Gallbladder cancer integrated bioinformatics analysis of protein profile data

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ABSTRACT
Aim: Identifying the critical genes that differentiate gall bladder cancer from a normal gall bladder and the related biological terms was the aim of this study.

Background: The molecular mechanism underlying gall bladder cancer (GBC) trigger and development still requires investigations. Potential therapeutic biomarkers can be identified through protein-protein interaction network prediction of proteome as a complementary study.

Methods: Here, a literature review of proteomics studies of gall bladder cancer from 2010 to 2019 was undertaken to screen differentially expressed proteins in this cancer. A network of 27 differentially expressed proteins (DEPs) via Cytoscape 3.7.1 and its plug-ins was constructed and analyzed.

Results: Ten proteins were introduced as hub-bottlenecks among which four were from DEPs. The gene ontology analysis also indicated that positive regulation of multi-organism process and regulation of response to biotic stimulus are the most disrupted biological processes of GBC considering their relationships with the DEPs.

Conclusion: ACTG, ALB, GGH, and DYNC1H1, and relative biological terms were introduced as drug targets and possible diagnostic biomarkers.

Keywords: Gallbladder cancer, Protein-protein interaction network analysis, Hub-bottleneck proteins, Biological process.
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Introduction
Introducing valuable biomarkers of gallbladder malignancy as the most known frequent type of the biliary tract cancer is essential for early diagnosis and treatment approaches. This type of cancer is recognized as the fifth frequent malignancy of the gastrointestinal tract (1). While it is a rare type of cancer, its incidence is high in some countries including China, Pakistan, India, and Chili (2-4). The mortality rate of this type of cancer is very high because of its latent phase of trigger and speedy metastatic behavior (5). In this regard, most of the patients diagnosed with this type of cancer are at their advance stage of their disease by the time of diagnosis (5). The first line therapy for this cancer is surgery as other approaches have shown to be ineffective. Nevertheless, surgery is not a suitable method for all patients and only limited numbers of them can benefit from this approach (5). By identifying biomarkers related to the early stage of GC, it is possible to better understand the mechanism of the disease as well as design proper treatments. One of the famous molecular applications for early stage biomarker discovery is proteomics study. Using this approach, a vast number of proteins with differential expression could be detected. These elements are vital
for cell biological processes and function. Thus, any dysregulation of these agents could culminate in malfunction of the cell system and consequently abnormal behavior in an organism. The altered expression proteins which are specific and sensitive are known as biomarkers that are valuable for drug targeting (6). However, the biomarkers are not only signal agents that cause the damage; there could be complex interactions between them and other molecules such as genes and metabolites inducing a major disruption in an organism. As such, detection of these connections and the most significant nodes in a system of interaction is worth further examination (7). One of the well-known interactions is called protein-protein interaction network. By analyzing a protein map, it is possible to identify the most key contributors of a network structure and strength as well as the pattern of that specific condition such as a disease (8). Hence, promising biomarkers of a cancer can be evaluated for their interaction properties in a network of PPI connections. In this integrative bioinformatics study, it is aimed to collect the proteomics biomarkers reported for gallbladder cancer and set a complementary analyses of interaction behaviors of these biomarkers, thereby introducing the most key players in this cancer in terms of interaction properties.

Methods

The studies of gall bladder cancer proteomics were searched through Google Scholar and PubMed sources. The keywords used for our search were “Proteomics” and “Gall bladder Cancer”. These investigations had been published from 2010 to 2019. The proteins highlighted by these studies were gathered and then chosen for further analysis. Cytoscape v.3.7.1 and String db were used to construct a network of GBC, while the topological features were analyzed by Network Analyzer (9, 10). Note that the parameters for network centrality analysis are degree and betweenness. Nodes with the highest values of degree and betweenness are called hubs and bottlenecks, respectively. Elements with both features are hub-bottlenecks which are the most central proteins of the PPI network. The enrichment analysis of DEPs is the next step which involves a biological process (BP), molecular function (MF), cellular component (CC), and KEGG pathways. Meanwhile, BP has been the designated analysis for differential expressed proteins via ClueGO+Clue Pedia (11, 12). The statistical criteria for this procedure include the minimum number of genes; 2 and percentage in term; 1, respectively. For group P value correction, Bonferroni step down was used while for enrichment/depletion, two-sided hypergeometric test was applied. Asterisk signs in the grouping terms indicate statistically significant groups. Two star implies highest significance values of that group, while no star indicates no statistical significance.

Results

Table 1 presents a total of 27 DEPs (characterized with about 50% expression change) from investigation of six proteomics which are the utilized samples related to the human gall bladder cancer.

| Row | Protein Name | Uniprot Accession | Expression |
|-----|--------------|-------------------|------------|
| 1   | ANXA4(5)     | P09525            | Up         |
| 2   | ACTA2 (5)    | P62736            | Down       |
| 3   | ALB(5)       | P02768            | Up         |
| 4   | Hsp90B(5)    | P08238            | Down       |
| 5   | Dynclh1(5)   | Q14204            | Down       |
| 6   | ACTG (5)     | P63261            | Up         |
| 7   | DES(13)      | P17661            | Down       |
| 8   | HTRA1(14)    | Q92743            | Down       |
| 9   | TALN(14)     | P37802            | Down       |
| 10  | CTSZ(14)     | Q9UBR2            | Up         |
| 11  | GM2A(14)     | P17900            | Up         |
| 12  | CTSH(14)     | P09668            | UP         |
| 13  | GGH(14)      | Q92820            | Up         |
| 14  | NAGA(14)     | P17050            | Up         |
| 15  | NEFH(14)     | P12036            | Down       |
| 16  | RUSU(14)     | Q15404            | Down       |
| 17  | MUC13(14)    | Q9H13R2           | Up         |
| 18  | NUCKS1(14)   | Q9H1E3            | Up         |
| 19  | DMBT1(14)    | Q9UGM3            | Up         |
| 20  | HMGB2(14)    | P26583            | Up         |
| 21  | LAMB3(14)    | Q13751            | Up         |
| 22  | PSAP(14, 15) | P07602            | Up         |
| 23  | MIF(15)      | P14174            | Up         |
| 24  | ANK3(16)     | Q12955            | Down       |
| 25  | FHL1(16)     | Q13642            | Down       |
| 26  | ANXA3(17)    | P12429            | Up         |
| 27  | S100A8(14)   | P05109            | Down       |

Cytoscape via String db interaction analysis of the integrated DEPs based on Table 1 is presented in Figure 1. Addition of 50 proteins from STRING database to the main network of 27 DEPs leads to participation of the isolated proteins in the interactome.
Figure 1. String network of 27 DEPs of GBC with 27 links; Seven proteins remained as individual proteins in the network.

Figure 2. String network of 77 nodes with 917 links; Nodes with the highest degree and betweenness values are larger in size and darker orange, respectively.
The resultant network with centrality values, analyzed and visualized via Network Analyzer, is shown in Figure 2.

MUC13 and RSU1 remain as individuals after addition of 50 neighbor proteins to the query proteins. Some nodes show larger centrality values in terms of degree and betweenness, suggesting that the network is scale free.

Centrality analysis of the second network conducted by the Network Analyzer, the Cytoscape plug-in as the hub-bottlenecks, is tabulated in Table 2.

According to Table 2, ten hub-bottlenecks of this network contain four DEPs called DYNC1H1, ACTG, GGH, and ALB. The other proteins, on the other hand, could also be important in the GBC pathogenicity. In this regard, literature review of all central proteins has been considered in our study. DYNC1H1, ACTG, and ALB are reported as DEPs of GBC by a proteomics approach (5). MAPK1 as the most highlighted hub is a key differential protein in transition from GBC non-invasive to invasion condition based on a proteomics study (18). ALB and ACTG have the highest BC and as mentioned they are also from DEPs.

Gene ontology analysis via ClueGO has been conducted for DEPs in terms of biological process identifications. The pie chart summarizes the terms in groups of BPs. Single asterisk (*) shows p<0.05 and Double asterisk (**) implies p<0.01. Groups without stars are not statistically significant which have been omitted in this analysis by setting the query to show only groups with P value less than 0.05, see Figure 3.

Eleven groups have been identified using ClueGO, among which one has one star suggesting it as significantly lower than 0.01. Positive regulation of

![Figure 3. A pie chart view of biological processes identified by ClueGO for 27 DEPs. The asterisks indicate the significance of the grouping.](image)

| Row | Display name | Protein name | Degree | BC  |
|-----|--------------|--------------|--------|-----|
| 1   | MAPK1        | Mitogen-Activated Protein Kinase 1 | 43     | 0.03|
| 2   | LYZ          | Lysozyme     | 41     | 0.04|
| 3   | DYNC1H1*     | Cytoplasmic dynein 1 heavy chain 1 | 40     | 0.02|
| 4   | ALB*         | Albumin      | 40     | 0.05|
| 5   | ACTB         | Beta-actin   | 40     | 0.03|
| 6   | ANXA2        | Annexin A2   | 40     | 0.02|
| 7   | GGH*         | Gamma-Glutamyl Hydrolase | 39 | 0.02|
| 8   | ACTR10       | Actin Related Protein 10 | 39     | 0.02|
| 9   | ACTG*        | Actin Gamma 1 | 38     | 0.05|
| 10  | INS          | Insulin      | 38     | 0.03|
multi-organism process and regulation of response to bionic stimulus are the most highlighted groups in the BP query of DEPs based on the number of gene participations. These two groups claim around 18% of all groups in terms of protein participations. Both of these groups are statistically highly significant based on the Group P value Corrected with Bonferroni step down as 0.005 and 0.00002. The second ranked groups in this query are positive regulation of prostaglandin secretion involved in immune response and neutrophil degranulation claiming 16% of all groups.

The list of proteins participating in the first and second groups include CTS, DMBT1, HMGB2, MIF, MUC13, NUCKS1, PSAP, S100A8, HTRA1, and CTS, DMBT1, HMGB2, HTRA1, MIF, MUC13, PSAP, S100A8, respectively. These proteins are CTSH, PSAP, and S100A8 where the first two are linked to six groups while the latter one is associated with five groups.

Discussion

GBC is the most fatal fast growing type of cancer of the biliary tract (19) where only 20% of these patients are diagnosed as un-metastatic (20). Molecular investigation could provide essential knowledge of this lethal cancer especially in terms of protein dysregulations and their crucial interactions. There are some proteomics studies available from 2010 to 2019 which identified many key proteins. We performed a meta-analysis of the 27 DEPs introduced by these studies via bioinformatics. Different patterns of expressions are assigned for these proteins in the GBC network. These proteins could provide insight into molecular mechanisms of GBC and more precisely via network analysis. A network analysis of DEPs showed that seven proteins including MUC13, NUCKS1, RSU1, HMGB2, DES, DMBT1, and LAMB3 remained as individual proteins in the first query. As we added 50 neighbor proteins to the query proteins, RSU1 and MUC13 remained as separate proteins. Thus, the rest of individual query proteins from the first network joined the main network of interactions. This demonstrates that these two latest mentioned proteins do not show definite interactions with the rest of DEPs.

As the analysis continued with the centrality identification of GBC, hub-bottlenecks, i.e. key proteins were introduced. Among the ten hub-bottlenecks, four proteins belonged to the differential expressed set of GBC. These proteins were DYNCH11, ACTG, GGH, and ALB with the first one being down-regulated while the rest up-regulated in GBC. It can be inferred that up-regulation is dominant among the DE hub-bottlenecks in this study.

Although MAPK1 does not belong to the list of DEPs in GBC, its phosphorylation has been mentioned in one proteomics study (18). This protein has been additionally reported for other types of tumors such as cervical, colon, as well as head and neck cancers (21-24). In addition, some relationships between MAPK1 and GBC have been reported by other studies as well promoting the idea that this protein may have a contribution to GBC initiation likewise (25, 26). However, additional examinations are required in this regard.

Lysozyme, the next ranked hub-bottleneck of the GBC network, has been reported with high expression in the sera of patients with malignancies including cancers of lung, melanoma (27), and breast carcinomas (28). This protein also shows metaplastic alterations in gallbladder cancer developments (29). DYNCH11 is the third ranked hub-bottleneck and as the first ranked in DE hub-bottleneck. This protein indicates down-regulation in GBC and is highlighted as diagnostic biomarker by a proteomics study (5). Further, this protein is important in other cancers as also indicated by literature reviews (30). This protein has many fundamental responsibilities in a cell one of which is contribution to mitotic process which plays a key role in cancer (31). Albumin the next ranked hub-bottleneck and second ranked as DE hub-bottleneck is up-regulated in GBC according to the same proteomics evaluation (5). ACTB, the beta actin, has some linkage to different types of tumors. This house-keeping element has mostly showed over-expression in different cancers (32). Regarding GBC, it is still to be investigated and might have some connections. The sixth ranked hub-bottleneck, ANXA2 regulation changes is pinpointed as high expression in GBC by
one immunohistochemical research (33). In addition, it is accounted as a trustworthy marker as an approach to screening and treatment follow-ups of many kinds of cancers (34, 35). Gamma-Glutamyl Hydrolase, the third ranked DE hub-bottleneck, is up-regulated in GBC as mentioned by a proteomics investigation (14). It is also highly expressed in other types of cancers as well (36-38). Considering ACTR10, the eighth ranked hub-bottleneck, there is no report concerning its relationship with either with gall bladder cancer or with any other cancer types. ACTG the ninth ranked hub-bottleneck and fourth among DE hub-bottlenecks, showed up-regulation in GBC assigned with a proteomics study (5). Its over-expression has been previously reported for skin cancer (39) as well. The crucial role of gamma actin in cancer is mitotic process and centrosome performance regulations (40). Finally, the last important hub-bottleneck of the network is INS, insulin, which is known as cancer metabolism promoter (41). Collectively, the DE proteins among the hub-bottlenecks, ACTR10 was the only protein not reported for any types of cancers while the rest, based on previous reports showed some connections with different kinds of cancers. Interestingly, MAPK1, ANXA2, and LYZ specified some alterations in GBC by other types of studies rather than proteomics. Thus, they could be important as well for GBC pathogenicity. The next step was to evaluate biological processes related to DEPs. Significant biological processes (p-value <0.01; labeled by ** and p-value, 0.05; labeled by * in the figure 3) were considered. In this way, the aberrant processes of gall bladder cancer could be explored. Indeed, changes in the DE proteins could result in the malfunction of the related biological processes, especially for the DE hub-bottlenecks, it has additional values. As mentioned before, in this investigation, there are some DE proteins based on meta-analysis of proteomics data that indicate centrality values in the network of GBC, including ACTG, ALB, GGH, and DYNC1H1. Hence, the expression changes of these central proteins could conclude in extensive abnormal behavior in our network and accordingly the development of GBC. Further, MAPK1 as the most central hub-bottleneck of the GBC network has previously been found as a DEP in invasion behavior of GBC which could also have a role in other stages of this cancer type that warrants additional analysis in this regard.

A panel of biomarkers could be more trustworthy than only one assigned biomarker. In this regard, proteins that have been reported by proteomics studies for GBC were gathered and those with greater importance in terms of interactions were introduced as ACTG, ALB, GGH, and DYNC1H1. This panel could be suggested as a promising therapeutic target for screening of GBC once confirmed by complementary evaluations. The follow up of patients and evaluation of treatment are the two important features of application of this finding if more investigation validates our outcomes.

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Conflict of interests

The authors declare that they have no conflict of interest.

References

1. Tan Y, Ma SY, Wang FQ, Meng HP, Mei C, Liu A, et al. Proteomic-based analysis for identification of potential serum biomarkers in gallbladder cancer. Oncol Rep 2011;26:853-9.

2. Wi Y, Woo H, Won YJ, Jang JY, Shin A. Trends in gallbladder cancer incidence and survival in Korea. Cancer Res Treat 2018;50:1444.

3. Jaruvongvanich V, Yang JD, Peeraphatdit T, Roberts LR. The incidence rates and survival of gallbladder cancer in the USA. Eur J Cancer Prev 2019;28:1-9.

4. Tuo JY, Zhang M, Zheng RS, Zhang SW, Li GC, Yang NN, et al. Report of incidence and mortality of gallbladder cancer in China, 2014. Zhonghua Zhong Liu Za Zhi 2018;40:894-899.

5. Nazemalhosseini-Mojarad E, Haghighi A, Taghipour N, Keshavarz A, Mohebi SR, Zali MR, et al. Subtype analysis of Cryptosporidium parvum and Cryptosporidium hominis isolates from humans and cattle in Iran. Vet Parasitol. 2011;179:250-2.

6. Alizadah AH, Ranjbar M, Ansari S, MirArab A, Alavian SM, Mohammad K, et al. Seroprevalence of hepatitis B in Nahavand, Islamic Republic of Iran. East Mediterr Health J. 2006;12:528-37.

7. Mahboubi M, Azodi MZ, Tavirani MR, Mansouri V, Ahmadi NA, Hamdigh M, et al. Protein-Protein Interaction Analysis of Common Top Genes in Obsessive-Compulsive Disorder (OCD) and Schizophrenia: Towards New Drug Approach Obsessive-Compulsive disorder (OCD) and Schizophrenia Comorbidity Gene Analysis. Iran J Pharm Res 2018; S17:173.
8. Tavirani M, Zamanian-Azodi M, Rezaei-Tavirani M, Vafaee R. Interaction network prediction and analysis of Anorexia Nervosa. Iran J Child Neurol 2019;13:45-54.

9. Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, et al. Cytoscape: a software environment for integrated models of biomolecular interaction networks. Genome Res 2003;13:2498-504.

10. Szklarczyk D, Morris JH, Cook H, Kuhn M, Wyder S, Simonovic M, et al. The STRING database in 2017: quality-controlled protein–protein association networks, made broadly accessible. Nucleic Acids Res 2017;45:D362-8.

11. Bindea G, Galon J, Mlecnik B. CluePedia Cytoscape plugin: pathway insights using integrated experimental and in silico data. Bioinformatics 2013;29:661-3.

12. Bindea G, Mlecnik B, Hackl H, Charoentong P, Tosolini M, Kirilovsky A, et al. ClueGO: a Cytoscape plug-in to decipher functionally grouped gene ontology and pathway annotation networks. Bioinformatics 2009;25:1091-3.

13. Bhunia S, Barbhuiya MA, Gupta S, Shrivastava BR, Tiwari PK. Abstract A14: Downregulation of desmin involves gene promoter hypermethylation in gallbladder cancer. AACR; 2018.

14. Sahasarabudhe NA, Barbhuiya MA, Bhunia S, Subbannayya T, Gowda H, Advani J, et al. Identification of prosaposin and transgelin as potential biomarkers for gallbladder cancer using quantitative proteomics. Biochem Biophys Res Commun 2014;446:863-9.

15. Barbhuiya MA, Subbannayya T, Leal-Rojas PA, Shahasarabudhe N, Renuse S, Patil AH, et al. Su1999 Proteomic Analysis of Gallbladder Cancer to Identify Biomarkers and Therapeutic Targets. Gastroenterology 2015;148:571.

16. Truckenmueller F, Goeppert B, Pusch S, Heinze I, Kirkpatrick J, Schirmacher P, et al., editors. Mass-spectrometric analysis of deregulated proteins in gallbladder carcinoma. Journal of hepatology; 2019: elsewhere science bv po box 211, 1000 ae amsterdam, netherlands.

17. Tan Y, Meng H, Wu Q, Wang F, Wu H. Proteomic study of gallbladder cancer, with special reference on the expression and significance of annexin A3. Zhonghua Bing Li Xue Za Zhi 2010;39:382-6.

18. Subbannayya T, Leal-Rojas P, Zhavoronkov A, Ozarov IV, Korzinkin M, Babu N, et al. PIM1 kinase promotes gallbladder cancer cell proliferation via inhibition of proline-rich Akt substrate of 40 kDa (PRAS40). J Cell Commun Signal 2019;13:163-77.

19. Bhunia S, Gupta S, Shrivastava BR, Tiwari PK. Identification of S100 calcium binding protein A9 as a prognostic biomarker in gallbladder cancer. Meta Gene 2018;18:62-7.

20. Rai R, Gosai S, Rao CV, Chandra V. Molecular Diagnosis of Gall Bladder Cancer. Molecular Diagnostics in Cancer Patient 2019;11-25.

21. Burotto M, Chiou VL, Lee JM, Kohn EC. The MAPK pathway across different malignancies: a new perspective. Cancer 2014;120:3446-56.

22. Li XW, Tuergan M, Abulizi G. Expression of MAPK1 in cervical cancer and effect of MAPK1 gene silencing on epithelial-mesenchymal transition, invasion and metastasis. Asian Pac J Trop Med 2015;8:937-43.

23. Najar AG, Pashaei-Asl R, Omidi Y, Farajinia S, Nourazarian AR. EGFR antisense oligonucleotides encapsulated with nanoparticles decrease EGFR, MAPK1 and STAT3 expression in a human colon cancer cell line. Asian Pac J Cancer Prev 2013;14:495-8.

24. Reyes-Gibby CC, Wang J, Silvas MRT, Yu R, Yeung S-CJ, Shete S. MAPK1/ERK2 as novel target genes for pain in head and neck cancer patients. BMC Genetics 2016;17:40.

25. Mohri D, Iijichi H, Miyabayashi K, Takahashi R, Kudo Y, Sasaki T, et al. A potent therapeutics for gallbladder cancer by combinatorial inhibition of the MAPK and mTOR signaling networks. J Gastroenterol 2016;51:711-21.

26. Buchegger K, Silva R, López J, Ili C, Araya JC, Leal P, et al. The ERK/MAPK pathway is overexpressed and activated in gallbladder cancer. Pathol Res Pract. 2017;213:476-82.

27. Lugter T, Koschoschka E, Sagaster P, Micksche M. Serum lysozyme levels in patients with solid tumors. Oncology 1979;36:15-8.

28. Serra C, Vizoso F, Alonso L, Rodríguez JC, González LO, Fernández M, et al. Expression and prognostic significance of lysozyme in male breast cancer. Breast Cancer Res 2002;4:R16.

29. Sai K, Onda M, Ozawa Y, Honjo T, Okuta T, Sumiyama Y. Relationship between metaplastic changes and occurrence of endocin cells, lysozyme and lactoferrin in gallbladder carcinoma. Ann Cancer Res Ther 1998;7:34-8.

30.Sucularli C, Arslantas M. Computational prediction and analysis of deleterious cancer associated missense mutations in DYNC1H1. Mol Cell Probes 2017;34:21-9.

31. Gong LB, Wen T, Li Z, Xin X, Che XF, Wang J, et al. DYNC1H1 Promotes the Proliferation and Migration of Gastric Cancer by Up-Regulating IL-6 Expression. Front Oncol 2019;9.

32. Guo C, Liu S, Wang J, Sun MZ, Greenaway FT. ACTB in cancer. Clin Chim Acta 2013;417:39-44.

33. Yang L, Yang Z, Tan X, Miao X. Expression of annexin A1 (ANXA1) and A2 (ANXA2) and its significance in benign and malignant lesions of gallbladder. Zhonghua zhong liu za zhi 2010;32:595-9.

34. Wang CY, Lin CF. Annexin A2: its molecular regulation and cellular expression in cancer development. Dis Markers 2014;2014.

35. Christensen MV, Høgdall CK, Jochumsen KM, Høgdall EV. Annexin A2 and cancer: A systematic review. Int J Oncol 2018;52:5-18.

36. Melling N, Rashed M, Schroeder C, Hube-Magg C, Kluth M, Lang D, et al. High-Level γ-Glutamyl-Hydrolase (GGH)
Expression is Linked to Poor Prognosis in ERG Negative Prostate Cancer. Int J Mol Sci 2017;18:286.

37. Shubbar E, Helou K, Kovács A, Nemes S, Hajizadeh S, Enerbäck C, et al. High levels of γ-glutamyl hydrolase (GGH) are associated with poor prognosis and unfavorable clinical outcomes in invasive breast cancer. BMC Cancer 2013;13:47.

38. Sadahiro S, Suzuki T, Tanaka A, Okada K, Saito G, Miyakita H, et al. Gene expression levels of gamma-glutamyl hydrolase in tumor tissues may be a useful biomarker for the proper use of S-1 and tegafur-uracil/leucovorin in preoperative chemoradiotherapy for patients with rectal cancer. Cancer Chemother Pharmacol 2017;79:1077-85.

39. Dong X, Han Y, Sun Z, Xu J. Actin Gamma 1, a new skin cancer pathogenic gene, identified by the biological feature-based classification. J Cell Biochem 2018;119:1406-19.

40. Po ‘uha ST, Kavallaris M. Gamma-actin is involved in regulating centrosome function and mitotic progression in cancer cells. Cell Cycle 2015;14:3908-19.

41. Iqbal MA, Siddiqui FA, Gupta V, Chattopadhyay S, Gopinath P, Kumar B, et al. Insulin enhances metabolic capacities of cancer cells by dual regulation of glycolytic enzyme pyruvate kinase M2. Mol Cancer 2013;12:72.