A Case Report of Persistent *Acinetobacter baumannii* Bloodstream Infection

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Abstract

**Introduction:** Bloodstream infection due to *Acinetobacter baumannii* is characterized as a nosocomial infection, particularly occurs in intensive care units (ICUs). It is an opportunistic bacterium and therefore, almost exclusively affects predisposed patients who have undergone invasive procedures.

**Case Presentation:** We present a detailed clinical course of a persistent blood-stream infection due to *A. baumannii*.

**Conclusions:** In this case, clinical remission was achieved after several treatments with combinations of sulbactam, containing cefoperazone sodium/sulbactam, carbapenems, minocycline, and aminoglycosides. Despite not achieving bacterial clearance, the treatment presumably reduced the number of bacteria in the blood. Our findings shed light on the treatment of persistent bloodstream infection due to *A. baumannii*.

**Keywords:** *Acinetobacter baumannii*, Bloodstream Infection, Treatment

1. Introduction

Bloodstream infection due to *Acinetobacter baumannii* is characteristically a nosocomial infection with a fatality rate of up to 37% (1). Suspicion of bloodstream infection warrants immediate blood tests, and if bloodstream infection is confirmed, pathogen-based treatment should be initiated immediately. Bloodstream infection, followed by respiratory tract and surgical wound infections, is the most significant infection caused by *A. baumannii* (2). It is an opportunistic agent and therefore, almost exclusively affects predisposed patients who have undergone invasive procedures. Currently, reports on persistent bloodstream infections caused by *A. baumannii* are rare. This report systematically describes the course of persistent bloodstream infection and provides some information regarding the treatment of *A. baumannii*.

2. Case Presentation

The patient was a 58-year-old male who complained of “recurrent fever of more than 18 months and persistent fever of more than 20 days after graft replacement for left common iliac artery aneurysm two years ago”. He was admitted to our hospital on November 23, 2010. In November 2008, he was found to have an intraabdominal mass in a medical examination. He was diagnosed with a “left common iliac artery aneurysm” after which he underwent graft replacement in November 2008. Six months after the surgery, he developed recurrent high fever and chills without an apparent source. All blood cultures revealed the growth of *A. baumannii*. He received several combinations of imipenem, cefoperazone sodium/sulbactam, minocycline, rifampicin, and other agents for 4 to 12 weeks. However, he still exhibited fever with an interval ranging from weeks to months. Blood cultures obtained during the symptom-free interval remained positive (for *A. baumannii*).

Twenty days before the admission, he again had a fever with body temperature as high as 42°C and chills accompanied by shortness of breath. Loxoprofen was helpful to reduce his body temperature to normal, but minocycline had no effect on lowering his body temperature or alleviating chills. Since the onset of his condition, he had no cough or expectoration, no chest tightness or shortness of breath, no urinary frequency, urgency, or dysuria, no abdominal pain, and no diarrhea. He ate and slept normally and had normal urine and bowel movements. His body weight did not decline significantly. Physical exami-
nation on the admission was as follows: The patient's general condition was fair. His temperature, pulse rate, respiratory rate, and blood pressure were 37°C, 86 beats per min, 20 breaths per min, and 120/70 mmHg, respectively. He was conscious, breathing steadily, and had no cyanosis of the lips. No lymph nodes were palpable on the cervical area. His lungs were clear to auscultation bilaterally without rales or rhonchi. No cardiac murmur was heard. An abdominal surgery scar and abdominal mass were visible on the otherwise soft abdomen. The mass was approximately 5 cm × 5 cm and palpable under the xiphoid process, which was soft and slightly tender. The liver and spleen were not clinically palpable. Both lower limbs and feet were positive in terms of edema.

Past medical history: Four years ago, the patient was found to have elevated blood sugar in a physical examination and was diagnosed with Type II diabetes. He received oral metformin, acarbose, and glimepiride, and his glycemic level was stable (fasting blood glucose: 6.0 - 8.0 mmol/L; 2 h postprandial blood glucose: 8.0 - 10.0 mmol/L). Laboratory indicators (November 23, 2010): white blood cell (WBC), 5.29 × 10⁹/L; hemoglobin (HGB), 62 g/L; platelet (PLT), 146 × 10⁹/L; erythropoietin, 72.4 mIU/mL; alanine transaminase (ALT), 5 U/L; aspartate transaminase (AST), 9 U/L; albumin (ALB), 28.4 g/L; serum creatinine (Cr), 45 µmol/L; serum urea nitrogen (BUN), 3.91 mmol/L; potassium ion (K⁺), 3.06 mmol/L; calcium ion (Ca²⁺), 1.90 mmol/L; magnesium ion (Mg²⁺), 0.73 mmol/L; 24 h urinary protein, 0.705 g/24 h; C-reactive protein (CRP), 92.4 mg/L; erythrocyte sedimentation rate (ESR, th), 82 mmH₂O.

Blood cultures: A. baumannii (+) for ten times; and almost resistant to imipenem. They were all resistant to ceftazidime, ciprofloxacin, cefepime, gentamicin, piperacillin, trimethoprim/sulamethoxazole, piperacillin/tazobactam, ceftriaxone (Figure 1). According to CLSI (2010), susceptibility breakpoints were as follows: ≤ 8.0 µg/mL (ceftazidime), ≤ 1.0 µg/mL (ciprofloxacin), ≤ 8.0 µg/mL (cefepime), ≤ 4.0 µg/mL (gentamicin), ≤ 16.0 µg/mL (piperacillin), ≤ 2.0/38.0 µg/mL (trimethoprim/sulamethoxazole), ≤ 16.0/4.0 µg/mL (piperacillin/tazobactam), and ≤ 8.0 µg/mL (ceftriaxone).

Cardiac Doppler and cardiac function tests revealed left ventricle enlargement, mild mitral regurgitation, and aortic valve degeneration; left ventricular systolic function was normal. The CTA (64-row) of the abdomen and the artery of lower extremity showed that the wall of the abdominal aorta was thickened and the formation of multiple calcified plaques could be seen (Figure 2). The inferior segment of the abdominal aorta and the near segment of both common iliac arteries, internal iliac arteries, and external iliac arteries were occlusive. The insertion of the blood vessel, which was from the inferior segment of the abdominal aorta to both external iliac arteries could be seen. The stoma between the insertion of blood vessel and abdominal aorta was a little narrow. The multiple calcified plaques could be seen on the wall of both femoral arteries, the lumens of which was not narrow obviously. The shape and situation of both popliteal arteries, anterior tibial arteries, and dorsal arteries of foot were normal. The wall of them was smooth without obvious abnormalities.

Treatment after admission: The patient had underlying diabetes and repeatedly contracted high fever and chills after graft replacement. He had been admitted to our hospital with a fever on three occasions, and all blood cultures were positive for A. baumannii (Figure 1). Therefore, the persistent bloodstream infection by A. baumannii was confirmed. He received comprehensive anti-infective therapy: intravenous cefoperazone sodium/sulbactam (1:1) 3.0 g q6h, oral minocycline 50 mg twice a day (bid), and imipenem 0.5g every 8 hours (q8h) for 6 weeks. After the antibiotics were discontinued, the patient did not complain of discomfort. His medical indicators after antibiotic discontinuation were as follows: WBC: 5.31 × 10⁹/L; RBC: 2.94 × 10¹²/L; HGB: 66 g/L; PLT: 169 × 10⁹/L; ALT: 4 U/L; AST: 11 U/L; ALB: 31.2 g/L; CRP: 13.1 mg/L; ESR (th): 15mmH₂O; Cr: 50 µmol/L; and BUN: 4.5 mmol/L. The patient maintained a stable body temperature, permitting his discharge.

3. Discussion

The history of diabetes and vascular graft implantation, long-term intermittent fever with chills, and positive blood cultures during three hospitalizations indicated infection with pan-resistant A. baumannii. Therefore, persistent bloodstream infection by A. baumannii is the likely culprit for this admission. Esophageal echocardiography was performed to eliminate the possibility of heart valve vegetations. The results suggest no suspicious echo in any of the valves or the left atrial appendage, thus excluding the possibility of infective endocarditis. Meanwhile, in light of the past graft implantation, imaging tests on the patient’s surgical site were performed several times to locate lesions that may require surgical intervention. The CTA (64-row) of the abdomen and the artery of lower extremity showed that the wall of the abdominal aorta was thickened and the formation of multiple calcified plaques could be seen. The inferior segment of the abdominal aorta and the near segment of both common iliac arteries, internal iliac arteries, and external iliac arteries were occlusive. The insertion of the blood vessel, which was from the inferior segment of the abdominal aorta to both external iliac arteries could also be seen. Based on the clinical manifestations and imaging results, the surgical consultation suggested

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that the postoperative changes of the lower abdominal aorta and bilateral iliac arteries did not suggest vascular graft infection, thus no surgical intervention was needed.

The blood culture results of the past three admissions are listed in Figure 1. The initial susceptibility result indicated that the *A. baumannii* strains were resistant or intermediate sensitive to most antimicrobial agents. However, after several hospitalizations and medications, the blood cultures and susceptibility tests suggested the presence of pan-resistant *A. baumannii*. The evolution of the resistance spectrum may be the result of drug selective pressure, which changes the mechanism of drug resistance (3). According to our previous study on multidrug-resistant *A. baumannii* (4), enzyme-producing and active efflux pump mechanisms may coexist. As drugs against pan-resistant *A. baumannii* are limited at present, it is important to use them in proper combinations to optimize drug synergy and prevent the emergence of resistance during the treatment (5). Typical pairs include the following: carbapenems + sulbactam; polymyxins + rifampicin; colistin + minocycline; colistin + doxycycline; and sulbactam + carbapenems or cefepime (6-8).

In this case, the patient received the comprehensive anti-infective therapy consisting of cefoperazone sodium/sulbactam + oral minocycline 50 mg bid + imipenem (Tienam). The treatment prolonged the symptom-free intervals decreased his body temperature to normal and improved his clinical symptoms. By the sixth week of medication, the intravenous antibiotics were discontinued. Blood culture was performed for bacteriological evaluation on the same day when the patient did not exhibit fever. To our disappointment, the blood culture of both upper extremities was still positive for *A. baumannii*. Although the course of the treatment and time of drug discontinuance are critical for complete remission, it is unclear whether treatment should be discontinued upon observing clinical improvement or waiting until bacteria elimination is confirmed. Considering the outcomes for this patient, we believe that clinical improvement is a more suitable indicator of the therapeutic effect. Furthermore, we stopped the use of antibiotics at the end of week 6 to prevent further drug resistance. In the viewpoint of the weak virulence of the strains, we provided adjuvant therapy to enhance the patient's immunity. Fortunately, he did not experience fever, even after discharge.

Although polymyxin plus doxycycline can be used for the treatment of pan-resistant *A. baumannii* (9, 10), they are not clinically available. In this case, clinical remission was achieved after several treatments with combinations of sulbactam, containing cefoperazone sodium/sulbactam, carbapenems, minocycline, and aminoglycosides. Despite not achieving bacterial clearance, the treatment presumably reduced the number of bacteria in the blood. Our findings shed light on the management of persistent bloodstream infection due to *A. baumannii*. 

![Figure 1. Blood cultures and sensitivity tests of Acinetobacter baumannii. Abbreviations: AMK, amikacin; CAZ, ceftazidime; CIP, ciprofloxacin; FEP, cefepime; GEN, gentamicin; IPM, imipenem; LVX, levofloxacin; PIP, piperacillin; SXT, trimethoprim/sulfamethoxazole; TZP, piperacillin/tazobactam; CRO, ceftriaxone; CSL, cefoperazone/sulbactam cefepime; MNO, minocycline. R, resistant; S, sensitive; I, intermediary.](image-url)
3.1. Conclusions

In this case, clinical remission was achieved after several treatments with combinations of sulbactam, containing cefoperazone sodium/sulbactam, carbapenems, minocycline, and aminoglycosides. Despite not achieving
bacterial clearance, the treatment presumably reduced the number of bacteria in the blood. Our findings shed light on the treatment of persistent bloodstream infection due to A. baumannii.

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Footnotes

Authors’ Contribution: Wan Zhu, Yang Wang, and Jing-ping Zhang drafted the manuscript. Wenxia Cao collected the data from the computerized medical database.

Conflict of Interests: The authors declare that they have no competing interests.

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References

1. Chiu CH, Lee YT, Wang YC, Yin T, Kuo SC, Yang YS, et al. A retrospective study of the incidence, clinical characteristics, identification, and antimicrobial susceptibility of bacteremic isolates of Acinetobacter ursingii. BMC Infect Dis. 2015;15:400. doi: 10.1186/s12879-015-1452-2. [PubMed: 26423424]. [PubMed Central: PMC4590286].

2. Cisneros JM, Rodriguez-Bano J. Nosocomial bacteremia due to Acinetobacter baumannii: Epidemiology, clinical features and treatment. Clin Microbiol Infect. 2002;8(1):59-71. [PubMed: 11955005].

3. Zilberberg MD, Nathanson BH, Sulham K, Fan W, Shorr AF. Multidrug resistance, inappropriate empiric therapy, and hospital mortality in Acinetobacter baumannii pneumonia and sepsis. Crit Care. 2005;20(1):211. doi: 10.1186/cc3192. [PubMed: 15979459]. [PubMed Central: PMC4946760].

4. Zhang JP, Zhu W, Tian SF, Chu YZ, Chen BY. Molecular characteristics and resistant mechanisms of imipenem-resistant Acinetobacter baumannii isolates in Shenyang, China. J Microbiol. 2010;48(5):689-94. doi: 10.1007/s12275-010-0137-3. [PubMed: 21046349].

5. Guan X, He L, Hu B, Hu J, Huang X; Chinese X. D. R. Consensus Working Group, et al. Laboratory diagnosis, clinical management and infection control of the infections caused by extensively drug-resistant Gram-negative bacilli: A Chinese consensus statement. Clin Microbiol Infect. 2016;22 Suppl 1:515–25. doi: 10.1016/j.cmi.2015.11.004. [PubMed: 26627340].

6. Bernabeu-Wittel M, Pichardo C, Garcia-Curiel A, Pachon-Ibanez ME, Ibanez-Martinez J, Jimenez-Mejias ME, et al. Pharmacokinetic/pharmacodynamic assessment of the in-vivo efficacy of imipenem alone or in combination with amikacin for the treatment of experimental multidrug-resistant Acinetobacter baumannii pneumonia. Clin Microbiol Infect. 2005;11(4):399-405. doi: 10.1111/j.1469-0691.2005.01095.x. [PubMed: 15760436].

7. Saballs M, Pujol M, Tubau F, Pena C, Montero A, Dominguez MA, et al. Rifampicin/imipenem combination in the treatment of carbapenem-resistant Acinetobacter baumannii infections. J Antimicrob Chemother. 2006;58(3):687-700. doi: 10.1093/jac/dkl274. [PubMed: 16895941].

8. Principe I, D'Arezoo S, Capone A, Petrosillo N, Visca P. In vitro activity of tigecycline in combination with various antimicrobials against multidrug-resistant Acinetobacter baumannii. Ann Clin Microbiol Antimicrob. 2009;8(18). doi: 10.1186/1476-0711-8-18. [PubMed: 19460166]. [PubMed Central: PMC2693502].

9. Garnacho-Montero J, Amaya-Villar R. Multiresistant Acinetobacter baumannii infections: Epidemiology and management. Curr Opin Infect Dis. 2000;13(4):332-9. doi: 10.1097/QCO.0b013e32833a8c2b. [PubMed: 2058874].

10. Michalopoulos A, Falagas ME. Treatment of Acinetobacter baumannii infections. Expert Opin Pharmacother. 2010;11(5):779-88. doi: 10.1517/14656566001398150. [PubMed: 2021064].