Lipoprotein (a) interactions with cholesterol-containing lipids on angiographic coronary collateralization in type 2 diabetic patients with chronic total occlusion

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Abstract

Background: We investigated whether or to what extent the interaction of lipoprotein (a) [Lp(a)] with cholesterol-containing lipids was associated with angiographic coronary collateralization in type 2 diabetic patients with chronic total occlusion.

Methods: Serum levels of Lp(a), total cholesterol, low-density lipoprotein–cholesterol (LDL-C), high-density lipoprotein–cholesterol (HDL-C), and triglyceride were determined and non-HDL-C was calculated in 706 type 2 diabetic and 578 non-diabetic patients with stable coronary artery disease and angiographic total occlusion of at least one major coronary artery. The degree of collaterals supplying the distal aspect of a total occlusion from the contra-lateral vessel was graded as poor (Rentrop score of 0 or 1) or good coronary collateralization (Rentrop score of 2 or 3).

Results: For diabetic and non-diabetic patients, Lp(a), total cholesterol, LDL-C, and non-HDL-C levels were higher in patients with poor coronary collateralization than in those with good collateralization, whereas HDL-C and triglyceride levels were similar. After adjustment for potential confounding factors, tertiles of Lp(a), total cholesterol, LDL-C and non-HDL-C remained independent determinants for poor collateralization. A significant interaction between Lp(a) and total cholesterol, LDL-C or non-HDL-C was observed in diabetic patients (all P interaction < 0.001) but not in non-diabetics. At high tertile of total cholesterol (≥ 5.35 mmol/L), LDL-C (≥ 3.36 mmol/L) and non-HDL-C (≥ 4.38 mmol/L), diabetic patients with high tertile of Lp(a) (≥ 30.23 mg/dL) had an increased risk of poor collateralization compared with those with low tertile of Lp(a) (< 12.66 mg/dL) (adjusted OR = 4.300, 3.970 and 4.386, respectively, all P < 0.001).

Conclusions: Increased Lp(a) confers greater risk for poor coronary collateralization when total cholesterol, LDL-C or non-HDL-C are elevated especially for patients with type 2 diabetes.

Keywords: Lipoprotein (a), Low-density lipoprotein, Non-high-density lipoprotein cholesterol, Coronary collateral circulation, Stable coronary artery disease

Background

Abundant evidence suggests that in the case of atherothrombotic coronary artery obstruction, coronary collateral formation compensates myocardial ischemia [1, 2] and improves patients’ clinical outcome and even survival [3, 4]. The mechanism of collateral vessel growth is complex involving arteriogenesis which pertains to the remodeling of preexisting arterial vessels through the anatomic increase in lumen area and wall thickness and angiogenesis defined as new capillaries that stem from...
cholesterol (HDL-C) is an established coronary risk factor with high levels and bioactivity of vascular endothelial growth factor [14, 16]. Hypercholesterolemia particularly with high levels of ischemic myocardium to pro-angiogenic factors [22, 23]. However, the impact of plasma lipid levels on coronary collateralization in diabetes remains unclear. In this study, we hypothesized that the interaction of genetic lipoprotein (a) [Lp(a)] with environmental cholesterol-containing lipids (i.e., total cholesterol, LDL-C, and non-HDL-C) is associated with angiographic coronary collateralization in type 2 diabetic patients with stable coronary artery disease and chronic total occlusion.

Methods

Study population

A total of 1655 consecutive patients with stable coronary artery disease and chronic total occlusion (≥ 3 months) of at least one major epicardial coronary artery between May 2010 and November 2018 were screened from the database of Shanghai Rui Jin Hospital PCI Outcomes Program. This angiographic inclusion criterion of study patients was used because a severe coronary artery obstruction was a prerequisite for spontaneous collateral formation [4]. The duration of coronary artery occlusion was estimated from the date of occurrence of myocardial infarction in the area of myocardium supplied by the occluded vessel, from an abrupt worsening of existing angina pectoris, or from information obtained from a previous angiogram. For the purpose of this study, 258 patients were excluded because of PCI within the last 3 months (n = 98), a history of coronary artery bypass grafting (CABG) (n = 93), renal failure requiring hemodialysis (n = 7), chronic heart failure with NYHA class III or IV (n = 26), pulmonary heart disease (n = 25) and malignant tumor or immune system disorders (n = 9), as these conditions could influence collateral formation. Patients with type 1 diabetes (n = 11) were excluded by measurement of C-peptide level [5]. One hundred and two patients were further excluded due to unavailability of lipid profile. Thus, the remaining 1284 patients were enrolled in the final analyses. Among them, 706 patients had type 2 diabetes and 578 were non-diabetics (Fig. 1).

The diagnosis of type 2 diabetes was made according to the criteria of the American Diabetes Association, including glycosylated hemoglobin (HbA1c) ≥ 6.5%, fasting plasma glucose concentration ≥ 7.0 mmol/L, 2-h postprandial glucose concentration ≥ 11.1 mmol/L, or a random plasma glucose ≥ 11.1 mmol/L in a patient with classic symptoms of hyperglycemia or hyperglycemic crisis [24]. Hypertension was defined as systolic blood pressure ≥ 140 mmHg and/or diastolic blood pressure ≥ 90 mmHg, or use of anti-hypertensive agents for controlling blood pressure [25]. Dyslipidemia was defined according to the Third Report of The National Cholesterol Education Program (NCEP) [26]. Stable angina was diagnosed according to the criteria recommended by the American College of Cardiology/American Heart Association [27].

the budding of preexisting capillary vessels. Angiogenesis is likely caused by a combination of mechanical (shear stress) and chemical factors (related to ischemia and genes activated by ischemia), whereas angiogenesis is thought to be related to tissue hypoxia and the chemical factors produced under these conditions [2, 3]. These processes of growth and maturation of coronary collateral vessels are also influenced by multiple clinical and biochemical factors, inflammatory cytokines, and growth factors [5–9].

Lipoprotein (a) [Lp(a)] which is genetically determined contains principally a cholesterol rich low-density lipoprotein particle, one molecule of apolipoprotein (apo) B-100 and an apo (a) [10], and represents an atherothrombogenic lipoprotein [11]. Although the distribution of serum Lp(a) levels is very skewed, elevated circulating Lp(a) has emerged as an independent and causal cardiovascular risk factor and an important predictor of adverse outcomes for both general and higher risk populations [12], especially when low-density lipoprotein cholesterol (LDL-C) levels are elevated [13]. Previous studies with a small sample size have suggested an inverse relation between serum levels of Lp(a) and development of coronary collateral circulation [14, 15], with high levels of Lp(a) associated with reduced production and bioactivity of vascular endothelial growth factor [14, 16]. Hypercholesterolemia particularly with high levels of LDL-C and/or low levels of high-density lipoprotein cholesterol (HDL-C) is an established coronary risk factor that induces endothelial cell dysfunction and impairs collateral vessel growth [17]. Non-high-density lipoprotein cholesterol (non-HDL-C)—the sum of cholesterol in other lipoproteins except for high-density lipoprotein—is closely associated with coronary atheroma progression and cardiovascular outcome, and has been proposed to improve risk estimation beyond total cholesterol and/or LDL-C [18, 19], especially for individuals with LDL-C levels that are not high or have already reached the treatment goal when the triglyceride level is elevated [20].

Diabetes mellitus represents a powerful independent risk factor for increased cardiovascular mortality associated with coronary artery disease, partly because of an impaired physiological adaptive response of coronary collateral circulation [5, 7]. Hyperglycemia induces microvascular rarefaction in the myocardium even without ischemia, and capillary density further decreased in chronic ischemia hearts [21]. Patients with type 2 diabetes often have substantially adverse functional and structural remodeling of the coronary arterioles and even amongst those without known coronary artery disease, suggesting that diabetes may destabilize microvascular vessels of the heart and impair the responsiveness of ischemic myocardium to pro-angiogenic factors [22, 23].
The study protocol was approved by the Institutional Review Board of Rui Jin Hospital, Shanghai Jiaotong University School of Medicine and clinical investigation was conducted according to the principle of the Declaration of Helsinki. Written informed consent was obtained from all patients.

### Biochemical investigation
Blood samples were obtained at the day of angiography in all patients after an overnight fasting. Serum levels of creatinine, lipid profiles (including triglyceride, total cholesterol, LDL-C, HDL-C, and non-HDL-C), glucose, and glycated hemoglobin (HbA1c) were determined with standard laboratory techniques [7–9]. Glomerular filtration rate (GFR) was estimated using the Chronic Kidney Disease Epidemiology Collaboration (CKD-EPI) equation:

$$GFR_{\text{EPI}} \text{ (mL/min/1.73 m}^2) = 141 \times \min (\text{creatinine/k, 1})^{\alpha} \times \max (\text{creatinine/k, 1})^{-1.209} \times 0.993^{\text{age}} \times 1.018 \text{ [if female]},$$

where $k$ is 0.7 for females and 0.9 for males, $\alpha$ is $-0.329$ for females and $-0.411$ for males, min indicates the minimum of creatinine/k or 1, and max indicates the maximum of creatinine/k or 1 [28]. Serum Lp(a) and high-sensitivity C-reactive protein (hsCRP) levels were assayed by ELISA (Biocheck Laboratories, Toledo, OH, USA).

### Angiography and collateral grading
Coronary angiography was performed through the femoral or radial access with 6 Fr diagnostic catheters. All angiograms were analyzed independently by two blinded interventional cardiologists. The degree of coronary artery disease was assessed according to lesion classification scheme of the American College of Cardiology/American Heart Association [29]. The presence and degree of coronary collateralization from the contralateral vessel (often via connections of the epicardial surface or intraventricular septum) was visually estimated using the Rentrop scoring system, as this angiographic assessment of coronary collaterals is routinely applied in clinical practice [7–9, 30]. In patients with more than one chronic total occlusion, the vessel with the highest collateral grade was chosen for analysis. In case of disagreement, the difference in interpretation was resolved by a third reviewer.
Statistical analysis
Data are presented as mean ± standard deviation (SD) and number (percentages). For continuous variables, differences between groups were evaluated by t test for normally distributed values; otherwise, the Mann–Whitney U test was applied. For categorical variables, differences between groups were evaluated with the Chi-square test. To examine the relationships between Lp(a) and other cholesterol-containing lipid profiles, we employed Pearson's correlation. The serum levels of Lp(a) and lipid profile (total cholesterol, LDL-C, non-HDL-C, HDL-C, triglyceride) were divided into 3 groups according to tertile distribution, respectively. Univariable and multivariable logistic regression analyses after adjustment for age, gender, body mass index (BMI), traditional risk factors for coronary artery disease including smoking, hypertension, hyperlipidemia and diabetes, prior myocardial infarction, multi-vessel disease, GFR, hsCRP, left ventricular ejection fraction and use of statins were performed to detect the relationship between poor collateralization and serum levels of Lp(a) and cholesterol-containing lipids (total cholesterol, LDL-C, HDL-C and non-HDL-C). All analyses used 2-sided tests with an overall significance level of alpha = 0.05. SPSS 20.0 for Windows (SPSS, Inc., Chicago, IL, USA) was used for statistical analyses.

Results
Baseline characteristics
Among overall 1284 patients, poor coronary collateralization occurred in 323 diabetic (45.8%) and 182 non-diabetic patients (31.5%), respectively (P < 0.001). Both diabetic and non-diabetic patients with poor coronary collateralization were older, females and cigarette smokers in higher percentage and had more dyslipidemia but were less hypertensive than those with good collateralization (for all comparisons, P < 0.05). Biochemical tests showed hsCRP levels were more elevated but GFR was lower in patients with poor collateralization. There were no significant differences in the severity of coronary artery disease and medical treatments between the two groups (Table 1).

Lp(a) and lipid profile
In diabetic and non-diabetic settings, patients with poor coronary collateralization had higher serum levels of Lp(a), total cholesterol, LDL-C, and non-HDL-C compared to those with good collateralization (P ≤ 0.001), but HDL-C and triglyceride levels were similar (Table 1). In diabetic patients, Lp(a) correlated with total cholesterol (adjusted r = 0.080, P = 0.035), LDL-C (adjusted r = 0.076, P = 0.045), non-HDL-C (adjusted r = 0.090, P = 0.017) and triglyceride (adjusted r = − 0.113,

P = 0.003) but was not related to HDL-C (P = 0.231) after adjustment for gender, age, BMI, risk factors for coronary artery disease (hypertension, dyslipidemia, smoking), prior myocardial infarction, multi-vessel disease, renal function, log-transferred hsCRP and left ventricular ejection fraction. In non-diabetic patients, such a significant correlation was not found (P = 0.053–0.087). After adjustment for these potential risk factors, tertiles of Lp(a) (adjusted OR = 1.366, 95% CI 1.108–1.684, P = 0.003 and adjusted OR = 1.432, 95% CI 1.119–1.831, P = 0.004), total cholesterol (adjusted OR = 1.814, 95% CI 1.393–2.361, P < 0.001 and adjusted OR = 1.820, 95% CI 1.358–2.440, P < 0.001), LDL-C (adjusted OR = 1.830, 95% CI 1.407–2.381, P < 0.001 and adjusted OR = 1.699, 95% CI 1.270–2.274, P < 0.001) and non-HDL-C (adjusted OR = 1.810, 95% CI 1.386–2.364, P < 0.001 and adjusted OR = 1.912, 95% CI 1.407–2.597, P < 0.001) remained independent determinants for poor collateralization in diabetic and non-diabetic patients (Table 2). Three lipid measurements (total cholesterol, LDL-C and non-HDL-C) with significant difference between poor and good collaterals (Table 1) were chosen for further adjustment, and Lp(a) was still independently associated with collateralization in diabetics and non-dietetics (Additional file 1: Table S1). The result patterns were similar in additional analysis with the quartiles of Lp(a), total cholesterol, LDL-C, non-HDL-C, HDL-C and triglyceride (Additional file 2: Table S2).

For patients with diabetes, there was a significant interaction between Lp(a) and total cholesterol, LDL-C or non-HDL-C in relation to poor coronary collateralization (all adjusted P interaction < 0.001). At high tertile of total cholesterol (≥ 5.35 mmol/L), LDL-C (≥ 3.36 mmol/L) and non-HDL-C (≥ 4.38 mmol/L), patients with high tertile of Lp(a) (≥ 30.23 mg/dL) had a significantly increased risk of poor collateralization compared with those with low tertile of Lp(a) (< 12.66 mg/dL) (adjusted OR = 4.300, 95% CI 2.095–8.826, adjusted OR = 3.970, 95% CI 1.918–8.216 and adjusted OR = 4.386, 95% CI 2.115–9.094, respectively, all P < 0.001) (Fig. 2). Furthermore, the additional inclusion of interaction of Lp(a) with total cholesterol, LDL-C and non-HDL-C provided better risk prediction of poor coronary collateralization with a significantly improved goodness-of-fit and predictive performance with an increase of Nagelkerke R² of 3.4% (< 0.001) for total cholesterol, 3.7% (P < 0.001) for LDL-C and 3.4% (P < 0.001) for non-HDL-C, respectively, and C statistic of 0.019 (95% CI 0.003–0.034, P = 0.016) for total cholesterol, 0.019 (95% CI 0.003–0.034, P = 0.016) for LDL-C and 0.018 (95% CI 0.003–0.033, P = 0.020) for non-HDL-C, respectively. However, there was no interaction of Lp(a) with HDL-C and triglyceride on coronary
Table 1 Baseline characteristic, biochemical assessment and medication in diabetic and non-diabetic patients with poor and good collateralization

|                          | Overall  | Diabetes | Non-diabetes |
|--------------------------|----------|----------|--------------|
|                          | Poor     | Good     | Poor         | Good         | P value |
|                          | collateralization | collateralization | collateralization | collateralization | |
| n = 505                  | n = 323  | n = 383  | n = 182      | n = 396      |         |
| Female, n (%)            | 137 (27.1) | 120 (15.4) | < 0.001     | 97 (30.0)    | 75 (19.6) | 0.001 |
| Age                      | 66.7 ± 10.2 | 62.6 ± 10.4 | < 0.001     | 66.9 ± 10.4  | 63.4 ± 10.3 | < 0.001 |
| BMI, kg/m²               | 25.2 ± 3.1 | 25.3 ± 3.2 | 0.330       | 25.2 ± 3.0  | 25.5 ± 3.3 | 0.358 |
| Hypertension, n (%)      | 313 (62.0) | 562 (72.1) | < 0.001     | 202 (62.5)  | 285 (74.4) | < 0.001 |
| Diabetes mellitus, n (%) | 323 (64.0) | 383 (49.2) | < 0.001     | 323 (100.0) | 383 (100.0) | –       |
| Dyslipidemia, n (%)      | 137 (27.1) | 110 (14.1) | < 0.001     | 101 (31.3)  | 76 (19.8)  | < 0.001 |
| Smoking, n (%)           | 214 (42.4) | 238 (30.6) | < 0.001     | 137 (42.4)  | 116 (30.3) | < 0.001 |
| Prior MI, n (%)          | 150 (29.7) | 187 (24.0) | 0.023       | 91 (28.2)   | 87 (22.7)  | 0.096 |
| Severity of CAD, n (%)   | 1.001     | 0.439    |            |             |          |
| 1-vessel                 | 88 (17.4) | 139 (17.8) | 0.848       | 46 (14.2)   | 65 (17.0)  | 0.321 |
| 2-vessel                 | 152 (30.1) | 221 (28.4) | 0.505       | 94 (29.1)   | 98 (25.6)  | 0.296 |
| 3-vessel                 | 265 (52.5) | 419 (53.8) | 0.645       | 183 (56.7)  | 220 (57.4) | 0.834 |
| Multi-vessel             | 417 (82.6) | 640 (82.2) | 0.848       | 277 (85.6)  | 318 (83.0) | 0.321 |
| SBP, mmHg                | 136.8 ± 22.3 | 138 ± 20.6 | 0.283       | 137.6 ± 22.5 | 140.2 ± 19.8 | 0.107 |
| DBP, mmHg                | 78.0 ± 12.9 | 82.3 ± 12.4 | < 0.001   | 78.2 ± 13.2 | 83.2 ± 12.4 | < 0.001 |
| FBG, mmol/L              | 6.09 ± 2.38 | 5.83 ± 2.02 | 0.044      | 6.69 ± 2.74 | 6.61 ± 2.54 | 0.693 |
| HbA1C, %                 | 6.62 ± 1.27 | 6.46 ± 1.35 | 0.027      | 7.07 ± 1.37 | 7.09 ± 1.58 | 0.899 |
| Triglyceride, mmol/L     | 1.74 ± 0.94 | 1.76 ± 1.11 | 0.786      | 1.86 ± 1.06 | 1.83 ± 1.18 | 0.715 |
| Total cholesterol, mmol/L| 5.00 ± 1.02 | 4.64 ± 1.08 | < 0.001   | 5.10 ± 1.00 | 4.77 ± 1.12 | < 0.001 |
| LDL-C, mmol/L            | 3.22 ± 0.81 | 2.94 ± 0.83 | < 0.001   | 3.27 ± 0.79 | 3.01 ± 0.88 | < 0.001 |
| HDL-C, mmol/L            | 0.98 ± 0.24 | 1.01 ± 0.25 | 0.098      | 0.98 ± 0.25 | 0.99 ± 0.24 | 0.304 |
| Non-HDL-C, mmol/L        | 4.02 ± 1.04 | 3.64 ± 1.08 | < 0.001   | 4.12 ± 1.02 | 3.78 ± 1.11 | < 0.001 |
| Lp(a), mg/dL             | 23.82 (12.42–44.10) | 16.87 (9.28–32.22) | < 0.001 | 23.76 (12.97–43.48) | 16.87 (9.30–31.72) | < 0.001 |
| BUN, mmol/L              | 5.8 ± 1.9    | 5.8 ± 2.0    | 0.730      | 6.0 ± 2.0    | 5.9 ± 2.0    | 0.558 |
| Serum creatinine, μmol/L | 88 ± 25     | 84 ± 29     | 0.007      | 90 ± 28     | 84 ± 30     | 0.003 |
| Uric acid, μmol/L        | 342 ± 85    | 340 ± 90    | 0.707      | 340 ± 87    | 330 ± 89    | 0.129 |
| GFR, ml/min/1.73 m²      | 75.4 ± 17.7 | 83.7 ± 19.4 | < 0.001   | 74.1 ± 18.6 | 83.2 ± 19.5 | < 0.001 |
| hsCRP, mmol/L            | 4.28 (1.61–7.68) | 2.72 (1.12–4.85) | < 0.001 | 4.52 (1.74–7.89) | 2.86 (1.32–5.07) | < 0.001 |
| LVEF, %                  | 57.4 ± 8.4   | 61 ± 8.2    | < 0.001   | 57.0 ± 8.4   | 60.6 ± 7.5   | < 0.001 |
| Medication, n (%)        |            |            |            |              |          |
| Antipsleot               | 374 (74.1)  | 552 (70.9)  | 0.212      | 236 (73.1)  | 274 (71.5)  | 0.652 |
| ACE inhibitors/ ARBs     | 312 (61.8)  | 476 (61.1)  | 0.807      | 194 (60.1)  | 235 (61.4)  | 0.725 |
| β-blockers               | 244 (48.3)  | 379 (48.7)  | 0.907      | 152 (47.1)  | 171 (44.6)  | 0.522 |
| CCBs                     | 152 (30.1)  | 208 (26.7)  | 0.185      | 101 (68.7)  | 107 (27.9)  | 0.333 |
| Diuretics                | 61 (12.1)   | 81 (10.4)   | 0.348      | 39 (12.1)   | 42 (11.0)   | 0.645 |
| Nitrates                 | 270 (53.5)  | 427 (54.6)  | 0.636      | 182 (56.3)  | 230 (60.1)  | 0.320 |
| Statins                  | 324 (64.2)  | 515 (66.1)  | 0.473      | 188 (58.2)  | 226 (59.0)  | 0.829 |

Data are mean ± SD or number (%)

ACE angiotensin converting enzyme, ARB angiotensin receptor blocker, BMI body mass index, BUN blood urea nitrogen, CAD coronary artery disease, CCB calcium channel blocker, DBP diastolic blood pressure, FBG fasting blood glucose, GFR estimated glomerular filtration rate, HbA1C glycated hemoglobin A1C, HDL-C high-density lipoprotein cholesterol, hsCRP high-sensitivity C reactive protein, LDL-C low-density lipoprotein cholesterol, Lp(a) lipoprotein a, LVEF left ventricular ejection fraction, MI myocardial infarction, SBP systolic blood pressure
### Table 2 Impact of lipid profile on poor collateralization in patients with and without diabetes

| Tertiles of lipid profile | Overall (n = 1284) | Diabetes (n = 706) | Non-diabetes (n = 578) |
|---------------------------|-------------------|-------------------|------------------------|
|                           | n                 | Poor/good Adjusted OR (95% CI) | P value | n | Poor/good Adjusted OR (95% CI) | P value | n | Poor/good Adjusted OR (95% CI) | P value |
| **Lp(a)**                 |                   |                   |           |   |                   |           |   |                   |           |
| Per tertile, mg/dL        |                   |                   |           |   |                   |           |   |                   |           |
| T1 < 12.66                | 426               | 1                 | 1.394 (1.192–1.629) | < 0.001* | 128/298 | 1.366 (1.108–1.684) | 0.003* | 1432/1191–1.831 | 0.004* |
| 12.66 ≤ T2 < 20.33        | 430               | 1                 | 1.170 (1.081–1.591) | 0.316 | 117/134 | 1.282 (1.054–1.926) | 0.231 | 59/139 | 1.040 (0.640–1.688) | 0.875 |
| T3 ≥ 20.23                | 409               | 1                 | 1.929 (1.413–2.633) | < 0.001 | 126/102 | 1.863 (1.227–2.830) | 0.004 | 75/106 | 2.010 (1.236–3.269) | 0.005 |
| **Total cholesterol**     |                   |                   |           |   |                   |           |   |                   |           |
| Per tertile, mmol/L       | 1.791 (1.475–2.174) | < 0.001* | 1.814 (1.393–2.361) | < 0.001* | 1.820 (1.358–2.440) | < 0.001* | |
| T1 < 4.26                 | 420               | 1                 | 66/129 | 1 | – | – | 57/168 | 1 | – |
| 4.26 ≤ T2 < 5.35          | 441               | 1                 | 109/129 | 1 | 1.993 (1.251–3.173) | 0.004 | 61/142 | 1.854–2.158 | 0.196 |
| T3 ≥ 5.35                 | 423               | 1                 | 148/125 | 1 | 3.337 (1.962–5.674) | < 0.001 | 64/86 | 3.663 (2.010–6.675) | < 0.001 |
| **LDL-C**                 |                   |                   |           |   |                   |           |   |                   |           |
| Per tertile, mmol/L       | 1.748 (1.441–2.122) | < 0.001* | 1.830 (1.407–2.381) | < 0.001* | 1.699 (1.270–2.274) | < 0.001* | |
| T1 < 2.58                 | 401               | 1                 | 61/138 | 1 | – | – | 54/148 | 1 | – |
| 2.58 ≤ T2 < 3.36          | 431               | 1                 | 116/115 | 1 | 2.426 (1.518–3.879) | < 0.001 | 55/145 | 1.052 (0.648–1.709) | 0.837 |
| T3 ≥ 3.36                 | 452               | 1                 | 146/130 | 1 | 3.509 (2.064–5.964) | < 0.001 | 73/103 | 3.092 (1.724–5.548) | < 0.001 |
| **Non-HDL-C**             |                   |                   |           |   |                   |           |   |                   |           |
| Per tertile, mmol/L       | 1.828 (1.498–2.230) | < 0.001* | 1.810 (1.386–2.364) | < 0.001* | 1.912 (1.407–2.597) | < 0.001* | |
| T1 < 3.30                 | 436               | 1                 | 71/131 | 1 | – | – | 60/174 | 1 | – |
| 3.30 ≤ T2 < 4.38          | 441               | 1                 | 104/133 | 1 | 1.683 (1.073–2.639) | 0.023 | 63/141 | 1.407 (0.891–2.222) | 0.143 |
| T3 ≥ 4.38                 | 407               | 1                 | 148/119 | 1 | 3.263 (1.911–5.571) | < 0.001 | 59/81 | 4.260 (2.245–8.086) | < 0.001 |
| **HDL-C**                 |                   |                   |           |   |                   |           |   |                   |           |
| Per tertile, mmol/L       | 0.907 (0.775–1.060) | 0.220 | 0.846 (0.686–1.045) | 0.121* | 0.994 (0.780–1.267) | 0.960* | |
| T1 < 0.86                 | 388               | 1                 | 117/112 | 1 | – | – | 55/104 | 1 | – |
| 0.86 ≤ T2 < 1.04          | 437               | 1                 | 101/133 | 1 | 0.808 (0.540–1.210) | 0.302 | 60/143 | 0.834 (0.514–1.354) | 0.463 |
| T3 ≥ 1.04                 | 459               | 1                 | 105/138 | 1 | 0.717 (0.471–1.093) | 0.122 | 67/149 | 0.975 (0.602–1.579) | 0.918 |
| **Triglyceride**          |                   |                   |           |   |                   |           |   |                   |           |
| Per tertile, mmol/L       | 1.075 (0.391–1.266) | 0.383* | 1.092 (0.880–1.352) | 0.428* | 1.075 (0.516–1.435) | 0.428* | |
| T1 < 1.28                 | 436               | 1                 | 96/122 | 1 | – | – | 62/156 | 1 | – |
| 1.28 ≤ T2 < 1.84          | 448               | 1                 | 99/135 | 1 | 0.947 (0.625–1.434) | 0.796 | 76/138 | 1.311 (0.840–2.046) | 0.232 |
| T3 ≥ 1.84                 | 400               | 1                 | 128/126 | 1 | 1.183 (0.771–1.817) | 0.442 | 44/102 | 1.072 (0.633–1.814) | 0.796 |

CI: confidence interval, HDL-C: High-density lipoprotein cholesterol, LDL-C: Low-density lipoprotein cholesterol, Lp(a): Lipoprotein a, OR: Odds ratio

* P for trend for tertiles of lipid profile

a Multiple adjustment for gender, age, body mass index, hypertension, diabetes, dyslipidemia, smoking, prior myocardial infarction, multi-vessel disease, glomerular filtration rate, log-transferred high-sensitivity C reactive protein and left ventricular ejection fraction.
Fig. 2  Percentage of poor coronary collateralization in relation to interaction between Lp(a) and total cholesterol, LDL-C, non-HDL-C or HDL-C in diabetics (a–d) and non-diabetics (e–h). Values are percentage of poor collateralization according to tertile distribution of Lp(a) (blue, green and red line for tertile 1, 2 and 3, respectively). P values for poor collateralization for each tertile of total cholesterol, LDL-C, non-HDL-C and HDL-C are given.
collateralization (adjusted P interaction = 0.857 and 0.941, respectively).

For non-diabetic patients, no such interactions between Lp(a) and cholesterol-containing lipids and triglyceride were observed (Fig. 2).

**Discussion**

The results of this large cohort study support the hypothesis that in patients with stable coronary artery disease and chronic total occlusion, increased Lp(a) confers greater risk for poor coronary collateralization when total cholesterol, LDL-C or non-HDL-C are elevated especially in patients with type 2 diabetes.

**Relation between Lp(a) and coronary collateralization in diabetes**

It is suggested that there is an inverse association between Lp(a) concentration and risk of type 2 diabetes, with a higher risk for type 2 diabetes at low Lp(a) concentrations—approximately < 7 mg/dL [31]. Concerning type 1 diabetes, no different levels of Lp(a) were found between patients with any degree of coronary stenosis and those without coronary disease [32]. The present finding on an inverse association between elevated Lp(a) levels and poor coronary collateral formation is consistent with previous reports examining the same phenomenon [14, 15]. Nevertheless, our study population was unique as all patients had stable coronary artery disease and chronic total occlusion. This study is also the first to investigate the relation of Lp(a) and its interactions with a broad spectrum of cholesterol-containing lipids including total cholesterol, LDL-C, HDL-C, and non-HDL-C on coronary collateralization in a large number of patients with type 2 diabetes. Our results showed that the prevalence of elevated Lp(a) levels was higher in patients with poor coronary collateralization as compared to those with good collateralization, and notably, tertiles of Lp(a) remained an independent determinant for poor collateralization even after adjustment for various confounding factors including cholesterol-containing lipid profiles. These observations support a notion that Lp(a) could be a biomarker of coronary collateral circulation in diabetic patients with stable coronary artery disease and chronic total occlusion. Although its physiological function is still not completely elucidated, Lp(a) is known to be highly concentrated in the arterial wall, carries cholesterol and binds atherosclerogenic oxidized phospholipids, which attracts inflammatory cells to vessel walls and leads to smooth muscle cell proliferation, consequently, contributing to the process of atherosclerosis [16, 19]. Several studies have shown that high levels of Lp(a) adversely affect collateral vessel growth by inducing endothelial cell dysfunction through various mechanisms [33]. Aras et al found a strong negative correlation between serum Lp(a) levels and vascular endothelial growth factor concentration in patients with chronic total coronary occlusion [14]. High levels of Lp(a) attenuate synthesis and/or release of vascular endothelial growth factor and decrease production of endothelium-derived nitric oxide, leading to impaired angiogenesis [15]. Using a Lp(a) transgenic mouse hindlimb ischemia model, Morishita et al revealed that a high serum Lp(a) concentration abolished collateral formation by inhibition of transforming growth factor-β activity, suggesting that Lp(a) might also decrease arteriogenesis [34].

**Lp(a) interactions with LDL-C and non-HDL-C on coronary collateralization in diabetes**

The major finding of this study is that the adverse effect of Lp(a) on coronary collateral development was aggravated by the presence of a high level of LDL-C or non-HDL-C in patients with diabetes. Our results showed that Lp(a) was significantly correlated with LDL-C or non-HDL-C, and individuals with high Lp(a) were more likely to have LDL-C > 3.36 mmol/L or non-HDL-C > 4.38 mmol/L, confirming the physiological link between Lp(a) and LDL-C or non-HDL-C. Furthermore, there was a synergistic effect of Lp(a) and LDL-C or non-HDL-C on collateral formation in patients with diabetes. In high tertile of LDL-C or non-HDL-C, diabetic patients with high tertile of Lp(a) had an approximately fourfold increased risk of poor coronary collateralization compared with those with low tertile of Lp(a). These observations suggest that Lp(a) might exert a more pronounced detrimental effect on coronary collateral formation in a high-risk lipid profile environment.

Although certain studies have not shown any significant difference in the prevalence of elevated LDL-C in patients with diabetes compared with non-diabetic counterparts [35], total LDL-C may be a misleading measurement in diabetes. This may be in part due to a significant shift to small dense LDL-C in diabetic dyslipidemia, which is associated with greater cardiovascular disease risk. In fact, the same level of LDL-C can be associated with greater serum levels of apo B and hence, more low-density lipoprotein particles. Whether the adverse effects of Lp(a) interactions with LDL-C on coronary collateralization in patients with diabetes is due to these changes remains unknown and requires further investigations.

Non-HDL-C encompasses all of the atherogenic apoB-containing lipoproteins (LDL-C, very low-density lipoprotein cholesterol, intermediate-density lipoprotein cholesterol) [18]. Achieved non-HDL-C levels reflect the full burden of cholesterol transported in atherogenic lipoproteins, and seem to be more closely associated with coronary atheroma progression than LDL-C even
among statin-treated individuals [36]. Recently, both the International Atherosclerosis Society and National Lipid Association have flagged non-HDL-C as the major form of atherogenic cholesterol and recommended using this parameter as the marker for the primary and secondary prevention of atherosclerotic cardiovascular disease [20, 37, 38]. The mechanism of elevated non-HDL-C on poor coronary collateralization is likely to be complex. Lipid profile impairment, especially hypercholesterolemia and high levels of LDL-C and non-HDL, is an established risk factor that induces endothelial cell dysfunction and impairs coronary collateral vessel growth [18]. Alvim et al found that hypercholesterolemia and high non-HDL-C levels were associated with increased arterial stiffness characterized by elevated systolic and pulse blood pressures and reduced diastolic blood pressure [39]. Baykan et al observed that increased arterial stiffness could cause decreased coronary perfusion, reduces shear stress, arteriogenesis, and thus collateral formation [40]. Our data were partially corroborated by the results of several population studies demonstrating that non-HDL-C is a better marker of cardiovascular disease risk than LDL-C alone [36, 41].

Relation between HDL-C and coronary collateralization in diabetes
It has been well recognized that HDL-C has anti-atherogenic, anti-inflammatory, and anti-oxidant properties [42]. Sumi et al. observed that reconstituted HDL-C directly stimulates endothelial progenitor cell differentiation via phosphatidylinositol 3-kinase/Akt pathway and enhances ischemia-induced angiogenesis [43]. The relation between serum HDL-C and coronary collateral formation remains controversial. Kadi et al found that in patients with stable coronary artery disease there was a positive relationship between HDL-C and angiographic collateral score, and low HDL-C is an independent determinant of poor coronary collateralization [44]. Recently, Hsu et al reported that the extent of coronary artery disease severity but not HDL-C level was the most powerful predictor of coronary collateral formation in 501 Chinese patients with stable coronary artery disease [45]. In the present study, we did not find an association between serum HDL-C level and coronary collateral score. This finding is in line with our previous observations that HDL-C functionality rather than quantity alone may reflect its overall effect [46, 47]. However, the detailed mechanism still needs further investigation.

Clinical implications
The present study demonstrates the robust associations between Lp(a) interactions with cholesterol-containing lipids and coronary collateral formation in patients with stable coronary artery disease and chronic total occlusion, which were not linear and limited to highest Lp(a) and LDL-C or non-HDL-C tertile. These observations emphasize the potential importance of LDL-C and non-HDL-C in pointing to patients at increased risk for Lp(a)-mediated disease, as well as preventative strategies to mitigate the risk conferred by elevated Lp(a) levels (e.g., LDL-C or non-HDL-C lowering) especially for patients with diabetes. First, our study substantiates the concept that LDL-C or non-HDL reduction with statin therapy remains the mainstay of pharmacotherapy for dyslipidemia and the percentage reduction in LDL-C lowering is strongly correlated with reduction in atherosclerotic cardiovascular disease risk and events [18–20]. Recent trials using cholesterol absorption blocker ezetimibe in combination with statins [48] and proprotein convertase subtilisin/kexin type 9 (PCSK-9) inhibitors [49] clearly show that greater LDL-C reduction on top of statin therapy provides added benefits, without attenuation in cardiovascular disease benefit in patients starting treatment with lower LDL-C. In addition, dipeptidyl peptidase-4 (DPP4) inhibitor anagliptin has been shown to have inhibitory effects on hepatic cholesterol synthesis and reduce LDL-C by 9.5 mg/dL over 12 weeks, regardless of the use of statins [50]. Second, our study highlights the need for Lp(a) lowering therapy [51–53]. Ezetimibe treatment either alone or in combination with a statin does not affect serum Lp(a) concentration [54] and fibrates have a significantly greater effect in reducing serum Lp(a) than statins. Addition of fibrates to statins can enhance the Lp(a)-lowering effect of statins [55]. Nicotinic acid has been shown to decrease Lp(a) levels by 15–30% and reduce cardiovascular events [56]. However, even slower release niacin is usually poorly tolerated because of gastrointestinal side effects, making it less popular with patients [57]. PCSK-9 inhibitors are promising as these agents could lower Lp(a) with various degree among patients [49, 51, 56]. Likewise, apo(a) antisense oligonucleotides is a new treatment option for lowering elevated Lp(a). Clinical trials of Lp(a) reduction using antisense oligonucleotides that lower apo(a) levels by 90% are underway. For some patients with diabetes and increased Lp(a), such therapies may enter the clinic soon [58, 59]. The long-term efficacy of lipoprotein apheresis on morbidity in patients with elevated Lp(a) and chronic ischemic heart disease on maximally tolerated lipid-lowering therapy is well documented [60].

Study limitations
We recognize that there are several limitations in our study. First, the study is cross-sectional for the point of coronary collateral investigation, thereby allowing us
to detect association, not to formulate causal link. In addition, the correlation of Lp(a) with total cholesterol and LDL-C was statistically significant but very weak. It remains uncertain whether we will be able to use the recent data for any recommendations, as the Lp(a) tertiles are completely not specific. Second, the presence and degree of collaterals were evaluated according to the Rentrop scoring system. Although this angiographic assessment of coronary collaterals is easily to be incorporated into the routine clinical practice, coronary collaterals may be more accurately assessed by collateral flow index with simultaneous measurement of aortic pressure and the distal pressure within the occluded segment of the culprit coronary artery [61].

Conclusions
In patients with stable coronary artery disease and chronic total occlusion, poor coronary collateralization seems tightly linked to Lp(a) interactions with total cholesterol, LDL-C, and non-HDL-C. Individuals with high level of Lp(a) (> 30.23 mg/dL) and total cholesterol (> 5.35 mmol/L), LDL-C (> 3.36 mmol/L) or non-HDL-C (> 4.38 mmol/L) may warrant aggressive lipid lowering therapy especially for those with type 2 diabetes.

Additional files

**Additional file 1:** Table S1. Logistic regression analyses for poor collateralization in patients with and without diabetes.

**Additional file 2:** Table S2. Impact of lipid profile on poor collateralization in patients with and without diabetes.

**Abbreviations**
apo: apolipoprotein; BMI: body mass index; CKD-EPI: chronic kidney disease epidemiology collaboration; GFR: glomerular filtration rate; HbA1c: glycated hemoglobin; HDL-C: high-density lipoprotein cholesterol; hsCRP: high-sensitivity C-reactive protein; LDL-C: low-density lipoprotein cholesterol; Lp(a): lipoprotein (a); PCSK9: proprotein convertase subtilisin/kexin type 9; SD: standard deviation.

**Acknowledgements**
Not applicable.

**Authors’ contributions**
YS, FHD, WFS wrote the article, substantially contributed to discussion of the content, and edited the manuscript. YS, SC, YD, XQW, ZKY, JH performed the experiments and researched data for the article. FHD analyze the data; LL, RYZ substantially contributed to discussion of the content and reviewed the manuscript. All authors read and approved the final manuscript.

**Funding**
This study was supported in part by the Research Foundation of Chinese National Natural Science (81670451, 81770437, 81770447), Shanghai Science & Technology Committee (14ZK11009), Medico-engineering Project (GY20160656), Talent Young Investigators (17XJ11009) of Shanghai Jiao Tong University School of Medicine and Shanghai Municipal Education Commission-Gaofeng Clinical Medicine Grant Support (20181801).

**Availability of data and materials**
Data generated or analyzed during this study are included in this published article.

**Ethics approval and consent to participate**
The study protocol was approved by the Institutional Review Board of Rui Jin Hospital Shanghai Jiaotong University School of Medicine. Written informed consent was obtained from all patients, and clinical investigation was conducted according to the principle of the Declaration of Helsinki.

**Consent for publication**
All authors consent this manuscript for publication.

**Competing interests**
The authors declare that they have no competing interests.

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Received: 21 April 2019   Accepted: 16 June 2019

**Published online:** 24 June 2019

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