Indeterminate thyroid cytology: detecting malignancy using analysis of nuclear images

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Abstract

**Background:** Thyroid nodules diagnosed as “Atypia of Undetermined Significance/Follicular Lesion of Undetermined Significance” (AUS/FLUS) or “Follicular Neoplasm/Suspected Follicular Neoplasm” (FN/SFN), according to Bethesda’s classification, represent a challenge in clinical practice. Computerized analysis of nuclear images (CANI) could be a useful tool for these cases. Our aim was to evaluate the ability of CANI to correctly classify AUS/FLUS and FN/SFN thyroid nodules for malignancy. **Methods:** We studied 101 nodules cytologically classified as AUS/FLUS (n=68) or FN/SFN (n=33) from 97 thyroidectomy patients. Slides with cytological material were submitted to manual selection and analysis of the follicular cell nuclei for morphometric and texture parameters using ImageJ software. The histologically benign and malignant lesions were compared for such parameters which were then evaluated for the capacity to predict malignancy using the *Classification and Regression Trees* Gini model. The Intraclass Coefficient of Correlation was used to evaluate method reproducibility. **Results:** In AUS/FLUS nodule analysis, the benign and malignant nodules differed for Entropy (p<0.05), while the FN/SFN nodules differed for Fractal analysis, coefficient of variation (CV) of roughness, and CV-Entropy (p<0.05). Considering the AUS/FLUS and FN/SFN nodules separately, it correctly classified 90.0% and 100.0% malignant nodules, with a correct global classification of 94.1% and 97%, respectively. We observed that reproducibility was substantially or nearly complete (0.61-0.93) in 10 of the 12 nuclear parameters evaluated. **Conclusion:** CANI demonstrated an high capacity for correctly classifying AUS/FLUS and FN/SFN thyroid
nodules for malignancy. This could be a useful method to help increase diagnostic accuracy in the indeterminate thyroid cytology.

**Introduction**

Thyroid cancer is the commonest endocrine neoplasia and its frequency is increasing (1). The method of choice for diagnosing these lesions is fine needle aspiration (FNA), with cytological analysis performed using the Bethesda Classification System (2). Despite this system has proven extremely valid and reliable (3), a considerable number of cases still have indeterminate cytological diagnosis. Of these “Atypia of Undetermined Significance/Follicular Lesion of Undetermined Significance (AUS / FLUS)” and “Follicular Neoplasm/Suspicious for a Follicular Neoplasm (FN / SFN)” categories are of particular concern as they are relatively frequent and widely vary in terms of malignancy prediction. In fact, malignancy rates of between 5% and 81% for AUS / FLUS and 11% and 65.7% for FN / SFN nodules have been reported (4).

Thus, studies using techniques with varying degrees of complexity and cost have been conducted to clarify the nature of AUS / FLUS and FN / SFN thyroid lesions, such as those using molecular techniques, for example (5). However, this methodology is not yet available to most populations. Thus, more accessible diagnostic methods have been studied, such as those that use morphological aspects. The presence of nuclear atypia, for example, has proven significant in differentiating between benign and malignant tumors (4) and represents a good indicator of malignancy in nodules with indeterminate cytological diagnosis (6). The problem emerging from this approach is
heterogeneity of the criteria used to define the presence of nuclear atypia (6), as well as other morphological parameters, and the expected inter-observer variation.

In this sense, the evaluation of nuclear morphometric and textural parameters by computerized image analysis has been successfully used in tumor tissues from different neoplasias, including thyroid (7), proving to be an accessible, reproducible and low-cost tool. Although other studies have used this technique on thyroid cytological material (8, 9, 10, 11, 12, 13), in general, they have not focused on the indeterminate category. In addition, sample numbers have been modest and some of the methodologies are too complex and costly (14).

We hypothesized that computerized analysis of nuclear images (CANI) is able to select cytologically indeterminate lesions for malignancy. The aim of this study was to evaluate the ability of CANI to correctly classify cytological material from AUS / FLUS and FN / SFN thyroid nodules for malignancy.

Materials and methods

Design, patients and histological/cytological material processing

This cross-sectional study evaluated cytological material from AUS / FLUS and FN / SFN thyroid nodules using CANI from follicular cells. Various nuclear morphometric and textural parameters were evaluated and compared with the final histological diagnosis of benignity or malignancy.

Stored cytological material was studied from 101 nodules cytologically classified as AUS / FLUS and FN / SFN from 97 patients consecutively submitted to thyroidectomy between 2009 and 2017. There were 87 women and
10 men with a median age (25 and 75 percentiles) of 58 (48.5; 61.5) years and largest nodule diameter of 1.5 (1.0; 3.0) cm. Sixty-eight patients presented AUS / FLUS and 32 presented FN / SFN nodules (3 patients had nodules of both classes and 1 patient presented 2 FN / SFN nodules). Thus, in total 68 (67.3%) AUS / FLUS and 33 (32.7%) FN / SFN nodules were studied.

Two experienced pathologists (MEAM and CCO) reviewed all the slides containing the cytological material stained with Giemsa, and the respective histological samples stained with hematoxylin-eosin (HE). Cytological diagnosis was classified as either AUS / FLUS or FN / SFN and histopathological as either benign or malignant according to recommendations (2, 15).

**Ethical aspects**

This study was approved by the Research Ethics Committee of Botucatu School of Medicine – Unesp – under protocol number 1.622.593, issued on July 05, 2016 (CAAE: 56858616.9.0000.5411). Consent was not required.

**Image acquisition and analysis**

Cytological slides were scanned and photographed (43x magnification; PANORAMIC MIDI II - 3D Histech, Japan; http://3dhistech.com/pannoramic_midi), to obtain two to three photos of the areas with the largest number of follicular cell nuclei, avoiding regions with overlapping, clustered or fragmented nuclei (8). The best photograph from each slide was chosen for analysis, according to the opinion of two experienced pathologists (MEAM and CCO), and randomly numbered.
The captured images were evaluated using free software (Image J; https://imagej.nih.gov/ij/), by two researchers who did not know the final histopathological diagnosis (blinded analysis). The program converts the colour image to one containing 256 shade greyscale (16 bits), allowing better chromatin analysis and attenuating colour differences between samples. Careful manual tracing of the boundaries of each nucleus in the digitized image was performed using drawing tools, followed by data extraction (Figure 1). The number of nuclei to be evaluated in each photo was determined through a pilot statistical study that estimated the minimum quantity from which the addition of more nuclei would not alter the subsequent statistical analysis. This study was based on the standard error stabilization of 5% for each variable, concluding that 60 nuclei per photo would be sufficient and thus warranting the precision of the measures, independently of the sampled area (7). As the nuclear contour of photographs containing cytological material is performed manually and could lead to inter-observer differences, method reproducibility was tested by analysing 20 slides by both evaluators.

For each selected nucleus, the program supplied a list of morphometric and textural parameters from which the most relevant for performing nuclear evaluation were chosen, according to the previously described (10, 16). Thus, morphometric evaluation included primary parameters such as Area, Perimeter, Circularity, Largest diameter (Feret), and the ratio between largest and smallest diameter (Aspect Ratio, AR). Textural evaluation included Mean, Median, and Standard Deviation of greyscale (STDEV), expressed in a 256 shade greyscale, in which higher numbers mean lighter nuclei (0= black; 255= white), and Roughness (RA), Solidity, Fractal Dimension (Fractal) and Entropy, which
measure the regularity of the grayscale distribution. Secondary indicators were also considered, such as coefficient of variation (CV) of Area, CV-Mean intensity, CV-STDEV, CV-Perimeter, CV-Circularity, CV-Feret, CV-Median intensity, CV-AR, CV-Round, CV-Solidity, CV-Fractal, CV-Entropy and CV-RA. The program expresses these parameters in pixels, which can then be converted into microns (µm) (7).

Comparison and classification of the lesions

The lesions with benign and malignant final histological diagnoses were compared using the above nuclear cytological parameters. The cytological material was subsequently classified for malignancy using a regression model. This classification was then compared for final histological diagnosis with the aim of evaluating the diagnostic capacity of the method.

Statistical analysis

Collected data were submitted to statistical analysis using SPSS/Windows (version 21). Numerical variables were described by medians and quartiles (p25-p75) and the Mann-Whitney and Tukey tests were used. Patient gender was described as a numerical value and percentage and the Fisher Exact Test was used. The adopted level of significance was 5.0 %.

Based on the cytological nuclear parameters, the tumors were classified using the CRT (Classification and Regression Tree) regression model with the Gini algorithm, allowing the construction of a predictive classification tree, which starts from a root and branches from nodes that use cut-offs for the analyzed parameters provided by the model itself (17). Initially, a classification tree was
prepared with all AUS / FLUS and FN / SFN nodules for malignancy and
geniness, using STDEV as the initial and Median as the final parameter. Then,
the AUS / FLUS and FN / SFN nodules were analyzed separately by
building a classification tree for each cytological class. In these last two
trees, the initial parameter was STDEV, with final parameters of Entropy and
AR in AUS / FLUS nodules, and Fractal in FN / SFN nodules. Validity of the
classification method was determined by calculating sensitivity, specificity,
positive predictive value (PPV), negative predictive value (NPV), accuracy and
area under the receiver operator characteristic (ROC) curve (AUC). The
reproducibility was tested using the intraclass correlation coefficient (ICC). Sixty
nuclei from each of the 20 randomly chosen slides were analyzed by both
evaluators. In this statistical tool, reliability can be classified as absent (0), poor
(0.01-0.19), weak (0.20-0.39), moderate (0.30-0.59), substantial (0.60-0.79) and
almost complete (≥0.80) (18, 19).

Results

Of the studied nodules, 72 (71.3%) were benign (28 adenomas, 17
hyperplasia, 15 colloid goiters, 10 Hashimoto's thyroiditis, 1 metaplasia and 1
parathyroid adenoma) and 29 (28.7%), malignant nodules (11 follicular variants
of papillary carcinoma - FVPC, 8 papillary carcinomas - PC, 5 follicular
carcinomas, 4 Hürthle cell carcinomas and 1 parathyroid carcinoma). There
were no cases of non-invasive follicular thyroid neoplasm with papillary-like
nuclear features (NIFTP). Twenty (29.4%) of the AUS / FLUS nodules and 9
(27.3%) of the FN / SFN nodules were diagnosed malignant on histological
analysis. Those with benign and malignant final histological diagnoses did not
differ according to gender \( (p = 0.99) \) or to largest nodular diameter \( (p = 0.90) \). Patients with benign nodules were older than those with malignant nodules \( (p = 0.03; \text{Table 1}) \).

Analysing AUS / FLUS and FN / SFN nodules together, there was a difference between benign and malignant lesions for STDEV and Entropy \( (p < 0.05; \text{Table 1 - supplementary material}) \). Isolated analysis of AUS / FLUS nodules revealed benign and malignant lesions differed only for Entropy \( (p < 0.05; \text{Table 2 - supplementary material}) \), while FN / SFN nodules differed in Fractal, CV-RA and CV-Entropy \( (p < 0.05; \text{Table 3 - supplementary material}) \).

By jointly assessing AUS / FLUS and FN / SFN nodules for nuclear parameters using the CRT model (Figure 2), a correct classification of 65 (90.3\%) benign and 24 (82.8\%) malignant nodules was obtained, with a correct global classification of 89 (88.1\%) cases (Table 4 - supplementary material). Thus, analysis sensitivity, specificity, AUC, NPV, and accuracy were greater than 80\% (Table 2). When AUS / FLUS nodules were assessed separately for lesion nature using the same model (Figure 3), 46 (95.8\%) benign and 18 (90.0\%) malignant nodules were correctly classified, with a correct overall classification of 94.1\% (Table 5 - supplementary material). Sensitivity, specificity, PPV, NPV, accuracy and AUC were 90\% or more (Table 2). In the isolated analysis of FN / SFN nodules (Figure 4), 23 (95.8\%) benign and 9 (100\%) malignant were correctly classified, resulting in a correct overall classification of 97\% (Table 6 - supplementary material), with specificity and PPV of 100\%. Sensitivity, NPV, accuracy and AUC were 90\% or more (Table 2).

Regarding method reproducibility, almost complete agreement was observed in parameters Mean, STDEV, Median and Circularity, substantial
agreement in Area, Perimeter, Feret, AR, Fractal and Entropy, and moderate agreement in Solidity and RA (Table 3).

Discussion

CANI has proven a useful technique in general cytohistological evaluation of thyroid nodules. However, in lesions classified by Bethesda as indeterminate and composed mainly of AUS / FLUS and FN / SFN, results have not been fully conclusive (12, 20, 21). Recently, Valentim et al., 2018, using CANI to evaluate thyroid tumor tissues which frequently present these cytological diagnoses, observed high percentages of sensitivity and final diagnostic specificity (7). Their findings prompted us to undertake the present study in which the same tool was used to evaluate smears from thyroid nodules with indeterminate cytological diagnoses in which high percentages of correct classification regarding the real nature of the lesion were also observed.

CANI has been explored in detecting nuclear differences between colloid nodules and FVPC, between adenomatous goiter and follicular carcinoma, and even between anaplastic carcinoma and other tumors (12, 21). In fact, sensitivities of up to 98% have been reported in the evaluation of several thyroid lesions that included mainly classic papillary carcinoma (20). However, few authors have performed specific cytological evaluation of AUS / FLUS and FN / SFN nodules, the precise critical point of the Bethesda classification system. For instance, Collins & Collins, in 2013, evaluated some AUS / FLUS cases using commercial software, and observed significant differences between benign and malignant lesions regarding the nucleus / cytoplasm ratio (14). These findings encourage the use of computerized systems to evaluate FN /
SFN thyroid lesions, as well as investigate the usefulness of other free software. Additionally, as cytoplasm seems more prone to artefacts appearing during the FNA procedure and smear execution, it would be interesting to consider whether nuclear evaluation *per se* would be sufficient.

In this present study, CANI was used to evaluate a considerable number of AUS / FLUS and FN / SFN thyroid nodules, and the vast majority of cases were correctly classified in relation to final histological diagnosis. When AUS / FLUS and FN / SFN nodules were considered together in the analysis, a correct global classification of 88% was obtained, with sensitivity and specificity higher than 80% in detecting malignancy. However, individual evaluation by cytological class would seem more appropriate, as in practice, the nodule under investigation would present either AUS / FLUS or FN / SFN diagnosis and not AUS / FLUS and FN / SFN. Interestingly, these individual evaluations produced even higher percentages, with rates of 90% or more for sensitivity, specificity, PPV, NPV, accuracy and AUC. For FN / SFN nodules, specificity and PPV reached 100%, correctly classifying all truly malignant lesions.

An interesting finding of this study was the similarity in malignancy rates, close to 30%, observed for AUS / FLUS and FN / SFN nodules. Although rates below 15% are generally attributed to the AUS / FLUS category, malignancy rates for this diagnostic class, particularly in cancer centres, have in fact been higher than previously estimated, and can reach almost 38% (22). It is very important to consider the frequency of malignancy of the institution when evaluating different diagnostic tests since this rate can influence the percentages related to the performance of these tests (6).
Limitations of this study could be possible inter-observer variability in results using this method, the time required for nuclear image analysis, the number of nodes required for the design and size of the classification trees, as well the lack of NIFTP cases, and the sample size. In regard to inter-observer variability, we observed high reproducibility levels using CANI in this study as there was almost complete agreement between both evaluators in 10 the 12 parameters analysed (7). However, the model was not validated for the entire cohort of patients in order to test that reproducibility. Furthermore, it would be of interest to see if the developed model remains highly accurate in predicting malignancy if used on a new cohort of patients in the same institution, and even more importantly, to see the validation of the model for a cohort of patients from an outside institution. Regarding the time required for nuclear analysis, it varies according to individual experience, decreasing as experience increases, up to about 30 minutes/case (7). As for number of nodes and classification tree size, very large trees with numerous nodes can be associated with overfitting and a lack of explanatory power (7). This did not prove to be a significant problem in this study as both number of nodes and tree sizes could only be considered small to moderate. As for sample size, it could have been larger. However, it was sufficient to allow developing evaluation algorithms which can be tested on bigger samples in the future. Studies with larger samples would probably include NIFTP cases (23), which were not found in our group of patients, and could have nuclear alterations, but turn out to be "non-malignant" in the final histological examination. In this sense, we compared recently the histological material of NIFTP, infiltrative FVPC, and encapsulated/well-demarcated FVPC with tumor capsular invasion, using CANI, and observed nuclear textural
differences between NIFTP and infiltrative FVPC, but not between the former and encapsulated/well-demarcated FVPC (24). In addition to significantly larger samples, it would also be interesting to apply this model to other Bethesda categories, such as benign, for example. Thus, further studies are needed to address these issues.

Despite the above-mentioned limitations, this study has the merit of having developed models to aid predicting malignancy in AUS / FLUS and FN / SFN thyroid nodules. This easily accessible, inexpensive, and reproducible methodology could be a useful diagnostic support tool in evaluating lesions of indeterminate cytological diagnosis.

In conclusion, our study showed that computerized analysis of nuclear images of cytological material, associated with statistical tree classification can effectively classify AUS / FLUS and FN / SFN thyroid nodules for malignancy.

**Author Disclosure Statement**

No competing financial interests exist.

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Legend figures

**Figure 1.** Photograph of cytologic smear under analysis by Image J program.

A: color photo; B: grayscale photo; C and D: selected nucleus.

**Figure 2.** Method of Classification and Regression Trees - CRT, with Gini Model, of all AUS/FLUS and FN/SFN thyroid nodules, benign or malignant, according to the cytological nuclear parameters evaluated. AUS/FLUS, Atypia of Undetermined Significance/Follicular Lesion of Undetermined Significance; FN/SFN, Follicular Neoplasm/Suspicious for a Follicular Neoplasm; STDEV: standard deviation.

**Figure 3.** Method of Classification and Regression Trees - CRT, with Gini Model, of AUS/FLUS thyroid nodules, benign or malignant, according to the cytological nuclear parameters evaluated. AR: Aspect Ratio; AUS/FLUS, Atypia of Undetermined Significance/Follicular Lesion of Undetermined Significance; CV: coefficient of variation; CV-RA: coefficient of variation of roughness; STDEV: standard deviation.

**Figure 4.** Method of Classification and Regression Trees - CRT, with Gini Model, of FN/SFN thyroid nodules, benign or malignant, according to the
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roughness;

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TABLE 1  General data of 101 thyroid nodules in 97 patients com aus/flus and fn/sfn thyroid nodules.

| Parameter           | Final histological diagnosis | p   |
|---------------------|------------------------------|-----|
|                     | Benign                      | Malignant |
| Patients* (n)       | 71                          | 26  | -   |
| Men [n (%)]†        | 7 (9.9)                     | 3 (11.5) | 0.99 |
| Age (years)‡        | 59 (48.9; 63.0)             | 54 (41.8; 58.8) | 0.03 |
| Larger diameter (cm)‡ | 1.5 (1.0; 3.0)              | 1.5 (0.8; 3.2) | 0.90 |

*Only the worst diagnosis was considered. † Fisher's Exact Test. ‡ Median (25th percentile, 75th percentile), Mann-Whitney U test. Significance: p < 0.05. AUS/FLUS, Atypia of Undetermined Significance/Follicular Lesion of Undetermined Significance; cm, centimeter; FN/SFN, Follicular Neoplasm/Suspicious for a Follicular Neoplasm; n, number; %, percentage.
TABLE 2 Validity of the use of computed nuclear morphometry analysis, with the classification and regression trees (crt) analysis, as a classificatory method.

| Bethesda categories | AUS/FLUS and FN/SFN | AUS/FLUS | FN/SFN |
|---------------------|----------------------|----------|--------|
| Sensitivity (%)     | 83.0                 | 90.0     | 90.0   |
| Specificity (%)     | 90.0                 | 95.8     | 100.0  |
| PPV (%)             | 77.4                 | 90.0     | 100.0  |
| NPV (%)             | 93.0                 | 95.8     | 95.8   |
| Accuracy (%)        | 88.1                 | 94.1     | 97.0   |
| Area under the ROC curve | 0.86               | 0.93     | 0.95   |

AUS/FLUS, Atypia of Undetermined Significance/Follicular Lesion of Undetermined Significance; FN/SFN, Follicular Neoplasm/Suspicious for a Follicular Neoplasm; NPV, negative predictive value; %; percentage; PPV, positive predictive value; ROC, receiver operator characteristic.
**TABLE 3** Intraclass correlation coefficient, between two examiners, observed for each of the cytological nuclear parameters evaluated.

| Parameter              | Intraclass correlation coefficient | 95% Confidence interval |
|------------------------|-----------------------------------|-------------------------|
| Area                   | 0.76                              | 0.48 – 0.90             |
| Mean gray intensity    | 0.93                              | 0.80 – 0.97             |
| STDEV                  | 0.83                              | 0.63 – 0.93             |
| Perimeter              | 0.77                              | 0.52 – 0.90             |
| Feret                  | 0.79                              | 0.54 – 0.91             |
| Median gray intensity  | 0.90                              | 0.76 – 0.96             |
| AR                     | 0.61                              | 0.25 – 0.83             |
| Circularity            | 0.93                              | 0.83 – 0.97             |
| Solidity               | 0.42                              | 0.03 – 0.72             |
| RA                     | 0.59                              | -0.1 – 0.85             |
| Fractal                | 0.73                              | 0.44 – 0.89             |
| Entropy                | 0.70                              | 0.38 – 0.87             |

AR, Aspect Ratio; RA, roughness; STDEV, standard deviation of gray intensity.
Supplementary material

Table 1. Morphological and textural parameters of all thyroid nodules analyzed (classes III and IV) *, according to the final histological diagnosis.

| Parameter                  | Final histological diagnosis |  |  |
|----------------------------|------------------------------|---|---|
|                            | **Benign**                   | **Malignant** | **P**   |
| Area (µm²)                 | 57.18 (45.75; 70.49)         | 57.87 (46.72; 81.83) | 0.281  |
| Mean gray intensity ††      | 29.96 (22.74; 40.86)         | 27.73 (22.32; 43.50) | 0.407  |
| STDEV                      | 14.10 (11.97; 17.38)         | 12.31 (11.04; 15.08) | 0.020  |
| Perimeter (µm)             | 28.03 (24.20; 31.23)         | 27.10 (21.36; 30.72) | 0.519  |
| Circularity                | 0.90 (0.89; 0.91)            | 0.89 (0.88; 0.91)   | 0.422  |
| Feret (µm)                 | 9.84 (8.62; 10.79)           | 9.41 (7.81; 10.93)  | 0.699  |
| Median gray intensity ††   | 26.50 (17.75; 35.50)         | 25.00 (19.00; 38.00) | 0.721  |
| AR                         | 1.25 (1.21; 1.30)            | 1.26 (1.20; 1.31)   | 0.744  |
| Round                      | 0.80 (0.77; 0.83)            | 0.80 (0.77; 0.84)   | 0.721  |
| Solidity †                 | 0.95 (0.95; 0.96)            | 0.95 (0.95; 0.96)   | 0.717  |
| Fractal                    | 2.44 (2.40; 2.47)            | 2.46 (2.42; 2.50)   | 0.107  |
| Entropy                    | 2.65 (2.22; 3.13)            | 2.31 (2.03; 2.71)   | 0.010  |
| RA/1000 ‡                  | 187.41 (169.71; 202.98)      | 186.39 (170.54; 199.02) | 0.875  |
| CV-Área                    | 26.11 (20.58; 30.52)         | 22.92 (20.72; 29.47) | 0.620  |
| CV-Mean gray intensity     | 33.06 (27.40; 42.37)         | 34.34 (26.90; 40.57) | 0.775  |
| CV-STDEV                   | 36.54 (28.83; 41.30)         | 34.87 (28.02; 42.14) | 0.543  |
| CV-Perimeter               | 13.06 (10.23; 15.33)         | 12.19 (10.67; 14.91) | 0.713  |
| CV-Circularity             | 5.75 (4.45; 6.57)            | 5.61 (4.12; 6.85)   | 0.702  |
| CV-Feret                   | 13.65 (11.36; 16.89)         | 13.34 (10.78; 15.33) | 0.431  |
| CV-Median gray intensity   | 38.13 (30.22; 47.49)         | 37.45 (30.11; 43.04) | 0.719  |
| CV-AR                      | 15.66 (12.83; 18.88)         | 15.71 (13.45; 18.53) | 0.887  |
| CV-Round                   | 13.88 (11.34; 16.16)         | 14.05 (11.97; 16.31) | 0.758  |
| CV-Solidity                | 1.29 (1.06; 1.67)            | 1.33 (0.93; 1.82)   | 0.713  |
| CV-Fractal                 | 1.62 (1.34; 2.04)            | 1.70 (1.19; 2.05)   | 0.724  |
| CV-Entropy                 | 0.48 (0.30; 0.76)            | 0.32 (0.25; 0.59)   | 0.079  |
| CV-RA     | 12.27 (9.01; 19.94) | 10.35 (7.70; 14.63) | 0.129 |
|-----------|-------------------|--------------------|-------|

*Median (percentile 25; percentile 75); Tukey test. Significance: p< 0.05. †Expressed in a scale of 256 shades of grey, in which higher numbers mean lighter nuclei (0= black; 255= white). AR: Aspect Ratio; CV: coefficient of variation; RA: roughness; STDEV: standard deviation of Gray intensity; µm: micrometer.
Supplementary material

Table 2. Morphological and textural parameters of the class III nodules analyzed *, according to the final histological diagnosis.

| Parameter                  | Final histological diagnosis |       |      |
|----------------------------|-------------------------------|-------|------|
|                            | Benign                        | Malignant | p    |
| Area (µm²)                 | 57.18 (46.37; 66.92)          | 53.37 (44.72; 75.76) | 0.995 |
| Mean gray intensity†       | 29.36 (21.93; 37.42)          | 28.11 (24.75; 44.33) | 0.747 |
| STDEV                      | 13.99 (11.83; 17.44)          | 12.35 (11.08; 14.93) | 0.085 |
| Perimeter (µm)             | 28.03 (24.00; 30.50)          | 27.13 (23.02; 29.27) | 0.686 |
| Circularity                | 0.90 (0.89; 0.91)             | 0.89 (0.88; 0.91)    | 0.448 |
| Feret (µm)                 | 9.84 (8.44; 10.54)            | 9.41 (8.41; 10.30)   | 0.893 |
| Median gray intensity†     | 25.50 (17.00; 34.00)          | 26.00 (22.00; 40.00) | 0.492 |
| AR                         | 1.24 (1.21;1.32)              | 1.25 (1.20; 1.36)    | 0.726 |
| Round                      | 0.81 (0.76; 0.83)             | 0.80 (0.74; 0.84)    | 0.736 |
| Solidity†                  | 0.95 (0.95; 0.95)             | 0.95 (0.94; 0.96)    | 0.670 |
| Fractal                    | 2.44 (2.40; 2.48)             | 2.44 (2.42; 2.47)    | 0.706 |
| Entropy                    | 2.65 (2.30; 3.12)             | 2.33 (2.06; 2.77)    | 0.025 |
| RA/1000                    | 188.91 (174.27; 203.78)       | 186.59 (164.08; 200.37) | 0.788 |
| CV-Área                    | 26.71 (20.58; 31.72)          | 26.18 (20.11; 30.13) | 0.686 |
| CV-Mean gray intensity     | 34.22 (28.28; 45.40)          | 37.17 (27.10; 43.88) | 0.968 |
| CV-STDEV                   | 36.62 (30.20; 41.66)          | 33.72 (28.38; 42.37) | 0.628 |
| CV-Perimeter               | 13.25 (10.71; 15.96)          | 12.84 (10.25; 15.32) | 0.518 |
| CV-Circularity             | 5.86 (4.31; 6.71)             | 5.42 (4.00; 6.85)    | 0.882 |
| CV-Feret                   | 14.27 (11.43; 17.58)          | 14.06 (10.61; 15.72) | 0.326 |
| CV-Median gray intensity   | 38.78 (32.17; 47.83)          | 38.55 (29.42; 45.65) | 0.861 |
| CV-AR                      | 15.54 (12.34; 19.44)          | 16.07 (13.53; 18.60) | 0.989 |
| CV-Round                   | 13.93 (11.62; 16.50)          | 14.22 (12.26; 16.59) | 0.686 |
| CV-Solidity                | 1.34 (1.10; 1.67)             | 1.40 (0.89; 1.81)    | 0.647 |
| CV-Fractal                 | 1.69 (1.34; 2.04)             | 1.75 (1.43; 2.23)    | 0.484 |
| CV-Entropy                 | 0.45 (0.30; 0.73)             | 0.53 (0.25; 0.75)    | 0.809 |
| CV-RA         | 12.05 (9.31; 17.10) | 11.54 (8.87; 20.90) | 0.809 |

*Median (percentile 25; percentile 75); Tukey test. Significance: p< 0.05. †Expressed in a scale of 256 shades of grey, in which higher numbers mean lighter nuclei (0= black; 255= white). AR: Aspect Ratio; CV: coefficient of variation; RA: roughness; STDEV: standard deviation of Gray intensity; µm: micrometer.
Supplementary material

Table 3. Morphological and textural parameters of the class IV nodules analyzed *, according to the final histological diagnosis.

| Parameter               | Final histological diagnosis |       |       | p     |
|-------------------------|------------------------------|-------|-------|-------|
|                         | Benign                       | Malignant |       |       |
| Area (µm²)              | 58.06 (46.68; 80.88)         | 81.83 (64.49; 142.70) | 0.075 |
| Mean gray intensity†    | 31.56 (24.91; 44.59)         | 22.32 (14.88; 33.89) | 0.066 |
| STDEV                   | 14.12 (12.87; 16.84)         | 11.57 (8.85; 16.54) | 0.196 |
| Perimeter (µm)          | 27.92 (24.75; 32.55)         | 25.92 (4.45; 31.17) | 0.518 |
| Circularity             | 0.90 (0.89; 0.90)            | 0.89 (0.88; 0.91) | 0.806 |
| Feret (µm)              | 9.83 (8.86; 11.46)           | 9.27 (1.58; 11.14) | 0.544 |
| Median gray intensity†  | 28.50 (22.50; 37.50)         | 19.00 (11.00; 31.00) | 0.157 |
| AR                      | 1.26 (1.21; 1.29)            | 1.28 (1.21; 1.28) | 0.936 |
| Round                   | 0.80 (0.78; 0.82)            | 0.78 (0.78; 0.84) | 0.919 |
| Solidity†               | 0.95 (0.95; 0.96)            | 0.95 (0.95; 0.96) | 0.215 |
| Fractal                 | 2.44 (2.41; 2.47)            | 2.51 (2.50; 2.53) | 0.032 |
| Entropy                 | 2.64 (2.17; 3.13)            | 2.16 (2.01; 2.54) | 0.157 |
| RA/1000**               | 184.19 (157.68; 200.87)      | 186.39 (170.54; 199.02) | 0.544 |
| CV-Área                 | 23.45 (20.47; 28.77)         | 22.48 (22.23; 28.37) | 0.903 |
| CV-Mean gray intensity  | 30.34 (25.78; 38.94)         | 33.13 (25.57; 34.64) | 0.936 |
| CV-STDEV                | 36.15 (28.26; 41.04)         | 37.02 (25.74; 39.51) | 0.628 |
| CV-Perimeter            | 11.64 (9.99; 14.78)          | 11.48 (11.08; 13.75) | 0.746 |
| CV-Circularity          | 5.48 (4.72; 6.29)            | 6.20 (4.84; 7.27) | 0.374 |
| CV-Feret                | 12.36 (11.14; 14.69)         | 12.37 (11.40; 14.72) | 0.840 |
| CV-Median gray intensity| 32.71 (28.55; 45.93)         | 36.28 (31.45; 38.01) | 0.872 |
| CV-AR                   | 15.98 (13.21; 17.66)         | 14.46 (12.17; 18.53) | 0.657 |
| CV-Round                | 13.82 (11.29; 15.69)         | 12.78 (10.30; 16.29) | 0.968 |
| CV- Solidity            | 1.24 (1.04; 1.66)            | 1.16 (1.08; 2.03) | 0.808 |
| CV-Fractal              | 1.57 (1.34; 2.05)            | 1.19 (1.00; 1.89) | 0.090 |
| CV-Entropy              | 0.50 (0.32; 0.92)            | 0.26 (0.25; 0.30) | 0.008 |
| CV-RA | 12.83 (8.92; 24.49) | 7.84 (6.93; 10.41) | 0.017 |
|-------|-------------------|-------------------|-------|

*Median (percentile 25; percentile 75); Tukey test. Significance: p< 0.05. †Expressed in a scale of 256 shades of grey, in which higher numbers mean lighter nuclei (0= black; 255= white). AR: Aspect Ratio; CV: coefficient of variation; RA: roughness; STDEV: standard deviation of Gray intensity; µm: micrometer.
Supplementary material

Table 4. Comparative analysis between Classification and Regression Trees (CRT), with Gini Model, and the final benign and malignant histological diagnosis, in class III and IV nodules.

| Real diagnosis | Classification by the CRT | Correct classification (%) |
|----------------|---------------------------|----------------------------|
|                | Benign (n) | Malignant (n) | |
| Benign         | 65         | 7             | 90.3                      |
| Malignant      | 5          | 24            | 82.8                      |
| Global classification (%) | 88.1 |

CRT, Classification and Regression Trees; n, number.
Table 5. Comparative analysis between Classification and Regression Trees (CRT), with Gini Model, and the final benign and malignant histological diagnosis, in class III nodules.

| Real diagnosis | Classification by the CRT | Correct classification (%) |
|----------------|---------------------------|----------------------------|
|                | Benign (n)                | Malignant (n)              |                             |
| Benign         | 46                        | 2                          | 95.8                       |
| Malignant      | 2                         | 18                         | 90.0                       |
| Global classification (%) |                 |                            | 94.1                      |

CRT, Classification and Regression Trees; n, number.
Supplementary material

Table 6. Comparative analysis between Classification and Regression Trees (CRT), with Gini Model, and the final benign and malignant histological diagnosis, in class IV nodules.

| Real diagnosis | Classification by the CRT | Correct classification (%) |
|----------------|---------------------------|---------------------------|
|                | Benign (n)                | Malignant (n)             |
| Benign         | 23                        | 1                         | 95.8                       |
| Malignant      | 0                         | 9                         | 100.0                      |
| **Global classification (%)** | **97.0** | **97.0**                  |

CRT, Classification and Regression Trees; n, number.