Effects of Conditioning Depolarization and Repetitive Stimulation on $Q_\beta$ and $Q_\gamma$ Charge Components in Frog Cut Twitch Fibers

CHIU SHUEN HUI and WEI CHEN

From the Department of Physiology and Biophysics, Indiana University Medical Center, Indianapolis, Indiana 46202

ABSTRACT Charge movement was measured in frog cut twitch fibers with the double Vaseline-gap technique. Steady-state inactivation of charge movement was studied by changing the holding potential from −90 mV to a level ranging from −70 to −30 mV. $Q_\beta$ and $Q_\gamma$ at each holding potential were separated by fitting the $Q-V$ plot with a sum of two Boltzmann distribution functions. At −70 mV $Q_\beta$ and $Q_\gamma$ were inactivated to 54.0% (SEM 2.2) and 82.7% (SEM 3.0) of the amounts at −90 mV. At holding potentials ≥ −60 mV, more $Q_\gamma$ was inactivated than $Q_\beta$, and at −30 mV $Q_\gamma$ was completely inactivated but $Q_\beta$ was not. There was no holding potential at which $Q_\beta$ was unaffected and $Q_\gamma$ was completely inactivated. The differences between the residual fractions of $Q_\beta$ and $Q_\gamma$ are significant at all holding potentials ($P < 0.001$–0.05). The plot of the residual fraction of $Q_\beta$ or $Q_\gamma$ versus holding potential can be fitted well by an inverted sigmoidal curve that is a mirror image of the activation curve of the respective charge component. The pair of curves for $Q_\gamma$ correlates well with those for tension generation or Ca release obtained by other investigators. The time courses of the inactivation of $Q_\beta$ and $Q_\gamma$ were studied by obtaining several $Q-V$ plots with conditioning depolarizations lasting 1–20 s and separating each $Q-V$ plot into $Q_\beta$ and $Q_\gamma$ components by fitting with a sum of two Boltzmann distribution functions. The inactivation time constant of $Q_\beta$ was found to be 5–10 times as large as that of $Q_\gamma$. During repetitive stimulation, prominent $I_\beta$ humps could be observed in TEST-minus-CONTROL current traces and normal $Q_\beta$ components could be separated from the $Q-V$ plots, whether 20 or 50 mM EGTA was present in the internal solution, whether 2 or 10 stimulations were used, and whether the stimuli were separated by 400 ms or 6 s. Repetitive stimulation slowed the kinetics of the $I_\beta$ hump and could shift the $Q-V$ curve slightly in the depolarizing direction in some cases, resulting in an apparent suppression of charge at the potentials that fall on the steep part of the $Q-V$ curve.

Address reprint requests to Dr. Chiu Shuen Hui, Department of Physiology and Biophysics, Indiana University Medical Center, 635 Barnhill Drive, Indianapolis, IN 46202.

Dr. Chen’s present address is Department of Surgery, University of Chicago, 5841 S. Maryland Ave., Chicago, IL 60637.
INTRODUCTION

The preceding paper (Hui and Chen, 1992) shows that 0.5–1.0 mM tetracaine can be used to dissect out a steeply voltage-dependent charge component from the charge versus voltage (Q-V) plots of the total charge measured from cut fibers, referred to as method 1. The results agree qualitatively with those obtained from intact fibers (Hui, 1983a, b), although the dose–response relationships of the effect differ quantitatively in the two preparations. The steeply voltage-dependent component dissected with the pharmacological approach agrees well with the components separated by three other existing, independent methods: method 2 (in intact fibers: Hui, 1983a; in cut fibers: Hui, 1991a), method 3 (in cut fibers: Hui and Chandler, 1990; in intact fibers: Hui, 1991a), and method 4 (in cut fibers: Hui and Chandler, 1991).

Originally, Adrian and Peres (1979) separated $Q_\theta$ and $Q_v$ in intact fibers by changing the holding potential and making use of the difference in steepness between the steady-state inactivation of the two components. They assumed that when the holding potential was set at -40 mV, $Q_\theta$ was completely mobile and $Q_v$ was completely inactivated. Hui (1983b) found some inconsistency between the amounts of $Q_\theta$ in intact fibers separated by their method and by method 2. Because of the inherent difficulties in measuring charge movement in intact fibers with the three-microelectrode technique, the inactivation curves for $Q_\theta$ and $Q_v$ obtained from intact fibers could be subjected to substantial uncertainties. With growing interest in the possibility that $Q_v$ might be the trigger for Ca release from the sarcoplasmic reticulum (SR) (Almers, 1978; Huang, 1982; Hui, 1983b, 1991a; Vergara and Caputo, 1983; Hui and Chandler, 1990, 1991), it is of interest to study the inactivation curves for $Q_\theta$ and $Q_v$ in greater detail and to investigate whether there exists a holding potential at which $Q_\theta$ and $Q_v$ can be separated entirely. Another aim of this work is to separate the inactivation time courses of $Q_\theta$ and $Q_v$ and see if they are different.

In contrast to the “trigger hypothesis” for $Q_v$, a “feedback hypothesis” has also been proposed. In the latter hypothesis, $Q_v$ arises as a result of Ca release, as suggested originally by Dr. Knox Chandler (see Discussions sections in Horowicz and Schneider, 1981, and in Hui, 1983b). One piece of evidence that might be consistent with the feedback hypothesis was provided by Garcia et al. (1990), who observed that the $I_v$ humps in charge movement traces from cut fibers disappeared when the Ca$^{2+}$ released from the SR into the myoplasm was chelated by a high [EGTA], and the effect was reversible. To understand the cause of the disappearance of $I_v$ humps, we loaded cut fibers with different [EGTA], and found that substantial $I_v$ humps still existed during repetitive stimulation.

A preliminary report of some of the findings in this paper has appeared (Chen and Hui, 1989).

METHODS

Solutions

All concentrations are in millimolar.

Relaxing solution. Solution A: 120 K'glutamate, 1 MgSO$_4$, 0.1 K$_2$-EGTA, and 5 K$_2$-PIPES, pH 7.0.

Internal solution. Solution B: 45.5 Cs'glutamate, 20 Cs$_2$-creatine phosphate, 20 Cs$_2$-EGTA, 6.8 MgSO$_4$, 5.5 Cs$_2$-ATP, 5 glucose, and 5 Cs$_2$-PIPES, pH 7.0. Solution C: 20 Cs$_2$-creatine phosphate, 50 Cs$_2$-EGTA, 6.8 MgSO$_4$, 5.5 Cs$_2$-ATP, 5 glucose, and 5 Cs$_2$-PIPES, pH 7.0.
Effects of Depolarization on \( Q_\theta \) and \( Q_v \)

External solution. Solution D: 120 TEA-Cl, 2.5 RbCl, 1.8 CaCl\(_2\), 2.15 Na\(_2\)HPO\(_4\), and 0.85 NaH\(_2\)PO\(_4\), pH 7.1.

TEA\(^+\) and Rb\(^+\) in solution D and Cs\(^+\) in solutions B and C were used to minimize K\(^+\) currents. 1 \( \mu \)M tetrodotoxin was added to solution D to block Na\(^+\) current. Solutions B and C contained no added Ca, except for the trace amount of Ca present in Cs-glutamate, estimated to be 60 \( \mu \)M.

For muscle and fiber preparation, see the preceding paper (Hui and Chen, 1992).

Pulse Protocol

The procedure for measuring charge movement at a holding potential of \(-90\) mV was described in the preceding paper (Hui and Chen, 1992). For a less negative holding potential, \( V_H \), the pulse protocol was modified, as illustrated in Fig. 1. Panel A shows the CONTROL pulse sequence. To compare data at this \( V_H \) with those at \(-90\) mV, the fiber was always held at \(-110\) mV for 240 ms before being stimulated by the CONTROL pulse sequence, consisting of two transitions from \(-110\) to \(-90\) mV, separated by 225 ms. The average of the two ON currents in the signal-averaged CONTROL current trace was used for the subtraction of the linear current components in the single-sweep TEST current trace (see Chandler and Hui, 1990, for the advantage of this protocol).

Fig. 1 B shows the TEST pulse sequence for the two-pulse protocol. The one-pulse protocol can be achieved by omitting the 100 ms post-pulse. The durations of the TEST pulses vary according to the level of depolarization (Figs. 2, 5, 8, 11, and 14) in order to minimize the activation of nonlinear ionic currents.

Sampling of the three analog signals, \( V_1 \), \( V_2 \), and \( I_p \), was started 40 ms before the rising edge of the CONTROL or TEST pulse and lasted until the end of the final interval at \(-90\) mV. The
duration of the final interval was adjusted such that the total sampling interval was either 768
or 1,024 ms. Each point in a current trace corresponds to 1 ms.

The two-pulse protocol should be distinguished from another pulse protocol, referred to as
the double-pulse protocol. The former protocol consists of a TEST pulse and a post-pulse with
no break in between, whereas the latter consists of two identical TEST pulses separated by a
repolarization period of 400 ms. In this paper, the latter pulse protocol was always applied
from a holding potential of -90 mV.

Data Analysis

The procedures for data analysis were also similar to those used in the preceding paper (Hui
and Chen, 1992). For convenience, Eqs. 1–4 in that paper are listed here again. The
steady-state $Q$-$V$ plot obtained with the one-pulse protocol was fitted by a sum of two
Boltzmann distribution functions with CONTROL charge correction:

$$Q(V) = \sum_{i=1}^{\gamma} Q_{i,\max} F_i(V)$$  (1)

$$F_i(V) = F_i(-90) - [F_i(-90) - F_i(-110)][V + 110]/20 - F_i(-110)$$  (2)

$$F_i(V) = 1 + \exp \left(-\frac{V - V_i}{k_i}\right)^{-1}$$  (3)

in which $Q_{i,\max}$ represents the maximum amount of charge, $V_i$ the equi-distribution potential,
and $k_i$ the voltage dependence (or inverse steepness) factor, for $i = \beta$ or $\gamma$. Also, the gap
correction procedure of Hui and Chandler (1990) was applied. Occasionally, a $Q$-$V$ plot was
fitted by a single Boltzmann distribution function, which is equivalent to dropping one of the
two terms on the right-hand side of Eq. 1.

When the two-pulse protocol was used, the $Q$-$V$ plot of the final OFF charge at -90 mV was
fitted by:

$$Q(V) = A + \rho Q_{\gamma,\max} \left[1 + \exp \left(-\frac{V - V_\gamma}{k_\gamma}\right)^{-1}\right]$$  (4)

with gap correction. $A$ and $\rho$ are constants independent of $V$ and have been expressed explicitly
in Hui and Chandler (1991).

In addition, the steady-state inactivation data of $Q_\beta$ or $Q_\gamma$ were fitted by the (normalized)
inverted sigmoidal function:

$$G_i(V) = \left[1 + \exp \left(-\frac{V - V_i}{k_i}\right)^{-1}\right]^{-1}$$  (5)

which is a mirror image of the (normalized) activation curve, Eq. 3 above.

RESULTS

Voltage-dependent Inactivation of $Q_\beta$ and $Q_\gamma$ in a Cut Fiber

In the experiment shown in Fig. 2, charge movement was studied with the one-pulse
protocol. Panel A shows a family of TEST-minus-CONTROL current traces taken at a
$V_H$ of -90 mV. The traces resemble those obtained previously under identical conditions (Hui
and Chandler, 1990; Hui, 1991a, b). At potentials $\leq -65$ mV only fast $I_\beta$ transients can be seen in the ON and OFF segments of the traces. At -60 mV
FIGURE 2. Effect of maintained depolarization on TEST-minus-CONTROL current in a cut fiber. Fiber identification: 89021. Diameter = 93 μm. Sarcomere length = 3.5 μm. Saponin treatment was applied to membrane segments in both end pools at time zero. After rinsing, the solutions in the end pools were replaced by solution B. The solution in the center pool was then changed to an isotonic TEA-Cl solution (solution D). At the 21st minute the voltage clamp was turned on and the holding potential was set at −90 mV. From the beginning to the end of the experiment, the holding current (bracketed at −90 mV) changed from −22 to −29 nA and \( r_d/(r_d + r_i) \) decreased from 0.989 to 0.988. The one-pulse protocol was used to obtain the traces shown in this figure. (A) Traces taken from the 57th to the 76th minute. At the 119th minute the holding potential was changed to −70 mV. (B) Traces taken from the 132nd to the 151st minute. At the 180th minute the holding potential was changed to −60 mV. (C) Traces taken from the 194th to the 213th minute. At the 242nd minute the holding potential was changed to −50 mV. (D) Traces taken from the 256th to the 275th minute. The numbers at the right show the potentials in millivolts during the TEST pulses (the same for both traces in the same row). Only representative traces are shown in each panel.
a broad $I_h$ hump begins to appear in the ON segment, but not in the OFF segment. At $-56$ mV the $I_h$ hump becomes very pronounced. With further depolarizations, the peak amplitude of the $I_h$ hump increases progressively and rises above the peak of the $I_r$ component. At $\geq -30$ mV the hump fuses with, and cannot be visually separated from, the early $I_r$ component.

After changing the $V_H$ to $-70$ mV, the traces in Fig. 2 B were taken. The $I_h$ humps in the ON segments of Fig. 2 A at $-60$ and $-56$ mV disappear in Fig. 2 B, whereas those at more depolarized potentials are suppressed and their time courses are prolonged. The amplitudes of the OFF transients are also suppressed. The traces in Fig. 2 C were taken after the $V_H$ was changed to $-60$ mV. Both ON and OFF transients are further suppressed and there is no sign of any $I_h$ hump in the traces. Surprisingly, the ON segments (OFF segments) show an outward (inward) transient followed by a slower inward (outward) transient. These biphasic characteristics of the current transients also exist in the traces of Fig. 2 B, but to a lesser extent. At a $V_H$ of $-50$ mV (Fig. 2 D), the fast transients in both ON and OFF segments are greatly suppressed and the slower transients greatly enhanced. This biphasic nature of the current transients has been observed previously by Hui and Milton (1987) in intact fibers and by Hui (1990) in cut fibers in the presence of D600, and also by Chen and Hui (1991b) in cut fibers in the presence of nifedipine.

To understand the origin of the biphasic appearance of charge movement, the CONTROL current traces at $V_H$'s of $-90$ and $-40$ mV are compared in Fig. 3. In the upper pair of traces, the ON and OFF transients at a $V_H$ of $-40$ mV show an extra slower component, marked by the arrowheads. A subtraction of the CONTROL current trace at a $V_H$ of $-40$ mV from that at $-90$ mV and removing the constant pedestal during the pulse.

![FIGURE 3. Effect of holding potential on the CONTROL current transient. Same fiber as in Fig. 2. The upper pair of superimposed traces shows the CONTROL current traces, one taken when the holding potential was at $-90$ mV and the other at $-40$ mV (marked by arrowheads). The difference trace at the bottom was obtained by subtracting the CONTROL trace at $-40$ mV from that at $-90$ mV and removing the constant pedestal during the pulse.](image-url)

The origin of the extra slower ON and OFF transients in the CONTROL current trace at a depolarized $V_H$ is not known with certainty. Since the inward and outward deflections in a difference trace are almost symmetrical, they could be capacitive in
nature. A likely candidate for this extra capacitive current is $I_a$ (i.e., $Q_a$ current), which is readily detectable when a fiber is depolarized, as described by Adrian et al. (1976) in intact fibers and Brum and Rios (1987) in cut fibers. When the scaled CONTROL current trace is subtracted from the TEST current trace to generate a TEST-minus-CONTROL current trace at a depolarized $V_H$, this presumed $I_a$ current gives rise to an inward deflection in the ON segment and an outward deflection in the OFF segment. Moreover, the kinetics of the $I_a$ component appears to be not very voltage dependent and is slower than that of the $I_B$ component, but could be comparable to that of the $I_e$ component at some potentials.

The ON and OFF transients in the TEST-minus-CONTROL current traces at a $V_H$ of $-90$ mV (Fig. 2A) do not show any biphasicity. If the slower transients of reverse polarity in the other panels of Fig. 2 are indeed caused by $I_a$ current, then it can be concluded that the amount of $Q_a$ is negligible in a normally polarized fiber, consistent with the suggestion given by Hui (1991b).

The biphasic nature of the ON and OFF transients in TEST-minus-CONTROL current traces at a $V_H$ of $> -90$ mV complicates the fitting of baselines to the ON and OFF segments. The slower $I_a$ transient in the OFF segment was removed, although not perfectly ideally, by fitting a sum of an exponential decay and a sloping baseline up to the 70th point or so in that segment (see Fig. 1 of Hui, 1990). No data analysis was performed on the ON segments.

To quantitate the inactivation of $Q_B$ and $Q_n$, the amounts of total OFF charge at various $V_H$'s are plotted against TEST pulse potential in Fig. 4. The maximum amount of total charge was progressively reduced when the $V_H$ was changed to a less negative level. Curves 1 and 2 were least-squares fitted to the data at $V_H$'s of $-90$ and $-70$ mV, respectively, according to Eq. 1, with gap correction. Curve 1 rises with a shallow slope at the foot and begins to rise steeply at $\sim -65$ mV due to the activation of $Q_o$. The top portion of the curve rises with a shallow slope due to the additional activation of $Q_B$. Curve 2 represents the $Q-V$ distribution at a $V_H$ of $-70$ mV. From the values of $q_{B,max}/C_m$ and $q_{n,max}/C_m$ listed in the figure legend, it appears that the change in $V_H$ decreased the amounts of $Q_B$ and $Q_n$ to $44.7$ and $67.6\%$ of control, respectively. Thus at a $V_H$ of $-70$ mV, more $Q_B$ was suppressed than $Q_n$ in this fiber.

When the $Q-V$ plots at $V_H$'s of $-60$, $-50$, and $-40$ mV were fitted by Eq. 1, the fitting routine did not converge. However, the plots were fitted well by a single Boltzmann distribution function, with CONTROL charge correction and gap correction, represented by curves 3–5. This suggests that probably only one charge component remained mobile at these more depolarized $V_H$'s. Since no $I_n$ humps can be visualized in the traces of Fig. 2, C and D (and at a $V_H$ of $-40$ mV; not shown) and the $k$ values for curves 3–5 are relatively large, it is reasonable to assume that the residual charge at $V_H$'s of $-60$, $-50$, or $-40$ mV belongs to $Q_B$. However, the value of $q_{B,max}/C_m$ for curve 3 is larger than that for curve 2, and the value of $k_B$ for curve 3 is smaller than that for curve 2 (see figure legend), suggesting that there might still be some residual $Q_o$ at a $V_H$ of $-60$ mV, but the amount was too small to be resolved by method 3. A remedy will be presented below, in connection with Figs. 5 and 6.

Under the assumption that all the residual charge at a $V_H$ of $-50$ or $-40$ mV is totally $Q_B$, the residual amounts of $Q_B$ were then estimated to be $36.4$ and $8.6\%$ of control, respectively, at these levels of $V_H$. It should be noted that, because of the presence of $Q_n$, the value of $C_m$ became larger as $V_H$ was changed to a less negative
level (see figure legend). This introduces extra decreases in the values of \( q_{B,max}/C_m \) and \( q_{y,max}/C_m \) in addition to inactivation.

Charge movement was also studied in the same fiber with the two-pulse protocol and the charge components were separated by method 4. Fig. 5A shows TEST-minus-CONTROL current traces taken at a \( V_H \) of \(-90 \) mV. The ON segments of the traces are identical to those in Fig. 2A, implying that the fiber was stable. After \( V_H \) was changed to \(-70 \) mV, the ON and OFF transients were suppressed (Fig. 5B). At a \( V_H \) of \(-60 \) mV the transients were further suppressed (Fig. 5C).

The amounts of OFF charge at various \( V_H \) are plotted against TEST pulse potential function, with CONTROL charge correction and gap correction, to the three data sets. The best fit parameters are:

| Curve | \( \epsilon_m \) | \( q_{B,max}/C_m \) | \( V_B \) | \( k_B \) | \( q_{y,max}/C_m \) | \( V_y \) | \( k_y \) |
|-------|-----------------|-----------------|--------|--------|-----------------|--------|--------|
| 1     | 0.143           | 10.0            | \(-36.9\) | 12.2   | 13.0            | \(-62.0\) | 1.7    |
| 2     | 0.148           | 4.5             | \(-39.0\) | 7.9    | 8.8             | \(-55.9\) | 2.6    |
| 3     | 0.158           | 6.7             | \(-46.4\) | 6.5    |                 |         |        |
| 4     | 0.175           | 3.6             | \(-44.5\) | 18.5   |                 |         |        |
| 5     | 0.195           | 0.9             | \(-54.0\) | 11.0   |                 |         |        |

in Fig. 6. Only data at TEST pulse potentials \( \leq -30 \) mV were included in the analysis. The reason for this restriction is given in the Methods section of the preceding paper (Hui and Chen, 1992). Curves 1-3 were fitted to the three sets of data according to Eq. 4, with gap correction. Since the points at a \( V_H \) of \(-50 \) mV are essentially flat, consistent with the absence of \( Q_y \) in curve 4 of Fig. 4, no curve was fitted to the points. The values of \( q_{y,max}/C_m \) listed in the legend of Fig. 6 show that \( Q_y \) at a \( V_H \) of \(-70 \) mV was suppressed to 69.3% of control, very close to the value of 67.6% obtained with method 3 in Fig. 4. The agreement of the percentages of \( Q_y \) remaining at a \( V_H \) of \(-70 \) mV obtained by methods 3 and 4 implies that the time
constant of $I_q$ during the post-pulse might not have changed when $V_H$ was changed from $-90$ to $-70$ mV. Thus, there is a good chance that the time constant might be similar at a $V_H$ of $-60$ mV.

Curve 3 of Fig. 6 reveals that $Q_v$ was not completely suppressed at a $V_H$ of $-60$ mV and the residual $Q_v$ was 5.3% of control. Thus, method 4 provides a more sensitive means to detect a small amount of $Q_v$ than method 3. Since the amount of $Q_v$ in curve 1 of Fig. 4 is 13.0 nC/µF, there ought to be 0.7 nC/µF of $Q_v$ in curve 3 of the same figure not resolvable by method 3. The amount of $Q_B$ in curve 3 should

![Figure 5. Effect of maintained depolarization on the final OFF current after a constant brief post-pulse. Same fiber as in Fig. 2. The two-pulse protocol was used to obtain the traces in this figure, with the post-pulse potential set at $-62$ mV. (A) TEST-minus-CONTROL current traces taken at a holding potential of $-90$ mV from the 83rd to the 105th minute. (B) Traces taken at a holding potential of $-70$ mV from the 155th to the 177th minute. (C) Traces taken at a holding potential of $-60$ mV from the 217th to the 239th minute. The numbers at the right show the potentials in millivolts during the TEST pulses (the same for both traces in A and B). Only representative traces are shown in each panel.](image-url)
therefore be 6.0 nC/µF, equivalent to 60.1% for the residual \( Q_B \) at a \( V_H \) of -60 mV. The example given above illustrates the usefulness of method 4 in supplementing method 3 in separating charge components, at least qualitatively. Unfortunately, the residual amount of \( Q_B \) at a \( V_H \) of -60 mV so obtained still turned out to be larger than that at a \( V_H \) of -70 mV. This could be due to an underestimation of the value of \( q_v,\max /\epsilon_m \) in curve 3 of Fig. 6 because of scatter of data.

The residual fractions of \( Q_B \) and \( Q_v \) were studied in 26 cut fibers at different \( V_H \)'s. Not all the levels of \( V_H \) were always used in each fiber. In experiments in which more than one \( V_H \) was used, the \( V_H \) was always changed monotonically in a depolarizing direction. The mean values for \( Q_B \) and \( Q_v \) decreased monotonically when the level of \( V_H \) became less negative. On average, more \( Q_B \) was inactivated than \( Q_v \) at a \( V_H \) of -70 mV, but more \( Q_v \) was inactivated than \( Q_B \) at all \( V_H \)'s \( \geq -60 \) mV. The differences are all statistically significant (\( P < 0.001 \) for -70 and -30 mV, <0.01 for -50 and -40 mV, and <0.05 for -60 mV with the two-tailed t test).

The mean residual fractions of \( Q_B \) and \( Q_v \) are plotted against \( V_H \) in Fig. 7. Curves 1 and 2 were fitted to the two sets of data according to Eq. 5. The best fit parameters are: for curve 1, \( V_v = -64.8 \) mV and \( k_v = 3.4 \) mV; and for curve 2, \( V_v = -62.2 \) mV and \( k_v = 13.0 \) mV. The values of \( k_v \) for the inactivation curves in Fig. 7

| Curve | \( \epsilon_m \) | \( q_v,\max /\epsilon_m \) | \( V_v \) | \( k_v \) |
|-------|----------------|----------------|--------|--------|
| 1     | 0.142          | 3.0            | -62.3  | 3.6    |
| 2     | 0.149          | 2.1            | -57.0  | 1.3    |
| 3     | 0.160          | 0.2            | -59.8  | 0.0    |

No curve was fitted to the \( \triangle \)s.
are very close to the values of $k_A$ and $k_B$ for the activation curves of $Q_B$ and $Q_A$ 2.9 and 11.0 mV given by Hui and Chandler (1990) or 2.7 and 10.7 mV given by Hui (1991a). Hence, the activation and inactivation curves of each charge component are mirror images of each other, the same as in intact fibers (Hui, 1983b).

From curve 2, the amount of mobile $Q_B$ at a $V_H$ of -90 mV is 0.89 of the asymptotic value. Strictly speaking, the amount should be exactly 1 by definition. In other words, the curve fitted to the open squares should be forced to pass through 1 at -90 mV. When this was done, the value of $k_B$ was increased somewhat, which even exaggerated the general conclusion that the inactivation curve of $Q_A$ is steeper than that for $Q_B$. Considering the scatter of data, this refinement does not appear to be important.

**Figure 7.** Steady-state inactivation curves of $Q_B$ and $Q_A$. □'s and •'s represent, respectively, the average fractions of $Q_B$ and $Q_A$ that remain mobile at various holding potentials. The fractions at -90 mV are unity, by definition, for both charge components. The error bar for $Q_A$ at -30 mV is absent because the SEM is smaller than the size of the symbol. The two inverted sigmoidal curves 1 and 2 were fitted to the two data sets according to Eq. 5. $n = 20, 5, 7, 5$, and 3 at $V_H = -70, -60, -50, -40$, and -30 mV, respectively.

**Reversibility of the Effect of Maintained Depolarization**

In all the experiments reported here, a constant TEST pulse to -45 mV was always applied before, during, and after a sequence of runs that produced a $Q-V$ plot to track the condition of the fiber at each $V_H$. In seven fibers the $V_H$ was returned to -90 mV after a period of maintained depolarization to investigate whether the inactivation of $Q_B$ and $Q_A$ was reversible. In one of the successful experiments, the control amount of OFF charge was 10.6 nC/μF. During a $V_H$ of -70 and -60 mV, the average amounts of OFF charge were 8.1 and 5.3 nC/μF, respectively. On changing the $V_H$ back to -90 mV, the average OFF charge was 8.7 nC/μF. In another fiber, the control amount of OFF charge was 14.0 nC/μF. During a $V_H$ of -40 mV, the average amount of OFF charge was 0.9 nC/μF. On changing the $V_H$ back to -90 mV, the average OFF charge was 11.0 nC/μF.

In the remaining five fibers, the amounts of OFF charge were less reversible even after a waiting period of 10–20 min upon repolarization. We believe that the
irreversibility observed was not likely to reflect a deterioration in the condition of the fibers, as judged from the relative stability of the linear cable parameters. There was an indication that part of the charge was still locked in the $Q_a$ state, because the TEST-minus-CONTROL current traces after repolarization still showed some residual biphasic appearance similar to that in the traces of Fig. 2, C and D. The degree of persistence of the charge to stay in the inactivated state might depend on the magnitude and the duration of the maintained depolarization. Thus, for a finite period after repolarization, there might still be some uncertainty concerning the state of the fiber. Because of this complication, we are reluctant to chronically depolarize a fiber and apply CONTROL pulses in the positive potential range, as done by Bruin and Rios (1987).

Inactivation Time Courses of $Q_B$ and $Q_V$

To study the inactivation time courses of $Q_B$ and $Q_V$, the family of TEST pulses that gave rise to one $Q-V$ plot was individually preceded by the same conditioning depolarizing pulse having fixed magnitude and duration, and by a 240-ms resting period at −90 mV. Fig. 8A shows TEST-minus-CONTROL current traces taken without conditioning depolarization. The traces in Fig. 8B were taken with TEST pulses preceded by a 1-s conditioning pulse to −40 mV. Although the amplitudes of the $I_B$ components and OFF transients in these traces are not very different from those in the corresponding traces of Fig. 8A, the $I_V$ humps are much less pronounced. This crude visual analysis provides a hint that depolarization to −40 mV for just 1 s might inactivate more $Q_V$ than $Q_B$.

Fig. 8, C and D, shows traces taken with TEST pulses preceded by a conditioning pulse to −40 mV lasting 5 and 20 s, respectively. As the duration of the conditioning pulse was increased, both the ON and OFF transients in the traces were suppressed progressively. There is no sign of any $I_V$ hump in the ON segments of the traces in Fig. 8D. The OFF transients in these traces decay faster than those in Fig. 8A, probably because the OFF transients in Fig. 8D consist purely of $I_B$, which decays faster than $I_V$ in cut fibers (Hui and Chandler, 1991). At the end of the experiment, the $V_{H}$ was changed to −40 mV and traces similar to those of Fig. 2D were taken (not shown). These traces provided information about the steady-state inactivation of $Q_B$ and $Q_V$ at −40 mV.

The amounts of OFF charge in the traces of Fig. 8, and other traces not shown, are plotted against TEST pulse potential in Fig. 9. Curves 1, 2, and 3 were fitted to the data taken without and with 1 and 5 s conditioning depolarization, respectively, according to Eq. 1, with gap correction. The values of $q_{i,\text{max}}/e_m$ listed in the figure legend show that 1 s conditioning depolarization to −40 mV changed the amounts of $Q_B$ and $Q_V$ to 108.1 and 73.1% of control values, and 5 s conditioning depolarization to 86.6 and 32.7%, respectively. The data taken with 20 s conditioning depolarization could not be fitted by Eq. 1 but were fitted well by a single Boltzmann distribution function with CONTROL charge correction and gap correction, represented by curve 4. Since the $I_V$ hump is absent in the traces of Fig. 8D, the residual charge in curve 4 is assumed to belong to $Q_B$ and is 53.8% of control.

The percentages of $Q_B$ and $Q_V$ that remained mobile are plotted against the duration of the conditioning depolarization in Fig. 10. The two smooth curves show
Figure 8. Effect of the duration of conditioning depolarization on TEST-minus-CONTROL current in a cut fiber. Fiber identification: 97171. Diameter = 99 µm. Sarcomere length = 3.5 µm. Saponin treatment was applied to membrane segments in both end pools at time zero. After rinsing, the solutions in the end pools were replaced by solution B. The solution in the center pool was then changed to an isotonic TEA-Cl solution (solution D). At the 22nd minute the voltage clamp was turned on and the holding potential was set at −90 mV. From the beginning to the end of the experiment the holding current changed from −24 to −30 nA and $r_e/(r_e + r_i)$ decreased from 0.987 to 0.985. The fiber was stimulated by the one-pulse protocol. (A) Traces recorded from the 59th to the 79th minute with TEST pulses alone. (B) Traces recorded from the 99th to the 136th minute with TEST pulses, each of which was preceded by a conditioning prepulse to −40 mV for 1 s. (C) Traces recorded from the 178th to the 211st minute with TEST pulses, each preceded by a conditioning prepulse to −40 mV for 5 s. (D) Traces recorded from the 167th to the 216th minute with TEST pulses, each preceded by a conditioning prepulse to −40 mV for 20 s. The numbers at the right show the potentials in millivolts during the TEST pulses (the same for both traces in the same row). Only representative traces are shown in each panel.
the decay time courses of $Q_B$ and $Q_s$. Curve 2 was obtained by fitting a single exponential, whereas curve 1 was obtained by fitting a single exponential plus 1.3 nC/µF, which is the value of $\frac{q_{B, \text{max}}}{C_m}$ obtained from the $Q-V$ plot at a $V_H$ of $-40$ mV. The time constants for the inactivation of $Q_B$ and $Q_s$ at $-40$ mV are 25 and $3.4$ s, respectively, in this fiber, confirming the above observation that $Q_s$ is inactivated faster than $Q_B$.

Similar experiments were performed on four other fibers and the inactivation time constant of $Q_B$ is longer than that of $Q_s$ in all fibers. In one of the four fibers, the inactivation time constants of $Q_B$ and $Q_s$ are 59 and $5.3$ s at $-40$ mV. Averaging between the two fibers, the inactivation time constants of $Q_B$ and $Q_s$ at $-40$ mV are $42$ and $4.4$ s. From two of the other fibers, the average values for the inactivation time constants of $Q_B$ and $Q_s$ at $-30$ mV are $29$ and $5.0$ s. The inactivation time constants of $Q_B$ and $Q_s$ at $-50$ mV were studied in only one fiber and the values are 45 and $9.7$ s. These values vaguely suggest a voltage-dependent decrease in inactivation time constant for either $Q_B$ or $Q_s$ when $V_H$ becomes less negative. However, more experiments will be required to establish the detailed voltage dependencies of the inactivation time constants.

![Figure 9. Effects of the duration of conditioning prepulse on steady-state voltage distributions of $Q_B$ and $Q_s$. Same fiber as in Fig. 8. O’s show data taken with TEST pulses alone, and △'s, ▾’s, and ▼’s with TEST pulses preceded by a conditioning prepulse to $-40$ mV for 1, 5, and 20 s, respectively. The points were obtained from time integrals of OFF current transients in TEST-minus-CONTROL traces, some of which are shown in Fig. 8. Curves 1, 2, and 3 were obtained by fitting Eq. 1, with gap charge correction and gap correction, to the three data sets. Curve 4 was obtained by fitting a single Boltzmann distribution function, with CONTROL charge correction and gap correction, to the other data set. The best fit parameters are:

| Curve | $q_{B, \text{max}}/C_m$ | $V_B$ | $k_B$ | $q_{s, \text{max}}/C_m$ | $V_s$ | $k_s$ |
|-------|----------------|--------|-------|----------------|--------|-------|
| 1     | 9.0           | $-32.4$| 12.8  | 12.6           | $-53.8$| 3.3   |
| 2     | 9.1           | $-35.1$| 10.1  | 9.2            | $-54.4$| 2.6   |
| 3     | 7.8           | $-51.8$| 8.6   | 4.1            | $-60.2$| 2.6   |
| 4     | 4.8           | $-62.9$| 7.6   |                |        |       |
Effect of Multiple Stimulation on $Q_\gamma$ in the Presence of 20 mM Internal EGTA

The last group of experiments was performed to investigate whether the $I_\gamma$ hump indeed disappears during repetitive stimulation. Several experiments were first carried out with 20 mM EGTA in the internal solution, the same as in most of our cut fiber experiments. In these experiments each CONTROL run was accompanied by two TEST runs separated by 6 s. The same CONTROL current trace was scaled and subtracted from the first and second TEST current traces to generate the TEST-minus-CONTROL current traces shown in the same row of Fig. 11, A and B, respectively. The $I_\gamma$ humps present in the first TEST runs at -50 and -45 mV disappear in the second TEST runs, whereas those at -40 and -35 mV are suppressed in the second TEST runs. Those at -30 and -20 mV appear to be affected very little.

To examine the change in shape of the $I_\gamma$ humps more closely, each trace in Fig. 11 B was subtracted from the corresponding trace in Fig. 11 A. The difference traces are shown in Fig. 11 C. The amplitude of the OFF transient increases from the first difference trace to the second and then decreases in the lower difference traces. More surprisingly, except for the first one, every difference trace seems to have more ON charge than OFF charge. These peculiarities can be explained by the analyses shown in the next two figures.

The third trace in Fig. 11 C appears to have the largest inequality between ON and OFF charge and is selected to illustrate the point. The trace is shown in expanded scale as the lower trace in Fig. 12. Line 1 was obtained by least-squares fitting a sloping straight line to the last 200 ms in the ON segment and corresponded to a
Figure 11. Effect of repeated stimulations on TEST-minus-CONTROL current in a cut fiber. Fiber identification: 03142. Diameter = 96 μm. Sarcomere length = 3.5 μm. Saponin treatment was applied to membrane segments in both end pools at time zero. After rinsing, the solutions in the end pools were replaced by solution B. The solution in the center pool was then changed to an isotonic TEA-Cl solution (solution D). At the 20th minute the voltage clamp was turned on and the holding potential was set at −90 mV. From the beginning to the end of the experiment the holding current changed from −14 to −20 nA and \( \tau_e/(r_e + r_i) \) decreased from 0.993 to 0.991. For each TEST potential shown at the right in millivolts, a CONTROL current trace was taken first and then two TEST current traces were elicited by identical TEST pulses applied 6 s apart. The fiber was allowed to rest for 5 min before another group of CONTROL and TEST current traces at another potential was taken. The TEST-minus-CONTROL current traces obtained by subtracting the CONTROL current trace from the first and second TEST current traces are shown in A and B, respectively. The difference of two traces in the same row of A and B is shown in the corresponding row of C. The amounts of OFF charge in the difference traces are (from top to bottom): 1.8, 4.9, 2.7, 1.0, 0.3, −0.1, and 1.7 nC/μF. A negative value implies that the amount of OFF charge in the second TEST run is larger than that in the first TEST run at the same potential. Traces were taken from the 55th to the 165th minute. Only representative traces are shown in each panel.
baseline one would pick visually. With line 1 as baseline, the ON charge is 9.8 nC/μF, which is much larger than the 2.7 nC/μF of OFF charge listed in the legend of Fig. 11. On the other hand, if the ionic current did not change from the first TEST run to the second TEST run, then the time axis, represented by the thin line numbered 2, should be the baseline for the ON segment. With this baseline, the ON charge is −4.9 nC/μF, indicating that the assumption that the ionic current did not change was incorrect.

Unfortunately, there is no objective approach to determine exactly by how much the ionic current had changed in the second TEST run. However, if line 2 is artificially shifted downward by 0.19 nA, the new baseline represented by line 3 gives an ON charge that matches the OFF charge exactly. A small shift of 0.19 nA in the maintained current is quite probable and line 3 actually fits the points at the end of the interval very well. If line 3 is indeed the baseline for the ON segment, then the difference current in this segment is biphasic. This could explain the presence of a negative phase in difference charge movement traces observed by Csernoch et al. (1989). This appearance has been observed on other occasions (Hui, 1991b; Hui and Chen, 1992). A likely explanation for the appearance is that the $I_v$ hump became very broad in the second TEST run and that the slow positive tail of the $I_v$ component (marked by the arrow in Fig. 12A) gave rise to the slow negative phase in the difference trace. This example shows that, in this fiber, even a pulse of 400 ms duration to −45 mV was not sufficiently long to permit a reliable fit of the ON baseline. One should therefore be particularly cautious in drawing information from ON segments of difference traces at potentials just above the activation threshold of $Q_v$. 

**Figure 12.** Change in time course of the $I_v$ hump during repetitive stimulation. Same fiber as in Fig. 11. The upper pair of traces is replotted from the third traces in Fig. 11, A and B, and superimposed on each other. The first 9 points in the ON transients and the first 15 points in the OFF transients are off scale. The lower trace is replotted from the third trace in Fig. 11 C. Line 1 was fitted by least squares to the last 200 ms of the ON segment. Line 2 is the time axis corresponding to zero current. Line 3 was obtained by shifting line 2 downward by 0.19 nA such that the net ON charge matches the OFF charge.
The amounts of OFF charge in the traces of Fig. 11, A and B, and others not shown, are plotted against TEST pulse potential in Fig. 13. Filled diamonds and open squares represent, respectively, the amounts of charge in the first and second TEST runs. From here on, the Q-V plot based on the ith TEST runs will be called the ith Q-V plot. The two Q-V plots in Fig. 13 appear to have similar shapes and magnitudes, but the second Q-V plot is displaced by a few millivolts to the right along the voltage axis, similar to the effect of 25 μM tetracaine (Fig. 3 in Hui and Chen, 1992). Curves 1 and 2 were fitted to the two data sets, according to Eq. 1, with gap correction. The best fit parameters listed in the figure legend show that \( V_i \) in curve 2 is 5.3 mV less negative than that in curve 1, consistent with the shift observed visually. \( q_{\text{v, max}}/e_m \) turns out to be 4.8 nC/μF smaller in curve 2 than in curve 1, but \( q_{\text{g, max}}/e_m \) is 4.8 nC/μF larger. This could be a real effect of double stimulation. Alternatively, part of the change could be due to scatter of data, as explained in the Discussion of the preceding paper (Hui and Chen, 1992). If the latter is true, the mere effect of double stimulation is a shift of the Q-V plot slightly to the right. The shift in the Q-V plot explains why the OFF charge is largest at −50 mV in the difference traces of Fig. 11 C; it is because that particular potential falls on the steepest part of the Q-V curve.

**Figure 13.** Effects of repeated stimulations on steady-state voltage distributions of \( Q_{\text{b}} \) and \( Q_v \). Same fiber as in Fig. 11. ◆'s and □'s were obtained from time integrals of OFF current transients in TEST-minus-CONTROL traces in Fig. 11, A and B, respectively, and other traces not shown. Curves 1 and 2 were obtained by fitting Eq. 1, with gap correction, to the two data sets. The best fit parameters are:

| Curve | \( q_{\text{b, max}}/e_m \) | \( V_{\text{b}} \) | \( k_{\text{b}} \) | \( q_{\text{v, max}}/e_m \) | \( V_v \) | \( k_v \) |
|-------|-----------------|--------|------|-----------------|--------|------|
| 1     | 10.8            | −29.0  | 7.3  | 17.9            | −53.1  | 3.0  |
| 2     | 15.6            | −33.7  | 12.8 | 13.6            | −47.8  | 2.2  |
Experiments of the same kind were performed on three other fibers. The average changes in the six Boltzmann parameters from the first $Q-V$ plot to the second are listed in the first row of Table II. The only statistically significant change was the increase in $V_v$. Thus, the changes in $q_{\beta, \text{max}}/C_m$ and $q_{\gamma, \text{max}}/C_m$ observed in Fig. 13 were probably due to scatter of data.

**Effect of Increasing [EGTA], from 20 to 50 mM on Charge Movement**

If $Q_v$ is a consequence of Ca release, the 20 mM EGTA in the internal solution might not be high enough to effectively deplete the SR of releasable Ca or to effectively prohibit the Ca$^{2+}$ released from the SR to reach the tubular membrane, thereby resulting in very little change in $Q_v$ during repetitive stimulation. To definitively rule out this possibility, additional experiments were carried out with all the glutamate in the internal solution replaced isosmotically by EGTA (solution C) and the [EGTA]$_i$ increased to 50 mM. Before the study of repetitive stimulation with this internal solution, experiments were carried out to check whether the increase in [EGTA]$_i$ had any direct effect on charge movement. Results from eight experiments are summarized in Table I. The results show that increasing [EGTA]$_i$ from 20 to 50 mM has minimal effect on the amounts or voltage distributions of $Q_\beta$ or $Q_v$. 50 mM EGTA did appear to increase the value of $k_v$ somewhat and make the $I_v$ hump less prominent, probably due to a broadening of the waveform of the hump.

### Table I

Comparison of $Q-V$ Distributions of $Q_\beta$ and $Q_v$ with 20 and 50 mM EGTA in the End-Pool Solution

|       | (1) $V_\beta$ | (2) $k_\beta$ | (3) $q_{\beta, \text{max}}/C_m$ | (4) $V_v$ | (5) $k_v$ | (6) $q_{\gamma, \text{max}}/C_m$ |
|-------|---------------|---------------|-----------------|-----------|-----------|-----------------|
| 20 mM EGTA |               |               |                 |           |           |                 |
| Mean  | -41.7         | 11.2          | 10.5            | -58.9     | 2.6       | 13.1            |
| SEM   | 3.4           | 1.9           | 1.0             | 1.9       | 0.3       | 1.0             |
| 50 mM EGTA |               |               |                 |           |           |                 |
| Mean  | -38.5         | 9.9           | 12.4            | -61.8     | 3.5       | 12.4            |
| SEM   | 3.2           | 0.9           | 1.4             | 1.2       | 0.3       | 1.3             |
| $P$   | >0.5          | >0.5          | >0.2            | >0.2      | <0.05     | >0.5            |

Results were collected from eight fibers. Columns 1–6 give the mean values and the SEMs of the best fit parameters obtained by fitting $Q-V$ plots with a sum of two Boltzmann distribution functions, with CONTROL charge correction and gap correction. The last row gives the significance of the differences between the two sets of values, with the two-tailed $t$ test.

the internal solution replaced isosmotically by EGTA (solution C) and the [EGTA]$_i$ increased to 50 mM. Before the study of repetitive stimulation with this internal solution, experiments were carried out to check whether the increase in [EGTA]$_i$ had any direct effect on charge movement. Results from eight experiments are summarized in Table I. The results show that increasing [EGTA]$_i$ from 20 to 50 mM has minimal effect on the amounts or voltage distributions of $Q_\beta$ or $Q_v$. 50 mM EGTA did appear to increase the value of $k_v$ somewhat and make the $I_v$ hump less prominent, probably due to a broadening of the waveform of the hump.

**Effect of Multiple Stimulation on $Q_v$ in the Presence of 50 mM Internal EGTA**

If $Q_v$ is a consequence of Ca release, a train of only two stimulations (Fig. 11) might not be sufficient to effectively deplete the SR of releasable Ca. In the following group
of experiments carried out with 50 mM internal EGTA, the number of stimulations at each TEST potential was increased to 3, 5, or 10. Results from one of the experiments are shown in Figs. 14–16.

In this experiment, a train of 10 TEST pulses was applied at each potential at a frequency of one per 6 s. Again, the same CONTROL current trace was used to subtract the linear currents from all 10 TEST current traces. Since 15 min of recovery time was allowed between each train of stimulations, charge movement was studied at fewer TEST potentials than with the usual one- or two-pulse protocol. Fig. 14 A shows TEST-minus-CONTROL current traces obtained from the first TEST runs. The traces resemble those recorded from fibers with 20 mM EGTA in the internal solution. The traces obtained from the second TEST runs (Fig. 14 B) still show very prominent I\textsubscript{v} humps. The changes in the shapes of the I\textsubscript{v} humps can be visualized more easily by taking pairwise differences of the traces in Fig. 14, A and B, as shown in Fig. 15 A. The amount of OFF charge in all the difference traces was < 1 nC/\mu F (see Fig. 15 legend). The biphasic ON transients can be explained by a broadening of the I\textsubscript{v} waveform (see text associated with Fig. 12).

The traces in Fig. 14 C were obtained from the third TEST runs. They look exactly the same as the traces from the second TEST runs. In fact, the difference traces between the second and third TEST runs (Fig. 15 B) are essentially flat. The traces obtained from all subsequent TEST runs are all identical to the corresponding traces from the third TEST run. Only the traces obtained from the tenth TEST runs are shown in Fig. 14 D. The difference traces obtained by subtracting the traces in Fig. 14 D from the corresponding traces in Fig. 14 C are also flat (not shown). The persistence of the I\textsubscript{v} humps even in the tenth stimulation is in contrast to the finding of Garcia et al. (1990), who observed a disappearance of the hump during repetitive stimulation. The 62.5 mM EGTA they used was slightly higher than the 50 mM used in this experiment. However, the fiber in this experiment was stimulated at a much higher frequency.

The amounts of OFF charge in the traces from the first, second, and tenth TEST runs (Fig. 14, A, B, and D, and others not shown) are plotted against TEST pulse potential in Fig. 16. Those from the third TEST runs (Fig. 14 C) are not shown to avoid overlap. The three Q-V plots are not very different from each other. Curves 1–3 were fitted to the three data sets, according to Eq. 1, with gap correction. The best fit parameters are listed in the figure legend. Compared with the values in curve 1, \( q_{\beta,\text{max}}/e_m \) in curve 2 is decreased to 85.9\%, \( \bar{V}_\beta \) is shifted by \(-0.6\) mV, \( k_\beta \) is decreased by \(1.1\) mV, \( q_{\gamma,\text{max}}/e_m \) is increased to 103.9\%, \( \bar{V}_\gamma \) is shifted by \(+1.1\) mV, and \( k_\gamma \) is increased by \(0.7\) mV. Also, \( q_{\beta,\text{max}}/e_m \) in curve 3 is decreased to 76.9\%, \( \bar{V}_\beta \) is shifted by \(+1.1\) mV, \( k_\beta \) is decreased by \(1.7\) mV, \( q_{\gamma,\text{max}}/e_m \) is increased to 101.0\%, \( \bar{V}_\gamma \) is shifted by \(+2.1\) mV, and \( k_\gamma \) is decreased by \(0.2\) mV.

Similar experiments were performed on six other fibers. In two of the experiments, trains of 10 TEST pulses were applied at each potential as in the experiment just shown, whereas in the other four experiments only trains of three or five TEST pulses were applied. The average changes in the Boltzmann parameters for the second, third, and tenth Q-V plots are listed in the second, third, and fourth rows of Table II. All the changes are statistically insignificant with the two-tailed t test, except for the
FIGURE 14. Effect of multiple stimulations on TEST-minus-CONTROL current in a cut fiber with 50 mM EGTA in the internal solution. Fiber identification: 04261. Diameter = 87 μm. Sarcomere length = 3.5 μm. Saponin treatment was applied to membrane segments in both end pools at time zero. After rinsing, the solutions in the end pools were replaced by solution C containing 50 mM EGTA. Then the solution in the center pool was changed to an isotonic TEA-Cl solution (solution D). At the 22nd minute the voltage clamp was turned on and the holding potential was set at -90 mV. From the beginning to the end of the experiment the holding current changed from -14 to -18 nA and $r_e/(r_e + r_i)$ remained constant at 0.992. For each TEST potential shown at the right in millivolts, a CONTROL current trace was taken first and then 10 TEST current traces were elicited by a train of identical TEST pulses at a rate of one per 6 s. The fiber was allowed to rest for 15 min before another group of CONTROL and TEST current traces at another potential was taken. The TEST-minus-CONTROL current traces obtained by subtracting the CONTROL current trace from the 1st, 2nd, 3rd, and 10th TEST current traces are shown in A, B, C, and D, respectively, in the same row. Traces were taken from the 60th to the 256th minute. Only representative traces are shown in each panel.
decrease in $q_{h,max}/e_m$ in the second $Q-V$ plots and the shift in $V_f$ in the third $Q-V$ plots, but even these two changes are marginally significant.

Although a train of $\leq 10$ stimulations had a negligible effect on $Q_m$, one could argue that the stimulation rate of once per 6 s could be too slow to bring about a noticeable effect. Four experiments were done to test this idea by making use of the double-pulse protocol, in which each pair of TEST pulses was applied with a 400-ms time gap in between. Interestingly, the $I_v$ hump in the ON segment of the second pulse was still almost as prominent as that of the first. Moreover, a steeply rising component still existed in the second $Q-V$ plot and was similar to that in the first $Q-V$ plot.

**Figure 15.** Difference current traces between two successive stimulations from a cut fiber with 50 mM EGTA in the internal solution. Same fiber as in Fig. 14. Each trace was obtained from the difference of the two corresponding traces in Fig. 14, A and B (A) or in Fig. 14, B and C (B). The amounts of OFF charge in the difference traces in A are (from top to bottom): 0.3, 0.6, 0.4, 0.0, 0.0, 0.1, and $-0.1$ nC/$\mu$F. A negative value implies that the amount of OFF charge in the second TEST run is larger than that in the first TEST run at the same potential. The amounts of OFF charge in the difference traces in B are all negligible.

**DISCUSSION**

*Steady-State Inactivation of $Q_h$ and $Q_v$ by Maintained Depolarization*

Steady-state inactivation of charge movement was studied in intact fibers by several investigators (Adrian and Almers, 1976; Rakowski, 1981; Hui, 1983b; Rakowski et al., 1985). In most of the studies (except Hui, 1983b) the amounts of charge were not separated into $Q_h$ and $Q_v$ components and so only inactivation curves of the total charge were obtained. It was found that the inactivation curve of the total charge was roughly a mirror image of the activation curve.
In this paper the inactivation of charge movement was studied in cut fibers and the differential inactivation properties of $Q_\beta$ and $Q_\gamma$ were separated. Fig. 7 shows that the inactivation of $Q_\beta$ is less voltage dependent than that of $Q_\gamma$, in good agreement with the results obtained from intact fibers (Hui, 1983b). The curves in the figure were fitted by assuming that the voltage dependence of inactivation of each charge component follows an inverse sigmoidal function. Under this assumption, the $k$ factors for the activation and inactivation curves of $Q_\beta$ are roughly the same, $\sim 11-13$ mV, and likewise for the curves of $Q_\gamma$, $\sim 3$ mV. This implies that, for each charge species, the activation and inactivation curves are mirror images of each other.

A consequence of the shallow inactivation curve of $Q_\beta$ is that it crosses the inactivation curve of $Q_\gamma$ at a potential $>-70$ mV. Thus, at $-70$ mV a larger proportion of $Q_\beta$ is inactivated than $Q_\gamma$. This implies that it is unlikely to have all $Q_\beta$ serve as voltage sensors for triggering Ca release and all $Q_\gamma$ arise as a result of the release, because it is difficult to imagine how a larger reduction in the amount of moveable sensors can lead to a smaller reduction in the quantity of end products. The shallow inactivation curve of $Q_\beta$ can be explained by a possible multi-component nature of $Q_\beta$ (see Discussion sections in Chen and Hui, 1991a; Hui, 1991b). Each component might have a different inactivation curve and some of the curves can be more steeply voltage dependent than the others. A summation of all the curves with a spread in the values of $V^*$ and $k^*$ can yield a shallow resultant inactivation curve.
Another consequence of the shallow inactivation curve of $Q_\beta$ is that the amount of moveable $Q_\beta$ does not reach full value at a $V_H$ of $-90$ mV. The curve predicts that when $V_H$ is made more negative $\frac{q_{\beta,max}}{C_m}$ should increase, which has indeed been observed by Hui (1991b).

Adrian and Peres (1979) separated $Q_\beta$ and $Q_\gamma$ by assuming that at $-40$ mV $Q_\gamma$ is completely inactivated, whereas $Q_\beta$ is fully mobile. Fig. 7 shows that $Q_\gamma$ is completely inactivated at $-40$ mV in our cut fibers, but only a small fraction of $Q_\beta$ remains mobile. At more negative levels of $V_H$, more $Q_\beta$ remains mobile but then $Q_\gamma$ is not completely inactivated. There is no $V_H$ at which the separation method of Adrian and Peres (1979) can yield $Q_\beta$ and $Q_\gamma$ components that are equivalent to those separated by any of the four methods used in the preceding paper (Hui and Chen, 1992).

### Table I

| i | [EGTA] | $s$ | $mV$ | $\Delta V_\beta$ | $\Delta V_\gamma$ | $\frac{q_{\gamma,max}}{C_m}$ ratio | $\frac{q_{\beta,max}}{C_m}$ ratio | $\Delta k_\beta$ | $\Delta k_\gamma$ | $n$ |
|---|---|---|---|---|---|---|---|---|---|---|
| 2 | 20 | 6 | 3.8 | 3.8 | 108 | 4.1 | 0.9 | 104 | 4 | 4 |
|   |   |   | ($>0.3$) | ($>0.2$) | ($>0.5$) | ($<0.02$) | ($>0.5$) | ($>0.8$) |   |   |
| 2 | 50 | 6 | 5.3 | 0.8 | 79 | 3.5 | 1.2 | 118 | 7 | 7 |
|   |   |   | ($>0.1$) | ($>0.5$) | ($<0.05$) | ($>0.05$) | ($>0.05$) | ($>0.5$) |   |   |
| 3 | 50 | 6 | 6.1 | 1.5 | 90 | 4.3 | 1.2 | 114 | 7 | 7 |
|   |   |   | ($>0.1$) | ($>0.5$) | ($<0.05$) | ($>0.05$) | ($>0.05$) | ($>0.5$) |   |   |
| 10 | 50 | 6 | 2.7 | 0.9 | 85 | 4.7 | 0.8 | 102 | 3 | 3 |
|   |   |   | ($>0.3$) | ($>0.5$) | ($>0.4$) | ($>0.3$) | ($>0.2$) | ($>0.8$) |   |   |

Two groups of experiments were performed. In the first group listed in the first row, pairs of two identical TEST pulses were applied at different potentials at a rate of one pair per 5 min. The second group listed in the second to fourth rows was performed on seven fibers. In three of the fibers, trains of 10 TEST pulses were applied at a rate of one train per 10 or 15 min. In the remaining four fibers, trains of three or five identical TEST pulses were applied at the same rate. Columns 1 and 2 give the values of $i$ for the $i$th $Q-V$ plot (see definition in text) and the concentrations of EGTA. Column 3 gives the time separation between successive TEST pulses in a pair or a train. Columns 4–9 give the average changes in the values of the Boltzmann parameters from the first $Q-V$ plot to the $i$th $Q-V$ plot. $\Delta V_\beta$ or $\Delta V_\gamma$ were calculated by subtracting the value in the first $Q-V$ plot from the corresponding value in the $i$th $Q-V$ plot. $\frac{q_{\gamma,max}}{C_m}$ ratio was calculated by dividing the value in the $i$th $Q-V$ plot by the corresponding value in the first $Q-V$ plot. The statistical significance of each change, estimated with the two-tailed $t$ test, is shown in parenthesis below each value. Column 10 gives the number of fibers included in the average.

### Inactivation Time Courses of $Q_\beta$ and $Q_\gamma$

Chandler et al. (1976) found that the total charge was inactivated with a time constant of 10–25 s at $-20$ mV and 1°C. Adrian and Almers (1976) found that the half-time for inactivation of the total charge was $\sim 2$ min at $-50$ mV and 2–6°C. Rakowski (1981) suggested a voltage dependence of the inactivation time constant for the total charge. These results were obtained from intact fibers. If the charge movement they measured contained $Q_\gamma$, the time constants they obtained should be lumped values for both $Q_\beta$ and $Q_\gamma$. In this work, $Q_\beta$ and $Q_\gamma$ were separated in cut fibers by method 3 and were found to follow different time courses in inactivation. At
-50 to -30 mV the inactivation time constant of \( Q_\beta \) is 5-10-fold as large as that of \( Q_r \). There is also a suggestion of both time constants becoming smaller at a less negative potential.

**Comparison with Inactivation of Tension Generation or Ca Indicator Signal**

To gain insight into the possible physiological role(s) of \( Q_\beta \) and \( Q_r \) in excitation-contraction coupling, it is of interest to compare the voltage dependencies of inactivation of \( Q_\beta \) and \( Q_r \) with those of tension and rise in [Ca\(^{2+}\)]. Inactivation of K contracture tension was found to be steeply voltage dependent and the inactivation curve was almost a mirror image of the activation curve (Hodgkin and Horowicz, 1960; Luttgau and Glitsch, 1976). Although the \( k \) factor of the inactivation curve was not determined, contracture tension inactivated almost fully over a 20-mV range of potential, which correlates much better with the inactivation curve of \( Q_r \) in Fig. 7 than that of \( Q_\beta \).

The time course of inactivation of Ca release has not yet been studied. This should be an important experiment providing useful information to support the role(s) of \( Q_\beta \) and \( Q_r \).

**Effect of Repetitive Stimulation on \( I_r \) Hump**

The results presented in this paper do not agree with the conclusion drawn by Garcia et al. (1990). With the experimental protocol used in this work, pronounced \( I_r \) humps could still be observed in TEST-minus-CONTROL current traces, and a substantial \( Q_r \) component could still be resolved in \( Q-V \) plots after repetitive stimulation. The \( I_r \) hump they recorded at -20 mV resembles the hump we obtained at \(-45 \) mV (Fig. 12). These potentials fall on the steeply rising parts of the respective \( Q-V \) plots. The disappearance of the \( I_r \) hump they observed could be caused merely by a shift of the \( Q_r-V \) plot to the right. Fig. 12 also shows that the difference current trace between the first and second TEST runs at such a potential might erroneously yield an apparent inequality between ON and OFF charge if the ON baseline is not fitted properly.

The exact cause for the voltage shift of the \( Q_r-V \) plot is not known. One possibility is that, on repolarization, \( Q_r \) requires some finite time to be restored to the normal resting state. Thus, a few seconds after repolarization, \( Q_r \) could be in a temporary off state that is different from the normal resting state. The voltage distribution of \( Q_r \) in that off state could follow a Boltzmann distribution shifted by a few millivolts from that for the normal resting state. Another possibility is that \( Q_r \) is made up of two components, the primary one being activated by the depolarizing pulses and the secondary one activated by the additional depolarization caused by the binding of the released Ca\(^{2+}\) on the myoplasmic face of the tubular membrane. In other words, the primary component is consistent with the trigger hypothesis and the secondary component is consistent with the feedback hypothesis. With the secondary compo-
When Ca release is suppressed, for example by repetitive stimulation, a small shift of $V_v$ to a less negative potential can be observed.

If $Q_v$ is completely caused by Ca release, then some features of our results are difficult to explain. For example, depletion of Ca$^{2+}$ in the SR should be more effective with 50 mM EGTA than with 20 mM in the end pool solution, but the shifts in the $Q-V$ plots with both concentrations of EGTA are comparable (Table II). The only sensible conclusion that can accommodate all the results is that there is a genuine, steeply voltage-dependent component of charge that has been convincingly dissected out from the total charge with four independent methods (Hui and Chen, 1992). Part of this component can be activated by Ca feedback, but that part is considered as secondary. Under normal conditions $I_v$ is manifested as a hump in some potential range, but its waveform can be broadened when the fiber is stimulated repeatedly at a relatively high rate. Although $Q_v$, serving as a voltage sensor, is not suppressed during repetitive stimulation, Ca release can be substantially reduced due to a depletion of releasable Ca in the SR.

This project was supported by grants from the National Institutes of Health (NS-21955) and the Muscular Dystrophy Association. C. S. Hui was a recipient of a Research Career Development Award (NS-00976) from the NIH and W. Chen was a recipient of a postdoctoral fellowship from the Indiana Heart Association.

Original version received 29 April 1991 and accepted version received 23 January 1992.

REFERENCES

Adrian, R. H., and W. Almers. 1976. Charge movement in the membrane of striated muscle. *Journal of Physiology*. 254:339–360.

Adrian, R. H., W. K. Chandler, and R. F. Rakowski. 1976. Charge movement and mechanical repriming in skeletal muscle. *Journal of Physiology*. 254:361–388.

Adrian, R. H., and A. R. Peres. 1979. Charge movement and membrane capacity in frog muscle. *Journal of Physiology*. 289:83–97.

Almers, W. 1978. Gating currents and charge movements in excitable membranes. *Review of Physiology and Biochemical Pharmacology*. 82:96–190.

Brum, G., and E. Eios. 1987. Intramembrane charge movement in frog skeletal muscle fibres: properties of charge 2. *Journal of Physiology*. 387:489–517.

Chandler, W. K., and C. S. Hui. 1990. Membrane capacitance in frog cut twitch fibers mounted in a double Vaseline-gap chamber. *Journal of General Physiology*. 96:225–256.

Chandler, W. K., R. F. Rakowski, and M. F. Schneider. 1976. A nonlinear voltage dependent charge movement in frog skeletal muscle. *Journal of Physiology*. 254:245–283.

Chen, W., and C. S. Hui. 1989. Effects of tetracaine and maintained depolarization on charge movement components in frog cut twitch fibers. *Biophysical Journal*. 55:239a. (Abstr.)

Chen, W., and C. S. Hui. 1991a. Existence of $Q_v$ in frog cut twitch fibers with little $Q_p$. *Biophysical Journal*. 59:503–507.

Chen, W., and C. S. Hui. 1991b. Differential blockage of charge movement components in frog cut twitch fibers by nifedipine. *Journal of Physiology*. 444:579–603.

Csernoch, L., I. Uribe, M. Rodriguez, G. Pizzaro, and E. Rios. 1989. $Q_v$ and Ca release flux in skeletal muscle fibers. *Biophysical Journal*. 55:88a. (Abstr.)
Garcia, J., G. Pizarro, E. Rios, and E. Stefani. 1990. Depletion of the SR reduces the delayed charge movement of frog skeletal muscle. Biophysical Journal. 57:341a. (Abstr.)

Hodgkin, A. L., and P. Horowicz. 1960. Potassium contractures in single muscle fibres. Journal of Physiology. 153:386–403.

Horowicz, P., and M. F. Schneider. 1981. Membrane charge movement in contracting and non-contracting skeletal muscle fibres. Journal of Physiology. 314:565–595.

Huang, C. L.-H. 1982. Pharmacological separation of charge movement components in frog skeletal muscle. Journal of Physiology. 324:375–387.

Hui, C. S. 1983a. Pharmacological studies of charge movement in frog skeletal muscle. Journal of Physiology. 337:509–529.

Hui, C. S. 1983b. Differential properties of two charge components in frog skeletal muscle. Journal of Physiology. 337:531–552.

Hui, C. S. 1990. D600 binding sites on voltage-sensors for excitation-contraction coupling in frog skeletal muscle are intracellular. Journal of Muscle Research and Cell Motility. 11:471–488.

Hui, C. S. 1991a. Comparison of charge movement components in intact and cut twitch fibers of the frog. Effects of stretch and temperature. Journal of General Physiology. 98:287–314.

Hui, C. S. 1991b. Factors affecting the appearance of the slow charge component in frog cut twitch fibers. Journal of General Physiology. 98:315–347.

Hui, C. S., and W. K. Chandler. 1990. Intramembranous charge movement in frog cut twitch fibers mounted in a double Vaseline-gap chamber. Journal of General Physiology. 96:257–297.

Hui, C. S., and W. K. Chandler. 1991. Comparison of $Q_\alpha$ and $Q_\gamma$ charge movement in frog cut twitch fibers. Journal of General Physiology. 98:429–464.

Hui, C. S., and W. Chen. 1992. Separation of charge movement components in frog cut twitch fibers with tetracaine. Critical comparison with other methods. Journal of General Physiology. 99:985–1016.

Hui, C. S., and R. L. Milton. 1987. Suppression of charge movement in frog skeletal muscle by D600. Journal of Muscle Research and Cell Motility. 8:195–208.

Luttgau, H. C., and H. G. Glitsch. 1976. Membrane physiology of nerve and muscle fibres. Fortschrritte der Zoologie. 24:1–132.

Rakowski, R. F. 1981. Immobilization of membrane charge in frog skeletal muscle by prolonged depolarization. Journal of Physiology. 317:129–148.

Rakowski, R. F., P. M. Best, and M. R. James-Kracke. 1985. Voltage dependence of membrane charge movement and calcium release in frog skeletal muscle fibres. Journal of Muscle Research and Cell Motility. 6:403–433.

Vergara, J., and C. Caputo. 1983. Effects of tetracaine on charge movements and calcium signals in frog skeletal muscle fibers. Proceedings of the National Academy of Sciences, USA. 80:1477–1481.