Dissecting Source-Sink Relationship of Subtending Leaf for Yield and Fiber Quality Attributes in Upland Cotton (\textit{Gossypium hirsutum} L.)

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Abstract: Photosynthesis as a source is a significant contributor to the reproductive sink affecting cotton yield and fiber quality. Moreover, carbon assimilation from subtending leaves adds up a significant proportion to the reproductive sink. Therefore, this study aimed to address the source-sink relationship of boll subtending leaf with fiber quality and yield related traits in upland cotton. A core collection of 355 upland cotton accessions was subjected to subtending leaf removal treatment effects across 2 years. The analysis of variance suggested a significant effect range in the source-sink relationship under subtending leaf removal effects at different growth stages. Further insight into the variation was provided by the correlation analysis and principal component analysis. A significant positive correlation between different traits was observed and the multivariate analysis including hierarchical clustering and principal component analysis (PCA) categorised germplasm accessions into three groups on the basis of four subtending leaf removal treatment effects across 2 years. A set of genotypes with the lowest and highest treatment effects has been identified. Selected accessions and the outcome of the current study may provide a basis for a further study to explore the molecular mechanism of source-sink relationship of boll subtending leaf and utilization of breeding programs focused on cotton improvement.

Keywords: source-sink relationship; subtending leaf; cotton boll; multivariate analysis; upland cotton

1. Introduction

Cotton, \textit{Gossypium} spp., is widely accepted as the major fiber source globally and is cultivated across most of the tropical and subtropical parts of the world [1]. Among the prominent cotton producers, China, USA, India, Pakistan, and Uzbekistan are considered major shareholders [2,3]. The major limitations in cotton productivity are the biotic and abiotic stresses, including drought, heat, salinity, insect pests, diseases, and weeds, particularly during the early development stages [4]. Cotton is generally considered a resilient crop against stressors. However, viral infestations such as the Cotton Leaf Curl Virus Disease (CLCuD) have become major threats during the last two decades due to heavy infestations across South Asia and some parts of the USA [5]. Additionally, cotton growth and development may be highly challenged due to future climate change scenarios [6]. Upcoming climate changes are likely to play a prominent role in the change in...
cotton crop productivity patterns either positively or/and negatively. Fluctuating temperatures may significantly influence cotton growth and development due to a higher impact on respiration, transpiration rate, and photosynthesis, ultimately influencing fruit production [7].

The fiber quality attributes, including fiber length, strength, fineness, uniformity, and elongation, are the key desirable factors for optimizing textile processing and producing high-quality textile products [8]. A higher lint production is also considered as one of the most critical factors among major cotton production systems focusing on enhancing profitability [9]. Several cotton yield components play a significant role in enhancing lint yield, including bolls per plant, boll weight, seeds per boll, and ginning out percentage [10,11]. During the recent era, most of the improved cotton varieties being cultivated face a narrow genetic base due to the extensive unidirectional artificial selection by breeders for yield enhancement [12]. It became a serious concern nowadays, as narrow or limited genetic diversity may lead to a deficient allelic availability for a continued genetic improvement of cotton [13,14]. Hence, it is worth understanding the genetic basis of various domesticated traits and maintaining diversity among species to be utilized as a source in future breeding programs aimed at cotton improvement [14–16].

Since fibers in cotton are exclusively acclaimed as natural fibers and are made up of cellulose, with a polysaccharide 1,4-d-glucopyranose structural unit [17]. It is assumed that there must be a fundamental linkage among photosynthate assimilation, storage, and plant growth in terms of their strong relation as a source-sink [18]. Hence, a potential connection between carbon utilization and transport from photosynthate assimilation source tissues to the non-photosynthetic tissues is considered a sink [19]. Furthermore, fiber quality in cotton, including length, strength, and fineness, is extensively influenced by the photosynthate concentration in assimilates. During favorable conditions in the presence of adequate nutrients, light, and moisture, the cotton boll under-development may be assumed to acquire almost 60% or more assimilates required for fiber growth solely through subtending the leaf nearest to it [20,21]. Translocation becomes essential through other plant parts to supplement the remaining 40% development [22]. Therefore, an early vegetative growth is preferred in order to overcome the increased competition of different bolls as sinks are required to get translocated during the upcoming reproductive phase. Many published reports concluded that until the 12th node, the vegetative growth rate is usually enhanced, resulting in the derived carbohydrate transport towards reproductive units (bolls), unless any significant translocation among branches is accomplished [23,24].

Around 60–87% carbon in mature cotton boll is detained through CO$_2$ assimilation across the boll development, and as this process is carried out, the subtending leaf has a significant contribution for photosynthate accumulation, ultimately leading to increased biomass in seed cotton [20,21,24]. In this process, bolls and subtending leaves are linked in a sink-and-source-relationship to utilize photosynthate accumulation [25,26]. We can interpret their relationship as coordination among vegetative and reproductive growth phases in cotton with a substantial impact on the yield and quality of cotton produce [27]. Numerous studies on source-sink relationships have reported that an earlier and efficient sink development may lead to a robust and more substantial reproductive growth potential [28]. Higher yields are attributed to the substantial photosynthesis rate within the functional leaves, which is considered a source activity and is essential and effective for efficient photosynthates distribution through the reproductive parts [29,30].

A competition prevails for photosynthates across different plant development stages (Stewart 1989). The leaf is assumed to play an essential role in plant growth and development and is known as “powerhouse” in the plants due to its functionality as food synthesis in the presence of chlorophyll (Gitelson et al. 2003). Numerous leaf traits have been reported to show a substantially significant role in yield [31]. Considering the cotton plant as a reference, the leaf life cycle starts with bud initiation, acting as a carbohydrate exporter within a few days [32]. As the boll emerges, it starts extracting its maximum nutrition mostly from its subtending leaf [33].
The multivariate analysis has been extensively used as an important and mostly primarily acceptable method to estimate genetic variability and diversity in a more accurate manner [34]. It can also determine possible patterns of dissimilarities and potential genetic relationships within a specific germplasm collection [35]. The multivariate analyses have been extensively utilized globally by different crop improvement programs [36] on various crops such as wheat [35], maize [37,38], and sorghum [39].

The present-day improved cotton cultivars are assumed to be developed using limited germplasm sources as parents [40]. Due to the lack of extensive studies to explore the relationship between source-sink so far, this has made it difficult for a clear understanding in this regard and we are still handicapped in developing an optimal rationale for source-sink relationship manipulations. Hence, the current study has been planned to investigate the source-sink relationship of subtending leaves with boll influencing fiber quality and yield traits of upland cotton accessions during the years 2018 and 2019. The main objectives of the current study were aimed at: (a) The assessment of variability and diversity of the studied germplasm in response to the source-to-sink relationship; (b) Exploration of genotypes expressing minimum and maximum source-sink dependency. This study will be helpful in providing a deep insight into the source-sink relationship of subtending leaf with its boll and effect on yield and fiber quality. It will also provide a baseline for possible source-to-sink manipulations for cotton improvement yield and fiber quality.

2. Materials and Methods

A diverse collection containing 355 upland cotton accessions taken from the gene bank of the Institute of Cotton Research of CAAS has been considered plant material and utilized for the study. The collection comprised 331 cultivars developed and originated from China, whereas 24 cultivars were introduced from different countries. The Chinese accessions have been further subdivided into the five groups concerning the ecological areas of their origin, including 162 accessions from the Yellow River region (YR), 51 accessions from the Yangtze River Region (YZR), 98 accessions from the northwest in the land region (NW), 20 accessions from the Liaoning province (LN), and the foreign group consisted of 20 accessions from the USA, while four accessions were from countries of Central Asia. These cotton accessions have been planted following a triplicated randomized complete blocked design (RCBD) in the factorial arrangement at the field area of CRI, Anyang Henan during the sowing season in 2018 and 2019. Sowing was carried out on 30 April during both years viz., 2018 and 2019. The plot size was maintained as 3.5 m length of each accession in each replication and for each treatment.

After germination, thinning was carried out to maintain the plant population by keeping 10 plants in each row. The chemical control was applied at peak flowering and the boll setting period, whereas all other agronomic and plant protection practices were practiced following local recommendations. Five guarded plants from each line were selected randomly and tagged to be considered for further study. The start of the blooming flower tagging in all the experimental plots was carried out beginning from July 10, onward to August 10. Each flower during this period has been tagged from its blooming date. The boll subtending leaf removal has been carried out for the tagged leaves, as they reach 35 the days’ bolls represented as T₁, 40 days’ bolls as T₂, 50 days’ bolls as T₃, 60 days’ bolls as T₄, and one check as the control where the subtending leaf was kept unremoved (Table 1). The subtending leaf removal was carried out by hand on 20 bolls from each replication per accession from each treatment from the five selected and tagged plants. On maturity, picking was carried out to pick 20 bolls from every five plants for each treatment. The fiber quality for the nine traits was recorded for the picked bolls using a high volume instrument (HVI) in the Laboratory of Quality and Safety Risk Assessment for Cotton Products (Anyang), Ministry of Agriculture, People’s Republic of China. The traits considered for the current study included boll weight (BW), seed weight (SW), ginning outturn (GOT%), fiber weight (FW), fiber length (FL), fiber uniformity (FU), fiber fineness (MIC), fiber strength (FS), and fiber elongations (FE).
Table 1. Description of subtending leaf removal of 355 cotton accessions to determine the source-sink relationship.

| Treatment | Description                      | Number of Entries |
|-----------|----------------------------------|-------------------|
| Control   | No Subtending Leaf Removal        | 355               |
| T1        | Subtending Leaf removal after 35 Days boll age | 355               |
| T2        | Subtending Leaf removal after 40 Days Boll age | 355               |
| T3        | Subtending Leaf removal after 50 Days boll age | 355               |
| T4        | Subtending Leaf removal after 60 Days Boll age | 355               |

The effect of source (subtending leaves) to sink (boll) was estimated by comparing different treatments (removal of subtending leaves at multiple growth stages as shown in Table 1) with the control. Additionally, differences were considered as the source-sink effect of the corresponding trait.

\[
\text{Subtending Leaf Effect (E)} = \text{Control} - \text{Treatment (Mean)}
\]

where \( E \) is the subtending leaf effect or tolerance indice, control is the treatment with unremoved subtending leaf, whereas treatment is used for the corresponding treatment with the removed subtending leaf on different dates post anthesis (DPA).

The effects E1, E2, E3, and E4 were calculated using the same formula as above with the recorded values for each trait under study and their corresponding control values across both years.

**Statistical Analysis**

The calculated source-sink effects of 355 accessions for the E1–E4 effects across 2 years, as means of the ramets obtained for each genotype, were subjected to a combined analysis of variance of genotypes and source-sink effects across years using the general linear model (GLM), since the missing data from different replications over the treatments and years did not permit the repeated measures ANOVA for each treatment and year of the experiment. Factors were analyzed individually without interactions to avoid model complications that make it inappropriate. The expected mean squares were based on a random effects model for genotypes, treatments, and years.

The multivariate analysis was performed including basic statistics, PCA, and cluster analysis categorizing genotypes, years, and treatments for the source-sink effect and its relationship to the various variables under study. All the analyses were carried out using two statistical software packages: SAS-JMP Pro 15 (SAS Institute Inc., Cary, NC, USA, 1989–2019) and R 3.4.0.

**3. Results**

The variation existing for the removal of subtending leaf effects in the core germplasm of upland cotton (355 accessions) was estimated using the mixed linear model. The four treatment effects of subtending leaf removal derived for each trait were compared to dissect the source-sink relationship of subtending leaf with the yield and fiber quality. The differences were then termed as a source-sink effect for each trait under study. The summary statistics of the data collected is revealed in Figure 1. It showed that a high range of values existed regarding the removal of subtending leaf treatment effects in year 2018 as compared to 2019. Furthermore, the yield related traits particularly SW, BW, FW, and GOT showed more variation effects across both years in comparison to the fiber quality traits.
The overdispersion parameter estimated by the maximum likelihood using the generalized linear model (GLM) for fiber quality and yield related traits in 355 upland cotton accessions, under four treatment effects (E1–E4) across 2 years (2018 and 2019), are represented as Supplementary Tables S1–S9. From these results, the effect summary and effect tests have been concluded, since ANOVA (Table 2) suggested significant L-R chi-square values along with Log Worth values as represented in the summary effect plots in Table 2 among the genotypes for all traits. Whereas, the treatment effects have been significant for yield traits viz., boll weight (BW), fiber weight (FW), ginning outturn (GOT), and seed weight (SW). In contrast, fiber quality traits viz., fiber length (FL), fiber uniformity (FU), micronaire (MIC), fiber strength (FS), and fiber elongation (FE) showed non-significant treatment effects for the source-sink relationship. Furthermore, ANOVA also depicted a significant variation within the growing seasons viz., 2018 and 2019 for BW, FW, SW, FS, and FE. The significant variation of traits in different growing seasons depicted a significant environmental influence. However, GOT, FL, FU, and MIC did not show significant differences in the two seasons as the source-sink effect.

3.1. Correlations

The correlation and its distribution related to the source-sink effect of the nine traits in question were estimated to reveal their corresponding relationships (Figure 2). The upper triangle of the correlogram depicted correlations among the traits of the source-sink effect, while the lower triangle exhibited a scatterplot matrix of their density distributions. Source-sink effects of investigated traits displayed highly significant (≤0.0001) and high positive correlations between BW and FW (0.88), BW and SW (0.90), as well as FW and SW (0.84). MIC exhibited a significant negative correlation with FL and FS. GOT displayed a highly significant negative correlation with FL. Furthermore, significant negative correlations were observed between various source-sink effects of fiber quality and yield related traits. At the diagonal of the matrix, all the traits represent a normal state of frequency distribution in the shape of an histogram, which fits the data as suitable for the generalized linear model (GLM) with a normal distribution option. The lower triangle is for the depiction of the pairwise density distribution of traits. The narrowness of the ellipse shows the degree of correlation between a particular pair of traits in a specific cell. The narrow shaped ellipse with a diagonally oriented distribution shows the correlation between the paired traits, and the fairly round shaped ellipse with no diagonally oriented distribution shows no correlation.
Table 2. Analysis of variance (ANOVA) using the generalized linear model for the source-sink effect (E1–E4) across 2 years 2018 and 2019.

| Trait | Source             | DF  | L-R Chi Square | Log Worth | Summary Graph | p-Value |
|-------|--------------------|-----|----------------|-----------|--------------|---------|
| BW    | Genotype           | 354 | 966.18469      | 57.516    |              | <0.0001 *|
|       | Treatment Effect   | 3   | 54.951574      | 11.153    |              | <0.0001 *|
|       | Year               | 1   | 3.1565769      | 1.121     |              | 0.0756  |
| FW    | Genotype           | 354 | 937.32891      | 53.560    |              | <0.0001 *|
|       | Treatment Effect   | 3   | 39.597259      | 7.887     |              | <0.0001 *|
|       | Year               | 1   | 0.7938999      | 0.428     |              | 0.3729  |
| GOT   | Genotype           | 354 | 794.35135      | 35.114    |              | <0.0001 *|
|       | Treatment Effect   | 3   | 14.489381      | 2.637     |              | 0.0023 *|
|       | Year               | 1   | 4.1162681      | 1.372     |              | 0.0425 *|
| SW    | Genotype           | 354 | 960.37802      | 56.714    |              | <0.0001 *|
|       | Treatment Effect   | 3   | 39.621111      | 7.892     |              | <0.0001 *|
|       | Year               | 1   | 6.0551756      | 1.858     |              | 0.0139 *|
| FL    | Genotype           | 354 | 1030.1228      | 66.517    |              | <0.0001 *|
|       | Treatment Effect   | 3   | 32.998526      | 6.492     |              | <0.0001 *|
|       | Year               | 1   | 0.6609049      | 0.381     |              | 0.4162  |
| FU    | Genotype           | 354 | 997.69022      | 61.912    |              | <0.0001 *|
|       | Treatment Effect   | 3   | 2.4583405      | 1.603     |              | 0.4829  |
|       | Year               | 1   | 5.0287601      | 0.316     |              | 0.0249 *|
| MIC   | Genotype           | 354 | 1081.4485      | 73.956    |              | <0.0001 *|
|       | Treatment Effect   | 3   | 6.4663942      | 1.041     |              | 0.0910  |
|       | Year               | 1   | 1.3158987      | 0.600     |              | 0.2513  |
| FS    | Genotype           | 354 | 1073.4359      | 72.783    |              | <0.0001 *|
|       | Treatment Effect   | 3   | 2.8515984      | 2.548     |              | 0.4151  |
|       | Year               | 1   | 8.9126018      | 0.382     |              | 0.0028 *|
| FE    | Genotype           | 354 | 964.29565      | 57.254    |              | <0.0001 *|
|       | Treatment Effect   | 3   | 3.6658411      | 5.525     |              | 0.2999  |
|       | Year               | 1   | 21.852712      | 0.523     |              | <0.0001 *|

BW: Boll weight; FW: Fiber weight; GOT: Ginning outturn; SW: Seed weight; FL: Fiber length; FU: Fiber uniformity; MIC: Micronair; FS: Fiber strength; and FE: Fiber elongation; DF: Degree-of-freedom; L-R Chi Square: Likelihood ratio for Chi Square. * Significant at p < 0.01.
Figure 2. Scatterplot matrix to visualize several attributes by pairwise dependencies of nine fiber quality and yield related traits. The upper triangle matrix represents the correlations among source-sink effects of nine studied traits. Histograms at diagonal depict the shape of frequency distribution for the data of investigated traits, whereas the lower triangle matrix reveals the bivariate density distribution with ellipses between each pair of traits. The legends at the top right corner of the color gradient (red to blue) and the size of circles show the amount of correlation and \( \log p \)-value for significance threshold, respectively.

3.2. Principal Component Analysis (PCA)

The principal component analysis (PCA) can be considered as the most acceptable method, utilizing a multivariate approach, carrying out a dimension reduction across independent and interdependent variables retaining only those characteristics from a dataset contributing to its variance [41]. PCA was performed based on the correlation among source-sink effects of yield components and fiber quality traits. Nine PCs have been extracted from nine studied traits (Table S10) through PCA. A total of four PCs have been detected with Eigen values above 1, whereas the remaining components depicted minimal Eigen values.

The first component PC1 contributes to various traits towards variability, concluding that the source-sink effect of FW has a major contribution towards positive loading vectors (0.96435), followed by the source-sink effect of BW (0.94493), FW (0.95579), SW (0.91334), and MIC (0.31909) within the first component. It can be concluded that four major variables mentioned have a strong correlation with the first PC. The source-sink effect of FW has been found to have a strong correlation to this principal component. Hence, we can assume that PC1 can ultimately be considered as a measure to the source-sink effect for yield related traits. However, the remaining traits can be seen with a minimal contribution with positive loadings. PC2 exhibited source-sink effects of FL (0.71787) followed by FS (0.65328), FU (0.54193), FS (65328), and FE (0.33166). PC3 was related to GOT (0.87543) followed by FU (0.32163). PC4 revealed a maximum contribution towards FE (0.70599) and MIC (0.37958) as positive loadings, whereas FS (−0.46201) as negative loadings, respectively (Table S11).

As shown by the principal component analysis results, a total of 59.787% variance has been explained by the first three components (PCs). The yield, as well as fiber quality traits, are represented through factor map squared cosines or squared coordinates. It is
As shown by the principal component analysis results, a total of 59.787% variance is determined that the high values of squared cosines give a satisfactory contribution to the particular variable. Figure 3 shows that all the yield related traits cover PC1 except for GOT. However, all the fiber quality traits fall in PC2 except MIC, which is in PC1. With regards to the treatment effects, E1 and E4 fell in PC1 while, E2 and E3 fell in PC2. Furthermore, both years got maximum values in PC3.

![Figure 3. Squared cosines associated with the principal components for the studied traits, treatment effects (E1–E4), and years 2018 and 2019.](image)

In this study, most of the variables got a place nearest to the correlation eclipse in the summary plot, as shown in Figure 4. A significant representation of different variables across the first PC is depicted by the length of originating vectors. Almost all the fiber quality traits viz., FL, FS, FU, and FE except MIC, lied in the direction of PC2, whereas GOT, BW, FW, SW, and MIC lied in the direction of PC1. MIC and GOT are very close to the center with minimum contributions.

![Figure 4. Summary plots with (left) biplot between PC1 and PC2 displaying the distribution of 355 upland accessions across treatment effects and years; (right) contribution of different traits in variation for genotypes, treatment effects, and years.](image)

A more elaborative depiction of the traits in three PCs has been presented in Figure 5. In the biplot of PC1 and PC3, fiber yield traits viz., BW, FW, and SW lied close to each other concluding the presence of a high correlation among them. Meanwhile, in PC2 and PC3 biplot, fiber quality traits viz., FS, FE, and FU exhibited more correlation among themselves. These variables can be considered as critical in order to explain the variability in the studied dataset. The fiber fineness as MIC has not shown any relationship with any PC (Figure 5).
The principal component biplot drawn for the studied material indicated considerable variability among the studied treatments (Figure 4). A plot between PC1 and PC2 related to yield and fiber quality traits in a collection of 355 accessions has been clustered into three distinct groups encountering four subtending leaf removal treatment effects (E1–E4). For further clarification and details, the sub-populations have been represented by three different colors based on the yield contribution and fiber quality traits under study. It has been observed that the accessions which have been placed together are considered to have high similarity with each other.

3.3. Cluster Analysis

A scatter plot matrix, represented in Figure 6, has been drawn using factor scores as PC1 and PC2, and depicted a clear pattern for grouping of genotypes by observing three major distinct groups through clustering. The factors’ correspondence was further subjected to agglomerative hierarchical clustering to work out the Euclidean distance matrix through Ward’s method, and a dendrogram has been constructed based on the result. Numerous methods are available for the estimation of diversity within germplasm subjected to various contrasting environments. The two-way clustering has been executed through the AHMC method, resulting in a two-way cluster diagram and constellation plots. A scatterplot matrix has also been drawn to determine the densities composition of the cotton germplasm under different subtending leaf removal treatments at different boll ages. Depending on the clustering, we identified 10 genotypes, five with the high effect of source (subtending leaf) removal and five with the least effect of source (subtending leaf) removal. The selected genotypes have been presented in Table 3. These genotypes
can be further utilized in breeding programs to manipulate the source-sink relationship of subtending leaf in upland cotton.

Figure 6. Hierarchical clustering of 355 upland cotton accessions for yield related and fiber quality traits based on treatment effects across years 2018 and 2019.
Table 3. Top five selected genotypes for high and low source-sink effects for yield related and fiber quality traits.

| Treat. | Source-Sink 2018 | Yield Quality | Yield 2019 | Quality |
|--------|------------------|---------------|------------|---------|
| E1     | Low source-sink effect | Deltapine15 han2490 103030 dezimian531 jinmian10 | 2011SS emian17 qing1 lu890 | xinluzhong41 |
|        |                  | chaoyangmian1 zhong416 zhongmiansuo7 FM1735 shan70 ekangmian6 jinmian10 | guannong1 zhongmiansuo50 2011SS | xinluzhong41 |
|        |                  | huiyuan717 liaoamian17 ningmian22 PB12-1-8 nongda94-7 | yumian21 xinluzao23 ganmian2 | ganmian3 xinluzhong34 |
|        | High source-sink effect | chaoyangmian1 yumian1 ekangmian6 Deltapine15 han2490 103030 dezimian531 jinmian10 | 2011SS emian17 qing1 lu890 | xinluzhong41 |
|        |                  | xiaozao5 yishuhong 102909 FM1735 Delfo97-047 jiangmian9 zhongchuan88 | xinluzhong47 zhongmiansuo43 dezhimian531 ganmian2 | ganmian2 |
| E2     | Low source-sink effect | PB12-1-10 xinluzao4 jinmian15 zhongmiansuo14 | j4025 xinluzhong47 zhongmiansuo43 | zhongmiansuo50 dezhimian531 ganmian2 |
|        |                  | kezi201 jinmian1 xinluzao26 FM1735 zhongmiansuo14 | xinluzao15 zhongmiansuo64 jinmian3 | xinluzhong41 |
|        | High source-sink effect | xiaozao2 xinluzao15 xinluzhong40 | xinluzao10 xinluzhong40 | xinluzhong40 xinluzhong40 |
|        |                  | xinluzao26 jinmian15 xinluzao26 zhongmiansuo14 | shan79 zhongmiansuo69 xinluzhong40 | xinluzhong40 |
| E3     | Low source-sink effect | xinluzao26 jinmian15 xinluzao26 zhongmiansuo14 | shan79 zhongmiansuo69 xinluzhong40 | xinluzhong40 |
|        |                  | xinluzao26 jinmian15 xinluzao26 zhongmiansuo14 | shan79 zhongmiansuo69 xinluzhong40 | xinluzhong40 |
|        | High source-sink effect | xinluzao26 jinmian15 xinluzao26 zhongmiansuo14 | shan79 zhongmiansuo69 xinluzhong40 | xinluzhong40 |
| E4     | Low source-sink effect | yunzaoN95 liaoyangduannj zhongh416 FM1735 xinluzao3 | lumina2153 xinluzao15 baimian17 zhongmiansuo74 | bai425 xinluzao41 zhongmiansuo50 jinmian23 qing1 |
|        |                  | yunzaoN95 liaoyangduannj zhongh416 FM1735 xinluzao3 | xinluzao10 xinluzhong40 | xinluzhong40 |
|        | High source-sink effect | zhongmiansuo74 kemian4 shiziao2 zhongmiansuo14 yunmian1 | edaimian xinluzao20 xinluzao10 xinluzhong40 | xinluzhong40 |
|        |                  | zhongmiansuo74 kemian4 shiziao2 zhongmiansuo14 yunmian1 | xia13-7 xinluzao20 xinluzhong40 | xinluzhong40 |

4. Discussion

The source-sink relationship in plants is critical for the end-product as yield, and is a complicated phenomenon affecting the cotton yield and fiber quality. To understand the dynamics of the source-sink relationship, the core germplasm of upland cotton accessions was evaluated. The subtending leaf was considered a primary source, affecting the yield and fiber quality in cotton [26]. The effects of subtending leaf removal were subjected to multivariate data across 2 years and four subtending removal treatment effects related to fiber quality and yield traits. In addition, the corresponding variation in core germplasm was evaluated using the generalized linear model (GLM), correlations, PCA, and clustering. It is generally assumed that the subtending leaf is the major contributor to biomass accumulation in cotton bolls [42]. Hence, subtending leaves act as a source for their corresponding bolls, acting as a sink. This source-sink relationship is reflected as the coordination of vegetative and reproductive growth, and cotton development influences yield and fiber quality in cotton [43]. Considering a very basic and simple example, we look at a plant with two organs viz., leaf and root. Leaves are involved in carrying out the photosynthesis process and can be considered the sole source of carbon, whereas roots
are sinks for carbon and dependent on the leaf to support their growth and development. In contrast, we can consider roots as sources of nutrients and nitrogen, which they get from the soil and provide to the leaves, acting as net sinks in this scenario. Each plant cell utilizes carbon for respiration, growth of essential metabolites, building up proteins, various enzymes, and other genetic materials [44]. Hence, we can assume that mature leaves involved actively in photosynthesis have a relatively small carbon-sink activity. On the contrary, roots have a small nitrogen-sink activity and higher nitrogen-source activity, making them net sources for nitrogen (Burnett 2019).

Moreover, checking the photosynthesis through subtending leaves has been created by shading across flowering and the boll formation stage [45–47]. However, no clear and state-of-the-art experiment was reported to determine the source-sink relationship between subtending leaves with cotton boll, which is ultimately crucial to fiber yield enhancement in cotton [48]. Previously, a few studies have reported that the variation of starch and sucrose in cotton fibers have a remarkable association with fiber quality [49]. The portioning of photosynthates during the boll development stage (ovule or the fiber development) were primarily from the subtending leaves of the boll (Ryser 1992), and the content of sucrose and starch in cotton subtending leaves was higher than hexose [49,50]. Therefore, it is pertinent to examine the content and variation of sucrose and starch that might affect the fiber quality. In our study, all the genotypes depicted considerable variations when analyzed for fiber quality traits of FE, FU, FS, MIC, and FL and yield related traits such as BW, FW, SW, and GOT. The correlation matrix is used to investigate the dependence between multiple variables [51]. The significant positive associations among fiber quality parameters, i.e., FU, FL, FS, FE, and MIC, were observed in our study, which is in line with previously published statistics [52,53]. Similar to earlier findings, the current investigation found negative correlations among yield related and fiber quality traits. For instance, highly significant negative correlations have been exhibited by FE with SW and FS with FW and GOT [54]. However, GOT% revealed a highly significant positive correlation with MIC, as reported previously [55,56]. MIC depicting a significant negative correlation with FL has also been reported previously [57,58]. One of the liable mechanisms behind such negative correlations is repulsive linkage [58]. In these instances, superior genotypes harboring desirable traits related to yield and fiber quality can be utilized as recurrent parents to engulf negative correlations in selection-breeding programs [54]. In previous findings, FS and MIC revealed a positive correlation with each other as in the current study [59]. A biplot analysis has been implemented to understand the multivariate relationships among fiber quality and yield contributing traits across 355 upland cotton accessions concerning the association between germplasm accessions and the observed traits.

5. Conclusions

In the current study, insight into the variation of source-sink effects suggested a significant range of variation, as well as a positive correlation between different traits in core germplasm accessions under treatment effects across years. The treatment effect through the principal component analysis and hierarchical clustering methods divided accessions into three groups, based on four treatments across 2-year data. Our results depicted a significant variation among the core germplasm for the treatment effect through yield traits as well as for fiber length, and provide a basis for further studies to explore the source-sink relationship of subtending leaf with yield and fiber quality. A set of cotton accessions with higher treatment effect, which is considered to have a strong source-sink relationship and lower compensation effect along with a set of genotypes exhibiting a lower treatment and higher compensation effect, has been selected from this study. The reported results and selected genotypes based on their source-sink relationships can be further utilized in breeding programs to understand and evaluate the molecular mechanism behind the source-sink relationship in terms of carbon accumulation.

Supplementary Materials: The following are available online at https://www.mdpi.com/article/10.3390/plants10061147/s1, Supplementary Tables S1 to S9 Over-dispersion parameter estimated by
Maximum Likelihood using Generalized Linear Model (GLM) for Fiber quality and yield contributing traits in 355 upland cotton accessions under four treatment Effects (E1-E4) across two years 2018 and 2019. Supplementary Table S10. Eigenvalues contributing to different PCs for studied traits related to Source-Sink Effects across years 2018 and 2019. Supplementary Table S10 Loading Matrix Associated with the principal components for studied traits, treatment effects (E1-E4) and years 2018 and 2019.

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**References**

1. Zhang, X.; Kong, X.; Zhou, R.; Zhang, Z.; Zhang, J.; Wang, L.; Wang, Q. Harnessing perennial and indeterminant growth habits for ratoon cotton (Gossypium spp.) cropping. Ecosyst. Health Sustain. 2020, 6, 1715264. [CrossRef]

2. Oerke, E.C. Crop losses to pests. J. Agric. Sci. 2005, 144, 31–43. [CrossRef]

3. Sarfraz, Z.; Iqbal, M.S.; Pan, Z.; Jia, Y.; He, S.; Wang, Q.; Qin, H.; Liu, J.; Liu, H.; Yang, J.; et al. Integration of conventional and advanced molecular tools to track footprints of heterosis in cotton. BMC Genom. 2018, 19, 776. [CrossRef] [PubMed]

4. Suzuki, N.; Rivero, R.M.; Shulaev, V.; Blumwald, E.; Mittler, R. Abiotic and biotic stress combinations. New Phytol. 2014, 203, 32–43. [CrossRef]

5. Veres, A.; Wyckhuys, K.A.; Kiss, J.; Tóth, F.; Burgio, G.; Pons, X.; Avilla, C.; Vidal, S.; Razeringer, J.; Bazok, R. An update of the Worldwide Integrated Assessment (WIA) on systemic pesticides. Part 4: Alternatives in major cropping systems. Environ. Sci. Pollut. Res. 2020, 27, 29867–29899. [CrossRef] [PubMed]

6. Bange, M.P.; Baker, J.T.; Bauer, P.J.; Broughton, K.J.; Constable, G.A.; Luo, Q.; Oosterhuis, D.M.; Osanai, Y.; Payton, P.; Tissue, D.T. Climate Change and Cotton Production in Modern Farming Systems; CABl: Wallingford, UK, 2016.

7. Jans, Y.; von Bloh, W.; Schaphoff, S.; Müller, C. Global cotton production under climate change—Implications for yield and water consumption. Hydrol. Earth Syst. Sci. 2020. [CrossRef]

8. Anandjiwala, R.D.; Barkhuysen, F.; Tembo, A. Major factors affecting conversion of cotton fibers into quality products. In Proceedings of the 48th N.Z. Plant Protection Conference, Hamilton, New Zealand, 8 January 1995; pp. 208–212.

9. Conaty, W.C.; Constable, G.A. Factors responsible for yield improvement in new Gossypium hirsutum L. cotton cultivars. Field Crops Res. 2020, 250, 107780. [CrossRef]

10. Tang, B.; Jenkins, J.; Watson, C.; McCarty, J.; Creech, R. Evaluation of genetic variances, heritabilities, and correlations for yield and fiber traits among cotton F2 hybrid populations. Euphytica 1996, 91, 315–322. [CrossRef]

11. Rahman, S.A.; Iqbal, M.S.; Riaz, M.; Mahmood, A.; Shahid, M.R.; Abbas, G.; Farooq, J. Cause and effect estimates for yield contributing and morphological traits in upland cotton (Gossypium hirsutum L.). J. Agric. Res. 2013, 51, 393–398.

12. Yang, W.; Feng, H.; Zhang, X.; Zhang, J.; Doonian, J.H.; Batchelor, W.D.; Xiong, L.; Yan, J. Crop phenomics and high-throughput phenotyping: Past decades, current challenges, and future perspectives. Mol. Plant 2020, 13, 187–214. [CrossRef]

13. Brown, W.L. Genetic diversity and genetic vulnerability—An appraisal. Econ. Bot. 1983, 37, 4–12. [CrossRef]

14. Sarfraz, Z.; Iqbal, M.S.; Geng, X.; Iqbal, M.S.; Nazir, M.F.; Ahmed, H.; He, S.; Jia, Y.; Pan, Z.; Sun, G. GWAS Mediated Elucidation of Heterosis for Metric Traits in Cotton (Gossypium hirsutum L.) Across Multiple Environments. Front. Plant Sci. 2021, 12, 566. [CrossRef]
15. Zhu, G.; Li, W.; Wang, G.; Li, L.; Si, Q.; Cai, C.; Guo, W. Genetic basis of fiber improvement and decreased stress tolerance in cultivated versus semi-domesticated upland cotton. Front. Plant Sci. 2019, 10, 1572. [CrossRef] [PubMed]
16. Raheel, F.; Shah, M.K.N.; Iqbal, M.S.; Noorka, I.R.; Sarfraz, Z. Genetic Variations and Cultivar Purity within the G. hirsutum Cultivars via RAPD Marker. Am. J. Plant Sci. 2016, 5, 392–404. [CrossRef]
17. Woodings, C. Regenerated Cellulose Fibres; Elsevier: Amsterdam, The Netherlands, 2001.
18. Dingkuhn, M.; Luquet, D.; Fabre, M.; Muller, B.; Yin, X.; Paul, M.J. The case for improving crop carbon sink strength or plasticity for a CO$_2$-rich future. Curr. Opin. Plant Biol. 2020, 56, 259–272. [CrossRef] [PubMed]
19. Falchi, R.; Bonghi, C.; Drincovic, M.F.; Famiani, F.; Lara, M.V.; Walker, R.P.; Vizzotto, G. Sugar metabolism in stone fruit: Source-sink relationships and environmental and agronomical effects. Front. Plant Sci. 2020, 11, 1820. [CrossRef]
20. Ashley, D.A. C-Labelled Photosynthesize Translocation and Utilization in Cotton Plants. J. Crop Sci. 1972, 12, 69–74. [CrossRef]
21. Constable, G.; Rawson, H. Carbon production and utilization in cotton: Inferences from a carbon budget. Funct. Plant Biol. 1980, 7, 539–553. [CrossRef]
22. Nandula, V.K.; Vencill, W.K. Herbicide absorption and translocation in plants using radioisotopes. Weed Sci. 2015, 63, 140–151. [CrossRef]
23. FitzSimons, T.R.; Oosterhuis, D.M. The Role of Temperature on the Diurnal Sucrose Source to Sink Balance. J. Cotton Sci. 2016, 20, 196–206.
24. Wullschleger, S.; Oosterhuis, D. Photosynthetic carbon production and use by developing cotton leaves and bolls. Crop Sci. 1990, 30, 1259–1264. [CrossRef]
25. Gwathmey, C.; Clement, J. Alteration of cotton source–sink relations with plant population density and meipiquat chloride. Field Crops Res. 2010, 116, 101–107. [CrossRef]
26. Liu, J.; Lv, F.; Chen, J.; Ma, Y.; Wang, Y.; Chen, B.; Zhang, L.; Zhou, Z. Photosynthetic characteristics of the subtending leaf of cotton boll at different fruiting branch nodes and their relationships with lint yield and fiber quality. Front. Plant Sci. 2015, 6, 747. [CrossRef]
27. Xie, Z.; Jiang, D.; Cao, W.; Dai, T.; Jing, Q. Relationships of endogenous plant hormones to accumulation of grain protein and starch in winter wheat under different post-anthesis soil water statuses. Plant Growth Regul. 2003, 41, 117–127. [CrossRef]
28. Pace, P.; Cralle, H.T.; Cothren, J.T.; Senseman, S.A. Photosynthesize and dry matter partitioning in short-and long-season cotton cultivars. Crop Sci. 1999, 39, 1065–1069. [CrossRef]
29. Richards, R. Selectable traits to increase crop photosynthesis and yield of grain crops. J. Exp. Bot. 2000, 51, 447–458. [CrossRef] [PubMed]
30. Fischer, R.A.; Rees, D.; Sayre, K.D.; Lu, Z.-M.; Condon, A.G.; Saavedra, A.L. Wheat yield progress associated with higher stomatal conductance and photosynthetic rate, and cooler canopies. Crop Sci. 1998, 38, 1467–1475. [CrossRef]
31. Karp, A.; Shield, I. Bioenergy from plants and the sustainable yield challenge. New Phytol. 2008, 179, 15–32. [CrossRef] [PubMed]
32. Atwell, B.J.; Kriedemann, P.E.; Turnbull, C.G. Plants in Action: Adaptation in Nature, Performance in Cultivation; Macmillan Education AU: London, UK, 1999.
33. Gao, H.; Li, N.; Li, J.; Khan, A.; Ahmad, I.; Wang, Y.; Wang, F.; Luo, H. Improving boll capsule wall, subtending leaves anatomy and photosynthetic capacity can increase seed cotton yield under limited drip irrigation systems. Ind. Crops Prod. 2021, 161, 113214. [CrossRef]
34. Malik, R.; Sharma, H.; Sharma, I.; Kundu, S.; Verma, A.; Sheoran, S.; Kumar, R.; Chatrath, R. Genetic diversity of agromorphological characters in Indian wheat varieties using GT biplot. Aust. J. Crop Sci. 2014, 8, 1266.
35. Ajmal, S.U.; Minhas, N.M.; Hamdani, A.; Shakir, A.; Zubair, M.; Ahmad, Z. Multivariate analysis of genetic divergence in wheat (Triticum aestivum) germplasm. Pak. J. Bot. 2013, 45, 1643–1648.
36. Babić, V.; Pajić, Z.; Prodanović, S.; Babić, M.; Filipović, M. Visual assessment of sweet maize lines phenotype, according to UPOV descriptor, as indicator of heterosis. Genetika 2010, 42, 313–322. [CrossRef]
37. Azad, M.; Biswas, B.; Alam, N.; Alam, S.S. Genetic diversity in maize (Zea mays L.) inbred lines. Agriculturists 2012, 10, 64–70. [CrossRef]
38. Lee, K.-M.; Herrman, T.J.; Lingenfelsler, J.; Jackson, D.S. Classification and prediction of maize hardness-associated properties using multivariate statistical analyses. J. Cereal Sci. 2005, 41, 85–93. [CrossRef]
39. Ali, M.A.; Jabran, K.; Awan, S.; Abbas, A.; Zulkifial, M.; Acet, T.; Rehan, M. Morpho-physiological diversity and its implications for improving drought tolerance in grain sorghum at different growth stages. Aust. J. Crop Sci. 2011, 5, 311.
40. Bowman, D.T.; May, O.L.; Calhoun, D.S. Genetic base of upland cotton cultivars released between 1970 and 1990. Crop Sci. 1998, 38, 1572. [CrossRef] [PubMed]
41. Ngai, E.; Cheng, T. Identifying potential barriers to total quality management using principal component analysis and correspondence analysis. Int. J. Qual. Reliab. Manag. 1997, 14, 391–408. [CrossRef]
42. Liu, J.; Ma, Y.; Lv, F.; Chen, J.; Zhou, Z.; Wang, Y.; Abudurezike, A.; Oosterhuis, D.M. Changes of sucrose metabolism in leaf subtending to cotton boll under cool temperature due to late planting. Field Crops Res. 2013, 144, 200–211. [CrossRef]
43. Ngai, E.; Cheng, T.; Yog, L.; Yao, Y.; Meng, Y.; Chen, B.; Zhou, Z. Sucrose metabolism in the subtending leaf to cotton boll at different fruiting branch nodes and the relationship to boll weight. J. Agric. Sci. 2014, 152, 790. [CrossRef]
44. Haidar, S.; Aslam, M.; Hassan, M.; Hassan, H.M.; Ditta, A. Genetic diversity among upland cotton genotypes for different economic traits and response to cotton leaf curl virus (CLCV) disease. Pak. J. Bot. 2012, 44, 1779–1784.
45. Bondada, B.R.; Oosterhuis, D.M. Canopy photosynthesis, specific leaf weight, and yield components of cotton under varying nitrogen supply. *J. Plant Nutr.* 2001, 24, 469–477. [CrossRef]

46. Pettigrew, W.; Meredith, W., Jr. Genotypic variation in physiological strategies for attaining cotton lint yield production. *J. Cotton Sci.* 2012, 16, 179–189.

47. Wang, J.; Duncan, D.; Shi, Z.; Zhang, B. WEB-based gene set analysis toolkit (WebGestalt): Update 2013. *Nucleic Acids Res.* 2013, 41, W77–W83. [CrossRef]

48. Chen, B.L.; Yang, H.K.; Ma, Y.N.; Liu, J.R.; Lv, F.J.; Chen, J.; Meng, Y.L.; Wang, Y.H.; Zhou, Z.G. Effect of shading on yield, fiber quality and physiological characteristics of cotton subtending leaves on different fruiting positions. *Photosynthetica* 2017, 55, 240–250. [CrossRef]

49. Pettigrew, A.M.; Woodman, R.W.; Cameron, K.S. Studying organizational change and development: Challenges for future research. *Acad. Manag. J.* 2001, 44, 697–713.

50. Zhao, D.; Oosterhuis, D. Cotton responses to shade at different growth stages: Nonstructural carbohydrate composition. *Crop Sci.* 1998, 38, 1196–1203. [CrossRef]

51. Li, J.; Ji, L. Adjusting multiple testing in multilocus analyses using the eigenvalues of a correlation matrix. *Heredity* 2005, 95, 221–227. [CrossRef]

52. Wan, Q.; Zhang, Z.; Hu, M.; Chen, L.; Liu, D.; Chen, X.; Wang, W.; Zheng, J. T 1 locus in cotton is the candidate gene affecting lint percentage, fiber quality and spiny bollworm (*Earias* spp.) resistance. *Euphytica* 2007, 158, 241–247. [CrossRef]

53. Herring, A.D.; Auld, D.L.; Ethridge, M.D.; Hequet, E.F.; Bechere, E.; Green, C.J.; Cantrell, R.G. Inheritance of fiber quality and lint yield in a chemically mutated population of cotton. *Euphytica* 2004, 136, 333–339. [CrossRef]

54. Clement, J.; Constable, G.; Stiller, W.; Liu, S. Negative associations still exist between yield and fibre quality in cotton breeding programs in Australia and USA. *Field Crops Res.* 2012, 128, 1–7. [CrossRef]

55. Saeed, F.; Farooq, J.; Mahmood, A.; Hussain, T.; Riaz, M.; Ahmad, S. Genetic diversity in upland cotton for cotton leaf curl virus disease, earliness and fiber quality. *Pak. J. Agric. Res.* 2014, 27, 226–236.

56. Farooq, J.; Anwar, M.; Riaz, M.; Farooq, A.; Mahmood, A.; Shahid, M.; Rafiq, M.; Ilahi, F. Correlation and path coefficient analysis of earliness, fiber quality and yield contributing traits in cotton (*Gossypium hirsutum* L.). *JAPS J. Anim. Plant Sci.* 2014, 24, 781–790.

57. Rao, K.; Mary, T. Variability, correlation and path analysis of yield and fibre traits in upland cotton. *J. Res. APAU* 1996, 24, 66–70.

58. Desalegn, Z.; Ratanadilok, N.; Kaveeta, R. Correlation and heritability for yield and fiber quality parameters of Ethiopian cotton (*Gossypium hirsutum* L.) estimated from 15 (diallel) crosses. *Agric. Nat. Resour.* 2009, 43, 1–11.

59. SHAHZAD, K.; Xue, L.; Tingxiang, Q.; Liping, G.; Huini, T.; Zhang, X.; Hailin, W.; Zhang, M.; Zhang, B.; Xiuqin, Q. Genetic analysis of yield and fiber quality traits in upland cotton (*Gossypium hirsutum* L.) cultivated in different ecological regions of China. *J. Cotton Res.* 2019, 2, 1–11. [CrossRef]