Auto-antibodies and their association with clinical findings in women diagnosed with microscopic colitis.

Roth, Bodil; Gustafsson, Rita; Ohlsson, Bodil

Published in: PLoS ONE

DOI: 10.1371/journal.pone.0066088

2013

Link to publication

Citation for published version (APA):
Roth, B., Gustafsson, R., & Ohlsson, B. (2013). Auto-antibodies and their association with clinical findings in women diagnosed with microscopic colitis. PLoS ONE, 8(6), [e66088].
https://doi.org/10.1371/journal.pone.0066088

General rights
Unless other specific re-use rights are stated the following general rights apply:
Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.
- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

Read more about Creative commons licenses: https://creativecommons.org/licenses/

Take down policy
If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.
Auto-Antibodies and Their Association with Clinical Findings in Women Diagnosed with Microscopic Colitis

Bodil Roth*, Rita J. Gustafsson†, Bodil Ohlsson*

Department of Clinical Sciences, Department of Gastroenterology, Skåne University Hospital, Malmö, Lund University, Lund, Sweden

Abstract

Background: Microscopic colitis (MC) is a disease manifested by diarrhoea and is divided into collagenous and lymphocytic colitis. The aetiology is unknown, but auto-immunity is suggested. Auto-antibodies have been only rarely examined in this entity. The aim of the study was to examine the prevalence of auto-antibodies, and to examine associations between the presence of antibodies and clinical findings.

Methods and Findings: Women with MC verified by biopsy and younger than 73 years, at any Department of Gastroenterology, in the district of Skåne, between 2002 and 2010 were invited to participate in this study. The patients were asked to complete both a questionnaire describing their medical history and the Gastrointestinal Symptom Rating Scale (GSRS). Blood samples were collected. Anti-nuclear antibodies (ANA), anti-neutrophil cytoplasmic antibodies (ANCA), anti-Saccharomyces cerevisiae cerevisiae antibodies (ASCA), and antibodies against glutamic acid decarboxylase (anti-GAD), islet antigens-like insulin 2 (anti-IA2), thyroid peroxidase (anti-TPO), and thyrotropin receptor (TRAK) were analysed. Of 240 women identified, 133 were finally included in the study, median age 63 (59–67) years. Apart from the MC diagnosis, 52% also suffered from irritable bowel syndrome, 31% from hypertension and 31% from allergy. The prevalence of ANA (14%), ASCA IgG (13%), and anti-TPO antibodies (14%) for these patients was slightly higher than for the general population, and were found together with other concomitant diseases. Patients had more of all gastrointestinal symptoms compared with norm values, irrespective of antibody expression.

Conclusions: Women with MC have a slightly increased prevalence of some auto-antibodies. These antibodies are not associated with symptoms, but are expressed in patients with concomitant diseases, obscuring the pathophysiology and clinical picture of MC.

Introduction

Microscopic colitis (MC) is a disease with watery diarrhoea without endoscopically inflamed colonic mucosa, and is divided into two different entities, collagenous colitis (CC) and lymphocytic colitis (LC). The diagnostic criterion for LC is >20 intraepithelial lymphocytes/100 enterocytes, reactive surface epithelium and mixed inflammatory infiltrate in the lamina propria. When also the subepithelial collagen band is >10 μm thick, the diagnosis CC is set [1]. The aetiology is unknown, but an auto-immune process has been proposed due to the responsiveness to corticosteroids, and a high frequency of a HLA haplotype and TNF alpha gene polymorphism (-308) associated with susceptibility to several auto-immune diseases [2]. Furthermore, other auto-immune diseases are found in 40% of these patients, thyroid diseases, rheumatologic diseases, diabetes mellitus, and coeliac disease being the most common [3,4]. Microscopic colitis may be a different entity in younger than in older patients, where drug treatment may be a considerable aetiology of the colitis, and therefore should in these cases rather be classified as a secondary disease [1].

A high prevalence of several auto-antibodies is found in patients with auto-immune diseases [5,6]. Antibodies against anti-neutrophil cytoplasmic antibodies (ANCA) are found in 40%-70% of patients with ulcerative colitis, and anti-Saccharomyces cerevisiae cerevisiae antibodies (ASCA) are found in 30%-70% of patients with Crohn’s disease [7,8]. Although MC is categorised as an inflammatory bowel disease (IBD) of auto-immune origin [9], only a few, small studies have been performed to examine the prevalence of auto-antibodies in this entity. No increased levels of rheumatoid factor or antibodies against thyroglobulin, microsomal antigen, endomysium and transglutaminase were found [3,4]. In CC, a tendency to increased anti-nuclear antibodies (ANA) was seen in one study, whereas increased levels of ANA, ANCA, and ASCA were seen in others [3,4,7]. One confounding factor may be that the level of auto-antibodies correlates with disease activity, and high values may only be detected in the active disease [10].

Type 1 diabetes mellitus is an auto-immune disease associated with MC [3,4]. Auto-antibodies that develop against islet antigens-like insulin 2 (IA2) and glutamic acid decarboxylase (GAD) are the...
markers of the disease, and are present in 70%–80% of cases [11,12]. In the majority of diabetes cases, immune reaction against islet antigens and consequent formation of auto-antibodies begins long before the disease is diagnosed clinically [13]. The prevalence of these antibodies in MC has been only little examined.

The aim of this study was to further examine the prevalence of auto-antibodies in a larger cohort of MC patients, and if present, to examine the association between the presence of antibodies to concomitant diseases and clinical findings.

Methods

Ethics Statement

The study protocol was approved by the Ethics Committee of Lund University, and all participants gave their written, informed consent when taking part in the study (LU 2009/565 and 2011/209).

Patients

Women who had been treated for MC at any outpatient clinic of the Departments of Gastroenterology, throughout the district of Skåne, between 2002 and 2010, were identified by a search for the ICD-10 classification for CC and LC (K52.8) in outpatient records and the local register at the Department of Pathology, Skåne University Hospital, Malmö. About one-third of the total number of the cases identified was not invited due to their being over 73 years of age, because they had many other concomitant diseases and drug therapies, which could obscure the picture with several cases of secondary MC [1]. Only the 240 patients (median 63 years, range 22–73 years) who had the diagnoses verified by colonic biopsy, and were 73 years of age or younger, were invited to participate in the present study. Altogether, 159 (median 63 years, range 22–73 years) of the 240 patients invited (66%) were recruited to the study and 134 patients (56%) also agreed to provide blood samples. One patient was excluded due to another IBD diagnosis a few weeks after the inclusion, leaving 158 (66%) to be included in the final calculations and laboratory analyses. These patients represent the majority of female cases of diagnosed MC in the southernmost districts of Sweden, younger than 73 years of age. As MC and auto-immunity are more frequent in women than in men [9,14], and as the health-related quality of life and the experience of symptoms differ between the genders [15], we chose to include only women in the study.

Patient Recruitment and Study Design

Between March and June 2011, invitations, including information on the study and questionnaires about social and life style factors, gastrointestinal symptoms, and medical condition and medication were sent by mail to all 240 women. A reminding letter was sent a month after the invitation letter to those who had not answered. They were also invited to provide blood samples for routine analyses according to standardised methods at the Division of Clinical Chemistry, Department of Laboratory Medicine in Skåne, at their local hospital. Analyses for IgG auto-antibodies were performed. ANA, ANCA, and ASCA were analysed at the Division of Immunology, Department of Laboratory Medicine in Skåne, Lund, and analyses of anti-IA2, anti-GAD and anti-TPO antibodies, and TRAK were performed at the Division of Clinical Chemistry, Department of Laboratory Medicine in Skåne, Malmö.

Medical records were scrutinised, and age, gastrointestinal symptoms, examinations, and treatments were recorded. Patients were divided into two groups. One group included patients with at least two episodes of watery diarrhoea; and/or those who were dependent on long-term treatment of corticosteroids to maintain remission; and/or those who had had two pathological intestinal mucosa biopsies (MC1, n = 73). The other group included patients who had concomitant coeliac disease [11], had undergone an acute gastroenteritis shortly before the diagnostic colonoscopy [4], had had only one episode of severe diarrhoea, or had had a normal biopsy after the initial pathological intestinal biopsy, and were in clinical remission (MC2, n = 60).

Immunological Analyses

Healthy blood donors served as controls for all immunological analyses, except for TRAK, for which control values in healthy controls are available by the same method [16]. Cut-off levels and prevalence figures in the healthy population are the reference values at the Department of Laboratory Medicine in Skåne after in-house examinations. Our reference values for anti-GAD and anti-IA2 antibodies have been published previously [17].

ANA and ANCA were analysed by indirect immunofluorescence (IIF) [18,19]. The cut-off value for ANA, >14 IU/ml, is set at the dilution where 5% of blood donors (50 men and 50 women) showed a positive ANA. The assay is calibrated against the international WHO reference for homogeneous ANA. Cytoplasmic (C)-and peri-nuclear (P)-ANCA have reference values of >10 IU/ml. The cut-off is determined by the supplier (Euro Immune AG, Lübeck, Germany) and of 50 healthy blood donors tested, none showed a positive result. ANA and ANCA were analysed in accordance with the International Organisation for Standardisation (ISO)/International Electrotechnical Commission (IEC) 17025 (General requirements for the competence for testing and calibration laboratories).

ASCA is analysed by an automated ELISA method for which the reference value is set to >10 IU/ml in accordance with the manufacturer’s recommendation (Orgentec Diagnostika, Alegria®, Mainz, Germany), and both IgA and IgG levels are given. Of 50 healthy blood donors tested at our laboratory, 8% were positive for both ASCA IgA and IgG, whereas other studies have found 0.6%–3.1% positive among healthy blood donors by the same method [20].

Antibodies against IA2 and GAD were analysed with a commercial ELISA kit (RSR Ltd, Cardiff, UK) according to the manufacturer’s manual for which the cut-off values are defined as >15 and >11 kIU/L, respectively [12]. In a cohort of 120 healthy Swedish women, median age 32 years, range 19–44 years, none was positive for anti-GAD antibodies and 31% were positive for anti-IA2 antibodies [17].

Anti-TPO antibodies are analysed by a chemiluminescence enzyme immunological method (Roche Diagnostics Limited, cobas®, West Sussex, UK) and TRAK by an RIA method (Thermo B.R.A.H.M.S., TRAK Human RIA) [16]. Anti-TPO antibodies are found in 7% of blood donors when the cut-off level is set to >70 kIU/L [21]. The reference value for TRAK with a sensitivity of 98.8% is set to >1.5 IU/L, and <1% of 282 healthy individuals of median age 45 years, range 20–73 years, were positive [16].

Questionnaires

A self-administered questionnaire about marital status, education, employment, smoking habits, wine consumption, physical activity, medical conditions and medication was completed by the patients. The patients were asked the questions: “Have you ever been treated for any of the following diseases, namely, hypertension, allergy, thyroid disorders, fracture, rheumatoid arthritis, asthma, gastric ulcer, ovari al inflammation, herpes infection,
endometriosis, Chlamydia Trachomatis, kidney stones, coeliac disease, diabetes mellitus type 1 or 2, myocardial infarction, intermittent claudication or stroke. A list of the drugs, vitamins and minerals currently being taken was completed. The participants were also asked about heredity for cancer, stroke, myocardial infarction, hypertension, diabetes mellitus, thyroid disorders, asthma, rheumatoid arthritis, IBD or coeliac disease.

Gastrointestinal symptom rating scale (GSRS). The GSRS is a Swedish, disease-specific and self-administered questionnaire, designed to evaluate perceived severity of gastrointestinal symptoms during the previous week. The questionnaire includes 15 items and uses a 7-grade Likert scale. This gives a total range value between 15 and 105 where the highest score (seven) represents the most pronounced symptoms and the lowest (one) no symptoms. The items are divided into five dimensions representing Reflux Syndrome (two items), Abdominal Pain Syndrome (three items), Constipation Syndrome (three items), Indigestion Syndrome (four items) and Diarrhoea Syndrome (three items). The data are presented as the average of the total score divided by the number of items. Norm values for the healthy, gender-matched population are available.

Rome III criteria. The patients completed an abbreviated version of the Rome III questionnaire, including only irritable bowel syndrome (IBS) symptoms. This questionnaire has been translated and validated into the Swedish language (Magnus Simrén and Anna Ryden). Patients who fulfilled the criteria in the Rome III questionnaire were classified as also suffering from IBS-like symptoms.

Table 1. Prevalence of auto-antibodies in microscopic colitis.

|                        | Controls | Microscopic colitis | Collagenous colitis | Lymphocytic colitis | P- value |
|------------------------|----------|---------------------|---------------------|---------------------|----------|
| **Hb (117–153 g/L)**   | n = 133  | 132.2 ± 9.3         | 132.2 ± 9.5         | 132.2 ± 9.0         | 0.987    |
| **Leucocytes (3.5–8.8×10^9/L)** |          | 6.9 ± 2.0           | 7.1 ± 1.9           | 6.8 ± 2.1           | 0.430    |
| **S-Albumin (36–48 g/L)** |          | 39.9 ± 3.3          | 39.9 ± 3.6          | 40.0 ± 3.0          | 0.794    |
| **CRP (<3.1 mg/L)**    |          | 2.0 (1.0–4.0)       | 4.0 (1.0–4.0)       | 1.6 (1.0–2.9)       | 0.014    |
| **P-Sodium (137–145 mmol/L)** |          | 140.0               | 141.0               | 140.0               | 0.111    |
| **P-Potassium (3.5–4.4 mmol/L)** |          | 3.9 ± 0.4           | 3.9 ± 0.3           | 3.9 ± 0.4           | 0.916    |
| **ANA (≤14 IU/ml) (n, %)** |          | (5)^a               | 19 (14)             | 8 (10)              | 11 (20)  | 0.140    |
| **C-ANCA (≤10 IU/ml) (n, %)** |          | (0)^b               | 1 (1)               | 1 (1)               | 0 (0)    | 1.000    |
| **P-ANCA (≤10 IU/ml) (n, %)** |          | (0)^b               | 7 (5)               | 4 (5)               | 3 (5)    | 1.000    |
| **ASCA IgA (≤10 IU/ml) (n, %)** |          | (8)^b               | 0 (0)               | 0 (0)               | 0 (0)    |
| **ASCA IgG (≤10 IU/ml) (n, %)** |          | (8)^b               | 17 (13)             | 7 (9)               | 10 (18)  | 0.187    |
| **Anti-GAD (≤11 kIU/L) (n, %)** |          | (0)^b               | 7 (5)               | 4 (5)               | 3 (5)    | 1.000    |
| **Anti-IA2 (≤15 kIU/L) (n, %)** |          | (31)^c              | 2 (2)               | 1 (1)               | 1 (2)    | 0.100    |
| **Anti-TPO (≤70 kIU/L) (n, %)** |          | (7)^c               | 18 (14)             | 9 (12)              | 9 (16)   | 0.444    |
| **TRAK (≤15 IU/L) (n, %)** |          | (1)^d               | 1 (1)               | 0 (0)               | 1 (2)    |

ANA = anti-nuclear antibodies, C-ANCA = cytoplasmic anti-neutrophil cytoplasmic antibodies, ASCA = anti-Saccharomyces cerevisiae antibodies, CRP = C-reactive protein, anti-IA2 = anti-islet antigens-like insulin 2, anti-GAD = anti-glutamic acid decarboxylase, Hb = haemoglobin, IgA = Immunoglobulin A, IgG = Immunoglobulin G, P = plasma, P-ANCA = peri-nuclear anti-neutrophil cytoplasmic antibodies, S = serum, anti-TPO = anti-thyroid peroxidase, TRAK = thyrotropin receptor antibodies. Reference values in brackets. n (%) = number (percent) of positive antibodies. Controls consisted of: a = healthy blood donors, 50 men and 50 women, b = 50 healthy blood donors, c = 120 healthy women, reference No 17, d = 282 healthy controls from reference No 16. Values are given as mean ± standard deviation or median (interquartile range). Student t- test, Mann-Whitney U- test or Fishers exact test were used for comparisons between collagenous colitis and lymphocytic colitis. P<0.01 was considered statistically significant.

doi:10.1371/journal.pone.0066088.t001

endometriosis, Chlamydia Trachomatis, kidney stones, coeliac disease, diabetes mellitus type 1 or 2, myocardial infarction, intermittent claudication or stroke. A list of the drugs, vitamins and minerals currently being taken was completed. The participants were also asked about heredity for cancer, stroke, myocardial infarction, hypertension, diabetes mellitus, thyroid disorders, asthma, rheumatoid arthritis, IBD or coeliac disease.

Gastrointestinal symptom rating scale (GSRS). The GSRS is a Swedish, disease-specific and self-administered questionnaire, designed to evaluate perceived severity of gastrointestinal symptoms during the previous week. The questionnaire includes 15 items and uses a 7-grade Likert scale. This gives a total range value between 15 and 105 where the highest score (seven) represents the most pronounced symptoms and the lowest (one) no symptoms. The items are divided into five dimensions representing Reflux Syndrome (two items), Abdominal Pain Syndrome (three items), Constipation Syndrome (three items), Indigestion Syndrome (four items) and Diarrhoea Syndrome (three items). The data are presented as the average of the total score divided by the number of items. Norm values for the healthy, gender-matched population are available.

Rome III criteria. The patients completed an abbreviated version of the Rome III questionnaire, including only irritable bowel syndrome (IBS) symptoms. This questionnaire has been translated and validated into the Swedish language (Magnus Simrén and Anna Ryden). Patients who fulfilled the criteria in the Rome III questionnaire were classified as also suffering from IBS-like symptoms.
Statistical Analyses

The data were analysed using the statistical software package SPSS for Windows® (Release 20.0; IBM). Values are given as mean ± standard deviation (SD) or median (interquartile range) depending on distribution norm. Gastrointestinal symptoms are given as the mean with a 95% confidence interval, because the reference values in reference No 25 are given in this form. All patients who expressed any form of measured auto-antibody were classified as antibody positive, and calculations of the association between antibodies and clinical features were performed both for each antibody and for the total presence of antibodies. Differences between groups were calculated by the 2-tailed Mann-Whitney U-test or Student t-test. The Spearman correlation test was used to examine correlations between groups. Fisher’s exact test was used to examine differences between the presence of auto-antibodies in diseases and heredity; differences in the prevalence of auto-antibodies and diseases between CC and LC; and differences in the prevalence of auto-antibodies between MC1 and MC2. Due to several hypotheses tested, p<0.01 was considered statistically significant.

Results

Patient Characteristics

In total, the 133 women with MC, who provided blood samples, were included in the study (median age 63 (59–67) years, range 27–73 years). CC was diagnosed in 77 patients (58%) (median age 64 (59–68) years, range 31–73 years) and LC in 56 patients (42%) (median age 63 (54–67) years, range 27–72 years). Of these 133, 69 patients (52%) also fulfilled the Rome III criteria for IBS, 40 of the patients (52%) with CC and 29 of the patients (52%) with LC. The duration of gastrointestinal symptoms at study inclusion was 8 (3–14) years. There was no correlation between duration of symptoms and age (correlation coefficient = −0.097, p = 0.286). The body mass index was 24.8 (22.7–28.7) kg/m². Of the patients, 25% were regular smokers, 8% smoked occasionally, 40% had never smoked and 27% had never smoked. There was no difference in smoking habits between CC and LC (p = 0.350). Routine analyses were within the reference values used in the laboratory in the vast majority of cases (Table 1).

Prevalence of Auto-antibodies

The highest prevalence of antibodies was found of ANA, ASCA IgG, and anti-TPO antibodies, where the prevalence was doubled in the patients compared to the general population, with no differences between CC and LC, or between MC1 and MC2 (Table 1 and Table 2). There was no difference between CC and LC in the total presence of auto-antibodies (p = 0.375). The median age in ANA-positive patients tended to be higher than in ANA-negative patients (67 (62–71) years and 63 (56–67) years, respectively, p = 0.013). In contrast, the median age tended to be lower in ASCA-positive patients compared to ASCA-negative patients (59 (46–66) years and 64 (59–68) years, respectively, p = 0.031). The relatively high prevalence of anti-TPO antibodies was not reflected by such a difference in age in patients who expressed antibodies compared to those who did not (60 (47–66) years and 64 (59–68) years, respectively, p = 0.097).

When examining the most prevalent concomitant diseases, excluding sporadic diseases, ANA-positive patients also suffered from hypertension (nine), allergy (seven), thyroid disorders (six), rheumatoid arthritis (three), and coeliac disease (three). Patients expressing ASCA IgG also suffered from hypertension (eight), rheumatoid arthritis (four), asthma (three), coeliac disease (three), and allergy (three). Anti-TPO antibodies were found together with thyroid disorders in seven patients, with allergy in four and with asthma and rheumatoid arthritis in three. Of the seven who expressed P-ANCA, four suffered from allergy, and three from thyroid disorders, rheumatoid arthritis, and hypertension, respectively. Of the seven patients who expressed anti-GAD antibodies, five also suffered from diabetes mellitus. One of the patients with anti-IgA2 antibodies also expressed anti-GAD antibodies, and suffered from diabetes mellitus. Only eight of the patients with any antibody had MC as a sole disease. The patient with the most concomitant diseases suffered from seven of these, apart from MC and IBS. There was no difference in the results of routine blood

Table 2. Prevalence of auto-antibodies in persistent (MC1) and transient microscopic colitis (MC2).

|          | MC1 | MC2 | P-value |
|----------|-----|-----|---------|
| Age (years) | 63 (57–67) | 64 (59–68) | 0.405 |
| Hb (117–153 g/L) | 132.3±6.8 | 132.1±9.9 | 0.916 |
| Leucocytes (3.5–8.8×10^9/L) | 7.0±2.3 | 6.9±1.5 | 0.906 |
| S-Albumin (36–48 g/L) | 40.1±3.5 | 39.7±3.1 | 0.494 |
| CRP (<3.1 mg/L) | 2.0 (1.0–4.0) | 2.0 (1.0–3.5) | 0.840 |
| P-Sodium (137–145 mmol/L) | 141.0 (139.0–142.0) | 140.0 (139.0–142.0) | 0.253 |
| P-Potassium (3.5–4.4 mmol/L) | 3.9±0.3 | 3.9±0.4 | 0.736 |
| ANA (≤14 IU/ml) | 12 (16.4) | 7 (11.7) | 0.619 |
| C-ANCA (≤10 IU/ml) | 1 (1.4) | 0 (0) | 1.000 |
| P-ANCA (≤10 IU/ml) | 5 (6.8) | 2 (3.3) | 0.463 |
| ASCA IgA (≤10 IU/ml) | 0 (0) | 2 (3.3) | 0.311 |
| ASCA IgG (≤10 IU/ml) | 11 (15.1) | 6 (10) | 0.602 |
| Anti-GAD (≤11 IU/L) | 5 (6.8) | 2 (3.3) | 0.460 |
| Anti-IgA (≤15 IU/L) | 0 (0) | 2 (3.3) | 0.100 |
| Anti-TPO (≤70 IU/L) | 9 (12.3) | 9 (15.0) | 0.799 |
| TRAK (≤1.5 IU/L) | 1 (1.7) | 1 (1.7) | 0.456 |

ANA = anti-nuclear antibodies, C-ANCA = cytoplasmic anti-neutrophil cytoplasmic antibodies, ASCA = anti-Saccharomyces cerevisiae antibodies, CRP = C-reactive protein, anti-IgA2 = anti-islet antigens-like insulin 2, anti-GAD = anti-glutamic acid decarboxylase, Hb = haemoglobin, IgA = Immunoglobulin A, IgG = Immunoglobulin G, P = plasma, P-ANCA = perinuclear anti-neutrophil cytoplasmic antibodies, S = serum, anti-TPO = anti-thyroid peroxidase, TRAK = thyrotropin receptor antibodies. Reference values in brackets. n (%) = number (percent) of patients with positive antibodies. Values are given as mean ± standard deviation or median (interquartile range). Student t-test, Mann-Whitney U-test or Fisher’s exact test. P<0.01 was considered statistically significant.

doi:10.1371/journal.pone.0066088.t002
Prevalence of Other Diseases

analyses between patients with and those without auto-antibodies (Table 3).

Prevalence of Gastrointestinal Symptoms

Patients with MC had more gastrointestinal symptoms than healthy females in all the dimensions Reflux Syndrome, Abdominal Pain Syndrome, Constipation Syndrome, Indigestion Syndrome, and Diarrhoea Syndrome (Table 5). There was no difference in symptoms between CC and LC (Table 5). There was no difference in disease duration and degree of gastrointestinal symptoms between patients with and those without any kind of auto-antibodies (Table 3).

Discussion

The main finding of this cross-sectional study was that the prevalence of common auto-antibodies such as ANA, P-ANCA, ASCA IgG, and anti-GAD and anti-TPO antibodies is slightly more prevalent in MC patients than in the general Swedish population, but they are nonetheless present in only a minority of cases. C-ANCA, ASCA IgA, anti-IA2 antibodies, and TRAK are only occasionally found. There was no difference in gastrointestinal symptoms between patients with and those without auto-antibodies. The antibodies were present in patients who also suffered from other concomitant diseases.

Auto-immunity has been suggested as a plausible aetiology of MC, as an association with the same HLA genotypes in MC as described in other known auto-immune diseases has been described [2,9]. However, the prevalence of C-ANCA, ASCA IgA, anti-IA2 antibodies, and TRAK was in the same range or lower as in the general population, according to our laboratory reference values of healthy blood donors, as has also been described in previous studies [3,7,11,13,16,17]. Patients predisposed to auto-immunity usually express higher levels of several antibodies in serum [5,6,7,8]. The prevalence of ANA, P-ANCA, ASCA IgG, and anti-GAD and anti-TPO antibodies were slightly increased compared to healthy controls, but not at all to the same magnitude as in patients with Crohn’s disease, ulcerative colitis, diabetes mellitus or thyroid disorders [7,8,12,27,28]. Furthermore, reference values are set in relation to the general population, including both genders and all ages. In this study of women of upper middle age, the prevalence of auto-antibodies is higher than in the general population [14]. World-wide, 12%–26% of healthy women and 3%–14% of healthy men have anti-TPO antibodies [28], and 6% of healthy women are considered positive for ANA [29]. The increased prevalence of antibodies in our present study can be explained by other concomitant diseases of auto-immune character, e.g. rheumatoid arthritis, coeliac disease, thyroid disorders and diabetes mellitus, and by our cohort being of upper middle-aged women. As our older MC patients had many concomitant diseases, not only of auto-immune origin, it is difficult to estimate the true association with auto-immune diseases. The associations observed could be due to secondary effects on the digestive tract of the diseases, or their treatment, and do not necessarily mean a common causal aetiology [1]. Smoking per se is associated with a higher prevalence of ANA [30], and this could explain the higher ANA positivity in our study with a high frequency of former and current smokers. Thus, our finding in the present study does not indicate that auto-immunity is the most important cause of MC.

Previously described associations between MC and drug treatment and smoking [31,32], and the higher age in the diseased patients [9], suggest environmental and circumulatory influences as important aetiological factors for MC [33]. Intestinal ischemia should also be considered, especially as smoking is a risk factor for the disease [32], and the patients are older, with a high prevalence of cardiovascular diseases [33]. The high prevalence of using

| Table 3. Laboratory values and gastrointestinal symptoms in relation to the presence of auto-antibodies. |
|---------------------------------------------------------------|
| Antibodies | No antibodies | P-value |
| Age (year) | 62 (58–68) | 63 (58–67) | 0.845 |
| Hb (117–153 g/L) | 132.5±3.9 | 132.0±9.1 | 0.764 |
| Leucocytes (3.5–8.8×10⁹/L) | 6.9±2.1 | 7.0±1.9 | 0.791 |
| Albumin (36–48 g/L) | 40.0±3.3 | 40.0±3.4 | 0.966 |
| CRP (<3.1 mg/L) | 2.0 (1.0–4.4) | 2.0 (1.0–3.5) | 0.056 |
| P-Sodium (137–145 mmol/L) | 141.0 (139.0–143.0) | 140.0 (139.0–142.0) | 0.253 |
| P-Potassium (3.5–4.4 mmol/L) | 3.9±0.3 | 3.9±0.4 | 0.907 |
| Duration (years) | 10.0 (5.8–16.0) | 6.0 (3.0–12.0) | 0.038 |
| Abdominal pain | 2.55 (2.28–2.83) | 2.63 (2.37–2.88) | 0.428 |
| Indigestion | 3.01 (2.76–3.26) | 2.92 (2.66–3.18) | 0.372 |
| Reflux | 2.50 (2.17–2.83) | 2.30 (2.04–2.56) | 0.195 |
| Constipation | 3.33 (2.92–3.74) | 3.40 (3.04–3.76) | 0.457 |
| Diarrhoea | 4.02 (3.68–4.35) | 3.90 (3.55–4.24) | 0.372 |
| GSRS Total | 8.92 (8.04–9.80) | 9.30 (8.69–9.92) | 0.299 |

Values are given as median (interquartile range) or mean with a 95% confidence interval. GSRS = Gastrointestinal Symptom Rating Scale. Mann-Whitney U-test or Student t-test. P<0.01 was considered statistically significant. doi:10.1371/journal.pone.0066088.t003
angiotensin-converting enzyme inhibitors or angiotensin II receptor antagonists and beta-blockers in the current study, may also affect intestinal blood circulation, with secondary tissue effects. Several conditions, including surgical manipulation of the tissue, *Helicobacter pylori*, infections and drugs may lead to intestinal lymphocytosis [1,34]. In clinical practice, patients with a histopathological diagnosis of MC are clinically classified as MC, irrespective of whether the lymphocytosis is primary or secondary. Furthermore, 63% of patients have had only a single attack of MC [35]. In other prospective studies, there were high relapse rates after completion of treatment with budesonide [36,37], and some patients have refractory and severe symptoms requiring surgery [9]. This must indicate the necessity to restrict our future classification of MC. The diagnosis of MC should probably include only true conditions of consistent, permanent intestinal lymphocytosis, without any other plausible explanations [1,33,38].

However, we did not find any differences between MC1 and MC2 in antibody prevalence.

In this study, the patients represent the vast majority of female MC patients younger than 73 years in Skåne, independent of whether they are treated at primary care or tertiary centres. This may explain the lower prevalence of auto-immune diseases and antibody prevalence compared to other studies representing tertiary centres, in which studies of only auto-immune concomitant disorders are reported [3,4,7,39]. Previous studies, in which a higher prevalence of coeliac disease has been described, patients were younger and were screened by endoscopic biopsies specifically to detect coeliac disease. Coeliac disease is characterised by villous atrophy in the small intestine, and the colonic lymphocytosis may reflect the same reaction of the large intestine, rather than a distinct disease [1,39]. Our patient cohort has a high prevalence of IBS-like symptoms, and among these patients there

| Table 4. The prevalence of different past and current diseases in microscopic colitis and its association with auto-antibodies (ab). |
| --- |
| **Collagenous colitis (CC)** | **Lymphocytic colitis (LC)** | **P-value** | **Microscopic colitis (MC)** | **P-value*** |
| n = 77 | n = 56 | CC vs LC | n = 133 | ab vs MC |
| Age (year) | 64 (59–68) | 63 (54–67) | 0.080 | 63 (59–67) | 0.845 |
| IBS | 40 (52) | 29 (52) | 1.000 | 69 (52) | 0.482 |
| Hypertension | 27 (35) | 14 (25) | 0.173 | 41 (31) | 0.033 |
| Allergy | 29 (38) | 12 (21) | 0.075 | 41 (31) | 1.000 |
| Thyroid disorder | 14 (18) | 16 (29) | 0.208 | 30 (23) | 0.085 |
| Fracture | 13 (17) | 14 (25) | 0.372 | 27 (20) | 0.176 |
| Rheumatoid arthritis | 13 (17) | 13 (23) | 0.500 | 26 (20) | 0.258 |
| Asthma | 15 (19) | 7 (12) | 0.343 | 22 (16) | 0.810 |
| Gastric ulcer | 11 (14) | 10 (18) | 0.808 | 21 (16) | 0.327 |
| Ovarial inflammation | 13 (17) | 6 (11) | 0.321 | 19 (14) | 0.323 |
| Herpes | 6 (8) | 13 (23) | 0.022 | 19 (14) | 0.330 |
| Endometriosis | 8 (10) | 5 (9) | 1.000 | 13 (10) | 0.235 |
| Chlamydia | 6 (8) | 6 (11) | 0.763 | 12 (9) | 0.762 |
| Kidney stones | 9 (12) | 2 (4) | 0.116 | 11 (8) | 0.346 |
| Cancer | 8 (10) | 3 (5) | 0.347 | 11 (8) | 0.115 |
| Coeliac disease | 2 (3) | 8 (14) | 0.017 | 10 (8) | 0.314 |
| Diabetes | 5 (6) | 5 (9) | 1.000 | 10 (8) | 0.514 |
| Type 1 | 2 (3) | 1 (2) | 3 (2) |
| Type 2 | 3 (4) | 4 (7) | 7 (5) |
| Myocardial infarction | 6 (8) | 2 (4) | 0.464 | 8 (6) | 1.000 |
| Intermittent claudication | 3 (4) | 3 (5) | 1.000 | 6 (4) | 0.222 |
| Stroke | 3 (4) | 2 (4) | 1.000 | 5 (4) | 0.646 |

n (%) = number (percent) of patients with the disease. IBS = Irritable bowel syndrome, *= Comparison of prevalence of diseases in the whole MC group between patients with or without any auto-antibodies. Mann-Whitney U-test and Fishers exact test. P ≤ 0.01 was considered statistically significant. doi:10.1371/journal.pone.0066088.t004
may be hidden some cases of un-investigated coeliac disease, not
diagnosed by serology. As IBS is most often found in younger
persons and is characterised by no organic findings [26], we have
chosen to call the presence of abdominal pain and bloating IBS-
like symptoms. These symptoms are non-specific, and may depend
on side effects of drug treatment rather than a different and
distinct disease.

The peak incidence of many of the concomitant diseases is at a
younger age, whereas the peak incidence of MC is at 65 years
[9,14,40]. One limitation of this and most other studies in this field
is the retrospective or cross-sectional character, making it difficult
to know which disease, or treatment, debuted first. Furthermore, if
the patient is not referred to colonoscopy, the diagnosis IBS is set
instead of MC. In many cases, the patients have had gastrointestinal
symptoms for several years prior to colonoscopy, making the
peak age of incidence falsely higher. Patients with concomitant
diseases, already treated for these at a hospital, are probably
referred for a colonoscopy sooner after debut of another symptom
than an otherwise healthy individual, and thus the diagnosis is
most often set in these cases.

The small differences between CC and LC in this study
corresponding concomitant diseases and serological markers, and in
our previous study concerning gastrointestinal symptoms and
psychological well-being [41], suggest that the two forms of the
disease are not markedly heterogenous. A transition between the
subtypes over time has been described, and that perhaps the two
forms should be considered as one entity aetiologically [38]. The
classification as CC and LC may reflect a histopathologic
classification rather than a clinically relevant distinction [33].

In conclusion, although they showed a slightly increased
prevalence of some auto-antibodies, the majority of our MC
patients did not express any of the antibodies measured. The
higher prevalence of auto-antibodies measured may be explained
by other, concomitant, auto-immune diseases, a high frequency of
smokers, and the composition of the cohort being of women of late
middle age. Future research in this field should focus on
prospective studies in that fraction of patients with MC only, to
find out whether auto-immunity is the aetiology in these cases.

Author Contributions
Conceived and designed the experiments: BR RG BO. Performed the
experiments: BR RG BO. Analyzed the data: BR RG BO. Contributed
reagents/materials/analysis tools: BR RG BO. Wrote the paper: BR RG
BO. The patients included: RG. Collected the blood samples and
data from the patients: BR. Performed the statistical analyses: BO.
Contributed to the manuscript with constructive criticism, and read and
approved the final manuscript: BR RG BO.

References
1. Carmack SW, Lash RH, Gulizia JM, Genta RM (2009) Lymphocytic disorders
of the gastrointestinal tract: a review for the practicing pathologist. Adv Anat
Pathol 16: 290–306.
2. Koskela RM, Karttunen TJ, Niemela SE, Lehtola JK, Ilonen J, et al. (2008)
Human leucocyte antigen and TNFalpha polymorphism association in
microscopic colitis. Eur J Gastroenterol Hepatol 20: 276–282.
3. Bohr J, Tysk C, Yang P, Danielsson D, Jarnerot G (1996) Autoantibodies and
immunoglobulins in collagenous colitis. Gut 39: 73–76.
4. Holstein A, Barneonit J, Plaschke A, Rosemeier D, Waelkens E, et al. (2006)
Autoantibody profiles in microscopic colitis. J Gastroenterol Hepatol 21: 1016–
1020.
5. Pilia S, Casini MR, Cambioli VM, Bba A, Civoli P, et al. (2011) Prevalence of
Type 1 diabetes autoantibodies (GAD and IAZ) in Sardinian children and
adolescents with autoimmune thyroiditis. Diabet Med 28: 896–899.
6. Op De Beeck K, Maes L, Van den Bergh K, Derua R, Waëlens E, et al. (2012)
Heterogeneous nuclear RNPs as targets of autoantibodies in systemic rheumatic
diseases. Arthritis Rheum 64: 213–221.
7. De urn RH, Taegar SR, Launders CJ, Sutherland LR, Shanahan F (1991) Anti-
neutralophil cytoplasmic antibodies in ulcerative colitis. Comparison with other
collitides/diarrhoeal illnesss. Gastroenterology 100: 1590–1596.
8. Prudhoe L, de Cruz P, Ng SC, Kamn MA (2012) Serological antibodies in
inflammatory bowel disease: a systematic review. Inflamm Bowel Dis 18: 1340–
1355.
9. Pardi DS, Kelly CP (2011) Microscopic Colitis. Gastroenterology 140: 1165–
1165.
10. Gal AA, Velasquez A (2002) Antineutrophil cytoplasmic autoantibody in the
absence of Wegeners granulomatosis or microscopic polyangiitis: implications for
the surgical pathologist. Mod Pathol 15: 197–200.
11. Bonifacio E, Bingley PJ (1997) Ilet autoimmune and their use in predicting
insulin-dependent diabetes. Acta Diabetol 34: 105–193.
12. Rahmati K, Lernmark A, Becker C, Folsyn-Zadura A, Larson K, et al. (2008) A comparison of serum and EDTA plasma in the measurement of glutamic acid decarboxylase autoantibodies (GADA) and autoantibodies to islet antigen-2 (IA-2A) using the RSR radioimmunoassay (RIA) and enzyme linked immunosorbent assay (ELISA) kits. Clin Lab 54: 227–235.

13. Tuomilehto J, Yliharsila H (1998) Antibodies as predictors of insulin-dependent diabetes mellitus before the clinical onset. Nutrition 14: 403–405.

14. Symmons DP (2002) Epidemiology of rheumatoid arthritis: determinants of onset, persistence and outcome. Best Pract Res Clin Rheumatol 16: 707–722.

15. Sirrén M, Abrahamsson H, Svedlund J, Bjo¨rnsson ES (2001) Quality of life in patients with irritable bowel syndrome seen in referral centers versus primary care: the impact of gender and predominant bowel pattern. Scand J Gastroenterol 36: 543–552.

16. Costagida S, Morgenhalter NG, Hoermann R, Badenhoop K, Struck J, et al. (1999) Second generation assay for thyrotropin receptor antibodies has superior diagnostic sensitivity for Graves’ disease. J Clin Endocrinol Metab 84: 90–97.

17. Persson A, Becker C, Hansson I, Nilsson A, To¨rn C (2010) Comparison of serum and EDTA plasma in the measurement of glutamic acid decarboxylase autoantibodies (GADA) and autoantibodies to islet antigen-2 in Whole Blood Eluates from Dried Blood Spots Using the RSR-Enzyme Linked Immunosorbsent Assay Kits and In-House Radioimmunoassays. Exp Diabetes Res. Online June 3.

18. Westman KWA, Selga D, Bygren P, Segelmark M, Baslund B, et al. (1998) Clinical evaluation of a capture ELISA for detection of proteinase-3 antineutrophil cytoplasmic antibody. Technical Note Kidney Int 53: 1239–1236.

19. Reichlin M (2006) Measurement and clinical significance of antinuclear antibodies. Up to date Online ver 15.1.

20. Mankaı¨ A, Sakly W, Thabet Y, Achour A, Manoubi W, et al. (2013) Anti-Saccharomyces cerevisiae antibody. Technical Note Kidney Int 53: 1230–1236.

21. www.analysporten-labmedicin.skane.se.

22. Svedlund J, Sjo ¨din I, Dotevall G (1988) GSRS – a clinical rating scale for upper gastrointestinal disease. Scand J Gastroenterol 31 Suppl 221: 8–13.

23. Dimenas E, Glise H, Hallerback B, Herqvist H, Svedlund J, et al. (1993) Quality of life in patients with upper gastrointestinal symptoms An improved evaluation of treatment regimens? Scand J Gastroenterol 28: 601–607.

24. Dimenas E, Glise H, Hallerback B, Herqvist H, Svedlund J, et al. (1995) Well-being and gastrointestinal symptoms among patients referred to endoscopy owing to suspected duodenal ulcer. Scand J Gastroenterol 30: 1046–1052.

25. Dimenas E, Carlsson G, Glise H, Israelsson B, Wilkudd J (1996) Relevance of norm values as part of the documentation of quality of life instruments for use in upper gastrointestinal disease. Scand J Gastroenterol 31 Suppl 221: 8–13.

26. Drossman DA, Corraziari E, Delvaux M, Spiller R, Talley NJ, et al. Rome III: The Functional Gastrointestinal Disorders. McLean, VA: Dgnoton Associates; 2006.

27. Weng J, Ekelund M, Lehto M, Li H, Elberg G, et al. (2002) Screening for MODY mutations, GAD antibodies, and Type 1 diabetes-associated HLA genotypes in women with gestational diabetes mellitus. Diabetes Care 25: 68–71.

28. Prummel MF, Wiersinga WM (2005) Thyroid peroxidase autoantibodies in euthyroid subjects. Best Pract Res Clin Endocrinol Metab 19: 1–15.

29. Matthiesen LS, Berg G, Emeruddh J, Skogh T (1999) A prospective study on the occurrence of autoantibodies in low-risk pregnancies. Eur J Obstet Gynecol Reprod Biol 83: 21–26.

30. Karabulut G, Kitapcioglu G, Inal V, Kalfa M, Yargucu F, et al. (2011) Cigarette smoking in primary Sjo¨gren’s syndrome: positive association only with ANA positivity. Mod Rheumatol 21: 602–607.

31. Fernandez-Banares F, Esee M, Espinos JC, Roinach M, Forne M, et al. (2007) Drug consumption and the risk of microscopic colitis. Am J Gastroenterol 102: 324–330.

32. Yen EF, Pokhrel B, Du H, New S, Bianchi L, et al. (2012) Current and past cigarette smoking significantly increase risk for microscopic colitis. Inflamm Bowel Dis 18: 1033–1041.

33. Bjernhak C, Engel PJ, Nielsen PL, Munck LK (2011) Microscopic colitis: clinical findings, topography and persistence of histopathological subgroups. Aliment Pharmacol Ther 34: 1225–1234.

34. Lee S, Ogihie RT, Dupre M, Gao ZH (2009) Intravascular lymphocytosis in acute appendicitis: potential mimicry of chronic lymphocytic leukaemia. Histopathology 55: 660–664.

35. Olesen M, Eriksson S, Bohr J, Jarnerot G, Tyk C (2004) Lymphocytic colitis: a retrospective clinical study of 199 Swedish patients. Gut 53: 536–541.

36. Miehle S, Madisch A, Karimi D, Wenschak S, Kuhlsch E, et al. (2009) Budesonide is effective in treating lymphocytic colitis: a randomized double-blind placebo-controlled study. Gastroenterology 136: 2092–2100.

37. Bonderup OK, Hansen JB, Teglbjaerg PS, Christensen LA, Fallangborge JF (2009) Long-term budesonide treatment of collagenous colitis: a randomised, double-blind, placebo-controlled study. Gastroenterology 136: 2092–2100.

38. Rasmussen MA, Munck LK (2012) Systematic review: are lymphocytic colitis and collagenous colitis two subtypes of the same disease-microscopic colitis? Aliment Pharmacol Ther 36: 79–90.

39. Stewart M, Andreasen GN, Urquhsani S, Beck PL, Storr M (2011) The association of celiac disease and microscopic colitis: a large population-based study. Aliment Pharmacol Ther 35: 1140–1149.

40. Dahlquist GG, Nystro ¨m L, Patterson CC; Swedish Childhood Diabetes Study Group (2011) Diabetes Incidence in Sweden Study Group. Incidence of type 1 diabetes in Sweden among individuals aged 0–34 years, 1983–2007: an analysis of time trends. Diabetes Care 34: 1754–1759.

41. Roth B, Ohiison B (2013) Gastrointestinal symptoms and psychological well-being in patients with microscopic colitis. Scand J Gastroenterol 48: 27–34.