Clinical and Clinicopathological Findings of Arthritic Camel Calf Associated with Mycoplasma Infection (Camelus Dromedarius)

Abstract

Objective: The present study aimed to investigate the clinical and clinicopathological findings of arthritic camel calf associated with mycoplasma infection in district areas of Saudi Arabia.

Methods: Forty-one camel calves from different farms with different sex and age from 1-12 months were used in this study. Thirteen camels did not have any clinical articular abnormalities while twenty eight camels had gross articular problems such as lameness and swollen in joints either monoarthritis or polyarthritis. The synovial fluid was extracted from the arthritic joints. Then, the concentration of TLC, RBCs, and TP were measured in samples. The mycoplasma isolates which were identified were further confirmed by disk growth inhibition test using a panel of specific antiserum against selected reference mycoplasma spp.

Results: Concentration of all measured parameters in arthritic joints were significantly higher than clinically healthy joints (p<0.05). The synovial fluid concentration of TLC, RBCs, and TP were 9525±526 cells/µl, 4804.4±91 cells/µl, and 2.820±104 g/dl in arthritic joints respectively. The most commonly isolated bacteria were Mycoplasma spp., followed by non-haemolytic streptococci spp. and Staphylococcus aureus.

Conclusion: This study gives us a spotlight on the significance of mycoplasma arthritis in camel calves and significant increase of acute phase proteins and inflammatory cells in the synovial fluid. Information about the normal values of these parameters and their changing patterns may help camel rearing systems during arthritis by assessing the health status of joints in the camels; in addition, the information about normal values can be diagnostically valuable when considering diseased animals.

Keywords: Camel calf; Arthritic joints; Mycoplasma spp.; Synovial fluid; Clinical study

Abbreviations: TLC: Total Leukocytic Count; RBCs: Red Blood Cells; TP: Total Protein; M. bovis: Mycoplasma bovis; WBC: White Blood Cell; TNCC: Total Nucleated Cell Count; SD: Standard Deviation; FPT: Failure of Passive Transfer; AST: Aspartate Aminotransferase; LDH: Lactate Dehydrogenase

Introduction

Camel is an economically important farm animal used for meat, milk and hair production in the Middle East and North Africa [1]. Mycoplasmas are highly contagious organism capable of auto-replication and difficult to culture and slow growing [2,3]. Camel joints are infected by variety of infectious diseases that may affect their racing performance. Mycoplasma spp. are considered the common causes of some diseases as arthritis, pneumonia and abortion [4]. Camel calves are more susceptible to mycoplasma infection and developing clinical signs [5]. Consequently, the pathogen cannot be detected during the incubation period. Moreover, the serological cross reactions among the Mycoplasma spp. are a critical problem [6,7]. Synovial fluid analysis is the common method for diagnosis of various joint diseases [8]. The most commonly isolated organisms are Staphylococcus spp., Escherichia coli, Arcanobacterium spp., Corynebacterium and less commonly Mycoplasma spp. [8]. Navel ill is considered the most common term which includes navel abscesses and umbilical hernia [9-11]. Reports on arthritis associated with mycoplasmosis in camels are scare. Consequently, the present study aimed to assess the clinical and clinicopathological findings associated with arthritis caused by mycoplasma spp. in camel calves in district areas in Saudi Arabia.

Materials and Methods

Study area

The present study was conducted in Four areas in central province in Saudi Arabia including (Shaqra, Hermila, Dorm and Al-Qassim region) which are rich in camel population. Each area had been visited two times per month to collect the samples (sy-
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Clinical examination

Competent clinical examination of each camel was done consistent with Radostitis et al. [12]. Data concerned with the case history, clinical findings for each camel under investigation were recorded [12]. A detailed clinical examination of the diseased camels, including examination of joints was carried out.

Synovial fluid collection

Five milliliters of synovial fluid were collected from each joint using a sterile syringe after joints anesthetized, clipped and scrubbed using povidone-iodine solution. The needle was inserted into the medial pouch of the joint. Only blood-free samples were included in the analysis. In cases that blood contamination was suspected based on visual examination, the sample was discarded and a second sampling was attempted at a remote site in the joint. The collected synovial fluid was placed in anti-coagulant-coated tubes and stored at -20°C until use [8].

Synovial fluid analysis

The color, viscosity, presence of flocules and degree of turbidity of aspirated fluid were noted immediately after collection. One direct smear was taken on glass slides and another smear was prepared from resuspended sediment. Slides were stained with Wright’s stain and examined for cell morphology, differential leucocytic count (DLC) and Total nucleated cell counts (TNCCs) using a haemocytometer (L.W. Scientific Inc., Tucker, GA, USA). Total protein (TP) was analyzed using a regular refractometer (Caesar Instrument Co., Taipei, Taiwan).

Serum analysis

Blood was withdrawn from the jugular vein, centrifuged for serum separation and stored at -20°C until use. Glucose, cholesterol, AST and LDH were determined by their commercial reagents in an autoanalyzer (Alcyon 300/300i, Abbott). Serum proteins were determined by using 7.5% acrylamide gel (Laemmli UK, 1970, HSI, 1993).

Isolation of mycoplasma spp.

Few drops of synovial fluid were inoculated into PPLO broth then incubated at 37°C for 24 hour; the incubated broth were plated on PPLO agar plates in humidified candle jar with low oxygen tension. The mycoplasma colonies were examined after 48 hours then daily up to 7-10 days for fried egg colonies [13].

Gross examination of joints

The affected joints were examined for pathological lesions such as thickening of the joint capsule, abnormalities in the synovial membrane and other cartilage deformities.

Bacterial culture

Few drops of synovial fluid were inoculated into colombia blood agar base (Difco, Detroit, MI, USA) enriched with 5% defibrinated sheep blood then cultured on MacConkey agar plates (Fluka, Buchs, Switzerland). The inoculated plates were incubated at 37°C for 48 hours. Growing colonies were picked up, inoculated on nutrient broth and then subcultured on tryptic soy agar (Difco). The bacterial isolates were classified by species according to Barrow & Feltham [14] and Quinn et al. [15]. Samples not showing any bacterial growth after incubation were 10-fold diluted (10⁻¹ to 10⁻⁶) in deionized water 10% (v/v) and cultured in agar and broth mycoplasma experience media (Reigate, Surrey, UK) using a standard procedure [3]. Broths were examined daily and for 1 week for signs of growth opalescence or changes of pH indicated by a color change in the media. Plates were also examined for 7 days under 35X magnifications for typical fried egg appearance.

Statistical analysis

SPSS software (SPSS for Windows, version 11.5, SPSS Inc, Chicago, Illinois) were used for data analyses. The level of significance was set at p<0.05.

Results

Physical examination

41 camel calves with arthritic joint varies from lameness with or without swelling of the joints, the swollen joint either single or multiple swollen joints. Camel calves were lame for a variable period of time. The age of these camel calves was 1-12 months. Lameness was graded on five scales. Scale 1 (normal gait) and scale 5 (severe lameness with arched back) according to Spercher et al. [16]. The joint swelling was assessed as mild, moderate and severe. Fetlock joint considered the most frequently affected joints (6 animals; 14.2%), the tarsal joint (3 animals; 7.3%), the elbow joint (3 animals; 10.7%), the stifle (3 animals; 10.7%), the fetlock and carpal (3 animals; 10.7%), fetlock and tarsal (2 animals; 7.4%), and elbow and carpal (4 animals; 14.2%) (Table 2). The mycoplasma arthritis showed polyarthritus or monarthritus with respiratory signs and lameness (Table 1). The affected joints were mildly to moderately swollen and painful to manipulation. Most of polyarthritus involved wound in joints may be fistulated and oozing some pus. The affected camel calves were feverish, anorexic, runny eye and in some cases were recumbent.

Joints examination

There was mild eburnation of the articular cartilage in 28 joints. The examined joints were inflamed, edematous with thickening of the synovial membrane. The thickening of joint capsule was observed in 12 joints. The most frequent affected joints were elbow and carpal in polyarthritus and fetlock and carpal in monoarthritus (Table 2 & Figure 1).
Table 1: The clinical finding associated with isolated mycoplasma in camel calf.

| Clinical Finding                  | Mycoplasma positive (n= 28) | Mycoplasma negative (n= 13) |
|-----------------------------------|-----------------------------|-----------------------------|
|                                   | Number | %     | Number | %     |
| Anorexic Calves                  | 25     | 89.2  | 8      | 61.5  |
| Fever                            | 19     | 67.8  | 6      | 46.1  |
| Depressed Calves                 | 18     | 64.2  | 5      | 38.4  |
| Cough                            | 19     | 67.8  | 4      | 30.7  |
| Runny Eye                        | 12     | 42.8  | 2      | 15.3  |
| Watery Nose                      | 18     | 64.2  | 6      | 46.1  |
| Monoarthritis                    | 19     | 67.8  | 2      | 0.15  |
| Polyarthritis                    | 9      | 32.1  | 2      | 0.15  |
| Lameness without Swelling in Joints | 0     | 0     | 9      | 69.2  |
| Recumbency                       | 17     | 60.7  | 7      | 53.8  |
| Wounds involved in the Joint     | 23     | 82.1  | 1      | 0.7   |

Figure 1: Mycoplasma infection in arthritic joints in camel calves. (A) showed polyarthritis (elbow and carpal), while (B and C) showed monoarthritis in stifle and fetlock joints respectively.

Table 2: The sites of joints affection associated with mycoplasma infection in arthritic joints in camel calves.

| Signs                  | Monoarthritis | Polyarthritis | Total |
|------------------------|---------------|---------------|-------|
|                        | Fetlock | Carpal | Tarsal | Elbow | Stifle | Fetlock and Carpal | Fetlock and Tarsal | Elbow and Carpal | No of animals | Percentages     |
| No of animals          | 6      | 4      | 3      | 3     | 3      | 3                | 2                | 4                | 28            | 21.40% 14.20% 10.70% 10.70% 10.70% 10.70% 7.40% 14.20% 100% |
| Percentages            | 21.40% | 14.20% | 10.70% | 10.70%| 10.70% | 10.70%           | 7.40%            | 14.20%           | 100%          |
Synovial fluid analysis

The macroscopic findings of collected synovial fluid showed turbidity, blood with occasional floccules. Concentration of synovial fluid parameters in arthritic joints was significantly higher than normal range ($p<0.05$). The TNCC and TP concentrations are presented in Table 3. Observation of altered ALT and LDH and rise in serum TP levels were interpreted as infection. In diseased animals glucose and total cholesterol values were low ($p<0.05$) and urea were high ($p<0.05$). Differences among ALT, creatinine, total protein, albumin were statistically not significant (Table 5).

Table 3: Synovial fluid analysis of arthritic joints in camel calves.

| Parameter          | Mycoplasma Negative (n = 13) | Mycoplasma Positive (n = 28) |
|--------------------|-------------------------------|------------------------------|
| TLC (cells/µl)     | 9525±526                      | 5472±396                     |
| Neutrophils (cells/µl) | 5472±396                     | 1804±1008                    |
| Lymphocytes (cells/µl) | 1896±109                     | 804.4±91                     |
| Mono/Macrophages (cells/µl) | 2.820±1.4                    |                              |

Table 4: Bacterial species isolated from arthritic joints of young camels (n = 41).

| Isolates                   | No. of samples |
|----------------------------|----------------|
| Mycoplasma Spp.            | 28             |
| Staphylococcus aureus      | 1              |
| Non-haemolytic streptococci | 3              |
| No growth                  | 9              |

Bacterial culture

Bacterial culture revealed seven isolates for mycoplasma. In four samples, no bacterial growth was observed as showed in Table 4.

Discussion

Infectious arthritis is considered the common cause of lameness in different animal species including camels. It caused edematous swelling of joints, thickening of the synovial membranes and changes in synovial fluid components. The joints in calves are mainly infected during bacteremia from a remote nidus of infection [17]. In the current study, the affected camel calves with mycoplasma arthritis had monoarthritis and polyarthritis. The fetlock joint was the most commonly affected joint, followed by the carpal in monoarthritic and elbow and carpal in polyarthritis [18,19]. Many of the diseased animals have swellings at the joints (monoarthritis/polyarthritis) which restrict the animal movements and causing respiratory manifestations [20,16]. The cytological and biochemical characteristics of synovial fluid in joints of healthy camel calves have been recently published [8,21]. The differential leucocyte count, TLC and red blood cell counts in synovial fluid of arthritic joints were apparently higher than normal range (Table 3). This is because of the variation in the degree of inflammation in various joints. In healthy camel calves, the TNCC, polymorphonuclear and monocellular leucocytes in the fetlock and tarsal joints were <500 cells/µl and 400 cells/µl respectively [8,21]. In inflamed joints, large molecules such as protein were increased in the synovial fluids due to increases vascular permeability which allowing influx of these molecules in the synovial fluid [17]. Total protein (TP) concentration in cattle with septic arthritis was found to be >4 g/dl [15]. Similarly, TP in affected joints of camels were increased, but to a lesser degree compared with that in cattle. In healthy young camels, TP concentration in the arthritic joints was reported to be 2.8 ± 1.4 g/dl [8,21]. The concentrations of...
TP in affected joints were depending on the degree of inflammation of synovial membrane and on the cause of the condition [22,23].

Bacterial culture is one of the most important clinical tools in the diagnosis of infectious arthritis. The limitation of the usefulness of routine synovial fluid culture is due to localization of bacteria in the synovial membrane, previous antibacterial administration and the inherited antibacterial properties of the synovial fluid [24,25].

Conclusion

In conclusion, this study gives us a spotlight on the significance of mycoplasma arthritis on camel calves and significant increase of acute phase proteins and inflammatory cells in the synovial fluid. Information about the normal values of these parameters and their changing patterns may help camel rearing systems during arthritis by assessing the health status of joints in the camels; in addition, the information about normal values can be diagnostically valuable when considering diseased animals.

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