Disinfection effect of microwave radiation on Bacillus subtilis as indicator organism on contaminated dental stone casts under dry and wet conditions

Desinfektionsleistung von Mikrowellen gegenüber Bacillus subtilis als Indikatororganismus auf kontaminierten Dentalabgüssen unter trockenen und feuchten Bedingungen

Abstract

Objective: The disinfection of dental stone casts using microwave radiation has been shown, but doubts remain regarding its efficacy under various conditions. The aim of the present study was to evaluate the efficacy of microwave disinfection on wet and dry dental stone casts contaminated by a resistant microorganism.

Material and methods: In this in vitro study, 34 stone half-casts were prepared, contaminated with Bacillus subtilis and divided into two groups. After drying the specimens of one group for 15 minutes using 450 W microwave energy, all the wet and dry specimens were exposed to 900 W microwave energy for 5 minutes. Specimens were then individually transferred to nutrient broth culture medium and after 10 minutes, one milliliter from each tube was cultured in nutrient agar media for 24 hours, and the colonies were counted in CFU/mL. Data were analyzed using multifactorial ANOVA and Bonferroni tests.

Results: Casts in both wet and dry groups were disinfected to a high level (6 log), with no statistically significant differences between them (P<0.05).

Conclusion: According to the results, microwave irradiation can disinfect dental stone casts to a high degree, irrespective of moisture level. However, the result should be confirmed by exploring with other species of resistant microorganisms.

Keywords: Bacillus subtilis, disinfection, stone cast, microwaves, moist and dry conditions

Zusammenfassung

Zielsetzung: Obwohl die Desinfektion von Dentalabgüssen mit Mikrowellen bereits gezeigt werden konnte, bestehen Zweifel hinsichtlich der Wirksamkeit unter verschiedenen Bedingungen. Daher sollte die Wirksamkeit vergleichend auf nassen und trockenen Dentalabgüssen nach Kontamination mit Bacillus subtilis in vitro geprüft werden.

Material und Methode: 34 Zahnabgüsse wurden präpariert, mit B. subtilis kontaminiert und in zwei Gruppen aufgeteilt. In der ersten Gruppe wurden die Prüfkörper mit 450 W Mikrowellenenergie für 15 min getrocknet. Danach wurden die getrockneten und parallel die feucht belassenen Prüfkörper für 5 min mit 900 W Mikrowellenenergie expo-niert, für 10 min in Nährbouillon überführt, davon 1 ml auf Agar ausplat-tiert und nach 24 h die Kolonien gezählt. Die Daten wurden nach mehrfaktorieller Varianzanalyse mittels ANOVA mit dem Bonferroni-Test analysiert.

Ergebnisse: In beiden Gruppen wurde eine Abtötung um 6-log ohne signifikanten Unterschied erreicht (P<0.05).
**Introduction**

Microwave disinfection has been shown to be a viable alternative technique to chemical disinfection, and many resistant microorganisms that cannot be eliminated even with longer contact times in disinfectant solutions can be effectively disinfected using a microwave oven [1], [2], [3]. In the case of contaminated stone casts, evidence shows that microwave irradiation at energy levels of 800–900 W for 5 minutes is also an effective disinfection method. Despite the inherent weakness and sensitivity of stone casts to loss of moisture [4], microwave radiation has little adverse effect on physical and mechanical properties of the stone [5], [6].

In order to recommend the use of microwave irradiation as an effective disinfection technique based on EN 1040 standards [7], it is necessary to evaluate the effect of 900 W microwave irradiation as a safe energy level against an optimum indicator bacteria such as *Bacillus subtilis* [8], along with the effect of wetting or drying of casts on disinfection efficacy. It was shown previously that in the case of contamination with *B. subtilis*, the stone cast could only be disinfected (but not sterilized) in a dry condition using a microwave energy level of 850 W for 10 minutes. The cast was sterilized only when immersed in water or NaCl solution while applying the same energy level and duration of irradiation [9]. This protocol, however, has detrimental effects on the dimensional accuracy and surface smoothness of the casts. Meanwhile, there is evidence showing no significant difference between dry and moist specimens in terms of eliminating non-indicator microorganisms at a 600 W energy level [10]. It seems that the testing conditions may have considerably influenced the outcomes of these studies, and deleterious boiling conditions may not necessarily be required for effective disinfection of the dental stone casts, which is enough prior to sterilization [7].

The present study was designed to evaluate the efficacy of microwave irradiation in adequately disinfecting stone casts contaminated by indicator bacteria under wet and dry conditions.

The hypotheses of the present study were:

1. Microwaves irradiation with 900 W energy levels can effectively disinfect casts contaminated with *B. subtilis*.
2. Wetting or drying of casts has no effect on the quality of disinfection.

**Materials and methods**

**Pilot study**

A pilot study was prepared to explore the effective energy level and exposure duration of microwave irradiation. For the experiment, spherical stone specimens 10 mm in diameter (n=205) were separated from top of the alveolar ridge of a stone dental cast which had spherical beads in the place of teeth. A model was prepared by adhering 10 similar plastic spherical beads on the alveolar crest of an edentulous dental arch model made of wax, converting it to an acrylic cast by the conventional flashing technique used for resin acrylic dentures, and duplicating the model by making a conventional irreversible hydrocolloid impression and pouring with dental stone type III. All procedures in the pilot and main experiments were carried out in a sterile and microbiologically safe manner: bowls, mixing spatulas and impression trays were disinfected with 70% ethanol, and tweezers, pincers, and wax knives were flame sterilized when necessary.

The stone specimens were inoculated by soaking in broth cultures containing $10^7$–$10^8$ CFU/mL *B. subtilis* (ATCC 5127) for 10 min, except for 2 negative control specimens. The specimens, after removing from broth, were divided into 20 study groups of specimens, each for different irradiation durations and irradiation powers with 3 specimens as positive controls. The irradiation of the experimental groups was carried out at energy levels of 300, 450, 600 and 900 W for 5, 7, 10, 15 and 20 minutes after the specimens were allowed to absorb the appearing broth on surface. After treatment and cooling, each of the experimental and control specimens were placed into separate tubes of nutrient broth for 10 minutes, and 1 mL aliquot from each tube was cultured in nutrient agar for 24 hours at 37°C. Colonies were counted as CFU/mL. The results were analyzed using multifactorial ANOVA and Bonferroni tests.

**Main study**

The dental stone cast specimens were fabricated by pouring silicone molds of a maxillary dental arch. Type IV stone (Elite Model, Zhermack, Italy) was used to pour 17 casts in a sterile manner based on manufacturer’s instructions. The casts were separated into halves before final setting, using a sterile stone knife for simple separation after hardening.
A standard strain of *B. subtilis* (ATCC 6633; provided by the Pasteur Institute, Tehran, Iran) was used in the present study, and a microbial suspension (10⁷–10⁸ CFU/mL) was prepared to contaminate the stone casts. The half-cast specimens were immersed in the microbial suspension for 10 minutes, except for one negative control specimen, which was included in the study without contamination to validate the sterility of the study procedures (Figure 1). After the specimens were retrieved from the suspension, they were randomly divided into two study groups of n=15 each. The specimens in one group were dried in a microwave oven (Samsung PG 3210, China) for 15 minutes at 450 W and a frequency of 2450 MHz. Then they were irradiated at 900 W for five minutes, similar to the wet group. The wet specimens were irradiated immediately after removal from the broth tubes and absorption of their surface liquid (Figure 1 and Figure 2).

Three positive control specimens were selected with contamination and without sterilization to confirm the vitality of microorganisms, the validity of the contamination the casts, and to determine the initial counts of microorganisms on the study specimens. After irradiation, each specimen was placed in a sterile beaker containing 100 mL of sterile nutrient broth (34 beakers total). The beakers were shaken with great care, and after 10 minutes, 0.01 mL aliquots were separately cultured on agar plates. To increase the accuracy of initial colony counts, contaminated broth was serially diluted from 10⁷–10¹ CFU/mL. All the plates were incubated at 37°C for 24 hours and *B. subtilis* colonies were counted in terms of CFU/mL (Figure 3). The results were analyzed using ANOVA and t-tests.

### Results

**Pilot study**

Table 1 and Figure 4 show that applying microwaves for 20 minutes resulted in 6-log reductions in viability on contaminated specimens, regardless of the energy level, but when irradiation duration was reduced to 15 minutes, 900 W were necessary to achieve this level of disinfection. The microbicidal effect (ME) in Table 1 was calculated by subtraction of the log₁₀ value of the counts after microwave treatment from the log₁₀ value of the counts without exposure to the disinfection procedures (the positive control). Energy level and irradiation duration produced significant effects individually and interacting with each other (P<0.001) (Data not shown).
Table 1: Mean counts (CFU/mL) and microbiocidal effect against *Bacillus subtilis* at different energy levels and irradiation duration

| IT± | MC+ | MC | ME± | MC | ME | MC | ME | MC | ME |
|-----|-----|----|-----|----|----|----|----|----|----|
| 0   | $5 \times 10^6$ | 0  | $5 \times 10^6$ | 0  | $5 \times 10^6$ | 0  | $5 \times 10^6$ | 0  | $5 \times 10^6$ | 0  |
| 5   | $5 \times 10^6$ | 4.63 | $5 \times 10^6$ | 0  | $5 \times 10^6$ | 0  | $5 \times 10^6$ | 0  | $5 \times 10^6$ | 0  |
| 10  | $3.2 \times 10^2$ | 5.19 | $1.31 \times 10^2$ | 4.57 | $1.69 \times 10^2$ | 4.46 | $4.9 \times 10^3$ | 3  | $7.7 \times 10^4$ | 4.8 |
| 15  | 0  | 6.69 | $3.5 \times 10^1$ | 5.14 | $4.1 \times 10^1$ | 5.07 | $7.7 \times 10^1$ | 4.8 | 0  | 6.69 |
| 20  | 0  | 6.69 | 0  | 6.69 | 0  | 6.69 | 0  | 6.69 | 0  | 6.69 |

±: Irradiation time (min), ∗: Mean count, #: Microbicidal effect
Main study

In the main experiment, microwave irradiation at a 900 W energy level was highly effective, as presented in Table 2. It shows a 6-log reduction in viability within 5 minutes, regardless of wet or dry conditions. The results of ANOVA and the t-test are depicted in Table 3 and Figure 5, and did not reveal significant differences in *B. subtilis* colony count between the dry and wet groups (P<0.05). Median CFU decreases of 6.31 and 6.67 log were observed in moist and dry conditions, respectively.

**Table 2: Colony count of *Bacillus subtilis* and microbial effect (ME) after irradiation of wet and dry casts at 900 W for 5 minutes**

| No± | Untreated | Microwave irradiated |
|-----|-----------|----------------------|
|     | MC*       | ME#                  |
| 0*  | 5.7×10³   |                      |
| 1   | 1.6×10¹   | 6.56                 |
| 2   | 1.1×10¹   | 6.72                 |
| 3   | 3×10¹     | 6.28                 |
| 4   | 1.5×10¹   | 6.58                 |
| 5   | 3.1×10¹   | 6.27                 |
| 6   | 2.1×10¹   | 6.44                 |
| 7   | 1.9×10¹   | 6.48                 |
| 8   | 1.2×10¹   | 6.68                 |
| 9   | 1.1×10¹   | 6.72                 |
| 10  | 3.8×10¹   | 6.18                 |
| 11  | 6×10¹     | 5.98                 |
| 12  | 8.9×10¹   | 5.81                 |
| 13  | 1.4×10¹   | 6.61                 |
| 14  | 3.3×10¹   | 6.24                 |
| 15  | 1.3×10¹   | 6.65                 |
| Median | 2.8×10¹   | 6.31                 |

* Positive control, ±: Specimens number, -: Microbial count, #: Microbicidal effect

**Table 3: Results of ANOVA for *Bacillus subtilis* counts (CFU/mL) after irradiation in wet and dry cast conditions**

|        | Number | Mean     | SD      | t       | Degrees of freedom | P   |
|--------|--------|----------|---------|---------|--------------------|-----|
| Wet    | 15     | 2.8×10¹  | 2.17×10¹| -1.32   | 28                 | 0.195|
| Dry    | 15     | 1.93×10¹  | 1.27×10¹|         |                    |     |

In addition, the evaluation of gypsum specimens at 3x magnification did not reveal any holes, cracks, or color changes on the surface.

**Discussion**

In spite of the high numbers of *B. subtilis* that were needed to show increased log reduction of microorganisms, the irradiation of contaminated dried and moistened stone casts resulted in a high level of disinfection. This striking 6-log decrease can be achieved in 5 minutes by a household microwave oven at 900 W, which microbicidally exceeds the requirements of EN1040 standards [7]. In the pilot experiment, this level of disinfection was not achieved using 900 W in less than 15 minutes; however
another study found that 850 W of microwave irradiation for 10 minutes resulted in sterilization with the stone cast immersed in water or a 40% concentration of NaCl solution [9]. This lethal effect on B. subtilis was previously achieved at a frequency of 2450 MHz with acrylic specimens, but using 650 W for 3 minutes under immersion in water [11]. Difference in effective energy levels and irradiation times in these experiments can be related to different strains of microorganisms and techniques used. The current experiment showed no significant difference in the efficacy of microwave radiation exposure on moistened and dried stone casts. This was also observed in another study with different microorganisms at 600 W [10]. The moistened casts had been irradiated after removing from broth media as soon as the surface moisture had been absorbed in that and current studies. In addition to the non-thermal effects of microwaves that affect the biochemistry of cells and can inhibit or eliminate microorganisms [12], the thermal effect of microwaves is the other factor that results in killing the microorganisms. The thermal effect requires the presence of polar molecules such as water or ionic molecules [13], and makes bactericide possible at lower energy levels. Due to drying of the specimens during the first minutes of irradiation and the absence of water molecules (no thermal effect), the total elimination of microorganisms takes longer. As observed in the current pilot study and in a study by Abbas et al. [9], immersed specimens can be sterilized by 10 minutes of microwave irradiation, in comparison to the dry group that was only disinfected. The mechanical and physical properties of stone casts when irradiated without immersion in water or other liquids at 800–900 W for 5 minutes were not detrimentally affected [5], [6], [14], but immersion in water or 40% NaCl during the irradiation resulted in boiling the cast, negatively affecting the surface hardness and smoothness, as well as dimensional accuracy, as has been shown previously [9], [15].

The microorganism selected in the present study is one of the most resistant pathogens; it can convert into the spore form to tolerate harsh conditions. Among some typical indicator bacteria such as Bacillus cereus, Staphylococcus aureus, Bacillus subtilis var. nigeri, Bacillus stearothermophilus, Bacillus pumilus E, B. subtilis is considered to be an optimum indicator to evaluate the efficiency of microwave radiation [8]. It is recommended that further studies should be carried out with other strains of B. subtilis and other resistant species that can be used as indicator bacteria.

**Conclusions**

Considering the conditions and limitations of the present study, it was shown that it is possible to disinfect casts contaminated with B. subtilis using microwave ovens. This technique can be useful in infection control, as it provides high-level disinfection of casts and dies.

**Notes**

**Competing interests**

The authors declare that they have no competing interests.

**Acknowledgement**

The authors are very grateful to the Research Vice Chancellor of Tabriz University of Medical Sciences for supporting this study.

**Ethical considerations**

All stages of this study were performed on a standard strain with in vitro conditions.

**References**

1. Walsh SE, Maillard JY, Russell AD. Ortho-phthalaldehyde: a possible alternative to glutaraldehyde for high level disinfection. J Appl Microbiol. 1999 Jun;86(6):1039-46. DOI: 10.1046/j.1365-2672.1999.00791.x
2. Berg E, Nielsen Ø, Skau N. High-level microwave disinfection of dental gypsum casts. Int J Prosthodont. 2005 Nov-Dec;18(6):520-5.
3. Najdovski L, Dragas AZ, Kotnik V. The killing activity of microwaves on some non-sporogenic and sporogenic medically important bacterial strains. J Hosp Infect. 1991;19(4):239-47. DOI: 10.1016/0195-6701(91)90241-Y
4. Luebke RJ, Chan KC. Effect of microwave oven drying on surface hardness of dental gypsum products. J Prosthet Dent. 1985;54(3):431-5. DOI: 10.1016/0022-3913(85)90566-9
5. Hasan H. The effect of microwave disinfection on tensile strength of dental gypsum. Al-Rafidain Dent J. 2008;8:213-7.
6. Silva MA, Vitti RP, Consani S, Sinhoreti MA, Consani RL. Linear dimensional change, compressive strength and detail reproduction in type IV dental stone dried at room temperature and in a microwave oven. J Appl Oral Sci. 2012 Sep-Oct;20(5):588-93. DOI: 10.1590/S1678-77572012000500016
7. antiseptics SP-ECda. Quantitative suspension test for the evaluation of basic bactericidal activity of chemical disinfectants and antiseptics – test method and requirements (phase1), Polish Committee for Standardization (PKN); 2000.
8. Wu Q. Effect of high-power microwave on indicator bacteria for sterilization. IEEE Trans Biomed Eng. 1996 Jul;43(7):752-4. DOI: 10.1109/10.503183
9. Abass S, Mahmood M, Khalaf B. Effect of microwave irradiation on disinfection, dimensional accuracy, and surface porosity of dental casts. Mustansiria Dent J. 2011;8(2):177-87.
10. Robati Anaraki M, Akhi MT, Pirzadeh T, Moslehfard E, Ghanati H, Mosavi A, et al. Efficacy of microwave disinfection on moist and dry dental stone casts with different irradiation times. Adv Biosci Clin Med. 2015;3(3):40.
11. Mima EG, Pavarina AC, Neppelenbroek KH, Vergani CE, Spolidorio DM, Machado AL. Effect of different exposure times on microwave irradiation on the disinfection of a hard chairside resin. J Prosthodont. 2008 Jun;17(4):312-7. DOI: 10.1111/j.1532-849X.2007.00277.x
12. Banik S, Bandyopadhyay S, Ganguly S. Bioeffects of microwave - a brief review. Bioresour Technol. 2003 Apr;87(2):155-9. DOI: 10.1016/S0960-8524(02)00169-4

13. Michaelson SM. Effects of exposure to microwaves: problems and perspectives. Environ Health Perspect. 1974 Aug;8:133-55. DOI: 10.2307/3428196

14. Robati Anaraki M, Moslehi Fard E, Bahari M, Mahboubi S. Effect of repeated microwave disinfection on surface hardness and dimensional accuracy of two dental stone materials. Adv Biosci Clin Med. 2015;3(1):17.

15. Onizuka T, Kamimura N, Kajiwara H, Nakashima A, Suenaga K, Jimi T. Influence of boiling water treatment on surface roughness and surface microstructure of set gypsum (dental stone). Dent Mater J. 1995 Dec;14(2):245-55. DOI: 10.4012/dmj.14.245

Corresponding author:
Tahereh Pirzadeh
Department of Microbiology, Medical School, Tabriz University of Medical Science, Golgasht Ave., Tabriz, Eastern Azerbaijan, Iran, Phone: +989141014516, Phone/Fax: +984133364661
t_pirzad@yahoo.com

Please cite as
Robati Anaraki M, Mahboubi S, Pirzadeh T, Lotfipour F, Torkamanzad N. Disinfection effect of microwave radiation on Bacillus subtilis as indicator organism on contaminated dental stone casts under dry and wet conditions. GMS Hyg Infect Control. 2017;12:Doc09. DOI: 10.3205/dgkh000294, URN: urn:nbn:de:0183-dgkh0002944

This article is freely available from
http://www.egms.de/en/journals/dgkh/2017-12/dgkh000294.shtml

Published: 2017-08-08

Copyright
©2017 Robati Anaraki et al. This is an Open Access article distributed under the terms of the Creative Commons Attribution 4.0 License. See license information at http://creativecommons.org/licenses/by/4.0/.