Table 3). At the end of 24hrs and 48hrs exposure time respectively, mortality rate increased in ascending order of concentrations. The 1000 mg/L concentration recorded the highest mortality rate of Anopheles species larvae of 100% followed by 800 mg/L with 86.7%, 600 mg/L had 65%, 400 mg/L showed 43.4% while 200 mg/L was 36.7%. The LC50 of methanol extract obtained at the end of 48 hours exposure time of the assay was 316.22 mg/L (Table 5). There was significant difference in the mortality rate of Anopheles species larvae across concentrations of methanol extract ($\chi^2 = 119.78$, df = 5, $P < 0.05$).

The Anopheles species larvae were tolerant (mortality rate <80%) to concentrations of methanol extract between 200 to 600 mg/L while they showed possible tolerance (mortality rate 80% to 97%) to 800 mg/L concentration, but were susceptible (mortality rate 100%) to concentration of 1000 mg/L.

### 3.3 Comparison of Mortality Rate in Relation to Extracts of *H. suaveolens* at the End of 48 Hours Exposure Time

Mortality rate of Anopheles species larvae was higher in all concentrations of methanol extract over the aqueous extract with the exception of 400 mg/L concentration in which aqueous extract had higher mortality rate (Table 4). However, there was no significant difference ($P > 0.05$) in the mortality rate of Anopheles species larvae between extracts of *H. suaveolens* for each concentration.

### 3.4 Preliminary Characterization of *H. suaveolens* Methanol Leaf Extracts Using Gas Chromatography and Mass Spectrometry (GC-MS)

The composition and identification of the main compounds (Table 5) present in the methanol leaf extracts of *H. suaveolens* revealed the presence of twelve compounds on the chromatogram. These compounds were identified through mass spectrometry attached with the gas chromatography. The gas chromatography revealed the concentrations of the various compounds as a function of retention time while the peak heights showed the relative concentrations of the components present in the leaves of the plant *H. suaveolens*. On the other hand, the mass spectrometer analyses the compounds at different times to identify their structures as well as their nature. The highest peak area is recorded in peak seven, having 33.33% with 21.378 minutes retention time (RT), and the compound name is Oleic Acid while the lowest is recorded in peak eleven having 0.56% with 24.810 minutes as retention time (RT) and the compound name is Decyl fluoride. The gas chromatography and mass spectrometry (GC-MS) spectrum of the extract revealed the presence of long chains of hydrocarbons.

![GC-MS Chromatogram](image-url)

*Fig. 2. Showed GC-MS chromatogram of the methanol extract of *H. suaveolens*. Twelve compounds were detected in the extract. The highest peak area is recorded in peak seven, having 33.33% with 21.378 minutes retention time (RT) and the compound name is oleic acid while the lowest is recorded in peak eleven having 0.56% with 24.810 minutes as retention time (RT) and the compound name is decyl fluoride.*
Table 1. Phytochemical screening of active ingredients of *H. suaveolens* from methanol and aqueous extracts

| S/No | Secondary metabolite | *H. suaveolens* Methanol extract | *H. suaveolens* Aqueous extract |
|------|-----------------------|----------------------------------|-------------------------------|
| 1    | Tannins               | +                                | ++                            |
| 2    | Saponins              | -                                | -                             |
| 3    | Alkaloids             | -                                | -                             |
| 4    | Terpenoids            | +++                              | +                             |
| 5    | Carbohydrates         | +                                | -                             |
| 6    | Cardiac glycosides    | +                                | -                             |
| 7    | Balsams               | ++                               | +                             |
| 8    | Resins                | ++                               | +                             |
| 9    | Flavonoids            | ++                               | +++                           |
| 10   | Steroids              | +                                | +                             |

**Key:** +++: Strongly positive, ++: Positive, +: Trace, -: Not detected

Table 2. Mean mortality rate of *Anopheles species* larvae across concentrations of aqueous extract of *H. suaveolens* in relation to time

| Concentration (Mg/L) | Log concentration | Mean No. of larvae exposed | Mean mortality rate 24hours | Mean mortality rate 48hours | % Mortality rate | Probit mortality |
|----------------------|-------------------|---------------------------|------------------------------|----------------------------|-----------------|-----------------|
| 0.0                  | 0.0               | 20                        | 0.00                         | 0.00                       | 0.0             | 0.0             |
| 200                  | 2.30              | 20                        | 1.00                         | 5.00                       | 25.0            | 4.32            |
| 400                  | 2.60              | 20                        | 3.67                         | 11.0                       | 55.0            | 5.12            |
| 600                  | 2.77              | 20                        | 5.00                         | 12.7                       | 63.5            | 5.40            |
| 800                  | 2.90              | 20                        | 6.67                         | 16.0                       | 80.0            | 5.84            |
| 1000                 | 3.00              | 20                        | 7.67                         | 19.0                       | 95.0            | 6.70            |

Table 3. Mean mortality rate of *Anopheles species* larvae across concentrations of methanol extract of *H. suaveolens* in relation to time

| Concentration (Mg/L) | Log concentration | Mean No. of larvae exposed | Mean mortality rate 24hours | Mean mortality rate 48hours | % Mortality rate | Probit mortality |
|----------------------|-------------------|---------------------------|------------------------------|----------------------------|-----------------|-----------------|
| 0.0                  | 0.0               | 20                        | 0.0                          | 0.0                        | 0.0             | 0.0             |
| 200                  | 2.30              | 20                        | 2.33                         | 7.33                       | 36.7            | 4.66            |
| 400                  | 2.60              | 20                        | 2.67                         | 8.67                       | 43.4            | 4.83            |
| 600                  | 2.77              | 20                        | 5.00                         | 13.00                      | 65.0            | 5.38            |
| 800                  | 2.90              | 20                        | 6.33                         | 17.33                      | 86.7            | 6.11            |
| 1000                 | 3.00              | 20                        | 6.67                         | 20.00                      | 100             | 8.71            |

Table 4. Mortality rate in relation to extracts of *H. suaveolens* at the end of 48 hours exposure time

| Concentration (mg/L) | % Mortality rate | $\chi^2$ | Df | P value |
|----------------------|------------------|----------|----|---------|
|                      | Aqueous extract  | Methanol extract |     |         |
| 200                  | 25.0             | 36.7     | 2.2186 | 1 | 0.1364  |
| 400                  | 55.0             | 43.4     | 1.3675 | 1 | 0.2422  |
| 600                  | 63.5             | 65.0     | 0.01751 | 1 | 0.8947  |
| 800                  | 80.0             | 86.7     | 0.26929 | 1 | 0.6038  |
| 1000                 | 95.0             | 100      | 0.12821 | 1 | 0.7203  |
4. DISCUSSION

The result of the Phytochemical screening (as shown in Table 1) revealed that Tannins, Terpenoids, Flavonoids, Steroids and Carbohydrates were present in both methanolic and aqueous extracts of *H. suaveolens*. Cardiac glycoside was detected only in methanolic extract while resins were positive in aqueous extract only. There was absence of saponins and alkaloids in both methanolic and aqueous extracts. It is in accordance to the work of [30] who recorded absence of saponins in *H. suaveolens* in a study carried out on the larvae of *Culex quinquefasciatus* to test the larvicidal activity of the plant, but detected strong presence of alkaloids using ethanol as the solvent for extraction. Also, [31] reported that *H. suaveolens* was devoid of saponins but tannins (0.520%) and phenols (0.050%) were present. Researchers such as [32] identified the above mentioned components except for saponins and alkaloids in eucalyptus and tamarind using methanol as solvent when tested against *Anopheles gambiae*.

These secondary metabolites are known to be effective against a wide range of insect pests as well as mosquito vectors [33]. The most reported insecticidal group of compounds from plants with larvicidal activity against various species of mosquito are steroids, flavonoids, phenols, tannins, terpenoids, carbohydrates and saponins [34,35]. These phytochemicals may be responsible for their insecticidal properties [36]. The failure of the larvae to moult and then develop into perfect pupae may be due to the delayed lethal effects of the phytocompounds which cause disturbances in the endocrine mechanism regulating metamorphosis [37]. In the screening of the phytochemicals, methanolic extract detected more metabolites than the aqueous extract which possibly suggest that it is more sensitive in detection of active ingredients in the plant used. This is in agreement with the findings of [38] who worked on the efficacy of aqueous and methanol extracts of the plant *Desmodium triflorum* for potential antibacterial activity. Their work confirmed that methanol extract detected more metabolites than aqueous extract. Therefore, the potent larvicidal activity of aqueous and methanolic extracts of *H. suaveolens* could be attributed to chemical compounds detected in this study.

The bioassay investigation revealed that the aqueous and methanolic extracts of *H. suaveolens* showed promising larvicidal activity against *Anopheles species*. This is in agreement with the study by [39] who tested the larvicidal activity of *H. suaveolens* acetone aerial extracts against the larvae of filarial vector *Culex quinquefasciatus*. On the contrary, *H. suaveolens* petroleum leaf extracts did not show any effective larvicidal activity against *Culex* species mosquito larvae [40]. Possibly this could be as a result of inability of the solvent to extract the major active ingredients needed for the purpose.

In aqueous extract, mortality of *Anopheles* species larvae recorded after 24 hours increased

| Peak No | RT (min) | Compound Name                        | Area (%) | Molecular Formula | Molecular Weight (g/mol) |
|---------|----------|--------------------------------------|----------|-------------------|--------------------------|
| 1       | 6.469    | 2-Hexanone, 3-methyl-4-methylene     | 3.57     | C₆H₁₂O             | 126                      |
| 2       | 7.203    | 4H-Pyran-4-one, 2,3-dihydro-3,5-dihydroxy-6-methyl | 6.06     | C₆H₈O₄             | 144                      |
| 3       | 8.636    | 4-Hepten-3-one, 4-methyl             | 7.0      | C₆H₁₂O             | 126                      |
| 4       | 12.395   | 1,3-Cyclopentanediol, trans         | 9.60     | C₆H₁₀O₂            | 102                      |
| 5       | 18.630   | n-Hexadecanoic acid                 | 9.01     | C₁₈H₃₂O₂           | 256                      |
| 6       | 20.570   | Octadecanoic acid, methyl ester     | 1.39     | C₁₉H₃₈O₂           | 298                      |
| 7       | 21.378   | Oleic Acid                          | 33.33    | C₁₈H₃₄O₂           | 282                      |
| 8       | 21.602   | Octadecanoic acid                   | 13.52    | C₁₈H₃₀O₂           | 284                      |
| 9       | 23.461   | Methoxyacetic acid, 2-methylphenyl ester | 4.31     | C₁₀H₁₂O₃           | 180                      |
| 10      | 24.616   | 9-Octadeconal                        | 1.17     | C₁₈H₃₄O₂           | 266                      |
| 11      | 24.810   | Decyl fluoride                       | 0.56     | C₁₀H₂₁F            | 160                      |
| 12      | 25.703   | 1,3-Cyclohexadiene-1-methanol, alpha,2,6,6- | 10.42    | C₁₁H₁₈O            | 166                      |

Table 5. Compound identified from the methanol leaf extract of *H. suaveolens*
as the concentrations of the extracts increased. This possibly suggest that the metabolites get to the target sites of Anopheles species larvae much faster with an increase in concentration of H. suaveolens aqueous extract. This is in agreement with the finding of [41] who assessed larvicidal activity of H. suaveolens and Balanites aegyptica leaves and root extracts against mosquito species. After 48 hours, observed mortality increased as concentration increased, with 95% mortality at the highest concentration. This is in agreement with the studies by [42] who recorded 95% larval mortality of aqueous extracts of Phyllanthus emblica (dry seed) on Aedes albopictus. This is also in accordance with the result of [43] who recorded higher mortality rate with increase in concentration against mosquito species. The Anopheles species larvae were tolerant to concentrations of aqueous extract between 200 to 600 mg/L while they showed possible tolerance between 800 to 1000 mg/L concentrations.

At 24 hours larval exposure to methanol extracts, the result demonstrated progressive increase in percentage mortality as concentrations increased (Table 3). After 48 hours, the methanol extract demonstrated significant larval mortality progressively as the concentration increased as well. A mortality of 100% was recorded in the highest concentration of 1000mg/L. This result is in agreement with the finding of [44] who reported 100% larval mortality of acetone aerial extract of Hyptis suaveolens at the highest concentration of 1000 mg/L against the larvae of filarial vector Culex quinquefasciatus say after 48 hours bioassay. Also, [45] recorded 100% mortality of stored product pests, Sitophilus oryzae, S. zeamais and Callosobruchus maculatus using methanol extract of H. suaveolens. A significant mortality of fourth instars larvae of Culex quinquefasciatus was also recorded with methanolic extract of Ciona celata with $L_C_{50}=95.63$ ppm and $L_C_{90}=242.16$ ppm [46,47] reported also that methanol leaf extract of Acalypha indica was more lethal to the egg and larvae of An. stephensi and Culex quinquefasciatus. In this study, the Anopheles species larvae were tolerant to concentrations of methanol extract between 200 to 600 mg/L while they showed possible tolerance to 800 mg/L concentration, but were susceptible to concentration of 1000 mg/L.

Among the twelve compound detected in the GC-MS chromatogram of the methanol extract of H. suaveolens, the highest peak area is recorded in peak seven, having 33.33% with 21.378 seconds retention time (RT), and the compound name is Oleic Acid. Hence, [48] suggested that the compound oleic acid, eicosyl ester from Thalictrum javanicum are effective source of mosquito larvicides as they reduce the activity of chitinase and ecdysone 20- monoxygenase in both A. aegypti and Culex quinquefasciatus [49] also reported that oleic and linoleic acid were quite potent against the fourth instar larvae of Aedes aegypti, Anopheles stephensi and Culex quinquefasciatus say after 24h bioassay. In the contrary [50] reported that the bioassay of oleic acid against a Subterranean Termite, Coptotermes gestroi wasmann and a drywood Termite, Cryptotermes cynocephalus light showed low termiticidal activity as it delivered low mortality in both species and generated lower protection against Cryptotermes cynocephalus.

5. CONCLUSION

The investigation established the potential of H. suaveolens methanol and aqueous leaf extracts against the fourth larval stage of Anopheles species. The types and levels of active components of the extracts may be responsible for their potential against the Anopheles species larvae. Therefore, it is evident from this present investigation that crude extracts from H. suaveolens have promising larvicidal efficacy. Since this present investigation was undertaken under Laboratory conditions, field application of this plant for mosquito vector control should be tested and further investigation should also be done on the effect of the extracts on non-target organisms.

ETHICAL APPROVAL

It is not applicable.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

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