An in silico LLPS perturbation approach in the design of a novel SARS-CoV-2 spike receptor-binding domain inhibitor

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Abstract

The reversible process where a homogenous fluid de-mixes into two distinctively separate liquid phases is referred to as LLPS (Liquid-liquid phase separation). The resulting liquid is made up of one dilute phase and one condensed phase. An increasing number of studies have shown that the liquid-liquid phase separation is an important principle that underlies intracellular organization in biological systems, forming liquid condensates without a membrane envelope, otherwise known as MLOs (membraneless organelles). Such organelles include the P bodies, nucleolus and stress granules. Moreover, the regulation of many other biological processes such as signal transduction, chromatin rearrangement and RNA metabolism have been linked to the liquid-liquid phase separation.

Dear Editor,

Background

The reversible process where a homogenous fluid de-mixes into two distinctively separate liquid phases is referred to as LLPS (liquid-liquid phase separation). The resulting liquid is made up of one dilute phase and one condensed phase. An increasing number of studies have shown that the liquid-liquid phase separation is an important principle that underlies intracellular organization in biological systems, forming liquid condensates without a membrane envelope, otherwise known as MLOs (membraneless organelles). Such organelles include the P bodies, nucleolus, and stress granules. Moreover, the regulation of many other biological processes such as signal transduction, chromatin rearrangement, and RNA metabolism has been linked to the liquid-liquid phase separation [1].

A growing number of studies in recent years have focused on the mechanism of phase separation of a variety of biomolecules. Such studies have demonstrated that some proteins, such as the RNA helicase DEAD-Box 4 (DDX4), P granule protein LAF-1, transactive response DNA-binding protein (TDP-43), and the RNA-binding FUS protein, can undergo liquid-liquid phase separation both in vitro and in vivo. The resultant liquid condensates from the liquid-liquid phase separation process generally are deemed as a product of multivalent weak interactions between the numerous interacting motifs in IDRs (intrinsically disordered regions) or multiple folded domains. The LCRs (low complexity regions), which are generally covered by the intrinsically disordered regions, are suggested to play very important roles in driving liquid-liquid phase separation through Pi-Pi, hydrophobic, cation-Pi, and electrostatic interactions. These regions exhibit an over-representation of specific amino acid residues compared to the proteome proportion, such as the proline-arginine (PR)/glycine-arginine (GR) repeats,
arginine-glycine-glycine (RGG) motifs, and prion-like domains [2].

The spike protein of the SARS-CoV-2 is an envelope glycoprotein that contributes mostly during the attachment process of the virus, its fusion, and host cell entry. It is also an important target for the development of vaccines, neutralizing antibodies, and inhibitors of viral entry. Its synthesis begins as a precursor protein which is cleaved into an amino-terminal S1 subunit composed of 700 amino acid residues and a carboxyl-terminal S2 subunit which is made up of 600 amino acid residues. Both residues respectively mediate the attachment and membrane fusion of the viral protein [3]. Our study is targeted at the design of a novel drug-like compound that can interfere with the molecular grammar that governs the liquid-liquid phase separation of the SARS-CoV-2 spike receptor-binding domain.

Methods
Trivedi et al. [4] described the phase separation-driven inner centromere localization of the chromosomal passenger complex (CPC) by borealin, where the two regions making up the central disordered regions of the protein (as predicted by catGRANULE to display a high phase separation propensity) were deleted. The variant protein complex lacking this predicted borealin disordered region was deficient in both DNA-induced and spontaneous phase separation. We harnessed this approach in the prediction of the mechanism of action of our designed novel SARS-CoV-2 spike receptor-binding domain inhibitor. The inhibitor was designed through the structural modification of 1,6-hexanediol (Fig. 1), an aliphatic alcohol which is known for its ability to disrupt many phase-separated cellular organelles through the inhibition of weak hydrophobic interactions. Treatment of the chromosomal passenger complex with this alcohol reduced its enrichment in the inner centromere and likewise disrupted the in vitro coacervation of the ISB [4].

Waghorne et al. [5] described the design of novel chemotherapeutic agents (N,N-bis(glycyl)amines) that were prepared through the reductive amination of hexoses and pentoses and were subsequently screened for anticancer activity against cancerous virus. The amination approach was applied in the development of our novel inhibitor as shown in Fig. 1.

Results and discussion
A comparative drug-likeness prediction test was carried out using in silico ADMET prediction tools (SwissADME and pkCSM) [6, 7]. The predictive outcome shows that both compounds share similar pharmacokinetic and toxicity properties (Tables 1 and 2; Fig. 2).

We directed a blind docking protocol towards the antiviral activity prediction of our novel compound. With this approach, the docked compound can interact with the most suitable region on the surface of the protein (Fig. 3). The interacting residues were then used in

Table 1 Drug-likeness prediction

| ADME parameters | C₆H₁₆O₂ | C₆H₁₅NO₂ |
|-----------------|---------|---------|
| Solubility      | Soluble | Soluble |
| GI absorption   | High    | High    |
| BBB permeant    | Yes     | No      |
| P-gp substrate  | No      | No      |
| Total clearance (log ml/min/kg) | 0.516 | 0.87 |

Table 2 Drug toxicity prediction

| Toxicity parameters | C₆H₁₆O₂ | C₆H₁₅NO₂ |
|---------------------|---------|---------|
| Hepatotoxicity      | Inactive| Inactive|
| Carcinogenicity     | Inactive| Inactive|
| Cytotoxicity        | Inactive| Inactive|
| Mutagenicity        | Inactive| Inactive|
the prediction of the phase separation perturbation property of the compound.

As depicted in Fig. 4, five residues were involved in the inter-model interaction: hydrophobic interaction with lysine and tyrosine, and hydrogen bonding with serine and glutamate, while arginine was involved in both forms of interaction.

The phase separation propensity of the wild-type SARS-CoV-2 spike receptor-binding domain was predicted using the catGRANULE algorithm [8] as described by Trivedi et al. [4]. Following the amino acid deletion approach as effected on borealin to give rise to a mutant, we generated a variant of the viral protein through the deletion of the interacting residues (Fig. 5) and subjected it to the catGRANULE algorithm. Results emanating from the predicted propensity score and profile showed that the drug interacted with important residues that drives the liquid-liquid phase separation, hence the functionality of the viral protein. The residues were also observed to fall within the intrinsically disordered region of the protein (Fig. 6).

Wang et al. [9] used an extensive mutagenesis approach in the identification of the sequence-encoded molecular grammar that underlies the driving forces of the FUS family proteins’ phase separation and concluded that it is preferentially governed by multivalent interactions among aromatic (tyrosine) and positively charged (arginine) residues. Based on the results from our in silico study, we hereby hypothesize that the molecular grammar governing the phase separation of the SARS-CoV-2 spike receptor-binding domain requires a more robust multivalent interaction, as predicted result revealed different residues with
property variations (positively charged, negatively charged, non-charged, and aromatic residues), but common to all is their polarity.

**Conclusion**

We have computationally identified a novel inhibitory drug-like compound (6-(aminooxy) hexan-1-ol) against the SARS-CoV-2 spike receptor-binding domain through its predicted interaction with the amino acid residues that drives the viral protein’s liquid-liquid phase separation. The compound was analyzed and predicted to possess satisfactory physicochemical, ADME, and toxicity properties. Findings from this study has shown that the
compound has demonstrated drug-like attributes of being a potential inhibitor to be optimized for trial against the SARS-CoV-2 spike receptor-binding domain. We however recommend further experimental assessment of the antiviral potential of this compound.

Abbreviations
ADME: Absorption-distribution-metabolism-excretion; ADMET: Absorption-distribution-metabolism-excretion-toxicity; BBB: Blood-brain barrier; CPC: Chromosomal passenger complex; DDX4: DEAD-Box 4; DNA: Deoxyribonucleic acid; GI: Gastrointestinal; IDRs: Intrinsically disordered regions; ISB: INCENP-survivin-borealin; LCRs: Low complexity regions; LLPS: Liquid-liquid phase separation; MLO: Membraneless organelle; PDB: Protein Data Bank; P-gp: P-glycoprotein; PR/GR: Proline-arginine/glycine-arginine; RGG: Arginine-glycine-glycine; RNA: Ribonucleic acid; SARS-CoV-2: Severe acute respiratory syndrome coronavirus 2; TDP-43: Transactive response DNA-binding protein-43

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Authors’ contributions
OAD: literature review and experimental design. DMS: manuscript editing and experimental design. TM: manuscript editing and writing. HOU: manuscript writing. SC: manuscript editing. JNA: literature review. GOI: literature review. All authors have read and approved the manuscript.

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References
1. Tatavosian R, Kent S, Brown K, Yao T, Duc HN, Huynh TN, Zhen CY, Ma B, Wang H, Ren X (2019) Nuclear condensates of the polycomb protein chromobox 2 (CBX2) assemble through phase separation. J Biol Chem 294:1451–1463
2. Li Q, Peng X, Li Y, Tang W, Zhu J a, Huang J, Qi Y, Zhang Z (2020) LLPSDB: a database of proteins undergoing liquid–liquid phase separation in vitro. Nucleic Acids Res 48(D1):D320–D327. https://doi.org/10.1093/nar/gkz778
3. Wang Q, Zhang Y, Wu L et al (2020) Structural and functional basis of SARS-CoV-2 entry by using human ACE2. Cell 181(4):894–904.e9. https://doi.org/10.1016/j.cell.2020.03.045
4. Trivedi P, Palomba F, Niedzialkowska E et al (2019) The inner centromere is a biomolecular condensate scaffolded by the chromosomal passenger complex. Nat Cell Biol 21:1127–1137. https://doi.org/10.1038/s41556-019-0337-6
5. Waghorne CL, Corkran HM, Hunt-Painter AA et al (2016) NN-Bis(glycyl)amines as anti-cancer drugs. Bioorg Med Chem 24(17):3932–3939. https://doi.org/10.1016/j.bmc.2016.04.016
6. Daina A, Michielin O, Zoete V (2017) SwissADME: a free web tool to evaluate pharmacokinetics, drug-likeness and medicinal chemistry.
friendliness of small molecules. Sci Rep 7:42717. https://doi.org/10.1038/srep42717 Published 2017 Mar 3

7. Pires DEV, Blundell TL, Ascher DB (2015) pkCSM: predicting small-molecule pharmacokinetic and toxicity properties using graph-based signatures. J Med Chem 58(9):4066–4072. https://doi.org/10.1021/acs.jmedchem.5b00104

8. Bolognesi B, Lorenzo Gotor N, Dhar R et al (2016) A concentration-dependent liquid phase separation can cause toxicity upon increased protein expression. Cell Rep 16(1):222–231. https://doi.org/10.1016/j.celrep.2016.05.076

9. Wang J, Choi JM, Holehouse AS et al (2018) A molecular grammar governing the driving forces for phase separation of prion-like RNA binding proteins. Cell 174(3):688–699.e16. https://doi.org/10.1016/j.cell.2018.06.006

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