Non-typhoidal *Salmonella* (NTS) is an important pathogen that causes gastroenteritis, bacteraemia, and focal infections. Herein, we present our experience with bloodstream infections caused by *Salmonella* in paediatric leukaemia patients, which has been reported for the first time in both Europe and the US. According to our research, NTS might be a cause of serious infections in paediatric haematology-oncology patients. Following a low bacterial diet and increasing the hygiene of both the children and their surroundings would be beneficial in preventing these infections.

**Key words:** acute lymphoblastic leukaemia, non-typhoidal salmonellosis, neutropaenia.

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**Non-typhoidal *Salmonella* bacteraemia in paediatric leukaemia patients**

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**Introduction**

Infections are the leading cause of morbidity and mortality in paediatric cancer patients. Paediatric cancer patients have an increased tendency for infections due to the effect of specific chemotherapeutics, their neutropaenic state, and underlying malignant disease [1]. Bloodstream infections (BSI) form the majority of these infections. In children with leukaemia, only 30–40% of the microbial causes of infections have been identified microbiologically [1, 2]. The distribution of BSIs may change from one centre to another. Some authors have reported the predominance of Gram-negative bacteria along with Gram-positive bacteria [1, 3]. The most commonly isolated Gram-negative pathogens are *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, and *Escherichia coli* [1, 3]. The *Salmonella* species have rarely been reported in English literature as a causative agent of febrile neutropaenia in children.

Herein, we present our experience with bloodstream infections caused by *Salmonella* in paediatric leukaemia patients, which has been reported for the first time in both Europe and the US.

**Material and methods**

This retrospective study was designed to evaluate *Salmonella* bacteraemia cases in febrile patients who were hospitalised at the Paediatric Haematology-Oncology and Infectious Disease units of Dr. Behçet Uz Children’s Hospital for treatment of malignancies from December 2008 to February 2014. The medical records and computerised microbiology laboratory records of 162 children with acute lymphoblastic and acute myeloblastic leukaemia who were hospitalised at the Department of Paediatric Haematology-Oncology and Infectious Disease in Dr. Behçet Uz Children’s Hospital were reviewed to identify patients who were diagnosed with salmonellosis.

Data on demographics, chemotherapy, symptoms, details of treatment, culture results, antimicrobial sensitivity data of the cultures, delay of chemotherapy, and outcome were recorded. Empiric therapy with broad-spectrum antibiotics was initiated upon the presence of fever (defined as a temperature ≥ 38°C on two occasions within a 12-hour period or a single measurement ≥ 38.3°C) during neutropaenia. Two blood cultures, one from peripheral veins and one through ports, were taken according to our infection control procedure from each patient who presented with a fever. Blood samples were aseptically collected for blood cultures in paediatric blood culture bottles (BacT/Alert PF – Biomerieux-France) from all the study patients. Each bottle contained 1–3 ml of blood and was incubated at 37°C for up to seven days. Once a positive culture bottle was detected, a Gram stain slide was prepared from the bot-
tle along with a subculture of a loopful of the positive blood culture bottle contents on blood agar, EMB agar, Chocolate agar, and Sabouraud agar (Salubris-Turkey) followed by incubation at 37°C for 18–24 hours. If non-lactose fermenting colonies were detected, then biochemical identification tests were performed using a VITEK II automated system (Biomerieux-France). Final identification of isolates was confirmed serologically according to the Kauffman-White classification using “RSHM” Salmonella antisera (Turkey). Antibiotic susceptibility testing was performed on pure culture using the broth dilution method by VITEK II automated microbiology system. The following antibiotics were tested: amikacin, ampicillin, gentamicin, meropenem, netilmicin, piperacillin, cefepime, cefoxitin, ceftriaxone, cefuroxime, ciprofloxacin, amoxicillin-clavulanate, imipenem, trimethoprim/sulfamethoxazole, and tigecycline. The results were interpreted using the Clinical and Laboratory Standards Institute (CLSI) standards. Sensitivity results were then reported as sensitive or resistant based on the CLSI criteria [4].

Results

Between October 2008 and February 2014, seven children with acute leukaemia had suffered from non-typoidal Salmonella (NTS)-associated bacteraemia. Among them four had associated positive stool cultures. Six of the patients were neutropaenic (85.7%), while one was not. The demographic/clinical features are reviewed in Table 1. The median age was four years (ranging from two years to 10 years). Six patients (85.7%) were male, while one (14.3%) was a female. All but one of the children, in whom the family refused treatment, was receiving chemotherapy during the course of the NTS infection. Among the six other patients, two were receiving induction chemotherapy, one was receiving reinduction therapy, and three were on maintenance chemotherapy according to the ALL-BFM protocol (Table 1).

All of the children with NTS bacteraemia had associated diarrhoea. They had all been hospitalised and received broad-spectrum antibiotics after samples for blood cultures were obtained (Table 1). Modification of antibiotics was done in four of the patients after blood culture or clinical deterioration. All of the patients recovered from NTS BSI without any complications including typhlitis or surgical complications. However, five of the patients had their chemotherapy delayed for at least seven days (ranging from seven to 18 days).

The Salmonella species isolated from the patients were as follows: Salmonella spp. (3), Salmonella group D (2) and Salmonella enterica ssp. arizonae (2). All the NTS species were susceptible to ceftriaxone and fluoroquinolones. The resistance to individual antimicrobials was found at vary-

Table 1. Demographic/clinical characteristics of the patients

| Age (years) | Gender | Primary diagnosis | Treatment phase at time of infection | Total leucocyte count (number/mm³) | Absolut neutrophil count (number/mm³) | Complaints       | Antibiotics usage (days) | Modification of antibiotics (days) | Delay of chemotherapy (days) | Culture results          | Culture obtained from                                      |
|-------------|--------|------------------|--------------------------------------|------------------------------------|--------------------------------------|-----------------|--------------------------|-------------------------------|-----------------------------|---------------------------|------------------------------------------------------------|
| 2           | F      | B-ALL SRG        | Induction                            | 65,000                             | 1,550                                | Diarrhoea       | Pip-taz (10)             | NR                           | None                        | Salmonella spp.          | Blood, portal cath                                         |
| 3           | M      | B-ALL SRG        | Maintenance                          | 680                                | 60                                   | Fever, diarrhoea, artralgia            | Pip-taz (14) | Metronid (7)             | Imipenem (5)         | 7                          | Salmonella spp.          | Blood, stool                                               |
| 10          | M      | B-ALL 3rd relapse| Refractory disease                   | 11,800                             | 880                                  | Fever, diarrhoea, vomiting             | Pip-taz (14) | NR                       | Not under treat.       | 7                          | Salmonella group D | Blood, portal cath                                         |
| 4           | M      | B-ALL SRG        | Maintenance                          | 680                                | 60                                   | Fever, diarrhoea                        | Pip-taz (7)  | NR                       | 7                          | Salmonella enterica ssp. arizonae | Blood, stool                                               |
| 4           | M      | B-ALL HRG        | Reinduction                          | 880                                | 50                                   | Fever, diarrhoea                        | Pip-taz (8) | Metronid (7)             | Meronem (10d)         | 18                         | Salmonella enterica ssp. arizonae | Blood, stool                                               |
| 9           | M      | B-All relapse    | Induction                            | 100                                | 20                                   | Fever, diarrhoea                        | Meropenem (10) | Metronid (7)             | Vanco (10d)          | 15                         | Salmonella group D | Blood, portal cath                                         |
| 10          | M      | B-ALL 2nd relapse| Waiting for BMT, maintenance         | 900                                | 300                                  | Fever, diarrhoea, artralgia             | Pip-taz (10) | Amikacin (10)           | Meronem (21)         | 14                         | Salmonella spp.          | Blood, stool                                               |

B-ALL – B-cell acute lymphoblastic leukaemia, SRG – standard-risk group, HRG – high-risk group, BMT – bone marrow transplant, Pip-taz – piperacillin-tazobactam, Metronid – metronidazole, Vanco – vancomycin, NR – not required, treat. – treatment
ing rates: ampicillin in two isolates (28.5%), nitrofurantoin in two isolates (28.5%), and amikacin in one isolate (14.5%).

Discussion

Non-typhoidal Salmonella species mainly cause self-limiting enterocolitis in immunocompetent individuals [5]. However, approximately 5% of these patients were reported to develop secondary bacteraemia, which was associated with a low mortality ratio, ranging between 1 and 5% [5, 6]. The risk of invasive infections with non-typhoidal species was generally associated with inherited or acquired immunodeficiency syndromes. In one study invasive disease was reported to occur in up to 47% of immunocompromised infants in Africa [7].

Since oncology-haematology patients were under intensive chemotherapy, the paediatric cancer patients were supposed to have invasive infections due to NTS. However, there are only two articles present in the English literature in which infections due to Salmonella species are reported. In one case series from Pakistan, Salmonella paratyphi B was reported as the most commonly isolated organism in paediatric neutropaenic febrile children [3]. Moreover, in one case report from India; a five-year-old male child who had developed Salmonella typhi arthritis of his left hip during neutropaenic phase was presented [8]. In one study from Spain, among 29 cases of NTS bacteraemia, three children with malignancy were present [9]. In one study from Poland, among a total of 30 adult patients with NTS, 12 of the patients had a malignancy [10]. Two of the patients with malignancy died due to NTS infection. To our knowledge this is the first case series of NTS as a causative agent of bacteraemia in paediatric malignancy patients. The transmission of NTS generally happens when people eat contaminated foods of animal origin such as meat or eggs. They can also be infected by ingesting organisms in animal faeces, either directly or in contaminated food or water. According to our observations, our patients and their parents did not strictly comply with a low-bacteria diet and generally preferred consuming foods from local traditional restaurants. Although for now there is no evidence for the benefits of low-bacteria diet in children and adults for the prevention of infection and related complications, our experience with NTS infections supports the assumption that a low-bacteria diet would be beneficial in preventing food-borne infections in neutropaenic patients [11]. The definition of a neutropaenic or low-bacteria diet is controversial; however, commonly prohibited foods are raw fresh fruits and vegetables. To reduce the risk of food-borne infections in neutropaenic patients and their surroundings would be beneficial in preventing infections in neutropaenic children with these kinds of pathogens.

The authors declare no conflict of interest.

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