A tick bite patient with fever and meningitis co-infected with *Rickettsia raoultii* and Tacheng tick virus 1: a case report

Yu Zhang1,2,3†, Liang Jiang2†, Yicheng Yang1,2†, Songsong Xie2, Wumei Yuan1 and Yuanzhi Wang1,3*

**Abstract**

**Background:** Increasing numbers of tick-borne pathogens are being discovered, including those that infect humans. However, reports on co-infections caused by two or more tick-borne pathogens are scarce.

**Case presentation:** A 38-year-old male farmer was bitten by a hard tick, presented with fever (37.7 °C), severe headache and ejection vomiting. Lumbar puncture was performed in the lateral decubitus. The cerebrospinal fluid (CSF) was clear, and analysis showed severe increased pressure (320 mm H2O), mild leukocytosis (126.0 × 10⁶/L, mononuclear cells accounting for 73%) and elevated total protein concentration (0.92 g/L). Bacterial cultures of CSF and blood were negative. The diagnosis of *Rickettsia raoultii* and Tacheng tick virus 1 (TcTV-1) co-infection was confirmed by amplifying four rickettsial genetic markers and the partial small (S) RNA segment of TcTV-1 from the patient’s blood. The patient gradually recovered after treatment with levofloxacin and ribavirin.

**Conclusions:** This is the first reported co-infection case with fever and meningitis caused by *R. raoultii* and TcTV-1. It is vital to screen for multiple pathogens in tick-bitten patients, especially in those with severe complex symptoms.

**Keywords:** *Rickettsia raoultii*, Tacheng tick virus 1, Meningitis, Fever, Case report

**Background**

Hard ticks (Acari: Ixodidae) are among the most versatile vectors, capable of transmitting several species of pathogens, including bacteria, protozoa, fungi, nematodes, and viruses, to humans, domestic and wildlife animals [1]. In Xinjiang Uygur Autonomous Region (XUAR, northwestern China), there are natural foci of multiple tick-borne diseases, such as spotted fever, Lyme borreliosis, Crimean-Congo hemorrhagic fever, and tick-borne encephalitis [2]. To date, ten spotted fever group *Rickettsia* (SFGR) were found in XUAR. Among these, *Rickettsia raoultii* was shown to be highly prevalent in northwestern China, i.e. 22.9% (404/1764) of ticks tested positive [3–6]. In previous studies, *R. raoultii* infections have also been increasingly detected in tick bite patients throughout China. Common nonspecific manifestations in 35 tick-bitten patients with mild to moderate or severe illness included fever (74.3%, 36.8–41.0 °C), malaise (71.4%), nausea (42.9%), myalgia (31.4%), lymphadenopathy (28.6%), vomiting (25.7%) and headache (14.3%). Only two patients (5.7%, 2/35) had meningeal syndrome [7–10].

In addition, some emerging bunyaviruses, such as Tacheng tick virus 1 (TcTV-1) and Tacheng tick virus 2 (TcTV-2), are suggested to caused human infections in XUAR [11, 12]. Epidemiological investigations showed that (i) 7.7% (26/339) of adult ticks tested positive for TcTV-1, and (ii) syndrome of an index patient infected with TcTV-1 included fever (as high as 39.0 °C), a local skin bulge (about 5 × 5 cm in size), rash, chill, muscle...
soreness and arthralgia. Moreover, the patient showed neurologic signs, such as headache and dizziness [11].

Previously, SFGR species were reported in co-infections with Borrelia burgdorferi, Anaplasma phagocytophilum, Orientia tsutsugamushi and herpes simplex virus 2 [13–16]. In this study, we reported for the first time a tick bite patient co-infected with R. raoultii and TcTV-1 in XUAR.

**Case presentation**

The patient was a 38-year-old previously healthy man, who was engaged in agricultural work. He reported that he was bitten by a tick on his left upper limb, at the tenth intercostal area of the left anterior axillary line on May 23, 2019. When he noticed and removed the tick, it was engorged. Two days later, a painful erythematous mass (about 6 × 6 mm in size) developed around the site of the tick bite. On May 30, he developed persistent fever (37.7 °C ± 0.2 °C) and headache, and consulted a doctor at a local medical center. He was treated with calcium gluconate (1 g/day intravenously) and amoxicillin (0.5 g thrice daily orally) for 3 days. On June 4, his headache became increasingly severe, and was accompanied with nausea, mild neck stiffness and intense vomiting (more than 20 times per night). On June 5, the patient visited the Manasi County Hospital, XUAR. The patient received the brain MRI examination with DWI, and no obvious abnormality was found. Clinical hemogram, biochemical tests showed that most indexes were within the normal range. However, the white blood cell count (16.35 × 10^9/L), absolute neutrophil count (13.25 × 10^9/L), C-reactive protein (10.45 mg/L), glutamic-pyruvic transaminase (73.00 U/L), glutamic-oxalacetic transaminase (40.50 U/L) and total bile acid (21.3 U/L) levels. He was transferred under coma to the Department of Emergency Surgery, the First Affiliated Hospital, School of Medicine, Shihezi University. The cerebrospinal pressure of the patient reached 320 mm H2O (normal range, 80–180 mm H2O) as measured with lumbar puncture examination. The protein concentration of cerebrospinal fluid (CSF) increased to 0.92 g/L (normal range, 0.15–0.45 g/L). The leukocyte count was 126.0 × 10^9/L (normal range, 0–8 × 10^9/L), of which mononuclear cells accounted for 73.0%. Other clinical data are shown in Table 1. Blood and CSF samples were negative in BacT/Alert blood culture system.

To search for more eventual co-infecting pathogens, the patient’ anticoagulated blood samples were collected upon admission (being diagnosed as acute meningitis). The DNA and RNA were extracted by a TIANgenomic DNA Kit and an TIANamp Genomic RNA Kit (Tiangen Biotech, Beijing, China), respectively. The complementary DNA (cDNA) was synthesized using the Revert Aid First Strand cDNA Synthesis Kit (Transgen Biotech, Beijing, China). Known tick-borne pathogens, including Rickettsia spp., Anaplasma spp., Ehrlichia spp., Babesia spp., Francisella spp., Borrelia spp., forest encephalitis virus, severe fever and thrombocytopenia syndrome virus, Crimean-Congo hemorrhagic fever virus, TcTV-1 and TcTV-2, were detected using nested PCR (nPCR) or reverse transcription-PCR (RT-PCR).

Four Rickettsia-specific genetic markers, including 543-, 445-, 435-, and 364-bp products of the genes encoding the cell surface antigen 1 (sca1), outer membrane proteins A (ompA), 17 kilodalton antigen (17-kDa), and mitochondrial 16S ribosomal DNA (16SrDNA), were amplified using previously described primers [17]. At the same time, the partial S segment of TcTV-1 (328 bp) was also screened by RT-PCR [11]. The PCR products were purified using the TIAN gel Midi Purification Kit (Tiangen, Beijing, China) and sequenced by Sanger di-deoxy sequencing method (Sangon Biotech, Shanghai, China). Each test was repeated three times. Obtained sequences corresponded to those of R. raoultii and TcTV-1 as revealed by BLAST search (http://www.ncbi.nlm.nih.gov/BLAST/). Phylogenetic trees were constructed using the Maximum-Likelihood method in MEGA 7.0 software (Figs. 1 and 2) [18]. The samples tested negative for other tick-borne pathogens. All sequences obtained in this study were deposited in the GenBank database [TcTV-1: MW752511; ompA: MW752512; sca1: MW752513; 17-kDa: MW752514; 16SrDNA: MW752515].

The patient was treated with intravenous levofloxacin (0.5 g/day) and ribavirin (0.5 g/day) for 8 days and supportive treatment (potassium chloride oral solution, intravenous 20% mannitol, vitamin C and B6 injections). His mental status and body temperature improved at day 3 during hospitalization. On June 15, the patient was discharged from hospital although the data of mild laboratory abnormalities persisted, including the elevated protein concentration of CSF and gamma-glutamyl transpeptidase levels of serum. The patient was put on continuous oral doxycycline (100 mg, twice daily) and ribavirin (0.45 g, thrice daily) treatment at home for 4 days. After a 14-month follow-up, the tick bite patient recovered with no other complications.

**Discussion and conclusions**

In this study, we reported a severe acute meningitis case in a tick bite patient co-infected with R. raoultii and TcTV-1. Clinical manifestations included fever (37.7 °C ± 0.2 °C), severe headache, persistently intense vomiting, mild neck stiffness and raised intracranial pressure (320 mm H2O) with mild mononuclear cells and elevated CSF protein levels. It is well known that spotted fever can cause central nervous system...
infection, presenting as fever, headache, vomiting, increased intracranial pressure and aseptic meningitis [19–21]. In similar studies, slightly increased intracranial pressure (200 mm H₂O) was reported in a case of neurologic abnormalities caused by *R. raoultii* in northwestern China after tick bite, resulting in right eyelid ptosis, lethargy, fever (38.0–41.0 °C), headache and CSF leukocytosis [9]. Elevated intracranial pressure (235 mm H₂O) have been reported in a case of Japanese spotted fever with central nervous system involvement and multiple organ failure [22]. In the present case, severe increased intracranial pressure (320 mm H₂O) was also present, complicated with persistently intense vomiting. This finding means that (i) CSF pressure measurement is necessary in spotted fever patients with neurologic signs, and (ii) some emerging tick-borne viruses should also be considered as co-infecting agents in all endemic areas of spotted fever.

### Table 1  Clinical laboratory tests and complications of the patient

| Laboratory findings, Signs | Results (Hospital admission) | Results (Hospital discharged) | Normal range |
|----------------------------|-------------------------------|-------------------------------|--------------|
| **Laboratory findings**    |                               |                               |              |
| Cerebrospinal fluid test   |                               |                               |              |
| Cerebrospinal pressure     | 320 mm H₂O                    | 220 mm H₂O                    | 80–180 mm H₂O|
| Leukocyte count            | 126.0 x 10⁹/L                 | 2.0 x 10⁹/L                   | 0–8 x 10⁹/L  |
| Protein concentration      | 0.92 g/L                      | 0.56 g/L                      | 0.15–0.45 g/L|
| Chloride                   | 116 mmol/L                    | 124 mmol/L                    | 120–130 mmol/L|
| Pandy test                 | Positive                      | Weakly positive               | Negative     |
| Glucose                    | 2.7 mmol/L                    | 2.86 mmol/L                   | 2.5–4.4 mmol/L|
| Monocytes account          | 73%                           | /                             | /            |
| Cerebrospinal fluid culture| Negative                      | /                             | Negative     |
| **Hemogram**               |                               |                               |              |
| A white blood cell count   | 16.35 x 10⁹/L                 | 12.6 x 10⁹/L                  | 4–10 x 10⁹/L |
| Neutrophil count           | 13.25 x 10⁹/L                 | 8.41 x 10⁹/L                  | 1.4–7 x 10⁹/L|
| Lymphocyte count           | 2.08 x 10⁹/L                  | 3.1 x 10⁹/L                   | 1.2–3.5 x 10⁹/L|
| Hemoglobin level           | 147 g/L                       | 134 g/L                       | 110–160 g/L  |
| Platelet count             | 300 x 10⁹/L                   | 326 x 10⁹/L                   | 100–300 x 10⁹/L|
| Erythrocyte sedimentation rate (ESR) | 24.00 mm/h | 13 mm/h                       | 0–15 mm/h    |
| **Blood biochemistry**     |                               |                               |              |
| Albumin                    | 43 g/L                        | 43 g/L                        | 40–55 g/L    |
| Total bilirubin            | 22.1 umol/L                   | 22.1 umol/L                   | 2–28 umol/L  |
| Glutamic-pyruvic transaminase (ALT) | 73 u/L   | 41.0 u/L                     | 0–40 u/L     |
| Glutamic-oxaloacetic transaminase (AST) | 40.5 u/L | 18.0 u/L                     | 0–40 u/L     |
| Gamma-glutamyl transpeptidase (GGT) | 127.0 u/L | 63.0 u/L                     | 12–43 u/L    |
| **Bacteriological examination** |                           |                               |              |
| Cerebrospinal fluid culture| Negative                      | /                             | Negative     |
| Blood culture              | Negative                      | /                             | Negative     |
| **Signs**                  |                               |                               |              |
| Fever                      |                               |                               |              |
| Temperature on admission   | 37.8 °C                       |                               |              |
| Highest temperature        | 37.9 °C                       |                               |              |
| **Complications**          |                               |                               |              |
| A painful erythematous mass| Yes                           |                               |              |
| Headache                   | Yes                           |                               |              |
| Mild neck stiffness        | Yes                           |                               |              |
| Nausea                     | Yes                           |                               |              |
| Vomiting                   | Yes                           |                               |              |
| Morbus asthenicus          | Yes                           |                               |              |
| Poor diet and sleep        | Yes                           |                               |              |

*“/” mean not detected*
Previously, an index patient single-infected with TcTV-1 also showed lymphocytic meningitis similar to that after SFGR infection [9, 11, 22]. In our study, the patient co-infected with TcTV-1 and R. raoultii presented increased intracranial pressure (320 mm H2O), resulting in severe headache and intense vomiting (> 20 times per night), which might account for overlapping clinical signs of the central nervous system between TcTV-1 and R. raoultii infections. In addition, there are major differences in therapeutic measures between infections caused by various tick-borne pathogens, as exemplified by R. raoultii and TcTV-1. Therefore, early diagnosis based on identification of multiple pathogens, is crucial in tick-bitten patients with severe clinical syndrome. Public health workers and physicians need to be more aware of the clinical complexity of tick-borne pathogen infections, especially in high-risk areas of tick-borne diseases.

There are several limitations to our study. Firstly, rickettsia and TcTV-1 markers should have also been detected in CSF samples, although cellular and

![Fig. 1 Phylogenetic tree of patient’s R. raoultii strain compared with available nucleotide sequences, selected using BLAST (http://blast.ncbi.nlm.nih.gov/Blast.cgi). The tree was constructed by maximum-likelihood (ML; 1000 bootstrap replicates) of concatenated sequence data of four genes (17-kDa-sca1-rrs-ompA) using Molecular Evolutionary Genetics Analysis (MEGA, version 7.0; http://www.megasoftware.net/). The concatenated sequence of R. raoultii are indicated by a black triangle (▲). The sequences of Rickettsia felis were used as the outgroup.](image-url)
biochemical characteristics of CSF were useful in early diagnosis. Secondly, bacterial and viral metagenomic analysis should have been performed in more clinical samples (e.g., pharyngeal swab, urine, and cerebrospinal fluid) associated with the tick-bite patients. In future cases, these would help us to better understand complex clinical manifestations due to infection of single or multiple tick-borne pathogens.

**Abbreviations**

XUAR: Xinjiang Uygur Autonomous Region; SFGR: Spotted fever group Rickettsia; 16S rDNA: Mitochondrial 16S ribosomal DNA; 17-kDa: 17 Kilodalton antigen; ompA: Outer membrane proteins A; sca1: Cell surface antigen 1; CSF: Cerebrospinal fluid; MEGA7.0: Molecular Evolutionary Genetics Analysis 7.0; PCR: Polymerase chain reaction; R. raoultii: Rickettsia raoultii; TcTV-1: Tacheng tick virus 1.

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**Authors’ contributions**

YZW conceived and designed the study, YCY and WMY critically revised the manuscript. YZ and LJ completed the experiment, conducted molecular analyses and drafted the manuscript. SSX treated the patient, and carried out the clinical diagnosis. All authors read and approved the final manuscript.

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**Availability of data and materials**

The datasets generated and analysed during the current study are available in the GenBank repository, Accession number to datasets: TCTV-1: MW752511; ompK: MW752512; sco1: MW752513, 16S rDNA: MW752514.

**Declarations**

**Ethics approval and consent to participate**

This study was approved by the First Affiliated Hospital of Shihezi University. Written informed consent was obtained for all research participants.

**Consent for publication**

Written informed consent was obtained from the patient for publication of this case report and any accompanying images. A copy of the written consent is available for review by the Editor of this journal.

**Competing interests**

The authors declare that they have no competing interests.

**Author details**

1School of Medicine, Shihezi University, Shihezi 832002, Xinjiang Uygur Autonomous Region, China. 2The First Affiliated Hospital, School of Medicine, Shihezi University, Shihezi 832002, Xinjiang Uygur Autonomous Region, China. 3NHC Key Laboratory of Prevention and Treatment of Central Asia High Incidence Diseases, First Affiliated Hospital, School of Medicine, Shihezi University, Shihezi City 832002, Xinjiang Uygur Autonomous Region, China.

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