Review

The Potential Role and Regulatory Mechanisms of MUC5AC in Chronic Obstructive Pulmonary Disease

Jingyuan Li 1 and Zuguang Ye 1,2,*

1 Tianjin State Key Laboratory of Modern Chinese Medicine, Tianjin University of Traditional Chinese Medicine, Tianjin 301617, China; lijingyuan886@126.com
2 Institute of Chinese Materia Medica, China Academy of Chinese Medical Sciences, Beijing 100700, China
* Correspondence: zgye@icmm.ac.cn; Tel./Fax: +86-10-8425-2805

Abstract: Chronic obstructive pulmonary disease (COPD) is associated with high morbidity and mortality globally. Studies show that airway mucus hypersecretion strongly compromises lung function, leading to frequent hospitalization and mortality, highlighting an urgent need for effective COPD treatments. MUC5AC is known to contribute to severe mucobronchial lung diseases, worsening COPD pathogenesis. Various pathways are implicated in the aberrant MUC5AC production and secretion MUC5AC. These include signaling pathways associated with mucus-secreting cell differentiation [nuclear factor-κB (NF-κB) and IL-13-STAT6-SPDEF], as well as epithelial sodium channel (ENaC) and cystic fibrosis transmembrane conductance regulator (CFTR), and signaling pathways related to mucus transport and excretion—ciliary beat frequency (CBF). Various inhibitors of mucus hypersecretion are in clinical use but have had limited benefits against COPD. Thus, novel therapies targeting airway mucus hypersecretion should be developed for effective management of mucobronchial lung disease. Here, we systematically review the mechanisms and pathogenesis of airway mucus hypersecretion, with emphasis on multi-target and multi-link intervention strategies for the elucidation of novel inhibitors of airway mucus hypersecretion.

Keywords: COPD; mucobronchial lung diseases; airway mucus hypersecretion; MUC5AC; cell differentiation

1. Introduction

COPD is characterized by high mortality and morbidity globally. The World Health Organization (WHO) predicts that COPD will become the third among the top causes of deaths by 2030 [1]. In 2017, 3.2 million deaths were attributed to COPD, globally. A 2012–2015 Chinese epidemiological survey found the prevalence of COPD in individuals older than 40 years to be 13.7% [2]. COPD progression leads to a significant reduction in the quality of life, worsening the healthcare burden.

COPD is considered a treatable and preventable disease which often presents with unyielding airflow limitation. It is often driven by chronic inflammatory response of the airways and lung tissue. The complex pathogenesis of COPD involves protease-antiprotease imbalance, oxidative stress and chronic inflammation. The disease is characterized by irreversible airway remodeling that involves thickening of the airway wall and the airway smooth muscle (ASM) layer, mucus hypersecretion and epithelial cells metaplasia [3]. Thus, COPD is also regarded as a mucobronchial disease accompanied by increased amounts of mucin, particularly MUC5AC that may uniquely modify mucus properties [4].

COPD risk factors include air pollution (PM2.5), tobacco smoking, exposure to biomass fuel, as well as genetic susceptibility. The 2018 China pulmonary health [CPH] epidemiological survey found that a smoking rate exceeding 20 pack-years increases the prevalence of COPD by 2-fold.
Air pollution affects the health of populations in developing countries, such as China. Long-term and short-term exposure to PM2.5 contributes to the development of COPD. Heavy exposure to PM2.5 increases the occurrence of COPD by 2-fold in never-smokers and in the general population in China [2]. Consumption of biomass fuels increases indoor air pollution which contributes to the development of COPD, according to other studies. Additionally, epigenetic and genetic factors have been implicated in the pathogenesis of COPD.

Efforts to control the public health effects of COPD include developing preventive or mitigation measures to improve patients’ quality of life [5]. Treatment of patients with stable COPD include long-term oxygen therapy, pulmonary rehabilitation, and smoking cessation. However, pharmacological agents are still the main for treatments for all COPD patients after smoking cessation, which are mainly inhaled. The long-acting muscarinic antagonist (LAMA) is the recommended initial treatment choice for patients with mild disease and no exacerbations [6]. In case of more severe dyspnea, severe airflow obstruction, and lung hyperinflation, combining LAMA with a selective long-acting beta2-agonist (LABA) is recommended [7–10]. If the symptoms and exacerbations persist (more than two exacerbations per year or one hospitalization for COPD), a LAMA, a LABA, and an inhaled corticosteroid (ICS) may be prescribed [11–14]. An array of other systemic therapies (azithromycin, roflumilast, xanthines, and antioxidants) may be considered as third-line treatments [1]. The use of antibiotics in for COPD exacerbations remains controversial [15].

Although smoking cessation, lung volume reduction surgery, oxygen inhalation, and drug therapy improve pulmonary function to an extent, reduce the frequency of acute exacerbation of chronic obstructive pulmonary disease (AECOPD) and enhance quality of life, they do not halt COPD progression [16]. Multiple studies indicate that airway mucus hypersecretion contributes to the rapid deterioration of lung function and progression of COPD, including hospitalization and mortality. COPD with airway mucus hypersecretion increases the risk of death by 3.5-fold relative to non-airway mucus hypersecretion disease [17].

Mucins have been linked to mucus hypersecretion [18,19]. Among them, MUC5AC is considered the most important one as it is overproduction in COPD results in mucus obstruction. Thus, MUC5AC is essential to the pathogenesis of COPD [20–22]. In this review, we discuss MUC5AC function, regulatory mechanisms, pathophysiological effects, and clinical significance in COPD.

2. The Biological Characteristics of Mucins

2.1. Classification of Mucins

The main airway mucus secretory cells are airway goblet cells, submucosal glands, and Clara cells. Ciliated cells are responsible for mucociliary clearance. All airway secretory cell types differentiate from airway epithelial cells [23–25]. The airway mucus consists of a sol layer and a gel layer that together form a defense barrier termed the “cilia-mucus blanket” [26,27]. The sol layer is mainly comprised of water and is close to the surface of epithelial cells. The gel layer, which lies above the sol layer, consists of water, mucins, lysozyme, lactoferrin, and various peptides, including mucin; the main source of mucoelasticity. Mucin may be membrane-bound and secreted [28]. Mucins fall into four classes based on structure (Table 1): (1) membrane-constrained mucins which contain tandem repeats (TR) include Muc20, Muc17, Muc18, Muc16, Muc15, Muc13, Muc12, Muc3B, Muc1, Muc3 and Muc4 [29–31]. (2) Cysteine-rich secretory mucins (containing TR sequences and include Muc6, Muc19, Muc5B, Muc5AC, and Muc2 [32,33]. (3) Cysteine deficient secretory mucins (containing TR sequences) consist of Muc7, Muc8 and Muc9 [30]. (4) Mucins without TR sequence include Muc18, Muc15, and Muc14 [33–36].
Table 1. Mucin-type classification.

| Character                                      | Mucin                                      | References |
|------------------------------------------------|--------------------------------------------|------------|
| Membrane-constrained mucins (containing tandem repeats, tandem repeats TR) | Muc16, Muc15, Muc13, Muc12, Muc3B, Muc1    | [29–31]    |
| Cysteine-rich secretory mucins (containing TR sequences) | Muc3, Muc4, Muc17, Muc18 and Muc20          | [32,33]    |
| Cysteine deficient secretory mucins (containing TR sequences) | Muc7, Muc8 and Muc9                        | [30]       |
| Mucins without TR sequence mucins               | Muc14, Muc15 and Muc18                     | [33–36]    |

Cysteine-rich secretory, or gelatinous secretory mucin gives mucus its characteristic viscoelasticity and this property is critical to clearance of xenobiotics by ciliated cells. MUC5AC and MUC5B are the most important and abundant mucins, comprising about 75% of mucins [4,37]. These mucins are abundantly expressed in normal and stressed states. MUC5AC is the 1st line macromolecular substance, and represents 95% of total mucin secretion in the airway epithelium. However, abnormal quantity and quality of MUC5AC negatively affects airway function and may cause serious airway disease. MUC5AC is mainly secreted by the goblet airway epithelium, and its expression changes significantly under stress such as air pollution (PM2.5), tobacco smoking, as well as exposure to biomass fuel [30,38].

2.2. Mucin Secretion and Transport

2.2.1. Airway Epithelial Cell Differentiation

The airway epithelium is comprised of the cartilaginous trachea that branches into primary bronchi, bronchioles, and alveoli, which gaseous exchange sites [39]. The tracheobronchial epithelium consists of multiple cell types, such as secretory, basal, and ciliated cells covering the basement membrane [40] (Figure 1). Under normal physiological conditions, the pseudostratified ciliated bronchial epithelium is comprised of normal proportions of goblet, ciliated and basal cells [40,41]. External insult, for example by tobacco smoke or microbial invasion, triggers a hyperplastic response by goblet cells, a decline in the number of ciliated cells, and ciliated cell dysfunction [42]. This transformation results in over-production of mucus and plays an important role in the COPD development.

Figure 1. The structure of airway epithelium. It is a pseudostratified columnar epithelium containing basal, secretory and ciliated cells. Airway epithelial cell differentiation is influenced by factors such as bacterial infection, inflammatory factors and tobacco exposure.

2.2.2. Mucus Hydration

To maintain viscoelasticity, mucus secreted into the extracellular space through exocytosis must absorb >100 times its mass in water. Because water passively shuttles through the cell membrane along the ion current, its amounts are determined by Cl\(^-\) release and Na\(^+\) uptake in the airway. In human airways, ENaC-mediated Na\(^+\) absorption is a critical regulator of mucosal hydration, leading to effective mucus clearance [43] (Figure 2). In healthy individuals, well balanced epithelial Na\(^+\) absorption and Cl\(^-\) secretion hydrates airway surfaces, promoting efficient mucociliary clearance (MCC) [44]. In the presence of muco-obstructive lung disease, an ion transport imbalance coupled with mucin hypersecretion elevates mucin concentration in the mucus layer, osmotically compresses the
periciliary layer (PCL), and compromises MCC [45,46]. Moreover, other ion channels, including CFTR, Ca$^{2+}$-activated Cl$^{-}$ channel (CaCC) and Slc26A9, affect Cl$^{-}$ release [47]. Additionally, oxidative stress is known to have acute and chronic effects on airway ion transport [48].

2.2.3. Mucus Transport

Ciliated cells are responsible for the proper mucus transport. This process depends on the number of ciliated cells and adequate ciliated cells function, including CBF, ciliary waveform, and cilia.

Figure 2. The process of mucin secretion and transport. There are three major steps in this process: (1) Goblet cell differentiation; (2) Dysfunction of chloride and sodium channels leads to mucus hydration; (3) Ciliated cell wobble and expel mucus.
orientation [49]. Ciliated cells, which have about 200 cilia each, swing cilia in forward in a directionally coordinated manner, driving the removal of microorganisms and particulate matter trapped by the mucus layer from airways. Thus, ciliated cells are constantly beating, pushing mucus up and out through throat (center). Hence, cell differentiation, hydration and ciliary activity are the main determinants of mucus clearance [50] (Figure 2).

3. Clinical Significance of Mucin in COPD

3.1. Mucus Constitutes a Natural Physical Barrier

Airway epithelial cells (AEC) protect the lung by acting both as a mechanical and an immunological barrier. The epithelial mucus layer is the 1st line of defense against bacterial, viral and other toxicants (tobacco smoke, biomass fuel, air pollution) invasion [51,52]. Through its dense network of tight junctions and transcellular adherens, AEC function as a physical barrier between the body and the environment. Hydration and cross-linking of mucins and mucin concentration determine mucus viscoelasticity and thus the firmness of the mucus mechanical barrier. Alterations in mucus surface hydration and mucus concentration occur with inhalation of large particles, as well as bacterial and viral infections. Mucus traps particles and a ciliary escalator that transports it towards the oropharynx for elimination through expectoration or swallowing. This system constitutes the 1st line of innate defense, and is considered the host defense mechanism for the lungs. This process is called mucociliary clearance [53,54].

3.2. Mucus Hypersecretion Increases Airway Resistance

Since airway area is the main determinant of airway resistance, the larger conducting airways are the chief site of airway resistance. Mucus hypersecretion increases the size of the mucus plug and causes its retention in the airways, reducing airway area. Along with the thickened airway walls, the mucus plugs increase airway resistance and decrease airflow through the lungs [55,56]. COPD patients manifest with elevated mucus secretory cells in the large airways and mucus accumulation in the epithelium and lumen of small airways. The level of mucus accumulation in the lumen of small airways closely correlates with the severity of airflow limitation in most COPD patients. Retention of small airway mucus and even formation of mucus plugs have been observed in the lung tissue of patients who have succumbed to COPD [57,58]. Patients with small airway mucus obstruction do not exhibit symptoms like chronic cough and expectoration, probably because they lack cough receptors [47]. Thus, some COPD patients exhibit airway mucus hypersecretion, aggravated airflow obstruction, high airway resistance, chronic cough, and expectoration.

3.3. Mucus Hypersecretion Contributes to Pathogen Colonization

Mucins are highly glycosylated, which not only contributes to the viscoelasticity of mucins but is also though to mediate host-pathogen interactions. Alterations in mucin glycan pattern may result in bacterial colonization [59,60]. MUC5AC is a huge multidomain oligomeric secretory molecule. It consists of a heavily O-glycosylated apoprotein core and extensive intramolecular disulfide bonds between the cysteine-rich amino and carboxy-sequence, and 7 potential N-glycosylation sites. This structure facilitates easy MUC5AC coupling with unique sialylation and sulfation patterns that contribute to host immunity and bacterial colonization. Highly sulfated mucus causes acidification of the airway microenvironment, which significantly reduces the bactericidal efficacy of commonly used antibiotics [61]. This makes it difficult to clear pathogenic bacteria following lower respiratory tract infection, resulting in colonization by the pathogen, persistent infection, and further mucus hypersecretion. These events promote airway mucus retention and viscosity, making hypersecretion an independent risk factor for COPD progression and mortality [62].
4. Signaling Pathways Associated with Airway Mucus Hypersecretion

Various signaling pathways have been implicated in the aberrant secretion and production of MUC5AC and can be grouped into the following based on mucin secretion and transport process: (1) signaling pathways associated with mucus secretion of cell differentiation. These include NF-κB and IL-13-STAT6-SPDEF signaling [63–65]. (2) Ion channel signaling pathways associated with mucus components. These include ENaC, CFTR, and oxidative stress pathways [66–68]. (3) Signaling pathways associated with mucus transport and excretion-CBF [69].

4.1. Signaling Pathways Associated with Mucus Secretion of Cell Differentiation

4.1.1. NF-κB Pathway

In COPD, extracellular signaling factors such as tumor necrosis factor-α (TNF-α) bind to cell surface receptors to induce a cascade of downstream responses. Receptor stimulation triggers IκB kinase (IKK) activation. IKK then phosphorylates serine at the regulatory site of the IκB subunit of the intracellular NF-κB, allowing the IκB subunit to be ubiquitinated and degraded [70–72]. This in turn liberates NF-κB dimers, followed by translocation of the free NF-κB into the nucleus, where it drives the expression of target genes, including MUC5AC.

A study on a mucus hypersecretory cell model by Su Ui Lee et al. found that verproside significantly antagonizes the TNF-α-induced MUC5AC expression by inhibiting NF-κB expression and phosphorylation of its upstream effectors, including IKKβ, TGF-β-activated kinase 1 (TAK1) and IκBα in NCI-H292 cells [73]. A study by Soyoung Kwa et al. revealed that resistin modulated the expression of mucin in human airway epithelial cells through mitogen-activated protein kinase (MAPK)/NF-κB signaling. However, NF-κB inhibition suppressed resistin-induced MUC5AC expression, but not MUC5B expression [74] (Figure 3).

![Figure 3. IL-13-STAT6-SPDEF and NF-κB signaling pathways associated with mucus hypersecretion.](image)

4.1.2. IL-13-STAT6-SPDEF Pathway

SPDEF is an important member of the E26 transformation-specific (ETS) family of transcription factors. SPDEF was initially thought to regulate prostate development and prostate cancer development [75]. However, recent studies show that SPDEF is also expressed in the respiratory system, where it modulates the differentiation of airway epithelial cells [76]. The IL-13-STAT6-SPDEF-MUC5AC signaling pathway is the main signaling pathway driving cell differentiation of epithelial cells into
goblet cells. A study by Seibold MA et al. found that SPDEF knockdown decreased IL-13-induced MUC5AC expression in human airway epithelial cells and enhanced Forkhead box A2 (FOXA2) expression, a transcription factor known to prevent mucus production [77]. A high throughput screening for transcriptional networks modulating differentiation of cultured airway epithelial cells identified RCM-1, a small nontoxic molecule as an inhibitor of excessive mucus production and goblet cell metaplasia in mice after exposure to allergens. This molecule suppressed signaling by IL-13 and STAT6, downmodulating expression of the STAT6 target genes FOXA2 and SPDEF, which modulate goblet cell differentiation [78]. (Figure 3)

4.2. Ion Transport Pathways Associated with Mucus Hypersecretion

The main determinants of effective clearance of airway mucus are sufficient ciliary beat frequency and adequate mucus hydration. Airway surface hydration is mainly governed by ENaC and CFTR ion transport pathways [44,66]. Oxidative stress has been found to cause acute and chronic alterations to airway ion transport. Numerous studies have implicated that oxidative stress is a contributor to the damage of ion transport channels. Antioxidant drugs that reduce mucus viscosity by acting on the disulfide bonds of respiratory mucin oligomers facilitate ion transport [79].

Cigarette smoke contributes to COPD pathogenesis by modifying CFTR function. The CFTR protein functions as an anion channel on the apical surface of airway epithelia and mutation in CFTR cause aberrant Cl− and Na+ transport, leading to airway mucus dehydration and hypersecretion. Impaired CFTR function compromises mucociliary clearance, leads to airway obstruction and chronic airway infection, with Gram-negative bacteria such as P. aeruginosa causing respiratory failure [80]. CFTR belongs to the ATP-binding cassette (ABC) subfamily of transporters. ABC proteins bind ATP and use the energy to drive the transport of various molecules across cell membranes. In CFTR, the interactions of ATP and CFTR’s nucleotide-binding domains control the opening and closing of the channel, rather than driving solute transport. Deletion of F508 on CFTR causes its retention in the endoplasmic reticulum, reducing CFTR open probability, and its residence time in the plasma membrane [81,82]. Upon CFTR activation, Cl− and HCO3− are extruded into the airway lumen, with Na+ and H2O following passively through the paracellular pathway. (Figure 2)

Recent studies suggest that aberrant electrolyte transport as a result of impaired CFTR channel function fails to create the acidic environment needed for phagolysosome activity, providing conducive conditions for bacteria reproduction [83,84].

ENaC-mediated Na+ absorption across the apical plasma membrane modulates mucosal hydration, directly contributing effective mucus clearance. Cl− secretion is facilitated by either CFTR, or CaCCs, including Anoctamin-1/TMEM16A. Ciliated cells co-express Anoctamin-1, CFTR, and ENaC, which clear mucus and maintain lung sterility [85]. ENaC is a heterotrimer of α, β, and γ-subunits. The α- and γ-subunits must undergo proteolytic cleavage to enable ENaC activation and Na+ conductance [86]. Ubiquitination of the α and γ ENaC subunits on their N-termini by the E3 ubiquitin ligase Nedd4-2 (NEDD4L) limits the channel’s half-life. Knockdown of NEDD4-2 in mice has been shown to upregulate ENaC, resulting in lung pathology. Upon ENaC activation, Na+ enters the blood from the airway lumen and Cl−/H2O moves in the same direction, reducing mucus volume [87]. Cl− is the most abundant anion in mucus, at about 120mM, and provides the driving force for osmotic-induced changes in mucus volume. In contrast, HCO3− concentration is about 30mM and may proportionally affect mucus volume. Additionally, HCO3− may play an important role in buffering mucus pH. Mouse models with airway-specific overexpression of ENaC or exhibiting airway surface dehydration (mucus hyperconcentration), impaired MCC and mucus plugging, mimic muco-obstructive lung disease sharing key features of CF and COPD [88].

Oxidative stress also influences COPD pathogenesis. Unlike lightly cross-linked mucus gels, the pathologic mucus in lung disease is not easily transported by the mucociliary escalator, causing its build up, lung infection atelectasis, and airflow obstruction [68,89]. In addition to the aforementioned cysteine-rich domains in mucin N and C termini, cysteine-rich regions are also abundant within
internal domains. These internal cysteine thiols add to the antioxidant effects of mucins. High levels of oxidative stress in COPD mucus might increase mucin oxidative cross linkage through disulfide bonds, resulting in highly cross linked, elastic mucus [90]. Thus, antioxidant therapy may be effective against airway mucus hypersecretion.

4.3. Ciliophagy and CBF

Ciliophagy is a cilia-specific form of autophagy that mediates lung damage from cigarette smoking. It causes cilia shortening and mucus hypersecretion, hence airway epithelial damage [91]. Significant ciliary shortening has been seen in COPD patients with a history of chronic smoking, resulting in impaired mucociliary clearance. Previous studies have shown that lung tissue from COPD patients accumulates autophagy which negatively regulates the formation in cilia [92]. Lam and colleagues showed that elevated autophagy in primary cultured epithelial cells under cigarette smoke (CS) exposure was accompanied by cilia shortening and increased localization of ciliary proteins, including Ifit88, Arl13, centrin 1, and pericentrin, to autophagosomes in mouse airways demonstrating that autophagy is increased in COPD cells under CS exposure, and that cilia shortening is caused by increased autophagic degradation of ciliogenic proteins. Additionally, genetic suppression of autophagy was found to be protective against CS-induced cilia shortening in vitro and in vivo [93].

5. Therapeutic Approaches and Targets of Mucus Hypersecretion

At present, drug therapy of airway mucus hypersecretion is mainly divided into 7 types, and the specific mechanism or targets and representative drugs are detailed in Table 2.

Table 2. Representative drugs and targets for the treatment of mucus hypersecretion.

| Link Acting on Airway Mucus Hypersecretion | Representative Compounds | TARGET or Mechanism | Reference |
|-------------------------------------------|--------------------------|---------------------|-----------|
| Epithelial Differentiation                | S-Allylmercapto-L-cysteine * | NF-κB               | [64]      |
|                                           | Obtusifolin *             |                     | [94]      |
|                                           | Flavonoid 7,4'-Dihydroxyflavone |                   | [94]      |
|                                           | Luteolin *                |                     | [63]      |
|                                           | Platycodon D(3) *         | IL-13-STAT6-SPDEF   | [63,97]   |
|                                           | Tiotropium **             |                     | [98]      |
| Ion Transport to Improve Hydration        | Roflumilast **            | CFTR                | [67]      |
|                                           | Ivacafitor ***            |                    | [67]      |
|                                           | Hypertonic saline**       | ENaC                | [99]      |
|                                           | N-acetylcysteine **       | Antioxidant         | [100]     |
|                                           | Carbocisteine **          | Thiol mucolytics    | [79]      |
| Mucus transport                           | Salmeterol and Formoterol ** | Alter ciliary beat frequency | [62,103,102] |

Note: (1) * Drugs in pre-clinical tests (2) ** most commonly used drugs. (3) *** Ivacaftor: limited to patients with CFTR mutations.

5.1. Mucus Hypersecretion Inhibitors Targeting Cell Differentiation

Various natural products exhibit potential therapeutic benefit against mucus hypersecretion. Most studies seeking to identify inhibitors of mucus hypersecretion target NF-κB [103]. Among natural mucoregulatory agents, S-Allylmercapto-L-cysteine (SAMC), Obtusifolin, Flavonoid 7,4'-Dihydroxyflavone(Flavonoid 7,4'-DHF), Luteolin and Platycodon D have been reported to be the most potent suppressors of MUC5AC expression in airway epithelial cells. A study by Min et al. found that SAMC, an organosulfur compound in garlic, modulates MUC5AC and aquaporin 5 (AQ5P) expression in a COPD model by regulating the NF-κB signaling. SAMC treatment in the 20–100 µM range restored lipopolysaccharides (LPS) -induced suppression of IκBα expression levels in SPC-A1 cells in a dose-dependent manner [64]. A study by Byung-Soo Choi and colleagues using NCI-H292 cells found that 10–50 µM obtusifolin inhibits phorbol 12-myristate 13-acetate (PMA)-induced NF-κB nuclear translocation [94]. Moreover, Flavonoid 7,4'-DHF, a triterpenoid found in G. uralensis, and Luteolin, a flavonoidal found in L. japonica Thunb have exhibited similar effects on cell models, and significantly suppress MUC5AC expression and mucus production in PMA stimulated NCI-H292 airway epithelial cells [63,95].
The root of *P. grandiflorum*, a common medicinal and edible plant in East Asia, is widely used to calm panting in traditional Chinese medicine (TCM). Among its multiple active ingredients, including steroidal saponins, flavonoids, phenolic acids, and sterols, platycodin D(3) is thought to be the main active compound against COPD. A Korean study by Seok JH et al. compared the effects of *P. grandiflorum* de Candolle (APG), platycodin D(3) and deapi-platycodin against airway mucin hypersecretion in PMA-stimulated NCI-H292 cells and sulfur dioxide-stimulated rat bronchitis pulmonary mucin hypersecretion in vivo. This study suggests that Platycodin D(3) could inhibit airway mucus hypersecretion through inhibition of MUC5AC production and promotion of MUC5AC secretion in NCI-H292 cells [96].

Acetylcholine, which is released by parasympathetic nerve fibers, is a classical neurotransmitter in airways that induces mucus secretion and bronchoconstriction via muscarinic receptors [104]. For this reason, anticholinergics are applied in the treatment of obstructive airway diseases [105]. The long-acting anticholinergic agent, tiotropium, has been proven to be effective against COPD. A role for acetylcholine in epithelial cell differentiation and goblet cell metaplasia has been proposed. Tiotropium inhibits ovalbumin-induced goblet cell metaplasia in mice and guinea pigs. Inhibition of endogenous acetylcholine by tiotropium does not affect epithelial cell differentiation after air exposure but inhibits and reverses IL-13-induced goblet cell metaplasia and MUC5AC expression. This effect of tiotropium is thought to be mediated via FOXA2 and FOXA3 [65]. A Chinese multicenter, randomized, double-blind, placebo-controlled clinical trial on patients with mild or moderate COPD treated with tiotropium, found that the annual forced expiratory volume in 1 s (FEV1) decline after bronchodilator use was significantly lower in the tiotropium-treated group relative to the placebo-treated group (29 ± 5 mL/year vs. 51 ± 6 mL/year) [97].

### 5.2. Mucus Hypersecretion Inhibitors Targeting Ion Transport

CFTR mutations cause cystic fibrosis (CF), an airway disease that phenotypically resembles COPD and is characterized by chronic bronchitis. Acquired CFTR dysfunction may influence COPD pathogenesis, making CFTR a potential anti-COPD target [81,82]. Ivacaftor, a CFTR activator, reverses CFTR dysfunction in vitro by activating wild-type CFTR-dependent, short circuit current following chronic exposure. A study by Raju et al. found that cigarette smoke extract (CSE) exposure reduces CFTR-dependent current in human bronchial epithelial (HBE) cells and human bronchi via a mechanism that may involve CSE-induced reduction in CFTR gating, decreasing CFTR open-channel. Ivacaftor, which reverses these adverse effects in vitro, is potentially beneficial for COPD patients with chronic bronchitis [67]. Roflumilast, a phosphodiesterase IV inhibitor, was the first oral COPD drug approved by the U.S. Food & Drug Administration (FDA). This drug was approved on 5 October 2018 for the treatment of COPD at all severities. Roflumilast significantly reduced the frequency of moderate and severe attacks in patients with previous hospitalization in large clinical trials [98]. An in vitro study by Schmid A et al. on differentiated, primary human bronchial epithelial cells found that roflumilast acts on CFTR to enhance the effects of salbutamol against reduced functional Cl⁻ conductivity caused by smoke. Moreover, roflumilast enhances forskolin-induced CBF stimulation in airway surface liquid (ASL) volume supplemented smoked [106].

Inhaled hypertonic saline (HS) is an effective therapy for muco-obstructive lung diseases. It aids epithelial lining fluid (ELF) hydration, which aids in mucociliary clearance, improving MCC, FEV1, and quality of life [107]. A clinical study by Ramsey et al. showed that HS inhalation acutely reduces non-cystic fibrosis bronchiectasis mucus concentration by up to 25% [99]. Some anti-inflammatory and antioxidation mucolytics, including carbocisteine and N-acetylcysteine are widely used to treat respiratory diseases with phlegm production due to their ability to facilitate sputum elimination. A randomized placebo-controlled clinical trial by Zhong NS et al., involving 709 patients, found that relative to placebo, administration of 1500 mg carbocisteine daily for a year significantly reduced the number of exacerbations per (1.01 [SE 0.06] vs. 1.35 [SE 0.06]). Indicating that carbocisteine can reduce yearly exacerbation rates in COPD patients, which is more critical than mucolysis for long-term
management of COPD [79]. A randomized, double-blind placebo-controlled trial by Zhong NS et al., involving 1006 patients found that in patients with moderate-to-severe COPD, long-term use of 600 mg N-acetylcysteine, twice daily, significantly reduces the frequency of AECOPD, especially moderately severe cases [100].

5.3. Mucus Hypersecretion Inhibitors Targeting Ciliary Wobble and Mucus Transport

Moderate-severe COPD is effectively managed using long-acting inhaled bronchodilators. The long-acting beta-agonists, salmeterol and formoterol, can clear mucus in patients or increase ciliary beat frequency in animal models [101]. An open-label, single-center, randomized, cross-over study by Thomas Meyer et al., involving 24 patients with mild-moderate COPD, found that a single daily dose of 12 mg formoterol enhanced mucus clearance within 14 days of treatment [102]. A trial by Piatti et al. evaluating the effects of salmeterol xynaphoate on CBF of nasal epithelium and rheological parameters of tracheobronchial mucus found that it induced ciliostimulation in COPD patients and significantly faster CBF [69].

6. Challenges Facing the Use of Mucus Hypersecretion Inhibitors in Humans and Future Directions in Drug Development

Accumulating evidence shows that mucus hypersecretion is a critical player in COPD, suggesting that drugs targeting mucus hypersecretion may be effective against COPD. Some common drugs, including tiotropium, roflumilast, HS, N-acetylcysteine, carbocisteine, salmeterol and formoterol, are currently used to treat mucus hypersecretion in COPD. However, mucus hypersecretion involves production, transport and secretion steps, that present multiple potential targets. Current strategies have limited capacity to inhibit mucus hypersecretion. For instance, several clinical trials have shown that carbocisteine and N-acetylcysteine reduce the frequency of AECOPD by approximately 28% and 33.7%, respectively [79,100]. However, side effects limit the widespread use of some drugs. For example, atomized inhalation of HS is associated with mild and transient risks including bronchospasm, cough, and breathlessness, limiting its use [108].

COPD causes high morbidity and mortality worldwide, highlighting the urgent need for effective therapeutic strategies. Thus, the search for novel drugs and targets remains an active area of research. Antibody therapies, such as human anti-IL-13 monoclonal antibody, targeting IL-13-STAT6-SPDEF signaling, may be effective against mucus hypersecretion [77].

Small molecules from natural products are an important resource of pharmaceutical ingredients. TCM or natural medicine is increasingly used and widely accepted for its advantages, including multiple active components, links and targets. Thus, it is crucial to explore TCM for mucus hypersecretion inhibitors.

Author Contributions: J.L. drafted the manuscript. Z.Y. contributed to the conception of the review and helped perform the constructive discussions. All authors have read and agreed to the published version of the manuscript.

Funding: This study was supported by the Chinese Major Scientific and Technological Special Project for Major New Drug Creation (No. 2017ZX09201002-007 and No. 2017ZX09201002-006).

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

ABC: ATP-binding cassette; AEC: Airway epithelial cells; AECOPD: acute exacerbation of chronic obstructive pulmonary disease; AQPS: aquaporin 5; ASL: airway surface liquid; ASM: airway smooth muscle; CaCC: Ca$^{2+}$-activated Cl$^{-}$ channel; CBF: ciliary beat frequency; CF: cystic fibrosis; CFTR: cystic fibrosis transmembrane conductance regulator; COPD: Chronic obstructive pulmonary disease; CPH: China pulmonary health; CS: cigarette smoke; CSE: cigarette smoke extract; ELF: epithelial lining fluid; ENaC: epithelial sodium channel; ETS: E26 transformation-specific; FDA: Food & Drug Administration; FEV1: forced expiratory volume in 1 s; Flavonoid 7,4′-DHF: Flavonoid 7,4′-Dihydroxyflavone; FOXA2: Forkhead box A2; HBE: human bronchial epithelial; HS: hypertonic saline; ICS: inhaled corticosteroid; IKK: IκB kinase; LABA: long-acting beta2-agonist; LAMA: long-acting muscarinic antagonist; LPS: lipopolysaccharides; MAPK: mitogen-activated protein kinase; MCC: mucociliary clearance; NEDD4L: N-termini by the E3 ubiquitin ligase Nedd4-2; NF-κB: nuclear factor-κB; PCL: periciliary
layer; PMA: phorbol 12-myristate 13-acetate; SAMC: S-Allylmercaptop-L-cysteine; SPDEF: SAM pointed domain containing E26 transformation-specific transcription factor; TAK1: TGF-β-activated kinase 1; TCM: traditional Chinese medicine; TNF-α: tumor necrosis factor-α; TR: tandem repeats; WHO: World Health Organization.

References
1. Celli, B.R.; Wedzicha, J.A. Update on Clinical Aspects of Chronic Obstructive Pulmonary Disease. *New Engl. J. Med.* 2019, 381, 1257–1266. [CrossRef] [PubMed]
2. Wang, C.; Xu, J.; Yang, L.; Xu, Y.; Zhang, X.; Bai, C.; Kang, J.; Ran, P.; Shen, H.; Wen, F.; et al. Prevalence and risk factors of chronic obstructive pulmonary disease in China (the China Pulmonary Health [CPH] study): A national cross-sectional study. *Lancet* 2018, 391, 1706–1717. [CrossRef]
3. Stylianou, P.; Clark, K.; Goopat, B.; Smallwood, D.; Brightling, C.E.; Amrani, Y.; Roach, K.M.; Bradding, P. Tensin1 expression and function in chronic obstructive pulmonary disease. *Sci. Rep.* 2019, 9, 18942. [CrossRef] [PubMed]
4. Boucher, R.C. Muco-Obstructive Lung Diseases. *New Engl. J. Med.* 2019, 380, 1941–1953. [CrossRef]
5. Funk, G.C.; Kirchheiner, K.; Burghuber, O.C.; Hartl, S. BODE index versus GOLD classification for explaining anxious and depressive symptoms in patients with COPD—A cross-sectional study. *Respir. Res.* 2009, 10, 1. [CrossRef]
6. Troosters, T.; Celli, B.; Lystig, T.; Kesten, S.; Mehra, S.; Tashkin, D.P.; Decramer, M. Tiotropium as a first maintenance drug in COPD: Secondary analysis of the UPLIFT trial. *Eur. Respir. J.* 2010, 36, 65–73. [CrossRef]
7. Horita, N.; Goto, A.; Shibata, Y.; Ota, E.; Nakashima, K.; Nagai, K.; Kaneko, T. Long-acting muscarinic antagonist (LAMA) plus long-acting beta-agonist (LABA) versus LABA plus inhaled corticosteroid (ICS) for stable chronic obstructive pulmonary disease (COPD). *Cochrane Database Syst. Rev.* 2017, 2, CD012066. [CrossRef]
8. Lipson, D.A.; Barnhart, F.; Brealey, N.; Brooks, J.; Criner, G.J.; Day, N.C.; Dransfield, M.T.; Halpin, D.; Han, M.K.; Jones, C.E.; et al. Once-Daily Single-Inhaler Triple versus Dual Therapy in Patients with COPD. *N. Engl. J. Med.* 2018, 378, 1671–1680. [CrossRef]
9. Cazzola, M.; Matera, M.G. POINT: Should LAMA/LABA Combination Therapy Be Used as Initial Maintenance Treatment for COPD? Yes. *Chest* 2018, 154, 746–748. [CrossRef]
10. Wedzicha, J.A.; Banerji, D.; Chapman, K.R.; Vestbo, J.; Roche, N.; Ayers, R.T.; Thach, C.; Fogel, R.; Patalano, F.; Vogelmeier, C.F.; et al. Indacaterol-Glycopyrronium versus Salmeterol-Fluticasone for COPD. *N. Engl. J. Med.* 2016, 374, 2222–2234. [CrossRef]
11. Nici, L.; Mannen, M.J.; Charbek, E.; Alexander, P.E.; Au, D.H.; Boyd, C.M.; Criner, G.J.; Donaldson, G.C.; Dreher, M.; Fan, V.; et al. Pharmacologic Management of Chronic Obstructive Pulmonary Disease. An Official American Thoracic Society Clinical Practice Guideline. *Am. J. Respir. Crit. Care Med.* 2020, 201, e56–e69. [CrossRef] [PubMed]
12. Zheng, Y.; Zhu, J.; Liu, Y.; Lai, W.; Lin, C.; Qiu, K.; Wu, J.; Yao, W. Triple therapy in the management of chronic obstructive pulmonary disease: Systematic review and meta-analysis. *BMJ* 2018, 363, k4388. [CrossRef] [PubMed]
13. Cazzola, M.; Matera, M.G.; Rogliani, P. Adding a LAMA to ICS/LABA Therapy: A Meta-analysis of Triple Combination Therapy in COPD. *Chest* 2019, 155, 758–770. [CrossRef] [PubMed]
14. Cazzola, M.; Rogliani, P.; Calzetta, L.; Matera, M.G. Triple therapy versus single and dual long-acting bronchodilator therapy in COPD: A systematic review and meta-analysis. *Eur. Respir. J.* 2018, 52, 1801586. [CrossRef]
15. Sapey, E.; Bafadhel, M.; Bolton, C.E.; Wilkinson, T.; Hurst, J.R.; Quint, J.K. Building toolkits for COPD exacerbations: Lessons from the past and present. *Thorax* 2019, 74, 898–905. [CrossRef]
16. Lee, S.W.; Shin, S.Y.; Park, T.S.; Choi, Y.Y.; Park, J.C.; Park, J.; Oh, S.Y.; Kim, N.; Lee, S.H.; Lee, J.S.; et al. Clinical Utility of Quantitative CT Analysis for Fissure Completeness in Bronchoscopic Lung Volume Reduction: Comparison between CT and Chartis™. *Korean J. Radiol.* 2019, 20, 1216–1225. [CrossRef]
17. Hogg, J.C.; Chu, F.S.; Tan, W.C.; Sin, D.D.; Patel, S.A.; Pare, P.D.; Martinez, F.J.; Rogers, R.M.; Make, B.J.; Criner, G.J.; et al. Survival after lung volume reduction in chronic obstructive pulmonary disease: Insights from small airway pathology. *Am. J. Respir. Crit. Care Med.* 2007, 176, 454–459. [CrossRef]
18. Saco, T.V.; Breitzig, M.T.; Lockey, R.F.; Kolliputi, N. Epigenetics of Mucus Hypersecretion in Chronic Respiratory Diseases. Am. J. Respir. Cell Mol. Biol. 2018, 58, 299–309. [CrossRef]
19. Song, J.; Cano-Rodriquez, D.; Winkle, M.; Gjaltema, R.A.; Goubert, D.; Jurkowski, T.P.; Heijink, I.H.; Rots, M.G.; Hylkema, M.N. Targeted epigenetic editing of SPDEF reduces mucus production in lung epithelial cells. Am. J. Physiol. Lung Cell Mol. Physiol. 2017, 312, L334–L347. [CrossRef]
20. Zhou, J.S.; Zhao, Y.; Zhou, H.B.; Wang, Y.; Wu, Y.F.; Li, Z.Y.; Xuan, N.X.; Zhang, C.; Hua, W.; Ying, S.M.; et al. Autophagy plays an essential role in cigarette smoke-induced expression of MUC5AC in airway epithelium. Am. J. Physiol. Lung Cell Mol. Physiol. 2016, 310, L1042–L1052. [CrossRef]
21. Radicioni, G.; Cao, R.; Carpenter, J.; Ford, A.A.; Wang, T.; Li, L.; Kesimer, M. The innate immune properties of airway mucosal surfaces are regulated by dynamic interactions between mucins and interacting proteins: The mucin interactome. Mucosal Immunol. 2016, 9, 1442–1454. [CrossRef]
22. Kato, K.; Chang, E.H.; Chen, Y.; Lu, W.; Kim, M.M.; Niibori, M.; Hecker, L.; Kim, K.C. MUC1 contributes to goblet cell metaplasia and MUC5AC expression in response to cigarette smoke in vivo. Am. J. Physiol. Lung Cell Mol. Physiol. 2020, 319, L82–L90. [CrossRef]
23. McCauley, K.B.; Alyssandratos, K.D.; Jacob, A.; Hawkins, F.; Caballero, I.S.; Vedaie, M.; Yang, W.; Slovak, K.J.; Morley, M.; Carraro, G.; et al. Single-Cell Transcriptomic Profiling of Pluripotent Stem Cell-Derived SCGB3A2+ Airway Epithelium. Stem Cell Rep. 2018, 10, 1579–1595.
24. Mihalj, M.; Bujak, M.; Butkovič, J.; Zubčić, Ž.; Tolušić Levak, M.; Čes, J.; Kopić, V.; Baus Lončar, M.; Mihalj, H. Differential Expression of TFF1 and TFF3 in Patients Suffering from Chronic Rhinosinusitis with Nasal Polyposis. Int. J. Mol. Sci. 2019, 20, 5461. [CrossRef]
25. Atkinson, J.J.; Adair-Kirk, T.L.; Kelley, D.G.; Demello, D.; Senior, R.M. Clara cell adhesion and migration to extracellular matrix. Respir. Res. 2008, 9, 1. [CrossRef]
26. Rojas, D.A.; Iturra, P.; Ménendez, A.; Ponce, C.A.; Bustamante, R.; Gallo, M.; Bórquez, P.; Vargas, S.L. Increase in secreted airway mucins and partial Muc5b STAT6/FoxA2 regulation during Pneumocystis primary infection. Sci. Rep. 2019, 9, 2078. [CrossRef]
27. Requena, S.; Ponomarchuk, O.; Castillo, M.; Rebik, J.; Brochiero, E.; Borejdo, J.; Gryczynski, I.; Dzyuba, S.V.; Gryczynski, Z.; Grygorczyk, R.; et al. Imaging viscosity of intragranular mucin matrix in cystic fibrosis cells. Sci. Rep. 2017, 7, 16761. [CrossRef] [PubMed]
28. Bae, C.H.; Kim, H.S.; Song, S.Y.; Kim, Y.D. Phorbol 12-Myristate 13-Acetate Induces MUC16 Expression via PKCδ and p38 in Human Airway Epithelial Cells. Clin. Exp. Otorhinolaryngol. 2012, 5, 161–169. [CrossRef] [PubMed]
29. Ali, M.S.; Pearson, J.P. Upper airway mucin gene expression: A review. Laryngoscope 2007, 117, 932–938. [CrossRef] [PubMed]
30. Rose, M.C.; Voynow, J.A. Respiratory tract mucin genes and mucin glycoproteins in health and disease. Physiol. Rev. 2006, 86, 245–278. [CrossRef]
31. Yonezawa, S.; Higashi, M.; Yamada, N.; Yokoyama, S.; Kitamoto, S.; Kitajima, S.; Goto, M. Mucins in human neoplasms: Clinical pathology, gene expression and diagnostic application. Pathol. Int. 2011, 61, 697–716. [CrossRef] [PubMed]
32. Al-Khayal, K.; Abdulla, M.; Al-Obaid, O.; Zubaidi, A.; Vaali-Mohammed, M.A.; Alsheikh, A.; Ahmad, R. Differential expression of mucins in Middle Eastern patients with colorectal cancer. Oncol. Lett. 2016, 12, 393–400. [CrossRef] [PubMed]
33. Kufe, D.W. Mucins in cancer: Function, prognosis and therapy. Nat. Rev. Cancer 2009, 9, 874–885. [CrossRef] [PubMed]
34. Williams, O.W.; Sharaifkhaneh, A.; Kim, V.; Dickey, B.F.; Evans, C.M. Airway mucus: From production to secretion. Am. J. Respir. Cell Mol. Biol. 2006, 34, 527–536. [CrossRef]
35. Hauber, H.P.; Foley, S.C.; Hamid, Q. Mucin overproduction in chronic inflammatory lung disease. Can. Respir. J. 2006, 13, 327–335. [CrossRef]
36. Kerschner, J.E.; Khampang, P.; Samuels, T. Extending the chinchilla middle ear epithelial model for mucin gene investigation. Int. J. Pediatr. Otorhinolaryngol. 2010, 74, 980–985. [CrossRef]
37. Chillappagari, S.; Preuss, J.; Licht, S.; Müller, C.; Mahavadi, P.; Sarode, G.; Vogelmeier, C.; Guenther, A.; Nahrlich, L.; Rubin, B.K.; et al. Altered protease and antiprotease balance during a COPD exacerbation contributes to mucus obstruction. Respir. Res. 2015, 16, 85. [CrossRef] [PubMed]
42. Hallstrand, T.S.; Hackett, T.L.; Altemeier, W.A.; Matute-Bello, G.; Hansbro, P.M.; Knight, D.A.
40. Zuyderduyn, S.; Ninaber, D.K.; Schrumpf, J.A.; van Sterkenburg, M.A.; Verhoosel, R.M.; Prins, F.A.;
57. Lennox, A.T.; Coburn, S.L.; Leech, J.A.; Heidrich, E.M.; Kleyman, T.R.; Wenzel, S.E.; Pilewski, J.M.;
46. Liu, C.; Li, Q.; Zhou, X.; Kolosov, V.P.; Perelman, J.M. Rhythmic pressure waves induce mucin5AC expression
38. Young, H.W.; Williams, O.W.; Chandra, D.; Brockway, B.; Que, L.; McQualter, J.L.; Stripp, B.R.
56. Li, L.D.; Crouzier, T.; Sarkar, A.; Dunphy, L.; Han, J.; Ribbeck, K. Spatial configuration and composition of
55. van Putten, J.; Strijbis, K. Transmembrane Mucins: Signaling Receptors at the Intersection of Inflammation
53. Kogiso, H.; Ikeuchi, Y.; Sumiya, M.; Hosogi, S.; Tanaka, S.; Shimamoto, C.; Inui, T.; Marunaka, Y.;
52. Eenjes, E.; Mertens, T.; Buscop-van Kempen, M.J.; van Wetering, S.; Rabe, K.F.; Hiemstra, P.S. IL-4 and IL-13 exposure during mucociliary differentiation of bronchial epithelial cells increases antimicrobial activity and expression of antimicrobial peptides. Respir. Res. 2011, 12, 59. [CrossRef]
41. Ray, S.; Chiba, N.; Yao, C.; Guan, X.; McConnell, A.M.; Brockway, B.; Que, L.; McQualter, J.L.; Stripp, B.R.
39. Lindström, M.; Falk, R.; Hjelte, L.; Philipson, K.; Svartengren, M. Long-term clearance from small airways in subjects with ciliary dysfunction. Respir. Res. 2006, 7, 79. [CrossRef]
44. Blé, F.X.; Cannet, C.; Collingwood, S.; Danahay, H.; Beckmann, N. ENaC-mediated effects assessed by MRI in a rat model of hypertonic saline-induced lung hydration. Br. J. Pharmacol. 2010, 160, 1008–1015. [CrossRef] [PubMed]
45. Vanaki, S.M.; Holmes, D.; Saha, S.C.; Chen, J.; Brown, R.J.; Jayathilake, P.G. Muco-ciliary clearance: A review of modelling techniques. J. Biomech. 2020, 99, 109578. [CrossRef]
46. Liu, C.; Li, Q.; Zhou, X.; Kolosov, V.P.; Perelman, J.M. Rhythmic pressure waves induce mucin5AC expression via an EGFR-mediated signaling pathway in human airway epithelial cells. DNA Cell Biol. 2013, 32, 423–429. [CrossRef]
47. Randell, S.H.; Boucher, R.C.; University of North Carolina Virtual Lung Group. Effective mucus clearance is essential for respiratory health. Am. J. Respir. Cell Mol. Biol. 2006, 35, 20–28. [CrossRef]
48. O’Grady, S.M. Oxidative stress, autophagy and airway ion transport. Am. J. Physiol. Cell Physiol. 2019, 316, C16–C32. [CrossRef]
49. Li, Y.Y.; Liu, J.; Li, C.W.; Subramaniam, S.; Chao, S.S.; Yu, F.G.; Cohen, N.A.; Li, S.; Wang, Y. Myrtol standardized DNA Cell Biol. 2006, 35, 20–28. [CrossRef]
50. Milara, J.; Armengot, M.; Bañuls, P.; Tenor, H.; Beume, R.; Artigues, E.; Cortijo, J. Roflumilast N-oxide, a PDE4 inhibitor, improves cilia motility and ciliated human bronchial epithelial cells compromised by cigarette smoke in vitro. Br. J. Pharmacol. 2012, 166, 2243–2262. [CrossRef]
51. Mijošek, V.; Lasitschka, F.; Warth, A.; Zabeck, H.; Dalpke, A.H.; Weitnauer, M. Endoplasmic Reticulum Stress Is A Danger Signal Promoting Innate Inflammatory Responses in Bronchial Epithelial Cells. J. Innate Immun. 2016, 8, 464–478. [CrossRef]
52. Eenjes, E.; Mertens, T.; Buscop-van Kempen, M.J.; van Wijck, Y.; Taube, C.; Rottier, R.J.; Hiemstra, P.S. A novel method for expansion and differentiation of mouse tracheal epithelial cells in culture. Sci. Rep. 2018, 8, 7349. [PubMed]
53. Kogiso, H.; Ikeuchi, Y.; Sumiya, M.; Hosogi, S.; Tanaka, S.; Shimamoto, C.; Inui, T.; Marunaka, Y.; Nakahari, T. Seihai-to (TJ-90)-Induced Activation of Airway Ciliary Beatings of Mice: Ca²⁺-Dependent Mechanisms. Cells 2017, 6, 1–15. [CrossRef] [PubMed]
58. Willemse, B.W.; ten Hacken, N.H.; Rutgers, B.; Postma, D.S.; Timens, W. Association of current smoking with airway inflammation in chronic obstructive pulmonary disease and asymptomatic smokers. *Respir. Res.* 2005, 6, 38. [CrossRef] [PubMed]

59. Quintana-Hayashi, M.P.; Mahu, M.; De Pauw, N.; Boyen, F.; Pasmans, F.; Martel, A.; Premaratne, P.; Fernandez, H.R.; Teymournejad, O.; Vande Maele, L.; et al. The levels of Brachyspira hyodysenteriae binding to porcine colonic mucins differ between individuals, and binding is increased to mucins from infected pigs with de novo MUC5AC synthesis. *Infect. Immun.* 2015, 83, 1610–1619. [PubMed]

60. Xu, X.; Chen, W.; Leng, S.; Padilla, M.T.; Saxton, B.; Hutt, J.; Tessema, M.; Kato, K.; Kim, K.C.; Belinsky, S.A.; et al. Muc1 knockout potentiates murine lung carcinogenesis involving an epiregulin-mediated EGFR activation feedback loop. *Carcinogenesis* 2017, 38, 604–614.

61. Venkatakrishnan, V.; Thaysen-Andersen, M.; Chen, S.C.; Nevalainen, H.; Packer, N.H. Cystic fibrosis and bacterial colonization define the sputum N-glycosylation phenotype. *Glycobiology* 2015, 25, 88–100. [CrossRef] [PubMed]

62. Mensa, J.; Trilla, A. Should patients with acute exacerbation of chronic bronchitis be treated with antibiotics? Advantages of the use of fluoroquinolones. *Clin. Microbiol. Infect.* 2006, 12, 42–54. [CrossRef] [PubMed]

63. Lee, H.; Seo, H.S.; Ryu, J.; Yoon, Y.P.; Park, S.H.; Lee, C.J. Luteolin inhibited the gene expression, production and secretion of MUC5AC mucin via regulation of nuclear factor kappa B signaling pathway in human airway epithelial cells. *Pulm. Pharmacol. Ther.* 2015, 31, 117–122. [CrossRef] [PubMed]

64. Yang, M.; Wang, Y.; Zhang, Y.; Zhao, Z.; Li, S.; Zhang, J.; Cao, X.; Zhang, D. S-allylmercapto-l-cysteine modulates MUC5AC and AQP5 secretions in a COPD model via NF-κB signaling pathway. *Int. Immunopharmacol.* 2016, 39, 307–313. [CrossRef] [PubMed]

65. Kistemaker, L.E.; Hiemstra, P.S.; Bos, I.S.; Bouwman, S.; van den Berge, M.; Hylkema, M.N.; Meurs, H.; Kerstjens, H.A.; Gosens, R. Tiotropium attenuates IL-13-induced goblet cell metaplasia of human airway epithelial cells. *Thorax* 2015, 70, 668–676. [CrossRef]

66. Åstrand, A.B.; Hemmerling, M.; Root, J.; Wingren, C.; Pesic, J.; Johansson, E.; Garland, A.L.; Ghosh, A.; Tarran, R. Linking increased airway hydration, ciliary beating, and mucociliary clearance through ENaC inhibition. *Am. J. Physiol. Lung Cell Mol. Physiol.* 2015, 308, L22–L32. [CrossRef]

67. Raju, S.V.; Lin, V.Y.; Liu, L.; McNicholas, C.M.; Karki, S.; Sloane, P.A.; Tang, L.; Jackson, P.L.; Wang, W.; Wilson, L.; et al. The Cystic Fibrosis Transmembrane Conductance Regulator Potentiator Ivacaftor Augments Mucociliary Clearance Abrogating Cystic Fibrosis Transmembrane Conductance Regulator Inhibition by Cigarette Smoke. *Am. J. Respir. Cell Mol. Biol.* 2017, 56, 99–108. [CrossRef]

68. Biswas, S.; Hwang, J.W.; Kirkham, P.A.; Rahman, I. Pharmacological and dietary antioxidant therapies for chronic obstructive pulmonary disease. *Curr. Med. Chem.* 2013, 20, 1496–1530. [CrossRef]

69. Piatti, G.; Ambrosetti, U.; Santus, P.; Allegra, L. Effects of salmeterol on cilia and mucus in COPD and pneumonia patients. *Pharmacol. Res.* 2005, 51, 165–168. [CrossRef]

70. Bae, G.D.; Park, E.Y.; Kim, K.; Jang, S.E.; Jun, H.S.; Oh, Y.S. Upregulation of caveolin-1 and its colocalization with cytokine receptors contributes to beta cell apoptosis. *Sci. Rep.* 2019, 9, 16785. [CrossRef]

71. Tsai, Y.C.; Wang, S.L.; Wu, M.Y.; Liao, C.H.; Lin, C.H.; Chen, J.J.; Fu, S.L.; Pillonl, A. Flavonoid Isolated from Aquilaria sinensis, Exhibits Anti-Inflammatory Activity In Vitro and In Vivo. *Molecules* 2018, 23, 3177. [CrossRef] [PubMed]

72. Lee, S.U.; Lee, S.; Ro, H.; Choi, J.H.; Ryu, H.W.; Kim, M.O.; Yuk, H.J.; Lee, J.; Hong, S.T.; Oh, S.R. Piscroside C inhibits TNF-α/NF-κB pathway by the suppression of PKCδ activity for TNF-RSC formation in human airway epithelial cells. *Phytomedicine* 2018, 40, 148–157. [CrossRef] [PubMed]

73. Lee, S.U.; Sung, M.H.; Ryu, H.W.; Lee, J.; Kim, H.S.; In, H.J.; Shin, D.H.; et al. Verproside inhibits TNF-α-induced MUC5AC expression through suppression of the TNF-α/NF-κB pathway in human airway epithelial cells. *Cytokine* 2016, 77, 168–175. [CrossRef] [PubMed]

74. Kwak, S.; Kim, Y.D.; Na, H.G.; Bae, C.H.; Song, S.Y.; Choi, Y.S. Resistin upregulates MUC5AC/B mucin gene expression in human airway epithelial cells. *Biochem. Biophys. Res. Commun.* 2018, 499, 655–661. [CrossRef]

75. Cheng, X.H.; Black, M.; Ustiyan, V.; Le, T.; Fulford, L.; Sridharan, A.; Medvedovic, M.; Kalinichenko, V.V.; Whitsett, J.A.; Kalin, T.V. SPDEF inhibits prostate carcinogenesis by disrupting a positive feedback loop in regulation of the Foxm1 oncogene. *PLoS Genet.* 2014, 10, e1004656. [CrossRef]
76. Park, K.S.; Korfhagen, T.R.; Bruno, M.D.; Kitzmiller, J.A.; Wan, H.; Wert, S.E.; Hershey, G.K.K.; Chen, G.; Whitsett, J.A. SPDEF regulates goblet cell hyperplasia in the airway epithelium. J. Clin. Investig. 2007, 117, 978–988. [CrossRef]

77. Seibold, M.A. Interleukin-13 Stimulation Reveals the Cellular and Functional Plasticity of the Airway Epithelium. Ann. Am. Thorac. Soc. 2018, 15, S98–S102. [CrossRef]

78. Sun, L.; Ren, X.; Wang, I.C.; Pradhan, A.; Zhang, Y.; Flood, H.M.; Han, B.; Whitsett, J.A.; Kalin, T.V.; Kalinchenco, V.V. The FOXM1 inhibitor RCM-1 suppresses goblet cell metaplasia and prevents IL-13 and STAT6 signaling in allergen-exposed mice. Sci. Signal. 2017, 10, eaai5853. [CrossRef]

79. Zheng, J.P.; Kang, J.; Huang, S.G.; Chen, P.; Yao, W.Z.; Yang, L.; Bai, C.X.; Wang, C.Z.; Wang, C.; Chen, B.Y.; et al. Effect of carbocisteine on acute exacerbation of chronic obstructive pulmonary disease (PEACE Study): A randomised placebo-controlled study. Lancet 2008, 371, 2013–2018. [CrossRef]

80. Wellmerling, J.H.; Chang, S.W.; Kim, E.; Osman, W.H.; Boyaka, P.N.; Borchers, M.T.; Cormet-Boyaka, E. Reduced expression of the ion channel CFTR contributes to airspace enlargement as a consequence of aging and in response to cigarette smoke in mice. Respir. Res. 2019, 20, 200–206. [CrossRef]

81. Patel, S.D.; Bono, T.R.; Rowe, S.M.; Solomon, G.M. CFTR targeted therapies: Recent advances in cystic fibrosis and possibilities in other diseases of the airways. Eur. Respir. Rev. 2020, 29, 190068. [CrossRef] [PubMed]

82. Fernandez, E.F.; De Santi, C.; De Rose, V.; Greene, C.M. CFTR dysfunction in cystic fibrosis and chronic obstructive pulmonary disease. Expert Rev. Respir. Med. 2018, 12, 483–492.

83. Han, M.K.; Huang, Y.J.; Lipuma, J.J.; Boushey, H.A.; Boucher, R.C.; Cookson, W.O.; Curtis, J.L.; Erb-Downward, J.; Lynch, S.V.; Sethi, S.; et al. Significance of the microbiome in obstructive lung disease. Thorax 2012, 67, 456–463.

84. Trinh, N.T.; Bilodeau, C.; Maille, É.; Ruffin, M.; Quintal, M.C.; Desrosiers, M.Y.; Rousseau, S.; Brochiero, E. Delterious impact of Pseudomonas aeruginosa on cystic fibrosis transmembrane conductance regulator function and rescue in airway epithelial cells. Eur. Respir. J. 2015, 49, 1590–1602.

85. Hahn, A.; Salomon, J.J.; Leitz, D.; Feigenbutz, D.; Korsch, L.; Lisewski, I.; Schrimpf, K.; Millar-Büchner, P.; Hahn, A.; et al. Expression and function of Anoctamin 1/TMEM16A calcium-activated chloride channels in airways of in vivo mouse models for cystic fibrosis research. Pflugers Arch. 2018, 470, 1335–1348.

86. Kitamura, K.; Tomita, K. Proteolytic activation of the epithelial sodium channel and therapeutic application of a serine protease inhibitor for the treatment of salt-sensitive hypertension. Clin. Exp. Nephrol. 2012, 16, 44–48. [PubMed]

87. Kimura, T.; Kawabe, H.; Jiang, C.; Zhang, W.; Xiang, Y.Y.; Lu, C.; Salter, M.W.; Brose, N.; Lu, W.Y.; Rotin, D. Deletion of the ubiquitin ligase Nedd4L in lung epithelia causes cystic fibrosis-like disease. Proc. Natl. Acad. Sci. USA 2011, 108, 3216–3221. [PubMed]

88. Nakashima, R.; Kamei, S.; Nohara, H.; Fujikawa, H.; Maruta, K.; Kawakami, T.; Eto, Y.; Takahashi, N.; Suico, M.A.; Takeo, T.; et al. HDAC6-dependent ciliophagy is involved in ciliary loss and cholangiocarcinoma growth and Hedgehog signaling. Lastres-Becker, I.; Cuadrado, A.; Garcia-Gonzalo, F.R. NRF2-dependent gene expression promotes ciliogenesis. Sci. Rep. 2019, 9, 1335–1348. [PubMed]

89. Song, Y.; Wang, W.; Xie, Y.; Xiang, B.; Huang, X.; Guan, W.; Zheng, J. Carbocisteine inhibits the expression of Muc5b in COPD mouse model. Drug Des. Dev. Ther. 2019, 13, 3259–3268.

90. Ehre, C.; Rushton, Z.L.; Wang, B.; Hothen, L.N.; Morrison, C.B.; Fontana, N.C.; Markovetz, M.R.; Delion, M.F.; Kato, T.; Villalon, D.; et al. An Improved Inhaled Mucolytic to Treat Airway Muco-obstructive Diseases. Am. J. Respir. Crit. Care Med. 2019, 199, 171–180.

91. Martin-Hurtado, A.; Martin-Morales, R.; Robledinos-Antón, N.; Blanco, R.; Palacios-Blanco, I.; Lastres-Becker, I.; Cuadrado, A.; García-Gonzalo, F.R. NRF2-dependent gene expression promotes ciliogenesis and Hedgehog signaling. Sci. Rep. 2019, 9, 13896. [CrossRef] [PubMed]

92. Peixoto, E.; Jin, S.; Thelen, K.; Biswas, A.; Richard, S.; Morleo, M.; Mansini, A.; Holtorf, S.; Carbone, F.; Pastore, N.; et al. HDAC6-dependent ciliophagy is involved in ciliary loss and cholangiocarcinoma growth in human cells and murine models. Am. J. Physiol. Gastrointest. Liver Physiol. 2020, 318, G1022–G1033. [PubMed]

93. Lam, H.C.; Cloonan, S.M.; Bhashyam, A.R.; Haspel, J.A.; Singh, A.; Sathirapongsasuti, J.F.; Cervo, M.; Yao, H.; Chung, A.L.; Mizumura, K.; et al. Histone deacetylase 6-mediated selective autophagy regulates COPD-associated cilia dysfunction. J. Clin. Investig. 2013, 123, 5212–5230. [CrossRef] [PubMed]
94. Choi, B.S.; Kim, Y.J.; Choi, J.S.; Lee, H.J.; Lee, C.J. Obtusifolin isolated from the seeds of Cassia obtusifolia regulates the gene expression and production of MUC5AC mucin in airway epithelial cells via affecting NF-kB pathway. *Phytother. Res.* 2019, 33, 3016. [CrossRef] [PubMed]

95. Liu, C.; Weir, D.; Busse, P.; Yang, N.; Zhou, Z.; Emala, C.; Li, X.M. The Flavonoid 7,4′-Dihydroxyflavone Inhibits MUC5AC Gene Expression, Production, and Secretion via Regulation of NF-κB, STAT6, and HDAC. *Phytother. Res.* 2015, 29, 925–932.

96. Ryu, J.; Lee, H.J.; Park, S.H.; Kim, J.; Lee, D.; Lee, S.K.; Kim, Y.S.; Hong, J.H.; Seok, J.H.; Lee, C.J. Effects of the root of Platycodon grandiflorum on airway mucin hypersecretion in vivo and platycodin D(3) and deapi-platycodin on production and secretion of airway mucin in vitro. *Phytomedicine* 2014, 31, 529–533.

97. Zhou, Y.; Zhong, N.S.; Li, X.; Chen, S.; Zheng, J.; Zhao, D.; Yao, W.; Zhi, R.; Wei, L.; He, B.; et al. Tiotropium in Early-Stage Chronic Obstructive Pulmonary Disease. *N. Engl. J. Med.* 2017, 377, 923–935.

98. Ramsey, K.A.; Chen, A.; Radicioni, G.; Lourie, R.; Martin, M.; Broomfield, A.; Schending, Y.H.; Hasnain, S.Z.; Radford-Smith, G.; Simms, L.A.; et al. Airway Mucus Hyperconcentration in Non-Cystic Fibrosis Bronchiectasis. *Am. J. Respir. Crit. Care Med.* 2020, 201, 661–670. [CrossRef]

99. Pudi, K.K.; Barnes, C.N.; Moran, E.J.; Haumann, B.; Kerwin, E. A 28-day, randomized, double-blind, placebo-controlled, parallel group study of nebulized revefenacin in patients with chronic obstructive pulmonary disease. *Respir. Res.* 2017, 18, 182. [CrossRef]

100. Meyer, T.; Reitmeir, P.; Brand, P.; Herpich, C.; Schulze, A.; Scheuch, G.; Newman, S. Effects of formoterol and tiotropium bromide on mucus clearance in patients with COPD. *Respir. Med.* 2011, 105, 900–906. [CrossRef] [PubMed]

101. Albertson, T.E.; Chenoweth, J.A.; Adams, J.Y.; Sutter, M.E. Muscarinic antagonists in early stage clinical development for the treatment of asthma. *Expert Opin. Investig. Drugs* 2017, 26, 35–49. [CrossRef] [PubMed]

102. Cross, N.J. Anticholinergic agents in asthma and COPD. *Eur. J. Pharmacol.* 2006, 533, 36–39. [CrossRef]

103. Samsuzzaman, M.; Uddin, M.S.; Shah, M.A.; Mathew, B. Natural inhibitors on airway mucin: Molecular insight into the therapeutic potential targeting MUC5AC expression and production. *Life Sci.* 2019, 231, 116485. [CrossRef] [PubMed]

104. Albertson, T.E.; Chenoweth, J.A.; Adams, J.Y.; Sutter, M.E. Muscarinic antagonists in early stage clinical development for the treatment of asthma. *Expert Opin. Investig. Drugs* 2017, 26, 35–49. [CrossRef] [PubMed]

105. Albertson, T.E.; Chenoweth, J.A.; Adams, J.Y.; Sutter, M.E. Muscarinic antagonists in early stage clinical development for the treatment of asthma. *Expert Opin. Investig. Drugs* 2017, 26, 35–49. [CrossRef] [PubMed]

106. Gross, N.J. Anticholinergic agents in asthma and COPD. *Eur. J. Pharmacol.* 2006, 533, 36–39. [CrossRef]

107. Schmid, A.; Bauml, N.; Ivomnet, P.; Dennis, J.S.; Campos, M.; Krick, S.; Salathe, M. Roflumilast partially reverses smoke-induced mucociliary dysfunction. *Respir. Res.* 2015, 16, 135. [CrossRef]

108. Valderramas, S.R.; Atallah, A.N. Effectiveness and safety of hypertonic saline inhalation combined with exercise training in patients with chronic obstructive pulmonary disease: A randomized trial. *Respir. Care* 2009, 54, 327–333.

109. Tarrant, B.J.; Le Maitre, C.; Romero, L.; Steward, R.; Button, B.M.; Thompson, B.R.; Holland, A.E. Mucoactive agents for chronic, non-cystic fibrosis lung disease: A systematic review and meta-analysis. *Respirology* 2017, 22, 1084–1092. [CrossRef]