The impact of topically applied preservation solutions on the respiratory epithelium of tracheal grafts submitted to cold ischemia: functional and morphological analysis

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OBJECTIVE: Advances in graft reepithelialization and revascularization have renewed interest in airway transplantation. This study aims to determine whether topically applied preservation solutions can ameliorate ischemic injury to tracheal grafts. We analyzed 1) the effects of cold ischemia on the mucociliary clearance of tracheal grafts and 2) the impact of topically applied preservation solutions on the effects of cold ischemia on mucociliary clearance.

METHOD: Tracheal segments (n = 217) from 109 male Wistar rats were harvested, submerged in low-potassium-dextran-glucose, histidine-tryptophan-ketoglutarate, or saline solution (saline group), and stored at 4°C for 6, 10, 16, or 24 hours. A control group (not submerged) was analyzed immediately after harvesting. In situ mucociliary transport and ciliary beating frequency were measured using a stroboscope. Epithelial integrity, cellular infiltration, and mucus storage were quantified by light microscopy and image analysis software, along with transmission electron microscopy.

RESULTS: 1) The effects of cold ischemia: in situ mucociliary transport and ciliary beating frequency were greater in the control group than after cold ischemia. Microscopic analysis results were similar between groups. 2) The effects of preservation solutions: there was no difference between the low-potassium-dextran-glucose, histidine-tryptophan-ketoglutarate, and saline groups in functional or light microscopy analysis. The saline group presented stronger signs of ischemic injury with transmission electron microscopy.

CONCLUSIONS: Cold ischemia diminished the mucociliary clearance of the tracheal respiratory epithelium. Topically applied preservation solutions did not ameliorate the injury caused by cold ischemia to the tracheal respiratory epithelium.

KEYWORDS: Trachea; Mucociliary Clearance; Organ Preservation Solutions.

INTRODUCTION

Tracheal transplantation remains a challenge (1). A large number of surgical techniques and implantable devices have been tested, with disappointing results (1,2). In spite of that, some recent findings on tracheal revascularization and graft reepithelialization have renewed interest in tracheal transplantation (2–5). However, for tracheal transplantation to succeed, the effects of ischemic injury secondary to harvesting and storage on tracheal tissues need to be better understood.

Because of the particular segmental pattern of tracheal vascularization, with no major tracheal vessels, the intravascular administration of preservation solutions into tracheal grafts is far from useful in a clinical scenario (2,6,7). Therefore, we previously investigated whether preservation solutions were effective when applied topically. Due to the small sample size, preliminary data were inconclusive in both measuring the effects of cold ischemia on the tracheal respiratory epithelium and defining whether...
topically applied preservation solutions could ameliorate those effects (8).

The trachea acts as both a mechanical conduit of environmental air to and from the lower respiratory tract and a defense organ through mucociliary clearance. Mucociliary clearance is a delicate mechanism through which mucus is propelled from the lower to the upper airways, carrying with it noxious organic and inorganic particles to be swallowed or expelled by coughing (9). Based on its delicate mechanism, we consider mucociliary clearance, especially mucociliary transportation, an excellent criterion by which to verify tracheal graft preservation. Our aims were 1) to analyze the effects of cold ischemia on the tracheal respiratory epithelium and 2) to analyze the effects of topically applied preservation solutions on tracheal respiratory epithelium submitted to cold ischemia.

**MATERIALS AND METHODS**

This research was approved by the Ethical Committee of our Institution. All animals received humane care in compliance with the ‘Principles of Laboratory Animal Care’ formulated by the National Society for Medical Research and the ‘Guide for the Care and Use of Laboratory Animals’ prepared by the Institute of Laboratory Animal Resources and published by the National Institutes of Health (NIH Publication No. 86–23, revised 1996).

We obtained 217 tracheal segments from 109 Wistar male rats (weight, 300±50 g). The rats were anesthetized with intraperitoneal pentobarbital (50 mg/kg) and euthanized by exsanguination. A median cervicosternotomy was performed, followed by tracheal harvesting. The trachea was sectioned at the midpoint between the cricoid cartilage and the carina, providing two tracheal segments from each rat. Immediately after harvesting, each tracheal segment was randomly allocated to be submerged in one of three solutions: saline solution (saline group); low-potassium- dextran-glucose (LPD-glucose) (Vitrolife AB, Sweden) (LPD group); or histidine-tryptophan-ketoglutarate (HTK) (Dr F. Köhler Chemie, Germany) (HTK group). Each tracheal segment was submerged in the solution within a sterile test tube. The submerged tracheal segments were immediately allocated to be stored at 4°C for 6, 10, 16, or 24 hours. Each tracheal segment was submitted to only one period of cold ischemia. At the end of the ischemic time (6, 10, 16, or 24 hours), the tracheal segments were incubated at room temperature for 20 minutes and submitted to the functional analyses of mucociliary clearance as described below. After functional analyses, the segments were prepared for morphological analyses as described below.

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**Morphological analysis**

Morphological analysis was performed using light microscopy and transmission electron microscopy. After functional analysis, tracheal segments were fixed in formalin and embedded in paraffin. Serial sections (0.5 μm thick) were cut with a microtome and stained with hematoxylin-eosin, alcian blue, and periodic acid-Schiff (PAS). A small sample of each tracheal segment was prepared for transmission electron microscopy.

**Light microscopy**

The hematoxylin-eosin-stained slides were used to determine the ischemic injury on the tracheal grafts. We analyzed ischemic injury through epithelial integrity and cellular infiltration. The analysis was performed using modified semi-quantitative scales for epithelial integrity and cellular infiltration based on a previous work on tracheal transplantation (12). Epithelial integrity was quantified with a semi-quantitative scale rated from 1 to 3: 1 (low integrity), epithelial lining up to 1/3 of the tracheal surface; 2 (moderate integrity), epithelial lining from 1/3 to 2/3 of the tracheal surface; and 3 (high integrity), epithelial lining of more than 2/3 of the tracheal surface. Cellular infiltration was quantified with a semi-quantitative scale rated from 1 to 3: 1 (mild infiltration), scattered and diffuse cellular infiltration, without islets of dense infiltrate; 2 (moderate infiltration), more intense cellular infiltration, with up to two islets of dense infiltrate; and 3 (severe infiltration), a thick layer of dense infiltrate or more than two islets of dense infiltrate. Each slide was distinctly quantified by three observers blinded to the groupings, and the final score was achieved with the concordance of at least two observers.

The alcian blue- and PAS-stained slides were used to evaluate intracellular mucus storage in the respiratory epithelium after cold ischemia: alcian blue stained the acidic mucus, and PAS stained the neutral mucus. The best image for each slide was digitized with a digital camera (AxioCam, Zeiss, Germany) attached to a light microscope (Axio Imager.A1, Zeiss, Germany). The area in the microphotograph that corresponded to the respiratory epithelium was delimited using image analysis software (AxioVision 4.7, Zeiss, Germany). Then, the software quantified the area stained by alcian blue and PAS in the delimited area. The
quantification was expressed as the percentage of the respiratory epithelial area stained by alcian blue and PAS.

Transmission electron microscopy
We obtained one sample from each tracheal segment. The samples were fixed in 2% glutaraldehyde and 0.1% tannic acid for 2 hours, followed by fixation in 1% osmium tetroxide for 1 hour. After dehydration in a series of ethanol solutions for 1 hour, the samples were embedded in acetone and resin for 3 hours. The samples were baked for 24 hours to be embedded in pure resin and stored. Only samples from the control group and the more affected tracheal segments, as indicated by functional and light microscopy analysis, were stained and inspected. Thick sections (0.5 μm) were stained with toluidine blue to determine the area of interest. Then, ultrathin sections (90 nm) were prepared and stained with uranyl acetate and lead citrate. The specimens were inspected and photographed under a transmission electron microscope (1200EXII, Jeol, Japan).

Data presentation and statistical analysis
The data obtained were analyzed from two perspectives according to the objectives of the study. To analyze the effects of cold ischemia on the tracheal respiratory epithelium (our first objective), we grouped the tracheal segments according to the duration of ischemia and performed the first statistical analysis. Therefore, we had five groups: control group (not submitted to ischemia), 6 hours, 10 hours, 16 hours, and 24 hours. The type of preservation solution used was not considered in this first analysis. Subsequently, we focused on the effects of the topically applied preservation solutions on the ischemic tracheal respiratory epithelium (our second objective). The second statistical analysis compared the data from the different preservation solutions (control group, LPD group, HTK group, and saline group), including all ischemic times in each preservation solution group but not including the ischemic time itself.

The distribution of the variables was evaluated using the Kolmogorov-Smirnov test. For variables following a normal distribution, the data were presented as the mean ± standard deviation and compared with an analysis of variance (ANOVA) with Bonferroni’s post-test. Despite analyzing two factors (ischemic time and preservation solutions), we did not perform a two-way ANOVA because analyzing two factors (ischemic time and preservation solutions) violated assumption of the normality and variance homogeneity. Variables were rescaled to a standard normal distribution using the Kolmogorov-Smirnov test. We performed a two-way ANOVA analysis of variance with Bonferroni’s post-test for the comparisons of the ischemic time itself. For variables not following a normal distribution, the data were compared with a non-parametric statistical analysis using the Mann-Whitney U test. The statistical analysis was performed with SPSS 13.0 software (SPSS Inc., Chicago, IL, USA).

**RESULTS**

Effects of cold ischemia on the tracheal respiratory epithelium
As shown in Table 1, there was a statistically significant difference between the control group and the groups submitted to cold ischemia in terms of in situ mucociliary transport (p < 0.003) (Figure 1A) and ciliary beating frequency (p < 0.03) (Figure 1B). In the morphological analysis, there was no statistically significant difference among the groups in either epithelial integrity or cellular infiltration (data not shown). Figure 2 illustrates how the semi-quantitative scores were calculated for some histological slides. Indeed, there was no statistically significant difference in the quantification of the area stained by alcian blue and PAS among the groups (Figure 1C and 1D).

**Effects of topically applied preservation solutions on the tracheal respiratory epithelium**
As shown in Table 2, in situ mucociliary transport was higher in the control group compared to the other groups (p < 0.0001) (Figure 3A). In situ mucociliary transport was also higher in the HTK group than the saline group (p < 0.006). However, there was no difference in mucociliary transport between the other groups in which tracheal segments were submerged in preservation solutions. The ciliary beating frequency in the control group was different compared to the LPD, HTK, and saline groups (p < 0.02). However, there was no difference among the LPD, HTK, and saline groups in the same variable (Figure 3B). In the morphological analysis, there was no statistically significant difference among the groups in either epithelial integrity or cellular infiltration (data not shown). There was no statistically significant difference in the quantification of the area stained by alcian blue and PAS among the groups (Figure 3C and 3D).

The transmission electron microscopy findings are shown in Figure 4. The LPD group showed nuclear shrinkage, most likely related to osmotic differences between the respiratory cells and the LPD-glucose solution. Some signs of cytoplasmic disruption and vacuolization were found (Figure 4B). The HTK group showed electron-dense crystals throughout the respiratory cells. The cilia coincided with those crystals. Some necrotic cells were found, which contained a large amount of those crystals. However, the ultrastructural architecture of the respiratory cells was preserved (Figure 4C). The saline group showed marked signs of loss of cellular integrity, cytoplasmic disruption, and intense vacuolization (Figure 4D).

**Table 1 - Effects of cold ischemia on mucociliary clearance.**

|                | Control (n = 08) | 6 hours (n = 51) | 10 hours (n = 51) | 16 hours (n = 50) | 24 hours (n = 57) | p-value |
|----------------|-----------------|-----------------|------------------|------------------|------------------|---------|
| MT (mm/min)    | 0.104 (0.106)   | 0.026 (0.047)   | 0.029 (0.051)    | 0.018 (0.033)    | 0.019 (0.035)    | 0.003   |
| CBF (Hz)       | 15.34 ± 3.37    | 13.09 ± 3.12    | 12.14 ± 4.03     | 12.86 ± 3.79     | 11.01 ± 5.02     | 0.03    |
| AB (%)         | 10.96 ± 4.51    | 6.90 ± 5.82     | 5.54 ± 5.66      | 6.15 ± 4.04      | 5.37 ± 4.32      | 0.08    |
| PAS (%)        | 16.55 ± 9.72    | 13.87 ± 8.65    | 11.24 ± 6.56     | 12.55 ± 8.20     | 11.44 ± 5.88     | 0.16    |

Data according to the ischemic time [as median (interquartile range) or the mean ± standard deviation]. MT = in situ mucociliary transport. CBF = ciliary beating frequency. AB = area of the respiratory epithelium stained with alcian blue. PAS = area of the respiratory epithelium stained with periodic acid-Schiff. mm/min = millimeters/minute. Hz = hertz.
DISCUSSION

Some advances, such as the use of angiogenic factors for tracheal graft revascularization and improved knowledge on the reepithelialization phenomenon after tracheal allograft transplantation, have renewed interest in clinical airway transplantation (3,4). More recently, the use of tissue-engineering techniques has produced promising results (5). However, although some successful case reports have been published, there is no universally accepted clinical approach for tracheal transplantation. We believe that the poor knowledge on the effects of ischemia on tracheal grafts is one of the most important obstacles for performing tracheal transplantation in a clinical setting. Research on tracheal graft ischemia is scarce, and the principles of tracheal graft preservation are based on extrapolations from research on the preservation of other solid organs, primarily the lung. To our knowledge, there are no data regarding the impact of cold ischemia on the mucociliary clearance of tracheal grafts.

Mucociliary clearance is the most important innate defense mechanism of the respiratory system (9). Mucociliary clearance relies on the production and secretion of mucus by the goblet cells of the respiratory epithelium and submucosal glands in conjunction with the propulsion of that mucus by the ciliated cells. The trachea plays an essential role in mucociliary clearance by transporting potentially dangerous organic and inorganic particles adhered to the mucus from the lower to the upper airways to be swallowed or expelled by coughing (13). In fact, the trachea acts as a defense organ by effecting mucociliary clearance. Therefore, the maintenance of tracheal mucociliary clearance appears to be an important prerequisite for tracheal graft preservation. Indeed, the adequacy of tracheal mucociliary clearance could mirror the adequacy of tracheal graft preservation.

The effects of ischemia on the respiratory epithelium of major airways have not been extensively studied. Wagner et al analyzed the effects of ischemia on mucociliary clearance in a model of selective bronchial artery occlusion in sheep (14). They found a decrease in mucociliary clearance at approximately 20 minutes after bronchial artery occlusion. In a previous work, our group demonstrated that mucociliary clearance was severely impaired by room temperature ischemia, even after ischemic periods as short as 6 hours (8). In that study, we found a trend toward worse ciliary beating function in tracheal grafts submitted to 24-hour cold ischemia compared to shorter periods of cold ischemia. We credited the absence of statistical significance to the
small sample size and proposed that another study with a larger sample size be performed to clarify this issue (8).

The present data show that cold ischemia significantly impairs the mucociliary clearance of tracheal grafts. Both *in situ* mucociliary transportation and ciliary beating frequency were adversely affected by cold ischemia. As stated above, we previously hypothesized that the longer the ischemic time, the stronger the impairment of tracheal mucociliary clearance. However, despite a trend toward worse functional results after longer ischemic times, the difference between mucociliary clearance in the shorter (6 hours) and longer (24 hours) ischemic groups was not statistically significant (Figure 1). This finding could be partially explained by the method of functional analysis of mucociliary clearance employed in this work. When submerging the tracheal grafts, the preservation solution remained in contact with the mucus of the respiratory epithelium for several hours. This contact could have affected the rheological properties of the mucus and influenced the *in situ* mucociliary transport and ciliary beating frequency measurements, especially in the groups with longer ischemic times. For instance, the contact between the mucus and the preservation solution could diminish the viscosity of the mucus, which could facilitate ciliary movement and compensate for the ciliary dysfunction caused by the ischemic injury.

The morphological analysis demonstrated that mucous storage in the respiratory epithelium was not affected by cold ischemia up to 24 hours. Mucus production and secretion are two important events of mucociliary clearance. Organic and inorganic noxious agents adhere to mucous and are expelled by ciliary beating. A number of factors can diminish the mucous storage of airway epithelium by inducing goblet cell secretion, e.g., irritant gases, inflammatory mediators, oxygen metabolites, and pH variations (15). In other solid organs, such as the liver and lung, cold ischemia has some effects that modify the intracellular environment, such as the production of reactive oxygen species (oxygen metabolites) and the induction of intracellular calcium overload through calcium release from intracellular depots (16). We hypothesized that the release of mucus could be affected by the cellular modifications in the goblet cells caused by cold ischemia. However, our data did not show any difference in mucous storage after up to 24 hours of cold ischemia.

The trachea has a simple anatomic and physiologic structure compared to other solid organs. The trachea is a

### Table 2 - Effects of topically applied preservation solutions on mucociliary clearance.

|                | Control group (n=8) | LPD group (n=86) | HTK group (n=81) | Saline group (n=42) | p-value |
|----------------|---------------------|------------------|------------------|--------------------|---------|
| MT (mm/min)    | 0.104 (0.106)       | 0.023 (0.039)    | 0.028 (0.045)    | 0.013 (0.018)      | 0.0001  |
| CBF (Hz)       | 15.34 ± 3.57        | 12.16 ± 4.54     | 12.50 ± 4.14     | 11.90 ± 3.22       | 0.02    |
| AB (%)         | 10.96 ± 4.51        | 5.94 ± 5.02      | 5.86 ± 4.71      | 6.11 ± 5.59        | 0.14    |
| PAS (%)        | 16.55 ± 9.72        | 12.30 ± 6.55     | 12.03 ± 7.94     | 12.27 ± 7.75       | 0.56    |

Data according to the preservation solution group [as the median (interquartile range) or the mean ± standard deviation]. MT = *in situ* mucociliary transport. CBF = ciliary beating frequency. AB = area of the respiratory epithelium stained with alcian blue. PAS = area of the respiratory epithelium stained with periodic acid-Schiff. mm/min = millimeters/minute. Hz = hertz.
semi-rigid tube, approximately 3 mm thick, whose lumen remains open even after extraction (6). Conversely, the tracheal arterial supply has a segmental pattern, without a major tracheal artery or vein. In the clinical environment, this anatomical arrangement makes the intravascular administration of preservation solutions into the trachea exceedingly difficult (2,7).

Based on the tracheal anatomy of both human and Wistar rats and on previous studies, we did not attempt to deliver preservation solutions through any intravascular route to fulfill the objectives of this study (17,18). Therefore, we tried to obviate the need for the intravascular administration of preservation solutions into the trachea exceedingly difficult (2,7).

In situ mucociliary transport. B) Ciliary beating frequency. C) Area stained by alcian blue. D) Area stained by PAS. LPD = low-potassium-dextran-glucose solution. HTK = histidine-tryptophan-ketoglutarate solution. mm/min = millimeters/minute. Hz = hertz. PAS = periodic acid-Schiff. * = statistical significance. NS = not statistically significant.

Figure 3 - Functional and morphological analysis according to the preservation solution. A) In situ mucociliary transport. B) Ciliary beating frequency. C) Area stained by alcian blue. D) Area stained by PAS. LPD = low-potassium-dextran-glucose solution. HTK = histidine-tryptophan-ketoglutarate solution. mm/min = millimeters/minute. Hz = hertz. PAS = periodic acid-Schiff. * = statistical significance. NS = not statistically significant.

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Our study has some limitations. The analysis was focused on the respiratory epithelium of the tracheal grafts. We did not analyze other tracheal tissues, such as the smooth muscle...
or the cartilage, which are also important for tracheal function. Another limitation is the sample size of the control group, which was smaller than the other groups. Although this difference could lead to bias when interpreting the statistical analysis, this bias did not occur in this study. Despite the larger dispersion measures in some variables of the control group, significant differences among the groups could be clearly observed. As another limitation, morphological analysis through transmission electron microscopy was performed in only a small proportion of samples. Therefore, only a qualitative analysis, with not statistical analysis, could be performed, despite the large differences among the groups.

Our conclusions are the following: 1) cold ischemia diminished the mucociliary clearance of the tracheal respiratory epithelium, and 2) topically applied preservation solutions did not ameliorate the ischemic injury caused by cold ischemia to the tracheal respiratory epithelium. Further studies that address the function and morphology of connective and muscular tissues after ischemia could elucidate whether those effects occur in other tracheal cells.

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**AUTHOR CONTRIBUTIONS**

Azevedo-Pereira AE designed the study, collected the data, and wrote the manuscript. Saka JA collected the data. Oliveira-Braga KA and Pazetti R designed the study and collected the data. Canzian M designed the study and collected the data. Pêgo-Fernandes PM designed the study and revised the manuscript. Jatene FB revised the manuscript.

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