The Evaluation of *Stachys lavandulifolia* Leave Extracts on Cysts of *G. lamblia*, in Vitro

Mohammad Barati,¹ Mahdi Fakhar,² Shirzad Gholami,³ Bahman Rahimi Esboei,⁴,⁵ and Taher Elmi⁵

¹Infectious Diseases Research Center, AJA University of Medical Sciences, Tehran, Iran
²Department of Parasitology, Molecular and Cell Biology Research Center, School of Medicine, Mazandaran University of Medical Sciences, Sari, Iran
³Department of Parasitology and Mycology, Toxoplasmosis Research Center, Mazandaran University of Medical Sciences, Sari, Iran
⁴Department of Medical Parasitology and Mycology, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran
⁵Department of Medical Parasitology and Mycology, School of Medicine, Iran University of Medical Sciences, Tehran, Iran

Corresponding author: Bahman Rahimi Esboei, Department of Medical Parasitology and Mycology, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran. Tel: +98-912199542, E-mail: bahman864@yahoo.com

Received 2017 August 06; Revised 2017 September 18; Accepted 2017 September 24.

Abstract

**Background:** Diarrheal disease is one of the most common problems that affected deployed military personnel during operations and maneuvers. *Giardia* (*G.*) *lamblia* is one of the causes of parasitic diarrhea in humans. There are very few therapeutics agents with unpleasant adverse effects available for treating giardiasis. *Stachys lavandulifolia* with polyphenolic structure is considered as an antimicrobial agent. In this study we evaluated the in vitro inhibitory activity of *Stachys lavandulifolia* leaves on cysts of *G. lamblia*.

**Methods:** *G. lamblia* cysts were isolated by the sucrose method. Various concentrations of the watery and n-hexane extracts (2.5, 5, 10, 25, 50 mg/mL respectively) were used for giardiacidal activity in 6 different times (1, 5, and 30 minutes, as well as 1, 3, and 6 hours).

**Results:** The watery and n-hexane extract revealed great activity when compared to the control group (*P* < 0.05). The watery extract at the concentration of 100 mg/mL killed 93% of cysts after 6 hours. The n-hexane extract at the concentration of 100 mg/mL killed 100% of cysts after 6 hours. Both extracts showed dose dependent antigiardial activity and the n-hexane extract was better than the watery extract. When the doses of the extracts and contact times are increased, a gradual increase in antiprotozoal activity was observed.

**Conclusions:** *Stachys lavandulifolia* leaves, as a safe herbal medicine could be confirmed for anti giardial activity.

**Keywords:** Herbal Medicines, In Vitro, *Giardia Lamblia*, *Stachys lavandulifolia*  

1. Background

*Giardia lamblia*, a microaerophilic, binucleated flagellar protozoan, is a world-wide cause of intestinal infection that results in severe and explosive diarrhoea. *G. lamblia* is considered as the most commonly detected intestinal parasite in humans in both developing and developed countries (1). About 200 million people have symptomatic giardiasis and 2 million cases of symptomatic giardiasis were reported in Asia, Africa, and Latin America; about 500,000 new cases are reported annually (2, 3).

*G. lamblia* has been a major cause of traveler’s diarrhea. According to studies, diarrheal disease is considered one of the most common problems throughout the world, especially in deployed military personnel during operation and maneuvers. Diarrhea leads to the inability of military personnel and to reduce the combat capability of the military units consequently (4-6). Therefore, diagnosis and treatment of the disease agents is necessary.

Manifestation of giardiasis in humans may range from asymptomatic, mild-to-moderate symptoms, to severe occurrences of inadequate absorption, gastrointestinal disorder, or allergic disease (7). Several treatments for giardiasis have been developed. Tinidazole, metronidazole, furazolidone, and quinacrine are the list of current medical drugs, the use of traditional medicine is very common among the Iranian people (9). The genus Stachys is belonging to family Labiatae and includes many species that grow in the wild. *Stachys lavandulifolia* (*S. lavandulifolia*), one of the species of Stachys, was distributed in the north and west of Iran (Azerbaijan, Gilan, Golestan, Khorasan, Mazandaran, and Tehran) (10). *S. lavandulifolia*, with aromatic structure, includes the flavonoids, terpenoids, and phenyl ethanoids (11). *S. lavandulifolia* is locally known as Mountain Tea and as a traditional medicine that is...
used for headaches, neuralgia, nervous conditions, treating wounds, skin inflammation and as tonic at dyspepsia, astringent, and anti diarrheal in Iran (11).

The decoction of the leaves of *S. lavandulifolia* has been used for treatment of skin infection, menorrhagia, and especially antibacterial (12). Furthermore, the investigations on antibacterial, anti-inflammatory, anti anxiety, anti nephritic, and anti hepatitis properties of *S. lavandulifolia* have shown optimum activity (13-16). Antimicrobial and antifungal activity of *S. lavandulifolia* have been approved in various studies, however, there is very few evidence on anti parasitical activity of *S. lavandulifolia* leaves. Morteza-Semnani et al., (2007) demonstrated antibacterial activity of Stachys spp. against some bacterial species (13). Sereshti (2012) established that ethanolic extract of *S. lavandulifolia* leaves have shown appropriate effects on *Trichomonas vaginalis*, a protozoan parasite growth in vitro (17). *S. lavandulifolia* hydro alcoholic extract of aerial parts has been investigated for anti leishmanniasis activity and the extract indicate acceptable activity (18). This study was designed to evaluate the anti giardiasis activity of watery and n-hexane extract of *S. lavandulifolia* leaves.

2. Methods

2.1. Plant Material

*S. lavandulifolia* dried powder of the leaves was purchased from the Giah Essence Phyto-Pharmaceutical Co., Gorgan, Iran.

2.2. Preparation of Watery Extract of *S. lavandulifolia*

100 g of powder was extracted by the soxhellation process using 200 mL of distilled water for 24 hours at room temperature. The extract was concentrated after evaporating the water under vacuum at a temperature of 40°C. The remained water extract was freeze-dried and the obtained dried extract powder was 12.5 g.

2.3. Preparation of N-Hexane Extract of *S. lavandulifolia*

The leaves of *S. lavandulifolia* were dried. The leaves were pulverized in a grinding mill and a total amount of 100 g of powder was soxhelated with n-hexane for 24 hours at room temperature. The extract was dried after evaporating the solvent under vacuum at a temperature of 40°C. Then-hexane extract was freeze-dried. The obtained dried extract powder was 13 g.

2.4. Collection of *Giardia lamblia* Cysts

*G. lamblia* cysts were isolated from stool samples by a highly purified cyst suspension with a simplified sucrose gradient method. A total of 18 stool samples were fragmented in normal saline and filtered through a 300 µm filter. Three mL of stool suspension were layered on 3 mL of 0.85 M sucrose and centrifuged at 600 g for 10 minutes at 4°C. Aspirated the cysts with a Pasteur pipette at the sucrose-water interface and washed 3 times with normal saline. Washed cysts were carefully added to the top of a discontinuous density gradient, consisting of 2, 3 mL layers of 0.85 M and 0.4 M sucrose. After centrifugation, cysts concentrated at the 0.85 - 0.4 M sucrose interface were collected and washed again and then stored at 4°C for future uses.

2.5. In Vitro Experimental Assay

Four different concentrations (1, 10, 50 and 100 mg mL\(^{-1}\)) of the watery extract and n-hexane extract of *S. lavandulifolia* were used for 1, 5, and 30 minutes, as well as 1, 3, and 6 hours. A total of 2 mL of each solution was placed in test tubes, then 10,000 washed cysts were added. The contents of the tubes were gently mixed. The tubes were then incubated at 37°C for 1, 5, and 30 minutes, as well as 1, 3, and 6 hours. At the end of each incubation time, the upper phase was carefully removed. A total of 2 mL of 0.1% eosin stain was then added to the remaining settled cysts and mixed slightly. The cysts were then smeared on a glass slide, covered with a cover glass, and examined under a light microscope. The percentages of dead cysts were determined by counting 1000 cysts. Non treated cysts were considered as a control group in each experiment. Triplicate trials were performed for each experiment.

2.6. Viability Test

The viability of the cysts was detected by 0.1% Eosin as a vital dye under a light microscope. The cysts with no absorbed dye were considered potentially viable and otherwise, they were recorded as dead.

2.7. Statistical Analysis

Statistical analysis was directed by a one-way ANOVA (analysis of variance) considering a level of significance of 95% (P < 0.05), with SPSS-18 software.

3. Results

The anti giardial activity of watery and n-hexane extract of *S. lavandulifolia* is shown in Tables 1 and 2. Both the watery and n-hexane extracts of *S. lavandulifolia* showed
significant ability to reduction of *G. lamblia* cysts in comparison to the control group (*P* < 0.05). No significant reduction was observed in incubation at 1, 10, 50, and 100 mg/mL concentration after 1, 5, and 30 minutes.

The watery extract at the concentration of 100 mg/mL killed 73% and 93% after 3 and 6 hours. The n-hexane extract at the concentration of 100 mg/mL killed 89% and 100% after 3 and 6 hours. In general cysts of *G. lamblia* appeared to be more sensible to the n-hexane extract in comparison to the watery extract, however, there was no significant difference observed. The best effect was observed at 100 mg/mL after 6 hours of exposure time. The effects of these extracts were found to be concentration dependent. The high dose of watery extract shows a 93% reduction in cysts after 6 hours. N-hexane extract, at the highest concentration, completely inhibit the cysts growth after 6 hours. The highest activity was recovered for both extract against *G. lamblia* cysts with an IC$_{50}$ of 1 mg/mL after 3 hours. A typical depiction of viable and dead cysts is shown in Figure 1.

Figure 1. The Effect of Watery and N-Hexane Extract of *S. lavandulifolia* on *G. lamblia* Cyst in Vitro

Cysts that absorbed dye were recorded as dead (A) and otherwise, with no absorbed dye were considered potentially viable (B).

4. Discussion

Giardiasis is the most prevalent protozoal infection in most sites of the world. Clinical signs of this disease are ranged from asymptomatic to severe manifestation (19). The current treatments of the giardiasis are either one of the family of nitroimidazoles (usually metronidazole), nitrofurans, quinacrine, or paromomycin (20). Increased side effects and resistance of the parasite to these synthetic and semi-synthetic agents in the treatment of giardiasis make it necessary to find new, safe, and effective therapeutic agents (21). Until yet, metronidazole, with a wide variety of adverse drug reactions is included in selected therapeutic regimes (22). Therefore, some medicinal herbs with therapeutic effects could be considered as a drug of choice for treatment of *G. lamblia* infection (23-25).

Several experimental studies attributed traditional medicine for their anti-giardial activities. Calzada et al. (2006) investigated the susceptibility of *G. lamblia* trophozoites on 26 plants used in Mexican traditional medicine. They demonstrated methanolic extract of *Dorstenia contrajerva*, *Senna villosa*, and *Ruta chalepensis* were the most active toward *Giardia lamblia* with the 50% inhibitory concentration (IC$_{50}$) < 38 µg/mL after 48 hours and the trophozoites appeared to be resistance to methanolic extract of *Allium sativum*, *Aloysia triphylla*, *Annona cherimola*, *Artemisia absinthium*, *Artemisia ludoviciana*, *Bocconia frutescens*, *Caesalpinia pulcherrima*, *Caricapa paya*, *Cocos nucifera*, *Chenopodium ambrosioides* (green), *Chenopodium ambrosioides* (red), *Chenopodium murale*, *Chiranthodendron pentadactylon*, *Chrysactinia Mexicana*, *Dichondra argentea*, *Geranium mexicanum*, *Hippocratea excelsa*, *Lippia alba*, *Lygodium venustum*, *Matricaria recutita*, *Ocimum basilicum*, *Punica granatum*, *Schinus molle*, and *Thymus vulgaris* (26). Rahimi-esboe (2013) offers the methanolic extract of *Sambucus ebulusas* a good agent for killing *G. lamblia* cysts in vitro (27). The efficacy of the plants depends on its ingredients; therefore, evaluations of the components of a plant should be the first step of antimicrobial effects studies. In a recent study, Gertrude et al., (2017) assessed the antiparasitic effects of Ganaian medicinal plants against *Giardia lamblia*, *Entamoeba histolytica*, and *Naegleria fowleri* in an vitro examination. They indicated that *A. glaberrima*, *M. nobilis*, *M. angolensis*, *U. fasciata* extractions, ethyl acetate fraction of the extract of *E. ivorense* bark, and xylopic acid had IC$_{50}$ values of 15.91, 44.25, 20.00, 35.86, 13.76, and 11.45 µg/mL, respectively, against *G. lamblia* in comparison to the positive control (IC$_{50}$ = 10.47 µM) (28).

Several investigations have indicated that extract of folklore plants with phenolic consistent shows great anti giardial effects (29, 30). Some constituents have been isolated from *S. lavandulifolia*, mainly flavonoids with high antibacterial, antifungal, and anti parasitid characteristics (10, 13, 17). N-hexane extract of *S. lavandulifolia* had higher activity in vitro on *G. lamblia* cysts rather than watery extract. It is important to point out which one of the extracts displayed acceptable anti giardial activity with mortality rates ranging from 93 to 100% at 100 mg/mL after 6 hours.

The results of this study indicated that *S. lavandulifolia*’s giardiacidal activities attributed to the phenolic group, present in its extract. These phenolic and flavonoid com-
Table 1. Antigiardial Activity of Watery Extract of S. lavandulifolia Leaves

| Contact Time | Concentration, mg/ml |
|--------------|----------------------|
|              | 1        | 10       | 50       | 100      |
| 1, min       | 19       | 23       | 23       | 34       |
| 5, min       | 23       | 23       | 26       | 34       |
| 30, min      | 25       | 27       | 36       | 41       |
| 1, h         | 39       | 39       | 43       | 47       |
| 3, h         | 51       | 59       | 67       | 73       |
| 6, h         | 69       | 79       | 89       | 93       |

Table 2. Antigiardial Activity of N-Hexane Extract of S. lavandulifolia Leaves on G. lambia Cysts

| Contact Time | Concentration, mg/ml |
|--------------|----------------------|
|              | 1        | 10       | 50       | 100      |
| 1, min       | 25       | 29       | 34       | 41       |
| 5, min       | 28       | 33       | 42       | 52       |
| 30, min      | 37       | 41       | 51       | 63       |
| 1, h         | 49       | 51       | 63       | 75       |
| 3, h         | 61       | 67       | 79       | 89       |
| 6, h         | 81       | 83       | 93       | 100      |

pounds exist in S. lavandulifolia are rich in phenolic ring-associated hydroxyl groups. The hydroxyl groups donate hydrogen atoms to link to a negative charged microbial plasma membrane, promote the leakage of intracellular constituent, and disruption of cell membrane. Due to the S. lavandulifolia’s anti parasitical properties, it could be suggested as a helpful agent for the parasitic diseases. In Conclusion, in our study, S. lavandulifolia revealed an acceptable inhibitory activity against lambia cysts. N-hexan extract from S. lavandulifolia had higher anti protozoal activity on growth of G. lambia cysts. Finding further studies are required of Stachys lavandulifolia to evaluate its parasiticidal activity against other parasitic infection.

Acknowledgments
We thank all of those who helped us conduct this project.

Footnote

Conflict of Interests: We declare that we had no conflict of interest.

References
1. Schantz PM. Parasitic zoonoses in perspective. Int J Parasitol. 1991;21(2):161-70. doi: 10.1016/0020-7519(91)90006-S. [PubMed: 1869350].
2. World Health Organization. An overview of selected curable sexually transmitted diseases. Global Program on AIDS. 1995;2-27.
3. Thompson RC. Giardiasis as a re-emerging infectious disease and its zoonotic potential. Int J Parasitol. 2000;30(12-13):1259-67. doi: 10.1016/S0020-7519(00)00127-2. [PubMed: 1103253].
4. Barati M, Dabbagh Moghaddam A, Khoshdel A, Iravani S, Salahi-Moghaddam A, Tootoonchian M. Spatial distribution of intestinal amebiasis in Iran army units by geographic information systems [In Persian]. Journal of Police Medicine. 2013;2.
5. Sanders JW, Putnam SD, Riddle DR, Johanputra NK, Jones J, et al. The epidemiology of self-reported diarrhea in operations Iraqi freedom and enduring freedom. Diagn Microbiol Infect Dis. 2004;50(2):89-93. doi: 10.1016/j.diagmicrobio.2004.06.008. [PubMed: 15474396].
6. Sanders JW, Putnam SD, Gould P, Kolisnyk J, Merced N, Barthel V, et al. Diarrheal illness among deployed U.S. military personnel during Operation Bright Star 2001-Egypt. Diagn Microbiol Infect Dis. 2005;52(2):85-90. doi: 10.1016/j.diagmicrobio.2005.02.005. [PubMed: 15964494].
7. Jimenez-Cardoso E, Eligio-Garcia L, Cortes-Campos A, Flores-Luna A, Valencia-Mayoral P, Loza-Chavez I. Changes in beta-giardin sequence of Giardia intestinalis sensitive and resistant to albendazole strains. Parasitol Res. 2009;105(1):25-33. doi: 10.1007/s00436-009-1363-7. [PubMed: 19284572].
8. Harris JC, Plummer S, Lloyd D. Antigiardial drugs. Appl Microbiol Biotechnol. 2003;57(5-6):614-9. [PubMed: 12788688].
9. Amin GR. Popular medicinal plants of Iran [In Persian]. Iranian Research Institute of Medicinal Plants Tehran; 1991.
10. Morteza-Semnani K, Saeedi M, Shahani S. Antioxidant activity of the methanolic extracts of some species of Phlomis and Stachys on sunflower oil. *Afr J Biotechnol*. 2006;5(24).

11. Pirbalouti AG, Mohammadi M. Phytochemical composition of the essential oil of different populations of Stachys lavandulifolia Vahl. *Asian Pac J Trop Biomed*. 2013;3(2):123-8. doi: 10.1016/S2221-0338(11)60036-2. [PubMed: 23593599].

12. Pirbalouti AG. Medicinal plants used in Chaharmahal and Bakhtyari districts of Iran. *Herba Polonica*. 2009;3(2):69-77.

13. Morteza-Semnani K, Saeedi M, Mahdavi M, Rahimi F. Antimicrobial effects of methanolic extracts of some species of stachys and phlomis [In Persian]. *J Mazandaran Univ Med Sci*. 2012;21(3):124-8. doi: 10.1016/j.procb.2011.12.028.

14. Jenabi E, Asltoghiri M, Hajiloomohajeran M, Torkamani M. Effect of *Stachys tibetica* essential oil in anxiety. *Eur J Integr Med*. 2012;4(2):169-76. doi: 10.1016/j.eujim.2012.01.005.

15. Laggoune S, Zeghib A, Kabouche A, Kabouche Z, Maklad YA, Leon F, et al. Components and antioxidant, anti-inflammatory, anti-ulcer and antinococeptive activities of the endemic species *Stachys mialhesi* de Noé. *Arab J Chem*. 2016;9(1):99-7. doi: 10.1016/j.arabjc.2013.01.005.

16. Sereshti M, Yousofi Darani H, Zebardast N, Rafean M, Manochehre Noé. *Afr J Pharm Pharmacol*. 2012;6(15):447–75. doi: 10.1128/CMR.14.3.447-475.2001. [PubMed: 11432808].

17. Asadi M, Bahrami S, Ansari Samani R, Pakniat N. Effect of hydroalcoholic extracts of *Stachys lavandulifolia* Vahl and *Mespilus germanica* L. on *Leishmania major* [In Persian]. *Afr J Biotechnol*. 2009;8(24):208-12. doi: 10.1016/j.ejpp.2008.10.009. [PubMed: 19010325].

18. Zhang YH, Xue MQ, Bai YC, Yuan HH, Zhao HL, Lan MB. 3,5-Dicaffeoylquinic acid isolated from *Artemisia argyi* and its ester derivatives exert anti-leucyl-tRNA synthetase of *Giardia lamblia* (GlLeuRS) and potential anti-giardial effects. *Fitoterapia*. 2012;83(7):1281-5. doi: 10.1016/j.fitote.2012.05.016. [PubMed: 22668971].

19. Calzada F, Ypez-Mula L, Aguilar A. In vitro susceptibility of *Entamoeba histolytica* and *Giardia lamblia* to plants used in Mexican traditional medicine for the treatment of gastrointestinal disorders. *J Ethnopharmacol*. 2006;108(3):367-70. doi: 10.1016/j.jep.2006.05.025. [PubMed: 16846708].

20. Rahimi-Esboei B, Ebrahimzadeh MA, Ghohami S, Falah-Omrani V. Anti-giardial activity of *Sambucus ebulus*. *Eur Rev Med Pharmacol Sci*. 2013;17(5):2047–50. [PubMed: 23884825].

21. Gertrude KD, Christian A, Yaw DB, Trpta B, Brian MS, James HMK, et al. In vitro activity of selected gharian medicinal plants against parasites: *Giardia lamblia*, *entamoeba histolytica* and *naegleria fowleri*. *Afr J Pharm Pharmacol*. 2017;11(11):279-83. doi: 10.5897/ajpp2017.4795.

22. Hill DR, Nash TE. *Principles and practice of infectious diseases*. 1995. *Giardia lamblia*.

23. Machado M, Dinis AM, Salgueiro L, Custodio JB, Cavaleiro C, Sousa MC. Anti-Giardia activity of *Syzygium aromaticum* essential oil and eugenol: effects on growth, viability, adherence and ultrastructure. *Exp Parasitol*. 2011;127(4):732-9. doi: 10.1016/j.exppara.2011.01.011. [PubMed: 21725680].

24. Hernandez F, Hernandez D, Zamora Z, Diaz M, Anchea O, Rodriguez S, et al. Giardia duodenalis: effects of an ozonized sunflower oil product (Oleozon) on in vitro trophozoites. *Exp Parasitol*. 2009;121(3):208-12. doi: 10.1016/j.exppara.2008.10.009. [PubMed: 19010325].

25. Adams RD. Biology of *Giardia lamblia*. *Clin Microbiol Rev*. 2001;14(3):447-75. doi: 10.1128/CMR.14.3.447-475.2001. [PubMed: 11432808].