BACH family members regulate angiogenesis and lymphangiogenesis by modulating VEGFC expression

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Abstract

Angiogenesis and lymphangiogenesis are key processes during embryogenesis as well as under physiological and pathological conditions. Vascular endothelial growth factor C (VEGFC), the ligand for both VEGFR2 and VEGFR3, is a central lymphangiogenic regulator that also drives angiogenesis. Here we report that members of the highly conserved BACH (BTB and CNC homology) family of transcription factors regulate VEGFC expression, through direct binding to its promoter. Accordingly, downregulation of bach2a hinders blood-vessel formation and impairs lymphatic sprouting in a vegfc-dependent manner during zebrafish embryonic development. In contrast, BACH1-overexpression enhances intratumor blood-vessel density and peritumoral lymphatic vessel diameter in ovarian and lung mouse tumor models. The effects on the vascular compartment correlate spatially and temporally with BACH1 transcriptional regulation of VEGFC expression. Altogether, our results uncover a novel role for the BACH/VEGFC signaling axis in lymphatic formation during embryogenesis and cancer, providing a novel potential target for therapeutic interventions.
Introduction

The blood and lymphatic networks are two evolutionarily conserved transport systems that provide complementary functions in the maintenance of tissue homeostasis. In particular, the formation of new blood and lymphatic vessels is a prerequisite for vertebrate embryonic and postnatal development. During embryogenesis, the blood circulatory system is first to emerge. As early as mouse embryonic day (E) 7.5, vasculogenesis, the formation of a primitive vascular plexus through proliferation, migration and differentiation of endothelial cells (ECs), takes place. Later on, this plexus undergoes massive remodeling via angiogenesis, which involves also arterio-venous differentiation (A. S. Chung & Ferrara, 2011). At E10.5, a distinct subpopulation of ECs within the cardinal vein commits to the lymphatic lineage, buds off and migrates to establish primitive lymph sacs, which through further sprouting give rise to the peripheral lymphatic system (Oliver & Alitalo, 2005; Semo, Nicenboim, & Yaniv, 2016). During adult life, the majority of blood and lymphatic vessels are quiescent, with the exception of female reproductive organs during the ovarian cycle and the placenta during pregnancy. Reactivation of angiogenesis and lymphangiogenesis, however, is a hallmark of pathological processes associated with wound healing, myocardial infarction (MI), allograft rejection, chronic inflammation, tumor progression and malignant cell dissemination (A. S. Chung & Ferrara, 2011; Oliver & Alitalo, 2005).

The VEGF family of growth factors and its receptors are central signaling pathways, controlling angiogenesis and lymphangiogenesis during development and adulthood. VEGFs exert their activity by binding to the tyrosine kinase receptors (vascular endothelial growth factor receptor-1) VEGFR-1, VEGFR-2, and VEGFR-3, expressed in blood and lymphatic endothelial cells (LECs) (Lohela, Bry, Tammela, & Alitalo, 2009). VEGFA, the ligand for VEGFR-1 and VEGFR-2, is best known for its role in angiogenesis, stimulating EC proliferation and migration and increasing vascular permeability. VEGFB, which also binds to VEGFR1, plays a role in cell survival and indirectly promotes VEGFA induced angiogenesis (Lal, Puri, & Rodrigues, 2018).

VEGFC is another critical player in VEGF signaling. This protein, which signals through the receptors VEGFR-3 and VEGFR-2, plays a key role across species, guiding both lymphatic and blood vasculature development and remodeling (Gore et al., 2011; Hogan, Bos, et al., 2009; Karkkainen et al., 2004; Kuchler et al., 2006; Lohela et al., 2009; Shin et al., 2016; Villefranc et al., 2013; Yaniv et al., 2006). VEGFC is expressed by endothelial and non-
endothelial cells, thus acting via both autocrine and paracrine signaling (Covassin, Villefranc, Kacergis, Weinstein, & Lawson, 2006; Helker et al., 2013; Khromova, Kopnin, Rybko, & Kopnin, 2012; Kodama et al., 2008; Lohela, Helotera, Haiko, Dumont, & Alitalo, 2008; Villefranc et al., 2013). Evidence for its necessity for proper lymph formation is revealed by the finding that ectopic expression of VEGFC in mouse ear or skin keratinocytes results in hyperplasia of lymphatic vessels. Similarly, mice and zebrafish lacking VEGFC fail to develop a lymphatic vasculature as initial sprouting and directed migration of lymphatic progenitors from the cardinal vein are arrested (Karkkainen et al., 2004; Kuchler et al., 2006; Shin et al., 2016; Villefranc et al., 2013; Yaniv et al., 2006). In addition to its predominant role in lymphangiogenesis, VEGFC was shown to induce angiogenesis upon overexpression in the mouse cornea, skin or ischemic hindlimb. Moreover, coronary vessel development is VEGFC dependent, and hearts devoid of VEGFC reveal dramatic delay in the formation of subepicardial sprouts (H. I. Chen et al., 2014). Recently, it has been shown by in vitro studies that the let-7a/TGFBR3 axis regulates angiogenesis through transcriptional regulation of VEGFC (Wang et al., 2019).

VEGFC has a critical role not only during development but also during tumor progression. In various human cancers, enhanced expression of VEGFC and higher levels of VEGFC in serum are commonly associated with tumor aggressiveness and lymph-node metastasis (Lohela et al., 2009; Su et al., 2007). In esophageal carcinoma, for instance, angiogenesis is driven via the phosphoinositide-phospholipase C-ε (PI-PLCε)/NF-κB signaling pathway by direct promotion of VEGFC transcription (Y. Chen et al., 2019). In xenograft or transgenic tumor models, stimulation of lymphangiogenesis by VEGFC promotes malignant cell dissemination (Stacker et al., 2014). Furthermore, blockade of VEGFC expression in tumor cells by stably transfected small interfering RNAs reduces lymphangiogenesis and lymph node metastasis of murine mammary cancers. Similarly, soluble VEGFR-3 protein has been shown to inhibit VEGFC-induced tumor lymphangiogenesis and metastatic spread in a breast cancer mouse model (Wissmann & Detmar, 2006).

A series of environmental and cellular factors were shown to modulate VEGFC expression. For instance, Interleukin-6 (IL-6) and interleukin-17 (IL-17) regulate VEGFC expression via the PI3K-Akt or ERK 1/2 pathways, whereas MicroRNA-1826 significantly down-regulates VEGFC expression in human bladder cancer (J. C. Chen, Chang, Hong, Yu, & Su, 2012). Vegfc mRNA levels are heightened in the adipose tissue of obese mice, pointing to

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adipocytes as a source of elevated VEGFC levels in obesity (Karaman et al., 2015). In adults, inflammation induces robust upregulation of VEGFC expression by macrophages (Baluk et al., 2005). Additionally, various hormones, transcription factors, metallothioneins and microenvironmental stresses (e.g., hyperthermia, oxidative stress, high salt) were shown to control VEGFC expression (J. C. Chen et al., 2012; Cohen et al., 2009; Gauvrit et al., 2018; Machnik et al., 2009; Sapoznik et al., 2009; Schuermann, Helker, & Herzog, 2015).

Here, we report that BACH transcription factors, which are known to be involved in various intracellular signaling pathways, modulate VEGFC expression during embryonic development and tumor progression, offering a better understanding of blood and lymph vessel formation during physiological and pathological conditions.

**Results**

**Spatial and temporal expression of bach2 during zebrafish development**

In order to explore the molecular mechanisms governing VEGFC expression, we searched for genes that co-express with *VEGFC* in human prostate cancer datasets (Glinsky, Glinskii, Stephenson, Hoffman, & Gerald, 2004). We identified 434 genes, on which we further applied the Promoters of Clusters (POC) analysis (Tabach et al., 2007). This method screens the promoters of genes with shared biological function against a library of transcription factor binding motifs and identifies those which are statistically over-represented. Five factors passed the basic significance threshold, out of which only two possessed binding sites within the *VEGFC* promoter. We then searched for evolutionarily conserved regions, which often represent potential DNA regulatory elements (Ovcharenko, Nobrega, Loots, & Stubbs, 2004). Comparison of the *VEGFC* promoters of human, mouse and zebrafish revealed two highly conserved BACH (BTB and CNC homology) binding sequences (Figure 1A).

The BACH family of transcription factors comprises two members, BACH1 and BACH2. In mammals, BACH1 is expressed ubiquitously and has been shown to act either as an activator or repressor of transcription and to be involved in oxidative stress, metabolism, cell transformation, neurodegenerative diseases, tumor expansion and metastatic spread (Igarashi & Watanabe-Matsui, 2014; J. Lee et al., 2014; J. Lee et al., 2019; Lignitto et al., 2019; Nakanome et al., 2013; Warnatz et al., 2011; Watari et al., 2008; Wiel et al., 2019; Zhou, Wu, Zhao, Chang, & Lu, 2016). BACH2 is a transcriptional repressor crucial for the terminal differentiation and maturation of both T and B lymphocytes (Sidwell & Kallies, 2016) and
its loss is associated with severe autoimmune diseases. BACH proteins are highly conserved in vertebrates, particularly in the functional (BTB and bZip) domains and the regions immediately surrounding them (Igarashi & Watanabe-Matsui, 2014). Zebrafish have four bach genes with homology to mammalian BACH: bach1a, bach1b, bach2a and bach2b (Fuse, Nakajima, Nakajima-Takagi, Nakajima, & Kobayashi, 2015; Luo et al., 2016; Zhang et al., 2014).

As a first step, we evaluated the expression pattern of the different bach transcripts in ECs isolated by Fluorescence-Activated Cell Sorting (FACS) from Tg(fli1:EGFP)y1 (Lawson & Weinstein, 2002) zebrafish embryos (L. Covassin et al., 2006). We found the transcripts of bach2a and bach2b to be more abundant than bach1b, while bach1a is barely detectable in the GFP-positive (GFP⁺) cell population, at either 21-24 hours post-fertilization (hpf) or 3 days post fertilization (dpf) (Figure 1B). Consequently, we focused our studies on bach2a and bach2b, as they are the major paralogs expressed in the endothelium. To assess the spatial and temporal expression patterns of bach2a and bach2b during zebrafish embryogenesis, we performed whole-mount in situ hybridization (WISH). Abundant expression of bach2a and bach2b was apparent in the somites and somite boundaries at 20 hpf and up to 48 hpf (Figure 1C). Furthermore, we detected strong enrichment of bach2a transcripts in myotomes and in several areas of the central nervous system (CNS), including the hindbrain, midbrain-hindbrain boundary, midbrain and forebrain (Figure 1- figure supplement 1).

Figure 1
Spatial and temporal expression of *bach2* paralog transcripts during zebrafish development.

(A) BACH putative binding sites are evolutionary conserved in the VEGFC promoter region. Numbering is from the ATG (translation initiation) due to the difference in length of the mouse's 5' UTR. An arrow indicates Transcription Start Site (TSS), and the location of the first exon is marked as a gray rectangle. The location of the BACH sites is as predicted by Genomatix Genome Analyzer MatInspector: Human (NM_005429.4 TSS at hg19, chr4:177713899 on the minus strand); Mouse (NM_009506.2 TSS at mm9, chr8:54077532 on the plus strand); Zebrafish (NM_205734.1 TSS at Zv9, chr1:39270725 on the minus strand). (B) Semi-quantitative RT-PCR analysis of the indicated genes in enriched GFP+ cells isolated by FACS from *Tg(fli1:EGFP)y1* embryos at two developmental time points, 21-24 hpf and 3 dpf (two independent experiments for each time point). (C) A lateral view of the trunk region of a wild-type zebrafish embryo at 20, 24, 30 and 48 hpf, detected with a specific *bach2a* or *bach2b* anti-sense mRNA probe. A red arrowhead indicates somite boundaries. Scale bar, 100 µm.

**Figure 1- figure supplement 1**

Expression of *bach2a* at different stages during zebrafish development.

Embryos at 20 (A), 24 (B), 30 (C, D) and 48 (E) hpf were examined by whole-mount in situ hybridization with an antisense *bach2a* riboprobe. Abbreviations: FB, forebrain; S, somite; TE, telencephalon; MB, midbrain; MHB, midbrain hindbrain boundary; HB, hindbrain; rHB, rostral hindbrain; SB, somite-boundaries; M, myotomes; OT, optic tectum. Scale bar, 500 µm.

*bach2a* is essential for developmental angiogenesis and lymphangiogenesis

We then assessed the contribution of BACH to angiogenesis and lymphangiogenesis during embryonic development and, specifically, investigated their putative role as regulators of *vegfc* expression and function during zebrafish vasculature formation. We used antisense morpholino oligonucleotides (MOs) to knockdown the expression of the two *bach2* paralogs and then surveyed the phenotypic changes at different stages of development. At 30 hpf, clear defects were observed in the primordial hindbrain channel (PHBC) in *bach2a* MO-
injected Tg(fli1:EGFP)y1 zebrafish embryos (Figures 2A and 2B). Similar phenotypes were detected in vegfc morphants (Figures 2A and 2B), in line with previous reports demonstrating impaired PHBC formation following the blocking of Vegfc-activated Vegfr3 signaling (L. D. Covassin et al., 2006; Hogan, Herpers, et al., 2009; Schuermann et al., 2015; Shin et al., 2016; Villefranc et al., 2013). In contrast to bach2a downregulation, no defects were detected in Tg(fli1:EGFP)y1 embryos either injected with different concentrations of bach2b MOs (up to 10 ng per embryo, Figures 2A and 2C) or subjected to a bach2b CRISPR guide RNA (bach2b gRNA, Figure 2-figure supplement 1A, 1B, Figure 2A and 2B). The vascular abnormalities detected upon bach2a and vegfc knockdown were accompanied by pericardial and body edema, as well as reduced blood flow (Figure 2-figure supplement 2), resembling the phenotypes observed in collagen and calcium-binding EGF domain-1 (ccbe1) (Hogan, Bos, et al., 2009) and vegfc (Karkkainen et al., 2004; Shin et al., 2016) mutants. In contrast, no morphological abnormalities were observed following bach2b knockdown (Figure 2-figure supplement 3A-3C).

To confirm the specificity of the bach2a morphant phenotype, we generated bach2a mutants using CRISPR/Cas9-mediated gene editing (Figure 2-figure supplement 2C-2E). To our surprise, homozygous bach2a mutants displayed no primordial hindbrain channel defects (bach2a\textsuperscript{mut/–}, Figures 2D and 2E). We therefore hypothesized that genetic compensation (El-Brolosy et al., 2019; El-Brolosy & Stainier, 2017; Rossi et al., 2015) through activation of bach2b could potentially account for the absence of angiogenic phenotypes in bach2a mutants. To address this possibility, we mated bach2a\textsuperscript{+/–} carriers and injected their progeny with either a sub-dose of bach2b MO or with bach2b gRNA that does not induce vascular malformations in WT embryos (Figures 2A-2C). Interestingly, PHBC formation defects were identified in ~25% of the embryos, which upon genotyping were found to carry the bach2a mutation (bach2a\textsuperscript{mut/–} + bach2b MO and bach2a\textsuperscript{mut/–} + bach2b gRNA, Figures 2D and 2E), as opposed to their wild-type siblings (bach2a\textsuperscript{mut+/–} + bach2b MO and bach2a\textsuperscript{mut+/–} + bach2b gRNA, Figure 2E). Conversely, following bach2a MO injection, bach2a wild-type siblings displayed PHBC formation defects while homozygous mutants appeared normal (Figures 2D and 2E), suggesting that the bach2a\textsuperscript{mut/–} mutants were less sensitive than their bach2a\textsuperscript{mut+/–} siblings to bach2a MO injections, and further confirming the specificity of both mutants and morphants. In order to ascertain whether the compensation mechanism involves upregulation of the paralogous bach2b gene, we assessed its expression in wild-type and bach2a mutants, by RT-qPCR. Surprisingly, while no significant differences in the levels of bach2b mRNA...
were detected (Figure 2- figure supplement 1F), we observed a slight elevation in \textit{bach2a} mRNA levels in homozygous mutants (Figure 2- figure supplement 1G), suggesting an alternative, yet unknown, mechanism of adaptive compensation following \textit{bach2a} perturbation.

**Figure 2**

\textit{bach2a} is essential for developmental angiogenesis in zebrafish embryos.  

(A) Confocal images of the primordial hindbrain channel (PHBC, white arrow) of 30-hpf \textit{Tg(fli1:EGFP)}\textsuperscript{y1} embryos injected with control MO (10 ng), \textit{bach2a} MO (3.75 ng), \textit{bach2b} MO (3.75 ng), \textit{bach2b} guide RNA (gRNA,125 ng ) or \textit{vegfc} MO (10 ng). Asterisk indicates absence of PHBC. (B) Percentage of 30-hpf \textit{Tg(fli1:EGFP)}\textsuperscript{y1} embryos with intact PHBC formation following injection with control MO (10 ng, n\textsubscript{Control MO}= 68), \textit{bach2a} MO (3.75 ng, n\textsubscript{bach2a MO}= 107; *P<0.0001), \textit{bach2b} MO (3.75 ng, n\textsubscript{bach2b MO}= 48), \textit{bach2b} guide RNA (gRNA, 125 ng, n\textsubscript{bach2b gRNA} = 42) or \textit{vegfc} MO (10 ng, n\textsubscript{vegfc MO} = 35; *P<0.0001). Error bars, mean ± s.e.m. (C) Percentage of 30-hpf \textit{Tg(fli1:EGFP)}\textsuperscript{y1} embryos with intact PHBC formation following injection with control MO (10 ng, n\textsubscript{Control MO} = 24 ) or an increased concentration of \textit{bach2b} MO (5 ng, n\textsubscript{bach2b MO 5ng} = 24) or (10 ng, n\textsubscript{bach2b MO 10ng} = 24). Error bars, mean ± s.e.m.; P>0.99999. (D) Confocal projection at 30 hpf \textit{Tg(fli1:EGFP)}\textsuperscript{y1} of homozygous \textit{bach2a} mutants (\textit{bach2a}\textsuperscript{mut/-}) from F2 \textit{bach2a}\textsuperscript{mut/+} incross. White arrow points at an intact PHBC detected in embryos injected with control MO (10 ng, \textit{bach2a}\textsuperscript{mut/-} + Control MO) and \textit{bach2a} MO (3.75 ng, \textit{bach2a}\textsuperscript{mut/-} + \textit{bach2a} MO). Asterisk indicates defects in PHBC development following injection with \textit{bach2b} MO (3.75 ng, \textit{bach2a}\textsuperscript{mut/-} + \textit{bach2b} MO) or \textit{bach2b} gRNA (125 ng, \textit{bach2a}\textsuperscript{mut/-} + \textit{bach2b} gRNA). Scale bar, 100 μm. (E) Percentage of randomly selected \textit{bach2a}\textsuperscript{-/-} F2 incross progeny at 30 hpf with an intact PHBC formation injected with control MO (10 ng, \textit{bach2a}\textsuperscript{mut} + Control MO = 50; P>0.99999), \textit{bach2a} MO (3.75 ng, \textit{bach2a}\textsuperscript{mut} + \textit{bach2a} MO = 75; *P<0.0002), \textit{bach2b} MO (3.75 ng, \textit{bach2a}\textsuperscript{mut} + \textit{bach2a} MO = 55; * P<0.012) or \textit{bach2b} gRNA (125 ng, \textit{bach2a}\textsuperscript{mut} + \textit{bach2b} gRNA = 137; * P<0.001). Following genotyping, offspring followed the expected Mendelian ratios of inheritance. Error bars, mean ± s.e.m.
CRISPR/Cas9-mediated editing of the bach2 paralog loci.

(A) Schematic illustration of the bach2b locus (XM_005160548.3) and the guide RNA targeting exon 4. The protospacer-adjacent motif (PAM) sequence is shown in red letters and the cleavage site is indicated by a red arrowhead. (B) Agarose gel electrophoresis of PCR products amplified from 24-hpf wild-type (WT) or bach2b gRNA (125 ng, bach2bmut)-injected Tg(fli1:EGFP)y1 embryos using primer sets which flank the gRNA cut sites. Arrows indicate the lengths of the resultant DNA fragments. (C) Schematic diagram of the bach2a locus (GRCz10, chr17:15666119-15734205). The CRISPR guide was designed to target exon 2, with the cleavage site (red arrowhead) located 115 bp downstream of the ATG. (D) Representative genotyping PCR of a wild-type (WT) and a CRISPR/Cas9-mediated homozygous bach2a knockout (bach2amut−/−) Tg(fli1:EGFP)y1 embryo using primer sets which flank the gRNA cut sites. Arrows indicate the lengths of the resultant DNA fragments. (E) PCR-sequencing chromatograph of the bach2a locus in a Tg(fli1:EGFP)y1 embryo (WT) aligned with a homozygous bach2a knockout (bach2amut−/−). CRISPR/Cas9-mediated genome modification at the zebrafish bach2a locus leads to the deletion of five base-pairs, which causes a frame shift in the protein sequence (translation shown below the DNA sequence). A stop codon (indicated by a dot in the translation) created 17 amino acids downstream of the deletion site, leading to the formation of a 57-amino-acid-long, truncated protein. (F) Quantitative RT-PCR validation ofbach2b expression conducted for wild-type (bach2aD−/−) and homozygous (bach2aD−/−) 6-dpf siblings obtained from F2 bach2a intercross. Error bars, mean ± s.e.m.; P=0.2466 (two independent experiments consisting of twenty single larvae in each group). (G) Semi-quantitative RT-PCR analysis of bach2a expression in wild-type (bach2aD+/+) and homozygous (bach2aD−/−) 6-dpf siblings obtained from F2 bach2a heterozygous intercross.
Knockdown of bach2a causes morphological defects in zebrafish embryos.

(A) Lateral-view confocal micrographs of full-body, 3-dpf Tg(fli1:EGFP)y1 zebrafish embryos following injection with control (10 ng, Control MO), bach2a (3.75 ng, bach2a MO) or vegfc (10 ng, vegfc MO) morpholino oligonucleotide, showing body curvature of bach2a morphants. Scale bar, 500 µm. (B) Percentage of 3-dpf Tg(fli1:EGFP)y1 zebrafish embryos with a normal morphology (no body curvature) following injection with control (10 ng, Control MO) or bach2a MO (3.75 ng, nControl MO = 46; nbach2a MO = 47 *P≤0.0005). (C) Confocal images of the heart morphology of 3-dpf Tg(fli1:EGFP)y1 zebrafish embryos following injection with control, bach2a or vegfc MO. H marks the heart and a white arrowhead, a pericardial edema. Scale bar, 500 µm. (D) Quantification of 2- and 3-dpf Tg(fli1:EGFP)y1 zebrafish embryos with a normal heart morphology following injection with control MO (10 ng) or vegfc MO (10 ng, nControl MO = 16; vegfc MO = 19; 3 dpf: nControl MO = 47; nbach2a MO = 58; *P<0.0001). (E) Percentage of 3-dpf Tg(fli1:EGFP)y1 zebrafish embryos with a normal heart morphology following injection with control MO (10 ng) or vegfc MO (10 ng, nControl MO = 16; vegfc MO = 19; 3 dpf: nControl MO = 47; nbach2a MO = 58; *P<0.0001). (F) Dorsal view of 5-dpf Tg(fli1:EGFP)y1 zebrafish larvae following injection with control MO, bach2a MO or vegfc MO. Red arrowhead indicates edema. Scale bar, 100 µm. (G) Percentage of 5-dpf Tg(fli1:EGFP)y1 zebrafish larvae with edema following injection with control MO (10 ng) or bach2a MO (3.75 ng, nControl MO-10ng = 22; nbach2a MO = 28; *P<0.05). (H) Quantification of 5-dpf Tg(fli1:EGFP)y1 zebrafish larvae with edema following injection with control MO (10 ng) or vegfc MO (10 ng, nControl MO = 22; vegfc MO = 53; *P<0.0001). (I) Percentage of 2- and 3-dpf Tg(fli1:EGFP)y1 zebrafish embryos with a normal blood flow rate following injection with control MO (10 ng) or bach2a MO (3.75, 2 dpf: nControl MO = 14; nbach2a MO = 19; 3 dpf: nControl MO = 47; nbach2a MO = 58; *P<0.05). (J) Percentage of 3-dpf Tg(fli1:EGFP)y1 zebrafish embryos with a normal blood flow rate following injection with control MO (10 ng) or vegfc MO (10 ng: nControl MO = 54; vegfc MO = 41; *P<0.004).
No morphological defects in *bach2b* zebrafish morphants.

(A) Percentage of 3-dpf *Tg(fli1:EGFP)y1* zebrafish embryos following injection with control MO (10 ng) or *bach2b* MO (3.75, 5 or 10 ng) showing normal heart morphology (nControl MO = 18; n*bach2b* MO-3.75ng = 30; n*bach2b* MO-5ng = 30; n*bach2b* MO-10ng = 10; *P*≥0.3878). (B) Quantification of 5-dpf *Tg(fli1:EGFP)y1* zebrafish larvae with edema following injection with control MO (10 ng) or *bach2b* MO (3.75, 5 or 10 ng) (nControl MO = 18; n*bach2b* MO-3.75ng = 30; n*bach2b* MO-5ng = 30; n*bach2b* MO-10ng = 10; *P*>0.9999) (C) Percentage of 3-dpf *Tg(fli1:EGFP)y1* zebrafish embryos with a normal blood flow rate following injection with control MO (10 ng) or *bach2b* MO (3.75, 5 or 10 ng) (nControl MO = 18; n*bach2b* MO-3.75ng = 30; n*bach2b* MO-5ng = 30; n*bach2b* MO-10ng = 10; *P*<0.9999).

We then asked whether *bach2a* is also involved in lymphatic vessel development. To answer this question, we analyzed the effects of *bach2a* down-regulation on the formation of parachordal cells (PACs), the building blocks of the thoracic duct (TD) and the trunk lymphatic system in zebrafish (Nicenboim et al., 2015; Yaniv et al., 2006). A significantly reduced number of PAC-containing segments was detected in *bach2a* MO-injected *Tg(fli1:EGFP)y1* embryos at 3 dpf (Figures 3A and 3B), recapitulating the *vegfc* MO induced phenotype (Figures 3A and 3C) and the previously reported phenotype of *vegfc* mutants (Villefranc et al., 2013). Additionally, *bach2a* morphants failed to express the lymphatic endothelial marker *lyve1*, but the expression of *vegfc* receptor *flt4* remained unchanged (Figures 3- figure supplement 1). To rule out the possibility that the observed phenotypes are a consequence of developmental delay, we assessed the formation of the TD at 4 dpf. MO-mediated downregulation of *bach2a* resulted in a significant decrease in TD formation (76%, Figures 4A and 4B) as compared to control MO-injected siblings. Similarly, 87% of *vegfc* morphants were devoid of a TD (Figures 4A and 4C), as previously reported (Shin et al., 2016; Villefranc et al., 2013). In contrast to *bach2a* morphants, *bach2b* MO (Figures 3A, 3D, 4A and 4D) and *bach2b* gRNA-injected *Tg(fli1:EGFP)y1* embryos (Figures 3A, 3E, 4A and 4E) exhibited no lymphatic defects. In line with the possible genetic compensation underlying the absence of a clear angiogenic phenotype at early developmental stages (Figures 2D and 2E), injection of either a sub-dose of *bach2b*
MO or bach2b gRNA into the progeny of bach2a\(^{+/-}\) intercross leads 23% of embryos to display significant lymphangiogenic defects, which were subsequently identified by genotyping as homozygous mutants (bach2a\(^{mut/-}\) + bach2b MO or bach2a\(^{mut/-}\) + bach2b gRNA) (Figures 3F, 3G and 4F and 4G). In contrast, control MO-injected bach2a\(^{mut/-}\) appeared normal (Figures 3F, 3G, 4F and 4G). Likewise, no lymphatic phenotype was detected in bach2a\(^{mut/-}\) embryos upon injection of bach2a MO (Figures 3F, 3G, 4F and 4G), indicating that bach2a MO has minimal off-target effects. Finally, cardiac and body edema, characteristic of lymphatic-related defects, were detected in bach2a mutants following bach2b MO or bach2b gRNA injection, but not upon bach2a MO administration (Figure 4- figure supplement 1). Remarkably, the phenotypic defects resulting from bach2a downregulation were more pronounced than those caused by the loss of vegfc, possibly due to the involvement of bach2a in additional signaling pathways (Zhou et al., 2016).

**Figure 3**

*bach2a* is essential for parachordal cell (PAC) development in zebrafish embryos.

(A) Confocal projection of the trunk of 3-dpf *Tg(fli1:EGFP)*\(^{y1}\) embryos showing PACs (white arrow) in control MO, bach2b or bach2b gRNA-injected embryos but not following injection with bach2a or vegfc MO (white asterisk). Scale bar, 100 \(\mu\)m. (B) Number of PAC-containing segments (mean ± s.e.m.) in 3-dpf *Tg(fli1:EGFP)*\(^{y1}\) zebrafish embryos injected with control MO (10 ng) or bach2a MO (3.75 ng, \(n_{Control MO} = 46\); bach2a MO, \(n_{bach2a MO} = 53\); *\(P<0.001\)). Error bars, mean ± s.e.m. (C) Number of PAC-containing segments
in vegfc MO-injected morphants (10 ng, n-Control MO = 53, n-vegfc MO = 41; \*P<0.0001). Error bars, mean ± s.e.m. (D) Number of PAC-containing segments in 3-dpf Tgf(fli1:EGFP)\(^{+/-}\) zebrafish embryos injected with indicated bach2b MO concentrations (3.75, 5 or 10 ng, n-Control MO = 18; n bach2b MO-3.75ng = 30; n bach2b MO-5ng = 30; n bach2b MO-10ng = 10; P>0.2819). Error bars, mean ± s.e.m. (E) Quantification of PAC-containing segments in embryos injected with bach2b gRNA (125 ng, n-Control MO = 35; n bach2b gRNA = 42; P=0.0615). Error bars, mean ± s.e.m. (F) Confocal projection of the trunk region showing PAC-containing segments in Tgf(fli1:EGFP)\(^{+/-}\)- homozygous bach2a mutants (bach2a\(^{mut/-}\)) from F2 bach2a\(^{mut+/-}\) incross. An asterisk indicates the absence of PACs in bach2a\(^{mut+/-}\) embryos injected with bach2b MO (bach2a\(^{mut+/-}\)+ bach2b MO) or bach2b gRNA (bach2a\(^{mut+/-}\)+ bach2b gRNA) and a white arrow, their presence. Scale bar, 100 \(\mu\)m. (G) Number of PAC-containing segments (mean ± s.e.m.) in 3-dpf embryos randomly selected from bach2a\(^{+/-}\) F2 incross progeny injected with control MO (10 ng, n bach2amut + Control MO = 50; P=0.5514), bach2a MO (3.75 ng, n bach2amut + bach2a MO = 75; \*P<0.0001), bach2b MO (3.75 ng, n bach2amut + bach2b MO = 55; \*P and **P<0.0222) or bach2b gRNA (125 ng, n bach2amut + bach2b gRNA = 137; \*P and **P<0.0001 ). Following genotyping, offspring followed the expected Mendelian ratios of inheritance.

**Figures 3- figure supplement 1**

**Expression of lyve1 is impaired in bach2a morphants.**

(A) Whole mount *in situ* hybridization comparing the expression pattern (red arrowhead) of *lyve1* in 24-hpf wild-type zebrafish embryos following injection with control or bach2a MO (red asterisk indicates absence of expression). (B) Whole mount *in situ* hybridization using *flt4* riboprobe of 24-hpf, wild-type zebrafish embryos following injection with control or bach2a MO. Scale bar, 100 \(\mu\)m.
**Figure 4**

*bach2a* is necessary for thoracic duct development in zebrafish larvae.

(A) Confocal images of the thoracic duct (TD) in 4-dpf control MO- (10 ng), *bach2a* MO- (3.75 ng), *vegfc* MO- (10 ng), *bach2b* MO- (10 ng) or *bach2a* gRNA-injected Tg(fli1:EGFP)Y1 larvae. A white arrow indicates the presence of a TD and a white asterisk, its absence. Scale bar, 20 µm. (B) Percentage of 4-dpf Tg(fli1:EGFP)Y1 larvae with an intact thoracic duct (TD) following injection with control MO (10 ng) or *bach2a* MO (3.75 ng, nControl MO = 46; n*bach2a* MO = 52). Error bars, mean ± s.e.m.; *P*<0.0001. (C) Percentage of larvae with a TD following injection with *vegfc* MO (10 ng, nControl MO = 46; n*vegfc*MO = 52). Error bars, mean ± s.e.m.; *P*<0.0001. (D) Quantification of *bach2b* morphants with an intact TD following injection with indicated *bach2b* MO concentrations (3.75, 5 or 10 ng, nControl MO = 18; n*bach2b* MO-3.75ng = 30; n*bach2b* MO-5ng = 30; n*bach2b* MO-10ng = 10; *P*≥0.1356). Error bars, mean ± s.e.m. (E) Percentage of TD-containing larvae injected with *bach2b* gRNA (125 ng, nControl MO = 35; n*bach2b* gRNA = 42; *P*=0.5461). Error bars, mean ± s.e.m. (F) Confocal images of a TD (white arrow) in 4-dpf Tg(fli1:EGFP)Y1 homozygous *bach2a* mutants (*bach2a*mut/−) derived from *bach2a*mut+/− F2 incross. An asterisk indicates absence of a TD and a white arrow, its presence. Scale bar, 20 µm. (G) Analysis of Tg(fli1:EGFP)Y1 4-dpf progeny obtained from F2 *bach2a* heterozygous intercross. Random selection from the pool of siblings injected at the one-cell stage with control MO (10 ng, n*bachmut* + Control MO = 50), *bach2a* MO (3.75 ng, n*bach2mut* + *bach2a* MO = 75), *bach2b* MO (3.75 ng, n*bach2mut* + *bach2b* MO = 55) or *bach2b* gRNA (125 ng, n*bach2mut* + *bach2b* gRNA = 137) were found to maintain, following genotyping, the expected Mendelian ratios of inheritance. Error bars, mean ± s.e.m.; *P*<0.0003.
Morphological abnormalities in homozygous bach2a mutants following bach2b knockdown.

(A) Fluorescent images of homozygous bach2a (bach2amut-/+) siblings obtained from F2 bach2a heterozygous intercross injected at the one-cell stage with control (10ng, Control MO) or bach2b MO (3.75 ng). Scale bar, 500 µm. (B) Bright-field images corresponding to the larvae presented in panel A. Red arrowhead indicates cardiac edema. Scale bar, 500 µm. (C) Lateral view of 6-dpf homozygous bach2a (bach2amut-/+) siblings obtained from F2 bach2a heterozygous intercross injected with control (10 ng, Control MO), bach2a (3.75 ng, bach2a MO), bach2b (3.75 ng, bach2b MO) morpholino or bach2b guide RNA (125 ng, bach2b gRNA). Red arrowhead indicates cardiac edema, while body edema is indicated by a black arrow. Scale bar, 500 µm.

Taken together, these findings demonstrate that knockdown of bach2a through MO injection results in the formation of defective blood and lymphatic vascular plexuses, despite the presence of the bach2b paralog, indicating that bach2a is a major contributor to this phenotype. However, knockout of bach2a expression (in mutants) triggers a functional compensation by bach2b, suggesting that the two bach2 genes may share partially overlapping functions, which allows them to compensate for each other's loss during blood and lymph vessel development. Accordingly, it was suggested that BACH1 and BACH2 act in a complementary manner to maintain normal alveolar macrophage function and surfactant homeostasis in the lung (Ebina-Shibuya et al., 2016). Similar phenotypic differences between mutants and transient knockdown animals were observed in various model systems, some of which were attributed to genetic compensation (El-Brolosy et al., 2019; El-Brolosy & Stainier, 2017; Rossi et al., 2015).
**BACH1 promotes angiogenesis and lymphangiogenesis during tumor expansion in mouse models**

In light of the perception that cancers frequently reactivate embryonic developmental signaling cascades to promote their expansion and aggressiveness, resulting in metastasis and poor patient outcome, we decided to evaluate the contribution of BACH to vascular remodeling during tumor progression. Emerging evidence point to BACH1, a ubiquitously expressed protein, as a tumor-promoting factor that acts via multiple intracellular signaling cascades (Alvarez & Woolf, 2011; Yun et al., 2011). Recently, higher levels of BACH1 were found to be associated with poor prognosis in human ovarian cancer (Han et al., 2019) and to promote lung cancer metastasis (Lignitto et al., 2019; Wiel et al., 2019). BACH1 has been established as a major regulator of breast cancer bone metastasis (Liang et al., 2012) and was postulated as a potential novel therapeutic candidate for cancer treatment (Davudian, Mansoori, Shajari, Mohammadi, & Baradaran, 2016; J. Lee et al., 2019). To assess whether enhanced expression of BACH1 in tumor cells can stimulate blood and lymphatic vessel expansion during tumor progression as well as metastatic spread, we analyzed various ovarian and lung mouse tumors (Figures 5 and Figure 5- figure supplement 1). Inoculation of human ovarian clear cell carcinoma ES2 cells into the peritoneal cavity of immune-deficient CD-1 nude female mice has been established as a model of metastatic ovarian cancer (Shaw, Senterman, Dawson, Crane, & Vanderhyden, 2004). In this study, we used this model to evaluate the effects of ectopic expression of *BACH1* in human ovarian ES2 carcinoma cells (Figure 5). Twenty days post-injection, metastases were detected in the diaphragm of animals injected with *BACH1*-overexpressing ES2 cells. An increase in CD34+ tumor-associated blood vessel density was apparent in diaphragm specimens derived from mice injected with *BACH1*-overexpressing cells, as compared to control ES2 cells (Figures 5A and 5B). Similar differences in the intratumoral blood vasculature were observed following subcutaneous inoculation of either *BACH1*-overexpressing ES2 cells (Figure 5- figure supplement 1A, 1B and 1C) or *Bach1*-overexpressing mouse D122 Lewis Lung Carcinoma cells (Eisenbach et al., 1984) (Figure 5- figure supplement 1E, 1F and 1G). These results indicate that, in addition to its role in embryonic vascular development, BACH1 promotes tumor angiogenesis.

Numerous reports suggest that, in addition to blood vessel formation, lymphangiogenesis and lymphatic vessel remodeling are pivotal events for tumor expansion and metastatic spread (Stacker et al., 2014). We thus examined the effects of BACH1 on the lymphatic vasculature
in various mouse tumor models. To gain insight into the 3D complexity of the lymphatic
network within intact tumors, we subjected ES2 subcutaneous xenografts to a modified
CLARITY (Clear, Lipid-exchanged, Anatomically Rigid, Imaging-compatible, Tissue
hYdrogel) technique (K. Chung et al., 2013; Hama et al., 2011; Oren et al., 2018), along with
LYVE1 immunofluorescence staining. A robust increase in lymphatic vessel density was
observed in BACH1-overexpressing tumors as compared to control-derived ES2 tumors
(Figure 5C, Figure 5- Movie 1 and Figure 5- Movie 2). Morphometric analyses indicate a
significant increase in the relative area fraction occupied by LYVE1+ tumor-associated
lymphatic vessels in diaphragm specimens removed from mice injected with BACH1-
overexpressing ES2 cells (Figures 5D and 5E). In addition, enlarged lymph vessels were
observed in the peritumoral region of subcutaneous tumors derived from BACH1-
overexpressing ES2 and D122 cells (Figure 5- figure supplement 1D and 1H, respectively).
Expansion of the lymph vessels in the diaphragms of BACH1-overexpressing tumors was
associated with increased cell infiltration (arrow, Figure 5D). Co-immunofluorescence using
antibodies directed against LYVE1 and Cytokeratin 7, an antigen expressed in ES2 cells
(Stimpfl et al., 1999), revealed that the metastasizing cells originated from the transplanted
ES2 tumor cells (arrow, Figure 5F). This potential of BACH1 to promote tumor cell invasion
was further confirmed by an in vitro invasion assay demonstrating a ~two-fold increase in
the invasion abilities of BACH1-overexpressing as compared to control ES2 cells (Figures
5G and 5H). These data indicate that in mouse models BACH1 remolds vascular
architecture and promotes metastatic spread of tumor cells via the lymphatic vessels.
**Figure 5**

**BACH1** promotes angiogenesis and lymphangiogenesis during ovarian tumor progression in mouse models.

*Ex vivo* analysis of subcutaneous xenografts and diaphragm specimens excised from CD-1 nude female mice implanted with control (Control) or **BACH1** ectopically expressing (**BACH1**) human ovarian clear cell carcinoma ES2 cells. (A) Immunofluorescence labeling of blood vessels using anti-CD34 antibodies in diaphragm specimens excised from mice injected intraperitoneally with control or **BACH1**-expressing human ES2 cells. Scale bar, 100 µm. (B) Morphometric analysis of the diaphragm relative region covered by CD34+ blood vessels. Diaphragms were excised from mice inoculated intraperitoneally with either control (Control, n=3) or **BACH1**-overexpressing (**BACH1**, n=7) ES2 cells (mean ± s.e.m.; *P* = 0.0304). (C) Confocal z-projection images of control and **BACH1**-overexpressing subcutaneous-ES2 ovarian carcinoma xenografts subjected to LYVE1 immunofluorescence staining along with a modified CLARITY technique. Images demonstrate the complexity of the lymphatic vasculature. Scale bar, 100 µm. 3D reconstructions of the stacks are available in Supplementary movies 1 and 2. (D) Lymphatic vessel immunostaining, using anti-LYVE1 antibodies, of diaphragm specimens excised from mice injected intraperitoneally with control or **BACH1**-overexpressing ES2 ovarian carcinoma cells. A black arrow indicates infiltration of cells into the lymphatic vessel and an asterisk, their absence. Scale bar, 100 µm. (E) Morphometric analysis of the diaphragm relative region covered by LYVE1+ lymph vessels. Diaphragms were excised from mice inoculated intraperitoneally
with either control (Control, n=6) or BACH1-overexpressing (BACH1, n=9) ES2 cells (mean ± s.e.m.; *P=0.0047). (F) Immunofluorescence double staining of LYVE1⁺ lymphatic vessels (red) and cytokeratin 7 (CK7, green) of a diaphragm excised from mouse inoculated intraperitoneally with either control or BACH1-overexpressing ES2 cells. Nuclei were counterstained with DAPI (blue). A white arrow indicates infiltration of tumor cells into the lymphatic vessels and an asterisk, their absence. Scale bar, 100 μm. (G) Transwell Matrigel invasion assay performed in vitro with ES2 ectopically expressing BACH1 and control cells. The crystal violet dye staining images of the lower chambers are shown. Scale bar, 100 μm. (H) Percentage of ES2 cells that invaded through the Matrigel matrix (as in panel g) normalized to total cell number (n = 2 for each group, in duplicates; mean ± s.e.m.; *P=0.0209).

Figure 5- figure supplement 1

BACH1 overexpression in mouse tumor models stimulates angiogenesis, lymphangiogenesis and induction of VEGFC expression.

Analysis of subcutaneous tumor specimens (two mouse models: human ovarian clear cell carcinoma ES2 cells in CD-1 nude mice and mouse D122 Lewis lung carcinoma cells in C57BL/6 black mice) excised from mice implanted with either ectopic overexpression of HA-BACH1 or control cells expressing an empty vector (Control). (A) Western blot detecting HA-BACH1 and β-Tubulin in protein extracts from control (-) or BACH1 (+)-ectopically-expressing ES2 cell. (B) Immunofluorescence labeling of blood vessels using anti-
CD34 antibodies of subcutaneous tumors excised from mice inoculated with control (Control) or BACH1 (BACH1)-ectopically-expressing human ES2 cells. (C) Morphometric analysis of the relative coverage of CD34+ blood vessels within the tumor area (n = 5 in each group; mean ± s.e.m.; *P<0.0001). (D) Immunohistochemical staining of lymphatic endothelial cells using anti–LYVE1 antibodies. Scale bar, 100 µm. (E) Western blot detecting HA-BACH1 and β-Tubulin in protein extracts from control (-) or Bach1 (+)-ectopically-expressing mouse D122 Lewis lung carcinoma cells. (F) Immunofluorescence labeling of blood vessels using anti-CD34 antibodies of subcutaneous tumor specimens excised from mice inoculated with control (Control) or Bach1 (Bach1)-ectopically-expressing mouse D122 Lewis lung carcinoma cells. (G) Morphometric comparison between the coverage of CD34+ blood vessels within the tumor area of control (Control, n = 9) or Bach1-overexpressing (Bach1, n = 8) D122 Lewis lung cells (mean ± s.e.m.; *P<0.0001). (H) Immunohistochemical staining of lymphatic endothelial cells using anti-LYVE1 antibodies. (I) Quantitative RT-PCR validation of Vegfc mRNA levels in Bach1-overexpressing mouse D122 Lewis lung tumors compared to control tumor (n = 6 per group; mean ± s.e.m.; *P<0.05). (J) Immunohistochemical labeling of tumor sections using anti-VEGFC antibodies and counterstaining with hematoxylin (blue). Scale bar, 100 µm.

**Figure 5- Movie 1**

**Lymphatic vessel complexity in a control tumor.**
A movie demonstrating the lymphatic vessel density in a control subcutaneous ES2 ovarian carcinoma xenograft subjected to LYVE1 immunofluorescence staining along with a modified CLARITY technique. Scale bar, 100 µm.

**Figure 5- Movie 2**

**Lymphatic vessel complexity in a BACH1-overexpressing tumor.**
A movie demonstrating the lymphatic vessel density in a subcutaneous-BACH1-overexpressing ES2 ovarian carcinoma xenograft subjected to LYVE1 immunofluorescence staining along with a modified CLARITY technique. Scale bar, 100 µm.

**BACH and VEGFC are functionally linked**
Next, we investigated the molecular mechanisms underlying BACH activity. Since BACH proteins can directly bind to the promoter of their target genes (Ogawa et al., 2001; Warnatz et al., 2011; Yun et al., 2011), we assessed the ability of the binding sites in the VEGFC promoter region identified *in silico* (Figures 1A and Figure 6A) to interact with BACH. Chromatin immunoprecipitation (ChIP) was performed on human ovarian clear cell carcinoma ES2 cells, overexpressing BACH1 tagged with a human influenza hemagglutinin (HA) tag (Figure 5- figure supplement 1A). We observed that specific sequences from both proximal and distal regulatory regions were enriched in the presence of antibodies against either HA or BACH1, indicating that both sites are transcriptionally functional (Figure 6B). To investigate the functional significance of two potential BACH binding sites, promoter-reporter constructs were engineered with deletion of the BACH binding sites, either separately or concurrently. Disruption of the proximal site led to a 23% increase in VEGFC
basal promoter activity (Figures 6C and 6D), supporting an inhibitory role. In contrast, deletion of the distal binding site individually or in combination with the proximal site resulted in an 86% reduction in VEGFC basal promoter activity (Figures 6C and 6D), suggesting a pivotal role in basal promoter functioning. Additionally, overexpression of BACH1 in human ovarian ES2 carcinoma cells induces a significant increase in VEGFC expression (Figure 6E), strongly supporting a genetic and functional link between these two factors.

Figure 6

BACH1 and VEGFC genetically interact.

(A) Conservation of BACH sites in human, mouse and zebrafish. The distal site is completely conserved. The proximal site is fully conserved between mouse and human, while there are three BACH sites at very close proximity in zebrafish. All proximal sites differ by one nucleotide from the consensus sequence. (B) Chromatin immunoprecipitation (ChIP) assay, followed by PCR measurements, was performed using primer mapping to the above human BACH proximal and distal regulatory sites and DNA precipitated with non-specific IgG, HA-tag or BACH1 antibodies. (C) Schematic representation of the wild-type human VEGFC promoter-driven luciferase (Luc) reporters (blue) (pVEGFCw-Luc) and of three constructs deleted either from proximal (nt. -623 to -603, pVEGFCPro-Luc) or distal (nt. -2074 to -2054, pVEGFCDis-Luc) BACH binding sites or a combination thereof (nt -623 to -603 and -2074 to -2054, pVEGFCProDis-Luc). Numbers refer to the nucleotide positions relative to ATG (translation initiation). TSS, transcription start site. (D) Quantification of dual-luciferase activity in human ES2 cells driven from pVEGFCw-Luc, pVEGFCPro-Luc, pVEGFCDis-Luc and pVEGFCProDis-Luc constructs. Relative luciferase activity is shown as a percentage of the pVEGFCw-Luc value (mean ± s.e.m., n = 3). *P<0.0001 (E) Immunofluorescence staining of human ES2 cells stably expressing either an empty pIRES vector (Control) or N-terminally HA-tagged BACH1 (BACH1) with antibodies directed against the HA tag (red, left panel) or against VEGFC (red, right panel). Nuclei were counterstained with DAPI (blue). Scale bar, 20 µm.

To investigate whether these results are recapitulated in vivo, we analyzed the effect of
*bach2a* knockdown on *vegfc* expression during zebrafish development. A marked downregulation of *vegfc* expression was observed in *bach2a* morphants (Figure 7A). This effect was specific for *bach2a*, as *vegfc* expression remained intact in *bach2b* MO-injected embryos (Figure 7- figure supplement 1). We then attempted to rescue the phenotypes of *bach2a* morphants by overexpressing *vegfc* mRNA. Co-injection of *in vitro* transcribed *vegfc* mRNA and *bach2a* MO partially restored PHBC formation at 30 hpf (Figures 7B and 7C). Similarly, the number of PAC-containing segments increased by approximately 50% following co-injection of *bach2a* MO and *vegfc* mRNA (Figures 7D and 7E) and a two-fold recovery in TD formation was detected (Figures 7F and 7G). In addition, pericardial and body edema, as well as reduced blood flow, were partially restored following an injection of *vegfc* mRNA (Figure 7- figure supplement 2).

Finally, we addressed the molecular basis of the BACH1 and VEGFC interaction during tumor progression in the various mouse tumor models. VEGFC expression was specifically elevated in BACH1-overexpressing ES2 and D122 tumors both at the protein (Figures 7H, and Figure 5- figure supplement 1J, respectively) and mRNA level (Figures 7I and Figure 5- figure supplement 1I), with no significant changes in the mRNA levels of either VEGFA or VEGFB (Figure 7- figure supplement 3). Similarly, the mRNA expression levels of *MMP1*, a recognized transcription target of BACH1 (Liang et al., 2012; Yun et al., 2011), were significantly elevated in subcutaneous *BACH1*-overexpressing ES2 xenografts (Figure 7I). Collectively, these results highlight BACH as a novel regulator of blood and lymphatic vessel formation during both embryonic development and mouse tumor expansion, placing it upstream of VEGFC in these cascades.
BACH mediates angiogenesis and lymphangiogenesis in a VEGFC-dependent manner.

(A) Whole-mount in situ hybridization of 24-hpf, wild-type zebrafish embryos demonstrating the expression of vegfc mRNA following injection with control MO (Control MO 10 ng, red arrowhead) and the absence of its expression in embryos injected with specific MO targeting bach2a (3.75 ng, bach2a MO, red asterisk). Scale bar, 100 µm. (B) Confocal images of 30-hpf Tg(fli1:EGFP)y1 embryos co-injected with specific MOs targeting bach2a (3.75 ng) and in vitro transcribed vegfc mRNA (800 pg, bach2a MO + vegfc mRNA) demonstrating the restoration of PHBC (white arrow). (C) Percentage of rescued PHBC defects in 30-hpf Tg(fli1:EGFP)y1 embryos following co-injection with bach2a MO (3.75 ng) and vegfc mRNA (800 pg), nControl MO = 56; n bach2a MO = 58; n bach2a MO + vegfc mRNA = 56. Error bars, mean ± s.e.m.; * or **P<0.0001. (D) Rescue of PAC development in 3-dpf Tg(fli1:EGFP)y1 embryos following the co-injection of bach2a MO and vegfc mRNA (bach2a MO + vegfc mRNA; PACs are indicated by a white arrow). Scale bar, 100 µm. (E) Quantification of the number of PAC-containing segments (mean ± s.e.m.) in 3-dpf Tg(fli1:EGFP)y1 embryos following the co-injection of bach2a MO (3.75 ng) and vegfc mRNA (nControl MO = 47; n bach2a MO = 27; n bach2a MO + vegfc mRNA = 54; * or **P<0.01). (F) TD formation in 4-dpf Tg(fli1:EGFP)y1 embryos co-injected with bach2a MO and vegfc mRNA (bach2a MO + vegfc mRNA; TD is indicated by a white arrow). Scale bar, 20 µm. (G) Percentage of 4-dpf Tg(fli1:EGFP)y1 embryos showing normal TD following bach2a MO and vegfc mRNA injection (bach2a MO + vegfc mRNA), nControl MO = 32; n bach2a MO = 65; n bach2a MO + vegfc mRNA = 68. Error bars, mean ± s.e.m.; * or **P<0.01 (H) Immunohistochemistry labeling of control (Control) or BACH1 (BACH1) ectopically expressing ES2 ovarian carcinoma xenograft specimens using anti-VEGFC antibodies.
and counterstaining with hematoxylin (blue). Black asterisk localizes the region magnified in the black frame. Scale bar, 100 µm. (I) Quantitative RT-PCR measurement of VEGFC and MMP1 mRNA expression in xenograft initiated either from control or BACH1-overexpressing ES2 cells (n=5 in each group; mean ± s.e.m.; *P<0.05).

Figure 7- figure supplement 1

Vegfc mRNA levels are not affected in bach2b morphants.

Whole mount in situ hybridization presenting the expressions of vegfc in 24-hpf embryos injected with control or bach2b MO. Presence of expression is indicated by a red arrowhead. Scale bar, 100 µm.

Figure 7- figure supplement 2

Vegfc mRNA partially rescues morphological defects in bach2a morphants.

(A) Quantification of 3-dpf Tg(fli1:EGFP)记 zebrafish embryos injected with control (10 ng), bach2a (3.75 ng) MO or co-injection with bach2a MO (3.75 ng) together with in vitro transcribed vegfc mRNA (800 pg) showing normal body morphology (nControl MO = 25; n bach2a MO = 41; n bach2a MO + vegfc mRNA = 43; * or **P<0.05). (B) Percentage of normal heart morphology (3.5 dpf, b: nControl MO = 54; n bach2a MO = 28; n bach2a MO + vegfc mRNA = 33; * or **P<0.05). (C) Percentage of rescued body edema in 5-dpf larvae (nControl MO = 25; n bach2a MO = 66; n bach2a MO + vegfc mRNA = 70; *P<0.0001). (D) Quantification of normal blood flow rate of 3-dpf embryos (nControl MO = 54; n bach2a MO = 35; n bach2a MO + vegfc mRNA = 28). *P<0.05.
The expression of VEGFA and VEGFB during tumor growth in mouse models is not BACH1 dependent. (A) Quantitative RT-PCR measurement of VEGFA expression conducted on xenograft initiated either from control or BACH1-overexpressing ES2 cell (n=5 in each group; mean ± s.e.m.; P = 0.6857). (B) Quantitative RT-PCR validation of VEGFB expression conducted either on control or BACH1-overexpressing ES2 xenografts (n = 5 in each group; mean ± s.e.m.; P=0.0556). (C) Quantitative RT-PCR measurement of Vegfa mRNA levels in Bach1-overexpressing mouse D122 Lewis lung tumors compared to control tumor (n = 6 per group; mean ± s.e.m.; P=0.2500). (D) Quantitative RT-PCR validation of Vegfb mRNA levels in Bach1-overexpressing mouse D122 Lewis lung tumors compared to control tumor (n = 6 per group; mean ± s.e.m.; P>0.9999).

The expression of BACH1 and VEGFC correlates during human cancer progression
Metastatic spread of cancer cells from primary solid tumors to sentinel lymph nodes and distant tissues and organs is one of the hallmarks of malignant neoplasms (Valastyan & Weinberg, 2011), responsible for the majority of human cancer-related deaths. Neoplastic cell dissemination may occur either via blood vessels or via the lymphatic system (Paduch, 2016). VEGFC is one of the key factors promoting malignant cell spread, as demonstrated both in mouse tumor models (Ma et al., 2018; Mandriota et al., 2001; Skobe et al., 2001) and during human cancer progression (J. C. Chen et al., 2012; H. Jiang, Shao, & Zhao, 2014; Rinderknecht & Detmar, 2008; Thiele & Sleeman, 2006). Similarly, it was shown that BACH1 promotes the metastasis of breast cancer through different molecular mechanisms (J. Lee et al., 2014; U. Lee et al., 2013; Liang et al., 2012) and its stabilization in lung...
adenocarcinoma is associated with increased metastatic dissemination and poor survival (Lignitto et al., 2019; Wiel et al., 2019). To substantiate the pathophysiological relevance of the interaction between $BACH1$ and $VEGFC$, we carried out an *in silico* analysis of publicly available gene-expression data generated by The Cancer Genome Atlas (TCGA) Research Network (https://cancergenome .nih.gov/). We analyzed RNA-sequencing-derived data regarding aberrant gene expression in specimens taken from melanoma and lung adenocarcinoma (LUAD) human cancer patients. Melanoma is one of the commonest forms of skin cancer, while LUAD is at present the most common lung cancer subtype among non-smokers. In both cancers, while the early-stage is curable by surgical resection, lymphatic metastasis results in poor prognosis. Interestingly, we found that $BACH1$ expression positively and significantly correlates with the expression of $VEGFC$ in human melanoma and LUAD cancer progression (Figure 8). Specifically, $BACH1$ and $VEGFC$ expression are significantly higher in samples from melanoma patients clinically diagnosed with lymph node metastatic spread, as compared to those with primary tumors (Figure 8A). Similarly, the expression of $BACH1$ and $VEGFC$ were significantly augmented in specimens derived from clinical stage III LUAD patients in comparison to stage I and II (Figure 8B). According to the tumor node metastasis (TNM) taxonomy classification, stage I refers to the early, non-metastatic stage, while stages II and III usually indicate the intermediate, regional lymphatic metastatic stages, of which stage III has a higher lymphatic metastasis degree than stage II. This correlation may indicate the potential ability of $BACH1$ and $VEGFC$ to promote cellular migration and cancer invasion. Hence, BACH1 and VEGFC may serve as candidate diagnostic biomarkers in cancer patients.
The expression of BACH1 and VEGFC correlates during human cancer progression.

(A) Correlation of BACH1 and VEGFC expression in specimens from melanoma patients clinically diagnosed with a primary tumor (n = 84) in comparison to those with metastatic stage melanoma (n = 356), as deduced from The Cancer Genome Atlas (TCGA) Research Network RNA sequencing (RNA-Seq.) data. Mean ± s.e.m.; *P≤0.0008. (B) Correlation of BACH1 and VEGFC expression in specimens from lung adenocarcinoma (LUAD) clinically diagnosed with primary tumor stages I (n = 292), II (n = 133) and III (n = 95), as analyzed from The Cancer Genome Atlas (TCGA) Research Network RNA-Seq. data. Mean ± s.e.m.; *P<0.02.

Discussion

VEGFC is a potent regulator of the growth and maintenance of blood and lymphatic vessels during embryonic development, tumor expansion and metastasis. The results presented here demonstrate that members of the BACH family regulate VEGFC expression, thereby promoting angiogenesis and lymphangiogenesis during zebrafish development and in ovarian and lung mouse tumor models.

BACH1 and BACH2 are important for the homeostasis of heme, an essential molecule for many biological functions (Igarashi & Watanabe-Matsui, 2014; Warnatz et al., 2011). Imbalanced levels of heme can cause oxidative stress when it reacts with molecular oxygen, which in turn disrupts various cellular signaling pathways. BACH1 was initially discovered as a physiological repressor of heme oxygenase-1 (HO-1), a rate-limiting enzyme in heme
catabolism stimulated by Nuclear factor erythroid 2-related factor 2 (NRF2) (Warnatz et al., 2011). Recent emerging evidence, however, indicate the widespread functions of BACH1 in diverse physiological and pathological settings, including hematopoiesis, inflammation, cardiovascular disease, aging, metabolic disorders and neurodegenerative diseases. BACH2 was established as a crucial factor for B and T cell memory differentiation. Nevertheless, the potential roles of these transcription factors during embryogenesis remain obscure (Sidwell & Kallies, 2016). Members of the BACH family possess both a BTB/POZ and a CNC-bZip functional domain, conferring them with protein-protein interaction and DNA-binding capabilities (Zhou et al., 2016) to the antioxidant response elements (ARE) (Zhou et al., 2016). The BTB/POZ domain, located at the N-terminus, mediates dimer formation (Igarashi & Watanabe-Matsui, 2014). Conversely, the bZip domain enables DNA binding and the formation of heterodimers with small musculoaponeurotic fibrosarcoma (sMaf) transcription factors (Davudian, Mansoori, et al., 2016). Three evolutionary conserved sMafs have been identified in mammals and an additional one in zebrafish, all displaying complex expression patterns during embryogenesis (Yamazaki, Katsuoka, Motohashi, Engel, & Yamamoto, 2012). While single sMaf knockout mice show no or a mild phenotype, triple-knockout embryos display severe growth retardation and liver hypoplasia, resulting in embryonic lethality at E13.5 (Yamazaki et al., 2012). Interestingly, a member of the large Maf family of transcription factors, mafba in zebrafish (Koltowska et al., 2015) and Mafb in mice (Dieterich et al., 2015), was shown to be activated by VEGFC. In zebrafish, mafba is crucial for the migration of lymphatic precursors after their initial sprouting from the cardinal vein (Koltowska et al., 2015). In lymphatic endothelial cells (LEC), MAFB promotes the expression of PROX1, KLF4, NR2F2, and SOX18, key transcription factors and markers of differentiated LECs. Furthermore, E14.5 Mafb−/− mouse embryos show impaired lymphatic patterning in the skin (Dieterich et al., 2015). Altogether, these studies suggest that the BACH and MAF transcription factor families play key roles during lymphangiogenesis. Uncovering the complexity within these molecular networks may be exploited for the understanding of EC differentiation and vascular development.

Four bach genes (bach1a, bach1b, bach2a and bach2b) with homology to mammalian BACH are present in the zebrafish genome. Phylogenetic analysis indicates that bach1a, bach1b and bach2a, bach2b belong to two distinct groups, bach1 and bach2, respectively, with bach2 possibly diverging earlier than bach1 (Luo et al., 2016). It was shown that, during embryogenesis, the two bach1 genes inhibit heme oxygenase 1a (hmox1a) induction
in zebrafish (Fuse et al., 2015). In a recent study, overexpression of bach1b was found to suppress developmental angiogenesis by inhibiting Wnt/β-catenin signaling (L. Jiang et al., 2017). Herein, we show that the bach2a gene controls vegfc expression, directing blood and lymphatic vascular development in zebrafish. Yet, the lack of full overlapping expression between bach2a and vegfc supports additional tissue-specific functions for each of these factors.

Accumulating data establish BACH1 as a critical facilitator of tumorigenesis and metastasis in breast (U. Lee et al., 2013), colon (Davudian, Shajari, et al., 2016), prostate (Shajari et al., 2018) ovarian (Han et al., 2019) and lung (Lignitto et al., 2019; Wiel et al., 2019) cancer. Elevated levels of BACH1 expression have been linked to a higher risk of breast cancer recurrence in patients (Liang et al., 2012), while association with metastatic spread and poorer prognosis has recently been suggested in the case of human ovarian cancer (Han et al., 2019) and lung adenocarcinoma (Lignitto et al., 2019; Wiel et al., 2019). Ectopic expression of BACH1 in breast cancer cells promotes malignancy and metastasis, while its knockdown suppresses these processes. BACH1 has been placed downstream of the Raf kinase inhibitory protein (RKIP), a tumor suppressor gene shown to inhibit invasion and bone metastasis in a breast cancer xenograft mouse model (U. Lee et al., 2013; Yun et al., 2011). Inactivation of RKIP during tumor expansion results in higher expression of BACH1 and its target genes C-X-C chemokine receptor type 4 (CXCR-4) and matrix metalloproteinase1 (MMP1), established drivers of tumor progression and metastasis (Foley & Kuliopulos, 2014; Mishan, Ahmadiankia, & Bahrami, 2016). Furthermore, ablation of BACH1 in human colon carcinoma (Davudian, Shajari, et al., 2016) or in prostate cancer cells (Shajari et al., 2018), prevents cell growth, migration and invasion in vitro, decreasing the expression of its main metastasis-related genes, MMP1, let-7a and CXCR4. Interestingly, cxcr4 is expressed in the somites and the endothelium of zebrafish embryos (Chong, Emelyanov, Gong, & Korzh, 2001), where we detect the expression of both bach2a and bach2b transcripts. cxcr4 was shown to be crucial for lateral aortae formation (Siekmann, Standley, Fogarty, Wolfe, & Lawson, 2009) and trunk lymphatic vascular network assembly (Cha et al., 2012) and a key modulator of vascular progenitor cells (Sainz & Sata, 2007). In addition to its contribution to development, CXCR4 was shown to play a role in carcinogenesis, tissue repair and other pathological circumstances (Kawaguchi, Zhang, & Nakanishi, 2019). VEGFC can thus be added to a growing list of tumor and metastatic proteins, including CXCR4 and MMP1, all of which are transcriptionally regulated by
BACH1. Including VEGFC as a target of BACH1 allows novel perspectives of the role of BACH1 in vascular development during embryogenesis and pathological conditions. During the past years, a substantial amount of *in vitro*, animal and small-size human studies established BACH proteins as a hub of critical transcriptional networks that govern key processes during normal physiology and disease states. Surprisingly, however, *Bach* single- (Muto et al., 2004; Sun et al., 2002) or double- (Ebina-Shibuya et al., 2016) knockout mice exhibit no phenotype at birth. This may result from the existence of a yet unknown compensatory mechanism similar to the one described here in zebrafish. Further understanding the mechanisms underlying BACH’s mode of action during embryogenesis, adult life and tumorigenesis would pave the way to resolving this ambiguity.

Altogether, our results highlight BACH as a novel regulator of angiogenesis and lymphangiogenesis during embryonic development as well as tumor progression. We show that BACH controls the expression of VEGFC, an established pro-lymphangiogenic and angiogenic growth factor. The expanded recognition of BACH1 activity, together with its inhibitory effect on NRF2, an important detoxifying and antioxidant factor, mark it as a potential therapeutic target. Hence, various direct or indirect promising BACH1 modulators have been developed (Arbiser, 2011; Banerjee, Basu, Arbiser, & Pal, 2013; Cuadrado et al., 2019). Accordingly, recent reports demonstrated that targeting of BACH1 and mitochondrial metabolism may serve as an effective therapy for triple negative breast cancer (J. Lee et al., 2019). In two new studies on lung cancer it was shown that therapeutic intervention that either destabilizes BACH1 (Lignitto et al., 2019) or disrupts its ability to induce glycolysis (Wiel et al., 2019) have the potential to inhibit BACH1 pro-metastatic activity. Targeting the BACH1/VEGFC signaling axis with these inhibitors may potentially be significant therapeutically for various blood and lymphatic vessels pathologies as well.

**Experimental Procedures**

**Bioinformatics analysis**

Transcription factor binding site analysis was performed using the Genomatix Genome Analyzer (Genomatix Software GmbH, Germany) MatInspector program (Cartharius et al., 2005). Promoter regions of the human, mouse and zebrafish Vegfc genes were extracted from the UCSC genome browser (Kent et al., 2002) as follows: Human-GRC37/hg19 chr4: 177,713,306-177,716,211; Mouse-GRCm38/mm10 chr8:54,075,150-54,077,946; Zebrafish Zv9/danRer7 chr1:39,270,126-39,273,225. The zebrafish *bach* sequences used are: *bach1a*: 
NM_001040313; bach1b: NM_001020663; bach2a: XM_680223 and bach2b: XM_677841.

**Zebrafish lines and husbandry**
The EK and Tg(fli1:EGFP)y1 (Lawson & Weinstein, 2002) zebrafish lines were maintained under standard conditions (Isogai, Lawson, Torrealday, Horiguchi, & Weinstein, 2003). All experiments were carried out according to the guidelines of the Weizmann Institute Animal Care and Use Committee.

**Fluorescence Activated Cell Sorting (FACS) and RT-PCR analysis**
Tg(fli1:EGFP)y1 embryos at 21-24 hpf and 3 dpf were collected and dissociated as previously described (Nicenboim et al., 2015). FACS of single cell suspensions was performed at 4°C using FACS Aria flow cytometer (Becton Dickinson, San Jose, CA). Total RNA was isolated from equal numbers of GFP+ and GFP− cells by PerfectPure RNA Cultured Cell kit (5 PRIME), according to the manufacturer's instructions. RNA (100 ng) from each sample was subjected to first-strand cDNA synthesis with Superscript III reverse transcriptase (Invitrogen) and random hexamers. All PCR conditions were optimized to produce a single product of the correct base pair size in the linear range of the reaction using the following set of primers: bach1a: 5’-TGTAAGACGGCGGAGTAAGA and 5’-CTTCAGCTGGTTGTG GTCT, bach1b: 5’-CTTCAGTGCTCGTGTGTCCA and 5’-TG TAGGCGAACTCCAGCA AG, bach2a: 5’-GACAGAACACGAGCCACTCA and 5’-AC AGCGCATGACATCTTTGG A, bach2b: 5’-TGACATCCTGAACCTTGAGTGT and 5’-CT GCACATCTCGACACACCT, fli1: 5’-CCGAGGTCTTCTGCTCTCACA and 5’-GGGAC TGGTCAGCGTGGAGAT, bactin: 5’-CGAGCAGGAGATGGGAACC and 5’-CAACGG AAACGCTCATTGC.

**Construction of the pCS2vegfcCDS plasmid**
The full-length coding region of zebrafish vegfc was amplified from cDNAs derived from 24 hpf embryos using a forward (5’-ATGCACTTATTTGGATTTTCT) and a reverse (5’-TTAGTCCAGTCTTCCCCAG) primers, and sub-cloned into the pCRII-TOPO cloning vector (Invitrogen). Following nucleotide sequence verification, a Gateway-compatible (Invitrogen) middle entry clone was generated using Gateway BP clonase (Invitrogen) mediated recombination. A pCS2vegfcCDS plasmid was produced using Gateway LR clonase (Invitrogen). Capped mRNA was transcribed from a Not1-linearised template using
Microinjection of zebrafish embryos

Morpholino antisense oligonucleotides (Gene Tools) were designed with sequences complementary to zebrafish bach2a and bach2b cDNA in a location downstream to the initiating start codon. The morpholino sequences were as follow: bach2a: (GenBank accession number XM_680223.4), 5’-TGTCAGGCTTCTCTCCATAGACAT-3’ and bach2b: (GenBank accession number XM_677841.2), 5’-CTTACAGTTACCTCATCCAGGACAT-3’. The morpholino targeting the zebrafish vegfc was previously described (Yaniv et al., 2006). As a control the Gene Tools standard control MO (5’-CCTCTTACCTAGTTACATTTATA-3’) was applied. Specific Morpholino oligonucleotides were injected into one-cell stage embryos at concentrations from 3.75 up to 10 ng and control MO at 10 ng per embryo. For rescue experiments pCS2vegfcCDS mRNA (800 pg per embryo) was simultaneously injected with the morpholino (3.75 ng per embryo). For quantification of the phenotypes, embryos from each group were randomly selected. Quantification was deduced from pools of at least 3 independent experiments.

Design and synthesis of guide RNA (gRNA)

The design of the bach2a and bach2b CRISPR guide was performed with CHOPCHOP (Montague, Cruz, Gagnon, Church, & Valen, 2014) (chopchop.rc.fas.harvard.edu). Potential off-target sequences were checked using the MIT CRISPR Design site (crispr.mit.edu) (Hsu et al., 2013). Oligonucleotides synthesized for the guide sequence 5’-GGACGTCTCTGTGACGTGA and 5’-CTGTGCCGAATTCCTGCGCA for bach2a and bach2b respectively, were cloned into the BsmBI site of the pT7-gRNA plasmid (a gift from Wenbiao Chen) (Addgene plasmid # 46759) (Jao, Wente, & Chen, 2013). Alt-R S.p. Cas9 Nuclease 3NLS (250 ng/µl, Integrated DNA Technologies) and gRNA (125 ng/µl) in 5 µl total volume were co-injected into one-cell-stage Tg(fli1:EGFP)y1 embryos. To detect mutagenic events, DNA was extracted from 24 hpf embryos, amplified using a set of primers 5’-AGCAAGGAATGTCTATGGAGGA and 5’-ATGAGTGACCTGTGGCA TCTGTGTC (235bp) for bach2a and 5’-CGCTCCATTGTACAGTTACATCCAGGACAT-3’ for bach2b. PCR products were separated on 3.5% MetaPhor Agarose gel (Lonza). When both genotyping and phenotypic analyses of single zebrafish embryos from heterozygous bach2a intercross was needed, the larval tail (6 dpf)
was used for genotyping, whereas RNA was extracted from the anterior part using NucleoSpin RNA plus XS RNA purification kit (Macherey-Nagel).

**Whole-mount in situ Hybridization**

Embryos were dechorionated and fixed overnight in 4% PFA at 4°C, at the appropriate time points. *In situ* hybridization was performed as previously described (Yaniv et al., 2006) using single-stranded digoxygenin-dUTP-labeled RNA probes transcribed by T7 RNA polymerase (Roche). The PCR generated probes were amplified with the following set of primers: *bach2a*: 5’-AAGAGTGAGCTAGAGGGCA and 5’-CGTTCTCTTGTTCGGG ATCTTG; *bach2b*: 5’-CTGCGCAGTGAGAGGACGG and 5’-GCTCCACCTCTTGCTTGAC; *vegfc*: 5’-CATCAGCAGTCATACATCAG and 5’-GTCCAGTCTTCCCAGTATG; *lyve1*: 5’-GGTTTGGTTGGTGAGGAGTCGAGTCCCTGTC; *flt4*: 5’-CTCGGAATGACATGTGGCTGG and 5’-CAGCCAGCGAGCACAAGC. After performance of a color reaction with alkaline phosphatase substrates (Roche), embryos were fixed in 4% PFA and washed in PBS supplemented with 0.1% Tween-20 (PBST). For tissue clarification embryos were mounted in glycerol and imaged using Leica M165 FC stereomicroscope.

**Cell culture**

Human ovarian clear cell carcinoma ES2 cell line (American Type Culture Collection) was cultured in DMEM supplemented with 10% fetal bovine serum (FBS). Mouse D122 Lewis lung carcinoma cell line (Eisenbach et al., 1984), kindly provided by Prof. Lea Eisenbach, Weizmann Institute of Science, Israel) was grown in DMEM containing 10% FBS, 1 mM sodium pyruvate and 1% nonessential amino acids. All cell lines were routinely tested for mycoplasma contamination using the EZ-PCR Mycoplasma Test Kit (Biological Industries).

**Establishment of stable BACH1-overexpressing cell pools**

Full-length coding region of the human BACH1 (*BACH1*, GenBank accession number NM_206866) and mouse Bach1 (*Bach1*, GenBank accession number BC057894.1) with an N-terminal hemagglutinin (HA) tag were reverse transcribed with SuperScript III Reverse Transcriptase (Invitrogen) and PCR-amplified using Phusion high-fidelity DNA polymerase (New England Biolabs) together with the following set of primers: *BACH1*: 5’-ATGTCTCTGTGAGAATCCTG and 5’-TTACTCAGTAGTAGACATTTATC;
Bach1: 5′-ATGTCTGTGAGTAGTAGTGCG and 5′-TTACTCGTCAGTAGTGCACTTG. The fragments were ligated into pCRII-TOPO and their sequence fidelity was confirmed by sequencing. Inserts were restricted and ligated into pEIRES expression vector containing the human EF-1a promoter (Hobbs, Jitrapakdee, & Wallace, 1998) to produce the pRESBACH1 and pRESBach1 constructs. ES2 and D122 cells were transfected with pRESBACH1 and pRESBach1 expression vectors respectively, using Lipofectamine 2000 reagent (Invitrogen) and selected with 2.5µg/ml puromycin (Sigma) as was previously described (Cohen et al., 2009). An average of 50 individual puromycin-resistant colonies were collected together and BACH1-overexpression was confirmed by immunoblotting. ES2 and D122 cells that were stably transfected with the pEIRES empty vector were used as controls.

Chromatin immunoprecipitation (ChIP) assay
The ChIP experiments were performed using the ChIP-IT-Express kit (Active Motif catalog no. 53009). Cells were fixed and cross-linked using 1% formaldehyde at room temperature for 10 min. Fixation was stopped by adding glycine. DNA extracted from nuclear fraction was subjected to enzymatic shearing for 35 min at 37°C to obtain mononucleosomes. The resulting chromatin preparation was immunoprecipitated with magnetic protein G coupled beads and 10 mg of either anti HA-tag (HA.11, Covance), Bach1 (Santa Cruz, sc-14700) or nonspecific-IgG antibody. DNA-protein crosslinking was reversed at 65°C (4 h), treated with Proteinase K (2 h, 42°C). The recovered DNA was then subjected to PCR using the following set of specific primers: BACH proximal site: 5′-GAGGGAGAGTGAGAGGGG and 5′-CGCAGGATCCTCCAGAGC; BACH distal site: 5′-CCGAGTCTGATGGGATGGAA and 5′-GCCTTTGTGTGATCACAGCCTTG.

Transient transfection and Luciferase Assay
The 5′-regulatory region of human VEGFC gene (NG_034216.1) encompassing 2274 nucleotides (-1 to -2274 in relation to ATG, Figure 1) was synthesized by GenScript. This fragment was then used as a template for deletion of either the proximal (nt -623 to -603, 5′-GGCGGCTACATGCCCCTG) or distal (nt -2074 to -2054, 5′-GAGAGTGAGATGCTAAGCACTG) BACH binding site individually or in combination. Subsequently, these synthesized fragments were sub-cloned into the pGL4.10[luc2] promoter-less vector (Promega) between KpnI and XhoI restriction sites to generate the pVEGFCwt-Luc (wild-type VEGFC promoter region), pVEGFCΔPro-Luc (VEGFC promoter region deleted of the
proximal BACH binding site), pVEGFCΔDis-Luc (VEGFC promoter region deleted of the distal BACH binding site) and pVEGFCΔProDis-Luc (VEGFC promoter region deleted of both proximal and distal BACH binding sites). All constructs were verified by DNA sequence analysis. For the luciferase reporter gene assay, ES2 cells plated in 24-well plates (40,000 cells per well) were co-transfected with the indicated pVEGFC-Luc construct in combination with the pRL-TK-Renilla (pRL-TK vector, Promega) luciferase internal control vector using Lipofectamine 2000 (Thermo Fisher Scientific), according to the manufacturer's instructions. After 48 h cells were lysed and luciferase activity was measured using the Dual-Luciferase Assay System (Promega). All luciferase data were corrected for transfection efficiency based on the Renilla internal control following by subtraction of the pGL4.10[luc2] activity background. The data were calculated as means ± standard error of three independent experiments, each performed in nine replications. Luciferase activities for each transfection are plotted as average fold-change in relation to the pVEGFCwt-Luc.

Western blot analysis
Protein extraction was performed in RIPA buffer (20 mM Tris, pH 7.4, 137 mM NaCl, 10% glycerol, 0.5% (wt/vol) sodium deoxycholate, 0.1% (wt/vol) sodium dodecyl sulfate (SDS), 1% Triton X-100, 2 mM EDTA) supplemented with 1 mM phenylmethylsulfonyl fluoride (PMSF) and protease inhibitor cocktail (Sigma). Lysates (20 µg/lane) were electrophoresed in SDS-PAGE under reducing conditions and transferred to a nitrocellulose (Whatman). Membranes were probed with anti HA-tag monoclonal antibody (Covance, HA.11, 1:100) while β-Tubulin (Santa Cruz, H-235, 1: 500) was used as a loading control. Appropriate HRP-conjugated anti-mouse or anti-rabbit secondary antibodies (Jackson ImmunoResearch, 1:10,000) were used respectively.

Cell invasion assay
The invasion potential of the cells was examined in vitro using BioCoat Matrigel Invasion Chamber (Corning). A total of 80x10^3 cells suspended in serum-free medium were seeded in the upper chamber. To initiate cell invasion medium supplemented with 10% FBS was added as a chemoattractant in the low chamber. Cells were incubated at 37°C for 20 h and invaded cells on the inferior surface of the inserts were fixed, washed, and stained with crystal violet (Sigma). Each assay was repeated two times in duplicates. For analysis the entire Matrigel surface area was imaged (Olympus SZX16 stereomicroscope,
2.5× magnification) and quantified using the ImageJ software. To ensure that the difference in invasion rate is not due to differential cell growth rate, total cell number validation was carried on a parallel plate. Cells were stained with crystal violet, lysed with 1% SDS, 0.1N NaOH solution followed by direct dye intensity measurement.

**Tumor initiation, histologic preparation, immunohistochemistry and morphometric analysis**

Subcutaneous tumors were generated by injecting 2×10^6 single-cell suspensions of cells in 100 µl PBS into a shaved lower right flank of seven-week old mice. D122 Control or Bach1- overexpressing cells were injected to male immunocompetent syngeneic C57BL/6 (Harlan Laboratories), whereas ES2 cells were injected to female immunodeficient CD1 nude (Harlan Laboratories) mice. Orthotopic metastatic tumor growth was initiated by injecting 1×10^6 ES2 cells intraperitoneally at a remote site in the abdomen of seven-week-old female CD1 nude mice (Harlan Laboratories). In all experiments, animals were randomly assigned to the control and overexpressing cells injected groups. Excised mouse tumors were rinsed in ice-cold PBS and then gradually fixed at 4°C in 2.5% and 1% PFA for 24 and 48 hours, respectively. Paraffin-embedded tissue was sectioned serially at 4µm thickness. The first slide was stained with hematoxylin and eosin, whereas other representative slides underwent immunohistochemical staining using the following antibodies: anti CD34 (Cedarlane Laboratories, 1:100), LYVE1 (Fitzgerald Industries, 1:100), cytokeratin 7 (Abcam, ab9021, 1:200) and VEGFC (Santa Cruz, H-190 1:200).

Morphometric blood and lymphatic vessel coverage analysis were performed on CD-34 and LYVE1 stained tumor sections, respectively. Images of CD-34-stained sections were captured with a fluorescence microscope (NI-U, Nikon, Tokyo, Japan), equipped with Plan Fluor objectives connected to CCD camera (DS-Ri1, Nikon). Digital images were collected and assembled using Adobe Photoshop (Adobe Systems). LYVE1 immunohistochemical-stained sections were scanned using the Panoramic Viewer. The density of vessels was evaluated using Image Pro Plus software (Media Cybernetics). For all analysis, investigators were blinded and unaware of group allocation. All animal experiments described in this study were performed according to the Guidelines the Weizmann Institute Animal Care and Use Committee. For immunostaining of ES2 cells, fixation was carried out in 4% paraformaldehyde for 10 minutes. Fixed cells were then blocked for unspecific staining in 0.3% Triton X-100, 10% horse serum for 90 min at RT, incubated with antibodies directed either against HA-tag (Sigma, 1:200) or VEGFC (Santa Cruz, H-190
1:200)) antibody for 2 h at room temperature and visualized by incubating the cells with goat anti-rabbit Cy3 (Jackson ImmunoResearch, 1:10,000) secondary antibody. Nuclei were counterstained with DAPI (Invitrogen).

**Quantitative Real-Time-PCR (qRT-PCR)**

Total RNA was isolated from cultured cells or tumor specimens by PerfectPure RNA Cultured Cell or Tissue kit (5 PRIME) respectively, according to the manufacturer's instructions. RNA (1 µg) from each sample was subjected to first-strand cDNA synthesis with the High-Capacity cDNA Reverse Transcription kit (Applied Biosystems) and random hexamers. Quantitative real-time PCR was conducted with LightCycler-FastStart DNA Master SYBR Green I kit (Roche) using a LightCycler 480 Real-Time PCR System (Roche). The relative expression level of each target gene was determined using glyceraldehyde-3 phosphate dehydrogenase (GAPDH) and beta-2-microglobulin (B2M) as reference genes. Primers used were as follows: human *BACH1*: 5'-TCTTCCAGAAG AGGTGACAGT and 5'-ACTCCACACATTTGCACACT; *VEGFA*: 5'-ATGCGGATC AAACCTCACC and 5'-T CTTTCTTTGGTCTGCATTAC; *VEGFB*: 5'- CCACCAG AGGAAAGTTGGTGC and 5'- ACAGCGCTGCACAGTCAC; *VEGFC*: 5'-GCCACGG CTATTGCAAGCAAAGAT and 5'-AGTTGAGGGTGGCCCTGGTCTCTCCT; *MMP1*: 5'- CTGCCACAACCTGCCAAATG and 5'-CTGTCCCTGAAACAGCCAGCTACTTA; *GAPDH*: 5'-AGGGCTGCTTTTAACTCTGGT and 5'-CCCCACTTGATTTTGGAG GGA; *B2M*: 5'-TTCTGGCCCTGGAGGCTATC and 5'-TCAGGAATTTGTGACTTC CATTTC; Mouse *Bach1*: 5'-TGACAGCGGAGTC CTGTTCTG and 5'-TTATCCGTTGG GCATTGAA; *Vegfa*: 5'-TCTTCAAGCCATCTGTGTG and 5'-GAGGTTTGGATCCGC ATAAATCTG; *Vegfb*: 5'-ACGATGGCGCTGGAATGTGTG and 5'-TGGTCTGGATTCA CATTGCC; *Vegfc*: 5'- GTAAAAACAAACTTTCTCCTACTC and 5'-TCTGGATCCBACH2; *Bactin*: 5'-TGACAGGATGCAGAAGGAGA and 5'-GCC TCCGATCCAGACAGTGG. Zebrafish *bach2b*; 5'-CAGCATGCCAGGAGGT and 5'-GAGTTT GATCCCGCATAATCTG; *Vegfb*: 5'-ACGATGGCGCTGGAATGTGTG and 5'-TGGTCTGG CATTCCAGCAGTC; *Gapdh*: 5'-GACCGCCGATCTTTCAGTTG and 5'-CTTCCCTATT CTCGGCCTTGGACTGT; *B2m*: 5'-CC CGCCTCACATTGAAATCC and 5'- GCCTGATG TATCGATGCACTGG. Zebriflower *bach2b*; 5'-CAGCATGCCAGGAGGT and 5'-A GTGATTGCTCTCGCCCCGC; *bactin*: 5'-TGCAGGATGCAAGGAGGA and 5'-GCC TCCGATCCAGACAGT.
Clearing and immunofluorescence staining

Excised tumors were fixed with 4% PFA for 1 week at 4°C, washed three times with phosphate-buffered saline (PBS), permeabilized with (0.2% Triton X-100 in PBS) for 4 h, and immersed overnight in blocking solution (PBS containing 0.05% Triton X-100 and 10% normal goat serum). For immunofluorescent staining tumors were incubated with primary mouse anti LYVE1 antibody (Fitzgerald Industries) diluted (1:250) in an antibody cocktail (50% blocking solution, 0.05% Triton X-100 in PBSX1) for 1 week at 4°C, washed 24 h (1% blocking solution, 0.05% Triton X-100 in PBS), and probed with an Alexa fluor-594 conjugated goat anti rabbit secondary antibody (Molecular Probes - Life Technologies, A11037, 1: 250) diluted in antibody cocktail for an additional week at 4°C. Following 24 h wash, tumors were subjected to a Whole Organ Blood and Lymphatic Vessels Imaging (WOBLI) clearing procedure (K. Chung et al., 2013; Hama et al., 2011; Oren et al., 2018). Briefly, tumors were re-fixed with 4% PFA for 24-72h at 4°C, transferred to hydrogel solution (4% Acrylamide, 0.025% Bisacrylamide, 0.25% Va-044 and 4% PFA in PBS) for 1 week and passively cleared (200 mM Boric acid and 4% SDS) for 2 weeks at 37°C. Subsequently, tumors were placed in Scale solution (4M Urea, 10% Glycerol and 0.1% Triton X-100) for 48 h. A sequence of 3D images and movies were acquired using Zeiss LSM710 confocal microscope (Zeiss, Oberkochen, Germany).

Microscopy

Images were acquired on Zeiss LSM 780 upright confocal microscope (Carl Zeiss) with a W-Plan Apochromat ×20 objective, NA 1.0. Fluorescent proteins were excited with single-photon laser (488nm). Alternatively, Leica TCS SP8 microscope, equipped with environmental control, 2 internal Hybrid (HyD) detectors and Acusto Optical Tunable Filter (Leica microsystems CMS GmbH, Germany) was used and excitation was done by using 488nm by Ar laser and emission was collected using the internal HyD detector at 510-625nm with gain of 100.

Statistical Analysis

Statistical analyses were performed with analytic computerized software (Statistix 8 Student Edition, Analytical Software, Tallahassee, FL, USA). Comparisons between treatment groups were performed with either ANOVA or Kruskal–Wallis nonparametric ANOVA (alternatively, t-test or Wilcoxon Rank Sum Test were used if only two groups were compared, respectively). ANOVA test was used to analyze normally distributed data
(evaluated by Shapiro–Wilk test) that had equal variances between groups (evaluated by Bartlett’s test), whereas Kruskal–Wallis nonparametric ANOVA was used to analyze data that were not normally distributed and/or had unequal variance between groups; when relevant, Tukey HSD test, or Mean Ranks test, respectively, were used for All-Pairwise Comparisons. Fisher's exact test analysis was used to compare proportional data. Differences were considered significant at $P < 0.05$. Unless otherwise noted, data are presented as mean ± s.e.m.

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