Current Proposed Mechanisms of Action of Intravenous Immunoglobulins in Inflammatory Neuropathies

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Abstract: Intravenous immunoglobulins (IVIg) have been shown in a number of trials, to be an effective treatment for the three main types of inflammatory neuropathies: Guillain-Barré Syndrome (GBS), chronic inflammatory demyelinating polyneuropathy (CIDP), and multifocal motor neuropathy (MMN). IVIg is thought to exert its immunomodulatory effects by affecting several components of the immune system including B-cells, T-cells, macrophages, complement, cytokines and cellular adhesion molecules. This article reviews the published evidence and the principal postulated mechanisms of action of intravenous immunoglobulins with special emphasis on inflammatory neuropathies.

Key Words: Intravenous immunoglobulins, mechanisms of action, inflammatory neuropathy, Guillain Barre syndrome, Chronic inflammatory demyelinating polyneuropathy, multifocal motor neuropathy.

INTRODUCTION

Intravenous immunoglobulins (IVIg) have been shown to be effective in a number of disorders with an underlying autoimmune basis. Amongst those, the inflammatory neuropathies, Guillain-Barré Syndrome (GBS), chronic inflammatory demyelinating polyneuropathy (CIDP), and multifocal motor neuropathy (MMN), have all been found in well-conducted randomised controlled trials (RCTs) to benefit from IVIg.

GBS is an acute polyradiculoneuropathy which could be demyelinating (as in the “AIDP”, i.e. “Acute Inflammatory Demyelinating Polyneuropathy” variant) or axonal (as in the “AMAN”, i.e. “Acute Motor Axonal Neuropathy” variant). Its incidence is estimated at 1 to 2 per 100,000. Respiratory involvement is common, and mortality is estimated at about 10%. Proximal and distal symmetric weakness with sensory symptoms but few sensory signs occur in the majority cases. Facial and bulbar weakness is not uncommon. Diagnosis is clinical but also relies on electrophysiology which may show signs of demyelination, and spinal fluid analysis typically shows a raised protein level with normal cellularity. The syndrome occurs in a post-infectious context in 75% of cases and probably results from cross-reactivity with the infectious agent and peripheral nerve sharing common epitopes to which autoantibody attack is directed. The most common infectious agent resulting in GBS is Campylobacter jejuni. Plasma exchanges were shown to be more effective than placebo for GBS in RCTs [50]. IVIg was subsequently found to be of equivalent effectiveness to plasma exchange [27] and has become standard treatment for GBS because of ease of administration and a comparatively better side-effect profile.

CIDP is an acquired heterogeneous disorder affecting sensory and motor peripheral nerves caused by a patchy demyelinating process, producing sensory loss and positive sensory symptoms as well as motor weakness. Its prevalence is of 3 to 5 per 100,000 and incidence of about 0.50 per 100,000/year. In its typical form, the disorder is symmetric and involves both proximal and distal limb regions. There are rarer atypical forms which can produce predominantly uni- or multifocal as well as distal involvement. Diagnosis relies on clinical features, and also mainly on electrophysiology, which allows demonstration of a demyelinating process, producing slowing of nerve conductions in various segments as well as conduction block. Cerebrospinal fluid protein level is raised in the majority of cases, and peripheral nerve histology, may show a demyelinating process with inflammatory features. Magnetic resonance imaging (MRI) can show thickened hyperintense nerve roots, trunks or plexi, consistent with the diagnosis. IVIg as well as steroids and plasma exchanges have been shown to be effective treatments for CIDP. The pathogenesis of CIDP is inflammatory, most probably autoimmune, involving both T cells and antibodies. The efficacy of IVIg in CIDP has been shown in different studies [26, 42].

MMN is a rare disorder with an estimated prevalence of 1 to 2 per 100,000 frequently occurring in middle-aged men. It predominantly affects the upper limbs with asymmetric weakness occurring in individual nerve distributions. There is typically no sensory involvement. The diagnosis is clinical and confirmed in the majority of cases by electrophysiology which shows evidence of motor conduction block. Anti-GM1 antibody positivity can be present in 30 to 80% of cases. IVIg is the only treatment so far, found effective in MMN [4, 19, 36, 66].

HISTORY OF USE OF INTRAVENOUS IMMUNOGLOBULIN

The two main components of the adaptive immunity are B-cells (which produce immunoglobulins) and T-cells (re-
sponsible for cell-mediated immunity). Immune deficiencies occurring in either of the components have been shown to increase susceptibility to bacterial, viral or fungal infections. Antibody therapy had been recognised as a useful tool since the late 19th century when von Behring described antibodies against diphtheria. Replacement of immunoglobulins, either subcutaneously or intravenously, has been described as an effective therapy for primary immunodeficiencies for almost 50 years [9, 10]. In 1981 it was observed that intravenous immunoglobulin (IVIg) therapy in children with hypogammaglobulinemia and coincidental idiopathic thrombocytopenic purpura, led to a reproducible increase in platelet count [29]. This led way to the use of IVIg in several other diseases with a confirmed or suspected autoimmune aetiology.

Multiple sclerosis was one of the first neurological disorders in which IVIg was used [56], although it is not currently recommended by many guidelines. Since then, several reports of the successful use of IVIg have been published in several neuromuscular diseases including myasthenia gravis (MG) [1, 16, 18], CIDP [72], Gullain-Barre syndrome [2, 32] and multifocal motor neuropathy [46].

Currently, the use of IVIg can be classified broadly into low dose replacement therapy (usually 0.3-0.5g/kg every 3-4 weeks) or high-dose therapy for the anti-inflammatory action (usually 1-3g/kg). The comparison of IVIg dosing of 1.2g/kg over 3 days or 2.4 g/kg over 6 days in GBS has shown a better outcome for the six-day regimen [49]. However in CIDP lower initiating and maintenance doses may be sufficient [48], although large scale randomised trials looking into the exact dosing requirement are lacking. The half-life of IgG in the circulation is approximately 4 weeks, thereby requiring repeated courses every 8-12 weeks.

MECHANISMS OF ACTION OF IVIg

Although the presence of "natural" antibodies capable of recognising foreign antigens could plausibly explain the role of IVIg in IgG replacement therapy, the precise mechanism of action by which IVIg exerts its immunomodulatory effects is not clearly understood. In inflammatory neuropathies there are several proposed pathophysiological mechanisms and a detailed review of these is beyond the scope of this article and has been dealt with elsewhere [25, 65, 67]. Pathological studies of nerve biopsies in CIDP and GBS reveal lymphocytic and macrophagic infiltrates in the endoneurium with deposits of IgM and complement components. The role of B-cells is clearly established in GBS where anti-ganglioside antibodies and complement activation have been demonstrated [67]. Lymphocytic infiltrates are predominantly T-cells recruited by chemokines and endothelial cell adhesion molecules. T-cells secrete matrix metalloproteases which break down endoneurial proteins. Macrophages are the predominant antigen presenting cells as demonstrated by increased expression of NFκB and the inflammatory cytokines, IL-6 and IL-1β. All these mechanisms are potentially modulated by IVIg and will be discussed in detail below. Paradoxically, plasma exchange which works theoretically opposite to IVIg by removing IgG from the body, seem to work in similar clinical situations.

1. Effect of IVIg on B-Cells and Antibodies

B-cells, which form 5-15% of the circulating lymphoid pool, are responsible for humoral immunity and acts against extracellular pathogens. B-cells are activated in response to a variety of stimuli and differentiate to form plasma cells. Plasma cells are usually restricted to the secondary lymphoid organs, comprising less than 0.1% of the lymphocytes in circulation. Autoreactive B-cells may be stimulated by either autoantigens or through non-specific polyclonal activation. Soluble immunoglobulins, produced by the plasma cells against autoantigens, are responsible for the majority of clinical features in antibody mediated autoimmune diseases.

The use of IVIg in acquired haemophilia has shown that the presence of anti-idiotype antibodies against the anti-haemophilic factor (factor VIII antibodies) leads to a therapeutic suppression of these autoantibodies [53, 61]. Anti-idiotypes against autoantibodies to thyroglobulin [17], ANCA [51], acetylcholine receptor [77], DNA [58], platelet glycoprotein IIb/IIIa [41], beta-2 glycoprotein-I [7] and intrinsic factor [52] have been shown to be present in IVIg preparations. The presence of anti-idiotypes prevents the binding of pathogenic autoantibodies to the target epitopes, thereby ameliorating the autoimmune symptoms. Anti-idiotype antibodies are likely to be involved in the therapeutic effects of IVIg in GBS and MMN, where antibodies against different gangliosides have been described.

Other B-cell mediated effects of IVIg include inhibition of antibody production [33], inhibition of B-cell differentiation [60], inhibition of production of interleukin-6 and tumour necrosis factor-α [63], induction of B-cell apoptosis [64], down-regulation of specific auto-reactive B-cells [70] and regulation of B-cell subsets expressing CD5 [69], thereby suppressing the auto-antibody producing CD20+ B1 cells. Even though the serum anti-GM1 antibody levels in seropositive MMN patients do not fall after IVIg infusion, the Fab portion of Ig in the IVIg has been shown to inhibit binding of anti-GM1 Ab to target antigens [39]. Although the efficacy of IVIg in IgM paraproteineic neuropathies is not well established, the levels of IgM myelin-associated glycoprotein (MAG) and sulfoglucuronyl paragloboside (SGPG) antibodies are reduced in occasional patients treated with this therapy [15].

FcRn (so named because it was initially identified in neonatal intestinal epithelium) is a protective receptor crucial for regulating the half life of IgG. In normal circumstances, IgG binds to FcRn and is protected from catabolism after IgG infusion, the Fab portion of Ig in the IVIg has been shown to inhibit binding of anti-GM1 Ab to target antigens [39]. Although the efficacy of IVIg in IgM paraproteineic neuropathies is not well established, the levels of IgM myelin-associated glycoprotein (MAG) and sulfoglucuronyl paragloboside (SGPG) antibodies are reduced in occasional patients treated with this therapy [15].

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2. Effect of IVIg on T-Cells

T-cells mediate the cell mediated immunity and are involved in the handling of intracellular pathogens, but also play a major role in the regulation of B-cell responses. This is mediated by the two sub-population of CD4+ T-cells or T-helper cells (Th cells): Th1 cells secreting interleukin-2 (IL-
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3. Effect of IVIg on the Complement System

The heat-labile component of serum that augmented its bactericidal properties is one of the main pathogenic pathways involved in antibody-mediated autoimmune diseases. The formation of immune complexes activate the classical complement cascade resulting in the production of membrane attack complexes (MAC) which are thought to induce organ specific tissue damage in a variety of autoimmune diseases, like myasthenia gravis, lupus nephritis and GBS.

The anti-inflammatory activity of IVIg is at least partly mediated by its ability to prevent the formation of MAC and subsequent tissue destruction. Antibodies against several components of the classical complement pathway have been identified in IVIg. They include antibodies against C1 [43], C3a [6], C3b or C4 [20]. In addition, high doses of IVIg are thought to enhance the degradation of C3b [38]. In-vitro increase of complement uptake has been demonstrated in GBS and MG [5] and complement is thought to be important in GBS and its variant, Miller-Fisher syndrome [22, 23, 74], in which complement therapies are under investigation [22, 23].

4. IVIg Mediated Fc Receptor Blockade on Macrophages

FcγR receptors on the surface of macrophages can mediate inflammatory pathways by activating (FcγRI or FcγRIIa) or inhibiting (FcγRII) different receptors [31]. IVIg may inhibit the FcγRI or FcγRII receptors or upregulate the FcγRII receptors [14]. In idiopathic thrombocytopenic purpura (ITP), IVIg is thought to inhibit platelet phagocytosis through the FcγRII receptor [54]. In GBS and CIDP, inhibition of macrophage function reduces the phagocytosis of antigen-presenting cells and antibody-mediated cellular cytotoxicity, thus inhibiting macrophage-mediated demyelination [13]. Similarly, an increase in ratio of FcγRII/FcγRII receptors on monocytes has been demonstrated a week after IVIg administration in patients with CIDP and MMN who were beginning to improve [11].

5. Effect of IVIg on Cytokines

Cytokines are proteins or glycoproteins involved in the signalling process during a variety of immune reactions. Dysregulation of the cytokine system has been proposed as one of the mechanisms of autoimmunity.

IVIg reduces the levels of circulating IL-1β in patients with GBS [57] and Kawasaki disease [37]. Thousand fold increase in the levels of IL-1 receptor antagonist have been shown after IVIg therapy [3]. However, cytokine modulation is unlikely to be the major mechanism of action of immunoglobulins, since IVIg remains functionally active in mice strains deficient in IL-1R, IL-4, IL-10, IFN-γR, IL-12β and TNF-α [12]. TNF-α mediated cytotoxicity is also inhibited by IVIg [59].

6. Effect of IVIg in Modulating Cell Migration

Leukocyte migration across biological barriers has been suggested to be an important mechanism in the causation of organ specific autoimmune diseases. IVIg is thought to modulate endothelial cell function by interacting with intracellular adhesion molecules (ICAM) [75]. A significant reduction in expression of ICAM-1 was seen in 8 out of 10 patients with MMN and CIDP, during the first week after infusion of IVIg [11].

Other possible mechanisms by which IVIg modulate cell migration include the presence of antibodies against integrins [34] and the arginine-glycine-asparagine (RGD) cell adhesion motifs [71]. Inhibition of the chemokine (C-C motif) receptor-5 (CCR-5) by IVIg prevents the entry of HIV into its target cells [8].

7. Effect of IVIg on Superantigens

Superantigens like bacterial enterotoxins and viruses stimulate the Vβ chains of the T-cell receptor triggering the production of cytokines and breaking immune tolerance. The role of IVIG against the β-chains of the T-cell receptor [40] may thus be relevant in the influence of relapses triggered by infections in MG and CIDP [14].

8. Other Mechanisms of Action of IVIg

In toxic epidermal necrolysis, apoptosis of keratinocytes is prevented by blocking CD95, the Fas death receptor [73].

There is considerable variation in the sugar moieties attached to the asparagine 297 (N297) residue of the Fe portion of IgG. It has been suggested that sialic acid rich IgG levels are reduced in acute phases of several autoimmune disease models. Infusion of IVIg, which is pooled from several donors, may restore the levels of sialic acid-rich IgG, thus inducing an anti-inflammatory action, possibly through novel receptors in regulatory macrophages. [30, 44, 45].

In addition to the above mentioned immune mediated effects, whether IVIg has a direct effect on remyelination [68] in diseases like GBS and CIDP is not very clearly understood.

MECHANISMS OF SIDE-EFFECTS PRODUCED BY IVIg

Generally the side-effects of IVIg are usually minor occurring in less than 1 in 10 patients. Myalgia, chills or chest pain may occur in the first hour of infusion, which can be reversed by temporarily stopping the infusion and restarting.
at a slower rate. Fatigue, nausea and headache may occur for up to 24 hours. The exact pathogenesis of these side-effects are not clearly understood, but activation of the complement pathway by aggregated immunoglobulins may be partly responsible. This may also be partly responsible for the infrequent side-effect of aseptic meningitis.

Increase in serum viscosity produced by IVIg leads to increased risk of thrombo-embolic events, especially in pre-existing hyperviscosity syndromes like high cholesterol levels, hypergammaglobulinemia or cryoglobulinemia. Similarly, rate of infusion has to be slowed in patients with fluid overload like congestive cardiac failure. Although theoretically possible, hemolysis due to anti-A/B IgG blood group antibodies is almost never seen in clinical practice.

Severe anaphylactic reactions are fortunately rare, but cause a potential problem in selective IgA deficiency, which occurs in 1:1000 patients. These patients may have anti-IgA antibodies, which cross-react with the small IgA content in the infused IVIg, causing macromolecular complexes. Unless in an emergency situation, it is a good practice to check the immunoglobulin levels prior to commencement of IVIg therapy.

Subcutaneous Immunoglobulin (SCIg) as an Alternative to IVIg

Side effects of IVIg are likely to be secondary to an allergic reaction to a large quantity of foreign protein. The administration of Ig by subcutaneous route (usually by a small portable pump at home) has been used mainly to avoid these systemic side effects, but also to reduce the cost and to maintain higher trough levels. SCIg has generally been employed in the treatment of primary immunodeficiencies. Some studies even suggest safer administration of SCIg in patients with IgA sensitisation and in pregnancy. SCIg has been shown to be effective in small series of patients with MMN[24] and CIDP [35]. However, SCIg has to be administered more frequently, can have local skin reactions and is contraindicated in patients on anticoagulant therapy and those with bleeding disorders and thrombocytopenia. Further large trials are required before this can be used as standard practice in inflammatory neuropathies.

Most Plausible Mechanisms of Action of IVIg in Inflammatory Neuropathies

1. Anti-idiotypic antibody production
2. Inhibition of complement pathway
3. Fc receptor modulation on macrophages and other effector cells
4. Suppression of pathogenic cytokines
5. Effects on cell migration by modulation of adhesion molecules
6. T-cell modulation
7. Direct effect on remyelination

SUMMARY

Several immune components are likely to be important in the pathogenesis of GBS, CIDP and MMN. Demyelination in GBS and CIDP may happen along the course of the nerve, which requires a breakdown of blood-nerve barrier, where T-cells may be important for the pathophysiology. Demyelination also occurs distally in the intramuscular part of the nerve, where the barrier is absent, possibly mediated by humoral factors. Early in the disease course in GBS, T-cells may play an important role in pathogenesis as evidenced by increase in soluble IL-2 receptors with a reduction in IL-2. Increased levels of IL-6, TNF-α and IFN-γ are found in serum and CSF with increasing severity of the disease. Evidence of peripheral T-cell activation in the form of increased expression of HLA-DR, membrane-bound IL-2 receptor and transferrin receptor are also noticed during the course of GBS.

The presence of auto-antibodies in several patients with GBS (plus its variants) and MMN, suggests that the effect on B-cells and the complement pathway might play a significant role in the therapeutic efficacy of IVIg in these conditions. However not all patients have a clearly detectable antibody and hence T-cells and cytokines are thought to have a significant pathogenic role, as discussed earlier.

In conclusion, there are several possible mechanisms of action of IVIg, none of which has been conclusively proven to be the dominating pathway. The main mechanisms which are likely to be responsible in inflammatory neuropathies are summarised in Box 1. It is possible that combinations of different mechanisms are involved in several autoimmune neurological diseases where IVIg remains a highly effective therapy.

REFERENCES

[1] (1984) High-dose intravenous gammaglobulin for myasthenia gravis. Lancet, 2, 809-810.
[2] (1997) Randomised trial of plasma exchange, intravenous immunoglobulin, and combined treatments in Guillain-Barre syndrome. Plasma Exchange/Sandoglobulin Guillain-Barre Syndrome Trial Group. Lancet, 349, 225-230.
[3] Aukrust, P., Froland, S.S., Liabakk, N.B., Muller, F., Nordoy, I., Haug, C., Espevik, T. (1994) Release of cytokines, soluble cytokine receptors, and interleukin-1 receptor antagonist after intravenous immunoglobulin administration in vivo. Blood, 84, 2136-2143.
[4] Azulay, J.P., Blin, O., Pouget, J., Boucrart, J., Bille-Turc, F., Carles, G., Serratrice, G. (1994) Intravenous immunoglobulin treatment in patients with motor neuron syndromes associated with anti-GM1 antibodies: a double-blind, placebo-controlled study. Neurology, 44, 429-432.
[5] Basta, M., Illa, I., Dalakas, M.C. (1996) Increased in vitro uptake of the complement C3b in the serum of patients with Guillain-Barre syndrome, myasthenia gravis and dermatomyositis. J Neuroimmunol., 71, 227-229.
[6] Basta, M., Van Goor, F., Lucchioli, S., Billings, E.M., Vortmeyer, A.O., Baranyi, L., Szebeni, J., Alving, C.R., Carroll, M.C., Berkower, I., Stojilkovic, S.S., Metcalfe, D.D. (2003) F(ab)2-mediated neutralization of C3a and C5a anaphylatoxins: a novel effector function of immunoglobulins. Nat. Med., 9, 431-438.
[7] Blank, M., Anafi, L., Zandman-Goddard, G., Krause, I., Goldman, S., Shalev, E., Cervera, R., Font, J., Fridkin, M., Thiesen, H.J., Shoenfeld, Y. (2007) The efficacy of specific IVIG anti-idiotypic antibodies in antiphospholipid syndrome (APS): trophoblast invasiveness and APS animal model. Int. Immunol., 19, 857-865.
[8] Bouhlaïl, H., Hocini, H., Quillinet-Gregoire, C., Donkova, V., Rose, S., Amara, A., Longhi, R., Haefliger-Cavillon, N., Beretta, A., Kaveri, S.V., Kazatchkine, M.D. (2001) Antibodies to C-C chemokine receptor 5 in normal human IgG block infection of macrophages and lymphocytes with primary R5-tropic strains of HIV-1. J. Immunol., 166, 7606-7611.
[9] Chapel, H.M. (1994) Consensus on diagnosis and management of primary antibody deficiencies. Consensus Panel for the Diagnosis
Current Proposed Mechanisms of Action

and Management of Primary Antibody Deficiencies.[erratum appears in BMJ 1994 Apr 2;308(6933):913]. BMJ, 308, 581-585.

Chapel, H.M., Spickett, G.P., Ericson, D., Engl, W., Eibl, M.M., Bjorkander, J. (2000) The comparison of the efficacy and safety of intravenous versus subcutaneous immunoglobulin replacement therapy. J. Clin. Immunol., 20, 94-100.

Cunha, A., Gregson, N.A., Hughes, R.A. (2003) Intravenous immunoglobulin modulates lymphocyte CD54 and monocyte Fc gammaRIII expression in patients with chronic inflammatory neuropathies. J. Neuroimmunol., 135, 91-95.

Crow, A.R., Song, S., Semple, J.W., Freedman, J., Lazarus, A.H. (2007) A role for IL-1 receptor antagonist or other cytokines in the acute therapeutic effects of IVIg? Blood, 109, 155-158.

Dalakas, M.C. (2002) Mechanisms of action of IVIg and therapeutic considerations in the treatment of acute and chronic demyelinating neuropathies. Neurology, 59, S13-21.

Dalakas, M.C. (2004) The use of intravenous immunoglobulin in the treatment of autoimmune neuromuscular diseases: evidence-based indications and safety profile. Pharmacol. Ther., 102, 177-193.

Dalakas, M.C., Quares, R.H., Farrer, R.G., Damбросia, J., Soueidan, S., Stein, D.P., Cuper, E., Sekul, E.A., Otero, C. (1996) A controlled study of intravenous immunoglobulin in demyelinating neuropathy with IgM gammopathy. Ann. Neurol., 40, 792-795.

Devathasan, G., Kueh, Y.K., Chong, P.N. (1984) High-dose intravenous gammaglobulin for myasthenia gravis. Lancet, 2, 809-810.

Dieitrich, G., Kazatchkine, M.D. (1990) Normal immunoglobulin G (IgG) for therapeutic use (intraG Ig) contain idiotypic specificities against an immunodominant, disease-associated, cross-reactive idiotype of human anti-thyroglobulin autoantibodies. J. Clin. Invest., 85, 620-625.

Fateh-Moghadam, A., Wick, M., Besinger, U., Geursen, R.G. (1984) High-dose intravenous gammaglobulin for myasthenia gravis. Lancet, 1, 848-849.

Federico, P., Zochodne, D.W., Hahn, A.F., Brown, W.F., Feasby, T.E. (2000) Multifocal motor neuropathy improved by IVIg: randomised, double-blind, placebo-controlled study. Neurology, 55, 1256-1262.

Frank, M.M., Basta, M., Fries, L.F. (1992) The effects of intravascular immune globulin in multi-focal motor neuropathy in a murine model of Miller Fisher syndrome. Brain, 115, 1197-1208.

Harbo, T., Andersen, H., Hess, A., Hansen, K., Sindrup, S.H., Jakobsen, J. (2009) Subcutaneous versus intravenous immunoglobulin in multifocal motor neuropathy: a randomized, single-blind cross-over trial. Eur. J. Neurol., 16, 631-638.

Hughes, R.A., Allen, D., Mackowska, A., Gregson, N.A. (2006) Pathogenesis of chronic inflammatory demyelinating polyradiculoneuropathy. J. Peripher. Nerv. Sys., 11, 30-46.

Hughes, R.A., Donofrio, P., Bril, V., Dalakas, M.C., Deng, C., Hanna, K., Hartung, H.P., Latov, N., Merkies, I.S., van Doorn, P.A. (2006) Intravenous immune globulin (10% caprate- chromatography purified) for the treatment of chronic inflammatory demyelinating polyradiculoneuropathy (ICE study): a randomised placebo-controlled trial. Lancet Neurol., 7, 136-144.

Hughes, R.A., Raphael, J.C., Swan, A.V., van Doorn, P.A. (2001) Intravenous immunoglobulin for Guillain-Barre syndrome. Cochrane Database Syst. Rev., CD002063.

Huet, V., Kaveri, S.V., Mouhoub, A., Dietrich, G., Mani, J.C., Klatzmann, D., Kazatchkine, M.D. (1994) Anti-CD4 activity of normal human immunoglobulin G for therapeutic use. (Intravenous immunoglobulin, IVIg). Therap. Immunol., 1, 269-277.

Imbach, P., Barandun, S., d’Apuzzo, V., Baunegartner, C., Hirt, A., Morell, A., Rossi, E., Schoni, M., Vest, M., Wagner, H.P. (1981)

High-dose intravenous gammaglobulin for idiopathic thrombocytopenic purpura in childhood. Lancet, 1, 1228-1231.

Kaneko, Y., Nimmerjahn, F., Ravetch, J.V. (2006) Anti-inflammatory activity of immunoglobulin G resulting from Fc sialylation. Science, 313, 670-673.

Kazatchkine, M.D., Kaveri, S.V. (2001) Immunomodulation of autoimmune and inflammatory diseases with intravenous immune globulin. N. Engl. J. Med., 345, 747-755.

Kleyweg, R.P., van der Meche, F.G., Meulestel, J. (1988) Treatment of Guillain-Barre syndrome with high-dose gammaglobulin. Neurology, 38, 1639-1641.

Kondo, N., Kasahara, K., Kameyama, T., Suzuki, Y., Shimozawa, N., Tomatsu, S., Nakashima, Y., Hori, T., Yamagishi, A., Ogawa, T. (1994) Intravenous immunoglobulin productions by suppressing C2(+) dependent signal transduction through Fc gamma receptors in B lymphocytes. Scand. J. Immunol., 40, 37-42.

Lapointe, B.M., Hers, L.M., Gill, V., Metz, L.M., Kubes, P. (2004) IVIg therapy in brain inflammation: etiology-dependent differential effects on leukocyte recruitment. Brain, 127, 2649-2656.

Lee, D.H., Linker, R.A., Paulus, W., Schneider-Gold, C., Chan, A., Gold, R. (2008) Subcutaneous immunoglobulin infusion: a new therapeutic option in chronic inflammatory demyelinating polyneuropathy. Muscle Nerve, 37, 406-409.

Leger, J.M., Chassande, B., Musset, L., Meinginer, V., Pouche, P., Vennmann, N. (2001) Intravenous immunoglobulin therapy in multifocal motor neuropathy: a double-blind, placebo-controlled study. Brain, 124, 145-153.

Leung, D.Y., Cotran, R.S., Kurt-Jones, E., Burns, J.C., Newburger, J.W., Pober, J.S. (1989) Endothelial cell activation and high inter-leukin-1 secretion in the pathogenesis of acute Kawasaki disease. Lancet, 2, 1298-1302.

Lutz, H.U., Stammler, P., Jelezarova, E., Nater, M., Spacht, P.J. (1996) High doses of immunoglobulin G attenuate immune aggression mediated complement activation by enhancing physiologic cleavage of C3b in C3b-IgG complexes. Blood, 88, 184-193.

Malik, U., Oleksiewicz, L., Latov, N., Cardo, L.J. (1996) Intravenous immunoglobulin inhibits binding of anti-GM1 to its target antigen. Ann. Neurol., 39, 136-139.

Marchalions, J.J., Kaymaz, H., Dedoeoglu, F., Schluter, S.F., Yocum, D.E., Edmundson, A.B. (1992) Human autoantibodies re-active with synthetic autoantigens from T-cell receptor beta chain. Proc. Natl. Acad. Sci. U.S.A., 89, 3325-3329.

Mehta, Y.S., Badakere, S.S. (1996) In-vitro inhibition of antiplatelet antibodies by intravenous immunoglobulins and Rh immune globulins. J. Postgraduate Med., 42, 46-49.

Mendell, J.R., Barohn, R.J., Freimer, M.L., Kissel, J.T., King, W., Nagarja, H.N., Rice, R., Campbell, W.W., Donofrio, P.D., Jackson, C.E., Lewis, R.A., Shy, M., Simpson, D.M., Parry, G.J., Rivner, M.H., Thornton, C.A., Bromberg, M., Tandan, R., Harati, U., Nicola, M., Pfeiffer, W., Vitetta, L., Nadel, R., Marchalions J.J., Kaymaz, H., Dedoeoglu, F., Schluter, S.F., Yocum, D.E., Edmundson, A.B. (1992) Human autoantibodies re-active with synthetic autoantigens from T-cell receptor beta chain. Proc. Natl. Acad. Sci. U.S.A., 89, 3325-3329.

Nimmerjahn, F., Antony, R.M., Ravetch, J.V. (2007) Agalactosylated IgG antibodies depend on cellular Fc receptors for in vivo activity. Proc. Natl. Acad. Sci. U.S.A., 104, 8433-8437.

Nimmerjahn, F., Ravetch, J.V. (2008) Anti-inflammatory actions of intravenous immunoglobulin. Ann. Rev. Immunol., 26, 513-533.

Nobile-Orazio, E., Terenghi, F. (2005) IVIg in idiopathic autoinflammatory diseases: analysis in the light of the latest results. J. Neurol., 252 (Suppl 1), 17-13.

Pashov, A., Kaveri, A., Kazatchkine, M.D., Bellon, B. (1996) Suppression of experimental autoimmune encephalomyelitis by intravenous immunoglobulin. Kazatchkine MD, Morell A, Eds. Intravenous immunoglobulin: research and therapy. Parthenon Publishing, New York, 317-318.

Rajabally, Y.A., Seow, H., Wilson, P. (2006) Dose of intravenous immunoglobulin in chronic inflammatory demyelinating polyneuropathy. J. Peripher. Nerv. Sys., 11, 325-329.

Raphael, J.C., Chevert, S., Harbou, M., Jars-Guieneuc, M.C. (2001) Intravenous immune globulins in patients with Guillain-
Barre syndrome and contraindications to plasma exchange: 3 days versus 6 days. J. Neurol. Neurosurg. Psychiatry, 71, 235-238.

[50] Raphael, J.C., Chevret, S., Hughes, R.A., Ananne, D. (2002) Plasma exchange for Guillain-Barre syndrome. Cochrane Database Syst. Rev., CD001798.

[51] Rossi, F., Jayne, D.R., Lockwood, C.M., Kazatchkine, M.D. (1991) Anti-idiotypes against anti-neutrophil cytoplasmic antigen autoantibodies in normal polyclonal IgG for therapeutic use and in the remission sera of patients with systemic vasculitis. Clin. Exp. Immunol., 83, 298-303.

[52] Rossi, F., Kazatchkine, M.D. (1989) Antiidiotypes against autoantibodies in pooled normal human polyclonal IgG. J. Immunol., 143, 4104-4109.

[53] Schuller, E., Govaerts, A. (1983) Fir therapeutic polyimmunoglobulin preparations for intravenous use prevent experimental autoimmune uveoretinitis. Int. Immunol., 5, 1559-1567.

[54] Samuelsson, A., Towers, T.L., Ravetch, J.V. (2001) Anti-inflammatory activity of IVIG mediated through the inhibitory Fc receptor. Science, 291, 484-486.

[55] Saoudi, A., Hurez, V., de Kozak, Y., Kuhn, J., Kaveri, S.V., Kazatchkine, M.D., Druet, P., Bellon, B. (1993) Human immunoglobulin preparations for intravenous use prevent experimental autoimmune uveoretinitis. Clin. Exp. Immunol., 74, 311-316.

[56] Schaller, E., Govaerts, A. (1983) First results of immunotherapy with immunoglobulin G in multiple sclerosis patients. Eur. Neurol., 22, 205-212.

[57] Sharief, M.K., Ingram, D.A., Swash, M., Thompson, E.J. (1999) Intravenous immunoglobulin reduces circulating proinflammatory cytokines in Guillain-Barre syndrome. Neurology, 52, 1833-1838.

[58] Silvestris, F., D’Amore, O., Caffi, P., Savino, L., Dammacco, F. (1996) Intravenous immune globulin therapy of lupus nephritis: use of pathogenic anti-DNA-reactive IgG. Clin. Exp. Immunol., 104 (Suppl 1), 91-97.

[59] Stangel, M., Schumacher, H.C., Ruprecht, K., Boegner, F., Marx, P. (1997) Immunoglobulins for intravenous use inhibit TNF alpha cytotoxicity in vitro. Immunol. Invest., 26, 569-578.

[60] Stohl, W., Elliot, J.E. (1996) In vitro inhibition by intravenous immunoglobulin of human T cell-dependent B cell differentiation induced by staphylococcal superantigens. Clin. Immunol. Immunopathol., 79, 122-133.

[61] Sultan, Y., Kazatchkine, M.D., Maisonneuve, P., Nydegger, U.E. (1984) Anti-idiotypic suppression of autoantibodies to factor VIII (antihaemophilic factor) by high-dose intravenous gammaglobulin. Lancet, 2, 765-768.

[62] Sundblad, A., Marcos, M.A., Malanchere, E., Castro, A., Haury, M., Huez, F., Nobrega, A., Freitas, A., Coutinho, A. (1994) Observations on the mode of action of normal immunoglobulin at high doses. Immunol. Rev., 139, 125-158.

[63] Toungouz, M., Denys, C.H., De Groote, D., Dupont, E. (1995) In vitro inhibition of tumour necrosis factor-alpha and interleukin-6 production by intravenous immunoglobulins. Br. J. Haematol., 89, 698-703.

[64] Toyoda, M., Pao, A., Petrosian, A., Jordan, S.C. (2003) Pooled human gammaglobulin modulates surface molecule expression and induces apoptosis in human B cells. Am. J. Transplant., 3, 156-166.

[65] Van Asseldonk, J.T., Franssen, H., Van den Berg-Vos, R.M., Wokke, J.H., Van den Berg, L.H. (2005) Multifocal motor neuropathy. Lancet Neurol., 4, 309-319.

[66] Van den Berg, L.H., Kerkhoff, H., Oey, P.L., Franssen, H., Mollée, L., Vermeulen, M., Jennekens, F.G., Wokke, J.H. (1995) Treatment of multifocal motor neuropathy with high dose intravenous immunoglobulins: a double blind, placebo controlled study. J. Neurol. Neurosurg. Psychiatry, 59, 248-252.

[67] van Doorn, P.A., Ruts, L., Jacobs, B.C. (2008) Clinical features, pathogenesis, and treatment of Guillain-Barre syndrome. Lancet Neurol., 7, 939-950.

[68] van Engelen, B.G., Miller, D.J., Pavelko, K.D., Hommes, O.R., Rodriguez, M. (1994) Promotion of remyelination by polyclonal immunoglobulin in Thiefer's virus-induced demyelination and in multiple sclerosis. J. Neurol. Neurosurg. Psychiatry, 57 (Suppl), 65-68.

[69] Vassilev, T., Gelin, C., Kaveri, S.V., Zilber, M.T., Bournsell, L., Kazatchkine, M.D. (1993) Antibodies to the CD5 molecule in normal human immunoglobulins for therapeutic use (intravenous immunoglobulins, IVlg). Clin. Exp. Immunol., 92, 369-372.

[70] Vassilev, T., Yamamoto, M., Aissouai, A., Bonnin, E., Berrih-Aknin, S., Kazatchkine, M.D., Kaveri, S.V. (1999) Normal human immunoglobulin suppresses experimental myasthenia gravis in SCID mice. Eur. J. Immunol., 29, 2436-2442.

[71] Vassilev, T.L., Kazatchkine, M.D., Van Huyen, J.P., Mekrache, M., Bonnin, E., Mani, J.C., Lecoubrier, C., Korinth, D., Baruch, D., Schriever, F., Kaveri, S.V. (1999) Inhibition of cell adhesion by antibodies to Arg-Gly-Asp (RGD) in normal immunoglobulin for therapeutic use (intravenous immunoglobulin, IVlg). Blood, 93, 3624-3631.

[72] Vermeulen, M., van der Meche, F.G., Speelman, J.D., Weber, A., Busch, H.F. (1985) Plasma and gamma-globulin infusion in chronic inflammatory polyneuropathy. J. Neurol. Sci., 70, 317-326.

[73] Viard, I., Wehrli, P., Bullani, R., Schneider, P., Holler, N., Salomon, D., Hunziker, T., Saurat, J.H., Tschopp, J., French, L.E. (1998) Inhibition of toxic epidermal necrolysis by blockade of CD95 with human intravenous immunoglobulin. Science, 282, 490-493.

[74] Willison, H.J., Halstead, S.K., Beveridge, E., Zitman, F.M., Greenfield, K.N., Morgan, B.P., Plomp, J.J. (2008) The role of complement and complement regulators in mediating motor nerve terminal injury in murine models of Guillain-Barre syndrome. J. Neuroimmunol., 201-202, 172-182.

[75] Xu, C., Potier, B., Van Huyen, J.P., Lucchiari, N., Michel, O., Chevalier, J., Kaveri, S. (1998) Modulation of endothelial cell function by normal polyclonal human intravenous immunoglobulins: a possible mechanism of action in vascular diseases. Am. J. Pathol., 153, 1257-1266.

[76] Yu, Z., Lennon, V.A. (1999) Mechanism of intravenous immune globulin therapy in antibody-mediated autoimmune diseases. N. Engl. J. Med., 340, 227-228.

[77] Zweiman, B. (1989) Theoretical mechanisms by which immunoglobulin therapy might benefit myasthenia gravis. Clin. Immunol. Immunopathol., 53, S83-91.