PNPLA3 rs738409 C>G Variant Influences the Association Between Visceral Fat and Significant Fibrosis in Biopsy-proven Nonalcoholic Fatty Liver Disease

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Abstract

Background and Aims: Intra-abdominal visceral fat accumulation and patatin-like phospholipase domain containing 3 (PNPLA3) rs738409 G/C gene polymorphism confer a greater susceptibility to nonalcoholic fatty liver disease (NAFLD). We examined whether the relationship between visceral fat accumulation and liver disease severity may be influenced by PNPLA3 rs738409 polymorphism. Methods: The variant of PNPLA3 rs738409 was genotyped within 523 Han individuals with biopsy-confirmed NAFLD. Visceral fat area (VFA) was measured by bioelectrical impedance. Significant liver fibrosis (SF), defined as stage F ≥2 on histology, was the outcome measure of interest. Results: The distribution of PNPLA3 genotypes was CC: 27.5%, CG: 48.2%, and GG: 24.3%. Higher VFA was associated with greater risk of having SF (adjusted-odds ratio [OR]: 1.03; 95% confidence interval [CI]: 1.02–1.04, p<0.05), independent of potential confounders. Among subjects with the same VFA level, the risk of SF was greater among carriers of the rs738409 G genotype than among those who did not. Stratified analysis showed that PNPLA3 rs738409 significantly influenced the association between VFA and SF. VFA remained significantly associated with SF only among the rs738409 G-allele carriers (adjusted-OR: 1.05; 95% CI: 1.03–1.08 for the GG group; and adjusted-OR:1.03; 95% CI: 1.01–1.04 for the GC group). There was a significant interaction between VFA and PNPLA3 rs738409 genotype (Pinteraction=0.004). Conclusions: PNPLA3 rs738409 G allele has a moderate effect on the association between VFA and SF in adult individuals with biopsy-proven NAFLD. Existence of the PNPLA3 rs738409 G allele and VFA interact to increase risk of SF.

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Introduction

Nonalcoholic fatty liver disease (NAFLD) is a major health problem that affects up to nearly 30% the world’s adults. PNPLA3 refers to a spectrum of progressive liver conditions, ranging from simple steatosis (NAFL) to steatohepatitis (NASH), with varying amounts of fibrosis, and cirrhosis. Convincing evidence shows that increased intra-abdominal visceral fat accumulation is a strong predictor for the development of significant fibrosis (SF) of the liver in NAFLD. Unlike subcutaneous adipose tissue, visceral adipose tissue is anatomically related to the liver through the portal vein, and so the liver is directly exposed to higher levels of free fatty acids as well as multiple adipokines/cytokines directly released from expanded visceral adipose tissue within the portal vein, thereby promoting the development of NAFLD. Therefore, visceral fat accumulation is a key target for ther-
It is known that NAFLD is a complex and heterogeneous disease. Studies show ~20% of adults may have NAFLD, in the absence of overweight or obese status. The patatin-like phospholipase domain containing-3 (PNPLA3) rs738409 C>G variant (wild-type to mutant) is one of the strongest genetic variants that is related to a greater susceptibility to developing NASH and cirrhosis. Previous studies show that individuals with NAFLD, who carry the PNPLA3 rs738409 G allele, do not have insulin resistance or other features of metabolic syndrome. Preliminary studies also suggest that the PNPLA3 rs738409 GG genotype is associated with a lower risk of type 2 diabetes and cardiovascular disease; this finding supports the notion that the pathophysiology of NAFLD may be different among subjects carrying this genetic variant.

Thus, considering the possible differences in the pathophysiology of metabolic-related vs. PNPLA3-related NAFLD, we have tested whether PNPLA3 rs738409 may influence the effect of visceral fat area (VFA) on risk of having SF, and whether there is interaction between visceral fat content and PNPLA3 rs738409 polymorphisms, to affect liver disease severity, within a well-identified cohort of subjects with biopsy-confirmed NAFLD.

**Methods**

**Research population**

This is a cross-sectional analysis of our well-characterized Prospective Epidemic Research Specifically Of NASH (PERSONS) cohort of 1,015 ethnic Han adults with suspected NAFLD (mainly based on abnormal serum liver enzyme levels and/or evidence of hepatic steatosis on imaging techniques), who were admitted to the First Affiliated Hospital of Wenzhou Medical University (China) from July 18, 2017 to December 4, 2019, and who accepted the offer to undergo liver biopsy. As detailed in Figure 1, 492 individuals were excluded for the following reasons: (1) hepatocyte steatosis ≤5% on histology (n=80); (2) excessive alcohol intake (>140 g/week for men and >70 g/week for women, respectively) (n=112); (3) other secondary causes for hepatic steatosis (n=228); and (4) missing data for PNPLA3 rs738409 genotype or Bioimpedance measurements using InBody 720 (n=72). As a consequence of these exclusion criteria, a total of 523 adult individuals with NAFLD were included in the final analysis.

The study protocol was approved by the ethics committee of the First Affiliated Hospital of Wenzhou Medical University (protocol number #2016-246, 1 December 2016). All participants signed a written informed consent to participate in this study.

**Laboratory and clinical data**

Samples of venous blood were collected from all patients after at fast of least 8 h. Biochemical parameters were evaluated by employing an automated analyzer (Abbott AxSYM; Abbott Laboratories, Lake Bluff, IL, USA) centrally. Homeostasis model assessment of resistance of insulin (HOMA-IR) was calculated as follows: fasting insulin (mIU/L)×glucose (mmol/L)/22.5. Body mass index (BMI) was calculated by dividing weight in kilograms by height in meters-squared. Obese/overweight status was identified as BMI ≥25 kg/m². Diagnostic criteria for hypertension and diabetes have been described in our previous studies. VFA was measured within 1 day of liver biopsy. A bioelectrical impedance analyzer (BIA) (InBody 720; Biospace, Seoul, South Korea) was
used to measure VFA. The aforementioned laboratory and anthropometric variables were collected for all participants within 1 day of liver biopsy examinations.

Liver biopsy
Liver biopsy procedures have been described in detail previously. Briefly, NAFLD was defined as histological evidence of >5% of steatotic hepatocytes. Subjects with a NAFLD activity score (NAS) ≥5 (having a score of at least 1 for each histological component of hepatic steatosis, lobular inflammation and ballooning) were diagnosed as having definite NASH. Fibrosis stages were graded from 0 to 4, based on the Brunt’s histological criteria. SF of the liver was defined as histological stage F ≥2.

Analysis of PNPLA3 rs738409 polymorphism
As described previously, the MassARRAY platform (Agena Bioscience, San Diego, CA, USA) was used to assess genotype of PNPLA3 rs738409. For this genotype, we used ~20 ng of genomic DNA obtained from peripheral blood leukocytes. Locus-specific PCR as well as primers for detection were designed by the accompanying Assay Design Suite v3.1. Matrix assisted laser desorption ionization-time of flight mass spectrometry was used for detection of allele type, followed by amplification of DNA by multiplex PCR.

Statistical analysis
Continuous variables were expressed as mean±standard deviation or median with interquartile range, based on whether the distribution was normal or skewed, and then compared using the unpaired Student’s t-test or the Mann Whitney test as appropriate. Categorical variables were expressed as proportions and compared using the chi-squared test or the Fisher’s exact test as appropriate. The chi-squared test was also used to test whether PNPLA3 rs738409 genotypes were in Hardy-Weinberg equilibrium. The association between VFA and presence of SF (defined as histological stage F ≥2 on liver histology) was tested by binary logistic regression analysis. In these regression models, the association between VFA and presence of SF persisted in the analysis adjusted for potential confounders, such as sex, age, obesity/overweight, hypertension, type 2 diabetes, HOMA-IR, and serum total cholesterol, triglyceride and albumin levels. Stratified and interaction analyses were also performed to examine the effect of PNPLA3 rs738409 polymorphism on the association between VFA and SF. All data were analyzed with the R statistical package (The R Foundation; http://www.r-project.org; version 3.4.3) and Empower (R) (www.empowerstats.com; X&Y Solutions, Inc., Boston, MA, USA).

Results

Baseline characteristics
A total of 523 Chinese individuals with biopsy-confirmed NAFLD were enrolled in this study. Subjects had a mean age of 42 years and 73.8% were men. In total, 102 (19.5%) of the subjects had SF (stage F ≥2 on liver histology). The prevalence rates of hypertension and type 2 diabetes were 24.1% and 25.8% respectively. The distribution of PNPLA3 rs738409 genotypes was as follows: 144 (27.5%) had CC genotype; 252 (48.2%) had GC genotype; and 127 (24.3%) had GG genotype, respectively. This genotype distribution did not deviate from Hardy-Weinberg equilibrium. The frequency of the PNPLA3 rs738409 G variant was 0.48, similar to a previous study from China (0.45). Table 1 summarizes the baseline characteristics of study participants, stratified by the PNPLA3 rs738409 polymorphism type. Carriers of the PNPLA3 GG genotype had a significantly higher prevalence of severe steatosis and definite NASH. The three groups were well comparable in terms of sex, age, adiposity measures (including VFA), HOMA-IR score and other metabolic parameters. Notably, as shown in Table 2, after stratifying by both PNPLA3 rs738409 polymorphism and SF, values of VFA were significantly greater only among carriers of the G allele, who also had SF.

PNPLA3 rs738409 polymorphism influences the association between VFA and SF
The smoothing spline curve, obtained by a generalized additive model, showed a linear association between VFA and risk of having SF. As shown in Figure 2, as VFA increased, the likelihood of SF also progressively increased; however, it is worth noting that carriers of the PNPLA3 CC genotype had a lower risk of SF than those carrying the PNPLA3 G allele. Among individuals with the same level of VFA, the risk of SF was higher among carriers of the rs738409 G genotype than among those who did not. The smoothing spline curve clearly suggested that the PNPLA3 rs738409 G allele increased the probability of SF with increasing levels of VFA. A threshold effect analysis was also performed to examine if the slight fall in the probability of SF in the CC group with increasing levels of VFA was not statistically significant.

Association between VFA and SF
As demonstrated in Figure 3, within a logistic regression model with the presence or absence of SF as the dependent variable, there was a significant positive association between VFA and SF (adjusted-OR: 1.03, 95% CI: 1.01–1.04 in the GG group; adjusted-OR: 1.05, 95% CI: 1.03–1.08 in the GC group; adjusted-OR: 1.01, 95% CI: 0.99–1.03) among carriers of the rs738409 G allele, who also had SF. The smoothing spline curve, obtained by a generalized additive model, showed a linear association between VFA and risk of having SF. As shown in Figure 2, as VFA increased, the likelihood of SF also progressively increased; however, it is worth noting that carriers of the PNPLA3 CC genotype had a lower risk of SF than those carrying the PNPLA3 G allele. Among individuals with the same level of VFA, the risk of SF was higher among carriers of the rs738409 G genotype than among those who did not. The smoothing spline curve clearly suggested that the PNPLA3 rs738409 G allele increased the probability of SF with increasing levels of VFA. A threshold effect analysis was also performed to examine if the slight fall in the probability of SF in the CC group with increasing levels of VFA was not statistically significant.

Association between VFA and SF in different subgroups
We examined the association between VFA and risk of having SF in individuals who were stratified either by different PNPLA3 genotypes (additive or dominant models) or by other established risk factors for SF (i.e. sex, age, BMI, hypertension, diabetes and HOMA-IR). As shown in Figure 3, the significant association between VFA and SF persisted in the analysis adjusted for potential confounders (adjusted-OR: 1.03, 95% CI: 1.01–1.04 in the GC group; adjusted-OR: 1.05, 95% CI: 1.03–1.08 in the GG group) but not in the CC group (adjusted-OR: 1.01, 95% CI: 0.99–1.03). It should be noted, there was a significant interaction of PNPLA3 rs738409 genotypes on the association between VFA and risk of SF (P_interaction=0.004). When this association was assessed in a dominant genetic model, the association between VFA and
SF remained statistically significant after controlling for potential confounding variables in the GC+GG group (adjusted-OR: 1.03, 95% CI: 1.02–1.05) but not in the CC group (adjusted-OR: 1.01, 95% CI: 0.99–1.03). In addition, there was a significant association between VFA and SF when we stratified subjects into two groups, i.e. the GG vs. GC+CC subgroups. Interestingly, there was an interaction between PNPLA3 rs738409 GG and VFA (P_interaction = 0.004). These results suggested that the PNPLA3 rs738409 G allele and VFA interacted to moderately increase the risk of having SF.

Table 1. Baseline characteristics of study participants, stratified by PNPLA3 rs738409 polymorphism

|                      | All, n=523 | CC, n=144 | CG, n=252 | GG, n=127 | p         |
|----------------------|------------|-----------|-----------|-----------|-----------|
| Demographics         |            |           |           |           |           |
| Age in years         | 42.5±12.0  | 44.0±12.2 | 41.5±12.1 | 42.6±11.5 | 0.154     |
| Men                  | 381 (72.8) | 104 (72.2)| 191 (75.8)| 86 (67.7) | 0.244     |
| Metabolic risk factors|           |           |           |           |           |
| BMI in kg/m²          | 26.9±3.7   | 27.0±4.5  | 27.0±3.5  | 26.7±3.2  | 0.758     |
| Overweight/obesity   | 360 (68.8) | 119 (69.4)| 174 (69.0%)| 86 (67.7) | 0.949     |
| VFA in cm²            | 102.8±25.9 | 103.9±25.2| 102.4±25.6| 102.3±27.5| 0.838     |
| Type 2 diabetes       | 135 (25.8) | 41 (28.5) | 62 (24.6) | 32 (25.2) | 0.687     |
| Hypertension          | 126 (24.1) | 37 (25.7) | 54 (21.4) | 35 (27.6) | 0.365     |
| Laboratory parameters |            |           |           |           |           |
| AST in U/L            | 34.0 (25.0–52.5) | 31.5 (24.0–48.0) | 34.0 (25.0–54.0) | 36.0 (26.0–57.0) | 0.116 |
| ALT in U/L            | 51.0 (30.0–88.0) | 44.5 (28.8–73.8) | 52.0 (30.0–91.2) | 55.0 (30.5–89.5) | 0.109 |
| GGT in U/L            | 52.0 (32.5–84.5) | 51.5 (30.8–81.2) | 55.0 (35.0–85.2) | 49.0 (31.5–83.0) | 0.491 |
| Total bilirubin in µmol/L | 12.0 (10.0–16.0) | 12.0 (9.8–16.0) | 13.0 (10.0–16.0) | 12.0 (10.0–16.5) | 0.730 |
| Albumin in g/L        | 45.9±4.0   | 46.2±3.5  | 45.9±3.8  | 45.7±5.0  | 0.589     |
| Glucose in mmol/L     | 5.3 (4.9–6.3) | 5.4 (5.0–6.4) | 5.3 (4.9–6.2) | 5.3 (4.9–6.1) | 0.281 |
| Insulin in mIU/L      | 14.6 (9.6–21.3) | 15.2 (9.4–21.3) | 14.3 (10.0–21.2) | 14.9 (9.8–21.2) | 0.949 |
| HOMA-IR               | 3.6 (2.3–5.3) | 3.6 (2.3–5.5) | 3.4 (2.4–5.2) | 3.6 (2.2–5.5) | 0.879 |
| Total cholesterol in mmol/L | 5.1 (4.4–5.9) | 5.1±1.2 | 5.2±1.1 | 5.2±1.2 | 0.516 |
| Triglycerides in mmol/L | 1.9 (1.4–2.8) | 2.0 (1.4–3.2) | 1.9 (1.4–2.8) | 1.8 (1.4–2.5) | 0.179 |
| HDL-cholesterol in mmol/L | 1.0 (0.9–1.1) | 1.0±0.2 | 1.0±0.2 | 1.0±0.3 | 0.163 |
| LDL-cholesterol in mmol/L | 3.0 (2.4–3.6) | 2.9±0.9 | 3.1±0.9 | 3.1±1.0 | 0.055 |
| Liver histology       |            |           |           |           |           |
| Steatosis grade       | <0.001     |           |           |           |           |
| 1                     | 222 (42.4) | 83 (57.6) | 101 (40.1) | 38 (29.9) |
| 2                     | 193 (36.9) | 47 (32.6) | 97 (38.5) | 49 (38.6) |
| 3                     | 108 (20.7) | 14 (9.7)  | 54 (21.4) | 40 (31.5) |
| Hepatocyte ballooning | 0.922      |           |           |           |           |
| 0                     | 78 (14.9)  | 22 (15.3) | 38 (15.1%) | 18 (14.2) |
| 1                     | 296 (56.6) | 85 (59.0) | 141 (56.0%) | 70 (55.1) |
| 2                     | 149 (28.5) | 37 (25.7) | 73 (29.0%) | 39 (30.7) |
| Lobular inflammation  | 0.148      |           |           |           |           |
| 0                     | 64 (12.2)  | 25 (17.4) | 29 (11.5) | 10 (7.9) |
| 1                     | 306 (58.5) | 76 (52.8) | 153 (60.7) | 77 (60.6) |
| 2                     | 145 (27.7) | 41 (28.5) | 68 (27.0) | 36 (28.3) |
| 3                     | 8 (1.5)    | 2 (1.4)   | 2 (0.8)   | 4 (3.1)   |
| Definite NASH         | 202 (38.6) | 47 (32.6) | 95 (37.7) | 60 (47.2) | 0.044     |
| SF (F ≥2 stage)       | 102 (19.5) | 22 (15.3) | 49 (19.4) | 31 (24.4) | 0.167     |

Data are presented as n (%) or mean±standard deviation. ALT, alanine aminotransferase; AST, aspartate aminotransferase; GGT, gamma-glutamyltransferase; HOMA-IR, homeostasis model assessment-insulin resistance; HDL, high-density lipoprotein; LDL, low-density lipoprotein; NASH, non-alcoholic steatohepatitis.
### Table 2. Baseline characteristics of study participants, stratified by both PNPLA3 rs738409 polymorphism and SF of the liver

|                | CC                     | GC                     | GG                     | p    |
|----------------|------------------------|------------------------|------------------------|------|
|                | No SF, n=122           | SF, n=22               | No SF, n=203           | SF, n=49 | p |
| Demographics   |                        |                        |                        |      |
|                | Age in years           | 43.6±12.4              | 45.6±11.2              | 0.484 | 41.10±11.3 | 33.4±15.1 | 0.232 | 41.6±10.1 | 45.8±14.9 | 0.076 |
|                | Men                    | 91 (74.6)              | 12 (59.1)              | 0.135 | 161 (79.3) | 30 (61.2) | 0.008 |
|                | BMI in kg/m²            | 26.9±4.1               | 27.9±6.5               | 0.341 | 26.6±3.1 | 28.3±4.5 | 0.002 |
| Metabolic risk factors |                |                        |                        |      | 26.2±2.9 | 28.4±3.3 | <0.001 |
| Overweight/obesity |                        | 85 (69.7)              | 15 (68.2)              | 0.889 | 139 (68.5) | 35 (71.4) | 0.688 |
|                | VFA in cm²              | 103.3±26.2             | 107.0±19.3             | 0.534 | 99.4±24.3 | 114.7±27.7 | <0.001 |
| Type 2 diabetes |                        | 35 (28.7)              | 6 (27.3)               | 0.892 | 46 (22.7) | 16 (32.7) | 0.145 |
| Hypertension    | 34 (27.9)              | 3 (13.6)               | 0.160                  |      | 41 (20.2) | 13 (26.5) | 0.332 |
| Laboratory parameters |          |                        |                        |      | 22 (22.9) | 13 (41.9) | 0.039 |
| AST in U/L      | 31.0 (23.2–43.0)       | 52.0 (35.0–75.8)       | <0.001                 |      | 32.0 (24.0–49.0) | 54.0 (32.0–76.0) | <0.001 |
|                | 39.0 (27.0–67.5)       | 74.5 (51.2–98.5)       | <0.001                 |      | 48.0 (28.0–86.5) | 72.0 (43.0–99.0) | 0.006 |
|                | 50.0 (29.2–78.0)       | 68.5 (37.2–118.2)      | 0.058                  |      | 53.0 (32.5–83.0) | 60.0 (42.0–99.0) | 0.074 |
| Total bilirubin in µmol/L | 12.0 (10.0–15.0) | 14.0 (9.2–17.0)        | 0.566                  |      | 13.0 (10.0–16.0) | 12.0 (9.0–15.0) | 0.135 |
|                | 46.0±3.6               | 47.0±2.8               | 0.214                  |      | 46.1±3.7 | 45.3±4.1 | 0.185 |
| Albumin in g/L  | 5.5 (5.0–6.6)          | 5.3 (4.8–6.0)          | 0.338                  |      | 5.2 (4.8–6.0) | 5.6 (4.9–7.9) | 0.012 |
| Glucose in mmol/L | 15.4 (9.2–21.3)       | 13.9 (10.0–21.6)       | 0.987                  |      | 13.0 (9.2–18.4) | 21.4 (15.2–32.6) | <0.001 |
|                | 3.7 (2.3–5.4)          | 3.4 (2.2–6.3)          | 0.769                  |      | 3.2 (2.3–4.5) | 5.3 (4.1–11.1) | <0.001 |
| Insulin in mIU/L |                        | 14.0 (9.8–19.7)        | 16.9 (10.1–23.2)       | 0.268 |
| HOMA-IR         | Total cholesterol in mmol/L |                        | 5.1±1.2               | 4.6±1.4 | 0.057 | 5.1±1.0 | 5.4±1.3 | 0.074 |
|                | 2.1 (1.5–3.2)          | 1.7 (1.2–2.4)          | 0.072                  |      | 1.9 (1.4–2.9) | 2.0 (1.3–2.8) | 0.965 |
| Triglycerides in mmol/L |                | 1.0±0.2               | 0.495                  |      | 1.0±0.2 | 1.0±0.3 | 0.675 |
| HDL-cholesterol in mmol/L | 2.9±0.9               | 2.6±1.1               | 0.128                  |      | 3.0±0.8 | 3.2±1.1 | 0.245 |
| LDL-cholesterol in mmol/L |                | 76 (62.3)              | 7 (31.8)               | 0.003 | 87 (42.9) | 14 (28.6) | <0.001 |
| Liver histology | 38 (31.1)              | 9 (40.9)               | 0.345                  |      | 40 (20.2) | 13 (26.5) | 0.332 |
| Steatosis grade | 0.003                  | <0.001                 |                         |      | 34 (35.4) | 4 (12.9) |
| 1              | 37 (38.5)              | 12 (38.7)              |                         |      | 19 (19.8) | 13 (41.9) | 0.014 |
| 2              | 16 (16.2)              | 34 (35.4)              |                         |      | 19 (19.8) | 13 (41.9) | 0.014 |
We examined the association between VFA with SF, stratified by a PNPLA3 rs738409 dominant model, in men and in women separately. As described in Table 3, in the unadjusted models, VFA was associated with an increased risk of having SF in both sexes. After adjustment for age, overweight/obese, type 2 diabetes and hypertension, levels of serum total cholesterol, triglycerides and albumin, and HOMA-IR (adjusted model 2), the association between VFA and SF remained statistically significant for both men and women (adjusted-OR: 1.03, 95% CI: 1.01–1.04 for men; adjusted-OR: 1.02, 95% CI: 1.0–1.05 for women). However, as also shown in Table 3, after further stratification by PNPLA3 rs738409 genotypes (CC vs. GC+GG groups), the significant association between VFA and SF disappeared among carriers of the rs738409 CC genotype (adjusted-OR: 1.01, 95% CI: 0.99–1.04 for men; adjusted-OR: 0.98, 95% CI: 0.95–1.02 for women). In contrast, the association between VFA and SF remained significant among carriers of the rs738409 G allele, even after adjustment for potential confounders in both sexes (adjusted-OR: 1.03, 95% CI: 1.02–1.05 for men; adjusted-OR: 1.04, 95% CI: 1.01–1.07 for women).

**Discussion**

In this large cross-sectional study of ethnic Han individuals with biopsy-confirmed NAFLD, we found that intra-abdominal VFA was significantly associated with greater risk of having SF, on liver histology. Notably, this significant association persisted even after adjusting for potential confounding variables, such as sex, age, obese/overweight status, hypertension, diabetes, HOMA-IR, and levels of plasma lipids and albumin. Furthermore, after further stratification by PNPLA3 rs738409 polymorphism type, the association between VFA and SF remained significant only among carriers of the PNPLA3 rs738409 G allele but not among those carrying the CC genotype, thereby suggesting that the PNPLA3 rs738409 G allele and VFA can interact to moderately increase the risk of having SF. Furthermore, with the same level of VFA, the risk of having SF was significantly lower among carriers of the rs738409 CC genotype than among those carrying the rs738409 G allele.

In the last decade, the close inter-relationship between intra-abdominal VFA and SF in people with NAFLD has drawn increasing attention. Unlike subcutaneous fat in the abdomen, intra-abdominal visceral fat accumulation (being connected directly to the liver via the portal vein) is closely related to the development and progression of NAFLD. In our study, we found that VFA was associated with greater risk of SF, independent of pre-existing diabetes or other metabolic syndrome features, especially among carriers of the PNPLA3 CG or GG genotypes. The precise mechanisms underpinning the association between increased VFA and greater risk of SF are not fully understood. However, in accordance with the so-called "portal theory", it has been proposed that expanded and dysfunctional visceral adipose tissue may release higher amounts of free fatty acids as well as multiple adipokines and pro-inflammatory cytokines into the liver via portal vein, thus promoting the development and progression of NAFLD.

We found that compared to NAFLD subjects carrying the PNPLA3 rs738409 CC genotype, VFA was independently related to a greater risk of having SF only among those carrying the PNPLA3 rs738409 G allele. It is known that the PNPLA3 rs738409 C>G variant (wild-type to mutant), leading to an isoleucine to methionine substitution at position 148 of the protein (I148M), is strongly associated...
with an increased risk of NAFLD progression. As a liver lipase with triglyceride hydrolase enzyme activity, this genetic variant leads to loss of function, thereby reducing the remodeling of polyunsaturated fatty acids and monounsaturated fatty acids, leading to their retention within the liver. Therefore, it is conceivable that the combination of increased VFA and PNPLA3 rs738409 C>G variant may promote the progression of NAFLD from simple steatosis to NASH and cirrhosis.

We believe that the presence of an interaction effect of PNPLA3 rs738409 G allele and VFA to moderately increase risk of SF, and the observed dissociation of VFA and SF among the carriers of the PNPLA3 rs738409 CC genotype are two interesting findings of our study. However, the specific reasons for these results are not entirely known. In particular, the effect as well as role of the PNPLA3 rs738409 G variant within adipose tissue are poorly understood. Recently, it has been shown that PNPLA3 mRNA was expressed abundantly within the liver and clearly detectable within the subcutaneous adipose tissue of individuals with severe obesity. Other investigators confirmed that PNPLA3 protein was found not just within the liver but also within adipose tissue. It has been reported that the PNPLA3 rs738409 C>G variant may alter lipid composition of adipose tissue in a similar way to that observed in the liver. An experimental study also suggested that overexpression of the PNPLA3 rs738409 G variant lead to greater VFA and insulin resistance compared to the wild-type protein in mice. Although it remains uncertain how the PNPLA3 rs738409 polymorphism may interact with VFA to increase hepatic fibrogenesis, our results support the existence of a cross-talk between VFA and PNPLA3 rs738409 polymorphism in risk of NAFLD progression.

In our study, we enrolled patients with biopsy-proven NAFLD who also had measurement of VFA and PNPLA3 single nucleotide polymorphism status. There was a significant trend for patients with more SF (stage 2 or more) to have increased VFA or be a carrier for the G allele. Consequently, we thought it would also be a valuable point if those with negative biopsies were analyzed to understand if the VFA was significantly different in this population as well as the status of PNPLA3. Unfortunately, we did not enroll such patients in our cohort. Further studies are required to address and resolve this point in the future.

There are several essential limitations within our investigation. First, the mutation rates for PNPLA3 rs738409 polymorphism vary among different ethnic populations, with the highest rates being in Asian and American individuals, intermediate rates in northern European Whites, and lowest rates in Blacks. For example, according to a previous study, the risk allele mutation for PNPLA3 was in 49% among Hispanics, followed by non-Hispanic Caucasians (23%) and African Americans (17%). As the participants in our study were all ethnic Han Chinese individuals, the findings of our study might not be generalizable to other ethnic groups.

Second, the cross-sectional design of our study does not
allow any firm conclusions about causality. However, since PNPLA3 rs738409 polymorphism is inherited, reverse causation does not apply. Third, VFA was not measured with computed tomography (CT) scan. However, VFA estimated by BIA has a good correlation with VFA measured with CT scanning. Finally, we did not have detailed information on physical activity levels and diet regimens of these participants. The beneficial effect of different exercise regimes, without caloric restriction, on VFA is well known for overweight or obese individuals.

In conclusion, our research showed that VFA is associated with greater risk of having SF, independent of potential confounding factors, especially among carriers of the PNPLA3 rs738409 G-allele, and there is an interaction of PNPLA3 rs738409 polymorphism and VFA to increase risk of SF in Chinese individuals with biopsy-proven NAFLD. Our gene-visceral fat interaction study suggests that the PNPLA3 rs738409 G-allele may moderately modulate the adverse effects of VFA on risk of SF in NAFLD. However, further research is needed to further corroborate these findings in other different cohorts of NAFLD patients.

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Conflict of interest

MHZ has been an associate editor of *Journal of Clinical and Translational Hepatology* since 2013. Other authors have no conflict of interests related to this publication.

Author contributions

Study concept and design (GL, MHZ), acquisition of data (GL, HLM, LJT, OYH, XYP, PWZ, RSR, KIZ), pathology analysis (GL, HLM), critical revision of the manuscript (GT, CDB), statistical analysis (SDC), drafting of the manuscript (GL, MHZ), critical revision of the manuscript for important intellectual content and approved the submission.

Ethics approval

The study was approved by the local ethics committee of our hospital.

Patient consent

Written informed consent was obtained from all study participants. Personal information and identifying record data were omitted and de-identified prior to analysis.

Data sharing statement

The data underlying the results of this study are available upon request because they contain potentially sensitive information. Interested researchers can contact the corresponding author for data access requests via email (zhengmh@wmu.edu.cn). This work is a part of the PERSONS study.

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Journal of Clinical and Translational Hepatology 2022 vol. 10(3) | 439–448
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