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DE ROO, Mathias, et al. Anesthetics rapidly promote synaptogenesis during a critical period of brain development. *PLOS ONE*, 2009, vol. 4, no. 9, p. e7043

PMID : 19756154
DOI : 10.1371/journal.pone.0007043
Anesthetics Rapidly Promote Synaptogenesis during a Critical Period of Brain Development

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Abstract

Experience-driven activity plays an essential role in the development of brain circuitry during critical periods of early postnatal life, a process that depends upon a dynamic balance between excitatory and inhibitory signals. Since general anesthetics are powerful pharmacological modulators of neuronal activity, an important question is whether and how these drugs can affect the development of synaptic networks. To address this issue, we examined here the impact of anesthetics on synapse growth and dynamics. We show that exposure of young rodents to anesthetics that either enhance GABAergic inhibition or block NMDA receptors rapidly induce a significant increase in dendritic spine density in the somatosensory cortex and hippocampus. This effect is developmentally regulated; it is transient but lasts for several days and is also reproduced by selective antagonists of excitatory receptors. Analyses of spine dynamics in hippocampal slice cultures reveals that this effect is mediated through an increased rate of protrusions formation, a better stabilization of newly formed spines, and leads to the formation of functional synapses. Altogether, these findings point to anesthesia as an important modulator of spine dynamics in the developing brain and suggest the existence of a homeostatic process regulating spine formation as a function of neural activity. Importantly, they also raise concern about the potential impact of these drugs on human practice, when applied during critical periods of development in infants.

Introduction

Formation, elimination and remodeling of excitatory synapses on dendritic spines are continuously active processes that shape the organization of synaptic networks during development. In vivo experiments have shown that these processes are developmentally regulated, and are under the control of experience-driven neuronal activity [1–5]. Accumulating experimental works demonstrate that, during critical periods of development, both environmental, genetic and pharmacological interference with physiological neuronal activity can markedly and permanently alter wiring patterns and, thereby, information processing in the central nervous system (CNS) [6–8]. An important parameter regulating these processes is the balance between excitation and inhibition [9]. Alteration of this balance through interference with the function of local inhibitory circuits determines the characteristics and spacing of input segregation for ocular dominance columns formation and also controls the onset of critical periods by regulating perisomatic GABA responses [10–12]. The level of inhibition present in developing cortical networks plays therefore an important role in fine-tuning cortical circuitry to experience [13]. In line, functional deficits in neurodevelopmental disorders, such as the Down and the Rett syndrome, or autism spectrum disorders have been proposed to be linked to a shift in the balance between excitation and inhibition in the CNS [14–17].

The majority of currently used general anesthetics potentiates neurotransmission via the GABA A receptor complex and/or inhibit glutamatergic signaling via the blockade of NMDA receptors [18]. Given the important role of GABAergic and glutamatergic signaling during brain maturation [19], an intriguing possibility is that exposure to general anesthetics during critical periods of development might interfere with neural circuitry assembly. We tested here this hypothesis by examining spine density and dynamics following application of anesthetics or by applying antagonists of excitatory receptors. Using in vivo and in vitro analyses, we find that these pharmacological approaches lead to a rapid regulation of spine and synapse number during critical periods of cortical development. We show that this effect (i) is produced through an enhanced rate of spine and filopodia growth and a better long-term stabilization of newly formed spines, (ii) is lasting and (iii) results in the formation of functional synapses. Altogether, these results reveal that general anesthetics-induced modulation of neural activity initiates substantial changes in synapse number and dynamics, shaping thereby cortical connectivity during critical periods of development. Importantly, these new data also raise essential questions with regard to the debate...
about the safety and cognitive consequences of administering anesthetics to young infants.

Results

To examine the role of the general anesthetics on spine dynamics, we carried out both in vivo and in vitro experiments using different pharmacological tools including anesthetics which either enhance GABAergic transmission (midazolam, propofol) or interfere with excitatory NMDA dependent responses (ketamine). For in vivo analyses, spine density and morphology were analyzed using the transgenic H-line mice expressing the yellow fluorescent protein (YFP) in distinct subsets of cortical and hippocampal neurons from the second postnatal week [20]. Mice were subjected to a 5 h anesthesia at different ages and then sacrificed, fixed through perfusion and spine characteristics analyzed.

In mice that did not undergo anesthesia, we found, consistent with previous reports [21–23], that there was a significant increase in protrusion density on tufted apical dendrites of layer 5 pyramidal neurons of the somatosensory cortex (SSC) between postnatal day (PND) 15 and 30 (Fig. 1a, b). Dendritic protrusion density increased by 38 ± 5% from 0.52 ± 0.02 to 0.72 ± 0.02 protrusions μm⁻¹ between PND 15 and PND 20 (Ctrl, open column: P<0.05), and then remained unchanged at PND 30 (0.69±0.07). At PND 15, 7.6±2% of dendritic protrusions were long and thin filopodia, and their proportion progressively decreased to 3.75±1% by PND 30 (P<0.001).

In marked contrast, animals sacrificed at the end of a 5 h general anesthesia made with midazolam (25 mg/kg) at PND 15 showed a dramatic, two-fold increase in protrusion density with regard to non-anesthetized mice (Fig. 1a, b). This density increase concerned both dendritic spines (0.40±0.01 in control vs 0.94±0.06 μm⁻¹ in midazolam group) and filopodia (0.04±0.02 in control vs 0.16±0.01 μm⁻¹ in midazolam group). The higher protrusion density observed in the midazolam group was associated with a significant decrease in mean spine head diameter (0.62±0.03 μm, control vs 0.41±0.01 μm, mida, P<0.001 Fig. 1c). This effect was primarily due to an important increase in the number of spines with smaller head diameters, representing, most probably, newly formed spine populations (Fig. 1d). Thus a 5 h-long anesthesia with midazolam at PND 15 was sufficient to profoundly increase protrusion density on tufted shafts of layer 5 pyramidal neurons.

We then tested whether anesthesia produced similar effects at different ages by exposing mice to a 5 h midazolam anesthesia at

Figure 1. Midazolam increases protrusion density and reduces protrusions head width in the somatosensory cortex in vivo. (a) Representative confocal images (3D volume rendering) showing apical dendritic shafts in control and midazolam-treated animals at PND 15 and PND 30. Scale bar: 5 μm. (b) Protrusion density of pyramidal neurons of the SCC of mice sacrificed at PND 15; 20 and 30 (ctrl; +: P<0.05, Kruskal-Wallis with Dunn post tests). The increase in protrusion density is age-dependent in midazolam injected animals (Mida) compared to control groups (n = 4 animals per group, 4726 protrusions; two-way ANOVA with Bonferroni post tests). (c) Mean dendritic spine head diameter for midazolam-injected (Mida) compared to non-anesthetized mice (Ctrl) at PND 15, 20 and 30 (n = 4 animals per group, 1695 protrusions; **P<0.01, ***P<0.001, two-way ANOVA with Bonferroni post tests). (d–f) Frequency distribution histogram of spine head diameter shows that the effects observed in (b, c) are primarily due to an increase in the number of spines with small heads (each thick mark on x axis is a 0.1 μm interval). doi:10.1371/journal.pone.0007043.g001
PND 20 and 30. As illustrated in Fig. 1b, midazolam still induced a significant, but smaller increase in protrusion density on apical tufted dendrites at PND 20 (30±2% at PND 20 vs 104±20% at PND 15; P<0.001). At PND 30 however, midazolam anesthesia no longer modified protrusion density (Fig. 1a, b). Mean spine head diameters were significantly reduced following a 5 h midazolam anesthesia at PND 20, and less at PND 30 (mean decrease at PND 15: 35±1.2%; at PND 20: 37±4%; at PND 30: 10±2%; P<0.01, Fig. 1c). At all age groups, midazolam-induced decrease in average spine head diameter was primarily due to an increase in the number of spines with smaller head diameters (Fig. 1d–f). Together, these data suggest that the impact of anesthesia on dendritic spines is dependent on developmental age.

We then wondered whether other anesthetics also reproduced these effects. For this, we used propofol (50 mg/kg), another GABA<sub>A</sub> receptor agonist, and ketamine (30 mg/kg), an NMDA receptor antagonist. Similarly to midazolam, these two drugs also induced an age-dependent significant increase in dendritic spine density (Fig. 2a, b, P<0.05) as well as a simultaneous decrease in average dendritic spine head diameter (Fig. 2c: P<0.05). Altogether, these results strongly suggest that several general anesthetics, affecting the excitation/inhibition balance through different mechanisms, can alter in a similar fashion the development of dendritic spines.

To examine if the anesthetics-induced changes were lasting, mice received midazolam anesthesia at PND 15 and were then sacrificed at PND 20 and 30. At PND 20, dendritic protrusion density on apical tufts was still significantly higher in animals that received midazolam 5 days before than in aged-matched non-anesthetized controls (Fig. 3a, P<0.05). At PND 30 however, no such difference could be detected. Note that the increase in spine density promoted by anesthesia at PND 15 is sufficiently lasting that it now suggests a pruning effect between PND 15 and PND 30, while protrusion density in non-anesthetized animals actually increased during this period. Anesthesia-induced changes in spine head diameter, observed following a 5 h midazolam anesthesia at
PND 15, was still present at PND 20 (P<0.001) and, although to a lesser extent, also at PND 30 (mean decrease compared to age-matched controls at PND 20: 30±4.5%; at PND 30: 23±2.1%; Fig. 3b). The changes induced by a general anesthesia in young animals may thus last for several days.

We then wondered whether these changes were specific to the SSC, and therefore evaluated the effects of these three anesthetics on the development of dendritic protrusions in another cortical region, area CA1 of the hippocampus, where cells also express YFP. At PND 15, we found that all anesthetics induced within 5 h a significant increase in protrusion density on apical dendrites of CA1 pyramidal neurons (Fig. 4a, b, P<0.05). The effect was associated with a concomitant increase in the percentage of spines with smaller head diameter (Fig. 4c, P<0.001). Similarly to observations made in the SSC, the effects of the three anesthetics on CA1 dendritic spines were dependent on the age of the animals (Fig. 4b), thus reproducing the changes observed in the SSC.

To further investigate the dynamics and mechanisms underlying these effects, and since in vivo imaging is precluded by the need of anesthesia, we performed repetitive confocal imaging of eGFP transected CA1 pyramidal neurons in 2–3 weeks old hippocampal slice cultures, a model also showing a developmental regulation of spine turnover [24]. We found that a 5-hour-long bath application of midazolam (30 μM) rapidly induced a significant increase in protrusion density that still persisted 48 hours after application (Figure 5a, b, P<0.001). To understand the dynamic changes underlying this effect we measured the proportion of newly formed and lost protrusions detected on identified dendritic segments between observations made at 0, 5, 24 and 48 hours. We found that midazolam triggered a striking increase in protrusion formation (Fig. 5c, 0–5 h, P<0.001), whereas the loss of protrusions remained unchanged (Fig. 5d, 0–5 h, P>0.05). This effect was however transient and limited to the presence of midazolam, since 24 h after removal of midazolam, the rate of new spine formation and loss was again similar to those found in non-treated controls (Fig. 5b, c: 24–48 h, P>0.05). We also determined the number of newly formed spines per μm after one hour-treatment with midazolam. This growth rate was more than 9 times higher compared to control conditions (26.8±9.7 vs 2.8±1.1 new spines per μm per hour, respectively, Fig. 5e). Interestingly, the proportion of filopodia among new protrusions was not changed and remained low (Fig. 5f, P>0.05). We then also analyzed how midazolam treatment affected spine stability by measuring the proportion of spines persisting over a period of 48 h. Pre-existing spines, present before the application of midazolam, were not affected in their stability by midazolam treatment, indicating that the drug probably did not modify pre-established circuits (Fig. 5g, P>0.05). However, spines formed in the presence of midazolam had a two times higher probability to remain stable than newly formed spines appearing during the same period in control untreated cultures (Fig. 5h, P<0.01). These data indicate that midazolam-induced increase in protrusions density is primarily due to a rapid increase in spinogenesis followed by an enhanced stabilization of newly formed protrusions. These combined, but transient effects probably account for the fact that the midazolam-induced increase in spine density lasted for several days.

We next tested if new spines formed following midazolam application could be functional. After bolus loading of cells with the calcium indicator Fluo-4 AM, functionality of the new spines detected 24 h after midazolam application was tested by stimulation of Schaffer collaterals and recordings of calcium signals in individual spines (See Methods). Out of 43 new spines tested (4 cells), 22 responded to stimulation by an increase in intracellular calcium (Fig. 6a, b). Spines that did not show calcium responses were thus either silent, non-functional or more simply, the presynaptic axon could not be stimulated through the depolarizing current pulses. Under control conditions, the same proportion of spines also failed to show calcium responses (Fig. 6a–c), indicating that newly formed spines have the same probability to be functional as pre-existing spines. Furthermore, the responses of newly formed spines were comparable in amplitude with responses of neighboring pre-existing spines (Fig. 6d), and were abolished by blocking fast glutamatergic transmission with AP5 and NBQX (Fig. 6e). Therefore, a significant fraction of spines formed after midazolam treatment are functional and integrated in the pre-existing network within 24 h, thus probably accounting for the long-lasting effect on spine density of anesthetics.

Finally, we evaluated whether exposure to specific inhibitors of ionotropic glutamate receptors also increased protrusion density. As seen on Figure 7a, in vivo intraperitoneal administration of the

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Figure 4. Effects of midazolam, propofol and ketamine on dendritic protrusions in the hippocampus in vivo. (a) 3D volume rendering of confocal microscopy images of apical dendrites of CA1 hippocampal neurons of PND 15 mice in control (left) and midazolam-anesthetized (right) conditions. Scale bars: 1 μm. (b) Protrusion head density of pyramidal neurons of mice sacrificed at PND 15; 20 and 30 just after a 5 h-anesthesia with midazolam (Mida) propofol (Propo) or ketamine (Keta) compared to control conditions (ctrl). (c) Protrusion head width in PND 15 mice CA1 neurons after 5 h-midazolam (Mida), propofol (Propo) or Ketamine (Keta) anesthesia compared to control group. (b, c): n = 4 animals per group, 1291 spines; ***: P<0.001, *: P<0.05, two-way ANOVA with Bonferroni post tests.
AMPA/kainate antagonist NBQX (30 mg/kg) to PND 15 mice rapidly and significantly increased the number of protrusions in the SSC (P<0.001). In line, in hippocampal slice culture, a 5 h incubation with NBQX (20 μM) also induced an increase in protrusion density (Fig. 7b, P<0.05). In addition, blockade of NMDA receptor with MK801 (100 μM) also increased protrusion density within 5 h (Fig. 7b, P<0.01) whereas simultaneous blockade of both excitatory and inhibitory activity by TTX (1 μM) did not reproduce the effect (Fig. 7b). Taken together, these results further suggest the importance of the balance between excitation and inhibition rather than activity per se in the regulation of spinogenesis.

Discussion

The present study provides evidence that exposure to general anesthetics rapidly induces dendritic spine growth and the
formation of functional synapses during critical periods of early postnatal life in the rodent brain. This is to our knowledge the first demonstration that a few hours of anesthesia, can almost double the spine density on cortical neurons and thus the complexity of cortical synaptic networks. The magnitude of the effect, its persistence over several days and its restriction to critical periods of development certainly raises important questions as to the mechanisms involved, but also regarding the possible functional consequences of such a massive synaptic reorganization. The fact that both anesthetics that enhance GABAA receptor-mediated inhibition and those blocking NMDA receptor-mediated excitation promoted spine growth suggests that the balance between excitation and inhibition was the major determinant of this synaptic growth process. This could be in line with previous results indicating changes in spine and synapse properties or number upon prolonged interference with excitatory or inhibitory receptors, [25–30]. However neither the context, nor the dynamics of these previously reported changes makes them really comparable to our study, and clearly more experiments will be required to identify the signaling mechanisms implicated in these regulations.

The increase in spine and synapse density reported here following reduction of excitation in the tissue points to similarities with the homeostatic scaling effect described for excitatory receptors where pharmacological blockade of excitation promotes expression of additional receptors at all synapses [31,32]. In both cases the regulations involved tend to maintain neuronal activity, either by increasing sensitivity to glutamate at existing synapses, or as shown here, by increasing synapse number. There are however also different specificities between these mechanisms. The synaptic changes reported here are surprisingly fast and predominantly observed during a critical period of development, unlike the receptor scaling effect and unlike the changes in spine density or properties reported following long-term, chronic blockade or genetic manipulation of NMDA or AMPA receptors [25,27,29]. Additionally, unlike receptor scaling [32], spine growth triggered...
by interference with the excitation/inhibition balance is achieved by blockade of both AMPA and NMDA receptors, indicating that it is very sensitive to NMDA dependent mechanisms. The fast regulation of synapse number reported here could thus maintain the level of excitatory activity required for ensuring plasticity-mediated mechanisms. This could be particularly important at times where synapse turnover is high and selection of correct inputs is a central issue. This could also provide some clue regarding the physiopathological mechanisms possibly relating modulation or shifts of the excitatory/inhibitory balance to neuropsychiatric disorders such as proposed in Down or Rett syndromes as well as in autism spectrum disorders [14–17].

There are two additional important aspects related to this homeostatic mechanism. First, the magnitude and rapidity of the effect produced probably account for some of the discrepancies reported in the literature concerning the development of spines during the first weeks of life. Recent studies using in vivo 2-photon imaging have reported mechanisms of spine pruning between the second, third and fourth week of life in mice [33]. This is however at variance with previous anatomical data obtained through classical staining and brain fixation methods that reported a continuous increase in spine density in many cortical regions during the same periods [21–23]. Interestingly, in vivo imaging studies have been carried out in mice that had undergone a long anesthesia for the preparation of 2-photon imaging approach. This manipulation probably boosted spine density in these young mice and, as shown by our data, analyses in animals anesthetized around PND 15 then suggest a subsequent pruning of spines over the next 2 weeks, while analyses of mice that did not undergo anesthesia conversely indicate a progressive increase in spine density. Accordingly, our data indicate that anesthetics in young animals may induce important structural modifications that might be source of misinterpretations regarding analyses of spine number or spine morphology.

The second important aspect of this work relates to the effects that anesthetics might produce in human clinical use when applied during critical periods of development in infants. Our work shows that all anesthetics tested, which all interfere with the excitation/inhibition balance, promote a rapid increase in spine synapse density, but also affect spine morphology. These two effects were lasting for several days in young mice, and certainly contributed to modify cortical networks since many new spines turn out to be functional synapses. Although the behavioral significance of these changes remains to be determined, they might raise concern about the millions of human infants that receive general anesthesia during this developmental period every year worldwide. Indeed, an increasing number of clinical reports suggest the possibility of adverse long-term neurocognitive outcome in the population of young infants undergoing anesthesia/surgery [34–36].

Altogether, this study demonstrates that exposure to general anesthetics during critical periods of development increases dendritic spine number and suggests a mechanism for the rapid modulation of synaptogenesis via the modulation of the excitation/inhibition balance by these drugs. This new mechanism is likely to play a critical role in the regulation of the formation of neural circuits and may help understand dysfunctions related to conditions under which alterations of the excitation/inhibition balance may occur.

Materials and Methods
Ethics statements
All procedures were done in accordance with the animal care and use guidelines of the Geneva Cantonal Veterinary Office and the University of Geneva.

Experimental animals and anesthesia procedure
Mice expressing the yellow fluorescent protein (YFP) in layer 5B cortical neurons (H-line) [20] [37] were purchased from the Jackson laboratory (Bar Harbor, Maine, USA). Animals were group-housed and bred in the animal facilities of the University of Geneva Medical School under light (12 h light/dark cycle) and temperature (22±2°C) controlled conditions. Food and water were available ad libitum. Every effort was made to minimize the number of animals used and their suffering.

General anesthesia was induced by intraperitoneal injection of midazolam (25 mg/kg), propofol (50 mg/kg) or ketamine (30 mg/kg). At these concentrations, these drugs induced deep sedation (absence of the righting reflex >10 s) for approximately 90 minutes. Subsequent injections of midazolam, propofol and ketamine at concentrations of 15 mg/kg, 25 mg/kg and 15 mg/kg respectively, allowed maintaining anesthesia for an additional 90–120 minutes. Thus, to provide a 5-hour-long anesthesia, animals received a total of 3 intraperitoneal injections of anesthetics at 90 minutes intervals. To ensure that this above-mentioned anesthesia procedure did not induce significant metabolic and/or respiratory compromise, we performed arterial blood sampling from the ascending aorta in three PND 16 animals of each anesthesia protocol at the end of the 5-hour-long anesthesia procedure. This analysis revealed that physiological parameters, such as pO2, pCO2, lactate and blood sugar concentrations remained within physiological range. Control sham-treated animals received 3 intraperitoneal injections of physiological saline at 90 minutes intervals and have undergone the same handling and maternal separation as anesthetized animals. In a set of preliminary experiments we determined that this “sham treatment paradigm” does not induce any changes in dendritic spine density or morphology in the somatosensory cortex compared with control non-treated animals sacrificed immediately following maternal separation. To avoid hypothermia, animals were kept during the whole procedure on a homeothermic blanket
system in order to maintain body temperature at 37°C (Harvard Apparatus, Holliston, Massachusetts, United States).

**Histological processing and immunohistochemistry**

Depending on experimental protocols, animals were sacrificed either immediately following the 5-hour-long anesthesia period or at later time points, as indicated, using a lethal dose of pentobarbital (100 mg/kg) followed by intracardiac perfusion of a cold (4°C) 0.9% saline and, subsequently, by a 4% paraformaldehyde solution. Brains were then extracted from the skull and post-fixed overnight in a 4% paraformaldehyde solution at 4°C. Following fixation, brains were washed 3×1 h in a 0.1 M phosphate-based saline (PBS; pH 7.2) solution and then cryoprotected with sucrose (30%). 20 μm-thick sections were cut using a cryostat (Leica, Wetzlar, Germany) and plated on gelatin-coated slides. To strengthen signal intensity of YFP-positive dendritic shafts, sections were incubated overnight at 4°C with a polyclonal rabbit anti-GFP antibody (1:1000; Molecular Probes, Invitrogen, Carlsbad, California, United States) in PBS/0.5% bovine serum albumine (BSA)/0.3% Triton X-100. Sections were then rapidly washed 3 times in PBS and incubated for 90 minutes with an Alexa-conjugated secondary antibody (Molecular Probes, Carlsbad, California, United States) diluted in PBS/0.5% bovine serum albumine (BSA).

**Organotypic hippocampal slice cultures**

Transverse hippocampal organotypic slice cultures (400 μm thick) from 6–7 day-old rats were prepared as described [38], using a protocol approved by the Geneva Veterinarian Office (authorization 31.1.1007/3129/0) and maintained for 11–18 days in a CO2 incubator at 33°C (authorization 31.1.1007/3129/0) and maintained mostly unchanged over the different days of observation. Control experiments showed that transfection and viability over periods of weeks. We focused on dendritic segments of about 40 μm in length and located between 100 and 350 μm from the soma on secondary dendrites using a 40× objective and a 10× additional zoom (final resolution: 25 pixels per micron; steps between scans: 0.4 μm). For calcium imaging of spine activity, new spines were identified 24 h after midazolam application in mRFP transfected cells. Cells were then loaded with the cell permeable calcium indicator Fluo-4 AM (F-14201, Invitrogen). For this, 50 μg of Fluo-4 AM was dissolved in 10 μl Phromine (F-127, Invitrogen) and then diluted in 90 μl of standard pipette solution (150 mM NaCl, 2.5 mM KCl, 10 mM Hepes) for a final dye concentration of 500 nM. A standard patch pipette was then filled with 10 μl of dye solution and placed at a distance of about 10 μm from the soma. Dye was ejected by short pulses of pressured air at a frequency of 3 per minute during half an hour. Calcium transients in identified new spines and neighboring pre-existing spines were then recorded using line scans through the spine heads obtained during application of stimulation pulses to Schaffer collaterals. Stimulating electrode was always placed at minimum at 500 μm from the dendritic tree of the Fluo-4 AM loaded cell in order to preclude direct electrical stimulation of the cell. In all experiments carried out, we also verified that spines activated by stimulation were always surrounded by other silent, non-activated spines in order to exclude global activation effects. Spines located too close to each other were excluded from analysis. Confocal aperture was set to the minimum during line scans, and matching with the mRFP fluorescence in the red channel was systematically checked. For each spine tested, 2 to 7 linescans of 8 seconds of duration were performed, with electrical stimulation pulse occurring automatically at 3 seconds. Calcium responses were measured as ΔF/Fo, where Fo is the mean fluorescence measured during the baseline.

**Data analysis**

In this study we refer to protrusions, whenever analyses were carried out by considering filopodia and spines. Filopodia were defined as protrusions devoid of enlargement at the tip (overall they represented less than 5% of total protrusion number), while we classified as spines all protrusions exhibiting an enlargement at the tip. All turnover and stability analyses were carried out by scrolling across single z-stacks of raw images using a plug-in specifically developed for OsiriX software [http://www.osirix-viewer.com] except for density analysis on fixed tissue, which was performed on the LSM image view software (Carl-Zeiss, Göttingen, Germany). The measures of density, turnover and stability were carried out by analyzing all protrusions, i.e. filopodia and spines. We counted protrusions located behind each other on z-stacks whenever distinction was possible. For turnover analysis, we counted as new protrusions all new structures (spines or filopodia) appearing between two observations (5 h or 24 h) and characterized by a length of >0.4 μm. All filopodia were counted as separate protrusions. For disappearances, we counted all protrusions (spines or filopodia) that could no longer be identified on the next observation. Dubious situations due to possible changes in protrusion shape, size or orientation were discarded, but overall accounted for only a small number of cases (less than 1%). To further ensure reliability of analyses, all measurements of spine turnover and stability were carried out blind by two experimenters. Comparisons of the analyses made in this way showed variations in the results that were less than 3%. Furthermore, we used high numbers of n for both cells and spines and labeled all new or lost protrusions directly on the raw data to allow multiple checks.

Due to the lack of survival of filopodia on several days, stability analyses only included pre-existing spines, i.e. spines present at the
beginning of the experiment. Note that for illustration purposes, images presented in figures are maximum intensity projections of z stacks with volume rendering, further treated with a Gaussian blur filter.

Statistics

All statistics are given with the standard error of the mean (s.e.m.). Normality was tested for each distribution (D’Agostino and Pearson test), and \( \alpha \) was set to 5\% for all tests.

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Acknowledgments

We thank Anthony Holtmaat for helpful discussions, Marlis Moosmayer, Beatrice King and Lorena Jourdain for technical support, and Joel Spaltenstein for the development of Osirix software plugin. The mRFP1 plasmid was a generous gift from AK Hadjantonakis and RY Tsien, University of California, San Diego.

Author Contributions

Conceived and designed the experiments: MDR PK DM LV. Performed the experiments: MDR PK AB IN PM LV. Analyzed the data: MDR LV. Contributed reagents/materials/analysis tools: DM LV. Wrote the paper: MDR AD JK DM LV.