Bovine Tropical Theileriosis (BTT), a tick borne lymphoproliferative disease of ruminants, caused by apicomplex parasite *Theileria annulata* inflicts significant impact on health and productivity of dairy animals (Pandey et al. 2017). In India, BTT accounts for loss of US$ 384.3 million annually (Minjauw and McLeod 2003). Calf theileriosis causes heavy mortality along with specific clinical manifestations (Singh et al. 2014) including exophthalmia (Sudan et al. 2012, Singh et al. 2015). *Theileria* piroplasms are difficult to find in stained blood smears especially in cases of chronic carriers (Jaiswal et al. 2016). Therefore, molecular detection based on DNA probes remains an important tool for sensitive diagnosis of chronic carrier states. The present study was designed with the objective of standardization and validation of a PCR assay for sensitive and specific detection of theileriosis in calves.

Collection of blood samples: Blood samples (1 ml aliquot from each animal) were collected in clean sterile vacutainer, containing ethylene diamine tetra acetic acid (EDTA), from the jugular veins of earlier theileriosis confirmed cattle (microscopic observation of blood smears) for isolation of positive controls and laboratory standardization of PCR assay. Later on, blood samples were collected from 71 cattle calves from various goshalas of Mathura-Vrindavan area alongside those which were brought to TVCC, DUVASU, Mathura. Peripheral blood smears were also made using standard procedures.

DNA isolation, primers selection and polymerase chain reaction (PCR): DNA was isolated using standard blood isolation kit (Puregene) following manufacturer’s protocol. Primers TAMS F/R were custom synthesized using sequence published elsewhere (Sudan et al. 2016). This pair of primers was used for the synthesis of the primary PCR amplification product of the gene encoding the 30-kDa major *T. annulata* merozoite surface antigen. Details for primers design including position of nucleotides, nucleotide sequences, and expected PCR products are shown in Table 1. The PCR reaction was set up into 25 µl volume containing 12.5 µl PCR Master Mix (0.05/µl Taq DNA polymerase in reaction buffer, 4 mM MgCl₂, 0.4 mM dATP, 0.4 mM dCTP, 0.4 mM dGTP, and 0.4 mM dTTP), 1.5 µl of each primer (TAMS F/R), 2 µl of the DNA template and total volume was made up to 25 µl using nuclease free water. The details of thermal cycling profiles are given in Table 1. The PCR amplicons were analyzed by agarose gel electrophoresis in 1.5 % agarose gel.

Sensitivity and specificity assay: Serial dilutions of the positive DNA was performed in order to obtain regular scale of dilution from 1/100 to 1/200,000. In order to check the specificity of these primers, PCR on *Trypanosoma evansi* and Babesia sp. using standard primers was also performed (Parashar et al. 2015, Sudans et al. 2017).

Statistical analysis: The results of the PCR on blood were compared with that of blood smear examination. The sensitivity and specificity were calculated and compared with that of blood smear examination (kept as gold standard) using online software (http://graphpad.com/quickcalcs/kapp1.cfm).

The PCR produced a 721 bp PCR product from all the positive samples which is highly specific for *T. annulata*. The serial dilution results revealed a very good response. The DNA from other blood parasites failed to produce any of the amplification products accounting for its high specificity.

When compared with blood smear examination, PCR on blood detected 12 positive cases in comparison to blood smear examination, which detected 8 positive cases. (Table 2). Keeping blood smear examination as a gold standard for detecting actual number of confirmed positive cases, PCR on blood was found to be 100% sensitive and 93.65% specific based on their kappa value predictions.

Under the field conditions, the animals having subclinical infection are the major source of infection to the ticks. The subclinical infection cannot be easily diagnosed on routine blood smear examination whereas PCR detects very minute infection of *T. annulata* and can be used as an excellent tool for the diagnosis of BTT. Discrimination of *T. annulata* from non-pathogenic *Theileria* species and other hemoparasites that may occur simultaneously in the same carrier animal is feasible using PCR (d’Oliveira et al. 1995).
optimum and accurate diagnosis using PCR. The primers used in the present study were derived from the gene encoding the 30-kDa major merozoite surface antigen (Sudan et al. 2016). These primers are considered to be the most ideal primers for the specific detection of Theileria annulata. The sensitivity and specificity of PCR was compared with blood microscopy based on kappa value predictions. A total of 8 samples were found positive by blood smear examination whereas PCR detected 12 infections. Blood smear examination was kept as a gold standard for detecting actual number of confirmed positive cases, for being 100% sensitive along with 93.65% specific, respectively, in detecting calf theileriosis. The described PCR-based assay provides a valuable tool to study the epidemiology of BTT in calves and some vital data regarding epidemiology of theileriosis in calves from semi-arid parts of India has been generated. Such a record for screening of calves for theileriosis is missing from Indian context.

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