An international survey on aminoglycoside practices in critically ill patients: the AMINO III study

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Abstract

Background: While aminoglycosides (AG) have been used for decades, debate remains on their optimal dosing strategy. We investigated the international practices of AG usage specifically regarding dosing and therapeutic drug monitoring (TDM) in critically ill patients. We conducted a prospective, multicentre, observational, cohort study in 59 intensive-care units (ICUs) in 5 countries enrolling all ICU patients receiving AG therapy for septic shock.

Results: We enrolled 931 septic ICU patients [mean ± standard deviation, age 63 ± 15 years, female 364 (39%), median (IQR) SAPS II 51 (38–65)] receiving AG as part of empirical (761, 84%) or directed (147, 16%) therapy. The AG used was amikacin in 614 (66%), gentamicin in 303 (33%), and tobramycin in 14 (1%) patients. The median (IQR) duration of therapy was 2 (1–3) days, the number of doses was 2 (1–2), the median dose was 25 ± 6, 6 ± 2, and 6 ± 2 mg/kg for amikacin, gentamicin, and tobramycin respectively, and the median dosing interval was 26 (23.5–43.5) h. TDM of $C_{\text{max}}$ and $C_{\text{min}}$ was performed in 437 (47%) and 501 (57%) patients, respectively, after the first dose with 295 (68%) patients achieving a $C_{\text{max}}$/MIC > 8 and 353 (71%) having concentrations above $C_{\text{min}}$ recommended thresholds. The ICU mortality rate was 27% with multivariable analysis showing no correlation between AG dosing or pharmacokinetic/pharmacodynamic target attainment and clinical outcomes.

Conclusion: Short courses of high AG doses are mainly used in ICU patients with septic shock, although wide variability in AG usage is reported. We could show no correlation between PK/PD target attainment and clinical outcome. Efforts to optimize the first AG dose remain necessary.

Trial registration Clinical Trials, NCT02850029, registered on 29th July 2016, retrospectively registered, https://www.clinicaltrials.gov

Keywords: Antibiotics, Aminoglycoside, ICU, Therapeutic drug monitoring, PK/PD

Background

Early administration of adequate doses of antibiotics is a key issue for the optimal management of sepsis in critically ill patients [1, 2]. Optimized dosing is likely to reduce mortality in patients with severe sepsis and septic shock [3]. Therefore, broad-spectrum antimicrobial therapy that combines more than one antimicrobial agent...
can be used to ensure appropriate therapy in the initial phase of severe sepsis [4]. Even though the benefit of combination therapy remains unclear, aminoglycosides (AG) are often given as part of empiric therapy for sepsis, especially when Gram-negative bacteria are suspected [5–7]. To achieve maximal efficacy, the optimal AG dosing regimen should reach a peak concentration to minimum inhibitory concentration (peak/MIC) ratio of at least 8–10, as this pharmacokinetic/pharmacodynamics (PK/PD) target has been associated with clinical cure [8–11]. On the other hand, the optimal AG dosing regimen should expose patients to the lowest risk of toxicity [12]. Therapeutic drug monitoring (TDM), trough concentration monitoring in particular, is encouraged in critically ill patients to guide re-dosing and to limit toxicity. Many factors influence the PK and PD of antimicrobials in critically ill patients. An increased volume of distribution, the occurrence of hypo-albuminemia, and renal and/or hepatic dysfunctions are frequently observed in those patients leading to altered PK [13]. Previous studies have reported that, for up to 30–40% of critically ill patients, AG plasma concentrations were lower than the targeted peak concentrations using standard dosing regimens [14–17]. Positive fluid balance during the previous day and low body mass index (BMI) have been reported as risk factors of insufficient peak plasma concentrations [17]. The use of higher amikacin dosing regimens (≥25 mg/kg) has been suggested by several authors and recommended by national guidelines [12, 14, 18]. However, practices of antimicrobial therapy and TDM may widely vary based on available resources and local guidelines. Moreover, as local ecology and MICs’ distribution may also differ among countries, this could lead to different practices of AG use among countries. The main objective of the study was to provide a detailed description of agents, dosing, and monitoring practices of AG in in a large cohort of critically ill patients from different countries receiving AG for a sepsis or a septic shock.

The secondary objectives were to report renal adverse events and clinical outcomes, and to assess the correlation between PK/PD target attainment and these outcomes (clinical success and ICU mortality).

Methods

This study is reported in accordance with the STrengthening the Reporting of OBServational studies in Epidemiology (STROBE) guidelines [19].

A prospective, international, observational cohort design was used. All participating ICUs obtained local ethics committees’ approval [ethics approval for France (Institutional Review Board of Nîmes University Hospital, France 15/04.08) [20]] and the study was registered in Clinical Trials (NCT02850029). Written informed consent was waived due to the non-interventional design of the study.

Study population

All adult patients (≥18 years) requiring ICU care and receiving AG (amikacin, gentamicin, and tobramycin) therapy for a sepsis according to the Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3) were included in the study [21].

Settings

Overall, 85 ICUs accepted to participate to the study, and 59 ICUs enrolled patients from May 2016 to August 2017 in five different countries: Australia (5), Greece (5), Chile (4), UK (9), and France (62).

Data collection and management

Participating site data

For each participating ICU, the following information were recorded: type of hospital, type of ICU, number of beds, number of physicians, number of admissions per year (on the last year), annual ICU mortality, national or local AG guidelines availability, and TDM availability.

Patient data

Data recorded for each patient included were: demographic data (age, gender, height, total body weight), clinical data (admission diagnosis, co-morbidities, APACHE II or SAPS II (Simplified Acute Physiology Score) at admission, SOFA (Sequential Organ Failure Assessment) score at inclusion), and biological data (serum creatinine concentration, albumin concentration, and 24-h urine output every day during AG therapy (where possible)). Additionally, dosing and TDM data when available (dose and frequency, time of dosing and sampling, therapy duration, maximal and minimal AG plasma concentrations, and co-administered antibiotics) and toxicity data (co-administered nephrotoxic drugs, renal function assessment using the Acute Kidney Injury Network (AKIN) criteria [22]) at the end of AG therapy were collected. Finally, infection data (known or presumed pathogen, known or likely MIC) and the following outcome data: clinical success, ICU and hospital length of stays (LOS), and ICU mortality were recorded. Clinical success was defined as the resolution of initial symptoms for which AG were prescribed, relapse was defined as the occurrence of a new infectious episode with the same identified pathogen, and superinfection was defined as the occurrence of a new infectious episode with a different pathogen).

The definition used to assess PK/PD outcome was as follows. PK/PD target attainment was defined as AG concentration after the first dose ≥8 times the MIC where...
AG plasma concentrations were available. Breakpoints from the EUCAST database were considered [23]. Where infections were polymicrobial, the MIC of the least susceptible pathogen was used in the analysis. Actual values of MIC were considered where available to assess “a posteriori” \( C_{\text{max}}/\text{MIC} \) ratio.

**Data management**

Data collection was performed by trained staff at each participating center. Data were entered into a structured electronic password-protected and secured web-based case report form (eCRF). The eCRF was developed using the open-source REDCap Data Management Platform hosted at the Nîmes University Hospital [24].

Data monitoring was handled by the coordinating center (Nîmes University Hospital, France). Outstanding queries regarding the completion of the CRF were undertaken with each participating center where necessary to ensure accuracy of data.

**Statistical analysis**

Descriptive statistics of the primary outcome were reported using mean, standard deviation, median, and interquartile range (25th–75th percentile) for continuous variables. For categorical variables, frequencies and proportions were given. Two secondary outcomes were explored separately: ICU mortality and clinical success.

We used multivariate regression models involving eight covariates: admission type (medical or surgery), age, gender, body mass index (BMI, kg m\(^{-2}\)), SOFA score on the day of AG initiation, creatinine plasma concentration (\( \mu \text{mol} \text{ L}^{-1} \)) on the day of AG initiation, bacteremia, and directed or empirical antibiotic treatment. A 9th variable related to AG treatment was considered. We constructed a model for each way we considered AG: the mg/kg dose using either the total body weight (TBW), the ideal body weight (IBW, thanks to the Lorentz’s formula), or the adjusted body weight [ABW: given by the TBW when the BMI was below 30 kg m\(^{-2}\) and when the BMI was over 30 kg m\(^{-2}\) calculated according to the following formula: IBW + 0.4 \times (TBW-IBW)], the peak concentration (mg L\(^{-1}\)), the attainment of a peak concentration between 60 and 80 mg L\(^{-1}\) for amikacin, and between 20 and 30 mg L\(^{-1}\) for gentamicin. Because practices could differ between hospitals, we needed a model accounting for correlation within centers. For this purpose, we chose a GEE (generalized estimating equations) model with an exchangeable correlation structure. This method allows parameters of generalized linear model to be estimated in clustered data and returns a marginal model that fits in with our purpose (i.e., estimating average response in overall population) [25, 26].

For continuous variables mean ± standard deviation or median [interquartile range] are given. For categorical variables, numbers (%) are given.

For the multivariate analysis based on the robust standard error provided by the Huber–White standard error and assuming the normal distribution of the regression coefficients [27]. To test the significance of OR, we performed Wald tests. All models were built in the whole population. A \( P \) value of 0.05 was considered to indicate statistical significance. All statistical analyses were performed using R software (version 3.6.0) [28].

**Results**

From May 2016 to August 2017, 971 patients were enrolled from 59 ICUs worldwide (Additional file 1: Figure S1). Of the 971 patients initially included in the study, 40 were excluded because of missing data leading to 931 studied patients.

**Participating site data**

Most of the ICUs were from university hospitals (38, 64%), 3 were medical ICUs, 14 were surgical, and 42 were mixed-ICUs. Description of the participating sites and their local protocols regarding AG therapy are described in Additional file 2: Table S1.

**Demographic data**

Patient demographic data at inclusion are presented in Table 1. The source of sepsis was pulmonary (405, 44%), abdominal (233, 25%), urinary (126, 14%) infection, or bacteremia (158, 17.0%). The most common microorganisms for which AG were prescribed are presented in Table 2. Among patients with microbiological samples drawn (\( n = 826, 89\% \)), 511 patients had positive cultures. MICs for all administered antibiotics were available in 135 patients with 127 (94%) receiving adequate

**Table 1** Patients’ characteristics at baseline

| Variables                      | Value |
|-------------------------------|-------|
| Age (years)                   | 63 ± 15 |
| Female                        | 364 (39%) |
| Weight (TBW, kg)              | 77 ± 20 |
| Height (cm)                   | 169 ± 9.5 |
| BMI (kg m\(^{-2}\))           | 27 ± 6.8 |
| Medical/surgical admission    | 512 (55%)/417 (44.8%) |
| APACHE                        | 22 [15,28] |
| SOFA                          | 8 [6,11] |
| Creatinine level (\( \mu \text{mol} \text{ L}^{-1} \)) | 133 ± 120 |
| Albumin (g L\(^{-1}\))        | 27 ± 7 |
| Protein (g L\(^{-1}\))        | 55.6 ± 12.3 |

For continuous variables mean ± standard deviation or median [interquartile range] are given. For categorical variables, numbers (%) are given.

TBW: total body weight; BMI: body mass index; ICU: intensive-care medicine.
antimicrobial therapy. In 22 (17%) patients, AG enabled to broaden spectrum activity.

**Primary outcome**
Among the 931 included patients, 14 (1%) received tobramycin, 303 (33%) received gentamicin, and 614 (66%) received amikacin. Practices of AG administration are described in Table 3. AG were prescribed empirically in 761 (82%) patients and as a combination therapy in 914 (98%) patients, mostly in association with ß-lactam antibiotics (890, 96%). The median duration of therapy was 2 ((IQR 1–3) days, the median number of AG doses per course was 2 ((IQR 1–2), and the median dosing interval was 26 (IQR 23.5–43.5) h. 483 patients (52%) received a second dose. TDM of C\text{max} and C\text{min} concentrations was performed in 437 (47%) and 501 (57%) patients for the first dose (Figs. 1 and 2). After the first AG dose, 295 (68%) patients achieved the PK/PD target of C\text{max}/MIC > 8 considering an MIC of 8 mg L\textsuperscript{−1} for amikacin ((241/341, 71%), and 2 for gentamicin (53/90, 59%) and tobramycin (1/6, 17%), whereas 353 (71%) patients had concentrations above C\text{min} recommended thresholds.

For 128 patients, an MIC was available for at least one pathogen, with a median of 2 [1; 4] mg L\textsuperscript{−1}. Among these 128 patients, 76 had both MIC and C\text{max} available, with a median C\text{max}/MIC ratio of 25.7 [16.7; 40.3] and for 65 patients C\text{max}/MIC ratio was above 8.

### Table 2 Microbiological data

| Variables | n |
|-----------|---|
| Microbiological sample (n = 930) | 826 (89%) |
| Number of pathogens identified (n = 511) | 317 (62%) |
| 1 | 121 (24%) |
| 2 and more | 73 (14%) |
| Pathogens (n = 798) | 64 (8%) |
| Gram-positive bacteria | 255 (32%) |
| Staphylococcus/S. aureus | 120 (15%)/84 (10.5%) |
| Streptococci | 64 (8%) |
| Enterococci | 67 (8.4%) |
| Gram-negative bacteria | 453 (58%) |
| Escherichia coli | 166 (21%) |
| Klebsiella spp. | 70 (8.8%) |
| Pseudomonas aeruginosa | 73 (9%) |
| Enterobacter spp. | 41 (5%) |
| Acinetobacter baumannii | 9 (1%) |
| Stenotrophomonas maltophilia | 5 (0.6%) |
| Anaerobes bacteria | 24 (3%) |
| Fungi/Candida | 33 (4%)/31 (3.9%) |

### Table 3 Aminoglycoside therapy characteristics

|                              | All (n = 931) | Amikacin (n = 614) | Gentamicin (n = 303) | Tobramycin (n = 14) |
|------------------------------|--------------|--------------------|----------------------|---------------------|
| Duration of therapy (days)   | 2 [1; 3]     | 2 [1; 3]           | 2 [1; 3]             | 2 [2; 4.75]         |
| Number of injections         | 2 [1; 2]     | 2 [1; 2]           | 2 [1; 2]             | 2 [1; 4]            |
| Second dose administered     | 483 (52%)    | 326 (53%)          | 164 (53%)            | 13 (93%)            |
| First dose in mg             | –            | 2000 [1500; 2250]  | 440 [320; 560]       | 500 [381; 555]      |
| First dose in mg per kg of TBW | –          | 26 [21.9; 29.4]    | 5.6 [4.3; 7.5]       | 6 [5.1; 7.1]        |
| First dose in mg per kg of ABW | –           | 267.8 [23.8, 29.4] | 6.2 [4.8; 7.8]       | 6.1[5.4; 7.1]       |
| Infusion duration (min)      | 30 [30; 30]  | 30 [30; 30]        | 30 [30; 30]          | 30 [30; 30]         |
| C\text{max} after first dose | 437/931 (47%) | 341/614 (56%)      | 90/303 (30%)         | 6/14 (43%)          |
| C\text{max} value (mg L\textsuperscript{−1}) | –         | 73 [57; 90]        | 21 [17; 25]          | 19 [14; 26]         |
| Delay end of infusion and C\text{max} sampling (min) | 30 [30; 50] | 30 [30; 50]        | 30 [30; 44]          | 30 [26; 37]         |
| Targeted C\text{max} attainment after first dose | 295/437 (68%) | 241/341 (71%)      | 53/90 (59%)          | 1/6 (17%)           |
| C\text{max} after first dose | 501/880 (57%) | 347/577 (60%)      | 145/289 (50%)        | 6/14 (43%)          |
| C\text{max} above threshold | 353/498 (71%) | 230/347 (66%)      | 118/145 (81%)        | 5/6 (83%)           |
| Time from first dose to C\text{min} (h) | 22.9 [20.4; 24] | 22.6 [20.5; 24]   | 23.4 [21.5; 24.1]    | 24.4 [23.6, 26.1]   |
| Interval after first dose (h) | 26 [23; 43] | 26.7 [23; 45]      | 24.8 [23; 34]        | 24.5 [23; 29]       |

For continuous variables, mean ± standard deviation or median (interquartile range) are given. For categorical variables, numbers (%) are given. The ABW was calculated as follow: IBW + 0.4 x (TBW-IBW). The C\text{max} attainment is defined as a C\text{max} above 20 mg L\textsuperscript{−1} for gentamicin and tobramycin or above 60 mg L\textsuperscript{−1} for amikacin. The C\text{min} thresholds considered are 0.5 mg L\textsuperscript{−1} for gentamicin and tobramycin and 2.5 mg L\textsuperscript{−1} for amikacin.
Secondary outcomes

Clinical response

Antimicrobial therapy was considered successful in 621/843 (74%) patients. Relapse and superinfection were observed in 42/843 (5%) and 71/843 (8%) patients, respectively, whereas in 17 patients, emergence of multi-drug resistant (MDR) pathogens was noted.

Renal function on AG therapy

Concomitantly to AG, 361 (39%) patients received at least one nephrotoxic agent among non-steroid anti-inflammatory drug, iodine, angiotensin-converting enzyme, and vancomycin. On the last day of therapy, AKIN score was ≥ 1 in 108/481 (22%) analysable patients.

Patient outcome

The ICU mortality rates were 27%, respectively. The median ICU and hospital LOS were 11 (IQR 5–25) and 24 (IQR 11–43) days, respectively.

Independent factors associated with ICU mortality and clinical success are presented in Tables 4 and 5. SOFA score was independently associated with the risk of clinical failure and death. Additionally, patient age was an independent risk factor associated with ICU mortality, whereas $C_{\text{max}}$ attainment was not independently associated with clinical success and ICU mortality. The
Table 4: Multivariate analysis of clinical success for all AG considering targeted C\text{\text{max}} (maximal concentration) attainment

| Clinical success | OR (95% CI) | P value |
|------------------|-------------|---------|
| Medical admission| 1.08 [0.67; 1.74] | 0.74 |
| Male             | 0.60 [0.32; 1.12]  | 0.11 |
| Age (years)      | 1.00 [0.99; 1.02]  | 0.67 |
| BMI (kg m\textsuperscript{-2}) | 1.01 [0.97; 1.04]  | 0.91 |
| SOFA score       | 0.85 [0.72; 0.91]  | <0.0001 |
| Serum creatinine (µmol L\textsuperscript{-1}) | 1.00 [0.99; 1.0]  | 0.52 |
| Empirical treatment | 0.71 [0.33; 1.53]  | 0.39 |
| Bacteremia       | 0.87 [0.49; 1.55]  | 0.64 |
| C\text{\text{max}} attainment | 1.24 [0.79; 1.94]  | 0.35 |

Italic values indicate P values <0.05

OR were calculated with a GEE model assuming an exchangeable correlation structure. P values result of a Wald test. The regression was based on 360 patients. The targeted C\text{\text{max}} was defined as a serum level above 60 mg L\textsuperscript{-1} for amikacin and 20 mg L\textsuperscript{-1} for gentamicin and tobramycin

OR: odds ratio; CI: confidence interval, BMI: body mass index, GEE: generalized estimating equations

Table 5: Multivariate analysis of ICU mortality for all AG considering targeted C\text{\text{max}} (maximal concentration) attainment

| ICU mortality | OR (95% CI) | P value |
|---------------|-------------|---------|
| Medical admission | 1.28 [0.75; 2.17] | 0.36 |
| Male          | 1.08 [0.73; 1.60]  | 0.69 |
| Age (years)   | 1.03 [1.01; 1.04]  | 0.001 |
| BMI (kg m\textsuperscript{-2}) | 1.00 [0.97; 1.04]  | 0.8 |
| SOFA score    | 1.18 [1.10; 1.26]  | <0.0001 |
| Serum creatinine (µmol L\textsuperscript{-1}) | 0.99 [0.99; 1.01]  | 0.32 |
| Empirical treatment | 0.81 [0.45; 1.48]  | 0.5 |
| Bacteremia    | 0.62 [0.31; 1.25]  | 0.18 |
| C\text{\text{max}} attainment | 0.78 [0.46; 1.31]  | 0.34 |

Italic values indicate P values <0.05

OR were calculated with a GEE model assuming an exchangeable correlation structure. P values result of a Wald test. The regression was based on 393 patients. The targeted C\text{\text{max}} was defined as a serum level above 60 mg L\textsuperscript{-1} for amikacin and 20 mg L\textsuperscript{-1} for gentamicin and tobramycin

OR: odds ratio; CI: confidence interval, BMI: body mass index, GEE: generalized estimating equations

Discussion

Main findings

Short course of high doses of AG is the dosing regimen extensively used at an international level with amikacin being the most commonly AG prescribed in this large cohort of septic patients. AG TDM is current practice for most of participating ICUs, although ICU patients for whom this monitoring is recommended vary between local guidelines. Finally, in the subgroup of patients for whom TDM was performed, we could find no correlation between C\text{\text{max}} target attainment and ICU mortality or clinical success (Fig. 2).

Relationship to previous papers

Although AG have been prescribed for decades, administering optimal dosing regimens in critically ill patients with sepsis or septic shock remain challenging due to several pathophysiological changes affecting drug PK and due to the emergence of resistance. Additionally, nephrotoxicity has been one of the reasons limiting AG use in critically ill patients with frequent compromised renal function. Aminoglycoside dose optimization can be defined as the dose having the highest likelihood of a good outcome and the lowest likelihood of toxicity. Consequently, doses have to be given once daily and should be stopped as early as possible [29]. Extended interval of high doses of AG has been suggested by several authors to maximize efficacy while limiting the probability of nephrotoxicity [17, 18]. The findings of the present study confirm that, in critically ill patients receiving AG therapy, this dosing regimen is widely used. Still, this higher dosing regimen is considered as an “off-label” dosing regimen in most countries. Rybak et al. have previously reported low prevalence of nephrotoxicity (1.2%) when short courses (3 days) of AG were administered [30]. In the present study, more than 20% of patients had persistent AKI at the end of treatment. Similar findings have been reported by Duszynska et al. in septic patients reporting that 24% of patients developed AKI during amikacin therapy [31]. Finally, the rate of nephrotoxicity attributable to AG therapy is difficult to evaluate as several confounders may influence the persistence of AKI in septic patients. Sepsis-related AKI ranges from 20 to 50%, independently of AG use [32].

Amikacin is the AG most being prescribed in this large cohort of critically ill patients. However, clinical practices widely vary among countries and local epidemiology. Indeed, given that the vast majority of ICUs were located in France, it may explain the use of amikacin in two thirds of patients in this cohort as amikacin is commonly prescribed in this country. Even though the number of patients receiving gentamicin was smaller in our cohort, it is interesting to note that the target attainment rate is lower (59%) than for amikacin (71%). The median dose of 6 mg kg\textsuperscript{-1} appears insufficient to achieve PK/PD target. Indeed, an increased volume of distribution (0.41 L kg\textsuperscript{-1}) of gentamicin has been observed in sepsis leading to gentamicin underexposure and increasing risk of different other models tested did not find a significant difference in the association between clinical outcomes and the variables of interest related to AG.
therapeutic failure [33]. Higher gentamicin loading doses should be given to critically ill patients as reported previously [34]. Similarly, in a previous single centre study, we showed that a gentamicin dosing regimen of 8 mg kg\(^{-1}\) of total body weight failed to reach the PK/PD target of at least 8 times the MIC in a significant proportion of critically ill patients considering an MIC of 2 mg L\(^{-1}\). Considering actual MIC, the proportion of patients experiencing underdosing could be lower as most of the identified pathogens in this study had an MIC < 2 mg L\(^{-1}\). Nevertheless, when AG are used as empirical therapy at the early phase of septic shock where bacterial inoculum is supposed to be high and antibiotic appropriateness is essential, the worst-case scenario (i.e., covering the least susceptible pathogen) should be considered to choose optimal dosing regimen to ensure therapeutic efficacy.

AG TDM has been advocated to optimize AG drug dosing in the setting of ICU patients targeting a \(C_{\text{max}}/\text{MIC}\) ratio of at least 8 to maximize the effect of treatment [35, 36]. In a previous study, Duszynska et al. showed that among septic patients, 83% required dose and/or interval adjustment during amikacin therapy [31]. Conversely to this study and others showing good clinical outcomes correlated to this PK/PD target, we failed to demonstrate any correlation between \(C_{\text{max}}\) target attainment and clinical outcome in the present study (Fig. 2) [37]. One potential explanation could be the wide use of combination therapy with beta lactams to treat severe infections. The impact of non-target attainment for AG could be minimized by the administration of appropriate combined antibiotic questioning the need for adding AG in this setting. However, it is noteworthy that AG were the only antibiotics effective against the causative agent for 22 patients. Also, recent guidelines on the treatment of sepsis have recommended using combination therapy, especially in patients with septic shock [38, 39]. Consequently, it is important to optimize AG dosing as if they were the only effective agent being used. Unfortunately, the small size of this cohort of patients for whom AG enabled to broaden spectrum activity precluded to identify any correlation between target attainment and clinical outcome. Another important finding is the significant proportion (67%) of patients that still did not achieve PK/PD targets in our cohort, even though high doses of AG were mostly used. Previously, some authors aimed to identify the main determinants of the first amikacin peak in critically ill patients [40]. Based on their findings, Boidin et al. supported the use of an amikacin dose \(\geq 37.8\) mg kg\(^{-1}\) of lean body weight (LBW) to optimize the attainment of \(C_{\text{max}} \geq 80\) mg L\(^{-1}\) after the first dose in critically ill patients [40]. As interindividual \(C_{\text{max}}\) variability has been observed in several previous studies, it is unlikely that the “one dose fits all” strategy would be the optimal strategy in this population. Conversely, TDM should be considered even when short courses of AG are administered. TDM has been shown to decrease hospital stay, and the incidence of nephrotoxicity, mortality, and costs [41]. However, TDM alone may not be sufficient to determine the appropriate dosing to be given. Individualized approaches using Bayesian forecasting and TDM algorithm should be implemented in daily practice to take into account the different factors affecting AG PK in septic patients [41, 42]. These approaches also offer the advantage to estimate area under the curve (AUC) in clinical practice with limited PK sampling. Estimates of \(C_{\text{max}}\) can vary substantially based on the duration of the infusion and the timing of \(C_{\text{max}}\) sampling, whereas AUC appears more reliable and stable measure. Additionally, AUC better reflects cumulative exposure over the entire dosing period and may be less sensitive to differences in concentration sampling times. Thus, AUC-guided TDM has been advocated as a more reliable indicator of bacterial killing and clinical efficacy for AG [43].

**Study limitations**
Several limitations of the AMINO III study needs to be acknowledged.

First, the distribution of the participating ICUs may not reflect the daily practice of all ICUs for each country as in some countries a small number of patients were included. Second, PK/PD outcome assessment was possible in less than half of the included patients. As a result, correlations with patient and site characteristics factors may have been underestimated. However, these findings highlight that, despite national or local recommendations, aminoglycoside TDM is not systematically performed in septic patients. Third, we only considered AG use in septic patients in the present study precluding to extrapolate these findings to other indications of AG therapy. Fourth, assessment of clinical success was not assessed by an independent committee; however, it was clearly predefined and standardized for improving data quality. Finally, ototoxicity was not assessed due to the observational design of the present study.

**Conclusion**

Short courses of high AG doses are mainly used in ICU patients with septic shock, although wide variability in AG usage is reported. In this cohort, we could show no correlation between \(C_{\text{max}}\) target attainment and ICU mortality or clinical success. Efforts to optimize the first AG dose and to perform TDM are still needed.

**Abbreviations**

AG: Aminoglycosides; TDM: Therapeutic drug monitoring; ICU: Intensive-care unit; PK/PD: Pharmacokinetics/pharmacodynamics; MIC: Minimal inhibitory
concentration; BMI: Body mass index; AKI: Acute Kidney Injury Network; CRF: Case report form; TBW: Total body weight; IBW: Ideal body weight; ABW: Adjusted body weight; $C_{\text{max}}$: Maximal plasma concentration; $C_{\text{AUC}}$: Minimal plasma concentration; MDR: Multidrug resistant; LBW: Lean body weight; AUC 1: Area under the curve.

**Supplementary Information**

The online version contains supplementary material available at https://doi.org/10.1186/s13613-021-00834-4.

**Additional file 1: Figure S1.** Representation of AMINO III study participating sites. Dark gray: study participating countries

**Additional file 2: Table S1.**

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**Authors' contributions**

CR, JYL, and JR contributed to study design, data collection, data analysis, and drafting the manuscript. GB, LE, DK, and JL contributed to study coordination, data collection, and data interpretation, and ML, LM, CB, JA, IB, JC, JB, JMC, and JA significantly contributed to data collection and data analysis. LE contributed to study coordination and data monitoring. BL contributed to study design, data analysis and statistical analysis. All authors read and approved the final manuscript. The Azurea group endorsed the conduct of the study.

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**Availability of data and materials**

The dataset used and analyzed during the current study are available from the corresponding author on reasonable request.

**Declarations**

**Ethics approval and consent to participate**

The study was approved by Institutional Review Board of Nîmes University Hospital, France (15/04.08). Written informed consent was waived due to the non-interventional design of the study.

**Consent for publication**

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**Competing interests**

The authors declare that they have no competing interests.

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**References**

1. Kollef MH, Sherman G, Ward S, Fraser VI. Inadequate antimicrobial treatment of infections: a risk factor for hospital mortality among critically ill patients. Chest. 1999;115(2):462–74.

2. Feucht D, Zanotti S, Taiberg L, et al. Duration of hypotension before initiation of effective antimicrobial therapy is the critical determinant of survival in human septic shock. Crit Care Med. 2006;34(6):1589–96.

3. Dellinger RP, Levy MM, Rhodes A, et al. Surviving sepsis campaign: international guidelines for management of severe sepsis and septic shock. 2012. Crit Care Med. 2013;41(2):580–637.
5. Pinder M, Bellomo R, Lipman J. Pharmacological principles of antibiotic prescription in the critically ill. Anaesth Intens Care. 2002;30(2):134–44.

6. Brunkhorst FM, Oppert M, Marx G, Bloos F, Ludwig K, Putensen C, Nierhaus A, Jaschinski U, Meier-Hellmann A, Weyland et al. Effect of empirical treatment with moxifloxacin and meropenem vs meropenem on sepsis-related organ dysfunction in patients with severe sepsis: a randomized trial. JAMA. 2012;307(22):2390–9.

7. Falagas ME, Koptides P. Old antibiotics for infections in critically ill patients. Curr Opin Crit Care. 2007;13(5):592–7.

8. Moore RD, Smith CR, Lietman PS. The association of aminoglycoside plasma levels with mortality in patients with gram-negative bacteremia. J Infect Dis. 1984;149(3):443–8.

9. Moore RD, Smith CR, Lietman PS. Association of aminoglycoside plasma levels with therapeutic outcome in gram-negative pneumonia. Am J Med. 1984;77(4):657–62.

10. Moore RD, Lietman PS, Smith CR. Clinical response to aminoglycoside therapy: importance of the ratio of peak-concentration to minimal inhibitory concentration. J Infect Dis. 1987;155(1):93–9.

11. Deziel-Evans LM, Murphy JE, Job ML. Correlation of pharmacokinetic indices with therapeutic outcome in patients receiving aminoglycosides. Clin Pharm. 1986;5(4):319–24.

12. Agence fran
cçaise de sécurité sanitaire des produits de s. Update on good use of injectable aminoglycosides, gentamicin, tobramycin, netilmicin, amikacin Pharmacological properties, indications, dosage, and mode of administration, treatment monitoring. Med Mal Infect. 2012;42:301–8.

13. Roberts JA, Lipman J. Pharmacokinetic cues for antibiotics in the critically ill patient. Crit Care Med. 2009;37(3):840–51 (quiz 859).

14. Taccone FS, Latreille PF, Sapnen H, Duguet T, Delattre I, Layeux B, De Backer D, Wittebole X, Wallenmâc P, Vincent JL, et al. Revisiting the loading dose of amikacin for patients with severe sepsis and septic shock. Crit Care. 2010;14(2):R53.

15. Galvez R, Luengo C, Cornejo R, Kosche J, Romero C, Tobar E, Illanes V, Llanos O, Castro J. Higher than recommended amikacin loading doses achieve pharmacokinetic targets without associated toxicity. Int J Antimicrob Agents. 2011;38(2):146–51.

16. Roger C, Nucci B, Molinari N, Bastide S, Saissi G, Pradel G, Barbar S, Aubert A, Brisset V, Boulanger C, Lloret S, Elotmani L, et al. Standard dosing of amikacin and gentamicin in critically ill patients results in variable and subtherapeutic concentrations. Int J Antimicrob Agents. 2015;46(1):21–7.

17. de Montmoreuill E, Bouadma L, Gault N, Mourvillier B, Mariotte E, Chemam S, Massias L, Papy E, Tubach F, Wolff M, et al. Predictors of insufficient amikacin peak concentration in critically ill patients receiving a 25 mg/kg total body weight regimen. Intensive Care Med. 2014;40(7):998–1005.

18. Roger C, Nucci B, Louart B, Friggeri A, Knani H, Evrard A, Lavigne JP, Initiative S. The strengthening the reporting of observational studies in epidemiology (STROBE) statement: guidelines for reporting observational studies. Lancet. 2007;370(9596):1453–7.

19. Toulouse E, Lafont B, Granier S, McGurk G, Bazin JE. French legal approach to patient consent in clinical research. Anaesth Crit Care Pain Med. 2007;26(6):1258–63.

20. Jager NG, van Hest RM, Lipman J, Taccone FS, Roberts JA. Therapeutic drug monitoring of anti-infective agents in critically ill patients. Expert Rev Clin Pharmacol. 2016;9(7):961–79.

21. Ruiz J, Ramirez P, Company MJ, Gordon M, Villarreal E, Concha P, Aroca M, Frasquet J, Remedios-Marques M, Castellanos-Ortega A. Impact of amikacin pharmacokinetic/pharmacodynamic index on treatment response in critically ill patients. J Glob Antimicrob Resist. 2018;12:58–63.

22. Galvez R, Luengo C, Cornejo R, Kosche J, Romero C, Tobar E, Illanes V, Llanos O, Castro J. Higher than recommended amikacin loading doses achieve pharmacokinetic targets without associated toxicity. Int J Antimicrob Agents. 2011;38(2):146–51.

23. Roger C, Nucci B, Molinari N, Bastide S, Saissi G, Pradel G, Barbar S, Aubert A, Brisset V, Boulanger C, Lloret S, Elotmani L, et al. Standard dosing of amikacin and gentamicin in critically ill patients results in variable and subtherapeutic concentrations. Int J Antimicrob Agents. 2015;46(1):21–7.

24. Galvez R, Luengo C, Cornejo R, Kosche J, Romero C, Tobar E, Illanes V, Llanos O, Castro J. Higher than recommended amikacin loading doses achieve pharmacokinetic targets without associated toxicity. Int J Antimicrob Agents. 2011;38(2):146–51.

25. Zeger SL, Liang KY. Longitudinal data analysis for discrete and continuous outcomes. Biometrics. 1986;42(1):121–30.

26. Gardiner JC, Luo Z, Roman J. Fixed effects, random effects and GEE: an international community of software platform partners. J Biomed Inform. 2019;55:103208.

27. White H. A Heteroskedasticity-Consistent Covariance Matrix Estimator and a Direct Test for Heteroskedasticity. Econometrica. 1980;48:817–38.

28. R Core Team (2019). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria URL https://www.R-project.org/

29. Drusiano GL, Louie A. Optimization of aminoglycoside therapy. Antimicrob Agents Chemother. 2011;55(6):2528–31.

30. Rybak MJ, Abate BJ, Kang SL, Rufking MJ, Lerner SA, Drusano GL. Prospective evaluation of the effect of an aminoglycoside dosing regimen on rates of observed nephrotoxicity and ototoxicity. Antimicrob Agents Chemother. 1999;43(7):1549–55.

31. Wegler M, Paquet JS, Moutier D, Kellou C, Hillel J. Pharmacokinetics of aminoglycosides in critically ill patients. J Antimicrob Chemother. 2009;63(3):572–9.

32. Bellomo R, Kellum JA, Ronco C, Wald R, Martensson J, Maiden M, Bagshaw SM, Glassford NJ, Lankadeva Y, Vaara ST, et al. Acute kidney injury in sepsis. Intensive Care Med. 2017;43(6):816–28.

33. Goncalves-Pereira J, Martins A, Povoa P. Pharmacokinetics of gentamicin in critically ill patients: pilot study evaluating the first dose. Clin Microbiol Infect. 2010;16(8):1258–63.

34. Rea RS, Capitanio B, Bies R, Bigos KL, Smith R, Lee H. Suboptimal aminoglycoside dosing in critically ill patients. Ther Drug Monit. 2008;30(6):674–81.

35. Jager NG, van Hest RM, Lipman J, Taccone FS, Roberts JA. Therapeutic drug monitoring of anti-infective agents in critically ill patients. Expert Rev Clin Pharmacol. 2016;9(7):961–79.

36. Jenkins A, Thomson AH, Brown NM, Semple Y, Sluman C, MacGowan A, Lovering AM, Wiffen PJ. Amikacin use and therapeutic drug monitoring in adults: do dose regimens and drug exposures affect either outcome or adverse events? A systematic review. J Antimicrob Chemother. 2016;71(10):2754–9.

37. Ruiz J, Ramirez P, Company MJ, Gordon M, Villarreal E, Concha P, Aroca M, Frasquet J, Remedios-Marques M, Castellanos-Ortega A. Impact of amikacin pharmacokinetic/pharmacodynamic index on treatment response in critically ill patients. J Glob Antimicrob Resist. 2018;12:90–5.

38. Rhodes A, Evans LE, Alhazzani W, Levy MM, Antonelli M, Ferrer R, Kumar A, Sevransky JE, Sprung CL, Nunnally ME, et al. Surviving Sepsis Campaign: International Guidelines for Management of Sepsis and Septic Shock: 2016. Crit Care Med. 2017;45(3):486–552.

39. Bretonniere C, Leone M, Milesi C, Allacouchiche B, Armand-Lefevre L, Balides O, Bouadma L, Decre D, Figueiredo S, Gauzit R, et al. Strategies to reduce curative antibiotic therapy in intensive care units (adult and paediatric). Intensive Care Med. 2015;41(7):1181–96.

40. Bodin C, Jenck S, Bourguignon L, Torkmani S, Rousse-Jean A, Leduc‑chowdri S, Mary J, Ammenouch N, Dupont H, Marcon F, et al. Determinants of amikacin first peak concentration in critically ill patients. Fundam Clin Pharmacol. 2018;32(6):669–77.

41. van Lent‑Evers NA, Mathot RA, Geus WP, van Hout BA, Vinkis AA. Impact of goal-oriented and model-based clinical pharmacokinetic dosing of aminoglycosides on clinical outcome: a cost-effectiveness analysis. Ther Drug Monit. 1999;21(1):63–73.

42. Venisse N, Bouamery A. Groupe Suivi Therapeutique Pharmacologique de la Societe Francaise de Pharmacologie et de T. [Level of evidence for therapeutic drug monitoring of aminoglycosides]. Therapie. 2011;66(1):39–44.

43. Craig WA. Optimizing aminoglycoside use. Crit Care Clin. 2011;27(1):107–21.

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