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Genome Analysis of Endobacterium cerealis, a Novel Genus and Species Isolated from Zea mays Roots in North Spain

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Abstract: In the present work, we analyse the genomic and phenotypic characteristics of a strain named RZME27T isolated from roots of a Zea mays plant grown in Spain. The phylogenetic analyses of 16S rRNA gene and whole genome sequences showed that the strain RZME27T clustered with the type strains of Neorhizobium galegae and Pseudorhizobium pelagicum from the family Rhizobiaceae. This family encompasses several genera establishing symbiosis with legumes, but the genes involved in nodulation and nitrogen fixation are absent in its genome. Nevertheless, genes related to plant colonization, such as those involved in motility, chemotaxis, quorum sensing, exopolysaccharide biosynthesis and hydrolytic enzymes production were found. The comparatively pangenomic analyses showed that 78 protein clusters present in the strain RZME27T were not found in the type strains of its closest relatives N. galegae and P. pelagicum. The calculated average nucleotide identity (ANI) values between the strain RZME27T and the type strains of N. galegae and P. pelagicum were 75.61% and 75.1%, respectively, similar or lower than those found for other genera from family Rhizobiaceae. Several phenotypic differences were also found, highlighting the absence of the fatty acid C19:0 cyclo ω8c and propionate assimilation. These results support the definition of a novel genus and species named Endobacterium cerealis gen. nov. sp. nov. whose type strain is RZME27T.

Keywords: Endobacterium gen. nov.; Endobacterium cerealis sp. nov.; Zea mays; Spain; endophytes; genome analysis

1. Introduction

The family Rhizobiaceae [1] belongs to the class Alphaproteobacteria and currently, this family comprises the classical genera Rhizobium [2], Agrobacterium [3], Sinorhizobium [4] (later transferred to Ensifer [5]), and Allorhizobium [6] as well as the more recently described genera Shinella [7], Ciceribacter [8], Neorhizobium [9], Pararhizobium [10], Pseudorhizobium [11], Gellertiella [12] and Georhizobium [13]. Several of these genera contain species originally isolated from legume nodules, such as Rhizobium, Ensifer (Sinorhizobium), Allorhizobium, Shinella, Neorhizobium or Pararhizobium [14–16] and from
plant tumours such as Agrobacterium [17]. However, other genera within this family have not been reported to date as legume endosymbionts, plant endophytes or plant pathogens, such as Ciceribacter, Pseudorhizobium, Gellertiella and Georhizobium [11–13]. Some genera contain species originally isolated from cereals, but to date only two species of the genus Rhizobium, Rhizobium zea [18] and Rhizobium wenxiniae [19], have been isolated from Zea mays (maize) roots.

The endophytic bacterial community of maize has been widely studied and several new endophytic species of this cereal have been described, but only two new genera, Dyadobacter [20] and Runella [21], were isolated from stems of Zea mays and they belong to the family Cytophagaceae within the Class Flavobacteria. In this work, we report the existence of a novel genus belonging to the family Rhizobiaceae, which was isolated from roots of Zea mays growing in a field of León province (Northern Spain), the largest and most productive area for maize cropping in Spain. The aims of this study were the genome analysis of this novel genus for which we proposed the name Endobacterium gen. nov. and the characterization through the analysis of genomic, chemotaxonomic and phenotypic traits of its first described species Endobacterium cerealis gen. nov. sp. nov., whose type strain, RZME27T, has been deposited in two international culture collections under the accession numbers LMG 31256T and CECT 9794T.

2. Materials and Methods

2.1. Strain Isolation

The strain RZME27T was isolated from roots of a Zea mays plant harvested six months after the sowing in a field in Riego de la Vega (León, NW Spain, 42°23′21″ N 5°58′56″ O). The roots were surface disinfected with 70% ethanol for 1 min, 2% NaClO for 3 min and 70% ethanol for 30 s. Then, the roots were rinsed five times with sterile distilled water and crushed into a tube containing 10 mL of sterile phosphate-buffered saline (PBS) at pH7, which was further maintained in shaking at 160 rpm during 1 h at room temperature. Then, six serial decimal dilutions were obtained and 100 µL aliquots were spread onto Tryptic Soy Agar (TSA; BD Difco, Franklin Lakes, NJ, USA) plates and incubated for 48 h at 28 °C. In the higher dilution, a mucoid colony was picked and transferred to a new TSA plate and the isolated strain was named RZME27T. In parallel, in order to ensure their complete external disinfection, some of the surface-disinfected root samples were incubated in the same medium, showing no bacterial growth around them.

2.2. 16S rRNA Gene Phylogenetic Analysis

The 16S rRNA gene of strain RZME27T was amplified and sequenced as previously described [22] at the Sequencing DNA Service of NUCLEUS, University of Salamanca (Spain). The 16S rRNA gene sequence obtained was compared with those held in GenBank [23] and for its phylogenetic analysis, the sequence was aligned with those of the type strains of closest-related species using the ClustalW program [24]. The phylogenetic distances were calculated according to Kimura’s two-parameter model [25]. The phylogenetic trees were inferred using the neighbour joining (NJ) and maximum likelihood (ML) algorithms [26,27] and MEGA 7.09 [28] was used for all phylogenetic analyses.

2.3. Genome Sequencing, Assembling, Annotation and Analysis

The genomic DNA from a pure culture of the strain RZME27T was purified using the DNeasy UltraClean Microbial DNA Isolation Kit (Qiagen, Venlo, Netherlands) following manufacturer’s protocol. Sequencing, upon preparation of pair-end libraries, was performed on Illumina MiSeq sequencing platform (2 × 250 bp). Sequencing data was assembled using Velvet 1.12.10 [29]. Annotation was undertaken using RAST 2.0 (Rapid Annotation using Subsystem Technology) [30,31] and the NCBI (National Center for Biotechnology Information) Prokaryotic Genome Annotation Pipeline (PGAP) (https://www.ncbi.nlm.nih.gov/genome/annotation_prok/) [32,33]. The draft genome sequence of the strain RZME27T was deposited in DDBJ/EMBL/GenBank under the BioProject PRJNA576662 (accession number WIXI00000000). Data from the type strains of related species and genera were
retrieved from public databases (Table S1). The circular genome map of the strain RZME27\textsuperscript{T} and the schematic map of BLAST (Basic Local Alignment Search Tool) comparison among closest type strain genomes was generated using the CGView server (http://cgview.ca/; accessed 17th March 2020) [34,35]. Phylogenomic analysis was conducted with the Type Strain Genome Server [36]. This web-server tool employs the Genome-BLAST Distance Phylogeny method (GBDP) [37] to compare whole genome sequences at nucleotide level, allowing to calculate dDDH value and construct the phylogram. The Average Nucleotide Identity using BLAST (ANIb) value was calculated with JSpeciesWS Online Service, which is based on BLAST+(version 2.2.29+) [38]. Carbohydrate-active enzyme analysis was performed using the dbCAN2 meta server (http://cys.bios.niu.edu/dbCAN2; accessed 22th March 2020) [39]. Secondary metabolite production potential was carried out analyzing the presence in the genome of biosynthetic gene clusters (BGCs) within the tool antiSMASH version 5.1.1 [40]. KofamKOALA tool was used to annotate the genome based on the KEGG (Kyotto Encyclopedia of Genes and Genomes) database [41] Comparative genome-wide analysis of orthologous clusters and gene ontology analysis among all predicted protein-coding genes was performed using OrthoVenn2 (https://orthovenn2.bioinfotoolkits.net/home; accessed 20th March 2020) [42]. The genomes of phylogenetically related strains were retrieved from GenBank (https://www.ncbi.nlm.nih.gov/genome; accessed 15th March 2020) (Supplementary Table S1).

2.4. Fatty Acid Analysis and Phenotypic Characterization

The cellular fatty acids were analysed by using the Microbial Identification System (MIDI; Microbial ID) Sherlock 6.1 and the library RTSBA6 according to the technical instructions provided by this system [43]. The strains RZME27\textsuperscript{T}, Neorhizobium galegae HAMBI 540\textsuperscript{T}, Pseudorhizobium pelagicum R1-200B4\textsuperscript{T} and Rhizobium leguminosarum USDA 2370\textsuperscript{T} were cultured aerobically on Tryptone Yeast agar (TY) plates [44] at 28 °C and cells were collected after 48 h incubation. Gram staining was carried out by the procedure described by Doetsch [45] after 24 h of incubation at 28 °C. The strain was grown on nutrient agar (NA; BD Difco, Franklin Lakes, NJ, USA) for 48 h at 22 °C to check for motility by phase-contrast microscopy using the hanging drop method. The flagellation type was determined by electron microscopy after 48 h of incubation of strain RZME27\textsuperscript{T} on Yeast Mannitol Agar (YMA; Laboratorios Microkit, Madrid, Spain) plates at 22 °C. The cells were gently suspended in sterile water and then stained with 2% uranyl acetate and examined at 80 kV with a JEOL 1010 transmission electron microscope equipped with a Gatan Bioscan 792 digital camera from the Microscopy Service of NUCLEUS, University of Salamanca (Spain). The phenotypic characterization was performed in this study using the API ID32GN and API 20NE systems (BioMérieux, Marcv L’Etoile, France) in the conditions indicated by the manufacturer and the results were read after 48 h of incubation at 28 °C. Growth temperature range was determined by incubating cultures in YMA medium [46] at 4, 15, 28, 37 and 45 °C. Growth pH range was determined in the same medium with final pH 5, 6, 7, 8 and 9. PCA buffer (phosphate citric acid buffer; Na\textsubscript{2}HPO\textsubscript{4} 0.4 M and citric acid 0.2 M) was used to adjust the pH from 5 and 6, phosphate buffer (Na\textsubscript{2}HPO\textsubscript{4} 0.2 M and NaH\textsubscript{2}PO\textsubscript{4} 0.2 M) was used for pH 7 and TE buffer 0.2 M was used for pH 8 and 9. Salt tolerance was tested in the same medium containing 0.5, 1, 1.5, 2, 2.5, 4 and 7% (w/v) NaCl. Catalase production was assayed by using 0.3% hydrogen peroxide with one colony taken from the TY plates. Oxidase activity was detected by using N,N,N′,N′-tetramethyl-1,4-phenylenediamine dihydrochloride. The strains Neorhizobium galegae HAMBI 540\textsuperscript{T}, Pseudorhizobium pelagicum R1-200B4\textsuperscript{T} and Rhizobium leguminosarum USDA 2370\textsuperscript{T} were used as reference

3. Results

3.1. 16S rRNA Gene Phylogenetic Analysis

The comparison against GenBank of the 16S rRNA gene showed that the strain RZME27\textsuperscript{T} belongs to the family Rhizobiaceae and that it is equidistant to different species of genera Neorhizobium and
Rhizobium, showing similarity values ranging from 97% to 98%. The type strains of the type species of these genera, N. galegae, and R. leguminosarum, showed 97.4% and 95.8% similarity, respectively, with respect to the strain RZME27\textsuperscript{T}. Similarity values higher than 95% were also presented by the type strains of P. pelagicum, Agrobacterium radiobacter and Pararhizobium giardinii (Supplementary Table S2). Similarity values equal or lower than 95% were found for the type strains of the remaining genera of the family Rhizobiaceae (Supplementary Table S2). Similarity values higher than 97% also were found between the type species of Pseudorhizobium and Neorhizobium and between the type species of Ensifer and those of Ciceribacter, Gellertiella and Pararhizobium (Supplementary Table S2). The NJ and ML phylogenetic analyses of the 16S rRNA gene showed that the new strain RZME27\textsuperscript{T} clustered with the type strains of N. galegae, P. pelagicum and R. leguminosarum (Figure 1).

![Figure 1](Image)

**Figure 1.** Neighbour Joining phylogenetic tree based on 16S rRNA gene sequences (1433 nt) showing the taxonomic location of the type strain of *Endobacterium cerealis* within the family *Rhizobiaceae*. Bootstrap values calculated for 1000 replications are indicated. Bar, 1 nt substitution per 1000 nt. Accession numbers from GenBank are given in brackets. The nodes marked with filled circles were also obtained with the maximum likelihood algorithm.

### 3.2. Genome Analyses

#### 3.2.1. Genome Properties

The draft genome of strain RZME27\textsuperscript{T} is composed by 51 contigs with a total genomic length of 6,114,623 bp and a G+C content of 58.80% (Table 1, Figure 2A). Genome annotation tools NCBI PGAP and RAST pipeline identified 5893 genes and 5836 coding sequences, respectively (Table 1).
Genome subsystem within a genomic island. The genes involved in flagellar biosynthesis were found siderophore production and symbiotic genes were not found in this genome.

Genes involved in exopolysaccharide production, genes codifying degradative enzymes. However, genes related with stress tolerance were also annotated with RAST, as choline dehydrogenase (EC.1.1.99.1) and betaine poly-gamma-glutamate biosynthesis or exportation, were detected. Some genes involved in osmotic biosynthesis, such as glycinebetaine.

LuxI and LuxR gene was located within a quorum sensing operon in the strain RZME27T. In addition, other genes related with lipopolysaccharide formation were identified. Several genes involved in exopolysaccharide biosynthesis are located in the genome of strain RZME27T, have been identified. The database shows that genes are distributed into 25 subsystems according to SEED viewer in RAST (Supplementary Table S3, Figure 2B).

Figure 2. Genome circular representation and subsystems category distribution of annotated genes of the *Endobacterium cerealis* RZME27T genome. (A) From the outer circle to the inner circle, the genomic map of RZME27T shows: contigs (light grey), open reading frames (ORFs; dark green), GC skew curves (+/−; green/purple), GC content (black), Start/Stop codons (dark blue/red), coding sequences (CDSs; blue), tRNAs (light green), rRNAs (violet) and tmRNAs (light blue). (B) Annotation of RZME27T genome via the RAST server. The panel shows the annotation coverage, the categories distribution and the feature counts.

Table 1. Genome properties of the *Endobacterium cerealis*, its closest related genera and the type species of the type genus of family *Rhizobiaceae*.

| Attributes                  | *E. cerealis* RZME27T | *P. pelagicum* RI-200B4T | *N. galegae* HAMBI 540T | *R. leguminosarum* USDA 2370T |
|-----------------------------|-----------------------|---------------------------|-------------------------|-------------------------------|
| Sequence size (bp)          | 6,114,623             | 5,134,606                 | 6,455,027               | 7,851,935                     |
| Number of contigs           | 51                    | 62                        | 2                       | 108                           |
| GC content (%)              | 58.8                  | 62.8                      | 61.3                    | 60.6                          |
| Longest contig size         | 649,048               | 392,382                   | 4,647,962               | 1,820,721                     |
| Shortest contig size        | 285                   | 4118                      | 1,807,065               | 502                           |
| N50 value                   | 345,671               | 184,300                   | 4,647,962               | 413,138                       |
| L50 value                   | 7                     | 11                        | 1                       | 5                             |
| Total number of genes       | 5893                  | 4509                      | 6248                    | 7753                          |
| Total CDSs                  | 5836                  | 4458                      | 6,181                   | 7699                          |
| Number of coding sequences/CDSs (with protein) | 5705 | 4417 | 6014 | 7312 |
| Pseudogenes                 | 131                   | 41                        | 167                     | 387                           |
| Number of RNAs (tRNAs, rRNAs, ncRNAs) | 57 (49, 4, 4) | 51 (47, 3, 1) | 67 (51, 9, 7) | 54 (46, 4, 4) |

3.2.2. Genome Mining

Using the RAST pipeline, we observed that genes are distributed into 25 subsystems according to SEED viewer in RAST (Supplementary Table S3, Figure 2B). However, this only represent the 23% of the genome, being the 77% of the genes unassigned to a subsystem. Nonetheless, we used manual mining to identify genes of interest within the genome of the strain RZME27T, even if these genes are not assigned to a subsystem. We identified genes involved in motility, chemotaxis and exopolysaccharide production, genes codifying degradative enzymes. However, genes related with siderophore production and symbiotic genes were not found in this genome.

Genes involved in pilus formation and in the type IV secretion system were found in the same genome subsystem within a genomic island. The genes involved in flagellar biosynthesis were found
in a genome region containing genes codifying for flagellar biosynthesis proteins (FliR or FlhAB), flagellar basal-body rod proteins (FlgABCDFG) and flagellar motor rotor proteins (MotAB and FliMN). These genes are located in a specific region with chemotaxis genes (che) regulated by the luxR gene, which is a transcriptional regulator sensible to quorum sensing molecules. In addition, a luxl gene was located within a luxR gene in a quorum sensing operon in the strain RZME27T.

Several genes involved in exopolysaccharide biosynthesis are located in the genome of strain RZME27T, such as exoD and exoZ. In addition, other genes related with lipopolysaccharide biosynthesis, such as wadC, and lipopolysaccharide exportation, such as lptABDFG, have been annotated. Also, genes codifying other capsule formation related enzymes, such as those involved in poly-gamma-glutamate biosynthesis or exportation, were detected. Some genes involved in osmotic stress tolerance were also annotated with RAST, as choline dehydrogenase (EC.1.1.99.1) and betaine aldehyde dehydrogenase (EC.1.2.1.8), involved in the synthesis of the osmoprotectant glycinebetaine.

Using the dbCAN2 meta server, we identified a total of 73 genes encoding glycosyl hydrolases (GH) and 60 genes encoding glycosyl transferases (GT), amongst other carbohydrate-related enzymes and activities (Supplementary Table S4). The dbCAN2 meta server annotation also revealed the existence of 24 identified signal peptides.

The genome analysis reveals that this strain harbors three genes involved in cellulose biosynthesis related to those forming the celABC operon of Rhizobium strains. These genes, encoding for a cellulose synthase catalytic subunit [Uridine 5'-diphosphate(UDP)-forming] (EC 2.4.1.12), a cyclic di-GMP binding protein precursor and a ß-1,4-glucanase (cellulase) (EC 3.2.1.4), showed similarities of 79.84, 66.43 and 68.70% with the ones located in the genome of Rhizobium etli CFN42T. Annotation using the KofamKOALA online tool also revealed the existence of complete metabolic pathways as well as many genes classified into pathways that are not complete (Supplementary Table S5). This annotation revealed the presence of high number of genes forming complete pathways related to carbohydrate metabolism. We also found genes potentially involved in the nitrate catabolism, but those genes are not forming a complete metabolic pathway. No genes involved in nodulation or nitrogen fixation were found using this annotation tool.

The potential production of secondary metabolites was scanned with antiSMASH, finding three possible gene clusters involved in the biosynthesis of a homoserine lactone, a terpene and a TfuA-related toxin, respectively. The first gene cluster shows between 100% and 63% similarity with those located in other strains from genus Rhizobium as Rhizobium sp. NFR07, Rhizobium leucanec CsPAO 29.8, Rhizobium freirei PRF 81T, Rhizobium tropici CIAT 899T and Rhizobium lusitanum P1-7T. The cluster related with terpene biosynthesis also reveals around 80% similarity with gene clusters found in strains of Neorhizobium galegae and Rhizobium sp. On the contrary, the gene cluster predicted to be implicated in the synthesis of a TfuA-related toxin revealed only low similarity (12%) with gene clusters found in other members of the genera Rhizobium and Ensifer.

3.2.3. Genetic Relatedness and Pangenome Analysis

The GBDP phylogenomic tree confirmed the phylogenetic position of the strain RZME27T derived from the 16S rRNA gene analysis showing that this strain belongs to a cluster together the type strains of N. galegae HAMBI 540T and P. pelagicum R1-200B4T (Figure 3). The ANIb values between the whole genome of the strain RZME27T and those of the type strains of the type species from the family Rhizobiaceae genera were lower than 76% (Table 2). The ANIb values between the strain RZME27T and its closest relatives N. galegae and P. pelagicum were 75.6% and 75.1%, respectively. Similar or slightly higher values were found between the type species of Ensifer and Pararhizobium, Pararhizobium and Shinella, Ensifer and Shinella and Pseudorhizobium and Neorhizobium (Table 2). These results support the affiliation to the strain RZME27T to a new genus within the family Rhizobiaceae.
Table 2. Calculated ANIb values for available genomes of the type strains from the type species of the genera included in the family *Rhizobiaceae* (the accession numbers for these genomes are in parentheses). *Endobacterium cerealis* RZME27T (WIXI00000000), *Neorhizobium galegae* HAMBI 540T (HG938353.1), *Pseudorhizobium pelagicum* R1-200B4T (JOKI01000001.1), *Rhizobium leguminosarum* USDA 2370T (QBLB00000000.1), *Georhizobium profundi* WS11T (CP032509.1), *Gellertiella hungarica* DSM 29853T (IMG Ga0373349), *Ciceribacter lividus* DSM 25528T (QPIX00000000.1), *Shinella granuli* DSM 18401T (SLVX00000000.1), *Ensifer adhaerens* Casida AT (CP015880.1), *Allorhizobium undicola* ORS 992T (HGXQ00000000.1), *Agrobacterium radiobacter* LMG 140T (MRDG01000013.1), *Pararhizobium giardinii* H152T (ARBG00000000.1).

| Species | RZME27T | HAMBI 540T | R1-200B4T | USDA 2370T | WS11T | DSM 29853T | DSM 25528T | DSM 18401T | Casida AT | ORS 992T | LMG 140T | H152T |
|---------|---------|------------|-----------|------------|-------|-----------|-----------|-----------|----------|----------|---------|-------|
| RZME17T | 100     |            |           |            |       |           |           |           |          |          |        |       |
| HAMBI 540T | 75.61  | 100        |           |            |       |           |           |           |          |          |        |       |
| R1-200B4T | 75.09  | 75.40      | 100       |            |       |           |           |           |          |          |        |       |
| USDA 2370T | 73.34  | 74.49      | 73.03     | 100        |       |           |           |           |          |          |        |       |
| WS11T | 70.91  | 70.23      | 70.69     | 70.24      | 100   |           |           |           |          |          |        |       |
| DSM 29853T | 72.78  | 72.34      | 73.02     | 72.56      | 70.75 | 100       |           |           |          |          |        |       |
| DSM 25528T | 73.82  | 73.47      | 74.19     | 73.15      | 70.87 | 73.33     | 100       |           |          |          |        |       |
| DSM 18401T | 73.14  | 73.53      | 73.40     | 73.92      | 71.08 | 73.55     | 73.70     | 100       |          |          |        |       |
| Casida AT | 72.75  | 73.20      | 73.03     | 73.64      | 70.87 | 72.82     | 73.04     | 76.03     | 100      |          |        |       |
| ORS 992T | 72.97  | 72.70      | 72.65     | 72.81      | 69.94 | 72.50     | 73.27     | 72.75     | 72.23    | 100      |        |       |
| DSM 30147T | 74.30  | 73.77      | 73.39     | 73.29      | 70.10 | 72.26     | 73.11     | 73.20     | 72.63    | 72.76    | 100    |       |
| H152T | 72.90  | 73.17      | 72.73     | 74.57      | 70.70 | 72.57     | 72.97     | 75.43     | 75.74    | 71.96    | 72.52  | 100    |
R. leguminosarum of RZME27

Possible gene clusters involved in the biosynthesis of a homoserine lactone, a terpene and a complete pathways related to carbohydrate metabolism. We also found genes potentially involved relevant difference with respect to the rest of its related reference strains (Table 3). Recorded in Table 4. The strain RZME27 T differs with respect to the type strain of description of the new taxon and the differences with respect to its phylogenetic related genera are strain RZME27 T and its closest relatives from the family Rhizobiaceae. The absence of C 19:0 cyclo 6c); * Fatty acids present in amounts lower than 1% in all species.

The genome analysis reveals that this strain harbors three genes involved in cellulose biosynthesis related to those forming the flagellum (Figure S1). The phenotypic characteristics of the strain RZME27 T are reported in the

Phylogeny method (GBDP) tool and retrieved from the TYGS website. The tree was inferred with FastME 2.1.6.1 from GBDP distances calculated from genome sequences. The branch lengths are scaled in terms of GBDP distance formula d5.

The genomic comparison among the genomes of the strains E. cerealis RZME27T, N. galegae HAMBI 540T, P. pelagicum R1-200B4T and R. leguminosarum USDA 2370T is depicted in Figure 4A, showing that RZME27T genome is closer to those of the type strains of N. galegae and P. pelagicum than to the genome of R. leguminosarum USDA 2370T.

Figure 3. Whole genome based phylogenomic tree constructed with the Genome-BLAST Distance Phylogeny method (GBDP) tool and retrieved from the TYGS website. The tree was inferred with FastME 2.1.6.1 from GBDP distances calculated from genome sequences. The branch lengths are scaled in terms of GBDP distance formula d5.

Figure 4. Comparative genomics among the genomes of E. cerealis RZME27T, N. galegae HAMBI 540T, P. pelagicum R1-200B4T and R. leguminosarum USDA 2370T. (A) Schematic representation of BLAST comparisons amongst the four type species. From the outer to the inner circle: RZME27T (pink), HAMBI 540T (blue), R1-200B4T (green) and USDA 2370T (gold). (B) The Venn diagram and the bar chart below show the number of shared and unique orthologous amongst the four genomes.
The results of the comparative pan-genomic analysis are shown in a Venn diagram including the strain RZME27^T and its close related species (Figure 4B). The analysis shows a “core” genome composed by 2360 clusters of orthologous, most of them annotated as clusters of proteins with functions associated to cellular metabolic processes, motility, colonization and membrane exchange specialized systems. We found 78 protein clusters, which have been identified only in the strain RZME27^T, whereas only 51 and 29 clusters were found, respectively, in its closest relative type strains of P. pelagicum and N. galegae (Figure 4B).

3.3. Fatty Acid Analysis and Phenotypic Characterization

The results of the fatty acid analysis are recorded in Table 3. The major fatty acids of the strain RZME27^T are those from summed feature 8 (C_{18:1}ω6c/C_{18:1}ω7c) and the C_{16:0} as occurs in its closest genera within family Rhizobiaceae. The absence of C_{19:0} cyclo ω8c in strain RZME27^T is the most relevant difference with respect to the rest of its related reference strains (Table 3).

Table 3. Cellular fatty acid composition of Endobacterium cerealis, its closest related genera and the type species of the type genus of family Rhizobiaceae.

| Fatty Acid  | E. cerealis  | N. galegae  | P. pelagicum  | R. leguminosarum  |
|------------|--------------|-------------|---------------|-------------------|
|            | RZME27^T     | HAMBI 540^T | R1-200B4^T    | USDA 2370^T      |
| C_{16:0}   | 8.86         | 11.58       | 7.69          | 7.18              |
| C_{17:0}   | 0.10         | 1.91        | nd            | nd                |
| C_{18:0}   | 2.08         | 2.28        | 2.26          | 12.71             |
| C_{16:0} 3OH | 2.14        | 2.54        | nd            | nd                |
| C_{18:0} 3OH | 1.30        | 1.60        | 1.49          | 0.99              |
| C_{18:1}ω7c 11-methyl | 0.38 | 0.55 | 3.04 | 7.06 |
| C_{19:0} cyclo ω8c | nd | 10.76 | 4.37 | 3.74 |
| Summed feature 2^1 | 5.18 | 4.63 | 4.41 | 3.59 |
| Summed feature 3^2 | 1.78 | 0.63 | 1.49 | 1.19 |
| Summed feature 8^3 | 77.70 | 61.14 | 73.84 | 62.77 |

1 Summed feature 2: (C_{14:0} 3OH/C_{16:1} iso l/C_{12:0} aldehyde); 2 Summed feature 3: (C_{16:1}ω7c/C_{16:1}ω6c); 3 Summed feature 8: (C_{18:1}ω7c/C_{18:1}ω6c); * Fatty acids present in amounts lower than 1% in all species are not shown.

The cells of strain RZME27^T are Gram negative straight rods and motile by means of one polar flagellum (Figure S1). The phenotypic characteristics of the strain RZME27^T are reported in the description of the new taxon and the differences with respect to its phylogenetic related genera are recorded in Table 4. The strain RZME27^T differs with respect to the type strain of N. galegae in the colonies colour, nitrate reduction and l-rhamnose assimilation, with respect to the type strain of P. pelagicum in assimilation of malate, inositol, melibiose, l-histidine and l-proline and with respect to the type strain of R. leguminosarum in nitrate reduction, growth in presence of 1% NaCl and at 37 °C, assimilation of 5-keto-gluconate and 3-hydroxybutyrate (Table 4).
Table 4. Differential phenotypic characteristics of Endobacterium cerealis, its closest related genera and the type species of the type genus of family Rhizobiaceae.

| Characteristics | \(E.\) cerealis \(\text{RZME27}^\text{T}\) | \(N.\) galegae \(\text{HAMBI 540}^\text{T}\) | \(P.\) pelagicum \(\text{R1-200B4}^\text{T}\) | \(R.\) leguminosarum \(\text{USDA 2370}^\text{T}\) |
|-----------------|-----------------|-----------------|-----------------|-----------------|
| Colony colour (on YMA) | white | white-pink | white | white-cream |
| Nitrate reduction | + | – | + | – |
| Growth in 1% NaCl | + | + | + | – |
| Growth in 4% NaCl | + | – | + | – |
| Growth in 7% NaCl | – | – | + | – |
| Growth at 37 °C | w | + | + | – |
| Assimilation of (API 20NE): | | | | |
| Malate | + | + | – | w |
| \(\alpha\)-rhamnose | + | – | + | + |
| d-ribose | w | + | – | + |
| Inositol | + | + | – | + |
| Melibiose | + | + | – | + |
| 5-keto-gluconate | + | + | + | – |
| 2-keto-gluconate | – | – | w | – |
| Propionate | + | – | – | – |
| 3-hydroxybutyrate | – | – | – | + |
| \(\alpha\)-histidine | + | + | – | + |
| \(\alpha\)-alanine | w | + | w | – |
| \(\alpha\)-serine | – | – | + | – |

+: positive, –: negative, w: weak.

4. Discussion

The family Rhizobiaceae mainly contains genera with strains nodulating legumes, but in the last years several of them have been isolated from sources different to the legume nodules [14–16]. This occurs in the case of the most recently described genera within the family, Pseudorhizobium, Gellertiella and Georhizobium isolated from water-related environments [11–13]. However, none of the Rhizobiaceae genera described to date were originally isolated from cereal related sources [11–17]. In this work we described the first new genus within family Rhizobiaceae isolated from roots of a cereal and whose type strain RZME27\(^\text{T}\) is to date the only available strain for this genus. Although, ideally the description of new taxa should be based on more than one strain [47], many recently described bacterial genera only contain the type strain, as occurs for example with the two most recently described genera in the family Rhizobiaceae, Gellertiella and Georhizobium, also isolated from sources other than legume nodules, whose descriptions were based on a single strain [12,13].

The definition of this new genus is based on the minimal standards for definition of new bacterial taxa which have been recently published [48] and include two successive steps, the first one based on the 16S rRNA gene analysis and the second one on the calculation of several parameters derived from genome analyses, known as OGRI [48]. From these parameters, the ANIb values have been taken as reference for genera differentiation, being the cut-off values previously established for those of the family Rhizobiaceae after the comparison of their genomes [11].

Following this scheme, we analysed the 16S rRNA gene of the strain RZME27\(^\text{T}\), which was phylogenetically equidistant to several species of genera Rhizobium and Neorhizobium.
(Supplementary Table S2) and the similarity values for this gene support the affiliation of this strain to a new genus since some of more recently proposed genus within the family *Rhizobiaceae* have similar or even higher similarity values in their 16S rRNA gene (Table 1). This occurs in the case of the genera *Pseudorhizobium* and *Neorhizobium* whose type species have 97.1% similarity and in the case of genera *Ciceribacter*, *Pararhizobium* and *Gellertiella* with respect to the genus *Ensifer*, with similarity values ranging from 97.5% to 97.7%. The description of all these genera was based on genetic analyses, which commonly included the phylogenies of the 16S rRNA and different housekeeping genes. Nevertheless, currently, the description of new taxa should be based on genome analyses and Chun et al. [48] proposed the minimal standards for the use of genome data for the taxonomy of Prokaryotes. Although they only give the ANI threshold values for species differentiation, they have been evaluated for genera delineation in different bacterial families. For example, in the family *Enterobacteriaceae* the different genera show ANI values ranging from 75 to 80% [49], including the recently described genus *Scandinaviun* which has a ANIb value of 79.2% with respect to its closest related genus [50]. In the case of the Family *Rhizobiaceae*, it should be highlighted that the description of the genus *Pseudorhizobium* was based on genome analysis [11] and in this work the ANIb values among genomes of different genera belonging to the family *Rhizobiaceae* and to other families from the order *Rhizobiales* were analysed [11]. According to the results of this previous work, the current limit of ANIb value for genus delineation within several families of the order *Rhizobiales* was around 76% [11]. Recently, the description of the new genus *Geo rhizobium* was also supported by the results of ANIb calculation, with values lower than 76% with respect to the other genera of family *Rhizobiaceae* [13]. The ANIb values obtained for RZME27 with respect to the type species of the genera included in the family *Rhizobiaceae* were lower than 76% in all cases, supporting the affiliation of this strain to a new genus within this family (Table 2).

Taking into account that this family contains several genera able establish nitrogen fixing symbiosis with legumes and that the new genus is related to *N. galegae*, which is able to induce effective nodules in *Galega* sp. [9], we searched for symbiotic genes involved in nodulation and nitrogen fixation, but we did not find them in the genome of the strain RZME27 as also occurred in *P. pelagicum* [11]. However, several genes involved in colonization were found in the genome of strain RZME27, such as those related with the type IV secretion system, located in a genomic island, which is widespread among endophytic bacteria [51,52]. We also identified genes involved in motility, such as those codifying flagellar biosynthesis proteins, located in a genome region, which also contains chemotaxis involved genes (*che*) in neighbour positions regulated by *luxR* genes. The presence of *luxR* genes, which are associated to chemotaxis, as well as flagellar proteins indicates that these functions are modulated by population density where quorum sensing response plays the main role [53]. The genome of strain RZME27 and those of its closest relatives, *N. galegae*, *P. pelagicum* and *R. leguminosarum*, also contain the *luxI* gene, found by manual mining and antiSMASH analysis, which encodes a homoserine lactone synthase, commonly found in Proteobacteria and necessary for the production of homoserine lactone-quorum sensing related molecules in these bacteria [54].

Several genes found in the strain RZME27 involved in exopolysaccharide biosynthesis and degradative hydrolytic enzymes production may also play a role in the establishment of bacteria in rhizosphere and endosphere environments, as has been reported for rhizobia and other bacterial endophytes [55,56]. Particularly, genes involved in cellulose biosynthesis and cellulase production found in the genomes of the strain RZME27 and the type strains of *N. galegae*, *P. pelagicum* and *R. leguminosarum* were similar to those of *celABC* operon of *Rhizobium* strains nodulating clover [57]. Remarkably, annotations showed the presence of genes involved the synthesis of glycine-betaine, one of the main osmoregulate molecules in bacteria [58,59], in the genomes of the strain RZME27 and its closest relatives *N. galegae* and *P. pelagicum*. The results obtained in the in vitro phenotypic tests confirmed that the strain RZME27 can grow under saline osmotic stress, as well as *N. galegae*, which also grows in these conditions [60], and as *P. pelagicum*, which was originally isolated from a saline environment [11].
Comparative analysis helps to understand relationships between different species, and their evolution and genomic adaptation throughout the identification of clusters of orthologous genes originated by vertical inheritance from a unique common ancestor [61,62]. The highest number of clusters was found in the pangenome of the type strains of R. leguminosarum and N. galegae, which are able to nodulate legumes, confirming that symbiotic rhizobia have very complex pangenomes and also these two strains shared the highest number of clusters (Figure 4B). The strains sharing the highest number of clusters were the type strains of N. galegae and R. leguminosarum, all of them isolated from plant sources (Figure 4B). However, P. pelagicum, which was isolated from a marine habitat, shares less protein clusters with the species inhabiting soil related niches, than these species among them (Figure 4B). Remarkably, the strain RZME27T has a higher number of unshared clusters with the type species of its related genera than these type species among themselves. Nevertheless, the total number of clusters shared by the analyzed strains was quite high taking into account their different ecological niches.

5. Conclusions

In this work, we reported the genomic and phenotypic characteristics of a strain named RZME27T isolated from Zea mays roots growing in a field located at León (Northern Spain). The results of 16S rRNA gene and genome analyses showed that this strain belongs to the family Rhizobiaceae with its closest related taxa belonging to genera Neorhizobium, Rhizobium and Pseudorhizobium. The genes involved in legume nodulation and nitrogen fixation were absent in the genome of the strain RZME27T; nevertheless, genes involved in plant colonization and genes involved in osmotolerance were found. The comparative pan-genomic analysis showed that 78 protein clusters present in the strain RZME27T were not found in the type strains of its closest relatives N. galegae and P. pelagicum, nor in the type species of the genus Rhizobium. The ANIb values between the genomes of the strain RZME27T and those the type species of Neorhizobium and Pseudorhizobium were similar or lower than those found between the genomes other genera from the family Rhizobiaceae recently described, supporting the affiliation of the strain RZME27T to a novel genus for which we propose the name Endobacterium cerealis gen. nov. sp. nov.

6. Description of Endobacterium gen. nov.

Endobacterium (En.do.bac’te.ri.um. Gr. pref. endo, within; L. neut. n. bacterium, a small rod; N.L. masc. n. Endobacterium, a rod isolated from the endosphere of Zea mays).

Aerobic, Gram negative, motile and non-sporulated rods forming white colonies on YMA. Optimal growth at 28 °C and pH 7. Catalase and oxidase were positive. Nitrate reduction is positive. Glucose fermentation is negative. Esculin hydrolysis and production of urease and β-galactosidase were positive. Production of indole, H₂S and arginine dehydrolase is negative. Growth was observed up to 4% (w/v) NaCl. The main fatty acids are C₁₈:₁ω₇c/C₁₈:₁ω₆c. The G+C content of genomic DNA of the type strain of the type species is 58.8 mol%. Delineation of the genus was determined by the phylogenetic information from 16S rRNA gene sequence and by the analysis of the complete genome sequence. The type species is Endobacterium cerealis.

7. Description of Endobacterium cerealis sp. nov.

Endobacterium cerealis (ce.re.al’is. N.L. masc. adj. cerealis, of or pertaining to cereal, because the type strain was isolated from the cereal Zea mays).

Gram-stain negative rods that are 0.7–0.8 µm wide and 2.5–3.0 µm long. They are motile by means of one polar flagellum. Colonies are small, pearl white colored in YMA at 28 °C, which is the optimal growth temperature. The optimal pH for growth is 7–7.5 and the optimal NaCl concentration is 1%. Growth was observed in presence of 3.5% (w/v) NaCl and it was weak in presence of 4% (w/v) NaCl. Growth was observed in a range from 15 to 37 °C and pH from 6 to 8. No growth observed at 40 °C or pH 5. Oxidase and catalase positive. Nitrate reduction is positive. Glucose
fermentation is negative. Esculin hydrolysis and production of urease and β-galactosidase were positive. Production of indole, H₂S and arginine dehydrolase is negative. Assimilation as sole carbon and energy source of d-glucose, d-mannose, l-rhamnose, N-acetyl-glucosamine, l-ribose, inositol, sucrose, maltose, mannitol, melibiose, l-fucose, l-sorbitose, l-arabinose, d, l-lactate, gluconate, malate, l-alanine, 5 keto-glucuronate, l-histidine and l-proline is positive. Assimilation of itaconate, suberate, malonate, glycogen, 3-hydroxybenzoate, salicine, propionate, caprate, acetate, valerate, citrate, adipate, phenylacetate, 2 keto-gluconate, l-serine, 4-hydroxybenzoate and 3-hydroxybutyrate is negative. The fatty acid profile consists of C₁₈:₁ ω₆c/C₁₈:₁ ω₇c (summed feature 8), C₁₆:₀, C₁₆:₀ 3OH, C₁₈:₀ 3OH, C₁₆:₁ 6c/C₁₆:₁ ω₇c (summed feature 3), C₁₄:₀ 3OH/C₁₆:₁ iso I (summed feature 2), C₉:₁ ω₇c 11-methyl, C₁₇:₀ and C₁₈:₀. The G+C content of the strain RZME₂₇ᵀ is 58.8 mol%.

The type strain RZME₂₇ᵀ (=LMG 31256ᵀ = CECT 9794ᵀ) was isolated from roots of Zea mays in Spain.

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