Hyperspectral Imaging Reveals Spectral Differences and Can Distinguish Malignant Melanoma from Pigmented Basal Cell Carcinomas: A Pilot Study

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Pigmented basal cell carcinomas can be difficult to distinguish from melanocytic tumours. Hyperspectral imaging is a non-invasive imaging technique that measures the reflectance spectra of skin in vivo. The aim of this prospective pilot study was to use a convolutional neural network classifier in hyperspectral images for differential diagnosis between pigmented basal cell carcinomas and melanoma. A total of 26 pigmented lesions (10 pigmented basal cell carcinomas, 12 melanomas in situ, 4 invasive melanomas) were imaged with hyperspectral imaging and excised for histopathological diagnosis. For 2-class classifier (melanocytic tumours vs pigmented basal cell carcinomas) using the majority of the pixels to predict the class of the whole lesion, the results showed a sensitivity of 100% (95% confidence interval 81–100%), specificity of 90% (95% confidence interval 60–98%) and positive predictive value of 94% (95% confidence interval 73–99%). These results indicate that a convolutional neural network classifier can differentiate melanocytic tumours from pigmented basal cell carcinomas in hyperspectral images. Further studies are warranted in order to confirm these preliminary results, using larger samples and multiple tumour types, including all types of melanocytic lesions.

Key words: deep learning; neural network; basal cell carcinoma; malignant melanoma.

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Basal cell carcinoma (BCC) is the most common skin cancer and the most frequently occurring form of all cancers among whites in the Western countries, with an estimated lifetime risk of ≥ 30% (1). The most common subtypes, nodular and superficial BCC, may occasionally contain melanin, referred to as “pigmented BCC”, which constitute ~7% of all BCCs (2). Dermoscopy uses a magnifying lens and polarized light, to assess different structures and colours of a lesion. In dermoscopy the pigment in BCC can be seen as blue-grey ovoid nests and globules, brown to blue-grey maple leaf-like structures, or small blue-grey dots (peppering) (2–4). Heavily pigmented BCCs may contain brown to black globules or dots, or blue-white veil-like structures, and are thus difficult to distinguish from melanocytic lesions (5). The pigmentation may cause confusion in diagnosing these lesions. Differential diagnoses for pigmented BCCs include melanoma in situ (MIS), invasive melanoma and benign pigmented lesions (2, 6). Usually, the clinical features suffice to establish the diagnosis of a pigmented BCC, but the diagnosis is not always clear-cut (Fig. 1). In uncertain cases diagnostic biopsy or excision is required.

Various non-invasive imaging modalities have been developed to aid in the diagnosis of BCC and to avoid unnecessary biopsies. These include reflectance confocal microscopy (RCM) and optical coherence tomography (OCT), as well as investigational methods still under development, such as Raman spectroscopy, high-resolution ultrasonography, and terahertz pulse imaging (4). Hyperspectral imaging (HSI) is a novel non-invasive imaging technique that measures the reflectance spectra of skin in vivo to identify different biological tissues (7, 8). We have previously shown that HSI can delineate the lesion borders of ill-defined BCCs and lentigo maligna (LM) more accurately than clinical examination, and can detect the invasive component in lentigo maligna melanoma (LMM) (9–11).

Convolutional neural networks (CNN) represent a form of artificial intelligence in decision-making and
are a strongly emerging concept in dermatological diagnostics. CNNs can classify pigmented and non-pigmented skin tumours from clinical and dermoscopic images with an accuracy similar to that of dermatologists (12–14). In addition to 2-dimensional (2D) images, a CNN classifier has been utilized with hyperspectral images of ex vivo malignant tumours (15, 16).

The aim of this pilot study was to use a combination of 3-, 2- and 1-dimensional (3D, 2D and 1D) convolutional layers in a neural network classifier to distinguish between pigmented BCCs and malignant melanocytic tumours.

MATERIALS AND METHODS

Participant recruitment

The study protocol complied with the principles of the Declaration of Helsinki and was approved by the local ethics committee of Tampere University Hospital. All participants gave their written informed consent. Voluntary patients were recruited from among those referred for suspected skin malignancies to the Department of Dermatology at Päijät-Häme Central Hospital, Lahti, Finland, between May 2014 and November 2017. Inclusion criteria were clinically pigmented tumours, which were histologically confirmed as BCC, MIS, or invasive malignant melanoma (MM).

Hyperspectral image acquisition and histopathological examination

All lesions were clinically examined with a dermatoscope (DermLite DL3, 3Gen, San Juan Capistrano, CA, USA) and photographed. For subsequent orientation purposes, a small mark was drawn on the healthy skin beside the lesion, using a black marker pen. Hyperspectral images were taken in vivo with an HSI camera prototype (VTT FPI VIS-VNIR Spectral Camera, VTT Technical Research Centre of Finland, Espoo, Finland). Spectral separation of the imager is based on Fabry-Pérot interferometer (FPI), which enables fast scanning in the spectral domain. The imager measures diffuse reflectance on wavebands from visible to near infrared light (500–850 nm) for every pixel of the image. The full width of half maximum (FWHM) of each waveband varies from 10 to 30 nm. The imager rapidly captures 76 wavebands within a few seconds in a 120 mm² field of view with a spatial resolution of 640 pixels/mm² (pixel = 125 × 125 μm). The imaging depth of hyperspectral imaging depends on the wavelength (17). In the wavelength range used (450–850 nm) the imaging depth varies in the range 0.5–5.0 mm. Because the whole wavelength range is used, its effect on the mean imaging depth is minor (17). The outcome is a hyperspectral image: a 3D data cube containing a reflectance spectrum for each pixel of the 2D image. The HSI technique is described in more detail elsewhere (7, 10).

After imaging, the lesions were surgically excised and sent for routine histopathological examination. The tissue specimens were fixed in 4% formalin, embedded in paraffin, sectioned using the traditional vertical bread-loaf technique, and stained with haematoxylin and eosin (H&E).

Data pre-processing and 3-dimensional convolutional neural network

Of the 76 wavebands, 66 were used for the analyses. Thus, HSI produced a raw data cube (size 240 × 320 × 66 pixels), which was calibrated to the radiance using Saari et al.'s method (18). A white reference target was captured for each data cube, and this was used to convert imaged radiance to reflectance R = I / I₀, where I is the imaged region of interest and I₀ is the data cube from the white reference. To reduce the effect of vignetting and lighting irregularities, each imaged spectrum was subtracted by its average white reference target was captured for each data cube, and this was calibrated to the radiance using Saari et al.'s method (18). A white

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RESULTS

Patient demographics and lesion characteristics

A total of 32 lesions fulfilling the inclusion criteria were imaged in Päijät-Häme Central Hospital, Lahti. Hyperspectral images of 6 lesions were excluded from the study because the HSI settings were not optimal at the time of imaging, leading to imaging artefacts (under-exposure or FPI switch inadvertently off). Hence, a total of 26 lesions from 24 patients were included in the study. Patient demographics and lesion characteristics are shown in Table I.

Fifteen patients were men and 9 women, mean age 73.1 (range 53–91) years. Thirteen had Fitzpatrick’s skin phototype II, 8 phototype III, and, in 3 patients, the phototype was not reported. Fourteen patients had a previous history of skin cancer, as follows: AK (n=8), Bowen’s disease (n=4), squamous cell carcinoma (n=3), BCC (n=9), and MIS or MM (n=4). Five patients had a history of other malignancy (2 prostate cancers, one tongue cancer, one lip cancer, and one hepatocellular and pancreatic carcinoma). Two of the patients were immunosuppressed: one patient was an organ-transplant receiver and used sirolimus for antirejection, and one patient used azathioprine for Crohn’s disease.

The imaged lesions were located on the face (n=8), neck (n=3), torso (n=10), upper extremity (n=4) and lower extremity (n=1). The mean ± standard deviation (SD) diameter of the lesions was 9.8 ± 2.9 mm (range 5.3–19 mm) and the mean ± SD area was 71.2 ± 46.9 mm² (range 11.0–247 mm²).

Histopathological diagnoses

The histopathological diagnoses of the imaged lesions were as follows: 10 BCC (6 nodular, 2 superficial, 2 both superficial and nodular), 12 MIS (all lentigo maligna subtype), and 4 MM (3 lentigo maligna subtypes and 1 superficial spreading subtype, Breslow depths in the range 0.3–1.4 mm).

Hyperspectral analysis

The hyperspectral analyses and lesion classification with CNN were conducted by a trained mathematician (IP). Due to limited sample size, all melanocytic tumours (MIS and MM) were first combined in single group for lesion classification. Each hyperspectral image was classified into one of the following: non-malignant (nBCC), melanocytic (MM), pigmented malignant melanoma in situ (MIS). The classification was applied to the whole lesion. The predicted label was determined by the majority voting of the CNN’s prediction of the class of each pixel. The classification result is shown in Table I.

| Lesion number | Age, years | Sex (F/M) | Lesion location | Size, mm | Histopathological diagnosis (n=true label) | Predicted label by CNN (2 classes) | Predicted label by CNN (3 classes) |
|---------------|------------|-----------|----------------|---------|------------------------------------------|-----------------------------------|-----------------------------------|
| 1             | 56         | F         | Eyelid         | 2 × 7   | nBCC | Pigmented BCC | Pigmented BCC |
| 2             | 82         | F         | Temple         | 11 × 12 | LMM (Breslow 1.5) | Melanocytic MM | Melanocytic MM |
| 3             | -          |          | Temple         | 5 × 6   | nBCC | Pigmented BCC | Pigmented BCC |
| 4             | M          | Upper back| 8 × 8          | LM      | Melanocytic | MIS | Melanocytic |
| 5             | M          | Groin     | 8 × 10         | n/sBCC | Melanocyticc | Pigmented BCC | Pigmented BCC |
| 6             | 69         | M         | Shoulder       | 15 × 21 | MM (Breslow 1.4) | Melanocytic | MM |
| 7             | 57         | F         | Breastbone     | 7 × 8   | sBCC | Pigmented BCC | Pigmented BCC |
| 8             | 73         | M         | Back           | 10 × 13 | sBCC | Pigmented BCC | Pigmented BCC |
| 9             | 81         | F         | Neck           | 6 × 13  | n/sBCC | Pigmented BCC | Pigmented BCC |
| 10            | 72         | F         | Nose           | 10 × 10 | nBCC | Pigmented BCC | Pigmented BCC |
| 11            | 53         | F         | Eyelid         | 4 × 12  | nBCC | Pigmented BCC | Pigmented BCC |
| 12            | 86         | F         | Upper back     | 7 × 12  | nBCC | Pigmented BCC | Pigmented BCC |
| 13            | 66         | M         | Shoulder       | 9 × 9   | LM | Melanocytic | MIS |
| 14            | 75         | M         | Shoulder       | 5 × 10  | nBCC | Pigmented BCC | Pigmented BCC |
| 15            | 91         | M         | Upper back     | 7 × 10  | LM | Melanocytic | MIS |
| 16            | 84         | M         | Chest          | 4 × 8   | LMM (Breslow 0.3) | Melanocytic | MM |
| 17            | 83         | M         | Chest          | 8 × 12  | LMM (Breslow 0.4) | Melanocytic | MM |
| 18            | 69         | M         | Forehead       | 8 × 10  | LM | Melanocytic | MIS |
| 19            | 80         | M         | Neck           | 8 × 12  | LM | Melanocytic | MIS |
| 20            | 62         | M         | Forearm        | 10 × 17 | LM | Melanocytic | MIS |
| 21            | 75         | M         | Chest          | 11 × 13 | LM | Melanocytic | MIS |
| 22            | 77         | F         | Forearm        | 9 × 12  | LM | Melanocytic | MIS |
| 23            | 77         | M         | Cheek          | 10 × 14 | LM | Melanocytic | MIS |
| 24            | 71         | F         | Lower back     | 6 × 8   | LM | Melanocytic | MIS |
| 25            | 79         | M         | Temple         | 8 × 8   | LM | Melanocytic | MIS |
| 26 b          | 70         | M         | Neck           | 6 × 8   | LM | Melanocytic | MIS |

The predicted label for the whole lesion was determined by the predicted class of the majority of pixels in the lesion area. The result of classification is presented for the study sample divided into 2 classes (pigmented basal cell carcinoma (BCC) and melanocytic tumours) and 3 classes (pigmented BCC, melanoma in situ (MIS) and malignant melanoma (MM)).

Lesions 2 and 3 belonged to the same patient. Lesions 25 and 26 belonged to the same patient. Convolutional neural networks (CNN) classification erroneous. nBCC: nodular basal cell carcinoma; sBCC: superficial basal cell carcinoma; LMM: lentigo maligna melanoma.
Fig. 2. Classification and likelihood maps of a histologically confirmed invasive melanoma (lentigo maligna subtype, Breslow depth 1.5 mm). The lesion was correctly classified as malignant melanoma (MM) by hyperspectral imaging using the majority of the pixels method. (A) Red, green and blue (RGB) image taken by hyperspectral camera. (B) Classification map showing most of the pixels representing invasive malignant melanoma (MM). Upper colour bar depicts the 4 classes in image B. (C) Likelihood map for the localization of MM. (D) Likelihood map for the localization of pigmented basal cell carcinoma (BCC). Only some scattered artefactual pixels are shown outside the lesion area. (E) Likelihood map for the localization of marker pen. (F) Likelihood map for the localization of healthy skin. Lower colour bar represents the certainty of the classifier in each of the likelihood maps. Red line shows the division of the hyperspectral image into halves, one of which was used for the training of the classifier and the other for the classifying task.

Fig. 3. Classification and likelihood maps of a histologically confirmed pigmented basal cell carcinoma (BCC). The lesion was correctly classified as BCC by hyperspectral imaging using the majority of pixels method. (A) Red, green and blue (RGB) image taken by hyperspectral camera. (B) Classification map showing most of the pixels classified as pigmented BCC. (C) Likelihood map for the localization of invasive melanoma (malignant melanoma; MM). (D) Likelihood map for the localization of pigmented basal cell carcinoma (BCC). (E) Likelihood map for the localization of marker pen. (F) Likelihood map for the localization of healthy skin. Lower colour bar represents the certainty of the classifier in each of the likelihood maps. Red line shows the division of the hyperspectral image into halves, one of which was used for the training of the classifier and the other for the classifying task.
Hyperspectral imaging can distinguish melanoma from pigmented BCC

pixel-wise by 3D CNN, which produced a classification map comprehending 4 classes (“pigmented basal cell carcinoma”, “melanocytic tumour”, “healthy skin” and “marker pen”) (Figs 2–4). The last layer of the classifier also created likelihood maps indicating how certain the CNN is about the classification. To obtain the classification for each pixel over each image, the maximum likelihood (the highest value of each pixel in likelihood maps) was used. Two different approaches were used for the analyses: (i) a majority of pixels classification; and (ii) a pixel-wise classification. The histopathological diagnoses of the excision specimens represented the true classification (true label) of the lesions.

In the majority of pixels classification, the classification maps were used to acquire an automated diagnosis for the imaged lesions, i.e. the whole lesion was predicted to belong to the same class as the majority of the pixels in the lesion area. In addition, the accuracy of CNN classification per individual pixel was computed in comparison with the manual annotations in the images (4 pixel classes, as discussed above). By using the pixel-wise classification the number of classified areas increased from 26 lesions to 164,144 classified pixels. The confusion matrices for both lesion and pixel-wise classifications are shown in Fig. 5.

For the majority of pixel classification, sensitivity was 100% (16/16) (95% CI 81–100%), specificity 90% (9/10) (95% CI 60–98%) and PPV 94% (16/17) (95% CI 73–99%) in differentiating melanocytic tumours from pigmented BCCs. Only one true pigmented BCC was falsely predicted as a melanocytic tumour, and all melanocytic tumours were correctly predicted according to their true label. In pixel-wise classification the sensitivity for differentiating the class “melanocytic tumours” from the class “pigmented BCC” was 99.98% (95% CI 99.95–99.99%), specificity 93.2% (95% CI 92.6–93.7%) and PPV 97.8% (95% CI 97.6–98.0%).

This study also tested the accuracy of the CNN classifier, by dividing the sample into 3 groups: pigmented BCC (n = 10), MIS (n = 12) and MM (n = 4), and then used the classification maps to acquire an automated diagnosis for every lesion in a manner similar to that described above. Classifier accuracy by individual pixels was also computed (5 pixel classes). In the lesion classification using the majority of pixels, calculated sensitivity was 100% (4/4) (95% CI 51–100), specificity 100% (22/22)
(95% CI 85–100) and PPV 100% (4/4) (95% CI 51–100) in distinguishing MM from other lesions (MIS and pigmented BCC). In pixel-wise classification between tumour classes only, the statistical characteristics for class “MM” were as follows: sensitivity 99.6% (95% CI 99.5–99.7), specificity 98.6% (95% CI 98.5–98.8) and PPV 97.3% (95% CI 97.0–97.6). The confusion matrices for 3-class lesion classification and 5-class pixel-wise classification are shown in Fig. 4. The result of the CNN classification for all lesions in comparison with true histopathological diagnoses is shown in Table I. The difference in the spectra between different lesion types and healthy skin is shown in Fig. S3.

DISCUSSION

To the best of our knowledge, this pilot study is the first to utilize in vivo hyperspectral imaging (HSI) and a deep learning CNN as a tumour classifier for skin tumours. The CNN lesion classifier using 2 different approaches (majority of pixels classifier and single pixel classifier) reached high sensitivity and specificity in distinguishing melanocytic tumours from pigmented BCCs.

HSI is a novel non-invasive optical imaging method that can be used to image tissues in vivo and ex vivo tissues rapidly in various medical applications. It produces a hyperspectral data cube in which different tissue types can be identified and visualized based on their specific reflectance spectra (8). Map-like images generated by HSI can be used to delineate malignant skin lesions preoperatively and to visualize invasive parts in large lesions to target biopsy sites (9–11).

Furthermore, HSI could be useful for dermatologists as a pre-surgical diagnostic aid for planning the surgical excision margins. HSI could be a helpful diagnostic tool even for inexperienced physicians, since it is user-independent and gives an automated most likely diagnosis.

No earlier reports on HSI combined with CNN for tumour classification were found in the literature. Among the available non-invasive diagnostic modalities, one reminiscent of the current method is MelaFind®, a multispectral imaging (MSI) system that acquires 10 wavebands in the range 430–950 nm, and uses automated algorithms based on linear classifiers for pattern recognition and differential diagnosis of pigmented lesions (24). MelaFind has demonstrated high sensitivity, of 82–98%, but only moderate specificity, of 8–52%, in recognizing melanomas (25–27). A limitation of MelaFind is that it may misclassify non-melanoma skin cancers, such as BCCs, because the technology is designed to assess overall structural disorganization of lesions, rather than atypical cellular features (28). Another non-invasive MSI technique is SIAscope (spectrophotometric intracutaneous analysis), which captures 8 narrowband spectral images in the range 400–1,000 nm to visualize the quantity and microscopic architecture of chromophores (melanin, blood and collagen) in the lesion area (29). However, SIAscope does not improve diagnostic accuracy over dermoscopy in the diagnosis of pigmented BCC (30). Tomatis et al. (31) developed an artificial neural network (ANN) classifier for 1,391 pigmented lesions imaged with MSI, which was able to distinguish melanoma from other lesions with a sensitivity of 80% and specificity of 76%. In another study, Carrara et al. (32) developed an ANN classifier for MSI data to differentiate between reassuring lesions and those requiring excision, using a dataset of 1,966 excised and 1,940 non-excised and clinically non-suspicious lesions, resulting in sensitivity of 88% and specificity of 80%. For comparison, in meta-analyses, dermoscopy has achieved sensitivity and specificity of 88% and 86%, respectively, for the diagnosis of melanoma (33), 82.4% and 83.5% for the diagnosis of LM and LMM (34), and 81.9% and 81.8% for the diagnosis of superficial BCC (35). However, the diagnostic accuracy of dermoscopy is user-dependent and depends on the experience of the clinician. Recently, deep learning CNN have been utilized to facilitate and automate diagnostic classification in clinical and dermoscopic images, reaching diagnostic accuracies similar to those achieved by expert dermatologists, with the area under the curve (AUC) of the receiver operating curve (ROC) ranging from 0.74 to 0.96 (12–14).

The results of the current study indicate that HSI might be capable of spectral differentiation at the cellular level and of recognizing the tumour cell of origin (melanocyte vs keratinocyte). The CNN classification maps in pixel-wise resolution can be thought to represent aggregates of cells in the microscopic structure of the skin. The hyperspectral image analysis appears, in most cases, to recognize the different cell types in pigmented BCC and melanocytic tumours regardless of the presence of melanin pigment. Both tumour types have increased amounts of melanin in the dermis, often histologically visualized in macrophages. Thus, the results of the current study show that HSI does not only use pigment as a chromophore, but also recognizes other characteristics beyond the pigment. Further verification of this is required in studies at the cellular level. There were some cases with pixels misclassified as melanoma (Fig. 4) within the pigmented BCCs, which could be due to the fact that, histologically, there are some entrapped melanocytes in the areas of pigmented BCCs (36).

The current study used a combination of 3D, 2D and 1D CNN for automated pixel-wise tumour classification in hyperspectral images of the skin. 3D CNN was used because the standard 2D CNN used with clinical and dermoscopic photographs may not suffice to utilize spectral data, which is 3-dimensional in nature. This method produced classification maps of imaged lesions that predict the lesion’s diagnosis pixel-wise (Figs 2 and 3). The classification map not only offers identification (the most likely diagnosis), but also delineation and map-like repre-
sentation of the lesion. Figs 2 and 3 show that there are a few misclassified pixels in the tumour area, but that, by far, most pixels are classified correctly, predicting the true histopathological diagnosis. In addition, at the periphery of the image in the area of healthy skin there are some rare misclassified pixels. Furthermore, the black hair has been classified as “marker pen”, since no separate class was created for hair. The few misclassified pixels could be explained by imaging errors, such as local increased noise or other perturbation in the hyperspectral image, or by inaccuracy in the CNN model. When classifying whole lesions using the majority of pixels method, the few misclassified pixels were averaged, leading to very high sensitivity and specificity for differential diagnosis of melanocytic tumours from BCCs. In the case of 3 tumour classes, the accuracy of the CNN classifier was also very high with all tumours classified correctly. The confusion matrices of individual pixel classification (Fig. 4) demonstrate the robustness of the CNN’s pixel-wise classification, but it should be noted that the pixel-wise accuracy is presented only in comparison with manually annotated pixel classes in images. The pixel-wise classification is fairly accurate in distinguishing tumour classes from each other, with high sensitivity and specificity for both 2-class and 3-class lesion classification. However, if all (4 or 5) pixel classes are observed, it can be seen that there is a portion of “healthy” annotated pixels that are assigned to tumour classes “pigmented BCC”, “melanocytic tumours”, “MIS” and “MM”, which are false positives in the whole imaged region and represent the inherent error in CNN classification. In this study setting, the majority of pixels method can be assumed to predict the diagnosis reliably, because we know a priori that no single BCC cells are situated inside a melanocytic tumour and vice versa. This is not necessarily the case when, for example, multiple melanocytic tumours are included in the classification task. In the development of the CNN classification method, this study strove for pixel-wise classification, because there are clinical situations in which a single skin lesion may include multiple tumour types (e.g. an invasive part of surrounding carcinoma in situ, or invasive melanoma arising in melanoma in situ or in pre-existing naevus). The CNN method used in the present study is novel and can be developed further. In addition, the classification of CNN improves with larger training datasets, which will require the collection of larger samples of different tumour types and from different locations on the body. With a larger training set the pixel-wise classification of CNN could be further improved to be more robust, thereby reducing the number of misclassified pixels.

Study limitations

This study has some limitations. Due to the pilot nature of the study, the sample size was small and halves of the images were used for the CNN training phase. Ideally, the CNN training should be conducted with a dataset separate from that used for the classification task. However, the small sample size was compensated by the pixel-wise analysis, in which the pixels were analysed separately and could be counted as separate lesions. This increases the reliability of the findings. It should be noted that benign tumours clinically mimicking pigmented BCCs (benign lentigines, seborrhoeic keratosis, benign naevi) were not included, and further studies on this topic are warranted. The CNN classification method used is novel and requires further development work. An HSI camera prototype was used in this study, which could be developed further to be more robust in order to avoid imaging noise and artefacts. There is currently no commercially available HSI device for skin imaging.

Conclusion

This is the first study to utilize hyperspectral images and deep-learning CNN to achieve automated diagnosis for melanocytic tumours and pigmented BCCs. The results will therefore serve as a basis for future studies.

The results of this study suggest that, with a larger sample and training dataset, the CNN could accurately and pixel-wise distinguish between malignant melanocytic tumours and pigmented BCCs. To confirm these preliminary results, future studies with larger samples and multiple tumour types, including all types of melanocytic lesions, are warranted.

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