Radiation-Induced Lymphopenia Risks of Photon Versus Proton Therapy for Esophageal Cancer Patients

Saba Ebrahimi, BSc1; Gino Lim, PhD1; Amy Liu, MSc2; Steven H. Lin, MD, PhD3; Susannah G. Ellsworth, MD4; Clemens Grassberger, PhD5; Radhe Mohan, PhD2; Wenhua Cao, PhD2

1Department of Industrial Engineering, University of Houston, Houston, TX, USA
2Department of Radiation Physics, The University of Texas MD Anderson Cancer Center, Houston, TX, USA
3Department of Radiation Oncology, The University of Texas MD Anderson Cancer Center, Houston, TX, USA
4Department of Radiation Oncology, Indiana University, Indianapolis, IN, USA
5Departments of Radiation Oncology, Massachusetts General Hospital, Boston, MA, USA

Abstract

**Purpose:** To assess possible differences in radiation-induced lymphocyte depletion for esophageal cancer patients being treated with the following 3 treatment modalities: intensity-modulated radiation therapy (IMRT), passive scattering proton therapy (PSPT), and intensity-modulated proton therapy (IMPT).

**Methods and Materials:** We used 2 prediction models to estimate lymphocyte depletion based on dose distributions. Model I used a piecewise linear relationship between lymphocyte survival and voxel-by-voxel dose. Model II assumes that lymphocytes deplete exponentially as a function of total delivered dose. The models can be fitted using the weekly absolute lymphocyte counts measurements collected throughout treatment. We randomly selected 45 esophageal cancer patients treated with IMRT, PSPT, or IMPT at our institution (15 per modality) to demonstrate the fitness of the 2 models. A different group of 10 esophageal cancer patients who had received PSPT were included in this study of in silico simulations of multiple modalities. One IMRT and one IMPT plan were created, using our standards of practice for each modality, as competing plans to the existing PSPT plan for each patient. We fitted the models by PSPT plans used in treatment and predicted absolute lymphocyte counts for IMRT and IMPT plans.

**Results:** Model validation on each modality group of patients showed good agreement between measured and predicted absolute lymphocyte counts nadirs with mean squared errors from 0.003 to 0.023 among the modalities and models. In the simulation study of IMRT and IMPT on the 10 PSPT patients, the average predicted absolute lymphocyte count (ALC) nadirs were 0.27, 0.35, and 0.37 K/µL after IMRT, PSPT, and IMPT treatments using Model I, respectively, and 0.14, 0.22, and 0.33 K/µL using Model II.

**Conclusions:** Proton plans carried a lower predicted risk of lymphopenia after the treatment course than did photon plans. Moreover, IMPT plans outperformed PSPT in terms of predicted lymphocyte preservation.

**Keywords:** IMRT; PSPT; IMPT; lymphopenia
Introduction

Radiation-induced lymphopenia (RIL), lymphocyte depletion, is a common toxicity of radiation therapy and is associated with worse outcomes in a number of solid tumors, including esophageal cancer [1–4]. Because lymphocytes have a substantial role in the body’s anticancer immune response, severe lymphopenia can reduce patients’ survival even in the early stages of tumor progression [4–8]. Multiple recent studies have shown that severe lymphopenia is strongly associated with poor treatment outcomes for patients with cervical [8, 9], pancreatic [10, 11], rectal [12], lung [5, 13], and esophageal [2, 14] cancers. Thus, preservation of the lymphocytes from radiation damage is crucial for the effectiveness of radiation therapy, and it is critical to understand the clinical and dosimetric factors affecting the severity and incidence of RIL and develop strategies for its mitigation.

Radiation-induced lymphopenia occurs presumably owing to the high radiosensitivity of lymphocytes. In conventional photon radiation therapy, the large low- and medium-dose bath expose substantial fractions of circulating lymphocytes. Clinical data show that dose distribution patterns and fractionation regimens significantly influence lymphocyte depletion [15, 16]. Dose distribution patterns from protons and photons can differ greatly, and the dosimetric advantages of the state-of-the-art proton therapy over photon therapy in terms of sparing of organs at risk and normal tissue have been demonstrated extensively [17, 18]. Furthermore, intensity-modulated proton therapy (IMPT) performs further better than intensity-modulated radiation therapy (IMRT) in terms of normal tissue sparing [19].

RIL risk likely depends on the treatment modality. Recent studies have reported greater lymphocyte depletion in patients treated with photon therapy than with proton therapy [2, 4, 13, 14, 20]. For example, Shiraishi et al [2] reported that proton beam therapy was associated with a lower risk of grade 4 lymphopenia compared with IMRT in esophageal cancer patients receiving neoadjuvant chemoradiotherapy.

In this study, we aimed to analyze probability of severe lymphopenia induced from radiation therapy using simple models of lymphocyte loss based on dose distribution patterns. Such models have the potential to help physicians identify patients at high risk of grade 4 lymphopenia based on the evaluation of dose distributions of treatment plans. In this regard, we modeled and compared expected absolute lymphocyte count (ALC) depletion kinetics in esophageal cancer patients treated with the following 3 different modalities: IMRT, passive-scattering proton therapy (PSPT), and IMPT.

Materials and Methods

In this section, we will describe 2 prediction models for lymphocyte depletion based on radiation doses and then report the patient selection for model validation as well as treatment planning of IMRT and IMPT for patients treated with PSPT.

ALC Depletion Prediction Using a Piecewise Linear Lymphocyte Survival Function

On the basis of the dose distributions in each patient, we can estimate the ALC during treatment by using a piecewise linear relationship between lymphocyte survival and dose per fraction for each modality. A piecewise linear function can be modeled by interpolating previous findings about radiation-induced lymphocyte death [21, 22]. Nakamura et al [22] reported the percentages of surviving lymphocytes as 90%, 50%, 10%, and 0% for radiation doses of 0.5 Gy, 2 Gy, 3 Gy, and 6 Gy or higher for each fraction, respectively. This estimation assumes that all circulating lymphocytes may receive dose by the end of treatment (after 28 fractions in this study). The lymphocyte survival probability for each voxel i, \( S_i \), after receiving a fractional dose \( d \) (in Gy), can be calculated using the following piecewise linear function:

\[
S_i(d) = \begin{cases} 
-0.2d + 1 & 0 \leq d < 0.5, \\
-0.26d + 1.03 & 0.5 \leq d < 2, \\
-0.4d + 1.3 & 2 \leq d < 3, \\
-0.03d + 0.2 & 3 \leq d < 6, \\
0 & 6 \leq d. \end{cases}
\]

We assume that the initial number of lymphocytes in 1 μL of body volume can be estimated by multiplying the pretreatment ALC value for each patient by the percentage of blood in 1 μL of body volume. We also assume that all fast and slowly circulating lymphocytes are distributed uniformly throughout the irradiated volume and receive dose during radiation therapy treatment. The average percentage of blood in the human body is 7% of body weight/volume [23], so the number of lymphocytes in 1 μL of body volume before treatment can be estimated as \( L_0 = \text{ALC}_0 \times 0.07 \) (cells ×1000/μL). Therefore, the total number of lymphocytes in a given patient’s body would be
where \( \vartheta \) is the volume of each voxel and \( N_v \) is the total number of voxels in the body for each patient.

The total number of lymphocytes remaining in the body volume after 1 fraction is calculated by summing the number of surviving lymphocytes in all voxels as follows:

\[
N_1 = \sum_{i=1}^{N_v} S_i(\vartheta) \vartheta L_0.
\]

To find the ALC value after the entire course of treatments, the probability of lymphocyte survival after delivering \( k \) fractions \((P_k)\) is then calculated using the ratio of the remaining lymphocytes to the initial number of lymphocytes in the body.

\[
P_k = \frac{\sum_{i=1}^{N_v} (S_i(\vartheta))_k \vartheta \times L_0}{\vartheta \times L_0 \times N_v} = \frac{\sum_{i=1}^{N_v} (S_i(\vartheta))_k}{N_v}
\]

Thus, the expected final value of ALC after \( k \) fractions \((k \geq 1)\) can be estimated using the following equation:

\[
ALC_k = R \times P_k \times ALC_0 \text{cells/\muL},
\]

where \( R \) is the fitting parameter and \( ALC_0 \) is the ALC value before treatment. \( R \) can be determined by fitting the model to ALC measurements collected from patients. It is also used to account for all other compounding contributing factors, such as tumor location and histology, treatment volume, chemo status, and so on.

Note that only weekly ALC measurements could be available in current clinical practice, rather than measurement per fraction. Therefore, there would be at most 5 or 6 weekly ALC data points available for patients receiving a conventional photon or proton treatment, such as in this study.

**ALC Prediction Using Exponential Curve Fitting**

Previous studies have shown that ALC loss follows exponential decay in setting of total body radiation in primates [24] and in humans (accidental exposure) [25]. An exponential fitting method was also used to study ALC loss in partial body radiation therapy [26]. As an alternative to the piecewise linear model described above, we can also model patient ALC using an exponential function of accumulated delivered dose. Specifically, the measured weekly ALC data points from RT patients can be used to fit an exponential function based on the total delivered dose to the body (ie, the sum of doses in all voxels in treatment field) after \( k \) fractions \((D_k > 0)\) as

\[
ALC(D_k) = a \cdot \exp(-b \cdot D_k) + c,
\]

where \( a \) is a fixed parameter indicating the initial ALC before starting the treatment \( ALC_0 \), \( b \) is an index of an individual patient’s lymphocytes sensitivity to dose, and \( c \) is added to the exponential function to account for the replenishment of lymphocytes after irradiation. To avoid a negative value of ALC, nonnegative constraint was placed on \( c \) (ie, \( c \geq 0 \)).

Using the weekly ALC data points and delivered dose in each week, we can determine the fitted values of the \( b \) and \( c \) parameters for each patient. In the planning study, we used the same parameters to predict ALC during treatment for PSPT, IMRT, and IMPT plans according to individual patient’s ALC baseline value \((a)\) and the delivered dose values of \( D_k \).

In addition, one could use the ALC measurement data from only the initial weeks of treatment (eg, 3 weeks) to fit the exponential ALC function of dose to predict the final ALC after the entire treatment course. The rationale for this approach was to determine whether patient-specific factors, including lymphocyte radiation sensitivity, derived from initial treatment fractions, is predictive of loss of lymphocytes by the end of treatment.

**Patient Selection and Treatment Planning**

The model validation study and the in silico simulation study of retrospective patient cohorts were approved by the institutional review board of The University of Texas MD Anderson Cancer Center. Forty-five esophageal cancer patients, who had been treated with IMRT, PSPT, or IMPT (15 per modality) with the same treatment prescription of delivering 50.4 Gy in 28 fractions, were randomly selected to validate the prediction models of lymphocyte depletion. Dose distributions used in their treatments and weekly ALC measurements throughout the treatment course (including baseline ALC before treatment) were collected from our database. All patients had at least 5 ALC data points.

Ebrahimi et al (2021), *Int J Particle Ther*
Ten other esophageal cancer patients treated with PSPT (50.4 Gy delivered in 28 fractions) were included in the modality comparison study. Five patients had 6 weekly measurements; 4 patients had 5 measurements; and 1 patient had only 4 measurements. ALC nadir was defined as the lowest value among the weekly measurements for each patient. The average ALC nadir for the 10 patients was 0.34 K/μL, ranging from 0.07 to 0.68 K/μL. Grade 4 lymphopenia (G4L) and grade 3 lymphopenia (G3L), according to the Common Toxicity Criteria for Adverse Events, version 5.0 (National Cancer Institute, Bethesda, MD), are defined as ALC < 200 cells/μL and ALC < 500 cells/μL, respectively. Three patients had grade 2 lymphopenia, 4 had grade 3, and 3 had grade 4. All patients received concurrent chemoradiation therapy, during which chemotherapy regimens were doublets of a taxane, fluorouracil, or platinum-based compound. Examples of the PSPT dose distributions for all patients can be found in Supplemental Figure S1.

We used MatRad [27], a research-oriented treatment planning system, to create IMRT and IMPT plans for each patient. The prescription dose to the clinical target volume was 50.4 Gy in 28 fractions for all patients. Six or seven beams were used for IMRT plans for these 10 patients based on our planning protocol in clinic. Two or 3 beams were used for IMPT plans with the same beam angles as those used for patients’ actual PSPT treatments. In optimization of the IMRT and IMPT plans, the same dosimetric criteria were used for each patient, but objective weights and constraints were adjusted, when necessary, to achieve the best possible target coverage and normal tissue sparing. For optimization and evaluation of PSPT and IMPT plans, we used a constant relative biological effectiveness of 1.1 [28]. All plans were normalized to have 95% of the planning target volume receive the prescription dose.

### Results

#### Model Validation

Dose distributions and ALC measurements of 15 patients per treatment modality (IMRT, PSPT, or IMPT) were used to validate both piecewise linear and exponential models. Results of ALC predictions are summarized in Table 1. The average mean body doses were 14.44, 7.37, and 6.12 Gy for IMRT, PSPT, and IMPT treatments, respectively. The average ALC nadir for the 15 patients treated with IMRT, PSPT, and IMPT was 0.17, 0.33, and 0.39 K/μL, respectively. The average predicted ALC nadirs after treatment were 0.15, 0.32, and 0.37 K/μL after IMRT, PSPT, and IMPT treatments using the piecewise linear model, respectively, and 0.12, 0.30, and 0.36 K/μL using the exponential model. The mean squared error was 0.005, 0.023, and 0.003 for IMRT, PSPT, and IMPT treatments based on the piecewise linear model, and 0.005, 0.005, and 0.004 based on the exponential model.

For each group of patients, \( \Delta ALC \) (baseline – nadir) from real values and predictions were also calculated (Supplemental Table S1). The average \( \Delta ALC \) from measurements were 1.25, 1.08, and 0.97 K/μL for IMRT, PSPT, and IMPT treatments, respectively; the average predicted \( \Delta ALC \) were 1.27, 1.09, and 0.98 K/μL from the piecewise linear model, and 1.30, 1.11, and 0.99 K/μL from the exponential model. Both measured and predicted ALC changes indicate patients receiving IMRT had the most lymphocyte reduction during treatment, while IMPT patients had the least lymphocyte reduction.

As the average baseline ALCs from the patient groups were different among RT modalities, we also compared normalized \( \Delta ALC \) (ie, \( \Delta ALC/baseline \ ALC \)) (Supplemental Table S2). The results also showed good agreement between real values and predictions from the 2 models. The mean squared errors for all treatment modalities and prediction models were less than 0.007, and the mean absolute errors were less than 0.067.
**Dosimetric Characteristics of IMRT, PSPT and IMPT Plans**

In the planning study of the 10 patients treated with PSPT, we first compared the dose distributions of the PSPT plans used for treatment and the IMRT and IMPT plans generated for this study, in terms of dose-volume metrics (eg, mean body dose, $V_{5\%}$, $V_{10\%}$, etc.). An example of the dose distributions of IMRT, PSPT, and IMPT plans for a patient (patient 5) can be found in the Supplement (Supplemental Figure S2). As expected, the radiation dose using the IMPT plan conformed more closely to the planning target volume than did the PSPT and IMRT plans, and the IMRT plan resulted in the largest dose baths to the body. The mean body doses, averaged among the 10 patients, were 7.46, 4.84, and 3.85 Gy for IMRT, PSPT, and IMPT plans, respectively. The fractions of the body volume that received different doses were consistently higher for photon therapy (IMRT) plans than for proton therapy (IMPT and PSPT) plans, especially at low doses, such as 5 or 10 Gy. For each dose-volume index, IMPT plans outperformed PSPT plans. More detailed comparison of body dose-volume metrics for the 10 patients among the 3 modalities can be found in Supplemental Figure S3.

**Lymphocyte Survival Based on Piecewise Linear Function of Voxel Dose**

Using the piecewise linear lymphocyte survival function, we estimated ALCs as a function of delivered radiation dose to each voxel in the patient body for IMRT, PSPT, and IMPT plans for each patient. Before the analysis, we determined the fitting parameter, $R$ in (5), to be 26%, based on the average of the measured ALC data for the 10 PSPT patients. The same $R$ values were used for all following ALC predictions.

The average predicted ALCs after treatment were 0.27 (95% CI [0.21, 0.33]), 0.35 (95% CI [0.27, 0.42]), and 0.37 K/µL (95% CI [0.29, 0.44]) for IMRT, PSPT, and IMPT plans, respectively. Figure 1A shows box plots of predicted ALCs at the end of treatment courses for the 3 treatment modalities. Figure 1B shows box plots of predicted ALC changes before and after treatment ($\Delta$ALC = baseline – nadir). Proton plans showed smaller ALC depletion than did photon plans, and IMPT had the highest ALC nadirs compared with the other 2 plans.

The predicted ALC changes for each of the 10 patients for the 3 modalities are shown in Figure 2A. These $\Delta$ALC values for IMRT plans were higher than those of IMPT and PSPT plans for all 10 patients and support the hypothesis that protons consistently cause less ALC depletion than do photons. Actual ALC changes from measured ALC data for PSPT treatments are indicated by black diamonds. Of note, we observed that the ALC estimates for PSPT plans were relatively close to the real measured data, with a mean absolute error of 0.075 and a mean squared error of 0.010. It reassures that this simple linear model of lymphocyte survival from radiation dose can provide good predictions.

Recent studies have reported a lower risk of grade 4 lymphopenia for patients treated with proton therapy compared with patients treated with photon therapy [2, 14, 20]. Figure 2B shows the predicted ALC nadirs for IMRT and IMPT treatments, which were calculated by subtracting the predicted ALC change from the measured ALC baseline, for each of the 10 patients. Figure 2B demonstrates the predicted ALC nadir if each patient had been treated by IMRT or IMPT instead of PSPT. Grade 4 lymphopenia occurred in patients 5, 7, and 8 after PSPT treatment. Based on the predicted ALC nadirs, grade 4 lymphopenia might have been avoided for patient 8 if they had been treated with IMPT instead. Patients 6 and 10 had grade 3 lymphopenia after PSPT; however, they may have developed grade 4 lymphopenia if they had been treated with IMRT. Patients 3 and 4 were predicted to have developed grade 3 lymphopenia if IMRT had been used instead of PSPT. In all cases IMPT would have reduced the risk of RIL.
Exponential Fitting

Weekly measurements of ALC and the total delivered dose for PSPT were used to fit an exponential curve (equation 6 above) for each patient. Figure 3A shows box plots of predicted posttreatment ALC values for all patients using this approach for all 3 modalities, and Figure 3B shows box plots of predicted ALC changes before and after treatment (ΔALC). Consistent with the predictions of the piecewise linear function approach, the predicted final ALC was the lowest for IMRT; IMPT was estimated to result in a higher ALC than PSPT. The fitted exponential curve and data points used for fitting for all patients are shown in Supplemental Figure S4. The average estimated ALCs after treatment, calculated using exponential fitting, were 0.14 (95% CI [0.08, 0.19]), 0.22 (95% CI [0.14, 0.30]), and 0.33 K/L (95% CI [0.19, 0.45]) for IMRT, PSPT, and IMPT plans, respectively, which shows the same trend as the previous approach.

A comparison of the predicted ALC change (baseline – nadir) using the exponential model is shown in Figure 4A. The ΔALC values for IMRT were higher than those for IMPT and PSPT for all 10 patients. Estimated ALCs had a mean absolute error of 0.125 and a mean squared error of 0.023, higher than for the piecewise linear method.
Figure 4B shows the predicted ALC nadirs for IMRT and IMPT treatments, similar to Figure 2B. For example, patients 7 and 8 may have avoided grade 4 lymphopenia if treated with IMPT, and patients 1 and 6 had grade 3 lymphopenia with PSPT but might have had grade 4 if treated with IMRT.

To predict lymphopenia in the early stages of treatment, we used the ALC-dose data for the first 3 weeks to estimate the parameters of the exponential ALC function. The fitted exponential curves using 3-week data for all patients are shown in Supplemental Figure S5. Figure 5A illustrates the measured ALC nadirs and the estimated final ALC based on 3-week and all-week data after PSPT treatment. ALC predictions based on 3-week data were lower than ALC predictions using all weekly data. Figure 5B shows an $R^2$ comparison between 2 exponential fittings, which was higher for the first exponential fitting for all 10 patients. The average $R^2$ values for exponential fitting using all weekly data and 3-week data were 93.5% and 90.0%, respectively.

Discussion

The present work suggests that IMPT would lead to less lymphocyte depletion than PSPT and that IMRT may produce the most lymphocyte depletion. This was demonstrated in both model validation study and treatment planning study. The 2 models of lymphocyte survival performed well in predicting the trend of ALC changes during RT treatment. It is worth noting that this study focuses on comparing RIL risks from different treatments for the same individual patient, rather than associating RIL with a specific treatment for prospective patients and analyzing a large cohort of patients. Studies like this can generate hypotheses for clinical trials for investigating the RIL risks associate with different radiation modalities and competing treatment plans. Although simple models, such as those used in this study, are entirely based on radiation dose and their accuracy in predicting lymphocyte changes may be limited, other recent studies have shown that these models (eg, effective dose to the circulating immune cells), could be clinically useful to evaluate susceptibility to severe lymphopenia for esophageal cancer patients [29, 30].

In particular, the limitation of the piecewise linear model is owing to the assumption that lymphocyte distribution throughout the treatment field is homogeneous. This is not the case in which organs (eg, spleen in the upper abdomen) may have a concentration of lymphocytes, and may have a greater influence on lymphocyte depletion than the body as a whole. Several studies have investigated blood flow models to estimate the dose to the circulating blood during treatment for some cancer.
sites, such as the lung and brain [31, 32]. These models could be used to determine more precise dose to circulating lymphocytes as an input to piecewise linear model to better understand radiation-induced lymphopenia in future studies. Nevertheless, it is reassuring that our simple model has a good predictive power and potential clinical usefulness.

The exponential lymphocyte survival model built from fitting measured ALC data was also able to predict ALC depletion during treatment. However, it was less accurate than the piecewise linear model for predicting the ALC nadir. For example, Supplemental Figure S6 shows the comparison of measured and predicted ALC nadirs based on the linear and exponential methods for all PSPT patients. Note that predictions were calculated for 28 fractions (5.6 weeks), but the measured nadirs only were from 4 to 6 weeks among patients. Two possible causes may be (1) the uncertainty in ALC measurements, especially near the end of the treatment course, when the ALC values can be very low, as a result of the rounding errors (ALK measurements in clinical laboratory reports are rounded to the nearest 100 cells/μL); and (2) the variability in time between radiation treatment fraction delivery and blood draw for ALC measurement. For example, spurious increases in ALC were seen for patients 2, 4, and 7 (see Supplemental Figure S4). To mitigate the sensitivity of this model to uncertain ALC data, it is crucial to ensure the accuracy and consistency of ALC measurement practice in future studies. In addition, to improve the predictive power, we will investigate approaches to incorporate preclinical predictors into the model in our future work, such as patient age and body mass index [33].

Although it is not clear that the exponential fitting approach will be of benefit in predicting ALC for new patients, as it requires large data sets, this approach is useful in comparative studies of different dose patterns for individual patients, such as in the present work. In addition, a recent study by Ellsworth et al [26] showed that ALC loss kinetics early in the course of treatment are strongly correlated with the extent of ALC depletion during radiation therapy and can be used to identify patients at high risk of developing severe radiation-induced lymphopenia. By only fitting the ALC data in initial weeks of treatment (of the first few fractions), one could estimate of ALC nadir early in treatment course based on the dose distribution and the consideration of individual patients’ lymphocyte sensitivity and make midcourse correction with adaptive plans if and when needed.

This study also motivates further studies to investigate the clinical factors that affect RIL risk of different radiation modalities. With help of research on continuing better understanding of lymphocyte distribution throughout the treatment field, radiation dose could be optimized accordingly to avoid lymphocyte killing. For example, IMPT and IMRT plans in this study were optimized using the same conventional dosimetric criteria as the PSPT plans. Additional immune sparing could be possible by optimizing plans with constraints on dose received by volumes of the body (and immune organs at risk, such as the spleen,
heart, etc.), which is most promising for IMPT due to its high complexity and flexibility in modulation. Such methods to enhance the ability of IMPT to minimize lymphopenia risk but without compromising tumor coverage and other normal tissues at risk will be studied in our future work.

Conclusions

This treatment planning study assessed RIL risk and the impact of different dose distributions of IMRT, PSPT, and IMPT on ALCs for 10 esophageal cancer patients. Two methods were proposed to estimate posttreatment ALC, and their reliability in predicting ALC were validated in separate patient groups for each treatment modality. Results from both approaches showed significant lymphocyte reduction associated with treatment. Proton plans showed a lower risk of lymphopenia after the treatment course than did photon plans, and IMPT plans outperformed PSPT plans in terms of lymphocyte preservation.

ADDITIONAL INFORMATION AND DECLARATIONS

Conflicts of Interest: The authors have no conflicts of interest to disclose.

Acknowledgments: The authors thank Dr Amy Ninetto in the Scientific Publications Department at MD Anderson for editing this manuscript.

Funding: This work was supported in part by the National Cancer Institute of the National Institutes of Health (2U19CA021239-35).

Ethical approval: All patient data has been collected under internal review board–approved protocol.

References

1. Sellins KS, Cohen JJ. Gene induction by gamma-irradiation leads to DNA fragmentation in lymphocytes. *J Immunol*. 1987; 139:3199–206.
2. Shiraishi Y, Fang P, Xu C, Song J, Krishnan S, Koay EJ, Mehran RJ, Hofstetter WL, Blum-Murphy M, Ajani JA, Komaki R, Minsky B, Mohan R, Hsu CC, Hobbs BP, Lin SH. Severe lymphopenia during neoadjuvant chemoradiation for esophageal cancer: a propensity matched analysis of the relative risk of proton versus photon-based radiation therapy. *Radiother Oncol*. 2018;128:154–60.
3. Holub K, Vargas A, Biete A. Radiation-induced lymphopenia: the main aspects to consider in immunotherapy trials for endometrial and cervical cancer patients. *Clin Transl Oncol*. 2020;22:2040–8.
4. Ellsworth SG. Field size effects on the risk and severity of treatment-induced lymphopenia in patients undergoing radiation therapy for solid tumors. *Adv Radiat Oncol*. 2018;3:512–9.
5. Campian JL, Ye X, Brock M, Grossman SA. Treatment-related lymphopenia in patients with stage III non-small-cell lung cancer. *Cancer Invest*. 2013;31:183–8.
6. Grossman SA, Ye X, Lesser G, Sloan A, Carraway H, Desideri S, Piantadosi S. Immunosuppression in patients with high-grade gliomas treated with radiation and temozolomide. *Clin Cancer Res*. 2011;17:5473–80.
7. Venkatesulu BP, Mallick S, Lin SH, Krishnan S. A systematic review of the influence of radiation-induced lymphopenia on survival outcomes in solid tumors. *Crit Rev Oncol Hematol*. 2018;123:42–51.
8. Cho O, Chun M, Chang S-J, Oh Y-T, Noh OK. Prognostic value of severe lymphopenia during pelvic concurrent chemoradiotherapy in cervical cancer. *Anticancer Res*. 2016;36:3541–7.
9. Wu ES, Oduyebo T, Cobb LP, Cholakian D, Kong X, Fader AN, Levinson KL, Tanner EJ, Stone RL, Piotrowski A, Grossman S, Long Roche K. Lymphopenia and its association with survival in patients with locally advanced cervical cancer. *Gynecol Oncol*. 2016;140:76–82.
10. Balmanoukian A, Ye X, Herman J, Laheru D, Grossman SA. The association between treatment-related lymphopenia and survival in newly diagnosed patients with resected adenocarcinoma of the pancreas. *Cancer Invest*. 2012;30:571–6.
11. Wild AT, Ye X, Ellsworth SG, Smith JA, Narang AK, Garrq T, Campian J, Laheru DA, Zheng L, Wolfgang CL, Tran PT, Grossman SA, Herman JM. The association between chemoradiation-related lymphopenia and clinical outcomes in patients with locally advanced pancreatic adenocarcinoma. *Am J Clin Oncol*. 2015;38:259–65.
12. Campian JL, Ye X, Sarai G, Herman J, Grossman SA. Severe treatment-related lymphopenia in patients with newly diagnosed rectal cancer. *Cancer Invest*. 2018;36:356–61.

13. Tang C, Liao Z, Gomez D, Levy L, Zhuang Y, Gebremichael RA, Hong DS, Komaki R, Welsh JW. Lymphopenia association with gross tumor volume and lung V5 and its effects on non-small cell lung cancer patient outcomes. *Int J Radiat Oncol Biol Phys*. 2014;89:1084–91.

14. Davuluri R, Jiang W, Fang P, Xu C, Komaki R, Gomez DR, Welsh J, Cox JD, Crane CH, Hsu CC, Lin SH. Lymphocyte nadir and esophageal cancer survival outcomes after chemoradiation therapy. *Int J Radiat Oncol Biol Phys*. 2017;99:128–35.

15. Cho Y, Park S, Byun HK, Lee CG, Cho J, Hong MH, Kim HR, Cho BC, Kim S, Park J, Yoon HL. Impact of treatment-related lymphopenia on immunotherapy for advanced non-small cell lung cancer. *Int J Radiat Oncol*. 2019;105:1065–73.

16. Wild AT, Herman JM, Dholakia AS, Moningi S, Lu Y, Rosati LM, Hacker-Prietz A, Assadi RK, Saeed AM, Pawlik TM, Jaffee EM, Laheru DA, Tran PT, Weiss MJ, Wolfgang CL, Ford E, Grossman SA, Ye X, Ellsworth SG. Lymphocytesparing effect of stereotactic body radiation therapy in patients with unresectable pancreatic cancer. *Int J Radiat Oncol*. 2016;94:571–9.

17. Blanchard P, Garden A, Gunn G, Rosenthal D, Morrison W, Hernandez M, Crutison J, Lee J, Ye R, Fuller C, Mohamed A, Hutcheson K, Holliday E, Thaker N, Sturgis E, Kies M, Zhu X, Mohan R, Frank S. Intensity modulated proton beam therapy (IMPT) versus Intensity modulated proton therapy (IMRT) for oropharynx cancer patients - a case matched analysis. *Radiother Oncol*. 2016;120:48–55.

18. Moreno AC, Frank SJ, Garden AS, Rosenthal DI, Fuller CD, Gunn GB, Reddy JP, Morrison WH, Williamson TD, Holliday EB, Phan J, Blanchard P. Intensity modulated proton therapy (IMPT) - the future of IMRT for head and neck cancer. *Oral Oncol*. 2019;88:66–74.

19. Moeller DS, Nordsmark M, Nyeng TB, Alber M, Hoffmann L. PO-0958: anatomical changes in oesophageal cancer patients: posterior beam IMPT is more robust than IMRT. *Radiother Oncol*. 2018;127:S525–6.

20. Routman DM, Garant A, Lester SC, Day CN, Harmsen WS, Sanhueza CT, Yoon HH, Neben-Wittich MA, Martenson JA, Haddock MG, Hallemier CL, Merrell KW. A comparison of grade 4 lymphopenia with proton versus photon radiation therapy for esophageal cancer. *Adv Radiat Oncol*. 2019;4:63–9.

21. Yovino S, Kleinberg L, Grossman SA, Narayanan M, Ford E. The etiology of treatment-related lymphopenia in patients with malignant gliomas: modeling radiation dose to circulating lymphocytes explains clinical observations and suggests methods of modifying the impact of radiation on immune cells. *Cancer Invest*. 2013;31:140–4.

22. Nakamura N, Kusunoki Y, Akiyama M. Radiosensitivity of CD4 or CD8 positive human T-lymphocytes by an in vitro colony formation assay. *Radiat Res*. 1990;123:224–7.

23. Cameron JR, Skofronick JG, Grant RM, Morin RL. Physics of the body. *Med Phys*. 2000;27:425–425.

24. Farese AM, Hankey KG, Cohen MV, MacVittie TJ. Lymphoid and myeloid recovery in rhesus macaques following total body x-irradiation. *Health Phys*. 2015;109:414–26.

25. Goans RE, Holloway EC, Berger ME, Ricks RC. Early dose assessment in criticality accidents. *Health Phys*. 2001;81:446–9.

26. Ellsworth SG, Yalamanchili A, Zhang H, Grossman SA, Hobbs R, Jin J-Y. Comprehensive analysis of the kinetics of radiation-induced lymphocyte loss in patients treated with external beam radiation therapy. *Radiat Res*. 2020;193:73–81.

27. Wieser H-P, Cisternas E, Wahl N, Ulrich S, Stadler A, Mescher H, Müller, L-R, Klinge T, Gabrys H, Burigo L, Mairani A, Ecker S, Ackermann B, Ellerbrock M, Parodi K, Jäkel O, Bangert M. Development of the open-source dose calculation and optimization toolkit MatRad. *Med Phys*. 2017;44:2556–68.

28. Paganetti H, Niemierko A, Ancukiewicz M, Gerweck LE, Goitein M, Loeffler JS, Suit HD. Relative biological effectiveness (RBE) values for proton beam therapy *Int J Radiat Biol*. 2002;53:407–21.

29. So TH, Chan SK, Chan WL, Choi H, Chiang CL, Lee V, Lam TC, Wong I, Law S, Kwong D, Kong F-M, Jin JY, Lam KO. Lymphopenia and radiation dose to circulating lymphocytes with neoadjuvant chemoradiation in esophageal squamous cell carcinoma. *Adv Radiat Oncol*. 2020;5:880–8.

30. Xu C, Jin J-Y, Zhang M, Liu A, Wang Jun, Mohan R, Kong F-M, Lin SH. The impact of the effective dose to immune cells on lymphopenia and survival of esophageal cancer after chemoradiotherapy. *Radiother Oncol*. 2020;146:180–6.
31. Hammi A, Paganetti H, Grassberger C. 4D blood flow model for dose calculation to circulating blood and lymphocytes. *Phys Med Biol*. 2020;65:055008.

32. Jin J-Y, Mereniuk T, Yalamanchali A, Wang W, Machtay M, Kong F-M, Ellsworth S. A framework for modeling radiation induced lymphopenia in radiotherapy. *Radiother Oncol*. 2020;144:105–13.

33. van Rossum PSN, Deng W, Routman DM, Liu AY, Xu C, Shiraishi Y, Peters M, Merrell KW, Hallemeier C, Mohan R, Lin SH. Prediction of severe lymphopenia during chemoradiation therapy for esophageal cancer: development and validation of a pretreatment nomogram. *Pract Radiat Oncol*. 2020;10:e16–26.