Prospective Association Between Inflammatory Markers and Progression of Coronary Artery Calcification in Adults With and Without Type 1 Diabetes

Amy C. Alman, PhD
Gregory L. Kinney, PhD, MPH
Russell P. Tracy, PhD
David M. Maahs, MD, PhD

John E. Hokanson, PhD
Marian J. Rewers, MD, PhD
Janet K. Snell-Bergeon, PhD, MPH

OBJECTIVE—The role of inflammation in the increased risk of cardiovascular disease in type 1 diabetes is unclear. We examined the association of inflammation and progression of coronary artery calcification (CAC)—a marker of subclinical atherosclerosis—in adults with and without type 1 diabetes.

RESEARCH DESIGN AND METHODS—A nested case-control study was performed within the prospective cohort of the Coronary Artery Calcification in Type 1 Diabetes (CACTI) study. Participants underwent two CAC measurements ~2.5 years apart. Case subjects (n = 204) were those with significant progression of CAC. Control subjects (n = 258) were frequency-matched to case subjects on diabetes status, sex, age, and baseline CAC status. Inflammatory marker assessments were performed on stored blood samples from baseline. A principal components analysis (PCA) was performed and a composite score derived from that analysis. The composite score was constructed by assigning a value of 1 for each PCA component where at least one of the markers exceeded the 75th percentile (range 0–4). Conditional logistic regression was used for the matching strategy.

RESULTS—The first two components of the PCA were modestly (odds ratio 1.38 [95% CI 1.08–1.77] and 1.27 [1.02–1.59], respectively) associated with CAC progression after adjustment for other risk factors. The composite score was more strongly associated with CAC progression for those with elevated markers in three or four of the principal components compared with those with none.

CONCLUSIONS—Measures of inflammation were associated with progression of CAC in a population of adults with and without type 1 diabetes.

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From the 1Department of Epidemiology and Biostatistics, College of Public Health, University of South Florida, Tampa, Florida; the 2Department of Epidemiology, Colorado School of Public Health, University of Colorado Denver, Aurora, Colorado; the 3Department of Pathology, University of Vermont, Burlington, Vermont; and the 4Barbara Davis Center, University of Colorado Denver, Aurora, Colorado. Corresponding author: Amy C. Alman, salman@health.usf.edu. Received 13 September 2012 and accepted 19 December 2012. DOI: 10.2337/dc12-1874

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Inflammation and CAC

the CACTI Study, a prospective cohort study examining the prevalence of CAC—a marker of subclinical atherosclerosis—in adults with type 1 diabetes and a comparable group of control subjects. Detailed descriptions of the study design have been published (22). Briefly, the full cohort consisted of 1,416 participants (652 with type 1 diabetes, 764 control subjects) who reported no history of CVD and were asymptomatic for CVD at enrollment. All study participants provided informed consent, and the protocol was reviewed and approved by the Colorado Multiple Institutional Review Board.

Examination measurements

Physical examination measurements included height, weight, waist and hip circumference, and systolic and diastolic blood pressure. BMI was calculated as weight in kilograms divided by the square of height in meters. Hypertension was defined as systolic blood pressure ≥140 mmHg or diastolic blood pressure ≥90 mmHg or treatment with antihypertensive medication. A fasting blood sample was collected and stored at −80°C until assayed for measurement of cholesterol (total and HDL) and triglyceride levels. Retinopathy was as-essed for measurement of cholesterol (total and HDL) and triglyceride levels. Physical examination measurements in-cluded height, weight, waist and hip circumference, and systolic and diastolic blood pressure. BMI was calculated as weight in kilograms divided by the square of height in meters. Hypertension was defined as systolic blood pressure ≥140 mmHg or diastolic blood pressure ≥90 mmHg or treatment with antihypertensive medication. A fasting blood sample was collected and stored at −80°C until assayed for measurement of cholesterol (total and HDL) and triglyceride levels. Retinopathy was as-

Subject selection

Selection of subjects for this nested case-control study occurred after the second visit, which was an average of 2.5 years (range 1.1–4.3) after the baseline. Eligible case subjects for this study were those with significant progression of CAC (n = 204), defined as an increase in volume of CAC between the baseline and follow-up visits of ≥2.5 square root transformed units. Significant regression would be a reduction in CAC volume of ≥2.5 square root transformed units, but this has not been observed in this population (CAC volume in 53 subjects decreased by less than that). Previous work has shown that this definition of change in CAC volume represents meaningful differences in CAC that are unlikely to be due to interscan variability (23,24). Prevalence of CAC at baseline was not a factor in whether a subject was eligible to be a case; thus, case subjects could have had no CAC at baseline.

Participants with no significant progression in the volume of CAC were eligible to be selected as a control subject. A binary indicator was used to identify the presence of CAC at baseline. Control subjects were frequency matched to case subjects on diabetes status, sex, and age group at the follow-up visit (<30, 30–39, 40–49, and ≥60 years) and the presence of CAC at baseline (n = 258).

Laboratory measurements

Laboratory assessments were performed in the laboratory of Dr. Russell P. Tracy at the University of Vermont on the stored plasma or serum specimens obtained at baseline. Plasma or serum was deemed optimal for different analytes. IL-6 was measured using a multiplex panel (Bio-Rad). TNF-α and sIL-1RA were measured on a cytokine panel (Millipore/Linco). Commercially available ELISA kits (R&D Systems, Alpco Diagnostics) were used to measure hsCRP, sIL-6R, soluble TNF-α receptor type II (sTNFR2), sIL-2R, IL-18, soluble intercellular adhesion molecule-1 (ICAM-1), P-selectin, soluble matrix metalloproteinase-3 (MMP-3), and osteoprotegerin (OPG). These biomarkers were chosen to represent different inflammatory pathways. Baseline measures for the inflammatory markers were used to examine the association of inflammation on the progression of CAC from baseline. Detailed information regarding the assays, range of detection, and standard values for each marker are presented in the Supplementary Data.

Statistical analyses

Distributions of all variables were examined to determine departure from normality. All of the inflammatory markers, with the exception of P-selectin, were skewed and were log-transformed for all analyses. Differences were compared by diabetes and case status. Parametric continuous data are presented as means ± SD. Nonparametric data are presented as the median and interquartile range, with the exception of the log-transformed inflammatory markers, which are presented as the geometric mean and the interquartile range. Categorical data are presented as the number of subjects and the percent. Statistical testing to detect differences between groups included the t test for parametric continuous data, the Wilcoxon rank sum test for nonparametric data, and the χ² test for categorical data.

Principal components analysis

Many cytokines exhibit pleiotropic and synergistic effects. Evaluating single independent markers would likely underestimate these effects; thus, a method that considers the combined influence of multiple markers, and therefore the overall inflammatory burden, would be preferable. However, combining markers into a composite measure needs to avoid overestimating the association due to the potentially high correlation between markers. We therefore used principal components analysis (PCA) with orthogonal rotation to derive uncorrelated linear transformations of the biomarkers. We considered the eigen values (≥1), scree plots, and interpretability (variables with factor loads ≥0.40) of the final solution in determining the minimum number of components to use for further analysis. Markers with factor loads ≥0.40 on multiple components (ICAM-1, OPG, and MMP-3) were dropped, and the PCA was run again. In the final PCA, the first four components explained 65% of the total variance and were used in the multivariate regression analysis.

Inflammatory marker composite score

To test a measure of the combined effects (inflammatory burden), a composite score
was constructed using the component interpretation from the PCA. Subjects were assigned a score of 1 for each component in which one or more of the markers with a factor load ≥0.40 in that component had a value exceeding the 75th percentile. The scores were added across the four components to arrive at the summed composite score. Composite scores could therefore range from 0 (in which none of the markers exceeded the 75th percentile) to 4 (in which at least one marker from each component exceeded the 75th percentile). In this way, the composite score captured subjects with high levels of inflammatory markers in multiple components, without inflating the score due to the correlation between markers within components. We used the distribution of the composite score to categorize the score into the following: 0, 1–2, and 3–4.

Multivariate modeling

Conditional logistic regression to adjust for the matching of case subjects and control subjects was used to determine the independent effect of the principal components and the composite score on the progression of CAC. All continuous variables were examined for the best functional form before model testing using the −2 log-likelihood ratio test and visual inspection of plots. Potential confounding variables were considered for inclusion in the models based on a priori criteria: significance in previous work, significant contribution to the model fit (P value of the Wald χ² < 0.05), or confounding the association between the main variable of interest and the outcome by more than 10%. The age-group, diabetes status, presence of CAC at baseline, and sex were included as the conditional variables to account for the matching. Systolic blood pressure, duration of diabetes, the number of years in school, age at baseline (continuous), HbA₁c, LDL cholesterol, and BMI were included in the final models. Additional variables that were considered for inclusion but did not meet the a priori criteria were race, Hispanic ethnicity, years of follow-up, smoking status, waist circumference, waist-to-hip ratio, intra-abdominal fat, subcutaneous fat, hypertension, retinopathy, nephropathy, total cholesterol, HDL, and triglycerides. Interaction terms were tested in the final models between diabetes status, sex, and race with the principal components and composite score to determine if diabetes status, sex, or race was a significant modifier of the relationship between the inflammatory markers and progression of CAC. All analyses were performed using SAS/STAT 9.2 software (SAS Institute Inc., Cary, NC).

RESULTS—Characteristics of the population stratified by diabetes status are reported in Table 1. Among subjects with type 1 diabetes, case subjects with significant CAC progression were older, had a longer duration of diabetes, higher HbA₁c, higher systolic and diastolic blood pressure, and higher triglycerides. Among subjects without diabetes, case subjects had a higher BMI, higher waist circumference, and higher measures of intra-abdominal and subcutaneous fat. Among both groups, the presence of CAC at baseline was significantly associated with CAC progression. No significant differences were found in either group for sex, race, smoking status, total cholesterol, HDL, or LDL.

Table 2 summarizes the distributions for the inflammatory markers stratified by diabetic status. Among subjects with diabetes, geometric means of sIL-2R, TNF-α, sTNFR2, sp-selectin, and MMP-3 were higher in case subjects than in control subjects. Geometric means of IL-6 and sIL-1RA were higher in case subjects than in control subjects among those without diabetes. The means of the other markers (hsCRP, sIL-6R, IL-18, ICAM-1, and OPG) were not significantly different or of borderline significance in both groups. Relative to subjects with a composite score of 0, case subjects were more likely to have elevated biomarkers in three or four components than control subjects in those with and without diabetes. The percentage of subjects who reported being ill within 24 h of the blood draw did not significantly differ by the composite score (P = 0.14; data not shown).

Table 3 reports the inflammatory marker patterns derived from the PCA. Values for sTNFR2, sIL-2R, and IL-18 were positively correlated with component 1; that is, subjects with higher scores on this component would have higher values of these inflammatory markers (factor loads of 0.84, 0.78, and 0.63, respectively). Similarly, scores on component 2 were correlated with the values of sIL-1RA and TNF-α (factor loads of 0.86, and 0.78, respectively). Finally, hsCRP and IL-6 were highly correlated with component 3 (factor loads of 0.84 and 0.78, respectively), and sp-selectin and sIL-6R were highly correlated with component 4 (factors loads of 0.83 and 0.58, respectively).

To determine the association of the inflammatory markers on progression of CAC, conditional logistic regression models were fit (as described above) for the principal components and the composite score as a measure of inflammatory burden. The results are presented in Table 4. In unadjusted analyses, components 1, 2, and 3 were modestly but significantly associated with progression of CAC. Component 4 was not significantly associated. After adjustment for age, systolic blood pressure, LDL, BMI, years in school, duration of diabetes, and HbA₁c, the association was similar for components 1 and 2, but the association was no longer statistically significant for component 3.

In separate models, the odds ratios (ORs) for the composite score were strongly associated with progression of CAC, even after adjustment, and showed a dose-response effect with the strongest effect of composite score 3–4. Interaction terms for diabetes, sex, and race with the principal components and for the composite score were tested in the final models. Diabetes, sex, and race were not significant modifiers (P > 0.05). Exclusion of subjects who reported being ill within 24 h of the blood draw did not substantially alter these results (data not shown).

CONCLUSIONS—These results demonstrate that markers of inflammation are prospectively associated with progression of CAC in a population of adults with and without type 1 diabetes. Two principal components were associated with the progression of CAC. The inflammatory markers that loaded strongly on these components were sTNFR2, sIL-2R, IL-18, sIL-1RA, and TNF-α. A composite score measuring inflammatory burden was also strongly associated with progression of CAC.

Prevalent coronary calcium has been associated with incident coronary heart disease events (12,13). Progression of CAC is significantly related to all-cause mortality independent of prevalent coronary calcium (14). No prospective studies have been published on the relationship between CAC and cardiac events in patients with type 1 diabetes; however, the EDC study found that CAC was strongly correlated with clinical CAD, myocardial infarction, and obstructive CAD (25). In subjects with type 1 diabetes, certain inflammatory markers (hsCRP, IL-6, and
Inflammation and CAC

Table 1—Characteristics of study subjects

| Variable                  | Type 1 diabetes (n = 306) | No diabetes (n = 156) |
|---------------------------|---------------------------|-----------------------|
|                           | Case subjects n = 137     | Control subjects n = 169 | P value |
|                           |                           |                       |         |
| Age (years)*              | 43 ± 8.1                  | 40 ± 7.7              | 0.0002  |
| Sex (% male)†             | 78 (56.9)                 | 91 (53.8)             | 0.59    |
| CAC present†              | 100 (73.0)                | 95 (56.2)             | 0.002   |
| Race†                     |                           |                       |         |
| White‡                    | 128 (93.4)                | 162 (96.4)            |         |
| Black                     | 3 (2.2)                   | 1 (0.6)               | 0.22    |
| Other                     | 6 (4.4)                   | 5 (3.0)               | 0.26    |
| Hispanic                  | 3 (2.2)                   | 7 (4.2)               | 0.34    |
| Years in school‡          | 16 (14–16)                | 16 (14–16)            | 0.93    |
| Years of follow-up*       | 2.5 ± 0.42                | 2.4 ± 0.34            | 0.06    |
| Smoking status†           |                           |                       |         |
| Never‡                    | 83 (61.0)                 | 108 (65.5)            |         |
| Past                      | 33 (24.3)                 | 43 (26.1)             | 1.00    |
| Current                   | 20 (14.7)                 | 14 (8.5)              | 0.10    |
| BMI (kg/m²)*              | 26.8 ± 4.7                | 26.5 ± 4.7            | 0.60    |
| Waist circumference (cm)* | 88.9 ± 12.7               | 87.3 ± 12.7           | 0.27    |
| Waist-to-hip ratio*       | 0.85 ± 0.08               | 0.83 ± 0.08           | 0.06    |
| Fat (mm³)§                 |                           |                       |         |
| Visceral                  | 39,423 (25,148–57,367)    | 36,720 (23,134–56,786) | 0.39   |
| Subcutaneous              | 138,157 (96,201–240,891)  | 135,761 (88,502–184,920) | 0.53 |
| Type 1 diabetes duration (years)*‖ | 29.2 ± 8.6 | 23.8 ± 9.0 | <0.0001 |
| HbA1c, (%)[|]              | 8.0 (7.2–8.9)             | 7.8 (7.0–8.4)         | 0.02    |
| Insulin dose (units/kg/day)*‖ | 0.62 ± 0.26             | 0.61 ± 0.26           | 0.88    |
| Retinopathy†              | 61 (46.6)                 | 44 (26.8)             | 0.004   |
| Nephropathy†              | 23 (16.8)                 | 12 (7.1)              | 0.008   |
| Hypertension†             | 89 (65.0)                 | 68 (40.2)             | <0.0001 |
| Blood pressure (mmHg)     |                           |                       |         |
| Systolic*                 | 125 ± 14.3                | 117 ± 14.1            | <0.0001 |
| Diastolic*                | 79 ± 9.2                  | 77 ± 8.3              | 0.03    |
| Cholesterol (mg/dL)*      |                           |                       |         |
| Total                     | 176 ± 31.3                | 176 ± 34.5            | 0.97    |
| HDL                       | 55 ± 16.9                 | 57 ± 17.8             | 0.43    |
| LDL                       | 102 ± 26.5                | 102 ± 29.1            | 0.96    |
| Triglycerides (mg/dL)§     | 83 (67–118)               | 74 (59–102)           | 0.02    |

*Data presented as mean ± SD, P value from t test. †Data presented as number (%), P value from χ² or Fisher exact test. ‡Referent category. §Data presented as median (interquartile range), P value from Wilcoxon rank sum test. ‖Among subjects with diabetes.

TNF-α) have been cross-sectionally associated with microvascular and macrovascular complications (5–8). In the EDC study, hsCRP was associated with an increased risk of CAD (9).

Only two prior studies from independent samples have examined the relationship between inflammation and progression of CAC (18,19) and did not find an association between hsCRP and progression (18) or incident (19) CAC after adjusting for other risk factors. Hokanson et al. (24) previously showed that interscan variability increases with increasing calcium volume scores, which may lead to biased estimates of the change in calcium scores over time. The definition of progression used for this report (≥2.5 square root transformed units) has been shown to be representative of actual progression and is <1% likely to be due to interscan variability (23,24). By using this approach, our study was better able to identify differences in markers of inflammation that were masked when using other approaches to assessing CAC progression.

A previous report from the CACTI cohort found that fibrinogen was significantly associated with the progression of CAC (OR 2.92 [95% CI 1.36–6.27]) in subjects with type 1 diabetes. In the nested case-control sample used for this study, log-transformed fibrinogen was not significantly associated with progression of CAC in the whole sample (P = 0.34) or in those with type 1 diabetes (P = 0.29; data not shown) after adjusting for other risk factors. The discrepancies are likely explained by the differences in the study designs between the two reports. The previous study involved samples from the entire cohort, and the definition of progression included...
subjects who developed clinical CAD during the follow-up period. For these reasons, we did not include fibrinogen in the PCA or models for this report. However, both reports lend support to the notion that an association exists between inflammation and progression of CAC.

TNF-R2 is typically expressed in immune system cells and is important for the activation and proliferation of certain types of T cells (26,27). The generation of the soluble receptor, sIL-2R, depends on cell activation (28), and serum levels have been used in studies as a measure of T-cell activation (29). IL-18 was initially described as a potent inducer of interferon-γ production and was subsequently realized to be important in T-cell differentiation (30). A previous analysis of data from a smaller nested case-control sample of the CACTI cohort (98 case subjects and 173 control subjects) found a significant association between sIL-2R and progression of CAC independently of other risk factors among subjects with and without type 1 diabetes (20). In the current study, these three cytokines loaded highly on component 1 and may reflect a T-cell activation construct associated with progression of CAC.

TNF-α and sIL-1RA loaded highly on component 2. sIL-1RA is produced primarily by the liver as an acute-phase protein and acts as a natural inhibitor of the effects of IL-1β (31). The diverse effects of TNF-α include cell death and proinflammatory changes to vascular endothelial cells, which promote leukocyte adhesion and thrombosis (27). IL-1 and TNF-α act synergistically to promote the inflammatory response (30); therefore, component 2, which is significantly associated with progression of CAC, may represent IL-1/TNF-α-mediated inflammatory processes.

hsCRP is an acute-phase protein produced in response to infection or physical trauma. Production of hsCRP is predominantly controlled by IL-6 (32). IL-6 is the primary regulator of the hepatic acute-phase response, is elevated in systemic inflammation, and increases with obesity (2). Some studies have not found any association between hsCRP and prevalence or severity of CAC (17,33,34) or CAC progression (18). The study by Kronmal et al. (19) found that the association between hsCRP and incident coronary calcium was attenuated by BMI. Other studies have also found an obesity effect, albeit specific to sex, where hsCRP was found to be associated with prevalent coronary calcium but not in women, after controlling for BMI (35,36). A recently published cross-sectional study by Raaz-Schrauder et al. (16) demonstrated that IL-6 was associated with prevalent coronary calcium but did not find a consistent association with hsCRP. Jenny et al. (15) found modest significant associations between prevalent coronary calcium and IL-6 and a

Table 2—Distribution of inflammatory markers

| Variable                  | Type 1 diabetes (n = 306) |   | No diabetes (n = 156) |   |
|---------------------------|---------------------------|---|-----------------------|---|
|                           | Case subjects n = 137     | Control subjects n = 169 | P value | Case subjects n = 67 | Control subjects n = 89 | P value |
| hsCRP (mg/L)*             | 1.6 (0.9–2.4)             | 1.4 (0.9–2.3)             | 0.26     | 1.4 (0.9–2.0)         | 1.2 (0.9–1.6)             | 0.05    |
| IL-6 (pg/mL)*             | 1.9 (1.3–2.9)             | 1.6 (1.0–2.6)             | 0.07     | 1.9 (1.2–2.8)         | 1.4 (0.9–2.2)             | 0.02    |
| sIL-6R (pg/mL)*           | 38,277 (29,695–47,933)    | 36,640 (31,183–45,637) | 0.23     | 37,437 (29,094–46,898) | 37,198 (29,879–45,142) | 0.90    |
| IL-18 (pg/mL)*            | 252.8 (194.9–316.2)       | 236.4 (175.5–305.2)       | 0.16     | 245.8 (197.4–293.3)   | 228.4 (170.3–301.0)       | 0.22    |
| sIL-1RA (pg/mL)*          | 7.6 (0.1–112.7)           | 7.7 (0.1–91.6)            | 0.99     | 15.6 (0.1–212.4)      | 3.7 (0.1–58.2)            | 0.005   |
| sIL-2R (pg/mL)*           | 1,047.5 (769.3–1,336.5)   | 889.0 (688.2–1,137.1)     | 0.001    | 746.5 (579.3–904.1)   | 683.3 (548.8–845.1)       | 0.13    |
| TNF-α (pg/mL)*            | 4.3 (2.6–8.2)             | 3.0 (2.0–6.2)             | 0.01     | 3.8 (2.2–7.3)         | 2.8 (2.1–5.8)             | 0.09    |
| sTNFR2 (pg/mL)*           | 2,955 (2,438–3,317)       | 2,576 (2,207–2,949)       | <0.0001  | 2,342 (2,003–2,660)   | 2,202 (1,926–2,592)       | 0.07    |
| ICAM-1 (ng/mL)*           | 273.7 (244.6–300.4)       | 266.8 (235.2–296.1)       | 0.35     | 260.3 (237.8–291.8)   | 256.1 (222.5–287.2)       | 0.68    |
| sP-selectin (ng/mL)†      | 109.1 ± 36                | 100.4 ± 33                | 0.03     | 94.7 ± 32             | 100.9 ± 32                | 0.24    |
| OPG (pmol/L)*             | 7.1 (6.0–8.2)             | 7.1 (6.1–8.5)             | 0.84     | 6.6 (5.5–7.7)         | 6.2 (5.2–7.6)             | 0.21    |
| MMP-3 (ng/mL)*            | 16.1 (9.7–24.8)           | 13.3 (8.9–19.6)           | 0.008    | 13.2 (10.2–18.1)      | 13.3 (8.9–19.7)           | 0.94    |
| Composite score ‡         | 0.006                     | 0.040                     | 0.04     | 0.030                 | 0.020                     | 0.28    |

*Data presented as geometric mean (interquartile range), P value from t test of log-transformed values. †Data presented as mean ± SD, P value from t test. ‡Data presented as number (%), P value from χ2 test. §Referent category.

Table 3—Inflammatory marker patterns derived by PCA

| Component 1 Markers | Component 2 Markers | Component 3 Markers | Component 4 Markers |
|---------------------|---------------------|---------------------|---------------------|
| sTNFR2              | sIL-1RA             | sCRP                | sP-selectin          |
| 0.84†               | 0.86†               | 0.84†               | 0.83†               |
| sIL-2R              | TNF-α               | IL-6                | sIL-6R              |
| 0.78†               | 0.78†               | 0.78†               | 0.58†               |
| IL-18               | IL-18               | sIL-1RA             | sIL-2R              |
| 0.63†               | 0.19                | 0.14                | 0.24                |
| TNF-α               | sP-selectin         | sIL-2R              | sTNFR2              |
| 0.30                | 0.12                | 0.13                | 0.12                |
| sIL-6R              | sTNFR2              | sTNFR2              | sTNFR2              |
| 0.21                | 0.12                | 0.12                | 0.11                |
| IL-6                | hsCRP               | sP-selectin         | IL-18               |
| 0.17                | 0.08                | 0.12                | -0.10               |
| sP-selectin         | sIL-6R              | IL-18               | IL-6                |
| -0.08               | -0.07               | 0.04                | 0.09                |
| HsCRP               | IL-6                | sIL-6R              | sIL-1RA              |
| 0.06                | 0.05                | -0.03               | -0.06               |
| sIL-1RA             | sIL-2R              | TNF-α               | HsCRP               |
| -0.01               | -0.03               | 0.00                | -0.02               |

*Pearson correlation coefficient between the inflammatory marker and the component. †Factor considered for interpretation of the component with load >0.4.
**Inflammation and CAC**

### Table 4—Multiple logistic regression of principal components with progression of CAC

| Variable                  | Crude* OR (95% CI) | Adjusted† OR (95% CI) |
|---------------------------|--------------------|-----------------------|
| **Model 1: Individual principal components** |                    |                       |
| Component 1               | 1.40 (1.13–1.74)   | 1.38 (1.08–1.77)      |
| Component 2               | 1.23 (1.01–1.50)   | 1.27 (1.02–1.59)      |
| Component 3               | 1.22 (1.00–1.49)   | 1.14 (0.90–1.46)      |
| Component 4               | 1.09 (0.90–1.33)   | 1.01 (0.80–1.27)      |
| **Model 2: Composite score** |                    |                       |
| Composite score           |                    |                       |
| 0                         | 1.00               | 1.00                  |
| 1–2                       | 1.77 (1.03–3.06)   | 1.94 (1.06–3.52)      |
| 3–4                       | 3.53 (1.86–6.67)   | 3.72 (1.80–7.69)      |  

P values for interactions terms: diabetes × principal components (P = 0.86; P = 0.23; P = 0.52; P = 0.11, respectively); diabetes × composite score (P = 0.48; P = 0.64, respectively). *Age-group, diabetes status, presence of CAC at baseline, and sex were included as the conditional variables to account for the matching. †Adjusted for systolic blood pressure, duration of diabetes (only in subjects with diabetes), years in school, age at baseline, HbA1c, LDL, and BMI. ‡Referent category.

In our study population, hsCRP and IL-6 were most highly correlated with component 3, which may represent a construct for the acute-phase response and which was not significantly associated with progression of CAC in the adjusted model (OR 1.14 [95% CI 0.90–1.46]). Although hsCRP and IL-6 were not independently associated with progression of CAC in the current study, they did contribute to the composite score, which had a stronger association with progression of CAC than the individual principal components.

IL-6/IL-6R trans-signaling has been shown in mice to promote endothelial adhesion molecules and macrophage infiltration into vascular lesions (37). P-selectin is an endothelial adhesion molecule that promotes leukocyte adhesion and migration (38). P-selectin and sIL-6R loaded highly on component 4 and may represent an endothelial adhesion and leukocyte migration component of CAC progression.

Similar to P-selectin, ICAM-1 promotes leukocyte adhesion and migration and has been suggested to be a biomarker of CAC burden (39). OPG is typically thought to inhibit atherosclerosis by acting as a decoy substrate for receptor activator of nuclear factor-κB ligand; however, in inflamed tissues, OPG enhanced MMP activity in vascular smooth muscle cells, which is important in atherosclerosis because MMPs degrade the matrix in atherosclerotic plaques (40). ICAM-1, MMP-3, and OPG loaded highly (factor loads >0.4) on multiple components, were dropped from the PCA analysis, and did not contribute to the composite score.

This study has several limitations. Not all potentially influential inflammatory markers were measured. However, owing to the common biologic pathways of many cytokines, we chose to combine markers into a composite score, which could strengthen a single measure with multiple possible pathways. These data were collected as part of a nested case-control study. Some bias might have existed in the selection of subjects for this sample. There were too few controls to get a true one-to-one match on age, sex, diabetes status, and baseline CAC. Frequency matching resulted in a sample of control subjects that did not exactly match the case subjects on these important variables. However, we adjusted for the sampling using conditional logistic regression and additionally controlled for age as a continuous variable in the final models.

In summary, this study demonstrated a significant prospective association between inflammatory markers and progression of CAC in a population of adults with and without type 1 diabetes. The PCA identified two components, potentially representing T-cell activation and IL-1/TNF-α-mediated inflammatory processes, that were independently but modestly associated with progression of CAC. Subjects with at least one elevated marker in three to four of the components were 3.7-times more likely to experience progression of CAC during the follow-up period. These results lend support to the idea that measurement of inflammatory markers may improve risk estimation of CVD progression. They also demonstrate that consideration should be made for inflammatory burden when considering the association of inflammatory markers and CAC. Considering the increased burden of CVD in the type 1 diabetic population, anti-inflammatory therapies could prove to be a valuable tool in this population.

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A.C.A. analyzed the data and wrote the manuscript. G.L.K. and J.K.S.-B. designed the case/control study, collected data, assisted in the analysis, and edited the manuscript. R.P.T. designed the case/control study, performed the marker assays, and edited the manuscript. D.M.M., J.E.H., and M.J.R. designed the case/control study and edited the manuscript. J.K.S.-B. is the guarantor for this work and, as such, had full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the analysis.

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