Comparison of Polypropylene and Ceramic Microfiltration Membranes Applied for Separation of 1,3-PD Fermentation Broths and Saccharomyces cerevisiae Yeast Suspensions

Wirginia Tomczak *† and Marek Gryta *†

Faculty of Chemical Technology and Engineering, West Pomeranian University of Technology in Szczecin, ul. Pułaskiego 10, 70-322 Szczecin, Poland
* Correspondence: tomczak.wirginia@gmail.com (W.T.); marek.gryta@zut.edu.pl (M.G.)
† New address: CEA, DEN/DEC, 13108 Saint-Paul-les-Durance, France.

Abstract: In recent years, microfiltration (MF) has gained great interest as an excellent technique for clarification of biological suspensions. This paper addresses a direct comparison of efficiency, performance and susceptibility to cleaning of the ceramic and polymeric MF membranes applied for purification of 1,3-propanediol (1,3-PD) fermentation broths and suspensions of yeast Saccharomyces cerevisiae. For this purpose, ceramic, titanium dioxide (TiO\textsubscript{2}) based membranes and polypropylene (PP) membranes were used. It has been found that both TiO\textsubscript{2} and PP membranes provide sterile permeate during filtration of 1,3-PD broths. However, the ceramic membrane, due to the smaller pore diameter, allowed obtaining a better quality permeate. All the membranes used were highly susceptible to fouling with the components of the clarified broths and yeast suspensions. The significant impact of the feed flow velocity and fermentation broth composition on the relative permeate flux has been demonstrated. Suitable cleaning agents with selected concentration and duration of action effectively cleaned the ceramic membrane. In turn, the use of aggressive cleaning solutions led to degradation of the PP membranes matrix. Findings of this study add to a growing body of literature on the use of ceramic and polypropylene MF membranes for the clarification of biological suspensions.

Keywords: 1,3-propanediol; ceramic membrane; clarification; fermentation broth; fouling; membrane cleaning; membrane wetting; microfiltration; polymeric membrane; yeast suspension

1. Introduction

Microfiltration (MF) is one of the oldest membrane technologies [1], which is characterized by operating pressure lower than 0.35 MPa and high permeate fluxes, mainly between $10^{-4}$ and $10^{-2}$ m/s [2]. Over the last two decades, many attempts have been made by researchers to comprehensively investigate the use of MF in the clarification of fermentation broths [3–18] and yeast suspensions [19–32]. It is due to the fact that separation of biological materials using conventional centrifugal methods is difficult and expensive [33]. In turn, MF is an effective technique for turbidity removal [34] and offers several remarkable advantages, such as low cost, easy operation and implementation, high productivity, absence of phase transition, non-use of additional solvents and so on [8].

Nowadays, advanced clarification of biological suspensions can be performed using a wide variety of commercially available MF membranes. Moreover, according to [35] the global market of MF membranes should increase from $2.4$ billion in 2018 to $3.7$ billion by 2023. Nevertheless, predicting the most suitable membrane for the identified application is a challenging task and requires a multi-criteria approach. Speed [36] has indicated that among the most important factors in selecting the right membrane are mechanical strength, resistance to cleaning chemicals, pore size and surface charge. Additionally, since the decrease in permeate flux is one of the most important economic criterions for a given
separation process, due attention should be paid to the membrane performance. It is well established that the fouling phenomenon is a complex physicochemical phenomenon that depends on the process operating parameters and the interactions between the membrane and feed components. Moreover, it leads to the necessity of frequent membrane cleaning. Consequently, the extensive knowledge of the membrane material and specification is a key factor in successfully realizing an efficient and economically viable MF process. Moreover, as it has been pointed out by Warsinger et al. [37], comparing different types of membrane materials leads to an understanding of their advantages and disadvantages.

In recent years, research on the separation processes using ceramic membranes has been receiving growing attention. Indeed, Li et al. [38] in the newly published review article have demonstrated that the publications focused on ceramic membranes have been expanding over the last 10 years, from less than 150 in 2007 to double that in 2018. It is due to the distinguished merits of ceramic membranes over their polymeric counterparts. As recognized in the literature [24,39–44], ceramic membranes offer mechanical strength and resistance to harsh chemical conditions. In addition, they can withstand temperatures of up to 500 °C [24]. The above-mentioned advantages of ceramic membranes enable their specialized use in extreme operating conditions. For instance, they allow for aggressive physical and chemical cleaning of the modules, which ensures the removal of irreversible fouling, without the risk of damaging membrane integrity. Moreover, ceramic membranes show excellent corrosion resistance [41] as well as inertness to microorganisms [24] and organic media [45]. The high reliability of ceramic membranes decreases the cleaning requirements, reduces the replacement of membrane modules and, thus, prolongs their operational life expectancy. Nevertheless, ceramic membranes are more prone to breakage than polymeric membranes [46], thus, they need to be handled carefully.

Nowadays, ceramic membranes are available in different configurations, among which tubular-shaped clearly dominate the field [47]. Generally, ceramic membranes have a multilayer structure consisting of one or more different inorganic materials. In industry, the composition of ceramic membranes is usually based on alumina (Al₂O₃), zirconia (ZrO₂), titania (TiO₂) or combination of these oxides [47] and their characteristics depend on the materials involved. Since the surface groups of ceramic and polymeric membranes are different, differences in their fouling are expected [46]. Indeed, ceramic membranes are generally hydrophilic in nature which ensures a lower protein adhesion than hydrophobic membranes, such as polypropylene and polyethylene [42]. Moreover, they have relatively narrow pore size distribution and higher porosity, resulting in better separation characteristics and a higher flux [46].

It is well recognized that the high prices of raw materials and manufacturing are identified as the main disadvantages of ceramic membranes. Indeed, Issaoui and Limousy [44] have noted that the price of an α-alumina porous tubular ceramic membrane with average pore diameters in the range from 1000 to 6000 nm ranges between $500 and $1000/m². Definitely, the cost of ceramic membranes is the main limiting factor in their large-scale industrial applications. In light of this, polymeric membranes can be considered an alternative.

The most commonly used polymers in membrane production are polypropylene (PP), polyvinylidene fluoride (PVDF), polyacrylonitrile (PAN), polyethersulfone (PES), cellulose acetate (CA), polyamide (PI) and polyvinylpyrrolidone (PVP) [48]. Among the main advantages of polymeric membranes are the low cost of production and possibility to synthesize novel polymers with well-defined structures [49]. In addition, they show minimal interaction with organic compounds [50]. However, due to their nature, they are characterized by a reduced chemical stability to high and low pH, which limits the possibilities of the chemical cleaning used [51]. It is an important issue which must be considered, since for cleaning membranes fouled by bacterial suspensions, the most commonly used are aggressive cleaning agents, such as: sodium hydroxide (NaOH) and sodium hypochlorite (NaOCl) [5]. Indeed, it has been widely reported [52–59] that chemical cleaning of polymeric membranes may lead to modification of their hydraulic performances, mechanical properties and physical structures. For instance, Malczewska and Zak [54]
have investigated the impact of NaOH, hydrochloric acid (HCl) and NaOCl solutions on the properties of a flat sheet PES ultrafiltration membrane. The authors have demonstrated that NaOCl led to degradation of the membrane most quickly. In addition, they found changes in the membrane surface properties, morphology and hydraulic performance caused by NaOCl, indicating that it could cause ageing of the membrane after a prolonged exposure. In turn, in [56] it has been shown that NaOCl may lead to ageing on the PVDF membrane after prolonged exposure and significant changes in its mechanical properties, hydraulic performance and chemical groups as well as physical structures.

As stated before, the membrane material is considered as one of the major factors playing a role on the membrane fouling phenomenon. Hence, in the last years, an important research effort has been made on the comparison of performance of ceramic and polymeric membranes used in various pressure driven separation processes, such as: MF of milk [51], surface water [46] and algal rich water [60], MF and ultrafiltration (UF) of ground water [61] and real produced water [62], UF of synthetic feedwater [63,64], oil-in-water emulsion [65] and cadmium(II) ions from aqueous solutions [49] as well as nanofiltration (NF) of trace organic chemicals [66] and acid mine waters [67]. For instance, Vatai et al. [65] have demonstrated the comparison of a ceramic ZrO$_2$ membrane (nominal pore size of 20 nm) with a polyaryletherketone (PAEK) membrane (MWCO of 100 kDa) applied for UF of oil-in-water emulsion. The authors have pointed out that, although oil rejection of a ceramic membrane was not satisfactory, its advantage over a polymer membrane in terms of productivity has been found. Moreover, the necessity of conducting the filtration process under optimal operating conditions, especially in the case of ceramic membranes, was emphasized. Also, the authors have highlighted their better mechanical, thermal and chemical stability. In turn, Kenari et al. [63] have investigated the UF process of synthetic water with using two types of membranes: a ceramic tubular membrane made of ZrO$_2$ and a support of $\alpha$-Al$_2$O$_3$ (MWCO of 150 kDa) and a polymeric hollow fiber membrane composed of a blend of PVP and PES (MWCO of 200 kDa). The authors have shown that, although both membranes used were similarly fouled during the investigation process, the physical cleaning was more effective for a ceramic membrane than a polymeric one.

It is essential to mention that in the available literature studies comparing the performance of polymeric and ceramic membranes used for clarification of biological suspensions have not been found. Therefore, much effort should be devoted to comparing performances of ceramic and polymeric membranes applied for MF of 1,3-propanediol (1,3-PD) fermentation broths and suspensions of yeast *Saccharomyces cerevisiae*. The motivation lies in the fact that 1,3-PD is one of the most important organic chemical materials which is being extensively applied in a range of industrial applications. For instance, it is used for synthesis of a biodegradable polymer known as polytrimethylene terephthalate (PTT) as well as production of detergents, cosmetics, composites and lubricants [23,68–71]. However, fermentation broths of 1,3-PD are complex media which, apart from the desired product, contain water, microbial cells, residual carbon source (e.g., glycerol) and various by-products (e.g., acetic acid, lactic acid, ethanol and 2,3-butanediol) [72]. Hence, in order to obtain the pure main product, the clarification of post-fermentation solution as the first step of downstream processing is required. In subsequent stages, metabolites are separated by the nanofiltration (NF) and reverse osmosis (RO) [73]. The spiral wound structure of membrane modules used in these processes requires careful removal of turbidity (Silt Density Index < 5) and sterility of the obtained MF permeate.

In turn, interest in the yeast filtration is related to the fact that *Saccharomyces cerevisiae* is the most intensively studied unicellular eukaryote and one of the main industrial microorganisms [74]. Indeed, its cytoplasm is a rich source of various bioproducts (e.g., proteins, polysaccharides and cytoplasmic enzymes) that are valuable in a massive variety of fields, such as the beverage and food industry, biotechnology and pharmacology [23]. Furthermore, *Saccharomyces cerevisiae* is one of the hosts for synthetic biology [20]. Importantly, this microorganism has well-defined granulometric properties, its suspension is
easily prepared feed and, hence, it is an excellent model system for studying the behavior and treatment of biological materials [21,24].

Given the background described above, this work addresses the direct comparison of efficiency, performance and susceptibility to cleaning of the ceramic and polymeric microfiltration membranes applied for clarification of 1,3-propanediol fermentation broths and aqueous suspensions of dry distillery yeast Saccharomyces cerevisiae-Bc 16a. For this purpose, the membranes made of TiO$_2$ and PP in different module configurations (tubular and capillary) were tested.

2. Materials and Methods

2.1. Feed Suspensions

The glycerol fermentation was carried out in a LiFlusGX bioreactor (Biotron Inc., Korea), using Citrobacter freundii and Lactobacillus casei bacteria isolated and characterized in Department of Biotechnology and Food Microbiology, Poznań University of Life Science (Poland). The process has been described in detail in our previous works [72,75]. The obtained 1,3-PD post-fermentation solutions were subsequently used as feed in the studied MF processes. Their compositions are presented in Table 1.

Table 1. The compositions of post-fermentation broths with Lactobacillus casei and Citrobacter freundii bacteria.

| Component [g/L]       | Lactobacillus casei | Citrobacter freundii |
|------------------------|----------------------|-----------------------|
| wetted mass            | 21.6–27.9            | 28–32.5               |
| meat extract           | 8                    | 1.5                   |
| yeast extract          | 5                    | 2                     |
| peptone K              | 10                   | 2.5                   |
| lactic acid            | 13.5–15.2            | 2.1–3.5               |
| 1,3-propanediol        | -                    | 18.9–22.5             |
| glycerol               | 3–4.7                | 1.7–3.8               |
| acetic acid            | 2–3.6                | 1.2–2.3               |
| formic acid            | 0.25–0.3             | 0.8–1.22              |
| succinic acid          | -                    | 0.7–2.13              |
| ammonium citrate       | -                    | -                     |
| K$_2$HPO$_4$·3H$_2$O   | 0.6                  | 3.4                   |
| KH$_2$PO$_4$           | -                    | 1.3                   |
| MgSO$_4$·7H$_2$O       | 0.4                  | 0.9                   |
| (NH$_4$)$_2$SO$_4$     | -                    | 2                     |
| CH$_3$COONa            | 1.5                  | -                     |
| CaCl$_2$·2H$_2$O       | -                    | 0.01                  |
| CoCl$_2$·6H$_2$O       | 0.004                | 0.002                 |

Suspensions of commercially available distillery yeast Saccharomyces cerevisiae-Bc 16a (Wytwórnia Drożdży Maszewo Łęborskie, Poland) were also used as feed in this study. The yeast concentrations of 0.1, 0.16 and 0.5 g/L were prepared by mixing dry yeast cells and distilled water at room temperature for 30 min.

2.2. Microfiltration Set-Up and Process Parameters

Microfiltration experiments were carried out in the installations (Figure 1), the schemes of which were described in detail in our previous studies [5,76].

In the present research, four different modules with MF membranes were used (Table 2). Modules M1 and M2, without an external shell, were tested in the installation shown in Figure 1a. In each of these modules, the four capillary membranes made of polypropylene were assembled. In turn, in the pilot installation presented in Figure 1b, tubular modules (diameter 12 mm) with the polypropylene membrane (module M3) or ceramic membrane (M4) were tested.
Due to the hydrophobic properties of polypropylene, water does not wet PP membranes, while water penetration into the pores (Liquid Enter Pressure—LEP) occurs under the pressure of 0.2–0.3 MPa. Therefore, prior to the MF experiments, a membrane wetting operation was required. For this purpose, the Accurel PP S6/2 membranes assembled in the modules M1 and M2 were submerged into ethanol for 15 min, and then the filtration of ethanol (100 mL) was conducted. Subsequently, in order to remove residual ethanol, the membranes were flushed by distilled water (2 L) and the permeate flux $J$ as a function of transmembrane pressure (TMP) was determined. Then, the filtration of distilled water under TMP equal to 0.03 MPa for 4 h was performed. Due to the instability of the recorded permeate flux values, the next day water was removed from the system, the membranes
were immersed in ethanol and the filtration was conducted for 15 min. Then, alcohol was rinsed with water and the relationship \( J = f(\text{TMP}) \) was studied. Subsequently, water filtration was continued under \( \text{TMP} = 0.03 \text{ MPa} \) for 2 h. Then, water was replaced with ethanol and the above-described operation was repeated two times. Since the obtained relationship between the permeate flux and \( \text{TMP} \) was repeatable, the studies on MF of biological suspensions were started. In turn, the Accurel PP V8/2 HF membrane assembled in the M3 module was wetted by filling the outer space of the module with ethanol and filtering it for 15 min. Subsequently, the water filtration was carried out for 30 min. In the next step, the water flux as a function of \( \text{TMP} \) was determined using a pressure ranging from 0.04 to 0.1 MPa. Finally, the water filtration was continued for several hours, obtaining a constant relationship \( J = f(\text{TMP}) \), independent of the feed flow rate.

Microfiltration experiments of 1,3-propanediol fermentation broths and yeast Saccharomyces cerevisiae suspensions have been carried out at constant temperature of 30 °C and across a wide range of process parameters. The \( \text{TMP} \) ranging from 0.02 to 0.08 MPa was used. In turn, the applied feed flow velocity \( V_F \) was from 0.8 to 11 m/s.

The permeate flux \( J \) was calculated based on the measured volume of collected permeate according to Equation (1), as follows:

\[
J = \frac{\Delta V}{\Delta t S}
\]

where \( \Delta V \) is the permeate volume (L) collected over \( \Delta t \) period (h), \( S \) is the total active membrane area (m²).

Fouling intensity was determined by measuring the relative permeate flux, defined as the ratio between the actual permeate flux \( J \) and maximum permeate flux \( J_0 \) for a clean membrane.

Cleaning efficiency was estimated from the ratio of water permeate fluxes after membrane cleaning to the maximum flux, obtained under the same operational conditions (temperature, transmembrane pressure and feed flow velocity).

2.3. Analytical Methods
Permeate and feed samples were analyzed in terms of compounds content, turbidity and number of bacteria. The analytical methods used for this purpose were described in detail in our previous work [5]. The morphology of the membrane surface was examined by scanning electron microscopy (SEM) (SU-70 and SU-8000, Hitachi High Technologies Co., Tokyo, Japan).

3. Results
3.1. Membranes Morphology and Maximum Performance
The polypropylene membranes used for the present studies were formed via TIPS method. It has been determined that, due to the interaction with the gelling bath, the porosity of the membrane surface (Figure 2a) slightly differs from that observed inside the wall (Figure 2b). In turn, the ceramic membrane manufactured by Tami Ind. had both a support layer and a thin active layer made of TiO₂ (Table 2). The structure of these layers was crystalline, ranging from a small size in the surface active layer (Figure 2c) to over 20 µm in the support layer (Figure 2d).

As reported in Section 2.2, an operation of wetting the PP membranes with ethanol was carried out prior to the MF experiments. Subsequently, in order to remove the solvent, the membranes were shortly rinsed with distilled water. However, the performed operation was not effective since the membranes regained their hydrophobic properties. Probably, the new PP membrane, after washing the ethanol with water, shows a strong tendency to push the water out of the pores and part of its volume is again filled by air, which blocks the flow of water through the pores. Indeed, after 2–3 h of the water filtration a rapid decline in the permeate flux was observed. For instance, for \( \text{TMP} \) equal to 0.05 MPa, a decrease in the M2 module performance from 630 to 112 L/m²h was reported.
Figure 2. SEM images of tested membranes. Accurel PP S6/2 membrane: (a) surface, (b) cross-section, ceramic membrane: (c) surface and (d) cross-section.

An important point which should be noted is that stable modules performance (Figure 3, lines L2) was obtained after 2–3 days of the rinsing, during which the operation of membrane wetting was repeated several times. Moreover, the results presented in Figure 3 show that the PP membranes performance after wetting depends on the pumping pressure of the rinse water. The M3 module (V8/2 HF membrane) was rinsed under much higher pressure (0.04–0.1 MPa) than the S6/2 membranes assembled in M1 and M2 modules (0.01–0.05 MPa). Probably, the use of higher TMP values prevents water from being pushed out of the pores and re-creating the gas phase in them, hence, the performance drop of the M3 module was much lower.

In turn, the performance of the ceramic membrane used was more stable. The membrane was conditioned by rinsing with distilled water for two days, with fluctuations of the permeate flux up to 30%. Finally, the performance of the M4 module stabilized after washing the membrane with 3% NaOH solution for 60 min, and then for the same period of time with 3% H₃PO₄ solution. It is worth noting that under the same operating conditions, the maximum permeate flux noted for the ceramic membrane (M4) is lower than that for the V8/2 membranes (M3) (Figure 3). This observation is related to the fact that the V8/2 membranes have a larger pore size (0.20 μm) than the ceramic membrane (0.14 μm) (Table 2). Obviously, membranes with larger pore size are more porous, which means they provide a higher permeate flux [31].

Figure 3 shows the stabilized performances obtained for the tested modules fed with distilled water. Unfortunately, the permeate flux decreases significantly during filtration of real solutions. Importantly, the intensity of the flux decline, apart from the properties of
membranes (hydrophilic or hydrophobic), is significantly influenced by the composition of the feed. For this reason, in order to investigate the intensity of the fouling phenomenon, in subsequent stages of the research, in addition to yeast suspension, real broths with different compositions (Table 1) were used.

\[\text{Figure 3.}\] The maximum permeate flux as a function of transmembrane pressure (TMP): 1—after one operation of membrane wetting, 2—stabilized \(J_0\) values after several operations of membrane wetting.

3.2. Filtration of Fermentation Broths with Citrobacter freundii Bacteria

The glycerol fermentation process was carried out for 48 h. Once the process run was complete, the obtained 1,3-propanediol post-fermentation solution was clarified using the MF membranes. Figure 4 shows the changes in the relative permeate flux of the M1 module during the filtration of solution with *Citrobacter freundii* bacteria. The process under TMP equal to 0.03 MPa and \(V_F\) of 0.8 m/s was conducted. It has been demonstrated that the permeate flux systematically decreased over 120 min. Obviously, it was caused by the membrane fouling, defined as “the accumulation of substances on the membrane surface and/or within the membrane pores, which results in deterioration of membrane performance” [77]. Subsequently, the solution was drained from the tank and the membrane module was flushed with 2 L of distilled water (TMP = 0). Then, the performance of the membrane for distilled water was controlled. It has been reported that the permeate flux during the MF of distilled water increased slightly, indicating that the presented method of the membrane cleaning was not efficient. Then, the MF process of the broth was resumed. It can be clearly observed that after three series of filtration, followed by systematically carried out water filtration, the maximum performance of the membrane module decreased by nearly 50%.

One membrane from the M1 module was collected and the surface morphology of the selected membrane was examined using scanning electron microscopy (SEM). Meanwhile, three membranes remaining in the M1 module were rinsed with 1% NaOH solution for 30 min. It is important to note that this operation allowed to almost completely recover the maximum permeate flux (Figure 4, CR point). The high effectiveness of NaOH solution is related to the fact that at caustic conditions, large organic particles (e.g., colloids and microbes) can be disintegrated into fine particles and soluble organic matters, while organic matters (e.g., proteins and carbohydrates) can be hydrolyzed and solubilized into small molecules [78]. It is worth noting that in the study [10], NaOH solution was successfully
used as a cleaning agent for microfiltration membranes made of CA and PVDF and a PES ultrafiltration membrane fouled with *Bacillus thuringiensis* fermentation broth.

![Figure 4](image-url)

**Figure 4.** Changes in relative permeate flux during microfiltration (MF) of 1,3-propanediol (1,3-PD) fermentation broth with *Citrobacter freundii*. Module M1, TMP = 0.03 MPa, V_F = 0.8 m/s, CR—chemical cleaning (1% NaOH).

Considering the application of NF/RO processes for MF permeate separation, the sterility and turbidity of filtered fermentation broths control the efficiency of the MF process. Therefore, in the present study these quality attributes of the obtained permeate were continuously controlled. It has been found that both the ceramic and PP membranes used provided a sterile permeate. Indeed, although the number of bacteria in the feed was of an order of 12 log CFU (colony-forming units-CFU/mL), no bacteria was detected in the permeate samples.

Regarding turbidity, during the MF process with the M1 module, the broth was recirculated through the system, which resulted in a continuous concentration of the feed and, consequently, an increase in its final turbidity to about 8000 NTU (Figure 5). Despite such a high value, the turbidity-free filtrate was obtained. It is worth noting that during the first series of measurements with the new membranes, the permeate turbidity increased to 3 NTU, while in the third series it stabilized at 1 NTU. The observed decrease of the NTU values during the MF process indicated the phenomenon of membranes fouling, which was confirmed by the observed decrease in module performance (Figure 4). Roughly speaking, the membrane efficiency can be increased due to the cake formation, as particles with diameters smaller than the pore size of the membrane are more susceptible to being captured by the cake [79]. An apparent decrease in the turbidity of permeate samples during the pressure driven separation processes of fermentation broths has also been reported in several previous studies [5,7,72,80].

Figure 6 shows the SEM image of the PP S6/2 membrane surface after the MF process of fermentation broth with *Citrobacter freundii* performed under TMP of 0.03 MPa and V_F of 0.8 m/s. It can be clearly observed that the surface of the membrane tested was completely covered by deposit in which the *Citrobacter freundii* cells can be seen. As it has been described in [81] they are rod shaped and typically range from 1 to 5 µm in length.
described in [81] they are rod shaped and typically range from 1 to 5 µm in length. It can be clearly observed that the surface of the membrane tested was completely covered by deposit in which the deposit formed during 6 h of MF process.

![SEM image of PP S6/2 membrane (module M1) surface covered by deposit after MF process](image)

**Figure 6.** SEM image of PP S6/2 membrane (module M1) surface covered by deposit after MF process of fermentation broth with *Citrobacter freundii*. Deposit formed during 6 h of MF process.

It is well known that the adverse effects caused by fouling may be reduced by operating the filtration process under an increased shear rate near the membrane. Generally, enhanced hydrodynamic conditions are applied in the systems equipped with ceramic membranes. Indeed, the increase of the permeate flux with increase of the shear rate has been reported in several previous works [5,7,8,27,80,82–84] where the use of ceramic MF and UF membranes in clarification of various biological suspensions has been studied. A similar pattern of results was obtained in the present work. In our research, the effect of the feed velocity during the MF of 1,3-PD fermentation broth with using the ceramic membrane (module M4) under TMP equal to 0.08 MPa has been investigated. It was found that increasing the feed flow velocity from 5.5 to 11 m/s allowed the increase of the steady state permeate flux from 180 to 228 L/m²h. It clearly indicates that two-fold increase of the feed velocity led to an increase of the relative flux from 0.3 to 0.38 (Figure 7). This observation can be explained by the fact that the higher feed crossflow velocity led to a reduction of the deposited particles and consequently an increase in the permeate flux.
However, it should be mentioned that in industry, the use of high crossflow velocities leads to high pressure losses and increased process costs [51].

![Graph showing the effect of feed flow velocity on the relative permeate flux during MF of 1,3-PD fermentation broth with Citrobacter freundii. Module M4, TMP = 0.08 MPa, J0 = 311 l/m²h.](image)

**Figure 7.** Effect of feed flow velocity on the relative permeate flux during MF of 1,3-PD fermentation broth with *Citrobacter freundii*. Module M4, TMP = 0.08 MPa, $J_0 = 311 \text{ l/m}^2\text{h}$.

Figure 8 shows the changes in the feed and permeate turbidity during the MF of 1,3-PD fermentation broth with *Citrobacter freundii* using the ceramic membrane.

![Graph showing changes in feed and permeate turbidity during MF of 1,3-PD fermentation broth with *Citrobacter freundii*. Module M4, TMP = 0.08 MPa.](image)

**Figure 8.** Changes in feed and permeate turbidity during MF of 1,3-PD fermentation broth with *Citrobacter freundii*. Module M4, TMP = 0.08 MPa.

It has been found that, likewise to the process with using the PP membranes, the obtained permeate was sterile and its turbidity decreased over the MF time. It has been demonstrated that the ceramic membrane used provided obtaining the permeate characterized by the turbidity equal to 0.2 NTU at the end of the trial. The most conspicuous observation to emerge from the data comparison was the quality of permeate obtained...
that the separation improvement was mainly due to internal membrane fouling. The turbidity of the permeate was independent of the feed flow velocity (Figure 8), indicating significant effects on the obtained permeate turbidity.

The filtration process of seawater collected from a seawater desalination reverse osmosis plant provided the permeate with 3.8 NTU and 4.8 NTU, equal to 640.44 ± 380.32 NTU. The authors have shown that the use of ceramic membranes with the pore sizes of 0.14 and 0.80 µm provided the permeate with 3.8 NTU and 4.8 NTU, respectively. On the other hand, our results differ from those of Ciu et al. [86] who studied the filtration process of seawater collected from a seawater desalination reverse osmosis plant. For this purpose, the authors have used ceramic membranes with pore sizes equal to 50, 200, 500 and 800 nm. They have indicated that the pore size of membranes has insignificant effects on the obtained permeate turbidity.

Moreover, results obtained in the present study (module M4) have shown that the turbidity of the permeate was independent of the feed flow velocity (Figure 8), indicating that the separation improvement was mainly due to internal membrane fouling.

Membrane chemical cleaning is considered the most effective method to recover the permeate flux. However, its efficacy depends on several factors, such as: the membrane and fouling type, choice of cleaning agents and their concentration as well as cleaning conditions (e.g., temperature, pressure, cross-flow velocity, pH, time) [78]. However, as stated before, prolonged exposure to chemical cleaning agents may lead to physical and chemical degradation of polymeric membranes, resulting in a change in their performance and selectivity. Therefore, in order to investigate the impact of multiple cleaning cycles on the polypropylene membranes’ properties, the long-term MF process of 1,3-PD fermentation broth with Citrobacter freundii with periodically rinsing the module with 1% NaOH solution has been conducted (Figure 9).

\[ \text{Figure 9.} \text{ Changes in relative permeate flux during the long-term MF process of 1,3-PD fermentation broth with Citrobacter freundii, with periodically cleaning the module with 1\% sodium hydroxide (NaOH) solution. Module M1, TMP = 0.03 MPa, } V_F = 0.8 \text{ m/s, CR—chemical cleaning.} \]

The clarification process under TMP of 0.03 MPa and \( V_F \) equal to 0.8 m/s has been conducted. Results obtained in our research revealed that cleaning allowed the recovery of the initial permeate flux in 60–80%. Importantly, the performance of MF with distilled water, conducted in the next step, allowed the complete recovery of the maximum permeability of the membranes. This noteworthy result suggests that the NaOH solution did not completely...
remove the deposit but allowed a part of it to be loosened. Hence, the water filtration provided to cleanse the deposit, especially, that accumulated in the membrane's pores.

It is necessary to mention that, although the multiple cleaning cycles have been performed, the recorded permeate turbidity for the last two measurement series (Figure 9) was lower than 1 NTU (Figure 10). What becomes apparent from the discussed results is that in the end of the process a better separation efficiency was obtained (Figure 5).

The SEM analysis showed slight surface degradation of the PP membranes (Figure 11). Since maintaining the good separation properties of membranes has been documented (Figure 10), it can be concluded that the cleaning with NaOH solution changed the top layer of membranes, while the pores were not significantly enlarged. However, in comparison with Figure 2a, an increase in the porosity of the membrane surface can be observed. It could facilitate the penetration of the foulants deeper into the membrane wall, which has resulted in an increase in the decline of relative flux observed for the last two measurements series, shown in Figure 9.

![Figure 10](image1.png)

**Figure 10.** Changes in feed and permeate turbidity during the last two measurement series of the MF of 1,3-PD fermentation broth with *Citrobacter freundii*. Module M1, TMP = 0.03 MPa, VF = 0.8 m/s.

The clarification process under TMP of 0.03 MPa and VF equal to 0.8 m/s has been conducted. Results obtained in our research revealed that cleaning allowed the recovery of the initial performance of the module required a more complex cleaning procedure (Figure 12). In order to determine the minimum cleaning time required to recover the maximum permeate flux, cycles of 5–10 minutes were performed. It has been found that recovering the initial performance of the module required a more complex cleaning procedure (Figure 12). In order to determine the minimum cleaning time required to recover the maximum permeate flux, cycles of 5–10 minutes were performed. Since maintaining the good separation properties of membranes has been documented (Figure 10), it can be concluded that the cleaning with NaOH solution changed the top layer of membranes, while the pores were not significantly enlarged. However, in comparison with Figure 2a, an increase in the porosity of the membrane surface can be observed. It could facilitate the penetration of the foulants deeper into the membrane wall, which has resulted in an increase in the decline of relative flux observed for the last two measurements series, shown in Figure 9.

![Figure 11](image2.png)

**Figure 11.** SEM image surface of Accurel PP S6/2 membrane. Module M1—membranes several times rinsed with 1% NaOH solution.
In regards to the ceramic membrane, it has been reported that its cleaning after the filtration of 1,3-PD fermentation broths with *Citrobacter freundii* (Figure 7, TMP = 0.08 MPa, \( V_F = 5.5 \) m/s) with only 1% NaOH did not provide satisfactory results. Indeed, it has been found that recovering the initial performance of the module required a more complex cleaning procedure (Figure 12). In order to determine the minimum cleaning time required to recover the maximum permeate flux, cycles of 5–10 min were performed.

![Figure 11. SEM image surface of Accurel PP S6/2 membrane. Module M1—membranes several times rinsed with 1% NaOH solution.](image)

In regards to the ceramic membrane, it has been reported that its cleaning after the filtration of 1,3-PD fermentation broths with *Citrobacter freundii* (Figure 7, TMP = 0.08 MPa, \( V_F = 5.5 \) m/s) with only 1% NaOH did not provide satisfactory results. Indeed, it has been found that recovering the initial performance of the module required a more complex cleaning procedure (Figure 12). In order to determine the minimum cleaning time required to recover the maximum permeate flux, cycles of 5–10 min were performed.

It has been demonstrated that the membrane rinsing with distilled water for 30 min allowed to recover the maximum membrane permeability in 40% (Figure 12). Then, in order to increase the cleaning efficiency, 3% NaOH solution was used. It has been found that after five cycles of caustic rinsing, the M4 module performance increased to 86% of its maximum value. However, the subsequent membrane rinsing and water filtration for 30 min did not allow improving the cleaning efficiency. This result is different from that obtained for PP membranes, where, after rinsing with NaOH solution, additional water filtration ensured the recovery of initial membranes permeability (Figure 9). In turn, the efficiency of the M4 module slightly increased after backwashing (25 min total, Figure 12, operations 10–14) and only membrane rinsing with 3% \( H_3PO_4 \) solution for 50 min allowed recovery of the initial module performance (Figure 12, operation 15–20).

The cleaning time determined for individual rinsing solutions (Figure 12) was applied for the procedure of the M4 module cleaning (Figure 13) after the filtration was performed at the feed flow velocity equal to 11 m/s (Figure 7). The efficiencies obtained for each stage of membrane cleaning, presented in Figure 13, were similar to the previous ones (Figure 12). However, the rinsing of the membrane with 3% NaOH solution for 25 min provided a lower recovery of the membrane performance, equal to 69% of its maximum value (instead of 86%). In turn, after 25 min of backwash, 71% of the maximum permeate flux has been noted. It has been determined that final cleaning with \( H_3PO_4 \) solution for 50 min ensured the complete recovery of the maximum module performance (Figure 13, operation 4).

![Figure 12. The efficiency of cleaning the ceramic membrane after the MF of 1,3-PD fermentation broth with *Citrobacter freundii* (\( V_F = 5.5 \) m/s. TMP = 0.08 MPa). Operation time: water—30 min, NaOH—5 min, backwash—5 min, acid—10 min.](image)
The use of the developed membrane cleaning procedure allowed to maintain the steady-state relative permeate flux equal to 0.2 for repeated MF process (Figure 14). A slight decrease in the initial permeate flux was observed, which was reduced by increasing the cleaning temperature from 30 °C to 45 °C and extending the contact time with the cleaning solutions (NaOH and H3PO4) to 60 min (Figure 14, series S6 and S7).

![Figure 13](image1.png)

**Figure 13.** The efficiency of cleaning the ceramic membrane after the MF of 1,3-PD fermentation broth with *Citrobacter freundii* (*V*<sub>F</sub> = 11 m/s. *TMP* = 0.08 MPa). Operation time: water—30 min, NaOH—25 min, backwash—25 min, acid—50 min.

The use of the developed membrane cleaning procedure allowed to maintain the steady-state relative permeate flux equal to 0.2 for repeated MF process (Figure 14). A slight decrease in the initial permeate flux was observed, which was reduced by increasing the cleaning temperature from 30 °C to 45 °C and extending the contact time with the cleaning solutions (NaOH and H3PO4) to 60 min (Figure 14, series S6 and S7).

![Figure 14](image2.png)

**Figure 14.** Changes in relative permeate flux during the long-term MF process of 1,3-PD fermentation broth with *Citrobacter freundii*. Module M4, *TMP* = 0.02 MPa, *VF* = 5.5 m/s, CR—chemical cleaning.

Summarizing, results obtained in the present study demonstrated that ceramic membranes can be effectively cleaned by suitable cleaning agents with selected concentration and duration of action. In turn, the use of aggressive solutions could cause rapid degradation of the polypropylene membrane matrix.
3.3. Filtration of Fermentation Broths with Lactobacillus casei Bacteria

It is well known that the intensity of membrane fouling depends on the composition and properties of a feed solution. Therefore, for comparative purposes, in the next step of our research, the MF of 1,3-PD fermentation broth with Lactobacillus casei was investigated. This feed contained more protein components than the broth with Citrobacter freundii (Table 1). It has been found that during the MF process of broth with Lactobacillus casei the reported relative flux was equal to 0.2 (Figure 15, S1 and S2). Thus, it was 10 percentage points lower than that obtained during the MF of broth with Citrobacter freundii (Figure 7), performed under the same values of TMP and \( V_F \), equal to 0.08 MPa and 5.5 m/s, respectively. These results indicate that increasing the concentration of foulants, such as meat and yeast extracts as well as peptone K in a feed, led to more intensive membrane fouling.

![Figure 15. Changes in relative permeate flux during the long-term MF process of 1,3-PD fermentation broth with Lactobacillus casei with periodically cleaning the module. Module M4, TMP = 0.08 MPa, \( V_F = 5.5 \) m/s, CR—chemical cleaning.](image)

As expected, cleaning the M4 module with distilled water did not significantly affect the performance of the process and the relative permeate flux remained at the value equal to 0.2 (Figure 15, S1 and S2).

A much higher module performance, with the stabilization of the relative permeate flux of 0.25, was obtained as a result of cyclic cleaning of the module with NaOH and acid solutions (Figure 15, Series 3–5). However, despite an extensive cleaning procedure, the performance of the M4 module only increased to 0.83 of its maximum value (Figure 16).

A similarly significant decline in the MF process performance with using PP membranes has been noted (Figure 17). Although the new M2 module for this investigation was used, the relative permeate flux was stabilized at 0.05 (Figure 17, S1). Membrane cleaning with distilled water led to a slight increase of the flux, however, the continued process of broth clarification led to the decline in relative flux again to the value of 0.05 (Figure 17, S2). For the next cleaning, 1% NaOH solution for 30 min was used, which allowed the recovery of the initial membrane performance. As a consequence, a lower decrease in the permeate flux during the next series of the MF process was obtained (Figure 17, S3).
crease in the permeate flux during the next series of the MF process was obtained (Figure 17, S2). For the next cleaning, 1% NaOH solution for 30 min was used, which allowed the recovery of the initial membrane performance. As a consequence, a lower decrease in the permeate flux was observed. If distilled water was used, the relative permeate flux was stabilized at 0.05 (Figure 17, S1). Membrane fouling with polypropylene membranes used for the clarification of fermentation broths with Saccharomyces cerevisiae leads to a greater reduction of the filtration process performance.

Membranes has been noted (Figure 17). Although the new M2 module for this investigation was used, the relative permeate flux was stabilized at 0.05 (Figure 17, S1). Membrane fouling with polypropylene membranes used for the clarification of fermentation broths with Lactobacillus casei with periodically cleaning the module. Module M2, TMP = 0.05 MPa, V_F = 5.5 m/s, CR—chemical cleaning.

A series of MF experiments conducted allowed for comparison the performances of polypropylene membranes used for the clarification of fermentation broths with Citrobacter freundii (module M1, Figure 9) and Lactobacillus casei (module M2, Figure 17). It was found that although the more favorable operating conditions were used for the process with the M2 module, a decrease in the permeate flux was greater. This finding confirmed the above-mentioned result indicating that the broth composition plays a key role in the intensity of membrane fouling, and the increased concentration of protein components in the feed leads to a greater reduction of the filtration process performance.
3.4. Filtration of Yeast Suspensions

It is well known that the design of the module and the turbulence of the feed low affect the intensity of the membrane fouling phenomenon. Hence, in the present study, the M3 and M4 modules with the same design and similar membranes diameters (Table 2) were applied for clarification of Saccharomyces cerevisiae yeast suspensions in the pilot installation (Figure 1b). It ensured investigating the M3 and M4 modules performance under similar hydrodynamic conditions.

Although the high flow velocities (Re number = 12,000) for the process with using the M3 module have been applied, during the MF process of the solution containing 0.1% yeast, the relative permeate flux decreased to a value below 0.06 (Figure 18, S1). It was noticed that rinsing the module with distilled water caused a slight increase in the membrane performance, while the use of chemical cleaning agents (1% NaOH for 20 min and 0.5% H₃PO₄ for 20 min) led to an increase of the relative flux to 0.8. However, continuation of the MF process with the membrane cleaned by this method resulted in a greater decline in the performance, below 0.04 of the maximum permeate flux (Figure 18, S2). It is worth noting that, typically, yeast cells are 5–10 µm in diameter [87,88], hence, they are retained by the membranes used in our research. However, the prepared feed solution, apart from yeast cells, contained other substances, e.g., proteins, which formed a deposit layer on the surface of the membranes (Figure 19). The presence of additional components resulted from the fact that freeze-dried distiller’s yeast was used in the study.

![Figure 18. Changes in relative permeate flux during the long-term MF process of yeast suspension. Module M3, TMP = 0.08 MPa, V_p = 5.5 m/s.](image)

In order to reduce the intensity of membrane fouling, backwash (1 min) every 10 min was performed. As a result, a two-fold increase in MF process performance was obtained (Figure 18, S3–S5). The discontinuation of backwash resulted in a significant drop in the module performance (Figure 18, S6). Increasing the yeast content in the feed led to a decrease of the process performance, although for both 0.16% and 0.5% solutions the declines in permeate flux were similar (Figure 18, S7 and S8). This finding is in agreement with that presented by Hassan et al. [20] who indicated that during the MF of Saccharomyces cerevisiae yeast suspensions, the steady-state permeate flux is independent from the cell initial concentration in the range from 6 to 10 g/L.
Figure 19. SEM image of PP S6/2 membrane (module M3) surface covered by biological deposit with yeast cells after MF process of yeast suspensions.

To limit decrease in the flux, backwashing again was applied which resulted in a slight increase in the performance (Figure 18, S8–S10). Indeed, the observed increases were not as significant as for the series S3–S5, also for the solution containing 0.1% yeast (Figure 18, S10). It should be emphasized that after each chemical cleaning, the module performance was recovered, which was even greater than the initial performance (Figure 20, CR1–CR5). SEM studies showed that such an increase could be due to slight degradation of the membrane matrix. Likewise to the previous finding (Figure 11), it was determined that the membrane surface porosity increased significantly. Indeed, the surface pores of 2–5 µm were observed (Figure 21a). In this case, after starting the MF process, the pores quickly filled with filtered biomass, which also resulted in a rapid decline in the permeate flux. Indeed, the penetration of biomass into the pores was confirmed by the observations of the membrane cross-section (Figure 21b). Despite the use of cleaning the membrane with alkaline and acid solutions, deposits were still found inside the pores up to a depth of 2 µm.

Figure 20. Effect of chemical membrane cleaning on the maximum permeate flux. Module M3. CR—chemical rinsing.
Regarding the ceramic membrane used, its performance was also very low. The noted relative permeate flux, similar to the PP membrane, was equal to 0.045 (Figure 22, S2). Although ceramic membranes exhibit good chemical resistance, it must be recognized that the use of the same chemical agents as for PP membrane (NaOH and H$_3$PO$_4$) was not efficient and led to a decline in the module performance after the first cleaning cycle (Figure 23, 1–3). Hence, additional operations were required. Finally, the initial membrane performance by cleaning with the HNO$_3$ solution as a final step was recovered.

Cleaning of the ceramic membrane carried out after the second series of MF measurements (Figure 22 S2) showed that HNO$_3$ solution is not sufficient to effectively clean the membrane, since its double use allowed to obtain the relative permeate flux equal to 0.7 (Figure 24). On the other hand, the use of NaOH solution with hypochlorite allowed increasing the value to 0.8, while the complete recovery of the membrane permeability was obtained after repeated cleaning with NaOH and H$_3$PO$_4$ solutions.
Although ceramic membranes exhibit good chemical resistance, it must be recognized that the use of the same chemical agents as for PP membrane (NaOH and H$_3$PO$_4$) was not efficient and led to a decline in the module performance after the first cleaning cycle (Figure 23). Hence, additional operations were required. Finally, the initial membrane performance by cleaning with the HNO$_3$ solution as a final step was recovered.

To sum up, our investigations have shown that fermentation broth components from ethanol production led to the intense membranes fouling, the removal of which required multi-step cleaning. It has been indicated that ceramic membranes, contrary to PP membranes, can be cleaned with aggressive solutions without any damage to the membrane matrix.
4. Conclusions

This paper outlines a direct comparison of efficiency, performance and susceptibility to cleaning of the ceramic and polymeric MF membranes applied for clarification of 1,3-propanediol fermentation broths and suspensions of yeast *Saccharomyces cerevisiae*. For this purpose, ceramic membrane made of TiO$_2$ (hydrophilic) and polypropylene (hydrophobic) membranes, in different modules configurations (tubular and capillary), were tested. The MF experiments were carried out at a constant temperature of 30 °C and a wide range of process parameters. The transmembrane pressure ranging from 0.02 to 0.08 MPa was used. In turn, the applied feed flow velocity was from 0.8 to 11 m/s.

Both the ceramic and PP membranes have demonstrated a very high efficiency in purifying the biological suspensions. Although the number of bacteria in the feed was of an order of 12 log CFU, the membranes ensured sterile permeate during the filtration of 1,3-PD post-fermentation solutions. Although during the broths separation the turbidity of the feed increased to 8000 NTU, the turbidity of the obtained permeate was in the range of 1–3 NTU.

Moreover, the results obtained in the present study offer vital evidence that the permeate quality is directly correlated to the membrane pore size. Indeed, since the ceramic membrane had a nominal pore size (0.14 µm) smaller than the PP membranes (0.20 µm), it allowed obtaining the permeate characterized by a lower turbidity, equal to 0.2 NTU. It has been shown that all the membranes used were very susceptible to fouling, both by the components of 1,3-PD fermentation broths and yeast suspensions. The significant impact of the feed flow velocity and the composition of the fermentation broths on the relative permeate flux has been pointed out. It has been demonstrated that the membrane properties (hydrophobic/hydrophilic) did not significantly affect the intensity of the fouling phenomenon. The obtained data show that the decrease in process performance was mainly dependent to the properties of the filter cake formed on the membranes surface. For the ceramic membrane, internal fouling also had a significant influence on the permeate flux decline.

Moreover, much attention has been paid to the development of effective and comprehensive cleaning procedures for the membranes used. It has been demonstrated that suitable cleaning agents with selected concentration and duration of action effectively cleaned both the ceramic and PP membranes. However, the use of aggressive solutions led to degradation of the PP membranes matrix.

Findings of this study add to a growing body of literature on the use of ceramic and polypropylene MF membranes for the clarification of biological materials. However, it should always be taken into account that the fermentation broths are very complex medium, the composition of which strictly depends on the carbon source, bacteria strain and fermentation conditions as well as several other factors. Therefore, the results of the presented work give an overall view of the MF process efficiency for biological suspensions, however, all the characteristic dependencies should be examined individually.

**Author Contributions:** Conceptualization, W.T. and M.G.; methodology, W.T. and M.G.; formal analysis, W.T.; investigation, W.T.; resources, M.G.; data curation, W.T.; writing—original draft preparation, W.T.; writing—review and editing, M.G.; visualization, W.T.; supervision, M.G.; project administration, M.G.; funding acquisition, M.G. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by the framework of the Project: Biotechnological Conversion of Glycerol to Polyols and Dicarboxylic Acids; (No. 01.01.02-00-074/09) co-funded by the European Union from the European Regional Development Funds within the framework of the Innovative Economy Operational Programme 2007–2013.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.
Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

Abbreviations

CA  cellulose acetate
CR  chemical cleaning (rinsing)
J  actual permeate flux
J₀  maximum permeate flux
MF  microfiltration
NTU  nephelometric turbidity units
PAN  polyacrylonitrile
PAEK  polyaryletherketone
PES  polyethersulfone
PI  polyamide
PP  polypropylene
PTT  polytrimethylene terephthalate
PVDF  polyvinylidenefluoride
PVP  polyvinylpyrrolidone
NF  nanofiltration
SEM  scanning electron microscopy
TMP  transmembrane pressure
UF  ultrafiltration
Vₑ  feed flow velocity
1,3-PD  1,3-propanediol

References

1. Belfort, G.; Davis, R.H.; Zydney, A.L. The Behavior of Suspensions and Macromolecular Solutions in Crossflow Microfiltration. *J. Membr. Sci.* 1994, 96, 1–58. [CrossRef]
2. Chew, J.W.; Kilduff, J.; Belfort, G. The Behavior of Suspensions and Macromolecular Solutions in Crossflow Microfiltration: An Update. *J. Membr. Sci.* 2020, 601, 117865. [CrossRef]
3. Białas, W.; Celińśka, E.; Dembczyński, R.; Szymanowska, D.; Nowacka, M.; Jesionowski, T.; Grajek, W. Cross-Flow Microfiltration of Fermentation Broth Containing Native Corn Starch. *J. Membr. Sci.* 2013, 427, 118–128. [CrossRef]
4. Kujundzic, E.; Greenberg, A.R.; Fong, R.; Moore, B.; Kujundzic, D.; Hernández, M. Biofouling Potential of Industrial Fermentation Broth Components during Microfiltration. *J. Membr. Sci.* 2010, 349, 44–55. [CrossRef]
5. Tomczak, W.; Gryta, M. Cross-Flow Microfiltration of Glycerol Fermentation Broths with Citrobacter Freundii. *Membranes* 2020, 10, 67. [CrossRef] [PubMed]
6. Thuy, N.T.H.; Boontawan, A. Production of Very-High Purity Succinic Acid from Fermentation Broth Using Microfiltration and Nanofiltration-Assisted Crystallization. *J. Membr. Sci.* 2017, 524, 470–481. [CrossRef]
7. Gryta, M.; Tomczak, W. Microfiltration of Post-Fermentation Broth with Backflushing Membrane Cleaning. *Chem. Pap.* 2015, 69. [CrossRef]
8. Yang, X.; Zhou, S.; Li, M.; Wang, R.; Zhao, Y. Purification of Cellulase Fermentation Broth via Low Cost Ceramic Microfiltration Membranes with Nanofibers-like Attapulgite Separation Layers. *Sep. Purif. Technol.* 2017, 175, 435–442. [CrossRef]
9. Jokić, A.; Pačin, I.; Grahovac, J.; Lukić, N.; Dodić, J.; Rončević, Z.; Šereš, Z. Improving Energy Efficiency of *Bacillus Velezensis* Broth Microfiltration in Tubular Ceramic Membrane by Air Sparging and Turbulence Promoter. *J. Chem. Technol. Biotechnol.* 2019. [CrossRef]
10. Marzban, R.; Saberi, F.; Shirazi, M.M.A. Microfiltration and Ultrafiltration of Bacillus Thuringiensis Fermentation Broth: Membrane Performance and Sporecrystal Recovery Approach. *Braz. J. Chem. Eng.* 2016, 33, 783–791. [CrossRef]
11. Sikder, J.; Pereira, C.; Palchoudhury, S.; Vohra, K.; Basumatary, D.; Pal, P. Synthesis and Characterization of Cellulose Acetate-Polysulfone Blend Microfiltration Membrane for Separation of Microbial Cells from Lactic Acid Fermentation Broth. *Desalination* 2009, 249, 802–808. [CrossRef]
12. Carrère, H.; Blaszczuk, F. Comparison of Operating Modes for Clarifying Lactic Acid Fermentation Broths by Batch Cross-Flow Microfiltration. *Process Biochem.* 2001, 36, 751–756. [CrossRef]
13. Davies, J.L.; Baganz, F.; Ison, A.P.; Lye, G.J. Studies on the Interaction of Fermentation and Microfiltration Operations: Erythromycin Recovery From Saccharopolyspora Erythraea Fermentation Broths. *Biotechnol. Bioeng.* 2000, 69, 429–439. [CrossRef]
14. Milcent, S.; Carrère, H. Clarification of Lactic Acid Fermentation Broths. *Sep. Purif. Technol.* 2001, 22–23, 393–401. [CrossRef]
15. Persson, A.; Jönsson, A.-S.; Zacchi, G. Separation of Lactic Acid-Producing Bacteria from Fermentation Broth Using a Ceramic Microfiltration Membrane with Constant Permeate Flow. *Biotechnol. Bioeng.* 2001, 72, 269–277. [CrossRef]

16. Persson, A.; Jönsson, A.-S.; Zacchi, G. Use of Microfiltration as First Step in Recovery of Protein a from Fermentation Broth. *Appl. Biochem. Biotechnol.* 2004, 112, 151–162. [CrossRef]

17. Toussaint, G.; Ding, L.H.; Jaffrin, M.Y.; Hassairi, I.; Nonus, M. Recovery of α-Agarase Enzyme from Fermentation Broths by Membrane Crossflow Filtration. *Sep. Sci. Technol.* 2000, 35, 795–809. [CrossRef]

18. Alexandri, M.; Schneider, R.; Venus, J. Membrane Technologies for Lactic Acid Separation from Fermentation Broths Derived from Renewable Resources. *Membranes* 2018, 8, 94. [CrossRef]

19. Gençal, Y.; Durmaz, E.N.; Çulfaz-Emecen, P.Z. Preparation of Patterned Microfiltration Membranes and Their Performance in Crossflow Yeast Filtration. *J. Membr. Sci.* 2015, 476, 224–233. [CrossRef]

20. Hassan, I.; Ennouri, M.; Lafforgue, C.; Schnitz, P.; Ayadi, A. Experimental Study of Membrane Fouling during Crossflow Microfiltration of Yeast and Bacteria Suspensions: Towards an Analysis at the Microscopic Level. *Membranes* 2013, 3, 44–68. [CrossRef]

21. Jokić, A.; Zavargo, Z.; Šereš, Z.; Tekić, M. The Effect of Turbulence Promoter on Cross-Flow Microfiltration of Yeast Suspensions: A Response Surface Methodology Approach. *J. Membr. Sci.* 2010, 350, 269–278. [CrossRef]

22. Lewis, W.J.T.; Chew, Y.M.J.; Bird, M.R. The Application of Fluid Dynamic Gauging in Characterising Cake Deposition during the Cross-Flow Microfiltration of a Yeast Suspension. *J. Membr. Sci.* 2012, 405–406, 113–122. [CrossRef]

23. Liu, D.; Savoire, R.; Vorobiev, E.; Lanoisellé, J.-L. Effect of Disruption Methods on the Dead-End Microfiltration Behavior of Yeast Suspension. *Sep. Sci. Technol.* 2010, 45, 1042–1050. [CrossRef]

24. Keskinler, B.; Yildiz, E.; Erhan, E.; Dogru, M.; Bayhan, Y.; Akay, G. Crossflow Microfiltration of Low Concentration-Nonliving Yeast Suspensions. *J. Membr. Sci.* 2004, 233, 59–69. [CrossRef]

25. Gabrus, E.; Szaniaw ska, D. Application of Backflushing for Fouling Reduction during Microfiltration of Yeast Suspensions. *Desalination* 2009, 240, 46–53. [CrossRef]

26. Stopka, J.; Bugan, S.; Broussous, L.; Schlosser, Š.; Larbot, A. Microfiltration of Beer Yeast Suspensions through Stamped Ceramic Membranes. *Sep. Purif. Technol.* 2001, 25, 535–543. [CrossRef]

27. Mores, W.D.; Davis, R.H. Yeast Fouulant Removal by Backpulses in Crossflow Microfiltration. *J. Membr. Sci.* 2002, 208, 389–404. [CrossRef]

28. Mores, W. Direct Visual Observation of Yeast Deposition and Removal during Microfiltration. *J. Membr. Sci.* 2001, 187, 219–230. [CrossRef]

29. Mahesh Kumar, S.; Roy, S. Filtration Characteristics in Dead-End Microfiltration of Living Saccharomyces Cerevisiae Cells by Alumina Membranes. *Desalination* 2008, 229, 348–361. [CrossRef]

30. Keskinler, B.; Akay, G.; Bayhan, Y.; Erhan, E. Effect of Ionic Environment on the Crossflow Microfiltration Behaviour of Yeast Suspensions. *J. Membr. Sci.* 2002, 206, 351–360. [CrossRef]

31. Coulson, J.M.; Richardson, J.F. *Coulson & Richardson’s Chemical Engineering*; Butterworth-Heinemann: Boston, MA, USA; ISBN 9780750644457.

32. Manni, A.; Achiou, B.; Karim, A.; Harrati, A.; Sadik, C.; Ouammou, M.; Alami Younssi, S.; El Bouari, A. New Low-Cost Ceramic Microfiltration Membrane Made from Natural Magnesite for Industrial Wastewater Treatment. *J. Environ. Chem. Eng.* 2020, 8, 103906. [CrossRef]

33. Available online: https://www.bccresearch.com/market-research/membrane-and-separation-technology/membrane-microfiltration.

34. Speed, D.E. Environmental aspects of planarization processes. In *Advances in chemical mechanical planarization (CMP)*; Elsevier: Amsterdam, The Netherlands, 2016; pp. 229–269. ISBN 978-0-08-100165-3.

35. Warsinger, D.M.; Chakraborty, S.; Tow, E.W.; Plumlee, M.H.; Bellona, C.; Loutatidou, S.; Karimi, L.; Mikelonis, A.M.; Achilli, A.; Ghassemi, A.; et al. A Review of Polymeric Membranes and Processes for Potable Water Reuse. *Prog. Polym. Sci.* 2018, 81, 209–237. [CrossRef] [PubMed]

36. Li, C.; Sun, W.; Lu, Z.; Ao, X.; Li, S. Ceramic Nanocomposite Membranes and Membrane Fouling: A Review. *Water Res.* 2020, 175, 115674. [CrossRef] [PubMed]

37. Gaspar, I.; Koris, A.; Bertalan, Z.; Vatai, G. Comparison of Ceramic Capillary Membrane and Ceramic Tubular Membrane with Inserted Static Mixer. *Chem. Pap.* 2011, 65. [CrossRef]

38. Kumar, C.M.; Roshni, M.; Vasanth, D. Treatment of Aqueous Bacterial Solution Using Ceramic Membrane Prepared from Cheaper Clays: A Detailed Investigation of Fouling and Cleaning. *J. Water Process Eng.* 2019, 29, 100797. [CrossRef]

39. Hubadillah, S.K.; Othman, M.H.D.; Matsuura, T.; Ismail, A.F.; Rahman, M.A.; Harun, Z.; Jaafar, J.; Nomura, M. Fabrications and Applications of Low Cost Ceramic Membrane from Kaolin: A Comprehensive Review. *Ceram. Int.* 2018, 44, 4538–4560. [CrossRef]

40. Baruah, G.L.; Nayak, A.; Belfort, G. Scale-up from Laboratory Microfiltration to a Ceramic Pilot Plant: Design and Performance. *J. Membr. Sci.* 2006, 274, 56–63. [CrossRef]
71. Kluge, M.; Pérocheau Arnaud, S.; Robert, T. 1,3-Propanediol and Its Application in Bio-Based Polyesters for Resin Applications. *Chem. Africa* **2019**, *2*, 215–221. [CrossRef]

72. Gryta, M.; Markowska-Szczupak, A.; Bastrzyk, J.; Tomczak, W. The Study of Membrane Distillation Used for Separation of Fermenting Glycerol Solutions. *J. Membr. Sci.* **2013**, *431*, 1–8. [CrossRef]

73. Bastrzyk, J.; Gryta, M.; Karakulski, K. Fouling of Nanofiltration Membranes Used for Separation of Fermented Glycerol Solutions. *Chem. Pap.* **2014**, *68*. [CrossRef]

74. Kavšček, M.; Stražar, M.; Curk, T.; Petrovič, U. Yeast as a Cell Factory: Current State and Perspectives. *Microb. Cell Factories* **2015**, *14*. [CrossRef]

75. Tomczak, W.; Gryta, M. The Application of Ultrafiltration for Separation of Glycerol Solution Fermented by Bacteria. *Pol. J. Chem. Technol.* **2013**, *15*, 115–120. [CrossRef]

76. Gryta, M.; Waszak, M.; Tomaszewsk, M. Studies of Polypropylene Membrane Fouling during Microfiltration of Broth with Citrobacter Freundii Bacteria. *Pol. J. Chem. Technol.* **2015**, *17*, 56–64. [CrossRef]

77. Nady, N.; Franssen, M.C.R.; Zuilhof, H.; Eldin, M.S.M.; Boom, R.; Schroën, K. Modification Methods for Poly(Arylsulfone) Membranes: A Mini-Review Focusing on Surface Modification. *Desalination* **2011**, *275*, 1–9. [CrossRef]

78. Wang, Z.; Ma, J.; Tang, C.Y.; Kimura, K.; Wang, Q.; Han, X. Membrane Cleaning in Membrane Bioreactors: A Review. *J. Membr. Sci.* **2014**, *468*, 276–307. [CrossRef]

79. Someya, M.; Higashino, K.; Imoto, Y.; Sakanakura, H.; Yasutaka, T. Effects of Membrane Filter Material and Pore Size on Turbidity and Hazardous Element Concentrations in Soil Batch Leaching Tests. *Chemosphere* **2020**, *128981*. [CrossRef]

80. Tomczak, W.; Gryta, M. Clarification of 1,3-Propanediol Fermentation Broths by Using a Ceramic Fine UF Membrane. *Membranes* **2020**, *10*, 319. [CrossRef] [PubMed]

81. Silva, J.R.; Cardoso, G.; Maciel, R.R.G.; de Souza, N.C. Morphological Alterations on Citrobacter Freundii Bacteria Induced by Erythrosine Dye and Laser Light. *Lasers Med. Sci.* **2015**, *30*, 469–473. [CrossRef]

82. Choi, H.; Zhang, K.; Dionysiou, D.D.; Oerther, D.B.; Sorial, G.A. Influence of Cross-Flow Velocity on Membrane Performance during Filtration of Biological Suspension. *J. Membr. Sci.* **2005**, *248*, 189–199. [CrossRef]

83. Jana, A.; Ghosh, S.; Majumdar, S. Energy Efficient Harvesting of *Arthrospira* Sp. Using Ceramic Membranes: Analyzing the Effect of Membrane Pore Size and Incorporation of Flocculant as Fouling Control Strategy. *J. Chem. Technol. Biotechnol.* **2018**, *93*, 1085–1096. [CrossRef]

84. Streit, F.; Athès, V.; Ghir, A.; Corrieu, G.; Béal, C. Microfiltration Conditions Modify Lactobacillus Bulgaricus Cryotolerance in Response to Physiological Changes. *Bioproc. Biosyst. Eng.* **2011**, *34*, 197–204. [CrossRef]

85. Mora, F.; Pérez, K.; Quezada, C.; Herrera, C.; Cassano, A.; Ruby-Figueroa, R. Impact of Membrane Pore Size on the Clarification Performance of Grape Marc Extract by Microfiltration. *Membranes* **2019**, *9*, 146. [CrossRef] [PubMed]

86. Cui, Z.; Peng, W.; Fan, Y.; Xing, W.; Xu, N. Ceramic Membrane Filtration as Seawater RO Pre-Treatment: Influencing Factors on the Ceramic Membrane Flux and Quality. *Desalin. Water Treat.* **2013**, *51*, 2575–2583. [CrossRef]

87. Nguyen, K.; Murray, S.; Lewis, J.A.; Kumar, P. Morphology, Cell Division, and Viability of Saccharomyces Cerevisiae at High Hydrostatic Pressure. Available online: https://arxiv.org/abs/1703.00547 (accessed on 7 January 2021).

88. Walker, G.M. *Yeast Physiology and Biotechnology*; J. Wiley & Sons: New York, NY, USA, 1998; ISBN 978-0-471-96447-6.