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Bridging the gap between striatal plasticity and learning
Elodie Perrin$^{1,2}$ and Laurent Venance$^{1,2}$

The striatum, the main input nucleus of the basal ganglia, controls goal-directed behavior and procedural learning. Striatal projection neurons integrate glutamatergic inputs from cortex and thalamus together with neuromodulatory systems, and are subjected to plasticity. Striatal projection neurons exhibit bidirectional plasticity (LTP and LTD) when exposed to Hebbian paradigms. Importantly, correlative and even causal links between procedural learning and striatal plasticity have recently been shown. This short review summarizes the current view on striatal plasticity (with a focus on spike-timing-dependent plasticity), recent studies aiming at bridging in vivo skill acquisition and striatal plasticity, the temporal credit-assignment problem, and the gaps that remain to be filled.

Introduction

The striatum receives topographic glutamatergic afferents from all cortical areas and from some thalamic nuclei [1] (Figure 1). It is an important site for action selection and procedural memory formation [2]. Since the demonstration by Yin and coll. [3*] of striatal plasticity following acquisition of a procedural skill, several studies have extended this pioneering work by assessing striatal plasticity across various learning tasks. This review aims at giving the current view on ex vivo striatal plasticity in the light of recent studies evidencing correlative or causal link between in vivo learning and striatal plasticity, in a physiological context. Here, ex vivo refers to brain slice recordings from animals subjected to training or treatment, as opposed to studies in which brain slices are examined in naïve animals to reveal plasticity mechanisms. Striatal plasticity has been a controversial field for at least two decades because of its great variety of results (reviewed in Refs. [4–8]) and the rise of back-and-forth investigations between in vivo and ex vivo bring a unique opportunity for a better understanding of striatal plasticity, and most importantly for bridging the gap between learning and striatal plasticity.

Striatal complexity

Three main reasons account for the diversity of results concerning striatal plasticity: the induction protocols (rate-coded versus time-coded and Hebbian versus non-Hebbian), the striatal heterogeneity, and some technical issues. Some critical technical issues are the age of the animals, the slice orientation (coronal versus sagittal versus horizontal), the location of the stimulation electrode (cortex versus corpus callosum versus striatum) and the rate of the extracellular and intracellular component washout (LTP being optimally observed under sufficient rates of superfusion and high resistance whole-cell recordings). Intermingled anatomo-functional compartments and neuronal units constitute the basis of the striatal heterogeneity: dorsolateral and dorsomedial striatum (DLS and DMS), and direct and indirect trans-striatal pathways, just to cite the main ones which can be assessed during recordings (Figure 1). DMS and DLS receive inputs from associative and sensorimotor cortices and encode for goal-directed behavior and skill acquisition, respectively [3*,9]. In rodents, striatal projection neurons (SPNs) belong either to the direct (d-SPNs) or indirect (i-SPNs) trans-striatal pathways and show distinct dopaminergic receptor expression, D1-class and D2-class receptors, respectively [10]. Recent studies show that d-SPNs and i-SPNs are engaged in a complementary and coordinated manner for action initiation and execution [11–13]. DMS/DLS and d-SPNs/i-SPNs are distinguished in the majority of the plasticity studies. Nevertheless, the third level of striatal structuration, the striosomes (patch)/matrix compartments [14], remains to be more documented for striatal plasticity expression. Another compartment has been recently added, the annular compartments, surrounding the striosomes [15] (Figure 1). Functionally, substance P increases dopamine release within the striosomes but decreases it in the annular compartment, and leaves dopamine unmodified in the matrix [15,16] suggesting distinct neuromodulation of striatal plasticity among these compartments.

Here, instead of recapitulating the plasticity observed in brain slices with the signaling pathways at play (reviewed in Refs. [4,6–8]), we opt for another angle: we first present recent in vivo studies establishing correlative and causal
links between learning and striatal plasticity, and then from these studies we discuss the conditions of emergence of bidirectional striatal plasticity.

From learning to striatal plasticity
Striatal plasticity has been assessed during goal-directed behavior, and across the early and late phases of procedural learning. The analysis of various parameters, used as proxies for synaptic plasticity, has been achieved either in vivo during behavioral tasks (analysis of the firing rate and activity coherence [9,13,17,18,19**,20]); measurement of opto-induced LFP [21**]); or ex vivo after behavioral training (NMDAR/AMPA ratio [3**,22*,23]; spontaneous-EPSCs; [24]; saturation/occlusion plasticity tests [25*,26]) (Figure 2). The link between the acquisition/consolidation of procedural learning and striatal plasticity was first shown by the combined analysis of in vivo firing rate and ex vivo NMDAR/AMPA ratio from mice subjected to an accelerating rotarod [3*]. In vivo analysis shows that DMS, but not DLS, displays increased activity during the early phases of skill acquisition whereas the reverse picture is obtained during the consolidation phases, that is DLS displays increased firing activity while DMS is back to naive levels. Interestingly, NMDAR/AMPA ratio varies only in DLS for the consolidation phase [3*], pointing to the non-NMDAR nature of the corticostriatal plasticity in DMS for the early phases. Ex vivo saturation/occlusion experiments after extended training show LTP at i-SPNs but not at d-SPNs, suggesting that LTP is induced at d-SPNs for the consolidation phase [3*]. Ex vivo AMPAR/NMDAR ratio analysis revealed that during T-maze task, LTP is engaged (but not LTD) in DMS in the early phase while LTD (but not LTP) is involved in the late phase, whereas in DLS, LTD is involved only in the late phase (but LTP in DLS was not explored) [25*]. After habit learning using the lever-pressing task (corresponding to the late phase described in [3*,25*]), ex vivo spontaneous-EPSC are specifically decreased in DLS i-SPNs (indicative of a postsynaptic LTD) [24]. In a serial order task, learning

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Schematic representation of the striatal heterogeneity and the anatomo-functional compartments of the dorsal striatum.
Schematic representation of the direct and indirect trans-striatal pathways of the basal ganglia. Striosomes are shown with black dots distributed between the dorsolateral striatum (blue) and the dorsomedial striatum (orange). Grouped black dots represent striosomes surrounded by the annular compartment (red line, [15]), whereas isolated black dots illustrate the exo-patch [14]. Striosomal SPNs mainly project to SNc whereas SPNs from the matrix belong to the direct or indirect pathway, identified respectively by the expression of D1 and D2 receptors. The direct and indirect pathways are represented, respectively, in green and purple. GPe, external segment of the globus pallidus; EP, entopeduncular nucleus; STN: subthalamic nucleus; SNr, substantia nigra pars reticulata; SNc, substantia nigra pars compacta.

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Figure 1

Bridging the gap between striatal plasticity and learning Perrin and Venance 105

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Current Opinion in Neurobiology 2019, 54:104–112
of a precise sequence depends on d-SPNs in the DLS and induces an increase of the AMPAR/NMDAR ratio at their synapses (but not at i-SPNs) [23]. In a goal-directed task, \textit{ex vivo} AMPAR/NMDAR ratio analysis reveals opposing plasticity in d-SPNs and i-SPNs in DMS, while no modification is observed in DLS [22*]. During the learning of a sensory discrimination task, LTP is detected \textit{in vivo} and \textit{ex vivo} in the auditory striatum [21**].

Learning abstract routines, such as neuropsychiatric skills, requires corticostriatal plasticity as revealed by \textit{in vivo} firing coherence between motor cortex and DLS [9,19**,20]. Firing rate in DLS and coherence in the theta band between motor (M1) cortex and DLS increases in the late phase of volitional modulation of M1 activity [9,19**]; these phenomena are NMDAR-mediated since no change occurs in NMDAR-knock-out mice [19**]. In addition, firing activity patterns of neuronal ensembles that trigger maximal dopamine release along trials of the neuropsychiatric task are selected and progressively shaped for optimized reinforcement learning [20].

It remains to get the full picture of the plasticities successively engaged in d-SPNs and i-SPNs in DLS and DMS from goal-directed to habit formation.

**From striatal plasticity to learning**

A reverse strategy consists in triggering LTP or LTD during a behavioral task to investigate causality between synaptic plasticity and behavioral modifications. NMDAR-LTP and endocannabinoid-LTD were induced by presynaptic optogenetic-stimulation associated with optogenetic SPN depolarization in DMS in operant alcohol self-administration [27**]. LTP and LTD induction promote respectively a long-lasting increase and decrease in alcohol-seeking behavior [27**]. This demonstrates a causal link between the polarity of an induced plasticity and its effect on behavior. Moreover, this strategy allows to test the plasticities identified in brain slices in SPNs or interneurons, and to investigate their \textit{in vivo} impact on synaptic transmission [21**] or behavior [27**].

**Emergence of bidirectional striatal plasticity: Hebbian mode is the key**

The observation that LTP can be induced \textit{in vivo} using a Hebbian paradigm changed the view of a LTD dominance in the striatum [28–30]. Since then, numerous studies have reported LTP (as well as LTD) depending on the stimulation protocol (for reviews see Refs. [4–8]). Nevertheless, striatal LTP still appears more capricious to induce than LTD. Interestingly, following \textit{in vivo} learning tasks, LTP is systematically detected [3*,21**,23,25*,26]. Obviously, the induction phase matters and LTP appears more likely induced upon Hebbian protocols. Hebbian plasticity relies on the quasi-coincident activity on either side of the synapse and spike-timing dependent-plasticity (STDP) protocols aim at mimicking such a Hebbian mode by pairing presynaptic stimulations with postsynaptic back-propagating action potentials [31,32]. Most of the striatal STDP studies report bidirectional (LTP and LTD) plasticity [33–39,40*,41–44] (Figure 3). Note that in rate-coded protocols, the removal of external magnesium or postsynaptic depolarization for inducing LTP aims at mimicking a...
Endocannabinoid-mediated and NMDAR-mediated LTP and LTD are induced depending on the number of pairings in a Hebbian paradigm (STDP).

Changes of the synaptic weight ($W_{\text{total}}$) when the number of pairings and the relative timing between presynaptic and postsynaptic timing ($\Delta t_{\text{STDP}}$) were varied. The color-map depicts the synaptic plasticity predicted by a mathematical model (from [36] and confirmed by experiments (illustrated by the color-coded points) (adapted from [35 and 36]). A small number of post-pre pairings ($\sim 10$–15) produces endocannabinoid-LTP (eCB-LTP), whereas a larger number of pairings ($>75$) induced NMDAR-LTP; for pre-post pairings, endocannabinoid-LTD (eCB-LTD) is induced from 50 pairings. The color bar indicates the color code. In the right panels, the black lines represent the simulation results and the black circles illustrate experimental results.

Therefore, we focus here on striatal STDP [33–39,40,41–46], which is a typical Hebbian paradigm. Bidirectional plasticity has been observed following STDP, that is LTD and LTP were observed depending on the order of pairings [33–36,39,41,43–47]. Interestingly, by varying the number and/or frequency of STDP pairings a complex plasticity landscape is obtained: from the ‘classical’ bidirectional NMDAR-LTP and endocannabinoid-LTD for $\sim 100$ pairings [34,36], to endocannabinoid-LTP for a lower number ($\sim 10$–15) of pairings [39,40,44,47] (Figure 3). It remains to determine the role of endocannabinoid-LTP in vivo and how it combines with NMDAR-LTP.

It should be noted that the plasticities observed in vitro (brain slices) and in vivo relate to distinct phases. Studies using brain slices investigate plasticity up to one hour post-protocol, thus referring to the early plasticity, whereas AMPA/NMDAR ratio and occlusion/saturation analysis are performed 1–3 days after the learning task, corresponding to the long-lasting phase of plasticity. Therefore, conclusions drawn from in vitro and ex vivo / in vivo plasticity are not straightforward, and it remains to determine whether similar signaling pathways are engaged during early and late plasticity phases.

In Hebbian plasticity, the association of two factors controls the synaptic strength, that is two inputs (and/or activity patterns) on the presynaptic and postsynaptic elements, with the addition of a third factor modulating plasticity [48]. Here, recent studies concerning GABA and dopamine acting as third factors for striatal STDP help to clarify the plasticity debate.

The conflicting observations of Hebbian [34,35] or anti-Hebbian striatal STDP (in brain slices [33,36,39,41,45] or in vivo [37]) are explained by the use (or lack of use) of GABA antagonists [38,32]. The appearance of tonic GABAergic signaling during development gates STDP polarity, promoting anti-Hebbian STDP in the adult striatum [43]. It remains to investigate pathological effects of tonic GABAergic transmission in striatal plasticity and procedural learning.
Dopamine is crucial for action selection and supervised learning [49]. Striatal Hebbian plasticity requires dopamine (brain slices [34,35,50]; *in vivo* [37,42]). A dendritic spine enlargement and an increase of calcium occur when dopamine is released concomitantly or after (~1 s) glutamate [51**] allowing *in vivo* STDP [37,42]. There are conflicting results concerning the involvement of D1R versus D2R in STDP (post-pre LTD and pre-post LTP requires D1R-activation but not D2R-activation [35]; LTP in d-SPNs is D1R-mediated and LTD in i-SPNs is D2R-mediated [34]; reviewed in Refs. [6–8]). In the absence of dopamine (and with GABA antagonists), D1SPNs show LTD instead of LTP with pre-post pairings, whereas D2-SPNs display LTP for both post-pre and pre-post pairings [34]. Methodological differences such as the location of the stimulation electrode (leading to different dopamine release [52*]) or the number and frequency of paired stimulations could account for differential activation of D1R and D2R and specific-regulation of the back-propagating action potential [5]. This is particularly illustrated by the fact that LTP induced by theta burst optogenetic stimulation is dependent on presynaptic NMDAR and BDNF [53] but not on dopamine, whereas LTP induced with electrical high-frequency stimulation is generally dopamine-dependent (reviewed in Refs. [4–8]).

In future studies it will be crucial to investigate in behaving animals the action of third factors [48] in Hebbian learning, like for the eligibility traces (see next chapter).

**Solving the temporal credit-assignment problem with eligibility traces and striatal plasticity**

The temporal credit-assignment problem questions the temporal link between the reward and the preceding action to allow reinforcement learning [49]. The existence of eligibility traces, originally brought by computational models [54–56], helps to solve the temporal credit-assignment problem. Eligibility traces are synaptic tags induced by Hebbian learning and are transformed into synaptic plasticity by the retroactive effect of neuromodulators. Theoretically, eligibility traces allow to keep a synaptic trace from the learning sequence, but not to promote plasticity *per se*, unless the reward signal occurs before extinction of eligibility traces (Figure 4). Therefore, eligibility traces temporally link the learning sequence with the reward allowing the induction of reinforcement learning via striatal plasticity. Structural plasticity, used as a proxy for synaptic plasticity, occurs exclusively when dopamine release happens 0.3–2 s after an STDP paradigm [51**] (Figure 4). D1R and dendritic PKA activation allow to bridge the action (glutamatergic inputs) and the subsequent reward (dopamine); PKA activation is short-lived because of the high phosphodiesterase-10 A activity in distal dendrites [51**].

Eligibility traces bridge the gap between learning sequence and subsequent reward to promote reinforcement learning.

Illustration of reinforcement learning allowed by short-lived eligibility traces at corticostriatal synapses. Eligibility traces are triggered following Hebbian sequence, which *per se* does not induce plasticity (illustrated here by the flat grey line coding for the synaptic weight). These eligibility traces (constituted by PKA activity controlled by phosphodiesterase-10 A in d-SPNs, as reported by [45]) can be transformed before their extinction into plasticity, if the teaching signal (dopamine in striatum) is delivered during the maintenance phase (3) of the eligibility traces; No plasticity is induced if dopamine is released either before (1), during the build-up (2) or after (4) eligibility traces (vertical bars illustrate the neuronal firing).

Eligibility traces also retroactive effects of on existing plasticity since dopamine delivered 2 s after cell-conditioning protocol (and importantly not before or during protocol) converts LTD in LTP [42,52*].

Therefore, the expression of eligibility traces and the delivering of a distal reward allow the expression of plasticity [51**] or even the conversion of a form of plasticity into another [42,52*].

**Future directions**

Among the striatal compartments, the striosome and the matrix remain the less documented in terms of
physiological role in striatal function. Thanks to new markers [14], in vivo two-photon monitoring during task performance showed overlapping responses of neurons belonging to the striosome and the matrix, with differential firing activity for reward coding [57]. Because of striosome/matrix differential inputs and outputs [14], dopaminergic [15,16] and endocannabinoid [58] regulation, specific plasticities with differential modulation are expected to occur.

Although differential polarity of STDP in GABAergic and cholinergic interneurons has been shown [59], the full picture of the interplay of plasticities at striatal circuits is just beginning to be understood. It remains to further investigate the underlying mechanisms of the plasticities occurring at the lateral connections [60,61] such as interneuron–interneuron, interneurons–SPNs and SPN–SPNs. For example, a study showing an LTD at inhibitory synapses between SPN–SPNs and fast-spiking interneuron–SPNs demonstrates that distinct endocannabinoid signaling pathways are engaged depending on membrane potentials (up versus down states) [62]. Additionally, with the discovery of long-range projecting corticostriatal GABAergic neurons modulating motor activity via differential action on d-SPNs and i-SPNs [63], it will be important to take into account the fact that cortical activation leads to the direct release not only of glutamate but also of GABA into the striatum. This changes our view on the striatal excitation-inhibition balance, and begs the question of whether plasticity, if any, occurs between these long-range GABAergic neurons and the d-SPNs and i-SPNs.

Determining the conditions of emergence of plasticity helps to better understand the striatal capability for storage and recall of information. Noisy STDP pairings shows that plasticity robustness depends on the signaling pathways: NMDAR-LTP is more fragile than endocannabinoid-plasticity [44]. Interestingly, resistance of NMDAR-LTP to noisy patterns is increased with higher frequency or number of pairings. In vivo Hebbian plasticity appears as a multivariate function of the number and frequency of pairings, but also the variability of the spike timing. In-vivo-like conditions for striatal plasticity, using naturalistic firing patterns of cortical/thalamic/striatal neurons recorded in learning tasks still need to be explored. Although STDP aims at mimicking Hebbian learning, reservations were expressed about its physiological validity [64]. Input-timing-dependent plasticity constitutes a Hebbian upgrade of STDP. It consists in paired activation of presynaptic inputs (distinct cortical areas and/or thalamic nuclei, for example), leading to subthreshold or suprathreshold activity in the postsynaptic neuron, as performed recently in avian basal ganglia [65].

Calcium imaging of i-SPNs and d-SPNs recorded ex vivo just after different phases of an operant lever-pressing task revealed that i-SPNs fired before d-SPNs in the goal-directed phase, whereas the reverse picture is observed during the habitual phase [13]. GABAergic fast-spiking interneurons become more excitable in habitual behavior [66,67] and could account for the reverse temporal order of firing between d-SPNs and i-SPNs. It remains to examine plasticity in DLS across goal-directed to habitual behavior at d-SPNs and i-SPNs, and also in GABAergic interneurons. Also, most of the studies focused on the involvement of NMDAR-LTP in learning. Based on the diversity of plasticities revealed by studies in brain slices (Figure 3), one needs to evaluate the role of the endocannabinoid-LTD and -LTP [39,40,47] across learning. Supporting this view, a recent study showed that endocannabinoids set the transition between goal-directed and habit formation via the control of cortico(orbital frontal cortex)-striatal synaptic weight [68]; the nature of the endocannabinoid plasticity at play at these synapses allowing the shift between goal-directed behavior to habits remains to be determined.

Attempts are made to link the complexity of striatal STDP and goal-directed behavior by elaborating computational models (for recent examples see [42,69]). In future studies, it will be necessary to upgrade the models with recent experimental findings, such as, to name a few, the lateral connections [60,61], the new faces of striatal STDP [39,40,47], the key role of striatal GABAergic interneurons in procedural learning [66,67] as well as the eligibility traces features [42,51,52].

A way to approach in vivo striatal plasticity during learning is to analyze the cortico-striatal synchronous oscillations. However, because of the absence of a laminar organization of the striatum, these oscillations can be contaminated by volume-conducted signals [70] leading to inaccurate interpretation. To overcome this, an elegant strategy consists in the specific-expression of channelrhodopsin in corticostriatal pyramidal cells and thus the unique possibility to estimate striatal opto-LFP changes in vivo during skill learning [21]. Another strategy is the use of fiber photometry to monitor upstream activity in cortical inputs arising from distinct cortices [71] (Figure 2). In vivo patch-clamp recordings in awake and behaving (head-fixed) rodents allows a single-cell resolution and the data collection of subthreshold and suprathreshold events [72]. This approach has been used for the analysis of the membrane potential dynamics during goal-directed behavior in d-SPNs and i-SPNs [73].

Although cortico-striatal and thalamo-striatal afferents equally contact SPNs [1,74], the thalamo-striatal plasticity has been the focus of a limited number of studies [1,75–78], despite the existence of a brain slice preparation preserving both cortico-striatal and thalamo-striatal connections [79]. Cortical and thalamic (parafascicular nucleus) inputs target evenly d-SPNs and i-SPNs.
[80,81], and thalamo-striatal NMDAR-LTD was observed in d-SPNs and i-SPNs [75–77]. Major differences between cortico-striatal and thalamo-striatal plasticity are expected because of different organizations of glutamatergic and dopaminergic synapses on cortico-striatal and thalamo-striatal pathways [74]. Thus, characterizing the thalamo-striatal plasticity repertoire and its putative interactions with cortico-striatal plasticity is of crucial importance to fully understand the role of the striatum in goal-directed and procedural learning.

The field of striatal plasticity has come to a new age in which the investigation of intrinsic, synaptic and structural plasticity at play across procedural learning (from goal-directed behavior to habits) and across the striatal anatomofunctional complexity has become possible in behaving rodents. A new period of (constructive) debates is expected not only since various forms of plasticity should arise depending not only on the striatal complexity but also on the behavioral task and the related learning phase (early versus late).

Conflict of interest statement
Nothing declared.

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References and recommended reading
Papers of particular interest, published within the period of review, have been highlighted as:
• of special interest
•• of outstanding interest

1. Hunnicutt BJ, Jongbloets BC, Birdsong WT, Gertz KJ, Zhong H, Mao T: A comprehensive excitatory input map of the striatum reveals novel functional organization. eLife 2016, 5.

2. Graybiel AM, Grafton ST: The striatum: where skills and habits meet. Cold Spring Harb Perspect Biol 2015, 7.

3. Yin HH, Mulcare SP, Hilário MRF, Clouse E, Holloway T, Davis MI, •• Hansson AC, Lovinger DM, Costa RM: Dynamic reorganization of striatal circuits during the acquisition and consolidation of a skill. Nat Neurosci 2009, 12:333-341.

In this seminal article, the authors provide the first evidence that synaptic plasticity (via AMPA/NMDAR ratio analysis and saturation plasticity experiments) can be detected in DLS following procedural learning (accelerating rotorad).

4. Di Filippo M, Picconi B, Tantucci M, Ghiglieri V, Bagetta V, Sgobio C, Tozzi A, Parnetti L, Calabresi P: Short-term and long-term plasticity at corticostriatal synapses: implications for learning and memory. Behav Brain Res 2009, 199:108-118.

5. Surmeier DJ, Plotkin J, Shen W: Synaptic plasticity in dorsal striatal circuitry controlling yaptic action selection. Curr Opin Neurobiol 2009, 19:621.

6. Lovinger DM: Neurotransmitter roles in synaptic modulation, plasticity and learning in the dorsal striatum. Neuropharmacology 2010, 58:951-961.

7. Lerner TN, Kreitzer AC: Neuromodulatory control of striatal plasticity and behavior. Curr Opin Neurobiol 2011, 21:322-327.

8. Cerovic M, d’Isla R, Tonini R, Brambilla R: Molecular and cellular mechanisms of dopamine-mediated behavioral plasticity in the striatum. Neurobiol Learn Mem 2013, 105:63-80.

9. Koralek AC, Costa RM, Carmena JM: Temporally precise cell-specific coherence develops in corticostriatal networks during learning. Neuron 2013, 79:865-872.

10. Calabresi P, Picconi B, Tozzi A, Ghiglieri V, Filippo MD: Direct and indirect pathways of basal ganglia: a critical reappraisal. Nat Neurosci 2014, 17:1022-1030.

11. Cui G, Jun SB, Jin X, Pham MD, Vogel SS, Lovinger DM, Costa RM: Concurrent activation of striatal direct and indirect pathways during action initiation. Nature 2013, 494:238-242.

12. Tercuapetla F, Jin X, Lima SQ, Costa RM: Complementary contributions of striatal projection pathways to action initiation and execution. Cell 2016, 166:703-715.

13. O’Hare JK, Ade KK, Sukharnakova T, Van Hooser SD, Palmer ML, Yin HH, Calakos N: Pathway-specific striatal substrates for habitual behavior. Neuron 2016, 89:472-473.

14. Smith JB, Klug JR, Ross DL, Howard CD, Hollon NG, Ko VI, Hoffman H, Callaway EM, Gerfen CR, Jin X: Gene-based dissection unveils the inputs and outputs of striatal patch and matrix compartments. Neuron 2016, 91:1089-1094.

15. Brimblecombe KR, Cragg SJ: Substance P weights striatal dopamine transmission differently within the striosome-matrix axis. J Neurosci 2015, 35:9017-9023.

16. Salinas AG, Davis MI, Lovinger DM, Mateo Y: Dopamine dynamics and cocaine sensitivity differ between striosome and matrix compartments of the striatum. Neuropharmacology 2016, 108:275-283.

17. Thorn CA, Graybiel AM: Differential entrainment and learning-related dynamics of spike and local field potential activity in the sensorimotor and associative striatum. J Neurosci 2014, 34:2845-2859.

18. Barnes TD, Mao J-B, Hu D, Kubota Y, Dreyer AA, Stamosulis C, Brown EN, Graybiel AM: Advance cueing produces enhanced action-boundary patterns of spike activity in the sensorimotor striatum. J Neurophysiol 2011, 105:1861-1878.

19. Koralek AC, Jin X, Long JD, Costa RM, Carmena JM: •• Corticostriatal plasticity is necessary for learning intentional neuropsychiatric skills. Nature 2012, 483:331-335.

Building-up on their seminal publication (Yinet et al., 2009), Costa’s lab demonstrate the involvement of dorsal striatum and of striatal synaptic plasticity in an abstract skill learning task using neuropsychiatric in the behaving rodents, that is volutional modulation of M1 neural activity using auditory feedback. Using this new goal-directed behavioral task, they identified an increased coherence of the spiking activity in the theta band between the motor cortex and the striatum during the late phase of learning, accompanied by an ex vivo modification of the NMDAR/AMPA ratio.

20. Athalye VR, Santos FJ, Carmena JM, Costa RM: Evidence for a neural law of effect. Science 2018, 359:1024-1029.

21. Xiong Q, Znamensky P, Zador AM: Selective corticostriatal •• plasticity during acquisition of an auditory discrimination task. Nature 2015, 521:348-351.

The authors show the specific-engagement of striatal synaptic plasticity in striatal neurons coding for low or high frequency in the acquisition of an auditory frequency discrimination task. They set-up an elegant technique to assess synaptic efficacy changes in behaving animals by estimating opto-LFP (with the expression of ChR2 in auditory cortex). This study paved the way for future works to assess synaptic plasticity in behaving animals during a learning task.

22. Shan Q, Ge M, Christie MJ, Balleine BW: The acquisition of goal-directed actions generates opposing plasticity in direct and indirect pathways in dorsomedial striatum. J Neurosci 2014, 34:9196-9201.

During learning of a goal-directed behavior, the authors show that opposing AMPA/NMDAR ratio modifications occurs in identified d-SPNs and i-SPNs in d HPMS whereas there is no change in the DLS: AMPA/ NMDAR ratio changes suggest that LTP and LTD are expressed in d- SPNs and i-SPNs, respectively (if both LTP and LTD are NMDAR-mediated).
23. Rothwell PE, Hayton SJ, Sun GL, Fucillo MV, Lim BK, Malenka RC: Input- and output-specific regulation of serial order performance by corticostriatal circuits. Neuron 2015, 88:345-356.

24. Shan Q, Christie MJ, Balleine BW: Plasticity in striatopallidal projection neurons mediates the acquisition of habitual actions. Eur J Neurosci 2015, 42:2097-2104.

25. Hawes SL, Evans RC, Unruh BA, Benkert EE, Gillani F, Dumas TC, Blackwell KT: Multimodal plasticity in dorsal striatum while learning a lateralized navigation task. J Neurosci 2015, 35:10535-10549.

The authors explore the engagement of synaptic, intrinsic and structural plasticity in DLS and DMS during learning in a T-maze task. This very impressive body of work shows region(DLS versus DMS)- and hemi-sphere(s versus contra-lateral)-specific synaptic modifications correlated with the learned turning direction and the learning stages (from naive to overtrained rats). Importantly, the authors do not only show the engagement of LTP but also that of LTD (occurring mainly in the late phases of learning).

26. Giordano N, Iemolo A, Mancini M, Cacace F, De Risi M, Latagliata EC, Ghiglioni V, Bellenchini GC, Puglisi-Allegra S, Calabresi P et al.: Motor learning and metaplasticity in striatal neurons: relevance for Parkinson’s disease. Brain J Neuro 2018, 141:505-520.

27. Ma T, Cheng Y, Hellard ER, Wang X, Lu J, Gao X, Huang COY, ** Wei X-Y, Ji-J-Y, Wang J: Bidirectional and long-lasting control of alcohol-seeking behavior by corticostriatal LTP and LTD. Nat Neurosci 2018, 21:373-383.

This work constitutes the first causal demonstration of striatal synaptic plasticity (in d-SPNs in DMS) in an operant task. The authors applied vivo Hebbian bidirectional plasticity and show that LTP and LTD have an opposing effect on alcohol-seeking behavior. Moreover, they show ex vivo that the same stimulation (presynaptic HFS) induces either endocannabinoid-LTD or NMDAR-LTP depending on the non-Hebbian or Hebbian mode.

28. Calabresi P, Pisani A, Mercuri NB, Bernardi G: The corticostriatal projection: from synaptic plasticity to dysfunctions of the basal ganglia. Trends Neurosci 1996, 19:19-24.

29. Lovering DM, Partridge JG, K-C Tang: Plastic control of striatal glutamatergic transmission by ensemble actions of several neurotransmitters and targets for drugs of abuse. Ann NY Acad Sci 2003, 1003:226-240.

30. Mahon S, Deniau J-M, Charpier S: Corticostriatal plasticity: life after the depression. Trends Neurosci 2004, 27:460-467.

31. Fino E, Venance L: Spike-timing dependent plasticity in the striatum. Front Synaptic Neurosci 2010, 2.

32. Feldman DE: The spike timing dependence of plasticity. Neuron 2012, 75:556-571.

33. Fino E, Gowinski J, Venance L: Bidirectional activity-dependent plasticity at corticostriatal synapses. J Neurosci 2005, 25:11279-11287.

34. Shen W, Flajolet M, Greengard P, Surmeier DJ: Dichotomous dopaminergic control of striatal synaptic plasticity. Science 2008, 321:848-851.

35. Pavlak V, Kerr JND: Dopamine receptor activation is required for corticostriatal spike-timing-dependent plasticity. J Neurosci Off J Soc Neurosci 2008, 28:2435-2446.

36. Fino E, Paille V, Cui Y, Moreira-Herreras T, Deniau J-M, Venance L: Distinct coincidence detectors govern the corticostriatal spike timing-dependent plasticity. J Physiol 2010, 588:3045-3062.

37. Schulz JM, Redgrave P, Reynolds JN: Cortico-striatal spike-timing dependent plasticity after activation of subcortical pathways. Front Synaptic Neurosci 2010, 2.

38. Paille V, Fino E, Du K, Morera-Herreras T, Perez S, Kotaletski JH, Venance L: GABAergic circuits control spike-timing-dependent plasticity. J Neurosci 2013, 33:9353-9363.

39. Cui Y, Paille V, Xu H, Genet S, Delord B, Fino E, Berry H, Venance L: Endocannabinoids mediate bidirectional striatal spike-timing-dependent plasticity. J Physiol 2015, 593:2833-2849.

40. Cui Y, Prokin I, Xu H, Delord B, Genet S, Venance L, Berry H: Endocannabinoid dynamics gate spike-timing dependent depression and potentiation. eLife 2016, 5.

In this study (and in Cui et al., 2015, and Xu et al., 2018), the authors report that a low number of paired corticostriatal stimulations induced a new form of synaptic plasticity at d-SPNs and i-SPNs in DLS, that is an endocannabinoid-LTP. This LTP is controlled by dopamine and is dependent on the presynaptic Pka. Endocannabinoid-LTP may be involved in quick synaptic adaptations during rapid learning.

41. Valtcheva S, Venance L: Astrocytes gate Hebbian synaptic plasticity in the striatum. Nat Commun 2016, 7.

42. Fisher SD, Robertson PB, Black MJ, Redgrave P, Sagar MA, Abraham WC, Reynolds JN: Reinforcement determines the timing dependence of corticostriatal synaptic plasticity in vivo. Nat Commun 2017, 8.

43. Valtcheva S, Paillé V, Dembistskaya Y, Perez S, Gangarossa G, Fino E, Venance L: Developmental control of spike-timing-dependent plasticity by tonic GABAergic signaling in striatum. Neuropharmacology 2017, 121:261-277.

44. Cui Y, Prokin I, Mendes A, Berry H, Venance L: Robustness of STDP to spike timing jitter. Sci Rep 2018, 8:8139.

45. Shindou T, Ochi-Shindou M, Wickens JR: A Ca2+ threshold for induction of spike-timing-dependent depression in the mouse striatum. J Neurosci 2011, 31:13015-13022.

46. Hawes SL, Gillani F, Evans RC, Benkert EA, Blackwell KT: Sensitivity to theta-burst timing permits LTP in dorsal striatal adult brain slice. J Neurophysiol 2013, 110:2027-2036.

47. Xu H, Perez S, Cornil A, Cui Y, Berry H, Venance L: Endocannabinoid-dopamine interactions mediate spike-timing dependent potentiation in the striatum. Nat Commun 2018, 9:4118 http://dx.doi.org/10.1038/s41467-018-06409-5.

48. Foncelle A, Mendes A, Jedrzejewska-Szczek J, Valtcheva S, Berry H, Blackwell KT, Venance L: Modulation of spike-timing dependent plasticity: towards the inclusion of a third factor in computational models. Front Comput Neurosci 2018, 12.

49. Schultz W: Neuronal reward and decision signals: from theories to data. Physiol Rev 2015, 95:853-951.

50. Kreitzer AC, Malenka RC: Dopamine modulation of state-dependent endocannabinoid release and long-term depression in the striatum. J Neurosci 2005, 25:10537-10545.

51. Yagishita S, Hayashi-Takagi A, Ellis-Davies GCR, UraKubo H, Ishii S, Kasai H: A critical time window for dopamine actions on the structural plasticity of dendritic spines. Science 2014, 345:1616-1620.

Yagishita et al. experimentally elucidate the temporal credit-assignment problem by showing that dopamine (reward) released up to 2s after glutamatergic input (the action to be rewarded) allows the expression of structural plasticity (spine enlargement). This mechanism is D1R-mediated and thus would occur at d-SPNs. The authors found that eligibility traces were kept short-lived because of the high phosphodies- terase-10A activity in the distal dendrites limiting PKA activation in a narrow time window.

52. Shindou T, Shindou M, Watanabe S, Wickens J: A silent eligibility trace enables dopamine-dependent synaptic plasticity for reinforcement learning in the mouse striatum. Eur J Neurosci 2018 http://dx.doi.org/10.1111/ejn.13921.

Using STDP paradigm in brain slices, the authors show that caged-dopamine delivered 2-4 seconds after glutamatergic inputs allows the conversion of LTD into LTP at corticostriatal synapses onto d-SPNs. Importantly, there was no shift of the polarity of plasticity when dopamine was released before or concomitantly with glutamate.

53. Park H, Popescu A, Poo M: Essential role of presynaptic NMDA receptors in activity-dependent BDNF secretion and corticostriatal LTP. Neuron 2014, 84:1009-1022.

54. Izhikevich EM: Solving the distal reward problem through linkage of STDP and dopamine signaling. Cereb Cortex 2007, 17:2443-2452.

55. Gerstner W, Lehmahn M, Liakoni V, Cornell D, Brea J: Eligibility traces and plasticity on behavioral time scales: experimental support of neoHebbian three-factor learning rules. Front. Neural Circuits 2018, 12.
data at the reinforcement-action interface. PLoS Biol 2015, 13: e1002034.

70. Lalia L, Rueda Orozco PE, Jurado-Parras M-T, Brovelli A, Robbe D: Local or not local: investigating the nature of striatal theta oscillations in behaving rats. eNeuro 2017, 4.

71. Kupferschmidt DA, Juczewski K, Cui G, Johnson KA, Lovinger DM: Parallel, but dissociable, processing in discrete corticostriatal inputs encodes skill learning. Neuron 2017, 96:476-489 e5.

72. Petersen CCH: Whole-cell recording of neuronal membrane potential during behavior. Neuron 2017, 95:1266-1281.

73. Sippy T, Lapray D, Crochet S, Petersen CCH: Cell-type-specific sensorimotor processing in striatal projection neurons during goal-directed behavior. Neuron 2015, 88:298-305.

74. Smith Y, Galvan A, Ellender TJ, Doig N, Villalba RM, Huerta-Ocampo I, Wichmann T, Bolam JP: The thalamostriatal system in normal and diseased states. Front Syst Neurosci 2014, 8.

75. Ding J, Peterson JD, Surmeier DJ: Corticostriatal and thalamostriatal synapses have distinctive properties. J Neurosci Off J Soc Neurosci 2008, 28:6483-6492.

76. Ellender TJ, Harwood J, Kosillo P, Capogna M, Bolam JP: Heterogeneous properties of central lateral and parafascicular thalamic synapses in the striatum. J Physiol 2013, 591:257-272.

77. Wu Y-W, Kim J-I, Tawfik VL, Lalchandani RR, Scherrer G, Ding JB: Input- and cell type-specific endocannabinoid-dependent LTD in the striatum. Cell Rep 2015, 10:75-87.

78. Cavaccini A, Gritt M, Giorgi A, Locarno A, Heck N, Migliarini S, Bertero A, Mercuri M, Margiani G, Trusel M et al.: Serotonergic signaling controls input-specific synaptic plasticity at cortical circuits. Neuron 2018, 98:801-816 e7.

79. Smeal RM, Gaspar RC, Keefe KA, Wilcox KS: A rat brain slice preparation for characterizing both thalamostriatal and corticostriatal afferents. J Neurosci Methods 2007, 159:224-235.

80. Doig NM, Moss J, Bolam JP: Cortical and thalamic innervation of direct and indirect pathway medium-sized spiny neurons in mouse striatum. J Neurosci 2010, 30:14610-14618.

81. Lei W, Deng Y, Liu B, Mu S, Guley NM, Wong T, Reiner A: Confocal laser scanning microscopy and ultrastructural study of VGLUT2 thalamic input to striatal projection neurons in rats. J Comp Neurol 2013, 521:1354-1377.