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Molecular Biomarkers of Embryonic Stem Cells

Takumi Miura, Akihiro Umezawa and Hidenori Akutsu

Department of Reproduction, National Research Institute for Child Health and Development, Tokyo, Japan

1. Introduction

Embryonic stem cells (ESCs) are pluripotent cells capable of both limitless self-renewal and differentiation into all embryonic lineages, and thus ESCs can give rise to any adult cell type. When ESCs are stably maintained in culture and their pluripotency is strictly enforced, they can serve as an unlimited source for tissue replacement in regenerative medicine for degenerative diseases such as neural disorders, heart disease, and type I diabetes. They also offer enormous potential for drug discovery and toxicology, human developmental biology, and cancer research. Studies of human ESCs (hESCs) biology have developed rapidly since the first reports of their derivation in 1998 (Thomson et al., 1998). Many studies have tried to manipulate the growth and differentiation conditions of hESCs with variable success (Biswa and Hutchins, 2007; Hoffman and Carpenter, 2005). hESCs have been differentiated into the derivatives of all three germ layers: ectoderm, mesoderm, and endoderm. Specifically, these derivatives include cardiomyocytes, neural cells, hepatocyte-like cells, endothelial cells, pancreatic hormone expressing endocrine cells, and hematopoietic progenitor cells (Barberi et al., 2007; Carpenter et al., 2003; D’Amour et al., 2006; Levenberg et al., 2007; Lu et al., 2007; Roy et al., 2006; Wang et al., 2007), and thus hESCs have great potential for use in regenerative medicine to restore heart disease, neuronal functions, hepatic disease, blood vessels, and type I diabetes. In addition, mouse ESCs (mESCs) can generate hepatocytes (Gouon-Evans et al., 2006; Soto-Gutierrez et al., 2007), insulin-producing cells (Schroeder et al., 2006), cerebellar neurons (Salero and Hatten, 2007), and even germ cells (West et al., 2006) in vitro, suggesting that hESC can be applied much more widely to regenerative medicine in the future. On October 2010, Geron corporation in United States announced plans to initiate the phase I clinical trial of hESC-derived oligodendrocyte progenitor cells. However, the clinical application of hESCs is restricted thus far for alleged ethical and scientific reasons. First, hESC research often faces opposition from those who object to the destruction of human embryos. Second, ESC therapy potentially poses the risk of tumorigenesis. ESCs frequently form teratocarcinomas when transplanted into mice. Moreover, the ability of ESCs to provide differentiated cells for regenerative medicine will require continual maintenance of the undifferentiated stem cells for long periods in culture. However, chromosomal stability during extended cell passage cannot be guaranteed, and recent cytogenetic studies of ESCs have revealed karyotypic aberrations (Baker et al., 2007). Third, cell replacement therapies have been limited by the availability of sufficient quantities of cells for transplantation. Although there are many reports describing a method to maintain ESC properties in culture, the large-scale culture of...
ESC lines is still problematic and susceptible to substantial challenges at present (Thomson, 2007). Fourth, the potential for immunorejection should be a concern in its therapeutic use, and thus histocompatible ESCs will be required. Genetically, matched pluripotent ESCs generated via somatic cell nuclear transfer or parthenogenesis are a potential source of patient-derived histocompatible cells and tissues for transplantation (Kim et al., 2007; Menendez et al., 2005; Yang et al., 2007). Selected hESCs can serve as a source of histocompatible tissues for transplantation (Kim et al., 2007). The largest impact on recent ESC biology is the generation of ESC-like cells termed “induced pluripotent stem cells (iPSCs)” from fibroblasts that are created by introducing four genes, Oct4, Sox2, c-Myc, and Klf4 (Takahashi and Yamanaka, 2006). Although the first report described the generation of mouse iPSCs, human iPSCs have also been generated by introducing the same four genes as the mouse iPSCs derived from adult human dermal fibroblasts (Takahashi et al., 2007) or the other distinct genes, Oct4, Sox2, Nanog, and LIN28, from human fetal fibroblasts (Yu et al., 2007). These cells could differentiate into cell types of the three germ layers in vitro and produce teratomas, suggesting that iPSCs have the potential to generate patient- and disease-specific stem cells. Despite the importance of our knowledge of ESCs both in cell biology and clinical medicine, the molecular mechanism underlying the cell biological characteristics of ESCs such as the mechanism that maintains pluripotency and that regulates ESC differentiation, remains largely unknown.

Recent we showed the function of CD9, which is highly expressed in undifferentiated state in ESCs, as well as in embryos using the conventional gene targeting strategy to reveal whether CD9 can serve as a molecular marker to detect, classify, and isolate a particular subpopulation of ESCs and to monitor their state of differentiation (Akutsu et al., 2009). This chapter also reviews other ESC molecular markers (Oct4, Sox2, Nanog, Klf4 and Rex1) in addition to CD9. The accumulation of these ES molecular marker studies will be provided a more detailed view of ESCs and facilitated our understanding of early embryonic development and cell-based therapies.

2. The membrane protein CD9

CD9 is a member of the transmembrane 4 superfamily, also known as the tetraspanin family. Most of these members are cell-surface proteins. CD9 is expressed on the cell surface of mouse and rat male germline stem cells and of neural stem cells. Therefore, CD9 may be involved in the common machinery in stem cells of many self-renewing tissues. In addition, CD9 is also involved in cell development, growth, motility, cell differentiation, and egg-sperm fusion (Hadjiajrgrou and Patterson, 1995; Kanatsu-Shinohara et al., 2004b; Kaprielian et al., 1995; Miyado et al., 2000; Miyado et al., 2008). The expression of CD9 in embryonic as well as adult stem cells populations may indicate a role of CD9 in stem cell self-renewal. Oka et al. have been reported that CD9 is highly expressed in undifferentiated ESCs but rapidly down-regulated after cells differentiation (Oka et al., 2002). Upon application of an antibody against CD9, mouse ESCs can not form compact ES-like colonies. Moreover, ESCs are dead in the presence of the anti-CD9 antibody. Therefore, CD9 may play a role in maintenance of undifferentiated mouse ESCs (Oka et al., 2002). Despite high potential role of CD9 in mouse ESCs, however, CD9 null mice are born healthy and grew normally. Therefore, the question whether CD9 has a role in pluripotent cells of the inner cell mass has not been addressed. Based on these findings, we recently reported that CD9 is dispensable for maintenance of an undifferentiated state and pluripotency (Figure 1)
Fig. 1. Generation of CD9 knockout ESCs.
A. Strategy for generation of CD9-KO ES cells. The schematic maps of the CD9 allele (top), the KO vector carrying the neomycin cassette (middle), and the KO allele generated by homologous recombination (bottom) were shown in scale. CD9-/- fertilized eggs can not be
obtained by a cross of CD9-/- female and male mice. To address this issue, we used ICSI to insert CD9-/- sperm directly into the cytoplasm of CD9-/- egg and bypass the fusion step. As a result, CD9-/- ES cells were successfully isolated from blastocyst of the CD9-/- fertilized egg. B. RT-PCR analysis of ES cell-marker genes. Transcripts of Oct4, Sox2, Nanog, and Rex1 were detected in both CD9-/- and CD9+/+ ESCs without feeder cells. A neomycin-resistance gene was targeted to delete a part of the third exon and all of the fourth exon of CD9. Therefore, neomycin gene was detected in CD9-/- ES cell lines. C. Alkaline phosphatase staining shows undifferentiated CD9-/- ESCs as well as CD9+/+ ESCs. Bar = 500 μm. D. CD9-/- and CD9+/+ ESCs were fixed and stained with antibodies against Oct4, Nanog and Sox2. Nuclei were counterstained with DAPI. Bar = 50 μm. E. Teratomas of CD9 knockout ESCs containing all three germ layers. F. Chimeric embryos derived from CD9 knockout ESCs. When EGFP-positive CD9-/- ESCs, which were homozygotes for the partially deleted CD9 allele and marked by the constitutively-active EGFP transgene, were injected into blastocysts, the embryos developed to chimeras at E13.5 in which widespread contributions of GFP-positive cells were observed in fluorescent stereomicroscopic observation (left panel). Right panel is the control embryo, showing an absence of fluorescence.

(Akutsu et al., 2009). In this report, we established mouse ESCs lacking CD9 by gene targeting. These CD9-/- ESCs exhibited the morphology and growth properties of ESCs, which express the ES marker factors Oct4, Sox2 and Nanog and have the ability to give rise to teratomas composed of tissues from all three germ layers. CD9-/- ESCs also generated mouse chimeras, contributing to various tissues. However, it has been reported that CD9 strongly expresses in mouse and human ES cells, suggesting that CD9 may be one marker of pluripotent stem cells (Nash et al., 2007; Oka et al., 2002). Therefore, our CD9 knockout ESCs may explain the role of CD9 as a hallmark trait of stem cells-self-renewal and differentiation capacity. Thus, we should consider that CD9 might be one of markers for identification of pluripotent stem cells without functional significance like Oct4.

3. The transcription factor OCT4

Oct4 (octamer-binding transcription factor 4) also known as POU5F1 (POU domain, class 5, transcription factor 1) is a protein that is expressed by all pluripotent cells during mouse embryogenesis, and is also abundantly expressed by undifferentiated mouse ESCs and ECC cell lines (Okamoto et al., 1990; Rosner et al., 1990; Scholer et al., 1989a; Scholer et al., 1989b), as well as in EGC cell lines (Donovan and de Miguel, 2003). So far, however, experiments show that Oct4 expression is generally weaker in germline stem cells (GSCs) ( Kanatsu-Shinohara et al., 2004a). Oct4 has also been established as a marker for human pluripotent ESCs. Therefore, downregulation of Oct4 is required for the differentiation of somatic lineages. Oct4-deficient mouse embryos only develop to a stage that looks like a blastocyst, and although cells are allocated to the interior, these blastocysts are actually only composed of trophectoderm cells. As these structures lack a genuine ICM, they cannot be used to produce ESC cell lines ( Nichols et al., 1998). Oct4 has therefore been viewed as being involved in preventing trophectoderm and perhaps somatic-cell differentiation from the ICM, as well as being crucial for maintaining the pluripotent state during embryonic development. Recently, it has been reported that Oct4 only is sufficient to reprogram human neural stem cells to pluripotency (Kim et al., 2009). Therefore, Oct4 is a master gene for
pluripotency. In mouse ESCs, the manipulation of Oct4 expression through inducible or repressible Oct4 transgenes indicates that the relative amount of Oct4 protein ultimately determines cell fate (Niwa et al., 2000). However, the target genes that are actually responsible for implementing Oct4 decisions are only partly known (Du et al., 2001; Saijoh et al., 1996). Similarly, the potential interactions of Oct4 with other (co)factors, except for Sox2 (SRY-related high-mobility group (HMG)-box protein-2; (Pevny and Lovell-Badge, 1997)), remain unclear.

4. The transcription factor SOX2

SRY (sex determining region Y)-box 2, also known as Sox2, is a transcription factor. Sox2 is a member of the HMG-domain DNA-binding-protein family that is implicated in the regulation of transcription and chromatin architecture (Pevny and Lovell-Badge, 1997). Sox2 forms a ternary complex with either Oct4 or the ubiquitous Oct1 protein on the enhancer DNA sequences of Fgf4 (Yuan et al., 1995). This allows Sox2 to participate in the regulation of the ICM and its progeny or derivative cells. Consistent with this role, Sox2 is expressed in ESCs, but it is also expressed in neural stem cells. Therefore, Sox2 is essential to maintain self-renewal of undifferentiated embryonic stem cells. When gene targeting was used to inactivate Sox2, the primitive ectoderm was defective, but it could be rescued (albeit only to survive longer) by the injection of wild-type ESCs into the Sox2-/- blastocysts (Avilion et al., 2003). Reduction of Sox2 expression induces mouse ESCs to differentiate cells into the trophoectoderm lineage, indicating that Sox2 function is essential for maintaining pluripotency. These results also are suggested by Sox2 ablation in vivo. Interestingly, the forced expression of Oct4 rescues the pluripotency of Sox2-null ESCs (Masui et al., 2007). These findings indicate that Sox2 has a unique function in maintaining the pluripotency of ESCs that is related to the transcriptional activation of Oct4.

5. The transcription factor NANOG

Nanog is a highly divergent homeodomain-containing protein commonly accorded a central position in the transcriptional network of pluripotency (Boyer et al., 2005; Cole et al., 2008; Loh et al., 2006; Wang et al., 2006). It is essential for early embryonic development (Mitsui et al., 2003). Undifferentiated, wild-type ESCs normally express Nanog. However, the physiological levels of Nanog in ESCs do not prevent their differentiation after LIF withdrawal. So, under physiological conditions, Nanog seems to be one of several factors that are expressed in pluripotent cells and are downregulated at the onset of differentiation. Nanog-/- mouse ESCs differentiate slowly into extra-embryonic endoderm lineages, which is consistent with the absence of a primitive ectoderm in Nanog-/- embryos that were analysed at E5.5 in vivo (Mitsui et al., 2003). So Nanog expression is responsible for the maintenance of a primitive ectoderm in the embryo. Unlike wild-type ESCs and those forced to express Oct4, mouse ESCs that are overexpressing Nanog are resistant, but not completely refractory, to the spontaneous differentiation that occurs after LIF withdrawal or by chemical induction (for example, after treatment with 3-methoxybenzamide or all-trans retinoic acid). The persistence of Nanog therefore seems to delay, rather than block, the differentiation of ESCs; that is, the threshold of differentiation is increased rather than abolished. In contrast to Nanog overexpression, the reduced expression seen in Nanog+/- ESCs results in labile pluripotency whereby spontaneous differentiation is more likely to
occur after longer times spent in culture (‘passages’) (Hatano et al., 2005). So, the amount of Nanog per cell is crucial for stably maintaining an undifferentiated state even in the presence of LIF. In addition, Nanog is not one of the Yamanaka 4 factors employed to reprogram mouse fibroblasts (Maherali et al., 2007; Okita et al., 2007; Takahashi et al., 2007; Takahashi and Yamanaka, 2006; Wernig et al., 2007). Moreover, addition of Nanog to these 4 factors has not been reported to increase efficiencies. However, Nanog is expressed weakly or not at all in incompletely reprogrammed cells that fail to activate properly the endogenous pluripotent transcriptional circuitry (Silva and Smith, 2008; Sridharan et al., 2009; Takahashi and Yamanaka, 2006). Selection or screening for activation of endogenous Nanog expression facilitates isolation of fully reprogrammed iPSCs that can contribute to adult chimeras and give germline transmission (Okita et al., 2007). Furthermore, in human cells Nanog does facilitate molecular reprogramming (Yu et al., 2007). It has also been shown that Nanog promotes the transfer of pluripotency after ES cell fusion (Silva et al., 2006). However, conditional gene deletion in ESCs revealed that Nanog is not essential for propagation of pluripotency ex vivo (Chambers et al., 2007). Nanog null ESCs are more prone to differentiate but can be maintained indefinitely. Moreover, they contribute extensively to somatic chimeras, similar to CD9 null ESCs.

6. Other transcriptional factors KLF4 and REX1

The mechanism by which Klf4 regulates ES cell self-renewal was first revealed by its identification as a highly up-regulated target gene of LIF signaling in ES cells (Li et al., 2005). ES cells overexpressing Klf4 had a great propensity for self-renewal based on secondary embryoid body (EB) formation. Klf4-transduced EBs expressed higher levels of Oct4, consistent with the notion that Klf4 regulates ES cell self-renewal (Li et al., 2005). The role of Klf4 in regulating pluripotency of ES cells is further revealed by global analysis of promoter occupancy by Yamanaka 4 factors (Kim et al., 2008a). The results identified a transcriptional hierarchy within the four reprogramming factors with both auto-regulatory and feed-forward regulation. In addition, the study indicated that Klf4 is an upstream regulator of a large feed-forward loop that contains Oct4, Sox2, and c-Myc, as well as other common downstream factors including Nanog (Kim et al., 2008a). Combining the results of these studies, it appears that Klf4 exerts a crucial role in somatic cell reprogramming and maintenance of ES cell self-renewal. On the other hand, Klf4 also exhibits both cytostatic and anti-apoptotic effect that is context-dependent. The ability of Klf4 in maintaining immortality of iPSCs maybe explained in part by the requirement of c-Myc as a member of reprogramming factor. Thus, in a manner similar to the cooperation between Klf4 and Ras to affect transformation (Rowland and Peeper, 2006), Klf4 and c-Myc cooperate to affect iPSC cell self-renewal. Thus, Klf4 may suppress apoptosis induced by c-Myc and c-Myc neutralizes Klf4’s cytostatic effect by suppressing p21WAF1/CIP1 (Yamanaka, 2007). In this manner, the balance between Klf4 and c-Myc might play a critical role in the establishment of an immortalized state of iPSCs. In addition, re-expression of Klf4 in an appropriate environment can regenerate the naïve ground state from mouse epiblast stem cells (EpiSCs), which are derived from columnar epithelial epiblast of the early post-implantation embryo (Brook and Gardner, 1997; Hanna et al., 2010; Hanna et al., 2009; Tesar et al., 2007). Therefore, the essential requirement of Klf4 for reprogramming of somatic cells has subsequently been substantiated (Di Stefano et al., 2009; Shi et al., 2008). On the other hand, Klf4+- mice were phenotypically and histologically normal (Katz et al., 2005; Segre et al., 1999). Klf4-/- mice
were born at the expected Mendelian ration. Therefore, Klf4 is also dispensable for maintenance of self-renewal and pluripotency of ESCs.

In addition to Oct4, Sox2, Nanog and Klf4, other putative transcription factors expressing pluripotent stem cells in stem-cell-specific manner have been also identified by several investigators. For example, Rex1 (for reduced expression-1, also known as Zfp42) was first identified a gene that expresses in F9 embryonal carcinoma (EC) cells and is down-regulated after retinoic acid (RA) treatment to induce differentiation (Hosler et al., 1989). This gene encodes a C2H2 zinc-finger protein that is closely similar to Yy1, an evolutionally-conserved component of polycomb-related complex 2 (Gordon et al., 2006). Its highly-specific expression in pluripotent stem cells has been confirmed in mouse and human ESCs (Eiges et al., 2001; Rogers et al., 1991), making it one of the most famous markers of pluripotency tested in various stem cells such as multipotent adult progenitor cells (Jiang et al., 2002) and amniotic fluid cells (Karlmark et al., 2005). Moreover, Rex1 is also known as a marker of the naïve ground state (Nichols and Smith, 2009). This has been argued that the blastocyst origin of human ESCs is evidenced by their expression of Rex1. However, its function in ESCs has not yet been characterized well although it has been reported that a targeted deletion of Rex1 results in loss of the ability to differentiate into visceral endoderm induced by RA in F9 EC cells (Thompson and Gudas, 2002), and that a gene silencing by RNA interference for Rex1 results in loss of capacity to self-renew in ESCs (Zhang et al., 2006). In addition, it has been recently reported that over-expression of Rex1 in ESCs neither induces differentiation in the presence of LIF nor maintains self-renewal in the absence of LIF. Rex1-/- ESCs can be established and contribute whole embryos after blastocyst injection, indicating that they possess proper pluripotency. Moreover, Rex1-/- mice were produced by the intercross of heterozygotes, and both male and female homozygotes were normal and fertile (Masui et al., 2008). These findings support that Rex1 is also dispensable for maintenance of pluripotency in ESCs.

7. Concluding remarks

ESCs can bring unique application to medical and pharmaceutical research. Of note, recent advances in ESC biology have led to the successful generation of iPSCs, which could solve many scientific and ethical problems associated with regenerative medicine and cell-based therapies for degenerative human diseases. Thus, the understanding molecular biomarkers for ESCs is becoming increasingly important for the detection, classification, and isolation of a particular population of ES/iPS cells, and for monitoring the state of differentiation. This chapter discusses that Oct4 only is functionally essential for maintenance of pluripotency in ESCs (Table 1). This is consistent with evidence that Oct4 alone is able to reprogram mouse and human neural stem cells (Kim et al., 2009; Kim et al., 2008b). Therefore, other molecular biomarker highly expressed in ESCs might be markers for identification of pluripotent stem cells without functional significance like Oct4.

Although we have mainly focused here on studies using mouse ESCs, it will be important to understand how these findings relate to human ES cell studies. Studies on human ESCs may be best compared with studies on pluripotent mouse EpiSC lines, which have been established from post-implantation embryos (Brons et al., 2007; Tesar et al., 2007). ESCs and EpiSCs differ from one another in their factor requirements in vitro and in their capacity to incorporate into developing chimaeras. The recent demonstration of revertibility of primed EpiSC state to naïve ESC state is reported in mouse and human (Hanna et al., 2010; Hanna et al., 2009). In the near future, naïve human ESCs will need to be generated from blastocyst.
Table 1. Function of the best-characterized ES cell markers

| Disrupted genes | Knockout ESCs | Passed pluripotency test | References |
|-----------------|---------------|--------------------------|------------|
| Oct4            | not established\(^a\) | N/D                      | Okamoto et al., 1990 |
| Sox2            | established\(^b\)   | chimera                  | Masui et al., 2007 |
| Nanog           | established      | chimera                  | Chambers et al., 2007 |
| Klf4            | established      | chimera                  | Nakatake et al., 2006 |
| Rex1            | established      | chimera                  | Masui et al., 2008 |
| CD9             | established      | chimera                  | Akutsu et al., 2009 |

\(^a\)Oct4 deficient blastocysts lack ICM.
\(^b\)Sox2 null ESCs are maintained by the forced expression of Oct4.

Table 1. Function of the best-characterized ES cell markers

Embryos. Because the naive human pluripotent stem cells will provide a critical tool to model the earliest steps in human embryonic development. Understanding how pluripotent molecular biomarker assemblies change as cells move from one pluripotent compartment to another will allow us to view how the dynamic alterations in cell phenotype that underlie developmental transitions are dictated, which will be surely enhanced our knowledge of ESCs and of early embryonic development and cell-based therapies.

8. References

Akutsu, H.; Miura, T.; Machida, M.; Birumachi, J.; Hamada, A.; Yamada, M.; Sullivan, S.; Miyado, K. & Umezawa, A. (2009). Maintenance of pluripotency and self-renewal ability of mouse embryonic stem cells in the absence of tetraspanin CD9. *Differentiation*, Vol. 78, pp. 137-142.

Avilion, A. A.; Nicolis, S. K.; Pevny, L. H.; Perez, L.; Vivian, N. & Lovell-Badge, R. (2003). Multipotent cell lineages in early mouse development depend on SOX2 function. *Genes Dev*, Vol. 17, pp. 126-140.

Baker, D. E.; Harrison, N. J.; Maltby, E.; Smith, K.; Moore, H. D.; Shaw, P. J.; Heath, P. R.; Holden, H. & Andrews, P. W. (2007). Adaptation to culture of human embryonic stem cells and oncogenesis in vivo. *Nat Biotechnol*, Vol. 25, pp. 207-215.

Barberi, T.; Bradbury, M.; Dincer, Z.; Panagiotakos, G.; Socci, N. D. & Studer, L. (2007). Derivation of engraftable skeletal myoblasts from human embryonic stem cells. *Nat Med*, Vol. 13, pp. 642-648.

Biswas, A. & Hutchins, R. (2007). Embryonic stem cells. *Stem Cells Dev*, Vol. 16, pp. 213-222.

Boyer, L. A.; Lee, T. I.; Cole, M. F.; Johnstone, S. E.; Levine, S. S.; Zucker, J. P.; Guenther, M. G.; Kumar, R. M.; Murray, H. L.; Jenner, R. G.; Gifford, D. K.; Melton, D. A.; Jaenisch, R. & Young, R. A. (2005). Core transcriptional regulatory circuitry in human embryonic stem cells. *Cell*, Vol. 122, pp. 947-956.

Brons, I. G.; Smithers, L. E.; Trotter, M. W.; Rugg-Gunn, P.; Sun, B.; Chua de Sousa Lopes, S. M.; Howlett, S. K.; Clarkson, A.; Ahrlund-Richter, L.; Pedersen, R. A. & Vallier, L.
(2007). Derivation of pluripotent epiblast stem cells from mammalian embryos. *Nature*, Vol. 448, pp. 191-195.

Brook, F. A. & Gardner, R. L. (1997). The origin and efficient derivation of embryonic stem cells in the mouse. *Proc Natl Acad Sci U S A*, Vol. 94, pp. 5709-5712.

Carpenter, M. K.; Rosler, E. & Rao, M. S. (2003). Characterization and differentiation of human embryonic stem cells. *Cloning Stem Cells*, Vol. 5, pp. 79-88.

Chambers, I.; Silva, J.; Colby, D.; Nichols, J.; Nijmeijer, B.; Robertson, M.; Vrana, J.; Jones, K.; Grotewold, L. & Smith, A. (2007). Nanog safeguards pluripotency and mediates germline development. *Nature*, Vol. 450, pp. 1230-1234.

Cole, M. F.; Johnstone, S. E.; Newman, J. J.; Kagey, M. H. & Young, R. A. (2008). Tcf3 is an integral component of the core regulatory circuitry of embryonic stem cells. *Genes Dev*, Vol. 22, pp. 746-755.

D’Amour, K. A.; Bang, A. G.; Eliazer, S.; Kelly, O. G.; Agulnick, A. D.; Smart, N. G.; Moorman, M. A.; Kroon, E.; Carpenter, M. K. & Baetge, E. E. (2006). Production of pancreatic hormone-expressing endocrine cells from human embryonic stem cells. *Nat Biotechnol*, Vol. 24, pp. 1392-1401.

Di Stefano, B.; Prigione, A. & Broccoli, V. (2009). Efficient genetic reprogramming of unmodified somatic neural progenitors uncovers the essential requirement of Oct4 and Klf4. *Stem Cells Dev*, Vol. 18, pp. 707-716.

Donovan, P. J. & de Miguel, M. P. (2003). Turning germ cells into stem cells. *Curr Opin Genet Dev*, Vol. 13, pp. 463-471.

Du, Z.; Cong, H.&Yao, Z. (2001). Identification of putative downstream genes of Oct-4 by suppression-subtractive hybridization. *Biochem Biophys Res Commun*, Vol. 282, pp. 701-706.

Eiges, R.; Schuldiner, M.; Drukker, M.; Yanuka, O.; Itskovitz-Eldor, J. & Benvenisty, N. (2001). Establishment of human embryonic stem cell-transfected clones carrying a marker for undifferentiated cells. *Curr Biol*, Vol. 11, pp. 514-518.

Gordon, S.; Akopyan, G.; Garban, H. & Bonavida, B. (2006). Transcription factor YY1: structure, function, and therapeutic implications in cancer biology. *Oncogene*, Vol. 25, pp. 1125-1142.

Gouon-Evans, V.; Boussemart, L.; Gadue, P.; Nierhoff, D.; Koehler, C. I.; Kubo, A.; Shafriz, D. A. & Keller, G. (2006). BMP-4 is required for hepatic specification of mouse embryonic stem cell-derived definitive endoderm. *Nat Biotechnol*, Vol. 24, pp. 1402-1411.

Hadjiargyrou, M. & Patterson, P. H. (1995). An anti-CD9 monoclonal antibody promotes adhesion and induces proliferation of Schwann cells in vitro. *J Neurosci*, Vol. 15, pp. 574-583.

Hanna, J.; Cheng, A. W.; Saha, K.; Kim, J.; Lengner, C. J.; Soldner, F.; Cassady, J. P.; Muffat, J.; Carey, B. W. & Jaenisch, R. (2010). Human embryonic stem cells with biological and epigenetic characteristics similar to those of mouse ESCs. *Proc Natl Acad Sci U S A*, Vol. 107, pp. 9222-9227.

Hanna, J.; Markoulaki, S.; Mitalipova, M.; Cheng, A. W.; Cassady, J. P.; Staerk, J.; Carey, B. W.; Lengner, C. J.; Foreman, R.; Love, J.; Gao, Q.; Kim, J. & Jaenisch, R. (2009). Metastable pluripotent states in NOD-mouse-derived ESCs. *Cell Stem Cell*, Vol. 4, pp. 513-524.
Hatano, S. Y.; Tada, M.; Kimura, H.; Yamaguchi, S.; Kono, T.; Nakano, T.; Suemori, H.; Nakatsuji, N. & Tada, T. (2005). Pluripotential competence of cells associated with Nanog activity. *Mech Dev*, Vol. 122, pp. 67-79.

Hoffman, L. M. & Carpenter, M. K. (2005). Characterization and culture of human embryonic stem cells. *Nat Biotechnol*, Vol. 23, pp. 699-708.

Hosler, B. A.; LaRosa, G. J.; Grippo, J. F. & Gudas, L. J. (1989). Expression of REX-1, a gene containing zinc finger motifs, is rapidly reduced by retinoic acid in F9 teratocarcinoma cells. *Mol Cell Biol*, Vol. 9, pp. 5623-5629.

Jiang, Y.; Jahagirdar, B. N.; Reinhardt, R. L.; Schwartz, R. E.; Keene, C. D.; Ortiz-Gonzalez, X. R.; Reyes, M.; Lenvik, T.; Lund, T.; Blackstad, M.; Du, J.; Aldrich, S.; Lisberg, A.; Low, W. C.; Largaespada, D. A. & Verfaillie, C. M. (2002). Pluripotency of mesenchymal stem cells derived from adult marrow. *Nature*, Vol. 418, pp. 41-49.

Kanatsu-Shinohara, M.; Inoue, K.; Lee, J.; Yoshimoto, M.; Ogonuki, N.; Miki, H.; Baba, S.; Kato, T.; Kazuki, Y.; Toyokuni, S.; Toyoshima, M.; Niwa, O.; Oshimura, M.; Heike, T.; Nakahata, T.; Ishino, F.; Ogura, A. & Shinohara, T. (2004a). Generation of pluripotent stem cells from neonatal mouse testis. *Cell*, Vol. 119, pp. 1001-1012.

Kanatsu-Shinohara, M.; Toyokuni, S. & Shinohara, T. (2004b). CD9 is a surface marker on mouse and rat male germline stem cells. *Biol Reprod*, Vol. 70, pp. 70-77.

Kaprielian, Z.; Cho, K. O.; Hadjiargyrou, M. & Patterson, P. H. (1995). CD9, a major platelet cell surface glycoprotein, is a ROCA antigen and is expressed in the nervous system. *J Neurosci*, Vol. 15, pp. 562-573.

Karlmark, K. R.; Freilinger, A.; Marton, E.; Rosner, M.; Lubec, G. & Hengstschlager, M. (2005). Activation of ectopic Oct-4 and Rex-1 promoters in human amniotic fluid cells. *Int J Mol Med*, Vol. 16, pp. 987-992.

Katz, J. P.; Perreault, N.; Goldstein, B. G.; Actman, L.; McNally, S. R.; Silberg, D. G.; Furth, E. E. & Kaestner, K. H. (2005). Loss of Klf4 in mice causes altered proliferation and differentiation and precancerous changes in the adult stomach. *Gastroenterology*, Vol. 128, pp. 935-945.

Kim, J.; Chu, J.; Shen, X.; Wang, J. & Orkin, S. H. (2008a). An extended transcriptional network for pluripotency of embryonic stem cells. *Cell*, Vol. 132, pp. 1049-1061.

Kim, J. B.; Greber, B.; Arauzo-Bravo, M. J.; Meyer, J.; Park, K. I.; Zaehres, H. & Scholer, H. R. (2009). Direct reprogramming of human neural stem cells by OCT4. *Nature*, Vol. 461, pp. 649-653.

Kim, J. B.; Zaehres, H.; Wu, G.; Gentile, L.; Ko, K.; Sebastiano, V.; Arauzo-Bravo, M. J.; Ruau, D.; Han, D. W.; Zenke, M. & Scholer, H. R. (2008b). Pluripotent stem cells induced from adult neural stem cells by reprogramming with two factors. *Nature*, Vol. 454, pp. 646-650.

Kim, K.; Lerou, P.; Yabuuchi, A.; Lengerke, C.; Ng, K.; West, J.; Kirby, A.; Daly, M. J. & Daley, G. Q. (2007). Histocompatible embryonic stem cells by parthenogenesis. *Science*, Vol. 315, pp. 482-486.

Levenberg, S.; Zoldan, J.; Basevitch, Y. & Langer, R. (2007). Endothelial potential of human embryonic stem cells. *Blood*, Vol. 110, pp. 806-814.

Li, Y.; McClinton, J.; Zhong, L.; Edenberg, H. J.; Yoder, M. C. & Chan, R. J. (2005). Murine embryonic stem cell differentiation is promoted by SOCS-3 and inhibited by the zinc finger transcription factor Klf4. *Blood*, Vol. 105, pp. 635-637.
Loh, Y. H.; Wu, Q.; Chew, J. L.; Vega, V. B.; Zhang, W.; Chen, X.; Bourque, G.; George, J.; 
Leong, B.; Liu, J.; Wong, K. Y.; Sung, K. W.; Lee, C. W.; Zhao, X. D.; Chiu, K. P.; 
Lipovich, L.; Kuznetsov, V. A.; Robson, P.; Stanton, L. W.; Wei, C. L.; Lim, B. & Ng, H. H. (2006). The Oct4 and Nanog transcription network regulates pluripotency in mouse embryonic stem cells. Nat Genet, Vol. 38, pp. 431-440.

Lu, S. J.; Feng, Q.; Caballero, S.; Chen, Y.; Moore, M. A.; Grant, M. B. & Lanza, R. (2007). Generation of functional hemangioblasts from human embryonic stem cells. Nat Methods, Vol. 4, pp. 501-509.

Maherali, N.; Sridharan, R.; Xie, W.; Utikal, J.; Eminli, S.; Arnold, K.; Stadtfeld, M.; 
Yachechko, R.; Tchieu, J.; Jaenisch, R.; Plath, K. & Hochedlinger, K. (2007). Directly reprogrammed fibroblasts show global epigenetic remodeling and widespread tissue contribution. Cell Stem Cell, Vol. 1, pp. 55-70.

Masui, S.; Nakatake, Y.; Toyoooka, Y.; Shimosato, D.; Yagi, R.; Takahashi, K.; Okochi, H.; 
Okuda, A.; Matoba, R.; Sharov, A. A.; Ko, M. S. & Niwa, H. (2007). Pluripotency governed by Sox2 via regulation of Oct3/4 expression in mouse embryonic stem cells. Nat Cell Biol, Vol. 9, pp. 625-635.

Masui, S.; Ohtsuka, S.; Yagi, R.; Takahashi, K.; Ko, M. S. & Niwa, H. (2008). Rex1/Zfp42 is dispensable for pluripotency in mouse ES cells. BMC Dev Biol, Vol. 8, pp. 45.

Menendez, P.; Bueno, C.; Wang, L. & Bhatia, M. (2005). Human embryonic stem cells: potential tool for achieving immunotolerance? Stem Cell Rev, Vol. 1, pp. 151-158.

Mitsui, K.; Tokuzawa, Y.; Itoh, H.; Segawa, K.; Murakami, M.; Takahashi, K.; Maruyama, M.; 
Maeda, M. & Yamanaka, S. (2003). The homeoprotein Nanog is required for maintenance of pluripotency in mouse epiblast and ES cells. Cell, Vol. 113, pp. 631-642.

Miyado, K.; Yamada, G.; Yamada, S.; Hasuwa, H.; Nakamura, Y.; Ryu, F.; Suzuki, K.; Kosai, K.; Inoue, K.; Ogura, A.; Okabe, M. & Mekada, E. (2000). Requirement of CD9 on the egg plasma membrane for fertilization. Science, Vol. 287, pp. 321-324.

Miyado, K.; Yoshida, K.; Yamagata, K.; Sakakibara, K.; Okabe, M.; Wang, X.; Miyamoto, K.; 
Akutsu, H.; Kondo, T.; Takahashi, Y.; Ban, T.; Ito, C.; Toshimori, K.; Nakamura, A.; 
Ito, M.; Miyado, M.; Mekada, E. & Umezawa, A. (2008). The fusing ability of sperm is bestowed by CD9-containing vesicles released from eggs in mice. Proc Natl Acad Sci U S A, Vol. 105, pp. 12921-12926.

Nakatake, Y.; Fukui, N.; Iwamatsu, Y.; Masui, S.; Takahashi, K.; Yagi, R.; Yagi, K.; Miyazaki, J.; Matoba, R.; Ko, M. S. & Niwa, H. (2006). Klf4 cooperates with Oct3/4 and Sox2 to activate the Lefty1 core promoter in embryonic stem cells. Mol Cell Biol, Vol. 26, pp. 7772-7782.

Nash, R.; Neves, L.; Faast, R.; Pierce, M. & Dalton, S. (2007). The lectin Dolichos biflorus agglutinin recognizes glycan epitopes on the surface of murine embryonic stem cells: a new tool for characterizing pluripotent cells and early differentiation. Stem Cells, Vol. 25, pp. 974-982.

Nichols, J. & Smith, A. (2009). Naive and primed pluripotent states. Cell Stem Cell, Vol. 4, pp. 487-492.

Nichols, J.; Zevnik, B.; Anastasiadis, K.; Niwa, H.; Klewe-Nebenius, D.; Chambers, I.; 
Scholer, H. & Smith, A. (1998). Formation of pluripotent stem cells in the mammalian embryo depends on the POU transcription factor Oct4. Cell, Vol. 95, pp. 379-391.
Niwa, H.; Miyazaki, J. & Smith, A. G. (2000). Quantitative expression of Oct-3/4 defines differentiation, dedifferentiation or self-renewal of ES cells. *Nat Genet*, Vol. 24, pp. 372-376.

Oka, M.; Tagoku, K.; Russell, T. L.; Nakano, Y.; Hamazaki, T.; Meyer, E. M.; Yokota, T. & Terada, N. (2002). CD9 is associated with leukemia inhibitory factor-mediated maintenance of embryonic stem cells. *Mol Biol Cell*, Vol. 13, pp. 1274-1281.

Okamoto, K.; Okazawa, H.; Okuda, A.; Sakai, M.; Muramatsu, M. & Hamada, H. (1990). A novel octamer binding transcription factor is differentially expressed in mouse embryonic cells. *Cell*, Vol. 60, pp. 461-472.

Okita, K.; Ichisaka, T. & Yamanaka, S. (2007). Generation of germline-competent induced pluripotent stem cells. *Nature*, Vol. 448, pp. 313-317.

Pevny, L. H. & Lovell-Badge, R. (1997). Sox genes find their feet. *Curr Opin Genet Dev*, Vol. 7, pp. 338-344.

Rogers, M. B.; Hosler, B. A. & Gudas, L. J. (1991). Specific expression of a retinoic acid-regulated, zinc-finger gene, Rex-1, in preimplantation embryos, trophoblast and spermatocytes. *Development*, Vol. 113, pp. 815-824.

Rosner, M. H.; Vigano, M. A.; Ozato, K.; Timmons, P. M.; Poirier, F.; Rigby, P. W. & Staudt, L. M. (1990). A POU-domain transcription factor in early stem cells and germ cells of the mammalian embryo. *Nature*, Vol. 345, pp. 686-692.

Rowland, B. D. & Peeper, D. S. (2006). KLF4, p21 and context-dependent opposing forces in cancer. *Nat Rev Cancer*, Vol. 6, pp. 11-23.

Roy, N. S.; Cleren, C.; Singh, S. K.; Yang, L.; Beal, M. F. & Goldman, S. A. (2006). Functional engraftment of human ES cell-derived dopaminergic neurons enriched by coculture with telomerase-immortalized midbrain astrocytes. *Nat Med*, Vol. 12, pp. 1259-1268.

Saijoh, Y.; Fujii, H.; Meno, C.; Sato, M.; Hirota, Y.; Nagamatsu, S.; Ikeda, M. & Hamada, H. (1996). Identification of putative downstream genes of Oct-3, a pluripotent cell-specific transcription factor. *Genes Cells*, Vol. 1, pp. 239-252.

Salero, E. & Hatten, M. E. (2007). Differentiation of ES cells into cerebellar neurons. *Proc Natl Acad Sci U S A*, Vol. 104, pp. 2997-3002.

Scholer, H. R.; Balling, R.; Hatzopoulos, A. K.; Suzuki, N. & Gruss, P. (1989a). Octamer binding proteins confer transcriptional activity in early mouse embryogenesis. *Embo J*, Vol. 8, pp. 2551-2557.

Scholer, H. R.; Hatzopoulos, A. K.; Balling, R.; Suzuki, N. & Gruss, P. (1989b). A family of octamer-specific proteins present during mouse embryogenesis: evidence for germline-specific expression of an Oct factor. *Embo J*, Vol. 8, pp. 2543-2550.

Schroeder, I. S.; Rolletschek, A.; Blyszczyk, P.; Kania, G. & Wobus, A. M. (2006). Differentiation of mouse embryonic stem cells to insulin-producing cells. *Nat Protoc*, Vol. 1, pp. 495-507.

Segre, J. A.; Bauer, C. & Fuchs, E. (1999). Klf4 is a transcription factor required for establishing the barrier function of the skin. *Nat Genet*, Vol. 22, pp. 356-360.

Shi, Y.; Desponts, C.; Do, J. T.; Hahm, H. S.; Scholer, H. R. & Ding, S. (2008). Induction of pluripotent stem cells from mouse embryonic fibroblasts by Oct4 and Klf4 with small-molecule compounds. *Cell Stem Cell*, Vol. 3, pp. 568-574.

Silva, J.; Chambers, I.; Pollard, S. & Smith, A. (2006). Nanog promotes transfer of pluripotency after cell fusion. *Nature*, Vol. 441, pp. 997-1001.

Silva, J. & Smith, A. (2008). Capturing pluripotency. *Cell*, Vol. 132, pp. 532-536.
Soto-Gutierrez, A.; Navarro-Alvarez, N.; Zhao, D.; Rivas-Carrillo, J. D.; Lebkowski, J.; Tanaka, N.; Fox, I. J. & Kobayashi, N. (2007). Differentiation of mouse embryonic stem cells to hepatocyte-like cells by co-culture with human liver nonparenchymal cell lines. *Nat Protoc*, Vol. 2, pp. 347-356.

Sridharan, R.; Tchieu, J.; Mason, M. J.; Yachechko, R.; Kuoy, E.; Horvath, S.; Zhou, Q. & Plath, K. (2009). Role of the murine reprogramming factors in the induction of pluripotency. *Cell*, Vol. 136, pp. 364-377.

Takahashi, K.; Tanabe, K.; Ohnuki, M.; Narita, M.; Ichisaka, T.; Tomoda, K. & Yamanaka, S. (2007). Induction of pluripotent stem cells from adult human fibroblasts by defined factors. *Cell*, Vol. 131, pp. 861-872.

Takahashi, K. & Yamanaka, S. (2006). Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell*, Vol. 126, pp. 663-676.

Tesar, P. J.; Chenoweth, J. G.; Brook, F. A.; Davies, T. J.; Evans, E. P.; Mack, D. L.; Gardner, R. L. & McKay, R. D. (2007). New cell lines from mouse epiblast share defining features with human embryonic stem cells. *Nature*, Vol. 448, pp. 196-199.

Thompson, J. R. & Gudas, L. J. (2002). Retinoic acid induces parietal endoderm but not primitive endoderm and visceral endoderm differentiation in F9 teratocarcinoma stem cells with a targeted deletion of the Rex-1 (Zfp-42) gene. *Mol Cell Endocrinol*, Vol. 195, pp. 119-133.

Thomson, H. (2007). Bioprocessing of embryonic stem cells for drug discovery. *Trends Biotechnol*, Vol. 25, pp. 224-230.

Thomson, J. A.; Itskovitz-Eldor, J.; Shapiro, S. S.; Waknitz, M. A.; Swiergiel, J. J.; Marshall, V. S. & Jones, J. M. (1998). Embryonic stem cell lines derived from human blastocysts. *Science*, Vol. 282, pp. 1145-1147.

Wang, J.; Rao, S.; Chu, J.; Shen, X.; Levasseur, D. N.; Theunissen, T. W. & Orkin, S. H. (2006). A protein interaction network for pluripotency of embryonic stem cells. *Nature*, Vol. 444, pp. 364-368.

Wang, Z. Z.; Au, P.; Chen, T.; Shao, Y.; Daheron, L. M.; Bai, H.; Arzigian, M.; Fukumura, D.; Jain, R. K. & Scadden, D. T. (2007). Endothelial cells derived from human embryonic stem cells form durable blood vessels in vivo. *Nat Biotechnol*, Vol. 25, pp. 317-318.

Wernig, M.; Meissner, A.; Foreman, R.; Brambrink, T.; Ku, M.; Hochedlinger, K.; Bernstein, B. E. & Jaenisch, R. (2007). In vitro reprogramming of fibroblasts into a pluripotent ES-cell-like state. *Nature*, Vol. 448, pp. 318-324.

West, J. A.; Park, I. H.; Daley, G. Q. & Geijsen, N. (2006). In vitro generation of germ cells from murine embryonic stem cells. *Nat Protoc*, Vol. 1, pp. 2026-2036.

Yamanaka, S. (2007). Strategies and new developments in the generation of patient-specific pluripotent stem cells. *Cell Stem Cell*, Vol. 1, pp. 39-49.

Yang, X.; Smith, S. L.; Tian, X. C.; Lewin, H. A.; Renard, J. P. & Wakayama, T. (2007). Nuclear reprogramming of cloned embryos and its implications for therapeutic cloning. *Nat Genet*, Vol. 39, pp. 295-302.

Yu, J.; Vodyanik, M. A.; Smuga-Otto, K.; Antosiewicz-Bourget, J.; Frane, J. L.; Tian, S.; Nie, J.; Jonsdottir, G. A.; Ruotti, V.; Stewart, R.; Slukvin, II & Thomson, J. A. (2007). Induced pluripotent stem cell lines derived from human somatic cells. *Science*, Vol. 318, pp. 1917-1920.
Yuan, H.; Corbi, N.; Basilico, C. & Dailey, L. (1995). Developmental-specific activity of the FGF-4 enhancer requires the synergistic action of Sox2 and Oct-3. *Genes Dev*, Vol. 9, pp. 2635-2645.

Zhang, J. Z.; Gao, W.; Yang, H. B.; Zhang, B.; Zhu, Z. Y. & Xue, Y. F. (2006). Screening for genes essential for mouse embryonic stem cell self-renewal using a subtractive RNA interference library. *Stem Cells*, Vol. 24, pp. 2661-2668.
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InTech Europe
University Campus STeP Ri
Slavka Krautzeka 83/A
51000 Rijeka, Croatia
Phone: +385 (51) 770 447
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中国上海市延安西路65号上海国际贵都大饭店办公楼405单元
Phone: +86-21-62489820
Fax: +86-21-62489821