Over the past decade, numerous publications have stressed the importance of IFN-I in the pathogenesis of systemic lupus erythematosus (SLE), but their assays typically used microarrays and/or quantitative PCR (qPCR) and thus appear to be laborious and not well suited for use in clinical practice. In the previous issue of *Arthritis Research & Therapy*, Li and colleagues [1] have investigated a much simpler methodology to measure the level of activation of IFN-I in patients with SLE. They measured the expression of CD64 (FcγRI) by flow cytometry on monocytes and demonstrated high levels in SLE compared with healthy controls. The authors also showed that CD64 levels correlated with IFN-stimulated gene (ISG) expression (by qPCR) and disease activity (by Systemic Lupus Erythematosus Disease Activity Index (SLEDAI)) and that CD64 was downregulated in four patients who received pulse methylprednisolone therapy. As expected, CD64 was induced in normal monocytes by IFN-I in vitro. Similar findings have been obtained in SLE monocytes by flow cytometry in another study focused on sialoadhesin (Siglec-1 or CD169), although this assay required indirect immunofluorescence staining [2]. Both these molecules appear promising as biomarkers of IFN-I activation in SLE.

The clinical significance of IFN-I pathway activation in SLE is multifaceted. First, the pathway has been implicated in the pathogenesis of the disease, and therefore targeted therapies against IFN-I are currently in clinical trials. Second, IFN-I activation may identify a subset of SLE patients with potential diagnostic, prognostic and therapeutic implications. Third, change in IFN-I activity levels may reflect change in disease activity and thus help clinical management of the disease. In this context, the data by Li and colleagues on CD64 expression have clinical relevance as they might facilitate research in all of the above areas with a simple tool.

The implication of IFN-I in SLE pathogenesis comes from multiple pieces of evidence, including genetic, gene expression association studies, and induction of SLE by therapeutic administration of IFN-I [3]. The potential mechanisms by which IFN-I may promote inflammation and autoimmunity have been reviewed recently and include activation of immature dendritic cells and break of peripheral tolerance, augmentation of humoral immunity pathways with production of pathogenic antibodies, induction of Th1 cells, chemokine production, and priming of myeloid cells for enhanced responses to inflammatory stimuli [4]. The emerging evidence about the pathogenic role of IFN-I in SLE led to the recent introduction in clinical trials of several anti-IFN antibodies. This exciting new era of IFN-I targeting should greatly benefit from simple biomarkers of the pathway's activity, such as CD64 and CD169 expression. This would facilitate stratification of SLE patients according to IFN-I activation, as well as monitoring of the degree of IFN-I inhibition during therapy - adequate to suppress disease activity, but not excessive to cripple immunosurveillance.

Cross-sectional studies, including our own and the study by Li and colleagues, have shown that IFN-I activation in SLE is present in about half of adult patients and it is associated with disease activity, renal involvement, as well as autoantibodies to dsDNA, and RNA
Binding Proteins (anti-Ro, La, Sm, RNP) [1,5-7]. This group of patients might represent a different subclass of the disease where the IFN-I pathway is dominant and therefore its therapeutic targeting most beneficial. The design of those studies, however, leaves open the possibility that some patients may be positive for IFN only intermittently, especially during disease flares, and again negative after aggressive therapy, such as with pulse methylprednisolone [1,5]. More research needs to be done in this area before conclusions can be drawn. We believe that although SLE patients should be stratified in clinical trials of anti-IFN therapy, patients negative for IFN-I activation should not be excluded from the trials.

Two longitudinal studies failed to show ability of IFN-I activation to parallel acute changes of disease activity [6,7]. However, one of those studies used microarray data (which is less accurate than qPCR) for their ISG score [7] and both had only few patients with more than two visits. Moreover, IFN-regulated chemokine levels did parallel disease activity in a larger study [8]. Interestingly, high baseline levels of these chemokines substantially increased the risk for lupus flare, especially a renal one, in the next year [8]. Flares were also increased for patients with high baseline ISG scores, but in a more delayed manner [6]. In our experience, about 30 to 40% of patients, followed longitudinally for at least 2 years, demonstrate parallel courses of SLEDAI and ISG scores [9]. Based on the above studies, it appears that the matter has not been resolved yet, but it is possible that ISG scores work as biomarkers of disease activity only in a subgroup of patients, and not necessarily the ones with high baseline IFN-I activity. Other pathway signatures or a combination of those might be eventually required to evaluate all patients [10].

Although measurement of monocyte CD64 and CD169 expression promises, it is likely not specific for either IFN-I or SLE disease activity. Similar to other ISGs, both can be induced by viral infection and IFN-γ, whereas CD64 expression may also be induced by IL-10 [1,11,12]. Thus, at this point, none of the above gene measurements can be expected to substitute for clinical judgment to differentiate lupus flare from infection. Furthermore, before these new monocyte cell surface markers can fulfill their promise for ease and efficiency of IFN-I detection, they will need to be validated against currently used IFN-I molecular assays (especially qPCR) in carefully conducted large longitudinal prospective studies of SLE patients.

Abbreviations
IFN = interferon; ISG = IFN-stimulated gene; qPCR = quantitative PCR; SLE = systemic lupus erythematosus; SLEDAI = Systemic Lupus Erythematosus Disease Activity Index.

Competing interests
KAK has applied for a patent for an interferon assay. GDK declares that he has no competing interests.

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