Nonclassical Islet Peptides: Pancreatic and Extrapancreatic Actions

Andrew English and Nigel Irwin
School of Pharmacy and Pharmaceutical Sciences, Ulster University, Coleraine, Northern Ireland, UK.

ABSTRACT: The pancreas has physiologically important endocrine and exocrine functions; secreting enzymes into the small intestine to aid digestion and releasing multiple peptide hormones via the islets of Langerhans to regulate glucose metabolism, respectively. Insulin and glucagon, in combination with ghrelin, pancreatic polypeptide and somatostatin, are the main classical islet peptides critical for the maintenance of blood glucose. However, pancreatic islets also synthesis numerous ‘nonclassical’ peptides that have recently been demonstrated to exert fundamental effects on overall islet function and metabolism. As such, insights into the physiological relevance of these nonclassical peptides have shown impact on glucose metabolism, insulin action, cell survival, weight loss, and energy expenditure. This review will focus on the role of individual nonclassical islet peptides to stimulate pancreatic islet secretions as well as regulate metabolism. In addition, the more recognised actions of these peptides on satiety and energy regulation will also be considered. Furthermore, recent advances in the field of peptide therapeutics and obesity-diabetes have focused on the benefits of simultaneously targeting several hormone receptor signalling cascades. The potential for nonclassical islet hormones within such combinational approaches will also be discussed.

KEYWORDS: β-cell, insulin secretion, nonclassical islet peptides, dual agonist, triagonist

Introduction
Metabolic abnormality can occur due to a range of genetic or acquired disorders including diabetes and obesity. These disorders involve a disturbance of metabolomics involved in energy balance regulation. Unfortunately, the prevalence of obesity and related type 2 diabetes mellitus (T2DM) is consistently increasing. The underlying pathophysiological feature of T2DM, and other related forms of diabetes, is primarily associated with a loss in function and/or mass of insulin secreting beta-cells (β-cells) of the endocrine pancreas. The pancreas is situated behind the peritoneum and consists of a head, body, and tail that is close to the duodenum. Most pancreatic mass (90%) is made of exocrine cells grouped in lobules and separated by connective tissue. The product of exocrine cells, an alkaline liquid concentrated with digestive enzymes, is drained via a pancreatic duct and enters the small intestine to aid digestion. However, throughout the exocrine tissue clusters of endocrine cells called ‘islets of Langerhans’ exist, these islets are known to play a vital role in controlling metabolism, and especially blood glucose equilibrium (Figure 1). The main hormones secreted from pancreatic islet cells are glucagon from alpha (α) cells, insulin from β-cells, somatostatin from delta (Δ) cells and pancreatic polypeptide (PP), with release believed to be under strict endocrine, paracrine, and neuronal control. However, several nonclassical islet peptides have also been described and shown to possess similarly important effects on islet function (Table 1). This is epitomised by recent acceptance of ghrelin as an islet-derived peptide secreted from epsilon (ε) cells with established intra-islet actions, as well as effects on metabolism beyond the pancreas. As such, although much research on pancreatic islets and T2DM treatments has focused on the classic peptide hormones, the role and therapeutic applicability of nonclassic islets peptide hormones also needs to be considered.

Classical Islet Peptides

As noted above, the endocrine pancreas secretes 5 peptide hormones, namely, glucagon, insulin, somatostatin, PP, and ghrelin, from specific cell types, that play a critical role in glucose homeostasis and energy metabolism (Table 1). A secretory interplay between glucagon and insulin balances blood glucose in healthy individuals. Glucagon is released in times of low glucose to promote hepatic glycogenolysis, whereas insulin is released in response to raised glucose to promote glycogenesis and lipogenesis. The underlying molecular processes of insulin exocytosis in response to glucose is well known and involves the closure of KATP channels, β-cell membrane depolarisation, calcium influx, and subsequent secretion of insulin. During the onset of diabetes this relationship is negatively affected and insulin secretion rates decrease, which is often combined with desensitisation of the insulin receptor (IR) located on peripheral tissues. As such, treatment of diabetes has focused predominantly on addressing the deficit in insulin secretion through administration of direct insulin secretagogues or insulin replacement therapy, as well as therapies aimed at desensitising the IR. In this regard, there are a variety of nonclassical islet peptides that can trigger or potentiate insulin secretion, improve β-cell survival or increase insulin sensitivity through modulation of G-protein coupled receptors (GPCRs) (Figure 1).
Nonclassical Islet Peptides – Pancreatic Effects

Incretin and related peptides

As well as the classical islet peptides, there are a number of other peptide hormones known to be synthesised and released within pancreatic islets, often termed ‘nonclassical’ islet peptides (Figure 1). The incretin hormones, namely glucagon-like peptide (GLP)-1 and glucose dependent insulinotropic peptide (GIP), are the most comprehensively studied nonclassical islet peptides. Typically, GLP-1 and GIP are considered to be endocrine L- and K-cell derived peptides released in response to nutrient ingestion. Elegant studies have described the secretory dynamics of these 2 hormones from the gut. However, both GIP and GLP-1 have also been shown to be synthesised and released locally within pancreatic islets, with important paracrine effects. Glucagon-like peptide-1 has direct and indirect actions on α-cells, directly inhibiting glucagon and indirectly increasing intra-islet regulation via somatostatin. GIP effects on glucagon are glucose dependent. As such, at low levels of glycaemia, glucagon release is increased by GIP with minimal effect on insulin, with this glucagonotropic effect disappearing at higher glucose levels. Mechanistically, GLP-1 and GIP potentiate insulin secretion by binding to specific GPCRs located on pancreatic β-cells to increase cAMP and activate Epac and protein kinase A (PKA). The process of GIP and GLP-1 induced insulin secretion is well described, primarily due to increased interest in the ‘incretin effect’ over the past decades, clearly defining how the intestinally released hormones potentiate insulin release following meals. These mechanisms supplement glucose-mediated effects, closing K<sup>+</sup>ATP and Kv channels and promoting depolarisation of the β-cell. Calcium channels are also sensitised via PKA and Epac. Together this facilitates intracellular calcium stores to be released, resulting in insulin secretion. Glucagon-like peptide-2 (GLP-2) is another proglucagon derived peptide, its principle role is to maintain intestinal mucosal villus epithelium and aid gut absorption. In addition, GLP-2 acts as an intestinal growth factor, with clinical application for short bowel syndrome. Notably, GLP-2 also has been demonstrated to possess proliferative and anti-apoptotic effects in β-cells, highlighting a role in glucose homeostasis. However, GLP-2 has no effect on insulin secretion from β-cells, or isolated mouse islets.

Peptide Tyrosine Tyrosine (PYY) is a 36 amino acid peptide hormone best known as being released postprandially from enteroendocrine L-cells, similar to GLP-1. Moreover, both hormones are expressed within pancreatic islets and released locally (Figure 1). Peptide Tyrosine Tyrosine and neuropeptide Y (NPY) both

*Incretin secreted by isolated islets, cell type unknown

Figure 1. Endocrine pancreatic cell types and their peptide secretions. Exocrine pancreatic acinar cells constitute most of the pancreatic tissue, these cells produce digestive enzymes which are transported via the pancreatic ducts. The endocrine pancreas is illustrated with all cell types; alpha, beta, delta, PP, and epsilon. The cells are arranged in compact islets and secrete a number of classical and ‘nonclassical’ peptides, as depicted. PP indicates pancreatic polypeptide.
activate neuropeptide Y receptors (NPYRs) and represent promising antidiabetes targets. Although the peptides activate the same family of receptors, their effects on islet secretions differ due to differences in receptor affinity and specificity. Early rodent studies indicated plasma insulin levels were reduced or unaffected by PYY. Further studies in isolated islets and human β-cells also revealed minimal effect of PYY on insulin secretion, although β-cell anti-apoptotic and proliferative effects were noted. As such, transgenic mouse models overexpressing PYY exhibited enhanced glucose facilitated insulin responses and amplified islet number. Consistent with these findings, knock out of PYY in the gut and pancreas causes disruption of islet morphology and decreased β-cell mass. In addition, streptozotocin-induced insulin deficiency and β-cell destruction is linked to reduced islet PYY levels, whereas hydrocortisone-induced increases of β-cell mass are linked to elevated PYY levels. Furthermore, in the developing foetal pancreas PYY is detectable at a very early stage, indicating potential effects on pancreatic islet development. For PYY effects, it is also important to note the distinct receptor specificity of the 2 main circulating molecules and pancreatic expression of these receptors. As such, PYY (1–36) binds and activates all NPYRs, whereas PYY (3–36) is an NPYR2-specific molecule. PYY can prevent glucagon release from isolated islets, although NPY receptors are thought to be specifically

Table 1. Classical and ‘nonclassical’ peptides secreted by the pancreas. Table includes the peptides main origin tissue (including gut and CNS) and cells of origin. The key secretory initiators are stated and the effect each peptide has on the pancreatic islet.

| Peptide | Origin | Regulation of Secretion | Effects on Islet |
|---------|--------|--------------------------|------------------|
| Insulin | Pancreas | β-cell | Plasma glucose, FFA, glucagon | Inhibits glucagon |
| Glucagon | Pancreas | α-cell | Amino acids released by digestion | Stimulates insulin secretion |
| Somatostatin | Pancreas | Δ-cell | Glucagon | Inhibits release of insulin and glucagon |
| Pancreatic polypeptide | Pancreas | PP-cell | To be determined | Inhibits insulin |
| Ghrelin | Pancreas | Epsilon cell | Dietary intake of nutrients | Inhibits insulin |

Nonclassical

| Peptide | Origin | Regulation of Secretion | Effects on Islet |
|---------|--------|--------------------------|------------------|
| GLP-1 | Pancreas/Gut | α-cell/Intestinal L-cell | Ingested glucose load | Increases insulin secretion, β-cell mass, inhibits glucagon, apoptosis |
| GLP-2 | Pancreas/Gut | α-cell/L-cell | Dietary intake of nutrients | Positive effects on β-cell proliferation and apoptosis |
| GIP | Pancreas/Gut | α-cell/Intestinal K-cell | Ingested glucose load | Causes insulin secretion, and β-cell proliferation |
| PYY | Pancreas/Gut | α-cell/Intestinal L-cell | Dietary intake of nutrients | β-cell proliferative and anti-apoptotic effects |
| Gastrin | Pancreas/Gut | β-cell/G-cell | Stomach distension, digested proteins | Causes insulin secretion |
| CCK | Pancreas/Gut | β-cell/EEC cell | Stomach distension, digested proteins | β-cell proliferative and anti-apoptotic effects |
| Xenin-25 | Pancreas/Gut | Islets/K-cell | Dietary intake of nutrients | Increases GIP-mediated insulin secretion |
| Secretin | Pancreas/Gut | β-cell/S-cells | Dietary intake of nutrients/Stomach acid | Inhibits glucagon |
| Urocortin3 | Pancreas | α-cell/β-cell | Plasma glucose (co-secreted with insulin) | Promotes insulin secretion |
| Amylin | Pancreas | β-cell | Dietary intake of nutrients | Inhibits insulin secretion and β-cell proliferation |
| Vaspressin | Pancreas/CNS | β-cell/M-cell | Response to hypertonicity | Promotes Insulin secretion, β-cell proliferative and anti-apoptotic effects |
| Oxytocin | Pancreas/CNS | β-cell/M-cell | Related to reproductive function | Promotes insulin secretion, β-cell proliferative and anti-apoptotic effects |

Abbreviations: CCK, cholecystokinin; CNS, central nervous system; EEC, enteroendocrine cells; FFA, free fatty acid; GIP, glucose dependent insulinotropic peptide; GLP, glucagon-like peptide; PYY, Peptide Tyrosine Tyrosine.
on β-cells. This would suggest PYY modulation of glucagon occurs via indirect pathways and could involve insulin or somatostatin release. The ability of PYY to restore deranged insulin and glucagon secretions in T2DM, is of clinical significance and these findings justify further investigation. Similar to PYY, the neurotransmitter NPY has also been linked to decreased insulin and glucagon release.

**Gastrin and cholecystokinin**

Receptors for gastrin and cholecystokinin (CCK), namely CCK1 receptors, are highly expressed on α- and β-cells and both are synthesised and secreted by islets and effect islet function. Hypergastrinaemia is linked to excessive insulin production and low blood glucose, and both hormones induce secretion with or without glucose. There is conflicting evidence on the effect of gastrin on glucagon secretion, with some models highlighting a stimulatory effect whereas others conflict these findings. Disruption of the gastrin gene in mice does not affect islet morphology or basal glucagon levels, but does cause substandard glucagon secretion in response to insulin caused hypoglycaemia. Gastrin in combination with epidermal growth factor has been found to cause β-cell regeneration in mice after near total destruction by alloxan. In respect to CCK, little effect on glucagon release has been recorded, but a close relationship to GLP-1 has been identified. Research has focused on an intra-islet loop based on local islet secretion of these peptides, whether GLP-1 and CCK originating from the gut have the same islet effects is unknown. Sulphation of CCK at a tyrosine located 7 residues from the C-terminus is crucial for CCK bioactivity, this post-translational modification allows for strengthened protein-protein interaction. Glucagon-like peptide-1 secreted from the α-cell targets the β-cell to stimulate CCK production, while CCK can also stimulate α-cells to release GLP-1 increasing β-cell survival and proliferation, in a positive feedback loop within β-cells. Mechanistically, CCK1 receptor stimulation phosphorylates PKA and phospholipase C (PLC) increasing intracellular calcium concentrations via the L-channel, this causes positive bias on PLC activity. Increased calcium and diacylglycerol produced by PLC activates protein kinase C which ultimately opens gated calcium channels. Further work is required to fully assess the therapeutic promise of both gastrin and CCK, especially in relation to potential off-target side effects.

**Urocortin3 and islet amyloid polypeptide**

Urocortin3 (Ucn3) is a peptide hormone expressed by mature β-cells, early work revealed insulin and glucagon release to increase in response to Ucn3 treatment. Interestingly, Ucn3 may also have benefits on plasma glucose and glucose tolerance. This has driven new studies to define the physiological role of Ucn3, and it is known that Ucn3 amplifies somatostatin release to modulate insulin and glucagon secretion. Somatostatin receptors (SSTRs) have been acknowledged on α- and β-cells. Rodent studies reveal inhibition of glucagon and insulin is facilitated through SSTRs 5 and 2, respectively; knockout of these receptors results in variation of basal and glucose stimulated insulin secretion. In T2DM, Ucn3 expression is reduced in β-cells and causes an unstable glycaemic state. Islet amyloid polypeptide (IAPP, amylin) is a peptide co-secreted from β-cells with insulin in response to glucose and fatty acids at a ratio of 15:1 (insulin:amylin). Currently, 3 amylin receptors have been identified, namely AMY1, AMY2, and AMY3 and activation by amylin is thought to regulate insulin secretion under certain disease states or conditions. Knockout studies in β-cells have revealed reduced amylin causes increased glucose-induced insulin secretion, whereas treatment of amylin at physiological levels inhibits insulin secretion. Amylin also controls β-cell proliferation in a glucose dependent manner. As such, at high glucose concentrations amylin inhibits proliferation and at low levels promotes β-cell proliferation.

**Xenin-25 and neurotensin**

Xenin-25 is a peptide hormone traditionally characterised as being secreted from entodocrine K-cells with GIP, but xenin-25, as well as the related neurotensin hormone, have also been found in pancreatic islets, alongside neurotensin receptors (NTSRs). The main pancreatic effects of xenin include increasing PP secretion and amplifying the effects of GIP, indirectly effecting insulin and glucagon release. Xenin-25 interacts with acetylcholine signalling pathways to increase β-cell insulin secretion, and this is thought to be independent of NTSRs. This may explain slight differences in the pancreatic effect of xenin-25 and neurotensin, despite both being NTSR ligands. Proliferation in rodent and human β-cells is enhanced by xenin-25 and neurotensin, suggesting the peptides as promising antidiabetic agents. The signalling mechanisms involved in insulin and glucagon secretion for xenin-25, differ between human and mouse models in response to co-treatment with GIP. In mice, positive effects of co-treatment were seen under hyperglycaemic conditions. In humans, effects on insulin and glucagon where specifically recorded in patients with reduced glucose tolerance, but not in patients with T2DM. Cholinergic signalling may act as a regulatory mechanism to increase insulin secretion, and failure is thought to lead to the development of T2DM. In mice, atropine treatment inhibited the effect of xenin on GIP-mediated insulin and glucagon secretion implicating cholinergic signalling whereas in humans there is no inhibition.

**Vasopressin, oxytocin, and secretin**

Vasopressin (AVP) and oxytocin (OT) are neuropeptides and components of a cohesive system, with synthesis also evidenced within the pancreas. Oxytocin receptor knockout (OTKO) mice have increased sensitivity to AVP and AVP causes a significant concentration dependent increase of insulin
secretion in rodent (BRIN BD11) and human (1.1B4) \( \beta \)-cells, but has no effect on glucagon secretion. AVP receptors; V1a, V1b, V2 are highly abundant on \( \beta \)-cells and antagonism of V1a, V1b has a negative effect on AVP-mediated insulin secretion. AVP also has positive effects on \( \beta \)-cell proliferation and survival. Oxytocin (OT) is known to stimulate insulin and glucagon secretion and have positive effects on insulin action, glucose intolerance and islet hypertrophy. Calcium-calmodulin kinase (Ca-CAMKK), phosphoinositide-3-kinase (PI3K) and AMP-activated protein kinase (AMPK) pathways may mediate these effects and serotonin (5-HT) and dopamine may have supplementary properties. Secretin is produced in the duodenum by \( S \)-cells and in the pancreas by secretin producing islets, it consists of 27 amino acids with a sequence similar to glucagon and GIP. Following an oral glucose load, secretin maintains blood glucose control through initiating insulin release, while other work has detailed suppressive effects on \( \alpha \)-cell function and glucagon release. Interestingly, secretin has also been found to enhance the effects of CCK to increase \( \beta \)-cell proliferation and mass.

**Nonclassical Islet Peptides – Appetite Regulation**

The peptide hormones discussed above are considered to be nonclassical islet peptides generated within the pancreas, that mediate numerous local biological actions (Table 1). However, as well as direct pancreatic effects, many of these hormones have well characterised effects outside of the pancreas, that could be linked to pancreatic or extra pancreatic sites of synthesis. Glucagon-like peptide-1 has a major effect on energy balance and appetite via a complex relationship that is linked to centrally mediated actions. Physiological effects of GLP-1 include decreased food intake, inhibited gastric emptying and reduced gastric motility. These effects are facilitated by binding to the GLP-1R and signalling the central nervous system (CNS) via the vagal nervous system. Glucagon-like peptide-1 is secreted by GCG neurons, which allow direct central activation of the GLP-1R in the NTS of the hypothalamus. GCG neurons signal afferent nerves to relay satiety information to the hypothalamus. Glucose dependent insulinoergic peptides’ physiological effects on appetite are somewhat disputed, in vitro work using mouse \( 3T3L1 \) adipocytes found GIP increases adipogenesis, whereas chronically reduced GIP in mice attenuates weight gain. However, transgenic overexpression of GIP, as well as GIP agonist treatment in mice reduces obesity related traits. Moreover, recent clinical studies employing co-activation of GLP-1 and GIP receptors have revealed that GIP can augment the body weight reducing effects of GLP-1.

As noted above, PYY circulates in distinct bioactive forms, with PYY (3-36) recognised as the foremost circulating PYY peptide postprandially. As well as pancreatic secretion, PYY is released from L-cells of the gut, 15 minutes after food intake, secretion is proportional to calories consumed. This would indicate that PYY release occurs prior to nutrients reaching the distal small intestine, suggesting secretion is mediated centrally. In addition, there is evidence of enteroendocrine PYY releasing cells in the upper ileum, with studies revealing such cells can express multiple gut hormones, which may also be important in this regard.

Rodent receptor KO studies reveal PYY (3-36) acts on the arcuate nucleus (ARC) in the CNS via the NPY2 receptor. Activation reduces appetite by increasing POMC/\( \alpha \)-MSH activity while suppressing NPY neurons. Glucagon-like peptide-1 administration is known to cause nausea and gastric complications, and clinical studies have revealed such side-effects are heightened after PYY(3-36) administration, indicating related effects on energy regulation. Although CCK increases gallbladder contraction, inhibits gastric emptying and reduces food intake, many of its effects on energy balance are due to activation of peripheral vagal afferent fibres. However, CCK is also believed to penetrate the blood brain barrier (BBB) to act directly in the CNS to effect appetite and digestion.

Xenin-25 decreases food intake when administered centrally, or peripherally at elevated doses, indicating action as a satiety factor. In animal studies, xenin-25 causes anorexia via activation of central NTRSR1. This has been supported by work in NTRSR1 KO mice, which were nonresponsive to xenin. The mechanisms involved in xenin appetite regulatory effects are poorly described, the extracellular signal-regulated kinases signalling cascade has been implicated, but is not essential for feeding suppression. In addition, a long acting xenin-25 analogue, xenin-25[Lys13PAL], lacked effects on energy intake and body weight, but did induce significant benefits on glucose homeostasis in mice.

Oxytocin and vasopressin secretion are controlled by signals that regulate appetite, and both hormones are released acutely following a meal. Oxytocin is well established to influence energy balance in vivo, suppressing carbohydrate intake and promoting movement and thermogenesis. In rodent brain tissue, OT cells express IRs and glucokinase, and can be activated by insulin and glucose. Further research identified leptin receptors, as well as receptors allied with other anorectic peptides such as \( \alpha \)-MSH, on OT producing cells. Ligands of these receptors signal the NTS including noradrenergic A2 cells, and these cells directly interact with OT cells. Other peptides, such as CCK, may support the energy balance effects of OT at central sites, but further studies are needed to elucidate exact mechanisms.

**Therapeutic Potential of Multiagonist Nonclassical Islet Peptides**

**Dual agonists with nonclassical islet hormone components**

Recent novel peptide-based approaches for the treatment of T2D and obesity have focused on simultaneous activation of multiple receptors, many of which are related to nonclassical islet peptide hormones (Table 2). Synthetic peptides acting...
as dual or triagonists at GPCRs have been predicted to have positive effects on energy balance expenditure, glucose homeostasis and appetite. Early dual compounds focused on GLP-1/glucagon co-agonism and activity was based on the naturally occurring peptide oxyntomodulin. Treatment improved maximum insulin secretion compared with exendin-4 and resulted in large decreases in food intake, adiposity and increased lipolysis. Weight loss effects have been linked to significant leptin sensitisation and was successfully translated into phase II clinical trials. It has been concluded that these peptides have a greater effect than either single hormone. MEDI0382 is a GLP-1/glucagon agonist displaying significant promise in numerous clinical studies, reducing fasting and postprandial glucose and body weight. MEDI0382 has consistently exhibited a mild adverse event profile. Another dual GLP-1/glucagon agonist (SAR425899) also has significant clinical promise, reducing HbA1c and body weight over 28 days, with mild adverse events (Table 2).

In recent years, dual incretin agonists have been developed, these have focused on manipulating GIPR and GLP-1R signalling to reduce food intake and adiposity, and promote combined benefits on pancreatic β-cells. A novel GIP/GLP-1 agonist (N-ac(DAla2)GIP/GLP-1-exe) activates both incretin receptors to cause concentration dependent insulin secretion from BRIN BD11 β-cells and isolated islets. Dual receptor activation results in increased weight loss and improved insulin sensitivity compared with mono treatment in mice. These results have been confirmed in further preclinical work and in clinical studies. In a recent Phase 2A clinical trial a GLP-1/GIP dual agonist (NNC0090-2746) with balanced affinity for GLP-1R and GIPR significantly reduced HbA1c and weight over 8 weeks in T2D patients compared with placebo. LY3298176 is a fatty acid adapted peptide with GLP-1 and GIP receptor activity, designed to be administered once weekly. Preclinically, LY3298176 causes glucose-dependent insulin secretion from islets isolated from wild-type, GIPR−/− and GLP-1R−/− mice, indicative of activity at both receptors. LY3298176 has successfully translated preclinical findings into human studies, imparting meaningful enhancement of glycaemic control and body weight specifically, reducing HbA1c and body weight greater than a benchmark group taking Dulaglutide. This highlights the importance of balancing receptor interaction when developing dual or triagonists. It is worth noting that there was a 30% nonresponse rate, but results would indicate that dual incretin agonists are more potent than established mono treatments (Table 2).

Table 2. Dual and triagonist peptides with classical/’nonclassical’ islet peptide components. Table includes peptide hormone components, compound name, main effects and relevant citation(s).

| PEPTIDES HORMONE COMPONENTS | COMPOUND NAME | MAIN EFFECTS | REFERENCE |
|-----------------------------|---------------|--------------|-----------|
| **DUAL AGONISTS**           |               |              |           |
| GLP-1/glucagon              | MEDI0382      | Reduces fasting and postprandial glucose/body weight | 123,124 |
| GLP-1/glucagon              | SAR425899     | Reduces HbA1c/body weight | 125 |
| GIP/GLP-1                   | N-ac(DAla2)GIP/GLP-1-exe | Increases insulin secretion/sensitivity; reduces body weight | 128 |
| GLP-1/GIP                   | NNC0090-2746 | Reduces HbA1c/body weight | 117 |
| GLP-1/GIP                   | LY3298176     | Increases insulin secretion; Reduces HbA1c/body weight | 93,129 |
| GLP-1/xenin                 | Exendin-4/Xenin-8-Gln | Increases insulin secretion/GIP action, reduces appetite | 131 |
| GLP-1/gastrin               | ZP3022        | Lowers HbA1c, improves glucose tolerance and β-cell neogenesis | 132 |
| GLP-1/PYY                   | EP45          | Lowers blood glucose, stimulates insulin secretion | 133,134 |
| **Triagonists**             |               |              |           |
| GIP/glucagon/GLP-1          | GIP-Oxm       | Improves glucose homeostasis, insulin secretion and body weight | 135 |
| YAG-glucagon                | Y1-dA2-I22-N17-V18-L27, G28,29-glucagon | Reduces plasma glucose, food intake and body weight; increases plasma insulin and insulin sensitivity | 136 |
| GLP-1/GIP/glucagon          | HM15211       | Improves weight loss, anti-inflammatory and anti-oxidative stress, neuroprotective | 142,143 |
| GLP-1/gastrin/xenin         | Exendin-4(Lys27 PAL)/gastrin/xenin-8-Gln | Decreases nonfasting HbA1c, food intake. Enhances insulin levels, glucose tolerance | 144 |
| Glugagon/GLP-1/NPY          | GGP817        | Unvalidated (FRET analysis) | 145 |

Abbreviations: FRET, fluorescence resonance energy transfer; GIP, glucose dependent insulinotropic peptide; GLP, glucagon-like peptide; PYY, Peptide Tyrosine Tyrosine.
A novel GLP-1/xenin hybrid also indicated significant therapeutic potential in mice, increasing insulin secretion, reducing appetite and improving GIP action.130 A GLP-1/gastrin dual agonist, ZP3022, also displayed multiple antidiabetic effects in vivo. These included lowering HbA1c, improving glucose tolerance and increasing β-cell neogenesis.131 Dual treatment of GLP-1 and PYY led to a synergistic reduction in food intake.132 A recent study characterised a chimeric peptide (EP45) that consists of sequences from exendin-4 (Ex-4) and PYY(3-36), multiple organs can be targeted with this peptide.133 (Table 2). Fluorescence resonance energy transfer (FRET) assays demonstrate EP45 stimulates cAMP production via the GLP-1R, while inhibiting cAMP production via the NPY2R.133 These findings would indicate EP45 potentiates blood glucose lowering effects by activating GLP-1R on β-cells, thus triggering glucose stimulated insulin secretion and upregulating insulin gene expression.133 EP45 could also activate receptors centrally to affect appetite and would have relevance for the treatment of T2D.

Triagonists With Nonclassical Islet Hormone Components

The positive results from dual agonist studies have driven the development of triagonists, such as GIP-oxyntomodulin (GIP-Oxm)134 and an adapted glucagon peptide (YAG-glucagon).135 Most recent work has focused on the incretin/glucagon system and triagonists have been developed to activate the GLP-1/ GIP/glucagon receptors.136 Glucagon-like peptide-1/glucose dependent insulinotropic peptide and glucagon independently stimulate complementary mechanisms in vivo to cause glucose-mediated insulin secretion,137 and reductions in body weight and cholesterol are greater than a dose matched liraglutide treatment.138,139 In a diabetic rodent model GLP1R-GGCR-GIPR tri-agonism had beneficial pancreatic effects, animals displayed improved glucose tolerance and reduced ad libitum blood glucose, with reduced insulin levels suggesting improved insulin sensitivity140 (Table 2). Peptide characterisation of triple-acting entities in nonhuman primates has been completed by Sanofi, Novo Nordisk and Hanmi, with Novo Nordisk and Hanmi conducting phase 1 clinical trials.129 A long acting GLP-1/GIP/glucagon agonist (HM15211) has a unique pharmacological profile, demonstrating benefits on weight loss and lipid profiles, while exhibiting no risk of hyperglycaemia. Interestingly, in mice HM15211 has also shown promise in treating nonalcoholic steatohepatitis (NASH), fibrosis141 and alzheimers disease142 (Table 2).

Other triagonists include an enzyme resistant GLP-1/gastrin/xenin peptide which has antidiabetic potential in respect to insulin secretion and enhancement of GIP action,143 as well as a triagonist (GGPS817) that activates the GluR, GLP-1R and NPY2R.144 This peptide has not been validated in vivo, but it is thought agonism of GluR would cause increased energy expenditure.145 GLP-1R activation would result in enhanced β-cell function and improved glucose homeostasis146 and NPY2R agonism would suppress appetite.95

Conclusion

Signalling processes within pancreatic islets are highly complicated,147,148 and our recent knowledge of an important role for numerous nonclassical islet peptides is these processes requires further detailed study. Understanding their physiology, and potential pathophysiology in diabetes, could open up new promising avenues for drug effective candidates.19 Both preclinical and clinical evidence demonstrate that metabolic profile can be improved by synergistic action of multiple peptide hormone receptor targets, many of which include nonclassical islet peptides.149 Such strategies hold great promise for obesity and related forms of diabetes, especially given the optimism generated through early clinical observations.

Author Contributions

AE: Wrote the first draft of the manuscript.
AE,NE: Jointly developed the structure of the paper.
AE, NE: Made critical revisions and approved final version.
All authors reviewed and approved the final manuscript.

ORCID iD

Andrew English https://orcid.org/0000-0003-1232-5585

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