Complete Genome Sequence of the Olive-Infecting Strain *Xylella fastidiosa* subsp. *pauca* De Donno

Annalisa Giampetruzzi,a Maria Saponari,b Rodrigo P. P. Almeida,c Salwa Essakhi,b Donato Boscia,b Giuliana Loconsole,a Pasquale Saldarelli,b

Department of Soil, Plant and Food Sciences, University of Bari Aldo Moro, Bari, Italy,a; Institute for Sustainable Plant Protection, National Research Council (CNR), Bari, Italy,b; Department of Environmental Science, Policy and Management, University of California, Berkeley, Berkeley, California, USAc

**ABSTRACT** We report here the complete and annotated genome sequence of the plant-pathogenic bacterium *Xylella fastidiosa* subsp. *pauca* strain De Donno. This strain was recovered from an olive tree severely affected by olive quick decline syndrome (OQDS), a devastating olive disease associated with *X. fastidiosa* infections in susceptible olive cultivars.

In 2013, *Xylella fastidiosa* was detected in olive trees (*Olea europaea* L.) in southern Italy (Apulia region). It represented the first outbreak of this quarantine pathogen under field conditions in the European Union, and it was the first documented event of widespread infections in this plant species. Infected trees exhibit a severe disease termed olive quick decline syndrome (OQDS). Symptoms of OQDS include yellow and brown lesions on leaf tips and margins, extensive branch and twig dieback, and subsequent tree mortality (1). Genome data and multilocus sequence typing (MLST) analyses (2, 3) showed that olive-infecting isolates of *X. fastidiosa* were genetically related to subspecies *pauca* and all harbored the sequence type 53 (ST53). In this report, we describe the complete and finished genome sequence of *X. fastidiosa* subsp. *pauca* strain De Donno, selected among the ST53-cultured isolates recovered from OQDS-affected olive trees. This strain was cultured in June 2014 from a symptomatic olive tree (40.011389 N 18.048056 E); when mechanically inoculated in different olive cultivars under experimental conditions, it caused symptoms identical to those observed in contaminated olive groves (4).

A combined strategy of sequencing by the HiSeq 4000 Illumina platform and PacBio RSII platform was performed. Illumina sequencing yielded a total of 5,700,601 2 × 150-bp high-quality paired reads, of which 1% (87,950 reads) low-quality reads were discarded. In parallel, 105,585 fastq reads, with a mean length of 8,527 bp (longest read, 56,602 bp), were obtained by PacBio sequencing. De novo hybrid genome assembly was done using both Illumina and PacBio data set with SPAdes version 3.9.0 (5, 6). The final assembly resulted in a single circular 2,508,465-nucleotide (nt) chromosome with 52% G+C content. In addition, a circular plasmid of 35,273 nt, named pXF-De Donno, with a G+C content of 49.6%, was also identified. Nucleotide coverage was, on average, 1,765.5 × for the plasmid (standard deviation [SD], 216.9 ×) and 636.5 × for the chromosome (SD, 76.2 ×). Functional annotation by submission to the NCBI Prokaryotic Genome Automatic Annotation Pipeline (PGAAP) resulted in the identification of 6 rRNA genes (2 operons), 49 tRNA loci, 2,381 genes, 2,322 protein-coding genes, 3 noncoding RNAs in the chromosome, and 39 protein-coding genes in the plasmid. The complete genome description of the strain De Donno offers insights into the biology of this devastating olive disease.
Accession number(s). This whole-genome shotgun project has been deposited in GenBank under the accession numbers CP020870 to CP020871. The version described in this paper is the first version.

ACKNOWLEDGMENTS

This work used the Vincent J. Coates Genomics Sequencing Laboratory at the University of California (UC), Berkeley, supported by the NIH S10 OD018174 Instrumentation Grant, for PacBio and Illumina library preparation.

The generated libraries were sequenced by the UC Davis Genome Center, UC Irvine. We thank Shana McDevitt (UC Facility Director) for technical assistance. The computational work has been executed on the equipment of “Rete di Laboratori Pubblici SELGE-Regione Puglia (cod. 14)” and on the IT resources made available by the ReCaS project (PONa3_00052).

The work was supported by funding from the European Union’s Horizon 2020 research and innovation program under grant agreement 635646: Pest Organisms Threatening Europe (POnTE).

REFERENCES

1. Saponari M, Boscia D, Nigro F, Martelli GP. 2013. Identification of DNA sequences related to Xylella fastidiosa in oleander, almond and olive trees exhibiting leaf scorch symptoms in Apulia (southern Italy). J Plant Pathol 95:659 – 668. https://doi.org/10.4454/JPP.V95I3.035.
2. Loconsole G, Saponari M, Boscia D, D’Attoma G, Morelli M, Martelli GP, Almeida RPP. 2016. Intercepted isolates of Xylella fastidiosa in Europe reveal novel genetic diversity. Eur J Plant Pathol 146:85–94. https://doi.org/10.1007/s10658-016-0894-x.
3. Giampetruzzi A, Chiumenti M, Saponari M, Donvito G, Italiano A, Loconsole G, Boscia D, Cariddi C, Martelli GP, Saldarelli P. 2015. Draft genome sequence of the Xylella fastidiosa CoDiRO strain. Genome Announc 3(1): e01538-14. https://doi.org/10.1128/genomeA.01538-14.
4. Saponari M, Boscia D, Altamura G, D’Attoma G, Cavalieri V, Loconsole G, Zicca S, Dongiovanni E, Palmisano F, Susca L, Morelli M, Potere O, Saponari A, Fumarola G, Di Carolo M, Tavano D, Savino VN, Martelli GP. 2016. Pilot project on Xylella fastidiosa to reduce risk assessment uncertainties, 13:EN-1013. EFSA Supporting Publication. https://doi.org/10.2903/sp.efsa.2016.EN-1013.
5. Antipov D, Korobeynikov A, McLean JS, Pevzner PA. 2016. hybridSPAdes: an algorithm for hybrid assembly of short and long reads. Bioinformatics 32:1009 –1015. https://doi.org/10.1093/bioinformatics/btv688.
6. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prijibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. doi.