Inositol (1,4,5)-trisphosphate (InsP$_3$)-gated Ca Channels from Cerebellum: Conduction Properties for Divalent Cations and Regulation by Intraluminal Calcium

ILYA BEZPROZVANNY and BARBARA E. EHRLICH

From the Laboratory of Molecular Hermeneutics, Departments of Medicine and Physiology, University of Connecticut, Farmington, Connecticut 06032-1305

ABSTRACT The conduction properties of inositol (1,4,5)-trisphosphate (InsP$_3$)-gated calcium (Ca) channels (InsP$_3$R) from canine cerebellum for divalent cations and the regulation of the channels by intraluminal Ca were studied using channels reconstituted into planar lipid bilayers. Analysis of single-channel recordings performed with different divalent cations present at 55 mM on the trans (intraluminal) side of the membrane revealed that the current amplitude at 0 mV and the single-channel slope conductance fell in the sequence: Ba (2.2 pA, 85 pS) > Sr (2.0 pA, 77 pS) > Ca (1.4 pA, 55 pS) > Mg (1.1 pA, 42 pS). The mean open time of the InsP$_3$R recorded with Ca (2.9 ms) was significantly shorter than with other divalent cations (~5.5 ms). The "anomalous mole fraction effect" was not observed in mixtures of divalent cations (Mg and Ba), suggesting that these channels are single-ion pores. Measurements of InsP$_3$R activity at different intraluminal Ca levels demonstrated that Ca in the submillimolar range did not potentiate channel activity, and that very high levels of intraluminal Ca (> 10 mM) decreased channel open probability 5-10-fold. When InsP$_3$R were measured with Ba as a current carrier in the presence of 110 mM cis potassium, a $P_{Ba}/P_{K}$ of 6.3 was estimated from the extrapolated value for the reversal potential. When the unitary current through the InsP$_3$R at 0 mV was measured as a function of the permeant ion (Ba) concentration, the half-maximal current occurred at 10 mM trans Ba.

The following conclusions are drawn from these data: (a) the conduction properties of InsP$_3$R are similar to the properties of the ryanodine receptor, another intracellular Ca channel, and differ dramatically from the properties of voltage-gated Ca channels of the plasma membrane. (b) The estimated size of the Ca current through the InsP$_3$R under physiological conditions is 0.5 pA, approximately
four times less than the Ca current through the ryanodine receptor. (c) The potentiation of InsP3R by intraluminal Ca in the submillimolar range remains controversial. (d) A quantitative model that explains the inhibitory effects of high trans Ca on InsP3R activity was developed and the kinetic parameters of InsP3R gating were determined.

**INTRODUCTION**

Inositol 1,4,5-trisphosphate (InsP3) is a second messenger used by many cell types to stimulate release of intracellular calcium (Ca) from intracellular Ca stores into the cytoplasm (Berridge, 1993; Berridge and Irvine, 1989) due to activation of InsP3-gated Ca channels (Bezprozvanny and Ehrlich, 1993a; Bezprozvanny, Watras, and Ehrlich, 1991a; Ehrlich and Watras, 1988; Ferris, Huganir, Supattapone, and Snyder, 1989; Maeda, Kawasaki, Nakade, Yokota, Taguchi, Kasai, and Mikoshiba, 1991; Watras, Bezprozvanny, and Ehrlich, 1991). InsP3 receptors (InsP3R) have been purified to homogeneity from several sources (Chadwick, Saito, and Fleischer, 1990; Maeda et al., 1991; Mourey, Verma, Supattapone, and Snyder, 1990; Supattapone, Worley, Baraban, and Snyder, 1986) and multiple isoforms of InsP3R have been cloned (Blondel, Takeda, Janssen, Seino, and Bell, 1993; Furuichi, Miyawaki, Wada, Maeda, and Mikoshiba, 1989; Kume, Muto, Aruga, Nakagawa, Michikawa, Furuichi, Nakade, Okano, and Mikoshiba, 1993; Mignery, Newton, Archer, and Sudhof, 1990; Sudhof, Newton, Archer, Ushkaryov, and Mignery, 1991). Experimental results using Ca release assays and single-channel recordings have indicated that InsP3R are regulated by a variety of cofactors and cellular processes including cytosolic and intraluminal free Ca levels, phosphorylation of the InsP3R, intracellular pH, and ATP levels (Bezprozvanny and Ehrlich, 1993a; Bezprozvanny et al., 1991a; Ferris, Cameron, Bredt, Huganir, and Snyder, 1991a; Ferris, Huganir, Bredt, Cameron, and Snyder, 1991b; Ferris, Huganir, and Snyder, 1990; Finch, Turner, and Goldin, 1991; Iino, 1990, 1991; Missiaen, De Smedt, Droogmans, and Casteels, 1992a; Nunn and Taylor, 1992; Supattapone, Danoff, Theibert, Joseph, Steiner, and Snyder, 1988a; Volpe and Alderson-Lang, 1990). Despite this wealth of information about InsP3R structure and regulation, very little is known about the conduction properties of InsP3-gated Ca channels.

On the other hand, the permeability and selectivity properties of another type of intracellular Ca channel, the ryanodine receptor (RyR), was characterized in detail (Anderson, Lai, Rousseau, Erickson, and Meissner, 1989; Lai, Erickson, Rousseau, Liu, and Meissner, 1988; Lindsay, Manning, and Williams, 1991; Lindsay and Williams, 1991; Liu, Lai, Rousseau, Jones, and Meissner, 1989; Smith, Imagawa, Ma, Fill, Campbell, and Coronado, 1988; Tinker, Lindsay, and Williams, 1992, 1993; Tinker and Williams, 1992; Williams, 1992). In these reports, the activity of purified cardiac and skeletal RyR was recorded in planar lipid bilayers under a variety of ionic conditions and, based on these data, a quantitative model of ion conduction in the RyR was developed (Tinker et al., 1992) using Eyring rate theory (described in Hille [1992]). Tinker et al. (1992) concluded that permeation of divalent cations through the RyR occurs in a single file with, at most, one ion able to occupy the conduction pathway at any time (Tinker et al., 1992; Tinker and Williams, 1992). This result is distinct from a well-established model of ion permeation through voltage-gated Ca
channels of the plasma membrane (Almers and McClesky, 1984; Hess and Tsien, 1984; Tsien, Hess, McCleskey, and Rosenberg, 1987), where binding of at least two Ca ions in the pore is necessary for passage of the Ca current. Is the RyR an exception among Ca channels or do intracellular Ca channels operate in a manner fundamentally different from Ca channels of the plasma membrane? To answer this question and to obtain more information about the permeability and selectivity properties of InsP3R, we studied the conduction of divalent cations through InsP3-gated channels from canine cerebellum. Channels were reconstituted into planar lipid bilayers, as was done previously to characterize InsP3R regulation by cytosolic Ca (Bezprozvanny et al., 1991a) and ATP (Bezprozvanny and Ehrlich, 1993a).

In the first part of this report we describe the conduction properties of InsP3R for divalent cations. In the second part, we examine the controversial topic concerning the regulation of InsP3R by intraluminal Ca (compare Combettes, Claret, and Champeil [1992] and Shuttleworth [1992] with Missiaen et al. [1992a] and Missiaen, De Smedt, Droogmans, and Casteels [1992b]). In the last part of the paper we estimate the size of the Ca current through a single InsP3-gated channel expected under physiological conditions.

Preliminary accounts of part of this work have been published in abstract form (Bezprozvanny and Ehrlich, 1993b; Bezprozvanny and Ehrlich, 1994).

**METHODS**

**Preparation of Microsomes from Canine Cerebellum**

Microsomal vesicles of canine cerebellum were obtained using gradient centrifugation procedures. Endoplasmic reticulum vesicles derived from canine cerebella were made as described previously (Watras et al., 1991). Briefly, cerebella, excised from anesthetized dogs, were minced, and then homogenized with a Brinkman Polytron in 4 vol of buffer A (5 mM NaNO3/100 μM EGTA/20 mM HEPES, pH 7.4). Another 4 vol of buffer A were added to the homogenate, and the suspension was centrifuged for 20 min at 4 k gmax (Beckman 35 Ti rotor, Beckman Instruments, Inc., Fullerton, CA). The supernatant fluid was filtered through two layers of cheese cloth, and the filtrate was centrifuged for 30 min at 90 k gmax (Beckman 60 Ti rotor, Beckman Instruments, Inc.). The pellet from the latter spin was resuspended in buffer B (0.6 M KCl/5 mM NaN3/20 mM NaP2O7/10 mM HEPES, pH 7.2) to a final concentration of 4 ml/g cerebellum, and centrifuged for 20 min at 4 k gmax (Beckman 60 Ti rotor, Beckman Instruments, Inc.). The resulting supernatant fluid was centrifuged for 30 min at 63 k gmax (Beckman 60 Ti rotor, Beckman Instruments, Inc.). The pellet from this last spin was resuspended in buffer C (10% sucrose, 10 mM MOPS, pH 7.0) and either frozen in liquid nitrogen (and stored at −80°C), or used immediately for reconstitution experiments.

**Planar Lipid Bilayers**

Cerebellar microsomes were fused with planar lipid bilayers, made from phosphatidylethanolamine and phosphatidylserine (75:25%; Avanti Polar Lipids, Inc., Birmingham, AL). Bilayers were formed by painting a solution of lipid in decane across a 100-μM hole in a Teflon sheet that bisected a lucite chamber. The hole was prepped with a phosphatidylcholine:phosphatidylserine mixture (75:25%) before formation of the membrane. Channel incorporation into the bilayer was followed as described previously (Bezprozvanny and Ehrlich, 1993a). Briefly, vesicles (~5 μg protein) were added to the cis chamber which contained 800 mM KCl, 250 mM...
Tris-HEPES, 10 mM CaCl₂, pH 7.35. The trans chamber, which was held at virtual ground, contained 55 mM of the hydroxide of the divalent cation under investigation (Mg, Ca, Sr, Ba) dissolved in 250 mM HEPES, pH 7.35. Solutions in both chambers were stirred until the activity of several potassium (K) and/or chloride (Cl) channels appeared in the current recordings; this activity was used as an indication of fusion of vesicles to the bilayer. Sometimes the KCl concentration in the cis chamber was further increased by addition of an aliquot of 3 M KCl in order to stimulate fusion of the vesicles with the bilayer. In some experiments, the bilayer was broken and reformed in the presence of microsomes in order to facilitate fusion. After fusion of microsomes, the cis chamber was perfused with 10 vol of Tris-HEPES solution (250 mM HEPES/110 mM Tris, pH 7.35) leaving the divalent cation on the trans side of the bilayer as the only small ion present in the system. In the series of experiments where InsP₃-gated channels were recorded in the presence of cytoplasmic K, the cis chamber was perfused with 110 mM KOH dissolved in 250 mM HEPES, pH 7.35 instead of the Tris-HEPES solution.

The vesicles fused to the bilayer with an orientation such that the cis side corresponded to the cytoplasmic side of the reticular membrane and the trans side corresponded to the lumen of the reticulum. After perfusion of the cis chamber, InsP₃-gated channels were activated by addition of 2 μM InsP₃ to the cis side. In all experiments in this report, InsP₃-gated channels were recorded in the presence of cytoplasmic K, the cis chamber was perfused with 110 mM KOH dissolved in 250 mM HEPES, pH 7.35 instead of the Tris-HEPES solution. Activity of InsP₃-gated channels was recorded using a bilayer clamp amplifier (BC-525A, Warner Instrument Corp., Hamden, CT) under voltage clamp conditions. The junction potential between the cis and trans chambers was measured using 3 M KCl 1% agarose bridge as the only electrical connection between cis and trans solutions. The junction potential was measured for each pair of solutions used and compensation for junction potential was done electronically during the experiments. For experiments with Tris (110 mM) in the cis chamber the junction potential was very close to 0 mV, and for experiments with K (110 mM) in the cis chamber it was ~2 mV.

In some cases the concentration of ions in the trans chamber was manipulated during the experiment. Because perfusion of the trans chamber often caused breakage of the bilayer, we changed ion concentrations in the trans compartment by addition of aliquots of a concentrated stock solution with stirring for at least 1 min. A volume of the trans solution equal to the added volume was withdrawn after each addition in order to keep the volume of solution in the trans compartment constant. If this protocol is followed, then the concentration of each ion species in the trans chamber after k increment additions of volume v (typically in the range 75–150 μl in our experiments) to the chamber with total volume V (3 ml in our experiments) is equal to:

\[
  c_k = (c_0 - s)[V/(V + v)]^k + s
\]

where \(c_k\) is the ion concentration in the trans compartment after \(k\) additions, \(c_0\) is the concentration of the same ion in the beginning of the experiment, \(s\) is the concentration of this ion in the stock solution added. This equation was used for calculation of the trans ion concentration when necessary.

In some experiments the free Ca concentration in the trans chamber containing a divalent cation at 55 mM (Ba, Sr, Mg) dissolved in HEPES and a Ca-EGTA mixture, had to be
determined. First, the level of contaminated Ca in the divalent cation/HEPES solution was determined using atomic absorption spectroscopy (Galbraith Laboratories Inc., Knoxville, TN). Then, the free trans Ca concentration was estimated using a computer program developed by Fabiato (1988) based on the total trans Ca concentration (contaminated and added) and using apparent stability constants for divalent cation-EGTA complexes equal to \(1.23 \times 10^7 \text{ M}^{-1}\) for Ca-EGTA, \(2.40 \times 10^4 \text{ M}^{-1}\) for Ba-EGTA, \(3.81 \times 10^4 \text{ M}^{-1}\) for Sr-EGTA, and \(70.8 \text{ M}^{-1}\) for Mg-EGTA at pH 7.35 and 20°C (Fabiato, 1988).

**Analysis**

Data were stored with a tape recorder and analyzed using a personal computer. For computer analysis using pClamp 5.5, data were filtered to 800 Hz by an 8-pole Bessel filter and digitized at 2 kHz. Acquired data were filtered using a digital Gaussian filter (pClamp 5.5) with a cutoff frequency of 500 Hz and events longer than 2 ms were used for construction of the dwell time histograms; the threshold used for channel analysis was adjusted manually based on the size of the current. Events longer than 4 ms were chosen for construction of amplitude histograms. Because channel openings had a decreased duration in the experiments with Ca as a current carrier (see Results), the data from these experiments were digitized at 4 kHz and events longer than 1.5 ms were used in the construction of dwell time distributions. Under the experimental conditions used in this report, InsP3-gated channels spent more than 90% of the open time in the main subconductance state (Watras et al., 1991); only openings to this level were considered in the analysis. Evidence for the presence of two to three functional channels in the bilayer was obtained in almost all experiments (e.g., Fig 1, traces 2 and 3). The number of active channels in the bilayer was estimated as a maximal number of simultaneously active channels during the course of an experiment (Horn, 1991). The open probability for one channel was calculated using the binomial distribution for levels 0, 1, and 2, and assuming that the channels were identical and independent (Colquhoun and Hawkes, 1983).

In some experiments, the permeability ratio of divalent (\(D^{2+}\), present in trans chamber) and monovalent (\(M^+\), present in cis chamber) cations (\(P_D/P_M\)) was estimated based on the extrapolated value of the reversal potential (\(E_{rev}\)). In our calculations we used the following modification of Goldman-Hodgkin-Katz equation (Hille, 1992):

\[
E_{rev} = \frac{RT}{2F} \ln \frac{4P_D[D^{2+}]}{P_M[M^+]} \tag{2}
\]

where \([D^{2+}]\) and \([M^+]\) are the divalent and monovalent cation concentrations respectively. The symbols \(R\), \(T\), and \(F\) have their usual meaning. As all experiments were conducted at room temperature (20°C), a value of 25.3 mV was used for \(RT/F\).

Where appropriate, average values are presented as mean ± SE (number of experiments).

**RESULTS**

*InsP3-gated Ca Channels from Canine Cerebellum in Planar Lipid Bilayers*

After fusion of canine cerebellar endoplasmic reticulum vesicles with planar lipid bilayers and perfusion of the cis (cytoplasmic) chamber with Tris-HEPES solution (see Methods), channel openings were not observed in the absence of InsP3 (Fig. 1, top trace). Addition of 2 µM InsP3 to the cis side of the membrane induced channel openings (Fig. 1, second and third traces). Because the currents are shown on a compressed time scale in Fig. 1, most of the channel openings are not resolved. When the time scale of the current recording is expanded 15-fold (Fig. 1, fourth trace),
most of the openings are completely resolved. Usually several InsP$_3$-gated channels were present in the bilayer (at least two channels are active in the experiment shown in Fig. 1). The activity of InsP$_3$-gated channels in all experiments in this paper was recorded under "optimal" conditions, i.e., after addition of 2 μM InsP$_3$ in the presence of 0.2 μM free Ca (Bezprozvanny et al., 1991a; Finch et al., 1991; Iino, 1990) and 0.5 mM ATP (Bezprozvanny and Ehrlich, 1993a; Ferris et al., 1990; Iino, 1991) on the cytoplasmic side of the membrane. Because multiple channels often incorporated into the membrane, the exact value for the single-channel open probability could not be obtained, but it was estimated to be in the range of 5–12% under the optimal conditions used assuming that the channels in the bilayer were

![Figure 1](image)

**FIGURE 1.** InsP$_3$-gated channels from endoplasmic reticulum of cerebellum in planar lipid bilayers. Calcium and ATP act as coagonists of the InsP$_3$R (Bezprozvanny and Ehrlich, 1993a; Bezprozvanny et al., 1991a) but were not able to activate the channels without InsP$_3$ present (first trace). Addition of 2 μM InsP$_3$ to the cis (cytoplasmic) side opened at least two channels incorporated in the membrane (second trace; channel opening are upwards). Barium, present at 55 mM on the trans (intraluminal) side, was used as the current carrier. Channel activity was recorded under voltage clamp conditions with a transmembrane potential of 0 mV. InsP$_3$-gated channels remained active for extended periods of time without desensitization (second and third traces are separated by 5 min). Most of the channel openings are not resolved due to the compressed time scale used. When the time scale is expanded 15 times (fourth trace), most of the channel openings are resolved. Heparin completely and irreversibly inhibited the activity of the InsP$_3$R (last trace). See text for details and the exact composition of experimental solutions. Experiment 20427IP3.

identical and independent (see Methods). In the experiment shown in Fig. 1, Ba was the current carrier (see below). Once activated by addition of InsP$_3$, channels maintained their activity for extended periods of time (from several minutes to 2 h in some experiments) with no signs of time-dependent channel inactivation (Fig. 1, second and third traces were separated by 5 min). Channel openings were completely abolished by the addition of heparin, a known inhibitor of InsP$_3$ binding and InsP$_3$-induced Ca release (Ghosh, Eis, Mullaney, Ebert, and Gill, 1988; Kobayashi, Somlyo, and Somlyo, 1988; Supattapone et al., 1988b), at concentrations as low as 0.5 μg/ml (Fig. 1, bottom trace). Ruthenium red (10 μM) or ryanodine (2 μM), which
are known to act on the RyR (Smith et al., 1988), had no effect on InsP₃-gated channel activity (data not shown).

Divalent Cation Permeation through InsP₃-gated Ca Channels

To study the permeation of InsP₃-gated Ca channels for the alkaline earth cations, a series of experiments was performed where the divalent cation under investigation was used as the permeant ion in the trans (intraluminal) chamber. The cis chamber in these experiments contained the same Tris-HEPES solution (see Methods) that was used in previous reports (Bezprozvanny and Ehrlich, 1993a; Bezprozvanny et al., 1991a) and in the experiment shown in Fig. 1. In agreement with the results obtained for another intracellular Ca channel, the RyR (Ashley, 1989; Smith, Coronado, and Meissner, 1986; Tinker and Williams, 1992; Williams, 1992), all four alkaline earth cations tested (Ba, Sr, Ca, Mg) carried measurable current through InsP₃-gated channels incorporated in planar lipid bilayers at a transmembrane potential of 0 mV (Fig. 2 A). With all four cations used, channel currents were qualitatively similar, although some quantitative differences were observed. Single-channel data analyses revealed differences in the unitary current amplitude (A–D in Fig. 2 B) and in the
TABLE I

| Ion | Current (pA) | Mean open time (ms) | Conductance (pS) |
|-----|--------------|---------------------|------------------|
| Ba  | 2.2 ± 0.1 (n = 8) | 6.4 ± 1.0 (n = 7) | 85 ± 3 (n = 4) |
| Sr  | 2.0 ± 0.1 (n = 4)  | 5.9 ± 1.6 (n = 3) | 77 ± 2 (n = 3)  |
| Ca  | 1.4 ± 0.1 (n = 6)  | 2.9 ± 0.2 (n = 5) | 55 ± 4 (n = 3)  |
| Mg  | 1.1 ± 0.1 (n = 4)  | 4.3 ± 0.3 (n = 4) | 42 ± 2 (n = 3)  |

mean open time of the channels (E–H in Fig. 2 B). The observed mean closed time of the channels varied dramatically from experiment to experiment, most probably reflecting the presence of a variable number of channels incorporated in the bilayer in different experiments. Therefore, a reliable estimate of the mean closed time and open probability variation as a function of the ion species used as a current carrier could not be obtained.

The properties of InsP₃-gated channels obtained in this series of experiments are summarized in Table I. At a transmembrane potential of 0 mV and with 55 mM divalent cation in the intraluminal (trans) chamber, the current amplitude ranged from 2.2 ± 0.1 pA (n = 8) with Ba as a current carrier to 1.1 ± 0.1 pA (n = 4) with Mg (Fig. 2, Table I) and fell in the sequence Ba > Sr > Ca > Mg. The mean open time of the channels was significantly smaller with Ca as a current carrier (2.9 ± 0.2 ms [n = 5]) than with other alkaline earths (~5.5 ms) (Fig. 2, Table I).

The single-channel conductance of the channels with different divalent cations was estimated from the slope of the current-voltage relationship between 0 and −40 mV. The planar lipid bilayers became unstable at transmembrane potentials below −40 mV and the current-voltage relationship was not linear at transmembrane potentials above 0 mV. It also was difficult to separate channel openings from the baseline noise at positive potentials, especially in experiments with Mg and Ca as a current carriers. Between 0 and −40 mV the current-voltage relationship was ohmic with all divalent cations tested (Fig. 3). The single-channel conductance (determined using least squares linear regression) ranged from 85 ± 3 pS (n = 4) for Ba to 42 ± 2 pS (n = 3).
for Mg and fell in the same sequence as the values of the current amplitude at 0 mV: 
Ba > Sr > Ca > Mg (Fig. 3, Table I).

In this series of experiments channels were recorded with Tris-HEPES solution on 
the cytoplasmic (cis) side of the membrane. No measurable currents in the opposite 
direction (carried by Tris$^+$ ions) through InsP$_3$-gated channels were observed even at 
potentials as high as +60 mV (data not shown). Therefore, a true value for the 
reversal potential could not be obtained from these experiments. A value for the 
reversal potential could not be obtained by extrapolating the linear segment of 
current-voltage relationship either because of the observed nonlinearity of the 
current-voltage relationship at positive potentials.

Mole Fraction Behavior of the Unitary Current through InsP$_3$-gated Channels

How many divalent cations can be present simultaneously in the permeation pathway 
of an InsP$_3$R? The "anomalous mole fraction effect" is one of the strongest pieces of 
evidence in favor of multiple occupancy of voltage-gated Ca channels (Almers and 
McClesky, 1984; Hess and Tsien, 1984; Tsien et al., 1987). To see if the anomalous 
mole fraction effect is observed for InsP$_3$R, we measured the activity of InsP$_3$-gated 
channels in planar lipid bilayers in the presence of varied molar ratios of Ba and Mg. 
Barium and Mg were chosen because the single-channel conductance and the unitary 
current amplitude at any given transmembrane potential differ most dramatically for 
this pair of ions (Figs. 2 and 3; Table I). To exclude potential masking effects of 
contaminating Ca, 1 mM EGTA was added to the trans chamber in these experi-
ments. In these experiments, InsP$_3$-gated channels were first recorded in the 
presence of 55 mM Ba/HEPES and 1 mM EGTA in the trans chamber and then a 
series of additions of a solution of 55 mM Mg/HEPES and 1 mM EGTA to the trans 
chamber were made. After each addition, an equal volume of solution was withdrawn 
in order to keep the total volume in the trans chamber constant. The resulting trans 
concentrations of Ba and Mg were calculated as described in Methods using Eq. 1. 
This protocol allowed us to measure the current amplitude of the same InsP$_3$-gated 
channel at different molar ratios of Ba to Mg (the molar fraction for Mg ranged from 
0 to 0.9) while the total concentration of divalent cations in the trans chamber was 
kept at 55 mM. In parallel experiments, channels were first recorded in the presence 
of 55 mM Mg/HEPES and 1 mM EGTA in the trans chamber and subsequent 
additions of 55 mM Ba/HEPES and 1 mM EGTA were performed, so that the molar 
fraction for Mg ranged from 1 to 0.1. The average sizes of the unitary currents at 
transmembrane potentials of 0 mV and −20 mV obtained in these experiments are 
plotted as functions of the Mg molar fraction in Fig. 4. The data were pooled from 
both series of experiments. The relatively small variation in the size of the current at 
any given Mg molar fraction provides evidence that Eq. 1 gives a reasonable 
estimation of the Mg and Ba concentrations during our experiments. It is apparent 
that both functions are monotonic; that is, the anomalous mole fraction effect was not 
observed for InsP$_3$R at either 0 mV or −20 mV for mixtures of Ba and Mg. Linear 
regression provided satisfactory fits to the data (Fig. 4), indicating that the affinities 
of the divalent cation binding site in the InsP$_3$R are similar for Ba and Mg. The
single-channel slope conductance can be estimated from the differences in the current amplitudes at 0 and -20 mV. From the data shown in Fig. 4, the slope conductance is a monotonic function of the Mg molar fraction; the slope conductance decreases from 73.5 to 38.5 pS as the Mg molar fraction increases from 0 to 1. These data are in good agreement with results obtained for the RyR from sheep cardiac muscle using a similar approach (Lindsay et al., 1991; Tinker and Williams, 1992).

The Effect of the Intraluminal Ca Level on the Activity of InsP3-gated Channels

Regulation of the InsP3R by intraluminal Ca was studied in a series of experiments where the activity of InsP3-gated channels was measured at different intraluminal (trans) Ca concentrations. Similar results were obtained using either Ba, Sr, or Mg as a current carrier. The use of divalent cations as current carriers in these experiments could compromise our results if these ions interact with intraluminal regulatory sites of the InsP3R. Unfortunately, it was not possible to conduct these experiments using monovalent cations due to the presence of K channels in the microsomal membrane.

![Figure 4](image)

**Figure 4.** Test for the presence of the anomalous mole fraction effect. The dependence of the elementary current at 0 mV (●) and at -20 mV (■) through InsP3-gated channels with a varying Mg to Ba molar ratio on the trans side of the membrane. The points are the means ± SEM of the current values obtained in three experiments in which the activity of the same InsP3R was recorded at different Mg to Ba molar ratios (two experiments where the Mg mole fraction was changed from 0 to 0.9, and one experiment where it was changed from 1 to 0.1). Mg and Ba concentrations were estimated using Eq. 1. See text for details.

Representative current traces from this series of experiments are shown on Fig. 5 A (Sr is the current carrier). In these experiments, channels were first recorded in the absence of Ca on the intraluminal side of the membrane (Fig. 5 A, top trace, 55 mM Sr/HEPES and 1 mM EGTA in the trans chamber, pCa of 5.5 based on the level of contaminated Ca in the Sr/HEPES solution) and then increasing amounts of Ca/HEPES solution were added to the trans chamber. The total concentrations of Ca, Sr, and EGTA were calculated using Eq. 1 (see Methods) after each addition of Ca/HEPES. Based on these values, the free Ca concentration in the trans compartment was estimated as described in Methods. This protocol allowed us to measure the activity of the same channel at different intraluminal free Ca concentrations from the micromolar range to several millimolar Ca (Fig. 5 A). Some decrease in single-channel current amplitude occurring in these experiments (Fig. 5 A, compare last two traces) could be attributed to the fact that at high trans Ca levels a substantial portion of the trans Sr was replaced by Ca ions. Because the conductance of the channels for Ca is less than that for Sr (Figs. 2 and 3), some reduction in current amplitude is
expected. During data analysis adjustments for these changes were accomplished by lowering the current threshold level.

The general result of these experiments is that the activity of InsPs-gated channels was not potentiated by an increase in trans Ca levels in the expected physiological range of intraluminal free Ca concentrations, in disagreement with the previously proposed model (Irvine, 1990). As a matter of fact, InsPs-gated channels were slightly inhibited by an increase in trans Ca (Fig. 5 B). On average, an elevation in the trans Ca level to 1 mM reduced the single-channel open probability to 67 ± 11% (n = 3) of its control level (Fig. 5 B). Further increases in trans Ca caused additional decreases in the channel's activity. At 10 mM trans Ca, the single-channel open probability was equal to only 33 ± 5% (n = 3) of its control level (Fig. 5 B). It is notable that a similar result was previously obtained with another intracellular Ca release channel, RyR (Ma, Fill, Knudson, Campbell, and Coronado, 1988). In this section we focused on the effects of intraluminal Ca below 10 mM. A more detailed consideration of the inhibitory effects on InsPs-gated channels caused by high intraluminal Ca levels (> 10 mM) is presented in the next section of this paper.
Effect of High (≥ 10 mM) Intraluminal Ca on the Activity of InsP₃-gated Channels: a Kinetic Model of InsP₃R Gating

To get a better understanding of the mechanism underlying the inhibitory effects of high (≥ 10 mM) intraluminal Ca on the gating of the InsP₃R, we conducted a series of experiments where channel activity was initially recorded in the absence of trans Ca (1 mM EGTA added) using 55 mM Sr as a current carrier. Then, trans Ca concentration was increased by serial addition of a solution containing 55 mM Ca/HEPES. After each addition, an equal volume of trans solution was withdrawn. By using this protocol the activity of the same channel was measured at various Ca to Sr molar ratios while the total concentration of divalent cations in the trans chamber was equal to 55 mM. The concentrations of Ca and Sr in the trans chamber were estimated using Eq. 1 (see Methods).

By using this method we were able to measure the activity of the same channel at Ca molar fractions in the trans chamber ranging from 0 to 0.8 (see Fig. 6A). The single-channel open probability in this series of experiments decreased 5–10-fold by the increase in the Ca mole fraction. Single-channel data analysis revealed that high levels of intraluminal Ca inhibit InsP₃-gated channels by decreasing both the frequency of channel openings and the mean open time of the channels (Fig. 6B). For the experiment shown on Fig. 6A, the mean open time of the channels decreased from 4.73 to 2.86 ms when the Ca mole fraction in the trans chamber was increased from 0 to 0.8 (Fig. 6B, left), approaching the values typically obtained in experiments where only Ca was the current carrier (Fig. 2, Table I). In the same experiment the observed mean closed time of the channels was simultaneously increased from 65 to 320 ms (Fig. 6B, middle, two channels are active in the membrane). Single-channel current amplitude at 0 mV was only slightly affected in this experiment, changing from 1.65 pA to 1.49 pA (Fig. 6B, right).

How is it possible to explain the inhibitory effects of high trans Ca on InsP₃-gated channels? We speculated that channel modulation could be explained as a result of an increase in the local Ca concentration in the vicinity of the channel pore on the cytoplasmic side of the membrane at every channel opening (Neher, 1986; Stern, 1992). This idea was inspired by a similar model that was recently suggested for the skeletal muscle RyR based on Ca-release measurements (Jong, Pape, Chandler, and Baylor, 1993). Because InsP₃R are inactivated by cytoplasmic free Ca above 1 μM (Bezprozvanny et al., 1991a; Finch et al., 1991; Iino, 1990) this local elevation of Ca, that can reach 100 μM at macromolecular distances from the channel pore for 2 pA of Ca current (Stern, 1992), can inactivate the channels prematurely, which would then cause a decrease in the mean open time in the presence of high trans Ca. If it takes a long time for the bound Ca to dissociate from the inhibitory site when compared with the rate of channel openings, then a decrease in the frequency of channel openings would be also observed. Because Sr, Ba, or Mg in the cis solution at concentrations up to 1 mM did not inhibit the gating of the InsP₃R (data not shown), these effects would be only observed in the presence of trans Ca. Indeed, when InsP₃-gated channels were recorded at different Mg to Ba molar ratios (see above), channel gating was not affected (data not shown). Therefore, the effects on open-channel probability and the channel's kinetic characteristics observed here are
specific for Ca. A direct test of the proposed model would have been to use a high concentration of BAPTA in order to buffer Ca in the vicinity of the channel pore (Stern, 1992). Unfortunately, BAPTA acts as potent inhibitor of InsP$_3$ binding to its receptor (Richardson and Taylor, 1993), a result we confirmed by finding that

![Figure 6](image)

**Figure 6.** Effect of varying the trans Ca molar fraction on the gating of the InsP$_3$R. (A) InsP$_3$-gated channels were recorded at various Ca to Sr molar ratios in the trans chamber in the same experiment; the total concentration of divalent cations in the trans chamber was equal to 55 mM. The concentrations of Ca and Sr in the trans chamber were estimated using Eq. 1 (see Methods). The molar fraction of Ca in the trans chamber is indicated on the right side of each current record. (B) Open-time distribution (left), observed closed time distribution (center), and current amplitude histogram at 0 mV (right) are shown for the same InsP$_3$R in the absence of added Ca in the trans chamber (top) and at a trans Ca molar fraction of 0.8 (bottom). Data for single-channel analysis are taken from the experiment shown in A. Histograms were constructed from 1,462 events in the absence of added trans Ca and 624 events at a trans Ca molar fraction of 0.8. Open and closed time distributions were fitted with single exponential functions. Experiment 40110Sr4.

InsP$_3$-gated channels were inactivated in the presence of 10 mM BAPTA (data not shown).

A quantitative model developed based on the assumption that trans Ca modulated InsP$_3$R activity due to the increase in the local Ca concentration on the cytoplasmic side and the kinetic parameters extracted from a fit to the experimental data are presented in the Appendix.
Measurements of InsP$_3$-gated Channels in the Presence of Cytosolic Potassium

One goal of our experiments is to estimate the size of the unitary current through the InsP$_3$R under physiological conditions. In all experiments presented to this point, InsP$_3$-gated channels were measured in the presence of Tris-HEPES solution on the cis side of the membrane and divalent cations dissolved in HEPES on the trans side. It was necessary to use this ionic composition because cerebellar microsomes contain K and Cl channels, as well as InsP$_3$R, and therefore it was necessary to avoid K and Cl ions to measure solely the activity of the InsP$_3$-gated channel. However, during InsP$_3$-induced Ca release in the intact cell it is the selectivity of the channels for K ions, not for Tris, which contributes to the size of the Ca current. Therefore, it is important to measure the unitary current through InsP$_3$-gated channels in the presence of cytosolic K.

To perform these experiments, we decided to take advantage of the fact that K channels from the endoplasmic reticulum are inhibited by Ba. Unfortunately, the potency of Ba as an inhibitor of K channels from cerebellum is not as great as it is for K channels from skeletal muscle sarcoplasmic reticulum (Coronado, Rosenberg, and Miller, 1980). The inhibitory effect of Ba on the K channels from cerebellar microsomes is similar to its effect on the K channels from aortic sarcoplasmic reticulum (Bezprozvanny, Benevolensky, and Naumov, 1991b). However, the activity of K channels was greatly diminished by the presence of 55 mM Ba on the trans side, especially at negative potentials due to the voltage dependence of the Ba block (Bezprozvanny et al., 1991b).

A representative experiment from this series is shown in Fig. 7. In the first part of the experiment (Fig. 7 A) InsP$_3$-gated channels were recorded at 0 mV as usual, after addition of 2 μM InsP$_3$, with 55 mM Ba in the trans chamber as the current carrier, and the cis chamber filled with the Tris-HEPES solution supplemented with 0.2 μM free Ca and 0.5 mM ATP. The cis compartment was subsequently perfused with 20 vol of 110 mM K/HEPES solution (pH 7.35), a concentration of K chosen to emulate the K content in the cytoplasm. Change in the junction potential caused by the change in the ionic composition of cis solution was compensated electronically after perfusion (see Methods). After adjustment of the free Ca concentration in the cytosolic compartment to 0.2 μM and addition of 0.5 mM ATP and 2 μM InsP$_3$, InsP$_3$-gated channels were once again observed (Fig. 7 B, first pair of traces). Some K channel activity was not completely inhibited by Ba, but this did not create a problem in data analysis because these channels were easily distinguished from InsP$_3$-gated channels due to the marked differences in channel amplitude and kinetic characteristics (Fig. 7 B, first pair of traces). InsP$_3$-gated channels were also recognized by their sensitivity to heparin (Fig. 7 B, second pair of traces), whereas the activity of K channels was not affected by heparin (Fig. 7 B, second pair of traces).

It is clear from Fig. 7 that the unitary current through InsP$_3$-gated channels at 0 mV is substantially larger in the presence of 110 mM K on the cis side than in the presence of Tris in the same concentration. On average, the mean single-channel current amplitude in this series of experiments was equal to 3.0 ± 0.1 pA (n = 3) with 110 mM K in the cytoplasmic solution. This value is 1.36 times larger than the value obtained with Ba as a current carrier in the presence of Tris on the cytoplasmic side.
The relatively small increase (8%) in channel slope conductance from 85 ± 3 pS (n = 4) for Ba vs Tris (see Fig. 3 and Table I) to 92 ± 3 pS (n = 3) for Ba vs K (Fig. 8) cannot account for the 36% difference in the single-channel amplitude at 0 mV. It should be pointed out that the current-voltage relationship is significantly nonlinear at positive potentials in the presence of Tris (see above) and that only the linear portion of the curve is shown in Fig. 3. Therefore, it is not correct to estimate the Ba to Tris permeability ratio \( P_{Ba}/P_{Tris} \) by extrapolating the linear portion of the current-voltage relationship. Although it appears that the current-voltage relationship has shifted to the right with K, the mostly likely explanation is a decrease in the curvature of the current-voltage relationship in the presence of K, as it was reported in similar experiments for the RyR (Smith et al., 1988; Tinker and Williams, 1992). The Ba to K permeability ratio \( P_{Ba}/P_{K} \) of the InsP₃R could be estimated using Eq. 2 (see Methods) from the reversal potential of current through the channels under the biionic conditions used in this series of experiments. Unfortunately, K channels were no longer sufficiently inhibited by Ba at positive transmembrane potentials due to the presence of heparin.

**Figure 7.** InsP₃-gated channels recorded in the presence of 110 mM cis K. In the first part of the experiment (A), the cis chamber was filled with Tris-HEPES solution supplemented with 0.2 μM Ca²⁺ and 0.5 mM ATP; the trans chamber contained 55 mM Ba/HEPES. In the absence of InsP₃, channel activity was not observed (A, first pair of traces). Addition of 2 μM InsP₃ induced channel activity (A, second pair of traces). After channel activation, the cis compartment was perfused with 20 vol of 110 mM K/HEPES solution. After adjustment of the free Ca concentration to 0.2 μM, addition of 0.5 mM ATP and 2 μM InsP₃, InsP₃R were reactivated (B, first pair of traces). The single-channel current amplitude was 1.3 times larger under these conditions (Ba vs K) compared with the first part of the experiment (Ba vs Tris). A residual activity of K channels was sometimes observed (B, first pair of traces). Heparin inhibited InsP₃-gated channels (B, second pair of traces) but did not affect K channels (last trace). Current was recorded at 0 mV. Replacement of Tris-HEPES by K/HEPES caused a 2-mV change in the junction potential that was calibrated before the experiment and compensated electronically (see Methods). Experiment 30804BaK.
to the voltage-dependent nature of the inhibition (Bezprozvanny et al., 1991b). Therefore, the true value of the reversal potential for InsP₃-gated channels under biionic conditions (Ba vs K) could not be obtained. However, when the activity of the purified RyR was recorded in planar lipid bilayers under similar conditions (Smith et al., 1988; Tinker and Williams, 1992), the single-channel current–voltage relationship was composed of two linear segments with the intersection at the reversal potential. Because the permeation pathways of InsP₃R and RyR seem to be constructed in a similar way (see Discussion), we assumed that the current-voltage relationship of the InsP₃R under biionic conditions is also ohmic on both sides of the reversal potential. If this is the case, then a value for the reversal potential ($E_{\text{rev}}$) of +32 mV is obtained by extrapolating the current-voltage relationship until it intersects with the voltage axis (Fig. 8). From these data and the ionic conditions used in this series of experiments (55 mM Ba trans vs 110 mM K cis), $P_{\text{Ba}}/P_{\text{K}}$ of 6.3 is estimated from Eq. 2 (see Methods). With the RyR, a value for $P_{\text{Ba}}/P_{\text{K}}$ of 6 was obtained (Smith et al., 1988; Tinker and Williams, 1992). Therefore, most likely both types of intracellular Ca release channels have similar selectivity for divalent cations versus monovalent cations.

**The Dependence of the Elementary Current on the Permeant Ion Concentration**

In all experiments presented so far, the concentration of permeant ion in the trans chamber was fixed at 55 mM. However, it is generally believed that the free Ca concentration in the lumen of the reticulum does not exceed the level of several millimolar. To estimate the size of the current through InsP₃R in the physiological situation we measured the amplitude of the divalent current through InsP₃-gated channels as a function of the permeant ion concentration. Fig. 9A shows the elementary currents recorded at a transmembrane potential of 0 mV at different trans Ba concentrations, indicated on the right side of each record in mM. In this series of experiments InsP₃-gated channels were first recorded in the presence of 5 mM Ba/HEPES on the trans side and then subsequent additions of aliquots of a 400 mM Ba/HEPES stock solution were made to the trans chamber. The Ba concentration in...
the *trans* compartment after each addition was estimated as described in Methods. The *cis* chamber contained Tris-HEPES solution that was not changed during an experiment. Using this protocol we obtained recordings of the same channel at *trans* Ba concentrations ranging from 5 mM to 72 mM (Fig. 9A) or up to 160 mM (data not shown).

The mean values of the elementary current obtained in this series of experiments are plotted as a function of the *trans* Ba concentration in Fig. 9B (filled circles). The

![Figure 9](image)

**Figure 9.** Dependence of the single-channel current amplitude at 0 mV on the Ba concentration in the *trans* chamber. (A) InsP3-gated channels were recorded at different *trans* Ba concentrations from 5 to 72 mM in the same experiment. The *trans* Ba concentrations, calculated using Eq. 1 after each addition of Ba/HEPES (see text for details), are indicated in millimolar on the right side of each current record. The *cis* chamber contained the same Tris-HEPES solution throughout the experiment. Experiment 31231Bai. (B) The amplitude of elementary current through the InsP3R at 0 mV is plotted as a function of the *trans* Ba concentration for the experiments with Tris-HEPES in the *cis* side (●) and for the experiments with symmetrical 110 mM K/HEPES (○). The symbols represent the mean value ± SEM (*n* = 3). The smooth curve is the best fit to the equation \( i = i_{\text{max}} / (1 + K_d / [\text{Ba}]) \), where \( i \) is the elementary current, \( i_{\text{max}} \) is the saturating value of \( i \), [Ba] is the *trans* Ba concentration, and \( K_d \) is the apparent dissociation constant. The best fit to the data (smooth curves) was obtained with \( i_{\text{max}} = 2.8 \) pA and \( K_d = 10.3 \) mM for the experiments with Tris-HEPES in the *cis* chamber and with \( i_{\text{max}} = 3.4 \) pA and \( K_d = 9.0 \) mM for the experiments with symmetrical 110 mM K/HEPES.

data from these experiments could be fit using the Michaelis-Menten equation with an apparent dissociation constant for Ba of 10.0 ± 0.2 mM (*n* = 3) and a maximal unitary current of 2.8 ± 0.2 pA (*n* = 3). The value of the apparent dissociation constant for Ba is close to the value for the \( K_d \) obtained in a similar series of experiments for cardiac L-type Ca channels (Hess, Lansman, and Tsien, 1986). Because Ca ions go through these channels under physiological conditions, we also measured the unitary current as a function of the *trans* Ca concentration (data not
shown). Unfortunately, it was not possible to analyze the data at *trans* Ca concentrations below 20 mM because channel openings are shorter in duration and the unitary current amplitude is smaller with Ca as a current carrier compared with Ba (Fig. 2, Table I). Because the size of the single-channel current at 20 mM *trans* Ca was equal to 85% of its maximal value, a reliable estimate of the apparent dissociation constant for Ca could not be obtained.

It is notable that the increase in the size of the unitary current through the InsP$_3$R is mainly due to changes in the driving force for Ba ions. The single-channel conductance at 5 mM *trans* Ba was equal to 71 ± 4 pS (n = 3, data not shown), 90% of its value at 55 mM intraluminal Ba. This result is in agreement with measurements of the single-channel conductance of sheep cardiac RyR, where the channel conductance in symmetrical 2 mM Ba was equal to 75% of its maximal value (Tinker and Williams, 1992). Therefore, both intracellular Ca channels are designed to have near maximal conductance at millimolar concentrations of divalent cations. Because the free Ca concentration in the lumen of the endoplasmic/sarcoplasmic reticulum is unlikely to exceed several millimolar, this feature optimizes the ability of these channels to pass ions in the physiological situation.

In the next series of experiments we made one more step towards measurements using physiological conditions. The activity of InsP$_3$-gated channels was recorded at 0 mV at different *trans* Ba concentrations in the presence of 110 mM symmetrical K. Because there is no driving force for K ions at 0 mV in these conditions, the recorded current is carried by divalent (Ba) ions. In the beginning of each experiment in this series the *cis* chamber contained 110 mM K/HEPES, pH 7.35 supplemented with 0.2 µM Ca$^{2+}$ and 0.5 mM ATP; the *trans* chamber was filled with 5 mM Ba/HEPES and 110 mM K/HEPES, pH 7.35. After addition of 2 µM InsP$_3$ to the *cis* chamber Ba current through InsP$_3$-gated channels was recorded. The Ba concentration in the *trans* chamber was then increased by subsequent additions of aliquots of 200 mM Ba/HEPES stock solution dissolved in 110 mM K/HEPES, pH 7.35. The Ba concentration in the *trans* compartment after each addition was estimated as described in Methods.

Using this protocol we obtained recordings of the same channel at a transmembrane potential of 0 mV and at *trans* Ba concentrations ranging from 5 mM to 72 mM in the presence of 110 mM K on both sides of the membrane. Mean values of the single-channel current amplitude at every *trans* Ba concentration obtained in this series of experiments are plotted in Fig. 9 B (open circles). The data were fit using the Michaelis-Menten equation with an apparent dissociation constant for Ba equal to 9.5 ± 1.0 mM (n = 3) and a maximal unitary current equal to 3.5 ± 0.1 pA (n = 3) (smooth curve on Fig. 9 B). It is obvious from Fig. 9 B that the amplitude of the single-channel current at every *trans* Ba concentration in this series of experiments (Fig. 9 B, open circles) is 20–30% larger than the size of the current measured in the presence of Tris in the *cis* compartment in the previous series of experiments (Fig. 9 B, filled circles). These data are in agreement with the results obtained in the preceding section of the paper, where the activity of InsP$_3$-gated channels was measured in the presence of cytosolic K. Only measurements at 0 mV could be performed in the presence of symmetrical K because K channels were not sufficiently inhibited at low Ba concentrations.
DISCUSSION

This study focused on a characterization of the conduction properties of InsP$_3$-gated Ca channels from cerebellum. The main issues addressed are: (a) measurements of the single-channel conductance, the unitary current at 0 mV, and the mean open time of the InsP$_3$R for different alkaline earth cations; (b) a test for the presence of the anomalous mole fraction effect in a mixture of divalent cations; (c) an evaluation of the effects of intraluminal Ca concentration on activity and kinetic characteristics of InsP$_3$-gated channels; (d) a description of how the unitary current varies with changes in the permeant ion (Ba) concentration; (e) measurements of InsP$_3$-gated channel activity in the presence of K. In the following discussion the results obtained will be evaluated and an effort to uncover some general themes will be made based on comparisons of our data with information available for another type of intracellular Ca channel, the RyR, and for the L-type Ca channel, the best studied example of voltage-gated Ca channels of the plasma membrane.

Conduction Properties of InsP$_3$ Receptor for Divalent Cations

The presence of Cl and K channels in the cerebellar microsomes used in our experiments imposed limits on the ion composition of the solutions that could be used in this study. For this reason we could not obtain as detailed and precise information about InsP$_3$R selectivity and permeation as was recently done with another type of intracellular Ca channel, the RyR, purified from either cardiac or skeletal muscle (Lindsay et al., 1991; Lindsay and Williams, 1991; Smith et al., 1988; Tinker et al., 1992; Tinker and Williams, 1992). A detailed analysis of InsP$_3$R selectivity properties must be postponed until reliable recordings from purified InsP$_3$R incorporated in the bilayer are obtained. However, some generalizations about intracellular Ca channel selectivity and permeation properties can still be made based on the information obtained for InsP$_3$R in the present study.

All four alkaline earth cations tested are able to pass through the InsP$_3$R with single-channel conductances that fall in the sequence Ba > Sr > Ca > Mg (Figs. 2 and 3). The same order of conductances was reported for the RyR (Tinker and Williams, 1992) although the absolute values of the single-channel conductance are approximately twice as large for the RyR. Both channels have a similar selectivity for divalent cations: the $P_{\text{Ba}}/P_{\text{K}}$ permeability ratio is 6.3 for InsP$_3$R (see Results), whereas reported values of $P_{\text{Divalent}}/P_{\text{K}}$ permeability ratios for the RyR range between 4 and 8 (Lindsay and Williams, 1991; Liu et al., 1989; Smith et al., 1988; Tinker and Williams, 1992). When channel permeation was studied with mixtures of divalent cations, the anomalous mole fraction effect was not observed (Fig. 4 and Tinker and Williams, 1992). It is premature to make a final conclusion that InsP$_3$R and RyR are single-ion channels based solely on the absence of the anomalous mole fraction effect in our experiments and in the experiments of Tinker and Williams (1992) because in some recording conditions this effect is difficult to observe even for known multi-ion channels (Yue and Marban, 1990). Nevertheless, it seems reasonable to assume that both intracellular Ca channels act as single-ion pores until evidence for multiple occupancy are obtained. The similarity in the conduction properties of InsP$_3$R and RyR is not surprising because the transmembrane domains of the InsP$_3$R and the
RyR are 40% homologous and both channels are organized as tetramers (Anderson et al., 1989; Chadwick et al., 1990; Furuichi et al., 1989; Mignery, Sudhof, Takei, and De Camilli, 1989; Mignery et al., 1990; Otsu, Willard, Khanna, Zorzato, Green, and MacLennan, 1990; Takeshima, Nishimura, Matsumoto, Ishida, Kangawa, Minamino, Matsuo, Ueda, Hanaoka, Hirose, and Numa, 1989). Thus, it seems reasonable to assume that the permeation pathways of the InsP₃ and RyR are constructed in a similar manner.

It is notable that the order of conductances for divalent cations, selectivity against monovalent cations, and the mechanism of ion permeation for both intracellular Ca channels differ dramatically from those obtained in studies of L-type voltage-gated Ca channels of the plasma membrane. L-type Ca channels are extremely selective against monovalent cations \( \frac{P_{\text{Divalent}}}{P_{\text{Monovalent}}} > 1,000 \) [Lee and Tsien, 1982, 1984]. To describe the function of the L-type channels a two-site model was proposed (Almers, McCleskey, and Palade, 1984; Hess and Tsien, 1984; Tsien et al., 1987) which explains Ca permeation through these channels as a result of electrical repulsion between Ca ions bound in the two intrapore binding sites. The strongest experimental evidence in support of this model is that with mixtures of cations the anomalous mole fraction effect is observed with the L-type Ca channel (Almers et al., 1984; Hess and Tsien, 1984). The sequence Ba > Ca = Sr >> Mg was reported for cardiac L-type channels (Hess et al., 1986) with Mg being practically impermeable. The ability of Mg ions to carry substantial currents through the RyR (Ashley, 1989; Smith et al., 1986, 1988; Tinker and Williams, 1992) and the InsP₃R (Fig. 2 and 3, Table I) is especially striking when the very high hydration energy and extremely slow substitution rate of water molecules in the inner hydration shell of Mg ions (Hille, 1992) is taken into consideration. One possible explanation of this observation is that when Mg ions pass through the selectivity filters of both intracellular Ca channels they are able to keep their inner shell of water molecules. This suggestion implies that the narrowest portion of the channel pore is at least 10 Å² for both intracellular channels. An even larger estimate of the pore size (40 Å²) was obtained for the RyR (Lindsay et al., 1991) based on the ability of large organic cations like Tris and TEA to permeate through these channels. The idea of a fairly wide pore fits with the observed order of conductances of these channels for divalent cations which corresponds to the order of mobilities of these ions in water.

From the data presented in this report and the studies of RyR permeation, both InsP₃R and RyR act as rather nonspecific cation-selective channels, permeable to Ca, K, and Na. However, under physiological conditions, K and Na are equally distributed across the uncharged membrane of sarcoplasmic reticulum (Somlyo, Shuman, and Somlyo, 1977). Most likely there is no gradient for K and Na across the membrane of endoplasmic reticulum as well. Therefore, the main current through RyR and InsP₃R is carried by Ca due to the large electrochemical gradient for Ca across the reticular membrane. The situation is quite different for voltage-gated Ca channels in the plasma membrane. In the operational voltage range of plasma membrane Ca channels, there is a large driving force for Na ions to enter the cell and for K ions to leave the cell due to concentration and electrical gradients across the plasma membrane. Therefore, voltage-gated Ca channels (for example, L-type) must
discriminate very well against Na and K ions in order to avoid large fluxes of monovalent cations at every opening of a Ca channel.

Are InsP$_3$ Receptors Potentiated by Submillimolar Intraluminal Ca?

Addition of submaximal InsP$_3$ concentrations causes partial discharge of Ca from the intracellular stores. This phenomenon is often called "quantal release" (Muallem, Pandol, and Beeker, 1989). Irvine (1990) suggested that regulation of the InsP$_3$R's sensitivity to InsP$_3$ by intraluminal Ca concentration may be responsible for quantal release. In support of this hypothesis, a decrease in the rate of InsP$_3$-induced Ca release upon store depletion was demonstrated using permeabilized rat hepatocytes (Nunn and Taylor, 1992) and smooth muscle cells (Missiaen et al., 1992a,b). In contrast, no changes in the InsP$_3$R sensitivity due to the depletion of Ca stores were observed by two other laboratories in experiments with either permeabilized rat hepatocytes (Combettes et al., 1992) or cells from the avian supraorbital nasal gland (Shuttleworth, 1992). The evaluation of these data is difficult, because the absolute levels of intraluminal free Ca concentration and their changes during the experiment are not measurable and can not be controlled. It is quite possible that different levels of intraluminal Ca concentrations were reached by different laboratories during the depletion process. Therefore, the experiments are difficult to compare and the data obtained are controversial. Hence, it is of interest to see whether the regulation of InsP$_3$R by intraluminal Ca would be evident in planar lipid bilayer experiments, a situation in which the free Ca concentration on both cytoplasmic and intraluminal sides of the membrane can be easily manipulated.

In our experiments using either Sr, Ba or Mg as a current carrier, potentiation of InsP$_3$-gated channel activity by intraluminal Ca was not observed (Fig. 5). In fact, some inhibitory effect of intraluminal free Ca in the millimolar range on InsP$_3$R activity was evident in our experiments (Fig. 5). This inhibitory action of elevated intraluminal Ca ($\geq 1$ mM; Fig. 5) is very similar to the inhibition of skeletal muscle RyR by intraluminal Ca (Ma et al., 1988). It seems that both intracellular Ca release channels are less likely to open when intraluminal Ca content is very high. Interestingly, a model with inhibition of InsP$_3$R by high intraluminal Ca levels was recently proposed as a mechanism of Ca oscillations in gonadotrophs (Stojilkovic, Kukuljan, Tomic, Rojas, and Catt, 1993). Thus, the inhibitory effect of intraluminal Ca on InsP$_3$R observed in our reconstitution experiments may be physiologically significant.

It follows from our experiments (Fig. 5) that the activity of the InsP$_3$R is not potentiated by intraluminal Ca. If this is the case, then how can the results of Missiaen et al. (1992a, b) obtained using permeabilized cells be explained? One possible explanation is that when intracellular stores are depleted in the experiments with permeabilized cells and the free intraluminal Ca concentration falls, the unitary Ca current through InsP$_3$R decreases below its normal value (Fig. 9) and the rate of InsP$_3$-induced Ca flux is diminished. Because of the high value for the apparent dissociation constant of the divalent cation binding site in the channel pore, equal to 10 mM for Ba (Fig. 9B), one can conclude that in the physiological range of intraluminal free Ca concentrations the elementary current through InsP$_3$-gated channels is a linear function of the free Ca concentration in the lumen of the reticulum. Therefore, the observed decrease in the rate of InsP$_3$-induced Ca release
upon store depletion could be caused not by inhibition of InsP3R but rather, by a decrease in the size of the unitary Ca current through InsP3R due to a diminished driving force for Ca ions across the reticular membrane. The best way to describe the ability of the InsP3R to release Ca as a function of the intraluminal Ca will be to calculate the product of the curves shown on Figs. 5 B and 9 B. When this is done, an “optimal” intraluminal Ca concentration of ~ 10 mM corresponding to the maximal rate of InsP3-induced Ca release is expected. Unfortunately, the free intraluminal Ca concentration is not known precisely and it is difficult to estimate changes in the degree of store depletion in the experiments with permeabilized cells. The lack of information regarding the intraluminal Ca concentration makes it difficult to predict InsP3R behavior in physiological experiments.

Although this explanation seems to be very plausible, other potential explanations for the observed discrepancy between our data and the results reported using permeabilized cells (Missiaen et al., 1992a, b; Nunn and Taylor, 1992) may include:

(a) There is a hypothetical “intraluminal regulatory site” that will bind other divalent cations. Because 55 mM Ba, Sr, or Mg must be present in the intraluminal compartment in order to measure channel currents in planar lipid bilayer experiments, the InsP3R could be maximally activated at any intraluminal Ca concentration due to the binding of other divalent cation to this site.

(b) Some accessory protein that is necessary for regulation of InsP3R by intraluminal Ca was lost at some point during the isolation and reconstitution of the InsP3R in the bilayer.

(c) Different isoforms of InsP3R may vary in their regulation by intraluminal Ca. Unfortunately, functional expression of different isoforms of the InsP3R has not been achieved to date and the distinctive functional characteristics of the InsP3R isoforms are not yet known. Thus, it is not possible to precisely identify the isoform used in our experiments. It is likely that cultured smooth muscle cells and hepatocytes may primarily express isoforms of the InsP3R that differ from the type I InsP3R which is the most abundant isoform found in cerebellum, the tissue used in our study.

It seems that additional experiments performed using a variety of methods will be necessary in order to clarify the issue of InsP3R regulation by intraluminal Ca. At this point, however, the mechanism for potentiation of InsP3R activity by increased intraluminal Ca remains controversial.

**Kinetic Model of InsP3 Receptor Gating**

When InsP3-gated channels were recorded using 55 mM Ca on the trans side as a current carrier, the mean open time of the channels was significantly shorter than with other divalent cations (Fig. 2, Table I). High Ca concentration on the trans side inhibited the activity of InsP3R by decreasing both the frequency of channel opening and the mean open time of the channels (Fig. 6). It is possible that these effects of high trans Ca concentration on InsP3R gating indicate the presence of an intraluminal inhibitory Ca binding site of InsP3R complex. However, we suggested an alternative explanation to the observed phenomenon (see Results and Appendix). We speculated that channel modulation by intraluminal Ca could be explained as a result of an increase in the local Ca concentration in the vicinity of the channel pore on the cytoplasmic side of the membrane every time a channel opens and passes Ca current
The local elevation of Ca on the cytoplasmic side of the membrane near the InsP$_3$R causes channel inactivation due to the bell-shaped dependence of InsP$_3$R activity on the free Ca level in the cytosol (Bezprozvanny et al., 1991a; Finch et al., 1991; Iino, 1990). An advantage of this model is that it is based on the known properties of InsP$_3$R and it is not necessary to introduce any new functional characteristics of InsP$_3$R, such as a hypothetical intraluminal Ca binding inhibitory site. Notably, a similar model was suggested recently for the skeletal muscle RyR based on Ca-release measurements (Jong et al., 1993).

Based on this hypothesis, we were able to develop the model of InsP$_3$R gating that was in quantitative agreement with the observed effects of intraluminal Ca on InsP$_3$R functional properties (see Results and Appendix). The four state model of InsP$_3$R gating that was used in our calculations is shown in Fig. 10. Although this model is simplified, it adequately reflects the essence of InsP$_3$R regulation by cytosolic Ca (Bezprozvanny et al., 1991a; Finch et al., 1991; Iino, 1990). Inactivation of an open channel by an increase in the local Ca concentration in the vicinity of an open-channel pore is included in the model by introducing the direct transition from $R^*C^+$ (open) state to $RC^+$ (closed and inactivated) state with a rate constant proportional to the local Ca concentration. The following estimates for the kinetic parameters of InsP$_3$R gating were obtained from the fit to the data (see Appendix):

$$\alpha = 224 \text{ s}^{-1}; \beta = 25 \text{ s}^{-1}; k_2 = 0.8 \text{ s}^{-1}; k_3 = 4 \mu\text{M}^{-1}\text{s}^{-1}; k_* = 183 \text{ s}^{-1} \quad (3)$$

Thus, not only the effects of intraluminal Ca on InsP$_3$R function can be explained by immediate negative feedback of elevated cytosolic Ca in the vicinity of a channel pore, but this hypothesis also allowed us to estimate the kinetic parameters of InsP$_3$R gating. Some predictions about the expected functional properties of the InsP$_3$R can be made based on the model of InsP$_3$R gating shown in Fig. 10 with the values of the rate constants from Eq. 3.
It is expected from the model of InsP₃R gating shown in Fig. 10, that if cytosolic Ca concentration is suddenly elevated to a value equal to \( c \) (\( c \gg k_2, k_3 \)), then the InsP₃R will be converted into the inactivated state \( RC^+C^- \) with a rate constant approximately equal to \( c k_3/(1 + f_3/e_0) \) (binding of Ca to the activating site is assumed to be much faster than to the inhibitory site). The time course of Ca-induced inactivation of the InsP₃R was studied in experiments with brain microsomes using a rapid superfusion system (Finch et al., 1991) and with permeabilized smooth muscle cells using caged Ca (Iino and Endo, 1992). In the experiments with brain microsomes (Finch et al., 1991) a conditioning pulse of 10 \( \mu \)M Ca²⁺ was given for varying durations and the microsomes were challenged with 1 \( \mu \)M InsP₃. The characteristic time for InsP₃R inactivation by Ca under these conditions was 580 ms (Finch et al., 1991). In the experiments with permeabilized smooth muscle cells (Iino and Endo, 1992) the cytosolic Ca concentration was suddenly elevated by flash photolysis of DM-nitrophen from ~270 nM to ~850 nM in the presence of 3 \( \mu \)M InsP₃ and immediate slowing of the rate of InsP₃-induced Ca release was observed. Unfortunately, it is not possible to estimate exactly the time constant of InsP₃R inactivation from the data of Iino and Endo (1992). Nonetheless, it is obvious that divergent values for the rate of InsP₃R inactivation by Ca have been obtained using different methods. Based on the rate constants from Eq. 3, the characteristic time of InsP₃R conversion to the inactivated state estimated for experimental conditions similar to those used with the brain microsomes should be 25 ms, and for those used with permeabilized smooth muscle cells should be 320 ms. Additional measurements of the rate of Ca-induced inactivation of the InsP₃R as a function of the Ca concentration with high temporal resolution are necessary to test the model.

The value of \( k_2 \) equal to only 0.8 s⁻¹ means that recovery of the InsP₃R from Ca-induced inactivation should take on average 1.25 s. Moreover, if the presence of several inhibitory sites in the InsP₃R complex is taken into consideration and it is assumed that binding of a Ca ion to at least one of these sites is sufficient to prevent channel opening, then this time will be even longer. For example, it is easily shown that for four inhibitory sites the average time of recovery is 2.6 s. Indeed, when the time course of InsP₃R recovery from Ca-induced inactivation was studied in Xenopus oocytes (Parker and Ivorra, 1990) or in brain synaptosomes (Finch et al., 1991), a characteristic time of recovery in the range of several seconds was measured, in agreement with our estimations.

The open-channel probability of the InsP₃R in planar lipid bilayer experiments is in the range 5–12% under optimal conditions (Fig. 1; Bezprozvanny and Ehrlich, 1993a; Bezprozvanny et al., 1991a; Watras et al., 1991). This number has to be viewed as an upper estimate because of the uncertainty about the number of active channels in the bilayer. It follows that InsP₃ is a partial agonist of the InsP₃R which means that InsP₃ has a low intrinsic efficacy (Bezprozvanny and Ehrlich, 1993a; Pallotta, 1991). One possible explanation for this fact comes from the model of InsP₃R gating shown in Fig. 10. Indeed, even at an optimal cytosolic Ca concentration, the InsP₃R spends just a fraction of its time in the \( RC^+ \) state, the only state from which the channel can open. Therefore, the intrinsic efficacy of InsP₃ is decreased. The intrinsic efficacy of InsP₃ is also affected by ATP (Bezprozvanny and Ehrlich, 1993a), an allosteric regulator of the InsP₃R. Although a high concentration of InsP₃
(2 μM) was used in our experiments, this concentration may not suffice to saturate InsP₃ binding sites completely (Bezprozvanny, Bezprozvannaya, and Ehrlich, 1994; Watras et al., 1991). In our model (Fig. 10) the effects of submaximal ATP and InsP₃ concentrations on channel gating are built into the rate constants α and β. All these factors influence the open probability of InsP₃R observed in our experiments. From the model of InsP₃R gating considered in this report (Fig. 10) a maximal open channel probability of only 3.6% (at 0.2 μM Ca²⁺) is calculated based on the rate constant estimations from Eq. 3, in agreement with the observed values.

The mean open time of the InsP₃R under physiological conditions can be estimated from the model shown in Fig. 10. The rate constant of the direct transition from R*C⁺ to RC⁺C⁻ (Fig. 10) is proportional to the local Ca concentration in the vicinity of a channel pore [Ca]⁺; the local Ca concentration in the vicinity of the pore is, in turn, proportional to the size of Ca current through InsP₃R (Eqs. A1 and A2). The size of the unitary Ca current through InsP₃R in the physiological situation is estimated to be ~ 0.5 pA (see below), four times less (x = 0.25) than the size of the single-channel current in the series of experiments used to estimate InsP₃R kinetic parameters (Fig. 6 B). According to the model shown in Fig. 10 the mean open time of the InsP₃R under physiological conditions is (Eq. A11) \( \tau_0 = \frac{1}{\alpha + \beta k_{-}} = 3.7 \text{ ms} \), if numerical values for α and \( k_{-} \) are taken from Eq. 3. Note that only one out of five channel closings occurs by direct transition from the open to the inactivated state because \( x k_{-} / (\alpha + x k_{-}) \approx 0.2 \) under physiological conditions. Thus, in the first approximation, the simplified model of channel gating that does not take this direct transition into account can be used for the analysis of InsP₃R behavior under physiological conditions as it was recently done in a paper devoted to the theoretical analysis of Ca wave propagation (Bezprozvanny, 1994).

**Physiological Size of Ca Current through Ins₃ Receptors**

To estimate the size of the unitary current through the RyR under physiological conditions, Tinker et al. (Tinker et al., 1992, 1993) developed a quantitative model for ion conduction through the RyR with the use of Eyring rate theory (Hille, 1992). This model allowed them to estimate the size of the unitary Ca current through the RyR to be ~ 2 pA at a transmembrane potential of 0 mV under physiological conditions (Tinker et al., 1992, 1993). Unfortunately, we were more limited in our study of InsP₃R permeation due to the presence of K and Cl channels in the membrane. Because we did not have the luxury of being able to create a mathematical model of the energetic profile of InsP₃R permeation pathway, we had to resort to approximations of the unitary Ca current through the InsP₃R with the ionic conditions expected in the physiological situation. Although the intraluminal free Ca concentration in the endoplasmic and sarcoplasmic reticulum is not precisely known, it is believed to be somewhere in the millimolar range. To compare InsP₃R and RyR properties we will estimate the size of the unitary Ca current through InsP₃R assuming that the intraluminal free Ca concentration is 2.5 mM and that 110 mM K is present on both sides of the reticular membrane; these are the same conditions used in the calculations of the Ca current through the RyR (Tinker et al., 1992, 1993). Because K, Na, and Cl ions are evenly distributed across the membrane of sarcoplasmic reticulum (Somlyo et al., 1977) there is no voltage across this mem-
brane. Most likely endoplasmic reticulum membrane is also uncharged and there-
fore, it is assumed in the following estimates that the transmembrane voltage is 0 mV.

In our experiments, the amplitude of the unitary current through InsPs-gated
channels was measured at a transmembrane potential of 0 mV and at different
intraluminal Ba concentrations (5–72 mM) in the presence of 110 mM symmetrical K
(Fig. 9 B, open circles). Because there was no driving force for K ions at 0 mV under
these conditions, the recorded current was carried by divalent (Ba) ions. The data
were fit using the Michaelis-Menten equation with an apparent dissociation constant
for Ba of 9.0 mM and a maximal unitary current of 3.4 pA (Fig. 9 B, smooth curve). If
these parameter values are used, then the unitary Ba current through InsPsR at 2.5
mM intraluminal Ba and 110 mM symmetrical K is equal to 0.74 pA.

In the series of experiments discussed above, Ba was the current carrier. However,
Ca ions go through InsPsR in cells. When InsPs-gated channels were recorded under
standard conditions, that is, in the presence of 55 mM divalent cation in the trans
chamber and Tris-HEPES solution in the cis chamber, the size of the Ca unitary
current was equal to 64% of the unitary Ba current. After introducing this correction,
the size of the unitary Ca current through the InsPsR at a transmembrane potential
of 0 mV under physiological conditions (2.5 mM intraluminal free Ca; 110 mM
symmetrical K) is equal to 0.47 pA, fourfold less than the Ca current through the
cardiac RyR under exactly the same conditions (Tinker et al., 1992, 1993). As it was
pointed out already, because of the high apparent $k_d$ for divalent cations, the
elementary Ca current through the InsPsR should be a linear function of the
intraluminal free Ca concentration in the physiological range. Of course, the
numbers obtained must be viewed as estimations until detailed studies of permeation
through the purified InsPsR for monovalent and divalent cations are conducted, the
intraluminal free Ca concentration is known, and the voltage across the membrane of
endoplasmic reticulum is measured.

How many Ca ions are released into the cytosol every time an InsPsR opens? The
mean open time of the InsPs-gated channels under physiological conditions was
estimated to be 3.7 ms (see preceding section). Taking this number and the size of
the Ca current of 0.47 pA estimated above, a value of 5,400 Ca ions/channel opening
is obtained. This number and estimates of InsPsR density should be used when
modeling spatiotemporal aspects of Ca signaling. It is notable that not only is the size
of the Ca current through the RyR fourfold higher, but the mean open time of RyR
type channels measured under similar conditions is ~20 ms (Smith et al., 1986), five
times longer than for the InsPsR. As a result, at every channel opening ~20 times
more Ca ions are released through the RyR than through the InsPsR. One can
speculate that the coexistence of two intracellular Ca channel types activated via
different mechanisms allows the cell to choose between two modes of Ca release: rapid dumping of accumulated Ca through the RyR channel or slow leakage through
the InsPsR. This suggestion is also supported by the observation that the RyR can be
activated to an open probability of virtually 100% (Smith et al., 1986), whereas the
InsPsR has never been observed with an open probability greater than 10% (Fig. 1;
Bezprozvanny and Ehrlich, 1993a; Bezprozvanny et al., 1991a; Watras et al., 1991).
In specialized cells such as cardiac and skeletal muscle cells that need very rapid Ca
release to initiate contraction in milliseconds, InsPsR are essentially absent, whereas
RyR are abundant (Anderson et al., 1989; Imagawa, Smith, Coronado, and Campbell, 1987; Lai et al., 1988). In contrast, smooth muscle cells, which control the steady level of tension by changing the rate of Ca efflux into the cytoplasm in minutes, employ predominantly InsP₃R for Ca transport (Chadwick et al., 1990; Walker, Somlyo, Goldman, Somlyo, and Trentham, 1987; Watras and Benevolensky, 1987). It is interesting to note that both receptor types are found in the brain (Bezprozvanny et al., 1991a; Ellisman, Deernick, Ouyang, Beck, Tanksley, Walton, Airey, and Sutko, 1990; McPherson, Kim, Valdivia, Knudson, Takekura, Franzini-Armstrong, Coronado, and Campbell, 1991; Ross, Meldolesi, Milner, Satoh, Supattapone, and Snyder, 1989; Supattapone et al., 1988b), suggesting additional complexity in neuronal Ca signaling.

**APPENDIX**

**Quantitative Model of InsP₃R Modulation by Intraluminal Ca**

In the results section the inhibitory effects of high intraluminal Ca levels on InsP₃R gating are described. A model was suggested explaining this effect as a result of an increase in the local Ca concentration in the vicinity of the pore on the cytoplasmic side of the membrane. In this Appendix, the predictions of the model are compared with the experimental data and kinetic parameters of InsP₃R gating are extracted from fits of the model to the data. In the vicinity of an open channel the free Ca concentration on the cytoplasmic side ([Ca]*) is equal to (Stern, 1992; Jong et al., 1993; Neher, 1986):

\[
[Ca]^* = \phi/2\pi Dr + [Ca]
\]  

(A1)

where \(\phi\) is the flux of Ca ions through the open channel, \(D\) is the diffusion constant for Ca ions in the \(cis\) solution, \(r\) is the distance from the mouth of a channel, and \([Ca]\) is the free Ca concentration in the \(cis\) chamber which is equal to 0.2 \(\mu M\) in our experiments. The value of \([Ca]^*\) is not affected by the presence of 1 mM EGTA in the \(cis\) chamber (Stern, 1992). The increase of Ca level near the mouth of channel to \([Ca]^*\) happens within a few microseconds after opening of a channel (Jong et al., 1993).

The flux of Ca ions through the open channel is equal to:

\[
\phi = i_{Ca}/2F
\]  

(A2)

where \(i_{Ca}\) is the Ca current through the channel and \(F\) is the Faraday constant. Let the parameter \(x\) be equal to the Ca molar fraction in the \(trans\) chamber:

\[
x = [Ca]_{trans}/([Ca]_{trans} + [Sr]_{trans}).
\]  

(A3)

Because InsP₃-gated channels seem to function as single ion pores, the affinity of the binding site for different divalent cations are similar (see discussion of Fig. 4), and the value of the single-channel current in our experiments did not change significantly with an increase in Ca \(trans\) molar fraction (Fig. 6 B, right):

\[
i_{Ca} = xi
\]  

(A4)
where \( i \) is the observed value of single-channel current through the InsP3R. From Eqs. A1, A2, and A4:

\[
[Ca]^* = \frac{i}{4\pi F D r} + [Ca]. \tag{A5}
\]

At macromolecular distances (\( r = 20 \) nm) for 2 pA of current (typical value in our experiments):

\[
i/4\pi F D r = 100 \mu M \gg 0.2 \mu M = [Ca]. \tag{A6}
\]

Therefore, in the first approximation:

\[
[Ca]^* = xc \tag{A7}
\]

\[
c = \frac{i}{4\pi F D r}
\]

where parameter \( c \) does not change with an increase in the \textit{trans} Ca mole fraction in our experiments. The conclusion from Eq. A7 is that in our experiments the free Ca concentration in the vicinity of an open channel on the \textit{cis} side is directly proportional to the Ca mole fraction on the \textit{trans} side.

In modeling InsP3R regulation by Ca, it is necessary to consider at least four distinct kinetic states of the InsP3R because of the bell-shaped Ca dependence of the channels (Bezprozvanny et al., 1991a; Finch et al., 1991; Iino, 1990). A simplified model of InsP3 channel regulation by Ca with one activating and one inhibitory site is (Bezprozvanny, 1994):

\[
R*C_+ \hspace{1cm} \beta \parallel \alpha
\]

\[
R \xrightarrow{k_i[Ca]} k_1[Ca] \xrightarrow{k_2} RC^+ \xrightarrow{k_3} RC^+C_-
\]

where \( R \) is the InsP3R, \( RC^+ \) is the InsP3R with Ca bound at the activating site, \( RC^+C_- \) is the InsP3R with Ca bound at both activating and inhibitory sites and \( R^*C^+ \) is the only open state of the channel. The main focus of this model presented here is to describe the regulation of InsP3R by Ca. Because our experiments were conducted in the presence of high concentrations of InsP3 (2 \( \mu M \)) and ATP (0.5 mM) it is assumed that both agonists (Bezprozvanny and Ehrlich, 1993a; Watras et al., 1991) are bound to the receptor in all four states of the model.

Inactivation of an open channel by an increase in the local Ca concentration in the vicinity of a channel pore is included in the model of channel gating by introducing the direct transition from the \( R^*C^+ \) state to the \( RC^+C_- \) state with the rate constant proportional to \([Ca]^*\):

\[
R^*C^+ \hspace{1cm} \beta \parallel \alpha \xrightarrow{k_i[Ca]^*} k_1[Ca]^* \xrightarrow{k_2} RC^+ \xrightarrow{k_3} RC^+C_-
\]

It is assumed that a channel in the \( RC^+C_- \) state can not open until Ca is dissociated.
from inhibitory site; that is, the transition from $R^{*}C^{+}$ to $RC^{+}C^{−}$ is irreversible. The proposed mechanism of InsP$_3$-gated channel inactivation by an increase in the local Ca concentration is identical to the mechanism suggested previously for the skeletal muscle RyR (Jong et al., 1993).

From Eq. A7:

$$k_1[Ca] = k_1x_C = x_k.$$ 

(A10)

$$\beta = k_1.$$ 

Therefore, the rate of transition from $R^{*}C^{+}$ to $RC^{+}C^{−}$ is directly proportional to the Ca mole fraction on the trans side in our experiments. This result is valid only if intraluminal Ca does not have other effects on InsP$_3$R gating besides those caused by alterations in the local cytosolic Ca concentration.

Using the method of Colquhoun and Hawkes (Colquhoun and Hawkes, 1982) it can be shown that for the model (Eq. A9) the mean open time of the channels and the observed mean closed time of the channels (the mean of all shut times) are equal to (taking Eq. A10 into consideration):

$$\tau_o = 1/(\alpha + x_k)$$

(A11)

$$\tau_c = (1 + k_3[Ca]/k_2 + k_5/k_4[Ca])\beta + x_k/[\alpha + x_k]k_2$$

(A12)

Let us introduce the parameter $\beta'$:

$$\beta' = \beta/(1 + k_3[Ca]/k_2 + k_5/k_4[Ca])$$

(A13)

Then, Eq. A12 becomes:

$$\tau_c = 1/\beta' + x_k/[\alpha + x_k]k_2$$

(A14)

As a result, both the mean open time of the channels (Eq. A11) and the observed mean closed time of the channels (Eq. A14) are now expressed as simple functions of the Ca mole fraction on the trans side ($x$). From Eq. A11, the mean rate of channel closing should be a linear function of Ca trans mole fraction:

$$\text{mean rate of channel closing} = 1/\tau_o = \alpha + x_k$$

(A15)

A reasonable fit to the experimental data (Fig. 11A) is provided by Eq. A15 assuming $\alpha = 224$ s$^{-1}$, $k_5 = 183$ s$^{-1}$.

The mean duration of channel shut times as a function of the Ca trans mole fraction was measured in the same experiment. According to the model (Eq. A9) the distribution of all shut times must be fit by the sum of three exponential functions. Therefore, it is not correct to use a single exponential fit to the distribution of the observed closed times in this context. We estimated the mean shut time by calculating the ratio of total shut time of the channels and the number of openings detected in the same segment of data. The value obtained was multiplied by the number of channels active in the membrane (equal to 2 in the experiment under consideration) in order to obtain a value for the mean shut time of a single channel. Because the single-channel open probability was low (<10%) in our experiments, simultaneous openings of several channels were very infrequent and only a small error was introduced by applying this procedure to multichannel recordings. Eq. A14 provides
an adequate fit to the data (Fig. 11 B) if $\beta' = 8.4 \text{ s}^{-1}$, $k_2 = 0.8 \text{ s}^{-1}$ and the parameters $\alpha$ and $k_4$ are taken from the fit to the mean open time (Fig. 11 A).

From the bell-shaped Ca dependence of InsP$_3$-gated channels, we previously estimated (Bezprozvanny et al., 1991a) that

$$k_5/k_4 = k_2/k_3 = k_3 = 0.2 \mu\text{M} = [\text{Ca}]$$

(A16)

where $k_2^+$ and $k_3^-$ are the affinities of the activating and inhibitory sites for Ca, respectively.

From Eqs. A16 and A13 and the estimation of $\beta' = 8.4$, s$^{-1}$ obtained above:

$$\beta = (1 + k_5[\text{Ca}]/k_2 + k_3/k_4[\text{Ca}])\beta' = 3\beta' = 25.2 \text{ s}^{-1}$$

(A17)

From Eq. A16 and the estimation of $k_2 = 0.8 \text{ s}^{-1}$,

$$k_3 = k_2/k_3^- = 4 \mu\text{M}^{-1} \text{s}^{-1}$$

(A18)

It seems reasonable to assume that $k_3 = k_1$, because both constants represent the rates of Ca binding to the same inhibitory site of the InsP$_3$R (assuming that the channel closes immediately after Ca is bound in the inhibitory site). If this is true, then for maximal $i_{\text{Ca}}$ current ($x = 1$; $i_{\text{Ca}} = i = 1.5 \text{ pA}$) from our estimation of $k_4 = 183 \text{ s}^{-1}$:

$$[\text{Ca}]^* = k_4/k_1 = 46 \mu\text{M}.$$  

(A19)

This value is in the range of local Ca concentrations expected for the macromo-
lecular distances from the mouth of an open channel passing 1.5 pA Ca current (Stern, 1992). Therefore, the model of channel inactivation by an increase in local Ca concentration described above makes physical sense.

Because the activity of the InsP$_3$R in our experiments was measured at optimal cytoplasmic Ca levels (0.2 μM Ca), additional increases in local Ca levels could only inhibit InsP$_3$R channel activity. Therefore, Ca-induced activation of the InsP$_3$R (Bezprozvanny et al., 1991a; Finch et al., 1991; Iino, 1990) was not a factor in the experiments testing the effects of intraluminal Ca on channel gating and no estimations of $k_4$ and $k_5$ can be obtained from the experimental data in this paper.

Finally, the following kinetic diagram of InsP$_3$R gating that is in agreement with our data can be drawn using the values of the rate constants obtained above assuming [Ca] = 0.2 μM, as it was in our experiments:

$$\begin{align*}
R^*C^+ & \xrightarrow{25.2 \text{ s}^{-1}} 224 \xrightarrow{185x} R C^+ \\
R C^+ & \xrightarrow{k_4 \text{Ca} \text{M}^{-1} \text{s}^{-1}} \xrightarrow{0.8} R C^+ C^- \\
R C^+ & \xrightarrow{k_5 \text{ s}^{-1}} R C^+ C^-
\end{align*}$$

(A20)

The estimates obtained for the rate constants are based on the assumption that the only mechanism responsible for the effects of trans Ca on the InsP$_3$R gating is an immediate negative feedback caused by an increase in the local Ca concentration in the vicinity of a channel pore. However, additional effects of intraluminal Ca on the gating of the channels can not be completely ruled out. It is necessary to note that several inhibitory and activating Ca binding sites are actually present in the channel complex (Bezprozvanny et al., 1991a). Therefore, a more complicated kinetic model may need to be considered for an exact determination of the rate constants. It was also assumed in our calculations that InsP$_3$ and ATP were bound to the receptor continuously, which probably is an oversimplification. However, the simplified model presented above adequately reflects the essence of InsP$_3$R regulation by cytoplasmic Ca and can explain the effects of intraluminal Ca on channel gating observed in our experiments.

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