**Research paper**

**Plasmodium species mixed infections in two areas of Manhiça District, Mozambique**

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We compared the distribution patterns of individual *Plasmodium* species and mixed-species infections in two geographically close endemic areas, but showing environmental differences. Comparisons concerned circulating *Plasmodium* infections in both human and mosquito vector populations in the dry and wet seasons, at a micro-epidemiological level (households). Both areas revealed a very high overall prevalence of infection, all year-round and in all age groups. *Plasmodium falciparum* was the predominant species, being found in the vast majority of infected individuals regardless of the presence of other species. *Plasmodium malariae* and *Plasmodium ovale* occurred almost exclusively in mixed infections. Seasonal variation in *P. malariae* prevalence was observed in one area but not in the other. A decrease in *P. malariae* prevalence concurred with a marked increase of *P. falciparum* prevalence. However this was strongly dependent on age and when analysing infections at the individual level, a different pattern between co-infecting species was unveiled. Regarding transmission patterns, in both areas, *P. falciparum* gametocyte predominated in single infections regardless of age and *P. malariae* gametocyte carriage increased when its overall prevalence decreased.

**Key words: Plasmodium, mixed infections, transmission, Mozambique**

1. **Introduction**

In natural parasite populations of *Plasmodium* interspecific (mixed) infections commonly occur. However as pointed out very recently by several authors [1, 2, 3] there has been only sporadic interest in this subject.

In clinical, microscopically-based diagnostics, mixed malaria infections are often missed but sensitive PCR-based techniques [4, 5] and/or longitudinal studies have demonstrated that mixed infections are remarkably common [6, 7]. Experimental studies with humans [reviewed in 8 and 9] and rodents [10, 11] indicated that different parasite species seem to interact, affecting mortality, pathology and infection dynamics. Effects of mixed infections on pathology were also recorded in field studies, namely attenuation of *falciparum* disease severity [12, 13, 14, 15].

When prevalence of mixed *Plasmodium* infections in humans is analysed in endemic areas, the pair *falciparum*- *vivax* seems to be under-represented [13, 16, 17]. In contrast, an excess of mixed infections is often correlated with the pair *falciparum*- *malariae* [6, 18, 19, 20]. Also, seasonal variation between individual species prevalence was observed between *P. falciparum* and *P. vivax* in Vanuatu [13, 21] and between *P. falciparum* and *P. malariae* in Nigeria [6] and Burkina Faso [22]. However, geographical differences in these distribution patterns have also been reported [8]. Whether this alternation really occurs and has a biological meaning or if it is due to a sampling artefact remains controversial.

In previous studies carried out in Guinea-Bissau [4, 20], we observed that households were likely to work as individual units of transmission. In addition, different associations of *Plasmodium* species seemed to occur in infections detected in the human host or in the mosquito vector [20].

In this context, the main aim of this study was to determine whether different patterns of mixed infections might be affected by even minor differences in environmental conditions. For the purpose, we compared prevalence of different *Plasmodium* species and mixed-infections in two very close areas, but with slightly distinct environmental characteristics. We also analysed infections in both human and mosquito vector populations at the household level in order to determine which parasite populations were actually being transmitted between the two hosts and to better assess possible differences in the transmission of the several *Plasmodium* species. Further, the same households were sampled in the dry and wet seasons to assess possible effects of different transmission levels on parasite transmission among vertebrate hosts and mosquito vectors.

2. **Materials and Methods**

**Study area**

The study took place in two areas of the Manhiça district, in southern Mozambique, east Africa - Manchiana and Ilha Josina. Both areas were subsequently included in an efficacy trial of a malaria vaccine candidate [23]. In both, climate is characterised by a hot and wet season (from October to May) and a dry and cold season (from June to September). Malaria is endemic with perennial transmission and marked seasonality. The average entomological inoculation rate in the district was estimated to be 15 infectious bites per person per year in 1998 [Aranda in 24]. *Anopheles funestus sensu lato* and *Anopheles gambiae sensu lato* are the main malaria vectors of the region [25].

Manchiana is a flat savannah area with moderate vegetation located approximately 10 km north of the
village of Manhiça. This sandy area has two main permanent mosquito-breeding sites located on the east and west borders and most of the population lives about 1Km away from them.

Ilha Josina is an island in the Incomati River, which crosses the district from north to south. It is a marshy area located about 40 km northeast from Manchiana. The access is made through a small bridge or boats. There are local abundant mosquito breeding sites in addition to the surrounding river and abundant vegetation.

In both areas, there are two health facilities offering basic services and an elementary mother and child health care unit. Population is mainly made up of farmers growing maize and beans and also selling reeds in Ilha Josina. Housing conditions are generally poor and typically made of reed with thatched or corrugated roofs.

**Sampling**

The investigation was approved by the Ethical Committee of the Faculty of Medicine, Eduardo Mondlane University, Maputo. The local village chief and each person (or parent) was informed of the nature and aims of the study and told that participation was voluntary. An informed consent was obtained from each household chief.

In both areas, sampling took place in the dry (August 2001) and wet (February 2002) seasons. In each season, collections were made in a total of 12 households in Manchiana (10 households surveyed in both seasons) and in a total of 13 households in Ilha Josina (11 households surveyed in both seasons). All houses were mapped and their inhabitants fully identified (including the room in which they slept). In each household, blood fed resting *Anopheles* females were collected in the morning (6.00-7.00 am), followed by blood samples collection (fingerprick) from all inhabitants, for microscopic detection of gametocytes and for PCR analysis.

Assuming that the extrinsic incubation period at an average temperature of 28°C is of 9-10 days for *P. falciparum*, 14-16 days for *P. malariae* and 12-14 days for *P. ovale* [26], mosquitoes were kept in paper-cups corresponding to each house/room during 8 days with glucose *ad libitum*. This should allow the development of oocysts from all species, resulting from infections acquired the night prior to collection. After this period dissections were performed to obtain midguts and salivary glands for PCR analysis. We assumed that parasites found in salivary glands were the result of previously acquired infections.

**Identification of Plasmodium species**

Detection of malaria infection and identification of *Plasmodium* species, both in blood samples and in mosquitoes, was carried out by nested-PCR amplification of the small subunit ribosomal RNA genes [5]. Parasite densities were determined in blood smears and asexual and sexual forms were recorded separately. Smears were Giemsa-stained and parasite densities were recorded as the number of parasites/µl of blood, assuming an average leukocyte count of 8000/µl (all smears were examined against 500 leukocytes prior to being declared negative). *Plasmodium falciparum* (F) and *P. malariae* (M) asexual parasitaemias were divided into groups according to densities: Low (F and M <80 parasites/µl); Moderate (F 80-3500 parasites/µl and M 80-650 parasites/µl); High (F >3500 parasites/µl and M >650 parasites/µl). Infections detected only by PCR were grouped as Sub-Patent (SP).

**Identification of mosquito species**

Species identification of the members of the *An. gambiae* and *An. funestus* complexes was performed by PCR according to previously described protocols [27, 28]. One microlitre of dilutions of 1:25 in ddH2O of each DNA sample extracted from individual mosquitoes was used as DNA template in separate PCR reactions.

**Statistical analysis**

Since collections were carried out in households (clusters) which involves a higher variability than random sampling [29], overall prevalence in each area and season and their confidence intervals were determined considering global and cluster variance. Design effects were also determined. Since these had values around 1.0, meaning that sampling was approximate to simple random sampling, other proportions were determined considering that data arose from random sampling. Comparison of prevalences was made using the Pearson χ² test. A significance level of 5% was considered. Calculations were made using Epi Info Version 6.04d and Statistica 5.0 for Windows.

### 3. Results

A total of 308 blood samples were collected from 213 asymptomatic people in both areas and seasons (70 people from Manchiana in each season; 81 and 87 people from Ilha Josina in the dry and wet season, respectively). Age of the population sampled varied between 1 and 82 years-old with nearly 55% being less than 14 years-old.

A total of 1342 mosquitoes were collected and kept for sporogenic development: 423 and 112 mosquitoes in Manchiana (dry and wet season, respectively), and 269 and 538 mosquitoes in Ilha Josina (dry and wet season, respectively).

**Table 1 – Prevalence of infection in Manchiana and Ilha Josina in the dry and wet seasons.**

| Area          | Manchiana | Ilha Josina |
|---------------|-----------|-------------|
| Season        | Dry       | Wet         | Dry       | Wet       |
|               | n = 70    | n = 70      | n = 81    | n = 87    |
| Overall       | 71 (52-91) | 80 (67-93)  | 91 (83-100) | 91 (85-96) |
| Single infections |         |             |           |           |
| F             | 60 63 51 69 |             |           |           |
| M             | 0 1 1 0   |             |           |           |
| Mixed infections |       |             |           |           |
| F+M           | 9 14 26 21 |             |           |           |
| F+O           | 0 1 9 0   |             |           |           |
| F+M+O         | 3 0 5 1   |             |           |           |
| Overall infection |   |             |           |           |
| F             | 71 79 90 91 |             |           |           |
| Overall infection M | |             |           |           |
| O             | 11 16 32 22 |             |           |           |
| Overall infection O | |             |           |           |
|               | 3 1 14 1   |             |           |           |

n: sample size; F: *P. falciparum*; M: *P. malariae*; O: *P. ovale*; F+M: mixed infection by *P. falciparum* and *P. malariae*; F+O: mixed infection by *P. falciparum* and *P. ovale*; F+M+O: mixed infection by *P. falciparum*, *P. malariae* and *P. ovale*. Ninety-five per cent Confidence Intervals of overall prevalence in square brackets (see Materials and Methods).
Infection in the human population

As expected, PCR analysis consistently revealed higher infection rates than optical microscopy (OM). This was especially evident for mixed infections (PCR versus OM - Manchiana: dry season 11% vs 4%; wet season 16% vs 10%; Ilha Josina: dry season 40% vs 7%; wet season 22% vs 14%) and overall prevalence of species other than P. falciparum (e.g. P. ovale was almost only detected by PCR).

In Manchiana the overall prevalence (determined by PCR) increased from 71% in the dry season to 80% in the wet season (Table 1). Plasmodium falciparum was the predominant species occurring in 100% and 98% of the infected population, either in single or mixed infections.

Prevalence of species other than P. falciparum and, consequently, of mixed infections also increased from 16% to 20% in the dry and wet seasons, respectively (Figure 1).

In Ilha Josina, an overall prevalence of 91% was maintained from one season to another (Table 1), but prevalence of mixed infections significantly decreased ($\chi^2=6.334, p=0.012$) in the wet season (43% in the dry and 24% in the wet season) (Figure 1). This was concomitant with lower P. malariae and P. ovale prevalences in the wet season (P. malariae: 32% and 22%, P. ovale: 14% and 1%, in the dry and wet seasons, respectively) (Table 1).

Overall prevalence of infection was high in all age groups. Among infected people in both areas and seasons, mixed infections were more prevalent in children less than 14 years old. From the dry to the wet season, a considerable decrease in prevalence of mixed infections occurred among adults (>14 years-old), being significantly different from the one observed in children (≤14 years-old) in Ilha Josina ($\chi^2=8.421, p=0.004$). In Manchiana, although not statistical significant, differences between age groups were still evident ($\chi^2=3.09, p=0.079$) (Figure 1).

Densities of P. falciparum asexual forms were higher in Ilha Josina than in Manchiana and decreased in the wet season in both areas (geometric mean and median - Manchiana: dry season 858.37 and 619.97 parasites/µl blood; wet season 256.06 and 136.68 parasites/µl blood;
Ilha Josina: dry season 1040.80 and 694.65 parasites/µl blood; wet season 467.92 and 426.04 parasites/µl blood.

In Manchiana this decrease was coincident with a lower prevalence of Moderate and High parasite densities and a higher prevalence of Low (and Sub-Patent) densities (Figure 2). However, in Ilha Josina, the prevalence of Low-density infections also increased but prevalence of Moderate and High densities remained unaltered. In both areas and both seasons, *P. falciparum* Moderate and High parasite densities predominated in children while Sub-Patent and Low densities predominated in adults (Figure 2).

**Figure 2** – Distribution of *P. falciparum* density groups (Neg: non-infected, SP: sub-patent, L: low, M: moderate, H: high) by age groups (1-5 years old: black bars; 6-14 years old: grey bars; > 14 years old: white bars) in both areas and both seasons. Prevalence of each density group in the total population is shown below.

Densities of asexual *P. malariae* were recorded in a very small number of samples but also seemed to decrease in the wet season (Table 2). However, patency increased in both areas (patent infections, considering the total number of *P. malariae* infections: Manchiana - 38% in the dry and 73% in the wet season; Ilha Josina - 23% in the dry and 68% in the wet season). Only one patent *P. ovale* infection was recorded (63 parasites/µl blood), in Ilha Josina, in the dry season.

**Table 2** Asexual forms densities (parasites/µl blood) and distribution of *P. malariae* density groups in both areas and both seasons.

| Area       | Density groups by age group | Dry season | Wet season |
|------------|-----------------------------|------------|------------|
|            | 1-5 years old               |            |            |
| Manchiana  | 7Neg, 2SP                   | 2Neg, 1L, 4M, 1H |
|            | 1SP, 2M, 1H                 | 2Neg, 1SP, 1L, 1M |
|            | 13Neg, 2SP                  | 10Neg, 2SP |
|            | Geometric mean              | 448.37 (n=3) | 225.44 (n=8) |
|            | Median                      | 285.71 (n=3) | 339.58 (n=8) |
| Ilha Josina| 2M, 1H                      | 1Neg, 2L, 3M, 4H |
|            | 2Neg, 9SP, 1L               | 2Neg, 4SP, 1L, 3M |
|            | 5Neg, 11SP, 1L, 1M          | 5Neg, 2SP |
|            | Geometric mean              | 391.25 (n=6) | 270.55 (n=13) |
|            | Median                      | 238.50 (n=6) | 334.00 (n=13) |

n: sample size; Neg: non-infected, SP: sub-patent, L: low, M: moderate, H: high
In mixed infections, for both areas and seasons, Moderate and High \( P. \) \( malariae \) densities occurred in individuals with Moderate and High \( P. \) \( falciparum \) densities (only two cases with Low and one case with Sub-Patent \( P. \) \( falciparum \) densities). All but one of these individuals were aged between 1 and 14 years-old. Similarly, \( P. \) \( ovale \) infections occurred mostly in children with Moderate and High \( P. \) \( falciparum \) parasitaemias.

Overall prevalence of gametocyte-carriers increased in the wet season in both areas (Figure 3). In Manchiana this increase reflects the increase of \( P. \) \( falciparum \) gametocyte prevalence while in Ilha Josina it reflects the increase of \( P. \) \( malariae \) gametocyte prevalence. No \( P. \) \( ovale \) gametocytes were detected. Gametocyte prevalence predominated in children, except in Ilha Josina, in the dry season (≤14 \textit{versus} >14 – Manchiana: dry season 36\% vs 0\%; wet season 53\% vs 21\%; Ilha Josina: dry season 18\% vs 31\%; wet season 48\% vs 8\%). \( P. \) \( falciparum \) gametocytes seemed to predominate in single infections, even after adjusting for age (gametocyte prevalence in single \textit{versus} mixed infections – Manchiana: dry season 75\% vs 25\%; wet season 71\% vs 29\%; Ilha Josina: wet season 69\% vs 31\%), except in Ilha Josina, in the dry season (33\% vs 67\%).

Figure 3 – Prevalence of gametocyte-carriers of each \( Plasmodium \) species (\( F – P. \) \( falciparum \); \( M – P. \) \( malariae \); without – prevalence of non-gametocyte carriers). Only population with patent parasitaemia was considered (Manchiana-dry season: \( P. \) \( falciparum \) – \( n=44 \); \( P. \) \( malariae \) – \( n=3 \); Manchiana-wet season: \( P. \) \( falciparum \) – \( n=43 \); \( P. \) \( malariae \) – \( n=8 \); Ilha Josina-dry season: \( P. \) \( falciparum \) – \( n=54 \); \( P. \) \( malariae \) – \( n=6 \); Ilha Josina-wet season: \( P. \) \( falciparum \) – \( n=66 \); \( P. \) \( malariae \) – \( n=13 \)).

Infection in the mosquito population

A sub-sample of 122 mosquitoes was identified to species by PCR. \( Anopheles \) \( funestus \) \textit{sensu stricto} was the predominant species, accounting for 116 (95\%) of the identifications. \( Anopheles \) \( arabiensis \) was the other species identified, but only in 6 specimens (5\%) collected from Manchiana in the wet season.

A total of 1337 midguts and 1341 salivary glands were analysed for the two seasons and study areas.

In Manchiana, for the dry season, midgut and salivary glands infection rates of 8\% (33/423, 95\% CI 4-12) and 10\% (43/423, 95\% CI 4-18), respectively, were determined. In the wet season, midgut infection rates were 9\% (10/110, 95\% CI -7-26) and 7\% (70\%) of the infected mosquitoes were collected from the same house. An infection rate of 1\% (1/112, 95\% CI -2-4) was determined in salivary glands.

In Ilha Josina, in the dry season, a midgut and salivary glands infection rate of 12\% (33/266, 95\% CI -9-34; with 23 of the infected mosquitoes were collected in only two houses) and 6\% (16/269, 95\% CI -7-19; with 15 of the infected mosquitoes were collected in a single house), respectively, were determined. In the wet season, midguts and salivary glands infection rates of 5\% (26/538, 95\% CI 2-7) and 4\% (23/537, 95\% CI -0.6-9; with 13 of the infected mosquitoes collected from the same house) respectively, were determined.

Regardless of site or season, \( P. \) \( falciparum \) single infections were highly predominant (94\%, 174/185). Other \( Plasmodium \) species single infections (4 \( P. \) \( malariae \) and 1 \( P. \) \( ovale \)) were found in midguts only from Manchiana, in the dry season. Mixed infections between \( P. \) \( falciparum \) and \( P. \) \( malariae \) were also detected in just a few midguts: 2 in Manchiana in the dry season and 4 in the Ilha Josina in the wet season.

4. Discussion

In the great majority of occasions, malaria control relies mainly on general measures that often fail to consider micro-epidemiological features that may influence its outcome. In this context, it may be particularly important to assess the prevalence of certain genotypes or mixed-species infections before control measures are implemented.

In light of this, the present study aimed to assess whether minor micro-epidemiological differences could relate to differential distribution of malaria parasites and we saw that seasonal variation in \( P. \) \( malariae \) prevalence was observed in one area but not in the other. A decrease in \( P. \) \( malariae \) prevalence concurred with a marked increase of \( P. \) \( falciparum \) prevalence but this was strongly dependent on age. Further, an increase of \( P. \) \( malariae \) gametocyte carriage was observed when its overall prevalence decreased. In both areas \( P. \) \( falciparum \) gametocytes predominated in single infections regardless of age.

The overall prevalence of infection was verified to be very high in both studied areas and was detected all year-round. In Ilha Josina, a marshy area with abundant
mosquito breeding sites, both higher infection prevalence and parasite densities occurred, which reflect conditions more prone to intense transmission. In this area, the prevalence of *P. malariae* and *P. ovale* (especially the latter) markedly decreased in the wet season while prevalence of single *P. falciparum* infections increased. This strongly suggests that an alternation in the relative contribution of each species to the total parasite indexes occurred in this area. This alternation is neither attributable to differences in vector behaviour since the same mosquito species were found in both areas, nor to drug use or access to health facilities since both areas have similar characteristics regarding these factors.

Previous studies [8, 30] have suggested that *P. malariae*, which is able to develop chronic infections, can be found at higher prevalences when transmission and overall infection rates are lower and it is alleviated from the suppressive effect of *P. falciparum*. A similar phenomenon may explain the increase in *P. ovale* from the wet to the dry season since this parasite is able to relapse after long periods of dormancy in the liver [31]. In Manchiana, a sandy area with fewer mosquito breeding sites, where transmission and prevalence of infection are lower, the establishment of *P. malariae* infections may be facilitated, while in Ilha Josina, the high transmission during the wet season favours *P. falciparum*, which transmits more rapidly and suppresses *P. malariae* to undetectable levels. However, the decrease in the prevalence of mixed infections in Ilha Josina was observed mainly in adults, strongly supporting the notion that suppression of other *Plasmodium* species by *P. falciparum* only occurs when a certain level of immunity is reached and is able to control both species parasitaemia [6]. This is corroborated by the fact that, in mixed infections, high parasitaemias of one species occurred almost exclusively in individuals, mainly children, who also presented high parasitaemias of the other species.

A more comprehensive analysis of infections in mosquitoes collected in the same households could have helped to determine which parasite populations were actually being transmitted between the two host populations and to better assess possible differences in the transmission of the several *Plasmodium* species. Unfortunately, species other than *P. falciparum* were found at very low prevalence in mosquitoes thus precluding any comparisons. Moreover, most of the data derives from one or two houses only which could have biased the data on mosquito infection rates. Alternatively, transmission may be assessed through the analysis of gametocyte-carriage. This analysis limits the amount of conclusive information since gametocyte-patency may not correlate directly to mosquito infections [20] and the detection threshold of OM seriously reduces the sample size. Sensitive molecular tools are only available for the detection of *P. falciparum* gametocytes [32, 33] but not the other species. Nonetheless, some interesting observations were done.

In Ilha Josina in the wet season, both the *P. malariae* patent parasitaemia and the proportion of *P. malariae* gametocyte-carriers increased, despite the reduction in *P. malariae* prevalence, which may reflect an enhancement of transmissibility due to the increased pressure exerted by *P. falciparum*. Once more this is only evident in individuals under 14 years-old, the most susceptible portion of population.

McKenzie et al. [34], following analysis of records obtained from adults sequentially or simultaneously inoculated with both species, reported that prior or concurrent *P. malariae* infection enhanced production of *P. falciparum* gametocytes. In contrast, Price et al. [35] reported that mixed *P. falciparum* + *P. vivax* infections among Thai individuals are associated with a reduction in the prevalence of *P. falciparum* gametocytes. In agreement with the latter report, in the present study we observed that *P. falciparum* gametocytes seemed more likely to be present in single than in mixed infections and that this was independent of individuals age. Collectively, the implications of these observations may have a profound repercussion in the outcome of disease control. If the simultaneous infection by other species inhibits *falciparum*-gametocyte production and if control measures affect and reduce more effectively *Plasmodium* species other than *P. falciparum*, this inhibitory effect may disappear.

The results of the present work suggest that slightly distinct environmentally characteristics affect transmission patterns, and may have an effect in the way different human malaria species establish in the human population. The relevance of these data should be taken into account when general control strategies are planned. Control interventions may alter the proportions of parasite populations in a given region, which could have consequences on the acquisition of protective immunity or malaria pathology.

In spite of the microepidemiological approach, the complexity of the three biological malaria entities (parasite, human and mosquito) raises questions that are difficult to address in a field setting. The use of experimental models in which some of the variables can be controlled would benefit future studies in humans.

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**Conflict of interest**

The authors have declared that no conflict of interest exists.

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