Associations of transcription factor 7-Like 2 (TCF7L2) gene polymorphism in patients of type 2 diabetes mellitus from Khyber Pakhtunkhwa population of Pakistan

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Abstract

Background: Type 2 diabetes mellitus (T2DM) is the most prevalent component of metabolic syndrome. Environmental factors and various complex genes like transcription factor 7-like 2 (TCF7L2) gene have involved in the disease development.

Objective: To determine TCF7L2 genetic association (rs7903146C/T and rs12255372G/T) in T2DM patients of Khyber Pakhtunkhwa population of Pakistan.

Subjects and methods: This study comprised of 176 subjects including 118 T2DM patients and 58 healthy controls. Genomic DNA was extracted and genotype of common variants (rs7903146 C/T and rs12255372 G/T) was carried out by amplification-refractory mutation system (ARMS)-PCR of sequence specific oligonucleotides.

Results: The distribution of genotype of TCF7L2 SNPs (rs7903146 C/T and rs12255372 G/T) was significantly associated with T2DM as compared to the controls (p <0.0001). The genetic models of the rs7903146 (C/T) and rs12255372 (G/T) SNPs were significantly associated between cases and controls (p <0.0001). On the other hand, the significant association was observed between the two SNPs and different biochemical parameters like serum fasting glucose, lipid profile, creatinine and blood HbA1c levels (p <0.05).

Conclusion: It is concluded that the SNPs of the TCF7L2 gene are significantly associated with T2DM disease susceptibility in the population of Khyber Pakhtunkhwa of Pakistan.

Keywords: T2DM; TCF7L2; Genetic association, ARMS-PCR, Single nucleotide Polymorphism (SNPs), Khyber Pakhtunkhwa.

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Background

Diabetes mellitus (DM) is one of the commonest metabolic disorder which is characterized by having persistent hyperglycemia due to the abnormalities in insulin secretion or resistance to the insulin action¹. The burden of DM is increasing in the developing countries including South Asian populations. Type 2 diabetes mellitus (T2DM) is the most frequent type of diabetes which can develop metabolic syndrome due to the basic dysfunctions of insulin (resistance or lack of secretion). Insulin resistance in T2DM along with hypertension, obesity and dyslipidemia is the major risk factor for metabolic syndrome. In Pakistan, there are conflicting reports for the prevalence of T2Dm which ranges from 7-19%²,³. In Khyber Pakhtunkhwa region, the prevalence reported for T2DM is about 9%³. T2DM is the multifactorial anomaly including numerous environmental, metabolic and complex genetic risk factors. Various studies have been conducted for the susceptibility of T2DM, but very few reports are available from the South Asian populations⁴. South Asian populations are very genetically heterogeneous and comprised of Pakistan, India and Bangladesh countries. In South Asians resident of UK, it has been demonstrated that genetic variants can affect more than 10% of population which is six times more type 2 diabetes than Caucasian population⁵. Several studies investigated the association of genetic variants including TCF7L2 that could developT2DM⁶,⁷. Genetic polymorphisms of the TCF7L2 have been strongly linked to T2DM susceptibility and more repro-
ducible association with disease than any other reported gene. \textit{TCF7L2} gene is localized on chromosome 10q25 and it encodes 215.9 kb nucleotide sequence. This gene plays role in Wnt-signaling pathway and affects the insulin resistance. Although, \textit{TCF7L2} is considered to play function in insulin secretions from pancreas but the exact mechanism for the gene involvement in diabetes development is unclear. Genetic polymorphism of \textit{TCF7L2} gene has been widely investigated in different populations like Chinese, White Europeans, Israeli, African-American, Argentinians, West.

Africans, Mexicans, Indians, Iranians, and Pakistani groups. On the other hand, some other studies described the lack of association between SNP variants with type 2 diabetes. From Pakistan, some previous reports documented the association of \textit{TCF7L2} SNPs with T2DM and some other data did not find the link to T2DM. Pakistani population is a complex ethnic group with different language speaking and cultures. Very few studies have been conducted for the role of \textit{TCF7L2} with T2DM in this population. Till present, no published data documented the role of \textit{TCF7L2} SNPs in diabetes risk for Pashtun language group of Pakistan. Therefore, this study was aimed to determine the association of common SNPs (rs7903146 C/T and rs12255372 G/T) of \textit{TCF7L2} with susceptibility of type 2 diabetes in population of Northern region of Pakistan from the Khyber-Pakhtunkhwa province.

**Subjects and methods**

Ethical approval of this research was granted from the institutional research board (IRB) of Lady Reading Hospital (LRH) and University of Peshawar, Pakistan (IBR/UoP/2017/7817). Helsinki guidelines (2008) were followed for sample collection of human subjects after written informed consent.

**Subject selection and sample collection**

Sample size calculation carried by online tool (The Survey System Creative, Research Systems). The sample size was calculated by the following formula keeping the confidence level equal to 95% and the margin of error equal to 7%. The calculated sample size for each group was 55 subjects. This study comprised of total 176 subjects including 118 T2DM patients and 58 healthy controls. The cases were recruited from Lady reading hospital, Peshawar and healthy controls were obtained from same ethnic region of Peshawar. T2DM cases were selected according to the prescribed criteria by American Diabetes Association (fasting plasma glucose (FPG) ≥ 126 mg/dL, random plasma glucose of 200 mg/dL or impaired oral glucose tolerance test OGTT (2-hour plasma glucose ≥ 200 mg/dL) and HbA1c level > 6.5%. The patients with related anomalies like type 1 diabetes, type 2 diabetes with complications, gestational diabetes mellitus and heart diseases were excluded from the study. The healthy controls who were apparently normal for fasting blood glucose recruited from the Peshawar region. The demographic data including height, weight, gender and family history was obtained from all the participants. Body mass index (BMI) was calculated. Five ml of blood was collected from each subject including patients and healthy controls. Two ml of the sample was transferred into EDTA vacutainer and three ml was put in serum separating vacutainer. The serum was obtained for biochemical analysis and the EDTA whole blood was used for genetic studies. Samples were stored at -20°C till further analyses.

Biochemical analysis and Genotyping of \textit{TCF7L2} gene

In all subjects, the biochemical analysis was carried out by measuring the serum fasting glucose and lipid profile (Total cholesterol, Triglyceride, LDL-cholesterol and HDL-cholesterol) by using Clinical Chemistry analyzer. Other biochemical tests including blood HbA1c and creatinine were also determined in all the participants. All the experiments were conducted according to the standard protocols by using commercially available kits. The whole blood of all the subjects was processed for extraction of total genomic DNA by using standard method of phenol–chloroform extraction. The concentration and purity of DNA samples were measured by Nano-drop spectrophotometry (OD at 260/280). Genotype analysis of common SNPs of \textit{TCF7L2} (rs7903146 C>T and rs12255372 G>T) was carried out by using amplification refractory mutation system (AMRS)-PCR techniques. Four primers were amplified to genotype each SNP, two outers and two inners (rs7903146 C/T; forward inner primer (C allele) 5’-CAATTAGAGCTAACATATTCTTTTTAGAGAC-3’; reverse inner primer (T allele) 5’-GTCCTCATAAGCCCAATTTAATTATA-GAA-3’; forward outer primer 5’-GTAATGCAGATGATCTGATGTAGATCTCTCT-3’; reverse outer primer 5’-AGAAAAAATACAAAGACATGCAAAGC-3’ and rs12255372 G/T; forward inner primer (T allele) 5’-CTGCCAAGCAGATAATCTCAGCGAAGTT-3’; reverse inner primer (G allele) 5’-GAGGCTGAGATATCAGAAATATGATC-3’; forward outer primer 5’-GGCTGTATGAGTCATTT-3’; reverse outer primer 5’-CTTCATGAGCTGATCAGAAATATGATC-3’; forward outer primer 5’-GGCTGTATGAGTCATTT-3’. The whole blood of all the subjects was processed for extraction of total genomic DNA by using standard method of phenol–chloroform extraction. The concentration and purity of DNA samples were measured by Nano-drop spectrophotometry (OD at 260/280). Genotype analysis of common SNPs of \textit{TCF7L2} (rs7903146 C>T and rs12255372 G>T) was carried out by using amplification refractory mutation system (AMRS)-PCR techniques. Four primers were amplified to genotype each SNP, two outers and two inners (rs7903146 C/T; forward inner primer (C allele) 5’-CAATTAGAGCTAACATATTCTTTTTAGAGAC-3’; reverse inner primer (T allele) 5’-GTCCTCATAAGCCCAATTTAATTATA-GAA-3’; forward outer primer 5’-GTAATGCAGATGATCTGATGTAGATCTCTCT-3’; reverse outer primer 5’-AGAAAAAATACAAAGACATGCAAAGC-3’ and rs12255372 G/T; forward inner primer (T allele) 5’-CTGCCAAGCAGATAATCTCAGCGAAGTT-3’; reverse inner primer (G allele) 5’-GAGGCTGAGATATCAGAAATATGATC-3’; forward outer primer 5’-GGCTGTATGAGTCATTT-3’; reverse outer primer 5’-CTTCATGAGCTGATCAGAAATATGATC-3’; forward outer primer 5’-GGCTGTATGAGTCATTT-3’; reverse outer primer 5’-CTTCATGAGCTGATCAGAAATATGATC-3’; forward outer primer 5’-GGCTGTATGAGTCATTT-3’. The whole blood of all the subjects was processed for extraction of total genomic DNA by using standard method of phenol–chloroform extraction. The concentration and purity of DNA samples were measured by Nano-drop spectrophotometry (OD at 260/280). Genotype analysis of common SNPs of \textit{TCF7L2} (rs7903146 C>T and rs12255372 G>T) was carried out by using amplification refractory mutation system (AMRS)-PCR techniques. Four primers were amplified to genotype each SNP, two outers and two inners (rs7903146 C/T; forward inner primer (C allele) 5’-CAATTAGAGCTAACATATTCTTTTTAGAGAC-3’; reverse inner primer (T allele) 5’-GTCCTCATAAGCCCAATTTAATTATA-GAA-3’; forward outer primer 5’-GTAATGCAGATGATCTGATGTAGATCTCTCT-3’; reverse outer primer 5’-AGAAAAAATACAAAGACATGCAAAGC-3’ and rs12255372 G/T; forward inner primer (T allele) 5’-CTGCCAAGCAGATAATCTCAGCGAAGTT-3’; reverse inner primer (G allele) 5’-GAGGCTGAGATATCAGAAATATGATC-3’; forward outer primer 5’-GGCTGTATGAGTCATTT-3’; reverse outer primer 5’-CTTCATGAGCTGATCAGAAATATGATC-3’; forward outer primer 5’-GGCTGTATGAGTCATTT-3’; reverse outer primer 5’-CTTCATGAGCTGATCAGAAATATGATC-3’; forward outer primer 5’-GGCTGTATGAGTCATTT-3’; reverse outer primer 5’-CTTCATGAGCTGATCAGAAATATGATC-3’; forward outer primer 5’-GGCTGTATGAGTCATTT-3’; reverse outer primer 5’-CTTCATGAGCTGATCAGAAATATGATC-3’; forward outer primer 5’-GGCTGTATGAGTCATTT-3’; reverse outer primer 5’-CTTCATGAGCTGATCAGAAATATGATC-3’; forward outer primer 5’-GGCTGTATGAGTCATTT-3’; reverse outer primer 5’-CTTCATGAGCTGATCAGAAATATGATC-3’; forward outer primer 5’-GGCTGTATGAGTCATTT-3’; reverse outer primer 5’-CTTCATGAGCTGATCAGAAATATGATC-3’; forward outer primer 5’-GGCTGTATGAGTCATTT-3’; reverse outer primer 5’-CTTCATGAGCTGATCAGAAATATGATC-3’; forward outer primer 5’-GGCTGTATGAGTCATTT-3’; reverse outer primer 5’-CTTCATGAGCTGATCAGAAATATGATC-3’; forward outer prime...
GATGATTGT-3'; reverse outer primer 5'-ACGCTTTGAAGGTAGAGAGGACACACT-3') as described earlier16. For polymerase chain reaction (PCR), total reaction volume was 20 µL containing Master-mix, each inner and outer primers, DNA template and nuclease free water. In thermal cycle, the protocol carries the following cycles; the initial denaturation at 95℃ for 5 minutes, then 35 cycles were repeated for denaturation at 94℃ for 30 seconds, annealing at 58℃ for 30 seconds and cyclic extension at 72℃ for 30 seconds and then one cycle of final extension at 72℃ for 10 minutes. The amplified products were resolved on 2% agarose gel and the bands were visualized by using ultraviolet (UV) documentation system. The inspection of each band was inferred to determine the genotype (homozygous or heterozygous) patterns.

Statistical analysis
Data analysis was performed by statistical packages for social sciences (SPSS) version 23. The equation of Hardy Weinberg equilibrium (HWE) was applied to calculate the frequencies of alleles and genotypes for each SNPs of the TCF7L2 gene. The analysis of continuous quantitative variables was done by independent t test and nominal variables by using Chi-square test (χ2). Two sided chi-square test was used to check the differences in variables and genotype frequencies between T2DM patients and controls. The association of SNPs and risk of T2DM was performed by calculating the odds ratio (OR) and confidence interval (95%) by regression analysis. Statistically, p value was considered significant if it was ≤ 0.05.

Results
In this study, 176 subjects (118 T2DM cases and 58 controls) were investigated for the genetic association with the disease. The mean demographic (age, BMI) and clinical parameters (fasting glucose, HbA1c, lipid profile, and creatinine) in subjects are presented in Table 1. Most of the variables were significantly different in T2DM cases as compared to controls (p <0.05), while BMI and serum HDL-cholesterol were not different between the groups (p >0.05).

Table 1. Comparison of demographic and clinical parameters between T2DM cases and controls of the study.

| Variables               | controls (n=58) | T2DM cases (n=118) | p-value  |
|-------------------------|-----------------|-------------------|----------|
| Age (years)             | 42.81 ± 9.82    | 57.48 ± 9.09      | <0.0001* |
| Body Mass Index (Kg/m²) | 24.31 ± 3.29    | 24.66 ± 4.09      | >0.567   |
| Serum fasting glucose (mg/dl) | 90.97 ± 6.55 | 246.53 ± 77.94     | <0.0001* |
| HbA1c (%)               | 5.24 ± 4.66     | 11.42 ± 2.76      | <0.0001* |
| Serum T. Cholesterol (mg/dl) | 156.31 ± 30.09 | 177.91 ± 48.02    | <0.002*  |
| Serum Triglyceride (mg/dl) | 145.31 ± 35.38 | 179.16 ± 66.44     | <0.0001* |
| Serum LDL-cholesterol (mg/dl) | 106.60 ± 25.23 | 121.23 ± 33.33     | <0.004*  |
| Serum HDL-cholesterol (mg/dl) | 33.85 ± 4.49 | 34.62 ± 8.09       | >0.496   |
| Serum Creatinine (mg/dl) | 0.73 ± 0.16     | 1.47 ± 2.11       | <0.010*  |

*variables were considered statistically significant when p value < 0.05.
The frequencies distribution of the genotypes and the alleles of TCF7L2 SNPs (rs7903146 and rs12255372) were determined by amplifying a sequence specific region in T2DM cases and controls (Table 2). For rs7903146 SNP, the most frequent genotype was CT (75.4%) in patients and was 46.5% in controls). While the allele frequency showed that C allele as most frequent in T2DM cases (52%) and in controls it was 64% (Figure 1). For rs12255372 SNP, the GG genotype distribution was 52.3% in cases and was 88.8% in controls. The G allele frequency was lower (74%) in cases and 93% in controls while, the T allele was found as minor allele (Figure 2) On the other hand, the CT and GT genotypes frequencies were significantly higher in Pakistani cases (75.4% and 44.7% respectively) than in controls (46.5% and 9.5% respectively). However, the frequency of the TT genotypes of both SNPs did not show any significant association with the disease risk (9.3%, 2.8% and 12%, 1.5% respectively).

Table 2. Frequencies of genotype and allele distribution of the TCF7L2 SNPs rs7903146 and rs12255372 in cases and controls.

| SNP       | Number of individuals | Genotype   | Frequency of cases (%) | Frequency of controls (%) | Allele Frequency of cases (%) | Allele Frequency of controls (%) |
|-----------|-----------------------|------------|------------------------|----------------------------|-------------------------------|---------------------------------|
| rs7903146 | (118/58)              | CC         | 18 (15.2)              | 24 (41.3)                  | C                             | 52                              |
|           |                       | CT         | 89 (75.4)              | 27 (46.5)                  | T                             | 47                              |
|           |                       | TT         | 11 (9.3)               | 7 (12)                     |                               |                                 |
| rs12255372| (105/63)              | GG         | 55 (52.3)              | 56 (88.8)                  | G                             | 74                              |
|           |                       | GT         | 47 (44.7)              | 6 (9.5)                    | T                             | 25                              |
|           |                       | TT         | 3 (2.8)                | 1 (1.5)                    |                               |                                 |

Fig 1. Allele and genotype of SNP rs7903146 of TCF7L2 in T2DM patients and controls.
The model analysis was studied to detect the association of TCF7L2 genetic polymorphism. For SNP rs7903146, association of rs7903146 polymorphism was found with T2DM in the different genetic models. In the co-dominant model, the heterozygous CT contributed 75% occurrence of type 2 diabetes (unadjusted OR = 4.395; 95% CI =2.081-9.2822; p <0.0001) and in the dominant model (CT/TT) imparted 85% risk (unadjusted OR =3.9216, 95% CI =1.9003-8.0929; p <0.0002) for the disease development. However, no association was observed with TT genotype (unadjusted OR =2.0952; 95% CI =0.6784-6.4707; p >0.1986) and recessive model (unadjusted OR =0.7490, 95% CI =0.2743-2.0454 and p >0.5728) (Table 3). After the adjustment for age, BMI, lipid profile, and other biochemical profiles, the frequency of T allele risk established the strong association of SNP rs7903146 between patients and controls (adjusted OR = 1.6244, 95% CI = 1.0269-2.5694; adjusted p <0.0381).

| Genetic Model/s | Genotype | controls n (%) | cases n (%) | p value | OR (95% CI) | Adjusted p value | Adjusted OR (95% CI) |
|-----------------|----------|----------------|-------------|---------|-------------|------------------|---------------------|
| Co-dominant     | CC (wild-type) | 24 (41.3) | 18 (15.2) | <0.0001 | 4.3951 (2.0810 - 9.2822) | <0.0381 | 1.6244 (1.0269 - 2.5694) |
|                 | CT (Heterozygous) | 27 (46.5) | 89 (75.4) | <0.0001 | 2.0952 (0.6784 - 6.4707) |     |                     |
|                 | TT (homozygous) | 7 (12) | 11 (9.3) | <0.0001 | 0.7490 (0.2743 - 2.0454) |     |                     |
| Dominant model  | CC | 24 (41.3) | 18 (15.2) | <0.0002 | 3.9216 (1.9003 - 8.0929) |     |                     |
|                 | CT-TT | 34 (58.6) | 100 (84.7) | <0.0002 | - | - |                     |
| Recessive       | TT | 7 (12) | 11 (9.3) | 0.5728 | 0.7490 (0.2743 - 2.0454) |     |                     |
|                 | CC-CT | 51 (87.9) | 107 (90.6) | 0.5728 | 3.5236 (1.8126 - 6.8497) |     |                     |
| Over dominant   | CT | 27 (46.5) | 89 (75.4) | <0.0002 | 3.5236 (1.8126 - 6.8497) |     |                     |
|                 | CC-TT | 31 (53.4) | 29 (24.5) | <0.0002 | - | - |                     |
The association of the co-dominant GT genotype (OR =7.9758; 95% CI =3.1544-20.1663; p <0.0001), the dominant model GG vs GT+TT (OR = 7.2727, 95% CI = 3.0344-17.4310 and p <0.0001), and the over-dominant model GT vs GG+TT (OR =7.6983, 95% CI =3.0526-19.4141 and p <0.0001) for the rs12255372 SNP were significantly different between cases and controls (Table 4). However, TT genotype was not associated with T2DM disease (OR =3.0545, 95% CI =0.3082-30.2721 and p =0.3400) and also the recessive model did not link with the phenotype (OR = 1.8235; p = 0.6063) in the studied subjects (Table 4). While, the T allele frequency of rs12255372 SNP was significantly linked between the cases and controls (p =0.0001; OR =4.9793; 95% CI = 2.2806-10.8713).

Table 4. The genetic model of rs12255372 SNP of TCF7L2 association in T2DM cases and controls.

| Genetic Model/s   | Genotype     | controls n (%) | cases n (%) | p value | OR (95% CI)       | Adjusted p value | Adjusted OR (95% CI) |
|-------------------|--------------|----------------|-------------|---------|-------------------|------------------|---------------------|
| Co-dominant       | GG (wild-type) | 56 (88.8)     | 55 (52.3)   | <0.0001 | 4.3951 (2.0810 - 9.2822) | < 0.0001 | 4.9793 (2.2806 - 10.8713) |
|                   | GT (Heterozygous) | 6 (9.5)       | 47 (44.7)   |         | 2.0952 (0.6784 - 6.4707) |
|                   | TT (homozygous) | 1 (1.5)       | 3 (2.8)     |         |                   |
| Dominant model    | GG            | 56 (88.8)     | 55 (52.3)   | <0.0001 | 3.9216 (1.9003 - 8.0929) | -     | -     |
|                   | GT-TT         | 7 (11.1)      | 50 (46.7)   |         |                   |
| Recessive         | TT            | 1 (1.5)       | 3 (2.8)     | 0.6063  | 0.7490 (0.2743 - 2.0454) | -     | -     |
|                   | GG-GT         | 62 (98.4)     | 102 (97.1)  |         |                   |
| Over dominant     | GT            | 6 (9.5)       | 47 (44.7)   | <0.0001 | 3.5236 (1.8126 - 6.8497) | -     | -     |
|                   | GG-TT         | 57 (90.4)     | 58 (55.2)   |         |                   |

Discussion
T2DM is a complex disease afflicted hundreds of millions in the world and it is increasing rapidly nowadays. This prevalence contributes to the growing urbanization of countries, the sedentary life styles, the environmental changes and the genetic factors. Multiple genes have been studied widely and considered as risk factors for developing T2DM. Among these, TCF7L2 gene has been elucidated as the strongest risk factor for developing T2DM\textsuperscript{25}.

In this study, the association was determined for common SNPs (rs7903146 C/T and rs12255372 G/T) of TCF7L2 gene with T2DM in Khyber Pakhtunkhwa population. Our results detected the significant association of heterozygous genotypes of both SNPs (CT and GT; p <0.00001) with T2DM susceptibility in T2DM cases. Furthermore, the T allele frequencies for SNPs were also significantly higher in cases than controls (p <0.000). Genetic variations in TCF7L2 gene has been investigated as risk of T2DM in the diverse populations. In the British ancestry, a study described the nucleotide variations of TCF7L2 were associated with high risk of disease due to the alterations in pro-insuliconcentrations and impaired function of pancreatic β-cells\textsuperscript{26}. The results of present study were in consistent to a previous study in which the T allele frequencies of the SNPs (rs7903146 and rs12255372) were significantly higher in diabetes patients as compared to controls (p <0.00004). Similar results have been reported for rs7903146 SNP in Asian Indian population with type 2 diabetes\textsuperscript{27} and with post-transplant diabetes mellitus\textsuperscript{28}. Recently, a meta-analysis study of Indian population described the positive correlation of rs7903146 SNP with gestational diabetes mellitus reported\textsuperscript{29}. On the other hand, the difference in genotype distribution of homozygous (TT) and heterozygous genotypes (GT and CT) were associated between T2DM cases and controls\textsuperscript{30}, while no association was established for TT genotype in this study. Due to the controversial reports for the involvement of TCF7L2 in T2DM progression but the precise mecha-
nism is still unknown. Though, there are reports suggesting genetic variants of TCF7L2 may influence the factors for T2DM development by changing the GLP-1 levels indirectly by inducing the gene from transcription factors.

A study from Scandinavian population demonstrated the association of T allele distribution with impaired secretion of insulin due to the proliferation beta cells of pancreas. Various meta-analyses demonstrated the association of common SNPs rs12255372 and rs7903146 as contributing factors for T2DM progression in diverse population like South Asian, Caucasian, East Asian and other ethnicities. On the other hand, various studies from local populations of different countries established the link for disease susceptibility and TCF7L2 variations. Although, the results of present study are in accordance to the previous studies but TT genotypes and recessive genetic models have been found associated in previous study by Wu et al., but other genetic models and the GT genotype did not find any association (p >0.05). The model analysis results of this study are comparable to the previous study in which co-dominant and over-dominant models were significantly associated with T2DM. Furthermore, there was found association between biochemical parameters and genetic polymorphism except BMI and HDL which are similar to the previous study. In contrary to the current results, several studies did not demonstrated the link between genetic polymorphism of TCF7L2 and T2DM in different populations and ethnic groups like Chinese population and other regions. The strength of studying genetic polymorphism of common variants in type 2 diabetes patients of different ethnic groups creates the opportunities to establish the biomarkers for diagnosis and disease management. There are some limitations in this study, sample size, population selection on the basis of ethnicity and advance technologies like DNA sequencing and genome wide sequencing may be helpful to document the genetic variants on large scale.

Conflict of interest
None declared.

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