Bone mass, body composition and vitamin D status of ARV-naïve, urban, black South African women with HIV infection, stratified by CD4 count

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Abstract
Summary This is the first report examining vitamin D status and bone mass in African women with HIV infection using dual-energy X-ray absorptiometry (DXA) with an appropriate HIV-negative control group. Unlike previous publications, it demonstrates no difference in bone mineral density (BMD) or vitamin D status in HIV-positive patients, at different disease stages, vs. HIV-negative subjects.

Introduction Low bone mass and poor vitamin D status have been reported among HIV-positive patients; suggesting HIV or its treatment may increase the risk of osteoporosis, a particular concern for women in countries with high HIV prevalence such as South Africa. We describe bone mass and vitamin D status in urban premenopausal South African women, who were HIV positive but not on antiretroviral therapy (ARV).

Methods This study is a cross-sectional measurement of BMD and body composition by DXA and vitamin D status by serum 25-hydroxyvitamin D (25(OH)D) concentration. Subjects were recruited into three groups: HIV negative (n=98) and HIV positive with preserved CD4 cell count (non-ARV; n=74) or low CD4 cell counts prior to ARV initiation (pre-ARV; n=75).

Results The mean (standard deviation (SD)) age of women was 32.1 (7.2) years. Mean CD4 (SD) counts (×10⁶/l) were 412 (91) and 161 (69) in non-ARV and pre-ARV groups (p<0.0001). Pre-ARV women were significantly lighter and had lower mean BMI than the other two groups (p<0.002). The pre-ARV group also had significantly less fat and lean mass compared with non-ARV and HIV-negative subjects (p≤0.05). After full adjustment, there were no significant differences in BMD at any site (p>0.05) between the groups, nor was vitamin D status significantly different between groups (p>0.05); the mean (SD) cohort 25(OH)D being 60 (18) nmol/l.

Conclusion Contrary to previous studies, these HIV-positive women did not have lower BMD or 25(OH)D concentrations than HIV-negative controls, despite the pre-ARV group being lighter with lower BMI.

Keywords Body composition · Bone mineral density · Dual energy X-ray absorptiometry · HIV infection · Vitamin D

Introduction

HIV infection and the use of antiretroviral (ARV) medication have been associated with low bone mineral density (BMD) and poor vitamin D status. In a meta-analysis, the prevalence of low BMD in HIV-positive individuals was three times higher than in HIV-negative controls [1–3]. Similarly, studies have described high prevalence of low 25-hydroxyvitamin D
(25(OH)D) concentrations in HIV-positive patients [4]. Some studies of the effects of HIV and/or its treatment on bone are limited by retrospective design, a preponderance of white, male subjects, and lack of HIV-negative controls [5] while others are prospective [6] and do include women [7, 8]. Other studies are limited by confounding by low body weight or other risk factors for low BMD, such as intravenous drug use (IDU), exposure to a large variety of ARV regimes and measurement of BMD and vitamin D status after varying duration of ARV exposure [6]. The few prospective studies focusing on women have also been limited by some of these aspects [6, 9], and as a result it is difficult to ascertain with certainty if HIV infection and/or its treatment or factors unrelated to HIV infection are contributing factors to the low bone mass and low vitamin D status described in the current literature. In contrast, there are data to suggest that after adjusting for body weight, BMD is normal or near normal, and that patients on ARV do not have increased rates of bone loss [10, 11]. As a result, there is not a definitive consensus on the contribution of HIV infection or ARV exposure on BMD in infected individuals.

In South Africa, estimates of HIV prevalence for 2010 are 10.5 % for the total population and 29.3 % for women attending antenatal clinics. The epidemic is described as “hyperendemic” because of the high prevalence and continuing drivers of transmission [12–14]. In South Africa, individuals generally become eligible for ARV treatment when their CD4 count is less than a nationally specified threshold. By 2009, 56 % of those requiring ARV were able to receive them, with the government intending to increase ARV coverage to 80 % by 2011 [12].

Vitamin D has well-known associations with bone health via its role in calcium and phosphate homeostasis, and vitamin D status is considered an important modulator of immune function by some authors [14–16]. In South Africa, adults are largely dependent on the cutaneous synthesis of vitamin D to maintain vitamin D status, as only small amounts of vitamin D are obtained from the diet due to limited food fortification. In Johannesburg (26° S latitude), there is sufficient ultraviolet B (UVB) radiation in sunshine throughout the year for dermal synthesis of vitamin D [17]. Nevertheless, vitamin D deficiency has been described in tropical/subtropical countries despite the potential for adequate skin exposure to UVB-containing sunshine [18].

The aim of the study presented here was to describe BMD, body composition and vitamin D status in South African women with and without HIV infection, prior to a planned longitudinal study of this cohort to chart the changes in these outcomes over time. We hypothesised that HIV-positive women with low CD4 counts, below the threshold that would make them eligible for ARV treatment, would have lower bone mass, less fat and muscle mass and inferior vitamin D status than HIV-positive women with preserved CD4 counts and HIV-negative women in South Africa.

Methods

Subjects

Urban, black, premenopausal, South African women (n=247) were recruited from clinics in Soweto, Greater Johannesburg and enrolled into the study between February and July 2010. Subjects were recruited from a voluntary counselling and testing clinic and local health clinics. The aim was to recruit 95 HIV-negative and 73 (±10) in each of two HIV-positive groups (with or without low CD4 counts). This sample size was based on calculations for the longitudinal study to detect a 2 % change in lumbar spine BMD, allowing for a between-individual coefficient of variation in BMD of 5 %, with 95 % confidence and 80 % power. For the study presented here, this sample size was sufficient for a comparison of three groups to allow the detection of mean differences between each pair of groups of around 0.4 standard deviation (SD) at 5 % significance and 80 % power. The study was approved by the University of the Witwatersrand Human Research Ethics Committee (HREC number: M101525) and the Gauteng Department of Health.

Eligible subjects were adult females (defined as aged greater than 18 years) and premenopausal (defined as regular menses). Other inclusion criteria included a documented negative HIV test within the last 12 weeks for HIV-negative women and a documented positive HIV test for all other women. Patient-retained clinic records were scrutinised whenever possible to confirm medical history, current CD4 count, prior exposure to ARVs and concurrent medication use. Exclusion criteria included conditions associated with abnormal bone metabolism or current use of medication likely to affect bone or vitamin D status such as bisphosphonates. Pregnant and lactating women were excluded as were those with an acute medical condition. The group with the lowest CD4 count were largely recruited after the other groups: May to June and February to April, respectively.

Study posters were displayed in the clinic and training sessions undertaken with clinic staff. Women who expressed an interest in the study underwent initial telephone screening, in their language, to ensure inclusion and exclusion criteria were met. Prior to enrolment, potential subjects completed a medical- and health-related questionnaire to assess past and current health status and medication use and to further assess compliance with inclusion and exclusion criteria. Women who remained eligible were enrolled in the full study after they had provided written consent.

The enrolled women consisted of HIV-negative (n=98) and HIV-positive (n=149) subjects. The HIV-positive women were recruited into two prespecified groups: those with relatively preserved CD4 counts (>350×10⁶ cells/l), not requiring ARV therapy (non-ARV group; n=74) and those with low CD4 counts (in the region of 200×10⁶ cells/l).
requiring ARV initiation (pre-ARV group; \( n = 75 \)) according to the current South Africa (SA) treatment guidelines [19]. HIV-negative status was confirmed using the Determine™ rapid HIV-antibody test (Alere San Diego, Inc., San Diego, CA, USA), while HIV-positive status was established using a second platform. HIV-positive women were either newly diagnosed or known to be HIV positive, but not on ARVs. All HIV-positive women provided an up-to-date (within 3 months) \( CD_4 \) count prior to enrolling into the study. All HIV-positive women were either newly diagnosed or known to be HIV positive, but not on ARVs. All HIV-positive women provided an up-to-date (within 3 months) \( CD_4 \) count prior to enrolling into the study. All HIV-positive women received SA standard of care with respect to ARV provision and clinical follow up. Women requiring urgent ARV initiation were managed in such a way that there would be no delay in ARV initiation if they were to participate in the study.

Women attended the Developmental Pathways for Health Research Unit (DPHRU) facility at the Chris Hani Baragwanath Academic Hospital, after an overnight fast and underwent phlebotomy, anthropometry, and dual-energy X-ray absorptiometry (DXA) assessment of bone mass and body composition. After phlebotomy, subjects were given breakfast and each received ZAR 50.00 (=US$6.25) for travel expenses.

Anthropometry

Height was measured to the nearest millimetre using a stadiometer (Holtain, Crosswell, UK). Weight was measured to the nearest 100 g using a digital scale (Tanita, TBF-410 MA Body Composition Analyzer, Tanita Corporation of America, Inc., Arlington Heights, IL, USA) with participants wearing light clothing and no shoes. BMI was calculated as the participant’s weight in kilograms divided by the square of their height in metres (in kilogram per square metre). Underweight, normal, overweight, and obese were defined as BMI <18.5, 18.5–24.9, 25–29.9, \( \geq 30.0 \) kg/m\(^2\), respectively [20].

Bone absorptiometry and body composition measurements

DXA was performed using a Hologic QDR 4500A DXA (model: Discovery W (S/N 71201) software version 12.5:7 Hologic, Inc., Waltham, MA, USA) according to standard procedures. Scans were conducted using the automatic scan mode, i.e. ‘array’, ‘fast array’ or ‘slow array’, depending on the weight of the subjects. Subjects wore light clothing having removed metal objects, jewellery, etc. DXA was used to measure bone mineral content (BMC, in grams), bone area (BA, in square centimetre) and areal BMD (in grams per square centimetre), of whole body (WB), total hip (TH), femoral neck (FN) and lumbar spine (LS). The coefficients of variation (CV\%) for repeated measurements of the manufacturer’s phantom were 0.3, 0.4 and 0.2 % for BA, BMC and BMD, respectively. The CV for repeated measurement by the DXA operator of the LS and TH BMD were 0.7 and 1.0 %, respectively. DXA scans for WB were analysed using WB less head (WBLH) as many women wore wigs and hair weaves that could not be removed prior to scanning. This artificial hair was of similar density to soft tissue and therefore could cause measurement artefact. Total fat and lean body mass (in grams) were also measured by DXA.

Laboratory analysis

Blood was collected for 25(OH)D analysis, measured by chemi-luminescent immunoassay (Liason) kit (DiaSorin Inc., Stillwater, MN, USA). The blood samples were allowed to clot for a minimum of 20 min at room temperature, and the serum was aliquoted and stored at \(-20 \) C until analysed. All samples were run in duplicate. The inter-assay CV for low and higher 25(OH)D controls was 10 and 9 %, respectively, whereas the intra-assay CV was 8 and 6 %, respectively. The DPHRU laboratory participates in the International Vitamin D External Quality Assessment Scheme and holds the certificate of proficiency [21].

Statistical analysis

Data were analysed using DataDesk 6.3.1 (Data Description Inc, Ithaca, NY, USA) and summary statistics were documented as mean (SD) or median (interquartile range), depending on the distribution. Comparisons were made between the three groups of women using hierarchical linear models; ANOVA (or ANCOVA) and Scheffé post hoc tests were used to compare group means (standard error (SE)). To consider the possible influence of group differences in bone and body size, bone mineral data were adjusted for age, weight, height and bone area, and bone area was adjusted for age, weight and height, using ANCOVA [16]. Preliminary plots of the relationship between fat mass and lean mass in this sample population demonstrated non-linearity. Regression of fat mass on lean mass in the HIV-negative control group with data in natural logarithms gave a power exponent 2.05±0.14. Consequently, a fat mass-to-lean square mass term was used to describe differences in body composition between the groups, and logarithmic regression was used to adjust fat mass for lean mass in statistical models. BMD SD scores (SDS) were generated using HIV-negative subjects as the reference population (ref) against which the SDS for each individual HIV-positive woman (i) was derived as follows: \([\text{BMD}_i – \text{mean BMD}_{\text{ref}}/\text{SD}_{\text{ref}}]\). A \( p \) value of \( \leq 0.05 \) was considered to be statistically significant.
Results

Subject characteristics

By design, the mean CD4 count \( \times 10^6 \) cells/l in the pre-ARV group was significantly lower than that in the non-ARV group (412 (91) and 161 (69), respectively, \( p < 0.0001 \)). The mode of acquisition of HIV-infection was via heterosexual transmission in all subjects, only one subject reporting IDU in the past (Table 1).

Mean age (SD) was 32.1 (7.2) years with HIV-negative women being significantly but only slightly younger than both groups of HIV-positive women. The age ranges were similar in the three groups (18–49, 22–48 and 19–47 years in HIV-negative, non-ARV and pre-ARV women, respectively). Median (IQR) gravidity was 2 (1; 3) with both HIV-positive groups having a higher median gravidity compared to the HIV-negative group.

Anthropometry and body composition

HIV-negative women tended to be shorter than both groups with HIV-infection \( p = 0.06 \), while HIV positive, pre-ARV women were significantly lighter than the other two groups \( p < 0.05 \). Median (IQR) BMI of the study cohort was 26.1 (22.4; 31) kg/m\(^2\) with BMI in pre-ARV women being significantly lower than in HIV-negative and non-ARV women. Combined overweight and obesity represented 65, 65 and

Table 1 Subject characteristics, anthropometric measurements and vitamin D status as measured by serum 25(OH)D

|                      | Group 1 HIV-negative \( n = 98 \) | Group 2 HIV-positive, non-ARV \( n = 74 \) | Group 3 HIV-positive, pre-ARV \( n = 75 \) | Group effect ANOVA \( p \) |
|----------------------|----------------------------------|---------------------------------|---------------------------------|------------------|
| Age (years)          | 30.0 (8.1)                       | 33.5 (6.1)\(^a\)                | 33.4 (6.5)\(^a\)                | 0.001            |
| HIV status           | Negative                         | Positive                        | Positive                        |                  |
| Current CD4 count \( \times 10^6 \) cells/l | ND                              | 412 (91)                        | 161 (69)\(^b\)                | <0.001           |
| Median (IQR)         | NA                               | 420 (127;409)                   | 175 (120;165)                   |                  |
| Min                  | NA                               | 240                             | 18                              |                  |
| Max                  | NA                               | 604                             | 275                             |                  |
| Gravidity median (IQR) | 1 (0;2)                         | 2 (2;3)\(^a\)                  | 2 (1;3)\(^a\)                  |                  |
| Range                | 0–5                             | 0–6                             | 0–6                             |                  |
| Current hormonal contraceptive use (%) | 34 (35.4)                       | 26 (36.6)                       | 25 (33.3)                       | 0.9              |
| Current smoking (%)  | 10.2                             | 13.5                            | 8                               | 0.2              |
| Height (cm)          | 157.6 (5.9)                      | 159.4 (5.9)                     | 159.2 (5.3)                     | 0.06             |
| Weight (kg)          | 69.7 (17.0)                      | 72.0 (17.4)                     | 62.3 (15.2)\(^cd\)            | <0.001           |
| BMI (kg/m\(^2\)) Median (IQR) | 27.3 (23.1;31.7)             | 27.8 (23.3;32.3)                | 23.5 (20.5;27.0)\(^de\)       | <0.001           |
| Overweight BMI >24.9 kg/m\(^2\), <30 kg/m\(^2\) (%) | 35                             | 28                              | 28                              | 0.001            |
| Obese BMI >30 kg/m\(^2\) (%) | 30                             | 37                              | 16                              |                  |
| Underweight BMI <18.5 kg/m\(^2\) (%) | 4                              | 1                               | 11                              |                  |
| WBLH Fat (kg)        | 26.1 (11.5)                      | 26.1 (9.8)                      | 19.7 (9.3)\(^bc\)             | <0.0001          |
| WBLH Lean (kg)       | 38.3 (60.8)                      | 39.5 (62.4)                     | 36.4 (48.1)\(^d\)            | 0.005            |
| Fat/lean\(^2\) (kg/kg\(^2\))* | 17.32 (4.80)                   | 15.92 (4.56)                    | 14.58 (5.47)\(^ef\)         | 0.002            |
| 25(OH)D (nmol/l)     | 59.7 (16.5)                      | 59.2 (16.5)                     | 61.6 (22.3)                     | 0.7              |
| 25(OH)D (nmol/l) >50 (%) | 73.5                          | 70.3                             | 66.7                            |                  |
| 25(OH)D (nmol/l) <50 (%) | 26.5                          | 29.7                             | 33.3                            |                  |
| 25(OH)D (nmol/l) <25 (%) | 1.0                             | 2.7                              | 5.3                             |                  |

All values are mean (SD) unless indicated. Letters are used to indicate significance of between-group differences as tested by ANOVA/Scheffé 25(OH)D 25 hydroxyvitamin D, ARV antiretroviral therapy, cm centimetres, IQR interquartile range, kg kilograms, SD standard deviation, WBLH whole body less head, ND not determined, NA not applicable

*Value multiplied by 1,000 to illustrate the relative differences in kilogram

\(^a\) Significantly different from group 1, \( p \leq 0.01 \)
\(^b\) Significantly different from group 2, \( p \leq 0.001 \)
\(^c\) Significantly different from group 1, \( p \leq 0.05 \)
\(^d\) Significantly different from group 1, \( p \leq 0.05 \)
\(^e\) Significantly different from group 2, \( p \leq 0.01 \)
\(^f\) Significantly different from group 1, \( p \leq 0.001 \)

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44% of subjects in HIV-negative, non-ARV and pre-ARV women, respectively, while underweight was present in 4, 1 and 11%, respectively (Table 1).

There were significant differences in fat mass between groups with pre-ARV women having significantly lower fat mass than non-ARV women ($p \leq 0.001$). Although lean mass was also lower in pre-ARV compared with non-ARV women ($p=0.005$) the pre-ARV group had lower fat mass-to-lean square mass ratio than the other two groups ($p=0.002$). When fully adjusting for lean mass using logarithmic regression, the pre-ARV group had significantly lower fat mass for their lean mass than the other two groups; such that for each unit of lean mass the pre-ARV group had a mean difference (SE) of 21 (5)% less fat than the controls, $p=0.0002$, and 16 (5)% less fat than the non-ARV group, $p=0.02$.

Bone measures

No significant differences in BMD at the TH, FN, LS and WBLH were found, and age and size adjustment did not reveal any differences between groups. When expressed as SD scores, there were no significant differences between pre-ARV and non-ARV groups in BMD for any site measured ($p>0.05$) and all the mean values were within a −0.5 SD of the HIV-negative reference group (Table 2). In addition, no significant differences were found in BMC values except at WBLH when fully adjusted for age, size and BA ($p=0.03$). Unadjusted BA was significantly greater in both groups of HIV-positive women than HIV-negative women at some sites but these differences disappeared after adjusting for age and size (see Electronic supplementary material (ESM) for BA and BMC data).

Vitamin D status

Mean (SD) 25(OH)D for the whole cohort was 60.1 (18.4) nmol/l and there were no significant differences between groups ($p>0.05$). 25(OH)D concentration was <50 nmol/l in 29.6% of individuals; with similar proportions in each of the groups in this category (26.5, 29.7 and 33.3% in HIV-negative, non-ARV and pre-ARV, respectively). Very few subjects had a 25(OH)D concentration <25 nmol/l (1.0, 2.7 and 5.3% in the three groups, respectively), despite the slightly greater number of pre-ARV subjects whose blood samples for 25(OH)D measurement were obtained during the winter months.

Discussion

The aim of this study was to determine whether South African HIV-positive women with preserved CD4 counts differed from those with low CD4 counts making them eligible for ARV and to compare each group with HIV-negative women. In this group of urban, South African women, pre-ARV women were significantly lighter than HIV-negative and non-ARV subjects and had lower fat mass than expected for their lean mass, raising the possibility that women with advancing HIV disease preferentially lose fat rather than lean mass. There were no significant differences between groups in BMC or BMD at any site before or after adjustment for age, BA, weight and height and the observed smaller BA in the HIV-negative women disappeared after adjustment for age, height and weight. There was no significant difference in vitamin D status between groups with the majority of subjects having a serum concentration >50 nmol/l.

The assessment of ‘optimal’ vitamin D status is problematic because varying cut-offs are used to define sufficiency, insufficiency and deficiency [22]. A concentration below 25 nmol/l is generally recognised as indicating an increased risk of rickets and osteomalacia [23]. The 2010 Institute of Medicine report considered that a blood 25(OH)D concentration of 20 ng/mL (50 nmol/l) to be sufficient for good bone health in ‘practically all individuals’ [24]. However, it noted that evidence was lacking to make a similar statement regarding non-skeletal health. In the context of HIV infection and ARV use, the optimal vitamin D status remains undefined because there

Table 2 BMD of the three groups of South African women

| BMD (g/cm²) | Group effect * | p |
|------------|----------------|---|
| Mean (SD)  |                |    |
|            | Group 1 HIV-negative $n=98$ | Group 2 HIV-positive, non-ARV $n=74$ | Group 3 HIV-positive, pre-ARV $n=75$ |
| Total Hip  | 1.013 (0.131)  | 0.985 (0.124)  | 0.988 (0.125)  | 0.3 |
| Femoral Neck | 0.930 (0.114) | 0.916 (0.125) | 0.923 (0.131) | 0.8 |
| Lumbar Spine | 1.018 (0.118) | 1.021 (0.109) | 1.006 (0.128) | 0.7 |
| WBLH | 0.958 (0.079) | 0.943 (0.071) | 0.947 (0.080) | 0.4 |

* Group effect by ANOVA. There were no significant differences between pairs of groups by Scheffé post hoc tests.

ARV antiretroviral therapy, BMD bone mineral density (in gram per square centimetre), SD standard deviation, WBLH whole body less head.
may be different requirements for maximal bone health and immune functioning compared with HIV-negative populations. However, in contrast to other reports [4, 25], in our study, there were no indications that HIV infection was associated with inferior vitamin D status because there were no significant differences in vitamin D status between the three groups, the distributions of 25(OH)D concentration were similar, and vitamin D status appeared to be generally adequate with very few women having a concentration <25 nmol/l.

Contrary to previous reports [9], we found no significant differences in BMD between either group of HIV-positive and HIV-negative women. Full adjustment for bone and body size did not alter these results. This lack of any differences is surprising as HIV-positive women with low CD4 counts, requiring ARV initiation, were significantly lighter, with lower fat and lean mass, than the other women. However, it may reflect the selection criteria for this study because despite recruiting women with low CD4 counts, of clinical concern, women with severe clinical disease received immediate ARV therapy and were thus excluded from the study. It may also be influenced by the fact that the subjects were not intravenous drug users and thus not exposed to the additional effect on BMD that this poses. Another limitation may be that the groups were different in terms of duration of hormonal contraception use, parity and total duration of lactation; however, at the time of the study, no women were pregnant or lactating. The findings are also limited by the fact that the sample of HIV-positive women was likely to be heterogeneous with respect to immune status and duration of infection. However, most other studies have also recruited HIV-positive subjects in a similar manner and this is unlikely to account for the different findings in our study.

The rates of combined overweight and obesity 65 % in HIV-negative and non-ARV subjects in this study were greater than the national average in South Africa of 51.5 % [26]; even women with advanced HIV-disease (pre-ARV group) had a combined overweight and obesity rate of 44 %. It is possible, therefore, that the typically high weight of South African women has a sparing effect on bone in those with HIV infection, even with CD4 counts below the threshold for initiation of ARV intervention.

Historically, being overweight has been viewed as protective against osteoporotic fracture, although evidence is emerging that overweight and obesity may be a risk factor for leg fragility fractures in women [27]. In the study population of younger black women in South Africa, there were no significant differences in BMD SD score, expressed relative to the HIV-negative group, according to HIV status at any site. The effects of HIV and its treatment on fracture risk in South Africa are unknown.

The lack of difference between the groups which is at variance from previously reported studies may be the result of true lack of effect of HIV infection or reflect important differences in bone response to HIV between black Africans and Caucasians. The study design in which two distinct groups of HIV-positive women, based on South African eligibility criteria for ARV treatment plus the inclusion of a HIV-negative control group strengthens the finding that HIV infection with varying degree of immunosuppression does not appear to be driving alterations in BMD or vitamin D status in these young, urban women. The high rates of overweight may be masking more dramatic differences in BMD and vitamin D in those subjects with advanced clinical HIV disease not included in this study. Further work is required to address the effects of ARV exposure on bone and vitamin D status as well as the relative effect of ‘traditional’ osteoporosis risk factors in this population. The data from this study provide an insight into bone health, body composition and vitamin D status in African women living with HIV. They challenge our own hypotheses and previously reported differences in BMD and vitamin D status in HIV-positive subjects living in developed countries and highlight the importance of studying subjects prior to ARV exposure.

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References

1. Brown TT, McComsey GA (2006) Osteopenia and osteoporosis in patients with HIV: a review of current concepts. Curr Infect Dis Rep 8(2):162–170
2. Brown TT, Qaqish RB (2006) Antiretroviral therapy and the prevalence of osteopenia and osteoporosis: a meta-analytic review. AIDS 20(17):2165–2174
3. Brown TT et al (2004) Reduced bone mineral density in human immunodeficiency virus-infected patients and its association with increased central adiposity and postload hyperglycemia. J Clin Endocrinol Metab 89(3):1200–1206
4. Welz T et al (2010) Efavirenz is associated with severe vitamin D deficiency and increased alkaline phosphatase. AIDS 24(12):1923–1928
5. Bonjoch A et al (2010) High prevalence of and progression to low bone mineral density in HIV-infected patients: a longitudinal cohort study. AIDS 24(18):2827–2833
6. Dolan SE, Kanter JR, Grinspoon S (2006) Longitudinal analysis of bone density in human immunodeficiency virus-infected women. J Clin Endocrinol Metab 91(8):2938–2945
7. Yin M et al (2005) Bone mass and mineral metabolism in HIV+ postmenopausal women. Osteoporos Int 16(11):1345–1352
8. Arnsten JH et al (2006) HIV infection and bone mineral density in middle-aged women. Clin Infect Dis 42(7):1014–1020
9. Dolan SE et al (2004) Reduced bone density in HIV-infected women. AIDS 18(3):475–483
10. Bolland MJ et al (2007) Low body weight mediates the relationship between HIV infection and low bone mineral density: a meta-analysis. J Clin Endocrinol Metab 92(12):4522–4528
11. Bolland MJ et al (2007) Bone mineral density remains stable in HAART-treated HIV-infected men over 2 years. Clin Endocrinol (Oxf) 67(2):270–275
12. Republic of South Africa. Country progress report on the declaration of commitment on HIV/AIDS 2010. Report – reporting period: January 2008 - December 2009. http://data.unaids.org/pub/report/2010/southafrica_2010_country_progress_report_en.pdf
13. Statistics South Africa (2010) Mid-year population estimates 2010: Pretoria South Africa. p. 1–16
14. Adams JS et al (2007) Vitamin D in defense of the human immune response. Ann N Y Acad Sci 1117:94–105
15. Conesa-Botella A et al (2009) Is vitamin D deficiency involved in the immune reconstitution inflammatory syndrome? AIDS Res Ther 6:4
16. Liu PT et al (2006) Toll-like receptor triggering of a vitamin D-mediated human antimicrobial response. Science 311(5768):1770–1773
17. Pettifor JM, Ross FP, Solomon L (1978) Seasonal variation in serum 25-hydroxycholecalciferol concentrations in elderly South African patients with fractures of femoral neck. Br Med J 1(6116):826–827
18. Schoenmakers I, Goldberg GR, Prentice A (2008) Abundant sunshine and vitamin D deficiency. Br J Nutr 99(6):1171–1173
19. National Department of Health South Africa (2010) Clinical guidelines for the management of HIV & AIDS in adults and adolescents. http://www.sahivsoc.org/upload/documents/Clinical_Guidelines_for_the_Management_of_HIV_AIDS_in_Adults_Adolescents_2010.pdf
20. WHO (2006) W.H.O. BMI classification
21. Poopedi MA, Norris SA, Pettifor JM (2011) Factors influencing the vitamin D status of 10-year-old urban South African children. Public Health Nutr 14(2):334–339
22. Prentice A, Goldberg GR, Schoenmakers I (2008) Vitamin D across the lifecycle: physiology and biomarkers. Am J Clin Nutr 88:500S–506S
23. Scientific Advisory Committee on Nutrition (2007) Update on vitamin D. Norwich: TSO (The Stationery Office)
24. Institute of Medicine (2010) Dietary reference intakes for calcium and vitamin D: National Academies Press
25. Van Den Bout-Van Den Beukel CJ et al (2008) Vitamin D deficiency among HIV type 1-infected individuals in the Netherlands: effects of antiretroviral therapy. AIDS Res Hum Retroviruses 24(11):1375–1382
26. Kruger HS et al (2011) Overweight among children decreased, but obesity prevalence remained high among women in South Africa, 1999–2005. Public Health Nutr 2012 Apr;15(4):594–9
27. Compston JE et al (2011) Obesity is not protective against fracture in postmenopausal women: GLOW. Am J Med 124(11):1043–1050