Novel Approaches in Addressing Ovarian Insufficiency in 2019: Are We There Yet?

Konstantinos Sfakianoudis1, Anna Rapani2,3, Sokratis Grigoriadis2,3, Dimitra Retsina2,4, Evangelos Maziotis2,3, Petroula Tsioulou2,3, Polina Giannelou1,2, Konstantinos Pantos1, Michael Koutsilieris2, Nikolaos Vlahos3, George Mastorakos4, and Mara Simopoulou2,3

Abstract
Ovarian insufficiency is described as a multifaceted issue typically encountered in the field of assisted reproduction. The three main identified diagnoses of ovarian insufficiency include premature ovarian failure (POF), poor ovarian response (POR), and advanced maternal age (AMA). Patient heterogeneity in the era of individualized medicine drives research forward leading to the emergence of novel approaches. This plethora of innovative treatments in the service of adequately managing ovarian insufficiency is called to undertake the challenge of addressing infertile patients exploring their reproductive options. This review provides an all-inclusive presentation and critical analysis on novel treatments that have not achieved routine clinical practice status yet, but have recently emerged as promising. In light of the lack of randomized controlled trials conveying safety and efficiency, clinicians are left puzzled in addressing the “how” and “for whom” these approaches may be beneficial. From ovarian injection employing platelet-rich plasma (PRP) or stem cells to artificial gametes and ovaries, ovarian transplantation, and mitochondrial replacement therapy, this descriptive review provides insight toward assisting the practitioner in decision making regarding these cutting-edge treatments. Biological mechanisms, invasiveness levels, efficiency, as well as possible complications, the current status along with bioethical concerns are discussed in the context of identifying future optimal treatment.

Keywords
ovarian insufficiency, premature ovarian failure, poor ovarian response, stem cells, platelet rich plasma, mitochondria replacement therapy

From the time of birth of the first in vitro fertilization (IVF) child in 1978 till today, the common denominator in research is the focus of clinicians’ efforts to override pathophysiological barriers leading to infertility and provide solutions. From controlled ovarian hyperstimulation (COH) protocols to the emergence of intracytoplasmic sperm injection (ICSI), embryo and gamete cryopreservation, and preimplantation genetic screening (PGS), these—once introduced as—novel approaches have been successfully established as clinical routine practice1.

Despite the fact that several infertile couples can be successfully treated, one category of infertile patients still remains highly challenging for clinicians. This category refers to patients diagnosed with ovarian insufficiency. Ovarian insufficiency is a term describing a wide range of pathophysiological conditions where ovarian function is

1 Centre for Human Reproduction, Genesis Athens Clinic, Athens, Greece
2 Department of Physiology, Medical School, National and Kapodistrian University of Athens, Athens, Greece
3 Assisted Reproduction Unit, 2nd Department of Obstetrics and Gynecology, Aretaieion Hospital, Medical School, National and Kapodistrian University of Athens, Athens, Greece
4 Unit of Endocrinology, Diabetes Mellitus and Metabolism, 2nd Department of Obstetrics and Gynecology, Aretaieion Hospital, Medical School, National and Kapodistrian University of Athens, Athens, Greece

Submitted: January 4, 2020. Revised: March 9, 2020. Accepted: April 2, 2020.

Corresponding Author:
Mara Simopoulou, Associate Professor of Experimental Physiology and Embryology, Sr. Clinical Embryologist and Geneticist, Laboratory of Experimental Physiology, Medical School, National and Kapodistrian University of Athens 75, Mikras Asias 11527, Athens, Greece. Email: marasimopoulou@hotmail.com
compromised. Several iatrogenic, pathophysiological, or physiological factors and processes can compromise ovarian function namely maternal age, autoimmune diseases, mutations in genes implicated in the regulation of ovarian function and development, chromosomal and developmental abnormalities, infectious diseases, and gonadotoxic treatments (e.g., chemotherapy or radiotherapy). In the context of assisted reproductive technology (ART), women presenting with ovarian insufficiency tend to produce oocytes of compromised quality, may be unable to ovulate normally, may not be responding to COH protocols or they may present with primary failure of ovarian function. Considering that ART success requires good quality oocytes that will in turn provide good quality embryos of high implantation potential, patients with ovarian insufficiency present with a significantly compromised reproductive potential.

The three main patient categories with ovarian insufficiency are defined as premature ovarian failure (POF), poor ovarian response (POR), and advanced maternal age (AMA). Due to the wide spectrum of pathophysiological conditions leading to ovarian failure, managing patients with ovarian insufficiency is highly perplexing. These challenges have served as a driver for the emergence of a new era in the field of reproductive medicine which has risen dynamically in the last decade, aiming to effectively address ovarian insufficiency in the context of treating infertility. Recent studies investigating the regenerative dynamic potential of the ovaries demonstrate that follicular growth could be stimulated when an adequate ovarian environment is restored. This restoration can be achieved employing factors of known regenerative potential such as stem cells, isolated growth factors, or platelet rich plasma (PRP). Additionally, recent advances in the field of stem cell biology are enabling the reprogramming of differentiated cells to stem cells and subsequently to gamete-like cells also known as artificial gametes. Last but not least, recent studies suggest that mitochondrial function is strongly related to oocyte aging and fertilization failure. These data, in combination with the recent technological development allowing nearly complete replacement of the cytoplasm of an oocyte, introduce the novel approach of mitochondrial replacement therapy for oocyte rejuvenation claiming to lead to an advanced cellular dynamic.

Our team of experts provides herein a timely and essential critical review analysis aiming to report on the novel approaches in addressing the challenging case of ovarian insufficiency, expressing considerations and concerns toward clinical application in humans. This review presents the prospective use of ovarian rejuvenation employing ovari an injection of stem cells, growth factors, and PRP, along with mitochondrial replacement therapy, and continuing to employment of artificial gametes, artificial ovaries, and ovarian transplantation. This review provides a comprehensive and critical analysis with regards to the novel treatments addressing ovarian insufficiency. The manuscript discusses the biological mechanisms entailed, the invasiveness and efficiency, along with the possible complications, the current status, and the bioethical implications of these novel approaches.

### Stem Cell Transplantation

Stem cell therapy has noted considerable success in the field of regenerative medicine during the last two decades. The term “stem cells” is used in order to describe a wide spectrum of undifferentiated cells of the human body having the ability to self-renewal, to proliferate and differentiate to several organ and tissue-specific cell types. Due to their developmental ability, stem cells may represent the "pin of the arrow" in the field of regenerative medicine today.

As anticipated, following successful implementation of stem cell therapy on various systems (as mentioned above), investigating the potential use of stem cell therapy in the context of treating the female reproductive system became the focal point of cutting edge research. This trend has further focused on the context of infertility related to ovarian insufficiency. The explosion of interest witnessed by the scientific community regarding the potential use of stem cell therapy in treating infertility is depicted in a recent systematic review published by Fazelli et al. in 2018. The results provided, indicate that more than of 11,400 published studies have investigated the potential use of stem cell as an effective therapy for several conditions leading to both male and female infertility. From endometrial rejuvenation in patients presenting with Asherman syndrome to restoration of ovarian function in poor responder patients, stem cells presented as an appealing therapeutic tool in the field of reproductive regenerative medicine.

Studies have hitherto focused on the use of the multipotent stem cells and particularly of the mesenchymal stem cells (MSCs) as a potential therapeutic tool for ovarian insufficiency. Their exceptional properties including undifferentiation, proliferation, and renewal capacity are of high significance rendering stem cells a remarkable “tool” in the service of transplantation and drug targeted therapy. Additionally, the unique profile of the multipotent stem cells reduces the rejection and tumorigenic risk and increases stem cell survival rate making therapy more efficient and safer.

Stem cell therapy could perhaps represent the missing piece of the puzzle in overcoming the challenges encountered in the management and treatment of these infertile couples. Multipotent stem cells can effectively induce tissue regeneration, mediated by the two exceptional and unique abilities characterizing them. On the one hand, these cells are able to migrate to damaged tissues and organs and differentiate to tissue-specific cells, promoting healing processes. On the other hand, several studies demonstrate that various multipotent stem cells have the ability to act as paracrine modulators. These cells are able to secrete a wide spectrum of growth factors, chemokines, and mitogenic proteins and via these are able to promote proliferation and...
vascularization. Among these factors are the transforming growth factor α (TGF-α), the TGF-β, the epidermal growth factor and the insulin-like growth factor 1 (IGF-1). Furthermore, multipotent stem cells are able to modulate immune response and present with anti-inflammatory properties. These cells are capable of secreting several growth factors and anti-inflammatory cytokines such as members of interleukin superfamily, tumor necrosis factor, and interferon γ (INF-γ). Moreover, several studies demonstrate that multipotent stem cells exert anti-apoptotic properties limiting tissue degeneration. In regard to the ovarian function, several studies demonstrate that the ovarian niche is able to attract several types of undifferentiated cells from other organs and tissues and especially from bone marrow. These cells are able to migrate into the ovaries supporting folliculogenesis in a procedure known as “homing.” Following the migration to ovaries, these cells secrete several growth factors and hormones orchestrating ovarian function. Considering the pathophysiological base of ovarian insufficiency involving degenerative phenomena leading to the collapse of the ovarian niche and the disruption of the molecular network controlling the ovarian vascularization, it is easy to understand the rationale behind the use of the multipotent stem cells as a potential therapeutic tool. Stem cells could restore ovarian function via the secretion of several hormones and growth factors inducing angiogenesis and tissue regeneration.

The potential therapeutic value of stem cells in ovarian failure was reported for the first time approximately 25 years ago. Sanders et al. published a retrospective observational study indicating that POF patients were able to achieve natural conceptions following bone marrow transplantation. Fast forward to today and this approach albeit promising still retains its experimental procedure status. Several other published reports indicated that ovarian function restoration could be achieved following bone marrow transplantation in some POF patients previously treated with high-dose of chemotherapy or/and total body irradiation. It has been voiced that the mechanism of action for bone marrow stem cells (BMSCs) is to migrate to ovaries through the circulation, exhibiting regenerative properties via the secretion of several hormones and growth factors. Following “homing,” BMSCs could restore the function of pre-existing undamaged follicles, leading finally to restoration of ovarian function.

Following the publication of aforementioned observations, numerous studies investigating the potential use of stem cells for ovarian insufficiency were conducted in animal models prior to initiating human studies. As reported, the compromised function of ovaries in POF or POR models was reversed, estrogen secretion from the compromised tissue was restored, and follicular growth was reactivated, while granulosa cell apoptosis was mitigated accompanied with the occurrence of angiogenesis. What is more, fertility restoration and live births were documented suggesting that BMSCs transplantation could be an effective treatment regarding ovarian function restoration in POF, POR, and AMA patients. Following the encouraging published data indicating the prospective use of BMSCs in addressing ovarian insufficiency, the therapeutic potential of other sources of stem cells indicating their ability to induce similar regenerative phenomena. The results from the aforementioned studies demonstrate that there are several easily accessible novel sources of stem cells that may be used toward ovarian rejuvenation.

Hitherto, clinical studies investigating the potential therapeutic value of these cells in humans are limited. This may be attributed to the fact that there are several limitations entailed in stem cell transplantation, albeit promising outcomes have been documented. In the study of Eddesy et al., 10 women presenting with idiopathic POF were subjected to MSC autologous ovarian transplantation. Recovery of menstruation was achieved 3 months following autologous MSC transplantation for two patients and a subsequent live birth of a healthy offspring has been documented. In another study published by Gabr et al., 30 women with idiopathic POF were also subjected to MSC autologous ovarian transplantation. Ovarian function restoration, including gonadotropin level reduction, and estrogen and anti-Müllerian hormone (AMH) level increase, was observed 1 month post-treatment. The therapeutic effectiveness was impressively sustained during a 48 week follow up. Three patients achieved pregnancy via IVF treatment and one patient reported a pregnancy following natural conception. In the prospective observational pilot study published by Herrera et al., the effectiveness of an autologous stem cell ovarian transplantation protocol (ASCOT protocol) was assessed in POR patients. This was the first clinical study investigating stem cell therapy for ovarian rejuvenation in POR patients. Following bone marrow mobilization with granulocyte-colony stimulating factor (G-CSF) administration and stem cell collection by apheresis from peripheral blood, the 17 patients participating in the study were submitted to CD133+ BMSC ovarian infusion via intra-arterial catheterization under ultrasonography guidance. The time frame between stem cell collection and stem cell infusion was less than 24 h for all patients. Two weeks following ASCOT treatment, a significant increase in the antral follicle count (AFC) was observed. As anticipated, an increase in AMH levels was also observed. Following COH treatment, an increase in the number of AFC and oocytes obtained was recorded in comparison to the patients’ previous medical history. However, the embryo euploidy rate was reported to be low. Five pregnancies were achieved, two following an IVF/embryo transfer (ET) cycle and three via natural conception. The potential effectiveness of stem cell therapy was also recorded in a 45 years old perimenopausal woman. Eight weeks following stem cell therapy, a significant improvement regarding AFC number and AMH levels was observed. The patient achieved pregnancy via an IVF-frozen cycle. The woman was subjected to noninvasive prenatal testing indicating a normal karyotype. A live birth was
finally achieved and the patient delivered via cesarean section at 38 weeks. No complications were observed in the pediatric follow up.

Stem cell therapy seems to be a promising approach. Nonetheless, in order to safely recruit it in clinical application toward treating ovarian insufficiency in the context of infertility, it should be highlighted that it is equally imperative to consider both strengths and weaknesses.

Despite the fact that mesenchymal stem cells are characterized by a low immunologic profile, allogeneic stem cell transplantation could lead to severe graft rejection, posing a risk for the patients' health. Studies in the field demonstrate that autologous stem cell transplantation is related with enhanced therapeutic efficiency compared to allogeneic transplantation in one out of the four women treated with allogeneic transplantation could face severe chronic gynecological graft disease. One of the antipodes, autologous stem cell transplantation could serve as a potential solution in an effort to eliminate the graft rejection risk. However, hitherto, the practice of autologous stem cell transplantation entails two levels of invasiveness. The first one refers to the harvesting procedures and methods of stem cell retrieval via bone marrow aspiration. This fact coupled with the second level of invasiveness encountered at the stage of treatment application entailing stem cell injection into the ovaries, performed via laparoscopy or intra-arterial catheterization, significantly increases concerns. Thus, it is of high significance to investigate the potential use of other sources of stem cells for autologous transplantation, except from BMSCs, namely endometrial stem cells (EnSCs) and adipose derived stem cells (ADSCs). These categories of stem cells have successfully been used as a potential therapeutic tool for addressing ovarian insufficiency in animal models. Nonetheless, studies in humans have not been conducted till date. An overview of the presented studies on the novel approach of stem cells administration indicating study design, methods employed, outcome, and any adverse effects reported is provided in Table 1.

Reporting on the safety of stem cell therapy in ovarian insufficiency, serious considerations should be highlighted, especially in light of the lack of published evidence addressing any side effects related to this novel approach. The most severe potential adverse effect is that unfortunately, intense cell proliferation events that occur following stem cell transplantation may induce malignant formation. It is well documented that long-term cultured mesenchymal stem cells could induce tumorigenesis and metastasis. Following stem cells, isolation from their source of origin, in vitro expansion of cell population is typically required to achieve a clinically suitable grade of stem cells. However, stem cells at higher passages could lead to malignant cell transformation. Considering the aforementioned, it is of added value to determine both the appropriate dose and the specific administration time following stem cell harvesting, in order to achieve a balance between safety and efficiency. It seems that the appropriate management of patient—defined by severity—requires an individualized standardization by the practitioner in the era of precision medicine. Nonetheless, it is imperative to thoroughly understand the underlying mechanisms regulating stem cell treatment.

Regarding the appropriate administration method for ovarian stem cell transplantation, data that have been obtained till date are inconclusive and lacking clarity. Two potential administration methods have been proposed namely stem cells injection into the ovarian cortex via laparoscopy and intra-arterial ovarian catheterization under ultrasonography guidance. Both of these methods appear to be equally efficient. Considering the level of invasiveness, intra-arterial catheterization may be viewed as more patient-friendly. However, in certain cohorts of patients such as AMA women, the practice of intra-arterial catheterization targeting atrophic ovaries of compromised volume due to aging renders the procedure challenging. Future randomized controlled trials (RCTs) comparing the two methods with respect to the patients’ pathophysiology will provide a deeper impact in categorically delineating this issue and addressing the benefits entailed.

In conclusion, robust data provided from future RCTs and well-designed clinical trials are vital in order to ascertain safe conclusions with respect to an array of crucial questions remaining unanswered and perhaps even undefined. This very fact justifies why beyond the promising nature of its application it still retains its experimental status. The appropriate dose and specific administration time following stem cell harvesting, the appropriate administration method, the duration of the recovery period and the potential treatment replication are all issues of value that merit further investigation. Furthermore, cost-effectiveness studies along with studies focusing on assessing patients’ behavioral patterns and response comparing stem cell therapy to the conventional treatment options should be submitted. Regarding the cost of stem cell therapy, it may be safe to conclude that it typically corresponds to a particularly high charge per treatment cycle one that could range from over two to fivefold higher than the cost for a conventional IVF cycle depending on the country where treatment is provided. This fact alone may raise considerable bioethical issues. Considering the multifaceted nature of ovarian insufficiency implicating several different pathophysiological conditions, meticulous observations and documentations are needed in order to determine the specific cohorts of patients for whom stem cell therapy may be efficient and safe. Till then, stem cell transplantation for ovarian rejuvenation should maintain the status and classification of an experimental treatment and practitioners should abstain from embarking on clinical application outside the scope of an approved research protocol.

Platelet-Rich Plasma Intra-Ovarian Infusion

PRP is comprised of crucial ingredients identified in repair mechanisms instigating fundamental physiological
| Publication          | Study design    | Type of ovarian insufficiency | Intervention                                                                 | Method employed                                        | Outcome                                                                 | Adverse effects                                                                 |
|----------------------|-----------------|------------------------------|-------------------------------------------------------------------------------|--------------------------------------------------------|--------------------------------------------------------------------------|--------------------------------------------------------------------------------|
| Salooja et al. (1994) | Retrospective observational study | Iatrogenic POF               | Treatment for AML with high-dose chemotherapy with autologous BM transplantation | Autologous BM transplantation                         | Cycle restoration in young patients (age 21–32); spontaneous pregnancies | NR                                                                           |
| Sanders et al. (1996) | Retrospective observational study | Iatrogenic POF               | High-dose chemotherapy with TBI and BM transplantation for aplastic anemia     | Autologous BM transplantation                         | Cycle restoration; ovulation; and spontaneous pregnancies               | Risk of spontaneous abortion; risk of preterm labor; and risk of SGA          |
| Salooja et al. (2001) | Retrospective observational study | Iatrogenic POF               | Chemotherapy, total body irradiation with peripheral blood or BM transplantation | Peripheral blood or BM transplantation autologous or autologous | Cycle restoration; ovulation; and spontaneous pregnancies               | High risk for maternal and fetal complications in allograft patients         |
| Johnson et al. (2005) | Animal study (mice) | Iatrogenic POF model; ataxia telangiectasia-mutated gene-deficient model | Mice sterilized by chemotherapy, as well as in ataxia telangiectasia-mutated gene-deficient mice | BM transplantation                                     | Cycle restoration; ovulation; and BM could sustain oocyte production in adulthood | NR                                                                           |
| Eggan et al. (2006)  | Animal study (mice) | Iatrogenic POF model         | Parabiotic mice treated with chemotherapy                                     | Parabiotic model                                       | Cycle restoration; ovulation; and BMSCs “homing”                         | NR                                                                           |
| Veitia et al. (2007)  | Case report      | Amenorrheic patient with Fanconi anemia | Allogeneic BM transplantation due to aplastic anemia                           | Allogeneic BM transplantation                         | Cycle restoration; ovulation; Hormone recovery; and spontaneous pregnancy | NR                                                                           |
| Lee et al. (2007)    | Animal study (mice) | Iatrogenic POF model         | Chemotherapy and BM transplantation                                           | Allogeneic BM transplantation via tail vein           | Fertility rescue in mice treated with nonlethal doses of chemotherapy    | NR                                                                           |
| Abd-Allah et al. (2013) | Animal study (rats) | Iatrogenic POF model         | Chemotherapy and BM transplantation                                           | Allogeneic BM transplantation via ear vein           | Hormone recovery; follicle number increase; and VEGF production          | NR                                                                           |
| Guo et al. (2013)    | Animal study (rats) | Iatrogenic POF model; perimenopausal model | Chemotherapy and GCs and BMSCs isolation                                      | Isolated GSs co-cultured with BMSCs                  | Increased GCs apoptosis in perimenopausal rats; apoptosis reduction following BMSCs co-culture | NR                                                                           |
| Lai et al. (2013)    | Animal study (mice) | Iatrogenic POF model         | Chemotherapy and hAFCs transplantation                                          | hAFCs injection into ovaries                         | Hormone recovery and GCs proliferation                                 | NR                                                                           |
| Sun et al. (2013)    | Animal study (mice) | Iatrogenic POF model         | Chemotherapy and ADSCs transplantation                                          | ADSCs transplantation via injection into ovaries or/and intravenous injection | Hormone recovery and follicle number increase and ovulation             | NR                                                                           |
| Liu et al. (2014)    | Animal study (rats) | Iatrogenic POF model         | Chemotherapy and BMSCs injection                                               | BMSCs injection via tail vain                         | Hormone recovery and follicle number increase                             | NR                                                                           |
| Lai et al. (2015)    | Animal study (mice) | Iatrogenic POF model         | Chemotherapy and EnSCs transplantation                                          | EnSCs injection via tail vain                         | Cycle restoration and GSCs depletion reduction                           | NR                                                                           |
| Edessy et al. (2016) | Prospective observational study | Idiopathic POF               | BM aspiration and autologous BMSCs transplantation                             | BMSCs injected into the ovaries via laparoscopy      | Cycle restoration; hormone recovery; spontaneous pregnancy; and live birth | NR                                                                           |
| Publication          | Study design         | Type of ovarian insufficiency | Intervention                                                                 | Method employed                                                                 | Outcome                                                                                                           | Adverse effects   |
|---------------------|----------------------|-------------------------------|------------------------------------------------------------------------------|--------------------------------------------------------------------------------|-------------------------------------------------------------------------------------------------------------------|------------------|
| Gabr et al. (2016)  | Prospective observational study | Idiopathic POF             | G-CSF administration for BM mobilization, BM aspiration, and autologous BMSCs transplantation | BMSCs injected into the ovaries in two different sides: BMSCs were laparoscopically injected into the ovarian cortex and into the ovarian artery | Cycle restoration; hormone recovery; follicular growth reactivation; and pregnancy via IVF or natural conception | NR               |
| Wang et al. (2019)  | Animal study (mice)  | Iatrogenic POF model         | Chemotherapy and MenSCs transplantation                                       | MenSCs injection via tail vein                                                 | Hormone recovery; follicular growth reactivation; and GCs apoptosis reduction                                   | NR               |
| Herraiz et al. (2018) | Animal study (mice)    | POR model                 | SCID ovarietomized mice xenotransplanted with ovarian cortical fragments from POR patients and treated with human BMSCs | Human C133 + BMSCs injection via tail vein                                      | Hormone recovery; follicular growth reactivation; and GCs apoptosis reduction; follicular growth reactivation; GCs apoptosis reduction; and spontaneous pregnancy | NR               |
| Mohamed et al. (2018) | Animal study (mice)    | Iatrogenic POF model         | Chemotherapy and BMSCs transplantation                                        | BMSCs intra-ovarian injection                                                  | Hormone recovery; follicular growth reactivation; and spontaneous pregnancy                                   | NR               |
| Herraiz et al. (2018) | Pilot study           | POR                         | G-CSF administration for BM mobilization, BM aspiration, and autologous stem cell ovarian transplantation protocol (ASCOT protocol) | BMSCs injected into the ovaries via intra-arterial catheterization under ultrasonography guidance | Hormone recovery; follicular growth reactivation; and pregnancy via IVF or natural conception                   | Low embryo euploidy rate |
| Mohamed et al. (2019) | Animal study (mice)    | Iatrogenic POF model         | Chemotherapy and UCMSCs transplantation                                        | UCMSCs ovarian transplantation via laparotomy                                   | Hormone recovery and spontaneous pregnancy                                                                     | NR               |
| Gupta et al. (2018) | Case report           | Perimenopausal women (AMA)  | BM aspiration, autologous BMSCs ovarian transplantation                        | BMSCs injected into the ovaries via laparoscopy                                 | Hormone recovery; follicular growth reactivation; pregnancy via IVF                                            | NR               |

ADSCs: adipose derived stem cells; AMA: advanced maternal age; AML: acute myeloid leukemia; BM: bone marrow; BMSC: bone marrow stem cells; GC: granulosa cells; G-CSF: granulocyte-colony stimulating factor; MenSCs: mesenchymal stem cells; NR: not reported; POF: premature ovarian failure; POR: poor ovarian response; SCID: severe combined immune deficiency; TBI: total-body irradiation; UCMSCs: umbilical cord-derived mesenchymal stem cells.
procedures in a healthy individual. Numerous physiological mechanisms are involved in the multifaceted phenomenon of tissue restoration and repair process, including proliferation, angiogenesis, programmed cell death, and cell migration, all of which are attributed to the valuable synergy of factors evoked by platelets namely vascular endothelial growth factor (VEGF), platelet-derived growth factor AB (PDGF-AB), and TGF-b1. Additionally, growth factors are coupled by the presence of chemokines and cytokines engaging in order for tissue healing to be orchestrated. The rationale behind implementing PRP in the context of managing pathologies affecting tissue integrity was based on the aforementioned key components originally verified in participating in the tissue repair process. Its application may extend to restore these physiological procedures in a system identified with pathologies compromising function.

PRP has noted considerable success in various systems. Following successful implementation of PRP in treating pathologies related to various systems, investigating the potential use of PRP in the context of the female reproductive system was anticipated. In light of the fact that monthly ovulation in the female reproductive system has been described as micro-trauma, the rational of recruiting PRP along with its vital components being involved in tissue repair, may be viewed as one way of justifying its application regarding ovarian function restoration.

Intraovarian injection of PRP in animal models and subsequent resurgence in oocyte production was initially introduced in literature describing the injection of growth factors in the ovary. These studies resulted in increased follicular development and a diminution in apoptotic events in animal models. This radical approach claims to trigger and development and a diminution in apoptotic events in animal induced in literature describing the injection of growth factors subsequent resurgence in oocyte production was initially introduced. Restoration of ovarian function was introduced by Sfakianoudis et al. Within the context of ovarian insufficiency until robust trials. Aside from the promising results already published acknowledging the need for well-designed clinical trials. Aside from the promising results already published for some cohorts of patients, there has been some concern voiced on the aftereffect of PRP application. The intense cell proliferation events that occur may induce malignant formations possibly due to differentiation of stem cells present in ovaries. Nevertheless, the presence of stem cells in ovaries represents a heated debate with opposing schools of thought emerging, supporting the norm of the restricted reserve of oocytes versus the revolutionary concept of

---

In an effort to elucidate all aspects regarding PRP application as a therapeutic option for ovarian insufficiency, it should be highlighted that both strengths and weaknesses are equally imperative to consider. PRP constitutes an autologous product minimizing the potential of side effects that may be related with the administration of a heterologous sample. Despite PRP application being described in the literature as a straightforward procedure, nonetheless, the procedure of PRP application entails predicted challenges with respect to its invasive nature and the fact that the clinical application of intraovarian injection requires meticulous handling and respective patient individualized standardization by the practitioner. For certain cohorts of patients such as menopausal women, the practice of intraovarian infusion targeting atrophic ovaries of compromised volume due to aging adds another level of complexity rendering the procedure rather demanding. What is more, multiple injection sites seem to be required, thus the level of invasiveness may be viewed as considerable. Regarding the cost of this novel approach, an affordable technique has been indicatively proposed for the preparation of blood samples with an estimated cost of $10. However, in clinical practice, the cost of this service is typically reaching over 100-fold of this corresponding to approximately 20% of a conventional IVF cycle cost depending on the country where treatment is provided. PRP injection remains an invasive procedure performed under ultrasound guidance with the anticipated risks not adequately elucidated. Lack of published evidence addressing any side effects is far from reassuring leading to the acknowledgment of the need for well-designed clinical trials. Aside from the promising results already published for some cohorts of patients, there has been some concern voiced on the aftereffect of PRP application. The intense cell proliferation events that occur may induce malignant formations possibly due to differentiation of stem cells present in ovaries. Nevertheless, the presence of stem cells in ovaries represents a heated debate with opposing schools of thought emerging, supporting the norm of the restricted reserve of oocytes versus the revolutionary concept of
| Publication                  | Study design          | Type of ovarian insufficiency                          | Intervention                                                                 | Method employed                                                                 | Outcome                                                                                                                                   | Adverse effects |
|-----------------------------|-----------------------|--------------------------------------------------------|------------------------------------------------------------------------------|--------------------------------------------------------------------------------|------------------------------------------------------------------------------------------------------------------------------------------|-----------------|
| Quintana et al. (2004)      | Animal study (mice)   | NA                                                     | Ovarian injection of vascular endothelial growth factor                      | Growth factor administration                                                   | Direct injection of VEGF into the mouse ovary results in the development of an enhanced vascular network promoting follicular development and diminishing apoptosis | NR              |
| Danfort et al. (2003)       | Animal study (rat)    | Immature ovaries                                       | Ovaries were injected with vascular endothelial growth factor                | Growth factor administration                                                   | VEGF stimulates preantral follicular development; exogenous estrogen up-regulates VEGF expression in the ovary and enhances early follicle growth | NR              |
| Sills et al. (2018)         | Pilot study           | Poor prognosis (patients over age 35 with at least one ovary, infertility duration of > 1 year, at least one prior failed (or canceled) IVF cycle, or amenorrhea for at least 3 months) | PRP activation and intraovarian injection                                   | PRP activation with calcium gluconate and disposition under the ovarian capsule | Improved ovarian function                                                                                                              | NR              |
| Sfakianoudis et al. (2018)  | Case report           | Premature menopause                                    | PRP intraovarian injection                                                  | PRP intraovarian injection                                                     | Hormonal restoration; natural cycle IVF attempt resulting to biochemical pregnancy                                                      | NR              |
| Sills et al. (2019)         | Questionnaire study   | Low ovarian reserve patients and/or patients with at least 1 failed IVF attempt | Ovarian PRP                                                                | Ovarian PRP                                                                  | Ovarian hormonal rejuvenation; reversing low or absent ovarian reserve                                                                 | NR              |
| Wang et al. (2019)          | EX vivo               | Mesenchymal stem cells from healthy individuals         | PRP as a proliferation and differentiation promoter                          | Mesenchymal stem cells proliferation and differentiation induced by PRP       | PRPs with different platelet concentrations exerts osteogenic, adipogenic, and chondrogenic differentiation                               | NR              |
| Farimani et al. (2019)      | Case series           | Poor ovarian response                                 | PRP intraovarian injection                                                  | PRP intraovarian injection                                                     | Improved oocyte yield; spontaneous conceptions and live birth                                                                          | NR              |
| Sfakianoudis et al. (2019)  | Case series           | Poor ovarian response                                 | PRP intraovarian injection                                                  | PRP intraovarian injection                                                     | Hormonal improvement; natural conception and live birth                                                                               | NR              |
| Pantos et al. (2019)        | Case series           | Premature ovarian failure                             | PRP intraovarian injection                                                  | PRP intraovarian injection                                                     | Restoration of menstruation; improvement in hormonal profile; natural conception                                                        | NR              |

IVF: in vitro fertilization; NR: not reported; PRP: platelet-rich plasma; VEGF: vascular endothelial growth factor.
ovarian germline stem cells capable to rejuvenate the ovarian tissue.

While investigating the possibilities of PRP application, precise primary and secondary outcomes should be decided upon, in order for a conclusive report on PRP’s effectiveness to be delivered. Undoubtedly, the molecular interaction needs to be elucidated prior to designing large RCTs that aim to convey safety in practice. It has been voiced that PRP’s concentration may play a key role in cell proliferation and differentiation of mesenchymal cells. Managing various pathologies may require different optimal concentrations. This may explain the heterogeneity in the results and outcomes documented and observed following PRP application amongst patients. A personalized approach taking into account the underlying pathology should be pursued. Several factors potentially affecting the end result namely time intervals between PRP applications, volume administered, as well as the threshold of the maximum number of allowed interventions should be addressed. Pondering on whether PRP has a place in clinical practice in addressing ovarian insufficiency, a study submitted the thesis that aside from promising outcomes in ovarian physiology, an essential improvement on the treated patients’ quality of life has been noted. This could be plausibly attributed to the hormonal rebalance recorded following intervention, encouraging the employment of PRP and extending its consequences beyond a strict physiological and molecular context.

The ovarian reserve issue may be a multifaceted matter, thus managing these patients may require a plethora of available tools and techniques. Hitherto, the complex dialog between PRP components and the ovary remains to be deciphered. In anticipation of solid molecular and clinical evidence to support the validity of the “ovarian rejuvenation” concept, PRP may face the risk of remaining a costly service. Nonetheless, preliminary data showcase its potential of being perceived as a ground-breaking medical innovation. Could that be enough to give it a try? Maybe. Nonetheless, we need to question whether evidence is accumulating to a satisfactory degree leaving the scientific community content in deciding to introduce routine clinical application of PRP for ovarian insufficiency.

Ovarian Tissue Transplantation

Ovarian tissue cryopreservation may not be considered as a unique novel approach. Since the first implementation, over 130 babies have been born. However, it is still considered to be an experimental approach. Various reports attempt to establish a commonly accepted technique for ovarian transplantation. Orthotopic transplantation may present as an optimal practice since the microenvironment around the transplanted tissue is the most suitable for follicular development. On the other hand, there is limited number of tissue fragments allowed to be transferred due to space constriction, while the level of invasiveness presents as a major limitation resulting in severe pelvic adhesions. It should be highlighted that natural conception may be achieved in cases of orthotopic transplantation. The method of transplanting ovarian tissue may be considered technically appealing since the utilized ovarian tissue is characterized by a solid extracellular matrix structure coupled with complex cellularity. Furthermore, biomaterials are introduced in order to enhance the structure and integrity of ovarian tissue by improving adhesion, proliferation, and differentiation of the cells involved. Natural biomaterials present with significant heterogeneity in their composition spectrum, while synthetic biomaterials’ ability to promote cellular and tissue remodeling is restricted. Thus, both sources present equally with weaknesses regarding the configuration of an optimal model for ovarian tissue transplantation.

Since undergoing autologous ovarian tissue transplantation requires collecting tissue prior to acknowledging and identifying signs of ovarian deficiency, this strategy is typically addressing strictly a particular cohort of patients. These patients are women that are destined to experience an anticipated ovarian shortage following cancer therapy, without excluding the patient group that will experience anticipated gamete exhaustion due to a prognosis or even a diagnosis of POF. In cases of a diagnosis of malignancy, ovarian tissue cryopreservation for subsequent transplantation is suggested, since it could be performed in a short time-frame without delaying the initiation of therapy. From the first attempts on ovarian tissue transplantation almost 20 years ago to numerous successful live births following this procedure published in literature since then, a new line of approaching ovarian deficiency has been introduced and established.

Valuable observations have been documented in literature reporting on restoration of fertility, ovarian activity reinstatement, endocrine function reactivation, and follicular recruitment. What is more, transplanting ovarian tissue has been proposed strictly on the grounds of ovarian rejuvenation leading to what may be described as a hormonal ovarian reboot irrespectively of pursuing fertility treatment. Ovarian tissue transplantation enabled a hormonal restoration lasting an impressive 7-year period, which was documented to be extended to a 12-year restoration period when the procedure was repeated. Nonetheless, one should not fail to highlight that this invasive procedure minimizes and potentially jeopardizes the current ovarian reserve for the patient, thus bioethical concerns are raised.

A major limitation to consider is that autologous transplantation entails the risk of reintroducing cancerous cells to the treated ovary. Moreover, immune rejections constitute a major complication for this line of approach, albeit the autologous source of tissue may contribute to overcoming this side effect. Ischemia has also been described as a detrimental side effect following ovarian tissue transplantation. The main cause of this event is the lack of vessel anastomosis during the tissue transplantation procedure. As a result, the transplanted ovarian tissue is exposed for a certain time period of 5 days in a hypoxic and ischemic environment.
which demands an urgent intervention to induce neovascularization. In this context, various approaches have been attempted to optimize the outcome including employment of growth factors as a supplementary tool to assist toward completion of neovascularization in a timely fashion. However, their application in human models is nowhere near consideration even though current experimentation on animals has contributed favorable results. Concerns over subsequent cancer diagnosis have emerged, nonetheless, they were considered as a low probability event when tissue is sourced from young women aged 20–25. The technique of cryopreservation and its respective efficiency should be further taken into account as a challenging part of the ovarian tissue transplantation approach. It is the actual cryopreservation technique that may perhaps result in compromised outcomes misleadingly attributed to the transplantation technique. Contemplating on the logistics of performing ovarian tissue transplantation for restoring endocrine function in menopausal patients, the procedure is less invasive since the graft site may not involve the pelvic cavity. Furthermore, it may be performed under local anesthetics, while in case of an ovarian neoplasm emergence, the ovarian tissue implant could be immediately extracted. An overview of the presented studies on the novel approach of ovarian tissue transplantation indicating study design, methods employed, outcome, and any adverse effects reported is provided in Table 3.

All of the above constitute vital indications in favor of this treatment option. Nonetheless, similarly to all hormone replacement approaches aiming to postpone the permanent hibernation of the ovaries for women reaching the end of their reproductive life-span, ovarian tissue transplantation entails risks. Apart from the concerns raised when rejuvenation is the primary patients’ interest, practitioners appear hesitant to embark on this suggested approach even in cases where patients are facing a reproductive dead-end. This may be attributed to the realization that ovarian transplantation has been around for several decades, failing nonetheless to be established as a foolproof approach in clinical practice one that could provide specialists with confidence regarding application.

### Artificial Ovary

An additional step in ovarian tissue preparation prior to implantation has been proposed in an effort to address the risk of reestablishing the treated disease. It includes the formation of an ovarian tissue scaffold where oocytes appear, grow, and subsequently “harvested.” Hitherto, this approach has been applied in animal models in which granulosa cell-oocyte complexes were collected and subjected to a 3D in vitro culture. Following standard proliferation, oocyte yield was higher than a conventional IVF approach could achieve. Likewise, constructing a 3D artificial ovary has been attempted in order to successfully incorporate, accommodate, and enable maturation of human oocytes. Most previous attempts have failed due to the common denominator of implementing a solid structure albeit lacking adequate mimicking of vital endocrine and paracrine pathways. Three ovarian cell types were included namely theca cells, granulosa, and oocytes. By including theca cells as a fundamental component of the 3D ovary, hormonal production was achieved. Interestingly, implementing collagen or alginate scaffolds failed to ascertain a similar endocrinology response. What is more, enhanced interaction between cell types was observed conspiring toward the development of a physiological model. Under these circumstances, maturation and development of the primordial oocytes may be enabled. The novelty behind this approach is indisputable; however, the fact that it constitutes what may be described as an “isolated” system lacking the complexity of numerous interactions with other physiological systems should be highlighted.

Bioengineering strategies to restore fertility and address ovarian insufficiently include transplantation of fresh or cryopreserved ovarian tissue and tissue engineering involving implementation of growth factors, stem cells, pluripotent stem cells, mesenchymal stem cells, and biomaterials. The percentage of cancer patient survivors is increasing, translating to patients who despite succeeding in overcoming a disease, they will be reproducibly challenged experiencing infertility strain and concerns attributed to gonadal injury during therapy. Numerous successful attempts at performing ovarian tissue cryopreservation and subsequent transplantation reveal valuable data on the efficiency of the approach. However, little is known in regard to whether this approach should be considered as a method to delay the onset of menopause—a trend that appears to be rapidly evolving. In regard to the concept of the artificial ovary, further considerations should be taken into account. An overview of the presented studies regarding the novel approach of artificial ovary indicating study design, methods employed, outcome, and any adverse effects reported is provided in Table 4. It should be highlighted that exposing sensitive tissue in a challenging environment may entail certain epigenetic risks which in turn may raise bioethical issues. How far is too far when the—once considered as science fiction—concept of harvesting gametes becomes a reality? Until strong evidence accumulates, the concept of artificial ovary clearly remains at the experimental stage. With RCTs being an extrapolation at best, clinical practice will have to wait.

### Artificial Gametes

There are four basic cell types employed in the formation of artificial gametes, including germ line stem cells, induced pluripotent stem cells (iPSCs), somatic cell nuclear transfer to embryonic stem cells (SCNT), and SCNT to donor oocytes. Successful implementation resulting to live birth following artificially generating gametes has only been reported in animal models, thus hitherto artificial gametes.
| Publication                  | Study design  | Type of ovarian insufficiency | Intervention                                                                 | Method employed                                                                 | Outcome                                                                 | Adverse effects                                                                 |
|-----------------------------|---------------|-------------------------------|------------------------------------------------------------------------------|---------------------------------------------------------------------------------|--------------------------------------------------------------------------|--------------------------------------------------------------------------------|
| Oktay and Karlikaya (2000)  | Case report   | Iatrogenic POF                | Salpingo-oophorectomy and ovarian tissue transplantation                     | Tissue sutured laparoscopically beneath left pelvic peritoneum                   | Long term graft survival, follicle emergence, ovulation and subsequent menstruation following ovarian stimulation | NR                                                                            |
| Donnez et al. (2004)        | Case report   | Iatrogenic POF                | Chemotherapy and ovarian tissue transplantation                               | Orthotopic autotransplantation of ovarian cortical tissue by laparoscopy        | Recovery of regular ovulatory cycles                                      | Risk for recurrence of disease following implantation, oocyte quality compromised by heterotopic site transplantation |
| Meirow et al. (2005)        | Case report   | Iatrogenic POF                | Chemotherapy and ovarian tissue transplantation                               | Strips of thawed ovarian tissue transplanted to the left ovary and small fragments injected into the right ovary | Menstruation and live birth following IVF employing a natural cycle       | Risk of grafting malignant cells                                             |
| Silber et al. (2005)        | Case report   | Premature menopause           | Ovarian transplantation between healthy monozygotic twins                     | Ovarian cortex graft                                                           | Live birth following natural conception                                  | Risk of follicular atresia                                                   |
| Demeestere et al. (2007)    | Case report   | Iatrogenic POF                | Chemotherapy and ovarian tissue transplantation                               | Orthotopic and heterotopic autotransplantation of cryopreserved ovarian tissue | Recovery of ovarian function and live birth                               | Reintroducing malignant cells and ischemic damage during revascularization; limited life span of the graft |
| Andersen et al. (2008)      | Case series   | Iatrogenic POF                | Autotransplantation of frozen/thawed ovarian tissue and assisted reproduction | Orthotopic and heterotopic autotransplantation of ovarian cortical tissue       | Restoration of menstrual cyclicity, conception, and live birth            | NR                                                                            |
| Schubert et al. (2008)      | Animal study  | Iatrogenic POF                | Xenotransplantation of ovarian tissue                                          | Fresh or frozen human ovarian cortex grafted into mice                          | Increase of primary and secondary follicles; decrease of primordial follicles | Massive oxidative stress                                                   |
| Van Eyck et al. (2009)      | Animal study  | Iatrogenic POF                | Laparoscopic surgery for nonovarian benign gynecological disorders and tissue transplantation | Frozen–thawed human ovarian tissue fragments grafted to intraperitoneal cavity | Graft revascularization                                                   | Ischemic tissue damage                                                      |
| Kim et al. (2012)           | Case series   | Iatrogenic POF                | Laparoscopic oophorectomy and laparotomy                                      | Heterotopic site transplantation                                               | Ovarian endocrine function restoration                                    | Primordial follicles cryoinjury                                              |
| Kong et al. (2017)          | Animal study  | Iatrogenic POF                | Bilateral ovariectomy performed on mice                                        | Bovine ovarian tissue xenotransplanted into mice                               | Administration of Ang-2 and VEGF prior to transplantation alleviates ischemic damage, maintains follicle normality and density; reduces levels of apoptosis and fibrosis in the grafts | NR                                                                            |

NR: not reported; POF: poor ovarian response.
constitute an experimental procedure. Based on observations in animal models where artificial gametes may be transferred to ovaries enabling natural conception, the scientific community cannot help but ponder whether this strategy may present as a useful tool in the hands of practitioners managing ovarian insufficiency in the distant or not too distant future.

The option of employing artificial gametes has been introduced as an alternative solution for patients lacking the potential to produce functional gametes such as women with premature ovarian insufficiency. In human models, the development of artificial oocytes has been described, while a successful fertilization has been confirmed in one case study. As reported by White et al., human oogonial stem cells (OSCs) were transplanted to human ovary tissue xenografted to mice following in vitro proliferation, resulting in the development of artificial oocytes. A challenging process of gamete formation originating from human embryonic stem cells has been described; however, this route was outlined as insufficient since differentiation of embryonic cells in culture systems entails certain difficulties to perform. Human amniotic stem cells have also been successfully recruited in an effort to form artificial gametes. Moreover, engaging human hepatic cell lines in order for germine cells to originate resulted in the formation of follicle-like structures, subsequent blastocyst-like structures, and finally germ cell/embryonic tumors following prolonged culture. An overview of the presented studies on the novel approach of stem cells administration indicating study design, methods employed, outcome, and any adverse effects reported is provided in Table 4.

It is vital for concerns on safety to be thoroughly addressed prior to even considering the route to clinical application. With RCTs being a mere hypothesis, it appears that the experimental stage regarding artificial gametes may take longer than anticipated. Despite the biological risks entailed, there are significant ethical concerns raised in a societal level that should be addressed prior to pursuing this option as a solid novel approach for management of ovarian insufficiency. A clear presentation of the patients for whom this approach may be beneficial and for whom access should be granted is imperative.

### Table 4. An Overview of Published Studies on the Novel Approach of Artificial Ovary Indicating Study Design, Methods Employed, Outcome, and Adverse Effects Reported.

| Publication          | Study design       | Type of ovarian insufficiency | Intervention                      | Method employed                      | Outcome                                      | Adverse effects |
|----------------------|--------------------|------------------------------|-----------------------------------|--------------------------------------|----------------------------------------------|-----------------|
| Pangas et al. (2003) | Animal study (mice)| Dissected ovaries            | Granulosa oocyte complexes isolated from mice | BMSCs injection via tail vein | Hormone recovery; Follicle number increase | NR              |
| Krotz et al. (2010) | Ex vivo study      | Oophorectomy for benign indications | Oophorectomy and artificial ovary creation | Theca and granulosa cells seeded into micro-molded gels and self-assembled into complex 3D microtissues | An artificial human ovary can be created with self-assembled human theca and granulosa cell microtissues | NR              |

BMSC: bone marrow stem cells; NR: not reported.

### Table 5. An Overview of Published Studies on the Novel Approach of Artificial Gametes Indicating Study Design, Methods Employed, Outcome, and Adverse Effects Reported.

| Publication          | Study design       | Type of ovarian insufficiency | Intervention                      | Method employed                      | Outcome                                      | Adverse effects |
|----------------------|--------------------|------------------------------|-----------------------------------|--------------------------------------|----------------------------------------------|-----------------|
| Aflatoonian et al. (2009) | Ex vivo study | Cell lines                   | Human embryonic stem cells        | Differentiation was induced          | Human primordial stem cells as well as postmeiotic spermatids were formed in vitro; steroid hormones production was detected | NR              |
| Cheng et al. (2012)  | Ex vivo study      | Human amniotic fluid stem cells | Oocytes isolated from healthy young women for sex reassignment purposes | Differentiation into oocyte-like cells Intraovarian oogonial stem cells injection and xenografting | Differentiation into oocyte-like cells Intraovarian oogonial stem cells injection and xenografting | NR              |
| White et al. (2012)  | Animal study (mice)|                         | Ovaries isolated from healthy young women for sex reassignment purposes | Human hepatic germ-line                          | Germ-line cells formation | NRLenovocyte-like cells, and zona pellucida-like morphology were observed | NR              |
| Ma et al. (2013)     | Ex vivo study      |                             | Germ-line cells formation         | Germ-line cells formation               | Follicle-like structures, generating oocyte-like cells, and subsequently developing into blastocyst-like structures in vitro were observed | Hepatic teratomas |

NR: not reported.
Mitochondrial Replacement Therapy

Mitochondria are identified as “the powerhouse of the cell.” The role of mitochondria in the development of the preimplantation embryo and its implantation potential has been studied for at least two decades. The processes of fertilization and proper embryonic development require great amounts of adenosine triphosphate (ATP) which is produced by mitochondria. Oocyte mitochondrial DNA (mtDNA) content has been associated with poorer oocyte quality and ovarian insufficiency. The prognostic value of the copy number has been a topic of controversy in literature since a number of studies observed a statistically significant higher number of mtDNA copies in embryos that resulted in implantation failure, whereas other studies observed no statistically significant difference.

Nonetheless, it is evident that maternal aging is the cause for mitochondrial dysfunction through swelling and cristae disruption in oocytes. The employment of mitochondria from a young healthy donor was hypothesized to provide the preimplantation embryo with a better and more potent developmental environment during the early stages. This may have served as the driver leading to early attempts of mitochondrial replacement therapy, which was initially approached by performing ooplasmic transfer, as first described by Cohen. Although ooplasmic transfer presented with success in enabling an improvement in embryo quality resulting to reports of clinical pregnancy and live birth, the Food and Drug Administration (FDA) decided to ban its application in 2001 due to ethical and medical concerns mainly related to heteroplasmy and interrelated adverse effects.

More than a decade following the FDA ban, novel approaches regarding mitochondrial transfer in order to achieve improved ovarian function have been implemented. To avoid concerns introduced by including a heterologous donor, the initial approach employed the autologous germ-line mitochondrial energy transfer (AUGMENT). The protocol of AUGMENT includes isolation of mitochondria from the patient’s OSCs, processed and injected into the patient’s own oocytes during ICSI.

The first application of AUGMENT was performed by Fakih et al. targeting women presenting with low quality oocytes and/or embryos, diagnosed with diminished ovarian reserve, ovulatory dysfunction, polycystic ovarian syndrome (PCOS), tubal factor, and endometriosis. A total of 93 women from two IVF centers were recruited. The results of the study supported a beneficial role for the AUGMENT as the clinical pregnancy rate achieved was 35% and 22% for each center respectively, in contrast to the 11% and 4% corresponding to the clinical pregnancy rate of the same patients prior to AUGMENT. In the same study, sibling oocytes from 25 patients were randomly allocated to two groups, one designed to undergo the AUGMENT therapy and a control group. Out of the 25 patients, two underwent embryo-transfer with embryos originating from the control group, for nine patients, embryo-transfer was not performed, and 14 patients underwent embryo-transfer with embryos originating from the AUGMENT group. Eight pregnancies were observed all originating from the experimental group. A second, albeit smaller study, conducted by Oktay et al. included 15 women undergoing an autologous germline mitochondrial energy transfer, described as autologous mitochondrial injection (AMI) by the authors. Of the 15 women recruited, only 10 underwent embryo-transfer, and 4 of them achieved a clinical pregnancy status.

These promising results were contradicted by the interim analysis of a discontinued large-scale triple blind RCT. Following the enrollment of about 31% of the study’s total population (n = 59/190), an interim analysis was performed. According to the results of the study, AUGMENT did not increase neither the fertilization rate nor the euploid blastocyst formation rate. On the contrary, blastocyst formation rate was decreased in the experimental group. Moreover, embryo grading according to Spanish Association for the study of Reproductive Biology (ASEBIR) criteria was poorer, and live birth rates were similarly lower in the experimental group. These observations in the interim analysis rendered the recruitment of more patients unnecessary and the study was terminated, although the clinical pregnancy and live birth rates presented in the study were better when compared to the other studies regarding the autologous mitochondrial transfer. It should be highlighted that patients included in the RCT were not necessarily of poor ovarian function as they were younger than 42 years old, with median AMH of 1.69 ng/ml, with an interquartile range from 0.97 to 3.24 ng/ml. This is a reason for caution in all studies employing autologous mitochondrial transfer. The population enrolled is mainly a mixed population with only a few participants being diagnosed with diminished or poor ovarian response. It may be possible that the autologous mitochondrial transfer is suitable for a specific population which still remains to be identified.

Apart from the autologous mitochondria transfer, heterologous mitochondrial transfer, known as mitochondrial replacement therapy (MRT), has been employed mainly on the grounds of avoiding transmission of mitochondrial diseases. There are four protocols available for MRT, the pronuclear transfer (PNT), the polar body transfer (PBT), the maternal spindle transfer (MST), and the germinal vesicle transfer (GVT). The three latter techniques require the employment of donor enucleated MII oocytes, whereas PNT requires the employment of a donor zygote following removal of the pronuclei and polar bodies. In PNT, both parental pronuclei are transferred to the donor zygote. GVT requires transfer of the maternal germinal vesicle enclosing the nuclear DNA to the donor enucleated oocyte, and PB requires the transfer of the polar body of the maternal MII oocyte in the enucleated donor oocyte. MST requires the transfer of the spindle transfer enclosing the nuclear DNA to an enucleated donor oocyte. PNT and PBT have been evaluated as possible methods for MRT with embryos.
reaching the blastocyst stage without progressing to implantation or pregnancy. Zhang et al. employed PNT to achieve a pregnancy for a 30-year-old woman with mitochondrial disease. The same group achieved a pregnancy employing MST, for a woman with Leigh syndrome. It should be mentioned that although initially a permission for long-term follow-up was granted, the parents of the offspring decided to forego the permission, thus pediatric follow-up data are not available. In the context of employing MRT, MST is the sole technique applied in an effort to address poor ovarian response. The clinical trial has so far resulted in one live birth, from a 32-year-old poor responder with four previous failed IVF attempts. This technique has raised considerable ethical concerns. The European Society of Human Reproduction and Embryology (ESHRE) issued a formal thesis on strong discouragement and disapproval regarding the technique’s application in the context of addressing poor ovarian response. Practitioners should refrain from adopting the clinical practice of MST for anything else other than to avoid transmission of a mitochondrial inherited disorder for which case no other alternative is viable. It is strictly this approach that has claimed regulation status by the Human Fertilisation and Embryology Authority (HFEA) in the United Kingdom. It is clear that in the context of ART treatment, there is a lack of robust or even adequate data indicating safety and effectiveness of the method, which currently maintains its experimental status. An overview of the presented studies on the novel approach of stem cells administration indicating study design, methods employed, outcome, and any adverse effects reported is provided in Table 6.

Long term follow-up of MRT is required in order to report on the safety and efficacy of the procedure or even to buttress the original hypothesis trail of thought driving its development. Conducting more RCTs is a requirement in the topic. The RCTs should present with strict inclusion and exclusion criteria, and should not employ a mixed population. It is of imperative importance to evaluate the safety and the efficiency of a technique that may present as one of the turning points of ART practice. Moreover, although MST appears to be the sole option in providing favorable results in humans, all protocols should be evaluated, prior to deciding on superiority in the application. Although heteroplasmity levels seem to be low by employing the novel MRT techniques, mito-nuclear interactions are a topic of research. Two meta-analyses performed in animal models have reported contradicting results. The first chronologically published meta-analysis has reported that mito-nuclear interactions do not pose a side-effect of MRT. The second meta-analysis has reported that MRT may routinely affect offspring biological and biochemical characteristics with large scale effect due to mito-nuclear interaction, which seem to be even greater in humans. It is of interest that the second meta-analysis has been questioned regarding its design by the author of the first. Until large scale human studies are performed, the scientific community ought to practice cautiousness and maintain a vigilant stance in order to limit possible and perhaps probable long-term side effects that currently may not be efficiently identified, understood, treated, or reversed. At the moment, the research developing and fueling this novel approach is clearly not validated to justify clinical application. Perhaps this may be a good time for the scientific community to re-examine the “certainties” that fueled the idea that replacing mitochondria may be the answer to ovarian insufficiency prior to proceeding with clinical application.

**Discussion**

The concept of ovarian reboot has recently attracted heated interest. This may be attributed to the increased prevalence of the diagnosis of ovarian insufficiency for cases that demand effective options on managing their subsequent infertility status. This increase may be justified on the grounds of the remarkable results and elevated survival rates of treating cancer patients in the era of precision medicine. These patients can now enjoy a prolonged life span which nonetheless may harbor infertility and reproductive dilemmas. On another note, the concept of delaying aging has been intensively promoted in media. This reality reinforces

---

**Table 6. An Overview of Published Studies on the Novel Approach of MRT Indicating Study Design, Methods Employed, Outcome, and Adverse Effects Reported.**

| Publication       | Study design       | Type of ovarian insufficiency | Intervention | Method employed | Outcome                        | Adverse effects |
|-------------------|--------------------|------------------------------|--------------|----------------|--------------------------------|----------------|
| Fakih et al.(2015) | Prospective cohort study | Diminished ovarian reserve | MRT          | AUGMENT         | Enhanced clinical pregnancy rate | NR             |
| Oktay et al.(2015) | Case series        | Mixed population             | MRT          | AUGMENT         | Enhanced clinical pregnancy rate | NR             |
| Labarta et al.(2019) | RCT               | Impaired embryo quality      | MRT          | AUGMENT         | No statistically significant improvement | Impaired blastocyst formation rate | NR             |
| ISRCTN 1455145    | Prospective cohort study | Impaired embryo quality      | MRT          | Maternal spindle transfer | Results have not been reported | NR             |

AUGMENT: autologous germline mitochondrial energy transfer; MRT: mitochondrial replacement therapy; NR: not reported; RCT: randomized controlled trial.
practitioners toward engaging and endeavoring novel approaches fueled by the patients’ request. On the antipode, understandably clinicians appear hesitant and practice skepticism in embarking on novel proposed options, since data are often conflicting and lack robustness. Nonetheless, reports in literature describing case series and pilot studies communicate to the scientific community that clinical application of novel—albeit not yet having reached routine clinical application status—approaches may present a real option. It may not be uncommon for certain practices to invest in optimizing a new line of treatment on premature ovarian insufficiency. The variety of approaches including stem cells, platelet-rich plasma, ovarian tissue transplantation, formation of artificial gametes, even the latest emerging idea of mitochondrial replacement technique calls for specialization and expertise by clinics and practitioners. A critical assessment providing a comparative perspective between all novel approaches, regarding issues of importance to the practitioner from cost and practical aspects of application to novelty, ethical concerns raised, and potential for acquiring future clinical routine practice is provided in Table 7. Despite the variation considering the respective unidentified molecular pathways leading to a compromised ovary, “could a successful application in restoring function translate to rewinding time?” “Could the resulting follicles and oocytes carry a distinctive element compromising their potential and rendering them present, but nonetheless of poor quality and potential?”

In order to properly assess these points of heated debate, certain guidelines should be proposed and established. The common denominator of all novel approaches in the service of overcoming ovarian insufficiency as critically presented herein lies in inconclusive but rather promising data. This very fact renders all of them to be at an experimental stage rather than of established clinical routine practice. Reestablishing fertility potential may be the main objective in published literature. Nonetheless, restoring fertile status following the diagnosis of ovarian insufficient may be defined by numerous criteria, enabled by various approaches, and confirmed employing different outcome measures depending on study design and population identity. Interestingly, and as shown herein, the perception of a successful outcome varies considerably amongst researchers. This may only be delineated by concrete studies examining outcomes on a clinical as well as on a basic research level. It is of high significance to highlight that all the approaches included herein have been proposed as promising techniques aiming to address the infertility related to ovarian insufficiency. Societies of Reproductive Medicine have issued in the past statements and guidelines, clearly shifting the status if a certain technique from being an experimental procedure to one classified as ascertaining the status of clinical routine practice. Such was the case for oocyte vitrification and warming121. Nonetheless, hitherto strong evidence is not in place for any of the presented techniques in order for the label of experimental procedure to be lifted. In the future, well-designed studies presenting clearly indicated outcome measures, limitations, and complications, while ascertaining

Table 7. A Critical Assessment Providing a Comparative Perspective between All Novel Approaches, Regarding Issues of Importance to the Practitioner from Cost and Practical Aspects of Application to Novelty, Ethical Concerns Raised, and Potential for Acquiring Future Clinical Routine Practice Status.

| Approach                        | Cost | Invasiveness of application regarding the patient | Adverse effects of application regarding the patient | Time frame of action | Novelty | Ethical concerns | Potential for acquiring status of clinical routine practice |
|---------------------------------|------|--------------------------------------------------|----------------------------------------------------|--------------------|--------|-----------------|----------------------------------------------------------|
| Stem cells                      | ++   | ++                                               | ++                                                 | +++                | +++    | ++              | +++                                                      |
| Platelet rich plasma            | ++   | ++                                               | ++                                                 | +++                | +++    | ++              | +++                                                      |
| Ovarian transplantation         | +++  | +++                                              | +                                                  | +++                | +++    | ++              | +++                                                      |
| Artificial ovary                | +++  | +                                                | +                                                  | +                  | +++    | +               | +                                                        |
| Artificial gametes              | +++  | +                                                | +                                                  | +                  | +++    | +               | +                                                        |
| Mitochondrial replacement therapy| +++  | +                                                | +                                                  | +                  | +++    | +               | +                                                        |

+: very low; ++: low; +++: moderate; ++++: high; ++++++: very high.
reproducibility may accelerate drawing a safe conclusion on whether introducing these methods to the clinical routine practice set up would be beneficial or detrimental. However, understanding the outcome of such approaches presents one side of the coin. Distinguishing the profiles of patients who respond to certain approaches and to what extent these approaches restore their ovarian function remains to be delineated in this era of individualized medicine. The molecular mechanism involved during the implementation of the aforementioned novel treatments may only be unraveled by basic research studies that could hold the potential to conclusively address the question “for whom these novel approaches may perform efficiently.” While estimating the cost and effectiveness of the aforementioned approaches, one should not fail to consider the financial and psychological cost entailed—on an individualized basis—in addressing ovarian insufficiency when opting to employ what may be described as the conventional and traditional approach of oocyte donation. The field of medical practice has experienced the phenomenon of embarking on novel ideas solely on the grounds of limited-yet-promising data accompanying a novel technique covering the unchartered territory. It is important to be provided with published data conveying transparency to enable a fair and impartial assessment, as not all of them may be perceived as equally promising. Taking into account the fact that there is an increase in the cost of health care set up extending to infertility treatment, evidence-based medicine in the field of ART is in the spotlight of research aiming to a more collected and bullet proof approach to manage pathologies such as ovarian insufficiency.

There may not be a superior method waiting to be discovered in addressing the issue of premature ovarian insufficiency especially in light of patient heterogeneity. Albeit novel available options may be constantly emerging, extreme caution should be practiced as they may be introduced lacking the platform of solid and well-designed studies that should buttress and confirm their value. Embracing novel approaches and acknowledging what drives their emergence is of pivotal importance. Faced with the promise of what presents as the holy grail in treating ovarian insufficiency one should fine-tune the sense of balance between practicing enthusiasm, patience, and caution. The reproduction endocrinology and infertility world will benefit from definitively connecting the dots between cause and effect, and concurring on safety and effectiveness while practicing the golden ratio between the patients’ desire for alternative treatment and providing them with a last resort option—one not surrounded by uncertainty—in the world of evidence-based medicine.

**Acknowledgments**

The present study was conducted as part of the M.Sc. program “Research in Female Reproduction” of Medical School, National and Kapodistrian University of Athens.

**Authors’ Contributions**

K.S., G.M., and M.S. conceived and designed the project. A.R., S.G., D.R., E.M., P.T., and P.G. performed the literature review. K.S., A.R., S.G., D.R., E.M., P.T., and P.G. contributed to drafting and editing the manuscript. K.P., N.V., G.M., M.K., and M.S. supervised the study and revised the manuscript. All authors approved the final draft.

**Ethical Approval**

Our institution does not require ethical approval for reporting review articles.

**Statement of Human and Animal Rights**

This article does not contain any studies with human or animal subjects.

**Statement of Informed Consent**

There are no human subjects in this article and informed consent is not applicable.

**Declaration of Conflicting Interests**

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

**Funding**

The author(s) received no financial support for the research, authorship, and/or publication of this article.

**ORCID iD**

Mara Simopoulou @ https://orcid.org/0000-0002-1000-9100

**References**

1. Wang J, Sauer MV. In vitro fertilization (IVF): a review of 3 decades of clinical innovation and technological advancement. Ther Clin Risk Manag. 2006;2(4):355.
2. Marder W, Fisseha S, Ganser MA, Somers EC. Ovarian Damage during chemotherapy in autoimmune diseases: broad health implications beyond fertility. Clin Med Insights Reprod Health. 2012;6:9–18.
3. Torrealday S, Kodaman P, Pal L. Premature ovarian insufficiency - an update on recent advances in understanding and management. F1000Research. 2017 [cited 2019 Oct 19];6. Available from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5710309/
4. Rudnicka E, Kruszwewska J, Klicka K, Kowalczyk J, Grymowicz M, Skórska J, Pięta W, Smolareczky R. Premature ovarian insufficiency – aetiology, epidemiology, and diagnostic evaluation. Przegląd Menopauzalny Menopause Rev. 2018; 17(3):105–108.
5. Ubaldi FM, Cimadomo D, Vairelli A, Fabozzi G, Venturella R, Maggiulli R, Mazzilli R, Ferrero S, Palagiano A, Rienzi L. Advanced maternal age in IVF: still a challenge? The present and the future of its treatment. Front Endocrinol. 2019;10:94.
6. Herrera S, Buigues A, Díaz-García C, Romeu M, Martínez S, Gómez-Seguí I, Simón C, Hsueh AJ, Pellicer A. Fertility rescue and ovarian follicle growth promotion by bone marrow stem cell infusion. Fertil Steril. 2018;109(3):90–918.
7. Pantos K, Simpoulopou M, Pantou A, Rapani A, Tsioulou P, Nitsos N, Syrkos S, Pappas A, Koutsilieris M, Sfakianoudis K. A case series on natural conceptions resulting in ongoing pregnancies in menopausal and prematurely menopausal women following platelet-rich plasma treatment. Cell Transplant. 2019;28(9–10):1333–1340.

8. Simpoulopou M, Sfakianoudis K, Tsioulou P, Rapani A, Gian nelou P, Kiriakopoulos N, Pantou A, Vlahos N, Anfandis G, Bolaris S, Pantos K, et al. What will the future hold for artificial organs in the service of assisted reproduction: prospects and considerations. Front Med. 2019;13(6):627–638. [cited 2019 Oct 19]. Available from https://doi.org/10.1007/s11684-019-0697-58.

9. Cozzolino M, Marin D, Sisti G. New Frontiers in IVF: mtDNA and autologous germline mitochondrial energy transfer. Reprod Biol Endocrinol. 2019;17(1):55.

10. Labarta E, de Los Santos MJ, Herraiz S, Romeu M, Buigues A, Martínez S, Díaz GC, Gómez Cervelló I, Gil-Sanchis C, Santamaría X, Cabanillas S, Díaz A, Fazeli Z, Abedindo A, Omrani MD, Ghaderian SMH. 2019. Oct 19]. Available from https://doi.org/10.1007/s11684-019-0697-58.

11. Fazeli Z, Abedindo A, Omrani MD, Ghaderian SMH. Mesenchymal stem cells (MSCs) therapy for recovery of fertility: a systematic review. Stem Cell Rev Rep. 2018;14(1):1–12.

12. Cervelló I, Gil-Sanchis C, Santamaria X, Cabanillas S, Díaz A, Faus A, Pellicer A, Simón C. Human CD133(+) bone marrow-derived stem cells promote endometrial proliferation in a murine model of Asherman syndrome. Fertil Steril. 2015;104(6):1552–1560.

13. Servis M, Conte F, Messina D, Conte G, Masiello A, Ragni M, Capasso R, Capobianco S, Fazio R, Vitale G, et al. Therapeutic application of multipotent stem cells in ovarian transplantation: increase reproductive potential in patients who are poor responders. Fertil Steril. 2018;110(3):496–505.

14. Mirzaei H, Sadri NJ, Salehi H, Stenvang J, Masoudifar A, Mirzaei HR, Jaafari MR. Therapeutic application of multipotent stem cells. J Cell Physiol. 2018;233(4):2815–2823.

15. Liao SY, Tse HF. Multipotent (adult) and pluripotent stem cells for heart regeneration: what are the pros and cons? Stem Cell Res Ther. 2013;4(6):151.

16. Haynesworth SE, Baber MA, Caplan AI. Cytokine expression by human marrow-derived mesenchymal progenitor cells in vitro: effects of dexamethasone and IL-1 alpha. J Cell Physiol. 1996;166(3):585–592.

17. Caplan AI, Bruder SP. Mesenchymal stem cells: building blocks for molecular medicine in the 21st century. Trends Mol Med. 2001;7(6):259–264.

18. Doorn J, Moll G, Le BK, van BC, de BJ. Therapeutic applications of mesenchymal stromal cells: paracrine effects and potential improvements. Tissue Eng Part B Rev. 2012;18(2):101–115.

19. Gneccini M, He H, Noisieux N, Liang OD, Zhang L, Morello F, Mu H, Melo LG, Pratt RE, Ingwall JS, Dzau VJ. Evidence supporting paracrine hypothesis for Akt-modified mesenchymal stem cell-mediated cardiac protection and functional improvement. FASEB J. 2006;20(6):661–669.

20. Murphy MB, Moncivais K, Caplan AI. Mesenchymal stem cells: environmentally responsive therapeutics for regenerative medicine. Exp Mol Med. 2013;45:e54.

21. Liu J, Zhang H, Zhang Y, Li N, Wen Y, Cao F, Ai H, Xue X. Homing and restorative effects of bone marrow-derived mesenchymal stem cells on cisplatin injured ovaries in rats. Mol Cells. 2014;37(12):865–872.

22. Sanders JE, Hawley J, Levy W, Gooley T, Buckner CD, Deeg HJ, Doney K, Storb R, Sullivan K, Witherspoon R, Appelbaum FR. Pregnancies following high-dose cyclophosphamide with or without high-dose busulfan or total-body irradiation and bone marrow transplantation. Blood. 1996;87(7):3045–3052.

23. Salooja N, Chatterjee R, McMillan AC, Kelsey SM, Newland AC, Milligan DF, Franklin IM, Hutchinson RM, Linch DC, Goldstone AH. Successful pregnancies in women following single autotransplant for acute myeloid leukemia with a chemotherapy ablation protocol. Bone Marrow Transplant. 1994;13(4):431–435.

24. Salooja N, Szydlo RM, Socie G, Rio B, Chatterjee R, Ljungan m P, Van Lint MT, Powles R, Jackson G, Hinterberger FM, Kolb HJ. Late effects working party of the European group for blood and marrow transplantation. Pregnancy outcomes after peripheral blood or bone marrow transplantation: a retrospective survey. Lancet Lond Engl. 2001;358(9278):271–276.

25. Johnson J, Bagley J, Skaznik WM, Lee H-J, Adams GB, Nii kura Y, Tschudy KS, Tilly JC, Cortes ML, Forkert R, Spitzer T, et al. Oocyte generation in adult mammalian ovaries by putative germ cells in bone marrow and peripheral blood. Cell. 2005;122(2):303–315.

26. Telfer EE, Gosden RG, Byskov AG, Spears N, Albertini D, Andersen CY, Anderson R, Braw TR, Clarke H, Gougeon A, McLaughlin E, et al. On regenerating the ovary and generating controversy. Cell. 2005;122(6):821–822.

27. Eggan K, Jurga S, Gosden R, Min IM, Wagers AJ. Ovulated oocytes in adult mice derive from non-circulating germ cells. Nature. 2006;441(7097):1109–1114.

28. Veitia RA, Gluckman E, Fellous M, Soulier J. Recovery of female fertility after chemotherapy, irradiation, and bone mar row allograft: further evidence against massive oocyte regeneration by bone marrow-derived germline stem cells. Stem Cells. 2005;23(5):1334–1335.

29. Yoon SY. Mesenchymal stem cells for restoration of ovarian function. Clin Exp Reprod Med. 2019;46(1):1–7.

30. He Y, Chen D, Yang L, Hou Q, Ma H, Xu X. The therapeutic potential of bone marrow mesenchymal stem cells in premature ovarian failure. Stem Cell Res Ther. 2018;9(1):263.

31. Edessy M, Hosni HN, Shady Y, Waf Y, Bakr S, Kamel M. Autologous stem cells therapy, The first baby of idiopathic premature ovarian failure. Acta Medica Int. 2016;3(1):19.

32. Gabr H, Wael A, Ahmed EG. Autologous stem cell transplantation in patients with idiopathic premature ovarian failure. J Tissue Sci Eng. 2016;7(3).
33. Gupta S, Lodha P, Karthick MS, Tandulwadkar SR. Role of autologous bone marrow-derived stem cell therapy for follicular recruitment in premature ovarian insufficiency: review of literature and a case report of world’s first baby with ovarian autologous stem cell therapy in a perimenopausal woman of age 45 year. J Hum Reprod Sci. 2018;11(2):125–130.

34. Tauchmanová L, Selleri C, De RG, Sammartino A, Di CC, Musella T, Martorelli C, Lombardi G, Rotoli B, Nappi C, Colao A. Estrogen-progesterin therapy in women after stem cell transplant: our experience and literature review. Menopause N. Y. N. 2007;14(2):320–330.

35. Lee H-I, Selesniemi K, Niikura Y, Niikura T, Klein R, Dombkowski DM, Tilly JL. Bone marrow transplantation generates immature oocytes and rescues long-term fertility in a preclinical mouse model of chemotherapy-induced premature ovarian failure. J Clin Oncol Off J Am Soc Clin Oncol. 2007;25(22):3198–3204.

36. Abd ASH, Shalaby SM, Pasha HF, El SAS, Raafat N, Shabrawy SM, Awad HA, Amer MG, Gharib MA, El Gendi EA, Raslan AA, et al. Mechanistic action of mesenchymal stem cell injection in the treatment of chemically induced ovarian failure in rabbits. Cytotherapy. 2013;15(1):64–75.

37. Guo J, Gao X, Lin Z, Wu W, Huang L, Dong H, Chen J, Lu J, Fu Y, Wang J, Ma Y, et al. BMSCs reduce rat granulosa cell apoptosis induced by cisplatin and perimenopause. BMC Dev Biol. 2013;13:18.

38. Lai D, Wang F, Chen Y, Wang L, Wang Y, Cheng W. Human amniotic fluid stem cells have a potential to recover ovarian function in mice with chemotherapy-induced sterility. BMC Dev Biol. 2013;13:34.

39. Sun M, Wang S, Li Y, Yu L, Gu F, Wang C, Yao Y. Adipose-derived stem cells improved mouse ovary function after chemotherapy-induced ovary failure. Stem Cell Res Ther. 2013;4(4):80.

40. Lai D, Wang F, Yao X, Zhang Q, Wu X, Xiang C. Human endometrial mesenchymal stem cells restore ovarian function through improving the renewal of germline stem cells in a mouse model of premature ovarian failure. J Transl Med. 2015;13:155.

41. Wang K, Li Z, Li J, Liao W, Qin Y, Zhang N, Huo X, Mao N, Zhu H. Optimization of the platelet-rich plasma concentration for mesenchymal stem cell applications. Tissue Eng Part A. 2019;25(5–6):333–351.

42. Mohamed SA, Shalaby S, Brakta S, Elam L, Elsharoud A, Al Hendy A. Umbilical cord blood mesenchymal stem cells as an infertility treatment for chemotherapy-induced premature ovarian insufficiency. Biomedicines. 2019;7(1):7.

43. Sfakianoudis K, Simopoulou M, Nitsos N, Rapani A, Pappas A, Pantou A, Chronopoulou M, Deligeoroglou E, Koutsilieris M, Pantos K. Autologous platelet-rich plasma treatment enables pregnancy for a woman in premature menopause. J Clin Med. 2018;8(1):1. [cited 2019 Sep 17];8(1). Available from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6352170/.

44. Costa V, McGregor M, Laneuville P, Brophy JM. The cost-effectiveness of stem cell transplantsations from unrelated donors in adult patients with acute leukemia. Value Health J Int Soc Pharmacoeconomics Outcomes Res. 2007;10(4):247–255.

45. Ashfaq K, Yahaya I, Hyde C, Andronis L, Barton P, Baylissa S, Chen Y F. Clinical effectiveness and cost-effectiveness of stem cell transplantation in the management of acute leukaemia: a systematic review. NIHR Journals Library; 2010. Available from https://www.ncbi.nlm.nih.gov/books/NBK56809/.

46. Gurtner GC, Werner S, Barrandon Y, Longaker MT. Wound repair and regeneration. Nature. 2008;453(7193):314–321.

47. Dhurat R, Sukesh MS. Principles and methods of preparation of platelet-rich plasma: a review and author’s perspective. J Cutan Aesthetic Surg. 2014;7(4):189.

48. Sánchez GDJ, Méndez BE, Trejo BNI. Platelet-rich plasma Peptides: key for regeneration. [Internet] Int J Pept. 2012 [cited 2019 Sep 11]. Available from https://www.hindawi.com/journals/ijpep/2012/532519/.

49. Amable PR, Carias RBV, Teixeira MVT, da Cruz PÍ, Corrêa ARJF, Granjeiro JM, Borojevic R. Platelet-rich plasma preparation for regenerative medicine: optimization and quantification of cytokines and growth factors. Stem Cell Res Ther. 2013;4(3):67.

50. Szafarowska M, Jerzak M. [Ovarian aging and infertility]. Ginekol Pol. 2013;84(4):298–304.

51. Choi J, Minn KW, Chang H. The efficacy and safety of platelet-rich plasma and adipose-derived stem cells: an update. Arch Plast Surg. 2012;39(6):585–592.

52. Mlynarek RA, Kuhn AW, Bedi A. Platelet rich plasma (PRP) in orthopedic sports medicine. Am J Orthop Belle Mead NJ. 2016;45(5):290–326.

53. Patel AN, Selzaman CH, Kumpati GS, McKellar SH, Bull DA. Evaluation of autologous platelet rich plasma for cardiac surgery: outcome analysis of 2000 patients. J Cardiothorac.Surg. 2016;11(1):62.

54. Frautsch RS, Hashem AM, Halaba C, Cakmakoglu C, Zins JE. Current evidence for clinical efficacy of platelet rich plasma in aesthetic surgery: a systematic review. Aesthet Surg J. 2017;37(3):353–362.

55. Merchán WH, Gómez LA, Chasoy ME, Alfonso RCA, Muñoz AL. Platelet-rich plasma, a powerful tool in dermatology. J Tissue Eng Regen Med. 2019;13(5):892–901.

56. Duffy DM, Ko C, Jo M, Brannstrom M, Curry TE. Ovulation: parallels with inflammatory processes. Endoc Rev. 2019;40(2):369–416.

57. Danforth DR, Arbogast LK, Ghosh S, Dickerman A, Rofagha ARJF, Granjeiro JM, Borojevic R. Platelet-rich plasma preparation for regenerative medicine: optimization and quantification of cytokines and growth factors. Stem Cell Res Ther. 2013;4(3):67.

58. Szafarowska M, Jerzak M. [Ovarian aging and infertility]. Ginekol Pol. 2013;84(4):298–304.

59. Choi J, Minn KW, Chang H. The efficacy and safety of platelet-rich plasma and adipose-derived stem cells: an update. Arch Plast Surg. 2012;39(6):585–592.

60. Mlynarek RA, Kuhn AW, Bedi A. Platelet rich plasma (PRP) in orthopedic sports medicine. Am J Orthop Belle Mead NJ. 2016;45(5):290–326.

61. Patel AN, Selzaman CH, Kumpati GS, McKellar SH, Bull DA. Evaluation of autologous platelet rich plasma for cardiac surgery: outcome analysis of 2000 patients. J Cardiothorac. Surg. 2016;11(1):62.

62. Frautsch RS, Hashem AM, Halaba C, Cakmakoglu C, Zins JE. Current evidence for clinical efficacy of platelet rich plasma in aesthetic surgery: a systematic review. Aesthet Surg J. 2017;37(3):353–362.

63. Merchán WH, Gómez LA, Chasoy ME, Alfonso RCA, Muñoz AL. Platelet-rich plasma, a powerful tool in dermatology. J Tissue Eng Regen Med. 2019;13(5):892–901.

64. Duffy DM, Ko C, Jo M, Brannstrom M, Curry TE. Ovulation: parallels with inflammatory processes. Endoc Rev. 2019;40(2):369–416.

65. Danforth DR, Arbogast LK, Ghosh S, Dickerman A, Rofagha ARJF, Granjeiro JM, Borojevic R. Platelet-rich plasma preparation for regenerative medicine: optimization and quantification of cytokines and growth factors. Stem Cell Res Ther. 2013;4(3):67.

66. Szafarowska M, Jerzak M. [Ovarian aging and infertility]. Ginekol Pol. 2013;84(4):298–304.

67. Choi J, Minn KW, Chang H. The efficacy and safety of platelet-rich plasma and adipose-derived stem cells: an update. Arch Plast Surg. 2012;39(6):585–592.

68. Mlynarek RA, Kuhn AW, Bedi A. Platelet rich plasma (PRP) in orthopedic sports medicine. Am J Orthop Belle Mead NJ. 2016;45(5):290–326.

69. Patel AN, Selzaman CH, Kumpati GS, McKellar SH, Bull DA. Evaluation of autologous platelet rich plasma for cardiac surgery: outcome analysis of 2000 patients. J Cardiothorac. Surg. 2016;11(1):62.

70. Frautsch RS, Hashem AM, Halaba C, Cakmakoglu C, Zins JE. Current evidence for clinical efficacy of platelet rich plasma in aesthetic surgery: a systematic review. Aesthet Surg J. 2017;37(3):353–362.

71. Merchán WH, Gómez LA, Chasoy ME, Alfonso RCA, Muñoz AL. Platelet-rich plasma, a powerful tool in dermatology. J Tissue Eng Regen Med. 2019;13(5):892–901.
poor responder patients. Gynecol Obstet Invest. 2019;84(1): 99–106.
60. Sills ES, Rickers NS, Li X, Palermo GD. First data on in vitro fertilization and blastocyst formation after intraovarian injection of calcium gluconate-activated autologous platelet rich plasma. Gynecol Endocrinol. Off J Int Soc Gynecol Endocrinol. 2018;34(9):756–760.
61. Pantos K, Nitos N, Kokkali G, Vaxevanoglou T, Markomichali C, Pantou A, Grammatis M, Lazaros L, Sfakianoudis K. Ovarian rejuvenation and folliculogenesis reactivation in peri-menopausal women after autologous platelet-rich plasma treatment. Helsinki, Finland: Human Reproduction.
62. Farimani M, Heshmati S, Poorolajal J, Bahmanzadeh M. A report on three live births in women with poor ovarian response following intra-ovarian injection of platelet-rich plasma (PRP). Mol Biol Rep. 2019;46(2):1611–1616.
63. Sills ES, Li X, Rickers NS, Wood SH, Palermo GD. Metabolic and neurobehavioral response following intraovarian administration of autologous activated platelet rich plasma: first qualitative data. Neuro Endocrinol Lett. 2019;39(6):427–433.
64. Machado ES, Leite R, dos Santos CC, Artuso GL, Gluszczak F, de Jesus LG, Caldas JMP, Bredemeier M, Machado ES, Leite R, dos Santos CC, et al. Turn down - turn up: a simple and low-cost protocol for preparing platelet-rich plasma. Clinics. 2019 [cited 2019 Sep 11];74. Available from http://www.scielo.br/scielo.php?script=sci_abstract&pid=S1807-59322019000100258&lng=en&nrm=iso&tlng=en.
65. Martinez ZMJ, Martí CAJ, Solá I, Expósito JA, Bolíbar I, Rodríguez L, García J, Zaror C. Autologous platelet-rich plasma for treating chronic wounds. Cochrane Database Syst. Rev. 2016;25(5):CD006899. [cited 2019 Oct 20];(5). Available from https://www.cochranelibrary.com/cdsr/doi/10.1002/14651858.CD006899.pub3/abstract.
66. Sills ES, Wood SH. Autologous activated platelet-rich plasma injection into adult human ovary tissue: molecular mechanism, analysis, and discussion of reproductive response. Biosci Rep. 2019. [cited 2019 Oct 20];39(6). Available from https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6549090/.
67. Hanna CB, Hennebold JD. Ovarian germline stem cells: an unlimited source of oocytes?. Fertil Steril. 2014;101(1): 20–30.
68. Amorim CA, Leonel ECR, Afifi Y, Coomarasamy A, Fishel S. Cryostorage and retransplantation of ovarian tissue as an infertility treatment. Best Pract Res Clin Endocrinol Metab. 2019;33(1):89–102.
69. Silber S. Ovarian tissue cryopreservation and transplantation: scientific implications. J Assist Reprod Genet. 2016;33(12): 1595–1603.
70. Donnez J, Dolmans MM. Ovarian cortex transplantation: 60 reported live births brings the success and worldwide expansion of the technique towards routine clinical practice. J Assist Reprod Genet. 2015;32(8):1167–1170.
71. Demestere I, Simon P, Emiliani S, Delbaere A, Englert Y. Orthotopic and heterotopic ovarian tissue transplantation. Hum Reprod Update. 2009;15(6):649–665.
72. Kuo C Y, Baker H, Fries MH, Yoo JJ, Kim PCW, Fisher JP. Bioengineering strategies to treat female infertility. Tissue Eng Part B Rev. 2016;23(3):294–306.
73. Oktay K, Karlikaya G. Ovarian function after transplantation of frozen, banked autologous ovarian tissue. N Engl J Med. 2000;342(25):1919.
74. Donnez J, Dolmans MM, Demydle D, Jadoul P, Pirard C, Squifflet J, Martinez MB, van Langendonckt A. Livebirth after orthotopic transplantation of cryopreserved ovarian tissue. Lancet Lond Engl. 2004;364(9443):1405–1410.
75. Meirov D, Levron J, Eldar Geva T, Hardan I, Fridman E, Zalel Y, SchiﬁE, Dor J. Pregnancy after transplantation of cryopreserved ovarian tissue in a patient with ovarian failure after chemotherapy. N Engl J Med. 2005;353(3):318–321.
76. Silber SJ, Lenahan KM, Levine DJ, Pineda JA, Gorman KS, Friez MJ, Crawford EC, Gosden RG. Ovarian transplantation between monoyzotic twins discordant for premature ovarian failure. N Engl J Med. 2005;353(1):58–63.
77. Demestere I, Simon P, Emiliani S, Delbaere A, Englert Y. Fertility preservation: successful transplantation of cryopreserved ovarian tissue in a young patient previously treated for Hodgkin’s disease. The Oncologist. 2007;12(12):1437–1442.
78. Andersen CY, Rosendahl M, Byskov AG, Loy A, Ottosen C, Dueholm M, Schmidt KLT, Andersen AN, Ernst E. Two successful pregnancies following autotransplantation of frozen/thawed ovarian tissue. Hum Reprod Oxf. 2008;23(10): 2266–2272.
79. Kim SS. Assessment of long term endocrine function after transplantation of frozen-thawed human ovarian tissue to the heterotopic site: 10 year longitudinal follow-up study. J Assist Reprod Genet. 2012;29(6):489–493.
80. Atala A. Tissue engineering of reproductive tissues and organs. Fertil Steril. 2012;98(1):21–29.
81. Schubert B, Canis M, Darcha C, Artonne C, Smitj S, Grizard G. Follicular growth and estradiol follow-up after xenografting of fresh and cryopreserved human ovarian tissue. Fertil Steril. 2008;89(6):1787–1794.
82. Van EAS, Jordan BF, Gallez B, Heiiler JF, Van Langendonckt A, Donnez J. Electron paramagnetic resonance as a tool to evaluate human ovarian tissue reoxygenation after xenografting. Fertil Steril. 2009;92(1):374–381.
83. Kong HS, Lee J, Youm HW, Kim SK, Lee JR, Suh CS, Kim SH. Effect of treatment with angioipoietin-2 and vascular endothelial growth factor on the quality of xenografted bovine ovarian tissue in mice. PLoS One. 2017;12(9):e0184546.
84. Donnez J, Dolmans MM. Natural hormone replacement therapy with a functioning ovary after the menopause: dream or reality? Reprod Biomed Online. 2018;37(3):359–366.
85. Pangas SA, Saudye H, Shea LD, Woodruff TK. Novel approach for the three-dimensional culture of granulosal cell-oocyte complexes. Tissue Eng. 2003;9(5):1013–1021.
86. Krotz SP, Morgan JR, Carson S. In vitro maturation of oocytes via the pre-fabricated self-assembled artificial human ovary. J Assist Reprod Genet. 2010;27(12):743–750.
87. Kim S, Lee Y, Lee S, Kim T. Ovarian tissue cryopreservation and transplantation in patients with cancer. Obstet Gynecol Sci. 2018;61(4):431–442.

88. Hendriks S, Dancet EAF, van Pelt AMM, Hamer G, Repping S. Artificial gametes: a systematic review of biological progress towards clinical application. Hum Reprod Update. 2015;21(3):285–296.

89. Moreno I, Míguez FJM, Simón C. Artificial gametes from stem cells. Clin Exp Reprod Med. 2015;42(2):33–44.

90. White YAR, Woods DC, Takai Y, Ishihara O, Seki H, Tilly JL. Oocyte formation by mitotically active germ cells purified from oocytes of reproductive-age women. Nat Med. 2012;18(3):413–421.

91. Aflatoonian B, Ruban L, Jones M, Aflatoonian R, Fazeli A, Moore HD. In vitro post-meiotic germ cell development from human embryonic stem cells. Hum Reprod Oxf Engl. 2009;24(12):3150–3159.

92. Cheng X, Chen S, Yu X, Zheng P, Wang H. BMP15 gene is activated during human amniotic fluid stem cell development into oocyte-like cells. DNA Cell Biol. 2012;31(7):1198–1204.

93. Ma Z, Liu R, Wang X, Huang M, Gao Q, Lu Y, Liu C. Spontaneous germline potential of human hepatic cell line in vitro. Mol Hum Reprod. 2013;19(4):216–226.

94. Hendriks S, Dondorp W, de Wert G, Hamer G, Repping S, Dancet EAF. Potential consequences of clinical application of artificial gametes: a systematic review of stakeholder views. Hum Reprod Update. 2015;21(3):297–309.

95. Segers S, Mertes H, de Wert G, Dondorp W, Pennings G. Balancing ethical pros and cons of stem cell derived gametes. Ann Biomed Eng. 2017;45(7):1620–1632.

96. Jansen RP, de Boer K. The bottleneck: mitochondrial imperatives in oogenesis and ovarian follicular fate. Mol Cell Endocrinol. 1998;145(1–2):81–88.

97. May PP, Chrétien MF, Jacques C, Vasseur C, Malthiery Y, Reynier P. Low oocyte mitochondrial DNA content in ovarian insufficiency. Hum Reprod Oxf Engl. 2005;20(3):593–597.

98. Selè E. Mitochondrial DNA as a biomarker for in-vitro fertilization outcome. Curr Opin Obstet Gynecol. 2016;28(3):158–163.

99. Victor AR, Brake AJ, Tyndall JC, Griffith DK, Zouves CG, Barnes FL, Viotti M. Accurate quantitation of mitochondrial DNA reveals uniform levels in human blastocysts irrespective of ploidy, age, or implantation potential. Fertil Steril. 2017;107(1):34–42.

100. Grindler NM, Moley KH. Maternal obesity, infertility and mitochondrial dysfunction: potential mechanisms emerging from mouse model systems. Mol Hum Reprod. 2013;19(8):486–494.

101. Cohen J, Scott R, Alikani M, Schimmel T, Munné S, Levron J, Wu L, Brenner C, Warner C, Willadsen S. Ooplasmic transfer in mature human oocytes. Mol Hum Reprod. 1998;4(3):269–280.

102. Barritt JA, Brenner CA, Malter HE, Cohen J. Mitochondria in human offspring derived from ooplasmic transplantation. Hum Reprod Oxf Engl. 2001;16(3):513–516.

103. Fakih MH, Shmoury ME, Szeptycki J, Cruz DB dela, Lux C de G, Verjee S, Burgess CM, Cohn GM, Casper R. The AUGMENT SM Treatment: physician reported outcomes of the initial global patient experience. 2015.

104. Oktay K, Baltaci V, Sonmez E, Turan V, Unsal E, Baltaci A, Aktuna S, Moy F. Oogonial precursor cell-derived autonomous mitochondria injection to improve outcomes in women with multiple IVF failures due to low oocyte quality: a clinical translation. Reprod Sci Thousand Oaks Calif. 2015;22(12):1612–1617.

105. Greenfield A, Braude P, Flinter F, Lovell BR, Ogilvie C, Perry ACf. Assisted reproductive technologies to prevent human mitochondrial disease transmission. Nat Biotechnol. 2017;35(11):1059–1068.

106. Herbert M, Turnbull D. Progress in mitochondrial replacement therapies. Nat Rev Mol Cell Biol. 2018;19(2):71–72.

107. Tachibana M, Kuno T, Yaegashi N. Mitochondrial replacement therapy and assisted reproductive technology: a paradigm shift toward treatment of genetic diseases in gametes or in early embryos. Reprod Med Biol. 2018;17(4):421–433.

108. Labarta E, de Los Santos MJ, Escribá MJ, Pellicer A, Herraz S. Mitochondria as a tool for oocyte rejuvenation. Fertil Steril. 2019;111(2):219–226.

109. Hyslop LA, Blakeley P, Craven L, Richardson J, Fogarty NME, Fragouli E, Lamb M, Wamaitha SE, Prathalingam N, Zhang Q, O’Keefe H, et al. Towards clinical application of pronuclear transfer to prevent mitochondrial DNA disease. Nature. 2016;534(7607):383–386.

110. Ma H, O’Neil RC, Martí GN, Hariraharan M, Zhang ZZ, He Y, Cinnioglu C, Kayali R, Kang E, Lee Y, Hayama T, et al. Functional human oocytes generated by transfer of polar body genomes. Cell Stem Cell. 2017;20(1):112–119.

111. Zhang J, Zhuang G, Geng Y, Grifo J, Acosta C, Shu Y, Liu H. Pregnancy derived from human zygote pronuclear transfer for a patient who had arrested embryos after IVF. Reprod Biomed Online. 2016;33(4):529–533.

112. Zhang J, Liu H, Luo S, Chavez BA, Liu Z, Yang M, Munné S, Konstantinidis M, Wells D, Huang T. First live birth using human oocytes reconstituted by spindle nuclear transfer for mitochondrial DNA mutation causing Leigh syndrome. Fertil Steril. 2016;106(3):e375–e376.

113. Zhang J, Liu H, Luo S, Chávez BA, Liu Z, Yang M, Merhi Z, Silber SJ, Munné S, Konstantinidis M, et al. Live birth derived from oocyte spindle transfer to prevent mitochondrial disease. Reprod Biomed Online. 2017;34(4):361–368.

114. Reardon S. Genetic details of controversial “three-parent baby” revealed. Nat News. 2017;544(7648):17.

115. ISRCTN - ISRCTN11455145: Spindle transfer for the treatment of infertility problems associated to poor egg quality: a pilot trial. [Internet] [cited 2019 Oct 20]. Available from http://www.isrctn.com/ISRCTN11455145.

116. Welcome. [Internet] [cited 2019 Oct 20]. Available from https://www.eshre.eu/Press-Room/ESHRE-News/target Text=–Spindle%20transfer%20in%20the%20treatment%20of%20infertility%3A%20an%20ESHRE%20Statement}
This treatment, also known as, aim of correcting cytoplasmic disorders.

117. Eyre WA. Mitochondrial Replacement Therapy: are mitochondrial interactions likely to be a problem? Genetics. 2017;205(4):1365–1372.

118. Dobler R, Dowling DK, Morrow EH, Reinhardt K. A systematic review and meta-analysis reveals pervasive effects of germline mitochondrial replacement on components of health. Hum Reprod Update. 2018;24(5):519–534.

119. Eyre WA. Mitochondrial replacement and its effects on human health. Hum Reprod Update. 2019;25(3):392–394.

120. Simopoulou M, Sfakianoudis K, Giannelou P, Pierouli A, Rapani A, Maziotis E, Galatis D, Bakas P, Vlahos N, Pantos K, Koutsilieris M. Treating infertility: current affairs of cross-border reproductive care. Open Med Wars Pol. 2019;14:292–299.

121. Practice committees of American society for reproductive medicine, society for assisted reproductive technology. Mature oocyte cryopreservation: a guideline. Fertil Steril. 2013;99(1):37–43.

122. Campbell KA, Saltzman BM, Mascarenhas R, Khair MM, Verma NN, Bach BR, Cole BJ. Does intra-articular platelet-rich plasma injection provide clinically superior outcomes compared with other therapies in the treatment of knee osteoarthritis? A systematic review of overlapping meta-analyses. Arthroscopy. 2015;31(11):2213–2221.