Seed-Mediated Preparation of Ag@Au Nanoparticles for Highly Sensitive Surface-Enhanced Raman Detection of Fentanyl

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Abstract: Bimetallic nanoparticles have received extensive attention due to their unique physical and chemical properties, including enhanced optical properties, chemical stability, and better catalytic activity. In this article, we have successfully achieved the controllable preparation of Ag@Au nanoparticles via a seed-mediated growth method. By regulating the amount of seeds—silver nanospheres—we realized that Ag@Au nanoparticles gradually changed from spherical to a sea-urchin-like structure. The structure and composition of the prepared nanoparticles were characterized via scanning electron microscopy (SEM), transmission electron microscopy (TEM), energy-dispersive X-ray spectroscopy (EDS), and high-angle circular dark field scanning transmission electron microscopy (HAADF-STEM). In addition, we use the prepared Ag@Au nanoparticles as a substrate material for highly sensitive surface-enhanced Raman spectroscopy (SERS). Using 4-aminothiophenol (4-ATP) as the test molecule, we explored the SERS enhancement effects of Ag@Au nanoparticles with different structures. Furthermore, we used Ag@Au nanoparticles for SERS to detect the drug fentanyl, and realized the label-free detection of fentanyl, with the lowest detection concentration reaching $10^{-7}$ M. This research not only provides a method for preparing bimetallic Ag@Au nanoparticles with different structures, but also provides a reference for the application of Ag@Au nanoparticles in the field of detection technology.

Keywords: Ag@Au nanoparticle; core–shell structure; sea-urchin-like structure; SERS; fentanyl

1. Introduction

Noble metal nanocrystals have received extensive attention due to their unique localized surface plasmon resonance characteristics. A large number of studies have reported the application of noble metal nanocrystals in the fields of sensing, catalysis, and detection [1–5]. The characteristics of metal nanomaterials depend largely on their size, structure, and composition. In the past few decades, much research has been devoted to the controllable preparation of noble metal nanocrystals with uniform morphology and size. Various preparation methods have been developed one after another, including the solvothermal method [6–8], electrochemical method [9,10], and photochemical method [11]. Many metal nanocrystals with various regular morphologies have also been prepared. For example, Mirkin et al. realized the transformation of silver nanocrystals from spherical to icosahedral [12]. Yan et al. successfully prepared branched gold nanocrystals with tunable local surface plasmon resonance (LSPR) characteristics, and used the prepared gold nanocrystals as a substrate to achieve quantitative detection of the heme concentration in the cytosol of human red blood cells [13]. By controlling the kinetics involved in the growth of preformed Rh cube seeds, Xia et al. achieved the synthesis of Rh nanocrystals, including cubes, cubic octahedrons, and octahedrons [14]. In addition to polyhedral nanocrystals with regular morphologies, researchers have found that nanomaterials with sharp protrusions and pores often exhibit specific properties. For example, core–satellite structures and sea-urchin-like...
structures with sharp protrusions promote electromagnetic field enhancement by generating multiple “hot spots”. This phenomenon has also aroused the interest of a large number of researchers.

In addition, researchers have found that alloy nanomaterials with different elemental compositions have the advantages of various nanomaterials, and show better performance than single-element nanomaterials in various fields. The preparation and performance of bimetallic nanoparticles is now an exciting research area, because they provide a new way to change the properties of the particles by mixing two metals in one particle. Especially for two kinds of metal nanoparticles with different advantages, we can realize the combination of their advantages through the controllable preparation of bimetallic nanoparticles, and avoid their respective disadvantages. For example, for two metal nanomaterials—gold and silver—studies have shown that silver nanomaterials have a larger excitation window—from blue to near-infrared—than gold nanoparticles (from red to near-infrared) [15]. In addition, it can be said that silver is a more effective optical material than gold. Due to its greater scattering contribution, which is related to the real part of its dielectric constant, the surface-enhanced Raman scattering (SERS) signal generated by silver is more than 100 times higher than that of similar gold nanostructures [16]. However, the size and shape of gold nanoparticles are easier to control, and have good chemical resistance—that is, higher biocompatibility [17]. Therefore, silver nanomaterials are used more in in vitro research, while gold nanoparticles are used more in in vivo research [18,19].

In order to make full use of the advantages of gold and silver nanoparticles, and avoid their disadvantages, we urgently need to develop an Ag@Au nanomaterial that can coat the surface of silver nanoparticles with a gold layer, or form an alloy of gold and silver on the surface of silver nanoparticles. In general, alloy nanoparticles are obtained via the simultaneous reduction of two metal precursors (Ag⁺ and Au³⁺). Conversely, the controlled growth of the metal on the preformed colloidal surface produces good results if the inner metal (the metal forming the colloid) has a greater surface free energy than the metal on its surface. However, because the surface free energy of gold is greater than that of silver (1.128 and 0.923 Jm⁻², respectively), it is easier to grow silver on the surface of gold nanoparticles. In fact, a number of studies have also reported that Au@Ag nanoparticles were obtained by growing silver layers on gold seeds [20,21]. For example, Xia et al. recently achieved regulation of the plasma performance of Au@Ag nanoparticles by adjusting the thickness of the silver layer grown on the surface of gold nanospheres and gold nanorods [22]. Cheng et al. adjusted the concentration of the surfactant cetyltrimethylammonium chloride (CTAC) to precisely control the position of the Ag coating on the convex gold nanoarrows. Three different nanostructures were obtained at low, medium, and high CTAC concentrations—namely, anisotropic coating, intermediate coating, and conformal coating, respectively [23]. Conversely, depositing gold onto silver nanoparticles by reducing gold precursors yields a solid solution of the two metals [24]. In general, the gold precursor will undergo a galvanic replacement reaction with the silver nanoparticles, so that the internal silver atoms are first replaced by silver ions, and then the silver ions are reduced to silver atoms by the reducing agent in the solution and deposited on the surface, so that the hollow structure is finally obtained [25–28]. As far as we know, only a few studies have reported the method of growing a gold layer on the surface of silver nanoparticles to prepare Ag@Au core–shell-structured nanoparticles. For example, Xia et al. successfully prepared Ag@Au nuclear sheath nanowires by depositing Au atoms on the surface of pre-synthesized Ag nanowires, which greatly improved their stability under different corrosive environmental conditions [29]. Kim et al. used sodium sulfite to selectively bind Au cations, and thereby reduce the reduction potential of Au (E° = 0.111 V). The gold(I) sulfite complex is relatively benign to the Ag nanowire surface, so no oxidative etching will occur; on the contrary, the Au coating is promoted, and finally Ag@Au nanowires are formed [30]. Kim et al. achieved gold-spiked coating of silver particles through controlled Ostwald ripening of small gold nanoparticles on the surface of larger silver particles [31].
In this article, in order to fully combine the advantages of gold and silver nanoparticles and avoid their disadvantages, we have developed a method for preparing Ag@Au nanoparticles. Ag@Au nanoparticles have both the excellent surface plasmon resonance characteristics of silver nanoparticles and the stability of gold. First, Au\textsuperscript{3+} is combined with Br\textsuperscript{−} in CTAB to form [AuBr\textsubscript{4}]\textsuperscript{−}, and then the addition of ascorbic acid reduces Au\textsuperscript{3+} to Au\textsuperscript{+}. Finally, an Au(I)–GSH complex is formed by combining Au\textsuperscript{+} with glutathione modified on the surface of silver nanoparticles, and then Au\textsuperscript{+} is reduced by glutathione in situ to grow gold on the surface of silver nanoparticles. By simply adjusting the amount of silver seeds added, we realized the transformation of the Ag@Au nanoparticle structure from spherical to a sea-urchin-like structure.

2. Materials and Methods

2.1. Materials

The hydrochloroauric acid trihydrate (HAuCl\textsubscript{4} 3H\textsubscript{2}O, 99.9% trace metals basis), hexadecyltrimethylammonium bromide (CTAB, AR, 99%), glutathione (Mw = 307.32, ≥98%), 4-aminothiophenol (4-ATP, GC, ≥98%), nitric acid (HNO\textsubscript{3}, AR), and hydrochloric acid (HCl, AR) were purchased from Aladdin. Silver nanospheres (diameter: 80 nm; purity: >99% concentration: 0.1 mg/mL, XFJ63) were purchased from Nanjing/Jiangsu XFNANO Materials Tec Co., Let. Fentanyl hydrochloride standard product was purchased from the Shanghai Institute of Criminal Science and Technology (Shanghai, China). All reagents were used directly, without further processing. All of the water used in the experiment was ultrapure water (18.2 MΩ·cm) purified using a Milli-Q Lab System (Nihon Millipore Ltd., Hangzhou, China). The flasks and glass slides used in the experiment were first washed with aqua regia (HCl:HNO\textsubscript{3} = 3:1) for 30 min, then washed with water (18.2 MΩ·cm) and absolute ethanol twice, and finally dried for use.

2.2. Synthesis of Ag@Au Nanoparticles

First, we mixed the silver nanoball seeds with 100 µL of 1 mM glutathione solution in a 1.5 mL centrifuge tube at room temperature; using an IKA shaker, we then vortexed it for 1 min, and incubated it for 1 h to make glutathione modified into silver nanospheres by the Ag–S bond. At the same time, we added 4.0 mL ultrapure water to a 20 mL glass flask, followed by 200 µL of 10 mM HAuCl\textsubscript{4} 3H\textsubscript{2}O solution and 0.8 mL of 0.1 M CTAB solution. The mixed solution was stirred at 500 rpm for 30 min at room temperature using an IKA magnetic stirring device, mixed well, and then 475 µL of 0.1 M ascorbic acid solution was added, stirring was continued at room temperature for 5 min, and the mixed solution was used as a growth solution. Finally, the silver nanospheres modified with glutathione were added to the growth solution, and the reaction was carried out at room temperature under magnetic stirring for 2 h. The resulting product was centrifuged at 5000 rpm for 10 min; the supernatant was removed, and washed twice with water and ethanol, respectively. In order to explore the effect of the amount of silver nanoball seeds added on the produced Ag@Au NPs, we added 40 µL, 80 µL, 160 µL, and 500 µL of silver nanosphere seeds to the reaction, without changing any of the other reaction conditions.

2.3. Characterization and SERS Test

The morphology and structure of the prepared nanoparticles were passed through a field emission scanning electron microscope (SEM, 3.0 kV, JEOL, JSM-6700F, Japan), transmission electron microscope (TEM, 100.0 kV, JEOL-2100F, Japan), high-resolution TEM (HRTEM, 200 kV, JEOL-2100F, Japan), and selected-area electron diffraction (SAED, JEOL-2100F, Japan) for characterization. The SERS test was conducted using a Raman spectrometer (Thermo Fisher DXR2xi, America) under 633 nm laser excitation; the laser power was 6 mW, and the acquisition time was 10 s. For surface-enhanced Raman spectroscopy testing, we first added 10 µL of Ag@Au nanoparticles to the centrifuge tube, and then added 50 µL of 4-ATP solutions of different concentrations, mixing well. The mixed solution was stored at room temperature for 30 min, and then 5 µL was dropped on a
glass slide, dried at room temperature, and used for surface-enhanced Raman spectroscopy (SERS) testing. The SERS test used a laser Raman spectrometer (Thermo Fisher DXR2xi, America) equipped with a microscope (50× objective lens) and a CCD detector; the laser source was 633 nm, the acquisition time was 10 s, and the laser power was 6 mW. We used the supporting software to analyze the Raman peak spectrum. The SERS test for the drug fentanyl followed the same process.

3. Results and Discussion

Characterization of Ag@Au NPs

We studied the morphology and size of Ag@Au nanoparticles via SEM and TEM. Figure 1a is the SEM image of Ag@Au nanoparticles prepared when the amount of silver seed added is 40 µL, and Figure 1b is the corresponding TEM image. As we can see, the surface of the prepared Ag@Au nanoparticles is distributed with dense, thorn-like structures. From the HRTEM image (Figure 1c) of the gold thorns grown on the surface, it can be seen that the lattice spacing is 0.24 nm, indicating that the exposed crystal faces of the gold thorns are mainly composed of {111} planes [4]. Figure 1d shows the corresponding HAADF map. We performed EDS characterization of the prepared Ag@Au nanoparticles, as shown in Figure 1e–g. Figure 1e,f are the distribution diagrams of silver and gold, respectively. Figure 1g shows the overlapping distribution of the two elements. From the EDS results, it can be seen that the atomic ratio of gold to silver is 68:32.

Figure 1. (a) SEM, (b) TEM, (c) HRTEM, and (d) HAADF images of as-prepared Ag@Au NPs, when the amount of Ag seed was 40 µL. (e–g) The corresponding EDS images.

Figure 2 shows the characterization results of Ag@Au nanoparticles obtained when the amount of silver seed added is 500 µL. Figure 2a,b show the SEM and TEM images of Ag@Au nanoparticles, respectively. From the figure, we can clearly see that the Ag@Au nanoparticles obtained still maintain a spherical structure, and have a uniform size distribution of about 95 nm. From its HRTEM image (Figure 2c), we can see that the lattice spacing of gold nanoparticles distributed on the surface is 0.24 nm, indicating that it is mainly composed of {111} planes. In addition, from the HAADF map (Figure 2d), it can be seen that only a small part of the Ag@Au nanoparticles are gray in the central area, indicating that the Ag@Au nanoparticles have formed a partially hollow structure, while the main part is still a solid structure. Furthermore, we collected the distribution of Au and Ag elements in Ag@Au nanoparticles via EDX, as shown in Figure 2e–g. From the figure, we can see that gold and silver form an alloy structure distributed over the entire surface.
Furthermore, we explored the effects of the amount of silver seeds on the Ag@Au nanoparticles. As shown in Figure 3, when there was no silver seed, we prepared a mesoporous gold structure (Figure 3a). When adding 40 µL of silver seeds, we prepared Ag@Au nanoparticles with dense, thorn-like structures on their surface (Figure 3b). When the amount of silver seeds was doubled (80 µL), the thorn-like structures on the surface of the Ag@Au nanoparticles were reduced (Figure 3c). When the amount of silver seeds was further increased to 160 µL, the thorn-like structures on the surface were further reduced (Figure 3d). When the amount of silver seed was increased to 500 µL, we achieved a spherical structure with shape retention (Figure 3e). According to the EDS characterization results of the nanoparticles prepared under different concentrations of silver seeds, we performed a statistical analysis of the proportions of gold and silver atoms, as shown in Figure 3f. The EDS diagrams of Ag@Au nanoparticles prepared under different silver seed conditions are shown in Figure A1. Analyzing the EDS results, when the amount of silver seeds added was 40, 80, 160, and 500 µL, the proportion of tightness in the prepared Ag@Au nanoparticles was 0.68, 0.44, 0.38, and 0.20, respectively. The corresponding proportions of silver were 0.32, 0.56, 0.62, and 0.80, respectively.

Based on the above experimental results, we can make the following inferences on the formation mechanism of Ag@Au nanoparticles: Figure 3g is a schematic diagram of the growth of Ag@Au nanoparticles. First, Au^{3+} ions in HAuCl₄·3H₂O solution combine with Br⁻ ions in CTAB to form [AuBr₄]⁻, and then ascorbic acid reduces [AuBr₄]⁻ to get Au⁺. This process is consistent with previous reports [32,33]. When silver seeds modified with glutathione were added, Au⁺ reduced glutathione adsorbed on the surface of silver seeds in situ to form Au₀. It can be seen from Figure 2D that Au⁺ inevitably had a galvanic replacement reaction with silver seeds (forming few hollow structures). However, the majority of Ag@Au nanoparticles are still solid, which also indicates that the glutathione on the surface of the silver seeds largely inhibits the galvanic replacement reaction. In addition, the reduction of glutathione also causes the gold ions to be reduced to gold atoms and deposited on the surface of the silver seeds, which also prevents the occurrence of the galvanic replacement reaction. Thus, when the amount of silver seeds is low, there are enough gold atoms to grow along the direction of glutathione, and eventually form sea-urchin-like Ag@Au nanoparticles. When the amount of silver seeds is greater, the Ag@Au spherical shell structure with the original shape will be formed. The above results show that we have successfully prepared Ag@Au nanoparticles. Compared with the previously reported Au@Ag core–shell structure and the Ag@Au hollow structure prepared by the
displacement reaction, the Ag@Au nanoparticles we prepared have a solid structure. A gold thorn structure grows on the surface of the seed, which realizes the combination of the advantages of gold and silver nanoparticles.

Figure 3. TEM images of as-prepared Ag@Au nanoparticles, when the amount of Ag seed added was (a) 0 µL, (b) 40 µL, (c) 80 µL, (d) 160 µL, and (e) 500 µL. (f) A statistical graph of the atomic ratio of Ag and Au in Ag@Au nanoparticles prepared with different amounts of silver seeds, according to the EDS results. (g) Schematic diagram of the growth process of Ag@Au nanoparticles. Scale bar: 100 nm.

Furthermore, we used the prepared Ag@Au nanoparticles as the substrate material for surface-enhanced Raman spectroscopy (SERS) for highly sensitive detection of the drug fentanyl. First, we explored the SERS enhancement effect of Ag@Au nanoparticles with different structures. Figure 4 shows the SERS spectra obtained with $10^{-4}$ M $p$-aminothiophenol (4-ATP) as the test molecule and Ag@Au nanoparticles with different structures as the base material. We can see that the order of SERS enhancement effects, from high to low, is spherical Ag@Au nanoparticles, sea-urchin-shaped Ag@Au nanoparticles, and mesoporous gold nanoparticles. Therefore, in the follow-up test, we use spherical Ag@Au nanoparticles as the enhancement reagent. The Raman characteristic peaks of 4-ATP molecules are located at 1076 cm$^{-1}$ and 1585 cm$^{-1}$ [34], and we see obvious SERS peaks at 1140 cm$^{-1}$, 1389 cm$^{-1}$, and 1432 cm$^{-1}$, which can be attributed to $\beta$CH + νCN, νNN + νCN, and νNN + $\beta$CH of $p,p'$-dimercaptoazobenzene (DMAB), respectively [4]. This indicates that the prepared nanomaterials have catalytic activity, so that 4-ATP is partially converted to DMAB by catalytic oxidation.
We used spherical Ag@Au nanoparticles to detect fentanyl. Figure 5a shows the Raman spectrum and SERS spectrum of the fentanyl standard substance. As shown in Figure 5a (black line), the Raman peaks of fentanyl solid powder are located at 621 cm$^{-1}$, 746 cm$^{-1}$, 831 cm$^{-1}$, 1002 cm$^{-1}$, 1030 cm$^{-1}$, 1201 cm$^{-1}$, 1447 cm$^{-1}$, and 1600 cm$^{-1}$, which is consistent with previous reports [35]. However, the peak at 621 cm$^{-1}$ in the SERS spectrum (red line) disappeared. This is because, for a group, the Raman peak of tensile vibration usually appears in a higher frequency range, and is less affected by the external environment. The Raman peak of deformation vibration is usually located in a lower frequency range, and is sensitive to environmental changes [36]. The peak at 621 cm$^{-1}$ is the bending vibration of C–C–C [35], which is easily affected by the SERS detection environment, and disappears. We assigned the SERS peak of fentanyl, as shown in Table A1. Furthermore we performed SERS tests on different concentrations of fentanyl, and the results are shown in Figure 5b. We can see that the lowest detection concentration is $10^{-7}$ M.

4. Conclusions

In this paper, we prepared Ag@Au nanoparticles via a seed-mediated growth method. By adjusting the amount of silver seeds, we realized the transformation of Ag@Au nanopar-
articles from a spherical to a sea-urchin-like structure, and proposed the possible growth mechanism. We further explored the SERS enhancement performance of the prepared nanoparticles, using 4-ATP as the test molecule. The results show that the order of SERS enhancement effects, from high to low, is spherical Ag@Au nanoparticles, sea-urchin-shaped Ag@Au nanoparticles, and mesoporous gold nanoparticles. Finally, with Ag@Au spherical nanoparticles as the enhancement material, we achieved the highly sensitive SERS detection of fentanyl, with the lowest detection concentration reaching $10^{-7}$.

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**Appendix A**

![Figure A1. TEM, HAADF, and EDS characterization of the prepared nanoparticles with different amounts of Ag seeds: (a1–a5) 40 µL, (b1–b5) 80 µL, (c1–c5) 160 µL, and (d1–d5) 500 µL.](image-url)
Table A1. Characteristic vibrations of fentanyl.

| Fentanyl | Δ(C–C–C), Δ(N–CH₂) |
|----------|---------------------|
| 746      | ν(N–C–C–C)          |
| 831      |                    |
| 1002     | Δ(CH) Ar           |
| 1030     | ν(C=C), Δ(CH₂) twisting in aliphatic ring |
| 1201     | Δ(C=C) benzyl stretch |
| 1447     | Δ(CH₂) scissoring   |
| 1600     | ν(C=C) Ar          |

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