An association study of SERPING1 gene and age-related macular degeneration in a Han Chinese population

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Purpose: Single nucleotide polymorphisms (SNPs) in the complement component 1 inhibitor (SERPING1) gene have been shown to be significantly associated with age-related macular degeneration (AMD) in Caucasian populations. A replication study of an association between these SNPs and AMD in a Chinese population is reported in this study.

Methods: Six SNPs, including rs2511990, rs1005510, rs11546660, rs2511989, rs2511988, and rs4926 in SERPING1 were genotyped in a Han Chinese subject group using the SNaPshot method of ABI. This subject group was composed of 194 patients with choroidal neovascularization (CNV or wet) AMD, 78 patients with soft drusen, and 285 matched controls. P values of the SNPs were calculated using an additive model. Haplotype frequencies between cases and controls were compared by χ2 analysis. The haplotype analysis was performed using Haploview 4.0.

Results: None of the six SNPs showed significant association with AMD. None of the major haplotypes were observed to be significantly associated with AMD or choroidal neovascularization AMD (CNV) after a stringent Bonferroni correction.

Conclusions: We demonstrate that SNPs in SERPING1 are not significantly associated with AMD in the mainland Han Chinese population.

Age-related macular degeneration (AMD) is a leading cause of blindness in the elderly population, characterized as chronic and progressive degeneration of photoreceptors, the underlying retinal pigment epithelium (RPE), Bruch’s membrane, and possibly, the choriocapillaris in the macula [1,2]. AMD is divided clinically into dry and wet AMD. Patients with dry AMD present with cellular debris (drusen) in or under the retinal pigment epithelium (RPE), irregularities in the pigmentation of the RPE, or geographic atrophy (GA). Patients with exudative or wet AMD are characterized by serous detachment of the RPE or choroidal neovascularization (CNV), or both [1,2]. Advanced AMD, including geographic atrophy or exudative disease, can cause severe vision loss.

It is believed that AMD is a complex disorder caused by the interaction of multiple genetic and environmental risk factors [3–7]. Identification of AMD related genes has been tremendously successful. Complement pathway genes, including complement factor H (CFH) [2,8–13], C2/CFB [14–16], and C3 [15–17], have been confirmed by many replication studies. The LOC387715/HTRA1 gene has also been verified as a major AMD locus in different populations [18–24]. Recently, SNPs in the serpin peptidase inhibitor, clade G (C1 inhibitor) member 1 (SERPING1) gene showed highly significant genotypic association with age-related macular degeneration in two Caucasian populations [25]. Unfortunately, this finding could not be replicated by other studies [26–34].

To further analyze the association of SERPING1 and AMD, we investigated the association between SNPs in this gene and AMD in a mainland Han Chinese population.

METHODS

Subjects: The Institutional Review Boards of the Sichuan Provincial People’s Hospital, Xinhua Hospital of Shanghai Jiao Tong University, and Zhongshan Ophthalmic Center, China approved this study. All subjects provided informed consent before participation in the study. AMD patients and normal age-matched controls, including individuals with a normal eye examination (individuals age 60 years or older with no drusen or RPE changes), were recruited in the ophthalmology clinic at Sichuan Provincial People’s Hospital, Xinhua Hospital of Shanghai Jiao Tong University, and Zhongshan Ophthalmic Center, China. All participants went through a standard examination protocol as in the...
previous description [19,24,27]. Grading was performed using a standard grid classification suggested by the International ARM Epidemiological Study Group for age-related maculopathy (ARM) and the age-related macular degeneration group [27]. All abnormalities in the macula were characterized according to the type, size and number of drusen, and hyperpigmentation or hypopigmentation, as well as AMD stages as defined by AREDS 1–5 stages. Patients with clinical features of AMD and CNV (CNV from other causes was excluded), with or without drusen, were diagnosed as wet AMD patients. Patients with only soft drusen were diagnosed as drusen (dry AMD) patients. In total, 194 wet AMD patients (Eight from Zhongshan Ophthalmic Center, 28 from Xinhua Hospital, 158 from of the Sichuan Provincial People’s Hospital), 78 soft drusen patients (all from of the Sichuan Provincial People’s Hospital), and 285 normal matched controls (Eight from Zhongshan Ophthalmic Center, 30 from Xinhua Hospital and 247 from of the Sichuan Provincial People’s Hospital) were recruited. In the normal matched controls, all individuals underwent an eye exam, no signs of early AMD, such as soft drusen or irregular pigmentation of the RPE in the macular area, were observed. Clinical information about the cases and controls is listed in Table 1.

**Genotyping:** Blood from each subject was drawn and collected in an EDTA-containing tube. Genomic DNA was extracted from the blood by a Gentra Puregene Blood DNA kit (Minneapolis, MN). SNP genotyping was performed by the dye terminator-based SNaPshot method (Applied Biosystems, Foster City, CA). SNP analysis was performed on the ABI 3130 genetic analyzer (Applied Biosystems). Genotypes of the SNPs were determined by Genemapper software (Applied Biosystems). All SNPs reported in this manuscript had a genotyping success rate >96 percent and accuracy as judged by random re-genotyping of 10 percent of the samples in the subject group. Six SNPs in SERPING1 were genotyped. The PCR and SNaPshot primers are listed in Table 2.

**Haplotype analysis:** Haplotype analysis was performed using Haploview 4.0. We performed the haplotype analysis following the instructions from the Broad Institute. If the genotype was not available, the genotype was set as 0.

**Statistical analysis:** Hardy–Weinberg equilibrium (HWE) for each SNP polymorphism was tested by the χ² test with df=1. P values of the SNPs were calculated using an additive model. Haplotype frequencies between cases and controls were compared by χ² analysis. The unadjusted odds ratios of the alleles and genotypes were estimated by the χ² test. All statistical analyses were performed using the software SPSS, (SPSS, Chicago, IL) version 10.0 [18–24].

### RESULTS

**Single nucleotide polymorphism analysis:** All six SNPs selected were successfully genotyped and all of these SNPs were within HWE in both case (p>0.001, Table 3) and control groups (p>0.05, Table 3). The SNP frequencies in this study were similar to those of Han Chinese Beijing (HCB) available...
in HapMap3 in the International HapMap Project. None of the six SNPs showed significant association with AMD or subphenotypes of AMD including wet AMD or soft drusen, which are landmarks of early AMD even before a stringent Bonferroni correction (p>0.05, Table 3). SNP rs2511989 was reported to be the most significant association of SNP in the SERPING1 gene with AMD in previous studies [25]. Although rs2511989 showed high polymorphism, no association between this SNP and AMD was observed in the Chinese population (p=0.76 for all AMD; p=0.61 for CNV AMD; p=0.77 for soft drusen).

**Haplotype association analysis:** We then performed haplotype analysis using Haploview 4.0, and 14, 15, and 14 haplotypes were observed in the AMD-control, wet AMD-control, and drusen-control groups, respectively. We found that haplotype TGTCGG and haplotype CGCCGG were shown to have a significant difference between both AMD-control (p=0.0064, p=0.006, respectively, Table 4) and wet AMD-control groups (p=0.0042, p=0.025, respectively, Table 4). The haplotype CGTGCC was shown to have a significant difference between both AMD-control (p=0.0102, Table 4) and drusen-control (p=0.032, Table 4). In addition, the haplotype CGTGGT was shown to have a significant difference between wet AMD and controls (p=0.026, Table 4). But none of the haplotypes were shown to have a significant difference between cases and controls (p>0.05, Table 4) after a stringent Bonferroni correction. On the other hand, the haplotype CGTGGCA was shown to be significantly associated with soft drusen in our subject group (p=7.87x10^-5; Table 4) with frequencies of 0.11 in cases and 0.03 in controls, even after a stringent Bonferroni correction (p=0.0011, Table 4). This haplotype conferred a 3.72-fold (95% CI: 1.83–7.54) increased likelihood of dry AMD (Table 4). Additionally, the haplotype CGTACA was also shown to have a significant difference between both all AMD-control and wet AMD-control groups (p=0.05, Table 4) after a stringent Bonferroni correction. However, the frequency of this haplotype was low and it was absent in the controls.

**DISCUSSION**

Although genes in complement pathways, including CFH, C2/ BF, and C3 [2, 8–17] and chr.10q26 (LOC387715/ HTRA1) [18–24], have been identified as related to AMD, these loci could not explain all genetic contributions to AMD, suggesting that additional genetic variants related to AMD have not yet been found. Based on the candidate gene approach, Ennis et al. [25] reported that SNPs in SERPING1 were significantly associated with AMD in two Caucasian populations. Additional evidence for SERPING1 involving AMD includes: 1) SERPing1 gene encoding C1INH plays an
important role in complement pathways, which have been confirmed to participate in the pathogenesis of AMD; and 2) SERPING1 was expressed in both retinal and RPE-choroid layers in RT–PCR and immunofluorescence studies [25,29]. AMD affection status was correlated with increased abundance of choroidal C1INH [29]. Complement activation pathways include lectin, classical and alternative pathways. SERPING1 encodes C1INH, an inhibitor of the classical and lectin pathways of complement activation. The classical complement pathway is initiated by the C1 complex, which comprises a C1q hexamer complex with a zymogenic (C1r)–(C1s) 2. SERPING1 irreversibly inhibits C1r and C1s, MASP-1 (mannan-binding lectin serine peptidase 1), and MASP-2 (mannan-binding lectin serine peptidase 2, the C1s ortholog in the lectin pathway), as well as modulating the complement activation through inhibition unrelated to proteases [30–33]. However, Park et al. [26] were unable to replicate the association between the genetic variation in SERPING1 and AMD in two large and well characterized Caucasian subject groups, and Allikmets et al. [34] were also unable to confirm that SERPING1 is involved in AMD.

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**Table 4. SERPING1 haplotype association with AMD in the Han Chinese subject group.**

| Type of AMD | Haplotype | Frequency | Haplotype association (p-value) | Bonferroni correction (p value) | Odds ratio (95% CI) |
|-------------|-----------|-----------|---------------------------------|---------------------------------|-------------------|
| All AMD     | H1:CATGTG 0.63 | 0.57 | 0.0609 |                                |                   |
|             | H2:CGTGTC 0.04 | 0.06 | 0.1091 |                                |                   |
|             | H3:GTGCG 0.06 | 0.02 | 0.0064 | 0.0900                         |                   |
|             | H4:GTGCA 0.03 | 0.04 | 0.3301 |                                |                   |
|             | H5:CGTGCG 0.04 | 0.03 | 0.2641 |                                |                   |
|             | H6:CGTGCG 0.02 | 0.05 | 0.0102 | 0.1428                         |                   |
|             | H7:CATGTA 0.02 | 0.03 | 0.0105 |                                |                   |
|             | H8:CATATG 0.02 | 0.03 | 0.5689 |                                |                   |
|             | H9:TATGTC 0.02 | 0.01 | 0.3679 |                                |                   |
|             | H10:CGGTCA 0.02 | 0.01 | 0.5900 |                                |                   |
|             | H11:CGTACA 0.03 | 0.00 | 0.0008 | 0.0112                         |                   |
|             | H12:CGTACG 0.01 | 0.02 | 0.4070 |                                |                   |
|             | H13:CGTGTA 0.01 | 0.02 | 0.0628 |                                |                   |
|             | H14:CGTCG 0.00 | 0.02 | 0.0060 | 0.0840                         |                   |
| Wet AMD     | H1:CATGTG 0.63 | 0.57 | 0.0717 |                                |                   |
|             | H2:CGTGTC 0.03 | 0.06 | 0.0946 |                                |                   |
|             | H3:GTGCA 0.04 | 0.04 | 0.7668 |                                |                   |
|             | H4:CTGCG 0.06 | 0.02 | 0.0042 | 0.0630                         |                   |
|             | H5:CGTGCG 0.02 | 0.05 | 0.0620 |                                |                   |
|             | H6:CATGTA 0.02 | 0.03 | 0.6366 |                                |                   |
|             | H7:CGTGCA 0.02 | 0.03 | 0.5447 |                                |                   |
|             | H8:CATATG 0.02 | 0.03 | 0.2741 |                                |                   |
|             | H9:GGTCG 0.02 | 0.01 | 0.5848 |                                |                   |
|             | H10:TATGTC 0.02 | 0.01 | 0.3616 |                                |                   |
|             | H11:CGTACG 0.01 | 0.02 | 0.6028 |                                |                   |
|             | H12:CGTCG 0.00 | 0.02 | 0.0256 | 0.3840                         |                   |
|             | H13:CGTACG 0.03 | 0.00 | 0.0003 | 0.0045                         |                   |
|             | H14:CGTCG 0.01 | 0.01 | 0.3513 |                                |                   |
|             | H15:GTGTA 0.00 | 0.02 | 0.0263 | 0.3950                         |                   |
| Drusen AMD  | H1:CATGTG 0.62 | 0.57 | 0.2702 |                                |                   |
|             | H2:CGTGTC 0.04 | 0.06 | 0.5894 |                                |                   |
|             | H3:CTGCG 0.11 | 0.03 | 7.87E-05 | 1.10E-03 | 3.72 (1.83–7.54) |
|             | H4:CTGTCG 0.01 | 0.05 | 0.0317 | 0.4438                         |                   |
|             | H5:GTGAC 0.01 | 0.04 | 0.0909 |                                |                   |
|             | H6:CATATG 0.04 | 0.03 | 0.6489 |                                |                   |
|             | H7:CTGCG 0.05 | 0.02 | 0.1275 |                                |                   |
|             | H8:CATGT 0.02 | 0.03 | 0.3929 |                                |                   |
|             | H9:CTGTA 0.01 | 0.02 | 0.6998 |                                |                   |
|             | H10:CGTCG 0.00 | 0.02 | 0.0894 |                                |                   |
|             | H11:TATGTC 0.02 | 0.01 | 0.3297 |                                |                   |
|             | H12:CGTCG 0.01 | 0.01 | 0.8976 |                                |                   |
|             | H13:CTGAC 0.01 | 0.02 | 0.4630 |                                |                   |
|             | H14:CTGTCG 0.00 | 0.01 | 0.2656 |                                |                   |
SNPs showed significant association with AMD and none of if especially of a different ethnicity, are important to determine SERPING1. It is really associated with AMD. None of the six SNPs showed significant association with AMD and none of the major haplotypes were observed to be significantly associated with AMD or choroid neovascularization AMD (CNV) after a stringent Bonferroni correction in our study, suggesting that SERPING1 may not be related to AMD in the Han Chinese population. In the haplotype analysis, none of the SNPs tagged the significant haplotypes. Because half of samples’ genotype data for rs11546660 and rs4926 was not available in the HapMap for the Chinese, we cannot compare the haplotype frequencies to those in the HapMap. Although four haplotypes including TGTGCG, CGCGCG, CGTGCG, and CGTGTA were shown to have significant associations with different subphenotypes of AMD, anymore after a stringent Bonferroni correction, the significant associations no longer existed, suggesting that these haplotypes were not specifically associated with AMD. Since the haplotype CGTACA was rare in all AMD (3%) and wet groups (3%), and absent in the drusen group and controls, we think that the significant association between this haplotype and AMD is not reliable. The haplotype CGTGCA was shown to be significantly associated with soft drusen in the subject group even after a stringent Bonferroni correction (p=0.0011, Table 4). Further replication studies are needed to clarify the current situation because of the limited number of soft drusen samples in this study.

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