The Effects of Sodium Selenite on Mitochondrial DNA Copy Number and Reactive Oxygen Species Levels of In Vitro Matured Mouse Oocytes

Nassim Ghorbanmehr, Ph.D.¹, ², Mojdeh Salehnia, Ph.D.³*, Mahboobeh Aminooshahi, Ph.D.³

1. Department of Anatomy, Faculty of Medical Sciences, Tarbiat Modares University, Tehran, Iran
2. Department of Biotechnology, Faculty of Biological Sciences, Alzahra University, Tehran, Iran

*Corresponding Address: P.O. Box: 14115-111, Department of Anatomy, Faculty of Medical Sciences, Tarbiat Modares University, Tehran, Iran

Email: salehnim@modares.ac.ir

Received: 3/Aug/2017, Accepted: 22/Oct/2017

Abstract

The aim of present study is to determine the effects of supplementation of oocyte maturation medium with sodium selenite (SS) on oocyte mitochondrial DNA (mtDNA) copy number and reactive oxygen species (ROS) levels.

Materials and Methods: In this experimental study germinal vesicle (GV), metaphase I (MI), and metaphase II (MII) stage oocytes were recovered from 6-8 week old female mice after superovulation. Some of the GV oocytes were cultured and matured in the presence and absence of SS. Then in vivo and in vitro matured (IVM) oocytes were subjected to mitochondria staining by MitoTracker green, ROS analysis, and mtDNA copy number determination using absolute real-time polymerase chain reaction (PCR).

Results: The maturation rate of GV oocytes to the MII stage significantly increased in the SS supplemented group (79.25%) compared to the control group (72.46%, P<0.05). The intensity of mitochondrial staining was not different among the studied groups, whereas the mitochondria distribution in the cytoplasm of the IVM oocytes showed some aggregation pattern. The in vivo obtained MI oocytes had lower ROS levels and higher mtDNA copy numbers than IVM-MII oocytes (P<0.05). The SS supplemented group had significantly lower ROS levels and higher mtDNA copy numbers than the non-treated group (P<0.05).

Conclusion: SS increased oocyte mtDNA copy number by decreasing oxidative stress. SS had an association with better oocyte developmental competence.

Keywords: In Vitro Maturation, mtDNA, Oocyte, Reactive Oxygen Species, Sodium Selenite

Citation: Ghorbanmehr N, Salehnia M, Aminooshahi M. The effects of sodium selenite on mitochondrial DNA copy number and reactive oxygen species levels of in vitro matured mouse oocytes. Cell J. 2018; 20(3): 396-402. doi: 10.22074/cellj.2018.5430.

Introduction

Mitochondria are multifunctional organelles with critical functions in ATP production, calcium homeostasis, and cell apoptosis (1). The localization and presence of these organelles are critical for successful fertilization (2). Different cell types have a variety of mitochondria that have their own genome-mitochondrial DNA (mtDNA) (3). There are various reports of mtDNA copy number in oocytes from different mammalian species (3-7). It is estimated that a human MII oocyte contains approximately 20000 to over 800,000 mtDNA (4, 5); this range is 11000-428,000 in mice matured and integrity of the cytoskeleton between in vitro matured (IVM) and naturally collected mouse oocytes (8).

Reactive oxygen species (ROS) is the byproduct of the oxidative phosphorylation chain system in mitochondria (9). Elevated levels of ROS cause oxidative stress and may lead to alterations in several redox pathways (10). The cells possess powerful enzymatic and non-enzymatic antioxidant defenses to protect against the damaging effects of ROS on DNA, lipids, and proteins (11). Excessive ROS or inadequate antioxidant protection within the cell results in oxidative stress. The cells can be protected against these effects by supplementation of culture media with antioxidants (12-14). Some antioxidants are synthesized by oocyte mitochondria (15). The exogenous antioxidant could improve oocyte and embryo development by decreasing ROS levels (13, 14, 16).

Selenium is a trace element present in the catalytic site of antioxidant enzymes such as glutathione peroxidase. In the form of sodium selenite (SS), it is used as a supplement in culture media and protects cells from oxidative damage (17). Abedelahi et al. (13, 16) have demonstrated that SS can improve the in vitro growth and maturation of mouse preantral follicles by reducing ROS levels and increasing the total antioxidant capacity and glutathione peroxidase activity of follicles. Similarly, Liu et al. (18) showed that SS significantly suppressed oxidative stress, by decreasing oxidative status of the cell and lipid peroxidation levels. Tareq et al. (19) reported an increase in the rate of maturation and embryo development in porcine oocytes cultured in the presence of selenium. The similar beneficial effect of SS in the culture media of porcine embryo was shown by a decrease in apoptosis and increase in expression of glutathione peroxidase (20).

Little is known about mtDNA copy number changes during in vitro maturation of mouse oocytes in the presence of SS as an antioxidant. The present study determined...
the effects of SS supplementation of oocyte maturation medium on oocyte maturation, mtDNA copy number, and ROS levels in comparison with in vitro collected oocytes.

**Materials and Methods**

Unless otherwise indicated, all chemicals were purchased from Sigma Aldrich (Germany). This experimental study used National Medical Research Institute (NMRI) female mice (n=48). The mice were housed in the Animal House at Tarbiat Modares University. The Ethical Committee of the Tarbiat Modares University approved this study (Ref No. 52.1637).

**Germinal vesicle oocyte collection**

Adult female mice 6-8 weeks old (n=38) were superovulated by intraperitoneal injection (i.p.) of 10 IU pregnant mare serum gonadotropin (PMSG, Folligon, Intervet, Australia). Female mice were killed by cervical dislocation 48 hours after the PMSG injection and the dissected ovaries were placed in α-minimal essential medium (α-MEM, Gibco, UK) supplemented with 5% heat-inactivated fetal bovine serum (FBS, Gibco, UK). Antral follicles were punctured with needles to release the oocytes and the cumulus cells were mechanically removed. Oocytes that had a prominent germinal vesicle (GV) and clear ooplasm with 90 µm diameter were selected and collected (n=817).

Some of GV oocytes (n=778) were subjected to in vitro maturation in SS supplemented and non-supplemented groups. The other GV oocytes were analyzed by mitochondrial staining (n=10) and mtDNA copy number analysis (n=29).

**In vivo metaphase I and metaphase II oocyte collection**

For harvesting the ovulated in vivo metaphase I (MI, OV-MI) and metaphase II (MII, OV-MII) oocytes, female mice (n=10) were superovulated by i.p. injection of 10 IU PMSG followed by another injection of 10 IU human chorionic gonadotropin (hCG, Sereno, Switzerland) 48 hours later. The oocytes were collected from the ampullary region of each oviduct 12-16 hours after the hCG injection. Cumulus cells were removed enzymatically by using 0.01% hyaluronidase. The oocytes with homogeneous and clear ooplasm that lacked any polar body or GV were considered OV-MI oocytes, those with one polar body were classified as OV-MII oocytes. The collected OV-MI were studied for mitochondrial staining using MitoTracker green (Molecular Probes, Invitrogen, Eugene, OR, USA) staining. We prepared a stock solution of MitoTracker green at a concentration of 1 mmol in DMSO and stored the solution at -20°C. The in vitro MI oocytes from both experimental groups and in vivo MII oocytes at the GV, MI, and MII stages (n=10 for each group and developmental stage) were stained with 0.2 mmol MitoTracker green in PBS at 37°C for 10 minutes. After washing in PBS, the oocytes were mounted on glass slides and observed under fluorescent microscope at the 490 wavelength (21). Then, a micrograph of each oocyte was prepared and imported into ImageJ software (National Institutes of Health, Bethesda, MD, USA). Next, we analyzed and compared the fluorescence intensity in different groups of oocytes.

**Visualization of the mitochondria using MitoTracker green**

The presence of viable mitochondria was identified by MitoTracker green (Molecular Probes, Invitrogen, Eugene, OR, USA) staining. We prepared a stock solution of MitoTracker green at a concentration of 1 mmol in DMSO and stored the solution at -20°C. The in vitro MI oocytes from both experimental groups and in vivo MII oocytes at the GV, MI, and MII stages (n=10 for each group and developmental stage) were stained with 0.2 mmol MitoTracker green in PBS at 37°C for 10 minutes. After washing in PBS, the oocytes were mounted on glass slides and observed under fluorescent microscope at the 490 wavelength (21). Then, a micrograph of each oocyte was prepared and imported into ImageJ software (National Institutes of Health, Bethesda, MD, USA). Next, we analyzed and compared the fluorescence intensity in different groups of oocytes.

**Reactive oxygen species analysis**

The collected in vitro and in vivo MII oocytes were washed twice with PBS and incubated in 40 mmol/L of tris-HCl buffer (pH=7.0) that contained 5 mmol/L 2,7’ dichlorodihydrofluorescein diacetate (DCF, Merck, Germany) at 37°C for 30 minutes (n=60 for each group for three repeats of 20 pooled oocytes per repeat). Next, the oocytes were sonicated at 50W for 2 minutes, and centrifuged at 4°C and 10000 g for 20 minutes. Then, the supernatant was monitored using a spectrofluorometer at 488 nm excitation and 525 nm emission (22). Data were expressed as µM H2O2 and the mean of the DCF fluorescence intensity. A standard curve was prepared by fluorescence intensity of different concentrations of H2O2.

**DNA extraction from individual oocytes**

We extracted DNA from completely denuded individual oocytes from all studied groups (n=15 for each developmental stage per group). A total of 10 µl of lysis solution that contained 50 mM tris-HCl (pH=8.5), 0.1 mM EDTA, 0.5% Tween-20, and 200 µg/ml proteinase K (Roche, Germany) were added to each tube followed by an overnight incubation at 55°C. The samples were heated to 95°C for 10 minutes to inactivate proteinase.
K. Each sample was used directly as template DNA for polymerase chain reaction (PCR).

**Primer design**

We sought to identify the unique regions of the mouse mitochondrial genome with no pseudogene in the nuclear DNA. The entire sequence of mouse mitochondrial DNA was obtained from NCBI (NC_005089.1). The FASTA format of this sequence was split into 200 bp fragments with 50 bp overlaps. These fragments were searched against the mouse nuclear genome using NCBI Blast. The unique regions of the mitochondrial genome that had no duplicate in the nuclear genome were identified and used for primer design. Specific primers (Table 1) were design using Primer3Plus (http://sourceforge.net/projects/primer3/) software and synthesized at MWG Germany.

**Preparation of standard dilutions**

In order to obtain standard curves we constructed standard DNA by cloning the PCR products. These products were amplified using the primer sets presented in Table 1 into the pTZ57R/T vector (Thermo Scientific Bio, USA). We used the MTF and MTR primers to amplify a 68 bp unique fragment of mtDNA. After electrophoresis, the amplified product was extracted from agarose gel by the Expin™ Combo GP kit (GeneAll Biotechnology, Korea) according to the manufacturer’s protocol. The extracted product was cloned into the vector pTZ57R/T (Thermo Scientific, USA), purified, and sequenced. The recombinant plasmid was linearized and cleaned up by a GeneAll kit (General Biosystem, Korea). The product underwent spectrophotometry. The concentration of recombinant plasmid was calculated and diluted to $3 \times 10^5$ copies/5 µl. We prepared four serial dilutions of standard DNA with at 1/10 standard concentration. These standard dilutions were kept at 4°C until analysis and used in real-time PCR for mtDNA copy number quantification.

**Quantification of mitochondria DNA copy number using real-time polymerase chain reaction**

Real-time PCR was performed to determine the total amount of mtDNA of each single oocyte in all study groups. Each reaction contained 10 µl of SYBR green master mix (Applied Biosystems, USA), 2 µl primer mix (MTF and MTR), 3 µl of sterile water, and 5 µl of oocyte DNA extract (5 µl of each total DNA sample). Each oocyte DNA extract was divided into two wells as duplicates. All real-time runs included four concentrations of serial standard dilutions in triplicate ($R^2 \geq 0.99$). To rule out cross contamination a "no template control" (NTC) was added to each single real-time run. The reactions were performed with an ABI 7500 instrument (Applied Biosystems, CA, USA). Each PCR reaction included an initial denaturation step of 95°C for 10 minutes, followed by 40 cycles of 95°C for 15 seconds, 60°C for 30 seconds, and 72°C for 30 seconds. A melting curve stage was included at the end of the run to confirm the absence of non-specific products and primer dimerization. The copy number of mtDNA for each oocyte was calculated from both duplicate wells.

**Statistical analysis**

Statistical analysis was performed using SPSS software (IBM SPSS statistics 22). All data were presented as mean ± SD. The normality of data was tested by the Kolmogorov-Smirnov test and the data of developmental rates of oocytes were compared by the t test. The mtDNA copy number and ROS level of oocytes were assessed by one-way ANOVA and Tukey’s HSD was used as the post hoc test. Statistical significance was $P<0.05$ for all analyses.

**Results**

**Maturation rate of oocytes**

Table 2 summarizes the maturation rates of GV oocytes. The percent of oocytes which matured in the presence of SS were 6.85 ± 1.28 for the MI stage and 79.25 ± 0.52 for the MII stage. In the absence of SS, the maturation rate for GV oocytes was 6.36 ± 1.50 for the MI stage and 71.32 ± 3.78 for the MII stage. These rates were significantly higher in the SS supplemented group compared to the non-treated control group ($P<0.05$).

| Group | Sodium selenite | Total number | Number of arrested GV (mean% ± SE) | Number of MI (mean% ± SE) | Number of MII (mean% ± SE) | Number of degenerated (mean% ± SE) |
|-------|----------------|--------------|-----------------------------------|--------------------------|---------------------------|----------------------------------|
| Control | - | 461 | 73 (16.67 ± 2.14) | 32 (6.36 ± 1.50) | 335 (71.32 ± 3.78) | 21 (5.63 ± 1.86) |
| Experiment | + | 317 | 38 (11.97 ± 1.54) | 22 (6.85 ± 1.28) | 251 (79.25 ± 0.52)* | 6 (1.90 ± 0.93) |

*; There was significant difference with the control group in the same column ($P<0.05$), GV; Germinal vesicle, MI; Metaphase I oocytes, and MII; Metaphase II oocytes.

Table 1: Mouse mitochondrial specific primer sequences

| Primer code | Primer sequence (5'-3') | Length (bp) |
|-------------|-------------------------|-------------|
| MTF         | GCTAGTGTGAGTGATAGGGTAG    | 20          |
| MTR         | CCAATACGCCCTGTAACAAC     | 22          |

Table 2: The maturation rates of GV oocytes in the presence and absence of sodium selenite
Mitochondrial distribution

We observed mitochondrial distribution of oocytes at different developmental stages in the study groups with a fluorescent microscope using MitoTracker green. The representative micrographs of these oocytes were shown in Figure 1 and 2. The mitochondrial distribution in the cytoplasm of in vivo obtained oocytes consisted of a homogenously diffused pattern (Fig.1). There were some aggregations of mitochondria within the IVM oocytes (Fig.2). We observed similar patterns of mitochondrial distribution in all IVM oocytes with and without SS supplementation (Fig.2A-D).

The florescent intensities with regards to mitochondrial staining (Fig.3A) in GV oocytes was 38.21 ± 0.40. In the presence of SS, it was 37.62 ± 1.24 for IVM-MI and 41.02 ± 0.72 for IVM-MII. In the absence of SS, this finding was 36.99 ± 1.13 for IVM-MI and 39.32 ± 1.12 for IVM-MII. Fluorescence intensity for OV-MI was 39.22 ± 0.72 and 41.69 ± 2.64 for OV-MII. There was no significant difference between the groups.
Fig. 3: The relative fluorescence intensity and reactive oxygen species (ROS) levels in oocytes at different developmental stages. A. Relative fluorescence intensity with MitoTracker green staining in oocytes at different developmental stages obtained from in vivo (OV) and in vitro (IVM) conditions. There was no significant difference between groups. In the presence of sodium selenite (SS+) and in the absence of sodium selenite (SS-). B. ROS levels in MII oocytes derived from in vivo (control) and in vitro conditions. The minimum level of ROS was demonstrated in the in vivo obtained oocytes (OV-MII). There was a significantly lower ROS level in IVM oocytes in the presence of sodium selenite (SS+) compared to the non-sodium selenite SS- treated group (P<0.05).

**Reactive oxygen species concentration**

The ROS levels in all studied MII oocytes (IVM and in vivo collected) were given in Figure 3. Data were shown as μM of H$_2$O$_2$. The level of ROS in IVM-MII oocytes in the presence of SS was 2.44 ± 0.10. In the group without SS, ROS was 3.08 ± 0.06 which was significantly lower in the SS supplemented group compared to the control group (P<0.05). The concentration of ROS in the OV-MII (2.17 ± 0.15) group was significantly lower than both IVM-MII oocyte groups (P<0.05).

**Mitochondrial DNA copy number**

The mean mtDNA copy number in single oocytes for all study groups is shown in Figure 4. This copy number in GV oocytes was 127,468.68 ± 1066.61. The copy number for OV-MI oocytes was 199,335.58 ± 28843.67, whereas for OV-MII oocytes it was 472,881.19 ± 28822.47. The IVM-MI oocytes in the SS supplemented groups had a mean mtDNA copy number of 168,244.12 ± 3759.48. IVM-MII oocytes in the SS supplemented groups had a mean mtDNA copy number of 349,414.2 ± 56027.22. In the group without SS these numbers were 137,223.5 ± 4285.05 (IVM-MI) and 238,720.16 ± 8267.06 (IVM-MII). Oocytes from the SS supplemented group had a significantly higher mtDNA copy number compared to the group without SS (P<0.05). All IVM oocytes had significantly lower mtDNA copy numbers than their respected in vivo obtained oocytes (P<0.05). There was a significantly greater mtDNA copy number for all MII oocytes compared to both GV and MI oocytes in the same group (P<0.05).

**Discussion**

The present study, similar to other investigations, showed that the developmental competence of IVM oocytes was lower than in vivo obtained oocytes (5, 6). However, we demonstrated the beneficial effects of supplementation of culture media with SS as an antioxidant on oocyte maturation by reducing the ROS levels. A similar effect of SS on follicular development and oocyte maturation in mice and bovines has been previously shown by other investigators (13, 16, 23). Selenium acts via intracellular signaling factors that include protein kinase C, nuclear factor-kappa B, and inhibitors of apoptosis proteins (24, 25). Many of the biological actions of selenium are attributed to its powerful antioxidant properties, including direct quenching of ROS and chelation of metal ions (25).

As our data demonstrated, all IVM-MII oocytes had...
higher ROS levels than in vivo obtained oocytes. This level in the SS treated group was lower than the non-treated group. Researchers previously reported increased ROS production during IVM of oocytes (23, 26). Our previous studies also showed that SS improved the in vitro development of follicles by increasing the follicular total antioxidant capacity level and decreasing the ROS level (13, 16). In agreement with this observation, Saito et al. (25) have reported that removal of selenium from the culture medium induced ROS production and cell death. Selenium suppresses oxidative stress by increasing the activity of antioxidant selenoenzymes and inhibits the activation of the PI3K/AKT and ERK signaling pathways stimulated by oxidative stress (18).

An in vivo study by Said et al. (27) showed that SS had a radioprotective effect and improved rat folliculogenesis through increasing ovarian granulosa cell proliferation, and decreasing lipid peroxidation and oxidative stress. ROS is generated during ATP production within the mitochondria and its high level causes oxidative damage of mtDNA (28). On the other hand, mitochondria are especially sensitive to oxidative stress because of its minimal DNA repair enzymes compared to genomic DNA (29). It has been demonstrated that oxidative stress induces mtDNA degradation (28). Research has shown that DNA damage can interfere with POLRMT RNA primer synthesis and disrupt pol γ processivity and affect mtDNA replication (30). In this regard, Ge et al. (8) concluded that the non-physiological condition of controlled ovarian stimulation and in vitro maturation treatments inhibited mtDNA replication, altered mitochondrial function, and increased ROS production. Therefore, damage to the mitochondria might partly explain the low efficiency of assisted reproductive techniques and high rate of embryonic loss associated with these clinical procedures.

Overall, our results revealed a significantly lower mtDNA copy number for all IVM oocytes (MI and MII) compared to in vivo matured oocytes. This might explain that changes in mtDNA copy number could interfere with normal oocyte and embryo development. Therefore, IVM oocytes have lower potential for fertilization and further development. Ge et al. (8) also detected significant differences in the mtDNA copy number and level of ROS in mouse oocytes obtained from in vitro and in vivo conditions.

Additionally, our data showed that mtDNA copy number of oocytes increased significantly from the GV (127,468) to the MII (472,881) stages. The average mtDNA copy number determined in the present study was close to other investigations (6, 7). In contrast, no significant increase in mtDNA copy number from GV to IVM derived MII oocytes were reported in ovines and humans (4, 31). Attempts to quantify the amount of mtDNA in oocytes using PCR-based methods showed highly variable results. This discrepancy in mtDNA within the oocytes could by mainly related to technical error, different sources of oocytes (pooled or single), and different developmental stages of oocytes in several species (3-5, 32, 33).

Our results, for the first time, demonstrated that supplementation of maturation medium with SS could increase the mtDNA copy number of MI and MII oocytes compared to the non-treated group. Perhaps, the mitochondrial biogenesis in oocytes was stimulated during IVM in the presence of SS and was associated with higher developmental competence of the oocytes. In agreement with this suggestion, it has been shown that mtDNA copy number could change in response to environmental signals such as temperature, energy deprivation, nutrients, and growth factors (34).

This study showed that the mitochondrial distribution in IVM oocytes had some aggregation in comparison with in vivo obtained oocytes; however, the intensity of mitochondrial staining did not differ in these studied groups. Similarly, Stojkovic et al. (21) showed that the mitochondrial clumps became larger after IVM of oocytes. Liu et al. (35) demonstrated that the distribution of mitochondria in IVM oocytes differed slightly from that of in vivo obtained oocytes. They concluded that this different pattern resulted in the reduced developmental potential of IVM oocytes. Insufficient culture conditions might prevent mitochondrial migration within the ooplasm and affect cytoplasmic maturation (36). Thus, proper distribution of mitochondria during IVM of oocytes is critical for further development. In this regard, Kim et al. (14) have reported that treating oocytes with antioxidant could improve cytoplasmic maturation and cause morphologically uniform distribution of mitochondria and lipid droplets in the cytoplasm.

Conclusion
SS increases oocyte mtDNA copy number by decreasing oxidative stress and is associated with better oocyte developmental competence.

Acknowledgements
This work was financially supported by Tarbiat Modares University as a post-Doctorate thesis and by the Iran National Science Foundation. We thank Mr. Pour Beyranvand for providing excellent technical assistance. There is no conflict of interest in this study.

Author’s Contributions
N.G.; Designed and performed the experiments, analyzed data and co-wrote the paper. M.S.; Supervised the research, designed the experiments and co-wrote the paper. M.A.; Performed experiments and analyzed data. All authors read and approved the final manuscript.

References
1. Jacobson J, Duchen MR. Interplay between mitochondria and cellular calcium signaling. Mol Cell Biochem. 2004; 256(1-2): 209-218.
2. Schatten H, Sun QY, Prather R. The impact of mitochondrial function/dysfunction on IVF and new treatment possibilities for infertility. Reprod Biol Endocrinol. 2014; 12: 111.
3. Sato D, Itami N, Tasaki H, Takeo S, Kuwayama T, Iwata H. Relationship between mitochondrial DNA copy number and SIRT1 expression in porcine oocytes. PLoS One. 2014; 9(4): e94488.
4. Barritt JA, Kokot M, Cohen J, Steuerwald N, Brenner CA. Quantification of human ooplasmic mitochondria. Reprod Biomed Online. 2002; 4(3): 243-247.

5. Santos TA, EI Shourbagy S, St John JC. Mitochondrial content reflects oocyte variability and fertilization outcome. Fertil Steril. 2006; 85(3): 584-591.

6. Cao L, Shiota H, Horii T, Nagao Y, Imai H, Abe K, et al. The mitochondrial bottleneck occurs without reduction of mtDNA content in female mouse germ cells. Nat Genet. 2007; 39(3): 386-390.

7. Wai T, Ao A, Zhang X, Cyr D, Dufort D, Shoubridge EA. The role of mitochondrial DNA copy number in mammalian fertility. Biol Reprod. 2010; 83(1): 52-62.

8. Ge H, Tollner TL, Hu Z, Da M, Li X, Guan H, et al. Impaired mitochondrial function in murine oocytes is associated with controlled ovarian hyperstimulation and in vitro maturation. Reprod Fertil Dev. 2012; 24(7): 945-952.

9. Chen Q, Vazquez EJ, Moghaddas S, Hoppel CL, Lesnfsky EJ. Production of reactive oxygen species by mitochondria: central role in oocyte signaling. J Biol Chem. 2003; 278(36): 36027-36031.

10. Agarwal A, Gupta S, Sekhon L, Shah R. Redox considerations in female reproductive function and assisted reproduction: from molecular mechanisms to health implications. Antioxid Redox Signal. 2008; 10(8): 1375-1403.

11. Birben E, Sahiner UM, Sackesen C, Erzurum SC, Kalayci O. Reactive oxygen species formation and oxidative stress in bovine granulosa cells: possible involvement of nitric oxide. Domest Anim Endocrinol. 2000; 18(1): 1-17.

12. Wai T, Ao A, Zhang X, Cyr D, Dufort D, Shoubridge EA. The role of mitochondrial DNA copy number in mammalian fertility. Biol Reprod. 2010; 83(1): 52-62.

13. Abedelahi A, Salehnia M, Allameh AA, Davoodi D. Sodium selenite caused by selenium deficiency and protective effect of antioxidants. J Biol Chem. 2003; 278: 39428-39434.

14. Cetica PD, Pintos NL, Dalvit GC, Beconi MT. Antioxidant enzyme activity and oxidative stress in bovine oocyte in-vitro maturation. IUBMB Life. 2001; 51(1): 57-64.

15. Said RS, Nada AS, El-Demerdish E. Sodium selenite improves folliculogenesis in radiation-induced ovarian failure: a mechanistic approach. PLoS One. 2012; 7(12): e50928.

16. Gopee NV, Johnson VJ, Sharma RP. Sodium selenite-induced apoptosis in murine B-lymphoma cells is associated with inhibition of protein kinase C-delta, nuclear factor kappab, and inhibitor of apoptosis protein. Toxicol Sci. 2004; 78(2): 204-214.

17. Stojkovic M, Tayefi-Nasrabadi H, Mohammadnejad D, Abedelahi A, et al. Oxidative stress and antioxidant defense. World Allergy Org J. 2012; 5(1): 9-19.

18. Farzollahi M, Tayefi-Nasrabadi H, Mohammadnejad D, Abedelahi A. Supplementation of culture media with vitamin E improves mouse antral follicle maturation and embryo development from vitrified ovarian tissue. J Obstet Gynaecol Res. 2016; 42(5): 526-535.

19. Santos TA, El Shourbagy S, St John JC. Mitochondrial content reflects oocyte variability and fertilization outcome. Fertil Steril. 2006; 85(3): 584-591.

20. Uhms SJ, Gupta MK, Yang JH, Lee SH, Lee HT. Selenium improves the developmental ability and reduces the apoptosis in porcine parthenotes. Mol Reprod Dev. 2007; 74(11): 1386-1394.

21. Stojkovic M, Machado SA, Stojkovic P, Zakhartchenko V, Hutzler P, Goncalves PB, et al. mitochondrial distribution and adenosine triphosphate content of bovine oocytes before and after in vitro maturation: correlation with morphological criteria and developmental capacity after in vitro fertilization and culture. Biol Reprod. 2001; 64(3): 904-909.

22. LeBel CP, Ischiropoulos H, Bondy SC. Evaluation of the probe 2′,7′-dichlorofluorescin as an indicator of reactive oxygen species formation and oxidative stress. Chem Res Toxicol. 1992; 5(2): 227-231.

23. Basini G, Tamanini C. Selenium stimulates estradiol production in bovine granulosa cells: possible involvement of nitric oxide. Domest Anim Endocrinol. 2000; 18(1): 1-17.

24. Gontijo NM, Johnson VJ, Sharma RP. Sodium selenite-induced apoptosis in murine B-lymphoma cells is associated with inhibition of protein kinase C-delta, nuclear factor kappab, and inhibitor of apoptosis protein. Toxicol Sci. 2004; 78(2): 204-214.

25. Saito Y, Yoshida Y, Akazawa T, Takahashi K, Niki E. Cell death caused by selenium deficiency and protective effect of antioxidants. J Biol Chem. 2003; 278: 39428-39434.