ALG3-CDG: a patient with novel variants and review of the genetic and ophthalmic findings

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Abstract

Background: ALG3-CDG is a rare autosomal recessive disease. It is characterized by deficiency of alpha-1,3-mannosyltransferase caused by pathogenic variants in the ALG3 gene. Patients manifest with severe neurologic, cardiac, musculoskeletal and ophthalmic phenotype in combination with dysmorphic features, and almost half of them die before or during the neonatal period.

Case presentation: A 23 months-old girl presented with severe developmental delay, epilepsy, cortical atrophy, cerebellar vermis hypoplasia and ocular impairment. Facial dysmorphism, clubfeet and multiple joint contractures were observed already at birth. Transferrin isoelectric focusing revealed a type 1 pattern. Funduscopy showed hypopigmentation and optic disc pallor. Profound retinal ganglion cell loss and inner retinal layer thinning was documented on spectral-domain optical coherence tomography imaging. The presence of optic nerve hypoplasia was also supported by magnetic resonance imaging. A gene panel based next-generation sequencing and subsequent Sanger sequencing identified compound heterozygosity for two novel variants c.116del p.(Pro39Argfs*40) and c.1060 C > T p.(Arg354Cys) in ALG3.

Conclusions: Our study expands the spectrum of pathogenic variants identified in ALG3. Thirty-three variants in 43 subjects with ALG3-CDG have been reported. Literature review shows that visual impairment in ALG3-CDG is most commonly linked to optic nerve hypoplasia.

Keywords: N-linked glycosylation, Congenital disorder of glycosylation, ALG3-CDG, Optic nerve hypoplasia, Arthrogryposis, Transferrin isoelectric focusing, Novel mutation
Background
Congenital disorders of glycosylation (CDG) are a rapidly expanding group of genetic multisystem diseases characterized by hypoglycosylation of glycoproteins and glycolipids. Based on the type of glycosidic bond, N- and O-glycosylation disorders are distinguished as two main subclasses of CDG [1].

ALG3-CDG (MIM #601,110) is a rare autosomal recessive disorder of protein N-glycosylation caused by the deficiency of alpha-1,3-mannosyltransferase, which adds one mannose residue in an alpha-1,3 linkage to Man5GlcNAc2-PP-Dol [2].

ALG3-CDG patients show various combinations of cardiac defects (obstructive cardiomyopathy, dilatation of aorta), lung hypoplasia, hepatomegaly, neurological impairment (cerebral and cerebellar atrophy, agenesia of corpus callosum, epilepsy) with developmental delay, musculoskeletal involvement (ulnar deviation, joint contractures, skeletal dysplasia, scoliosis), urogenital abnormalities (nephrocalcinosis, renal cysts, hydronephrosis), microcephaly, visual impairment and dysmorphic features (down slanting palpebral fissures, hypertelorism, high nasal bridge, anteverted nares, large and thick low-set ears with abnormal pinnae, micrognathia, thin lips, inverted nipples). Forty-two cases of ALG3-CDG have been reported to date [2–18].

In this study, we characterize in detail the first Czech patient with ALG3-CDG and provide a review of the literature on the identified disease-causing variants and reported ocular findings.

Case presentation
The study was approved by the Ethics committee of the General University Hospital in Prague and adhered to the tenets of the Helsinki Declaration.

The patient was born at 34 6/7 weeks of gestation, with a birth weight of 2.190 kg (34th percentile, -0.41 SD), length of 45 cm (46th percentile, -0.1 SD) and head circumference of 30 cm (16th percentile, -1 SD), following a pregnancy with polyhydramnion and discrete pericardial effusion. She is the first child of non-consanguineous Czech parents. Her early postnatal adaptation was uneventful (Apgar score 8-9-9), but already at birth multiple joint contractures and craniofacial dysmorphic features were noticed. The otoacoustic emissions were absent. Abdominal ultrasound documented multiple renal cysts.

At 3 months of age, the girl was admitted to the hospital with recurrent apnoeic episodes, congenital laryngeal stridor and micro-aspirations. Oral feeding was found to be risky and gastrostomy was placed. On clinical examination she presented with central hypotonia, plagiocephaly, abnormal subcutaneous fat distribution around the neck, port-wine stain on the forehead and craniofacial dysmophia (Fig. 1A, 1B). Length (55 cm, 12th percentile) and weight (4.66 kg, 39th percentile) were normal but there was severe progressive microcephaly (head circumference 34.5 cm, <1st percentile, -3.49 SD). Laboratory investigations showed antithrombin (35 %, controls 80–140 %) and factor XI deficiency (42 %, controls 55–135 %).

Neurological examination showed spastic tetraplegia, hyporeflexia and developmental delay. Although clinical seizures were not noticed, electroencephalography (EEG) was severely abnormal showing high amplitude slow spike-wave complexes (SSWC) and slow background activity. Anticonvulsant therapy was therefore initiated. The brain magnetic resonance imagining showed cerebral and cerebellar atrophy, cavum septum pellicudum, enlarged 4th ventricle and cisterna magna, delayed myelinization, and optic nerve hypoplasia.

Ophthalmic examination showed hypopigmentation of the fundus and pale optic discs. Visual and brain stem auditory evoked potentials (VEP, BAEP) were both pathological with prolonged latencies of wave N70 and waves III, V, respectively. Cardiological examination was normal apart from insignificant patent arterial duct and patent foramen ovale.

At 12 months of age, epilepsy was not compensated despite combined therapy with levetiracetam and phenobarbital. Add-on therapy with topiramate was started. At that time, although the development was severely delayed, the patient began to fix objects and to react to sounds and voices of family members, gurgle and lift up her head.

Despite gastrostomy feeding, all her anthropometric parameters severely declined with length 66 cm (<1st percentile, -2.35 SD), weight 7.38 kg (6th percentile, -1.96 SD) and head circumference 38 cm (<1st percentile, -5.17 SD). EEG at 14 months showed similar pattern to the earlier recordings with slow background activity and multiple SSWC.

At the last follow-up, the girl was 23 months-old, severe developmental delay was apparent, seizures were ameliorated on triple anticonvulsant medication. Her recurrent micro-aspirations were life threatening. She showed further deceleration of the anthropometric data: length 77 cm (<1st percentile, -2.69 SD), weight 10.2 kg (6th percentile, -1.96 SD) and head circumference 40 cm (<1st percentile, -5.53 SD). On physical examination, flexed joints, long fingers with ulnar deviation, adducted thumbs and laterally deviated toes were observed (Fig. 1C-1E).

We performed esophagogastroduodenoscopy that revealed esophagitis and duodenal ulcer. Repeated abdominal ultrasonography documented multiple cysts in both kidneys which were of stable size since birth, other abdominal organs were without abnormalities.

Ocular examination including spectral-domain optical coherence tomography (SD-OCT) (Spectralis, Heidelberg Engineering GmbH, Heidelberg, Germany) revealed
profound loss of retinal ganglion cells and inner retinal layer thinning with preservation of the external limiting membrane, ellipsoid zone and interdigitation zone (Fig. 1 F-I). Fundus appeared hypopigmented and optic discs were pale as documented using ultra-wide field camera (Clarus 700, Carl Zeiss Meditec AG, Jena, Germany). Visual activity could not be measured.

Transferrin isoelectric focusing was performed as described [19] and showed a type 1 pattern.

DNA from the proband and her parents was extracted from peripheral leukocytes. Targeted sequencing of 260 genes related to neuromuscular diseases and arthrogryposis was performed by hybrid capture enrichment (Roche NimbleGen, Pleasanton, CA, USA) and massive parallel sequencing (NextSeq, Illumina, San Diego, CA, USA) as described [20]. Sequencing data were analysed with Sequence Pilot software (JSI medical systems, Ettenheim, Germany). Conventional Sanger sequencing
was used to verify the presence of presumed causal variants and for targeted screening in first-degree relatives. The effect of one missense mutation was predicted using five computational tools: Sorting Intolerant from Tolerant (SIFT) [21], Polyphen-2 [22], Mutation Taster2 [23], M-CAP [24] and CADD [25]. The pathogenicity of the detected variants was evaluated according to the American College of Medical Genetics and Genomics (ACMG) recommendations [26].

Two ALG3 variants (NM_005787.6) c.116del p.(Pro39Argfs*40) and c.1060 C>T p.(Arg354Cys) were identified, neither of them has been reported. Segregation analysis confirmed their trans position, the c.116del was inherited from the mother while the c.1060 C>T from the father. All five prediction tools used evaluated the missense variant as deleterious or likely deleterious. Based on the available evidence both variants were classified as pathogenic according to the ACMG criteria.

**Discussion**

In this study we describe clinical, biochemical, and molecular genetic findings of an unreported ALG3-CDG patient.

### Table 1 Summary of ocular and vision-related findings in patients with ALG3-CDG

| Reference       | Age at evaluation | Clinical findings                                                                 |
|-----------------|-------------------|-----------------------------------------------------------------------------------|
| Korner [2]      | 5 y               | Coloboma of the iris, optic nerve atrophy                                         |
| Denecke [4]     | 6 y               | Rotational and horizontal nystagmus; unable to fixate, recognize objects, pupillary light reflex present, reduced amplitudes on ERG (at 6 m) |
| Pregnancy (19 w)|                   |                                                                                   |
| Schollen [5]    | 4 y               | Blindness, optic nerve atrophy                                                    |
| Sun [6]         | Shortly after birth (36 w) | Bilateral optic nerve atrophy; abnormal pupillary light reflex                     |
| Kranz [7]       | 9 y               | Cortical blindness, strabismus, decreased amplitudes of both rods and cones on ERG |
|                 | 7 y               | Strabismus; deposition of abnormal metabolic products in conjunctiva possibly indicating retinal involvement, decreased amplitudes of both rods and cones on ERG |
| Rimella-Le-Huu [8] | 15 m | Poor visual contact; latent nystagmus; hypopigmentation of the retina, optic nerve atrophy |
| Riess [9]       | 15 y              | Cortical visual impairment, divergent strabismus, myopia, mild optic nerve atrophy, normal retina, no cataracts, no nystagmus. |
|                 | 21 y              | Cortical visual impairment, strabismus, horizontal nystagmus, mild optic nerve atrophy, normal retina, no cataracts. |
| Lepais [10]     | MTP (25 w)        | Ocular proptosis, corneal opacities                                              |
|                 | Shortly after birth (36 w) | Bilateral congenital cataract                                                      |
| Fiumara [11]    | 2 y               | Initial signs of chorioretinal dystrophy, i.e., optic nerve atrophy               |
| Barba [12]      | 5 y               | Poor eye contact                                                                  |
|                 | 6 y               | Poor eye contact                                                                  |
| Alsubhi [13]    | Neonatal          | NR                                                                                |
|                 | Neonatal          | NR                                                                                |
|                 | Neonatal          | NR                                                                                |
|                 | Neonatal          | NR                                                                                |
|                 | Neonatal          | NR                                                                                |
|                 | Neonatal          | NR                                                                                |
|                 | Neonatal          | NR                                                                                |
| Himmelreich [14]| 2 m               | Descending eyelid axes, not evident fixation                                      |
|                 | 2 m               | Descending eyelid axes                                                           |
|                 | 3 y               | NR                                                                               |
|                 | 16 m              | Horizontal nystagmus and difficult fixation (noted at 3.5 m), on MRI (at 11 m) hypogenesis of the anterior optic pathways (optic nerve and chiasma), small papillae (at 11 m), able to see smaller objects |
| Bian [15]       | MTP (28 w)        | Eyelid ptosis                                                                    |
|                 | MTP (22 w)        | NR                                                                               |
| Paketci [16]    | 4.5 m             | Poor eye contact, deviations in the eyes                                           |
|                 | 2 m               | Poor eye contact                                                                  |

**Table 1 Summary of ocular and vision-related findings in patients with ALG3-CDG (Continued)**

| Reference       | Age at evaluation | Clinical findings                                                                 |
|-----------------|-------------------|-----------------------------------------------------------------------------------|
| Ferrer [17]     | Fetal demise (24 4/7 w) | NR                                                                                |
|                 | Stillbirth (30 1/7 w) |                                                                                   |
| Alsharhan [18]  | 17 y              | Strabismus, myopia, thick eyebrows and eyelashes                                  |
|                 | 5 y               | Optic nerve atrophy, epicanthal folds, long eyelashes, telecanthus                |
|                 | 2 y               | Epicanthal folds, cortical blindness, small optic nerves chiasm and tracks        |
|                 | 7 y               | Strabismus, myopia, impaired visual awareness                                     |
|                 | Stillbirth        | Down-slanting palpebral fissures                                                  |
|                 | 1 y               | Optic nerve atrophy                                                              |
|                 | 30 y              | Strabismus, epicanthal folds                                                      |
|                 | 38 y              | Down-slanting palpebral fissures                                                  |
|                 | 36 y              | Not detected                                                                      |
|                 | Neonatal          | NR                                                                                |
| Current study   | 23 m              | Poor fixation, down-slanting palpebral fissures, hypopigmented fundus, retinal ganglion cell loss, optic nerve hypoplasia |

ERG electroretinography, MRI magnetic resonance imaging, MTP medical termination of pregnancy, NR not reported, m months, w weeks, y years Ocular features reported including negative findings as provided in the original reports are shown. In most patients detailed ophthalmic examination has not been reported thus the presence of other phenotypes cannot be excluded Ophthalmic examination in patients reported by Alsharhan [19] was assumed to be done at the same time as Nijmegen Pediatric CDG Rating Scale evaluation
| Reference               | DNA change       | Protein change                              | Zygosity | gnomAD allele frequency | No of affected subjects | Origin and/or ethnicity |
|------------------------|------------------|---------------------------------------------|----------|-------------------------|-------------------------|-------------------------|
| Korner [2]             | c.353G>A         | p.(Gly118Asp)                               | HOM      | 0                       | 1                       | German                  |
| Denecke [3], Denecke [4]| c.165 C>T        | p.Val54Thrfs*13                             | HOM      | 0                       | 2                       | Italian                 |
| Schollen [5]           | c.796 C>T        | p.(Arg266Cys)                               | HOM      | 1/248,496               | 1                       | White                   |
| Sun [6]                | c.512G>A         | p.(Arg171Gln)                               | HOM      | 9/279,202               | 1                       | Dominican Republic      |
| Kranz [7]              | c.211T>C         | p.(Trp71Arg)                               | HET      | 0                       | 2                       | White                   |
| Rimella-Le-Huu [8]     | c.116 C>T        | p.(Pro39Leu)                               | HET      | 0                       | 1                       | Swiss/Italian           |
| Riss [9]               | c.206T>C         | p.Ile69Thr                                 | HET      | 0                       | 2                       | Vietnamese              |
| Lepais [10]            | c.286G>A         | p.(Gly96Arg)                               | HOM      | 5/248,964               | 2                       | Turkish                 |
| Fiumara [11]           | c.564_566del     | p.(Leu190del)                              | HET      | 0                       | 1                       | NR                      |
| Barba [12]             | c.1 A>G          | p.(Met1?)                                  | HOM      | 1/189,664               | 1                       | NR                      |
| Alsubhi [13]           | c.165 C>T        | p.Val54Thrfs*13                             | HET      | 0                       | 1                       | Turkish                 |
| Himmelreich [14]       | c.165 C>T        | p.Val54Thrfs*13                             | HOM      | 0                       | 1                       | Iraqi                   |
| Alsubhi [13]           | c.165 C>T        | p.Val54Thrfs*13                             | HOM      | 0                       | 1                       | Albanian                |
| Farolfi et al. BMC Ophthalmology (2021) 21:249 | c.165 C>T | p.Val54Thrfs*13 | HOM | 0 | 1 | Turkish |
| Lepais [10]            | c.286G>A         | p.(Gly96Arg)                               | HOM      | 5/248,964               | 2                       | Turkish                 |
| Fiumara [11]           | c.1188G>A        | p.(Trp394*)                                | HET      | 0                       | 1                       | Chinese                 |
| Bian [15]              | c.512G>T         | p.(Arg171Leu)                               | HET      | 1/247,844               | 2                       | Chinese                 |
| Paketoč [16]           | c.165 C>T        | p.Val54Thrfs*13                             | HOM      | 0                       | 2                       | NR                      |
| Alsharhan [18]         | c.566T>C         | p.(Leu219Pro)                               | HET      | 0                       | 1                       | White                   |
| Fiumara [11]           | c.796 C>T        | p.(Arg266Cys)                               | HOM      | 1/248,496               | 1                       | Ecuador                 |
| Ferrer [17]            | c.1188G>A        | p.(Trp394*)                                | HET      | 0                       | 2*                      | Pakistani               |
| Alsharhan [18]         | c.566T>C         | p.(Leu219Pro)                               | HET      | 0                       | 1                       | White                   |
| Fiumara [11]           | c.796 C>T        | p.(Arg266Cys)                               | HOM      | 1/248,496               | 1                       | Ecuador                 |
| Paketoč [16]           | c.796 C>T        | p.(Arg266Cys)                               | HOM      | 1/248,496               | 1                       | Ecuador                 |
| Alsharhan [18]         | c.991 C>T        | p.(Gln331*)                                | HET      | 0                       | 1                       | African American        |
| Paketoč [16]           | c.914 C>A        | p.(Ala305Asp)                               | HET      | 2/247,836               | 1                       | Arabic                  |
| Alsharhan [18]         | c.512G>T         | p.(Arg171Leu)                               | HET      | 9/279,202               | 1                       | Arabic                  |
| Current study          | c.116del         | p.Pro39Argfs*40                             | HET      | 0                       | 1                       | Saudi Arabian           |
| Current study          | c.1060 C>T       | p.(Arg540Cys)                               | HET      | 6/248,692               | 0                       | Czech                   |

HET heterozygous, HOM homozygous, N no, NR not reported, Y yes

*Predicted at protein level to be silent, i.e. p.(=), the variant was however shown at cDNA level to lead to deletion of 37 bp (r.160_196del) with aberrant splicing and initiation of premature termination codon

bAt cDNA level leading to exon 2 deletion (r.197_296del)

Both also carried a homozygous variant of uncertain significance c.944C>G p.(Ser315Cys) in COG5; OMIM # 6136122

The variant is listed in the Euroglycanet network database (http://www.euroglycanet.org)

Each row represents a single family. Information on ethnicity and origin is as complete as it was possible to extract from published studies. Mutation description follows Human Genome Variation Society guidelines and NM_005787.6 was taken as the reference sequence. Allele frequency was mined from gnomAD v2.1.1
We also review ocular and molecular genetic findings in all 43 individuals with ALG3-CDG identified to date.

Our case adds to the previously published phenotype data several novel aspects. In earlier reports pale or small optic disc was observed in 12 patients suggesting optic nerve atrophy or hypoplasia [2, 5, 6, 8, 9, 11, 14, 18]. Our case report is however the first clearly documenting optic nerve and retinal layers pathology by ocular imaging methods.

Clinical course
Out of the 43 reported individuals 17 had a lethal phenotype with prenatal manifestation, medical termination of pregnancy and/or early neonatal death. Three further patients died of respiratory failure during pneumonia in childhood (aged 1.8; 3.5 and 6 years) [11, 12, 14] and one of multiorgan failure (aged 1 year) [18]. Neurological symptoms have recently been summarized in 26 patients, and a ketogenic diet was suggested as a therapeutic option for intractable epilepsy [16]. Dysmorphic and musculoskeletal features have been also recently reviewed in 19 patients [15].

The longest living patient is 38-years-old male suffering from moderate intellectual disability, intractable seizures, feeding difficulties, facial dysmorphism, hearing loss, cardiac abnormalities, osteopenia/osteoporosis, nocturnal apnoea, scoliosis, hypothyroidism [18].

Ocular phenotype
Presumably because of poor compliance due to young age at examination and developmental delay available ophthalmic data in ALG3-CDG are limited (Table 1). Most often poor eye fixation, strabismus and/or nystagmus have been noted. However, the likely underlying cause was only reported in few patients and comprised optic nerve atrophy/hypoplasia observed in 11 patients including the current study confirming deficiency of retinal ganglion cells by SD-OCT, and signs of choroidal dystrophy in three patients, of these one had also optic atrophy [11]. Electoretinography was reported in three patients; decreased amplitudes of both rod and cone responses were observed [4, 7].

Less common ocular features of ALG3-CDG were congenital cataract found in one individual [10] and iris coloboma present in another patient [2]. One fetus examined after pregnancy termination at week 25 was noted to have proptosis and corneal opacities [10]. Another fetus terminated in week 28 had eyelid ptosis [15].

Disease-causing variants
Sequence variants previously reported as causing ALG3-CDG (reference sequence NM 005787.6) were searched in the literature. The population frequency of the variants was retrieved from the Genome Aggregation Database v2.1.1 from 125,748 human exomes and 15,708 genomes [27]. In total 33 pathogenic/likely pathogenic variants in ALG3 have been found in 43 individuals with ALG3-CDG from 29 families (Table 2). Most subjects carried a unique disease-causing variant(s), only three mutations have been identified recurrently. Homozygous variants were mainly linked to consanguinity when this information was available [2–18].

Conclusions
CDG pose a diagnostic challenge due to their high phenotypic variability. Combination of severe neurologic and visual impairment with dysmoria, arthrogryposis and other congenital malformations should raise suspicion of a CDG syndrome.

Abbreviations
ACMG: American College of Medical Genetics and Genomics; ALG3: Asparagine-linked glycosylation 3; BAEP: Brain stem auditory evoked potentials; CDG: Congenital disorders of glycosylation; gnomAD: Genome aggregation database; EEG: Electroencephalography; MIM: Mendelian Inheritance in Man; SD: Standard deviation; SD-OCT: Spectral-domain optical coherence tomography; SSWC: Slow spike-wave complexes; VEP: Visual evoked potentials

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Authors’ contributions
MF and AC conducted the review and wrote a substantial part of the manuscript including the figure and tables. TH, MF, BK, and PL examined the patient. TH and PL contributed to the manuscript with advice and critical comments, supervised the manuscript preparation. HH and NO performed the biochemical analyses. JZ did the molecular genetic study. The manuscript was reviewed and revised by all the authors. The author(s) read and approved the final manuscript.

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Availability of data and materials
All data generated or analysed during this study are included in this published article.

Declarations
Ethics approval and consent to participate
The study was approved by the Ethics committee of the General University Hospital in Prague (reference no. 34/19; date 23.5.2019). Written informed consent was obtained from the patient’s parents before inclusion.

Consent for publication
We obtained from the patient’s parents written informed consent for publication of this Case report and any accompanying images. A copy of the written consent is available for review by the Editor of this journal.

Competing interests
The authors declare that they have no competing interests.
epilepsy with ketogenic diet therapy in twins with ALG3-CDG. Brain Dev 2020, 42(7):539–545.

17. Ferrer A, Starosta RT, Ranatunga W, Unger D, Kozicz T, Klee E, Rust LM, Wick M, Morava E: Fetal glycosylation defect due to ALG3 and CDG variants detected via amniocentesis: Complex glycosylation defect with embryonic lethal phenotype. Mol Genet Metab 2020, 131(4):424–429.

18. Alsharhan H, Ng BG, Daniel EJ, Friedman J, Pivnick EJ, Al-Hashem A, Faqeeh EA, Liu P, Engelhardt NM, Keller KN et al: Expanding the phenotype, genotype and biochemical knowledge of ALG3-CDG. J Inherit Metab Dis 2021.

19. Guiflard M, Wada Y, Hansikova H, Yuasa I, Vesela K, Ondrušková N, Kadoya M, Janssen A, Van den Heuvel UP, Morava E et al: Transferin mutations at the glycosylation site complicate diagnosis of congenital disorders of glycosylation type I. J Inherit Metab Dis 2011, 34(4):901–906.

20. Stehlikova K, Skalova D, Zidkova J, Haberlova J, Vohanka S, Mazanec R, Mrazova L, Vondracek P, Olejskova H, Zamecnik J et al: Muscular dystrophies and myopathies: the spectrum of mutated genes in the Czech Republic. Clin Genet 2017, 91(3):463–469.

21. Sm NL, Kumar P, Hu J, Henikoff S, Schneider G, Ng PC: SFT: web server: predicting effects of amino acid substitutions on proteins. Nucl Acid Res 2012, 40(Web Server issue):W452–457.

22. Adzhubei IA, Schmidt S, PESHKIN L, Ramensky VE, Gerasimova A, Bork P, Kondrashov AS, Sunyaev SR: A method and server for predicting damaging missense mutations. Nat Methods 2010, 7(4):248–249.

23. Schwarz JM, Cooper DN, Schuelke M, Seelow D: MutationTaster2: mutation prediction for the deep-sequencing age. Nat Methods 2014, 11(4):361–362.

24. Jagadeesh KA, Wenger AM, Berger MJ, Guturu H, Stenson PD, Cooper DN, Bernstein JA, Bejerano G: M-CAP eliminates a majority of variants of uncertain significance in clinical exomes at high sensitivity. Nat Genet 2016, 48(12):1581–1586.

25. Rentzsch P, Witten D, Cooper GM, Shendure J, Kircher M: CADP: predicting the deleteriousness of variants throughout the human genome. Nucl Acid Res 2019, 47(D1):D886–D894.

26. Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, Grody WW, Hegde M, Lyon E, Spector E et al: Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med 2015, 17(5):402–404.

27. Karczewski KJ, Francioli LC, Tiao G, Cummings BB, Affoldi J, Wang Q, Collins RL, Larichia KM, Ganna A, Bimbam DP et al: The mutational constraint spectrum quantified from variation in 141,456 humans. Nature 2020, 581(7809):434–443.

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